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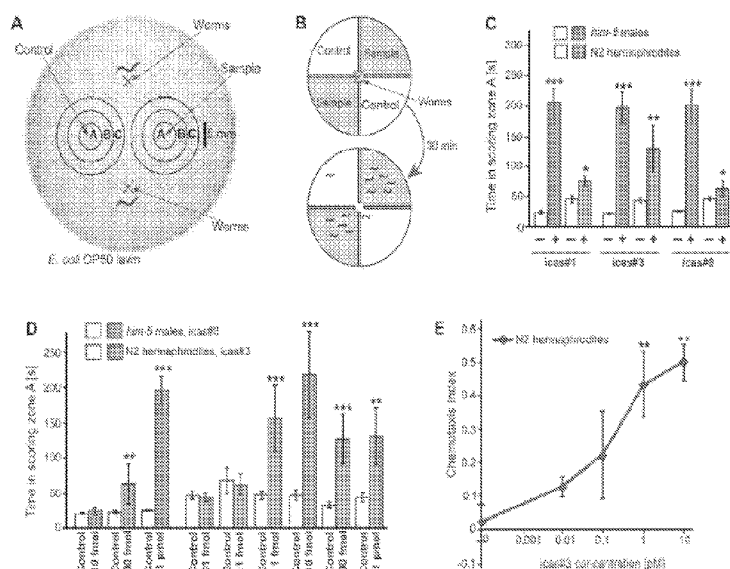
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(54) Title: SMALL MOLECULE COMPOUNDS THAT CONTROL PLANT- AND INSECT-PATHOGENIC NEMATODES

(57) Abstract: The present invention relates to methods of modifying nematode behavior or using certain isolated modulator compounds. Also disclosed are methods of promoting or inhibiting reproduction in a nematode population, methods of promoting or inhibiting nematode aggregation at a first location, and methods of treating or preventing parasite infection of a plant.



Figures 1A-E



NO, NZ, OM, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

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SMALL MOLECULE COMPOUNDS THAT CONTROL PLANT- AND INSECT-PATHOGENIC NEMATODES

[0001] This invention was made with U.S. Government support under Grant Nos. GM088290, GM085285, and T32GM008500 awarded by the National Institutes of Health. The U.S. Government has certain rights in this invention.

FIELD OF THE INVENTION

[0002] The present invention relates to methods of modifying nematode behavior.

BACKGROUND OF THE INVENTION

[0003] Communication among individuals of a species relies on a number of different sensory inputs including chemical, mechanical, auditory, or visual cues (EDWARD O. WILSON, *SOCIOBIOLOGY: THE NEW SYNTHESIS* (25th anniv. ed. 2000)). Chemical signaling is perhaps the most ancient form of interorganismal communication (EDWARD O. WILSON, *SOCIOBIOLOGY: THE NEW SYNTHESIS* (25th anniv. ed. 2000); Wyatt, *Nature* 457:262-63 (2009)), and analysis of the chemical signals and the behaviors they mediate is of great significance for understanding the ecological and evolutionary dynamics of intra- and inter-specific interactions.

[0004] Nematodes are among the most abundant animals in the world, by individual count (SIEVERT LORENZEN: *THE PHYLOGENETIC SYSTEMATICS OF FREELIVING NEMATODES* (The Ray Society, London, 1994)). They inhabit sulfurous sediment, deep-sea trenches, mammals, insects, plants, arctic ice, and many other habitats, making them one of the most successful groups of animals on earth (Nussbaumer et al., *Aquat. Microb. Ecol.* 34:239-46 (2004); ISTVÁN ANDRASSY: *EVOLUTION AS A BASIS FOR THE SYSTEMATIZATION OF NEMATODES* (Budapest, 1976); Tietjen, *Deep-Sea Res.* 36:1579-94 (1989); Aben-Athar, *Rev. Bras. Med.* 2:89-94 (1951); Lutz, *Weitere Mittheilungen* 3: 425-28 (1888); Blaxter, *Science* 282:2041-46 (1998)). Many nematode behaviors have been studied, such as mate finding in roots, bacteria, sand, agar, and intestines (DONALD L. LEE: *THE BIOLOGY OF NEMATODES* (CRC Press, London, 2002)). Little is known about nematode pheromone systems. Although there have been many attempts to identify nematode pheromones (DONALD L. LEE: *THE BIOLOGY OF NEMATODES* (CRC Press, London, 2002)), identification has only been successful in very few species (Jaffe et al., *J. Chem. Ecol.* 15:2031-43 (1989); Srinivasan et

al., *Nature* 454:1115-18 (2008); Jeong, et al., *Nature* 433:541-45 (2005); Butcher et al., *Nat. Chem. Biol.* 7:420-22 (2007); Pungaliya et al., *PNAS* 19:7708-13 (2009)).

[0005] The free-living nematode *C. elegans* is used extensively as a model system for social behaviors such as foraging, population density sensing, mating, and aggregation (de Bono & Maricq, *Annu. Rev. Neurosci.* 28:451-501 (2005)). Entomopathogenic nematodes (EPN), such as *Heterorhabditis* spp. and *Steinernema* spp. are obligate insect parasites that kill and consume their hosts with the aid of symbiotic bacteria (Kaya & Gaugler, *Annu. Rev. Entomol.* 38:181-206 (1993)). *C. elegans* is typically found in decomposing plant material (Barriere & Felix, *Curr. Biol.* 15:1176-84 (2005)). It completes its life cycle within 3.5 days under standard laboratory conditions. When the conditions are not favorable for normal growth and development, such as high temperature, high density, or low food availability, it forms dauer larvae (alternative 3rd larvae). This specialized life stage, which does not feed and is resistant to stressful conditions (Golden & Riddle, *Dev. Biol.* 102:368-78 (1984); Golden & Riddle, *Science* 218:578-80 (1982); Golden & Riddle, *PNAS* 81:819-23 (1984)), is analogous to infectious juveniles (IJs) of entomopathogenic nematodes and second stage juveniles (J2) of plant parasitic root-knot nematodes (PAUL DE LEY: A QUICK TOUR OF NEMATODE DIVERSITY AND THE BACKBONE OF NEMATODE PHYLOGENY (WormBook, ed. The *C. elegans* Research Community, 2006)) (*Meloidogyne* spp) that are equally adapted to survive conditions unfavorable for normal growth and development. Upon depletion of food resources or when overcrowded, IJs of entomopathogenic nematodes and dauer larvae of the free living nematode *Caenorhabditis elegans* display a dispersal behavior (Golden & Riddle, *Dev. Biol.* 102:368-78 (1984); Rolston et al., *J. Nematol.* 38:221-28 (2006); Abebe et al., *J. Exp. Biol.* 213:3223-29 (2010)).

[0006] For *C. elegans*, entry of early larval stages (L1) into the dauer stage is regulated by a pheromone, called daumone (Jeong et al., *Nature* 433:541-45 (2005)). Subsequent to the identification of daumone, many related compounds have been found in *C. elegans* (Butcher et al., *Nat. Chem. Biol.* 3:420-22 (2007); Butcher et al., *PNAS* 105:14288-92 (2008); Srinivasan et al., *Nature* 454:1115-18 (2008); Butcher et al., *Org. Lett.* 11:3100-03 (2009); Pungaliya et al., *PNAS* 106:7708-13 (2009); von Reuss et al., *J. Am. Chem. Soc.* 134(3):1817-24 (2012)). All of these compounds have the unusual dideoxysugar ascarylose and are as a group called ascarosides.

[0007] Studies in *C. elegans* have shown that this family of small-molecule pheromones regulates gender specific attraction, repulsion, aggregation, olfactory plasticity,

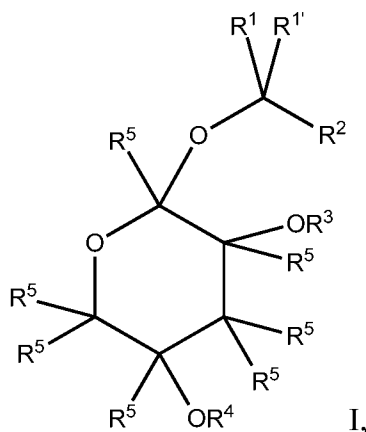
and entry into dauer (a stress-resistant life stage), collectively demonstrating that ascarosides mediate a wide range of *C. elegans* behaviors (Srinivasan et al., *Nature* 454:1115-18 (2008); Macosko et al., *Nature* 458:1171-75 (2009); Yamada et al., *Science* 329:1647-50 (2010); Butcher et al., *Nat. Chem. Biol.* 3:420-22 (2007)). Because ascarosides have not yet been found in any other animal phylum (Bartley et al., *J. Nat. Prod.* 59(10):921-26 (1996)), ascarosides may comprise a nematode-specific chemical code that may regulate important cues for multiple nematode species. Despite their importance for many aspects of *C. elegans* biology, however, knowledge of ascaroside structures, biosynthesis, and homeostasis, as well as the extent to which they may be produced and/or affect other nematodes, remained incomplete.

[0008] The present invention is directed to overcoming these and other deficiencies in the art.

SUMMARY OF THE INVENTION

[0009] A first aspect of the present invention relates to a method of modifying nematode behavior. This method involves administering one or more isolated modulator compounds to the nematode under conditions effective to modify nematode behavior, where the one or more modulator compounds is selected from the group consisting of:

(i) a compound of Formula I:

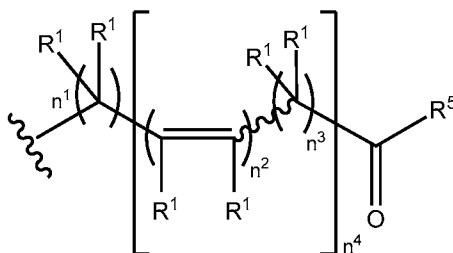


where:

R¹ is H, -C(R)₃, -OR, -N(R)₂, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl; where each R is independently H, halogen, an alkyl, or an alkenyl;

R^1 is absent, H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl; where each R is independently H, halogen, an alkyl, or an alkenyl;

R^2 is a moiety of formula



where:

each R^1 is independently H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl; where each R is independently H, halogen, an alkyl, or an alkenyl;

R^5 is H, $-OH$, $-OR^6$, $-OCR^6R^7R^8$, $-CR^6R^7R^8$, $-NH_2$, $-NHR^6$, $-NR^6R^7$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, an arylalkyl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R^3 or R^4 of another unit of Formula I;

where:

R^6 and R^7 are each independently H, $-CR_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, or a cycloalkenyl, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, or cycloalkenyl is optionally substituted with one or more substituents independently selected from the group consisting of $-OR^8$, $-C(O)R^8$, $-NHC(O)R^8$, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl;

where:

each R is independently H, halogen, an alkyl, or an alkenyl; and R^8 is H, $-C(R)_3$, $-[C(R)_2]_{n4}NHC(O)R^9$, $-[C(R)_2]_{n4}C(O)(NH)R^9$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl,

a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR^9$, $-C(O)R^9$, $-NHC(O)R^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms;

where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR$, $-C(O)R$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; and

R^8 is H, $-C(R)_3$, $-[C(R)_2]_{n^4}NHC(O)R^9$, $-[C(R)_2]_{n^4}C(O)(NH)R^9$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a

cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}^9$, $-\text{C}(\text{O})\text{R}^9$, $-\text{NHC}(\text{O})\text{R}^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms; where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{N}(\text{R})_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{C}(\text{O})\text{R}$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30;

n^1 , n^2 , and n^3 are each independently an integer of 0 to 30;

n^4 is an integer of 1 to 30; and

the sum of n^1 , each n^2 , and each n^3 is 1 to 30;

R^3 and R^4 are each independently H, $-\text{CR}^6\text{R}^7\text{R}^8$, $-\text{C}(\text{O})\text{R}^8$, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R^5 of another unit of Formula I; where:

R^6 and R^7 are each independently H, $-\text{CR}_3$, $-\text{OR}$, $-\text{N}(\text{R})_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a

heterocyclyl, a cycloalkyl, or a cycloalkenyl, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, or cycloalkenyl is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{OR}^8$, $-\text{C}(\text{O})\text{R}^8$, $-\text{NHC}(\text{O})\text{R}^8$, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl;

where:

each R is independently H, halogen, an alkyl, or an alkenyl; and

R^8 is H, $-\text{C}(\text{R})_3$, $-\text{[C}(\text{R})_2\text{]}_{n4}\text{NHC}(\text{O})\text{R}^9$, $-\text{[C}(\text{R})_2\text{]}_{n4}\text{C}(\text{O})(\text{NH})\text{R}^9$, $-\text{OR}$, $-\text{N}(\text{R})_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}^9$, $-\text{C}(\text{O})\text{R}^9$, $-\text{NHC}(\text{O})\text{R}^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms; where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{N}(\text{R})_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{C}(\text{O})\text{R}$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; and

R^8 is H, $-C(R)_3$, $-[C(R)_2]_{n^4}NHC(O)R^9$, $-[C(R)_2]_{n^4}C(O)(NH)R^9$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR^9$, $-C(O)R^9$, $-NHC(O)R^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms;

where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR$, $-C(O)R$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; and

each R^5 is independently H, $-OH$, $-OR^6$, $-OCR^6R^7R^8$, $-CR^6R^7R^8$, $-NH_2$, $-NHR^6$, $-NR^6R^7$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, an arylalkyl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R^3 or R^4 of another unit of Formula I;

where:

R^6 and R^7 are each independently H, $-CR_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a

heterocyclyl, a cycloalkyl, or a cycloalkenyl, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, or cycloalkenyl is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{OR}^8$, $-\text{C}(\text{O})\text{R}^8$, $-\text{NHC}(\text{O})\text{R}^8$, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl;

where:

each R is independently H, halogen, an alkyl, or an alkenyl; and

R^8 is H, $-\text{C}(\text{R})_3$, $-\text{[C}(\text{R})_2\text{]}_{n4}\text{NHC}(\text{O})\text{R}^9$, $-\text{[C}(\text{R})_2\text{]}_{n4}\text{C}(\text{O})(\text{NH})\text{R}^9$, $-\text{OR}$, $-\text{N}(\text{R})_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}^9$, $-\text{C}(\text{O})\text{R}^9$, $-\text{NHC}(\text{O})\text{R}^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms; where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{N}(\text{R})_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{C}(\text{O})\text{R}$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; and

R^8 is H, $-C(R)_3$, $-[C(R)_2]_{n^4}NHC(O)R^9$, $-[C(R)_2]_{n^4}C(O)(NH)R^9$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR^9$, $-C(O)R^9$, $-NHC(O)R^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms;

where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR$, $-C(O)R$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; and

(ii) a compound comprising:

at least one nucleobase,

at least one fatty acid,

at least one amino acid, and

at least one sugar;

wherein the at least one nucleobase, the at least one fatty acid, the at least one amino acid, and the at least one sugar are linked by covalent bonds;

and

wherein the compound has a molecular weight of less than about 2,000 g/mol;

with the proviso that the nematode is not a *Caenorhabditis* species.

[0010] A second aspect of the present invention relates to a method of promoting or inhibiting reproduction in a nematode population. This method involves administering one or more isolated modulator compounds to the population under conditions effective to promote or inhibit reproduction in the nematode population, where the one or more isolated modulator compounds is selected from the group consisting of ascr#1; ascr#3; ascr#7; ascr#8; ascr#9; ascr#10; ascr#16; ascr#18; ascr#20; ascr#22; a combination of ascr#9 and ascr#10; a combination of ascr#9 and ascr#16; a combination of ascr#9 and ascr#18; a combination of ascr#9 and ascr#20; and a combination of ascr#9 and ascr#22.

[0011] A third aspect of the present invention relates to a method of promoting or inhibiting nematode aggregation at a first location. This method involves:

- (i) contacting the first location with one or more isolated modulator compounds under conditions effective to promote or inhibit nematode aggregation at the first location, or
- (ii) contacting a second location with one or more isolated modulator compounds under conditions effective to promote or inhibit nematode aggregation at the first location, where the first location and the second location are spaced to permit said contacting at the second location to have an effect on nematode aggregation at the first location;

where the one or more isolated modulator compounds is selected from the group consisting of ascr#1; ascr#2; ascr#3; ascr#7; ascr#8; ascr#9; ascr#10; ascr#11; ascr#16; ascr#18; ascr#20; ascr#22; icas#9; oscr#10; a combination of ascr#9 and ascr#10; a combination of ascr#9 and ascr#16; a combination of ascr#9 and ascr#18; a combination of ascr#9 and ascr#20; a combination of ascr#9 and ascr#22; a combination of ascr#2, ascr#3, ascr#8, and icas#9; a combination of ascr#9, ascr#3, ascr#8, and icas#9; a combination of ascr#9 and oscr#10; and a combination of ascr#9, oscr#10, and ascr#11.

[0012] A fourth aspect of the present invention relates to a method of treating or preventing parasite infection of a plant. This method involves:

- (i) contacting the plant with one or more isolated modulator compounds under conditions effective to treat or prevent parasite infection of the plant, or

(ii) contacting a second location with one or more isolated modulator compounds under conditions effective to treat or prevent parasite infection of the plant, where the plant and the second location are spaced to permit said contacting the second location to have an effect on parasite infection of the plant; where the parasite is a nematode or an insect and the one or more isolated modulator compounds is selected from the group consisting of ascr#1; ascr#2; ascr#3; ascr#7; ascr#8; ascr#9; ascr#10; ascr#11; ascr#16; ascr#18; ascr#20; ascr#22; icas#9; oscr#10; a combination of ascr#9 and ascr#10; a combination of ascr#9 and ascr#16; a combination of ascr#9 and ascr#18; a combination of ascr#9 and ascr#20; a combination of ascr#9 and ascr#22; a combination of ascr#2, ascr#3, ascr#8, and icas#9; a combination of ascr#9, ascr#3, ascr#8, and icas#9; a combination of ascr#9 and oscr#10; and a combination of ascr#9, oscr#10, and ascr#11.

BRIEF DESCRIPTION OF THE DRAWINGS

[0013] Figures 1A–E show that indole ascarosides attract *C. elegans* hermaphrodites and males. Figure 1A is a schematic representation of the bioassay used to measure attraction behavior in worms. Zone A is the region where the sample or control solution is applied. The X denotes the initial position of the assayed worms. Figure 1B is a schematic representation of a quadrant chemotaxis assay. An X denotes the spot where washed worms are placed at the beginning of the assay. The shaded regions of the quadrant plate indicate the agar containing the chemical, whereas the white regions denote control agar. The number of animals in each quadrant was counted after 30 minutes and a chemotaxis index was computed (see Example 9). The chemotaxis index for the schematic is 0.84. Figure 1C shows that icas#1, icas#3, and icas#9 are attractive to both *C. elegans* sexes. All three compounds were assayed at 1 pmol using N2 hermaphrodites and *him-5* males. Open bars: no compound (solvent vehicle) controls. Figure 1D is a chart of the dose dependence of icas#3 response for N2 hermaphrodites and *him-5* males in the spot attraction assay ($*p < 0.01$, $**p < 0.001$, $***p < 0.0001$, unpaired *t* test with Welch's correction). Figure 1E is a graph of the dose dependence of icas#3 attraction for N2 hermaphrodites in the quadrant chemotaxis assay (one-factor ANOVA with Dunnett's post-test, $**p < 0.01$).

[0014] Figures 2A–E demonstrate that the response to icas#3 in N2 hermaphrodites is mediated by ASK sensory neurons and the downstream AIA interneurons. Figure 2A is a

schematic representation of the connectivity of the ASK sensory neuron to other neurons. The primary synaptic output of ASK is the AIA interneuron. Figure 2B shows that attraction of hermaphrodites to *icas#3* is dependent on the ASK sensory neurons and the AIA interneurons. Ablation of the RMG interneuron does not affect attraction of N2 or *ncs-1::gfp* hermaphrodites to *icas#3* ($*p < 0.01$, $***p < 0.0001$, unpaired Student's *t* test with Welch's correction). Figure 2C is a graph of the aggregation of N2 and ASK-ablated hermaphrodites at low worm density (20 worms per plate). ASK-ablated worms do not aggregate in response to *icas#3*. *icas#3* induces G-CaMP fluorescence signals in AIA interneurons (Figure 2D). The colored traces represent fluorescence changes in the AIA neurons of individual animals upon exposure to 1 μ M *icas#3*. The black traces represent fluorescent changes of individual animals upon exposure to buffer. The grey shading indicates presence of *icas#3*, $n = 10$ animals. Figure 2E is a graph of the average AIA fluorescence change in animals exposed to either buffer or *icas#3* ($**p < 0.01$, unpaired Student's *t* test with Welch's correction). Error bars indicate standard error of mean (S.E.M).

[0015] Figures 3A–D show that social and solitary wild-type hermaphrodites are attracted to *icas#3*, but not to non-indole ascarosides. Solitary and social wild-type hermaphrodites are not attracted to a physiological *ascr#2,3,5* mixture in the spot attraction assays, in contrast to *npr-1(ad609)* mutant worms (Figure 3A) ($***p < 0.0001$, unpaired *t* test with Welch's correction). In the quadrant chemotaxis assay, hermaphrodites from all tested strains are attracted to 1 pM *icas#3* and repelled by a physiological mixture of non-indole ascarosides (10 nM *ascr#2,3,5*), except for *npr-1(ad609)* mutant worms, which are also attracted to the *ascr#2,3,5* blend (Figure 3B) (chemotaxis after 30 minutes (for chemotaxis indices at 15 minutes, see Figure 5C). Figure 3C is a graph of the dose-dependence of *icas#3* attraction for social hermaphrodites in the quadrant chemotaxis assay. Figures 3B–C: $*p < 0.05$, $**p < 0.01$, one-factor ANOVA with Dunnett's post-test. Figure 3D shows that social wild-type hermaphrodites and *npr-1(ad609)* mutant worms are attracted to *icas#3* in the spot attraction assay ($**p < 0.001$, $***p < 0.0001$, unpaired *t* test with Welch's correction).

[0016] Figures 4A–C relate to an emerging model for a modular language of signaling molecules. Figure 4A shows that *icas#3* and *ascr#3* are competing signals for N2 hermaphrodites. Mixtures of 120 fmol *ascr#3* and 10 fmol *icas#3* (Condition 1) attract worms to zone A, whereas larger amounts of a mixture of the same ratio (Condition 2; 12 pmol *ascr#3* and 1 pmol *icas#3*) deter worms from zone A and instead attract to the periphery (zones B and C). In experiments with Condition 2, only one worm entered the treated zone A,

whereas 31 worms entered control zone A ($***p < 0.0001$, unpaired Student's *t* test with Welch's correction). Figure 4B shows that synergistic blends of non-indole ascarosides induce dauer at nanomolar to micromolar concentrations and function as a male attractant at picomolar to nanomolar concentrations, whereas indole ascarosides icas#3 and icas#9 act as hermaphrodite attractants and aggregation signals at femtomolar to picomolar concentrations. Figure 4C shows the modular assembly of *C. elegans* signaling molecules, based on building blocks derived from tryptophan ("head group"), fatty acids ("lipid side chain"), *p*-aminobenzoic acid (PABA, "terminus"), and carbohydrate metabolism ("ascarylose").

[0017] Figures 5A–B show that indole ascarosides are strong hermaphrodite attractants. In the spot attraction assay, N2 hermaphrodites are strongly attracted to low concentrations of icas#9, whereas males are not attracted ($***p < 0.0001$, unpaired Student's *t* test with Welch's correction) (Figure 5A). Figure 5B is a graph of quadrant chemotaxis indices of N2 and CB4856 hermaphrodites on plates containing 1 pM icas#3 with or without food. In the quadrant chemotaxis assay, hermaphrodites from all tested strains are attracted to 1 pM icas#3 and repelled by a physiological mixture of non-indole ascarosides (10 nM of each ascr#2,3,5), except for *npr-1(ad609)* mutant worms, which are also attracted to the ascr#2,3,5 blend (Figure 5C) (chemotaxis after 15 minutes (for chemotaxis indices at 30 minutes, see Figure 3B), $*p < 0.05$, $**p < 0.01$, one-factor ANOVA with Dunnett's post-test).

[0018] Figures 6A–E relate to aggregation and locomotory changes in response to icas#3. Figure 6A shows the aggregation behavior of solitary and social hermaphrodites on icas#9 plates at low worm density (20 worms per 5 cm plate) ($*p < 0.05$, $**p < 0.01$, one-factor ANOVA with Dunnett's post-test). Figure 6B shows the aggregation of *him-5* males on plates containing 100 fM or 100 pM of icas#3 ($**p < 0.01$, one-factor ANOVA with Dunnett's post-test). Figure 6C shows that *daf-22* hermaphrodites are attracted to icas#3 in the spot attraction assay ($*p < 0.01$, $**p < 0.01$, $***p < 0.0001$, unpaired Student's *t*-test with Welch's correction). Figure 6D shows the forward velocity (velocity of worms during the worm's forward movement) of N2 hermaphrodites at different icas#3 concentrations. Figure 6E shows the number of reversals per minute of N2 hermaphrodites at different icas#3 concentrations. Figure 6D–E: $*p < 0.05$, $**p < 0.01$, one-factor ANOVA with Dunnett's post-test.

[0019] Figures 7A–B show that ASK ablation affects icas#3-dependent locomotory behavior of hermaphrodites. ASK-ablated hermaphrodites do not display reduced forward velocity upon exposure to icas#3 (Figure 7A). Reversal frequency of ASK ablated worms

does not increase in response to icas#3 (Figure 7B). $**p < 0.001$, unpaired t test with Welch's correction).

[0020] Figures 8A–D show that indole ascarosides mediate aggregation behavior in *C. elegans*. Figure 8A shows the aggregation behavior of solitary and social hermaphrodites at low worm densities (20 worms per plate) on different concentrations of icas#3. Figure 8B shows the aggregation behavior of solitary and social hermaphrodites at high worm densities (~120 worms per plate) on different concentrations of icas#3. Figure 8C shows the aggregation of *daf-22* hermaphrodites at low worm density on two different concentrations of icas#3. Figure 8D shows the mean stopped duration of N2 hermaphrodites at different icas#3 concentrations. Figures 8A–D: $*p < 0.05$, $**p < 0.01$ one-factor ANOVA with Dunnett's post-test.

[0021] Figures 9A–G relate to the identification of indole ascarosides as *daf-22*-dependent metabolites. Figure 9A shows the chemical structures of important ascarosides (Srinivasan et al., *Nature* 454:1115-18 (2008), which is hereby incorporated by reference in its entirety). Figure 9B is a schematic representation of Differential Analysis via 2D-NMR spectroscopy (DANS). Comparison of wild-type NMR spectra with *daf-22* mutant NMR spectra enabled detection of spectroscopic signals that may represent *daf-22*-dependent signaling molecules. Figure 9C is a small section of the actual wild-type and *daf-22* NMR spectra used for DANS. Signals of indole carboxylic acid are present in both spectra (left box in each panel), whereas another indole-derived signal (right box in each panel) is only present in the wild-type, but not the *daf-22* spectrum. (Figure 9D is an HPLC-MS-based comparison of the wild-type and *daf-22* metabolomes. Ion chromatograms obtained for wild-type show peaks for the molecular ions of five different indole ascarosides which are absent from the *daf-22* chromatograms. Figure 9E shows the structures of identified indole ascarosides. Figure 9F shows the structures of additional non-indole ascarosides identified in this study. Figure 9G is a graph of the relative amounts of indole ascarosides icas#3 and icas#9 and non-indole ascarosides ascr#3 and ascr#9 secreted by *C. elegans* N2 grown in liquid culture, as determined by HPLC-MS analyses of media extracts (normalized to concentration of ascr#3; $n = 5$, $\pm$ SEM). For mass spectrometric quantification of indole and non-indole ascarosides, standard mixtures of authentic reference compounds were used.

[0022] Figure 10 is the central section of the dqfCOSY NMR spectrum (CDCl_3 , 600 MHz) of the wild-type (N2) metabolome fraction containing *daf-22* dependent indole derivatives (boxes).

[0023] Figures 11A–C are the ^1H NMR spectrum (CD_3OD , 600 MHz) (Figure 11A), ^1H , ^{13}C -HSQC spectrum (CD_3OD , 600 MHz) (Figure 11B), and ^1H , ^{13}C -HMBC spectrum (CD_3OD , 600 MHz) (Figure 11C) of synthetic icas#3.

[0024] Figures 12A–E relate to HPLC-MS identification of indole ascarosides. They show the electrospray ionization MS spectra (negative ion mode) of icas#9, icas#7, icas#1, icas#3, and icas#10, respectively.

[0025] Figures 13 A–H relate to HPLC-MS analysis of the biosynthetic origin of indole ascarosides. They show the HPLC-MS ion chromatograms (acquired using negative-ion electrospray ionization and single-ion recording mode) of whole-body extracts of *C. elegans* cultivated in CeMM medium with a 1:1 mixture of L-[2,4,5,6,7- D_5]-tryptophan and L-tryptophan showing [D_5]- and [H]-isotopomers of icas#9 (Figures 13A–B), icas#1 (Figures 13E–F), icas#3 (Figures 13C–D), and icas#10 (Figures 13G–H), respectively.

[0026] Figures 14A–B also show that indole ascarosides mediate aggregation behavior in *C. elegans*. Figure 14E shows the aggregation (arrow) of N2 hermaphrodites (20 worms per plate) on plates containing 10 pM of icas#3 compared to behavior on control plates. Figure 14F shows the aggregation of N2 hermaphrodites (~120 worms per plate) on plates containing 1 pM of icas#3 compared to behavior on control plates.

[0027] Figures 15A–B are the ^1H NMR spectrum (Figure 15A) and ^{13}C NMR spectrum (Figure 15B) of Methyl 5-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)pentanoate (**3**). ^1H NMR: 400 MHz, acetone- d_6 ; ^{13}C NMR: 100 MHz, acetone- d_6 .

[0028] Figures 16A–B are the ^1H NMR spectrum (Figure 16A) and ^{13}C NMR spectrum (Figure 16B) of 5-(3'*R*,5'*R*-dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)pentanoic acid (**oscr#9**). ^1H NMR: 400 MHz, methanol- d_4 ; ^{13}C NMR: 100 MHz, methanol- d_4 .

[0029] Figures 17A–B are the ^1H NMR spectrum (Figure 17A) and ^{13}C NMR spectrum (Figure 17B) of Methyl 9-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)nonanoate (**5**). ^1H NMR: 400 MHz, acetone- d_6 ; ^{13}C NMR: 100 MHz, acetone- d_6 .

[0030] Figures 18A–B are the ^1H NMR spectrum (Figure 18A) and ^{13}C NMR spectrum (Figure 18B) of 9-(3'*R*,5'*R*-dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)nonanoic acid (**oscr#10**). ^1H NMR: 600 MHz, methanol- d_4 ; ^{13}C NMR: 100 MHz, methanol- d_4 .

[0031] Figure 19 is the ^1H NMR spectrum (500 MHz, chloroform- d_1) of Ethyl (8*R*)-hydroxy-(3*R*)-*tert*-butyldimethylsilyloxynonanoate (**9**).

[0032] Figure 20 is the ^1H NMR spectrum (400 MHz, chloroform- d_1) of Ethyl (8*R*)-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxynonanoate (**10**).

[0033] Figures 21A–D are the ^1H NMR spectrum (Figure 21A), ^{13}C NMR spectrum (Figure 21B), HSQC spectrum (Figure 21C), and HMBC spectrum (Figure 21D) of (8*R*)-(3'*R*,5'*R*-dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxynonanoic acid (**bhas#10**). ^1H NMR: 500 MHz, methanol- d_4 ; ^{13}C NMR: 125 MHz, methanol- d_4 ; HSQC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 ; HMBC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 .

[0034] Figures 22A–B are the ^1H NMR spectrum (Figure 22A) and ^{13}C NMR spectrum (Figure 22B) of (8*R*)-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)non-1-ene (**12**). ^1H NMR: 400 MHz, acetone- d_6 ; ^{13}C NMR: 100 MHz, acetone- d_6 .

[0035] Figure 23 is the ^1H NMR spectrum (400 MHz, chloroform- d_1) of (12*R*)-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-*tert*-butyldimethylsilyloxytridec-5-enoic acid ethyl ester (**13**).

[0036] Figure 24 is the ^1H NMR spectrum (400 MHz, chloroform- d_1) of (12*R*)-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-*tert*-butyldimethylsilyloxytridecanoic acid ethyl ester (**14**).

[0037] Figure 25 is the ^1H NMR spectrum (400 MHz, chloroform- d_1) of (12*R*)-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxytridecanoic acid ethyl ester (**15**).

[0038] Figures 26A–D are the ^1H NMR spectrum (Figure 26A), dqfCOSY spectrum (Figure 26B), HSQC spectrum (Figure 26C), and HMBC spectrum (Figure 26D) of (12*R*)-(3'*R*,5'*R*-dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxytridecanoic acid (**bhas#22**). ^1H NMR: 600 MHz, methanol- d_4 ; dqfCOSY: 600 MHz, methanol- d_4 ; HSQC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 ; HMBC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 .

[0039] Figures 27A–B are the ^1H NMR spectrum (Figure 27A) and ^{13}C NMR spectrum (Figure 27B) of 11-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)undec-1-ene (**17**). ^1H NMR: 400 MHz, acetone- d_6 ; ^{13}C NMR: 100 MHz, acetone- d_6 .

[0040] Figure 28 is the ^1H NMR spectrum (400 MHz, chloroform- d_1) of 15-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-*tert*-butyldimethylsilyloxypentadec-5-enoic acid ethyl ester (**18**).

[0041] Figure 29 is the ^1H NMR spectrum (400 MHz, chloroform- d_1) of 15-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-*tert*-butyldimethylsilyloxypentadecanoic acid ethyl ester (**19**).

[0042] Figure 30 is the ^1H NMR spectrum (400 MHz, chloroform- d_1) of 15-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxypentadecanoic acid ethyl ester (**20**).

[0043] Figures 31A–D are the ^1H NMR spectrum (Figure 31A), dqfCOSY spectrum (Figure 31B), HSQC spectrum (Figure 31C), and HMBC spectrum (Figure 31D) of 15-(3'*R*,5'*R*-dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxypentadecanoic acid (**bhos#26**). ^1H NMR: 400 MHz, methanol- d_4 ; dqfCOSY: 600 MHz, methanol- d_4 ; HSQC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 ; HMBC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 .

[0044] Figures 32A–D are the ^1H NMR spectrum (Figure 32A), dqfCOSY spectrum (Figure 32B), HSQC spectrum (Figure 32C), and HMBC spectrum (Figure 32D) of 9-(5'*R*-(1*H*-indole-3-carbonyloxy)-3'*R*-hydroxy-6'*S*-methyl-tetrahydro-(2*H*)-pyran-2'-yloxy)nonanoic acid (**icos#10**). ^1H NMR: 600 MHz, methanol- d_4 ; dqfCOSY: 600 MHz, methanol- d_4 ; HSQC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 ; HMBC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 .

[0045] Figures 33A–B are the ^1H NMR spectrum (Figure 33A) and ^{13}C NMR spectrum (Figure 33B) of 4-*tert*-butyldimethylsilyloxybenzoic acid (**22**). ^1H NMR: 400 MHz, chloroform- d_1 ; ^{13}C NMR: 100 MHz, chloroform- d_1 .

[0046] Figures 34A–C are the ^1H NMR spectrum (Figure 34A), dqfCOSY spectrum (Figure 34B), and HMBC spectrum (Figure 34C) of (8*R*)-(3'*R*-hydroxy-5'*R*-(4-hydroxybenzoyloxy)-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)non-(2*E*)-enoic acid (**hbas#3**). ^1H NMR: 600 MHz, methanol- d_4 ; dqfCOSY: 600 MHz, methanol- d_4 ; HMBC: 600 MHz for ^1H , 151 MHz for ^{13}C , methanol- d_4 .

[0047] Figures 35A–B are the ^1H NMR spectrum (Figure 35A) and dqfCOSY spectrum (Figure 35B) of (8*R*)-(3'*R*-hydroxy-5'*R*-(*E*)-(2-methylbut-2-enoyloxy)-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)non-(2*E*)-enoic acid (**mbas#3**). ^1H NMR: 600 MHz, methanol- d_4 ; dqfCOSY: 600 MHz, methanol- d_4 .

[0048] Figures 36A–D are the ^1H NMR spectrum (Figure 36A), dqfCOSY spectrum (Figure 36B), HMQC spectrum (Figure 36C), and HMBC spectrum (Figure 36D) of 2-(8*R*)-(3'*R*,5'*R*-di-hydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)nonanoyl-3,4,5-trihydroxy-6-

hydroxymethyl-(2*H*)-tetrahydropyran (glas#10). ¹H NMR: 600 MHz, methanol-*d*₄; dqfCOSY: 600 MHz, methanol-*d*₄; HMQC: 600 MHz for ¹H, 151 MHz for ¹³C, methanol-*d*₄; HMBC: 600 MHz, methanol-*d*₄.

[0049] Figures 37A–K show the HPLC-ESI-MS data of: (ω-1)-oxygenated ascarosides (ascr) (Figure 37A), (ω)-oxygenated ascarosides (oscr) (Figure 37B), (ω-1)-oxygenated β-hydroxyascarosides (bhas) (Figure 37C), (ω)-oxygenated β-hydroxyascarosides (bhos) (Figure 37D), (ω-1)-oxygenated indole ascarosides (icas) (Figure 37E), (ω)-oxygenated indole ascarosides (icos) (Figure 37F), (ω-1)-oxygenated indole β-hydroxy ascarosides (ibha) (Figure 37G), (ω)-oxygenated indole β-hydroxy ascarosides (ibho) (Figure 37H), glucosyl ascaroside esters (glas) (Figure 37I), ascr#8 and 4-(4-hydroxybenzoyl)- and 4-(2-(*E*)-methyl-2-butenoyl)-ascarosides (hbas and mbas) (Figure 37J), and ascr#2 and ascr#6.1 (Figure 37K). * = confirmed using synthetic standards; ^s SMID: Small Molecule Identifier for small molecules identified from *C. elegans* and other nematodes. The SMID database (www.smid-db.org) is an electronic resource maintained by Frank C. Schroeder and Lukas Mueller at the Boyce Thompson Institute in collaboration with Wormbase (www.wormbase.org). The purpose of this database is to introduce searchable, gene-style identifiers, “SMIDs”, for all small molecules newly identified from *C. elegans* and other nematodes.

[0050] Figure 38 shows the HPLC elution profiles of ascarosides identified in wild-type and mutant excretome extracts of *C. elegans* (Δ indicates components with (*E*)-configured α,β-unsaturated sidechains).

[0051] Figures 39A–B are sections of dqfCOSY spectra (600 MHz, methanol-*d*₄) of mbas#3-enriched fraction from wild-type *C. elegans* media extracts (Figure 39A) and synthetic mbas#3 (Figure 39B) showing characteristic signals for methyl groups of the ascarylose ring and the side chain (blue), the allylic methyl group of the tiglate unit (red), and the pH dependant signal for the side chain double bond (green).

[0052] Figures 40A–B are sections of dqfCOSY spectra (600 MHz, methanol-*d*₄) of hbas#3-enriched fraction from wild-type *C. elegans* media extracts (Figure 40A) and synthetic hbas#3 (Figure 40B) showing characteristic signals for methyl groups of the ascarylose ring and the side chain (blue), the *para*-substituted 4-hydroxybenzoyl unit (red), and the side chain double bond (green).

[0053] Figures 41A–B are enlarged sections of dqfCOSY spectra (600 MHz, methanol-*d*₄) of hbas#3-enriched fraction from wild-type *C. elegans* media extracts (Figure

41A) and synthetic hbas#3 (Figure 41B) showing characteristic signals for methyl groups of the ascarylose ring (blue), and the ascarylose spin system (black).

[0054] Figure 42 is the dqfCOSY spectrum (600 MHz, methanol-*d*₄) of oscr#9-enriched fraction from *acox-1(ok2257)* media extracts.

[0055] Figures 43A–B are sections of dqfCOSY spectra (600 MHz, methanol-*d*₄) of glas#10-enriched fraction from *acox-1(ok2257)* worm pellet extracts (Figure 43A) and synthetic glas#10 (Figure 43B), showing characteristic signals for methyl groups of the ascarylose ring and the side chain (blue), the anomeric hydrogen of the glucose unit (red), the glucose spin system (green), and the ascarylose spin system (black).

[0056] Figures 44A–P are MS/MS product ion spectra of ascaroside standards. ascr#5 shows *m/z* 147 [C₆H₁₁O₄] for ascarylose and *m/z* 73 for the C₃-sidechain and/or the ascarylose derived C₃O₂H₅ fragment of identical molecular composition (Figure 44A). oscr#9 (Figure 44B) and ascr#9 (Figure 44C) show *m/z* 99 [C₅H₇O₂] for the C₅-sidechain, *m/z* 147 [C₆H₁₁O₄], and *m/z* 73 for the ascarylose derived C₃O₂H₅ fragment. ascr#7 shows *m/z* 125 [C₇H₉O₂] for the C₇-sidechain, *m/z* 111 [C₆H₇O₂] and *m/z* 73 for the ascarylose derived C₃O₂H₅ fragment (Figure 44D). ascr#1 shows *m/z* 127 [C₇H₁₁O₂] for the C₇-sidechain and *m/z* 73 for the ascarylose derived C₃O₂H₅ fragment (Figure 44E). ascr#3 shows *m/z* 111 [C₆H₇O₂] and *m/z* 73 for the ascarylose derived C₃O₂H₅ fragment (Figure 44F). ascr#10 (Figure 44G) and oscr#10 (Figure 44H) show *m/z* 173 [C₉H₁₅O₃] and 155 [C₉H₁₃O₂] for the C₉-sidechain and *m/z* 73 for the ascarylose derived C₃O₂H₅ fragment. β-hydroxy ascarosides such as bhas#22 (C₁₃-sidechain) (Figure 44I) and bhos#26 (C₁₅-sidechain) (Figure 44J) show side chain specific fragment ions at *m/z* 185 [C₁₁H₂₁O₂] and *m/z* 213 [C₁₃H₂₅O₂], respectively, which originate from loss of ascarylose and acetic acid. Furthermore, bhas#22 and bhos#26 show an intensive product ion at *m/z* 59 [C₂H₃O₂] for acetate, as well as the diagnostic ion at *m/z* 73 for the ascarylose derived C₃O₂H₅ fragment. Indole ascarosides such as icas#9 (Figure 44K) and icas#1 (Figure 44L) show side chain specific fragment ions for the corresponding ascarosides at *m/z* 247 [C₁₁H₁₉O₆] and *m/z* 275 [C₁₃H₂₃O₆], respectively, which originate from loss of the indole carbonyl unit [C₉H₅NO]. Furthermore, icas#9 and icas#1 show *m/z* 160 [C₉H₆NO₂] for indole carboxylate ions. Ascaroside product ions of indole ascarosides show additional fragment ions corresponding to the fragmentation pattern of non-indole ascarosides, such as the diagnostic fragment ion at *m/z* 73 for the ascarylose derived C₃O₂H₅ fragment. icas#3 (Figure 44M) shows *m/z* 301 [C₁₅H₂₅O₆] from loss of indole carbonyl unit [C₉H₅NO], along with *m/z* 160 [C₉H₆NO₂] for indole carboxylate ions.

mbas#3 (Figure 44N)) shows m/z 301 $[C_{15}H_{25}O_6]$ and m/z 283 $[C_{15}H_{23}O_5]$, which originate from loss of tigloyl $[C_5H_6O]$ and tiglate $[C_5H_8O_2]$ units, respectively. Furthermore, mbas#3 shows m/z 99 $[C_5H_7O_2]$ for tiglate ions. The diagnostic fragment ion at m/z 73 for the ascarylose derived $C_3O_2H_5$ fragment is of low intensity. hbas#3 (Figure 44O)) shows m/z 301 $[C_{15}H_{25}O_6]$ which originates from loss of a hydroxybenzoyl unit $[C_7H_4O]$. Furthermore, hbas#3 shows m/z 137 $[C_7H_5O_3]$ and m/z 93 $[C_6H_5O]$ for hydroxybenzoate and phenolate ions. The ascaroside diagnostic fragment ion at m/z 73 for the ascarylose derived $C_3O_2H_5$ fragment is of low intensity. PABA linked ascr#8 (Figure 44P)) shows characteristic fragment ions at m/z 136 $[C_7H_6NO_2]$ and m/z 92 $[C_6H_6N]$ for PABA and anilid ions, respectively, along with the diagnostic fragment ion at m/z 73 for the ascarylose derived $C_3O_2H_5$ fragment.

[0057] Figure 45 shows the alignment of human MFE-2 isoform 2 (NP_000405.1) with *C. elegans* MAOC-1 (NP_495494.1 (red)) and DHS-28 (NP_509146.1; oxidoreductase domain (green), SCP-2 sterol carrier protein domain (yellow)) performed using ClustalW. Amino acids constituting the catalytic sites are marked with green and red boxes and the peroxisomal targeting signal in black. Identical amino acids are marked in grey and similar amino acids are marked in light grey.

[0058] Figures 46A–C relate to the identification of new ascarosides in *C. elegans*. Figure 46A is the LC-MS total ion current (TIC) chromatogram of wild-type *C. elegans* excretome (ESI⁺). Figure 46B illustrates the MS/MS fragmentation of ascarosides. Figure 46C is the LC-MS/MS screen (precursors of m/z 73) of wild-type excretome, and reveals known ascarosides (black), new homologs (C4, C6, C8, C11, and C12), new (ω)-oxygenated isomers (C5 ω , C9 ω , and C11 ω) and new 4'-acylated derivatives (hbas#3 and mbas#3) (* non-ascarosides). The highly polar ascr#5 elutes at 6.5 min, outside of the shown retention time range.

[0059] Figures 47A–C show ascarosides identified in wild-type, *acox-1*, *maoc-1*, *dhs-28*, and *daf-22* worms via LC-MS/MS. Figure 47A shows (ω -1)-oxygenated ascarosides, Figure 47B shows (ω)-oxygenated ascarosides, and Figure 47C shows examples for 4'-acylated derivatives. The stereochemistry of compounds that were not synthesized for syntheses and a complete list of the 146 characterized ascarosides) was proposed as shown based on analogy and HPLC-MS retention times (see Figure 38).

[0060] Figures 48A–B relate to attraction to hbas#3. Wild-type (N2) hermaphrodites are attracted to hbas#3 in the spot attraction assay in a dose dependent manner (Figure 48A).

Figure 48B shows the dose dependence of hbas#3 attraction for wild-type hermaphrodites in the quadrant chemotaxis assay.

[0061] Figures 49A–B relate to ascaroside biogenesis. Figure 49A shows the proposed roles of peroxisomal β -oxidation enzymes ACOX-1, MAOC-1, DHS-28, and DAF-22 in ascaroside biosynthesis. Figure 49B shows the gene structures of *acox-1*, *maoc-1*, *dhs-28*, and *daf-22* mutant alleles, showing point mutations (vertical bars) and deletions (horizontal bars).

[0062] Figures 50A–E show the amount of (ω -1)-oxygenated ascarosides in wild-type (Figure 50E) and β -oxidation mutants (*acox-1* (Figure 50A), *maoc-1* (Figure 50B), *dhs-28* (Figure 50C), and *daf-22* (Figure 50D) with saturated (blue), α,β -unsaturated (red), and β -hydroxylated (green) side chains.

[0063] Figure 51 shows the alignment of *C. elegans* ACOX-1 isoform a.1 with other peroxisomal acyl-CoA oxidases performed using ClustalW. Identical amino acids are marked in grey, similar amino acids are marked in light grey, and the peroxisomal targeting signal is marked in black.

[0064] Figures 52A–B show the amount of (ω)-oxygenated ascarosides in wild-type (Figure 52A) and *acox-1* mutants (Figure 52B) (see also Figures 53A–E).

[0065] Figures 53A–E are the profiles of (ω)-oxygenated ascaroside in wild-type (N2) (Figure 53A) and β -oxidation mutants (*acox-1* (Figure 53B), *maoc-1* (Figure 53C), *dhs-28* (Figure 53D), *daf-22* (Figure 53E) showing saturated (blue), α,β -unsaturated (red), and β -hydroxylated (green) ascarosides. (For (ω -1)-oxygenated ascarosides, see Figures 50A–E).

[0066] Figures 54A–B relate to indole ascaroside biosynthesis. Relative abundance of (ω -1) and (ω)-oxygenated C₅-ascarosides ascr#9 and oscr#9 and their corresponding indole ascarosides icas#9 and icos#9 in *acox-1* indicates that indole-3-carbonyl attachment is highly specific (Figure 54A). Figure 54B includes the LC-MS ion traces of *daf-22(m130)* excretome following incubation with a 1:1 mixture of ascr#10 and oscr#10, showing a preference for indole-3-carbonyl attachment to the (ω -1)-oxygenated ascr#10.

[0067] Figure 55 shows the relative abundance of ascarosides ascr#3 and ascr#9 and their corresponding indole ascarosides icas#3 and icas#9 in wild-type excretome extracts, and indicates that indole attachment is highly dependent on side chain length.

[0068] Figures 56A–B relate to the ascaroside profiles. Analysis of worm body ascaroside profiles (Figure 56A) reveals ascaroside glucosides (e.g. glas#10) and indicates preferential excretion of unsaturated ascarosides. Ratio of (ω -1) to (ω)-linked saturated

ascarosides in *dhs-28* mutant excretome (Figure 56B) shows strong dependence on nutritional conditions (for β -hydroxy ascarosides see Figures 59A–C), reflecting starvation dependence of *ascr#3/ascr#5* ratio in wild-type *C. elegans* (see Figure 60) (Welch's t-test, *: $P < 0.05$; ** $P < 0.001$).

[0069] Figures 57A–C are LC-MS/MS chromatograms (precursor ions of $m/z = 73$) of *acox-1(ok2257)* worm body extracts showing glucosyl esters *glas#10*, *glas#18*, and *glas#22* and the corresponding non-glycosylated ascarosides *ascr#10*, *ascr#18*, and *ascr#22*.

[0070] Figure 58 shows the differential excretion of (ω)-oxygenated ascarosides by wild type *C. elegans* (For (ω -1)-oxygenated ascarosides see Figure 56A).

[0071] Figures 59A–C show the ratio of (ω -1) to (ω)-linked ascarosides in *dhs-28(hj8)* mutant worms and show strong dependence on nutritional conditions. Figure 59A is the ratio of β -hydroxy ascarosides in *dhs-28(hj8)* excretome extracts. Figure 59B is the ratio of β -hydroxy ascarosides in *dhs-28(hj8)* worm body extracts. Figure 59C is the ratio of saturated ascarosides in *dhs-28(hj8)* worm body extracts (saturated ascarosides in *dhs-28(hj8)* excretome extracts are shown in Figure 56B).

[0072] Figure 60 shows the starvation dependence of *ascr#3/ascr#5* ratio in wild type excretome.

[0073] Figures 61A–B relate to ascaroside biogenesis. Figure 61A shows the modular assembly of ascarosides from amino acid (green), fatty acid (blue), and carbohydrate (red) building blocks. Figure 61B shows a model for ascaroside biogenesis. Chain elongation of fatty acids (by putative elongase homologs *elo-1-9²⁵*) (Agbaga et al., *PNAS* 105:12843–48 (2008), which is hereby incorporated by reference in its entirety) is followed by (ω -1)- or (ω)-oxygenation of VLCFAs and ascarylose attachment. The resulting VLCAs enter peroxisomal β -oxidation via ACOX-1, MAOC-1, DHS-28, and DAF-22 producing short chain ascarosides, which are linked to amino acid-derived moieties and other building blocks.

[0074] Figure 62 is the alignment of *Yersinia pseudotuberculosis* CDP-3, 6-dideoxy-D-glycero-D-glycero-4-hexulose-5-epimerase or *ascE* (AAA88702.1) with *C. elegans* homolog C14F11.6 (CCD64543.1) performed using ClustalW. Identical amino acids are marked in grey and similar amino acids are marked in light grey.

[0075] Figures 63A–D relate to the quantification and response of *S. feltiae* to the *C. elegans* dispersal blend. Figure 63A shows the structures of the ascarosides for the dispersal blend. Figure 63B shows the quantification of dispersing *C. elegans* using Image J. (the second column shows inverted pictures and the third column shows the counted nematodes).

Figure 63C shows the contribution of individual ascarosides to the activity of the synthetic blend. ascr#2 (3.68 pmol/ul), ascr#3 (0.165 pmol/ul), ascr#8 (0.25 pmol/ul), and icas#9 (0.005 pmol/ul). Seven experiments were done for each treatment. +, present and -, absent. Student's t-test, unpaired ($p < 0.05$). Figure 63D shows the *S. feltiae* response to the *C. elegans* dispersal blend visualized within an entire plate.

[0076] Figures 64A–D relate to the dispersal assay. Figure 64A shows naturally dispersing infective juveniles (IJ) of *S. feltiae* from an insect cadaver. Figure 64B shows approximately 300 IJs placed on an agar plate in water. Figure 64C shows IJs treated with either water (control) or insect cadaver extract. Images are representative of six experiments for each treatment. Figure 64D shows the dispersal assay on the same plate. The illustration represents two experiments. Behavior is temperature and season dependent. Assays were conducted at RT ($22 \pm 0.5^\circ\text{C}$) or under temperature controlled conditions.

[0077] Figure 65 shows the structures of several ascarosides.

[0078] Figures 66A–E show that an ascaroside blend regulates *C. elegans* dispersal behavior, and the dispersal blend is recognized by other nematodes. Figure 66A shows the identification of the dispersal blend. Images (~250 nematodes) are representative of 9, 10, and 11 experiments of control (0.25% *E. coli* (HB101)), synthetic blend with 0.25% *E. coli* (HB101), and dauer supernatant, respectively. Figure 66B shows quantification using Image J (<http://rsbweb.nih.gov/ij/download.html>). Control vs synthetic blend student's t-test unpaired ($p < 0.02$). Figure 66C shows the analysis of host cadavers infected by entomopathogenic nematodes or root knot nematodes for components of the *C. elegans* dispersal blend. Figure 66D shows the *S. feltiae* IJs' (~250) response to the dispersal blend; four experiments for each treatment. Figure 66E shows the response of root-knot J2s (*Meloidogyne* spp., mixture of *M. incognita*, *M. javanica*, and *M. floridensis*) to the *C. elegans* dispersal blend. The data represent 19 and 20 experiments from control water and *C. elegans* dispersal blend, respectively. At 2 h, using a student's t-test, unpaired, $p < 0.007$.

[0079] Figures 67A–E show the activity guided fractionation of the *S. feltiae* dispersal pheromone. Figure 67A shows the response to reverse phase (C18) chromatography fractions of insect cadaver extract. The image represents two independent experiments. The estimated physiologically relevant concentration was used in the assays; Frc, fraction. Figure 67B relates to the testing of physiologically relevant concentration of ascarosides found in Frc A (ascr#9, 40.3 pmol/ μl and ascr#11, 1.3 pmol/ μl). Image represents three experiments. Figure 67C shows that ascr#2 and ascr#9 are structural analogs. Natural and synthetic ascr#9 were

tested in combination with fraction B and C. Image represents four experiments. Figure 67D shows the structure of ascr#11. It (1.3 pmol/ μ l) is also sufficient to cause dispersal in combination with fractions B and C. Image represents three experiments.

[0080] Figures 68A–C are LC-MS ion chromatograms of fraction A (Figure 68A–B) and fraction B (Figure 68C). First panel shows ascr#11 at m/z 279, second panel shows ascr#9 at m/z 293.

[0081] Figure 69 shows the ascr#9 and ascr#11 profile during *S. feltiae* development. For each time point, six insect cadavers were analyzed by LC-MS. For the 0 time point, 4 uninfected larvae were analyzed. * detected but not quantifiable.

[0082] Figure 70 shows that ascr#9 can replace the function of ascr#2 in the *C. elegans* dispersal blend. Representative pictures are presented. Negative control using 0.25% *E. coli* HB101 (3 experiments), ascr#9 substitution in the *C. elegans* dispersal blend (6 experiments), and positive controls: the synthetic blend dispersal blend (7 experiments) and positive control dauer liquid culture (3 experiments).

[0083] Figures 71A–B relate to the dispersal blend of the phylogenetically related nematodes species. Figure 71A is the phylogenetic tree for entomopathogenic nematodes, plant parasitic nematodes, and *C. elegans*. The figure is adapted from *C. elegans* and the biology of nematodes (PAUL DE LEY: A QUICK TOUR OF NEMATODE DIVERSITY AND THE BACKBONE OF NEMATODE PHYLOGENY (WormBook, ed. The *C. elegans* Research Community, 2006), which is hereby incorporated by reference in its entirety). Red color indicates the example of genera. Figure 71B shows the presence of ascr#9 and ascr#11 in host insect cadaver of *Steinernema* spp. and *Heterorhabditis* spp. For each species, four insect (*G. mellonella*) cadavers infected with both *Steinernema* spp. or *Heterorhabditis* spp. were analyzed by LC-MS for ascr#9 and ascr#11 profiles.

[0084] Figures 72A–D show the structure and HPLC-ESI-MS data of (ω -1)-oxygenated saturated ascarosides (Figures 72A), (ω -1)-oxygenated unsaturated ascarosides (Figure 72B), (ω)-oxygenated saturated ascarosides (Figure 72C), and indole ascaroside icas#9 and ascr#2 (Figure 72D). ¹SMID: Small Molecule IDentifier for small molecules identified from *C. elegans* and other nematodes.

[0085] Figure 73 shows the HPLC retention times of ascarosides (ascr's) and omega-ascarosides (oscr's). Side chains range from 3–17 carbons, derived from analysis of wild-type and peroxisomal beta-oxidation mutants of *C. elegans* and synthetic standards.

[0086] Figures 74A–B show the identification of ascarosides in worm water extract of adult *S. glaseri*. Figure 74A is the total ion current (TIC) chromatogram from LC-MS/MS; screen for precursor ions of m/z 73.0 reveals ascaroside signals. Figure 74B is the corresponding ion traces for ascr#9, ascr#12, ascr#1, and ascr#14 from LC-MS measurement.

[0087] Figures 75A–K are NMR spectra of components in ascaroside synthesis, as follows.

NMR Conditions	Component	Figure
^1H NMR (400 MHz, acetone- d_6)	methyl 5-hydroxypentanoate	Figure 75A
^{13}C NMR (100 MHz, acetone- d_6)	methyl 5-hydroxypentanoate	Figure 75B
^1H NMR (400 MHz, chloroform- d_1)	methyl 9-hydroxynonanoate	Figure 75C
^1H NMR (400 MHz, acetone- d_6)	5-(3' <i>R</i> ,5' <i>R</i> -dibenzoyloxy-6' <i>S</i> -methyl-(2 <i>H</i>)-tetrahydropyran-2-yloxy)-pentanoic acid methyl ester	Figure 75D
^{13}C NMR (100 MHz, acetone- d_6)	5-(3' <i>R</i> ,5' <i>R</i> -dibenzoyloxy-6' <i>S</i> -methyl-(2 <i>H</i>)-tetrahydropyran-2-yloxy)-pentanoic acid methyl ester	Figure 75E
^1H NMR (400 MHz, methanol- d_4)	5-(3' <i>R</i> ,5' <i>R</i> -dihydroxy-6' <i>S</i> -methyl-(2 <i>H</i>)-tetrahydropyran-2-yloxy)-pentanoic acid	Figure 75F
^{13}C NMR (100 MHz, methanol- d_4)	5-(3' <i>R</i> ,5' <i>R</i> -dihydroxy-6' <i>S</i> -methyl-(2 <i>H</i>)-tetrahydropyran-2-yloxy)-pentanoic acid	Figure 75G
^1H NMR (400 MHz, acetone- d_6)	9-(3' <i>R</i> ,5' <i>R</i> -dibenzoyloxy-6' <i>S</i> -methyl-(2 <i>H</i>)-tetrahydropyran-2-yloxy)-nonanoic acid methyl ester	Figure 75H
^{13}C NMR (100 MHz, acetone- d_6)	9-(3' <i>R</i> ,5' <i>R</i> -dibenzoyloxy-6' <i>S</i> -methyl-(2 <i>H</i>)-tetrahydropyran-2-yloxy)-nonanoic acid methyl ester	Figure 75I
^1H NMR (600 MHz, methanol- d_4)	9-(3' <i>R</i> ,5' <i>R</i> -dihydroxy-6' <i>S</i> -methyl-(2 <i>H</i>)-tetrahydropyran-2-yloxy)-nonanoic acid	Figure 75J
^{13}C NMR (100 MHz, methanol- d_4)	9-(3' <i>R</i> ,5' <i>R</i> -dihydroxy-6' <i>S</i> -methyl-(2 <i>H</i>)-tetrahydropyran-2-yloxy)-nonanoic acid	Figure 75K

[0088] Figures 76A–D relate to the structure (Figure 76A), detection (Figures 76B–C), and synthesis (Figure 76D) of ascarosides. Both ascarosides (ascr) and indole ascarosides (icas) have been previously described ascarosides in *C. elegans*. In addition to (ω -1)-oxygenated ascr, their (ω)-oxygenation isomers (oscr) were identified in this HPLC-MS screen. Figure 76B shows the HPLC-MS identification of short and medium chain ascarosides in conditioned worm water obtained from free-living *C. elegans* (mixed-stage), insect parasitic *S. glaseri* (adult), and rat parasitic *N. brasiliensis* (adult). HPLC-MS analysis of *C. elegans* exudate shows known ascarosides (ascr#1, #3, #7, and icas#9), along with the

corresponding homologs (ascr#9, #10, #12, #14, and #18), and (ω)-oxygenated isomers (oscr#9, #10, #18). The highly polar ascr#5 elutes at 6.5 minutes, outside of the shown retention time range. Cross species comparison of molecular ion signals and their HPLC-retention times indicated that ascarosides abundantly produced by *C. elegans* are also found in many other nematode species, including *S. glaseri* and *N. brasiliensis* (peaks representing compounds also abundant in *C. elegans* shown in blue). Figure 76C shows the HPLC-MS identification of medium and long chain ascarosides in *P. strongyloides* (mixed-stage) and *H. bacteriophora* (adult). ascr#18 (blue) is produced in significant amounts by *C. elegans*, whereas longer chain homologs (red peaks) are abundantly produced by *P. strongyloides* and *H. bacteriophora*, but not *C. elegans*. Figure 76D shows the synthesis of (ω)-oxygenated ascarosides, oscr#9 and oscr#10.

[0089] Figure 77 is a heat map showing that ascarosides are produced by a wide range of nematode species. It is based on results from the HPLC-MS analysis of worm media samples obtained from incubating worms for several hours in S Basal, ddH₂O, or DMEM. Parasitic species indicated as infective juveniles (IJ) and adults (A) were collected separately; all other samples were obtained from mixed stage cultures. Colors in this heat map represent relative abundance of ascarosides detected by HPLC-MS, as indicated in the bar diagram on the right. For example, of the ascarosides detected in *S. glaseri* infective juveniles, the total was composed of 5.7% ascr#11, 92.2% ascr#9, 2.1% ascr#12, and 0.1% ascr#10.

[0090] Figure 78 shows that nematodes respond uniquely to areas conditioned by ascarosides. Several nematode species were scored for their response to areas conditioned with three concentrations of synthesized ascarosides (0.6 μ L of 1nM, 1 μ M, and 1mM, corresponding to 0.6 fmol, 0.6 pmol, and 0.6 nmol, respectively, per scoring area). Their occupancy in the conditioned region was compared to their occupancy in the control region for 20 minutes; 10 individuals per trial. Experiments in which nematodes that spent significantly more time in regions conditioned with ascaroside are highlighted in green. Experiments in which nematodes that spent significantly more time in control regions are highlighted in red, indicating avoidance of the ascaroside. Error bars, S.D. Statistical significance for each value was calculated in comparison to the response to water, shown first in each set. P-values were determined using an unpaired Student's t-test. ** $P < 0.01$, * $P < 0.05$.

[0091] Figure 79 relates to the retention assay using software that detects the presence of worms in both regions and provides an output of the percentage of each scoring region

occupied by worms, which is calculated once per second during the entire trial. The output is a plot of worm occupancy ratio vs. time for both scoring regions (right panel). The scoring index described by Bargmann et al., *Cell* 74:515-27 (1993), which is hereby incorporated by reference in its entirety, was adopted.

[0092] Figures 80A–C relate to the attraction assay. *C. elegans* males demonstrate differential long-range attraction to different nematode species and ascarosides. Long-range attraction of *C. elegans* males to conspecific hermaphrodites was established by scoring chemotaxis to a hermaphrodite or ascaroside-conditioned point source on a 10 cm agar plate (Figure 80A). Hermaphrodites were suspended in M9 buffer and placed within a copper cylinder on the agar plate for 6 hours before they were removed and the paralyzing agent sodium azide was added. The same was done with an M9 buffer control in a copper cylinder, on the opposite side. *C. elegans* males were placed in the center of the plate and allowed to wander until they became paralyzed in either spot. An Attraction Index adapted from Bargmann et al., *Cell*. 74:515-27 (1993), which is hereby incorporated by reference in its entirety, was then calculated, comparing worms that became paralyzed in the control vs. cue region. Figure 80B shows that *C. elegans* males are attracted to a point source conditioned by 50 conspecific hermaphrodites. They demonstrate partial attraction to equal numbers of *P. redivivus* females, but not to *S. carpocapsae*, *P. pacificus*, or *P. strongyloides* females/hermaphrodites. Figure 80C shows that *C. elegans* males can detect and chemotax towards a point source conditioned by 25 hermaphrodites over 6 hours. They chemotax towards a point source of ascr#2 and ascr#3, but not ascr#8. Error bars, S.D. Statistical significance for each value was calculated in comparison to the control, shown in Figure 80C. P-values were determined using an unpaired Student's t-test. ** P < 0.01, * P < 0.05.

[0093] Figure 81 shows that ascarosides do not increase mating to conspecific hermaphrodites, nor do they induce mating to different nematode species. A mating response assay (Peden et al., *Curr. Biol.* 15:394-404 (2005), which is hereby incorporated by reference in its entirety) was adapted, quantifying the percentage of males that successfully placed their ventral tail on hermaphrodites for 10 consecutive seconds, within the span of a 3-minute trial. Error bars, S.D.

[0094] Figure 82 shows the similar assembly of signaling molecules in nematodes and bacteria. *N*-acyl homoserine lactones (AHLs) are a family of small-molecules that mediate bacterial quorum sensing. AHLs are based on a homoserine lactone and feature species-specific variations in the *N*-acyl chain (Dickschat, *Nat. Prod. Rep.* 27:343-69 (2010),

which is hereby incorporated by reference in its entirety). Ascarosides are assembled in a very similar fashion, based on the dideoxysugar ascarylose scaffold to which a variable lipid chain is attached (Edison, *Curr. Opin. Neurobiol.* 4:378-88 (2009), which is hereby incorporated by reference in its entirety). They both play significant roles in mediating important survival strategies.

[0095] Figure 83 is a list of ascaroside compounds detected in *M. javanica* and/or *M. hapla*.

[0096] Figure 84 shows the high-resolution mass spectrometry data of ascaroside signaling molecules identified from root knot nematodes.

[0097] Figures 85A–E are HPLC-MS chromatograms of extracts from root knot nematodes (*M. javanica*, *M. floridensis*, *M. incognita*) demonstrating the presence of ascarosides ascr#10 (Figure 85A), ascr#16 (Figure 85B), ascr#18 (Figure 85C), ascr#20 (Figure 85D), and ascr#22 (Figure 85E).

[0098] Figures 86A–E are HPLC-MS chromatograms of extracts from *M. javanica* (J2 larvae) demonstrating the presence of ascarosides ascr#10 (Figure 86A), ascr#16 (Figure 86B), ascr#18 (Figure 86C), ascr#20 (Figure 86D), and ascr#22 (Figure 86E).

[0099] Figures 87A–E are HPLC-MS chromatograms of extracts from Northern Root-Knot Nematode *M. hapla* (J2 larvae) demonstrating the presence of ascarosides ascr#10 (Figure 87A), ascr#16 (Figure 87B), ascr#18 (Figure 87C), ascr#20 (Figure 87D), and ascr#22 (Figure 87E).

[0100] Figures 88A–E are HPLC-MS chromatograms of synthetic ascaroside standards (ascr#10 (Figure 88A), ascr#16 (Figure 88B), ascr#18 (Figure 88C), ascr#20 (Figure 88D), and ascr#22 (Figure 88E)).

DETAILED DESCRIPTION OF THE INVENTION

[0101] Disclosed herein are methods of modifying nematode behavior using certain isolated modulator compounds.

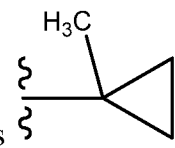
DEFINITIONS

[0102] As used herein, the following terms, unless otherwise indicated, shall be understood to have the following meanings. If not defined otherwise herein, all technical and scientific terms used herein have the same meaning as is commonly understood by one of

ordinary skill in the art to which this invention belongs. In the event that there is a plurality of definitions for a term herein, those in this section prevail unless stated otherwise.

[0103] The term “alkyl” refers to an aliphatic hydrocarbon group which may be a linear, branched, or cyclic hydrocarbon structure or combination thereof. Representative alkyl groups are those having 24 or fewer carbon atoms, for instance, methyl, ethyl, n-propyl, i-propyl, n-butyl, s-butyl, t-butyl, n-pentyl, i-pentyl, n-hexyl, and the like. Lower alkyl refers to alkyl groups having about 1 to about 6 carbon atoms in the chain. Branched alkyl means that one or more lower alkyl groups such as methyl, ethyl, or propyl are attached to a linear alkyl chain.

[0104] The statement that alkyl is intended to include linear, branched, or cyclic hydrocarbon structures and combinations thereof means that an “alkyl” group also includes

the following combination of linear and cyclic structural elements  (and similar combinations).

[0105] “Alkenyl” means an alkyl, as defined above, containing at least one double bond between adjacent carbon atoms. Alkenyls include both *cis* and *trans* isomers. Branched alkenyl means that one or more lower alkyl groups such as methyl, ethyl, or propyl are attached to a linear alkenyl chain. Representative straight chain and branched alkenyls are those having about 2 to about 6 carbon atoms in the chain, for instance, ethylenyl, propylenyl, 1-butenyl, 2-butenyl, isobutylenyl, 1-pentenyl, 2-pentenyl, 3-methyl-1-butenyl, 2-methyl-2-butenyl, 2,3-dimethyl-2-butenyl, and the like.

[0106] The term “halogen” refers to fluoro, chloro, bromo, and iodo.

[0107] The term “haloalkyl” refers to a branched or straight-chain alkyl as described above, substituted with one or more halogens.

[0108] The term “haloalkenyl” refers to a branched or straight-chain alkenyl as described above, substituted with one or more halogens.

[0109] The term “aryl” means an aromatic monocyclic or multi-cyclic (polycyclic) ring system of 6 to about 19 carbon atoms, for instance, about 6 to about 10 carbon atoms, and includes arylalkyl groups. Representative aryl groups include, but are not limited to, groups such as phenyl, naphthyl, azulenyl, phenanthrenyl, anthracenyl, fluorenyl, pyrenyl, triphenylenyl, chrysenyl, and naphthacenyl.

[0110] The term “arylalkyl” means an alkyl residue attached to an aryl ring. Examples are benzyl, phenethyl, and the like.

[0111] The term “heteroaryl” means an aromatic monocyclic or multi-cyclic ring system of about 5 to about 19 ring atoms, for instance, about 5 to about 10 ring atoms, in which one or more of the atoms in the ring system is/are element(s) other than carbon, for example, nitrogen, oxygen, and/or sulfur. As is well known to those skilled in the art, heteroaryl rings have less aromatic character than their all-carbon counter parts. Thus, for the purposes of the invention, a “heteroaryl” group need only have some degree of aromatic character. For instance, in the case of multi-cyclic ring systems, only one of the rings needs to be aromatic for the ring system to be defined as “heteroaryl”. Exemplary heteroaryls contain about 5 to 6 ring atoms. The prefix aza, oxa, thia, or thio before heteroaryl means that at least a nitrogen, oxygen, or sulfur atom, respectively, is present as a ring atom. A nitrogen, carbon, or sulfur atom in the heteroaryl ring may be optionally oxidized; the nitrogen may optionally be quaternized. Representative heteroaryls include, but are not limited to, purinyl, pyridyl, 2-oxo-pyridinyl, pyrimidinyl, pyridazinyl, pyrazinyl, triazinyl, furanyl, pyrrolyl, thiophenyl, pyrazolyl, imidazolyl, oxazolyl, isoxazolyl, thiazolyl, isothiazolyl, triazolyl, oxadiazolyl, thiadiazolyl, tetrazolyl, indolyl, isoindolyl, benzofuranyl, benzothiophenyl, indolyl, 2-oxoindolyl, dihydrobenzofuranyl, dihydrobenzothiophenyl, indazolyl, benzimidazolyl, benzooxazolyl, benzothiazolyl, benzoisoxazolyl, benzoisothiazolyl, benzotriazolyl, quinolyl, isoquinolyl, quinazolinyl, cinnolyl, pthalazinyl, quinoxalinyl, and the like.

[0112] The terms “cycloalkyl” and “cycloalkenyl” refer to a non-aromatic, saturated (cycloalkyl) or unsaturated (cycloalkenyl), mono- or multi-cyclic ring system of about 3 to about 8 carbon atoms, for instance, about 5 to about 7 carbon atoms. Exemplary cycloalkyl and cycloalkenyl groups include, without limitation, cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl, norbornyl, cyclopropenyl, cyclobutenyl, cyclopentenyl, cyclohexenyl, cyclophenyl, anti-bicyclopropane, syn-tricyclopropane, and the like.

[0113] As used herein, “heterocycle” or “heterocyclyl” refers to a stable 3- to 18-membered ring (radical) which is saturated, unsaturated, or aromatic, and which consists of carbon atoms and from one to five heteroatoms selected from the group consisting of nitrogen, oxygen and sulfur. For purposes of this invention, the heterocycle may be a monocyclic, bicyclic, or a polycyclic ring system, which may include fused, bridged, or spiro ring systems, including bicyclic rings in which any of the above heterocycles are fused to a benzene ring. The nitrogen, carbon, or sulfur atoms in the heterocycle may be optionally

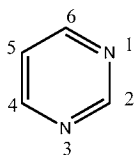
oxidized; the nitrogen atom may be optionally quaternized; and the ring may be partially or fully saturated. The heterocycle may be attached via any heteroatom or carbon atom.

Heterocycles include heteroaryls as defined below. Examples of such heterocycles include, without limitation, morpholinyl, pyrrolidinonyl, pyrrolidinyl, piperidinyl, piperizynyl, hydantoinyl, valerolactamyl, oxiranyl, oxetanyl, tetrahydrofuranyl, tetrahydropyranyl, tetrahydropyridinyl, tetrahydroprimidinyl, tetrahydrothiophenyl, tetrahydrothiopyranyl, tetrahydropyrimidinyl, tetrahydrothiophenyl, tetrahydrothiopyranyl, and the like. Further heterocycles and heteroaryls are described in Katritzky et al., eds., *Comprehensive Heterocyclic Chemistry: The Structure, Reactions, Synthesis and Use of Heterocyclic Compounds*, Vol. 1-8, Pergamon Press, N.Y. (1984), which is hereby incorporated by reference in its entirety.

[0114] The term “acyl” refers to groups of from 1 to 8 carbon atoms of a straight, branched, or cyclic configuration, saturated, unsaturated, or aromatic, and combinations thereof, attached to the parent structure through a carbonyl functionality. One or more carbons in the acyl residue may be replaced by nitrogen, oxygen, or sulfur as long as the point of attachment to the parent remains at the carbonyl. Examples include acetyl (Ac), benzoyl, propionyl, isobutyryl, t-butoxycarbonyl, benzyloxycarbonyl, and the like.

[0115] The term “amino acid” refers to the fragment of an amino acid that remains following amide bond formation via reaction of the amino acid carboxyl group with an amino group of another molecule. The amino acid can be in D- or L-configuration. Suitable amino acids include α -amino acids, β -amino acids, γ -amino acids, δ -amino acids, and ϵ -amino acids, and include not only natural amino acids (*i.e.*, those found in biological systems, including the twenty amino acids found in natural proteins), but also naturally-occurring variants of such amino acids, as well as synthetic amino acids and their analogues known to those skilled in the art. Exemplary amino acids include the twenty natural amino acids, 4-hydroxyproline, hydroxylysine, demosine, isodemosine, 3-methylhistidine, norvalin, beta-alanine, gamma-aminobutyric acid, citrulline, homocysteine, homoserine, ornithine, and methionine sulfone.

[0116] The term “pyrimidine” refers to a heteroaromatic compound containing a benzene ring with two carbon atoms replaced by two nitrogen atoms (diazine). For instance, the following moiety having the carbon atoms at positions 1 and 3 replaced by nitrogen atoms



is considered a pyrimidine:

. This term, as it is defined herein, also includes its

isomeric forms of diazine, such as pyridazine, with the nitrogen atoms in positions 1 and 2; and pyrazine, with the nitrogen atoms in positions 1 and 4. The term “pyrimidine” also generally includes its analogues and derivatives. For instance, the natural nucleobases, cytosine (C), thymine (T), and uracil (U), are pyrimidine derivatives. The term “purine” refers to a heteroaromatic compound containing a pyrimidine ring fused to an imidazole ring. The term “purine” also generally includes its analogues and derivatives. For instance, the natural nucleobases, adenine (A) and guanine (G). Other examples of naturally occurring purine derivatives are hypoxanthine, xanthine, theobromine, caffeine, uric acid, and isoguanine. Exemplary purines and pyrimidines include those disclosed in U.S. Patent No. 3,687,808; Concise Encyclopedia Of Polymer Science And Engineering, pages 858-859; Kroschwitz, J. I., ed. John Wiley & Sons, 1990; and Englisch et al., *Angewandte Chemie*, International Edition, 1991, 30, 613, each of which is hereby incorporated by reference in its entirety.

[0117] The term “nucleobase” includes all natural and synthetic nucleobases as well as universal nucleobases. Typical natural nucleobases include adenine, guanine, cytosine, uracil, and thymine. Synthetic nucleobases typically include inosine, xanthine, hypoxanthine, nubularine, isoguanisine, or tubercidine. As used herein, a universal nucleobase is any modified, unmodified, naturally occurring or non-naturally occurring nucleobase that can substitute for more than one of the natural nucleobases. Universal bases typically contain an aromatic ring moiety that may or may not contain nitrogen atoms and generally use aromatic ring stacking to stabilize an oligonucleotide duplex. Some universal bases can be covalently attached to the C-1' carbon of a pentose sugar to make a universal nucleotide. Some universal bases do not hydrogen bond specifically with another nucleobase. Some universal bases base pair with all of the naturally occurring nucleobases. Some universal bases may interact with adjacent nucleotide bases on the same nucleic acid strand by hydrophobic stacking. Exemplary universal nucleobases include, but are not limited to, 2,4-difluorotoluene, nitropyrrolyl, nitroindolyl, 8-aza-7-deazaadenine, 4-fluoro-6-methylbenzimidazole, 4-methylbenzimidazole, 3-methyl isocarbostyrylyl, 5-methyl isocarbostyrylyl, 3-methyl-7-propynyl isocarbostyrylyl, 7-azaindolyl, 6-methyl-7-azaindolyl, imidizopyridinyl, 9-methyl-imidizopyridinyl, pyrrolopyridinyl, isocarbostyrylyl, 7-propynyl isocarbostyrylyl, propynyl-7-azaindolyl, 2,4,5-trimethylphenyl, 4-methylindolyl, 4,6-dimethylindolyl, phenyl, naphthalenyl, anthracenyl, phenanthracenyl, pyrenyl, stilbenzyl, tetracenyl, pentacenyl, and structural derivatives thereof.

[0118] Suitable nucleobases include, but are not limited to, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 5-halouracil and cytosine, 5-propynyl uracil and cytosine, 6-azo uracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 5-halouracil, 5-(2-aminopropyl)uracil, 5-amino allyl uracil, 8-halo, amino, thiol, thioalkyl, hydroxyl and other 8-substituted adenines and guanines, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine, 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil and 5-propynylcytosine, dihydrouracil, 3-deaza-5-azacytosine, 2-aminopurine, 5-alkyluracil, 7-alkylguanine, 5-alkyl cytosine, 7-deazaadenine, N6,N6-dimethyladenine, 2,6-diaminopurine, 5-amino-allyl-uracil, N3-methyluracil, substituted 1,2,4-triazoles, 2-pyridinone, 5-nitroindole, 3-nitropyrrole, 5-methoxyuracil, uracil-5-oxyacetic acid, 5-methoxycarbonylmethyluracil, 5-methyl-2-thiouracil, 5-methoxycarbonylmethyl-2-thiouracil, 5-methylaminomethyl-2-thiouracil, 3-(3-amino-3-carboxypropyl)uracil, 3-methylcytosine, 5-methylcytosine, N4-acetyl cytosine, 2-thiocytosine, N6-methyladenine, N6-isopentyladenine, 2-methylthio-N-6-isopentenyladenine, N-methylguanines, and O-alkylated bases. Further purines and pyrimidines include those disclosed in U.S. Pat. No. 3,687,808; Concise Encyclopedia Of Polymer Science And Engineering, pages 858-859; Kroschwitz, J. I., ed. John Wiley & Sons, 1990; and Englisch et al., Angewandte Chemie, International Edition, 1991, 30, 613, each of which is hereby incorporated by reference in its entirety.

[0119] The term “nucleoside” refers to a compound comprising a nucleobase, as defined herein, linked to a pentose at the 1'-position. When the nucleobase is a purine derivative or analogue, the pentose is typically attached to the nucleobase at the 9-position of the purine derivative or analogue. When the nucleobase is a pyrimidine derivative or analogue, the pentose is typically attached to the nucleobase at the 1-position of the pyrimidine (*e.g.*, Kornberg and Baker, DNA Replication, 2nd Ed., Freeman, San Francisco, 1992, which is hereby incorporated by reference in its entirety). When a nucleoside is present in R³, R⁴, or R⁵ herein, the nucleoside may be connected to the neighboring atom(s) through any atom on the nucleobase or pentose.

[0120] The term “fatty acid” generally refers to a carboxylic acid with an aliphatic tail (chain). The aliphatic chain can be between about 2 and about 36 carbon atoms in length. Fatty acids can be saturated, unsaturated, or polyunsaturated. The aliphatic chain can be a linear or a branched chain. The term “fatty acid” may be used herein to refer to a “fatty acid

derivative” which can include one or more different fatty acid derivatives, or mixtures of fatty acids derivatives. Exemplary fatty acids include unsaturated fatty acids, saturated fatty acids, and diacids; mono-, di-, and tri-glycerides of ascarosides that have a carboxylic acid functionality; hydroxy acids, ω hydroxy acids, ω -1 hydroxy acids, di-hydroxy fatty acids (*e.g.*, dihydroxy fatty acids that are omega- or omega-1 hydroxylated, as well as alpha- or beta-hydroxylated fatty acids).

[0121] The term “sugar” refers to a compound which is either a carbohydrate per se made up of one or more monosaccharide units having at least 5 carbon atoms (which may be linear, branched, or cyclic) with an oxygen, nitrogen, or sulfur atom bonded to each carbon atom; or a compound having as a part thereof a carbohydrate moiety made up of one or more monosaccharide units each having at least 5 carbon atoms (which may be linear, branched or cyclic), with an oxygen, nitrogen or sulfur atom bonded to each carbon atom. Representative sugars include the mono-, di-, tri-, and oligosaccharides containing from about 4–9 monosaccharide units, and polysaccharides such as starches, glycogen, cellulose, and polysaccharide gums. Exemplary monosaccharides include C_5 and above (*e.g.*, C_5 – C_8 or C_5 – C_6) sugars; di- and trisaccharides include sugars having two or three monosaccharide units (*e.g.*, C_5 – C_8 or C_5 – C_8).

[0122] The term “monosaccharide” means a sugar molecule having a chain of 3–10 carbon atoms in the form of an aldehyde (aldose) or ketone (ketose). Suitable monosaccharides include both naturally occurring and synthetic monosaccharides. Suitable monosaccharides include trioses, such as glycerose and dihydroxyacetone; tetroses such as erythrose and erythrulose; pentoses, such as xylose, arabinose, ribose, xylulose ribulose; methyl pentoses (6-deoxyhexoses), such as rhamnose and fucose; hexoses, such as ascarlose, glucose, mannose, galactose, fructose, and sorbose; and heptoses, such as glucoheptose, galamannoheptose, sedoheptulose, and mannoheptulose. Exemplary monosaccharides embrace radicals of allose, altrose, arabinose, cladinose, erythrose, erythrulose, fructose, D-fucitol, L-fucitol, fucosamine, fucose, fuculose, galactosamine, D-galactosaminitol, N-acetyl-galactosamine, galactose, glucosamine, N-acetyl-glucosamine, glucosaminitol, glucose, glucose-6-phosphate, gulose glyceraldehyde, L-glycero-D-mannose-heptose, glycerol, glycerone, gulose, idose, lyxose, mannosamine, mannose, mannose-6-phosphate, psicose, quinovose, quinovasamine, rhamnitol, rhamnosamine, rhamnose, ribose, ribulose, sedoheptulose, sorbose, tagatose, talose, tartaric acid, threose, xylose, and xylulose.

The monosaccharide can be in D- or L- configuration. A typical monosaccharide used herein is hexose.

[0123] The monosaccharide may further be a deoxy sugar (alcoholic hydroxy group replaced by hydrogen), amino sugar (alcoholic hydroxy group replaced by amino group), a thio sugar (alcoholic hydroxy group replaced by thiol, or C=O replaced by C=S, or a ring oxygen of cyclic form replaced by sulfur), a seleno sugar, a telluro sugar, an aza sugar (ring carbon replaced by nitrogen), an imino sugar (ring oxygen replaced by nitrogen), a phosphano sugar (ring oxygen replaced with phosphorus), a phospho sugar (ring carbon replaced with phosphorus), a C-substituted monosaccharide (hydrogen at a non-terminal carbon atom replaced with carbon), an unsaturated monosaccharide, an alditol (carbonyl group replaced with CHOH group), aldonic acid (aldehydic group replaced by carboxy group), a ketoaldonic acid, a uronic acid, an aldaric acid, and so forth. Amino sugars include amino monosaccharides, such as galactosamine, glucosamine, mannosamine, fucosamine, quinovasamine, neuraminic acid, muramic acid, lactosedi-amine, acosamine, bacillosamine, daunosamine, desosamine, forosamine, garosamine, kanosamine, kansosamine, mycaminose, mycosamine, perosamine, pneumosamine, purpurosamine, rhodosamine. It is understood that the monosaccharide and the like can be further substituted.

[0124] The terms “disaccharide”, “trisaccharide”, and “polysaccharide” embrace radicals of abequose, acra-bose, amicitose, amylopectin, amylose, apiose, arcanose, ascarylose, ascorbic acid, boivino-se, cellobiose, cellotriose, cellulose, chacotriose, chalcose, chitin, colitose, cyclodextrin, cymarose, dextrin, 2-deoxyribose, 2-deoxyglucose, diginose, digitalose, digitoxose, evalose, evemitrose, fructoologosa-cc-haride, galto-oligosaccharide, gentianose, gentiobiose, glucan, glucogen, glycogen, hamamelose, heparin, inulin, isolevoglucosenone, isomaltose, isomaltotriose, isopanose, kojibiose, lactose, lactosamine, lactosedi-amine, laminarabiose, levoglucosan, levoglucosenone, β -maltose, maltriose, mannan-oligosaccharide, manninotriose, melezitose, melibiose, muramic acid, mycarose, mycinose, neuraminic acid, nigerose, nojirimycin, moviose, oleandro-se, panose, paratose, planteose, primeverose, raffinose, rhodino-se, rutinose, sarmentose, sedoheptulose, solatriose, sophorose, stachyose, streptose, sucrose, α , α -trehalose, trehalosamine, turanose, tyvelose, xylobiose, umbelliferose, and the like. Further, it is understood that the “disaccharide”, “trisaccharide”, and “polysaccharide” and the like can be further substituted. Disaccharide also includes amino sugars and their derivatives, particularly, a mycaminose derivatized at the C-4' position or a 4 deoxy-3-amino-glucose derivatized at the C-6' position.

[0125] The term “polycyclic” or “multi-cyclic” used herein indicates a molecular structure having two or more rings, including, but not limited to, fused, bridged, or spiro rings.

[0126] The above “alkyl”, “alkenyl”, “cycloalkyl”, and “cycloalkenyl” radicals, as well as the ring system of the above aryl, heterocyclyl, or heteroaryl groups, may be optionally substituted.

[0127] The term “substituted” or “optionally substituted” is used to indicate that a group may have a substituent at each substitutable atom of the group (including more than one substituent on a single atom), provided that the designated atom’s normal valency is not exceeded and the identity of each substituent is independent of the others. In accordance with the present invention, up to three H atoms in each residue can be replaced with alkyl, halogen, haloalkyl, alkenyl, haloalkenyl, cycloalkyl, cycloalkenyl, hydroxy, alkoxy, acyl, carboxy, carboalkoxy (also referred to as alkoxycarbonyl), carboxamido (also referred to as alkylaminocarbonyl), cyano, carbonyl, nitro, amino, alkylamino, dialkylamino, mercapto, alkylthio, sulfoxide, sulfone, acylamino, amidino, aryl, heteroaryl, heterocyclyl, aryloxy, heteroaryloxy, a purine or pyrimidine or an analogue or derivative thereof (as defined in “nucleobase”), or a sugar such as a monosaccharide having 5 or 6 carbon atoms (as defined in “monosaccharide”). “Unsubstituted” atoms bear all of the hydrogen atoms dictated by their valency. When a substituent is keto (*i.e.*, =O), then two hydrogens on the atom are replaced. Combinations of substituents and/or variables are permissible only if such combinations result in stable compounds; by “stable compound” or “stable structure” is meant a compound that is sufficiently robust to survive isolation to a useful degree of purity from a reaction mixture, and formulation into an efficacious agent.

[0128] In the characterization of some of the substituents, certain substituents may combine to form rings. Unless stated otherwise, it is intended that such rings may exhibit various degrees of unsaturation (from fully saturated to fully unsaturated), may include heteroatoms, and may be substituted with other substituent groups as described above.

[0129] The compounds described herein may contain one or more asymmetric centers and may thus give rise to enantiomers, diastereomers, and other stereoisomeric forms. Each chiral center may be defined, in terms of absolute stereochemistry, as (R)- or (S)-. The present invention is meant to include all such possible isomers, as well as mixtures thereof, including racemic and optically pure forms. Optically active (R)- and (S)-, (-)- and (+)-, or (D)- and (L)-isomers may be prepared using chiral synthons or chiral reagents, or resolved

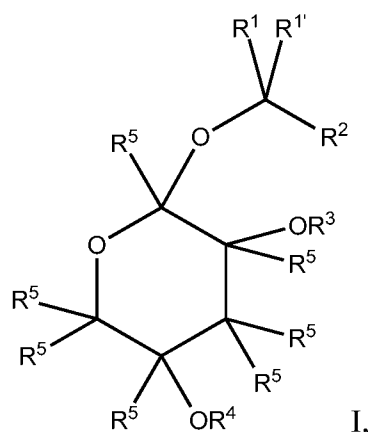
using conventional techniques. When the compounds described herein contain olefinic double bonds or other centers of geometric asymmetry, and unless specified otherwise, it is intended that the compounds include both E and Z geometric isomers. Likewise, all tautomeric forms are also intended to be included. The configuration of any carbon-carbon double bond appearing herein is selected for convenience only and is not intended to designate a particular configuration; thus a carbon-carbon double bond depicted arbitrarily herein as *trans* may be Z, E, or a mixture of the two in any proportion.

[0130] The term “compounds of the invention,” and equivalent expressions, are meant to embrace the prodrugs, the pharmaceutically acceptable salts, the oxides, the solvates, *e.g.* hydrates, and inclusion complexes of that compound, where the context so permits, as well as any stereoisomeric form, or a mixture of any such forms of that compound in any ratio, unless otherwise specified. Inclusion complexes are described in Remington, The Science and Practice of Pharmacy, 19th Ed. 1:176-177 (1995), which is hereby incorporated by reference in its entirety. The most commonly employed inclusion complexes are those with cyclodextrins, and all cyclodextrin complexes, natural and synthetic, are specifically encompassed within the claims. Thus, in accordance with some embodiments of the invention, a compound as described herein, including in the contexts of pharmaceutical compositions, methods of treatment, and compounds per se, is provided as the salt form. Similarly, reference to intermediates, whether or not they themselves are claimed, is meant to embrace their salts, and solvates, where the context so permits. For the sake of clarity, particular instances when the context so permits are sometimes indicated in the text, but these instances are purely illustrative and it is not intended to exclude other instances when the context so permits.

[0131] The “quaternization” of any basic nitrogen-containing groups of the compounds disclosed herein is also contemplated. The basic nitrogen can be quaternized with any agents known to those of ordinary skill in the art including, for example, lower alkyl halides, such as methyl, ethyl, propyl and butyl chloride, bromides and iodides; dialkyl sulfates including dimethyl, diethyl, dibutyl and diamyl sulfates; long chain halides such as decyl, lauryl, myristyl and stearyl chlorides, bromides and iodides; and aralkyl halides including benzyl and phenethyl bromides. Water or oil-soluble or dispersible products may be obtained by such quaternization.

[0132] A first aspect of the present invention relates to a method of modifying nematode behavior. This method involves administering one or more isolated modulator

compounds to the nematode under conditions effective to modify nematode behavior. In this aspect of the present invention, the one or more modulator compounds is (i) a compound of Formula I:



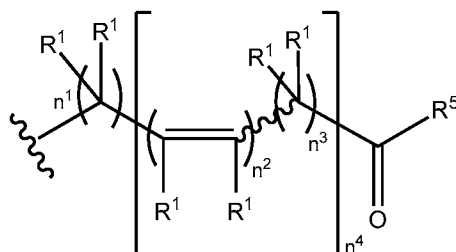
where:

R^1 is H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl;

where each R is independently H, halogen, an alkyl, or an alkenyl;

$R^{1'}$ is absent, H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl; where each R is independently H, halogen, an alkyl, or an alkenyl;

R^2 is a moiety of formula



where:

each R^1 is independently H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl; where each R is independently H, halogen, an alkyl, or an alkenyl;

R^5 is H, $-OH$, $-OR^6$, $-OCR^6R^7R^8$, $-CR^6R^7R^8$, $-NH_2$, $-NHR^6$, $-NR^6R^7$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, an arylalkyl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R^3 or R^4 of another unit of Formula I;

where:

R^6 and R^7 are each independently H, $-CR_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, or a cycloalkenyl, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, or cycloalkenyl is optionally substituted with one or more substituents independently selected from the group consisting of $-OR^8$, $-C(O)R^8$, $-NHC(O)R^8$, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl;

where:

each R is independently H, halogen, an alkyl, or an alkenyl; and

R^8 is H, $-C(R)_3$, $-[C(R)_2]_{n4}NHC(O)R^9$, $-[C(R)_2]_{n4}C(O)(NH)R^9$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR^9$, $-C(O)R^9$, $-NHC(O)R^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms;

where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR$, $-C(O)R$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a

heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and n^4 is an integer of 1 to 30; and R^8 is H, $-C(R)_3$, $-[C(R)_2]_{n^4}NHC(O)R^9$, $-[C(R)_2]_{n^4}C(O)(NH)R^9$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR^9$, $-C(O)R^9$, $-NHC(O)R^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms;

where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR$, $-C(O)R$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30;

n^1 , n^2 , and n^3 are each independently an integer of 0 to 30;

n^4 is an integer of 1 to 30; and

the sum of n^1 , each n^2 , and each n^3 is 1 to 30;

R^3 and R^4 are each independently H, $-CR^6R^7R^8$, $-C(O)R^8$, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl,

an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R⁵ of another unit of Formula I;

where:

R⁶ and R⁷ are each independently H, -CR₃, -OR, -N(R)₂, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, or a cycloalkenyl, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, or cycloalkenyl is optionally substituted with one or more substituents independently selected from the group consisting of -OR⁸, -C(O)R⁸, -NHC(O)R⁸, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl;

where:

each R is independently H, halogen, an alkyl, or an alkenyl; and

R⁸ is H, -C(R)₃, -[C(R)₂]_{n4}NHC(O)R⁹, -[C(R)₂]_{n4}C(O)(NH)R⁹, -OR, -N(R)₂, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of -C(R)₃, -OR⁹, -C(O)R⁹, -NHC(O)R⁹, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms;

where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R⁹ is H, -C(R)₃, -OR, -N(R)₂, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of -C(R)₃, -OR, -C(O)R, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a

cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; and

R^8 is H, $-C(R)_3$, $-[C(R)_2]_{n^4}NHC(O)R^9$, $-[C(R)_2]_{n^4}C(O)(NH)R^9$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR^9$, $-C(O)R^9$, $-NHC(O)R^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms; where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR$, $-C(O)R$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; and

each R^5 is independently H, $-OH$, $-OR^6$, $-OCR^6R^7R^8$, $-CR^6R^7R^8$, $-NH_2$, $-NHR^6$, $-NR^6R^7$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, an arylalkyl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R^3 or R^4 of another unit of Formula I;

where:

R^6 and R^7 are each independently H, $-CR_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a

cycloalkyl, or a cycloalkenyl, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, or cycloalkenyl is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{OR}^8$, $-\text{C}(\text{O})\text{R}^8$, $-\text{NHC}(\text{O})\text{R}^8$, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl;

where:

each R is independently H, halogen, an alkyl, or an alkenyl; and

R^8 is H, $-\text{C}(\text{R})_3$, $-\text{[C}(\text{R})_2\text{]}_{n4}\text{NHC}(\text{O})\text{R}^9$, $-\text{[C}(\text{R})_2\text{]}_{n4}\text{C}(\text{O})(\text{NH})\text{R}^9$, $-\text{OR}$, $-\text{N}(\text{R})_2$,

halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}^9$, $-\text{C}(\text{O})\text{R}^9$, $-\text{NHC}(\text{O})\text{R}^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms;

where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{N}(\text{R})_2$, halogen, an alkyl, a haloalkyl, an

alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{C}(\text{O})\text{R}$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; and

R^8 is H, $-\text{C}(\text{R})_3$, $-\text{[C}(\text{R})_2\text{]}_{n4}\text{NHC}(\text{O})\text{R}^9$, $-\text{[C}(\text{R})_2\text{]}_{n4}\text{C}(\text{O})(\text{NH})\text{R}^9$, $-\text{OR}$, $-\text{N}(\text{R})_2$,

halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl,

a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}^9$, $-\text{C}(\text{O})\text{R}^9$, $-\text{NHC}(\text{O})\text{R}^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms; where:

each R is independently H, halogen, an alkyl, or an alkenyl;

R^9 is H, $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{N}(\text{R})_2$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, where the alkyl, alkenyl, aryl, heteroaryl, heterocyclyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-\text{C}(\text{R})_3$, $-\text{OR}$, $-\text{C}(\text{O})\text{R}$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, and a cycloalkyl; where each R is independently H, halogen, an alkyl, or an alkenyl; and

n^4 is an integer of 1 to 30; or

(ii) a compound comprising:

at least one nucleobase,

at least one fatty acid,

at least one amino acid, and

at least one sugar;

where the at least one nucleobase, the at least one fatty acid, the at least one amino acid, and the at least one sugar are linked by covalent bonds; and

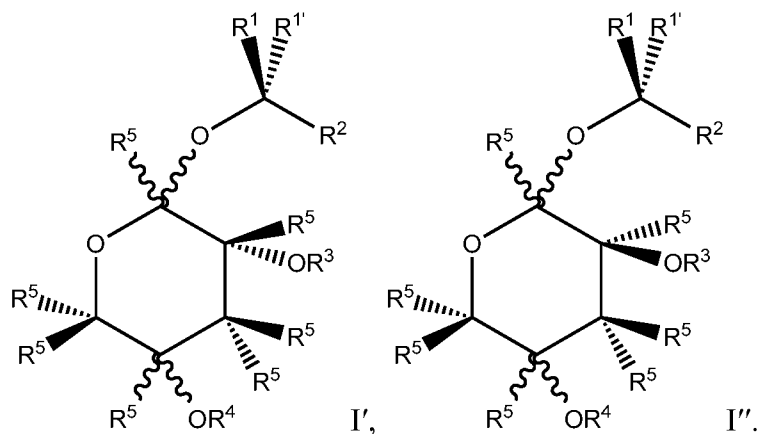
where the compound has a molecular weight of less than about 2,000 g/mol;

with the proviso that the nematode is not a *Caenorhabditis* species.

[0133] As will be understood by the skilled artisan, administering according to this aspect of the present invention may include directly contacting the nematode with the one or more modulator compounds, and/or contacting a nematode-susceptible location with the one

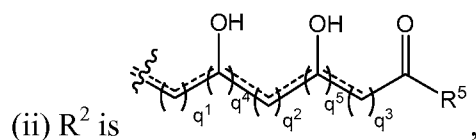
or more modulator compounds, as long as the one or more modulator compounds is administered such that it is able to affect nematode behavior.

[0134] In at least one embodiment of this aspect of the present invention, the one or more modulator compounds is a compound of Formula I. Suitable modulator compounds according to this embodiment include one or more isolated modulator compounds of Formula I' or Formula I'':



Suitable modulator compounds according to this embodiment also include one or more isolated modulator compounds of Formula I in which at least one of the following conditions is met:

(i) R¹ is independently -C(R')₃, -OR, -N(R)₂, halogen, a haloalkyl, an alkenyl, or a haloalkenyl; wherein each R is independently H, halogen, an alkyl, or an alkenyl, and each R' is independently halogen or an alkenyl; and
R^{1'} is independently absent, H, -C(R)₃, -OR, -N(R)₂, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl;



wherein:

each is independently a single or double bond;

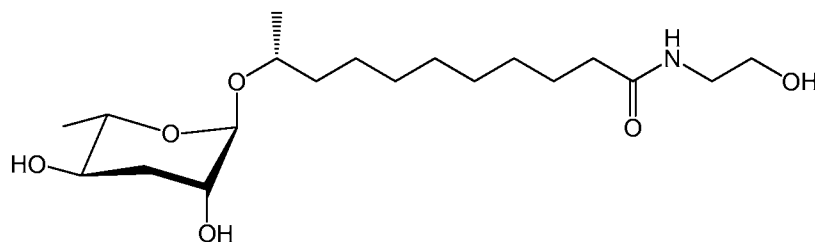
q^1, q^2 , and q^3 are each independently an integer of 1 to 26;

q^4 and q^5 are each independently an integer of 0 to 26;

the sum of q^1, q^2, q^3, q^4 , and q^5 is less than or equal to 28; and

the sum of q^4 and q^5 is greater than or equal to 2;

- (iii) R^5 is H, $-OR^6$, $-OCR^6R^7R^8$, $-CR^6R^7R^8$, $-NH_2$, $-NR^6R^7$, halogen, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, an arylalkyl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, or a nucleoside;
- (iv) R^5 is a bond connecting to R^3 or R^4 of another unit of Formula I, forming a compound containing at least two units of Formula I;
- (v) R^3 and R^4 are each independently $-CR^6R^7R^8$, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, or $-(CO)R^8$ wherein R^8 is H, $-C(R)_3$, $-[C(R)_2]_{n4}NHC(O)R^9$, $-[C(R)_2]_{n4}C(O)(NH)R^9$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, a cycloalkyl, a cycloalkenyl, a purine, a pyrimidine, or a monosaccharide having 5 or 6 carbon atoms, wherein the alkyl, cycloalkyl, cycloalkenyl, purine, pyrimidine, or monosaccharide is optionally substituted with one or more substituents independently selected from the group consisting of $-C(R)_3$, $-OR^9$, $-C(O)R^9$, $-NHC(O)R^9$, halogen, an alkyl, a haloalkyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, and a monosaccharide having 5 or 6 carbon atoms;
- (vi) the compound is a compound of Formula I'';
- with the proviso that the compound is not



[0135] In at least one embodiment of this aspect of the present invention, the one or more modulator compounds is a compound comprising at least one nucleobase, at least one fatty acid, at least one amino acid, and at least one sugar.

[0136] In this and all aspects of the present invention, a single modulator compound or a combination of modulator compounds can be administered.

[0137] In this aspect of the present invention, the nematode is preferably selected from the group consisting of plant-parasitic nematodes and entomopathogenic nematodes.

[0138] Suitable plant-parasitic nematodes according to this aspect of the present invention include *Acontylus*, *Afenestrata*, *Aglenchus*, *Allotrichodorus*, *Allotylenchus*, *Amplimerlinius*, *Anguina*, *Antarctenchus*, *Antarctylus*, *Aorolaimus*, *Aphasmatylenchus*, *Aphelenchoides*, *Apratylenchoides*, *Atalodera*, *Atetylenchus*, *Atylenchus*, *Axodorylaimellus*,

Axonchium, Bakernema, Basiria, Basirienchus, Bellodera, Belondira, Belonolaimus, Bitylenchus, Blandicephalonema, Boleodorus, Brachydorus, Bursadera, Bursaphelenchus, Cacopaurus, Cactodera, Caloosia, Campbellenchus, Carphodorus, Cephalenchus, Chitinotylenchus, Coslenchus, Criconema, Criconemella, Criconemoides, Crossonema, Cryphodera, Cucullitylenchus, Cynipanguina, Discocriconemella, Ditylenchus, Dolichodera, Dolichodorus, Dolichorhynchus, Dorylaimellus, Duotylenchus, Ecphyadophora, Ecphyadophoroides, Epicharinema, Eutylenchus, Geocenamus, Globodera, Gracilacus, Gracilancea, Halenchus, Helicotylenchus, Hemicriconemoides, Hemicycliophora, Heterodera, Hirschmanniella, Hoplolaimus, Hoplotylus, Hylonema, Immanigula, Irantylenchus, Laimaphelenchus, Lelenchus, Longidorella, Longidorus, Loofia, Macrotriphurus, Malenchus, Meloidodera, Meloidoderella, Meloidoderita, Meloidogyne, Meloinema, Merlinius, Mesocriconema, Metaxonchium, Miculenchus, Mitranema, Monotrichodorus, Morulaimus, Mukazia, Nacobbodera, Nacobbus, Nagelus, Neodolichodorus, Neodolichorhynchus, Neopsilenchus, Neothada, Nimigula, Nothocriconema, Nothocriconemoides, Ogma, Opailaimus, Paralongidorus, Pararotylenchus, Paratrichodorus, Paratrophurus, Paratylenchus, Pateracephalonema, Phallaxonchium, Pleurotylenchus, Polenchus, Pratylenchoides, Pratylenchus, Probelondira, Pseudhalenchus, Psilenchus, Pterotylenchus, Punctodera, Quinisulcius, Radopholus, Rhizonema, Rotylenchulus, Rotylenchus, Sarisodera, Sauertylenchus, Scutellonema, Scutylenchus, Senegalonema, Siddiqia, Sphaeronema, Subanguina, Swangeria, Sychnotylenchus, Syncheilaxonchium, Telotylenchus, Tetylenchus, Thada, Thecavermiculatus, Trichodorus, Trichotylenchus, Triversus, Trophonema, Trophotylenchulus, Trophurus, Tylenchocriconema, Tylenchorhynchus, Tylenchulus, Tylenchus, Tylodorus, Verutus, Xenocriconemella, Xiphinema, and Zygotylenchus.

[0139] Suitable entomopathogenic nematodes according to this aspect of the present invention include *Steinernema abbasi*, *Steinernema aciari*, *Steinernema affine*, *Steinernema akhursti*, *Steinernema anatoliense*, *Steinernema apuliae*, *Steinernema arenarium*, *Steinernema ashiuense*, *Steinernema asiaticum*, *Steinernema australe*, *Steinernema backanese*, *Steinernema bedding*, *Steinernema biocornutum*, *Steinernema ceratophorum*, *Steinernema cholashansense*, *Steinernema citrae*, *Steinernema cubanum*, *Steinernema cumgarensense*, *Steinernema diaprepsi*, *Steinernema eapokense*, *Steinernema everestense*, *Steinernema feltiae*, *Steinernema glaseri*, *Steinernema guangdongense*, *Steinernema hebeinse*, *Steinernema hermaphroditum*, *Steinernema ichnusae*, *Steinernema intermedium*,

Steinernema jolietii, *Steinernema kariii*, *Steinernema khoisanae*, *Steinernema kraussei*, *Steinernema kushidai*, *Steinernema leizhouense*, *Steinernema lici*, *Steinernema litorale*, *Steinernema longicaudatum*, *Steinernema monticolum*, *Steinernema neocurtillae*, *Steinernema oregonense*, *Steinernema pakistanense*, *Steinernema phyllogphagae*, *Steinernema puertoricense*, *Steinernema rarum*, *Steinernema riobrave*, *Steinernema ritteri*, *Steinernema robustispiculum*, *Steinernema sangi*, *Steinernema sasonense*, *Steinernema scapterisci*, *Steinernema scarabaei*, *Steinernema schliemanni*, *Steinernema siamkayai*, *Steinernema sichuanense*, *Steinernema silvaticum*, *Steinernema tami*, *Steinernema texanum*, *Steinernema thanhi*, *Steinernema websteri*, *Steinernema weiseri*, *Steinernema yirgalemense*, *Heterorhabditis amazonensis*, *Heterorhabditis bacteriophora*, *Heterorhabditis baujardi*, *Heterorhabditis downesi*, *Heterorhabditis floridensis*, *Heterorhabditis indica*, *Heterorhabditis marelatus*, *Heterorhabditis megidis*, *Heterorhabditis mexicana*, *Heterorhabditis taysearae*, *Heterorhabditis zealandica*, and *Heterorhabditis sonorensis*.

[0140] Nematode behavior according to this aspect of the present invention includes reproduction, development, dauer formation, aggregation, attraction, repulsion, dispersal, deterrence, feeding, host finding, and host invasion.

[0141] A second aspect of the present invention relates to a method of promoting or inhibiting reproduction in a nematode population. This method involves administering one or more isolated modulator compounds to the population under conditions effective to promote or inhibit reproduction in the nematode population. In this aspect of the present invention, the one or more isolated modulator compounds is selected from the group consisting of ascr#1; ascr#3; ascr#7; ascr#8; ascr#9; ascr#10; ascr#16; ascr#18; ascr#20; ascr#22; a combination of ascr#9 and ascr#10; a combination of ascr#9 and ascr#16; a combination of ascr#9 and ascr#18; a combination of ascr#9 and ascr#20; and a combination of ascr#9 and ascr#22.

[0142] Suitable isolated modulator compounds for promoting reproduction according to this aspect of the present invention include ascr#1, ascr#3, ascr#7, ascr#8, and ascr#10. These modulator compounds can be used to attract male and/or female nematodes, thereby increasing the opportunity for mating.

[0143] Suitable isolated modulator compounds for inhibiting reproduction according to this aspect of the present invention include ascr#9; ascr#10; ascr#16; ascr#18; ascr#20; ascr#22; a combination of ascr#9 and ascr#10; a combination of ascr#9 and ascr#16; a combination of ascr#9 and ascr#18; a combination of ascr#9 and ascr#20; and a combination

of ascr#9 and ascr#22. These modulator compounds can be used to repel male and/or female nematodes, thereby reducing the opportunity for mating.

[0144] Suitable nematodes according to this aspect of the present invention include *Meloidogyne javanica*, *Meloidogyne floridensis*, *Meloidogyne incognita*, *Meloidogyne hapla*, *Steinernema glaseri*, *Heterorhabditis bacteriophora*, *Heterorhabditis zealandica*, and *Heterorhabditis floridensis*.

[0145] A third aspect of the present invention relates to a method of promoting or inhibiting nematode aggregation at a first location. This method involves (i) contacting the first location with one or more isolated modulator compounds under conditions effective to promote or inhibit nematode aggregation at the first location, or (ii) contacting a second location with one or more isolated modulator compounds under conditions effective to promote or inhibit nematode aggregation at the first location, where the first location and the second location are spaced to permit said contacting at the second location to have an effect on nematode aggregation at the first location. In this aspect of the present invention, the one or more isolated modulator compounds is selected from the group consisting of ascr#1; ascr#2; ascr#3; ascr#7; ascr#8; ascr#9; ascr#10; ascr#11; ascr#16; ascr#18; ascr#20; ascr#22; icas#9; oscr#10; a combination of ascr#9 and ascr#10; a combination of ascr#9 and ascr#16; a combination of ascr#9 and ascr#18; a combination of ascr#9 and ascr#20; a combination of ascr#9 and ascr#22; a combination of ascr#2, ascr#3, ascr#8, and icas#9; a combination of ascr#9, ascr#3, ascr#8, and icas#9; a combination of ascr#9 and oscr#10; and a combination of ascr#9, oscr#10, and ascr#11.

[0146] Suitable isolated modulator compounds for promoting aggregation according to this aspect of the present invention include ascr#1, ascr#3, ascr#7, ascr#8, and ascr#10. These modulator compounds can be used to attract male and/or female nematodes, thereby increasing aggregation.

[0147] Suitable isolated modulator compounds for inhibiting aggregation according to this aspect of the present invention include ascr#2; ascr#3; ascr#8; ascr#9; ascr#10; ascr#11; ascr#16; ascr#18; ascr#20; ascr#22; icas#9; oscr#10; a combination of ascr#9 and ascr#10; a combination of ascr#9 and ascr#16; a combination of ascr#9 and ascr#18; a combination of ascr#9 and ascr#20; a combination of ascr#9 and ascr#22; a combination of ascr#2, ascr#3, ascr#8, and icas#9; a combination of ascr#9, ascr#3, ascr#8, and icas#9; a combination of ascr#9 and oscr#10; and a combination of ascr#9, oscr#10, and ascr#11. These modulator

compounds can be used to repel male, female, and/or larval nematodes, thereby decreasing aggregation.

[0148] In at least one embodiment of this aspect of the present invention, the first location can be contacted with one or more modulator compounds that attract desirable nematodes to the location.

[0149] In at least one embodiment of this aspect of the present invention, the first location can be contacted with one or more modulator compounds that repel undesirable nematodes already at the first location or deter undesirable nematodes from approaching the first location.

[0150] In at least one embodiment of this aspect of the present invention, the second location can be contacted with one or more modulator compounds that attract undesirable nematodes to the second location and away from the first location. In these embodiments, the second location can optionally be contacted with a toxin that is harmful to the nematode. Suitable nematode toxins include, without limitation, chitosan, abamectin, neem cake, marigold extract, nematophagus fungi, Aldicarb, and Dazomet.

[0151] In at least one embodiment of this aspect of the present invention, the second location can be contacted with one or more modulator compounds that repel/deter desirable nematodes from the second location to the first location.

[0152] Embodiments in which both the first location and the second location are contacted with one or more modulator compounds are also contemplated. For example, the first location can be contacted with one or more modulator compounds that attract desirable nematodes to the first location while the second location is contacted with one or more modulator compounds that repel/deter desirable nematodes away from the second location to the first location. Alternatively or additionally, the first location can be contacted with one or more modulator compounds that repel/deter undesirable nematodes from the first location while the second location is contacted with one or more modulator compounds that attract undesirable nematodes away from the first location.

[0153] As will be apparent to one of ordinary skill in the art, when the second location is contacted with the one or more modulator compounds, the second location should be spaced close enough to the first location for the one or more modulator compounds to have an effect on aggregation at the first location. Preferably, the second location is between about 1 cm and about 200 cm away from the first location. As will be apparent to the skilled artisan, suitable distances will depend, *inter alia*, upon the size of the plant and include, for example,

between about 1 cm and about 50 cm, between about 2 cm and about 50 cm, between about 2 cm and about 200 cm, between about 50 cm and about 200 cm, about 1 cm, about 2 cm, about 50 cm, and about 200 cm.

[0154] By way of example, one or more modulator compounds that repel/deter undesirable nematodes can be placed at a perimeter around the first location (*e.g.*, a location susceptible to or harboring nematodes), to prevent undesirable nematodes outside the perimeter from moving toward the first location. Alternatively, one or more modulator compounds that repel/deter desirable nematodes can be placed at a perimeter around the first location, to prevent desirable nematodes within the perimeter from moving away from, or escaping from within the perimeter of, the first location. In the latter example, the methods of the present invention can be used in concert with applying desirable nematodes themselves to the first location.

[0155] In one embodiment of this aspect of the present invention the first location is a plant (including, *e.g.*, plant parts such as roots). In another embodiment, the first location is a population of insects. Embodiments in which the first location is a population of insects at a plant are also contemplated. Other suitable first locations include soil (*e.g.*, adjacent to plant roots), sand, loam, artificial culture substrates, peat, and moss.

[0156] When the first location is at a plant, desirable nematodes are those that are beneficial to the plant, and undesirable nematodes are those that are harmful to the plant. When the first location is at a population of insect pests (*e.g.*, insects that are harmful to plants), desirable nematodes are those that are harmful to the insects, and undesirable nematodes are those that are beneficial to the insects. When the first location is at a population of insects that are beneficial (*e.g.*, plant-beneficial insects), desirable nematodes are those that are beneficial to the insects, and undesirable nematodes are those that are harmful to the insects.

[0157] Nematodes harmful to plants according to this and all aspects of the present invention include those that infect the plant itself, as well as those that infect insects that are beneficial to the plant. Nematodes that infect plants and/or infect plant-beneficial insects according to the present invention include *Meloidogyne javanica*, *Meloidogyne floridensis*, *Meloidogyne incognita*, *Meloidogyne hapla*, *Steinernema riobrave*, *Steinernema diaprepesi*, *Steinernema carpocapse*, *Steinernema feltiae*, *Steinernema glaseri*, *Heterorhabditis bacteriophora*, *Heterorhabditis zealandica*, and *Heterorhabditis floridensis*.

[0158] Plant-beneficial nematodes according to this and all aspects of the present invention include those that infect plant parasitic insects and/or plant parasitic nematodes. Nematodes that infect plant parasitic insects and/or plant parasitic nematodes according to the present invention include *Steinernema riobrave*, *Steinernema diaprepesi*, *Steinernema carpocapse*, *Steinernema feltiae*, *Steinernema glaseri*, *Heterorhabditis bacteriophora*, *Heterorhabditis zealandica*, and *Heterorhabditis floridensis*.

[0159] Nematodes harmful to insects, *i.e.*, entomopathogenic nematodes, are those that infect the insect. In some embodiments, the entomopathogenic nematodes infect plant-parasitic insects. In other embodiments, the entomopathogenic nematodes infect plant-beneficial insects. Entomopathogenic nematodes that infect plant-parasitic insects according to the present invention include *Steinernema riobrave*, *Steinernema diaprepesi*, *Steinernema carpocapse*, *Steinernema feltiae*, *Steinernema glaseri*, *Heterorhabditis bacteriophora*, *Heterorhabditis zealandica*, and *Heterorhabditis floridensis*. Entomopathogenic nematodes that infect plant-beneficial insects according to the present invention include *Steinernema riobrave*, *Steinernema diaprepesi*, *Steinernema carpocapse*, *Steinernema feltiae*, *Steinernema glaseri*, *Heterorhabditis bacteriophora*, *Heterorhabditis zealandica*, and *Heterorhabditis floridensis*.

[0160] Suitable plants according to this and all aspects of the present invention involving plants include, without limitation, dicots and monocots. More particularly, useful crop plants can include, without limitation, alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, green bean, wax bean, lima bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, sugar beet, parsnip, turnip, cauliflower, broccoli, turnip, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, and sugarcane. Examples of suitable ornamental plants include, without limitation, *Arabidopsis thaliana*, Saintpaulia, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

[0161] Suitable plant-parasitic insects according to this and all aspects of the present invention involving insects include, without limitation, European corn borer, beet armyworm, cabbage looper, corn ear worm, fall armyworm, diamondback moth, cabbage root maggot, onion maggot, seed corn maggot, pickleworm (melonworm), pepper maggot, and tomato pinworm. European corn borer is a major pest of corn (dent and sweet corn) but also feeds on over 200 plant species, including green beans, wax beans, lima beans, soybeans, peppers,

potato, tomato, and many weed species. Additional insect larval feeding pests which damage a wide variety of vegetable crops include, without limitation, beet armyworm, cabbage looper, corn ear worm, fall armyworm, diamondback moth, cabbage root maggot, onion maggot, seed corn maggot, pickleworm (melonworm), pepper maggot, and tomato pinworm. Collectively, this group of insect pests represents the most economically important group of pests for vegetable production worldwide.

[0162] Contacting according to this and all aspects of the present invention can be carried out through a variety of procedures that will be apparent to the skilled artisan. Suitable application methods include high or low pressure spraying, immersion, atomizing, foaming, fogging, coating, and encrusting. Other suitable application procedures can be envisioned by those skilled in the art.

[0163] The modulator compounds of the present invention can be applied to the first and/or second location in accordance with the present invention alone or in a mixture with other materials. Alternatively, the modulator compounds of the present invention can be applied separately with other materials being applied at different times. Suitable other materials include effective amounts of other agricultural or horticultural chemicals, such as herbicides (*e.g.*, glyphosate), insecticides (when used against harmful insects), nematicides (when used against undesirable nematodes), molluscicides, bactericides, acaricides, fungicides, and/or plant growth regulators.

[0164] In at least one embodiment, contacting includes direct application to a plant. All or part of a plant including, without limitation, leaves, stems, roots, propagules (*e.g.*, cuttings), fruit, etc., may be contacted with the one or more modulator compounds. Contacting may also be carried out indirectly, via application, *e.g.*, to soil or other plant substrates. Applying the one or more modulator compounds to a plant may be carried out at a rate of about 0.1 to 10,000 g/ha of modulator compound. Preferably, application to a plant is carried out at a rate of about 10 to 1,000 g/ha of modulator compound.

[0165] A fourth aspect of the present invention relates to a method of treating or preventing parasite infection of a plant. This method involves: (i) contacting the plant with one or more isolated modulator compounds under conditions effective to treat or prevent parasite infection of the plant, or (ii) contacting a second location with one or more isolated modulator compounds under conditions effective to treat or prevent parasite infection of the plant, where the plant and the second location are spaced to permit said contacting the second location to have an effect on parasite infection of the plant. In this aspect of the present

invention, the parasite is a nematode or an insect and the one or more isolated modulator compounds is selected from the group consisting of ascr#1; ascr#2; ascr#3; ascr#7; ascr#8; ascr#9; ascr#10; ascr#11; ascr#16; ascr#18; ascr#20; ascr#22; icas#9; oscr#10; a combination of ascr#9 and ascr#10; a combination of ascr#9 and ascr#16; a combination of ascr#9 and ascr#18; a combination of ascr#9 and ascr#20; a combination of ascr#9 and ascr#22; a combination of ascr#2, ascr#3, ascr#8, and icas#9; a combination of ascr#9, ascr#3, ascr#8, and icas#9; a combination of ascr#9 and oscr#10; and a combination of ascr#9, oscr#10, and ascr#11.

[0166] Treating and/or preventing parasite infection according to this aspect of the present invention includes reducing the degree of infection already present in the plant as well as preventing infection from occurring in the first place. When the parasite is a nematode, treating and/or preventing includes the use of one or more modulator compounds that affect the nematode parasite itself, and/or using one more modulator compounds that affect nematodes that are harmful to the nematode parasite. When the parasite is an insect, treating and/or preventing includes the use of one or more modulator compounds that affect nematodes that are harmful to the insect parasite.

[0167] As will be apparent to the skilled artisan, various embodiments noted above for contacting the first and/or second location with one or more modulator compounds are also relevant here, the first location being the subject plant in this aspect of the present invention.

[0168] In at least one embodiment of this aspect of the present invention, the parasite is a nematode. Suitable nematode parasites according to this aspect of the present invention include *Meloidogyne javanica*, *Meloidogyne floridensis*, *Meloidogyne incognita*, and *Meloidogyne hapla*. Suitable nematodes that are harmful to nematode parasites include *Steinernema riobrave*, *Steinernema diaprepesi*, *Steinernema carpocapse*, *Steinernema feltiae*, *Steinernema glaseri*, *Heterorhabditis bacteriophora*, *Heterorhabditis zealandica*, and *Heterorhabditis floridensis*.

[0169] In at least one embodiment of this aspect of the present invention, the parasite is an insect. Suitable insects according to this aspect of the present invention include the plant-parasitic insects identified above. Suitable entomopathogenic nematodes according to this aspect of the present invention include *Steinernema riobrave*, *Steinernema diaprepesi*, *Steinernema carpocapse*, *Steinernema feltiae*, *Steinernema glaseri*, *Heterorhabditis bacteriophora*, *Heterorhabditis zealandica*, and *Heterorhabditis floridensis*.

[0170] Suitable plants according to this aspect of the present invention include those identified above.

[0171] The present invention may be further illustrated by reference to the following examples.

EXAMPLES

[0172] The following Examples are intended to illustrate, but by no means are intended to limit, the scope of the present invention as set forth in the appended claims.

Example 1—Analytical Instrumentation and Procedures

[0173] Nuclear magnetic resonance (NMR) spectra were recorded on a Varian INOVA 600 NMR (600 MHz for ^1H , 151 MHz for ^{13}C). NMR-spectra were processed using Varian VNMR and MestreLabs MestReC software packages. Additional processing of bitmaps derived from NMR spectra was performed using Adobe Photoshop CS3 as described in Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety. High-performance liquid chromatography-mass spectrometry (HPLC–MS) was performed using an Agilent 1100 Series HPLC system equipped with a diode array detector and connected to a Quattro II spectrometer (Micromass/Waters). Data acquisition and processing were controlled by MassLynx software. Flash chromatography was performed using a Teledyne ISCO CombiFlash system.

Example 2—*C. elegans* Strains and General Culture Methods

[0174] All strains were maintained at 20°C, unless mentioned otherwise, on NGM agar plates, made with Bacto agar (BD Biosciences) and seeded with OP50 bacteria grown overnight. *C. elegans* variety N2 Bristol and males from the *him-5(e1490)* strain CB1490 were used for the attraction bioassays and the automated tracker experiments. The *him-5(e1490)* mutant segregates XO male progeny by X chromosome nondisjunction during meiosis (Hodgkin et al., *Genetics* 91:67-94 (1979), which is hereby incorporated by reference in its entirety). The transgenic strain PS6025 *qrIs2[sra-9::mCasp1]*, which expresses mammalian caspase in the ASK neuron under the influence of the *sra-9* promoter (gift of Tokumitsu Wakabayashi, Iwate University), was used for genetic ablation of the ASK neuron. Other strains used are as follows: CB4856, *C. elegans* Hawaii isolate (Hodgkin & Doniach, *Genetics* 146:149-64 (1997), which is hereby incorporated by reference in its entirety); RC301, *C. elegans* Freiburg isolate (de Bono & Bargmann, *Cell* 94:679-89 (1998); Hodgkin & Doniach, *Genetics* 146:149-64 (1997), which are hereby incorporated by

reference in their entirety); DA609 *npr-1(ad609)*; CX4148 *npr-1(ky13)* (de Bono & Bargmann, *Cell* 94:679-89 (1998), which is hereby incorporated by reference in its entirety); CX9740 *C. elegans* (N2); *kyEx2144 [ncs-1::GFP]* (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety); N2; *Ex(gcy-28::dp::mec-4D)* (Shinkai et al., *J. Neurosci.* 31:3007-15 (2011), which is hereby incorporated by reference in its entirety); CX10981 *kyEx2866 [“ASK::GCaMP2.2b” sra-9::GCaMP2.2b SL2 GFP, ofm-1::GFP]* (*ASK imaging line*); CX11073 *kyEx2916 [“AIA::GCaMP2.2b” T01A4.1::GCaMP2.2b SL2 GFP, ofm-1::GFP]* (*AIA imaging line*) (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety); DR476 *daf-22(m130)* (Golden & Riddle, *Mol. Gen. Genet.* 198:534-36 (1985), which is hereby incorporated by reference in its entirety); and *daf-22(ok693)* (Butcher et al., *PNAS* 106:1875-79 (2009), which is hereby incorporated by reference in its entirety).

Example 3—*C. elegans* Metabolite Naming

[0175] All newly identified ascarosides are named with four letter “SMID”s (SMID: Small Molecule IDentifiers), for instance, “icas#3” or “ascr#10.” The SMID database, which is hereby incorporated by reference in its entirety, is an electronic resource maintained by Frank Schroeder and Lukas Mueller at the Boyce Thompson Institute in collaboration with Paul Sternberg and WormBase. This database catalogues newly identified *C. elegans* small molecules from *C. elegans* and other nematodes, assigns a unique four-letter SMID (a searchable, gene-style Small Molecule IDentifier) to the small molecule identified, and includes a list of other names and abbreviations used in the literature for each compound.

Example 4—Preparation of Metabolite Extracts

[0176] Metabolite extracts were prepared according to a method previously described in Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety, which was modified as follows.

[0177] Worms (N2 or *daf-22*) from three 10 cm NGM plates were washed using M9-medium into a 100 mL S-medium pre-culture, where they were grown for 5 days at 22°C on a rotary shaker. Concentrated *E. coli* OP50 derived from 1 L of bacterial culture (grown for 16 hours in LB media) was added as food at days 1 and 3. Subsequently, the pre-culture was divided equally into four 1 L Erlenmeyer flask containing 400 mL of S-medium for a combined volume of 425 mL of S-medium, which was then grown for an additional 10-day period at 22°C on a rotary shaker. Concentrated OP50 derived from 1 L of bacterial culture

was added as food every day from days 1 to 9. Subsequently, the cultures were centrifuged and the supernatant media and worm pellet were lyophilized separately. The lyophilized materials were extracted with 95% ethanol (250 mL, 2 times) at room temperature for 12 hours. The resulting yellow suspensions were filtered. The filtrate was then evaporated *in vacuo* at room temperature, producing media and worm pellet metabolite extracts.

Example 5—Chromatographic Fractionation

[0178] The media metabolite extract from two cultures was adsorbed on 6 g of octadecyl-functionalized silica gel and dry-loaded into an empty 25 g RediSep *Rf* sample loading cartridge. The adsorbed material was then fractionated via a reversed-phase RediSep *Rf* GOLD 30 g HPLC18 column using a water-methanol solvent system. The solvent starts with 100% water for 4 minutes, followed by a linear increase of methanol content up to 100% methanol at 42 minutes, which was continued up until 55 minutes. The eight fractions generated from this fractionation were evaporated *in vacuo*. The residue was analyzed by HPLC-MS and two-dimensional nuclear magnetic resonance (2D-NMR) spectroscopy.

Example 6—Mass Spectrometric Analysis

[0179] Worm media extracts or metabolite fractions derived from the chromatographic fractionation were resuspended in 1.5 ml methanol and centrifuged at 2,000 *g* for 5 minutes. The supernatant was submitted to HPLC-MS analyses. HPLC was performed with an Agilent Eclipse XDB-C18 column (9.4 × 250 mm, 5 μm particle diameter). A 0.1% acetic acid–acetonitrile solvent gradient was used, starting with an acetonitrile content of 5% for 5 minutes, which was increased to 100% over a period of 40 minutes. Mass spectrometry was performed using electrospray ionization in either negative or positive ion mode.

Example 7—*C. elegans* Axenic Cultures and Biosynthetic Studies

[0180] Axenic *in vitro* cultures of *C. elegans* (N2, Bristol) were established as described by Nass & Hamza, *Curr. Protoc. Toxicol.* 31:1.9.1–1.9.18 (2007), which is hereby incorporated by reference in its entirety, using the *C. elegans* Maintenance Medium (CeMM, Lu & Goetsch, *Nematologica* 39:303-11 (1993), which is hereby incorporated by reference in its entirety) with cholesterol (5 mg/l) instead of sitosterol and nucleoside-5-phosphates. After 21 days, the cultures were centrifuged. The resulting supernatant media and worm pellet were lyophilized separately. The lyophilized worm pellets (1.2–2.0 mg) were extracted with 2 ml methanol, filtered, and then concentrated *in vacuo*. The lyophilized worm media were

extracted with ethyl acetate–methanol (95:5, 100 mL, two times), filtered, and then concentrated *in vacuo*. Residues were taken up in 150 µL methanol and investigated by high-performance liquid chromatography-electrospray ionization tandem-mass spectrometry (HPLC-ESI-MS). For the application experiment, 50 mL CeMM medium was supplemented with 9.2 mg L-[2,4,5,6,7-D₅]-tryptophan from Cambridge Isotope Laboratories.

Example 8—Spot Attraction Assays

[0181] The spot attraction assays were conducted according to a previously reported method (Pungaliya et al., *PNAS* 106:7708-13 (2009); Srinivasan et al., *Nature* 454:1115-18 (2008), which are hereby incorporated by reference in their entirety). For both *C. elegans* hermaphrodites and males, 50–60 worms were harvested daily at the fourth larval stage (L4) and stored segregated by sex at 20°C overnight, to be used as young adults the following day. For the competition experiments, two conditions were used: 120 nM ascr#3 and 10 nM icas#3 (Condition 1), or 12 µM ascr#3 and 1 µM icas#3 (Condition 2) in water containing 10% ethanol. Aliquots were stored at –20°C in 20 µL tubes. 10% ethanol in water was used as control.

Example 9—Quadrant Chemotaxis Assays

[0182] Chemotaxis to both non-indole and indole ascarosides was assessed on 10 cm four-quadrant petri plates (Wicks et al., *Dev. Biol.* 221:295-307 (2000), which is hereby incorporated by reference in its entirety). Each quadrant was separated from adjacent ones by plastic spacers (Figure 1B). Pairs of opposite quadrants were filled with nematode growth medium (NGM) agar containing either indole ascarosides or non-indole ascarosides at different concentrations. Animals were washed gently in a S-basal buffer, placed in the center of a four-quadrant plate with ascarosides in alternating quadrants, and scored after 15 minutes and 30 minutes. A chemotaxis index was calculated as: (the number of animals on ascaroside quadrants minus the number of animals on buffer quadrants) / (total number of animals).

Example 10—Measurement of Locomotory Parameters

[0183] Reversal frequency and velocity were measured using an automated worm-tracking system (Pungaliya et al., *PNAS* 106:7708-13 (2009); Srinivasan et al., *Nature* 454:1115-18 (2008), which are hereby incorporated by reference in their entirety).

Example 11—Aggregation Assays

[0184] Aggregation behavior of worms was measured using assays reported in de Bono & Bargmann, *Cell* 94:679-89 (1998), which is hereby incorporated by reference in its

entirety. Aggregation assays were conducted on standard NGM plates. Plates containing indole ascarosides were made by adding the indole ascaroside stock solution to the NGM media before they were poured onto the plates. These plates were dried at room temperature for 2 to 3 days. Control plates were treated similarly, except that ethanol solutions instead of icas solutions were added to the plates, corresponding to the amount of ethanol introduced via the icas solutions. Final ethanol concentrations of the plates were below 0.1% for all conditions. After drying, both control plates and plates containing indole ascarosides were seeded with 150 μ l of an overnight culture of *E. coli* OP50 using a micropipette, and were allowed to dry for 2 days at room temperature.

[0185] For “low worm density” experiments, 20 worms were placed onto the lawn and left at 20°C for 3 hours. For “high worm density” experiments, approximately 120 worms were placed onto the bacterial lawn and left at 20°C for 3 hours. Aggregation behavior was quantified as the number of animals that were in touch with two or more animals at greater than 50% of their body length.

Example 12—Calcium Imaging and Analysis

[0186] Transgenic lines that express the genetically encoded Ca^{2+} sensor in ASK (*kyEx2866*) and AIA (*kyEx2916*) (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety) were used for calcium imaging. Young adults or adult worms were inserted into an “Olfactory chip” microfluidic device (Chronis et al., *Nat. Methods* 4:727-31 (2007), which is hereby incorporated by reference in its entirety). Dilutions of icas#3 were done with S-basal buffer (with no cholesterol). As stock solutions of icas#3 contained small amounts of ethanol, equivalent amounts of ethanol were added to the S-basal control flow.

[0187] Imaging was conducted using an inverted Zeiss microscope equipped with an Andor camera. Exposure time for image acquisition was 300 milliseconds. Before imaging the ASK neuron, the worm was exposed to blue light for 3 minutes as ASK responds to the blue light itself. This step is necessary for the neuron to adapt to the blue light that is used for Ca^{2+} measurements. Movies were analyzed using custom-made Matlab scripts. The first peak of fluorescence immediately after exposure to buffer or icas#3 was chosen for calculating the average change in fluorescence upon exposure to either buffer or icas#3. The value for this maximum was then subtracted from the mean fluorescence during the 5 seconds before the delivery of icas#3/buffer (corresponding to the region between 5 to 10 seconds in Figure 2D).

Example 13—Statistical Analysis

[0188] Unpaired Student's *t* tests with Welch's correction was used: (i) for comparing attraction of hermaphrodites and males on indole ascarosides (*: $p < 0.01$, **: $p < 0.001$, ***: $p < 0.0001$) (Figures 1C–1D, 3A, 3D, 4A, 5A, and 6C); (ii) to compare reversals between unablated and ASK ablated lines (*: $p < 0.01$, **: $p < 0.001$) (Figures 7A–B); (iii) to compare the attraction of wild-type worms to the genetically ablated lines for ASK and AIA as well as the ASI and RMG neuron ablations (***: $p < 0.0001$) (Figure 2B); and (iv) for comparing G-CaMP fluorescence changes to buffer and icas#3 (**: $p < 0.001$) (Figure 2E). One-factor ANOVA followed by Dunnett's post-test was used: (i) for comparing the quadrant chemotaxis indices of the various strains (*: $p < 0.05$, **: $p < 0.01$) (Figures 1E and 3B–C); (ii) for comparing aggregation of solitary, social worms and *Cel-daf-22* on plates containing indole ascarosides (*: $p < 0.05$, **: $p < 0.01$) (Figures 5C, 6A–B, and 8A–C); (iii) to compare stopped duration of worms on plates with indole ascarosides (*: $p < 0.05$, **: $p < 0.01$) (Figure 8D); and (iv) to compare velocities and reversal frequencies on plates with indole ascarosides (*: $p < 0.05$, **: $p < 0.01$) (Figures 6D–E). All error bars indicate standard error of mean (S.E.M).

Example 14—Calculation of Number of Molecules of icas#3 in One L4 Worm Volume at 100 fM

[0189] The number of molecules (*n*) of icas#3 in one L4 worm volume at 100 fM (femtomolar) was calculated using equation (1):

$$n = V_{YA} * c * N_A \quad (1)$$

where:

V_{YA} = volume of *C. elegans* young adult (Knight et al., *Evol. Dev.* 4:16-27 (2002), which is hereby incorporated by reference in its entirety)

c = concentration (100 fM)

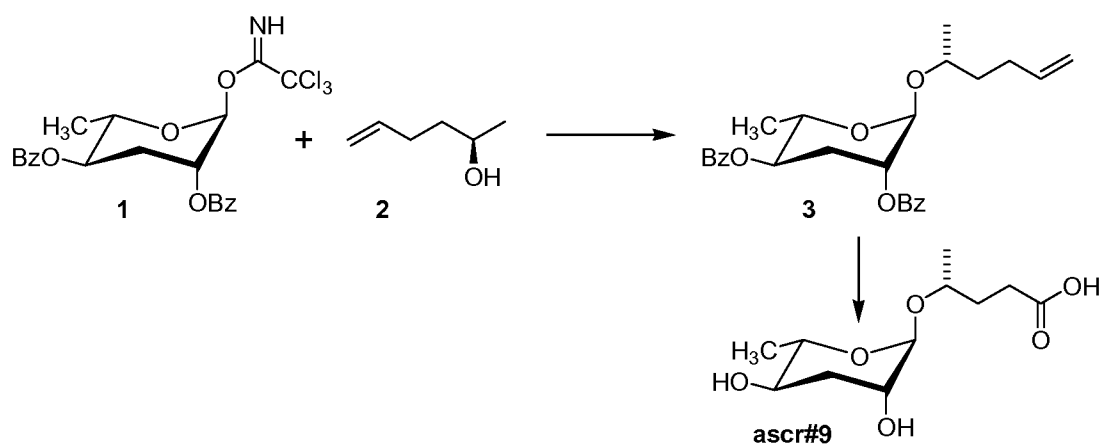
N_A = Avogadro's number

[0190] In this case, $V_{YA} = 3 * 10^6 \mu m^3 = 3 * 10^{-9} L$ and $c = 10 fM = 10^{-14} M$. Solving for *n*, $n = V_{YA} * c * N_A = 3 * 10^{-9} L * 10^{-14} mol * L^{-1} * 6.022 * 10^{23} mol^{-1} = 18$ molecules.

Therefore, there are 18 icas#3 molecules contained in one worm volume of agar at an icas#3 concentration of 10 fM.

Example 15—Chemical Syntheses

[0191] Samples of indole ascarosides icas#1, icas#7, icas#3, and icas#9 for use in biological assays and as standards for HPLC-MS were prepared via chemical synthesis. Detailed synthetic procedures and NMR-spectroscopic data are set forth in Examples 16–21.

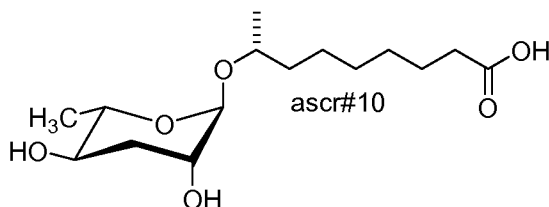
Example 16—Synthesis of Ascaroside ascr#9

[0192] To prepare ascr#9, (*R*)-hex-5-en-2-ol **2** (32 mg, 0.32 mmol) (prepared as described in Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety, using the conditions described for the synthesis of ascr#6) was coupled to trichloroacetimide **1** (150 mg, 0.3 mmol, Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety). The resulting glycoside **3** (92 mg, 0.21 mmol) was dissolved in acetone (2 ml) and treated with 2 ml of a 1 M solution of potassium permanganate. After 30 minutes, the reaction mixture was poured into a mixture of ice-cold saturated aqueous sodium chloride solution (5 ml), acetic acid (0.1 ml), and dichloromethane (DCM) (5 ml). The organic phase was separated and the aqueous phase was extracted with two 5 ml-portions of DCM. The combined organic extracts were dried over sodium sulfate, evaporated to dryness, and re-dissolved in a mixture of 0.5 M aqueous lithium hydroxide (2 ml) and dioxane (6 ml). The mixture was stirred for 3 hours at 70 °C, then cooled to 23 °C and acidified with 0.2 M aqueous hydrochloric acid. The mixture was evaporated to dryness and purified via Combiflash column chromatography using a methanol–DCM solvent gradient, yielding 15.6 mg (0.063 mmol) of pure ascr#9 as a viscous oil.

[0193] ^1H (600 MHz) and ^{13}C (126 MHz) NMR spectroscopic data for ascr#9 were acquired in methanol- d_4 . Chemical shifts are referenced to $(\text{CD}_2\text{HOD}) = 3.31$ ppm and $(\text{CD}_2\text{HOD}) = 49.05$ ppm. Coupling constants are given in Hertz [Hz]. ^1H NMR (600 MHz,

CD₃OD): δ 4.65 (s, 1H), 3.84 (m, 1H), 3.72 (m, 1H), 3.61 (dq, 1H, $J = 9.4$ Hz, $J = 6.1$ Hz), 3.51 (ddd, 1H, $J = 11.2$ Hz, $J = 9.5$ Hz, $J = 4.5$ Hz), 2.43 (m, 2H), 1.95 (dt, 1H, $J = 13.1$ Hz, $J = 3.5$ Hz), 1.71-1.87 (m, 3H), 1.22 (d, 3H, $J = 6.1$ Hz), 1.15 (d, 3H, $J = 6.1$ Hz) ppm; ¹³C NMR (126 MHz, CD₃OD): δ 174.5, 97.3; 71.5, 71.4, 69.9, 68.4, 36.0, 33.5, 31.3, 19.1, 18.1 ppm; ESI-MS (m/z): [M-H] 247.2.

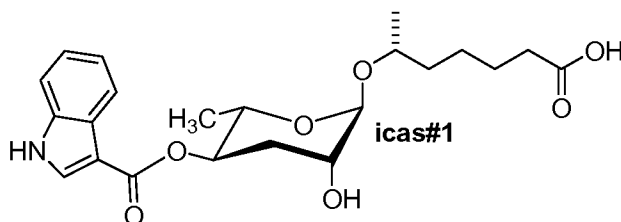
Example 17—Synthesis of Ascaroside ascr#10



[0194] To prepare ascr#10, a stirred solution of ascr#3 (3.2 mg, 10.6 μ mol, Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety) in 10 ml of ethanol was hydrogenated using palladium on activated carbon (10% Pd, 1 atmosphere H₂ pressure) at 23 °C for 18 hours. After completion, the reaction was evaporated to dryness. The residue was filtered over a short pad of silica using a 1:8 (v/v) mixture of methanol and DCM, yielding 3.0 mg (9.9 μ mol) of pure ascr#10.

[0195] ¹H (600 MHz) and ¹³C (126 MHz) NMR spectroscopic data for ascr#10 were acquired in methanol-*d*₄. Chemical shifts were referenced to (CD₂HOD) = 3.31 ppm and (CD₂HOD) = 49.05 ppm. Coupling constants were given in Hertz [Hz]. ¹H NMR (600 MHz, CD₃OD): δ 4.64 (s, 1H), 3.78 (m, 1H), 3.71 (m, 1H), 3.63 (dq, 1H, $J = 9.3$ Hz, $J = 6.2$ Hz), 3.51 (ddd, 1H, $J = 11.2$ Hz, $J = 9.3$ Hz, $J = 4.6$ Hz), 2.27 (t, 2H, $J = 7.4$ Hz), 1.94 (dt, 1H, $J = 13.0$ Hz, $J = 3.7$ Hz), 1.77 (ddd, 1H, $J = 13.3$ Hz, $J = 11.5$ Hz, $J = 3.0$ Hz), 1.61 (m, 2H), 1.56 (m, 1H), 1.46 (m, 1H), 1.32-1.38 (m, 6H), 1.21 (d, 3H, $J = 6.2$ Hz), 1.12 (d, 3H, $J = 6.1$ Hz) ppm; ¹³C NMR (126 MHz, CD₃OD): δ 177.7, 97.3, 72.3, 71.0, 69.8, 68.1, 38.1, 38.2, 35.7, 34.9, 30.0, 26.5, 26.0, 19.0, 18.0 ppm; ESI-MS (m/z): [M-H] 303.2.

Example 18—Synthesis of Indole Carboxy Ascaroside icas#1



[0196] To prepare icas#1, ascr#1 was first converted into the corresponding methyl ester. Ascr#1 (10 mg, 0.036 mmol), prepared as reported in Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety, was dissolved in a mixture of toluene (1 mL) and methanol (1 mL). To this mixture, a solution of trimethylsilyldiazomethane (TMS-diazomethane) (2 M solution in hexane, 50 μ L, 0.1 mmol) was added. After stirring for 20 minutes at 23 °C, excess TMS-diazomethane was destroyed by addition of acetic acid (40 μ L) and the solvents were removed *in vacuo*, yielding the methyl ester of ascr#1 (10.3 mg, 0.035 mmol) as a viscous oil.

[0197] Next, a solution of indole-3-carbonyl chloride was prepared. A well-stirred suspension of indole-3-carboxylic acid (67.7 mg, 0.42 mmol) in dry DCM (2 ml) containing a small amount of dimethylformamide (DMF) (20 μ L) was treated with oxalyl chloride (0.84 mmol, 107 mg, 72 μ L) at 0 °C. Following addition to the oxalyl chloride, the mixture was stirred for 20 minutes at 23 °C, producing a clear, slightly yellow solution. This solution was evaporated to dryness *in vacuo* at 0.1 Torr to ensure removal of excess oxalyl chloride, and was subsequently re-dissolved in 2 ml of dry DCM.

[0198] The sample of ascr#1 methyl ester (10.3 mg, 0.035 mmol) was dissolved in 1 ml of dry DCM, to which diisopropylethylamine was added (129 mg, 1 mmol). The resulting solution was equipped with an effective stir bar and cooled to -20 °C. Subsequently, the solution of indole-3-carboxylic acid chloride was added drop-wise over a period of 10 minutes with vigorous stirring. The well-stirred reaction was gradually warmed to -7 °C, at which temperature ice cold saturated aqueous sodium bicarbonate solution (2 ml) was added. The biphasic mixture was allowed to warm to 20 °C, and was extracted three times with ethyl acetate. The combined ethyl acetate extracts were evaporated *in vacuo* and subjected to column chromatography on silica gel using 0–10% methanol in DCM.

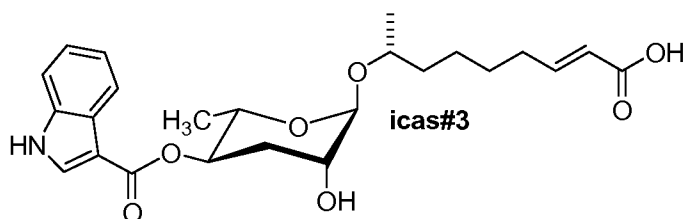
[0199] Fractions containing the bis-2,4-*O*-(indole-3-carbonyl)-derivative of the ascr#1 methyl ester were combined, evaporated to dryness, and treated with a mixture of 3 ml aqueous 0.5 M lithium hydroxide solution and 7 ml dioxane at 67 °C for 3 hours. Subsequently, the reaction mixture was cooled to 23 °C, neutralized by addition of 0.2 M aqueous hydrochloric acid, and evaporated *in vacuo*. The residue was purified by HPLC with an Agilent Eclipse XDB C-18 column (25 cm $\times$ 9.4 mm, 5 μ m particle diameter). Acetonitrile and 0.1% aqueous acetic acid were used as solvents, increasing the percentage of acetonitrile from 15% at 0 minute to 85% at 30 minutes. Icas#1-containing fractions were evaporated, yielding 5.8 mg (0.014 mmol) of the target compound as a wax-like white solid.

[0200] ^1H (600 MHz), ^{13}C (126 MHz), and HMBC NMR spectroscopic data for icas#1 were obtained using methanol- d_4 and are shown in Table 1 below. Chemical shifts are referenced to (CD_2HOD) = 3.31 ppm and (CD_2HOD) = 49.05 ppm. Coupling constants are given in Hertz [Hz]; *: interchangeable.

Table 1. NMR spectroscopic data of icas#1

Position	^{13}C [ppm]	^1H [ppm]	^1H - ^1H -coupling constants
1	177.6		
2	35.1	2.35	$J_{2,3} = 7.4$
3	26.6*	1.45-1.70	
4	26.2*	1.45-1.70	
5	38.1	1.45-1.70	
6	72.7	3.86	
7	19.4	1.17	$J_{6,7} = 6.1$
1'	97.7	4.75	
2'	69.6	3.79	
3'	33.5	2.01 (ax)	$J_{3'\text{ax}, 3'\text{eq}} = 13.0$, $J_{3'\text{ax}, 4'} = 11.4$, $J_{2', 3'\text{ax}} = 2.9$
		2.21 (eq)	$J_{2', 3'\text{eq}} = 3.2$, $J_{3'\text{eq}, 4'} = 4.7$
4'	70.6	5.12	$J_{4', 5'} = 9.6$
5'	68.8	4.05	$J_{5', 6'} = 6.3$
6'	18.3	1.24	
2''	133.5	7.97	
3''	108.4		
3''-COO	166.5		
3a''	127.3		
4''	121.9	8.02	$J_{4'', 5''} = 7.2$
5''	122.7	7.16	
6''	123.8	7.29	
7''	113.1	7.44	$J_{6'', 7''} = 7.9$
7a''	138.2		

Example 19—Synthesis of Indole Carboxy Ascaroside icas#3



[0201] To prepare icas#3, ascr#3 was first converted into the corresponding methyl ester. Ascr#3 (5.2 mg, 0.017 mmol), prepared as reported in Pungaliya et al., *PNAS*

106:7708-13 (2009), which is hereby incorporated by reference in its entirety, was dissolved in a mixture of toluene (1 mL) and methanol (1 mL). To this mixture, a solution of TMS-diazomethane (2 M solution in hexane, 25 μ L, 0.05 mmol) was added. After stirring for 20 minutes at 23 °C, excess TMS-diazomethane was destroyed by addition of acetic acid (30 μ L) and the solvents were removed *in vacuo*, yielding the methyl ester of ascr#3 (5.3 mg, 0.017 mmol) as a viscous oil.

[0202] The sample of ascr#3 methyl ester (10.3 mg, 0.035 mmol) was reacted with indole carbonyl chloride as described in Example 18 for the preparation of icas#1, using proportionally smaller amounts of all reagents. Purification of the crude reaction products via HPLC using the conditions described in Example 18 yielded icas#3 (2.3 mg, 5.2 μ mol) as a colorless oil.

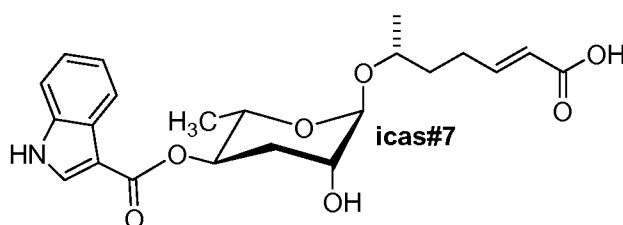
[0203] ^1H (600 MHz), ^{13}C (126 MHz), and HMBC NMR spectroscopic data for icas#3 were obtained using methanol- d_4 and are shown in Table 2 below. Chemical shifts are referenced to (CD_2HOD) = 3.31 ppm and (CD_2HOD) = 49.05 ppm. Coupling constants are given in Hertz [Hz].

Table 2. NMR spectroscopic data of icas#3

Position	^{13}C [ppm]	^1H [ppm]	^1H - ^1H -coupling constants	Relevant HMBC correlations
1	173.7			
2	126.7	5.82	$J_{2,3} = 15.1$	
3	147.8	6.79	$J_{3,4} = 6.9$	C-1, C-4, C-5
4	33.6	2.23		C-2, C-3, C5, C-6
5	29.9	1.54		C-4, C-6
6	27.2	1.51		
7	38.7	1.53		C-5, C-6, C-8
8	73.1	3.83		C-6, C-7, C-9
9	19.9	1.15	$J_{8,9} = 6.1$	C-7, C-8
1'	98.0	4.73		C-3', C-5', C-8
2'	70.1	3.77		C-4'
3'	33.8	1.98 (ax)	$J_{3'\text{ax},3'\text{eq}} = 13.0$, $J_{3'\text{ax},4'} = 11.4$, $J_{2',3'\text{ax}} = 2.9$	
		2.19 (eq)	$J_{2',3'\text{eq}} = 3.2$, $J_{3'\text{eq},4'} = 4.7$	C-4'
4'	70.9	5.09	$J_{4',5'} = 9.6$	C-6'
5'	69.1	4.02	$J_{5',6'} = 6.3$	C-4'
6'	18.8	1.21		C4', C-5'
2''	133.9	7.94		C-3'', C-3a'', C-7a''
3''	108.8			
3''-COO	167.0			

3a''	127.9			
4''	122.3	7.99	$J_{4'',5''} = 7.4$	C-6'', C-7a'', 3''-COO
5''	123.0	7.15		C-3a'', C-7''
6''	124.2	7.18		C-4'', C-7a''
7''	113.5	7.42	$J_{6'',7''} = 7.9$	C-3a'', C-5''
7a''	138.8			

Example 20—Synthesis of Indole Carboxy Ascaroside icas#7



[0204] A standard sample of icas#7 (120 μ g) was obtained from ascr#7 (0.5 mg, Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety) as described in Example 18 for the preparation of icas#1 from ascr#1.

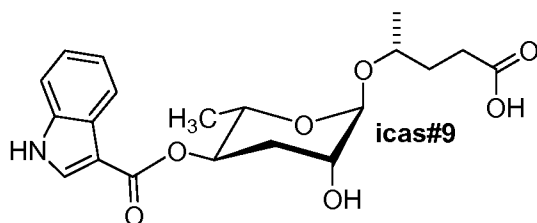
[0205] ^1H (600 MHz) NMR spectroscopic data for icas#7 were obtained using methanol- d_4 and are shown in Table 3 below. Chemical shifts are referenced to (CD_2HOD) = 3.31 ppm. Coupling constants are given in Hertz [Hz].

Table 3. NMR spectroscopic data of icas#7

Position	^1H [ppm]	^1H - ^1H -coupling constants
2	5.84	$J_{2,3} = 15.3$
3	6.99	$J_{3,4} = 6.8$
4	2.40	
	2.33	
5	1.70	
	1.65	
6	3.91	
7	1.18	$J_{6,7} = 6.1$
1'	4.75	
2'	3.80	
3'	2.03 (ax)	$J_{3'\text{ax},3'\text{eq}} = 13.0$, $J_{3'\text{ax},4'} = 11.4$, $J_{2',3'\text{ax}} = 2.9$
	2.21 (eq)	$J_{2',3'\text{eq}} = 3.2$, $J_{3'\text{eq},4'} = 4.7$
4'	5.12	$J_{4',5'} = 9.6$
5'	4.07	$J_{5',6'} = 6.3$
6'	1.24	
2''	7.97	
4''	8.04	$J_{4'',5''} = 7.5$

5"	7.15	
6"	7.16	
7"	7.43	$J_{6'',7''} = 7.9$

Example 21—Synthesis of Indole Carboxy Ascaroside icas#9



[0206] Icas#9 was obtained from ascr#9 as described in Example 18 for the preparation of icas#1 from ascr#1. NMR-spectroscopic data are in agreement with published data (Butcher et al., *Org. Lett.* 11:3100-03 (2009), which is hereby incorporated by reference in its entirety).

Example 22—Identification of the Indole Ascaroside icas#3 Via Comparative Metabolomics

[0207] All currently known small-molecule pheromones in *C. elegans* are derived from peroxisomal β -oxidation of long-chained fatty acids via DAF-22, a protein with strong homology to human sterol carrier protein SCPx (Pungaliya et al., *PNAS* 106:7708-13 (2009); Butcher et al., *PNAS* 106:1875-79 (2009), which are hereby incorporated by reference in their entirety). It was suspected that putative aggregation pheromones may be derived from the same pathway, suggesting that *daf-22* mutants would not produce them. In that case, a spectroscopic comparison of the wild-type metabolome with that obtained from *daf-22* mutant worms should reveal candidate compounds for attraction or aggregation signals.

[0208] In a previous study, an NMR spectroscopy-based technique termed Differential Analysis of NMR spectra ("DANS") was used to compare the wild-type metabolome with that of *daf-22* mutant worms (Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety). This comparison had led to the identification of ascr#6–8, of which ascr#8 is a major component of the male-attracting signal (Pungaliya et al., *PNAS* 106:7708-13 (2009), which is hereby incorporated by reference in its entirety). Based on NMR spectra with improved signal-to-noise ratio, a more detailed comparison of wild type and *daf-22*-mutant metabolomes was conducted, revealing several indole-containing compounds in the wild-type metabolome that were not produced by *daf-22* worms (Figures 9B–C). The established role of DAF-22 in pheromone biosynthesis

(Pungaliya et al., *PNAS* 106:7708-13 (2009); Butcher et al., *PNAS* 106:1875-79 (2009); Golden & Riddle, *Mol. Gen. Genet.* 198:534-36 (1985), which are hereby incorporated by reference in their entirety) suggests that these indole derivatives may represent a previously unrecognized family of signaling molecules.

[0209] To clarify the structures and biological roles of the *daf-22*-dependent indole derivatives, complete identification of these indole derivatives was pursued via NMR spectroscopy-guided fractionation of the wild-type metabolome. Reverse-phase chromatography produced eight metabolite fractions, which were analyzed by two-dimensional NMR spectroscopy. The NMR spectra revealed the presence of *daf-22*-dependent indole derivatives in two fractions, which were selected for additional NMR-spectroscopic and mass spectrometric studies. These analyses indicate that the most abundant *daf-22*-dependent indole derivative consists of an indole-carboxy unit linked to ascarlose bearing a 9-carbon unsaturated side-chain identical to that found in the known *icas#3* (Srinivasan et al., *Nature* 454:1115-18 (2008), which is hereby incorporated by reference in its entirety) (see NMR and MS data in Examples 18–19, Figures 10–13). Based on its structural relationship to the known *icas#3*, this newly identified metabolite was named indole carboxy ascaroside “*icas#3*” (Figure 9E).

Example 23—Icas#3 Is Part of a Larger Family of Tryptophan-Derived Small Molecules

[0210] Some other *daf-22*-dependent indole compounds detected by DANS were investigated to determine whether they also represent indole ascarosides. A mass spectrometric (MS) approach was employed for this investigation, as analysis of the mass spectra of *icas#3* had revealed a characteristic MS fragmentation pattern (loss of the indole-3-carboxy moiety, Figure 12) that enabled a screen for related compounds.

[0211] MS screening for compounds with similar fragmentation profiles indicates that *icas#3* is a member of a larger series of indole ascarosides featuring side chains with five to nine carbons (Figures 9D–E). The most abundant components of this family of indole ascarosides are *icas#3*, *icas#9*, and *icas#10*, which are accompanied by smaller amounts of *icas#1* and *icas#7* (Figure 9E). All of these compounds represent new metabolites, except for *icas#9*, which has been reported to possess moderate dauer-inducing activity and is unique among known dauer pheromones in producing a bell-shaped response curve (Butcher et al., *Org. Lett.* 11:3100-03 (2009), which is hereby incorporated by reference in its entirety).

[0212] Two new non-indole ascarosides were also detected: ascr#9, which features a saturated 5-carbon side chain, and ascr#10, which features a saturated 9-carbon side chain, representing the saturated analog of the known ascr#3 (Figure 9F).

[0213] The MS analyses further revealed that the indole ascarosides' quantitative distribution is distinctly different from that of the corresponding non-indole ascarosides, suggesting that incorporation of the indole unit is strongly regulated. The most abundant indole ascaroside, icas#3, is accompanied by 10- to 40-fold larger amounts of the corresponding non-indole ascaroside, ascr#3; whereas icas#9 is more abundant than the corresponding ascr#9 (Figure 9G).

[0214] To determine the biosynthetic origin of the indole ascarosides and to exclude the possibility that they are produced by the *E. coli* food source, axenic (bacteria-free) *in vitro* cultures of *C. elegans* (N2) were established using the chemically defined CeMM medium (Lu & Goetsch, *Nematologica* 39:303-11 (1993); Nass & Hamza, *Curr. Protoc. Toxicol.* 31:1.9.1-1.9.18 (2007), which are hereby incorporated by reference in their entirety). HPLC-MS analysis of the axenic cultures revealed the presence of icas#1, icas#3, icas#9, and icas#10, thus indicating that indole ascarosides are produced by *C. elegans* without participation of dietary bacteria. Use of a 1:1 mixture of L-[2,4,5,6,7-D₅]-tryptophan and L-tryptophan in the axenic medium resulted in production of [D₅]-icas#1, [D₅]-icas#3, [D₅]-icas#9, and [D₅]-icas#10, along with equivalent amounts of the unlabelled compounds (Figure 13). In conclusion, these biochemical studies established the tryptophan origin of the indole-3-carboxy moiety in the indole ascarosides and indicate that these compounds are products of a strongly regulated biosynthetic pathway.

Example 24—Indole Ascarosides Strongly Attract Hermaphrodites

[0215] The addition of an indole-3-carboxy moiety to the ascarosides represents a significant structural change. This chemical difference may indicate signaling functions for these compounds distinct from those of their non-indole cognates.

[0216] Three synthetic indole ascarosides of varying side-chain lengths, icas#1, icas#3, and icas#9, were tested in the spot attraction assay to demonstrate social functions of small molecules (Pungaliya et al., *PNAS* 106:7708-13 (2009); Srinivasan et al., *Nature* 454:1115-18 (2008), which are hereby incorporated by reference in their entirety) (Figure 1A). All three tested indole ascarosides, icas#1, icas#3, and icas#9, attracted both males and hermaphrodites at high concentrations (Figure 1C). Testing the most abundant indole ascaroside, icas#3, over a broader range of concentrations revealed that, at low

concentrations, icas#3 was strongly attractive to hermaphrodites, whereas males were no longer attracted (Figure 1D). Similarly, hermaphrodites, but not males, were strongly attracted to low concentrations of icas#9 (Figure 5A).

[0217] Hermaphrodite attraction to icas#3 was further investigated using a quadrant chemotaxis bioassay as reported in Macosko et al., *Nature* 458:1171-75 (2009); Wicks et al., *Dev. Biol.* 221:295-307 (2000), which are hereby incorporated by reference in their entirety (Figure 1B). In contrast to the spot attraction assay, which measures attraction to a point source of compounds, the quadrant chemotaxis assay measures aggregation of hermaphrodites on plate sections with well-defined compound concentration (Macosko et al., *Nature* 458:1171-75 (2009); Wicks et al., *Dev. Biol.* 221:295-307 (2000), which are hereby incorporated by reference in their entirety). It was found that concentrations as low as 1 pM icas#3 resulted in a strong attraction of hermaphrodites (Figure 1E), both in the presence and absence of food (Figure 5B).

[0218] The biological role of icas#3 thus starkly differs from that of the corresponding non-indole ascaroside ascr#3, which strongly attracts males but repels hermaphrodites (Pungaliya et al., *PNAS* 106:7708-13 (2009); Srinivasan et al., *Nature* 454:1115-18 (2008), which are hereby incorporated by reference in their entirety). The results herein show that by simply attaching an indole-3-carboxy group to the 4-position of the ascarylose, the strongly male-attracting ascr#3 is converted into a signal that primarily attracts hermaphrodites. The difference in the amounts at which ascr#3 and icas#3 are produced by the worms corresponds to their relative potency: the male-attracting ascr#3, which is of much lower potency than icas#3, is produced in much higher concentrations than the highly potent hermaphrodite attractant icas#3 (Figure 9G).

Example 25—Solitary and Social Wild-Type Hermaphrodites Are Attracted to icas#3, but not to Non-Indole Ascarosides

[0219] The results from the spot attraction and quadrant chemotaxis assays indicate that hermaphrodites are strongly attracted to indole ascarosides, suggesting that these compounds regulate *C. elegans* aggregation behavior. *C. elegans* exhibits natural variation in its foraging behavior, with some strains (e.g., the common laboratory strain N2) dispersing individually on a bacterial lawn, whereas most wild-type strains (e.g., RC301 and CB4856 (Hawaii)) accumulate and aggregate where bacteria are the most abundant (de Bono & Bargmann, *Cell* 94:679-89 (1998); Hodgkin & Doniach, *Genetics* 146:149-64 (1997), which are hereby incorporated by reference in their entirety). These variants are referred to as

“solitary” and “social,” respectively (de Bono & Bargmann, *Cell* 94:679-89 (1998); Rogers et al., *Nat. Neurosci.* 6:1178-85 (2003), which are hereby incorporated by reference in their entirety). These differences in foraging and aggregation behavior are associated with two different alleles of the neuropeptide Y-like receptor NPR-1 (de Bono & Bargmann, *Cell* 94:679-89 (1998); Rogers et al., *Nat. Neurosci.* 6:1178-85 (2003), which are hereby incorporated by reference in their entirety), which differ at a single amino acid position: solitary strains such as N2 express a high-activity variant of NPR-1 (215-valine), whereas aggregating strains such as CB4856 express a low-activity variant of NPR-1 (215-phenylalanine) (de Bono & Bargmann, *Cell* 94:679-89 (1998); Rogers et al., *Nat. Neurosci.* 6:1178-85 (2003), which are hereby incorporated by reference in their entirety). The strong loss-of-function mutants *npr-1(ad609)* and *npr-1(kyl3)*, which were generated in the N2 background, also show a high tendency to aggregate (de Bono & Bargmann, *Cell* 94:679-89 (1998); Hodgkin & Doniach, *Genetics* 146:149-64 (1997), which are hereby incorporated by reference in their entirety).

[0220] A previous study showed that loss of function of *npr-1* affects the hermaphrodite’s response to non-indole ascarosides (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety). While wild-type (N2) worms expressing the high-activity variant of NPR-1 are repulsed by non-indole ascarosides, *npr-1(ad609)* mutants showed attraction to a near-physiological mixture of the most abundant non-indole ascarosides, ascr#2, ascr#3, and ascr#5 (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety).

[0221] Both the quadrant chemotaxis and spot attraction assays confirmed the attraction of *npr-1(ad609)* hermaphrodites to ascr#2/3/5 mixtures; however, hermaphrodites of the two tested social wild-type strains (RC301 and CB4856) showed no attraction in either assay (Figures 3A–B). In contrast, both social wild-type strains (RC301 and CB4856) as well as *npr-1(ad609)* hermaphrodites were strongly attracted to icas#3, in both the quadrant chemotaxis and spot-attraction assays (Figures 3B–D and 5B–C). These results indicate that icas#3 functions as a hermaphrodite attractant in both solitary and social *C. elegans* strains.

Example 26—Femtomolar Concentrations of Indole Ascarosides Promote *C. elegans* Aggregation

[0222] How a constant background concentration of indole ascarosides affects hermaphrodite behavior was tested. Aggregation of solitary N2 worms and several social strains (including the social wild-type strain CB4856 and two *npr-1* loss-of-function mutants)

were measured in response to icas#3 using two different conditions: “high worm density,” with 120 worms per 5 cm plate; and “low worm density,” with 20 worms per 5 cm plate.

[0223] At low worm density, a very strong increase in aggregation was observed at concentrations as low as 10 fM (femtomolar) icas#3 for both solitary and social hermaphrodites (Figures 8A and 14E). Aggregation of N2 hermaphrodites increased as much as 4-fold at 1 pM icas#3, with higher icas#3 concentrations producing less aggregation. Similarly, the naturally occurring social strain CB4856 displayed a bell-shaped response curve with maximal aggregation at 1 pM of icas#3 and lower aggregation not significantly different from the control at 1 nM of icas#3 (Figure 8A). In contrast, icas#3 increased aggregation of *npr-1(ad609)* hermaphrodites over the entire tested concentration range, without a drop off at higher concentrations (Figure 8A). At high worm density, up to a 3-fold increase in aggregation of N2 hermaphrodites was observed on icas#3 plates (Figures 8B and 14F), whereas hermaphrodites from all three tested social strains showed nearly complete aggregation even in the absence of icas#3, which precluded detection of any additional aggregation-promoting effect of icas#3 (Figure 8B). These results show that icas#3 increases hermaphrodite aggregation even in the absence of a concentration gradient of this compound, and that solitary and social strains are similarly affected. Similarly, the second-most abundant indole ascaroside, icas#9, increased aggregation of both solitary and social hermaphrodites (Figure 6A).

[0224] The effect of icas#3 on aggregation of males, which generally tend to aggregate in the absence of hermaphrodites (Gems & Riddle, *Genetics* 154:1597-610 (2000), which is hereby incorporated by reference in its entirety), was also investigated. The aggregation of *him-5* males on icas#3 plates was found to be significantly increased (Figure 6B).

[0225] These results show that indole ascarosides promote aggregation behavior even in the absence of a concentration gradient, suggesting that sensing of icas#3 and icas#9 affects the response to other aggregation-promoting (chemical or other) signals or conditions. For example, secretion of additional indole ascarosides by the worms on plates containing exogenous icas#3 could contribute to the observed increase in aggregation. To investigate this possibility, *daf-22* hermaphrodites in the aggregation assay were tested. *daf-22* hermaphrodites did not produce indole ascarosides but responded to icas#3 in both the spot attraction and quadrant chemotaxis assay as strongly as N2 worms (Figures 3B and 5C). *daf-*

22 hermaphrodites were found to show less aggregation than N2 worms at 1 pM icas#3 but not at 10 pM icas#3 (Figure 8C). These results suggest that secretion of additional indole ascarosides or other *daf-22*-dependent compounds by the worms may contribute to aggregation on icas#3 plates; however, other factors, such as low oxygen levels or contact with other worms (Rogers et al., *Curr. Biol.* 16:649-59 (2006); Chang et al., *Publ. Lib. Sci. Biol.* 4(9):e274 (2006); Chang & Bargmann, *PNAS* 105:7321-26 (2008), which are hereby incorporated by reference in their entirety), may be more important. Furthermore, changes in locomotory behavior on icas#3 plates could affect the level of aggregation (Rogers et al., *Curr. Biol.* 16:649-59 (2006), which is hereby incorporated by reference in its entirety). Using an automated machine-vision system to track worm movement (Cronin et al., *BMC Genet.* 6:5 (2005), which is hereby incorporated by reference in its entirety), aggregation-inducing concentrations of icas#3 were found to strongly increase mean stopped duration and affect other locomotory parameters (Figures 6D–E and 8D). These changes in worm locomotion, in conjunction with other aggregation-mediating factors, may contribute to the observed increase in aggregation on icas plates.

Example 27—Response to icas#3 Requires the Sensory Neuron ASK and the Interneuron AIA

[0226] The amphid single-ciliated sensory neurons type K (ASK) play an important role in mediating *C. elegans* behaviors. Previous work has shown that the ASK neurons are required for behavioral responses of males and hermaphrodites to non-indole ascarosides (Srinivasan et al., *Nature* 454:1115-18 (2008); Macosko et al., *Nature* 458:1171-75 (2009), which are hereby incorporated by reference in their entirety). ASK sensory neurons are connected via anatomical gap-junctions to the RMG interneuron, which has been shown to act as a central hub regulating aggregation and related behaviors based on input from ASK and other sensory neurons (Macosko et al., *Nature* 458:1171-75 (2009); White et al., *Philos. Trans. R. Soc. London [Biol]* 314:1-340 (1986), which are hereby incorporated by reference in their entirety) (Figure 2A).

[0227] To investigate the neural circuitry required for icas#3-mediated hermaphrodite attraction and aggregation, tests were conducted to determine whether the ASK neurons are required for these behaviors. Worms lacking the ASK neurons due to cell-specific expression of mammalian caspase in the developing neurons (Tokumitsu Wakabayashi, Iwate University Japan) were used for these tests. Ablation of ASK sensory neurons resulted in a near complete loss of attraction to icas#3 (Figure 2B). In contrast, ablation of the ASI neurons,

which, like the ASK neurons partake in dauer pheromone sensing, had no significant effect on *icas#3*-mediated attraction in hermaphrodites (Figure 2B). Further, ablation of both ASI and ASK neurons did not result in a more significant loss of attraction compared to ASK ablation alone, suggesting that the ASK sensory neurons are required for sensing *icas#3* (Figure 2B).

[0228] Tests were also conducted to determine whether the ASK neurons are required for *icas#3*-mediated aggregation. The results show that hermaphrodites lacking the ASK neurons did not aggregate in response to *icas#3* at any of the tested concentrations (Figure 2C). Locomotory analysis of ASK-ablated hermaphrodites on *icas#3* plates showed neither increased reversal frequency nor decreased velocity, as observed for wild-type worms (Figures 7A–B).

[0229] Tests were then conducted to determine whether the RMG interneuron is required for *icas#3*-mediated behaviors. The cell position of the RMG interneuron in wild-type worms and in a transgenic strain expressing *ncs-1::gfp* (a gift from the Bargmann lab) was identified using differential interference contrast (DIC) microscopy (Sulston et al., *Dev. Biol.* 100:64-119 (1983), which is hereby incorporated by reference in its entirety). This transgene expresses GFP in the RMG interneuron and a few other sensory neurons (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety). It was found that ablation of the RMG interneuron in both wild-type and *ncs-1::gfp* worms did not affect *icas#3* response in the spot attraction assay (Figure 2B). These results indicate that the RMG interneuron is not required for transduction of *icas#3*-derived attraction signals from the ASK sensory neurons, contrary to the behavioral effects of non-indole ascarosides, which require both the ASK sensory neurons and the RMG interneuron (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety).

[0230] Given this observation, tests were also conducted to determine which interneuron downstream of ASK is required for response to *icas#3*. According to the wiring diagram of *C. elegans*, the primary synaptic output of the ASK neuron is the AIA interneuron (White et al., *Philos. Trans. R. Soc. London [Biol]* 314:1-340 (1986), which is hereby incorporated by reference in its entirety). A transgenic line expressing a hyperactive form of MEC-4 in the AIA interneuron (a gift from the Ishihara lab, Japan) (Shinkai et al., *J. Neurosci.* 31:3007-15 (2011), which is hereby incorporated by reference in its entirety) was used to test whether the AIA interneuron is required for sensing *icas#3*. Expression of MEC-4, a DEG/ENaC sodium channel, causes neuronal toxicity in *C. elegans*, thereby resulting in

the loss of the AIA neuron (Harbinder et al., *PNAS* 94:13128-33 (1997), which is hereby incorporated by reference in its entirety). The AIA-deficient worms did not show any attraction to icas#3, suggesting that the AIA interneurons are required for the icas#3 response. Hence the neural circuitry required for attraction to icas#3 is different from that of the non-indole ascarosides.

[0231] Because behavioral assays showed that the ASK and AIA neurons participate in sensing icas#3, tests were conducted to determine whether icas#3 elicits calcium flux in these neurons. To measure Ca^{2+} flux, transgenic lines expressing the genetically encoded calcium sensors (GCaMP) in these neurons (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety) were used. The “Olfactory chip” was used to restrain the worms and apply ON and OFF steps of icas#3 while imaging from these neurons (Chronis et al., *Nat. Methods* 4:727-31 (2007), which is hereby incorporated by reference in its entirety). There was no detection of Ca^{2+} transients in ASK neurons even when a wide range of concentration ranging from 1 pM to 1 μM was applied. Calcium responses in the AIA interneuron, which is the primary synaptic target of the ASK neuron (White et al., *Philos. Trans. R. Soc. London [Biol]* 314:1-340 (1986), which is hereby incorporated by reference in its entirety), was then monitored. The results showed that icas#3 elicited significantly increased G-CaMP fluorescence in the AIA neurons (Figures 2D–E), similar to the results reported for stimulation of AIA interneurons with a mixture of three non-indole ascarosides (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety). These results show that the ASK sensory neurons are required for the response to icas and that this response is transduced via the AIA interneuron.

Example 28—Icas#3 and ascr#3 Are Competing Signals

[0232] Previous studies have shown that high dauer-inducing concentrations of ascr#3 strongly deter both social and solitary hermaphrodites (Srinivasan et al., *Nature* 454:1115-18 (2008); Macosko et al., *Nature* 458:1171-75 (2009), which are hereby incorporated by reference in their entirety). To investigate whether addition of ascr#3 would affect icas#3-mediated attraction of hermaphrodites, mixtures containing these two compounds in a near-physiological ratio of 12:1 (ascr#3:icas#3) were tested in a modified spot attraction assay, in which N2 hermaphrodite attraction was scored to three concentric zones A–C (Figure 1A). The results show that at the lower of the two concentrations tested (120 fmol ascr#3 and 10 fmol icas#3), the presence of ascr#3 did not interfere with icas#3-

mediated attraction, whereas higher concentrations of ascr#3 resulted in strong repulsion, even in the presence of proportionally increased icas#3 concentrations (12 pmol ascr#3 and 1 pmol icas#3, Figure 4A). Following retreat from the outer edge of zone A, many worms remained “trapped” in a circular zone B surrounding central zone A, repulsed by the high concentration of the icas#3/ascr#3 blend inside zone A, but attracted by the lower concentrations of the icas#3/ascr#3 blend that diffused into zone B. These results show that the repulsive effect of ascr#3 prevails at high concentrations of physiological icas#3/ascr#3 mixtures, whereas attraction by icas#3 dominates at lower concentrations.

Discussion of Examples 1–28

[0233] Indole ascarosides are the first *C. elegans* pheromones shown to strongly attract wild-type hermaphrodites and promote aggregation. The indole ascarosides fit the broad definition of aggregation pheromones in that they attract and/or arrest conspecifics to the region of release irrespective of sex (EDWARD O. WILSON, *SOCIOBIOLOGY: THE NEW SYNTHESIS* (The Belknap Press of Harvard University, Cambridge, Mass., 25th Anniv. ed. 2000); Shorey, *Annu. Rev. Entomol.* 18:349-80 (1973); Wertheim et al., *Annu. Rev. Entomol.* 50:321-46 (2005), which are hereby incorporated by reference in their entirety). In promoting these behaviors, the indole ascarosides are active at such low (femtomolar) concentrations that the worm’s behavioral response must result from sensing of only a few molecules. For example, at an icas#3 concentration of 10 fM, there are only about 20 icas#3 molecules contained in a cylinder corresponding to the length and diameter of an adult hermaphrodite. Given their high specific activity, it is reasonable that indole ascarosides are of much lower abundance than non-indole ascarosides.

[0234] The indole ascarosides’ strongly attractive properties suggest that these compounds serve to attract conspecifics to desirable environments such as food sources. However, the results from the competition experiments indicate that attraction of hermaphrodites by icas#3 can be counteracted by high concentrations of ascr#3, which are repulsive to hermaphrodites (Srinivasan et al., *Nature* 454:1115-18 (2008), which is hereby incorporated by reference in its entirety). The competition experiments further show that at low concentrations of a physiological blend of icas#3 and ascr#3, the attractive properties of icas#3 dominate, whereas at high concentrations of the same blend, the repulsion by ascr#3 becomes dominant (Figure 4A). These findings suggest that under dauer-inducing conditions with high population density, the associated high concentrations of ascr#3 promote dispersal (Srinivasan et al., *Nature* 454:1115-18 (2008), which is hereby incorporated by reference in

its entirety), whereas low population density and correspondingly lower concentrations of ascr#3 result in attraction mediated by icas#3. Therefore, icas' and ascr's can represent opposing stimuli regulating population density and level of aggregation. In turn, population density, food availability, and other environmental factors may affect relative rates of the biosyntheses of ascr's and icas' as part of a regulatory circuit.

[0235] Indole ascarosides affect aggregation behavior even in the absence of a concentration gradient: very low background concentrations (fM-pM) of icas#3 and icas#9 strongly increased the propensity of hermaphrodites (and males) to aggregate. This finding suggests that sensing of icas#3 and icas#9 increases susceptibility for aggregation-promoting (chemical or other) signals or conditions, such as additional quantities of icas secreted by the worms on the plate.

[0236] Aggregation in *C. elegans* is known to depend on a diverse set of genetic factors and environmental conditions, including food availability and oxygen concentration, suggesting the existence of neuronal circuitry that integrates inputs from different sources (de Bono & Bargmann, *Cell* 94:679-89 (1998); Cheung et al., *Curr. Biol.* 15:905-17 (2005); Coates & de Bono, *Nature* 419:925-29 (2002); de Bono et al., *Nature* 419:899-903 (2002); Gray et al., *Nature* 430:317-22 (2004), which are hereby incorporated by reference in their entirety). Aggregation and attraction signals originating from several different sensory neurons, including the oxygen-sensing URX-neurons and the ascr-sensing ASK neurons, have recently been shown to converge on the RMG interneuron, which is proposed to act as a central hub coordinating these behaviors (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety). The RMG interneuron is the central site of action of the neuropeptide-Y receptor homolog NPR-1, which distinguishes solitary strains (high NPR-1 activity) from social strains (low NPR-1 activity) (de Bono & Bargmann, *Cell* 94:679-89 (1998); Rogers et al., *Nat. Neurosci.* 6:1178-85 (2003), which are hereby incorporated by reference in their entirety). In social *npr-1(lf)* mutant hermaphrodites, oxygen-sensing URX neurons promote aggregation at the edges of the bacterial lawn, whereas solitary N2 hermaphrodites do not respond to oxygen gradients. Similarly, repulsion by ascr depends on NPR-1, as solitary hermaphrodites are repelled by ascr, whereas social *npr-1(lf)* hermaphrodites display either greatly diminished repulsion or weak attraction (Macosko et al., *Nature* 458:1171-75 (2009), which is hereby incorporated by reference in its entirety).

[0237] In contrast, *icas#3* was shown to promote hermaphrodite attraction and aggregation in both social and solitary strains. *Icas#3* attracted solitary N2 as well as social *npr-1(lf)* hermaphrodites and increased hermaphrodite aggregation in the solitary strain N2, the social wild-type strains RC301 and CB4856 (Hawaii) carrying a low-activity variant of NPR-1, and the two tested *npr-1* null alleles. The finding that *icas#3*-mediated attraction and aggregation were not reduced by high NPR-1 activity suggests that these *icas#3*-mediated behaviors rely on signaling pathways distinct from those controlling aggregation responses to other types of stimuli, such as low oxygen levels. This is supported by the observation that hermaphrodites lacking the RMG interneuron, which coordinates other aggregation responses via NPR-1, were still attracted to *icas#3*. Furthermore, *icas#3*-mediated aggregation differed from NPR-1-dependent aggregation behavior in that aggregation of worms on *icas#3* plates was more dynamic and not restricted to the edge of the bacterial lawn where oxygen is limited. Worm velocity was not significantly reduced at the *icas#3* concentrations that induce maximal aggregation (1–10 pM, Figure 6D), and *icas#3*-mediated aggregation was associated with less clumping (average clump size 3–5 worms) than that found for aggregating NPR-1 mutant worms (average clump size 6–16 worms) (Rogers et al., *Curr. Biol.* 16:649-59 (2006), which is hereby incorporated by reference in its entirety). These observations show that *icas#3*-mediated aggregation is phenotypically distinct from aggregation behaviors controlled by NPR-1 and the RMG interneuron.

[0238] *Icas#3*-mediated attraction and aggregation were shown to depend on the ASK neurons, similar to hermaphrodite repulsion and male attraction by *ascr* (Srinivasan et al., *Nature* 454:1115-18 (2008), which is hereby incorporated by reference in its entirety), confirming the central role of this pair of sensory neurons for perception of different types of pheromones in *C. elegans* (Figures 2A–E). Further, *icas#3* responses were shown to depend on the AIA interneurons and did not require the RMG interneuron. Therefore, it appears that the sensory neuron ASK participates in perception of two different types of pheromones, *ascr*'s and *icas*', and that these signals are transduced via two different neurophysiological pathways as part of a complex neural and genetic circuitry integrating a structurally diverse array of pheromone signals.

[0239] Calcium transients have been recorded from amphid sensory neurons in response to non-indole ascarosides; however, the reported changes in G-CaMP fluorescence are relatively small (on the order of about 20%) (Macosko et al., *Nature* 458:1171-75 (2009); Kim et al., *Science* 326:994-98 (2009), which are hereby incorporated by reference in their

entirety). It has been recently reported that the non-indole ascaroside ascr#5 does not elicit calcium transients in the ASI sensory neurons, although the ASI neurons function as sensors of ascr#5 and express the ascr#5-receptors *srg-36* and *srg-37* (McGrath et al., *Nature* 477:321-25 (2011), which is hereby incorporated by reference in its entirety). Similarly, there was no detection of significant Ca^{2+} transients in the ASK neurons in response to a wide range of concentrations of icas#3. Perhaps any icas#3-elicited Ca^{2+} signals in this neuron are even weaker than those of non-indole ascarosides, as icas#3 is active at extremely low concentrations (femtomolar to low picomolar). Further, there may be involvement of additional neurons in icas#3 signaling, given that the ASK neurons are postsynaptic to a number of other sensory neurons (White et al., *Philos. Trans. R. Soc. London [Biol]* 314:1-340 (1986), which is hereby incorporated by reference in its entirety). In particular, as shown herein, icas#3 elicited significant changes in G-CaMP fluorescence in the AIA interneurons, which are the primary postsynaptic targets of the ASK sensory neurons (Figures 2D–E).

[0240] The identification of indole ascarosides as aggregation signals reveals an unexpected complexity of social signaling in *C. elegans*. The results herein indicate that ascarylose-derived small molecules (icas' and ascr's) serve at least three distinct functions in *C. elegans*: dauer induction, male attraction, and hermaphrodite social signaling (Figure 4B). Previous studies have shown that ascr's often have more than one function. For example, ascr#3 plays significant roles for both dauer signaling and male attraction (Butcher et al., *Nat. Chem. Biol.* 3:420-22 (2007); Srinivasan et al., *Nature* 454:1115-18 (2008), which are hereby incorporated by reference in their entirety). The results herein demonstrate that specific structural variants of ascarylose-derived small molecules are associated with specific functions (Figure 4C). Addition of an indole carboxy group to ascr's changes the signaling properties such that the indole-modified compounds can have signaling effects very different from those of the unmodified compounds: icas#3 strongly attracts hermaphrodites and promotes aggregation, whereas ascr#3 repulses hermaphrodites and attracts males. It has also been shown herein that, in addition to structural variation, distinct signaling functions are associated with different concentration windows: while high nanomolar concentrations of ascr's are required for dauer formation, low nanomolar to high picomolar concentrations of ascr's promote male attraction; and picomolar to femtomolar concentrations of icas' promote hermaphrodite attraction and aggregation (Figure 4B).

[0241] Social signaling in *C. elegans* thus appears to be based on a modular language of small molecules, derived from combinatorial assembly of several structurally distinct

building blocks (Figure 4C). Different combinations of these building blocks serve different, occasionally overlapping, signaling functions. The results for the relative abundance of ascr's and icas' with identical side chains (Figure 9G) indicate that integration of the different building blocks is carefully controlled. Biochemically, the building blocks are derived from three basic metabolic pathways: carbohydrate metabolism, peroxisomal fatty-acid β -oxidation, and amino acid metabolism. These structural observations raise the possibility that social signaling via small molecules transduces input from the overall metabolic state of the organism. Food availability and nutrient content in conjunction with other environmental factors may control ascr and icas biosynthesis pathways to generate specific pheromone blends that differentially regulate aggregation, mate attraction, and developmental timing. The expansive vocabulary of a modular chemical language would make it possible for different nematodes to signal conspecifically as well as interspecifically, but it was not known whether nematode species other than *C. elegans* rely on ascarylose-based small molecules for chemical communication. Lipid-derived glycosides of ascarylose have been identified from several other nematode species (Bartley et al., *J. Nat. Prod.* 59:921-26 (1996), which is hereby incorporated by reference in its entirety), which may indicate that nematodes may have the ability to produce ascr- or icas-like compounds.

[0242] The identification of indole ascarosides as attraction and aggregation signals has demonstrated that *C. elegans* aggregation behavior depends on dedicated chemical signals produced by conspecifics and not just shared preference for specific environmental conditions. *C. elegans* social signaling thus appears to be significantly more highly evolved than previously suspected.

Example 29—Analytical Instrumentation

[0243] NMR spectra were recorded on a Varian INOVA 600 (600 MHz for ^1H , 151 MHz for ^{13}C), INOVA 500 (500 MHz for ^1H , 125 MHz for ^{13}C), or INOVA 400 (400 MHz for ^1H , 100 MHz for ^{13}C) spectrometer. NMR spectra were processed using Varian VNMR, MestreLabs MestReC and Mnova software packages.

[0244] Low-resolution HPLC–MS and HPLC–MS/MS were performed using an Agilent 1100 Series HPLC system equipped with a diode array detector and connected to a Quattro II mass spectrometer (Micromass/Waters). High-resolution MS/MS was performed using an LTQ Orbitrap Velos Hybrid Fourier transform (FT) mass spectrometer (Thermo Scientific, Cornell University Life Sciences Core Laboratories Center). High-resolution HPLC-MS was performed using a Waters nanoACQUITY UPLC System equipped with a

Waters Acquity UPLC HSS C-18 column (2.1 × 100 mm, 1.8 µm particle diameter) connected to a Xevo G2 QToF Mass Spectrometer. MassLynx software was used for MS data acquisition and processing.

[0245] Flash column chromatography was performed using a Teledyne ISCO CombiFlash system. HPLC fractionation was performed using the Agilent 1100 Series HPLC system equipped with an Agilent Eclipse XDB-C18 column (9.4 × 250 mm, 5 µm particle diameter) coupled to a Teledyne ISCO Foxy 200 fraction collector.

Example 30—*C. elegans* Strains and General Culture Methods

[0246] *C. elegans* variety Bristol, strain N2 (wild type), *acox-1(ok2257)*, *dhs-28(hj8)*, *maoc-1(hj13)*, *maoc-1(ok2645)*, *daf-22(m130)*, *daf-22(ok693)*, F58F9.7(*tm4033*), C48B4.1(*ok2619*), F59F4.1(*ok2119*), and F08A8.3(*tm5192*) were maintained at 20 °C on NGM agar plates, made with Bacto agar (BD Biosciences) and seeded with *E. coli* OP50 grown overnight.

Example 31—Preparation of Metabolite Extracts

[0247] Wild-type (N2, Bristol) or *acox-1(ok2257)*, *maoc-1(hj13)*, *dhs-28(hj8)*, and *daf-22(ok693)* mutant worms from four 10 cm NGM plates were washed using M9-medium into a 100 mL S-medium pre-culture where they were grown for four days at 22°C on a rotary shaker at 220 revolutions per minute (rpm). Concentrated *E. coli* OP50 derived from 1 L of bacterial culture was added as food at days 1 and 3. Subsequently, each pre-culture was divided equally into four 500 mL Erlenmeyer flasks containing 100 mL of S-medium on day 4. Two of these cultures, labeled non-starved (NS), were grown for 5 days at 22°C on a rotary shaker, and fed with concentrated OP50 derived from 500 mL of bacterial culture every day from day 1 to day 4. The remaining two cultures of each set, labeled starved (S), were fed once with concentrated OP50 derived from 500 mL of bacterial culture on day 1, and grown for an additional 9 days at 22°C on a rotary shaker without food. Subsequently, the cultures were harvested on day 5 for NS and day 10 for S, and centrifuged. The resulting supernatant media and worm pellets were frozen over dry ice–acetone slush and lyophilized separately. The lyophilized materials from the supernatant were extracted with 150 mL of 95% ethanol at room temperature for 16 hours. The worm pellets were crushed with ~2 g of granular NaCl using a mortar and pestle, and extracted with 100 mL of 100% ethanol at room temperature for 16 hours. The resulting suspensions were filtered. The filtrate was evaporated in vacuum

at room temperature, producing media metabolite (the worm “excretome”) extracts and worm pellet metabolite extracts.

Example 32—Ascaroside Feeding Experiment with *daf-22(m130)*

[0248] Ascaroside feeding experiments were performed with the *daf-22(m130)* mutant, which is less sensitive to growth defects due to added ascarosides than the *daf-22(ok693)* mutant. HPLC-MS analysis of *daf-22(m130)* showed similar ascaroside profiles as *daf-22(ok693)*, notably a total lack of short chain ascarosides with chain length less than 12 carbons. Non-starved cultures of *daf-22 (m130)* were grown as described in Example 31, but with the addition of 5 μ M per culture of either *oscr#3* or a 1:1 mixture of *oscr#10* and *oscr#10*, added on day 1 after pre-culture splitting.

Example 33—Sample Preparation

[0249] Media or worm pellet metabolite extracts were resuspended in ~15 mL methanol and centrifuged. The supernatants were then collected, concentrated in vacuum at room temperature, resuspended in 1 mL methanol, and centrifuged. 30 μ L of the resulting extract was directly injected for LC-MS/MS analysis.

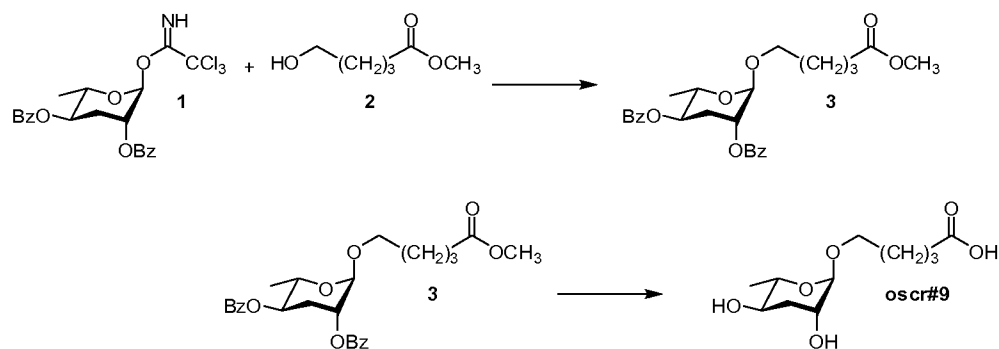
Example 34—Mass Spectrometric Analysis

[0250] Low resolution HPLC-MS/MS profiling was performed using the Agilent 1100 Series HPLC system equipped with an Agilent Eclipse XDB-C18 column (9.4 $\times$ 250 mm, 5 μ m particle diameter) connected to the Quattro II mass spectrometer using a 10:1 split. A 0.1% acetic acid–acetonitrile solvent gradient was used at a flow rate of 3.6 ml/min, starting with an acetonitrile content of 5% for 5 minutes, which was increased to 100% over a period of 40 minutes. Metabolite extracts were analyzed by HPLC-ESI-MS in negative and positive ion modes, using a capillary voltage of 3.5 kV and a cone voltage of -40 V and +20 V, respectively. HPLC-MS/MS screening for precursor ions of $m/z = 73.0$ (negative mode) and neutral loss of 130.0 (positive mode) was performed using argon as collision gas at 2.1 mtorr and 30 eV. Ascaroside fragmentation was further analyzed by high-resolution MS/MS using the LTQ Orbitrap. To confirm the elemental composition of new compounds, mutant metabolome samples and fractions were additionally analyzed by high-resolution HPLC-MS using the Xevo G2 QTof.

Example 35—Synthesis of Ascaroside *oscr#9*

[0251] Ascaroside *oscr#9* was prepared as shown in Scheme 2 below.

Scheme 2



[0252] To prepare **3**, a solution of 2,4-di-*O*-benzoyl-ascarylose-1-(2,2,2-trichloroacetimide) (**1**, 132 mg, 263 μ mol, Butcher et al., *Nat. Chem. Biol.* 3:420–22 (2007), which is hereby incorporated by reference in its entirety) and methyl 5-hydroxypentanoate (**2**, 125 mg, 950 μ mol, Huckstep et al., *Synthesis* 10:881–82 (1982), which is hereby incorporated by reference in its entirety) in dry DCM (3 ml) at 0°C was treated with trimethylsilyloxy triflate (5 μ l). After 3 hours, the solution was washed with saturated aqueous NaHCO₃ solution (0.5 ml), dried over Na₂SO₄, and concentrated in vacuum. Flash column chromatography on silica using a gradient of 20–40% ethyl acetate in hexanes afforded Methyl 5-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)pentanoate (**3**) (66.8 mg, 142 μ mol, 47%) as a colorless oil. ¹H NMR (400 MHz, acetone-*d*₆): δ (ppm) 1.28 (d, *J* = 6.2 Hz, 3H), 1.67–1.80 (m, 4H), 2.23 (m, 1H), 2.40 (t, *J* = 6.9 Hz, 2H), 2.48 (m, 1H), 3.58 (m, 1H), 3.64 (s, 3H), 3.83 (m, 1H), 4.13 (dq, *J* = 9.8 Hz, *J* = 6.0 Hz, 1H), 4.87 (s.br, 1H), 5.15 (ddd, *J* = 11.0 Hz, *J* = 10.4 Hz, *J* = 4.5 Hz, 1H), 5.18 (s.br, 1H), 7.50–7.60 (m, 4H), 7.62–7.72 (m, 2H), 8.05 (d, *J* = 7.5 Hz, 2H), 8.11 (d, *J* = 7.5 Hz, 2H); ¹³C NMR (100 MHz, acetone-*d*₆): δ (ppm) 18.3, 22.5, 29.6, 30.4, 34.0, 51.5, 67.5, 67.9, 71.4, 71.5, 97.0, 129.4, 129.5, 130.3, 130.4, 131.0, 131.0, 134.1, 134.2, 165.9, 166.0, 174.0. See Figures 15A–B.

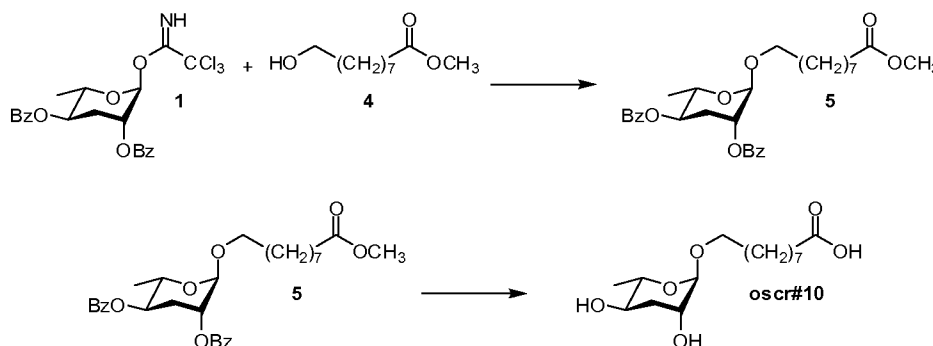
[0253] To prepare oscr#9, a solution of **3** (66.8 mg, 142 μ mol) in dry THF (0.5 ml) was added to a mixture of LiOH (28 mg, 1.4 mmol) and water (0.6 ml) in 1,4-dioxane (4 ml). After stirring at 66°C for 2 hours, the solution was acidified with glacial acetic acid and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–30% methanol in DCM containing 1% acetic acid afforded 5-(3'*R*,5'*R*-Dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)pentanoic acid (oscr#9) (26 mg, 105 μ mol, 74%) as a colorless oil. ¹H NMR (400 MHz, methanol-*d*₄): δ (ppm) 1.22 (d, *J* = 6.0 Hz, 3H), 1.58–1.72 (m, 4H), 1.77 (ddd, *J* = 13.1 Hz, *J* = 11.1 Hz, *J* = 3.2 Hz, 1H), 1.95 (ddt, *J* = 13.1 Hz, *J* = 3.7 Hz, *J* = 0.9 Hz, 1H), 2.33 (t, *J* = 7.2 Hz, 2H), 3.43 (dt, *J* = 9.6 Hz, *J* = 6.0 Hz, 1H), 3.47–3.59 (m, 2H), 3.71 (dt, *J* = 9.8 Hz, *J* = 6.2 Hz, 1H), 3.77 (m, 1H), 4.50 (s, 1H); ¹³C NMR (100

MHz, methanol-*d*₄): δ (ppm) 18.1, 23.0, 30.1, 34.7, 36.0, 67.9, 68.3, 69.4, 70.9, 100.4, 177.5. See Figures 16A–B.

Example 36—Synthesis of Ascaroside oscr#10

[0254] Ascaroside oscr#10 was prepared as shown in Scheme 3 below.

Scheme 3



[0255] To prepare **5**, a solution of 2,4-di-*O*-benzoyl-ascarylose-1-(2,2,2-trichloroacetimide) (**1**, 132 mg, 263 μ mol, Butcher et al., *Nat. Chem. Biol.* 3:420–22 (2007), which is hereby incorporated by reference in its entirety) and methyl 9-hydroxynonanoate (**4**, 112.8 mg, 600 μ mol, Kai et al., *Tetrahedron* 64:6760–69 (2008), which is hereby incorporated by reference in its entirety) in dry DCM (3 ml) at 0°C was treated with trimethylsilyloxy triflate (5 μ l). After 3 hours, the solution was washed with saturated aqueous NaHCO₃ solution (0.5 ml), dried over Na₂SO₄, and concentrated in vacuum. Flash column chromatography on silica using a gradient of 20–40% ethyl acetate in hexanes afforded Methyl 9-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)nonanoate (**5**) (99.3 mg, 190 μ mol, 72%) as a colorless oil. ¹H NMR (400 MHz, acetone-*d*₆): δ (ppm) 1.28 (d, *J* = 6.2 Hz, 3H), 1.30 – 1.40 (m, 6H), 1.40 – 1.49 (m, 2H), 1.56 – 1.72 (m, 2H), 2.22 (ddd, *J* = 13.6 Hz, *J* = 11.5 Hz, *J* = 3.2 Hz, 1H), 2.30 (t, *J* = 7.5 Hz, 2H), 2.46 (m, 1H), 3.55 (dt, *J* = 9.8 Hz, *J* = 6.5 Hz, 1H), 3.60 (s, 3H), 3.81 (dt, *J* = 9.6 Hz, *J* = 6.6 Hz, 1H), 4.13 (dq, *J* = 9.7 Hz, *J* = 6.2 Hz, 1H), 4.86 (s.br, 1H), 5.15 (ddd, *J* = 11.4 Hz, *J* = 9.8 Hz, *J* = 4.6 Hz, 1H), 5.18 (m, 1H), 7.50 – 7.60 (m, 4H), 7.63 – 7.71 (m, 2H), 8.04 (m, 2H), 8.11 (m, 2H); ¹³C NMR (100 MHz, acetone-*d*₆): δ (ppm) 18.3, 25.6, 26.8, 29.7, 29.9, 30.0, 30.2, 30.4, 34.4, 51.4, 67.4, 68.2, 71.4, 71.5, 97.0, 129.4, 129.5, 130.2, 130.3, 130.9, 131.0, 134.1, 134.2, 165.9, 165.9, 174.3. See Figures 17A–B.

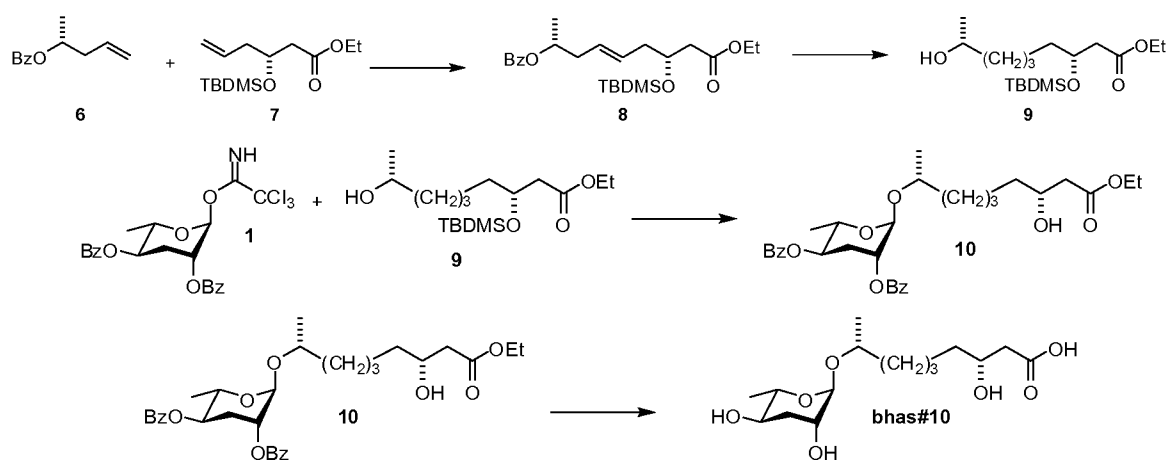
[0256] To prepare ascr#10, a solution of **5** (99.3 mg, 190 μ mol) in THF (500 μ l) was added to a mixture of LiOH (38 mg, 1.9 mmol) and water (800 μ l) in 5 ml 1,4-dioxane (5 ml). After stirring at 66°C for 3 hours, the solution was acidified with acetic acid and

concentrated in vacuum. Flash column chromatography on silica gel using a gradient of 5–30% methanol in DCM containing 1% glacial acetic acid afforded 9-(3'*R*,5'*R*-Dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)nonanoic acid (oscr#10) (49 mg, 161 μ mol, 85%) as a colorless oil. ^1H NMR (400 MHz, methanol- d_4): δ (ppm) 1.22 (d, $J = 6.1$ Hz, 3H), 1.32–1.43 (m, 8H), 1.56–1.63 (m, 4H), 1.77 (ddd, $J = 13.1$ Hz, $J = 11.1$ Hz, $J = 3.2$ Hz, 1H), 1.96 (ddt, $J = 13.1$ Hz, $J = 3.7$ Hz, $J = 0.9$ Hz, 1H), 2.28 (t, $J = 7.4$ Hz, 2H), 3.41 (dt, $J = 9.6$ Hz, $J = 6.2$ Hz, 1H), 3.49–3.59 (m, 2H), 3.68 (dt, $J = 9.8$ Hz, $J = 5.5$ Hz, 1H), 3.76 (m, 1H), 4.49 (s, 1H); ^{13}C NMR (100 MHz, methanol- d_4): δ (ppm) 17.3, 25.2, 26.4, 28.0, 29.3, 29.5, 29.6, 30.5, 34.1, 61.1, 67.4, 68.5, 69.9, 99.4, 176.8. See Figures 18A–B.

Example 37—Synthesis of Ascaroside bhas#10

[0257] Ascaroside bhas#10 was prepared as shown in Scheme 4 below.

Scheme 4



[0258] To prepare **9**, a solution of **6** (366 mg, 1.91 mmol, Guan & Greenberg, *J. Am. Chem. Soc.* 131:15225–31 (2009), which is hereby incorporated by reference in its entirety) and **7** (104 mg, 380 μ mol, Evans & Andrews, *Angew. Chem. Int. Ed.* 47:5426–29 (2008), which is hereby incorporated by reference in its entirety) in dry DCM (10 ml) was treated with 1,4-benzoquinone (4 mg, 38 μ mol) in DCM (0.5 ml) and stirred for 10 minutes. A solution of Grubbs 2nd generation catalyst (16 mg, 19 μ mol) in DCM (0.5 ml) was added. The resulting mixture was stirred at 40°C. After 20 hours, the mixture was filtered over a small layer of silica and concentrated in vacuum. Flash column chromatography on silica using a gradient of 0–20% ethyl acetate in hexane afforded a mixture of the desired product **8** and the homodimer of **6**. The mixture was not purified further; instead, the crude mixture (160 mg) was dissolved in ethanol (2 ml), treated with Pd/C (15 mg, 10%, w/w), and hydrogenated for 40 hours. The resulting mixture was filtered, concentrated in vacuum, and purified by flash

column chromatography on silica gel using a gradient of 10–30% ethyl acetate in hexane to afford Ethyl (8*R*)-hydroxy-(3*R*)-*tert*-butyldimethylsilyloxynonanoate (**9**) (48 mg, 144 μ mol, 38% over two steps) as a colorless oil. ^1H NMR (500 MHz, chloroform- d_1): δ (ppm) 0.03 (s, 3H), 0.06 (s, 3H), 0.86 (s, 9H), 1.19 (d, $J = 6.2$ Hz, 3H), 1.26 (t, $J = 7.2$ Hz, 3H), 1.29–1.47 (m, 6H), 1.47–1.53 (m, 2H), 2.40 (dd, $J = 14.6$ Hz, $J = 5.7$ Hz, 1H), 2.44 (dd, $J = 14.6$ Hz, $J = 7.0$ Hz, 1H), 3.75–3.83 (m, 1H), 4.09–4.15 (m, 3H). See Figure 19.

[0259] To prepare **10**, a solution of 2,4-di-O-benzoyl-ascarylose-1-(2,2,2-trichloroacetimide) (**1**, 62 mg, 120 μ mol, Butcher et al., *Nat. Chem. Biol.* 3:420–22 (2007), which is hereby incorporated by reference in its entirety) in dry DCM (2 ml) at -10°C was treated with **9** (47 mg, 141 μ mol) and trimethylsilyloxy triflate (10 μ l). After 3.5 hours, the solution was washed with saturated aqueous NaHCO_3 solution (0.5 ml), dried over Na_2SO_4 , and concentrated in vacuum. Flash column chromatography on silica using a gradient of 10–40% ethyl acetate in hexane afforded Ethyl (8*R*)-(3'*R*,5'*R*-Dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxynonanoate (**10**) (4.0 mg, 7.2 μ mol, 6%) as a colorless oil. ^1H NMR (400 MHz, chloroform- d_1): δ (ppm) 1.19 (d, $J = 6.1$ Hz, 3H), 1.27 (t, $J = 7.2$ Hz, 3H), 1.28 (d, $J = 6.4$ Hz, 3H), 1.33–1.72 (m, 8H), 2.20 (ddd, $J = 14.3$ Hz, $J = 11.6$ Hz, $J = 3.2$ Hz, 1H), 2.42 (dd, $J = 16.5$ Hz, $J = 9.0$ Hz, 1H), 2.38–2.45 (m, 1H), 2.52 (dd, $J = 16.5$ Hz, $J = 3.0$ Hz, 1H), 3.00 (d, $J = 3.9$ Hz, 1H), 3.80–3.89 (m, 1H), 3.98–4.07 (m, 1H), 4.11 (dq, $J = 9.7$ Hz, $J = 6.1$ Hz, 1H), 4.17 (q, $J = 7.2$ Hz, 2H), 4.95 (s.br, 1H), 5.12–5.22 (m, 2H), 7.43–7.50 (m, 4H), 7.55–7.62 (m, 2H), 8.05 (d, $J = 7.5$ Hz, 2H), 8.11 (d, $J = 7.5$ Hz, 2H). See Figure 20.

[0260] To prepare bhas#10, a solution of **10** (4 mg, 7.2 μ mol) in THF (150 μ l) was treated with LiOH (7 mg, 290 μ l) in water (100 μ l) and 1,4-dioxane (250 ml) at 67°C for 5 hours. The reaction mixture was acidified with acetic acid (100 μ l), concentrated in vacuum, treated with methanol (2 ml), and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–25% methanol in DCM with 0.5% glacial acetic acid afforded (8*R*)-(3'*R*,5'*R*-Dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxynonanoic acid (bhas#10) (1.5 mg, 4.7 μ mol; 65%) as a colorless oil.

[0261] ^1H (600 MHz), ^{13}C (151 MHz), and HMBC NMR spectroscopic data for bhas#10 were obtained using methanol- d_4 and are shown in Table 4 below. Chemical shifts are referenced to $(\text{CD}_2\text{HOD}) = 3.31$ ppm and $(\text{CD}_2\text{HOD}) = 49.05$ ppm. See Figures 21A–D.

Table 4. NMR spectroscopic data of bhas#10

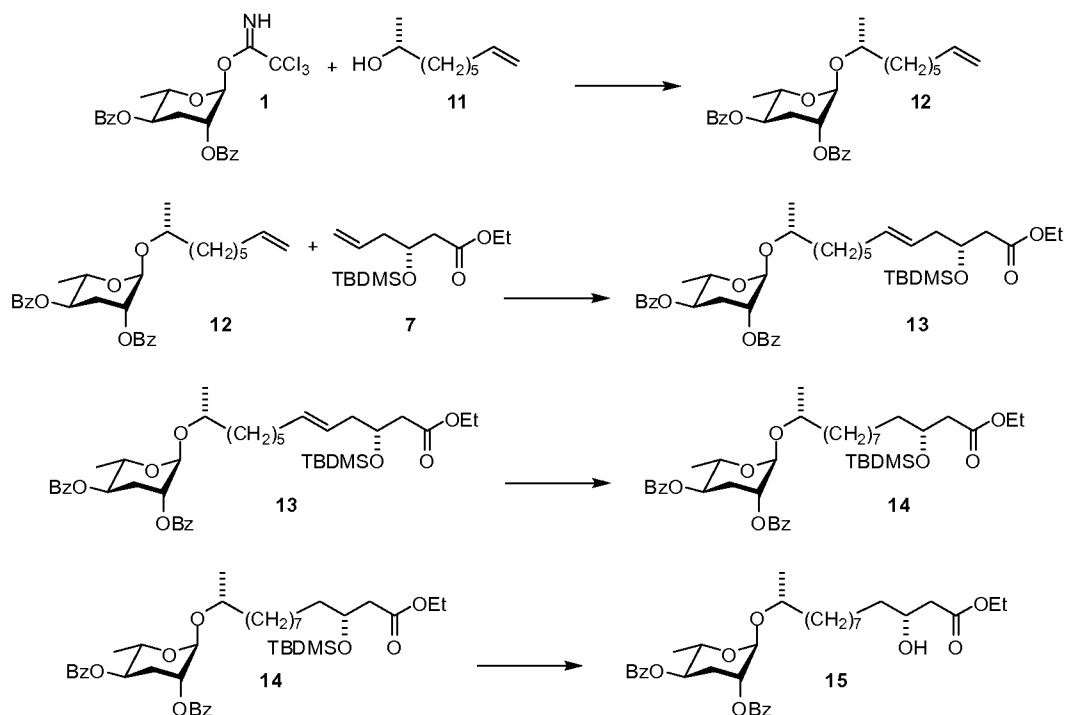
Position	$\delta^{13}\text{C}$ [ppm]	$\delta^1\text{H}$ [ppm]	^1H - ^1H -coupling constants [Hz]	Relevant HMBC correlations
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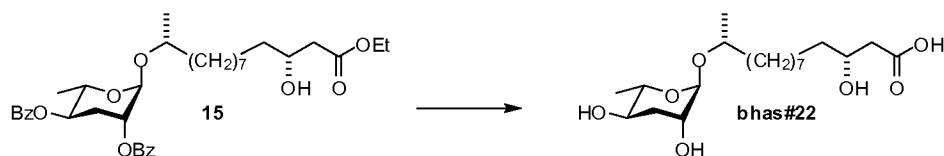
1	178.5			
2	44.8	2.23-2.33 2.34-2.44		
3	70.0	3.89-3.98		
4	38.0	1.44-1.52		
5-6	26.8, 27.0	1.33-1.52		
7	38.4	1.44-1.63		C6, C8,
8	72.5	3.78		C1', C-7, C-9
9	19.3	1.12	$J_{8,9} = 6.2$	C-8, C-7
1'	97.5	4.64		C-3', C-5', C-8
2'	70.0	3.71		
3'	35.9	1.77 (<i>ax</i>)	$J_{3'ax,3'eq} = 13.2$, $J_{3'ax,4'} = 11.5$, $J_{2',3'ax} = 3.1$	C-4', C-5'
		1.95 (<i>eq</i>)	$J_{2',3'eq} = 3.2$, $J_{3'eq,4'} = 4.7$	C-1', C-2', C-4', C-5'
4'	68.4	3.51	$J_{4',5'} = 9.4$	C-3', C-5', C-6'
5'	71.2	3.63		C-1', C-3', C-4', C-6'
6'	18.1	1.22	$J_{5',6'} = 6.2$	C4', C-5'

Example 38—Synthesis of Ascaroside bhas #22

[0262] Ascaroside bhas#22 was prepared as shown in Scheme 5 below.

Scheme 5





[0263] To prepare **12**, a solution of 2,4-di-*O*-benzoyl-ascarylose-1-(2,2,2-trichloroacetimide) (**1**, 132 mg, 263 μ mol, Butcher et al., *Nat. Chem. Biol.* 3:420–22 (2007), which is hereby incorporated by reference in its entirety) in dry DCM (3 ml) at 0°C was treated with (8*R*)-hydroxynon-1-ene (**11**, 85.2 mg, 600 μ mol, Ferrie et al., *Synlett* 18:2891–93 (2007), which is hereby incorporated by reference in its entirety) and trimethylsilyloxy triflate (5 μ l). After 3 hours, the solution was washed with saturated aqueous NaHCO₃ solution (0.5 ml), dried over Na₂SO₄, and concentrated in vacuum. Flash column chromatography on silica using a gradient of 10–30% ethyl acetate in hexanes afforded (8*R*)-(3'*R*,5'*R*-Dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)non-1-ene (**12**) (71.0 mg, 148 μ mol, 56%) as a colorless oil. ¹H NMR (400 MHz, acetone-*d*₆): δ (ppm) 1.20 (d, *J* = 6.1 Hz, 3H), 1.27 (d, *J* = 6.3 Hz, 3H), 1.33–1.72 (m, 8H), 2.09 (m, 2H), 2.23 (ddd, *J* = 13.5 Hz, *J* = 11.4 Hz, *J* = 3.2 Hz, 1H), 2.47 (m, 1H), 3.91 (m, 1H), 4.20 (dq, *J* = 9.6 Hz, *J* = 6.1 Hz, 1H), 4.93 (ddt, *J* = 10.2 Hz, *J* = 2.2 Hz, *J* = 1.3 Hz, 1H), 5.01 (s.br, 1H), 5.02 (ddt, *J* = 17.1 Hz, *J* = 2.2 Hz, *J* = 1.6 Hz, 1H), 5.13 (m, 1H), 5.16 (ddd, *J* = 11.3 Hz, *J* = 9.8 Hz, *J* = 4.6 Hz, 1H), 5.84 (ddt, *J* = 17.1 Hz, *J* = 10.3 Hz, *J* = 6.8 Hz, 1H), 7.50–7.60 (m, 4H), 7.63–7.71 (m, 2H), 8.04 (m, 2H), 8.12 (m, 2H); ¹³C NMR (100 MHz, acetone-*d*₆): δ (ppm) 18.3, 19.5, 26.3, 29.7, 29.7, 30.4, 34.4, 37.8, 67.7, 71.5, 72.1, 72.9, 94.4, 114.8, 129.4, 129.5, 130.2, 130.4, 131.0, 131.0, 134.1, 134.2, 139.8, 165.9, 166.0. See Figures 22A–B.

[0264] To prepare **13**, a solution of **12** (38 mg, 80 μ mol) and **7** (65 mg, 240 μ mol, Evans & Andrews, *Angew. Chem. Int. Ed.* 47:5426–29 (2008), which is hereby incorporated by reference in its entirety) in dry DCM (2 ml) was treated with 1,4-benzoquinone (1 mg, 8 μ mol) in DCM (0.5 ml) and stirred for 10 minutes. A solution of Grubbs 2nd generation catalyst (3 mg, 4 μ mol) in DCM (0.5 ml) was added. The resulting mixture was stirred at 40°C. After 20 hours, the mixture was filtered over a small layer of silica and concentrated in vacuum. Flash column chromatography on silica using a 5:1 mixture of hexanes and ethyl acetate afforded Ethyl (12*R*)-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-*tert*-butyldimethylsilyloxytridec-5-enoate (**13**) (17.0 mg, 23 μ mol, 29%) as a colorless oil. ¹H NMR (400 MHz, chloroform-*d*₁): δ (ppm) 0.03 (s, 3H), 0.06 (s, 3H), 0.86 (s, 9H), 1.19 (d, *J* = 6.2 Hz, 3H), 1.25 (t, *J* = 7.1 Hz, 3H), 1.28 (d, *J* = 6.3 Hz, 3H), 1.32–1.44 (m, 4H), 1.44–1.52 (m, 2H), 1.61–1.68 (m, 2H), 1.99–2.06 (m, 2H), 2.17–2.22 (m, 3H), 2.38–2.43

(m, 3H), 3.84 (m, 1H), 4.06-4.18 (m, 4H), 4.95 (s, 1H), 5.14 (s.br, 1H), 5.18 (dt, $J = 4.2$ Hz, $J = 10.6$ Hz, 1H), 5.39 (dt, $J = 15.2$ Hz, $J = 6.8$ Hz, 1H), 5.47 (dt, $J = 15.2$ Hz, $J = 6.3$ Hz, 1H), 7.43-7.49 (m, 4H), 7.56-7.61 (m, 2H), 8.04 (d, $J = 7.4$ Hz, 2H), 8.11 (d, $J = 7.2$ Hz, 2H). See Figure 23.

[0265] To prepare **14**, a solution of **13** (14.2 mg, 18.9 μmol) in methanol (1.5 ml) was treated with Pd/C and hydrogenated for 24 hours. The mixture was filtered and concentrated in vacuum to afford Ethyl (12*R*)-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-*tert*-butyldimethylsilyloxytridecanoate (**14**) (12.8 mg, 17.0 μmol , 90%) as a colorless oil. ^1H NMR (600 MHz, chloroform- d_1): δ (ppm) 0.03 (s, 3H), 0.05 (s, 3H), 0.86 (s, 9H), 1.19 (d, $J = 6.1$ Hz, 3H), 1.25 (t, $J = 7.1$ Hz, 3H), 1.28 (d, $J = 6.3$ Hz, 3H), 1.29-1.40 (m, 10H), 1.42-1.53 (m, 4H), 1.58-1.68 (m, 2H), 2.21 (ddd, $J = 14.0$ Hz, $J = 11.7$ Hz, $J = 2.8$ Hz, 1H), 2.37-2.45 (m, 3H), 3.84 (m, 1H), 4.08-4.15 (m, 4H), 4.95 (s, 1H), 5.14 (s.br, 1H), 5.18 (dt, $J = 4.2$ Hz, $J = 10.6$ Hz, 1H), 7.43-7.49 (m, 4H), 7.56-7.61 (m, 2H), 8.04 (d, $J = 7.4$ Hz, 2H), 8.11 (d, $J = 7.2$ Hz, 2H). See Figure 24.

[0266] To prepare **15**, a solution of **14** (19.5 mg, 26.8 μmol) in acetonitrile (1 ml) was treated with 40% aqueous hydrofluoric acid (10 μl) in acetonitrile (100 μl). After stirring for 1 hour, the solution was treated with NaHCO_3 (100 mg) for 15 minutes, dried over Na_2SO_4 , and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–80% ethyl acetate in hexanes afforded Ethyl (12*R*)-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxytridecanoate (**15**) (12.0 mg, 19.6 μmol ; 73%) as a colorless oil. ^1H NMR (501 MHz, chloroform- d_1): δ (ppm) 1.19 (d, $J = 6.1$ Hz, 3H), 1.27 (t, $J = 7.2$ Hz, 3H), 1.28 (d, $J = 6.4$ Hz, 3H), 1.30-1.55 (m, 14H), 1.60-1.68 (m, 2H), 2.21 (ddd, $J = 13.5$ Hz, $J = 11.6$ Hz, $J = 3.1$ Hz, 1H), 2.38 (dd, $J = 16.4$ Hz, $J = 9.2$ Hz, 1H), 2.41 (m, 1H), 2.49 (dd, $J = 16.3$ Hz, $J = 3.1$ Hz, 1H), 3.84 (m, 1H), 3.99 (m, 1H), 4.12 (dq, $J = 9.8$ Hz, $J = 6.2$ Hz, 1H), 4.17 (q, $J = 7.1$ Hz, 2H), 4.95 (s, 1H), 5.15 (s.br, 1H), 5.18 (ddd, $J = 11.2$ Hz, $J = 9.9$ Hz, $J = 4.5$ Hz, 1H), 7.43 – 7.49 (m, 4H), 7.56 – 7.61 (m, 2H), 8.04 (m, 2H), 8.11 (m, 2H). See Figure 25.

[0267] To prepare bhas#22, a solution of **15** (12 mg, 19.6 μmol) in THF (1 ml) was treated with LiOH (15 mg) in water (200 μl) and 1,4-dioxane (2 ml) at 67°C for 3 hours. The reaction mixture was acidified with acetic acid (100 μl), concentrated in vacuum, treated with methanol (2 ml), and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–30% methanol in DCM with 0.2% acetic acid afforded (12*R*)-(3'*R*,5'*R*-

Dihydroxy-6'-*S*-methyl-(2*H*)-tetrahydropyran- 2'-yloxy)-(3*R*)-hydroxytridecanoic acid (bhas#22) (7.3 mg, 19.4 μ mol; 99 %) as a colorless oil.

[0268] ^1H (600 MHz), ^{13}C (151 MHz), and HMBC NMR spectroscopic data for bhas#22 were obtained using methanol- d_4 and are shown in Table 5 below. Chemical shifts were referenced to (CD_2HOD) = 3.31 ppm and (CD_2HOD) = 49.05 ppm. See Figures 26A–D.

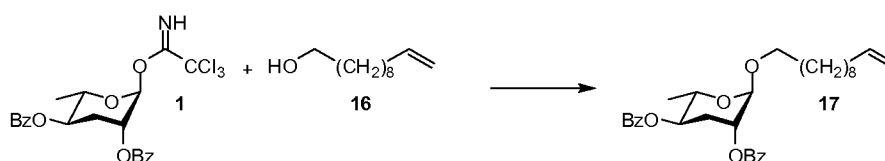
Table 5. NMR spectroscopic data of bhas#22

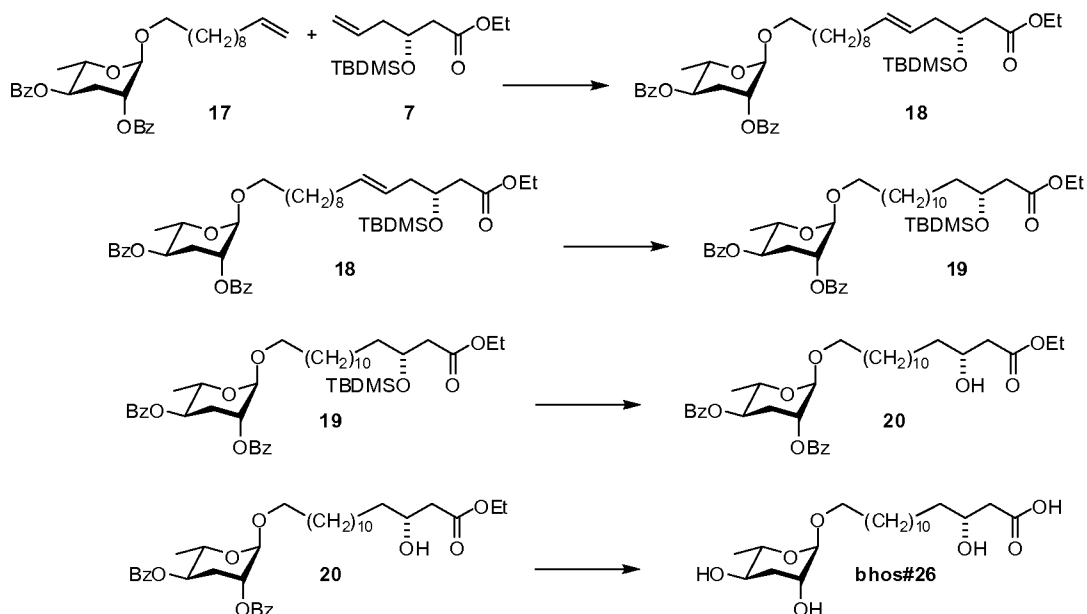
Position	$\delta^{13}\text{C}$ [ppm]	$\delta^1\text{H}$ [ppm]	^1H - ^1H -coupling constants [Hz]	Relevant HMBC correlations
1	176.4			
2	43.4	2.34, 2.42	$J_{2,2} = 15.2$, $J_{2,3} = 8.1$	C-1, C-3, C-4
3	69.2	3.95		C-4, C-5
4	37.8	1.45		
5-10	26.4 - 30.4	1.26-1.39		
11	38.0	1.40-1.50		C9, C10, C12, C13
12	72.2	3.78		C1', C-9, C-10, C11
13	18.9	1.12	$J_{12,13} = 6.2$	C-11, C-12
1'	97.2	4.64		C-2', C-3', C-5', C-12
2'	69.6	3.71		C-3', C-4'
3'	35.5	1.77 (<i>ax</i>)	$J_{3'\text{ax},3'\text{eq}} = 13.1$, $J_{3'\text{ax},4'} = 11.4$, $J_{2',3'\text{ax}} = 3.0$	C-2', C-4', C-5'
		1.95 (<i>eq</i>)	$J_{2',3'\text{eq}} = 3.2$, $J_{3'\text{eq},4'} = 4.7$	C-1', C-2', C-4', C-5'
4'	68.0	3.52	$J_{4',5'} = 9.6$	C-1', C-3', C-6'
5'	70.9	3.63	$J_{5',6'} = 6.3$	C-1', C-3', C-4', C-6'
6'	17.8	1.21		C4', C-5'

Example 39—Synthesis of Ascarolside bhos#26

[0269] Ascaroside bhos#26 was prepared as shown in Scheme 6 below.

Scheme 6





[0270] To prepare **17**, a solution of 2,4-di-*O*-benzoyl-ascarylose-1-(2,2,2-trichloroacetimide) (**1**, 132 mg, 263 μ mol, Butcher et al., *Nat. Chem. Biol.* 3:420–22 (2007), which is hereby incorporated by reference in its entirety) in dry DCM (3 ml) at 0°C was treated with 11-hydroxyundec-1-ene (**16**, 102 mg, 600 μ mol) and trimethylsilyloxy triflate (5 μ l). After 3 hours, the solution was washed with saturated aqueous NaHCO₃ solution (0.5 ml), dried over Na₂SO₄, and concentrated in vacuum. Flash column chromatography on silica using gradient of 10–30% (v/v) ethyl acetate in hexanes afforded 11-(3',5'-Dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)undec-1-ene (**17**) (92.3 mg, 182 μ mol, 69%) as a colorless oil. ¹H NMR (400 MHz, acetone-*d*₆): δ (ppm) 1.28 (d, *J* = 6.2 Hz, 3H), 1.30–1.49 (m, 11H), 1.63–1.72 (m, 2H), 2.03 (m, 2H), 2.22 (ddd, *J* = 13.5 Hz, *J* = 11.3 Hz, *J* = 3.2 Hz, 1H), 2.46 (m, 1H), 3.55 (dt, *J* = 9.7 Hz, *J* = 6.5 Hz, 1H), 3.80 (dt, *J* = 9.8 Hz, *J* = 6.7 Hz, 1H), 4.13 (dq, *J* = 9.8 Hz, *J* = 6.3 Hz, 1H), 4.86 (s.br, 1H), 4.90 (ddt, *J* = 10.2 Hz, *J* = 2.2 Hz, *J* = 1.3 Hz, 1H), 5.98 (ddt, *J* = 17.1 Hz, *J* = 2.2 Hz, *J* = 1.6 Hz, 1H), 5.15 (ddd, *J* = 11.3 Hz, *J* = 9.8 Hz, *J* = 4.6 Hz, 1H), 5.18 (m, 1H), 5.80 (ddt, *J* = 17.1 Hz, *J* = 10.3 Hz, *J* = 6.8 Hz, 1H), 7.49–7.59 (m, 4H), 7.62–7.71 (m, 2H), 8.04 (m, 2H), 8.11 (m, 2H); ¹³C NMR (100 MHz, acetone-*d*₆): δ (ppm) 18.28, 26.88, 29.67, 29.83, 30.10, 30.16, 30.28, 34.46, 67.43, 68.25, 71.43, 71.53, 97.00, 114.66, 129.42, 129.48, 130.23, 130.35, 130.95, 130.96, 134.08, 134.19, 139.78, 165.89, 165.91. See Figures 27A–B.

[0271] To prepare **18**, a solution of **17** (30.8 mg, 60 μ mol) and **7** (50 mg, 180 μ mol, Evans & Andrews, *Angew. Chem. Int. Ed.* 47:5426–29 (2008), which is hereby incorporated by reference in its entirety) in dry DCM (2 ml) was treated with 1,4-benzoquinone (1 mg, 8

μmol) in DCM (0.5 ml) and stirred for 10 minutes. A solution of Grubbs 2nd generation catalyst (3 mg, 4 μmol) in DCM (0.5 ml) was added. The resulting mixture was stirred at 40°C. After 20 hours, the mixture was filtered over a small layer of silica and concentrated in vacuum. Flash column chromatography on silica using a 5:1 mixture of hexanes and ethyl acetate afforded Ethyl 15-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-*tert*-butyldimethylsilyloxypentadec-5-enoate (**18**) (14.2 mg, 18.9 μmol , 31%) as a colorless oil. ¹H NMR (400 MHz, chloroform-*d*₁): δ (ppm) 0.03 (s, 3H), 0.06 (s, 3H), 0.86 (s, 9H), 1.25 (t, *J* = 7.2 Hz, 3H), 1.30 (d, *J* = 6.2 Hz, 3H), 1.27-1.43 (m, 12H), 1.61-1.68 (m, 2H), 1.95-2.02 (m, 2H), 2.16-2.25 (m, 2H), 2.35-2.46 (m, 3H), 3.50 (dt, *J* = 9.6 Hz, *J* = 6.5 Hz, 1H), 3.76 (dt, *J* = 9.6 Hz, *J* = 6.8 Hz, 1H), 4.04-4.18 (m, 4H), 4.82 (s, 1H), 5.18 (ddd, *J* = 11.2 Hz, *J* = 9.9 Hz, *J* = 4.7 Hz, 1H), 5.21 (s.br, 1H.), 5.37 (dt, *J* = 15.2 Hz, *J* = 7.0 Hz, 1H), 5.45 (dt, *J* = 15.3 Hz, *J* = 6.9 Hz, 1H), 7.43-7.50 (m, 4H), 7.56-7.61 (m, 2H), 8.04 (m, 2H), 8.11 (m, 2H). See Figure 28.

[0272] To prepare **19**, a solution of **18** (14.2 mg, 18.9 μmol) in methanol (1.5 ml) was treated with Pd/C (10 mg, 10%, w/w) and hydrogenated for 24 hours. The mixture was filtered and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–80% ethyl acetate in hexanes afforded Ethyl 15-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-*tert*-butyldimethylsilyloxypentadecanoate (**19**) (12.8 mg, 17.0 μmol , 90%) as a colorless oil. ¹H NMR (400 MHz, chloroform-*d*₁): δ (ppm) 0.03 (s, 3H), 0.06 (s, 3H), 0.86 (s, 9H), 1.25 (t, *J* = 7.2 Hz, 3H), 1.30 (d, *J* = 6.2 Hz, 3H), 1.25-1.36 (m, 14H), 1.36-1.42 (m, 2H), 1.45-1.51 (m, 2H), 1.61-1.68 (m, 2H), 2.21 (ddd, *J* = 14.3 Hz, *J* = 11.4 Hz, *J* = 3.2 Hz, 1H), 2.35-2.46 (m, 3H), 3.50 (dt, *J* = 9.6 Hz, *J* = 6.5 Hz, 1H), 3.76 (dt, *J* = 9.6 Hz, *J* = 6.8 Hz, 1H), 4.04-4.18 (m, 4H), 4.82 (s, 1H), 5.18 (ddd, *J* = 11.2 Hz, *J* = 9.9 Hz, *J* = 4.7 Hz, 1H), 5.20 (s.br, 1H.), 7.43-7.50 (m, 4H), 7.56-7.61 (m, 2H), 8.04 (m, 2H), 8.11 (m, 2H). See Figure 29.

[0273] To prepare **20**, a solution of **19** (9.2 mg, 12.2 μmol) in acetonitrile (2 ml) was treated with 40% hydrofluoric acid (20 μl). After stirring for 1 hour, the solution was treated with NaHCO₃ (100 mg) for 15 minutes, dried over Na₂SO₄, and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–80% ethyl acetate in hexanes afforded Ethyl 15-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxypentadecanoate (**20**) (4.4 mg, 6.9 μmol ; 57%). ¹H NMR (400 MHz, chloroform-*d*₁): δ (ppm) 1.27 (t, *J* = 7.2 Hz, 3H), 1.30 (d, *J* = 6.2 Hz, 3H), 1.25-1.36 (m, 16H), 1.36-1.42 (m, 2H), 1.45-1.51 (m, 2H), 1.61-1.68 (m, 2H), 2.21 (ddd, *J* = 14.3 Hz, *J* = 11.4 Hz, *J* = 3.2 Hz,

1H), 2.39 (dd, $J = 16.4$ Hz, $J = 9.0$ Hz, 1H), 2.41 (m, 1H), 2.50 (dd, $J = 16.4$ Hz, $J = 3.0$ Hz, 1H), 3.50 (dt, $J = 9.6$ Hz, $J = 6.5$ Hz, 1H), 3.76 (dt, $J = 9.6$ Hz, $J = 6.8$ Hz, 1H), 3.99 (m, 1H), 4.07 (dq, $J = 10.0$ Hz, $J = 6.2$ Hz, 1H), 4.17 (q, $J = 7.1$ Hz, 2H), 4.82 (s, 1H), 5.18 (ddd, $J = 11.2$ Hz, $J = 9.9$ Hz, $J = 4.7$ Hz, 1H), 5.20 (s.br, 1H), 7.43-7.50 (m, 4H), 7.56-7.61 (m, 2H), 8.04 (m, 2H), 8.11 (m, 2H). See Figure 30.

[0274] To prepare bhos#26, a solution of **20** (4.4 mg, 6.9 μ mol) in THF (1 ml) was treated with a solution of LiOH (5 mg) in water (200 μ l) and 1,4-dioxane (2 ml) and stirred at 67°C. After 3 hours, the solution was acidified with glacial acetic acid (50 μ l) and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–50% methanol in DCM with 0.2% acetic acid afforded 15-(3'*R*,5'*R*-Dihydroxy-6'-methyl-(2*H*)-tetrahydropyran-2'-yloxy)-(3*R*)-hydroxypentadecanoic acid (bhos#26) (2.3 mg, 5.7 μ mol; 83%).

[0275] ^1H (600 MHz), ^{13}C (151 MHz), and HMBC NMR spectroscopic data for bhos#26 were obtained using methanol- d_4 and are shown in Table 6 below. Chemical shifts are referenced to (CD_2HOD) = 3.31 ppm and (CD_2HOD) = 49.05 ppm. See Figures 31A–D.

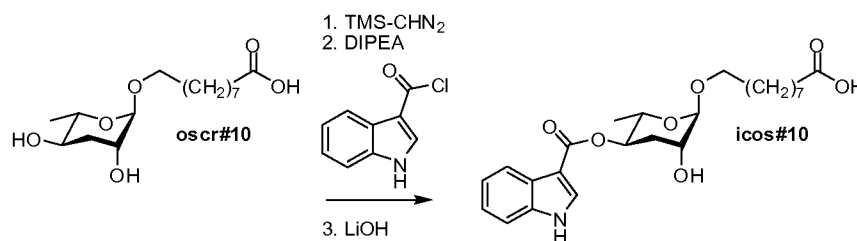
Table 6. NMR spectroscopic data of bhos#26

Position	$\delta^{13}\text{C}$ [ppm]	$\delta^1\text{H}$ [ppm]	^1H - ^1H -coupling constants [Hz]	Relevant HMBC correlations
1	180.0			
2	45.0	2.23, 2.33	$J_{2,2} = 16.0$, $J_{2,3} = 7.0$	C-1, C-3, C-4
3	70.0	3.88		C-4, C-5
4	37.7	1.44, 1.59		
5-12	30.40-30.50	1.30-1.59		
13	27.2	1.37		
14	30.5	1.57		
15	68.0	3.41, 3.68	$J_{15,15} = 9.6$ Hz, $J_{15,14} = 6.3$	C-1', C-14,
1'	100.0	4.49		C-2', C-3', C-5', C-15
2'	69.1	3.76		C-3', C-4'
3'	35.6	1.77 (<i>ax</i>)	$J_{3'ax,3'eq} = 13.1$, $J_{3'ax,4'} = 11.3$, $J_{2',3'ax} = 3.1$	C-2', C-4', C-5'
		1.95 (<i>eq</i>)	$J_{2',3'eq} = 3.2$, $J_{3'eq,4'} = 4.7$	C-2', C-4', C-5'
4'	68.0	3.51	$J_{4',5'} = 9.6$	C-5', C-6'
5'	70.5	3.56	$J_{5',6'} = 6.1$	C-3', C-4', C-6'
6'	17.8	1.23		C4', C-5'

Example 40—Synthesis of Ascaroside icos#10

[0276] Ascaroside icos#10 was prepared as shown in Scheme 7 below.

Scheme 7



[0277] To prepare icos#10, a solution of oscr#10 (12 mg, 39.5 μ mol) in a mixture of methanol (1 ml) and toluene (1 ml) was treated with 2.0 M TMS-diazomethane in diethyl ether (23 μ l, 46 μ mol). After stirring for 30 minutes, excess reagent was quenched by addition of acetic acid (20 μ l) and the solution was concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–10% methanol in DCM afforded the methyl ester (11.3 mg, 35.5 μ mol, 90%) as a colorless solid.

[0278] A solution of the methyl ester in DCM (1 ml) at -20°C was treated with DIPEA (175 μ l, 1 mmol) and indolecarboxylic acid chloride. Indolecarboxylic acid chloride was freshly prepared by treatment of indole-3-carboxylic acid (68 mg, 420 μ mol) in DCM (2 ml) at 0°C with DMF (10 μ l) and SOCl₂ (72 μ l, 840 μ mol). After stirring the reaction mixture for 20 minutes at room temperature, the solution was concentrated in vacuum and DCM (2 ml) was added drop wise. The solution was allowed to reach -7°C and was then quenched with saturated aqueous NaHCO₃ solution (2 ml). The aqueous phase was extracted with DCM (2 ml, three times). The combined organic phases were dried over Na₂SO₄ and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–10% methanol in DCM afforded an isomeric mixture of indole carboxylate esters (8.4 mg, 18.2 μ mol, 51%). The resulting isomeric mixture of indole carboxylate esters was dissolved in THF (1 ml) and treated with a solution of LiOH (2.8 mg, 116 μ mol) in water (0.5 ml) and 1,4-dioxane (2 ml) at 67°C. After stirring for 2 hours, the solution was acidified with acetic acid (30 μ l) and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–20% methanol in DCM containing 0.2% acetic acid afforded an isomeric mixture of icos#10 isomers. HPLC afforded pure samples of 9-(5'*R*-((1*H*)-Indole-3-carboxyloxy)-3'*R*-hydroxy-6'*S*-methyl-tetrahydro-(2*H*)-pyran-2'-yloxy)nonanoic acid (icos#10) (0.6 mg, 1.3 μ mol, 7%) and its isomer (0.3 mg, 0.67 μ mol, 3.7%).

[0279] ^1H (600 MHz), ^{13}C (151 MHz), and HMBC NMR spectroscopic data for icos#10 were obtained using methanol- d_4 and are shown in Table 7 below. Chemical shifts are referenced to (CD_2HOD) = 3.31 ppm and (CD_2HOD) = 49.05 ppm. See Figures 32A–D.

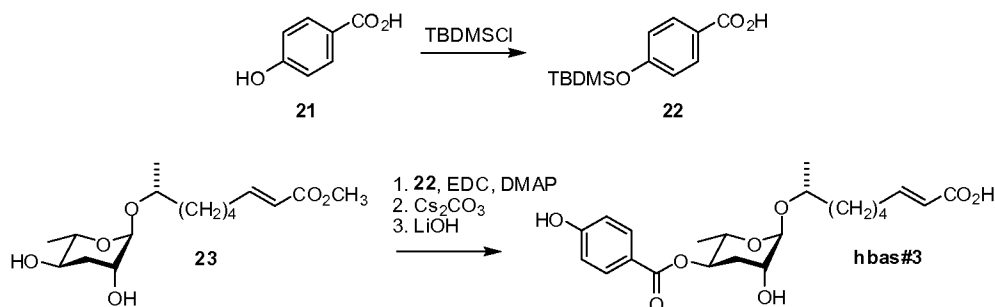
Table 7. NMR spectroscopic data of icos#10

Position	$\delta^{13}\text{C}$ [ppm]	$\delta^1\text{H}$ [ppm]	^1H - ^1H -coupling constants [Hz]	Relevant HMBC correlations
1	176.4			
2	35.5	2.26	$J_{2,3} = 7.5$	C-1, C-3, C-4
3	26.1	1.62		
4-5	30.1	1.38		
6	21.0	1.38		
7	27.1	1.43		
8	30.4	1.65		
9	68.2	3.49, 3.77	$J_{9,9} = 9.7$, $J_{9,8} = 6.5$	C-1'
1'	100.2	4.60		C-3', C-9
2'	68.8	3.85		
3'	33.2	2.02 (<i>ax</i>)	$J_{3'\text{ax},3'\text{eq}} = 13.1$, $J_{3'\text{ax},4'} = 11.3$, $J_{2',3'\text{ax}} = 3.0$	C-4', C-5'
		2.21 (<i>eq</i>)	$J_{3'\text{ax},3'\text{eq}} = 12.9$, $J_{3'\text{eq},4'} = 4.0$	C-1', C-4', C-5'
4'	70.3	5.13	$J_{4',5'} = 9.8$	C-5', C-6', C3''
5'	68.2	4.00	$J_{5',6'} = 6.3$	
6'	17.8	1.25		C4', C-5'
2''	133.1	7.96		C-3'', C3a'', C7a''
3''	108.1			
3''-CO	166.0			
3a''	137.9			
4''	121.5	8.02		C5'', C3a''
5''	123.4	7.20		C4'', C3a''
6''	122.3	7.20		C7'', C7a''
7''	112.7	7.45		C6'', C7a''
7a''	127.0			

Example 41—Synthesis of Ascaroside hbas#3

[0280] Ascaroside hbas#3 was prepared as shown in Scheme 8 below.

Scheme 8



[0281] To prepare **22**, a solution of 4-hydroxybenzoic acid (**21**, 1.52 g, 10 mmol) in DMF (7 ml) was treated with DIPEA (5.2 ml, 30 mmol) and *tert*-butyldimethylsilyl chloride (3.7 g, 24.5 mmol). After 12 hours, the mixture was brought to a pH of 4 by addition of 1 M H_3PO_4 . The mixture was extracted twice with hexanes (15 ml). The organic phase was washed twice with water (15 ml), dried over Na_2SO_4 , and concentrated in vacuum. The residue (3.7 g) was dissolved in THF (10 ml), and treated with water (7 ml) and glacial acetic acid (21 ml). After stirring for 90 minutes, the mixture was added to ice water and extracted twice with a 1:1 mixture (*v/v*) of diethyl ether and hexanes (30 ml). The organic phase was washed with water (30 ml), dried over Na_2SO_4 , and concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–20% methanol in DCM containing 0.2 % acetic acid afforded 4-*tert*-butyldimethylsilyloxybenzoic acid (**22**, 1.42 g, 5.3 mol, 53%) as a white solid. ^1H NMR (400 MHz, chloroform-*d*₁): δ (ppm) 0.24 (s, 6H), 0.99 (s, 9H), 6.89 (d, $J = 8.8$ Hz, 2H), 8.02 (d, $J = 8.8$ Hz, 2H); ^{13}C NMR (100 MHz, chloroform-*d*₁): δ (ppm) - 4.22, 18.40, 25.73, 120.08, 122.39, 132.46, 132.46, 161.01, 172.23. See Figures 33A–B.

[0282] To prepare hbas#3, a solution of ascaroside#3 methyl ester (**23**, 5.7 mg, 18 μmol , Srinivasan et al., *Pub. Lib. Sci. Biol.* 10:e1001237 (2012), which is hereby incorporated by reference in its entirety) in dry DCM (500 μl) was treated with **22** (11.0 mg, 41 μmol), 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDC, 9.0 mg, 47 μmol), and 4-dimethylaminopyridine (DMAP, 6.2 mg, 51 μmol). After stirring for 48 hours, the solution was concentrated in vacuum and treated with H_2O (500 μL). The resulting products were extracted with DCM (1 ml, three times), dried over Na_2SO_4 , and concentrated in vacuum. Flash column chromatography on silica using a gradient of 0–30% methanol in DCM afforded an isomeric mixture of 4-*tert*-butyldimethylsilyloxybenzoyl-ascaroside#3 methyl esters (7.3 mg, 12.9 μmol ; 72%).

[0283] For deprotection of the aromatic *tert*-butyldimethylsilyloxy group (Jiang & Wang, *Tetrahedron Lett.* 44:3859–61 (2003), which is hereby incorporated by reference in its

entirety), a mixture of (4-*tert*-butyldimethylsilyloxybenzoyl)-ascaroside#3 methyl esters (6.0 mg, 10.7 μmol) in DMF (360 μl) was treated with Cs_2CO_3 (1.9 mg, 5.4 μmol) in H_2O (36 μl) and stirred for 3.5 hours. The resulting products were extracted with DCM (1 ml, twice), dried over Na_2SO_4 , and concentrated in vacuum. Flash column chromatography on silica gel using a gradient of 0–30% methanol in DCM afforded an isomeric mixture of 4-hydroxybenzoyl-ascaroside#3 methyl esters (4.5 mg, 10.0 μmol ; 93%). For cleavage of the methyl ester group, a mixture of 4-hydroxybenzoyl-ascaroside#3 methyl esters (4.5 mg, 10.0 μmol) in THF (100 μl) was treated with LiOH (2.3 mg) in H_2O (30 μl) and 1,4-dioxane (500 μl) at 67°C. After 2 hours, the reaction was quenched by addition of glacial acetic acid (50 μl). The solution was concentrated in vacuum. Flash column chromatography on silica using a gradient of 5–20% methanol in DCM with 0.2% acetic acid afforded an isomeric mixture of hbas#3 isomers (1.2 mg, 2.8 μmol , 28%). HPLC afforded pure samples of (8*R*)-(3'*R*-Hydroxy-5'*R*-(4-hydroxybenzoyloxy)-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)non-(2*E*)-enoic acid (hbas#3) (0.6 mg, 1.4 μmol ; 14%) and its isomer (0.6 mg, 1.4 μmol ; 14%).

[0284] ^1H (600 MHz), ^{13}C (151 MHz), and HMBC NMR spectroscopic data for hbas#3 were obtained using methanol- d_4 and are shown in Table 8 below. Chemical shifts are referenced to $(\text{CD}_2\text{HOD}) = 3.31$ ppm and $(\text{CD}_2\text{HOD}) = 49.05$ ppm. See Figures 34A–C.

Table 8. NMR spectroscopic data of hbas#3

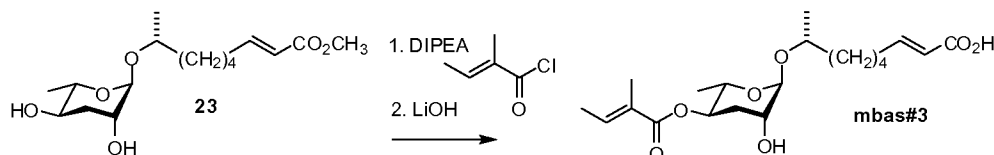
Position	$\delta^{13}\text{C}$ [ppm]	$\delta^1\text{H}$ [ppm]	^1H - ^1H -coupling constants [Hz]	Relevant HMBC correlations
1	171.7			
2	124.4	5.83	$J_{2,3} = 15.6$	C-4
3	149.0	6.90	$J_{3,4} = 7.0$	C-1, C-4, C-5
4	33.1	2.26		C-2, C-3, C5, C-6
5, 6	29.2, 26.5	1.40 – 1.54		C-4, C-6
7	38.1	1.53, 1.62		C-5, C-6, C-8
8	72.7	3.83		C-6, C-7, C-9
9	19.4	1.16	$J_{8,9} = 6.1$	C-7, C-8
1'	97.5	4.73		C-3', C-5', C-8
2'	69.5	3.77		C-4'
3'	33.1	1.95 (<i>ax</i>)	$J_{3'\text{ax},3'\text{eq}} = 12.9$, $J_{3'\text{ax},4'} = 11.2$, $J_{2',3'\text{ax}} = 2.9$	C-4', C-5'
		2.15 (<i>eq</i>)	$J_{2',3'\text{eq}} = 3.2$, $J_{3'\text{eq},4'} = 4.7$	C-2', C-4', C-5'
4'	71.5	5.05	$J_{4',5'} = 9.6$	C-5', C-6', C-7''
5'	68.4	3.98	$J_{5',6'} = 6.3$	C-4', C-6'
6'	18.1	1.18		C4', C-5'
7''-COO	167.2			
1''	122.1			

2'',6''	132.8	7.85	$J = 8.9$	C-1'',C-3'',5'',C-4'', C-7''
3'',5''	116.2	6.83		C-2'',6'', C-4''
4''	163.6			

Example 42—Synthesis of Ascaroside mbas#3

[0285] Ascaroside mbas#3 was prepared as shown in Scheme 9 below.

Scheme 9



[0286] To prepare mbas#3, a solution of ascaroside#3 methyl ester (**23**, 10 mg, 31.4 μmol , Srinivasan et al., *Pub. Lib. Sci. Biol.* 10:e1001237 (2012), which is hereby incorporated by reference in its entirety) in dry DCM (1 ml) at 0°C was treated with DIPEA (110 μl , 630 μmol). (*E*)-2-methylbut-2-enoyl acid chloride (35 μl , 316 μmol) in DCM (0.5 ml) was added drop wise. After stirring at 0°C for 1 hour, the solution was allowed to return to room temperature and treated with saturated aqueous NaHCO_3 solution (0.5 ml). The product was extracted with ethyl acetate, dried over Na_2SO_4 , and concentrated in vacuum. Flash chromatography on silica using a gradient of 5–25% ethyl acetate in hexanes afforded the di-tiglate ester (5.2 mg, 10.8 μmol , 34%) as a yellowish solid.

[0287] The product (4.5 mg, 9.4 μmol) was dissolved in THF (1 ml) and treated with LiOH (0.5 mg, 22 μmol) in water (100 μl) and 1,4-dioxane (2 ml). After stirring at 67°C for 3 hours, the reaction was quenched by addition of glacial acetic acid (100 μl). The solution was concentrated in vacuum. The residue was dissolved in methanol and concentrated in vacuum. Flash column chromatography on silica gel using a gradient of 0–20% methanol in DCM containing 0.2% acetic acid afforded a mixture of mbas#3 isomers (2.1 mg, 5.45 μmol). HPLC provided a pure sample of (8*R*)-(3'*R*-Hydroxy-5'*R*-(*E*)-(2-methylbut-2-enoyloxy)-6'*S*-methyl-(2*H*)-tetrahydropyran-2'-yloxy)non-(2*E*)-enoic acid (mbas#3) (1.2 mg, 3.1 μmol ; 33% yield) identical to the natural product from *C. elegans*.

[0288] ^1H (600 MHz), ^{13}C (151 MHz), and HMBC NMR spectroscopic data for mbas#3 were obtained using methanol- d_4 and are shown in Table 9 below. Chemical shifts are referenced to (CD_2HOD) = 3.31 ppm and (CD_2HOD) = 49.05 ppm. See Figures 35A–B.

Table 9. NMR spectroscopic data of mbas#3

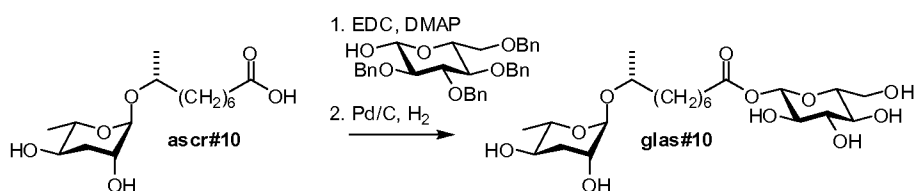
Position	$\delta^{13}\text{C}$ [ppm]	$\delta^1\text{H}$ [ppm]	^1H - ^1H -coupling constants	Relevant HMBC
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			[Hz]	correlations
1	171.7			
2	124.4	5.82	$J_{2,3} = 15.6$	
3	149.0	6.88	$J_{3,4} = 7.0$	
4	33.0	2.23		C-5, C-6
5, 6	29.3, 26.4	1.42 – 1.55		
7	37.9	1.60 1.52		
8	72.4	3.81		
9	18.9	1.14	$J_{8,9} = 6.1$	C-7, C-8
1'	97.4	4.70		C-3', C-5', C-8'
2'	69.3	3.73		
3'	32.8	1.85 (<i>ax</i>)	$J_{3'ax,3'eq} = 12.9$, $J_{3'ax,4'} = 11.2$, $J_{2',3'ax} = 2.9$	
		2.07 (<i>eq</i>)	$J_{2',3'eq} = 3.2$, $J_{3'eq,4'} = 4.7$	
4'	71.4	4.90	$J_{4',5'} = 9.6$	
5'	68.2	3.89	$J_{5',6'} = 6.1$	
6'	17.8	1.13		C-4', C-5'
1''-COO	168.5			
2''	129.2			
3''	138.8	6.85	$J_{3'',4''} = 7.0$	
4''	14.2	1.81		C-2'', C-3''
5''	11.8	1.82		C-1'', C-2'', C-3''

Example 43—Synthesis of Ascaroside glas#10

[0289] Ascaroside glas#10 was prepared as shown in Scheme 10 below.

Scheme 10



[0290] To prepare glas#10, a solution of ascr#10 (3 mg, 9.9 μ mol) in dry DMF (2 ml) was treated with 2,3,4,6-tetra-*O*-benzyl-D-glucose (11 mg, 20 μ mol), DMAP (12.2 mg, 100 μ mol), and EDC (19.2 mg, 100 μ mol). After stirring at room temperature for 18 hours, the solution was concentrated in vacuum. The residue was treated with aqueous acetic acid (200

μl), concentrated, and purified by flash column chromatography on silica using a gradient of 5–50% methanol in DCM.

[0291] The product was dissolved in ethanol (1 ml), treated with Pd/C (10 mg, 10% Pd (w/w)), and hydrogenated for 24 hours. The mixture was filtered and concentrated in vacuum. HPLC provided pure samples of β-D-glucosyl-ascaroside#10 (1.5 mg, 3.22 μmol, 33%) and α-D-glucosyl-ascaroside#10 (1.2 mg, 2.58 μmol, 26%).

[0292] ¹H (600 MHz), ¹³C (151 MHz), and HMBC NMR spectroscopic data for glas#10 were obtained using methanol-*d*₄ and are shown in Table 10 below. Chemical shifts are referenced to (CD₂HOD) = 3.31 ppm and (CD₂HOD) = 49.05 ppm. See Figures 36A–D.

Table 10. NMR spectroscopic data of glas#10

Position	δ ¹³ C [ppm]	δ ¹ H [ppm]	¹ H- ¹ H-coupling constants [Hz]	Relevant HMBC correlations
1	173.8			
2	34.7	2.41	J _{2,2} = 15.4, J _{2,3} = 7.8	C-1
3	25.5	1.64		C-1, C-2
4		1.37		
5, 6	29.8, 30.1	1.30-1.50		
7	26.6	1.45 – 1.55		
8	72.3	3.78		
9	19.2	1.12	J _{8,9} = 6.1	C-8, C-7
1'	97.4	4.60		C-9, C-3', C-5'
2'	69.8	3.71		
3'	35.8	1.77 (<i>ax</i>)	J _{3'<i>ax</i>,3'<i>eq</i>} = 13.0, J _{3'<i>ax</i>,4'} = 11.4, J _{2',3'<i>ax</i>} = 2.9	
		1.95 (<i>eq</i>)	J _{2',3'<i>eq</i>} = 3.2, J _{3'<i>eq</i>,4'} = 4.7	
4'	68.2	3.52	J _{4',5'} = 9.5	
5'	71.0	3.63	J _{5',6'} = 6.3, J _{5',4'} = 9.3	
6'	18.0	1.22		C-5', C-4'
1''	95.4	5.47	J _{1'',2''} = 8.1	C-1
2''	73.8	3.32	J _{2'',3''} = 9.1	
3''	77.8	3.41	J _{3'',4''} = 9.7	
4''	70.9	3.35	J _{4'',5''} = 9.7	
5''	78.7	3.36		
6''	62.2	3.67	J _{6'',6''} = 12.0, J _{6'',5''} = 4.8	
		3.83	J _{6'',6''} = 12.0, J _{6'',5''} = 1.9	

Example 44—Identification and Quantification of Ascarosides

[0293] For the identification of ascarosides detected in wild type and mutants (ascr, oscr, bhas, bhos, icas, icos, ibha, ibho and glas), HPLC-retention times were plotted versus *m/z* (or chain length). (For HPLC-EMI-MS data, see Figure 37A ((ω -1)-oxygenated ascarosides (ascr)), Figure 37B ((ω)-oxygenated ascarosides (oscr)), Figure 37C ((ω -1)-oxygenated β -hydroxyascarosides (bhas)), Figure 37D ((ω)-oxygenated β -hydroxyascarosides (bhos)), Figure 37E ((ω -1)-oxygenated indole ascarosides (icas)), Figure 37F ((ω)-oxygenated indole ascarosides (icos)), Figure 37G ((ω -1)-oxygenated indole β -hydroxy ascarosides (ibha)), Figure 37H ((ω)-oxygenated indole β -hydroxy ascarosides (ibho)), Figure 37I (glucosyl ascaroside esters (glas)), Figure 37J (ascr#8, and 4-(4-hydroxybenzoyl)- and 4-(2-(*E*)-methyl-2-butenoyl)-ascarosides (hbas and mbas)), and Figure 37K (ascr#2 and ascr#6.1).) Components belonging to a homologous series exhibited almost linear elution profiles (Figure 38), indicating that components within a series share the same relative stereochemistry. The structure and stereochemistry of the various series were then identified based on the (1) isolation of representative examples and NMR analysis (for example see Figures 39A–B, 40A–B, 41A–B, 42, and 43A–B), (2) comparison of representative examples with synthetic standards, (3) molecular formula as established from high-resolution MS, (4) characteristic MS/MS fragmentation (see Figures 44A–P), and (5) HPLC-retention times that matched retention time values extrapolated from those of the synthetic samples. The (*E*)-configuration of α,β -unsaturated ascarosides was established by comparison with ascr#3, (*Z*)-configured ascr#3, and ascr#7, and was also suggested by the stereoselectivity of acyl-CoA-oxidase (ACOX) activity. The (3*R*)-stereochemistry of the β -hydroxyascarosides (bhas and bhos series) was deduced from comparison with synthetic standards of bhas#10, bhas#22, and bhos#26 as representative examples, and was also suggested from the sequence homology of MAOC-1 and DHS-28 with (*R*)-selective MFE-2 (Figure 45).

[0294] Quantification of ascarosides was performed by integration of LC-MS signals from the corresponding ion traces. Ascaroside concentrations were calculated using response factors determined for synthetic standards of ascr#1, ascr#3, ascr#5, ascr#7, ascr#9, ascr#10, oscr#9, oscr#10, bhas#22, bhos#26, icas#3, icas#9, icos#10, and glas#10. For most compounds, mass spectrometer response was roughly linear (less than 10% error) for amounts of 1 pmol to 2 nmol per injection. Response factors for ascarosides that were not

synthesized were extrapolated based on data observed for the available standards. Generally, strong differences were observed between the response factors of short-chained members of each series (side chains less than C7), whereas differences between response factors of longer-chained homologs were small. Since not all short-chained members of all series were synthesized, the systematic errors of the absolute amounts reported for some short-chain ascarosides could be larger than for longer-chained compounds.

[0295] In order to roughly account for culture duration and worm biomass, ascaroside content of the excretome and worm pellet samples were recorded in fmol ascarosides produced per hour of culture time per mg of worm pellet dry weight. All quantitative data reported in Figures 37–62 were derived from at least two independent biological repeats.

Example 45—Spot Attraction Assays

[0296] Attraction assays with hbas#3 were conducted as previously reported (Srinivasan et al., *Nature* 454:1115–18 (2008); Pungaliya et al., *Proc. Natl. Acad. Sci. U.S.A.* 106:7708–13 (2009), which are hereby incorporated by reference in their entirety). For the attraction assays, 50–60 hermaphrodite worms were harvested daily at the early fourth larval stage (L4) and stored at 20°C overnight to be used as young adults the following day. Hbas#3 was dissolved in water containing 10% ethanol. Aliquots were stored at -20 °C in 20 µL tubes. 10% ethanol in water was used as control.

Example 46—Quadrant Assays for Measuring Chemotaxis

[0297] Chemotaxis to hbas#3 was assessed on 10 cm four-quadrant petri plates (Wicks et al., *Dev. Biol.* 221:295–307 (2000), which is hereby incorporated by reference in its entirety). Each quadrant was separated from adjacent ones by plastic spacers. Pairs of opposite quadrants were filled with nematode growth medium (NGM) agar containing different concentrations of hbas#3. Animals were washed gently in an S-basal buffer and placed in the center of a four-quadrant plate with ascarosides in alternating quadrants, and scored after 15 minutes and 30 minutes. A chemotaxis index was calculated as: (the number of animals on ascaroside quadrants minus the number of animals on buffer quadrants)/(total number of animals).

Example 47—Statistical Analyses

[0298] Unpaired student's *t* tests with Welch's correction were used for comparing ascaroside profiles between wild-type and mutant metabolomes and for comparing attraction of hermaphrodites on hbas#3 (*: $P < 0.05$, **: $P < 0.001$, ***: $P < 0.0001$). One-factor ANOVA

followed by Dunnett's post-test (*: $P < 0.05$, **: $P < 0.01$) was used for comparing the chemotaxis indices of the different concentrations of hbas#3.

Example 48—LC-MS/MS Reveals New Ascarosides in Wild Type *C. elegans*

[0299] It was desirable to develop an analysis method that would (1) facilitate sensitive detection and quantitation of the known ascarosides in the metabolomes of different *C. elegans* strains and mutants and (2) aid with the discovery of new ascaroside derivatives.

[0300] Because of the vast complexity of the *C. elegans* metabolome, HPLC-MS analysis of metabolite extracts results in extremely crowded chromatograms that are difficult to interpret (Figure 46A). However, it seemed likely that structurally related ascarosides would exhibit characteristic MS/MS fragmentation patterns, which provide a much more selective and sensitive means for their detection. Therefore, ESI-MS/MS fragmentation of a series of synthetic ascarosides was investigated.

[0301] It was found that with negative-ion electrospray ionization (ESI⁻), ascarosides gave rise to an intense and highly diagnostic product ion at m/z 73.02939 [$C_3H_5O_2$], which originated from the ascarylose unit (Figures 44A–P and 46B). This detection method proved suitable for all known ascarosides, except for ascr#2 and ascr#4, which do not ionize well under ESI⁻ conditions. For detection of ascarosides that ionize only under ESI⁺ conditions, neutral loss of 130.0663 amu [$C_6H_{10}O_3$] due to cleavage of the ascarylose unit was monitored. However, it was found that ESI⁺ MS/MS detection of ascarosides was generally less sensitive than ESI⁻ MS/MS detection, as ascarosides fragment less predictably under ESI⁺ conditions. Tests were conducted then to determine whether a screen for precursor ions of m/z 73 could be used to detect known as well as yet unidentified ascarosides in the *C. elegans* wild-type metabolome. Liquid culture metabolite extracts, which contain accumulated excreted metabolites from large numbers of worms (the worm “excretome”), were used. The resulting HPLC-MS/MS chromatograms showed a large number of well-resolved peaks, most of which were found to represent ascarosides, including several families of previously undetected compounds.

[0302] The identities of the known ascarosides were confirmed using synthetic standards. In addition, it was found that the known saturated ascarosides ascr#1, ascr#9, and ascr#10 are accompanied by substantial quantities of homologs with six- to fifteen-carbon side chains (Figures 46C and 47A). Identification of this homologous series was based on high-resolution MS/MS, LC-retention times, and the synthesis of representative members (see Example 44 and Figure 38]). The LC-MS/MS screen further revealed that ascarosides

with side chains of 5 to 11 carbons were accompanied by smaller quantities of slightly less polar isomers. These ascaroside isomers were produced more abundantly by *acox-1* mutant worms.

[0303] Several additional peaks in the wild-type MS/MS chromatograms could not be assigned to any of the known ascaroside classes. For two of these compounds, MS/MS product ions at m/z 301.1651 [$C_{15}H_{25}O_6$] suggest that they represent ascr#3 derivatives. The putative ascr#3 derivatives were isolated by preparative HPLC and analyzed using 2D NMR spectroscopy (dqfCOSY, see Figures 39A–B, 40A–B, and 41A–B), which confirmed that these compounds were ascr#3-based and further indicated the presence of a 4-hydroxybenzoyl or (*E*)-2-methyl-2-butenoyl (tigloyl) moiety attached to the 4-position of the ascarylose (Figure 47C). These structural assignments were corroborated via total synthesis of authentic samples (see Examples 41–42). In analogy to the recently reported indole-3-carboxy derivative of ascr#3 (“icas#3”) (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety), the 4-hydroxybenzoyl and (*E*)-2-methyl-2-butenoyl derivatives of ascr#3 were named with SMIDs, “hbas#3” and “mbas#3”, respectively. Hbas#3 and mbas#3 were the first ascarosides to incorporate 4-hydroxybenzoyl and (*E*)-2-methyl-2-butenoyl moieties, which, in analogy to the indole-3-carboxy moiety in icas#3 (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety), could be derived from amino acid precursors. Because of its structural similarity to the aggregation-inducing indole ascarosides (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety), hbas#3 was tested for its effect on worm behavior. It was found that hbas#3 strongly attracted *C. elegans* at concentrations as low as 10 femtomolar (see Figures 48A–B), which exceeds the potency of any previously known *C. elegans* small molecule signal (Srinivasan et al., *Nature* 454:1115–18 (2008); Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which are hereby incorporated by reference in their entirety). Low femtomolar activity is unusual for small molecule signals in animals, but matches that of some classes of peptide hormones (Rittschof & Cohen, *Peptides* 25:1503–16 (2004); Gozes et al., *CNS Drug Rev.* 11:353–68 (2005), which are hereby incorporated by reference in their entirety).

Example 49—Peroxisomal β -Oxidation in Ascaroside Biosynthesis

[0304] Comparative LC-MS/MS was used to investigate ascaroside biogenesis. Studies have indicated that the side chains of the ascarosides are derived from peroxisomal β -

oxidation of longer-chained precursors and that the acyl-CoA oxidase ACOX-1 participates in the first step of ascaroside side chain β -oxidation, introducing α,β -unsaturation (Figures 49A–B) (Joo et al., *J. Biol. Chem.* 285:29319–25 (2010), which is hereby incorporated by reference in its entirety). In vertebrates as well as in *Drosophila*, the next two steps in peroxisomal β -oxidation—hydration of the double bond and subsequent dehydrogenation to the β -ketoacyl-CoA ester are catalyzed by one protein, e.g., the multifunctional enzyme type 2 (MFE-2). These two enzymatic functions appear to be separated in *C. elegans* such that the dehydratase and dehydrogenase are distinct proteins (Figure 45) (Haataja et al., *Biochem. J.* 435:771–81 (2011), which is hereby incorporated by reference in its entirety). Studies have shown that *C. elegans* DHS-28, a protein with homology to the (*R*)-selective β -hydroxyacyl-CoA dehydrogenase domain of human MFE-2, likely participates in converting β -hydroxyacyl-CoA-derivatives into the corresponding β -ketoacyl-CoA intermediates, which are subsequently cleaved by the β -ketoacyl-CoA thiolase DAF-22 (Butcher et al., *PNAS* 106:1875–79 (2009); Joo et al., *Biochem. J.* 422:61–71 (2009), which are hereby incorporated by reference in their entirety). However, it remained unclear which enzyme serves as the enoyl-CoA hydratase that catalyzes the essential second step of the β -oxidation cascade.

[0305] Using the MS/MS-based ascaroside screening method described herein, the ascaroside profiles of *acox-1(ok2257)*, *dhs-28(hj8)*, and *daf-22(ok693)* mutant worms were re-investigated. Additionally, another study indicated that *maoc-1* encoded a peroxisomal 2-enoyl-CoA hydratase (Zhang et al., *PNAS* 107:4640–45 (2010), which is hereby incorporated by reference in its entirety), which was hypothesized to participate in the hydration step of ascaroside β -oxidation. Thus, the excretomes of several other peroxisomal mutants, including *maoc-1(hj13)* worms, were analyzed.

Example 50—Side-Chain Functionalization Precedes β -Oxidation

[0306] LC-MS/MS analysis of the excretome of *acox-1(ok2257)* mutant worms revealed that abundance of the α,β -unsaturated ascr#3, the dominating component of wild-type media, was greatly reduced (Figures 50A–E). This decrease in ascr#3 and other α,β -unsaturated ascarosides did not appear to be the result of overall down regulation in ascaroside production, but instead was accompanied by accumulation of a series of saturated ascarosides. For example, ascr#10, the dihydro-derivative and direct precursor of ascr#3, was 13.6 times more abundant in *acox-1(ok2257)* than in the wild-type excretome. The corresponding homologs with 11- and 13-carbon side chains, ascr#18 and ascr#22, were 29

times and 66 times more abundant in *acox-1* than in the wild-type excretome, respectively. The build-up of longer-chained saturated ascarosides in the *acox-1(ok2257)* excretome confirmed the importance of ACOX-1 in α,β -dehydrogenation of the ascaroside side chain (Figure 49A). Because ascaroside biosynthesis is not abolished in *acox-1(ok2257)* worms, it seems likely that other, yet unidentified ACOX-enzymes contribute to peroxisomal β -oxidation of long chain ascaroside precursors. However, LC-MS/MS analysis of the excretome of several other peroxisomal *acox* mutants (see Examples 29–34 and 44–47 and Figure 51]) revealed largely wild-type like ascaroside profiles.

[0307] Further analysis of the *acox-1(ok2257)* excretome revealed the complete absence of ascr#5, one of the major dauer inducing ascarosides produced abundantly in wild type worms (Figure 52]). Ascr#5 differs from all other previously identified ascarosides in that its side chain is attached to the ascarylose sugar via the terminal carbon (“ ω linkage”), and not the penultimate carbon (“ $\omega-1$ linkage”) (Figure 47B). Instead of ascr#5, large quantities of a new homologous series of saturated ascarosides in *acox-1(ok2257)* were detected, smaller amounts of which were also present in the wild-type excretome (Figure 52]). The most abundant component of this series of isomers was isolated via preparative HPLC, and identified by NMR spectroscopy to be an ω -linked ascaroside with a C₅ side chain (Figures 42 and 47B). This suggests that the additional series of compounds observed in *acox-1* represents a series of ω -linked saturated ascarosides (Figure 37B), which were confirmed by synthesis of C₅ and C₉ ω -linked ascarosides (see Examples 35–36). To differentiate the (ω)-linked ascarosides from their previously described ($\omega-1$)-linked isomers, the newly found (ω)-linked compounds were named with the SMID “oscr”, e.g., the synthesized (ω)-linked isomers of ascr#9 and ascr#10 were named oscr#9 and oscr#10 (Figure 47]).

[0308] Thus, production of (ω)-linked ascr#5 was abolished in *acox-1(ok2257)* worms, whereas production of longer chain homologs with 5–13 carbon side chains, e.g., oscr#9, was starkly upregulated (Figure 52). These results indicate that β -oxidation in *acox-1(ok2257)* worms is strongly dependent on whether the side chain is ($\omega-1$)- or (ω)-functionalized. Chain shortening of ($\omega-1$)-oxygenated substrates appeared to stall at a chain length of 9 carbons as in ascr#10, whereas (ω)-oxygenated substrates were processed for two additional rounds of β -oxidation to afford large quantities of (ω)-oxygenated oscr#9 featuring a 5-carbon side chain. This suggests that side chain oxygenation precedes peroxisomal β -oxidation. In contrast, the time point of ascarylose attachment seemed less certain. The

absence of any (ω -1)- or (ω)-hydroxylated fatty acids in the investigated *C. elegans* mutant metabolome samples suggests a biosynthetic model in which very long-chain ascarosides serve as substrates for peroxisomal β -oxidation. However, it may be possible that β -oxidation occurs prior to ascarylose attachment.

Example 51—MAOC-1 and DHS-28 Are Functional Homologs of Human MFE-2

[0309] In contrast to wild-type and *acox-1(ok2257)* worms, short-chain ($< C_9$) ascarosides were not detected in *maoc-1(hj13)* and *dhs-28(hj8)* worms, which instead accumulated several series of (ω -1)- and (ω)-oxygenated medium and long chain ascarosides ($\geq C_9$). The ascaroside profile of the *maoc-1(hj13)* excretome was dominated by α,β -unsaturated ascarosides such as ascr#21 (C_{13}) and ascr#25 (C_{15}) (Figures 50A–E), supporting the hypothesis that MAOC-1 functions as an enoyl-CoA hydratase in the ascaroside biosynthetic pathway (Figure 49A). In addition, the *maoc-1(hj13)* excretome contained smaller amounts of the corresponding saturated ascarosides, along with a third homologous series of compounds, whose high-resolution mass spectrometry (HRMS) and MS/MS fragmentation suggested that they represent a series of long chain β -hydroxylated ascarosides (Figures 37C–D, 44A–P, and 50A–E). These structural assignments were confirmed via synthesis of representative members of this series (see Examples 37–39), the C_9 and C_{13} β -hydroxylated (ω -1)-ascarosides bhas#10 and bhas#22, as well as the C_{15} β -hydroxylated (ω)-ascaroside bhos#26 (SMIDs for (ω -1)- and (ω)-oxygenated β -hydroxylated ascarosides: “bhas” and “bhos”). Since β -hydroxylated ascarosides are putative products of enoyl-CoA hydratases such as MAOC-1, their presence in the excretomes of both studied *maoc-1* mutant strains, *maoc-1(hj13)*, which carries a point mutation in the active site (D186N), and the 2110 base pair deletion mutant *maoc-1(ok2645)* (Figure 49B) suggests that additional enoyl-CoA hydratases may participate in ascaroside biosynthesis.

[0310] Similar to the results for *maoc-1(hj13)*, the *dhs-28(hj8)* ascaroside profile was found to be dominated by compounds with side chains ranging from C_9 – C_{21} (Figures 50A–E). However, in contrast to *maoc-1(hj13)*, saturated and α,β -unsaturated ascarosides constituted a relatively smaller portion of total ascarosides in *dhs-28(hj8)*. Further, (ω -1)- and (ω)-oxygenated β -hydroxy ascarosides (bhas and bhos) with odd numbered side chains from C_{13} – C_{21} were found to be major components (Figures 50A–E and 53A–E), which is consistent with the proposed biosynthetic role of DHS-28 as a β -hydroxyacyl-CoA dehydrogenase (Butcher et al., *PNAS* 106:1875–79 (2009); Joo et al., *Biochem. J.* 422:61–71 (2009), which are hereby incorporated by reference in their entirety). Analysis of the *daf*-

22(*ok693*) excretome revealed the absence of all ascarosides with side chains shorter than 12 carbons, as reported earlier (Pungaliya et al., *PNAS* 106:7708–13 (2009); Butcher et al., *PNAS* 106:1875–79 (2009); Joo et al., *Biochem. J.* 422:61–71 (2009), which are hereby incorporated by reference in their entirety) (Figures 50A–E and 53A–E). The *daf-22(ok693)* excretome contained small amounts of homologous series of (ω -1)- and (ω)-oxygenated long chain ascarosides featuring saturated (ascr and oscr) and β -hydroxylated side chains (bhas and bhos). In addition, the *daf-22(ok693)* excretome contained very long chain ascarosides (VLCA, $>C_{22}$) with side chain lengths of up to 33 carbons, as reported earlier (Pungaliya et al., *PNAS* 106:7708–13 (2009); Zagoriy et al., *Chem. Biodivers.* 7:2016–22 (2010), which are hereby incorporated by reference in their entirety).

Example 52—Identification of New Indole Ascarosides

[0311] Indole-3-carbonylated ascarosides were much less abundant than the corresponding unfunctionalized ascarosides and have been shown to function as highly potent aggregation signals (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety). LC-MS/MS screening revealed several new types of indole ascarosides (Figure 37E–H and 47C). The results of synthetic samples of icas#3, icas#9, and icas#1 (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety) show that indole ascarosides exhibit a characteristic fragmentation pattern that includes neutral loss of 143 amu [C_9H_5NO] due to cleavage of the indole-3-carbonyl unit, as well as the ascaroside-diagnostic product ion at m/z 73 (see Figures 44A–P). LC-MS/MS screening for components with this fragmentation pattern revealed that known (ω -1)-oxygenated isomers icas#1, icas#9, and icas#10 in *acox-1(ok2257)* are accompanied by a series of (ω)-oxygenated indole ascarosides (SMID: icos#1, icos#9, and icos#10), which was confirmed by chemical synthesis of icos#10 as a representative example (see Example 40). Peroxisomal β -oxidation mutants that do not produce short chain ascarosides (<9 carbon side chains), for example *maoc-1* and *dhs-28* worms, also did not produce any of the corresponding indole ascarosides. Instead, the *maoc-1* and *dhs-28* excretomes contained significant amounts of (ω -1)- and (ω)-oxygenated long chain indole ascarosides (SMIDs icas and icos) and indole β -hydroxy ascarosides (SMIDs ibha and ibho) with side chains from 9–17 carbons (see Figures 37E–H).

Example 53—Indole Ascaroside Biogenesis

[0312] Experiments with deuterium-labeled tryptophan and axenic *in vitro* cultures have shown that the indole-3-carbonyl moiety of indole ascarosides originates from L-tryptophan (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety). A similar L-tyrosine or L-phenylalanine origin seems likely for the 4-hydroxybenzoyl moiety of hbas#3, whereas the tigloyl group of mbas#3 could be derived from L-isoleucine (Attygalle et al., *J. Chem. Ecol.* 33:963–70 (2007), which is hereby incorporated by reference in its entirety). However, it remained unclear at what stage in ascaroside biosynthesis the indole-3-carbonyl moiety is attached.

[0313] Comparison of ascaroside and indole ascaroside profiles revealed that indole ascaroside biosynthesis is tightly controlled. For example, it was found that in *acox-1* mutants, the (ω)-ascaroside ascr#9 was over 100 times more abundant than the (ω -1)-ascaroside ascr#9, whereas (ω -1)-indole ascaroside icas#9 was much more prominent than (ω)-indole ascaroside icos#9 (Figure 54A). Similarly, icas#3 and icas#9, the dominating indole ascarosides in the wild type excretome, were produced in roughly equal amounts, whereas ascr#3 was about 20 times more abundant than ascr#9 (Figure 55). This strong dependence of indole ascaroside biosynthesis on side chain length and (ω -1)- vs. (ω)-oxygenation suggests that these compounds originate from specific attachment of an indole-3-carbonyl unit to the corresponding non-indole ascaroside.

[0314] To test whether non-indole ascarosides serve as precursors for indole ascarosides, *daf-22(m130)* worms (which are devoid of all short-chained indole and non-indole ascarosides) were incubated with a 1:1 mixture of ascr#10 and oscr#10 for 5 days. Subsequent analysis by LC-MS shows partial conversion into icas#10 and icos#10 (Figure 54B), indicating that non-indole ascarosides serve as specific precursors to their corresponding indole derivatives. Moreover, conversion of (ω -1)-ascaroside ascr#10 was preferred over conversion of (ω)-ascaroside oscr#10, reflecting the ratios of these compounds in wild type and *acox-1* mutants. Similarly, *daf-22(m130)* worms were found to convert added ascr#3 into icas#3. Further conversion of indole or non-indole ascarosides into shorter chain derivatives (e.g., ascr#1 or icas#1) was not observed. These results indicate that attachment of an L-tryptophan-derived indole-3-carbonyl unit represents the final step in indole ascaroside biosynthesis.

Example 54—Ascaroside Excretion Is Selective

[0315] Despite detailed investigations of ascaroside function, little was known about how ascarosides are released and transported from their site of biosynthesis. The ascaroside profiles of the wild-type excretomes (liquid culture supernatant extracts) and worm body metabolomes (worm pellet extracts) were compared to identify possible non-excreted ascaroside derivatives and to determine quantitative differences. Ascaroside profiles of worm pellet extracts differed significantly from those excreted into the media, indicating that ascarosides are differentially released by *C. elegans* (Figure 56A). In the worm pellets, the most abundant ascarosides, such as ascr#3 in wild-type and ascr#10 in *acox-1* worms, were accompanied by significantly more polar derivatives, which were absent from the media extracts (Figure 57). MS/MS analysis suggested that these components represent ascaroside *O*-glycoside esters. Isolation of the putative ascr#10 glycoside from *acox-1(ok2257)* worm pellet extracts and subsequent NMR spectroscopic analysis (Figures 43A–B) indicate esterification of ascr#10 with the anomeric hydroxy group of β -glucose, which was subsequently confirmed via synthesis (SMID for 1- β -D-glucosyl ascr#10: glas#10, Figure 56A). The fact that large quantities of highly polar glas#10 and other ascaroside glucosides (see Figure 37I) were retained in the worm bodies but not excreted suggests that they represent transport or storage forms of the ultimately excreted signaling molecules.

[0316] In addition, saturated ascarosides were retained in the worm bodies to a much greater extent than their α,β -unsaturated derivatives (Figure 56A). Differential release was also observed for (ω)-oxygenated components (Figure 58). Therefore, it appears that *C. elegans* exhibits remarkable control over the release of ascaroside signals.

Example 55—Nutritional State Affects Ascaroside Biosynthesis

[0317] Ascaroside biosynthesis has been reported to depend on various environmental factors, including food availability (Butcher et al., *PNAS* 106:1875–79 (2009), which is hereby incorporated by reference in its entirety), developmental stage (Kaplan et al., *Publ. Lib. Sci. ONE* 6:e17804 (2011), which is hereby incorporated by reference in its entirety), and temperature (Joo et al., *J. Biol. Chem.* 285:29319–25 (2010), which is hereby incorporated by reference in its entirety).

[0318] LC-MS was used to investigate the effect of nutritional state on ascaroside biosynthesis by comparing the excretomes of well-fed and starved cultures of wild-type and mutant worms. The results indicate that the ratio of (ω -1) to (ω)-linked ascarosides strongly

depends on nutritional state. Production of long-chain (ω -1)-oxygenated ascarosides in starved cultures of *dhs-28* was about 5 times higher than those of well fed cultures (Figure 56B and 59A–C). Similarly, starved wild-type worms excreted significantly larger amounts of the (ω -1)-linked ascr#3 than well-fed worms, relative to the amounts of (ω)-linked ascr#5 (Figure 60). Both ascr#5 and ascr#3 are major components of the dauer pheromone; however, they affect worm behavior differently (Jeong et al., *Nature* 433:541–45 (2005); Srinivasan et al., *Nature* 454:1115–18 (2008); Butcher et al., *PNAS* 105:14288–92 (2008); Macosko et al., *Nature* 458:1171–75 (2009), which are hereby incorporated by reference in their entirety).

[0319] Therefore, it appears that ascaroside signaling is actively regulated in response to changes in nutrient availability via modulation of (ω -1)- and (ω)-functionalization of very long-chain fatty acids upstream of peroxisomal β -oxidation. Together with the recent finding that (ω)- and (ω -1)-functionalized ascarosides are sensed by different families of G-protein coupled receptors (Kim et al., *Science* 326:994–98 (2009); McGrath et al., *Nature* 477:321–25 (2011), which are hereby incorporated by reference in their entirety), these results suggest that (ω)- and (ω -1)-functionalized ascarosides target separate downstream signaling pathways.

Discussion of Examples 29–55

[0320] Ascarosides play important roles for several different aspects of *C. elegans* biology. This functional diversity is paralleled by corresponding structural diversity and a complex biosynthetic pathway. The MS/MS-based study described in Examples 34–43 and Figures 37A–37K revealed 146 ascarosides, including 124 previously unreported analogs, which show several unexpected features including (ω)-oxygenation of the fatty acid-derived side chains, 4-hydroxybenzoylation or (*E*)-2-methyl-2-butenoylation of the ascarylose unit, and glucosyl esters. Most ascarosides occur as members of homologous series of compounds with (ω -1)- or (ω)-linked saturated, α,β -unsaturated, or β -hydroxylated side chains ranging from 3 to 21 (occasionally more) carbons. Importantly, only a few members of each series are produced abundantly in wild type nematodes, and incorporation of specific structural features such as modification in position 4 of the ascarylose (e.g., as in indole ascarosides) or the incorporation of α,β -unsaturation appears to be tightly controlled.

[0321] Given their assembly from carbohydrate, lipid, and amino acid-derived building blocks, the ascarosides appear as a modular library of small molecule signals that integrate inputs from three basic metabolic pathways (Figure 61A). The ascarosides then transduce input from these pathways via their diverse signaling functions in *C. elegans*.

behavior and development, including dauer formation, mate attraction, hermaphrodite repulsion, and aggregation (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012); Edison, A. S., *Curr. Opin. Neurobiol.* 19:378–88 (2009), which are hereby incorporated by reference in their entirety). Their specific biosyntheses suggest that many of the newly identified ascarosides also contribute to known or as yet undetermined functions in *C. elegans*. As an example, hbas#3 was shown to act as an attraction signal whose potency exceeds that of all previously known small molecules in this model organism.

[0322] A working model for ascaroside biogenesis is proposed in Figure 61B. The finding that ascarosides are members of several homologous series with side chains up to 21 (and more) carbons suggests their origin from peroxisomal β -oxidation of very long chain precursors. Previous studies reported the presence of very long chain ascarosides (VLCA) with C₂₉ and C₃₁ side chains in wild-type and *daf-22* mutants, which could represent precursors or intermediates in ascaroside biosynthesis (Pungaliya et al., *PNAS* 106:7708–13 (2009); Zagoriy et al., *Chem. Biodivers.* 7:2016–22 (2010), which are hereby incorporated by reference in their entirety). Alternatively, very long chain fatty acids (VLCFAs) could undergo peroxisomal β -oxidation prior to (ω -1)- or (ω)-functionalization and subsequent attachment of the ascarylose. The observation that the *acox-1* mutation affected (ω -1)- and (ω)-oxygenated ascarosides differently suggests that (ω -1)- and (ω)-functionalization of VLCFA precursors occur upstream of their breakdown by peroxisomal β -oxidation. Given the large range in side chain lengths, it seems likely that the resulting (ω -1)- and (ω)-hydroxy VLCFAs are linked to ascarylose prior to entering the β -oxidation pathway, though the presence of a promiscuous ascarosyl transferase may also be possible (Figure 61B).

[0323] Chain shortening of VLCA can then progress via repetitive cycles of peroxisomal β -oxidation. The results from the LC-MS/MS screening allows for the proposal of precise roles for enzymes participating in each of the four-step β -oxidation cycle: the acyl-CoA oxidase ACOX-1, enoyl-CoA hydratase MAOC-1, β -hydroxyacyl-CoA dehydrogenase DHS-28, and β -ketoacyl-CoA thiolase DAF-22. Mutations in *acox-1*, *maoc-1*, and *dhs-28* were shown to result in specific changes of the corresponding ascaroside profiles, in agreement with their proposed functions.

[0324] The acyl-CoA oxidase ACOX-1 has been the subject of a previous study that indicates that mutations in *acox-1* primarily affect the biosynthesis of ascr#2 and ascr#3, but not of ascr#1 (Joo et al., *J. Biol. Chem.* 285:29319–25 (2010), which is hereby incorporated by reference in its entirety). However, the results described in Example 50 indicate that *acox-*

l(ok2257) mutants have a reduced ability to process C₉ (ω-1)-functionalized ascarosides, resulting in diminished production of all shorter-chained ascarosides and build-up of C₉ and longer chained saturated ascarosides. Mutations of *maoc-1* (as well as *dhs-28* and *daf-22*) have been shown to result in expansion of intestinal lipid droplets and cause an increase in fasting- and lipolysis-resistant triglycerides (Zhang et al., *PNAS* 10:4640–45 (2010), which is hereby incorporated by reference in its entirety). The results described in Example 51 show that MAOC-1 participates in ascaroside biosynthesis, acting as the previously unidentified enoyl-CoA hydratase. These findings further demonstrate that hydration of enoyl-CoAs and dehydrogenation of β-hydroxyacyl-CoAs in *C. elegans* are catalyzed by two distinct enzymes, MAOC-1 and DHS-28, as had been indicated by their homology to separate functional domains of human MFE-2 (Zhang et al., *PNAS* 10:4640–45 (2010), which is hereby incorporated by reference in its entirety).

[0325] The results described in Examples 52-53 further show that attachment of the tryptophan-derived indole-3-carbonyl unit in indole ascarosides likely represents the last step in their biosynthesis, and that this step is highly specific. As attachment of an indole-3-carbonyl group to ascarosides can dramatically alter their biological function, such tight regulation makes sense. For example, indole-3-carbonyl addition to the dauer-inducing and strongly repulsive signal *ascr#3* results in the potent hermaphrodite attractant *icas#3* (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety).

[0326] The biosynthesis of ascarylose in *C. elegans* has not been investigated. However, the detection of ascarosides in axenic *C. elegans* cultures demonstrates that *C. elegans* produces ascarylose endogenously (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety). Ascarylose biosynthesis in bacteria is well understood and the *C. elegans* genome includes several homologs of bacterial genes in this pathway, for example *ascE* from *Yersinia pseudotuberculosis* (Figure 62) (Thorson et al., *J. Bacteriol.* 176:5483-93 (1994), which is hereby incorporated by reference in its entirety), providing potential entry points for the study of ascarylose biosynthesis and its regulation in nematodes. Moreover, the oxidases catalyzing (ω-1)- or (ω)-functionalization of VLCFA precursors remain to be identified. The finding that the ratio of (ω-1)/(ω)-oxygenated ascarosides in wild type and peroxisomal β-oxidation mutants was strongly affected by starvation indicates that this step may encode information about nutritional state. In addition, ascaroside excretion was shown to be surprisingly

specific. Given the high sensitivity and selectivity of LC-MS/MS, ascaroside profiling using the method described in Examples 29-55 can aid in identifying additional genes and environmental factors that participate in ascaroside biosynthesis and homeostasis.

Example 56—Dauer Production

[0327] *C. elegans* were cultured under dauer inducing liquid culture conditions (Kaplan et al., *Publ. Lib. Sci. ONE* 6:e17804 (2011), which is hereby incorporated by reference in its entirety) in S-complete with 20,000 worms/ml and 0.5% (wet weight) *E. coli* (HB101). Nematodes were incubated at 22°C in a shaker (250 rpm) for 112 hours after feeding L1 larvae nematodes. Thereafter, nematodes were treated with 1% sodium dodecyl sulphate (SDS) for 15 minutes. Surviving nematodes were allowed to be separated from dead nematodes on an agar plate prior to collection. After removal of the dead nematodes by vacuum, dauer animals were collected using M9 buffer and placed at 4°C.

Example 57—Rearing of *S. feltiae*

[0328] *S. feltiae* was ordered from ARBICO Organics (Tucson, AZ). *G. mellonella* larvae (Wax worms, Grubco, Hamilton, OH) were infected with 50 *S. feltiae* dauer juveniles per larvae. After two days, the infected larvae were placed into new 6 cm diameter petri dishes and the white trap method was used to collect infective juveniles (IJs) (LAWRENCE LACEY: MANUAL OF TECHNIQUES IN INSECT PATHOLOGY (1997), which is hereby incorporated by reference in its entirety).

[0329] *S. feltiae* IJs were verified by polymerase chain reaction (PCR) using species specific primers from the ITS rDNA region as described in Campos-Herrera et al., *Ann. Appl. Biol.* 158:55-68 (2011), which is hereby incorporated by reference in its entirety. PCR amplifications were performed in an MJ Research PTC 200 Peltier Thermal Cycler. Amplifications were conducted as described in Campos-Herrera et al., *Ann. Appl. Biol.* 158:55-68 (2011), which is hereby incorporated by reference in its entirety, in a 25 µL final volume containing 1 µL DNA template, using sterile deionized water and DNA prepared from *Steinernema riobrave* as negative controls. Cycling parameters were 94°C for 15 minutes followed by 35 cycles of denaturation at 94°C for 30 seconds, annealing at 59°C for 20 seconds, and extension at 72°C for 20 seconds with a final extension of 72°C for 10 minutes. Amplicon sizes were verified through electrophoresis on a tris-acetate-EDTA (TAE) 2% agarose gel and visualized in the UVP BioDoc-it™ System.

[0330] The *S. feltiae* primer produced specific amplification for all samples containing IJs from lab populations in conventional PCR. The primers showed no amplification for *S. riobrave* and deionized water controls.

Example 58—Rearing of Root Knot Nematodes

[0331] Infected tomato plants were collected from field sites in Florida. Roots were inspected for root knot infection, and root knot nematode eggs were collected as described by in Hussey & Barker, *Plant Dis. Rep.* 57:1025-28 (1973), which is hereby incorporated by reference in its entirety, with modifications. The infected roots were treated with 1% bleach for 2 minutes. Eggs released from egg mass matrices were collected with a nested filter system (85 µm) to collect plant debris and with 25 µm nylon filters (Nytex) to collect eggs. Eggs were washed thoroughly with MILLI-Q water and placed at room temperature for 3 days onto a 20 µm filter on an 8 cm diameter petri dish with a small amount of water to hatch.

[0332] Root-knot nematodes extracted from infested tomato roots were identified based on morphology and isozyme phenotyping for esterase (EST) and malate dehydrogenase (MDH). Morphological identifications were conducted using perineal patterns of mature females is described in JOSEPH NEAL SASSER & CATHY CAMERON CARTER: AN ADVANCED TREATISE ON MELOIDOGYNE (1985), which is hereby incorporated by reference in its entirety. Briefly, isozyme phenotyping was conducted using 25 young egg-laying females. Extracts of females dissected directly from the root system were run on two polyacrylamide gel electrophoresis (PAGE) (Brito et al., *Nematology* 10:757-66 (2008), which is hereby incorporated by reference in its entirety). One gel was stained for both MDH and EST activities (KENNETH REECE BARKER, CATHY CAMERON CARTER & JOSEPH NEAL SASSER: AN ADVANCED TREATISE ON MELOIDOGYNE (1985), which is hereby incorporated by reference in its entirety), whereas a second was stained only for EST.

Example 59—Identification of *C. elegans* Dispersal Blend

[0333] Liquid cultures that induced 60% dauer (2 experiments) and 40% dauer after 67 hours of feeding L1s were analyzed using LC-MS. Four ascarosides were common to all three liquid media. The concentrations of each were measured from the liquid cultures that produced 60% dauers.

Example 60—Dispersal Assay

[0334] *S. feltiae* IJs were washed with MILLI-Q water three times and incubated in 6 cm petri dishes for 36 hours with a small amount (4–5 ml) of MILLI-Q water. Nematodes were placed the following day on a 10.7 g/L agar with 1010 g/cm² gel strength (PhytoTechnology Lab. Shawnee Mission, KS). Nematode behavior was assayed on multiple plates with internal plate replicates to rule out the possibility that behavior was affected by plate composition. Approximately 300 IJs in 10 µl water were placed on an agar medium and the test compounds or extracts (1–2 µl) were placed into the nematode suspension. Upon absorption (~15 minutes) of the liquid, the freely moving nematodes were video recorded for 5–10 minutes. Dispersal behavior is temperature and season dependent. During winter, the assay is effective at room temperature (22 ± 1°C). During summer, the assay requires a temperature controlled environment due to the effects on nematode behavior above 23°C.

[0335] *C. elegans* dauer juveniles were washed with MILLI-Q water 3 times, placed into 6 cm petri dishes with a small amount of water, and rested overnight. Approximately 200–300 nematodes in 10 µl of water were placed on an agar plate, to which 2 µl of treatment was added. The liquid culture that produced 60% dauer animals was centrifuged, filtered with a 0.45 µm filter, and used as a positive control for dispersal. Thereafter, media were lyophilized and resuspended in MILLI-Q water 5 times. 2 µl to 10 µl of nematode suspension was used for the assay. As a negative control, 0.5 % *E. coli* (HB101) was prepared in S-complete, lyophilized, and adjusted to the final volume of 0.25 % *E. coli* in the assay. The dispersal behavior was observed for 12–15 minutes.

[0336] To assay the dispersal of root knot nematodes, the root knot nematodes that hatched within 1–3 days were collected and washed with MILLI-Q water 3 times using 10 µm nylon filters (Nytex). Thereafter, they were placed into a 1.5 ml Eppendorf tube. The nematodes in 10 µl water were placed on an agar plate. Nematodes found at locations away from where they were originally placed were counted at 1 hour and 2 hours. Each treatment was normalized using the total number of nematodes that were deployed. For each treatment, 20 experiments were conducted on three different days. The nematode density was ~30 per plate in 14 experiments and 100 per plate in 6 experiments. The experiments were conducted in the morning.

Example 61—Quantifying of *C. elegans* Dispersal

[0337] Nematodes were quantified using Image J software (Image Processing and Analysis in Java, National Institutes of Health). The number of nematodes visualized and counted using Image J is illustrated in Figure 63B. Nematodes inside the circle were subtracted manually from the total number of worms counted because of the location of their placement. Each treatment was replicated 9–11 times. The graph was normalized to the positive control, the dauer conditioned medium.

Example 62—Purification of the *S. feltiae* Dispersal Blend

[0338] Activity-guided fractionation was conducted as reported in Srinivasan et al., *Nature* 454:1115-18 (2008), which is hereby incorporated by reference in its entirety, with modifications. A total of 33 insect host cadavers (*G. mellonella* larvae) were placed into 70% EtOH and stored at -20°C until extraction. The insect cadavers were homogenized using 1 g of ceramic zirconium beads (1.25 mm) (ZIRMIL) in 2 ml tubes for 37 seconds using a Precellys24 homogenizer. Samples were centrifuged for 15 minutes at 18400 relative centrifugal force (rcf), and the supernatant was lyophilized and resuspended in MILLI-Q water. The dispersal activity of nematodes was tested using the dispersal assay described in Example 60 and a physiologically relevant concentration of insect host cadaver extract or fractionated extract. To facilitate calculations for physiologically relevant concentration of the ascarosides, wax worm volume was estimated at ~200 µl; the average weight of wax worms was 232 ± 57 mg (n=19).

[0339] The first reverse-phase solid-phase extraction was performed using Sep-Pak Plus C18 cartridges (Waters corporation, Milford, MA). The initially collected flow through was termed Fraction A. Thereafter, the column was washed with water, collected, and saved. Subsequently, the column was eluted with 50% (Fraction B) and 90% (Fraction C) MeOH. The fractions were tested for dispersal activity both individually and in combination. Also, individual fractions were analyzed by LC-MS. Fraction A contained ascr#9, which was collected by LC-MS and tested for activity with Fraction B + C.

Example 63—Time Course Study of *S. feltiae* Ascaroside Production

[0340] The one-on-one assay method (LAWRENCE LACEY: MANUAL OF TECHNIQUES IN INSECT PATHOLOGY (1997), which is hereby incorporated by reference in its entirety) was used. One wax worm was placed in one well of a 24 well plate. For infection, 50 *S. feltiae* IJ were placed in 25 µl of water in each well. After 24 hours, the plates were examined for

infected insect larvae. Those that were infected were placed in a white trap (LAWRENCE LACEY: MANUAL OF TECHNIQUES IN INSECT PATHOLOGY (1997), which is hereby incorporated by reference in its entirety) and those that were infected after 48 hours were placed on a separate white trap. Initial sampling was made after 48 hours. Thereafter, samples were taken every day for 9 days and once at day 14. For each experiment, samples comprised six insect host cadavers. The insect cadavers were placed into a 2 ml Eppendorf tube with 1 ml of water. The cadavers were punctured with a needle to allow dissipation of internal contents and then vortexed. Samples were centrifuged at 3380 rcf for 10 minutes, and 0.5–1 ml of the supernatant was recovered. The supernatant was frozen at -20°C and then lyophilized. Thereafter, it was resuspended in 200 µl of 50% MeOH and diluted 1:1 with 0.1% formic acid. Sample pH was 4.3. At this point, 20 µl of each sample was subjected to LC-MS for analysis of *ascr#9*.

Example 64—*Ascr#9* in Insect Host Cadavers of *Steinernema* spp and *Heterorhabditis* spp.

[0341] Insect hosts (*G. mellonella*) were infected with *H. bacteriophora*, *H. zealandica*, *H. floridensis*, *S. carpocapsae*, *S. riobrave*, or *S. diaprepesi*. When nematodes began to emerge from insect cadavers, they were placed into 1.5 ml of 70% EtOH and stored at -20°C until use. Thereafter, insect cadavers were homogenized using 1 g of ceramic zirconium beads (1.25 mm) (ZIRMIL) in 2 ml tubes for 39 seconds using a Precellys24 homogenizer. The homogenized cadavers were centrifuged at 3380 rcf for 10 minutes. The supernatant was diluted with 1 ml of HPLC water, placed at -20°C, and then placed into a SpeedVac® (Speed Vac Plus SC210A, Savant) overnight. Each cadaver extract was re-suspended in 1 ml of 50% MeOH and centrifuged at 18400 rcf for 15–20 minutes. Thereafter, samples were diluted in a 1:1 ratio with 0.1% formic acid, yielding a sample pH of 4.2. Presence or absence of *arc#9* was determined by LC-MS.

Example 65—LC-MS Analysis

[0342] Ascaroside analysis was carried out using the method reported in Kaplan et al., *Publ. Lib. Sci. ONE* 6:e17804 (2011), which is hereby incorporated by reference in its entirety.

Example 66—Dispersal Assay

[0343] For entomopathogenic nematodes (EPN), insect cadavers are known to promote dispersal behavior of the IJ stage (Shapiro-Ilan et al., *J. Invertebr. Pathol.* 83:270-72

(2003), which is hereby incorporated by reference in its entirety). An assay was therefore developed to identify compounds in consumed insect cadavers that promote dispersal. Approximately 300 IJs of *S. feltiae* in 10 µl of water were placed on an agar plate (Figure 64B). Then, either 2 µl water (Figure 64C) or an aqueous extract of *Galleria mellonella* larval cadavers infected with *S. feltiae* (Figure 64D) was placed into the nematode suspension. After absorption of water into the media, nematodes were able to move freely. The water-treated nematodes remained near the site of deployment with no dispersal behavior. In contrast, cadaver extract-treated nematodes were very active and moved away from the point of release. This was the same distinct dispersal behavior observed for nematodes leaving a cadaver. LC-MS analyses of the cadaver extract revealed the presence of an ascaroside (ascr#9) (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety), supporting the hypothesis that *S. feltiae* might utilize ascarosides as dispersal signals.

Example 67—*C. elegans* Dispersal Is Regulated by a Blend of Ascarosides

[0344] The dispersal bioassay in Example 66 was then investigated to determine whether it could also be used to test for dispersal of *C. elegans* dauer larvae. For this assay, a growth medium from a *C. elegans* liquid culture that had produced 60% dauer larvae was used. Following removal of all nematodes, this dauer-conditioned medium strongly induced dispersal behavior in the *C. elegans* dauer. Using LC-MS, the dauer forming medium was found to contain four known ascarosides (ascr#2, ascr#3, ascr#8, and icas#9) (Butcher et al., *Nat. Chem. Biol.* 3:420-22 (2007); Pungaliya et al., *PNAS* 106:7708-13 (2009); Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which are hereby incorporated by reference in their entirety) (Figure 63A and 65), which were shown previously to individually promote dauer entry in *C. elegans* (Butcher et al., *Nat. Chem. Biol.* 3:420-22 (2007); Butcher et al., *Org. Lett.* 11:3100-03 (2009); Pungaliya et al., *PNAS* 106:7708-13 (2009), which are hereby incorporated by reference in their entirety). The concentrations of the ascarosides in the dauer forming culture medium were estimated to be ascr#2 (3.68 pmol/µl), ascr#3 (0.165 pmol/µl), ascr#8 (0.25 pmol/µl), and icas#9 (0.005 pmol/µl).

[0345] A synthetic blend of these ascarosides was then tested for dispersal activity, using the dauer conditioned medium as a positive control. The media was estimated to contain approximately half of the original 0.5% *E. coli* (HB101) food source. Thus 0.25% *E. coli* was added to the synthetic test samples as well as to a water control to prevent food searching behavior induced by the absence of food. The number of dispersing nematodes was

normalized to the percent of the positive control response. In the presence of just the food (negative control), approximately 35% of the dauer larvae left the release location. However, with the addition of the synthetic ascaroside blend, nearly twice as many nematodes (62%) moved away from the release location (Figures 63B and 66A–B). Tested individually at physiological concentration, ascr#8 (50%) and ascr#2 (40%) gave the strongest response, but all four were less active than the blend (Figure 63C). This suggests that *C. elegans* dauer larvae were able to perceive and respond to single components of the dispersal blend, but the complete four-component blend was necessary to restore the activity.

Example 68—EPNs and Plant Parasitic Nematodes Sense the *C. elegans* Dispersal Blend

[0346] It was hypothesized that many nematode species might be able to sense and respond to signals released by other nematode species. Thus, ascaroside released by *C. elegans* could function as valid avoidance signals. In the dispersal assay, IJs of *S. feltiae* exhibited no noticeable movement when exposed to water, but were very active and moved away from the release location when exposed to the *C. elegans* dispersal blend (Figures 63D and 66C). The mobile J2 form of wild root knot nematodes (a mixture of *Meloidogyne* spp; *M. javanica*, *M. floridensis*, and *M. incognita*) isolated from tomato roots were also tested for response to the *C. elegans* dispersal blend (Figure 66D). Again, more nematodes (10–12%) left the area where the *C. elegans* blend was introduced as compared to the control. This suggests that at least some nematode species can respond to dispersal blends of distantly related nematode species.

Example 69—The Major Components of *S. feltiae* and *C. elegans* Blends Are Structural Analogs

[0347] For characterization of the *S. feltiae* dispersal pheromone, insect host cadavers were extracted with 70% EtOH, fractionated by reverse phase (C18) chromatography, and assayed (Figures 67A–D). The bioassay revealed that a combination of all three fractions (A, B, and C) was necessary for activity (Figure 67A). Analysis of the fractions by LC-MS (Figures 68A–C) revealed 2 ascarosides in fraction A, which were identified to be ascr#9 and ascr#11 with the help of a synthetic standard (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012); von Reuss et al., *J. Am. Chem. Soc.* 134(3):1817-24 (2012), which are hereby incorporated by reference in their entirety). Bioassays showed that ascr#9 and ascr#11 did not have activity by themselves at physiologically relevant concentrations (Figure 67B). This is because the natural fraction A was inactive when tested alone (Figure 67A). However, combining synthetic ascr#9 at the biologically relevant concentration (40

pmol/ μ l) with natural fractions B and C restored the original dispersal activity (Figure 67C), confirming ascr#9 as an active component of the *S. feltiae* dispersal blend. The second ascaroside found in fraction A, ascr#11, also restored the full activity when combined with fractions B and C (Figure 67D), indicating that either ascr#9 or ascr#11 were by themselves sufficient to reconstitute full dispersal activity in combination with fractions B and C.

[0348] A developmental profile has been previously established (Kaplan et al., *Publ. Lib. Sci. ONE* 6:e17804 (2011), which is hereby incorporated by reference in its entirety) for the principle *C. elegans* dispersal pheromone component, ascr#2. The primary ascaroside of fraction A in the *S. feltiae* dispersal blend, ascr#9, is a structural analog to ascr#2 (Figure 67C). The accumulation of ascr#9 in *S. feltiae* infected insect cadavers was thus analyzed in a time course experiment (Figure 69). The results show that ascr#9 had a very similar accumulation profile as previously established for ascr#2 in *C. elegans*. Furthermore, the accumulation of ascr#9 was highest directly prior to IJ dispersal and remained constant for at least 6 days in the IJ-depleted cadaver. This is very similar to observation of ascr#2 and dauer formation for *C. elegans* (Kaplan et al., *Publ. Lib. Sci. ONE* 6:e17804 (2011), which is hereby incorporated by reference in its entirety). In contrast, the ascr#11 profile (Figure 69) differed from that of ascr#9. It initially became detectable but not quantifiable at day 8 (dispersal day), and could not be quantified in the insect host cadaver until day 14, which is 6 days after dispersal.

[0349] Ascr#9 was also tested and was found to be able to substitute for its structural analog ascr#2 in the *C. elegans* dispersal blend (Figure 70). Thus, a cross species perception, as found with *S. feltiae*, may also be true for *C. elegans*.

Example 70—Ascr#9 Could Be a Common Component in Dispersal Blends of Phylogenetically Related EPNs

[0350] In beetles and flies, phylogenetically related species share components in their aggregation pheromone blends (Symonds & Elgar, *Proc. Biol. Sci.* 271:839-46 (2004); Symonds & Wertheim, *J. Evol. Biol.* 18:1253-63 (2005), which are hereby incorporated by reference in their entirety). To further test to what degree dispersal blend components were shared by phylogenetically related nematode species, insect host cadavers infected with *Steinernema* spp. and *Heterorhabditis* spp. were analyzed for presence of ascr#9, ascr#2 and ascr#11 (Figures 71A–B).

[0351] The insect host cadavers for both species were found to contain ascr#9 (Figures 71A–B), which suggests that ascr#9 may be used by a broad range of EPN species

as part of their dispersal blends. Ascr#11 was found in all tested *Steinernema* spp., but not in *Heterorhabditis* spp, suggesting that ascr#11 may be specific for *Steinernema* spp.

Discussion of Examples 56–70

[0352] For *C. elegans*, shared and unique compositions of ascarosides regulate different behaviors. For example, the mating and dispersal blends share ascr#2, ascr#3, and ascr#8 (Srinivasan et al., *Nature* 454:1115-18 (2008); Pungaliya et al., *PNAS* 106:7708-13 (2009); Kaplan et al., *Publ. Lib. Sci. ONE* 6:e17804 (2011), which are hereby incorporated by reference in their entirety). The *C. elegans* mating blend is characterized by a unique component, ascr#4 (Srinivasan et al., *Nature* 454:1115-18 (2008), which is hereby incorporated by reference in its entirety), and the dispersal blend has a unique component, icas#9 (Srinivasan et al., *Publ. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety). It is therefore thought that this can be common for many nematode species.

[0353] It was not yet clear whether *Meloidogyne* spp. also utilizes dispersal blends, but it has been shown herein that these nematode species can detect and respond to signals released by other nematode species that are indicative of decomposing and decaying plant material. The *C. elegans* dispersal blend shows similar activity in developmentally analogous stages of phylogenetically related species, suggesting that it can be used to identify genetic targets as well as to formulate dispersal blends for control of species parasitic to plants, humans, and livestock. Examples 56–70 not only demonstrate that several nematode species utilize species-specific small molecule signals to regulate dispersal behavior, but also that nematode dispersal behavior may be broadly induced by interspecies communication.

Example 71—Retention Assay

[0354] OP50 *E. coli* was grown on a standard 5 cm agar plate (made with standard Nematode Growth Medium). The bacterial lawn was 16 mm in diameter and was grown overnight at 20°C before being used in trials. Two 4 mm spots (0.6 µL) were placed on opposite sides of the bacterial lawn (using a transparent template to guide spot placement), and several minutes elapsed for the liquid to settle in before placing nematodes down on the assay. Recording began immediately upon worm placement. 0.6 µL of the control was placed on one side of the lawn, and 0.6 µL of the experimental cue was placed on the other side of the lawn, changing the location of the cue throughout trials between left/right and top/bottom to avoid bias.

[0355] Nematodes were isolated by gender at the L4 stage the day before being used in trials as developed adults. Worms were evenly divided and placed at two points equidistant from the foci of the scoring region. Trials were recorded for 20 minutes, and frames were collected for analysis at 1 frame per second. Results were averaged from at least three different trials. For every nematode species in this study, a different total number of worms (using water in both scoring regions) was tested to determine the minimum number of worms necessary for consistent unbiased results over a 20 minute trial. The total number of worms used in the multiple species assays depended on that species' optimal parameters. 10 worms were used for *P. redivivus* males and females, 20 worms were used for *C. elegans* males and *O. dolichuridae* males, and 14 worms were used for *S. glaseri* males. The Automated Software was used to compare worm occupancy in each scoring region over time, and the Chemotaxis Index (Bargmann et al., *Cell* 74:515-27 (1993), which is hereby incorporated by reference in its entirety) was adapted to score preference or avoidance to each ascaroside.

Example 72—Attraction Assay

[0356] 10 cm standard chemotaxis plates were prepared (Bargmann et al., *Cell* 74:515-27 (1993), which is hereby incorporated by reference in its entirety) and stored at 4°C until the day before use. Standard 1 cm-diameter copper pipes were sectioned into 1 cm-tall segments to use for the holding chambers. The females/hermaphrodites were isolated overnight, washed 3 times in M9 buffer, and then allowed to wander on a plate pre-conditioned with 50uL of 1000× streptomycin for 2 hours to kill any bacteria that might produce a false positive attractant. The nematodes were then washed 3 times, and the supernatant from the last wash was placed in a copper chamber, as the control, 1 cm from the border of the 10 cm plate. The nematodes were suspended in M9 within the copper chamber, 1 cm from the border opposite the control chamber. After 6 hours, they were subsequently removed and replaced with 3 µL of 1M sodium azide. 100 *C. elegans* adult males were isolated overnight, washed several times in ddH₂O, and placed at the center of the agar plate. After several hours, males paralyzed within 2 cm of each region were scored. The Chemotaxis Index (Bargmann et al., *Cell* 74:515-27 (1993), which is hereby incorporated by reference in its entirety) was then used to score male chemotaxis to either point source.

Example 73—Mating Assay

[0357] The mating assay described in Peden et al., *Curr. Biol.* 15:394-404 (2005), which is hereby incorporated by reference in its entirety, was adapted. Briefly, 40 young adult hermaphrodites/females were placed on an 8 mm bacterial lawn. *C. elegans* L4 males were isolated overnight. 5 males were placed on the hermaphrodite/female-rich plate. The number of males that responded to a hermaphrodite within a 3-minute period were counted (a single male only being counted once). Response was defined when a male stopped at the hermaphrodite and affixed their ventral tail on the potential mate for more than 10 consecutive seconds. They were scored for their Response Efficiency, also adapted from Peden et al., *Curr. Biol.* 15:394-404 (2005), which is hereby incorporated by reference in its entirety, which was calculated using the formula: Response Efficiency = [# males responding/5 males] × 100%.

Example 74—Automated Software (for Retention Assay)

[0358] A video camera attached to the microscope produced a digital video stream, which was then analyzed. The ratio of the time the average worm spent in each region of interest was calculated for every trial. For ease of implementation, all worms in a single experiment were assumed to be roughly the same size. Thus worm pixels were counted instead of whole worms, allowing fractions of a worm in the region of interest to be taken into account. It also eliminated the need for a shape-based worm identification algorithm, and allowed each frame to be analyzed independently. A band-pass filter was applied to each frame to eliminate the effect of uneven lighting and also accentuate the worms against the background. The worm was then identified after thresholding the filtered image. Throughout each experiment, the locations and sizes of the regions of interest were known. Through the filtering described above, it is known which pixels are occupied by worms and which ones are not. Thus, the ratio of worm-pixels to all pixels inside the region of interest can be calculated to produce the worm-occupancy ratio. This calculation was done for every frame, giving a plot output of worm-occupancy ratio vs. time for each region.

Example 75—HPLC analysis.

[0359] A Thermo Finnigan LCQ Deca XP Max was used with electrospray ionization in positive ion mode in the 50-1000 AMU range. The Thermo Separation spectra HPLC system consisted of a P4000 quaternary pump, an AS 3000 auto sampler, and a UV 6000 diode array detector. The solvents were water with (a) 0.1% formic acid and (b) 90%

acetonitrile–10% water with 10mM ammonium formate. The column temperature was maintained at 60°C and a solvent flow of 1.0 ml/min. The reversed phase column (PLRP-S Polymeric reversed phase column, 250x4.6mmid, Varian Inc.) was eluted with a solvent composition starting with 90:10 (a,b) for 2 minutes, followed by a gradient to 5:95 in 20 minutes and an additional 5 minutes at 5:95. Ultraviolet absorption was monitored at 190 and 400 nm. Solvent flow between the ultraviolet detector and mass spectroscopy electrospray interface split 9:1 with a low-volume micro needle P450 splitter valve (Upchurch Scientific, Oak Harbor, WA), making it possible to obtain spectra of eluted compounds and simultaneously collect 90% of the injected material for bioassay. The purified peak with retention time of 9.7 minutes was active to males in the bioassay. LC-MS with chemical ionization (positive mode) shows that the major peak had an m/z of 294(M+NH₄). This was confirmed to be ascr#1 using NMR analysis.

Example 76—Construction of the Phylogenetic Tree:

[0360] Small subunit ribosomal DNA (SSU rDNA) sequences for this analysis were obtained from GenBank. The sequences were first aligned using MUSCLE and then subsequently trimmed to facilitate comparison of sequences with varying lengths. The trimmed alignment resulted in 690 characters represented for all taxa. The Neighbor-Joining (N-J) analysis was done using the “Dnadist” and “Neighbor” programs from the PHYLIP 3.68 package, using “Consense” to produce a majority-rule consensus tree.

Example 77—Culture of Nematodes and Collection of Worm Excretions and Secretions

[0361] *C. elegans*, *Rhabditis* sp., *O. tipulae*, *C. sp.7*, *P. pacificus*, *P. redivivus*, *Koernia* sp., and *P. strongyloides* were maintained on standard 6 cm agar plates with *E. coli* OP50. To grow large amounts, they were grown in liquid culture—S complete medium with *E. coli* HB101 at 20°C at 250 rpm in an incubator shaker. The worms were exposed to several wash and filtration steps, using S Basal to remove bacteria. The worms were collected between the washes by centrifugation at 3000 rpm for 5 minutes. The worms were incubated at 1 worm/μL in S Basal for 6 hours. The worms were then filtered out and the remaining supernatant was passed through a 0.22 μm filter before being stored at -20°C.

[0362] *S. carpocapsae*, *S. glaseri*, *S. riobrave*, and *H. bacteriophora* were maintained as reported in Hallem, et al., *Curr. Biol.* 21(5):377-83 (2011), which is hereby incorporated by reference in its entirety. Briefly, several last-instar *Galleria mellonella* larvae were placed on a 6 cm Petri dish with a 55 mm Whatman 1 filter paper. Approximately 500-1000 IJs were

placed on this filter paper. After 7 days, the insect cadavers were placed on an inverted 6 cm Petri plate carrying a Whatman 1 filter paper and immersed in a small volume of ddH₂O in a 10cm Petri dish. Emerging IJs were collected and washed several times as described above, before being incubated in ddH₂O at 1 worm/ μ L for 6 hours. Adults were dissected out of insect cadavers after 3 days post-infection, and washed/incubated as described above.

[0363] *N. brasiliensis*, *A. suum*, *P. penetrans*, and *Romanomermis* spp. were provided by the collaborators identified in Table 11 below. All strains used in this Example are listed. Species without a strain identification are labeled “no designation”.

Table 11. Source of nematode strains

Species	Strain	Source or References
<i>Nippostrongylus brasiliensis</i>	no designation	Edward G. Platzer
<i>Heterorhabditis bacteriophora</i>	M31e	Sternberg collection
<i>Rhabditis</i> sp.	AF5	Caenorhabditis Genetics Center
<i>Oscheius tipulae</i>	PS1305	Sternberg collection
<i>Caenorhabditis elegans</i>	N2	Sternberg collection
<i>Caenorhabditis</i> sp.7	JU1325	Marie-Anne Felix
<i>Ascaris suum</i>	no designation	Robin Gasser
<i>Pristionchus pacificus</i>	RS2333	Ralf Sommer
<i>Koernia</i> sp.	RS1982	Ralf Sommer
<i>Pratylenchus penetrans</i>	3/C	Antoon T. Ploeg
<i>Panagrellus redivivus</i>	PS2298	Sternberg collection
<i>Pelodera strongyloides</i>	DF5022	Caenorhabditis Genetics Center
<i>Steinernema carpocapsae</i>	ALL	Sternberg collection
<i>Steinernema riobrave</i>	no designation	Sternberg collection
<i>Steinernema glaseri</i>	VII	S. Patricia Stock
<i>Romanomermis iyengari</i>	no designation	Edward G. Platzer
<i>Romanomermis culicivorax</i>	3B4	Edward G. Platzer

[0364] *N. brasiliensis* was incubated in 0.85% saline at 1 worm/5 μ L (given their large size) at 30°C. *A. suum* were washed in sterile saline before being incubated in DMEM for 3, 6, and 13 hours at 37°C. The supernatants were tested individually and then subsequently pooled for combined analysis. *P. penetrans* and *Romanomermis* spp. were incubated in ddH₂O at 20°C at 250 rpm overnight.

Example 78—Worm Sample Preparation

[0365] Worm water samples were lyophilized, extracted with 1-10 ml methanol twice, and filtered over cotton wool. Extracts were concentrated in vacuum. Resulting residues were resuspended in 150 μ L methanol and then filtered.

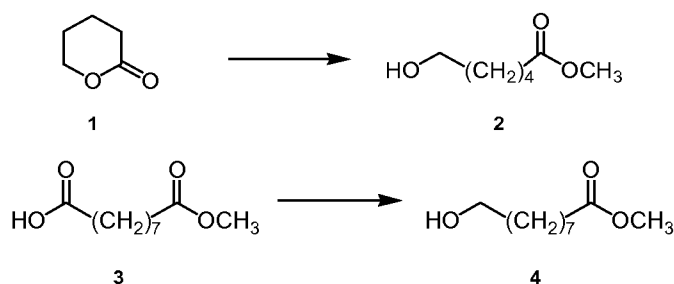
Example 79—MS Analysis

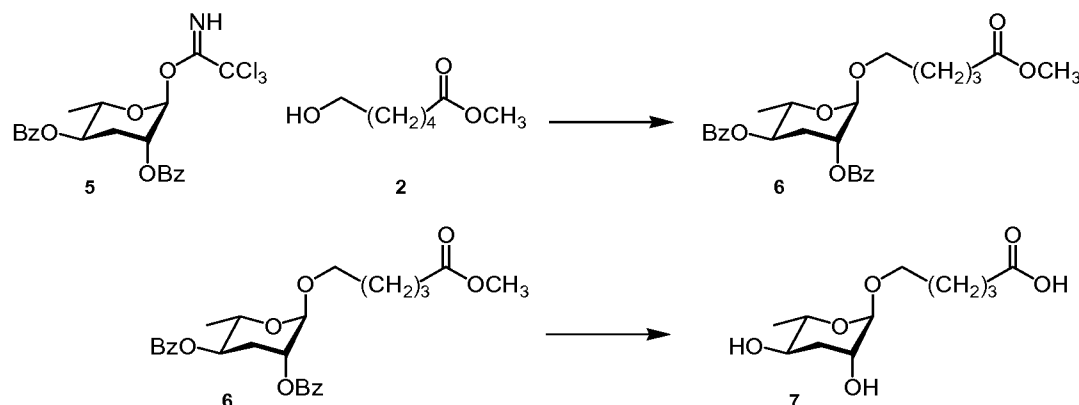
[0366] HPLC-MS was performed using an Agilent 1100 Series HPLC system equipped with an Agilent Eclipse XDB-C18 column (9.4×250 mm, 5 μ m particle diameter) connected to a Quattro II spectrometer (Micromass/Waters) using a 10:1 split. A 0.1% acetic acid–acetonitrile solvent gradient was used at a flow rate of 3.6 ml/min, starting with an acetonitrile content of 5% for 5 minutes and then increased to 100% over a period of 40 minutes. Metabolite extracts were analyzed by HPLC-ESI-MS in negative and positive ion modes using a capillary voltage of 3.5 kV and a cone voltage of -40 V and +20 V, respectively. Ascarosides were identified by comparison of quasi molecular ion signals (Figures 72A–D) and HPLC retention times (Figure 73) with those of a library of 150 components identified from wild-type and peroxisomal beta-oxidation mutants of *C. elegans*. To confirm the assignment of ascaroside structures, MS/MS screen was used for precursor ions of $m/z = 73.0$ (Figures 74A–B), because previous work indicated that all ascarosides afforded an intensive $C_3H_5O_2$ product ion upon MS/MS (argon as collision gas at 2.1 mtorr and 30 eV). Quantification of ascarosides was performed by integration of LC-MS signals from the corresponding ion-traces. Relative ascaroside concentrations were finally calculated using response factors determined for synthetic standards of ascr#1, ascr#2, ascr#3, ascr#5, ascr#7, ascr#9, ascr#10, oscr#9, oscr#10, and icas#9. Response factors for the ascarosides which have not been synthesized yet were extrapolated based on data observed for the available standards.

Example 80—Synthesis of Ascaroside oscr #9

[0367] Ascaroside oscr#9 was prepared as shown in Scheme 11 below.

Scheme 11





[0368] To prepare **2**, freshly distilled δ -valerolactone (**1**) (856 mg) in methanol (17 ml) was treated with concentrated H_2SO_4 (100 μl) and refluxed for 12 hours. The solution was cooled to -20°C , treated with NaHCO_3 (80 mg), and stirred for 10 minutes. The reaction mixture was filtered and the solvent was removed in vacuum. The residue was taken up in DCM (10 ml), dried over Na_2SO_4 , filtered, and concentrated in vacuum to give methyl 5-hydroxypentanoate (**2**) (Huckstep et al., *Synthesis* 10:881-82 (1982), which is hereby incorporated by reference in its entirety) (1071 mg, 94%) as a colorless oil, which was used directly without any further purification. ^1H NMR (400 MHz, acetone- d_6): δ 1.55 (2H, m), 1.68 (2H, m), 2.35 (2H, t, $J = 7.4$ Hz), 3.57 (2H, m), 3.64 (3H, s); ^{13}C NMR (100 MHz, acetone- d_6): δ 22.2, 32.9, 34.1, 51.5, 61.9, 174.2. See Figures 75A–B.

[0369] To prepare **4**, a solution of nonanedioic acid monomethyl ester (**3**) (923 mg, 4.6 mmol) in dry THF (3 ml) at -20°C was treated with 1 M BH_3 in THF (4.6 ml, 4.6 mmol) over 10 minutes. After stirring at room temperature for 4 hours, the reaction was quenched with 0.77 M aqueous K_2CO_3 solution (10 ml) at 0°C . The product was extracted with diethyl ether (3×20 ml), washed with saturated aqueous NaCl solution, dried over Na_2SO_4 , and concentrated in vacuum to afford methyl 9-hydroxynonanoate (**4**) (Kai K. et al., *Tetrahedron* 64:6760-69 (2008), which is hereby incorporated by reference in its entirety) (850 mg, 99% yield) as a colorless oil, which was used directly without any further purification. ^1H NMR (400 MHz, chloroform- d_1): δ 1.27-1.37 (8H, m), 1.50-1.66 (4H, m), 2.29 (2H, t, $J = 7.5$ Hz), 3.62 (2H, t, $J = 6.5$ Hz), 3.66 (3H, s). See Figure 75C.

[0370] To prepare **6**, a solution of 2,4-di-O-benzoyl-ascarylose-1-(2,2,2-trichloroacetimidate) (**5**) (11116) (132 mg, 263 μmol) and **2** (125 mg, 950 μmol) in dry DCM (3 ml) at 0°C was treated with trimethylsilyloxy triflate (5 μl). After 3 hours, the solution was washed with saturated aqueous NaHCO_3 solution (0.5 ml), dried over Na_2SO_4 , and concentrated in vacuum. Flash column chromatography on silica gel using a 20–40% (v/v)

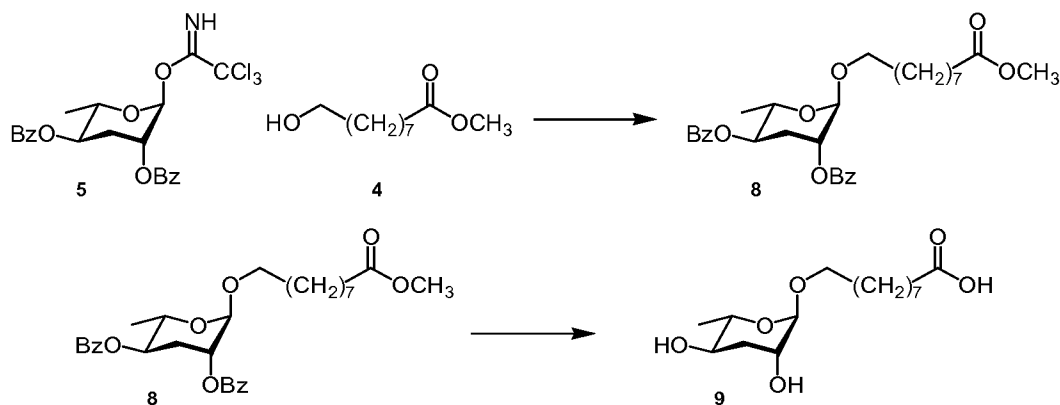
ethyl acetate gradient in *n*-hexane afforded 5- (3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2-yloxy)-pentanoic acid methyl ester (**6**) (66.8 mg, 142 μ mol, 47%) as a colorless oil. ^1H NMR (400 MHz, acetone- d_6): δ 1.28 (3H, d, J = 6.2 Hz), 1.67-1.80 (4H, m), 2.23 (1H, m), 2.40 (2H, t, J = 6.9 Hz), 2.48 (1H, m), 3.58 (1H, m), 3.64 (3H, s), 3.83 (1H, m), 4.13 (1H, dq, J = 9.8 Hz, J = 6.0 Hz), 4.87 (1H, s.br), 5.15 (1H, ddd, J = 11.0 Hz, J = 10.4 Hz, J = 4.5 Hz), 5.18 (1H, s.br), 7.50-7.60 (4H, m), 7.62-7.72 (2H, m), 8.05 (2H, d, J = 7.5 Hz), 8.11 (2H, d, J = 7.5 Hz); ^{13}C NMR (100 MHz, acetone- d_6): δ 18.3, 22.5, 29.6, 34.0, 51.5, 67.5, 67.9, 71.4, 71.5, 97.0, 129.4, 129.5, 130.03, 130.4, 131.0 (2X), 134.1, 134.2, 165.9, 166.0, 174.0. See Figures 75D–E.

[0371] To prepare oscr#9 (**7**), a solution of **6** (66.8 mg, 142 μ mol) in dry THF (0.5 ml) was added to a mixture of LiOH (28 mg, 1.4 mmol) and water (0.6 ml) in 1,4-dioxane (4 ml). After stirring at 66°C for 2 hours, the solution was acidified with glacial acetic acid and concentrated in vacuum. Flash column chromatography on silica gel using a 5–30% (*v/v*) methanol gradient in DCM containing 1% glacial acetic acid afforded 5-(3'*R*,5'*R*-dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2-yloxy)-pentanoic acid (oscr#9) (**7**) (26 mg, 105 μ mol, 74%) as a colorless oil. ^1H NMR (400 MHz, methanol- d_4): δ 1.22 (3H, d, J = 6.0 Hz), 1.58-1.72 (4H, m), 1.77 (1H, ddd, J = 13.1 Hz, J = 11.1 Hz, J = 3.2 Hz), 1.95 (1H, ddt, J = 13.1 Hz, J = 3.7 Hz, J = 0.9 Hz), 2.33 (2H, t, J = 7.2 Hz), 3.43 (1H, dt, J = 9.6 Hz, J = 6.0 Hz), 3.47 – 3.59 (2H, m), 3.71 (1H, dt, J = 9.8 Hz, J = 6.2 Hz), 3.77 (1H, m), 4.50 (1H, s); ^{13}C NMR (100 MHz, methanol- d_4): δ 18.1, 23.0, 30.1, 34.7, 36.0, 67.9, 68.3, 69.4, 70.9, 100.4, 177.5; ESI-MS (negative mode) m/z = 247.1 [$\text{M}-\text{H}$]. See Figures 75F–G.

Example 81—Synthesis of Ascaroside oscr #10

[0372] Ascaroside oscr#10 was prepared as shown in Scheme 12 below.

Scheme 12



[0373] To prepare **8**, a solution of 2,4- di-*O*-benzoyl-ascarylose-1-(2,2,2-trichloroacetimide) (**5**) (132 mg, 263 μ mol, Hallem, et al., *Curr. Biol.* 21(5):377-83 (2011), which is hereby incorporated by reference in its entirety) and **4** (112.8 mg, 600 μ mol) in dry DCM (3 ml) at 0°C was treated with trimethylsilyloxy triflate (5 μ l). After 3 hours, the solution was washed with saturated aqueous NaHCO₃ solution (0.5 ml), dried over Na₂SO₄, and concentrated in vacuum. Flash column chromatography on silica gel using 20–40% (v/v) ethyl acetate gradient in *n*-hexane afforded 99.3 mg 9-(3'*R*,5'*R*-dibenzoyloxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2-yloxy)-nonanoic acid methyl ester (**8**) (190 μ mol, 72% yield) as a colorless oil. ¹H NMR (400 MHz, acetone-*d*₆): δ 1.28 (3H, d, 6.2 Hz), 1.30 – 1.40 (6H, m), 1.40 – 1.49 (2H, m), 1.56 – 1.72 (2H, m), 2.22 (1H, ddd, *J* = 13.6 Hz, *J* = 11.5 Hz, *J* = 3.2 Hz), 2.30 (2H, t, *J* = 7.5 Hz), 2.46 (1H, m), 3.55 (1H, dt, *J* = 9.8 Hz, *J* = 6.5 Hz), 3.60 (3H, s), 3.81 (1H, dt, *J* = 9.6 Hz, *J* = 6.6 Hz), 4.13 (1H, dq, *J* = 9.7 Hz, *J* = 6.2 Hz), 4.86 (1H, s.br), 5.15 (1H, ddd, *J* = 11.4 Hz, *J* = 9.8 Hz, *J* = 4.6 Hz), 5.18 (1H, m), 7.50 – 7.60 (4H, m), 7.63 – 7.71 (2H, m), 8.04 (2H, m), 8.11 (2H, m); ¹³C NMR (100 MHz, acetone-*d*₆): δ 18.3, 25.6, 26.8, 29.7, 29.9, 30.0, 30.2, 30.4, 34.4, 51.4, 67.4, 68.2, 71.4, 71.5, 97.0, 129.4, 129.5, 130.2, 130.3, 130.9, 131.0, 134.1, 134.2, 165.9 (2C), 174.3. See Figures 75H–I.

[0374] To prepare oscr#10, a solution of **8** (99.3 mg, 190 μ mol) in THF (500 μ l) was added to a mixture of LiOH (38 mg, 1.9 mmol) and water (800 μ l) in 5 ml 1,4-dioxane (5 ml). After stirring at 66°C for 3 hours, the solution was acidified with acetic acid and concentrated in vacuum. Flash column chromatography on silica gel using a 5–30% (v/v) methanol gradient in DCM containing 1% glacial acetic acid afforded 9-(3'*R*,5'*R*-dihydroxy-6'*S*-methyl-(2*H*)-tetrahydropyran-2-yloxy)-nonanoic acid (oscr#10) (**9**) (49 mg, 161 μ mol, 85%) as a colorless oil. ¹H NMR (400 MHz, methanol-*d*₄): δ 1.22 (3H, d, *J* = 6.1 Hz), 1.32–1.43 (8H, m), 1.56–1.63 (4H, m), 1.77 (1H, ddd, *J* = 13.1 Hz, *J* = 11.1 Hz, *J* = 3.2 Hz), 1.96 (1H, ddt, *J* = 13.1 Hz, *J* = 3.7 Hz, *J* = 0.9 Hz), 2.28 (2H, t, *J* = 7.4 Hz), 3.41 (1H, dt, *J* = 9.6 Hz, *J* = 6.2 Hz), 3.49 – 3.59 (2H, m), 3.68 (1H, dt, *J* = 9.8 Hz, *J* = 5.5 Hz), 3.76 (1H, m), 4.49 (1H, s); ¹³C NMR (100 MHz, methanol-*d*₄): δ 17.3, 25.2, 26.4, 28.0, 29.3, 29.5, 29.6, 30.5, 34.1, 61.1, 67.4, 68.5, 69.9, 99.4, 176.8; ESI-MS (negative ion mode) *m/z* = 303.2 [M-H]. See Figures 75J–K.

Example 82—Many Nematode Species Produce Ascarosides

[0375] A mass spectrometry-based screen was initiated for ascarosides (Figures 76A–D) in a varied selection of nematode species, including both free-living and parasitic (plant,

insect, and mammal) nematodes. Previous studies have shown that *C. elegans* ascaroside production is developmental stage-specific (Kaplan et al., *Pub. Lib. Sci. ONE* 6(3):e17804 (2011), which is hereby incorporated by reference in its entirety). To obtain comprehensive samples, exudates from a developmentally mixed culture were tested, thus containing an accumulation of ascarosides released by all larval and adult stages. When feasible, parasitic infective juveniles and adults were tested separately, for the purpose of identifying ascarosides uniquely associated with host seeking or mate finding, respectively. Nematode secretions were analyzed via HPLC-MS using a protocol optimized for the detection of ascarosides in complex metabolome samples (Example 79). These analyses revealed the presence of ascarosides in most of the nematode samples. Samples were compared with mass spectroscopic data from a library of 150 ascarosides previously identified from both wild-type and mutant strains of *C. elegans*. These analyses revealed the presence of ascarosides in most of the analyzed nematode samples.

[0376] Ascarosides generally occurred as mixtures, including compounds with saturated and unsaturated side chains. Side-chain lengths were highly variable and ranged from the shorter chains also found in *C. elegans* to compounds with considerably longer side chains in *Pelodera strongyloides* and *Heterorhabditis bacteriophora*. As in *C. elegans*, most of the identified ascarosides bear the ascarylose sugar at the penultimate carbon of the side chain (“ ω -1-functionalization”), except for *Caenorhabditis sp. 7* and *Rhabditis sp.*, which produce ascarosides in which the ascarylose is attached to the terminal carbon of the side chain (“ ω -functionalization”, e.g. oscr#9 and oscr#10 in Figure 76D). Indole ascarosides, e.g. icas#9 (Figure 76A), which were recently shown to act as aggregation signals in *C. elegans* (Srinivasan et al., *Pub. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety), were found only in *Caenorhabditis spp.* However, in *C. elegans*, indole ascaroside concentrations were much lower than those of simple ascarosides (“ascr” and “oscr” in Figure 76A). As the available metabolite sample sizes for most of the species studied herein were much smaller than those available for *Caenorhabditis spp.*, it is possible that indole ascarosides could not be detected in some nematodes because the amounts were too small for detection by HPLC-MS.

[0377] Ascaroside profiles generally varied between species (Figure 77). However, species from very different ecologies in some cases produced the same ascarosides. For example, ascr#9 is produced by the compost-dwelling *Panagrellus redivivus*, *Rhabditis sp.*, and *Caenorhabditis spp.*; soil-dwelling *Oschieus tipulae*; insect-associated *Pristionchus*

pacificus; and insect-infective *Heterorhabditis bacteriophora*, *Steinernema carpocapsae*, *Steinernema riobrave*, and *Steinernema glaseri*.

[0378] No known ascarosides were detected in 3 of the 17 analyzed nematode species, namely *Pratylenchus penetrans*, *Ascaris suum*, and *Romanomermis* species. It is possible that these species produce only very small quantities of ascarosides, or that they produce ascarosides with unexpected structural features that could not be detected with the used method, given that ascarosides similar to those identified from wild-type and mutant strains of *C. elegans* were screened for. Previous studies have reported the presence of lipid-like long-chain ascarosides in *Ascaris sum*; however, they were limited to oocytes and eggs (Bartley et al., *J. Nat. Prod.* 59(10):921-26 (1996); Tarr, *Comp. Biochem. Physiol.* 46B:167-76 (1973), which are hereby incorporated by reference in their entirety). Given their highly lipophilic character, these compounds are poorly soluble in aqueous media, which explains their absence in the above analysis herein, and thus are unlikely to serve as pheromones. Mermithid species, such as *Romanomermis spp.*, have the unique ability to store a lifetime supply of lipid-composed sustenance (Platzer, *J. Am. Mosquito Contr. Association Bulletin* 7:58-64 (2007); Ittycheriah et al., *Nematologica* 23:165-71 (1977), which are hereby incorporated by reference in their entirety). It is possible that they have a different mechanism of fatty acid metabolism or perhaps ascaroside synthesis has not yet evolved, given that the *Romanomermis spp.* were the most ancestral species tested.

Example 83—Ascarosides Mediate Distinct Nematode Behaviors

[0379] Nematode behavioral responses to a series of synthesized ascarosides were tested using several different behavior assays. A retention assay (Figure 78) using automated software to detect preference or avoidance of regions conditioned with individual ascarosides (Figure 79) was used. Nematode species in the HPLC/MS study were screened for compatibility with this bioassay, which requires adequate movement across a bacterial lawn. Parasitic nematodes were largely excluded due to their requirement of substrates similar to their host environment. This investigation was further limited to males, because in most species, males are much more mobile than females or hermaphrodites, with the exception of *P. redivivus* females, which move well and were therefore included. *Osccheius tipulae* was excluded due to the fact that males are only occasionally present in this hermaphroditic species; instead, the closely related male–female species, *Osccheius dolichuroides*, was added. In a previous study, samples of ~1 pmol (from 1 μ M concentration) of ascr#2 and ascr#3 proved to be the most active concentration for attraction in *C. elegans* (Srinivasan et al.,

Nature 454:1115-18 (2008), which is hereby incorporated by reference in its entirety). Thus, available synthetic ascarosides were tested at the concentration of 1nM, 1 μ M, and 1mM.

[0380] Several of the tested nematode species preferred areas conditioned by many of the same ascarosides, particularly ascr#1, ascr#3, ascr#7, ascr#8, ascr#9, and ascr#10 (Figure 78). However, activity thresholds varied between species. For example, *C. elegans* males preferred 1 μ M-conditioned regions of ascr#10, whereas *S. glaseri* and *P. redivivus* males only preferred 1 mM-conditioned regions of ascr#10. Furthermore, several nematode species responded to ascarosides that were not produced by their own species, suggesting possible interspecies communication. One of the tested species, *O. dolichuroides*, did not respond to any of the assayed ascarosides. It is possible that this species responds to other ascarosides or that specific environmental conditions are required to elicit its response to ascarosides.

[0381] Further, *P. redivivus* males and females were observed to respond to different sets of ascarosides, a phenomenon previously described in *C. elegans* (Srinivasan et al., *Nature* 454:1115-18 (2008), which is hereby incorporated by reference in its entirety). These gender-specific responses demonstrate that ascarosides may serve different roles both between and within species. For example, icas#9 acted as an aggregating stimulant in *C. elegans* (Srinivasan et al., *Pub. Lib. Sci. Biol.* 10(1):e1001237 (2012), which is hereby incorporated by reference in its entirety); however, it repelled *P. redivivus* females and elicited no response from *P. redivivus* males. Because icas#9 was not produced by *P. redivivus*, it could serve as an interspecies cue.

[0382] The ability of *C. elegans* hermaphrodites to attract *C. elegans* males from a distance was also tested. Previous studies have reported that *C. elegans* hermaphrodites failed to attract males on a 5 cm plate containing a point source conditioned by a single hermaphrodite or on an agar-mounted slide holding hermaphrodite-incubated supernatant (Bargmann et al., *Cell.* 74:515-27 (1993), which is hereby incorporated by reference in its entirety). In the experiments described herein, both the diameter of the assay arena and the number of hermaphrodites were expanded, using a 10 cm plate and testing 1–300 hermaphrodites (Figure 80A). The results demonstrate that *C. elegans* hermaphrodites indeed attracted males from a distance (Figure 80B). The ability of *C. elegans* males to chemotax towards 50 *C. elegans* hermaphrodites was compared to that of equal numbers of females or hermaphrodites from other species. Because it was found that *C. elegans* males were attracted to 7 different ascarosides (Figure 78), it was predicted that species producing these

ascarosides would attract *C. elegans* males, whereas those producing few or none of these ascarosides would fail to attract *C. elegans* males. The results herein show that *C. elegans* males demonstrated partial attraction to *P. redivivus* females (Figures 80A–C), whose species produced 3 of the 7 known *C. elegans* male attractants (Figure 77) (Srinivasan et al., *Nature* 454:1115-18 (2008); Pungaliya et al., *PNAS* 19:7708-13 (2009), which are hereby incorporated by reference in their entirety). It was also found that *C. elegans* males did not show long-range attraction to females or hermaphrodites from the species *S. carpocapsae*, *P. pacificus*, or *P. strongyloides*, which produced two, one, and none of the known *C. elegans* male attractants, respectively. These findings provide further evidence for the possibility of ascaroside-mediated interactions between different nematode species. The individual roles of three previously described *C. elegans* mate-finding pheromones (ascr#2, ascr#3, and ascr#8) (Srinivasan et al., *Nature* 454:1115-18 (2008); Butcher et al., *Nat. Chem. Biol.* 7:420-22 (2007), which are hereby incorporated by reference in their entirety) were also tested to elicit male attraction from a distance. The results show that ascr#2 and ascr#3 elicited *C. elegans* male attraction, with amounts as low as 500 attomol and 5 femtomol, respectively, whereas ascr#8 was not active in this assay. These results suggest that ascr#2 and ascr#3 serve to attract *C. elegans* males from a distance, whereas ascr#8 serves to hold males within vicinity upon arrival to a point source.

[0383] Mating between *C. elegans* males and conspecific hermaphrodites as well as females/hermaphrodites of different nematode species were also investigated. The mating assay described in Peden & Barr, *Curr. Biol.* 15:394-404 (2005), which is hereby incorporated by reference in its entirety, which scores the percentage of males that attempt to copulate with hermaphrodites/females, was adapted (Figure 81). The investigation was to test whether *C. elegans* would mate with distantly related nematode species. The results show that they did not attempt to copulate with *P. redivivus*, *S. carpocapsae*, *P. pacificus*, or *P. strongyloides* for 10 seconds (Figure 81). Furthermore, the addition of a blend of ascarosides did not induce mating to any of these nematode species. Previous studies indicated that *C. elegans* male mating behavior was guided primarily by hermaphrodite recognition via mechanosensory or local chemical cues (Liu et al., *Neuron* 14:79-89 (1995), which is hereby incorporated by reference in its entirety). The results herein suggest that ascaroside sex pheromones serve to assist *C. elegans* mate location, but do not affect physical mating.

Discussion of Examples 71–83

[0384] The findings of Examples 71–83 indicate that ascarosides comprise a broad nematode lexicon, given their widespread production and recognition by different nematode species. These findings are evocative of bacterial quorum sensing, where acyl homoserine lactones (AHLs) are both produced and sensed by many species of Gram-negative bacteria (Miller et al., *Annu. Rev. Microbiol.* 55:165-99 (2001), which is hereby incorporated by reference in its entirety) (Figure 82). Different AHLs are structurally very similar but have species-specific variations in the N-acyl chain (Miller et al., *Annu. Rev. Microbiol.* 55:165-99 (2001), which is hereby incorporated by reference in its entirety). Ascarosides are organized in a very similar fashion: they are composed of the same ascarylose sugar ring but have variations in the attached fatty acid side chain (Dickschat, *Nat. Prod. Rep.* 27:343-69 (2010), which is hereby incorporated by reference in its entirety) (Figure 82). The results herein indicate that nematodes secrete combinations from the same repertoire of ascarosides to present a specific chemical signature to their surrounding environment.

[0385] The shared structural organization and broad-reaching nature of both ascarosides and AHLs provide new insight into the syntax of biochemical communication networks. Many bacterial behaviors are mediated by quorum sensing, such as bioluminescence, biofilm formation, virulence factor expression, antibiotic production, and motility (Boyer & Wisnieski-Dye, *FEMS Microbiol. Ecol.* 1:1-19 (2009), which is hereby incorporated by reference in its entirety). Similarly, many *C. elegans* behaviors are mediated by ascarosides, such as mate finding, repulsion, aggregation, olfactory plasticity, and entry into a diapausal life stage (Srinivasan et al., *Nature* 454:1115-18 (2008); Jeong, et al., *Nature* 433:541-45 (2005); Butcher et al., *Nat. Chem. Biol.* 7:420-22 (2007); Pungaliya et al., *PNAS* 19:7708-13 (2009); Niblack et al., *Annu. Rev. Phytopathol.* 44:283-303 (2006); Macosko et al., *Nature* 458:1171-75 (2009); Yamada et al., *Science* 329:1647-50 (2010); Butcher et al., *Nat. Chem. Biol.* 3:420-22 (2007); Golden & Riddle, *Science* 218:578-80 (1982), which are hereby incorporated by reference in their entirety).

[0386] Furthermore, the ascaroside profile of *C. elegans* changes in response to growth and environmental perturbation (Kaplan et al., *Pub. Lib. Sci. ONE* 6(3):e17804 (2011), which is hereby incorporated by reference in its entirety). Considering the broad diversity of nematodes and nematode behaviors, the modular nature of ascarosides seems an appropriate strategy for using a single mechanism to convey the changing needs of an individual or population. Molecules that are structurally similar to bacterial AHLs can be

synthesized to develop a new class of antimicrobial drugs that interfere with bacterial communication (Schauder et al., *Genes Dev.* 15:1468-80 (2001), which is hereby incorporated by reference in its entirety). Similar studies in nematodes can enable the design of synthetic ascaroside blends to interfere with nematode reproduction and survival in parasite-host models.

[0387] Because nematodes are currently responsible for about \$100 billion of losses to global agriculture annually (JOSEPH A. VEECH: VISTAS ON NEMATOLOGY (Society of Nematologists, Inc., Hyatsville, MD, 1987), which is hereby incorporated by reference in its entirety) and infect 3 billion humans worldwide (Blumenthal & Davis, *Nat. Genet.* 36:1246-47 (2004), which is hereby incorporated by reference in its entirety), there exists significant need for parasitic control. The concept of using species-specific pheromones for the management of insect pests has driven over 50 years of insect pheromone research with > 100 pheromones utilized in pest management (Witzgall et al., *J. Chem. Ecol.* 36:80-100 (2010), which is hereby incorporated by reference in its entirety). However, pheromone-mediated interruption has only been practically applied to a single nematode pheromone, vanillic acid, towards control of the soybean parasitic nematode *Heterodera glycines* (Meyer et al., *J. Nematol.* 29:282-88 (1997), which is hereby incorporated by reference in its entirety). This is most likely attributed to the lack of nematode pheromone identification, aside from those discovered in *H. glycines* and *C. Elegans* (Jaffe et al., *J. Chem. Ecol.* 15:2031-43 (1989); Srinivasan et al., *Nature* 454:1115-18 (2008); Jeong, et al., *Nature* 433:541-45 (2005); Butcher et al., *Nat. Chem. Biol.* 7:420-22 (2007); Pungalija et al., *PNAS* 19:7708-13 (2009), which are hereby incorporated by reference in their entirety). The discovery of this broad-reaching, modular library of nematode pheromones can undoubtedly prove useful towards this necessary effort, while interpretation of the syntax can provide new insight into the evolution of biochemical signatures.

Example 84—Ascaroside Production in Root Knot Nematodes

[0388] Ascarosides ascr#10, ascr#16, ascr#18, ascr#20, and ascr#22 were detected in certain species of *Meloidogyne* plant parasitic nematodes (Figure 83) and identified using high-resolution mass spectrometry (Figures 84–88). It is predicted that these ascarosides and/or structural derivatives thereof (alone, in concert, and/or in concert with ascr#9 or other ascarosides) act as dispersants and repellants in *Meloidogyne* species. (It is also predicted that ascr#18 and ascr#22 and/or structural derivatives thereof (alone, in concert, and/or in concert with ascr#9 or other ascarosides) act as dispersants and repellants in *Heterorhabditis*.)

Example 85 – Ascaroside function

[0389] Various ascarosides and ascaroside blends were administered to various nematodes described below (Table 12). As outlined below, the ascarosides and ascaroside blends resulted in various nematode behaviors, such as attracting a male nematode, or promoting dauer formation. A very strong result of affecting nematode behavior is designated in the table as “strong.”

Table 12: Ascarosides/ascaroside blends function

ASCAROSIDE FUNCTION**Nematodes**

Model Nematodes: *Caenorhabditis elegans*, *Pristionchus pacificus*, *Panagrellus redivivus*

Plant-Parasitic: Root-Knot Nematodes (RKN), *Meloidogyne* spp. (second stage juveniles)

Entomopathogenic (insect-parasitic): *Heterorhabditis* spp., *Steinernema* spp., *Steinernema feltiae* (infectious juveniles), *S. glaseri*

	Repel Males	Repel Females/Herm.	Dispersal (of dauer larvae, infectious juveniles)	Attract Males	Attract Females/Herm.	Promote Dauer Formation
<i>C. elegans</i>		<i>ascr#3</i> <i>ascr#5</i> Blends <i>ascr#2/ascr#3/ascr#5</i> ([strong]) <i>ascr#3/icas#3</i> ([strong])	<i>ascr#2</i> <i>ascr#8</i> Blends <i>ascr#2or9/ascr#3/ascr#8/icas#9</i>	<i>ascr#1</i> <i>ascr#2</i> <i>ascr#3</i> <i>ascr#7</i> <i>ascr#8</i> <i>ascr#9</i> <i>ascr#10</i> <i>icas#1</i> ([strong]) <i>icas#3</i> ([strong]) <i>icas#9</i> ([strong]) <i>hbas#3</i> ([strong]) Blends <i>ascr#2/ascr#3</i> ([strong]) <i>ascr#2/ascr#3/ascr#4</i> ([strong]) <i>ascr#2/ascr#3/ascr#8</i> ([very strong])	Indoles <i>icas#1</i> ([strong]) <i>icas#3</i> ([strong]) <i>icas#9</i> ([strong]) <i>hbas#3</i> ([very strong]) Blends <i>ascr#3/icas#3</i>	<i>ascr#1</i> <i>ascr#2</i> ([strong]) <i>ascr#3</i> ([strong]) <i>ascr#4</i> ([strong]) <i>ascr#5</i> ([strong]) <i>ascr#8</i> ([strong]) <i>icas#9</i> Blends <i>ascr#2/a</i> <i>scr#3/ascr#5</i> ([strong]) <i>ascr#2/a</i> <i>scr#3/ascr#5/ascr#8</i>
<i>C. brenneri</i> <i>C. remanei</i>				Blends <i>ascr#2/ascr#3/ascr#4</i>		
<i>C. briggsae</i> <i>C. japonica</i>				Blends <i>ascr#2/ascr#3/ascr#4</i>		
<i>Panagrellus redivivus</i>	<i>oscr#10</i>	<i>icas#9</i>		<i>ascr#1</i> <i>ascr#3</i> <i>ascr#7</i> <i>ascr#8</i> <i>ascr#9</i> <i>ascr#10</i> <i>oscr#9</i>	<i>ascr#8</i>	

	Repel Males	Repel Females/Herm.	Dispersal (of dauer larvae, infectious juveniles)	Attract Males	Attract Females/Herm.	Promote Dauer Formation
<i>Meloidogyne javanica</i> <i>M. floricola</i> <i>M. incognita</i> <i>M. hapla</i>		ascr#9 ascr#10* ascr#16* ascr#18* ascr#20* ascr#22* Blends of the above including ascr#9 ([strong])*	ascr#9 ascr#10* ascr#16* ascr#18* ascr#20* ascr#22* Blends ascr#2or9/ascr#3/ ascr#8/icas#9			
<i>Steinernema riobrave</i> <i>S. diaprepesi</i> <i>S. carpocapse</i> <i>S. feltiae</i>			ascr#9 ascr#11 ascr#2 ascr#3 ascr#8 ascr#9 icas#9 Blends ascr#2or9/ascr#3/ ascr#8/icas#9 ascr#9/ ascr#11 ascr#9/oscr#10 ascr#9/oscr#10/ascr#11			
<i>S. glaseri</i>				ascr#1 ascr#3 ascr#7 ascr#8 ascr#10		
<i>Heterorhabditis bacteriophora</i> <i>H. zealandica</i> <i>H. floricola</i>		ascr#18* ascr#22*	ascr#9 ascr#18* ascr#22*			

Example 86 – Ascarosides produced

[0390] Examples of various ascarosides that are produced by species of nematodes as outlined below (Table 13). Examples of sources of the ascarosides are provided by the species of nematode as well as the type of nematode.

Table 13: Ascarosides produced by species of nematode

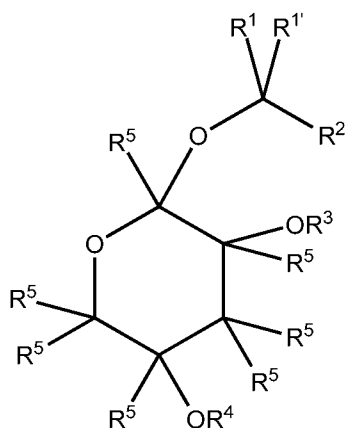
Type	Species	Ascarosides Produced
	<i>Koernia</i> sp.	ascr#1, ascr#3, ascr#14
	<i>Rhabditis</i> sp.7	Adult: ascr#1, ascr#3, ascr#7, ascr#9, ascr#10, ascr#12, ascr#14, ascr#18, icas#9, oscr#9 Dauer: ascr#1, ascr#3, ascr#10, ascr#12, ascr#14, ascr#18, icas#9

Animal parasite (rodent)	<i>Nippostrongylus brasiliensis</i>	ascr#1, ascr#3, ascr#10 Adult: ascr#3, ascr#7, ascr#10 IJ: ascr#3, ascr#10
Animal parasite (skin)	<i>Pelodera strongyloides</i>	ascr#18, ascr#20, ascr#22, ascr#24, ascr#26
Insect parasite	<i>Heterorhabditis bacteriophora</i>	ascr#9, ascr#12, ascr#18, ascr#20, ascr#22, ascr#24, ascr#26
Insect parasite	<i>Steinernema carpocapsae</i>	Adult: ascr#1, ascr#9, ascr#10, ascr#11, ascr#12, ascr#18, ascr#22 IJ: ascr#9, ascr#10
Insect parasite	<i>Steinernema glaseri</i>	Adult: ascr#1, ascr#9, ascr#10, ascr#12, ascr#14 IJ: ascr#9, ascr#10, ascr#11, ascr#12
Insect parasite	<i>Steinernema riobrave</i>	IJ: ascr#9, ascr#10, ascr#14, ascr#16, ascr#18
Insect associated	<i>Pristionchus pacificus</i>	ascr#1, ascr#9, ascr#12
Compost dweller	<i>Panagrellus redivivus</i>	ascr#1, ascr#3, ascr#9, ascr#10, ascr#14, ascr#18
Compost dweller	<i>Rhabditis sp.</i>	ascr#1, ascr#9, ascr#10, ascr#12, ascr#9, ascr#10
Soil dweller	<i>Oscheius tipulae</i>	ascr#9, ascr#12

[0391] Although preferred embodiments have been depicted and described in detail herein, it will be apparent to those skilled in the relevant art that various modifications, additions, substitutions, and the like can be made without departing from the spirit of the invention and these are therefore considered to be within the scope of the invention as defined in the claims which follow.

CLAIMS

1. A method of modifying behavior of a nematode, comprising:
providing a composition comprising a compound of the formula:



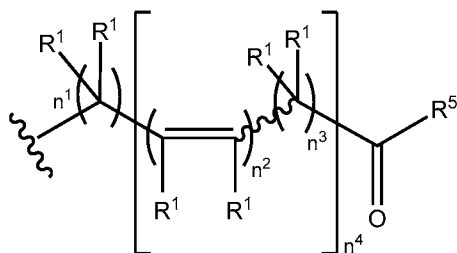
(Formula I), or a pharmaceutical equivalent, derivative,

analog and/or salt thereof; and

administering an effective dosage of the composition to the nematode;

wherein the nematode is in contact with a plant.

2. The method of claim 1, wherein R^1 is H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl; wherein each R is independently H, halogen, an alkyl, or an alkenyl.
3. The method of claim 1, wherein $R^{1'}$ is absent, H, $-C(R)_3$, $-OR$, $-N(R)_2$, halogen, an alkyl, a haloalkyl, an alkenyl, or a haloalkenyl; wherein each R is independently H, halogen, an alkyl, or an alkenyl.
4. The method of claim 1, wherein R^2 is a moiety of the formula:



(Formula II), or a pharmaceutical equivalent,

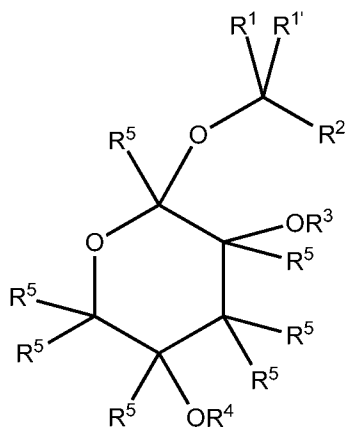
derivative, analog and/or salt thereof.

5. The method of claim 1, wherein R^3 is H, $-CR^6R^7R^8$, $-C(O)R^8$, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R^5 of another unit of Formula I.
6. The method of claim 1, wherein R^4 is H, $-CR^6R^7R^8$, $-C(O)R^8$, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R^5 of another unit of Formula I.
7. The method of claim 1, wherein R^5 is H, $-OH$, $-OR^6$, $-OCR^6R^7R^8$, $-CR^6R^7R^8$, $-NH_2$, $-NHR^6$, $-NR^6R^7$, halogen, an alkyl, a haloalkyl, an alkenyl, a haloalkenyl, an aryl, a heteroaryl, an arylalkyl, a heterocyclyl, a cycloalkyl, a cycloalkenyl, an acyl, an amino acid, a nucleoside, a monosaccharide having 5 or 6 carbon atoms, or a bond connecting to R^3 or R^4 of another unit of Formula I.
8. The method of claim 1, wherein the nematode is not a *Caenorhabditis* species.
9. The method of claim 1, wherein the nematode is selected from the group consisting of *Acuarioidea*, *Aelurostrongylus*, *Aelurostrongylus abstrusus*, *Amidostomatidae*, *Amidostomum*, *Ancylostoma braziliense*, *Ancylostoma caninum* (dog hookworm), *Ancylostoma ceylanicum*, *Ancylostoma duodenale*, *Ancylostoma tubaeforme*, *Ancylostomatidae*, *Ancylostomatinae*, *Angiostrongylidae*, *Angiostrongylus*, *Aproctoidea*, *Ascaridia galli*, *Ascaris lumbricoides*, *Ascaris suum*, *Brevistriatinae*, *Brugia malayi*, *Brugia timori*, *Bunostominae*, *Camallanoidea*, *Carolinensis minutus*, *Chabertia*, *Chabertia ovina*, *Chabertiidae*, *Cloacina*, *Cloacinidae*, *Cooperia*, *Cooperia pectinata*, *Cooperia punctata*, *Cooperiidae*, *Cosmocercoidea*, *Crenosoma*, *Crenosomatidae*, *Cyathostoma*, *Cyathostominae*, *Cyclodontostomum*, *Cylicocycylus nassatus*, *Cystocaulus*, *Cystocaulus ocreatus*, *Deletrocephalidae*, *Deletrocephalus*, *Diaphanocephalidae*, *Diaphanocephaloidea*, *Dictyocaulidae*, *Dictyocaulinae*, *Dictyocaulus arnfeldi*, *Dictyocaulus filaria*, *Dictyocaulus osleri*, *Dictyocaulus viviparus*, *Dictyocausus viviparous*, *Didelphostrongylus*, *Dioctophyma renale*,

Diectophymatoidea, *Diplotriaeñoidea*, *Dirofilaria immitis*, *Dracunculoidea*,
Dromaeostrongylidae, *Elaeophora scheideri*, *Elaphostrongylinae*, *Filarinema*,
Filarioidea, *Filaroides*, *Filaroididae*, Gapeworm, *Ghathostomatoidea*,
Globocephaloidinae, *Gongylonema pulchrum*, *Gyalocephalinae*, *Habronema*,
Habronematoidea, *Haemonchidae*, *Haemonchinae*, *Haemonchus contortus*,
Haemonchus placei, *Halocercus*, *Heligmonellidae*, *Heligmonellinae*, *Heligmonoides*
speciosus, *Heligmosomatidae*, *Heligmosomidae*, *Heligmosomoidea*,
Heligmosomoides, *Herpetostrongylidae*, *Herpetostrongylinae*, *Heterakoidea*,
Hovorkonema, *Hypodontus*, *Kalicephalus*, *Labiomultiplex*, *Labiosimplex*,
Labiostrongylus, *Libyostrongylinae*, *Loa loa*, *Longistriata*, *Mackerrastrongylidae*,
Macropostrongylus, *Mansonella ozzardi*, *Mansonella perstans*,
Mansonella streptocerca, *Marshallagia*, *Metastrongylidae*, *Metastrongyloidea*
(lungworms), *Metastrongyloidea* sp. RJ-2010, *Metastrongylus*, *Metastrongylus apri*,
Metastrongylus asymmetricus, *Metastrongylus confusus*, *Metastrongylus elongates*,
Metastrongylus pudendotectus, *Metastrongylus salmi*, *Molineidae*, *Molineoidea*,
Monilonema, *Muellerinae*, *Muellerius capillaris*, *Muspiceoidea*, *Nematodirinae*,
Nematodirus battus, *Neoheligionella granjoni*, *Nicollina*, *Nicollinidae*,
Nippostrongylinae, *Nippostrongylus brasiliensis*, *Oesophagostomum*,
Oesophagostomum columbianum, *Oesophagostomum radiatum*, *Ohbayashinema*,
Oncocerca volvulus, *Orientostrongylus ezoensis*, *Oslerus*, *Oslerus osleri*, *Ostertagia*
ostertagi, *Ostertagia venulosum*, *Otostrongylus*, *Oxyuroidea*, *Papillostrongylus*,
Paraelaphostrongylus tenuis, *Parafilaroides*, *Parazoniolaimus*, *Physalopteroidea*,
Potorostrongylus, *Protostrongylidae*, *Protostrongylinae*, *Pseudaliidae*, *Pseudalius*,
Rictularioidea, *Rugopharynx*, *Setaria cervi*, *Seuratoidea*, *Skrjabingylus*, *Spiruroidea*,
Stenurus, *Stephanofilaria stilesi*, *Stephanuridae*, *Stephanurus*, *Strongylida*,
Strongylida sp. AM-2008, *Strongylidae*, *Strongylinae*, *Strongyloidea*, *Strongyloides*
papillosus, *Subuluroidea*, *Syngamidae*, *Syngamus*, *Teladorsagia circumcincta*,
Ternidens, *Tetrabothriostrongylus*, *Thelazioidea*, *Torynurus*, *Toxocara canis*,
Toxocara cati, *Toxocara vitulorum*, *Trichinella spiralis*, *Trichinelloidea*,
Trichostrongylus axei, *Trichostrongylus colubriformis*, *Trichostrongylus vitrinus*,
Trichostrongylidae, *Trichostrongylinae*, *Trichostrongyloidea*, *Trichuris suis*,
Troglostrongylus, *Umingmakstrongylus pallikuukensis*, *Unclassified*
Metastrongyloidea, *unclassified Protostrongylidae*, *unclassified Strongylida*,

unclassified Trichostrongylidae, Varestrongylinae, Wucheria bancrofti,
and *Zoniolaimus*.

10. The method of claim 1, wherein the compound includes at least one nucleobase.
11. The method of claim 1, wherein the compound includes at least one fatty acid.
12. The method of claim 1, wherein the compound includes at least one amino acid.
13. The method of claim 1, wherein the compound includes at least one sugar.
14. The method of claim 1, wherein the nematode behavior is selected from the group consisting of reproduction, development, dauer formation, aggregation, attraction, repulsion, dispersal, deterrence, feeding, host finding, and host invasion.
15. The method of claim 1, wherein the plant is selected from the group consisting of dicots, monocots, crop plants, alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, green bean, wax bean, lima bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, sugar beet, parsnip, turnip, cauliflower, broccoli, turnip, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, sugarcane, ornamental plants, *Arabidopsis thaliana*, Saintpaulia, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.
16. A method of promoting or inhibiting reproduction in a nematode in contact with a plant, comprising:
providing a composition comprising a compound of the formula:



(Formula I), or a pharmaceutical equivalent, derivative,

analog and/or salt thereof; and

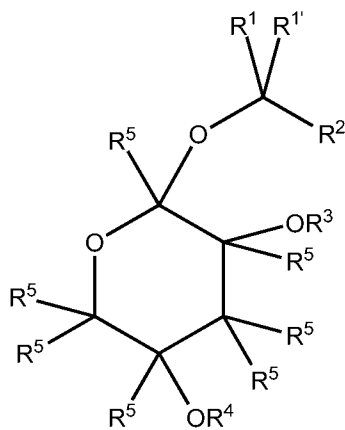
administering an effective dosage of the composition to the nematode in contact with the plant.

17. The method of claim 16, wherein the composition comprises one or more compounds selected from the group consisting of ascr#1, ascr#2, ascr#3, ascr#7, ascr#8, ascr#9, ascr#10, ascr#11, ascr#12, ascr#14, ascr#16, ascr#18, ascr#20, ascr#22, ascr#24, ascr#26, icas#9, hbas#3, and oscr#9.
18. The method of claim 16, wherein the composition comprises ascr#1, ascr#2, ascr#3, ascr#4, ascr#5, ascr#7, ascr#8, ascr#10, icas#9, hbas#3, and combinations thereof.
19. The method of claim 18, wherein the nematode is *Caenorhabditis elegans*.
20. The method of claim 16, wherein the composition comprises ascr#2, ascr#3, ascr#4, and combinations thereof.
21. The method of claim 20, wherein the nematode is *Caenorhabditis brenneri*, *Caenorhabditis remanei*, *Caenorhabditis briggsae*, or *Caenorhabditis japonica*.
22. The method of claim 16, wherein the composition comprises ascr#1, ascr#3, ascr#7, ascr#8, ascr#9, ascr#10, oscr#9, oscr#10, icas#9, and combinations thereof.
23. The method of claim 22, wherein the nematode is *Panagrellus redivivus*.

24. The method of claim 16, wherein the composition comprises ascr#2, ascr#3, ascr#8, icas#9, ascr#9, ascr#10, ascr#16, ascr#18, ascr#20, ascr#22, and combinations thereof.
25. The method of claim 24, wherein the nematode is *Meloidogyne javanica*, *Meloidogyne floridensis*, *Meloidogyne incognita*, or *Meloidogyne hapla*.
26. The method of claim 16, wherein the composition comprises ascr#9, ascr#11, and combinations thereof.
27. The method of claim 26, wherein the nematode is *Steinernema riobrave*, *Steinernema diaprepesi*, or *Steinernema carpocapse*.
28. The method of claim 16, wherein the composition comprises ascr#2, ascr#3, ascr#8, ascr#9, icas#9, ascr#10, ascr#11, and combinations thereof.
29. The method of claim 28, wherein the nematode is *Steinernema feltiae*.
30. The method of claim 16, wherein the composition comprises ascr#1, ascr#3, ascr#7, ascr#8, ascr#10, and combinations thereof.
31. The method of claim 30, wherein the nematode is *Steinernema glaseri*.
32. The method of claim 16, the composition comprises ascr#9, ascr#18, ascr#22, and combinations thereof.
33. The method of claim 32, wherein the nematode is *Heterorhabditis bacteriophora*, *Heterorhabditis zealandica*, or *Heterorhabditis floridensis*.
34. The method of claim 16, wherein the composition comprises ascr#1, ascr#3, ascr#7, ascr#10, ascr#18, ascr#20, ascr#22, ascr#24, ascr#26, and combinations thereof.
35. The method of claim 34, wherein the nematode is *Nippostrongylus brasiliensis* or *Pelodera strongyloides*.
36. A method of promoting or inhibiting nematode aggregation at a first location in a plant, said method comprising:

- (i) contacting the first location with one or more isolated modulator compounds under conditions effective to promote or inhibit nematode aggregation at the first location, or
- (ii) contacting a second location with one or more isolated modulator compounds under conditions effective to promote or inhibit nematode aggregation at the first location, wherein the first location and the second location are spaced to permit said contacting at the second location to have an effect on nematode aggregation at the first location;

wherein the compound comprises the formula:



(Formula I), or a pharmaceutical equivalent, derivative, analog and/or salt thereof.

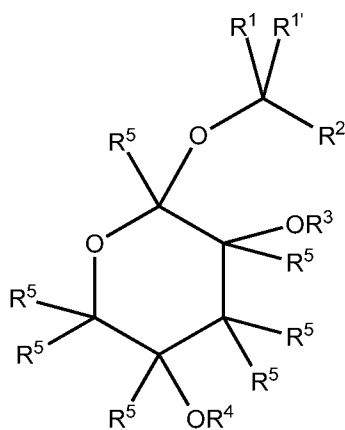
37. The method of claim 36, wherein the one or more isolated modulator compounds is selected from the group consisting of ascr#1, ascr#2, ascr#3, ascr#7, ascr#8, ascr#9, ascr#10, ascr#11, ascr#12, ascr#14, ascr#16, ascr#18, ascr#20, ascr#22, ascr#24, ascr#26, icas#9, hbas#3, and oscr#9.

38. The method of claim 36, wherein the second location is in the plant.

39. The method of claim 36, wherein the plant is selected from the group consisting of dicots, monocots, crop plants, alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, green bean, wax bean, lima bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, sugar beet, parsnip, turnip, cauliflower, broccoli, turnip, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry,

grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, sugarcane, ornamental plants, *Arabidopsis thaliana*, Saintpaulia, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.

40. The method of claim 36, wherein said contacting comprises high pressure spraying, low pressure spraying, immersion, atomizing, foaming, fogging, coating, or encrusting.
41. The method of claim 36, further comprising contacting the second location with a toxin that is harmful to the nematode.
42. The method of claim 36, wherein the first location is at a population of insects.
43. The method of claim 42, wherein the insects are selected from the group consisting of European corn borer, beet armyworm, cabbage looper, corn ear worm, fall armyworm, diamondback moth, cabbage root maggot, onion maggot, seed corn maggot, pickleworm, pepper maggot, and tomato pinworm.
44. A method of reducing the likelihood or preventing parasite infection of a plant, said method comprising:
providing a composition comprising a compound of the formula:



(Formula I), or a pharmaceutical equivalent, derivative,

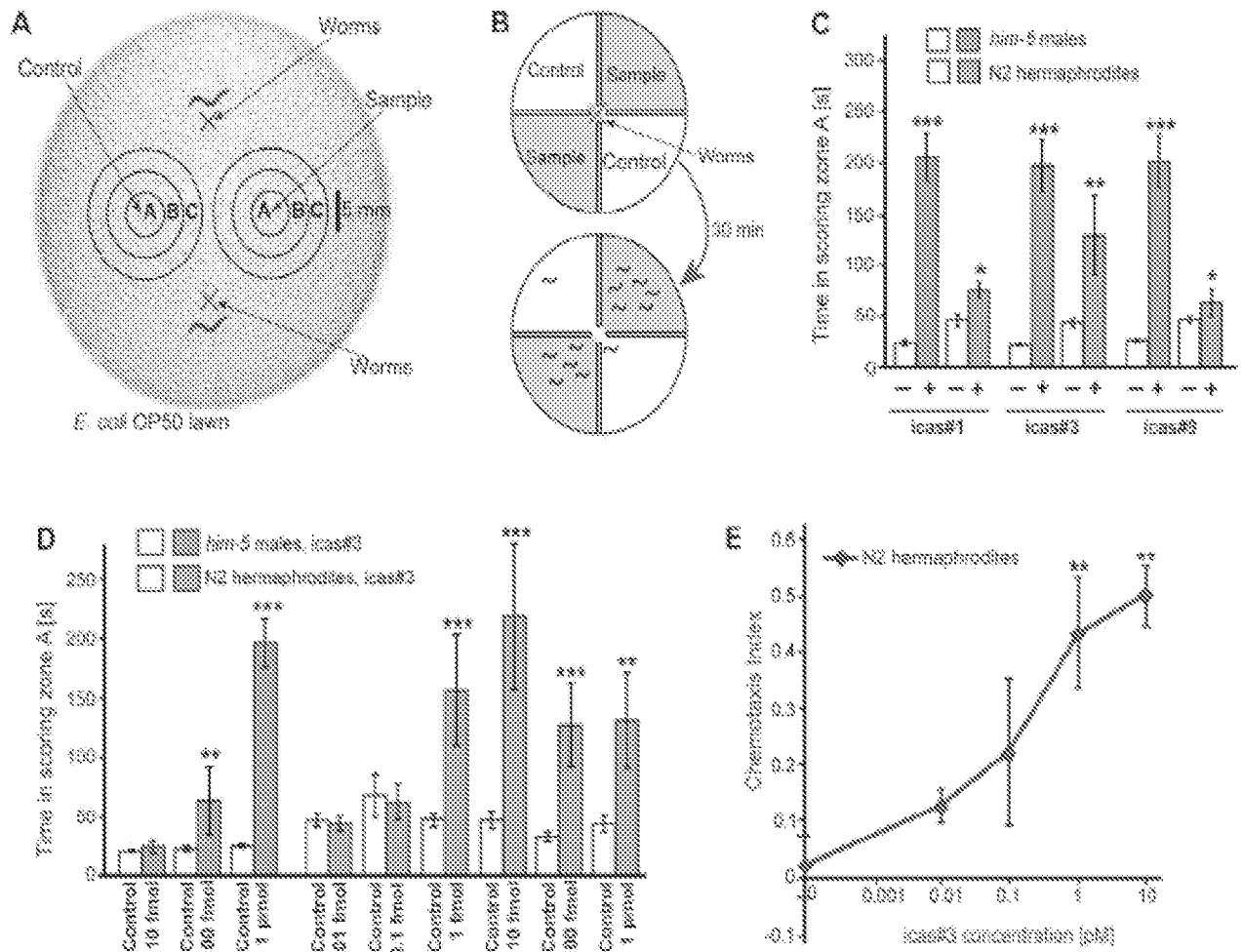
analog and/or salt thereof; and

administering an amount of the composition to the plant sufficient to reduce the likelihood or prevent parasite infection of the plant.

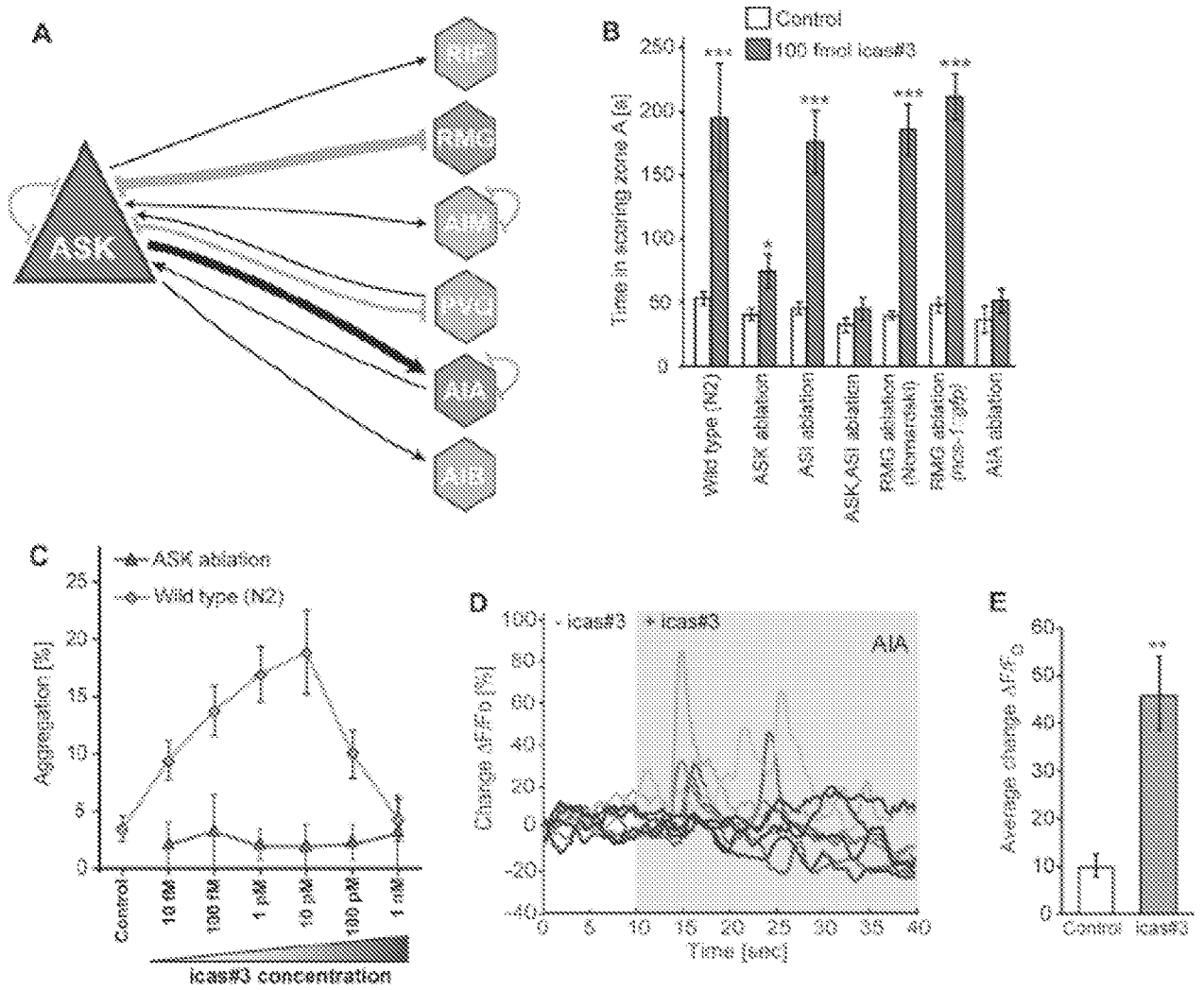
45. The method of claim 44, wherein the parasite is a nematode or an insect.
46. The method of claim 44, wherein the composition comprises one or more compounds selected from the group consisting of ascr#1, ascr#2, ascr#3, ascr#7, ascr#8, ascr#9, ascr#10, ascr#11, ascr#12, ascr#14, ascr#16, ascr#18, ascr#20, ascr#22, ascr#24, ascr#26, icas#9, hbas#3, and oscr#9.
47. The method of claim 44, wherein the plant is selected from the group consisting of dicots, monocots, crop plants, alfalfa, rice, wheat, barley, rye, cotton, sunflower, peanut, corn, potato, sweet potato, bean, green bean, wax bean, lima bean, pea, chicory, lettuce, endive, cabbage, brussel sprout, beet, sugar beet, parsnip, turnip, cauliflower, broccoli, turnip, radish, spinach, onion, garlic, eggplant, pepper, celery, carrot, squash, pumpkin, zucchini, cucumber, apple, pear, melon, citrus, strawberry, grape, raspberry, pineapple, soybean, tobacco, tomato, sorghum, sugarcane, ornamental plants, *Arabidopsis thaliana*, Saintpaulia, petunia, pelargonium, poinsettia, chrysanthemum, carnation, and zinnia.
48. The method of claim 44, wherein the parasite is an insect.
49. The method of claim 44, wherein the insect is selected from the group consisting of European corn borer, beet armyworm, cabbage looper, corn ear worm, fall armyworm, diamondback moth, cabbage root maggot, onion maggot, seed corn maggot, pickleworm, pepper maggot, and tomato pinworm.
50. The method of claim 44, wherein the parasite is selected from the group consisting of *Acuarioidea*, *Aelurostrongylus*, *Aelurostrongylus abstrusus*, *Amidostomatidae*, *Amidostomum*, *Ancylostoma braziliense*, *Ancylostoma caninum* (dog hookworm), *Ancylostoma ceylanicum*, *Ancylostoma duodenale*, *Ancylostoma tubaeforme*, *Ancylostomatidae*, *Ancylostomatinae*, *Angiostrongylidae*, *Angiostrongylus*, *Aproctoidea*, *Ascaridia galli*, *Ascaris lumbricoides*, *Ascaris suum*, *Brevistriatinae*, *Brugia malayi*, *Brugia timori*, *Bunostominae*, *Camallanoidea*, *Carolinensis minutus*, *Chabertia*, *Chabertia ovina*, *Chabertiidae*, *Cloacina*, *Cloacinidae*, *Cooperia*, *Cooperia pectinata*, *Cooperia punctata*, *Cooperiidae*, *Cosmocercoidea*, *Crenosoma*,

Crenosomatidae, *Cyathostoma*, *Cyathostominae*, *Cyclodontostomum*,
Cylicocyclus nassatus, *Cystocaulus*, *Cystocaulus ocreatus*, *Deletrocephalidae*,
Deletrocephalus, *Diaphanocephalidae*, *Diaphanocephaloidea*, *Dictyocaulidae*,
Dictyocaulinae, *Dictyocaulus arnfeldi*, *Dictyocaulus filaria*, *Dictyocaulus osleri*,
Dictyocaulus viviparus, *Dictyocaulus viviparus*, *Didelphostrongylus*, *Dioctophyma*
renale, *Dioctophymatoidea*, *Diplotrriaenoidea*, *Dirofilaria immitis*, *Dracunculoidea*,
Dromaeostrongylidae, *Elaeophora scheideri*, *Elaphostrongylinae*, *Filarinema*,
Filarioidea, *Filaroides*, *Filaroididae*, *Gapeworm*, *Ghathostomatoidea*,
Globocephaloidinae, *Gongylonema pulchrum*, *Gyalocephalinae*, *Habronema*,
Habronematoidea, *Haemonchidae*, *Haemonchinae*, *Haemonchus contortus*,
Haemonchus placei, *Halocercus*, *Heligmonellidae*, *Heligmonellinae*, *Heligmonoides*
speciosus, *Heligmosomatidae*, *Heligmosomidae*, *Heligmosomoidea*,
Heligmosomoides, *Herpetostrongylidae*, *Herpetostrongylinae*, *Heterakoidea*,
Hovorkonema, *Hypodontus*, *Kalicephalus*, *Labiomultiplex*, *Labiosimplex*,
Labiostrongylus, *Libyostrongylinae*, *Loa loa*, *Longistriata*, *Mackerrastrongylidae*,
Macropostrongylus, *Macropostrongylus*, *Mansonella ozzardi*, *Mansonella perstans*,
Mansonella streptocerca, *Marshallagia*, *Metastrongylidae*, *Metastrongyloidea*
(lungworms), *Metastrongyloidea sp. RJ-2010*, *Metastrongylus*, *Metastrongylus apri*,
Metastrongylus asymmetricus, *Metastrongylus confusus*, *Metastrongylus elongates*,
Metastrongylus pudendotectus, *Metastrongylus salmi*, *Molineidae*, *Molineoidea*,
Monilonema, *Muellerinae*, *Muellerius capillaris*, *Muspiceoidea*, *Nematodirinae*,
Nematodirus battus, *Neohelimonella granjoni*, *Nicollina*, *Nicollinidae*,
Nippostrongylinae, *Nippostrongylus brasiliensis*, *Oesophagostomum*,
Oesophagostomum columbianum, *Oesophagostomum radiatum*, *Ohbayashinema*,
Oncocerca volvulus, *Orientostrongylus ezoensis*, *Oslerus*, *Oslerus osleri*, *Ostertagia*
ostertagi, *Ostertagia venulosum*, *Otostrongylus*, *Oxyuroidea*, *Papillostrongylus*,
Paraelaphostrongylus tenuis, *Parafilaroides*, *Parazoniolaimus*, *Physalopteroidea*,
Potorostrongylus, *Protostrongylidae*, *Protostrongylinae*, *Pseudaliidae*, *Pseudalius*,
Rictularioidea, *Rugopharynx*, *Setaria cervi*, *Seuratoidea*, *Skrjabingylus*, *Spiruroidea*,
Stenurus, *Stephanofilaria stilesi*, *Stephanuridae*, *Stephanurus*, *Strongylida*,
Strongylida sp. AM-2008, *Strongylidae*, *Strongylinae*, *Strongyloidea*, *Strongyloides*
papillosus, *Subuluroidea*, *Syngamidae*, *Syngamus*, *Teladorsagia circumcincta*,
Ternidens, *Tetrabothriostrongylus*, *Thelazioidea*, *Torynurus*, *Toxocana canis*,

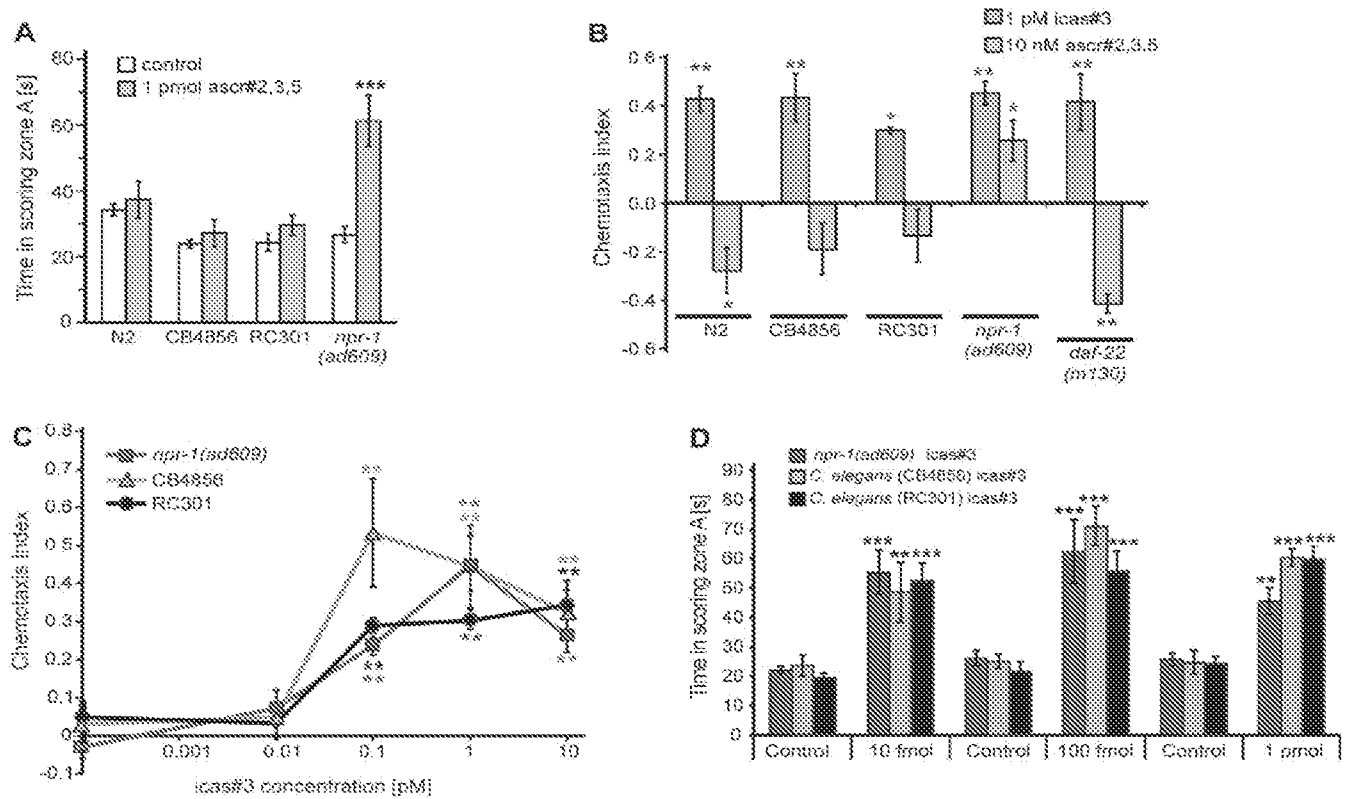
Toxocara cati, *Toxocara vitulorum*, *Trichinella spiralis*, *Trichinelloidea*,
Trichostronglyus axei, *Trichostronglyus colubriiformis*, *Trichostronglyus vitrinus*,
Trichostrongylidae, *Trichostrongylinae*, *Trichostrongyloidea*, *Trichuris suis*,
Troglostrongylus, *Umingmakstrongylus pallikuukensis*, *Unclassified*
Metastrongyloidea, *unclassified Protostrongylidae*, *unclassified Strongylida*,
unclassified Trichostrongylidae, *Varestrongylinae*, *Wucheria bancrofti*, and
Zoniolaimus.



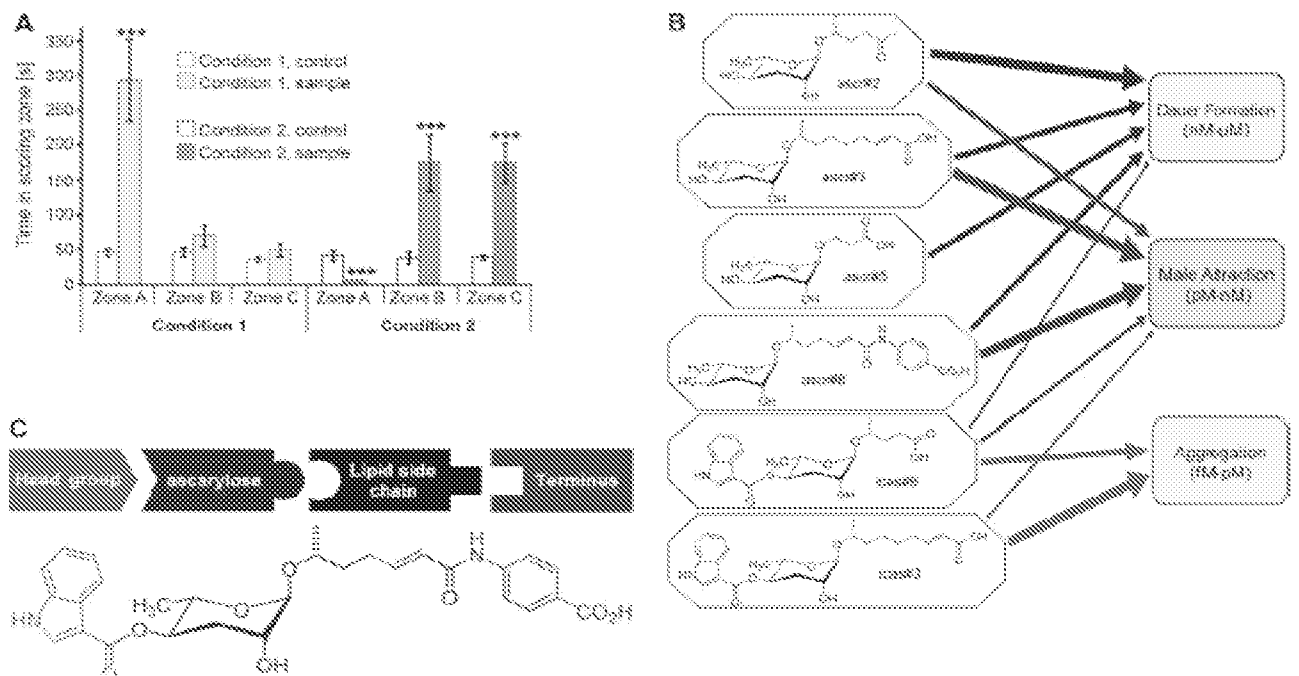
Figures 1A–E



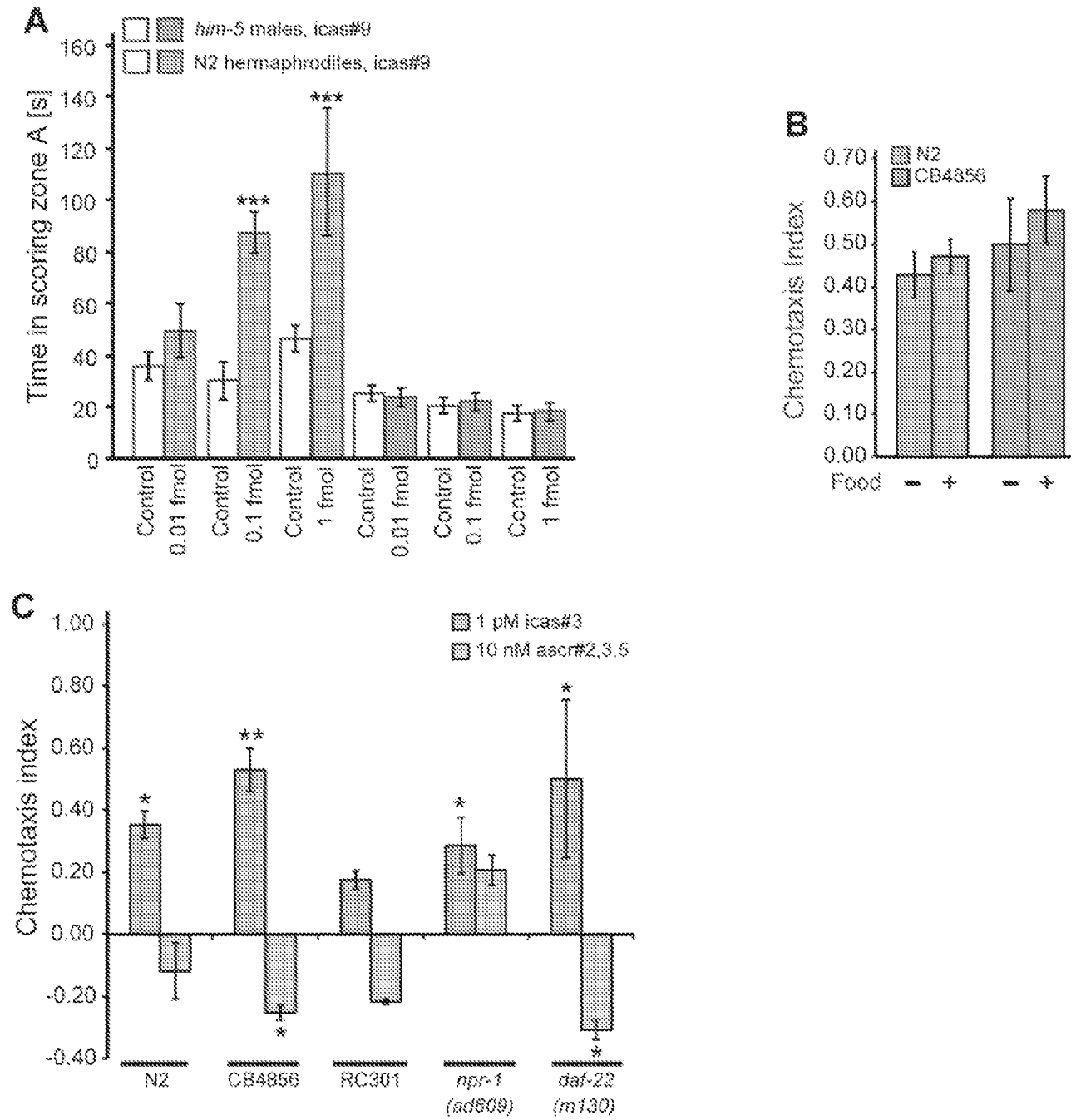
Figures 2A–E



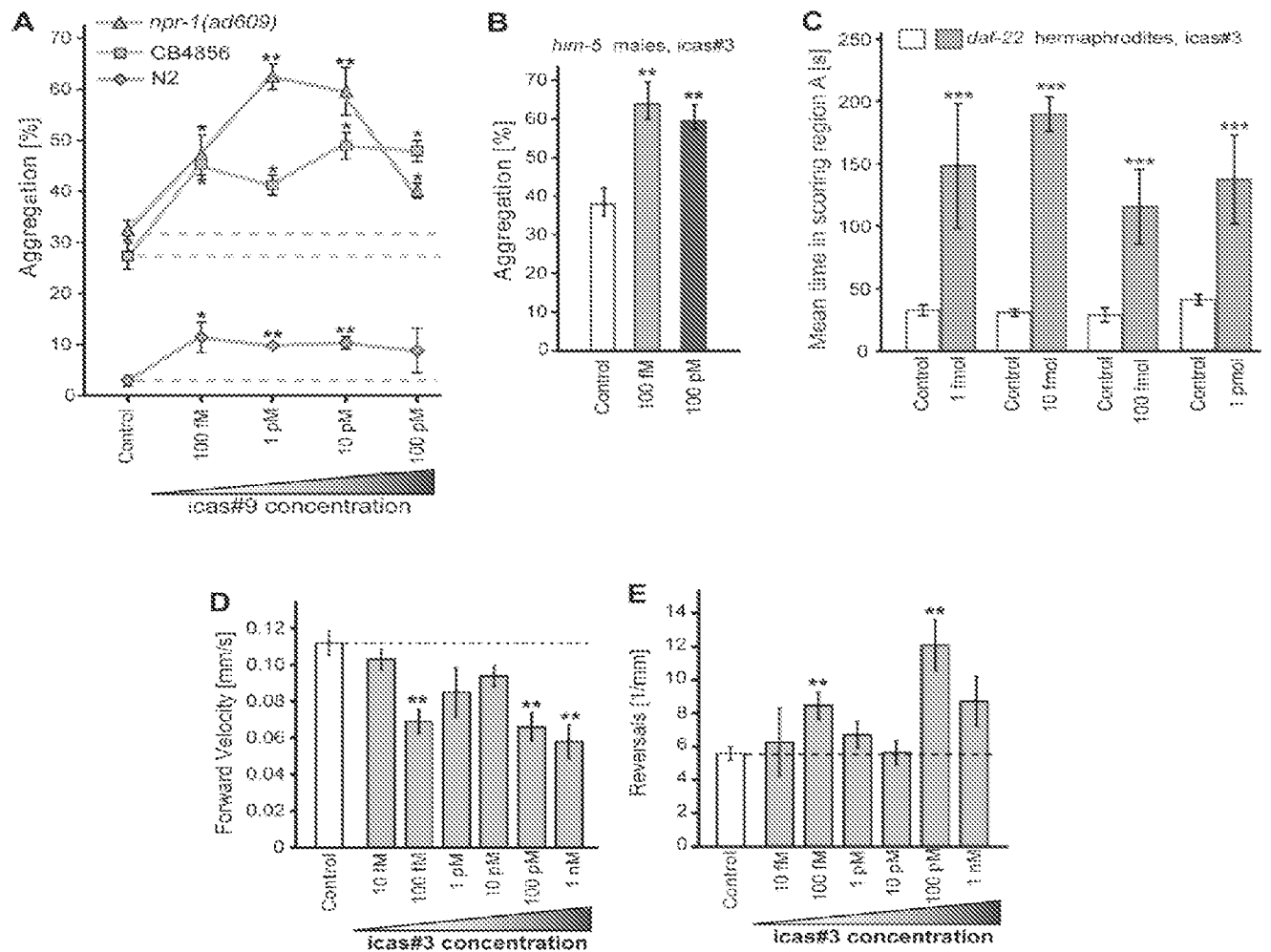
Figures 3A–D



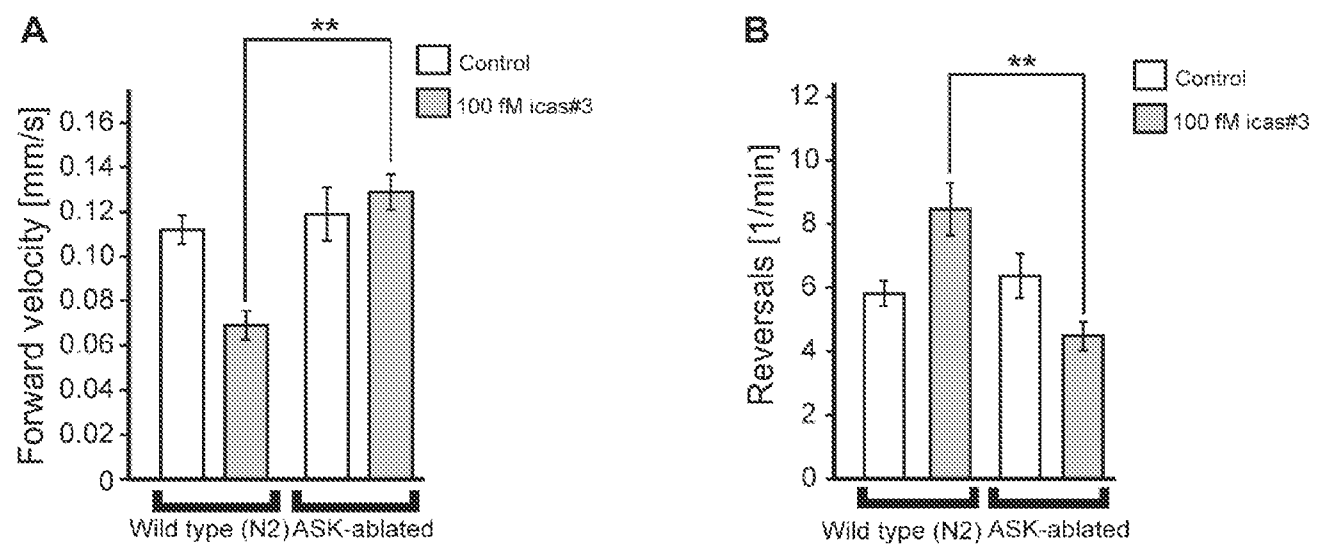
Figures 4A–C



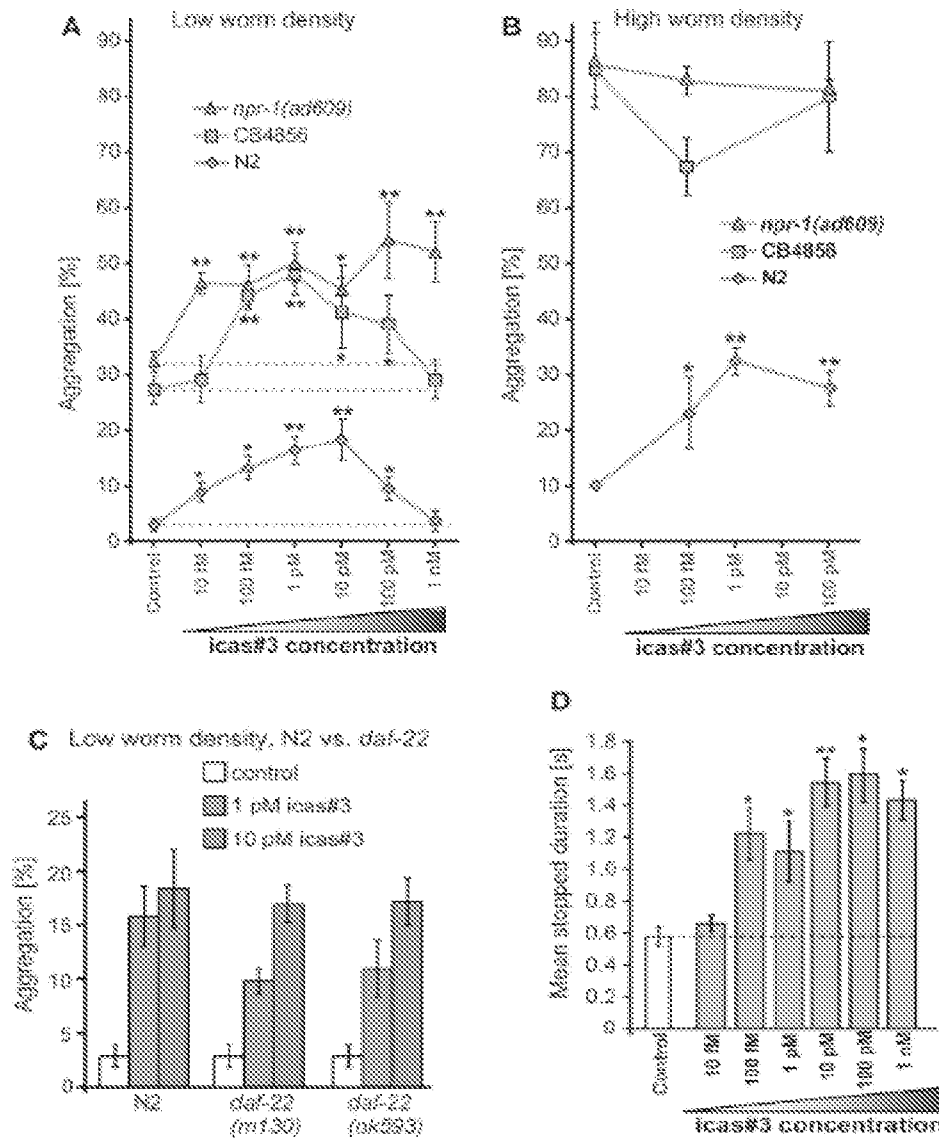
Figures 5A–C



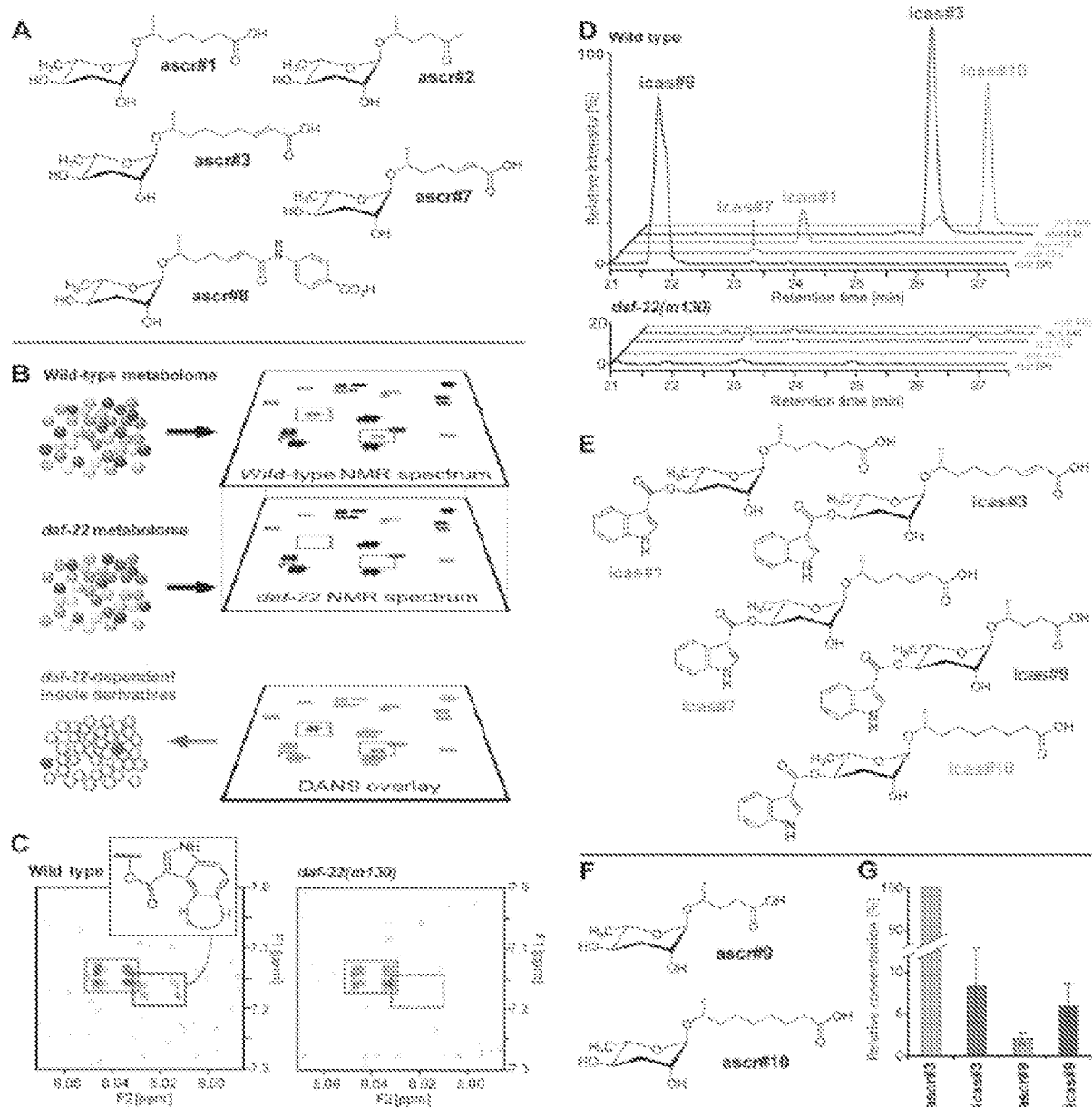
Figures 6A–E



Figures 7A–B



Figures 8A-D



Figures 9A–G

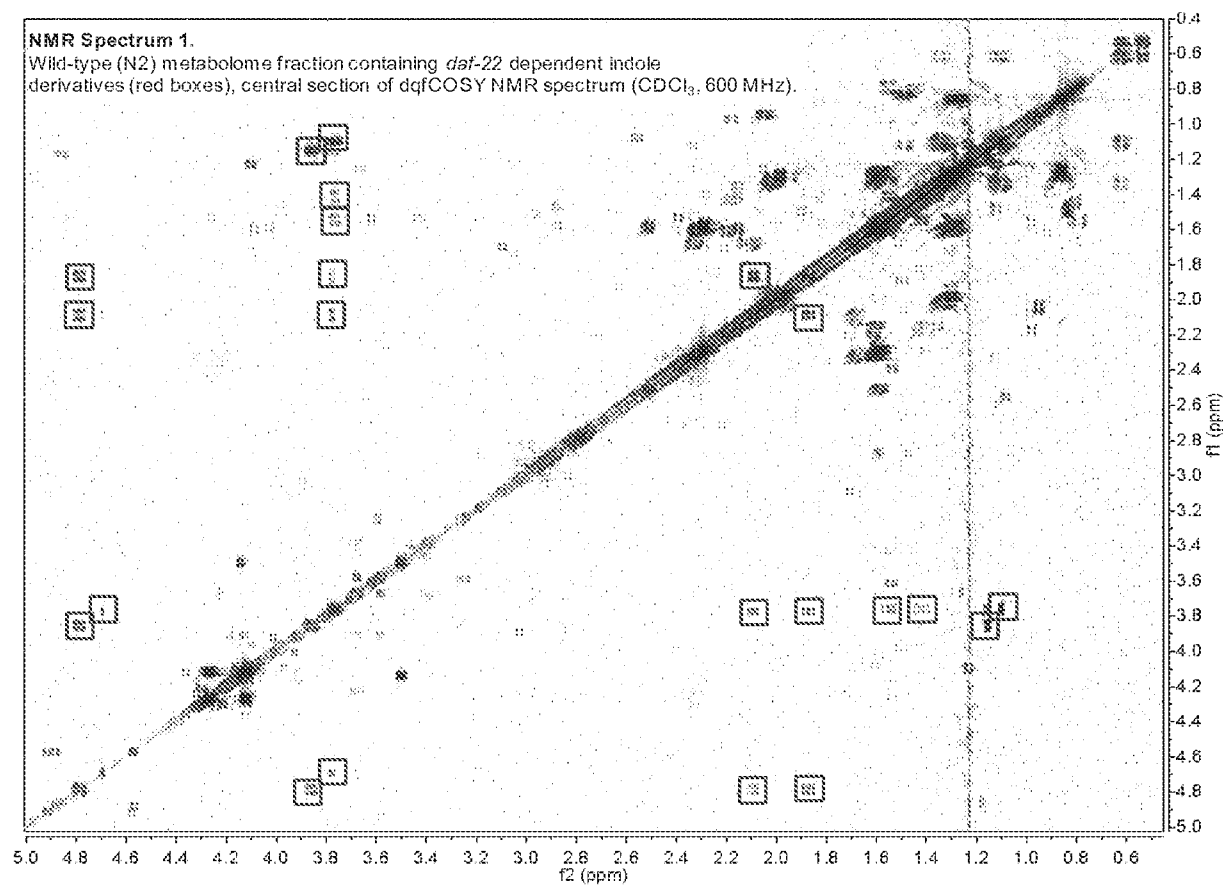
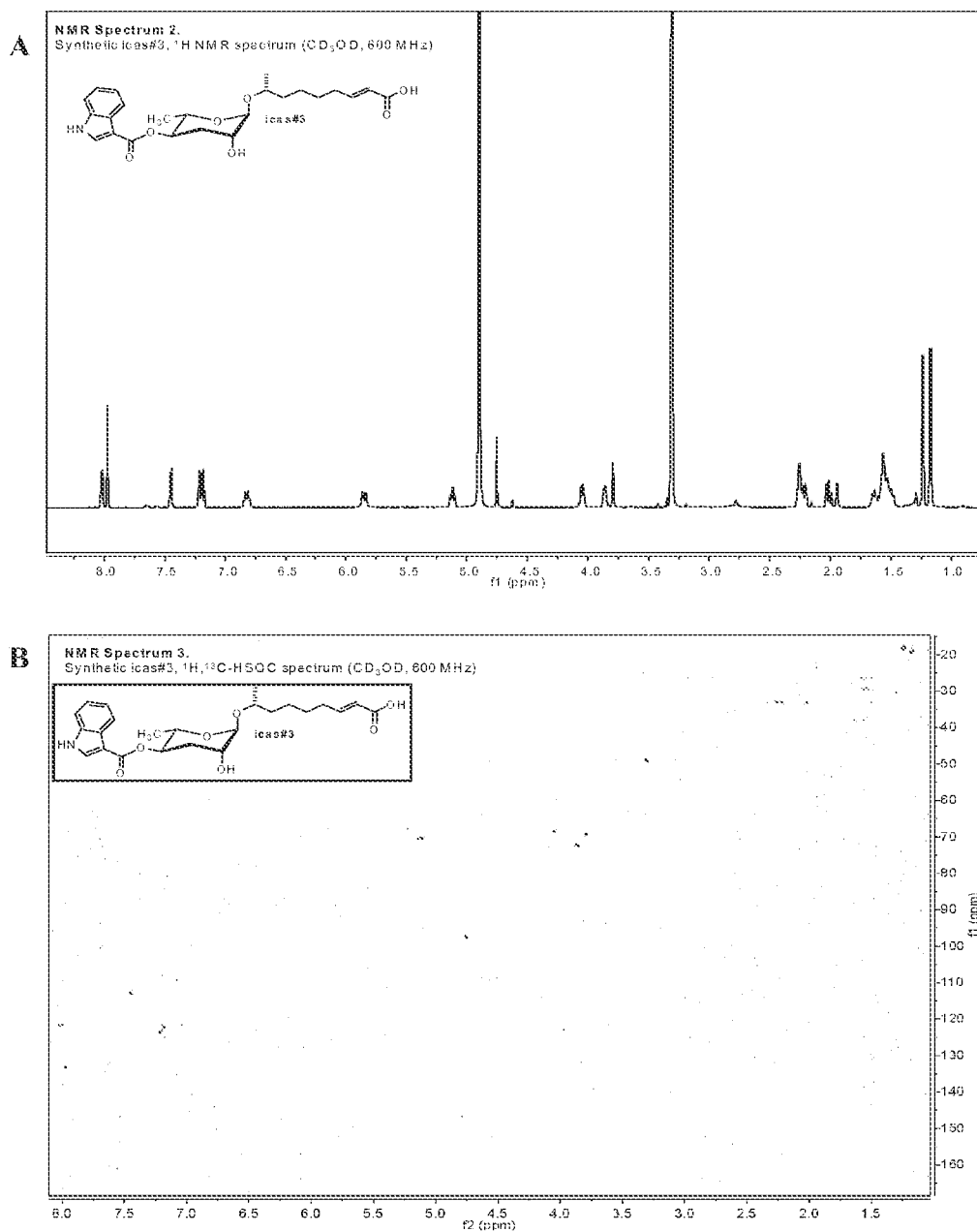
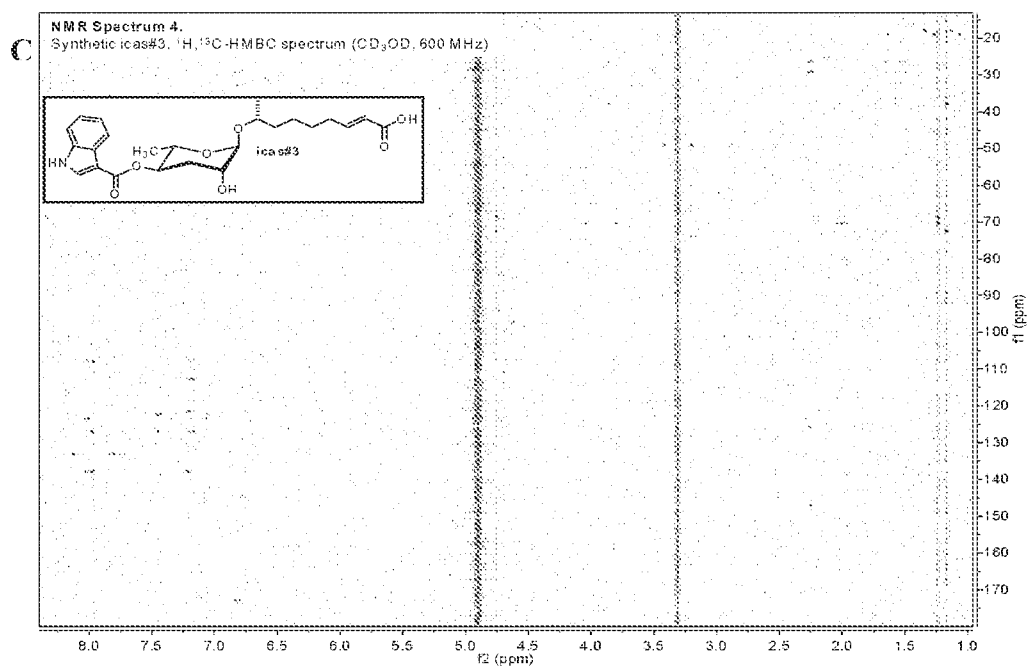


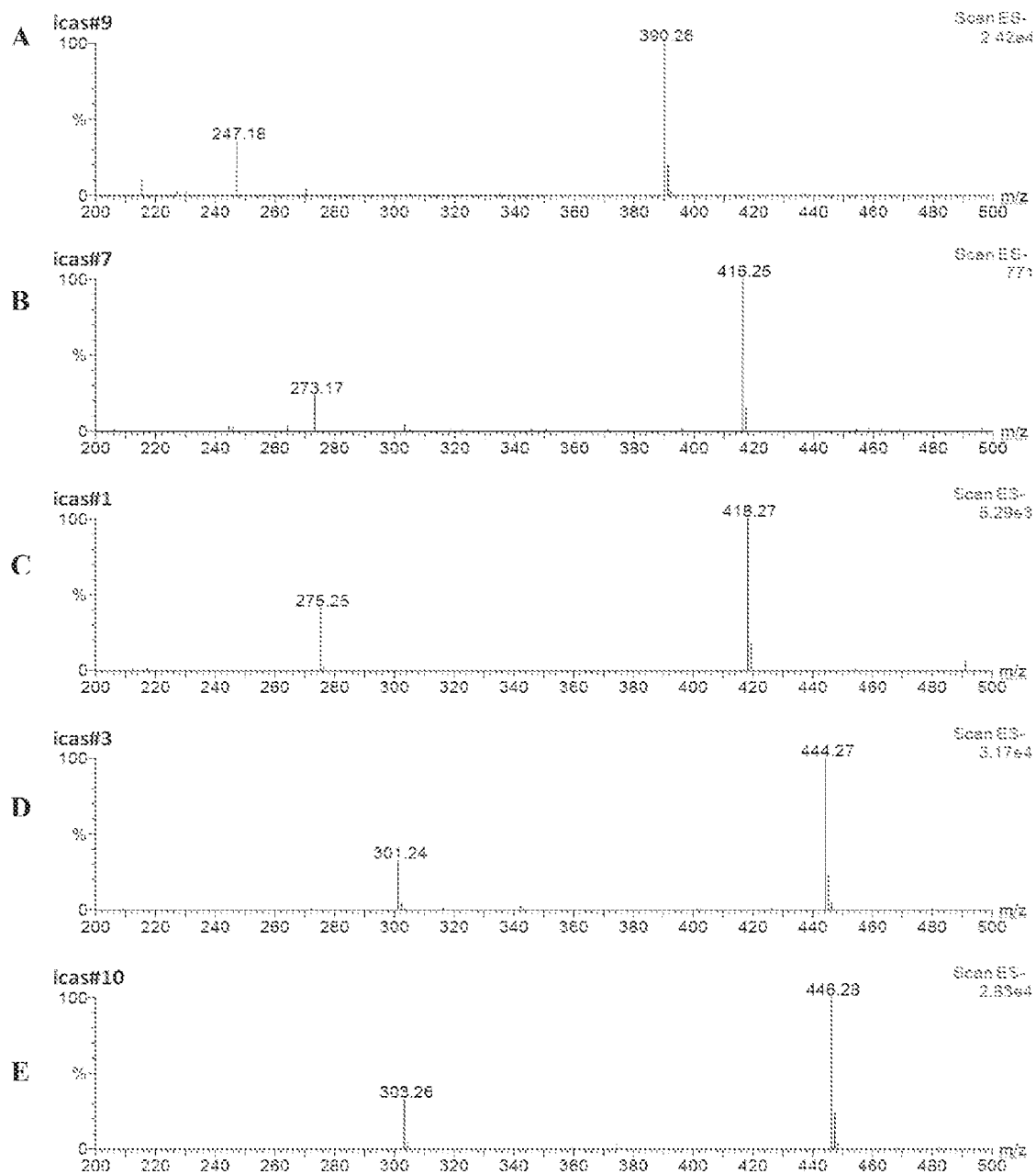
Figure 10



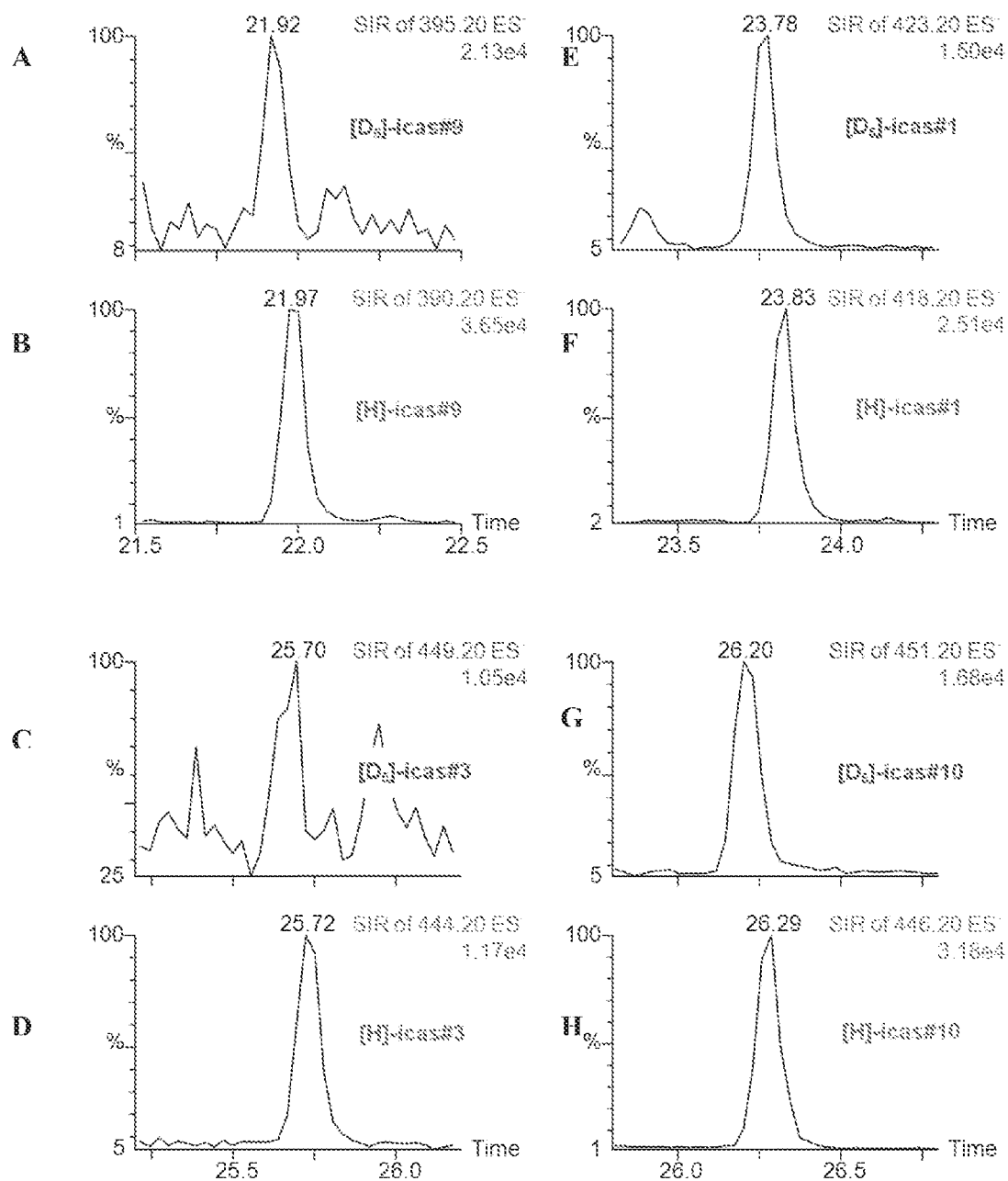
Figures 11A-B



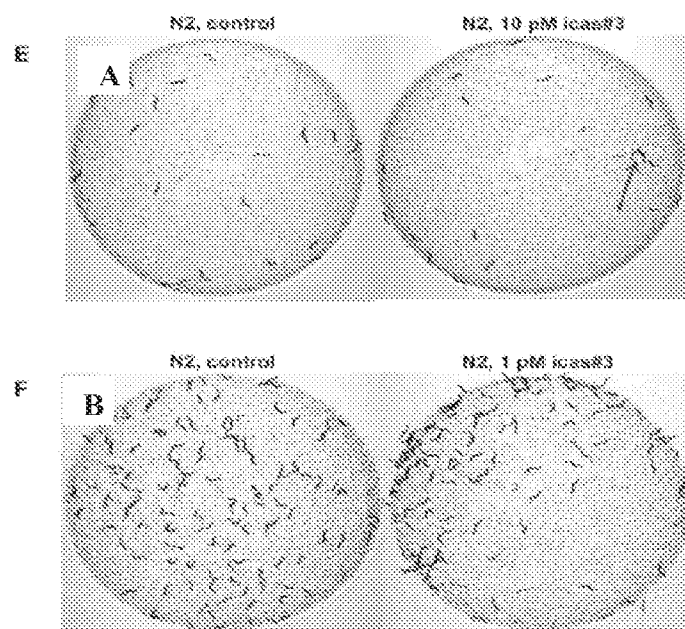
Figures 11-C



Figures 12A-E



Figures 13A-H



Figures 14A–B

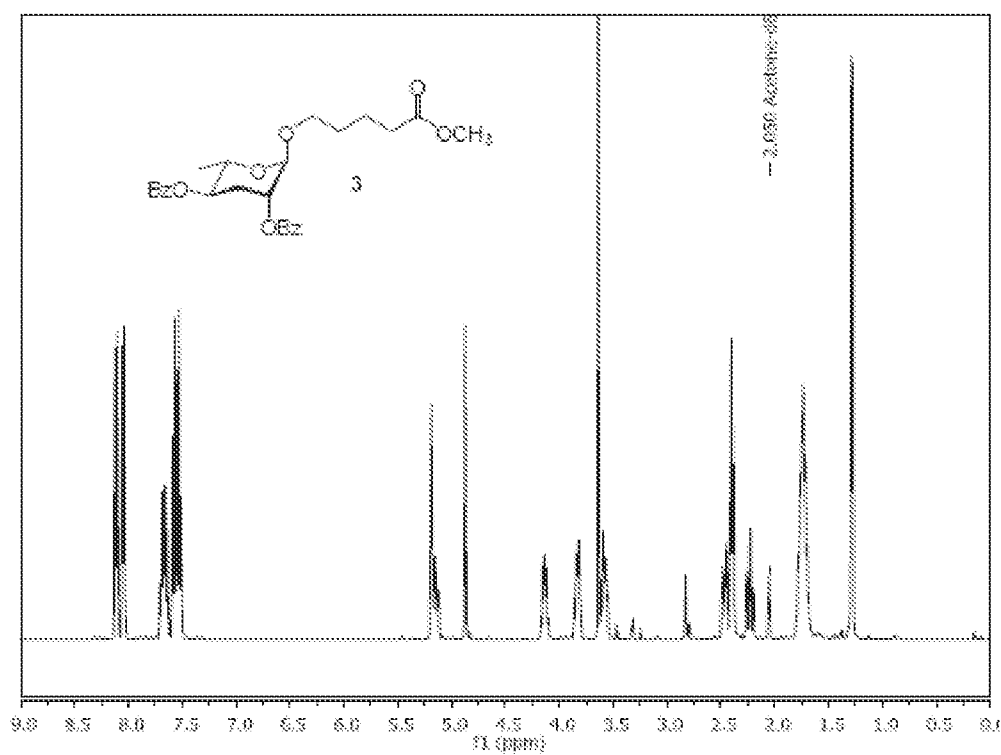


Figure 15A

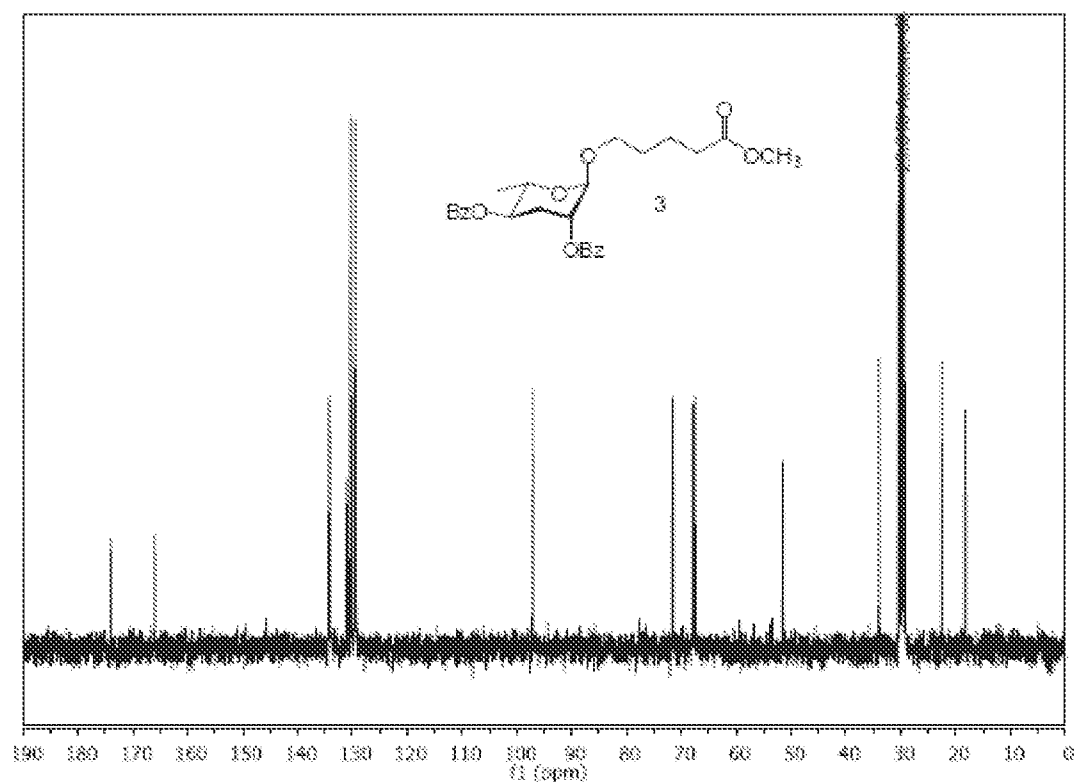


Figure 15B

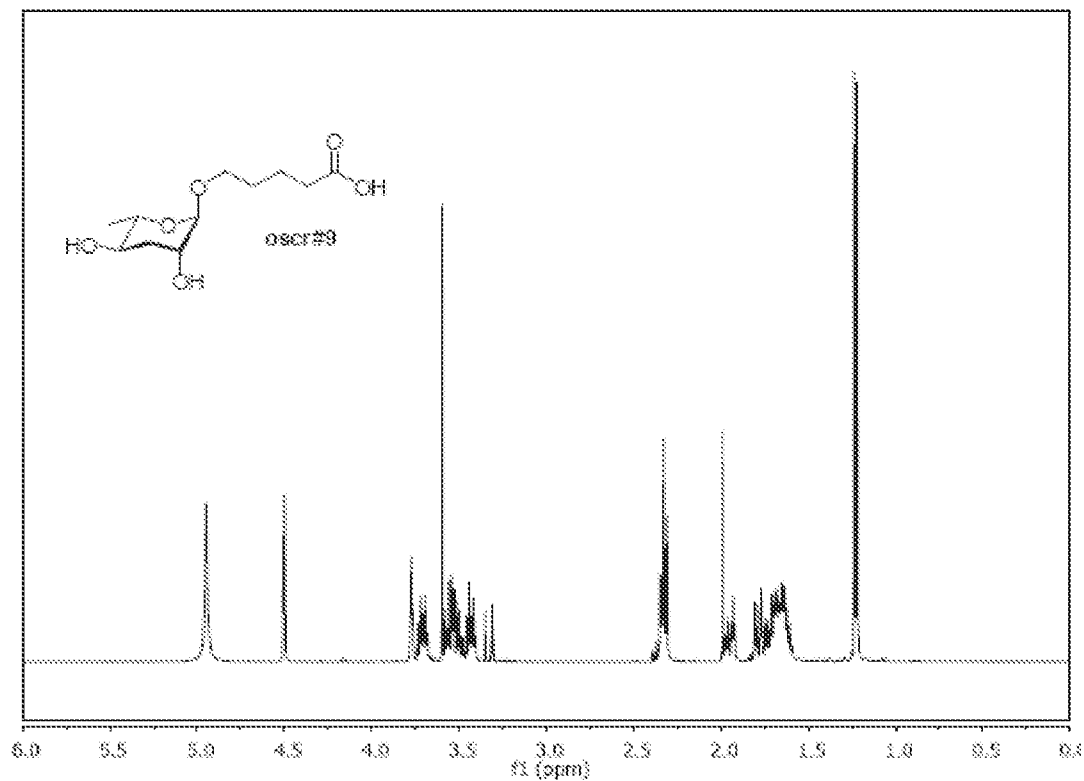


Figure 16A

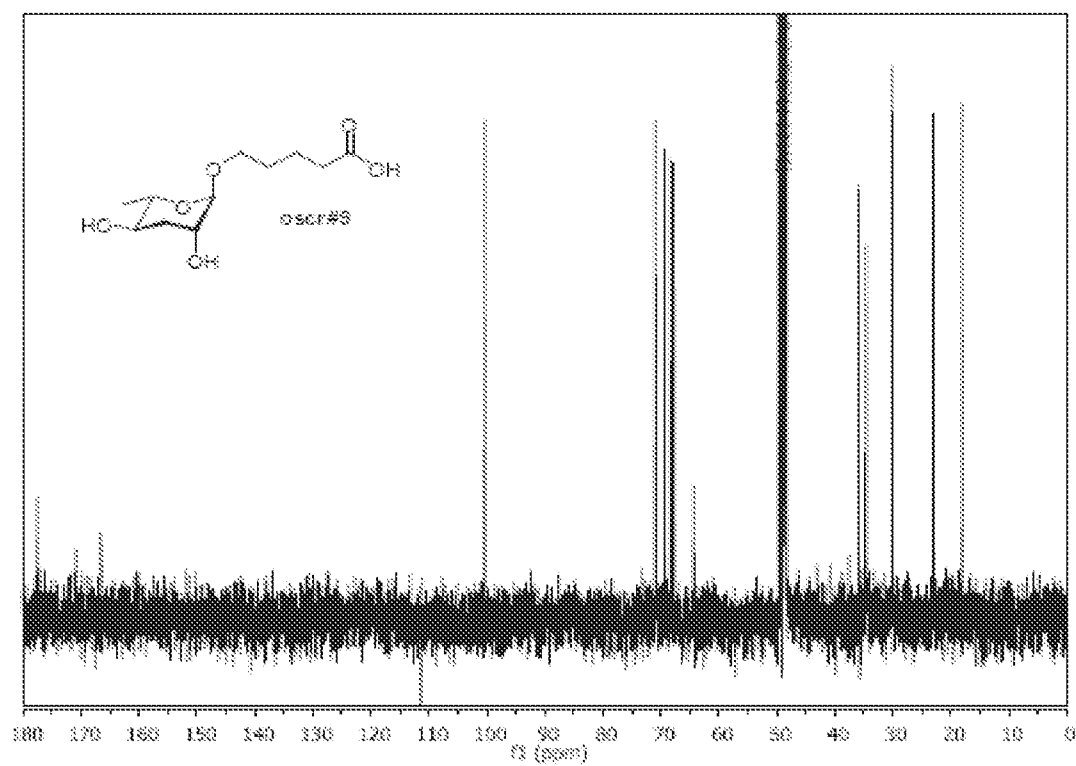


Figure 16B

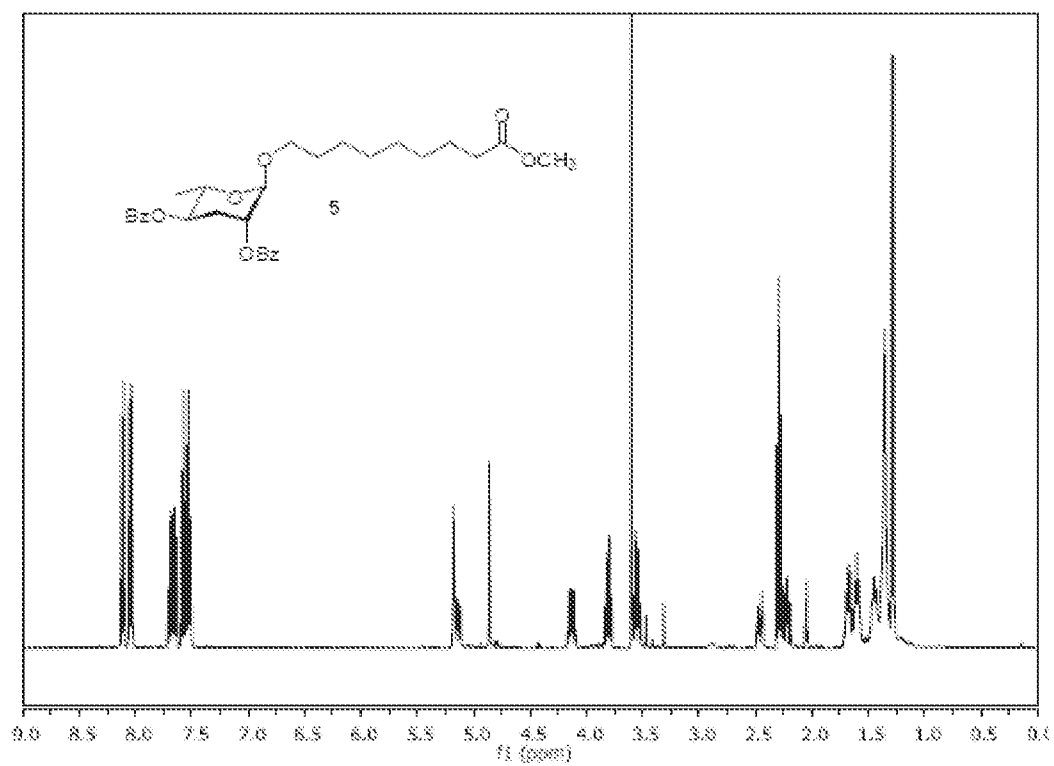


Figure 17A

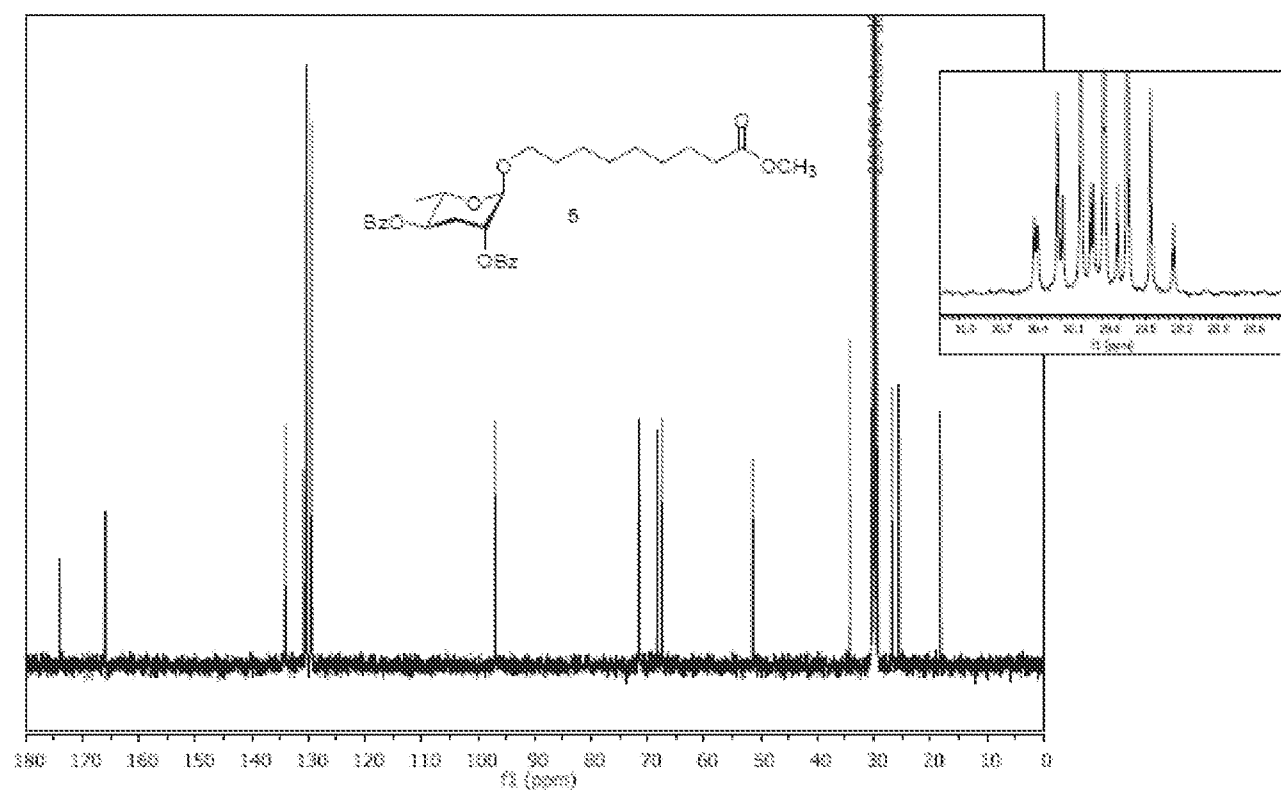


Figure 17B

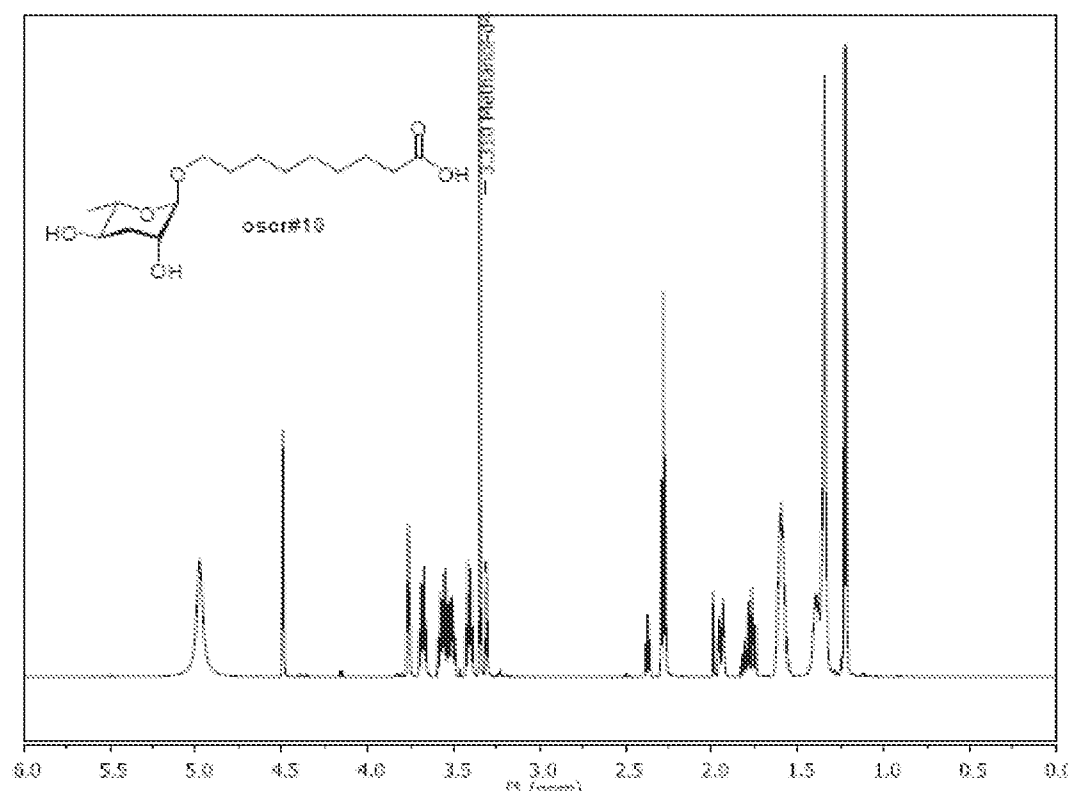


Figure 18A

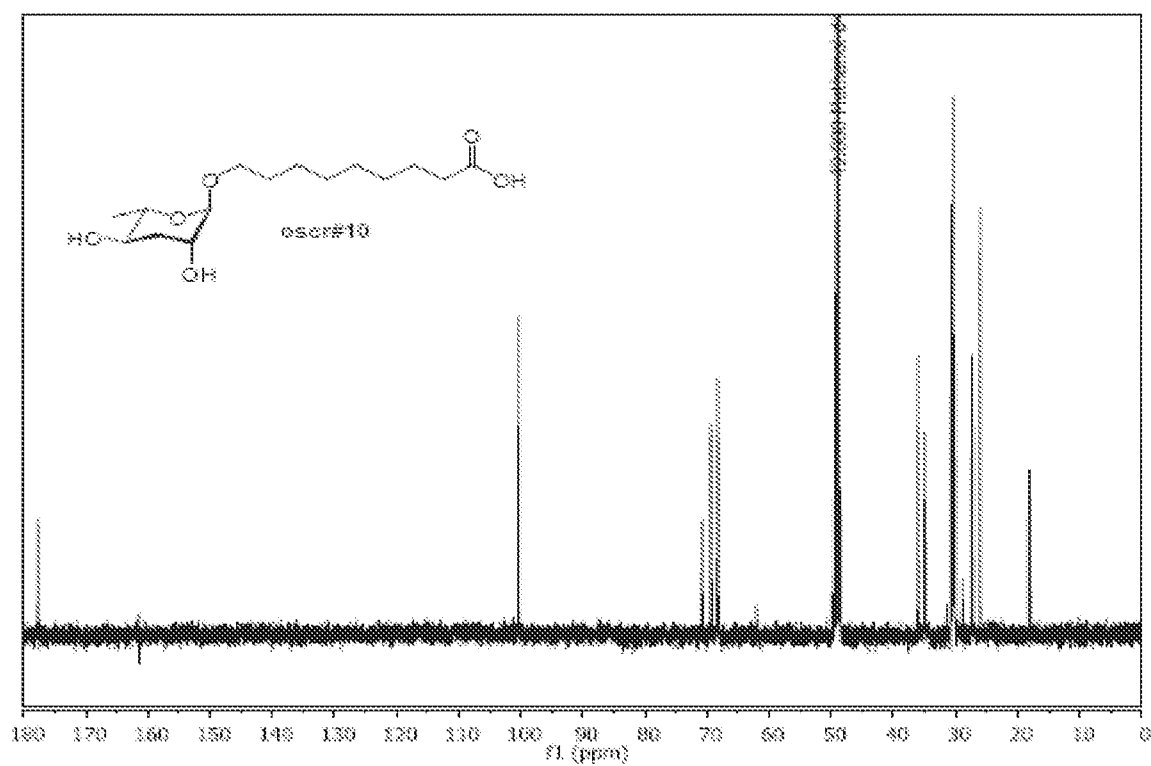


Figure 18B

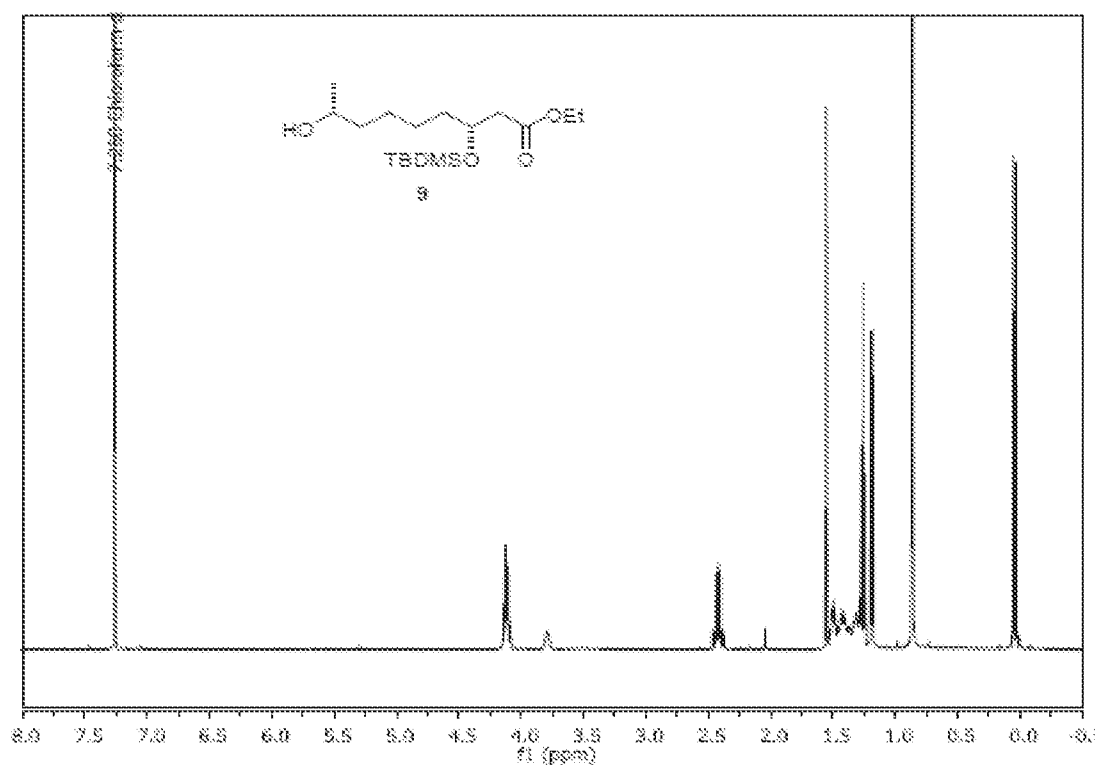


Figure 19

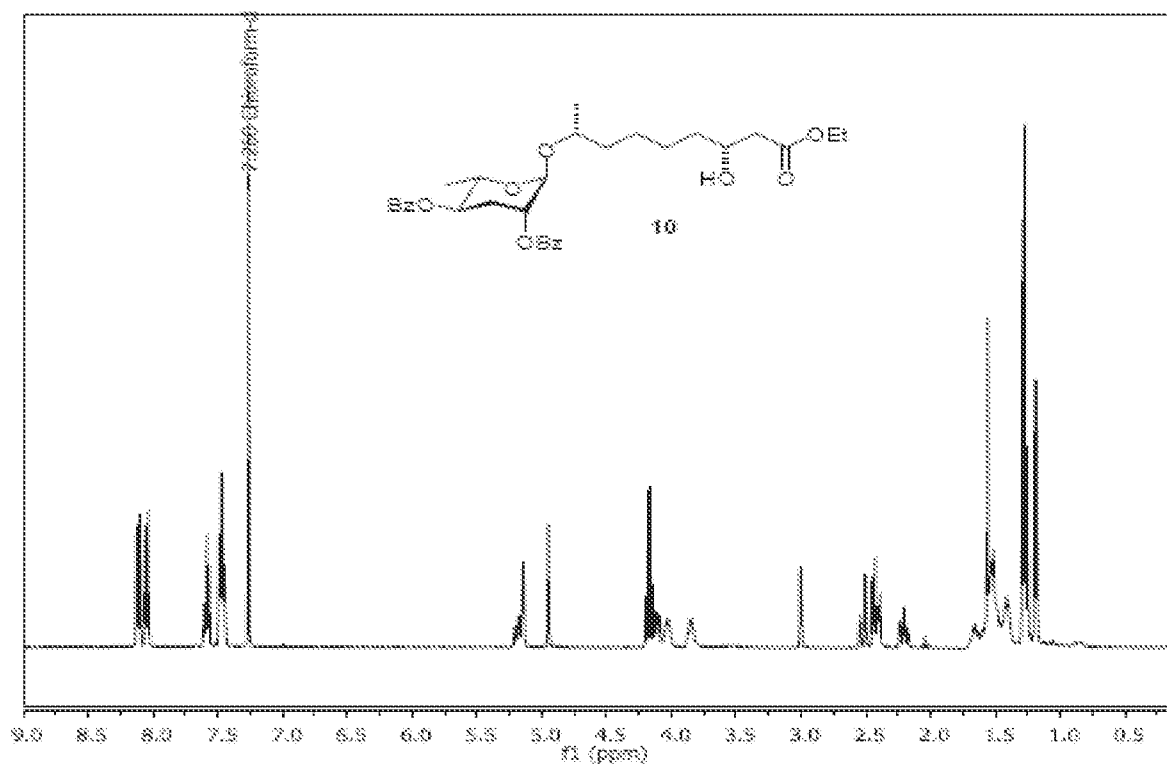


Figure 20

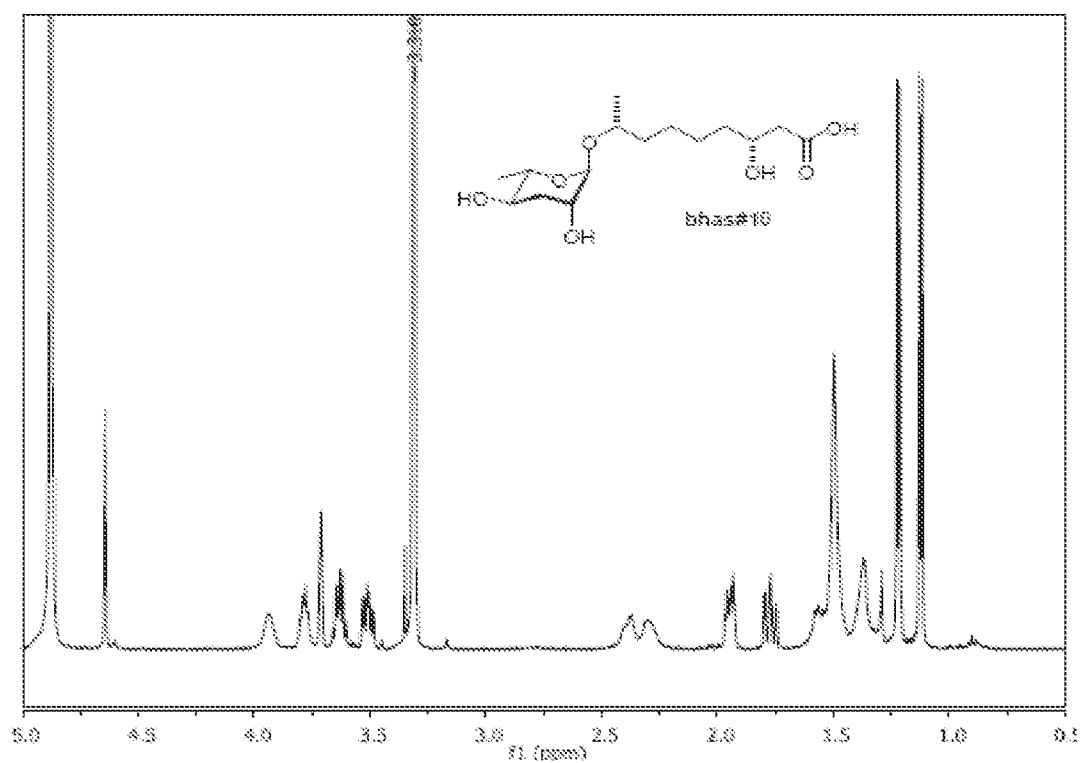


Figure 21A

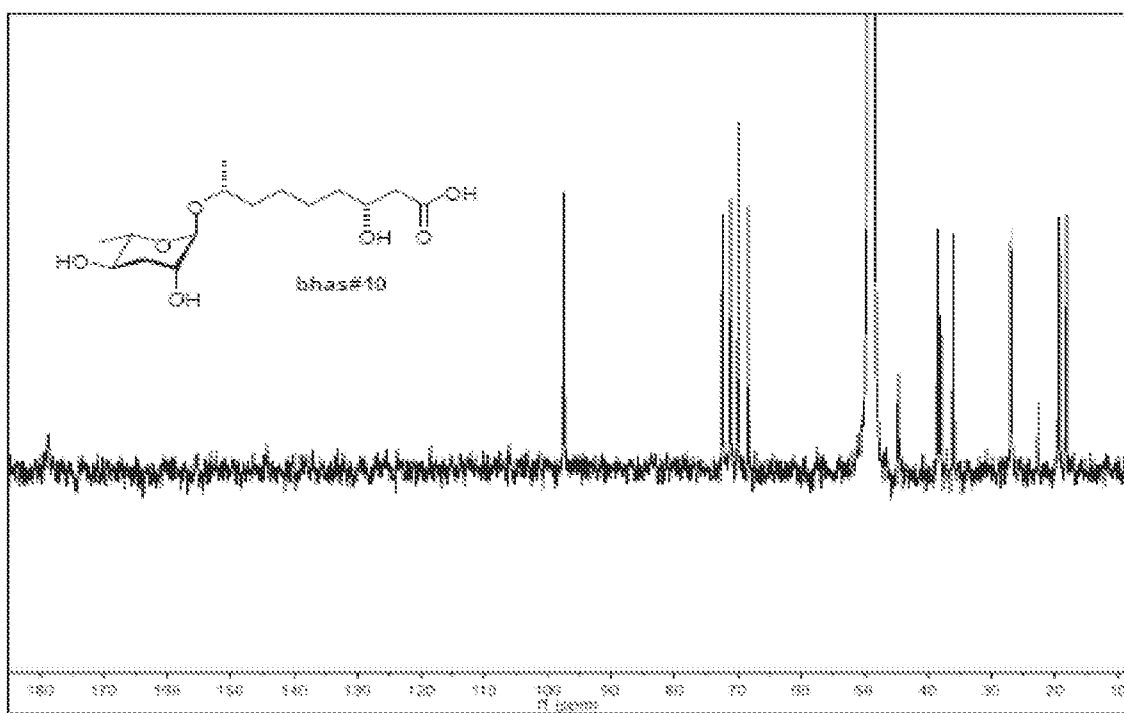


Figure 21B

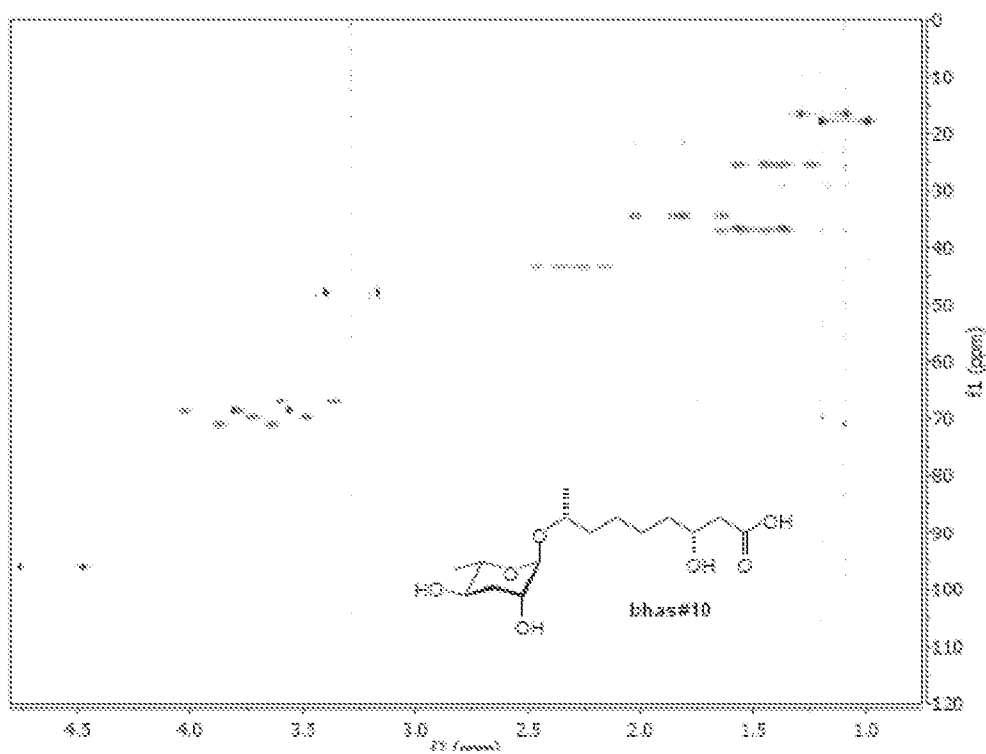


Figure 21C

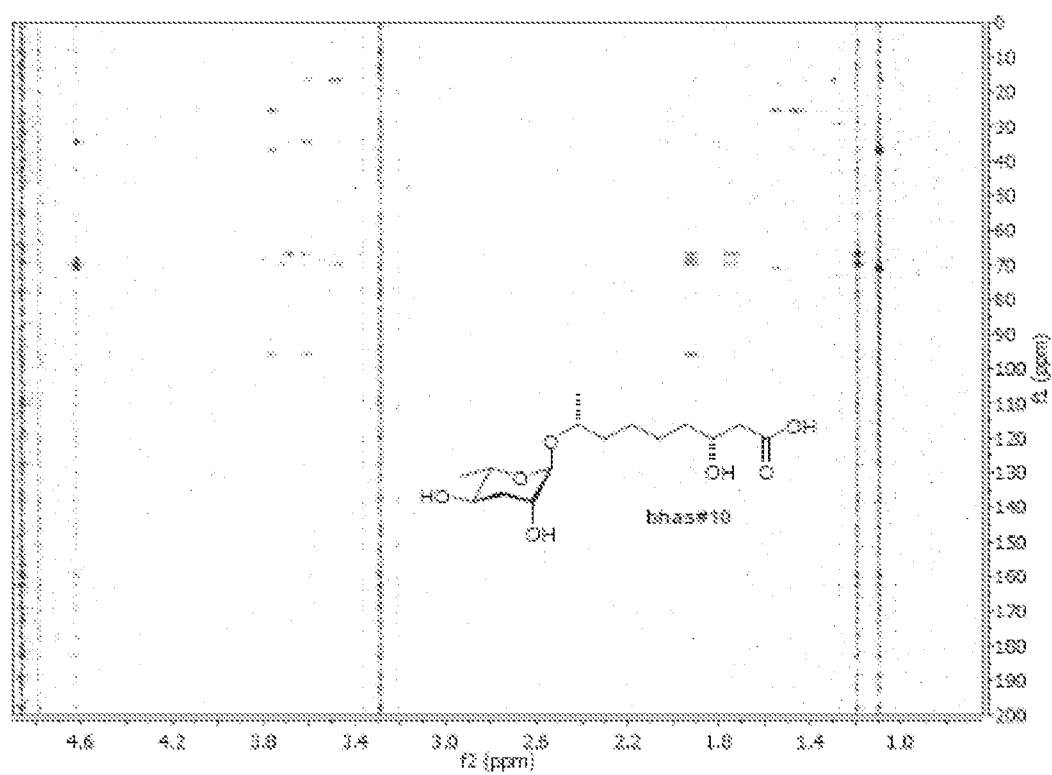


Figure 21D

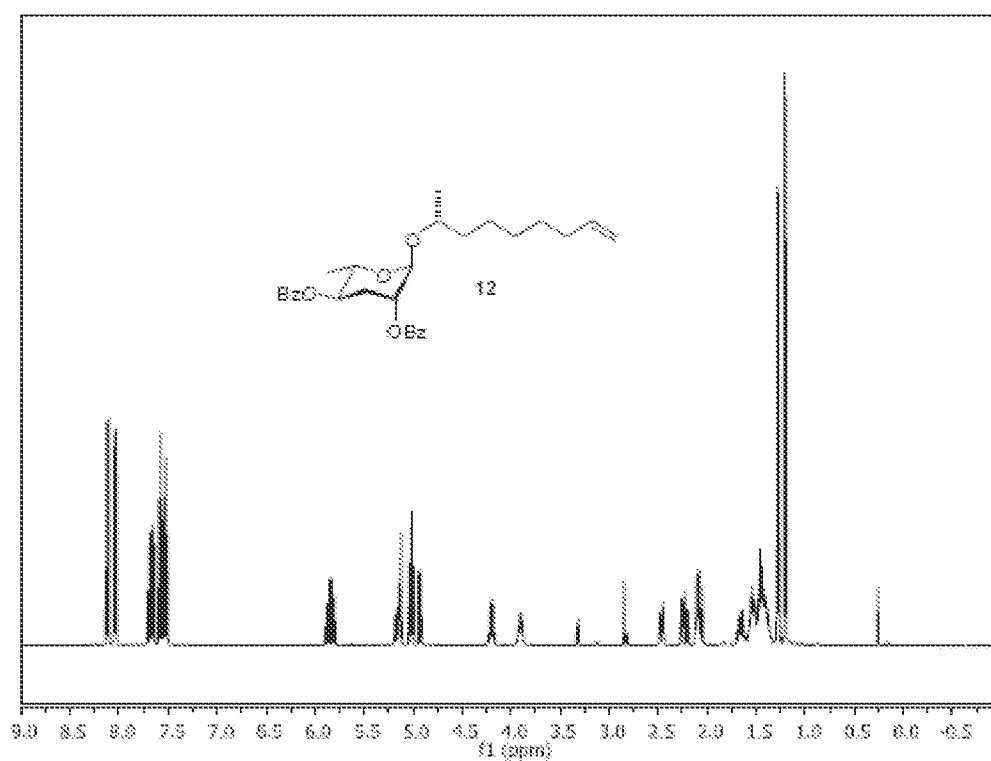


Figure 22A

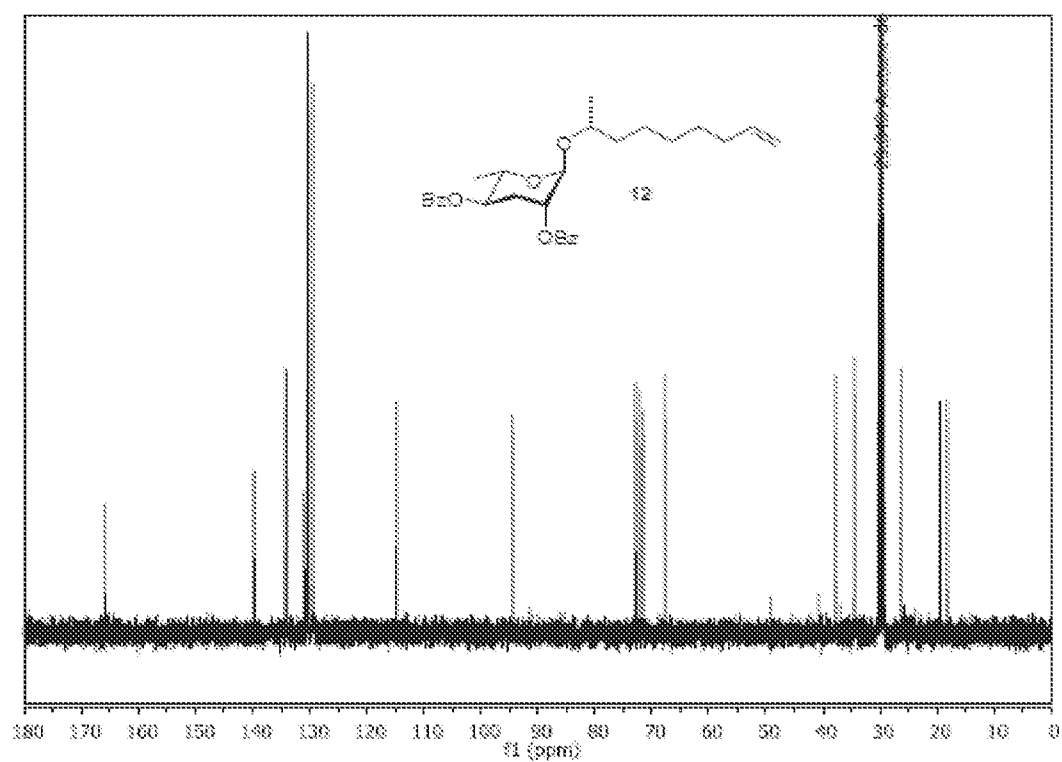


Figure 22B

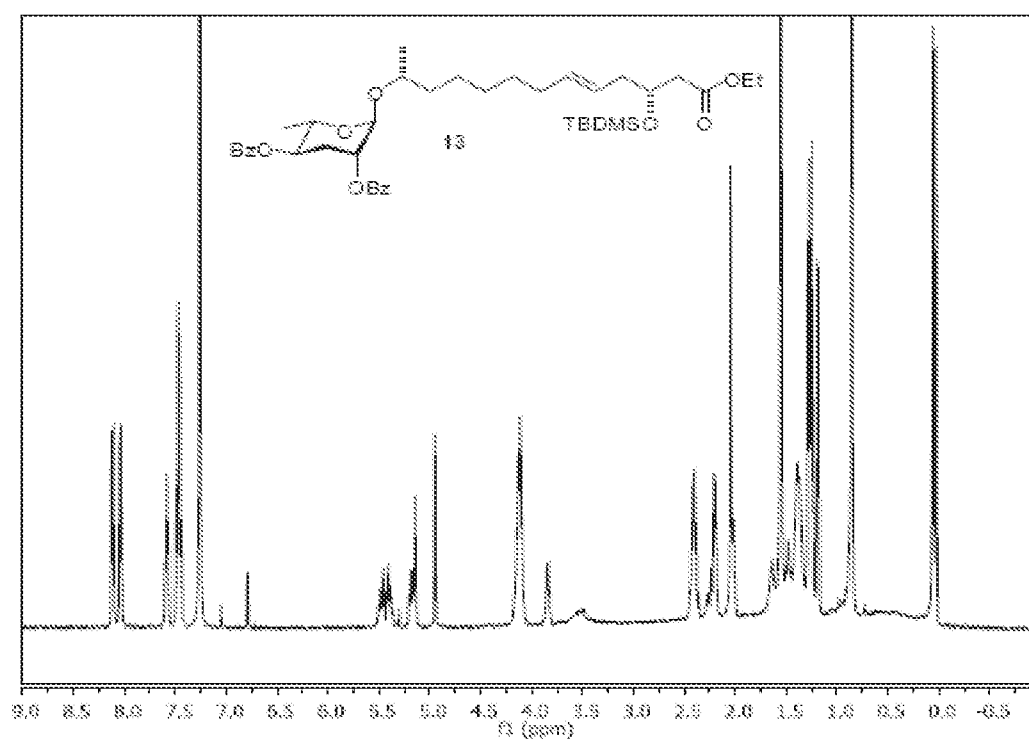


Figure 23

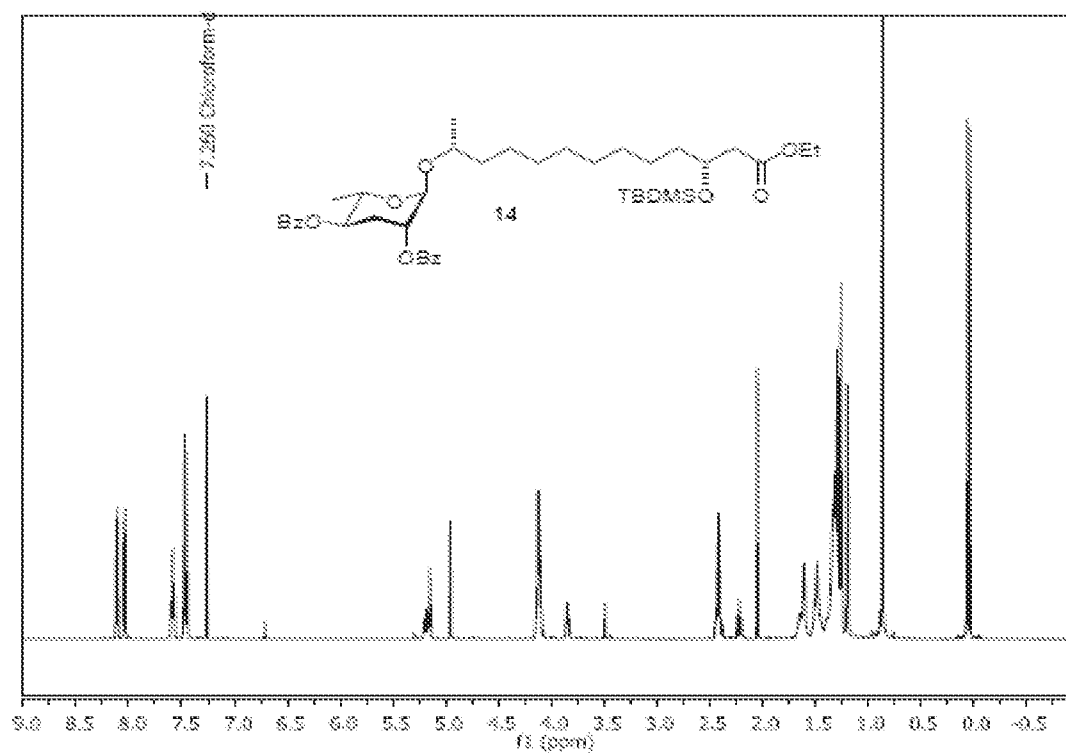


Figure 24

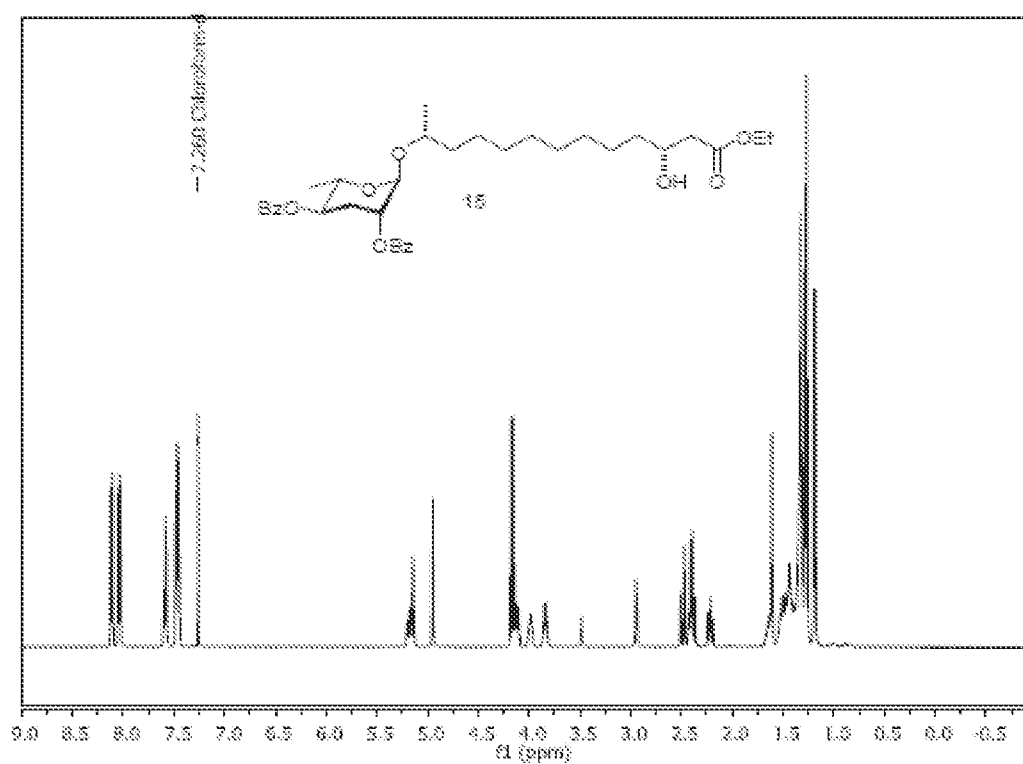


Figure 25

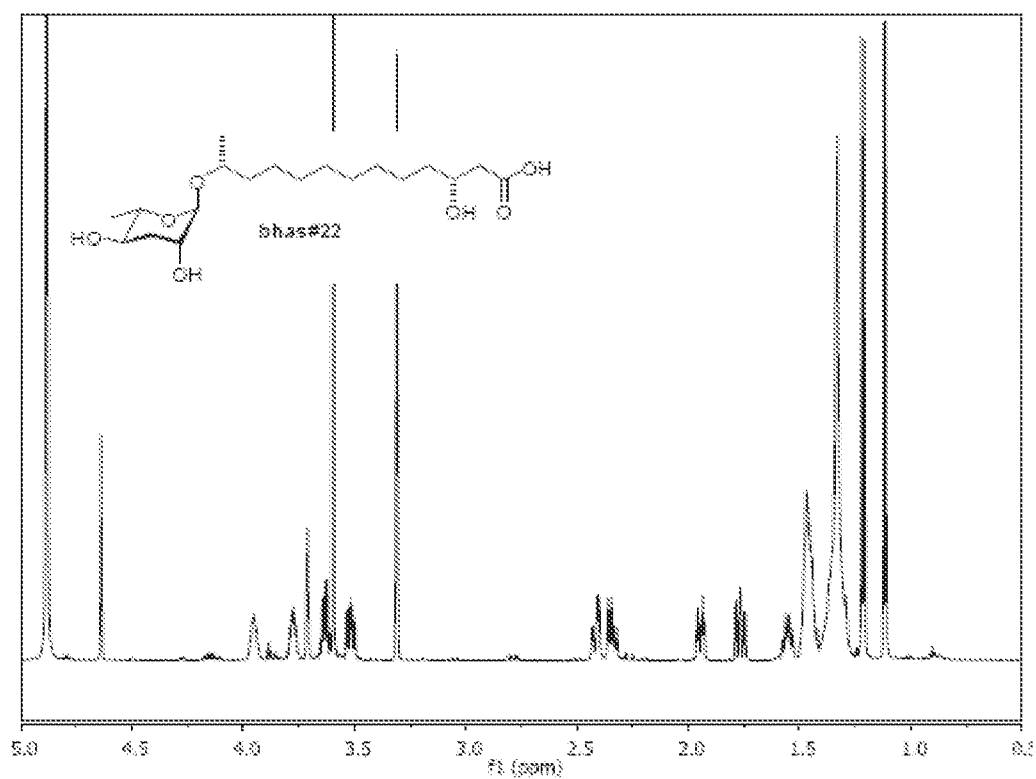


Figure 26A

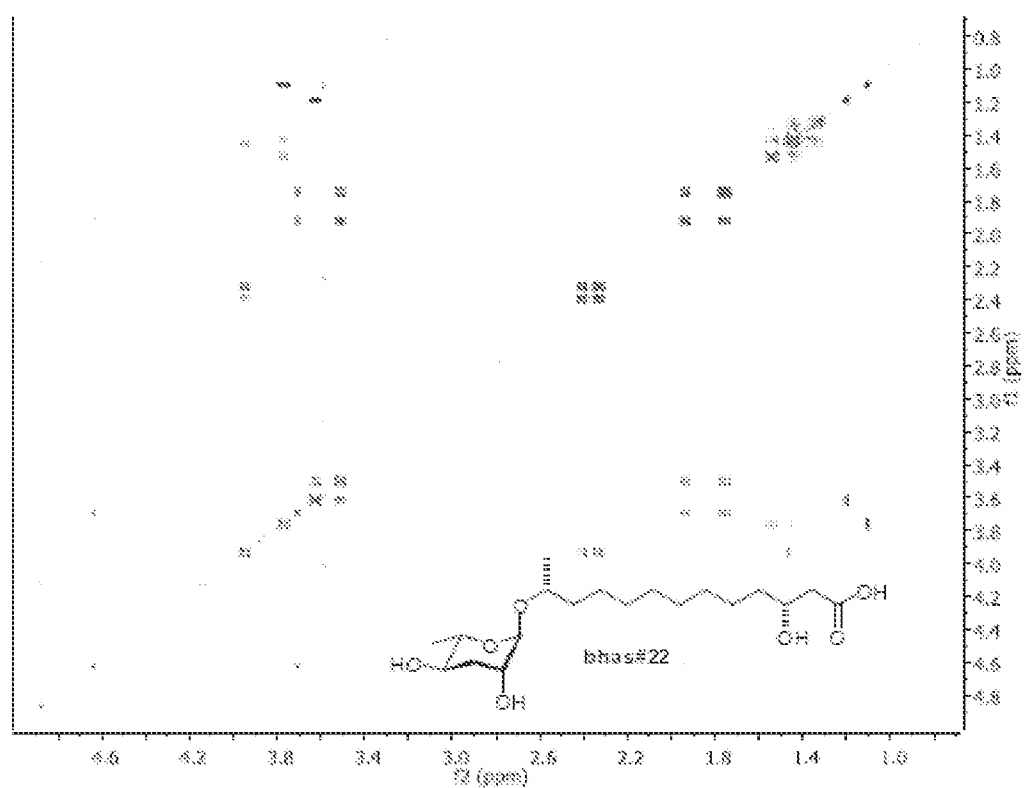


Figure 26B

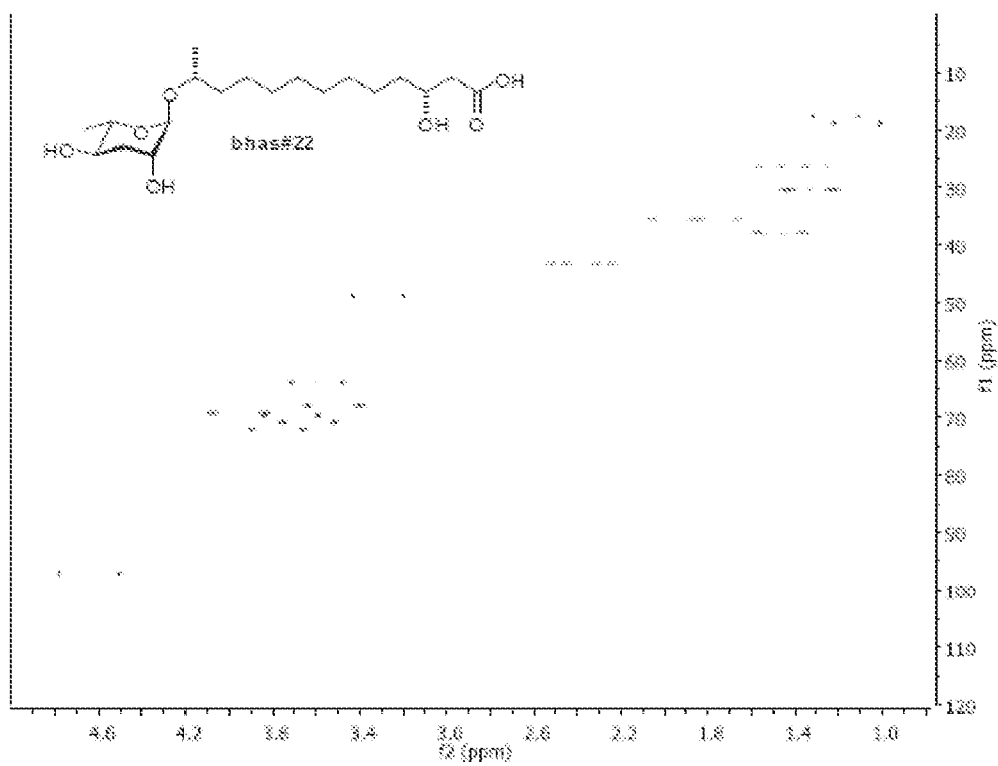


Figure 26C

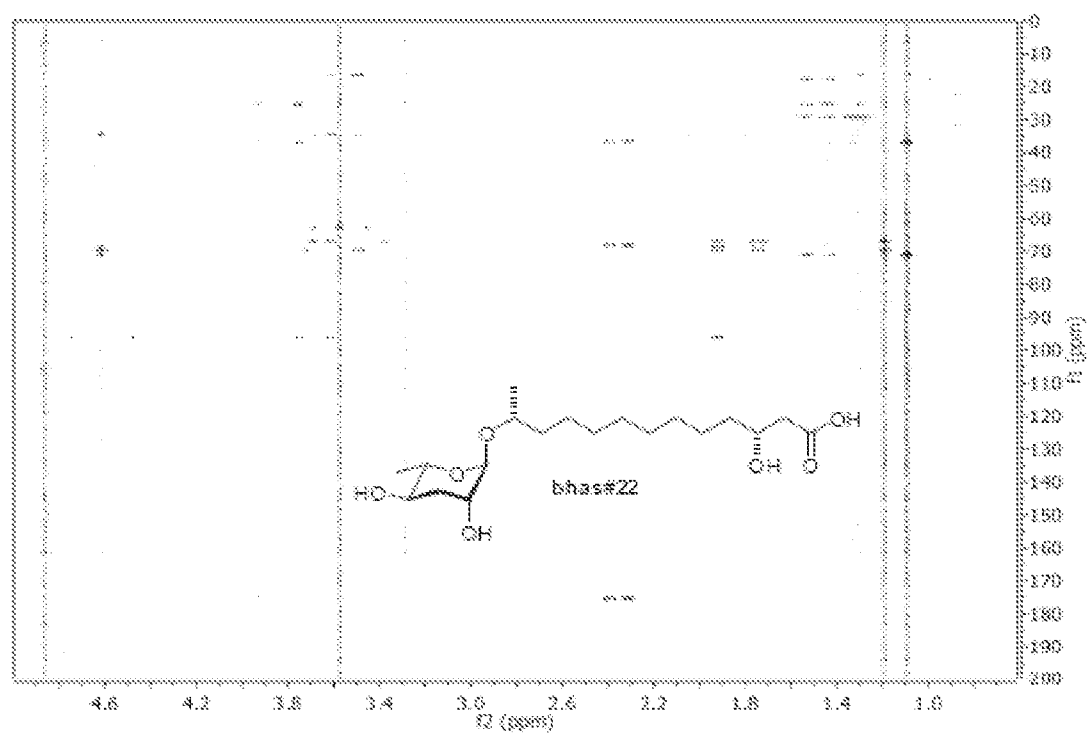


Figure 26D

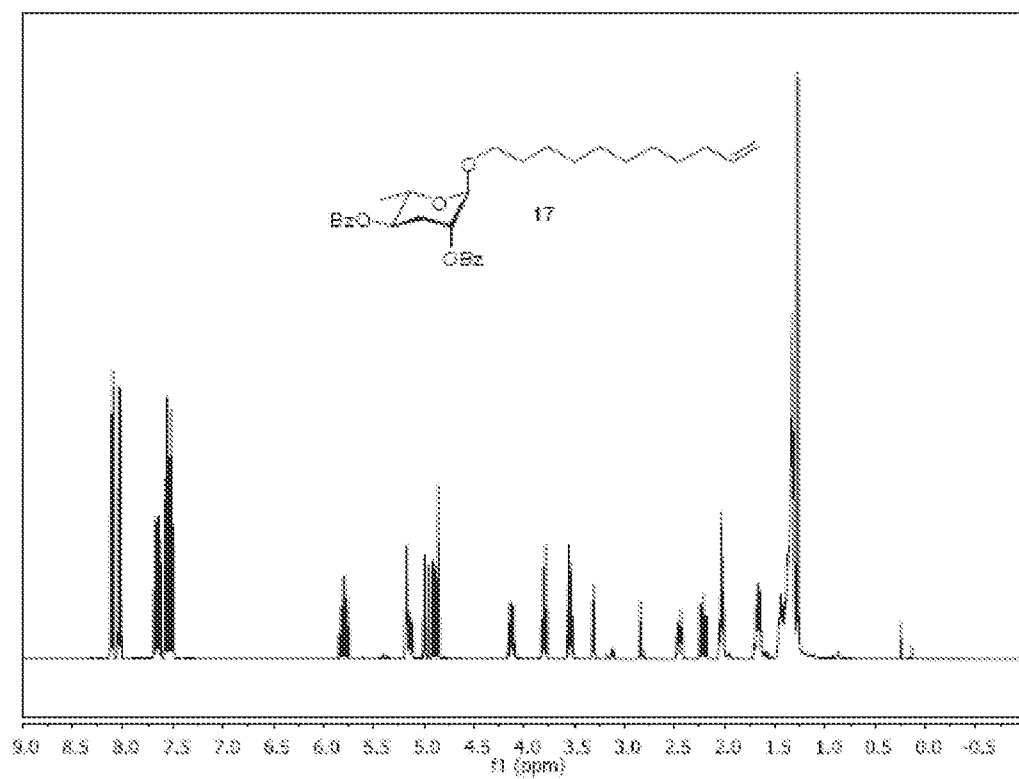


Figure 27A

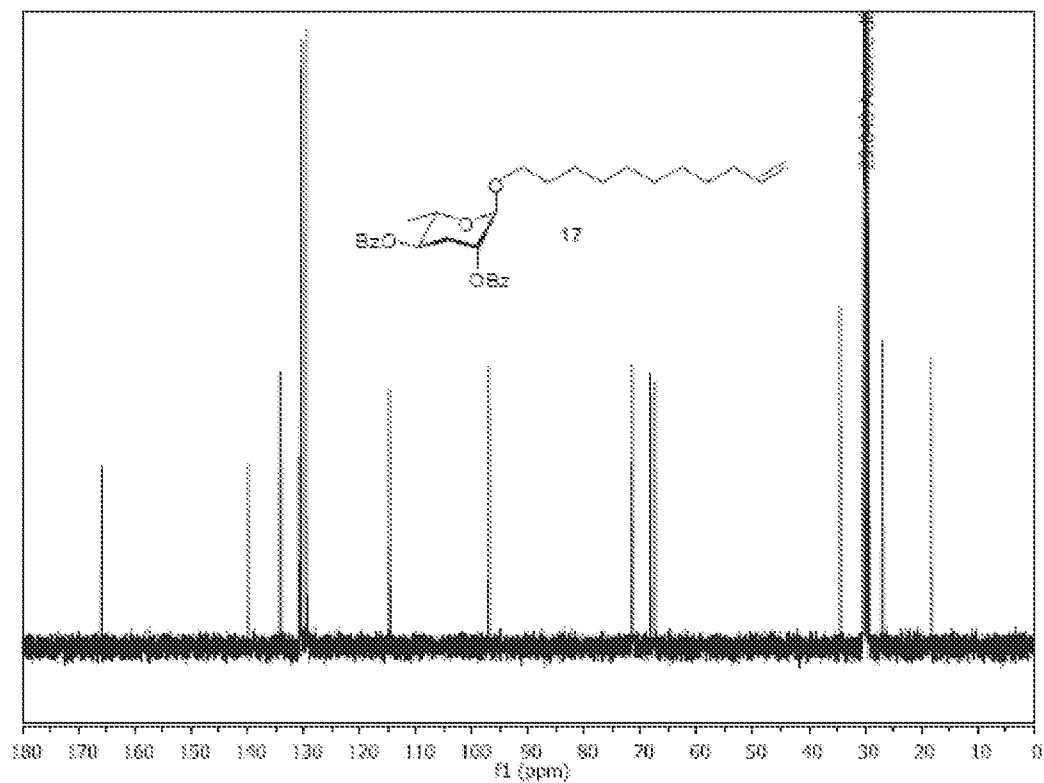


Figure 27B

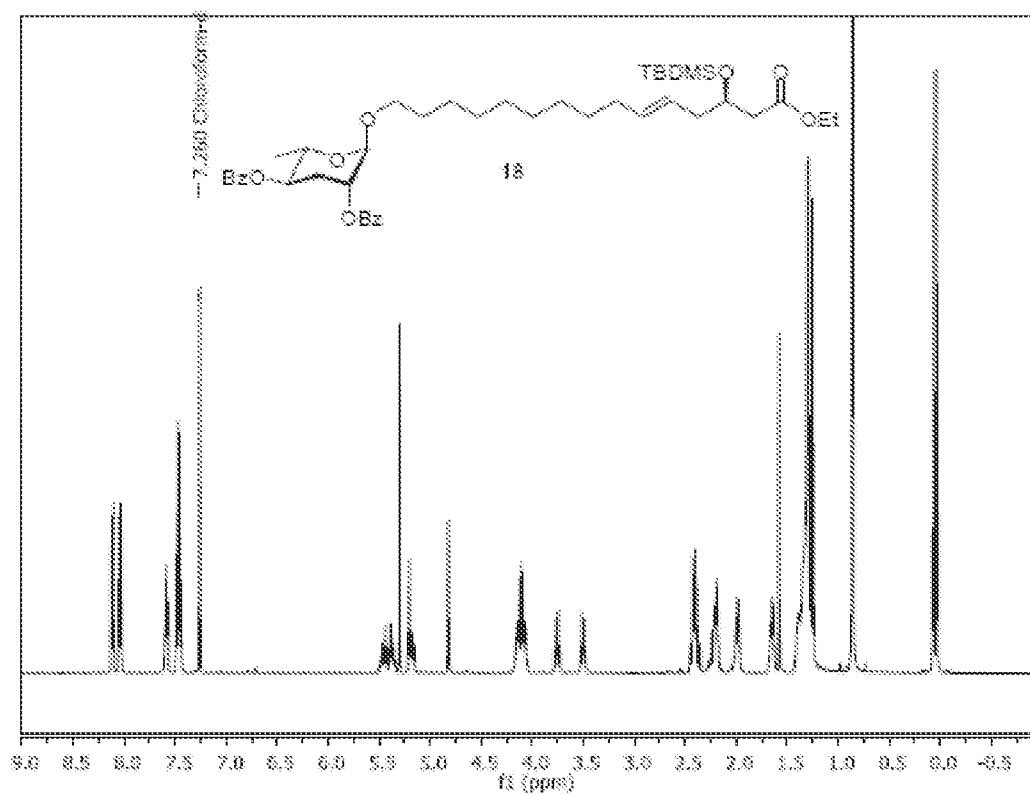


Figure 28

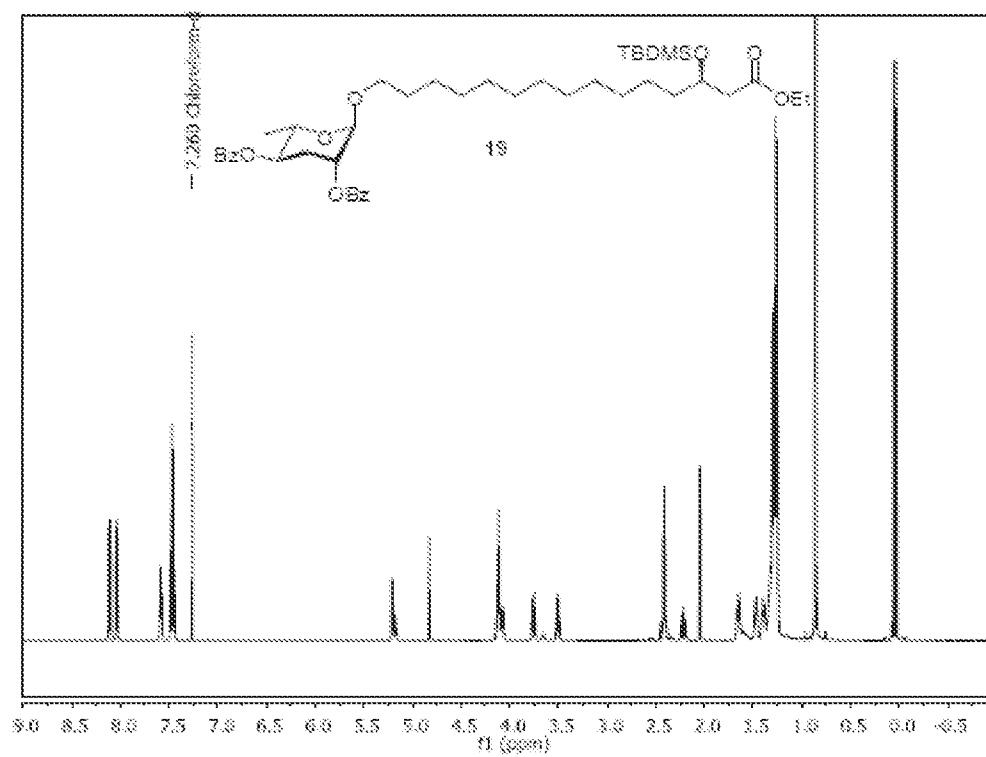


Figure 29

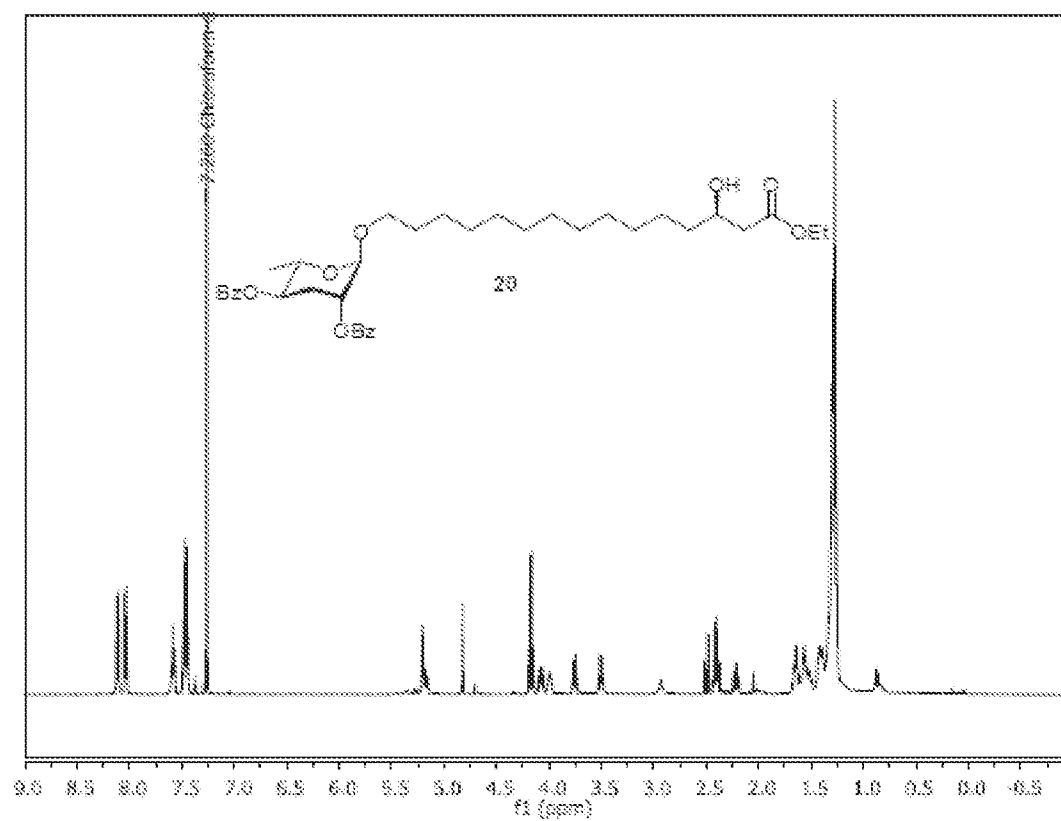


Figure 30

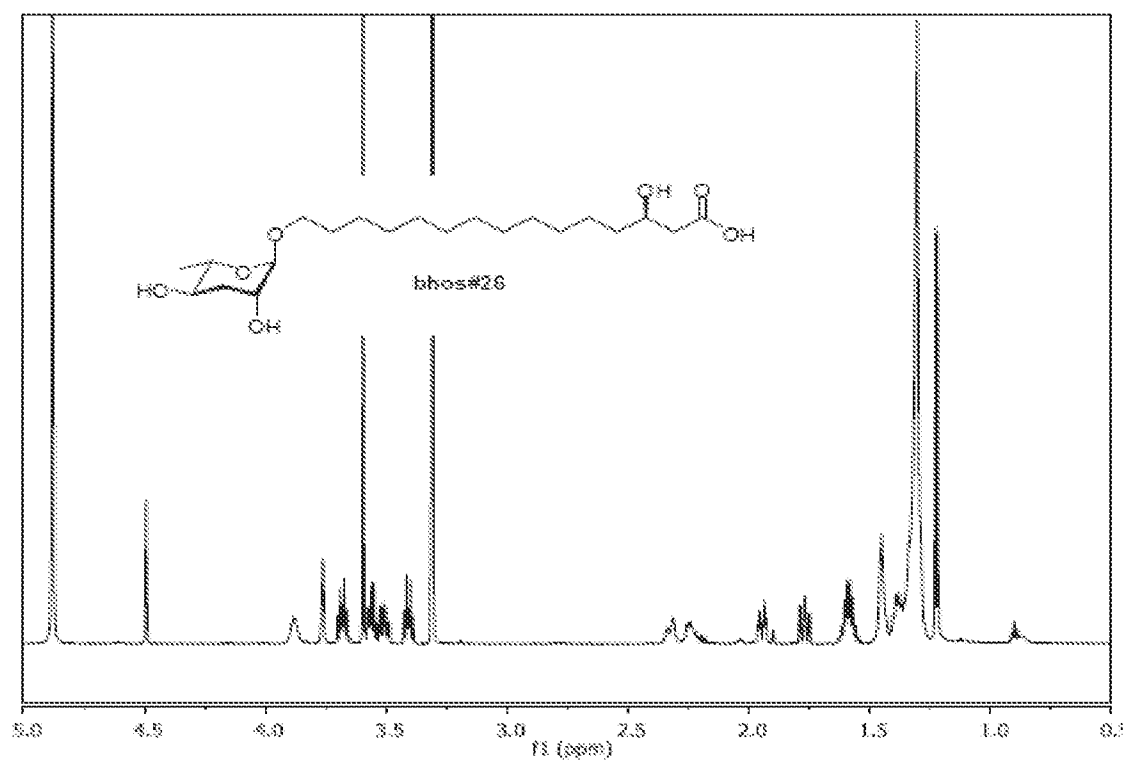


Figure 31A

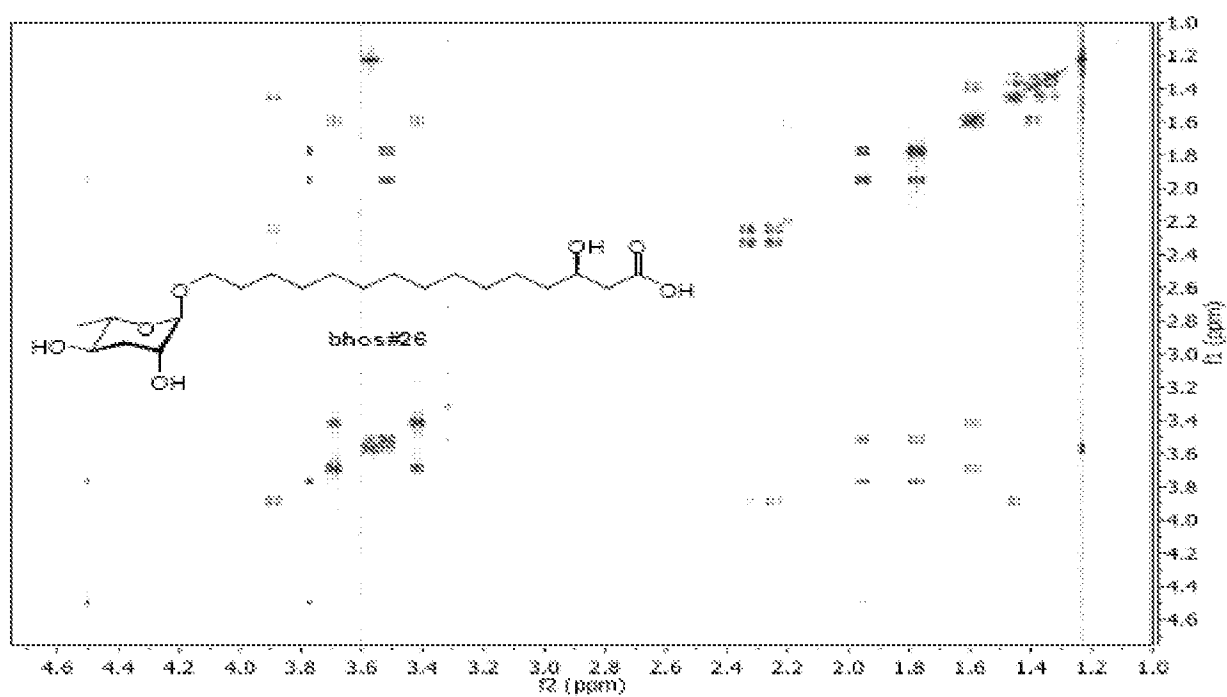


Figure 31B

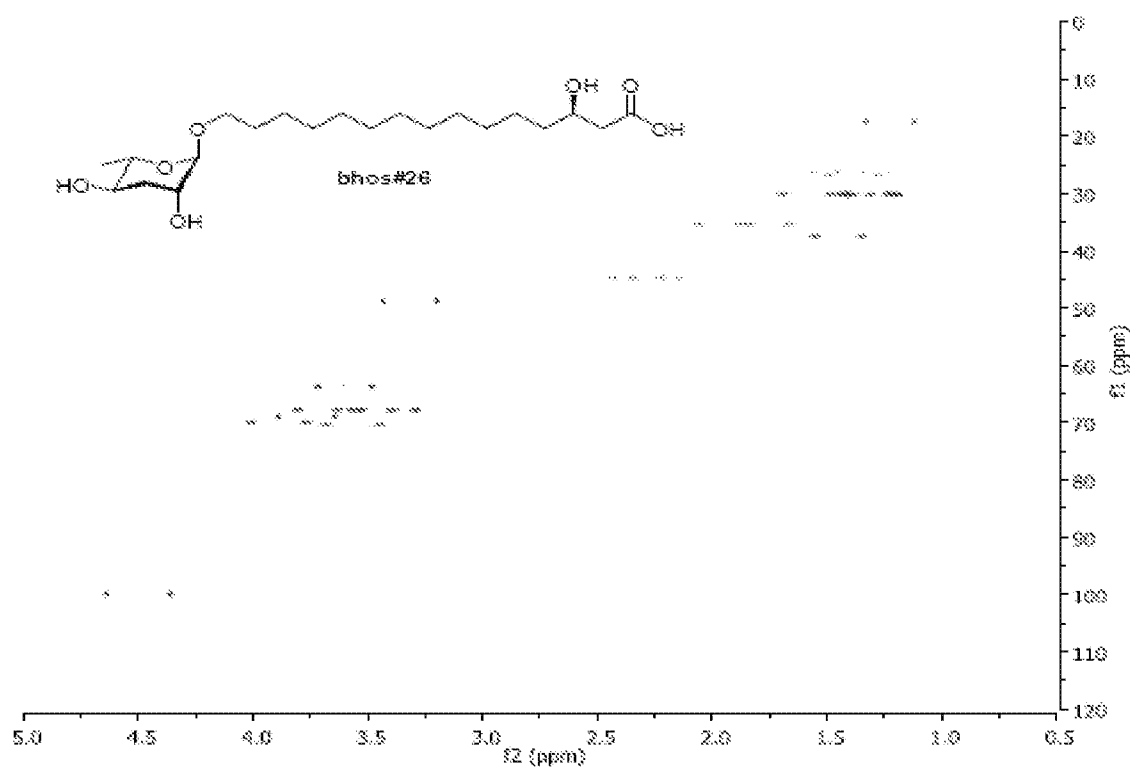


Figure 31C

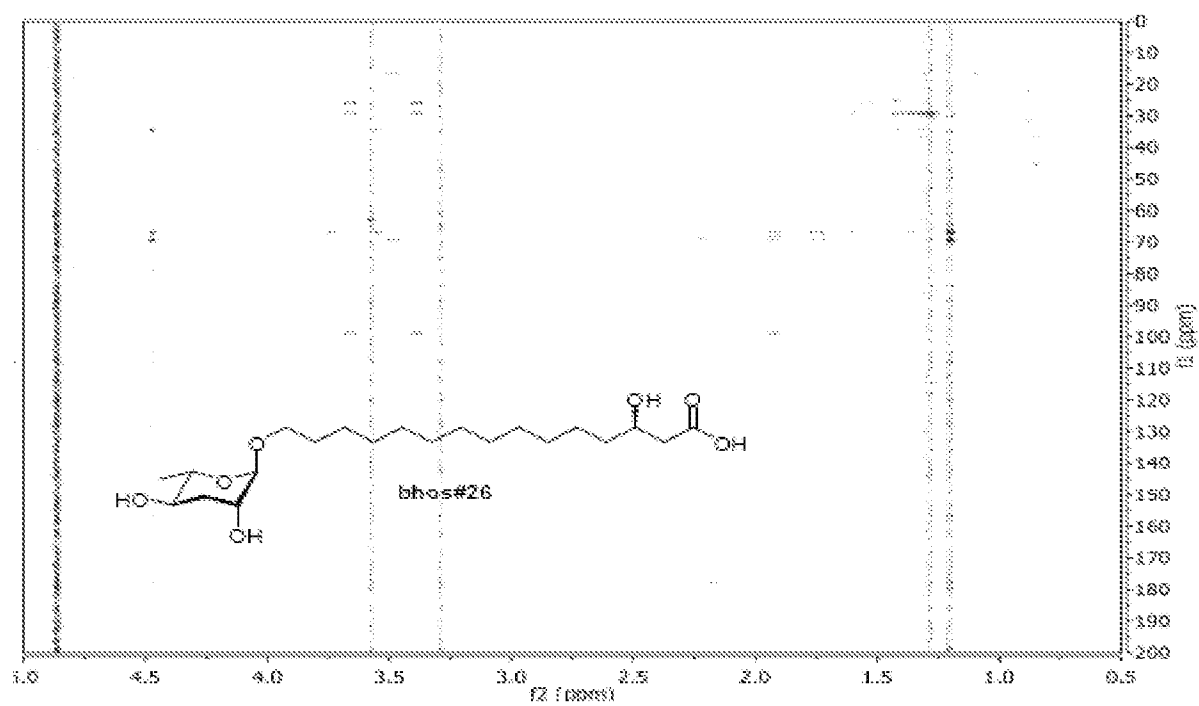


Figure 31D

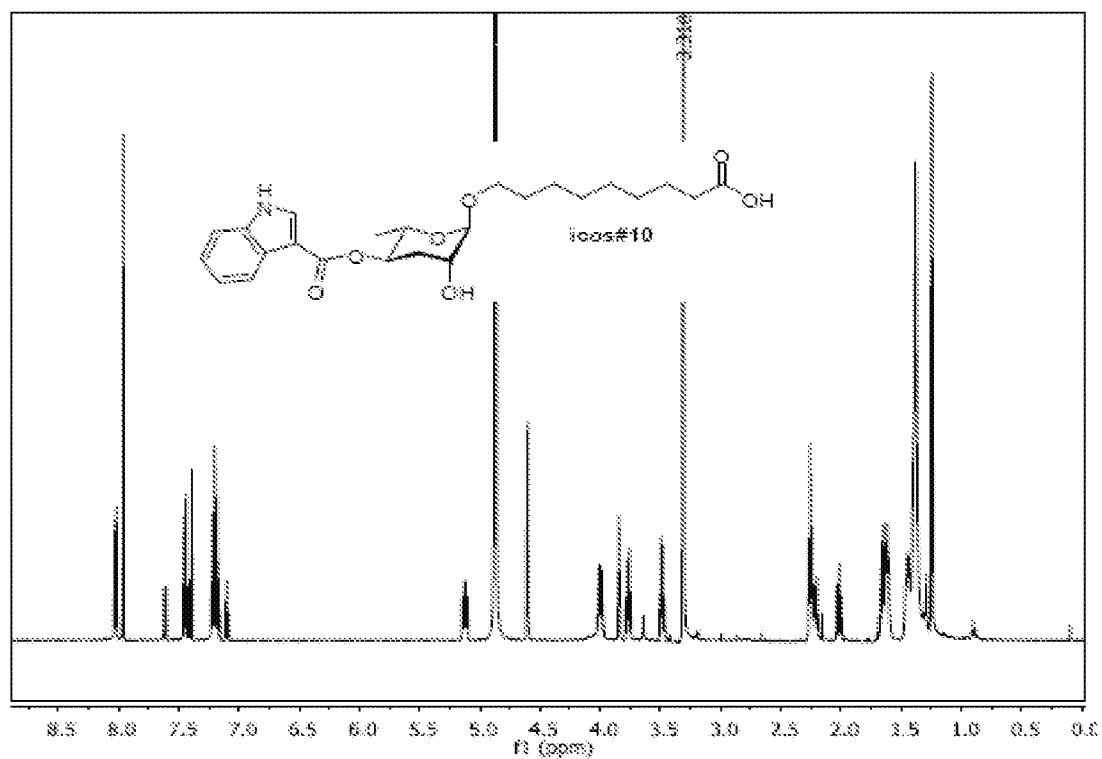


Figure 32A

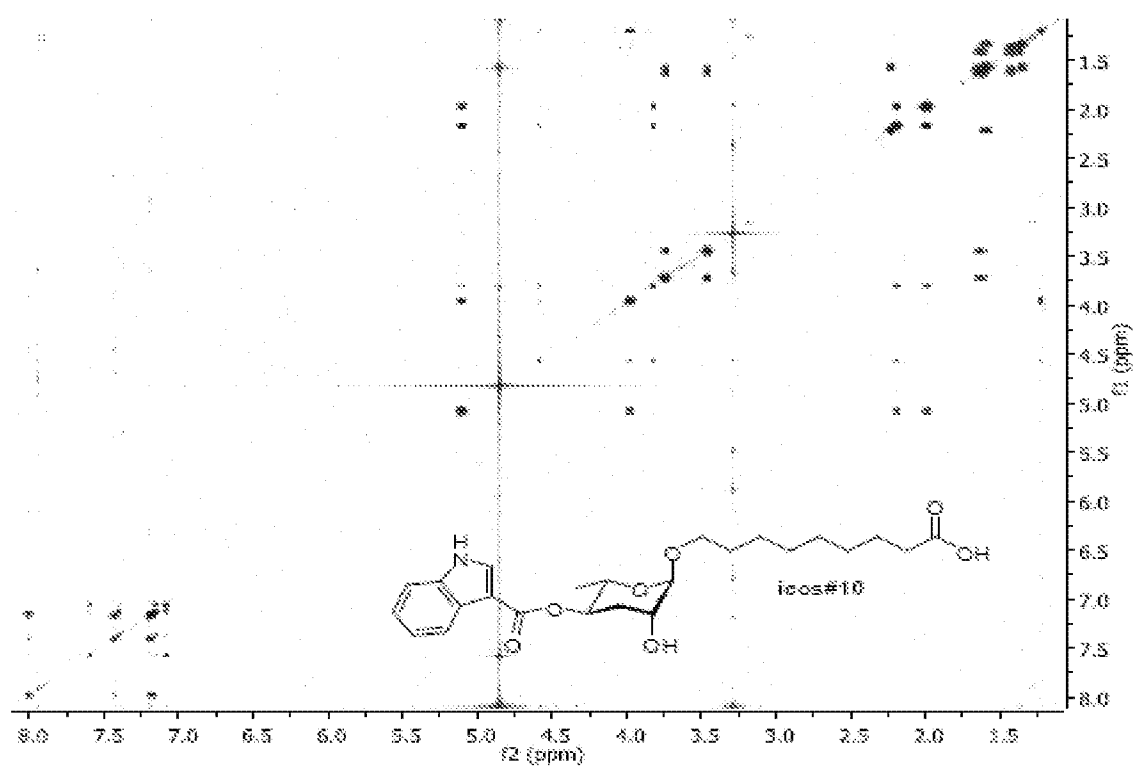


Figure 32B

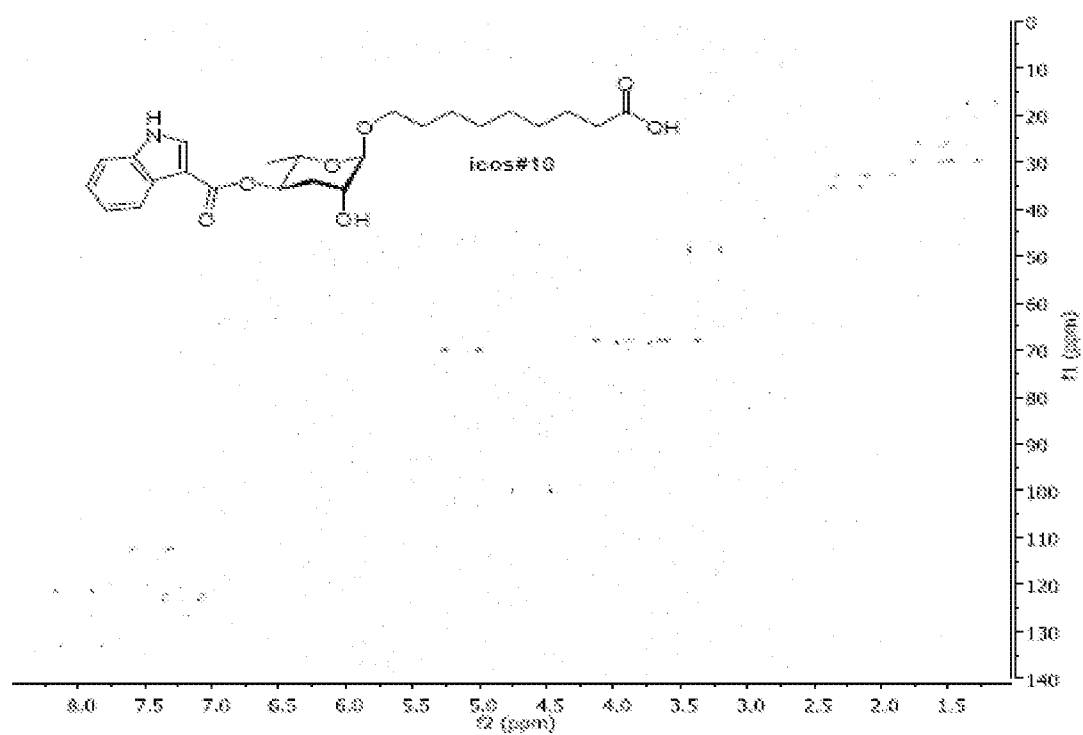


Figure 32C

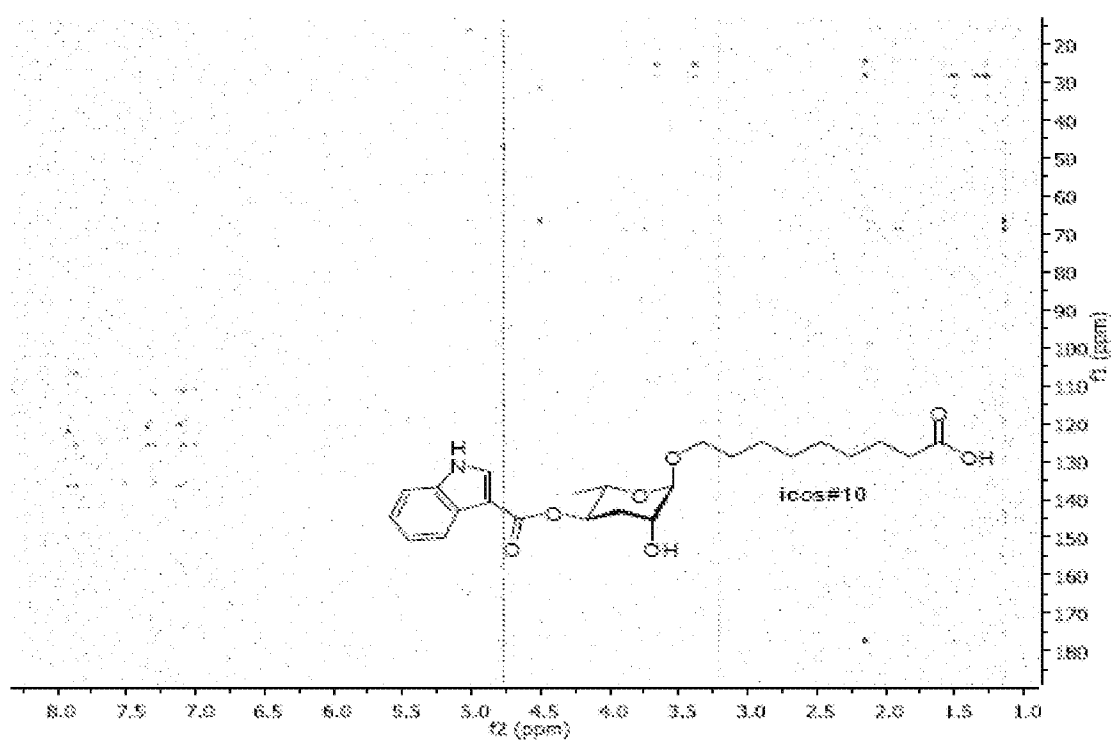


Figure 32D

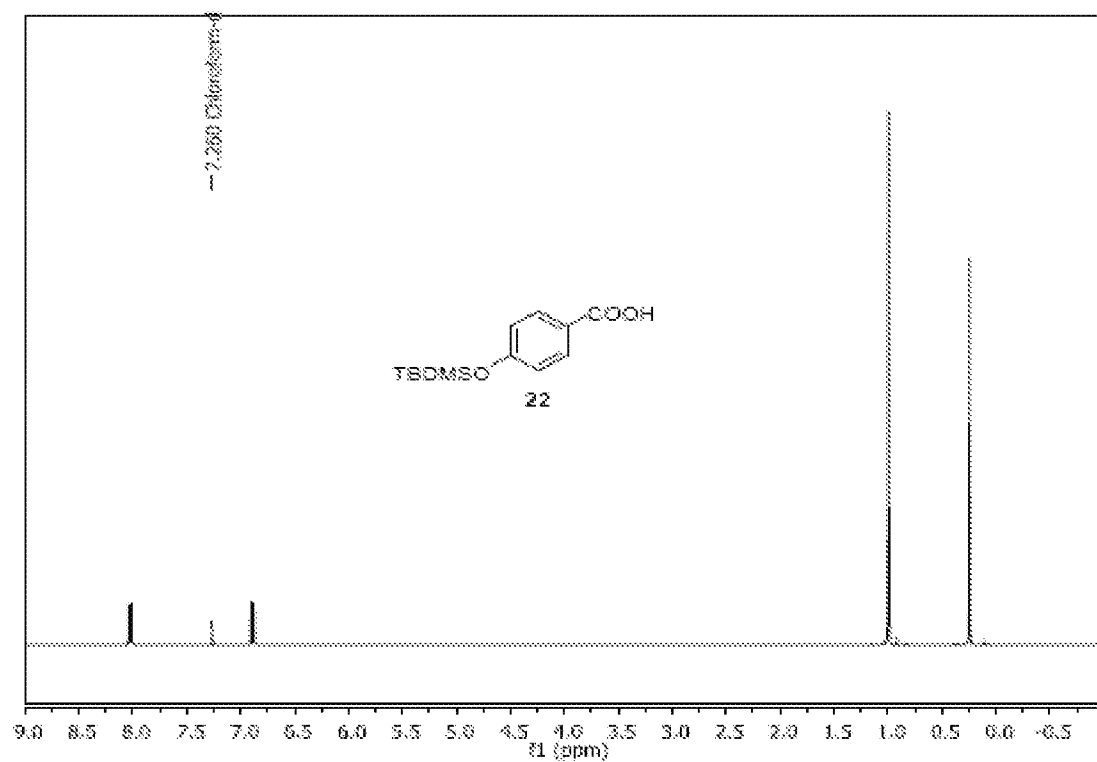


Figure 33A

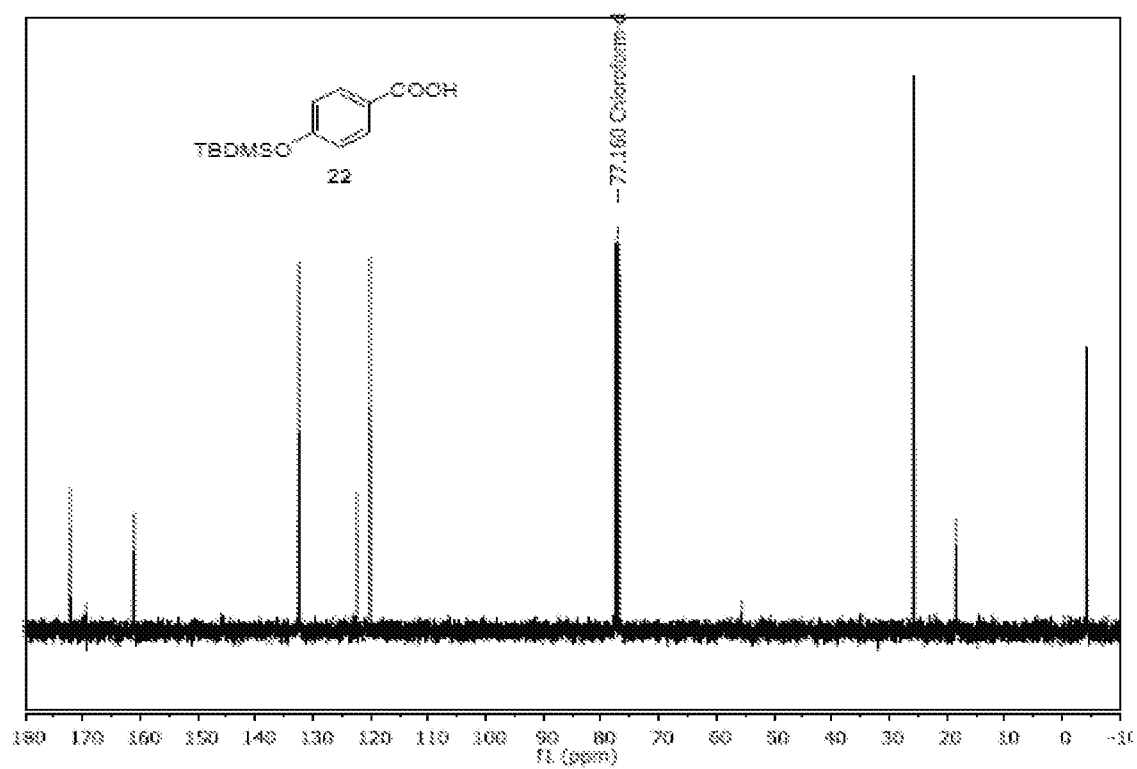


Figure 33B

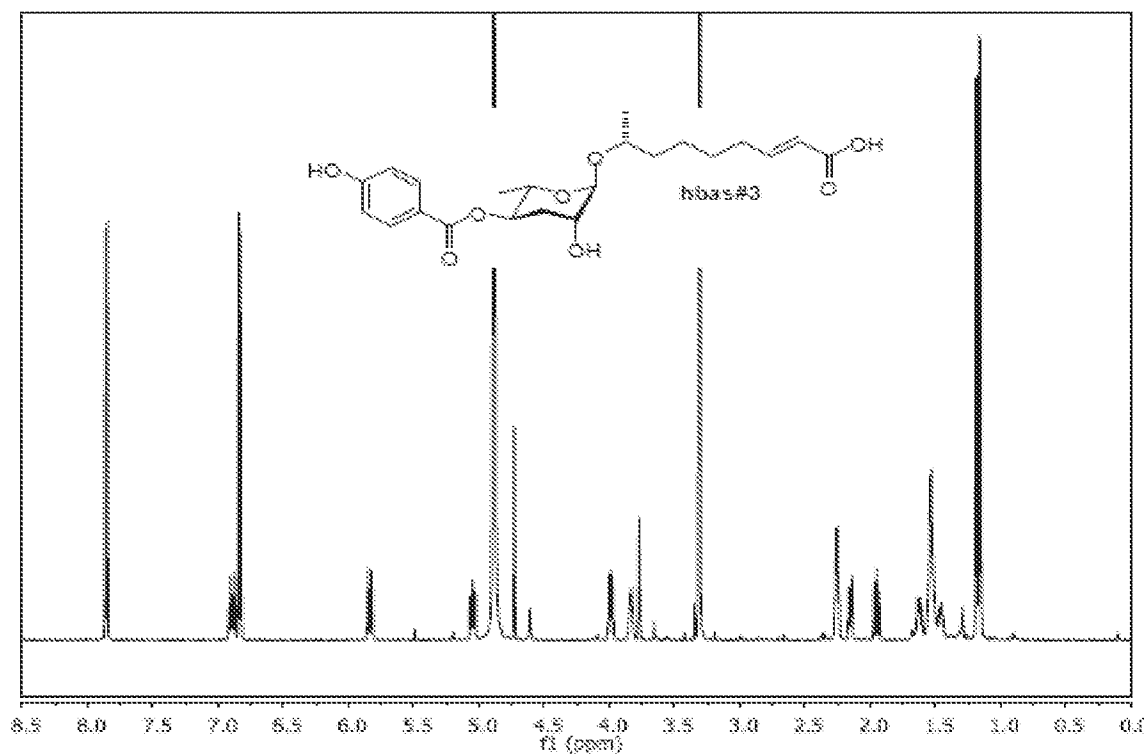


Figure 34A

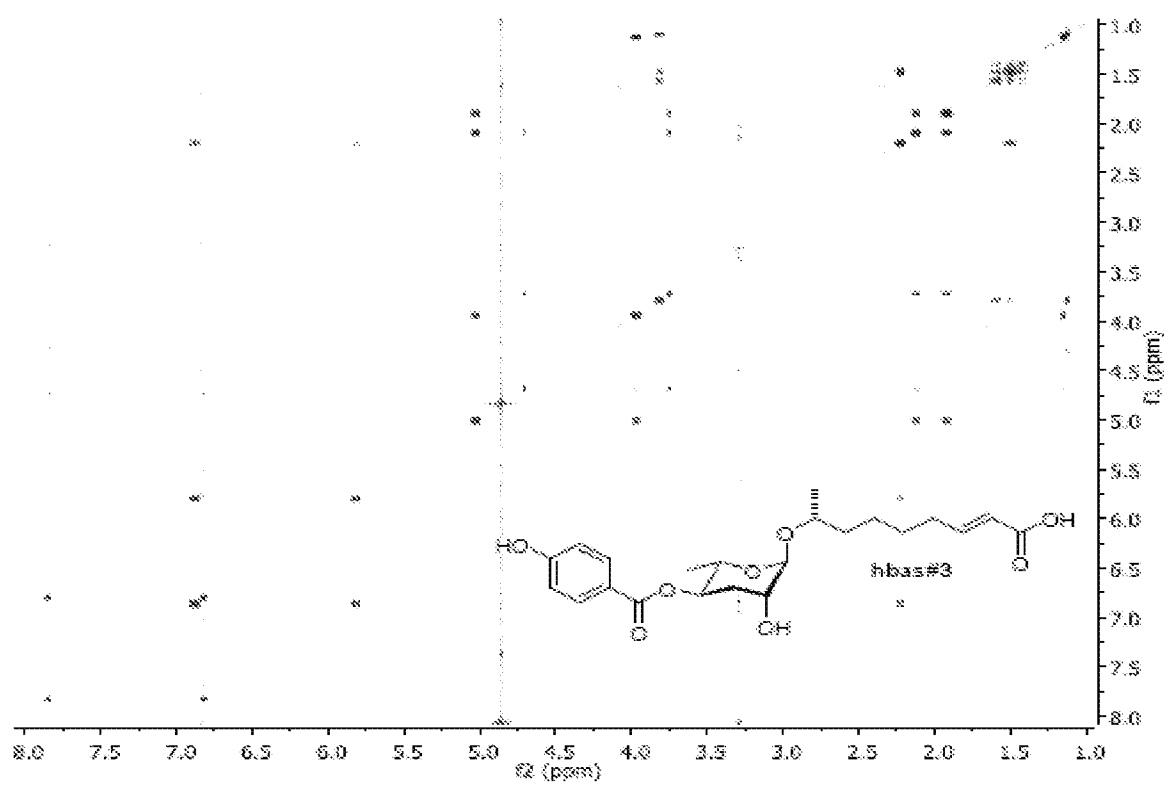


Figure 34B

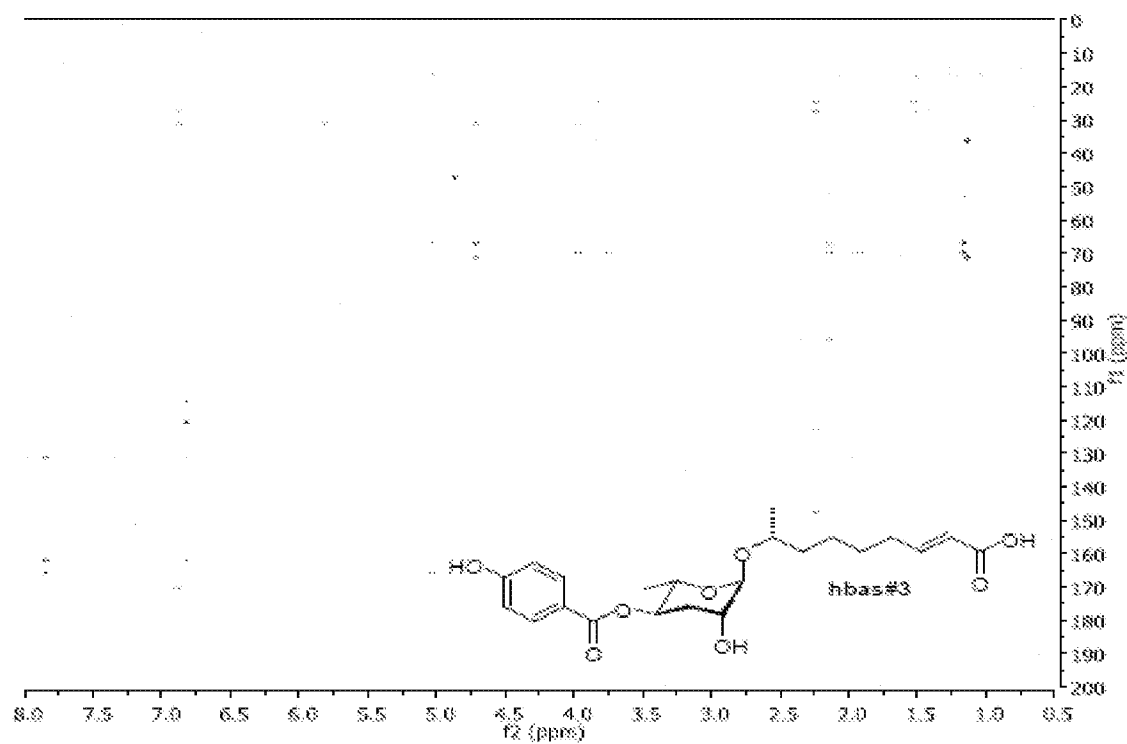


Figure 34C

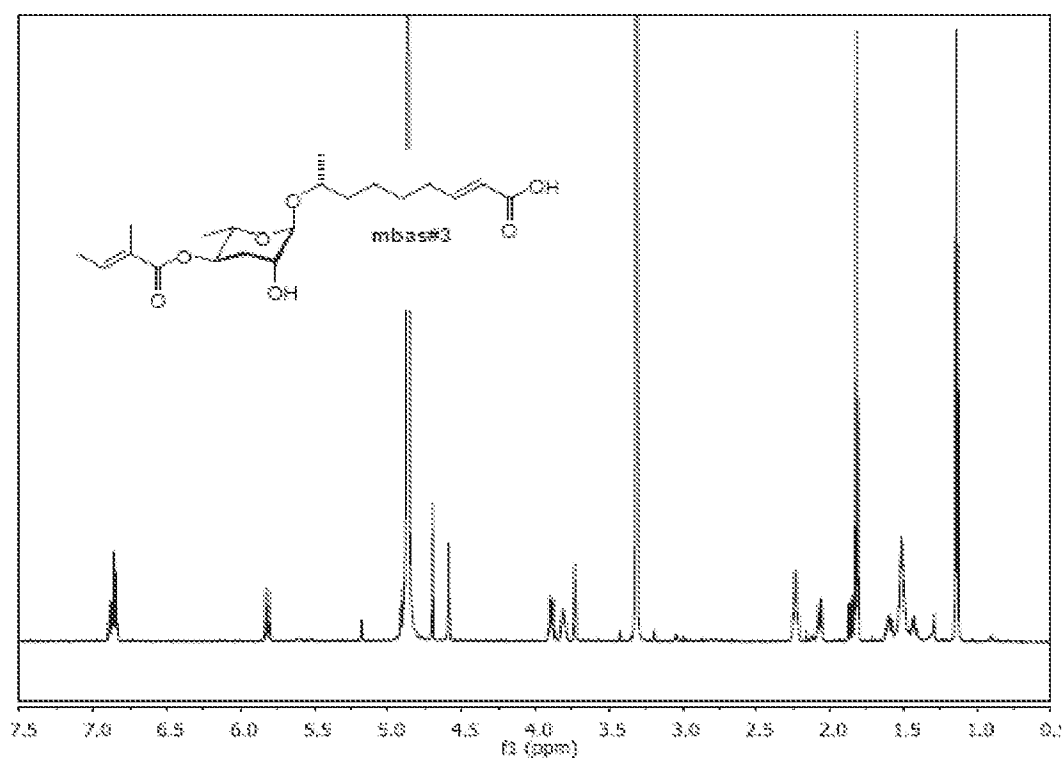


Figure 35A

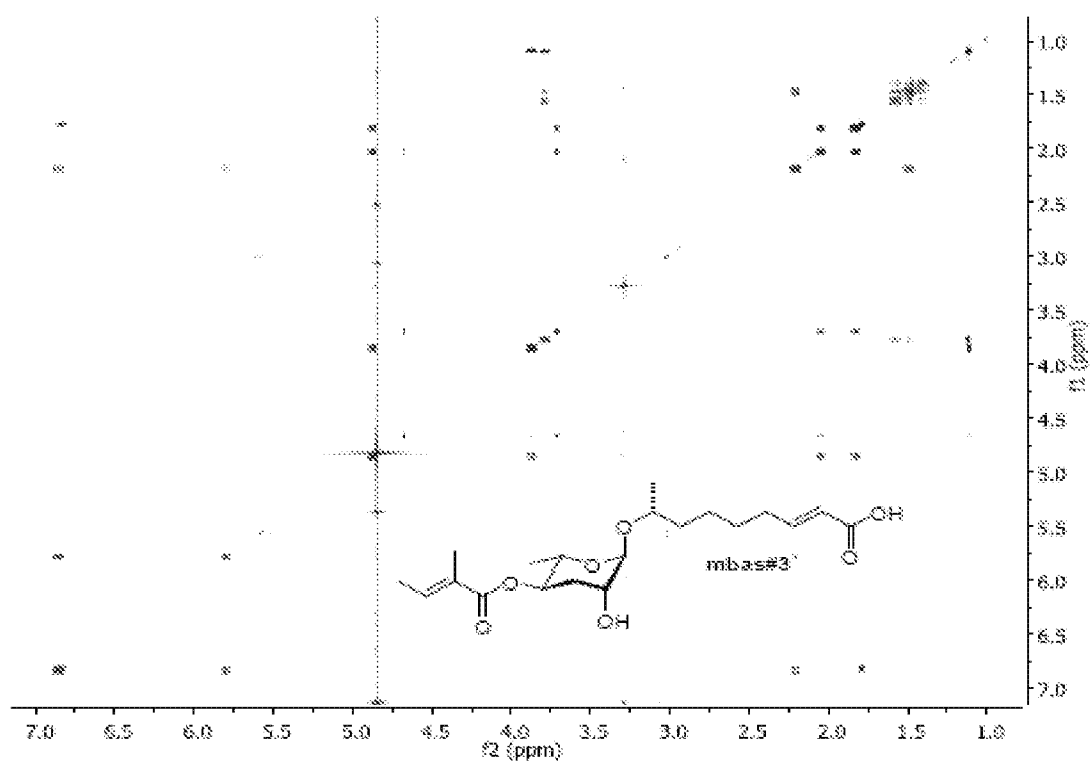


Figure 35B

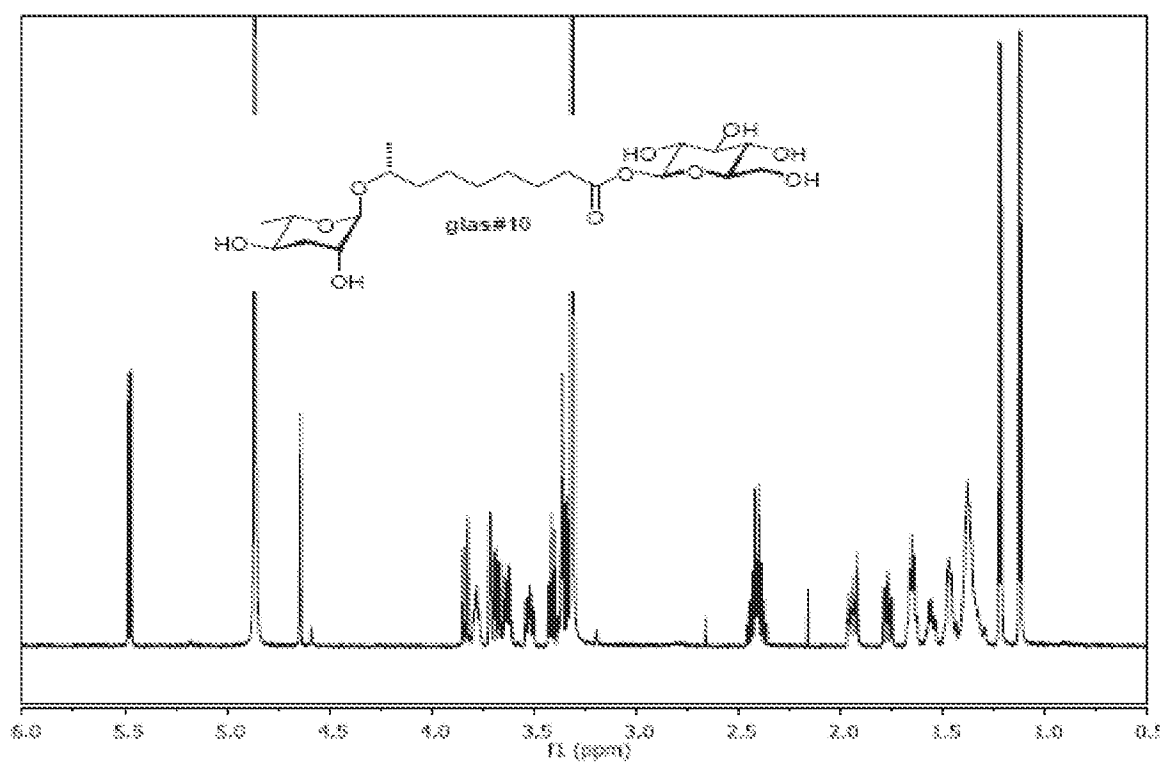


Figure 36A

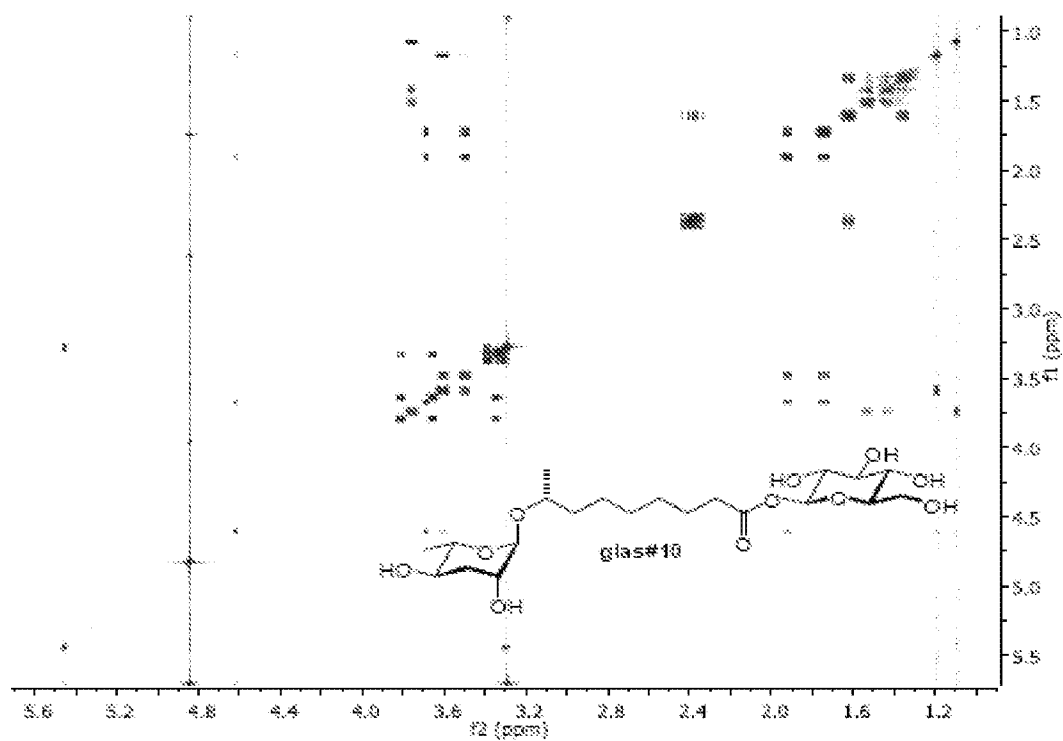


Figure 36B

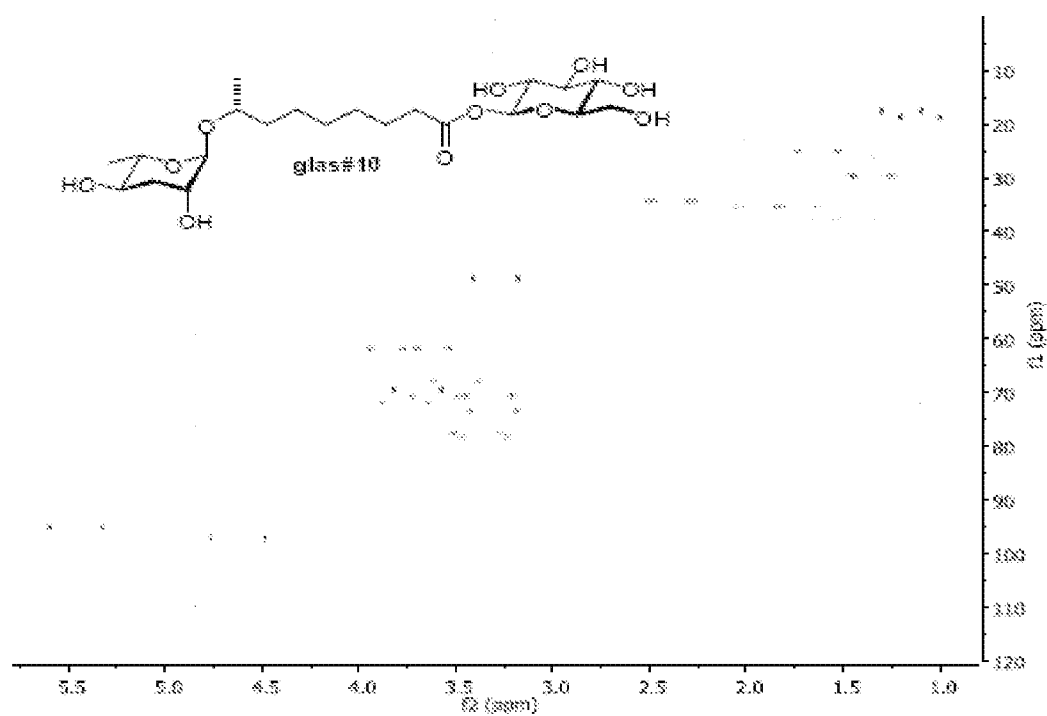


Figure 36C

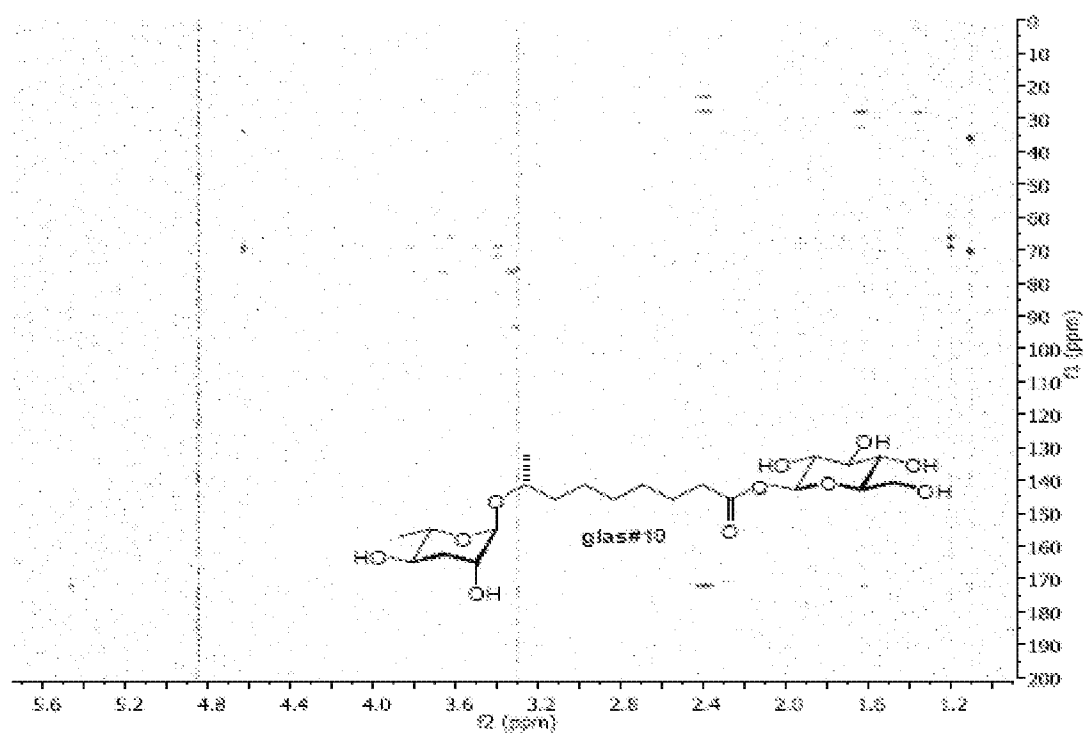
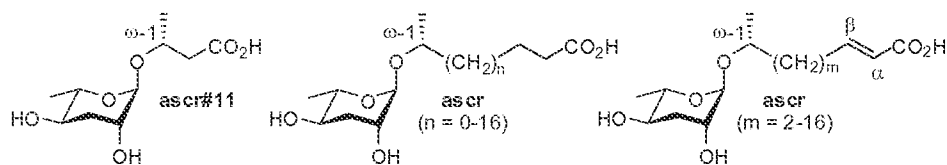


Figure 36D



Side chain length (n, m)	SMID [*]	Molecular formula	Molecular weight amu	m/z M-H-calculated	m/z M-H-observed	Retention time min ±SD
C4	ascr#11 [*]	C10H18O6	234.1103	233.1025	233.1031	10.21 ±0.03
C5, n = 0	ascr#9 [*]	C11H20O6	248.1260	247.1182	247.1189	11.69 ±0.01
C6, n = 1	ascr#12	C12H22O6	262.1416	261.1338	261.1343	13.09 ±0.02
ΔC7, m = 2	ascr#7 [*]	C13H22O6	274.1416	273.1339	273.1337	13.96 ±0.07
C7, n = 2	ascr#1 [*]	C13H24O6	276.1573	275.1495	275.1497	14.52 ±0.04
ΔC8, m = 3	ascr#13	C14H24O6	288.1573	287.1495	287.1481	15.61 ±0.18
C8, n = 3	ascr#14	C14H26O6	290.1729	289.1651	289.1647	15.96 ±0.03
ΔC9, m = 4	ascr#3 [*]	C15H26O6	302.1729	301.1651	301.1652	16.90 ±0.02
C9, n = 4	ascr#10 ^{*†}	C15H28O6	304.1886	303.1808	303.1800	17.44 ±0.02
ΔC10, m = 5	ascr#15	C16H28O6	316.1886	315.1808	315.1817	18.34 ±0.02
C10, n = 5	ascr#16	C16H30O6	318.2042	317.1964	317.1959	18.98 ±0.05
ΔC11, m = 6	ascr#17	C17H30O6	330.2042	329.1964	329.1957	19.75 ±0.02
C11, n = 6	ascr#18	C17H32O6	332.2199	331.2121	331.2130	20.43 ±0.02
ΔC12, m = 7	ascr#19	C18H32O6	344.2199	343.2121	343.2120	21.36 ±0.03
C12, n = 7	ascr#20	C18H34O6	346.2355	345.2277	345.2278	21.97 ±0.03
ΔC13, m = 8	ascr#21	C19H34O6	358.2355	357.2277	357.2273	22.83 ±0.02
C13, n = 8	ascr#22	C19H36O6	360.2512	359.2434	359.2437	23.58 ±0.02
ΔC14, m = 9	ascr#23	C20H36O6	372.2512	371.2434	371.2444	24.46 ±0.01
C14, n = 9	ascr#24	C20H38O6	374.2668	373.2590	373.2596	25.29 ±0.03
ΔC15, m = 10	ascr#25	C21H38O6	386.2668	385.2590	385.2598	26.15 ±0.02
C15, n = 10	ascr#26	C21H40O6	388.2825	387.2747	387.2743	27.09 ±0.02
ΔC16, m = 11	ascr#27	C22H40O6	400.2825	399.2747	399.2734	27.89 ±0.03
C16, n = 11	ascr#28	C22H42O6	402.2981	401.2903	401.2901	28.97 ±0.04
ΔC17, m = 12	ascr#29	C23H42O6	414.2981	413.2903	413.2891	29.80 ±0.03
C17, n = 12	ascr#30	C23H44O6	416.3138	415.3060	415.3067	30.96 ±0.03
C18, m = 13	ascr#31	C24H44O6	428.3138	427.3060	427.3075	31.78 ±0.03
C18, n = 13	ascr#32	C24H46O6	430.3294	429.3216	429.3221	33.02 ±0.02
ΔC19, m = 14	ascr#33	C25H46O6	442.3294	441.3216	441.3215	33.74 ±0.03
C19, n = 14	ascr#34	C25H48O6	444.3451	443.3373	443.3374	35.12 ±0.08
ΔC20, m = 15	ascr#35	C26H48O6	456.3451	455.3373	455.3371	35.59 ±0.06
C20, n = 15	ascr#36	C26H50O6	458.3607	457.3529	457.3501	37.14 ±0.07

$\Delta C21$, m = 16	ascr#37	C27H50 O6	470.3607	469.3529	469.3519	37.71 $\pm$ 0.13
C21, n = 16	ascr#38	C27H52 O6	472.3764	471.3686	471.3697	39.15 $\pm$ 0.06

² Pungalaya et al., *Proc. Nat'l Acad. Sci. U.S.A.* 106:7708--13 (2009), which is hereby incorporated by reference in its entirety.

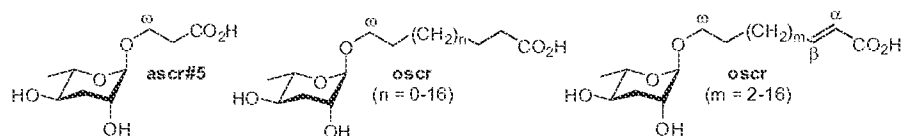
⁴ Srinivasan et al., *Pub. Lib. Sci. Biol.* 10:e1001237 (2012), which is hereby incorporated by reference in its entirety.

⁵ Jeong et al., *Nature* 433:541--45 (2005), which is hereby incorporated by reference in its entirety.

⁶ Butcher et al., *Nat. Chem. Biol.* 3:420--22 (2007), which is hereby incorporated by reference in its entirety.

⁷ Butcher et al., *Proc. Nat'l Acad. Sci. U.S.A.* 106:1875--79 (2009), which is hereby incorporated by reference in its entirety.

Figure 37A

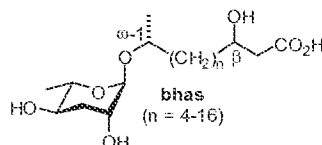


Side chain length (n, m)	SMID ⁵	Molecular formula	Molecular weight amu	m/z M-H-calculated	m/z M-H-observed	Retention time min ±SD
C3	ascr#5 *8	C9H16O6	220.0947	219.0869	219.0871	6.53 ±0.07
C5, n = 0	oscr#9 *	C11H20O6	248.1260	247.1182	247.1192	11.88 ±0.02
C6, n = 1	oscr#1 2	C12H22O6	262.1416	261.1338	261.1345	13.40 ±0.02
ΔC7, m = 2	oscr#7	C13H22O6	274.1416	273.1339	273.1350	14.67 ±0.02
C7, n = 2	oscr#1	C13H24O6	276.1573	275.1495	275.1503	14.95 ±0.03
C8, n = 3	oscr#1 4	C14H26O6	290.1729	289.1651	289.1672	16.45 ±0.02
ΔC9, n = 4	oscr#3	C15H26O6	302.1729	301.1651	301.1636	17.58 ±0.03
C9, n = 4	oscr#1 0*	C15H28O6	304.1886	303.1808	303.1814	18.00 ±0.02
ΔC10, m = 5	oscr#1 5	C16H28O6	316.1886	315.1808	315.1816	18.91 ±0.06
C10, n = 5	oscr#1 6	C16H30O6	318.2042	317.1964	317.1967	19.48 ±0.03
ΔC11, m = 6	oscr#1 7	C17H30O6	330.2042	329.1964	329.1956	20.39 ±0.01
C11, n = 6	oscr#1 8	C17H32O6	332.2199	331.2121	331.2124	20.98 ±0.08
ΔC12, m = 7	oscr#1 9	C18H32O6	344.2199	343.2121	343.2125	21.86 ±0.06
C12, n = 7	oscr#2 0	C18H34O6	346.2355	345.2277	345.2302	22.54 ±0.03
ΔC13, m = 8	oscr#2 1	C19H34O6	358.2355	357.2277	357.2271	23.41 ±0.02
C13, n = 8	oscr#2 2	C19H36O6	360.2512	359.2434	359.2452	24.19 ±0.02
ΔC14, m = 9	oscr#2 3	C20H36O6	372.2512	371.2434	371.2436	25.04 ±0.03
C14, n = 9	oscr#2 4	C20H38O6	374.2668	373.2590	373.2589	25.91 ±0.02
ΔC15, m = 10	oscr#2 5	C21H38O6	386.2668	385.2590	385.2567	26.74 ±0.01
C15, n = 10	oscr#2 6	C21H40O6	388.2825	387.2747	387.2739	27.73 ±0.02
ΔC16, m = 11	oscr#2 7	C22H40O6	400.2825	399.2747	399.2728	28.54 ±0.03
C16, n = 11	oscr#2 8	C22H42O6	402.2981	401.2903	401.2905	29.67 ±0.02
ΔC17, m = 12	oscr#2 9	C23H42O6	414.2981	413.2903	413.2900	30.42 ±0.02
C17, n = 12	oscr#3 0	C23H44O6	416.3138	415.3060	415.3080	31.68 ±0.04
ΔC18, m = 13	oscr#3 1	C24H44O6	428.3138	427.3060	427.3053	32.44 ±0.02
C18, n = 13	oscr#3 2	C24H46O6	430.3294	429.3216	429.3207	33.78 ±0.02
ΔC19, m = 14	oscr#3 3	C25H46O6	442.3294	441.3216	441.3218	34.44 ±0.05
C19, n = 14	oscr#3 4	C25H48O6	444.3451	443.3373	443.3372	35.86 ±0.05
ΔC20, m = 15	oscr#3 5	C26H48O6	456.3451	455.3373	455.3384	36.23 ±0.05
C20, n = 15	oscr#3 6	C26H50O6	458.3607	457.3529	457.3545	37.96 ±0.05
ΔC21, m = 16	oscr#3 7	C27H50O6	470.3607	469.3529	469.3504	38.08 ±0.19

C21, n = 16	oscr#3 8	C27H52 O6	472.3764	471.3686	471.3679	40.19 ±0.13
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⁸ Butcher et al., *Proc. Nat'l Acad. Sci. U.S.A.* 105:14288–92 (2008), which is hereby incorporated by reference in its entirety.

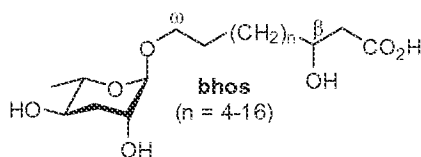
Figure 37B



Side chain length (n)	SMID ⁵	Molecular formula	Molecular weight amu	m/z M-H-calculated	m/z M-H-observed	Retention time min ±SD
C ₉ , n = 4	bhas#1 0*	C ₁₅ H ₂₈ O ₇	320.1835	319.1757	319.1764	13.98 ±0.03
C ₁₀ , n = 5	bhas#1 6	C ₁₆ H ₃₀ O ₇	334.1992	333.1913	333.1909	15.55 ±0.03
C ₁₁ , n = 6	bhas#1 8	C ₁₇ H ₃₂ O ₇	348.2148	347.2070	347.2058	16.72 ±0.02
C ₁₂ , n = 7	bhas#2 0	C ₁₈ H ₃₄ O ₇	362.2305	361.2226	361.2235	18.02 ±0.02
C ₁₃ , n = 8	bhas#2 2*	C ₁₉ H ₃₆ O ₇	376.2461	375.2383	375.2371	19.39 ±0.02
C ₁₄ , n = 9	bhas#2 4 ⁷	C ₂₀ H ₃₈ O ₇	390.2618	389.2539	389.2537	20.74 ±0.03
C ₁₅ , n = 10	bhas#2 6 ⁷	C ₂₁ H ₄₀ O ₇	404.2774	403.2696	403.2693	22.19 ±0.03
C ₁₆ , n = 11	bhas#2 8 ⁷	C ₂₂ H ₄₂ O ₇	418.2931	417.2852	417.2852	23.66 ±0.03
C ₁₇ , n = 12	bhas#3 0 ⁷	C ₂₃ H ₄₄ O ₇	432.3087	431.3009	431.3009	25.32 ±0.02
C ₁₈ , n = 13	bhas#3 2	C ₂₄ H ₄₆ O ₇	446.3244	445.3165	445.3164	27.07 ±0.03
C ₁₉ , n = 14	bhas#3 4	C ₂₅ H ₄₈ O ₇	460.3400	459.3322	459.3316	28.97 ±0.02
C ₂₀ , n = 15	bhas#3 6	C ₂₆ H ₅₀ O ₇	474.3557	473.3478	473.3471	30.97 ±0.03
C ₂₁ , n = 16	bhas#3 8	C ₂₇ H ₅₂ O ₇	488.3713	487.3635	487.3629	33.12 ±0.06

⁷ Butcher et al., *Proc. Nat'l Acad. Sci. U.S.A.* 106:1875–79 (2009), which is hereby incorporated by reference in its entirety.

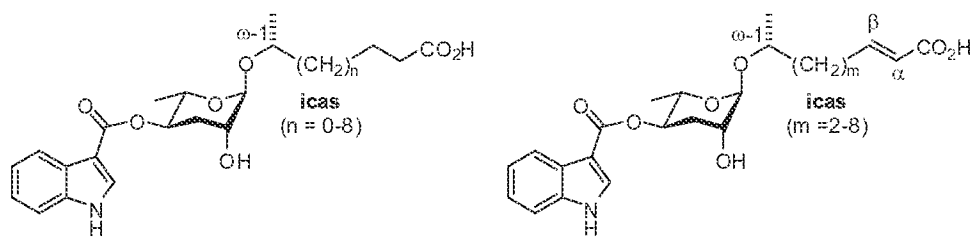
Figure 37C



Side chain length (n)	SMID ⁵	Molecular formula	Molecular weight amu	m/z M-H-calculated	m/z M-H-observed	Retention time min ±SD
C ₉ , n = 4	bhos#1 0	C ₁₅ H ₂₈ O ₇	320.1835	319.1757	319.1781	14.44 ±0.01
C ₁₀ , n = 5	bhos#1 6	C ₁₆ H ₃₀ O ₇	334.1992	333.1913	333.1902	15.82 ±0.01
C ₁₁ , n = 6	bhos#1 8	C ₁₇ H ₃₂ O ₇	348.2148	347.2070	347.2054	17.17 ±0.02
C ₁₂ , n = 7	bhos#2	C ₁₈ H ₃₄ O ₇	362.2305	361.2226	361.2224	18.51 ±0.01

	0					
C ₁₃ , n = 8	bhos#2	C ₁₉ H ₃₅ O ₇	376.2461	375.2383	375.2370	19.86 ±0.02
C ₁₄ , n = 9	bhos#2	C ₂₀ H ₃₈ O ₇	390.2618	389.2539	389.2537	21.23 ±0.03
C ₁₅ , n = 10	bhos#2	C ₂₁ H ₄₀ O ₇	404.2774	403.2696	403.2691	22.70 ±0.01
C ₁₆ , n = 11	bhos#2	C ₂₂ H ₄₂ O ₇	418.2931	417.2852	417.2849	24.28 ±0.06
C ₁₇ , n = 12	bhos#3	C ₂₃ H ₄₄ O ₇	432.3087	431.3009	431.3005	25.95 ±0.07
C ₁₈ , n = 13	bhos#3	C ₂₄ H ₄₆ O ₇	446.3244	445.3165	445.3164	27.73 ±0.02
C ₁₉ , n = 14	bhos#3	C ₂₅ H ₄₈ O ₇	460.3400	459.3322	459.3318	29.68 ±0.03
C ₂₀ , n = 15	bhos#3	C ₂₆ H ₅₀ O ₇	474.3557	473.3478	473.3473	31.76 ±0.03
C ₂₁ , n = 16	bhos#3	C ₂₇ H ₅₂ O ₇	488.3713	487.3635	487.3640	33.96 ±0.08

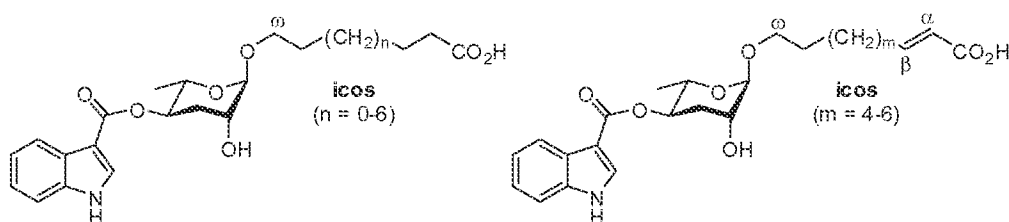
Figure 37D



Side chain length (n, m)	SMID ⁵	Molecular formula	Molecular weight amu	m/z M-H--calculated	m/z M-H--observed	Retention time min ±SD	W T (N 2)	acox-1 (ok255 7)	maoc -1 (hj13)	dhs-28 (hj8)	daf-22 (ok69 3)
C ₅ , n = 0	icas#9	C ₂₀ H ₂₅ NO ₇	391.1631	390.1553	390.1555	20.18 ±0.02	+	+	-	-	-
C ₆ , n = 1	icas#1	C ₂₁ H ₂₇ NO ₇	405.1788	404.1709	404.1703	21.02 ±0.04	+	+	-	-	-
ΔC ₇ , m = 2	icas#7	C ₂₂ H ₂₇ NO ₇	417.1788	416.1709	416.1719	21.74 ±0.03	+	-	-	-	-
C ₇ , n = 2	icas#1	C ₂₂ H ₂₉ NO ₇	419.1944	418.1866	418.1864	22.06 ±0.03	+	+	-	-	-
C ₈ , n = 3	icas#1	C ₂₃ H ₃₁ NO ₇	433.2101	432.2022	432.2013	23.20 ±0.04	+	+	-	-	-
ΔC ₉ , m = 4	icas#3	C ₂₄ H ₃₁ NO ₇	445.2101	444.2022	444.2029	23.94 ±0.03	+	-	+	-	-
C ₉ , n = 4	icas#1	C ₂₄ H ₃₃ NO ₇	447.2257	446.2179	446.2185	24.55 ±0.03	+	+	+	+	-
ΔC ₁₀ , m = 5	icas#1	C ₂₅ H ₃₅ NO ₇	459.2257	458.2179	458.2198	25.21 ±0.05	-	-	+	+	-
C ₁₀ , n = 5	icas#1	C ₂₅ H ₃₇ NO ₇	461.2414	460.2335	460.2369	25.89 ±0.04	+	+	+	+	-
ΔC ₁₁ , m = 6	icas#1	C ₂₆ H ₃₅ NO ₇	473.2414	472.2335	472.2344	26.68 ±0.04	-	-	+	+	-
C ₁₁ , n = 6	icas#1	C ₂₆ H ₃₇ NO ₇	475.2570	474.2492	474.2494	27.44 ±0.03	+	+	+	+	-
ΔC ₁₂ , m = 7	icas#1	C ₂₇ H ₃₇ NO ₇	487.2570	486.2492	486.2486	28.20 ±0.06	-	-	+	+	-
C ₁₂ , n = 7	icas#2	C ₂₇ H ₃₉ NO ₇	489.27265	488.2648	488.2628	28.98 ±0.03	-	+	+	+	-
ΔC ₁₃ , m = 8	icas#2	C ₂₈ H ₃₉ NO ₇	501.27265	500.2648	500.2640	29.71 ±0.04	-	-	+	+	-
C ₁₃ , n = 8	icas#2	C ₂₈ H ₄₁ NO ₇	503.28830	502.2805	502.2807	30.66 ±0.03	-	+	+	+	-

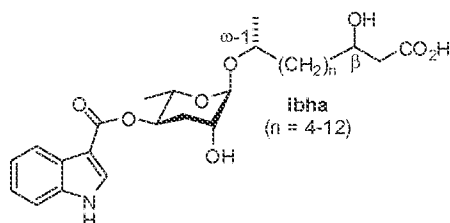
⁴ Srinivasan et al., *Pub. Lib. Sci. Biol.* 10:e1001237 (2012), which is hereby incorporated by reference in its entirety.⁵ Butcher et al., *Org. Lett.* 11:3100-03 (2009), which is hereby incorporated by reference in its entirety.

Figure 37E



Side chain length (n, m)	SMID ^S	Molecular formula	Molecular weight amu	m/z M-H-calculated	m/z M-H-observed	Retention time min ±SD	W T (N 2)	acox-1 (ok255 7)	maoc -1 (hj13)	dhs-28 (hj8)	daf-22 (ok69 3)
C ₅ , n = 0	icos#9	C ₂₀ H ₂₅ NO ₇	391.1631	390.1553	390.1541	20.48 ±0.03	-	+	-	-	-
C ₇ , n = 2	icos#1	C ₂₂ H ₂₉ NO ₇	419.1944	418.1866	418.1858	22.63 ±0.03	+	+	-	-	-
ΔC ₉ , m = 4	icos#3	C ₂₄ H ₃₁ NO ₇	445.2101	444.2022	444.2049	24.66 ±0.04	-	-	+	-	-
C ₉ , n = 4	icos#1 0*	C ₂₄ H ₃₃ NO ₇	447.2257	446.2179	446.2171	25.29 ±0.03	-	+	+	+	-
ΔC ₁₀ , m = 5	icos#1	C ₂₅ H ₃₃ NO ₇	459.2257	458.2179	458.2170	26.02 ±0.05	-	-	+	+	-
C ₁₀ , n = 5	icos#1 6	C ₂₅ H ₃₅ NO ₇	461.2414	460.2335	460.2350	26.73 ±0.04	-	-	+	+	-
ΔC ₁₁ , m = 6	icos#1	C ₂₆ H ₃₅ NO ₇	473.2414	472.2335	472.2325	27.45 ±0.03	-	-	+	+	-
C ₁₁ , n = 6	icos#1 8	C ₂₆ H ₃₇ NO ₇	475.2570	474.2492	474.2490	28.21 ±0.04	-	+	+	+	-

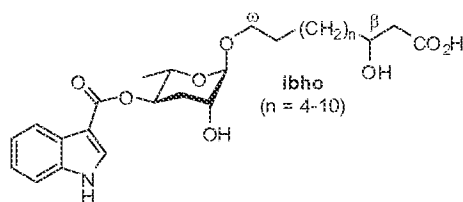
Figure 37F



Side chain length (n)	SMID ^S	Molecular formula	Molecular weight amu	m/z M-H-calculated	m/z M-H-observed	Retention time min ±SD	W T (N 2)	acox-1 (ok255 7)	maoc -1 (hj13)	dhs-28 (hj8)	daf-22 (ok69 3)
C ₉ , n = 4	ibha# 10	C ₂₄ H ₃₅ NO ₈	463.2206	462.2128	462.2113	20.72 ±0.03	-	-	-	+	-
C ₁₀ , n = 5	ibha# 16	C ₂₅ H ₃₅ NO ₈	477.2363	476.2284	476.2251	21.81 ±0.04	-	-	-	+	-
C ₁₁ , n = 6	ibha# 18	C ₂₆ H ₃₇ NO ₈	491.2519	490.2441	490.2476	23.08 ±0.04	-	-	+	+	-
C ₁₂ , n = 7	ibha# 20	C ₂₇ H ₃₉ NO ₈	505.2676	504.2597	504.2582	24.38 ±0.03	-	-	+	+	-
C ₁₃ , n = 8	ibha# 22	C ₂₈ H ₄₁ NO ₈	519.2832	518.2754	518.2726	25.86 ±0.03	-	-	+	+	-
C ₁₄ , n = 9	ibha# 24	C ₂₉ H ₄₃ NO ₈	533.2989	532.2910	532.2938	27.30 ±0.04	-	-	-	+	-
C ₁₅ , n = 10	ibha# 26	C ₃₀ H ₄₅ NO ₈	547.3145	546.3067	546.3033	28.96 ±0.04	-	-	-	+	+
C ₁₆ , n =	ibha#	C ₃₁ H ₄₇ NO ₈	561.3302	560.3223	560.3201	30.63	-	-	-	+	-

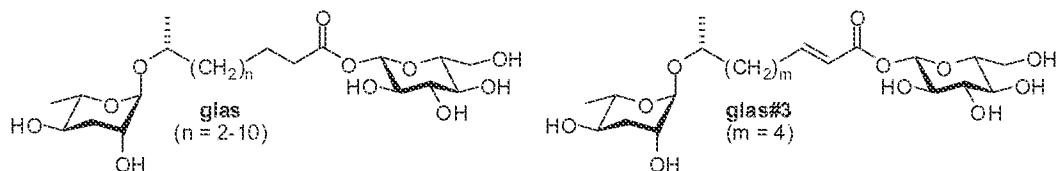
11	28					±0.05					
C ₁₇ , n =	ibha#	C ₃₂ H ₄₅ NO ₈	575.3458	574.3380	574.3347	32.46	-	-	-	+	-
12	30					±0.01					

Figure 37G



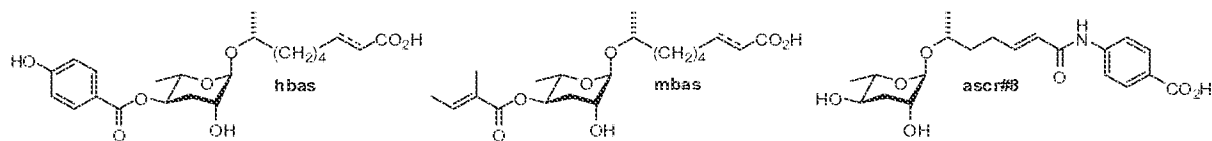
Side chain length (n)	SMID ^s	Molecular formula	Molecular weight amu	m/z M-H-calculated	m/z M-H-observed	Retention time min ±SD	W T (N2)	acox-1 {ok25 57}	ma oc-1 {hj 13}	dhs-28 {hj8}	daf-22 {ok6 93}
C ₉ , n = 4	ibho# 10	C ₂₄ H ₃₃ NO ₈	463.2206	462.2128	462.2114	21.39 ±0.03	-	-	-	+	-
C ₁₀ , n = 5	ibho# 16	C ₂₅ H ₃₅ NO ₈	477.2363	476.2284	476.2252	22.56 ±0.03	-	-	-	+	-
C ₁₁ , n = 6	ibho# 18	C ₂₆ H ₃₇ NO ₈	491.2519	490.2441	490.2438	23.79 ±0.03	-	-	+	+	-
C ₁₂ , n = 7	ibho# 20	C ₂₇ H ₃₉ NO ₈	505.2676	504.2597	504.2580	25.15 ±0.03	-	-	-	+	-
C ₁₃ , n = 8	ibho# 22	C ₂₈ H ₄₁ NO ₈	519.2832	518.2754	518.2719	26.58 ±0.03	-	-	-	+	-
C ₁₄ , n = 9	ibho# 24	C ₂₉ H ₄₃ NO ₈	533.2989	532.2910	532.2886	27.94 ±0.03	-	-	-	+	-
C ₁₅ , n = 10	ibho# 26	C ₃₀ H ₄₅ NO ₈	547.3145	546.3067	546.3094	29.65 ±0.03	-	-	-	+	+

Figure 37H



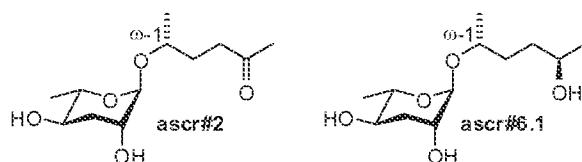
Side chain length (n, m)	SMID ⁵	Molecular formula	Molecular weight amu	<i>m/z</i> M+Cl ⁻ calculated	<i>m/z</i> M+Cl ⁻ observed	Retention time min ±SD	W T (N2)	1 acox-1 (ok25 57)	ma oc-1 (hj 13)	dhs-28 (hj8)	daf-22 (ok6 93)
C ₇ , n = 2	glas#1	C ₁₉ H ₃₂ O	438.2101	473.1795	473.1803	11.99 ±0.04	+	-	-	-	-
ΔC ₉ , m = 4	glas#3	C ₂₁ H ₃₆ O	464.2258	499.1952	499.1932	14.64 ±0.03	+	-	-	-	-
C ₉ , n = 4	glas#1 0*	C ₂₃ H ₃₈ O	466.2414	501.2108	501.2112	15.05 ±0.03	+	+	-	-	-
C ₁₀ , n = 5	glas#1 6	C ₂₂ H ₄₀ O	480.2571	515.2265	515.2269	16.19 ±0.05	-	+	-	-	-
C ₁₁ , n = 6	glas#1 8	C ₂₃ H ₄₂ O	494.2727	529.2421	529.2402	17.34 ±0.04	+	+	-	-	-
C ₁₂ , n = 7	glas#2 0	C ₂₄ H ₄₄ O	508.2884	543.2578	543.2551	18.41 ±0.04	-	+	-	-	-
C ₁₃ , n = 8	glas#2 2	C ₂₅ H ₄₆ O	522.3040	557.2734	557.2720	19.49 ±0.05	-	+	-	-	-
C ₁₄ , n = 9	glas#2 4	C ₂₆ H ₄₈ O	536.3197	571.2891	571.2896	20.59 ±0.04	-	+	-	-	-
C ₁₅ , n = 10	glas#2 6	C ₂₇ H ₅₀ O	550.3353	585.3047	585.3095	21.77 ±0.04	-	+	-	-	-

Figure 37I



Side chain length	SMID ⁵	Molecular formula	Molecular weight amu	<i>m/z</i> M-H ⁺ calculated	<i>m/z</i> M-H ⁺ observed	Retention time min ±SD	W T (N2)	1 acox-1 (ok25 57)	ma oc-1 (hj13)	dhs-28 (hj8)	daf-22 (ok6 93)
ΔC ₇	ascr#8* 2	C ₂₀ H ₂₇ N O ₇	393.1788	392.1709	392.1712	16.77 ±0.04	+	-	-	-	-
ΔC ₉	hbas#3 *	C ₂₂ H ₃₀ O	422.1941	421.1862	421.1866	22.41 ±0.03	+	-	-	-	-
C ₉	hbas#1 0	C ₂₂ H ₃₂ O	424.2097	423.2019	423.2018	22.94 ±0.04	-	+	-	-	-
ΔC ₉	mbas# 3*	C ₂₀ H ₃₂ O	384.2148	383.2070	383.2079	25.66 ±0.04	+	-	-	-	-
C ₉	mbas# 10	C ₂₀ H ₃₄ O	386.2305	385.2226	385.2239	26.38 ±0.04	-	+	-	-	-

Figure 37J



Side chain length	SMID ^S	Molecular formula	Molecular weight amu	<i>m/z</i> M+Na+ calculated	<i>m/z</i> M+Na+ observed	Retention time min ±SD
C ₆	_{*6} ascr#2	C ₁₂ H ₂₂ O ₅	246.1467	269.1365	269.1371	13.06 ±0.03
C ₆	_{*2} ascr#6	C ₁₂ H ₂₄ O ₅	248.1624	271.1521	271.1532	12.72 ±0.04

². Pungaliya et al., *Proc. Natl. Acad. Sci. U.S.A.* 106:7708--13 (2009), which is hereby incorporated by reference in its entirety.

⁶. Butcher et al., *Nat. Chem. Biol.* 3:420–22 (2007), which is hereby incorporated by reference in its entirety.

Figure 37K

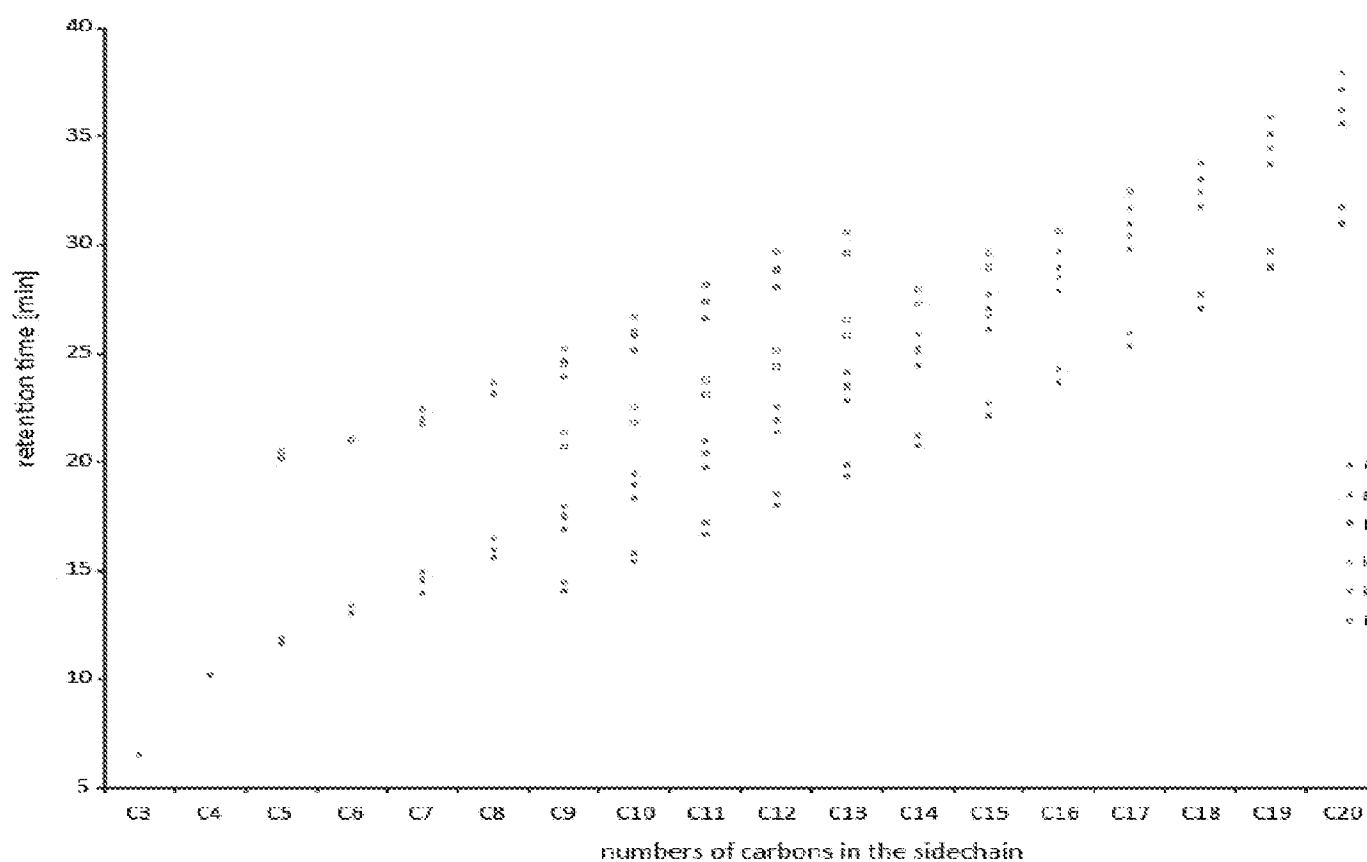
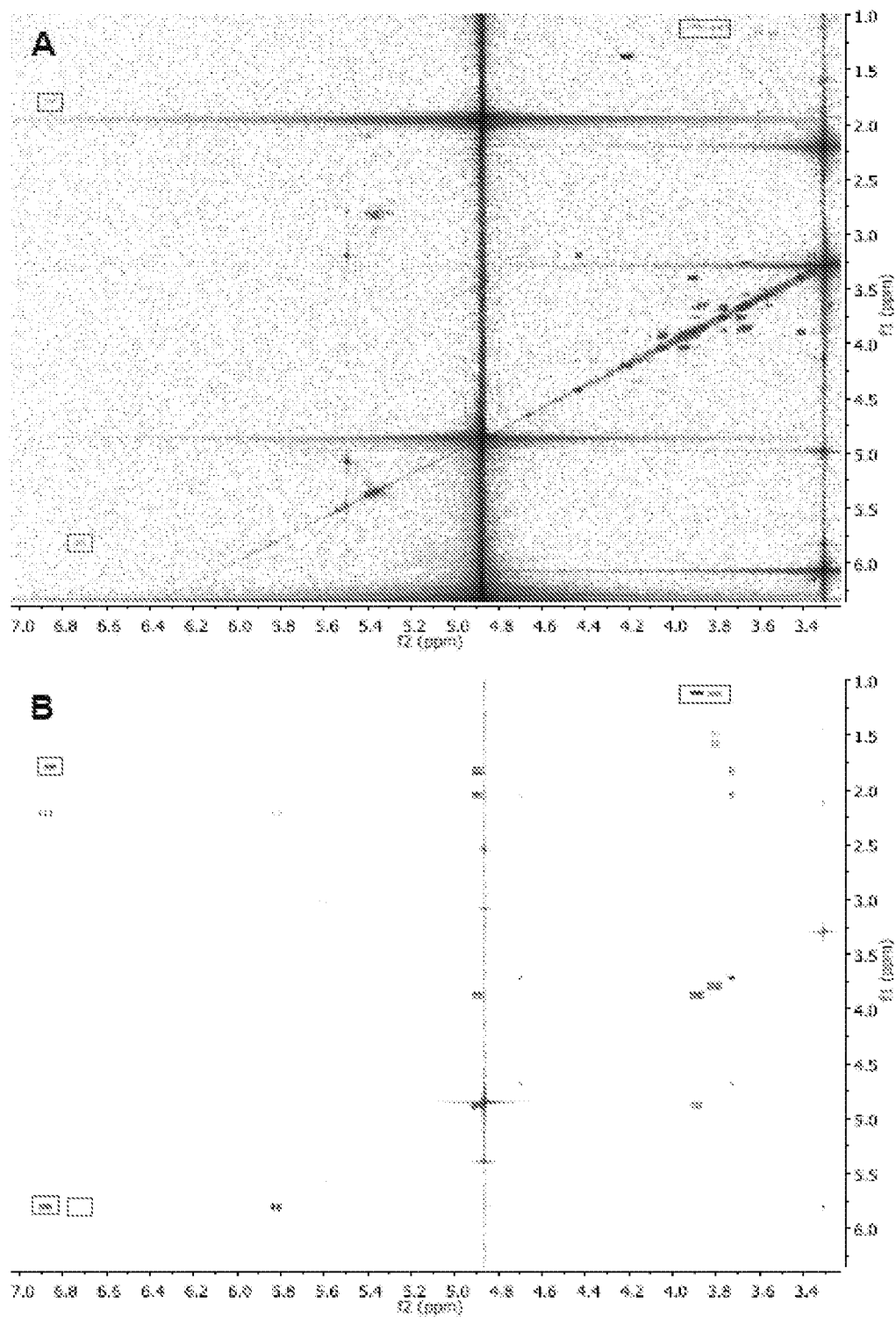
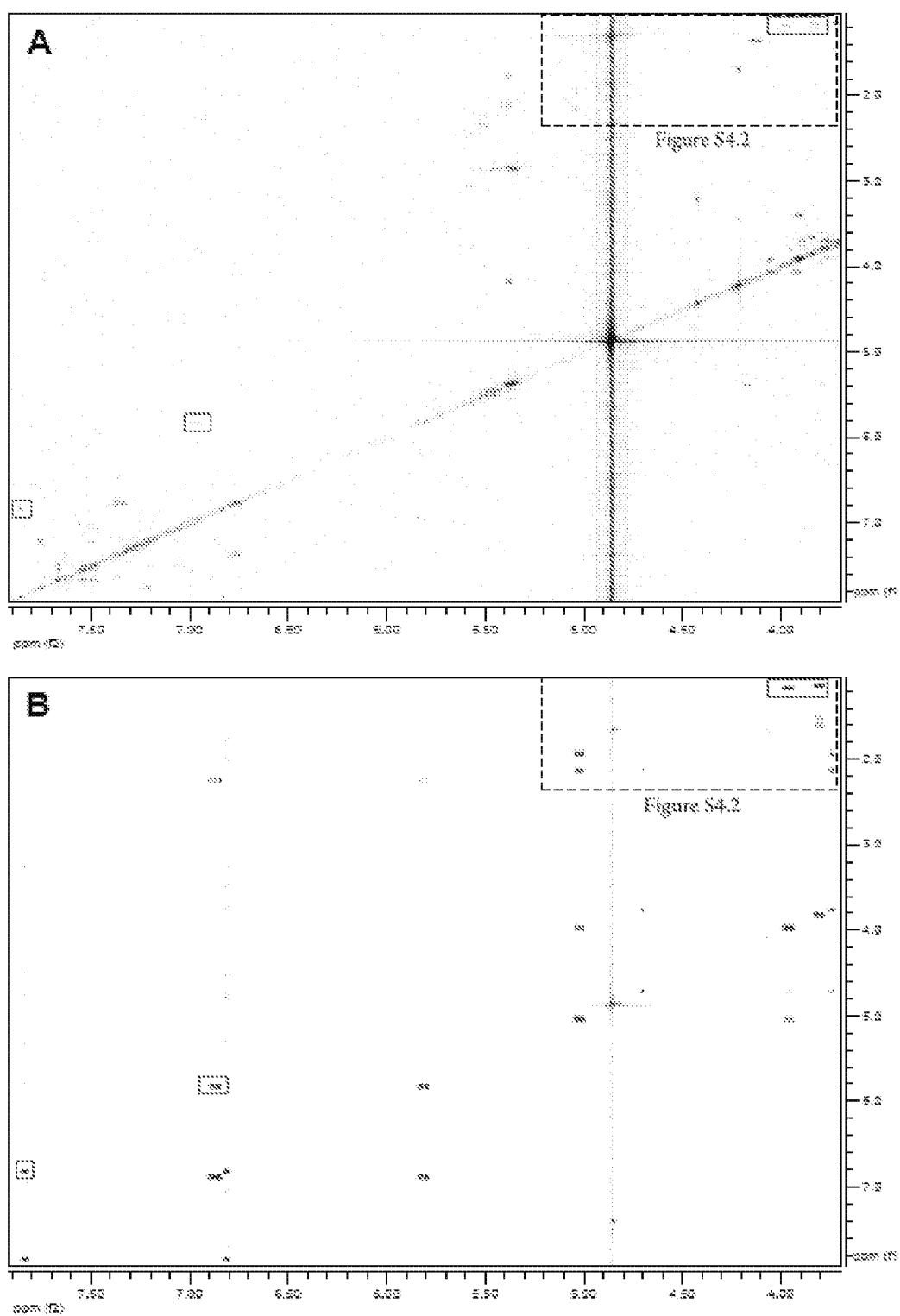


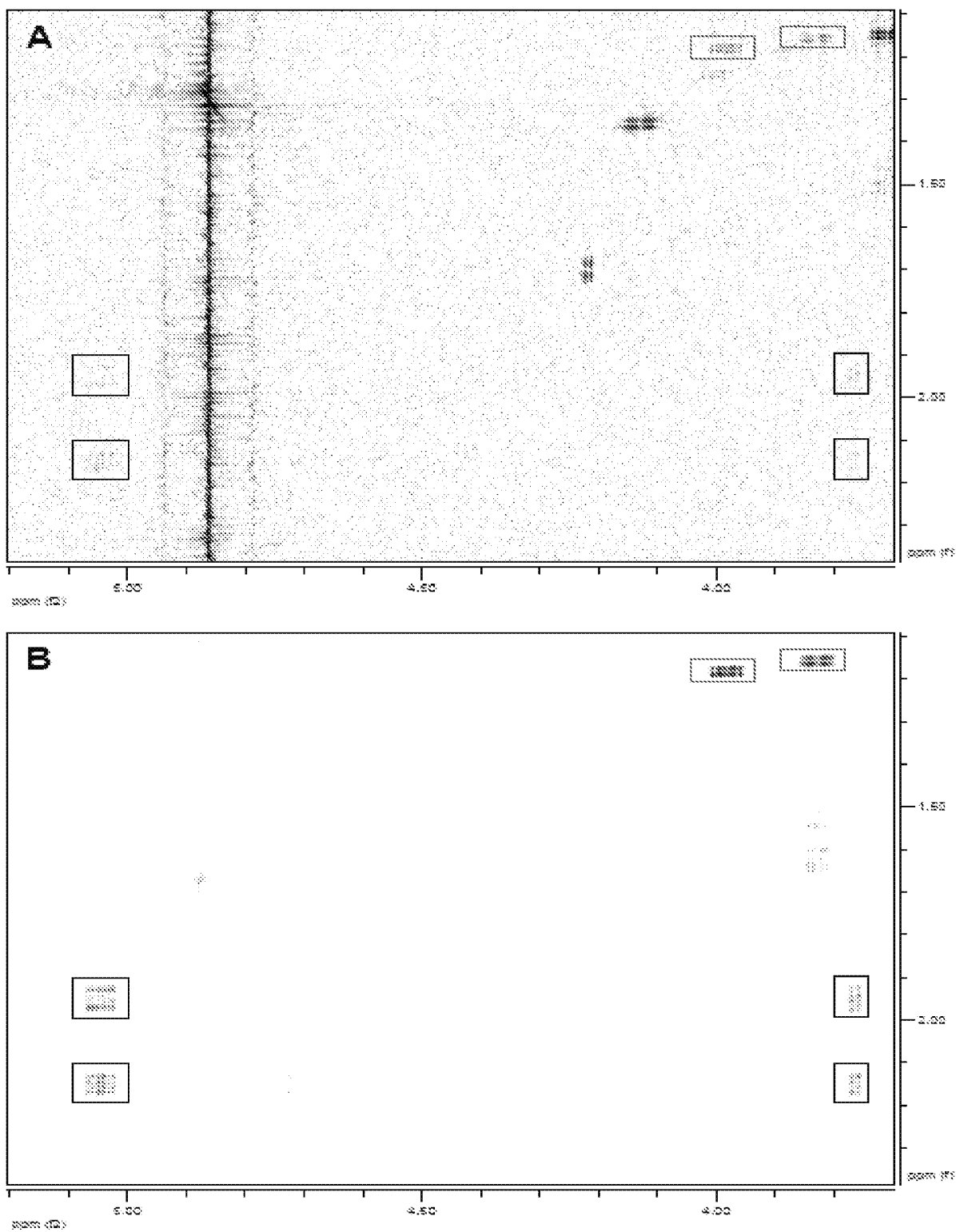
Figure 38



Figures 39A-B



Figures 40A–B



Figures 41A-B

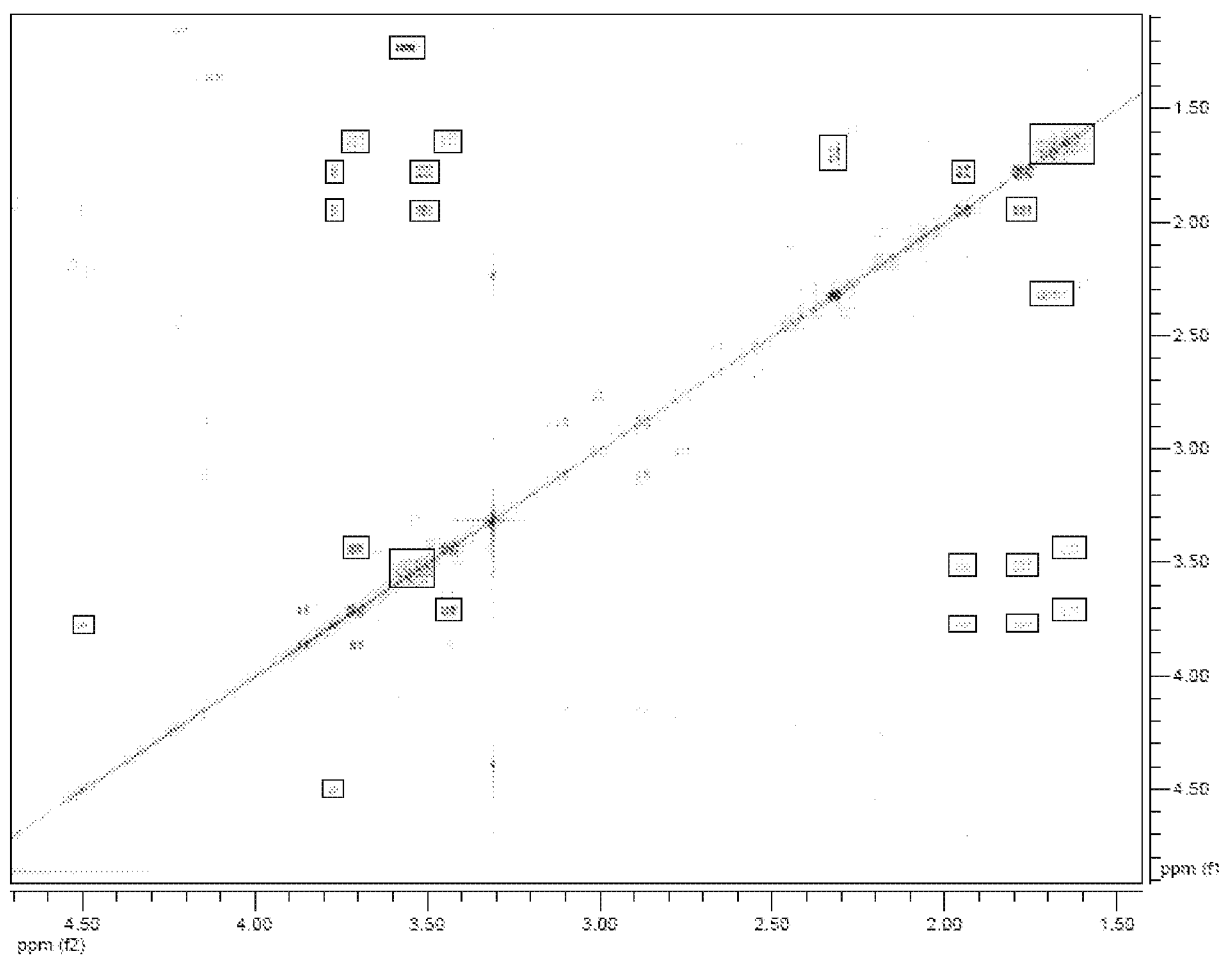
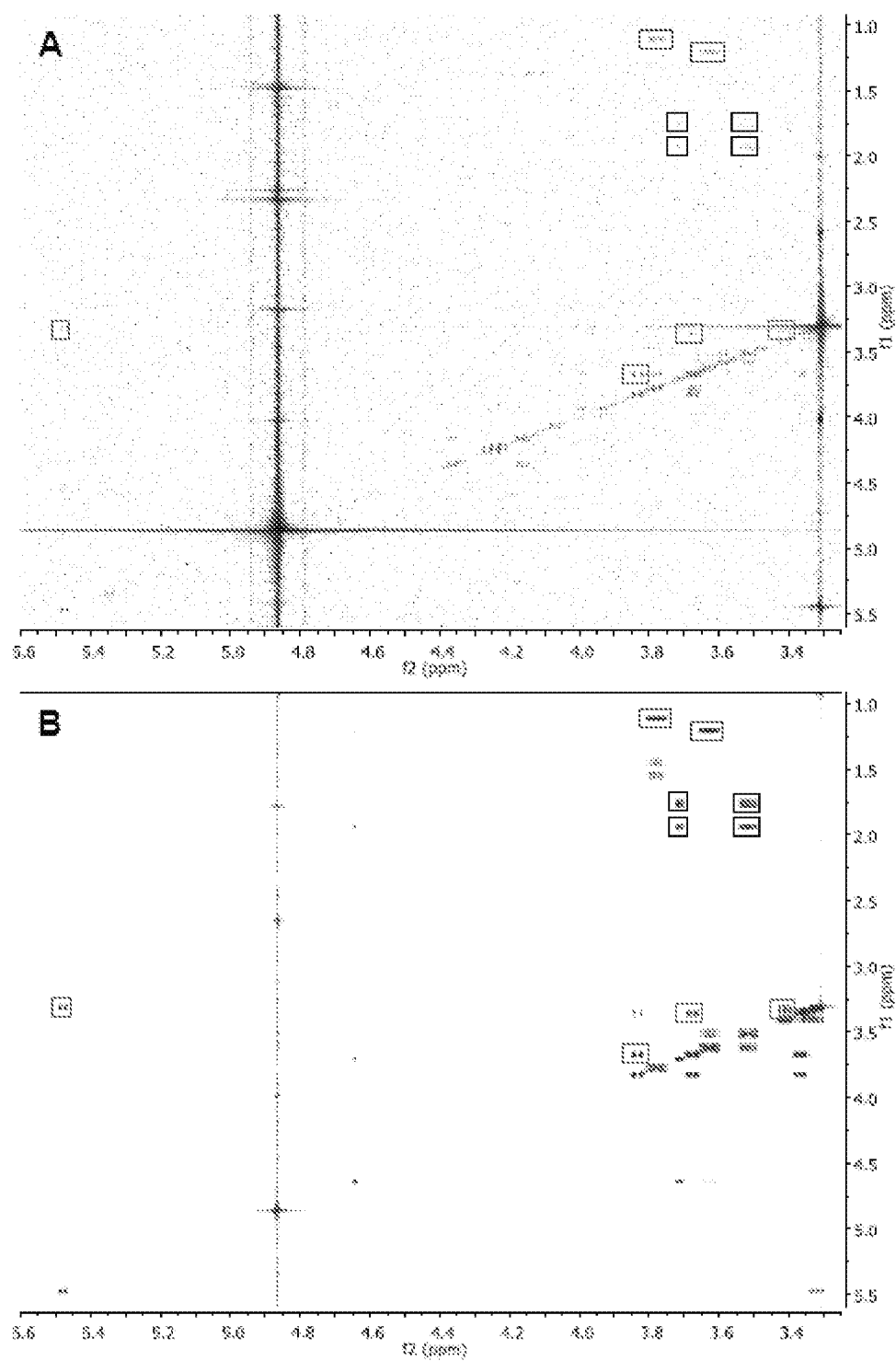
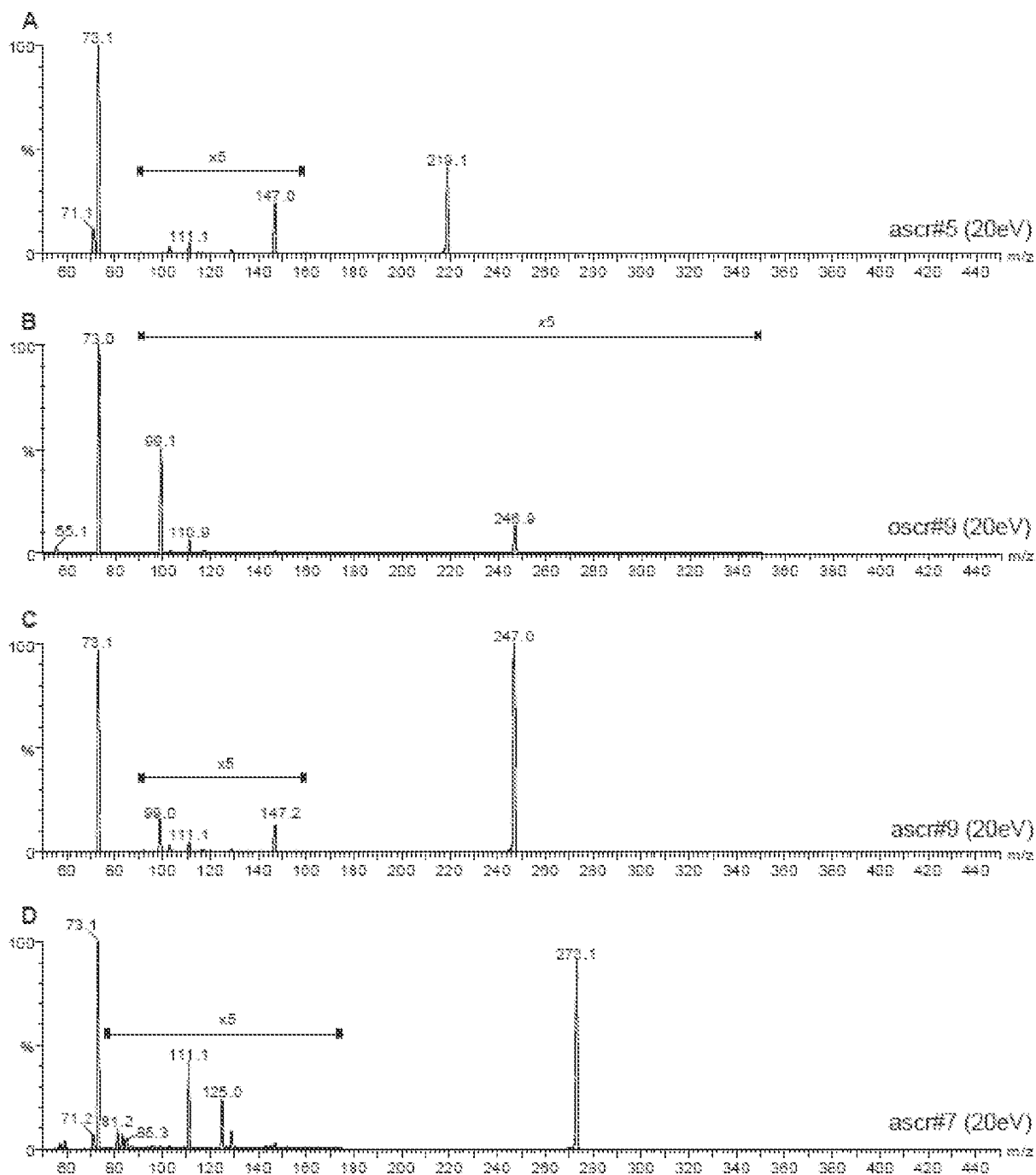


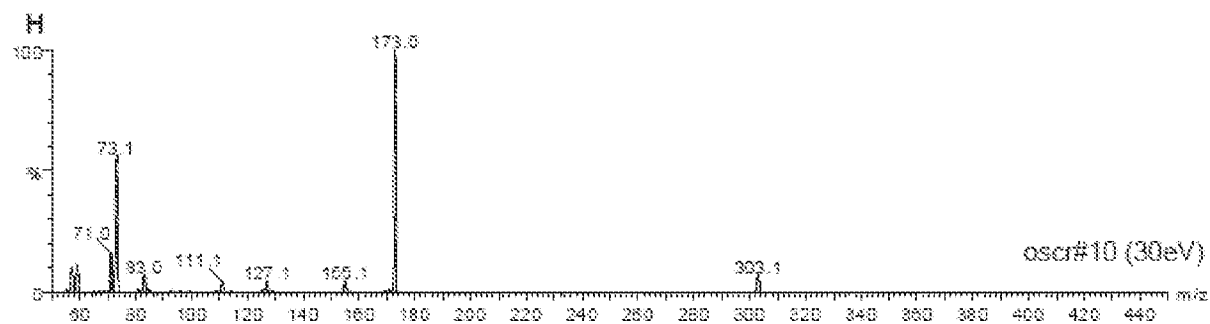
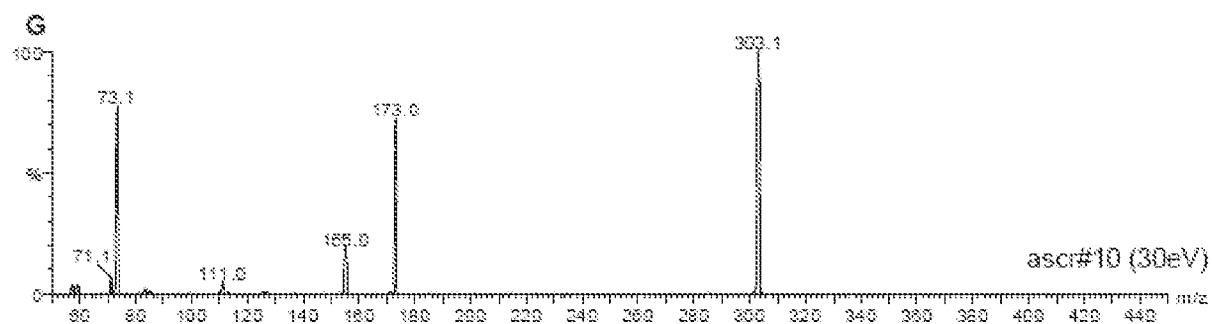
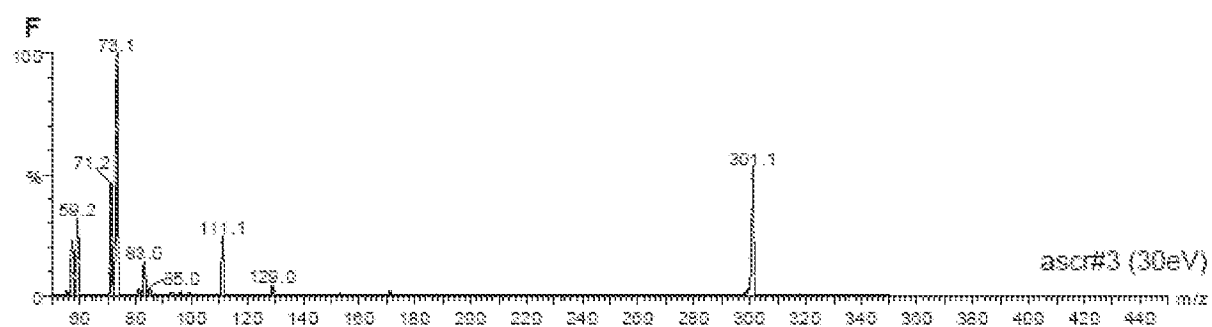
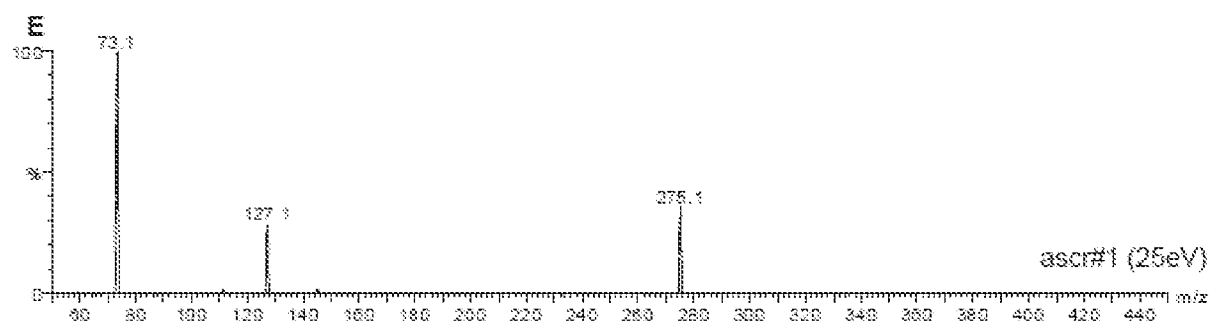
Figure 42



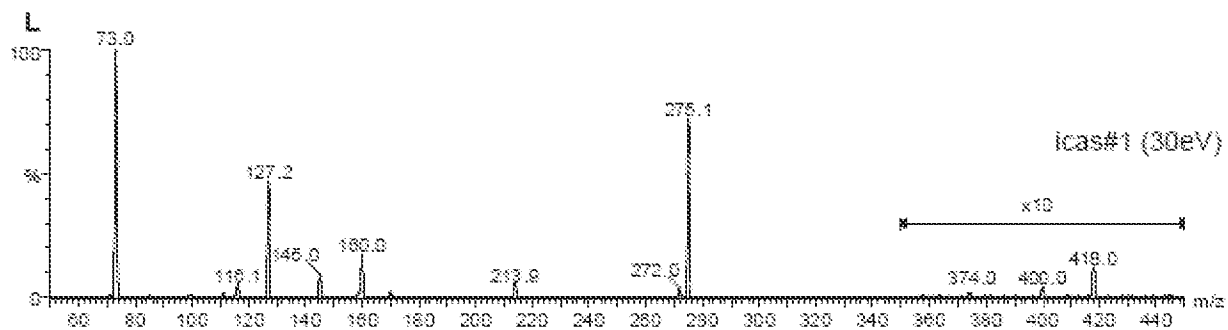
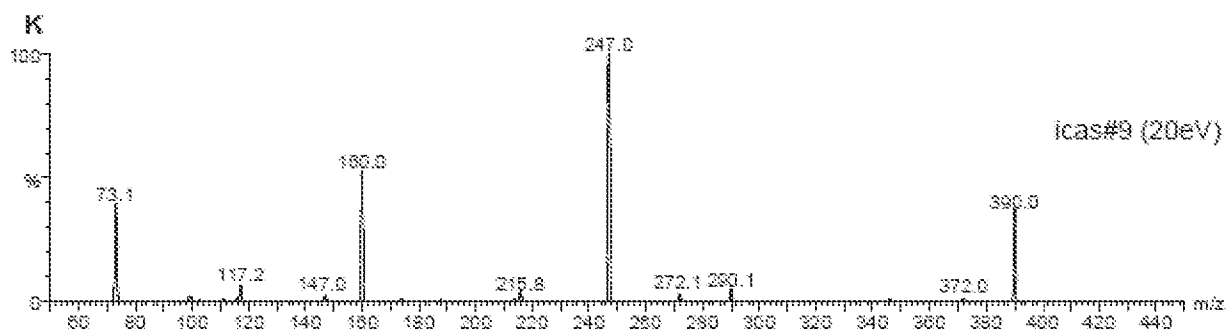
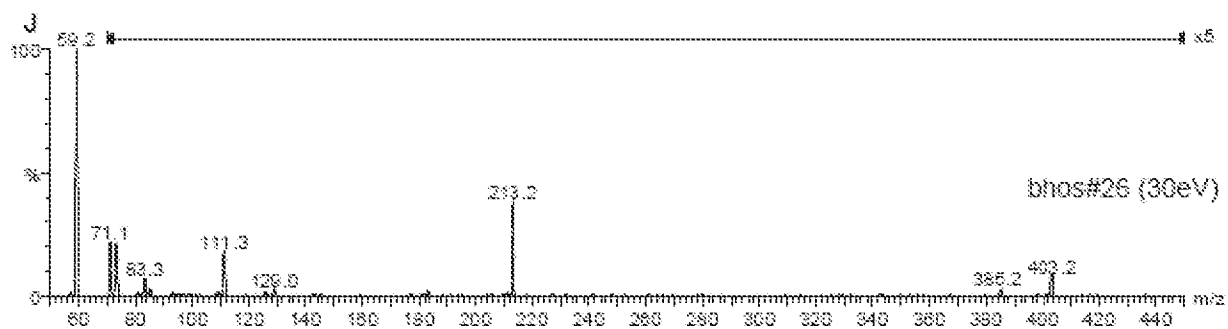
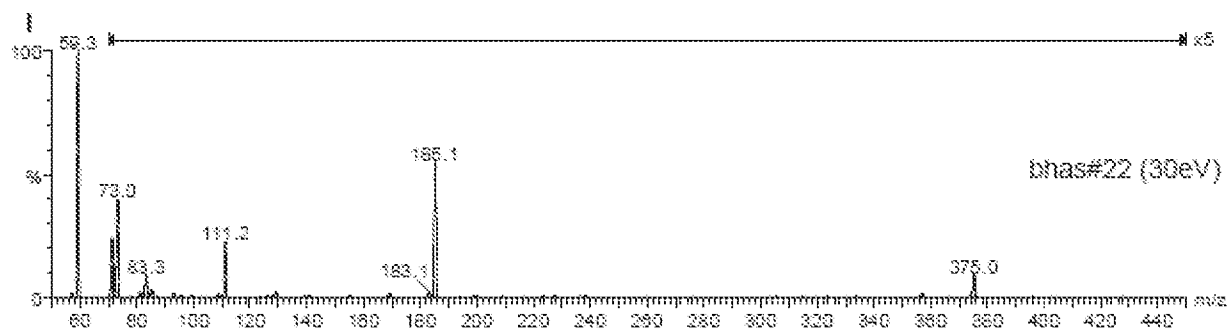
Figures 43A-B



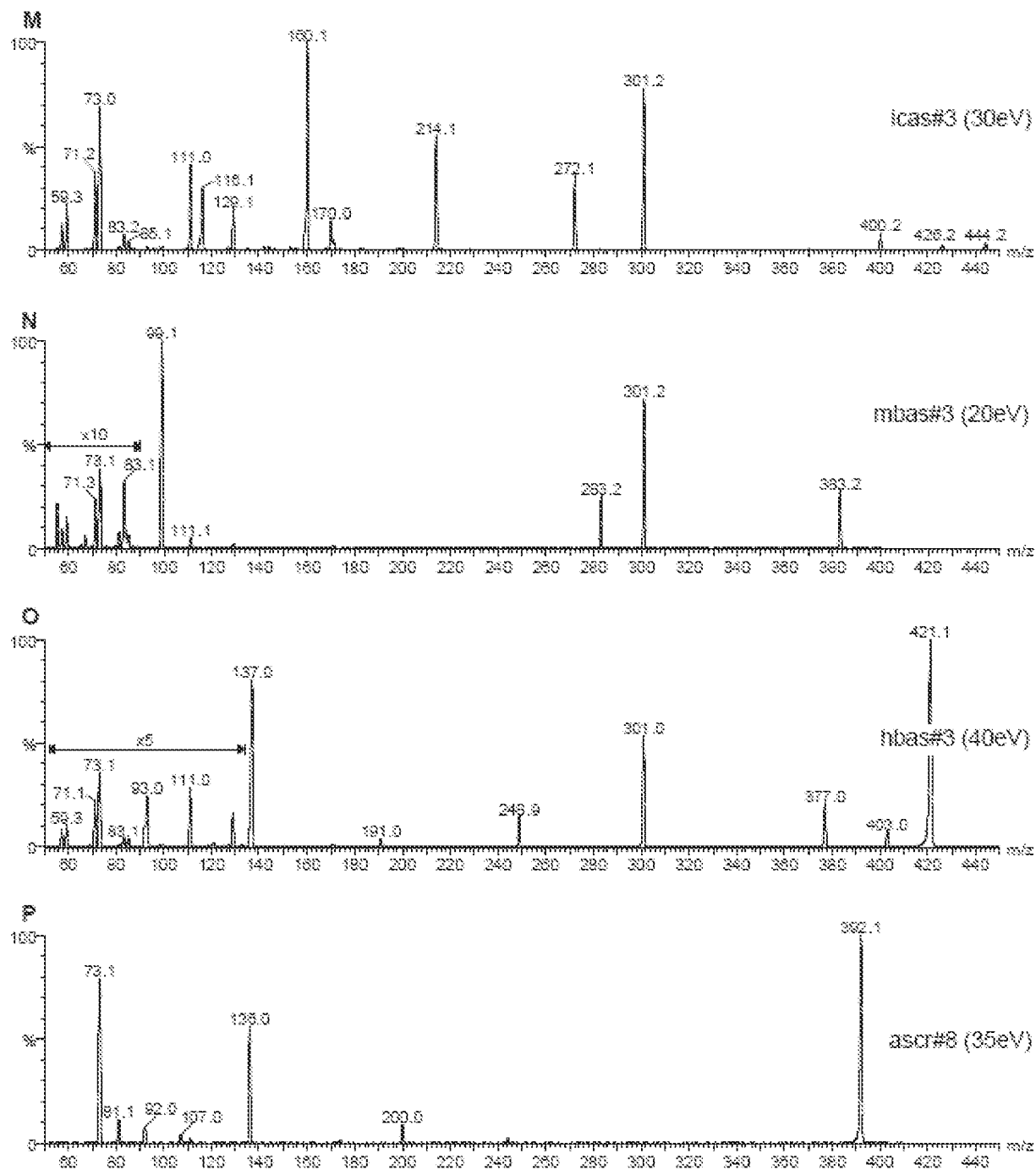
Figures 44A–D



Figures 44E--H



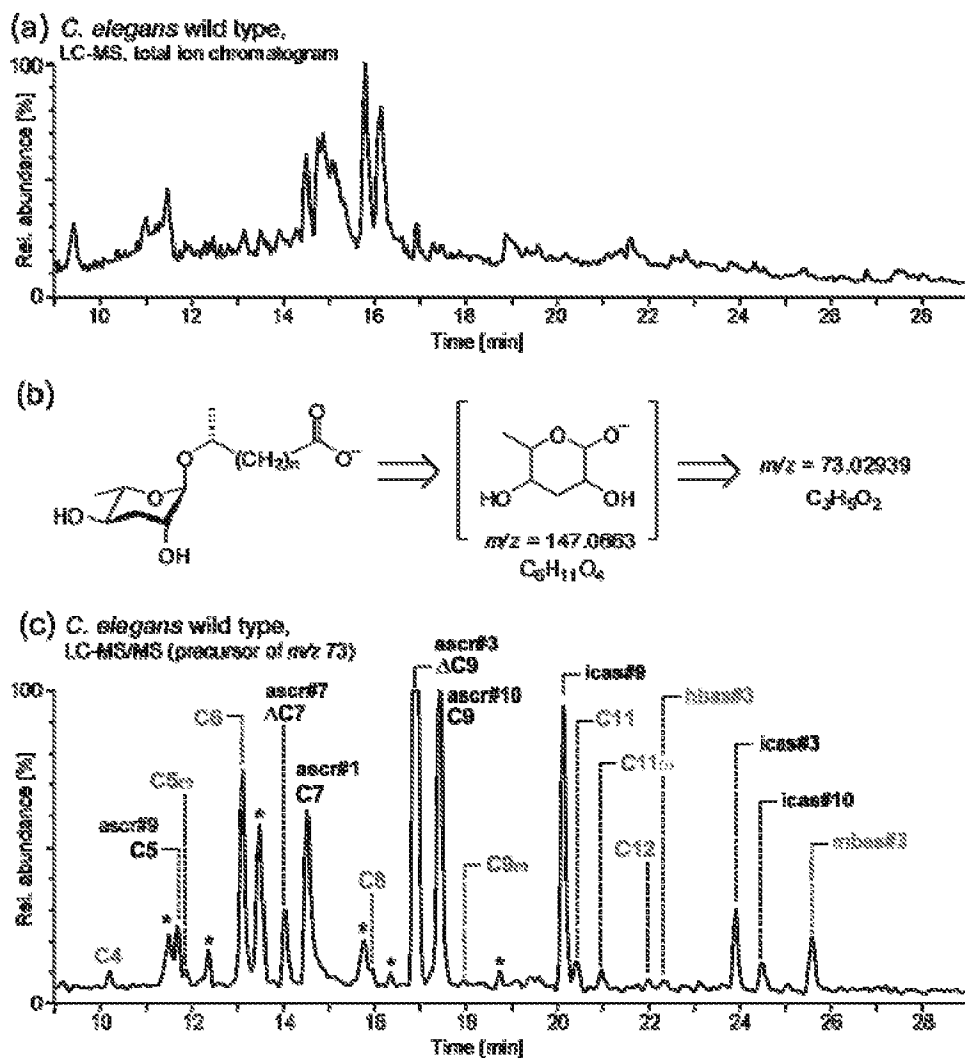
Figures 44I-L



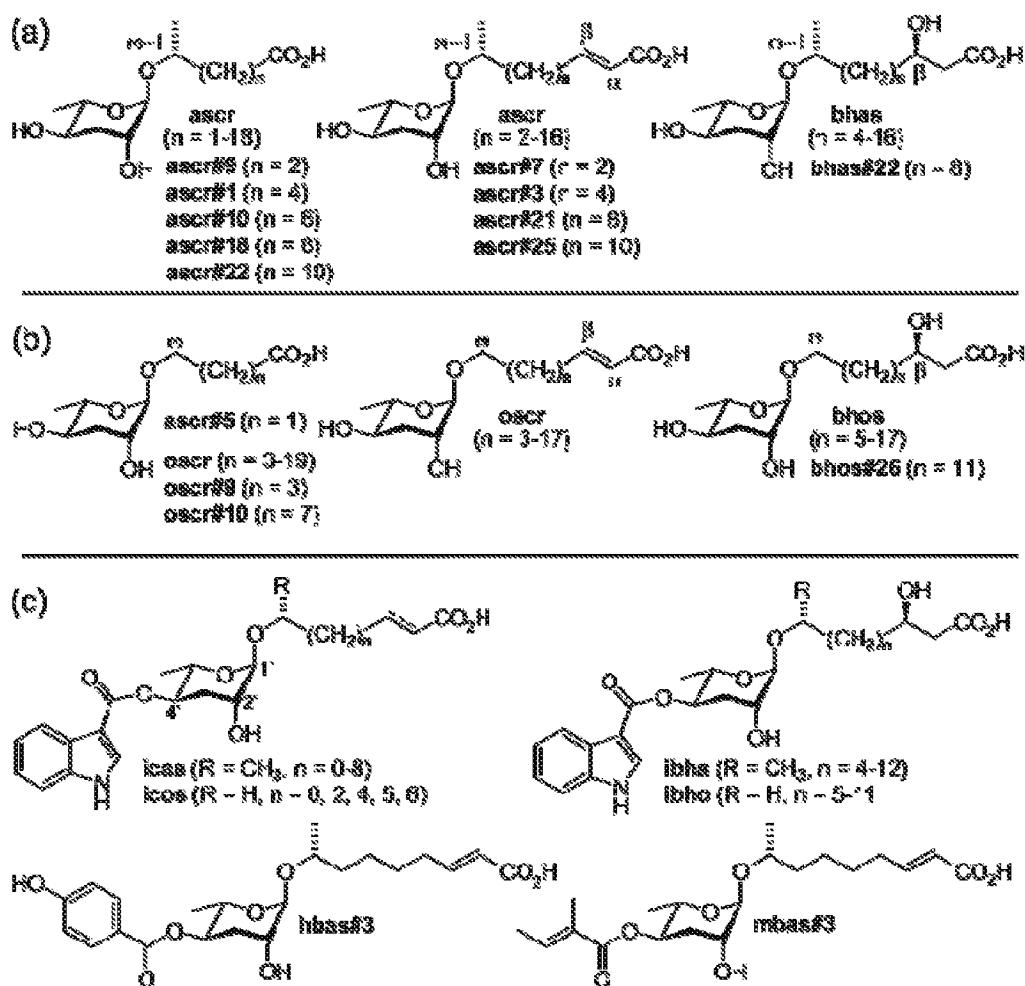
Figures 44M-P

A DHS-28 C.elegans	-----	76
B NFE-2 (H.sapiens)	MSSELESTGNTLTATACGLSPATATLRISALVNTGSGSTTPKNGLLASGWEELSPFSAATGNTS	76
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	146
B NFE-2 (H.sapiens)	SESEKNTKILAEKSHINVTNNAALSLGATAPISDEINQIEHVELPGEFLTPRAMEKNGQVSLIDTS	146
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	214
B NFE-2 (H.sapiens)	SLNIVTETICQVSAKLCGLGLANLATEGPGSNFQNLADPGKSTTQREILATLQEPVQNLAL	214
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	284
B NFE-2 (H.sapiens)	SEKCTGSLERMAKSLQLEKILASVPGQNHSMPEAWAATKNSDDEASNGQKCEKCCILIEVDS	284
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	354
B NFE-2 (H.sapiens)	NDELEGTQNHESPTITETGSGGQMLPPEMAITELTETALGQVSLINIDITLTPKSSVSG	354
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	424
B NFE-2 (H.sapiens)	TPGILIQSMNGGARIQSEKPAFVLAQSLRLVTPRAGSLTETALQKAGSSVSLDNLKES	424
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	494
B NFE-2 (H.sapiens)	KELIKHQKELVSGPPEELIKWVAVAGSPKALTEKSLQALAKL--K--KELIKHQKELV	494
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	564
B NFE-2 (H.sapiens)	APQFQILQKQIFGKARDLQPPKLVKVFPEKIALPKKSTPAVLDKESQKELKSLVQKELV	564
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	634
B NFE-2 (H.sapiens)	SKAVLLAPPTSPKLTGGGKQLEVRIILGPPQDICEVQVQVAFKHKHSTIGAKTILKPG	634
C MAOC-1 C.elegans	-----	
A DHS-28 C.elegans	-----	704
B NFE-2 (H.sapiens)	STKQKAT--PQTIILSGEKEVLELQPAIFGSLAPQNTLSPKLGKIDVAKI--	704
C MAOC-1 C.elegans	-----	

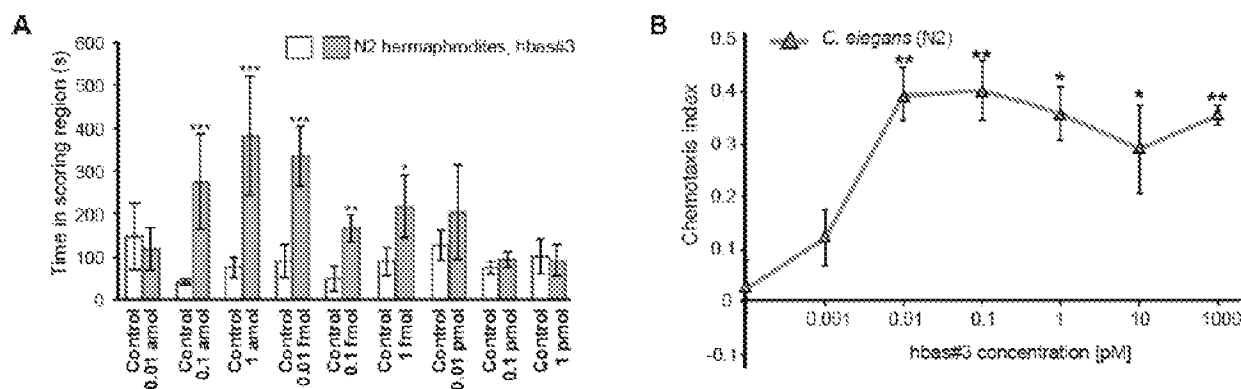
Figure 45



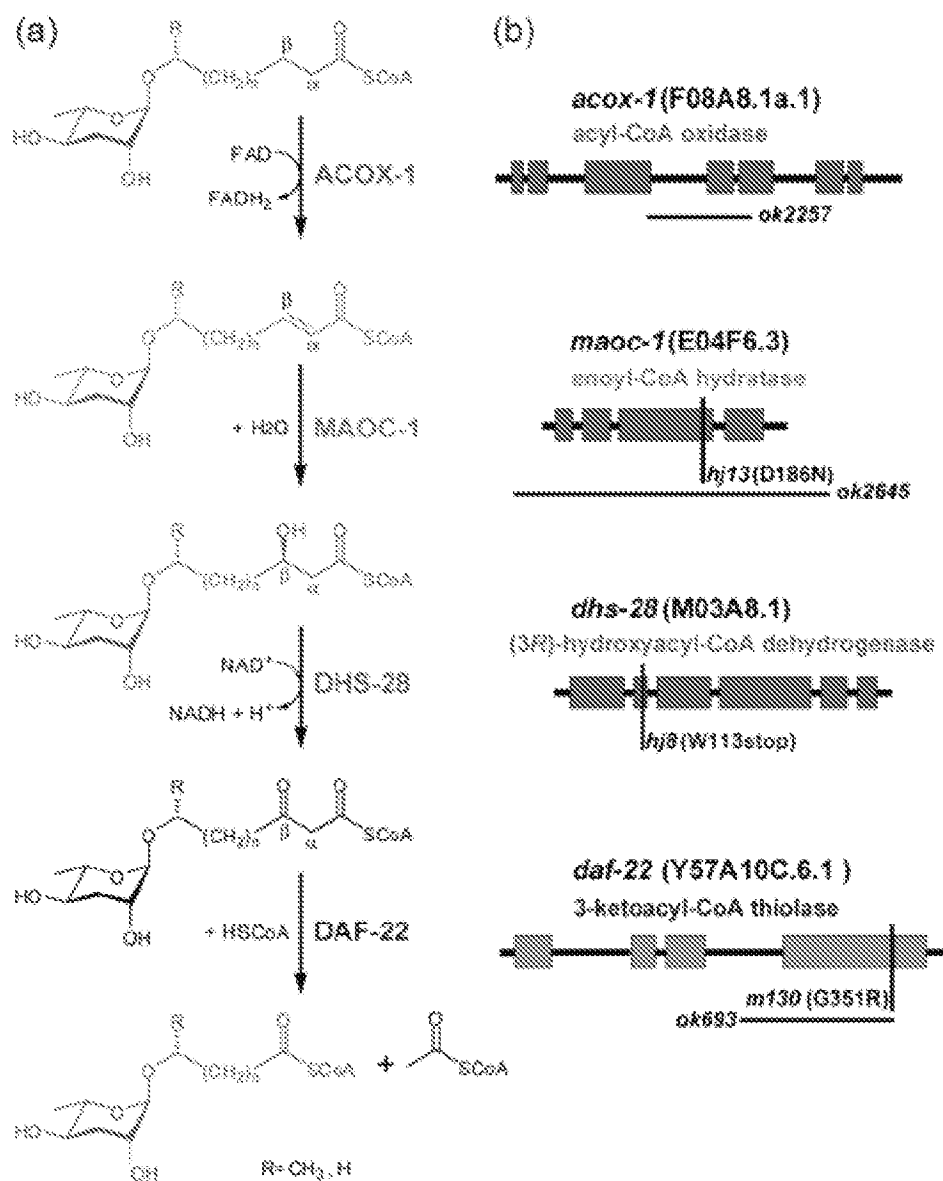
Figures 46A–C



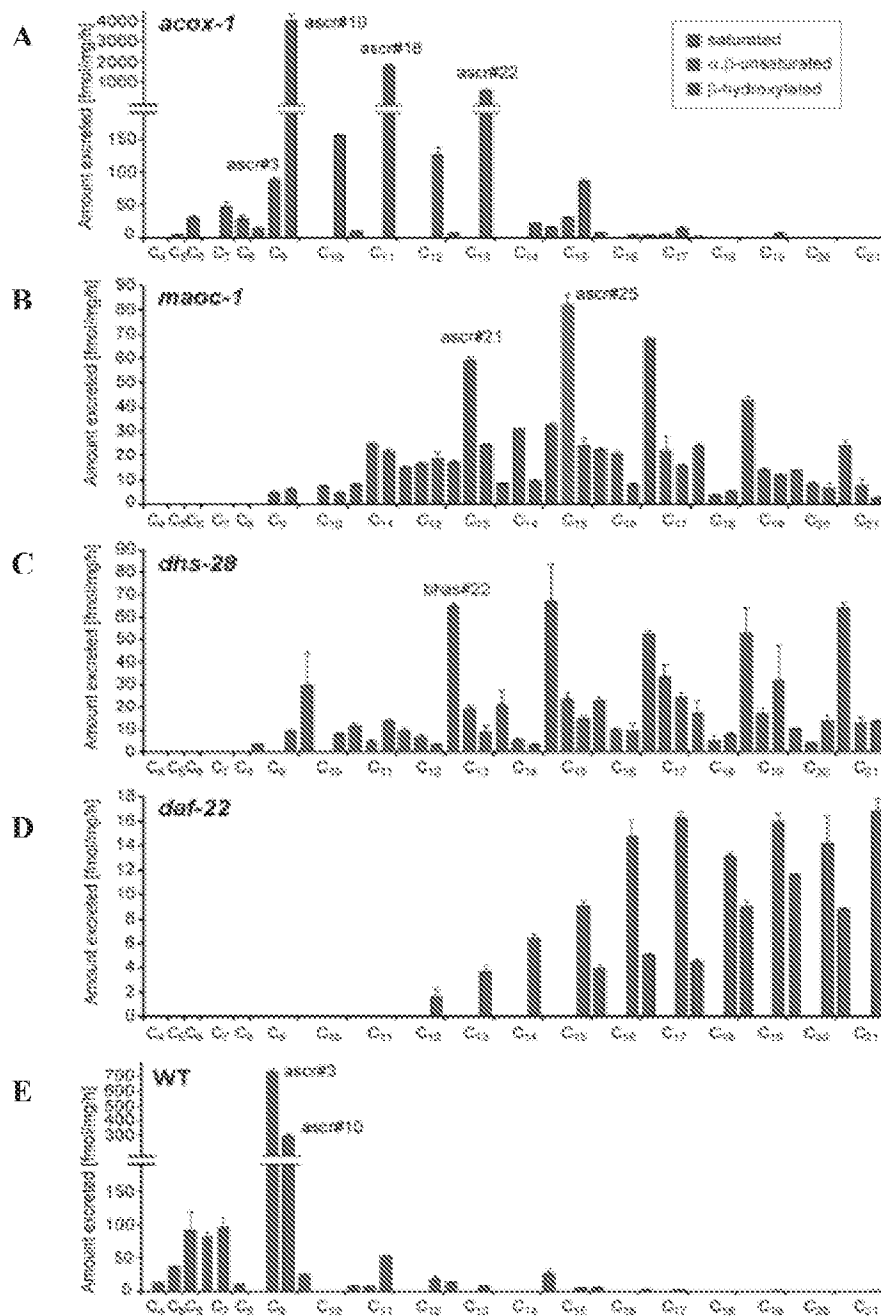
Figures 47A-C



Figures 48A-B



Figures 49A–B



Figures 50A–E

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ACOX-1      MVHLNPTTCEP...LE...HARD...CEMPAP...ETATAP...AEIPE... 60
F08A9.3     ---MSR...C...S...E...K...RERIAS...P...TES...RIHE... 66
F89F4.1     ---MSRW...C...V...E...E...E...E...E...E...E... 67
C48B4.1     -MFLNWL...C...C...E...E...E...E...E...E...E... 68

ACOX-1      M...E...E...E...E...E...E...E...E...E...E...E... 119
F08A9.3     M...E...E...E...E...E...E...E...E...E...E...E... 118
F89F4.1     V...E...E...E...E...E...E...E...E...E...E...E... 116
C48B4.1     I...E...E...E...E...E...E...E...E...E...E...E... 118

ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 179
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 176
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 176
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 176

ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 239
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 236
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 236
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 236

ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 299
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 296
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 296
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 296

ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 359
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 356
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 356
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 356

ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 419
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 416
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 416
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 416

ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 479
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 476
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 476
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 476

ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 539
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 536
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 536
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 536

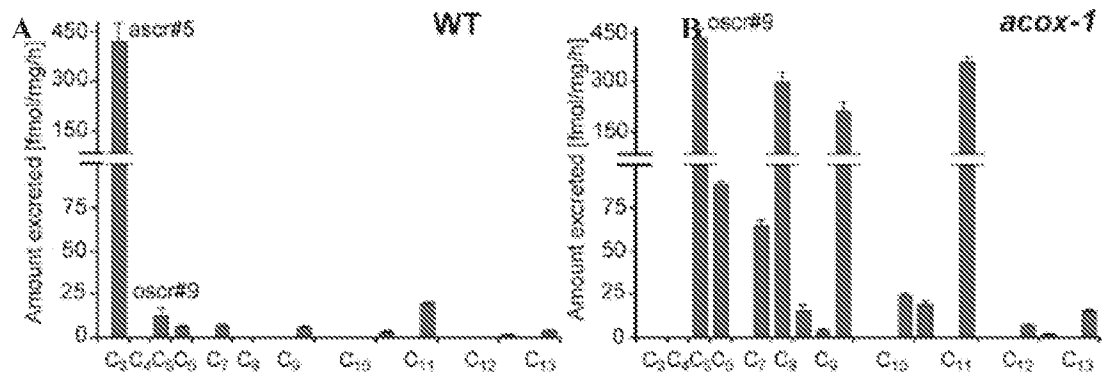
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F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 596
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 596
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 596

ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 659
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 656
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 656
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 656

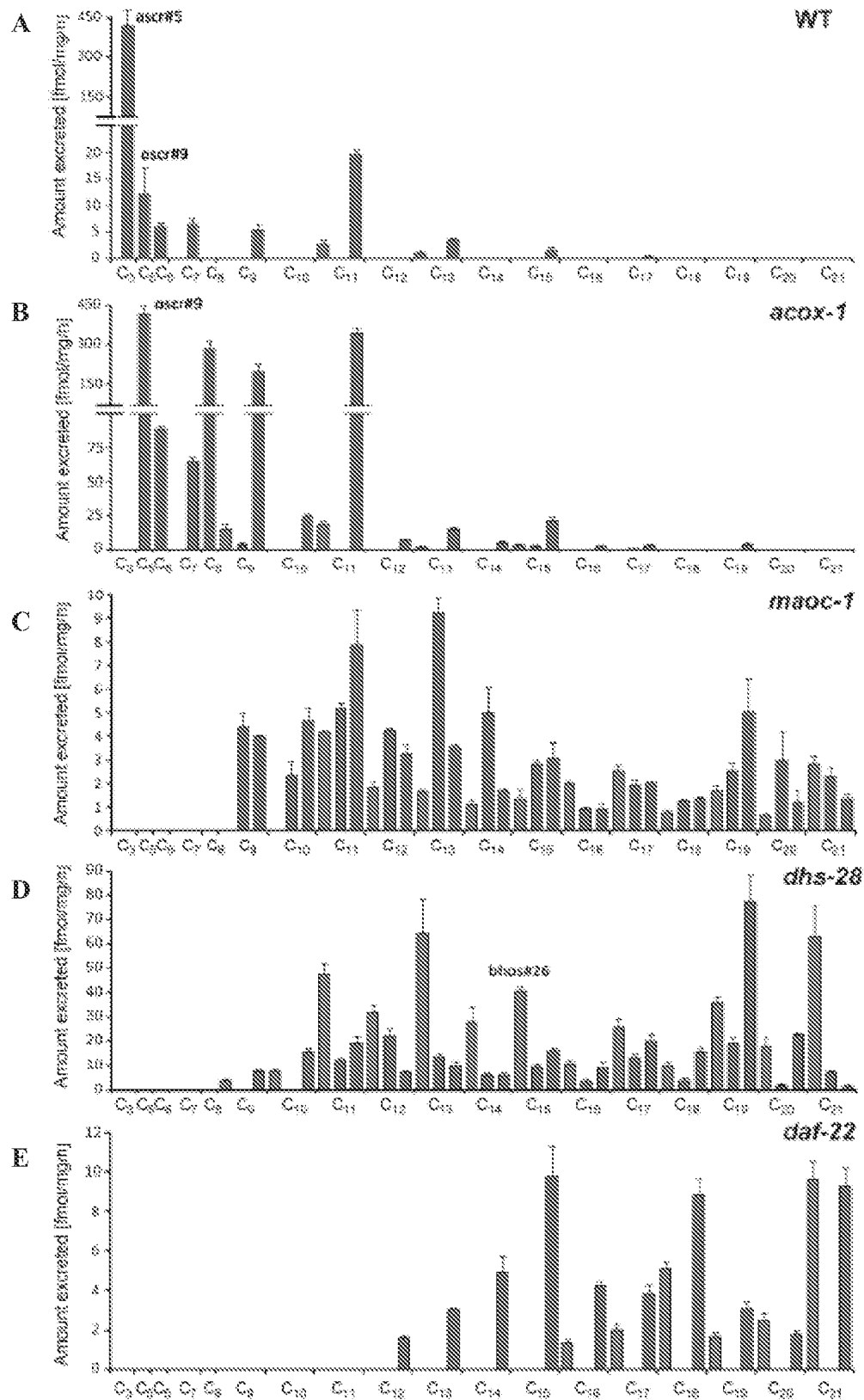
ACOX-1      E...E...E...E...E...E...E...E...E...E...E...E... 674
F08A9.3     E...E...E...E...E...E...E...E...E...E...E...E... 660
F89F4.1     E...E...E...E...E...E...E...E...E...E...E...E... 667
C48B4.1     E...E...E...E...E...E...E...E...E...E...E...E... 669

```

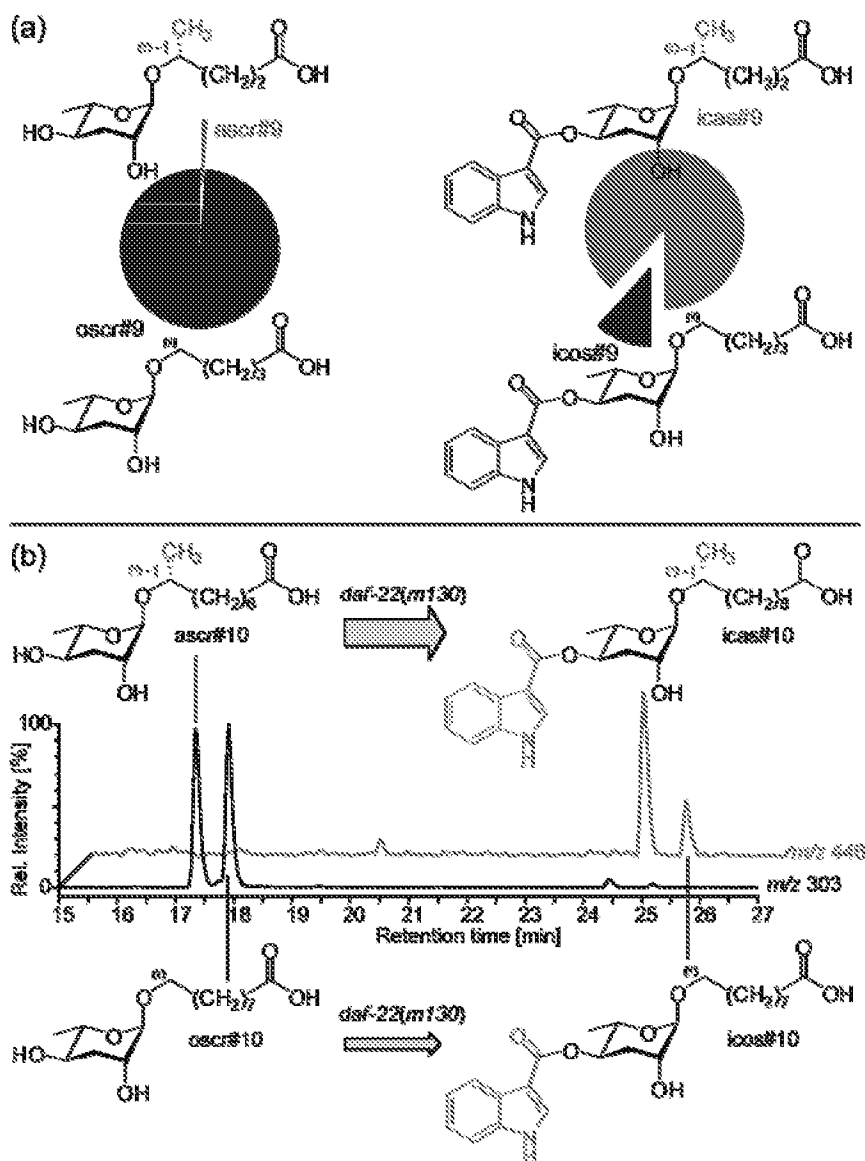
Figure 51



Figures 52A-B



Figures 53A-E



Figures 54A–B

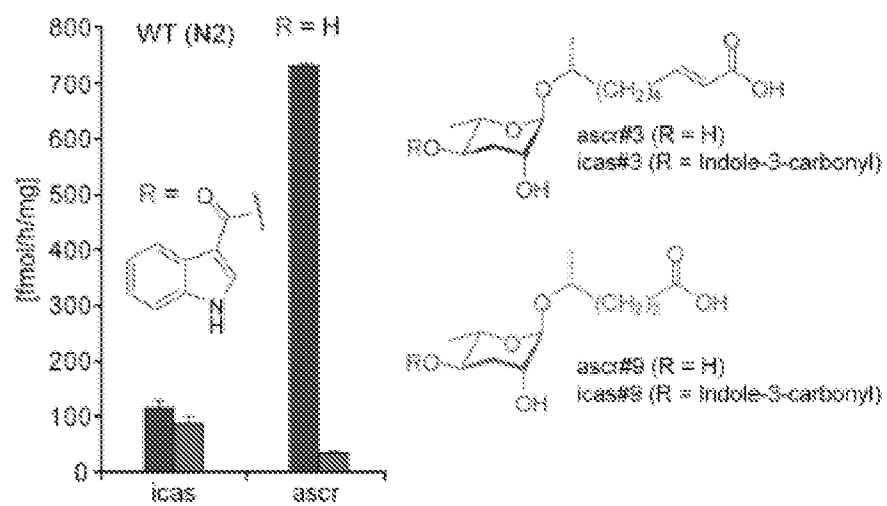
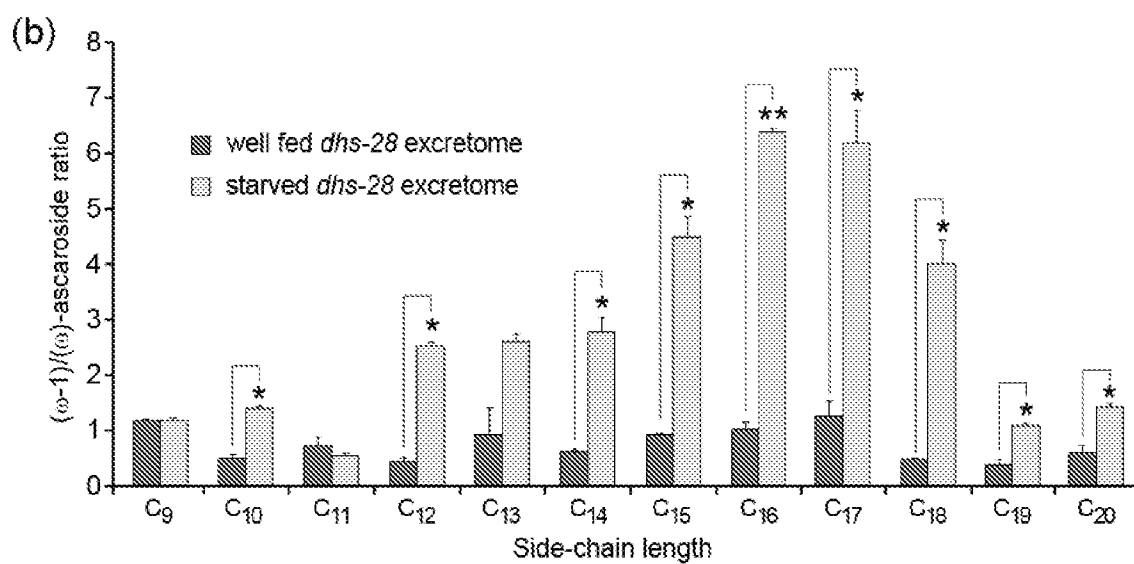
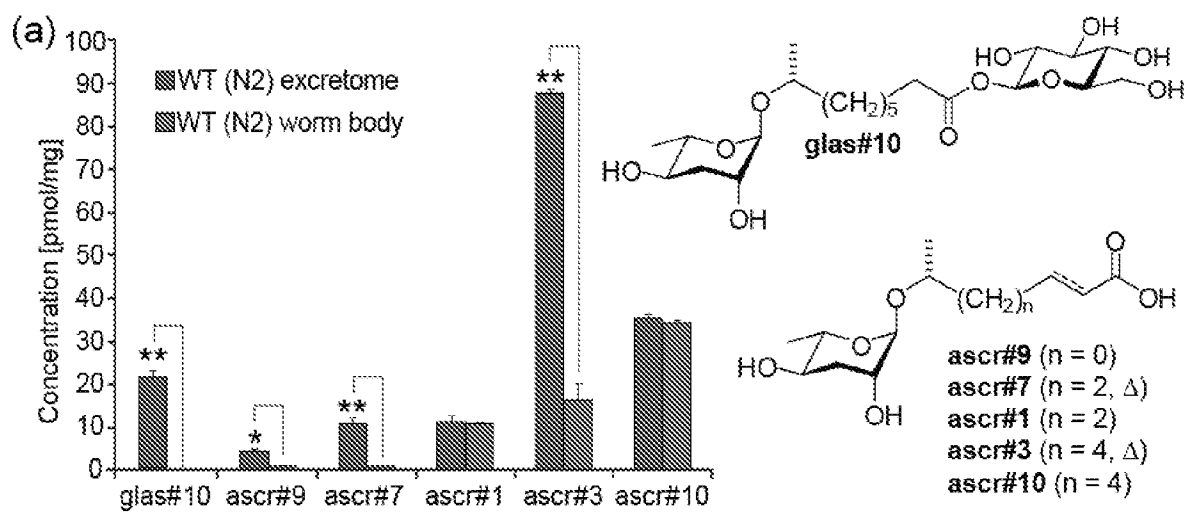
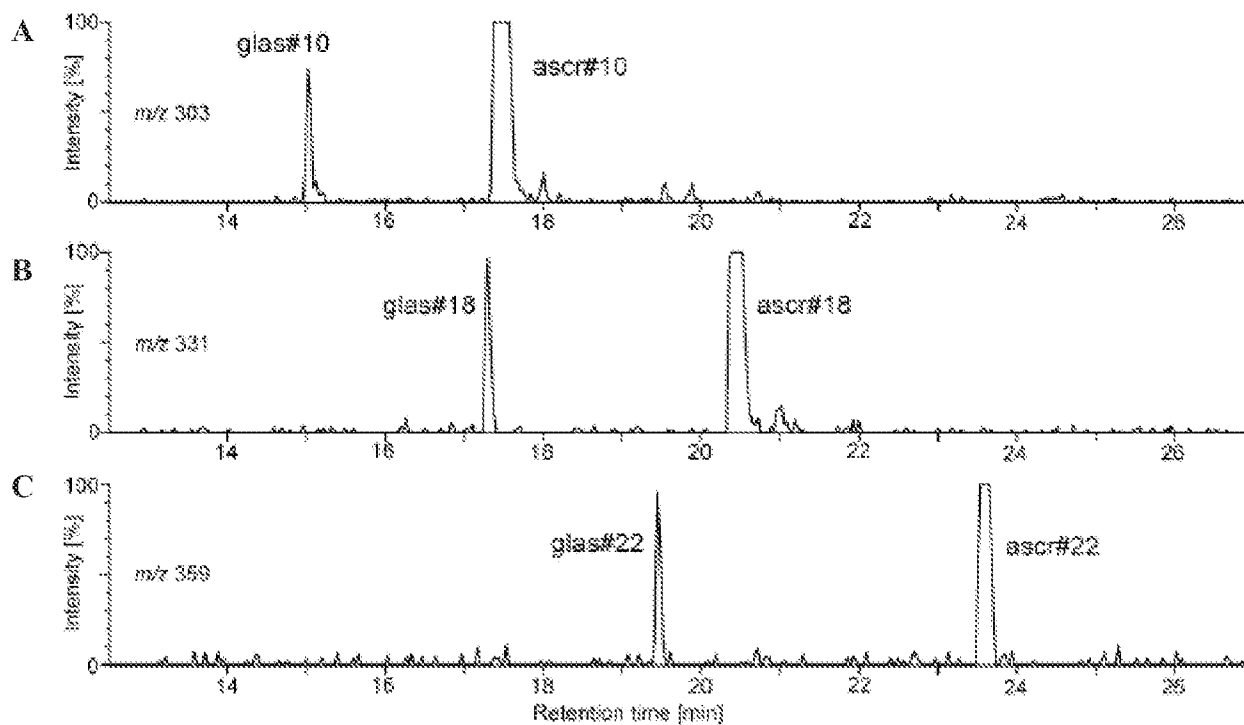


Figure 55



Figures 56A–B



Figures 57A-C

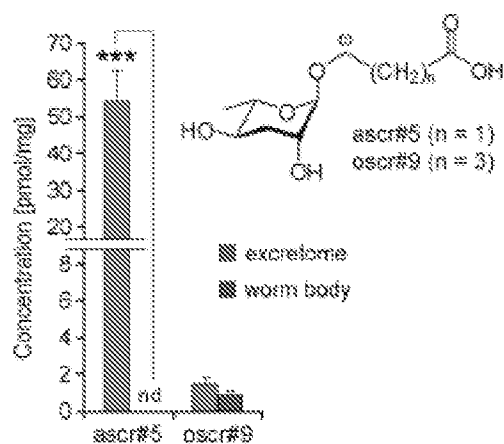
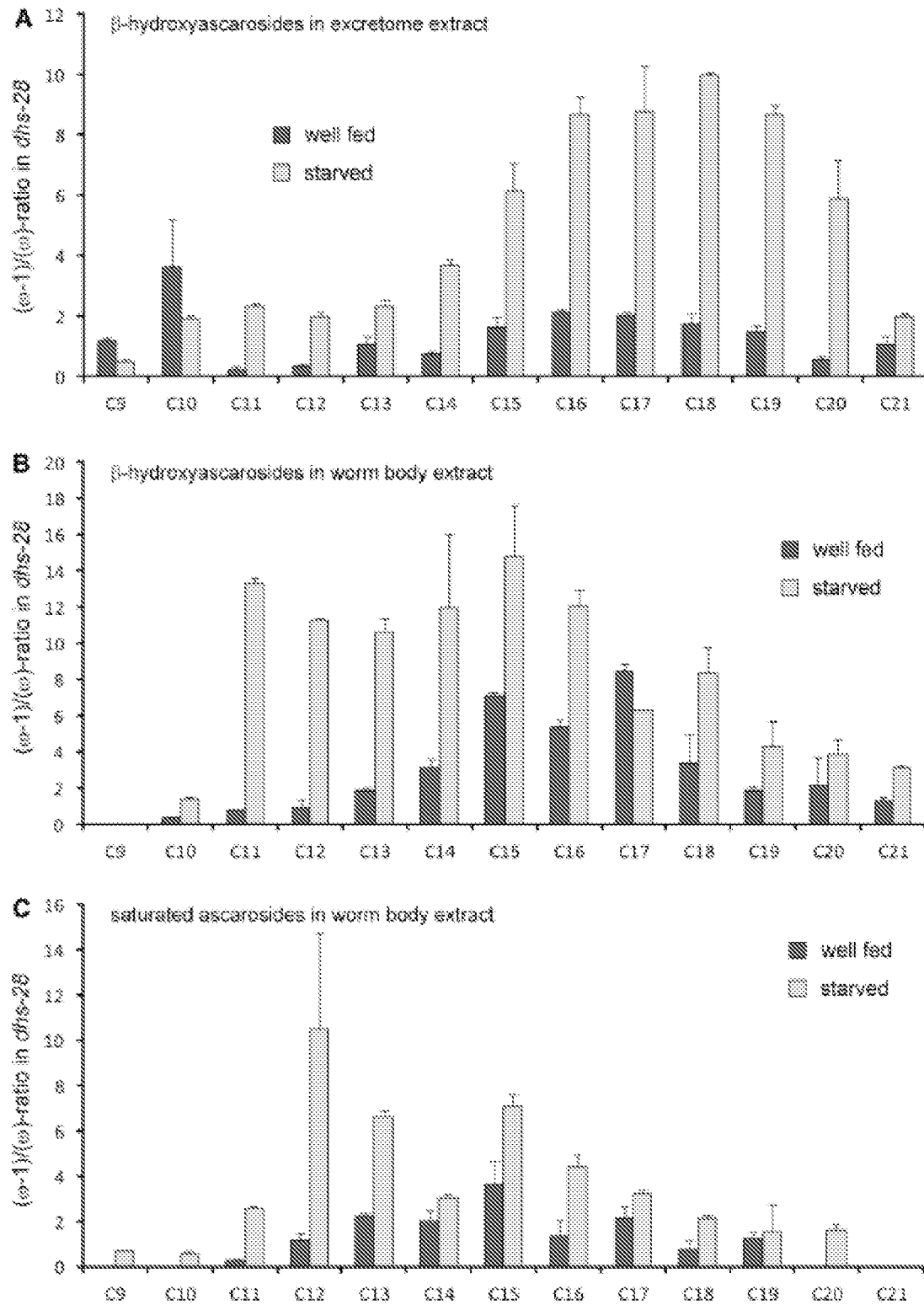


Figure 58



Figures 59A-C

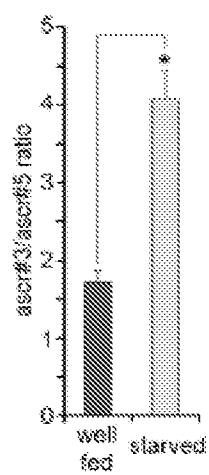
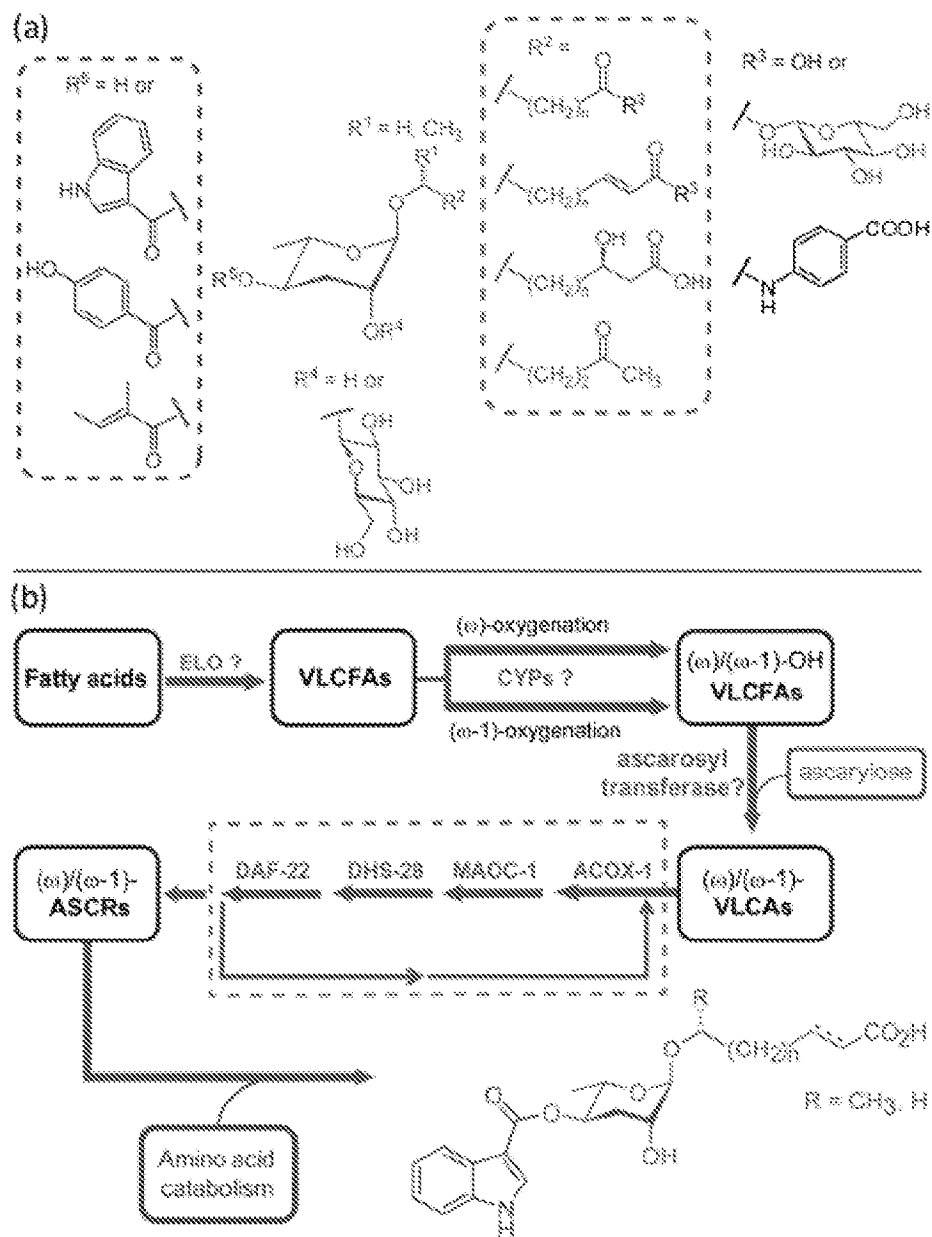


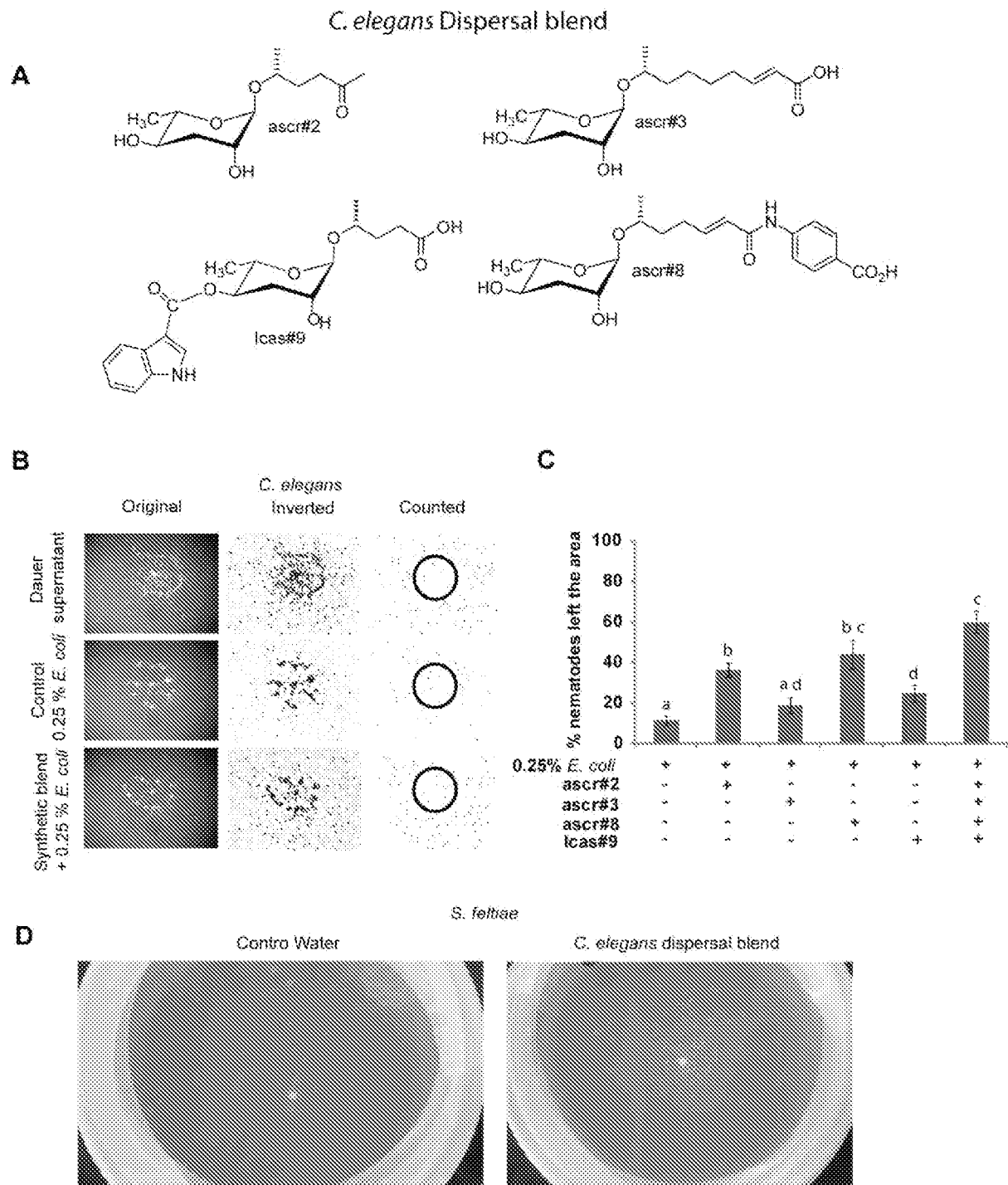
Figure 60



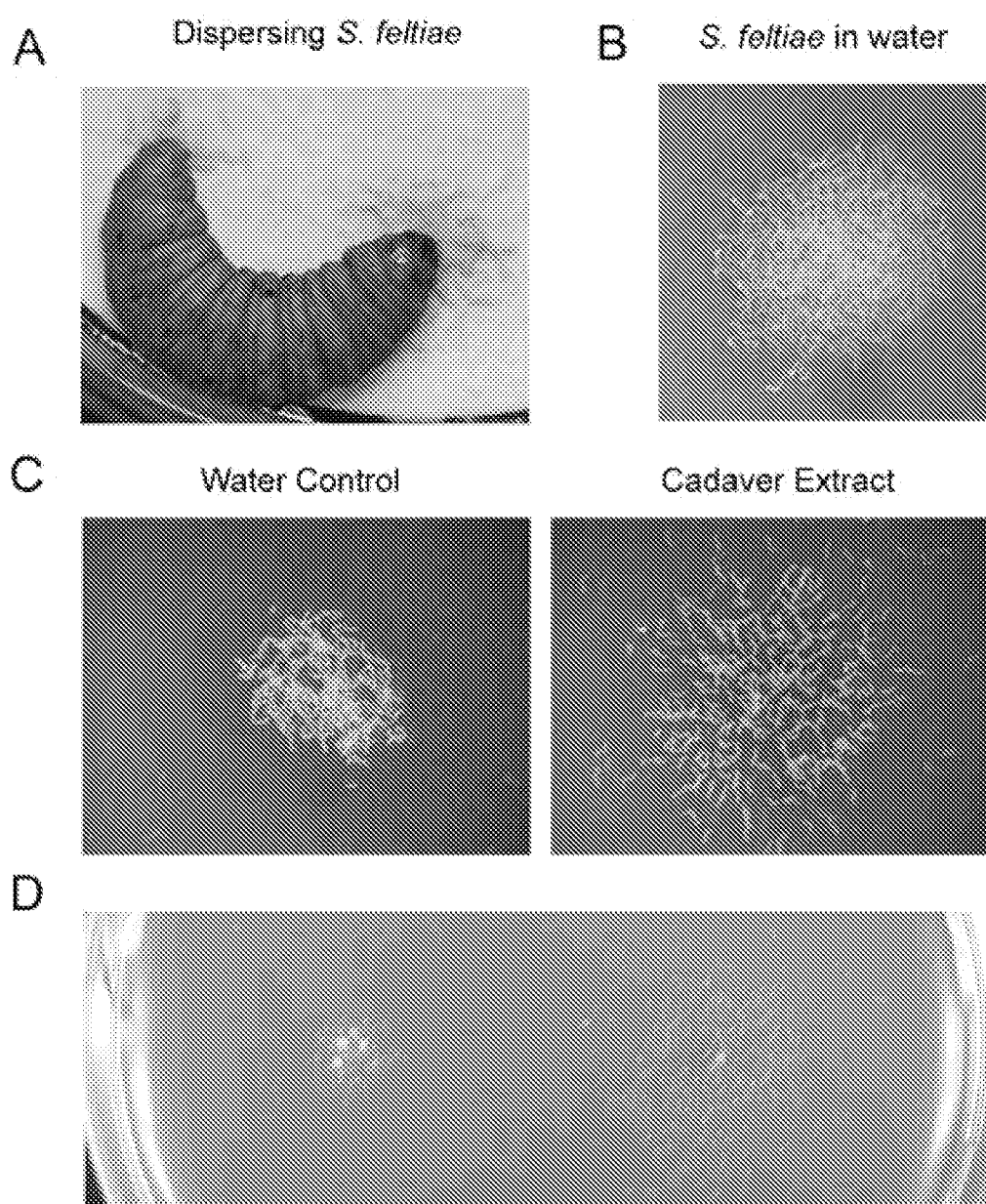
Figures 61A–B

[illegible]

Figure 62



Figures 63A–D



Figures 64A–D

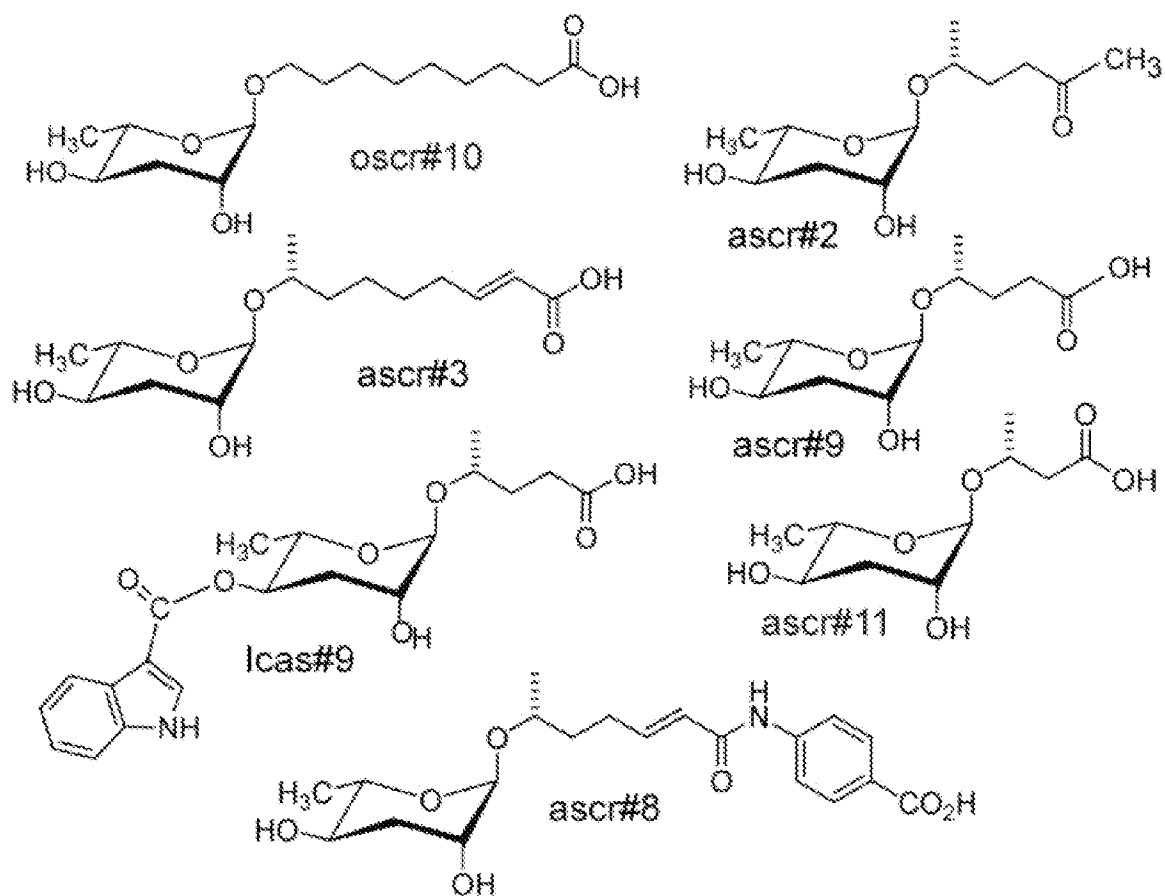
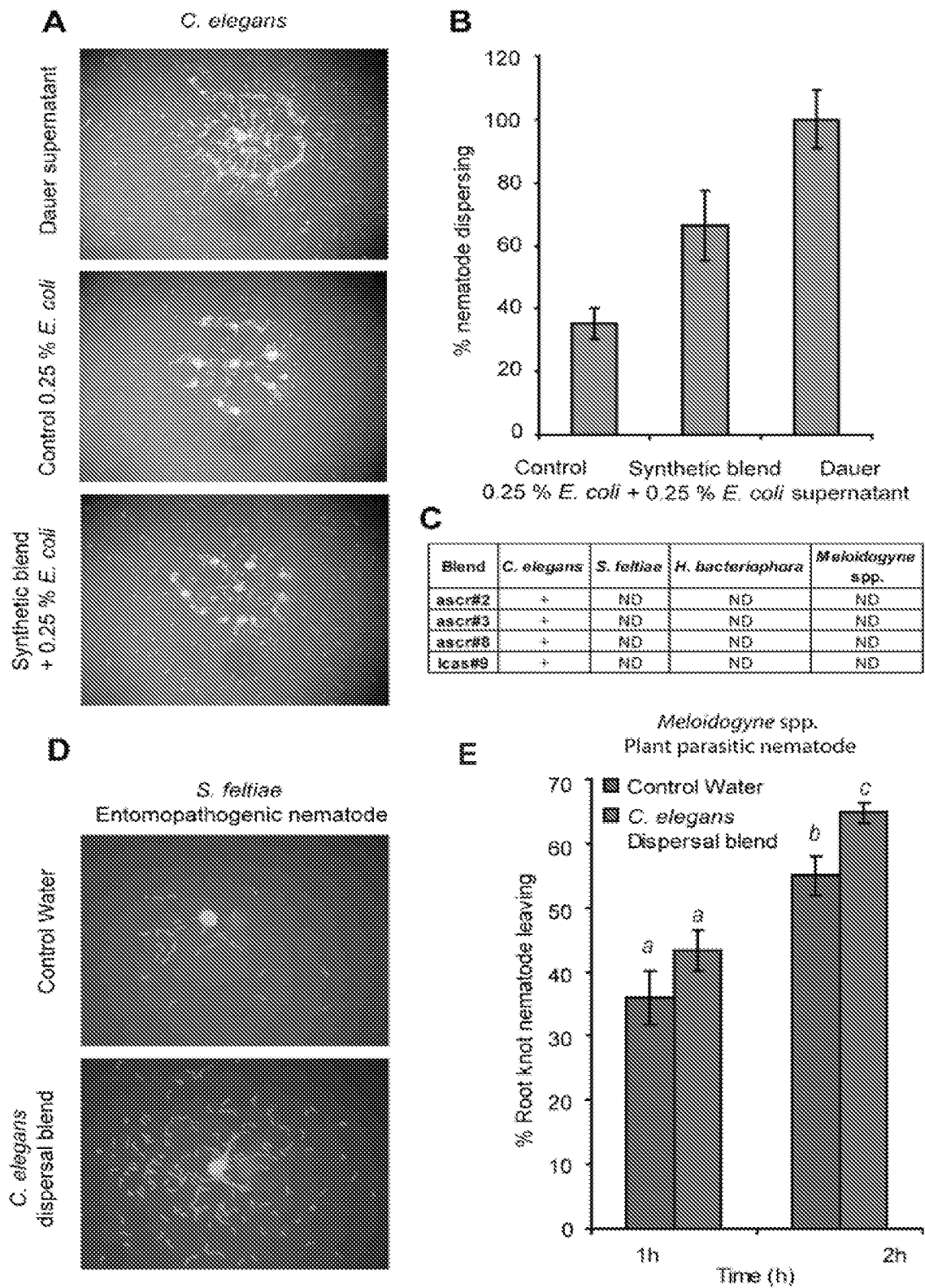
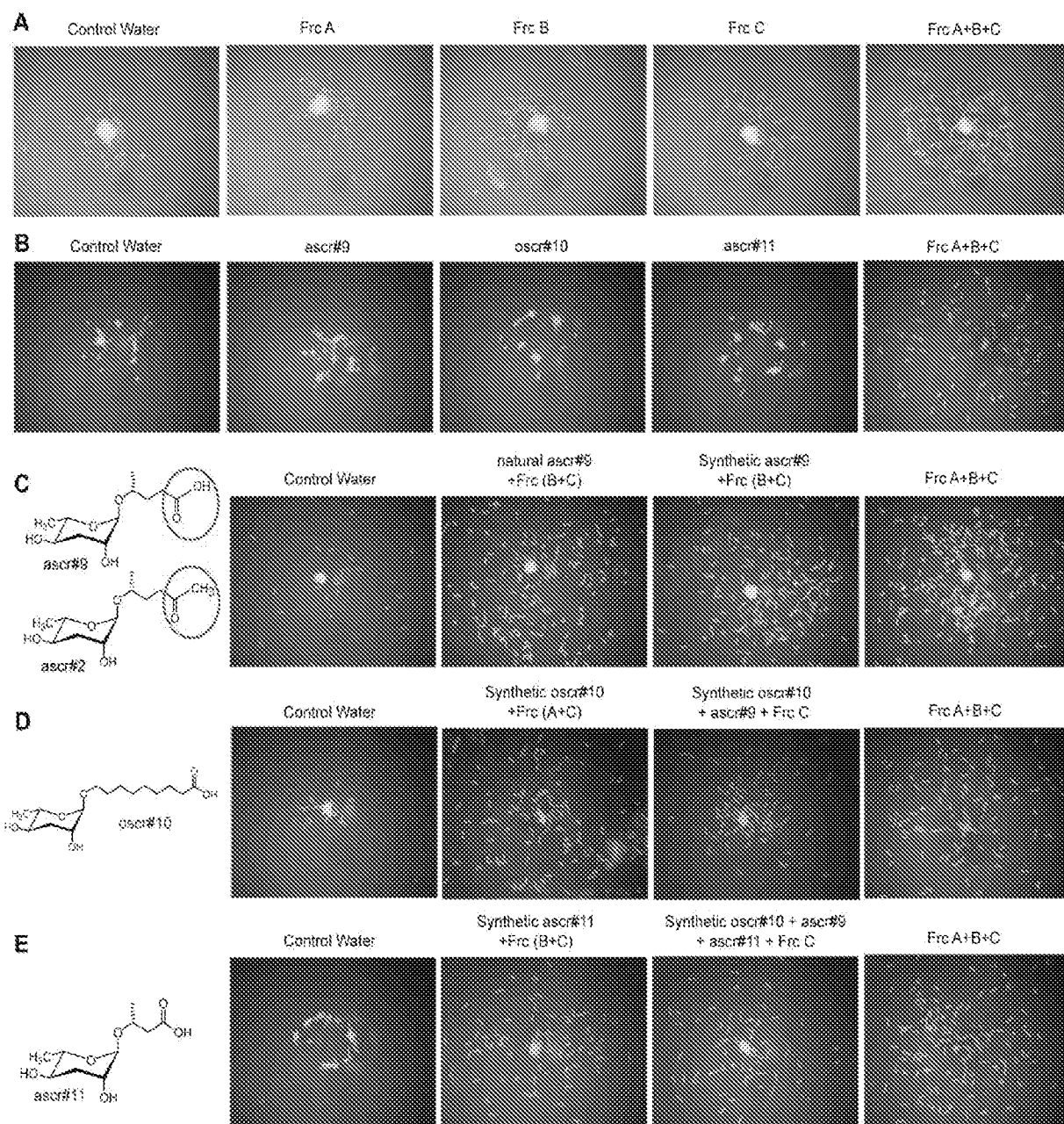


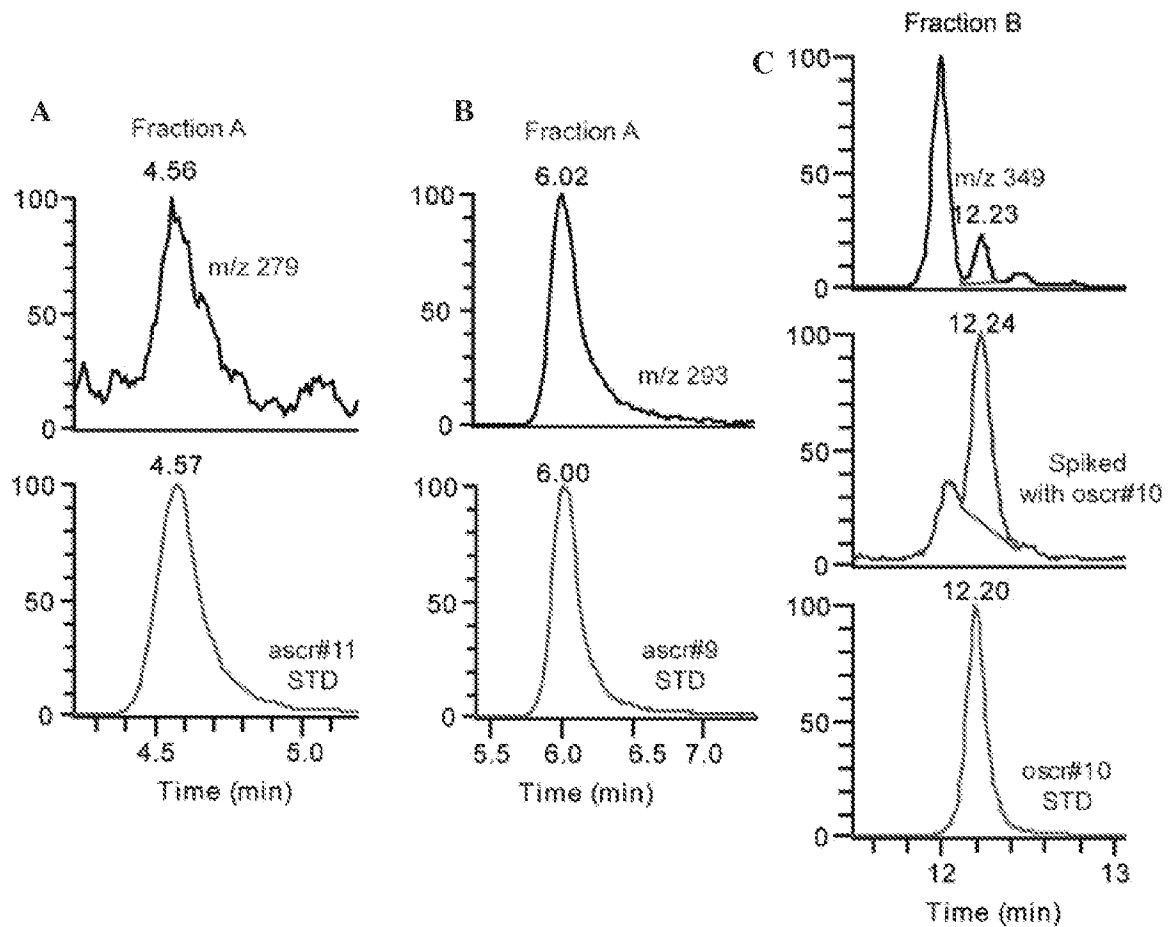
Figure 65



Figures 66A-E



Figures 67A–E



Figures 68A-C

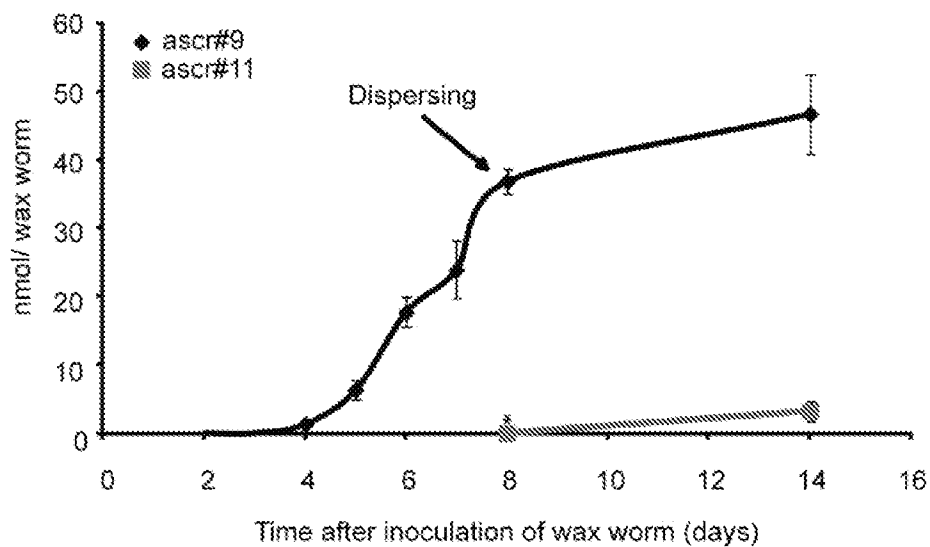


Figure 69

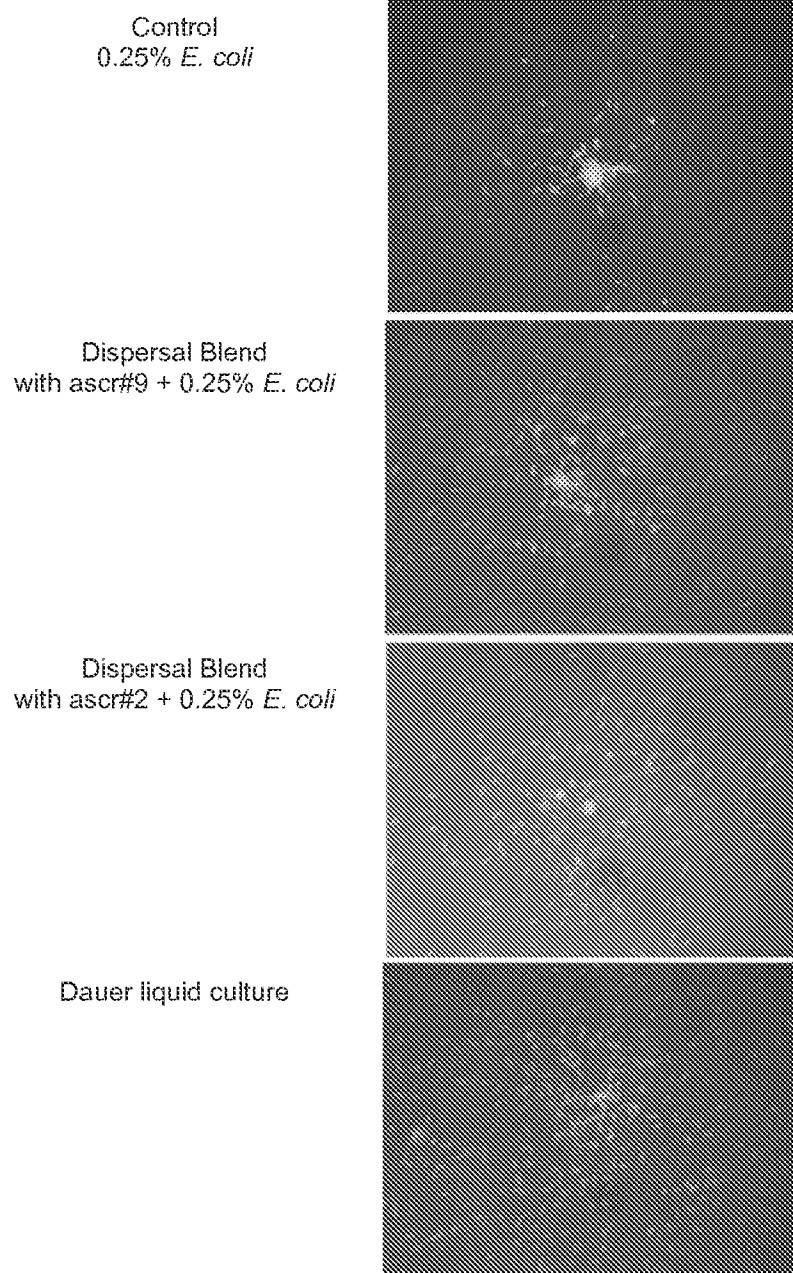
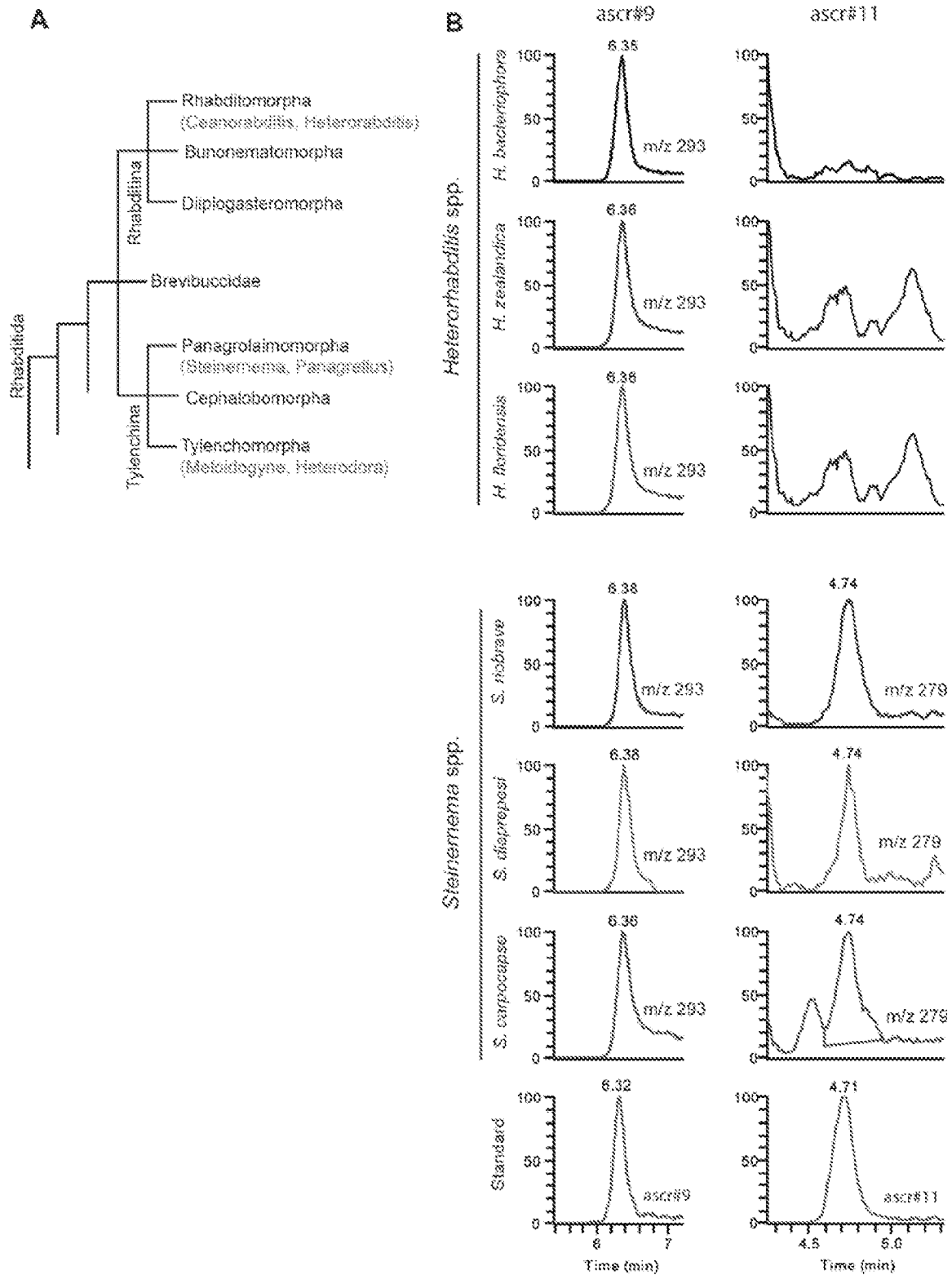
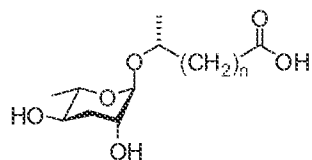


Figure 70

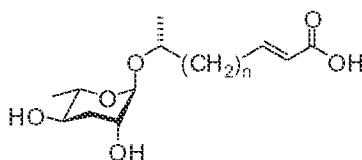


Figures 71A–B



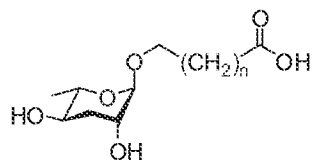
Side chain (n)	SMID ¹	Molecular formula	Molecular mass g/mol	m/z M-H ⁺
1	ascr#1 1	C ₁₀ H ₁₈ O ₆	234.11034	233.1 0
2	ascr#9	C ₁₁ H ₂₀ O ₆	248.12599	247.1 2
3	ascr#1 2	C ₁₂ H ₂₂ O ₆	262.14164	261.1 3
4	ascr#1	C ₁₃ H ₂₄ O ₆	276.15729	275.1 5
5	ascr#1 4	C ₁₄ H ₂₆ O ₆	290.17294	289.1 7
6	ascr#1 0	C ₁₅ H ₂₈ O ₆	304.18859	303.1 8
7	ascr#1 6	C ₁₆ H ₃₀ O ₆	318.20424	317.2 0
8	ascr#1 8	C ₁₇ H ₃₂ O ₆	332.21989	331.2 1
9	ascr#2 0	C ₁₈ H ₃₄ O ₆	346.23554	345.2 3
10	ascr#2 2	C ₁₉ H ₃₆ O ₆	360.25119	359.2 4
11	ascr#2 4	C ₂₀ H ₃₈ O ₆	374.26684	373.2 6
12	ascr#2 6	C ₂₁ H ₄₀ O ₆	388.28249	387.2 8

Figure 72A



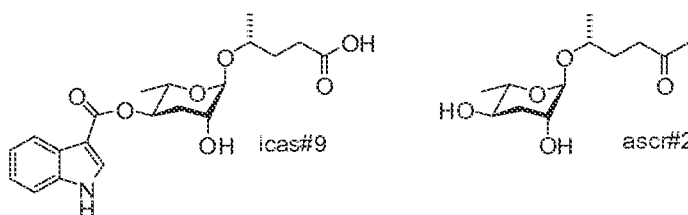
Side chain (n)	SMID ¹	Molecular formula	Molecular mass g/mol	m/z M-H ⁺
2	ascr# 7	C ₁₃ H ₂₂ O ₆	274.14164	273.1 3
4	ascr# 3	C ₁₅ H ₂₆ O ₆	302.17294	301.1 7

Figure 72B



Side chain (n)	SMID ¹	Molecular formula	Molecular mass g/mol	m/z M-H ⁺
1	ascr#5	C ₉ H ₁₆ O ₆	220.09479	219.09
3	oscr#9	C ₁₁ H ₂₀ O ₆	248.12599	247.12
7	oscr#10	C ₁₅ H ₂₈ O ₆	304.18859	303.18

Figure 72C



SMID ¹	Molecular formula	Molecular mass g/mol	m/z
icas#9	C ₂₀ H ₂₅ NO ₇	391.16310	390.16 M-H
ascr#2	C ₁₂ H ₂₂ O ₅	246.14672	269.14 M+Na

Figure 72D

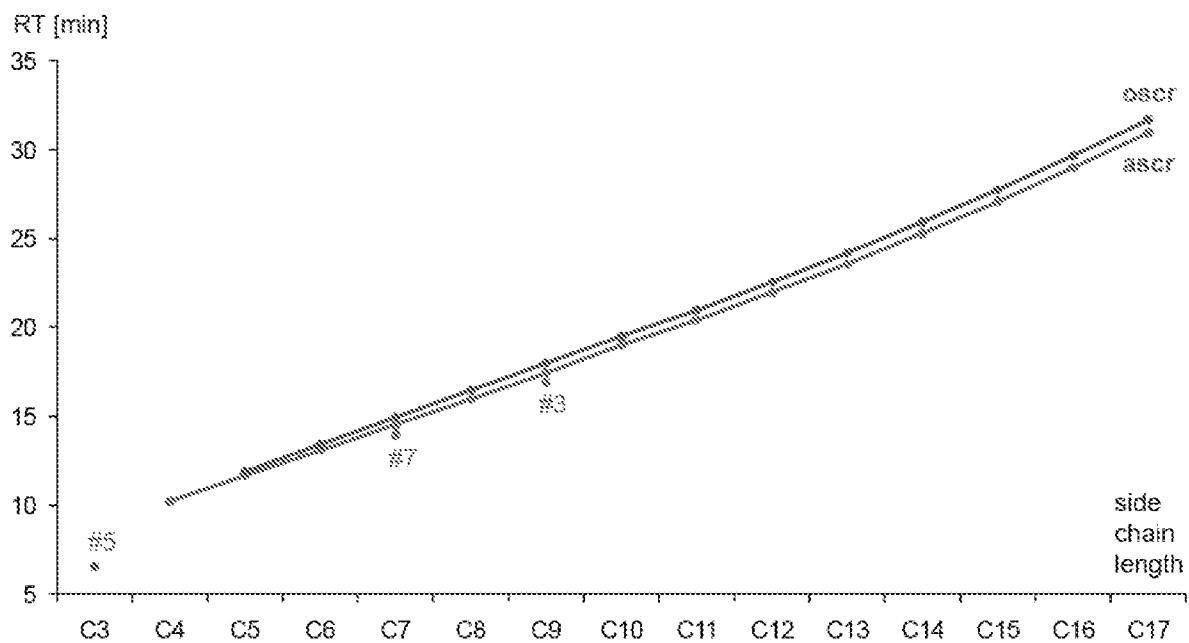
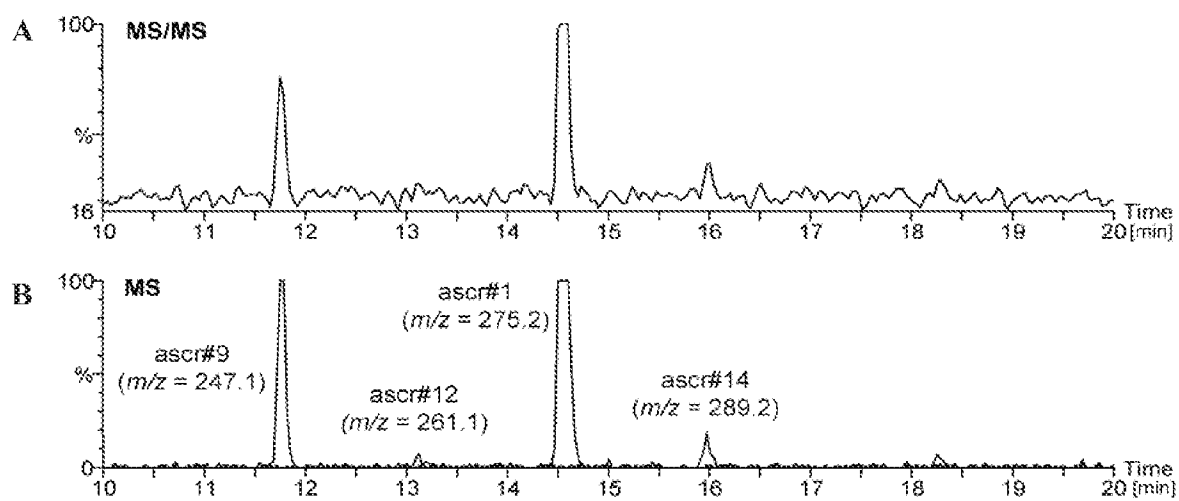


Figure 73



Figures 74A-B

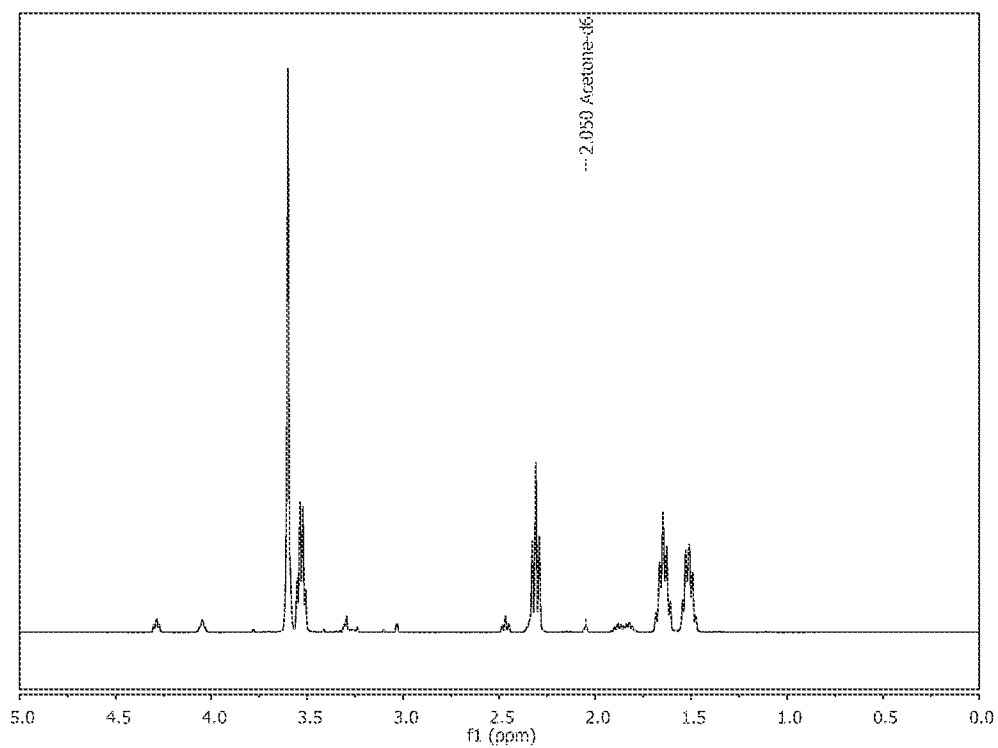


Figure 75A

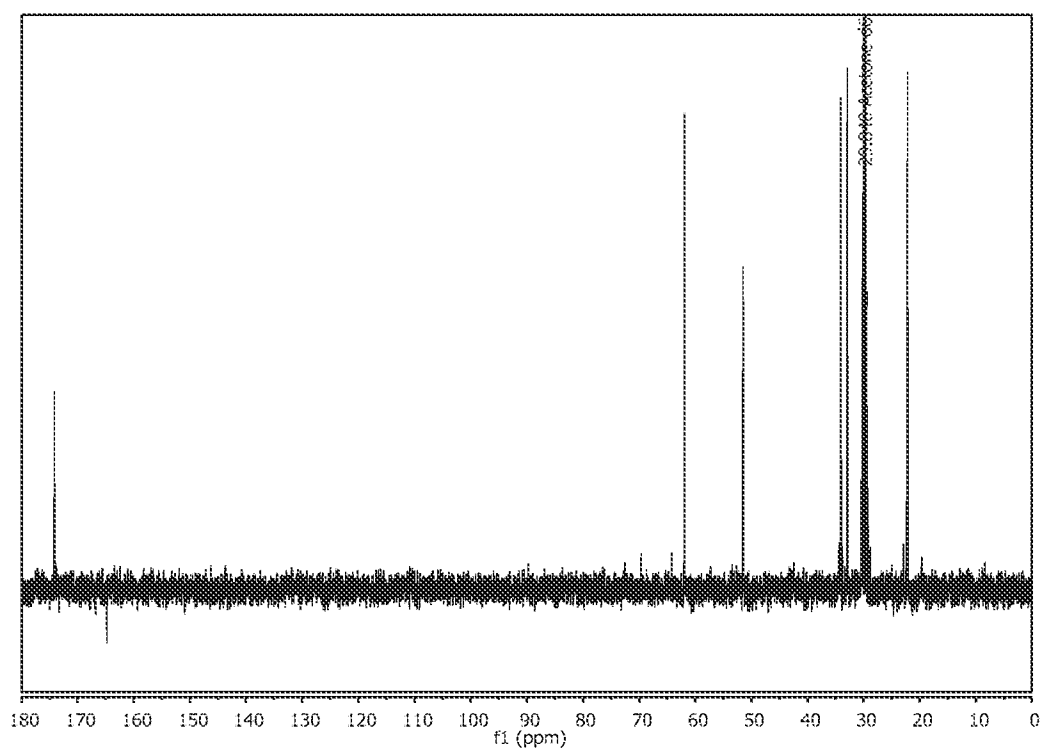


Figure 75B

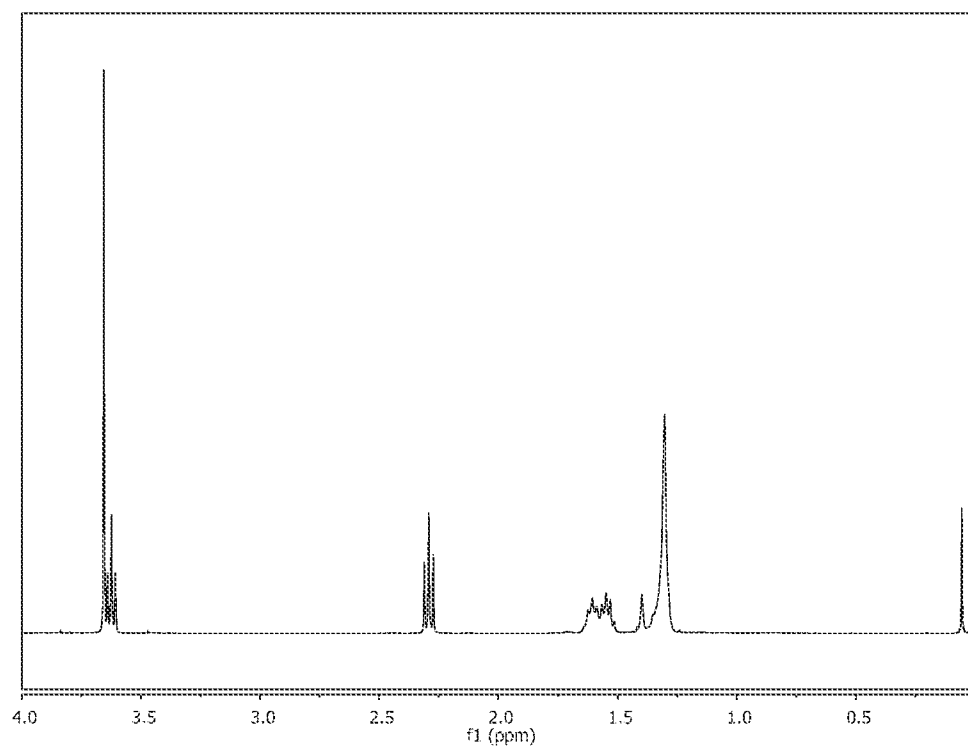


Figure 75C

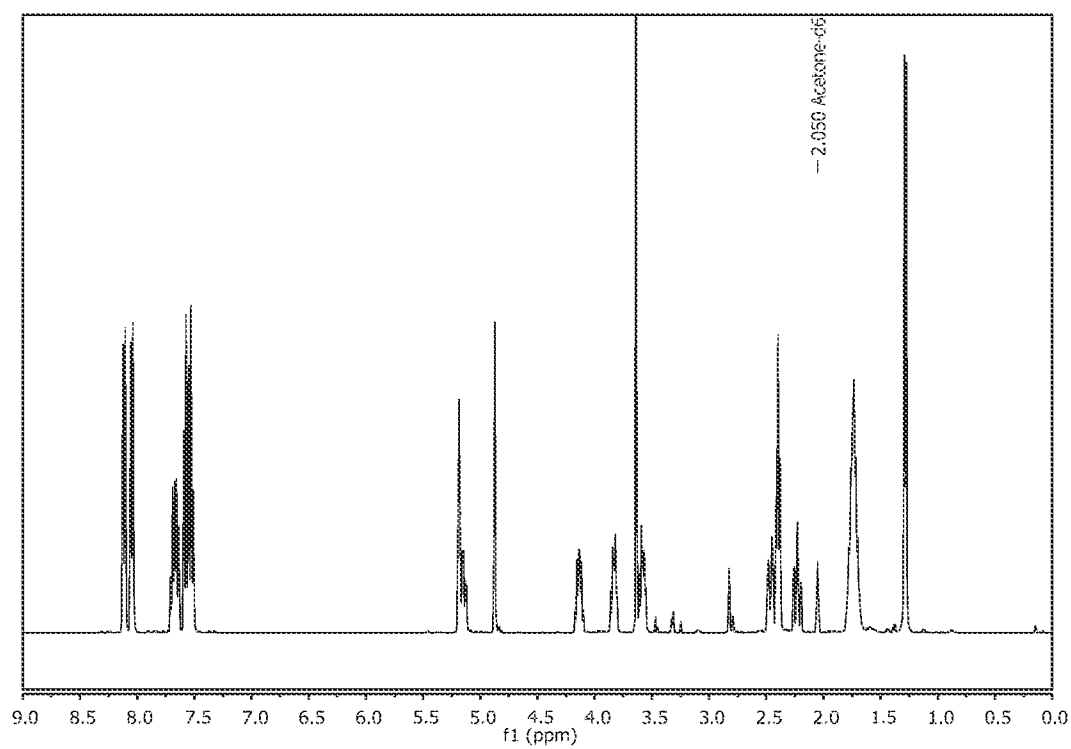


Figure 75D

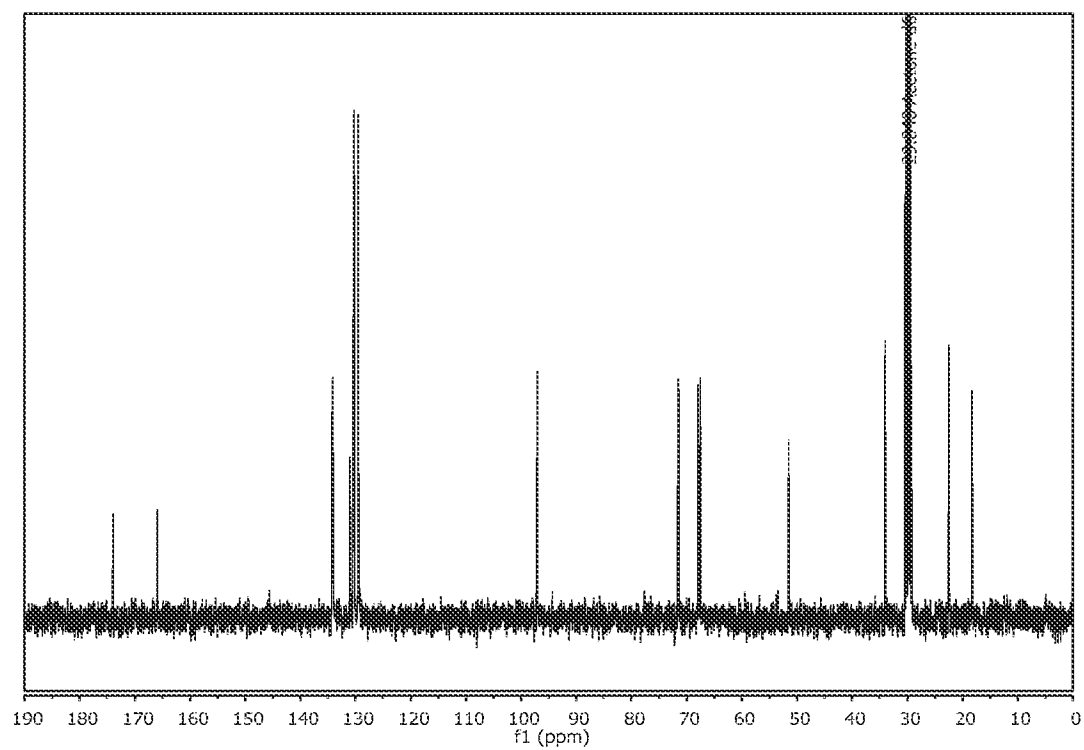


Figure 75E

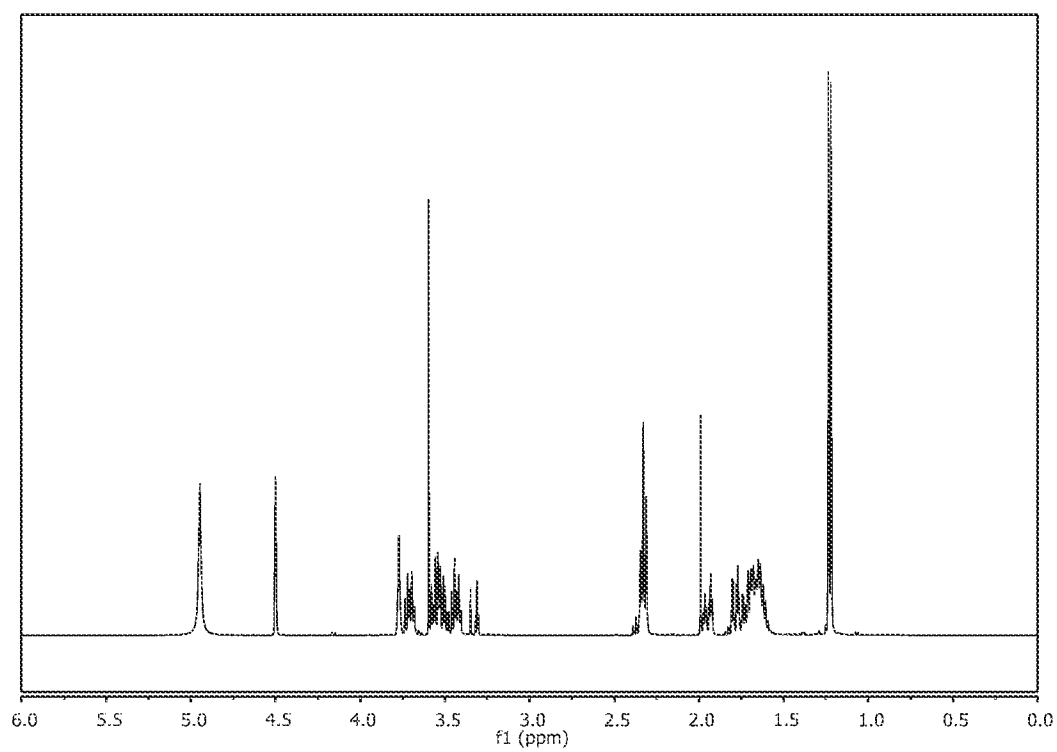


Figure 75F

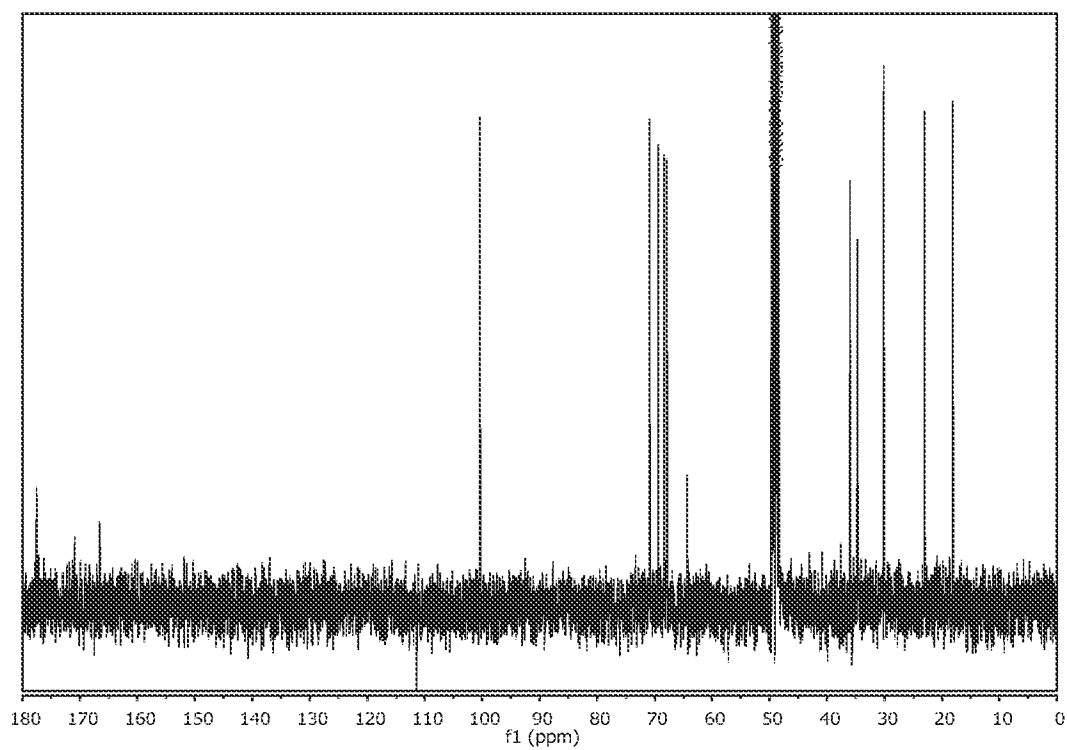


Figure 75G

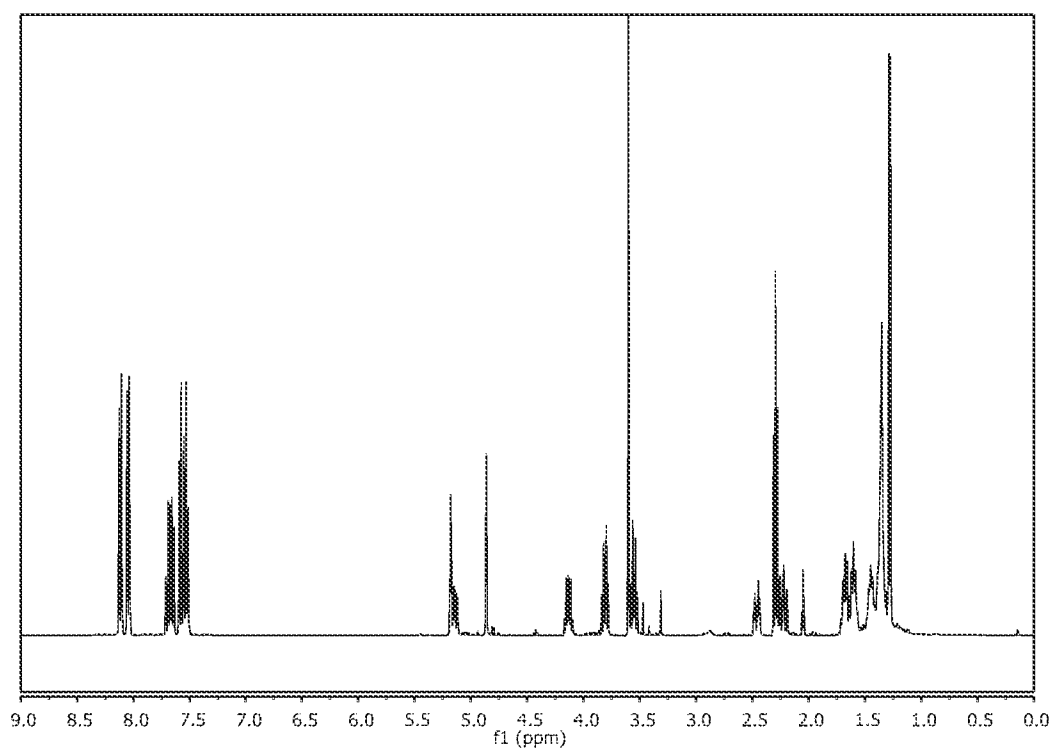


Figure 75H

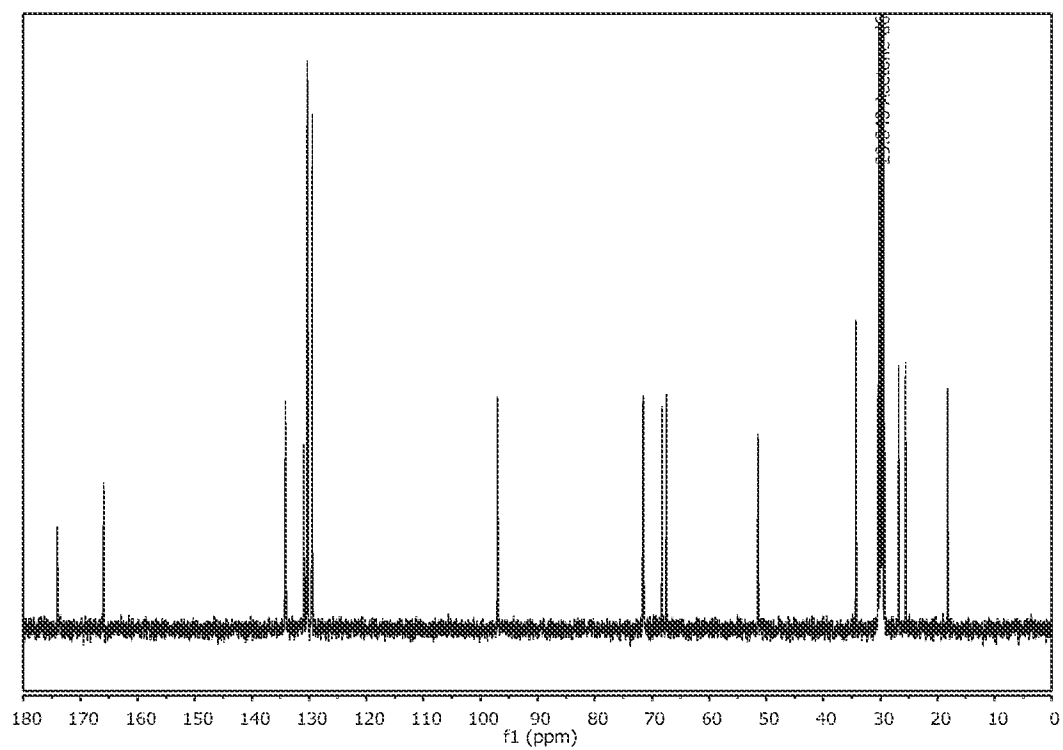


Figure 75I

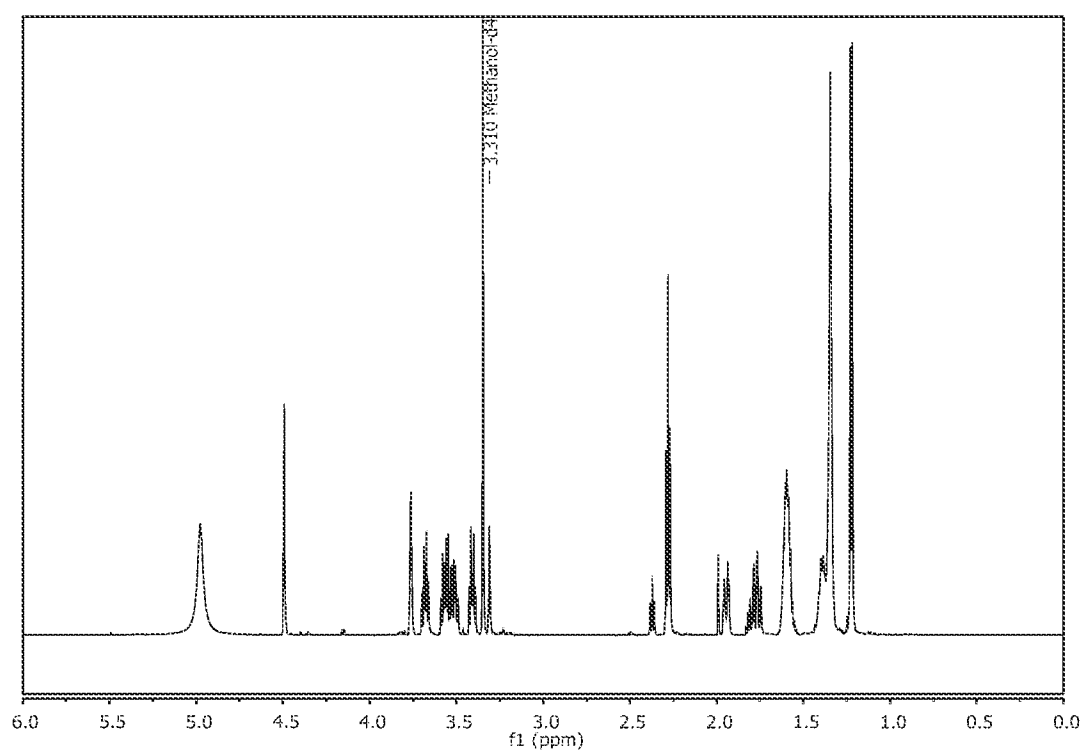


Figure 75J

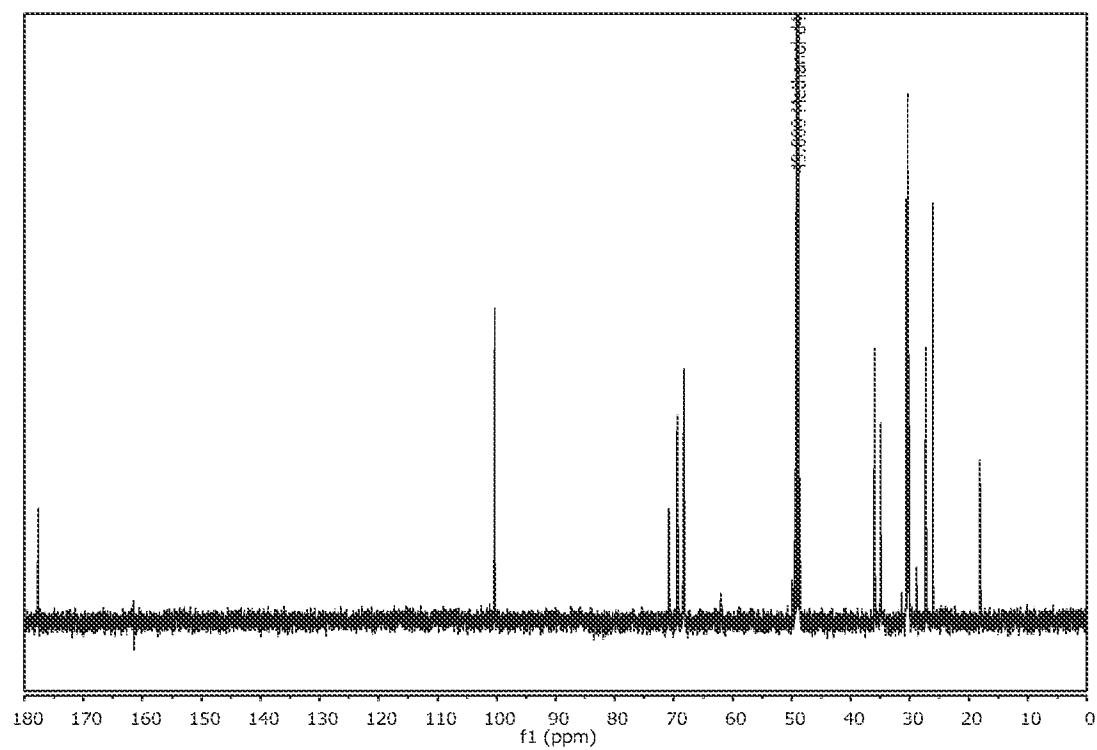
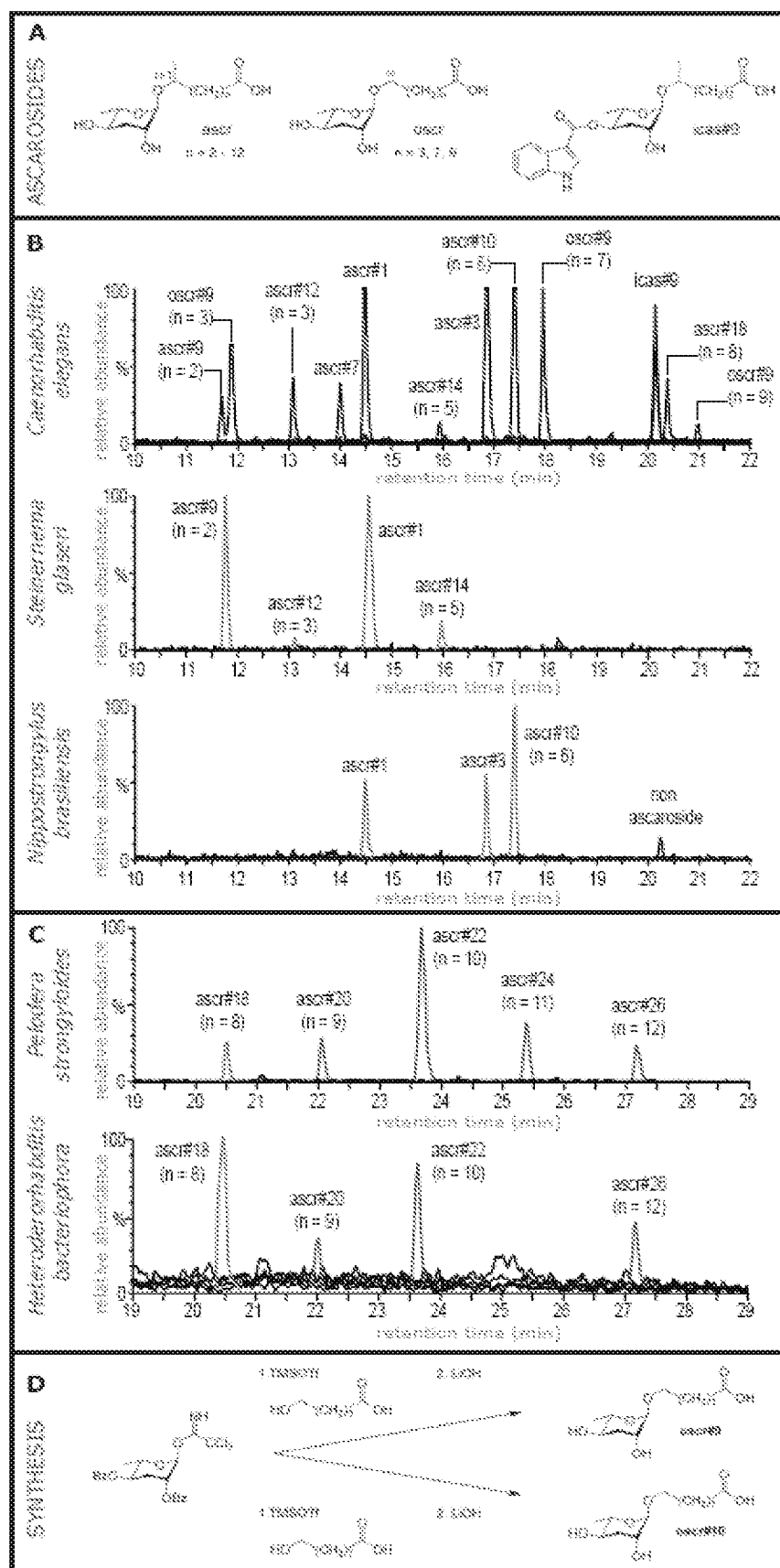


Figure 75K



Figures 76A–D

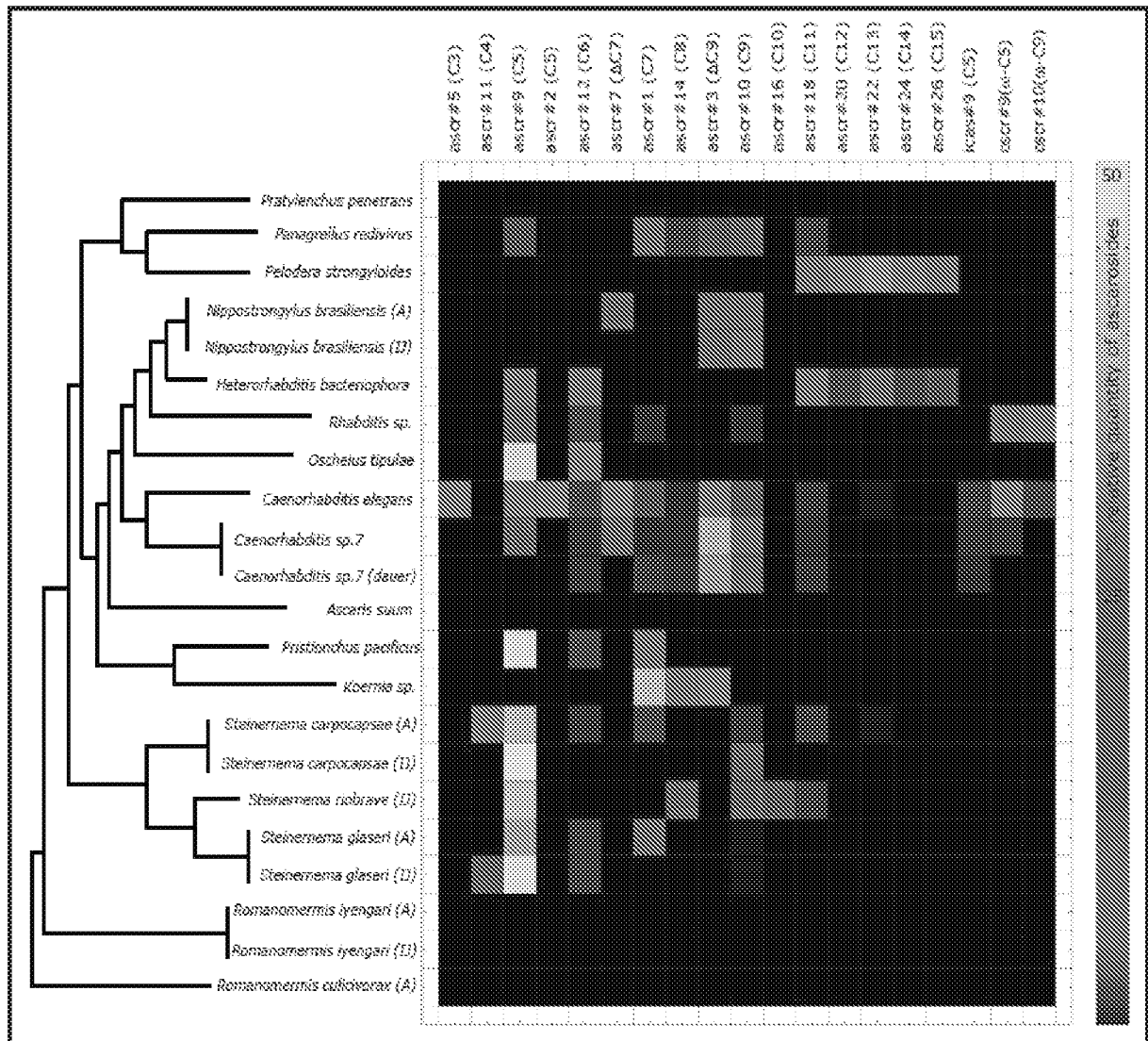


Figure 77

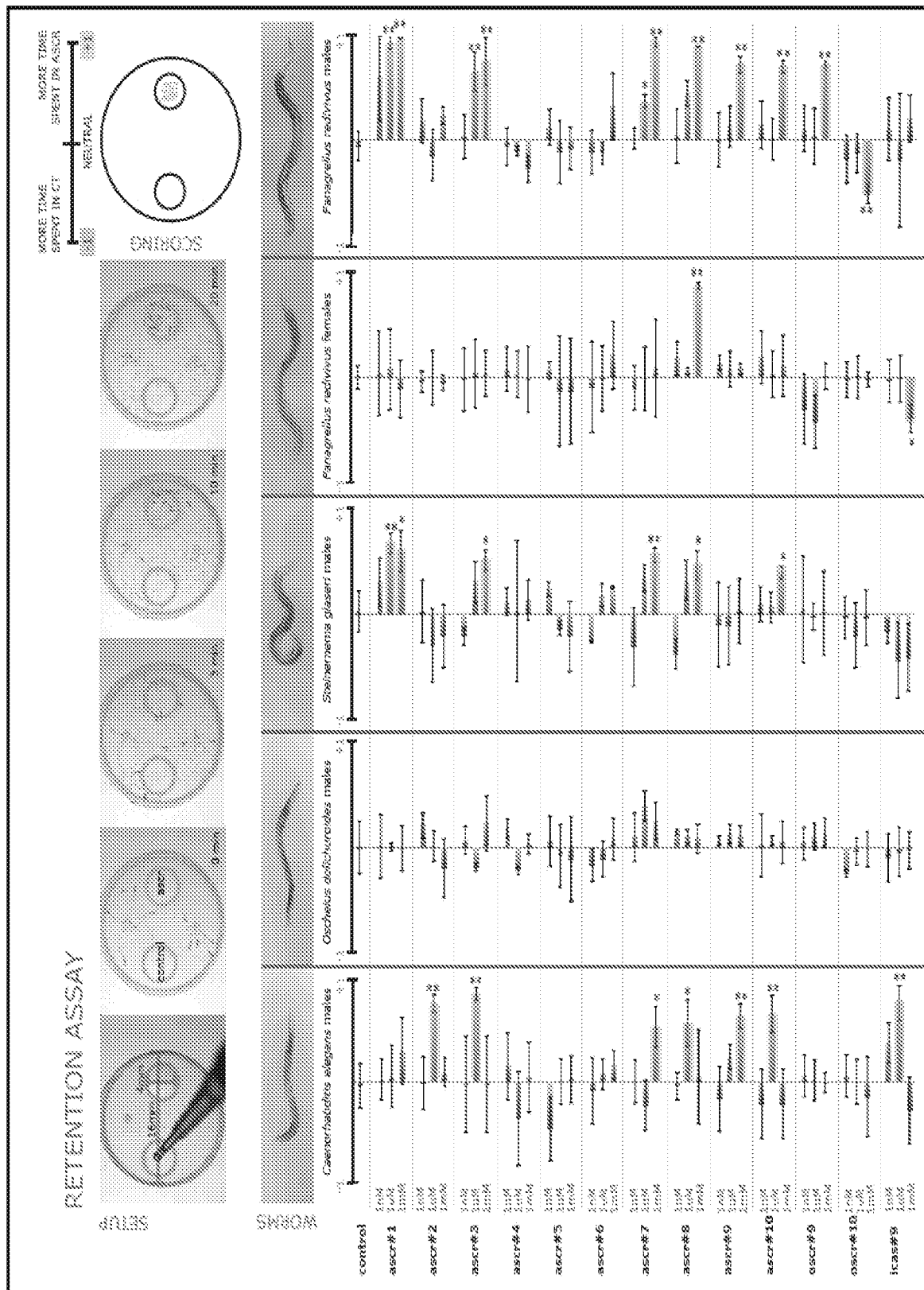


Figure 78

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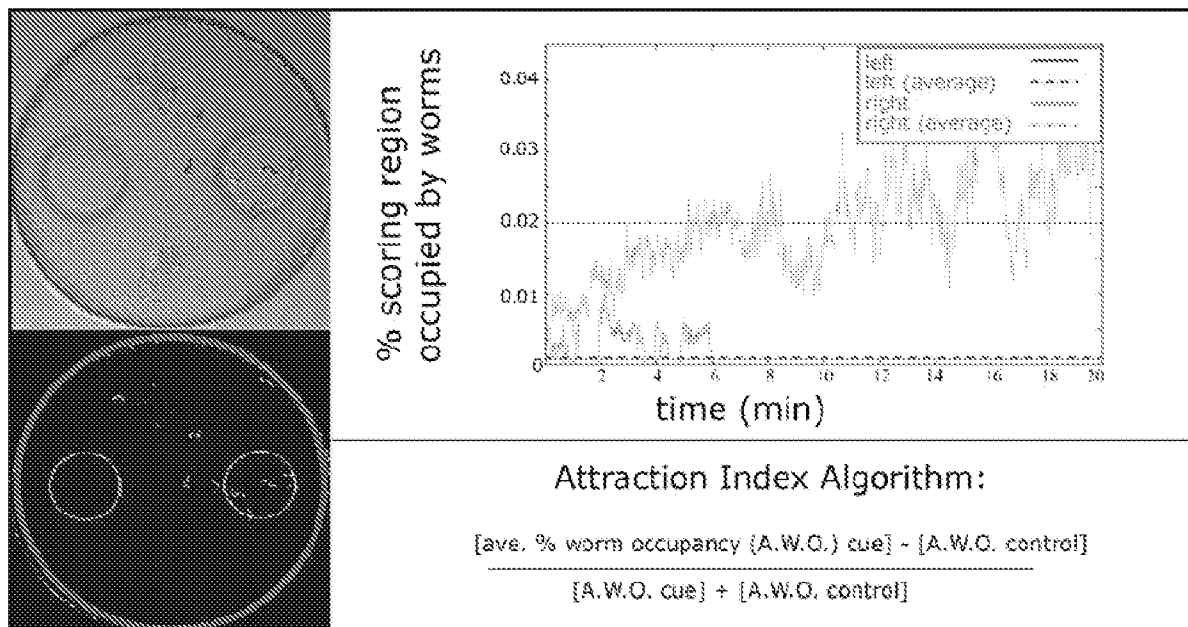


Figure 79

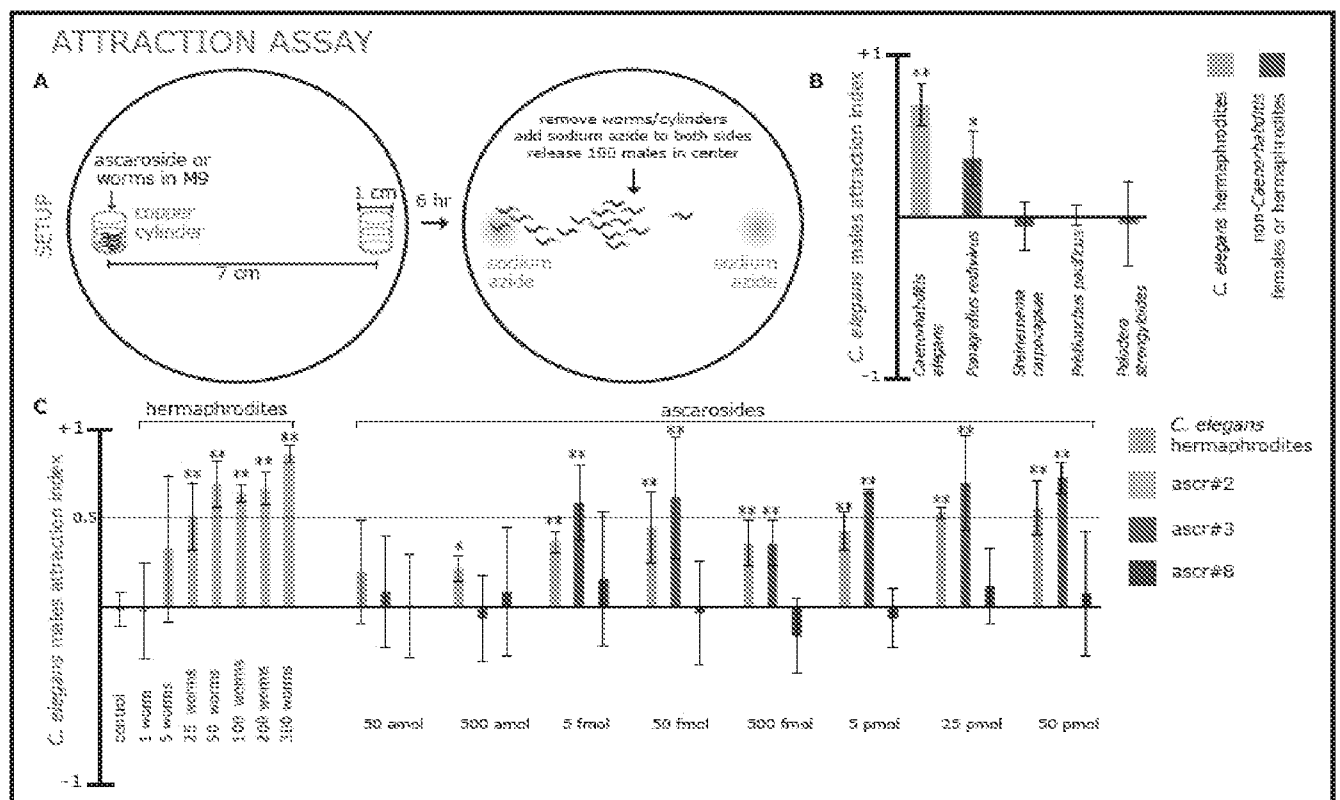


Figure 80

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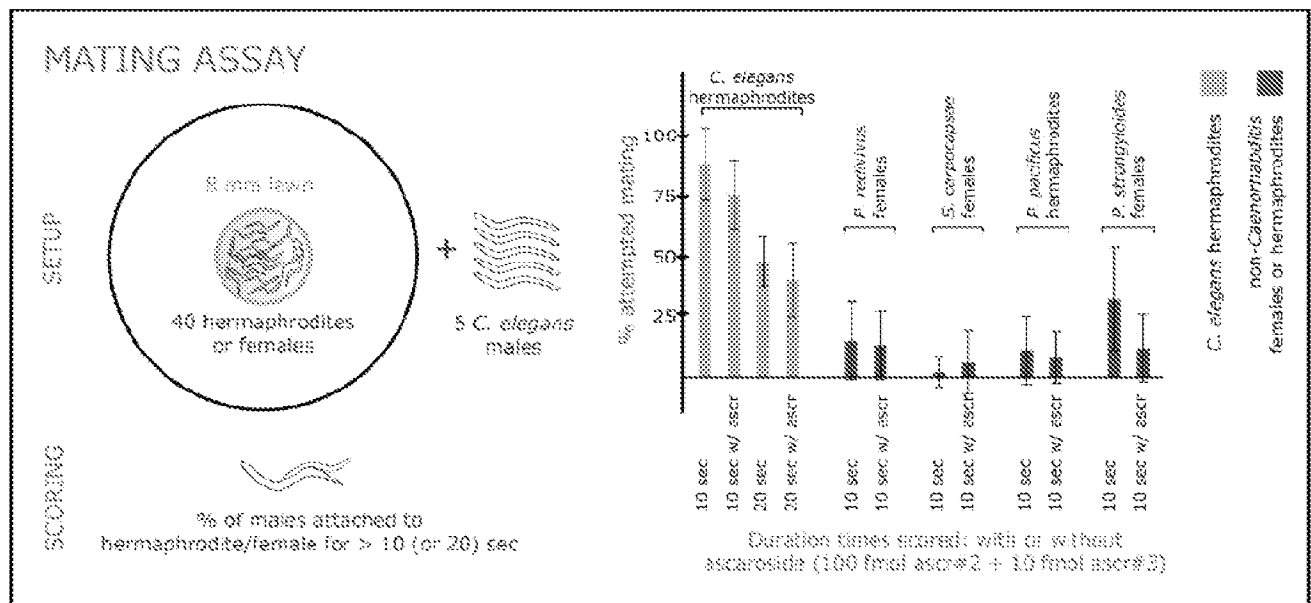


Figure 81

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	NEMATODE PHEROMONES ASCAROSIDES	BACTERIA QUORUM SENSING MOLECULES N-ACYL HOMOSERINE LACTONES
STRUCTURE		
FUNCTION	holding, avoidance, aggregation, dauer formation, olfactory plasticity, long-range attraction ¹⁵⁻¹⁸	bioluminescence, virulence, antibiotic production, biofilm formation, motility ²²
DISTRIBUTION	diverse range of nematode species not yet discovered in other animal phyla²⁰ cross-species interaction (<i>S. glaseri</i> does not make, but responds to ascr#3)	diverse range of gram-negative bacteria species²³ not yet discovered outside Proteobacteria phylum²⁸ cross-species interaction (<i>S. enterica</i> does not make, but responds to AHLs)²⁹

Figure 82

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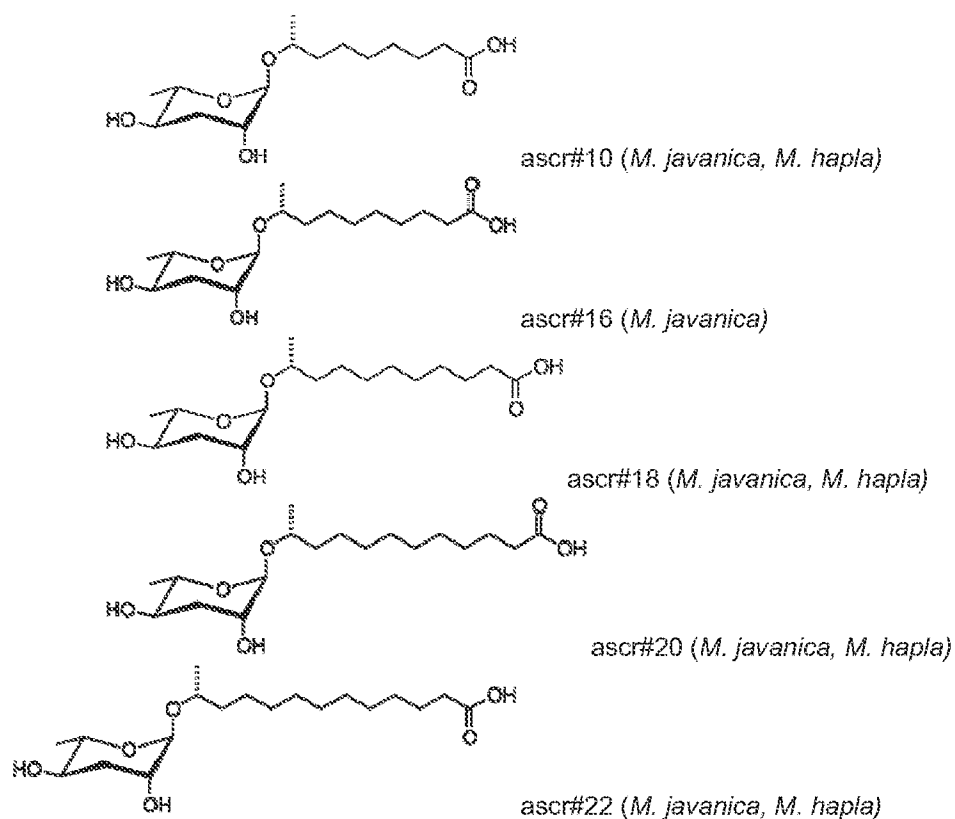
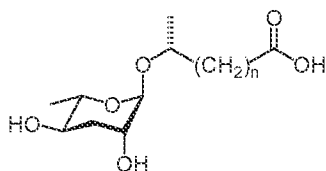


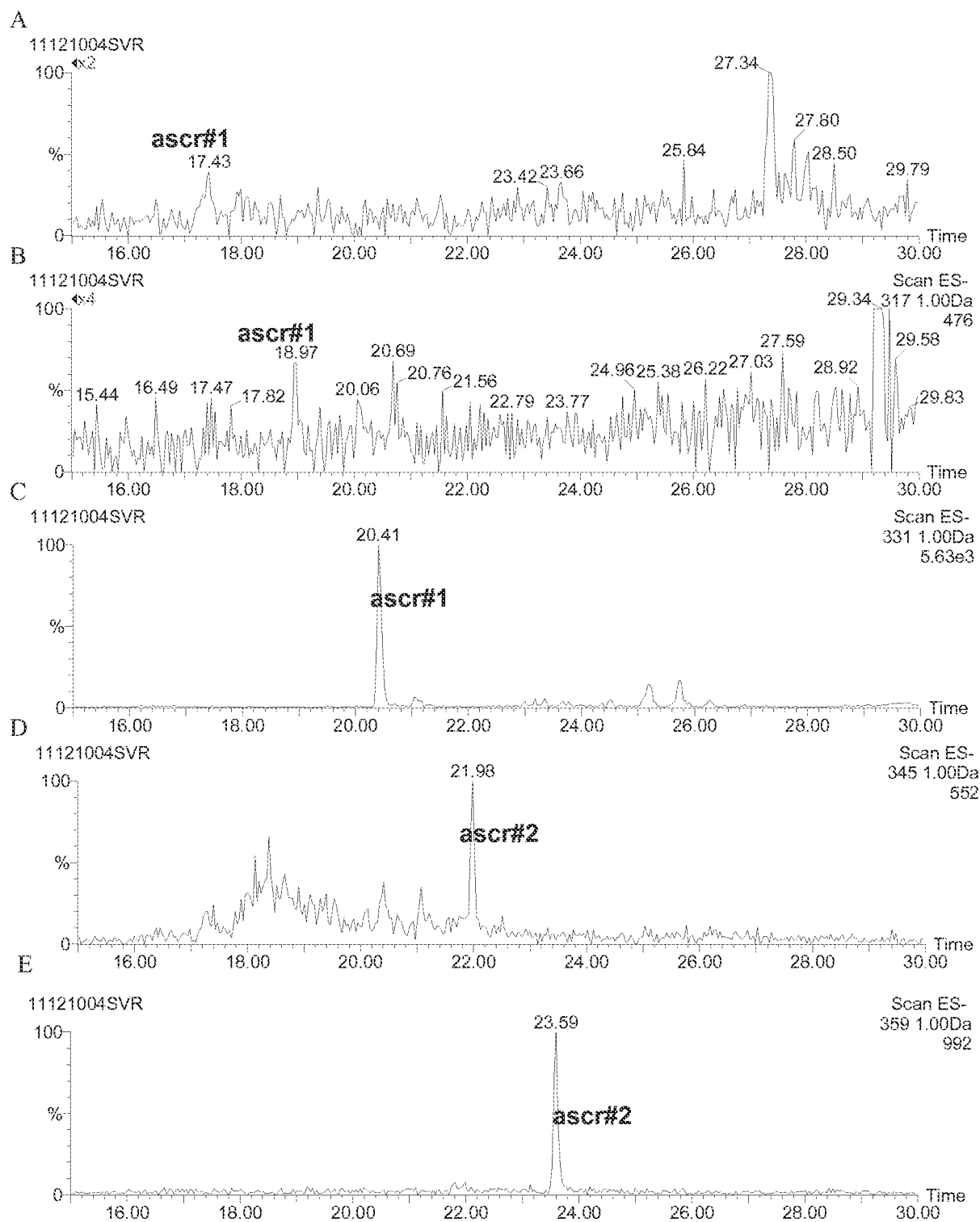
Figure 83



SMID	n	m/z calc	<i>M. hapla</i> m/z obs	<i>M. javanica</i> m/z obs
ascr#10	6	303.1808	303.1819	303.1813
ascr#16	7	317.1964		317.1959
ascr#18	8	331.2121	331.2117	331.2129
ascr#20	9	345.2277	345.2283	345.2267
ascr#22	10	359.2434	359.2434	359.2429

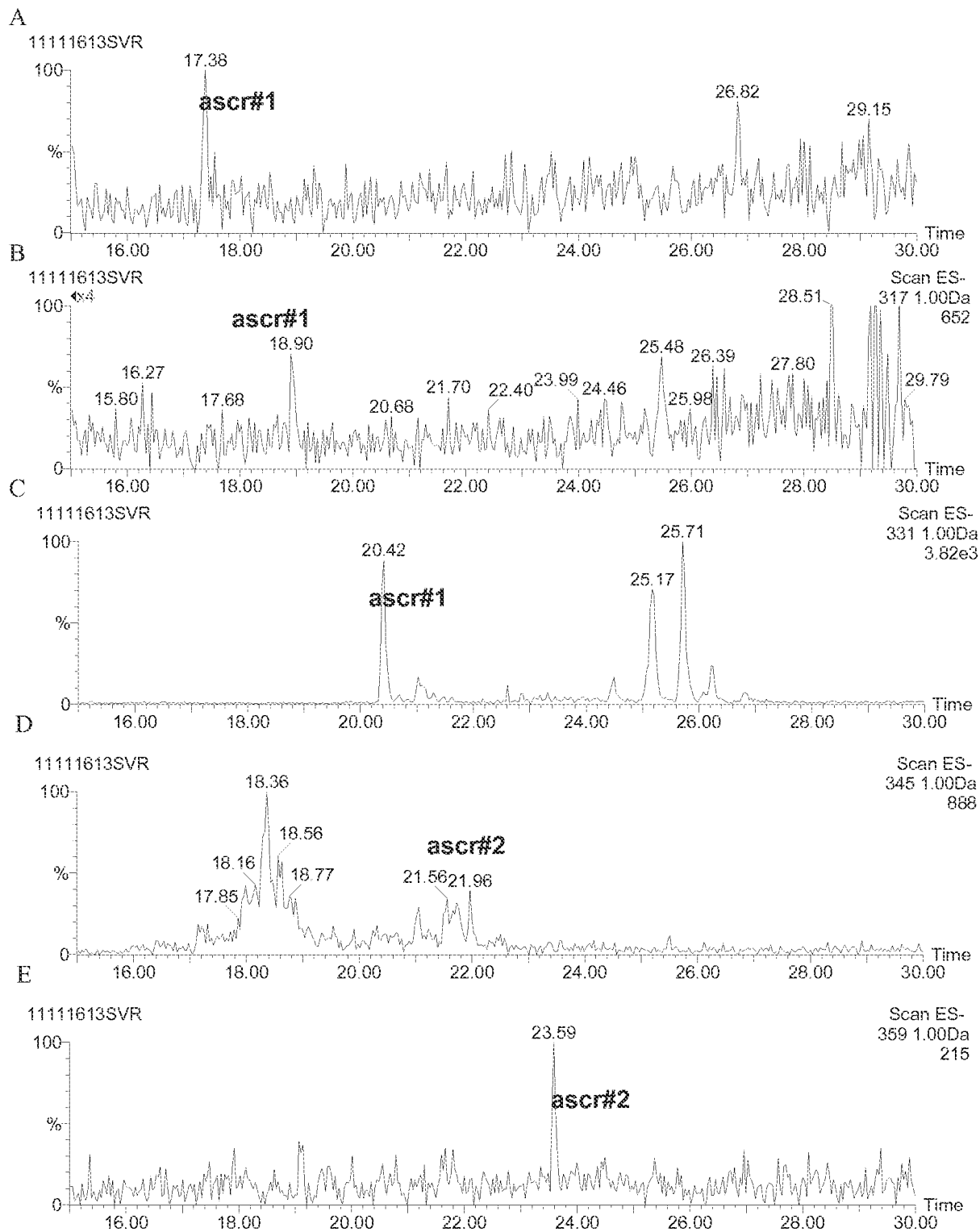
Figure 84

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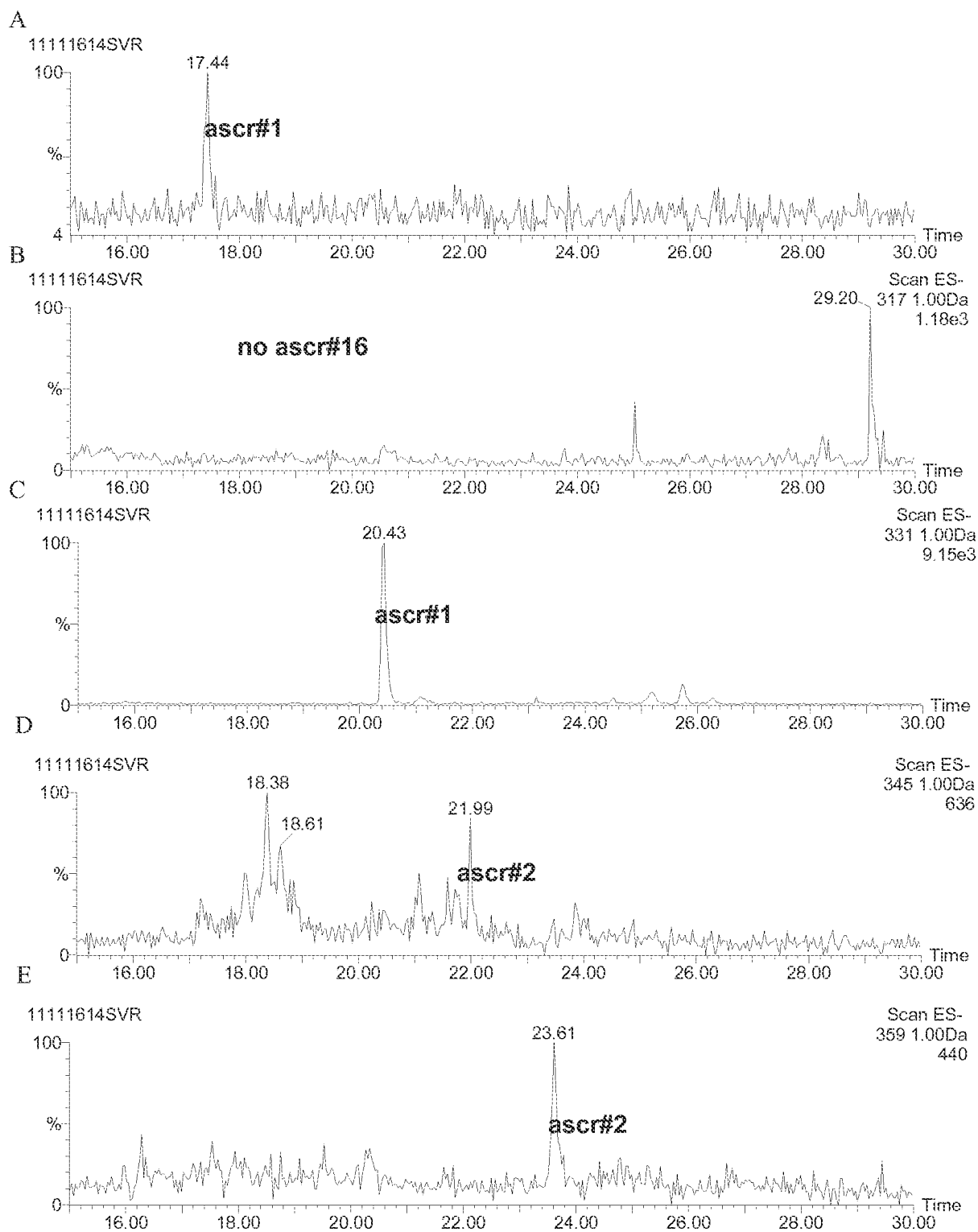
Figures 85A-E

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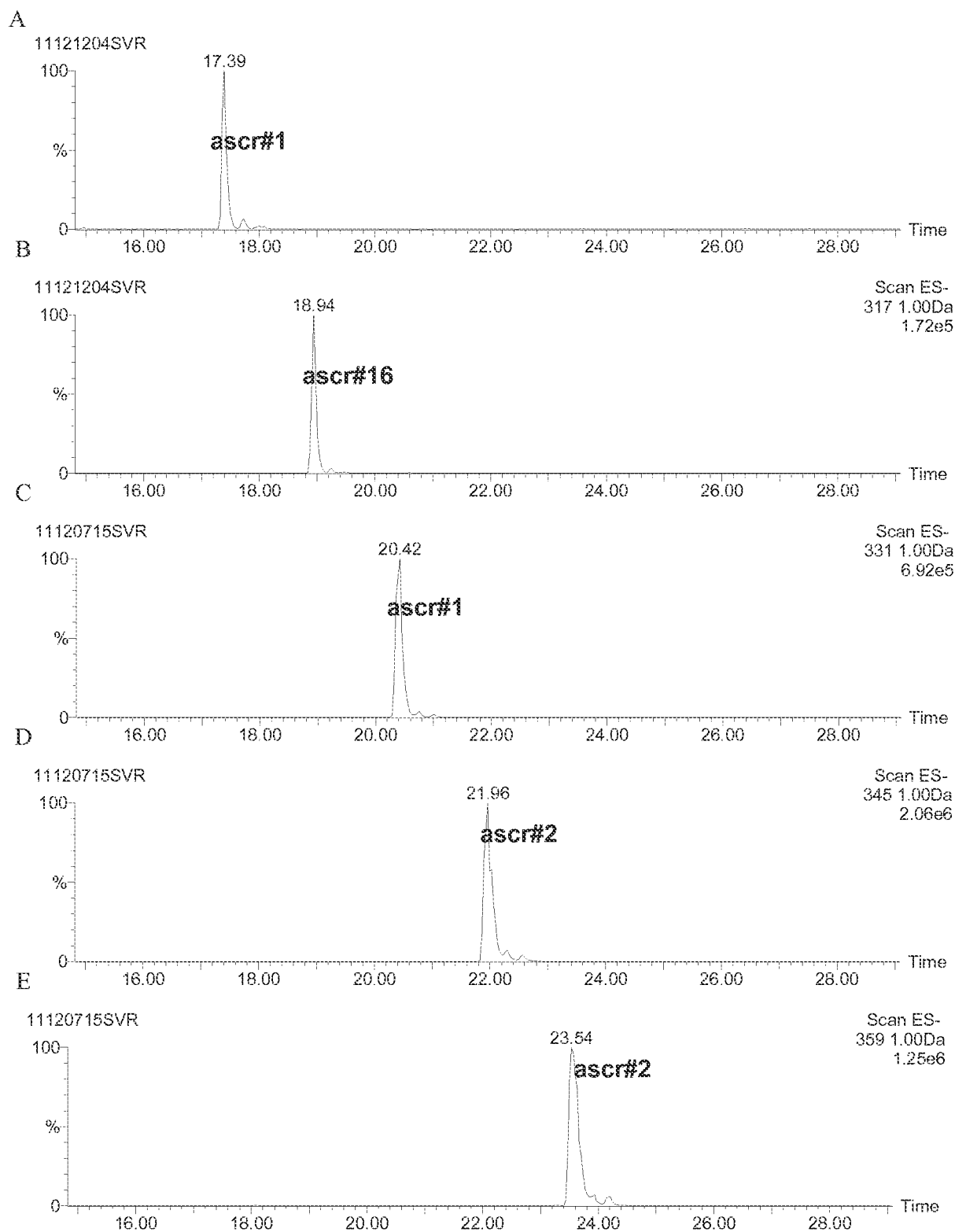
Figures 86A-E

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Figures 87A-E

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Figures 88A-E