

(19) **DANMARK**



Patent- og
Varemærkestyrelsen

(10) **DK/EP 3577219 T3**

(12) Oversættelse af
europæisk patentskrift

-
- (51) Int.Cl.: **C 12 N 9/26 (2006.01)**
- (45) Oversættelsen bekendtgjort den: **2023-11-13**
- (80) Dato for Den Europæiske Patentmyndigheds bekendtgørelse om meddelelse af patentet: **2023-08-23**
- (86) Europæisk ansøgning nr.: **18708589.9**
- (86) Europæisk indleveringsdag: **2018-01-30**
- (87) Den europæiske ansøgnings publiceringsdag: **2019-12-11**
- (86) International ansøgning nr.: **EP2018052197**
- (87) Internationalt publikationsnr.: **WO2018141707**
- (30) Prioritet: **2017-02-01 IN 201741003686** **2017-03-27 EP 17163114**
- (84) Designerede stater: **AL AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO PL PT RO RS SE SI SK SM TR**
- (73) Patenthaver: **Novozymes A/S, Krogshøjvej 36, 2880 Bagsværd, Danmark**
- (72) Opfinder: **Andersen, Carsten, Krogshøjvej 36, 2880 Bagsværd, Danmark**
DAMAGER, Iben, Krogshøjvej 36, 2880 Bagsværd, Danmark
GHADIYARAM, Chakshusmathi, Plot No. 32, 47-50 EPIP Area, Whitefield, Bangalore 560066, Indien
SAINATHAN, Rajendra, Kulothungan, Plot No. 32, 47-50 EPIP Area, Whitefield, Bangalore 560066, Indien
IYER, Padma, Venkatachalam, Plot No. 32, 47-50 EPIP Area, Whitefield, Bangalore 560066, Indien
- (54) Benævnelse: **ALFA-AMYLASE-VARIANTER**
- (56) Fremdragne publikationer:
EP-A1- 2 540 824
EP-A2- 3 121 270
WO-A1-2013/001078

Reference to a Sequence Listing

This patent contains a Sequence Listing.

BACKGROUND OF THE INVENTION

5

Field of the Invention

The present invention relates to variants of an alpha-amylase, polynucleotides encoding the variants, nucleic acid constructs, expression vectors and host cells comprising a polynucleotide of the invention and methods of producing the variants.

10

Description of the Related Art

Alpha-amylases (alpha-1,4-glucan-4-glucanohydrolases, E.C. 3.2.1.1) constitute a group of enzymes, which catalyses hydrolysis of starch and other linear and branched 1,4-glucosidic oligo- and polysaccharides.

15

There is a long history of industrial use of alpha-amylases in several known applications such as detergent, baking, brewing, starch liquefaction and saccharification e.g. in preparation of high fructose syrups or as part of ethanol production from starch. These and other applications of alpha-amylases are known and utilize alpha-amylases derived from microorganisms, in particular bacterial alpha-amylases.

20

Among the first bacterial alpha-amylases to be used were an alpha-amylase from *B.licheniformis*, also known as Termamyl which have been extensively characterized and the crystal structure has been determined for this enzyme. Alkaline amylases, such as AA560, form a particular group of alpha-amylases that have found use in detergents. Many of these known bacterial amylases have been modified in order to improve their functionality in a particular application.

25

Bacillus amylases, such as Termamyl, AA560 (WO 2000/060060) and SP707 (described by Tsukamoto et al., 1988, *Biochem. Biophys. Res. Comm.* 151: 25-31) form a particular group of alpha-amylases that have found use in detergents. These amylases have been modified to improve the stability in detergents. WO 96/23873 e.g. disclose to delete the amino acids 181+182 or the amino acids 183+184 of SP707 (SEQ ID NO: 7 of WO 96/23873) to improve the stability of this amylase. WO 96/23873 further discloses to modify the SP707 amylase by substituting M202 with e.g. a leucine to stabilize the molecule towards oxidation. Thus, it is known to modify amylases to improve certain properties.

30

For environmental reasons it has been increasingly important to lower the temperature in washing, dishwashing and/or cleaning processes. However, most enzymes including amylases have a temperature optimum which is above the temperature usually used in low temperature

35

washing. Alpha-amylase is a key enzyme for use in detergent compositions and its use has become increasingly important for removal of starchy stains during laundry washing or dishwashing. Therefore, it is important to find alpha-amylase variants, which retain their wash performance, stain removal effect and/or activity when the temperature is lowered. However, despite the efficiency of current detergent enzyme compositions, there are many stains that are difficult to completely remove. These problems are compounded by the increased use of low (e.g., cold water) wash temperatures and shorter washing cycles. Thus, it is desirable to have amylolytic enzymes that can function under low temperature and at the same time preserve or increase other desirable properties such as specific activity (amylolytic activity), stability and/or wash performance.

It is the object of the present invention to provide alpha-amylase variants which have improved wash performance at low temperature compared to the parent alpha-amylase of SEQ ID NO: 2.

Summary of the Invention

The present invention relates to a variant of a parent alpha-amylase, wherein said variant comprises

(i) substitutions at positions corresponding to positions G109A and G7A of the parent amino acid sequence set forth in SEQ ID NO: 2,

(ii) said variant has at least 90% but less than 100% sequence identity with the parent amino acid sequence set forth in SEQ ID NO: 2, and

(iii) said variant has alpha-amylase activity,

wherein said variant has an improved wash performance at a low temperature between 10-35°C compared to the parent amino acid sequence set forth in SEQ ID NO: 2.

The present invention also relates to a polynucleotide encoding a variant according to the invention, a nucleic acid construct comprising the polynucleotide encoding the variant according to the invention, an expression vector comprising the polynucleotide encoding the variant according to the invention, and a host cell comprising the polynucleotide encoding the variant according to the invention.

The present invention also relates to a method of producing an alpha-amylase variant, comprising (a) cultivating the host cell of the invention under conditions suitable for expression of the variant, and (b) recovering the variant.

5

DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to variants of a parent alpha-amylase, wherein said variant comprises

(i) substitutions at positions corresponding to positions G109A and G7A of the parent amino acid sequence set forth in SEQ ID NO: 2,

10

(ii) said variant has at least 90% but less than 100% sequence identity with the parent amino acid sequence set forth in SEQ ID NO: 2, and

(iii) said variant has alpha-amylase activity,

wherein said variant has an improved wash performance at a low temperature between 10-35°C compared to the parent amino acid sequence set forth in SEQ ID NO: 2.

15

Definitions

Allelic variant: The term “allelic variant” means any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequences. An allelic variant of a polypeptide is a polypeptide encoded by an allelic variant of a gene.

20

The term “alpha-amylase” (alpha-1,4-glucan-4-glucanohydrolase, E.C. 3.2.1.1) constitutes a group of enzymes which catalyzes hydrolysis of starch and other linear and branched 1,4-glycosidic oligo- and polysaccharides. For purposes of the present invention, alpha-amylase activity is determined according to the procedure described in Example section.

25

The term ‘amino acid’ as used herein includes the standard twenty genetically-encoded amino acids and their corresponding stereoisomers in the ‘D’ form (as compared to the natural ‘L’ form), omega-amino acids other naturally-occurring amino acids, unconventional amino acids (e.g. α , α -disubstituted amino acids, N-alkyl amino acids, etc.) and chemically derivatised amino acids. Chemical derivatives of one or more amino acids may be achieved by reaction with a functional side group. Such derivatised molecules include, for example, those molecules in which free amino groups have been derivatised to form amine hydrochlorides, *p*-toluene sulphonyl groups, carboxybenzoxy groups, *t*-butyloxycarbonyl groups, chloroacetyl groups or formyl groups. Free

30

carboxyl groups may be derivatised to form salts, methyl and ethyl esters or other types of esters and hydrazides. Free hydroxyl groups may be derivatised to form O-acyl or O-alkyl derivatives. Also included as chemical derivatives are those peptides which contain naturally occurring amino acid derivatives of the twenty standard amino acids. For example: 4-hydroxyproline may be substituted for proline; 5-hydroxylysine may be substituted for lysine; 3-methylhistidine may be substituted for histidine; homoserine may be substituted for serine and ornithine for lysine. Derivatives also include peptides containing one or more additions or deletions as long as the requisite activity is maintained. Other included modifications are amidation, amino terminal acylation (e.g. acetylation or thioglycolic acid amidation), terminal carboxylamidation (e.g. with ammonia or methylamine), and the like terminal modifications.

When an amino acid is being specifically enumerated, such as 'alanine' or 'Ala' or 'A', the term refers to both L-alanine and D-alanine unless explicitly stated otherwise. Other unconventional amino acids may also be suitable components for polypeptides of the present invention, as long as the desired functional property is retained by the polypeptide. For the peptides shown, each encoded amino acid residue, where appropriate, is represented by a single letter designation, corresponding to the trivial name of the conventional amino acid. In one embodiment, the polypeptides of the invention comprise or consist of L-amino acids.

The term "cDNA" means a DNA molecule that can be prepared by reverse transcription from a mature, spliced, mRNA molecule obtained from a eukaryotic or prokaryotic cell. cDNA lacks intron sequences that may be present in the corresponding genomic DNA. The initial, primary RNA transcript is a precursor to mRNA that is processed through a series of steps, including splicing, before appearing as mature spliced mRNA.

The term "coding sequence" means a polynucleotide, which directly specifies the amino acid sequence of a variant. The boundaries of the coding sequence are generally determined by an open reading frame, which begins with a start codon such as ATG, GTG or TTG and ends with a stop codon such as TAA, TAG, or TGA. The coding sequence may be a genomic DNA, cDNA, synthetic DNA, or a combination thereof.

The term "control sequences" means nucleic acid sequences necessary for expression of a polynucleotide encoding a variant of the present invention. Each control sequence may be native (*i.e.*, from the same gene) or foreign (*i.e.*, from a different gene) to the polynucleotide encoding the variant or native or foreign to each other. Such control sequences include, but are not limited to, a leader, polyadenylation sequence, propeptide sequence, promoter, signal peptide sequence, and transcription terminator. At a minimum, the control sequences include a promoter, and transcriptional and translational stop signals. The control sequences may be provided with linkers for the purpose of introducing specific restriction sites facilitating ligation of the control sequences with the coding region of the polynucleotide encoding a variant.

The terms “enhanced wash performance” or “improved wash performance” mean the ability of the polypeptide of the invention to provide a cleaning effect (e.g. stain removal) in a wash process, such as laundry or dishwashing, is improved compared to that of the parent alpha-amylase of SEQ ID NO:2. Wash performance may be determined using methods well known in the art, such as using an automatic mechanical stress assay (AMSA). It will be appreciated by persons skilled in the art that the enhanced wash performance may be achieved under only some or perhaps all wash conditions, for example at wash temperatures of 20°C or higher (such as at 40°C).

The term “enzyme detergency benefit” used herein, refers to the advantageous effect an enzyme may add to a detergent compared to the same detergent without the enzyme. Important detergency benefits which can be provided by enzymes are stain removal with no or very little visible soils after washing and/or cleaning, prevention or reduction of re-deposition of soils released in the washing process (an effect that also is termed anti-redeposition), restoring fully or partly the whiteness of textiles which originally were white but after repeated use and wash have obtained a greyish or yellowish appearance (an effect that also is termed whitening). Textile care benefits, which are not directly related to catalytic stain removal or prevention of re-deposition of soils, are also important for enzyme detergency benefits. Examples of such textile care benefits are prevention or reduction of dye transfer from one fabric to another fabric or another part of the same fabric (an effect that is also termed dye transfer inhibition or anti-backstaining), removal of protruding or broken fibers from a fabric surface to decrease pilling tendencies or remove already existing pills or fuzz (an effect that also is termed anti-pilling), improvement of the fabric-softness, colour clarification of the fabric and removal of particulate soils which are trapped in the fibers of the fabric or garment. Enzymatic bleaching is a further enzyme detergency benefit where the catalytic activity generally is used to catalyze the formation of bleaching component such as hydrogen peroxide or other peroxides.

The term “expression” includes any step involved in the production of a variant including, but not limited to, transcription, post-transcriptional modification, translation, post-translational modification, and secretion.

The term “expression vector” means a linear or circular DNA molecule that comprises a polynucleotide encoding a variant and is operably linked to control sequences that provide for its expression.

The term “fragment” means a polypeptide having one or more (e.g., several) amino acids absent from the amino and/or carboxyl terminus of the polypeptide of SEQ ID NO:2; wherein the fragment has alpha-amylase activity.

The term “host cell” means any cell type that is susceptible to transformation, transfection, transduction, or the like with a nucleic acid construct or expression vector comprising a

polynucleotide of the present invention. The term “host cell” encompasses any progeny of a parent cell that is not identical to the parent cell due to mutations that occur during replication.

The term “Intensity value” as used herein, refers to the wash performance measurement. It is measured as the brightness expressed as the intensity of the light reflected from the sample when illuminated with white light. When the sample is stained the intensity of the reflected light is lower, than that of a clean sample. Therefore, the intensity of the reflected light can be used to measure wash performance, where a higher intensity value correlates with higher wash performance. Color measurements are made with a professional flatbed scanner (Kodak iQsmart, Kodak) used to capture an image of the washed textile. To extract a value for the light intensity from the scanned images, 24-bit pixel values from the image are converted into values for red, green and blue (RGB). The intensity value (Int) is calculated by adding the RGB values together as vectors and then taking the length of the resulting vector:

$$Int = \sqrt{r^2 + g^2 + b^2}$$

The terms “Delta intensity” or “Delta intensity value” are defined herein as the result of an intensity measurement of a test material, e.g. a swatch CS-28 (Center For Testmaterials BV, P.O. Box 120, 3133 KT Vlaardingen, the Netherlands) or a hard surface. The swatch is measured with a portion of the swatch, washed under identical conditions, as background. The delta intensity is the intensity value of the test material washed with amylase subtracting the intensity value of the test material washed without amylase.

The term “improved property” as used herein, refers to a characteristic associated with a variant that is improved compared to the parent. Such improved properties include, but are not limited to, wash performance, thermal activity, thermostability, and stability under storage conditions and chemical stability. The improved property may be any of those herein defined and described, such as stability.

The term “improved wash performance” is defined herein as displaying an alteration of the wash performance of an amylase of the present invention relative to the wash performance of the amylase of SEQ ID NO: 2. The alteration may e.g. be seen as increased stain removal. Improved wash performance is determined according to Example 1. The wash performance is improved if the Improvement Factor (IF) is above 1.0, preferably above 1.05 in one or more of the conditions listed in example 1 either in model detergent A at 20°C where the alpha-amylase variant concentration is 0.2 mg/L, or in model detergent A at 40°C where the alpha-amylase variant concentration is 0.05 mg/L, or in model detergent J at 20°C where the alpha-amylase variant concentration is 0.2 mg/L, or in model detergent J at 30°C where the alpha-amylase variant

concentration is 0.05 mg/L or in Detergent K at 20°C where the alpha-amylase variant concentration is 0.2 mg/L. The wash conditions are described in the Example section.

The term “wash performance” includes cleaning in general e.g. hard surface cleaning as in dish wash, but also wash performance on textiles such as laundry, and also industrial and institutional cleaning. Improved wash performance may be measured by comparing the delta intensities as described in the definition herein.

The term “isolated” means a substance in a form or environment which does not occur in nature. Non-limiting examples of isolated substances include (1) any non-naturally occurring substance, (2) any substance including, but not limited to, any enzyme, variant, nucleic acid, protein, peptide or cofactor, that is at least partially removed from one or more or all of the naturally occurring constituents with which it is associated in nature; (3) any substance modified by the hand of man relative to that substance found in nature; or (4) any substance modified by increasing the amount of the substance relative to other components with which it is naturally associated (e.g., multiple copies of a gene encoding the substance; use of a stronger promoter than the promoter naturally associated with the gene encoding the substance). An isolated substance may be present in a fermentation broth sample. In one aspect, the present invention relates to an isolated alpha-amylase variant.

Isolated polynucleotide: The term “isolated polynucleotide” means a polynucleotide that is modified by the hand of man. In one aspect, the isolated polynucleotide is at least 1% pure, e.g., at least 5% pure, at least 10% pure, at least 20% pure, at least 40% pure, at least 60% pure, at least 80% pure, at least 90% pure, and at least 95% pure, as determined by agarose electrophoresis. The polynucleotides may be of genomic, cDNA, RNA, semisynthetic, synthetic origin, or any combinations thereof.

Isolated variant: The term “isolated variant” means a variant that is modified by the hand of man. In one aspect, the variant is at least 1% pure, e.g., at least 5% pure, at least 10% pure, at least 20% pure, at least 40% pure, at least 60% pure, at least 80% pure, and at least 90% pure, as determined by SDS-PAGE.

Low temperature: “Low temperature” is a temperature of 5-40°C, preferably 5-35°C, preferably 5-30°C, more preferably 5-25°C, more preferably 5-20°C, most preferably 5-15°C, and in particular 5-10°C. In a preferred embodiment, “Low temperature” is a temperature of 10-35°C, preferably 10-30°C, or 10-25°C, or 10-20°C, or 10-15°C.

The term “mature polypeptide” means a polypeptide in its final form following translation and any post-translational modifications, such as N-terminal processing, C-terminal truncation, glycosylation, phosphorylation, *etc.* It is known in the art that a host cell may produce a mixture of two or more different mature polypeptides (*i.e.*, with a different C-terminal and/or N-terminal amino acid) expressed by the same polynucleotide.

The term “mature polypeptide coding sequence” means a polynucleotide that encodes a mature polypeptide having alpha-amylase activity.

The term “mutant” means a polynucleotide encoding a variant.

5 The term “mutation”, in the context of the polypeptides of the invention, means that one or more amino acids within the reference amino acid sequence (*i.e.* SEQ ID NO:1) are altered by substitution with a different amino acid or by deletion. Additionally, the mutation may correspond to an insertion of one or more extra amino acid(s) within the reference amino acid sequence.

10 The term “nucleic acid construct” means a nucleic acid molecule, either single- or double-stranded, which is isolated from a naturally occurring gene or is modified to contain segments of nucleic acids in a manner that would not otherwise exist in nature or which is synthetic, which comprises one or more control sequences. The term nucleic acid construct is synonymous with the term “expression cassette” when the nucleic acid construct contains the control sequences required for expression of a coding sequence of the present invention.

15 The term “operably linked” means a configuration in which a control sequence is placed at an appropriate position relative to the coding sequence of a polynucleotide such that the control sequence directs expression of the coding sequence.

The term “parent” or “parent alpha-amylase” means an alpha-amylase to which an alteration is made to produce the enzyme variants of the present invention. The parent may be a naturally occurring (wild-type) polypeptide or a variant thereof. For example, the parent may be the alpha-amylase of SEQ ID NO:1 (known as SP722). Alternatively, it may also mean the alpha-amylase of SEQ ID NO: 2.

The relatedness between two amino acid sequences or between two nucleotide sequences is described by the parameter “sequence identity”.

25 For purposes of the present invention, the sequence identity between two amino acid sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, *J. Mol. Biol.* 48: 443-453) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice *et al.*, 2000, *Trends Genet.* 16: 276-277), preferably version 5.0.0 or later. The parameters used may be gap open penalty of 10, gap extension penalty of 0.5, and the EBLOSUM62 (EMBOSS version of BLOSUM62) substitution matrix. The output of Needle labeled “longest identity” (obtained using the –nobrief option) is used as the percent identity and is calculated as follows:

$$(\text{Identical Residues} \times 100) / (\text{Length of Alignment} - \text{Total Number of Gaps in Alignment})$$

35 Alternatively, the parameters used may be gap open penalty of 10, gap extension penalty of 0.5, and the EDNAFULL (EMBOSS version of NCBI NUC4.4) substitution matrix. The output of Needle labeled “longest identity” (obtained using the –nobrief option) is used as the percent identity and is calculated as follows:

(Identical Deoxyribonucleotides x 100)/(Length of Alignment – Total Number of Gaps in Alignment)

Starch removing process: The expression “starch removing process” relates to any kind of process whereby starch is removed (or converted) such as in washing processes where starch is removed from textile e.g. textile cleaning such as laundry. A starch removing process could also be hard surface cleaning such as dish wash or it could be cleaning processes in general such as industrial or institutional cleaning. The expression also comprises other starch removing processes or starch conversion, ethanol production, starch liquefaction, textile desizing, paper and pulp production, beer making and detergents in general.

Substantially pure polynucleotide: The term “substantially pure polynucleotide” means a polynucleotide preparation free of other extraneous or unwanted nucleotides and in a form suitable for use within genetically engineered polypeptide production systems. Thus, a substantially pure polynucleotide contains at most 10%, at most 8%, at most 6%, at most 5%, at most 4%, at most 3%, at most 2%, at most 1%, and at most 0.5% by weight of other polynucleotide material with which it is natively or recombinantly associated. A substantially pure polynucleotide may, however, include naturally occurring 5'- and 3'- untranslated regions, such as promoters and terminators. It is preferred that the substantially pure polynucleotide is at least 90% pure, e.g., at least 92% pure, at least 94% pure, at least 95% pure, at least 96% pure, at least 97% pure, at least 98% pure, at least 99% pure, and at least 99.5% pure by weight. The polynucleotides of the present invention are preferably in a substantially pure form.

Substantially pure variant: The term “substantially pure variant” means a preparation that contains at most 10%, at most 8%, at most 6%, at most 5%, at most 4%, at most 3%, at most 2%, at most 1%, and at most 0.5% by weight of other polypeptide material with which it is natively or recombinantly associated. Preferably, the variant is at least 92% pure, e.g., at least 94% pure, at least 95% pure, at least 96% pure, at least 97% pure, at least 98% pure, at least 99%, at least 99.5% pure, and 100% pure by weight of the total polypeptide material present in the preparation. The variants of the present invention are preferably in a substantially pure form. This can be accomplished, for example, by preparing the variant by well known recombinant methods or by classical purification methods.

The term “subsequence” means a polynucleotide having one or more (e.g., several) nucleotides absent from the 5' and/or 3' end of a mature polypeptide coding sequence; wherein the subsequence encodes a fragment having alpha-amylase activity.

Textile: Textile sample CS-28 (rice starch on cotton) is obtained from Center For Testmaterials BV, P.O. Box 120, 3133 KT Vlaardingen, the Netherlands.

The term “textile care benefits”, as used herein, is defined as not being directly related to catalytic stain removal or prevention of re-deposition of soils, are also important for enzyme detergency benefits. Examples of such textile care benefits are prevention or reduction of dye

transfer from one textile to another textile or another part of the same textile (an effect that is also termed dye transfer inhibition or anti-backstaining), removal of protruding or broken fibers from a textile surface to decrease pilling tendencies or remove already existing pills or fuzz (an effect that also is termed anti-pilling), improvement of the textile-softness, color clarification of the textile and removal of particulate soils which are trapped in the fibers of the textile. Enzymatic bleaching is a further enzyme detergency benefit where the catalytic activity generally is used to catalyze the formation of bleaching component such as hydrogen peroxide or other peroxides or other bleaching species.”

The term “variant” means a polypeptide having alpha-amylase activity comprising a mutation, *i.e.*, a substitution, insertion, and/or deletion, at one or more (*e.g.*, several) positions relative to the ‘parent’ alpha-amylase of SEQ ID NO:1 or SEQ ID NOs: 2, 3, 4, 5, 6, 7, or 8. A substitution means replacement of the amino acid occupying a position with a different amino acid; a deletion means removal of the amino acid occupying a position; and an insertion means adding an amino acid adjacent to and immediately following the amino acid occupying a position. The variants of the present invention have at least 20%, *e.g.*, at least 40%, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 95%, or at least 100% of the alpha-amylase activity of the mature polypeptide of SEQ ID NO: 1 or SEQ ID NO: 2.

The term “wild-type” alpha-amylase means an alpha-amylase expressed by a naturally occurring microorganism, such as a bacterium, yeast, or filamentous fungus found in nature.

Conventions for Designation of Variants

The polypeptides of the invention having alpha-amylase activity correspond to variants of an alpha-amylase derived from *Bacillus*, as shown in SEQ ID NO: 2.

SEQ ID NO: 1

HHNGTNGTMMQYFEWHL PNDGNHWNRLRDDASNLRNRGITAIWIPPAWKGT SQNDVGYGAY
DLYDLGEFNQKGTVRTKYGTRSQLESIAIHALKNNGVQVYGDVVMNHKGGADATENVLAVEVN
PNNRNQEISGDY TIEAWTKFDFPGRGNTYSDFKWRWYHFDGVDWDQSRQFQNRIYKFRGDG
KAWDWEVDSENGNYDYL MYADVMDHPEVVNELRRWGEWYTNLNL DGFRI DAVKHIKYSF
TRDWLTHVRNATGKEMFAVAEFWKNDLGALENYLNKTNWNH SVFDVPLHYNLYNASNSGGN
YDMAKLLNGTVVQKHPMHAVTFVDNHDSQPGESLESFVQEWFKPLAYALILTREQGYPSV FY
GDYYGIPTHSVPAMKAKIDPILEARQNFA YGTQH DYFDH HNIIGWTREGNTTHPNSGLATIMSD
GPGGEKWMYVGQNKAGQVWH DITGNKPGTVTINADGWANFSVNGGSVSIWKR

For the purposes of the present invention, the polypeptide disclosed in SEQ ID NO: 1 is used to determine the corresponding amino acid residue in another alpha-amylase polypeptide.

However, the skilled person would recognize that the sequence of SEQ ID NO: 2 may also be used to determine the corresponding amino acid residue in another alpha-amylase polypeptide. The amino acid sequence of another alpha-amylase is aligned with the mature polypeptide disclosed in SEQ ID NO: 1, and based on the alignment, the amino acid position number corresponding the any amino acid residue in the mature polypeptide disclosed in SEQ ID NO: 1 is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, *J. Mol. Biol.* 48: 443-453) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice *et al.*, 2000, *Trends Genet.* 16: 276-277), preferably version 5.0.0 or later. The parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EBLOSUM62 (EMBOSS version of BLOSUM62) substitution matrix.

Identification of the corresponding amino acid residue in another alpha-amylase can be determined by an alignment of multiple polypeptide sequences using several computer programs including, but not limited to, MUSCLE (multiple sequence comparison by log-expectation; version 3.5 or later; Edgar, 2004, *Nucleic Acids Research* 32: 1792-1797), MAFFT (version 6.857 or later; Katoh and Kuma, 2002, *Nucleic Acids Research* 30: 3059-3066; Katoh *et al.*, 2005, *Nucleic Acids Research* 33: 511-518; Katoh and Toh, 2007, *Bioinformatics* 23: 372-374; Katoh *et al.*, 2009, *Methods in Molecular Biology* 537: 39-64; Katoh and Toh, 2010, *Bioinformatics* 26: 1899-1900), and EMBOSS EMMA employing ClustalW (1.83 or later; Thompson *et al.*, 1994, *Nucleic Acids Research* 22: 4673-4680), using their respective default parameters.

When the other alpha-amylase has diverged from the mature polypeptide of SEQ ID NO: 1 such that traditional sequence-based comparison fails to detect their relationship (Lindahl and Elofsson, 2000, *J. Mol. Biol.* 295: 613-615), other pairwise sequence comparison algorithms can be used. Greater sensitivity in sequence-based searching can be attained using search programs that utilize probabilistic representations of polypeptide families (profiles) to search databases. For example, the PSI-BLAST program generates profiles through an iterative database search process and is capable of detecting remote homologs (Atschul *et al.*, 1997, *Nucleic Acids Res.* 25: 3389-3402). Even greater sensitivity can be achieved if the family or superfamily for the polypeptide has one or more representatives in the protein structure databases. Programs such as GenTHREADER (Jones, 1999, *J. Mol. Biol.* 287: 797-815; McGuffin and Jones, 2003, *Bioinformatics* 19: 874-881) utilize information from a variety of sources (PSI-BLAST, secondary structure prediction, structural alignment profiles, and solvation potentials) as input to a neural network that predicts the structural fold for a query sequence. Similarly, the method of Gough *et al.*, 2000, *J. Mol. Biol.* 313: 903-919, can be used to align a sequence of unknown structure with the superfamily models present in the SCOP database. These alignments can in

turn be used to generate homology models for the polypeptide, and such models can be assessed for accuracy using a variety of tools developed for that purpose.

For proteins of known structure, several tools and resources are available for retrieving and generating structural alignments. *E.g.* the SCOP superfamilies of proteins have been structurally aligned, and those alignments are accessible and downloadable. Two or more protein structures can be aligned using a variety of algorithms such as the distance alignment matrix (Holm and Sander, 1998, *Proteins* 33: 88-96) or combinatorial extension (Shindyalov and Bourne, 1998, *Protein Engineering* 11: 739-747), and implementation of these algorithms can additionally be utilized to query structure databases with a structure of interest in order to discover possible structural homologs (*e.g.*, Holm and Park, 2000, *Bioinformatics* 16: 566-567).

In describing the alpha-amylase variants of the present invention, the nomenclature described below is adapted for ease of reference. The accepted IUPAC single letter or three letter amino acid abbreviation is employed.

Substitutions. For an amino acid substitution, the following nomenclature is used: Original amino acid, position, substituted amino acid. Accordingly, the substitution of *e.g.* threonine at position 226 with alanine is designated as "Thr226Ala" or "T226A". Multiple mutations are separated by addition marks ("+"), *e.g.*, "Gly205Arg + Ser411Phe" or "G205R + S411F", representing substitutions at positions 205 and 411 of glycine (G) with arginine (R) and serine (S) with phenylalanine (F), respectively.

Deletions. For an amino acid deletion, the following nomenclature is used: Original amino acid, position, *. Accordingly, the deletion of glycine at position 181 is designated as "Ser181*" or "S181*". Multiple deletions are separated by addition marks ("+"), *e.g.*, "Ser181* + Thr182*" or "S181* + T182*".

Insertions. For an amino acid insertion, the following nomenclature is used: Original amino acid, position, original amino acid, inserted amino acid. Accordingly, the insertion of lysine after *e.g.* glycine at position 195 is designated "Gly195GlyLys" or "G195GK". An insertion of multiple amino acids is designated [Original amino acid, position, original amino acid, inserted amino acid #1, inserted amino acid #2; etc.]. For example, the insertion of lysine and alanine after glycine at position 195 is indicated as "Gly195GlyLysAla" or "G195GKA".

In such cases the inserted amino acid residue(s) are numbered by the addition of lower case letters to the position number of the amino acid residue preceding the inserted amino acid residue(s). In the above example, the sequence would thus be:

<u>Parent:</u>	<u>Variant:</u>
195	195 195a 195b
G	G - K - A

Multiple modifications. Variants comprising multiple modifications are separated by addition marks (“+”), e.g., “Arg170Tyr+Gly195Glu” or “R170Y+G195E” representing a substitution of arginine and glycine at positions 170 and 195 with tyrosine and glutamic acid, respectively.

Different modifications. Where different alterations can be introduced at a position, the different alterations are separated by a comma, e.g., “Arg170Tyr,Glu” represents a substitution of arginine at position 170 with tyrosine or glutamic acid. Thus, “Tyr167Gly,Ala + Arg170Gly,Ala” designates the following variants:

“Tyr167Gly+Arg170Gly”, “Tyr167Gly+Arg170Ala”, “Tyr167Ala+Arg170Gly”, and “Tyr167Ala+Arg170Ala”.

Parent alpha-amylases

The parent alpha-amylase may be a polypeptide with at least 90% sequence identity with the polypeptide set forth in SEQ ID NO: 2.

In one aspect, the parent alpha-amylase has a sequence identity to the polypeptide of SEQ ID NO: 2 of at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99, or 100%, which has alpha-amylase activity. In one aspect, the amino acid sequence of the parent alpha-amylase differs by no more than ten amino acids, e.g. by five amino acids, by four amino acids, by three amino acids, by two amino acids, and by one amino acid from the polypeptide of SEQ ID NO: 2.

The parent alpha-amylase preferably comprises or consists of the amino acid sequence of SEQ ID NO: 2. In another embodiment, the parent alpha-amylase is an allelic variant of the polypeptide of SEQ ID NO: 2.

The parent may be obtained from microorganisms of any genus. For purposes of the present invention, the term “obtained from” as used herein in connection with a given source shall mean that the parent encoded by a polynucleotide is produced by the source or by a cell in which the polynucleotide from the source has been inserted. In one aspect, the parent is secreted extracellularly.

The parent may be a bacterial alpha-amylase. For example, the parent may be a gram-positive bacterial polypeptide such as a *Bacillus*, *Clostridium*, *Enterococcus*, *Geobacillus*, *Lactobacillus*, *Lactococcus*, *Oceanobacillus*, *Staphylococcus*, *Streptococcus*, or *Streptomyces* alpha-amylase, or a gram-negative bacterial polypeptide such as a *Campylobacter*, *E. coli*, *Flavobacterium*, *Fusobacterium*, *Helicobacter*, *Ilyobacter*, *Neisseria*, *Pseudomonas*, *Salmonella*, or *Ureaplasma* alpha-amylase.

In one aspect, the parent is a *Bacillus alkalophilus*, *Bacillus amyloliquefaciens*, *Bacillus brevis*, *Bacillus circulans*, *Bacillus clausii*, *Bacillus coagulans*, *Bacillus firmus*, *Bacillus lautus*,

Bacillus lentus, *Bacillus licheniformis*, *Bacillus megaterium*, *Bacillus pumilus*, *Bacillus stearothermophilus*, *Bacillus subtilis*, or *Bacillus thuringiensis* alpha-amylase.

In another aspect, the parent is a *Streptococcus equisimilis*, *Streptococcus pyogenes*, *Streptococcus uberis*, or *Streptococcus equi* subsp. *Zooepidemicus* alpha-amylase.

5 In another aspect, the parent is a *Streptomyces achromogenes*, *Streptomyces avermitilis*, *Streptomyces coelicolor*, *Streptomyces griseus*, or *Streptomyces lividans* alpha-amylase.

It will be understood that for the aforementioned species, the invention encompasses both the perfect and imperfect states, and other taxonomic equivalents, e.g., anamorphs, regardless of the species name by which they are known. Those skilled in the art will readily
10 recognize the identity of appropriate equivalents.

Strains of these species are readily accessible to the public in a number of culture collections, such as the American Type Culture Collection (ATCC), Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH (DSMZ), Centraalbureau Voor Schimmelcultures (CBS), and Agricultural Research Service Patent Culture Collection, Northern Regional Research Center
15 (NRRL).

The parent may be identified and obtained from other sources including microorganisms isolated from nature (e.g., soil, composts, water, etc.) or DNA samples obtained directly from natural materials (e.g., soil, composts, water, etc.) using the above-mentioned probes. Techniques for isolating microorganisms and DNA directly from natural habitats are well known
20 in the art. The polynucleotide encoding a parent may then be derived by similarly screening a genomic or cDNA library of another microorganism or mixed DNA sample. Once a polynucleotide encoding a parent has been detected with a probe(s), the polynucleotide may be isolated or cloned by utilizing techniques that are known to those of ordinary skill in the art (see, e.g., Sambrook *et al.*, 1989, *supra*).

25 The parent may be a hybrid polypeptide in which a portion of one polypeptide is fused at the N-terminus or the C-terminus of a portion of another polypeptide.

The parent may also be a fused polypeptide or cleavable fusion polypeptide in which one polypeptide is fused at the N-terminus or the C-terminus of another polypeptide. A fused polypeptide is produced by fusing a polynucleotide encoding one polypeptide to a polynucleotide
30 encoding another polypeptide. Techniques for producing fusion polypeptides are known in the art, and include ligating the coding sequences encoding the polypeptides so that they are in frame and that expression of the fused polypeptide is under control of the same promoter(s) and terminator. Fusion proteins may also be constructed using intein technology in which fusions are created post-translationally (Cooper *et al.*, 1993, *EMBO J.* 12: 2575-2583; Dawson *et al.*, 1994,
35 *Science* 266: 776-779).

A fusion polypeptide can further comprise a cleavage site between the two polypeptides. Upon secretion of the fusion protein, the site is cleaved releasing the two polypeptides. Examples of cleavage sites include, but are not limited to, the sites disclosed in Martin *et al.*, 2003, *J. Ind. Microbiol. Biotechnol.* 3: 568-576; Svetina *et al.*, 2000, *J. Biotechnol.* 76: 245-251; Rasmussen-Wilson *et al.*, 1997, *Appl. Environ. Microbiol.* 63: 3488-3493; Ward *et al.*, 1995, *Biotechnology* 13: 498-503; and Contreras *et al.*, 1991, *Biotechnology* 9: 378-381; Eaton *et al.*, 1986, *Biochemistry* 25: 505-512; Collins-Racie *et al.*, 1995, *Biotechnology* 13: 982-987; Carter *et al.*, 1989, *Proteins: Structure, Function, and Genetics* 6: 240-248; and Stevens, 2003, *Drug Discovery World* 4: 35-48.

10 Variants

The present invention provides variants of a parent alpha-amylase, wherein said variant comprises

- (i) substitutions at positions corresponding to positions G109A and G7A of the parent amino acid sequence set forth in SEQ ID NO: 2,
- (ii) said variant has at least 90% but less than 100% sequence identity with the parent amino acid sequence set forth in SEQ ID NO: 2, and
- (iii) said variant has alpha-amylase activity,

wherein said variant has an improved wash performance at a low temperature between 10-35°C compared to the parent amino acid sequence set forth in SEQ ID NO: 2.

In an embodiment, the variant has at least 95%, such as at least 96%, at least 97%, at least 98%, and at least 99%, but less than 100%, sequence identity with the mature polypeptide of SEQ ID NO: 2.

In one embodiment, the number of modifications in the variants of the present invention is 1 to 20, e.g., 1 to 10 and 1 to 5, such as 1, 2, 3, 4, 5, 6, 7, 8, 9 or 10 modifications.

In one preferred embodiment, the variant comprises modifications in the positions corresponding to the positions of the amino acid sequence set forth in SEQ ID NO: 2, selected from the group consisting of:

H1*+G7A+G109A+N280S+E391A; H1*+G7A+G109A+N280S+W284H+K320A+M323N+E391A;
 H1*+G7A+G109A+N280S+E391A; H1*+G7A+G109A+N280S+K320A+E391A;
 H1*+G7A+G109A+N280S+M323S+E391A; H1*+G7A+G109A+N280S+M323N+E391A;
 H1*+G7A+G109A+N280S+W284F+E391A; H1*+G7A+G109A+N280S+W284R+E391A;
 H1*+G7A+G109A+N280S+K320A+M323S+E391A; H1*+G7A+G109A+W284R+E391A; and
 H1*+G7A+G109A+N280S+K320A+M323N+E391A.

In one embodiment, the invention relates to variants of SEQ ID NO: 2 comprising corresponding to H1*+G7A+G109A+N280S+E391A; and H1*+G7A+G109A+N280S+W284H+M323N+E391A, wherein numbering is according to SEQ ID NO: 1, and wherein the variant has at least 90% sequence identity to SEQ ID NO: 2.

5 In one embodiment, the variant of the invention further comprises a modification in one or more positions selected from the group of 140, 181, 182, 183, 184, 195, 206, 243, 260, 304, and 476. In a particular embodiment, the variant of the invention comprises one or more further modifications selected from the group of W140Y/F, R181*, G182*, D183*, G184*, N195F/Y, I206Y/F, Y243F, E260A/D/C/Q/L/M/F/P/S/W/V/G/H/I/K/N/R/T/Y, G304R/K/E/Q, and
10 G476E/Q/R/K. In one preferred embodiment, the variant according to the invention further comprises substitutions at two, three or four positions selected from the group consisting of G304R, W140YF, E260GHIKNPRTY and G476EQRK. In a more preferred embodiment, the substitutions at the two, three or four positions are selected from the group consisting of G304R, W140Y, E260G and G476K.

15 In one embodiment, the variant of the invention comprises the modifications corresponding to H1*+G7A+G109A+W140Y+D183*+G184*+N195F+I206Y+Y243F+E260G+N280S+G304R+E391A+G476K, and H1*+G7A+G109A+W140Y+D183*+G184*+N195F+I206Y+Y243F+E260G+N280S+W284H+G3
20 04R+M323N+E391A+G476K, wherein numbering is according to SEQ ID NO: 1, the variant has at least 90% sequence identity to SEQ ID NO: 2, and is a variant of SEQ ID NO: 1

Essential amino acids in a parent can be identified according to procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells,
25 1989, *Science* 244: 1081-1085). In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for alpha-amylase activity to identify amino acid residues that are critical to the activity of the molecule. See also, Hilton *et al.*, 1996, *J. Biol. Chem.* 271: 4699-4708. The active site of the alpha-amylase or other biological interaction can also be determined by physical analysis of structure, as determined by
30 such techniques as nuclear magnetic resonance, crystallography, electron diffraction, or photoaffinity labeling, in conjunction with mutation of putative contact site amino acids. See, for example, de Vos *et al.*, 1992, *Science* 255: 306-312; Smith *et al.*, 1992, *J. Mol. Biol.* 224: 899-904; Wlodaver *et al.*, 1992, *FEBS Lett.* 309: 59-64. The identities of essential amino acids can also be inferred from analysis of identities with polypeptides that are related to the parent.

Polynucleotides

The present invention also relates to isolated polynucleotides that encode any of the variants of the present invention.

Nucleic Acid Constructs

The present invention also relates to nucleic acid constructs comprising a polynucleotide encoding a variant of the present invention. In an embodiment, the nucleic acid constructs comprising a polynucleotide encoding a variant of the present invention are operably linked to one or more (several) control sequences that direct the expression of the coding sequence in a suitable host cell under conditions compatible with the control sequences.

A polynucleotide may be manipulated in a variety of ways to provide for expression of a variant. Manipulation of the polynucleotide prior to its insertion into a vector may be desirable or necessary depending on the expression vector. The techniques for modifying polynucleotides utilizing recombinant DNA methods are well known in the art.

The control sequence may be a promoter sequence, which is recognized by a host cell for expression of the polynucleotide. The promoter sequence contains transcriptional control sequences that mediate the expression of the variant. The promoter may be any nucleic acid sequence that shows transcriptional activity in the host cell including mutant, truncated, and hybrid promoters, and may be obtained from genes encoding extracellular or intracellular polypeptides either homologous or heterologous to the host cell.

Examples of suitable promoters for directing the transcription of the nucleic acid constructs of the present invention in a bacterial host cell are the promoters obtained from the *Bacillus amyloliquefaciens* alpha-amylase gene (amyQ), *Bacillus licheniformis* alpha-amylase gene (amyL), *Bacillus licheniformis* penicillinase gene (penP), *Bacillus stearothermophilus* maltogenic amylase gene (amyM), *Bacillus subtilis* levansucrase gene (sacB), *Bacillus subtilis* xylA and xylB genes, *E. coli* lac operon, *Streptomyces coelicolor* agarase gene (dagA), and prokaryotic beta-lactamase gene (Villa-Kamaroff et al., 1978, Proc. Natl. Acad. Sci. USA 75: 3727-3731), as well as the tac promoter (DeBoer et al., 1983, Proc. Natl. Acad. Sci. USA 80: 21-25). Further promoters are described in "Useful proteins from recombinant bacteria" in Gilbert et al., 1980, Scientific American 242: 74-94; and in Sambrook et al., 1989, supra.

Examples of suitable promoters for directing the transcription of the nucleic acid constructs of the present invention in a filamentous fungal host cell are the promoters obtained from the genes for *Aspergillus nidulans* acetamidase, *Aspergillus niger* neutral alpha-amylase, *Aspergillus niger* acid stable alpha-amylase, *Aspergillus niger* or *Aspergillus awamori* glucoamylase (glaA), *Aspergillus oryzae* TAKA amylase, *Aspergillus oryzae* alkaline protease, *Aspergillus oryzae* triose phosphate isomerase, *Fusarium oxysporum* trypsin-like protease (WO 96/00787), *Fusarium venenatum* amyloglucosidase (WO 00/56900), *Fusarium venenatum*

Daria (WO 00/56900), *Fusarium venenatum* Quinn (WO 00/56900), *Rhizomucor miehei* lipase, *Rhizomucor miehei* aspartic proteinase, *Trichoderma reesei* beta-glucosidase, *Trichoderma reesei* cellobiohydrolase I, *Trichoderma reesei* cellobiohydrolase II, *Trichoderma reesei* endoglucanase I, *Trichoderma reesei* endoglucanase II, *Trichoderma reesei* endoglucanase III, *Trichoderma reesei* endoglucanase IV, *Trichoderma reesei* endoglucanase V, *Trichoderma reesei* xylanase I, *Trichoderma reesei* xylanase II, *Trichoderma reesei* beta-xylosidase, as well as the NA2-tpi promoter (a modified promoter including a gene encoding a neutral alpha-amylase in *Aspergilli* in which the untranslated leader has been replaced by an untranslated leader from a gene encoding triose phosphate isomerase in *Aspergilli*; non-limiting examples include modified promoters including the gene encoding neutral alpha-amylase in *Aspergillus niger* in which the untranslated leader has been replaced by an untranslated leader from the gene encoding triose phosphate isomerase in *Aspergillus nidulans* or *Aspergillus oryzae*); and mutant, truncated, and hybrid promoters thereof.

In a yeast host, useful promoters are obtained from the genes for *Saccharomyces cerevisiae* enolase (ENO-1), *Saccharomyces cerevisiae* galactokinase (GAL1), *Saccharomyces cerevisiae* alcohol dehydrogenase/glyceraldehyde-3-phosphate dehydrogenase (ADH1, ADH2/GAP), *Saccharomyces cerevisiae* triose phosphate isomerase (TPI), *Saccharomyces cerevisiae* metallothionein (CUP1), and *Saccharomyces cerevisiae* 3-phosphoglycerate kinase. Other useful promoters for yeast host cells are described by Romanos et al., 1992, *Yeast* 8: 423-488.

The control sequence may also be a suitable transcription terminator sequence, which is recognized by a host cell to terminate transcription. The terminator sequence is operably linked to the 3'-terminus of the polynucleotide encoding the variant. Any terminator that is functional in the host cell may be used.

Preferred terminators for filamentous fungal host cells are obtained from the genes for *Aspergillus nidulans* anthranilate synthase, *Aspergillus niger* alpha-glucosidase, *Aspergillus niger* glucoamylase, *Aspergillus oryzae* TKA amylase, and *Fusarium oxysporum* trypsin-like protease.

Preferred terminators for yeast host cells are obtained from the genes for *Saccharomyces cerevisiae* enolase, *Saccharomyces cerevisiae* cytochrome C (CYC1), and *Saccharomyces cerevisiae* glyceraldehyde-3-phosphate dehydrogenase. Other useful terminators for yeast host cells are described by Romanos et al., 1992, *supra*.

The control sequence may also be a suitable leader sequence, a nontranslated region of an mRNA that is important for translation by the host cell. The leader sequence is operably linked to the 5'-terminus of the polynucleotide encoding the variant. Any leader sequence that is functional in the host cell may be used.

Preferred leaders for filamentous fungal host cells are obtained from the genes for *Aspergillus oryzae* TAKA amylase and *Aspergillus nidulans* triose phosphate isomerase.

Suitable leaders for yeast host cells are obtained from the genes for *Saccharomyces cerevisiae* enolase (ENO-1), *Saccharomyces cerevisiae* 3-phosphoglycerate kinase, *Saccharomyces cerevisiae* alpha-factor, and *Saccharomyces cerevisiae* alcohol dehydrogenase/glyceraldehyde-3-phosphate dehydrogenase (ADH2/GAP).

The control sequence may also be a polyadenylation sequence, a sequence operably linked to the 3'-terminus of the variant-encoding sequence and, when transcribed, is recognized by the host cell as a signal to add polyadenosine residues to transcribed mRNA. Any polyadenylation sequence that is functional in the host cell may be used.

Preferred polyadenylation sequences for filamentous fungal host cells are obtained from the genes for *Aspergillus nidulans* anthranilate synthase, *Aspergillus niger* glucoamylase, *Aspergillus niger* alpha-glucosidase, *Aspergillus oryzae* TAKA amylase, and *Fusarium oxysporum* trypsin-like protease.

Useful polyadenylation sequences for yeast host cells are described by Guo and Sherman, 1995, *Mol. Cellular Biol.* 15: 5983-5990.

The control sequence may also be a signal peptide coding region that encodes a signal peptide linked to the N-terminus of a variant and directs the variant into the cell's secretory pathway. The 5'-end of the coding sequence of the polynucleotide may inherently contain a signal peptide coding region naturally linked in translation reading frame with the segment of the coding region that encodes the variant. Alternatively, the 5'-end of the coding sequence may contain a signal peptide coding region that is foreign to the coding sequence. The foreign signal peptide coding region may be required where the coding sequence does not naturally contain a signal peptide coding region. Alternatively, the foreign signal peptide coding region may simply replace the natural signal peptide coding region in order to enhance secretion of the variant. However, any signal peptide coding region that directs the expressed variant into the secretory pathway of a host cell may be used.

Effective signal peptide coding sequences for bacterial host cells are the signal peptide coding sequences obtained from the genes for *Bacillus* NCIB 11837 maltogenic amylase, *Bacillus licheniformis* subtilisin, *Bacillus licheniformis* beta-lactamase, *Bacillus stearothermophilus* alpha-amylase, *Bacillus stearothermophilus* neutral proteases (nprT, nprS, nprM), and *Bacillus subtilis* prsA. Further signal peptides are described by Simonen and Palva, 1993, *Microbiological Reviews* 57: 109-137.

Effective signal peptide coding sequences for filamentous fungal host cells are the signal peptide coding sequences obtained from the genes for *Aspergillus niger* neutral amylase, *Aspergillus niger* glucoamylase, *Aspergillus oryzae* TAKA amylase, *Humicola insolens* cellulase,

Humicola insolens endoglucanase V, Humicola lanuginosa lipase, and Rhizomucor miehei aspartic proteinase.

Useful signal peptides for yeast host cells are obtained from the genes for Saccharomyces cerevisiae alpha-factor and Saccharomyces cerevisiae invertase. Other useful
5 signal peptide coding sequences are described by Romanos et al., 1992, supra.

The control sequence may also be a propeptide coding region that encodes a propeptide positioned at the N-terminus of a variant. The resultant polypeptide is known as a proenzyme or propolypeptide (or a zymogen in some cases). A propolypeptide is generally inactive and can be converted to an active polypeptide by catalytic or autocatalytic cleavage of the propeptide from
10 the propolypeptide. The propeptide coding region may be obtained from the genes for Bacillus subtilis alkaline protease (aprE), Bacillus subtilis neutral protease (nprT), Myceliophthora thermophila laccase (WO 95/33836), Rhizomucor miehei aspartic proteinase, and Saccharomyces cerevisiae alpha-factor.

Where both signal peptide and propeptide regions are present at the N-terminus of a
15 variant, the propeptide region is positioned next to the N-terminus of the variant and the signal peptide region is positioned next to the N-terminus of the propeptide region.

It may also be desirable to add regulatory sequences that allow the regulation of the expression of the variant relative to the growth of the host cell. Examples of regulatory systems are those that cause the expression of the gene to be turned on or off in response to a chemical
20 or physical stimulus, including the presence of a regulatory compound. Regulatory systems in prokaryotic systems include the lac, tac, and trp operator systems. In yeast, the ADH2 system or GAL1 system may be used. In filamentous fungi, the Aspergillus niger glucoamylase promoter, Aspergillus oryzae TKA alpha-amylase promoter, and Aspergillus oryzae glucoamylase promoter may be used. Other examples of regulatory sequences are those that allow for gene
25 amplification. In eukaryotic systems, these regulatory sequences include the dihydrofolate reductase gene that is amplified in the presence of methotrexate, and the metallothionein genes that are amplified with heavy metals. In these cases, the polynucleotide encoding the variant would be operably linked with the regulatory sequence.

30 Expression Vectors

The present invention also relates to recombinant expression vectors comprising a polynucleotide of the present invention. In an embodiment, the recombinant expression vectors comprising a polynucleotide of the present invention comprises a promoter, and transcriptional and translational stop signals. The various nucleotide and control sequences may be joined
35 together to produce a recombinant expression vector that may include one or more (several) convenient restriction sites to allow for insertion or substitution of the polynucleotide encoding the

variant at such sites. Alternatively, the polynucleotide may be expressed by inserting the polynucleotide or a nucleic acid construct comprising the polynucleotide into an appropriate vector for expression. In creating the expression vector, the coding sequence is located in the vector so that the coding sequence is operably linked with the appropriate control sequences for expression.

5 The recombinant expression vector may be any vector (e.g., a plasmid or virus) that can be conveniently subjected to recombinant DNA procedures and can bring about the expression of the polynucleotide. The choice of the vector will typically depend on the compatibility of the vector with the host cell into which the vector is to be introduced. The vector may be a linear or closed circular plasmid.

10 The vector may be an autonomously replicating vector, *i.e.*, a vector that exists as an extrachromosomal entity, the replication of which is independent of chromosomal replication, e.g., a plasmid, an extrachromosomal element, a minichromosome, or an artificial chromosome. The vector may contain any means for assuring self-replication. Alternatively, the vector may be one that, when introduced into the host cell, is integrated into the genome and replicated together with
15 the chromosome(s) into which it has been integrated. Furthermore, a single vector or plasmid or two or more vectors or plasmids that together contain the total DNA to be introduced into the genome of the host cell, or a transposon, may be used.

The vector preferably comprises one or more (several) selectable markers that permit easy selection of transformed, transfected, transduced, or the like cells. A selectable marker is a
20 gene the product of which provides for biocide or viral resistance, resistance to heavy metals, prototrophy to auxotrophs, and the like.

Examples of bacterial selectable markers are the *dal* genes from *Bacillus licheniformis* or *Bacillus subtilis*, or markers that confer antibiotic resistance such as ampicillin, chloramphenicol, kanamycin, or tetracycline resistance. Suitable markers for yeast host cells are ADE2, HIS3,
25 LEU2, LYS2, MET3, TRP1, and URA3.

The vector preferably comprises an element(s) that permits integration of the vector into the host cell's genome or autonomous replication of the vector in the cell independent of the genome.

For integration into the host cell genome, the vector may rely on the polynucleotide's
30 sequence encoding the variant or any other element of the vector for integration into the genome by homologous or non-homologous recombination. Alternatively, the vector may comprise additional nucleotide sequences for directing integration by homologous recombination into the genome of the host cell at a precise location(s) in the chromosome(s). To increase the likelihood of integration at a precise location, the integrational elements should contain a sufficient number
35 of nucleic acids, such as 100 to 10,000 base pairs, 400 to 10,000 base pairs, and 800 to 10,000 base pairs, which have a high degree of identity to the corresponding target sequence to enhance

the probability of homologous recombination. The integrational elements may be any sequence that is homologous with the target sequence in the genome of the host cell. Furthermore, the integrational elements may be non-encoding or encoding nucleotide sequences. On the other hand, the vector may be integrated into the genome of the host cell by non-homologous recombination.

For autonomous replication, the vector may further comprise an origin of replication enabling the vector to replicate autonomously in the host cell in question. The origin of replication may be any plasmid replicator mediating autonomous replication that functions in a cell. The term "origin of replication" or "plasmid replicator" means a nucleotide sequence that enables a plasmid or vector to replicate *in vivo*.

More than one copy of a polynucleotide of the present invention may be inserted into the host cell to increase production of a variant. An increase in the copy number of the polynucleotide can be obtained by integrating at least one additional copy of the sequence into the host cell genome or by including an amplifiable selectable marker gene with the polynucleotide where cells containing amplified copies of the selectable marker gene, and thereby additional copies of the polynucleotide, can be selected for by cultivating the cells in the presence of the appropriate selectable agent.

The procedures used to ligate the elements described above to construct the recombinant expression vectors of the present invention are well known to one skilled in the art (see, e.g., Sambrook *et al.*, 1989, *supra*) to obtain substantially pure variants.

Host Cells

The present invention also relates to recombinant host cells, comprising a polynucleotide of the present invention. In an embodiment, the recombinant host cells, comprising a polynucleotide of the present invention, are operably linked to one or more (several) control sequences that direct the production of a variant of the present invention. A construct or vector comprising a polynucleotide is introduced into a host cell so that the construct or vector is maintained as a chromosomal integrant or as a self-replicating extra-chromosomal vector as described earlier. The term "host cell" encompasses any progeny of a parent cell that is not identical to the parent cell due to mutations that occur during replication. The choice of a host cell will to a large extent depend upon the gene encoding the variant and its source.

The host cell may be any cell useful in the recombinant production of a variant, e.g., a prokaryote or a eukaryote.

The prokaryotic host cell may be any gram-positive or gram-negative bacterium. Gram-positive bacteria include, but are not limited to, *Bacillus*, *Clostridium*, *Enterococcus*, *Geobacillus*, *Lactobacillus*, *Lactococcus*, *Oceanobacillus*, *Staphylococcus*, *Streptococcus*, and *Streptomyces*.

Gram-negative bacteria include, but are not limited to, *Campylobacter*, *E. coli*, *Flavobacterium*, *Fusobacterium*, *Helicobacter*, *Ilyobacter*, *Neisseria*, *Pseudomonas*, *Salmonella*, and *Ureaplasma*.

The bacterial host cell may be any *Bacillus* cell, including, but not limited to, *Bacillus alkalophilus*, *Bacillus amyloliquefaciens*, *Bacillus brevis*, *Bacillus circulans*, *Bacillus clausii*,
 5 *Bacillus coagulans*, *Bacillus firmus*, *Bacillus lautus*, *Bacillus lentus*, *Bacillus licheniformis*, *Bacillus megaterium*, *Bacillus pumilus*, *Bacillus stearothermophilus*, *Bacillus subtilis*, and *Bacillus thuringiensis* cells.

The bacterial host cell may also be any *Streptococcus* cell, including, but not limited to, *Streptococcus equisimilis*, *Streptococcus pyogenes*, *Streptococcus uberis*, and *Streptococcus equi* subsp. *Zooepidemicus* cells.
 10

The bacterial host cell may also be any *Streptomyces* cell, including, but not limited to, *Streptomyces achromogenes*, *Streptomyces avermitilis*, *Streptomyces coelicolor*, *Streptomyces griseus*, and *Streptomyces lividans* cells.

The introduction of DNA into a *Bacillus* cell may, for instance, be effected by protoplast
 15 transformation (see, e.g., Chang and Cohen, 1979, *Mol. Gen. Genet.* 168: 111-115), by using competent cells (see, e.g., Young and Spizizen, 1961, *J. Bacteriol.* 81: 823-829, or Dubnau and Davidoff-Abelson, 1971, *J. Mol. Biol.* 56: 209-221), by electroporation (see, e.g., Shigekawa and Dower, 1988, *Biotechniques* 6: 742-751), or by conjugation (see, e.g., Koehler and Thorne, 1987, *J. Bacteriol.* 169: 5271-5278). The introduction of DNA into an *E. coli* cell may, for instance, be
 20 effected by protoplast transformation (see, e.g., Hanahan, 1983, *J. Mol. Biol.* 166: 557-580) or electroporation (see, e.g., Dower *et al.*, 1988, *Nucleic Acids Res.* 16: 6127-6145). The introduction of DNA into a *Streptomyces* cell may, for instance, be effected by protoplast transformation and electroporation (see, e.g., Gong *et al.*, 2004, *Folia Microbiol. (Praha)* 49: 399-405), by conjugation (see, e.g., Mazodier *et al.*, 1989, *J. Bacteriol.* 171: 3583-3585), or by transduction (see, e.g.,
 25 Burke *et al.*, 2001, *Proc. Natl. Acad. Sci. USA* 98: 6289-6294). The introduction of DNA into a *Pseudomonas* cell may, for instance, be effected by electroporation (see, e.g., Choi *et al.*, 2006, *J. Microbiol. Methods* 64: 391-397) or by conjugation (see, e.g., Pinedo and Smets, 2005, *Appl. Environ. Microbiol.* 71: 51-57). The introduction of DNA into a *Streptococcus* cell may, for instance, be effected by natural competence (see, e.g., Perry and Kuramitsu, 1981, *Infect. Immun.* 32:
 30 1295-1297), by protoplast transformation (see, e.g., Catt and Jollick, 1991, *Microbios* 68: 189-2070, by electroporation (see, e.g., Buckley *et al.*, 1999, *Appl. Environ. Microbiol.* 65: 3800-3804) or by conjugation (see, e.g., Clewell, 1981, *Microbiol. Rev.* 45: 409-436). However, any method known in the art for introducing DNA into a host cell can be used.

The host cell may also be a eukaryote, such as a mammalian, insect, plant, or fungal
 35 cell.

Fungal cells may be transformed by a process involving protoplast formation, transformation of the protoplasts, and regeneration of the cell wall in a manner known *per se*. Suitable procedures for transformation of *Aspergillus* and *Trichoderma* host cells are described in EP 238023 and Yelton *et al.*, 1984, *Proc. Natl. Acad. Sci. USA* 81: 1470-1474. Suitable methods for transforming *Fusarium* species are described by Malardier *et al.*, 1989, *Gene* 78: 147-156, and WO 96/00787. Yeast may be transformed using the procedures described by Becker and Guarente, *In* Abelson, J.N. and Simon, M.I., editors, *Guide to Yeast Genetics and Molecular Biology, Methods in Enzymology*, Volume 194, pp 182-187, Academic Press, Inc., New York; Ito *et al.*, 1983, *J. Bacteriol.* 153: 163; and Hinnen *et al.*, 1978, *Proc. Natl. Acad. Sci. USA* 75: 1920.

Methods of Production

The present invention also relates to methods of producing a variant, comprising: (a) cultivating a host cell of the present invention under conditions suitable for the expression of the variant; and (b) recovering the variant. Accordingly, the present invention relates to methods of producing a variant, comprising (a) cultivating a host cell comprising an expression vector or a polynucleotide encoding variant comprising a modification at one or more positions corresponding to positions 109, 1, 7, 280, 284, 320, 323, and 391 of the amino acid sequence set forth in SEQ ID NO: 1, and optionally in one or more positions corresponding to positions 140, 181, 182, 183, 184, 195, 206, 243, 260, 304, and 476 of the amino acid sequence as set forth in SEQ ID NO: 1, under conditions suitable for the expression of the variant; and (b) recovering the variant.

In one aspect, the present invention relates to a method of obtaining an alpha-amylase variant, comprising introducing into a parent alpha-amylase a modification at one or more positions corresponding to positions selected from the group consisting of 109, 7, 1, 391, 280, 284, 320, and 323 of the amino acid sequence set forth in SEQ ID NO: 1, and optionally in one or more positions corresponding to positions selected from the group consisting of 140, 181, 182, 183, 184, 195, 206, 243, 260, 304, and 476 of the amino acid sequence as set forth in SEQ ID NO: 1, wherein each modification is independently a substitution or deletion, and the variant has alpha-amylase activity; and recovering the variant.

The host cells are cultivated in a nutrient medium suitable for production of the variant using methods known in the art. For example, the cell may be cultivated by shake flask cultivation, or small-scale or large-scale fermentation (including continuous, batch, fed-batch, or solid state fermentations) in laboratory or industrial fermentors performed in a suitable medium and under conditions allowing the polypeptide to be expressed and/or isolated. The cultivation takes place in a suitable nutrient medium comprising carbon and nitrogen sources and inorganic salts, using procedures known in the art. Suitable media are available from commercial suppliers or may be

prepared according to published compositions (e.g., in catalogues of the American Type Culture Collection). If the variant is secreted into the nutrient medium, the variant can be recovered directly from the medium. If the variant is not secreted, it can be recovered from cell lysates.

The variant may be detected using methods known in the art that are specific for the variants. These detection methods may include use of specific antibodies, formation of an enzyme product, or disappearance of an enzyme substrate. For example, an enzyme assay may be used to determine the activity of the variant.

The variant may be recovered by methods known in the art. For example, the variant may be recovered from the nutrient medium by conventional procedures including, but not limited to, collection, centrifugation, filtration, extraction, spray-drying, evaporation, or precipitation.

The variant may be purified by a variety of procedures known in the art including, but not limited to, chromatography (e.g., ion exchange, affinity, hydrophobic, chromatofocusing, and size exclusion), electrophoretic procedures (e.g., preparative isoelectric focusing), differential solubility (e.g., ammonium sulfate precipitation), SDS-PAGE, or extraction (see, e.g., *Protein Purification*, J.-C. Janson and Lars Ryden, editors, VCH Publishers, New York, 1989) to obtain substantially pure variants.

In an alternative aspect, the variant is not recovered, but rather a host cell of the present invention expressing a variant is used as a source of the variant.

Compositions

A variant of the invention may be comprised in compositions enriched in such a variant. The term "enriched" means that the alpha-amylase activity of the composition has been increased, e.g., with an enrichment factor of 1.1.

The composition may comprise a variant of the invention as the major enzymatic component, e.g., a mono-component composition. Alternatively, the composition may comprise multiple enzymatic activities, such as an aminopeptidase, amylase, carbohydrase, carboxypeptidase, catalase, cellulase, chitinase, cutinase, cyclodextrin glycosyltransferase, deoxyribonuclease, esterase, alpha-galactosidase, beta-galactosidase, glucoamylase, alpha-glucosidase, beta-glucosidase, haloperoxidase, invertase, laccase, lipase, mannosidase, oxidase, pectinolytic enzyme, peptidoglutaminase, peroxidase, phytase, polyphenoloxidase, proteolytic enzyme, ribonuclease, transglutaminase, or xylanase. The additional enzyme(s) may be produced, for example, by a microorganism belonging to the genus *Bacillus*, e.g. *Bacillus licheniformis* and *Bacillus subtilis*, or the genus *Aspergillus*, e.g., *Aspergillus aculeatus*, *Aspergillus awamori*, *Aspergillus foetidus*, *Aspergillus fumigatus*, *Aspergillus japonicus*, *Aspergillus nidulans*, *Aspergillus niger*, or *Aspergillus oryzae*; *Fusarium*, e.g., *Fusarium bactridioides*, *Fusarium cerealis*, *Fusarium crookwellense*, *Fusarium culmorum*, *Fusarium*

graminearum, *Fusarium graminum*, *Fusarium heterosporum*, *Fusarium negundi*, *Fusarium oxysporum*, *Fusarium reticulatum*, *Fusarium roseum*, *Fusarium sambucinum*, *Fusarium sarcochroum*, *Fusarium sulphureum*, *Fusarium toruloseum*, *Fusarium trichothecioides*, or *Fusarium venenatum*; *Humicola*, e.g., *Humicola insolens* or *Humicola lanuginosa*; or *Trichoderma*, e.g., *Trichoderma harzianum*, *Trichoderma koningii*, *Trichoderma longibrachiatum*, *Trichoderma reesei*, or *Trichoderma viride* or any other host cell herein described.

The compositions may be prepared in accordance with methods known in the art and may be in the form of a liquid or a dry composition. For instance, the composition may be in the form of a granulate or a microgranulate. The variant may be stabilized in accordance with methods known in the art.

The above alpha-amylase variants of the invention may typically be a component in a cleaning composition, such as a detergent composition, e.g., a laundry detergent composition or a dishwashing detergent composition. Especially preferred is a liquid laundry detergent composition.

Such cleaning compositions comprise a cleaning/detergent adjunct, preferably a mixture of components. Typically, the cleaning adjunct will be present in the composition in an amount from 0.001 to 99.9 wt%, more typically from 0.01 to 80 wt% cleaning adjunct.

The composition may comprises one or more surfactants, which may be non-ionic including semi-polar and/or anionic and/or cationic and/or zwitterionic and/or ampholytic and/or semi-polar nonionic and/or mixtures thereof. The surfactants are typically present at a level of from 0.1% to 60% by weight or from 0.5 to 50 wt% or 1 to 40 wt% of the composition.

Uses

The alpha-amylase variants of the invention are useful in detergent compositions, laundry washing, dishwashing and/or cleaning processes at low temperature.

Method of Use

Alpha-amylase variants of the invention may be used in methods for cleaning and/or treating a situs inter alia a surface or fabric. In one aspect, such method comprises the steps of optionally washing and/or rinsing said surface or fabric, contacting said surface or fabric with any consumer product disclosed in this specification then optionally washing and/or rinsing said surface or fabric.

As used herein, washing includes but is not limited to, scrubbing, and mechanical agitation. Drying of such surfaces or fabrics may be accomplished by any one of the common means employed either in domestic or industrial settings. Such means include but are not limited to

forced air or still air drying at ambient or elevated temperatures at pressures between 5 and 0.01 atmospheres in the presence or absence of electromagnetic radiation, including sunlight, infrared, ultraviolet and microwave irradiation. In one aspect, said drying may be accomplished at temperatures above ambient by employing an iron wherein, for example, said fabric may be in direct contact with said iron for relatively short or even extended periods of time and wherein pressure may be exerted beyond that otherwise normally present due to gravitational force. In another aspect, said drying may be accomplished at temperatures above ambient by employing a dryer. Apparatus for drying fabric is well known and it is frequently referred to as a clothes dryer. In addition to clothes such appliances are used to dry many other items including towels, sheets, pillowcases, diapers and so forth and such equipment has been accepted as a standard convenience in many nations of the world substantially replacing the use of clothes lines for drying of fabric. Most dryers in use today use heated air which is passed over and or through the fabric as it is tumbled within the dryer. The air may be heated, for example, either electronically, via gas flame, or even with microwave radiation. Such air may be heated from about 15°C to about 400°C, from about 25°C to about 200°C, from about 35°C to about 100°C, or even from about 40°C to about 85°C and used in the dryer to dry a surface and/or a fabric. As will be appreciated by one skilled in the art, the cleaning compositions of the present invention are ideally suited for use in laundry applications. Accordingly, the present invention includes a method for laundering a fabric. The method comprises the steps of contacting a fabric to be laundered with a said cleaning laundry solution comprising at least one embodiment of Applicants' cleaning composition, cleaning additive or mixture thereof. The fabric may comprise most any fabric capable of being laundered in normal consumer or institutional use conditions. The solution preferably has a pH of from about 8 to about 10.5. The compositions may be employed at concentrations of from about 500 ppm to about 15,000 ppm in solution. The water temperatures typically range from about 5 °C to about 90 °C. The water to fabric ratio is typically from about 1:1 to about 30:1.

In the following is exemplified detergent compositions.

Table 1: Examples of a granular laundry detergent compositions designed for hand washing or top-loading washing machines. Each granular laundry detergent composition is numbered 1 to 6 in order to differentiate between the different compositions.

	1 (wt %)	2 (wt %)	3 (wt %)	4 (wt %)	5 (wt %)	6 (wt %)
Linear alkylbenzenesulfonate	20	22	20	15	20	20
C ₁₂₋₁₄ Dimethylhydroxyethyl ammonium chloride	0.7	0.2	1	0.6	0.0	0

AE3S	0.9	1	0.9	0.0	0.5	0.9
AE7	0.0	0.0	0.0	1	0.0	3
Sodium tripolyphosphate	5	0.0	4	9	2	0.0
Zeolite A	0.0	1	0.0	1	4	1
1.6R Silicate (SiO ₂ :Na ₂ O at ratio 1.6:1)	7	5	2	3	3	5
Sodium carbonate	25	20	25	17	18	19
Polyacrylate MW 4500	1	0.6	1	1	1.5	1
Random graft copolymer ¹	0.1	0.2	0.0	0.0	0.0	0.0
Carboxymethyl cellulose	1	0.3	1	1	1	1
Protease (Savinase®, 32.89 mg active/g)	0.1	0.1	0.1	0.1		0.1
Lipase - Lipex® (18 mg active /g)	0.03	0.07	0.3	0.1	0.07	0.4
*Amylase of the present invention (mg active)	0.63	1.0	2.0	0.44	0.88	0.3
Fluorescent Brightener 1	0.06	0.0	0.06	0.18	0.06	0.06
Fluorescent Brightener 2	0.1	0.06	0.1	0.0	0.1	0.1
DTPA	0.6	0.8	0.6	0.25	0.6	0.6
MgSO ₄	1	1	1	0.5	1	1
Sodium Percarbonate	0.0	5.2	0.1	0.0	0.0	0.0
Sodium Perborate Monohydrate	4.4	0.0	3.85	2.09	0.78	3.63
NOBS	1.9	0.0	1.66	0.0	0.33	0.75
TAED	0.58	1.2	0.51	0.0	0.015	0.28
Sulphonated zinc phthalocyanine	0.0030	0.0	0.0012	0.0030	0.0021	0.0
S-ACMC	0.1	0.0	0.0	0.0	0.06	0.0
Direct Violet 9	0.0	0.0	0.0003	0.0005	0.0003	0.0
Acid Blue 29	0.0	0.0	0.0	0.0	0.0	0.0003
Sulfate/Moisture	Balance					

*Amylase of the present invention is shown as mgs of active enzyme per 100g of detergent.

Table 2: Other examples (numbered 7 to 12 below in table 2) of granular laundry detergent

compositions designed for front-loading automatic washing machines.

	7 (wt%)	8 (wt%)	9 (wt%)	10 (wt%)	11 (wt%)	12 (wt%)
Linear alkylbenzenesulfonate	8	7.1	7	6.5	7.5	7.5
AE3S	0	4.8	0	5.2	4	4
C12-14 Alkylsulfate	1	0	1	0	0	0
AE7	2.2	0	3.2	0	0	0
C ₁₀₋₁₂ Dimethyl hydroxyethylammonium chloride	0.75	0.94	0.98	0.98	0	0
Crystalline layered silicate (δ - Na ₂ Si ₂ O ₅)	4.1	0	4.8	0	0	0
Zeolite A	5	0	5	0	2	2
Citric Acid	3	5	3	4	2.5	3
Sodium Carbonate	15	20	14	20	23	23
Silicate 2R (SiO ₂ :Na ₂ O at ratio 2:1)	0.08	0	0.11	0	0	0
Soil release agent	0.75	0.72	0.71	0.72	0	0
Acrylic Acid/Maleic Acid Copolymer	1.1	3.7	1.0	3.7	2.6	3.8
Carboxymethylcellulose	0.15	1.4	0.2	1.4	1	0.5
Protease - Purafect® (84 mg active/g)	0.2	0.2	0.3	0.15	0.12	0.13
Lipase - Lipex® (18.00 mg active/g)	0.05	0.15	0.1	0	0	0
Cellulase - Celluclean™ (15.6 mg active/g)	0	0	0	0	0.1	0.1
*Amylase of the present invention (mg active)	4.0	2.0	1.0	0.7	6.0	3.0
Amylase ⁴	0.15	0.04	0.03	-	0.01	0.16
TAED	3.6	4.0	3.6	4.0	2.2	1.4
Percarbonate	13	13.2	13	13.2	16	14
Na salt of Ethylenediamine-N,N'- disuccinic acid, (S,S) isomer (EDDS)	0.2	0.2	0.2	0.2	0.2	0.2
Hydroxyethane di phosphonate	0.2	0.2	0.2	0.2	0.2	0.2

(HEDP)						
MgSO ₄	0.42	0.42	0.42	0.42	0.4	0.4
Perfume	0.5	0.6	0.5	0.6	0.6	0.6
Suds suppressor agglomerate	0.05	0.1	0.05	0.1	0.06	0.05
Soap	0.45	0.45	0.45	0.45	0	0
Sulphonated zinc phthalocyanine (active)	0.0007	0.0012	0.0007	0	0	0
S-ACMC	0.01	0.01	0	0.01	0	0
Direct Violet 9 (active)	0	0	0.0001	0.0001	0	0
Sulfate/ Water & Miscellaneous	Balance					

*Amylase of the present invention is shown as mgs of active enzyme per 100g of detergent.

Table 3: Examples of Heavy Duty Liquid laundry detergent compositions (numbered 13 to 18)

	13 (wt%)	14 (wt%)	15 (wt%)	16 (wt%)	17 (wt%)	18 (wt%)
C ₁₂₋₁₅ Alkylethoxy(1.8)sulfate	14.7	11.6		16.31		17.29
C _{11.8} Alkylbenzene sulfonate	4.3	11.6	8.3	7.73	11.7	7.73
C ₁₆₋₁₇ Branched alkyl sulfate	1.7	1.29		3.09		3.3
C ₁₂₋₁₄ Alkyl -9-ethoxylate	0.9	1.07		1.31		1.31
C ₁₂ dimethylamine oxide	0.6	0.64		1.03		1.03
Citric acid	3.5	0.65	3	0.66	2.27	0.67
C ₁₂₋₁₈ fatty acid	1.5	2.32	3.6	1.52	0.82	1.52
Sodium Borate (Borax)	2.5	2.46	1.2	2.53		2.53
Sodium C ₁₂₋₁₄ alkyl ethoxy 3 sulfate			2.9		3.9	
C ₁₄₋₁₅ alkyl 7-ethoxylate			4.2		1.9	
C ₁₂₋₁₄ Alkyl -7-ethoxylate			1.7		0.5	
Ca chloride dihydrate					0.045	
Ca formate	0.09	0.09		0.09		0.09
A compound: bis((C ₂ H ₅ O)(C ₂ H ₄ O) _n)(CH ₃)-N ⁺ - C _x H _{2x} -N ⁺ -(CH ₃)- bis((C ₂ H ₅ O)(C ₂ H ₄ O) _n); n is 20 to 30; x is 3 to 8, optionally sulphated or sulphonated			1.2		0.66	

Random graft co-polymer ¹		1.46	0.5		0.83	
Ethoxylated Polyethylenimine ²	1.5	1.29		1.44		1.44
Diethylene triamine pentaacetic acid	0.34	0.64		0.34		0.34
Diethylene triamine penta (methylene phosphonic acid)			0.3		0.3	
1-hydroxyethylidene-1,1-diphosphonic acid					0.18	
Dihydroxybenzene-3,5-disulfonic acid disodium salt hydrate						0.19
Tinopal AMS-GX		0.06				0.29
Tinopal CBS-X	0.2	0.17		0.29		
Tinopal TAS-X B36					0.091	
Amphiphilic alkoxyated grease cleaning polymer ³	1.28	1	0.4	1.93		1.93
CHEC			0.2			
Ethanol	2	1.58	1.6	5.4	1.2	3.57
Propylene Glycol	3.9	3.59	1.3	4.3		3.8
Diethylene glycol	1.05	1.54		1.15		1.15
Polyethylene glycol	0.06	0.04		0.1		0.1
*Amylase of the present invention (mg active)	15.0	10.0	5.0	8.0	4.25	11.7
Amylase ⁴	0.01	0.1	0.15	0.12	-	0.05
Monoethanolamine	3.05	2.41	0.4	1.26	0.31	1.13
NaOH	2.44	1.8		3.01	3.84	0.24
Sodium Cumene Sulphonate			1		0.95	
Sodium Formate		0.11		0.09	0.2	0.12
Water, Aesthetics (Dyes, perfumes) and Minors (Enzymes including lipase, protease, additional amylase each at 0.2% active protein, solvents, structurants)	balance					

¹ Random graft copolymer is a polyvinyl acetate grafted polyethylene oxide copolymer having a

polyethylene oxide backbone and multiple polyvinyl acetate side chains. The molecular weight of the polyethylene oxide backbone is about 6000 and the weight ratio of the polyethylene oxide to polyvinyl acetate is about 40 to 60 and no more than 1 grafting point per 50 ethylene oxide units.

5 ² Polyethylenimine (MW = 600) with 20 ethoxylate groups per -NH.

³ Amphiphilic alkoxyated grease cleaning polymer is a polyethylenimine (MW = 600) with 24 ethoxylate groups per -NH and 16 propoxylate groups per -NH

*Amylase of the present invention is shown as mgs of active enzyme per 100g of detergent.

10 Table 4: Examples of Heavy Duty Liquid laundry detergent composition (numbered 19 to 21)

	19 (wt%)	20 (wt%)	21 (wt%)
Sodium Alkylbenzene sulfonate	21.0	10.2	3.53
C ₁₂₋₁₈ Alkyl 1.5-9-ethoxylate	18.0	6.32	0.88
Branched Alkyl Sulfate			2.44
Sodium Alkyl ethoxy 1-3 sulfate		1.17	14.81
Citric Acid		3.14	2.05
C ₁₂ Dimethylamine oxide			0.56
C ₁₂₋₁₈ Fatty acid	15.0	2.59	1.48
Protease (Purafect Prime®, 40.6mg active/g)	1.5	0.52	1.64
Mannanase (Mannaway®, 11mg active/g)	0.1	0.06	
Xyloglucanase (Whitezyme®, 20mg active/g)	0.2	0.06	
Lipase (ILipex)	0.1	0.2	0.05
*Amylase of the present invention (mg active)	5.9	2.3	12.8
bis((C ₂ H ₅ O)(C ₂ H ₄ O) _n)(CH ₃)-N ⁺ -C _x H _{2x} -N ⁺ -(CH ₃)-bis((C ₂ H ₅ O)(C ₂ H ₄ O) _n), wherein n = from 20 to 30, and x = from 3 to 8, optionally sulphated or sulphonated	2.0	0.63	
Random graft co-polymer ¹		1.07	
Ethoxylated Polyethylenimine ²	0.8		1.51
Amphiphilic alkoxyated polymer ³			
Amylase ⁴			
Phosphonated chelant	0.8	0.41	0.53
Hydrotrope		0.93	

Brightener	0.2	0.09	0.19
Ethoxylated thiophene Hueing Dye	0.004		
Minors: dyes, perfume, perfume micro capsules, enzymes, enzyme stabilizers, solvents, structurants, pH modifying agents	Balance	Balance	Balance

*Amylase of the present invention is shown as mgs of active enzyme per 100g of detergent.

**Based on total cleaning and/or treatment composition weight, a total of no more than 7% water.

Raw Materials and Notes For Composition Examples 1-21

- 5 Linear alkylbenzenesulfonate having an average aliphatic carbon chain length $C_{11}-C_{18}$
 C_{12-18} Dimethylhydroxyethyl ammonium chloride
 AE3S is C_{12-15} alkyl ethoxy (3) sulfate
 AE7 is C_{12-15} alcohol ethoxylate, with an average degree of ethoxylation of 7
 AE9 is C_{12-16} alcohol ethoxylate, with an average degree of ethoxylation of 9
 - 10 HSAS is a mid-branched primary alkyl sulfate with carbon chain length of about 16-17 as disclosed in US 6,020,303 and US 6,060,443
 Polyacrylate MW 4500 is supplied by BASF
 Carboxymethyl cellulose is Finnfix® V supplied by CP Kelco, Arnhem, Netherlands
 CHEC is a cationically modified hydroxyethyl cellulose polymer.
 - 15 Phosphonate chelants are, for example, diethylenetetraamine pentaacetic acid (DTPA)
 Hydroxyethane di phosphonate (HEDP)
 Savinase®, Natalase®, Stainzyme®, Lipex®, Celluclean™, Mannaway® and Whitezyme® are all products of Novozymes, Bagsvaerd, Denmark.
 Purafect®, Purafect Prime® are products of Genencor International, Palo Alto, California, USA
 - 20 Fluorescent Brightener 1 is Tinopal® AMS, Fluorescent Brightener 2 is Tinopal® CBS-X, Direct Violet 9 is Pergasol® Violet BN-Z NOBS is sodium nonanoyloxybenzenesulfonate
 TAED is tetraacetylenediamine
 S-ACMC is carboxymethylcellulose conjugated with C.I. Reactive Blue 19 product name AZO-CM-CELLULOSE
 - 25 Soil release agent is Repel-o-tex® PF
 Acrylic Acid/Maleic Acid Copolymer is molecular weight 70,000 and acrylate: maleate ratio 70:30
 EDDS is a sodium salt of ethylenediamine-N,N'-disuccinic acid, (S,S) isomer Suds suppressor agglomerate is supplied by Dow Corning, Midland, Michigan, USA
 HSAS is mid-branched alkyl sulfate
 - 30 Liquitint® Violet CT is supplied by Milliken, Spartanburg, South Carolina, USA
- ¹ Random graft copolymer is a polyvinyl acetate grafted polyethylene oxide copolymer having a

polyethylene oxide backbone and multiple polyvinyl acetate side chains. The molecular weight of the polyethylene oxide backbone is about 6000 and the weight ratio of the polyethylene oxide to polyvinyl acetate is about 40 to 60 and no more than 1 grafting point per 50 ethylene oxide units.

5 ² Polyethyleneimine (MW = 600) with 20 ethoxylate groups per -NH.

³Amphiphilic alkoxyated polymer is a polyethylenimine (MW 600), prepared from a polymer that is derivatised to contain 24 ethoxylate groups per -NH and 16 Propoxylate groups per -NH.

Amylase⁴ is any of a) to k) herein (mg active protein).

10 Table 5: Examples of Unit Dose Laundry detergent compositions (numbered 22 to 26). Such unit dose formulations can comprise one or multiple compartments.

	22 (wt%)	23 (wt%)	24 (wt%)	25 (wt%)	26 (wt%)
Alkylbenzene sulfonic acid	14.5	14.5	14.5	14.5	14.5
C ₁₂₋₁₈ alkyl ethoxy 3 sulfate	7.5	7.5	7.5	7.5	7.5
C ₁₂₋₁₈ alkyl 7-ethoxylate	13.0	13.0	13.0	13.0	13.0
Citric Acid	0.6	0.6	0.6	0.6	0.6
Fatty Acid	14.8	14.8	14.8	14.8	14.8
*Amylase of this invention (mg active)	6	12	8	2	10
Ethoxylated Polyethylenimine ¹	4.0	4.0	4.0	4.0	4.0
Protease (Purafect Prime®, 40.6 mg active/g)	1.4	2.0	0.9	1.2	0
Cellulase (Celluclean, active protein)	0.1	0.2			0.1
Amylase ⁴ (active protein) a) to k) herein	0.1	0.05	0.1	0.2	0.1
Hydroxyethane diphosphonic acid	1.2	1.2	1.2	1.2	1.2
Brightener	0.3	0.3	0.3	0.3	0.3
P-diol	15.8	13.8	13.8	13.8	13.8
Glycerol	6.1	6.1	6.1	6.1	6.1
MEA	8.0	8.0	8.0	8.0	8.0
TIPA	-	-	2.0	-	-
TEA	-	2.0	-	-	-
Cumene sulphonate	-	-	-	-	2.0
cyclohexyl dimethanol	-	-	-	2.0	-
Water	10	10	10	10	10

Structurant	0.14	0.14	0.14	0.14	0.14
Perfume	1.9	1.9	1.9	1.9	1.9
Buffers (monoethanolamine)	To pH 8.0				
Solvents (1,2 propanediol, ethanol)	To 100%				

*Amylase of the present invention is shown as mgs of active enzyme per 100g of detergent.

¹ Polyethylenimine (MW = 600) with 20 ethoxylate groups per -NH.

Table 6: Example of Multiple Compartment Unit Dose Composition (numbered 27). Multiple compartment unit dose laundry detergent formulations of the present invention are provided below. In these examples the unit dose has three compartments, but similar compositions can be made with two, four or five compartments. The film used to encapsulate the compartments is polyvinyl alcohol.

Base composition 1	27 (wt%)
Glycerol (min 99)	5.3
1,2-propanediol	10.0
Citric Acid	0.5
Monoethanolamine	10.0
Caustic soda	-
Dequest 2010	1.1
Potassium sulfite	0.2
*Amylase of this invention (mg active)	8.0
Nonionic Marlipal C24EO7	20.1
HLAS	24.6
Optical brightener FWA49	0.2
C12-15 Fatty acid	16.4
Polymer Lutensit Z96	2.9
Polyethyleneimine ethoxylate PEI600 E20	1.1
MgCl ₂	0.2
Solvents (1,2 propanediol, ethanol)	To 100%

Table 7: Multi-compartment formulations (numbered as composition 1 or 2 and with a compartment designation [A, B, or C]).

Composition	<u>1</u>		<u>2</u>
-------------	----------	--	----------

Compartment	A	B	C		A	B	C
Volume of each compartment	40 ml	5 ml	5 ml		40 ml	5 ml	5 ml
Active material in Wt. %							
Perfume	1.6	1.6	1.6		1.6	1.6	1.6
Dyes	< 0.01	< 0.01	< 0.01		< 0.01	< 0.01	< 0.01
TiO ₂	0.1	-	-		-	0.1	-
Sodium Sulfite	0.4	0.4	0.4		0.3	0.3	0.3
Acusol 305	1.2				2	-	-
Hydrogenated castor oil	0.14	0.14	0.14		0.14	0.14	0.14
<u>Base Composition 1</u>	Add to 100%	Add to 100%	Add to 100%		Add to 100%	Add to 100%	Add to 100%

*Amylase of the present invention is shown as mgs of active enzyme per 100g of detergent.

The dimensions and values disclosed herein are not to be understood as being strictly limited to the exact numerical values recited. Instead, unless otherwise specified, each such dimension is intended to mean both the recited value and a functionally equivalent range surrounding that value. For example, a dimension disclosed as "40 mm" is intended to mean "about 40 mm".

10 EXAMPLES

pNP-G7 assay for determination of alpha-amylase activity

The alpha-amylase activity may be determined by a method employing the G7-pNP substrate. G7-pNP which is an abbreviation for 4,6-ethylidene(G₇)-*p*-nitrophenyl(G₁)- α ,D-maltoheptaoside, a blocked oligosaccharide which can be cleaved by an endo-amylase, such as an alpha-amylase. Following the cleavage, the alpha-Glucosidase included in the kit digest the hydrolysed substrate further to liberate a free PNP molecule which has a yellow color and thus can be measured by visible spectrophotometry at $\lambda=405\text{nm}$ (400-420 nm.). Kits containing G7-pNP substrate and alpha-Glucosidase is manufactured by Roche/Hitachi (cat. No.11876473).

20 REAGENTS:

The G7-pNP substrate from this kit contains 22 mM 4,6-ethylidene- G7-pNP and 52.4 mM HEPES (2-[4-(2-hydroxyethyl)-1-piperaziny]-ethanesulfonic acid), pH 7.0) .

The alpha-Glucosidase reagent contains 52.4 mM HEPES, 87 mM NaCl, 12.6 mM MgCl₂, 0.075 mM CaCl₂, ≥ 4 kU/L alpha-glucosidase).

The substrate working solution is made by mixing 1 mL of the alpha-Glucosidase reagent with 0.2 mL of the G7-pNP substrate. This substrate working solution is made immediately before use.

Dilution buffer: 50 mM MOPS, 0.05% (w/v) Triton X100 (polyethylene glycol p-(1,1,3,3-tetramethylbutyl)-phenyl ether (C₁₄H₂₂O(C₂H₄O)_n (n = 9-10))), 1mM CaCl₂, pH8.0.

PROCEDURE:

The amylase sample to be analyzed was diluted in dilution buffer to ensure the pH in the diluted sample is 7. The assay was performed by transferring 20µl diluted enzyme samples to 96 well microtiter plate and adding 80µl substrate working solution. The solution was mixed and pre-incubated 1 minute at room temperature and absorption is measured every 20 sec. over 5 minutes at OD 405 nm.

The slope (absorbance per minute) of the time dependent absorption-curve is directly proportional to the specific activity (activity per mg enzyme) of the alpha-amylase in question under the given set of conditions. The amylase sample should be diluted to a level where the slope is below 0.4 absorbance units per minute.

Automatic Mechanical Stress Assay (AMSA) for laundry

In order to assess the wash performance in laundry washing experiments are performed, using the Automatic Mechanical Stress Assay (AMSA). With the AMSA, the wash performance of a large quantity of small volume enzyme-detergent solutions can be examined. The AMSA plate has a number of slots for test solutions and a lid firmly squeezing the laundry sample, the textile to be washed against all the slot openings. During the washing time, the plate, test solutions, textile and lid are vigorously shaken to bring the test solution in contact with the textile and apply mechanical stress in a regular, periodic oscillating manner. For further description see WO02/42740 especially the paragraph "Special method embodiments" at page 23-24.

General wash performance description

A test solution comprising water (10°dH), detergent, e.g. 5.1 g/L European liquid detergent as described below and the enzyme of the invention, e.g. at concentration of 0, 0.8 and/or 1.2 mg enzyme protein/L, is prepared. Fabrics stained with starch (e.g. CS-28 from Center For Testmaterials BV, P.O. Box 120, 3133 KT, Vlaardingen, The Netherlands) is added and washed for 20 minutes at 20°C. After thorough rinse under running tap water and drying in the dark, the light intensity or reflectance values of the stained fabrics are subsequently measured as

a measure for wash performance. The test with 0 mg enzyme protein/L is used as a blank to obtain a delta remission value. Preferably mechanical action is applied during the wash step, e.g. in the form of shaking, rotating or stirring the wash solution with the fabrics.

5 The AMSA wash performance experiments were conducted under the experimental conditions specified below:

Table 8: AMSA experimental conditions

Laundry liquid detergent dosage	5.7 g/L European (EU) liquid detergent (cf. Example 1A), or 0.8 g/L Northern America (US) liquid detergent (cf. Example 1B)
Test solution volume	160 micro L
pH	as is
Wash time	5-20 minutes, preferably 20 minutes
Temperature	15-40°C, preferably 20°C
Water hardness	10°dH, $\text{Ca}^{2+}:\text{Mg}^{2+}:\text{HCO}_3^- = 3:1:6$
Enzyme concentration in test solution	0.8 and 1.2 Mg/L
Test material	CS-28 (Rice starch on cotton)

10 Amylase dilution buffer: Amylase was diluted in ultrapure water (MilliQ water) with a small concentration of calcium (0.1 mM) to stabilize the amylase during storage and 0.01 % Triton X-100 to reduce risk of adsorption of enzyme protein to containers and pipettes.

Water hardness was adjusted to 10°dH by addition of CaCl_2 , MgCl_2 , and NaHCO_3 ($\text{Ca}^{2+}:\text{Mg}^{2+}:\text{HCO}_3^- = 3:1:4.5$) to the test system. After washing the textiles were flushed in tap water and dried.

15 The wash performance is measured as the brightness of the color of the textile washed. Brightness can also be expressed as the intensity of the light reflected from the sample when illuminated with white light. When the sample is stained the intensity of the reflected light is lower, than that of a clean sample. Therefore, the intensity of the reflected light can be used to measure wash performance.

20 Color measurements are made with a professional flatbed scanner (Kodak iQsmart, Kodak, Midtager 29, DK-2605 Brøndby, Denmark), which is used to capture an image of the washed textile.

To extract a value for the light intensity from the scanned images, 24-bit pixel values from the image are converted into values for red, green and blue (RGB). The intensity value (Int) is calculated by adding the RGB values together as vectors and then taking the length of the resulting vector:

$$Int = \sqrt{r^2 + g^2 + b^2}$$

Results of the AMSA laundry test of different variants are shown in Table 1 and 2. In the result the index is 100. The performance result of the parent alpha-amylase is assigned the value of 100 and the results of the variants are compared to this value.

TOM wash performance

Water hardness was adjusted to the strength described below by addition of CaCl_2 , MgCl_2 and NaHCO_3 . Wash solutions were prepared with desired amount of detergent, temperature and water hardness in a bucket as described below. Detergent was dissolved during magnet stirring for 10 minutes (wash solution was used within 30 to 60 min after preparation).

Temperature and rotation (rpm) in the water bath in the Terg-O-toMeter were set according to the settings below in Table 2. When temperature was adjusted according to settings (tolerance is $\pm 0.5^\circ\text{C}$) wash solution was added to TOM beaker according to the amount described below.

Agitation in the beaker was at 200 rpm. 2 handmade rice starch swatches (HM CS-28), 2 handmade tapioca starch swatches (HM CS-29) and ballast were added to each of the beakers and wash carried out according to time stated below. Swatches were rinsed in cold tap water for 5 minutes and placed in a washing bag and rinsed in washing machine (AEG ÖKO LAVAMAT 86820) on "STIVN" program. The swatches were sorted and left to dry between filter paper in a drying cupboard without heat overnight.

Textile sample HM CS-28 (rice starch on cotton, 5x5 cm, starch applied in 2.5 cm in diameter circle) and HM CS-29 (tapioca starch on cotton, 5x5 cm, starch applied in 2.5 cm in diameter circle) were obtained from Center for Testmaterials BV, P.O. Box 120, 3133 KT Vlaardingen, the Netherlands.

White knitted cotton was used as ballast and was obtained from Warwick Equest Ltd, Unit 55, Consett Business Park, Consett, County Durham, DH8 6BN UK.

Table 9: Experimental conditions

	European conditions using WE SUD	European conditions using WE HDL
--	-------------------------------------	-------------------------------------

Detergent dosage	1.87 g/L	5.30 g/L
Enzyme concentration in wash solution	0.065 mg enzyme protein/L	0.2 mg enzyme protein/L
Water hardness	20.6°dH (Ca ²⁺ :Mg ²⁺ :HCO ₃ ⁻ = 4:1:7.5)	
Test solution volume	1000 ml	
Wash time	5-15 minutes, preferably 15 minutes	
Rotation	200 rpm	
pH	as is	
Temperature	15-40°C, preferably at 15°C	

Detergents and test materials were as follows:

Laundry liquid detergent	European (WE) conditions: WE SUD as described in Example 2A below and WE HDL as described in Example 2B below (Detergent K).
Test material	HM CS-28 (Rice starch on cotton, 5x5 cm swatch with starch applied in 2.5 cm in diameter circle), HM CS-29 (tapioca starch on cotton, 5x5 cm swatch with starch applied in 2.5 cm in diameter circle).
Ballast	White knitted cotton in size 5x5cm added to a total weight of 40 g (40 g including all swatches i.e. ballast and test material).

The wash performance was measured as the brightness of the color of the textile washed expressed in remission values (REM). Remission measurements were made using a Macbeth 7000 Color Eye spectrophotometer. Each of the dry swatches was measured. As there is a risk of interference from the back-ground, the swatches were placed on top of 2 layers of fabric during the measurement of the remission. The remission was measured at 460 nm. The UV filter was not included. An average result for remission for the swatches was calculated.

The wash performance of different variants is shown in Table 5 as Improvement Factor (IF) and is calculated as shown below:

$$IF = \frac{REM_{\text{Variant}} - REM_{\text{Blank}}}{REM_{\text{Reference enzyme}} - REM_{\text{Blank}}}$$

Example 1**Wash performance of alpha-amylases using Automatic Mechanical Stress Assay**

In order to assess the wash performance of the alpha-amylases in a detergent base composition, washing experiments may be performed using Automatic Mechanical Stress Assay (AMSA). With the AMSA test the wash performance of a large quantity of small volume enzyme-detergent solutions can be examined. The AMSA plate has a number of slots for test solutions and a lid firmly squeezing the textile swatch to be washed against all the slot openings. During the washing time, the plate, test solutions, textile and lid are vigorously shaken to bring the test solution in contact with the textile and apply mechanical stress in a regular, periodic oscillating manner. For further description see WO 02/42740, especially the paragraph "Special method embodiments" at page 23-24.

General wash performance description

A test solution comprising water (6°dH or 15°dH), 0.79 g/L detergent, e.g., model detergent J as described below, and the enzyme of the invention at concentration of 0 or 0.2 mg enzyme protein/L, is prepared. Fabrics stained with starch (CS-28 from Center For Test materials BV, P.O. Box 120, 3133 KT, Vlaardingen, The Netherlands) is added and washed for 10 minutes at 20°C and 40°C, or alternatively 10 minutes at 20°C and 30°C as specified in the examples. After thorough rinse under running tap water and drying in the dark, the light intensity values of the stained fabrics are subsequently measured as a measure for wash performance. The test with 0 mg enzyme protein/L is used as a blank and corresponds to the contribution from the detergent. Preferably mechanical action is applied during the wash step, e.g. in the form of shaking, rotating or stirring the wash solution with the fabrics. The AMSA wash performance experiments may be conducted under the experimental conditions specified below:

Table A: Experimental condition

Detergent	Liquid Model detergent J (see Table B)
Detergent dosage	0.79 g/L
Test solution volume	160 micro L
pH	As is
Wash time	10 minutes
Temperature	20°C or 30°C
Water hardness	6°dH

Enzyme concentration in test	0.2 mg enzyme protein/L and 0,05 mg enzyme protein/L
Test material	CS-28 (Rice starch cotton)

Table B: Model detergent J

Compound	Content of compound (% w/w)	% active component (% w/w)
LAS	5.15	5.00
AS	5.00	4.50
AEOS	14.18	10.00
Coco fatty acid	1.00	1.00
AEO	5.00	5.00
MEA	0.30	0.30
MPG	3.00	3.00
Ethanol	1.50	1.35
DTPA (as Na5 salt)	0.25	0.10
Sodium citrate	4.00	4.00
Sodium formate	1.00	1.00
Sodium hydroxide	0.66	0.66
H ₂ O, ion exchanged	58.95	58.95

Water hardness was adjusted to 6°dH by addition of CaCl₂, MgCl₂, and NaHCO₃ (Ca²⁺:Mg²⁺:HCO₃⁻ = 2:1:4.5) to the test system. After washing the textiles were flushed in tap water and dried.

5

Table C: Experimental condition

Detergent	Liquid Model detergent A (see Table D)
Detergent dosage	3.33 g/L
Test solution volume	160 micro L
pH	As is
Wash time	10 minutes
Temperature	20°C or 40°C
Water hardness	15°dH
Enzyme concentration in test	0.2 mg enzyme protein/L, 0.05 mg enzyme protein/L
Test material	CS-28 (Rice starch cotton)

Table D: Model detergent A

Compound	Content of compound (% w/w)	% active component (% w/w)
LAS	12.00	11.60
AEOS, SLES	17.63	4.90
Soy fatty acid	2.75	2.48
Coco fatty acid	2.75	2.80
AEO	11.00	11.00
Sodium hydroxide	1.75	1.80
Ethanol / Propan-2-ol	3.00	2.70/0.30
MPG	6.00	6.00
Glycerol	1.71	1.70
TEA	3.33	3.30
Sodium formate	1.00	1.00
Sodium citrate	2.00	2.00
DTMPA	0.48	0.20
PCA	0.46	0.18
Phenoxy ethanol	0.50	0.50
H ₂ O, ion exchanged	33.64	33.64

Water hardness was adjusted to 15°dH by addition of CaCl₂, MgCl₂, and NaHCO₃

- 5 (Ca²⁺:Mg²⁺:HCO₃⁻ = 4:1:7.5) to the test system. After washing the textiles were flushed in tap water and dried.

Table E: Experimental condition

Detergent	[Detergent Composition K]
Detergent dosage	5.3 g/L
Test solution volume	160 micro L
pH	As is
Wash time	10 minutes
Temperature	20°C or 40°C
Water hardness	15°dH
Enzyme concentration in test	0.2 mg enzyme protein/L, 0.05 mg enzyme protein/L

Test material	CS-28 (Rice starch cotton)
---------------	----------------------------

Table F: Detergent K (as stated in the table 11 below)

Compound	Content of compound (wt% active)
Sodium alkylbenzene sulfonate	8.7
Sodium alkyl ethoxy 3 sulfate	1.0
C12-18 alkyl 1.5-7-ethoxylate	5.3
Citric Acid	3.1
Optical Brightener	0.05
Polypropylene Glycol	1.1
Phosphonated chelant	0.5
Minors (dyes perfumes, enzymes, enzyme stabilisers, solvents, structurants, polymers) and water	to 100%

Water hardness was adjusted to 15°dH by addition of CaCl₂, MgCl₂, and NaHCO₃

- 5 (Ca²⁺:Mg²⁺:HCO₃⁻ = 4:1:7.5) to the test system. After washing the textiles were flushed in tap water and dried.

10 The wash performance is measured as the brightness expressed as the intensity of the light reflected from the sample when illuminated with white light. When the sample is stained the intensity of the reflected light is lower, than that of a clean sample. Therefore, the intensity of the reflected light can be used to measure wash performance.

Color measurements are made with a professional flatbed scanner (EPSON Expression 10000XL, EPSON) used to capture an image of the washed textile.

15 To extract a value for the light intensity from the scanned images, 48→24 Bit Color pixel values from the image are converted into values for red, green and blue (RGB). The intensity value (Int) is calculated by adding the RGB values together as vectors and then taking the length of the resulting vector:

$$Int = \sqrt{r^2 + g^2 + b^2}$$

20

The wash performance of the variants according to the invention are shown in the tables below. Table 3 shows the results obtained from the experiment accessing the wash performance in model detergents A (Table D) and J (Table B) in different concentrations (0.05mg enzyme/L

detergent and 0.2mg enzyme/L detergent), and at different temperatures (20°C and 40°C). Table 4 shows the results obtained from the experiment accessing the wash performance in detergent K (Table F) in different concentrations (0.05mg enzyme/L detergent and 0.2mg enzyme/L detergent) and at different temperatures (20°C and 40°C).

Table 10: Results of wash performance in Model detergents

Mutations	0.05mg/L -A-20C	0.2mg/L -A-20C	0.05mg/L -A-40C	0.2mg/L -A-40C	0.05mg/L -J-20C	0.2mg/L -J-20C	0.05mg/L -J-30C	0.2mg/L -J-30C
SEQ ID NO: 2 + H1*+G7A+G109A+N280S+W284H+K320A+M323N+E391A	4.7	1.5	2.3	1.1	2.7	1.1	3.5	1.5
SEQ ID NO: 2+ G7A+W284H+K320A+M323N	4.7	1.4	1.8	1.0	3.0	1.1	2.0	1.2
SEQ ID NO: 2+ G7A+K320A+M323N	4.3	1.6	2.0	1.0	2.0	1.1	2.0	1.2
SEQ ID NO: 2+ K320A	4.7	1.3	2.3	1.0	3.7	1.0	4.5	1.5
SEQ ID NO: 2+ G7A+K320A	4.3	1.4	2.3	1.1	3.3	1.1	2.5	1.3
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+E391A	1.7	1.2	1.3	1.3	1.5	1.3	1.1	1.4
SEQ ID NO: 2+ H1*+G109A+N280S+W284H+E391A	2.3	1.7	2.8	1.4	5.0	2.0	5.5	1.8
SEQ ID NO: 2+ H1*+G109A+N280S+E391A	0.8	1.0	1.0	1.1	1.0	1.1	1.3	1.1
SEQ ID NO: 2+ H1*+G109A+N280S+M323S+E391A	1.3	1.9	1.5	1.3	1.5	1.5	1.6	1.6
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+K320A+E391A	1.5	1.2	1.3	1.2	1.2	1.1	1.7	1.1
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+M323S+E391A	1.3	1.3	1.5	1.1	1.0	1.0	1.2	1.1
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+M323N+E391A	1.4	1.3	1.3	1.2	1.0	1.2	1.6	1.1
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+W284F+E391A	1.3	1.4	1.3	1.2	0.9	1.2	1.6	1.0
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+W284R+E391A	1.1	1.3	1.0	1.0	0.7	1.2	1.4	0.9

SEQ ID NO: 2 + H1*+G7A+G109A+N280S+K320A+M323S+E391A	1.1	1.3	0.9	1.1	0.5	1.2	0.8	1.0
SEQ ID NO: 2 + H1*+G7A+G109A+W284R+E391A	1.0	1.2	0.9	1.1	0.6	1.1	1.1	1.0
SEQ ID NO: 2 + H1*+G7A+G109A+N280S+K320A+M323N+E391A	1.0	1.2	0.8	1.0	0.8	1.2	0.9	1.0

Mutations	0.05mg/L-A-20C	0.2mg/L-A-20C	0.05mg/L-A-40C	0.2mg/L-A-40C	0.05mg/L-J-20C	0.2mg/L-J-20C	0.05mg/L-J-30C	0.2mg/L-J-30C
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+W284H+K320A+M323N+E391A	4.7	1.5	2.3	1.1	2.7	1.1	3.5	1.5
SEQ ID NO: 2+ G7A+W284H+K320A+M323N	4.7	1.4	1.8	1.0	3.0	1.1	2.0	1.2
SEQ ID NO: 2+ G7A+K320A+M323N	4.3	1.6	2.0	1.0	2.0	1.1	2.0	1.2
SEQ ID NO: 2+ K320A	4.7	1.3	2.3	1.0	3.7	1.0	4.5	1.5
SEQ ID NO: 2+ G7A+K320A	4.3	1.4	2.3	1.1	3.3	1.1	2.5	1.3
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+E391A	1.7	1.2	1.3	1.3	1.5	1.3	1.1	1.4
SEQ ID NO: 2+ H1*+G109A+N280S+W284H+E391A	2.3	1.7	2.8	1.4	5.0	2.0	5.5	1.8
SEQ ID NO: 2+ H1*+G109A+N280S+E391A	0.8	1.0	1.0	1.1	1.0	1.1	1.3	1.1

SEQ ID NO: 2+ H1*+G109A+N280S+M323S+E391A	1.3	1.9	1.5	1.3	1.5	1.5	1.6	1.6
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+K320A+E391A	1.5	1.2	1.3	1.2	1.2	1.1	1.7	1.1
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+M323S+E391A	1.3	1.3	1.5	1.1	1.0	1.0	1.2	1.1
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+M323N+E391A	1.4	1.3	1.3	1.2	1.0	1.2	1.6	1.1
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+W284F+E391A	1.3	1.4	1.3	1.2	0.9	1.2	1.6	1.0
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+W284R+E391A	1.1	1.3	1.0	1.0	0.7	1.2	1.4	0.9
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+K320A+M323S+E391A	1.1	1.3	0.9	1.1	0.5	1.2	0.8	1.0
SEQ ID NO: 2+ H1*+G7A+G109A+W284R+E391A	1.0	1.2	0.9	1.1	0.6	1.1	1.1	1.0
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+K320A+M323N+E391A	1.0	1.2	0.8	1.0	0.8	1.2	0.9	1.0

Table 11: Results of wash performance in detergent K

Mutations	0.05mg/L-Ariel-20C	0.2mg/L-Ariel-20C	0.05mg/L-Ariel-40C	0.2mg/L-Ariel-40C
Reference - SEQ ID NO: 2	1.0	1.0	1.0	1.0
SEQ ID NO: 2+ H1*+G109A+N280S+E391A	1.1	1.0	2.0	1.1
SEQ ID NO: 2+ H1*+G7K+G109A+N280S+E391A	1.2	1.6	0.9	1.1
SEQ ID NO: 2+ H1*+G7E+G109A+N280S+E391A	1.2	1.6	0.6	1.0
SEQ ID NO: 2+ H1*+G7N+G109A+N280S+E391A	0.9	1.6	1.0	1.1

SEQ ID NO: 2+ H1*+G7Q+G109A+N280S+E391A	1.6	1.6	1.1	1.1
SEQ ID NO: 2+ H1*+G7L+G109A+N280S+E391A	1.6	1.6	1.4	1.3
SEQ ID NO: 2+ H1*+G7D+G109A+N280S+E391A	1.9	1.6	1.5	1.3
SEQ ID NO: 2+ H1*+G109A+N280S+K320A+E391A	0.4	0.9	1.0	1.3
SEQ ID NO: 2+ H1*+G109A+N280S+K320M+E391A	1.5	1.1	1.8	1.3
SEQ ID NO: 2+ H1*+G109A+N280S+K320T+E391A	1.3	1.1	1.5	1.1
SEQ ID NO: 2+ H1*+G109A+N280S+K320V+E391A	1.0	0.9	1.7	1.1
SEQ ID NO: 2+ H1*+G109A+N280S+M323R+E391A	0.7	0.9	1.3	1.0
SEQ ID NO: 2+ H1*+G109A+N280S+K320S+E391A	1.8	1.3	1.8	1.3
SEQ ID NO: 2+ H1*+G109A+N280S+E391V	1.3	1.1	2.0	1.3
SEQ ID NO: 2+ H1*+G109A+W284R+E391A	1.2	1.3	1.2	1.2
SEQ ID NO: 2+ H1*+G109A+W284F+E391A	1.7	1.3	1.8	1.2
SEQ ID NO: 2+ H1*+G109A+N280S+K320A+M323S+E391A	2.0	1.4	1.4	1.2
SEQ ID NO: 2+ H1*+G109A+N280S+W284F+E391A	0.9	0.9	1.3	1.0
SEQ ID NO: 2+ H1*+G109A+N280S+M323N+E391A	2.0	1.6	2.0	1.3
SEQ ID NO: 2+ H1*+G109A+N280S+M323K+E391A	2.3	1.6	2.0	1.6
SEQ ID NO: 2+ H1*+G109S+N280S+E391A	2.5	1.3	2.3	1.1
SEQ ID NO: 2+ H1*+G109A+W284H+E391A	1.5	1.2	1.3	1.1
SEQ ID NO: 2+ H1*+G109A+N280S+K320A+M323N+E391A	1.6	1.1	1.2	1.1
SEQ ID NO: 2+ H1*+G7A+G109A+N280S+E391A	1.5	1.4	1.2	1.2

As can be seen from Table 10 and Table 11, all the tested variants have an improved wash performance compared to the reference (SEQ ID NO: 2) in at least one of the tested conditions.

Example 2 - Wash performance in TOM of alpha-amylases in liquid detergent K

The wash performance of the tested variant and corresponding parent alpha-amylase (SEQ ID NO: 2) were tested as described above for TOM scale wash. The detergent used was detergent K. The results are given as (performance of variant minus performance of blank) divided by (performance of parent minus performance of blank).

Table 12: Wash performance in TOM scale

	Detergent K [WE HDL]	
	IF HM CS-28	IF HM CS-29
Reference /SEQ ID NO: 2)	1,00	1,00
SEQ ID NO: 2 + H1*+G109A+W284H+E391A	1,28	1,58
SEQ ID NO: 2 + H1*+G109A+N280S+K320A+M323N+E391A	1,13	1,61
SEQ ID NO: 2 + H1*+G7A+G109A+N280S+E391A	1,22	2,02

Table 13: Wash performance in TOM scale

1L, 5min wash 0.13 mg enzyme protein/L	Det. K	
	IF HM CS-29	IF HM CS-26
Reference SEQ ID NO: 2	1,00	1,00
SEQ ID NO: 2 + H1*+G109A+W284H+E391A	1,00	1,00
SEQ ID NO: 2 + H1*+G109A+N280S+K320A+M323N+E391A	1,08	1,11
SEQ ID NO: 2 + H1*+G7A+G109A+N280S+E391A	1,10	1.13

Table 14: Wash Performance in Full Scale Washing Machine Test

Full scale, 0.072 mg enzyme protein/L	Det. K	
	IF HM CS-29	IF HM CS-26
Reference SEQ ID NO: 2	1,00	1,00
SEQ ID NO: 2 + H1*+G109A+W284H+E391A	1,07	1,02
SEQ ID NO: 2 + H1*+G109A+N280S+K320A+M323N+E391A	1,22	1,32
SEQ ID NO: 2 + H1*+G7A+G109A+N280S+E391A	1,32	1.41

KRAV

1. Variant af en stam-alfa-amylase, hvor varianten omfatter

(i) substitutioner i positioner svarende til positioner G109A og G7A i aminosyre-stamsekvensen ifølge SEQ ID NO: 2,

(ii) varianten har mindst 90% men mindre end 100% sekvensidentitet med aminosyre-stamsekvensen ifølge SEQ ID NO: 2, og

(iii) varianten har alfa-amylase-aktivitet,

hvor varianten har en forbedret vaskeydelse ved en lav temperatur mellem 10-35°C sammenlignet med aminosyre-stamsekvensen ifølge SEQ ID NO: 2.

2. Variant ifølge krav 1, hvor varianten har mindst 95%, mindst 96%, mindst 97%, mindst 98%, eller mindst 99% men mindre end 100% sekvensidentitet med aminosyresekvensen i stam-alfa-amylasen ifølge SEQ ID NO: 2.

3. Variant ifølge et hvilket som helst af de foregående krav, hvor varianten omfatter modifikationer i positionerne svarende til positionerne i aminosyresekvensen ifølge SEQ ID NO: 2, valgt fra gruppen bestående af:

H1*+G7A+G109A+N280S+E391A;

H1*+G7A+G109A+N280S+W284H+K320A+M323N+E391A;

H1*+G7A+G109A+N280S+E391A;

H1*+G7A+G109A+N280S+K320A+E391A;

H1*+G7A+G109A+N280S+M323S+E391A;

H1*+G7A+G109A+N280S+M323N+E391A;

H1*+G7A+G109A+N280S+W284F+E391A;

H1*+G7A+G109A+N280S+W284R+E391A;

H1*+G7A+G109A+N280S+K320A+M323S+E391A;

H1*+G7A+G109A+W284R+E391A; og

H1*+G7A+G109A+N280S+K320A+M323N+E391A.

4. Variant ifølge et hvilket som helst af de foregående krav, hvor stam-alfa-amylasen er aminosyresekvensen ifølge SEQ ID NO: 2, eller en hvilken som helst alfa-amylase med mindst 90%, såsom mindst 92%, såsom mindst 95%, såsom mindst 97%, såsom mindst 98%, såsom mindst 99% eller 100% sekvensidentitet med en hvilken som helst af aminosyresekvenserne ifølge SEQ ID NO: 2.

5. Variant ifølge et hvilket som helst af de foregående krav, hvor stam-alfa-amylasen omfatter eller består af aminosyresekvensen ifølge SEQ ID NO: 1 og 2.
6. Polynukleotid, der koder for varianten ifølge et hvilket som helst af de foregående krav.
7. Nukleinsyrekonstrukt, der omfatter polynukleotidet ifølge krav 6.
8. Ekspressionsvektor, der omfatter polynukleotidet ifølge krav 6.
9. Værtscelle, der omfatter polynukleotidet ifølge krav 6.
10. Fremgangsmåde til fremstilling af en alfa-amylase-variant, hvilken fremgangsmåde omfatter:
 - a. dyrkning af værtscellen ifølge krav 9 under betingelser, der er egnet til ekspresion af varianten; og
 - b. indvinding af varianten.