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(54) Titre : COMPOSITIONS IMMUNOGENES POUR CHLAMYDIA PNEUNOMIAE
(54) Title: IMMUNOGENIC COMPOSITIONS FOR CHLAMYDIA PNEUNOMIAE

(57) **Abrégé/Abstract:**

The invention relates to polypeptides for use as an autotransporter antigen. The invention further relates to methods and uses of a polypeptide for an autotransporter function in preparation of a medicament for the prevention or treatment of a Chlamydia pneumoniae infection or for the preparation of an assay for the diagnosis of a Chlamydia pneumoniae infection in an individual. Also, a method is provided for raising an immune response in an individual by administering to the individual a polypeptide for use as an autotransporter antigen.



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(54) Title: IMMUNOGENIC COMPOSITIONS FOR CHLAMYDIA PNEUMONIAE

(57) Abstract: The invention relates to polypeptides for use as an autotransporter antigen. The invention further relates to methods and uses of a polypeptide for an autotransporter function in preparation of a medicament for the prevention or treatment of a Chlamydia pneumoniae infection or for the preparation of an assay for the diagnosis of a Chlamydia pneumoniae infection in an individual. Also, a method is provided for raising an immune response in an individual by administering to the individual a polypeptide for use as an autotransporter antigen.



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IMMUNOGENIC COMPOSITIONS FOR *CHLAMYDIA PNEUMONIAE*

This application claims the benefit of U.S. Provisional Application 60/542,832, filed March 2, 2004; U.S. Provisional Application 60/643,110, filed January 12, 2005; and
 5 U.S. Provisional Application 60/644,552, filed January 19, 2005, all of which are incorporated herein in their entireties.

All documents cited herein are incorporated by reference in their entirety.

10 Field

The invention is in the field of immunology and vaccinology. In particular, it relates to immunogenic compositions comprising combinations of immunogenic molecules from *Chlamydia pneumoniae*.

15 Background Art

The bacteria of the genus *Chlamydia* (and *Chlamydophila*, according to the recently proposed but still controversial re-classification of *Chlamydiaceae* (Bush *et al* (2001) Int J Syst Evol Microbiol 51: 203-20; Everett *et al* (1999) Int J Syst Bacteriol 49: Pt2 415-40; Schachter *et al* (2001) Int J Syst Evol Microbiol 51: 249, 251-3) are obligate
 20 intracellular parasites of eukaryotic cells, which have a unique biphasic life cycle involving two pleiomorphic developmental forms: an extracellular, metabolically inert, spore-like, infectious form (the elementary bodies, EBs) and an intracellular, non-infectious, replicative form (the reticulate bodies, RBs) which remains contained in a specialized cytoplasmic compartment (the *Chlamydial* inclusion). The EBs are
 25 responsible for the initial attachment to host cell surface and the establishment of the cytoplasmic inclusion where EBs can differentiate to RBs and thus initiate the replicative stage. Eventually RBs revert to infectious EB forms able to start new replicative cycles in neighbouring host cells.

As *Chlamydia* infection is an intracellular infection, the currently accepted paradigm is that effective anti-*Chlamydial* immunisation would require both an adequate T-cell response and high serum levels of neutralising antibodies and that "an ideal vaccine should induce long lasting (neutralising) antibodies and a cell mediated immunity that can quickly respond upon exposure to *Chlamydia*". Several sometimes contradictory
 35 studies have indicated that both CD4+ and CD 8 positive T cells have a role in *Chlamydial* clearance (Loomis and Starnback (2002) Curr Opin Microbiol 5: 87-91). Indeed, there now appears to be a prevailing consensus that specific CD4+ T cells and B cells are critical to the complete clearance of intracellular *Chlamydia* and for mediating recall immunity to *Chlamydia* infection (see Igiertseme, Black and Caldwell
 40 (2002) Biodrugs 16: 19-35 and Igiertseme *et al* (1999) Immunology 98: 510-519).

Whilst it is now possible to carry out searches of the whole *Chlamydia pneumoniae* genome, there is still insufficient information available on parallel proteome characterisation. By way of example, while sequence data is available for many of the
 45 *Chlamydia pneumoniae* antigens, there is insufficient characterisation of the *Chlamydia* antigens in terms of their immunological and/or biological function. By way of example, whilst applications such as WO 99/28475 and WO 99/27105 disclose sequence information, there is no characterisation of these sequences in terms of their immunological and/or biological function. In contrast, WO 02/02404
 50 provides information on the immunogenicity and immunoaccessibility of certain

Chlamydia proteins and highlights that (i) current genomic annotations and/or (ii) predictions based on cellular location and/or cellular function based on *in-silico* analyses may not always be accurate.

- 5 Applicants have recently engaged in a whole-genome search (Montigiani *et al* (2002) Infection and Immunity 70:368-379) for possible vaccine candidates among proteins potentially associated with the outer membrane of *C.pneumoniae*. For this study, mouse antisera was prepared against over 100 recombinant His-tagged or
- 10 Glutathione-S-transferase (GST) fusion proteins encoded by genes predicted by *in silico* analyses to be peripherally located in the *Chlamydial* cell. From this screening study, 53 recombinant proteins derived from the genome of *Chlamydia* (*Chlamydophila pneumoniae* (CPn) were described which induced mouse antibodies, capable of binding, in a FACS assay, to the surface of purified CPn cells.
- 15 The scope of the Montigiani study (*ibid*) was restricted to checking if polyclonal antisera produced in mice against the recombinantly expressed antibodies to CPn antigens were capable of binding to the surface of the CPn cells. No studies were carried out to test whether antisera against the recombinant FACS positive antigens were capable of interfering with EB *in vitro* infectivity of host cells – that is, whether
- 20 the murine antibodies raised against the recombinantly expressed antigens could inhibit CPn infectivity *in vitro* to an extent greater than 50%, a property that common practice qualifies such antigens as “neutralising”.
- 25 Indeed, so far, only few *C. pneumoniae* antigens with ‘neutralizing’ properties have been described in the literature: notably, a protein identified as 76-kDa-homolog protein (Perez-Melgosa *et al* (1994) Infect Immunity 62: 880-6), the surface-exposed outer membrane proteins MOMP (Wolf *et al* (2001) Infect Immun 69: 3082-91), PorB (Kawa *et al* (2002) J Immunol 168 : 5184-91 and Kubo *et al* (2000) Mol Microbiol 38 : 772-80), and very recently also the Pmp21 member of the *Chlamydia*-specific
- 30 polymorphic family of outer membrane proteins (A.Szczepek, personal communication). All these proteins were in fact selected in the earlier FACS-based screening study (Montigiani *et al* (2002) *ibid*). It can be however noted that outer membrane antigens, as it is the case for MOMP and PorB, could possibly present some kind of practical problems for a recombinant vaccine development project. For
- 35 instance both MOMP and PorB are integral membrane proteins which appear to require a native conformation to maintain neutralizing epitopes which are discontinuous and conformation-dependent. The production of such proteins may require special processing steps (refolding) which could be undesirable in the preparation of an hypothetical vaccine. Other general problems may arise from the
- 40 extent of allelic variation, and from regulated proteins which are not always expressed in all *Chlamydial* cell or all *Chlamydial* isolates.

Thus, it is desirable to provide improved compositions capable of eliciting an immune response upon exposure to *Chlamydia pneumoniae* proteins. It is also desirable to

- 45 provide improved compositions comprising one or more combinations of two or more selected CPn proteins with complementary immunological and/or biological profiles capable of providing immunity against *Chlamydial* induced disease and/or infection (such as in prophylactic vaccination) or (b) for the eradication of an established chronic *Chlamydial* infection (such as in therapeutic vaccination).

50

Brief description of the drawings and tables

- Figure 1A.** Assay of *in vitro* neutralization of *C.pneumoniae* infectivity for LLC-MK2 cells by polyclonal mouse antisera to recombinant *Chlamydial* proteins.
- Figure 1B** shows serum titres giving 50% neutralization of infectivity for 10 *C.pneumoniae* recombinant antigens. Each titer was assessed in 3 separate experiments (SEM values shown).
- Figure 2** shows immunoblot analysis of two dimensional electrophoretic maps of *C.pneumoniae* EBs using the immune sera described in the text.
- Figure 3** shows mean numbers of *C.pneumoniae* IFU recovered from equivalent spleen samples from immunized and mock-immunized hamsters following a systemic challenge.
- Figure 4** shows flow cytometric analysis of splenocytes from DNA-immunized HLA-A2 transgenic and non transgenic mice.
- Figure 5** shows a flow cytometric analysis of splenocytes from transgenic and non transgenic mice infected with *C. pneumoniae* EBs.
- Figure 6** shows an alignment of the proteins in the 7105-7110 protein family.
- Figure 7** shows an N-terminal alignment of Cpn0794 – Cpn0799.
- Figure 8** shows a protein encoded by Cpn0796 and demonstrates a C-terminal domain comprising approximately residues from 1 to 648.
- Figure 9** shows an alignment of the C-terminal (beta barrel) domains of the proteins encoded by the *C.pneumoniae* genes Cpn0795 and Cpn0796.
- Table I** shows a summary of data and properties of the *C.pneumoniae* antigens described in the text.
- Table 2** shows results from hamster mouse model studies for hypothetical proteins.
- Table 3** shows expressed genes of CPn EB selected by microarray.
- Table 4** shows *C. pneumoniae* selected peptides: protein sources and HLA-A2 stabilization assay.
- Table 5** shows ELISPOT assay with CD8+ T cells from DNA immunised HLA-A2 transgenic mice.
- Table 6** shows IFN- γ production from splenocytes of DNA immunized HLA-A2 transgenic and non transgenic mice.

Summary of the Invention

5 The present invention relates to a polypeptide for use as an autotransporter antigen, the polypeptide comprising: (a) an amino acid sequence selected from the group consisting of SEQ ID NO: 54, SEQ ID NO: 6, SEQ ID NO: 55, SEQ ID NO: 56, SEQ ID NO: 78, and SEQ ID NO: 79, (b) an amino acid sequence having at least 50% sequence identity to an amino acid sequence of (a); or (c) an amino acid sequence comprising one or more fragments of at least 7 consecutive amino acids from an amino acid sequence of (a) or combinations thereof.

10 The present invention also relates to the use of a polypeptide in the preparation of a medicament for the prevention or treatment of a *Chlamydia pneumoniae* infection in an individual. For example, the use of the polypeptide may be as an autotransporter protein which immunoreacts with seropositive serum of an individual infected with *Chlamydia pneumoniae*.

15 The present invention further relates to a method of eliciting an immune response in an individual comprising administering to the individual a polypeptide comprising (a) an amino acid sequence selected from the group consisting of SEQ ID NO: 54, SEQ ID NO: 6, SEQ ID NO: 55, SEQ ID NO: 56, SEQ ID NO: 78, and SEQ ID NO: 79 (b) an amino acid sequence having at least 50% sequence identity to an amino acid sequence of (a), or (c) an amino acid sequence comprising one or more fragment of at least 1, 2, 3, 4, 5, 6, or 7 amino acids from an amino acid sequence of (a) or mixtures thereof.

20 Also, a method is provided for diagnosing an immune response in an individual comprising (a) contacting a biological sample obtained from the individual with a binding agent that binds to a polypeptide with an autotransporter function, (b) detecting in the biological sample the amount of the polypeptide that binds to the binding agent; and (c) comparing the amount of the polypeptide to a predetermined cut-off value and thereby determining the presence of an immune response in the individual.

30 A composition for eliciting an immune response in a subject comprising two or more *Chlamydia pneumoniae* autotransporter proteins or immunogenic fragments thereof is also provided. The composition may further comprise one or more immunostimulants.

35 Also provided is a polypeptide for use as an autotransporter antigen comprising an amino acid sequence corresponding to SEQ ID NO: 86, an amino acid sequence having at least 50% sequence identity to SEQ ID NO: 86, or an amino acid sequence comprising one or more fragments of at least 7 consecutive amino acids of SEQ ID NO: 86.

40 The present invention relates to a composition comprising a first biological molecule from a *Chlamydia pneumoniae* bacterium and a second biological molecule from a *Chlamydia pneumoniae* bacterium. The first biological molecule is selected from the group consisting of SEQ ID No 1 to SEQ ID No 86, or the group consisting of SEQ ID No. 1 to 41.

45 The composition may also contain the second biological molecule being selected from the group consisting of SEQ ID No 1 to SEQ ID No. 86 or SEQ ID No 1 to SEQ ID No 41.

The composition may also comprise two or more biological molecules selected from the group consisting of SEQ ID Nos 1-41.

5 The composition may also comprise one or more biological molecules selected from the group consisting of SEQ ID Nos 1-41 combined with one or more biological molecules selected from the group consisting of SEQ ID Nos 42-86.

10 The composition according to any one of the previous claims further comprising an adjuvant such as an ADP-ribosylating exotoxin or a derivative thereof or an adjuvant is selected from the group consisting of cholera toxin (CT), Escherichia heat-labile enterotoxin (LT) and mutants thereof having adjuvant activity.

15 A vaccine and use of the vaccine is also provided comprising the composition of the present invention. The vaccine may be used in the preparation of a medicament for the prevention or treatment of a *Chlamydia* infection and may be administered mucosally, intra-nasally or intra-vaginally, for example.

20 Further, a method is provided for treating a *Chlamydia* infection in a host subject wherein the method comprises the administration of a safe and effective amount of a vaccine.

25 In another aspect of the invention, an immunogenic composition is provided comprising a combination of *Chlamydia pneumoniae* antigens, the combination comprising at least one *Chlamydia pneumoniae* antigen associated with elementary bodies of *Chlamydia pneumoniae* and at least one *Chlamydia pneumoniae* antigen associated with reticulate bodies of *Chlamydia pneumoniae*.

30 In another aspect of the invention, an immunogenic composition is provided comprising a combination of *Chlamydia pneumoniae* antigens, the combination comprising at least one *Chlamydia pneumoniae* antigen of a first antigen group and at least one *Chlamydia pneumoniae* antigen of a second antigen group, said first antigen group comprising a Type III secretion system (TTSS) protein and said second antigen group comprising a Type III secretion system (TTSS) effector protein.

35 In yet another aspect of the invention, an immunogenic composition is provided comprising a combination of *Chlamydia pneumoniae* antigens comprising at least one *Chlamydia pneumoniae* antigen that is conserved over at least two serovars.

40 In still another aspect of the invention, an immunogenic composition is provided comprising a combination of *Chlamydia pneumoniae* antigens, the combination eliciting a *Chlamydia pneumoniae* specific TH1 immune response and a *Chlamydia pneumoniae* specific TH2 immune response.

45 The present invention further provides a method of monitoring the efficacy of treatment of a patient infected with *Chlamydia pneumoniae* comprising determining the level of *Chlamydia pneumoniae* specific antibody in the patient after administration of an immunogenic composition of the present invention to the patient.

Description of the Invention

The present invention provides compositions comprising a first biological molecule from a *Chlamydia pneumoniae* bacterium and a second biological molecule from a *Chlamydia pneumoniae* bacterium. The term "biological molecule" includes proteins, antigens and nucleic acids. The compositions may also comprise further biological molecules preferably also from *Chlamydia pneumoniae*. That is to say, the compositions may comprise two or more biological molecules (eg. 3, 4, 5, 6, 7, 8 etc.) at least two of which are from a *Chlamydia pneumoniae* bacterium (eg. 3, 4, 5, 6, 7, 8 etc.). Such compositions include those comprising (i) two or more different *Chlamydia pneumoniae* proteins; (ii) two or more different *Chlamydia pneumoniae* nucleic acids, or (iii) mixtures of one or more *Chlamydia pneumoniae* protein and one or more *Chlamydia pneumoniae* nucleic acid.

In one aspect of the present invention, an immunogenic composition is provided comprising a combination of at least one antigen that elicits a *Chlamydia pneumoniae* specific TH1 immune response (such as a cell mediated or cellular immune response) and at least one antigen that elicits a *Chlamydia pneumoniae* specific TH2 response (such as a humoral or antibody response). The immunogenic composition may further comprise a TH1 adjuvant and a TH2 adjuvant.

In another aspect of the present invention, an immunogenic composition is provided comprising a combination of *Chlamydia pneumoniae* antigens comprising at least one *Chlamydia pneumoniae* antigen that is conserved over at least two serovars.

In yet another aspect of the present invention, an immunogenic composition is provided comprising a combination of at least one antigen that elicits a *Chlamydia pneumoniae* specific TH1 immune response and at least one antigen that elicits a *Chlamydia pneumoniae* specific TH2 immune response, the combination comprising at least one *Chlamydia pneumoniae* antigen that is conserved over at least two serovars.

In another aspect of the present invention, the immunogenic composition comprising at least one antigen that elicits a *Chlamydia pneumoniae* specific TH1 immune response and at least one antigen that elicits a *Chlamydia pneumoniae* specific TH2 immune response preferably comprises a combination of *Chlamydia pneumoniae* antigens comprising at least one *Chlamydia pneumoniae* antigen associated with the EB of *Chlamydia pneumoniae* and at least one *Chlamydia pneumoniae* antigen associated with the RB of *Chlamydia pneumoniae*. Still further such combinations can comprise EB and/or RB antigens from one serovar combined with RB and/or EB antigens from at least one other serovar.

In an additional aspect of the present invention, a kit is provided comprising a combination of *Chlamydia pneumoniae* antigens wherein at least one of the *Chlamydia pneumoniae* antigens is associated with the EB of *Chlamydia pneumoniae* and at least one of the *Chlamydia pneumoniae* antigens is associated with the RB of *Chlamydia pneumoniae*. The kit may further include a TH1 adjuvant, a TH2 adjuvant and instructions.

The present invention further provides methods of eliciting a *Chlamydia* specific immune response by administering an immunogenic composition of this invention.

The present invention further provides a method of monitoring the efficacy of treatment of a subject infected with *Chlamydia pneumoniae* comprising determining the level of Chlamydia specific antibody or Chlamydia specific effector molecule in the subject after administration of an immunogenic composition of this invention.

5

In one preferred embodiment the first and second biological molecules are from different *Chlamydia pneumoniae* species (for example, from different *Chlamydia pneumoniae* serovars) but they may be from the same species. The biological molecules in the compositions may be from different serogroups or strains of the same species. The first biological molecule is preferably selected from the group consisting of SEQ ID Nos 1-86. More preferably, it is selected from the group consisting of SEQ IDs 1-41 and/or SEQ ID Nos 42-86. It is preferably a purified or isolated biological molecule. The second biological molecule is preferably selected from the group consisting of SEQ ID Nos 1-86. More preferably, it is selected from the group consisting of SEQ IDs 1-41 and/or SEQ ID Nos 42-86. It is preferably a purified or isolated biological molecule. Specific compositions according to the invention therefore include those comprising: two or more biological molecules selected from the group consisting of SEQ ID Nos 1-41; one or more biological molecules selected from the group consisting of SEQ IDs 1-41 combined with one or more biological molecules selected from the group consisting of SEQ IDs 42-86. One or both of the first and second biological molecules may be a *Chlamydia pneumoniae* biological molecule which is not specifically disclosed herein, and which may not have been identified, discovered or made available to the public or purified before this patent application was filed.

In another embodiment, a combination of *Chlamydia pneumoniae* antigens is provided, the combination comprising at least one Type III Secretion System (TTSS) protein and at least one Type III Secretion System (TTSS) secreted or effector protein or fragment thereof. There are many methods for identifying TTSS proteins (i.e., TTSS proteins associated with the Chlamydial TTSS machinery). TTSS is a complex protein secretion and delivery machine or apparatus, which may be located, either wholly or partially, on the Elementary Body (EB) and which allows an organism, such as Chlamydia, to maintain its intracellular niche by injecting proteins, such as bacterial effector proteins (which may act as anti-host virulence determinants) into the cytosol of a eukaryotic cell in order to establish the bacterial infection and to modulate the host cellular functions. TTSS proteins exposed on the EB surface may play a role in adhesion and/or uptake into host cells.

By way of background information, the TTSS is a complex protein secretion and delivery machine or apparatus, which may be located on the Elementary Body (EB) and which allows an organism, such as Chlamydia, to maintain its intracellular niche by injecting proteins, such as bacterial effector proteins (which may act as anti-host virulence determinants) into the cytosol of a eukaryotic cell in order to establish the bacterial infection and to modulate the host cellular functions. These injected proteins (the TTSS effector proteins) can have various effects on the host cell which include but are not limited to manipulating actin and other structural proteins and modification of host cell signal transduction systems. The injected (or translocated) proteins or substrates of the TTSS system may also be processed and presented by MHC-class I molecules.

Not all the proteins secreted by a Type III secretion system are delivered into the host cell or have effector function. Although the Elementary Body (EB) is regarded as “metabolically inert”, it has been postulated that the Chlamydial TTSS system located on the (EB) is triggered by membrane contact and is capable of releasing pre-formed “payload” proteins. The current hypothesis is that Type Three Secretion System (TTSS) becomes active during the intracellular phase of the chlamydial replicative cycle for the secretion of proteins into the host cell cytoplasm and for the insertion of chlamydial proteins (like the Inc set) into the inclusion membrane that separates the growing chlamydial microcolony from the host cell cytoplasm (see Montigiani et al (2002) Infection and Immunity 70(1); 386-379).

Proteins may be expressed and secreted by 2 hours (early cycle) after infection while the expression of other early and mid cycle Type III specific genes are not detectable until 6-12 hours (mid cycle). After 16-20 hours, the RBs begin to differentiate into EBs, and by 48-72 hours, the EBs predominate within the inclusion. Host cell lysis results in the release of the EBs to the extracellular space where they can infect more cells. For purposes of this description, an early gene is one that is expressed (in terms of mRNA expression) early in infection, an intermediate gene is one that is expressed in the mid-cycle after infection and a late gene is one which is expressed during the terminal transition of RBs to EBs. There may be a time lag between surface expression of early, mid and late stage proteins and their transcriptional and translational profiles because mRNA abundance may not always correlate with protein abundance.

In one example, the present invention may comprise TTSS effector proteins. The TTSS effector proteins as described are associated with the RB form of *Chlamydia pneumoniae* and may be identified, for example, using immunofluorescence microscopy (see Bannantine et al, Infection and Immunity 66(12); 6017-6021). Effector antibodies to putative Chlamydial TTSS effector proteins secreted by the TTSS machinery may be micro-injected into host cells at specified time points during *Chlamydia pneumoniae* infection (e.g., early, mid or late cycle). Host cell reaction to *Chlamydia pneumoniae* (e.g., actin remodeling, inhibition of endosomal maturation, host lipid acquisition, and MHC Class I and Class II molecule downregulation) associated with *Chlamydia pneumoniae* entry into host cells is then observed. Based on these temporal observations, TTSS effector proteins (RB-associated *Chlamydia pneumoniae* proteins) may be detected.

A specific composition of the present invention may comprise a combination of *Chlamydia pneumoniae* antigens, said combination consisting of two, three, four, five or all six *Chlamydia pneumoniae* antigens of a first antigen group, said first antigen group consisting of: (1) pmp2; (2) pmp10; (3) Enolase; (4) OmpH-like protein; and (5) the products of CPn specific genes CPn0759 and CPn0042. These antigens are referred to herein as the ‘first antigen group’.

Preferably, the composition of the invention comprises a combination of *Chlamydia pneumoniae* antigens, said combination selected from the group consisting of: (1) pmp2 and pmp10; (2) pmp2 and Enolase; (3) pmp2 and OmpH-like protein; (4) pmp2 and CPn0759; (5) pmp2 and CPn0042; (6) pmp10 and Enolase; (7) pmp10 and

OmpH-like protein; (8) pmp10 and CPn0759; (9) pmp10 and CPn0042; (10) Enolase and OmpH-like protein (11) Enolase and CPn0759; (12) Enolase and CPn0042; (13) OmpH-like protein and CPn0759 (14) OmpH-like protein and CPn0042; (15) CPn0759 and CPn0042; (16) pmp2 and pmp10 and Enolase; (17) pmp2 and pmp10 and OmpH-like protein; (18) pmp2 and pmp10 and CPn0759; (19) pmp2 and pmp10 and CPn0042; (20) pmp2 and Enolase and OmpH-like protein; (21) pmp2 and Enolase and CPn0759; (22) pmp2 and Enolase and CPn0042; (23) pmp2 and OmpH-like protein and CPn0759; (24) pmp2 and OmpH-like protein and CPn0042; (25) pmp2 and CPn0759 and CPn0042; and (26) pmp10 and Enolase and OmpH-like protein; (27) pmp10 and Enolase and CPn0759; (28) pmp10 and Enolase and CPn0042; (29) Enolase and OmpH-like protein and CPn0759; (30) Enolase and OmpH-like protein and CPn0042; (31) OmpH-like protein and CPn0759 and CPn0042.

Preferably, the composition of *Chlamydia pneumoniae* antigens consists of pmp2, pmp10, Enolase, OmpH-like protein and CPn0759.

Preferably, the composition of *Chlamydia pneumoniae* antigens consists of pmp2, pmp10, Enolase, OmpH-like protein and CPn0042.

Preferably, the composition of *Chlamydia pneumoniae* antigens consists of pmp2, pmp10, Enolase, OmpH-like protein and CPn0759 and CPn0042.

The invention also provides for a slightly larger group of 12 *Chlamydia pneumoniae* antigens that are particularly suitable for immunisation purposes, particularly when used in combinations. (This second antigen group includes the six *Chlamydia pneumoniae* antigens of the first antigen group). These 12 *Chlamydia pneumoniae* antigens form a second antigen group of (1) pmp2; (2) pmp10; (3) Enolase; (4) OmpH-like protein; (5) CPn0759; (6) CPn0042; (7) ArtJ; (8) HtrA; (9) AtoS; (10) OmcA; (11) CPn0498; and (12) CPn0525. These antigens are referred to herein as the 'second antigen group'.

The invention therefore provides a composition comprising a combination of *Chlamydia pneumoniae* antigens, said combination selected from the group consisting of two, three, four, five, six, seven, eight, nine, ten, eleven, or twelve *Chlamydia pneumoniae* antigens of the second antigen group. Preferably, the combination is selected from the group consisting of two, three, four or five *Chlamydia pneumoniae* antigens of the second antigen group. Still more preferably, the combination consists of six *Chlamydia pneumoniae* antigens of the second antigen group. Each of the *Chlamydia pneumoniae* antigens of the first and second antigen group are described in more detail below.

(1) Pmp10 (CPn0449)

One example of a pmp10 protein is set forth as SEQ ID NO: 1 below (GenBank Accession No.GI:14195016). Preferred pmp10 proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 1; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 1, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These pmp2 proteins include variants (e.g. allelic variants, homologs, orthologs, paralog,

- mutants, *etc.*) of SEQ ID NO: 1. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 1. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 1. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 1

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1 MKSQFSWLVL SSTLACFTSC STVFAATAEN IGPSDSFDGS TNTGTYTPKN TTTGIDYTLT
61 GDITLQNLGD SAALTKGCFS DTESLSFAG KGYSLSFLNI KSSAEGAALS VTSDKNLSLT
121 GFSSLTFLAA PSSVITTPSG KGAVKCGDL TFDNNGTILE KQDYCEENG AISTKNLSLK
181 NSTGSISFEG NKSSATGKKG GAICATGTVD ITNNTAPTLE SNNIAEAAGG AINSTGNCTI
15 241 TGNTSLVFSE NSVTATAGNG GALSGDADVT ISGNQSVTFE GNQAVANGGA IYAKKLTLAS
301 GGGGGISFSN NIVQGTAGN GGAISILAAG ECSLSAEAGD ITFNGNAIVA TTPQTTRKNS
361 IDIGSTAKIT NLRAISGHSI FFYDEPITANT AADSTDITLNL NKADAGNSTD YSGSIVFSGE
421 KLSSEDEAKVA DNLTSTLKQP VTLTAGNLVL KRGVTLDTKG FTQTAGSSVI MDAGTTLKAS
481 TEEVTLTGLS IPVDSLGEK KVVIAASAAS KNVALSGPIL LLDNQGNAYE NHDLGKTQDF
20 541 SFVQLSALGT ATTTDVPAVP TVATPTHYGY QGTWGMTWVD DTASTPKTKT ATLAWTNTGY
601 LPNPERQGPI VPNSLWGSFS DIQAIQGVIE RSALTLCSDR GFWAAGVANF LDKDKKGEKR
661 KYRHKSGGYA IGGAAQTCSE NLISFAFCQL FGSDKDFLVA KNHTDTYAGA FYIQHITECS
721 GFIGCLLDKL PGWSHKPLV LEGQLAYSHV SNDLTKKYTA YPEVKGSWGN NAFNMMLGAS
781 SHSYPEYLHC FDTYAPYIKL NLTYIRQDSF SEKGTEGRSF DDSNLFNLSL PIGVKFEKFS
25 841 DCNDFS YDLT LSYVPDLIRN DPKCTTALVI SGASWETIAN NLARQALQVR AGSHYAFSPM
901 FEVLGQFVFE VRGSSRIYNV DLGGKFQF

(2) *Pmp2 = Polymorphic Outer Membrane Protein G Family (CPn 0013)*

- One example of a pmp2 protein is disclosed as SEQ ID NO^s: 139 and 140 in WO 02/02606. {GenBank accession number: gi|4376270|gb|AAD18172.1 'CPn0013'; SEQ ID NO: 2 below}. Preferred pmp2 proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 2; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 1, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These pmp2 proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of SEQ ID NO: 2. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 1. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 2. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 2

1 MKIPLRFLLI SLVPTLSMSN LLGAATTEEL SASNSFDGTT STTSFSSKTS
51 SATDGTNYVF KDSVVIENVP KTGETQSTSC FKNDAAAGDL NFLGGGFSFT
101 FSNIDATTAS GAAIGSEAN KTVTLSGFSA LSFLKSPAST VTNGLGAINV
50 151 KGNLSLLDND KVLIQDNFST GDGGAINCAG SLKIANNKSL SFIGNSSSTR
201 GGAIHTKNLT LSSGGETLFQ GNTAPTAAGK GGAIAIADSG TLSISGDSGD
251 IIFEGNTIGA TGTVSHSAID LGTSAKITAL RAAQGHTIYF YDPITVTGST
301 SVADALNINS PDTGDNKEYT GTIVFSGEKL TEAEAKDEKN RTSKLLQNVA
351 FKNGTVVLKG DVVLSANGFS QDANSKLIMD LGTSLVANTE SIELTNLEIN
55 401 IDSLRNGKKI KLSAATAQKD IRIDRPVLA ISDESFYQNG FLNEDHSYDG
451 ILELDAGKDI VISADSRSID AVQSPYGYQG KWTINWSTDD KKATVSWAQ
501 SFNPTEAQEA PLVPNLLWGS FIDVRSFQNF IELGTEGAPY EKRFVWAGIS

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551 NVLHRSGREN QRKFRHVSGG AVVGASTRMP GGDTLSLGFA QLFARDKDYF
601 MNTNFAKTYA GSLRLQHDAS LYSVVSILLG EGGLREILLP YVSKTLPCSF
651 YGQLSYGHTD HRMKTESLPP PPPTLSTDHT SWGGYVWAGE LGTRVAVENT
701 SGRGFFQEYT PFVKVQAVYA RQDSFVELGA ISRDFSDSLH YNLAIPLGIK
751 LEKRFAEQYY HVVAMYSADV CRSNPKCTTT LLSNQGSWKT KGSNLARQAG
801 IVQASGFRSL GAAAEELFGNF GFEWRGSSRS YNVDAGSKIK F*

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(3) *Enolase (Cpn0800)*

One example of an 'Eno' protein is disclosed as SEQ ID NO^s: 93 and 94 in WO 02/02606. {GenBank accession number: gi|4377111|gb|AAD18938.1| 'Cpn0800'; SEQ ID NO: 3 below}. Preferred Eno proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 2; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 2, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These Eno proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of SEQ ID NO: 3. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 3. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 3. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

25 *SEQ ID No 3*

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1 MFEAVIADIQ AREILDSRGY PTLHVKVTTT TGSVGEARVP SGASTGKKEA
51 LEFRDTSRPR YQKGVLQAV KNVKEILFPL VKGCSVYEQS LIDSLMMDSD
101 GSPNKETLGA NAILGVSLAT AHAAAATLRR PLYRYLGGCF ACSLPCPMNN
30 151 LINGGMHADN GLEFQEFMIR PIGASSIKEA VNMGADVFTT LKKLLHERGL
201 STGVGDEGGF APNLASNEEA LELLLLAIEK AGFTPGKDIS LALDCAASSF
251 YNVKTGTIDG RHYEQIAIL SNLCDRYPID SIEDGLAEDD YDGWALLTEV
301 LGEKVQIVGD DLFVTNPILI LEGISNGLAN SVLIKPNQIG TLTETVYAIK
35 351 LAQMAGYTTI ISHRSGETTD TTIADLAVAF NAGQIKTGSL SRSERVAKYN
401 RLMEIEEELG SEAIFTDSNV FSYEDSEE*

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(4) *OmpH-like outer membrane protein (CPn0301)*

One example of 'OmpH-like' protein is disclosed as SEQ ID NO^s: 77 & 78 in WO 02/02606. {GenBank accession number: gi|4376577|gb|AAD18450.1| 'CPn0301'; SEQ ID NO: 4 below}. Preferred OmpH-like proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 4; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 3, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These OmpH-like proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of SEQ ID NO: 4. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 4. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably 19 or more, to remove the signal peptide) from the N-terminus of SEQ ID NO: 4. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide as described above, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 4

1 MKKLLFSTFL LVLGSTSAAH ANLGYVNLKR CLEESDLGKK ETEELEAMKQ
 51 QFVKNAEKIE EELTSIYNKL QDEDYMESLS DSASEELRKK FEDLSGEYNA
 5 YQSQYYQSIN QSNVKRIQKL IQEVKIAAES VRSKEKLEAI LNEEAVLAIA
 151 PGTDKTTEII AILNESFKKQ N*

(5) CPn0042 (Hypothetical)

One example of hypothetical protein is set forth as SEQ ID NO: 5 below.

10 GenBank accession number: gi|4376296|gb|AAD18195.1. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 5; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 5, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These Hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 5. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 5. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably 19 or more, to remove the signal peptide) from the N-terminus of SEQ ID NO: 5. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 5

1 MEEVSEYLQQ VENQLESCSK RLTKMETFAL GVRLEAKEEI ESIILSDVFN RFEVLCRDIE
 61 DMLSRVEEIE RMLRMAELPL LPIKEALTKA FVQHNSCKEK LTKVEPYFKE SPAYLTSEER
 30 121 LQSLNQTLQR AYKESQKVSQ LESEVRACRE QLKDQVRQFE TQGVSLIKEE ILFVTSTFRT
 181 KFSYHSFRLH VPCMRLYEEY YDDIDLERTR ARWMAMSERV RDAFQAFQEM LKEGLVEEAQ
 241 ALRETEYWLY REERKSKKKH

(6) CPn0795 (Hypothetical)

35 One example of hypothetical protein is disclosed as SEQ ID NO^s: 63 & 64 in WO 02/02606. {GenBank accession number: gi|4377106|gb|AAD18933.1| 'CPn0795'; SEQ ID NO: 6 below). As the examples demonstrate, we have shown for the first time that CPn0795 and related proteins in the group Cpn0794 – Cpn0799 have a secreted autotransporter function. It has been shown that proteins secreted by the autotransporter secretion mechanism possess an overall unifying structure, including an amino-terminal leader peptide (for secretion across the inner membrane), the secreted mature protein (or passenger domain), and a dedicated C-terminal domain, which forms a pore in the outer membrane through which the passenger domain passes to the cell surface. It is likely that requirements for secretion across the outer membrane are contained within a single molecule and secretion is an energy-independent process. Structural properties of the proteins may be confined by the size of the pore considering the biophysical constraints that may be imposed on secretion.

50 The autotransporter, or type V, secretion system is a dedicated protein translocation mechanism which allows the organism to secrete proteins to and beyond the bacterial surface. The secretion mechanism and the ability to develop a new autotransporter protein simply by a single recombination event have presented bacteria with abundant

opportunities to increase the efficiency of secretion of proteins that were developed as periplasmic or exported virulence factors.

5 In one model of autotransporter (type V) secretion mechanism, proteins are exported by the autotransporter secretion mechanism and are translated as a polypeptide possessing domains. The autotransporters possess an overall unifying structure comprising three functional domains: the amino-terminal leader sequence, the secreted mature protein (passenger domain) and a carboxy-terminal (beta-) domain that forms a beta-barrel pore to allow secretion of the passenger protein. The leader
10 sequence directs secretion via the sec apparatus and is cleaved at the inner membrane by a signal peptidase releasing the remaining portion of the molecule into the periplasm. Once in the periplasm the β -domain assumes a biophysically favored state characterized by a β -barrel shaped structure which inserts itself into the outer membrane to form a pore. After insertion into the outer membrane the passenger domain is translocated to the bacterial cell surface where it may remain intact or
15 undergo processing. A processed protein may be released into the extracellular milieu or remain associated with the bacterial cell surface. (Henderson and Nataro, "Virulence Functions of Autotransporter Proteins", Infection and Immunity, Vol. 69, No. 3, March 2001, pages 1231-1243).

20 Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 6; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID
25 NO: 6, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These Hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 6. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 6. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or
30 more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably 19 or more, to remove the signal peptide) from the N-terminus of SEQ ID NO: 6. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain). As the Examples
35 demonstrate, we have shown for the first time that Cpn0795 appears to be present and accessible to antibodies on the surface of the infectious EB form which makes this protein a good component of an immunogenic composition or vaccine.

40 Table 1 of this application demonstrates that Cpn0795 (SEQ ID NO: 6) a Cpn specific hypothetical protein is a FACS positive protein which demonstrates significant immunoprotective activity in a hamster spleen model of *Chlamydia pneumoniae* infection. We have found evidence to demonstrate that other Cpn proteins in this group of Cpn specific hypothetical proteins have now been found to have a secreted autotransporter function. These proteins, which are absent from *Chlamydia*
45 *trachomatis* include: gi/4377105 (Cpn0794), gi/4377106 (Cpn0795), gi/4377107 (Cpn0796), gi/4377108 (Cpn0797), gi/4377109 (Cpn0798), gi/4377110 (Cpn0799).

SEQ ID No 6

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(7) ArtJ arginine periplasmic-binding protein (CPn 0482)

One example of 'ArtJ' protein is disclosed as SEQ ID NO^s: 73 & 74 in WO 02/02606. {GenBank accession number: gi|4376767|gb|AAD18622.1| 'CPn0482'; SEQ ID NO: 7 below}. Preferred ArtJ proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 7; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 7, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These ArtJ proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 7. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 7. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 7. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain). The ArtJ protein may be bound to a small molecule like arginine or another amino acid.

SEQ ID No 7

1 MIKQIGRFFR AFIFIMPLSL TSCESKIDRN RIWIVGTNAT YPPFEYVDAQ
51 GEVVGFDIDL AKAISEKLGK QLEVREFAFD ALILNLKKHR IDAILAGMSI
101 TPSRQKEIAL LPYYGDEVQE LMVVSQRSLE TPVLPLTQYS SVAVQTGTFO
151 EHYLLSQPGI CVRSFDSTLE VIMEVRYGKS PVAVLEPSVG RVVLKDFPNL
201 VATRLELPPE CWVLGCGLGV AKDRPEEIQT IQQAITDLKS EGVIQSLTKK
WQLSEVAYE*

(8) HtrA DO Serine Protease (CPn0979)

One example of an 'HtrA' protein is disclosed as SEQ ID NO^s: 111 & 112 in WO 02/02606. {GenBank accession number: gi|4377306|gb|AAD19116.1| 'CPn0979'; SEQ ID NO: 8 below}. Preferred HtrA proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 8; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 8, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These HtrA proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 8. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 8. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably at least 16 to remove the signal peptide) from the N-terminus of SEQ ID NO: 8. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above, of a cytoplasmic

domain, of a transmembrane domain, or of an extracellular domain). In relation to SEQ ID NO: 8, distinct domains are residues: 1-16; 17-497; 128-289; 290-381; 394-485; and 394-497.

5 SEQ ID No 8

1 MITKQLRSWL AVLVGSSLLA LPLSGQAVGK KESRVSELPQ DVLLKEISGG
 51 FSKVATKATP AVVYIESFPK SQAVTHPSPG RRGPYENPFD YFNDEFFNRF
 101 FGLPSQREKP QSKEAVRG TG FLVSPDGYIV TNNHVVEDTG KIHVTLHDGQ
 151 KYPATVIGLD PKTDLAVIKI KSONLPYLSF GNSDHLKVGD WAIAIGNPFG
 10 201 LQATVTVGVI SAKGRNQLHI ADFEDFIQTD AAINPGNSGG PLLNIDGQVI
 251 GVNTAIVSGS GGYIGIGFAI PSLMANRIID QLIRDGQVTR GFLGVTLOPI
 301 DAELAACYKL EKVGALVTD VVKGSPADKA GLKQEDVIA YNGKEVDSLS
 351 MFRNAVSLMN PDTRIVLKVV REGKVIEIPV TVSQAPKEDG MSALQRVGIR
 401 VQNLTPETAK KLGIAPETKG ILIISVEPGS VAASSGIAPG QLILAVNRQK
 15 451 VSSIEDLNRT LKDSNNENIL LMVSQGDVIR FIALKPEE*

(9) *AtoS two-component regulatory system sensor histidine kinase protein (CPn0584)*

One example of 'AtoS' protein is disclosed as SEQ ID NO^s: 105 & 106 in WO 02/02606. {GenBank accession number: gi|4376878|gb|AAD18723.1| 'CPn0584';
 20 SEQ ID NO: 9 below}. Preferred AtoS proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 9; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 9, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40,
 25 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These AtoS proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 9. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 9. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8,
 30 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 9. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

35 SEQ ID No 9

1 MNVPDSKNLH PPAYELLEIK ARITQSYKEA SAILTAIPDG ILLLSETGHF
 51 LICNSQAREI LGIDENLEIL NRSFTDVLDP TCLGFSIQEA LESLKVPKTL
 101 RLSLCKESKE KEVELFIRKN EISGYLFIQI RDRSDYKQLE NAIERYKNIA
 151 ELGKMTATLA HEIRNPLSGI VGFASILKKE ISSPRHQRL SSIISGTRSL
 40 201>NNLVSSMLEY TKSQPLNLKI INLQDFSSSL IPLLSVSFPN CKFVREGAQP
 251 LFRSIDPDRM NSVVWNLVKN AVETGNSPIT LTLHTSGDIS VTNPGTIPSE
 301 IMDKLFTPEF TTKREGNGLG LAEAQKIIRL HGGDIQLKTS DSAVSFFIII
 351 PELLAALPKE RAAS*

45 (10) *OmcA 9kDa-cysteine-rich lipoprotein(CPn0558)*

One example of 'OmcA' protein is disclosed as SEQ ID NO^s: 9 & 10 in WO 02/02606. {GenBank accession number: gi|4376850|gb|AAD18698.1| 'CPn0558', 'OmcA', 'Omp3'; SEQ ID NO: 10 below}. Preferred OmcA proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g.
 50 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 10; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 10, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These OmcA proteins include variants (e.g. allelic variants, homologs, orthologs,
 55 paralogs, mutants, etc.) of SEQ ID NO: 10. Preferred fragments of (b) comprise an

- epitope from SEQ ID NO: 10. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably 18 or more to remove the signal peptide) from the N-terminus of SEQ ID NO: 10. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain). The protein may be lipidated (e.g. by a *N*-acyl diglyceride), and may thus have a N-terminal cysteine.

10 *SEQ ID No 10*

1 MKKAVLIAAM FCGVVSLSSC CRIVDCCFED PCAPSSCNPC EVIRKKERSC
51 GGNACGSYVP SCSNPGSTE CNSQSPQVKG CTSPDGRCKQ *

15 *(11) CPn0498 (Hypothetical)*

- One example of a hypothetical protein is set forth as SEQ ID NO: 11 below. (GenBank Accession No. GI:4376784; AAD18638.1). Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 11; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 11, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These Hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 11. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 11. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably 18 or more to remove the signal peptide) from the N-terminus of SEQ ID NO: 11. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain). The protein may be lipidated (e.g. by a *N*-acyl diglyceride), and may thus have a N-terminal cysteine.

SEQ ID No 11

- 35
1 MNRRKARWV ALFAMTALIS VGCCPWSQAK SRCSIDKYIP VVNRLLEVCG LPEAENVEDL
61 IESSSAWVLT PEERFSGELV SICQVKDEHA FYNDLSLLHM TQAVPSYSAT YDCAVVFGGP
121 LPALRQRLDF LVREWQRGVR FKKIVFLCGE RGRYQSIEEQ EHFFDSRYNP FPTEENWESG
181 NRVTPSSEEE IAKFVWMQML LPRAWRDSTS GVRVTFLLAK PEENRVVANR KDTLLLFERSY
40 241 QEAFPGRVLF VSSQPFIGLD ACRVGQFFKG ESYDLAGPGF AQGVLYKHYA PRICLHTLAE
301 WLKETNGCLN ISEGCFG

(12) CPn 0525 (hypothetical)

- One example of 'Cpn0525' protein is disclosed as SEQ ID NO^s: 117 & 118 in WO 02/02606. {GenBank accession number: gi|4376814|gb|AAD18665.1| 'CPn0525', SEQ ID NO: 12 below}. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 12; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 12, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These OmcA proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs,

mutants, *etc.*) of SEQ ID NO: 12. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 12. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably 18 or more to remove the signal peptide) from the N-terminus of SEQ ID NO: 12. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 12

```

1  MHDALLSILA IQELDIKMIR LMRVKKEHQK ELAKVQSLKS DIRRKVQEKE
51  LEMENLKTQI RDGENRIQEI SEQINKLENQ QAAVKKMDEF NALTQEMTTA
101 NKERRSLEHQ LSDLMDKQAG GEDLIVSLKE SLASTENSSS VIEKEIFESI
151 KKINEEGKAL LEQRTELKHA TNPILLSIYE RLLNNKKDRV VVPIENRVCS
201 GCHIVLTPQH ENLVRKKDRL IFCEHCSRIL YWQESQVNAQ ENSTAKRRRR
251 RAAV*

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Third Antigen Group

The immunogenicity of other *Chlamydia pneumoniae* antigens may be improved by combination with two or more *Chlamydia pneumoniae* antigens from either the first antigen group or the second antigen group. Such other *Chlamydia pneumoniae* antigens include a third antigen group consisting of (1) LcrE, (2) DnaK, (3) Omp85 homolog, (4) Mip-like; (5) OmcB (6) MurG (7) Cpn0186 and (8) fliY. These antigens are referred to herein as the "third antigen group".

(13) LcrE low calcium response E protein (CPn0324)

One example of a 'LcrE' protein is disclosed as SEQ ID NO^s: 29 & 30 in WO 02/02606. {GenBank accession number: gi|4376602|gb|AAD18473.1| 'CPn0324'; SEQ ID NO: 13 below}. Preferred LcrE proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 13; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 13, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These LcrE proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of SEQ ID NO: 13. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 13. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 13. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 13

```

1  MAASGGTGGL GGTQGVNLAA VEAAAADADA AEVVASQEGS EMNMIQQSQD
51  LTNPAAATRT KKKEEFQTL ESRKKGEAGK AEKKSESTEE KPDTDLADKY
101 ASGNSEISGQ ELRGLRDAIG DDASPEDILA LVQEKIKDPA LQSTALDYLV
151 QTTPPSQGKL KEALIQARNT HTEQFGRTAI GAKNILEFASQ EYADQLNVSP
201 SGLRSLYLEV TGDTHTCDSL LSMLQDRYTY QDMAIVSSFL MKGMATELKR
251 QGPYVPSAQL QVLMETRNIL QAVLTSYDYF ESRVPILLDS LKAEGIQTPS
301 DLNFKVAES YHKIINDKFP TASKVEREVR NLIGDDVDSV TGVNLFFSA
351 LRQTSSRLFS SADKRQQLGA MIANALDAVN INNEDYPKAS DFPKPYPWS*

```

(14) DnaK heat-shock protein 70 (chaperone) (CPn0503)

One example of 'DnaK' protein is disclosed as SEQ ID NO^s: 103 & 104 in WO 02/02606. {GenBank accessionnumber: gi|4376790|gb|AAD18643.1| 'CPn0503'; SEQ ID NO: 14 below. Preferred DnaK proteins for use with the invention comprise

5 an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 14; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 14, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These DnaK proteins include

10 variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of SEQ ID NO: 14. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 14. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 14. Other

15 fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 14

20

1	MSEHKKSSKI	IGIDLGTTNS	CVSVMEGGQA	KVITSSEGTR	TTPSIVAFKG
51	NEKLVGIPAK	RQAVTNPEKT	LGSTKRFIGR	KYSEVASEIQ	TVPYTVTSGS
101	KGDAVFEVDG	KQYTPEEIGA	QILMKMKETA	EAYLGGETVTE	AVITVPAYFN
151	DSQRASTKDA	GRIAGLDVVR	IIPEPTAAAL	AYGIDKVGDK	KIAVFDLGGG
201	TFDISILEIG	DGVFEVLSTN	GDTLGGLGDDF	DEVIKWMIE	EFKKQEGIDL
25	251	SKDNMALQRL	KDAAEKAKIE	LSGVSSSTEIN	QPFITMDAQG
	301	AQFEKLAASL	IERTKSPCIK	ALSDAKLSAK	DIDVLLVGG
	351	VKELEFGKEPN	KGVPNPDEVVA	IGAAIQGGVL	GGEVKDVL
	401	TLGGVMTTLV	ERNTTIPTQK	KQIFSTAADN	QPAVTIVVLQ
	451	EIGRFDLTDI	PPAPRGHPQI	EVSFDIDANG	IFHVSADVA
30	501	ASSGLQDEEI	QRMVRDAEIN	KEEDKKRREA	SDAKNEADSM
	551	YKEQIPETLV	KEIEERIENV	RNALKDDAPI	EKIKEVTEDL
	601	MQSQSASAAA	SSAANAKGGP	NINTEDLKKH	SFSTKPPSNN
	651	DVEIIDNDDK*			

35

(15) Omp85 homolog (Cpn0300)

One example of an Omp85 Homolog protein is disclosed as SEQ ID NO^s: 147 & 148 in WO 02/02606. {GenBank accession number: gi|4376576|gb|AAD18449.1| 'CPn0300'; SEQ ID NO: 15 below}. Preferred Omp85 proteins for use with the

40 invention comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 15; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 15, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These DnaK proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of SEQ ID NO: 15. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 15. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the

45 N-terminus of SEQ ID NO: 15. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

50

SEQ ID No 15

1 MLIMRNKVIL QISILALIQT PLTLFSTKVK KEGHVVDSDI TIITEGENAS
 5 51 NKHPLPKLKT RSGALFSQLD FDEDLRILAK EYDSVEPKVE FSEGKTNIAL
 101 HLIAPPSIRN IHISGNQVVP EHKILKTLQI YRNDLFEREK FLKGLDDLRT
 151 YYLKRGYFAS SVDYSLEHNQ EKGHIDVLIK INEGPCGKIK QLTFSGISRS
 201 EKSDIQEFIQ TKQHSTTTTSW FTGAGLYHPD IVEQDSLAIK NYLHNNGYAD
 251 AIVNSHYDLD DKGNNILLYMD IDRGSRYTLG HVHIQGFVVL PKRLIEKQSQ
 10 301 VGPNDLYCPD KIWDGAHKIK QTYAKYGYIN TNVDVLFIPH ATRPIYDVTY
 351 EVSEGSPYKV GLIKITGNTH TKSDVILHET SLFPGDTFNR LKLEDTEQRL
 401 RNTGYFQSVS VYTVRSQLDP MGNADQYRDI FVEVKETTTG NLGLFLGFSS
 451 LDNLFGGIEL SESNFDLFGA RNIFSKGFRG LRGGEHLFL KANFGDKVTD
 501 YTLKWTKPHF LNTPWILGIE LDKSINRALS KDYAVQTYGG NVSTTYILNE
 15 551 HLKYGLEYRG SQTSLHEKRR FLLGPNIDSN KGFVSAAGVN LNYDSVDSR
 601 TPTTGIRGGV TFEVSGLGGT YHFTKLSLNS SIYRKLTRKG ILKIKGEAQF
 651 IKPYSNTTAE GVPVSEFFEL GGETTVRGYK SFIIGPKYSA TEPQGGSSSL
 701 LISEEFQYPL IRQPNISAFV FLDSGFVGLQ EYKISLKDLR SSAGFGLRFD
 751 VMNNVPVMLG FGWPFRTPTET LNGEKIDVSQ RFFFAFGGMF *

20

(16) Mip-like FKBP-type peptidyl-prolyl cis-trans (CPn0661)

One example of a Mip-like protein is disclosed as SEQ ID NO^s: 55 & 56 in WO 02/02606. {GenBank accession number: gi|4376960|gb|AAD18800.1| 'CPn0661'; SEQ ID NO: 16 below}. Preferred Mip-like proteins for use with the invention
 25 comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 16; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 16, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These mip-
 30 like proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 16. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 16. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus
 35 of SEQ ID NO: 16. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

40 **SEQ ID No 16**

1 MNRRWNVLVA TVALALSVAS CDVRSKDKDK DQGSLEVEYKD NKDTNDIELS
 51 DNQKLSRTFG HLLARQLRKS EDMFFDIAEV AKGLQAEIVC KSAPLTETEV
 101 EEKMAEVQKL VFEKSKENL SLAEKFLKEN SKNAGVVEVQ PSKLOYKIIK
 45 151 EGAGKAISGK PSALLHYKGS FINGQVFSSS EGNNEPILLP LGQTIPGFAL
 201 GMQGMKEGET RVLVIHPDLA YGTAGQLPPN SLLIFEINLI QASADEVAHV
 251 PQEGNQGE*

50 **(17) OmcB 60 kDa Cysteine rich OMP (CPn0557)**

One example of an OmcB protein is disclosed as SEQ ID NO^s: 47 & 48 in WO 02/02606. {GenBank accession number: gi|4376849|gb|AAD18697.1| 'CPn0557'; SEQ ID NO: 17 below}. Preferred OmcB proteins for use with the invention
 55 comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 17; and/or (b) which is a fragment of at least n

consecutive amino acids of SEQ ID NO: 17, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These OmcB proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 17. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 17. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 17. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain)..

SEQ ID No 17

1	MSKLIRRVVT	VLALTSMA SC	FASGGIEAAV	AESLITKIVA	SAETKPAPVP
51	MTAKKVRLVR	RNKQPVEQKS	RGAFCDKEFY	PCEEGRCQPV	EAQQESC YGR
101	LYSVKVNDDC	NVEICQSVPE	YATVGSPYPI	EILAIGKKDC	VDVVITQQLP
151	CEAEFVSSDP	ETTP TSDGKL	VWKIDRLGAG	DKCKITVWVK	PLKEGCCFTA
201	ATVCACPELR	SYTKCGQPAI	CIKQEGPDCA	CLRCPVCYKI	EVVNTGSAIA
251	RNVTVDNPVP	DGYSHASGQR	VLSFNLGDMR	PGDKKVFTVE	FCPQRRGQIT
301	NVATVTYCGG	HKCSANVTTV	VNEPCVQVNI	SGADWSYVCK	PVEYSISVSN
351	PGDLVLHDVV	IQDTLP SGVT	VLEAPGGEIC	CNKVWVRIKE	MCPGETLQFK
401	LVVKAQVPGR	FTNQVA VTSE	SNCGTCTSCA	ETTTHWKGLA	ATHMCVLDTN
451	DPICVGENTV	YRICVTNRGS	AEDTNVSLIL	KFSKELQPIA	SSGPTKGTIS
501	GNTVVFDALP	KLGSKESVEF	SVTLKGIAPG	DARGEAILSS	DTLTSPVSDT
551	ENTHVVY*				

(18) MurG peptidoglycan transferase protein (CPn0904)

One example of a 'MurG' protein is disclosed as SEQ ID NO^s: 107 & 108 in WO 02/02606. {GenBank accession number: gi|4377224|gb|AAD19042.1| 'CPn0904'; SEQ ID NO: 18 below}. Preferred MurG proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 18; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 18, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These MurG proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 18. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 18. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 18. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain). The MurG may be lipidated e.g. with undecaprenyl.

SEQ ID No 18

1	MMKKIRKVAL	AVGGSGGHIV	PALSVKEAFS	REGIDVLLLG	KGLKNHPSLQ
51	QGISYREIPS	GLPTVLNPIK	IMSRTL SLCS	GYLKARKELK	IFDPDLVIGF
101	GSYHSLP VLL	AGLSHKIPLF	LHEQNLVPGK	VNQLFSRYAR	GIGVNFSPVT
151	KHFRCPAEEV	FLPKRSFSLG	SPMMKRCTNH	TPTICVVGGS	QGAQILN TCV
201	PQALVKLVNK	YPNMYVHHIV	GPKSDVMKVQ	HVYNRGEVLC	CVKPFEEQLL
251	DVLLAADLVI	SRAGATILEE	ILWAKVPGIL	IPYPGAYGHQ	EVNAKFFVDV
301	LEGGTMILEK	ELTEKLLVEK	VTFALDSHNR	EKQRNSLAAY	SQQRSTKTFH
351	AFICECL*				

(19) CPn0186 (Hypothetical)

One example of a hypothetical protein is set forth as SEQ ID NO: 19 below}.

- (GenBank Accession No. GI:4376456; AAD18339.1). Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 19; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 19, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 19. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 19. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 19. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 19

20
 1 MSSPVNNTPS APNIPIPAPT TPGIPTTKPR SSFIEKVIIIV AKYILFAIAA TSGALGTILG
 61 LSGALTPGIG IALLVIFVVS MVLLGLILKD SISGGEERRL REEVSRFTSE NQRLTVITTT
 121 LETEVKDLKA AKDQLTLEIE AFRNENGNLK TTAEDLEEQV SKLSEQLEAL ERINQLIQAN
 181 AGDAQEISSE LKKLISGWDS KVVEQINTSI QALKVLLGQE WVQEAQTHVK AMQEIQIALQ
 25 241 AEILGMHNQS TALQKSVENL LVQDQALTRV VGELLESENK LSQACSALRQ EIEKLAQHET
 301 SLQQRIDAML AQEQNLAEQV TALEKMKQEA QKAESEFIAC VRDRTEGRRE TTPPTTPVVE
 361 GDESQEEDEG GTPPVSQPSS PVDRATGDGQ

(20) FliY Glutamine Binding Protein (CPn0604)

- 30 One example of a hypothetical protein is set forth as SEQ ID NO^s: 11 & 12 in WO 02/02606. {GenBank accession number: gi|4376900|gb|AAD18743.1| 'CPn0604'; SEQ ID NO: 20 below}. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 20; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 20, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 20. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 20. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 20. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 20

50
 1 MKIKFSWKVN FLICLLAVGL IFFGCSRVRK EVLVGRDATW FPKQFGIYTS
 51 DTNAFLNDLV SEINYKENLN INIVNQDWVH LFENLDDKKT QGAFTSVLPT
 101 LEMLEHYQFS DPILLTGPVL VVAQDSPYQS IEDLKGRLLG VYKFDSSVLV
 151 AQNIPDAVIS LYQHVPIALE ALTSNCYDAL LAPVIEVTAL IETAYKGRLL
 201 IISKPLNADG LRLAILKGTN GDLLEGFNAG LVKTRRSKY DAIKQRYRLP

The immunogenicity of other *Chlamydia pneumoniae* antigens may be improved by combination with two or more *Chlamydia pneumoniae* antigens from either the first antigen group or the second antigen group or the third antigen group. Such other *Chlamydia pneumoniae* antigens include a fourth antigen group consisting one or more members of the PMP family. These antigens are referred to herein as the "fourth antigen group". Each of the *Chlamydia pneumoniae* antigens of the fourth antigen group is described in more detail below.

Fourth Antigen Group

(21) Polymorphic Membrane Proteins (PMP)

A family of twenty one *Chlamydia pneumoniae* genes encoding predicted polymorphic membrane proteins (PMP) have been identified (*pmp1* to *pmp21*).

Pmp1 (CPn0005)

One example of a *Pmp1* protein is set forth as SEQ ID NO^s: 41 & 42 in WO 02/02606. {GenBank accession number: gi|4376260|gb|AAD18163.1 'CPn0005'; SEQ ID NO: 21 below}.

SEQ ID No 21

1	MRFSLCGFPL	VFSFTLLSVF	DTSLSATTIS	LTPEDSFHGD	SQNAERSYNV
51	QAGDVYSLTG	DVSISNVDNS	ALNKACFNV	SGSVTFAGNH	HGLYFNNISS
101	GTTKEGAVLC	CQDPQATARF	SGFSTLSFIQ	SPGDIKEQGC	LYSKNALMLL
151	NNYVVRFEQN	QSKTKGGAIS	GANVTIVGNY	DSVSFYQNAA	TFGGAIHSSG
201	PLQIAVNQAE	IRFAQNTAKN	SGGGALYSDG	DIDIDQNAYV	LFRENEALTT
251	AIGKGGAVCC	LPTSGSSTPV	PIVTFSDNKQ	LVFERNHSIM	GGGAIYARKL
301	SISSGGPTLF	INNISYANSQ	NLGGAIADT	GGEISLSAEK	GTITFQGNRT
351	SLPFLNGIHL	LQNAKFLKLQ	ARNGYSIEFY	DPITSEADGS	TQLNINGDPK
401	NKEYTGTLIF	SGEKSLANDP	RDFKSTIPQN	VNLSAGYLV	KEGAEVTVSK
451	FTQSPGSHLV	LDLGTKLIAS	KEDIAITGLA	IDIDSLSSSS	TAAVIKANTA
501	NKQISVTDSI	ELISPTGNAY	EDLRMRNSQT	FPLLSLEPGA	GGSVTVTAGD
551	FLPVSPHYGF	QGNWKLAWTG	TGNKVGEFFW	DKINYKPRPE	KEGNLVPNIL
601	WGNVAVDVRSL	MQVQETHASS	LQTDRLGLWID	GIGNFFHVSA	SEDNIRYRHN
651	SGGYVLSVNN	EITPKHYTSM	AFSOLFSDRK	DYAVSNNEYR	MYLGSYLYQY
701	TTSILGNIFRY	ASRNPVNVVG	ILSRRLQNP	LMIFHFLCAY	GHATNDMKT
751	YANFPMVKNS	WRNNCWAIEC	GGSMPLLVFE	NGRLFQGAIP	FMKLQLVYAY
801	QGDFKETTDAD	GRRFSNGSLT	SISVPLGIRF	EKLALSQDVL	YDFSFSYIPD
851	IFRKDPSCEA	ALVISGDSWL	VPAAHVSRHA	FVGSGTGRYH	FNDYTELLCR
901	GSIECRPHAR	NYNINCGSKF	RF*		

Pmp 4 (CPn0017)

One example of a *Pmp 4* protein is designated SEQ ID NO: 22. The sequence for *pmp4* protein can be found at AE001587.1 GI:4376271.

Pmp 6 (CPn0444)

One example of a *Pmp 6* protein is set forth as SEQ ID NO^s 31 & 32 in WO 02/02606. {GenBank accession number: gi|4376727|gb|AAD18588.1| 'CPn0444'; SEQ ID NO: 23 below}.

SEQ ID No 23

1	MKYSLEWLLT	SSALVFSLHP	LMAANTDLSS	SDNYENGSSG	SAAFTAKETS
51	DASGTTYTLT	SDVSITNVSA	ITPADKSCFT	NTGGALSFGV	ADHSLVLQTI

5 101 ALTHDGAAIN NTNTALSFSG FSSLLIDSAP ATGTSGGKGA ICVTNTEGGT
 151 ATFTDNASVT LQKNTSEKDG AAVSAYSIDL AKTTTAAALLD QNTSTKNGGA
 201 LCSTANTTVQ GNSGTVTFSS NTATDKGGGI YSKEKDSTLD ANTGVVTFKS
 251 NTAKTGGAWS SDDNLALTGN TQVLFQENKT TGSAAQANNP EGC GGAICCY
 301 LATATDKTGL AISQNQEMSF TSNTTTANGG AIYATKCTLD GNTTLTFDQN
 351 TATAGCGGAI YTETEDFSLK GSTGTVTFST NTAKTGGALY SKGNSSLTGN
 401 TNLLFSGNKA TGPSNSSANQ ECGGAILAF IDSGSVSDKT GLSIANNQEV
 451 SLTSNAATVS GGAIYATKCT LTGNGSLTFD GNTAGTSGGA IYTETEDFTL
 10 501 TGSTGTVTF S TNTAKTGGAL YSKGNNSLSG NTNLLFSGNK ATGPSNSSAN
 551 QEGCGGAILS FLESASVSTK KGLWIEDNEN VSLSGNTATV SGGAIYATKC
 601 ALHGNTTLTF DGNTAETAGG AIYTETEDFT LTGSTGTVTF STNTAKTAGA
 651 LHTKGNTSFT KNKALVFSGN SATATATTTT DQEGCGGAIL CNISESDIAT
 701 KSLTLTENES LSFINTAKR SGGGIYAPKC VISGSESINF DGNTAETSGG
 751 AIYSKNLSIT ANGPVSFTNN SGGKGGAIYI ADSGELSLEA IDGDITFSGN
 15 801 RATEGTSTPN SIHLGAGAKI TKLAAAPGHT IYFYDPITME APASGGTIEE
 851 LVINPVVKAI VPPPQPKNGP IASVPVPVA PANPNTGTIV FSSGKLPSQD
 901 ASIPANTTTI LNQKINLAGG NVVLKEGATL QVYSFTQQPD STVFMDAGTT
 951 LETTTTNTND GSIDLKNLSV NLDALDGKRM ITIAVNSTSG GLKISGDLKF
 20 1001 HNNESGFYDN PGLKANLNL PFLDLSSTSGT VNLDDFNPIP SSMAAPDYGY
 1051 QGSWTLVPKV GAGGKVTLVA EWQALGYTPK PELRATLVPN SLWNAYVNIH
 1101 SIQQEIATAM SDAPSHPGIW IGGIGNAFHQ DKQKENAGFR LISRGYIVGG
 1151 SMTTPQEYTF AVAFSOLFQK SKDYVVSDIK SQVYAGSLCA QSSYVIPLHS
 1201 SLRRHVLSKV LPELPGETPL VLHGQVSYGR NHHNMTTKLA NNTQKSDWD
 1251 SHSFAVEVGG SLPVDLNYRY LTSYSPYVKL QVSVNQKGF QEVAADPRIF
 25 1301 DASHLVNVI PMGLTFKHES AKPPSALLLT LGYAVDAYRD HPHCLTSLTN
 1351 GTSWSTFATN LSRQAFFAEA SGHLKLLHGL DCFASGSC EL RSSRSYNAN
 1401 CGTRYSF*

30 *Pmp 7 (CPn0445)*

One example of a Pmp 7 protein is set forth as SEQ ID NO^s 153 & 154 in WO 02/02606. {GenBank accession number: gi|4376728|gb|AAD18589.1| 'CPn0445'; SEQ ID NO: 24 below}.

35 *SEQ ID No 24*

1 MKSSVSWLFF SSIPLFSSLS IVAAEVTLD SNN SYDGSNG TTFTVFSTTD
 51 AAAGTTYSL SDVSFQNAGA LGIPLASGCF LEAGGDLTFQ GNQHALKFAF
 101 INAGSSAGTV ASTSAADKNL LFNDFSRLSI ISCP SLLLSP TGQCALKSVG
 40 151 NLSLTGNSQI IFTQNFSSDN GGVINTKNFL LSGTSQFASF SRNQAF TGKQ
 201 GGVVYATGTI TIENSPGIVS FSQNLAKGSG GALYSTDNCS ITDNFQVIFD
 251 GNSAWAAQA QGGAICCTTT DKTVTLTGNK NLSFTNNTAL TYGGAISGLK
 301 VSISAGGPTL FQSNISGSSA GQGGGGAINI ASAGELALSA TSGDITFNNN
 351 QVTNGSTSTR NAINIIDTAK VTSIRAATGQ SIYFYDPITN PGTAASTDTL
 45 401 NLNLADANSE IEYGGAI VFS GEKLSPT EKA IANVTSTIR QPAVLARGDL
 451 VLRDGVTVTF KDLTQSPGSR ILM DGGTTLS AKEANLSLNG LAVNLSSL DG
 501 TNKAALKTEA ADKNISLSGT IALIDTEGSF YENHNLKSAS TYPLLELT TA
 551 GANGTITLGA LSTLT LQEP E THYGYQGNWQ LSWANATSSK IGSINWTRTG
 601 YIPSPERKSN LPLNSLWGNF IDIRSINQLI ETKSSGEPFE RELWLSGIAN
 50 651 FFYRDSMPTR HGFRHISGGY ALGITATTPA EDQLTFAFCQ LFARDRNHIT
 701 GKNHGD TYGA SLYFHHT EGL FDIANFLWGK ATRAPWVLSE ISQIIPLSFD
 751 AKFSYLHTDN HMKTY YTDNS IIKG SWRND A FCADLGASLP FVISVPYLLK
 801 EVEPFVKVQY IYAHQQDFYE RHAEGRAF NK SELINVEIPI GVTFERDSKS
 851 EKGTYDLTLM YILDAYRRNP KCQTS LIASD ANWMAYGTNL ARQGF SVRAA
 55 901 NHFQVNP HME IFGQFAFEVR SSSRNYNTNL GSKFCF*

Pmp 8 (CPn0446)

One example of a Pmp 8 protein is set forth as SEQ ID NO^s 45 & 46 in WO 02/02606. {GenBank accession number: gi|4376729|gb|AAD18590.1| 'CPn0446'; SEQ ID NO: 25 below}.

SEQ ID No 25

	1	<u>MKIPLHKLLI</u>	<u>SSTLVTPILL</u>	<u>SIATYGADAS</u>	<u>LSPTDSFDGA</u>	<u>GGSTFTPST</u>
	51	ADANGTNYVL	SGNVYINDAG	KGTALTGCCF	TETTGDLTFT	GKGYSFSFNT
5	101	VDAGSNAGAA	ASTTADKALT	FTGFSNLSFI	AAPGTTVASG	KSTLSSAGAL
	151	NLTDNGTILF	SQNVSNNEANN	NGGAITTKTL	SISGNTSSIT	FTSNSAKKLG
	201	GAIYSSAAAS	ISGNTGQLVF	MNNKGETGGG	ALGFEASSSI	TQNSSLFFSG
	251	NTATDAAGKG	GAIYCEKTGE	TPTLTISGNK	SLTFAENSSV	TQGGAIKAHG
	301	LDLSAAGPTL	FSNNRCGNTA	AGKGAIAIA	DSGSLSLSAN	QGDITFLGNT
10	351	LTSTSAPTST	RNAIYLGSSA	KITNLRAAQG	QSIYFYDPIA	SNTTGASDVL
	401	TINQPDSNSP	LDYSGTIVFS	GEKLSADEAK	AADNFTSILK	QPLALASGTL
	451	ALKGNVELDV	NGFTQTEGST	LLMQPGTKLK	ADTEAISLTK	LVVDLSALEG
	501	NKSVSIETAG	ANKTITLTSP	LVFQDSSGNF	YESHTINQAF	TQPLVVFTAA
	551	TAASDIYIDA	LLTSPVQTPE	PHYGYQGHWE	ATWADTSTAK	SGTMTWVTG
15	601	YNPNPERRAS	VVPDSLWASF	TDIRTLQQIM	TSQANSIYQQ	RGLWASGTAN
	651	FFHKDKSGTN	QAFRHKSYGY	IVGGSAAEDFS	ENIFSVAFQC	LFGKDKDLFI
	701	VENTSHNYLA	SLYLQHRAFL	GGLPMPSFGS	ITDMLKDIPL	ILNAQLSYSY
	751	TKNDMDTRYT	SYPEAQGSWT	NNSGALELGG	SLALYLPKEA	PFFQGYFPFL
20	801	KFQAVYSRQQ	NFKESGAEAR	AFDDGDLVNC	SIPVGIRLEK	ISEDEKNNFE
	851	ISLAYIGDVY	RKNPRSRTSL	MVSGASWTSL	CKNLARQAFL	ASAGSHLTLS
	901	PHVELSGEAA	YELRGSAGHIY	NVDCGLRYSF	*	

Pmp 9 (CPn0447)

One example of a Pmp 9 protein is set forth as SEQ ID NO^s 33 & 34 in WO 02/02606. {GenBank accession number: gi|4376731|gb|AAD18591.1| 'CPn0447'; SEQ ID NO: 26 below}.

SEQ ID No 26

30	1	<u>MKSSLHWFLI</u>	<u>SSSLALPLSL</u>	<u>NFSAFAAVVE</u>	<u>INLGPTNSFS</u>	<u>GPGTYTPPAQ</u>
	51	TTNADGTIYN	LTGDVSITNA	GSPTALTASC	FKETTGNLSF	QGHGYQFLLQ
	101	NIDAGANCTF	TNTAANKLLS	FSGFSYLSLI	QTTNATTGTG	AIKSTGACSI
	151	QSNYSCYFGQ	NFSNDNGGAL	QGSSISLSLN	PNLTFAKNKA	TQKGGALYST
	201	GGITINNTLN	SASFSENTAA	NNGGAIYTEA	SSFISSNKAI	SFINNSVTAT
35	251	SATGGAIYCS	STSAPKPVLT	LSDNELNFI	GNTAITSGGA	IYTDNLVLSS
	301	GGPTLFKNNS	AIDTAAPLGG	AIAIADSGSL	SLSALGGDIT	FEGNTVVKGA
	351	SSSQTTTRNS	INIGNTNAKI	VQLRASQGNT	IYFYDPITTS	ITAALSDALN
	401	LNGPDLAGNP	AYQGTIVFSG	EKLSEAEAAE	ADNLKSTIQQ	PLTLAGGQLS
	451	LKSGVTLVAK	SFSQSPGSTL	LMDAGTTLET	ADGITINNLV	LNVDLSKETK
40	501	KATLKATQAS	QVTLTSGSL	LVDPSPGNVYE	DVSWNNPQVF	SCLTLTADDP
	551	ANIHITDLAA	DPLEKNPIHW	GYQGNWALSW	QEDTATKSKA	ATLTWTKTGY
	601	NPNPERRGTL	VANTLWGSFV	DVRSIQQLVA	TKVRQSQETR	GIWCEGISNF
	651	FHKDSTKINK	GFRHISAGYV	VGATTTLASD	NLITAAFCQL	FGKDRDHFIN
	701	KNRASAYAAS	LHLQHLATLS	SPSLRLRYLPG	SESEQPVLEF	AQISYIYSKN
45	751	TMKTYTQAP	KGESSWYNDG	CALELASSLP	HTALSHEGLE	HAYFPFIKVE
	801	ASYIHQDSFK	ERNITLVRSE	DSGDLINVS	PIGITFERFS	RNERASYEAT
	851	VIYVADVYRK	NPDCTTALLI	NNTSWKTTGT	NLSRQAGIGR	AGIFYAFSPN
	901	LEVTSNLSME	IRGSSRSYNA	DLGGKFQF*		

50 Pmp 11 (CPn0451)

One example of a Pmp 11 protein is set forth as SEQ ID NO^s 115 & 116 in WO 02/02606. {GenBank accession number: gi|4376733|gb|AAD18593.1| 'CPn0451'; SEQ ID NO: 27 below}.

55 SEQ ID No 27

	1	<u>MKTSIPWVLV</u>	<u>SSVLAFSCHL</u>	<u>QSLANEELLS</u>	<u>PDDSFNGNID</u>	<u>SGTFTPKTSA</u>
	51	TTYSLTGDVF	FYEPGKGTPL	SDSCFKQTTD	NLTFLNGHGS	LTFGFIDAGT
60	101	HAGAAASTTA	NKNLTFSGFS	LLSFDSSPST	TVTTGQGTLS	SAGGVNLENI
	151	RKLVVAGNFS	TADGGAIKGA	SFLLTGTSGD	ALFSNNSSST	KGGAIATTAG
	201	ARIANNTGYV	RFLSNIASTS	GGAIDDEGTS	ILSNNKFLYF	EGNAAKTTGG
	251	AICNPKASGS	PELIISNNKT	LIFASNVAET	SGGAIHAKKL	ALSSGGFTEF

5 301 LRNNVSSATP KGGAISIDAS GELSLSAETG NITFVRNTLT TTGSTDTPKR
 351 NAINIGSNGK FTELRAAKNH TIFYDYDPTS EGTSSDVLKI NNGSAGALNP
 401 YQGTILFSGE TLTADDELKVA DNLKSSFTQP VSLSGGKLLL QKGVTLLEST
 451 FSQEAGSLLG MDSGTTLSTT AGSITITNLG INVDSLGLKQ PVSLTAKGAS
 501 NKVIVSGKLN LIDIEGNIYE SHMFSDHQLF SLLKITVDAD VDTNVDISSL
 551 IPVPAEDPNS EYGFQGWNV NWTDTATNT KEATATWTKT GFVPSPERKS
 601 ALVCNTLWGV FTDIRSLQQL VEIGATGMEH KQGFVSSMT NFLHKTGDEN
 651 RKGFRHTSGG YVIGGSAHTP KDDLFTFAFC HLFARDKDCF IAHNNSRTYG
 10 701 GTLFFKHSHT LQPQNYLRLG RAKFSESAIE KFPREIPLAL DVQVSFSHSD
 751 NRMETHYTSL PESEGSWSNE CIAGGIGLDL PFVLSNPHPL FKTFIPQMKV
 801 EMVYVSQNSF FESSSDGRGF SIGRLLNLSI PVGAKFVQGD IGDSYTYDLS
 851 GFFVSDVYRN NPQSTATLVM SPDSWKIRGG NLSRQAFLLR GSNNYVYNSN
 901 CELFGHYAME LRGSSRYNV DVGTKLRF*

15 *Pmp 12 (CPn0452)*

One example of a Pmp 12 protein is set forth as SEQ ID NO^s 51 & 52 in WO 02/02606. {GenBank accession number: gi|4376735|gb|AAD18594.1 'CPn0452'; SEQ ID NO: 28 below).

20 *SEQ ID No 28*

25 1 MTILRNFLTC SALFLALPAA AQVVYLHESD GYNGAINNKS LEPKITCYPE
 51 GTSYIFLDDV RISNVKHDQE DAGVFINRSG NLFFMGNRCN FTFHNLMTG
 101 FGAAISNRVG DTTLTLSNFS YLAFTSAPLL PQQGAIYSL GSVMIENSEE
 151 VTFCGNYSSW SGAAIYTPYL LGSKASRPSV NLSGNRYLVF RDNVSQGYGG
 201 AISTHNLTLT TRGPSCFENN HAYHDVNSNG GAIAIAPGGS ISISVKSGDL
 251 IFKGNTASQD GNTIHNSIHL QSGAQFKNLR AVSESGVYFY DPISHSESHK
 301 ITDLVINAPE GKETYEGTIS FSGLCLDDHE VCAENLTSTI LQDVTLAGGT
 351 LSLSDGVTLQ LHSFKQEASS TLTMSPGTTL LCSGDARVQN LHILIEDTDN
 30 401 FVPVRIRAED KDALVSLEKL KVAFEAYWSV YDFPQFKEAF TIPLLELLGP
 451 SFDSLILGET TLERTQVTTE NDAVRGFWSL SWEYPPSLD KDRRITPTKK
 501 TVFLTWNPEI TSTP*

Pmp 13 (CPn0453)

35 One example of a Pmp 13 protein is set forth as SEQ ID NO^s 3 & 4 in WO 02/02606. {GenBank accession number: gi|4376736|gb|AAD18595.1 'CPn0453'; SEQ ID NO: 29 below}.

SEQ ID No 29

40 1 MKTSIRKFLI STTLAPCFAS TAFTVEVIMP SENFDGSSGK IFPYTTLSDP
 51 RGTLCIFSGD LYIANLDNAI SRTSSSCFSN RAGALQILGK GGVFSFLNIR
 101 SSADGAAISS VITQNPELCP LSFSGFSQMI FDNCELSLSD TSASNVIPHA
 151 SAIYATTPML FTNNDLILFQ YNRSAGFGAA IRGTSITIEN TKKSLLFNGN
 45 201 GSISNGGALT GSAAINLINN SAPVIFSTNA TGIYGGAIYL TGGSMITSGN
 251 LSGVLVFNNS SRSGGAIYAN GNVTFNNNSD LTFQNTASP QNSLPAPTPP
 301 PTPPAVTPLL GYGGAIFCTP PATPPPTGVS LTISGENSVT FLENIASEQG
 351 GALYGKKISI DSNKSTIFLG NTAGKGGAIA IPESGELSLS ANQGDILFNK
 401 NLSITSGTPT RNSIHFGKDA KFATLGATQG YTLYFYDPIT SDDLASAASAA
 50 451 ATVVVNPKAS ADGAYSGTIV FSGETLTATE AATPANATST LNQKLELEGG
 501 TLALRNGATL NVHNFTQDEK SVVIMDAGTT LATTNGANNT DGAITLNKLV
 551 INLDSLDTGK AAVNVQSTN GALTISGTLG LVKNSQDCCD NHGMFNKDLQ
 601 QVPILELKAT SNTVTTTDFS LGTNGYQQSP YGYQGTWEFT IDTTTHTVTG
 651 NWKKTGYLPH PERLAPLIPN SLWANVIDLR AVSQASAADG EDVPGKQLSI
 55 701 TGITNFFHAN HTGDARSYRH MGGGYLINTY TRITPDAAALS LGFGQLFTKS
 751 KDYLVGHGHS NVYFATVYSN ITKSLFGSSR FFSGGTSRVT YSRSNEKVKT
 801 SYTKLPKGRC SWSNNCWLGE LEGNLPITLS SRILNLKQII PFVKAEVAYA
 851 THGGIQENTP EGRIFGHGHL LNVAVPVGVR FGKNSHNRPD FYTIIIVAYAP
 901 DVYRHNPD CD TTPPINGATW TSIGNNLTRS TLLVQASSHT SVNDVLEIFG
 60 951 HCGCDIRRTS RQYTLDIGSK LRF*

Pmp 14 (CPn0454)

One example of a Pmp 14 protein is set forth as SEQ ID NO^s 35 & 36 in WO 02/02606. {GenBank accession number: gi|4376737|gb|AAD18596.1 'CPn0454'; SEQ ID NO: 30 below}.

5 *SEQ ID No 30*

	1	<u>MPLSFKSSSF</u>	<u>CLLACLCSAS</u>	<u>CAFAETRLGG</u>	NFVPPITNQG	EEILLTSDFV
	51	CSNFLGASFS	SSFINSSSNL	SLLGKGLSLT	FTSCQAPTNS	NYALLSAAET
10	101	LTFKNFSSIN	FTGNQSTGLG	GLIYGKDIVF	QSIKDLIFTT	NRVAYSPASV
	151	TTSATPAITT	VTTGASALQP	TDSLTVENIS	QSIKFFGNLA	NFGSAISSSP
	201	TAVVKFINNT	ATMSFSHNFT	SSGGGVIYGG	SSLLFENNSG	CIIFTANSCV
	251	NSLKGVTSS	GTALGSGGA	ICIPTGTFEL	KNNQKCTFS	YNGTPNDAGA
	301	IYAETCNIVG	NQGALLDSN	TAARNGGAIC	AKVLNIQGRG	PIEFSRNRAE
15	351	KGGAIFIGPS	VGDPKQTST	LILASEGDI	AFQGNMLNTK	PGIRNAITVE
	401	AGGEIVLSA	QGGSLVFDY	PITHSLPTTS	PSNKDITINA	NGASGSVVFT
	451	SKGLSSTELL	LPANTTTILL	GTVKIASGEL	KITDNAVNV	LGFATQSGSQ
	501	LTLGSGGTLG	LATPTGAPAA	VDFTIGKLAF	DPFSFLKRDF	VSASVNAGTK
	551	NVTLTGALVL	DEHDVTDLYD	MVSLQTPVAI	PIAVFKGATV	TKTGFPDGEI
20	601	ATPSHYGYQG	KWSYTWSRPL	LIPAPDGGFP	GGSPSANTL	YAVWNSDTLV
	651	RSTYILDPER	YGEIVSNSLW	ISFLGNQAFS	DILQDVLLID	HPGLSITAKA
	701	LGAYVEHTPR	QGHEGFSGRY	GGYQAALSMN	YTDHTTLGLS	FGQLYGKTNA
	751	NPYDSRCSEQ	MYLLSFFGQF	PIVTQKSEAL	ISWKAAYGYS	KNHLNTTYLR
	801	PDKAPKSQGG	WHNNSYYVLI	SAEHPFLNWC	LLTRPLAQAW	DLSGFISAEF
25	851	LGGWQSKFTE	TGDLQRSFSR	GKGYNVSLPI	GCSSQWFTEP	KKAPSTLTIK
	901	LAYKPDIIYRV	NPHNIVTVVS	NQESTSISGA	NLRRHGLEFVQ	IHDVVDLTED
	951	TQAFLNYTFD	GKNGFTNHRV	STGLKSTF*		

Pmp 15 (CPn0466)

One example of a Pmp 15 protein is set forth as SEQ ID NO^s 5 & 6 in WO 02/02606. {GenBank accession number: gi|4376751|gb|AAD18608.1 'CPn0466'; SEQ ID NO: 31 below}.

SEQ ID No 31

35	1	<u>MRFFCFGMLL</u>	<u>PFTFVLANEG</u>	<u>LQLPLETYIT</u>	LSPEYQAAPQ	VGFTHNQNQD
	51	LAIVGNHNDF	ILDYKYYRSN	GGALTCKNLL	ISENIGNVFF	EKNVCPNSGG
	101	AIYAAQNCTI	SKNQNYAFTT	NLVSDNPTAT	AGSLLGGALF	AINCSITNNL
	151	GQGTFFVDNLA	LNKGGALYTE	TNLSIKDNKG	PIIHKQNRAL	NSDSLGGGIY
40	201	SGNSLNIEN	SGAIQITSNS	SGSGGGIFST	QTLTISSNKK	LIEISENSAF
	251	ANNYGSNFNP	GGGGLTTTFC	TILNNREGVL	FNNNQSQSNG	GAIHAKSIII
	301	KENGPVYFLN	NTATRGGALL	NLSAGSGNGS	FILSADNGDI	IFNNNTASKH
	351	ALNPPYRNAI	HSTPNMNLQI	GARPGYRVLF	YDPIEHELPS	SFPIELNFET
	401	GHTGTVLFSG	EHVHQNFTE	MNFFSYLRNT	SELRQGVLA	EDGAGLACYK
	451	FFQRGGTLLL	GQGAUITTAG	TIPTPSSTPT	TVGSTITLNH	IAIDLPSILS
45	501	FQAQAPKIWI	YPTKTGSTYT	EDSNPTITIS	GTLTLRNSNN	EDPYDSL DLS
	551	HSLEKVPLLY	IVDVAQKIN	SSQLDLSTLN	SGEHYGYQGI	WSTYWVETTT
	601	ITNPTSLLGA	NTKHKL LLYAN	WSPLGYRPHP	ERRGEFITNA	LWQSAYTALA
	651	GLHSLSSWDE	EKGHAASLQG	IGLLVHQKDK	NGFKGFRSHM	TGYSATTEAT
	701	SSQSPNFSLG	FAQFFSKAKE	HESQNSTSSH	HYFSGMCIEN	TLFKEWIRLS
50	751	VSLAYMFTSE	HTHTMYQGLL	EGNSQGSFHN	HTLAGALSCV	FLPQPHGESL
	801	QIYPFITALA	IRGNLA AFQE	SGDHAREFSL	HRPLTDVSLP	VGIRASWKNH
	851	HRVPLVWLTE	ISYRSTLYRQ	DPELHSLKLLI	SQGTWTTQAT	PVTYNALGIK
	901	VKNTMQVFPPK	VTLSLDYSAD	ISSSTLSHYL	NVASMRF*	

55 *Pmp 16 (CPn0467)*

One example of a Pmp 16 protein is set forth as SEQ ID NO^s 7 & 8 in WO 02/02606. {GenBank accession number: gi|4376752|gb|AAD18609.1| 'CPn0467'; SEQ ID NO: 32 below}.

SEQ ID No 32

60	1	MFGMTPAVYS	LQTDLSLEKFA	LERDEEFRTS	FPLLDLSLSTL	TGFSPITTFV
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51 GNRHNSSQDI VLSNYKSIDN ILLWTSAGG AVSCNNFLLS NVEDHAFFSK
 101 NLAIGTGGAI ACQGACTITK NRGPLIFFSN RGLNNASTGG ETRGGAIACN
 151 GDFTISQNGG TFYFVNNSVN NWGGALSTNG HCRIQSNRAP LLFFNNTAPS
 201 GGGALRSENT TISDNTRPIY FKNNGCNGG AIQTSVTVAI KNNSGSVIFN
 251 NNTALSGSIN SGNGSGGAIY TTNLSIDNPN GTILFNNNYC IRDGGAICTQ
 301 FLTIKNSGHV YFTNNQGNWG GALMLLQDST CLLFAEQGNI AFQNNNEVFLT
 351 TFGRYNAIHC TPNSNLQLGA NKGYTTFDFF PIEHQHPTTN PLIFNPNANH
 401 QGTILFSSAY IPEASDYENN FISSSKNTSE LRNGVLSIED RAGWQFYKFT
 451 QKGGILKLGH AASIATTANS ETPSTSVGSQ VIINNLAJNL PSILAKGKAP
 501 TLWIRPLQSS APFTEDNNPT ITLSGPLTLL NEENRDPYDS IDLSEPLQNI
 551 HLLSLSDVTA RHINTDNFHP ESLNATEHYG YQGIWSPYVW ETITTTNNAS
 601 IETANTLYRA LYANWTPLGY KVNPEYQGDJ ATTPPLWQSFH TMFSLLRSYN
 651 RTGDSDIERP FLEIQGIADG LFFVHQNSIPG APGFRIQSTG YSLQASSETS
 701 LHQKISLGFA QFFTRTKEIG SSNNVSAHNT VSSLYVELPW FQEAFASTSTV
 751 LAYGYGDHHL HSLHPSHQEQ AEGTCYSHTL AAAIGCSFPW QQKSYLHLSP
 801 FVQAIAIRSH QTAEEIGDN PRKFVSQKPF YNLTPLPLGIQ GKWQSKFHPV
 851 TEWTLLELSYQ PVLYQQNPQI GVTLLASGGS WDILGHNYVR NALGYKVHNO
 901 TALFRSLDLF LDYQGSVSSS TSTHHLQAGS TLKF*

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Pmp 18 (CPn0471)

One example of a Pmp 18 protein is set forth as SEQ ID No 33 below {GenBank accession number: gi|4376753|gb|AAD18610.1| 'CPn0471'.

25 ***SEQ ID No 33***

1 MQNNRSLSKS SFFVGALILG KTTILLNATP LSDYFDNQAN QLTTLFPLID TLTNMTYPYSH
 61 RATLFGVRDD TNQDIVLDHQ NSIESWFENF SQDGGALSCK SLAITNTKNQ ILFLNSFAIK
 121 RAGAMYVNGN FDLSENHSGI IFSGNLSFPN ASNFADTCTG GAVLCSKNVT ISKNQGTAYF
 181 INNKAASSGG AIQAAIINIK DNTGPCLFFN NAAGGTAGGA LFANACRIEN NSQPIYFLNN
 241 QSGLGGAIRV HQECILTKNT GSVIFNNNFA MEADISANHS SGGAIYCISC SIKDNPGIAA
 301 FDNNTAARDG GAICTQSLTI QDSGPVYFTN NQGTWGGAIM LRQDGAFTLF ADQGDIIFYN
 361 NRHFKDTFSN HVSVNCTRNV SLTVGASQGH SATFYDPILQ RYTIQNSIQK FNPNPHEHGT
 421 ILFSSTYIPD TSTSRDDFIS HFRNHIGLYN GTLALEDRAE WKVYKFDQFG GTLRLGSRV
 481 FSTTDEEQSS SSVGSVININ NLAINLPSIL GNRVAPKLWI RPTGSSAPYS EDNNPIINLS
 541 GPLSLLDEN LDYDTADLA QPIAEVPLLY LLDVTAKHIN TDNFYPEGLN TTQHYGYQGV
 601 WSPYWIETIT TSDTSSEDTV NTLHRQLYGD WPTPGYKVN ENKGDIALSA FWQSFHNLFA
 661 TLRVQTQQGQ IAPTASGEAT RLFVHQNSNN DAKGFHMEAT GYSLGTTSTN ASNHSFGVNF
 721 SQLFSNLYES HSDNSVASHT TTVALQINNP WLQERFSTSA SLAYSYSNHH IKASGYSGKI
 781 QTEGKCYSTT LGAALSCSL LQWRSRPLHF TPFQIAIAVR SNQTAFAQESG DKARKFSVHK
 841 PLYNLTVPLG IQSAWESKFR LPTYWNIELA YQPVLYQNP EINVSLESSG SSWLLSGTTL
 901 ARNAIAFKGR NQIFIFPKLS VFLDYQGSVS SSTTTHYLHA GTTFKF

45 ***Pmp 19 (CPn0539)***

One example of a Pmp 19 protein is set forth as SEQ ID No 34 below {GenBank accession number: gi|4376829|gb|AAD18679.1 'CPn0539'; SEQ ID NO: 34 below}.

SEQ ID No 34

50 1 MKQMRLWGFL FLSSFCQVSY LRANDVLLPL SGIHSGEDLE LFTLRSSSPT KTTYSLRKDF
 61 IVCDFAGNSI HKPGAAFLNL KGDLEFFINST PLAALTFKNI HLGARGAGLF SESNVTFKGL
 121 HSLVLENNES WGGVLTTSGL LSFNNNTSVL CQNNISYGP GALLLQGRKS KALFFRDNRG
 181 TILFLKNKAV NQDESHPGYG GAVSSISPGS PITFADNQE LFOENEGELG GAIYNDQGA
 241 TFENNQTTTS FFSNKASFGG AVYSRYCNLY SQWGDTLFTK NAAAKVGGAI HADYVHIRDC
 301 KGSIVFEENS ATAGGAIAN AVCDINAQGP VRFINNSALG LNGGAIYMQA TGSILRLHAN
 361 QGDIEFCGNK VRSQFHSHIN STSNFTNNAI TIQGAFFES LSANEGHRIC FYPPIISATE
 421 NYNSLYINHQ RLLEAGGAVI FSGARLSPEH KKENKNKTSI INQPVRLCSG VLSIEGGAIL
 481 AVRSFYQEGG LLALGPGSKL TTQGKNSEKD KIVITNLGFN LENLDSSDPA EIRATEKASI
 541 EISGVPRVYG HTESFYENHE YASKPYTTSI ILSAKKLVT PSRPEKDIQN LIIAESEYMG
 601 YGYQGSWEFS WSPNDTKEKK TIIASWTPTG EFSLDPKRRG SFIPTTLWST FSGLNIASNI
 661 VNNNYLNNSE VIPLQHLCHF GGPVYQIMEQ NPKQSSNNLL VQHAGHNVA RIPFSFNTIL
 721 SAALTQLFSS SSQONVADKS HAQILIGTVS LNKSQWALS RSSFSYTEDS QVMKHVFPYK
 781 GTSRGSWRNY GWSGSGVMSY AYPKGIRYLK MTPFVDLQYT KLVQNPVET GYDPRYFSS

841 EMTNLSLPIG IALEMRFIGS RSSLFLOVST SYIKDLRRVN PQSSASLVLN HYTWDIQGVF
 901 LGKEALNITL NSTIKYKIVT AYMGISSTQR EGSNLSANAH AGLSLSF

As the Examples demonstrate, we and others have demonstrated (Grimwood et al
 5 (2001), Infection and Immunity 69(4), 2383-2389) using Flow cytometry (FACS)
 analyses and Western Blot analyses that PMP19 does not appear to be surface
 exposed. However, high levels of mRNA expression is nevertheless observed in gene
 microarray analysis of pmp19 (CPn0539).

10 *Pmp 20 (CPn0540)*

One example of a Pmp 20 protein is set forth as SEQ ID NO^s 119 & 120 in WO
 02/02606. {GenBank accession number: gi|4376830|gb|AAD18680.1 'CPn0540';
 SEQ ID NO: 35 below}.

15 *SEQ ID No 35*

	1	<u>MKWL</u> <u>PATAVF</u>	<u>AAVL</u> <u>PALTA</u> <u>F</u>	GDPASVEIST	SHTGSGDPTS	DAALTGFTQS
	51	STETDGTTYT	IVGDITFSTF	TNIPVPVVP	DANDSSSNSS	KGGSSSSGAT
	101	SLIRSSNLHS	DFDFTKDSVL	DLYHLFFPSA	SNTLNPALLS	SSSSGGSSSS
20	151	SSSSSSGSAS	AVVAADPKGG	AAFYSNEANG	TLTFTTDSGN	PGSLTLQNLK
	201	MTGDGAAIYS	KGPLVFTGLK	NLTFTGNESQ	KSGGAAYTEG	ALTQQAIVEA
	251	VTFTGNTSAG	QGGAIYVKEA	TLFNALDSLK	FEKNTSGQAG	GGIYTESTLT
	301	ISNITKSIEF	ISNKASVPAP	APEPTSPAPS	SLINSTTIDT	STLQTRAASA
	351	TPAVAPVAAV	TPTPISTQET	AGNGGAIYAK	QGISISTFKD	LTFKSNSASV
25	401	DATLTVDSST	IGESGGAIFA	ADSIQIQQCT	GTTLEFSGNTA	NKSGGGIYAV
	451	GQVTLEDIAN	LKMTNNTCKG	EGGAIYTKKA	LTINNGAILT	TFSGNTSTDN
	501	GGAIFAVGGI	TLSDLVEVRF	SKNKTGNYS	PITKAASNTA	PVVSSSTTAA
	551	SPAVPAAAAA	PVTNAAKGGA	LYSTEGTLVS	GITSILSFEN	NECQNQGGGA
	601	YVTKTFQCS	SHRLQFTSNK	AADEGGGLYC	GDDVTLTNLT	GKTLFQENSS
30	651	EKHGGGLSLA	SGKSLTMTSL	ESFCLNANTA	KENGGGANVP	ENIVLTFTYT
	701	PTPNEPAPVQ	QPVYGEALVT	GNTATKSGGG	IYTKNAAFSN	LSSVTFDQNT
	751	SSENGGALLT	QKAADKTDCS	FTYITNVNIT	NNTATGNNGG	IAGGKAHFDR
	801	IDNLTVQSNQ	AKKGGGVYLE	DALILEKVIT	GSVSQNTATE	SGGGIYAKDI
	851	QLQALPGSFT	ITDNKVETSL	TTSTNLYGGG	IYSSGAVTLT	NISGTFGITG
35	901	NSVINTATSQ	DADIQGGGIY	ATTSLSINQC	NTPILFSNNS	AATKKTSTTK
	951	QIAGGAIFSA	AVTIENNSQP	IIFLNNSAKS	EATTAATAGN	KDSCGGAIAA
	1001	NSVTLTNNPE	ITFKGNYAET	GGAIGCIDLT	NGSPPRKVSI	ADNGSVLFQD
	1051	NSALNRGGAI	YGETIDISRT	GATFIGNSSK	HDGSAICCST	ALTLAPNSQL
	1101	IFENNKVTET	TATTKASINN	LGAAIYGNNE	TSDVTISLSA	ENGSIFFKNN
40	1151	LCTATNKYCS	IAGNVKFTAI	EASAGKAISF	YDAVNVSTKE	TNAQELKLNE
	1201	KATSTGTILF	SGELHENKSY	IPQKVTFAHG	NLILGKNAEL	SVVSFTQSPG
	1251	TTITMGPGSV	LSNHSKEAGG	IAINNVIIDF	SEIVPTKDNA	TVAPPTLKL
	1301	SRTNADSKDK	IDITGTVTLL	DPNGNLYQNS	YLGEDRDITL	FNIDNSASGA
	1351	VTATNVTLQG	NLGAKKGYLG	TWNLDPNSSG	SKIILKWTFD	KYLRWPYIPR
45	1401	DNHFIYINSIW	GAQNSLVTVK	QGILGNMLNN	ARFEDPAFNN	FWASAIGSFL
	1451	RKEVSRNSDS	FTYHGRGYTA	AVDAKPRQEF	ILGAAFSQVF	GHAESYHLD
	1501	NYKHKSGSHS	TQASLYAGNI	FYFPAIRSRP	ILFQGVATYG	YMQHDTTTY
	1551	PSIEEKNMAN	WDSIAWLFDL	RFSVDLKEPQ	PHSTARLTFY	TEAEYTRIRQ
	1601	EKFTELDYDP	RSFSACSYGN	LAIPTGFSVD	GALAWREIIL	YNKVSAAAYLP
50	1651	VILRNNPKAT	YEVLSSTKEG	NVVNVLPTRN	AARAEVSSQI	YLGSYWTLYG
	1701	TYTIDASMNT	LVQMANGGIR	FVF*		

Pmp21 (CPn0963)

One example of a Pmp 21 protein is set forth as SEQ ID NO^s 83 & 84 in WO
 55 02/02606. {GenBank accession number: gi|4377287|gb|AAD19099.1| 'CPn0963';
 SEQ ID NO: 36 below}.

SEQ ID No 36

60	1	<u>MVAKKTVRSY</u>	<u>RSSF</u> <u>SHSVIV</u>	<u>AILS</u> <u>AGIAFE</u>	<u>AHSL</u> <u>HSSELD</u>	LGVFENKQFEE
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	51	HSAHVEEAQT	SVLKGS DPNV	PSQKESEKVL	YTQVPLTQGS	SGESLDLADA
	101	NFLEHFQHLF	EETTVFGIDQ	KLVWSDLDTR	NFSQPTQEPD	TSNAVSEKIS
	151	SDTKENRKDL	ETEDPSKKSG	LKEVSSDLPK	SPETAVAAIS	EDLEISENIS
5	201	ARDPLQGLAF	FYKNTSSQSI	SEKSSSFQGI	IFSGSGANS	LGFENLKAPK
	251	SGAAVYSDRD	IVFENLVKGL	SFISCESLED	GSAAGVNIVV	THCGDVTITD
	301	CATGLDLEAL	RLVKDFSRGG	AVFTARNHEV	QNNLAGGILS	VVGNGGAIIV
	351	EKNSAEKSNG	GAFACGSFVY	SNNENTALWK	ENQALSGGAI	SSASDIDIQG
	401	NCSAIEFSGN	QSLIALGEHI	GLTDFVGGGA	LAAQGTTLTR	NNAVVCVKKN
10	451	TSKTHGGAIL	AGTVDLNETI	SEVAFKQNTA	ALTGGALSAN	DKVIIANNFG
	501	EILFEQNEVR	NHGGAIYCGC	RSNPKLEQKD	SGENINIIGN	SGAITFLKNK
	551	ASVLEVMTQA	EDYAGGGALW	GHNVLDSNS	GNIQFIGNIG	GSTFWIGEYV
	601	GGGAILSTDR	VTISNNSGDV	VFKGNKGQCL	AQKYVAPQET	APVESDASST
	651	NKDEKSLNAC	SHGDHYPPKT	VEEVPPSL	EEHPVVSSTD	IRGGGAILAQ
15	701	HIFITDNTGN	LRFSGNLGGG	EESSTVGDLA	IVGGGALLST	NEVNVCSNQ
	751	VVFSDNVTSN	GCDSSGAILA	KKVDISANHS	VEFVSNGSGK	FGGAVCALNE
	801	SVNITDNGSA	VSFSKNRTRL	GGAGVAAPQG	SVTICGNQGN	IAFKENFVFG
	851	SENQRSGGGA	IIANSSVNIQ	DNAGDILFVS	NSTGSYGGAI	FVGSLVASEG
	901	SNPRTLITIG	NSGDILFAKN	STQTAASLSE	KDSFGGGAIY	TQNLKIVKNA
20	951	GNVSFYGNRA	PSGAGVQIAD	GGTVCLEAFG	GDILFEGNIN	FDGSFNAILH
	1001	CGNDSKIVEL	SAVQDKNIIF	QDAITYEENT	IRGLPKDVS	PLSAPSLIFN
	1051	SKPQDDSAQH	HEGTIRFSRG	VSKIPQIAAI	QEGTLALSQN	AELWLGLKQ
	1101	ETGSSIVLSA	GSILRIFDSQ	VDSSAPLPTE	NKEETLVSA	VQINMSSPTP
	1151	NKDKAVDTPV	LADIISITVD	LSSFVPEQDG	TLPLPPEIII	PKGTLKLSNA
25	1201	IDLKIIDFTN	VGYNHALLS	SHKDIPLISL	KTAEGMTGTP	TADASLSNIK
	1251	IDVSLPSITP	ATYGHTGVWS	ESKMEDGRLV	VGWQPTGYKL	NPEKQGALVL
	1301	NNLWSHYTDL	RALKQEIFAH	HTIAQRMELD	FSTNVWGSGL	GVVEDCQNI
	1351	EFDGFKHHLT	GYALGLDTQL	VEDFLIGGCF	SQFFGKTESQ	SYKAKNDVKS
	1401	YMGAAYAGIL	AGPWLIKAF	VYGNINNDLT	TDYGTLGIST	GSWIGKGFIA
30	1451	GTSIDYRYIV	NPRRFISAIV	STVVPFVEAE	YVRIDLPEIS	EQGKEVRTFQ
	1501	KTRFENVAIP	FGFALEHAYS	RGSRAEVNSV	QLAYVFDVYR	KGPVSLITLK
	1551	DAAYSWKSYG	VDIPCKAWKA	RLSNNTWNS	YLSTYLAFNY	EWREDLIAYD
	1601	FNGGIRIIF*				

Preferred PMP proteins for use with the invention comprise an amino acid sequence:

35 (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to one of the polypeptide sequences set forth for the pmp proteins above and/or (b) which is a fragment of at least *n* consecutive amino acids of one of the polypeptide sequences set forth above wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 40 60, 70, 80, 90, 100, 150, 200, 250 or more). These PMP proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of the polypeptide sequences set forth above. Preferred fragments of (b) comprise an epitope from one of the polypeptide sequences set forth above. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or 45 more) from the N-terminus of one of the polypeptide sequences set forth above. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

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Fifth Antigen Group

The immunogenicity of other *Chlamydia pneumoniae* antigens may be improved by combination with two or more *Chlamydia pneumoniae* antigens from either the first antigen group or the second antigen group or the third antigen group or the fourth antigen group. Such other *Chlamydia pneumoniae* antigens include a fifth antigen 55 antigen group consisting one or more cell surface exposed proteins. These antigens are referred to herein as the "fifth antigen group". Each of the *Chlamydia pneumoniae* antigens of the fifth antigen group is described in more detail below.

(37) PorB Outer Membrane Protein B (CPn0854)

One example of a PorB protein is set forth as SEQ ID NO^s: 67 & 68 in WO 02/02606. {GenBank accession number: gi|4377170|gb|AAD18992.1| 'CPn0854'; SEQ ID NO: 37 below}. Preferred PorB proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 37; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 37, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These PorB proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 37. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 37. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 37. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 37

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1  MNSKMLKHLR LATLSFSMFF GIVSSPAVYA LGAGNPAAPV LPGVNPEQTG
51  WCAFQLCNSY DLFAALAGSL KFGFYGDYVF SESAHITNVP VITSVTTSQT
101 GTTPTITSTT KNVDFDLNNS SISSSCVFAT IALQETSPAA IPLLDIAFTA
151 RVGGLKQYYR LPLNAYRDFT SNPLNAESEV TDGLIEVQSD YGIVWGLSLQ
201 KVLWKDGVSF VGVSADYRHG SSPINYIIVY NKANPEIYFD ATDGNLSYKE
251 WSASIGISTY LNDYVLPYAS VSIGNTSRKA PSDSFTELEK QFTNFKFKIR
301 KITNFDRVNF CFGTTCCISN NFYYSVEGRW GYQRAINITS GLQF*
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(38) 76kDa Protein Homolog (CPn0728)

One example of a 76kDa Protein Homolog protein is set forth as SEQ ID NO^s: 13 & 14 in WO 02/02606. {GenBank accession number: gi|4377033|gb|AAD18867.1| 'CPn0728'; SEQ ID NO: 38 below}. Preferred 76kDa proteins homologs for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 38; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 38, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These 76kDa protein homologs include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 38. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 38. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 38. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 38

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1  MVNPIGPGPI DETERTPPAD LSAQGLEASA ANKSAEAQRI AGAEAKPKES
51  KTDSVERWSI LRSVNALMS LADKLGIASS NSSSSTSRSA DVDSTTATAP
101 TPPPTTFDDY KTQAQTAYDT IFTSTSLADI QAALVSLQDA VTNIKDTAAT
151 DEETAIAAEW ETKNADAVKV GAQITELAKY ASDNQAILDS LGKLTSEFDLL
201 QAALLQSVAN NNKAAELLKE MQDNPVVPKG TPAIAQSLVD QTDATATQIE
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251 KDGNAIRDAY FAGQNASGAV ENAKSNNSIS NIDSAKAAIA TAKTQIAEAQ
 301 KKFPDSPILQ EAEQMVIQAE KDLKNIKPAD GSDVPNPGTT VGGSKQQGSS
 351 IGSIRVSMLL DDAENETASI LMSGFRQMIH MFNTENPDSQ AAQQELAAQA
 401 RAAKAAGDDS AAAALADAQK ALEAALGKAG QQQGILNALG QIASAAVVSA
 451 GVPPAAASSI GSSVKQLYKT SKSTGSDYKT QISAGYDAYK SINDAYGRAR
 501 NDATRDVINN VSTPALTRSV PRARTEARGP EKTDQALARV ISGNSRTLGD
 551 VYSQVSALQS VMQIIQSNPQ ANNEEIRQKL TSAVTKPPQF GYPYVQLSND
 601 STQKFIKLE SLFAEGSRTA AEIKALSFET NSLFIQQVLV NIGSLYSGYL
 651 Q*

(39) *OmpA conserved outer membrane protein (CPn0695)*

One example of an OmpA conserved outer membrane protein protein is set forth as SEQ ID NO^s: 59 & 60 in WO 02/02606. {GenBank accession number: gi|4376998|gb|AAD18834.1| 'CPn0695'; SEQ ID NO: 39 below}. Preferred ompA proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 39; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 39, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 39. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 39. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 39. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 39

1 MKKLLKSALL SAAFAGSVGS LQALPVGNPS DPSLLIDGTI WEGAAGDPCD
 51 PCATWCDAIS LRAGFYGDYV FDRILKVDAP KTFMSGAKPT GSAAANYTTA
 101 VDRPNPAYNK HLHDAEWFN AGFIALNIWD RFDVFCTLGA SNGYIRGNST
 151 AFNLVGLFGV KGTTVNANEL PNVSLSNGVV ELYTDTSFWS SVGARGALWE
 201 CGCATLGAEF QYAQSKPKVE ELNVICNVSQ FSVNPKPKGYK GVAFFPLPTDA
 251 GVATATGTKS ATINYHEWQV GASLSYRLNS LVPYIGVQWS RATFDADNIR
 301 IAQPKLPTAV LNLTAWNPSL LGNATALSTT DSFSDFMQIV SCQINKFKSR
 351 KACGVTVGAT LVDADKWSLT AEARLINERA AHVSGQFRF*

(40) *PepA (CPn0385)*

One example of a PepA protein protein is set forth as SEQ ID NO^s: 99 & 100 in WO 02/02606. {GenBank accession number: gi|4376664|gb|AAD18529.1 'CPn0385'; SEQ ID NO: 40 below}. Preferred PepA proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 40; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 40, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These PepA proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 40. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 40. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 40. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 40

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1  MVLFHAQASG RNRVKADAIV LPFWHFKDAK NAASFEEAEFE PSYLPALLENF
51  QGKTGEIELL YSSPKAKEKR IVLLGLGKNE ELTSDVVFQT YATLTRVLRK
101 AKCSTVNIIL PTISELRISA EEFLVGLSSG ILSLNYDYPR YNKVDRNLET
151 PLSKVTVIGI VPKMADAIFR KEAAIFEGVY LTRDLVNRNA DEITPKKLAE
201 VALNLGKEFP SIDTKVLGKD AIAKEKMGLL LAVSKGSCVD PHFIVVRYQG
251 RPKSKDHTVL IGKGVTFDSG GLDLKPGKSM LTMKEDMAGG ATVLGILSAL
301 AVLELPINVT GIIPATENAI DGASYKMGDV YVGMSGLSVE ICSTDAEGRL
351 ILADAITYAL KYCKPTRIID FATLTGAMVV SLGEEVAGFF SNNDVLAEDL
401 LEASAETSEP LWRLPLVKKY DKTLHSDIAD MKNLGSNRAG AITAALFLQR
451 FLEESSVAWA HLDIAGTAYH EKEEDRYPKY ASGFGVRSIL YYLENSLSK*

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(41) Conserved Outer Membrane Protein (Cpn0278)

One example of a conserved outer membrane protein protein is set forth as SEQ ID NO: 41 below. GenBank Accession No. GI:4376552; AAD18427.1. Preferred conserved outer membrane proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 41; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 41, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These conserved outer membrane proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 41. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 41. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 41. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 41

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1  MKKKLSLLVG LIFVLSSCHK EDAQNKIRIV ASPTPHAEEL ESLQEEAKDL GIKLKILPVD
61  DYRIPNRLLL DKQVDANYFQ HQAFLDDECE RYDCKGELVV IAKVHLEPQA IYSKKHSSLE
121 RLKSQKKLTI AIPVDRTNAQ RALHLLEECG LIVCKGPANL NMTAKDVCCK ENRSINILEV
181 SAPLLVGSLP DVDAAVIPGN FAIAANLSPK KDSLCLLEDLS VSKYTNLVVI RSEDVGSPKM
241 IKLQKLFQSP SVQHFFDTKY HGNILTMTQD NG

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Sixth Antigen Group

The immunogenicity of other *Chlamydia pneumoniae* antigens may be improved by combination with two or more *Chlamydia pneumoniae* antigens from either the first antigen group or the second antigen group or the third antigen group or the fourth antigen group or the fifth antigen group. Such other *Chlamydia pneumoniae* antigens include a sixth antigen group consisting one or more FACS positive CPn antigens. These antigens are referred to herein as the "sixth antigen group". Each of the *Chlamydia pneumoniae* antigens of the sixth antigen group is described in more detail below.

(42) Predicted Omp (CPn0020)

One example of a predicted Omp protein is set forth as SEQ ID NO^s: 91 & 92 in WO 02/02606. {GenBank accession number gi|4376272|gb|AAD18173.1: 'CPn0020';

SEQ ID NO: 42 below). Preferred Omp proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 42; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 42, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These Omp proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 42. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 42. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 42. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

15 **SEQ ID No 42**

1	MKRCFLFLAS FVLMGSSADA	LTHQEAVKKK	NSYLSHFKSV	SGIVTIEDGV
51	LNIHNNLRIQ	ANKVYVENTV	GQSLKLVAHG	NVMVNYRAKT
101	DTDSCLLTNG	RFAMYPWFLG	GSMITLTPET	IVIRKGYIST
20	151	SGDYLEYSSD	SLLSIGKTTL	RVCRIPIFL
	201	TGGFLGSYLG	MSYSPISRKH	FSSTFFLDSF
	251	NVFNMKSYYA	HRLAIDMAEA	HDRYRLHGDF
	301	TVADIFPNNF	MLKNTGPTRV	DCTWNDNYFE
	351	YLTLRQYPIS	IYNTGVYLEN	IVECGYLNFA
25	401	LHKTVPPLPIG	TLSSTLGSSL	IYSDVPEIS
	451	SYIQRRHIIE	PFVTFITETR	PLAKNEDHYI
	501	VLSKTNPRFP	RIHAKLWTH	ILSNTESKPT
	551	DAEWIWKHC	WDHMNIRWEW	IGNDNVAMTL
	601	DVSRPIDQLL	DSPLSDHRNL	ILGKLFVRPH
30	651	YLEYQMILGT	KIFEHWQLYG	VYERREADSR
			FFFLKLDKP	KKPPF*

(43) Predicted Omp (CPn0021)

One example of a predicted Omp protein is set forth as SEQ ID NO^s: 49 & 50 in WO 02/02606. {GenBank accession number gi|4376273|gb|AAD18174.1: 'CPn0021'; SEQ ID NO: 43 below}. Preferred Omp proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 43; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 43, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 43. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 43. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 43. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

50 **SEQ ID No 43**

1	MGLFHLTLFG LLLCSLPISL	VAKFPESVGH	KILYISTQST	QQALATYLEA
51	LDAYGDHDF	VLRKIGEDYL	KQSIHSSDPQ	TRKSTIIGAG
101	LSQAMETADP	LQQLLVLSAV	SGHLGKTSDD	LLFKALASPY
55	151	LANLKNTKVI	DHLHSFIHKL	PEEIQCLSAA
			IFLRLETEES	DAYIRDLLAA

5
201 KKSARSATA LQIGFYQQR FLPTLRNLLT SASPDQDEAI LYALGKLKDG
251 QSYYNIKKQL QKPDVDVTLA AAQALIALGK EEDALPVIKK QALEERPRAL
301 YALRHLPSEI GIPIALPIFL KTKNSEAKLN VALALLELG C DTPKLLLEYIT
351 ERLVQPHYNE TLALSFSKGR TLQNWKRVI IVPQDPQERE RLLSTTRGLE
401 EQILTFLFRL PKEAYLPCIY KLLASQKTQL ATTAISFLSH TSHQEALDLL
451 FQAAKLPGEP IIRAYADLAI YNLTKDPEKK RSLHDYAKKL IQETLLFVDT
501 ENQRPHPSMP YLRYQVTPES RTKLMLDILE TLATSKSSED IRLLIQLMTE
551 GDAKNFPVLA GLLIKIVE*

10 **(44) Oligopeptide Binding Protein Oppa-1 Lipoprotein (CPn0195)**

One example of an oligopeptide binding protein is set forth as SEQ ID NO^s: 23 and 24 in WO 02/02606. {GenBank accession number gi|4376466|gb|AAD18348.1: 'CPn0195'; SEQ ID NO: 44 below}. Preferred oligopeptide binding proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 44; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 44, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 44. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 44. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 44. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 44

30
1 MRKISVGICI TILLSLSVVL QGCKESSHSS TSRGELAINI RDEPRSLDPR
51 QVRLISEISL VKHIYEGLVQ ENNLSGNIEP ALAEDYSLSS DGLTYTFKLK
101 SAFWSNGDPL TAEDFIESWK QVATQEVSGI YAFALNPIKN VRKIQEGHLS
151 IDHFGVHSPN ESTLVVTLES PTSHFLKLLA LPVFFPVHKS QRTLQSKSLP
201 IASGAFYPKN IKQKQWIKLS KNPHYNNQSQ VETKTITIH F IPDANTAACL
35
251 FNQGKLNWQG PPWGERIPQE TLSNLQSKGH LHSFDVAGTS WLTFNINKFP
301 LNNMKLREAL ASALDKEALV STIFLGRAKT ADHLLPTNIH SYPEHQKQEM
351 AQRQAYAKKL FKEALEELQI TAKDLEHLNL IFPVSSSASS LLVQLIREQW
401 KESLGFAIPI VGKEFALLQA DLSSGNFSLA TGGWFADFAD PMAFLTIFAY
451 PSGVPPYAIN HKDFLEILQN IEQEODHQKR SELVSQASLY LETFHIIEPI
40
501 YHDAFQFAMN KKLSNLGVSP TGVVDFRYAK EN*

(45) CHLPS 43 kDa Protein Homologue-1 (CPn0562)

One example of a CHLPS protein is set forth as SEQ ID NO: 45 below. GenBank
45 Accession No. GI:4376854; AAD18702.1. Preferred CHLPS proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 45; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 45, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These CHLPS proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 45. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 45. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or
55 one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the

N-terminus of SEQ ID NO: 45. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

5 SEQ ID No 45

1 MSIAIAREQY AAILDMHPKP SIAMFSSEQA RTSWEKRQAH PYLYRLLEII WGVVKFLLGL
61 IFFIPLGLFW VLQKICQNF I LLGAGGWIFR PICRDSNLLR QAYAARLFSA SFQDHVSSVR
121 RVCLQYDEVF IDGLELRPN AKPDRWMLIS NGNSDCLEYR TVLQGEKDWI FRIAEESQSN
181 ILIFNYPGVM KSQGNITRNN VVKSQACVR YLRDEPAGPQ ARQIVAYGYS LGASVQAEAL
241 SKEIADGSDS VRWFVVKDRG ARSTGAVAKQ FIGSLGVWLA NLTHWNINSE KRSKDLHCPE
301 LFIYGKDSQG NLIGDGLFCK ETCFAAPFLD PKNLEECGSK KIPVAQTGLR HDHILSDDVI
361 KEVAGHIQRH FDN

15

(46) YscJ (Yop translocation J protein) (CPn0828)

One example of a YscJ protein is set forth as SEQ ID NO^s: 109 and 110 in WO 02/02606. {GenBank accession number gi|4377140|gb|AAD18965.1| 'CPn0828'; SEQ ID NO: 46 below. Preferred YscJ proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 46; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 46, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These YscJ proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 46. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 46. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 46. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 46

35 1 MVRRSISFCL FFLMTLLCCT SCNSRSLIVH GLPGREANEI VVLLVSKGVA
51 AOKLPQAAAA TAGAATEQMW DIAVPSAQIT EALAILNQAG LPRMKGTSLL
101 DLFAKQGLVP SELQEKIRYQ EGLSEQMAST IRKMDGVVDA SVQISFTTEN
151 EDNLPLTASV YIKHRGVLDN PNSIMVSKIK RLIASAVPGL VPENVSVVSD
201 RAAYSDITIN GPWGLTEEID YVSVWGIIA KSSLTKFRLI FYVLILILFV
40 251 ISCGLLWVIW KTHTLIMTMG GTKGFENPTP YTKNALEAKK AEGAAADKEK
301 KEDADSQGES KNAETSDKDS SDKDAPEGSN EIEGA*

(47) Hypothetical (CPn 0415)

One example of a hypothetical protein is set forth as SEQ ID NO^s: 101 and 102 in WO 02/02606. {GenBank accession number gi|4376696|gb|AAD18559.1| 'CPn0415'; SEQ ID NO: 47 below}. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 47; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 47, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 47. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 47. Other preferred fragments lack one or

- more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 47. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 47

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101
151
201
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301
351
401

MTLIEFVIIIV WCNAFLIKLC VIMGLQSRLO HCIEVSQNSN FDSQVKQFIY
ACQDKTLRQS VLKIFRYHPL LKIHDIAAV YLLMALEEGE DLGLSFLNVQ
QYPSGAVELF SCGGFPWKGL PYPAEHAIEFG LLLLQIAEFY EESQAYVSKM
SHFQQALFDH QGSVFPSLWS QENSRLLEKEK TTLSQSFLFQ LGMQIHPEYS
LEDPALGFWM QRTRSSSAFV AASGCQSSLG AYSSGDVGVV AYGPCSGDIS
DCYYFGCCGI AKEFVCQKSH QTTEISFLTSTGKPHPRNTG FSYLRDSYVH
LPIRCKITIS DKQYRVHAAL AEATSAMTFS IFCKGKNCQV VDGPRLRSCS
LDSYKGPGND IMILGENDAI NIVSASPYME IFALQGKEKF WNADFLINIP
YKEEGVMLIF EKKVTSEKGR FFTKMN*

(48) Hypothetical (CPn0514)

- 20 One example of a hypothetical protein is set forth as SEQ ID NO^s: 87 and 88 in WO 02/02606. {GenBank accession number gi|4376802|gb|AAD18654.1| 'CPn0514'; SEQ ID NO: 48 below}. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 48; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 48, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 48. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 48. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 48. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 48

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45
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51
101
151
201
251

MSNQLQPCIS LGCVSYINSF PLSLQLIKRN DIRCVLAPPA DLLNLLIEGK
LDVALTSSLG AISHNLGYVP GFGIAANQRI LSVNLYAAPT FFNSPQPRIA
ATLESRSSIG LLKVLCRHLW RIPTPHILRF ITTKVLRQTP ENYDGLLLIG
DAALQHPVLP GFVTYDLASG WYDLTKLPFV FALLHSTSW KEHPLPNLAM
EEALQQFESS PEEVLKEAHQ HTGLPPSLLO EYYALCQYRL GEEHYESFEK
FREYYGTLYQ QARL*

(49) Hypothetical (CPn0668)

- 50 One example of a hypothetical protein is set forth as SEQ ID NO^s: 57 and 58 in WO 02/02606. {GenBank accession number gi|4376968|gb|AAD18807.1 'CPn0668'; SEQ ID NO: 49 below}. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 49; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 49, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These

hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 49. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 49. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 49. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

10 **SEQ ID No 49**

1 **MKFLLYVPLL LVLVSTGCDA** KPVSEFEPFSG KLSTQRFEPQ HSAEEYFSQG
 51 QEFLKKGNER KALLCFGIIT HHEPRDILRN QAQYLIGVCY FTQDHPDLAD
 101 KAFASYLQLP DAEYSEELFQ MKYATAQRFA QGKRKRICRL EGFPKLMNAD
 151 EDALRIYDEI LTAFPSKDLG AQALYSKAAL LIVKNDLTEA TKTLKKLTLO
 201 FPLHLSSEA FVRLSEIYLQ QAKKEPHNLQ YLHFAKLNEE AMKKQHPNHP
 251 LNEVVSANVG AMREHYARGL YATGRFYEEK KKAEEANIYY RTAITNYPDT
 301 LLVAKCQKRL DRISKHTS*

20 **(50) Hypothetical (CPn0791)**

One example of a hypothetical protein is set forth as SEQ ID NO^s: 123 and 124 in WO 02/02606. {GenBank accession number gi|4377101|gb|AAD18929.1| 'CPn0791'; SEQ ID NO: 50 below}. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 50; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 50, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 50. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 50. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 50. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 50

40 1 MYSCYSKGIS HNYLLHPMSR LDIFVFDSL I ANQDQNLLEE IFCSEDTVLF
 51 KAYRTTALQS PLAANKLNIA RKVANYILAD NGEIDTVKLV EAIHHL SQCT
 101 YPLGPHRHNE AQDREHLLKM LKALKENPKL KESIKTLFVP SYSTIQNLIR
 151 HTLALNPQTI LSTIHVRQAA LTALFTYLRQ DVGSCFATAP AILIHQEYPE
 201 RFLKDLNDLI SSGKLSRIVN QREIAVPINL SGCIGELFKP LRILDLYPDP
 45 251 LVKLSSSPGL KKAFAANLI ETLGDSEAQI QQLLSHQYLM QKLQNVHETL
 301 TANDIIKSTL LHYYQLQEST VRAIFFKEGL FSKEQVAFST QHPRELSEIQ
 351 RVYHYLHAYE EAKSAFIHDT QNPLLKAWAY TLATLADASQ PTISNHIRLA
 401 LGWKSEDPHS LVSLVTHFVE EEVENIRILV QQCEQTYHEA RSQLEYIEGR
 451 MRNPLNNQDS QILTMDHMRF RQELNKALYE WDSAQEKAKK FLHLPEFLLS
 50 501 FYTKQIPLYF RSSYDAFIQE FAHLYANAPA GFRILFTHGR THPNTWSPIY
 551 SINEFIREFLS EFFTSTESL LGKHAVINLE KETSRLVHNI TAMLHTDVFQ
 601 EALLTRILEA YQLPVPPSIL NHLDQLSQTP WVYVSGGTVD TLLLDYFESS
 651 EPLTLTEKHP ENPHELAIFY ADALKDLPTG IKSYLEEGSH SLLSSSPTHV
 701 FSIIAGSPLF REAWDNDWYS YTWLRDVVVK QHQDFLQDTI LPQLSIYAFI
 55 751 ENFCNKYALQ HVVHDFHDFC SDHSLTLPEL YDKGSRFLSS LFTKDKTVAL
 801 IYIRRLLYLM VREVPYVSEQ QLPEVLDNVS SYLGISSRIT YEKFRSLIEE
 851 TIPKMTLLSS ADLRHIYKGL LMQSYQKIYT EEDTYLRLTT AMRHHNLAYP

901 APLLFADSNW PSIFYFGFILN PGTTEIDLWK FNYAGLQGQP LDNIQELFAT
 951 SRPWTLYANP IDYGMPPPPG YRSRLPKEFF *

(51) Hypothetical (CPn0792)

- 5 One example of a hypothetical protein is set forth as SEQ ID NO^s: 61 and 62 in WO 02/02606. {GenBank accession number gi|4377102|gb|AAD18930.1| 'CPn0792'; SEQ ID NO: 51 below}. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 51; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 51, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 51. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 51. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 51. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 51

25	1	MKHTFTKRVL	FFFFLVIPIPI	LLLNLMMVVG	FSFSAAKANL	VQVLHTRATN
	51	LSIEFEKKLT	IHKLELDRLA	NTLALKSYAS	PSAEPYAQAY	NEMMALSNNTD
	101	FSLCLIDPFD	GSVRTKNPGD	PFIRYLKQHP	EMKKKLSAAV	GKAFLLTIPG
	151	KPLLHYLILV	EDVASWDSTT	TSGLLVSFYP	MSFLQKDLFQ	SLHITKGNIC
	201	LVNKYGEVLF	CAQDSESEFV	FSLDLPNLPQ	FQARSPSAIE	IEKASGILGG
30	251	ENLITVSINK	KRYLGLVLNK	IPIQGTYTLS	LVPVSDLIQS	ALKVPLNICEF
	301	FYVLAFLLMW	WIFSKINTKL	NKPLQELTFC	MEAARGNHN	VRFEPQPYGY
	351	EFNELGNIFN	CTLLLLLNSI	EKADIDYHSG	EKLQKELGIL	SSLQSALLSP
	401	DFPTFPKVTF	SSQHLRRRQL	SGHFNGWTVQ	DGGDTLLGII	GLAGDIGLPS
	451	YLYALSARSL	FLAYASSDVS	LQKISKDTAD	SFSKTTEGNE	AVVAMTFIKY
35	501	VEKDRSLELL	SLSEGAPTMF	LQRGESFVRL	PLETHQALQP	GDRLICLTGG
	551	EDILKYFSQL	PIEELLKDPL	NPLNTENLID	SLTMMLNNET	EHSADGTLTI
	601	LSFS*				

(52) Hypothetical (CPn0820)

- 40 One example of a hypothetical protein is set forth as SEQ ID NO^s: 113 and 114 in WO 02/02606. {GenBank accession number gi|4377132|gb|AAD18958.1| 'CPn0820'; SEQ ID NO: 52 below}. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 52; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 52, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 52. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 52. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 52. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 52

5
1 MCNSIAMKKQ KRGFVLMELL MSFTLIALLL GTLGFWYRKI YTVQKQKERI
51 YNFYIEESRA YKQLRTLFSM SLSSSYEPPG SLFSLIFDRG VYRDPKLAGA
101 VRASLHHDTK DQRLELRICN IKDQSYFETQ RLLSHVTHV LSFQRNPDPE
151 KLPETIALTI TREPKAYPPR TLTYQFAVGK*

(53) Hypothetical (CPn0126)

10 One example of a hypothetical protein is set forth as SEQ ID NO: 53 below.
GenBank Accession No. GI:4376390; AAD18279.1 Preferred hypothetical proteins
for use with the invention comprise an amino acid sequence: (a) having 50% or more
identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%,
15 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 53; and/or (b) which is a
fragment of at least *n* consecutive amino acids of SEQ ID NO: 53, wherein *n* is 7 or
more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200,
250 or more). These hypothetical proteins include variants (*e.g.* allelic variants,
homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 53. Preferred fragments
of (b) comprise an epitope from SEQ ID NO: 53. Other preferred fragments lack one
20 or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-
terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or
more) from the N-terminus of SEQ ID NO: 53. Other fragments omit one or more
domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of
a transmembrane domain, or of an extracellular domain).

SEQ ID No 53

1 MVFSYYCMGL FFFSGAIISS GLLVSLGVGL GLSVLGVL LLAGLLLFKI QSMLEVPKA
61 PDLLEDAS ERLRVKASRS LASLPKEISQ LESYIRSAAN DLNTIKTWPH KDQRLVETVS
121 RKLERLAAAQ NYMISELCEI SEILEEEHH LILAQESLEW IGKSLFSTFL DMESFLNLSH
181 LSEVRPYLAV NDPRLLEITE ESWEVVSFHI NVTSAFKKAQ ILFKNNEHSR MKKKLESVQE
241 LLETFIYKSL KRSYRELGL SEKMRIHDN PLFPWVQDQQ KYAHAKNEFG EIARCLEEFEE
301 KTFFWLDEEC AISYMDWDF LNESIQNKKS RVDRDYISTK KIALKDRART YAKVLLLEENP
361 TTEGKIDLQD AQRAFERQSQ EFTLTLEHTET KVRLEALQQC FSDLREATNV RQVRFTNSEN
421 ANDLKESFEK IDKERVRYQK EQRLYWETID RNEQELREEI GESLRLQNRK KGYRAGYDAG
481 RLKGLLRQWK KNLRDVEAHL EDATMDFEHE VSKSELCSVR ARLEVLEEEEL MDMSPKVADI
541 EELLSYEERC ILPIRENLER AYLYQYKNCSE ILSKAKFFFP EDEQLLVSEA NLREVGAQLK
601 QVQGKCQERA QKFAIFEKHI QEQKSLIKEQ VRSFDLAGVG FLKSELLSIA CNLYIKAVVK
661 ESIPVDVPCM QLYYSYEDN EAVVRNRLN MTERYQNFKR SLNSIQFNGD VLLRDPVYQP
721 EGHETRLKER ELQETTLSC KLVQAQDRLS ELESRLSRR

(54) Hypothetical (CPn0794)

One example of a hypothetical protein is set forth as SEQ ID NO: 54 below.
GenBank Accession No. GI:4377105; AAD18932.1. Preferred hypothetical proteins
45 for use with the invention comprise an amino acid sequence: (a) having 50% or more
identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%,
96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 54; and/or (b) which is a
fragment of at least *n* consecutive amino acids of SEQ ID NO: 54, wherein *n* is 7 or
more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200,
50 250 or more). These hypothetical proteins include variants (*e.g.* allelic variants,
homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 54. Preferred fragments
of (b) comprise an epitope from SEQ ID NO: 54. Other preferred fragments lack one
or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-
terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or
55 more) from the N-terminus of SEQ ID NO: 54. Other fragments omit one or more

domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 54

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1  MSLYQKWWNS  QLKKSLEYST  VAALIFMIPS  QESFADSLID  LNLGLDPSVE  CLSGDGAFSV
61  GYFTKAGSTP  VEYQPFKYDV  SKKTFTILSV  ETANQSGYAY  GISYDGTITV  GTCSLGAGKY
121  NGAKWSADGT  LTPLTGITGG  TSHTEARAIS  KDTQVIEGFS  YDASGQPKAV  QWASGATTVT
181  QLADISGGSR  SSYAYAISDD  GTIIVGSMES  TITRKTAVK  WVNNVPTYLG  TLGGDASTGL
241  YISGDGTIVV  GAANTATVTN  GNQESHAYMY  KDNQMKD

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(55) Hypothetical (CPn0796)

One example of a hypothetical protein is set forth as SEQ ID NO: 55 below. GenBank Accession No. GI:4377107; AAD18934.1. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 55; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 55, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 55. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 55. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 55. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain). Cpn0796 may be secreted from *C. pneumoniae* and is localized in the membrane of Chlamydia in young inclusions whereas an N-terminal part of Cpn0796 is secreted into the host cell cytoplasm at later times. Cpn0796 was proposed to be an autotransporter and it is the first example of secretion into the host cell cytoplasm of a proposed Chlamydia autotransporter. The finding in the host cell cytoplasm of Cpn0796 suggests that an unknown transport mechanism exists for translocation over the inclusion membrane (Vandahl, "Proteome analysis of Chlamydia pneumoniae – proteins at the Chlamydia-host cell Interface," Abstract of PhD Dissertation, Dan Med Bull 2004: 51:306).

SEQ ID No 55

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1  MQPCLNMSIV  RNSALPLPCL  SRSETFKKVR  SHMKFMKVL  PWIYRKDLWV  TAFLLTAIPG
61  SFAHTLVDA  GEPRHAAQAT  GVSGDGKIVI  GMKVPDDPFA  ITVGFQYIDG  HLQPLEAVRP
121  QCSVYPNGIT  PDGTVIVGTN  YAIGMGSAV  KWNVGKVS  ELPDLTLD  ASAVSADGRV
181  IGGNRNINLG  ASVAVKWEDD  VITQLPSLPD  AMNACVNGIS  SDGSIIVGTM  VDVSWRNTAV
241  QWIGDQLSVI  GTLGGTTSVA  SAISTDGTVI  VGGSENADSQ  THAYAYKNGV  MSDIGTLGGF
301  YSLAHAVSSD  GSVIVGVSTN  SEHRYHAFQY  ADGQMVDLGT  LGGPESYAQG  VSGDGKVI  V
361  RAQVPSGDWH  AFLCPFQAPS  PAPVHGGSTV  VTSQNPRGMV  DINATYSSLK  NSQQQLQRLL
421  IQHSAKVESV  SSGAPSFTSV  KGAISKQSPA  VQNDVQKGT  FLSYRSQVHGN  VQNQQLLTGA
481  FMDWKLASAP  KCGFKVALHY  GSQDALVERA  ALPYTEQGLG  SSVLSGFGGQ  VQGRYDFNLG
541  ETVVLQPFMG  IQVLHLSREG  YSEKNVRFPV  SYDSVAYSAA  TSFMGAHVFA  SLSPKMSTAA
601  TLGVERDLNS  HIDEFKGSVS  AMGNFVLENS  TVSVLRPFAS  LAMYDVRQQ  QLVTLVVMN
661  QQPLTGTL  SLVSQSSYNLSE

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One preferred protein for use with the invention comprises an N-terminal peptide of Cpn0796 that may be secreted to be exposed on the bacterial cell surface and can also become detached via a proteolytic event. In one embodiment, the N-terminal peptide of Cpn0796 may form a beta-propeller structural conformation. One example of the

N-terminal peptide of Cpn0796 is set forth as SEQ ID NO: 86 below. The N-terminal peptide of Cpn0796 for use with the invention may comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 86; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 86, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of SEQ ID NO: 86. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 86. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 86.

SEQ ID NO: 86

HTLVDIAGEPRHAAQATGVSGDGKIVIGMKVPDDPFAITVGFQYIDGHLQPLEAVRPQCSVYPNG
ITPDGTVIVGTNYAIGMGSAVKWVNGKVSELPMLPDTLDSVASAVSADGRVIGGNRNINLGASV
AVKWEDDVITQLPSLPDAMNACVNGISSDGSIIIVGTMVDVSWRNTAVQWIGDQLSVIGTLGGTTS
VASAISTDGTIVGGSENADSQTHAYAYKNGVMSDIGTLGGFYSLAHAVSSDGSVIVGVSTNSEH
RYHAFQYADGQMVDLGLTGGPESYAQGVSGDGKIVIGRAQVPSGDWHAFLC

(56) Hypothetical (CPn0797)

One example of a hypothetical protein is set forth as SEQ ID NO: 56 below. GenBank Accession No. GI:4377108; AAD18935.1 Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 56; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 56, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralogs, mutants, *etc.*) of SEQ ID NO: 56. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 56. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 56. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 56

1 MSKKIKVLGH LTLCTLFRGV LCAAALSNIG YASTSQESPY QKSIEDWKGY TFTDLELLSK
61 EGWSEAHAVS GNGSRIVGAS GAGQGSVTAV IWESHLIKHL GTLGGEASSA EGISKDGEVV
121 VGWSDTREGY THAFVFDGRD MKDLGTLGAT YSVARGVSGD GSIIVGVSAT ARGEDYGWQV
181 GVKWEKGKIK QLKLLPQGLW SEANAISEDG TVIVGRGEIS RNHIVAVKWN KNAVYSLGTL
241 GGSVASAEAI SANGKVIVGW STTNNGETHA FMHKDETMHD LGTLGGGFSV ATGVSADGRA
301 IVGFSAVKTG EIHAFYYAEG EMEDLTTLGG EEARVFDISS EGNDIIGSIK TDAGAERAYL
361 FHIHK

(76) Oligopeptide Binding Protein Oppa-2 Lipoprotein (CPn0196)

One example of an oligopeptide binding protein is set forth as SEQ ID NO^s: 127 and 128 in WO 02/02606. {GenBank accession number GI:4376467; AAD18349.1

‘CPn0196’; SEQ ID NO: 76 below}. Preferred oligopeptide binding proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 76; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 76, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO 76. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 76. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 76. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

15

SEQ ID No 76

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1  mlrffavfis tllwltsgcs psqsskgifv vnmkemprrsl dpgktrliad qtlmrhlyeg
61 lveehsqnge ikpalaesyt isedgtrytf kiknilwsng dpltaqdfvs swkeilkeda
121 ssvlyafllp iknaraifdd tespenlgvr aldkrhleiq letpcahflh fltlpiffpv
181 hetlrnysts feempitcga frpvslekgl rhlhleknpmy hnksrvklhk iivqfisnan
241 taailfkhhk ldwqgppwge pippeisasl hqddqlfslp gasttwllfn iqkkpwnnak
301 lrkalslaid kdmlltkvvyq glaepthdil hprlypgtyp erkrqneril eaqqfleeal
361 delqmtredl eketltfstf sfsygricqm lreqwkkvkl ftipivggef ftiqknfleg
421 nysltvnqwt aafidpmsyl mifanpggis pyhlqdashfq tllikitqeh kkhlnqlili
481 ealdylehch ileplchpnl rialnknkn fnlfvrtsd frfiekli

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Seventh Antigen Group

The immunogenicity of other *Chlamydia pneumoniae* antigens may be improved by combination with two or more *Chlamydia pneumoniae* antigens from either the first antigen group or the second antigen group or the third antigen group or the fourth antigen group or the fifth antigen group or the sixth antigen group. Such other *Chlamydia pneumoniae* antigens include a seventh antigen group consisting one or more hypothetical proteins (ie proteins which, for example, have no known cellular location and/or function. These antigens are referred to herein as the “seventh antigen group”. Each of the *Chlamydia pneumoniae* antigens of the seventh antigen group is described in more detail below.

(57) Hypothetical (CPn0331)

One example of a hypothetical protein is set forth as SEQ ID NO: 57 below. GenBank Accession No. GI:4376609; AAD18480.1. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 57; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 57, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 57. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 57. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 57. Other fragments omit one or more

domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 57

5 1 MAVSGGGGVQ PSSDPGKWNP ALQGEQAE GP SPLKESIFSE TKQASSAAKQ ESLVRSGSTG
 61 MYATESQINK AKYRKAQDRS STSPKSKLKG TFSKMRA SVQ GFMSGFGSRA SRVSAKRASD
 121 SGEGETSLLPT EMDVALKKGN RISPEMQGFF LDASGMGGSS SDISQLSLEA LKSSAFSGAR
 181 SLSLSSSESS SVASFGSFQK AIEPMSEK V NAWTVARLGG EMVSSLLDPN VETSSSLVRRRA
 241 MATGNEGMID LSDLGQEEVS TAMTSPRAVE GKVKVSSSDS PEANPTGIPN SNTLERAEKE
 10 301 AEKQESREQL SEDQMMLARA MAGLLTGAAP QEVLSNSVWS GPSTVFPPPK FSGTLPTQRS
 361 GDKSKHKSPG IEKSTNHTNF SPLREGTVKS AEVKSLEPHPE SMYRFPKDSI VSREEPEAVV
 421 KESTAFKNPE NSSQNFLPIA VESVFPKESG TGGALGSDAV SSSYHFLAQR GVSLLAPLPR
 481 ATDDYKEKLE AHKGPGGPPD PLIYQYRNVA VEPPIVLRSP QPFSGSSRLS VQKPEAASV
 541 HDDGGGGNSG GFSGDQRRGS SGQKASRQEK KGKKLSTDI

(58) Hypothetical (CPn0234)

One example of a hypothetical protein is set forth as SEQ ID NO: 58 below. GenBank Accession No. gi|4376508|gb| AAD18387.1 Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more
 20 identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 58; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 21, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (*e.g.* allelic variants,
 25 homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 58. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 58. Other preferred fragments lack one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 58. Other fragments omit one or more
 30 domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 58

35 1 MLQSCCKALL SIVVSILAFH PIPGMGVEAK SGFLGKVKGW FSKKEIQEEA RILPVKDSLS
 61 WKRYDYTSSS GFSVEFPGE P DHSGQIVEVP QSEITIRYDT YVTETHPDNT VYVVSWEYEP
 121 EKVDISRPEL NLQEGFSGMM QALPESQVLF MQARQIQGHK ALEFWIVCED VYFRGMLISV
 181 NHTLYQVFMV YKNKNPQALD KEYEAFSQSF KITKIREPRT IPSSVKKKVS L

(59) Hypothetical (CPn0572)

40 One example of a hypothetical protein is set forth as SEQ ID NO: 59 below. Genbank Accession No. gi|4376866|gb| AAD18712.1. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (*e.g.* 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 59; and/or (b) which is a
 45 fragment of at least *n* consecutive amino acids of SEQ ID NO: 59, wherein *n* is 7 or more (*e.g.* 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (*e.g.* allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 59. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 59. Other preferred fragments lack one
 50 or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (*e.g.* 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 59. Other fragments omit one or more domains of the protein (*e.g.* omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 59

5 1 MAAPINQFST TTQITQTGQT TTTTTVGSLG EHSVTTTGSG AAAQTSQTVT LIADHEMQEI
 61 ASQDGSVVSF SAEHSFSTLP PETGSGVATA QSAQSAGLFS LSGRTQRRDS EISSSSDGSS
 121 ISRTSSNASS GETSRAESSP DLGDLDSLGS SERAEGAEGP EGPGLPEST IPHYDPTDKA
 181 SILNFLKNPA VQKMQTKGG HFVYVDEARS SFIFVRNGDW STAESIKVSN AKTKENITKP
 241 ADLEMCIAKF CVGYETIHS DWTGRVKPTME ERSATGNYN HLMLSMKFKT AVVYGPWNAK
 10 301 ESSSGYTPSA WRRGAKVETG PIWDDVGGLK GINWKTTPAP DFSFINETPG GGAHSTSTHG
 361 PGTPVGATVV PNVNVNLGGI KVDLGGINLG GITTNVTTEE GGTNITSTK STSTDDKVS I
 421 TSTGSQSTIE EDTIQFDDPG QGEDDNAIPG TNTPPPPGPP PNLSSSRLLT ISNASLNQVL
 481 QNVRQHLNTA YDSNGNSVSD LNQDLGQVVK NSENGVNFPT VILPKTTGDT DPSGQATGGV
 541 TEGGGHIRNI IQRNTQSTGQ SEGATPTPQP TIAKIVTSLR KANVSSSSVL PQPQVATTIT
 601 PQARTASTST TSIGTGTEST STTSTGTGTG SVSTQSTGVG TPTTTTTRSTG TSATTTTSSA
 15 661 STQTPQAPLP SGTRHVATIS LVRNAAGRSI VLQGGGRSQS FPIPPSGTGT QNMGAQLWAA
 721 ASQVASTLGQ VVNQAATAGS QPSSRRSSPT SPRRK

Eighth Antigen Group

20 The immunogenicity of other *Chlamydia pneumoniae* antigens may be improved by
 combination with two or more *Chlamydia pneumoniae* antigens from either the first
 antigen group or the second antigen group or the third antigen group or the fourth
 antigen group or the fifth or the sixth antigen group or the seventh antigen group.
 Such other *Chlamydia pneumoniae* antigens include an eighth antigen group consisting
 25 one or more FACS positive CPn antigens. These antigens are referred to herein as the
 “eight antigen group”. Each of the *Chlamydia pneumoniae* antigens of the eight
 antigen group is described in more detail below.

(60) Low Calcium Response Protein H (CPn0811)

30 One example of a Low Calcium Response Protein H is set forth as SEQ ID NO: 60
 below. Genbank Accession No. GI:4377123; AAD18949.1. Preferred low calcium
 response proteins for use with the invention comprise an amino acid sequence: (a)
 having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%,
 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 60; and/or
 (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 60,
 35 wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90,
 100, 150, 200, 250 or more). These low calcium response proteins include variants
 (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 60.
 Preferred fragments of (b) comprise an epitope from SEQ ID NO: 60. Other preferred
 fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or
 40 more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8,
 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 60. Other fragments
 omit one or more domains of the protein (e.g. omission of a signal peptide, of a
 cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 60

50 1 mskpsprnan qpqkpsasfn kktrsr lael aaqkkakadd leqvhvppte eeikkalgni
 61 feglslngldl qqilglldyl leeytvayt fysqgkynea vglfql laaa qpqnykymlg
 121 lsscyhqlhl yneaafgffl afdagpdnpi ppyyiadsll klqqpeesnn fldvtmdicg
 181 nnpefkilke rcqimkqsie kqmagetkka ptkkpagksk tttknksgkk r

(61) Yop Proteins Translocation Protein T (CPn0823)

One example of a Yop Proteins Translocation Protein T is set forth as SEQ ID NO: 61
 below. Genbank Accession No. GI:4377135; AAD18960.1. Preferred Yop proteins

for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 61; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 61, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These Yop proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 61. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 61. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 61. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

15 **SEQ ID No 61**

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1 mgislpelfs nlgsayldyi fqhppayvws vfllllarll pifavapflg aklfpspiki
61 gislswlaii fpkvladtqi tnymdnnlfy vllvkemiig ivigfvlafp fyaaqsagsf
121 itnqqgiqgl egatslisie qtsphgilyh yfvtiifwlv gghrivisll lqtlevipih
20 181 sffpaemmsl sapiwitmik mcqlclvmti qlsapaalam lmsdlflgii nrmapqvqvi
241 yllsalkafm glfltlaww fiikqidyft lawfkevpim llgsnpqvl

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(62) Yop Proteins Translocation Protein J

One example of a Yop Proteins Translocation Protein J is set forth as SEQ ID NO: 62 below Genbank Accession No. GI:4377140; AAD18965.1. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 62; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 62, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 62. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 62. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 62. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

40 **SEQ ID No 62**

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1 mvrrsisfcl fflmtllcct scnsrslivh glpgreanei vlllvskgva aqklpqaaaa
61 tagaateqmw diavpsaqit ealailnqag lprmkgtsl1 dlakqglvp selqekiryq
121 eglseqmast irkmdgvvda svqisftten ednlpltasv yikhrgvldn pnsimvskik
45 181 rliasavpgl vpenvsyvds raaysditin gpwglteeid yvsvwgiila kssltkfrli
241 fyvlililfv iscgllwviw kthtlimtmg gtkgffnptp ytknaleakk aegaaadkek
301 kedadsqges knaetsdkds sddapegsn eiega

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(63) OmpA (CPn0695)

One example of an OmPA encoded (MOMP) protein is set forth as SEQ ID NO: 63 below Genbank Accession No. GI:4376998; AAD18834.1. Preferred OmpA proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%,

- 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 63; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 63, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These OmpA proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 63. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 63. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 63. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 63

```

15      1 mkkllksall saafagsvgs lqalpvgngps dpsllidgti wegaagdpcd pcatwcdais
      61 lragfygdyv fdrilkvdap ktfsmgakpt gsaaanytta vdrnpaynk hlhdaewftn
      121 agfialniwd rfdvfctlga sngyirngst afnlvglfgv kgttvnanel pnvslsngvv
      181 elytdtsfsw svgargalwe cgcatlgaef qyaqskpkve elnvicnvsq fsvnkpkyk
      241 gvafplptda gvatatgtks atinyhewqv gaslsyrlns lvpvlgvqws ratfdadnir
20      301 iaqpkltav lnltawnpsl lgnatalstt dsfsdfmqiv scqinkfksr kacgvtvgat
      361 lvdadkwslt aearlinera ahvsgqfrf

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(64) Hypothetical (CPn0210)

- One example of a Hypothetical Protein is set forth as SEQ ID NO: 64 below Genbank Accession No. GI:4376482; AAD18363.1. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 64; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 64, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, *etc.*) of SEQ ID NO: 64. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 64. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 64. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 64

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      1 mlvelealkr efahlkdqkp tsdqeitsly qcldhlefvl lglgqdkflk atededvlfe
      61 sqkaidawna lltkardvlg lgdigaiyqt ieiflgaylsk vnrrafcias eihflktair
      121 dlnayylldf rwpckieef vdwgndcvei akrklctfek etkelnesll reehamekcs
45      181 iqdlqrklsd iielhdvsl fcfsktpsge eyqkdelyqs rlryllllye ytllicktstd
      241 fgeqarakee firekfsll lekgikqtke lefaiakskl ergclvmrky eaaakhslds
      301 mfeeetvksp rkdte

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(65) Low Calcium Response Locus Protein H (CPn1021)

- One example of a Low Calcium Response Protein H is set forth as SEQ ID NO: 65 below Genbank Accession No. GI:4377352; AAD19158.1. Preferred low calcium response proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%,

93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 65; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 65, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These low calcium response proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 65. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 65. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 65. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 65

1 mshlnyllek iaasskedfp fpddlesyle gyvpdknial dtyqkifkis sedlekvyke
61 gyhayldkdy aksitvfrwl vffnfpvskf wfslgaslmh seqysqalha ygvtaavlrdk
121 dpyphyayyi cytlitnehee aekalemawv raqhkplyne lkeeildirk hk

Ninth Antigen Group

The immunogenicity of other *Chlamydia pneumoniae* antigens may be improved by combination with two or more *Chlamydia pneumoniae* antigens from either the first antigen group or the second antigen group or the third antigen group or the fourth antigen group or the fifth antigen group or the sixth antigen group or the seventh antigen group or the eighth antigen group. Such other *Chlamydia pneumoniae* antigens include a ninth antigen group. These antigens are referred to herein as the "ninth antigen group". Each of the *Chlamydia pneumoniae* antigens of the ninth antigen group is described in more detail below.

(66) Low Calcium Response Protein D (CPn0323)

One example of a Low Calcium Response Protein D is set forth as SEQ ID NO: 66 below Genbank Accession No. GI:4376601; AAD18472.1. Preferred low calcium response proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 66; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 66, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These low calcium response proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 66. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 66. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 66. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 66

1 mnkllnfvsr tlggdtalnm inkssdlila lwmmgvvlmi iiplpppivd lmitinlsis
61 vflmlvalyi psalqlsvfp slllittmfr lginissrqr illkayaghv iqafgdfvvg
121 gnyvvgfiif liitiiqfiv vtkgaervae vaarfrldam pgkqmaidad lragmidatq
181 ardakraiqk eselygamdg amkfikgdiv agivislili vggltigvam hgmdlaqaah
241 vytllsigdg lvsqipslli altagivttr vssdkntnlg keistqlvke pralllagaa

5
301 tlgvgffkgf plwsfsilal ifvalgilll tkksaagkkg ggsgasttvv aagdgaatvg
361 dnpddysl tl pvilelgkdl skliqhktks ggsfvddmip kmrqalyqdi girypgihr
421 tdspslegyd ymillnevy vrgkipphhv ltnevednls rynlpfityk naaglpaw
481 sedakailek aaikywtple viilhllyff hkssqeflgi qevrsmiefm ersfpdlvke
541 vtrliplqkl teifkrlvqe qisikdlrti leslsewaqt ekdtvlltey vrsslklyis
601 fkfsqgqsai svylldpeie emirgaikqt sagsylaldp dsvnlilksm rntitptpag
661 gqppvlltai dvrryvrkli etefpdiavi syqeilpeir iqplgriqif

(67) CHLPS 43kDa Protein Homolog-1 (CPn0062)

10 One example of a CHLPS 43kDa Protein Homolog-1 is set forth as SEQ ID NO: 67
below Genbank Accession No. GI:4376318; AAD18215.1. Preferred CHLPS
proteins for use with the invention comprise an amino acid sequence: (a) having 50%
or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%,
15 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 67; and/or (b) which is a
fragment of at least n consecutive amino acids of SEQ ID NO: 67, wherein n is 7 or
more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200,
250 or more). These CHLPS proteins include variants (e.g. allelic variants, homologs,
orthologs, paralogs, mutants, etc.) of SEQ ID NO: 67. Preferred fragments of (b)
comprise an epitope from SEQ ID NO: 67. Other preferred fragments lack one or
20 more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-
terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or
more) from the N-terminus of SEQ ID NO: 67. Other fragments omit one or more
domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of
a transmembrane domain, or of an extracellular domain).

25

SEQ ID No 67

30
1 mmsskrtski avlsilltft hsigfanans svglgtvyit sevvkkpqkg serkqakkep
61 rarkgylvps srlsaraqk mknssrkess ggceisians tprsvklrrn kraeqkaakq
121 gfsafsnltl ksllpklpks qktsiherek atsrfrvnesq lssarkryct pssaapslfl
181 eteivrapve rtkelq dnei hipvvqvqn pkeqntkttk qlasqasiqq segteqslre
241 laqgaslpvl vrsnpevsq rkeellkel vaerrqckrk svrqalears ltkkvarggs
301 vtstlrydpe kaaeiksrrn ckvspeareq kyssckrdar angkqdkttp sedasqeeqq
361 tgaglvkrtp ksqvasnaqn fyrnsknnti dsyltanqys csseetdwpc sscvskrrth
35 421 nsisvctmvv tviamivgal iianatesqt tsdptpptpt p

(68) Hypothetical (CPn0169)

40 One example of a CHLPS 43kDa Protein Homolog-1 is set forth as SEQ ID NO: 68
below Genbank Accession No. GI:4376437; AAD18322.1. Preferred CHLPS
proteins for use with the invention comprise an amino acid sequence: (a) having 50%
or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%,
95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 68; and/or (b) which is a
fragment of at least n consecutive amino acids of SEQ ID NO: 68, wherein n is 7 or
more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200,
45 250 or more). These CHLPS proteins include variants (e.g. allelic variants, homologs,
orthologs, paralogs, mutants, etc.) of SEQ ID NO: 68. Preferred fragments of (b)
comprise an epitope from SEQ ID NO: 68. Other preferred fragments lack one or
more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-
terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or
50 more) from the N-terminus of SEQ ID NO: 68. Other fragments omit one or more
domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of
a transmembrane domain, or of an extracellular domain).

SEQ ID No 68

1 mknvgsecsq plvmelntqp lrrnlcesrlv kitsfviall alvvggitlta lagagilsfl
 5 61 pwlvlgivlv vlcalflflfs ykfcpikeig vvyntdsqih qwfqkqrnk d lekat enpel
 121 fgenraednn rsarsqvket lrdcdgnvlk kiyernldvl lfmnwvpktm ddvdpseds
 181 irtvişcykl ikackpefrs lisellramq sglgllsr cs ryqeraktvs hkda plfcpt
 241 hsy yrdgylt plragpryii nr ai

(69) PmpD family (CPn0963)

10 One example of a PmpD protein is set forth as SEQ ID NO: 69 below Genbank
 Accession No. GI:4377287; AAD19099.1. Preferred PmpD proteins for use with the
 invention comprise an amino acid sequence: (a) having 50% or more identity (e.g.
 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%,
 98%, 99%, 99.5% or more) to SEQ ID NO: 69; and/or (b) which is a fragment of at
 15 least n consecutive amino acids of SEQ ID NO: 69, wherein n is 7 or more (e.g. 8, 10,
 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more).
 These PmpD proteins include variants (e.g. allelic variants, homologs, orthologs,
 paralog s, mutants, etc.) of SEQ ID NO: 69. Preferred fragments of (b) comprise an
 epitope from SEQ ID NO: 69. Other preferred fragments lack one or more amino
 20 acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or
 one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the
 N-terminus of SEQ ID NO: 69. Other fragments omit one or more domains of the
 protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a
 transmembrane domain, or of an extracellular domain).

SEQ ID No 69

1 mvakktvrsy rssfshsviv ailsagiafe ahs l hsseld lgvfnkqfee hsahveeaqt
 30 61 svlkgdpvn psqkesekvl ytqvpl tqgs sgesldlada n flehfqhl f eettvfgidq
 121 klw sldl dtr n fsqptqep d tsn avsekis sdtkenrkdl etedpskks g lkevssdlpk
 181 spetavaais edleisenis ardplqglaf fykntssqsi sekdssfqgi ifsgsgang s
 241 lgfenlkapk sgaavysdrd ivfenlvkgl sfiscesled gsaagvnivv thcgdvtltd
 301 catgldleal rlvkdf srgg avftarnhev qnnlaggils vvg nkgai v v eknsaeksng
 361 gafacgsfvy snnentalwk enqalsggai ssasdidig g ncsaiefsgn qslialgehi
 35 421 gltdfv ggg a laa qgtltlr nnavvqc vkn tskthggail agtvdlneti sevafkqnta
 481 altggalsan dkvi iannfg eilfegnev r nhggaiycgc rsnpkleqkd sgeniniign
 541 sgaitflknk asvlevmtqa edyagggalw ghnvll dsns gniqfignig gstfwigeyv
 601 gggailstdr vtisnns gdv vfkgnkgqcl aqkyvapqet apvesdasst nkdekslnac
 661 shgdhyppkt veeevpsll eehpvvsstd irgggailaq hifitdntgn lrfsgnlggg
 40 721 eesstv gdl a ivgggallst nevnvcsnqn vvf sdnvtsn gcdsggaila kkvdisanhs
 781 vefvsngsgk fggavcalne svnitdngsa vsfsknrtrl ggagvaapqg svticgnqgn
 841 iafenfvfg senqrs gga i ianssvniq dnagdilfvs nstgsyggai fvgslvaseg
 901 snprtltitg nsgdilfakn stqtaaslse kdsfgggaiy tqnlkivkna gnv sfygnra
 961 psgagvqiad ggtvcleafg gdilfegn in fdgsfnaihl cgndskivel savqd kniif
 45 1021 qdaityeent irglpdkdvs plsapslifn skpqddsagh hegtirfsrg vskipqiaai
 1081 qegtlalsqn aelwlaglkq etgssivlsa gsilrifdsq vdssaplpte nkeetlv sag
 1141 vqinmssptp nkdkavdtpv ladiisitvd lssfvpeqdg tlp lppeiii pkgtklhsna
 1201 idlkiidptn vgyenhalls shkdiplisl ktaegmtgtp tadaslsnik idvslpsitp
 1261 atyghtgvws eskmedgrlv vgwqptgykl npekqgalvl nnlwshytdl ralkq eifah
 50 1321 htiaqrmeld fstnvwgsgl gvvedcqni g efdgfkhhlt gyalgltdql vedfliggcf
 1381 sqffgktesq sykakndvks ymgaayagil agpwlikgaf vygninndlt tdygtlgist
 1441 gswigkgfia gtsidyryiv nprrfisaiv stvvpfveae yvridlpeis eggkevrtfq
 1501 ktrfenvaip fgfalehays rgsraevns v qlayvfdvyr kgpvslitlk daayswksy g
 1561 vdipckawka rlsnntewns ylstylafny ewredliayd fnggiriif

Tenth Antigen Group

The immunogenicity of other *Chlamydia pneumoniae* antigens may be improved by
 combination with two or more *Chlamydia pneumoniae* antigens from either the first

antigen group or the second antigen group or the third antigen group or the fourth antigen group or the fifth antigen group or the sixth antigen group or the seventh antigen group or the eighth antigen group or the ninth antigen group. Such other *Chlamydia pneumoniae* antigens include a tenth antigen group. Each of the

5 *Chlamydia pneumoniae* antigens of the tenth antigen group is described in more detail below.

(70) OmpH-like outer membrane protein (CPn0301)

One example of 'OmpH-like' protein is disclosed as SEQ ID NO^s: 77 & 78 in WO

10 02/02606. {GenBank accession number: gi|4376577|gb|AAD18450.1| 'CPn0301'; SEQ ID NO: 70 below and SEQ ID No 4 above}. Preferred OmpH-like proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 4; and/or (b) which is a

15 fragment of at least *n* consecutive amino acids of SEQ ID NO: 3, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These OmpH-like proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 4. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 4. Other preferred fragments lack one or

20 more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably 19 or more, to remove the signal peptide) from the N-terminus of SEQ ID NO: 4. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above, of a cytoplasmic domain, of a

25 transmembrane domain, or of an extracellular domain).

SEQ ID No 70

30

1	<u>MKKLLFSTFL</u>	<u>LVLGSTSAAH</u>	<u>ANLGYVNLKR</u>	CLEESDLGKK	ETEELEAMKQ
51	QFVKNAEKIE	EELTSIYNKL	QDEDYMESLS	DSASEELRKK	FEDLSGEYNA
101	YQSQYYQSIN	QSNVKRIQKL	IQEVKIAAES	VRSKEKLEAI	LNEEAVLAIA
151	PGTDKTTEII	AILNESFKKQ	N*		

(71) L7/L12 Ribosomal Protein (CPn0080)

35 One example of an L7/L12 Ribosomal protein is set forth as SEQ ID No 71 below {GenBank accession number: GI:4376338; AAD18233.1}. 'CPn0080'; SEQ ID NO: 71 below. Preferred L7/L12 proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more)

40 to SEQ ID NO: 71; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 71, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These L7/L12 ribosomal proteins include variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, etc.) of SEQ ID NO: 71. Preferred fragments of (b) comprise an epitope from SEQ ID

45 NO: 71. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more; preferably 19 or more, to remove the signal peptide) from the N-terminus of SEQ ID NO: 71. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide as described above,

50 of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 71

1 mttesletlv eklsnltvle lsqkkllee kwdvtasapv vavaagggge apvaaeptef
 5 61 avtledvpad kkigvlkvvr evtglalkea kemteglpkt vkektksda edtvkklqda
 121gakasfkgl

(72) AtoS two-component regulatory system sensor histidine kinase protein (CPn0584)

One example of 'AtoS' protein is disclosed as SEQ ID NO^s: 105 & 106 in WO
 10 02/02606. {GenBank accession number: gi|4376878|gb|AAD18723.1| 'CPn0584';
 SEQ ID NO: 72 below and SEQ ID No 9 above}. Preferred AtoS proteins for use
 with the invention comprise an amino acid sequence: (a) having 50% or more identity
 (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%,
 15 98%, 99%, 99.5% or more) to SEQ ID NO: 72; and/or (b) which is a fragment of at
 least *n* consecutive amino acids of SEQ ID NO: 72, wherein *n* is 7 or more (e.g. 8, 10,
 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more).
 These AtoS proteins include variants (e.g. allelic variants, homologs, orthologs,
 paralog, mutants, etc.) of SEQ ID NO: 72. Preferred fragments of (b) comprise an
 epitope from SEQ ID NO: 72. Other preferred fragments lack one or more amino
 20 acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or
 one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the
 N-terminus of SEQ ID NO: 72. Other fragments omit one or more domains of the
 protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a
 transmembrane domain, or of an extracellular domain).

25

SEQ ID No 72

1 MNVPDSKNLH PPAYELLEIK ARITQSYKEA SAILTAIPDG ILLSETGHF
 51 LICNSQAREI LGIDENLEIL NRSFTDVLDP TCLGFSIQEA LESLKVPKTL
 101 RLSLCKESKE KEVELFIRKN EISGYLFIQI RDRSDYKQLE NAIERYKNIA
 151 ELGKMTATLA HEIRNPLSGI VGFASILKKE ISSPRHQRL SSIISGTRSL
 201 NNLVSSMLEY TKSQPLNLKI INLQDFESSL IPLLSVSFPN CKFVREGAQP
 251 LFRSIDPDRM NSVWNLVKN AVETGNPIT LTLHTSGDIS VTNPGTIPSE
 301 IMDKLFTPF TTKREGNGLG LAEAQKIIRL HGGDIQLKTS DSAVSFFIII
 35 351 PELLAALPKE RAAS*

(73) OmcA 9kDa-cysteine-rich lipoprotein (CPn0558)

One example of 'OmcA' protein is disclosed as SEQ ID NO^s: 9 & 10 in WO
 02/02606. {GenBank accession number: gi|4376850|gb|AAD18698.1| 'CPn0558',
 40 'OmcA', 'Omp3'; SEQ ID NO: 73 below and SEQ ID No 10 above}. Preferred
 OmcA proteins for use with the invention comprise an amino acid sequence: (a)
 having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%,
 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 73; and/or
 (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 73,
 45 wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90,
 100, 150, 200, 250 or more). These OmcA proteins include variants (e.g. allelic
 variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 73. Preferred
 fragments of (b) comprise an epitope from SEQ ID NO: 73. Other preferred fragments
 lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from
 50 the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20,
 25 or more; preferably 18 or more to remove the signal peptide) from the N-terminus
 of SEQ ID NO: 73. Other fragments omit one or more domains of the protein (e.g.
 omission of a signal peptide as described above, of a cytoplasmic domain, of a

transmembrane domain, or of an extracellular domain). The protein may be lipidated (e.g. by a *N*-acyl diglyceride), and may thus have a N-terminal cysteine.

SEQ ID No 73

5

1 MKKAVLIAAM FCGVVSLSSC CRIVDCCFED PCAPSSCNPC EVIRKKERSC
51 GGNACGSYVP SCSNPGSTE CNSQSPQVKG CTSPDGRCKQ *

(74) Hypothetical (CPn0331)

10 One example of a hypothetical protein is set forth as SEQ ID NO: 74 below and SEQ ID No 57 above. Genbank Accession No. GI:4376609; AAD18480.1. Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 74; and/or
15 (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 74, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 74. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 74. Other preferred fragments
20 lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 74. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

25

SEQ ID NO 74

1 mavsggggvq pssdpgkwnp alqgeqaegp splkesifse tkqassaakq eslvrsgstg
61 myatesqink akyrkaqdrs stspksklkg tfskmravsq gfmsgfgsra srvsakrasd
121 sgegtsllpt emdvalkkgn rispemqgff ldasgmggss sdisqlslea lkssafsgar
181 slslsssess svasfgsfqk aiepmseekv nawnvarlwg emvsslldpn vetsslvrra
241 matgnegmid lsdlgqeevs tamtsprave gkvkvssdsd peanptgipn sntleraeke
301 aekgesreql sedqmmlara maglltgaap qevlsnsvws gpstvfpppk fsgtlptqrs
361 gdkskhkspg iekstnhtnf splregtvks aevkslphpe smyrfpkdsi vsreepeavv
421 kestafknpe nssqnflpia vesvfpkesg tggalgdsav sssyhflaqr gvsllaplpr
35 481 atddykekle ahkgpggppd pliyqyrnva veppivlrsp qpfsgssrls vqgkpeaasv
541 hddggggngsg gfsqdqrrgs sgqkasrkek kgkklstdi

(75) PmpD family (CPn0963)

40 One example of a PmpD protein is set forth as SEQ ID NO: 75 below Genbank Accession No. GI:4377287; AAD19099.1. Preferred PmpD proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 75; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 75, wherein *n* is 7 or more (e.g. 8, 10,
45 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 75. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 75. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or
50 more) from the N-terminus of SEQ ID NO: 75. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 75

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5      1 mvakktvrsy rssfshsviv ailsagiafe ahslhsseld lgvfknkqfee hsahveeaqt
      61 svlkgdvpvn psqkesekvl ytvvpltggs sgesldlada nflehfgqlf eettvfgidq
121    klvwsdldtr nfsqptqepd tsnavseki sdtkenrkd etedpskks lkevssdlpk
181    spetavaais edleisenis ardplqglaf fykntssqsi sekdssfqqi ifsgsgansg
241    lgfenlkapk sgaavysdrd ivfenlvkgl sfiscesled gsaagvnivv thcgdvtltd
10     301 catgldleal rlvkdfsrgg avftarnhev qnnlaggils vvgknkgaivv eknsaeksng
      361 gafacgsfvy snnentalwk enqalsggai ssasdidigg ncsaiefsgn qslialgehi
      421 gltdfvvgga laaaggtltlr nnavvqcvcn tskthggail agtvdlneti sevafkqnta
      481 altggalsan dkviianfng eilfeqnevr nhggaiycgc rsnpkleqkd sgeniniign
      541 sgaitflknk asvlevmtqa edyagggalw ghnvllsdsns gniqfignig gstfwigeyv
15     601 gggailstdr vtisnsgdv vfkgnkgqcl aqkyvapqet apvesdasst nkdekslnac
      661 shgdhyppkt veeevppsll eehpvvsstd irgggailaq hifitdntgn lrfsgnlggg
      721 eesstvgdla ivgggallst nevnvcnqn vvfndvtsn gcdsggaila kkvdisanhs
      781 vefvsngsgk fggavcalne svnitdngsa vsfsknrtrl ggagvaapqg svticgnqgn
      841 iafkenfvfg senqrsggga iianssvniq dnagdilfvs nstgsyggai fvgslvaseg
20     901 snprtltitg nsdildfakn stqtaaslse kdsfgggaiy tqnlkivkna gnvsfygnra
      961 psgagvqiad ggtvcleafg gdilfegnin fdgsfnaihl cgndskivel savqdkniif
1021   qdaityeent irglpdkdvs plsapslifn skpqddsagh hegtirfsrg vskipqiaai
1081   qegtlalsqn aelwlaglkq etgssivlsa gsilrifdsq vdssaplpte nkeetlv sag
1141   vqinmssptp nkdkavdtpv ladiisitvd lssfvpeqdg tlplppeiii pkgtklhsna
1201   idlkiidptn vgyenhalls shkdiplisl ktaegmtgtp tadaslsnik idvslpsitp
25     1261 atyghtgws eskmedgrlv vgwqptgykl npekqgalvl nnlwshytdl ralkgeifah
      1321 htiaqrmeld fstnvwsgsl gvvedcqng efdgfkhhlt gyalgldtql vedfliggcf
      1381 sqffgktesq sykakndvks ymgaayagil agpwlikgaf vygninnldt tdygtlgist
      1441 gswigkgfia gtsidryyiv nprrfisaiv stvvpfveae yvridlpeis eggkevrtfq
30     1501 ktrfenvaip fgfalehays rgsraevnsv qlayvfdvyr kgpvslitlk daayswksyg
      1561 vdipckawka rlsnntewns ylstylafny ewredliayd fnggiriif

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(76) Hypothetical (CPn0798)

One example of a hypothetical protein is set forth as SEQ ID NO: 78 below.

GenBank Accession No. GI:4377109; AAD18936 Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 78; and/or (b) which is a fragment of at least *n* consecutive amino acids of SEQ ID NO: 78, wherein *n* is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 78. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 78. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 78. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 78

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50     1 mkktccqnyr sigvvfsvvl fvlttqtlfa ghfidigtsg lyswargvsg dgrvvvgyeg
      61 gnafkyvdge kflleglvpr sealvfkasy dgsviigisd qdpscravkw vngalvdldi
121    fsegmqsfae gvssdgktiv gclysddtet nfavkwdetg mvvlpnlped rhscawdase
181    dgsvivgdam gseeiakavy wkdgqhlls nipgakrssa havskdgsfi vgefiseene
55     241 vhafvyhngv ikdigtlggd ysvatgvsrd gkvivghstr tdgeyrafky vdgrmidlgt
      301 lggsasfafg vsddgktivg kfetelgech afiylld

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(77) Hypothetical (CPn0799)

One example of a hypothetical protein is set forth as SEQ ID NO: 79 below. GenBank Accession No. GI: 15618708; AAD18937 Preferred hypothetical proteins for use with the invention comprise an amino acid sequence: (a) having 50% or more identity (e.g. 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, 99.5% or more) to SEQ ID NO: 79; and/or (b) which is a fragment of at least n consecutive amino acids of SEQ ID NO: 79, wherein n is 7 or more (e.g. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). These hypothetical proteins include variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.) of SEQ ID NO: 79. Preferred fragments of (b) comprise an epitope from SEQ ID NO: 79. Other preferred fragments lack one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the C-terminus and/or one or more amino acids (e.g. 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25 or more) from the N-terminus of SEQ ID NO: 79. Other fragments omit one or more domains of the protein (e.g. omission of a signal peptide, of a cytoplasmic domain, of a transmembrane domain, or of an extracellular domain).

SEQ ID No 79

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1 maaikqilrs mlsqssslwmv lfslyslsgy cyvitdkped dfhsssavkw dhwgkttlsr
20 61 lsnkkasaka vsgtgattvg fikdtwsrty avrwnywgk elptsswvkk skatgissdg
121 siiagivene lsqsfavtwk nnemyllpst wavqskaygi ssdgsvivgs akdawsrtfa
181 vkwtgheaqv lpvgwavksv ansvsangsi ivgsvqdasg ilyavkwegn tithlgtlgg
241 ysaiakavsn ngkvivgrse tyygevhafc hkngvmsdlg tlggsysaak gvsatgkviv
25 301 gmsttangkl hafkyvggrm idlgeyswke acanavsidg eiivgvqse

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Preferably the composition of the invention comprises a combination of CPn antigens selected from the group consisting of: (1) CPn0301 and CPn0080; (2) CPn 0584 and CPn 0558; and (3) CPn 0331 and CPN 0963. Preferably the composition comprises a combination of any one or more of groups (1), (2) and (3).

Even more preferably, the composition of the present invention comprises a combination of CPn antigens selected from the group consisting of (1) CPn0385, CPn0324, CPn 0503, CPn0525 and CPn 0482. Preferably the composition is administered in the presence of alum and/or cPG.

The invention thus includes a composition comprising a combination of *Chlamydia pneumoniae* antigens, said combination selected from the group consisting of two, three, four, five or six *Chlamydia pneumoniae* antigens of the first antigen group and two, three, four, five, or six *Chlamydia pneumoniae* antigens of the second antigen group. Preferably, the combination is selected from the group consisting of three, four, five or six *Chlamydia pneumoniae* antigens from the first antigen group and three, four, five or six *Chlamydia pneumoniae* antigens from the second antigen group. Still more preferably, the combination consists of six *Chlamydia pneumoniae* antigens from the first antigen group and three, four, five or six, *Chlamydia pneumoniae* antigens from the second antigen group.

The invention further includes a composition comprising a combination of *Chlamydia pneumoniae* antigens, said combination selected from the group consisting of two, three, four, five or six, *Chlamydia pneumoniae* antigens of the second antigen group and two, three, four, five, six, seven or eight *Chlamydia pneumoniae* antigens of the third antigen group. Preferably, the combination is selected from the group consisting of three, four, five or six *Chlamydia pneumoniae* antigens from the second antigen

group and three, four, five, six, seven or eight *Chlamydia pneumoniae* from the third antigen group. Still more preferably, the combination consists of six *Chlamydia pneumoniae* antigens from the second antigen group and three, four, five, six, seven or eight *Chlamydia pneumoniae* antigens of the third antigen group.

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There is an upper limit to the number of *Chlamydia pneumoniae* antigens which will be in the compositions of the invention. Preferably, the number of *Chlamydia pneumoniae* antigens in a composition of the invention is less than 20, less than 19, less than 18, less than 17, less than 16, less than 15, less than 14, less than 13, less than 12, less than 11, less than 10, less than 9, less than 8, less than 7, less than 6, less than 5, less than 4, or less than 3. Still more preferably, the number of *Chlamydia pneumoniae* antigens in a composition of the invention is less than 6, less than 5, or less than 4. The *Chlamydia pneumoniae* antigens used in the invention are preferably isolated, i.e., separate and discrete, from the whole organism with which the molecule is found in nature or, when the polynucleotide or polypeptide is not found in nature, is sufficiently free of other biological macromolecules so that the polynucleotide or polypeptide can be used for its intended purpose.

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In either of the above combinations, preferably the composition comprises one or more *Chlamydia pneumoniae* antigens from the fourth antigen group which includes porB. Or, alternatively, in either of the above combinations, preferably the *Chlamydia pneumoniae* antigens from the fourth antigen group includes one or more members of the pmp3 family.

20

Other aspects of the present invention are presented in the accompanying claims and in the following description and drawings. These aspects are presented under separate section headings. However, it is to be understood that the teachings under each section are not necessarily limited to that particular section heading.

25

Before describing the present invention in detail, it is to be understood that this invention is not limited to particularly exemplified molecules or process parameters as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments of the invention only, and is not intended to be limiting. In addition, the practice of the present invention will employ, unless otherwise indicated, conventional methods of virology, microbiology, molecular biology, recombinant DNA techniques and immunology all of which are within the ordinary skill of the art. Such techniques are explained fully in the literature. See, e.g., Sambrook, *et al.*, *Molecular Cloning: A Laboratory Manual* (2nd Edition, 1989); *DNA Cloning: A Practical Approach*, vol. I & II (D. Glover, ed.); *Oligonucleotide Synthesis* (N. Gait, ed., 1984); *A Practical Guide to Molecular Cloning* (1984); and *Fundamental Virology*, 2nd Edition, vol. I & II (B.N. Fields and D.M. Knipe, eds.).

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All publications, patents and patent applications cited herein, whether *supra* or *infra*, are hereby incorporated by reference in their entirety. It must be noted that, as used in this specification and the appended claims, the singular forms "a", "an" and "the" include plural referents unless the content clearly dictates otherwise. All scientific and technical terms used in this application have meanings commonly used in the art unless otherwise specified. As used in this application, the following words or phrases have the meanings specified.

45

The term "comprising" means "including" as well as "consisting" *e.g.* a composition "comprising" X may consist exclusively of X or may include something additional *e.g.* X + Y.

5

The term "about" in relation to a numerical value x means, for example, $x \pm 10\%$.

References to a percentage sequence identity between two amino acid sequences means that, when aligned, that percentage of amino acids are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in section 7.7.18 of *Current Protocols in Molecular Biology* (F.M. Ausubel *et al.*, eds., 1987) Supplement 30. A preferred alignment is determined by the Smith-Waterman homology search algorithm using an affine gap search with a gap open penalty of 12 and a gap extension penalty of 2, BLOSUM matrix of 62. The Smith-Waterman homology search algorithm is disclosed in Smith & Waterman (1981) *Adv. Appl. Math.* 2: 482-489.

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IMMUNE RESPONSE

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The mechanism by which the immune system controls disease includes the induction of neutralising antibodies via humoral immunity and the generation of T-cell responses via cellular immunity. As used herein, the term "immune response" against an antigen refers to the development in a host mammalian subject of a humoral and/or a cellular immune response against that antigen.

25

As used herein, the term "humoral immune response" refers to an immune response mediated by antibody molecules. The antibodies generated by humoral immunity are primarily effective against extracellular infectious agents.

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SEQ ID Nos 1-86 in the compositions of the invention may be supplemented or substituted with an antibody that binds to the protein. This antibody may be monoclonal or polyclonal.

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As used herein, the term "cell mediated immune (CMI) response" is one mediated by T-lymphocytes and/or other white blood cells. The CMI immune mechanisms are generally more effective against intracellular infections and disease because the CMI mechanisms prime T cells in a way that, when an antigen appears at a later date, memory T cells are activated to result in a CMI response that destroys target cells that have the corresponding antigen or a portion thereof on their cell surfaces, and thereby the infecting pathogen. The CMI response is focused on the destruction of the source of infection mediated by either effector cells that destroy infected cells of the host by direct cell-to-cell contact and/or by the release of molecules, such as cytokines, that possess anti-viral activity. Thus the CMI response, which is characterised by a specific T lymphocyte cellular response, is crucial to produce resistance to diseases caused by cancer, viruses, pathogenic and other intracellular microorganisms.

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In one aspect of the present invention, an immunogenic composition is provided comprising a combination of at least one antigen that elicits a *Chlamydia pneumoniae* specific Th1 immune response (such as a cell mediated or cellular immune response) and at least one antigen that elicits a *Chlamydia pneumoniae* specific Th2 response (such as a humoral or antibody response). The immunogenic composition may further comprise a Th1 adjuvant and a Th2 adjuvant.

In one embodiment, the invention provides a composition comprising a combination of *Chlamydia pneumoniae* antigens that elicit at least a *Chlamydia pneumoniae* specific Th1 immune response. As an example, the combination of *Chlamydia pneumoniae* antigens may include at least one antigen associated with reticulate bodies (RBs) of *Chlamydia pneumoniae*, including but not limited to antigens expressed, exposed on or translocated into, through or across on the inclusion membrane, antigens expressed, secreted, released or translocated into the cytosol of host cells, or antigens processed or degraded in the cytosol of host cells and/or expressed, exposed or presented on the surface of the host cell. The compositions of the invention will preferably elicit both a cell mediated immune response as well as a humoral immune response in order to effectively address a *Chlamydia* intracellular infection. This immune response will preferably induce long lasting (eg neutralising) antibodies and a cell mediated immunity that can quickly respond upon exposure to *Chlamydia*.

The invention also comprises an immunogenic composition comprising one or more immunoregulatory agents. Preferably, one or more of the immunoregulatory agents include an adjuvant. The adjuvant may be selected from one or more of the group consisting of a Th1 adjuvant and Th2 adjuvant, further discussed below. The adjuvant may be selected from the group consisting of a mineral salt, such as an aluminium salt and an oligonucleotide containing a CpG motif. Most preferably, the immunogenic composition includes both an aluminium salt and an oligonucleotide containing a CpG motif. Use of the combination of a mineral salt, such as an aluminium salt, and an oligonucleotide containing a CpG motif provide for an enhanced immune response. This improved immune response is wholly unexpected and could not be predicted from the use of either agent alone. The invention therefore includes an oligonucleotide containing a CpG motif, a mineral salt such as an aluminium salt, and an antigen, such as a *Chlamydia pneumoniae* antigen.

T CELLS IMPLICATED IN THE CMI RESPONSE

At least two special types of T cells are required to initiate and/or to enhance CMI and humoral responses. The antigenic receptors on a particular subset of T cells which express a CD4 co-receptor can be T helper (Th) cells or CD4 T cells (herein after called T helper cells) and they recognise antigenic peptides bound to MHC class II molecules. In contrast, the antigenic receptors on a particular subset of T cells which express a CD8 co-receptor are called Cytotoxic T lymphocytes (CTLs) or CD8+ T cells (hereinafter called CD8+ T cells) and they react with antigens displayed on MHC Class I molecules.

HELPER T CELLS

Helper T cells or CD4+ cells can be further divided into two functionally distinct subsets: Th1 and Th2 which differ in their cytokine and effector function. Th1 and Th2 responses have been shown to be regulated not only in a positive but also in a negative way such that Th1 cellular responses are augmented by Th1 cytokines such as IL-2, IL-12 and IFN-gamma and decreased by Th2 cytokines such as IL-4 and IL-10. In contrast, antibody responses are enhanced by Th2 cytokines such as IL-4 and IL-10 but are downregulated by Th1 cytokines such as IFN-gamma and another cytokine IL-12 that enhances IFN-gamma and is produced by monocytes. Thus, classic Th1 cytokines such as IFN-gamma, IL-2 and IL-12 can be regarded as immune

co-factors that induce an effective inflammatory response. In contrast, the classic Th2 cytokines such as IL-4 and IL-10 can be regarded as cytokines that will suppress a severe inflammatory response.

5 CD8+ T CELLS

CD8+ T cells may function in more than one way. The best known function of CD8+ T cells is the killing or lysis of target cells bearing peptide antigen in the context of an MHC class I molecule. Hence the reason why these cells are often termed cytotoxic T lymphocytes (CTL). However, another function, perhaps of greater protective
10 relevance in certain infections is the ability of CD8+ T cells to secrete interferon gamma (IFN-gamma). Thus assays of lytic activity and of IFN-gamma release are both of value in measuring CD8+ T cell immune response (eg in an ELISPOT assay as set forth below). In infectious diseases there is evidence to suggest that CD8+ T cells can protect by killing an infectious agent comprising an infectious antigen at the
15 early stages of a disease before any symptoms of disease are produced.

ENHANCED CMI RESPONSE

The present invention concerns methods, processes and compositions capable of enhancing and/or modulating the CMI response in a host subject against a target
20 antigen. As used herein, the term "enhancing" encompasses improvements in all aspects of the CMI response which include but are not limited to a stimulation and/or augmentation and/or potentiation and/or up-regulation of the magnitude and/or duration, and/or quality of the CMI response to an antigen or a nucleotide sequence encoding an antigen of interest. By way of example, the CMI response may be
25 enhanced by either (i) enhancing the activation and/or production and/or proliferation of CD8+ T cells that recognise a target antigen and/or (ii) shifting the CMI response from a Th2 to a Th1 type response. This enhancement of the Th1 associated responses is of particular value in responding to intracellular infections because, as explained above, the CMI response is enhanced by activated Th1 (such as, for example, IFN-
30 gamma inducing) cells.

Such an enhanced immune response may be generally characterized by increased titers of interferon-producing CD4⁺ and/or CD8⁺ T lymphocytes, increased antigen-specific CD8+ T cell activity, and a T helper 1-like immune response (Th1) against
35 the antigen of interest (characterized by increased antigen-specific antibody titers of the subclasses typically associated with cellular immunity (such as, for example IgG2a), usually with a concomitant reduction of antibody titers of the subclasses typically associated with humoral immunity (such as, for example IgG1)) instead of a T helper 2-like immune response (Th2).

40 The enhancement of a CMI response may be determined by a number of well-known assays, such as by lymphoproliferation (lymphocyte activation) assays, CD8+ T cell assays, or by assaying for T-lymphocytes specific for the epitope in a sensitized subject (see, for example, Erickson *et al.* (1993) J. Immunol. 151: 4189-4199; and
45 Doe *et al.* (1994) Eur. J. Immunol. 24: 2369-2376) or CD8+ T cell ELISPOT assays for measuring Interferon gamma production (Miyahara *et al* PNAS(USA) (1998) 95: 3954-3959).

ENHANCED T-CELL RESPONSE

As used herein, the term "enhancing a T-cell response" encompasses improvements in all aspects of the T-cell response which include but are not limited to a stimulation and/or augmentation and/or potentiation and/or up-regulation of the magnitude and/or duration, and/or quality of the T-cell response to an antigen (which may be repeatedly administered) or a nucleotide sequence encoding an antigen. The antigen may be a *Chlamydia* antigen, preferably a *Chlamydia pneumoniae* antigen. By way of example, the T-cell response may be enhanced by either enhancing the activation and/or production and/or distribution and/or proliferation of the induced T-cells and/or longevity of the T-cell response to T-cell inducing/modulating antigen or nucleotide sequence encoding an antigen. The enhancement of the T-cell response in a host subject may be associated with the enhancement and/or modulation of the Th1 immune response in the host subject.

The enhancement of the T-cell response may be determined by a number of well-known assays, such as by lymphoproliferation (lymphocyte activation) assays, CD8+ T-cell cytotoxic cell assays, or by assaying for T-lymphocytes specific for the epitope in a sensitized subject (see, for example, Erickson *et al.* (1993) J. Immunol. 151: 4189-4199; and Doe *et al.* (1994) Eur. J. Immunol. 24: 2369-2376) or CD8+ T-cell ELISPOT assays for measuring Interferon gamma production (Miyahara *et al* PNAS(USA) (1998) 95: 3954-3959).

Activated Th1 cells enhance cellular immunity (including an increase in antigen-specific CTL production) and are therefore of particular value in responding to intracellular infections. Activated Th1 cells may secrete one or more of IL-2, IFN-gamma, and TNF-beta. A Th1 immune response may result in local inflammatory reactions by activating macrophages, NK (natural killer) cells, and CD8 cytotoxic T cells (CTLs). A Th1 immune response may also act to expand the immune response by stimulating growth of B and T cells with IL-12. Th1 stimulated B cells may secrete IgG2a.

Activated Th2 cells enhance antibody production and are therefore of value in responding to extracellular infections. Activated Th2 cells may secrete one or more of IL-4, IL-5, IL-6, and IL-10. A Th2 immune response may result in the production of IgG1, IgE, IgA and memory B cells for future protection.

ANTIGEN

Each disease causing agent or disease state has associated with it an antigen or immunodominant epitope on the antigen which is crucial in immune recognition and ultimate elimination or control of a disease causing agent or disease state in a host. In order to mount a humoral and/or cellular immune response against a particular disease, the host immune system must come in contact with an antigen or an immunodominant epitope on an antigen associated with that disease state.

As used herein, the term "antigen" refers to any agent, generally a macromolecule, which can elicit an immunological response in an individual. The term "antigen" is used interchangeably with the term "immunogen". The immunological response may be of B- and/or T-lymphocytic cells. The term may be used to refer to an individual macromolecule or to a homogeneous or heterogeneous population of antigenic macromolecules. As used herein, "antigen" is used to refer to a protein molecule or

portion thereof which contains one or more antigenic determinants or epitopes. As used herein, the term "antigen" means an immunogenic peptide or protein of interest comprising one or more epitopes capable of inducing a CMI response to an infectious *Chlamydia* pathogen. The antigen can include but is not limited to an auto-antigen, a self-antigen, a cross-reacting antigen, an alloantigen, a tolerogen, an allergen, a haptén, an immunogen or parts thereof as well as any combinations thereof.

EPITOPE

As used herein, the term "epitope" generally refers to the site on an antigen which is recognised by a T-cell receptor and/or an antibody. Preferably it is a short peptide derived from or as part of a protein antigen. However the term is also intended to include peptides with glycopeptides and carbohydrate epitopes. Several different epitopes may be carried by a single antigenic molecule. The term "epitope" also includes modified sequences of amino acids or carbohydrates which stimulate responses which recognise the whole organism. It is advantageous if the selected epitope is an epitope of an infectious agent, such as a *Chlamydia* bacterium, which causes the infectious disease.

SEQ ID Nos 1-86 in the compositions of the invention may be supplemented or substituted with molecules comprising fragments of SEQ ID Nos 1-86. Such fragments may comprise at least n consecutive monomers from the molecules and, depending on the particular sequence, n is either (i) 7 or more for protein molecules (eg. 8, 18, 20 or more), preferably such that the fragment comprises an epitope from the sequence, or (ii) 10 or more for nucleic acid molecules (eg 15, 18, 20, 25, 30, 35, 40 or more).

SOURCE OF EPITOPES

The epitope can be generated from knowledge the amino acid and corresponding DNA sequences of the peptide or polypeptide, as well as from the nature of particular amino acids (e.g., size, charge, etc.) and the codon dictionary, without undue experimentation. See, e.g., Ivan Roitt, *Essential Immunology*, 1988; Kendrew, *supra*; Janis Kuby, *Immunology*, 1992 e.g., pp. 79-81. Some guidelines in determining whether a protein will stimulate a response, include: Peptide length—preferably the peptide is about 8 or 9 amino acids long to fit into the MHC class I complex and about 13-25 amino acids long to fit into a class II MHC complex. This length is a minimum for the peptide to bind to the MHC complex. It is preferred for the peptides to be longer than these lengths because cells may cut peptides. The peptide may contain an appropriate anchor motif which will enable it to bind to the various class I or class II molecules with high enough specificity to generate an immune response (See Bocchia, M. *et al*, Specific Binding of Leukemia Oncogene Fusion Protein Peptides to HLA Class I Molecules, *Blood* 85:2680-2684; Englehard, VH, Structure of peptides associated with class I and class II MHC molecules *Ann. Rev. Immunol.* 12:181 (1994)). This can be done, without undue experimentation, by comparing the sequence of the protein of interest with published structures of peptides associated with the MHC molecules. Thus, the skilled artisan can ascertain an epitope of interest by comparing the protein sequence with sequences listed in the protein data base.

T CELL EPITOPES

Preferably one or more antigens of the present invention contain one or more T cell epitopes. As used herein, the term "T cell epitope" refers generally to those features

of a peptide structure which are capable of inducing a T cell response. In this regard, it is accepted in the art that T cell epitopes comprise linear peptide determinants that assume extended conformations within the peptide-binding cleft of MHC molecules (Unanue *et al.* (1987) *Science* 236: 551-557). As used herein, a T cell epitope is generally a peptide having at least about 3-5 amino acid residues, and preferably at least 5-10 or more amino acid residues. However, as used herein, the term "T cell epitope" encompasses any MHC Class I-or MHC Class II restricted peptide. The ability of a particular T cell epitope to stimulate/enhance a CMI response may be determined by a number of well-known assays, such as by lymphoproliferation (lymphocyte activation) assays, CD8+ T-cell cytotoxic cell assays, or by assaying for T-lymphocytes specific for the epitope in a sensitized subject. See, e. g., Erickson *et al.* (1993) *J. Immunol.* 151: 4189-4199; and Doe *et al.* (1994) *Eur. J. Immunol.* 24: 2369-2376 or CD8+ T-cell ELISPOT assays for measuring Interferon gamma production (Miyahara *et al* PNAS(USA) (1998) 95: 3954-3959).

CD8+ T-CELL EPITOPES

Preferably the antigens of the present invention comprise CD8+ T-cell inducing epitopes. A CD8+ T-cell -inducing epitope is an epitope capable of stimulating the formation, or increasing the activity, of specific CD8+ T-cells following its administration to a host subject. The CD8+ T-cell epitopes may be provided in a variety of different forms such as a recombinant string of one or two or more epitopes. CD8+ T-cell epitopes have been identified and can be found in the literature, for many different diseases. It is possible to design epitope strings to generate CD8+ T-cell response against any chosen antigen that contains such CD8+ T-cell epitopes. Advantageously, CD8+ T-cell inducing epitopes may be provided in a string of multiple epitopes which are linked together without intervening sequences so that unnecessary nucleic acid material is avoided.

T HELPER EPITOPES

Preferably the antigens of the present invention comprise helper T lymphocyte epitopes. Various methods are available to identify T helper cell epitopes suitable for use in accordance herewith. For example, the amphipathicity of a peptide sequence is known to effect its ability to function as a T helper cell inducer. A full discussion of T helper cell-inducing epitopes is given in U.S. Patent 5,128,319, incorporated herein by reference.

B CELL EPITOPES

Preferably the antigens of the present invention comprise a mixture of CD8+ T-cell epitopes and B cell epitopes. As used herein, the term "B cell epitope" generally refers to the site on an antigen to which a specific antibody molecule binds. The identification of epitopes which are able to elicit an antibody response is readily accomplished using techniques well known in the art. See, e. g., Geysen *et al.* (1984) *Proc. Natl. Acad. Sci. USA* 81: 3998-4002 (general method of rapidly synthesizing peptides to determine the location of immunogenic epitopes in a given antigen); U. S. Patent No. 4,708,871 (procedures for identifying and chemically synthesizing epitopes of antigens); and Geysen *et al.* (1986) *Molecular Immunology* 23: 709-715 (technique for identifying peptides with high affinity for a given antibody).

COMBINATION OF EPITOPES

In a preferred embodiment of the present invention, the antigen or antigen combination comprises a mixture of a CD8+ T-cell -inducing epitopes and a T helper cell-inducing epitopes.

- 5 As is well known in the art, T and B cell inducing epitopes are frequently distinct from each other and can comprise different peptide sequences. Therefore certain regions of a protein's peptide chain can possess either T cell or B cell epitopes. Therefore, in addition to the CD8+ T-cell epitopes, it may be preferable to include one or more epitopes recognised by T helper cells, to augment the immune response
10 generated by the CD8+ T-cell epitopes.

- The mechanism of enhancing a CD8+ T-cell induced response *in vivo* by T helper cell inducing agents is not completely clear. However, without being bound by theory, it is likely that the enhancing agent, by virtue of its ability to induce T helper cells, will
15 result in increased levels of necessary cytokines that assist in the clonal expansion and dissemination of specific CD8+ T-cells. Regardless of the underlying mechanism, it is envisioned that the use of mixtures of helper T cell and CD8+ T-cell -inducing antigen combinations of the present invention will assist in the enhancement of the CMI response. Particularly suitable T helper cell epitopes are ones which are active
20 in individuals of different HLA types, for example T helper epitopes from tetanus (against which most individuals will already be primed). It may also be useful to include B cell epitopes for stimulating B cell responses and antibody production. Synthetic nucleotide sequences may also be constructed to produce two types of immune responses: T cell only and T cell combined with a B cell response.

25

IMMUNODOMINANT EPITOPE

- When an individual is immunized with an antigen or combination of antigens or nucleotide sequence or combinations of nucleotide sequences encoding multiple epitopes of a target antigen, in many instances the majority of responding T
30 lymphocytes will be specific for one or more linear epitopes from that target antigen and/or a majority of the responding B lymphocytes will be specific for one or more linear or conformational epitopes for the antigen or combination of antigens.. For the purposes of the present invention, then, such epitopes are referred to as "immunodominant epitopes". In an antigen having several immunodominant
35 epitopes, a single epitope may be the most dominant in terms of commanding a specific T or B cell response.

- As the Examples show, at least sixteen peptides of the present invention were recognised by IFN-gamma positive CD8+ T cell populations which were actually
40 expanded as a result of bacterial infection.

ADJUVANTS

- The compositions of the present invention may be administered in conjunction with other immunoregulatory agents. In particular, the compositions of the present
45 invention may be administered with an adjuvant.

The inclusion of an adjuvant and in particular, a genetic adjuvant may be useful in further enhancing or modulating the CMI response. An adjuvant may enhance the CMI response by enhancing the immunogenicity of a co-administered antigen in an

immunized subject, as well inducing a Th1-like immune response against the co-administered antigen which is beneficial in a vaccine product.

5 An immune response and particularly a CMI response may be refined, by the addition of adjuvants to combinations of antigens or nucleotide sequences encoding combinations of antigens which lead to particularly effective compositions for eliciting a long lived and sustained enhanced CMI response.

10 As used herein, the term "adjuvant" refers to any material or composition capable of specifically or non-specifically altering, enhancing, directing, redirecting, potentiating or initiating an antigen-specific immune response.

15 The term "adjuvant" includes but is not limited to a bacterial ADP-ribosylating exotoxin, a biologically active factor, immunomodulatory molecule, biological response modifier or immunostimulatory molecule such as a cytokine, an interleukin, a chemokine or a ligand or an epitope (such as a helper T cell epitope) and optimally combinations thereof which, when administered with an antigen, antigen composition or nucleotide sequence encoding such antigens enhances or potentiates or modulates the CMI response relative to the CMI response generated upon administration of the antigen or combination of antigens alone. The adjuvant may be any adjuvant known in the art which is appropriate for human or animal use.

25 Immunomodulatory molecules such as cytokines (TNF-alpha, IL-6, GM-CSF, and IL-2), and co-stimulatory and accessory molecules (B7-1, B7-2) may be used as adjuvants in a variety of combinations. In one embodiment GM-CSF is not administered to subject before, in or after the administration regimen. Simultaneous production of an immunomodulatory molecule and an antigen of interest at the site of expression of the antigen of interest may enhance the generation of specific effectors which may help to enhance the CMI response. The degree of enhancement of the CMI response may be dependent upon the specific immunostimulatory molecules and/or adjuvants used because different immunostimulatory molecules may elicit different mechanisms for enhancing and/or modulating the CMI response. By way of example, the different effector mechanisms/immunomodulatory molecules include but are not limited to augmentation of help signal (IL-2), recruitment of professional APC (GM-CSF), increase in T cell frequency (IL-2), effect on antigen processing pathway and MHC expression (IFN-gamma and TNF-alpha) and diversion of immune response away from the Th1 response and towards a Th2 response (LTB) (see WO 97/02045). Unmethylated CpG containing oligonucleotides (see WO96/02555) are also preferential inducers of a Th1 response and are suitable for use in the present invention.

45 Without being bound by theory, the inclusion of an adjuvant is advantageous because the adjuvant may help to enhance the CMI response to the expressed antigen by diverting the Th2 response to a Th1 response and/or specific effector associated mechanisms to an expressed epitope with the consequent generation and maintenance of an enhanced CMI response (see, for example, the teachings in WO 97/02045).

50 The inclusion of an adjuvant with an antigen or nucleotide sequence encoding the antigen is also advantageous because it may result in a lower dose or fewer doses of the antigen/antigenic combination being necessary to achieve the desired CMI response in

the subject to which the antigen or nucleotide sequence encoding the antigen is administered, or it may result in a qualitatively and/or quantitatively different immune response in the subject. The effectiveness of an adjuvant can be determined by administering the adjuvant with the antigen in parallel with the antigen alone to animals and comparing antibody and/or cellular-mediated immunity in the two groups using standard assays such as radioimmunoassay, ELISAs, CD8+ T-cell assays, and the like, all well known in the art. Typically, the adjuvant is a separate moiety from the antigen, although a single molecule (such for example, CTB) can have both adjuvant and antigen properties.

As used herein, the term "genetic adjuvant" refers to an adjuvant encoded by a nucleotide sequence and which, when administered with the antigen enhances the CMI response relative to the CMI response generated upon administration of the antigen alone.

Bacterial ADP-ribosylating toxins and detoxified derivatives thereof may be used as adjuvants in the invention. Preferably, the protein is derived from *E. coli* (i.e., *E. coli* heat labile enterotoxin "LT"), cholera ("CT"), or pertussis ("PT").

In one preferred embodiment, the genetic adjuvant is a bacterial ADP-ribosylating exotoxin.

ADP-ribosylating bacterial toxins are a family of related bacterial exotoxins and include diphtheria toxin (DT), pertussis toxin (PT), cholera toxin (CT), the *E. coli* heat-labile toxins (LT1 and LT2), *Pseudomonas* endotoxin A, *Pseudomonas* exotoxin S, *B. cereus* exoenzyme, *B. sphaericus* toxin, *C. botulinum* C2 and C3 toxins, *C. limosum* exoenzyme, as well as toxins from *C. perfringens*, *C. spiriforma* and *C. difficile*, *Staphylococcus aureus* EDIN, and ADP-ribosylating bacterial toxin mutants such as CRM₁₉₇, a non-toxic diphtheria toxin mutant (see, e.g., Bixler *et al.* (1989) *Adv. Exp. Med. Biol.* 251:175; and Constantino *et al.* (1992) *Vaccine*). Most ADP-ribosylating bacterial toxins are organized as an A:B multimer, wherein the A subunit contains the ADP-ribosyltransferase activity, and the B subunit acts as the binding moiety. Preferred ADP-ribosylating bacterial toxins for use in the compositions of the present invention include cholera toxin and the *E. coli* heat-labile toxins.

Cholera toxin (CT) and the related *E. coli* heat labile enterotoxins (LT) are secretion products of their respective enterotoxic bacterial strains that are potent immunogens and exhibit strong toxicity when administered systemically, orally, or mucosally. Both CT and LT are known to provide adjuvant effects for antigen when administered via the intramuscular or oral routes. These adjuvant effects have been observed at doses below that required for toxicity. The two toxins are extremely similar molecules, and are at least about 70-80% homologous at the amino acid level.

Preferably the genetic adjuvant is cholera toxin (CT), enterotoxigenic *E. Coli* heat-labile toxin (LT), or a derivative, subunit, or fragment of CT or LT which retains adjuvanticity. In an even more preferred embodiment, the genetic adjuvant is LT. In another preferred embodiment, the genetic adjuvant may be CTB or LTB.

Preferably the enterotoxin is a non-toxic enterotoxin.

The use of detoxified ADP-ribosylating toxins as mucosal adjuvants is described in WO 95/17211 and as parenteral adjuvants in WO 98/42375. The toxin or toxoid is

preferably in the form of a holotoxin, comprising both A and B subunits. Preferably, the A subunit contains a detoxifying mutation; preferably the B subunit is not mutated. Preferably, the adjuvant is a detoxified LT mutant such as LT-K63, LT-R72, and LTR192G. The use of ADP-ribosylating toxins and detoxified derivatives thereof, particularly LT-K63 and LT-R72, as adjuvants can be found in the following references each of which is specifically incorporated by reference herein in their entirety (Beignon, *et al.* Infection and Immunity (2002) 70(6):3012 – 3019; Pizza, *et al.*, Vaccine (2001) 19:2534 – 2541; Pizza, *et al.*, Int. J. Med. Microbiol (2000) 290(4-5):455-461; Schariton-Kersten *et al.* Infection and Immunity (2000) 68(9):5306 – 5313; Ryan *et al.* Infection and Immunity (1999) 67(12):6270 – 6280; Partidos *et al.* Immunol. Lett. (1999) 67(3):209 – 216; Peppoloni *et al.* Vaccines (2003) 2(2):285 – 293; and Pine *et al.* J. Control Release (2002) 85(1-3):263 – 270). Numerical reference for amino acid substitutions is preferably based on the alignments of the A and B subunits of ADP-ribosylating toxins set forth in Domenighini *et al.*, Mol. Microbiol (1995) 15(6):1165 – 1167, specifically incorporated herein by reference in its entirety.

By way of further example, at least one of the enterotoxin subunit coding regions may be genetically modified to detoxify the subunit peptide encoded thereby, for example wherein the truncated A subunit coding region has been genetically modified to disrupt or inactivate ADP-ribosyl transferase activity in the subunit peptide expression product (see, for example, WO 03/004055).

Thus, these results demonstrate that this genetic adjuvant is particularly desirable where an even more enhanced CMI response is desired. Other desirable genetic adjuvants include but are not limited to nucleotide sequences encoding IL-10, IL-12, IL-13, the interferons (IFNs) (for example, IFN-alpha, IFN-ss, and IFN-gamma), and preferred combinations thereof. Still other such biologically active factors that enhance the CMI response may be readily selected by one of skill in the art, and a suitable plasmid vector containing same constructed by known techniques.

Preferred further adjuvants include, but are not limited to, one or more of the following set forth below:

35 *Mineral Containing Compositions*

Mineral containing compositions suitable for use as adjuvants in the invention include mineral salts, such as aluminium salts and calcium salts. The invention includes mineral salts such as hydroxides (*e.g.* oxyhydroxides), phosphates (*e.g.* hydroxyphosphates, orthophosphates), sulphates, *etc.* (*e.g.* see chapters 8 & 9 of ref. Bush and Everett (2001) Int J Syst Evol Microbiol 51: 203-220), or mixtures of different mineral compounds, with the compounds taking any suitable form (*e.g.* gel, crystalline, amorphous, *etc.*), and with adsorption being preferred. The mineral containing compositions may also be formulated as a particle of metal salt. See WO 00/23105.

Aluminum salts may be included in immunogenic compositions and/or vaccines of the invention such that the dose of Al³⁺ is between 0.2 and 1.0 mg per dose.

Preferably the adjuvant is alum, preferably an aluminium salt such as aluminium hydroxide (AlOH) or aluminium phosphate or aluminium sulfate. Still more preferably the adjuvant is aluminium hydroxide (AlOH).

- 5 Preferably a mineral salt, such as an aluminium salt, is combined with and another adjuvant, such as an oligonucleotide containing a CpG motif or an ADP ribosylating toxin. Still more preferably, the mineral salt is combined with an oligonucleotide containing a CpG motif.

10 *Oil-Emulsions*

Oil-emulsion compositions suitable for use as adjuvants in the invention include squalene-water emulsions, such as MF59 (5% Squalene, 0.5% Tween 80, and 0.5% Span 85, formulated into submicron particles using a microfluidizer). See WO90/14837. See also, Frey et al., "Comparison of the safety, tolerability, and immunogenicity of a MF59-adjuvanted influenza vaccine and a non-adjuvanted influenza vaccine in non-elderly adults", *Vaccine* (2003) 21:4234-4237. MF59 is used as the adjuvant in the FLUAD™ influenza virus trivalent subunit vaccine.

20 Particularly preferred adjuvants for use in the compositions are submicron oil-in-water emulsions. Preferred submicron oil-in-water emulsions for use herein are squalene/water emulsions optionally containing varying amounts of MTP-PE, such as a submicron oil-in-water emulsion containing 4-5% w/v squalene, 0.25-1.0% w/v Tween 80™ (polyoxyethylsorbitan monooleate), and/or 0.25-1.0% Span 85™ (sorbitan trioleate), and, optionally, N-acetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1'-2'-dipalmitoyl-*sn*-glycero-3-hydroxyphosphoryloxy)-ethylamine (MTP-PE), for example, the submicron oil-in-water emulsion known as "MF59" (International Publication No. WO90/14837; US Patent Nos. 6,299,884 and 6,451,325, incorporated herein by reference in their entireties; and Ott et al., "MF59 -- Design and Evaluation of a Safe and Potent Adjuvant for Human Vaccines" in *Vaccine Design: The Subunit and Adjuvant Approach* (Powell, M.F. and Newman, M.J. eds.) Plenum Press, New York, 1995, pp. 277-296). MF59 contains 4-5% w/v Squalene (e.g. 4.3%), 0.25-0.5% w/v Tween 80™, and 0.5% w/v Span 85™ and optionally contains various amounts of MTP-PE, formulated into submicron particles using a microfluidizer such as Model 110Y microfluidizer (Microfluidics, Newton, MA). For example, MTP-PE may be present in an amount of about 0-500 µg/dose, more preferably 0-250 µg/dose and most preferably, 0-100 µg/dose. As used herein, the term "MF59-0" refers to the above submicron oil-in-water emulsion lacking MTP-PE, while the term MF59-MTP denotes a formulation that contains MTP-PE. For instance, "MF59-100" contains 100 µg MTP-PE per dose, and so on. MF69, another submicron oil-in-water emulsion for use herein, contains 4.3% w/v squalene, 0.25% w/v Tween 80™, and 0.75% w/v Span 85™ and optionally MTP-PE. Yet another submicron oil-in-water emulsion is MF75, also known as SAF, containing 10% squalene, 0.4% Tween 80™, 5% pluronic-blocked polymer L121, and thr-MDP, also microfluidized into a submicron emulsion. MF75-MTP denotes an MF75 formulation that includes MTP, such as from 100-400 µg MTP-PE per dose.

Submicron oil-in-water emulsions, methods of making the same and immunostimulating agents, such as muramyl peptides, for use in the compositions, are described in detail in International Publication No. WO90/14837 and US Patent Nos. 6,299,884 and 6,451,325, incorporated herein by reference in their entireties.

Complete Freund's adjuvant (CFA) and incomplete Freund's adjuvant (IFA) may also be used as adjuvants in the invention.

Saponin Formulations

5 Saponin formulations, may also be used as adjuvants in the invention. Saponins are a heterologous group of sterol glycosides and triterpenoid glycosides that are found in the bark, leaves, stems, roots and even flowers of a wide range of plant species. Saponin from the bark of the *Quillaia saponaria* Molina tree have been widely studied as adjuvants. Saponin can also be commercially obtained from *Smilax ornata* (sarsapilla), *Gypsophilla paniculata* (brides veil), and *Saponaria officianalis* (soap root). Saponin adjuvant formulations include purified formulations, such as QS21, as well as lipid formulations, such as ISCOMs. Saponin compositions have been purified using High Performance Thin Layer Chromatography (HP-LC) and Reversed Phase High Performance Liquid Chromatography (RP-HPLC). Specific purified fractions using these techniques have been identified, including QS7, QS17, QS18, QS21, QH-A, QH-B and QH-C. Preferably, the saponin is QS21. A method of production of QS21 is disclosed in U.S. Patent No. 5,057,540. Saponin formulations may also comprise a sterol, such as cholesterol (see WO 96/33739). Combinations of saponins and cholesterol can be used to form unique particles called Immunostimulating Complex (ISCOMs). ISCOMs typically also include a phospholipid such as phosphatidylethanolamine or phosphatidylcholine. Any known saponin can be used in ISCOMs. Preferably, the ISCOM includes one or more of Quil A, QHA and QHC. ISCOMs are further described in EP 0 109 942, WO 96/11711 and WO 96/33739. Optionally, the ISCOMS may be devoid of additional detergent. See WO 00/07621.

A review of the development of saponin based adjuvants can be found in Barr *et al* (1998) Advanced Drug Delivery Reviews 32: 247-271 and Sjolander *et al* (1998) Advanced Drug Delivery Reviews (1998) 32: 321-338.

30 *Virosomes and Virus Like Particles (VLPs)*
Virosomes and Virus Like Particles (VLPs) can also be used as adjuvants in the invention. These structures generally contain one or more proteins from a virus optionally combined or formulated with a phospholipid. They are generally non-pathogenic, non-replicating and generally do not contain any of the native viral genome. The viral proteins may be recombinantly produced or isolated from whole viruses. These viral proteins suitable for use in virosomes or VLPs include proteins derived from influenza virus (such as HA or NA), Hepatitis B virus (such as core or capsid proteins), Hepatitis E virus, measles virus, Sindbis virus, Rotavirus, Foot-and-Mouth Disease virus, Retrovirus, Norwalk virus, human Papilloma virus, HIV, RNA-phages, Q β -phage (such as coat proteins), GA-phage, fr-phage, AP205 phage, and Ty (such as retrotransposon Ty protein p1). VLPs are discussed further in WO 03/024480, WO 03/024481; Niikura *et al* Virology (2002) 293:273 – 280; Lenz *et al* Journal of Immunology (2001) 5246 – 5355; Pinto, *et al* Journal of Infectious Diseases (2003) 188:327 – 338; and Gerber *et al* Journal of Virology (2001) 75(10):4752 – 4760; Virosomes are discussed further in, for example, Gluck *et al* Vaccine (2002) 20:B10 –B16.

Bacterial or Microbial Derivatives

Adjuvants suitable for use in the invention include bacterial or microbial derivatives such as:

- 5 *Non-toxic derivatives of enterobacterial lipopolysaccharide (LPS)*
Such derivatives include Monophosphoryl lipid A (MPL) and 3-O-deacylated MPL (3dMPL). 3dMPL is a mixture of 3 De-O-acylated monophosphoryl lipid A with 4, 5 or 6 acylated chains. A preferred "small particle" form of 3 De-O-acylated monophosphoryl lipid A is disclosed in EP 0 689 454. Such "small particles" of
- 10 3dMPL are small enough to be sterile filtered through a 0.22 micron membrane (see EP 0 689 454). Other non-toxic LPS derivatives include monophosphoryl lipid A mimics, such as aminoalkyl glucosaminide phosphate derivatives *e.g.* RC-529. See Johnson *et al.* (1999) *Bioorg Med Chem Lett* 9:2273-2278.

Lipid A Derivatives

Lipid A derivatives include derivatives of lipid A from *Escherichia coli* such as OM-174. OM-174 is described for example in Meraldi *et al.* Vaccine (2003) 21:2485 – 2491; Pajak, *et al* Vaccine (2003) 21:836 – 842.

5

Immunostimulatory oligonucleotides

Immunostimulatory oligonucleotides suitable for use as adjuvants in the invention include nucleotide sequences containing a CpG motif (a sequence containing an unmethylated cytosine followed by guanosine and linked by a phosphate bond).
10 Bacterial double stranded RNA or oligonucleotides containing palindromic or poly(dG) sequences have also been shown to be immunostimulatory.

The CpG's can include nucleotide modifications/analogs such as phosphorothioate modifications and can be double-stranded or single-stranded. Optionally, the
15 guanosine may be replaced with an analog such as 2'-deoxy-7-deazaguanosine. See Kandimalla, *et al* Nucleic Acids Research (2003) 31(9): 2393 – 2400; WO 02/26757 and WO 99/62923 for examples of possible analog substitutions. The adjuvant effect of CpG oligonucleotides is further discussed in Krieg Nature Medicine (2003) 9(7): 831 – 835; McCluskie, *et al* FEMS Immunology and Medical Microbiology (2002)
20 32:179 – 185; WO 98/40100, U.S. Patent No. 6,207,646, U.S. Patent No. 6,239,116, and U.S. Patent No. 6,429,199.

The CpG sequence may be directed to TLR9, such as the motif GTCGTT or TTCGTT. See Kalman *et al* (1999) (Nature Genetics 21: 385-389). The CpG
25 sequence may be specific for inducing a Th1 immune response, such as a CpG-A ODN, or it may be more specific for inducing a B cell response, such a CpG-B ODN. CpG-A and CpG-B ODNs are discussed in Blackwell, *et al* J. Immunol. (2003) 170(8):4061 – 4068; Krieg BBRC (2003) 306:948 – 953; and WO 01/95935. Preferably, the CpG is a CpG-A ODN.

30

Preferably, the CpG oligonucleotide is constructed so that the 5' end is accessible for receptor recognition. Optionally, two CpG oligonucleotide sequences may be attached at their 3' ends to form "immunomers". See, for example, Kandimalla, *et al* (2003) 31(part 3):664 – 658; Bhagat *et al* BBRC (2003) 300:853 – 861 and WO 03/035836.

35

Preferably the adjuvant is CpG. Even more preferably, the adjuvant is Alum and an oligonucleotide containing a CpG motif or ALOH and an oligonucleotide containing a CpG motif.

Human Immunomodulators

Human immunomodulators suitable for use as adjuvants in the invention include cytokines, such as interleukins (*e.g.* IL-1, IL-2, IL-4, IL-5, IL-6, IL-7, IL-12, *etc.*), interferons (*e.g.* interferon- γ), macrophage colony stimulating factor, and tumor
40 necrosis factor.

45

ADP-ribosylating toxins and detoxified derivatives thereof.

Bacterial ADP-ribosylating toxins and detoxified derivatives thereof may be used as adjuvants in the invention. Preferably, the protein is derived from *E. coli* (*i.e.*, *E. coli* heat labile enterotoxin "LT), cholera ("CT"), or pertussis ("PT"). The use of
50 detoxified ADP-ribosylating toxins as mucosal adjuvants is described in WO95/17211

- and as parenteral adjuvants in WO98/42375. Preferably, the adjuvant is a detoxified LT mutant such as LT-K63, LT-R72, and LTR192G. The use of ADP-ribosylating toxins and detoxified derivatives thereof, particularly LT-K63 and LT-R72, as adjuvants can be found in the following references, each of which is specifically incorporated by reference herein in their entirety: Beignon, et al., "The LTR72 Mutant of Heat-Labile Enterotoxin of Escherichia coli Enhances the Ability of Peptide Antigens to Elicit CD4+ T Cells and Secrete Gamma Interferon after Coapplication onto Bare Skin", *Infection and Immunity* (2002) 70(6):3012-3019; Pizza, et al., "Mucosal vaccines: non toxic derivatives of LT and CT as mucosal adjuvants", *Vaccine* (2001) 19:2534-2541; Pizza, et al., "LTK63 and LTR72, two mucosal adjuvants ready for clinical trials" *Int. J. Med. Microbiol* (2000) 290(4-5):455-461; Scharton-Kersten et al., "Transcutaneous Immunization with Bacterial ADP-Ribosylating Exotoxins, Subunits and Unrelated Adjuvants", *Infection and Immunity* (2000) 68(9):5306-5313; Ryan et al., "Mutants of Escherichia coli Heat-Labile Toxin Act as Effective Mucosal Adjuvants for Nasal Delivery of an Acellular Pertussis Vaccine: Differential Effects of the Nontoxic AB Complex and Enzyme Activity on Th1 and Th2 Cells" *Infection and Immunity* (1999) 67(12):6270-6280; Partidos et al., "Heat-labile enterotoxin of Escherichia coli and its site-directed mutant LTK63 enhance the proliferative and cytotoxic T-cell responses to intranasally co-immunized synthetic peptides", *Immunol. Lett.* (1999) 67(3):209-216; Peppoloni et al., "Mutants of the Escherichia coli heat-labile enterotoxin as safe and strong adjuvants for intranasal delivery of vaccines", *Vaccines* (2003) 2(2):285-293; and Pine et al., (2002) "Intranasal immunization with influenza vaccine and a detoxified mutant of heat labile enterotoxin from Escherichia coli (LTK63)" *J. Control Release* (2002) 85(1-3):263-270. Numerical reference for amino acid substitutions is preferably based on the alignments of the A and B subunits of ADP-ribosylating toxins set forth in Domenighini et al., *Mol. Microbiol* (1995) 15(6):1165-1167, specifically incorporated herein by reference in its entirety.
- Preferably the adjuvant is an ADP-ribosylating toxin and an oligonucleotide containing a CpG motif (see for example, WO 01/34185)
- Preferably the adjuvant is a detoxified ADP-ribosylating toxin and an oligonucleotide containing a CpG motif.
- Preferably the detoxified ADP-ribosylating toxin is LTK63 or LTK72.
- Preferably the adjuvant is LTK63. Preferably the adjuvant is LTK72.
- Preferably the adjuvant is LTK63 and an oligonucleotide containing a CpG motif.
- Preferably the adjuvant is LTK72 and an oligonucleotide containing a CpG motif.

Bioadhesives and Mucoadhesives

- Bioadhesives and mucoadhesives may also be used as adjuvants in the invention. Suitable bioadhesives include esterified hyaluronic acid microspheres (Singh *et al.* (2001) *J. Cont. Rel.* 70:267-276) or mucoadhesives such as cross-linked derivatives of poly(acrylic acid), polyvinyl alcohol, polyvinyl pyrrolidone, polysaccharides and carboxymethylcellulose. Chitosan and derivatives thereof may also be used as adjuvants in the invention. See for example, WO99/27960.

Microparticles

- Microparticles may also be used as adjuvants in the invention. Microparticles (*i.e.* a particle of ~100nm to ~150µm in diameter, more preferably ~200nm to ~30µm in diameter, and most preferably ~500nm to ~10µm in diameter) formed from materials that are biodegradable and non-toxic (*e.g.* a poly(α -hydroxy acid), a

polyhydroxybutyric acid, a polyorthoester, a polyanhydride, a polycaprolactone, *etc.*), with poly(lactide-co-glycolide) are preferred, optionally treated to have a negatively-charged surface (*e.g.* with SDS) or a positively-charged surface (*e.g.* with a cationic detergent, such as CTAB).

5

Liposomes

Examples of liposome formulations suitable for use as adjuvants are described in U.S. Patent No. 6,090,406, U.S. Patent No. 5,916,588, and EP 0 626 169.

10 *Polyoxyethylene ether and Polyoxyethylene Ester Formulations*

Adjuvants suitable for use in the invention include polyoxyethylene ethers and polyoxyethylene esters (WO99/52549). Such formulations further include polyoxyethylene sorbitan ester surfactants in combination with an octoxynol (WO01/21207) as well as polyoxyethylene alkyl ethers or ester surfactants in
15 combination with at least one additional non-ionic surfactant such as an octoxynol (WO01/21152). Preferred polyoxyethylene ethers are selected from the following group: polyoxyethylene-9-lauryl ether (laureth 9), polyoxyethylene-9-stearyl ether, polyoxyethylene-8-stearyl ether, polyoxyethylene-4-lauryl ether, polyoxyethylene-35-lauryl ether, and polyoxyethylene-23-lauryl ether.

20

Polyphosphazene (PCPP)

PCPP formulations are described, for example, in Andrianov *et al* Biomaterials (1998) 19(1 – 3):109 – 115; Payne *et al* Adv. Drug. Delivery Review (1998) 31(3):185 – 196.

25

Muramyl peptides

Examples of muramyl peptides suitable for use as adjuvants in the invention include N-acetyl-muramyl-L-threonyl-D-isoglutamine (thr-MDP), N-acetyl-normuramyl-L-alanyl-D-isoglutamine (nor-MDP), and N-acetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1'-2'-dipalmitoyl-*sn*-glycero-3-hydroxyphosphoryloxy)-ethylamine (MTP-
30 PE).

Imidazoquinolone Compounds

Examples of imidazoquinolone compounds suitable for use as adjuvants in the invention include Imiquimod and its homologues, described further in Stanley, "Imiquimod and the imidazoquinolones: mechanism of action and therapeutic potential" Clin Exp Dermatol (2002) 27(7):571 – 577; and Jones, "Resiquimod 3M", Curr Opin Investig Drugs (2003) 4(2):214 – 218. The invention may also comprise combinations of aspects of one or more of the adjuvants identified above. For example, the following
40 adjuvant compositions may be used in the invention:

- (1) a saponin and an oil-in-water emulsion (WO99/11241);
- (2) a saponin (*e.g.*, QS21) + a non-toxic LPS derivative (*e.g.*, 3dMPL) (see WO 94/00153);
- (3) a saponin (*e.g.*, QS21) + a non-toxic LPS derivative (*e.g.*, 3dMPL) + a
45 cholesterol;
- (4) a saponin (*e.g.* QS21) + 3dMPL + IL-12 (optionally + a sterol (WO98/57659); combinations of 3dMPL with, for example, QS21 and/or oil-in-water emulsions (European patent applications 0835318, 0735898 and 0761231).

- (5) SAF, containing 10% Squalane, 0.4% Tween 80, 5% pluronic-block polymer L121, and thr-MDP, either microfluidized into a submicron emulsion or vortexed to generate a larger particle size emulsion.
- (6) RibiTM adjuvant system (RAS), (Ribi Immunochem) containing 2% Squalene, 0.2% Tween 80, and one or more bacterial cell wall components from the group consisting of monophosphorylipid A (MPL), trehalose dimycolate (TDM), and cell wall skeleton (CWS), preferably MPL + CWS (DetoxTM); and
- (7) one or more mineral salts (such as an aluminum salt) + a non-toxic derivative of LPS (such as 3dPML).
- (7) combinations of 3dMPL with, for example, QS21 and/or oil-in-water emulsions (See European patent applications 0835318, 0735898 and 0761231);
- (8) one or more mineral salts (such as an aluminum salt) + a non-toxic derivative of LPS (such as 3dPML); and
- (9) one or more mineral salts (such as an aluminum salt) + an immunostimulatory oligonucleotide (such as a nucleotide sequence including a CpG motif).

Aluminium salts and MF59 are preferred adjuvants for parenteral immunisation. Mutant bacterial toxins are preferred mucosal adjuvants. Bacterial toxins and bioadhesives are preferred adjuvants for use with mucosally-delivered vaccines, such as nasal vaccines.

The composition may include an antibiotic.

Preferably the compositions of the present invention are administered with alum and/or CpG sequences.

25 *Nucleic Acid*

The antigens or epitopes of the present invention may be administered as nucleotide sequences encoding the antigens or epitopes. As used herein, the term nucleotide sequence refers to one of more nucleotide sequences which encode one or more epitopes which are used in the compositions or combinations of the present invention. The term "nucleotide sequence (NOI)" is synonymous with the term "polynucleotide" or "nucleic acid". The NOI may be DNA or RNA of genomic or synthetic or of recombinant origin. The NOI may be double-stranded or single-stranded whether representing the sense or antisense strand or combinations thereof. For some applications, preferably, the NOI is DNA. For some applications, preferably, the NOI is prepared by use of recombinant DNA techniques (e.g. recombinant DNA). For some applications, preferably, the NOI is cDNA. For some applications, preferably, the NOI may be the same as the naturally occurring form.

40 The term "nucleic acid" includes DNA and RNA, and also their analogues, such as those containing modified backbones (e.g. phosphorothioates, *etc.*), and also peptide nucleic acids (PNA), *etc.* The invention includes nucleic acid comprising sequences complementary to those described above (e.g. for antisense or probing purposes).

45 Nucleic acid according to the invention can be prepared in many ways (e.g. by chemical synthesis, from genomic or cDNA libraries, from the organism itself, *etc.*) and can take various forms (e.g. single stranded, double stranded, vectors, probes, *etc.*). They are preferably prepared in substantially pure form (*i.e.* substantially free from other *Chlamydial* or host cell nucleic acids).

The invention provides a process for producing nucleic acid of the invention, comprising the step of amplifying nucleic acid using a primer-based amplification method (*e.g.* PCR).

- 5 The invention provides a process for producing nucleic acid of the invention, comprising the step of synthesising at least part of the nucleic acid by chemical means.

VECTOR

- 10 In one embodiment of the present invention, an antigen or antigenic combination or NOI encoding same is administered directly to a host subject. In another embodiment of the present invention, a vector comprising an NOI is administered to a host subject. Preferably the NOI is prepared and/or administered using a genetic vector. As it is well known in the art, a vector is a tool that allows or facilitates the transfer of an
15 entity from one environment to another. In accordance with the present invention, and by way of example, some vectors used in recombinant DNA techniques allow entities, such as a segment of DNA (such as a heterologous DNA segment, such as a heterologous cDNA segment), to be transferred into a host and/or a target cell for the purpose of replicating the vectors comprising the NOI of the present invention and/or
20 expressing the antigens or epitopes of the present invention encoded by the NOI. Examples of vectors used in recombinant DNA techniques include but are not limited to plasmids, chromosomes, artificial chromosomes or viruses. The term "vector" includes expression vectors and/or transformation vectors. The term "expression vector" means a construct capable of *in vivo* or *in vitro/ex vivo* expression. The term
25 "transformation vector" means a construct capable of being transferred from one species to another.

NAKED DNA

- 30 The vectors comprising the NOI of the present invention may be administered directly as "a naked nucleic acid construct", preferably further comprising flanking sequences homologous to the host cell genome. As used herein, the term "naked DNA" refers to a plasmid comprising the NOI of the present invention together with a short promoter region to control its production. It is called "naked" DNA because the plasmids are not carried in any delivery vehicle. When such a DNA plasmid enters a host cell, such
35 as a eukaryotic cell, the proteins it encodes are transcribed and translated within the cell.

VIRAL VECTORS

- 40 Alternatively, the vectors comprising the NOI of the present invention may be introduced into suitable host cells using a variety of viral techniques which are known in the art, such as for example infection with recombinant viral vectors such as retroviruses, herpes simplex viruses and adenoviruses. The vector may be a recombinant viral vectors. Suitable recombinant viral vectors include but are not limited to adenovirus vectors, adeno-associated viral (AAV) vectors, herpes-virus vectors, a retroviral vector,
45 lentiviral vectors, baculoviral vectors, pox viral vectors or parvovirus vectors (see Kestler *et al* 1999 Human Gene Ther 10(10):1619-32). In the case of viral vectors, administration of the NOI is mediated by viral infection of a target cell.

TARGETED VECTOR

The term "targeted vector" refers to a vector whose ability to infect or transfect or transduce a cell or to be expressed in a host and/or target cell is restricted to certain cell types within the host subject, usually cells having a common or similar phenotype.

5 EXPRESSION VECTOR

Preferably, the NOI of the present invention which is inserted into a vector is operably linked to a control sequence that is capable of providing for the expression of the antigens or epitopes by the host cell, i.e. the vector is an expression vector. The agent produced by a host cell may be secreted or may be contained intracellularly depending on the NOI and/or the vector used. As will be understood by those of skill in the art, expression vectors containing the NOI can be designed with signal sequences which direct secretion of the NOI through a particular prokaryotic or eukaryotic cell membrane.

FUSION PROTEINS

15 The *Chlamydia pneumoniae* antigens used in the invention may be present in the composition as individual separate polypeptides, but it is preferred that at least two (i.e. 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20) of the antigens are expressed as a single polypeptide chain (a 'hybrid' polypeptide). Hybrid polypeptides offer two principal advantages: first, a polypeptide that may be unstable or poorly expressed on its own can be assisted by adding a suitable hybrid partner that overcomes the problem; second, commercial manufacture is simplified as only one expression and purification need be employed in order to produce two polypeptides which are both antigenically useful.

25 The hybrid polypeptide may comprise two or more polypeptide sequences from the first antigen group. Accordingly, the invention includes a composition comprising a first amino acid sequence and a second amino acid sequence, wherein said first and second amino acid sequences are selected from a *Chlamydia* bacterium, preferably a *Chlamydia pneumoniae* antigen or a fragment thereof of the first antigen group. Preferably, the first and second amino acid sequences in the hybrid polypeptide comprise different epitopes.

35 The hybrid polypeptide may comprise two or more polypeptide sequences from the second antigen group. Accordingly, the invention includes a composition comprising a first amino acid sequence and a second amino acid sequence, wherein said first and second amino acid sequences are selected from a *Chlamydia pneumoniae* antigen or a fragment thereof of the second antigen group. Preferably, the first and second amino acid sequences in the hybrid polypeptide comprise difference epitopes.

40 The hybrid polypeptide may comprise one or more polypeptide sequences from the first antigen group and one or more polypeptide sequences from the second antigen group. Accordingly, the invention includes a composition comprising a first amino acid sequence and a second amino acid sequence, said first amino acid sequence selected from a *Chlamydia pneumoniae* antigen or a fragment thereof from the first antigen group and said second amino acid sequence selected from a *Chlamydia* bacterium, preferably a *Chlamydia pneumoniae* antigen or a fragment thereof from the second antigen group. Preferably, the first and second amino acid sequences in the hybrid polypeptide comprise difference epitopes.

- The hybrid polypeptide may comprise one or more polypeptide sequences from the first antigen group and one or more polypeptide sequences from the third antigen group or the fourth antigen group or the fifth antigen group or the sixth antigen group or the seventh antigen group or the eighth antigen group or the ninth antigen group or the tenth antigen group. Accordingly, the invention includes a composition comprising a first amino acid sequence and a second amino acid sequence, said first amino acid sequence selected from a *Chlamydia pneumoniae* antigen or a fragment thereof from the first antigen group and said second amino acid sequence selected from a *Chlamydia pneumoniae* antigen or a fragment thereof from the third antigen group or the fourth antigen group or the fifth antigen group or the sixth antigen group or the seventh antigen group or the eighth antigen group or the ninth antigen group or the tenth antigen group. Preferably, the first and second amino acid sequences in the hybrid polypeptide comprise difference epitopes.
- The hybrid polypeptide may comprise one or more polypeptide sequences from the second antigen group and one or more polypeptide sequences from the third antigen group or the fourth antigen group or the fifth antigen group or the sixth antigen group or the seventh antigen group or the eighth antigen group or the ninth antigen group or the tenth antigen group. Accordingly, the invention includes a composition comprising a first amino acid sequence and a second amino acid sequence, said first amino acid sequence selected from a *Chlamydia pneumoniae* antigen or a fragment thereof from the second antigen group and said second amino acid sequence selected from a *Chlamydia pneumoniae* antigen or a fragment thereof from the third antigen group or the fourth antigen group or the fifth antigen group or the sixth antigen group or the seventh antigen group or the eighth antigen group or the ninth antigen group or the tenth antigen group. Preferably, the first and second amino acid sequences in the hybrid polypeptide comprise difference epitopes.
- Hybrids consisting of amino acid sequences from two, three, four, five, six, seven, eight, nine, or ten *Chlamydia pneumoniae* antigens are preferred. In particular, hybrids consisting of amino acid sequences from two, three, four, or five *Chlamydia pneumoniae* antigens are preferred. Different hybrid polypeptides may be mixed together in a single formulation. Within such combinations, a *Chlamydia pneumoniae* antigen may be present in more than one hybrid polypeptide and/or as a non-hybrid polypeptide. It is preferred, however, that an antigen is present either as a hybrid or as a non-hybrid, but not as both.
- Two-antigen hybrids for use in the invention may comprise any one of the combinations disclosed above.
- Hybrid polypeptides can be represented by the formula $\text{NH}_2\text{-A-}\{-\text{X-L-}\}_n\text{-B-COOH}$, wherein: X is an amino acid sequence of a *Chlamydia pneumoniae* antigen or a fragment thereof from the first antigen group, the second antigen group or the third antigen group or the fourth antigen group or the fifth antigen group or the sixth antigen group or the seventh antigen group or the eighth antigen group or the ninth antigen group or the tenth antigen group.; L is an optional linker amino acid sequence; A is an optional N-terminal amino acid sequence; B is an optional C-terminal amino acid sequence; and n is 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14 or 15.

If a -X- moiety has a leader peptide sequence in its wild-type form, this may be included or omitted in the hybrid protein. In some embodiments, the leader peptides will be deleted except for that of the -X- moiety located at the N-terminus of the hybrid protein *i.e.* the leader peptide of X₁ will be retained, but the leader peptides of X₂ ... X_n will be omitted. This is equivalent to deleting all leader peptides and using the leader peptide of X₁ as moiety -A-.

For each *n* instances of {-X-L-}, linker amino acid sequence -L- may be present or absent. For instance, when *n*=2 the hybrid may be NH₂-X₁-L₁-X₂-L₂-COOH, NH₂-X₁-X₂-COOH, NH₂-X₁-L₁-X₂-COOH, NH₂-X₁-X₂-L₂-COOH, *etc.* Linker amino acid sequence(s) -L- will typically be short (*e.g.* 20 or fewer amino acids *i.e.* 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples comprise short peptide sequences which facilitate cloning, poly-glycine linkers (*i.e.* comprising Gly_{*n*} where *n* = 2, 3, 4, 5, 6, 7, 8, 9, 10 or more), and histidine tags (*i.e.* His_{*n*} where *n* = 3, 4, 5, 6, 7, 8, 9, 10 or more). Other suitable linker amino acid sequences will be apparent to those skilled in the art. A useful linker is GSGGGG (SEQ ID No 77), with the Gly-Ser dipeptide being formed from a *Bam*HI restriction site, thus aiding cloning and manipulation, and the (Gly)₄ tetrapeptide being a typical poly-glycine linker.

-A- is an optional N-terminal amino acid sequence. This will typically be short (*e.g.* 40 or fewer amino acids *i.e.* 39, 38, 37, 36, 35, 34, 33, 32, 31, 30, 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples include leader sequences to direct protein trafficking, or short peptide sequences which facilitate cloning or purification (*e.g.* histidine tags *i.e.* His_{*n*} where *n* = 3, 4, 5, 6, 7, 8, 9, 10 or more). Other suitable N-terminal amino acid sequences will be apparent to those skilled in the art. If X₁ lacks its own N-terminus methionine, -A- is preferably an oligopeptide (*e.g.* with 1, 2, 3, 4, 5, 6, 7 or 8 amino acids) which provides a N-terminus methionine.

-B- is an optional C-terminal amino acid sequence. This will typically be short (*e.g.* 40 or fewer amino acids *i.e.* 39, 38, 37, 36, 35, 34, 33, 32, 31, 30, 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples include sequences to direct protein trafficking, short peptide sequences which facilitate cloning or purification (*e.g.* comprising histidine tags *i.e.* His_{*n*} where *n* = 3, 4, 5, 6, 7, 8, 9, 10 or more), or sequences which enhance protein stability. Other suitable C-terminal amino acid sequences will be apparent to those skilled in the art. Most preferably, *n* is 2 or 3.

The invention also provides nucleic acid encoding hybrid polypeptides of the invention. Furthermore, the invention provides nucleic acid which can hybridise to this nucleic acid, preferably under "high stringency" conditions (*e.g.* 65°C in a 0.1xSSC, 0.5% SDS solution).

The NOI of the present invention may be expressed as a fusion protein comprising an adjuvant and/or a biological response modifier and/or immunomodulator fused to the antigens or epitopes of the present invention to further enhance and/or augment the CMI response obtained. The biological response modifier may act as an adjuvant in the sense of providing a generalised stimulation of the CMI response. The antigens or epitopes may be attached to either the amino or carboxy terminus of the biological response modifier.

METHODS OF MAKING

Polypeptides of the invention can be prepared by various means (*e.g.* recombinant expression, purification from cell culture, chemical synthesis, *etc.*) and in various forms
 5 (*e.g.* native, fusions, non-glycosylated, lipidated, *etc.*). They are preferably prepared in substantially pure form (*i.e.* substantially free from other *Chlamydial* or host cell proteins).

The invention also provides a process for producing a polypeptide of the invention, comprising the step of culturing a host cell transformed with nucleic acid of the
 10 invention under conditions which induce polypeptide expression. The invention provides a process for producing a polypeptide of the invention, comprising the step of synthesising at least part of the polypeptide by chemical means. The invention further provides a process for producing a composition according to the invention comprising the step of bringing one or more of SEQ IDs 1-86 into combination with
 15 one or more other of SEQ IDs 1-86

Strains

Preferred polypeptides of the invention comprise an amino acid sequence found in *C.pneumoniae* serovars, or in one or more of an epidemiologically prevalent serotype.
 20 Where hybrid polypeptides are used, the individual antigens within the hybrid (*i.e.* individual -X- moieties) may be from one or more strains. Where $n=2$, for instance, X_2 may be from the same strain as X_1 or from a different strain. Where $n=3$, the strains might be (i) $X_1=X_2=X_3$ (ii) $X_1=X_2\neq X_3$ (iii) $X_1\neq X_2=X_3$ (iv) $X_1\neq X_2\neq X_3$ or (v) $X_1=X_3\neq X_2$, *etc.*

25

Heterologous host

Whilst expression of the polypeptides of the invention may take place in *Chlamydia*, the invention preferably utilises a heterologous host. The heterologous host may be prokaryotic (*e.g.* a bacterium) or eukaryotic. It is preferably *E.coli*, but other suitable
 30 hosts include *Bacillus subtilis*, *Vibrio cholerae*, *Salmonella typhi*, *Salmonella typhimurium*, *Neisseria lactamica*, *Neisseria cinerea*, *Mycobacteria* (*e.g.* *M.tuberculosis*), yeasts, *etc.*

Details as to how the molecules which make up the SEQ IDs 1-86 can be produced and
 35 used can be found from the relevant international applications such as WO 00/37494, WO 02/02606 and WO 03/049762 and WO 03/068811 and these details need not be repeated here. Where the composition includes a protein that exists in different nascent and mature forms, the mature form of the protein is preferably used. For example, the
 40 mature form of the *Chlamydia pneumoniae* protein lacking the signal peptide may be used

ADMINISTRATION

Compositions of the invention will generally be administered directly to a patient. Direct delivery may be accomplished by parenteral injection (*e.g.* subcutaneously,
 45 intraperitoneally, intravenously, intramuscularly, or to the interstitial space of a tissue), or by rectal, oral (*e.g.* tablet, spray), vaginal, topical, transdermal {*e.g.* see WO99/27961} or transcutaneous {*e.g.* WO02/074244 and WO02/064162} intranasal {*e.g.* see WO03/028760} ocular, aural, pulmonary or other mucosal administration. The invention may be used to elicit systemic and/or mucosal immunity.

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The compositions of the present invention may be administered, either alone or as part of a composition, via a variety of different routes. Certain routes may be favoured for certain compositions, as resulting in the generation of a more effective immune response, preferably a CMI response, or as being less likely to induce side effects, or as being easier for administration.

By way of example, the compositions of the present invention may be administered via a systemic route or a mucosal route or a transdermal route or it may be administered directly into a specific tissue. As used herein, the term "systemic administration" includes but is not limited to any parenteral routes of administration.

In particular, parenteral administration includes but is not limited to subcutaneous, intraperitoneal, intravenous, intraarterial, intramuscular, or intrasternal injection, intravenous, intraarterial, or kidney dialytic infusion techniques. Preferably, the systemic, parenteral administration is intramuscular injection.

In one preferred embodiment of the method, the compositions of the present invention are administered via a transdermal route. While it is believed that any accepted mode and route of immunization can be employed and nevertheless achieve some advantages in accordance herewith, the examples below demonstrate particular advantages with transdermal NOI administration. In this regard, and without being bound by theory, it is believed that transdermal administration of a composition may be preferred because it more efficiently activates the cell mediated immune (CMI) arm of the immune system.

The term "transdermal" delivery intends intradermal (e.g., into the dermis or epidermis), transdermal (e.g., "percutaneous") and transmucosal administration, i.e., delivery by passage of an agent into or through skin or mucosal tissue. See, e.g., *Transdermal Drug Delivery: Developmental Issues and Research Initiatives*, Hadgraft and Guy (eds.), Marcel Dekker, Inc., (1989); *Controlled Drug Delivery: Fundamentals and Applications*, Robinson and Lee (eds.), Marcel Dekker Inc., (1987); and *Transdermal Delivery of Drugs*, Vols. 1-3, Kydonieus and Berner (eds.), CRC Press, (1987). Thus, the term encompasses delivery of an agent using a particle delivery device (e.g., a needleless syringe) such as those described in U.S. Patent No. 5,630,796, as well as delivery using particle-mediated delivery devices such as those described in U.S. Patent No. 5,865,796.

As used herein, the term "mucosal administration" includes but is not limited to oral, intranasal, intravaginal, intrarectal, intratracheal, intestinal and ophthalmic administration.

Mucosal routes, particularly intranasal, intratracheal, and ophthalmic are preferred for protection against natural exposure to environmental pathogens such as RSV, flu virus and cold viruses or to allergens such as grass and ragweed pollens and house dust mites. The enhancement of the immune response, preferably the CMI response will enhance the protective effect against a subsequently encountered target antigen such as an allergen or microbial agent.

In another preferred embodiment of the present invention, the compositions of the present invention may be administered to cells which have been isolated from the host subject. In this preferred embodiment, preferably the composition is administered to professional antigen presenting cells (APCs), such as dendritic cells. APCs may be derived from a host subject and modified *ex vivo* to express an antigen of interest and

then transferred back into the host subject to induce an enhanced CMI response. Dendritic cells are believed to be the most potent APCs for stimulating enhanced CMI responses because the expressed epitopes of the antigen of interest must be acquired, processed and presented by professional APCs to T cells (both Th1 and Th2 helper
 5 cells as well as CD8+ T-cells) in order to induce an enhanced CMI response.

PARTICLE ADMINISTRATION

Particle-mediated methods for delivering the compositions of the present invention are known in the art. Thus, once prepared and suitably purified, the above-described
 10 antigens or NOI encoding same can be coated onto core carrier particles using a variety of techniques known in the art. Carrier particles are selected from materials which have a suitable density in the range of particle sizes typically used for intracellular delivery from a gene gun device. The optimum carrier particle size will, of course, depend on the diameter of the target cells.

15 By "core carrier" is meant a carrier on which a guest antigen or guest nucleic acid (e.g., DNA, RNA) is coated in order to impart a defined particle size as well as a sufficiently high density to achieve the momentum required for cell membrane penetration, such that the guest molecule can be delivered using particle-mediated techniques (see, e.g., U.S. Patent No. 5,100,792). Core carriers typically include
 20 materials such as tungsten, gold, platinum, ferrite, polystyrene and latex. See e.g., *Particle Bombardment Technology for Gene Transfer*, (1994) Yang, N. ed., Oxford University Press, New York, NY pages 10-11. Tungsten and gold particles are preferred. Tungsten particles are readily available in average sizes of 0.5 to 2.0 microns in diameter. Gold particles or microcrystalline gold (e. g., gold powder A1570, available from Engelhard Corp., East Newark, NJ) will also find use with the present invention. Gold particles provide uniformity in size (available from Alpha
 25 Chemicals in particle sizes of 1-3 microns, or available from Degussa, South Plainfield, NJ in a range of particle sizes including 0.95 microns). Microcrystalline gold provides a diverse particle size distribution, typically in the range of 0.5-5 microns. However, the irregular surface area of microcrystalline gold provides for highly efficient coating with nucleic acids. A number of methods are known and have been described for coating or precipitating NOIs onto gold or tungsten particles. Most such methods generally combine a predetermined amount of gold or tungsten with
 30 plasmid DNA, CaCl₂ and spermidine. The resulting solution is vortexed continually during the coating procedure to ensure uniformity of the reaction mixture. After precipitation of the NOI, the coated particles can be transferred to suitable membranes and allowed to dry prior to use, coated onto surfaces of a sample module or cassette, or loaded into a delivery cassette for use in particular gene gun instruments.

40 The particle compositions or coated particles are administered to the individual in a manner compatible with the dosage formulation, and in an amount that will be effective for the purposes of the invention. The amount of the composition to be delivered (e. g., about 0.1 mg to 1 mg, more preferably 1 to 50 ug of the antigen or
 45 allergen, depends on the individual to be tested. The exact amount necessary will vary depending on the age and general condition of the individual to be treated, and an appropriate effective amount can be readily determined by one of skill in the art upon reading the instant specification.

50 HOST MAMMALIAN SUBJECT

As used herein, the term "host mammalian subject" means any member of the subphylum cordata, including, without limitation, humans and other primates, including non-human primates such as chimpanzees and other apes and monkey species; farm animals such as cattle, sheep, pigs, goats and horses; domestic mammals
 5 such as dogs and cats; laboratory animals including rodents such as mice, rats and guinea pigs; birds, including domestic, wild and game birds such as chickens, turkeys and other gallinaceous birds, ducks, geese, and the like. The terms do not denote a particular age. Thus, both adult and newborn individuals are intended to be covered. The methods described herein are intended for use in any of the above vertebrate
 10 species, since the immune systems of all of these vertebrates operate similarly. If a mammal, the subject will preferably be a human, but may also be a domestic livestock, laboratory subject or pet animal.

The mammal is preferably a human. Where the vaccine is for prophylactic use, the human is preferably a child (e.g. a toddler or infant) or a teenager; where the vaccine
 15 is for therapeutic use, the human is preferably a teenager or an adult. A vaccine intended for children may also be administered to adults e.g. to assess safety, dosage, immunogenicity, etc.

PREVENT AND/OR TREAT

20 The invention also provides the use of the compositions of the invention in the manufacture of a medicament for raising an immune response in a mammal. The medicament is preferably a vaccine and to the preparation of a vaccine to prevent and/or treat an disorder associated with a *Chlamydia* bacterium. It is to be appreciated that all references herein to treatment include curative, palliative and
 25 prophylactic treatment.

The administration of antigenic combinations of the present invention or a composition comprising the NOI encoding the antigenic combinations may be for either "prophylactic" or "therapeutic" purpose. As used herein, the term "therapeutic"
 30 or "treatment" includes any of following: the prevention of infection or reinfection; the reduction or elimination of symptoms; and the reduction or complete elimination of a pathogen. Treatment may be effected prophylactically (prior to infection) or therapeutically (following infection).

35 Prophylaxis or therapy includes but is not limited to eliciting an effective immune response, preferably a CMI immune response and/or alleviating, reducing, curing or at least partially arresting symptoms and/or complications resulting from a T cell mediated immune disorder. When provided prophylactically, the composition of the present invention is typically provided in advance of any symptom. The prophylactic
 40 administration of the composition of the present invention is to prevent or ameliorate any subsequent infection or disease. When provided therapeutically, the composition of the present invention is typically provided at (or shortly after) the onset of a symptom of infection or disease. Thus the composition of the present invention may be provided either prior to the anticipated exposure to a disease causing agent or
 45 disease state or after the initiation of an infection or disease.

Whether prophylactic or therapeutic administration (either alone or as part of a composition) is the more appropriate will usually depend upon the nature of the disease. By way of example, immunotherapeutic composition of the present
 50 invention could be used in immunotherapy protocols to actively inducing immunity

by vaccination. This latter form of treatment is advantageous because the immunity is prolonged. On the other hand a vaccine composition will preferably, though not necessarily be used prophylactically to induce an effective CMI response against subsequently encountered antigens or portions thereof (such as epitopes) related to the target antigen.

These uses and methods are preferably for the prevention and/or treatment of a disease caused by a *Chlamydia* (e.g. trachoma, pelvic inflammatory disease, epididymitis, infant pneumonia, atherosclerosis, cardiovascular disease *etc.*). The compositions may also be effective against *C.pneumoniae*.

PROPHYLACTICALLY OR THERAPEUTICALLY OR IMMUNOLOGICALLY EFFECTIVE AMOUNT

The composition dose administered to a host subject, in the context of the present invention, should be sufficient to effect a beneficial prophylactic or therapeutic immune response, preferably a CMI response in the subject over time.

The invention also provides a method for raising an immune response in a mammal comprising the step of administering an effective amount of a composition of the invention. The immune response is preferably protective and preferably involves antibodies and/or cell-mediated immunity. The method may raise a booster response.

As used herein, the term "prophylactically or therapeutically effective dose" means a dose in an amount sufficient to elicit an enhanced immune response, preferably a CMI response to one or more antigens or epitopes and/or to alleviate, reduce, cure or at least partially arrest symptoms and/or complications from a T cell mediated immune disorder.

Immunogenic compositions used as vaccines comprise an immunologically effective amount of antigen(s), as well as any other components, as needed. By 'immunologically effective amount', it is meant that the administration of that amount to an individual, either in a single dose or as part of a series, is effective for treatment or prevention. This amount varies depending upon the health and physical condition of the individual to be treated, age, the taxonomic group of individual to be treated (e.g. non-human primate, primate, *etc.*), the capacity of the individual's immune system to synthesise antibodies, the degree of protection desired, the formulation of the vaccine, the treating doctor's assessment of the medical situation, and other relevant factors. It is expected that the amount will fall in a relatively broad range that can be determined through routine trials.

The mammal is preferably a human. Where the vaccine is for prophylactic use, the human is preferably a child (e.g. a toddler or infant) or a teenager or an adult; where the vaccine is for therapeutic use, the human is preferably a teenager or an adult. A vaccine intended for children may also be administered to adults e.g. to assess safety, dosage, immunogenicity, *etc.* Preferably, the human is a teenager. More preferably, the human is a pre-adolescent teenager. Even more preferably, the human is a pre-adolescent female or male. Preferably the pre-adolescent male or female is around 9-12 years of age.

One way of assessing the immunogenicity of the component proteins of the immunogenic compositions of the present invention is to express the proteins recombinantly and to screen patient sera or mucosal secretions by immunoblot or by protein or DNA microarray. A positive reaction between the protein and the patient serum indicates that the patient has previously mounted an immune response to the protein in question- that is, the protein is an immunogen. This method may also be used to identify immunodominant proteins.

One way of checking efficacy of therapeutic treatment involves monitoring *Chlamydia* infection after administration of the composition of the invention. One way of checking efficacy of prophylactic treatment involves monitoring immune responses against the *Chlamydia* antigen, such as the *Chlamydia pneumoniae* antigen in the compositions of the invention after administration of the composition. For example, checking efficacy of prophylactic treatment may involve monitoring immune responses both systemically (such as monitoring the level of IgG1 and IgG2a production) and mucosally (such as monitoring the level of IgA production) against the *Chlamydia pneumoniae* antigens in the compositions of the invention after administration of the composition. Typically, serum *Chlamydia* specific antibody responses are determined post-immunization but pre-challenge whereas mucosal *Chlamydia* specific antibody body responses are determined post-immunization and post-challenge.

These uses and methods are preferably for the prevention and/or treatment of a disease caused by *Chlamydia pneumoniae* (e.g. pneumonia, bronchitis, pharyngitis, sinusitis, erythema nodosum, asthma, atherosclerosis, stroke, myocardial infarctions, coronary artery disease, etc.).

The vaccine compositions of the present invention can be evaluated in *in vitro* and *in vivo* animal models prior to host, e.g., human, administration. For example, *in vitro* neutralization by Peterson *et al* (1988) is suitable for testing vaccine compositions directed toward *Chlamydia*, preferably *Chlamydia pneumoniae*.

One example of such an *in vitro* test is described as follows. Hyper-immune antisera is diluted in PBS containing 5% guinea pig serum, as a complement source. *Chlamydia pneumoniae* (10^4 IFU; inclusion forming units) are added to the antisera dilutions. The antigen-antibody mixtures are incubated at 37°C for 45 minutes and inoculated into duplicate confluent Hep-2 or HeLa cell monolayers contained in glass vials (e.g., 15 by 45 mm), which have been washed twice with PBS prior to inoculation. The monolayer cells are infected by centrifugation at 1000X g for 1 hour followed by stationary incubation at 37°C for 1 hour. Infected monolayers are incubated for 48 or 72 hours, fixed and stained with *Chlamydia* specific antibody, such as anti-MOMP. Inclusion-bearing cells are counted in ten fields at a magnification of 200X. Neutralization titer is assigned on the dilution that gives 50% inhibition as compared to control monolayers/IFU.

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- The efficacy of immunogenic compositions can also be determined in vivo by challenging animal models of *Chlamydia pneumoniae* infection, e.g., guinea pigs or mice, with the immunogenic compositions. The immunogenic compositions may or may not be derived from the same serovars as the challenge serovars. Preferably the immunogenic compositions are derivable from the same serovars as the challenge serovars. More preferably, The serovars of the present invention are obtainable from clinical isolates or from culture collections such as the American Tissue Culture Collection (ATCC).
- 10 In vivo efficacy models include but are not limited to: (i) A murine infection model using human *Chlamydia pneumoniae* serotypes; (ii) a murine disease model which is a murine model using a mouse-adapted *Chlamydia pneumoniae* strain, such as the *Chlamydia pneumoniae* mouse pneumonitis (MoPn) strain also known as *Chlamydia muridarum*; and (iii) a primate model using human *Chlamydia pneumoniae* isolates.
- 15 The MoPn strain is a mouse pathogen while human *Chlamydia pneumoniae* serotypes are human pathogens (see for example, Brunham et al (2000) J Infect Dis 181 (Suppl 3) S538-S543; Murrin et al (2000) J Infect Dis 181 (Suppl 3) S544-S551 and Read et al (2000) NAR 28(6); 1397-1406). As the Examples demonstrate, human *Chlamydia pneumoniae* serotypes can be used in mouse models although they normally require high inocula or pretreatment with progesterone. Progesterone is generally used because it seems to render the epithelium more susceptible to chlamydial infection (see Pal et al 2003 Vaccine 21: 1455-1465). On the other hand, MoPn, which was originally isolated from mouse tissues, is thought to be a natural murine pathogen and thus offers an evolutionarily adapted pathogen for analysis of host-pathogen interactions. Although the MoPn serovar is thought to have a high degree of DNA homology to the human *Chlamydia* serovars, it may also have some unique properties (see for example, Pal et al (2002) Infection and Immunity 70(9); 4812-4817).
- 30 By way of example, in vivo vaccine compositions challenge studies can be performed in the murine model of *Chlamydia pneumoniae* (Morrison et al 1995). A description of one example of this type of approach is as follows. Female mice 7 to 12 weeks of age receive 2.5 mg of depoprovera subcutaneously at 10 and 3 days before vaginal infection. Post-vaccination, mice are infected in the genital tract with 1,500 inclusion-forming units of *Chlamydia pneumoniae* contained in 5ml of sucrose-phosphate-glutamate buffer, pH 7.4. The course of infection is monitored by determining the percentage of inclusion-bearing cells by indirect immunofluorescence with *Chlamydia pneumoniae* specific antisera, or by a Giemsa-stained smear from a scraping from the genital tract of an infected mouse. The presence of antibody titers in the serum of a mouse is determined by an enzyme-linked immunosorbent assay. The immunogenic compositions of the present invention can be administered using a number of different immunization routes such as but not limited to intra-muscularly (i.m.), intra-peritoneal (i.p.), intra-nasal (i.n.), sub-cutaneous (s.c) or transcutaneous (t.c) routes. Generally, any route of administration can be used provided that the desired immune response at the required mucosal surface or surfaces is achieved. Likewise, the challenge serovars may be administered by a number of different routes. Typically, the challenge serovars are administered mucosally, such as but not limited to an intra-nasal (i.n) challenge.
- 50 Alternative in-vivo efficacy models include guinea pig models. For example, in vivo vaccine composition challenge studies in the guinea pig model of *Chlamydia*

pneumoniae infection can be performed. A description of one example of this type of approach follows. Female guinea pigs weighing 450 – 500 g are housed in an environmentally controlled room with a 12 hour light-dark cycle and immunized with vaccine compositions via a variety of immunization routes. Post-vaccination, guinea pigs are infected in the genital tract with the agent of guinea pig inclusion conjunctivitis (GPIC), which has been grown in HeLa or McCoy cells (Rank et al. (1988)). Each animal receives approximately 1.4×10^7 inclusion forming units (IFU) contained in 0.05 ml of sucrose-phosphate-glutamate buffer, pH 7.4 (Schacter, 1980). The course of infection monitored by determining the percentage of inclusion-bearing cells by indirect immunofluorescence with GPIC specific antisera, or by Giemsa-stained smear from a scraping from the genital tract (Rank et al 1988). Antibody titers in the serum is determined by an enzyme-linked immunosorbent assay.

Compositions of the invention will generally be administered directly to a patient. Direct delivery may be accomplished by parenteral injection (e.g. subcutaneously, intraperitoneally, intravenously, intramuscularly, or to the interstitial space of a tissue), or mucosally, such as by rectal, oral (e.g. tablet, spray), vaginal, topical, transdermal (See e.g. WO99/27961) or transcutaneous (See e.g. WO02/074244 and WO02/064162), intranasal (See e.g. WO03/028760), ocular, aural, pulmonary or other mucosal administration.

DOSAGE

Prophylaxis or therapy can be accomplished by a single direct administration at a single time point or multiple time points. Administration can also be delivered to a single or to multiple sites. Some routes of administration, such as mucosal administration via ophthalmic drops may require a higher dose. Those skilled in the art can adjust the dosage and concentration to suit the particular route of delivery.

Dosage treatment can be a single dose schedule or a multiple dose schedule. multiple doses may be used in a primary immunisation schedule and/or in a booster immunisation schedule. in a multiple dose schedule the various doses may be given by the same or different routes e.g. a parenteral prime and mucosal boost, a mucosal prime and parenteral boost, *etc.*

35 HOMOLOGUES

SEQ IDs 1-86 in the compositions of the invention may be supplemented or substituted with molecules comprising sequences homologous (ie. sharing sequence identity) to SEQ ID Nos 1-86.

40 Proteins (including protein antigens) as used in the invention (as encoded by the NOI) may have homology and/or sequence identity with naturally occurring forms. Similarly coding sequences capable of expressing such proteins will generally have homology and/or sequence identity with naturally occurring sequences. Techniques for determining nucleic acid and amino acid "sequence identity" also are known in the art. Typically, such techniques include determining the nucleotide sequence of the mRNA for a gene and/or determining the amino acid sequence encoded thereby, and comparing these sequences to a second nucleotide or amino acid sequence.

50 In general, "identity" refers to an exact nucleotide-to-nucleotide or amino acid-to-amino acid correspondence of two polynucleotides or polypeptide sequences,

respectively. Two or more sequences (polynucleotide or amino acid) can be compared by determining their "percent identity." The percent identity of two sequences, whether nucleic acid or amino acid sequences, is the number of exact matches between two aligned sequences divided by the length of the shorter sequences and multiplied by 100.

An approximate alignment for nucleic acid sequences is provided by the local homology algorithm of Smith and Waterman, *Advances in Applied Mathematics* 2: 482-489 (1981). This algorithm can be applied to amino acid sequences by using the scoring matrix developed by Dayhoff, *Atlas of Protein Sequences and Structure*, M. O. Dayhoff ed., 5 suppl. 3: 353-358, National Biomedical Research Foundation, Washington, D. C., USA, and normalized by Gribskov, *Nucl. Acids Res.* 14 (6): 6745-6763 (1986). An exemplary implementation of this algorithm to determine percent identity of a sequence is provided by the Genetics Computer Group (Madison, WI) in the "BestFit" utility application. The default parameters for this method are described in the Wisconsin Sequence Analysis Package Program Manual, Version 8 (1995) (available from Genetics Computer Group, Madison, WI). A preferred method of establishing percent identity in the context of the present invention is to use the MPSRCH package of programs copyrighted by the University of Edinburgh, developed by John F. Collins and Shane S.

Sturrok, and distributed by IntelliGenetics, Inc. (Mountain View, CA). From this suite of packages the Smith-Waterman algorithm can be employed where default parameters are used for the scoring table (for example, gap open penalty of 12, gap extension penalty of one, and a gap of six). From the data generated the "Match" value reflects "sequence identity." Other suitable programs for calculating the percent identity or similarity between sequences are generally known in the art, for example, another alignment program is BLAST, used with default parameters. For example, BLASTN and BLASTP can be used using the following default parameters: genetic code = standard; filter = none; strand = both; cutoff = 60; expect = 10; Matrix = BLOSUM62 ; Descriptions = 50 sequences; sort by = HIGH SCORE; Databases = non-redundant, GenBank + EMBL + DDBJ + PDB + GenBank CDS translations + Swiss protein + Spupdate + PIR. Details of these programs can be found at the following internet address: <http://www.ncbi.nlm.gov/cgi-bin/BLAST>.

Alternatively, homology can be determined by hybridization of polynucleotides under conditions which form stable duplexes between homologous regions, followed by digestion with single-stranded-specific nuclease (s), and size determination of the digested fragments. Two DNA, or two polypeptide sequences are "substantially homologous" to each other when the sequences exhibit at least about 80%-85%, preferably at least about 90%, and most preferably at least about 95%-98% sequence identity over a defined length of the molecules, as determined using the methods above.

As used herein, substantially homologous or homologous also refers to sequences showing complete identity to the specified DNA or polypeptide sequence. DNA sequences that are substantially homologous or homologous can be identified in a Southern hybridization experiment under, for example, stringent conditions, as defined for that particular system. For example, stringent hybridization conditions can include 50% formamide, 5x Denhardt's Solution, 5x SSC, 0.1% SDS and 100 pg/ml

denatured salmon sperm DNA and the washing conditions can include 2x SSC, 0.1% SDS at 37 C followed by 1x SSC, 0.1% SDS at 68 C. Defining appropriate hybridization conditions is within the skill of the art.

- 5 Preferably the degree of identity is preferably greater than 50% (eg. 65%. 80%. 90%. or more) and include mutants and allelic variants. Sequence identity between the proteins is preferably determined by the Smith-Waterman homology search algorithm as implemented in the MPSRCH program (Oxford. Molecular). using an affine gap search with parameters gap open penalty=12 and gap extension penalty=1.

10 SEQ IDs 1-86 in the compositions of the invention may be supplemented or substituted with nucleic acid which can hybridise to the *Chlamydia* nucleic acid. preferably under "high stringency" conditions (c. 65 C in an 0.1 x SSC, 0.5% SDS solution).

15 *Hypothetical Protein*

As used herein, the term "hypothetical protein" refers to a protein which lacks a known cellular location or a known cellular function. Typically, a hypothetical protein lacks significant homologies with known well characterised proteins.

20 COMPOSITIONS

The invention also provides the compositions of the invention for use as medicaments (eg. as immunogenic compositions or vaccines) or as diagnostic reagents for detecting a *Chlamydia* infection in a host subject. It also provides the use of the compositions in the manufacture of: (i) a medicament for treating or preventing infection due to *Chlamydia pneumoniae* bacteria; (ii) a diagnostic reagent for detecting the presence of *Chlamydia Pneumoniae* bacteria or of antibodies raised against *Chlamydia Pneumoniae* bacteria; and/or (iii) a reagent which can raise antibodies against *Chlamydia pneumoniae* bacteria.

30 The invention also provides a method of treating a patient, comprising administering to the patient a therapeutically effective amount of a composition according to the invention.

35 The present invention provides compositions that are useful for preventing and/or treating T cell mediated immune disorders. In one embodiment, the composition is a pharmaceutical composition. In another preferred embodiment, the composition is an immunotherapeutic composition. In an even more preferred embodiment, the composition is a vaccine composition.. The composition may also comprise a carrier such as a pharmaceutically or immunologically acceptable carrier. Pharmaceutically acceptable carriers or immunologically acceptable carriers are determined in part by the particular composition being administered as well as by the particular method used to administer the composition. Accordingly, there is a wide variety of suitable formulations of pharmaceutical compositions or vaccine compositions or immunotherapeutic compositions of the present invention.

45 *Immunogenic compositions and medicaments*

50 Compositions of the invention are preferably immunogenic compositions, and are more preferably vaccine compositions. The pH of the composition is preferably between 6 and 8, preferably about 7. The pH may be maintained by the use of a

buffer. The composition may be sterile and/or pyrogen-free. The composition may be isotonic with respect to humans.

- 5 Vaccines according to the invention may either be prophylactic (*i.e.* to prevent infection) or therapeutic (*i.e.* to treat infection), but will typically be prophylactic. Accordingly, the invention includes a method for the therapeutic or prophylactic treatment of *Chlamydia pneumoniae* infection in an animal susceptible to *Chlamydia* infection comprising administering to said animal a therapeutic or prophylactic amount of the immunogenic compositions of the invention. Preferably, the
- 10 immunogenic composition comprises a combination of *Chlamydia pneumoniae* antigens, said combination selected from the group consisting of two, three, four, five or all six *Chlamydia pneumoniae* antigens of the first antigen group. Still more preferably, the combination consists of all six *Chlamydia pneumoniae* antigens of the first antigen group.
- 15 Alternatively, the immunogenic composition comprises a combination of *Chlamydia pneumoniae* antigens, said combination selected from the group consisting of two, three, four, five, six, seven, eight, nine, ten, eleven, or twelve *Chlamydia pneumoniae* antigens selected from the first antigen group and the second antigen group.
- 20 Preferably, the combination is selected from the group consisting of three, four, or five *Chlamydia pneumoniae* antigens selected from the second antigen group. Still more preferably, the combination consists of five *Chlamydia pneumoniae* antigens selected from the second antigen group.
- 25 Alternatively, the immunogenic composition comprises a combination of *Chlamydia pneumoniae* antigens, said combination consisting of two, three, four, or five *Chlamydia pneumoniae* antigens of the first antigen group and one, two, three, four, five or six *Chlamydia pneumoniae* antigens of the third antigen group. Preferably, the combination consists of three, four or five *Chlamydia pneumoniae* antigens of the first
- 30 antigen group and one, two, three, four, five or six *Chlamydia pneumoniae* antigens of the third antigen group.
- Alternatively, the immunogenic composition comprises a combination of *Chlamydia pneumoniae* antigens, said combination consisting of two, three, four, five, six, seven, eight, nine, ten, eleven or twelve *Chlamydia pneumoniae* antigens of the first antigen group and the second antigen group and one, two, three, four, five or six *Chlamydia pneumoniae* antigens of the third antigen group. Preferably, the combination is selected from the group consisting of three, four, or five *Chlamydia pneumoniae* antigens from the second antigen group and three, four or five *Chlamydia pneumoniae* antigens from the third antigen group. Still more preferably, the combination consists of five
- 35 *Chlamydia pneumoniae* antigens from the second antigen group and three, four or five *Chlamydia pneumoniae* antigens of the third antigen group.
- 40 In certain embodiments, the composition comprises molecules from different *Chlamydia* species. In some embodiments, the composition may comprise molecules from different serogroups and/or strains of the same *Chlamydia* species. Further embodiments comprise mixtures of one or more *Chlamydia* molecules from different strains.
- 45 Many proteins are relatively conserved between different species serogroups and
- 50

strains of *Chlamydia trachomatis* and *Chlamydia pneumoniae*. To ensure maximum cross-strain recognition and reactivity, regions of proteins that are conserved between different *Chlamydia* species, serogroups and strains can be used in the compositions of the present invention. The invention therefore provides proteins which comprise stretches of amino acid sequence that are shared across the majority of *Chlamydia* strains. Preferably, therefore, the composition comprises a protein comprising a fragment of a *Chlamydia pneumoniae* protein (preferably a protein from SEQ ID Nos 1-86 or more preferably SEQ ID Nos 1-41 wherein said fragment consists of n consecutive conserved amino acids.

Further antigens

The compositions of the invention may further comprise antigen derived from one or more sexually transmitted diseases in addition to *Chlamydia trachomatis*. Preferably the antigen is derived from one or more of the following sexually transmitted diseases: *N.gonorrhoeae* {e.g. i, ii, iii, iv}; human papiloma virus; *Treponema pallidum*; herpes simplex virus (HSV-1 or HSV-2); HIV (HIV-1 or HIV-2); and *Haemophilus ducreyi*.

A preferred composition comprises: (1) at least t of the *Chlamydia pneumoniae* antigens from either the first antigen group or the second antigen group, where t is 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or 13, preferably t is five; (2) one or more antigens from another sexually transmitted disease. Preferably, the sexually transmitted disease is selected from the group consisting of herpes simplex virus, preferably HSV-1 and/or HSV-2; human papillomavirus; *N.gonorrhoeae*; *Treponema pallidum*; and *Haemophilus ducreyi*. These compositions can thus provide protection against the following sexually-transmitted diseases: *Chlamydia*, genital herpes, genital warts, gonorrhoea, syphilis and chancroid (see Stephens et al (1998) Science 282: 754-759).

Where a saccharide or carbohydrate antigen is used, it is preferably conjugated to a carrier protein in order to enhance immunogenicity (For example, Ramsay *et al.* (2001) *Lancet* 357(9251):195-196; Lindberg (1999) *Vaccine* 17 Suppl 2:S28-36; Buttery & Moxon (2000) *J R Coll Physicians Lond* 34:163-168; Ahmad & Chapnick (1999) *Infect Dis Clin North Am* 13:113-133; Goldblatt (1998) *J. Med. Microbiol.* 47:563-567; European patent 0 477 508; US Patent No. 5,306,492; International patent application WO98/42721; *Conjugate Vaccines* (eds. Cruse *et al.*) ISBN 3805549326, particularly vol. 10:48-114; and Hermanson (1996) *Bioconjugate Techniques* ISBN: 0123423368 or 012342335).

Preferred carrier proteins are bacterial toxins or toxoids, such as diphtheria or tetanus toxoids. The CRM₁₉₇ diphtheria toxoid is particularly preferred (*Research Disclosure*, 453077 (Jan 2002)). Other carrier polypeptides include the *N.meningitidis* outer membrane protein EP-A-0372501), synthetic peptides (EP-A-0378881, EP-A-0427347), heat shock proteins (WO93/17712, WO94/03208) pertussis proteins (WO98/58668, EP-A-0471177) protein D from *H.influenzae* (WO00/56360) cytokines (WO91/01146), lymphokines, hormones, growth factors, toxin A or B from *C.difficile* (WO00/61761) iron-uptake proteins WO01/72337) *etc.* Where a mixture comprises capsular saccharides from both serogroups A and C, it may be preferred that the ratio (w/w) of MenA saccharide:MenC saccharide is greater than 1 (e.g. 2:1, 3:1, 4:1, 5:1, 10:1 or higher). Different saccharides can be conjugated to the same or

different type of carrier protein. Any suitable conjugation reaction can be used, with any suitable linker where necessary.

5 Toxic protein antigens may be detoxified where necessary *e.g.* detoxification of pertussis toxin by chemical and/or genetic means. Where a diphtheria antigen is included in the composition it is preferred also to include tetanus antigen and pertussis antigens. Similarly, where a tetanus antigen is included it is preferred also to include diphtheria and pertussis antigens. Similarly, where a pertussis antigen is included it is preferred also to include diphtheria and tetanus antigens.

10 Antigens in the composition will typically be present at a concentration of at least 1 µg/ml each. In general, the concentration of any given antigen will be sufficient to elicit an immune response against that antigen. As an alternative to using protein antigens in the composition of the invention, nucleic acid encoding the antigen may be used Robinson & Torres (1997) *Seminars in Immunology* 9:271-283; Donnelly *et al.* (1997) *Annu Rev Immunol* 15:617-648; Scott-Taylor & Dalgleish (2000) *Expert Opin Investig Drugs* 9:471-480; Apostolopoulos & Plebanski (2000) *Curr Opin Mol Ther* 2:441-447; Ilan (1999) *Curr Opin Mol Ther* 1:116-120; Dubensky *et al.* (2000) *Mol Med* 6:723-732; Robinson & Pertmer (2000) *Adv Virus Res* 55:1-74; Donnelly *et al.* (2000) *Am J Respir Crit Care Med* 162(4 Pt 2):S190-193 and Davis (1999) *Mt. Sinai J. Med.* 66:84-90. Protein components of the compositions of the invention may thus be replaced by nucleic acid (preferably DNA *e.g.* in the form of a plasmid) that encodes the protein.

25 DISEASE STATES

The compositions of the present invention may be used to prevent and/or treat disorders such as but not limited to: pneumonia, cardiovascular diseases, atherosclerosis, bronchitis, pharyngitis, laryngitis, sinusitis, obstructive lung diseases, asthma, chronic obstructive pulmonary disease, reactive arthritis, otitis media, abdominal aortic aneurysm, erythema nodosum, Reiter syndrome, sarcoidosis, Alzheimer's disease, multiple sclerosis, lymphogranuloma venereum, ocular trachoma, pelvic inflammatory disease, inclusion conjunctivitis, genital trachoma, infant pneumonitis, incipient trachoma, keratitis, papillary hypertrophy, corneal infiltration, vulvovaginitis, mucopurulent rhinitis, salpingitis, cervicitis, cervical follicles, prostatitis, proctitis, urethritis, lymphogranule inguinale, climatic bubo, tropical bubo, and/or esthiomene.

FORMULATIONS

40 *Chlamydial* infections affect various areas of the body and so the compositions of the invention may be prepared in various forms. For example, the compositions may be prepared as injectables, either as liquid solutions or suspensions. Solid forms suitable for solution in, or suspension in, liquid vehicles prior to injection can also be prepared (*e.g.* a lyophilised composition). The composition may be prepared for topical administration *e.g.* as an ointment, cream or powder. The composition may be prepared for oral administration *e.g.* as a tablet or capsule, as a spray, or as a syrup (optionally flavoured). The composition may be prepared for pulmonary administration *e.g.* as an inhaler, using a fine powder or a spray. The composition may be prepared as a suppository or pessary. The composition may be prepared for nasal, aural or ocular administration *e.g.* as drops. The composition may be in kit form, designed such that a combined composition is reconstituted just prior to

administration to a patient. Such kits may comprise one or more antigens in liquid form and one or more lyophilised antigens.

Further components of the composition

- 5 The composition of the invention will typically, in addition to the components mentioned above, comprise one or more 'pharmaceutically acceptable carriers', which include any carrier that does not itself induce the production of antibodies harmful to the individual receiving the composition. Suitable carriers are typically large, slowly metabolised macromolecules such as proteins, polysaccharides, polylactic acids, polyglycolic acids, polymeric amino acids, amino acid copolymers, and lipid aggregates (such as oil droplets or liposomes). Such carriers are well known to those of ordinary skill in the art. The vaccines may also contain diluents, such as water, saline, glycerol, *etc.* Additionally, auxiliary substances, such as wetting or emulsifying agents, pH buffering substances, and the like, may be present. A thorough discussion of pharmaceutically acceptable excipients is available in Gennaro (2000) *Remington: The Science and Practice of Pharmacy*. 20th ed., ISBN: 0683306472.

- 20 The biological molecules of the present invention be formulated into a pharmaceutical composition or an immunotherapeutic composition or a vaccine composition. Such formulations comprise biological molecules combined with a pharmaceutically acceptable carrier, such as sterile water or sterile isotonic saline. Such formulations may be prepared, packaged, or sold in a form suitable for bolus administration or for continuous administration. Injectable formulations may be prepared, packaged, or sold in unit dosage form, such as in ampoules or in multi-dose containers containing a preservative. Formulations include, but are not limited to, suspensions, solutions, emulsions in oily or aqueous vehicles, pastes, and implantable sustained-release or biodegradable formulations. Such formulations may further comprise one or more additional ingredients including, but not limited to, suspending, stabilizing, or dispersing agents. In one embodiment of a formulation for parenteral administration, the active ingredient is provided in dry (for eg, a powder or granules) form for reconstitution with a suitable vehicle (e. g., sterile pyrogen-free water) prior to parenteral administration of the reconstituted composition. The pharmaceutical compositions may be prepared, packaged, or sold in the form of a sterile injectable aqueous or oily suspension or solution. This suspension or solution may be formulated according to the known art, and may comprise, in addition to the active ingredient, additional ingredients such as the dispersing agents, wetting agents, or suspending agents described herein. Such sterile injectable formulations may be prepared using a non-toxic parenterally-acceptable diluent or solvent, such as water or 1,3-butane diol, for example. Other acceptable diluents and solvents include, but are not limited to, Ringer's solution, isotonic sodium chloride solution, and fixed oils such as synthetic mono-or di-glycerides. Other parentally-administrable formulations which are useful include those which comprise the active ingredient in microcrystalline form, in a liposomal preparation, or as a component of a biodegradable polymer systems. Compositions for sustained release or implantation may comprise pharmaceutically acceptable polymeric or hydrophobic materials such as an emulsion, an ion exchange resin, a sparingly soluble polymer, or a sparingly soluble salt.

KITS

- 50 Also included in the invention is a kit for enhancing a CMI response to the biological molecules of the present invention. Such a kit may comprise an antigenic composition

or nucleotide sequence encoding same. The kit may also include an adjuvant, preferably a genetic adjuvant is administered with or as part of the biological molecule and instructions for administering the biological molecule. Other preferred components of the kit include an applicator for administering the biological molecule.

- 5 As used herein, the term "applicator" refers to any device including but not limited to a hypodermic syringe, gene gun, particle acceleration device, nebulizer, dropper, bronchoscope, suppository, impregnated or coated vaginally-insertable material such as a tampon, douche preparation, solution for vaginal irrigation, retention enema preparation, suppository, or solution for rectal or colonic irrigation for applying the
10 NOI either systemically or mucosally or transdermally to the host subject.

- The invention also provides for a kit comprising comprising a combination of *Chlamydia pneumoniae* antigens. The combination of *Chlamydia pneumoniae* antigens may be one or more of the immunogenic compositions of the invention. The
15 kit may further include a second component comprising one or more of the following: instructions, syringe or other delivery device, adjuvant, or pharmaceutically acceptable formulating solution. The invention also provides a delivery device pre-filled with the immunogenic compositions of the invention.

20 EXAMPLES

- The following invention will now be further described only by way of example in which reference is made to the following Figures. The following examples are presented only to illustrate the present invention and to assist one of ordinary skill in making and using the same. The examples are not intended in any way to otherwise
25 limit the scope of the invention. Efforts have been made to ensure accuracy with respect to numbers used (*e.g.*, amounts, temperatures, etc.), but some experimental error and deviation should, of course, be allowed for.

- Figure 1A.** Assay of *in vitro* neutralization of *C.pneumoniae* infectivity for LLC-MK2
30 cells by polyclonal mouse antisera to recombinant *Chlamydial* proteins. Results are shown as reduction in the number of inclusions obtained when monolayers were infected with antiserum-treated infectious EBs, as compared to inclusion numbers given by untreated EBs. Percent reduction values are plotted against the reciprocal of the corresponding serum dilution. For each dilution inclusion counts were corrected for
35 background inhibition of infectivity observed with the corresponding dilution of the pre-immune serum. The figure shows results obtained with serial dilutions of antibodies raised against a 'neutralizing' antigen (♥), a 'non-neutralizing' FACS- positive antigen (v), and against the GST polypeptide, used in the fusion constructs, alone (σ).

- Figure 1B** shows serum titres giving 50% neutralization of infectivity for the 10
40 *C.pneumoniae* recombinant antigens described in the text. Each titer was assessed in 3 separate experiments (SEM values shown).

- Figure 2** shows immunoblot analysis of two dimensional electrophoretic maps of
45 *C.pneumoniae* EBs using the imune sera described in the text. Immunoblots were obtained from either of two EB gels (panels A and B at the top) covering different pH intervals, according to which of the two allowed the best detection of a given antigen. The arrows in the HtrA immunoblot show which of the signals had a corresponding stained spot in the gel (arrows in panel A) which was subjected to MALDI-TOF

identification. The two patterns in the HtrA blot are both suggestive of typical electrophoretic 'trains' composed of single charge variants of the same protein.

5 **Figure 3** shows mean numbers of *C.pneumoniae* IFU recovered from equivalent spleen samples from immunized and mock-immunized hamsters following a systemic challenge. Standard deviation values are shown above the bars. Antigens which induced significant protection are highlighted with an asterisk above the corresponding bar. All antigens were delivered in Freund's adjuvant. n.i. = non immunized controls

10 **Figure 4** shows flow cytometric analysis of splenocytes from DNA-immunized HLA-A2 transgenic and non transgenic mice. Groups of 4 mice were immunized 3 times i.m. with 50µg of plasmid DNA expressing *C. pneumoniae* Low Calcium Response Protein H. IFN-γ production from splenocytes was monitored following either a 6h
15 (ex-vivo) or a 6 day (restimulated) pulse with peptide CH-6 (10µg/ml). Equal numbers of gated live lymphocyte cells were acquired with a LSRII FACS System (Becton Dickinson) and percentages of IFN-γ producing CD8⁺ T cells were calculated using DIVA Software (Becton Dickinson).

20 **Figure 5** shows a flow cytometric analysis of splenocytes from transgenic and non transgenic mice infected with *C. pneumoniae* EBs. (A) HLA-A2 transgenic mice were intranasally infected twice with 5x10⁵ *C. pneumoniae* FB/96 EBs and splenocytes were stimulated for 6 days in the presence of relevant peptides before
25 determining IFN-γ production by CD8⁺ T cells as described in the legend of Figure 4. (B) HLA-A2 transgenic and non transgenic mice were infected together with the same EBs preparation and CD8⁺ T cells were subjected to FACS analysis as reported in (A).

30 **Table I** shows a summary of data and properties of the *C.pneumoniae* antigens described in the text. The neutralization titer is reported as the reciprocal of the antiserum dilution causing a 50% reduction in the number of inclusions in the *in vitro* infectivity assay. For the hamster model data the statistical significance of the results was evaluated by a two-tailed Student's t-test: significant data (p ≤ 0.05) are
35 highlighted with an asterisk. ND = not detected.

Table 2 shows results from hamster mouse model studies for hypothetical proteins.

40 **Table 3** shows expressed genes of CPn EB selected by microarray.

Table 4 shows *C. pneumoniae* selected peptides: protein sources and HLA-A2 stabilization assay.

45 **Table 5** shows ELISPOT assay with CD8⁺ T cells from DNA immunised HLA-A2 transgenic mice.

Table 6 shows IFN-γ production from splenocytes of DNA immunized HLA-A2 transgenic and non transgenic mice.

50 **METHODS AND MATERIALS (Examples 1-4) (see Reference Section 1)**

C.pneumoniae EB purification

C.pneumoniae FB/96, a clinical isolate obtained from a patient with pneumonia at the Sant'Orsola Polyclinic, Bologna, Italy, was grown in LLC-MK2 cells seeded in individual wells of a six-well plastic plate (7). Cells were harvested 72 hr after infection with a sterile rubber, sonically disrupted and the elementary bodies (EB) purified by gradient centrifugation as described (26). Purified *Chlamydiae* were resuspended in sucrose-phosphate-glutamic acid (SPG) transport buffer, and stored in 0.5 ml aliquots, at -80°C until used. When required, prior to storage, EB infectivity was heat-inactivated by 3 hour incubation at 56°C.

Expression and purification of recombinant proteins

Open reading frames (ORFs), selected from the *C. pneumoniae* CWL029 genome sequence (16), were PCR-cloned into plasmid expression vectors and purified from *E.coli* cultures, essentially as previously described (25). Recombinant *Chlamydial* proteins were obtained as GST fusion proteins by using pGEX-KG derived vectors (12) in *E. coli* BL21 (Novagen). PCR primers were designed so as to amplify genes without the N-terminal signal peptide coding sequence. When a signal peptide or processing site was not clearly predictable, the ORF sequence was cloned as annotated by Kalman and coworkers (16). Recombinant *E.coli* cells were grown in LB medium (500 ml), containing 100 µg/ml Ampicillin, and grown at 37°C until OD₆₀₀ = 0.5, and then induced with 1 mM IPTG. Cells were collected by centrifugation 3 hr after induction and broken in a French Press (SLM Aminco, Rochester, NY). After centrifugation at 30.000 g, the supernatants were loaded onto Glutathione Sepharose 4B columns (Amersham Pharmacia Biotech) and column bound proteins were eluted with 50 mM Tris-HCl, 10 mM reduced glutathione, pH 8.0. Protein concentrations in the samples were determined using the Bradford method.

Preparation of mouse antisera

Groups of four 5/6-week old CD1 female mice (Charles River, Como, Italy) were immunized intraperitoneally at day 1 with 20µg of protein in Complete Freund's adjuvant (CFA) and boosted at day 15 and 28 with 20µg of recombinant protein in Incomplete Freund's adjuvant (IFA). Pre-immune and immune sera were prepared from blood samples collected on days 0, 27 and 42. In order to reduce the amount of antibodies possibly elicited by contaminating *E. coli* antigens, the immune sera were incubated overnight at 4°C with nitrocellulose strips adsorbed with a total protein extract from *E. coli* BL21.

Flow cytometry assays

Analyses were performed essentially as previously described (25). Gradient purified, heat-inactivated EBs (2x10⁵ cells) from *C.pneumoniae* FB/9, resuspended in phosphate-saline buffer (PBS), 0.1% bovine serum albumin (BSA), were incubated for 30 min. at 4°C with the specific mouse antisera (standard dilution 1:400). After centrifugation and washing with 200 µl of PBS-0.1% BSA, the samples were incubated for 30 minutes at 4°C with Goat Anti-Mouse IgG, F(ab)'₂-specific, conjugated with R-Phycoerythrin (Jackson ImmunoResearch Laboratories Inc.). The samples were washed with PBS-0.1%BSA, resuspended in 150 µl of PBS-0.1%BSA and analysed by Flow Cytometry using a FACSCalibur apparatus (Becton Dickinson, Mountain View, CA). Control samples were similarly prepared. Positive control antibodies were: i), a commercial anti-*C.pneumoniae* specific monoclonal antibody

(Argene Biosoft, Varilhes, France) and, ii), a mouse polyclonal serum prepared by immunizing mice with gradient purified *C.pneumoniae* EBs. Background control sera were obtained from mice immunized with the purified GST peptide used in the fusion constructs (GST-fusions control). FACS data were analysed using the Cell Quest Software (Becton Dickinson, Mountain View, CA). The shift between the background control histogram and the immune serum testing histogram was taken as a measure of antibody binding to the EB cell surface. The Kolmorov-Smirnov (K-S) two-sample test (44) was performed on the two overlapped histograms. The D/s(n) values (an index of dissimilarity between the two curves) are reported as "K-S score" in Table 1.

2D Western Blot analysis of immune sera, and Mass Spectrometry

Gradient purified *C. pneumoniae* EBs were washed with 5 mM Tris-HCl pH 7.5, 0.1 mM EDTA, 10% glycerol, centrifuged 15 min. at 13 000 x g and pellets were resuspended in reswelling solution (7 M urea, 2 M thiourea, 2% (w/v) CHAPS, (2%w/v) ASB14, 2% (v/v) IPG buffer pH 3-10 NL, or pH 4-7, 2 mM TBP, 65 mM DTT). Protein samples (200 or 20 µg of protein for Coomassie Blue stained reference gels, or gels to be processed for immunoblotting, respectively) were adsorbed overnight on Immobiline DryStrips (7 cm, pH 3-10 NL, or pH 4-7). Electrofocusing was performed in an IPGphor Isoelectric Focusing Unit (Amersham Biosciences, Uppsala, Sweden). The focused strips were equilibrated as described (15) and loaded on linear 9-16.5 % acrylamide gradients (7x 4 cm, 1.5 mm thick), for SDS-PAGE separation in a Mini Protean III Cell (Bio-Rad, Hercules, CA). Gels were stained with colloidal Coomassie Blue (Novex, San Diego, CA) (4) and the protein maps so obtained were scanned with a Personal Densitometer SI (Molecular Dynamics) at 12 bits and 50 mm per pixel.

For Western Blot analyses, the proteins separated in the 2DE maps were transferred onto nitrocellulose membranes, overnight at 30 Volts, using a Protean III apparatus (BioRad, Hercules, CA). Membranes were stained with a 0.05% (w/v) CPTS (Copper(II) phthalocyanine-3,4',4'',4'''-tetrasulfonic acid tetrasodium salt) in 12 mM HCl, and marked peripherally with 8 India-ink dots to provide anchors for subsequent image superimposition and matching. After scanning and image acquisition, the membranes were destained with 0.5 M NaHCO₃, incubated with the mouse sera to be analyzed (either pre-immune or specific immune sera, diluted 1:1000), and then with a peroxidase-conjugated anti-mouse antibody (Amersham Biosciences, Uppsala, Sweden). After washing with PBS, 0.1% Tween-20, blots were developed using the Opti-4CN Substrate Kit (Biorad, Hercules, CA), and the images of the immunostained blots again acquired as above. Images were analysed with the computer program Image Master 2D Elite, version 4.01 (Amersham Biosciences, Uppsala, Sweden). Superimposition and matches between Western-blot membranes and Coomassie stained gels were performed as follow. First, the CPTS-stained membrane image and the immunostained blot image were superimposed using the peripheral dot marks. Then, the sum image so obtained was superimposed to the Coomassie stained protein map using the CPTS stained CPn proteins as anchors. The areas on the Coomassie stained map corresponding to immunostained spots on the blot were excised from the preparative gel for protein identification. Protein sample were dried in a vacuum centrifuge, and in-gel digested, for 2h at 37°C, with an excess of porcine Trypsin (Promega, Madison, WI), in 100 mM ammonium bicarbonate. Tryptic peptides were desalted and concentrated using Zip-Tip (Millipore, Bedford, MA). Peptides were directly eluted and loaded onto a SCOUT 384 Anchor Chip multiprobe plate (400 µm,

5 Bruker Daltonics, Bremen, Germany) with a solution of 2-5 dihydroxybenzoic acid (5g/l), in 50% acetonitrile, 0.1% trifluoroacetic acid. Spectra were acquired on a Bruker Biflex III matrix-assisted laser desorption ionization-time of flight (MALDI-TOF) apparatus. Resulting values for monoisotopic peaks were used for database searches using the Mascot software (32), as available at the website <http://www.matrixscience.com/>.

In vitro neutralization assays

10 *In vitro* neutralization assays were performed on LLC-MK2 (Rhesus monkey kidney) epithelial cell cultures. Serial four-fold dilutions of mouse immune and corresponding preimmune sera were prepared in sucrose-phosphate-glutamic acid buffer (SPG). Mouse polyclonal sera to whole EBs were used as positive control of neutralization, whereas SPG buffer alone was used as negative control of neutralization (control of infection). Purified infectious EBs from the *C.pneumoniae* FB/96 were diluted in 15 SPG buffer to contain 2.5×10^7 IFU/ml, and 10ul of EBs suspension were added to each serum dilution in a final volume of 100ul. Antibody-EB interaction was allowed to proceed for 30 min at 37°C on a slowly rocking platform. The 100ul of reaction mix of each sample was used to inoculate PBS-washed LLC-MK2 confluent monolayers (in triplicate for each serum dilution), in a 24-well tissue culture plate, 20 and centrifuged at 805 x g for 1 hour at 37°C. After centrifugation Eagle's minimal essential medium containing Earle's salts, 20% fetal bovine serum and 1ug/ml cycloheximide was added. Infected cultures were incubated at 37°C in 5%CO₂ for 72 hours. The monolayers were fixed with methanol and the *Chlamydial* inclusions were detected by staining with mouse anti-*Chlamydia* fluorescein-conjugated monoclonal 25 antibody (Merifluor *Chlamydia*, Meridian Diagnostics, Inc.) and quantified by counting 10 fields per well at a magnification of 40X. The inhibition of infectivity due to EBs interaction with the immune sera was calculated as percentage reduction in mean IFU number as compared to the SPG (buffer only)/EBs control. In this calculation the IFU counts obtained with immune sera were corrected for background 30 inhibition of infection due to the corresponding pre-immune mouse serum. According to common practice, the sera were considered as "neutralizing" if they could cause a 50% or greater reduction in infectivity. The corresponding neutralizing titer was defined as the serum dilution at which a 50% reduction of infectivity was observed. Experimental variability was evaluated by calculating the standard error of 35 measurement (SEM), from three titration experiments for each recombinant antigen, as shown in Fig. 1B.

In vivo screening

40 *In vivo* evaluation was performed using a hamster model of systemic infection, as recently described (34). Essentially, adult (10-11 week old) Syrian hamsters (Morini, S. Polo D'Enza, Italy), previously immunized with the recombinant vaccine candidates were challenged systemically with infectious Cpn elementary bodies (EB). Protection was assessed by determining the levels of viable EB recovered from the spleen, as compared to non-immunized animals. Statistical significance of the results 45 was evaluated by a two-tailed Student's t-test.

Groups of 8 hamsters were immunized subcutaneously with recombinant antigens, or only injected with buffer for the control group, at days 0, 7, and 21. For each immunization 20 ug protein 1:1 diluted with Freund's complete adjuvant (first dose) 50 and Freund's incomplete adjuvant (booster doses) was injected. At day 35 post-

infection the hamsters were anaesthetised with Ketamine and inoculated intraperitoneally and intranasally with 0.1 ml of *C.pneumoniae* EB suspension (1.0×10^8) at each site. Animals were sacrificed seven days after infection. The spleen was weighed, and homogenized in a mortar to obtain a 10% (wt/vol) suspension in cold SPG buffer. Tissue suspensions were centrifuged at $300 \times g$ for 10 min at 4°C to remove coarse debris. The clarified homogenates (0.2 ml) were inoculated in duplicate onto LLC-MK2 cells seeded in plastic individual well of a 24 well plate, incubated at 37°C for 72 h and fixed in acetone before detection and counting of numbers of *Chlamydial* inclusions per well by immunofluorescence microscopy. The protocol was approved by the ethical committee of the University of Bologna.

Example 1 (in vitro studies)

Screening antisera for in vitro neutralizing properties

Following a genome-wide screening for proteins likely to be localized on the cell surface of *C. pneumoniae*, we recently reported (25) that antisera to 53 recombinant *Chlamydial* antigens were capable to bind in a FACS assay, the surface of *Chlamydial* cells. In order to check whether some of the FACS positive antigens were capable of interfering with EB *in vitro* infectivity, we raised mouse antisera against the recombinant FACS positive antigens and assessed the effect of each antiserum on the infectivity of purified EBs with respect to monolayers of LLC-MK2 cells. Infectious EB were first incubated with the antiserum and then used to infect cell monolayers in 24-well multititer plates. In parallel, control samples were similarly processed in which the EBs were: i), either treated with buffer only, or, ii), treated with the same dilutions of the corresponding preimmune mouse sera.

Results I

Using this assay, 10 sera have so far proved to effectively neutralize *in vitro* infectivity to an extent greater than 50%, a property that common practice qualifies such antigens as “neutralising” (Figure 1). These 10 sera were obtained by mouse immunization with recombinant proteins derived from the following *C.pneumoniae* genes:

- *pmp10* and *pmp2*, encoding two members of the heterogeneous *Chlamydial* PMP family of polymorphic membrane proteins;
- *artJ*, encoding a putative extracellular solute (possibly Arginine) binding protein of an aminoacid transport system;
- *eno*, encoding a protein homologous to bacterial enolases, glycolytic enzymes which can be found also on the bacterial surface;
- *htrA*, encoding a putative chaperone with heat-shock inducible protease activity;
- the Cpn0301 “hypothetical” gene, encoding a protein homologous to the ompH family of bacterial proteins, some members of which have been shown to be chaperones involved in outer membrane biosynthesis;
- two Cpn-specific “hypothetical” genes Cpn0795 and Cpn0042;
- *omcA* encoding a 7-9 kDa protein annotated as an outer membrane protein; and
- *atoS* a putative sensor member of a transport system.

As shown in Figure 1 and summarized in Table I, OmpH, enolase and Cpn0795 appeared to induce the highest neutralizing sera, with titers above 400. By contrast, Pmp2, ArtJ and Cpn0042 induced titers equal or less than 100, while the remaining 4 antigens, Pmp10, HtrA, AtoS and OmcA showed intermediate titers.

Example 2 (in vivo studies)**Evaluation of antisera specificity by 2D immunoblot analysis of Cpn protein extracts**

5 In order to investigate if the neutralizing activity observed in the *in vitro* infection of LLC-MK2 monolayers was actually due to the binding of the antibodies to the selected *C.pneumoniae* proteins, rather than to possible cross-reactions with other antigens, we assessed the specificity of the antisera by immunoblot analysis of two dimensional electrophoretic maps of EB proteins.

10

In particular, this analysis was carried out on six antigens (Pmp2, Pmp10, Eno, ArtJ, HtrA and OmpH-like) known to be visible in the 2D maps of EB total proteins (Montigiani *et al.*, 2002 Infection and Immunity 70: 368-379). Total EB proteins were resolved by 2D-electrophoresis using two different pH intervals (pH 3-10 non linear, and pH 4-7, respectively) since previous experiments had shown that some of the proteins under study were better detected using one rather than the other of the above pH intervals. For each pH interval four gels were run in parallel. One gel was stained with Coomassie Blue to visualize the protein spots, while the other gels were blotted on nitrocellulose filters and stained with one of the selected sera at 1000-fold dilution. Subsequently, the images of the immunostained blots (Fig.2, panels c to h) were superimposed to the corresponding Coomassie Blue-stained gel to identify the spots which had reacted with a given antiserum. The matching protein spots were excised and processed for peptide identification by MALDI-TOF analysis.

Results 2

25 In all six maps the immunoreactive protein species in the excised gel area were found to contain peptides from the expected *Chlamydial* protein. Even when the serum reacted with more than one electrophoretic protein species, the mass spectra of all spots which could be detected in the Coomassie Blue stains 2DE map were always consistent with the same polypeptide being present as multiple electrophoretic species.

Interestingly, the immunoblot obtained with the HtrA antiserum showed two sets of 4 spots arranged as two typical electrophoretic 'trains' at two different molecular weights. On the Coomassie Blue stained gel it was possible to identify 4 corresponding spots, 3 in the upper train and 1 in the lower Mw set. MS analysis identified all of them as products of Cpn HtrA gene. Interestingly the lower Mwt species missed 3 N-terminal tryptic peptides, detected in the higher Mw spot series, and mapping within the first 100 aa of the ORF. These results suggest that HtrA was present in the EB protein sample both as a full *htrA* product, and as a discrete species missing a short N-terminal peptide, possibly as a result of some post-translational processing.

Discussion of Results 2

45 In the analysis of data which are based on polyclonal antibody reactivity one should consider that cross-reactions due to epitope mimickry are always difficult to exclude. The problem of antisera specificity was addressed in this work by 2D immunoblotting and identification of the reacting electrophoretic species by mass spectrometry analysis. This approach was possible for 6 of the 10 antisera, i.e. those corresponding to proteins previously identified by mass spectrometry (MALDI-TOF) analysis on 2D electrophoretic maps of *C.pneumoniae* EB proteins (25, 42) (Table 1, and Figure 2).

50

The probability of fortuitous cross-reactions between unrelated *Chlamydial* protein species was minimized by the results of the immunoblotting analyses which showed that out of ca 300 protein spots in a map, all those reacting with the tested antisera were consistent with the expected antiserum specificity. Obviously, since during 2-D electrophoresis conformational epitopes are generally lost, structure-dependent cross-reactions cannot be ruled out in this type of analysis.

Example 3

In vivo evaluation of the in vitro neutralizing antigens in a hamster model of systemic infection

We have recently described a new hamster model of systemic *Chlamydia pneumoniae* infection in which replicating *Chlamydia* disseminate through macrophages and accumulate in the spleen (34). We therefore asked the question whether the *in vitro* neutralizing antigens we identified would also have protective activity *in vivo* using this model. To this aim, the 10 *in vitro* neutralizing recombinant antigens were used to immunize 8 hamsters with 3 subcutaneous injections over a three-week period, and challenged with 2×10^8 Cpn EBs two weeks later. Spleen infection was assessed 7 days after challenge. The difference between the mean number of infectious *Chlamydiae* recovered from control animals and the mean number of *Chlamydiae* recovered from animals immunized with the recombinant *Chlamydial* antigens, was taken as a measure of protection specifically induced by the putative vaccine candidate.

Results 3

The results of spleen protection observed for the various antigens in repeated experiments are shown in Figure 3 and reported as percentage values in Table 1. Six out of ten antigens, Pmp2, Pmp10, Enolase, the OmpH-like protein, and the products of the *C.pneumoniae*-specific genes Cpn0759 and Cpn0042, showed a statistically significant protective activity, with a reduction in IFU recovered from the spleens of immunized animals higher than 80% with respect to mock-immunized controls.

A limit of the hamster model is that, because of the absence of immunological reagents, the relative contribution of humoral and cell-mediated immunity cannot be assessed. However, we asked the question whether recombinant antigens could elicit also in the hamster neutralizing antibodies with sufficiently high titers. Therefore we tested the sera from hamsters immunized with Pmp2 and enolase, two of the most protective antigens, in the *in vitro* neutralization assay. Both antigens had a neutralizing titer of approximately 100 (data not shown).

Summary of Results 3

In conclusion, a considerable proportion (60%) of the *in vitro* neutralizing antigens were also protective in the hamster *in vivo* model and our data suggest that antibody-mediated neutralization could play a role at least in this model of systemic infection.

Discussion of Results 3

Beside assaying the *in vitro* neutralization properties of the selected subset of 10 FACS-positive antigens, we also assessed the performance of these antigens in protecting against *C. pneumoniae* infection in an animal model of systemic infection recently described in the hamster (34). This evaluation addressed the capability that the recombinant antigens would have of inducing a protective response against naturally replicating *Chlamydiae* (rather than EB's purified from *in vitro* cultures

grown under artificial conditions) and in the context of a systemic infection. In fact the hamster model we used, while it does not model the typical respiratory infection considered to be the predominant route by which *C. pneumoniae* infects humans, it nevertheless simulates a situation of systemic invasion which could be preliminary to the establishment of *C. pneumoniae* chronic infection considered by several authors as being associated to the development or progression of cardiovascular disease, and other chronic degenerative diseases. Notably, a limit of any hamster model is the current lack of hamster-specific immunological reagents which would allow the analysis of cell mediated immune responses. However, in the case of systemic infections, by common wisdom, neutralizing antibodies are likely to have a protective action. The finding that 6 of the 10 '*in vitro* neutralizing' antigens had also a >80% protective action *in vivo*, and that a measurable neutralizing activity was also found in the sera of immunized hamsters, suggests that a specific antibody mediated immunity could at least partially contribute to the animal protection here described.

Example 4

Two 'hypothetical' proteins 6784 and 6814 (encoded by the ORFs Cpn0498 and Cpn0525) yielded FACS-positive sera which, however, were not able to neutralize host cell infection *in vitro*. However, these antigens performed remarkably well in the hamster-spleen test.

Table 2

Gene/ORF ID in CWL029	Protein ID	Recombination Fusion Type	Annotation	Reciprocal of 50% neutralisation titre	% Protection in the hamster spleen test (ref 34)
Cpn0498	4376784	GST	Hypothetical protein	0	94
CPn0525	4376814	GST	Hypothetical protein (similarity to CT398)	0	97
CPn0324		HIS	Low Calcium Response Element (LcrE)		Completely protected 8 of 16 animals and reduced the infectivity titres of the eight positive animals

Discussion of Results 4

Interestingly, whilst antiserum against CPn0525 gave negative *in vitro* results (ie no neutralising activity), the CPn0525 protein gave 97 per cent protection from spleen infection in an *in vivo* hamster immunisation assay (ie a positive *in vivo* result).

- 5 Likewise, whilst antiserum against Cpn0498 gave negative *in vitro* results (ie no neutralising activity), the CPn0498 protein gave 94 per cent protection from spleen infection in an *in vivo* hamster immunisation assay. Thus a positive signal obtained in the FACS assay does not guarantee a corresponding positive *in vitro* neutralization activity and conversely a negative neutralization activity does not mean that a positive
- 10 *in vivo* result can be excluded.

General Discussion of Results 1-4*Strategy for identification of Chlamydia pneumoniae antigens of interest*

- Our strategy was based on the following experimental steps: 1) analysis of
- 15 *Chlamydia* genome sequence to select putative membrane-associated antigens, 2) cloning, expression and purification selected antigens, 3) preparation of antigen-specific sera by mouse immunization with the purified antigens, 4) FACS analysis of *Chlamydia* EBs using the mouse sera to identified surface-exposed antigens, 5) “*in vitro* neutralization” assay to test whether antibodies elicited by a given antigen can
- 20 interfere with the process of eukaryotic cell infection, and 6) use of appropriate animal model to test the capacity of selected antigens to confer protection against *Chlamydia* challenge.

- As recently described by Montigiani *et al* ((2002) Infection and Immunity 70: 368-379) from the initial screening of the *C.pneumoniae* genome, a panel of mouse sera was prepared against over 170 recombinant His-tagged or GST-fusion proteins encoded by genes or “open reading frames” somehow predicted to be peripherally located in the *Chlamydial* cell. When these antibodies were tested in a FACS assay for their ability to bind the surface of purified *C.pneumoniae* EBs, a list of 53 “FACS-positive” sera was obtained. The corresponding putative surface antigens were then
- 30 further assessed for their capability of inducing neutralizing antibodies. This part of the work involved testing which of the sera contained antibodies capable of interfering with the process of *in vitro* infection of epithelial cell cultures. In the *in vitro* “neutralization” assay purified infectious EBs are incubated with progressive
- 35 dilutions of the immune sera and, in parallel, dilutions of the corresponding pre-immune sera, and of sera against non *Chlamydia* control antigens.

- Cell cultures are infected in the presence of cycloheximide, which inhibits host cell protein synthesis and favours *Chlamydial* intracellular growth with the consequent
- 40 formation of typical cytoplasmic inclusions which can be stained with *Chlamydia* specific fluorescence-labeled monoclonal antibodies and counted with an UV light microscope. Working with appropriate pathogen-to-host cell ratios, it can be reasonably assumed that the number of detected cytoplasmic inclusion is proportional to the number of infectious *Chlamydiae* in the original sample. So a reduction in
- 45 inclusion numbers caused by the presence of an antigen-specific antiserum, as compared to the numbers obtained with control sera, gives a measure of the capability of a given antigen to elicit antibodies which can inhibit some stage of the *Chlamydial* infection process. According to common convention, an anti-serum is labelled as ‘neutralizing’ when the reduction of infectivity is equal or greater than 50%, and the

serum dilution yielding a 50% reduction in infectivity is referred to as the 50% end-point neutralization titer.

5 Some of the results obtained by screening the panel of recombinant antigens with the *C.pneumoniae* *in vitro* neutralization assay confirm that some of the listed antigens, like the members of the family of heterogeneous polymorphic membrane proteins (PMP), which, on the basis of published literature data, could be reasonably expected to be surface-exposed and possibly induce neutralizing antibodies. However, there are also proteins which could be considered so far only hypothetical, and proteins
10 which just on the basis of their current functional annotation could not be at all expected to be found on the bacterial surface. Using an *in vitro* neutralising assay, it was found that sera to 10 CPn antigens have so far proved to effectively neutralize *in vitro* infectivity to an extent greater than 50%, a property that common practice qualifies such antigens as “neutralising” (Figure 1). These 10 sera were obtained by
15 mouse immunization with recombinant proteins derived from the *C.pneumoniae* genes listed below.

Using a recently described *in vivo* model of systemic infection (hamster model), hamsters immunised with 6 of the *in vitro* neutralising antigens, when challenged with
20 CPn EBs, showed a greater than 80% reduction of spleen infection as compared with non-immunised controls.

Characterisation of 10 CPn proteins

The proteins identified by the present work can be divided in 3 groups:

- 25 • proteins which have an annotation compatible with (could be reasonably expected to have) an expected/predicted exposure on the *Chlamydial* cell surface and with the possibility that antibodies binding to them may actually interfere with host cell attachment and entry (ie proteins which could possibly induce neutralising antibodies)
- 30 • proteins which by homology with other gram-negative bacteria could be expected to have a periplasmic exposure (ie would not be expected at all to be found on the bacterial cell surface); and
- proteins which are still labelled as ‘hypothetical’ (ie cellular location and/or cellular function not known)

Group 1

(Pmp proteins (pmp2 and pmp10), OmcA and OmpH)

Pmp proteins (pmp2 and pmp10)

The first group includes the 2 polymorphic outer membrane proteins (Pmp’s) Pmp2 and Pmp10 (10, 11, 14, 30), the outer membrane protein OmpH-like, and OmcA, which is annotated (*Chlamydia* Genome Project at <http://Chlamydia-www.berkeley.edu:4231/>) as “predicted 9-kD cysteine-rich, outer membrane protein, lipoprotein”. The Pmp family of *Chlamydia*-specific proteins is generally thought to comprise probable pathogenicity factors, with an autonomous secretion capacity (autotransporters), important for adhesion to host cells and are generally considered as
40 promising vaccine candidates. However, apart from very recent unpublished results on Pmp21, this is the first time that antisera to recombinant Pmp’s are reported to have neutralizing properties.

OmcA

OmcA is the product of a gene co-transcribed in the same operon with the 60 kDa OmcB cystein-rich protein which is a major structural component of the *Chlamydial* outer membrane and a major immunogen in human *C. trachomatis* infections. OmcB and OmcA are likely to interact in some as yet unknown outer membrane structure, so it is possible that antibodies to OmcA can interfere with EB infectivity.

OmpH

Finally, the *Chlamydial* OmpH is probably a member of the OmpH (Skp) family of proteins which have been reported to have chaperonin activities in other bacteria very important for the correct biosynthesis of the outer membrane. These proteins appear to cooperate in this task with HtrA (see below). In fact, in *E.coli* single KO mutants of either OmpH (Skp) or HtrA (DegP) are still viable, but double mutants do not grow (37). It should be pointed out that even if the amino acid sequences of the ompH-like proteins of *Chlamydia* (all *C.pneumoniae* and *C.trachomatis* or *C.caviae* variants) line-up very well with the rest of the bacterial OmpH proteins, they are the only ones to be acidic, whereas the rest of the family comprises mostly very basic proteins (including some with histone like behaviour, at least *in vitro*). One could speculate that if the chaperone activity is maintained also in the ompH like *Chlamydial* proteins, this may be related to some *Chlamydial* peculiarity.

Second Group of Selected Proteins***(ArtJ, AtoS, HtrA and Enolase)***

The second group, which represents a somehow surprising finding, includes ArtJ, AtoS, HtrA and Enolase. If the current annotation (justified by analogy with homologous genes in other bacteria) is correct, all these proteins would be expected to have a periplasmic location in gram-negative bacteria, and to be surface-exposed only in a gram-positive bacterium. It is possible that owing to their atypical life cycle, requiring an efficient passage from a dormant spore-like status (the EB) to an active form needing to adapt quickly to host-cell responses to invasion, *Chlamydiae* in fact display some sensors directly on the outer surface of their infectious form.

ArtJ

In the case ArtJ – for which we have data supporting both antigen expression and serum specificity – the hypothesis of an atypical situation peculiar to *Chlamydia* is supported by the anomalous gene set-up resulting from the present analysis of the *Chlamydia* genomes. ArtJ is so annotated by analogy with the ART transport systems of *E.coli* which has 5 genes organized in two operons (24) : artPIQM and artJ which are responsible for the arginine transport. In Cpn however the artPIQM genes are absent and therefore it appears that *Chlamydial* ArtJ operates in a molecular context which is different from its *E.coli* model and must be peculiar to this species.

HtrA

HtrA (DegP), which in other bacteria has a complex hexameric structure, has been described as having multiple functions (3, 5, 18, 19, 27, 38) : a chaperonin assisting a correct outer membrane biogenesis, inducible protease for the elimination of misfolded membrane proteins, and also a sensor of 'stress' conditions. In *Chlamydia* none of these properties has been demonstrated yet, however we find that in purified EB HtrA is present in two forms one of which appears to be processed by being deprived of the N-terminal fragment. This fragment, if aligned with the homologous

HtrA sequence from *Thermoloba maritima* (18), would comprise a predicted loop acting as a structural lid controlling the access to the protease active. So it appears tempting to speculate that HtrA could have a similar protease activity and the two forms identified on the 2-D map represent the active and inactive species. Interestingly, the *C. trachomatis* HtrA ortholog is recognized by human sera from patients who had a *Chlamydial* genital infection (35), and a similarly HtrA is one of the antigens in the immunoproteome of *Helicobacter pylori* (13). Furthermore the homologue protein in *Haemophilus influenzae* is a protective antigen in both a passive infant rat model of bacteremia and the active chinchilla model of otitis media (23).

Enolase

Also in the second group of proteins expected to be located elsewhere than the cell surface, is Cpn enolase. This protein aligns with the well known family of conserved glycosylases, which are essentially cytoplasmic enzymes, but in *Streptococci* enolase has been shown to have also a cell surface location, and extracellular matrix binding properties (1, 28, 29)). Interestingly, Gaston and colleagues (8) also showed that in patients with reactive arthritis induced by *C. trachomatis*, enolase induces specific CD4⁺ T-cell responses. Furthermore, a clone responding to the enolase *C. trachomatis* ortholog, responded also to *C. pneumoniae* EBs, and, since no proliferative response could be observed using a fungal or a mammalian enolase, the authors of this study concluded that the CD4 T-cell stimulating epitope must be *Chlamydia* specific.

Third Group of Proteins

(unknown cellular location and/or cellular function, Cpn0795, CPn0042)

The third of the 3 groups in which we propose to divide, just for the sake of discussion, the 10 neutralizing antigens above described, comprises two proteins which are still annotated in public *Chlamydial* databases as the hypothetical products of two CPn-specific genes: Cpn0759 and Cpn0042. The Cpn0759 gene is the second gene in a cluster of 6 Cpn-specific hypothetical genes (from Cpn0794 to Cpn0799) immediately upstream of the enolase gene. With the exception of Cpn0759 the products of all the other genes in the cluster share similarities of 30 to 40% over long stretches of amino acids. The Cpn0042 gene encodes a hypothetical protein, with 4 coiled-coil regions, which has been described as a member of a new family of hypervariable outer membrane proteins (33). Interestingly, the hypervariability of these proteins could be due to a strand-slippage mechanism induced by the presence of a poly(C) stretch within the coding region of the corresponding genes, a mechanism already described in the Pmp's family for the *pmp10* gene (30). However, as indicated by their annotation, the function of these proteins is still unknown, and our observations provide the first experimental indication of a possible function related to the *Chlamydial* infection process.

Table 1 of this application demonstrates that Cpn0795 (SEQ ID NO: 6) a Cpn specific hypothetical protein is a FACS positive protein which demonstrates significant immunoprotective activity in a hamster spleen model of *Chlamydia pneumoniae* infection. We have found evidence to demonstrate that other Cpn proteins in this group of Cpn specific hypothetical proteins have now been found to have a secreted autotransporter function. These proteins, which are absent from *Chlamydia trachomatis* include: gi/4377105 (Cpn0794), gi/4377106 (Cpn0795), gi/4377107 (Cpn0796), gi/4377108 (Cpn0797), gi/4377109 (CPn0798), gi/4377110 (Cpn0799).

Fig. 6 shows an alignment of the proteins in the 7105-7110 protein family. This Alignment shows a new family of proteins expected to constitute a system of antigens probably delivered on the Cpn surface or secreted by a type V (autotransporter) secretion mechanism. This alignment was generated as follows:

5

Imperfect repeats were identified which allowed the alignment of the genes. Molecular modelling has also demonstrated that the C-terminal ends of 7106 and 7107 can be predicted to fold in a beta-barrel structure which can form a translocation pore for secretion across the outer membrane.

10

Cpn0794 = 7105 = FACS positive

Cpn0795 = 7106 = FACS positive

Cpn0796 = 7107 = FACS positive

Cpn0797 = 7108 = FACS positive

Cpn0798 = 7109 = No data available

15

Cpn0799 = 7110 = No data available

(Reference for FACS positive data = Montigiani et al (2002) Infect Immun 70(1) 368-79)

Operon1 = 0794, 0795, 0796 Operon2 = 0797, 0798

20

Cpn0795 and Cpn0796 have C terminal ends that may form transmembrane pores (see alignment, FIG. 9). CPn0794, Cpn0797, Cpn0798, and Cpn0799 have N-terminal ends indicating that all proteins have N-terminal and C-terminal ends.

Fig. 7 shows alignment of Cpn0794 – Cpn 0799. Proteins encoded by the genes Cpn0794, Cpn0795, Cpn0796, and Cpn0797 have been identified as likely to be exposed on the surface of the chlamydia cell and as possible vaccine candidates. These proteins are shown to be actually expressed by Cpn in vivo (WB data and FACS data). In the case of Cpn0797 we also showed that the level of expression in CPn EBs is high enough to be detected by mass spectrometry analysis on 2DE maps of protein extracts (see Montigiani et al.)

30

Following these observations, it is seen that the proteins encoded by Cpn0794, Cpn0796 and Cpn0797 proteins can be aligned according to a set of imperfect repeats present within their aminoacid sequences (see FIG. 7) , whereas the putative product of CPn0795 can be mostly aligned to the C-terminal portion of the Cpn0796 protein.

35

Furthermore, proteins encoded by genes Cpn0798 or Cpn0799 can also be aligned to the above proteins according to the above mentioned repeated sequence motifs (see FIG. 7).

Overall alignment of the 6 genes demonstrates that the genes encode for a family of functionally-related proteins.

Furthermore, in silico analysis of the protein encoded by Cpn0796, which encompasses the entire alignment of all the proteins in this family demonstrates that a functional precursor with the aminoacid sequence reported below:

45

SEQ ID NO: 80

50 MKFMKVLTPWIYRKDLWVTAFLLTAIPGSFAHTLVDIAGEPRHAAQATGVSGDGKIVIGMKVPDDPFA
ITVGFQYIDGHLQPLEAVRPQCSVYPNGITPDGTVIVGTNYAIGMGSAVKWVNGKVSELPMLPDTLD

SVASAVSADGRVIGGNRNINLGASVAVKWEDDVITQLPSLPDAMNACVNGISSDGSIIIVGTMVDVSWR
 NTAVQWIGDQLSVIGTLGGTTSVASAISTDGTIVVGGSENADSQTHAYAYKNGVMSDIGTLGGFYSLA
 HAVSSDGSVIVGVSTNSEHRYHAFQYADGQMVDLGTGGPESYAQGVSGDGKIVIGRAQVPSGDWHAF
 LCPFQAPSPAPVHGGSTVVTSONPRGMVDINATYSSLKNSQQQLQRLLIQHSAKVESVSSGAPSFTSV
 5 KGAIKQSPAVQNDVQKGTFLSYRSQVHGNVQNQQLLTGAFMDWKLASAPKCGFKVALHYGSQDALVE
 RAALPYTEQGLGSSVLSGFGGQVQGRYDFNLGETVVLQPFMGIQVLHLSREGYSEKNVRFPVSYDSVA
 YSAATSFMGHVFASLSPKMSTAATLGVERDLNSHIDEFKGSVSAMGNFVLENSTVSVLRPFASLAMY
 YDVRQQQLVTLVVMNQQLTGTLSLVSQSSYNLSF

10 Processing sites that assist in the secretion of the polypeptide from the cytoplasm and
 its release into the periplasm are located after aminoacid 31 (based on PSORT
 prediction and/or after aminoacid 47 similar to experimentally determined processing
 sites in other bacterial autotransporter molecules (e.g. BrkA from B.pertussis). Hence,
 15 the mature form of the Cpn0796 product is as follows:

SEQ ID NO: 81

HTLVDIAGEPRHAAQATGVSGDGKIVIGMKVPDDPFAITVGFQYIDGHLQPLEAVRPQCSVYPNGITP
 D
 20 GTVIVGTNYAIGMGSVAVKWVNGKVSELPMLPDTLDSVASAVSADGRVIGGNRNINLGASVAVKWEDD
 VITQLPSLPDAMNACVNGISSDGSIIIVGTMVDVSWRNTAVQWIGDQLSVIGTLGGTTSVASAISTDGT
 VIVGGSENADSQTHAYAYKNGVMSDIGTLGGFYSLAHAVSSDGSVIVGVSTNSEHRYHAFQYADGQMV
 DLGTGGPESYAQGVSGDGKIVIGRAQVPSGDWHAF LCPFQAPSPAPVHGGSTVVTSONPRGMVDINA
 25 TYSSLKNSQQQLQRLLIQHSAKVESVSSGAPSFTSVKGAIKQSPAVQNDVQKGTFLSYRSQVHGNVQ
 NQQLLTGAFMDWKLASAPKCGFKVALHYGSQDALVERAALPYTEQGLGSSVLSGFGGQVQGRYDFNLG
 ETVVLQPFMGIQVLHLSREGYSEKNVRFPVSYDSVAYSAAATSFMGHVFASLSPKMSTAATLGVERDL
 NSHIDEFKGSVSAMGNFVLENSTVSVLRPFASLAMYDVRQQQLVTLVVMNQQLTGTLSLVSQSSY
 NLSF

30 Or

SEQ ID NO: 82

TGVSGDGKIVIGMKVPDDPFAITVGFQYIDGHLQPLEAVRPQCSVYPNGITPDGTVIVGTNYAIG
 35 MGSVAVKWVNGKVSELPMLPDTLDSVASAVSADGRVIGGNRNINLGASVAVKWEDDVITQLPSLP
 DAMNACVNGISSDGSIIIVGTMVDVSWRNTAVQWIGDQLSVIGTLGGTTSVASAISTDGTIVVGG
 ENADSQTHAYAYKNGVMSDIGTLGGFYSLAHAVSSDGSVIVGVSTNSEHRYHAFQYADGQMVDLG
 TLGGPESYAQGVSGDGKIVIGRAQVPSGDWHAF LCPFQAPSPAPVHGGSTVVTSONPRGMVDINA
 40 TYSSLKNSQQQLQRLLIQHSAKVESVSSGAPSFTSVKGAIKQSPAVQNDVQKGTFLSYRSQVHG
 NVQNQQLLTGAFMDWKLASAPKCGFKVALHYGSQDALVERAALPYTEQGLGSSVLSGFGGQVQGR
 YDFNLGETVVLQPFMGIQVLHLSREGYSEKNVRFPVSYDSVAYSAAATSFMGHVFASLSPKMSTA
 ATLGVERDLNSHIDEFKGSVSAMGNFVLENSTVSVLRPFASLAMYDVRQQQLVTLVVMNQQLT
 GTLSLVSQSSYNLSF

45 In silico analysis of the protein encoded by Cpn0796 also demonstrates a C-terminal
 domain comprising approximately residues from 1 to 648. FIG. 8 illustrates Cpn0796.
 As shown in FIG. 8, Cpn0796 forms a beta-barrel structure and is capable of forming
 a pore across the bacterial outer membrane (OM). As is typical of 'autotransporter'
 molecules, after being secreted across the bacterial inner membrane into the periplasm
 50 through an N-terminal signal peptide mechanism, the molecule may form a pore in the
 OM through which the N-terminal domain may pass (the 'passenger' domain) to the
 outside of the bacterial cell. Also, these molecules may either remain anchored to the
 bacterial surface or undergo a proteolytic cut which releases the 'passenger domain'

or a portion of it into the medium surrounding the bacterial cell an example of which is represented in the following sequence:

SEQ ID NO: 83

5 MKFMKVLTPWIYRKDLWVTAFLLTAIPGSFAHTLVDIAGEPRHAAQATGV
 SGDGIKIVIGMKVPDDPFAITVGFQYIDGHLQPLEAVRPQCSVYPNGITPD
 GTVIVGTNYAIGMGSAVKWVNGKVSELPMLPDTLDSVASAVSADGRVIG
 GNRNINLGASVAVKWEDDVITQLPSLPDAMNACVNGISSDGSIIIVGTMVD
 10 VSWRNTAVQWIGDQLSVIGTLGGTTSVASAISTDGTIVIVGGSENADSQTH
 AYAYKNGVMSDIGTLGGFYSLAHAVSSDGSVIVGVSTNSEHRYHAFQYAD
 GQMVDLGTLLGGPESYAQGVSGDGKIVIGRAQVPSGDWHAFLCPFQAPSPA
 PVHGGSTVVTSONPRGMVDINATYSSLKNSQQQLQ

15 RLLIQHSAKVESVSSGAPSFTSVKGAIKQSPAVQNDVQKGTFLSYRSQVHGNVQNQQLLTGAFM
DWKLASAPKCGFKVALHYGSQDALVERAALPYTEQGLGSSVLSGFGGQVQ
GRYDFNLGETVVLQPFMGIQVLHLSREGYSEKNVRFPVSYDSVAYSATS
FMGAHVFAVSLSPKMSTAATLGVERDLNSHIDEFKGSVSAMGNFVLENSTV
SVLRPFASLAMYDVRQQQLVTLVVMNQQLTGTLSLVSQSSYNLSF

20 Also shown in FIG. 8, amino acid residues 365-385 represent an alpha helix conformation that spans the beta barrel pore

25 The N-terminal passenger domain may be cleaved via a specific proteolytic action from the membrane-anchored pore structure. A linker domain comprising the peptide sequence PSPAPV (**SEQ ID NO: 84**) as shown in bold in the following sequence illustrates a site at which cleavage of the N-terminal passenger domain may occur:

SEQ ID NO: 85

30 HTLVDIAGEPRHAAQATGVSGDGKIVIGMKVPDDPFAITVGFQYIDGHLQPLEAVRPQCSVYPNG
 ITPDGTIVIVGTNYAIGMGSAVKWVNGKVSELPMLPDTLDSVASAVSADGRVIGGNRNINLGASV
 AVKWEDDVITQLPSLPDAMNACVNGISSDGSIIIVGTMVDVSWRNTAVQWIGDQLSVIGTLGGTTS
 VASAISTDGTIVIVGGSENADSQTHAYAYKNGVMSDIGTLGGFYSLAHAVSSDGSVIVGVSTNSEH
 35 RYHAFQYADGQMVDLGTLLGGPESYAQGVSGDGKIVIGRAQVPSGDWHAFLCPFQ**PSPAPV**HGGS
 TVVTSONPRGMVDINATYSSLKNSQQQLQRLLIQHSAKVESVSSGAPSFTSVKGAIKQSPAVQN
 DVQKGTFLSYRSQVHGNVQNQQLLTGAFMDWKLASAPKCGFKVALHYGSQDALVERAALPYTEQG
 LGSSVLSGFGGQVQGRYDFNLGETVVLQPFMGIQVLHLSREGYSEKNVRFPVSYDSVAYSATSF
 MGAHVFAVSLSPKMSTAATLGVERDLNSHIDEFKGSVSAMGNFVLENSTVSVLRPFASLAMYDVR
 40 QQQLVTLVVMNQQLTGTLSLVSQSSYNLSF

45 The N-terminal peptide may be secreted to be exposed on the bacterial cell surface and can also become detached via the proteolytic event described above. The peptide may form a structural conformation known as beta-propellers indicated in the following sequence:

SEQ ID NO: 86

50 HTLVDIAGEPRHAAQATGVSGDGKIVIGMKVPDDPFAITVGFQYIDGHLQPLEAVRPQCSVYPNG
 ITPDGTIVIVGTNYAIGMGSAVKWVNGKVSELPMLPDTLDSVASAVSADGRVIGGNRNINLGASV
 AVKWEDDVITQLPSLPDAMNACVNGISSDGSIIIVGTMVDVSWRNTAVQWIGDQLSVIGTLGGTTS
 VASAISTDGTIVIVGGSENADSQTHAYAYKNGVMSDIGTLGGFYSLAHAVSSDGSVIVGVSTNSEH
 RYHAFQYADGQMVDLGTLLGGPESYAQGVSGDGKIVIGRAQVPSGDWHAFLC

Furthermore, the N-terminal passenger domain can also possess a specific protease activity, such as a serine protease-like activity. In addition to acting on a variety of substrates, the protease activity may act on the membrane anchored form of the molecule such that the N-terminal passenger domain is cleaved off from the surface of the chlamydial cell. The serine protease like activity is supported by the presence of a consensus serine protease triad of adequately spaced amino acid residues (namely H, D and S) which can be located on the virtual structure of the 'passenger' domain modelled on a set of experimentally-determined templates, e.g. 1nr0 (PDB identification code)

Based on the above analysis, the gene Cpn0796 gene encodes for a protein which promotes its own secretion on the EB surface and may also mediate or promote its own release into the surrounding medium. The secreted passenger peptide has several activities, including:

1. actin binding peptide, part of a chlamydial surface layer, and instrumental to the process of establishing the host cell infection
2. specific protease activity within the host cell cytoplasm instrumental to the intracellular survival of infecting chlamydiae.
3. specific activity within the host cell cytoplasm to down regulate expression of selected genes, either by repressing their transcription and/or by repressing their translation (m-RNA degradation)
4. cooperation with the products of genes Cpn0794, 0795, 0797, 0798, 0799
5. another function of the above N-terminal beta propeller domain is the regulation/ modulation of the activity of a cytosolic protease of the host cell in order to alter host cell properties in favour of chlamydial development, survival or persistence. See Fulop V, Bocskei Z, Polgar L. in "Prolyl oligopeptidase: an unusual beta-propeller domain regulates proteolysis." Cell. 1998 Jul 24;94(2):161-70.

The proteins encoded by Cpn0794, Cpn0797, Cpn0798, Cpn0799 - all comprising variants of the above described Cpn0796 structure - also provide beta propeller structures with activities similar and/or complementary to the ones described above.

Thus, a family of proteins cooperating to a common function either by generating - through events of site specific recombination - new molecules with structures and activities similar to the above described Cpn0796 product, OR by independently contributing to a multi-protein structure requiring a coordinated action of several related components.

FIG. 9 illustrates an alignment of the C-terminal domains of the proteins encoded by C.pneumoniae genes Cpn0795 and Cpn0796. As seen in FIG. 9, beta barrel domains of Cpn0795 or Cpn0796 include MKDLGTLGG (SEQ ID NO: 87), SXDGK (SEQ ID NO: 88) VIVG (SEQ ID NO: 89), VIXG (SEQ ID NO: 90) or HAF (SEQ ID NO: 91).

Fourth Group of Proteins

Cpn0498

So in this case the triple parallel-screening evaluation, with two positive and one negative result, brought to the identification of a previously unknown antigen (ie an

antigen with unknown biological function) having, according to current views, just the desirable basic properties in terms of antigenic function of a vaccine candidate. Further characterization of Cpn antigen data is included in *Finco et al.*, "Identification of New Potential Vaccine Candidates Against *Chlamydia pneumoniae* by Multiple Screenings," Vaccine, 23 (2005) 1178-1188, incorporated herein in its entirety.

Example 5

Background

The main stages in the *Chlamydial* life cycle are:

- 10 (i) the binding to the host cell surface and entry into the cytoplasm through a specialised vacuole (the *Chlamydial* inclusion) by an extracellular spore-like infective form, called the elementary body (EB); and
- (ii) the conversion of the EB to a non-infective replicative form called a reticulate body (RB) that replicates by binary fission a number of times
- 15 within the inclusion to form a microcolony.

The sets of genes which are expressed in the various stages of the *Chlamydial* life cycle and the signals that trigger the passage from one stage to the next are largely unknown and still need investigation.

20 Protein microarrays are used for high throughput protein analysis by detecting proteins and monitoring their expression levels. Through use of protein microarrays, complex screening of thousands of proteins and interactions with proteins may be performed in parallel. A protein array typically includes a surface, such as glass, membrane, microtiter wells, mass spectrometer plates, beads or other particles, for

25 binding ligands, proteins, or antibodies. For example, antibodies may be bound to the microarray to form a capture array. The capture array may be contacted with a biological sample to quantify the proteins in the biological sample. Also, proteins may be bound to the microarray and contacted with a biological sample to quantify protein-protein or protein-ligand interactions. Thus, protein microarrays may also be

30 used in diagnostics in which multiple immunoassays may be conducted in parallel such that levels of proteins in different samples may be quantified and compared for applications in the treatment or diagnosis of disease.

For example, in a capture array, antibodies are bound to the microarray and exposed

35 to a biological sample. Proteins and ligands that bind to the antibody array may be detected by direct labelling of the bound proteins. If a higher sensitivity or specificity is desired, a sandwich technique may be employed in which pairs of antibodies are directed to the same protein ligand. This technique is particularly useful if the amount of protein to be detected is low or if there are modifications to the protein. In addition,

40 the use of sandwich assays minimizes the risk of cross-reactivity in highly multiplexed assays by providing dual level target recognition, i.e. two levels of specificity for each locus in the array. Alternatively, the bound proteins may be detected via label-free detection methods such as including mass spectrometry, surface plasmon resonance and atomic force microscopy. This technique is useful if

45 modification or alteration of the protein is to be avoided.

Also, Large-scale functional chips containing large numbers of immobilized purified proteins may be used to assay a wide range of biochemical functions, such as protein interactions with other proteins, drug-target interactions, enzyme-substrates, etc. Such

50 proteins may be purified from an expression library, for example, and the protein

array can be used to screen libraries to select specific binding partners, including antibodies, synthetic scaffolds, peptides and aptamers. In this way, 'library against library' screening can be carried out, such as screening of drug candidates in combinatorial chemical libraries against an array of protein targets identified from genome projects.

Protein microarray technology permits analysis of the proteins themselves rather than inferring protein function, interactions and characteristics through mRNA expression. In many cases, mRNA expression does not correlate accurately with protein abundance. Furthermore, mRNA expression analysis does not provide sufficient information on protein-protein interaction or post-translational modifications. Thus, direct analysis of proteins via protein microarrays provides an advantage by providing more accurate information of proteins and protein-protein interactions that may not be readily available through measurement of mRNA expression.

Current DNA microarray techniques permit profiling of gene expression at the mRNA level as a function of the cellular state. This can lead to the identification of genes or clusters of genes whose up- or down-regulation is associated to a particular state of the cell and to the identification of therapeutically relevant targets. Using this technology, DNA fragments representing specific portions of all genes belonging to a given organism (the fragments can be derived from cDNA libraries or can be obtained by PCR amplification and chemical synthesis) are chemically bound to the surfaces of solid supports (chips) at high densities and in an ordered manner. Currently up to 10,000 DNA fragments or 250,000 oligonucleotides can be spotted onto a single squared centimetre of chip surface. The DNA chips are then utilised to define which of the spotted genes are transcriptionally active in a particular cellular population. To do so, RNA is prepared, labelled with fluorescent dyes and finally hybridised to the DNA fragments fixed to the surface of the chip. By using an appropriate computer-assisted fluorescence detector, the fluorescence signals emitted by each spot upon excitation with a laser beam is carefully quantified to define the transcription activity of all the arrayed genes.

CPn DNA microarrays have been developed to look at the transcriptional events which occur when a given CPn pathogen gets into contact with the host cells, both in vivo and in vitro settings. DNA chips carrying the entire genome of a particular bacterium, such as the CPn bacterium can be prepared in a very short period of time so that whole genome expression analysis can be determined.

Experimental Methodology
Specifically, a genomic DNA (open reading frame probes) microarray approach for gene expression in CPn bacteria was adopted. In this respect, an array was prepared for the analysis of the CPn life cycle on the basis of the published annotation of the complete genome. The Chlamydia DNA chips carry about 1000 PCR-derived DNA fragments, which have an average size of 400-700bp and correspond to internal portions of all CPn annotated genes.

Results 5

Table 3(i)-(xi) shows transcriptional activity for expressed genes for CPn EB selected by microarray. The data in Tables 3(i)-(iv) shows different mRNAs in order of abundance from cells in their infectious "spore-like" (EB) form. Data in Tables 3(v)-

(xi) correlates and summarizes mRNA expression levels of genes for CPn. The cells were used at the end of their cycle where gene expression is likely to be at its highest. As values less than approximately 10000 is likely to be background, the top set of proteins (approx top 30) with more intense signals are likely to be the most interesting proteins.

A review of the values for the expressed genes indicates that three of the FACS positive CPn antigens (CPn0331 (hypothetical), CPn0234 (hypothetical) and CPn0572 (hypothetical) are all highly expressed genes.

Table 3(v)-(xi) shows the transcriptional activity for expressed genes for CPn EB selected by microarray. The Table shows different mRNA in order of abundance from cells in their infectious "spore-like" (EB) form. The cells were used at the end of the cycle where gene expression is likely to be at its highest. A review of Table 3(i)-(iv) and (v)-(xi) indicates that CPn antigens CPn0558 (OmcA), CPn0331 (hypothetical), CPn0539 (Pmp19), CPn0234 (Hypothetical) and CPn0572 (Hypothetical) are all relatively highly expressed genes.

Where possible, attempts were made to place the transcriptional activities disclosed in Table 3(v)-(xi) in the context of the Chlamydia developmental cycle. In this respect, Chlamydia late gene products have been described more frequently than early gene products. This is primarily because of the presence of late gene products in EBs but not RBs and that it is easier to study EBs rather than RBs.

In addition, late gene functions appear to be predominantly those associated with the terminal differentiation of RBs back to EBs (Shaw et al., Mol Microbiology 37(4), 2000, 913-925). Late gene products appear to function in the termination of bacterial cell division and constitute structural components and remodelling activities involved in the formation of the cross-linked outer membrane complex that functions in the attachment and invasion of new host cells. By way of example, an important aspect of the secondary differentiation process (RB to infectious EB) is the expression of genes that encode proteins that form the highly disulfide cross-linked bacterial outer membrane (OM) complex. It is thought that several late cycle genes encode proteins with potential roles in the formation and maturation of the OM complex, a crucial step in the development of infectious EBs (see Belland et al., PNAS (USA) 100(14), 2003, 8478-83). The late genes omcA and omcB encode two cysteine-rich OM proteins that interact with the major OM protein (OmpA) to form this complex. A key protein component of the OM complex, the OmcB protein, has been found to undergo post-translational proteolytic processing. We have found that OmcB and OmcA show high levels of transcriptional activity (see top of Table 3(ii)). Cpn 0384 whose CT equivalent is CT046 (hctB) has been shown to be associated with differentiation from RB to EB (see Belland et al., PNAS (USA) 100(14), 2003, 8478-83). We also found Cpn0384 to have relatively high levels of transcriptional activity (again see top of Table 3(v)-(xi)). Other Cpn antigens thought to be involved in the Type III secretion system were found to have moderate expression levels in terms of transcriptional activity. This finding appears to be in line with published commentary where it is thought that while transcription of the two putative structural components of the Type III secretion system (yscJ and yscN (Cpn669)) begins at mid-cycle, export of effector molecules may be at a different stage of the developmental cycle.

Table 3(v)-(xi) indicates that high transcriptional activity was observed for Cpn0539 (CT412) which corresponds with a 98Kda protein known either as PmpA or Pmp19. Even though the Pmp19 protein demonstrates relatively "high" levels of transcriptional activity, this result is interesting because mRNA abundance for pmp19 does not seem to correlate with protein abundance. In this respect, results from our laboratory have shown that (i) Pmp19 was not detected in either 2D maps, Western Blots or FACS analysis studies which suggests that the pmp19 protein either is not surface exposed even though high levels of mRNA are expressed or that (ii) Pmp19 protein is expressed but processed or degraded by proteolytic digestion rendering it undetectable by immunoblot analysis. The results in our laboratory are confirmed by others. In this respect, Grimwood et al (2001) *Infection and Immunity* 69(4) 2383-2389 have shown that transcriptional profiles were detected for each of the *Chlamydia pneumoniae* 21 Pmp genes demonstrating that all pmp genes are transcribed during infection. Since each of the Pmp genes was transcribed, Grimwood et al (2001) evaluated protein expression by immunoblotting of *Chlamydia pneumoniae* CWL029 EB lysates using peptide-specific antisera. Interestingly, no Pmp-specific reactivity was detected for sera from either PmpA (Pmp19) or PmpB/C and PmpD gene by immunoblot analysis regardless of high anti-peptide reactivity. This result suggested that these proteins either are not stable or not translated. These findings demonstrate that there appears to be a variation in Pmp expression for the *Chlamydia pneumoniae* family of 21 polymorphic membrane proteins (Pmps) which are predicted to be localised to the bacterial outer membrane. The function of Chlamydial Pmps remains unknown, although based on sequence prediction and experimental testing, these Pmps are regarded as surface proteins and thus, likely to be critical for Chlamydial virulence. Like the Inclusion (Inc) Membrane proteins, the Pmp proteins are regarded, at present, as unique to the Chlamydiae family (see Rockey et al (2000) *Infection and Immunity* 69(10) 5473-5479). The findings disclosed here and by others, such as Grimwood et al, demonstrates that the *Chlamydia* organism appears to expend a considerable metabolic cost in Pmp transcription, such as Pmp19 transcription, despite the potential lack of production of a functional Pmp proteins, such as the Pmp19 protein.

Materials and Methods (Examples 6-8) (Reference Section II)

T cell Epitope prediction and peptide synthesis

T cell epitope prediction was carried out on the genomic sequence of *C. pneumoniae* CWL029 strain (Accession numbers NC_000922 or AE001363) using the BIMAS algorithm [24]. Synthetic peptides (purity > 80%) were synthesized by Primm Srl (Milan, Italy), suspended in 100% DMSO and kept at -20° C before use.

RMA-S/A2 cell line and HLA-A2 transgenic and non transgenic mice

The T cell lymphoma murine cell line RMA-S stably transfected with HLA-A2 (RMA-S/A2, H-2^b, TAP2⁻), was kindly provided by Dr. Barnaba, Università degli Studi "La Sapienza", Rome, Italy, and cultured at 37° C in RPMI-1640 (GIBCO) supplemented with heat inactivated 10% FCS, 100 IU/ml penicillin/streptomycin, 2 mM L-glutamine (GIBCO) and 5×10⁻⁵ M 2-ME (Sigma). H2-b HLA-A2 transgenic mice [35] were housed in a pathogen-free environment and screened for HLA-A2 expression by FCM carried out on total blood samples using the BB7.2 anti-A2 mAb [48]. Only mice with percentages of A2 expressing cells higher than 70-80 % were

used for DNA immunization and *C. pneumoniae* infection experiments. Animals which showed no HLA-A2 expression were mated in order to obtain an HLA-A2 non transgenic population, to be used as a control in the experiments.

5 **Epitope stabilization assay**

RMA-S/A2 cells ($3-5 \times 10^5$ /well) were seeded in serum-free RPMI medium, supplemented with human $\beta 2$ microglobulin (3 μ g/ml, Sigma), without or with the test peptide (10 μ M). Following overnight incubation at 26°C in humidified 5% CO₂ atmosphere, cells were shifted to 37° C for 2 h before determining the HLA-A2 expression level at the cell surface using the BB7.2 anti-A2 mAb and a PE-conjugated anti-mouse IgG (Jackson ImmunoResearch). Fluorescence intensity on living cells, which did not incorporate propidium iodide, was analyzed by FCM. As controls, corresponding samples without peptide and samples with peptide but treated only with the anti-mouse secondary antibody, were used.

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Infection and DNA immunization of HLA-A2 transgenic and non transgenic mice

Transgenic mice were intranasally infected twice with a month interval, using 5×10^5 *C. pneumoniae* FB/96 EBs [4] diluted in 50 μ l of PBS. *C. pneumoniae* antigen coding genes were amplified by PCR using FB/96 genomic DNA, cloned into plasmid pcmvKaSF2120 [49] and verified by DNA sequence analysis. Fifty μ g of endotoxin free recombinant plasmid DNA, diluted in Dulbecco's phosphate buffer (GIBCO), were injected into the tibialis muscle of mice at days 0, 21 and 35.

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25 **CD8⁺ T cells isolation and IFN- γ determination by ELISpot assay**

Splenocytes from DNA immunized mice were prepared one week after the third immunization using Cell Strainer (Falcon) filters. Following red blood cells lysis, CD8⁺ T cells from spleen cells suspensions were enriched by positive selection using magnetic activated cell sorting (MACS-Miltenyi Biotec) with CD8a (Ly-2) microbeads. CD8⁺ T cells purity was higher than 90%, as determined by FCM. Multiscreen 96-well nitrocellulose plates (Millipore) were coated with 5 μ g/ml of the anti-mouse IFN- γ antibody (R4-6A2, PharMingen) in 100 μ l of carbonate buffer, pH 9.2. After overnight incubation at 4°C, plates were saturated at 37°C with 200 μ l of BSA (1%) in PBS for 2 h. Purified CD8⁺ (5×10^4) were added in a total volume of 200 μ l/well in the presence of irradiated (3,000 rad) spleen cells from non immunized HLA-A2 transgenic mice as a source of antigen-presenting cells (2×10^5 /well), 10 μ g/ml of peptide and 10U/ml of human r-IL-2 (Chiron Corporation). After incubation for 20 h at 37° C, 5% CO₂, plates were washed and developed for bound IFN- γ using sequential incubations with biotinylated antimouse IFN- γ (XMB1.2, PharMingen), ExtrAvidin-alkaline phosphatase and substrate BCIP/NBT (Sigma) dissolved in water. Spots were enumerated in each well using a dissecting microscope. Medium containing an irrelevant peptide or without peptide was used as negative controls, while positive controls were represented by CD8⁺ T cells treated with ConA (5 μ g/ml).

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In vitro cultures and flow cytometric analysis of splenocytes from transgenic and non transgenic mice infected with *C. pneumoniae*

Splenocytes from infected mice were isolated one week after the second infection with *C. pneumoniae* Ebs. For *ex vivo* analysis of IFN- γ production, 2×10^6 splenocytes

were seeded in the presence of the test peptide (10 µg/ml) and anti-mouse CD28 antibody (1 µg/ml, PharMingen) as co-stimulus. After a two h incubation at 37° C, 5 % CO₂, Brefeldin A (10 µg/ml, Sigma) was added and the incubation was extended for 4 additional hours. Following two washes with PBS, cells were permeabilized, fixed and stained both with anti-murine-IFN-γ-(PE), anti-murine CD8 (APC) and anti-murine-CD69 (FITC) and the corresponding isotypes. Positive controls for cytokine production were performed on cells stimulated with anti-mouse CD3 and CD28 antibodies (1 µg/ml, PharMingen). Cells cultured either in the absence of peptide or pulsed with the HepB negative control peptide were used as negative controls. All samples were analyzed using a FACS LSRII flow cytometer (Becton Dickinson). For analysis of IFN-γ production by short term T cell lines, 5-10x10⁶ splenocytes from infected mice were cultured for 6 days in the presence of the test peptide (20 µg/ml), with rIL-2 (10 µg/ml) being added after the first two days. At the end of the incubation period, cells were washed twice in RPMI, pulsed again for 6 h in the presence of the test peptide (10 µg/ml), 1x10⁵ freshly prepared CD8 depleted antigen presenting cells from HLA-A2 transgenic mice (irradiated at 3000 rad) and anti-mouse CD28 antibody (1 µg/ml, PharMingen) as co-stimulus. After a two h incubation at 37° C, 5 % CO₂, Brefeldin A (10 µg/ml, Sigma) was added, the incubation was extended for 4 additional hours and IFN-γ production was analyzed by FCM.

Example 6

In silico analysis of *Chlamydia pneumoniae* genome and prediction of HLA-A2 T cell epitopes

The genome of the *Chlamydia pneumoniae* CWL029 strain was used to predict 9mer peptide sequences with high probability to bind class I HLA-A2 molecules. The analysis was carried out using the predictive algorithm available at the NIH Bioinformatics & Molecular Analysis Section Web server (<http://bimas.cit.nih.gov/>), which ranks potential MHC binders according to the predictive half-time dissociation of peptide/MHC complexes [24]. Since some *Chlamydial* proteins have been reported to induce autoimmune responses [25-28], we restricted our search to a subset of proteins, distinctive of the *Chlamydia* genus, consisting of 13 protein identified as members of the type III secretion system, 17 Polymorphic Membrane Proteins (PMP) and 19 additional proteins, 5 of which already identified as EB surface antigens [4]. The predicted binding score of 157.22, obtained for the well characterized HIV-1 p17 gag epitope ⁷⁷SLYNTVATL⁸⁵ [29], was taken as an arbitrary cut-off for peptide selection. A total of 55 potential *C. pneumoniae*-derived T cell epitopes, belonging to 31 different proteins, were identified (Table I), which had predicted binding scores ranging from 156.77 to 42,485.263

In vitro binding of peptides to HLA-A2

The capacity of the selected peptides to bind to HLA-A2 was assessed using an *in vitro* MHC class I stabilization assay, carried out with the murine transporter associated with antigen processing (TAP)-deficient cell line RMA-S/A2, stably transfected with the human class I A2 gene. MHC class I molecules, cultured at 37° C, are unstably expressed on the cell surface of TAP-deficient cells [30-32]. Culturing the cells at 37° C in the presence of binding peptides, results in formation of a more stable MHC/peptide complex which can be monitored by flow cytometric analysis. RMA-S/A2 cells were therefore cultured overnight at 26° C in the presence of the

test peptides, shifted to 37° C for 2 hours and the surface level of stabilized A2 molecules was quantified by direct staining with an anti-HLA-A2 specific mAb. Two known HLA-A2 restricted CTL epitopes were used as positive controls for binding to A2, the HIV-1 p17 gag peptide [29] and the influenza matrix M1 protein peptide FluMP58 [33], while the Hepatitis B virus envelope antigen peptide HbenvAg125 (HepB) was used as a negative control [34].

Results 6

The binding results obtained are shown in Table 4 and allowed the identification of 15 peptides with a net mean fluorescence intensity (Net MFI) higher than 92.3, corresponding to the value obtained with the HIV-1 p17 gag positive control peptide, 8 peptides with a Net MFI intermediate between the values 92.3 and 63.1, obtained with the two positive control peptides, and 12 peptides with a Net MFI ranging between 29.6 and 63. Fifteen of the *in silico* predicted peptides (27.2 %) did not confer stabilization to the A2 molecules, showing a Net MFI lower than 14, obtained with the HepB negative control peptide.

Example 7

Some HLA-A2 binders are recognized by CD8⁺ T cells from DNA-immunized transgenic mice

The *in vitro* assay with RMA-S/A2 cells allowed the definition of a set of peptides which were able to bind and stabilize the HLA-A2 molecules on the cell surface. To gain information about the possibility that the predicted epitopes could indeed be generated by *in vivo* processing of the antigens from which they were derived, peptide recognition by CD8⁺ T cells was studied under conditions in which the related complete antigen was intracellularly expressed and presented *in vivo*. The full-length ORF sequences coding for 13 *Chlamydial* proteins, including a total of 24 predicted peptides, were cloned into a suitable DNA expression vector and each recombinant plasmids was used to immunize distinct groups of transgenic mice expressing a chimeric class I molecule composed of the α 1 and α 2 domains of HLA-0201 and the α 3 domains, transmembrane and cytoplasmic, of H-2K^b [35].

The ORF sequences were selected among those containing either one or more epitopes positive in the *in vitro* assay or a combination of positive and negative epitopes. The ORF sequence corresponding to the outer membrane protein A (OMPA, CPn 0695) was included in this analysis, since human MHC-I-restricted epitopes have already been reported for this protein in *C. trachomatis* [18;36]. One coding sequence, related to gene CPn 0131 was chosen, which included four epitopes, all negative in the *in vitro* stabilization assay. After three immunization cycles, transgenic mice were sacrificed, spleen CD8⁺ T cells were isolated, stimulated for 20 hour with the corresponding peptide and *ex vivo* IFN- γ production was assessed using an enzyme-linked immunospot (ELISpot) assay.

Results 7

DNA-mediated expression of the ORFs including peptides CH-6 (CPn 0811), CH-7 (CPn 0623), CH-10 (CPn 0828), CH-13 (CPn 0695, OMPA) and CH-37 (CPn 0210) were associated with numbers of spot forming cells (SFC) significantly higher than those obtained with the HepB unrelated peptide, whereas some peptides related to antigens coded by genes CPn 0131, CPn 0323 and CPn 0062 induced SFC values only 2-3 times higher than the HepB control peptide (Table 5). Peptides related to

antigens coded by genes CPn 0132, CPn 0322, CPn 0325, CPn 0415 and CPn 0728 did not induce any IFN- γ production (data not shown).

Example 8

5 To test the capacity of peptides to amplify specific CD8⁺ T cell populations *in vitro*, some of these plasmids were used to repeat the DNA immunization experiment and to determine by flow cytometry the intracellular IFN- γ production by CD8⁺ T cells, both *ex vivo* and after a 6 day stimulation in the presence of the relevant peptides. In the attempt to establish a direct correlation between IFN- γ production by CD8⁺ T cells and HLA-A2 specific restriction, the experiment was carried out with both transgenic and non transgenic syngenic mice. The plasmids used contained genes CPn 0695, CPn 0811 and CPn 0823, including peptides CH-13, CH-6 and CH-7 respectively, which were highly positive in the *in vitro* binding and in the ELISpot assays and gene CPn 0323, including six different peptides, all of them with ELISpot values slightly higher than background

Results 8

20 The results of the experiment are summarized in Table 6, while representative dot plots from flow cytometric analysis of splenocytes stimulated with peptide CH-6 are shown in Fig. 4. When fresh spleen cells of DNA-immunized transgenic mice were pulsed with the tested peptides, only CH-6 or CH-7 induced relative fold increase (RFI) values about 5 times higher than those obtained pulsing the same cells with the HepB negative control peptide (Table 6, 4.58 and 5.2 RFI respectively).

25 When short term T cell lines (TCLs) instead of fresh splenocytes were used, a larger panel of peptides were able to trigger a significantly higher IFN- γ production by CD8⁺ T cells (Table 6). In fact, in addition to peptides CH-6 and CH-7, also peptides CH-13, CH-44, CH-45 and CH-46 were recognized by CD8⁺ T cell populations significantly larger than those induced by pulsing the same cells with the HepB peptide (RFI > 5). Importantly, peptide-induced IFN- γ production, appeared to be largely HLA-A2-dependent, since when the same experiments were carried out with non transgenic mice, the RFI values obtained were reliably lower (Table 6). The fact that non transgenic and transgenic spleen cells were both efficiently activated using the polyclonal stimulus (anti-CD3/anti-CD28), reinforced the hypothesis that the lower CD8⁺ T cells induction in non transgenic mice was due to the absence of specific interactions between the peptides and the human HLA-A2 molecules.

CD8⁺ T cells of transgenic mice infected with *C. pneumoniae* recognize HLA-A2 binders *in vivo*

40 It has been recently shown that infection of mice with *C. pneumoniae* elicits a pathogen-specific murine class I-restricted immune response [22]. Therefore, we asked whether any of the A2 *in vitro* binders could be recognized by specific CD8⁺ T cells that are clonally selected during the immune response raised against the corresponding native antigen in *C. pneumoniae* infected cells.

45 To address this issue, HLA-A2 transgenic mice were intranasally infected with a non lethal dose of *C. pneumoniae* EBs and challenged with an equal dose of bacteria one month later, before being sacrificed to obtain splenocytes that were used to measure IFN- γ production by CD8⁺ T cells. Since no appreciable IFN- γ -production could be observed if splenocytes from infected mice were tested directly *ex vivo* (data not

shown), spleen cells were cultured with each individual peptide or with the HepB irrelevant peptide for 6 days. The resulting short-term TCLs were then pulsed again for 6 hours with the same peptides and intracellular IFN- γ production by CD8⁺ T cells was assessed. The results obtained with 40 tested peptides are shown in Fig. 5A.

5 Sixteen peptides (CH-2, CH-7, CH-8, CH-10, CH-13, CH-15, CH-20, CH-21, CH-28, CH-35, CH-37, CH-45, CH-46, CH-47, CH-50 and CH-55) elicited the strongest CD8⁺ responses (1 to 7.1 % of IFN- γ -producing CD8⁺ T cells), while 19 peptides elicited low but consistent responses (percentages of CD8⁺/IFN- γ ⁺ T cells between 0.3 and 0.9). Five peptides did not induce percentages of IFN- γ -producing CD8⁺ T

10 cells significantly higher than those observed in response to the HepB control peptide.

When eight among the most reactive peptides were used again to pulse splenocytes of both transgenic and non transgenic mice infected with *C. pneumoniae*, seven of them were recognized by specific CD8⁺/IFN- γ ⁺ T cell populations present only in the

15 transgenic mice, while peptide CH-7 was recognized by CD8⁺ T cells present in both mice groups (Fig. 5B).

General Discussion of Results in Examples 6-8

In this work we have described peptides derived from *C. pneumoniae* antigens identified as putative T cell epitopes recognized by the common human class I MHC A2 haplotype.

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Understanding *C. pneumoniae*-specific CD8⁺ T cell-mediated immune response and designing protective vaccines rely on the possibility of identifying bacterial antigens or epitopes recognized by CD8⁺ T cells. Whereas the induction of a CTL-dependent immune response is predictable in response to pathogens which replicate in the cellular cytosol, providing antigens which can enter the cellular MHC-I presentation pathway, in the case of *Chlamydiae* it is not immediately obvious which antigens are made available to the proteasome and how they reach the cytosol, since these bacteria

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30 have a stringent intravacuolar localization inside the infected cell.

We have chosen an *in vivo* system based on HLA-A2 transgenic mice to test which of the predicted peptides could be recognized by specific CD8⁺ T cells following either DNA immunization with individual antigen coding genes or infection with *C. pneumoniae*. Our choice of a murine model is supported by previously published data. Wizel *et al.* [22], recently reported the first evidence that CD8⁺ T cells specific for *C. pneumoniae* antigens are induced in infected mice, and identified bacterial-derived murine MHC-I-restricted T cell epitopes able to trigger either lysis of *C. pneumoniae* infected cells or *in vitro* inhibition of the pathogen intracellular growth. These

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40 findings seem to confirm that some *C. pneumoniae* antigens can indeed reach the cytosol of infected cells and enter the MHC-I presentation pathway, i.e. during remodeling that occurs during *Chlamydia* replication or following autolysis of developing bacterial particles [22].

Furthermore, Kuon *et al.* [42] recently reported the identification of 11 *C. trachomatis*-derived HLA-B27-restricted peptides, capable of stimulating CD8⁺ T cells obtained from patients with *Chlamydia*-induced reactive arthritis. Importantly, 8 of them overlapped those selected by analyzing splenocytes of HLA-B27 transgenic mice infected with *C. trachomatis*, indicating that antigen processing can be closely

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reproduced using the murine animal model, although differences between murine and human antigen processing and T cell repertoires have been hypothesized [43].

5 The experiment which we have performed with *C. pneumoniae* infected A2 transgenic mice revealed that at least 16 peptides were recognized by IFN- γ -positive CD8⁺ T cell populations, which were actually expanded as a consequence of bacterial infection, since we could not detect IFN- γ production pulsing spleen cells from non infected transgenic mice with the same peptides (data not shown). These results suggest that the corresponding *Chlamydial* antigen may be able to enter the MHC-I presentation
10 pathway. The finding that a number of these peptides can also be recognized by specific CD8⁺ T cells when the corresponding protein is separately expressed by DNA immunization, strongly reinforces the hypothesis that the responses observed with the infected mice are indeed specific for the *in silico* predicted peptide epitopes and their corresponding antigens. Importantly, the comparisons of peptide-induced IFN- γ -
15 positive CD8⁺ T cells in A2 transgenic and non transgenic mice, either immunized with DNA or infected with *C. pneumoniae*, indicate that this recognition event is largely A2-specific.

Though, we cannot rule out the possibility that some of the tested peptides are also able to interact with the murine class-I MHC molecules, as suggested by the result
20 obtained with CH-7 in infected non transgenic mice (Fig. 5) and by the RFI values obtained with CH-7, CH-8 and CH-13 in DNA-immunized non transgenic mice (Table 6).

Both with DNA immunization and bacterial infection, we were able to show that the
25 OMPA-derived CH-13 peptide induces a specific CD8⁺ T cell response in A2 transgenic mice. These results appear to validate the choice of this animal model, since our observation that OMPA can enter the MHC-I presentation pathway correlates with the previous identification of HLA-A2-restricted and of murine MHC-I-restricted epitopes in OMPA proteins of *C. trachomatis* [18] and of *C.*
30 *pneumoniae* [23] respectively. With the exception of CH-13 and CH-17, all the other peptides recognized by CD8⁺ T cells of infected mice belong to *C. pneumoniae* antigens for which neither human nor murine T cell epitopes have been identified [22;23]. Interestingly, a couple of positively reacting peptides (CH-50 and CH-55) belong to the group of polymorphic outer membrane proteins [44;45], while most of
35 the others are part of the group of Type III secretion system-related proteins [45;46]. Peptides CH-7 and CH-8, both included in protein T of the *Yersinia* outer protein (Yop) system [47] and CH-10, included in protein J, which is part of the same translocation system, appear to be particularly reactive in the assay with the infected mice (Fig. 5A).

40 This is also true for other peptides included in antigens which are again related to the type III secretion system, such as CH-45, CH-46, and CH-47, all present in the low calcium response protein D. Intriguingly, CH-8, which is the most reactive peptide in the assay with the infected mice, does not seem to be recognized by a specific T cell
45 population when the corresponding antigen is expressed by DNA immunization (Tables 5 and 6). This may be due to different factors, i.e. low *in vivo* expression level of the injected DNA or altered protein conformation.

On the other hand, we should also consider the possibility that, following infection of
50 mice with *C. pneumoniae*, this peptide is recognized by a CD8⁺ T cell population

which is instead specific for an epitope derived from an unidentified *C. pneumoniae* antigen having a closely related sequence. Contrarily to CH-8, stimulation of spleen cells from infected transgenic mice with peptide CH-6 did not allow the detection of IFN- γ^+ /CD8 $^+$ T cells (Fig. 5A), but the same peptide was clearly reactive in the DNA immunization experiments (Tables 5 and 6). This may suggest that Low Calcium Response Protein H is not available for the cellular proteasome, but we could also assume either that the amount of peptide available to the MHC-presenting machinery is not sufficient to induce a cell response which is detectable with our assay, or that the reacting CD8 $^+$ T cell population does not expand over the detection limit of our assay.

On the whole, the results presented here allowed the identification of a number of antigens which may be available for proteasome-mediated processing in the course of *C. pneumoniae* infections, proposing them as possible targets for a HLA-A2-dependent cellular immune response. Further analysis will be required to verify if the specifically induced CD8 $^+$ T cells are able to recognize and induce lysis of peptide pulsed or *C. pneumoniae* infected mammalian cells and, possibly, to correlate the identified T cell epitopes with CD8 $^+$ T cell populations naturally induced in *C. pneumoniae* infected patients. Importantly, the results obtained with DNA-mediated expression of distinct antigens, can represent an initial step towards the definition of a significant set of *C. pneumoniae* HLA-A2-restricted epitopes, which could eventually be combined in DNA minigenes in the attempt to induce a CTL-dependent anti-*Chlamydia* protective immune response

25 Example 9

Immunizations with Combinations of the First Antigen Group

The five antigens of the first antigen group (OmpH-like protein, pmp10, pmp2, Enolase, OmpH-like, CPn0042 and CPn00795) were prepared as described in the Materials and Methods Section above for Examples 1-4. The antigens are expressed and purified. Compositions of antigen combinations are then prepared comprising five antigens per composition (and containing 15 μ g of each antigen per composition). CD1 mice are divided into seven groups (5-6 mice per group for groups 1 through 4; 3 to 4 mice for groups 5, 6 and 7), and immunized as follows:

Group	Immunizing Composition	Route of Delivery
1	Mixture of 5 antigens (15 μ g/each) + CFA	Intra-peritoneal
2	Mixture of 5 antigens (15 μ g/each) + AIOH (200 μ g)	Intra-peritoneal
3	Mixture of 5 antigens (15 μ g/each) + AIOH (200 μ g) + CpG (10 μ g)	Intra-peritoneal
4	Complete Freund's Adjuvant (CFA)	Intra-peritoneal
5	Mixture of 5 antigens (5 μ g/each) + LTK63 (5 μ g)	Intranasal
6	AIOH (200 μ g) + CpG (10 μ g)	Intra-peritoneal
7	LTK63 (5 μ g)	Intranasal

Mice are immunized at two week intervals. Two weeks after the last immunization, all mice are challenged by intravaginal infection with *Chlamydia pneumoniae* serovars.

40 Experiment 9 was repeated with another group of CPn antigens. These were:

CPn0385 (PepA), CPn0324 (LcrE), CPn0503 (DnaK), CPn0525 (Hypothetical) and CPn0482 (ArtJ). These antigens are combined and administered with and without alum and CpG as described in Experiment 9.

5 Summary

Applicants have identified a number of CPn proteins with desirable immunological and/or biological properties. Specifically, at least twelve CPn proteins have been identified which are capable of inducing the production of antibodies, which can neutralise, in a dose-dependent manner, the infectivity of *C. pneumoniae* in *in vitro* cell cultures. The induction of neutralising antibodies is important because it prevents infectious EBs from invading human tissues. Furthermore, at least six of these CPn proteins were also capable of attenuating *Chlamydial* (*C. pneumoniae*) infection in a *in vivo* hamster model. In addition, some of these CPn proteins were also capable of inducing not only adequate T-cell responses but also high serum levels of neutralising antibodies.

Apart from very recent unpublished results on pmp21, this is the first time that antisera to recombinant pmgs (pmp2 and pmp10) are reported to have neutralising properties. Interestingly, whilst antiserum against CPn0525 gave negative *in vitro* results (ie no neutralising activity), the CPn0525 protein gave 97 per cent protection from spleen infection in an *in vivo* hamster immunisation assay (see Table 2) (ie a positive *in vivo* result). Likewise, whilst antiserum against Cpn0498 gave negative *in vitro* results (ie no neutralising activity), the CPn0498 protein gave 94 per cent protection from spleen infection in an *in vivo* hamster immunisation assay (ie a positive *in vivo* result). Thus a positive signal obtained in the FACS assay does not guarantee a corresponding positive *in vitro* neutralization activity and conversely a negative neutralization activity does not mean that a positive *in vivo* result can be excluded.

Some of the results obtained by screening the panel of recombinant antigens with the *C.pneumoniae in vitro* neutralization assay are shown in Table 2. Just by a cursory look at the 'current annotation' column it can be seen that both in Table 1 and 2 are listed antigens, like the members of the family of heterogeneous polymorphic membrane proteins (PMP), which, on the basis of published literature data, could be reasonably expected to be surface-exposed and possibly induce neutralizing antibodies, but there are also proteins which could be considered so far only hypothetical, and proteins which just on the basis of their current functional annotation could not be at all expected to be found on the bacterial surface.

The characterisation for the first time of some of these CPn proteins in terms of not only neutralising properties but also different score profiles in a panel of screening tests is an important contribution to the art because it facilitates the selective combination of CPn antigens with particular immunological and biological properties.

In conclusion, this paper describes a group of recombinant antigens which can induce antibodies inhibiting the infectivity of *C. pneumoniae in vitro* and have protective effects *in vivo*.

All publications mentioned in the above specification are herein incorporated by reference. Various modifications and variations of the described methods and system of the invention will be apparent to those skilled in the art without departing from the

scope and spirit of the invention. Although the invention has been described in connection with specific preferred embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention
5 which are obvious to those skilled in molecular biology or related fields are intended to be covered by the present invention.

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CPn ID	Protein	Description	Accession	Length	Score	Fig2	yes	no	Score	Score	Score
CPn0449	0010-GST	Pmp 10, family of polymorphic outer membrane proteins	1-25	94503	5.22	40.24	Fig2	yes	287	92	0.0340*
CPn0013	6270-GST	Pmp 2, family of polymorphic outer membrane proteins	1-19	67459	5.84	34.99	Fig2	yes	87	92	0.0338*
CPn0482	6767-GST	ArtJ, Arginine binding protein?	7(*)	29070	5.45	20.16	Fig2	yes	105	43	0.2918
CPn0800	7111-GST	Eno, Enolase	no	46104	4.66	11.64	Fig2	yes	413	89	0.0005*
CPn0979	7306-GST	HtrA, chaperonin, serine protease	1-20	50142	6.39	28.67	Fig2	yes	249	0	0.9833
CPn301	6577-GST	OmpH-like outer membrane protein	1-21	17264	4.75	10.05	Fig2	yes	847	84	0.0067*
CPn0042	6296-GST	Cpn specific hypothetical, several possible paralogs, including the FACS+ Cpn0126	no	31.1	5.40	22.77	ND	no	80	84	0.0002*
CPn0795	7106-GST	Cpn specific hypothetical	no	39.5	7.50	14.84	ND	no	487	89	0.0109*
CPn0584	6878-GST	AtoS; Two-component sensor histidine kinase, AtoS-related	no	40.4	7.50	14.61	ND	no	270	74	0.3457
CPn0558	6850-GST	OmcA - predicted 9 kD cysteine rich, outer membrane protein, lipoprotein?	lipoprotein?	7.4	7.56	20.45	ND	no	197	64	0.2061

TABLE 1

TABLE 3 (i)

Expressed genes of <i>Chlamydia pneumoniae</i> EB selected by microarray	Annotation	CPMedia
CPN gene		
DN1+2		56951.62
CPn0332	CHLIR T2 Protein	52466.90
ompB	ompB 60 kDa Cysteine-Rich OMP	43304.22
ompA	ompA 9 kDa Cysteine-Rich Lipoprotein	42530.16
ompA	ompA Major Outer Membrane Protein	40478.10
CPn0678	hypothetical protein	34457.06
hcrB	hcrB Histone-like Protein-2	33501.69
CPn0331	CT082 hypothetical protein	33219.36
lorH_1	lorH_1 Low Calcium Response Protein H	27994.28
CPn0474	CT365 hypothetical protein	26041.66
CPn0443	CT005 hypothetical protein	25788.78
CPn0808	CT579 hypothetical protein	25269.04
groEL_1	groEL_1 Heat Shock Protein-60	25020.29
CPn0499	hypothetical protein	23843.93
yyjD	yyjD conserved hypothetical protein	23573.29
CPn0677	hypothetical protein	22986.01
DN9+10		22691.56
CPn0333	LutB Protein	22416.03
CPn0369	CT058 hypothetical protein	21736.31
CPn0473	hypothetical protein	21552.78
pmp_19	pmp_19 polymorphic membrane protein A Family	19039.90
CPn0728	CHLIR 76 kDa Homolog 1 (CT622)	18847.95
CPn0809	CT578 hypothetical protein	18822.27
CPn0676	Similarity to CT695	17430.80
CPn0524	hypothetical protein	14635.06
CPn1916	CT858 hypothetical protein CPAF protease 1	14296.87
CPn0234	CT181 hypothetical protein	14240.84
CPn0588	CT469 hypothetical protein	14094.71
CPn0665	CT288 hypothetical protein	13905.70
flaH	flaH ATP-dependent zinc protease	13661.27
CPn0428	hypothetical protein	12804.32
CPn0370	CT058 hypothetical protein	12727.99
CPn0810	CT577 hypothetical protein	12334.96
CPn0875	CT734 hypothetical protein	11123.85
CPn0472	hypothetical protein	10901.14
yifF	yifF Cationic Amino Acid Transporter	10803.42
CPn0537	CT814.1 hypothetical protein	10288.45
CPn0467	hypothetical protein	10234.67
tpsD	tpsD Sigma-28/WigG Family	9680.56
argJ	argJ Arginine Periplasmic Binding Protein	9553.60
CPn0329	Phospholipase D Superfamily	9017.13
yecA_2	yecA_2 Transport Permease	8825.55
CPn0572	CT456 hypothetical protein	8664.88
CPn0796	hypothetical protein FACS 4	8663.74
dagA_2	dagA_2 D-Alanine/Glycine Permease	8448.78
xeiD	xeiD Integrase/recombinase	8202.45
CPn0726	CT620 hypothetical protein	8175.83
pgi	pgi Glucose-6-P Isomerase	8055.55
thrS	thrS Threonyl-tRNA Synthetase	7987.45
CPn0720	CT659 hypothetical protein	7766.32
CPn0589	CT470 hypothetical protein	7758.09
CPn1004	CT847 hypothetical protein	7735.45
CPn0383	CT047 hypothetical protein	7292.22
necB	necB AcCoA Carboxylase/Transferase Beta	7137.78
CPn1005	CT848 hypothetical protein	7135.44
CPn0933	predicted disulfide bond isomerase	7119.73
clpP_1	clpP_1 CLP Protease	7039.67
CPn0964	hypothetical protein	6960.16
CPn0338	CT814 hypothetical protein	6882.66
pmp_15	pmp_15 Polymorphic Outer Membrane Protein B Family	6837.11
yscN	yscN Yop N (Flagellar-Type ATPase)	6752.80
yscN	yscN Yop N (Flagellar-Type ATPase)	6752.80
tlyC_2	tlyC_2 CBS Domains (Hemolysin homolog)	6692.52
dnjK	dnjK Heat Shock Protein-70	6667.15
secD/secE	secD/secE Protein Export Proteins SecD/SecE (fusion)	6650.81
vacB	vacB Ribonuclease family	6535.30
pmp_2	pmp_2 Polymorphic Outer Membrane Protein G Family	6452.21
recA	recA RecA recombination protein	6369.57
arcD	arcD Arginine/Oribthine Antipporter	6360.19
dsbD	dsbD Thiodisulfide Interchange Protein	6356.90
pmp_6	pmp_6 Polymorphic Outer Membrane Protein G/I Family	6321.42

TABLE 3 (ii)

nqrA	nqrA Ubiquinone Oxidoreductase, Alpha	6319.39
gyrB_2	gyrB_2 DNA Gyrase Subunit B	6257.92
CPn0001	CT001 hypothetical protein	6197.90
mdhC	mdhC Malate Dehydrogenase	6185.13
lerD	lerD Low Calcium Response D	6173.29
ygeA	ygeA rRNA Methyltransferase	6160.83
dagA_1	dagA_1 D-Ala/Gly Permease	6131.94
CPn0105	CT016 hypothetical protein	6102.59
ompB	ompB Outer Membrane Protein B (porB)	6032.94
yehF	yehF GTP Binding Protein	6024.47
CPn0768	CT668 hypothetical protein	6020.13
glyA	glyA Serine Hydroxymethyltransferase	5975.37
glyA	glyA Serine Hydroxymethyltransferase	5973.37
gpdA	gpdA Glycerol-3-P Dehydrogenase	5957.93
CPn0814	CT573 hypothetical protein	5949.06
pmp_12	pmp_12 Polymorphic Outer Membrane Protein (truncated) A/I Family	5942.77
CPn0507	CT421.1 hypothetical protein	5893.30
pmp_7	pmp_7 Polymorphic Outer Membrane Protein G Family	5881.74
yscC	yscC YopC/Gen Secretion Protein D	5874.56
glgB	glgB Glucan Branching Enzyme	5860.93
CPn0557	CT356 hypothetical protein	5800.47
CPn0925	CT779 hypothetical protein	5795.07
yscJ	yscJ Yeh Translocation J	5794.03
CPn0512	CT425 hypothetical protein	5771.00
pmp_8	pmp_8 Polymorphic Outer Membrane Protein G Family	5762.89
glxX	glxX Glutaryl-tRNA Synthetase	5753.53
CPn0629	hypothetical protein	5743.24
r13	r13 L3 Ribosomal Protein	5696.59
flhN	flhN Flagellar Motor Switch Domain/YscO family	5691.86
CPn0497	CT388 hypothetical protein	5674.36
tal	tal Transaldolase	5654.13
ldh	ldh Lactate Dehydrogenase	5644.61
fabI	fabI Enoyl-Acyl-Carrier Protein Reductase	5629.90
CPn0017		5609.25
yzbB	yzbB ABC transporter permease	5608.13
fisY	fisY Cell Division Protein FisY	5606.91
pmp_21	pmp_21 Putative Outer Membrane Protein D Family	5604.24
CPn0727	CT619 hypothetical protein	5599.19
pmp_13	pmp_13 Polymorphic Outer Membrane Protein G Family	5583.84
clpC	clpC ClpC Protease	5577.87
CPn0107	CT058 hypothetical protein	5564.97
CPn0927	CHLPS 43 kDa protein homolog 2	5564.93
clpB	clpB Clp Protease ATPase	5558.20
dacF	dacF D-Ala-D-Ala Carboxypeptidase	5557.25
CPn0243	hypothetical protein	5542.02
ahpC	ahpC Thio-specific Antioxidant (TSA) Peroxidase	5528.77
proS	proS Prolyl tRNA Synthetase	5528.37
gphD	gphD CHLTP Plasmid Paralog	5527.11
yagE	yagE YagE family	5514.15
CPn0590	CT471 hypothetical protein	5498.93
CPn0832	CT563 hypothetical protein	5498.31
CPn0807	CT380 hypothetical protein	5497.50
CPn0929	CHLPS 43 kDa protein homolog 4	5497.45
dhli	dhli DNA Ligase	5470.60
folD	folD Methylene Tetrahydrofolate Dehydrogenase	5469.26
r11	r11 L1 Ribosomal Protein	5468.39
r11	r11 L1 Ribosomal Protein	5468.39
CPn0483	hypothetical protein	5434.74
clpP_2	clpP_2 Clp Protease Subunit	5418.10
rhoA	rhoA RNA Polymerase Alpha	5409.04
fabH	fabH Oxoacyl Carrier Protein Synthase III	5408.47
fusA	fusA Elongation Factor G	5403.23
CPn0220	hypothetical protein	5400.77
yebC	yebC YebC family hypothetical protein	5399.50
CPn1033	CT373 hypothetical protein	5396.91
gapA	gapA Glyceraldehyde-3-P Dehydrogenase	5377.30
appD	appD ABC ATPase/Dipeptide Transport	5345.31
CPn0404	hypothetical protein	5322.62
pmp_14	pmp_14 Polymorphic Outer Membrane Protein H Family	5296.75
CPn0753	hypothetical protein	5290.01
CPn0462	hypothetical protein	5289.16
CPn0381	CT326 similarity	5285.39
CPn0374	CT056 hypothetical protein	5269.65

TABLE 3 (iii)

CPn0799	hypothetical protein	5269.61
CPn0862	CT289 hypothetical protein	5249.79
alsS	alsS, Alanine tRNA Synthetase	5243.94
CPn0126	hypothetical protein	5240.36
rs4	rs4, S4 Ribosomal Protein	5239.14
CPn0623	CT504 hypothetical protein	5234.03
glgC	glgC, Glucose-1-P Adenyltransferase	5214.97
CPn0365	hypothetical protein	5195.21
CPn0458	hypothetical protein	5178.06
CPn0066	hypothetical protein	5156.84
CPn0742	CT635 hypothetical protein	5133.02
CPn0461	hypothetical protein	5118.37
soxB	soxB, Protease	5106.34
CPn0675	CT696 hypothetical protein	5104.52
CPn0405	CT102 hypothetical protein	5095.20
lpxA	lpxA, Acyl-Carrier UDP-GlcNAc O-Acyltransferase	5072.90
pmp_17.3	pmp_17.3, Polymorphic Outer Membrane Protein (Frame-shift with CPn0469)	5068.28
pepF	pepF, Oligopeptidase	5065.00
uhpC	uhpC, Hexosphosphate Transport	5054.37
CPn0703	CT671 hypothetical protein	5063.25
CPn0874	CT733 hypothetical protein	5047.64
pepA	pepA, Leucyl Aminopeptidase A	5044.06
lpxA	lpxA, Lipote Synthetase	5008.60
CPn0514	CT427 hypothetical protein	5007.93
nbaA/nbaB	nbaA/nbaB, GTP Cyclohydrolase & DHBP Synthase	5005.81
dat	dat, dUTP Nucleotidohydrolase	4982.57
CPn0928	CHLPS 43 kDa protein homolog 3	4982.06
CPn0794	hypothetical protein FACS+ selected by Rita?	4965.51
CPn0798	hypothetical protein	4957.17
pnp	pnp, Polynucleotide Nucleotidyltransferase	4954.15
fabG	fabG, Oxonol (Carrier Protein) Reductase	4947.36
CPn0513	Fe-S oxidoreductase	4935.65
folP	folP, Dihydropteridine Synthase	4934.67
yhlH	yhlH, Dicarboxylase Translocator	4929.02
pdaA/pdaB	pdaA/pdaB (pyruvate), Oxoisovalerate Dehydrogenase Alpha & Beta Fusion	4916.85
pyrH	pyrH, UMP Kinase	4914.40
abcX	abcX, ABC Transporter ATPase	4897.52
sucA	sucA, Oxoglutarate Dehydrogenase	4895.62
yacB	yacB, predicted phosphatase/kinase	4888.84
CPn0595	CT476 hypothetical protein	4877.72
CPn0014		4876.89
phnP	phnP, Metal Dependent Hydrolase	4866.38
tyrP_2	tyrP_2, Tyrosine Transport	4861.62
CPn0559	CT444.L hypothetical protein	4860.36
CPn0878	SBT Domain protein	4858.94
ygg4	ygg4, Phosphate Permease	4856.29
lufA	lufA, Elongation Factor Tu	4842.50
lpxA	lpxA, Lipamide Dehydrogenase	4831.72
CPn0397	PE2C phosphatase family	4831.19
flvY	flvY, Glutamate Binding Protein	4828.17
phoH	phoH, ATPase	4814.17
glcT	glcT, Glutamate Symport	4807.24
lepA	lepA, GTPase	4801.42
rho	rho, Transcription Termination Factor	4800.55
yehM	yehM, Sulfate Transporter	4793.79
pmp_16	pmp_16, Polymorphic Outer Membrane Protein E Family	4791.71
adt_2	adt_2, ADP/ATP Translocase	4773.25
CPn0486	hypothetical proline permease	4763.62
trxB	trxB, Thioredoxin Reductase	4749.81
aroG	aroG, Deoxyheptarate Aldolase	4747.65
yggV	yggV, Hypothetical Protein	4722.09
eybS	eybS, Cysteinyl tRNA Synthetase	4700.25
CPn0690	ABC Transporter Membrane Protein	4695.46
nqr2	nqr2, NADH (Ubiquinone) Dehydrogenase	4691.83
pmp_11	pmp_11, Polymorphic Outer Membrane Protein G Family	4690.71
CPn0035	CT339 hypothetical protein	4686.43
CPn0797	hypothetical protein FACS+ selected by Rita?	4681.52
yacT	yacT, Omp85 Analog	4680.61
greA	greA, Transcription Elongation Factor	4676.81
edsA	edsA, Phosphatidate Cytidylyltransferase	4673.11
CPn0803	CT584 hypothetical protein	4667.94
CPn0021	Predicted-OMP [leader peptide]	4639.48
isp	isp, Tail-Specific Protease	4639.10

TABLE 3(IV)

rl6	rl6 L6 Ribosomal Protein	4636.73
gpfB	gpfB HSP-70 Cofactor	4635.78
fabF	fabF Acyl Carrier Protein Synthase	4633.09
mgfE	mgfE Mg++ Transporter (CBS Domain)	4611.86
himD	himD Integration Host Factor Alpha	4600.92
CPn0422	CT273 hypothetical protein	4600.34
CPn0692	ABC Transporter	4591.16
CPn0415	CT266 hypothetical protein	4586.28
aspC	aspC Aspartate Aminotransferase	4534.97
ded	ded dCTP Deaminase	4533.88
CPn0010.1	frame shift with CPn0910	4530.44
atoC	atoC 2-Component Regulator	4528.67
galB	galB (Pet112) Glu-tRNA-Gln Amidotransferase (B Subunit)	4510.59
CPn0827	CT560 hypothetical protein	4507.66
yccA	yccA YccA hypothetical protein	4491.46
metG	metG Methionyl-tRNA Synthetase	4490.20
atpA	atpA ATP Synthase Subunit A	4483.96
CPn1054	hypothetical protein	4483.87
rl5	rl5 L5 Ribosomal Protein	4482.91
xseA	xseA Exodeoxyribonuclease VII	4469.21
CPn0859	CT718 hypothetical protein	4460.34
CPn0449	pmp_10 (frameshift)	4458.00
nqr5	nqr5 NADH (Ubiquinone) Reductase 5	4453.91
CPn0041	hypothetical protein	4453.05
accA	accA AcCoA Carboxylase/Transferase Alpha	4443.85
oppA_2	oppA_2 Oligopeptide Binding Protein	4442.36
valS	valS Valyl-tRNA Synthetase	4442.14
CPn0456	hypothetical protein	4436.51
CPn0681	CT691 hypothetical protein	4426.13
dnaA_2	dnaA_2 Replication Initiation Factor	4424.21
CPn0087	hypothetical protein	4423.42
yseT	yseT YopT Translocation T	4421.69
rs3	rs3 S3 Ribosomal Protein	4418.77

Chlamydia Further Filing/
Microarray Expressed Gene Table
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TABLE 3(v)-(xi)

CPn	CT	Protein Function	MRNA levels	Stage in the Chlamydial developmental cycle	Reference (see legend)	EB Surface exposed	RB secreted
	CT081	CHLTR T2 protein		Very Late	Nicholson		
0557	CT443	OmcB	43304.22	Late Gene	Belland Shaw	EB Surface exposed	
0558	CT444	OmcA	42530.16	Late Gene	Belland	Yes FACS pos In vivo protective with CT467	
0695	CT681	MOMP		Mid Late II		FACS positive	
0384	CT046	hctB		Late Gene	Belland		
0331	CT082	Hypothetical	33219.36	Late	Nicholson	FACS positive	
0811	CT576	LcrH		Late Gene	Belland		
0474	CT365	Hypothetical		Immediate/Early	Belland		
0443	CT005						
0808	CT579	Hypothetical		Late Gene	Belland		Type III secretion cluster (WO 02/082091)
0134	CT110	GroEL Heat shock protein (Hsp-60)		Immediate Early gene Midcycle Midlate I	(see Table 1 of Belland et al (2003) Nicholson	FACS positive	
0499		Cpn hypothetical protein					
		Yyd conserved hypothetical protein					
		CPn specific protein					
0333	CT080	ItuB		Late Gene	Belland		
0369	CT058						
0539	CT412	Pmp19	19039.90			FACS positive	
0728	CT622						
0809	CT578			Late Gene	Belland		
0676	CT695						
1016	CT858	predicted Protease containing IRBP					Secreted Protein (WO 02/082091) Epitope

Chlamydia Further Filing
Microarray Expressed Gene Table
15-9-04 and 17-12-04

TABLE 3(v)-(xi)

0234	CT181	Hypothetical				FACS positive	
0588	CT469	Hypothetical					
0065	CT288						
0998	CT841						Epitope (WO 02/082091)
0369	CT058						
0810	CT577	Hypothetical					Type III secretion cluster (WO 02/082091)
0875	CT734	Hypothetical		Immediate Early	Belland		Secreted Protein (WO 02/082091)
0127	CT034						
0538	CT814			Late	Belland		
0482	CT381	ArtJ				FACS positive	
0329	CT154-158	CT157 Phospholipase D Superfamily				FACS positive	
0572	CT456	Hypothetical (now TarP)	8664.88			FACS positive	CT456 is secreted from Chlamydia by a Type III secretion system (TTSS) (Clifton et al 2004) and is translocated into the cytoplasm of the host cell. New function assigned = TarP = translocated actin-recruiting phosphoprotein CT456 gene is transcribed from mid to late cycle in the Chlamydia developmental cycle CT456 may vary across the serovars because it has tandem repeats
0876	CT735						

Chlamydia Further Filing/
Microarray Expressed Gene Table
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TABLE 3(v)-(xi)

1024	CT864						
0726	CT620						
1025	CT378						
0482	CT581						
0720	CT659			Late	Belland		
0589	CT470						
1004	CT847			Late	Belland		
0383	CT047						
0058	CT293						
1005	CT848						
0933	CT783	predicted disulfide bond isomerase					Secreted Protein (likely to be a Type III secretion protein) (WO 02/082091)
0520	CT431						
0538	CT814						
0466	CT869	PmpE				FACS positive	
0707	CT669	yscN					Type III secretion protein
0707	CT669	yscN		Mid-cycle	Shaw		
0503	CT396	DnaK	6667.15	Late	Nicholson	FACS positive	
0564	CT448						
0504	CT397						
0453	CT871	PmpG		Late	Belland	FACS positive	
0762	CT650	recA					Secreted Protein (WO 02/082091)
1031	CT374						
0786	CT595						
0743	CT634						
0715	CT661						
0001	CT001			Late	Nicholson		
1028	CT376						
0323	CT090						
0885	CT742						
0876	CT735						
0105	CT016	Hypothetical				FACS positive	Secreted Protein (see WO 02/48185), pg 36 (ie secreted by a Type III apparatus)
0854	CT713	PorB		Midlate II	Nicholson	FACS positive	

Chlamydia Further Filing/
Microarray Expressed Gene table
15-9-04 and 17-12-04

TABLE 3(v)-(xi)

0321	CT092						
0708	CT668	Hypothetical					Secreted Protein (WO 02/082091)
0520	CT432						
0521	CT432						
0855	CT714						
0814	CT573						
0506	CT421						
0702	CT674						
0475	CT866						
1057	CT356						
0925	CT779						
0828	CT559	YscJ				FACS positive	
0512	CT425						
0560	CT445						
0647	CT528						
0704	CT672			mid late I	Nicholson		
0497	CT388						
0083	CT313						
0919	CT773						
0406	CT104						
1012	CT854						
0880	CT739						
0963	CT812	PmpD				FACS positive	
0727	CT619						
0437	CT286	ClpC					Secreted Protein (WO 02/082091)
0369	CT058						
0134	CT110	Hsp-60 (omp2) Chaperonin		Immediate Early gene Midcycle Midlate I	(see Table 1 of Belland et al (2003) Nicholson	FACS positive	
0672	CT551						
0778	CT603						
0500	CT393						
0804	CT583						
0590	CT471						
0822	CT565						
0807	CT580						
0149	CT146						
0335	CT078						

Chlamydia Further Filing/
Microarray Expressed Gene Table
15-9-04 and 17-12-04

TABLE 3(v)-(xi)

0078	CT318						
0078	CT318						
0520	CT431						
0626	CT507						
0298	CT239			Mid late	Nicholson		
0550	CT437						
0573	CT457						
1032	CT373						
0624	CT505						
0682	CT690						
0381	CT326						
0374	CT056						
0062	CT289						
0598	CT479						
0733	CT626						
0623	CT504						
0607	CT489						
0742	CT635	Hypothetical				FACS Positive	
0613	CT494			mid late II	Nicholson		
0675	CT696						
00405	CT105						
0650	CT531						
0136	CT112						
0665	CT544						
0705	CT671	Hypothetical				FACS Positive	WO 02/48185 CT671 = secreted protein, pg 36 (ie secreted by a Type III apparatus) Epitope
0874	CT733						
0385	CT045	PepA	5044.06	Mid late I	Nicholson	FACS Positive	
0832	CT558						
0514	CT427						
0904	CT761	MurG	5005.81	See biogenome paper 2004 - MurG was consistently selected across the Serovars		FACS Positive	
0059	CT292	dut					Secreted Protein (WO 02/082091)

Chlamydia Further Filing/
Microarray Expressed Gene Table
15-9-04 and 17-12-04

TABLE 3(v)-(xi)

0999	CT842						
0296	CT237						
0513	CT426						
0758	CT613						
0207	CT204						
0304	CT245						
0698	CT678						
0191	CT130						
0378	CT054						
0611	CT492						
0584	CT467	AtoS	4877.72	Cross-reactivity between CT and CPn strains Ie CT467 and its CPn homologue are neutralising for their own species but are also cross-protective		FACS Positive	
0479	CT380						
0970	CT818						
0558	CT444	Omca		Late gene (see Table 1 of Belland et al (2003))		FACS Positive In vivo protective effect with CT444 and CT467	
0878	CT737						
0680	CT692						
0074	CT322						
0833	CT557						
0793	CT588						
0604	CT486						
0106	CT015						
0528	CT401						
0359	CT064						
0610	CT491			Mid late II	Nicholson		
1014	CT856						
0466	CT869?						
0351	CT065	ADP/ATP Translocase					
0314	CT099						
0484	CT382						
0932	CT782						
0690	CT686						
0427	CT278						
0453	CT871	pmpG		Late gene	(see Table 1 of Belland et	FACS Positive	

Chlamydia Further Filing/
Microarray Expressed Gene Table
15-9-04 and 17-12-04
TABLE 3(v)-(xi)

					al (2003)		
0035	CT339						
0300	CT241						
0741	CT636						
0567	CT451						
0803	CT584						
0555	CT441						
0633	CT514						
0502	CT395						
0916	CT770						
0286	CT194						
0416	CT267						
0422	CT273						
0692	CT684						
0415	CT266	Hypothetical				FACS Positive	
0495	CT390						
0392	CT039						
0586	CT468						
0004	CT004						
0827	CT560	Hypothetical protein					Type III secretion cluster (WO 02/082091)
0734	CT627						
0122	CT032						
0657	CT537						
0635	CT516						
1062	CT329						
0859	CT718						
0430	CT281						
0414	CT265	AccA					Secreted Protein (WO 02/082091)
0599	CT480	Oligopeptide binding lipoprotein (Oppa)		Immediate early gene	(see Table 1 Belland et al 2003)	FACS Positive	
0094	CT302						
0681	CT691	Hypothetical					Secreted Protein (WO 02/082091) Epitope
0309	CT250						
0823	CT564						
0641	CT522						

Table 4. *C. pneumoniae* selected peptides: protein sources and HLA-A2 stabilization assay

Peptide	Sequence	CPn ^{a)}	Protein	Group ^{b)}	Score ^{c)}	Net MFI ^{d)}
HepB	¹²⁵ TAFHQTLQD ¹³³		Hepatitis B virus envelope antigen		=====	14.0±24.4
GAG	⁷⁷ SLYNTVATL ⁸⁵		HIV-1 gag		157.22	92.4±23.8
IMa	⁵⁸ GILGFVFTL ⁶⁶		Influenza virus matrix M1		550.92	63.1±18.1
CH1	³¹ 5QLLEDEGKEL ³²³	0322	Yop proteins translocation protein U	Type III	324.06	74.0±22.6
CH2	⁴³³ ILLNEVPYV ⁴⁴¹	0323	Low calcium response protein D	Type III	5534.14	140.5±36.1
CH3	³⁴³ VLNLFSSAL ³⁵¹	0324	Low calcium response protein E	Type III	262.20	40.1±23.1
CH4	⁷ QLLESLAPL ¹⁵	0325	Secretion chaperone	Type III	745.35	120.1±25.2
CH5	²⁷¹ SILELLQFV ²⁷⁹	0702	Probable Yop proteins translocation protein C	Type III	1835.22	85.5±34.4
CH6	⁷⁸ YLLEEIVTV ⁸⁷	0811	Low calcium response protein H	Type III	11162.99	148.5±38.9
CH7	⁸³ YMDNNLFYV ⁹¹	0823	Yop proteins translocation protein T	Type III	6781.36	164.1±24.3
CH8	²⁵⁴ FLTLAWWF ²⁶²	0823	Yop proteins translocation protein T	Type III	3365.36	144.1±22.1
CH10	²¹⁴ GLTEEIDYV ²²²	0828	Yop proteins translocation protein J	Type III	1767.58	144.0±37.9
CH12	⁷⁹ WLVEFNPFV ⁸⁷	1021	Low calcium response locus protein H	Type III	6686.72	50.1±22.2
CH13	⁶⁹ YVFDRILKV ⁷⁷	0695	Outer membrane protein A	Ch spec	976.76	139.0±36.7
CH14	⁴⁰⁶ VMLIFEKKV ⁴¹⁴	0415	CT2 66 hypothetical protein	Cpn spec	1200.64	74.1±20.2
CH15	¹²⁷⁰ YLTSYSPYV ¹²⁷⁸	0444	Polymorphic outer membrane protein G/I family	Pmp	1759.66	138.1±23.5
CH16	¹⁵³⁰ VQLAYVFDV ¹⁵³⁸	0963	Putative outer membrane protein D family	Pmp	591.70	48.1±19.1
CH17	³⁰⁸ ILQEAQMV ³¹⁶	0728	76 kDa homolog_1	Ch spec	484.77	202.1±24.2
CH18	⁷¹ IALLVIFV ⁷⁹	0186	Similar to CT119 IncA	Ch spec	445.80	46.1±22.3
CH19	¹³²⁷ LLLTGAYV ¹³³⁵	0444	Polymorphic outer membrane protein G/I family	Pmp	437.48	56.1±21.6
CH20	¹⁴⁶ ALMLNNYV ¹⁵⁴	0005	Polymorphic outer membrane protein G family	Pmp	1415.38	142.5±38.6
CH21	⁶¹⁴ TLWGSFVDV ⁶²²	0447	Polymorphic outer membrane protein G/I family	Pmp	1096.83	121.1±18.0
CH22	¹⁵⁶⁶ WLFDLRFV ¹⁵⁷⁴	0540	Polymorphic membrane protein B family Pmp	28150.17	68.5±11.0	
CH24	⁴⁹⁰ LIQETLLFV ⁴⁹⁹	0021	Predicted OMP	Ch spec	843.21	105.1±20.8
CH28	⁴⁵ RLLEIIWGV ⁵³	0062	CHLPS 43 kDa protein homolog_1	Cpn spec	18200.54	99.5±15.0
CH29	²⁸³ YLMQKLQNV ²⁹⁶	0791	CT 590 hypothetical protein	Ch spec	2722.68	108.5±12.1
CH30	⁵²⁰ FLQRGESFV ⁵²⁸	0792	CT 589 hypothetical protein	Ch spec	759.66	105.1±8.1
CH31	⁴⁰¹ WLLRDDWLL ⁴⁰⁹	0009	hypothetical	Cpn spec	2726.91	101.1±16.3
CH32	¹⁸⁷ KLWEWLGYL ¹⁹⁵	0041	hypothetical	Cpn spec	4184.21	72.1±12.0
CH33	⁶⁸ LLMLAISLV ⁷⁵	0131	hypothetical	Cpn spec	1006.20	18.0±1.4
CH34	²⁰¹ KLLKDHF ²⁰⁹	0132	hypothetical	Cpn spec	1604.53	85.1±4.9
CH35	⁵⁶ ILSFLPWL ⁶⁴	0169	hypothetical	Cpn spec	886.78	90.6±5.7
CH36	¹⁴⁹ LLLIFNNYL ¹⁵⁷	0170	hypothetical	Ch spec	2808.32	41.0±19.8
CH37	¹²⁶ YLLDFRWPL ¹³⁴	0210	hypothetical	Cpn spec	42485.26	97.1±17.7
CH38	³⁷⁴ NLLKRWQFV ³⁸²	0352	hypothetical	Cpn spec	2406.15	64.1±13.4
CH39	³⁷⁸ FLLRHLSSV ³⁸⁶	0355	hypothetical	Cpn spec	2722.68	88.1±6.4
CH41	¹⁶² KLSEQLAL ¹⁷⁰	0186	Similar to CT119 IncA	Ch spec	345.48	51.6±27.6
CH42	²¹⁴ KVLLGQEWV ²²²	0186	Similar to CT119 IncA	Ch spec	212.39	16.0±37.5
CH43	³¹⁵ NLAEQVTAL ³²³	0186	Similar to CT119 IncA	Ch spec	201.44	71.6±34.6
CH44	¹²³ YVVGFIIFL ¹³¹	0323	Low calcium response protein D	Type III	413.32	25.0±16.3
CH45	³² WMMGVVLM ⁴⁰	0323	Low calcium response protein D	Type III	294.95	8.1±18.4
CH46	⁵⁶ NLSISVFL ⁶⁴	0323	Low calcium response protein D	Type III	284.97	18.0±26.2
CH47	¹¹⁰ VIQAFGDFV ¹¹⁸	0323	Low calcium response protein D	Type III	166.49	23.0±32.5
CH48	⁶³⁵ YLALDPDSV ⁶⁴³	0323	Low calcium response protein D	Type III	156.77	74.6±34.4
CH49	¹⁴⁹ KMSHFQQAL ¹⁵⁷	0415	CT2 66 hypothetical protein	Cpn spec	205.19	29.0±33.9
CH50	¹¹⁸⁷ SLCAQSSYV ¹¹⁹⁵	0444	Polymorphic outer membrane protein G/I family	Pmp	382.53	45.1±22.6
CH51	¹³⁶⁰ NLSRQAFFA ¹³⁶⁸	0444	Polymorphic outer membrane protein G/I family	Pmp	158.47	25.0±34.5
CH52	⁶⁷⁶ SLLEEHPV ⁶⁸⁶	0963	Putative outer membrane protein D family	Pmp	432.59	43.6±21.9
CH53	¹³⁰² NLWSHYTDL ¹³¹⁰	0963	Putative outer membrane protein D family	Pmp	265.96	1.6±24.7
CH54	³⁷⁷ ALWKENQAL ³⁸⁵	0963	Putative outer membrane protein D family	Pmp	177.30	45.6±21.9
CH55	⁵⁶⁸ ALWGHNVLL ⁵⁷⁶	0963	Putative outer membrane protein D family	Pmp	177.30	47.6±29.0
CH56	³³³ NLAGGILSV ³⁴¹	0963	Putative outer membrane protein D family	Pmp	159.97	29.6±29.0
CH57	⁸⁶ FVSKFWFSL ⁹⁴	1021	Low calcium response locus protein H	Type III	322.16	16.0±0.1
CH58	⁷³ SITVFRWL ⁸¹	1021	Low calcium response locus protein H	Type III	272.55	30.0±20.5
CH59	⁵⁸ YLIVFVLT ⁶⁶	0131	hypothetical	Cpn spec	419.44	29.0±10.6
CH60	⁴² VMLFIGLGV ⁵⁰	0131	hypothetical	Cpn spec	315.95	28.0±2.8
CH61	⁷⁶ VLFLIRSV ⁸⁴	0131	hypothetical	Cpn spec	201.24	14.0±2.8
CH62	³⁹⁷ FLFQLGMQI ⁴⁰⁵	0415	CT2 66 hypothetical protein	Ch spec	177.56	30.0±12.0

^{a)} Gene sequence designation as annotated from the genome sequence of Cpn strain CWL029 (<http://chlamydia-berkeley.edu:4231>)

^{b)} Type III: type III secretion system; Ch and Cpn spec: Chlamydia and *C. pneumoniae* specific; Pmp: Polymorphic membrane protein

^{c)} Calculated using the BIMAS algorithm

^{d)} Mean Fluorescence Intensity of cells with peptide - M Mean Fluorescence Intensity of cells without peptide ± Standard Deviation calculated on three experiments

Table 5. ELISpot assay with CD8⁺ T cells from DNA immunized HLA-A2 transgenic mice

Protein	Gene	Peptide	SFC ^{a)}
Hypothetical	CPn 0131	medium	13
		HepB	47
		CH 33	33
		CH 59	53
		CH 60	80
		CH 61	40
Hypothetical	CPn 0210	medium	7
		HepB	13
		CH 37	120
LCR Protein D	CPn 0323	medium	27
		HepB	27
		CH 2	93
		CH 44	80
		CH 45	87
		CH 46	40
		CH 47	80
		CH 48	60
CHLPS 43 kDa	CPn 0062	medium	33
		HepB	27
		CH 28	93
OMP A	CPn 0695	medium	13
		HepB	33
		CH 13	727
LCR Protein H	CPn 0811	medium	13
		HepB	27
		CH 6	213
Yop pt protein T	CPn 0823	medium	7
		HepB	47
		CH 7	493
		CH 8	53
Yop pt protein J	CPn 0828	medium	20
		HepB	60
		CH 10	247

a)

SFC = Spot Forming Colonies/10⁶ CD8 cells

Table 6. IFN- γ production from splenocytes of DNA immunized HLA-A2 transgenic and non transgenic mice

Protein	Gene	Peptide	Ex vivo RFI ^{a)}		stTCLs ^{b)} RFI ^{a)}	
			A2 ⁻ ^{c)}	A2 ⁺ ^{c)}	A2 ⁻ ^{c)}	A2 ⁺ ^{c)}
LCR Protein D	CPn 0323	CH 2	0,05	1,11	0,79	2,57
		CH 44	3,30	1,78	0,73	6,86
		CH 45	1,00	1,56	0,47	4,71
		CH 46	0,90	1,44	0,41	9,00
		CH 47	1,00	1,78	1,17	1,14
		CH 48	1,30	1,67	0,11	1,29
		CD3+CD28	134,00	90,55		
OMP A	CPn 0695	CH 13	3,29	2,54	23,42	209,81
		CD3 + CD28	248,71	73,23		
LCR Protein H	CPn 0811	CH 6	1,00	4,58	1,53	31,56
		CD3 + CD28	290,83	96,10		
Yop pt Protein T	CPn 0823	CH 7	1,20	5,20	11,69	94,57
		CH 8	2,00	1,60	16,81	28,21
		CD3 + CD28	247,60	91,00		

^{a)} Relative Fold Increase: ratio between the percentage of IFN- γ ⁺/CD8⁺ cells obtained with the tested peptide (or the CD3/CD28 co-stimulus) and the HepB negative control peptide

^{b)} Short term T cell lines

^{c)} HLA-A2 non transgenic and transgenic mice

CLAIMS:

1. A polypeptide for use as an autotransporter antigen, the polypeptide comprising:
 - 5 (a) an amino acid sequence selected from the group consisting of SEQ ID NO: 54, SEQ ID NO: 6, SEQ ID NO: 55, SEQ ID NO: 56, SEQ ID NO: 78, and SEQ ID NO: 79,
 - (b) an amino acid sequence having at least 50% sequence identity to an amino acid sequence of (a); or
 - 10 (c) an amino acid sequence comprising one or more fragments of at least 7 consecutive amino acids from an amino acid sequence of (a) or combinations thereof.
2. The polypeptide of claim 1 where use is as an antigen for raising a Chlamydia pneumoniae specific immune response
- 15 3. The polypeptide of claim 2 wherein the use is for raising a systemic immune response in an individual infected with Chlamydia pneumoniae.
4. The polypeptide of any one of claims 1-3 which is secreted into the cytoplasm of the host cell through a Type V autotransporter secretion system mechanism.
- 20 5. The polypeptide of any one of claims 1-3 wherein the polypeptide is selected from the group consisting of SEQ ID NO: 54, SEQ ID NO: 6, SEQ ID NO: 55, SEQ ID NO: 56, SEQ ID NO: 78, and SEQ ID NO: 79 and share one or more common N-terminal sequence motifs selected from the group consisting of G, DG, VG, G, AV, G, IVG, GTLGG, S, IVG, and M.
- 25 6. The polypeptide of claim 5 wherein the common N-terminal sequence motif is selected from the group consisting of GTLGG, S, IVG and M.
- 30 7. The polypeptide of any one of claims 1-3 for use in diagnosis.
8. The use of a polypeptide according to any one of claims 1-3 in the preparation of a medicament for the prevention or treatment of a Chlamydia pneumoniae infection in an individual.
- 35 9. The use according to claim 8 wherein the use is of an autotransporter protein which immunoreacts with seropositive serum of an individual infected with Chlamydia pneumoniae.
- 40 10. The use of a polypeptide according to any one of claims 1-3 in the preparation of an assay for the diagnosis of a Chlamydia pneumoniae infection in an individual.
- 45 11. A method of eliciting an immune response in an individual comprising administering to the individual a polypeptide comprising:
 - (a) an amino acid sequence selected from the group consisting of SEQ ID NO: 54, SEQ ID NO: 6, SEQ ID NO: 55, SEQ ID NO: 56, SEQ ID NO: 78, and SEQ ID NO: 79,
 - 50

(b) an amino acid sequence having at least 50% sequence identity to an amino acid sequence of (a), or

(c) an amino acid sequence comprising one or more fragment of at least 1, 2, 3, 4, 5, 6, or 7 amino acids from an amino acid sequence of (a) or mixtures thereof.

5

12. A method of diagnosing an immune response in an individual comprising:

(a) contacting a biological sample obtained from the individual with a binding agent that binds to a polypeptide according to any one of claims 1-3;

10 (b) detecting in the biological sample the amount of the polypeptide that binds to the binding agent; and

(c) comparing the amount of the polypeptide to a predetermined cut-off value and thereby determining the presence of an immune response in the individual.

15 13. The method of claim 11 or claim 12 wherein the polypeptide is defined according to any one of claims 1-3.

14. The method of claim 11 or the use of according to any one of claims 2-6 or 8-9 wherein the polypeptide.

20 15. A composition for eliciting an immune response comprising one or more Chlamydia pneumoniae autotransporter proteins or immunogenic fragments thereof and one or more immunostimulants.

25 16. The composition according to claim 15 wherein the Chlamydia pneumoniae autotransporter protein or the immunogenic fragment thereof comprises:

(a) an amino acid sequence selected from the group consisting of SEQ ID NO: 54, SEQ ID NO: 6, SEQ ID NO: 55, SEQ ID NO: 56, SEQ ID NO: 78, SEQ ID NO: 86 and SEQ ID NO: 79;

30 (b) an amino acid sequence having at least 50% sequence identity to an amino acid sequence of (a); or

(c) an amino acid sequence comprising one or more fragments of at least 1, 2, 3, 4, 5, 6 or 7 amino acids from an amino acid sequences of (a) or combinations thereof.

35 17. The composition according to claim 15 or 16 wherein the protein or immunogenic fragment thereof is defined according to any one of claims 1-3.

40 18. A composition for eliciting an immune response in a subject comprising two or more Chlamydia pneumoniae autotransporter proteins or immunogenic fragments thereof.

19. The composition according to claim 18 wherein the Chlamydia pneumoniae autotransporter protein or the immunogenic fragment thereof comprises:

45 (a) an amino acid sequence selected from the group consisting of SEQ ID NO: 54, SEQ ID NO: 6, SEQ ID NO: 55, SEQ ID NO: 56, SEQ ID NO: 78, SEQ ID NO: 86 and SEQ ID NO: 79;

(b) an amino acid sequence having at least 50% sequence identity to an amino acid sequence of (a); or

(c) an amino acid sequence comprising one or more fragments of at least 1, 2, 3, 4, 5, 6 or 7 amino acids from an amino acid sequences of (a) or combinations thereof.

5 20. The composition according to claim 18 or 19 wherein the composition further comprises one or more immunostimulants.

10 21. A method of making a composition according to any one of claims 15 or 16 wherein the method comprises combining one or more Chlamydia pneumoniae autotransporter proteins or immunogenic fragments thereof with one or more immunostimulants.

15 22. A method of making a composition according to claim 18 or 19 wherein the method comprises combining two or more Chlamydia pneumoniae autotransporter proteins or immunogenic fragments thereof.

20 23. The method according to claim 22 wherein the method comprises adding one or more immunostimulants to the Chlamydia pneumoniae autotransporter proteins or immunogenic fragments thereof.

25 24. A Chlamydia pneumoniae autotransporter protein selected from the group consisting of Cpn0794, Cpn0795, Cpn0796, Cpn0797, CPn0798 and Cpn0799 or an immunogenic fragment thereof wherein the autotransporter protein an amino acid motif comprising IVG, A, LGG and S.

 25. The autotransporter protein according to claim 24 wherein the repeat amino acid motif comprises IVG, A, LGG and S.

30 26. A polypeptide for use as an autotransporter antigen comprising an amino acid sequence corresponding to SEQ ID NO: 86, an amino acid sequence having at least 50% sequence identity to SEQ ID NO: 86, or an amino acid sequence comprising one or more fragments of at least 7 consecutive amino acids of SEQ ID NO: 86.

35 27. The polypeptide of claim 26 where use is as an antigen for raising a Chlamydia pneumoniae specific immune response

40 28. The polypeptide of claim 2 wherein the use is for raising a systemic immune response in an individual infected with Chlamydia pneumoniae.

 29. The use of a polypeptide according to any one of claims 26-28 in the preparation of a medicament for the prevention or treatment of a Chlamydia pneumoniae infection in an individual.

45 30. A method of raising an immune response in an individual, the method comprising administering to the individual a polypeptide comprising an amino acid sequence corresponding to SEQ ID NO: 86, an amino acid sequence having at least 50% sequence identity to SEQ ID NO: 86, or an amino acid sequence comprising one or more fragments of at least 7 consecutive amino acids of SEQ ID NO: 86.

50

31. A method of diagnosing an immune response in an individual, the method comprising:

(a) contacting a biological sample obtained from an individual with a binding agent that binds to a polypeptide defined in any one of claims 26-28;

5 (b) detecting in the sample the amount of the polypeptide that binds to the binding agent; and

(c) comparing the amount of polypeptide to a predetermined cut-off value and thereby determining the presence of an immune response in the individual.

Figure 1A

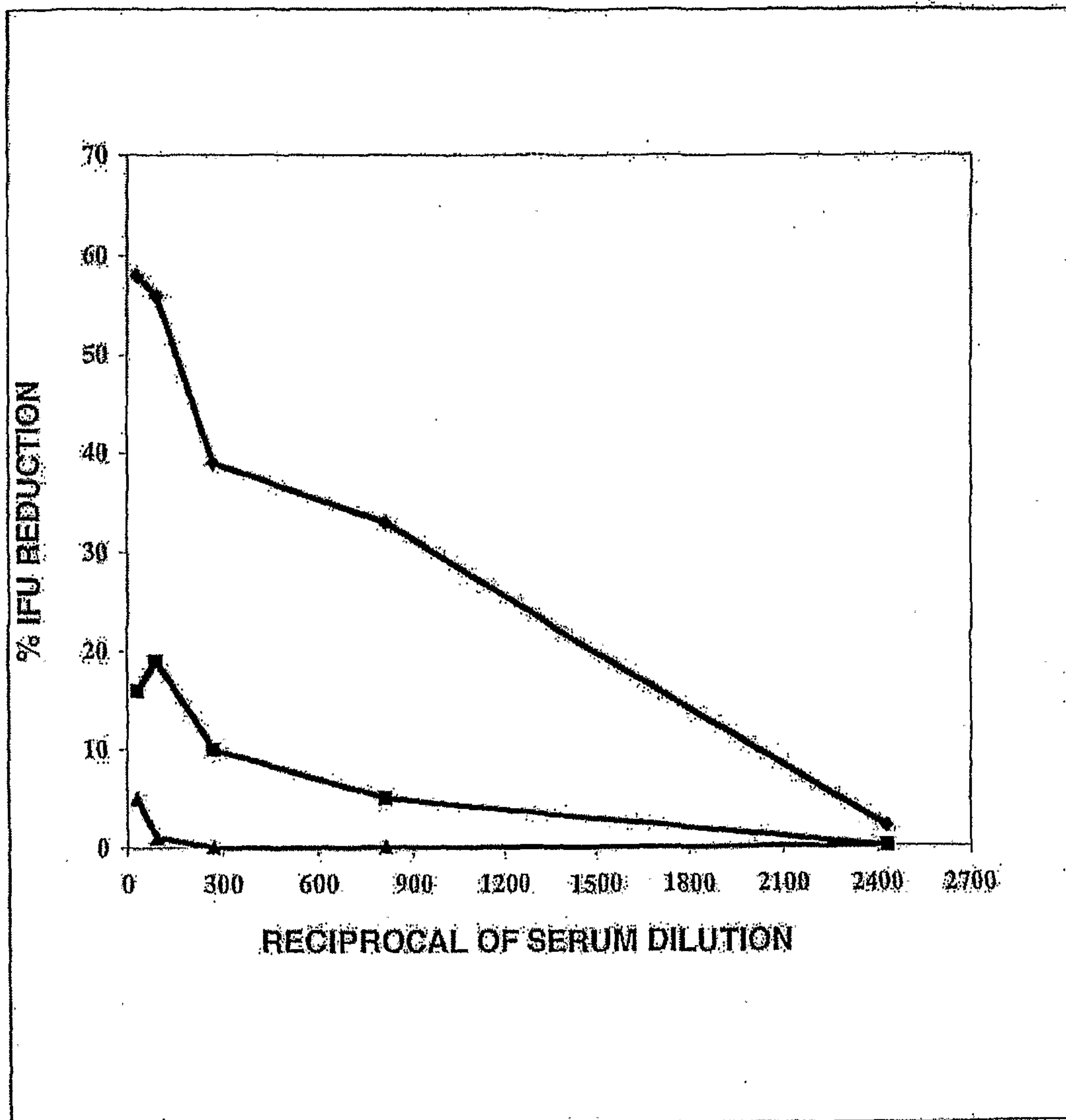
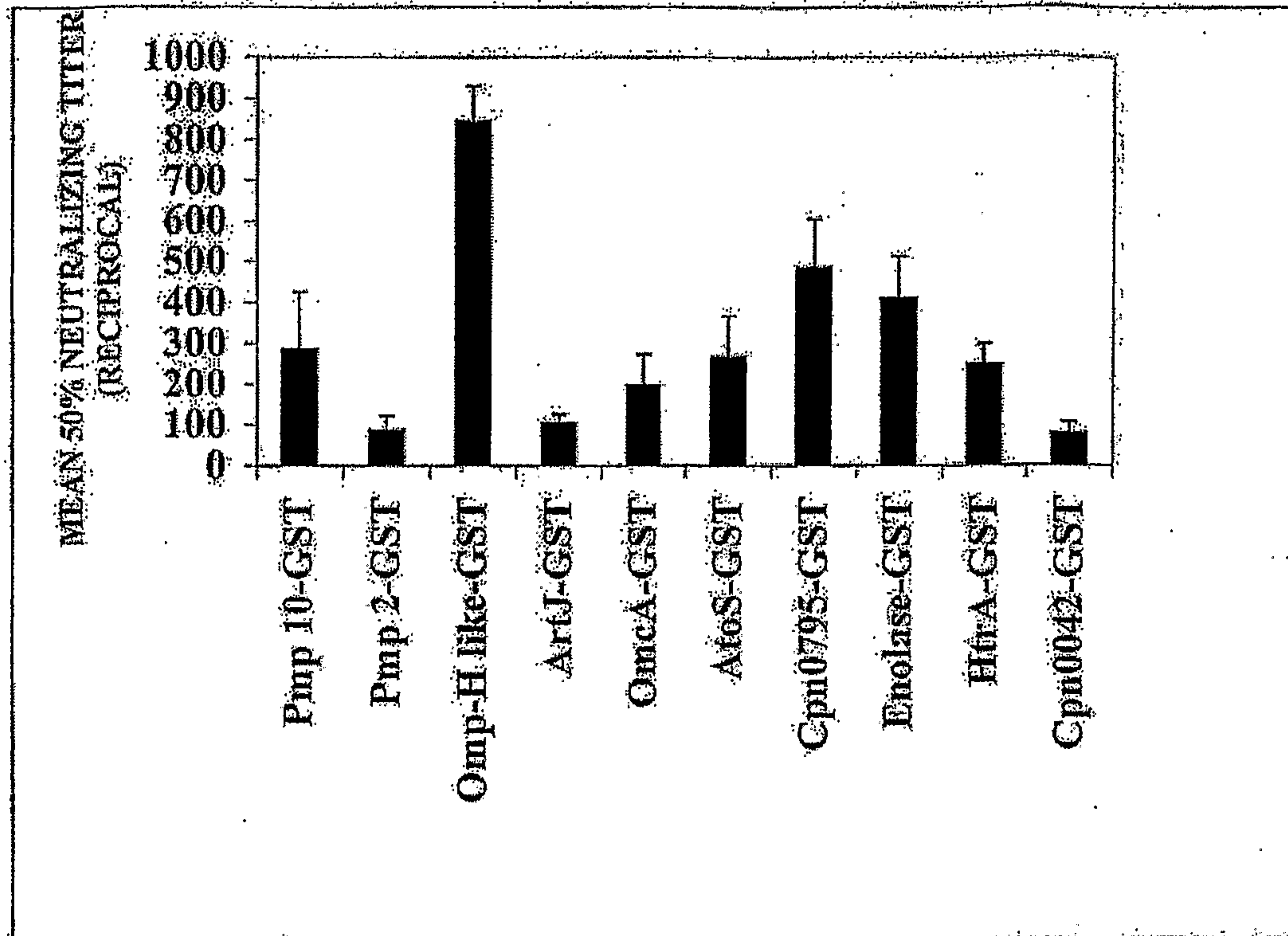


Figure 1B



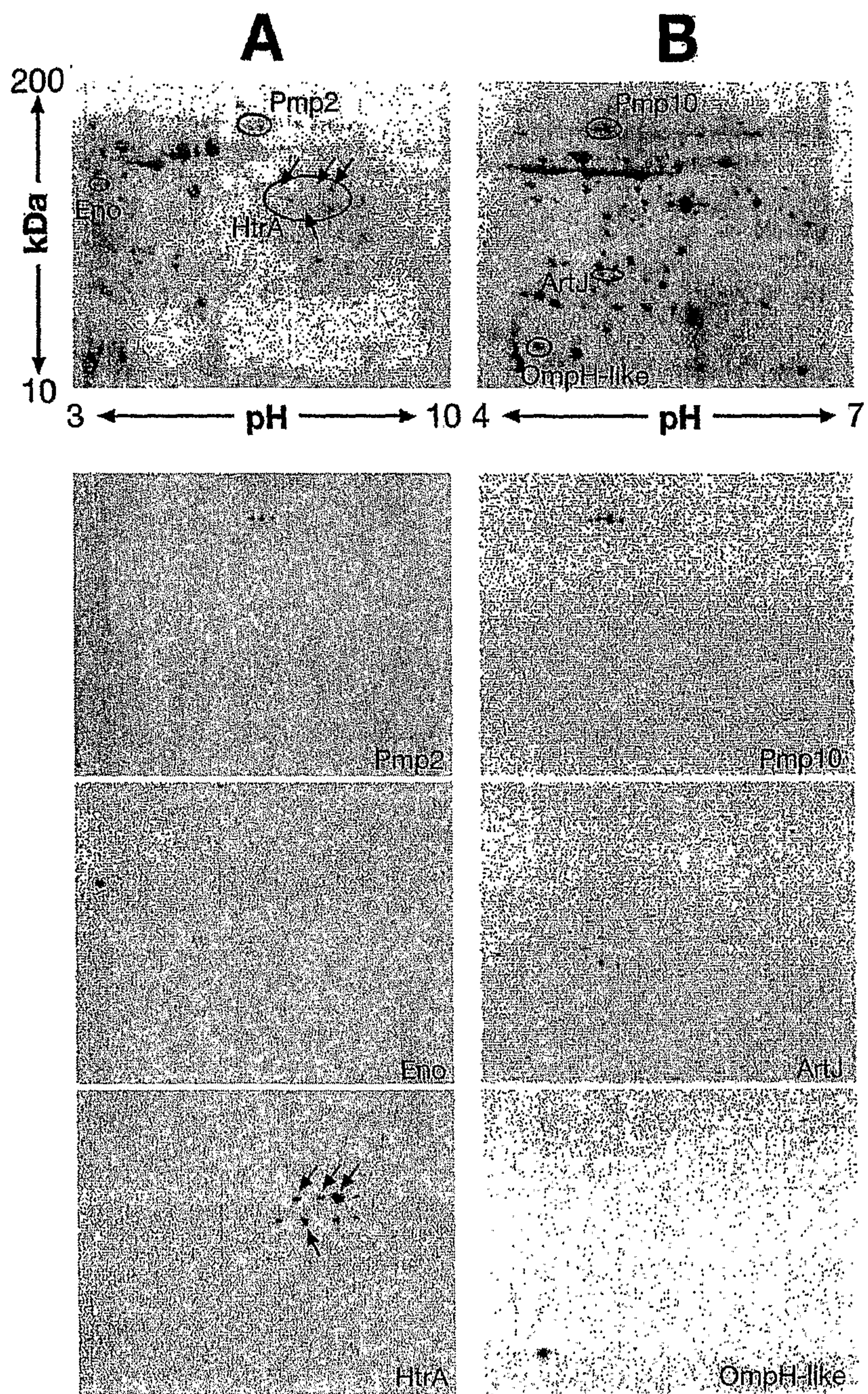


Figure 2

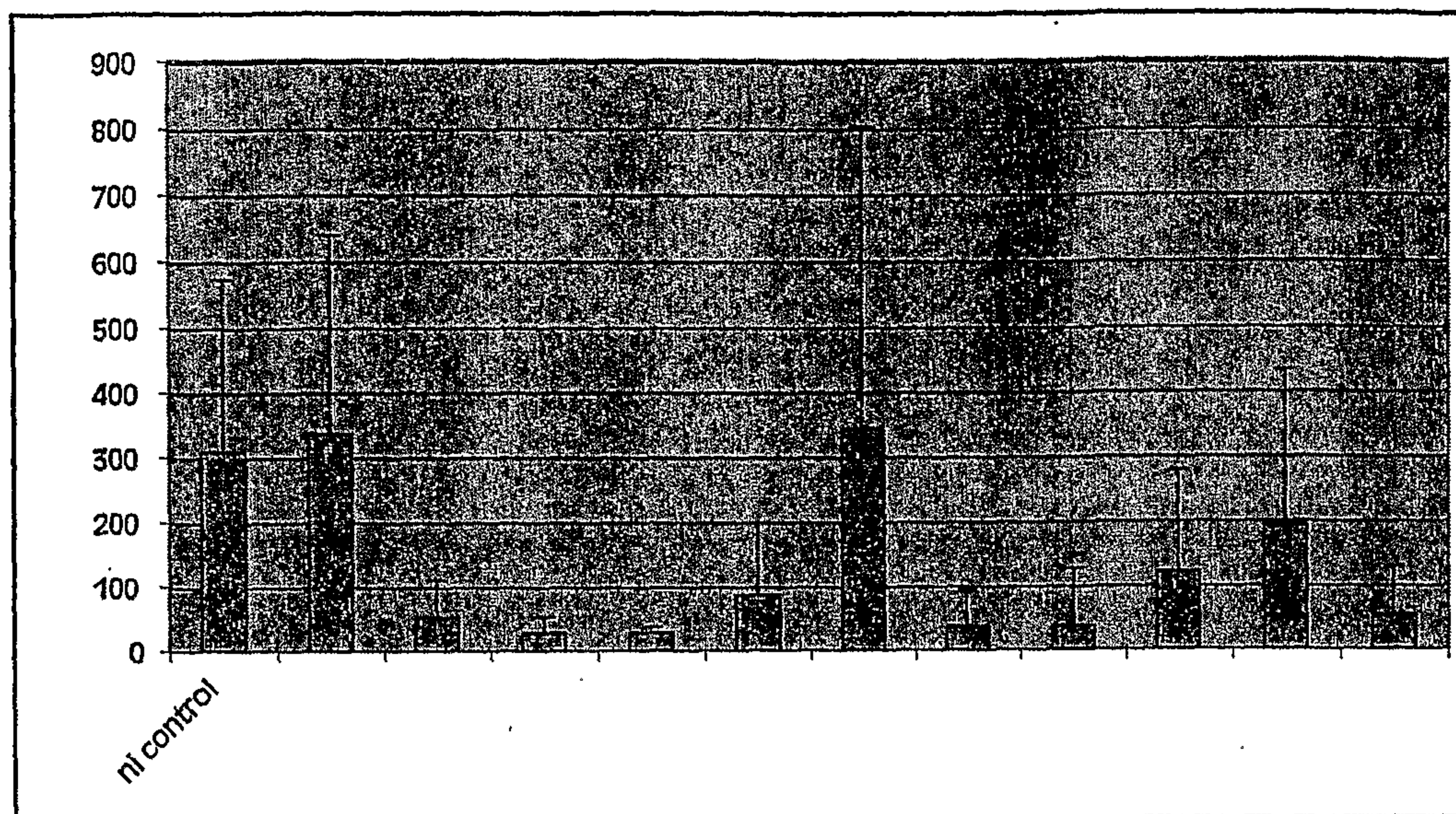


Figure 3. Mean numbers of *C.pneumoniae* IFU recovered from equivalent spleen samples from immunized and mock-immunized hamsters following a systemic challenge. Standard deviation values are shown above the bars. Antigens which induced significant protection are highlighted with an asterisk above the corresponding bar. All antigens were delivered in Freund's adjuvant. n.i. = non immunized controls

Figure 4

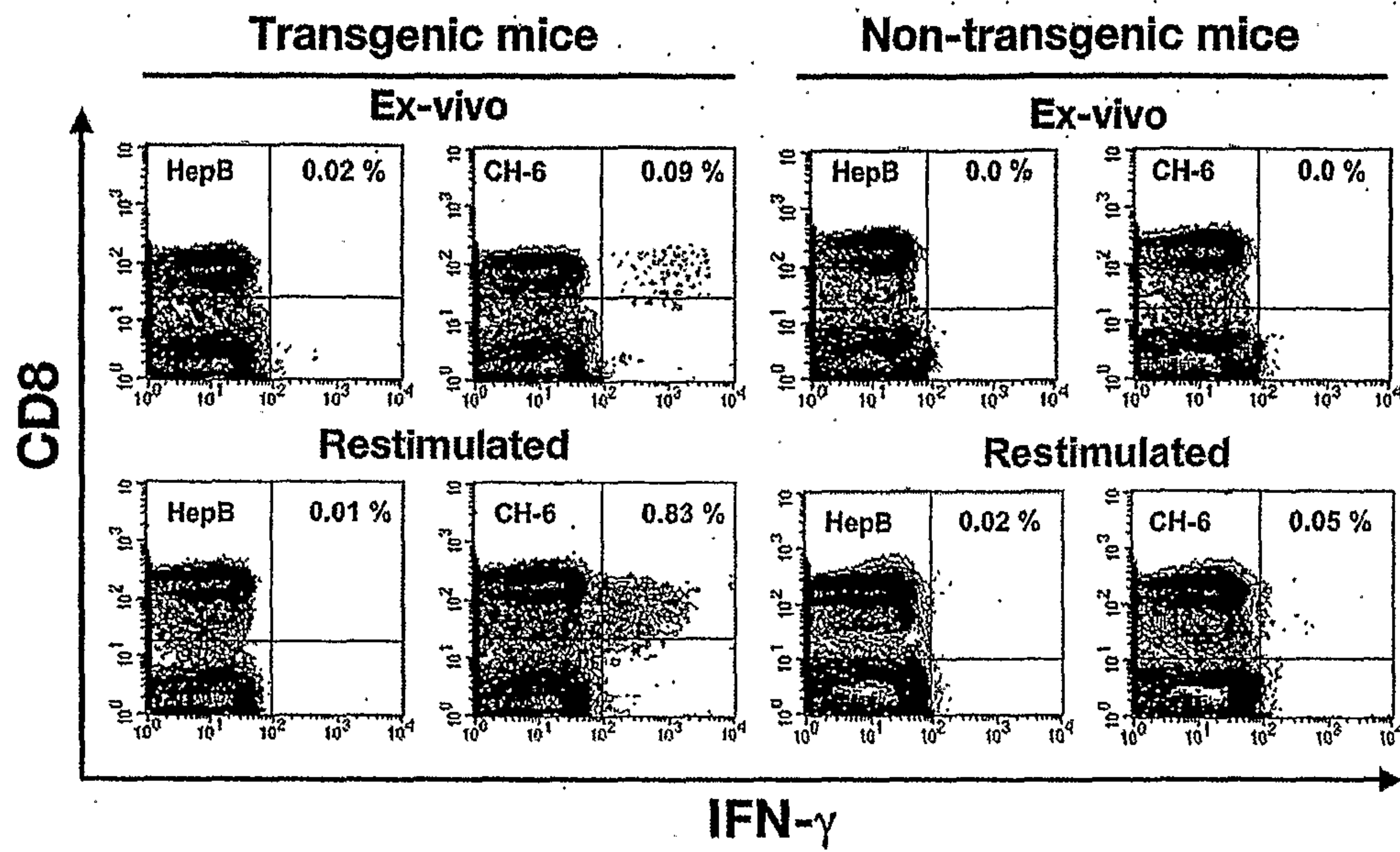
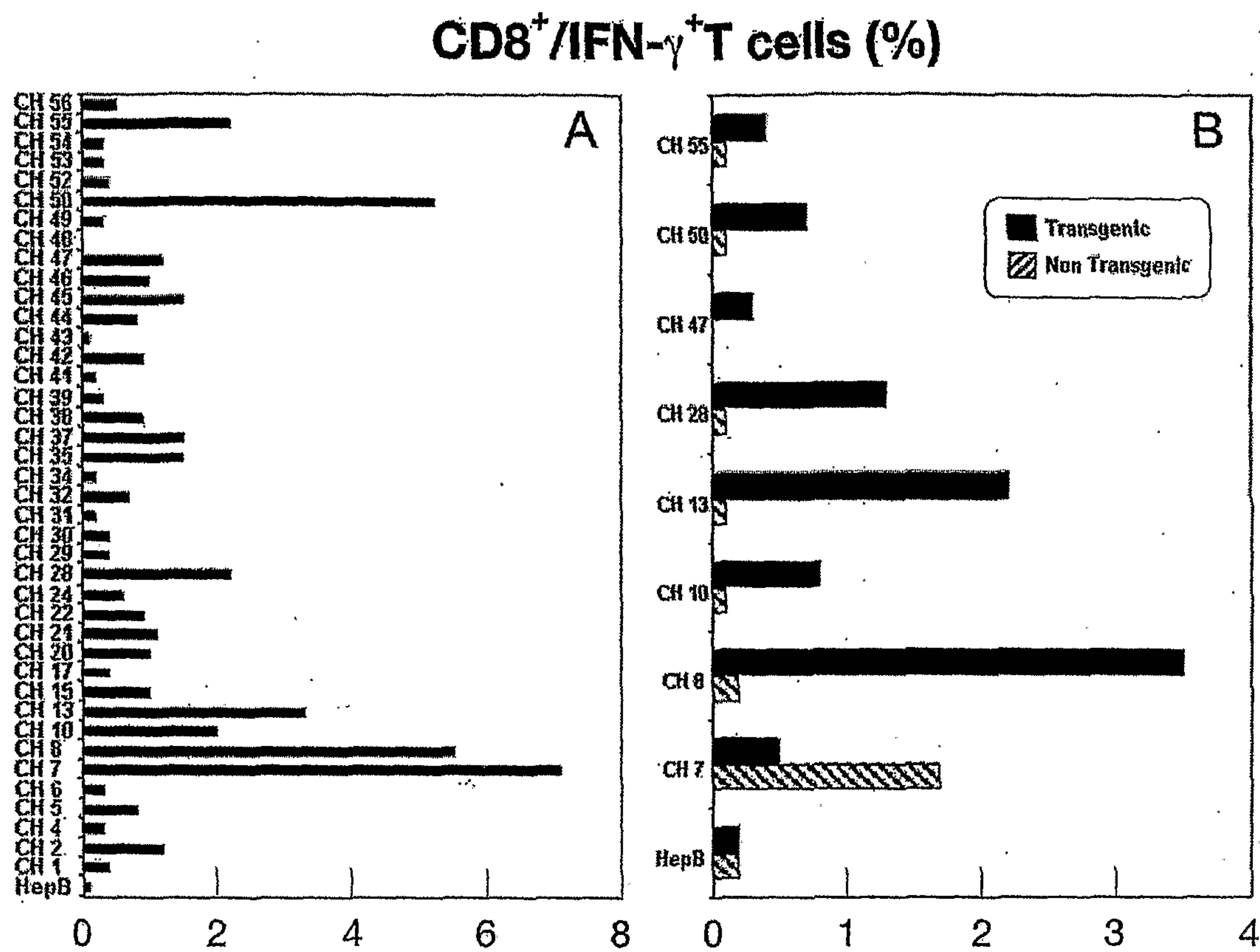
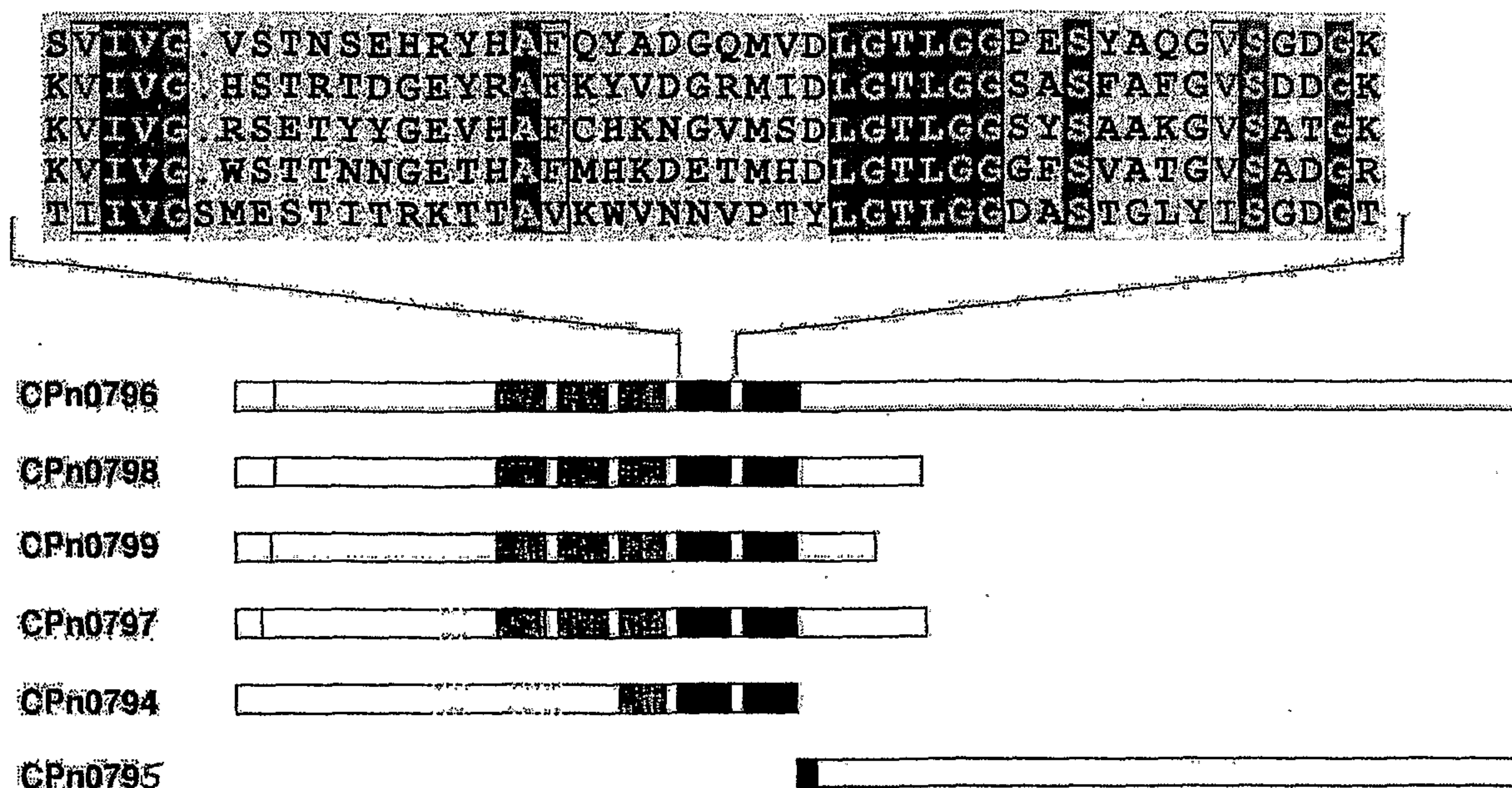


Figure 5





This Alignment shows a new family of proteins expected to constitute a system of antigens probably delivered on the Cpn surface or secreted by a type V (autotransporter) secretion mechanism

Fig. 6

Cpn0796

```

1  MKFMKVLTPW IYRKDLWVTA FLLTAIPGSF AHTLVDLIA   TGV
51  SGDGKIVIGM KVPDDPFAIT VGFOYIDGHL QPLEAVRPQC SVYPNGITPD
101 GTVIVGTNYA IGMGSVAVKW VNGKVSELP LPDTLDSVAS AVSADGRVIG
151 GNRNINLGAS VAVKWEDDVI TQLPSLPDAM NACVNGISSD GSIIVGTMVD
201 VSWRNTAVQW IGDQLSVIGT LGTTTSVASA ISTDGTIVIG GSENADSQTH
251 AYAYKNGVMS DIGTLGGFYS LAHAVSSDGS VIVGVSTNSE HRYHAFQYAD
301 GQMVDLGLTG GPESYAQCVS GDGKVIVGRA QVPSGDWHAF LCPFQAPSPA
351 PVHGGSTVVT' SQNPRGMVDI NATYSSLKNS QQQLQRLLIQ HSAKVESVSS
401 GAPSFTSVKG AISKQSPAVQ NDVOKGTFLS YRSQVHGNVQ NQOLLTGAFM
451 DWKLASAPKC GFKVALHYGS QDALVERAAL PYTEQGLGSS VLSGFGGQVQ
501 GRYDFNLGET VVLQPFMGIO VLHLSREGYS EKNVREFVSY DSVAYSAATS
551 FMGAHVFEASL SPKMSTAATL GVERDLNSHI DEFKGSVSAM GNFVLENSTV
601 SVLRPFASIA MYYDVROOQL VTLSVVMNOQ PLTGTLSLVS QSSYNLSF

```

Leader peptide	1-31
Or, alternatively:	
Leader peptide 2	1-47
b-propeller domain	32-342
Or, alternatively:	
b-propeller domain2	48-342
linker region	343-364
helix	365-385
beta barrel	386-648

FIG. 8

Alignment of the C-terminal (beta barrell) domains of the proteins encoded by the
C.pneumoniae genes Cpn0795 (7016) and Cpn0796 (7107)

```

7106      -----TSSTAKTVSPDGKVLGERSQIADGSWHAFMCHTDFSSNNVLFDLDN
7107      YADGO      PESYAQGVSGDGKVLVCGRAQVPSGDWHAFLCPFQAPSPAPVHGGST
              * *****..* *: ** *****:***:.....*:***: * :..* :..* ..

7106      TYKTLRENGRQLNSIFNLQNMMLQRASDHEFTEFGRSNIALGAGLYVNALQNLPSNLAAQ
7107      VVTSQ--NPRGMVDINATYSSLKNSQQQLQRLLIQHSKVESVSSGAPSFTSVKGAISKQ
              . .: . * * : . * . : : . : : : : * . . . . : : . : : *

7106      YFGIAYKIRPKYRLGVFLDHNFSHVPPNNFNVSHNRLWMGAFIGWQSDALGSSVKVSFG
7107      SP----AVQNDVQKGTFLSYRSQVHG----NVQNQQLLTGAFMDWKLASAPKCGFKVALH
              . : : : . : * . . . . * . . . * . : : * * : : * . . . * : :

7106      YGKQKATITREQLENTEAGSGESHFEGVAAQIEGRYGKSLGGHVVRVQPFLGLQFVHITRK
7107      YGSQDALVERAALPYTEQGLGSSVLSGFGGQVQGRYDFNLGETTVVLQPFMGIQVLHLSRE
              **.*.* : * * ** * *.* :.*...*:***. ** * :***:***:***:***:

7106      EYTENAVQFPVHYDPIDYSTGVVYLIGISHIALVDSLHVGTMRMGMEQNFAAHTDRFSGSI
7107      GYSEKNVRFPVSYDSVAYSAAATSFMGAHVFASLSPKMSTAATLGVERDLNSHIDEFKGSV
              *:*: *:*** **.: **:.. ::* . : * .: .: :*:***: :* *.*.*:

7106      ASIGNFVFEKLDVTHTRAFAEMRVNYELPYLQSLNLILRVNQPLQGVMGFSSDLRYALG
7107      SAMGNFVLENSTVSVLRPFASLAMYDVRQQQLVTLVVMNQPLTGTLVSLVSQSSYNLS
              :*:***:***: *: *.*.*: : *: : * :.* : :*** **.*.: :* * *.

7106      F
7107      F
              *

```

FIG. 9