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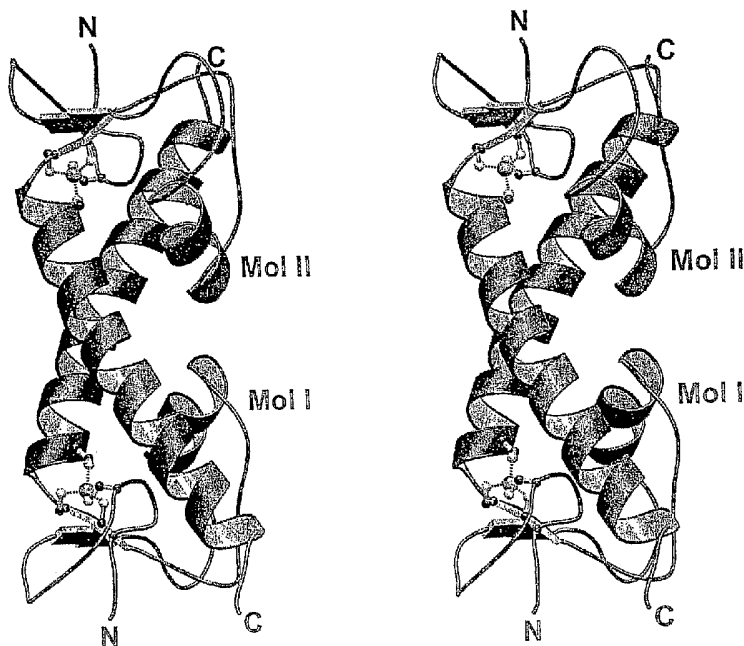
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(54) Title: METHOD FOR OBTAINING AGENTS AGAINST INSECTS BY USING THE STRUCTURAL COORDINATES OF
ZAD_{Grau}



(57) Abstract: The present invention discloses a method for obtaining agents interacting with a ZAD_{Grau} molecule by using the structural coordinates obtained from crystals of ZAD_{Grau}. These agents can serve as insecticides.



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METHOD FOR OBTAINING AGENTS AGAINST INSECTS BY USING THE
STRUCTURAL COORDINATES OF ZAD_{Grau}

The present invention relates to methods for obtaining agents
5 against insects by using the structural coordinates of a zinc
finger-associated domain (ZAD_{Grau}) of the transcription factor
Grauzone (Grau). This invention is also related to crystals
comprising ZAD_{Grau} molecules and to methods of obtaining 3-
dimensional models of ZAD_{Grau}-like molecules of unknown struc-
10 ture by using the structural coordinates of ZAD_{Grau}.

Resistance to antiparasitic compounds among insects is a
wide-spread problem. A large group of insecticides, the orga-
nophosphates and the carbamates inactivate the insecticidal
15 acetylcholinesterase, thereby terminating synaptic transmis-
sion through hydrolysis of the neurotransmitter acetylcho-
line, leading to paralysis and death of the insects. Insensi-
tivity of the insects towards the organophosphates and carba-
mates was shown to be conferred by a single amino-acid sub-
20 stitution in the acetylcholinesterase (Weill et al., 2003).
Therefore there is a continuous need to reveal new targets for
the development of new insecticides. These new targets are
preferably proteins, performing vital functions for the in-
sects.

25 Recent mutant analysis showed, that the proteins encoded by
three genes of the fruitfly *Drosophila melanogaster*, grauzone
(grau), Zeste-white-5 (Zw5) and Serendipity- δ (Sry- δ) carry
vital functions (Chen et al., 2000; Crozatier et al., 1992;
30 Gasner et al., 1999). These proteins all belong to a large
group of so-called C₂H₂ zinc finger proteins, which contain
zinc ions, which are usually coordinated by two cysteine and
two histidine residues. Zinc-finger proteins are defined by
the thirty amino-acid C₂H₂ motive (Miller et al., 1985; Pav-
35 latich and Pabo, 1991). The coordination of a zinc ion by
four amino acids plays a crucial role for the functional fold
of these proteins and is part of a distinct protein domain,

the so-called zinc finger domain. A large number of zinc finger domains function as DNA-binding domains in transcription factors, (for example Pavletich and Pabo, 1991; Rosenberg et al., 1986; Wolfe et al., 2000). Zinc finger domains can also
5 mediate interactions of proteins containing the zinc finger domains with RNA, other proteins and small molecules (Krishna et al., 2003; Makay and Crossley, 1998; McCarty et al., 2003). The zinc finger proteins often contain additional protein domains and primary sequence motifs such as the mammalian Krüppel associated box (KRAB) or the insect zinc finger-associated domain (ZAD) (Lespinet and Aravind, 2002), which
10 can be characterised by a conserved N-terminal primary sequence motif, containing two invariant pairs of cysteines (see for example the primary sequence alignment in Figure 1A). The KRAB domain acts as a transcriptional repressor module (Margolin et al., 1994; Peng et al., 2000), whereas the
15 function of the zinc finger-associated domain is unknown.

Computer-assisted primary sequence alignments revealed that
20 more than ninety proteins, containing zinc finger-associated domains are present in *Drosophila melanogaster*, among them the above-mentioned Grauzone, Zeste-white-5 and Serendipity- δ (Chung et al., 2002). Grauzone was shown to be necessary for the proper transcriptional activation of the gene *cortex*,
25 coding for a Cdc20-like APC subunit (Chu et al., 2001). Maternal loss of function mutations of *grau* cause an irregular growth arrest in meiosis II, whereas homozygous *grau* mutant embryos, which have received maternal *grau* activity, develop into normal adults. The mutant phenotype of *grau* suggests
30 that the transcriptional factor Grauzone is exclusively required during oogenesis and *cortex* is its only target gene (Chen et al., 2000).

The mutational analysis revealing the vital function of Serendipity- δ and Zeste-white-5 for *Drosophila melanogaster*,
35 showed that single amino-acid mutations within the respective ZADs of both proteins were responsible for the lethality, im-

plying that conserved amino-acids within the ZAD are essential for the function of these proteins (single amino acid mutations C7T in Serendipity- δ , Crozatier et al., 1992; and R4G in Zeste-white-5, Gazner et al., 1999). These results
5 strongly emphasise the crucial function of ZADs within ZAD-containing zinc finger proteins. ZADs do not exist e.g. in vertebrates and are lineage specific to insects. Therefore ZAD-containing zinc finger proteins might represent novel targets for insecticides.

10 The utilisation of the ZAD-containing proteins as a target for insecticides is hampered by the fact that no details about the function or the 3-dimensional structure of the ZAD are known. In particular, ZADs were identified as a primary
15 sequence motif by computer-assisted sequence alignments, but up to now it is unclear, if they encompass protein domains with an independent fold, whose 3-dimensional structure can be analysed. Similarly the biological function of ZADs is unknown.

20 The present invention meets these needs by providing a method for identifying an agent which interacts with a ZAD_{Grau} molecule using the structural coordinates of ZAD_{Grau} according to the base claim 1. Favourable embodiments of the invention are
25 subjects of further claims.

For the first time the invention provides a method for obtaining an agent which interacts with a ZAD_{Grau} molecule or a molecule with a similar fold to ZAD_{Grau}, comprising the following steps:
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a) Using the structural coordinates of:

i) at least a portion of the structural coordinates of ZAD_{Grau} shown in Figure 6 or

35 ii) at least a portion of a structural similar molecule of ZAD_{Grau}, having a root mean square deviation from the backbone of ZAD_{Grau} amino-acids of not more than 1.5 Å,

to generate a 3-dimensional model of the portion of ZAD_{Grau},

b) employing the 3-dimensional model and at least one molecular modeling technique to select or design an interacting agent.

The inventors were, for the first time, able to crystallise isolated ZADs from the protein Grauzone of *Drosophila melanogaster*. A subsequent successful x-ray crystallographic analysis of the protein crystals revealed the 3-dimensional structure of ZAD_{Grau} and showed that ZAD molecules are protein domains with an independent fold, comprising a globular portion structured around a zinc ion at the N-terminus of the protein and a long α -helix at the C-terminus (for details see for example figures 3A to 3C, 4A, 4B and example 3).

In ZAD_{Grau} crystals two ZADs are associated through a 2-fold axis in a head-to-tail fashion, interacting with each other. Major parts of the contact interface are build up by the long α -helix at the C-terminus of the protein (for details see for example figures 4A, 4B and example 3). Therefore the crystal structure unexpectedly suggests that ZAD modules in ZAD-containing proteins function as protein interacting modules, mediating contacts between different proteins e.g. two Grauzone proteins. The dimerization of two ZAD molecules, which was found in ZAD crystals, was also independently confirmed by other biochemical experiments like glutaraldehyde cross-linking experiments and gel filtration experiments in conjunction with multi-angle-laser-light scattering (for details see for example figures 5A, 5B and example 4). The crystallographic data and additional biochemical evidence therefore indicate that ZAD_{Grau} is a protein-protein interaction module with the capability to form homodimers and suggests that ZAD provides a taxon specific means for the assembly of ZAD-containing zinc finger transcription factor complexes.

Consequently the crystal structure of ZADs provides the possibility to obtain agents interacting with ZAD and interfering with the biological function of ZAD, namely the ability to provide protein-protein interactions. Since mutational
5 analysis suggests a vital role of ZADs for insects like *Drosophila melanogaster*, this method can identify potential insecticides.

In the first step a) of the method of the invention at least
10 a portion of the structural coordinates, which were determined by the inventors and are shown in Figure 6, is used to generate a 3-dimensional model of a portion of ZAD_{Grau}. This model can function as a potential binding site for the interacting agent. The knowledge about the exact 3-dimensional geometry of that portion of ZAD is a crucial prerequisite for
15 the selection or the design of agents, interacting with this binding site or interfering with the folding of ZAD. Agents interacting with at least portions of ZAD_{Grau} have to fulfil two requirements. First the agent has to be able to physically and structurally bind to the portion of ZAD_{Grau}. Binding
20 between the agent and the portion of ZAD_{Grau} can occur via salt bridges, hydrogen bonding interactions, hydrophobic interactions or van der Waals interactions. Secondly, the agent must be capable of assuming a conformation that enables association with the portion of ZAD_{Grau} and preferably shows no
25 sterical repulsion.

In step b) of the method of the invention, the 3-dimensional model generated in step a) and at least one molecular modeling
30 technique is used to select or design an interacting agent.

In general molecular modeling involves the computer-assisted calculation, representation and processing of realistic 3-
35 dimensional models of molecules and their physical-chemical properties. Several approaches are possible for the use of the crystal structure of ZAD_{Grau} for identifying interacting

agents by molecular modeling techniques. It is possible, for example, to use empirical forcefield methods to describe interactions between atoms in the form of analytical functions an empirical parameters. Using this method it is also possible to calculate binding and nonbinding interactions between atoms. The Insight II/Discover program package (Accelrys; San Diego, CA) supports different forcefields, for example CVFF (consistent valence forcefield) (Dauber-Osguthorpe et al., 1988), which might help to position potential interacting agents or small chemical entities into binding sites or binding pockets of 3-dimensional portions of the ZAD_{Grau} molecule.

The method of positioning or docking these molecules to binding sites of ZAD_{Grau} may also be accomplished by using software such as Quanta or SYBYL. After the docking operation an energy-minimization using molecular dynamics with standard molecular mechanic forcefields might be useful.

Step b) can further comprise analysing the results of the docking operation to evaluate the interaction between the agent and the portion of ZAD or the portion of the structural similar molecule. This variant of the method of the invention enables the determination of the theoretical binding energy between the agent selected or designed by a molecular modeling technique and the portion of ZAD.

Therefore, computer modeling techniques enable an expert to evaluate the potential binding of agents or small chemical entities to ZAD_{Grau} prior to the actual synthesis of these agents. If the results of the computer modeling techniques suggest insufficient interaction between an agent and ZAD_{Grau}, synthesis and testing of this agent in an assay is omitted.

The positioning of small chemical entities or agents to portions of ZAD_{Grau}, which is also called docking, might also be achieved by other specialized computer programs like:

AUTODOCK (Goodsell, D.S. and A.J. Olsen "Automated Docking of Substrates to Proteins by Simulated Annealing" Proteins: Structure Function and Genetics, 8, pages 195 to 202, 1990).

5

DOCK (Kuntz, I.D. et al., "A Geometric Approach to Macromolecule-Ligand Interactions", Journal of Molecular Biology, 161, pages 269 to 288, 1982).

10 GRID (Goodford, P.J. "A Computational Procedure for Determining Energetically Favorable Binding Sites on Biologically Important Macromolecules", Journal of Medicinal Chemistry, 28, pages 849 to 857, 1985).

15 FLEXX (M. Rary, Kramer B., Lengauer T., Klebe G. "A Fast Flexible Docking Method Using an Incremental Construction Algorithm", Journal of Molecular Biology, 261, pages 470 to 489, 1996). This program uses a special algorithm for docking small chemical compounds into binding sites, using an incremental construction algorithm. Conformational flexibility of
20 the ligands is incorporated by generating many low-energy confirmations of the chemical compounds.

It is also possible to use other molecular modeling techniques. E.g. individual chemical entities which were found to
25 interact with portions of ZAD_{Grau} can also be assembled into a single interacting agent by using for example the following software packages:

30 CAVEAT (Bartlett, P.A. et al. "CAVEAT: A Program to Facilitate the Structure-derived Design of Biologically Active Molecules", in "Molecular Recognition in Chemical and Biological Problems", special publication of the Royal Chemical Society, 78, pages 82 to 196, 1989).

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This program accesses data bases of different compounds which can act as "linkers" in order to connect different chemical

entities which were found to bind to structural portions of ZAD_{Grau}.

5 In accordance with this invention it is also possible to define novel agents in their entirety or by assembly of smaller portions of agents which were selected to bind to ZAD_{Grau} by using for example the software package:

10 LUDI (Bohm, H.J. "The Computer Program LUDI: A New Method for the De Novo Design of Enzyme Inhibitors", J. Comput. Aided Mol. Des., 6, pp 61 to 78, 1992).

This program uses a big library of different linkers to connect different interaction sites.

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According to this invention, molecules which are structurally similar to ZAD_{Grau} have a primary sequence identity of at least 30% to ZAD_{Grau} and form the N-terminal domain of an insect C₂H₂ zinc finger protein. Alternatively structurally similar molecules can be identified by using profile hidden Markow models e.g. with the HMMER software package and the Wise software package as described below.

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25 According to the crystal structure of ZAD_{Grau}, conserved amino acids in ZAD modules can be grouped into three classes: 1. those supporting zinc coordination (C4, C7, C53, C56), 2. those supporting the overall fold, and 3. those supporting dimerization. Class 1 is not an interesting target for ligand design since the coordination of a zinc ion by four cysteines is a common motif in many proteins and not specific to the zinc finger associated domains. Among the second class of residues the sequence H37-V42 is completely conserved, e.g. between *Drosophila melanogaster* and *Anopheles gambiae* ZAD_{Grau} with the exception of F38 (*Drosophila*), which is still conserved as a hydrophobic residue. These residues form a continuous cluster in the globular portion of ZAD_{Grau}, which is well accessible to the solvent and thus suitable for the de-

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sign of a ligand interacting with this region. In addition, the surface patch outlined by these residues is continuous with that formed by a second cluster of conserved residues, F63-V68 of *Drosophila* ZAD_{Grau}. Several residues in this latter
5 cluster belong to the class 3 of conserved residues, i.e. they mediate important interactions between the molecules forming the ZAD dimers. In particular F63 and F66, which are identical in ZAD_{Grau} from *Drosophila* and *Anopheles*, play important roles in inter-molecular stacking interactions.

10 Again, the surface patch outlined by the latter group of residues is well accessible to the solvent in the ZAD_{Grau} monomer structure. Therefore, the inventors propose to base the design of ligands on the groups of residues H37-V42 and F63-V68 (*Drosophila* numbering). A ligand spanning the two re-
15 gions or parts of both regions may concomitantly interfere with folding and dimerization of ZADs. Additional residues can be identified in the crystal structure of ZAD_{Grau}, which possibly can be assigned to class 2 as well as class 3, because these residues might be important for the folding of
20 the protein and also support dimerization. These residues are R5, L6, E47, Q59, I70, Q74 and Y77.

The primary sequence identity can be determined using algorithms like Clustal W 1.81 (Thompson et al., 1994). This high
25 percentage of homology suggests an overall 3-dimensional fold of these proteins, which is very similar to ZAD_{Grau}. Experts in the field of protein X-ray crystallography generally accept, that a different set of structural coordinates, e.g. obtained from a different crystal form of the molecule, or
30 crystals of a different ZAD_{Grau} construct (e.g. in combination with other parts of the Grauzone protein), or of a different ZAD alone or in combination with other parts of a protein molecule, still exhibits the same fold as that of ZAD_{Grau} described herein, if the backbone coordinates of the residues
35 comprising the regular secondary structure elements of ZAD-Grau (α -helices and β -strands) do not deviate by more than 1.5 Å. The technical term "root mean square deviation" de-

scribes the square root of the arithmetic mean of the squares of the deviation from the mean. It describes the medium variation in the coordinates of the backbone atoms of a protein. Superposition of different sets of structural coordinates can e.g. be performed by using programs like DALI (Holm, L. , Sander, C. Protein structure comparison by alignment of distance matrices. J. Mol. Biol. 233: 123-138, 1993).

In a preferred embodiment of the method of the invention the structural coordinates of at least two of the following ZAD_{Grau} amino-acids shown in Figure 6 are used in step a): R5, L6, E47, Q59, I70, Q74, Y77 and the groups H37-V42 and F63-V68 (sequence numbering corresponding to that of ZAD_{Grau}). It is also possible to use a portion of a structurally similar molecule of ZAD_{Grau}, which has a root mean square deviation from the backbone atoms of the above-mentioned amino-acids of not more than 1.5 Å. The above mentioned amino-acids preferably define the protein-protein interaction site of ZAD_{Grau} or are important for the fold of the ZAD molecule (for details see for example figure 4B or the example 3). Preferably the structural coordinates of at least one of the following amino acids are used in step a): arginine 5, phenylalanine 63, phenylalanine 66, isoleucine 70, because these residues are well conserved among the different ZAD molecules (see for example figures 1A and 1B). Agents interacting with some of these amino acids preferably can interfere with the protein-protein interaction properties of ZAD_{Grau} and can therefore present highly valuable insecticides.

In another favorable embodiment of the method of the invention the method further comprises the following steps:

c) Synthesizing said interacting agent and

d) bringing said agent in contact with the binding site of the molecule or structural similar molecule of ZAD to detect the interaction.

These additional steps can provide a valuable verification of the binding of agents, which were selected by computer-based modeling techniques. Step d) can preferably comprise orally
5 administering the interacting agent to flies, due to the ease of handling preferably *Drosophila melanogaster* and detecting the rate of dying or infertile flies in an essay. It is also possible to detect the interaction of the interacting agent with ZAD_{Grau} by e.g. gel filtration chromatography, UV/VIS
10 spectroscopy, fluorescence spectroscopy, ESR spectroscopy, NMR, isothermal titration calorimetric or "pull down" assays, wherein e.g. a ZAD_{Grau}-glutathione-S-transferase fusion protein is brought in contact with the interacting agent, then precipitated with glutathione-beads and the interacting agent
15 afterwards is detected in the pellet, thereby directly monitoring the interaction between ZAD_{Grau} and the interacting agent.

In a further advantageous embodiment of the method of the invention step b) comprises screening a library of individual
20 chemical entities for the agent by performing a computer-assisted docking operation between the individual chemical entities and the binding site. This can preferably be done by using computer-based libraries of small molecules and by
25 docking, or fitting these molecules into the binding sites of ZAD_{Grau}.

In another embodiment of the invention, step b) comprises the design of the interacting agent by assembling individual
30 chemical entities, which were selected by computer-based modeling techniques to bind to the binding sites of ZAD_{Grau} into a single interacting agent and docking this single interacting agent to the binding site using computational means. This method can be used for "De Novo" synthesis of new interacting
35 agents.

Step b) can further comprise performing an energy minimisation or molecular dynamics operation using computational means to improve the interaction between the agent and the portion of the ZAD molecule or the portion of the molecule with the similar fold. This can be done for example by using the above-mentioned molecular mechanic forcefields to evaluate the best interaction between the agent and the portion of ZAD.

10 In a preferred variant of the method of the invention, the interacting agent is selected or designed in step b) from a molecule comprising an overall structure similar to ZAD_{Grau}. Since ZAD_{Grau} can interact with itself, putative inhibitors of the biological function of ZAD preferably mimic the overall
15 structure of ZAD_{Grau}.

An agent comprising an overall structure similar to ZAD_{Grau} might exhibit e.g. the same backbone structure as ZAD_{Grau}, or might have functional groups, mimicking amino-acids important for the dimerisation-site, protein-interaction site of ZAD_{Grau} or for the fold of the ZAD_{Grau} molecule. Examples of such amino acids are the above mentioned R5, L6, E47, Q59, I70, Q74, I77 and the groups H37-V42 and F63-V68. Synergetic effects might appear, when residues mimicking both of the above
20 mentioned class 2 and class 3 residues are present in the interacting agent, thereby interfering with the folding of ZAD and the dimerization site.

An interacting agent can therefore be selected or designed from peptidomimetics. These molecules can for example be backbone isosters, having a similar backbone to ZAD_{Grau}. It is also possible to select or design so-called "topographical mimetics", which are non-peptide structures that mimic the binding interactions between two ZAD_{Grau} molecules. These peptidomimetics can comprise non-peptide ligands binding to
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ZAD_{Grau}, as well as compounds comprising both natural amino-acids as well as non-natural elements for example non-natural amino-acids. The peptidomimetics can therefore be compounds with structures in between a true peptide and a complete peptide mimetic, which entirely consists of non-natural elements. Methods of designing peptidomimetics, mimicking the structure of natural peptide compounds, are for example disclosed in the following publications, which are hereby incorporated in their entirety:

Hruby, V.J., Balse, P.M., "Conformational and Topographical Considerations in Designing Agonist Peptidomimetics from Peptide Leads", Current Medicinal Chemistry, 7, pp 945 to 970 (2000).

Kieber-Emmons, T., Murali, R., Greene, M.I. "Therapeutic Peptides and Peptidomimetics", Current Opinion in Biotechnology, 8, pp 435 to 441, (1997).

Wiley, R.A., Rich, D.H., "Peptidomimetics Derived from Natural Products", Medicinal Research Reviews, 13, pp 327 to 384, (1993).

Kemp, D.S., "Peptidomimetics and the Template Approach to Nucleation of Beta-Sheets and Alpha-Helices in Peptides", Trends in Biotechnology, 8, pp 249 to 255 (1999).

Alternatively the interacting agent is selected or designed by linking functional groups mimicking important side chains for the dimerization of ZAD_{Grau} by so-called spacer groups. These spacer groups could comprise the already above-mentioned "linkers" which for example are a part of the software package LUDI.

Yet another object of the invention is a method of obtaining a 3-dimensional model of a ZAD_{Grau}-like molecule of unknown structure, comprising the following step:

A1) using

- 5 - computational means,
 - structural and/or sequence information of the ZAD_{Grau}-like molecule of unknown structure and
 - at least a portion of the structural coordinates of the ZAD_{Grau} molecule shown in Figure 6,
- 10 to obtain the 3-dimensional model of the ZAD_{Grau}-like molecule.

This method of the invention is very useful for the determination of the 3-dimensional structures of ZAD_{Grau}-like molecules, which exhibit a high primary sequence homology to ZAD_{Grau} and therefore exhibit a similar 3-dimensional fold. Structural information about the ZAD_{Grau}-like molecule of unknown structure can for example be obtained in the form of x-ray reflection intensities from crystals of the ZAD_{Grau}-like molecule. It is also possible to use primary sequence information of the ZAD_{Grau}-like molecule in this method of the invention.

ZAD_{Grau}-like molecules of unknown 3-dimensional structure preferably exhibit a primary sequence similarity (homology) of at least 30%. ZAD_{Grau}-like molecules comprise short around 90 amino acid protein modules found as N-terminal domains of C₂H₂ zinc finger proteins in insects. They contain four invariant cysteine residues.

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ZAD_{Grau}-like molecules can preferably also be identified by using e.g. the HMMER package 2.1.1 (Eddy, S.R. (1998) Profile hidden Markov models. *Bioinformatics*, 14, 755-763) to construct a profile hidden Markov model (HMM). Data obtained from the HMM can then be used to search DNA-databases employ-

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ing e.g. the Wise package 2.2.0 (Birney E., Thompson J.D. and Gibson T.J. (1996) PairWise and SearchWise: finding the optimal alignment in a simultaneous comparison of a protein profile against all DNA translation frames. Nucleic Acids Res.

5 24(14):2730-9).

Preferably the primary sequences of the proteins are aligned for maximum homology. Homologous protein variants comprise a large number of matching amino-acids, which can be aligned
10 and have similar structural or chemical properties. In general similar amino acids can be divided into different groups depending on the nature of their side chains. Similar amino acids having basic side chains comprise for example lysine, arginine, histidine. Similar amino-acids with acidic side
15 chains comprise for example aspartic acid or glutamic acid. Amino-acids with uncharged polar side chains are, example given, glycine, asparagine, glutamine, serine, threonine, tyrosine or cysteine. Similar amino-acids with non-polar side chains are for example alanine, valine, leucine, isoleucine,
20 proline, phenylalanine, methionine or tryptophane. Similar amino-acids with aromatic side chains are e.g. tyrosine, phenylalanine, tryptophane or histidine. In homologous primary protein sequences large regions of similar amino-acids which can be aligned, are often interrupted by regions where
25 non-conservative changes occur, for example replacement of a leucine with an aspartic acid. Further, homologous protein variants may also include amino-acid deletions or insertions or both.

30 Preferably step A1) of the method of the invention comprises the following steps:

A11) Obtaining x-ray diffraction data of a crystallised ZAD-Grau-like molecule,

A12) using the portion of the structural coordinates of the ZAD_{Grau} molecule to obtain an electron density map of the ZAD_{Grau}-like molecule, and

- 5 A13) building of a 3-dimensional model of the ZAD_{Grau}-like molecule into the electron density map.

This method uses x-ray diffraction data from a ZAD-like molecule of unknown 3-dimensional structure and e.g. molecular replacement techniques in order to obtain a 3-dimensional model of the ZAD_{Grau}-like molecule of unknown structure.

Molecular replacement methods involve the computer-assisted calculation of patterson vector maps, which show the vectorial distances between atomic positions in the 3-dimensional structures. During the process of molecular replacement rotation and translation of this patterson map is performed within the unit cell of the crystal of unknown 3-D structure in order to superimpose the known 3-dimensional structure of ZAD_{Grau} onto the ZAD_{Grau}-like molecule. Software packages for molecular replacement are for example programs like Molrep or AmoRe (A.Vagin, A.Teplyakov, MOLREP: an automated program for molecular replacement., J. Appl. Cryst. 30, 1022-1025 (1997) and J. Navaza, AMoRe: an automated package for molecular replacement Acta Cryst. A50, 157-163 (1994)). A more detailed description of molecular replacement is given in chapter 10 "Molecular Replacement" in Drenth, "Principles of Protein X-Ray Crystallography", 1994, Springer, pp 213 to 234, which is hereby incorporated in its entirety.

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By using these molecular replacement methods a preliminary electron density map of the ZAD_{Grau}-like molecule of unknown 3-dimensional structure can be obtained. A 3-dimensional model of this ZAD_{Grau}-like molecule can then be built in step A13) of the method of the invention into the electron density

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map. This often involves the building of a preliminary, for example, polyalanine model into the electron density map and then iterative manually or computer-assisted building and refinement of additional elements, e.g. amino acid side chains
5 in order to obtain a more accurate model of the ZAD_{Grau}-like molecule.

In an alternative method step A1) can also comprise using a homology modeling program and the primary sequence of the
10 ZAD_{Grau}-like molecule in order to obtain the 3-dimensional model of the ZAD_{Grau}-like molecule.

In general homology modeling uses known primary sequence information and also known structural information in order to
15 predict an unknown 3-dimensional structure of a protein. Software packages for homology modeling of proteins comprise for example SYBYL or Swiss model (Tripos, St. Louis, MO; SWISS-MODEL: an automated protein homology-modeling server. Nucleic Acids Res. (2003) 31:3381-3385).

20 During the molecular replacement process or homology modeling process a primary sequence alignment between ZAD_{Grau} and the ZAD_{Grau}-like molecule with the unknown 3-dimensional structure might be generated. As mentioned above this involves the
25 alignment of matching homologous amino acids. For example it is possible to adjust the 3-dimensional model of ZAD_{Grau} according to the primary sequence alignment between ZAD_{Grau} and the ZAD_{Grau}-like molecule whose 3-dimensional structure has to be determined. This can be done for example by changing the
30 amino acid side chains of the 3-dimensional model of ZAD_{Grau}, for example to alanine where non-conservative changes occur. This model can then be used e.g. as a search model for the rotation and translation searches of the molecular replacement method.

Using the molecular replacement method or the homology modeling method of the invention, the 3-dimensional structures of ZAD_{Grau}-like molecules for example from *Anopheles gambiae* or from other ZADs of *Drosophila melanogaster* can be determined.

5 In general the 3-D structures of ZAD_{Grau}-like molecules having a sequence homology of at least 30 % to ZAD_{Grau} can be determined with this method of the invention.

Since the inventors revealed, that ZADs contain a zinc ion,
10 coordinated by four invariant cysteines, it is also possible to determine the structure of ZAD_{Grau}-like molecules of unknown 3-dimensional structure by multiple wavelength anomalous dispersion (MAD) using the anomalous scattering of this zinc-atom of ZADs. MAD is described e.g. in chapter 9 "Anomalous Scattering in the Determination of the Protein Phase An-
15 gels and the Absolute Configuration" in Drenth, "Principles of Protein X-Ray Crystallography", 1994, Springer, pp 201 to 212, which is hereby incorporated in its entirety.

20 Another embodiment of the invention are crystals comprising ZAD molecules. These crystals preferably diffract x-rays to a resolution of at least 3 Å and can be used to determine the 3-dimensional structures of the ZAD molecules in the crystal. Preferably the crystal comprises ZAD_{Grau} molecules with the
25 structural coordinates shown in Figure 6 or structurally similar molecules of ZAD_{Grau} comprising structural coordinates with a root mean square deviation from the backbone atoms of the coordinates shown in Figure 6 of not more than 1.5 Å.

30 According to another preferred embodiment of the invention a computer system for generation and computational utilization of a 3-dimensional model of ZAD is provided, comprising the following components:

35 I) A computer-readable medium having stored thereon

- i) at least a portion of the structural coordinates of all the ZAD_{Grau} amino-acids shown in Figure 6 or
- ii) at least a portion of a structurally similar molecule of ZAD_{Grau}, having a root mean square deviation from the backbone of the ZAD_{Grau} amino-acids of not more than 1.5 Å, or
- iii) x-ray diffraction data of ZAD_{Grau} or a structurally similar molecule of ZAD_{Grau}, having a root mean square deviation from the backbone of the ZAD_{Grau} amino-acids of not more than 1.5 Å,

II) computing subroutines for the generation and computational utilization of the said structural coordinates, and

III) a central processing unit for performing said computing subroutines.

Such a computer system is useful for analyzing the x-ray diffraction intensities of crystals comprising ZAD molecules. The computer system of the invention can also be used for the determination of the 3-dimensional structure of ZAD_{Grau}-like molecules by using the 3-dimensional coordinates of ZAD_{Grau} by using e.g. molecular modeling or molecular replacement subroutines.

The computing subroutines of component II) are preferably selected from a group consisting of x-ray data processing, reduction, indexing, merging, intensity scaling and truncation. The subroutines are for example included in programs like DENZO, MOSFLM, the CCP4-package or X-PLOR. Subroutines can also include programs for molecular replacement and molecular modeling, for example the above-mentioned SYBYL, Swiss model, AmoRE or molrep.

In the following the invention will be explained in more detail by the figures and the embodiments.

Figure 1 depicts a primary sequence alignment between the ZAD from Grauzone, its closest paralogue CG15073 in the *Drosophila melanogaster* genome and an *Anopheles gambiae* homologue.

5 Figure 1B shows a primary sequence alignment between ZAD_{Grau} and 32 other homologous proteins from *Drosophila melanogaster* and *Anopheles gambiae*.

10 Figures 2A to 2C depict biochemical and biophysical experiments revealing that ZAD_{Grau} comprises a zinc coordinating fold and an electron density map around the zinc center in ZAD_{Grau}.

15 Figures 3A to 3C show the 3-D structure and topology of ZAD_{Grau}.

Figures 4A and 4B show stereo ribbon plots of ZAD_{Grau} dimers in the crystal.

20 Figures 5A and 5B depict additional biochemical experiments showing that ZAD_{Grau} dimerizes in solution.

Figure 6 shows the structural coordinates of ZAD_{Grau}.

25 Example 1: Cloning of ZAD_{Grau} and Sample preparation:

The DNA coding for Grauzone ZAD residues 2-90 was PCR-amplified, cloned into the NotI/EcoRI sites of the pGEXT-3 vector and recombinantly expressed as a GST-fusion protein in
30 a BL21(DE3) *E. coli* strain. Cells were grown to an OD₆₀₀ 0.7-1.0 at 30°C, shifted to 20°C and induced with 0.7 mM IPTG over night. Cells were harvested by centrifugation and resuspended in phosphate-buffered saline containing Complete™ protease inhibitor tabs (Roche) and traces of lysozyme. Cells

were lysed by sonication and the insoluble fraction was removed by centrifugation. GST-ZAD_{Grau} was captured using Glutathione (GSH)-Sepharose 4B (Amersham) beads in disposable columns following the manufacturer's suggestions. Samples were
5 eluted with 50mM Tris-HCl, pH 8.0, 50mM GSH. Thrombin was directly added to the eluate to 1unit/100µg protein and incubated overnight at room temperature. The protein identity was confirmed by peptide finger printing and subsequently purified with a 1ml Ressource-Q 15µm anion exchange column applying a linear 20ml salt gradient (20mM Tris-HCl, pH 8.0, 0-1M
10 NaCl), a second GSH-column and a Superdex 25/60 75pg gel filtration (GF) column (GF buffer: 10mM Tris pH 7.5, 100mM NaCl, 5mM DTT). Samples were concentrated to 8mg/mL via 20ml 5000Da molecular weight cutoff concentrators (Vivascience).

15
Example 2: Crystallisation of ZAD_{Grau}, X-ray fluorescence spectra and data collection:

Crystallization screens were carried in the sitting drop vapor diffusion format on 24-well Cryschem platesTM (Hampton
20 Research, Laguna Niguel, USA). 1-4µl of an 8mg/ml ZAD_{Grau} solution in GF buffer were combined with 1µl reservoir buffer and equilibrated against a 400µl reservoir. Crystals grew at 22°C with 0.2M ammonium acetate, 0.1M sodium citrate, pH 5.6,
25 and 30% polyethylene glycol 4000 in 3-10 days.

After transfer into Paratone-N (Hampton Research) and removal of residual mother liquor, crystals could be shock frozen in a liquid nitrogen stream. Diffraction data were collected at
30 100K on the HASYLAB beamline BW6 (DESY, Hamburg, Germany; http://www-hasyllab.desy.de/facility/experimental_stations/stations/BW6.htm).

X-ray fluorescence spectra of native crystals clearly indicated zinc emission lines. Figure 2B depicts a X-ray emission
35

spectrum of a native ZAD_{Grau} crystal irradiated with X-radiation of $\lambda = 1.0\text{\AA}$. The peaks in the X-ray emission spectrum are denoted as follows: I - ZnK α line; II - ZnK β line; III - Compton scattering; IV - Elastic scattering. A single
5 ZAD_{Grau} crystal yielded complete anomalous data sets at the f'' -maximum of the K-edge ($\lambda = 1.2828\text{\AA}$) and at a remote wavelength ($\lambda = 1.0500\text{\AA}$). The data were recorded on a MarResearch (Norderstedt, Germany) CCD detector and processed with the HKL package ((Otwinowski and Minor, 1997). The table shows
10 the crystallographic data.

Table: Crystallographic Data

Dataset		Remote	Peak
<i>Data Collection</i>			
Spacegroup		P4 ₁ 2 ₁ 2	
Unit Cell Lengths (Å)	a	48.7	
	b	48.7	
	c	82.1	
Wavelength (Å)		1.0500	1.2828
Resolution (Å)		50.0-2.0	50.0-2.5
Unique Reflections		6907	6089
Redundancy		4.6	2.6
Completeness (%)		97.0	96.3
		(99.7)	(98.5)
I/ σ (I)		36.7	28.2
		(6.3)	(14.2)
R _{sym} ^a (%)		4.2	3.1
		(44.7)	(8.2)
<i>Phasing</i>			
Resolution (Å)		20.0-2.1	
Heavy Atom Sites		1	
R _{cullis} ^b	Centrics	0.50	
	Acentrics	0.61	
Phasing Power ^c	Centrics	1.67	
	Acentrics	1.97	
FOM ^d	Before DM ^e	0.42	
	After DM	0.78	
<i>Refinement</i>			
Resolution (Å)		15.0-2.0	
Number of Reflections		6590	
Number of Non-Hydrogen Atoms			
	Protein	644	
	Zn ²⁺ Ions	1	
	Water Oxygens	56	
R _{work} ^f (%)		24.1	
		(29.6)	
R _{free} ^f (%)		26.7	
		(33.7)	
RMSD ^g From Ideal Geometry			
	Bond Lengths (Å)	0.009	
	Bond Angles (°)	1.20	
Average B-Factors (Å ²)			
	Protein	50.6	
	Zn ²⁺ Ion	37.2	
	Water Oxygens	80.3	
	Wilson B-Factor	52.6	
RMSD B-Factors (Å ²)			
	Main Chain Bonds	1.9	
	Main Chain Angles	3.3	
	Side Chain Bonds	3.5	
	Side Chain Angles	5.2	
Ramachandran Analysis (%)			
	Preferred	91.9	
	Additionally Allowed	6.8	
	Disallowed	1.4	
Mean Residual Error (Å)		0.15	

^a $R_{\text{sym}}(I) = (\sum_{hkl} \sum_i [|I_i(hkl) - \langle I(hkl) \rangle|] / \sum_{hkl} \sum_i [I_i(hkl)])$; $I_i(hkl)$ - intensity of the i^{th} measurement of hkl ; $\langle I(hkl) \rangle$ - average value of hkl for all i measurements

^c $R_{\text{Cullis}} = \sum_{hkl} [||F_{\text{PH}} \pm F_{\text{P}}| - |F_{\text{H,calc}}||] / \sum_{hkl} [|F_{\text{PH}} \pm F_{\text{P}}|]$; $(F_{\text{PH}} + F_{\text{P}})$ if signs are opposite, $(F_{\text{PH}} - F_{\text{P}})$ if equal

^c Phasing Power = $(\sum_n [|F_{\text{h}}|^2] / \sum_n [|E|^2])^{1/2}$; $\sum_n |E^2|$ = lack of closure error = $\sum_n [|F_{\text{PH}}|(\text{obs}) - |F_{\text{PH}}|(\text{calc})]^2$

^d FOM = Figure of merit = $[|F(hkl)_{\text{best}}|] / |F(hkl)|$; $F(hkl)_{\text{best}} = \sum_{\alpha} [P(\alpha) F_{\text{hkl}}(\alpha)] / \sum_{\alpha} [P(\alpha)]$

^e DM - density modification (solvent flattening)

^f $R_{\text{work}} = \sum_{hkl} [||F_{\text{obs}}| - k|F_{\text{calc}}||] / \sum_{hkl} [|F_{\text{obs}}|]$; $R_{\text{free}} = \sum_{hkl \in \text{CT}} [||F_{\text{obs}}| - k|F_{\text{calc}}||] / \sum_{hkl \in \text{CT}} [|F_{\text{obs}}|]$; $hkl \in \text{CT}$ - test set

^g RMSD - root-mean-square deviation

Data for the last 0.05Å in parentheses

Example 3: Structure solution of ZAD_{Grau}, model building and refinement:

If not mentioned otherwise, programs from the CCP4 collection (Project., 1994) were employed for the structure solution.

The location of a single Zn^{2+} ion could be deduced from anomalous difference Patterson maps at the f'' -maximum (program RSPS) or by direct methods (SHELX, <http://shelx.uni-ac.gwdg.de/SHELX/>) and was used to phase the data with the remote data set as the reference (MLPHARE). The hand of the heavy atom was revealed during solvent flattening (DM).

Prominent secondary structure elements were manually fitted to the electron density with MAIN (Turk, 1996). Subsequently, these regions were decorated with side chains guided by the prominent features of the aromatic residues and by four cysteine residues, which coordinated the Zn^{2+} ion. The connecting loops were sequentially traced by alternating refinement of the partial model (CNS) (Brunger et al., 1998) and model building guided by the experimental and $2F_o - F_c$ electron density maps. Refinement included standard procedures of

positional and B-factor optimization, a round of simulated annealing and positioning of 56 water molecules into spherical peaks of the $F_o - F_c$ maps. The final step was a TLS refinement with REFMAC5 with the structure divided into four rigid bodies (the Zn^{2+} ion, residues 2-19 and 51-54, residues 20-

50, and residues 55-81). No data within 15.0-2.0Å were excluded and during all refinement steps the same set of reflections (5%) was used to monitor the R_{free} -factor (see Table). The geometry of the final model was analyzed with PROCHECK ((Laskowski et al., 1993); see Table), secondary structure elements were extracted with PROMOTIF (<http://www.biochem.ucl.ac.uk/bsm/promotif/promotif.html>).

ZAD is a Zinc-Binding Protein Module

Based on the four conserved cysteine residues, ZADs have been suggested to comprise metal-binding modules (Chung et al., 2002; Lander et al., 2001; Lespinet and Aravind, 2002). Because metal association is expected to contribute significantly to the stability of the fold, heat denaturation experiments were performed in the presence and absence of EDTA. In the absence of EDTA, ZAD_{Grau} was stable for at least 15 minutes at 60°C, whereas in the presence of EDTA (5mM) a considerable portion of the protein precipitated when heated above 50°C for 15 minutes (see Figure 2A). In these experiments care was taken to exclude divalent metal ions from the purification procedure. The result therefore suggests that ZAD_{Grau} contains an endogenous metal center that confers stability on its structure. For EDTA-dependent precipitation studies, 10μM ZAD_{Grau} samples in GF buffer either lacking EDTA or containing 5mM EDTA were incubated for 15min at room temperature, 37°C, 50°C and 60°C. Samples were pelleted and the supernatant was analyzed by SDS-PAGE.

Figure 2A shows a denaturing SDS-PAGE with the portion of ZAD_{Grau} remaining soluble after the above mentioned EDTA-dependent precipitation studies. The samples were incubated at the indicated temperatures in the absence (-EDTA) and in the presence of 5mM EDTA (+EDTA). Reduction of the soluble

fraction is clearly visible when ZAD_{Grau} is heated above 50°C in presence of the chelator EDTA.

In order to test the proposal, that ZAD_{Grau} contains a zinc metal center, the inventors performed an X-ray fluorescence scan. With this approach, the characteristic emission lines for zinc were observed (see Figure 2B and example 2). This result and the finding that the anomalous diffraction from the zinc center could be exploited to solve the structure with a native crystal (see below) demonstrate that ZAD_{Grau} includes a zinc ion.

Structure Solution and Quality of the Model

The primary sequence of ZAD_{Grau} is shown in Figure 1A. This Figure 1A shows an alignment of the Grauzone ZAD with its closest paralog, CG15073, in the *Drosophila melanogaster* genome and with a putative *Anopheles gambiae* homolog. Asterisks mark 100% conservation in all three ZADs. The grey boxes show conserved hydrophobic or aromatic amino acid residues. The secondary structure elements of ZAD_{Grau} are shown as arrows (β -sheets) or tubes (α -helices).

The crystal structure of ZAD_{Grau} was solved *de novo* by a two-wavelength multiple anomalous dispersion experiment around the K-edge of an endogenous Zn²⁺ ion. A segmented polyaniline model could be manually fitted to the experimental electron density map and allowed the subsequent incorporation of side chains. The sequence assignment was guided by well-defined aromatic residues and by the two pairs conserved cysteines (C4, C7, C53 and C56; see Figures 1A, 2C), which coordinated the Zn²⁺ ion.

Figure 2C shows a stereo plot of the electron density maps around the zinc center in ZAD_{Grau}. The experimental MAD elec-

tron density map (1 sigma) is shown around the zinc-coordinating cysteines. Around the Zn^{2+} -ion an anomalous difference Fourier map (10 sigma), generated with the anomalous differences at the peak wavelength and the phases obtained after solvent flattening is shown. The zinc-coordinating cysteine residues are labeled and shown in ball-and-stick. If not mentioned otherwise, all structural figures were prepared with Bobscript (<http://www.strubi.ox.ac.uk/bobscript/>) and rendered with Raster3D (Merritt and Bacon, 1997)

Three regions in flexible loops (V11-C17, D22-E27 and E41-E47; see Figures 1A, 3A) initially displayed weak densities and were fitted during the subsequent rounds of refinement and manual model building. These loop regions, which are made up of non-conserved residues, lack substantial intra- and interchain contacts. Thus, they gained above-average temperature factors during the refinement. The main chain electron density of the molecule remained fragmented around position S12-A14 and D22 in the final $2F_o - F_c$ map, consistent with high internal flexibility and lack of crystal packing contacts in these regions. All other side chains of the refined model, except those of some hydrophilic surface residues, were entirely covered by the final $2F_o - F_c$ map. No electron density developed for the C-terminal nine residues (positions 82-90; Figure 1A) of ZAD_{Grau} and the five N-terminal vector-derived residues that are contained in the recombinant protein. The final model therefore encompasses the residues D2-S81 of ZAD_{Grau} that could be traced unambiguously. Presumably because of the considerable fraction of flexible residues at the termini, which could not be accounted for in the final model, the refinement converged with an R-factor/ R_{free} -factor of 24.1%/26.7%. Convergence at these numbers is consistent with a rather high B-factor extracted from a Wilson plot ($52.6 \text{ \AA}^2$), which is comparable to the averaged B-factor of the final structure ($52.9 \text{ \AA}^2$).

During all refinement steps 5% of the reflections were set aside to monitor the R_{free} -factor (see Table). Of the final model, 91.9% of the residues resided in the preferred regions of the Ramachandran plot, 6.8% in the additionally allowed areas. Only the single residue K29, which was well defined in both the experimental and the final model-derived maps assumed an unconventional ϕ/ψ conformation even after manual interference. The mean positional error of the model was estimated at 0.15Å (Luzzati, 1952).

Structural Properties of ZAD_{Grau}

The structured portion of ZAD_{Grau} resembles the letter 'b'. Its approximate dimensions are 60Å x 30Å x 30Å (Figure 3A and 3B). Figure 3A shows a topological diagram of the ZAD_{Grau} structure identifying the origin of the four zinc-coordinating cysteines and the borders of the secondary structure elements (numbers). The Figures 3B and 3C depict two orthogonal stereo ribbon plots of the ZAD_{Grau} structure. In the side view of Figure 3B the structure resembles the letter 'b'. N- and C-termini, secondary structure elements and residues of the zinc center are labeled. The Zn²⁺ ion and its coordinating cysteine side chains are shown in ball-and stick.

The N-terminal body of the 'b' comprises a globular fold structured around a zinc ion. The C-terminal stem is formed by a long α -helix (α 2, positions 54-80) that contains almost one third of all residues of the domain. Residues 82-90 are not included in the ZAD consensus sequence (Chung et al., 2002) and are disordered in the present structure. Presumably, they provide a flexible linker to the remainder of the molecule, i.e. an acidic region followed by an array of eight

C₂H₂ zinc fingers in the case of Grauzone (Chen et al., 2000).

The arrangement of the different ZAD structural elements is summarized in Figures 3A-3C. It shows that the N-terminus of ZAD_{Grau} is folded into a hairpin motif (D2-V11; Figure 3A-3C). It contains two zinc-coordinating cysteine residues (C4 and C7; Figure 1A), which are conserved in all ZADs so far identified (Chung et al., 2002). Replacement of C7 by a tyrosine residue has been described as a partially lethal mutation of Sry- δ (Croizatier et al., 1992). This observation implies that C7 is essential for ZAD function and supports the argument that C7 is directly involved in zinc coordination. The N-terminal hairpin is followed by a loop (L1: S12-M16) extending into a two-stranded antiparallel β -sheet (β 1: C17-Q19, β 2: V51-C53). The β -sheet is interspersed by a long loop-helix-loop insertion (L2: I20-K29, α 1: V30-H37, L3: F38-K50). The C-terminal helix, α 2, is directly linked to the second β -strand with a slanting angle of $\sim 80^\circ$ between the helix axis and the average long axis of the β -sheet. Helix α 1 of the L2- α 1-L3 motif is positioned perpendicular to both the long axis of the sheet and the axis of helix α 2, wedging between the two motifs. The conserved C53 and C56 residues reside in the C-terminal part of strand β 2 and the first turn of helix α 2, respectively, and complete the zinc coordination sphere. The fold of ZAD_{Grau} appears to be critically dependent on the zinc coordination. This aspect of the structure (see Figures 3A-3C) is consistent with the results obtained with EDTA-dependent precipitation of ZAD_{Grau}. The two pairs of coordinating cysteines are approximately 50 residues apart. Zinc-coordination, therefore, links the β 2- α 2 transition region at the center of the molecule with the N-terminus of the domain likely to solidify the structure. In this view, the position of the zinc ion allows the fold to constrain the slanting an-

gle of the C-terminal helix, $\alpha 2$, with respect to the long axis of the β -sheet (see Figures 3B-3C). The relative orientation of this long helix and the β -sheet is further defined by helix $\alpha 1$, which rests with one of its surfaces on the end of the β -sheet distal to the zinc ion and the adjacent loop regions. With a neighboring surface, helix $\alpha 1$ is in contact with the N-terminal and central portions of helix $\alpha 2$. Both the $\beta 1$ -sheet and the $\alpha 1$ - $\alpha 2$ associations are based on extensive hydrophobic contacts forming a considerable hydrophobic core ($\alpha 1$: V29, V33, L34, H37, F38; $\beta 1/\beta 2$: L18, I20, I52; $\alpha 2$: W57, V60, F63, H64, Y67; Figures 1A, 3A-3C). In addition, the conserved H37 residue (atom NE2) of helix $\alpha 1$ hydrogen bonds at the edge of this core to the conserved H64 (atom ND1) of helix $\alpha 2$. In conclusion, the L2/ $\alpha 1$ /L3 region between the two central β -strands and the zinc ion provide platforms which orient the C-terminal helix relative to the globular portion of the ZAD_{Grau} molecule.

To further support the overall structure of ZAD_{Grau} obtained by X-ray crystallography, the inventors took circular dichroism spectra of the molecule in solution (data not shown). For far UV-CD measurements, the sample was dialyzed against 10mM Na₂HPO₄, pH 7.9. The spectra (190-260nm) were recorded with a JASCO J-720 spectropolarimeter (Jasco Corporation, Tokyo, Japan) at 20°C in a 0.1cm cuvette with a scanning speed of 50nm/min. 30 scans were averaged and baseline corrected. The spectra were analyzed using the neural network-based CD deconvolution software CDNN v2.1 (Böhm et al., 1992).

The calculated secondary structure content (45-50% helix, 5-10% strands) obtained from these recordings compare favorably to those seen in the crystal structure (37% helix, 6% strands). The conservation of length, predicted secondary

structure, arrangement of the annotated secondary structure elements and the critical amino acid residues of ZADs (Chung et al., 2002; Figure 1A) suggest that the present ZAD_{Grau} crystal structure provides a prototype for ZAD folding.

5

Crystal Structure Suggests ZAD_{Grau} Homodimers

The evolutionary restriction of ZADs to certain C₂H₂ zinc finger genes in insects, the chromosomal clustering of the majority of these ZAD-containing genes and the unique folding characteristics of the domain next to the DNA-binding domain of transcriptional regulators such as Grauzone strongly argue that the ZAD is associated with a specific and distinct biological function. However, the mere similarity to the fold group of the TCZFs does not allow any conclusions about the specific role of ZAD, because treble-clef motifs embody functions as diverse as binding to nucleic acids, proteins as well as small ligands and some may even exert enzymatic activity (Grishin, 2001).

20

However, the crystal structure of ZAD_{Grau} clearly supports a model, in which ZAD represents a protein-protein interaction module involved in homodimerization or interaction with other proteins. Figures 4A and 4B show that in the crystal two ZAD_{Grau} molecules are associated through a twofold axis in an isologous head-to-tail fashion. Figure 4A depicts a stereo ribbon plot of a ZAD_{Grau} dimer as seen in the crystal. The two subunits are denoted Mol I and Mol II and the two zinc centers are depicted in ball-and-stick. N- and C-termini are labeled. The orientation of the rear subunit is the same as in Figure 3B. Figure 4B shows a stereo representation of residue interactions in the dimer interface, 90° from the view in Figure 4A. The two ZAD_{Grau} subunits are rendered semi-transparent to clearly reveal the interacting residues in

30

ball-and-stick. All residues and the molecular termini are labeled.

As revealed by the protein-protein interaction server
5 (<http://www.biochem.ucl.ac.uk/bsm/PP/server/>), the dimer contact buries $\sim 1000\text{\AA}^2$ of accessible surface area (ΔASA). This value seems on the lower side when compared to known homodimers (Jones and Thornton, 1996). However, the interaction surface of ZAD_{Grau} covers a total of close to 20% of the en-
10 tire surface area. More importantly, a large number of amino acid residues, which are strongly conserved among ZAD family members (for details see Chung et al., 2002), are responsible for the contacts between the two subunits (see Figure 4B). When the conserved residues are mapped on the surface of
15 ZAD_{Grau}, it becomes obvious that the largest conserved surface patch closely coincides with the presumed dimer interface. In particular, hydrophobic residues of the long C-terminal helix $\alpha 2$ (F63, F66, I70, Y77; Figure 1A) build up major parts of the contact interface (Figure 4B). As a consequence, the di-
20 mer interface is composed of 72.5% of the non-polar amino acid residues. The presumed dimer interface is thus designed very differently from the remainder of the surface, which is lined with polar residues and exhibits a highly negative electrostatic potential. In the region where the tip of helix
25 $\alpha 2$ from one molecule contacts the globular portion of the other subunit, some intermolecular hydrogen bonds are observed as well (Q74-R5; Y77-E47; Figure 4B).

Because of the involvement of its hydrophobic side chains,
30 the presumed homodimerization mode provides a straightforward explanation for the amphipathic design and the unusual length of helix $\alpha 2$ (Figure 3A). Since, in addition, because both helix $\alpha 2$ and the globular portion of ZAD are involved in this dimerization mode, the importance of restraining their rela-

tive orientations by the inserted L2/ α 2/L3 module and the zinc coordination becomes obvious. Further supporting its significance, the present dimerization may explain the lethal phenotype observed with the conserved R4 (corresponding to R5 in Grauzone) mutated to glycine in Zw5 (Gaszner et al., 1999). Its side chain is positioned by ionic interactions with D46 to engage in a hydrogen bond with Q74 of the neighboring molecule (Figure 4). In any case, the analysis of the crystal packing strongly suggests a functional homodimerization of ZAD_{Grau}.

Besides the above symmetrical contact between two ZAD_{Grau} molecules, other crystal contacts were observed. Some of these are not functional because of an unreasonably small Δ ASA. Other contacts did not involve conserved residues. One rather intimate alternative association takes place through the crystallographic fourfold screw axis. However, this symmetry element gives rise to heterologous contacts, leaving the bonding potentials of ZAD unsaturated. The latter interaction mode could therefore lead to the formation of larger oligomers. The inventors therefore examined the arrangement of ZAD in solution.

Example 4: Examination of the dimerization of ZAD_{GRAU} in solution by Chemical cross-linking and Multi-angle-laser-light scattering:

The quaternary structure of ZAD_{Grau} in solution was investigated by two approaches. First, the inventors performed chemical crosslinking experiments with glutaraldehyde (see Figure 5A). The crosslinking reactions were carried out at 25°C in 20mM Tris-HCl, pH 7.5, 100mM NaCl with 15 μ M samples of ZAD_{Grau}, GST (26kDa), and SNAP-25 (25kDa) (Fasshauer et al., 1999) and 1mM glutaraldehyde. The reactions were stopped at selected time points by boiling in SDS sample buffer.

In contrast to SNAP-25, which was reported not to self-interact (Fasshauer et al., 1999), and in parallel with the dimerizing GST, ZAD_{Grau} could be efficiently crosslinked to the dimer state, but no higher oligomers were observed. Figure 5A shows the results from chemical crosslinking with glutaraldehyde displayed on a denaturing polyacrylamide gel. Crosslinking took place for the times indicated above the lanes. SNAP-25 served as a negative control, GST as a positive control. Molecular weight markers are given on the left (kDa).

Secondly, multi-angle-laser-light-scattering following size exclusion chromatography yielded strong evidence for homodimerization in solution (see Figure 5B). The experiment was performed on a HR-10/30 Superdex-200 size exclusion column (Amersham) connected to a UV spectrometer and the Dawn and Optilab instruments XY (Wyatt Technology Corp.). 200µl of a 25µM ZAD_{Grau} sample were chromatographed in 20mM Tris-HCl, pH 8.0, 150mM NaCl. The UV₂₈₀ absorption, the light scattering at 632.8nm and the differential refraction of the elution profile were monitored. Spectra were analyzed with the Astra software package (Wyatt, 1993). The inventors also checked ZAD_{Grau} migration on an analytical Superdex-75 column in comparison to protein molecular weight standards.

Figure 5B shows the results from a gel filtration chromatography run combined with multi-angle-laser-light scattering analysis of the emerging peak. Emergence of protein from the column was detected by absorbance at 280nm (black trace, left y-axis), readout from the light scattering of the single, symmetrical peak is depicted as a gray line 10 (right y-axis).

ZAD_{Grau} eluted as a single symmetrical peak from various size exclusion columns with different optimal separation regimes. Comparison of the elution times with those of reference proteins was consistent with ZAD_{Grau} dimers (not shown). The
5 scattering signal at 632.8nm across the elution peak indicated a molecular weight of 21.11kDa, which matched the theoretical dimer mass (21.14kDa) almost perfectly (Figure 5B). This high congruence and the lack of a monomer or a trimer signal, strongly suggested a monodisperse dimer solution.
10 Collectively, our results provide strong evidence for ZAD_{Grau} homodimerization under near-physiological salt conditions. They leave the twofold symmetrical association of ZAD_{Grau} in the crystal (see Figures 4A and 4B) as the only dimerization mode, which is consistent with all observations. Because
15 ZAD_{Grau} is an independently folding unit, dimerization is expected to prevail within the context of the full-length Grauzone transcription factor. By extension of this finding, a general function of ZADs could be to provide dimerization modules that mediate homodimer and/or heterodimer formation
20 among closely related members of the ZAD-transcription factor family.

In support of this observation, homodimerization was also reported for the ZAD-containing transcription factor Sry-
25 δ (Payre et al., 1997; Ruez et al., 1998). It is noteworthy, that Sry- δ comprises a deviated version of ZAD family members and could thus not be considered as prototype for ZAD function (Chung et al., 2002).

30 ZAD_{Grau} Lacks DNA-Binding Features

The majority of zinc finger proteins were shown to be nucleic acid binding proteins (Berg and Shi, 1996; Laity et al., 2001; Pavletich and Pabo, 1991; Wolfe et al., 2000). Because

of this amply documented function, the inventors asked whether the ZAD_{Grau} structure has features to support nucleic acid binding as well. In order to obtain first hints, the inventors compared the ZAD with the structure of the C-terminal treble-clef module of the chicken erythroid transcription factor GATA-1 (Tjandra et al., 1997), a representative of sequence-specific DNA-binding TCZFs (treble-clef zinc finger). The two molecules display opposite electrostatic surface potentials. Whereas the TCZF of GATA-1 exhibits an almost continuously electropositive surface potential that facilitates a close association with the negatively-charged sugar-phosphate backbone of the DNA, the ZAD_{Grau} is almost completely wrapped into negative potential (not shown). This observation strongly argues against DNA binding features of ZAD.

Superimposition of one subunit of a ZAD_{Grau} dimer on the structure of DNA-bound GATA-1 TCZF (Tjandra et al., 1997) revealed that the second ZAD_{Grau} module faces the minor groove of the DNA and would thus collide with the DNA backbone (not shown). Furthermore, Grauzone mutants, which lack the ZAD-containing region of the protein, had unaltered DNA-binding properties *in vitro*, whereas the part of Grauzone protein that contains the ZAD in the absence of C₂H₂ zinc fingers had no DNA-binding activity (Chen et al., 2000), supporting the argument that ZAD_{Grau} has no DNA-binding features. Since the key positions and features of the amino acid residues are conserved among ZADs (for an alignment see Chung et al., 2002), it appears unlikely that any of them carries DNA-binding properties. The inventors therefore propose that ZADs, in general, are protein interaction modules involved in homodimerization and/or the formation of protein complexes.

To further support this conclusion, the inventors also compared the ZAD_{Grau} dimer with the DNA-bound dimer of other members of the TCZF fold group, the glucocorticoid receptors

(Luisi et al., 1991; Grishin, 2001). In these receptor molecules, the DNA binding helix equivalent to ZAD helix $\alpha 2$ is employed as a recognition element, which is positioned in the DNA major groove. Two DNA binding domains interact through
5 their C-terminal helical extension, which have no equivalent in ZADs. These C-terminal extensions yield a spacing between the two recognition helices from neighboring subunits, which is large enough to position them both into neighboring turns of the DNA major groove. Therefore, the dimerization mode of
10 ZADs is fundamentally different from the interaction mode seen in the DNA binding domains of glucocorticoid receptors, consistent with these molecules serving different functions.

Example 5: Identification of a putative Grauzone ZAD homologue in the *Anopheles gambiae* genome:
15

The inventors searched the whole *Anopheles gambiae* genome using the ZAD HMM described in Chung et al. 2002 and the Wise package 2.2.0 (Birney et al., 1996). All identified ZADs were
20 aligned with all *Drosophila melanogaster* ZADs using ClustalW 1.8.1. (Thompson et al., 1994), false positive hits were eliminated. The alignment was used to construct a neighbor-joining tree with ClustalW. Based on this tree the inventors identified the closest relative of the Grauzone ZAD in the
25 *Anopheles gambiae* genome. The alignment of the Grauzone ZAD with the closest paralog, CG15073, of *Drosophila melanogaster* and the putative *Anopheles* homologue was performed with ClustalW using default parameters (see Figure 1A).

30 Figure 1B shows a primary sequence alignment of ZAD_{Grau} denoted as Grauzone with homologous protein domains from *Drosophila melanogaster* or *Anopheles gambiae*. The primary sequences of ZAD_{Grau} from *D.melanogaster* and the putative ZAD from *A.gambiae* (denoted as Grauzone *A.gambiae*) exhibit a high

degree of primary sequence homology of around 50% and a primary sequence identity of around 38% as determined with the program EMBOSS-Align (Rice, P. Longden, I. and Bleasby, A.

(2000) "EMBOSS: The European Molecular Biology Open Software Suite" Trends in Genetics 6(6.):276-277). The alignment was
5 done with the program ClustalW 1.81 (Thompson et al., 1994). The four cysteines coordinating the zinc are shown with a grey background. Similar residues are shown in grey letters.

10 Figure 6 shows the structural coordinates of ZAD_{Grau} as determined by the above-described x-ray structure analysis (see example 3). The structure coordinates refer to mathematical coordinates derived from mathematical equations related to the patterns obtained on diffraction of a monochromatic beam
15 of x-rays by the atoms of the crystal of ZAD_{Grau}. The diffraction data are used to calculate an electron density map of the asymmetric unit of the crystal. The electron density map shows the electron distribution within the crystal and can be used to build a 3-dimensional model of ZAD_{Grau}. It is common
20 knowledge of experts in the field of protein x-ray crystallography, that structurally similar proteins exhibiting the same overall 3-dimensional fold have a root mean square deviation from the backbone atoms of the coordinates of ZAD_{Grau} shown in this Figure 6 of not more than 1.5 Å. In Figure 6
25 the structural coordinates of ZAD_{Grau} are given as a table of structural coordinates in protein data bank (PDB) format, including the values X, Y, Z and B for each of the atoms in the amino-acids in ZAD_{Grau}. The term "atom type" refers to the type of atom of the respective amino acid, for example the
30 nitrogen-atom of the amide backbone (N), C-α (CA), C-β (CB), C-γ (CG), C-δ (CD) atoms or oxygen atoms. The term "residue" refers to the type of amino-acid given in the three letter code, to which the aforementioned atom belongs, for example aspartate (ASP) or isoleucine (ILE). The technical terms "X,

Y, Z" refer to the crystallographically defined atomic position of the atoms calculated with respect to a specially defined crystallographic origin. The term "B" refers to a thermal factor, that measures the mean displacement of the atom
5 around its atomic center in crystals of ZAD_{Grau} .

The scope of protection of the invention is not limited to the examples given hereinabove. The invention is embodied in each novel characteristic and each combination of characteristics,
10 istics, which includes every combination of any features which are stated in the claims, even if this combination of features is not explicitly stated in the claims.

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Claims

1. Method for obtaining an agent, which interacts with a
ZAD_{Grau} molecule or a molecule with similar fold to ZAD_{Grau},
5 comprising the following steps:
 - a) using the structural coordinates of
 - i) at least a portion of the structural coordinates of the
ZAD_{Grau} amino acids shown in Fig. 6, or
 - ii) at least a portion of a structurally similar molecule of
10 ZAD_{Grau}, having a root mean square deviation from the back-
bone of the said ZAD_{Grau} amino acids of not more than 1.5
Å,
to generate a three-dimensional model of the portion of
ZAD_{Grau} or the molecule with similar fold to ZAD_{Grau},
 - 15 b) employing the three-dimensional model and at least one mo-
lecular modeling technique to select or design an inter-
acting agent.
2. Method according to claim 1,
20 - wherein in step a) the structural coordinates of
 - i) at least two of the following ZAD_{Grau} amino acids shown
in Fig. 6:
- R5, L6, E47, Q59, I70, Q74, Y77, H37-V42 and F63-V68, or
 - ii) a portion of a structurally similar molecule of ZAD_{Grau},
25 having a root mean square deviation from the backbone
atoms of said amino acids of not more than 1.5 Å
is used.
3. Method according to one of the claims 1 or 2, further com-
30 prising the following steps:
 - c) synthesising said interacting agent and
 - d) bringing said agent in contact with the binding site of
the molecule or structural similar molecule to detect the
interaction.
- 35 4. Method according to one of the claims 1 to 3,
- wherein step b) comprises:

- screening a library of individual chemical entities for the agent by performing a computer assisted docking operation between the individual chemical entities and the portion of the three-dimensional model.

5

5. Method according to one of the claims 1 to 3,

- wherein step b) comprises:
 - designing of the agent by assembling individual chemical entities, binding to the portion of the
- 10 three-dimensional model into a single interacting agent and docking this single interacting agent to the portion of the three-dimensional model using computational means.

15 6. Method according to one of the claims 4 or 5,

- wherein step b) further comprises:
 - analysing the results of the docking operation to evaluate the interaction between the agent and the
- 20 portion of the three-dimensional model of the molecule or structural similar molecule.

7. Method according to one of the claims 4 to 6,

- wherein step b) further comprises:
 - performing an energy minimisation or molecular dynamics operation using computational means to improve
- 25 the interaction between the agent and the portion of the three-dimensional model.

8. Method according to claim 3,

- 30
- wherein step d) comprises orally administering the interacting agent to flies and detecting the rate of dying or infertile flies.

9. Method according to one of the claims 1 to 8,

- 35
- wherein in step b) the interacting agent is selected or designed from a molecule comprising an overall structure similar to ZAD_{Grau}.

10. Use of an agent obtaining by a method according to one of the claims 1 to 9 as an insecticide.

5 11. A Method of obtaining a three-dimensional model of an ZAD_{Grau}-like molecule of unknown structure, comprising the following step:

A1) using

- computational means,
 - 10 - structural and/or sequence information of the ZAD_{Grau}-like molecule of unknown structure and
 - at least a portion of the structural coordinates of the ZAD_{Grau} molecule shown in Fig. 6
- to obtain the three-dimensional model of the ZAD_{Grau}-like molecule.
- 15

12. Method according to the previous claim,

- wherein step A1) comprises the following steps:

- A11) obtaining x-ray diffraction data of a crystallized ZAD-
20 _{Grau}-like molecule,
- A12) using the portion of the structural coordinates of the ZAD_{Grau} molecule to obtain an electron density map of the ZAD_{Grau}-like molecule, and
- A13) building of a three-dimensional model of the ZAD_{Grau}-like
25 molecule into the electron density map.

13. Method according to claim 11,

- wherein step A1) comprises:

- using a homology modeling program and the primary se-
30 quence of the ZAD_{Grau}-like molecule to obtain the three-dimensional model of the ZAD_{Grau}-like molecule.

14. Method according to one of the claims 11 to 13,

- wherein step A1) comprises:
- 35 - generating a primary sequence alignment between ZAD-_{Grau} and the ZAD_{Grau}-like molecule and adjusting a three-dimensional model of ZAD_{Grau} according to the

primary sequence alignment and using this model as a portion of the structural coordinates of the ZAD molecule.

- 5 15. Method according to one of the previous claims 11 to 14,
- wherein the ZAD_{Grau}-like molecule is selected from the following molecules:
- ZAD-like molecules from *A. gambiae*, *D. melanogaster*.
- 10 16. A crystal comprising ZAD molecules.
17. The crystal according to the previous claim,
- further diffracting X-rays to a resolution of at least 3 Å.
- 15 18. The crystal according to claim 17,
- wherein the ZAD molecules are ZAD_{Grau}.
19. The crystal according to claim 17,
20 - wherein the ZAD molecules comprise
i) ZAD_{Grau} molecules with the structural coordinates shown in Fig. 6, or
ii) structurally similar molecules of ZAD_{Grau}, comprising structural coordinates with a root mean square
25 deviation from the backbone atoms of the coordinates shown in Fig. 6 of not more than 1.5 Å.
20. A computer-system for generation and computational utilization of a three-dimensional model of ZAD, comprising
30 the following components:
I) a computer readable medium having stored thereon
i) at least a portion of the structural coordinates of all the ZAD_{Grau} amino acids shown in Fig. 6, or
ii) at least a portion of a structurally similar molecule of
35 ZAD_{Grau}, having a root mean square deviation from the backbone of the said ZAD_{Grau} amino acids of not more than 1.5 Å, or

iii) x-ray diffraction data of ZAD_{Grau} or a structurally similar molecule of ZAD_{Grau},

II) computing subroutines for the generation and computational utilization of the said structural coordinates,

5 III) a central processing unit for performing said computing subroutines.

21. The computer-system according to the previous claim,

- 10 - wherein the computing subroutines of component II) are selected from the group consisting of
- x-ray data processing, reduction, indexing, merging, intensity scaling, truncation,
 - molecular replacement, molecular modeling.

Fig. 1 A

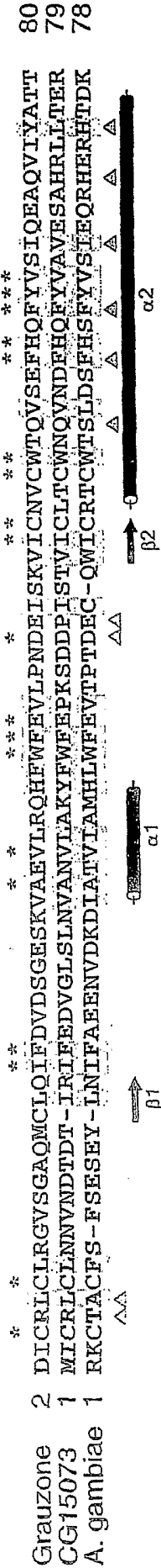


Fig. 1B

	1	20	40	60	80
X -09 CG11696	MICRLCLM	---DAEH---	GVPIFGQEP	PMGPAHRQ	LAELIERHLLVLAEND
X -10 CG11695	MICRLCLD	---DAEH---	SVPIFDQDD	SGDPVPSN	LAELIEKHLQLVLLPND
2R-07 CG15073	MICRLCLN	NVNDTDT	---TRIFEDV	---LSINVAN	VLAKYFWFEPEKSD
2R-08 Grauzone	DIICRLCLR	GVSGAQM	---CLQIFDV	---GESKVAE	VLROHFWFEVLPND
2L-11 CG17568	NWRLCAK	---DDAHG---	NVKVQMN	---ENSQGNW	DNVLLMAIRKYFEVQMLED
3R-13 CG6254	QWRLCSK	---EHPT---	NQNVFR	---EAKQNSW	ASTWMTAMTIGKYFWVDIKRED
2L-12 CG10366	QWRLCAK	---DDVQG---	NLNVFIKNE	DSGEAASME	LATAIGKYFWVNITRGE
X -01 CG2711	LNCRTCTR	ACKLH	---DDVQG---	NLNVFIKNE	DSGEAASME
X -02 CG2712	ALCRVCHT	ASPRC	---DDVQG---	NLNVFIKNE	DSGEAASME
3L-02 CG10270	TVCRTCLS	TVDDS	---AAYDLFR	---LAKKLCV	CTSLSVQAD
3L-03 CG10269	SLCRICLN	HLQND	---AAYDLFR	---LAKKLCV	CTSLSVQAD
3L-01 CG10274	AVCRICLN	RLPEN	GGAGSFDLFI	---TPG---	LAKKLCVCTSLAVEQSD
3R-36 CG4639	TLICRLCL	KEHQA	---YAFD---	EDDTQLS	---LPEVRLMA
2L-01 CG18555	NKCRVCLAS	---	KNMVNIFE	---RQDLPV	SIAMHIIECTGFVEKGD
2L-02 CG3485	DRCRVCLG	VS	---ENVNIFE	---KQDLVSV	IAQVISECTDCDIKEGD
3L-10 CG7204	SICRVCLVS	Y	---ENVNIFE	---KQDLVSV	IAQVISECTDCDIKEGD
2L-05 CG15436	EICRVCM	IS	---GKLVNIFE	---RRTRVSI	AEAMIAOCTGFVEKGD
3R-34 CG4413	VICRVCLN	NPSEG	---EELHDI	FE	---TAST---
3R-35 CG4936	IVCRVCLQ	QP	---KEHMAS	IFND	---DSEK---
3R-05 CG8145	NVCRVCG	RSRLCP	---KAVELFR	---KSLKLF	NS
3R-07 CG8159	NVCRVCA	SSTDNS	---MPKNLF	HI	---
3R-22 CG18764	LQCRTCG	SITVYK	---TPSNLF	FE	---
3R-44 CG1792	RNCRTCG	LFIFCS	---NPVNIF	DL	---
3R-29 CG17802	PQCRICG	DLITYQ	---KSLNLF	DL	---
3R-06 CG11762	IVCRVCG	RQKICE	---KSLNLF	DL	---
3R-08 CG9793	KVCRTCG	KRTNAE	---RSLNIFE	K	---
3R-09 CG9797	STCRVCA	KYASNK	---RSPKLF	ER	---
3R-15 CG6689	OKCRTCYN	DFTADF	---RAKDLF	DP	---
3R-27 CG17803	SKCRTCFR	IISRHE	---DAQDLY	DR	---
3R-23 CG18476	VICRTCD	STDDN	---LKLATP	---	---
3R-37 CG10669	RTCRVCG	CGDDGR	---YWIETP	---	---
3R-38 CG11902	DVCRTCAV	QNSQ	---LLPLST	C	---
Grauzone	RKCTACFS	---FSESEY	---LNIFAE	EN	---
A. gambiae					

block 4

r3

block 3

r2

block 2

r1

block 1

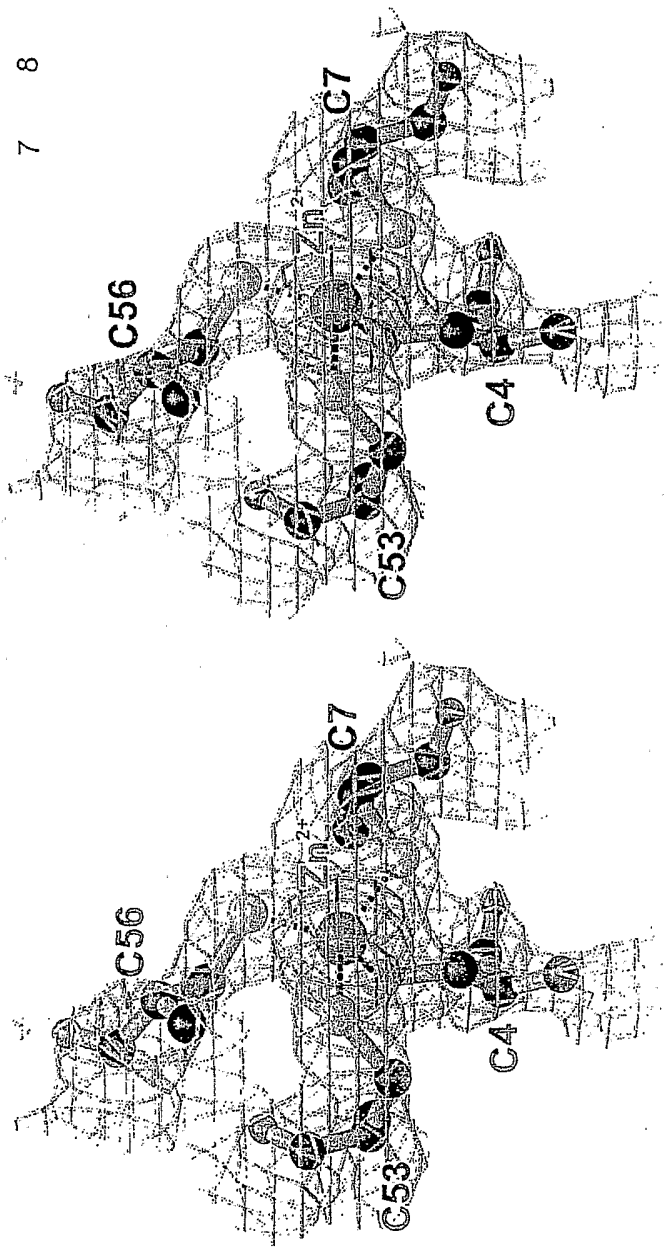
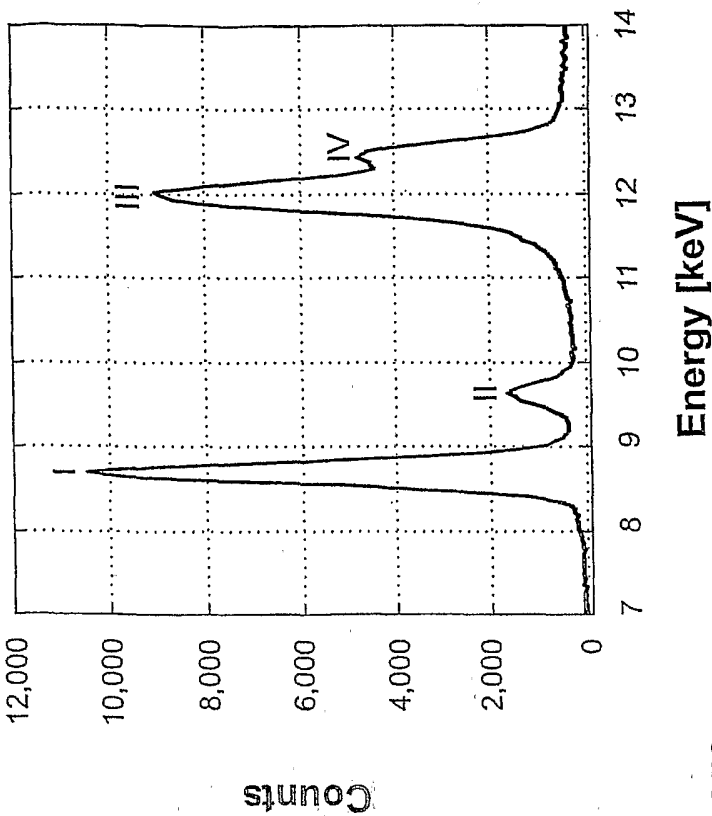


Fig. 3

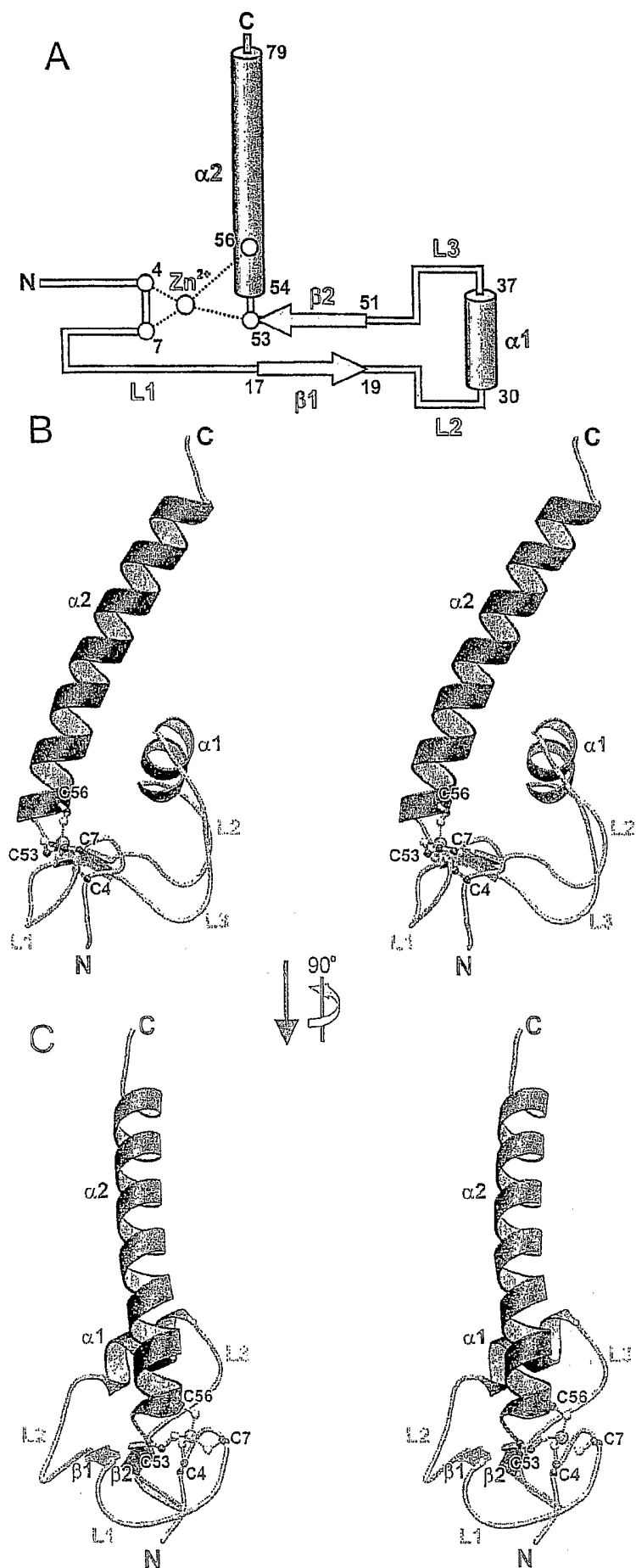


Fig. 4

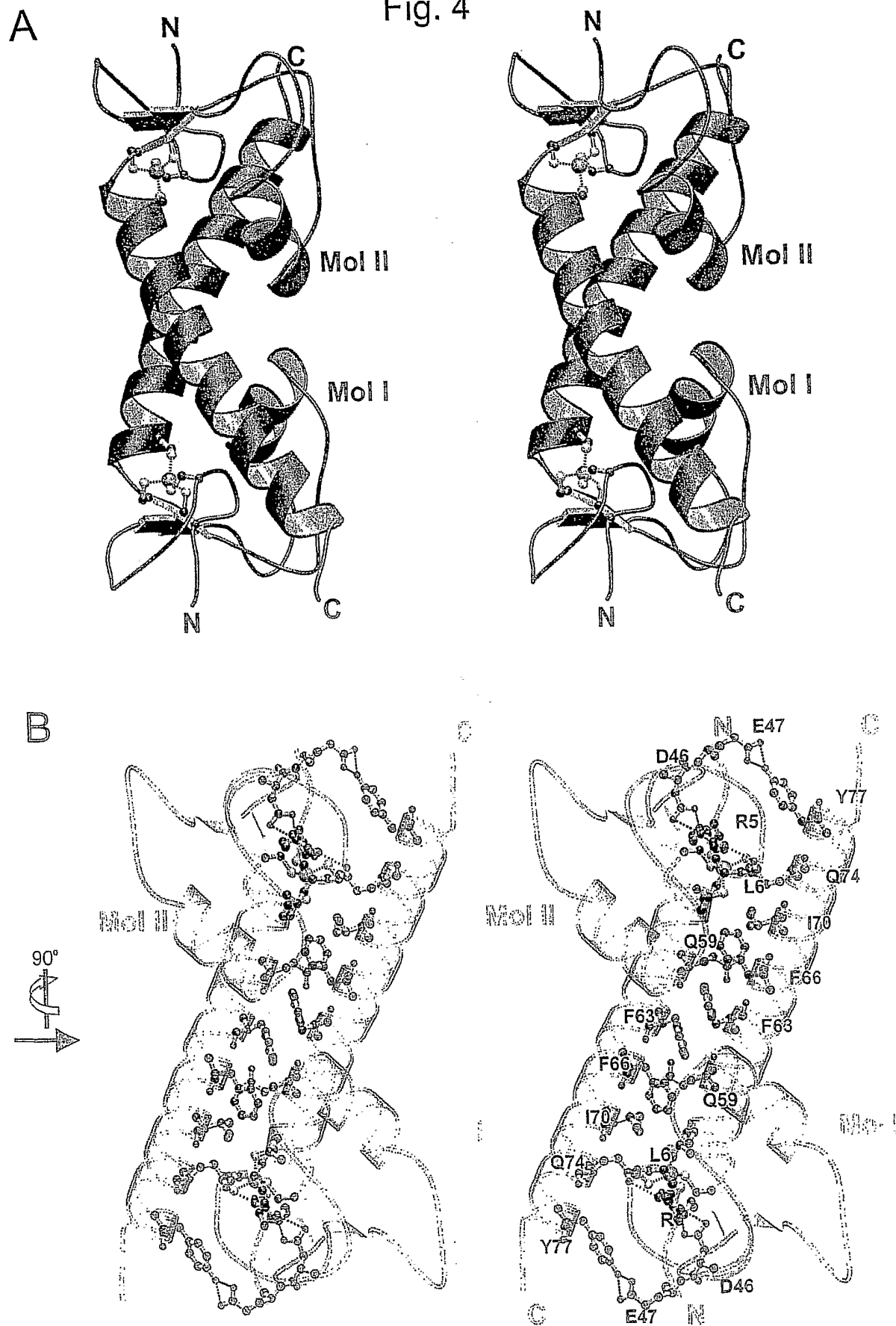


Fig. 5A

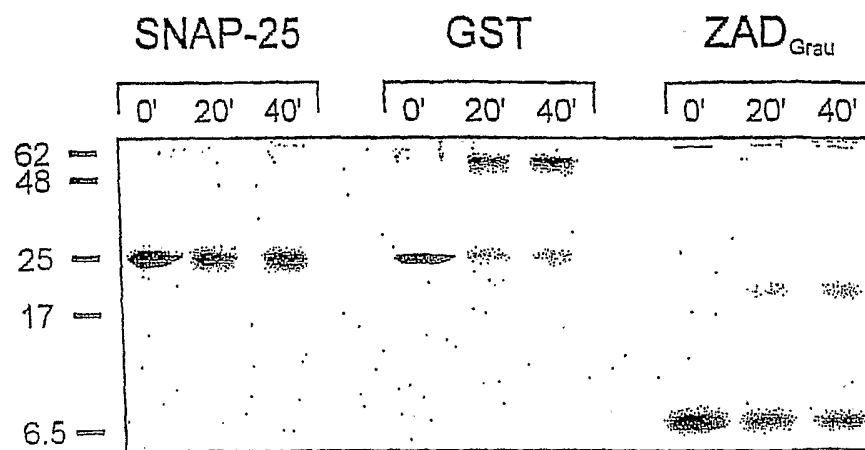
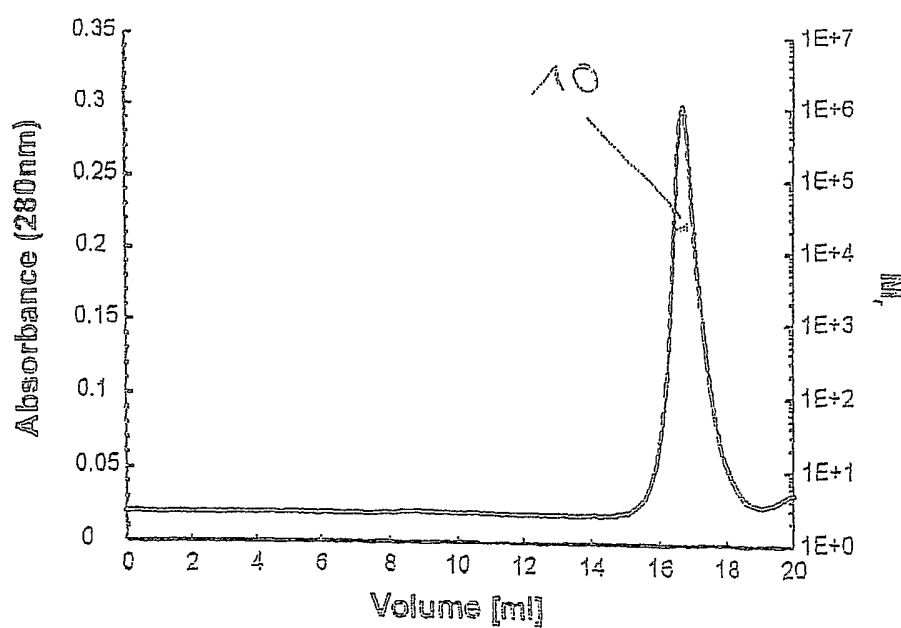


Fig. 5B



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Fig. 6

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CRYST1  48.660  48.660  82.123  90.00  90.00  90.00 P 41 21 2
SCALE1  0.020551  0.000000  0.000000  0.000000
SCALE2  0.000000  0.020551  0.000000  0.000000
SCALE3  0.000000  0.000000  0.012177  0.000000

```

	atom type	resi- due		X	Y	Z	B	
ATOM	1	N	ASP A	2	4.127	9.804	15.493	1.00 60.05 A
ATOM	2	CA	ASP A	2	4.430	10.543	16.755	1.00 59.95 A
ATOM	3	CB	ASP A	2	3.158	11.194	17.327	1.00 61.69 A
ATOM	4	CG	ASP A	2	3.397	12.615	17.849	1.00 66.39 A
ATOM	5	OD1	ASP A	2	2.414	13.394	17.912	1.00 69.89 A
ATOM	6	OD2	ASP A	2	4.522	13.041	18.219	1.00 71.87 A
ATOM	7	C	ASP A	2	5.059	9.598	17.779	1.00 56.76 A
ATOM	8	O	ASP A	2	5.001	8.379	17.634	1.00 56.57 A
ATOM	9	N	ILE A	3	5.658	10.168	18.815	1.00 52.94 A
ATOM	10	CA	ILE A	3	6.392	9.378	19.801	1.00 50.35 A
ATOM	11	CB	ILE A	3	7.887	9.266	19.337	1.00 50.48 A
ATOM	12	CG1	ILE A	3	8.412	7.854	19.556	1.00 52.49 A
ATOM	13	CD1	ILE A	3	7.972	6.873	18.495	1.00 53.33 A
ATOM	14	CG2	ILE A	3	8.797	10.289	20.016	1.00 52.35 A
ATOM	15	C	ILE A	3	6.229	9.977	21.199	1.00 46.02 A
ATOM	16	O	ILE A	3	6.093	11.194	21.341	1.00 45.03 A
ATOM	17	N	CYS A	4	6.197	9.129	22.223	1.00 42.18 A
ATOM	18	CA	CYS A	4	6.133	9.608	23.604	1.00 39.45 A
ATOM	19	CB	CYS A	4	5.819	8.449	24.571	1.00 39.90 A
ATOM	20	SG	CYS A	4	5.905	8.787	26.359	1.00 31.86 A
ATOM	21	C	CYS A	4	7.472	10.289	23.940	1.00 39.86 A
ATOM	22	O	CYS A	4	8.541	9.716	23.744	1.00 39.17 A
ATOM	23	N	ARG A	5	7.412	11.531	24.403	1.00 39.50 A
ATOM	24	CA	ARG A	5	8.604	12.275	24.799	1.00 39.00 A
ATOM	25	CB	ARG A	5	8.208	13.662	25.318	1.00 39.03 A
ATOM	26	CG	ARG A	5	9.389	14.540	25.765	1.00 40.88 A
ATOM	27	CD	ARG A	5	10.255	15.051	24.624	1.00 43.49 A
ATOM	28	NE	ARG A	5	11.434	15.773	25.103	1.00 46.29 A
ATOM	29	CZ	ARG A	5	12.573	15.193	25.483	1.00 47.52 A
ATOM	30	NH1	ARG A	5	13.585	15.942	25.895	1.00 48.38 A
ATOM	31	NH2	ARG A	5	12.705	13.877	25.466	1.00 45.55 A
ATOM	32	C	ARG A	5	9.466	11.553	25.855	1.00 38.16 A
ATOM	33	O	ARG A	5	10.684	11.700	25.861	1.00 38.85 A
ATOM	34	N	LEU A	6	8.835	10.777	26.733	1.00 37.56 A
ATOM	35	CA	LEU A	6	9.549	10.073	27.788	1.00 36.87 A
ATOM	36	CB	LEU A	6	8.660	9.886	29.022	1.00 35.21 A
ATOM	37	CG	LEU A	6	8.989	10.808	30.176	1.00 36.58 A
ATOM	38	CD1	LEU A	6	8.778	12.246	29.755	1.00 40.92 A
ATOM	39	CD2	LEU A	6	8.116	10.445	31.371	1.00 35.17 A
ATOM	40	C	LEU A	6	10.120	8.741	27.330	1.00 37.32 A
ATOM	41	O	LEU A	6	11.337	8.574	27.347	1.00 34.58 A
ATOM	42	N	CYS A	7	9.254	7.794	26.959	1.00 37.92 A
ATOM	43	CA	CYS A	7	9.682	6.418	26.634	1.00 39.16 A
ATOM	44	CB	CYS A	7	8.584	5.392	26.987	1.00 41.01 A
ATOM	45	SG	CYS A	7	7.048	5.513	26.037	1.00 40.02 A
ATOM	46	C	CYS A	7	10.098	6.247	25.174	1.00 42.35 A
ATOM	47	O	CYS A	7	10.714	5.240	24.808	1.00 39.52 A
ATOM	48	N	LEU A	8	9.749	7.242	24.360	1.00 43.99 A
ATOM	49	CA	LEU A	8	10.093	7.294	22.939	1.00 47.28 A
ATOM	50	CB	LEU A	8	11.607	7.422	22.759	1.00 45.89 A
ATOM	51	CG	LEU A	8	12.230	8.608	23.505	1.00 46.45 A
ATOM	52	CD1	LEU A	8	13.733	8.470	23.537	1.00 47.98 A
ATOM	53	CD2	LEU A	8	11.843	9.928	22.894	1.00 46.08 A
ATOM	54	C	LEU A	8	9.528	6.122	22.134	1.00 49.95 A
ATOM	55	O	LEU A	8	10.025	5.809	21.053	1.00 50.23 A

	atom type	resi- due		X	Y	Z	B	
ATOM	56	N	ARG A	9	8.473	5.504	22.662	1.00 53.95 A
ATOM	57	CA	ARG A	9	7.827	4.372	22.022	1.00 57.13 A
ATOM	58	CB	ARG A	9	7.204	3.437	23.055	1.00 57.78 A
ATOM	59	CG	ARG A	9	8.154	2.410	23.665	1.00 60.08 A
ATOM	60	CD	ARG A	9	7.424	1.262	24.364	1.00 66.01 A
ATOM	61	NE	ARG A	9	6.412	0.653	23.485	1.00 71.08 A
ATOM	62	CZ	ARG A	9	5.218	0.175	23.866	1.00 73.60 A
ATOM	63	NH1	ARG A	9	4.404	-0.341	22.954	1.00 73.09 A
ATOM	64	NH2	ARG A	9	4.825	0.195	25.137	1.00 75.92 A
ATOM	65	C	ARG A	9	6.747	4.917	21.109	1.00 59.85 A
ATOM	66	O	ARG A	9	6.343	6.070	21.249	1.00 58.67 A
ATOM	67	N	GLY A	10	6.282	4.079	20.182	1.00 63.33 A
ATOM	68	CA	GLY A	10	5.299	4.478	19.192	1.00 65.73 A
ATOM	69	C	GLY A	10	3.971	4.921	19.774	1.00 68.43 A
ATOM	70	O	GLY A	10	3.373	4.209	20.577	1.00 67.38 A
ATOM	71	N	VAL A	11	3.527	6.112	19.375	1.00 72.73 A
ATOM	72	CA	VAL A	11	2.187	6.608	19.700	1.00 76.73 A
ATOM	73	CB	VAL A	11	2.132	8.167	19.743	1.00 77.44 A
ATOM	74	CG1	VAL A	11	0.691	8.694	19.579	1.00 78.14 A
ATOM	75	CG2	VAL A	11	2.755	8.681	21.033	1.00 77.21 A
ATOM	76	C	VAL A	11	1.178	6.048	18.687	1.00 79.37 A
ATOM	77	O	VAL A	11	1.293	6.264	17.474	1.00 78.68 A
ATOM	78	N	SER A	12	0.196	5.329	19.222	1.00 82.65 A
ATOM	79	CA	SER A	12	-0.825	4.649	18.435	1.00 85.39 A
ATOM	80	CB	SER A	12	-1.760	3.896	19.386	1.00 85.86 A
ATOM	81	OG	SER A	12	-1.064	2.870	20.076	1.00 87.33 A
ATOM	82	C	SER A	12	-1.620	5.632	17.576	1.00 86.73 A
ATOM	83	O	SER A	12	-1.729	5.463	16.359	1.00 86.95 A
ATOM	84	N	GLY A	13	-2.158	6.662	18.224	1.00 88.54 A
ATOM	85	CA	GLY A	13	-2.945	7.686	17.553	1.00 89.65 A
ATOM	86	C	GLY A	13	-2.778	9.039	18.220	1.00 90.39 A
ATOM	87	O	GLY A	13	-2.266	9.978	17.607	1.00 90.82 A
ATOM	88	N	ALA A	14	-3.203	9.115	19.482	1.00 90.79 A
ATOM	89	CA	ALA A	14	-3.143	10.335	20.307	1.00 90.86 A
ATOM	90	CB	ALA A	14	-3.435	11.580	19.491	1.00 91.08 A
ATOM	91	C	ALA A	14	-4.087	10.285	21.510	1.00 90.47 A
ATOM	92	O	ALA A	14	-3.947	11.080	22.440	1.00 90.74 A
ATOM	93	N	GLN A	15	-5.055	9.373	21.484	1.00 89.49 A
ATOM	94	CA	GLN A	15	-5.948	9.176	22.621	1.00 88.68 A
ATOM	95	CB	GLN A	15	-7.325	8.767	22.136	1.00 89.58 A
ATOM	96	CG	GLN A	15	-8.426	9.326	22.979	1.00 92.04 A
ATOM	97	CD	GLN A	15	-9.649	9.545	22.167	1.00 95.11 A
ATOM	98	OE1	GLN A	15	-10.265	8.585	21.716	1.00 97.57 A
ATOM	99	NE2	GLN A	15	-9.997	10.808	21.933	1.00 98.05 A
ATOM	100	C	GLN A	15	-5.444	8.137	23.617	1.00 86.16 A
ATOM	101	O	GLN A	15	-6.217	7.608	24.416	1.00 86.26 A
ATOM	102	N	MET A	16	-4.160	7.812	23.533	1.00 83.00 A
ATOM	103	CA	MET A	16	-3.462	7.165	24.629	1.00 80.24 A
ATOM	104	CB	MET A	16	-2.628	5.988	24.111	1.00 81.05 A
ATOM	105	CG	MET A	16	-3.422	4.697	23.923	1.00 83.73 A
ATOM	106	SD	MET A	16	-2.506	3.224	24.460	1.00 88.16 A
ATOM	107	CE	MET A	16	-1.046	3.335	23.429	1.00 87.21 A
ATOM	108	C	MET A	16	-2.566	8.211	25.293	1.00 76.23 A
ATOM	109	O	MET A	16	-1.880	7.906	26.255	1.00 74.85 A
ATOM	110	N	CYS A	17	-2.623	9.449	24.795	1.00 71.74 A
ATOM	111	CA	CYS A	17	-1.635	10.482	25.099	1.00 67.91 A
ATOM	112	CB	CYS A	17	-0.836	10.804	23.842	1.00 67.67 A
ATOM	113	SG	CYS A	17	0.175	9.427	23.299	1.00 67.86 A
ATOM	114	C	CYS A	17	-2.227	11.781	25.619	1.00 64.31 A

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	atom type	resi- due		X	Y	Z	B	
ATOM	115	O	CYS A 17	-3.320	12.193	25.222	1.00	64.70 A
ATOM	116	N	LEU A 18	-1.476	12.420	26.509	1.00	59.23 A
ATOM	117	CA	LEU A 18	-1.731	13.780	26.941	1.00	56.14 A
ATOM	118	CB	LEU A 18	-1.403	13.958	28.428	1.00	55.36 A
ATOM	119	CG	LEU A 18	-2.282	13.202	29.424	1.00	55.81 A
ATOM	120	CD1	LEU A 18	-2.002	11.701	29.410	1.00	55.00 A
ATOM	121	CD2	LEU A 18	-2.109	13.766	30.829	1.00	55.54 A
ATOM	122	C	LEU A 18	-0.825	14.671	26.118	1.00	53.54 A
ATOM	123	O	LEU A 18	0.310	14.302	25.814	1.00	51.17 A
ATOM	124	N	GLN A 19	-1.326	15.840	25.742	1.00	52.69 A
ATOM	125	CA	GLN A 19	-0.478	16.859	25.140	1.00	52.24 A
ATOM	126	CB	GLN A 19	-1.300	17.888	24.351	1.00	52.87 A
ATOM	127	CG	GLN A 19	-2.141	17.305	23.226	1.00	53.35 A
ATOM	128	CD	GLN A 19	-1.311	16.781	22.078	1.00	53.77 A
ATOM	129	OE1	GLN A 19	-0.533	17.520	21.469	1.00	52.38 A
ATOM	130	NE2	GLN A 19	-1.488	15.506	21.763	1.00	56.34 A
ATOM	131	C	GLN A 19	0.259	17.524	26.287	1.00	51.03 A
ATOM	132	O	GLN A 19	-0.358	17.977	27.246	1.00	49.00 A
ATOM	133	N	ILE A 20	1.586	17.522	26.221	1.00	51.07 A
ATOM	134	CA	ILE A 20	2.390	18.219	27.218	1.00	51.49 A
ATOM	135	CB	ILE A 20	3.908	17.902	27.058	1.00	51.23 A
ATOM	136	CG1	ILE A 20	4.164	16.397	27.255	1.00	49.93 A
ATOM	137	CD1	ILE A 20	5.535	15.918	26.813	1.00	49.46 A
ATOM	138	CG2	ILE A 20	4.736	18.716	28.056	1.00	47.20 A
ATOM	139	C	ILE A 20	2.107	19.705	27.062	1.00	54.37 A
ATOM	140	O	ILE A 20	1.857	20.404	28.032	1.00	54.41 A
ATOM	141	N	PHE A 21	2.127	20.151	25.814	1.00	57.90 A
ATOM	142	CA	PHE A 21	1.793	21.517	25.430	1.00	61.34 A
ATOM	143	CB	PHE A 21	3.028	22.189	24.817	1.00	59.88 A
ATOM	144	CG	PHE A 21	4.178	22.344	25.783	1.00	57.44 A
ATOM	145	CD1	PHE A 21	5.264	21.474	25.742	1.00	53.83 A
ATOM	146	CE1	PHE A 21	6.330	21.616	26.633	1.00	52.84 A
ATOM	147	CZ	PHE A 21	6.306	22.627	27.585	1.00	52.22 A
ATOM	148	CE2	PHE A 21	5.223	23.498	27.638	1.00	53.82 A
ATOM	149	CD2	PHE A 21	4.169	23.355	26.739	1.00	54.93 A
ATOM	150	C	PHE A 21	0.636	21.507	24.420	1.00	65.60 A
ATOM	151	O	PHE A 21	0.449	20.534	23.690	1.00	66.15 A
ATOM	152	N	ASP A 22	-0.138	22.584	24.373	1.00	70.75 A
ATOM	153	CA	ASP A 22	-1.235	22.689	23.406	1.00	74.59 A
ATOM	154	CB	ASP A 22	-2.568	22.229	24.031	1.00	75.00 A
ATOM	155	CG	ASP A 22	-3.700	22.100	23.002	1.00	74.99 A
ATOM	156	OD1	ASP A 22	-3.765	21.061	22.304	1.00	74.01 A
ATOM	157	OD2	ASP A 22	-4.576	22.980	22.835	1.00	72.50 A
ATOM	158	C	ASP A 22	-1.346	24.114	22.880	1.00	77.38 A
ATOM	159	O	ASP A 22	-1.819	25.002	23.586	1.00	77.23 A
ATOM	160	N	VAL A 23	-0.849	24.315	21.657	1.00	81.50 A
ATOM	161	CA	VAL A 23	-1.135	25.496	20.832	1.00	84.24 A
ATOM	162	CB	VAL A 23	-2.480	25.310	20.094	1.00	85.04 A
ATOM	163	CG1	VAL A 23	-2.691	26.420	19.108	1.00	86.19 A
ATOM	164	CG2	VAL A 23	-2.529	23.956	19.386	1.00	85.96 A
ATOM	165	C	VAL A 23	-1.160	26.834	21.588	1.00	86.03 A
ATOM	166	O	VAL A 23	-2.009	27.047	22.456	1.00	86.39 A
ATOM	167	N	ASP A 24	-0.268	27.753	21.211	1.00	88.17 A
ATOM	168	CA	ASP A 24	-0.014	28.960	22.004	1.00	89.51 A
ATOM	169	CB	ASP A 24	-1.172	29.970	21.912	1.00	89.79 A
ATOM	170	CG	ASP A 24	-0.803	31.216	21.112	1.00	90.98 A
ATOM	171	OD1	ASP A 24	-1.252	32.322	21.488	1.00	92.25 A
ATOM	172	OD2	ASP A 24	-0.072	31.188	20.096	1.00	91.13 A
ATOM	173	C	ASP A 24	0.236	28.483	23.428	1.00	89.65 A
ATOM	174	O	ASP A 24	-0.551	28.715	24.348	1.00	89.93 A

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	atom type	resi- due		X	Y	Z	B	
ATOM	175	N	SER A 25	1.355	27.789	23.577	1.00	89.90 A
ATOM	176	CA	SER A 25	1.629	26.989	24.756	1.00	89.63 A
ATOM	177	CB	SER A 25	2.502	25.795	24.361	1.00	90.03 A
ATOM	178	OG	SER A 25	1.926	25.089	23.272	1.00	90.77 A
ATOM	179	C	SER A 25	2.260	27.845	25.857	1.00	88.72 A
ATOM	180	O	SER A 25	1.734	28.912	26.178	1.00	88.90 A
ATOM	181	N	GLY A 26	3.378	27.397	26.429	1.00	87.66 A
ATOM	182	CA	GLY A 26	3.926	28.014	27.623	1.00	86.17 A
ATOM	183	C	GLY A 26	3.014	27.697	28.793	1.00	84.40 A
ATOM	184	O	GLY A 26	1.795	27.852	28.693	1.00	84.01 A
ATOM	185	N	GLU A 27	3.596	27.251	29.903	1.00	82.05 A
ATOM	186	CA	GLU A 27	2.823	26.856	31.078	1.00	80.39 A
ATOM	187	CB	GLU A 27	1.935	28.009	31.581	1.00	80.87 A
ATOM	188	CG	GLU A 27	2.613	29.375	31.620	1.00	82.54 A
ATOM	189	CD	GLU A 27	1.647	30.496	31.950	1.00	85.05 A
ATOM	190	OE1	GLU A 27	1.093	30.489	33.070	1.00	86.97 A
ATOM	191	OE2	GLU A 27	1.439	31.382	31.091	1.00	85.38 A
ATOM	192	C	GLU A 27	1.974	25.628	30.739	1.00	77.04 A
ATOM	193	O	GLU A 27	1.097	25.688	29.875	1.00	77.16 A
ATOM	194	N	SER A 28	2.238	24.514	31.411	1.00	72.50 A
ATOM	195	CA	SER A 28	1.578	23.257	31.064	1.00	69.06 A
ATOM	196	CB	SER A 28	2.543	22.382	30.261	1.00	68.90 A
ATOM	197	OG	SER A 28	2.078	21.044	30.185	1.00	67.75 A
ATOM	198	C	SER A 28	1.017	22.458	32.245	1.00	65.71 A
ATOM	199	O	SER A 28	0.019	21.746	32.078	1.00	67.71 A
ATOM	200	N	LYS A 29	1.634	22.579	33.418	1.00	59.43 A
ATOM	201	CA	LYS A 29	1.379	21.693	34.567	1.00	55.74 A
ATOM	202	CB	LYS A 29	-0.108	21.540	34.914	1.00	56.61 A
ATOM	203	CG	LYS A 29	-0.831	22.865	35.149	1.00	60.80 A
ATOM	204	CD	LYS A 29	-2.022	22.699	36.105	1.00	62.69 A
ATOM	205	CE	LYS A 29	-2.466	24.030	36.702	1.00	63.22 A
ATOM	206	NZ	LYS A 29	-2.485	25.121	35.680	1.00	61.81 A
ATOM	207	C	LYS A 29	2.011	20.333	34.342	1.00	50.02 A
ATOM	208	O	LYS A 29	2.742	19.862	35.192	1.00	48.69 A
ATOM	209	N	VAL A 30	1.729	19.698	33.201	1.00	45.10 A
ATOM	210	CA	VAL A 30	2.390	18.448	32.846	1.00	40.76 A
ATOM	211	CB	VAL A 30	1.862	17.836	31.535	1.00	39.38 A
ATOM	212	CG1	VAL A 30	2.743	16.678	31.083	1.00	38.09 A
ATOM	213	CG2	VAL A 30	0.437	17.346	31.717	1.00	38.45 A
ATOM	214	C	VAL A 30	3.884	18.717	32.734	1.00	39.23 A
ATOM	215	O	VAL A 30	4.676	17.998	33.330	1.00	34.91 A
ATOM	216	N	ALA A 31	4.267	19.773	32.017	1.00	37.89 A
ATOM	217	CA	ALA A 31	5.687	20.098	31.861	1.00	36.58 A
ATOM	218	CB	ALA A 31	5.876	21.286	30.937	1.00	38.90 A
ATOM	219	C	ALA A 31	6.358	20.365	33.205	1.00	35.61 A
ATOM	220	O	ALA A 31	7.483	19.941	33.438	1.00	36.22 A
ATOM	221	N	GLU A 32	5.649	21.074	34.073	1.00	34.56 A
ATOM	222	CA	GLU A 32	6.118	21.468	35.391	1.00	34.27 A
ATOM	223	CB	GLU A 32	5.028	22.319	36.042	1.00	36.13 A
ATOM	224	CG	GLU A 32	5.411	23.052	37.315	1.00	45.30 A
ATOM	225	CD	GLU A 32	4.314	24.005	37.781	1.00	54.51 A
ATOM	226	OE1	GLU A 32	3.680	24.662	36.921	1.00	56.34 A
ATOM	227	OE2	GLU A 32	4.078	24.087	39.013	1.00	60.06 A
ATOM	228	C	GLU A 32	6.420	20.241	36.251	1.00	32.64 A
ATOM	229	O	GLU A 32	7.425	20.188	36.951	1.00	30.26 A
ATOM	230	N	VAL A 33	5.556	19.234	36.170	1.00	31.17 A
ATOM	231	CA	VAL A 33	5.726	18.010	36.953	1.00	30.24 A
ATOM	232	CB	VAL A 33	4.444	17.174	36.940	1.00	29.99 A
ATOM	233	CG1	VAL A 33	4.673	15.814	37.531	1.00	29.90 A
ATOM	234	CG2	VAL A 33	3.360	17.892	37.722	1.00	29.92 A

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		atom type	resi- due		X	Y	Z	B	
ATOM	235	C	VAL A 33		6.887	17.198	36.413	1.00	31.55 A
ATOM	236	O	VAL A 33		7.709	16.710	37.171	1.00	31.64 A
ATOM	237	N	LEU A 34		6.958	17.062	35.102	1.00	31.46 A
ATOM	238	CA	LEU A 34		8.066	16.358	34.464	1.00	32.67 A
ATOM	239	CB	LEU A 34		7.876	16.331	32.951	1.00	33.11 A
ATOM	240	CG	LEU A 34		6.696	15.532	32.429	1.00	32.39 A
ATOM	241	CD1	LEU A 34		6.585	15.733	30.920	1.00	37.14 A
ATOM	242	CD2	LEU A 34		6.871	14.107	32.762	1.00	36.20 A
ATOM	243	C	LEU A 34		9.414	17.019	34.765	1.00	33.32 A
ATOM	244	O	LEU A 34		10.406	16.334	34.968	1.00	33.57 A
ATOM	245	N	ARG A 35		9.443	18.339	34.823	1.00	31.78 A
ATOM	246	CA	ARG A 35		10.682	19.062	35.108	1.00	34.58 A
ATOM	247	CB	ARG A 35		10.485	20.552	34.923	1.00	35.54 A
ATOM	248	CG	ARG A 35		10.584	21.003	33.491	1.00	42.87 A
ATOM	249	CD	ARG A 35		10.845	22.473	33.349	1.00	46.05 A
ATOM	250	NE	ARG A 35		9.620	23.240	33.532	1.00	52.68 A
ATOM	251	CZ	ARG A 35		8.807	23.620	32.543	1.00	54.66 A
ATOM	252	NH1	ARG A 35		7.709	24.306	32.831	1.00	56.50 A
ATOM	253	NH2	ARG A 35		9.089	23.352	31.269	1.00	57.59 A
ATOM	254	C	ARG A 35		11.184	18.799	36.528	1.00	33.68 A
ATOM	255	O	ARG A 35		12.385	18.806	36.767	1.00	35.08 A
ATOM	256	N	GLN A 36		10.244	18.580	37.446	1.00	33.18 A
ATOM	257	CA	GLN A 36		10.539	18.278	38.843	1.00	33.98 A
ATOM	258	CB	GLN A 36		9.260	18.290	39.681	1.00	34.34 A
ATOM	259	CG	GLN A 36		8.692	19.660	39.900	1.00	39.56 A
ATOM	260	CD	GLN A 36		7.537	19.669	40.884	1.00	41.72 A
ATOM	261	OE1	GLN A 36		7.708	19.338	42.060	1.00	40.43 A
ATOM	262	NE2	GLN A 36		6.360	20.060	40.405	1.00	43.50 A
ATOM	263	C	GLN A 36		11.214	16.943	39.064	1.00	33.01 A
ATOM	264	O	GLN A 36		11.918	16.800	40.040	1.00	32.32 A
ATOM	265	N	HIS A 37		10.966	15.959	38.192	1.00	32.52 A
ATOM	266	CA	HIS A 37		11.457	14.601	38.393	1.00	33.06 A
ATOM	267	CB	HIS A 37		10.271	13.648	38.466	1.00	34.20 A
ATOM	268	CG	HIS A 37		9.276	14.054	39.497	1.00	24.88 A
ATOM	269	ND1	HIS A 37		9.574	14.069	40.835	1.00	25.41 A
ATOM	270	CE1	HIS A 37		8.529	14.506	41.511	1.00	26.25 A
ATOM	271	NE2	HIS A 37		7.567	14.784	40.652	1.00	22.09 A
ATOM	272	CD2	HIS A 37		8.019	14.526	39.385	1.00	26.03 A
ATOM	273	C	HIS A 37		12.444	14.070	37.377	1.00	35.50 A
ATOM	274	O	HIS A 37		12.953	12.988	37.570	1.00	34.62 A
ATOM	275	N	PHE A 38		12.712	14.805	36.304	1.00	34.41 A
ATOM	276	CA	PHE A 38		13.683	14.359	35.295	1.00	37.50 A
ATOM	277	CB	PHE A 38		12.969	13.966	33.992	1.00	37.88 A
ATOM	278	CG	PHE A 38		12.060	12.786	34.165	1.00	33.62 A
ATOM	279	CD1	PHE A 38		10.743	12.970	34.510	1.00	31.08 A
ATOM	280	CE1	PHE A 38		9.920	11.891	34.725	1.00	33.15 A
ATOM	281	CZ	PHE A 38		10.413	10.613	34.598	1.00	33.10 A
ATOM	282	CE2	PHE A 38		11.738	10.412	34.260	1.00	29.70 A
ATOM	283	CD2	PHE A 38		12.554	11.496	34.056	1.00	33.78 A
ATOM	284	C	PHE A 38		14.788	15.414	35.067	1.00	39.75 A
ATOM	285	O	PHE A 38		14.650	16.559	35.484	1.00	41.92 A
ATOM	286	N	TRP A 39		15.873	14.989	34.420	1.00	43.79 A
ATOM	287	CA	TRP A 39		17.144	15.748	34.325	1.00	47.59 A
ATOM	288	CB	TRP A 39		18.361	14.794	34.407	1.00	47.78 A
ATOM	289	CG	TRP A 39		18.343	13.796	33.307	1.00	52.09 A
ATOM	290	CD1	TRP A 39		19.036	13.849	32.128	1.00	53.75 A
ATOM	291	NE1	TRP A 39		18.699	12.775	31.338	1.00	57.17 A
ATOM	292	CE2	TRP A 39		17.774	12.007	31.995	1.00	56.79 A
ATOM	293	CD2	TRP A 39		17.519	12.626	33.236	1.00	55.50 A
ATOM	294	CE3	TRP A 39		16.577	12.035	34.101	1.00	56.33 A

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	atom type	resi- due		X	Y	Z	B	
ATOM	295	CZ3 TRP A	39	15.943	10.870	33.706	1.00	57.38 A
ATOM	296	CH2 TRP A	39	16.222	10.283	32.465	1.00	58.15 A
ATOM	297	CZ2 TRP A	39	17.128	10.838	31.594	1.00	57.00 A
ATOM	298	C TRP A	39	17.224	16.571	33.040	1.00	49.87 A
ATOM	299	O TRP A	39	17.997	17.522	32.954	1.00	50.49 A
ATOM	300	N PHE A	40	16.438	16.192	32.043	1.00	52.96 A
ATOM	301	CA PHE A	40	16.341	16.948	30.797	1.00	56.31 A
ATOM	302	CB PHE A	40	16.150	16.016	29.579	1.00	56.17 A
ATOM	303	CG PHE A	40	15.052	14.999	29.736	1.00	55.22 A
ATOM	304	CD1 PHE A	40	13.728	15.336	29.501	1.00	55.38 A
ATOM	305	CE1 PHE A	40	12.715	14.389	29.645	1.00	57.09 A
ATOM	306	CZ PHE A	40	13.024	13.106	30.015	1.00	52.76 A
ATOM	307	CE2 PHE A	40	14.348	12.758	30.245	1.00	54.74 A
ATOM	308	CD2 PHE A	40	15.348	13.697	30.097	1.00	54.62 A
ATOM	309	C PHE A	40	15.215	17.983	30.878	1.00	59.42 A
ATOM	310	O PHE A	40	14.438	18.004	31.831	1.00	58.67 A
ATOM	311	N GLU A	41	15.153	18.849	29.870	1.00	64.01 A
ATOM	312	CA GLU A	41	14.176	19.933	29.828	1.00	67.03 A
ATOM	313	CB GLU A	41	14.857	21.232	29.360	1.00	68.29 A
ATOM	314	CG GLU A	41	13.946	22.449	29.183	1.00	72.51 A
ATOM	315	CD GLU A	41	13.107	22.781	30.413	1.00	76.50 A
ATOM	316	OE1 GLU A	41	12.201	23.638	30.299	1.00	78.93 A
ATOM	317	OE2 GLU A	41	13.342	22.194	31.491	1.00	77.62 A
ATOM	318	C GLU A	41	12.987	19.581	28.932	1.00	67.98 A
ATOM	319	O GLU A	41	13.112	18.824	27.966	1.00	67.60 A
ATOM	320	N VAL A	42	11.828	20.128	29.278	1.00	69.60 A
ATOM	321	CA VAL A	42	10.620	19.944	28.495	1.00	71.46 A
ATOM	322	CB VAL A	42	9.502	19.274	29.330	1.00	72.11 A
ATOM	323	CG1 VAL A	42	8.648	18.379	28.441	1.00	72.52 A
ATOM	324	CG2 VAL A	42	10.084	18.469	30.500	1.00	71.39 A
ATOM	325	C VAL A	42	10.173	21.317	27.982	1.00	72.72 A
ATOM	326	O VAL A	42	9.779	22.182	28.767	1.00	72.25 A
ATOM	327	N LEU A	43	10.255	21.520	26.669	1.00	74.60 A
ATOM	328	CA LEU A	43	10.002	22.841	26.080	1.00	75.99 A
ATOM	329	CB LEU A	43	11.322	23.537	25.701	1.00	76.30 A
ATOM	330	CG LEU A	43	12.609	22.710	25.571	1.00	77.70 A
ATOM	331	CD1 LEU A	43	12.568	21.826	24.336	1.00	79.41 A
ATOM	332	CD2 LEU A	43	13.835	23.609	25.530	1.00	77.50 A
ATOM	333	C LEU A	43	9.058	22.784	24.873	1.00	76.63 A
ATOM	334	O LEU A	43	9.003	21.776	24.166	1.00	75.72 A
ATOM	335	N PRO A	44	8.320	23.872	24.644	1.00	78.58 A
ATOM	336	CA PRO A	44	7.348	23.951	23.545	1.00	79.91 A
ATOM	337	CB PRO A	44	7.103	25.458	23.406	1.00	80.11 A
ATOM	338	CG PRO A	44	7.335	26.013	24.769	1.00	79.60 A
ATOM	339	CD PRO A	44	8.347	25.119	25.434	1.00	78.80 A
ATOM	340	C PRO A	44	7.839	23.365	22.215	1.00	81.21 A
ATOM	341	O PRO A	44	7.099	22.595	21.596	1.00	81.86 A
ATOM	342	N ASN A	45	9.055	23.724	21.799	1.00	82.01 A
ATOM	343	CA ASN A	45	9.623	23.253	20.525	1.00	82.69 A
ATOM	344	CB ASN A	45	10.893	24.043	20.117	1.00	82.98 A
ATOM	345	CG ASN A	45	11.866	24.289	21.278	1.00	84.22 A
ATOM	346	OD1 ASN A	45	12.874	23.595	21.416	1.00	85.82 A
ATOM	347	ND2 ASN A	45	11.580	25.301	22.090	1.00	85.65 A
ATOM	348	C ASN A	45	9.859	21.733	20.491	1.00	82.85 A
ATOM	349	O ASN A	45	8.950	20.982	20.128	1.00	83.75 A
ATOM	350	N ASP A	46	11.054	21.286	20.881	1.00	82.38 A
ATOM	351	CA ASP A	46	11.430	19.872	20.877	1.00	81.57 A
ATOM	352	CB ASP A	46	10.596	19.071	21.901	1.00	81.95 A
ATOM	353	CG ASP A	46	11.384	18.701	23.148	1.00	81.75 A
ATOM	354	OD1 ASP A	46	12.589	18.383	23.035	1.00	79.94 A

	atom type	resi- due		X	Y	Z	B	
ATOM	355	OD2 ASP A	46	10.875	18.702	24.289	1.00	81.57 A
ATOM	356	C ASP A	46	11.340	19.215	19.493	1.00	80.57 A
ATOM	357	O ASP A	46	10.251	18.991	18.962	1.00	80.96 A
ATOM	358	N GLU A	47	12.500	18.899	18.925	1.00	78.83 A
ATOM	359	CA GLU A	47	12.594	17.995	17.779	1.00	77.54 A
ATOM	360	CB GLU A	47	14.078	17.841	17.380	1.00	78.40 A
ATOM	361	CG GLU A	47	14.421	16.783	16.322	1.00	80.80 A
ATOM	362	CD GLU A	47	15.313	15.645	16.834	1.00	82.13 A
ATOM	363	OE1 GLU A	47	15.599	14.719	16.044	1.00	83.06 A
ATOM	364	OE2 GLU A	47	15.738	15.660	18.012	1.00	80.83 A
ATOM	365	C GLU A	47	11.952	16.630	18.116	1.00	74.85 A
ATOM	366	O GLU A	47	11.508	15.903	17.222	1.00	74.39 A
ATOM	367	N ILE A	48	11.864	16.324	19.412	1.00	71.77 A
ATOM	368	CA ILE A	48	11.480	15.002	19.897	1.00	69.02 A
ATOM	369	CB ILE A	48	11.920	14.794	21.387	1.00	69.24 A
ATOM	370	CG1 ILE A	48	13.262	15.469	21.713	1.00	69.06 A
ATOM	371	CD1 ILE A	48	14.410	14.999	20.912	1.00	69.28 A
ATOM	372	CG2 ILE A	48	11.951	13.313	21.725	1.00	70.19 A
ATOM	373	C ILE A	48	9.975	14.781	19.804	1.00	66.61 A
ATOM	374	O ILE A	48	9.493	13.980	19.002	1.00	65.29 A
ATOM	375	N SER A	49	9.246	15.503	20.646	1.00	63.61 A
ATOM	376	CA SER A	49	7.832	15.249	20.887	1.00	61.20 A
ATOM	377	CB SER A	49	7.598	13.795	21.310	1.00	61.78 A
ATOM	378	OG SER A	49	6.232	13.548	21.582	1.00	59.95 A
ATOM	379	C SER A	49	7.316	16.175	21.978	1.00	58.78 A
ATOM	380	O SER A	49	8.075	16.669	22.814	1.00	59.16 A
ATOM	381	N LYS A	50	6.014	16.407	21.954	1.00	56.45 A
ATOM	382	CA LYS A	50	5.350	17.252	22.939	1.00	54.69 A
ATOM	383	CB LYS A	50	5.033	18.620	22.322	1.00	56.11 A
ATOM	384	CG LYS A	50	4.354	18.564	20.954	1.00	58.46 A
ATOM	385	CD LYS A	50	4.153	19.945	20.350	1.00	61.80 A
ATOM	386	CE LYS A	50	3.598	19.844	18.932	1.00	62.90 A
ATOM	387	NZ LYS A	50	3.162	21.159	18.370	1.00	64.95 A
ATOM	388	C LYS A	50	4.103	16.517	23.455	1.00	51.75 A
ATOM	389	O LYS A	50	3.128	17.123	23.872	1.00	49.88 A
ATOM	390	N VAL A	51	4.185	15.189	23.431	1.00	48.31 A
ATOM	391	CA VAL A	51	3.123	14.293	23.870	1.00	45.46 A
ATOM	392	CB VAL A	51	2.494	13.607	22.630	1.00	45.91 A
ATOM	393	CG1 VAL A	51	2.893	12.135	22.500	1.00	46.83 A
ATOM	394	CG2 VAL A	51	0.986	13.775	22.615	1.00	49.53 A
ATOM	395	C VAL A	51	3.734	13.273	24.848	1.00	40.72 A
ATOM	396	O VAL A	51	4.908	12.939	24.735	1.00	39.17 A
ATOM	397	N ILE A	52	2.947	12.800	25.801	1.00	37.74 A
ATOM	398	CA ILE A	52	3.361	11.746	26.722	1.00	35.24 A
ATOM	399	CB ILE A	52	3.716	12.352	28.128	1.00	36.66 A
ATOM	400	CG1 ILE A	52	4.245	11.266	29.081	1.00	36.29 A
ATOM	401	CD1 ILE A	52	4.728	11.781	30.422	1.00	35.48 A
ATOM	402	CG2 ILE A	52	2.526	13.095	28.752	1.00	36.92 A
ATOM	403	C ILE A	52	2.250	10.677	26.803	1.00	37.13 A
ATOM	404	O ILE A	52	1.075	10.990	27.003	1.00	35.15 A
ATOM	405	N CYS A	53	2.621	9.416	26.638	1.00	35.70 A
ATOM	406	CA CYS A	53	1.652	8.357	26.687	1.00	37.30 A
ATOM	407	CB CYS A	53	2.250	7.043	26.180	1.00	37.29 A
ATOM	408	SG CYS A	53	3.318	6.207	27.334	1.00	37.09 A
ATOM	409	C CYS A	53	1.092	8.240	28.099	1.00	37.01 A
ATOM	410	O CYS A	53	1.729	8.625	29.072	1.00	33.87 A
ATOM	411	N ASN A	54	-0.118	7.730	28.204	1.00	36.02 A
ATOM	412	CA ASN A	54	-0.787	7.602	29.494	1.00	37.66 A
ATOM	413	CB ASN A	54	-2.242	7.116	29.297	1.00	39.17 A
ATOM	414	CG ASN A	54	-3.008	6.935	30.623	1.00	48.07 A

	atom type	resi- due		X	Y	Z	B	
ATOM	415	OD1 ASN A	54	-3.035	5.838	31.208	1.00	54.87 A
ATOM	416	ND2 ASN A	54	-3.660	8.006	31.081	1.00	56.22 A
ATOM	417	C ASN A	54	-0.004	6.662	30.415	1.00	34.59 A
ATOM	418	O ASN A	54	-0.062	6.815	31.632	1.00	33.54 A
ATOM	419	N VAL A	55	0.716	5.695	29.836	1.00	32.04 A
ATOM	420	CA VAL A	55	1.507	4.743	30.616	1.00	32.93 A
ATOM	421	CB VAL A	55	2.117	3.627	29.716	1.00	33.55 A
ATOM	422	CG1 VAL A	55	3.159	2.810	30.479	1.00	34.83 A
ATOM	423	CG2 VAL A	55	1.022	2.707	29.170	1.00	37.11 A
ATOM	424	C VAL A	55	2.625	5.452	31.389	1.00	30.48 A
ATOM	425	O VAL A	55	2.883	5.156	32.562	1.00	30.85 A
ATOM	426	N CYS A	56	3.286	6.389	30.734	1.00	30.56 A
ATOM	427	CA CYS A	56	4.342	7.147	31.382	1.00	30.20 A
ATOM	428	CB CYS A	56	5.252	7.864	30.371	1.00	32.07 A
ATOM	429	SG CYS A	56	6.377	6.772	29.432	1.00	31.19 A
ATOM	430	C CYS A	56	3.712	8.151	32.321	1.00	29.45 A
ATOM	431	O CYS A	56	4.201	8.323	33.431	1.00	27.34 A
ATOM	432	N TRP A	57	2.625	8.801	31.894	1.00	28.32 A
ATOM	433	CA TRP A	57	2.024	9.839	32.718	1.00	28.77 A
ATOM	434	CB TRP A	57	0.978	10.676	31.959	1.00	29.49 A
ATOM	435	CG TRP A	57	0.286	11.703	32.852	1.00	29.82 A
ATOM	436	CD1 TRP A	57	-1.033	11.709	33.210	1.00	29.95 A
ATOM	437	NE1 TRP A	57	-1.286	12.766	34.048	1.00	26.98 A
ATOM	438	CE2 TRP A	57	-0.134	13.477	34.247	1.00	25.01 A
ATOM	439	CD2 TRP A	57	0.880	12.841	33.514	1.00	29.02 A
ATOM	440	CE3 TRP A	57	2.173	13.381	33.566	1.00	29.10 A
ATOM	441	CZ3 TRP A	57	2.396	14.526	34.331	1.00	28.08 A
ATOM	442	CH2 TRP A	57	1.363	15.114	35.059	1.00	27.48 A
ATOM	443	CZ2 TRP A	57	0.097	14.610	35.026	1.00	26.70 A
ATOM	444	C TRP A	57	1.454	9.252	34.002	1.00	28.36 A
ATOM	445	O TRP A	57	1.541	9.892	35.062	1.00	26.54 A
ATOM	446	N THR A	58	0.893	8.042	33.937	1.00	29.72 A
ATOM	447	CA THR A	58	0.348	7.431	35.150	1.00	30.72 A
ATOM	448	CB THR A	58	-0.539	6.140	34.883	1.00	35.87 A
ATOM	449	OG1 THR A	58	-0.952	5.599	36.144	1.00	38.43 A
ATOM	450	CG2 THR A	58	0.243	4.973	34.246	1.00	38.11 A
ATOM	451	C THR A	58	1.470	7.198	36.182	1.00	31.67 A
ATOM	452	O THR A	58	1.281	7.468	37.373	1.00	26.68 A
ATOM	453	N GLN A	59	2.640	6.761	35.707	1.00	28.82 A
ATOM	454	CA GLN A	59	3.783	6.484	36.569	1.00	28.69 A
ATOM	455	CB GLN A	59	4.878	5.764	35.777	1.00	27.98 A
ATOM	456	CG GLN A	59	4.474	4.423	35.178	1.00	38.30 A
ATOM	457	CD GLN A	59	5.345	3.276	35.630	1.00	44.73 A
ATOM	458	OE1 GLN A	59	5.444	2.993	36.823	1.00	55.93 A
ATOM	459	NE2 GLN A	59	5.966	2.598	34.676	1.00	54.06 A
ATOM	460	C GLN A	59	4.332	7.784	37.146	1.00	25.88 A
ATOM	461	O GLN A	59	4.668	7.864	38.312	1.00	26.24 A
ATOM	462	N VAL A	60	4.408	8.826	36.332	1.00	25.91 A
ATOM	463	CA VAL A	60	4.966	10.099	36.815	1.00	26.71 A
ATOM	464	CB VAL A	60	5.265	11.055	35.625	1.00	25.67 A
ATOM	465	CG1 VAL A	60	5.655	12.399	36.128	1.00	29.26 A
ATOM	466	CG2 VAL A	60	6.376	10.478	34.765	1.00	27.94 A
ATOM	467	C VAL A	60	4.019	10.781	37.795	1.00	25.89 A
ATOM	468	O VAL A	60	4.438	11.294	38.832	1.00	24.66 A
ATOM	469	N SER A	61	2.739	10.822	37.446	1.00	25.08 A
ATOM	470	CA SER A	61	1.739	11.502	38.264	1.00	24.86 A
ATOM	471	CB SER A	61	0.427	11.694	37.491	1.00	25.44 A
ATOM	472	OG SER A	61	-0.170	10.458	37.166	1.00	21.37 A
ATOM	473	C SER A	61	1.484	10.778	39.581	1.00	27.08 A
ATOM	474	O SER A	61	1.287	11.433	40.606	1.00	24.37 A

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	atom type	resi- due		X	Y	Z	B	
ATOM	475	N	GLU A 62	1.444	9.438	39.555	1.00	27.62 A
ATOM	476	CA	GLU A 62	1.402	8.654	40.795	1.00	27.21 A
ATOM	477	CB	GLU A 62	1.301	7.162	40.517	1.00	28.94 A
ATOM	478	CG	GLU A 62	-0.097	6.775	40.076	1.00	31.90 A
ATOM	479	CD	GLU A 62	-0.276	5.284	39.863	1.00	38.79 A
ATOM	480	OE1	GLU A 62	-1.405	4.897	39.499	1.00	35.88 A
ATOM	481	OE2	GLU A 62	0.698	4.515	40.055	1.00	41.82 A
ATOM	482	C	GLU A 62	2.616	8.944	41.670	1.00	26.16 A
ATOM	483	O	GLU A 62	2.470	9.171	42.870	1.00	25.07 A
ATOM	484	N	PHE A 63	3.808	8.974	41.093	1.00	26.92 A
ATOM	485	CA	PHE A 63	4.982	9.288	41.921	1.00	25.43 A
ATOM	486	CB	PHE A 63	6.305	9.074	41.218	1.00	25.39 A
ATOM	487	CG	PHE A 63	7.471	9.448	42.077	1.00	26.29 A
ATOM	488	CD1	PHE A 63	7.784	8.689	43.214	1.00	24.68 A
ATOM	489	CE1	PHE A 63	8.830	9.049	44.046	1.00	27.13 A
ATOM	490	CZ	PHE A 63	9.567	10.163	43.758	1.00	28.48 A
ATOM	491	CE2	PHE A 63	9.262	10.927	42.640	1.00	27.33 A
ATOM	492	CD2	PHE A 63	8.208	10.584	41.820	1.00	26.92 A
ATOM	493	C	PHE A 63	4.932	10.693	42.465	1.00	27.74 A
ATOM	494	O	PHE A 63	5.297	10.934	43.619	1.00	28.35 A
ATOM	495	N	HIS A 64	4.491	11.641	41.642	1.00	29.18 A
ATOM	496	CA	HIS A 64	4.434	13.040	42.053	1.00	28.44 A
ATOM	497	CB	HIS A 64	4.046	13.939	40.897	1.00	29.27 A
ATOM	498	CG	HIS A 64	4.088	15.385	41.254	1.00	30.82 A
ATOM	499	ND1	HIS A 64	5.219	16.150	41.099	1.00	32.31 A
ATOM	500	CE1	HIS A 64	4.974	17.378	41.516	1.00	36.19 A
ATOM	501	NE2	HIS A 64	3.734	17.427	41.967	1.00	32.97 A
ATOM	502	CD2	HIS A 64	3.166	16.185	41.831	1.00	32.80 A
ATOM	503	C	HIS A 64	3.468	13.269	43.205	1.00	31.21 A
ATOM	504	O	HIS A 64	3.767	14.009	44.133	1.00	33.81 A
ATOM	505	N	GLN A 65	2.314	12.616	43.149	1.00	29.00 A
ATOM	506	CA	GLN A 65	1.346	12.661	44.226	1.00	29.56 A
ATOM	507	CB	GLN A 65	0.149	11.797	43.858	1.00	31.04 A
ATOM	508	CG	GLN A 65	-1.116	12.205	44.506	1.00	35.67 A
ATOM	509	CD	GLN A 65	-2.245	11.286	44.129	1.00	40.39 A
ATOM	510	OE1	GLN A 65	-2.077	10.066	44.117	1.00	42.90 A
ATOM	511	NE2	GLN A 65	-3.393	11.862	43.796	1.00	45.31 A
ATOM	512	C	GLN A 65	1.982	12.153	45.524	1.00	30.23 A
ATOM	513	O	GLN A 65	1.785	12.727	46.600	1.00	29.41 A
ATOM	514	N	PHE A 66	2.740	11.068	45.418	1.00	29.42 A
ATOM	515	CA	PHE A 66	3.482	10.554	46.558	1.00	29.13 A
ATOM	516	CB	PHE A 66	4.169	9.233	46.200	1.00	28.55 A
ATOM	517	CG	PHE A 66	5.053	8.703	47.288	1.00	24.10 A
ATOM	518	CD1	PHE A 66	4.510	8.230	48.463	1.00	30.55 A
ATOM	519	CE1	PHE A 66	5.330	7.734	49.466	1.00	30.72 A
ATOM	520	CZ	PHE A 66	6.691	7.733	49.291	1.00	21.57 A
ATOM	521	CE2	PHE A 66	7.234	8.201	48.125	1.00	26.90 A
ATOM	522	CD2	PHE A 66	6.430	8.685	47.136	1.00	26.41 A
ATOM	523	C	PHE A 66	4.518	11.578	47.041	1.00	26.89 A
ATOM	524	O	PHE A 66	4.581	11.887	48.218	1.00	30.73 A
ATOM	525	N	TYR A 67	5.305	12.089	46.118	1.00	28.32 A
ATOM	526	CA	TYR A 67	6.365	13.074	46.381	1.00	28.45 A
ATOM	527	CB	TYR A 67	6.961	13.473	45.036	1.00	28.67 A
ATOM	528	CG	TYR A 67	8.148	14.442	44.987	1.00	30.80 A
ATOM	529	CD1	TYR A 67	9.451	13.954	44.910	1.00	32.40 A
ATOM	530	CE1	TYR A 67	10.548	14.795	44.814	1.00	35.98 A
ATOM	531	CZ	TYR A 67	10.357	16.166	44.726	1.00	41.70 A
ATOM	532	OH	TYR A 67	11.475	16.977	44.592	1.00	42.68 A
ATOM	533	CE2	TYR A 67	9.065	16.688	44.768	1.00	37.43 A

	atom type	resi- due		X	Y	Z	B	
ATOM	534	CD2	TYR A 67	7.958	15.813	44.867	1.00	35.11 A
ATOM	535	C	TYR A 67	5.834	14.286	47.137	1.00	32.00 A
ATOM	536	O	TYR A 67	6.492	14.786	48.061	1.00	33.13 A
ATOM	537	N	VAL A 68	4.633	14.739	46.760	1.00	35.21 A
ATOM	538	CA	VAL A 68	3.979	15.902	47.366	1.00	34.67 A
ATOM	539	CB	VAL A 68	2.765	16.398	46.501	1.00	36.68 A
ATOM	540	CG1	VAL A 68	1.882	17.375	47.275	1.00	38.44 A
ATOM	541	CG2	VAL A 68	3.269	17.033	45.210	1.00	36.52 A
ATOM	542	C	VAL A 68	3.574	15.585	48.795	1.00	34.19 A
ATOM	543	O	VAL A 68	3.726	16.428	49.691	1.00	34.51 A
ATOM	544	N	SER A 69	3.120	14.357	49.036	1.00	32.54 A
ATOM	545	CA	SER A 69	2.780	13.939	50.382	1.00	31.04 A
ATOM	546	CB	SER A 69	2.016	12.597	50.389	1.00	32.13 A
ATOM	547	OG	SER A 69	2.891	11.472	50.413	1.00	34.21 A
ATOM	548	C	SER A 69	4.016	13.891	51.271	1.00	29.19 A
ATOM	549	O	SER A 69	3.958	14.281	52.441	1.00	28.11 A
ATOM	550	N	ILE A 70	5.137	13.452	50.710	1.00	29.43 A
ATOM	551	CA	ILE A 70	6.394	13.402	51.452	1.00	27.93 A
ATOM	552	CB	ILE A 70	7.475	12.597	50.674	1.00	26.22 A
ATOM	553	CG1	ILE A 70	7.017	11.136	50.497	1.00	24.66 A
ATOM	554	CD1	ILE A 70	6.616	10.456	51.780	1.00	25.71 A
ATOM	555	CG2	ILE A 70	8.813	12.643	51.393	1.00	25.20 A
ATOM	556	C	ILE A 70	6.888	14.810	51.776	1.00	27.85 A
ATOM	557	O	ILE A 70	7.328	15.065	52.901	1.00	27.78 A
ATOM	558	N	GLN A 71	6.817	15.731	50.820	1.00	28.62 A
ATOM	559	CA	GLN A 71	7.259	17.097	51.083	1.00	29.96 A
ATOM	560	CB	GLN A 71	7.100	18.026	49.883	1.00	31.44 A
ATOM	561	CG	GLN A 71	7.276	19.495	50.333	1.00	36.64 A
ATOM	562	CD	GLN A 71	7.290	20.509	49.218	1.00	43.75 A
ATOM	563	OE1	GLN A 71	7.911	20.289	48.187	1.00	44.24 A
ATOM	564	NE2	GLN A 71	6.633	21.655	49.444	1.00	44.25 A
ATOM	565	C	GLN A 71	6.476	17.676	52.241	1.00	31.45 A
ATOM	566	O	GLN A 71	7.049	18.294	53.135	1.00	31.53 A
ATOM	567	N	GLU A 72	5.159	17.472	52.230	1.00	33.36 A
ATOM	568	CA	GLU A 72	4.318	17.994	53.291	1.00	35.02 A
ATOM	569	CB	GLU A 72	2.841	17.729	53.007	1.00	36.73 A
ATOM	570	CG	GLU A 72	1.965	18.866	53.515	1.00	47.50 A
ATOM	571	CD	GLU A 72	0.521	18.452	53.690	1.00	54.31 A
ATOM	572	OE1	GLU A 72	-0.153	18.244	52.652	1.00	60.10 A
ATOM	573	OE2	GLU A 72	0.080	18.324	54.860	1.00	55.68 A
ATOM	574	C	GLU A 72	4.709	17.452	54.667	1.00	32.88 A
ATOM	575	O	GLU A 72	4.798	18.211	55.621	1.00	34.81 A
ATOM	576	N	ALA A 73	4.968	16.151	54.766	1.00	32.54 A
ATOM	577	CA	ALA A 73	5.371	15.548	56.037	1.00	30.37 A
ATOM	578	CB	ALA A 73	5.444	14.037	55.904	1.00	32.23 A
ATOM	579	C	ALA A 73	6.707	16.098	56.521	1.00	29.96 A
ATOM	580	O	ALA A 73	6.898	16.365	57.714	1.00	30.04 A
ATOM	581	N	GLN A 74	7.631	16.287	55.587	1.00	28.00 A
ATOM	582	CA	GLN A 74	8.948	16.803	55.918	1.00	27.28 A
ATOM	583	CB	GLN A 74	9.878	16.655	54.719	1.00	26.25 A
ATOM	584	CG	GLN A 74	10.083	15.229	54.259	1.00	25.33 A
ATOM	585	CD	GLN A 74	10.866	14.368	55.243	1.00	27.94 A
ATOM	586	OE1	GLN A 74	11.699	14.878	55.980	1.00	30.36 A
ATOM	587	NE2	GLN A 74	10.595	13.062	55.246	1.00	29.65 A
ATOM	588	C	GLN A 74	8.876	18.270	56.348	1.00	25.81 A
ATOM	589	O	GLN A 74	9.610	18.702	57.229	1.00	26.25 A
ATOM	590	N	VAL A 75	8.009	19.036	55.693	1.00	29.55 A
ATOM	591	CA	VAL A 75	7.835	20.448	56.022	1.00	29.58 A
ATOM	592	CB	VAL A 75	6.926	21.140	54.970	1.00	29.35 A

	atom type	resi- due		X	Y	Z	B	
ATOM	593	CG1 VAL A	75	6.358	22.439	55.483	1.00	30.85 A
ATOM	594	CG2 VAL A	75	7.710	21.401	53.672	1.00	27.74 Å
ATOM	595	C VAL A	75	7.288	20.580	57.443	1.00	31.40 A
ATOM	596	O VAL A	75	7.814	21.350	58.255	1.00	31.99 A
ATOM	597	N ILE A	76	6.246	19.817	57.746	1.00	33.18 A
ATOM	598	CA ILE A	76	5.640	19.840	59.073	1.00	34.24 A
ATOM	599	CB ILE A	76	4.386	18.948	59.144	1.00	34.84 A
ATOM	600	CG1 ILE A	76	3.251	19.561	58.315	1.00	33.43 A
ATOM	601	CD1 ILE A	76	2.043	18.657	58.136	1.00	35.21 A
ATOM	602	CG2 ILE A	76	3.933	18.757	60.613	1.00	36.16 A
ATOM	603	C ILE A	76	6.653	19.431	60.128	1.00	35.07 A
ATOM	604	O ILE A	76	6.777	20.092	61.154	1.00	33.49 A
ATOM	605	N TYR A	77	7.419	18.432	59.918	1.00	35.31 A
ATOM	606	CA TYR A	77	8.400	17.948	60.883	1.00	36.86 A
ATOM	607	CB TYR A	77	9.035	16.644	60.384	1.00	37.48 A
ATOM	608	CG TYR A	77	9.986	15.987	61.365	1.00	38.43 A
ATOM	609	CD1 TYR A	77	9.515	15.112	62.342	1.00	36.57 A
ATOM	610	CE1 TYR A	77	10.385	14.499	63.241	1.00	40.56 A
ATOM	611	CZ TYR A	77	11.744	14.762	63.169	1.00	43.20 A
ATOM	612	OH TYR A	77	12.623	13.986	63.891	1.00	46.81 A
ATOM	613	CE2 TYR A	77	12.235	15.630	62.207	1.00	41.59 A
ATOM	614	CD2 TYR A	77	11.357	16.236	61.311	1.00	39.76 A
ATOM	615	C TYR A	77	9.483	18.992	61.154	1.00	36.62 A
ATOM	616	O TYR A	77	10.039	19.035	62.251	1.00	36.68 A
ATOM	617	N ALA A	78	9.989	19.578	60.073	1.00	36.79 A
ATOM	618	CA ALA A	78	11.039	20.587	60.156	1.00	37.14 A
ATOM	619	CB ALA A	78	11.636	20.832	58.777	1.00	36.37 A
ATOM	620	C ALA A	78	10.587	21.909	60.776	1.00	38.16 A
ATOM	621	O ALA A	78	11.418	22.703	61.223	1.00	39.75 A
ATOM	622	N THR A	79	9.280	22.153	60.783	1.00	40.05 A
ATOM	623	CA THR A	79	8.740	23.391	61.340	1.00	41.57 A
ATOM	624	CB THR A	79	7.765	24.079	60.351	1.00	40.83 A
ATOM	625	OG1 THR A	79	6.897	23.105	59.757	1.00	36.18 A
ATOM	626	CG2 THR A	79	8.541	24.785	59.250	1.00	38.62 A
ATOM	627	C THR A	79	8.060	23.233	62.701	1.00	45.70 A
ATOM	628	O THR A	79	7.735	24.234	63.346	1.00	46.79 A
ATOM	629	N THR A	80	8.124	22.023	63.249	1.00	50.79 A
ATOM	630	CA THR A	80	7.519	21.728	64.546	1.00	55.30 A
ATOM	631	CB THR A	80	6.543	20.526	64.449	1.00	56.08 A
ATOM	632	OG1 THR A	80	5.449	20.861	63.584	1.00	58.45 A
ATOM	633	CG2 THR A	80	5.989	20.159	65.821	1.00	58.58 A
ATOM	634	C THR A	80	8.587	21.458	65.610	1.00	58.07 A
ATOM	635	O THR A	80	9.690	21.007	65.293	1.00	57.95 A
ATOM	636	N SER A	81	8.316	21.889	66.841	1.00	60.75 A
ATOM	637	CA SER A	81	9.247	21.688	67.949	1.00	62.77 A
ATOM	638	C SER A	81	9.559	20.213	68.187	1.00	63.63 A
ATOM	639	O SER A	81	8.658	19.374	68.262	1.00	65.12 A
ATOM	640	CB SER A	81	8.717	22.330	69.234	1.00	63.10 A
ATOM	641	OG SER A	81	8.871	23.739	69.211	1.00	50.00 A
ATOM	642	OXT SER A	81	10.722	19.856	68.349	1.00	63.85 A
ATOM	643	OH2 HOH S	1	-2.914	8.154	33.667	1.00	90.61 S
ATOM	644	OH2 HOH S	2	-4.387	19.111	23.885	1.00	76.69 S
ATOM	645	OH2 HOH S	3	-4.056	12.730	34.911	1.00	56.05 S
ATOM	646	OH2 HOH S	4	5.517	5.678	39.727	1.00	50.96 S
ATOM	647	OH2 HOH S	5	8.659	22.343	37.799	1.00	63.30 S
ATOM	648	OH2 HOH S	6	-9.337	6.242	23.325	1.00	68.03 S
ATOM	649	OH2 HOH S	7	8.514	24.262	35.630	1.00	71.25 S
ATOM	650	OH2 HOH S	8	2.068	2.597	33.720	1.00	63.48 S
ATOM	651	OH2 HOH S	9	10.301	18.127	64.791	1.00	77.40 S

	atom type	resi- due		X	Y	Z	B	
ATOM	652	OH2 HOH S	10	19.207	13.105	28.671	1.00	74.34 S
ATOM	653	OH2 HOH S	11	10.308	19.599	43.117	1.00	73.01 S
ATOM	654	OH2 HOH S	12	4.528	15.757	19.357	1.00	71.32 S
ATOM	655	OH2 HOH S	13	2.033	22.579	20.388	1.00	89.40 S
ATOM	656	OH2 HOH S	14	-2.272	12.999	17.174	1.00	81.98 S
ATOM	657	OH2 HOH S	15	2.375	19.350	23.183	1.00	84.49 S
ATOM	658	OH2 HOH S	16	2.250	19.132	50.313	1.00	98.46 S
ATOM	659	OH2 HOH S	17	9.032	18.824	46.946	1.00	74.33 S
ATOM	660	OH2 HOH S	18	-4.401	5.825	19.722	1.00	84.93 S
ATOM	661	OH2 HOH S	19	13.240	21.205	68.567	1.00	84.72 S
ATOM	662	OH2 HOH S	20	16.110	19.764	18.709	1.00	96.22 S
ATOM	663	OH2 HOH S	21	-2.611	24.956	26.103	1.00	89.53 S
ATOM	664	OH2 HOH S	22	-0.722	11.429	48.410	1.00	92.30 S
ATOM	665	OH2 HOH S	23	13.889	16.325	55.811	1.00	76.97 S
ATOM	666	OH2 HOH S	24	11.095	3.106	26.502	1.00	66.01 S
ATOM	667	OH2 HOH S	25	3.448	3.607	39.616	1.00	89.93 S
ATOM	668	OH2 HOH S	26	7.272	10.967	15.674	1.00	90.84 S
ATOM	669	OH2 HOH S	27	0.505	8.271	44.201	1.00	63.98 S
ATOM	670	OH2 HOH S	28	14.395	22.680	61.089	1.00	57.97 S
ATOM	671	OH2 HOH S	29	-2.187	29.657	25.873	1.00	72.40 S
ATOM	672	OH2 HOH S	30	7.086	1.616	32.377	1.00	81.89 S
ATOM	673	OH2 HOH S	31	13.814	19.890	34.383	1.00	86.24 S
ATOM	674	OH2 HOH S	32	-5.611	5.646	32.880	1.00	76.11 S
ATOM	675	OH2 HOH S	33	2.995	9.509	52.422	1.00	76.29 S
ATOM	676	OH2 HOH S	34	3.017	5.911	22.593	1.00	92.04 S
ATOM	677	OH2 HOH S	35	10.939	27.230	24.442	1.00	81.68 S
ATOM	678	OH2 HOH S	36	3.191	26.519	21.460	1.00	99.55 S
ATOM	679	OH2 HOH S	37	-1.524	3.517	32.315	1.00	76.66 S
ATOM	680	OH2 HOH S	38	5.617	17.448	63.962	1.00	88.16 S
ATOM	681	OH2 HOH S	39	1.030	4.765	43.012	1.00	84.79 S
ATOM	682	OH2 HOH S	40	5.927	2.544	27.239	1.00	76.11 S
ATOM	683	OH2 HOH S	41	-2.051	8.847	38.098	1.00	88.77 S
ATOM	684	OH2 HOH S	42	8.966	19.098	72.583	1.00	85.19 S
ATOM	685	OH2 HOH S	43	12.361	6.052	19.758	1.00	89.74 S
ATOM	686	OH2 HOH S	44	-2.228	2.421	38.528	1.00	87.34 S
ATOM	687	OH2 HOH S	45	0.385	24.006	27.199	1.00	78.61 S
ATOM	688	OH2 HOH S	46	11.556	17.089	57.937	1.00	69.99 S
ATOM	689	OH2 HOH S	47	0.602	25.020	37.284	1.00	91.85 S
ATOM	690	OH2 HOH S	48	-3.151	6.487	38.364	1.00	87.28 S
ATOM	691	OH2 HOH S	49	2.334	16.716	18.256	1.00	75.37 S
ATOM	692	OH2 HOH S	50	1.628	14.232	54.161	1.00	72.05 S
ATOM	693	OH2 HOH S	51	3.903	31.001	34.266	1.00	83.60 S
ATOM	694	OH2 HOH S	52	11.253	12.666	16.859	1.00	84.77 S
ATOM	695	OH2 HOH S	53	0.354	17.688	19.388	1.00	95.71 S
ATOM	696	OH2 HOH S	54	-2.532	17.997	51.721	1.00	86.09 S
ATOM	697	OH2 HOH S	55	2.673	10.273	13.485	1.00	80.18 S
ATOM	698	OH2 HOH S	56	12.361	25.391	32.181	1.00	91.50 S
ATOM	699	ZN ZN Z	100	5.669	6.865	27.310	1.00	37.16 Z

INTERNATIONAL SEARCH REPORT

International Application No
PCT/EP2004/007934

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C07K14/435 G06F19/00 G01N33/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C07K G06F G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, BIOSIS, EMBASE, INSPEC, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	CHEN BIN ET AL: "Completion of meiosis in Drosophila oocytes requires transcriptional control by Grauzone, a new zinc finger protein" DEVELOPMENT (CAMBRIDGE), vol. 127, no. 6, March 2000 (2000-03), pages 1243-1251, XP002268985 ISSN: 0950-1991 cited in the application the whole document figures 2,3,5,7 ----- -/--	3,8, 16-21



Further documents are listed in the continuation of box C.



Patent family members are listed in annex.

° Special categories of cited documents:

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *G* document member of the same patent family

Date of the actual completion of the international search

20 January 2005

Date of mailing of the international search report

31/01/2005

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Authorized officer

Jenkins, G

INTERNATIONAL SEARCH REPORT

International Application No
PCT/EP2004/007934

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	KRISHNA S SRI ET AL: "Structural classification of zinc fingers." NUCLEIC ACIDS RESEARCH, vol. 31, no. 2, 15 January 2003 (2003-01-15), pages 532-550, XP002268986 ISSN: 0305-1048 (ISSN print) cited in the application the whole document figures 1-10	3,8, 16-21
A	GRISHIN NICK V: "Treble clef finger: A functionally diverse zinc-binding structural motif" NUCLEIC ACIDS RESEARCH, vol. 29, no. 8, 15 April 2001 (2001-04-15), pages 1703-1714, XP002268987 ISSN: 0305-1048 cited in the application the whole document figures 1-6	3,8, 16-21
A	FORSTER MARK J: "Molecular modelling in structural biology" MICRON, vol. 33, no. 4, 2002, pages 365-384, XP002268988 ISSN: 0968-4328 the whole document	3,8, 16-21
A	CHUNG HO-RYUN ET AL: "Genomic expansion and clustering of ZAD-containing C2H2 zinc-finger genes in Drosophila." EMBO REPORTS, vol. 3, no. 12, December 2002 (2002-12), pages 1158-1162, XP008027064 ISSN: 1469-221X (ISSN print) cited in the application the whole document	3,8, 16-21
T	JAUCH RALF ET AL: "The zinc finger-associated domain of the Drosophila transcription factor grauzone is a novel zinc-coordinating protein-protein interaction module." STRUCTURE (CAMBRIDGE, MASS.: 2001) UNITED STATES NOV 2003, vol. 11, no. 11, November 2003 (2003-11), pages 1393-1402, XP002268990 ISSN: 0969-2126 the whole document	3,8, 16-21

INTERNATIONAL SEARCH REPORT

International application No.
PCT/EP2004/007934

Box II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claims Nos.: 10
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
see FURTHER INFORMATION sheet PCT/ISA/210
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box II.2

Claims Nos.: 10

Present claim 10 relates to the use of compounds defined by reference to a desirable characteristic or property, namely that they bind the fold of the ZADgrau protein.

Claim 10 covers all compounds having this characteristic or property, whereas the application provides no support within the meaning of Article 6 PCT, nor disclosure within the meaning of Article 5 PCT, for any such compounds. In the present case, claim 10 so lacks support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Independent of the above reasoning, the claim also lacks clarity (Article 6 PCT). An attempt is made to define the compounds by reference to a result to be achieved. Again, this lack of clarity in the present case is such as to render a meaningful search over the whole of the claimed scope impossible. Consequently, this claim has not been searched.

The applicant's attention is drawn to the fact that claims relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure. If the application proceeds into the regional phase before the EPO, the applicant is reminded that a search may be carried out during examination before the EPO (see EPO Guideline C-VI, 8.5), should the problems which led to the Article 17(2) declaration be overcome.