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(54) **YEAST DISPLAY SYSTEMS**

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(57) **ABSTRACT**

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The present invention relates to the field of protein display libraries and library screening. In preferred embodiments, the present invention provides a three component system for display comprising a cell surface molecule, an adapter molecule and a display molecule.

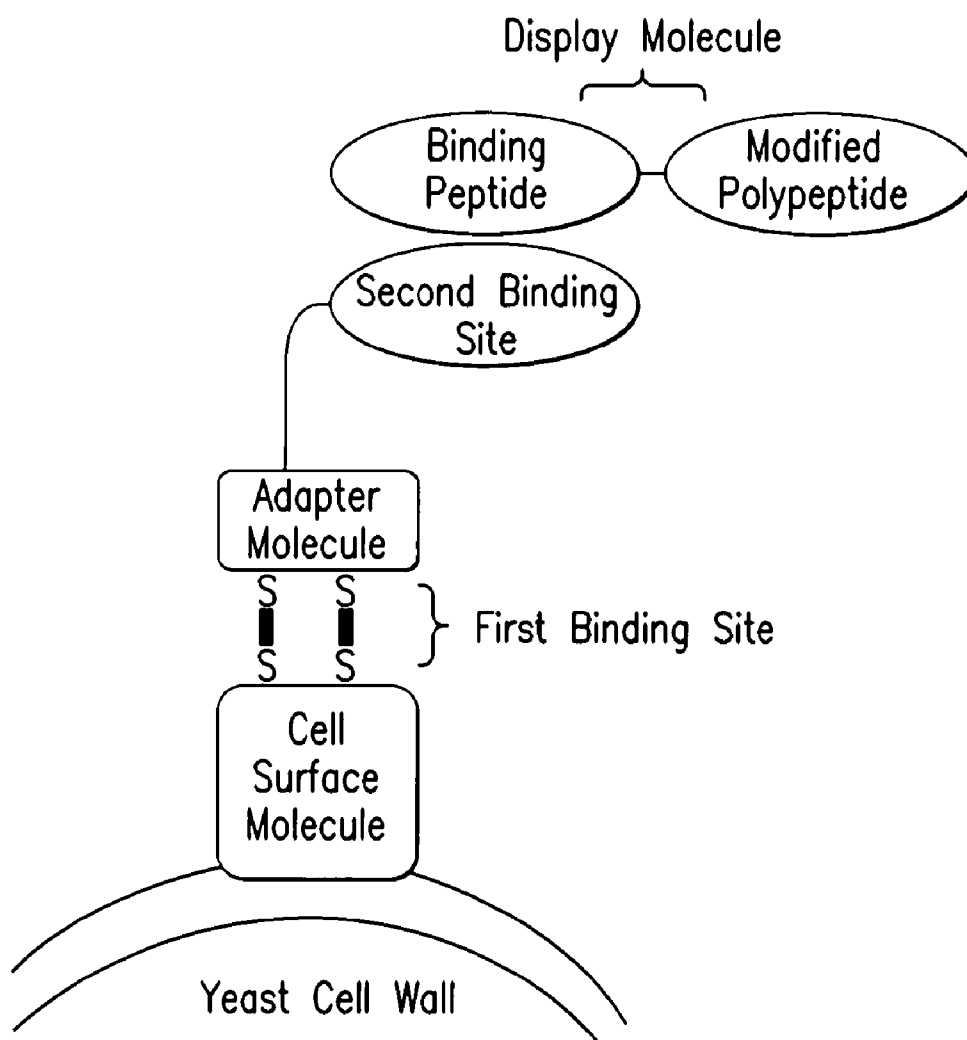


FIG. 1

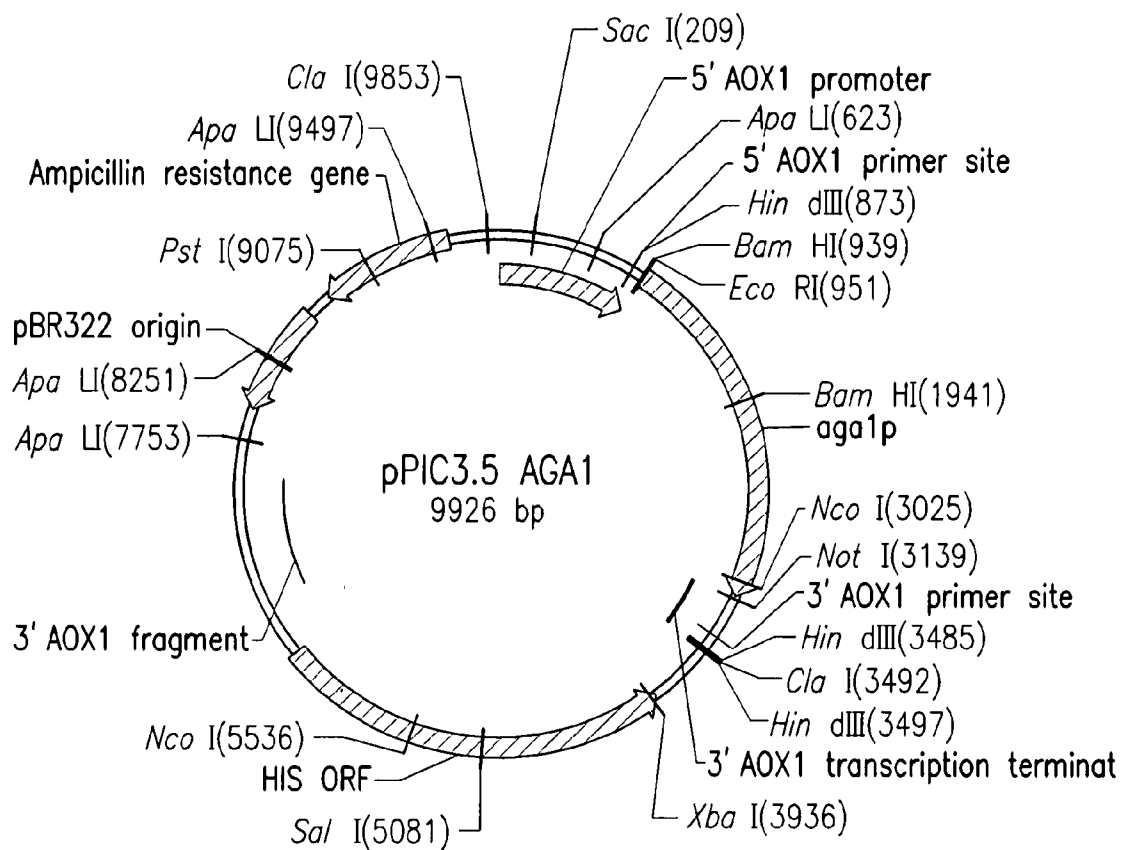


FIG.2

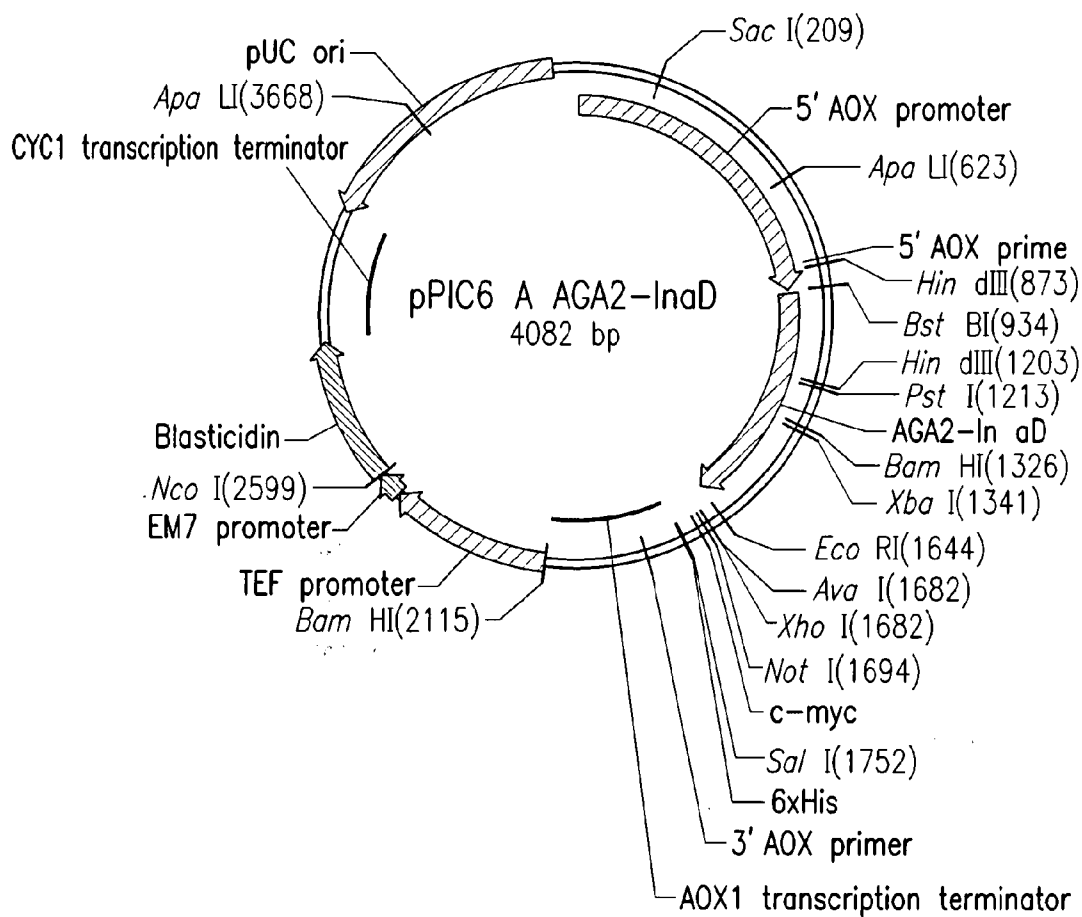


FIG.3

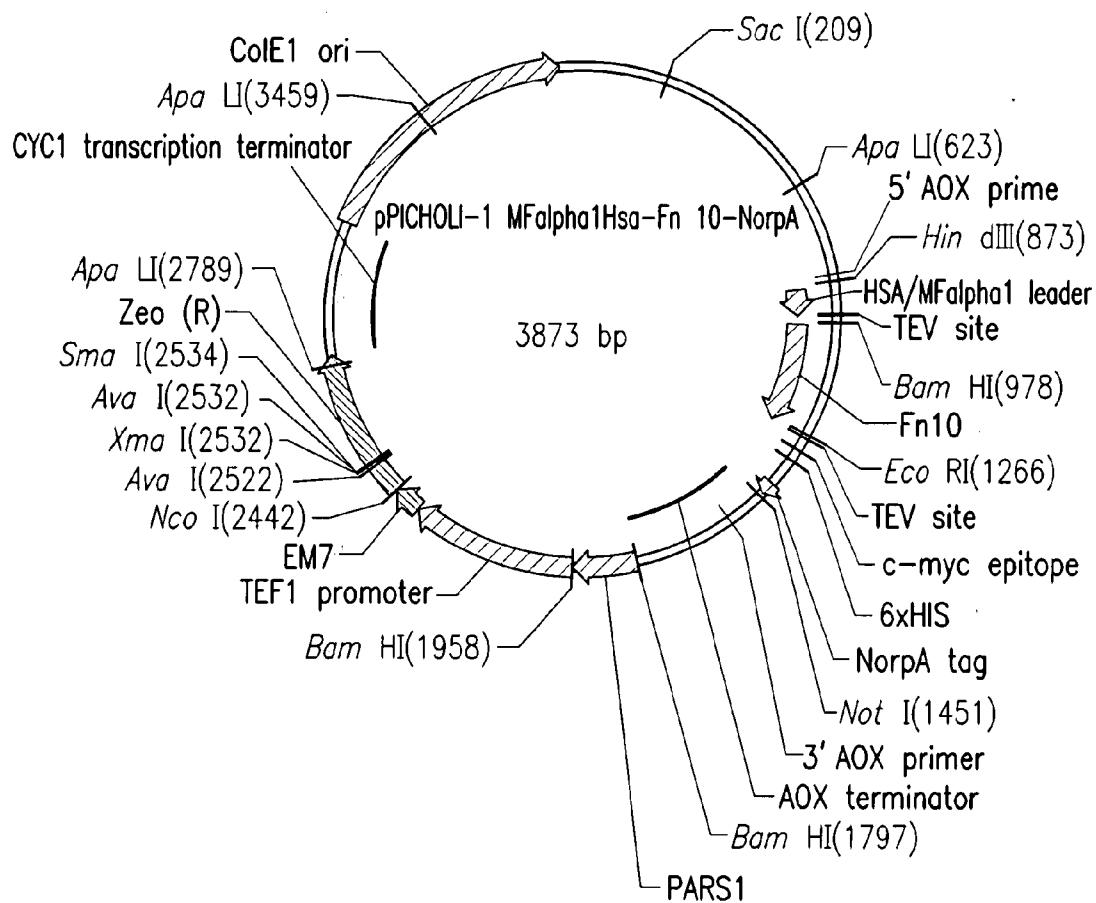


FIG.4

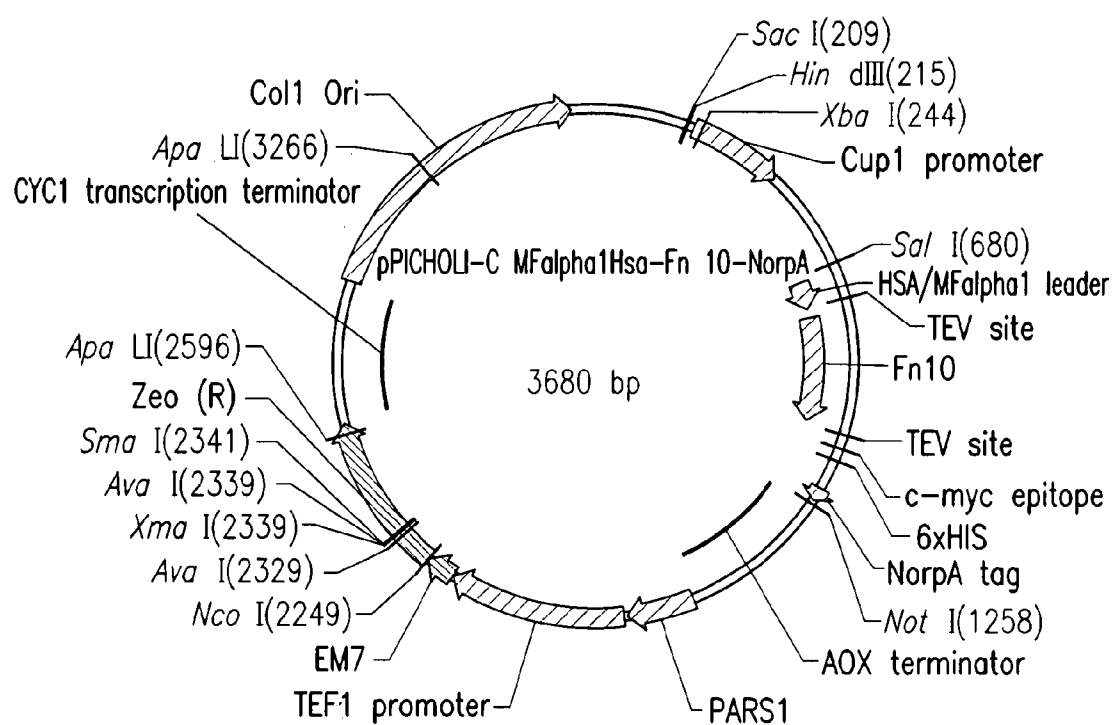


FIG.5

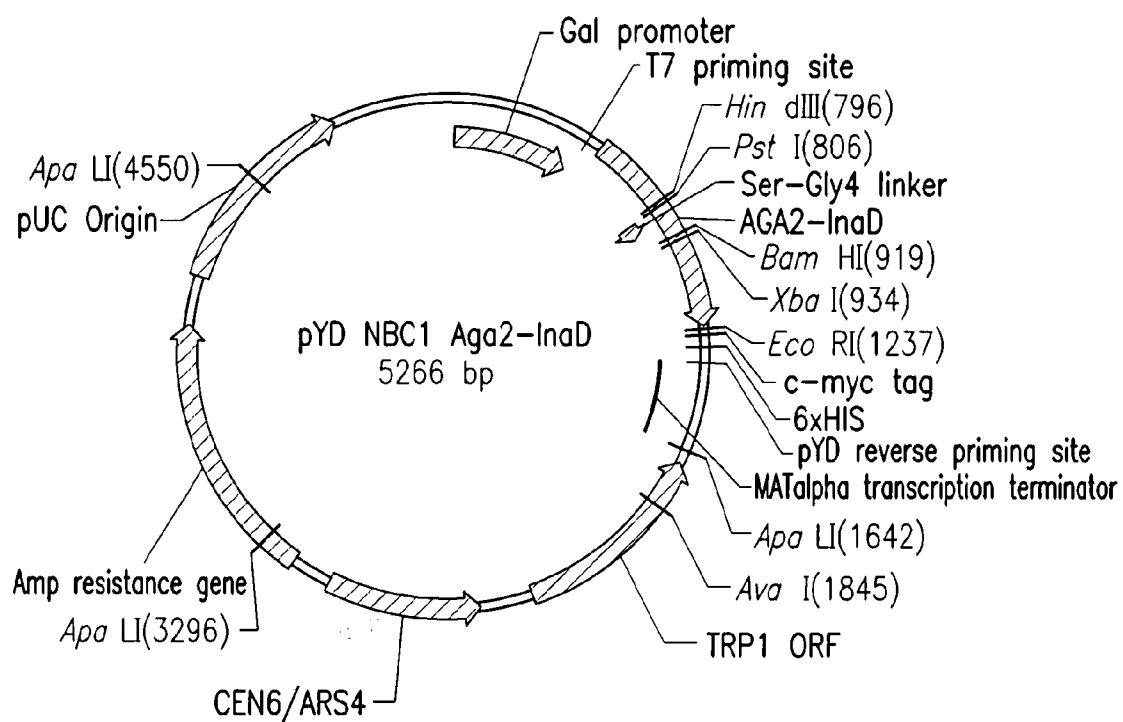


FIG.6

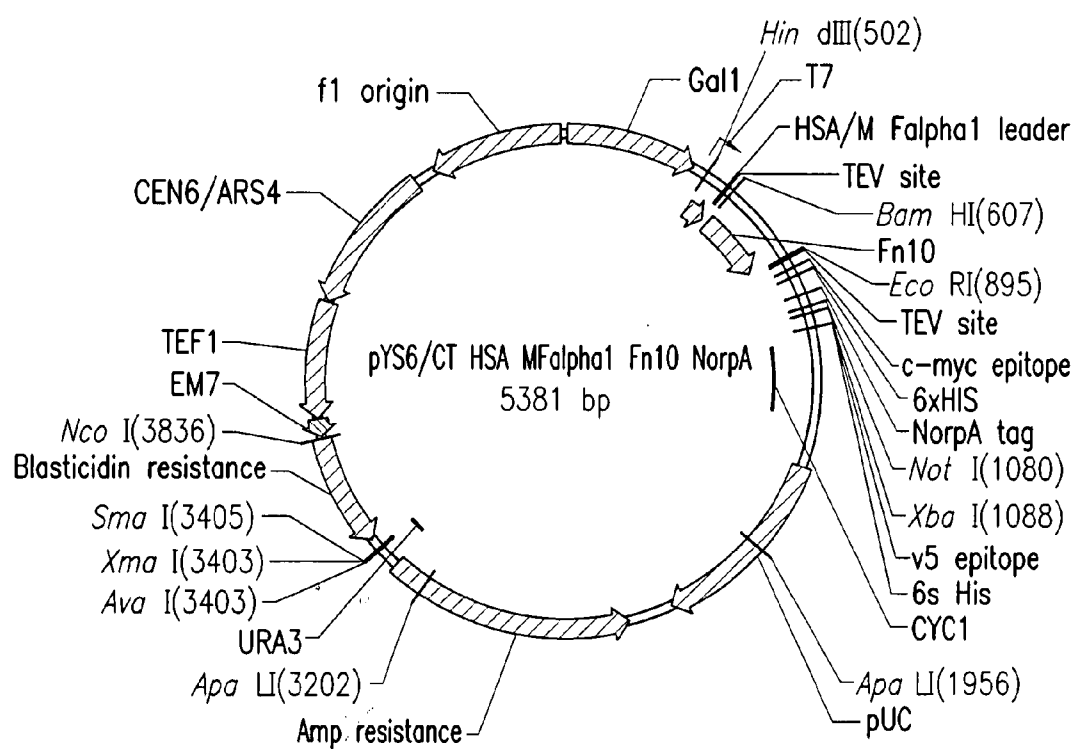


FIG.7



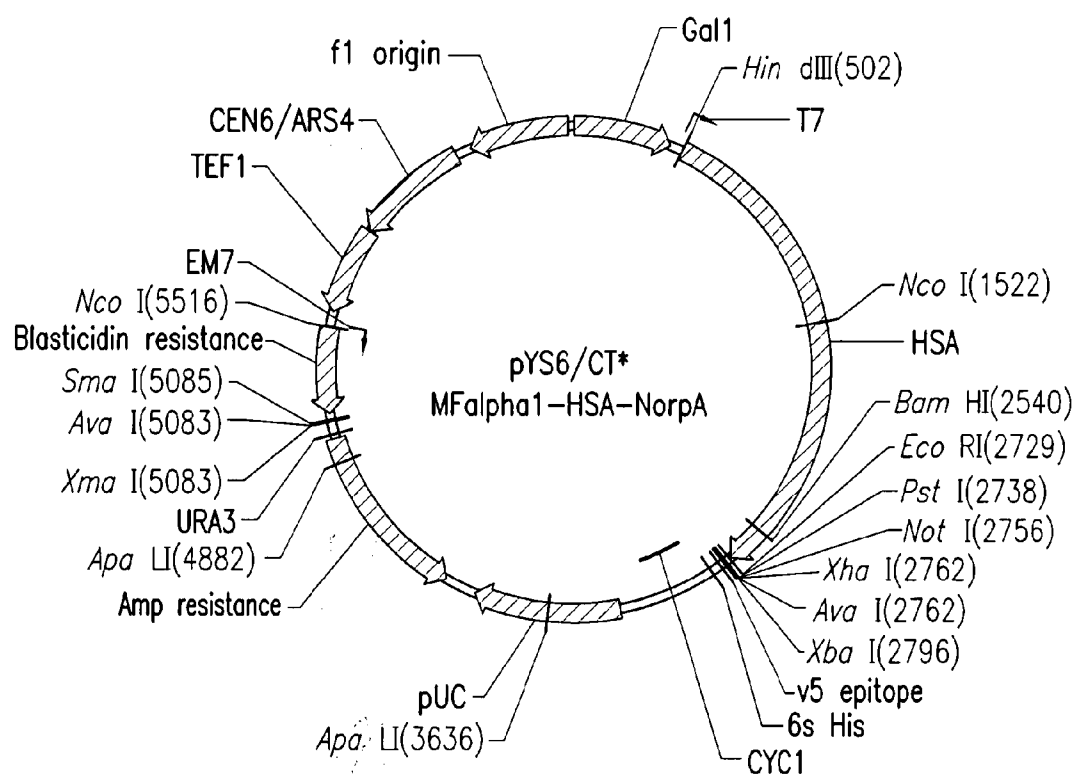


FIG.8

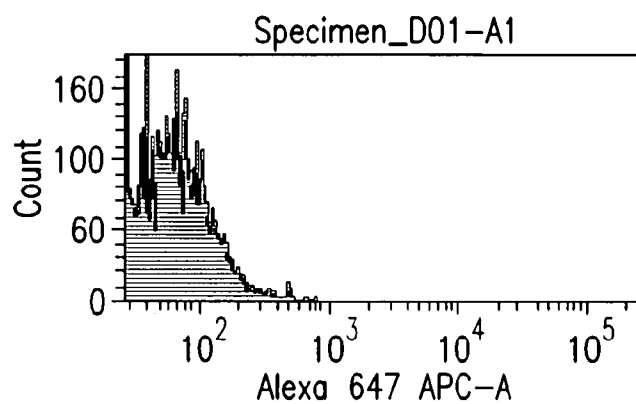


FIG.9A

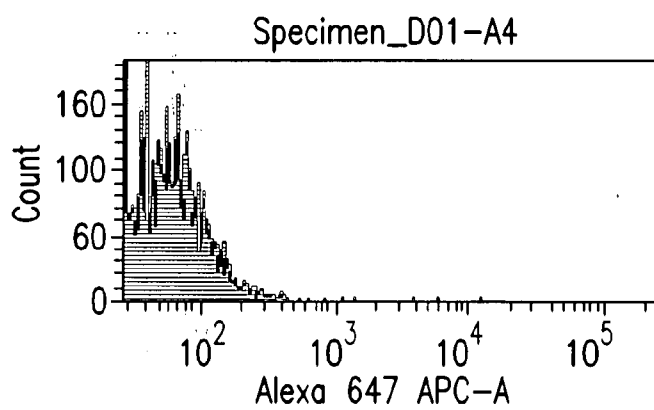


FIG.9B

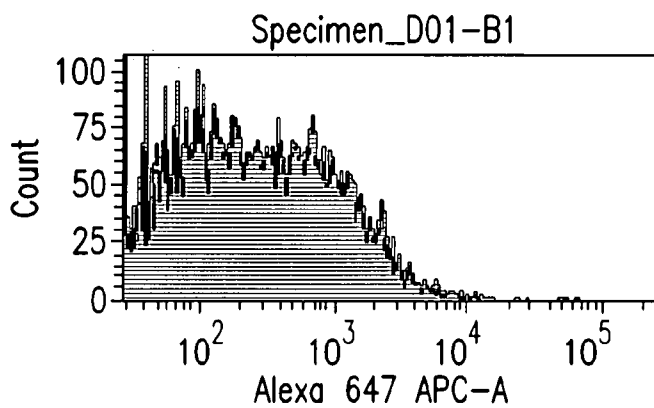


FIG.9C

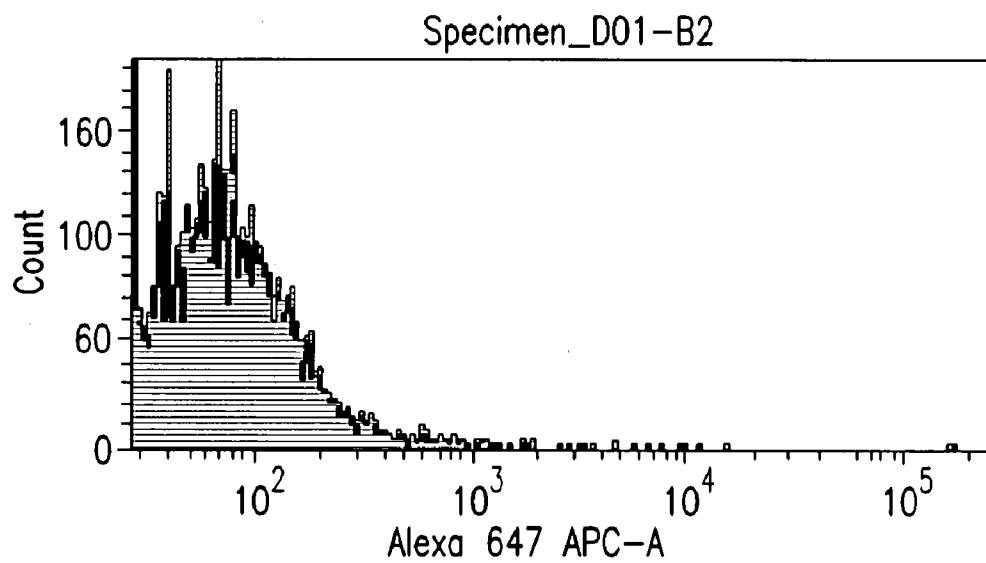


FIG.9D

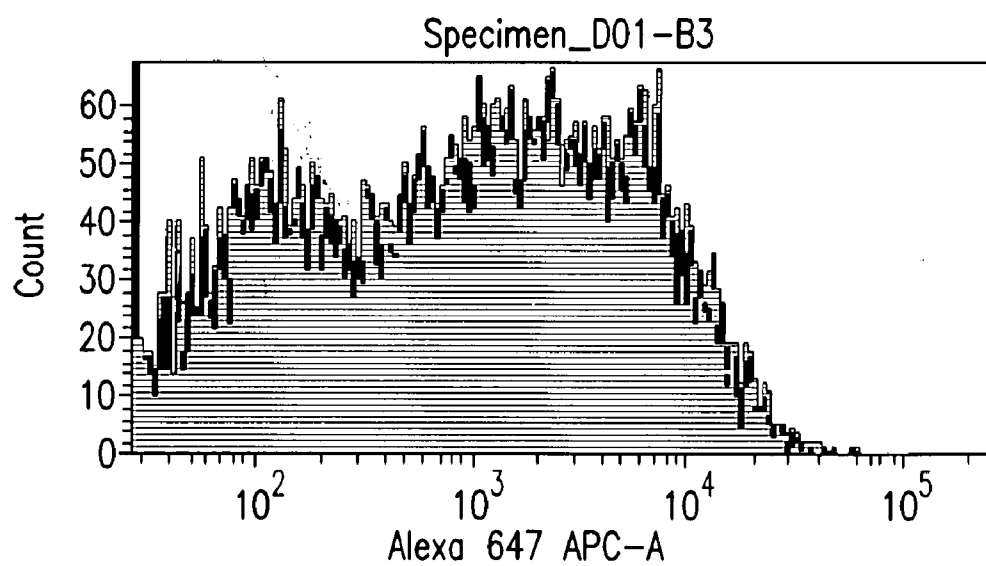


FIG.9E



FIG. 10A



FIG. 10B

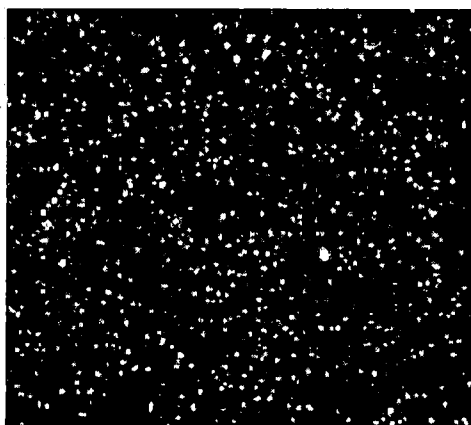


FIG. 10C

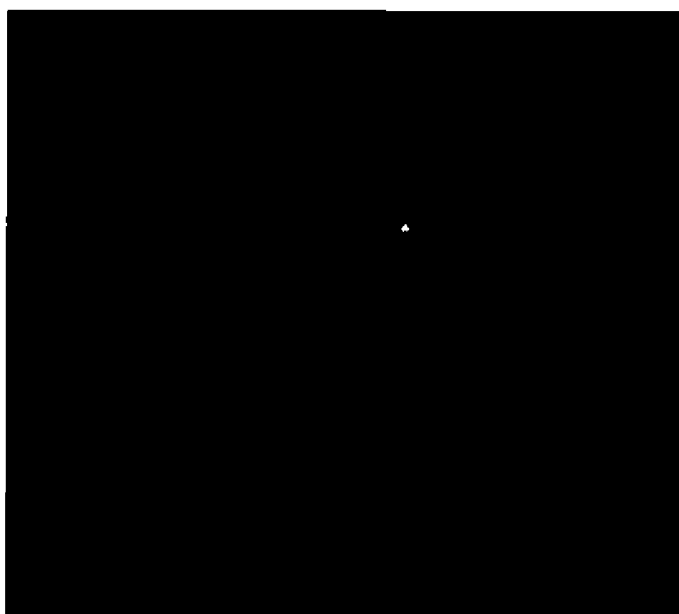


FIG. 10D

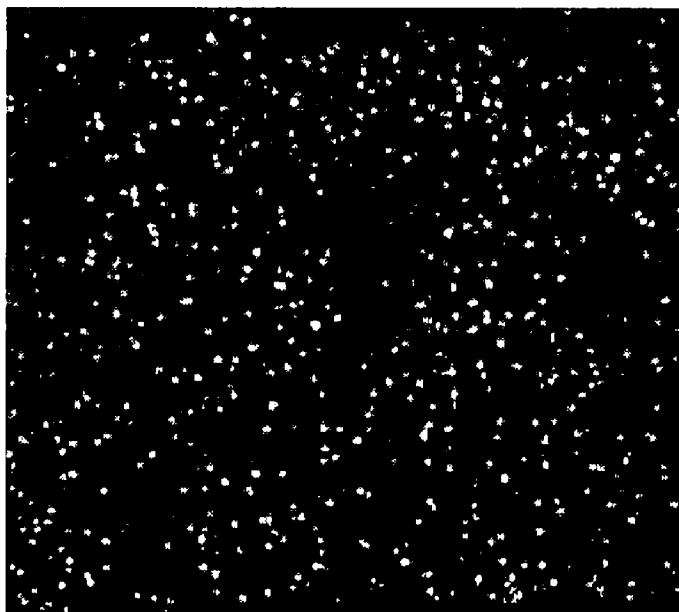


FIG. 10E

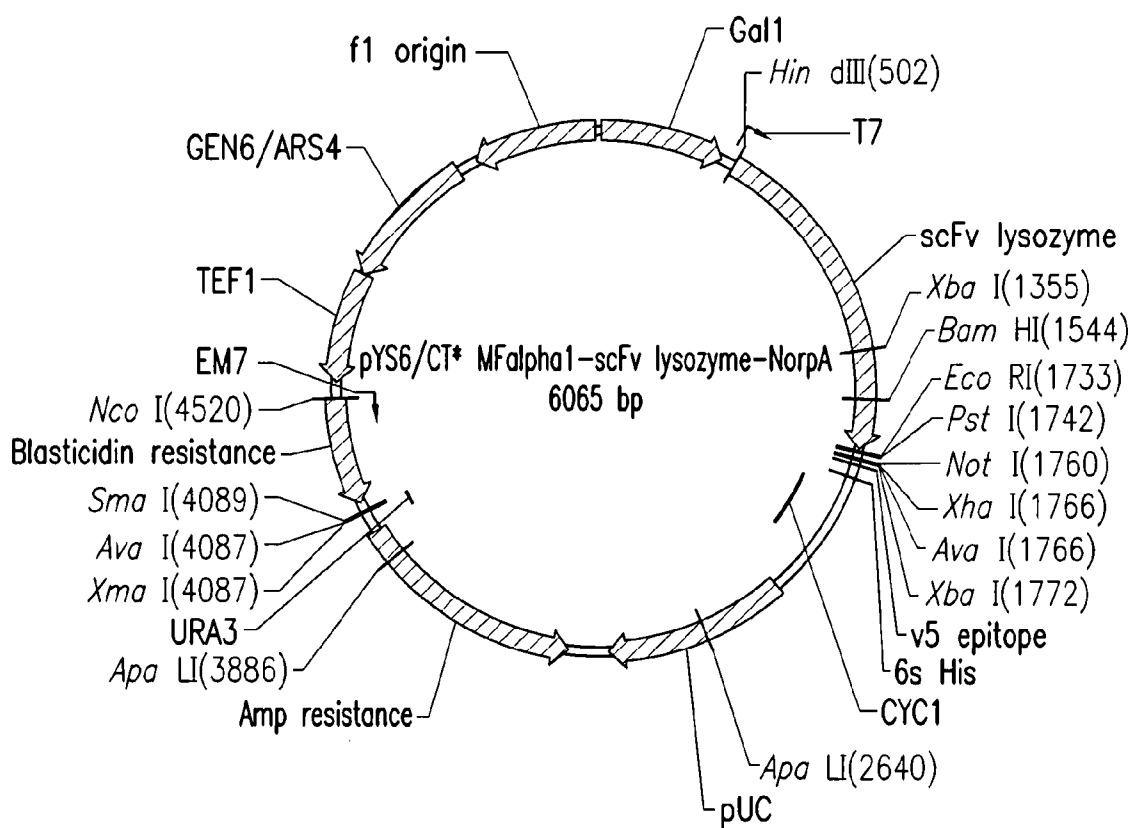


FIG. 11

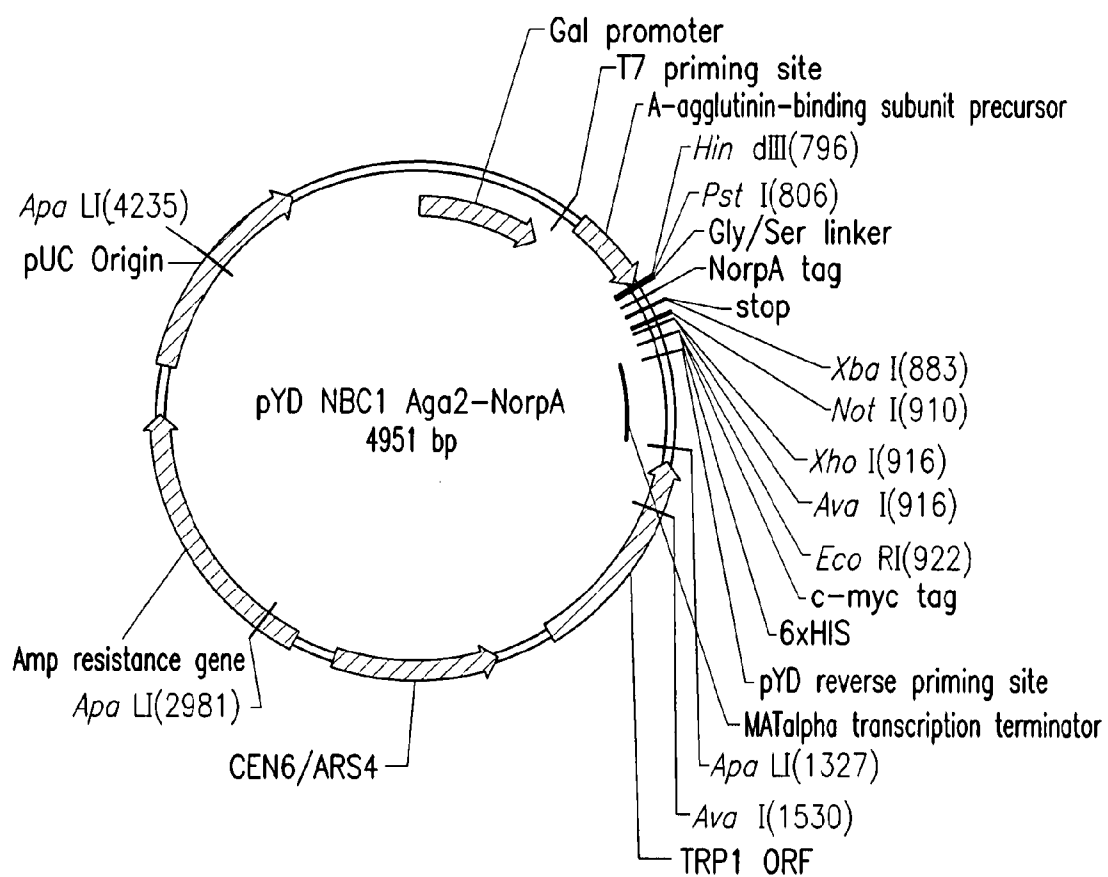


FIG.12

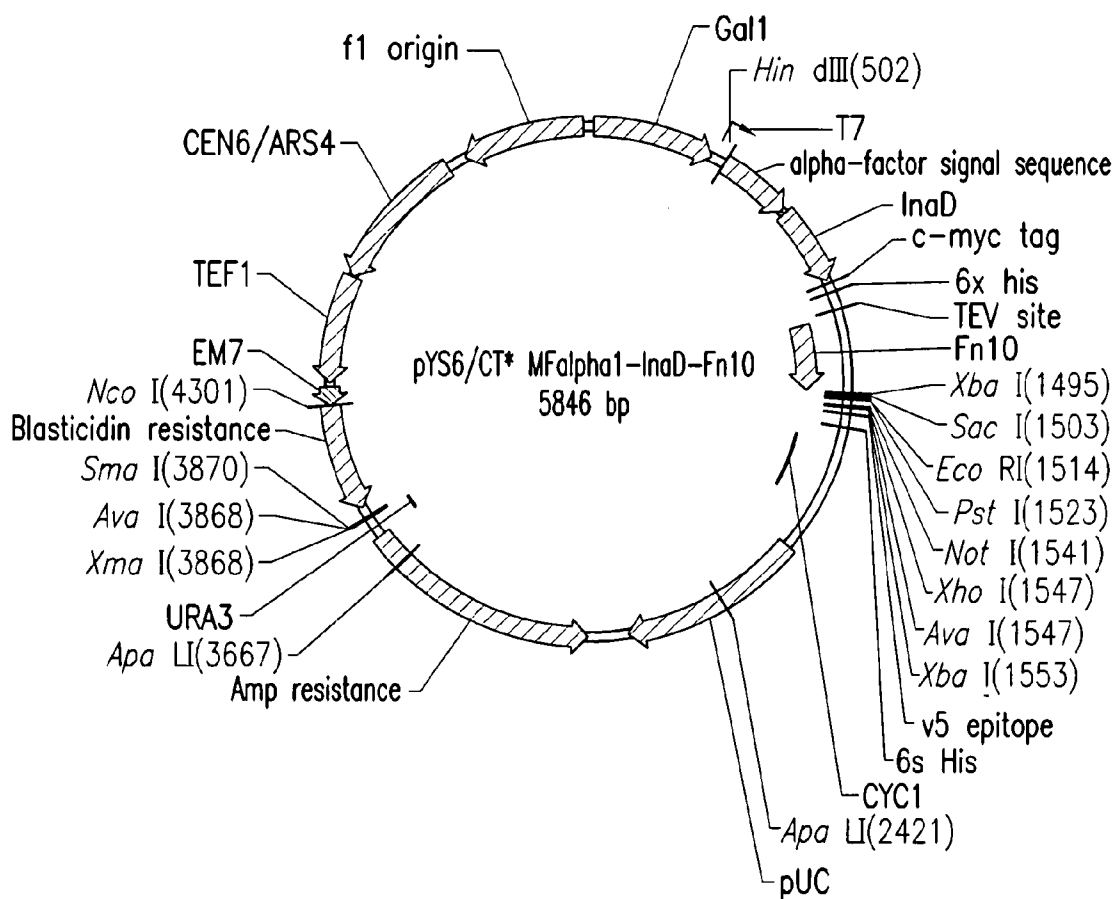


FIG.13



## YEAST DISPLAY SYSTEMS

### FIELD

**[0001]** The present invention relates to the field of protein display libraries and library screening. More specifically, the present invention relates to the production of proteins for display on cell surfaces.

### BACKGROUND

**[0002]** Protein binding domains can be predicted from sequence data, however re-designing proteins with improved or altered binding affinities often requires testing of a number of variants of the re-designed protein. Currently the best method for obtaining proteins with desired binding affinities is to generate and screen a protein library including such variants that can include rationally redesigned proteins, randomly altered proteins, or a combination thereof. Libraries of many types of protein, such as immunoglobulins and scaffold proteins and receptors or receptor ligands have successfully been constructed and screened for binding affinity.

**[0003]** There are many methods to screen libraries, but one of the most common methods is the phage display method, which comprises fusion of a protein library to the coat proteins of filamentous phage e.g., (Huse et al., '89; Clackson et al., '91); Marks et al., '92). Fusions are made most commonly to a minor coat protein, called the gene III protein (pIII), which is present in three to the copies at the tip of the phage. The fused library is then displayed on the surface of the viral particle. The phage display library can then be screened against an immobilized target protein. However, one major drawback of this method is that target proteins that bind the library with very high affinity are not always identified because the conditions required to elute the bound phase usually denature the phage particle such that it becomes impossible to identify the protein of interest. Another drawback of phage display libraries is the requirement that the target protein be immobilized on a solid surface, which can lead to difficulties in determining the actual affinity of a target protein for the phage display protein. Furthermore, some proteins of interest require post-translational modifications, such as glycosylation, methylation, or disulfide binding, that cannot be achieved when expressed in a phage particle.

**[0004]** An alternative method for screening protein libraries is to display the library on the surface of bacterial cells. This method solves many of the drawbacks associated with phage display but has its own problems. One problem with bacterial display is that the bacterial capsule can cause steric hindrance to proteins displayed on the bacterial surface. Also, bacteria do not contain the machinery to properly fold eukaryotic proteins, so the protein of interest may not always be expressed within the bacterium. Similar to the problem in phage, bacteria cannot provide post-translational modifications, like disulfide binding, to a eukaryotic protein.

**[0005]** Wittrup et al. (U.S. Pat. Nos. 6,699,558 and 5,696,251) have developed a method for a yeast cell display library. This is a two component system, wherein the first component involves expressing one subunit of the yeast mating adhesion protein, agglutinin, which is anchored to the yeast cell wall. The second component involves expressing a protein library fused to a second subunit of the agglutinin protein which forms high affinity disulfide bonds to the first agglutinin subunit. The protein library fused to the agglutinin is thus displayed on the surface of the cell. The library can then be

screened. This method allows for the proper folding and post-translational modification of eukaryotic proteins.

**[0006]** Rakestraw et al. (PCT/US20081003978) have developed a three component system for displaying a protein library on the surface of yeast cells. The first component involves expressing a protein library fused to a biotin-binding peptide, the second component involves modifying the yeast cell wall to express biotin, and the third component involves binding avidin to the biotin expressed on the cell surface. The fused protein library is then biotinylated and secreted from the yeast cell and binds to the avidin on the yeast cell surface, thus displaying the protein library on the surface of the yeast cell. One potential drawback of this system is that avidin non-specifically binds all biotin. Another potential drawback is that avidin contains four binding sites, which may cause steric hindrance thus preventing the biotinylated protein library from binding to the cell surface bound avidin. Similarly, the avidin molecule may bind the biotin fated protein library before binding the biotinylated yeast cell wall, thereby hindering the binding of the avidin to the yeast cell wall. Additionally, this method contains the added complication of having to biotinylate the protein library within the yeast cell. This necessary extra step requires further modification to the yeast cell. It is well established that the more modifications are made to a biological system, the less likely it is that the system will behave as designed. In addition, since avidin/streptavidin is multivalent, care must be taken to not cross-link the biotinylated cells. Finally, biotin/streptavidin and biotin/avidin are used in a number of commercially available labeling kits. Such kits would be difficult to use in such a biotin/avidin display system.

**[0007]** The prior art lacks a simple, efficient system capable of specifically binding a secreted protein library using an adapter molecule that binds to the protein library and to the surface of a eukaryotic cell through different binding moieties.

### SUMMARY OF THE INVENTION

**[0008]** The present invention meets this need by providing methods and compositions as disclosed through out the specification. One aspect includes host cells with a cell surface molecule attached to the surface of the cell, an adapter molecule comprising a first binding site and a second binding site and a display molecule comprising a modified polypeptide where the first binding site binds specifically to the cell surface molecule and cannot bind to the display molecule, the second binding site binds specifically to the display molecule and cannot bind to the cell surface molecule, and adapter molecule is not a component of the modified polypeptide. In certain embodiments, the host cell has a plurality of display molecules. In other embodiments which may be combined with the preceding embodiments, the host cell surface molecule may be covalently linked to the first binding site. In other embodiments which may be combined with the preceding embodiments, the host cell surface molecule may be covalently linked to the first binding site through a disulfide bond. In other embodiments which may be combined with any of the preceding embodiments, the host cell surface molecule may include a first agglutinin which may be Aga1p. In other embodiments which may be combined with any of the preceding embodiments, the host cell surface molecule may be attached to the cell membrane via a GPI anchor. In other embodiments which may be combined with any of the preceding embodiments, wherein the first binding site comprises a

second agglutinin which may be Aga2p. In other embodiments which may be combined with any of the preceding embodiments, the second binding site may be covalently linked to the display molecule. In other embodiments which may be combined with any of the preceding embodiments, the second binding site may be covalently linked to the display molecule through disulfide bonds. In other embodiments which may be combined with any of the preceding embodiments, the second binding site includes a PDZ domain which may be the PDZ domain of InaD which may have the amino acid sequence of SEQ ID NO: 8. In other embodiments which may be combined with any of the preceding embodiments, the display molecule includes a NorpA ligand which may be at the C-terminus which may have the amino acid sequence of SEQ ID NO: 9. In other embodiments which may be combined With any of the preceding embodiments except where the second binding site includes a PDZ domain or where the display molecule comprises a NorpA ligand, the display molecule includes a PDZ domain which may be the PDZ domain of InaD which may have the amino acid sequence of SEQ ID NO: 8. In other embodiments which may be combined with any of the preceding embodiments except where the second binding site includes a PDZ domain or where the display molecule comprises a NorpA ligand, the second binding site includes a NorpA ligand which may be at the C-terminus which may have the amino acid sequence or SEQ ID NO: 9. In other embodiments which may be combined with any of the preceding embodiments, wherein the modified polypeptide may be a scaffold protein, a signal transduction protein, an antibody, an immunoglobulin, an immunoadhesin, a receptor, a ligand an oncoprotein, a transcription factor, or an enzyme. In other embodiments which may be combined with any of the preceding embodiments, display molecule may be a fibronectin polypeptide which may include an F10 polypeptide. In other embodiments which may be combined with any of the preceding embodiments, the display molecule includes a secretion signal peptide may be an MFalpha, secretion signal sequence, a glucoamylase, an Aga2 secretion signal sequence, an Flo1p secretion signal sequence, an invertase secretion signal sequence, or an acid phosphatase secretion signal sequence. In other embodiments which may be combined with any of the preceding embodiments, the secretion signal peptide may be an MFalpha/HSA hybrid leader peptide. In other embodiments which may be combined with any of the preceding embodiments, expression of the display molecule is under the control of a first inducible promoter which may be an AOX 1 promoter, a Cup 1 promoter, or a Gal promoter. In other embodiments which may be combined with any of the preceding embodiments, the expression of the adapter molecule is under the control of a second inducible promoter which may be an AOX 1 promoter, a Cup 1 promoter, or a Gal promoter, in other embodiments which may be combined with any of the preceding embodiments, the host cell may be a yeast cell which may *Pichia pastoris* or *Saccharomyces cerevisiae*.

**[0009]** Another aspect includes libraries of host cells which include least two host cells in accordance with the preceding aspect and any and all of its embodiments where each host cell includes a different modified polypeptide.

**[0010]** Yet another aspect includes methods for displaying a modified polypeptide which includes (a) providing a host cell comprising a cell surface molecule attached to the surface of the cell and a first nucleic acid encoding a display polypeptide comprising a modified polypeptide, (b) contacting the

host cell with an adapter molecule comprising a first binding site and a second binding site under conditions wherein the first binding site binds to the cell surface molecule, and then (c) incubating, the host cell under conditions wherein the host cell exports the display polypeptide outside the host cell under conditions wherein the second binding site binds to the display polypeptide, where the first binding site binds specifically to the cell surface molecule and cannot bind to the display molecule, the second binding site binds specifically to the display polypeptide and cannot bind to the cell surface molecule, and the adapter molecule is not a component of the modified polypeptide. In other embodiments, the host cell may display at least  $10^2$ , at least  $10^3$ , at least  $10^4$ , or at least  $10^5$  modified polypeptides. In other embodiments which may be combined with the preceding embodiments, the host cell surface molecule may be covalently linked to the first binding site. In other embodiments which may be combined with the preceding embodiments, the host cell surface molecule may be covalently linked to the first binding site through a disulfide bond. In other embodiments which may be combined with any of the preceding embodiments, the host cell surface molecule may include a first agglutinin which may be Aga1p. In other embodiments Which may be combined with any of the preceding embodiments, the host cell surface molecule may be attached to the cell membrane via a GPI anchor. In other embodiments which may be combined with any of the preceding embodiments, wherein the first binding site comprises a second agglutinin which may be Aga2p. In other embodiments which may be combined with any of the preceding embodiments, the second binding site may be covalently linked to the display molecule. In other embodiments which may be combined with any of the preceding embodiments, the second binding site may be covalently linked to the display molecule through disulfide bonds. In other embodiments which may be combined with any of the preceding embodiments, the second binding site includes a PDZ domain which may be the PDZ domain of InaD which may have the amino acid sequence of SEQ ID NO: 8. in other embodiments which may be combined with any of the preceding embodiments, the display molecule includes a NorpA ligand which may heat the C-terminus which may have the amino acid sequence of SEQ ID NO: 9. In other embodiments which may be combined with any of the preceding embodiments except where the second binding site includes a PDZ domain or where the display molecule comprises a NorpA ligand, the display molecule includes a PDL domain which may be the PDZ domain of InaD which may have the amino acid sequence of SEQ ID NO: 8. In other embodiments which may be combined with any of the preceding embodiments except where the second binding site includes a PDZ domain or where the display molecule comprises a NorpA ligand, the second binding site includes a NorpA ligand which may be at the C-terminus which may have the amino acid sequence of SEQ ID NO: 9. In other embodiments which may be combined with any of the preceding embodiments, wherein the modified polypeptide may be a scaffold protein, a signal transduction protein, an antibody, an immunoglobulin, an immunoadhesin, a receptor, a ligand, an oncoprotein, a transcription factor, or an enzyme. In other embodiments which may be combined with any of the preceding embodiments, display molecule may be a fibronectin polypeptide which may include an F10 polypeptide. In other embodiments which may be combined with any of the preceding embodiments, the display molecule includes a secretion signal pep-

tide may be an MFalpha secretion signal sequence, a glucoamylase an Aga2 secretion signal sequence, an Flo1p secretion signal sequence, an invertase secretion signal sequence, or an acid phosphatase secretion signal sequence. In other embodiments which may be combined with any of the preceding embodiments, the secretion signal peptide may be an MFalpha/HSA hybrid leader peptide. In other embodiments which may be combined with any of the preceding embodiments, expression of the display molecule is under the control of a first inducible promoter which may be an AOX 1 promoter, a Cup 1 promoter, or a Gal promoter. In other embodiments which may be combined with any of the preceding embodiments, the expression of the adapter molecule is under the control of a second inducible promoter which may be an AOX 1 promoter, a Cup 1 promoter, or a Gal promoter. In other embodiments which may be combined with any of the preceding embodiments, the host cell may be a yeast cell which may be *Pichia pastoris* or *Saccharomyces cerevisiae*.

[0011] Still another aspect includes methods for generating a host cell display library which includes introducing into a plurality of host cells a display library of first nucleic acids each encoding a display polypeptide comprising a modified polypeptide, wherein at least two of the introduced first nucleic acids encode different modified polypeptides, wherein each host cell comprises a second nucleic acid which encodes a cell surface polypeptide and a third nucleic acid which encodes an adapter molecule comprising a first binding site and a second binding site where the first binding site binds to the cell surface molecule but not the display polypeptide, the second binding site binds to the display polypeptide but not the cell surface molecule, and the adapter molecule is not as component of the modified polypeptide. This aspect may be combined with any of the embodiments of the preceding aspects.

[0012] The foregoing are non-limiting examples of the present invention. Additional aspects and embodiments may be found throughout the specification.

#### BRIEF DESCRIPTION OF THE DRAWINGS

[0013] FIG. 1 shows a schematic of the three component system including the cell surface molecule, the adapter molecule including the first and second binding sites and the display molecule with the binding partner for the second binding site (a binding polypeptide in this embodiment) and a modified polypeptide as the molecule being displayed. In this embodiment, the host cell is a yeast cell.

[0014] FIG. 2 shows the pPIC3.5 AGA1 vector for yeast expression of the Aga1p as the cell surface molecule.

[0015] FIG. 3 shows the pPIC6 A AGA2-InaD vector for yeast expression of the Aga2p-InaD fusion polypeptide as the adapter molecule.

[0016] FIG. 4 shows the pPICHOL1-1 MFalpha1 Hsa-Fn10-NorpA vector for yeast expression of the MFalpha1Hsa-Fn10-NorpA fusion polypeptide as the display molecule where the NorpA is the binding partner of the second binding site of the adapter molecule (i.e., InaD) and the fibronectin F10 domain is modified polypeptide.

[0017] FIG. 5 shows the pPICHOL1-C MFalpha1 Hsa-Fn10-NorpA vector for yeast expression of the MFalpha1Hsa-Fn10-NorpA fusion polypeptide as the display molecule where the NorpA is the binding partner of the second binding site of the adapter molecule (i.e., InaD) and the fibronectin HO domain is modified polypeptide.

[0018] FIG. 6 shows the pYD NBC1 Aga2-InaD vector for yeast expression of the Aga2p-InaD fusion polypeptide as the adapter molecule.

[0019] FIG. 7 shows the pYS HSA MFalpha1 Fn10 NorpA vector for yeast expression of the MFalpha1HSA-Fn10-NorpA fusion polypeptide as the display molecule where the NorpA is the binding partner of the second binding site of the adapter molecule (i.e., InaD) and the fibronectin F10 domain is modified polypeptide.

[0020] FIG. 8 shows the pYS MFalpha1 HSA NorpA vector for yeast expression of the MFalpha1-HSA-NorpA fusion polypeptide as the display molecule where the NorpA is the binding partner of the second binding site of the adapter molecule (i.e., InaD) and the HSA is the displayed polypeptide.

[0021] FIG. 9A-E are FACS analysis graphs showing protein surface expression of yeast cells expressing either fibronectin (pYS6/CT HSA MFalpha1 Fn10 NorpA) or HSA (pYS6/CT MFalpha1-HSA-NorpA) in which the yeast cells are stained with anti-myc antibody and APC labeled secondary anti-mouse antibody. FIG. 9A is the a control with unstained yeast cells; FIG. 9B are uninduced yeast cells expressing fibronectin; FIG. 9C are induced yeast cells expressing fibronectin showing a shift in the curve; FIG. 9D are uninduced yeast cells expressing HSA; and FIG. 9E are induced yeast cells expressing HSA showing a shift in the curve.

[0022] FIG. 10A-E are images of FMAT analysis of yeast cells expressing either fibronectin (pYS6/CT HSA MFalpha1 Fn 10 NorpA) or HSA (pYS6/CT HSA-NorpA) in which the yeast cells are stained with anti-myc mouse monoclonal antibody and APC labeled secondary anti-mouse antibody and subjected to FMAT confocal fluorescence. FIG. 10A is the a control with unstained yeast cells; FIG. 10B are uninduced yeast cells expressing fibronectin; FIG. 10C are induced yeast cells expressing fibronectin appearing as white spots; FIG. 10D are uninduced yeast cells expressing HSA; and FIG. 10E are induced yeast cells expressing HSA appearing, as white spots.

[0023] FIG. 11 shows the pYS MFalpha1 scFv lysozyme NorpA vector for yeast expression of the MFalpha1 scFv lysozyme NorpA fusion polypeptide as the display molecule where the NorpA is the binding partner of the second binding site of the adapter molecule (i.e., InaD) and the lysozyme is modified polypeptide.

[0024] FIG. 12 shows a reversed system in which the pYD NBC1 Aga2-NorpA vector is used for yeast expression of the NBC1 Aga2-NorpA,

[0025] FIG. 13 shows a reversed system in which the pYS6/CT\*MFalpha1-InaD-Fn10 vector is used for yeast expression of the MFalpha1-InaD-Fn10.

#### DETAILED DESCRIPTION OF THE INVENTION

##### Definitions

[0026] As used herein the term "host cell" refers to a eukaryotic cell that has been modified to express a cell surface molecule, an adapter molecule, and a display molecule. Furthermore, it should be understood that the host cell secretes or excretes the display molecule prior to binding the display molecule to the adapter molecule on the surface of the host cell.

[0027] As used herein the term "cell surface molecule" refers to a peptide, polypeptide, binding domain, ligand, lipid,

or carbohydrate that is directed to the extracellular surface of the host cell. The cell surface molecule may be anchored to the cell surface by covalent binding or non-covalent binding. The cell surface molecule may include a phospholipid, carbohydrate or protein through which it attaches to the surface of the host cell. The cell surface molecule may be a polypeptide that binds to, or is conjugated to a phospholipid, carbohydrate, or a polypeptide on the surface of the cell. For example, the polypeptide may use a phosphatidyl-inositol-glycan (GPI) anchor to attach to the surface of the host cell, such as  $\alpha$ -agglutinins,  $\alpha$ -agglutinins, and flocculins. The cell surface molecule may also be a transmembrane protein with a binding domain located on the surface of the host cell that can bind to the first binding site of the adapter molecule.

**[0028]** As used herein the term “adapter molecule” refers to a peptide, polypeptide, binding domain, ligand, lipid, or carbohydrate or combination of the foregoing that has two distinct binding sites. The adapter molecule has a binding site that specifically binds the cell surface molecule and a second distinct binding site that specifically binds the display molecule. Without limiting the invention, the two binding sites of the polypeptide may be polypeptide domains each with its own binding affinity to a different molecule that are fused together. For example, the polypeptide may be an  $\alpha$ -agglutinin subunit, such as Aga2p, fused to a PDZ domain, or it may be a flocculin, such as Flo1, fused to a PDZ domain.

**[0029]** As used herein the term “first binding site” refers to a region of the adapter molecule that specifically recognizes and binds at least a portion of the cell surface molecule. For example, the first binding site may comprise a peptide, polypeptide, binding domain, ligand, lipid, or carbohydrate or combination thereof that specifically binds to a cell surface molecule which could include, without limitation, a peptide, binding domain, ligand, protein, lipoprotein, lipid, or carbohydrate. More specifically, but without limiting the invention, the first binding site may refer to the Aga2p subunit of  $\alpha$ -agglutinin that specifically binds to the Aga1p subunit of  $\alpha$ -agglutinin through disulfide bonds. In general, any molecular binding partners may be used for the first binding site and the corresponding portion of the cell surface molecule. Examples include  $\text{Ni}^{2+}$  ions and polyhistidine tags, sugar residues and Concanavalin A, p-aminophenyl- $\beta$ -D-thiogalactoside and  $\beta$ -galactosidase (Germino et al., Proc. Natl. Acad. Sci. USA 80:6848 (1983), glutathione and glutathione-S-transferase (Smith, D. B. and Johnson, K. S. Gene 67:311 (1988)); staphylococcal protein A and IgG (Uhlen, M. et al. Gene 23:369 (1983)), calmodulin nickel-binding proteins (CNBP) and calmodulin agarose Stofko-Hahn R. E. et al. FEBS Lett. 302(3):274-278); streptavidin or avidin and biotin (Takashi, S. and Dale, G. L., Proc. Natl. Acad. Sci. USA. 85:1647-1651 (1988)); amylase and maltose-binding protein domain from the malE gene of *E. coli* (Bach, H. et al, J. Mol. Biol. 312:79-93 (2001)), any epitope and its corresponding antibody (Sec, Kolodziej, P. A. and Young, R. A Methods Enzymol. 194:508-519 (1991), e.g., the FLAG<sup>TM</sup> octapeptide or antidigoxigenin antibody and digoxigenin).

**[0030]** As used herein the term “second binding site” refers to a region of the adapter molecule that specifically recognizes and binds the display molecule. For example, the second binding site may comprise a peptide, polypeptide, binding domain, ligand, lipid, or carbohydrate that specifically binds to a peptide, ligand, protein, lipoprotein, lipid, or carbohydrate comprising the display molecule. More specifically, but without limitations, the second binding site may

refer to a PMZ domain that specifically binds to a NorpA ligand. Any of the binding pairs suitable for the first binding site may also be used for the second binding site so long as they do not recognize the same partners.

**[0031]** As used herein the term “display molecule” refers to a molecule that can be localized to the surface of the host cell via binding of the adapter molecule on the surface of the host cell. The display molecule will typically comprise the molecule (or library of molecules) to be displayed and a binding partner that is specifically bound by the second binding site of the adapter molecule. In certain instances the molecule to be displayed and the binding partner may be one in the same. By way of example, the display molecule may comprise a peptide, polypeptide, binding domain, ligand, lipid, or carbohydrate or combination thereof. It should be understood that the display molecule is expressed or otherwise generated within the host cell and is secreted or excreted out of the cell so as to be displayed on the surface of said cell. The display molecule may comprise a library of varied molecules that can be screened for binding to a target or for improved or altered activity. In certain embodiments, the library may comprise modified polypeptides. The display molecule may also comprise a tag or peptide that can be labeled so as to detect binding of the display molecule to the cell surface, or sort host cells displaying said molecule.

**[0032]** As used herein the term “modified polypeptide” refers to any polypeptide of interest that is fused to a peptide, polypeptide, binding domain, ligand, lipid, or carbohydrate that specifically binds to a peptide, ligand, protein, lipoprotein, lipid, or carbohydrate comprising the second binding site of the adapter molecule, and is displayed on the surface of the host cell (and is therefore a component of the display molecule). Non-limiting examples of the modified polypeptide are scaffold proteins, signal transduction proteins, antibodies, immunoglobulins, immunoadhesins, receptors, ligands, oncoproteins, transcription factors, and enzymes,

**[0033]** As used herein, the term “plurality of display molecules” refers to at least two copies of the display molecule displayed on the surface of host cells. In certain instances, each unique display molecule is displayed by a different host cell.

**[0034]** As used herein the term “component of the modified polypeptide” refers to any naturally occurring binding partners of any fragment of the modified peptide. Non-limiting examples include an immunoglobulin light chain binding to an immunoglobulin heavy, a biotin molecule binding avidin, two subunits of  $\alpha$ -hemoglobin dimerizing, a myosin heavy chain binding to a myosin light chain, two monomers of glycophorin. A dimerizing, or two monomers of any naturally occurring dimer protein binding to one another.

**[0035]** As used herein the term “library of host cells” refers to a plurality of host cells, wherein each host cell comprises a non-identical modified polypeptide that is displayed on the surface of the cell.

**[0036]** As used herein the term “non-identical modified polypeptide” refers to the amino acid sequence of at least two modified polypeptides, wherein each amino acid sequence comprises amino acid substitutions, insertions, or deletions which differentiate one modified polypeptide displayed on the surface of a host cell from another modified polypeptide displayed on the surface of a second host cell.

**[0037]** As used herein the term “Fn10” refers to the tenth type III domain of human fibronectin.

**[0038]** Display Molecules

**[0039]** The display molecules may be used to display any molecule that may be expressed or otherwise generated in a host cell. Non limiting examples of such molecules follow.

**[0040]** Antibody Scaffold

**[0041]** The display molecules may be immunoglobulins. Methods of generating libraries of immunoglobulins were initially developed to display the immunoglobulins via phage, but additional methods of display have since been developed. All of the methods of generating libraries for these alternative methods of display may be adapted to allow display using the methods disclosed herein. Exemplary method of generating libraries of antibodies may be found in U.S. Pat. Nos. 5,223,409; 5,403,484; and 5,571,698 to Ladner et al.; U.S. Pat. No. 5,427,908 and 5,580,717 to Dower et al.; U.S. Pat. Nos. 5,969,108 and 6,172,197 to McCafferty et al.; and U.S. Pat. Nos. 5,885,793; 6,521,404; 6,544,731; 6,555,313; 6,582,915 and 6,593,081 to Griffiths et al.

**[0042]** Non-antibody Scaffold

**[0043]** Known non-immunoglobulin frameworks or scaffolds which may be displayed using the methods disclosed herein include, but are not limited to, fibronectins (Compound Therapeutics, Inc., Waltham, Mass.), ankyrin (Molecular Partners AG, Zurich, Switzerland), domain antibodies (Domantis, Ltd (Cambridge, Mass.) and Ablynx nv (Zwijnaarde, Belgium)), lipocalin (Anticalin) (Pieris Proteolab AG, Freising, Germany), small modular immunopharmaceuticals (Trubion Pharmaceuticals Inc., Seattle, Wash.), maxybodies (Avidia, Inc. (Mountain View, Calif.)), Protein A (Affibody AG, Sweden) and affilin (gamma-crystallin or ubiquitin) (Scil Proteins GmbH, Halle, (Germany), protein epitope munches (Polyphor Ltd, Allschwil, Switzerland).

**[0044]** (i):Fibronetins

**[0045]** The adnectin scaffolds are based on fibronectin type III domain (e.g., the tenth module of the fibronectin type III (10 Fn3 domain). The fibronectin type III domain has 7 or 8 beta strands which are distributed between two beta sheets, which themselves pack against each other to form the core of the protein, and further containing loops (analogous to CDRs) which connect the beta strands to each other and are solvent exposed. There are at least three such loops at each edge of the beta sheet sandwich, where the edge is the boundary of the protein perpendicular to the direction of the beta strands. (U.S. Pat. No. 6,673,901).

**[0046]** These fibronectin-based scaffolds are not an immunoglobulin, although the overall fold is closely related to that of the smallest functional antibody fragment, the variable region of the heavy chain, which comprises the entire antigen recognition unit in camel and llama IgG. Because of this structure, the non-immunoglobulin fibronectin molecule mimics antigen binding properties that are similar in nature and affinity to those of antibodies. These scaffolds can be used in a loop randomization and shuffling strategy in vitro that is similar to the process of affinity maturation of antibodies in vivo. These fibronectin-based molecules can be used as scaffolds where the loop regions of the molecule can be replaced with CDRs of the disclosure using standard cloning techniques.

**[0047]** (ii) Ankyrin—Molecular Partners

**[0048]** The technology is based on using proteins with ankyrin derived repeat modules as scaffolds for bearing vari-

able regions which can be used for binding to different targets. The ankyrin repeat module is a 33 amino acid polypeptide consisting of two anti-parallel  $\alpha$ -helices and a  $\beta$ -turn. Binding of the variable regions is mostly optimized by using ribosome display.

**[0049]** (iii) Maxybodies/Avimers—Avidia

**[0050]** Avimers are derived from natural A-domain containing protein such as LRP-1. These domains are used by nature for protein-protein interactions and in human over 250 proteins are structurally based on A-domains. Avimers consist of a number of different “A-domain” monomers (2-10) linked via amino acid linkers. Avimers can be created that can bind to the target antigen using the methodology described in, for example, 20040175756; 20050053973; 20050048512; and 20060008844.

**[0051]** (vi) Protein A—Affibody

**[0052]** Affibody® affinity ligands are small, simple proteins composed of a three-helix bundle based on the scaffold of one of the IgG-binding domains of Protein A. Protein A is a surface protein from the bacterium *Staphylococcus aureus*. This scaffold domain consists of 58 amino acids, 13 of which are randomized to generate Affibody® libraries with a large number of ligand variants (See e.g., U.S. Pat. No. 5,831,012). Affibody® molecules mimic antibodies, they have a molecular weight of 6 kDa, compared to the molecular weight of antibodies, which is 150 kDa. In spite of its small size, the binding site of Affibody molecules is similar to that of an antibody.

**[0053]** (v) Anticalins—Pieris

**[0054]** Anticalins® are products developed by the company Pieris Proteolab AG. They are derived from lipocalins, a widespread group of small and robust proteins that are usually involved in the physiological transport or storage of chemically sensitive or insoluble compounds. Several natural lipocalins occur in human tissues or body liquids.

**[0055]** The protein architecture is reminiscent of immunoglobulins, with hypervariable loops on top of a rigid framework. However, in contrast with antibodies or their recombinant fragments, lipocalins are composed of a single polypeptide chain with 160 to 180 amino acid residues, being just marginally bigger than a single immunoglobulin domain.

**[0056]** The set of four loops, which makes up the binding pocket, shows pronounced structural plasticity and tolerates a variety of side chains. The binding site can thus be reshaped in a proprietary process in order to recognize prescribed target molecules of different shape with high affinity and specificity.

**[0057]** One protein of lipocalin family, the bilin-binding protein (BBP) of Pieris *Brassicae* has been used to develop anticalins by mutagenizing the set of four loops. One example of a patent application describing “anticalins” is PCT WO199916873.

**[0058]** (vi) Affilin—Scil Proteins

**[0059]** Affilin™ molecules are small non-immunoglobulin proteins which are designed for specific affinities towards proteins and small molecules. New Affilin™ molecules can be very quickly selected from two libraries, each of which is based on a different human derived scaffold protein.

**[0060]** Affilin™ molecules do not show any structural homology to immunoglobulin proteins. Scil Proteins employs two Affilin™ scaffolds, one of which is gamma crystalline, a human structural eye lens protein and the other is “ubiquitin” superfamily proteins. Both human scaffolds are very small, show high temperature stability and are almost resistant to pH changes and denaturing agents. This high

stability is mainly due to the expanded beta sheet structure of the proteins. Examples of gamma crystalline derived proteins are described in WO200104144 and examples of "ubiquitin-like" proteins are described in WO2004106368.

**[0061]** (vii) Protein Epitope Mimetics (PEM)

**[0062]** PEM are medium-sized, cyclic, peptide-like molecules (MW 1-2kDa) mimicking beta-hairpin secondary structures of proteins, the major secondary structure involved in protein-protein interactions.

**[0063]** Non-scaffold

**[0064]** In addition to scaffolds which are useful for de novo generation of molecule with specific affinity, the methods disclosed herein may be used to display any other biological molecule that can be expressed or otherwise generated in the host cell. Libraries of such biological molecules, particularly polypeptides which can be encoded by polynucleotides for easy expression by the host cell, may be screened for improved characteristics of interest such as improved binding between receptor and ligand where the receptor or the ligand are part of the display molecule or improved enzymatic activity where the enzyme is part of the display molecule.

**[0065]** Expression Systems

**[0066]** Expression vectors may be used to express one or more of the cell surface molecules, the adapter molecule and the display molecule, in the host cell. Expression vectors for eukaryotic host cells typically include (i) eukaryotic DNA elements that control initiation of transcription, such as a promoter, (ii) eukaryotic DNA elements that control the processing of transcripts, such as a transcription termination/polyadenylation signal sequence, and (iii) optionally eukaryotic DNA elements that control replication in the eukaryotic host cell if the vector is to be independently replicated (e.g., non-integrating vectors). To ease construction of such expression vectors, the vectors may optionally include (iv) prokaryotic DNA elements coding for a bacterial replication origin and an antibiotic resistance marker to provide for the growth and selection of the expression vector when manipulating the vector in the bacterial host cell. Appropriate eukaryotic expression vectors for use with fungal, yeast, and mammalian cellular hosts are known in the art, and are described in, for example, Powell et al. (Cloning Vectors: A Laboratory Manual, Elsevier, N.Y., 1985).

**[0067]** Yeast host cells are of particular interest and include *Saccharomyces cerevisiae*, *Pichia pastoris*, and *Pichia methanolica*. These vectors include YIp-based vectors, such as YIp5, YRp vectors, such as YRp17, YE<sub>p</sub> vectors such as YE<sub>p</sub>13 and YC<sub>p</sub> vectors, such as YC<sub>p</sub>19. A number of vectors exist for the expression of recombinant proteins in yeast. Other example of the YE<sub>p</sub> vectors include YE<sub>p</sub>24, YE<sub>p</sub>51, and YE<sub>p</sub>52, which are cloning and expression vehicles useful in the introduction of genetic constructs into *S. cerevisiae* (see, e.g., Broach et al. (1983) in Experimental Manipulation of Cleric Expression, ed. M. Inouye Academic Press, p 83). These vectors are also shuttle vectors in that they can replicate in *E. coli* due the presence of the pBR322 ori, and in *S. cerevisiae* due to the replication determinant of the yeast 2 micron plasmid.

**[0068]** Suitable promoters for function in yeast include the promoters for metallothionein, 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem. 255, 2073 (1980) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Req. 7, 149 (1968); and Holland et al, Biochemistry 17, 4900 (1978)), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phospho-fruc-

tokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phospho-glucose isomerase, and glucokinase. Suitable vectors and promoters for use in yeast expression are further described in R. Hitzeman et al., EP073,657. Other suitable promoters for expression in yeast include the promoters from GAL1 (galactose), PGK (phosphoglycerate kinase), ADH (alcohol dehydrogenase), AOX1 (alcohol oxidase), HIS4 (histidinol dehydrogenase), and the like. Many yeast cloning vectors readily available and can be modified following the above discussion. Still other promoters, which have the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytocrome C, acid phosphatase, degradative enzymes associated with nitrogen metabolism, and the afore-mentioned metallothionein and glyceraldehyde-3-phosphate dehydrogenase, as well as enzymes responsible for maltose and galactose utilization. Finally, promoters that are active in only one of the two haploid mating types may be appropriate in certain circumstances. Among these haploid-specific promoters, the pheromone promoters MF $\alpha$ 1 and MF $\alpha$ 1 are of particular interest.

**[0069]** Secretion from yeast host cells of the components including the adapter molecule (if produced in the host cell) and the display molecule may be increased by use any available secretion signal sequences of yeast proteins. One example is the leader sequence of a precursor of yeast mating pheromone,  $\alpha$ -factor, which has also been used to direct secretion of heterologous proteins in yeast (See, e.g., Valenzuela, P., eds pp. 269-280, Butterworths, London; Brake, A. J. (1990) Meth. Enzymol. 185, 408-441). The  $\alpha$ -factor leader sequence, in addition to the N-terminal signal peptide of 17 residues, includes a hydrophilic pro-region which contains 72 residues and bears three sites of N-linked glycosylation. The pro-region is extensively glycosylated in the ER and Golgi and is cleaved by Kex2 endopeptidase in the late Golgi compartment. The presence of the pro-region at the N-terminus is believed to allow some heterologous proteins to pass the quality control in the ER and to reach the periplasm.

**[0070]** Another example is the leader sequence from yeast invertase (MLLQAFLFLLAGFAAKISADAHKS) (SEQ ID NO: 1). This leader sequence has been demonstrated to be cleaved from nascent heterologous peptide upon entrance into the endoplasmic reticulum. The enzyme responsible for cleavage of the pre sequence, Kex2, resides in the trans Golgi. A further example is the signal sequence of yeast acid phosphatase which may be used to direct the secretion of the components disclosed herein.

**[0071]** Methods for transforming *S. cerevisiae* cells with exogenous DNA and producing recombinant polypeptides therefrom are disclosed by, for example, Kawasaki, U.S. Pat. No. 4,599,311, Kawasaki et al., U.S. Pat. No. 4,931,373, Brake, U.S. Pat. No. 4,870,008, Welch et al, U.S. Pat. No. 5,037,743, and Murray et al U.S. Pat. No. 4,845,075. Transformed cells are selected by phenotype determined by the selectable, marker, commonly drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in *Saccharomyces cerevisiae* is the POT1 vector system disclosed by Kawasaki et al. (U.S. Pat. No. 4,931,373), which allows transformed cells to be selected by growth in glucose containing media.

**[0072]** Transformation systems for other yeasts, including *Hansenula polymorpha*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*, *Kluyveromyces fragilis*, *Ustilago may-*

*dis*, *Pichia pastoris*, *Pichia methanolica*, *Pichia guillermoidii* and *Candida maltosa* are known in the art. See, for example, Gleeson et al., J. Gen. Microbiol. 132:3459 (1986), and Cregg, U.S. Pat. No. 4,882,279. *Aspergillus* cells may be utilized according to the methods of McKnight et al., U.S. Pat. No. 4,935,349. Methods for transforming *Acremonium chrysogenum* are disclosed by Sumino et al., U.S. Pat. No. 5,162,228. Methods for transforming *Neurospora* are disclosed by Laambowitz, U.S. Pat. No. 4,486,533.

[0073] For example, the use of *Pichia methanolica* as host for the production of recombinant proteins is disclosed by Raymond, U.S. Pat. No. 5,716,808 Raymond, U.S. Pat. No. 5,736,383, Raymond et al., Yeast 14:11-23 (1998), and in international publication Nos. WO 97/17450, WO 97/17451, WO 98/02536, and WO 98/02565. DNA molecules for use in transforming *P. methanolica* will commonly be prepared as double-stranded, circular plasmids, which are preferably linearized prior to transformation. For polypeptide production in *P. methanolica* it is preferred that the promoter and terminator in the plasmid be that of a *P. methanolica* gene, such as a *P. methanolica* alcohol utilization gene (AUG1 or AUG2). Other useful promoters include those of the dihydroxyacetone synthase (DHAS), formate dehydrogenase (FMD), and catalase (CAT) genes. To facilitate integration of the DNA into the host chromosome, it is preferred to have the entire expression segment of the plasmid flanked at both ends by host DNA sequences. For large-scale, industrial processes where it is desirable to minimize the use of methanol, it is preferred to use host cells in which both methanol utilization genes (AUG1 and AUG2) are deleted. For production of secreted proteins host cells deficient in vacuolar protease genes (PEP4 and PRB1) are preferred. Electroporation is used to facilitate the introduction of a plasmid containing DNA encoding a polypeptide of interest into *P. methanolica* cells. *P. methanolica* cells can be transformed by electroporation using an exponentially decaying, pulsed electric field having a field strength of from 2.5 to 4.5 kV/cm, preferably about 3.75 kV/cm, and a time constant (t) of from 1 to 40 milliseconds, most preferably about 20 milliseconds.

[0074] For use of mammalian host cells, mammalian expression vectors are also well known in the art and may be used as well. Examples of suitable mammalian host cells include African green monkey kidney cells (Vero; ATCC CRL 1587), human embryonic kidney cells (293-HEK; ATCC CRL 1573), baby hamster kidney cells (BHK-21, BHK-570; ATCC CRL8544, ATCC CRL 10314), canine kidney cells (MDCK; ATCC CCL 34), Chinese hamster ovary cells (CHO-K1; ATCC (CCL61; CHO DG44 (Chasin et al., Som. Cell. Molec. Genet. 12:555, 1986)), rat pituitary cells (GH1; ATCC CCL82), HeLa S3 cells (ATCC, CCL2.2), rat hepatoma cells (H4-II-E, ATCC CRL 1548) SV40-transformed monkey kidney cells (COS-1 ATCC CRL 1650) and murine embryonic cells (NIH-3T3; ATCC CRL 1658).

## EXAMPLES

[0075] The following provides non-limiting examples of the systems, compositions and methods disclosed herein. Proteins can be displayed on the surface of yeast cells by utilizing the PDZ domain of the yeast InaD protein and the C-terminal 5 amino acids of the yeast NorpA protein. This three component protein display system consists of a vector expressing the protein to be displayed with a secretion signal fused at its N-terminus, and the NorpA ligand fused at the C-terminus; a second vector expressing an adapter protein

that can bind specifically to a yeast cell wall protein, and which is fused to the PDZ domain of InaD, which binds specifically to the NorpA ligand; and a third vector that express a yeast cell wall protein that binds specifically to the adapter protein. This system has been adapted for use in *Saccharomyces cerevisiae* and *Pichia pastoris*.

### Example 1

#### Yeast Display Using InaD/NorpA Interaction in *Pichia pastoris*

[0076] The protein display system for *P. pastoris* was developed to display a fibronectin type 111 domain (Fn10), by fusing a hybrid secretion sequence (MFalpha/HSA) or a yeast leader sequence (MFalpha1) at N-terminus of Fn10 and fusing the NorpA ligand to its C-terminus. Once expressed, the Fn10 was secreted from the cell and the NorpA ligand bound specifically to the PDZ domain of InaD through disulfide bonds. The InaD was fused to the C-terminus of the Aga2p protein. The Aga2p-InaD fusion protein served as the adapter protein, and the N-terminal Aga2p bound Aga1p, which was immobilized on the surface of the cell. Aga2p bound specifically to Aga1p through disulfide bonds.

[0077] The three component system consisting of the Fn10-NorpA fusion protein, the

[0078] Aga2p-InaD fusion protein, and the Aga1p cell surface protein were cloned into pPIC expression vectors, under the control of an inducible promoter. The inducible promoter used was the AOX1 promoter, which is induced by methanol. Thus when methanol was added to yeast cells transformed with the vectors, the three proteins were expressed. Aga1p as expressed on the surface of the cell, Aga2p-InaD was localized to the cell surface where the N-terminal region of Aga2p-InaD bound to Aga1p. Fn10-NorpA was localized to the secretory pathway, was secreted from the cell, and bound InaD via the C-terminal NorpA ligand (FIG. 1). The system can also be switched such that the InaD is fused to Aga1 and NorpA is fused to Aga2p (See FIGS. 12 and 13).

[0079] A c-myc epitope was fused between the NorpA ligand and the C-terminus of Fn10. The c-myc epitope allowed for the detection of the displayed fibronectin by using a c-myc antibody. The fluorescently labeled c-myc antibody bound to the c-myc epitope on the surface of the cell was detected by fluorescence-activated cell sorting (FA(S)).

[0080] Strains and Media

[0081] The *Escherichia coli* Top10 strain (Invitrogen Carlsbad, Calif.) was used as the host strain for recombinant DNA manipulation. The *P. pastoris* GS115 strain (Invitrogen Co., Carlsbad, Calif.) was used for the production of the fusion protein AGA2-InaD and HSA/MFalpha1-Fn10-NorpA. *E. coli* as cultivated in LB medium (1% tryptone, 0.5% yeast extract, and 0.5% sodium chloride) containing 100 ug/mL ampicillin. *P. pastoris* was cultivated in BMGY medium (1% yeast extract, 2% peptone, 100 mM potassium phosphate buffer (pH 6.0), 1.34% yeast nitrogen base,  $4 \times 10^{-5}$ % biotin, and 1% glycerol), and BMMY medium (1% yeast extract, 2% peptone, 100 mM potassium phosphate buffer (pH 6.0), 1.34% yeast nitrogen base,  $4 \times 10^{-5}$ % biotin, and 0.5-2.0% methanol).

[0082] Construction of Expression Plasmids

[0083] The gene corresponding to AGA1 as synthesized by Geneart and subcloned into pPIC3.5 (Invitrogen). The resulting vector was named pPIC3.5-AGA1 (FIG. 2). The AGA2-InaD anchor gene was synthesized by Geneart (Germany) and



subcloned into expression vector pPIC6a (Invitrogen) using BstI and EcoRI restriction sites. The resulting vector was named pPIC6-AGA2-InaD (FIG. 4). The fibronectin construct consists of the MFalpha1/HSA hybrid leader followed by the fibronectin fused at the C-terminus to the NorpA ligand sequence. The complete gene was synthesized by Geneart (Germany) and subcloned into pPICHOLI-1 (Mobictec). The resulting vector was named pPICHOLI-1 MFalpha1Hsa-Fn10-NorpA (FIG. 6).

**[0084]** Yeast Transformation.

**[0085]** Electro-competent *P. pastoris* GS115 (Invitrogen) strain was prepared according to the protocol specified by the supplier and co-transformed with SalI-digested pPIC3.5-AGA1, pPIC6-AGA2-InaD, and pPICHOLI-1 MFalpha1Hsa-Fn10-NorpA.

**[0086]** Cultivation Conditions

**[0087]** The yeast transformants were precultivated in BMGY medium containing 100 ug/ml Zeocin and 200 ug/ml Blasticidin at 30° C. for 16 hr, and used to inoculate 200 ml of BMGY medium (containing 100 ug/ml Zeocin and 200 ug/ml Blasticidin) in a 1 l baffled flask to give an initial OD<sub>600</sub> value of 0.1. After 24 hr. of cultivation, the culture was centrifuged at 1000 g for 10 min, and resuspended in BMMY medium (+100 ug/ml Zeocin and 200 ug/ml Blasticidin) containing 0.5%, 1.0%, or 2.0% methanol. To maintain the induction of the fusion proteins, 100% methanol was added every 24 hr. to the culture to the final concentrations mentioned above. Analysis of displayed fibronectins on the surface of yeast is performed using FACS and anti-myc antibody.

#### Example 2

##### Switch System to Secrete or Display Fibronectins on the Surface of *Pichia pastoris*

**[0088]** One variant of the above display system enables the choice between secretion and display of proteins from *P. pastoris*. To achieve this, the fibronectin construct consisting of the MFalpha1/HSA hybrid leader followed by fibronectin fused at the C-terminus to the NorpA ligand sequence is cloned into pPICHOLI-C instead of pPICHOLI-1. The resulting vector is named pPICHOLI-C MFalpha1Hsa-Fn10-NorpA (FIG. 8). The key difference between the two vectors is the promoter, which in pPICHOLI-1 is the AOX1 promoter induced by methanol, and in the pPICHOLI-C is the Cup1 promoter induced by copper. To display the fibronectin on the surface of *P. pastoris*, AGA1 and AGA2-InaD are induced with methanol, while pPICHOLI-C is induced with copper. This allows for the capture of the secreted fibronectin on the surface of yeast mediated through the tight InaD/NorpA interaction. For secretion of the fibronectin without displaying the protein on the surface of yeast, induction with copper is sufficient. Without the induction of AGA1 and AGA2-InaD (driven by methanol) the binding partner for NorpA (AGA1/AGA2-InaD) is not present on the surface of yeast and therefore the fibronectin will be secreted.

#### Example 3

##### Yeast Display Using InaD/NorpA Interaction in *Saccharomyces cerevisiae*

**[0089]** This example describes using the InaD/NorpA system with other yeast strains such as *Saccharomyces cerevisiae*

**[0090]** Strains and Media

**[0091]** *Escherichia coli* Top10 (Invitrogen, Carlsbad, Calif.) was used as the host strain for recombinant DNA manipulation. The *S. cerevisiae* strain EBY100 (Invitrogen Co., Carlsbad, Calif.) was used for the production of the fusion proteins AGA2-InaD and MSalpha1/HSA-Fn10-NorpA or pYS6CT\*MFalpha1-HSA-NorpA. *E. coli* was cultivated in LB medium (1% tryptone, 0.5% yeast extract, and 0.5% sodium chloride) containing 100 ug/mL ampicillin or 100 ug/ml Blasticidin. EBY 100 was cultivated in CM medium-URA.

**[0092]** Construction of Expression Plasmids

**[0093]** The InaD anchor gene was synthesized by Geneart (Germany) and subcloned in frame with the AGA2 anchor protein into the expression vector pYD NBC1 (derivative of pYD1Invitrogen) using HindIII and EcoRI restriction sites. The resulting vector was named pYD\_NBC1 AGA2-InaD (FIG. 10). The fibronectin construct consists of the MFalpha1/HSA hybrid leader sequence followed by the fibronectin fused at its C-terminus to the NorpA ligand. The complete gene was synthesized by Geneart (Germany) and subcloned into pYS6CT (Invitrogen), in which the origin of replication had been replaced by the CEN6/ARS4 region. The resulting vector was named pYS6CT\_HSA\_MFalpha1\_Fn10\_NorpA (FIG. 12).

**[0094]** Plasmids were isolated from *E. coli* and the sequence confirmed. The purified plasmids were then co-transformed into EBY 100 and plated out on selective media consisting of CM-TRP, +200 ug/ml Blasticidin. Transformed colonies appeared within 2 days and were tested for display of Fibronectin by FACS analysis using an anti-myc antibody (cccc).

#### Example 4

##### Yeast Display Using Flo1-InaD/NorpA in *Pichia pastoris*

**[0095]** This example describes the use of an alternative expression system, Flo1, which is used with InaD/NorpA in *Pichia pastoris*.

**[0096]** Strains and Media

**[0097]** The *Escherichia coli* Top10 strain (Invitrogen Carlsbad, Calif.) is used as the host strain for recombinant DNA manipulation. The *P. pastoris* GS115 strain (Invitrogen Co., Carlsbad, Calif.) is used for the production of the fusion protein Flo1-InaD and HSA/MFalpha1-Fn10-NorpA. *E. coli* was cultivated in LB medium (1% tryptone, 0.5% yeast extract, and 0.5% sodium chloride) containing 100 ug/mL ampicillin. *P. pastoris* was cultivated in BMGY medium (1% yeast extract, 2% peptone, 100 mM potassium phosphate buffer (pH 6.0), 1.34% yeast nitrogen base, 4x10<sup>-5</sup>% biotin, and 1% glycerol), and BMMY medium (1% yeast extract, 2% peptone, 100 mM potassium phosphate buffer (pH 6.0), 1.34% yeast nitrogen base, 4x10<sup>-5</sup>% biotin, and 0.5-2.0% methanol).

**[0098]** Construction of Expression Plasmids

**[0099]** The gene for Flo1, fused at the C-terminus to the PDZ domain of InaD, is synthesized by Geneart (Germany) and cloned into pPIC3.5 (Invitrogen) using a 5' EcoRI site and a 3'NotI site. The resulting plasmid is named pPIC3.5-Flo1-InaD. Expression of the fused protein is driven by the methanol inducible promoter AOX1. The fibronectin construct consists of the MFalpha1/HSA hybrid leader followed by the fibronectin fused at the C-terminus to the NorpA ligand sequence. The complete gene is synthesized by Geneart (Ger-



man/) and subcloned into pPICHOLI-1 (Mobitec). The resulting vector is named pPICHOLI-1 MFalpha1Hsa-Fn10-NorpA. Expression of the fibronectin construct is driven by the methanol inducible promoter AOX1.

**[0100]** Yeast Transformation.

**[0101]** Electro-competent *P. pastoris* GS115 (Invitrogen) strain is prepared according to the protocol specified by the supplier and co-transformed with SalI-digested pPIC3.5-Flo1-InaD, and pPICHOLI-1 MFalpha1 Hsa-Fn10-NorpA,

**[0102]** Cultivation Conditions

**[0103]** The yeast transformants are precultivated in BMGY medium containing 100 ug/ml Zeocin and 200 ug/ml Blastidicin at 30° C. for 16 hr. and used to inoculate 200 ml of BMGY medium (containing 100 ug/ml Zeocin and 200 ug/ml Blastidicin) in a 1 l baffie flask to give an initial OD<sub>600</sub> value of 0.1. After 24 hr. of cultivation, the culture is centrifuged at 1000 g for 10 min. and resuspended in BMMY medium (+100 ug/ml Zeocin and 200 ug/ml Blastidicin) containing 0.5%, 1 or 2.0% methanol. To maintain the induction of the fusion proteins, 100% methanol is added every 24 hr. to the culture to the final concentrations mentioned above. Analysis of displayed fibronectins on the surface of yeast is performed using FACS and anti-myc antibody.

#### Example 5

##### Screening of a Fibronectin Library

**[0104]** Fibronectin Library Display

**[0105]** A fibronectin library is generated by methods well known in the art, including the method disclosed in U.S. Pat. No. 6,673,901. Other methods, such as use of error prone PCR, use of random priming techniques, or use of computational techniques are well known in the art and can also be used. The fibronectin library is designed with appropriate restriction enzyme cleavage sites in order to clone the library into yeast expression vectors.

**[0106]** The fibronectin library is displayed on a plurality of *P. pastoris* cells as described above in Example 1. The fibronectin library is modified to contain an MFalpha/HSA hybrid leader sequence fused to the N-terminus and a NorpA ligand sequence fused to the C-terminus. The modified fibronectin library is then cloned into the pPICHOLI-1 vector. As in the examples above, the expression of the fibronectin library is under the control of the AOX1 promoter. *P. pastoris* cells are transformed with the pPICHOLI-1 vectors expressing the fibronectin library and the vectors expressing Aga1p and Aga2p-InaD. Expression of the components is induced by the addition of methanol to the cells, and the fibronectin library is displayed on a plurality of *P. pastoris* cells.

**[0107]** Screening of Display Library

**[0108]** The yeast display fibronectin library is screened for binding to a target protein of interest using one of many methods known in the art. For example, the target protein is contacted with the yeast display fibronectin library under conditions that allow for the specific binding of the target protein to any members of the library. All bound target protein is immobilized on the surface of a yeast cell. All unbound target protein is washed off. The bound target protein is fluorescently labeled by methods well known in the art, such as fluorescently labeled antibodies specific for the target protein. The labeled target protein, now immobilized on the surface of a yeast cell, is then detected using flow cytometry, i.e. FACS. Yeast cells that bind the labeled target protein will fluoresce and are sorted from those yeast cells that do not bind the target

protein. The sorted yeast cells that have bound the target protein are clonally expanded, and the clone line or line containing members of the fibronectin library that bind the target protein are determined.

**[0109]** Example 6

##### Screening of a Protein Library

**[0110]** Protein Library Display

**[0111]** A protein library is generated by methods well known in the art, such as use of error prone PCR, use of random priming techniques, or use of computational techniques. The protein library is designed with appropriate restriction enzyme cleavage sites in order to clone the library into yeast expression vectors.

**[0112]** The cloned protein library is displayed on a plurality of *P. pastoris* cells as described above in Example 1. The protein library is modified to contain an MFalpha/HSA hybrid leader sequence fused to the N-terminus and a NorpA ligand sequence fused to the C-terminus. The modified protein library is then cloned into the pPICHOLI-1 vector. As in the examples above, the expression of the protein library is under the control of the AOX1 promoter. *P. pastoris* cells are transformed with the pPICHOLI-1 vectors expressing the protein library and the vectors expressing Aga1p and Aga2p-InaD. Expression of the components is induced by the addition of methanol to the cells, and the protein library is displayed on a plurality of *P. pastoris* cells.

**[0113]** Screening of Display Library

**[0114]** The yeast display protein library is screened for binding to a target protein of interest using one of many methods known in the art. For example, the target protein is contacted with the yeast display protein library under conditions that allow for the specific binding of the target protein to any members of the library. All bound target protein is now immobilized on the surface of a yeast cell. All unbound target protein is washed off. The bound target protein is fluorescently labeled by methods well known in the art, such as fluorescently labeled antibodies specific for the target protein. The labeled target protein, now immobilized on the surface of a yeast cell, is then detected using flow cytometry, i.e. FACS. Yeast cells that bind the labeled target protein will fluoresce and are sorted from those yeast cells that do not bind the target protein. The sorted yeast cells that have bound the target protein are clonally expanded, and the clone line or line containing members of the protein library that bind the target protein are determined.

#### Example 7

##### Screening of a Fibronectin or HSA Libraries

##### Fibronectin or HSA Library Display

**[0115]** A fibronectin or HSA library is generated by methods well known in the art, such as use of error prone PCR, use of random priming techniques, or use of computational techniques. The libraries are designed with appropriate restriction enzyme cleavage sites in order to clone the libraries into yeast expression vectors (See FIG. 8 and SEQ ID NO: 10).

**[0116]** The cloned fibronectin library or HSA library is displayed on a plurality of *P. pastoris* cells as described above in Example 1. The fibronectin library is modified to contain an MFalpha/HSA hybrid leader sequence fused to the N-terminus and a NorpA ligand sequence fused to the C-terminus (pYS HSA\_MFalpha1 Fn10 NorpA). The HSA library is

modified to contain a MFalpha CT (C-Terminal) It is a left-over of the original Invitrogen vector used for the constructions. If you look at the vector map (e.g. FIG. 13) you can see a c-terminal v5 and 6xhis sequence of the c-terminus of the insert. I've placed a stop in front of it and it is not translated in the final displayed protein leader sequence (pYS6/CT HSA-NorpA). The modified fibronectin or HSA library is then cloned into the pYS vector. The expression of the fibronectin or HSA library is under the control of the T7 promoter. *P. pastoris* cells are transformed with the pYS vectors expressing the fibronectin or HSA library and the vectors expressing Aga1p and Aga2p-InaD. Expression of the components is induced by the addition of methanol to the cells, and, the fibronectin or HSA library is displayed on a plurality of *P. pastoris* cells.

[0117] (a) FACS Analysis of Protein Surface Expression.

[0118] Yeast cells expressing either fibronectin (pYS HSA\_MFalpha1 Fn10 NorpA) or HSA (pYS6/CT HSA-NorpA) were stained with anti-myc antibody, followed by APC labeled secondary anti-mouse antibody and then subjected to FACS analysis. The results of the analysis are shown in FIGS. 9A-E. Specifically, FIG. 9A is the control sample showing unstained yeast cells; FIG. 9B is a sample of uninduced yeast cells expressing fibronectin; FIG. 9C is a sample of induced yeast cells expressing fibronectin, showing a shift of cells compared with the uninduced cells; FIG. 9D is a sample of uninduced yeast cells expressing HSA; and FIG. 9E is a sample of induced yeast cells expressing HSA, again showing a shift of cells compared with the uninduced cells. These results clearly demonstrate that the yeast display system is able to express fibronectin molecules, and proteins, such as HSA.

[0119] b) Fluorometric Microvolume Assay Technology (FMAT, Perkin Elmer) Analysis of Yeast Expressing Fibronectin or HSA

[0120] Yeast cells expressing either fibronectin (plasmid) or HSA (plasmid) were also analyzed by FMAT by staining with anti-myc antibody and APC labeled secondary anti-mouse antibody. The samples were then subjected to FMAT confocal fluorescence microscopy and shown in FIGS. 10A-E. Those colonies that express fibronectin or HSA appear as white dots against a black background. Specifically, FIG. 10A is the control sample showing unstained yeast cells and appears entirely black; FIG. 10B is a sample of uninduced yeast cells expressing fibronectin. The uninduced yeast cells do not produce fibronectin and are not detected (image appears black); FIG. 10C is a sample of induced yeast cells expressing fibronectin. In this instance, induction leads to the fibronectin with a myc tag being expressed and detected using the anti-myc antibody. Subsequent detection with the APC secondary anti-mouse antibody and FMAT confocal fluorescence microscopy results in visible white colonies being detected. FIG. 10D is a sample of uninduced yeast cells expressing HSA, as before the uninduced yeast cells do not produce fibronectin and the image appears black; and FIG. 10E is a sample of induced yeast cells expressing HSA, again showing small white colonies compared with the uninduced cells. These results further confirm that the yeast display system is able to express fibronectin molecule and proteins, such as HSA.

#### Example 8

##### Screening of Single Chain Fv Libraries

##### Single Chain Fv Library Display

[0121] A single chain scFv library is generated by methods well known in the art, such as use of error prone PCR, use of

random priming techniques, or use of computational techniques. The libraries are designed with appropriate restriction enzyme cleavage sites in order to clone the libraries into yeast expression vectors (See FIG. 11 and SEQ ID NO 11).

[0122] The cloned scFv lysozyme library is displayed on a plurality of *P. pastoris* cells as described above in Example 1. The scFv library is modified to contain an MFalpha leader sequence fused to the N-terminus and a NorpA ligand sequence fused to the C-terminus (pYS6/CT\* MFalpha1-scFv lysozyme-NorpA). The modified scFv lysozyme library is then cloned into the pYS vector. The expression of the scFv lysozyme is under the control of the 17 promoter. *P. pastoris* cells are transformed with the pYS vectors expressing the scFv lysozyme library and the vectors expressing Aga1p and Aga2p-InaD. Expression of the components is induced by the addition of methanol to the cells, and the fibronectin or HSA library is displayed on a plurality of *P. pastoris* cells.

[0123] (a) FACS Analysis of Protein Surface Expression.

[0124] Yeast cells expressing the scFv lysozyme were stained with anti-myc antibody, followed by APC labeled secondary anti-mouse antibody and then subjected to FACS analysis.

[0125] (b) FMAT Analysis of Yeast Expressing Fibronectin or HSA

[0126] Yeast cells expressing the scFv lysozyme were also analyzed by FMAT by staining with anti-myc antibody and APC labeled secondary anti-mouse antibody. The samples were then subjected to FMAT confocal fluorescence microscopy.

#### Example 9

##### Screening of Protein Libraries with a Reverse System

##### Protein Library Display

[0127] In the Example, the yeast display system described herein is reversed such that NorpA is fused to Aga2 and the InaD is fused to Aga1. A protein library is generated by methods well known in the art, such as use of error prone PCR, use of random priming techniques, or use of computational techniques. The libraries are designed with appropriate restriction enzyme cleavage sites in order to clone the libraries into yeast expression vectors (See FIGS. 12 and 13 and SEQ ID NOs: 12 and 13).

[0128] The protein display system for *P. pastoris* was developed to display a fibronectin type III domain (Fn10), by fusing a leader sequence (MFalpha1) at N-terminus of InaD and fusing the Fn10 to its C-terminus. One expressed, the Fn10 was secreted from the cell and the PDZ domain of InaD bound to the NorpA ligand through disulfide bonds. The NorpA as fused to the C-terminus of the Aga2 protein. The Aga2p-NorpA fusion protein served as the adapter protein, and the N-terminal Aga2p bound Aga1p, which was immobilized on the surface of the cell. Aga2p bound specifically to Aga1p through disulfide bonds.

[0129] The three component system consisting of the Aga2p-NorpA fusion protein, the Aga1-InaD fusion protein, and the Aga1p cell surface protein were cloned into pPD and pYS expression vectors respectively, under the control of a Gal inducible promoter.

[0130] The inducible promoter used was the Gal1 promoter, which is induced by galactose. Thus when galactose was added to yeast cells transformed with the vectors, the three proteins were expressed. Aga1p as expressed on the surface of the cell. Aga2p-NorpA as localized to the cell surface where the N-terminal region of Aga2p-NorpA bound to Aga1p. Fn10-InaD was localized to the secretory pathway, Was secreted from the cell, and bound NorpA via the C-terminal InaD ligand.

pPIC3.5 AGA1 (956 bp-31336 bp, direct) 242aa (SEQ ID NO 2)  
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LITSSIKMST KNVATSVSTS TVESSYACST CAETSHSYSS VQTASSSSVT  
QQTSTKSWV SSMTTSDDEF NKHATGKYHV TSSGTSTIST SVSEATSTSS  
IDSESQEQSS HLLSTSVLSS SLSATLSSD STILLFSSVS SLSVEQSPVT  
TLQISSTSEI LQPTSSTAIA TISASTSSLs ATSISTPSTS VESTIESSSL  
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GGSGGGGSAS MTGGQQMGRE NLYFQGVPGS SVVSRAGELI HMVTLDKTGK  
KSPGICIVRG EVKDSPNTKT TGIFIKGIVP DSPAHL CGRL KVGDRILSLN  
GKDVRNSTEQ AVIDLIKEAD FKIELEIQTF DK

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YTITVYAVTG RGDSPASSKP ISINYRTEFE NLYFQGGGGG GEQKLISEED  
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pPICHOLI-1-C MfalphaHsa-Fn10-NorPA (691 bp-1248 bp, direct) 62aa (SEQ ID NO: 5)  
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LLISWDAPAV TVRYRITYG ETGGNSPVQE FTVPGSKSTA TISGLKPGVD  
YTITVYAVTG RGDSPASSKP ISINYRTEFE NLYFQGGGGG GEQKLISEED  
LHHHHHPST PPTPSPSTPP TPSPSYKTQG KTEFCA

pYD\_NBC1\_Aga2-InaD (534 bp-1235 bp, direct) 78aa (SEQ. ID NO: 6)  
MQLLRCSIF SVIASVLAQE LTTICEQIPS PTLESTPYSL STTTILANGK  
AMQGVFEYYK SVTFVSNCGS HPSTTSKGSP INTQYVFKLL QASGGGGSGG  
GGSGGGGSAS MTGGQQMGRE NLYFQGVPGS SVVSRAGELI HMVTLDKTGK  
KSPGICIVRG EVKDSPNTKT TGIFIKGIVP DSPAHL CGRL KVGDRILSLN  
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LHHHHHPST PPTPSPSTPP TPSPSYNTQG KTEFCA  
InaD PDZ domain amino acid sequence (InaD as 11-107)  
(SEQ ID NO: 8)  
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LCGRLLKVGDR ILSLNGKDV NRSTEQAVIDL KEADFKIEL EIQTDFK  
Norpa C-terminal 11 amino acids including EFCA motif  
(SEQ ID NO: 9)  
YKTQGKTEFC A  
pYS6/CT\* MFalpha HSA-Norpa  
(SEQ ID NO: 10)  
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pYS6/CT\* MFalpha-scFv lysozyme-Norpa  
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pYD\_NBC1\_Aga2-Norpa  
(SEQ ID NO: 12)  
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FEYKSVTFVSNCGSHPSTTSKGSPI NTQYVFKLLQASGGGGSGGGG  
SYKTQGKTEFCA  
pYS6/Cf\* MFalpha-InaD-Fn10 (507 bp-1487 bp, direct) 109aa  
(SEQ ID NO: 13)  
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Ala	Thr	Ser	Thr	Ser	Ser	Ile	Asp	Ser	Glu	Ser	Gln	Glu	Gln	Ser	Ser	545	550	555	560
His	Leu	Leu	Ser	Thr	Ser	Val	Leu	Ser	Ser	Ser	Ser	Leu	Ser	Ala	Thr	565	570	575	
Leu	Ser	Ser	Asp	Ser	Thr	Ile	Leu	Leu	Phe	Ser	Ser	Val	Ser	Ser	Leu	580	585	590	
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Val Glu Ser Thr Ile Glu Ser Ser Ser Leu Thr Pro Thr Val Ser Ser 645 650 655		
Ile Phe Leu Ser Ser Ser Ser Ala Pro Ser Ser Leu Gln Thr Ser Val 660 665 670		
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Leu Ile Ser Glu Glu Asp Leu His His His His His His Pro Ser Thr		
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Ser Val Ser Asp Val Pro Arg Asp Leu Glu Val Val Ala Ala Thr Pro		
	35	40 45
Thr Ser Leu Leu Ile Ser Trp Asp Ala Pro Ala Val Thr Val Arg Tyr		
	50	55 60
Tyr Arg Ile Thr Tyr Gly Glu Thr Gly Gly Asn Ser Pro Val Gln Glu		
	65	70 75 80



<400> SEQUENCE: 6

Met 1	Gln	Leu	Leu	Arg 5	Cys	Phe	Ser	Ile	Phe 10	Ser	Val	Ile	Ala	Ser 15	Val
Leu	Ala	Gln	Glu 20	Leu	Thr	Thr	Ile	Cys 25	Glu	Gln	Ile	Pro	Ser 30	Pro	Thr
Leu	Glu	Ser 35	Thr	Pro	Tyr	Ser	Leu 40	Ser	Thr	Thr	Thr	Ile 45	Leu	Ala	Asn
Gly 50	Lys	Ala	Met	Gln	Gly 55	Val	Phe	Glu	Tyr	Tyr	Lys 60	Ser	Val	Thr	Phe
Val 65	Ser	Asn	Cys	Gly	Ser 70	His	Pro	Ser	Thr	Thr 75	Ser	Lys	Gly	Ser	Pro 80
Ile	Asn	Thr	Gln	Tyr 85	Val	Phe	Lys	Leu 90	Leu	Gln	Ala	Ser	Gly 95	Gly	Gly
Gly	Ser	Gly 100	Gly	Gly	Gly	Ser	Gly	Gly 105	Gly	Gly	Ser	Ala	Ser 110	Met	Thr
Gly	Gly	Gln 115	Gln	Met	Gly	Arg	Glu 120	Asn	Leu	Tyr	Phe	Gln 125	Gly	Val	Pro
Gly 130	Ser	Ser	Val	Val	Ser	Arg 135	Ala	Gly	Glu	Leu	Ile 140	His	Met	Val	Thr
Leu 145	Asp	Lys	Thr	Gly	Lys 150	Lys	Ser	Phe	Gly	Ile 155	Cys	Ile	Val	Arg	Gly 160
Glu	Val	Lys 165	Asp	Ser	Pro	Asn	Thr	Lys	Thr 170	Thr	Gly	Ile	Phe	Ile 175	Lys
Gly	Ile	Val 180	Pro	Asp	Ser	Pro	Ala	His 185	Leu	Cys	Gly	Arg	Leu 190	Lys	Val
Gly	Asp	Arg 195	Ile	Leu	Ser	Leu	Asn 200	Gly	Lys	Asp	Val	Arg 205	Asn	Ser	Thr
Glu 210	Gln	Ala	Val	Ile	Asp	Leu 215	Ile	Lys	Glu	Ala	Asp 220	Phe	Lys	Ile	Glu
Leu 225	Glu	Ile	Gln	Thr	Phe 230	Asp	Lys								

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<210> SEQ ID NO 7
<211> LENGTH: 186
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: pYS6/CT_HSA_MFalpha1_Fn10_NorpA

<400> SEQUENCE: 7

Met Lys Trp Val Ser Phe Ile Ser Leu Leu Phe Leu Phe Ser Ser Ala
 1             5             10             15

Tyr Ser Arg Ser Leu Asp Lys Arg Glu Asn Leu Tyr Phe Gln Gly Gly
 20             25             30

Ser Val Ser Asp Val Pro Arg Asp Leu Glu Val Val Ala Ala Thr Pro
 35             40             45

Thr Ser Leu Leu Ile Ser Trp Asp Ala Pro Ala Val Thr Val Arg Tyr
 50             55             60

Tyr Arg Ile Thr Tyr Gly Glu Thr Gly Gly Asn Ser Pro Val Gln Glu
 65             70             75             80

Phe Thr Val Pro Gly Ser Lys Ser Thr Ala Thr Ile Ser Gly Leu Lys
 85             90             95

Pro Gly Val Asp Tyr Thr Ile Thr Val Tyr Ala Val Thr Gly Arg Gly
100            105            110

Asp Ser Pro Ala Ser Ser Lys Pro Ile Ser Ile Asn Tyr Arg Thr Glu
115            120            125

Phe Glu Asn Leu Tyr Phe Gln Gly Ser Gly Gly Gly Glu Gln Lys
130            135            140

Leu Ile Ser Glu Glu Asp Leu His His His His His His Pro Ser Thr
145            150            155            160

Pro Pro Thr Pro Ser Pro Ser Thr Pro Pro Thr Pro Ser Pro Ser Tyr
165            170            175

Lys Thr Gln Gly Lys Thr Glu Phe Cys Ala
180            185

<210> SEQ ID NO 8
<211> LENGTH: 97
<212> TYPE: PRT
<213> ORGANISM: Unknown
<220> FEATURE:
<223> OTHER INFORMATION: InaD PDZ domain amino acid sequence

<400> SEQUENCE: 8

Ala Gly Glu Leu Ile His Met Val Thr Leu Asp Lys Thr Gly Lys Lys
 1             5             10             15

Ser Phe Gly Ile Cys Ile Val Arg Gly Glu Val Lys Asp Ser Pro Asn
 20             25             30

Thr Lys Thr Thr Gly Ile Phe Ile Lys Gly Ile Val Pro Asp Ser Pro
 35             40             45

Ala His Leu Cys Gly Arg Leu Lys Val Gly Asp Arg Ile Leu Ser Leu
 50             55             60

Asn Gly Lys Asp Val Arg Asn Ser Thr Glu Gln Ala Val Ile Asp Leu
 65             70             75             80

Ile Lys Glu Ala Asp Phe Lys Ile Glu Leu Glu Ile Gln Thr Phe Asp
 85             90             95

Lys

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<210> SEQ ID NO 9  
 <211> LENGTH: 11  
 <212> TYPE: PRT  
 <213> ORGANISM: Unknown  
 <220> FEATURE:  
 <223> OTHER INFORMATION: NorpA C-terminal

<400> SEQUENCE: 9

Tyr Lys Thr Gln Gly Lys Thr Glu Phe Cys Ala  
 1 5 10

<210> SEQ ID NO 10  
 <211> LENGTH: 737  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: pYS6/CT\* MFalpha HSA-NorpA

<400> SEQUENCE: 10

Met Arg Phe Pro Ser Ile Phe Thr Ala Val Leu Phe Ala Ala Ser Ser  
 1 5 10 15

Ala Leu Ala Ala Pro Val Asn Thr Thr Thr Glu Asp Glu Thr Ala Gln  
 20 25 30

Ile Pro Ala Glu Ala Val Ile Gly Tyr Ser Asp Leu Glu Gly Asp Phe  
 35 40 45

Asp Val Ala Val Leu Pro Phe Ser Asn Ser Thr Asn Asn Gly Leu Leu  
 50 55 60

Phe Ile Asn Thr Thr Ile Ala Ser Ile Ala Ala Lys Glu Glu Gly Val  
 65 70 75 80

Ser Leu Glu Lys Arg Glu Ala Glu Ala Ala Ser Asp Ala His Lys Ser  
 85 90 95

Glu Val Ala His Arg Phe Lys Asp Leu Gly Glu Glu Asn Phe Lys Ala  
 100 105 110

Leu Val Leu Ile Ala Phe Ala Gln Tyr Leu Gln Gln Cys Pro Phe Glu  
 115 120 125

Asp His Val Lys Leu Val Asn Glu Val Thr Glu Phe Ala Lys Thr Cys  
 130 135 140

Val Ala Asp Glu Ser Ala Glu Asn Cys Asp Lys Ser Leu His Thr Leu  
 145 150 155 160

Phe Gly Asp Lys Leu Cys Thr Val Ala Thr Leu Arg Glu Thr Tyr Gly  
 165 170 175

Glu Met Ala Asp Cys Cys Ala Lys Gln Glu Pro Glu Arg Asn Glu Cys  
 180 185 190

Phe Leu Gln His Lys Asp Asp Asn Pro Asn Leu Pro Arg Leu Val Arg  
 195 200 205

Pro Glu Val Asp Val Met Cys Thr Ala Phe His Asp Asn Glu Glu Thr  
 210 215 220

Phe Leu Lys Lys Tyr Leu Tyr Glu Ile Ala Arg Arg His Pro Tyr Phe  
 225 230 235 240

Tyr Ala Pro Glu Leu Leu Phe Phe Ala Lys Arg Tyr Lys Ala Ala Phe  
 245 250 255

Thr Glu Cys Cys Gln Ala Ala Asp Lys Ala Ala Cys Leu Leu Pro Lys  
 260 265 270

Leu Asp Glu Leu Arg Asp Glu Gly Lys Ala Ser Ser Ala Lys Gln Arg  
 275 280 285

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Leu	Lys	Cys	Ala	Ser	Leu	Gln	Lys	Phe	Gly	Glu	Arg	Ala	Phe	Lys	Ala	290	295	300
Trp	Ala	Val	Ala	Arg	Leu	Ser	Gln	Arg	Phe	Pro	Lys	Ala	Glu	Phe	Ala	305	310	315
Glu	Val	Ser	Lys	Leu	Val	Thr	Asp	Leu	Thr	Lys	Val	His	Thr	Glu	Cys	325	330	335
Cys	His	Gly	Asp	Leu	Leu	Glu	Cys	Ala	Asp	Asp	Arg	Ala	Asp	Leu	Ala	340	345	350
Lys	Tyr	Ile	Cys	Glu	Asn	Gln	Asp	Ser	Ile	Ser	Ser	Lys	Leu	Lys	Glu	355	360	365
Cys	Cys	Glu	Lys	Pro	Leu	Leu	Glu	Lys	Ser	His	Cys	Ile	Ala	Glu	Val	370	375	380
Glu	Asn	Asp	Glu	Met	Pro	Ala	Asp	Leu	Pro	Ser	Leu	Ala	Ala	Asp	Phe	385	390	395
Val	Glu	Ser	Lys	Asp	Val	Cys	Lys	Asn	Tyr	Ala	Glu	Ala	Lys	Asp	Val	405	410	415
Phe	Leu	Gly	Met	Phe	Leu	Tyr	Glu	Tyr	Ala	Arg	Arg	His	Pro	Asp	Tyr	420	425	430
Ser	Val	Val	Leu	Leu	Leu	Arg	Leu	Ala	Lys	Thr	Tyr	Glu	Thr	Thr	Leu	435	440	445
Glu	Lys	Cys	Cys	Ala	Ala	Ala	Asp	Pro	His	Glu	Cys	Tyr	Ala	Lys	Val	450	455	460
Phe	Asp	Glu	Phe	Lys	Pro	Leu	Val	Glu	Glu	Pro	Gln	Asn	Leu	Ile	Lys	465	470	475
Gln	Asn	Cys	Glu	Leu	Phe	Glu	Gln	Leu	Gly	Glu	Tyr	Lys	Phe	Gln	Asn	485	490	495
Ala	Leu	Leu	Val	Arg	Tyr	Thr	Lys	Lys	Val	Pro	Gln	Val	Ser	Thr	Pro	500	505	510
Thr	Leu	Val	Glu	Val	Ser	Arg	Asn	Leu	Gly	Lys	Val	Gly	Ser	Lys	Cys	515	520	525
Cys	Lys	His	Pro	Glu	Ala	Lys	Arg	Met	Pro	Cys	Ala	Glu	Asp	Tyr	Leu	530	535	540
Ser	Val	Val	Leu	Asn	Gln	Leu	Cys	Val	Leu	His	Glu	Lys	Thr	Pro	Val	545	550	555
Ser	Asp	Arg	Val	Thr	Lys	Cys	Cys	Thr	Glu	Ser	Leu	Val	Asn	Arg	Arg	565	570	575
Pro	Cys	Phe	Ser	Ala	Leu	Glu	Val	Asp	Glu	Thr	Tyr	Val	Pro	Lys	Glu	580	585	590
Phe	Asn	Ala	Glu	Thr	Phe	Thr	Phe	His	Ala	Asp	Ile	Cys	Thr	Leu	Ser	595	600	605
Glu	Lys	Glu	Arg	Gln	Ile	Lys	Lys	Gln	Thr	Ala	Leu	Val	Glu	Leu	Val	610	615	620
Lys	His	Lys	Pro	Lys	Ala	Thr	Lys	Glu	Gln	Leu	Lys	Ala	Val	Met	Asp	625	630	635
Asp	Phe	Ala	Ala	Phe	Val	Glu	Lys	Cys	Cys	Lys	Ala	Asp	Asp	Lys	Glu	645	650	655
Thr	Cys	Phe	Ala	Glu	Glu	Gly	Lys	Lys	Leu	Val	Ala	Ala	Ser	Gln	Ala	660	665	670
Ala	Leu	Gly	Leu	Gly	Ser	Glu	Asn	Leu	Tyr	Phe	Gln	Gly	Ser	Gly	Gly	675	680	685

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Gly Gly Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu His His His His  
690 695 700

His His His His Pro Ser Thr Pro Pro Thr Pro Ser Pro Ser Thr Pro  
705 710 715 720

Pro Thr Pro Ser Pro Ser Tyr Lys Thr Gln Gly Lys Thr Glu Phe Cys  
725 730 735

Ala

<210> SEQ ID NO 11  
<211> LENGTH: 405  
<212> TYPE: PRT  
<213> ORGANISM: Artificial Sequence  
<220> FEATURE:  
<223> OTHER INFORMATION: pYS6/CT\* MFalpha1-scFv lysozyme-NorpA

<400> SEQUENCE: 11

Met Arg Phe Pro Ser Ile Phe Thr Ala Val Leu Phe Ala Ala Ser Ser  
1 5 10 15

Ala Leu Ala Ala Pro Val Asn Thr Thr Thr Glu Asp Glu Thr Ala Gln  
20 25 30

Ile Pro Ala Glu Ala Val Ile Gly Tyr Ser Asp Leu Glu Gly Asp Phe  
35 40 45

Asp Val Ala Val Leu Pro Phe Ser Asn Ser Thr Asn Asn Gly Leu Leu  
50 55 60

Phe Ile Asn Thr Thr Ile Ala Ser Ile Ala Ala Lys Glu Glu Gly Val  
65 70 75 80

Ser Leu Glu Lys Arg Glu Ala Glu Ala Ala Ser Gln Val Lys Leu Gln  
85 90 95

Gln Ser Gly Ala Glu Leu Val Lys Pro Gly Ala Ser Val Lys Leu Ser  
100 105 110

Cys Thr Ala Ser Gly Phe Asn Ile Lys Asp Thr Tyr Met His Trp Val  
115 120 125

Lys Gln Arg Pro Glu Gln Gly Leu Glu Trp Ile Gly Arg Ile Asp Pro  
130 135 140

Ala Asn Gly Asn Thr Lys Tyr Asp Pro Lys Phe Gln Gly Lys Ala Thr  
145 150 155 160

Ile Thr Ala Asp Thr Ser Ser Asn Thr Ala Tyr Leu Gln Leu Ser Ser  
165 170 175

Leu Thr Ser Glu Asp Thr Ala Val Tyr Tyr Cys Ala Arg Trp Asp Trp  
180 185 190

Tyr Phe Asp Val Trp Gly Gln Gly Thr Thr Val Thr Val Ser Ser Gly  
195 200 205

Gly Gly Gly Ser Gly Gly Gly Gly Ser Gly Gly Gly Gly Ser Asp Ile  
210 215 220

Glu Leu Thr Gln Ser Pro Ser Ser Met Tyr Thr Ser Leu Gly Glu Arg  
225 230 235 240

Val Thr Ile Thr Cys Lys Ala Ser Gln Asp Ile Asn Ser Tyr Leu Arg  
245 250 255

Trp Phe Gln Gln Lys Pro Gly Lys Ser Pro Lys Thr Leu Ile Tyr Tyr  
260 265 270

Ala Thr Ser Leu Ala Asp Gly Val Pro Ser Arg Phe Ser Gly Ser Gly  
275 280 285

Ser Gly Gln Asp Tyr Ser Leu Thr Ile Ser Ser Leu Glu Ser Asp Asp

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290	295	300
Thr Thr Thr Tyr Tyr Cys Leu Gln His Gly Glu Ser Pro Tyr Thr Phe 305 310 315 320		
Gly Gly Gly Thr Lys Leu Glu Ile Lys Arg Ala Ala Glu Gln Lys 325 330 335		
Leu Ile Ser Glu Glu Asp Leu Asn Gly Ser Glu Asn Leu Tyr Phe Gln 340 345 350		
Gly Ser Gly Gly Gly Gly Glu Gln Lys Leu Ile Ser Glu Glu Asp Leu 355 360 365		
His His His His His His His Pro Ser Thr Pro Pro Thr Pro Ser 370 375 380		
Pro Ser Thr Pro Pro Thr Pro Ser Pro Ser Tyr Lys Thr Gln Gly Lys 385 390 395 400		
Thr Glu Phe Cys Ala 405		

<210> SEQ ID NO 12  
 <211> LENGTH: 114  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: pYD\_NBC1\_Aga2-NorpA

<400> SEQUENCE: 12

Met Gln Leu Leu Arg Cys Phe Ser Ile Phe Ser Val Ile Ala Ser Val 1 5 10 15
Leu Ala Gln Glu Leu Thr Thr Ile Cys Glu Gln Ile Pro Ser Pro Thr 20 25 30
Leu Glu Ser Thr Pro Tyr Ser Leu Ser Thr Thr Thr Ile Leu Ala Asn 35 40 45
Gly Lys Ala Met Gln Gly Val Phe Glu Tyr Tyr Lys Ser Val Thr Phe 50 55 60
Val Ser Asn Cys Gly Ser His Pro Ser Thr Thr Ser Lys Gly Ser Pro 65 70 75 80
Ile Asn Thr Gln Tyr Val Phe Lys Leu Leu Gln Ala Ser Gly Gly Gly 85 90 95
Gly Ser Gly Gly Gly Gly Ser Tyr Lys Thr Gln Gly Lys Thr Glu Phe 100 105 110
Cys Ala

<210> SEQ ID NO 13  
 <211> LENGTH: 327  
 <212> TYPE: PRT  
 <213> ORGANISM: Artificial Sequence  
 <220> FEATURE:  
 <223> OTHER INFORMATION: pYS6/CT\* MFalpha1-InaD-Fn10

<400> SEQUENCE: 13

Met Arg Phe Pro Ser Ile Phe Thr Ala Val Leu Phe Ala Ala Ser Ser 1 5 10 15
Ala Leu Ala Ala Pro Val Asn Thr Thr Thr Glu Asp Glu Thr Ala Gln 20 25 30
Ile Pro Ala Glu Ala Val Ile Gly Tyr Ser Asp Leu Glu Gly Asp Phe 35 40 45
Asp Val Ala Val Leu Pro Phe Ser Asn Ser Thr Asn Asn Gly Leu Leu

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50	55	60
Phe Ile Asn Thr Thr	Ile Ala Ser Ile Ala Ala	Lys Glu Glu Gly Val
65	70	75 80
Ser Leu Glu Lys Arg	Glu Ala Glu Ala Ala	Ser Ala Gly Glu Leu Ile
	85	90 95
His Met Val Thr Leu	Asp Lys Thr Gly Lys Lys	Ser Phe Gly Ile Cys
	100	105 110
Ile Val Arg Gly Glu	Val Lys Asp Ser Pro Asn Thr	Lys Thr Thr Gly
	115	120 125
Ile Phe Ile Lys Gly	Ile Val Pro Asp Ser Pro	Ala His Leu Cys Gly
	130	135 140
Arg Leu Lys Val Gly	Asp Arg Ile Leu Ser	Leu Asn Gly Lys Asp Val
	145	150 155 160
Arg Asn Ser Thr Glu	Gln Ala Val Ile Asp	Leu Ile Lys Glu Ala Asp
	165	170 175
Phe Lys Ile Glu Leu	Glu Ile Gln Thr Phe Asp	Lys Ser Gly Gly Gly
	180	185 190
Gly Glu Gln Lys Leu	Ile Ser Glu Glu Asp	Leu His His His His
	195	200 205
His Pro Ser Thr Pro	Pro Thr Pro Ser Pro	Ser Thr Pro Pro Thr Pro
	210	215 220
Ser Pro Glu Asn Leu	Tyr Phe Gln Gly Val	Ser Asp Val Pro Arg Asp
	225	230 235 240
Leu Glu Val Val Ala	Ala Thr Pro Thr Ser	Leu Leu Ile Ser Trp Asp
	245	250 255
Ala Pro Ala Val Thr	Val Arg Tyr Tyr Arg	Ile Thr Tyr Gly Glu Thr
	260	265 270
Gly Gly Asn Ser Pro	Val Gln Glu Phe Thr	Val Pro Gly Ser Lys Ser
	275	280 285
Thr Ala Thr Ile Ser	Gly Leu Lys Pro Gly	Val Asp Tyr Thr Ile Thr
	290	295 300
Val Tyr Ala Val Thr	Gly Arg Gly Asp Ser	Pro Ala Ser Ser Lys Pro
	305	310 315 320
Ile Ser Ile Asn Tyr	Arg Thr	
	325	

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1. A library of host cells, wherein each host cell comprising:

- (a) a cell surface molecule attached to the surface of the cell,
- (b) an adapter molecule comprising a first binding site and a second binding site, and
- (c) a display molecule comprising a modified polypeptide; wherein the first binding site binds specifically to the cell surface molecule and cannot bind to the display molecule and the second binding site binds specifically to the display molecule and cannot bind to the cell surface molecule, and wherein the adapter molecule is not a component of the modified polypeptide.

2. The host cells of claim 1, further comprising a plurality of display molecules.

3. The host cells of claim 1, wherein the host cell surface molecule is covalently linked to the first binding site.

4. The host cells of claim 1, wherein the host cell surface molecule is covalently linked to the first binding site through a disulfide bond.

5. The host cells of claim 1, wherein the host cell surface molecule comprises a first agglutinin that is Aga1p, and wherein the first binding site comprises a second agglutinin that is Aga2p.

6. (canceled)

7. The host cells of claim 1, wherein the host cell surface molecule is attached to the cell membrane via a GPI anchor.

8. (canceled)

9. (canceled)

10. The host cells of claim 1, wherein the second binding site is covalently linked to the display molecule.

11. The host cells of claim 1, wherein the second binding site is covalently linked to the display molecule through disulfide bonds.

12. The host cells of claim 1, wherein the second binding site comprises a PDZ domain of InaD.

13. (canceled)

14. The host cells of claim 12, wherein the display molecule comprises a C-terminal NorpA ligand.

15. The host cells of claim 1, wherein the display molecule comprises a PDZ domain of InaD.

16. (canceled)

17. The host cells of claim 15, wherein the second binding site comprises a C-terminal NorpA ligand.

18. The host cells of claim 1, wherein the modified polypeptide is selected from the group consisting of: a scaffold protein, a signal transduction protein, an antibody, an immunoglobulin, an immunoadhesin, a receptor, a ligand, an oncoprotein, a transcription factor, an enzyme, and a fibronectin polypeptide.

19. The host cells of claim 18, wherein the display molecule is a fibronectin polypeptide.

20. The host cells of claim 19, wherein the fibronectin polypeptide comprises an F10 polypeptide.

21. The host cells of claim 1, wherein the display molecule comprises a secretion signal peptide.

22. The host cells of claim 21, wherein the secretion signal peptide comprises an MFalpha/HSA hybrid leader peptide, or an MFalpha leader peptide.

23. (canceled)

24. The host cells of claim 1, wherein expression of the display molecule is under the control of a first inducible promoter selected from the group consisting of an AOX1 promoter, a CUP 1 promoter, and a Gal 1 promoter.

25. (canceled)

26. (canceled)

27. The host cells of claim 1, wherein the expression of the adapter molecule is under the control of a second inducible promoter selected from the group consisting of an AOX1 promoter, a CUP 1 promoter, and a Gal 1 promoter.

28. (canceled)

29. (canceled)

30. The host cells of claim 1, wherein the host cell is a yeast cell.

31. (canceled)

32. (canceled)

33. A method for displaying a modified polypeptide comprising:

(a) providing a host cell comprising a cell surface molecule attached to the surface of the cell and a first nucleic acid encoding a display polypeptide comprising a modified polypeptide,

(b) contacting the host cell with an adapter molecule comprising a first binding site and a second binding site under conditions wherein the first binding site binds to the cell surface molecule, and

(c) incubating the host cell under conditions wherein the host cell exports the display polypeptide outside the host cell under conditions wherein the second binding site binds to the display polypeptide,

wherein the first binding site binds specifically to the cell surface molecule and cannot bind to the display molecule and the second binding site binds specifically to the display polypeptide and cannot bind to the cell surface molecule, and wherein the adapter molecule is not a component of the modified polypeptide.

34. The method of claim 33, wherein the host cell displays at least  $10^2$ , at least  $10^3$ , at least  $10^4$ , or at least  $10^5$  modified polypeptides.

35. (canceled)

36. (canceled)

37. (canceled)

38. (canceled)

39. (canceled)

40. (canceled)

41. (canceled)

42. (canceled)

43. (canceled)

44. (canceled)

45. (canceled)

46. (canceled)

47. (canceled)

48. (canceled)

49. (canceled)

50. (canceled)

51. (canceled)

52. (canceled)

53. (canceled)

54. (canceled)

55. (canceled)

56. (canceled)

57. (canceled)

58. (canceled)

59. (canceled)

60. (canceled)

61. (canceled)

62. (canceled)

63. (canceled)

64. A method for generating a host cell display library comprising:

introducing into a plurality of host cells a display library of first nucleic acids each encoding a display polypeptide comprising a modified polypeptide, wherein at least two of the introduced first nucleic acids encode different modified polypeptides,

wherein each host cell comprises a second nucleic acid which encodes a cell surface polypeptide and a third nucleic acid which encodes an adapter molecule comprising a first binding site and a second binding site, wherein the first binding site binds to the cell surface molecule but not the display polypeptide and the second binding site binds to the display polypeptide but not the cell surface molecule, and wherein the adapter molecule is not a component of the modified polypeptide.

\* \* \* \* \*