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# (12) United States Patent

Chiang et al.

### (54) INACTIVATED CANINE INFLUENZA VACCINES AND METHODS OF MAKING AND USES THEREOF

(71) Applicant: **BOEHRINGER INGELHEIM ANIMAL HEALTH USA INC.**,

Duluth, GA (US)

(72) Inventors: Yu-Wei Chiang, Athen, GA (US);

**David Cureton**, Alpharetta, GA (US); **Herve Poulet**, Sainte Foy-les Lyon

(FR)

(73) Assignee: **BOEHRINGER INGELHEIM ANIMAL HEALTH USA INC.**,

Duluth, GA (US)

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(51) Int. Cl.

A61K 39/145 (2006.01)

A61P 31/16 (2006.01)

G01N 33/569 (2006.01)

A61K 39/12 (2006.01)

A61K 39/00 (2006.01)

(52) U.S. Cl.

(58) Field of Classification Search

None

See application file for complete search history.

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(Continued)

Primary Examiner — Shanon A. Foley (74) Attorney, Agent, or Firm — Judy Jarecki-Black; Suzanne Shope

### (57) ABSTRACT

The present invention relates to canine influenza virus strains, and vaccines and compositions. The present invention also relates to reagents and methods allowing their detection, methods of vaccination as well as methods of producing these reagents and vaccines.

### 13 Claims, 21 Drawing Sheets

Specification includes a Sequence Listing.

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Figure 1A

SEQ ID NO:	Type	Gene Description
1	DNA	DNA encoding HA protein from #4 isolate (in eggs) of H3N2 CIV
2	Protein	HA protein from #4 isolate (in eggs) of H3N2 CIV
3	DNA	DNA encoding HA protein from #4 isolate (in MDCK cells) of H3N2 CIV
4	Protein	HA protein from #4 isolate (in MDCK cells) of H3N2 CIV
5	DNA	DNA encoding HA protein from #8 isolate (in eggs) of H3N2 CIV
6	Protein	HA protein from #8 isolate (in eggs) of H3N2 CIV
7	DNA	DNA encoding NA protein from #4 isolate (in eggs) of H3N2 CIV
8	Protein	NA protein from #4 isolate (in eggs) of H3N2 CIV
9	DNA	DNA encoding NA protein from #4 isolate (in MDCK cells) of H3N2 CIV
10	Protein	NA protein from #4 isolate (in MDCK cells) of H3N2 CIV
11	DNA	DNA encoding NA protein from #8 isolate (in eggs) of H3N2 CIV
12	Protein	NA protein from #8 isolate (in eggs) of H3N2 CIV
13	oligo	H3N2 NA FWD: 5'-GGG ACC ACG CTG AAC AAT AA-3'
14	oligo	H3N2 NA REV: 5'-TGA AAC GGA ACA CCC AAC TC -3'
15	oligo	H3N8 NA FWD: 5'-GTT CGC CCT CAG AAT GTA GAA-3'
16	oligo	H3N8 NA REV: 5'-CCT ATA CGG ACT TCG ATC CTT TAT T-3'
17	oligo	Ca.H3N2.HA.390R
18	oligo	Ca.H3N2.HA.259F
19	oligo	Ca.H3N2.HA.743F
20	oligo	Ca.H3N2.HA.1276F
21	oligo	Ca.H3N2.HA.1569F
22	oligo	Ca.H3N2.HA.871R
23	oligo	Ca.H3N2.HA.1408R
24	oligo	Ca.H3N2.NA.429F
25	oligo	Ca.H3N2.NA.484R
26	oligo	Ca.H3N2.NA.975F
27	oligo	Ca.H3N2.NA.1023R
28	oligo	Ca.H3N2.1229F
29	oligo	Ca.H3N2.1279R

Figure 1B

SEQ ID	type	Gene Description
NO:		
30	DNA	DNA encoding HA protein of H3N8 CIV strain A/Ca/CT/85863/11
		(GenBank KM359807.1)
31	protein	HA protein of H3N8 CIV strain A/Ca/CT/85863/11 (GenBank
		AIN25426.1)
32	DNA	DNA encoding NA protein of H3N8 CIV strain A/Ca/CT/85863/11
33	protein	NA protein of H3N8 CIV strain A/Ca/CT/85863/11
34	DNA	DNA encoding HA protein of H3N8 CIV strain
		A/canine/NY/120106.2/2011 (GenBank KM359803.1)
35	protein	HA protein of H3N8 CIV strain A/canine/NY/120106.2/2011
		(GenBank AIN25422.1)
36	DNA	DNA encoding NA protein of H3N8 CIV strain
		A/canine/NY/120106.2/2011 (GenBank KM359831.1)
37	protein	NA protein of H3N8 CIV strain A/canine/NY/120106.2/2011
		(GenBank AIN25464.1)
38	DNA	DNA encoding HA protein of H3N8 CIV strain WY/86033/07
39	protein	HA protein of H3N8 CIV strain WY/86033/07
40	DNA	DNA encoding NA protein of H3N8 CIV strain WY/86033/07
41	protein	NA protein of H3N8 CIV strain WY/86033/07
42	DNA	DNA encoding NA protein (SEQ ID NO:37) of H3N8 CIV strain
		A/canine/NY/120106

Figure 2

Real-time RT-PCR genotyping of CIVs

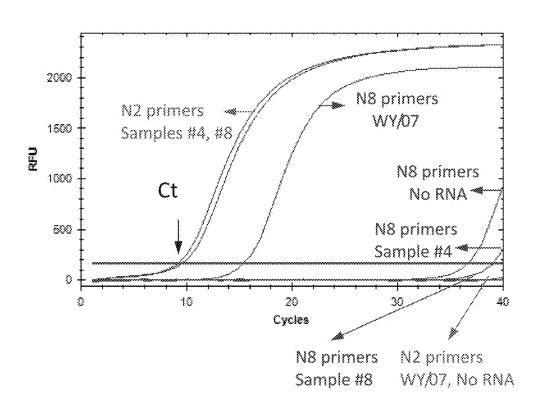
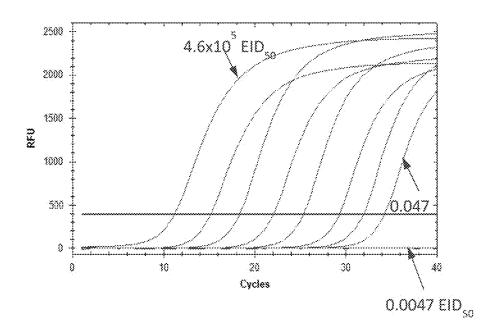
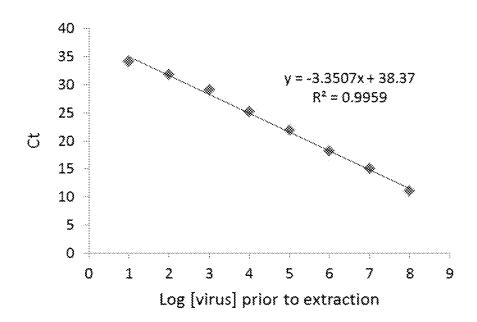


Figure 3
Sensitivity of the subtyping assay



Detection limit =  $0.047 \text{ EID}_{50}$  units

Figure 4
Efficiency of the subtyping assay



Amplification factor = 1.99 Efficiency = 98.81

Figure 5A

Protein sequence alignment of HA protein of H3N2 and sequence identity

Protein sequence alignment of HA protein of H3N2 and sequence iden  SEQ ID NO:2 (1) MRTVIALSY BOLDS ON LONG NAME OF THE PROTEIN SEQ ID NO:4 (1) MRTVIALSY BOLDS ON LONG NAME OF THE PROTEIN SERVICE OF THE PRO	50
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	and the second second
750 TD NO C (1)	
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101	150
SEQ ID NO:2 (101) DEFENSIVATION FOR ASSETLED TESTIVAS	ercora.
SEQ ID NO:4 (101) DIEVERSNAFONCYPYDVPDYASLKSTVASSGTLEFTTEGETWAG	
SEQ ID NO:6 (101) DIFVENSNAFON YPYLVFOYASIRSIVASSGTLEFTTEGETWAD	mosic
151	200
SEQ ID NO:2 (151) GSGACKPGPANSFFSBLAMLTWSSNTYFVLBVTMFNRNNFDKLYI	<b>XGVIIII</b>
SEQ ID NO:4 (151) GGGACKEGPANSFESRI.WELTKSGNEEPVINVTMPNNMMFURLYI	ACVER
SEQ ID NO:6 (151) SSCACERGEANSFESBLANDETS SONTYPYLDYTHERNNNFDELYT 201	%GV88 250
SEQ ID NO:2 (201) STRONG SIXIOAS A STRESON DE SERVICES	13080
SEO ID NO:4 (201) PSTNOSCTSLYTOASCRYKVSTRKSOUTTIPNICSRELVEGUSCR	7 (1786)
SEQ ID NO:6 (201) PSINOROTSIATOASCAN® STRENOCTI PRICERPLANCESA	130
251	300
SEQ ID NO:2 (251) TIVEPSSILVINSMONLIAPROYFEMBLOKSSIMESDAPITETS	SCITE
SEQ ID NO:4 (251) TIVERSULLVINGESSLIAFRS YEVMELGESSLIMESDAPIDTOLS	ECITE
SEQ ID NO:6 (251) TIVKPGDILVINSNGNLIAPRGYFKMHIGKSSIMRSDAFIDTCIS 301	ECITP 350
SEQ ID NO:2 (301) WSSIPNERFICAVARITYGACEKVAQATIKLATONANVELEQIE	53 80 80 80 80 80 80 80 80 80 80 80 80 80
	33.8KBA
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SEQ ID NO:4 (351) AGE 128 ACONV. WY SER STORY CARD.	
SEQ ID NO:6 (351) AGREEMENT WEST SERVING TO A ADDRESS OF THE SERVING THE SERVI	
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SEQ ID NO:2 (401) IEXTNEXFHQIEREFTEVECKIQDLERY/EDTKYDLWSYNAELLY	
SEQ ID NO:4 (401) TEXTMENSIONERS EVEN COLLERS VED TX VOLUMEN NASILLA	
SEQ ID NO:6 (401) TEXTNEXFROIENEESEVSCRIQUIESEVEUTKVOLWSYMAELDV. 451	84.839Q E.O.O.
	500
SEQ ID NO:4 (451) NIIILT SEMNKLER BLEEN BLEEN BLOCK IN NEW ACTE. SEQ ID NO:6 (451) NIIILT SEMNKLEEN TRACLER NAEDMINGER KINKED NA IE.	9200000 93500
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SEQ ID NO:6 (551) IMWACQRONIRONICI	
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SEQ ID NO:2 v. SEQ ID NO:6: 99.6%	
SEQ ID NO:4 v. SEQ ID NO:6: 99.6%	

Figure 5B DNA sequence alignment of polynucleotide encoding HA protein of H3N2 and sequence identity

				A protein of 115142 and sequence identity	^
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				101 150	ä
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SEQ	ID	NO:3	(101)		Í
SEQ	ID	NO:5	(101)		Š
				151 200	j
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		NO:3	(151)	GARAGO ANG CARAGO ANG TAO TAO TAO ANG ANG TAO	ř.
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		NO:1	(201)	ATGC ACAATO COL AAGATTCTTGATGGGAGGGACTCACACTAATA	Š
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# Figure 5C

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SEQ	ID	NO:1	(651)		330
SEQ	ID	NO:3	(651)	RCTCHARTCTCTROLLEGERARGCTRACTCRACTCRACTCRACTCRACTCRACTCRACT	77763
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SEQ	ID	NO:5	(701)		90
				751	300
SEQ	ID	NO:1	(751)		CI
		NO:3	(751)		XX
SEQ	ID	NO:5	(751)	ACAATAG CAAACTIGGAACATAG COTAATAAA AACAGTAATG AAA	CT
					350
SEQ	ID	NO:1	(801)	AATOGOTOOTOGAGATACTTOAAAATGCACATTGGGAAAAGCTGAA	XX.
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SEQ	ID	NO:3	(1201)	ATTICAAAAAACGAATGA AAGTI CCATCAAATTI AAAAAGAGTTTTI C	338
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		NO:1	(1251)		:AG
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### Figure 5D

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SEQ ID NO:1 (1301) TAXABLE AGAINST TAXABABLE AGAINST TAXABABABLE AGAINST TAXABABABLE AGAINST TAXABABABLE AGAINST TAXABAB
SEQ ID NO:3 (1301) TAGAICTTT STOCT TAGAAACCAS
SEQ ID NO:5 (1301) TAGATCTTTGGTCTTACAATGCCACCTTCTTTTGCCTTACAAAACCAG
                                                                                                   1351
SEQ ID NO:1 (1351) AACACATTGATTGACTGATTCAGAAATGAACAAATTGTTTGAAAAGGAC
SEQ ID NO:3 (1351) AACACAATTGATTTARCTGATTCAGAAATGAACAAATTGTTTGAAAAAGAC
SEQ ID NO:5 (1351) AACACAATTGATTTAACTGATTCAGAAATCAACAAATTCTTTGAAAAGAC
                                                                                                    1401
SEQ ID NO:1 (1401) TAGGAGGCAATTGAGGGAAAATGCTGAAGACATGGCCAATGGCTGCTTCA
SEQ ID NO:5 (1401) TAGGAGGGAATTGAGGGAAATT TAAGAATGGGAATGGCGAATGGCTTCA
                                                                                                    1451
SEQ ID NO:1 (1451) ACATA ACCOUNTS AND ACCOUN
SEQ ID NO:3 (1451) AGATATACCACRACTCTTGCATAGAATCGATTAGAAACGGA
SEQ ID NO:1 (1501) ACTIVACIAN ATATATANA ATATATANA AGATOAN AGTOAN AATOGOTTONA
SEQ ID NO:3 (1501) ACTTATGA CATAACATATATAGAGATGAGGCAGTGAACAATGGSTTCCA
                                                             (1501) ACTIVATOR CATAR ATATATA ASATOR CANCAR ARATOSTICA
SEQ ID NO:5
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                                                            (1551)
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SEQ ID NO:5
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Sequence identity:
SEQ ID NO:1 v. SEQ ID NO:3: 99.9%
SEQ ID NO:1 v. SEQ ID NO:5: 99.9%
SEQ ID NO:3 v. SEQ ID NO:5: 99.8%
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Figure 5E

Protein seque	Protein sequence alignment of HA protein of H3N8 and sequence identity							
	1 50							
~	(1) MKTTIILILITHWA <b>X</b> ONPISSKHTATICISHHAVANSTLVKTMSDDQTE (1) MKTTIILILITHWA <b>X</b> SONPISSKNTATICISHHAVANSTLVKTMSDDQTE							
<del></del>	(1) (1) NATT 11 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1							
SEQ ID NO:31 (	51) VIA 11.00 VIA							
SEQ ID NO:35 (S	51) VIKVIII VS INGENIORES (M. 1888) INDENIORES (M.							
SEQ ID NO:39 (	51) VTNATELVOSISMOKIONESTRADORNOTTLIDAMLODPHODAFOYESWO 101 150							
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SEQ ID NO:39 (15	51) SGACKEGSADSFFSRINWITKSSSSYFTINVTMENN#NFOKLY1WGIHHE 201 250							
	251 300							
	51) IVEOLIA INCOMENIA APPEARANTE A PROPERTI DE LA PROPERTI DEL PROPERTI DE LA PROPERTI DE LA PROPERTI DE LA PROPERTI DEL PROPERTI DE LA PROPE							
	51) IVKOGOTANINOMIVARKOTEKIA 🛊 SISS 🛊 SIVELIT VEKITEM							
	51) IVERCOLLMINS ONL VAPROTE IN TOX 55 <b>W</b> 5 VELO I CONTROL 350							
SEQ ID NO:31 (30								
	O1) GSTSNERFFORVARVTYGRUFRYTRONTLKLATGMENVFEROTEGIFGAT O1) GSTSNERFFORVARVTYGRUFRYTRONTLKLATGMENVFEROTEGIFGAT							
SEQ ID NO:31 (35	351 400							
	51)							
	51) AGETENGWEGWEGWEGERYONSESTGOADLKSTOAALIGENGKLNEVE 401 450							
SEQ ID NO:31 (40	Ol) ERTHER BOTTERE SEVEERIOD ERTYED KIDLESYNAELIVALENDE							
SEQ ID NO:35 (40	01) ERTNERFROIEKETSEVE <b>S</b> RIOOLEKYVELTRIOLKSYNAELLVALEROH							
SEQ ID NO:39 (40	01) BRINEKFHOIEKEFSEVE <b>S</b> RIODLENYVEDIKIDL®SYNAELIVALENOM 451 500							
SEQ ID NO:31 (45	51) TIDETOAKONEERE REGISENAR MCDC EKIYHKUDAACTESIATU							
SEQ ID NO:35 (45	51) TIOLOGA MIKLEEKTHELLEMAE MONOFELYHK INACTESIETTI							
SEQ ID NO:39 (45	51) TIDETDAKANKIFERTARQIRENARDMGGGGEKIYHKUDNACTESIRTÖ 501 550							
SEQ ID NO:31 (50	O1) YORYTYKARARIYAT KAYELKO YAYATI WISEATOLEI IL YYILGEL							
	01) YORY OF A STREET BOOK OF STREET BOOK OF STREET							
SEQ ID NO:39 (50	D1) YDHYIYADEA NAFOIKOVELKOVADWILWISFAISCFIICVVLLGFI 551 565							
	51) MMACQMCMCMCCT							
	51) MWACQKIKIKONICI							
	51) MWACQNGNINCNICI							
Sequence identity								
SEQ ID NO:31 v. SI								
SEQ ID NO:31 v. SI	EQ ID NO:39: 97.9%							
SEQ ID NO:35 v. SI	EQ ID NO:39: 97.7%							

Figure 5F **DNA sequence alignment of polynucleotide encoding HA protein of H3N8 and sequence identity** 

sequence identity					
050	T D	270.20	147	1	50
		NO:30	(1)		1 ( . 23 32 23 1
		NO:34	(1)		
SEQ	TD.	NO:38	(1)	51	100
		NO:30	(51)		A200
SEQ	ID	NO:34	(51)	AAA COA A CAA CAA AA AA AA AA AA AA AA AA AA A	4TG
SEQ	ID	NO:38	(51)		arc
				101	150
SEQ	ID	NO:30	(101)		SAG
SEQ	ID	NO:34	(101)		3A0
SEQ	ID	NO:38	(101)		3AG
					200
		MO:30	(151)		X27.0
		NO:34	(151)	T , $T$	93°C
SEQ	ID	NO:38	(151)	201	830 250
SEO	ID	NO:30	(201)		833
		NO:34	(201)	CANCARATO AT ATAKA (A CATACATOCA A CANATTOCA CATTA AT A C	ATG
,		NO:38	(201)	A	ATG
~			, ,	251	300
SEQ	ID	NO:30	(251)		3A.C
SEQ	ID	NO:34	(251)		3AC
SEQ	ID	NO:38	(251)	CAATOO TAOO AGACOO CACOO GACOO CITTI CAGTATGAGACTIO	3AC
				301	350
SEQ	ID	NO:30	(301)	TOTAL AREA CARCARTOTTA AREA TO A TOTAL AREA TO	::A:T
SEQ	ID	NO:34	(301)		28.7
SEQ	ID	NO:38	(301)		CAT
				351	400
SEQ	ID	NO:30	(351)		108
		NO:34	(351)		rc <b>o</b>
SEQ	ID	NO:38	(351)		19A
				401	450
		NO:30	(401)		AGA
		NO:34	(401)		403A
SEQ	ID	NO:38	(401)		868
		00			500
		NO:30	(451)		388 3
		NO:34	(451)		SBB
SEQ	ΙĐ	NO:38	(451)	Eng.	8888 C C A
CEO.	TT	NO:30	(501)	501	550 ****
					1 3 3 3 3 4 4 4 4 4 4 4 4 4 4 4 4 4 4 4
_		NO:34	(501)	TAN	2000 2000
ನಿಲ್ಲಿ	±Đ	NO:38	(501)	551	600
SEO	TD	NO:30	(551)		
		NO:34	(551)	TAA AATGAAATTI GACAAG TATA AT TUU AATTA ATCAC	
		NO:38	(551)		00A
Oug			1007)		650
SEO	TD	NO:30	(601)	200	ace Acer
		NO:34	(601)	Transference	
		NO:38	(601)	A2.470.52	000000000000000000000000000000000000000
~ ~~ ><	~		(002)		occurrence.

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# Figure 5G

Figure 5G					
				651 700	
		NO:30	(651)		
1.77		NO:34	(651)	T T	
SEQ	ID	NO:38	(651)	A.	
				701 750	
		NO:30	(701)		
		NO:34	(701)		
SEQ	ID	NO:38	(701)	G:	
				751 800	
		NO:30	(751)		
*		NO:34	(751)		
SEQ	TD	NO:38	(751)	801 850	
CEO	TD	NO:30	(801)	801 850	
		NO:34	(801)	T	
		NO:34	(801)		
Dug	2.0	110.00	(001)	851 900	
SEQ	ID	NO:30	(851)		
		NO:34	(851)		
		NO:38	(851)		
				901 950	
SEQ	ID	NO:30	(901)		
SEQ	ID	NO:34	(901)		
SEQ	ID	NO:38	(901)		
				951 1000	
,		NO:30	(951)		
		NO:34	(951)		
SEQ	1D	NO:38	(951)	TOSA A A TOCOCO A STATE TO A SOCIAL ANALES TITTA A SOCIAL ACTOR	
ana	TO	MO - 20	(1001)	1001 1050	
		NO:30	(1001) (1001)		
		NO:38	(1001)	GATUACOANTO TA CASARA ANA CASARA TOTO A CASARA	
OLOQ	1.10	NOTO	(1001)	1051 1100	
SEO	ID	NO:30	(1051)		
		NO:34	(1051)		
		NO:38	(1051)		
				1101 1150	
SEQ	ID	NO:30	(1101)		
SEQ	ID	NO:34	(1101)		
SEQ	ID	NO:38	(1101)		
				1151 1200	
		NO:30	(1151)		
		NO:34	(1151)		
SEQ	ID	NO:38	(1151)		
OB O	7 5	NO - 20	(1001)	1201 1250	
		NO:30	(1201) (1201)		
		NO:34	(1201)		
Uni	لطند	110.00	(1201)	1251 1300	
SEO	ΤĐ	NO:30	(1251)	1231 1300	
		NO:34	(1251)		
		NO:38	(1251)		
			,,		

### Figure 5H

rigure 3ri						
				1301 1350		
SEQ	ID	NO:30	(1301)			
_		NO:34	(1301)	AT A		
SEQ	ID	NO:38	(1301)			
				1351 1400		
		NO:30	(1351)			
		NO:34	(1351)			
SEQ	ID	NO:38	(1351)	A A A A A A A A A A A A A A A A A A A		
				1401 1450		
_		NO:30	(1401)			
		NO:34	(1401)	AAA		
SEQ	ID	NO:38	(1401)	ACCURACIAGA CALACA CONSTRUCTION OF CONSTRUCTIO		
				1451 1500		
		NO:30	(1451)			
		NO:34	(1451)	G.		
SEQ	ID	NO:38	(1451)	TO A AAR		
ar o	are sin	270 00	/25011	1550		
		NO:30	(1501)			
		NO:34	(1501)			
SEQ	TD	NO:38	(1501)	1551 1600		
OTO	ŦΡ	NO:30	(1551)	1551		
		NO:34	(1551)			
		NO:38	(1551)	G		
ನಿಭಿಲ್ಲ	117	140.20	(1001)	G.		
				1601 1650		
SEO	τĐ	NO:30	(1601)	1001		
		NO:34	(1601)			
		NO:38	(1601)			
			(,	1651 1695		
SEO	ID	NO:30	(1651)			
		NO:34	(1651)			
SEQ	ID.	NO:38	(1651)			
-						
		ce ide				
SEQ	ID	NO:30	v. SEQ	ID NO:34: 99.3%		
				ID NO:38: 98.1%		
SEQ	ID	NO:34	v. SEQ	ID NO:38: 97.8%		

### Sequence identity between HA proteins of H3N2 and H3N8

SEQ ID NO:	2	4	6	31	35	39
2		99.6%	99.6%	84.4%	84.8%	85.4%
4			99.6%	84.8%	84.8%	85.3%
6				85.0%	85.0%	85.5%
31					98.9%	97.9%
35						97.7%
39						

Figure 5I

<u>Protein se</u>	quence	alignment of NA protein of H3N2 and sequence identity
070 70 110 6		1 50
SEQ ID NO:8	(1)	
SEQ ID NO:10	(1)	
SEQ ID NO:12	(1)	51 100
SEQ ID NO:8	(51)	
SEQ ID NO:10	(51)	
SEQ ID NO:12	(51)	101 150
SEQ ID NO:8	(101)	
SEQ ID NO:10	(101)	
SEQ ID NO:12	(1.01)	SKONSTRLEAGGETWYTREPYVSCOHSKCYQFALGQGTTLKNKHENSTIH 151 200
SEQ ID NO:8	(151)	
SEQ ID NO:10	(151)	DRISHBILLBURELGVPFHLGIXQVCIANGSSSCHDGXAMLHVCVIGDDRN
SEQ ID NO:12	(151)	DRTSHRTLLMNELGVEFHLGTKOVCIAWSSSCHEGKAWLHVCVTGDERN 201 250
SEQ ID NO:8	(201)	
SEQ ID NO:10	(201)	
SEQ ID NO:12	(201)	ATASEVYNUMLVOSIGSWSRNILBTGESECVCTRCTCTVVMTDGSASCRA 251 300
SEQ ID NO:8	(251)	
SEO ID NO:10	(251)	
SEQ ID NO:12	(251)	
SEQ ID NO:8	(301)	
SEO ID NO:10	(301)	
SEQ ID NO:12	(301)	
SEQ ID NO:8	(351)	
SEQ ID NO:10	(351)	
SEQ ID NO:12	(351)	450 450
SEQ ID NO:8	(401)	
SEQ ID NO:10	(401)	
SEQ ID NO:12	(401)	NNWSCYST FYEORSCYNPORYEL IROS QUEEN WMTSN STV FCCTS 451 469
SEQ ID NO:8	(451)	
SEQ ID NO:10	(451)	CTYCTCCWFDGANINGNET
SEQ ID NO:12	(451)	
Sequence iden		
		ID NO:10: 100.0%
		ID NO:12: 100.0%
SEQ ID NO:10	v. SEQ	ID NO:12: 100.0%

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Figure 5J

DNA sequence alignment of polynucleotide encoding NA protein of H3N2 and sequence identity

	1 Sequence identity					
CEO	TD	NIO . 7	723	1 30		
		NO:7	(1)			
			, ,	ATGAACCAAATGAAAAGATAATAGCAATAGGTCTGGTTTTTAACCAT		
SEQ	2.17	NO:11	(1)	51 100		
SEO	TD	NO:7	(51)	51 100		
		NO:9	(51)			
		NO:11	(51)			
onă		110.11	(01)	101 150		
SEO	TD	NO:7	(101)			
		NO:9	(101)			
		NO:11	(101)			
<u>sc</u>			(/	151 200		
SEQ	ID	NO:7	(151)			
SEQ	ID	NO:9	(151)			
SEQ	ID	NO:11	(151)	GTGCCGTGTAAACCAATCATAATAGAAAGGAACATAACAGAGGTAGTATA		
				201 250		
SEQ	ID	NO:7	(201)			
SEQ	ID	NO:9	(201)			
SEQ	ID	NO:11	(201)			
				251 300		
		NO:7	(251)			
		NO:9	(251)			
SEQ	ID	NO:11	(251)			
				301 350		
-		NO:7	(301)			
		NO:9	(301)			
SEQ	ΙD	NO:11	(301)	351 400		
SEO	ID	NO:7	(351)			
SEQ	ID	NO:9	(351)			
SEQ	ID	NO:11	(351)			
				401 450		
SEQ	ID	NO:7	(401)			
SEQ	ID	NO:9	(401)			
SEQ	ID	NO:11	(401)			
				451 500		
		No:7	(451)			
		NO:9	(451)			
SEQ	TD	NO:11	(451)	F01 550		
ono	rr	4TO 6 "7	(501)	501 550		
		NO:7	, ,			
		NO:9	(501)			
೨೬೪	ıυ	NO:11	(501)	551 600		
SEO	TD	NO:7	(551)	331 800		
		NO:9	(551)			
		NO:11	(551)	ACCATIC CARACTER OF COTTACATOTT TO TOTAC ACCATICATION OF CARACTER		
~~~	~~		(~~~/	601 650		
SEQ	ID	NO:7	(601)			
		NO:9	(601)	CONTRACTOR		
		NO:11	(601)	COLACTIONTACTOR TO THE ATAMECIA AT CONTROL OF THE ACAD AT THE COLO		

				Figure 5K
				651 700
SEQ	ID	NO:7	(651)	ATO TO COMPANY OF CASE AND AREAST CASE AND AREAST CONTINUES.
SEQ	ID	NO:9	(651)	
SEQ	ID	NO:11	(651)	701 750
SEQ	ID	NO:7	(701)	
SEQ	ID	NO:9	(701)	
SEQ	ID	NO:11	(701)	751 800
SEQ	ID	NO:7	(751)	
SEQ	ID	NO:9	(751)	
SEQ	ID	NO:11	(751)	801 850
SEQ	ID	NO:7	(801)	
		NO:9	(801)	ATTOTOAGGACTOCTOAAGACATAGAGGAATGTTCCTGTTATCCCGGAT
SEQ	ID	NO:11	(801)	ATTUTCAGGGAGTGCTCAACACATAGAGGAATGTTCCTGTTATCCCGAT 851 900
SEO	ID	NO:7	(851)	
		NO:9	(851)	
SEQ	ID	NO:11	(851)	ATCCAAATGTTAGATGTGTTTGCAGAGACAATTGGAAGGGTTCCAATAG 901 950
SEO	ID	NO:7	(901)	
		NO:9	(901)	
		NO:11	(901)	951 1000
SEO	ID	NO:7	(951)	
		NO:9	(951)	
		NO:11	(951)	
<u>F</u>			(,	1001 1050
SEO	ID	NO:7	(1001)	
		NO:9	(1001)	GCASTAACTOCAAGGATCCTAATAATGAGAGAGAGGGAATCCAGGAGTGAAC
SEQ	ID	NO:11	(1001)	CASTAACTGCAAGSATCTAATAATCAGAGGGAATCCAGGAGTAA 1051 1100
SEQ	ID	NO:7	(1051)	
SEQ	ID	NO:9	(1051)	
SEQ	ID	NO:11	(1051)	1101 1150
SEQ	ID	NO:7	(1101)	
SEQ	ID	NO:9	(1101)	
SEQ	ID	NO:11	(1101)	1151 1200
SEQ	ID	NO:7	(1151)	
SEQ	ID	NO:9	(1151)	
SEQ	ΙĎ	NO:11	(1151)	1201 1250
SEO	ID	NO:7	(1201)	
		NO:9	(1201)	
		NO:11	(1201)	1251 1300
SEO	TD	NO:7	(1251)	
		NO:9	(1251)	
		NO:11	(1251)	TOTTAN ASSETT TOTTAN TO ACTUATION GOGACO CACARON A

### Figure 5L

		1301 1350
SEQ ID NO:7	(1301)	
SEQ ID NO:9	(1301)	
SEQ ID NO:11	(1301)	
		1351 1400
SEQ ID NO:7	(1351)	
SEQ ID NO:9	(1351)	
SEQ ID NO:11	(1351)	COTACO A CARA A CONTRATO CATO CATO CATO CATO CATO CATO CATO C
		1401
SEQ ID NO:7	(1401)	
SEQ ID NO:9	(1401)	
SEQ ID NO:11	(1401)	

### Sequence identity

SEQ ID NO:7 v. SEQ ID NO:9: 100.0% SEQ ID NO:7 v. SEQ ID NO:11: 99.9% SEQ ID NO:9 v. SEQ ID NO:11: 99.9%

# DNA sequence alignment of polynucleotide encoding NA protein of H3N8 and sequence identity

	1 50
SEQ ID NO:32 (1)	ATTO ACCOMAND AND AND AND AND AND AND AND AND AND
SEQ ID NO:36 (1)	
SEQ ID NO:40 (1)	ATOMA <b>S</b>
SEQ ID NO:42 (1)	A STANA COMMANDA A A A A A A A A A A A A A A A A A A
	51 100
SEQ ID NO:32 (51)	
SEQ ID NO:36 (51)	
SEQ ID NO:40 (51)	ATTAATCATTAATGTCATTTTCCATGTAGTCAGCATTATAGTAACAGTAC
SEQ ID NO:42 (51)	
	101 150
SEQ ID NO:32 (101)	
SEQ ID NO:36 (101)	
SEQ ID NO:40 (101)	The Control of the Co
SEQ ID NO:42 (101)	
	151 200
SEQ ID NO:32 (151)	
SEQ ID NO:36 (151)	
SEQ ID NO:40 (151)	ACCOMANDA CONTRACTOR ACCOMANDA CONTRACTOR ACCOMANDA CONTRACTOR ACCOMANDA CONTRACTOR ACCOMANDA CONTRACTOR ACCOM
SEQ ID NO:42 (151)	
	201 250
SEQ ID NO:32 (201)	CAR COMMISSION OF THE STATE OF
SEQ ID NO:36 (201)	
SEQ ID NO:40 (201)	
SEQ ID NO:42 (201)	
	251 300
SEQ ID NO:32 (251)	
SEQ ID NO:36 (251)	
SEQ ID NO:40 (251)	ATAA S
SEQ ID NO:42 (251)	

# Figure 5M

				Figure 5M
				301 350
SEQ	ID	NO:32	(301)	
SEQ	ID	NO:36	(301)	
SEQ	ID	NO:40	(301)	
SEO	ID	NO:42	(301)	
~ ~			. ,	351 400
SEO	ID	NO:32	(351)	
		NO:36	(351)	
		NO:40	(351)	G
		NO:42	(351)	
			, ,	401 450
SEQ	ID	NO:32	(401)	
SEQ	ID	NO:36	(401)	
SEO	ID	NO:40	(401)	
		NO:42	(401)	
			(/	451 500
SEO	TD	NO:32	(451)	
		NO:36	(451)	
		NO:40	(451)	TT
		NO:42	(451)	
Sug	1.0	1,0.12	(151)	501 550
SEO	ΤD	NO:32	(501)	
		NO:36	(501)	
		NO:40	(501)	GATTER STATE OF THE STATE OF TH
		NO:42	(501)	
Ong	11	110172	(301)	551 600
SEO	TD	NO:32	(551)	
		NO:36	(551)	
		NO:40	(551)	C
		NO:42	(551)	
SHY	1. 1.	110,42	(001)	601 650
8 P/O	TD	NO:32	(601)	001
		NO:36	(601)	
		NO:40	(601)	
SEQ	±υ	NO:42	(601)	651 700
SEQ	ΤĐ	NO:32	(651)	700
		NO:36	(651)	
		NO:40	(651)	AG TO AT A TO THE A SAME OF A A AG TO AT A TO THE A A A A A A
		NO:42	(651)	
Quit	4.17	NO. 32	(001)	701 750
SEO	TD	NO:32	(701)	ACACTETTATTO TAATOR TOO TOO TOO AAATA CAAA
		NO:36	(701)	GAGACTGTTATTCGGTAATGACTGATGGACCGGCAAATAGGCAASCT <b>A</b> AT
		NO:40	(701)	GAA
		NO:42	(701)	GACACTOTTATTOGGTAATGACTGATGGACCGGAAATAGGCAAGCT
V HV	# # <i>\</i>	1VV . 34	(101)	751 800
SEO	TD	NO:32	(751)	731 800
		NO:36	(751)	
		NO:40	(751)	TATA GALANGA TATA TATA GALANGA TATA TATA GALANGA TATA TATA GALANGA TATA TATA TATA TATA TATA TATA TATA
-		NO:42	(751)	
JEQ	.±1)	110.4Z	(191)	

## Figure 5N

		801 850
SEQ ID NO:32	(801)	
SEQ ID NO:36	(801)	AACTTTCAATGGGGGACACATAGAGGAGTGTTCTTGTTACCCCAATGAAG
SEQ ID NO:40	(801)	AAGTTT AATGGGGACACATAGAGGAGTGTTTTTTTTTTAGCCAATGAAG
SEQ ID NO:42	(801)	
		851 900
SEQ ID NO:32	(851)	
SEQ ID NO:36	(851)	
SEQ ID NO:40	(851)	(G, A, A, G, G, G, A, A, G, G, G, A, A, G,
SEQ ID NO:42	(851)	
		901 950.
SEQ ID NO:32	(901)	
SEQ ID NO:36	(901)	
SEQ ID NO:40	(901)	
SEQ ID NO:42	(901)	
00	1054	951 1000
SEQ ID NO:32	(951)	
SEQ ID NO:36	(951)	
SEQ ID NO:40	(951)	
SEQ ID NO:42	(951)	1001 1050
SEQ ID NO:32	(1001)	1001 1050
**	(1001)	
***	(1001)	G ATTACA TO THE CONTRACTOR OF
**	(1001)	
DDZ ID MO: 45	(1001)	1051 1100
SEQ ID NO:32	(1051)	
	(1051)	
**	(1051)	A
	(1051)	
	,	1101 1150
SEQ ID NO:32	(1101)	AAGAT AAGATTI GAAATAA TAAAAA TAA GCCCCCAA AAGAA
SEQ ID NO:36	(1101)	
SEQ ID NO:40	(1101)	aagat caggatti gaaataataaaaan caggati 👣 gaa agaa agaa Ca
SEQ ID NO:42	(1101)	
		1151 1200
SEQ ID NO:32	(1151)	T. A. C.
SEQ ID NO:36	(1151)	
	(1151)	
SEQ ID NO:42	(1151)	
		1201 1250
~	(1201)	
	(1201)	
	(1201)	CATATA CTĞACAAAAAAGCCA G
SEQ ID NO:42	(1201)	1202
ODO TO NO.00	(1001)	1251 1300
	(1251)	TTTOCTOCCTOTTTCTOCCTTGAAATGATTAGAGGTAAACCTGAAGAA TTTGGTCCCCTGTTTCTGGGTTGAAATGATTAGAGGTAAACCTGAAGAA
	(1251)	
	(1251)	
SEQ ID NO:42	(1251)	

### Figure 50

SEQ ID NO:32 (130 SEQ ID NO:36 (130 SEQ ID NO:40 (130 SEQ ID NO:42 (130	1) CARTATATORUS TACACS 1) CARTATATORUS TACACS 1) CARTATATORUS CONTRACACS	1350 CATTORA TATANA ATANA ATAN
SEQ ID NO:32 (135 SEQ ID NO:36 (135	1) &&&##################################</th><th></th></tr><tr><td>SEO ID NO:40 (135</td><td colspan=3>D NO:40: 97.6%</td></tr><tr><td>SEQ ID NO:42 (135</td><td>* 1000000000000000000000000000000000000</td><td>1</td></tr><tr><td>3EQ ID NO.42 (133</td><td>1)</td><td></td></tr><tr><td></td><td>1401</td><td></td></tr><tr><th>SEO ID NO:32 (140</th><th>3656666666666666666666666666</th><th></th></tr><tr><td>SEO ID NO:36 (140</td><td>* 18880000000000000000000000000000000000</td><td></td></tr><tr><td>SEQ ID NO:40 (140</td><td>* 1000000000000000000000000000000000000</td><td></td></tr><tr><td>SEO ID NO:42 (140</td><td>* * ***********************************</td><td></td></tr><tr><td>, , , , , , , , , , , , , , , , , , , ,</td><td>T /</td><td></td></tr><tr><td>Sequence identity</td><td></td><td></td></tr><tr><td>SEQ ID NO:32 v. SEC</td><td>OID NO:36: 99.5%</td><td></td></tr><tr><th>SEQ ID NO:32 v. SEQ</th><th>) ID NO:40: 97.6%</th><th></th></tr><tr><th>SEQ ID NO:32 v. SEQ</th><th>-</th><th></th></tr><tr><th>SEQ ID NO:36 v. SEC</th><th></th><th></th></tr><tr><th></th><th>•</th><th></th></tr><tr><th>SEQ ID NO:36 v. SEC</th><th>•</th><th></th></tr><tr><td>SEQ ID NO:40 v. SEC</td><td>Q ID NO:42: 98.0%</td><td></td></tr></tbody></table>	

# Protein sequence alignment of NA protein of H3N8 and sequence identity

				1
SEQ	ID	NO:33	(1)	
SEQ	ID	NO:37	(1)	
SEQ	ID	NO:41	(1)	MORPOGRAL AL GEALLGILLIA MALLIA SALITA SALITA DEL MORRETTE LO YMERLIA
				51 100
SEQ	ID	NO:33	(51)	The state of the s
SEQ	ID	NO:37	(51)	
SEQ	ID	NO:41	(51)	$\ddot{\mathrm{T}}$
				101 150
SEQ	ID	NO:33	(101)	
SEQ	ID	NO:37	(101)	
SEQ	ID	NO:41	(101)	
				151 200
SEQ	ID	NO:33	(151)	
SEQ	ID	NO:37	(151)	
SEQ	ID	NO:41	(151)	
				201 250
SEQ	ID	NO:33	(201)	
SEQ	ID	NO:37	(201)	
SEQ	ID	NO:41	(201)	

Apr. 14, 2020

### Figure 5P

		251 300
SEQ ID NO:33	(251)	
SEQ ID NO:37	(251)	
SEQ ID NO:41	(251)	
		301 350
SEQ ID NO:33	(301)	
SEQ ID NO:37	(301)	
SEQ ID NO:41	(301)	
		351 400
SEQ ID NO:33	(351)	$\Gamma$
SEQ ID NO:37	(351)	
SEQ ID NO:41	(351)	
		401. 450
SEQ ID NO:33	(401)	
SEQ ID NO:37	(401)	
SEQ ID NO:41	(401)	
		451 470
SEQ ID NO:33	(451)	
SEQ ID NO:37	(451)	
SEQ ID NO:41	(451)	

### Sequence identity

SEQ ID NO:33 v. SEQ ID NO:37: 99.6% SEQ ID NO:33 v. SEQ ID NO:41: 98.3% SEQ ID NO:37 v. SEQ ID NO:41: 98.7%

### Sequence identity between NA proteins of H3N2 and H3N8

SEQ ID NO:	8	10	12	33	37	41
8		100%	100%	43.5%	43.8%	43.7%
10			100%	43.5%	43.8%	43.7%
12				44.1%	44.4%	44.3%
33					99.6%	98.3%
37						98.7%
41						

### INACTIVATED CANINE INFLUENZA VACCINES AND METHODS OF MAKING AND USES THEREOF

# CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority to U.S. provisional application 62/185,266 filed on Jun. 26, 2015 and U.S. provisional application 62/298,285 filed on Feb. 22, 2016.

### FIELD OF THE INVENTION

The present invention relates to new canine influenza virus strains, and vaccines and compositions. The present invention also relates to reagents and methods allowing their detection, methods of vaccination as well as methods of producing these reagents and vaccines.

### BACKGROUND OF THE INVENTION

Influenza virus is a member of Orthomyxoviridae family (Murphy and Webster, Orthomyxoviruses, Fields Virology, Third Edition, vol. 1, pp. 1397-1445, 1996). There are three types of influenza viruses designated A, B, and C. The influenza virion contains a segmented negative-sense RNA genome. The influenza virion includes the following proteins: hemagglutinin (HA), neuraminidase (NA), matrix (M1), proton ion-channel protein (M2), nucleoprotein (NP), 30 polymerase basic protein 1 (PB1), polymerase basic protein 2 (PB2), polymerase acidic protein (PA), and nonstructural protein 2 (NS2) proteins. The NP and the matrix protein M1 are used to classify the influenza virus into group A, B or C.

The HA and NA proteins are envelope glycoproteins. The 35 HA protein is responsible for virus attachment and penetration of the viral particles into the cell and contains the major immunodominant epitopes for virus neutralization and protective immunity. Both HA and NA proteins are considered the most important components for prophylactic influenza 40 vaccines. To date, eighteen different HA subtypes and eleven different NA subtypes have been identified (Tong et al., 2013, PLoS Pathogens, Vol. 9 (10), New World Bats harbor diverse influenza A viruses).

Globally, influenza is the most economically significant 45 respiratory disease in humans, pigs, horses and poultry. Influenza virus is known for its continuous genetic and antigenic changes, which impede effective control of the virus. Of particular concern for prevention of epidemics and pandemics is the emergence of a new subtype of the virus by 50 genetic re-assortment or inter-species transmission.

Recently, influenza outbreaks have occurred in species, e.g., feline and canine, which historically do not carry influenza virus. During 2004 and 2005, outbreaks of respiratory disease in racing greyhounds caused by infection with 55 influenza virus occurred in Florida, eastern and western Iowa, and Texas. Molecular and antigenic analyses of three influenza viruses isolated from outbreaks of severe respiratory disease in racing greyhounds revealed that they are closely related to H3N8 equine influenza virus (Crawford et 60 al., Science, 2005, 310 (5747):482-485; PLOS Pathogens, 2014, 10 (10), e1004455).

U.S. Pat. No. 8,246,962 and Song et al. (2008 Emerg. Infect. Dis., 2008, 14, pp. 741-746) reported an infection of an H3N2 subtype influenza virus in a pet dog in the Republic 65 of Korea. The case was caused by a H3N2 avian-origin canine influenza virus (CIV), which infected dogs success-

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fully through nasal inoculation or contact (respiratory fluid exchange) under experimental conditions.

Li et. al. (Infection, Genetics and Evolution, 2010, 10 (8): 1286-1288) reported four sporadic cases of H3N2 canine influenza in Southern China, which were identified from sick dogs from May 2006 to October 2007. The evolutionary analysis showed that all eight segments of these four viruses are avian-origin and phylogenetically closely related to the H3N2 canine influenza viruses reported earlier in South Korea.

H3N2 canine influenza can be transmitted to cats and cause severe respiratory disease in cats (Song et al., J. Gen. Virol. 2011, 92:23050-2355).

On Mar. 26, 2015, the Chicago Tribune reported an outbreak of canine influenza caused by a new strain of virus identified as H3N2. As of today, the virus has now been detected in dogs in 30 states, including Illinois, Ohio, Wisconsin, Indiana, Iowa, Minnesota, Michigan, California, Massachusetts, Texas, New Jersey, South Dakota, and Georgia.

There were no vaccines available in the U.S. for H3N2 CIV as of June 2015. The H3N8 CIV vaccines currently marketed in the U.S. are unlikely to protect dogs against H3N2 CIV due to the genetic divergence in the HA proteins encoded by the two virus subtypes. Although both viruses encode H3 subtype HAs, the proteins only share about 77% sequence identity at the amino acid level.

Accordingly, there is an urgent need for an effective vaccine against H3N2 influenza infection in canines and felines.

### SUMMARY OF THE INVENTION

The present invention relates to an inactivated/killed canine influenza virus (CIV) composition or vaccine. In particular, the invention provides the canine influenza virus strains under the ATCC deposit Nos. PTA-122265 and PTA-122266, or any descendant or progeny of the strains. The canine influenza viruses are H3N2 strains isolated in the US.

In a particular embodiment, the inactivated vaccines comprise an adjuvant. The adjuvant may be any substance which increases and/or augments the elicited immune response, as compared to inactivated vaccine alone.

The invention also provides a polyvalent composition or vaccine that is effective to protect animals against a variety of CIV infections. In particular, the invention provides a polyvalent composition or vaccine comprising an inactivated H3N2 CIV and inactivated H3N8 CIV.

Methods for protection in animals against CIV infections and for reducing viral shedding in CIV infected animals are provided.

The invention also offers the possibility of diagnosing the presence of the canine influenza virus according to the invention in dogs. Its subject is therefore diagnostic tests and methods relating thereto using reagents which will be described below.

Kits comprising the inactivated canine Influenza strains and instructions for use are also provided.

These and other embodiments are disclosed or are obvious from and encompassed by, the following Detailed Description.

### BRIEF DESCRIPTION OF THE DRAWINGS

The following detailed description, given by way of example, and which is not intended to limit the invention to

specific embodiments described, may be understood in conjunction with the accompanying figures, incorporated herein by reference, in which:

FIGS. 1A-B are tables showing the SEQ ID NO assigned to each DNA and protein sequence.

FIG. 2 depicts real-time RT-PCR genotyping of CIVs.

FIG. 3 depicts the sensitivity of the subtyping assay.

FIG. 4 depicts the efficiency of the subtyping assay.

FIG. 5A-5P provide the sequence alignments of DNA and proteins.

#### DETAILED DESCRIPTION

It is noted that in this disclosure and particularly in the claims, terms such as "comprises", "comprised", "compris- 15 ing" and the like can have the meaning attributed to it in U.S. Patent law; e.g., they can mean "includes", "included", "including", and the like; and that terms such as "consisting essentially of" and "consists essentially of" have the meaning ascribed to them in U.S. Patent law, e.g., they allow for 20 elements not explicitly recited, but exclude elements that are found in the prior art or that affect a basic or novel characteristic of the invention.

The singular terms "a," "an," and "the" include plural referents unless context clearly indicates otherwise. Similarly, the word "or" is intended to include "and" unless the context clearly indicate otherwise. The word "or" means any one member of a particular list and also includes any combination of members of that list.

The term "animal" is used herein to include all mammals, 30 birds and fish. The animal as used herein may be selected from the group consisting of equine (e.g., horse), canine (e.g., dogs, wolves, foxes, coyotes, jackals), feline (e.g., lions, tigers, domestic cats, wild cats, other big cats, and other felines including cheetahs and lynx), bovine (e.g., 35 cattle, buffalos), swine (e.g., pig), ovine (e.g., sheep), caprine (e.g., goats), camelids (e.g., lamas), avian (e.g., chicken, duck, goose, turkey, quail, pheasant, parrot, finches, hawk, crow, ostrich, emu and cassowary), primate (e.g., prosimian, tarsier, monkey, gibbon, ape), humans, and fish. The term 40 "animal" also includes an individual animal in all stages of development, including embryonic and fetal stages.

The terms "polypeptide" and "protein" are used interchangeably herein to refer to a polymer of consecutive amino acid residues.

The term "nucleic acid", "nucleotide", and "polynucleotide" are used interchangeably and refer to RNA, DNA, cDNA, or cRNA and derivatives thereof, such as those containing modified backbones. It should be appreciated that the invention provides polynucleotides comprising 50 sequences complementary to those described herein. The "polynucleotide" contemplated in the present invention includes both the forward strand (5' to 3') and reverse complementary strand (3' to 5'). Polynucleotides according to the invention can be prepared in different ways (e.g. by 55 chemical synthesis, by gene cloning etc.) and can take various forms (e.g. linear or branched, single or double stranded, or a hybrid thereof, primers, probes etc.).

The term "genomic DNA" or "genome" is used interchangeably and refers to the heritable genetic information of 60 a host organism. The genomic DNA comprises the DNA of the nucleus (also referred to as chromosomal DNA) but also the DNA of the plastids (e.g., chloroplasts) and other cellular organelles (e.g., mitochondria). The genomic DNA or genome contemplated in the present invention also refers to 65 the RNA of a virus. The RNA may be a positive strand or a negative strand RNA. The term "genomic DNA" contem-

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plated in the present invention includes the genomic DNA containing sequences complementary to those described herein. The term "genomic DNA" also refers to messenger RNA (mRNA), complementary DNA (cDNA), and complementary RNA (cRNA).

The term "gene" is used broadly to refer to any segment of polynucleotide associated with a biological function. Thus, genes or polynucleotides include introns and exons as in genomic sequence, or just the coding sequences as in cDNAs, such as an open reading frame (ORF), starting from the start codon (methionine codon) and ending with a termination signal (stop codon). Genes and polynucleotides can also include regions that regulate their expression, such as transcription initiation, translation and transcription termination. Thus, also included are promoters and ribosome binding regions (in general these regulatory elements lie approximately between 60 and 250 nucleotides upstream of the start codon of the coding sequence or gene; transcription terminators (in general the terminator is located within approximately 50 nucleotides downstream of the stop codon of the coding sequence or gene). Gene or polynucleotide also refers to a nucleic acid fragment that expresses mRNA or functional RNA, or encodes a specific protein, and which includes regulatory sequences.

As used herein, the term "antigen" or "immunogen" means a substance that induces a specific immune response in a host animal. The antigen may comprise a whole organism, killed, attenuated or live; a subunit or portion of an organism; a recombinant vector containing an insert with immunogenic properties; a piece or fragment of DNA capable of inducing an immune response upon presentation to a host animal; a polypeptide, an antigen, an epitope, a hapten, or any combination thereof. Alternately, the immunogen or antigen may comprise a toxin or antitoxin.

The term "immunogenic protein or peptide" as used herein includes polypeptides that are immunologically active in the sense that once administered to the host, it is able to evoke an immune response of the humoral and/or cellular type directed against the protein. Preferably the protein fragment is such that it has substantially the same immunological activity as the total protein. Thus, a protein fragment according to the invention comprises or consists essentially of or consists of at least one epitope or antigenic determinant. An "immunogenic" protein or polypeptide, as used herein, includes the full-length sequence of the protein, analogs thereof, or immunogenic fragments thereof. By "immunogenic fragment" is meant a fragment of a protein which includes one or more epitopes and thus elicits the immunological response described above. Such fragments can be identified using any number of epitope mapping techniques, well known in the art. For example, linear epitopes may be determined by e.g., concurrently synthesizing large numbers of peptides on solid supports, the peptides corresponding to portions of the protein molecule, and reacting the peptides with antibodies while the peptides are still attached to the supports. Similarly, conformational epitopes are readily identified by determining spatial conformation of amino acids such as by, e.g., x-ray crystallography and 2-dimensional nuclear magnetic resonance.

The term "immunogenic protein or peptide" further contemplates deletions, additions and substitutions to the sequence, so long as the polypeptide functions to produce an immunological response as defined herein. The term "conservative variation" denotes the replacement of an amino acid residue by another biologically similar residue, or the replacement of a nucleotide in a nucleic acid sequence such that the encoded amino acid residue does not change or is

another biologically similar residue. In this regard, particularly preferred substitutions will generally be conservative in nature, i.e., those substitutions that take place within a family of amino acids. For example, amino acids are generally divided into four families: (1) acidic—aspartate and 5 glutamate; (2) basic—lysine, arginine, histidine; (3) nonpolar—alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan; and (4) uncharged polarglycine, asparagine, glutamine, cysteine, serine, threonine, tyrosine. Phenylalanine, tryptophan, and tyrosine are some- 10 times classified as aromatic amino acids. Examples of conservative variations include the substitution of one hydrophobic residue such as isoleucine, valine, leucine or methionine for another hydrophobic residue, or the substitution of one polar residue for another polar residue, such as 15 the substitution of arginine for lysine, glutamic acid for aspartic acid, or glutamine for asparagine, and the like; or a similar conservative replacement of an amino acid with a structurally related amino acid that will not have a major effect on the biological activity. Proteins having substan- 20 tially the same amino acid sequence as the reference molecule but possessing minor amino acid substitutions that do not substantially affect the immunogenicity of the protein are, therefore, within the definition of the reference polypeptide. All of the polypeptides produced by these modifi- 25 cations are included herein. The term "conservative variation" also includes the use of a substituted amino acid in place of an unsubstituted parent amino acid provided that antibodies raised to the substituted polypeptide also immunoreact with the unsubstituted polypeptide.

An "immunological response" to a composition or vaccine is the development in the host of a cellular and/or antibody-mediated immune response to a composition or vaccine of interest. Usually, an "immunological response" includes but is not limited to one or more of the following 35 effects: the production of antibodies, B cells, helper T cells, and/or cytotoxic T cells, directed specifically to an antigen or antigens included in the composition or vaccine of interest. Preferably, the host will display either a therapeutic or protective immunological response such that resistance to 40 new infection will be enhanced and/or the clinical severity of the disease reduced. Such protection will be demonstrated by either a reduction or lack of symptoms normally displayed by an infected host, a quicker recovery time and/or lowered pathogen loads in the infected host.

As used herein, the term "inactivated vaccine" means a vaccine composition containing an infectious organism or pathogen that is no longer capable of replication or growth. The pathogen may be bacterial, viral, protozoal or fungal in origin. Inactivation may be accomplished by a variety of 50 methods including freeze-thawing, chemical treatment (for example, treatment with thimerosal, formalin, BPL (beta-propiolactone) or BEI (binary ethylenimine), sonication, radiation, heat or any other conventional means sufficient to prevent replication or growth of the organism while main- 55 taining its immunogenicity.

The terms "polyvalent vaccine or composition", "combination or combo vaccine or composition" and "multivalent vaccine or composition" are used interchangeably to refer to a composition or vaccine containing more than one composition or vaccines. The polyvalent vaccine or composition may contain two, three, four or more compositions or vaccines. The polyvalent vaccine or composition may comprise recombinant viral vectors, active or attenuated or inactivated/killed wild-type viruses, or a mixture of recombinant viral vectors and wild-type viruses in active or attenuated or killed forms.

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In one embodiment, the present invention encompasses a novel inactivated/killed CIV composition or vaccine. The CIV strains are the H3N2 subtype newly identified in the USA. The inactivation may be the chemical inactivation that produces enumerable structural changes, including for example, formation of new chemical bonds via chemical crosslinking, irreversible chemical alteration of the nucleic acid and protein coat.

In another embodiment, the present invention provides a bivalent composition or vaccine comprising an inactivated H3N2 CIV and inactivated H3N8 CIV.

One embodiment of the invention provides the genomic DNA and gene sequences, and encoded protein sequences of CIV H3N2 and H3N8 strains.

In another embodiment, the invention provides the sequences for HA proteins or antigens. In one aspect of the embodiment, the HA proteins have the polypeptide sequence as set forth in SEQ ID NO:2, 4, 6, 31, 35, and 39. In another aspect, the HA proteins have at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, 99.1%. 99.2%, 99.3%, 99.4%, 99.5%, 99.6%, 99.7%, 99.8% or 99.9% sequence identity to a polypeptide having the sequence as set forth in SEQ ID NO:2, 4, 6, 31, 35, and 39. In yet another aspect, the HA proteins are encoded by the polynucleotides having at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, 99.1%. 99.2%, 99.3%, 99.4%, 99.5%, 99.6%, 99.7%, 99.8% or 99.9% sequence identity to a polynucleotide having the sequence as set forth in SEQ ID NO:1, 3, 5, 30, 34 and 38.

In another embodiment, the invention provides the sequences for NA proteins or antigens. In one aspect of the embodiment, the NA proteins have the polypeptide sequence as set forth in SEQ ID NO:8, 10, 12, 33, 37 and 41. In another aspect, the NA proteins have at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, 99.1%. 99.2%, 99.3%, 99.4%, 99.5%, 99.6%, 99.7%, 99.8% or 99.9% sequence identity to a polypeptide having the sequence as set forth in SEQ ID NO:8, 10, 12, 33, 37 and 41. In yet another aspect, the NA proteins are encoded by the polynucleotides having at least 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, 99.1%. 99.2%, 99.3%, 99.4%, 99.5%, 96%, 97%, 98%, 99%, 99.1%. 99.2%, 99.3%, 99.4%, 99.5%, 99.6%, 99.7%, 99.8% or 99.9% sequence identity to a polynucleotide having the sequence as set forth in SEQ ID NO:7, 9, 11, 32, 36, 40 and 42.

In one embodiment, the CIV stains may comprise an HA gene encoding an HA protein aforementioned. In another embodiment, the CIV strains may comprise an NA gene encoding an NA protein aforementioned.

In one embodiment, the invention provides CIV strains under the ATCC deposit numbers PTA-122265 and PTA-122266, or any parent, descendant or progeny of the deposited strains.

In one embodiment, the composition or vaccine of the invention includes a live CIV H3N2 strain. The CIV H3N2 strains were initially isolated from 1-14 year old dogs with symptoms of severe respiratory disorder in Illinois, USA. The strains were deposited at ATCC (American Type Culture Collection; 10801 University Boulevard, Manassas, VA 20110) on Jun. 25, 2015 and were accorded Accession Nos. PTA-122265 and PTA-122266.

In another embodiment, the present invention contemplates preparation and isolation of a progeny or descendant of a CIV H3N2 virus that has been deposited on Jun. 25, 2015 at ATCC under the Accession Numbers PTA-122265 and PTA-122266. The invention therefore extends to CIV H3N2 virus strains which are derived from the deposited strains through propagation or multiplication in an identical

or divergent form, in particular descendants which possess the essential characteristics of the deposited strains. Upon continued propagation, the strains may acquire mutations most of which will not alter the properties of these strains significantly. The progeny or descendant may comprise a polynucleotide encoding an HA protein having at least 99.6% sequence identity to the sequence as set forth in SEO ID NO:2, 4, or 6, or a polynucleotide encoding an NA protein having the sequence as set forth in SEQ ID NO:8, 10 or 12. The progeny or descendant may comprise a polynucleotide encoding an HA protein having at least 99.0% or 99.6% sequence identity to the sequence as set forth in SEQ ID NO:2, 4, or 6 and the HA protein may differ at amino acid position 62, 219 or 249 of a full-length HA0 protein (i.e. 15 uncleaved and containing the signal peptide). The amino acid at position 62, 219, or 249 of a full-length HA0 protein may be any amino acid denoted in Table 1 below.

TABLE 1

Amino Acid				
Abbreviation	Amino Acid	Abbreviation	Amino Acid	
A (Ala)	Alanine	L (Leu)	Leucine	
R (Arg)	Arginine	K (Lys)	Lysine	
D (Asp)	Aspartic acid	M (Met)	Methionine	
N (Asn)	Asparagine	F (Phe)	Phenylalanine	
C (Cys)	Cysteine	P (Pro)	Proline	
Q (Gln)	Glutamine	S (Ser)	Serine	
E (Glu)	Glutamic acid	T (Thr)	Threonine	
G (Gly)	Glycine	W (Trp)	Tryptophan	
H (His)	Histidine	Y (Tyr)	Tyrosine	
I (Île)	Isoleucine	V (Val)	Valine	

The inactivated pathogen or organism can be concentrated by conventional concentration techniques, in particular by ultrafiltration, and/or purified by conventional purification means, in particular using chromatography techniques including, but not limited to, gel-filtration or by ultrafiltration. As used herein, the term "immunogenicity" means capable of producing an immune response in a host animal against an antigen or antigens. This immune response forms the basis of the protective immunity elicited by a vaccine against a specific infectious organism.

Further, methods which are well known to those skilled in the art can be used to determine protein purity or homogeneity, such as polyacrylamide gel electrophoresis of a sample, followed by visualizing a single polypeptide band on a staining gel. Higher resolution may be determined using HPLC or other similar methods well known in the art.

In one embodiment, the invention provides for the administration of a therapeutically effective amount of a vaccine or composition for the delivery and expression of a CIV H3N2 antigen in a target cell. In another embodiment, the invention provides for the administration of a therapeutically effective 55 amount of a vaccine or composition for the delivery and expression of CIV H3N2 and CIV H3N8 antigens in a target cell. Determination of the therapeutically effective amount is routine experimentation for one of ordinary skill in the art. In one embodiment, the vaccine or composition comprises 60 an inactivated/killed CIV H3N2 strains, and a pharmaceutically or veterinarily acceptable carrier, adjuvant, vehicle or excipient. In another embodiment, the vaccine or composition comprises an inactivated/killed H3N2 CIV and inactivated/killed H3N8 CIV strains, and a pharmaceutically or 65 veterinarily acceptable carrier, adjuvant, vehicle or excipient. In an embodiment, the pharmaceutically or veterinarily

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acceptable carrier, vehicle, adjuvant, or excipient facilitates transfection and/or improves preservation of the virus or protein.

The pharmaceutically or veterinarily acceptable carriers, adjuvants, vehicles, or excipients are well known to the one skilled in the art. For example, a pharmaceutically or veterinarily acceptable carrier or vehicle or excipient can be a 0.9% NaCl (e.g., saline) solution or a phosphate buffer. Other pharmaceutically or veterinarily acceptable carrier, adjuvant, vehicle, or excipients that can be used for methods of this invention include, but are not limited to, poly-(Lglutamate) or polyvinylpyrrolidone. The pharmaceutically or veterinarily acceptable carrier, adjuvant, vehicle, or excipients may be any compound or combination of compounds facilitating the administration of the vector (or protein expressed from an inventive vector in vitro); the carrier, vehicle, adjuvant, or excipient may facilitate transfection and/or improve preservation of the vector (or protein). Doses and dose volumes are herein discussed in the general description and can also be determined by the skilled artisan from this disclosure read in conjunction with the knowledge in the art, without any undue experimentation.

The cationic lipids containing a quaternary ammonium salt which are advantageously but not exclusively suitable for plasmids, are advantageously those having the following formula:

$$\begin{array}{c} \text{CH}_{3} \\ \text{R}_{1} \text{---} \text{O} \text{---} \text{CH}_{2} \text{---} \text{CH} \text{---} \text{CH}_{2} \text{---} \overset{\text{CH}_{3}}{\text{---}} \text{R}_{2} \text{---} \text{X} \\ \text{OR}_{1} \\ \text{CH}_{3} \end{array}$$

35 in which  $R_1$  is a saturated or unsaturated straight-chain aliphatic radical having 12 to 18 carbon atoms,  $R_2$  is another aliphatic radical containing 2 or 3 carbon atoms and X is an amine or hydroxyl group, e.g. the DMRIE. In another embodiment the cationic lipid can be associated with a 40 neutral lipid, e.g. the DOPE.

Among these cationic lipids, preference is given to DMRIE (N-(2-hydroxyethyl)-N,N-dimethyl-2,3-bis(tetradecyloxy)-1-propane ammonium; WO96/34109), advantageously associated with a neutral lipid, advantageously DOPE (dioleoyl-phosphatidyl-ethanol amine; Behr J. P., 1994, Bioconjugate Chemistry, 5, 382-389), to form DMRIE-DOPE.

The composition or vaccine mixture with the adjuvant is formed extemporaneously and contemporaneously with administration of the preparation or shortly before administration of the preparation; for instance, shortly before or prior to administration, the composition or vaccine-adjuvant mixture is formed, so as to give enough time prior to administration for the mixture to form a complex, e.g. between about 10 and about 60 minutes prior to administration, such as approximately 30 minutes prior to administration.

When DOPE is present, the DMRIE:DOPE molar ratio is advantageously about 95:about 5 to about 5:about 95, more advantageously about 1:about 1, e.g., 1:1.

The immunogenic compositions and vaccines according to the invention may comprise or consist essentially of one or more adjuvants. Suitable adjuvants for use in the practice of the present invention are (1) polymers of acrylic or methacrylic acid, maleic anhydride and alkenyl derivative polymers, (2) immunostimulating sequences (ISS), such as oligodeoxyribonucleotide sequences having one ore more

non-methylated CpG units (Klinman D. M. et al., Proc. Natl. Acad. Sci., USA, 1996, 93, 2879-2883; WO98/16247), (3) an oil in water emulsion, such as the SPT emulsion described on p 147 of "Vaccine Design, The Subunit and Adjuvant Approach" published by M. Powell, M. Newman, Plenum 5 Press 1995, and the emulsion MF59 described on p 183 of the same work, (4) cation lipids containing a quaternary ammonium salt, (5) cytokines, (6) aluminum hydroxide or aluminum phosphate or (7) saponin, (8) Dimethyldioctade-cyl ammonium bromide (Vaccine Design p. 157), (9) Aridine (Vaccine Design p. 148) other adjuvants discussed in any document cited and incorporated by reference into the instant application, or (8) any combinations or mixtures thereof.

The oil in water emulsion (3), can be based on: light liquid paraffin oil (European pharmacopoeia type), isoprenoid oil such as squalane, squalene, oil resulting from the oligomerization of alkenes, e.g. isobutene or decene, esters of acids or alcohols having a straight-chain alkyl group, such as vegetable oils, ethyl oleate, propylene glycol, di(caprylate/20 caprate), glycerol tri(caprylate/caprate) and propylene glycol dioleate, or esters of branched, fatty alcohols or acids, especially isostearic acid esters.

The oil is used in combination with emulsifiers to form an emulsion. The emulsifiers may be nonionic surfactants, such 25 as: esters of on the one hand sorbitan, mannide (e.g. anhydromannitol oleate), glycerol, polyglycerol or propylene glycol and on the other hand oleic, isostearic, ricinoleic or hydroxystearic acids, said esters being optionally ethoxylated, or polyoxypropylene-polyoxyethylene copolymer 30 blocks, such as Pluronic, e.g., L121. Some of the emulsions, such as TS6, TS7, TS8 and TS9 emulsions, are described in U.S. Pat. Nos. 7,608,279 and 7,371,395.

The polymers of acrylic or methacrylic acid (1) are preferably crosslinked, in particular with polyalkenyl ethers 35 of sugars or polyalcohols. These compounds are known under the term carbomer (Pharmeuropa vol. 8, No. 2, June 1996). Persons skilled in the art can also refer to U.S. Pat. No. 2,909,462 describing such acrylic polymers crosslinked with a polyhydroxylated compound having at least 3 40 hydroxyl groups, preferably not more than 8, the hydrogen atoms of at least three hydroxyls being replaced with unsaturated aliphatic radicals having at least 2 carbon atoms. The preferred radicals are those containing 2 to 4 carbon atoms, e.g. vinyls, allyls and other ethylenically unsaturated groups. 45 The unsaturated radicals may themselves contain other substituents, such as methyl. The products sold under the name CARBOPOL™ (BF Goodrich, Ohio, USA) are particularly appropriate. They are crosslinked with an allyl sucrose or with allylpentaerythritol. Among them, there may 50 be mentioned CARBOPOLTM 974P, 934P and 971 P.

Among the copolymers of maleic anhydride and of alkenyl derivative, the EMA<sup>TM</sup> copolymers (Monsanto) which are copolymers of maleic anhydride and of ethylene, which are linear or crosslinked, for example crosslinked with 55 divinyl ether, are preferred.

The proportions of adjuvant which are useful are well known and readily available to the one skilled in the art. By way of example, the concentration of polymers of acrylic or methacrylic acid or of anhydride maleic and alkenyl copolymers in the final vaccine composition will be from 0.01% to 1.5% W/V, more particularly from 0.05 to 1% W/V, preferably from 0.1 to 0.4% W/V.

In one embodiment, the adjuvant may include TS6 (U.S. Pat. No. 7,371,395), LR2, LR3 and LR4 (U.S. Pat. No. 65 7,691,368), TSAP (US20110129494), TRIGEN<sup>TM</sup> (Newport Labs), synthetic dsRNAs (e.g. poly-IC, poly-ICLC [HILTO-

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NOL®]), and MONTANIDETM adjuvants (W/O, W/O/W, O/W, IMS and Gel; all produced by SEPPIC).

In yet another embodiment, the adjuvant may include interleukin-2 (IL-2), IL-12, interferon  $\alpha$  (IFN $\alpha$ ), polyinosinic and polycytidylic acid, and cytidine-phosphate-guanosine oligodeoxynucleotides (CpG ODN), which are known to significantly enhance CMI response to CIV vaccines (Vet. Immuno. and Immunopath. Vol. 129, Issues 1-2, 15 May 2009, Pages 1-13).

Optionally the vaccine used according to the method of the invention may contain a cytokine. The cytokine may be present as a protein or as a gene encoding this cytokine inserted into a recombinant viral vector. The cytokines may be selected among the pig cytokines.

In a specific embodiment, the pharmaceutical composition is directly administered in vivo. Advantageously, the pharmaceutical and/or therapeutic compositions and/or formulations according to the invention comprise or consist essentially of or consist of an effective quantity to elicit a therapeutic response of one or more expression vectors and/or polypeptides as discussed herein; and, an effective quantity can be determined from this disclosure, including the documents incorporated herein, and the knowledge in the art, without undue experimentation.

The composition or vaccine may contain a dose from about 10<sup>2</sup> to about 10<sup>20</sup>, about 10<sup>3</sup> to about 10<sup>18</sup>, about 10<sup>4</sup> to about 10<sup>16</sup>, about 10<sup>5</sup> to about 10<sup>12</sup> VLPs (virus like particles) produced in vitro or in vivo from a viral vector, a plasmid, or baculovirus. The viral vector may be titrated based on any virus titration methods including, but not limited to, FFA (Focus Forming Assay) or FFU (Focus Forming Unit), TCID<sub>50</sub> (50% Tissue Culture Infective Dose), PFU (Plaque Forming Units), and FAID<sub>50</sub> (50% Fluorescent Antibody Infectious Dose), and the VLPs produced in vitro can be titrated by hemagglutination assay, ELISA, and electron microscopy. Other methods may also be applicable depending on the type of VLP.

The composition or vaccine may contain from about  $10^{2.0}$  to about  $10^{10.0}$  TCID<sub>50</sub>/dose, from about  $10^{4.5}$  to about  $10^{8.0}$  TCID<sub>50</sub>/dose and from about  $10^{5.5}$  to about  $10^{6.5}$  TCID<sub>50</sub>/dose. The composition or vaccine may contain equivalent TCID<sub>50</sub> in the case of inactivated/killed composition or vaccine.

The dose volumes can be between about 0.1 and about 10 ml, between about 0.2 and about 5 ml.

When the antigen relates to hemagglutinin, such as inactivated influenza vaccines, the dosage is measured in hemagglutination units (HAUs). In an embodiment, the dosage may be from about 655 to about 65,500 HAU/dose.

It should be understood by one of skill in the art that the disclosure herein is provided by way of example and the present invention is not limited thereto. From the disclosure herein and the knowledge in the art, the skilled artisan can determine the number of administrations, the administration route, and the doses to be used for each injection protocol, without any undue experimentation.

The present invention contemplates at least one administration to an animal of an efficient amount of the therapeutic composition made according to the invention. The animal may be male, female, pregnant female and newborn. This administration may be via various routes including, but not limited to, intramuscular (IM), intradermal (ID) or subcutaneous (SC) injection or via intranasal or oral administration. The therapeutic composition according to the invention can also be administered by a needleless apparatus (as, for example with a Pigjet, Biojector or Vitaj et apparatus (Bioject, Oreg., USA)).

The composition or vaccine is administered to a dog or a cat (about six to eight-week old). A booster administration can be done if necessary around 2 to 8 weeks after the first administration.

Liquid jet needle-free injectors are devices performing injections of a certain amount of liquid under high pressure through a minute orifice. In an embodiment, the needle-free injection is a DERMA-VAC NF transdermal vaccinator system.

The volume of dose injected may be from about 0.1 ml to about 4.0 ml, about 0.1 ml to about 2 ml, about 0.1 ml to about 1 ml, from about 0.2 ml to about 0.8 ml, about 0.2 ml to about 0.5 ml. By definition, the volume of one dose means the total volume of vaccine administered at once to one animal.

Optionally, the administration can be repeated, as booster administration, at suitable intervals if necessary or desirable, e.g. from about 2 to about 8 weeks after the first administration, and preferably from about 2 to about 4 weeks after 20 the first administration. A booster administration can also be repeated every 6 months or every year.

Another object of the invention is the use of an efficient amount of the composition or vaccine as described above and of an acceptable vehicle or diluent, for the preparation 25 of a vaccine designed to be administered to an animal using a liquid jet needle-free injector as described above, and resulting in eliciting a safe and protective immune response against influenza infection.

In one embodiment of the invention, a prime-boost regimen can be employed, which is comprised of at least one primary administration and at least one booster administration using at least one common polypeptide, antigen, epitope or immunogen. Typically the same composition or vaccine is used as the primary administration and the boost. This administration protocol is called "prime-boost". However, different compositions or vaccines may be used as the prime administration and the boost.

Another object is a vaccination kit or set, comprising at least one vaccine vial containing the vaccine of the present 40 invention, and operatively assembled to perform the administration of the vaccine to an animal of the canine family. Such vaccination kit or set is able to elicit a safe and protective immune response against influenza infection.

The invention also offers the possibility of diagnosing the 45 presence of the CIV H3N2 or H3N8 strains according to the invention in dogs and cats. Its subject is therefore diagnostic tests and methods relating thereto using reagents which are described below.

A first reagent relates to the DNA sequences disclosed 50 here and their fragments, which may be used as probes or primers in well-known hybridization or PCR (Polymerase Chain Reaction) techniques.

A second reagent relates to the polypeptides encoded by these sequences from the virus, or synthesized by the 55 chemical route according to conventional techniques for peptide synthesis.

A third and fourth reagent relate to polyclonal and monoclonal antibodies which may be produced according to the customary techniques from the virus, the polypeptides or 60 fragments, extracted or encoded by the DNA sequences.

These second, third and fourth reagents may be used in a diagnostic method, a subject of the invention, in which a test is carried out, on a sample of physiological fluid (blood, plasma, serum and the like) or a sample of tissue (ganglia, 65 liver, lungs, kidneys and the like) obtained from a dog or a cat to be tested, for the presence of an antigen specific for a

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CIV H3N2 or H3N8 according to the invention, by seeking to detect either the antigen itself, or antibodies directed against this antigen.

The antigens and antibodies according to the invention may be used in any known laboratory diagnostic technique. However, it will be preferable to use them in techniques which can be used directly in the field by the veterinary doctor, the breeder or the owner of the animal. Persons skilled in the art have available a range of laboratory and field techniques and are therefore in the perfect position to adapt the use of this antigen and/or antibodies as diagnostic reagent(s).

The diagnostic techniques which will be used within the framework of the present invention are PCR and RT-PCR.

The subject of the invention is also a diagnostic kit comprising the reagents and/or polyclonal or monoclonal antibodies specific for H3N2 or H3N8 CIV antigen. The diagnostic kit may comprise primers designed based on the DNA sequences disclosed herein and reagents for PCR assay.

The method of diagnosing the presence of a CIV H3N2 or H3N8 strain using PCR comprises the steps of: a) generating a cDNA of the influenza virus genome isolated from an animal; b) exposing the cDNA to a primer pair comprising a forward primer and a reverse primer in a real-time polymerase chain reaction (PCR) to yield an amplicon, wherein the primer pair is specific to the H3N2 or H3N8 CIV genome; c) performing sequencing on the amplicon; and d) analyzing and comparing the sequence of the amplicon with known H3N2 or H3N8 CIV sequences.

The method of diagnosing the presence of an H3N2 or H3N8 CIV strain using real-time RT-PCR comprises the steps of: a) extracting genomic RNA from the influenza virus isolated from an animal; b) exposing the RNA to a primer pair comprising a forward and a reverse primer in a real-time reverse transcription-polymerase chain reaction (RT-PCR), wherein the primer pair is specific to the H3N2 or H3N8 CIV genome; c) analyzing and comparing the RT-PCR curve thereby characterizing the CIV strain.

The forward primer and reverse primer may be designed based on the HA and NA polynucleotide sequences disclosed herein. The primers may be the primers having the sequence as defined in SEQ ID NO:13-29 (FIG. 1 and Table 3)

The diagnostic techniques which will be used within the framework of the present invention include Western blotting, immunofluoroescence, ELISA and immunochromatography.

Accordingly, it is preferably sought to detect specific antibodies in the sample by an indirect test, by competition or by displacement. To do this, the antigen itself is used as diagnostic reagent, or a fragment of this antigen, conserving recognition of the antibodies. The labelling may be advantageously a labelling with peroxidase or a special labelling, preferably with colloidal gold.

It may also be desired to detect the antigen itself in the sample with the aid of a labelled antibody specific for this antigen. The labelling is advantageously as described above.

By antibody specific for the antigen which can be used in particular in competition or displacement or for the detection of the antigen itself, there is understood monoclonal or polyclonal antibodies specific for the antigen, fragments of these antibodies.

Another feature of the invention is the production of polyclonal or monoclonal antibodies specific for the antigen in accordance with the invention, it being possible for these antibodies to then be used in particular as diagnostic reagent

for the detection of the antigen in a sample of physiological fluid or in a tissue sample, or even for the detection of antibodies present in such a sample or specimen. The invention also includes the immunologically functional fragments of these antibodies.

Antibodies can be prepared by the customary techniques. Reference may be made in particular to Antibodies, A Laboratory Manual, 2014, Cold Spring Harbor Laboratory, USA

The subject of the invention is also a preparation, pure or  $^{10}$  partially pure, or even crude, of monoclonal or polyclonal antibodies specific for the antigen, especially mouse or rabbit antibodies.

The invention will now be further described by way of the following non-limiting examples.

### **EXAMPLES**

Construction of DNA inserts, plasmids and recombinant viral vectors was carried out using the standard molecular <sup>20</sup> biology techniques described by J. Sambrook et al. (Molecular Cloning: A Laboratory Manual, 4th edition, Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 2014).

Example 1 Culture, Isolation and Genotyping of Canine Influenza H3N2 Strains

#### Virus Isolation

Nasal swabs were obtained from pet dogs that exhibited symptoms of acute respiratory disease at veterinary clinics 30 in Chicago, Ill. Swabs were transferred to vials containing transport Minimal essential medium (MEM) supplemented with 2% fetal bovine serum (HyClone) and 1× Antibiotic-Antimycotic (Gibco). Vials were placed at 4° C. and shipped cold by overnight courier within 4 days.

Infectious virus was isolated from clinical samples as follows. Samples were vortexed, and swabs were pressed against the vial walls to remove absorbed medium. The medium in each vial was filtered through a 0.4 µM filter, and 100 μL of filtered medium was inoculated into Madin-Darby 40 Canine Kidney (MDCK) cells (ATCC) or 10 day old specific pathogen free (SPF) embryonated chicken eggs (Merial, Inc., Gainesville). Cell cultures were incubated at 37° C., and cell supernatants were harvested when a majority of cells exhibited virus-induced cytopathic effect (CPE). Inocu- 45 lated eggs were incubated at 37° C. for 72 hours and subsequently chilled at 4° C. for at least 6 hours. To identify eggs that contained influenza virus, allantoic fluid from each inoculated egg was subjected to hemagglutination (HA) assay using 0.5% chicken red blood cells (RBCs) (Merial, 50 Inc., Gainesville). Eggs containing >2 HA units per 50 μL of allantoic fluid were considered HA-positive, and the full volume of allantoic fluid in such eggs was removed, aliquoted, and frozen at -70° C. Five of twenty-four samples inoculated into eggs showed positive HA titers with values 55 ranging from 32-256 HA units/50 μL. Two of the five samples (virus isolates ID #'s 4 and 8) were chosen for further vaccine development purposes and deposited at ATCC on Jun. 25, 2015 under accession number PTA-122265 (virus ID #4) and PTA-122266 (virus ID #8). Viral RNA detection and virus subtyping

A real-time RT-PCR assay was developed to test for the presence and subtype of influenza virus in egg allantoic fluid. Primers pairs that specifically anneal to the N2 or N8 neuraminidase (NA) segment genomic RNA encoded by 65 H3N2 and H3N8 subtype canine influenza viruses (CIV) were designed based on sequence alignments of NA gene

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sequences from CIVs available in GenBank. Forward primer SEQ ID NO:13 and reverse primer SEQ ID NO:14 were specific for the N2 NA gene of H3N2 CIV. Forward primer SEQ ID NO:15 and reverse primer SEQ ID NO:16 were specific for the N8 NA gene of H3N8 CIV.

RNA extracted from 140 μL of egg allantoic fluid (QIAamp Viral RNA Mini Kit; Qiagen) was tested for the presence of N2 and N8 genomic RNA using a QuantiTect SYBR Green RT-PCR Kit (Qiagen). Assay results (see FIG. 2 and Table 2) showed that the N2-specific primers produced amplification products with cycle threshold (C<sub>t</sub>) values of 9.1-35, while the N8 specific primer pairs produced products with C<sub>t</sub> values >39 (the limit of detection). Conversely, RNA from eggs inoculated with a H3N8 subtype CIV (A/canine/Wyoming/86033/2007) yielded a product with a C<sub>t</sub> value of 15.4 when tested with N8-specific primers, but N2-specific primers did not yield detectable products. Thus, the virus isolated from the nasal swabs contained N2 but not N8 NA genomic RNA, suggesting that the isolated virus was H3N2 CIV.

TABLE 2

	Virus	Primer	
RNA source	subtype	target	$C_t$
None	N/A	N2 NA	ND
None	N/A	N8 NA	ND
A/Ca/WY/86033/07	H3N8	N2 NA	ND
A/Ca/WY/86033/07	H3N8	N8 NA	15.4
#4	H3N2	N2 NA	9.1
#4	H3N2	N8 NA	39.1
#8	H3N2	N2 NA	9.5
#8	H3N2	N8 NA	ND

\*ND-not detected

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FIGS. 3 and 4 show the sensitivity and efficiency of the subtyping assay. The assay is highly sensitive for H3N2 CIV as it is capable of detecting <0.01 EID50 units of virus (i.e. it can detect less than one infectious unit of virus). The assay is also highly efficient. FIG. 4 shows that the assay is 98.81% efficient on a scale from 1-100%. The amplification factor of the assay is 1.99, close to the perfect score of 2.0 (amplification factors are calculated in such a way that a value of 2.0 is considered perfect).

Example 2 Virus Propagation and Genomic RNA

Virus propagation in eggs

The 50% egg infectious dose (EID<sub>50</sub>) assay was used to determine the titer of infectious virus present in the virus isolates. Virus samples were diluted in 10-fold increments from 10<sup>-1</sup> to 10<sup>-9</sup>, and five eggs per dilution were inoculated with 100 µL per egg of the 10<sup>-4</sup> to 10<sup>-9</sup> dilutions. Eggs were sealed and incubated for 72 hours at 37° C. Eggs were then chilled at 4° C. for at least 6 hours, and allantoic fluid from each egg was tested for the presence of virus by HA assay. The HA titers ranged from 64-256 per 50 µL of allantoic fluid for viruses #4 and #8, and the EID<sub>50</sub>/mL titer was 8.3 (#4) and 8.1 (#8) as calculated using the Spearman-Karber formula.

Large-scale virus stocks were prepared by inoculating each of sixty eggs with  $100\,\mu L$  of a  $10^{-5}$  virus dilution. Eggs were incubated at  $37^{\circ}$  C. for 72 hours and chilled as before. Allantoic fluid was collected from each egg and pooled. Virus HA and EID<sub>50</sub> titers in the pooled fluids were 16 and 7.5 (#4) or 8 and 7.3 (#8). Virus stocks were aliquoted and stored in liquid nitrogen.

Virus Propagation in Madin-Darby Canine Kidney (MDCK)

MDCK cells (ATCC) approved for vaccine derivation purposes by the USDA were seeded such that cell monolayers were ~90% confluent at the time of infection. Prior to infection, cells were washed twice with MEM lacking serum 16

Consensus sequences for each gene were generated from the assemblies in Sequencher 5.1 and designated SEQ ID NOs: 1, 3, 5, 7, 9 and 11 as shown in FIGS. 1 and 5. The corresponding HA and NA protein sequences are designated SEQ ID NOs:2, 4, 6, 8, 10 and 12 as shown in FIGS. 1 and

TABLE 3

	primers for seque	ncing HA and NA genes
SEQ ID NO:		
	HA sequer	ncing primers
17	Ca.H3N2.HA.390R	GAACTCCAATGTGCCTGATG
18	Ca.H3N2.HA.259F	CTGCACACTAATAGATGCCCTA
19	Ca.H3N2.HA.743F	CCAATCTGGCAGAATAAGCG
20	Ca.H3N2.HA.1276F	GAAGGGAGGATTCAAGACCTT
21	Ca.H3N2.HA.1569F	GGTTCCAGATCAAAGGTGTT
22	Ca.H3N2.HA.871R	CATTCGGAAATGCAGGTGTCA
23	Ca.H3N2.HA.1408R	TCAGCATTTTCCCTCAATTGC
	NA gene seq	uencing primers
24	Ca.H3N2.NA.429F	GGG ACC ACG CTG AAC AAT AA
25	Ca.H3N2.NA.484R	TGA AAC GGA ACA CCC AAC TC
26	Ca.H3N2.NA.975F	TCA GGA CTT GTT GGC GAT AC
27	Ca.H3N2.NA.1023R	TCC TGG ATT CCC TCT CTC ATT A
28	Ca.H3N2.1229F	ACT GGT CTG GTT ATT CTG GTA TTT
29	Ca.H3N2.1279R	TCT TGT GGC CCT CCT CTT AT

and inoculated with virus using a multiplicity of infection 30 Analysis of the Sequences (M.O.I.) of 0.001 in a low inoculum volume. Samples were incubated at 37° C. for 1 hour with intermittent rocking, and media containing 1 ug/mL of TPCK-treated trypsin (Sigma-Aldrich) was added. Cultures were incubated at 37° C. and monitored for the appearance of virus-induced cytopathic 35 effect (CPE). Under these conditions, the virus caused visible cell rounding and dissociation of cells from the substrate after 24-hours. Cell supernatants were harvested when approximately 90% of the cells had dissociated from the substrate. Supernatants were clarified by centrifugation 40 at 1,000×g, and virus titers in the clarified supernatants were measured by HA and 50% tissue culture infectious dose (TCID<sub>50</sub>) assays. After a single passage in MDCK cells, virus #4 achieved titers of 64 HA units/50 µL and 7.8 TCID<sub>50</sub> units/mL, and virus #8 replicated to titers of 64-128 45 HA units/50 µL and 8.6 TCID<sub>50</sub> units/mL. These cell stocks were subsequently used to produce pre-master seed stock material (virus #4, 64-128 HA units/50 μL and 8.4 TCID<sub>50</sub> units/mL) and purified antigen for raising virus-specific antisera in rabbits (virus #8, 128 HA units/50 µL and 8.7 50 TCID<sub>50</sub> units/mL).

#### Viral genomic RNA sequencing

Genomic RNA was extracted from virus propagated in eggs and MDCK cells. The full-length HA and NA genes were reverse transcribed and amplified by PCR using a 55 One-Step RT-PCR Kit (Qiagen) and universal primers that anneal to the termini of each gene segment (Hoffman et al., 2001 Arch Virol Universal primer set for the full-length amplification of all influenza A viruses). The resulting RT-PCR products were separated by electrophoresis in a 60 0.8% agarose gel and purified using a QIAquick Gel Extraction Kit (Qiagen). Purified cDNAs were sequenced by conventional Sanger sequencing methods using HA and NA gene specific primers as shown in Table 3 below. Contiguous sequence assemblies with 2-fold or greater coverage were 65 generated by aligning overlapping individual sequences using Sequencher 5.1 software (Gene Codes Corporation).

The three HA protein sequences obtained from the CIV H3N2 strains isolated in the US share 99.6% sequence identity (FIG. 5). These HA proteins vary in the amino acid composition at positions 62, 219 and 249 of the immature (full-length) HA0 protein.

The three NA protein sequences obtained from the CIV H3N2 strains isolated in the US are identical, though the polynucleotides encoding the NA proteins differ slightly with 99.9% sequence identity (FIG. 5). The database search didn't detect any CIV H3N2 NA proteins having greater than 98% sequence identity to the NA protein sequences generated from the CIV H3N2 strains isolated in the US.

Example 3 Production of Virus-Specific Antibodies

### Virus purification

MDCK cells in 225 cm<sup>2</sup> flasks were washed twice with 20 mL MEM per flask and inoculated with cell passage 1 of virus #8 at an MOI of 0.001 in a low inoculum volume. Cultures were incubated at 37° C. for 1 hour with intermittent rocking. Medium containing 1 ug/mL of TPCK-treated trypsin was added to each flask, and the flasks were incubated at 37° C. for 48 hours. Cell supernatants were harvested, and 540 mL of virus culture was clarified by centrifugation at 5,000×g for 10 min. using a Beckman JA-10 rotor. To pellet the virus, the clarified supernatant was centrifuged at 90,000×g for 2 hours at 4° C. using a SW32 Ti rotor. The pellet supernatant was aspirated, and 0.5 mL PBS was added to each virus pellet. Following overnight incubation on ice, the pellets were resuspended by gentle pipetting and pooled. The resuspended pellet material was loaded onto a sucrose step gradient containing equal volumes of 20%, 30%, 40%, and 55% w/v sucrose in PBS, and the gradient was centrifuged overnight at 100,000×g in a SW32Ti rotor at 4° C. The virus band was removed from the gradient and diluted in 5 volumes of PBS. Virus in the suspension was pelleted by centrifugation at 100,000×g in a

SW32Ti rotor for 2 hours at 4° C. The pellet supernatant was aspirated, and the virus pellet was incubated in 1 mL of PBS at 4° C. for 48 hours. The pellet was then resuspended by gentle pipetting and titered. HA assays showed that approximately 80% of the HA units present in the clarified cell supernatant were recovered as purified material following the procedure described above.

#### Virus Inactivation

Five hundred thousand HA units of virus was diluted to a final volume of 20 mL in PBS. A chemical reaction using 10 formaldehyde was done to inactivate the viruses. Formaldehyde (2% v/v in PBS, Thermo Scientific) was added to a final concentration of 0.02%, and the solution was incubated at 22° C. for 13 hours while rotating at 90 rpm. Residual formaldehyde was neutralized by addition of sodium 15 bisulfite at a 1:1 molar ratio of sulfite ion to formaldehyde. Virus inactivation was confirmed by sequential passage of the undiluted virus suspension on MDCK cells. HA assays performed on the virus suspension before and after inactivation showed that the inactivation process did not reduce 20 the quantity of HA units when compared to that of the starting material.

Other inactivation methods involving chemical reactions, such as BPL (betapropiolactone) and BEI (binary ethylenimine) are used to inactivate the viruses.

These chemical inactivation methods produce enumerable structural changes, including for example, formation of new chemical bonds via chemical crosslinking, irreversible chemical alteration of the nucleic acid and protein coat (Uittenbogaard, 2011, Journal of Biological Chemistry, 286 <sup>30</sup> (42): pp36198-36214; Gard, Bull. Wld Hlth Org., 1957, 17, 979-989).

### Production of antibodies

Purified, inactivated H3N2 CIV #8 was mixed at a 1:1 ratio with TiterMax Gold adjuvant (Sigma-Aldrich). Ten <sup>35</sup> female specific pathogen free New Zealand White rabbits 10-16 weeks of age (Harlan) were injected subcutaneously in the hind quarter with 3 or 4 doses of 1,500 HA units of virus per dose. One week after the last vaccine dose, animals were anesthetized, and blood was collected. Serum samples <sup>40</sup> were tested for serum neutralizing antibody titers against H3N2 CIV.

### Example 4 Viral Antigenic Analysis

### Viral antigenic analysis

Antibodies that bind to the HA protein on the surface of influenza virus particles can inhibit particle-induced red blood cell (RBC) agglutination in the context of an HA assay if the antibodies are present in sufficient quantities. Thus, the 50 hemagglutination inhibition (HAI) assay is routinely used to (i) measure the quantity or dilution of an antibody required to inhibit RBC agglutination by a given virus strain (i.e. HAI titer), and (ii) to evaluate the antigenic relatedness of HA proteins present in virus particles from two or more different 55 virus strains. Tests of antigenic relatedness are performed with antisera raised against one virus strain (i.e. the homologous strain) and test the capacity of that antisera to inhibit RBC agglutination by heterologous virus strains. A heterologous virus is typically considered antigenically distinct 60 from the homologous virus if the HAI titer for the heterologous virus is >8-fold less than the HAI titer for the homologous virus.

An HAI assay employing serum from dogs vaccinated with a canarypox vector expressing the HA protein from 65 A/equine/Ohio/1/2003 H3N8 virus was used to test the antigenic relatedness of H3N8 and H3N2 CIV. This serum

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was previously determined to have an HAI titer of 512 when tested against A/canine/Colorado/8880/2006, a H3N8 canine influenza virus. The H3N2 CIV isolate #4 (egg passage 2) and A/canine/Colorado/8880/2006 were diluted to 8 HA units/25 uL in PBS and incubated with 2-fold dilutions of serum (25 µL per dilution) for 1 hour at 22° C. An equal volume (50 uL) of 0.5% chicken RBCs was added to each sample, and the solutions were incubated for an additional hour at 22° C. Antibodies in the serum failed to inhibit RBC agglutination by H3N2 CIV at all dilutions tested (1:8-1:2048, HAI titer <8). In contrast, the serum inhibited RBC agglutination by A/canine/Colorado/8880/ 2006 at a maximal dilution of 1:512 (HAI titer 512). Thus, antibodies in this serum that recognize the HA protein of a H3N8 CIV do not bind to the HA protein of H3N2 CIV with high affinity, suggesting that the viruses are antigenically distinct.

# Example 5 Production of Vaccine Active Ingredient and Preparation of Vaccines

To produce CIV H3N2 active ingredient (AI) for the preparation of inactivated CIV H3N2 vaccine, MDCK cells are used to amplify the CIV H3N2 virus in roller bottles for multiple passages according to standard virus production procedures.

The vaccine is prepared by mixing the CIV H3N2 active ingredient with an adjuvant. The adjuvant is aluminum hydroxide (or aluminum phosphate) or LR4 emulsion (U.S. Pat. No. 7,691,368) which contains 12.5% of "incomplete LR emulsion" and 87.5% of aqueous phase (containing the active ingredient).

# Example 6 Evaluation of H3N2 Monovalent Vaccine Safety and Immunogenicity in Animals

Vaccines were formulated as monovalent, aqueous solutions containing a 1× (1000 HAU) or 2× (1920 HAU) dose of whole, inactivated H3N2 CIV (PTA-122265 (virus ID #4)) and aluminum hydroxide (adjuvant). The placebo vaccine lacked antigen but contained adjuvant at the same concentration as the test vaccines.

Injection site reactions and clinical signs

Thirty CIV seronegative six-week old, commercial source beagles were randomized to three vaccination groups, containing 10 dogs each. Dogs in Groups 1 and 2 received 2× and 1× doses of the H3N2 CIV vaccine, respectively (Table 4). Dogs in Group 3 received a placebo vaccine. Each dog was vaccinated twice, 21 days apart, with 1 mL of vaccine, subcutaneously over the scapula. The dogs were monitored for injection site reactions and temperature elevations for three days after each vaccination and on days 14 and 27 (day prior to challenge). Blood samples were collected on days 0 (prior to vaccination), 7, 14, 21 (prior to vaccination), 27, and 35.

TABLE 4

Study Groups							
Group	Vaccine	Vaccination Route	Vaccination Frequency	No. of dogs	Challenge		
1	2X CIV	Subcutaneous	Twice,	10	H3N2 CIV		
2	1X CIV	Subcutaneous	21 days apart Twice, 21 days apart	10	H3N2 CIV		

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TABLE 4-continued

Study Groups							
Group	Vaccine	Vaccination Route	Vaccination Frequency	No. of dogs	Challenge		
3	Placebo	Subcutaneous	Twice, 21 days apart	10	H3N2 CIV		

Seven days after the second vaccination, the dogs were challenged with virulent H3N2 CIV (PTA-122266 (virus ID #8)) via aerosolization in a closed chamber using a commercial nebulizer. During challenge, dogs were randomized to challenge chambers using treatment group and litter as blocking factors such that each treatment group, litter, and sex (if possible) was represented in each challenge chamber run. After challenge, dogs were randomized to post-challenge (PC) pens such that each pen contained dogs from all three treatment groups and each chamber run from the challenge phase.

The dogs were observed for cough, fever, mucopurulent nasal discharge, and other clinical signs for seven days. Nasal swabs for virus isolation were collected on days 3, 4, and 5 PC. A dog was classified as having disease due to CIV  $\,^{25}$ if it developed cough in addition to either fever or mucopurulent nasal discharge. A dog was considered febrile when the rectal temperature was ≥39.7° C. and 0.5° C. above baseline (day 0 rectal temperature). The challenge was considered valid when at least 60% of the placebo vaccinated dogs developed disease due to CIV.

Following each vaccination of dogs with the H3N2 CIV vaccine at 1× and 2× doses, no systemic or injection site adverse reactions were observed. Prior to challenge, no dogs 35 n\*: number of dogs (out of 10 dogs) with symptoms. in any group showed clinical signs of disease. After challenge, the first clinical signs of disease (cough, mucopurulent nasal discharge, and fever) were observed two days post-challenge (DPC), similar to dogs with natural and experimental CIV infection (Table 5).

TABLE 5

Number of dogs expressing clinical signs of CIV disease post-challenge						
Vaccine Group	Cough	Fever	Mucopurulent Nasal Discharge			
2X CIV (n = 10)	0	1	0			
1X  CIV  (n = 10)	0	0	0			
Placebo (n = 10)	9	5	10			

Cough and mucopurulent nasal discharge were the most predominant clinical signs of disease, with most dogs in the placebo group coughing for 2 days and showing mucopurulent discharge for 4 days (Tables 6 and 7). Ninety percent 55 (90%) of the dogs in the placebo group were observed with cough, and 100% of the placebo group developed mucopurulent nasal discharge. No animals in the 2×CIV or 1×CIV vaccine groups developed cough or mucopurulent discharge. The days with highest frequency were 3 DPC for cough (8 60 dogs) and 4 DPC for mucopurulent nasal discharge (10 dogs). Two days post-challenge was the day with the highest frequency of fever with 6 febrile dogs—five in the placebo group and one in the 2×CIV vaccine group. Most dogs had one instance of fever, while one dog in the placebo group had two days of fever (Table 8). Rectal temperatures for dogs with fever ranged from 39.7° C. to 40.4° C.

20

TABLE 6

Total days with cough					
	Number of days of cough				
Vaccine Group	2	3	4	5	
2X CIV (n* = 0) 1X CIV (n = 0) Placebo (n = 9)	  6	_ _ 1	_ _ 1	_ _ 1	

n\*: number of dogs (out of 10 dogs) with symptom.

TABLE 7

15	Total days with mucopurulent nasal discharge							
		Number	of days of m	ıucopurulei	nt nasal di	ischarge		
	Vaccine Group	2	3	4	5	6		
20	2X CIV (n* = 0)	_	_	_	_	_		
-0	1X CIV (n = 0) Placebo $(n = 10)$	2	2	3	1	2		

n\*: number of dogs (out of 10 dogs) with symptoms.

TABLE 8

	Total days with fever					
Number of days of fever						
)	Vaccine Group	1	2			
•	2X CIV (n* = 1) 1X CIV (n = 0) Placebo (n = 5)	1 - 4	<u>_</u> 1			

Ninety percent (90%) of the dogs in the placebo group met the case definition for clinical disease, thereby validating the challenge. The incidence of disease in dogs administered the H3N2 CIV vaccines was significantly reduced compared to the placebo vaccinated dogs (p=0.0001) (Table 9). Clinical disease was prevented in 100% of the CIV vaccinates, underscoring the effectiveness of the vaccines at preventing clinical disease.

TABLE 9

2X dose and 1X dose H3N2 Vaccine Group Prevented Fractions for CIV disease						
Vaccine Group	Number of dogs with disease	P-value (Fisher's Exact Test)	Prevented Fraction (95% CI)*			
2X CIV (n = 10)	0	0.0001	1.00			
Placebo $(n = 10)$	9		(0.67, 1.00)			
1X CIV (n = 10)	0	0.0001	1.00 (0.67, 1.00)			

95% CI\*: 95% confidence interval

### Nasal shedding and serology

Fluids expressed from nasal swabs were inoculated into the allantoic cavity of 3 embryonic chicken eggs. Eggs were incubated for 3 days at 37° C. and tested for the presence of virus by HA assay. Specimens were considered positive for virus if at least 1 of 3 eggs contained detectable virus. Specimens were considered negative for virus if at least 2 eggs were viable at the time of testing, and no eggs con-

tained detectable virus. Positive samples and samples for which 2 or 3 eggs were inviable at the time of testing were tested by  ${\rm TCID}_{50}$  assay.

Heat-inactivated serum samples were incubated with 2.5% v/v washed cRBCs for 30 min at room temperature, and cRBCs were pelleted by centrifugation. The pellet supernatant was used to perform the HAI assay using a final concentration of 0.25% v/v cRBCs. The HAI titer of each sample was recorded as the reciprocal of the highest serum dilution that completely inhibited cRBC agglutination by 8 HA units of H3N2 CIV. A titer of <4 was considered negative for CIV serum antibody.

Nasal swabs were collected to gauge the capability of the test vaccines to prevent viral shedding after challenge. In this study, the collection days also corresponded with the days of highest frequency of cough and mucopurulent nasal discharge. All dogs in the placebo group exhibited nasal shedding of CIV, and shedding was significantly reduced in the 2× and 1× dose groups on each collection day (Table 10). The reduction in shedding in the test vaccine groups equated to an 80% preventable fraction (Table 11). Similarly, the CIV vaccines administered in this study reduced shedding to undetectable levels in 100% of the CIV vaccinated dogs at 5 DPC.

TABLE 10

Incidence of nasal CIV shedding						
_	Number of dogs with positive nasal shedding					
Vaccine Group	3 DPC	4 DPC	5 DPC			
	(Day 31)	(Day 32)	(Day 33)			
2X CIV (n = 10)	0 <sup>§a</sup>	2 <sup>§</sup> <sup>b</sup>	0 <sup>§a</sup>			
1X CIV (n = 10)	1 <sup>§c</sup>	1 <sup>§c</sup>	0 <sup>§a</sup>			
Placebo (n = 10)	10	10	10			

§significant difference between test vaccine and placebo

TABLE 11

2X dose and 1X dose H3N2 Vaccine Group Prevented Fractions for nasal shedding					
Vaccine Group	Number of dogs with positive nasal shedding	P-value (Fisher's Exact Test)	Prevented Fraction (95% CI)		
2X CIV (n = 10) Placebo (n = 10)	2 10	0.0007	0.80 (0.46, 0.96)	50	
1X  CIV  (n = 10)	2	0.0007	0.80 (0.46, 0.96)		

This study is consistent with the observation that dogs not 55 showing clinical signs of disease may still shed virus. The study found that 90% of the placebo dogs exhibited clinical disease, while 100% of the placebo dogs shed infectious virus. This result emphasizes the importance of limiting virus shedding as part of strategies to control virus trans- 60 mission

All animals were seronegative prior to the initiation of the study, and all dogs in the placebo group were seronegative prior to challenge (Table 12). On day 21, the proportion of dogs that seroconverted in the 2×CIV group was significantly higher than in the placebo group (p=0.0031), and on day 27, all dogs in the 2× and 1×CIV groups had serocon-

verted. Hence, seroconversion correlated with protection of the vaccinated dogs from clinical disease.

TABLE 12

3	Dogs seropositive (≥4) for CIV antibodies								
VaccineNumber of dogs seroconverted									
	Group	Day 0*	Day 7	Day 14	Day 21*	Day 27#	Day 35 <sup>†</sup>		
10	2X CIV	0	0	1	7 <sup>§a</sup>	10 <sup>§b</sup>	10		
	(n = 10) 1X CIV	0	0	0	2	$10^{\S b}$	10		
	(n = 10) Placebo (n = 10)	0	0	0	0	0	6		
15	(H = 10)								

<sup>\*</sup>Vaccination days-sample collected prior to vaccination

30

This study demonstrates that vaccination of dogs with an inactivated H3N2 CIV vaccine was well tolerated, produced seroconversion, and successfully protected 100% of vaccinates from clinical disease caused by challenge with virulent virus. Notably, the vaccines also reduced nasal shedding of infectious challenge virus, substantiating their use as safe and effective measures to alleviate signs of disease caused by CIV and potentially control virus transmission.

Example 6 Virus Propagation and Genomic RNA of H3N8 CIV Strains

Viruses from H3N8 CIV strains CT/85863/11, NY/120106/11 and WY/86033/07 were isolated in specific pathogen free embryonic chicken eggs and subsequently propagated in eggs or MDCK cells at 37° C. for 72 hours (eggs) or 36-48 hours (cells). Viruses were titrated by HA or TCID<sub>50</sub> assay as described for H3N2 CIV. The nucleotide sequences of the viral HA and NA genes from each virus strain were determined as for H3N2 CIV using the gene specific primers in FIG. 1.

The HA and NA polynucleotide and protein sequences for H3N8 CIV strains CT/85863/11, NY/120106/11 and WY/86033/07 are designated SEQ ID NOs:30-35 and 37-42 as shown in FIGS. 1 and 5.

# Example 7 Preparation of Bivalent H3N2 and H3N8 CIV Vaccines

Viruses used in vaccine preparation were produced by infection of confluent monolayers of MDCK cells in roller bottle cultures at MOIs ranging from 0.0001 to 0.00001 in 200 mL of MEM supplemented with 1-2 ug/mL of porcineorigin trypsin (Sigma-Aldrich) per roller bottle. Cultures were incubated at 37° C. and 0.5 rpm and harvested 24 hours (H3N2 CIV) or 36-42 hours (H3N8 CIV) after inoculation of the cells with virus. Cell debris was removed from the virus suspension by centrifugation at 1,000×g or filtration through membranes containing 5 µM diameter pores. To inactivate infectious virus in the suspensions, formaldehyde was added to a final (v/v) concentration of 0.04%, and the reactions were incubated for 24 hours at 37° C. Inactivated virus preparations were stored at 4° C. until further use. In some cases, inactivated viral antigens were concentrated by ultrafiltration using polysulfone membranes containing 10

 $<sup>^{</sup>a}$ p < 0.0001

 $<sup>^{</sup>b}$ p = 0.0007

 $<sup>^{</sup>c}p = 0.0001$ 

<sup>\*</sup>One day prior to challenge

<sup>†</sup>Seven days post-challenge

<sup>§</sup>significant difference between test vaccine and placebo

 $<sup>^{</sup>a}p = 0.0031$ 

 $<sup>^{</sup>b}p < 0.0001$ 

Bivalent vaccines were prepared by mixing the desired quantity of each inactivated vaccine antigen with an adjuvant overnight at 4° C. The adjuvant is aluminum hydroxide for aluminum phosphate) or LR4 emulsion (U.S. Pat. No. 7,691,368) which contains 12.5% of "incomplete LR emulsion" and 87.5% of aqueous phase (containing the active ingredient). Prepared vaccines were aliquoted into glass vials, sealed, and stored at 4° C. until use.

Example 8 Evaluation of H3N2/H3N8 Bivalent Vaccine Safety and Immunogenicity in Animals

Example 8.1 Efficacy of a Canine Influenza Virus H3N2/H3N8 Combination Vaccine Against H3N8 Challenge in Dogs

The goal of the study is to evaluate the serological response, adverse events (interference) and efficacy of an 20 injectable canine influenza virus H3N2/H3N8 combination vaccine in a vaccination-challenge model in dogs.

Thirty CIV seronegative six-week (+/-6 days) old, commercial source beagles were randomized to four vaccination groups as shown in Table 14 in one study. Dogs in Groups A and B received 1× and 2× doses of the H3N2/H3N8 combo CIV vaccine, respectively (Tables 13 and 14). Dogs in group C received a dose of H3N2 CIV vaccine. Dogs in Group D received a placebo vaccine. In another study, thirty CIV seronegative six-week (+/-6 days) old, commercial source beagles were randomized to two vaccination groups as shown in Table 15. Dogs in Group A received one dose of the H3N2/H3N8 combo CIV vaccine. Dogs in Group B received a placebo vaccine. Each dog was vaccinated twice, 21 days apart, with 1 mL of vaccine, subcutaneously. The dogs were monitored for injection site reactions and tem- 35 perature elevations for three days after each vaccination and on days 7, 14, 21 (prior to vaccination), 22-24, 28 (prior to challenge) and 38 (end of study). Blood samples were collected on days 0 (prior to vaccination), 7, 14, 21 (prior to vaccination), 28 (prior to challenge), and 38 (end of study).

**24**TABLE 14-continued

Study design							
Group	Vaccine	Dose Volume	Route of Adminis- tration	Frequency of Administration	Number of animals		
В	High dose	1 ml	SC	Twice,	5		
С	H3N2/H3N8 CIV H3N2	1 ml	sc	21 days apart Twice, 21 days apart	10		
D	Control	1 ml	SC	Twice, 21 days apart	5		

TABLE 15

Study design								
Group	Vaccine	Dose Volume	Route of Adminis- tration	Frequency of Administration	Number of animals			
A	H3N2/H3N8*	1 ml	SC	Twice,	15			
В	Control**	1 ml	SC	21 days apart Twice, 21 days apart	15			

H3N2/H3N8\*: Inactivated H3N2 and H3N8 adjuvanted with Al(OH)<sub>3</sub>; 1000 HAU of H3N2 and 750 HAU of H3N8 per dose; 1 ml = 1 dose Control\*\*: PBS adjuvanted with Al(OH)<sub>3</sub>; 1 ml = 1 dose

Seven days after the second vaccination (day 28), the dogs were challenged with virulent H3N8 CIV strains WY/86033/07 and NY/120106/11 via aerosolization in a closed chamber using a commercial nebulizer. During challenge, dogs were randomized to challenge chambers using treatment group and litter as blocking factors such that each treatment group, litter, and sex (if possible) was represented in each challenge chamber run. After challenge, dogs were randomized to post-challenge (PC) pens such that each pen contained dogs from all three treatment groups and each chamber run from the challenge phase.

TABLE 13

Vaccine formulation					
	H3N2/H3N8 Low dose(1X H3N2 and 1X H3N8)	H3N2/H3N8 High dose (1X H3N2 and 2X H3N8)	CIV H3N2	Control	
Vaccine formulation	Inactivated H3N2* and H3N8** adjuvanted with Al(OH) <sub>3</sub>	Inactivated H3N2 and H3N8 adjuvanted with Al(OH) <sub>3</sub>	Inactivated H3N2 adjuvanted with Al(OH) <sub>3</sub>	PBS adjuvanted with Al(OH) <sub>3</sub>	
dose	1 /2	\ /2	1000 HAU of H3N2 per dose; 1 ml = dose	1 ml = 1 dose	

H3N2\*: H3N2 CIV strain PTA-122265 (virus ID #4) H3N8\*\*: H3N8 CIV strain A/Ca/CT/85863/11

TABLE 14

Study design						
Group	Vaccine	Dose Volume	Route of Adminis- tration	Frequency of Administration	Number of animals	
A	Low dose H3N2/H3N8	1 ml	SC	Twice, 21 days apart	10	

The dogs were observed for cough, fever, mucopurulent nasal discharge, and other clinical signs for ten days. Nasal swabs for virus isolation were collected three to six times after PC. A dog was classified as having disease due to CIV if it developed cough in addition to either fever or mucopurulent nasal discharge. A dog was considered febrile when the rectal temperature was ≥39.7° C. and 0.5° C. above baseline (day 0 rectal temperature). The challenge was considered valid when at least 60% of the placebo vaccinated dogs developed disease due to CIV.

The results show that vaccination of dogs with an inactivated H3N2/H3N8 combination vaccine is well tolerated, produced seroconversion, and successfully protected vaccinates from clinical disease caused by challenge with virulent H3N8 virus. The H3N2/H3N8 combination vaccine also demonstrates the lack of interference between the inactivated H3N2 vaccine and the inactivated H3N8 vaccine when formulated together in a combination vaccine.

Example 8.2 Efficacy of a Canine Influenza Virus H3N2/H3N8 Combination Vaccine Against H3N2 Challenge in Dogs

The goal of the study is to evaluate the efficacy of an injectable canine influenza virus H3N2/H3N8 combination 15 vaccine against lung lesions induced by CIV H3N2 in a vaccination-challenge model.

Thirty CIV seronegative six-week (+/-6 days) old, commercial source beagles were randomized to two vaccination groups as shown in Table 16. Dogs in Group A received one 20 dose of the H3N2/H3N8 combo CIV vaccine. Dogs in Group B received a placebo vaccine. Each dog was vaccinated twice, 21 days apart, with 1 mL of vaccine, subcutaneously. The dogs were monitored for injection site reactions and temperature elevations for three days after each vaccination and on days 7, 14, 21 (prior to vaccination), 22-24, 28 (prior to challenge), 29-35, and 36 (end of study). Blood samples were collected on days 0 (prior to vaccination), 7, 14, 21 (prior to vaccination), 28 (prior to challenge), and 38 (end of study).

TABLE 16

Study design						
Group	Vaccine	Dose Volume	Route of Adminis- tration	Frequency of Administration	Number of animals	
A	H3N2/H3N8* Low dose	1 ml	SC	Twice, 21 days apart	15	

**26**TABLE 16-continued

	Study design					
Group	Vaccine	Dose Volume	Route of Adminis- tration	Frequency of Administration	Number of animals	
В	Control**	1 ml	SC	Twice, 21 days apart	15	

H3N2/H3N8\*: Inactivated H3N2 and H3N8 adjuvanted with Al(OH)<sub>3</sub>; 1000 HAU of H3N2 and 750 HAU of H3N8 per dose; 1 ml = 1 dose Control\*\*: PBS adjuvanted with Al(OH)<sub>3</sub>; 1 ml = 1 dose

One week after the second vaccination the dogs are challenged with CIV H3N2 (PTA-122266 (virus ID #8) via aerosolization as described in Example 8.1. The dogs are observed for cough, fever, mucopurulent nasal discharge, and other clinical signs for seven days. Nasal swabs for virus isolation are collected three to six times after PC. Data are analyzed based on the criteria set out in Example 8.1.

The results show that vaccination of dogs with an inactivated H3N2/H3N8 combination vaccine is well tolerated, produced seroconversion, and successfully protected vaccinates from clinical disease caused by challenge with virulent H3N2 virus. The H3N2/H3N8 combination vaccine also demonstrates the lack of interference between the inactivated H3N2 vaccine and the inactivated H3N8 vaccine when formulated together in a combination vaccine.

Having thus described in detail embodiments of the present disclosure, it is to be understood that the disclosure defined by the above examples is not to be limited to particular details set forth in the above description as many apparent variations thereof are possible without departing from the spirit or scope of the present disclosure.

All documents cited or referenced herein ("herein cited documents"), and all documents cited or referenced in herein cited documents, together with any manufacturer's instructions, descriptions, product specifications, and product sheets for any products mentioned herein or in any document incorporated by reference herein, are hereby incorporated herein by reference, and may be employed in the practice of the invention.

SEQUENCE LISTING

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Pro	290				Ile	295					300	Asn	Ī		Ile
305			-		Phe 310					315			-	-	320
-		-	-	325	Lys				330	-				335	
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Ser Asn Ser Thr Ile His Asp Arg Thr Ser His Arg Thr Leu Leu Met

Asn Glu Leu Gly Val Pro Phe His Leu Gly Thr Lys Gln Val Cys Ile

155

165 170 175

130 135

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															ccgta		240
	_			=		_			_					_	ctttc		300
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Cys Tyr Gln Phe Al	la Leu Gly Gln 135	Gly Thr Thr	Leu Asn Asn 140	Lys His	
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Asn Glu Leu Gly Val Pro Phe His Leu Gly Thr Lys Gln Val Cys Ile \$165\$

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	)> SE	-														
				_											cagta caaaat	60 120
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Asp Ile Trp Val Thr Arg Glu Pro Tyr Val Ser Cys Asp His Ser Lys 115 120 125	

Ser Asn Ser Thr Ile His Asp Arg Thr Ser His Arg Thr Leu Leu Met

Asn Glu Leu Gly Val Pro Phe His Leu Gly Thr Lys Gln Val Cys Ile \$165\$

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Ser	Phe	Phe	Ser	Arg 165	Leu	Asn	Trp	Leu	Thr 170	Lys	Ser	Gly	Ser	Ser 175	Tyr		
Pro	Thr	Leu	Asn 180	Val	Thr	Met	Pro	Asn 185	Asn	Lys	Asn	Phe	Asp 190	Lys	Leu		
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Gln	His 450	Thr	Ile	Asp	Leu	Thr 455	Asp	Ala	Glu	Met	Asn 460	Lys	Leu	Phe	Glu	
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Сув	Phe	Lys	Ile	Tyr 485	His	Lys	Сув	Asp	Asn 490	Ala	Cys	Ile	Glu	Ser 495	Ile	
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Trp	Ile 530	Leu	Trp	Ile	Ser	Phe 535	Ala	Ile	Ser	Cys	Phe 540	Leu	Ile	Cys	Val	
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Trp Ser Ala Thr Ala Cys His Asp Gly Lys Lys Trp Met Thr Val Gly 180 185 190

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His Ala Val Ala As<br/>n Gly Thr Leu Val Lys Thr Met Ser Asp Asp Gl<br/>n  $\,$ 

Ile Glu Val Thr Asn Val Thr Glu Leu Val Gln Ser Ile Ser Met Gly 55

Lys Ile Cys Asn Lys Ser Tyr Arg Val Leu Asp Gly Arg Asn Cys Thr 65 70 75 80

Leu Ile Asp Ala Met Leu Gly Asp Pro Gln Cys Asp Ala Phe Gln Tyr

Glu Ser Trp Asp Leu Phe Ile Glu Arg Ser Asn Ala Phe Ser Asn Cys 105

Tyr Pro Tyr Asp Ile Pro Asp Tyr Ala Ser Leu Arg Ser Ile Val Ala 120

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Gln	His 450	Thr	Ile	Asp	Leu	Thr 455	Asp	Ala	Glu	Met	Asn 460	ГÀа	Leu	Phe	Glu
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<223> OTHER INFORMATION: DNA encoding HA protein of H3N8 CIV strain WY/86033/07

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Phe Val Ile			Val Ser 120	Cys Ser	Pro Ser 125	Glu Cys	Arg				
Thr Phe Phe	e Leu Thr	Gln Gly S 135	Ser Leu	Leu Asn	Asp Lys 140	His Ser	Asn				
Gly Thr Ile	e Lys Asp	Arg Ser I 150	Pro Tyr	Arg Thr 155	Leu Met	Ser Val	Lys 160				
Ile Gly Gli	n Ser Pro 165	Asn Val 1	Tyr Gln	Ala Arg 170	Phe Glu	Ser Val 175					
Trp Ser Ala	a Thr Ala 180	Cys His A	Asp Gly 185	Lys Lys	Trp Met	Thr Val	Gly				
Val Thr Gly	-		Ala Ile 200	Ala Val	Val Asn 205	Tyr Gly	Gly				
Val Pro Val	l Asp Ile	Ile Asn S 215	Ser Trp	Ala Gly	Asp Ile 220	Leu Arg	Thr				
Gln Glu Se: 225		Thr Cys I	Ile Lys	Gly Asp 235	Cys Tyr	Trp Val	Met 240				
Thr Asp Gl	y Pro Ala 245	Asn Arg (	Gln Ala	Glu Tyr 250	Arg Ile	Phe Lys 255					
Lya Aap Gl	y Arg Val 260	Ile Gly (	Gln Thr 265	Asp Ile	Ser Phe	Asn Gly 270	Gly				

His Ile Glu Glu Cys Ser Cys Tyr Pro Asn Glu Gly Lys Val Glu Cys

-concinued
275 280 285
Ile Cys Arg Asp Asn Trp Thr Gly Thr Asn Arg Pro Ile Leu Val Ile 290 295 300
Ser Ser Asp Leu Ser Tyr Thr Val Gly Tyr Leu Cys Ala Gly Ile Pro 305 310 315 320
Thr Asp Thr Pro Arg Gly Glu Asp Ser Gln Phe Thr Gly Ser Cys Thr 325 330 335
Ser Pro Leu Gly Asn Lys Gly Tyr Gly Val Lys Gly Phe Gly Phe Arg 340 345 350
Gln Gly Thr Asp Val Trp Ala Gly Arg Thr Ile Ser Arg Thr Ser Arg 355 360 365
Ser Gly Phe Glu Ile Ile Lys Ile Arg Asn Gly Trp Thr Gln Asn Ser 370 375 380
Lys Asp Gln Ile Arg Arg Gln Val Ile Ile Asp Asp Pro Asn Trp Ser 385 390 395 400
Gly Tyr Ser Gly Ser Phe Thr Leu Pro Val Glu Leu Thr Lys Lys Gly 405 410 415
Cys Leu Val Pro Cys Phe Trp Val Glu Met Ile Arg Gly Lys Pro Glu 420 425 430
Glu Thr Thr Ile Trp Thr Ser Ser Ser Ile Val Met Cys Gly Val 435 440 445
Asp His Lys Ile Ala Ser Trp Ser Trp His Asp Gly Ala Ile Leu Pro 450 455 460
Phe Asp Ile Asp Lys Met 465 470
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aatatcattc tccatgtagt cagcattata gtaacagtac tggtcctcaa taacaataga 120
acagatetga aetgeaaagg gaegateata agagagtaea atgaaacagt aagagtagaa 180
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gcaattette	cctttgacat	cgataagatg				1410		

What we claim is:

- derived from canine isolates (H3N2 CIV), wherein the H3N2 CIV comprises,
  - a) a polynucleotide encoding an HA protein having at least 99.6% sequence identity to the sequence as set forth in SEQ ID NO:2, 4, or 6; and/or
  - b) a polynucleotide encoding an NA protein having at least 99% sequence identity to the sequence as set forth in SEQ ID NO:8, 10 or 12; and/or
  - c) an HA polynucleotide having at least 99.2% sequence identity to the sequence as set forth in SEQ ID NO:1; 30 or having at least 99.6% sequence identity to the sequence as set forth in SEQ ID NO:3; or having at least 99.9% sequence identity to the sequence as set forth in SEQ ID NO:5; and/or
  - d) an NA polynucleotide having at least 99.9% sequence  $^{35}$ identity to the sequence set forth in SEQ ID NO:7 or 9.
- 2. The composition of claim 1, wherein the H3N2 CIV is deposited at ATCC under the deposit number of PTA-122265 or PTA-122266 or is a progeny or descendant of PTA-122265 or PTA-122266 wherein the progeny or 40 descendant comprises a polynucleotide encoding an HA protein having at least 99.6% sequence identity to the sequence as set forth in SEQ ID NO:2, 4, or 6.
- 3. The composition of claim 1, wherein the H3N2 CIV is deposited at ATCC under the deposit number of PTA- 45 122265 or PTA-122266, or is a progeny or descendant of PTA-122265 or PTA-122266, wherein the progeny or descendant comprises a polynucleotide encoding an NA protein having at least 99% sequence identity to the sequence as set forth in SEQ ID NO:8, 10, or 12.
- 4. The composition of claim 1, wherein the H3N2 CIV is inactivated.
- 5. The composition of claim 1, wherein the composition further comprises an inactivated H3N8 CIV.
- 6. The composition of claim 5, wherein the H3N8 CIV 55 comprises a polynucleotide encoding an HA protein having at least 98% sequence identity SEQ ID NO:31, 35, or 39.
- 7. The composition of claim 5, wherein the H3N8 CIV comprises polynucleotide encoding an NA protein having at least 98% sequence identity SEQ ID NO:33, 37, or 41.

- 8. The composition of claim 1, wherein the composition 1. A composition comprising an H3N2 influenza virus 20 or vaccine further comprises one or more pharmaceutically or veterinarily acceptable carrier, adjuvant, vehicle, or excinient.
  - **9**. A method of vaccinating an animal, or for inducing an immunogenic or protective response against influenza virus infection in canines (CIV), or for reducing CIV viral shedding in an infected animal, comprising at least one administration of the composition of claim 1.
  - 10. The method of claim 9, wherein the animal is a canine or feline.
  - 11. A diagnostic method of the infection of H3N2 influenza virus in dogs or cats (H3N2 CIV), wherein a sample of physiological fluid or a dog or cat tissue sampling and a diagnostic reagent specific to H3N2 CIV are put together, and the potential presence of H3N2 CIV antigen, antibody or nucleic acid is revealed within this sample or sampling, wherein the diagnostic reagent specific to H3N2 CIV comprises
    - a) a polynucleotide encoding an HA protein having at least 99.6% sequence identity to the sequence as set forth in SEQ ID NO:2, 4, or 6; and/or
    - b) a polynucleotide encoding an NA protein having at least 99% sequence identity to the sequence as set forth in SEQ ID NO:8, 10 or 12; and/or
    - c) an HA polynucleotide having at least 99.2% sequence identity to the sequence as set forth in SEQ ID NO:1; or having at least 99.6% sequence identity to the sequence as set forth in SEQ ID NO:3; or having at least 99.9% sequence identity to the sequence as set forth in SEQ ID NO:5; and/or
    - d) an NA polynucleotide having at least 99.9% sequence identity to the sequence set forth in SEQ ID NO:7 or 9.
  - 12. The diagnostic method of claim 11, wherein the diagnostic reagent comprises a specific H3N2 CIV antigen selected from the group consisting of HA and NA, and wherein the antigen allows the detection of antibodies within the sample or sampling as being H3N2 CIV.
  - 13. The diagnostic method of claim 11, wherein the method is Western blotting, immunofluoroescence, ELISA or immunochromatography.