(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization

International Bureau





(10) International Publication Number WO 2016/193542 A1

(43) International Publication Date 8 December 2016 (08.12.2016)

(51) International Patent Classification: C08B 3/10 (2006.01) COSL 1/10 (2006.01) C08B 11/02 (2006.01) COSL 1/28 (2006.01) C08B 1/02 (2006.01)

(21) International Application Number:

PCT/FI2016/050375

(22) International Filing Date:

30 May 2016 (30.05.2016)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

20155406

29 May 2015 (29.05.2015)

FΙ

- (71) Applicant: TEKNOLOGIAN TUTKIMUSKESKUS VTT OY [-/FI]; Vuorimiehentie 3, 02150 Espoo (FI).
- (72) Inventors: ROPPONEN, Jarmo; c/o VTT, P.O. Box 1000, 02044 Vtt (FI). TALJA, Riku; c/o VTT, P.O. Box 1000, 02044 Vtt (FI). WILLBERG-KEYRILÄINEN, Pia; c/o VTT, P.O. Box 1000, 02044 Vtt (FI). HARLIN, Ali; c/o VTT, P.O. Box 1000, 02044 Vtt (FI). ASIKAIN-EN, Sari; c/o VTT, P.O. Box 1000, 02044 Vtt (FI). VAR-TIAINEN, Jari; c/o VTT, P.O. Box 1000, 02044 Vtt (FI).

- (74) Agent: SEPPO LAINE OY; Itämerenkatu 3 A, 00180 Helsinki (FI).
- (81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.
- (84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Published:

with international search report (Art. 21(3))



MOLAR MASS CONTROLLED CELLULOSE

FIELD

[0001] The present invention relates to affordable cellulose raw material with a novel beneficial method of preparing molar mass controlled transportable feed stock for added value applications in biomaterials manufacturing, including thermoplastic, dispersion able or dissolving derivatives.

5

25

30

BACKGROUND

- 10 [0002] Cellulose is the most abundant renewable organic polymer on the earth and hence can be regarded as important raw material for several industries such as textiles, papers, foods, cosmetics and biomaterials (Edgar et al., 2001). Cellulose is a linear polymer that consists of $\beta(1\rightarrow 4)$ linked D-glucose units. Hydroxyl groups of cellulose forms strong inter and intra molecular hydrogen bonds and van der Waals interactions 15 forming a resistant and stiff microfibril network. This structure is not uniform, and both highly ordered (crystalline) regions and regions with a low degree of order can be found. The relative proportion of these regions depends on the raw material and the treatments to which the cellulose has been produced (Klemm et al., 2002). Moreover, these regions caused the limited solubility of cellulose and make it difficult for solvents and reagents to 20 access areas within the cellulose fibres. As known this network is poorly reactive requiring a large excess of chemicals or demanding process conditions.
 - [0003] The modification of cellulose can be done either via homogeneous or heterogeneous procedures. In most cases, cellulose esters are produced industrially under heterogeneous conditions. Reaction rates and final degree of substitution (DS) in heterogeneous reactions are hindered by low accessibility of solid cellulose to the esterification reagents (Wei *et al.*, 2007). In order to have homogeneous chemical reaction cellulose need to be first dissolved. In order to achieve uniform chemical reactions or solubilization of cellulosic substrates, it is important to have accessibility high enough. However, due to the high crystallinity, cellulose can be only dissolved in limited solvents at low concentrations. To achieve chemical reactions efficient enough, the native cellulose

5

10

15

20

need to be first activated by disrupting inter- and intramolecular hydrogen bonding making structure accessible to further action of reactants. This can be achieved by varying degrees of chemical, enzymatic or mechanical activation.

[0004] The chemical activation can be achieved by different methods like using water, solvents, dilute acids and bases. As a result of chemical activation the cellulose structure becomes less ordered leading to an increase of the active surface area and thus increasing the number of available hydroxyl groups and the accessibility to chemicals. Well known method to disrupt fibrillar aggregation, and increase surface accessibility is to use fluids with a higher swelling power, such as dilute caustic soda (6-10%), dilute quaternary bases or aqueous zinc chloride. Disruption of the crystalline structure, such as with liquid ammonia or 20% caustic soda, which induces cellulose-I to cellulose-II (also regenerated cellulose) crystal modification. Treatments by acid hydrolysis and oxidation, thermal and mechanical treatments by grinding, ultrasonic treatment and freeze-drying and enzymatic treatment are also activation methods but they can degrade the molecules to a certain extent.

[0005] The enzymatic activation of cellulose can be done by using different cellulases which hydrolyse the 1,4-β-D-glucosidic bonds of the cellulose chain. There are three major groups of cellulases: endoglucanases, cellobiohydrolases or exoglucanases, and glucosidases. These enzymes can act alone on the cellulose chain or together degrading efficiently cellulose structure generating mainly glucose or cellobiose units.

[0006] The mechanical activation of the cellulose fibres is well known method in the pulp and paper industry. Depending on how and in which conditions the mechanical processing has done it can enhance fiber-fiber bonding, to cut or make the fibres stronger and to change cellulose structure.

25 [0007] Crepy *et al.* (2009) describe a method for synthetizing plastic materials by the internal plasticization of cellulose with fatty acids. Modifications were done in homogenous solvent under microwave irradiation, which is not currently industrially feasible. The method does not cover both homogeneous and heterogeneous reactions for molar mass controlled hydrolyzed cellulose.

30 [0008] For the application point of view cellulose solubility and mechanical properties need to be tailor without affecting its natural performance too much. For

5

15

instance, to avoid tedious recycling processes of solvents and huge excess of chemicals in chemical activation cellulose reactivity need to be increase without losing its good mechanical properties to obtain high quality cellulose based materials which are suitable for various applications. One way to increase reactivity of the cellulose is to decrease its molar mass in controlled manner.

SUMMARY OF THE INVENTION

- [0009] The invention is defined by the features of the independent claims. Some specific embodiments are defined in the dependent claims.
- 10 **[0010]** According to a first aspect of the present invention, there is provided a cheap and available raw material for biomaterial manufacturing.
 - [0011] According to a second aspect of the present invention, there is provided a hydrolyzed and reactive molar mass controlled cellulose, having for example excellent thermoplastic properties, which is thus usable after functionalization in various applications, such as composites, films, foams, encapsulation, packaging, textiles and non-wovens.
 - [0012] These and other aspects, together with the advantages thereof over known solutions are achieved by the present invention, as hereinafter described and claimed.
- [0013] The method of improving the reactivity of cellulose according to an embodiment of the present invention is mainly characterized by what is stated in the characterizing part of claim 1.
 - [0014] The cellulose ester according to an embodiment of the present invention is mainly characterized by what is stated in the characterizing part of claim 8.
- [0015] The cellulose ether according to an embodiment of the present invention is mainly characterized by what is stated in the characterizing part of claim 11.
 - [0016] The method of producing films from the molar mass controlled cellulose according to embodiments of the present invention is characterized in claims 13 and 15.

[0017] Considerable advantages are obtained by means of the invention. For example, the modification of hydrolyzed cellulose as herein described provides processable thermoplastic materials without using any external plasticizers. Additionally, this material forms mechanically strong films with excellent WVTR properties and good heat-sealability. Applying such hydrolyzed cellulose also provides benefits such as utilization of a non-food and recycled raw material source and recyclability. Furthermore, molar mass controlled cellulose coatings applied on a CNF-film provide fully cellulosic and thus also fully bio-based films with high smoothness on both surfaces.

5

15

[0018] Next, the present technology will be described more closely with reference to certain embodiments.

EMBODIMENTS

- [0019] The present technology provides means to convert inactive cellulose to more reactive form and to an easier functionalization in order to produce thermoprocessable cellulose products.
- [0020] "Long-chain fatty acid modification" herein means chain length of fatty acid substituents \geq C6, such as C6-C30 and more preferably C6-C18. Such modification may for example be esterification or etherification.
- [0021] FIGURE 1 illustrates a reaction scheme for synthesizing cellulose ester 20 samples.
 - [0022] FIGURE 2 illustrates a reaction scheme for synthesizing cellulose ether samples.
 - [0023] FIGURE 3 is a photo showing translucent and flexible rod produced with microcompounder from a processable cellulose palmitate ester at a temperature of 200 °C.
- 25 [0024] FIGURES 4A, 4B and 4C are diagrams showing the mechanical properties (E-modulus, tensile strength and tensile strain at break) of an example cellulose ester films.
 - [0025] FIGURE 5 is a diagram showing the water vapour transmission rates for example cellulose ester films.

WO 2016/193542 5 PCT/FI2016/050375

[0026] FIGURE 6 is a SEM-image (1000 x 1000) showing the advantageous effect of molar mass controlled cellulose (MMCC) coating in significantly decreasing surface porosity and roughness of CNF-films.

[0027] FIGURE 7 is similarly a SEM-image (5000 x 5000) showing the advantageous effect of molar mass controlled cellulose coating in significantly decreasing surface porosity and roughness of CNF-films.

[0028] One aspect of the present invention is a method of improving the reactivity of cellulose and preparing a transportable form of the previous for preparation of thermoplastic, dispersion able or dissolving derivatives, wherein molar mass and molar mass distribution of the cellulose is controlled uniformly to a range between 30 and 300 kDa therefore providing reactive and processable cellulose.

10

15

20

25

30

[0029] According to an embodiment, the method of improving the reactivity of cellulose comprises controlling (i.e. decreasing) the molar mass of a cellulose raw material via hydrolysis, excluding total hydrolysis, and by performing a long-chain (chain length between C6 and C30, such as C6-C18) fatty acid modification for the molar mass controlled cellulose.

[0030] According to one embodiment, the method comprises controlling (i.e. decreasing) the molar mass of a cellulose raw material via hydrolysis, excluding total hydrolysis, and by performing a hydroxyalkylation modification, e.g. hydroxypropylation, hydroxyethylation or hydroxybutylation for the molar mass controlled cellulose.

[0031] According to another embodiment of the invention, the hydrolysis is controlled so that the average molecular mass of the cellulose is reduced at least 60 % but not more than 85 % from the molecular mass of the starting raw material. It is preferred that the hydrolysis is controlled so that after the hydrolysis the average molecular mass of the cellulose is between 30 to 300 kDa, preferably between 40 to 200 kDa. It should be noted that the molar mass of the cellulose is indeed controlled, whereby the cellulose is not subjected to total hydrolysis.

[0032] As an example, the inventors prepared cellulose palmitates by heterogeneous esterification in pyridine and homogeneous esterification in the solvent of DMAc/LiCl. To see difference in reactivity, esterification was tested for both native softwood sulfite pulp and molar mass controlled cellulose. In addition, not only to focus cellulose reactivity, the

5

20

25

30

inventors also tested these derivatives as potential thermoplastic materials as well in applications where good and stable water vapor barrier properties are needed. It is described herein how the cellulose raw material molar mass has significant effect to cellulose reactivity. Using hydrolyzed cellulose as a starting material, better reaction efficiency was obtained without losing the good properties of the cellulose esters.

- [0033] As another example, commercial softwood sulphite dissolving grade pulp was treated with ozone to decrease the degree of polymerization. After the ozone treatment the pulp was subjected to hydrogen peroxide treatment aiming to further decrease the degree of polymerization and to reduce the content of carbonyl groups of the pulp.
- 10 **[0034]** Thus, according to one embodiment, the cellulose raw material is selected from native softwood pulp, native hardwood pulp, annual plant pulps such as bamboo pulp or straw pulp, softwood sulphite dissolving grade pulp, hardwood sulphite dissolving grade pulp, ozone treated hydrolyzed pulp or enzyme treated pulp.
- [0035] According to a further embodiment cellulose is hydrolyzed and thus activated by enzymatic treatment, ozone treatment, hydrogen peroxide treatment, alkaline treatment, or other chemical treatment, before performing a long chain fatty acid modification, such as an esterification or hydroxyalkylation.
 - [0036] According to one embodiment the long-chain fatty acid modification comprises either heterogeneous esterification or homogeneous esterification of the cellulose.
 - [0037] In case of homogenous esterification, before step a) the cellulose raw material is dissolved into LiCl/DMAc solution.
 - [0038] According to one embodiment the long chain fatty acid modification comprises heterogeneous etherification of the molar mass controlled cellulose, for example by hydroxyalkylation. The hydroxyalkylation reaction of the activated cellulose was herein subjected in alkaline conditions either with or without inert solvent such as toluene.
 - [0039] The target substitution level can be adjusted based on reaction conditions such as the amount of reagents and reaction time. The purity of washed esters was confirmed by FT-IR and NMR analysis to verify that all unreacted propylene oxide had been removed.

WO 2016/193542 7 PCT/FI2016/050375

[0040] With the MS level 0.7 or higher transparent film can be obtained when activated molar mass controlled cellulose were used.

[0041] The target degree of substitution for the cellulose esters was found to be (DS > 0.7) for both heterogeneous esterification and homogeneous esterification in the solvent of DMAc/LiCl. DMAc/LiCl -system can be used to dissolve cellulose without any degradation of cellulose. The purity of washed esters was confirmed by FT-IR and NMR analysis to verify that all unreacted fatty acid had been removed.

5

10

15

20

25

30

[0042] Thus, according to one embodiment, a cellulose ester and a cellulose ether obtained by the method as herein described has chain length between C2 and C30, such as C6-C18, and has a total degree of substitution (DS) from 0.7 to 3.

[0043] According to DS values of synthesized cellulose palmitate, it can be concluded that the molar mass of starting material has a significant effect to the end product degree of substitution. Both in homogeneous and heterogeneous method, DS values increased when molar masses decreased when similar reaction conditions were used. When pulps 1-3 were used in homogeneous system, degrees of substitution were > 0.7 and the formed cellulose esters were soluble to chloroform. In that case films could be prepared by simple solvent-casting and these films have high flexibility and optical transparency. Cellulose esters, which were synthesized in heterogeneous system using hydrolysed cellulose pulp showed also high DS and transparent films could be obtained. However, in that case, the cellulose esters were not fully soluble to chloroform due the uneven distribution of the palmitate functionalization and therefore some insoluble cellulose fibrils were observed.

[0044] The tests demonstrates that using the molar mass controlled cellulose as a starting materials, plastic films can be prepared with much lower DS values without external activation or plasticisation. In such case a smaller amount of reagents is needed, which in turn decreases the production costs.

[0045] The processability of the cellulose esters was tested with microcompaunder. Translucent and very flexible rod was formed (FIG. 3) and also fiber spinning is possible for the processed sample.

[0046] The cellulose palmitate films were also analysed by their contact angle values to determine their hydrophobicity. Contact angles indicate the cellulose ester films degree of wetting when liquid and solid phases are in interaction. When the contact angles are

high (> 90°), the films have low wettability. All measured contact angles were between 97 and 107°. That means that all our cellulose esters are hydrophobic materials.

[0047] The mechanical properties, E-modulus (E, MPa), tensile strength (σ_R MPa) and tensile strain at break (ϵ_R , %), of cellulose ester films were determined by tensile testing (FIG. 4). On the basis of the results it can be concluded that the DS has an effect to the mechanical properties of associated cellulose esters. In general, the higher DS is the better mechanical properties cellulose ester films have.

5

15

20

25

30

[0048] According to a further embodiment, a cellulose ester has a Young's modulus value of at least 300 MPa, more preferably at least 400 MPa.

10 [0049] Water vapour transmission rate and water vapour permeability were determined by measuring the amount of water vapour transmitted through the cellulose film. According to the WVTR values (FIG. 5), it was concluded that these cellulose esters have excellent water vapour permeability with quite low DS values.

[0050] According to one embodiment, a method of producing thermoformable cellulose ester or ether films is characterized by preparing the films from the purified cellulose esters or ethers without using any external plasticizers. However, plasticizers may be used for optimization causes.

[0051] Printed electronics, sensors, solar cells and diagnostics are examples or areas in which the acceptable average surface roughness (Ra) with 1000 x 1000 μm scan area is below 100 nm or even below 20 nm. CNF fine structure (micro- and nanoscale) has crucial role in surface smoothness. Tempo-oxidized CNF with very homogenous and transparent appearance forms smoother films as compared to CNF with opaque and slightly agglomerated structure. It is general knowledge in the art that nanocellulose films typicalle have high porosity. Such micro/nano -roughness, which is more or less natural behavior of any surfaces formed from imperfect dispersions, is in the present invention overcome by using thin coating solutions with zero or close to zero porosity i.e. molar mass controlled cellulose coatings (FIGS. 6 and 7).

[0052] Thus, according to an embodiment, a method of producing multilayered film structure comprises coating CNF film from both sides by immersing molar mass controlled C6-C18 cellulose into a multilayered, such as two or three layered, cellulose film structure without using external plasticizers. However, plasticizers may be used for optimization

WO 2016/193542 9 PCT/FI2016/050375

causes.

5

10

15

20

25

30

[0053] According to another embodiment, the produced cellulose ester films or multilayer CNF films are heat-sealable. The film structure needs to have a dry thickness of at least 10 μ m, more preferably at least 20 μ m and most suitably at least 30 μ m, in order to ensure the heat-sealability.

[0054] It is to be understood that the embodiments of the invention disclosed are not limited to the particular structures, process steps, or materials disclosed herein, but are extended to equivalents thereof as would be recognized by those ordinarily skilled in the relevant arts. It should also be understood that terminology employed herein is used for the purpose of describing particular embodiments only and is not intended to be limiting.

[0055] Reference throughout this specification to one embodiment or an embodiment means that a particular feature, structure, or characteristic described in connection with the embodiment is included in at least one embodiment of the present invention. Thus, appearances of the phrases "in one embodiment" or "in an embodiment" in various places throughout this specification are not necessarily all referring to the same embodiment. Where reference is made to a numerical value using a term such as, for example, about or substantially, the exact numerical value is also disclosed.

[0056] As used herein, a plurality of items, structural elements, compositional elements, and/or materials may be presented in a common list for convenience. However, these lists should be construed as though each member of the list is individually identified as a separate and unique member. Thus, no individual member of such list should be construed as a de facto equivalent of any other member of the same list solely based on their presentation in a common group without indications to the contrary. In addition, various embodiments and example of the present invention may be referred to herein along with alternatives for the various components thereof. It is understood that such embodiments, examples, and alternatives are not to be construed as de facto equivalents of one another, but are to be considered as separate and autonomous representations of the present invention.

[0057] Furthermore, the described features, structures, or characteristics may be combined in any suitable manner in one or more embodiments. In the following description, numerous specific details are provided, such as examples of lengths, widths,

WO 2016/193542 10 PCT/FI2016/050375

5

10

15

20

25

shapes, etc., to provide a thorough understanding of embodiments of the invention. One skilled in the relevant art will recognize, however, that the invention can be practiced without one or more of the specific details, or with other methods, components, materials, etc. In other instances, well-known structures, materials, or operations are not shown or described in detail to avoid obscuring aspects of the invention.

[0058] While the forgoing examples are illustrative of the principles of the present invention in one or more particular applications, it will be apparent to those of ordinary skill in the art that numerous modifications in form, usage and details of implementation can be made without the exercise of inventive faculty, and without departing from the principles and concepts of the invention. Accordingly, it is not intended that the invention be limited, except as by the claims set forth below.

[0059] The verbs "to comprise" and "to include" are used in this document as open limitations that neither exclude nor require the existence of also un-recited features. The features recited in depending claims are mutually freely combinable unless otherwise explicitly stated. Furthermore, it is to be understood that the use of "a" or "an", that is, a singular form, throughout this document does not exclude a plurality.

INDUSTRIAL APPLICABILITY

[0060] At least some embodiments of the present invention find industrial application in biomaterial manufacturing, such as in producing transparent and colorless films, for example for heat-sealable packaging applications, in foam applications, in electrical coating applications, in medical applications, in composite applications, in non-woven applications, fiber spinning and thermoforming, as well as in/for textiles, fibers, filaments, yarns and cloths. Furthermore, molar mass controlled cellulose coatings applied on a CNF-film provide fully cellulosic and thus also fully bio-based films with high smoothness on both surfaces, applicable for example in printed electronics, sensors, solar cells and diagnostics.

WO 2016/193542 11 PCT/FI2016/050375

EXAMPLE 1 - Synthesis of molar mass controlled cellulose

Ozone treatment

5

10

15

20

Z-stage was performed in a plastic flow through-reactor in medium consistency. Pulp was added into the reactor, and water was charged and the pulp was mixed when the water addition was done. Initial pH was adjusted with H₂SO₄ and oxygen flow through the reactor was started.

After 10 minutes ozone generator was started (160 A). Ozone generation (about 192 mg/min) in the carrier oxygen gas was first stabilized for 5 minutes. After stabilization ozone flow was lead to the potassium iodide solution (10 min), and after that gas flow was lead into the reactor. Pulp was mixed all the time during the ozone charging. Mixing was started already one minute before the charging and continued for one minute after the charging was finished. Ozone flow was lead again into potassium iodide solution (10 min), and pulp was rinsed with oxygen flow 10 minutes after the reaction time. Ozone formation was determined from potassium iodide solution by titration with Na₂S₂O₃.

Hydrogen peroxide treatment

P-stage was performed in Teflon coated medium consistency reactor. Preheated pulp was added into the reactor, and after that, reagents were charged, suspension was mixed and initial pH was measured. Mixing speed was 210 r/min. After reaction time pH was measured from the pulp in the reaction temperature, and residual hydrogen peroxide content of the filtrate was determined.

25 Alkaline extraction treatment

E-stage was carried out in a plastic jar. Pulp and chemicals was preheated to the reaction temperature and initial pH was measured. Pulp suspension was mixed every 15 minutes. After the reaction time pH was measured from the pulp in the reaction temperature.

30 Pulp washing

Washing between stages was a standard laboratory washing: Pulp was diluted to 5% consistency with deionized water, which temperature was the same as that of the preceding bleaching stage. After dewatering, the pulp was washed two times with cold deionized water with amount equivalent to ten times the absolutely dry pulp amount.

WO 2016/193542 12 PCT/FI2016/050375

The intrinsic viscosity of the pulp was determined by a standard ISO 5351-1. The viscosity test is a means for determining the extent of cellulose degradation produced by cooking and bleaching. The limiting viscosity number of cellulose is determined in dilute cupriethylene-diamine (CED) solution. First the pulp sample is continuously shaken in flask containing deionized water and copper pieces until the sample has been completely disintegrated. Then the CED solution is added and shaking is continued until the sample has been dissolved. After this the efflux time of the sample is determined with viscometer. The measurement programme of the viscometer gives automatically the intrinsic viscosity value of the sample.

10

15

20

5

Dissolving cellulose to DMAc/LiCl system

Cellulose was dissolved by a method described by Sjöholm et al 2000. Cellulose (2-5 wt-%) was added via solvent-exchange (water/methanol/DMAc sequence) to 5% LiCl/DMAc solution. The mixture was heated to 80 °C for 2 hours and allowed to slowly cool to room temperature. A uniformly transparent cellulose solution was observed.

Preparation of cellulose palmitate using homogenous method

The homogenous esterifications of the cellulose were conducted by using a method, in which cellulose was first dissolved via solvent-exchange to DMAc/5% LiCl solution. Then anhydrous pyridine (3.6 equivalents to cellulose AGU) was mixed with cellulose solution. Finally palmitoyl chloride (3.0 equivalents to cellulose AGU) was added slowly to the cellulose mixture. The mixture was then warmed to 60 °C and mixing was continued for 16 h at a constant temperature. The product was then precipitated with ethanol, filtered and additionally washed with ethanol and acetone.

25

30

Preparation of cellulose palmitate using heterogenous method

Cellulose and anhydrous pyridine (20 equivalents to cellulose AGU) were mixed together and palmitoyl chloride (3 equivalents to cellulose AGU) was added slowly to the cellulose mixture. The mixture was stirred either overnight at 60 °C or 5h at 100 °C temperature. The product was then precipitated using ethanol, filtered and additional washed with ethanol and with acetone.

WO 2016/193542 13 PCT/FI2016/050375

EXAMPLE 2 – Properties of molar mass controlled cellulose

Molar mass controlled cellulose prepared as described above was characterized by using methods generally known in the art. Table 1 shows properties of the initial pulp, and after ozone, hydrogen peroxide and alkaline treatments. Based on SEC measurements hydrolysis was successful in reducing molar mass from 520 kDa to 58 kDa with lower polydispersity.

Table 1.

5

	Softwood	Pulp 1	Pulp 2	Pulp 3
	sulphite pulp			
Mn, kDa	56	28	15	13
Mw, kDa	520	185	80	58
PD	9.3	5.8	6.3	4.3

The degree of substitution (DS) of the samples was analysed using solid state ¹³C CP/MAS NMR spectroscopy by comparing the carbonyl carbon integrals with cellulose C1 signal integral with the aid of signal deconvolution. According to the NMR, the DS values of prepared cellulose esters ranged from 0.2 to 1.3 (Table 2).

Table 2.

Entry	Cellulose	Method	Amount ^c	DS ^d
			(molar ratio)	
1	D	homogenous	3:1	0.2
2	Z1	homogenous	3:1	1.0
3	Z1	homogenous ^b	3:1	1.1
4	ZP2	homogenous	3:1	1.3
5	ZP2	homogenous	1.5:1	0.1
6	D	heterogenous	3:1	0.8
7 ^e	Z1	heterogenous	3:1	0.5
8	Z1	heterogenous	3:1	1.0
9	Z1	heterogenous	6:1	1.3
10	ZP2	heterogenous	3:1	1.2

WO 2016/193542 14 PCT/FI2016/050375

11	ZP2	heterogenous	6:1	1.2
12	ZP3	homogenous ^a	3:1	1.3
13	ZP3	homogenous ^a	3:1	1.3
14	ZP3	homogenous ^a	3:1	1.2
15	ZP3	homogenous ^a	3:1	0.9
16	ZP3	homogenous ^a	3:1	1.0
17	ZP3	homogenous ^a	3:1	0.9
18	ZP3	homogenous ^a	3:1	0.8

Reactions were conducted at 100 °C for 5h.

10

15

EXAMPLE 3 – Heat sealability of molar mass controlled cellulose in single and multilayered coatings

Heat sealability of the films was determined using the sealing strength tester (Labormaster HTC 3000, Willi Kopp, Germany). The sealing strength was measured after sealing at 170 °C or 200 °C with a sealing force of 850 kPa, a sealing time of 3 s, a delay time of 20 s, and a peeling rate of 12 m/min. Width of the sample strips was 2 cm. Table 3 presents the heat seal strength of molar mass controlled cellulose ester films expressed as N/m.

Table 3.

Film	Sealing temperature	Sealing strength (N/m)	Standard
	(°C)		deviation (N/m)
C6	170	132	30
C8	170	183	24

^a 2 wt-% cellulose in 5% LiCl/DMAc solution

^b 5 wt-% cellulose in 5% LiCl/DMAc solution

^c Molar ratio of palmitoyl chloride vs anhydroglucose unit (AGU)

⁵ d according to ¹³C CP/MAS NMR

e 60°C, 16h

WO 2016/193542 15 PCT/FI2016/050375

C10	170	0	0
C14	200	265	170
C16	200	290	127
C18	200	0	0

n=2-3

5

10

15

20

Cellulose nanofibrils (CNF) films plasticized with 30% sorbitol were coated (both sides) by immersing in cellulose palmitate WLL VII94B, 2 % in choloroform after which they were dried at ambient conditions. The thickness of CNF film was measured to be 40 um and cellulose palmitate/CNF/cellulose palmitate was 70 um. WVTR of CNF decreased from 850 to 90 g/m²/d by cellulose palmitate coating. OTR of cellulose palmitate decreased from 36000 to 55 cm³/m²/d by incorporating with CNF. OTR of CNF film decreased slightly due to cellulose palmitate coating most likely because of hydrophobic cellulose palmitate layer protects CNF film form swelling.

CNF films were pre-activated using Tantec corona and then coated with C6 (10%), C8 (10%), C14 (7.5%) and C16 (10%) in chloroform using Mayer bar (wet film deposit 100 μ m). Coatings were applied either once (dry thickness 10 μ m) or three times (dry thickness 30 μ m).

Heat sealability of the coated films was determined using the sealing strength tester (Labormaster HTC 3000, Willi Kopp, Germany). The sealing strength was measured after sealing at 170 °C or 200 °C with a sealing force of 850 kPa, a sealing time of 10 s, a delay time of 20 s, and a peeling rate of 0.2 m/min. Width of the sample strips was 5 cm. Table 4 presents the heat seal strength of multilayered films expressed as N/m.

Table 4.

Coated film	Sealing temperature	Sealing strength (N/m)	Standard
	(°C)		deviation (N/m)
CNF + C6 10μm	170	0	0
CNF + C6 30µm	170	56	9
CNF + C8 10μm	170	0	0
CNF + C8 30µm	170	56	18

WO 2016/193542 16 PCT/FI2016/050375

CNF + C14 10μm	200	0	0
CNF + C14 30μm	200	65	25
CNF + C16 10μm	200	0	0
CNF + C16 30μm	200	95	31

n=4-6

5 CITATION LIST

Non-patent literature:

Edgar K., Buchanan C., Debenham J., Rundquist .P, Seiler B., Shelton M., Tindall S., *Prog. Polym. Sci.* 2001, 26:1605-1688.

Klemm D., Schmauder H.-P., Heinze T., Cellulose, Biopolymers, 2002. 6: p. 275-319.

10 Wei Y., Cheng F., Hou G., J. Sci. Ind. Res. 2007, 66: 1019-1024.

Crepy L., Chaveriat L., Banoub J., Martin P., Joly N., ChemSusChem, 2009, 2(2), pp. 165-170.

Sjöholm E, Gustafsson K, Erikssin B, Brown W, Colmsjö A (2000) Aggregation of cellulose in lithium chloride/N,N-dimethylacetamide, Polymers 41:153-161

CLAIMS:

5

10

15

30

1. A method of improving the reactivity of cellulose and preparing a transportable form of the previous for preparation of thermoplastic, dispersion able or dissolving derivatives, **characterized** in that molar mass and molar mass distribution of the cellulose is controlled uniformly to a range between 30 and 300 kDa providing reactive and processable cellulose.

- 2. The method of claim 1, **characterized** by controlling molar mass of cellulose raw material via hydrolysis, excluding total hydrolysis.
- 3. The method of claim 1 or 2, **characterized** by selecting cellulose raw material from native softwood pulp, native hardwood pulp, annual plant pulp, softwood sulphite dissolving grade pulp, hardwood sulphite dissolving grade pulp, ozone treated hydrolyzed pulp or enzyme treated pulp
- 4. The method of any preceding claim, **characterized** by controlling hydrolysis in such manner that after the hydrolysis molar mass and molar mass distribution of the cellulose is in a range between 40 and 200 kDa.
- 5. The method of any preceding claims, **characterized** in that the cellulose is hydrolyzed by enzymatic treatment, ozone treatment, hydrogen peroxide treatment, alkaline treatment or other chemical treatment, before performing a long chain fatty acid modification.
- 6. The method of any preceding claims, **characterized** in that a long chain fatty acid modification comprises heterogeneous or homogenous esterification of the molar mass controlled cellulose.
 - 7. The method of any preceding claims, **characterized** in that a long chain fatty acid modification comprises heterogeneous etherification of the molar mass controlled cellulose.
 - 8. A molar mass controlled cellulose ester having chain length between C6 and C18 and a total degree of substitution (DS) from 0.7 to 3.

- 9. The ester of claim 8 having a Young's modulus value of at least 300 MPa, more preferably at least 400 MPA.
- 10. The ester of claim 8 or 9 produced by the method of any of claims 1 to 6.

5

- 11. A molar mass controlled cellulose ether having chain length between C6 and C18 and a total degree of substitution (DS) from 0.7 to 3.
- 12. The ether of claim 11 produced by the method of any of claims 1 to 6.

10

20

30

- 13. A method of producing thermoformable cellulose ester films, **characterized** by preparing the films from the purified molar mass controlled C6-C18 cellulose esters or ethers, obtained by the method of any of claims 1 to 6.
- 15 14. The method of claim 13, **characterized** in that no external plasticizers are used.
 - 15. A method of producing multilayered film structure, **characterized** by coating a CNF film from both sides by immersing molar mass controlled C6-C18 cellulose obtained by the method of any of claims 1 to 6 into a multilayered, such as two or three layered, cellulose film structure.
 - 16. The method of claim 15, **characterized** in that no external plasticizers are used.
- 17. A heat-sealable film structure having a dry thickness of at least 10 μm and produced by the method of any of claims 13 to 16.
 - 18. Use of the molar mass controlled cellulose material, obtained by the method of claims 1-6, 13-14, or 15-16, in film applications, heat-sealable packaging applications, foam applications, electronical coating applications, medical applications, composite applications, non-woven applications, fiber spinning and thermoforming, in/for textiles, fibers, filaments, yarns and cloths, and in printed electronics, sensors, solar cells and diagnostics.

 $R^{=}$ COCH₃, CO (CH₂)_n CH₃ or H where n = 2 30

FIG. 1

 $R^{=-}CH_{2}CH_{2}OR_{1}R^{"-}OR_{1}H$ $R^{'=}HOTCH_{2}CH_{2}OR_{1}R^{"-}R^{"}$ $R^{"=}HCH_{3}OT(CH_{2})nCH_{3}$ $Whrere n=_{1}20$

FIG. 2

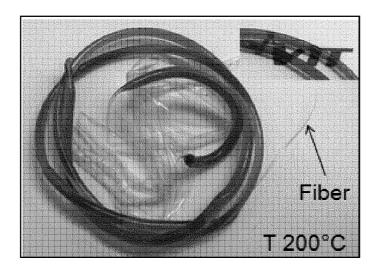


FIG. 3
SUBSTITUTE SHEET (RULE 26)

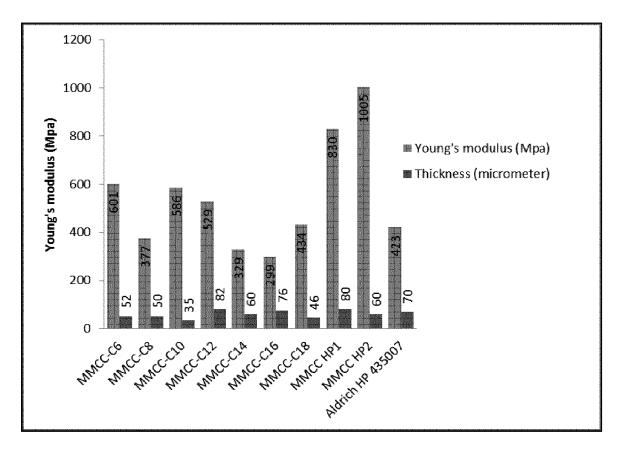


FIG. 4A

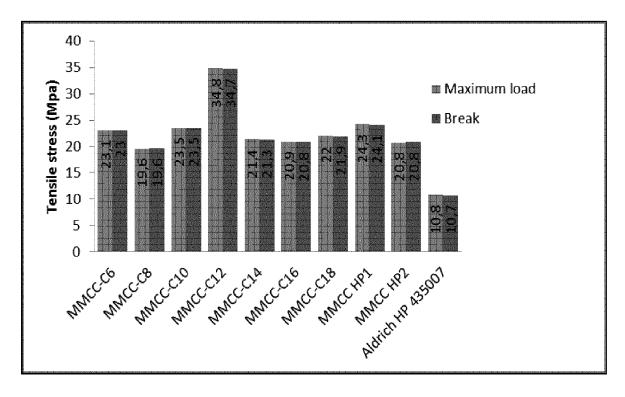


FIG. 4B SUBSTITUTE SHEET (RULE 26)

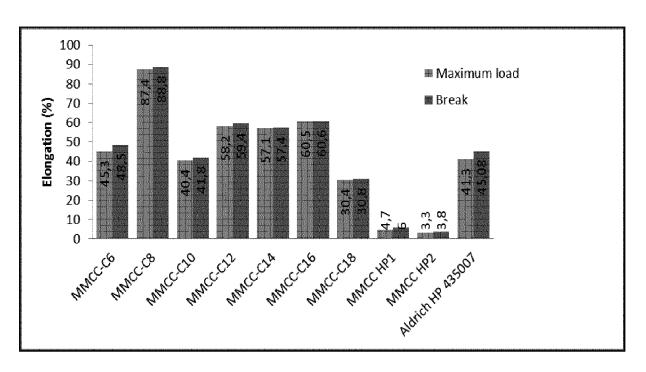


FIG. 4C

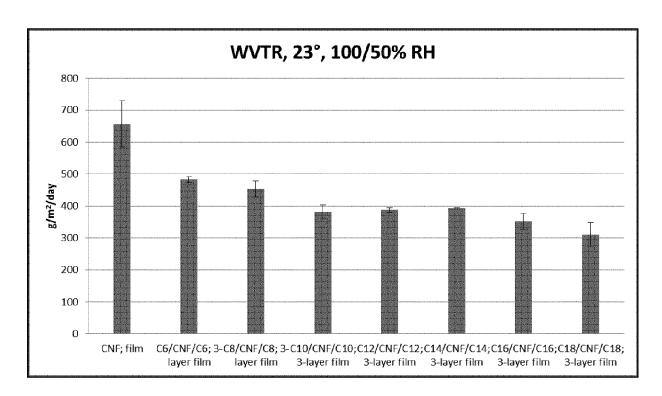


FIG. 5

WO 2016/193542 PCT/FI2016/050375

4/4

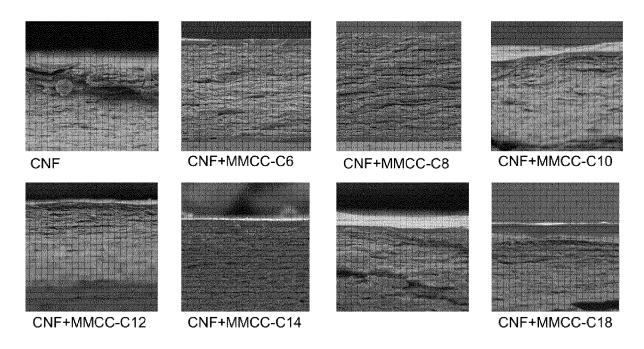


FIG. 6

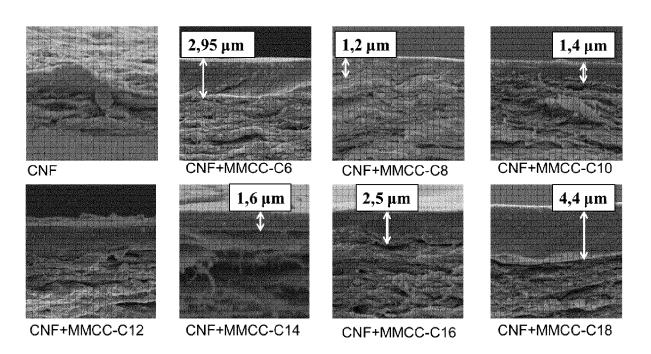


FIG. 7

INTERNATIONAL SEARCH REPORT

International application No.

PCT/FI2016/050375

A. CLASSIFICATION OF SUBJECT MATTER

See extra sheet

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC: C08B, C08L

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

FI, SE, NO, DK

Electronic data base consulted during the international search (name of data base, and, where practicable, search terms used)

EPODOC, WPIAP, EPO-Internal full-text databases, Full-text translation databases from Asian languages, PRH Internal, XPESP, XPIPCOM, XPMISC, XPOAC, XPRD, BIOSIS, COMPDX, EMBASE, MEDLINE, PUBCOMP, PUBSUBS, TDB, NPL

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Х	WO 2010019245 A1 (EASTMAN CHEM CO [US]) 18 February 2010 (18.02.2010) paragraphs [0011], [0051], [00183], example 12, claims	1, 3, 6, 8-16, 18
Х	EP 2489681 A1 (SHINETSU CHEMICAL CO [JP]) 22 August 2012 (22.08.2012) examples, claims	1-5, 7
Х	US 4228277 A (LANDOLL LEO M) 14 October 1980 (14.10.1980) examples, claims	1, 7
X	WO 02086206 A1 (WEYERHAEUSER CO [US]) 31 October 2002 (31.10.2002) examples, tables, claims 50 and 69	1-4

×	Further doc	uments are listed in the continuation of Box C.		X	See patent family annex.	
*	Special catego	ries of cited documents:	"T"		locument published after the inter	
"A"	document defi to be of particu	ning the general state of the art which is not considered alar relevance			nd not in conflict with the application inciple or theory underlying the inciple or theory underlying the inciple application.	
"E"	earlier applicate	tion or patent but published on or after the international	"X"		nent of particular relevance; the ci dered novel or cannot be consider	
"L"	document which	ch may throw doubts on priority claim(s) or which		when	the document is taken alone	
		blish the publication date of another citation or other	"Y"		nent of particular relevance; the ci	
"O"	special reason	(as specified) rring to an oral disclosure, use, exhibition or other mea	ac		nsidered to involve an inventive st	
ייקיי		lished prior to the international filing date but later than			ined with one or more other such obvious to a person skilled in the	
Г	the priority dat	1	"&"	_	ment member of the same patent fa	
	tare parently than	1	α	docum	ment member of the same patent is	anniy
Date of	of the actual c	ompletion of the international search	Date	of mai	ling of the international searc	ch report
	14 Se	eptember 2016 (14.09.2016)			16 September 2016 (1	6.09.2016)
Nam	e and mailing	address of the ISA/FI	Auth	orized	lofficer	
	_	nd Registration Office	Λn	tti Hoil	kkala	
		il-00101 HELSINKI, Finland	All	ווטדו וווי	nnaia	

Telephone No. +358 9 6939 500

Facsimile No. +358 9 6939 5328

INTERNATIONAL SEARCH REPORT

International application No.

PCT/FI2016/050375

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT			
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No	
X A	WO 2013021099 A1 (TEKNOLOGIAN TUTKIMUSKESKUS VTT [FI]) 14 February 2013 (14.02.2013) claims	17 1-16, 18	

INTERNATIONAL SEARCH REPORT Information on Patent Family Members

Patent document cited in search report	Publication date	Patent family members(s)	Publication date
WO 2010019245 A1	18/02/2010	CN 101657428 A	24/02/2010
		CN 101657470 A	24/02/2010
		CN 101657470 B	05/09/2012
		CN 101668779 A	10/03/2010
		CN 101668779 B	20/11/2013
		CN 102124031 A	13/07/2011
		CN 102124031 B	14/05/2014
		CN 102603643 A	25/07/2012
		CN 102675207 A	19/09/2012
		CN 102977216 A	20/03/2013
		CN 103467606 A	25/12/2013
		CN 103923352 A	16/07/2014
		EP 2118070 A1	18/11/2009
		EP 2118143 A1	18/11/2009
		EP 2121766 A1	25/11/2009
		EP 2247622 A1	10/11/2010
		EP 2313438 A1	27/04/2011
		EP 2313439 A1	27/04/2011
		JP 2010518244 A	27/05/2010
		JP 5558832 B2	23/07/2014
		JP 2014012852 A	23/01/2014
		JP 5860014 B2	16/02/2016
		JP 2010518166 A	27/05/2010
		JP 2010518245 A	27/05/2010
		JP 2011530643 A	22/12/2011
		KR 20090109556 A	20/10/2009
		KR 101551500 B1	08/09/2015
		KR 20090109106 A	19/10/2009
		KR 20090109107 A	19/10/2009
		KR 20110043757 A	27/04/2011
		US 2008194834 A1	14/08/2008
		US 7919631 B2	05/04/2011
		US 2008194808 A1	14/08/2008
		US 8148518 B2	03/04/2012
		US 2008194807 A1	14/08/2008
		US 8153782 B2	10/04/2012
		US 2009203898 A1	13/08/2009
		US 8158777 B2	17/04/2012
		US 2011306760 A1	15/12/2011
		US 8273872 B2	25/09/2012
		US 2011213138 A1	01/09/2011
		US 8354525 B2	15/01/2013
		US 2013190485 A1	25/07/2013
		US 9175096 B2	03/11/2015
		US 2010029927 A1	04/02/2010
		US 2012095207 A1	19/04/2012

INTERNATIONAL SEARCH REPORT Information on Patent Family Members

Patent document cited in search report	Publication date	Patent family members(s)	Publication date
		US 2012101269 A1	26/04/2012
		US 2012123112 A1	17/05/2012
		US 2012142910 A1	07/06/2012
		US 2016108137 A1	21/04/2016
		WO 2008100566 A1	21/08/2008
		WO 2008100569 A1	21/08/2008
		WO 2008100577 A1	21/08/2008
		WO 2009102306 A1	20/08/2009
		WO 2010019244 A1	18/02/2010
EP 2489681 A1	22/08/2012	CN 102643352 A	22/08/2012
1000017(1	OO;O 1	CN 102643352 B	19/11/2014
		IN 453DE2012 A	05/06/2015
		JP 2012172037 A	10/09/2012
		JP 5671367 B2	18/02/2015
		KR 20120095798 A	29/08/2012
		MX 20120093798 A	30/08/2012
		US 2012214981 A1	23/08/2012
		US 9371399 B2	21/06/2016
US 4228277 A	14/10/1980	US 4228277 B1	20/10/1992
		CA 1140541 A	01/02/1983
		DE 3004161 A1	21/08/1980
		DE 3004161 C2	28/06/1990
		GB 2043646 A	08/10/1980
		GB 2043646 B	08/12/1982
		JP S55110103 A	25/08/1980
		JP H0128041 B2	31/05/1989
		NL 8000786 A	14/08/1980
		NL 188649 B	16/03/1992
		NL 188649 C	17/08/1992
WO 02086206 A1	31/10/2002	AT 225418 T	15/10/2002
		AT 255178 T	15/12/2003
		AT 311487 T	15/12/2005
		AT 311488 T	15/12/2005
		AT 335873 T	15/09/2006
		AT 376597 T	15/11/2007
		AT 384809 T	15/02/2008
		AT 497035 T	15/02/2011
		AU 2893099 A	11/10/1999
		AU 5339901 A	26/11/2001
		AU 5363101 A	07/11/2001

INTERNATIONAL SEARCH REPORT Information on Patent Family Members

Patent document cited in search report	Publication date	Patent family members(s)	Publication date
		AU 5583901 A	20/11/2001
		BR 0110198 A	11/02/2003
		BR 0110198 B1	06/09/2011
		BR 0110662 A	25/03/2003
		BR 0110890 A	30/12/2003
		BR 0110890 B1	22/02/2012
		BR 0305986 A	17/08/2004
		BR 0305986 B1	02/04/2013
		BR 0305987 A	17/08/2004
		BR 0305988 A	17/08/2004
		BR 0305991 A	17/08/2004
		BR 0305991 B1	26/11/2013
		BR 9711352 A	18/01/2000
		BR 9908775 A	14/11/2000
		BR 9908775 B1	05/05/2009
		CA 2264180 A1	26/02/1998
		CA 2264180 C	01/09/2009
		CA 2323437 A1	23/09/1999
		CA 2323437 C	18/01/2005
		CA 2405091 A1	01/11/2001
		CA 2405091 C	30/06/2009
		CA 2406517 A1	22/11/2001
		CA 2406517 C	30/06/2009
		CA 2406550 A1	15/11/2001
		CA 2406550 C	25/08/2009
		CA 2452944 A1	02/07/2004
		CA 2452944 C	28/11/2006
		CA 2453241 A1	02/07/2004
		CA 2453241 C	29/01/2008
		CA 2453246 A1	02/07/2004
		CA 2453246 C	26/02/2008
		CA 2453276 A1	02/07/2004
		CA 2453276 C	31/10/2006
		CA 2641970 A1	26/02/1998
		CA 2641972 A1	26/02/1998
		CA 2641972 C	13/11/2012
		CN 1238015 A	08/12/1999
		CN 1081684 C	27/03/2002
		CN 1293723 A	02/05/2001
		CN 1108401 C	14/05/2003
		CN 1425081 A	18/06/2003
		CN 1191395 C	02/03/2005
		CN 1446954 A	08/10/2003
		CN 1200152 C	04/05/2005
		CN 1446953 A	08/10/2003
		CN 1219918 C	21/09/2005
		CN 1446955 A	08/10/2003

INTERNATIONAL SEARCH REPORT Information on Patent Family Members

Patent document cited in search report	Publication date	Patent family members(s)	Publication date
		CN 1219919 C	21/09/2005
		CN 1522317 A	18/08/2004
		CN 1224736 C	26/10/2005
		CN 1356412 A	03/07/2002
		CN 1230579 C	07/12/2005
		CN 1517487 A	04/08/2004
		CN 1244734 C	08/03/2006
		CN 1519424 A	11/08/2004
		CN 1306111 C	21/03/2007
		CN 1348023 A	08/05/2002
		CN 1356413 A	03/07/2002
		CN 1356414 A	03/07/2002
		CN 1432087 A	23/07/2003
		CN 100402741 C	16/07/2008
		CN 1505703 A	16/06/2004
		CN 100343427 C	17/10/2007
		CN 1517490 A	04/08/2004
		CN 1517491 A	04/08/2004
		CN 100460594 C	11/02/2009
		DE 60122169 D1	21/09/2006
		DE 60122169 T2	23/08/2007
		DE 60131077 D1	06/12/2007
		DE 60302539 D1	05/01/2006
		DE 60302539 T2	13/07/2006
		DE 60302540 D1	05/01/2006
		DE 60302540 T2	10/08/2006
		DE 60302540 T3	17/07/2008
		DE 60303900 D1	04/05/2006
		DE 60303900 T2	12/10/2006
		DE 60318810 D1	13/03/2008
		DE 60318810 T2	21/05/2008
		DE 69716092 D1	07/11/2002
		DE 69716092 T2	30/01/2003
		DE 69913117 D1	08/01/2004
		DE 69913117 T2	26/08/2004
		DE 69943167 D1	10/03/2011
		EP 0920548 A1	09/06/1999
		EP 0920548 B1	02/10/2002
		EP 1068376 A1	17/01/2001
		EP 1068376 B1	26/11/2003
		EP 1285110 A1	26/02/2003
		EP 1287191 A1	05/03/2003
		EP 1287191 B1	24/10/2007
		EP 1311717 A2	21/05/2003
		EP 1311717 A2 EP 1311717 B1	09/08/2006
		EP 1362935 A1	19/11/2003
		EP 1362935 B1	26/01/2011

INTERNATIONAL SEARCH REPORT Information on Patent Family Members

Patent document cited in search report	Publication date	Patent family members(s)	Publication date
		EP 1402092 A1	31/03/2004
		EP 1435403 A1	07/07/2004
		EP 1435403 B1	30/11/2005
		EP 1435404 A1	07/07/2004
		EP 1435404 B1	08/03/2006
		EP 1437428 A1	14/07/2004
		EP 1437428 B1	30/11/2005
		EP 1437428 B2	27/02/2008
		EP 1441050 A1	28/07/2004
		EP 1441050 B1	23/01/2008
		ES 2185045 T3	16/04/2003
		ES 2214015 T3	01/09/2004
		ES 2259751 T3	16/10/2006
		ES 2265428 T3	16/02/2007
		HK 1023377 A1	26/07/2002
		JP 2004211281 A	29/07/2004
		JP 3965153 B2	29/08/2007
		JP 2004211283 A	29/07/2004
		JP 3965154 B2	29/08/2007
		JP 2004211282 A	29/07/2004
		JP 4012878 B2	
			21/11/2007
		JP 2004211280 A	29/07/2004
		JP 4015109 B2	28/11/2007
		JP 2001501260 A	30/01/2001
		JP 4018152 B2	05/12/2007
		JP 2007046223 A	22/02/2007
		JP 4134209 B2	20/08/2008
		JP 2003531313 A	21/10/2003
		JP 4593865 B2	08/12/2010
		JP 2012046861 A	08/03/2012
		JP 5491477 B2	14/05/2014
		JP 2014074261 A	24/04/2014
		JP 5752215 B2	22/07/2015
		JP 2002506931 A	05/03/2002
		JP 2003532806 A	05/11/2003
		JP 2003533602 A	11/11/2003
		JP 2009019326 A	29/01/2009
		KR 20000068304 A	25/11/2000
		KR 100471549 B1	07/03/2005
		KR 20040062397 A	07/07/2004
		KR 100547710 B1	31/01/2006
		KR 20030092092 A	03/12/2003
		KR 100558502 B1	07/03/2006
		KR 20040062398 A	07/07/2004
		KR 100564515 B1	29/03/2006
		KR 20040062400 A	07/07/2004
		KR 100564516 B1	29/03/2006

INTERNATIONAL SEARCH REPORT Information on Patent Family Members

Patent document cited in search report	Publication date	Patent family members(s)	Publication date
		KR 20040062399 A	07/07/2004
		KR 100566200 B1	29/03/2006
		KR 20030004395 A	14/01/2003
		KR 100750008 B1	16/08/2007
		KR 20030013421 A	14/02/2003
		KR 100834248 B1	30/05/2008
		PA 02010371	25/04/2003
		PA 02011104	10/03/2003
		PA 02011317	25/04/2003
		PA 04000070	08/07/2004
		PA 04000071	08/07/2004
		PA 04000072	08/07/2004
		PA 04000073	08/07/2004
		RU 2003131266 A	10/05/2005
		RU 2268327 C2	20/01/2006
		TW 452615 B	01/09/2001
		TW 561205 B	11/11/2003
		TW 573088 B	21/01/2004
		TW 200426260 A	01/12/2004
		TW 1237072 B	01/08/2005
		TW 200424371 A	16/11/2004
		TW I238861 B	01/09/2005
		TW 200413582 A	01/08/2004
		TW I251040 B	11/03/2006
		TW 200420799 A	16/10/2004
		TW I259859 B	11/08/2006
		TW 1282829 B	21/06/2007
		US 6210801 B1	03/04/2001
		US 6221487 B1	24/04/2001
		US 6235392 B1	22/05/2001
		US 6306334 B1	23/10/2001
		US 6331354 B1	18/12/2001
		US 2002034638 A1	21/03/2002
		US 6440523 B1	27/08/2002
		US 2002076556 A1	20/06/2002
		US 6440547 B1	27/08/2002
		US 2002064654 A1	30/05/2002
		US 6444314 B1	03/09/2002
		US 2001028955 A1	11/10/2001
		US 6471727 B2	29/10/2002
		US 2002088572 A1	11/07/2002
		US 6491788 B2	10/12/2002
		US 6511930 B1	28/01/2003
		US 2002081428 A1	
		US 6514613 B2	27/06/2002
			04/02/2003
		US 2002155292 A1	24/10/2002
		US 6528163 B2	04/03/2003

INTERNATIONAL SEARCH REPORT Information on Patent Family Members

Patent document cited in search report	Publication date	Patent family members(s)	Publication date
		US 6596033 B1	22/07/2003
		US 6605350 B1	12/08/2003
		US 2003186053 A1	02/10/2003
		US 6685856 B2	03/02/2004
		US 2003186054 A1	02/10/2003
		US 6686039 B2	03/02/2004
		US 2003186055 A1	02/10/2003
		US 6686040 B2	03/02/2004
		US 2002037407 A1	28/03/2002
		US 6692827 B2	17/02/2004
		US 2002060382 A1	23/05/2002
		US 6706237 B2	16/03/2004
		US 2002036070 A1	28/03/2002
		US 6706876 B2	16/03/2004
		US 2002160186 A1	31/10/2002
		US 6773648 B2	10/08/2004
		US 2003183351 A1	02/10/2003
		US 6797113 B2	28/09/2004
		US 2003025252 A1	06/02/2003
		US 6861023 B2	01/03/2005
		US 2002148050 A1	17/10/2002
		US 7067444 B2	27/06/2006
		US 2002041961 A1	11/04/2002
		US 7083704 B2	01/08/2006
		US 2003025251 A1	06/02/2003
		US 7090744 B2	15/08/2006
		WO 0181664 A1	01/11/2001
		WO 0186043 A1	15/11/2001
		WO 0188236 A2	22/11/2001
		WO 9807911 A1	26/02/1998
		WO 9947733 A1	23/09/1999
		ZA 9902023 B	27/09/1999
		ZA 200309826 B	13/08/2004
		ZA 200309829 B	22/06/2004
		ZA 200309831 B	12/08/2004
		ZA 200309833 B	11/08/2004
WO 2013021099 A1	14/02/2013	EP 2688914 A2	29/01/2014
		EP 2739782 A1	11/06/2014
		US 2014088252 A1	27/03/2014
		US 2014230691 A1	21/08/2014
		WO 2012127119 A2	27/09/2012

INTERNATIONAL SEARCH REPORT

	PC1/F12016/050375
CLASSIFICATION OF SUBJECT MATTER	
IPC C08B 3/10 (2006.01) C08B 11/02 (2006.01) C08B 1/02 (2006.01) C08L 1/10 (2006.01) C08L 1/28 (2006.01)	