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- (54) **Emberi lambda variábilis régiókat és egér konstans régiókat tartalmazó könnyűláncot exprimáló egerek**

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(54) **MICE EXPRESSING A LIGHT CHAIN WITH HUMAN LAMBDA VARIABLE AND MOUSE CONSTANT REGIONS**

TRANSGENE MAUS, DIE IMMUNOGLOBULIN LEICHTE KETTEN MIT HUMANER LAMBDA VARIABLEN UND MAUS KONSTANTEN REGIONEN EXPRIMIERT

SOURIS EXPRIMANT UNE CHAÎNE LÉGÈRE D'IMMUNOGLOBULIN COMPRENANT UNE RÉGION VARIABLE LAMBDA HUMAINE ET UNE RÉGION CONSTANTE MURINE

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- **GORMAN J R ET AL: "THE LGK 3' ENHANCER INFLUENCES THE RATIO OF LGK VERSUS LGL B LYMPHOCYTES", IMMUNITY, CELL PRESS, US, vol. 5, no. 3, 1 September 1996 (1996-09-01), pages 241-252, XP000891305, ISSN: 1074-7613, DOI: 10.1016/S1074-7613(00)80319-2**

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Description**FIELD**

5 **[0001]** Genetically modified mice that comprise an unrearranged human λ immunoglobulin light chain variable gene segment (hV λ) and a human λ joining gene segment (hJ λ) operably linked with an intact mouse lambda light chain constant region (C λ). Genetically modified mice that express antibodies that comprise an immunoglobulin light chain derived from a hV λ , a hJ λ and a mouse C λ gene.

10 **BACKGROUND**

[0002] Mice that express antibodies that are fully human, or partly human and partly mouse, are known in the art. For example, transgenic mice that express fully human antibodies from transgenes containing human light and heavy chain immunoglobulin variable region genes have been reported. Genetically modified mice that comprise a replacement of the endogenous mouse heavy chain variable region (HCVR) gene segments and kappa (κ) light chain variable region (LCVR) gene segments with human HCVR and LCVR gene segments and that make chimeric antibodies with a chimeric human/mouse kappa chain are known as well.

15 **[0003]** Antibody light chains are encoded by one of two separate loci: kappa (κ) and lambda (λ). Mouse antibody light chains are primarily of the κ type. The ratio of κ to λ light chain usage in humans is about 60:40, whereas in mice it is about 95:5. Biased usage of κ light chains in mice is reportedly sustained in genetically modified mice capable of expressing fully or partly human antibodies. Thus, mice that express fully or partly human antibodies appear to be constrained in lambda variable usage.

20 **[0004]** There is a need in the art to generate lambda variable regions, whether mouse or human, for use in making epitope-binding proteins. There is a need in the art for mice that express fully or partly human antibodies, wherein the mice display an increased lambda variable (V λ) usage.

25 **[0005]** There is a need in the art for mice that express fully or partly human antibodies, wherein the mice display an increased λ variable (V λ) usage. WO 00/26373 refers to murine expression of human Ig λ locus.

30 **[0006]** WO 2010/039900 relates to knock-in non-human cells and mammals having a genome encoding chimeric Ig chains and antibodies and methods for producing such knock-in cells and mammals. A knock-in mouse having a genome comprising a human VH segment, a human JH gene segment and a human DH gene segment and a portion of a syngeneic Ig heavy chain locus, is disclosed, where the human heavy chain locus replaces all or portion of an endogenous heavy chain locus so that the animal is capable of producing chimeric heavy chains. In specific embodiments the mouse genome further comprises a human IgA light chain locus, or a portion thereof, and an Ig λ 3'LCR, where the human Ig λ light chain locus replaces all or portion of an endogenous Ig light chain locus.

35 **SUMMARY**

[0007] The invention in its broadest sense is as defined in the independent claims. Genetically modified mice, embryos, cells, tissues, as well as nucleic acid constructs for modifying mice, and methods and compositions for making and using them, are disclosed. Mice and cells that generate human λ variable regions in the context of a λ light chain, from an endogenous mouse light chain locus, are also provided as recited in the claims. Also disclosed are methods for making antibodies that comprise lambda variable regions. Methods for selecting heavy chains that express with cognate lambda variable regions are also disclosed.

45 **[0008]** Chimeric and human antigen-binding proteins (e.g., antibodies), and nucleic acids encoding them, are disclosed that comprise somatically mutated variable regions, including antibodies that have light chains comprising a variable domain derived from a human V λ and a human J λ gene segment fused to a mouse lambda light chain constant domain.

[0009] The present invention provides a mouse comprising an endogenous λ (lambda) light chain allele comprising an unrearranged human lambda variable (hV λ) and an unrearranged human lambda joining (hJ λ) gene segment, where the mouse comprises a deletion of a first V λ -J λ -C λ gene cluster of said endogenous mouse A light chain allele and replacement, in whole or in part, of the V λ -J λ gene segments of the second V λ -J λ -C λ gene cluster of said allele with said human V λ and J λ gene segments which are operably linked to an intact mouse lambda constant (C λ) region gene of said second V λ -J λ -C λ gene cluster so the mouse expresses a light chain derived from a hV λ , a hJ λ and a mouse C λ gene, where the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele.

55 **[0010]** The invention further provides:

an isolated cell that expresses a light chain derived from a hV λ , a hJ λ and a mouse C λ gene, wherein the cell is from, or obtainable from, the mouse of any one of the preceding claims, and wherein the cell comprises a rearranged

human V λ and a rearranged human J λ gene segment, and a deletion of a first V λ -J λ -C λ gene cluster of said endogenous mouse λ light chain allele and replacement, in whole or in part, of the V λ -J λ gene segments of the second V λ -J λ -C λ gene cluster of said allele with said human V λ and J λ gene segments which are operably linked to an intact mouse C λ constant region gene of said second V λ -J λ -C λ gene cluster, and wherein the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele;

an isolated mouse embryonic stem (ES) cell comprising an endogenous λ (lambda) light chain allele comprising an unrearranged human V λ and an unrearranged human J λ gene segment, where the cell comprises a deletion of a first V λ -J λ -C λ gene cluster of said endogenous mouse A light chain allele and a replacement, in whole or in part, of the V λ -J λ gene segments of the second V λ -J λ -C λ gene cluster of said allele with said human V λ and J λ gene segments which are operably linked to an intact mouse C λ constant region gene of said second V λ -J λ -C λ gene cluster, where the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele;

a mouse embryo comprising, made from, or obtainable from, the mouse ES cell of the invention, wherein the mouse embryo comprises an endogenous A (lambda) light chain allele comprising an unrearranged human V λ and an unrearranged human J λ gene segment, where the cell comprises a deletion of a first V λ -J λ -C λ gene cluster of said endogenous mouse λ light chain allele and a replacement, in whole or in part, of the V λ -J λ gene segments of the second V λ -J λ -C λ gene cluster of said allele with said human V λ and J λ gene segments which are operably linked to an intact mouse C λ constant region gene of said second V λ -J λ -C λ gene cluster, where the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele;

a hybridoma that expresses a light chain derived from a hV λ , a hJ λ and a mouse C λ gene wherein the hybridoma is from, or obtainable from, the mouse of the invention, wherein the cell comprises a rearranged human V λ and a rearranged human J λ gene segment, and a deletion of a first V λ -J λ -C λ gene cluster of said endogenous mouse A light chain allele and replacement, in whole or in part, of the V λ -J λ gene segments of the second V λ -J λ -C λ gene cluster of the allele with said human V λ and J λ gene segments which are operably linked to an intact mouse C λ constant region gene of said second V λ -J λ -C λ gene cluster, and wherein the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele; and

use of the B cell of the invention to make a hybridoma that expresses a light chain derived from a hV λ , a hJ λ and a mouse C λ gene, wherein the hybridoma is from, or obtainable from, the mouse of the invention, wherein the cell comprises a rearranged human V λ and a rearranged human J λ gene segment, and a deletion of a first V λ -J λ -C λ gene cluster of said endogenous mouse λ light chain allele and replacement, in whole or in part, of the V λ -J λ gene segments of the second V λ -J λ -C λ gene cluster of said allele with said human V λ and J λ gene segments which are operably linked to an intact mouse C λ constant region gene of said second V λ -J λ -C λ gene cluster and wherein the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele.

[0011] The present invention further provides a method of making an antibody in a mouse comprising: (a) exposing a mouse of the invention to an antigen; (b) allowing the mouse to develop an immune response to the antigen; and (c) isolating from the mouse of (b) an antibody that specifically recognizes the antigen, or isolating from the mouse of (b) a cell comprising an immunoglobulin domain that specifically recognizes the antigen, or identifying in the mouse of (b) a nucleic acid sequence encoding a heavy and/or light chain variable domain that binds the antigen, wherein the antibody comprises a light chain derived from a hV λ , a hJ λ and a mouse C λ gene.

[0012] Antibodies are disclosed that comprise (a) a human heavy chain variable domain (hV_H) fused to a mouse heavy chain constant region, and (b) a human V λ fused to a mouse C_L domain; including wherein one or more of the variable domains are somatically mutated, e.g., during antibody or immune cell selection in a mouse of the invention. The unrearranged hV λ and unrearranged hJ λ are operably linked with an intact mouse λ constant region (C λ).

[0013] Also disclosed is a mouse that comprises in its germline, at an endogenous mouse light chain locus, a human λ light chain variable region sequence, wherein the human lambda variable region sequence is expressed in a light chain that comprises a mouse immunoglobulin lambda constant region gene sequence.

[0014] The endogenous mouse light chain locus is a λ locus.

[0015] In one aspect, the mouse lacks an endogenous light chain variable sequence at the endogenous mouse light chain locus.

[0016] In one aspect, all or substantially all endogenous mouse light chain variable region gene segments are replaced with one or more human λ variable region gene segments.

[0017] The human λ light chain variable region sequence comprises a human J λ sequence. In one aspect, the human

J λ sequence is selected from the group consisting of J λ 1, J λ 2, J λ 3, J λ 7, and a combination thereof.

[0018] In one aspect, the human λ light chain variable region sequence comprises a fragment of cluster A of the human light chain locus. In a specific aspect, the fragment of cluster A of the human λ light chain locus extends from hV λ 3-27 through hV λ 3-1.

[0019] In one aspect, the human λ light chain variable region sequence comprises a fragment of cluster B of the human light chain locus. In a specific aspect, the fragment of cluster B of the human λ light chain locus extends from hV λ 5-52 through hV λ 1-40.

[0020] In one aspect, the human λ light chain variable region sequence comprises a genomic fragment of cluster A and a genomic fragment of cluster B. In a one aspect, the human λ light chain variable region sequence comprises at least one gene segment of cluster A and at least one gene segment of cluster B.

[0021] In one aspect, more than 10% of the light chain naïve repertoire of the mouse is derived from at least two hV λ gene segments selected from 2-8, 2-23, 1-40, 5-45, and 9-49. In one aspect, more than 20% of the light chain naïve repertoire of the mouse is derived from at least three hV λ gene segments selected from 2-8, 2-23, 1-40, 5-45, and 9-49. In one aspect, more than 30% of the light chain naïve repertoire of the mouse is derived from at least four hV λ gene segments selected from 2-8, 2-23, 1-40, 5-45, and 9-49.

[0022] Also disclosed is a genetically modified mouse, wherein the mouse comprises an unrearranged immunoglobulin V λ and a J λ gene segment operably linked to a mouse light chain locus that comprises an intact mouse C λ gene.

[0023] The V λ and J λ gene segments are human gene segments.

[0024] The endogenous mouse light chain locus is a λ light chain locus.

[0025] In one aspect, the mouse further comprises a replacement of one or more heavy chain V, D, and/or J gene segments with one or more human V, D, and/or J gene segments at an endogenous mouse heavy chain immunoglobulin locus.

[0026] In one aspect, the mouse comprises an unrearranged human immunoglobulin λ light chain variable gene segment (V λ) and a λ joining gene segment (J λ) at an endogenous mouse λ light chain locus that comprises an intact mouse C λ gene.

[0027] The λ light chain variable gene locus (the " λ locus") comprises at least one hV λ gene segment and at least one human J λ (hJ λ) gene segment. In another aspect, the λ locus comprises up to four hJ λ gene segments.

[0028] In one aspect, the V λ locus comprises a plurality of hV λ s. In one aspect, the plurality of hV λ s are selected so as to result in expression of a λ light chain variable region repertoire that reflects about 10%, about 20%, about 30%, about 40%, about 50%, about 60%, about 70%, about 80%, or about 90% or more of the V λ usage observed in a human. In one aspect, the V λ locus comprises gene segments hV λ 1-40, 1-44, 2-8, 2-14, 3-21, and a combination thereof.

[0029] In one aspect, the hV λ s include 3-1, 4-3, 2-8, 3-9, 3-10, 2-11, and 3-12. In a specific aspect, the V λ locus comprises a contiguous sequence of the human λ light chain locus that spans from V λ 3-12 to V λ 3-1. In one aspect, the V λ locus comprises at least 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12 hV λ s. In a specific aspect, the hV λ s include 3-1, 4-3, 2-8, 3-9, 3-10, 2-11, and 3-12. In a specific aspect, the V λ locus comprises a contiguous sequence of the human λ locus that spans from V λ 3-12 to V λ 3-1. In a specific aspect the endogenous κ locus is deleted in part or completely.

[0030] In one aspect, the V λ locus comprises 13 to 28 or more hV λ s. In a specific aspect, the hV λ s include 2-14, 3-16, 2-18, 3-19, 3-21, 3-22, 2-23, 3-25, and 3-27. In a specific aspect the endogenous κ locus is deleted in part or completely.

[0031] In one aspect, the V λ locus comprises 29 to 40 hV λ s. In a specific embodiment, all or substantially all sequence between hV λ 1-40 and hV λ 3-29 in the genetically modified mouse consists essentially of a human λ sequence of approximately 959 bp found in nature (e.g., in the human population) downstream of the hV λ 1-40 gene segment (downstream of the 3' untranslated portion), a restriction enzyme site (e.g., PI-SceI), followed by a human λ sequence of approximately 3,431 bp upstream of the hV λ 3-29 gene segment found in nature. In a specific aspect, the endogenous mouse κ locus is deleted in part or completely.

[0032] The V λ locus comprises at least one hJ λ . In one aspect, the V λ locus comprises a plurality of hJ λ s. In one aspect, the V λ locus comprises at least 2, 3, 4, 5, 6, or 7 hJ λ . In a specific aspect, the V λ locus comprises four hJ λ . In a specific aspect, the four hJ λ s are hJ λ 1, hJ λ 2, hJ λ 3, and hJ λ 7. In one aspect, the V λ locus comprises one hJ λ . In a specific aspect, the one hJ λ , is hJ λ 1. In a specific aspect, the endogenous κ locus is deleted in part or completely.

[0033] The V λ locus comprises at least one hV λ , at least one hJ λ , and an intact mouse C λ gene. In a specific aspect, the intact mouse C λ gene is C λ 2.

[0034] In one aspect, the mouse comprises a replacement of endogenous mouse V λ gene segments at the endogenous mouse λ locus with one or more human V λ gene segments at the endogenous mouse λ locus, wherein the hV λ gene segments are operably linked to an intact mouse C λ region gene, such that the mouse rearranges the hV λ gene segments and expresses a reverse chimeric immunoglobulin light chain that comprises a hV λ domain and a mouse C λ .

In a specific aspect, the intact mouse C λ gene is C λ 2. In one aspect, 90-100% of unrearranged mouse V λ gene segments are replaced with at least one unrearranged hV λ gene segment. In a specific aspect, all or substantially all of the endogenous mouse V λ gene segments are replaced with at least one unrearranged hV λ gene segment. In one aspect, the replacement is with at least 12, at least 28, or at least 40 unrearranged hV λ gene segments. In one embodiment,

the replacement is with at least 7 functional unrearranged hV λ gene segments, at least 16 functional unrearranged hV λ , gene segments, or at least 27 functional unrearranged hV λ , gene segments. In one embodiment, the mouse comprises a replacement of all mouse J λ gene segments with at least one unrearranged hJ λ gene segment. In one aspect, the at least one unrearranged hJ λ gene segment is selected from J λ 1, J λ 2, J λ 3, J λ 4, J λ 5, J λ 6, J λ 7, and a combination thereof.

[0035] Also disclosed is a genetically modified mouse, wherein the mouse comprises no more than two light chain alleles, wherein the light chain alleles comprise (a) an unrearranged immunoglobulin human V λ and a J λ gene segment at an endogenous mouse light chain locus that comprises an intact mouse C λ gene; and, (b) an unrearranged immunoglobulin V λ and a J λ gene segment at an endogenous mouse light chain locus that comprises a mouse C λ gene.

[0036] In one aspect, the endogenous mouse light chain locus is a κ locus. In another aspect, the endogenous mouse light chain locus is a λ locus.

[0037] In one aspect, the no more than two light chain alleles are selected from a κ allele and a λ allele, and two λ alleles. In a specific aspect, one of the two light chain alleles is a λ allele that comprises an intact C λ 2 gene.

[0038] In one aspect, the mouse comprises one functional immunoglobulin light chain locus and one nonfunctional light chain locus, wherein the functional light chain locus comprises an unrearranged immunoglobulin human V λ and a J λ gene segment at an endogenous mouse λ light chain locus that comprises an intact mouse C λ gene. In one aspect, the intact C λ gene is C λ 2.

[0039] In one aspect, the mouse further comprises at least one immunoglobulin heavy chain allele. In one aspect, the at least one immunoglobulin heavy chain allele comprises a human V μ gene segment, a human D μ gene segment, and a human J μ gene segment at an endogenous mouse heavy chain locus that comprises a human heavy chain gene that expresses a human/mouse heavy chain. In a specific aspect, the mouse comprises two immunoglobulin heavy chain alleles, and the mouse expresses a human/mouse heavy chain.

[0040] In one aspect, the mouse comprises a first light chain allele that comprises an unrearranged hV κ and an unrearranged hJ κ , at an endogenous mouse κ locus that comprises an endogenous C κ gene; and a second light chain allele that comprises an unrearranged hV λ and an unrearranged hJ λ , at an endogenous mouse λ locus that comprises an intact endogenous C λ gene as recited in the claims. In a specific aspect, the first and the second light chain alleles are the only functional light chain alleles of the genetically modified mouse. In one aspect, the intact endogenous C λ gene is C λ 2.

[0041] In one aspect, the mouse comprises six immunoglobulin alleles, wherein the first allele comprises an unrearranged immunoglobulin V λ and J λ gene segment at an endogenous mouse κ light chain locus that comprises a mouse C κ gene, the second comprises an unrearranged immunoglobulin V κ and J κ gene segment at an endogenous mouse κ light chain locus that comprises a mouse C κ gene, the third comprises an unrearranged immunoglobulin V λ and J λ gene segment at an endogenous mouse λ light chain locus that comprises an intact mouse C λ gene as recited in the claims, the fourth and fifth each independently comprise an unrearranged V μ and D μ and J μ gene segment at an endogenous mouse heavy chain locus that comprises a mouse heavy chain gene, and the sixth comprises either (a) an unrearranged immunoglobulin V λ and J λ gene segment at an endogenous mouse λ light chain locus that comprises a mouse C λ gene, (b) a λ locus that is nonfunctional, or (c) a deletion in whole or in part of the λ locus.

[0042] In one aspect, the first allele comprises an unrearranged hV λ and hJ λ . In one aspect, the second allele comprises an unrearranged hV κ and hJ κ . The third allele comprises an unrearranged hV λ and hJ λ . In one aspect, the fourth and fifth each independently comprise an unrearranged hV μ and hD μ and hJ μ . In one aspect, the sixth allele comprises an endogenous mouse λ locus that is deleted in whole or in part.

[0043] In one aspect, the mouse comprises six immunoglobulin alleles, wherein the first allele comprises an unrearranged immunoglobulin V λ and J λ gene segment at an endogenous mouse λ light chain locus that comprises an intact mouse C λ gene, the second comprises an unrearranged immunoglobulin V λ and J λ gene segment at an endogenous mouse λ light chain locus that comprises a mouse C λ gene, the third comprises an unrearranged immunoglobulin V κ and J κ gene segment at an endogenous mouse κ light chain locus that comprises a mouse C κ gene, the fourth and fifth each independently comprise an unrearranged V μ and D μ and J μ gene segment at an endogenous mouse heavy chain locus that comprises a mouse heavy chain gene, and the sixth comprises either (a) an unrearranged immunoglobulin V κ and J κ gene segment at an endogenous mouse κ light chain locus that comprises a mouse C κ gene, (b) a κ locus that is nonfunctional, or (c) a deletion of one or more elements of the κ locus.

[0044] In one aspect, the first allele comprises an unrearranged hV λ , and hJ λ , gene segment. In one aspect, the second allele comprises an unrearranged hV λ and hJ λ gene segment. In one aspect, the third allele comprises an unrearranged hV κ and hJ κ gene segment. In one aspect, the fourth and fifth each independently comprise an unrearranged hV μ and hD μ and hJ μ gene segment. In one aspect, the sixth allele comprises an endogenous mouse κ locus that is functionally silenced.

[00445] In one aspect, the genetically modified mouse comprises a B cell that comprises a rearranged antibody gene comprising a rearranged hV λ domain operably linked to an intact mouse C λ domain. In a specific embodiment, the mouse C λ domain is derived from a C λ 2 gene.

[00446] Also disclosed is a genetically modified mouse that expresses an antibody comprising a light chain that comprises a rearranged human V λ -J λ sequence and a mouse C λ sequence

[00447] Also disclosed is a mouse that expresses an antibody comprising a light chain derived from a hV λ , a hJ λ and a mouse C λ gene. In a specific aspect, the C λ region is C λ 2. In a specific aspect the antibody further comprises a heavy chain comprising a variable domain derived from a human V, a human D and a human J gene segment, and a heavy chain constant domain derived from a mouse heavy chain constant region gene. In one aspect, the mouse heavy chain constant region gene comprises a hinge-CH₂-CH₃ sequence of a heavy chain constant domain. In another aspect, the mouse heavy chain constant region gene comprises a CH₁-hinge-CH₂-CH₃ sequence of a heavy chain constant domain. In another aspect, the mouse heavy chain constant region gene comprises a CH₁-CH₂-CH₃-CH₄ sequence of a heavy chain constant domain. In another aspect, the mouse heavy chain constant region gene comprises a CH₂-CH₃-CH₄ sequence of a heavy chain constant domain.

[00448] In one aspect, the mouse expresses an antibody comprising a light chain that comprises a rearranged immunoglobulin λ light chain variable region comprising a human V λ /J λ sequence selected from 3-1/1, 3-1/7, 4-3/1, 4-3/7, 2-8/1, 3-9/1, 3-10/1, 3-10/3, 3-10/7, 2-14/1, 3-19/1, 2-23/1, 3-25/1, 1-40/1, 1-40/2, 1-40/3, 1-40/7, 7-43/1, 7-43/3, 1-44/1, 1-44/7, 5-45/1, 5-45/2, 5-45/7, 7-46/1, 7-46/2, 7-46/7, 9-49/1, 9-49/2, 9-49/7 and 1-51/1.

[00449] Also disclosed is a mouse that expresses an antibody comprising (a) a heavy chain comprising a heavy chain variable domain derived from an unrearranged human heavy chain variable region gene segment, wherein the heavy chain variable domain is fused to a mouse heavy chain constant (C_H) region; and, (b) a light chain comprising a light chain variable domain derived from an unrearranged hV λ and a hJ λ wherein the light chain variable domain is fused to an intact mouse C λ region.

[00500] In one aspect, the mouse comprises (i) a heavy chain locus that comprises a replacement of all or substantially all functional endogenous mouse V, D and J gene segments with all or substantially all functional human V, D, and J gene segments, a mouse C_H gene, (ii) a first λ light chain locus comprising a replacement of all or substantially all functional endogenous mouse V λ and J λ gene segments with all, substantially all, or a plurality of, functional hV λ and hJ λ gene segments, and an intact mouse C λ gene, (iii) a second λ light chain locus comprising a replacement of all or substantially all functional endogenous mouse V λ and J λ gene segments with all, substantially all, or a plurality of, functional hV λ and hJ λ gene segments, and an intact mouse C λ gene. In a specific aspect, the intact mouse C λ gene is C λ 2.

[00501] In one aspect, the mouse comprises a deletion of a C κ gene and/or a V κ and/or a J κ gene segment. In one embodiment, the mouse comprises a nonfunctional κ light chain locus.

[00502] Also disclosed is a genetically modified mouse that expresses an antibody, wherein greater than 10%, greater than 15%, greater than 20%, greater than 25%, greater than 30%, greater than 35%, greater than 40%, greater than 60%, greater than 70%, greater than 80%, or greater than 90% of total IgG antibody produced by the mouse comprises a λ -derived variable domain, and wherein the mouse expresses antibodies comprising a κ -derived variable domain fused with a mouse C κ region. In specific aspects, about 15-40%, 20-40%, 25-40%, 30-40%, or 35-40% of total antibody produced by the mouse comprises a λ -derived variable domain.

[00503] In one aspect, the λ -derived variable domain is derived from a hV λ and a hJ λ . In a specific aspect, the λ -derived variable region is in a light chain that comprises a mouse C λ region. In another specific aspect, the C λ region is a C λ 2 region. Also disclosed is an isolated DNA construct that comprises an upstream homology arm and a downstream homology arm, wherein the upstream and the downstream homology arms target the construct to a mouse κ locus, and the construct comprises a functional unrearranged hV λ segment and a functional unrearranged hJ λ segment, and a selection or marker sequence.

[00504] Also disclosed is an isolated DNA construct, comprising, from 5' to 3' with respect to the direction of transcription, a targeting arm for targeting a mouse λ sequence upstream of mouse V λ 2, a selection cassette flanked 5' and 3' with recombinae recognition sites, and a targeting arm for targeting a mouse λ sequence 3' of mouse J λ 2. In one aspect, the selection cassette is a Frt^red Hyg-TK cassette. In one aspect, the 3' targeting arm comprises mouse C λ 2, J λ 4, C λ 4, and mouse enhancer 2.4.

[00505] Also disclosed is an isolated DNA construct, comprising, from 5' to 3' with respect to the direction of transcription, a targeting arm for targeting the mouse λ locus 5' with respect to V λ 1, a selection cassette flanked 5' and 3' with recombinae recognition sites, and a 3' targeting arm for targeting a mouse λ sequence 3' with respect to mouse C λ 1. In one aspect, the selection cassette is a loxed neomycin cassette. In one aspect, the 3' targeting arm comprises the mouse λ 3' enhancer and mouse λ 3' enhancer 3.1.

[00506] Also disclosed is an isolated DNA construct, comprising from 5' to 3' with respect to the direction of transcription, a targeting arm for targeting the mouse λ locus 5' with respect to V λ 2, a selection cassette flanked 5' and 3' with recombinae recognition sites, and a 3' targeting arm for targeting a mouse λ sequence 3' with respect to mouse J λ 2

and 5' with respect to mouse Cλ2. In one aspect, the selection cassette is a Frt⁺ hygromycin-TK cassette. In one aspect, the 3' targeting arm comprises the mouse Cλ2-Jλ4-Cλ4 gene segments and mouse λ enhancer 2.4.

[0057] Also disclosed is an isolated DNA construct, comprising, from 5' to 3' with respect to the direction of transcription, a targeting arm for targeting the mouse λ locus 5' with respect to V λ 2, a selection cassette flanked 5' and 3' with recombinaase recognition sites, a human genomic fragment comprising a contiguous region of the human λ light chain locus from hV λ 3-12 downstream to the end of hJ λ 1, and a 3' targeting arm for targeting a mouse λ sequence 3' with respect to mouse J λ 2. In one aspect, the selection cassette is a Frt^red neomycin cassette. In one aspect, the 3' targeting arm comprises the mouse C λ 2-J λ 4-C λ 4 gene segments and mouse λ enhancer 2.4.

[0058] Also disclosed is an isolated DNA construct, comprising a contiguous region of the human λ light chain locus from hV λ 3-12 downstream to the end of hJ λ 1.

[0059] Also disclosed is an isolated DNA construct, comprising, from 5' to 3' with respect to the direction of transcription, a targeting arm for targeting the mouse λ locus 5' with respect to $V\lambda 2$, a selection cassette flanked 5' and 3' with recombinase recognition sites and a human genomic fragment comprising a contiguous region of the human λ light chain locus from hV λ 3-27 downstream to the end of hV λ 2-8. In one aspect, the selection cassette is a Frt^red hygromycin cassette. In one aspect, the human genomic fragment comprises a 3' targeting arm. In a specific aspect, the 3' targeting arm comprises about 53 kb of the human λ light chain locus from hV λ 3-12 downstream to the end of hV λ 2-8.

[0060] Also disclosed is an isolated DNA construct, comprising a contiguous region of the human λ light chain locus from hV λ 3-27 downstream to the end of hV λ 3-12.

[0061] Also disclosed is an isolated DNA construct, comprising, from 5' to 3' with respect to the direction of transcription, a targeting arm for targeting the mouse λ locus 5' with respect to V λ 2, a selection cassette flanked 5' and 3' with recombinase recognition sites, a first human genomic fragment comprising a contiguous region of the human λ light chain locus from hV λ 5-52 downstream to the end of hV λ 1-40, a restriction enzyme site, and a second human genomic fragment comprising a contiguous region of the human λ light chain locus from hV λ 3-29 downstream to the end of hV λ 82K. In one aspect, the selection cassette is a Frt^{ed} neomycin cassette. In one aspect, the restriction enzyme site is a site for a homing endonuclease. In a specific aspect, the homing endonuclease is PI-SceI. In one aspect, the second human genomic fragment is a 3' targeting arm. In a specific aspect, the 3' targeting arm comprises about 27 kb of the human λ light chain locus from hV λ 3-29 downstream to the end of hV λ 82K.

[0062] Also disclosed is an isolated DNA construct, comprising a contiguous region of the human λ light chain locus from hV λ 5-52 downstream to the end of hV λ 1-40.

[0063] Also disclosed is an isolated DNA construct, comprising, from 5' to 3' with respect to the direction of transcription, a targeting arm for targeting the mouse κ locus 5' with respect to the endogenous V κ gene segments, two juxtaposed recombinae recognition sites, a selection cassette 3' to the juxtaposed recombinae recognition sites, and a 3' targeting arm for targeting a mouse κ sequence 5' with respect to the κ light chain variable gene segments. In one aspect, the juxtaposed recombinae recognition sites are in opposite orientation with respect to one another. In a specific aspect, the recombinae recognition sites are different. In another specific aspect, the recombinae recognition sites are a *loxP* site and a *lox511* site. In one aspect, the selection cassette is a neomycin cassette.

[0064] Also disclosed is an isolated DNA construct, comprising, from 5' to 3' with respect to the direction of transcription, a targeting arm for targeting the mouse κ locus 5' with respect to the mouse J κ gene segments, a selection cassette, a recombinase recognition site 3' to the selection cassette, and a 3' targeting arm for targeting a mouse κ sequence 3' with respect to the mouse J κ gene segments and 5' to the mouse κ intronic enhancer. In one aspect, the selection cassette is a hygromycin-TK cassette. In one aspect, the recombinase recognition site is in the same direction with respect to transcription as the selection cassette. In a specific aspect, the recombinase recognition site is a *loxP* site.

[0065] Also disclosed is an isolated DNA construct, comprising, from 5' to 3' with respect to the direction of transcription, a first mouse genomic fragment comprising sequence 5' of the endogenous mouse V κ gene segments, a first recombina-
 se recognition site, a second recombina-
 se recognition site, and a second mouse genomic fragment comprising sequence
 3' of the endogenous mouse J κ gene segments and 5' of the mouse κ intronic enhancer.

[0066] Also disclosed is a genetically modified mouse, wherein the genetic modification comprises a modification with one or more of the DNA constructs described above or herein.

[0067] Also disclosed is the use of an isolated DNA construct to make a mouse as described herein. Also disclosed is use of an isolated DNA construct as described herein in a method for making an antigen-binding protein.

[0068] Also disclosed is a non-human stem cell that comprises a targeting vector that comprises a DNA construct as described above and herein. Also disclosed is a non-human stem cell, wherein the non-human stem cell is derived from a mouse described herein.

[0069] In one aspect, the non-human stem cell is an embryonic stem (ES) cell. In a specific aspect, the ES cell is a mouse ES cell.

[0070] Also disclosed is the use of a non-human stem cell as described herein to make a mouse as described herein.

[0071] Also disclosed is a mouse embryo, wherein the mouse embryo comprises a genetic modification as disclosed

herein. In one aspect, a host mouse embryo that comprises a donor ES cell is disclosed, wherein the donor ES cell comprises a genetic modification as described herein. In one aspect, the mouse embryo is a pre-morula stage embryo. In a specific aspect, the pre-morula stage embryo is a 4-cell stage embryo or an 8-cell stage embryo. In another specific aspect, the mouse embryo is a blastocyst.

[0072] Also disclosed is the use of a mouse embryo as described herein to make a mouse as described herein. In one aspect, use of a mouse embryo as described herein to make an antigen-binding protein is disclosed.

[0073] Also disclosed is a non-human cell, wherein the non-human cell comprises a rearranged immunoglobulin light chain gene sequence derived from a genetically modified mouse as described herein. In one aspect, the cell is a B cell. In one aspect, the cell is a hybridoma. In one aspect, the cell encodes an immunoglobulin light chain variable domain and/or an immunoglobulin heavy chain variable domain that is somatically mutated.

[0074] Also disclosed is a non-human cell, wherein the non-human cell comprises a rearranged immunoglobulin light chain gene sequence derived from a genetically modified mouse as described herein. In one aspect, the cell is a B cell. In one aspect, the cell is a hybridoma. In one aspect, the cell encodes an immunoglobulin light chain variable domain and/or an immunoglobulin heavy chain variable domain that is somatically mutated.

[0075] Also disclosed is the use of a non-human cell as described herein to make a mouse as described herein. In one aspect, use of a non-human cell as described herein to make an antigen-binding protein is disclosed.

[0076] Also disclosed is a mouse B cell that expresses an immunoglobulin light chain that comprises (a) a variable region derived from a hV λ gene segment and a hJ λ gene segment; and, (b) a mouse C λ gene. In a specific aspect, the C λ gene is C λ 2. In one aspect, the mouse B cell further expresses a cognate heavy chain that comprises (c) a variable region derived from a hV μ , a hD μ , and (d) a hJ μ segment. In one aspect, the B cell does not comprise a rearranged λ gene. In another aspect, the B cell does not comprise a rearranged κ gene.

[0077] Also disclosed is a method for making an antibody in a genetically modified mouse, comprising: (a) exposing a genetically modified mouse to an antigen, wherein the mouse has a genome comprising at least one hV λ , and at least one hJ λ at an endogenous light chain locus, wherein the endogenous light chain locus comprises a mouse C λ gene; (b) allowing the genetically modified mouse to develop an immune response to the antigen; and, (c) isolating from the mouse of (b) an antibody that specifically recognizes the antigen, or isolating from the mouse of (b) a cell comprising an immunoglobulin domain that specifically recognizes the antigen, wherein the antibody comprises a light chain derived from a hV λ , a hJ λ and a mouse C λ gene.

[0078] [Deleted]

[0079] [Deleted]

[0080] In one aspect, a method for making an antibody in a genetically modified mouse is disclosed, comprising: (a) exposing a genetically modified mouse to an antigen, wherein the mouse has a genome comprising at least one hV λ at a λ light chain locus and at least one J λ at the λ light chain locus, wherein the λ light chain locus comprises an intact mouse C λ gene; (b) allowing the genetically modified mouse to develop an immune response to the antigen; and, (c) isolating from the mouse of (b) an antibody that specifically recognizes the antigen, or isolating from the mouse of (b) a cell comprising an immunoglobulin domain that specifically recognizes the antigen, or identifying in the mouse of B a nucleic acid sequence encoding a heavy and/or light chain variable domain that binds the antigen, wherein the antibody comprises a light chain derived from a hV λ , a hJ λ and a mouse C λ gene.

[0081] The λ light chain constant gene is an intact mouse C λ gene. In a specific aspect, the intact mouse C λ gene is selected from C λ 1, C λ 2 and C λ 3. In a more specific aspect, the mouse C λ gene is C λ 2.

[0082] Also disclosed is a method for making a rearranged antibody gene in a genetically modified mouse, comprising: (a) exposing a genetically modified mouse to an antigen, wherein the genetic modification comprises a hV λ and a hJ λ , at an endogenous light chain locus, wherein the endogenous light chain locus comprises an intact mouse C λ gene; and, (b) identifying a rearranged immunoglobulin gene in said mouse, wherein the rearranged immunoglobulin gene comprises a λ light chain variable region gene segment and a C λ gene or functional fragment thereof.

[0083] In one aspect, the method further comprises cloning a nucleic acid sequence encoding a heavy and/or light chain variable region from the mouse, wherein the heavy and/or light chain variable region is from an antibody that comprises a human V λ and a mouse C λ .

[0084] [Deleted]

[0085] [Deleted]

[0086] [Deleted]

[0087] [Deleted]

[0088] In one aspect, a method for making a rearranged antibody gene in a genetically modified mouse is disclosed, comprising: (a) exposing a genetically modified mouse to an antigen, wherein the genetic modification comprises a hV λ and a hJ λ , at a mouse λ light chain locus, wherein the λ light chain locus comprises an intact mouse C λ gene; and, (b) identifying a rearranged immunoglobulin gene in said mouse, wherein the rearranged immunoglobulin gene comprises a λ light chain variable region gene segment and a C λ gene.

[0089] The λ light chain constant gene is an intact mouse C λ gene.

[0090] In one aspect, the method further comprises cloning a nucleic acid sequence encoding a heavy and/or light chain variable region from the mouse, wherein the heavy and/or light chain variable region is from an antibody that comprises a human V λ and a mouse C λ .

[0091] Also disclosed is a method for making an antibody, comprising exposing a mouse as described herein to an antigen, allowing the mouse to mount an immune response that comprises making an antibody that specifically binds the antigen, identifying a rearranged nucleic acid sequence in the mouse that encodes heavy chain and a rearranged nucleic acid sequence in the mouse that encodes a cognate light chain variable domain sequence of an antibody, wherein the antibody specifically binds the antigen, and employing the nucleic acid sequences of the heavy and light chain variable domains fused to human constant domains to make a desired antibody, wherein the desired antibody comprises a light chain that comprises a V λ domain fused to a C λ domain. The V λ domain is human and the C λ domain is a mouse C λ domain.

[0092] [Deleted]

[0093] In one aspect, a method for making an antibody is disclosed, comprising exposing a mouse as described herein to an antigen, allowing the mouse to mount an immune response that comprises making an antibody that specifically binds the antigen, identifying a rearranged nucleic acid sequence in the mouse that encodes a heavy chain variable domain and a rearranged nucleic acid sequence that encodes a cognate light chain variable domain sequence of an antibody, wherein the antibody specifically binds the antigen, and employing the nucleic acid sequences fused to nucleic acid sequences that encode a human heavy chain constant domain and a human light chain constant domain to make an antibody derived from human sequences, wherein the antibody that specifically binds the antigen comprises a light chain that comprises a human V λ domain fused to a mouse C λ region.

[0094] In one aspect, the mouse C λ region is selected from C λ 1, C λ 2 and C λ 3. In a specific aspect, the mouse C λ region is C λ 2.

[0095] In one aspect, a method for making a rearranged antibody light chain variable region gene sequence is disclosed, comprising (a) exposing a mouse as described herein to an antigen; (b) allowing the mouse to mount an immune response; (c) identifying a cell in the mouse that comprises a nucleic acid sequence that encodes a rearranged human V λ domain sequence fused with a mouse C λ domain, wherein the cell also encodes a cognate heavy chain comprising a human V H domain and a mouse C H domain, and wherein the cell expresses an antibody that binds the antigen; (d) cloning from the cell a nucleic acid sequence encoding the human V λ domain and a nucleic acid sequence encoding the cognate human V H domain; and, (e) using the cloned nucleic acid sequence encoding the human V λ domain and the cloned nucleic acid sequence encoding the cognate human V H domain to make a fully human antibody.

[0096] [Deleted]

[0097] In one aspect, a method for making a rearranged antibody light chain variable region gene sequence is disclosed, comprising (a) exposing a mouse as described herein to an antigen; (b) allowing the mouse to mount an immune response to the antigen; (c) identifying a cell in the mouse that comprises DNA that encodes a rearranged human V λ domain sequence fused with a mouse C λ domain, wherein the cell also encodes a cognate heavy chain comprising a human V H domain and a mouse C H domain, and wherein the cell expresses an antibody that binds the antigen; (d) cloning from the cell a nucleic acid sequence encoding the rearranged human V λ domain and a nucleic acid sequence encoding the cognate human V H domain; and, (e) using the cloned nucleic acid sequence encoding the human V λ domain and the cloned nucleic acid sequence encoding the cognate human V H domain to make a fully human antibody. In one aspect, the mouse C λ domain is mouse C λ 2.

[0098] Also disclosed is a genetically modified mouse that expresses a human λ -derived light chain fused to an endogenous light chain constant region (C λ), wherein the mouse, upon immunization with antigen, makes an antibody comprising a human V λ domain fused to a mouse C λ domain. In a specific aspect, the C λ domain is C λ 2.

[0099] Also disclosed in a genetically modified mouse comprising a modified endogenous λ light chain locus as described herein, that expresses a plurality of immunoglobulin λ light chains associated with a plurality of immunoglobulin heavy chains. In one aspect, the heavy chain comprises a human sequence. In various aspects, the human sequence is selected from a variable sequence, a C H 1, a hinge, a C H 2, a C H 3, and a combination thereof. In one aspect, the plurality of immunoglobulin λ light chains comprises a human sequence. In various aspects, the human sequence is selected from a variable sequence, a constant sequence, and a combination thereof. In one aspect, the mouse comprises a disabled endogenous immunoglobulin locus and expresses the heavy chain from a transgene or extrachromosomal episome. In one aspect, the mouse comprises a replacement at an endogenous mouse locus of some or all endogenous mouse heavy chain gene segments (*i.e.*, V, D, J), and/or some or all endogenous mouse heavy chain constant sequences (*e.g.*, C H 1, hinge, C H 2, C H 3, or a combination thereof), and/or some or all endogenous mouse light chain sequences (*e.g.*, V, J, constant, or a combination thereof), with one or more human immunoglobulin sequences.

[0100] Also disclosed is a mouse suitable for making antibodies that have a human λ -derived light chain wherein all or substantially all antibodies made in the mouse are expressed with a human λ -derived light chain. In one aspect, the human λ -derived light chain is expressed from an endogenous light chain locus.

[0101] Also disclosed is a method for making a λ -derived light chain for a human antibody, comprising obtaining from

a mouse as described herein a light chain sequence and a heavy chain sequence, and employing the light chain sequence and the heavy chain sequence in making a human antibody.

[0102] Also disclosed is, a method for making an antigen-binding protein, comprising exposing a mouse as described herein to an antigen; allowing the mouse to mount an immune response; and obtaining from the mouse an antigen-binding protein that binds the antigen, or obtaining from the mouse a sequence to be employed in making an antigen-binding protein that binds the antigen.

[0103] Also disclosed is a cell derived from a mouse as described herein. In one aspect, the cell is selected from an embryonic stem cell, a pluripotent cell, an induced pluripotent cell, a B cell, and a hybridoma.

[0104] Also disclosed is, a cell that comprises a genetic modification as described herein. In one aspect, the cell is a mouse cell. In one aspect, the cell is selected from a hybridoma and a quadroma. In one aspect, the cell expresses an immunoglobulin light chain that comprises a human λ variable sequence fused with a mouse λ constant sequence.

[0105] Also disclosed is, a tissue derived from a mouse as described herein.

[0106] Also disclosed is, use of a mouse or a cell as described herein to make an antigen-binding protein. In one aspect, the antigen-binding protein is a human protein. In one aspect, the human protein is a human antibody.

[0107] Also disclosed is an antigen-binding protein made by a mouse, cell, tissue, or method as described herein. In one aspect, the antigen-binding protein is a human protein. In one aspect, the human protein is a human antibody.

[0108] [Deleted]

BRIEF DESCRIPTION OF THE FIGURES

[0109]

FIG. 1 shows a detailed illustration, not to scale, of the human λ light chain locus including the clusters of $V\lambda$ gene segments (A, B and C) and the $J\lambda$ and $C\lambda$ region pairs (J-C pairs)

FIG. 2 shows a general illustration, not to scale, of a targeting strategy used to inactivate the endogenous mouse λ light chain locus.

FIG. 3 shows a general illustration, not to scale, of a targeting strategy used to inactivate the endogenous mouse κ light chain locus.

FIG. 4A shows a general illustration, not to scale of an initial targeting vector for targeting the endogenous mouse λ light chain locus with human λ light chain sequences including 12 $hV\lambda$ gene segments and $hJ\lambda 1$ gene segment (12/1- λ Targeting Vector).

FIG. 4B shows a general illustration, not to scale, of four initial targeting vectors for targeting the endogenous mouse κ light chain locus with human λ light chain sequences including 12 $hV\lambda$, gene segments and $hJ\lambda 1$ gene segment (12/1- κ Targeting Vector), 12 $hV\lambda$, gene segments and $hJ\lambda 1$, 2, 3 and 7 gene segments (12/4- κ Targeting Vector), 12 $hV\lambda$ gene segments, a human $V\kappa$ - $J\kappa$ genomic sequence and $hJ\lambda 1$ gene segment (12(κ)1- κ Targeting Vector) and 12 $hV\lambda$ gene segments, a human $V\kappa$ - $J\kappa$ genomic sequence and $hJ\lambda 1$, 2, 3 and 7 gene segments (12(κ)4- κ Targeting Vector).

FIG. 5A shows a general illustration, not to scale, of a targeting strategy for progressive insertion of 40 $hV\lambda$ gene segments and a single $hJ\lambda$ gene segment into the mouse λ light chain locus.

FIG. 5B shows a general illustration, not to scale, of a targeting strategy for progressive insertion of 40 $hV\lambda$ gene segments and a single $hJ\lambda$ gene segment into the mouse κ locus.

FIG. 6 show a general illustration, not to scale, of the targeting and molecular engineering steps employed to make unique human λ - κ hybrid targeting vectors for construction of a hybrid light chain locus containing a human κ intergenic sequence, multiple $hJ\lambda$, gene segments or both.

FIG. 7A shows a general illustration, not to scale, of the locus structure for a modified mouse λ light chain locus containing 40 $hV\lambda$ gene segments and a single $hJ\lambda$ gene segment operably linked to the endogenous $C\lambda 2$ gene.

FIG. 7B shows a general illustration, not to scale, of the locus structure for four independent, modified mouse κ light chain loci containing 40 $hV\lambda$ gene segments and either one or four $hJ\lambda$ gene segments with or without a contiguous human $V\kappa$ - $J\kappa$ genomic sequence operably linked to the endogenous $C\kappa$ gene.

FIG. 8A shows contour plots of $Ig\lambda^+$ and $Ig\kappa^+$ splenocytes gated on $CD19^+$ from a wild type mouse (WT), a mouse homozygous for 12 $hV\lambda$ and four $hJ\lambda$ gene segments including a human $V\kappa$ - $J\kappa$ genomic sequence (12 $hV\lambda$ - $V\kappa$ $J\kappa$ -4 $hJ\lambda$) and a mouse homozygous for 40 $hV\lambda$ and one $hJ\lambda$ gene segment (40 $hV\lambda$ -1 $hJ\lambda$).

FIG. 8B shows the total number of $CD19^+$ B cells in harvested spleens from wild type (WT), mice homozygous for 12 $hV\lambda$ and four $hJ\lambda$ gene segments including a human $V\kappa$ - $J\kappa$ genomic sequence (12 $hV\lambda$ - $V\kappa$ $J\kappa$ -4 $hJ\lambda$) and mice homozygous for 40 $hV\lambda$ and one $hJ\lambda$ gene segment (40 $hV\lambda$ -1 $hJ\lambda$).

FIG. 9A, in the top panel, shows contour plots of splenocytes gated on singlets and stained for B and T cells ($CD19^+$ and $CD3^+$, respectively) from a wild type mouse (WT) and a mouse homozygous for 40 $hV\lambda$ and four $J\lambda$ gene segments including a human $V\kappa$ - $J\kappa$ genomic sequence (40 $hV\lambda$ - $V\kappa$ $J\kappa$ -4 $hJ\lambda$). The bottom panel shows contour plots

of splenocytes gated on CD19⁺ and stained for Igλ⁺ and Igκ⁺ expression from a wild type mouse (WT) and a mouse homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ). **FIG. 9B** shows the total number of CD19⁺, CD19⁺Igκ⁺ and CD19⁺Igλ⁺ B cells in harvested spleens from wild type mice (WT) and mice homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ).

FIG. 9C shows contour plots of splenocytes gated on CD19⁺ and stained for immunoglobulin D (IgD) and immunoglobulin M (IgM) from a wild type mouse (WT) and a mouse homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ). Mature (72 for WT, 51 for 40hVλ-VκJκ-4hJλ) and transitional (13 for WT, 22 for 40hVλ-VκJκ-4hJλ) B cells are noted on each of the contour plots.

FIG. 9D shows the total number of CD19⁺ B cells, transitional B cells (CD19⁺IgM^{hi}IgD^{lo}) and mature B cells (CD19⁺IgM^{lo}IgD^{hi}) in harvested spleens from wild type mice (WT) and mice homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ).

FIG. 10A, in the top panel, shows contour plots of bone marrow stained for B and T cells (CD19⁺ and CD3⁺, respectively) from a wild type mouse (WT) and a mouse homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ). The bottom panel shows contour plots of bone marrow gated on CD19⁺ and stained for ckit⁺ and CD43⁺ from a wild type mouse (WT) and a mouse homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ). Pro and Pre B cells are noted on the contour plots of the bottom panel.

FIG. 10B shows the number of Pro (CD19⁺CD43⁺ckit⁺) and Pre (CD19⁺CD43⁻ckit⁺) B cells in bone marrow harvested from the femurs of wild type mice (WT) and mice homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ).

FIG. 10C shows contour plots of bone marrow gated on singlets stained for immunoglobulin M (IgM) and B220 from a wild type mouse (WT) and a mouse homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ). Immature, mature and pro/pre B cells are noted on each of the contour plots.

FIG. 10D shows the total number of immature (B220^{int}IgM⁺) and mature (B220^{hi}IgM⁺) B cells in bone marrow isolated from the femurs of wild type mice (WT) and mice homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ).

FIG. 10E shows contour plots of bone marrow gated on immature (B220^{int}IgM⁺) and mature (B220^{hi}IgM⁺) B cells stained for Igλ and Igκ expression isolated from the femurs of a wild type mouse (WT) and a mouse homozygous for 40 hVλ and four Jλ gene segments including a human Vκ-Jκ genomic sequence (40hVλ-VκJκ-4hJλ).

FIG. 11 shows a nucleotide sequence alignment of the Vλ-Jλ-Cκ junction of eighteen independent RT-PCR clones amplified from splenocyte RNA of mice bearing human λ light chain gene sequences at an endogenous mouse κ light chain locus. A6 = SEQ ID NO:57; B6 = SEQ ID NO:58; F6 = SEQ ID NO:59; B7 = SEQ ID NO:60; E7 = SEQ ID NO:61; F7 = SEQ ID NO:62; C8 = SEQ ID NO:63; E12 = SEQ ID NO:64; 1-4 = SEQ ID NO:65; 1-20 = SEQ ID NO:66; 3B43 = SEQ ID NO:67; 5-8 = SEQ ID NO:68; 5-19 = SEQ ID NO:69; 1010 = SEQ ID NO:70; 11A1 = SEQ ID NO:71; 7A8 = SEQ ID NO:72; 3A3 = SEQ ID NO:73; 2-7 = SEQ ID NO:74. Lower case bases indicate non-germline bases resulting from either mutation and/or N addition during recombination. Consensus amino acids within the Framework 4 region (FWR4) encoded by the nucleotide sequence of hJλ1 and mouse Cκ are noted at the bottom of the sequence alignment.

FIG. 12 shows a nucleotide sequence alignment of the Vλ-Jλ-Cκ junction of twelve independent RT-PCR clones amplified from splenocyte RNA of mice bearing human λ light chain gene sequences including a contiguous human Vκ-Jκ genomic sequence at an endogenous mouse κ light chain locus. 5-2 = SEQ ID NO:87; 2-5 = SEQ ID NO:88; 1-3 = SEQ ID NO:89; 4B-1 = SEQ ID NO:90; 3B-5 = SEQ ID NO:91; 7A-1 = SEQ ID NO:92; 5-1 = SEQ ID NO:93; 4A-1 = SEQ ID NO:94; 11A-1 = SEQ ID NO:95; 5-7 = SEQ ID NO:96; 5-4 = SEQ ID NO:97; 2-3 = SEQ ID NO:98. Lower case bases indicate non-germline bases resulting from either mutation and/or N addition during recombination. Consensus amino acids within the Framework 4 region (FWR4) encoded by the nucleotide sequence of each human Jλ and mouse Cκ are noted at the bottom of the sequence alignment.

FIG. 13 shows a nucleotide sequence alignment of the Vλ-Jλ-Cλ junction of three independent RT-PCR clones amplified from splenocyte RNA of mice bearing human λ light chain gene sequences at an endogenous mouse λ light chain locus. 2D1 = SEQ ID NO:101; 209 = SEQ ID NO:102; 3E15 = SEQ ID NO:103. Lower case bases indicate non-germline bases resulting from either mutation and/or N addition during recombination. Consensus amino acids within the Framework 4 region (FWR4) encoded by the nucleotide sequence of hJλ1 and mouse Cλ2 are noted at the bottom of the sequence alignment.

DETAILED DESCRIPTION

[0110] The invention in its broadest sense is as defined in the independent claims

[0111] The term "contiguous" includes reference to occurrence on the same nucleic acid molecule, *e.g.*, two nucleic acid sequences are "contiguous" if they occur on the same nucleic molecule but are interrupted by another nucleic acid sequence. For example, a rearranged V(D)J sequence is "contiguous" with a constant region gene sequence, although the final codon of the V(D)J sequence is not followed immediately by the first codon of the constant region sequence.

5 In another example, two V gene segment sequences are "contiguous" if they occur on the same genomic fragment, although they may be separated by sequence that does not encode a codon of the V region, *e.g.*, they may be separated by a regulatory sequence, *e.g.*, a promoter or other noncoding sequence. In one embodiment, a contiguous sequence includes a genomic fragment that contains genomic sequences arranged as found in a wild-type genome.

[0112] The phrase "derived from" when used concerning a variable region "derived from" a cited gene or gene segment includes the ability to trace the sequence back to a particular unrearranged gene segment or gene segments that were rearranged to form a gene that expresses the variable domain (accounting for, where applicable, splice differences and somatic mutations).

[0113] The phrase "functional" when used concerning a variable region gene segment or joining gene segment refers to usage in an expressed antibody repertoire; *e.g.*, in humans V λ gene segments 3-1, 4-3, 2-8, *etc.* are functional, whereas V λ gene segments 3-2, 3-4, 2-5, *etc.* are nonfunctional.

[0114] A "heavy chain locus" includes a location on a chromosome, *e.g.*, a mouse chromosome, wherein in a wild-type mouse heavy chain variable (V_H), heavy chain diversity (D_H), heavy chain joining (J_H), and heavy chain constant (C_H) region DNA sequences are found.

[0115] A " κ locus" includes a location on a chromosome, *e.g.*, a mouse chromosome, wherein in a wild-type mouse κ variable (V κ), κ joining (J κ), and κ constant (C κ) region DNA sequences are found.

[0116] A " λ locus" includes a location on a chromosome, *e.g.*, a mouse chromosome, wherein in a wild-type mouse λ variable (V λ), λ joining (J λ), and λ constant (C λ) region DNA sequences are found.

[0117] The term "unrearranged" includes the state of an immunoglobulin locus wherein V gene segments and J gene segments (for heavy chains, D gene segments as well) are maintained separately but are capable of being joined to form a rearranged V(D)J gene that comprises a single V(D),J of the V(D)J repertoire.

Mice Expressing Human λ Variable Domains

[0118] Mice that express antibodies that are fully human, or partly human and partly mouse, have previously been reported. VELOCIMMUNE® genetically engineered mice comprise a replacement of unrearranged V(D)J gene segments at endogenous mouse loci with human V(D)J gene segments. VELOCIMMUNE® mice express chimeric antibodies having human variable domains and mouse constant domains (see, *e.g.*, US Pat. No. 7,605,237). Most other reports concern mice that express fully human antibodies from fully human transgenes in mice that have disabled endogenous immunoglobulin loci.

[0119] Antibody light chains are encoded by one of two separate loci: kappa (κ) and lambda (λ). Mouse antibody light chains are primarily of the κ type. Mice that make mouse antibodies, and modified mice that make fully human or chimeric human-mouse antibodies, display a bias in light chain usage. Humans also exhibit light chain bias, but not so pronounced as in mice; the ratio of κ light chains to λ light chains in mice is about 95:5, whereas in humans the ratio is about 60:40. The more pronounced bias in mice is not thought to severely affect antibody diversity, because in mice the λ variable locus is not so diverse in the first instance. This is not so in humans. The human λ light chain locus is richly diverse.

[0120] The human λ light chain locus extends over 1,000 kb and contains over 80 genes that encode variable (V) or joining (J) segments (FIG. 1). Within the human λ light chain locus, over half of all observed V λ domains are encoded by the gene segments 1-40, 1-44, 2-8, 2-14, and 3-21. Overall, about 30 or so of the human V λ gene segments are believed to be functional. There are seven J λ gene segments, only four of which are regarded as generally functional J λ gene segments, J λ 1, J λ 2, J λ 3, and J λ 7.

[0121] The λ light chain locus in humans is similar in structure to the κ locus of both mice and humans in that the human λ light chain locus has several variable region gene segments that are capable of recombining to form a functional light chain protein. The human λ light chain locus contains approximately 70 V gene segments and 7 J λ -C λ gene segment pairs. Only four of these J λ -C λ gene segment pairs appear to be functional. In some alleles, a fifth J λ -C λ gene segment pair is reportedly a pseudo gene (C λ 6). The 70 V λ gene segments appear to contain 38 functional gene segments. The 70 V λ sequences are arranged in three clusters, all of which contain different members of distinct V gene family groups (clusters A, B and C; FIG. 1). This is a potentially rich source of relatively untapped diversity for generating antibodies with human V regions in non-human animals.

[0122] In stark contrast, the mouse λ light chain locus contains only two or three (depending on the strain) mouse V λ region gene segments (FIG. 2). At least for this reason, the severe κ bias in mice is not thought to be particularly detrimental to total antibody diversity.

[0123] According published maps of the mouse λ light chain locus, the locus consists essentially of two clusters of gene segments within a span of approximately 200 kb (FIG.2). The two clusters contain two sets of V, J, and C genes

that are capable of independent rearrangement: $V\lambda 2-J\lambda 2-C\lambda 2-J\lambda 4-C\lambda 4$ and $V\lambda 1-J\lambda 3-C\lambda 3-J\lambda 1-C\lambda 1$. Although $V\lambda 2$ has been found to recombine with all $J\lambda$ gene segments, $V\lambda 1$ appears to exclusively recombine with $C\lambda 1$. $C\lambda 4$ is believed to be a pseudo gene with defective splice sites.

[0124] The mouse κ light chain locus is strikingly different. The structure and number of gene segments that participate in the recombination events leading to a functional light chain protein from the mouse κ locus is much more complex (FIG. 3). Thus, mouse λ light chains do not greatly contribute to the diversity of an antibody population in a typical mouse.

[0125] Exploiting the rich diversity of the human λ light chain locus in mice would likely result in, among other things, a source for a more complete human repertoire of light chain V domains. Previous attempts to tap this diversity used human transgenes containing chunks of the human λ light chain locus randomly incorporated into the mouse genome (see, e.g., US 6,998,514 and US 7,435,871). Mice containing these randomly integrated transgenes reportedly express fully human λ light chains, however, in some cases, one or both endogenous light chain loci remain intact. This situation is not desirable as the human λ light chain sequences contend with the mouse light chain (κ or λ) in the expressed antibody repertoire of the mouse.

[0126] In contrast, the inventors describe genetically modified mice that are capable of expressing one or more λ light chain nucleic acid sequences directly from an endogenous lambda mouse light chain locus. Genetically modified mice capable of expressing human λ light chain sequences from an endogenous λ locus may be further bred to mice that comprise a human heavy chain locus and thus be used to express antibodies comprising V regions (heavy and λ light) that are fully human. The V regions express with mouse constant regions. In various aspects, no endogenous mouse immunoglobulin gene segments are present and the V regions express with human constant regions. These antibodies would prove useful in numerous applications, both diagnostic as well as therapeutic.

[0127] Many advantages can be realized for various aspects of expressing binding proteins derived from human $V\lambda$ and $J\lambda$ gene segments in mice. Advantages can be realized by placing human λ sequences at the endogenous λ light chain locus. Antibodies made from such mice can have light chains that comprise human $V\lambda$ domains fused to a mouse mouse $C\lambda$ region. The mice will also express human $V\lambda$ domains that are suitable for identification and cloning for use with human C_L regions, specifically $C\kappa$ and/or $C\lambda$ regions. Because B cell development in such mice is otherwise normal, it is possible to generate compatible $V\lambda$ domains (including somatically mutated $V\lambda$ domains) in the context of either $C\lambda$ or $C\kappa$ regions.

[0128] Genetically modified mice are described that comprise an unrearranged human $V\lambda$ gene segment and an unrearranged human $J\lambda$ at an endogenous immunoglobulin λ light chain locus. Mice that express antibodies that comprise a light chain having a human $V\lambda$ domain fused to a $C\lambda$ region are described.

Approaches to Engineering Mice to Express Human $V\lambda$ Domains

[0129] Various approaches to making genetically modified mice that make antibodies that contain a light chain that has a human $V\lambda$ domain fused to an endogenous $C\lambda$ region are described. Genetic modifications are described that, in various aspects, comprise a deletion of one or both endogenous light chain loci. For example, to eliminate mouse λ light chains from the endogenous antibody repertoire a deletion of a first $V\lambda-J\lambda-C\lambda$ gene cluster and replacement, in whole or in part, of the $V\lambda-J\lambda$ gene segments of a second gene cluster with human $V\lambda-J\lambda$ gene segments can be made as recited in the claims. Genetically modified mouse embryos, cells, and targeting constructs for making the mice, mouse embryos, and cells are also disclosed.

[0130] The deletion of one endogenous $V\lambda-J\lambda-C\lambda$ gene cluster and replacement of the $V\lambda-J\lambda$ gene segments of another endogenous $V\lambda-J\lambda-C\lambda$ gene cluster employs a relatively minimal disruption in natural antibody constant region association and function in the animal, in various aspects, because endogenous $C\lambda$ genes are left intact and therefore retain normal functionality and capability to associate with the constant region of an endogenous heavy chain. Thus, in such aspects the modification does not affect other endogenous heavy chain constant regions dependent upon functional light chain constant regions for assembly of a functional antibody molecule containing two heavy chains and two light chains. Further, in various aspects the modification does not affect the assembly of a functional membrane-bound antibody molecule involving an endogenous heavy chain and a light chain, e.g., a h $V\lambda$ domain linked to a mouse $C\lambda$ region. Because at least one functional $C\lambda$ gene is retained at the endogenous locus, animals containing a replacement of the $V\lambda-J\lambda$ gene segments of an endogenous $V\lambda-J\lambda-C\lambda$ gene cluster with human $V\lambda-J\lambda$ gene segments should be able to make normal λ light chains that are capable of binding antigen during an immune response through the human $V\lambda-J\lambda$ gene segments present in the expressed antibody repertoire of the animal.

[0131] A schematic illustration (not to scale) of a deleted endogenous mouse $V\lambda-J\lambda-C\lambda$ gene cluster is provided in FIG. 2. As illustrated, the mouse λ light chain locus is organized into two gene clusters, both of which contain function gene segments capable of recombining to form a function mouse λ light chain. The endogenous mouse $V\lambda 1-J\lambda 3-C\lambda 3-J\lambda 1-C\lambda 1$ gene cluster is deleted by a targeting construct (Targeting Vector 1) with a neomycin cassette flanked by recombination sites. The other endogenous gene cluster ($V\lambda 2-V\lambda 3-J\lambda 2-C\lambda 2-J\lambda 4-C\lambda 4$) is deleted in part by a targeting construct (Targeting Vector 2) with a hygromycin-thymidine kinase cassette flanked by recombination sites. In this second

targeting event, the C λ 2-J λ 4-C λ 4 endogenous gene segments are retained. The second targeting construct (Targeting Vector 2) is constructed using recombination sites that are different than those in the first targeting construct (Targeting Vector 1) thereby allowing for the selective deletion of the selection cassette after a successful targeting has been achieved. The resulting double-targeted locus is functionally silenced in that no endogenous λ light chain can be produced.

5 This modified locus can be used for the insertion of human V λ and J λ gene segments to create an endogenous mouse λ locus comprising human V λ and J λ gene segments, whereby, upon recombination at the modified locus, the animal produces λ light chains comprising rearranged human V λ and J λ gene segments linked to an endogenous mouse C λ gene segment.

[0132] Genetically modifying a mouse to render endogenous λ gene segments nonfunctional, in various aspects, results in a mouse that exhibits exclusively κ light chains in its antibody repertoire, making the mouse useful for evaluating the role of λ light chains in the immune response, and useful for making an antibody repertoire comprising V κ domains but not V λ domains.

[0133] A genetically modified mouse that expresses a hV λ linked to a mouse C λ gene having been recombined at the endogenous mouse λ light chain locus can be made by any method known in the art. A schematic illustration (not to scale) of the replacement of the endogenous mouse V λ 2-V λ 3-J λ 2 gene segments with human V λ and J λ gene segments is provided in FIG. 4A. As illustrated, an endogenous mouse λ light chain locus that had been rendered nonfunctional is replaced by a targeting construct (12/1- λ Targeting Vector) that includes a neomycin cassette flanked by recombination sites. The V λ 2-V λ 3-J λ 2 gene segments are replaced with a genomic fragment containing human λ sequence that includes 12 hV λ gene segments and a single hJ λ gene segment.

[0134] Thus, this first approach positions one or more hV λ gene segments at the endogenous λ light chain locus contiguous with a single hJ λ gene segment (FIG. 4A).

[0135] Further modifications to the modified endogenous λ light chain locus can be achieved with using similar techniques to insert more hV λ gene segments. For example, schematic illustrations of two additional targeting constructs (+16- λ and +12- λ Targeting Vectors) used for progressive insertion of additional human hV λ gene segments are provided in FIG. 5A. As illustrated, additional genomic fragments containing specific human hV λ gene segments are inserted into the modified endogenous λ light chain locus in successive steps using homology provided by the previous insertion of human λ light chain sequences. Upon recombination with each targeting construct illustrated, in sequential fashion, 28 additional hV λ gene segments are inserted into the modified endogenous λ light chain locus. This creates a chimeric locus that produces a λ light chain protein that comprises human V λ -J λ gene segments linked to a mouse C λ gene.

[0136] The above approaches to insert human λ light chain gene segments at the mouse λ locus, maintains the enhancers positioned downstream of the C λ 2-J λ 4-C λ 4 gene segments (designated Enh 2.4, Enh and Enh 3.1 FIG. 4A and FIG. 5A). This approach results in a single modified allele at the endogenous mouse λ light chain locus (FIG. 7A).

[0137] Also disclosed are compositions and methods for making a mouse that expresses a light chain comprising hV λ and J λ gene segments operably linked to a mouse C λ gene segment, including compositions and method for making a mouse that expresses such genes from an endogenous mouse λ light chain locus. The methods include selectively rendering one endogenous mouse V λ -J λ -C λ gene cluster nonfunctional (e.g., by a targeted deletion), and employing a hV λ and J λ gene segments at the endogenous mouse λ light chain locus to express a hV λ domain in a mouse.

[0138] Also disclosed is a second approach where human λ light chain gene segments may be positioned at the endogenous κ light chain locus. The genetic modification, in various aspects, comprises a deletion of the endogenous κ light chain locus. For example, to eliminate mouse κ light chains from the endogenous antibody repertoire a deletion of the mouse V κ and J κ gene segments can be made. Genetically modified mouse embryos, cells, and targeting constructs for making the mice, mouse embryos, and cells are also disclosed.

[0139] For the reasons stated above, the deletion of the mouse V κ and J κ gene segments employs a relatively minimal disruption. A schematic illustration (not to scale) of deleted mouse V κ and J κ gene segments is provided in FIG. 3. The endogenous mouse V κ and J κ gene segments are deleted via recombinase-mediated deletion of mouse sequences position between two precisely positioned targeting vectors each employing site-specific recombination sites. A first targeting vector (J κ Targeting Vector) is employed in a first targeting event to delete the mouse J κ gene segments. A second targeting vector (V κ Targeting Vector) is employed in a second, sequential targeting event to delete a sequence located 5' of the most distal mouse V κ gene segment. Both targeting vectors contain site-specific recombination sites thereby allowing for the selective deletion of both selection cassettes and all intervening mouse κ light chain sequences after a successful targeting has been achieved. The resulting deleted locus is functionally silenced in that no endogenous κ light chain can be produced. This modified locus can be used for the insertion of hV λ and J λ gene segments to create an endogenous mouse κ locus comprising hV λ and J λ gene segments, whereby, upon recombination at the modified locus, the animal produces λ light chains comprising rearranged hV λ and J λ gene segments operably linked to an endogenous mouse C κ gene segment. Various targeting vectors comprising human λ light chain sequences can be used in conjunction with this deleted mouse κ locus to create a hybrid light chain locus containing human λ gene segments operably linked with a mouse C κ region.

[0140] Thus, a second approach positions one or more human V λ gene segments are positioned at the mouse κ light

chain locus contiguous with a single human J λ gene segment (12/1- κ Targeting Vector, FIG. 4B).

[0141] Modifications to this approach can be made to add gene segments and/or regulatory sequences to optimize the usage of the human λ light chain sequences from the mouse κ locus within the mouse antibody repertoire.

[0142] Also disclosed is a third approach where, one or more hV λ gene segments are positioned at the mouse κ light chain locus contiguous with four hJ λ gene sequences (12/4- κ Targeting Vector FIG. 4B).

[0143] In a third approach, one or more hV λ gene segments are positioned at the mouse κ light chain locus contiguous with a human κ intergenic sequence and a single hJ λ gene sequence (12(κ)1- κ Targeting Vector, FIG. 4B).

[0144] Also disclosed is a fourth approach, one or more hV λ gene segments are positioned at the mouse κ light chain locus contiguous with a human κ intergenic sequence four hJ λ gene sequences (12(κ)4- κ Targeting Vector FIG. 4B).

[0145] All of the above approaches to insert human λ light chain gene segments at the mouse κ locus, maintain the κ intronic enhancer element upstream of the C κ gene (designated E κ i, FIG. 4B and FIG. 5B) and the 3' κ enhancer downstream of the C κ gene (designated E κ 3', FIG. 4B and FIG. 5B). The approaches result in four separate modified alleles at the endogenous mouse κ light chain locus (FIG. 7B).

[0146] [Deleted]

Lambda Domain Antibodies from Genetically Modified Mice

[0147] Mice comprising human λ sequences at the mouse λ light chain locus will express a light chain that comprises a hV λ region fused to a mouse C λ region. These are advantageously bred to mice that (a) comprise a functionally silenced light chain locus (e.g., a knockout of the endogenous mouse κ or λ light chain locus); (b) comprise an endogenous mouse λ light chain locus that comprises hV and hJ gene segments operably linked to an endogenous mouse C λ gene as recited in the claims; (c) comprise an endogenous mouse κ light chain locus that comprises hV κ and hJ κ gene segments operably linked to an endogenous mouse C κ gene; and, (d) a mouse in which one κ allele comprises hV κ s and hJ κ s; the other κ allele comprising hV λ s and hJ λ s; one λ allele comprising hV λ s and hJ λ s and one λ allele silenced or knocked out, or both λ alleles comprising hV λ s and hJ λ s; and, two heavy chain alleles that each comprise hV μ s, hD μ s, and hJ μ s.

[0148] The antibodies that comprise the hV λ domains expressed in the context of either C λ are used to make fully human antibodies by cloning the nucleic acids encoding the hV λ domains into expression constructs that bear genes encoding human C λ . Resulting expression constructs are transfected into suitable host cells for expressing antibodies that display a fully hV λ domain fused to hC λ .

EXAMPLES

[0149] The following examples are provided so as to describe how to make and use methods and compositions of the invention, and are not intended to limit the scope of what the inventors regard as their invention. Unless indicated otherwise, temperature is indicated in Celsius, and pressure is at or near atmospheric.

Example I

Deletion of the Mouse Immunoglobulin Light Chain Loci

[0150] Various targeting constructs were made using VELOCIGENE® technology (see, e.g., US Pat. No. 6,586,251 and Valenzuela et al. (2003) High-throughput engineering of the mouse genome coupled with high-resolution expression analysis, Nature Biotech. 21(6):652-659) to modify mouse genomic Bacterial Artificial Chromosome (BAC) libraries to inactivate the mouse κ and λ light chain loci.

[0151] **Deletion of the mouse λ light chain locus.** DNA from mouse BAC clone RP23-135k15 (Invitrogen) was modified by homologous recombination to inactivate the endogenous mouse λ light chain locus through targeted deletion of the V λ -J λ -C λ gene clusters (FIG. 2).

[0152] Briefly, the entire proximal cluster comprising V λ 1-J λ 3-C λ 3-J λ 1-C λ 1 gene segments was deleted in a single targeting event using a targeting vector comprising a neomycin cassette flanked by *loxP* sites with a 5' mouse homology arm containing sequence 5' of the V λ 1 gene segment and a 3' mouse homology arm containing sequence 3' of the C λ 1 gene segment (FIG. 2, Targeting Vector 1).

[0153] A second targeting construct was prepared to precisely delete the distal endogenous mouse λ gene cluster containing V λ 2-J λ 2-C λ 2-J λ 4-C λ 4 except that the targeting construct contained a 5' mouse homology arm that contained sequence 5' of the V λ 2 gene segment and a 3' mouse homology arm that contained sequence 5' to the endogenous C λ 2 gene segment (FIG. 2, Targeting Vector 2). Thus, the second targeting construct precisely deleted V λ 2-J λ 2, while leaving C λ 2-J λ 4-C λ 4 intact at the endogenous mouse λ locus. ES cells containing an inactivated endogenous λ locus (as described above) were confirmed by karyotyping and screening methods (e.g., TAQMAN®) known in the art. DNA

was then isolated from the modified ES cells and subjected to treatment with CRE recombinase thereby mediating the deletion of the proximal targeting cassette containing the neomycin marker gene, leaving only a single *loxP* site at the deletion point (FIG. 2, bottom).

[0154] Deletion of the mouse κ light chain locus. Several targeting constructs were made using similar methods described above to modify DNA from mouse BAC clones RP23-302g12 and RP23-254m04 (Invitrogen) by homologous recombination to inactivate the mouse κ light chain locus in a two-step process (FIG. 3).

[0155] Briefly, the $J\kappa$ gene segments (1-5) of the endogenous mouse κ light chain locus were deleted in a single targeting event using a targeting vector comprising a hygromycin-thymidine kinase (hyg-TK) cassette containing a single *loxP* site 3' to the hyg-TK cassette (FIG. 3, $J\kappa$ Targeting Vector). The homology arms used to make this targeting vector contained mouse genomic sequence 5' and 3' of the endogenous mouse $J\kappa$ gene segments. In a second targeting event, a second targeting vector was prepared to delete a portion of mouse genomic sequence upstream (5') to the most distal endogenous mouse $V\kappa$ gene segment (FIG. 3, $V\kappa$ Targeting Vector). This targeting vector contained an inverted *lox511* site, a *loxP* site and a neomycin cassette. The homology arms used to make this targeting vector contained mouse genomic sequence upstream of the most distal mouse $V\kappa$ gene segment. The targeting vectors were used in a sequential fashion (*i.e.*, $J\kappa$ then $V\kappa$) to target DNA in ES cells. ES bearing a double-targeted chromosome (*i.e.*, a single endogenous mouse κ locus targeted with both targeting vectors) were confirmed by karyotyping and screening methods (*e.g.*, TaqmanTM) known in the art. DNA was then isolated from the modified ES cells and subjected to treatment with Cre recombinase thereby mediating the deletion of endogenous mouse $V\kappa$ gene segments and both selection cassettes, while leaving two juxtaposed *lox* sites in opposite orientation relative to one another (FIG. 3, bottom; SEQ ID NO:1).

[0156] Thus, two modified endogenous light chain loci (κ and λ) containing intact enhancer and constant regions were created for progressively inserting unrearranged human λ germline gene segments in a precise manner using targeting vectors described below.

Example II

Replacement of Mouse Light Chain Loci with a Human λ Light Chain Mini-Locus

[0157] Multiple targeting vectors were engineered for progressive insertion of human λ gene segments into the endogenous mouse κ and λ light chain loci using similar methods as described above. Multiple independent initial modifications were made to the endogenous light chain loci each producing a chimeric light chain locus containing hV λ and J λ gene segments operably linked to mouse light chain constant genes and enhancers.

[0158] A human λ mini-locus containing 12 human V λ and one human J λ gene segment. A series of initial targeting vectors were engineered to contain the first 12 consecutive human V λ gene segments from cluster A and a hJ λ 1 gene segment or four hJ λ gene segments using a human BAC clone named RP11-729g4 (Invitrogen). FIGs. 4A and 4B show the targeting vectors that were constructed for making an initial insertion of human λ light chain gene segments at the mouse λ and κ light chain loci, respectively.

[0159] For a first set of initial targeting vectors, a 124,125 bp DNA fragment from the 729g4 BAC clone containing 12 hV λ gene segments and a hJ λ 1 gene segment was engineered to contain a *PI-SceI* site 996 bp downstream (3') of the hJ λ 1 gene segment for ligation of a 3' mouse homology arm. Two different sets of homology arms were used for ligation to this human fragment; one set of homology arms contained endogenous mouse λ sequences from the 135k15 BAC clone (FIG. 4A) and another set contained endogenous κ sequence 5' and 3' of the mouse $V\kappa$ and $J\kappa$ gene segments from mouse BAC clones RP23-302g12 and RP23-254m04, respectively (FIG. 4B).

[0160] For the 12/1- λ Targeting Vector (FIG. 4A), a *PI-SceI* site was engineered at the 5' end of a 27,847 bp DNA fragment containing the mouse C λ 2-J λ 4-C λ 4 and enhancer 2.4 of the modified mouse λ locus described in Example 1. The ~28 kb mouse fragment was used as a 3' homology arm by ligation to the ~124 kb human λ fragment, which created a 3' junction containing, from 5' to 3', a hJ λ 1 gene segment, 996 bp of human λ sequence 3' of the hJ λ 1 gene segment, 1229 bp of mouse λ sequence 5' to the mouse C λ 2 gene, the mouse C λ 2 gene and the remaining portion of the -28 kb mouse fragment. Upstream (5') from the human V λ 3-12 gene segment was an additional 1456 bp of human λ sequence before the start of the 5' mouse homology arm, which contained 23,792 bp of mouse genomic DNA corresponding to sequence 5' of the endogenous mouse λ locus. Between the 5' homology arm and the beginning of the human λ sequence was a neomycin cassette flanked by *Frt* sites.

[0161] Thus, the 12/1- λ Targeting Vector included, from 5' to 3', a 5' homology arm containing ~24 kb of mouse λ genomic sequence 5' of the endogenous λ locus, a 5' *Frt* site, a neomycin cassette, a 3' *Frt* site, ~123 kb of human genomic λ sequence containing the first 12 consecutive hV λ gene segments and a hJ λ 1 gene segment, a *PI-SceI* site, and a 3' homology arm containing ~28 kb of mouse genomic sequence including the endogenous C λ 2-J λ 4-C λ 4 gene segments, the mouse enhancer 2.4 sequence and additional mouse genomic sequence downstream (3') of the enhancer 2.4 (FIG. 4A).

[0162] In a similar fashion, the 12/1- κ Targeting Vector (FIG. 4B) employed the same ~124 human λ fragment with the

exception that mouse homology arms containing mouse κ sequence were used such that targeting to the endogenous κ locus could be achieved by homologous recombination. Thus, the 12/1- κ Targeting Vector included, from 5' to 3', a 5' homology arm containing ~23 kb of mouse genomic sequence 5' of the endogenous κ locus, an I-CeuI site, a 5' Frt site, a neomycin cassette, a 3' Frt site, ~124 kb of human genomic λ sequence containing the first 12 consecutive hV λ gene segments and a hJ λ 1 gene segment, a PI-SceI site, and a 3' homology arm containing ~28 kb of mouse genomic sequence including the endogenous the mouse C κ gene, E κ i and E κ 3' and additional mouse genomic sequence downstream (3') of E κ 3' (FIG. 4B, 12/1- κ Targeting Vector).

[0163] Homologous recombination with either of these two initial targeting vectors created a modified mouse light chain locus (κ or λ) containing 12 hV λ gene segments and a hJ λ 1 gene segment operably linked to the endogenous mouse light chain constant gene and enhancers (C κ or C λ 2 and E κ i/E κ 3' or Enh 2.4/Enh 3.1) gene which, upon recombination, leads to the formation of a chimeric λ light chain.

[0164] A human λ mini-locus with 12 human V λ and four human J λ gene segments. In another approach to add diversity to a chimeric λ light chain locus, a third initial targeting vector was engineered to insert the first 12 consecutive human V λ gene segments from cluster A and hJ λ 1, 2, 3 and 7 gene segments into the mouse κ light chain locus (FIG. 4B, 12/4- κ Targeting Vector). A DNA segment containing hJ λ 1, J λ 2, J λ 3 and J λ 7 gene segments was made by *de novo* DNA synthesis (Integrated DNA Technologies) including each J λ gene segment and human genomic sequence of ~100 bp from both the immediate 5' and 3' regions of each J λ gene segment. A PI-SceI site was engineered into the 3' end of this ~1 kb DNA fragment and ligated to a chloroamphenicol cassette. Homology arms were PCR amplified from human λ sequence at 5' and 3' positions relative to the hJ λ 1 gene segment of the human BAC clone 729g4. Homologous recombination with this intermediate targeting vector was performed on a modified 729g4 BAC clone that had been previously targeted upstream (5') of the human V λ 3-12 gene segment with a neomycin cassette flanked by Frt sites, which also contained an I-CeuI site 5' to the 5' Frt site. The double-targeted 729g4 BAC clone included from 5' to 3' an I-CeuI site, a 5' Frt site, a neomycin cassette, a 3' Frt site, a ~123 kb fragment containing the first 12 hV λ gene segments, a ~1 kb fragment containing human J λ 1, 2, 3 and 7 gene segments, a PI-SceI site, and a chloroamphenicol cassette. This intermediate targeting vector was digested together with I-CeuI and PI-SceI and subsequently ligated into the modified mouse BAC clone (described above) to create the third targeting vector.

[0165] This ligation resulted in a third targeting vector for insertion of human λ sequences into the endogenous κ light chain locus, which included, from 5' to 3', a 5' mouse homology arm containing ~23 kb of genomic sequence 5' of the endogenous mouse κ locus, an I-CeuI site, a 5' Frt site, a neomycin cassette, a 3' Frt site, a ~123 kb fragment containing the first 12 hV λ gene segments, a ~1 kb fragment containing hJ λ 1, 2, 3 and 7 gene segments, a PI-SceI site and a 3' homology arm containing ~28 kb of mouse genomic sequence including the endogenous the mouse C κ gene, E κ i and E κ 3' and additional mouse genomic sequence downstream (3') of E κ 3' (FIG. 4B, 12/4- κ Targeting Vector). Homologous recombination with this third targeting vector created a modified mouse κ light chain locus containing 12 hV λ gene segments and four hJ λ gene segments operably linked to the endogenous mouse C κ gene which, upon recombination, leads to the formation of a chimeric human λ /mouse κ light chain.

[0166] A human λ mini-locus with an integrated human κ light chain sequence. In a similar fashion, two additional targeting vectors similar to those engineered to make an initial insertion of human λ gene segments into the endogenous κ light chain locus (FIG. 4B, 12/1- κ and 12/4- κ Targeting Vectors) were engineered to progressively insert human λ light chain gene segments using uniquely constructed targeting vectors containing contiguous human λ and κ genomic sequences. These targeting vectors were constructed to include a ~23 kb human κ genomic sequence naturally located between human V κ 4-1 and J κ 1 gene segments. This human κ genomic sequence was specifically positioned in these two additional targeting vectors between human V λ and human J λ gene segments (FIG. 4B, 12(κ)1- κ and 12(κ)4- κ Targeting Vectors).

[0167] Both targeting vectors containing the human κ genomic sequence were made using the modified RP11-729g4 BAC clone described above (FIG. 6). This modified BAC clone was targeted with a spectinomycin selection cassette flanked by NotI and AsiSI restriction sites (FIG. 6, top left). Homologous recombination with the spectinomycin cassette resulted in a double-targeted 729g4 BAC clone which included, from 5' to 3', an I-CeuI site, a 5' Frt site, a neomycin cassette, a 3' Frt site, a ~123 kb fragment containing the first 12 hV λ gene segments, a NotI site about 200 bp downstream (3') to the nonamer sequence of the hV λ 3-1 gene segment, a spectinomycin cassette and an AsiSI site. A separate human BAC clone containing human κ sequence (CTD-2366j12) was targeted two independent times to engineer restriction sites at locations between hV κ 4-1 and hJ κ 1 gene segments to allow for subsequent cloning of a ~23 kb fragment for ligation with the hV λ gene segments contained in the double targeted modified 729g4 BAC clone (FIG. 6, top right).

[0168] Briefly, the 2366j12 BAC clone is about 132 kb in size and contains hV κ gene segments 1-6, 1-5, 2-4, 7-3, 5-2, 4-1, human κ genomic sequence down stream of the V κ gene segments, hJ κ gene segments 1-5, the hC κ and about 20 kb of additional genomic sequence of the human κ locus. This clone was first targeted with a targeting vector containing a hygromycin cassette flanked by Frt sites and a NotI site downstream (3') of the 3' Frt site. The homology arms for this targeting vector contained human genomic sequence 5' and 3' of the V κ gene segments within the BAC clone such that

upon homologous recombination with this targeting vector, the V κ gene segments were deleted and a NotI site was engineered ~133 bp downstream of the hV κ 4-1 gene segment (FIG. 6, top right). This modified 2366j12 BAC clone was targeted independently with two targeting vectors at the 3' end to delete the hJ κ gene segments with a chloroamphenicol cassette that also contained either a hJ λ 1 gene segment, a PI-SceI site and an AsiSI site or a human λ genomic fragment containing four hJ λ gene segments (*supra*), a PI-SceI site and an AsiSI site (FIG. 6, top right). The homology arms for these two similar targeting vectors contained sequence 5' and 3' of the hJ κ gene segments. Homologous recombination with these second targeting vectors and the modified 2366j12 BAC clone yielded a double-targeted 2366j12 clone which included, from 5' to 3', a 5' Frt site, a hygromycin cassette, a 3' Frt site, a NotI site, a 22,800 bp genomic fragment of the human κ locus containing the intergenic region between the V κ 4-1 and J κ 1 gene segments, either a hJ λ 1 gene segment or a human λ genomic fragment containing hJ λ 1, J λ 2, J λ 3 and J λ 7, a PI-SceI site and a chloroamphenicol cassette (FIG. 6, top right). Two final targeting vectors to make the two additional modifications were achieved by two ligation steps using the double-targeted 729g4 and 2366j12 clones.

[0169] Double targeted 729g4 and 2366j12 clones were digested with NotI and AsiSI yielding one fragment containing the neomycin cassette and hV λ gene segments and another fragment containing the ~23 kb genomic fragment of the human κ locus containing the intergenic region between the V κ 4-1 and J κ 1 gene segments, either a hJ λ 1 gene segment or a genomic fragment containing hJ λ 1, J λ 2, J λ 3 and J λ 7 gene segments, the PI-SceI site and the chloroamphenicol cassette, respectively. Ligation of these fragments generated two unique BAC clones containing from 5' to 3' the hV λ gene segments, the human κ genomic sequence between the V κ 4-1 and J κ 1 gene segments, either a hJ λ 1 gene segment or a genomic fragment containing hJ λ 1, J λ 2, J λ 3 and J λ 7 gene segments, a PI-SceI site and a chloroamphenicol cassette (FIG. 6, bottom). These new BAC clones were then digested with I-CeuI and PI-SceI to release the unique fragments containing the upstream neomycin cassette and the contiguous human λ and κ sequences and ligated into a modified mouse BAC clone 302g12 which contained from 5' to 3' mouse genomic sequence 5' of the endogenous κ locus, an I-CeuI site, a 5' Frt site, a neomycin cassette, a 3' Frt site, hV λ gene segments (3-12 to 3-1), a NotI site ~200 bp downstream of V λ 3-1, ~23 kb of human κ sequence naturally found between the human V κ 4-1 and J κ 1 gene segments, either a hJ λ 1 gene segment or a genomic fragment containing hJ λ 1, J λ 2, J λ 3 and J λ 7 gene segments, the mouse E κ i, the mouse C κ gene and E κ 3' (FIG. 4, 12hV λ -V κ J κ -hJ λ 1 and 12hV λ -V κ J κ -4hJ λ Targeting Vectors). Homologous recombination with both of these targeting vectors created two separate modified mouse κ light chain loci containing 12 hV λ gene segments, human κ genomic sequence, and either one or four hJ λ gene segments operably linked to the endogenous mouse C κ gene which, upon recombination, leads to the formation of a chimeric human λ /mouse κ light chain.

Example III

Engineering Additional Human V λ Genes Segments Into a Human λ Light Chain Mini-Locus

[0170] Additional hV λ gene segments were added independently to each of the initial modifications described in Example 2 using similar targeting vectors and methods (FIG. 5A, +16- λ Targeting Vector and FIG. 5B, +16- κ Targeting Vector).

[0171] Introduction of 16 additional human V λ gene segments. Upstream (5') homology arms used in constructing targeting vectors for adding 16 additional hV λ gene segments to the modified light chain loci described in Example 2 contained mouse genomic sequence 5' of either the endogenous κ or λ light chain loci. The 3' homology arms were the same for all targeting vectors and contained human genomic sequence overlapping with the 5' end of the human λ sequence of the modifications as described in Example 2.

[0172] Briefly, two targeting vectors were engineered for introduction of 16 additional hV λ gene segments to the modified mouse light chain loci described in Example 2 (FIG. 5A and 5B, +16- λ or +16- κ Targeting Vector). A ~172 kb DNA fragment from human BAC clone RP11-761113 (Invitrogen) containing 21 consecutive hV λ gene segments from cluster A was engineered with a 5' homology arm containing mouse genomic sequence 5' to either the endogenous κ or λ light chain loci and a 3' homology arm containing human genomic λ sequence. The 5' mouse κ or λ homology arms used in these targeting constructs were the same 5' homology arms described in Example 2 (FIG. 5A and 5B). The 3' homology arm included a 53,057 bp overlap of human genomic λ sequence corresponding to the equivalent 5' end of the ~123 kb fragment of human genomic λ sequence described in Example 2. These two targeting vectors included, from 5' to 3', a 5' mouse homology arm containing either ~23 kb of genomic sequence 5' of the endogenous mouse κ light chain locus or ~24 kb of mouse genomic sequence 5' of the endogenous λ light chain locus, a 5' Frt site, a hygromycin cassette, a 3' Frt site and 171,457 bp of human genomic λ sequence containing 21 consecutive hV λ gene segments, ~53 kb of which overlaps with the 5' end of the human λ sequence described in Example 3 and serves as the 3' homology arm for this targeting construct (FIG. 5A and 5B, +16- λ or +16- κ Targeting Vectors). Homologous recombination with these targeting vectors created independently modified mouse κ and λ light chain loci each containing 28 hV λ gene segments and a hJ λ 1 gene segment operably linked to endogenous mouse constant genes (C κ or C λ 2) which, upon recombination, leads to the formation of a chimeric light chain.

[0173] In a similar fashion, the +16- κ Targeting Vector was also used to introduce the 16 additional hV λ gene segments to the other initial modifications described in Example 2 that incorporated multiple hJ λ gene segments with and without an integrated human κ sequence (FIG. 4B). Homologous recombination with this targeting vector at the endogenous mouse κ locus containing the other initial modifications created mouse κ light chain loci containing 28 hV λ gene segments and hJ λ 1, 2, 3 and 7 gene segments with and without a human V κ -J κ genomic sequence operably linked to the endogenous mouse C κ gene which, upon recombination, leads to the formation of a chimeric λ - κ light chain.

[0174] **Introduction of 12 additional human V λ gene segments.** Additional hV λ gene segments were added independently to each of the modifications described above using similar targeting vectors and methods. The final locus structure resulting from homologous recombination with targeting vectors containing additional hV λ gene segments are shown in FIG. 7A and 7B.

[0175] Briefly, a targeting vector was engineered for introduction of 12 additional hV λ gene segments to the modified mouse κ and λ light chain loci described above (FIG. 5A and 5B, +12- λ or 12- κ Targeting Vectors). A 93,674 bp DNA fragment from human BAC clone RP11-22118 (Invitrogen) containing 12 consecutive hV λ gene segments from cluster B was engineered with a 5' homology arm containing mouse genomic sequence 5' to either the endogenous mouse κ or λ light chain loci and a 3' homology arm containing human genomic λ sequence. The 5' homology arms used in this targeting construct were the same 5' homology arms used for the addition of 16 hV λ gene segments described above (FIG. 5A and 5B). The 3' homology arm was made by engineering a PI-SceI site ~3431 bp 5' to the human V λ 3-29P gene segment contained in a 27,468 bp genomic fragment of human λ sequence from BAC clone RP11-761113. This PI-SceI site served as a ligation point to join the ~94 kb fragment of additional human λ sequence to the -27 kb fragment of human λ sequence that overlaps with the 5' end of the human λ sequence in the previous modification using the +16- λ or +16- κ Targeting Vectors (FIG. 5A and 5B). These two targeting vectors included, from 5' to 3', a 5' homology arm containing either -23 kb of mouse genomic sequence 5' of the endogenous κ light chain locus or -24 kb of mouse genomic sequence 5' of the endogenous λ light chain locus, a 5' Frt site, a neomycin cassette, a 3' Frt site and 121,188 bp of human genomic λ sequence containing 16 hV λ gene segments and a PI-SceI site, -27 kb of which overlaps with the 5' end of the human λ sequence from the insertion of 16 additional hV λ gene segments and serves as the 3' homology arm for this targeting construct (FIG. 5A and 5B, +12- λ or 12- κ Targeting Vectors). Homologous recombination with these targeting vectors independently created modified mouse κ and λ light chain loci containing 40 hV λ gene segments and human J λ 1 operably linked to the endogenous mouse constant genes (C κ or C λ 2) which, upon recombination, leads to the formation of a chimeric light chain (bottom of FIG. 5A and 5B).

[0176] In a similar fashion, the +12- κ Targeting Vector was also used to introduce the 12 additional hV λ gene segments to the other initial modifications that incorporated multiple hJ λ gene segments with and without an integrated human κ sequence (FIG. 4B). Homologous recombination with this targeting vector at the endogenous mouse κ locus containing the other modifications created a mouse κ light chain locus containing 40 hV λ gene segments and hJ λ 1, 2, 3 and 7 gene segments with and without a human V κ -J κ genomic sequence operably linked to the endogenous mouse C κ gene which, upon recombination, leads to the formation of a chimeric λ - κ light chain.

Example IV

Identification of targeted ES cells Bearing Human λ Light Chain Gene Segments

[0177] Targeted BAC DNA made according to the foregoing Examples was used to electroporate mouse ES cells to create modified ES cells for generating chimeric mice that express human λ light chain gene segments. ES cells containing an insertion of unrearranged human λ light chain gene segments were identified by a quantitative TAQMAN® assay. Specific primers sets and probes were design for insertion of human λ sequences and associated selection cassettes (gain of allele, GOA), loss of endogenous mouse sequences and any selection cassettes (loss of allele, LOA) and retention of flanking mouse sequences (allele retention, AR). For each additional insertion of human λ sequences, additional primer sets and probes were used to confirm the presence of the additional human λ sequences as well as the previous primer sets and probes used to confirm retention of the previously targeted human sequences. Table 1 sets forth the primers and associated probes used in the quantitative PCR assays. Table 2 sets forth the combinations used for confirming the insertion of each section of human λ light chain gene segments in ES cell clones.

[0178] ES cells bearing the human λ light chain gene segments are optionally transfected with a construct that expresses FLP in order to remove the Frt'd neomycin cassette introduced by the insertion of the targeting construct containing human V λ 5-52 - V λ 1-40 gene segments (FIG. 5A and 5B). The neomycin cassette may optionally be removed by breeding to mice that express FLP recombinase (e.g., US 6,774,279). Optionally, the neomycin cassette is retained in the mice.

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Table 1

Primer	SEQIDNO:	Probe	SEQIDNO:
hL2F	2	hL2P	24
hL2R	3		
hL3F	4	hL3P	25
hL3R	5		
NeoF	6	NeoP	26
NeoR	7		
61hJ1F	8	61hJ1P	27
61hJ1R	9		
67hT1F	10	67hT1P	28
67hT1R	11		
67hT3F	12	67hT3P	29
67hT3R	13		
HygF	14	HygP	30
HygR	15		
MKD2F	16	MKD2P	31
MKD2R	17		
MKP8F	18	MKP8P	32
MKP8R	19		
MKP15F	20	MKP15P	33
MKP15R	21		
MK20F	22	-	-
MKP4R	23		
68h2F	34	68h2P	38
68h2R	35		
68h5F	36	68h5P	39
68h5R	37		
mL1F	75	mL1P	83
mL1R	76		
mL2F	77	mL2P	84
mL2R	78		
mL11F	79	mL11P	85
mL11R	80		
mL12F	81	mL12P	86
mL12R	82		

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Table 2

Modification	Assay	Forward/Reverse Primer Set	Probe	Sequence Location
Insertion of 12 hV λ & hJ λ 1	GOA	hL2F/hL2R	hL2P	hV λ 3-12- hV λ 3-1
		hL3F/hL3R	hL3P	
		61hJ1F/61hJ1R	61hJ1P	hJ λ sequence
		NeoF/NeoR	NeoP	Neomycin cassette
	LOA	MK20F/MKP4R	-	<i>lox</i> 511/ <i>lox</i> P sequence of inactivated κ locus
		HygF/HygR	HygP	Hygromycin cassette from inactivated λ locus
		mL1F/mL1R	mL1P	Mouse V λ 1-C λ 1 Cluster
		mL2F/mL2R	mL2P	
		mL11 F/mL11 R	mL11P	Mouse V λ 2-C λ 2 Cluster
		mL12F/mL12R	mL12P	
	AR/LOA	MKD2F/MKD2R	MKD2P	Mouse sequence in 5' V κ locus
		MKP15F/MKP15R	MKP15P	Mouse sequence in 3' V κ locus
Insertion of 16 hV λ	GOA	67hT1 F/67hT1 R	67hT1P	hV λ 3-27 - hV λ 3-12
		67hT3F/67hT3R	67hT3P	
		HygF/HygR	HygP	Hygromycin cassette
	LOA	NeoF/NeoR	NeoP	Neomycin cassette
		mL1 F/mL1 R	mL1P	Mouse V λ 1-C λ 1 Cluster
		mL2F/mL2R	mL2P	
		mL11F/mL11R	mL11P	Mouse V λ 2-C λ 2 Cluster
		mL12F/mL12R	mL12P	
	AR	hL2F/hL2R	hL2P	hV λ 3-12-hV λ 3-1
		hL3F/hL3R	hL3P	
	AR/LOA	MKD2F/MKD2R	MKD2P	Mouse sequence in 5' V κ locus
		MKP15F/MKP15R	MKP15P	Mouse sequence in 3' V κ locus
Insertion of 12 hV λ	GOA	68h2F/68h2R	68h2P	hV λ 5-52-hV λ 1-40
		68h5F/68h5R	68h5P	
		NeoF/NeoR	NeoP	Neomycin cassette
	LOA	HygF/HygR	HygP	Hygromycin cassette
		mL1F/mL1R	mL1P	Mouse V λ 1-C λ 1 Cluster
		mL2F/mL2R	mL2P	
		mL11F/mL11R	mL11P	Mouse V λ 2-C λ 2 Cluster
		mL12F/mL12R	mL12P	
	AR	hL2F/hL2R	hL2P	hV λ 3-12-hV λ 3-1
		hL3F/hL3R	hL3P	
		67hT1F/67hT1R	67hT1P	hV λ 3-27- hV λ 3-12
		67hT3F/67hT3R	67hT3P	

(continued)

Modification	Assay	Forward/Reverse Primer Set	Probe	Sequence Location
	AR/LOA	MKD2F/MKD2R	MKD2P	Mouse sequence in 5' V κ locus
		MKP15F/MKP15R	MKP15P	Mouse sequence in 3' V κ locus

Example V**Generation of Mice Expressing Human λ Light Chains From an Endogenous Light Chain Locus**

[0179] Targeted ES cells described above were used as donor ES cells and introduced into an 8-cell stage mouse embryo by the VELOCIMOUSE® method (see, e.g., US Pat. No. 7,294,754 and Poueymirou et al. (2007) F0 generation mice that are essentially fully derived from the donor gene-targeted ES cells allowing immediate phenotypic analyses Nature Biotech. 25(1):91-99. VELOCIMICE® (F0 mice fully derived from the donor ES cell) independently bearing human λ gene segments were identified by genotyping using a modification of allele assay (Valenzuela *et al.*, *supra*) that detected the presence of the unique human λ gene segments (*supra*).

[0180] κ : λ light chain usage of mice bearing human λ light chain gene segments. Mice homozygous for each of three successive insertions of hV λ gene segments with a single hJ λ gene segment (FIG. 5B) and mice homozygous for a first insertion of hV λ gene segments with either a single hJ λ gene segment or four human J λ gene segments including a human V κ -J κ genomic sequence (FIG. 4B) were analyzed for κ and λ light chain expression in splenocytes using flow cytometry.

[0181] Briefly, spleens were harvested from groups of mice (ranging from three to seven animals per group) and grinded using glass slides. Following lysis of red blood cells (RBCs) with ACK lysis buffer (Lonza Walkersville), splenocytes were stained with fluorescent dye conjugated antibodies specific for mouse CD19 (Clone 1 D3; BD Biosciences), mouse CD3 (17A2; Biolegend), mouse Ig κ (187.1; BD Biosciences) and mouse Ig λ (RML-42; Biolegend). Data was acquired using a BD™ LSR II flow cytometer (BD Biosciences) and analyzed using FLOWJO™ software (Tree Star, Inc.). Table 3 sets forth the average percent values for B cells (CD19⁺), κ light chain (CD19⁺Ig κ ⁺Ig λ ⁻) and λ light chain (CD19⁺Ig κ ⁻Ig λ ⁺) expression observed in splenocytes from groups of animals bearing each genetic modification.

[0182] In a similar experiment, B cell contents of the splenic compartment from mice homozygous for a first insertion of 12 hV λ and four hJ λ gene segments including a human V κ -J κ genomic sequence operably linked to the mouse C κ gene (bottom of FIG. 4B) and mice homozygous for 40 hV λ and one hJ λ gene segment (bottom of FIG. 5B or top of FIG. 7B) were analyzed for Ig κ and Ig λ expression using flow cytometry (as described above). FIG. 8A shows the Ig λ and Ig κ expression in CD19⁺ B cells for a representative mouse from each group. The number of CD19⁺ B cells per spleen was also recorded for each mouse (FIG. 8B).

[0183] In another experiment, B cell contents of the spleen and bone marrow compartments from mice homozygous for 40 hV λ and four hJ λ gene segments including a human V κ -J κ genomic sequence operably linked to the mouse C κ gene (bottom of FIG. 7B) were analyzed for progression through B cell development using flow cytometry of various cell surface markers.

[0184] Briefly, two groups (N=3 each, 9-12 weeks old, male and female) of wild type and mice homozygous for 40 hV λ and four hJ λ gene segments including a human V κ -J κ genomic sequence operably linked to the mouse C κ gene were sacrificed and spleens and bone marrow were harvested. Bone marrow was collected from femurs by flushing with complete RPMI medium (RPMI medium supplemented with fetal calf serum, sodium pyruvate, Hepes, 2-mercaptoethanol, non-essential amino acids, and gentamycin). RBCs from spleen and bone marrow preparations were lysed with ACK lysis buffer (Lonza Walkersville), followed by washing with complete RPMI medium. 1x10⁶ cells were incubated with anti-mouse CD16/CD32 (2.4G2, BD Biosciences) on ice for 10 minutes, followed by labeling with a selected antibody panel for 30 min on ice.

[0185] Bone marrow panel: anti-mouse FITC-CD43 (1B11, BioLegend), PE-ckit (2B8, BioLegend), PeCy7-IgM (II/41, eBioscience), PerCP-Cy5.5-IgD (11-26c.2a, BioLegend), APC- B220 (RA3-6B2, eBioscience), APC-H7-CD19 (ID3, BD) and Pacific Blue-CD3 (17A2, BioLegend).

[0186] Bone marrow and spleen panel: anti-mouse FITC-Ig κ (187.1, BD), PE-Ig λ (RML-42, BioLegend), PeCy7-IgM (II/41, eBioscience), PerCP-Cy5.5-IgD (11-26c.2a, BioLegend), Pacific Blue-CD3 (17A2, BioLegend), APC- B220 (RA3-6B2, eBioscience), APC-H7-CD19 (ID3, BD).

[0187] Following staining, cells were washed and fixed in 2% formaldehyde. Data acquisition was performed on a FACSCANTOII™ flow cytometer (BD Biosciences) and analyzed with FLOWJO™ software (Tree Star, Inc.). FIGs. 9A-9D show the results for the splenic compartment of one representative mouse from each group. FIGs. 10A-10E show

the results for the bone marrow compartment of one representative mouse from each group. Table 4 sets forth the average percent values for B cells (CD19⁺), κ light chain (CD19⁺IgK⁺Ig λ ⁻) and λ light chain (CD19⁺Ig κ ⁻Ig λ ⁺) expression observed in splenocytes from groups of animals bearing various genetic modifications. Table 5 sets forth the average percent values for B cells (CD19⁺), mature B cells (B220^{hi}IgM⁺), immature B cells (B220^{int}IgM⁺), immature B cells expressing κ light chain (B220^{int}IgM⁺Ig κ ⁺) and immature B cells expressing λ light chain (B220^{int}IgM⁺Ig λ ⁺) observed in bone marrow of wild type and mice homozygous for 40 hV λ and four hJ λ gene segments including a human V κ -J κ genomic sequence operably linked to the mouse C κ gene. This experiment was repeated with additional groups of the mice described above and demonstrated similar results (data not shown).

Table 3

Genotype	% B cells	% Ig κ ⁺	% Ig λ ⁺
Wild Type	46.2	91.0	3.6
12 hV λ -hJ λ 1	28.3	10.4	62.5
12 hV λ -V κ J κ -hJ λ 1	12.0	11.0	67.5
12 hV λ -V κ J κ -4hJ λ	41.8	17.2	68.4
28 hV λ -hJ λ 1	22.0	13.3	51.1
40 hV λ -hJ λ 1	28.2	24.3	53.0

Table 4

Genotype	% B cells	% Ig κ ⁺	% Ig λ ⁺
Wild Type	49.8	91.2	3.5
40 hV λ -V κ J κ -4hJ λ	33.3	41.6	43.1

Table 5

Genotype	% B cells	% Mature B cells	% Immature B cells	% Immature Ig κ ⁺ B cells	% Immature Ig λ ⁺ B cells
Wild Type	62.2	9.2	12.0	79.0	8.84
40hV λ -V κ J κ -4hJ λ	60.43	2.59	7.69	38.29	43.29

[0188] Human V λ gene usage in mice bearing human λ light chain gene segments. Mice heterozygous for a first insertion of human λ sequences (hV λ 3-12 - hV λ 3-1 and hJ λ 1, FIG. 5B) and homozygous for a third insertion of human λ sequences (hV λ 5-52 - hV λ 3-1 and hJ λ 1, FIG. 5B) were analyzed for human λ light chain gene usage by reverse-transcriptase polymerase chain reaction (RT-PCR) using RNA isolated from splenocytes.

[0189] Briefly, spleens were harvested and perfused with 10 mL RPMI-1640 (Sigma) with 5% HI-FBS in sterile disposable bags. Each bag containing a single spleen was then placed into a STOMACHER™ (Seward) and homogenized at a medium setting for 30 seconds. Homogenized spleens were filtered using a 0.7 μ m cell strainer and then pelleted with a centrifuge (1000 rpm for 10 minutes) and RBCs were lysed in BD PHARM LYSE™ (BD Biosciences) for three minutes. Splenocytes were diluted with RPMI-1640 and centrifuged again, followed by resuspension in 1 mL of PBS (Irvine Scientific). RNA was isolated from pelleted splenocytes using standard techniques known in the art.

[0190] RT-PCR was performed on splenocyte RNA using primers specific for human hV λ gene segments and the mouse C κ gene (Table 6). PCR products were gel-purified and cloned into pCR2.1-TOPO TA vector (Invitrogen) and sequenced with primers M13 Forward (GTAAAACGAC GGCCAG; SEQ ID NO: 55) and M13 Reverse (CAGGAAACAG CTATGAC; SEQ ID NO:56) located within the vector at locations flanking the cloning site. Eighty-four total clones derived from the first and third insertions of human λ sequences were sequenced to determine hV λ gene usage (Table 7). The nucleotide sequence of the hV λ -hJ λ 1-mC κ junction for selected RT-PCR clones is shown in FIG. 11.

[0191] In a similar fashion, mice homozygous for a third insertion of human λ light chain gene sequences (*i.e.* 40 hV λ gene segments and four hJ λ gene segments including a human V κ -J κ genomic sequence, bottom of FIG. 7B) operably

linked to the endogenous mouse C κ gene were analyzed for human λ light chain gene usage by RT-PCR using RNA isolated from splenocytes (as described above). The human λ light chain gene segment usage for 26 selected RT-PCR clones are shown in Table 8. The nucleotide sequence of the hV λ -hJ λ -mC κ junction for selected RT-PCR clones is shown in FIG. 12.

[0192] In a similar fashion, mice homozygous for a first insertion of human λ light chain gene segments (12 hV λ gene segments and hJ λ 1, FIG. 4A & FIG. 5A) operably linked to the endogenous mouse C λ 2 gene were analyzed for human λ light chain gene usage by RT-PCR using RNA isolated from splenocytes (as described above). The primers specific for hV λ gene segments (Table 6) were paired with one of two primers specific for the mouse C λ 2 gene; C λ 2-1 (SEQ ID NO:104) or C λ 2-2 (SEQ ID NO:105).

[0193] Multiple hV λ gene segments rearranged to h λ 1 were observed from the RT-PCR clones from mice bearing human λ light chain gene segments at the endogenous mouse λ light chain locus. The nucleotide sequence of the hV λ -hJ λ -mC λ 2 junction for selected RT-PCR clones is shown in FIG. 13.

Table 6

5' hV λ Primer	Sequence (5'-3')	SEQ ID NO:
VLL-1	CCTCTCCTCC TCACCCTCCT	40
VLL-1n	ATGRCCDGST YYYCTCTCCT	41
VLL-2	CTCCTCACTC AGGGCACA	42
VLL-2n	ATGGCCTGGG CTCTGCTSCT	43
VLL-3	ATGGCCTGGA YCSCTCTCC	44
VLL-4	TCACCATGGC YTGGRYCYCM YTC	45
VLL-4.3	TCACCATGGC CTGGGTCTCC TT	46
VLL-5	TCACCATGGC CTGGAMTCYT CT	47
VLL-6	TCACCATGGC CTGGGCTCCA CTA CTT	48
VLL-7	TCACCATGGC CTGGACTCCT	49
VLL-8	TCACCATGGC CTGGATGATG CTT	50
VLL-9	TAAATATGGC CTGGGCTCCT CT	51
VLL-10	TCACCATGCC CTGGGCTCTG CT	52
VLL-11	TCACCATGGC CCTGACTCCT CT	53
3' Mouse C κ Primer	Sequence (5'-3')	SEQ ID NO:
mIgKC3'-1	CCCAAGCTTA CTGGATGGTG GGAAGATGGA	54

Table 7

hV λ	Observed No. of Clones
3-1	2
4-3	3
2-8	7
3-9	4
3-10	3
2-14	1
3-19	1
2-23	7
3-25	1
1-40	9

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(continued)

hV λ	Observed No. of Clones
7-43	2
1-44	2
5-45	8
7-46	3
9-49	6
1-51	3

Table 8

Clone	hV λ	hJ λ
1-3	1-44	7
1-5	1-51	3
2-3	9-49	7
2-5	1-40	1
2-6	1-40	7
3b-5	3-1	7
4a-1	4-3	7
4a-5	4-3	7
4b-1	1-47	3
5-1	3-10	3
5-2	1-40	7
5-3	1-40	7
5-4	7-46	2
5-6	1-40	7
5-7	7-43	3
6-1	1-40	1
6-2	1-40	2
6-7	1-40	3
7a-1	3-10	7
7a-2	9-49	2
7a-7	3-10	7
7b-2	7-43	3
7b-7	7-46	7
7b-8	7-43	3
11 a-1	5-45	2
11 a-2	5-45	7

[0194] FIG. 11 shows the sequence of the hV λ -hJ λ 1-mC κ junction for RT-PCR clones from mice bearing a first and third insertion of hV λ gene segments with a single hJ λ gene segment. The sequences shown in FIG. 11 illustrate unique

rearrangements involving different hV λ gene segments with hJ λ 1 recombined to the mouse C κ gene. Heterozygous mice bearing a single modified endogenous κ locus containing 12 hV λ gene segments and hJ λ 1 and homozygous mice bearing two modified endogenous κ loci containing 40 hV λ gene segments and hJ λ 1 were both able to produce human λ gene segments operably linked to the mouse C κ gene and produce B cells that expressed human λ light chains. These rearrangements demonstrate that the chimeric loci were able to independently rearrange human λ gene segments in multiple, independent B cells in these mice. Further, these modifications to the endogenous κ light chain locus did not render any of the hV λ gene segments inoperable or prevent the chimeric locus from recombining multiple hV λ and a hJ λ (J λ 1) gene segment during B cell development as evidenced by 16 different hV λ gene segments that were observed to rearrange with hJ λ 1 (Table 7). Further, these mice made functional antibodies containing rearranged human V λ -J λ gene segments operably linked to mouse C κ genes as part of the endogenous immunoglobulin light chain repertoire.

[0195] FIG. 12 shows the sequence of the hV λ -hJ λ -mC κ junction for selected RT-PCR clones from mice homozygous for 40 hV λ and four hJ λ gene segments including a human V κ -J κ genomic sequence. The sequences shown in FIG. 12 illustrate additional unique rearrangements involving multiple different hV λ gene segments, spanning the entire chimeric locus, with multiple different hJ λ gene segments rearranged and operably linked to the mouse C κ gene. Homozygous mice bearing modified endogenous κ loci containing 40 hV λ and four hJ λ gene segments were also able to produce human λ gene segments operably linked to the mouse C κ gene and produce B cells that expressed human λ light chains. These rearrangements further demonstrate that the all stages of chimeric loci were able to independently rearrange human λ gene segments in multiple, independent B cells in these mice. Further, these additional modifications to the endogenous κ light chain locus demonstrates that each insertion of human λ gene segments did not render any of the hV λ and/or J λ gene segments inoperable or prevent the chimeric locus from recombining the hV λ and J λ gene segments during B cell development as evidenced by 12 different hV λ gene segments that were observed to rearrange with all four hJ λ gene segments (Table 8) from the 26 selected RT-PCR clone. Further, these mice as well made functional antibodies containing human V λ -J λ gene segments operably linked to mouse C κ regions as part of the endogenous immunoglobulin light chain repertoire.

[0196] FIG. 13 shows the sequence of the hV λ -hJ λ -mC λ 2 junction for three individual RT-PCR clones from mice homozygous for 12 hV λ gene segments and hJ λ 1. The sequences shown in FIG. 13 illustrate additional unique rearrangements involving different hV λ gene segments, spanning the length of the first insertion, with hJ λ 1 rearranged and operably linked to the mouse C λ 2 gene (2D1 = V λ 2-8J λ 1; 2D9 = V λ 3-10J λ 1; 3E15 = V λ 3-1J λ 1). One clone demonstrated a nonproductive rearrangement due to N additions at the hV λ -hJ λ junction (2D1, FIG.13). This is not uncommon in V(D)J recombination, as the joining of gene segments during recombination has been shown to be imprecise. Although this clone represents an unproductive recombinant present in the light chain repertoire of these mice, this demonstrates that the genetic mechanism that contributes to junctional diversity among antibody genes is operating normally in these mice and leading to an antibody repertoire containing light chains with greater diversity.

[0197] Homozygous mice bearing modified endogenous λ loci containing 12 hV λ gene segments and hJ λ 1 were also able to produce human λ gene segments operably linked to an endogenous mouse C λ gene and produce B cells that expressed reverse chimeric λ light chains containing hV λ regions linked to mouse C λ regions. These rearrangements further demonstrate that human λ light chain gene segments placed at the other light chain locus (*i.e.*, the λ locus) were able to independently rearrange human λ gene segments in multiple, independent B cells in these mice. Further, the modifications to the endogenous λ light chain locus demonstrate that the insertion of human λ gene segments did not render any of the hV λ and/or hJ λ 1 gene segments inoperable or prevent the chimeric locus from recombining the hV λ and hJ λ 1 gene segments during B cell development. Further, these mice also made functional antibodies containing human V λ -J λ gene segments operably linked to a mouse C λ region as part of the endogenous immunoglobulin light chain repertoire.

[0198] As shown in this Example, mice bearing human λ light chain gene segments at the endogenous κ and λ light chain loci are capable of rearranging human λ light chain gene segments and expressing them in the context of a mouse C κ and/or C λ region as part of the normal antibody repertoire of the mouse because a functional light chain is required at various checkpoints in B cell development in both the spleen and bone marrow. Further, early subsets of B cells (*e.g.*, pre-, pro- and transitional B cells) demonstrate a normal phenotype in these mice as compared to wild type littermates (FIGs. 9D, 10A and 10B). A small deficit in bone marrow and peripheral B cell populations was observed, which may be attributed to a deletion of a subset of auto-reactive immature B cells and/or a suboptimal association of human λ light chain with mouse heavy chain. However, the Ig κ /Ig λ usage observed in these mice demonstrates a situation that is more like human light chain expression than that observed in mice.

Example VI

Breeding of Mice Expressing Human λ Light Chains From an Endogenous Light Chain Locus

[0199] To optimize the usage of the human λ gene segments at an endogenous mouse light chain locus, mice bearing

the unrearranged human λ gene segments are bred to another mouse containing a deletion in the opposing endogenous light chain locus (either κ or λ). For example, human λ gene segments positioned at the endogenous κ locus would be the only functional light chain gene segments present in a mouse that also carried a deletion in the endogenous λ light chain locus. In this manner, the progeny obtained would express only human λ light chains as described in the foregoing examples. Breeding is performed by standard techniques recognized in the art and, alternatively, by commercial companies, e.g., The Jackson Laboratory. Mouse strains bearing human light chain gene segments at the endogenous κ locus and a deletion of the endogenous λ light chain locus are screened for presence of the unique reverse-chimeric (human-mouse) λ light chains and absence of endogenous mouse λ light chains.

[0200] Mice bearing an unrearranged human λ light chain locus are also bred with mice that contain a replacement of the endogenous mouse heavy chain variable gene locus with the human heavy chain variable gene locus (see US 6,596,541, Regeneron Pharmaceuticals, the VELOCIMMUNE® genetically engineered mouse). The VELOCIMMUNE® mouse includes, in part, having a genome comprising human heavy chain variable regions operably linked to endogenous mouse constant region loci such that the mouse produces antibodies comprising a human heavy chain variable region and a mouse heavy chain constant region in response to antigenic stimulation. The DNA encoding the variable regions of the heavy chains of the antibodies can be isolated and operably linked to DNA encoding the human heavy chain constant regions. The DNA can then be expressed in a cell capable of expressing the fully human heavy chain of the antibody. Upon a suitable breeding schedule, mice bearing a replacement of the endogenous mouse heavy chain locus with the human heavy chain locus and an unrearranged human λ light chain locus at the endogenous κ light chain locus is obtained. Antibodies containing somatically mutated human heavy chain variable regions and human λ light chain variable regions can be isolated upon immunization with an antigen of interest.

Example VII

Generation of Antibodies From Mice Expressing Human Heavy Chains and Human λ Light Chains

[0201] After breeding mice that contain the unrearranged human λ light chain locus to various desired strains containing modifications and deletions of other endogenous Ig loci (as described above), selected mice are immunized with an antigen of interest.

[0202] Generally, a VELOCIMMUNE® mouse containing one of the single rearranged human germline light chain regions is challenged with an antigen, and lymphatic cells (such as B-cells) are recovered from serum of the animals. The lymphatic cells may be fused with a myeloma cell line to prepare immortal hybridoma cell lines, and such hybridoma cell lines are screened and selected to identify hybridoma cell lines that produce antibodies containing human heavy chain and human λ light chain that are specific to the antigen used for immunization. DNA encoding the variable regions of the heavy chains and the λ light chains may be isolated and linked to desirable isotypic constant regions of the heavy chain and light chain. Due to the presence of the additional hV λ gene segments as compared to the endogenous mouse λ locus, the diversity of the light chain repertoire is dramatically increased and confers higher diversity on the antigen-specific repertoire upon immunization. The resulting cloned antibody sequences may be subsequently produced in a cell, such as a CHO cell. Alternatively, DNA encoding the antigen-specific chimeric antibodies or the variable domains of the light and heavy chains may be isolated directly from antigen-specific lymphocytes (e.g., B cells).

[0203] Initially, high affinity chimeric antibodies are isolated having a human variable region and a mouse constant region. As described above, the antibodies are characterized and selected for desirable characteristics, including affinity, selectivity, epitope, etc. The mouse constant regions are replaced with a desired human constant region to generate the fully human antibody containing a somatically mutated human heavy chain and a human λ light chain derived from an unrearranged human λ light chain locus of the invention. Suitable human constant regions include, for example wild type or modified IgG1, IgG2, IgG3, or IgG4.

SEQUENCE LISTING

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EP 2 480 675 B1

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EP 2 480 675 B1

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EP 2 480 675 B1

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	ggggacgagg ccgattatta ctgcggaaca tgggatagca gcctgagtgc ttatgtcttc	360
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40	ccaccatcca gtgagcagtt a	441

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EP 2 480 675 B1

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EP 2 480 675 B1

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EP 2 480 675 B1

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    gctgaggacg aggctgatta ttactgctgc tcatatgcag gtagtagcac ttatgtcttc      360
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<220>
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    caccaggca aagcccccaa actcatgatt tatgagggtca gtaatcggcc ctcagggggt      180
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<400> 88

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EP 2 480 675 B1

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<210> 93

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Claims

1. A mouse comprising an endogenous λ (lambda) light chain allele comprising an unrearranged human lambda variable (hV λ) and an unrearranged human lambda joining (hJ λ) gene segment, where the mouse comprises a deletion of a first V λ -J λ -C λ gene cluster of said endogenous mouse λ light chain allele and replacement, in whole or in part, of the V λ -J λ gene segments of the second V λ -J λ -C λ gene cluster of said allele with said human V λ and J λ gene segments which are operably linked to an intact mouse lambda constant (C λ) region gene of said second V λ -J λ -C λ gene cluster so the mouse expresses a light chain derived from a hV λ , a hJ λ and a mouse C λ gene, where the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele.
2. The mouse of claim 1, wherein the endogenous λ (lambda) light chain allele comprises a plurality of human V λ gene segments.
3. The mouse of any one of the preceding claims, wherein the endogenous λ (lambda) light chain allele comprises:
 - (a) at least 12 unrearranged human V λ gene segments; or
 - (b) 13 to 28 unrearranged human V λ gene segments.
4. The mouse of claim 3, wherein the human V λ gene segments include
 - (a) hV λ 3-1, hV λ 4-3, hV λ 2-8, hV λ 3-8, hV λ 3-10, hV λ 2-11, and hV λ 3-12; or
 - (b) hV λ 2-14, hV λ 3-16, hV λ 2-18, hV λ 3-19, hV λ 3-21, hV λ 3-22, hV λ 2-23, hV λ 3-25, and hV λ 3-27.
5. The mouse of claim 1, wherein the endogenous λ (lambda) light chain allele comprises one unrearranged J λ gene segment which is hJ λ 1.
6. The mouse of claim 1, wherein the endogenous λ (lambda) light chain allele comprises four unrearranged human J λ gene segments.
7. The mouse of claim 6, wherein the four unrearranged human J λ gene segments are J λ 1, J λ 2, J λ 3 and J λ 7.
8. The mouse of claim 1, wherein the endogenous λ (lambda) light chain allele lacks an endogenous V λ gene segment.
9. The mouse of any one of claims 1 to 3, 5 and 8, wherein the endogenous κ (kappa) light chain allele is deleted.
10. The mouse of claim 1, wherein the endogenous mouse λ light chain allele comprises:
 - (a) a contiguous sequence of the human λ light chain locus that spans from V λ 3-12 to V λ 3-1;
 - (b) a contiguous sequence of the human λ light chain locus that spans from V λ 3-29 to V λ 3-1; or
 - (c) a contiguous sequence of the human λ light chain locus that spans from V λ 3-29 to V λ 3-1 and a contiguous sequence of the human λ light chain locus that spans from V λ 5-52 to V λ 1-40.
11. An isolated cell that expresses a light chain derived from a hV λ , a hJ λ and a mouse C λ gene, wherein the cell is from, or obtainable from, the mouse of any one of the preceding claims, and wherein the cell comprises a rearranged human V λ and a rearranged human J λ gene segment, and a deletion of a first V λ -J λ -C λ gene cluster of said endogenous mouse λ light chain allele and replacement, in whole or in part, of the V λ -J λ gene segments of the second V λ -J λ -C λ gene cluster of said allele with said human V λ and J λ gene segments which are operably linked to an intact mouse C λ constant region gene of said second V λ -J λ -C λ gene cluster, and wherein the endogenous

enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele.

12. The isolated cell of claim 11, wherein the cell is a B cell.

13. An isolated mouse embryonic stem (ES) cell comprising an endogenous λ (lambda) light chain allele comprising an unrearranged human $V\lambda$ and an unrearranged human $J\lambda$ gene segment, where the cell comprises a deletion of a first $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster of said endogenous mouse λ light chain allele and a replacement, in whole or in part, of the $V\lambda$ - $J\lambda$ gene segments of the second $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster of said allele with said human $V\lambda$ and $J\lambda$ gene segments which are operably linked to an intact mouse $C\lambda$ constant region gene of said second $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster, where the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ **light** chain allele.

14. A mouse embryo comprising, made from, or obtainable from, the mouse ES cell of claim 13, wherein the mouse embryo comprises an endogenous λ (lambda) light chain allele comprising an unrearranged human $V\lambda$ and an unrearranged human $J\lambda$ gene segment, where the cell comprises a deletion of a first $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster of said endogenous mouse λ light chain allele and a replacement, in whole or in part, of the $V\lambda$ - $J\lambda$ gene segments of the second $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster of said allele with said human $V\lambda$ and $J\lambda$ gene segments which are operably linked to an intact mouse $C\lambda$ constant region gene of said second $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster, where the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ **light** chain allele.

15. Use of the B cell of claim 12 to make a hybridoma that expresses a light chain derived from a **hV** λ a **hJ** λ and a mouse $C\lambda$ gene, wherein the hybridoma is from, or obtainable from, the mouse of any one of claims 1 to 10, wherein the cell comprises a rearranged human $V\lambda$ and a rearranged human $J\lambda$ gene segment, and a deletion of a first $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster of said endogenous mouse λ light chain allele and replacement, in whole or in part, of the $V\lambda$ - $J\lambda$ gene segments of the second $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster of said allele with said human $V\lambda$ and $J\lambda$ gene segments which are operably linked to an intact mouse $C\lambda$ constant region gene of said second $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster and wherein the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele.

16. A hybridoma that expresses a light chain derived from a **hV** λ a **hJ** λ and a mouse $C\lambda$ gene wherein the hybridoma is from, or obtainable from, the mouse of any one of claims 1 to 10, wherein the cell comprises a rearranged human $V\lambda$ and a rearranged human $J\lambda$ gene segment, and a deletion of a first $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster of said endogenous mouse λ light chain allele and replacement, in whole or in part, of the $V\lambda$ - $J\lambda$ gene segments of the second $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster of the allele with said human $V\lambda$ and $J\lambda$ gene segments which are operably linked to an intact mouse $C\lambda$ constant region gene of said second $V\lambda$ - $J\lambda$ - $C\lambda$ gene cluster, and wherein the endogenous enhancers Enh 2.4, mouse lambda 3' enhancer (Enh) and Enh 3.1 are maintained intact in said λ light chain allele.

17. A method of making an antibody in a mouse comprising:

- (a) exposing a mouse according to claim 1 to an antigen;
- (b) allowing the mouse to develop an immune response to the antigen; and
- (c) isolating from the mouse of (b) an antibody that specifically recognizes the antigen, or isolating from the mouse of (b) a cell comprising an immunoglobulin domain that specifically recognizes the antigen, or identifying in the mouse of (b) a nucleic acid sequence encoding a heavy and/or light chain variable domain that binds the antigen, wherein the antibody comprises a light chain derived from a **hV** λ , a **hJ** λ and a mouse $C\lambda$ gene.

Patentansprüche

1. Maus, umfassend ein endogenes λ (Lambda)-leichte-Kette-Allel, umfassend ein umgestelltes humanes Lambda-variables(**hV** λ)- und ein nichtumgestelltes humanes Lambda-Joining(**hJ** λ)-Gensegment, wobei die Maus eine Deletion eines ersten $V\lambda$ - $J\lambda$ - $C\lambda$ -Genclusters des endogenen Maus- λ -leichte-Kette-Allels und eine Ersetzung, ganz oder teilweise, der $V\lambda$ - $J\lambda$ -Gensegmente des zweiten $V\lambda$ - $J\lambda$ - $C\lambda$ -Genclusters des Allels mit den humanen $V\lambda$ - und $J\lambda$ -Gensegmenten umfasst, welche mit einem intakten Maus- λ -konstante($C\lambda$)-Regionen des zweiten $V\lambda$ - $J\lambda$ - $C\lambda$ -Genclusters operativ verknüpft sind, sodass die Maus eine leichte Kette exprimiert, die aus einem **hV** λ -, einem **hJ** λ - und einem Maus- $C\lambda$ -Gen stammt, wobei die endogenen Enhancer Enh 2.4, Maus-Lambda-3'-Enhancer (Enh) und Enh 3.1 in dem λ -leichte-Kette-Allel intakt gehalten werden.

2. Maus gemäß Anspruch 1, wobei das endogene λ (Lambda)-leichte-Kette-Allel eine Vielzahl humaner V λ -Gensegmente umfasst.
- 5 3. Maus gemäß einem der vorangehenden Ansprüche, wobei das endogene λ (Lambda)-leichte-Kette-Allel Folgendes umfasst:
 - (a) mindestens 12 nichtumgestellte humane V λ -Gensegmente, oder
 - (b) 13 bis 28 nichtumgestellte humane V λ -Gensegmente.
- 10 4. Maus gemäß Anspruch 3, wobei die humanen V λ -Gensegmente die Folgenden umfassen:
 - (a) hV λ 3-1, hV λ 4-3, hV λ 2-8, hV λ 3-8, hV λ 3-10, hV λ 2-11 und hV λ 3-12 oder
 - (b) hV λ 2-14, hV λ 3-16, hV λ 2-18, hV λ 3-19, hV λ 3-21, hV λ 3-22, hV λ 2-23, hV λ 3-25 und hV λ 3-27.
- 15 5. Maus gemäß Anspruch 1, wobei das endogene λ (Lambda)-leichte-Kette-Allel ein nichtumgestelltes J λ -Gensegment umfasst, welches hJ λ 1 ist.
6. Maus gemäß Anspruch 1, wobei das endogene λ (Lambda)-leichte-Kette-Allel vier nichtumgestellte humane J λ -Gensegmente umfasst.
- 20 7. Maus gemäß Anspruch 6, wobei die vier nichtumgestellten humanen J λ -Gensegmente J λ 1, J λ 2, J λ 3 und J λ 7 sind.
8. Maus gemäß Anspruch 1, wobei dem endogenen λ (Lambda)-leichte-Kette-Allel ein endogenes V λ -Gensegment fehlt.
- 25 9. Maus gemäß einem der Ansprüche 1 bis 3, 5 und 8, wobei das endogene κ (Kappa)-leichte-Kette-Allel deletiert ist.
10. Maus gemäß Anspruch 1, wobei das endogene Maus- λ -leichte-Kette-Allel folgendes umfasst:
 - 30 (a) eine zusammenhängende Sequenz des humanen λ -leichte-Kette-Locus, die von V λ 3-12 bis V λ 3-1 reicht,
 - (b) eine zusammenhängende Sequenz des humanen λ -leichte-Kette-Locus, die von V λ 3-29 bis V λ 3-1 reicht, oder
 - (c) eine zusammenhängende Sequenz des humanen λ -leichte-Kette-Locus, die von V λ 3-29 bis V λ 3-1 reicht, und eine zusammenhängende Sequenz des humanen λ -leichte-Kette-Locus, die von V λ 5-52 bis V λ 1-40 reicht.
- 35 11. Isolierte Zelle, die eine leichte Kette exprimiert, die aus einem hV λ -, einem hJ λ - und einem Maus-C λ -Gen stammt, wobei die Zelle aus der Maus gemäß einem der vorangehenden Ansprüche stammt, oder daraus erhältlich ist, und wobei die Zelle ein umgestelltes humanes V λ - und ein umgestelltes humanes J λ -Gensegment, und eine Deletion eines ersten V λ -J λ -C λ -Genclusters des endogenen Maus- λ -leichte-Kette-Allels, und eine Ersetzung, ganz oder
- 40 teilweise, der V λ -J λ -Gensegmente des zweiten V λ -J λ -C λ -Genclusters des Allels mit den humanen V λ - und J λ -Gensegmenten umfasst, welche mit einem intakten Maus-C λ -konstante-Region-Gen des zweiten V λ -J λ -C λ -Genclusters operativ verknüpft sind, und wobei die endogenen Enhancer Enh 2.4, Maus-Lambda-3'-Enhancer (Enh) und Enh 3.1 in dem λ -leichte-Kette-Allel intakt gehalten werden.
- 45 12. Isolierte Zelle gemäß Anspruch 11, wobei die Zelle eine B-Zelle ist.
13. Isolierte embryonale Mausstamm(ES)-Zelle, umfassend ein endogenes λ (Lambda)-leichte-Kette-Allel, umfassend ein nichtumgestelltes humanes V λ - und ein nichtumgestelltes humanes J λ -Gensegment, wobei die Zelle eine Deletion eines ersten V λ -J λ -C λ -Genclusters des endogenen Maus- λ -leichte-Kette-Allels, und eine Ersetzung, ganz oder teilweise, der V λ -J λ -Gensegmente des zweiten V λ -J λ -C λ -Genclusters des Allels mit den humanen V λ - und J λ -Gensegmenten umfasst, welche mit einem intakten Maus-C λ -konstante-Region-Gen des zweiten V λ -J λ -C λ -Genclusters operativ verknüpft sind, wobei die endogenen Enhancer Enh 2.4, Maus-Lambda-3'-Enhancer (Enh) und Enh 3.1 in dem λ -leichte-Kette-Allel intakt gehalten werden.
- 50 14. Mausedembryo, umfassend die, hergestellt aus, oder erhältlich aus der Maus-ES-Zelle gemäß Anspruch 13, wobei der Mausedembryo ein endogenes λ (Lambda)-leichte-Kette-Allel umfasst, umfassend ein nichtumgestelltes humanes V λ - und ein nichtumgestelltes humanes J λ -Gensegment, wobei eine Zelle des Embryos eine Deletion eines ersten V λ -J λ -C λ -Genclusters des endogenen Maus- λ -leichte-Kette-Allels, und eine Ersetzung, ganz oder teilweise, der
- 55

Vλ-Jλ-Gensegmente des zweiten Vλ-Jλ-Cλ-Genclusters des Allels mit den humanen Vλ- und Jλ-Gensegmenten umfasst, welche mit einem intakten Maus-Cλ-konstante-Region-Gen des zweiten Vλ-Jλ-Cλ-Genclusters operativ verknüpft sind, wobei die endogenen Enhancer Enh 2.4, Maus-Lambda-3'-Enhancer (Enh) und Enh 3.1 in dem λ-leichte-Kette-Allel intakt gehalten werden.

15. Verwendung der B-Zelle gemäß Anspruch 12 zur Herstellung eines Hybridoms, das eine leichte Kette exprimiert, die aus einem hVλ-, einem hJλ- und einem Maus-Cλ-Gen stammt, wobei das Hybridom aus der Maus gemäß einem der Ansprüche 1 bis 10 stammt, oder daraus erhältlich ist, wobei die Zelle ein umgestelltes humanes Vλ- und ein umgestelltes humanes Jλ-Gensegment, und eine Deletion eines ersten Vλ-Jλ-Cλ-Genclusters des endogenen Maus-λ-leichte-Kette-Allels, und eine Ersetzung, ganz oder teilweise, der Vλ-Jλ-Gensegmente des zweiten Vλ-Jλ-Cλ-Genclusters des Allels mit den humanen Vλ- und Jλ-Gensegmenten umfasst, welche mit einem intakten Maus-Cλ-konstante-Region-Gen des zweiten Vλ-Jλ-Cλ-Genclusters operativ verknüpft sind, und wobei die endogenen Enhancer Enh 2.4, Maus-Lambda-3'-Enhancer (Enh) und Enh 3.1 in dem λ-leichte-Kette-Allel intakt gehalten werden.

16. Hybridom, welches eine leichte Kette exprimiert, die aus einem hVλ-, einem hJλ- und einem Maus-Cλ-Gen stammt, wobei das Hybridom aus der Maus gemäß einem der Ansprüche 1 bis 10 stammt, oder daraus erhältlich ist, wobei die Zelle ein umgestelltes humanes Vλ- und ein umgestelltes humanes Jλ-Gensegment, und eine Deletion eines ersten Vλ-Jλ-Cλ-Genclusters des endogenen Maus-λ-leichte-Kette-Allels, und eine Ersetzung, ganz oder teilweise, der Vλ-Jλ-Gensegmente des zweiten Vλ-Jλ-Cλ-Genclusters des Allels mit den humanen Vλ- und Jλ-Gensegmenten umfasst, welche mit einem intakten Maus-Cλ-konstante-Region-Gen des zweiten Vλ-Jλ-Cλ-Genclusters operativ verknüpft sind, und wobei die endogenen Enhancer Enh 2.4, Maus-Lambda-3'-Enhancer (Enh) und Enh 3.1 in dem λ-leichte-Kette-Allel intakt gehalten werden.

17. Verfahren zur Herstellung eines Antikörpers in einer Maus, welches Folgendes umfasst:

- (a) Exponieren einer Maus gemäß Anspruch 1 gegenüber einem Antigen,
- (b) es der Maus gestatten, eine Immunantwort gegen das Antigen zu entwickeln, und
- (c) Isolieren aus der Maus gemäß (b) eines Antikörpers, der spezifisch das Antigen erkennt, oder Isolieren aus der Maus gemäß (b) eine Zelle, die eine Immunglobulin-domäne umfasst, die spezifisch das Antigen erkennt, oder Identifizieren in der Maus gemäß (b) einer Nucleinsäuresequenz, die eine Schwere- und/oder leichte-Kette-variable-Domäne codiert, die das Antigen bindet, wobei der Antikörper eine leichte Kette umfasst, die aus einem hVλ-, einem hJλ- und einem Maus-Cλ-Gen stammt.

Revendications

1. Souris comportant un allèle de chaîne légère λ (lambda) endogène comprenant un segment de gène de région variable lambda humaine (hVλ) non-réarrangé et un segment de gène de jonction lambda humaine (hJλ) non-réarrangé, laquelle souris comporte une délétion d'un premier cluster de gènes Vλ-Jλ-Cλ dudit allèle de chaîne légère λ endogène de souris et le remplacement, total ou partiel, des segments de gènes Vλ-Jλ du deuxième cluster de gènes Vλ-Jλ-Cλ dudit allèle par lesdits segments de gènes Vλ et Jλ humains qui sont opérationnellement reliés à un gène de région constante lambda (Cλ) de souris intact dudit deuxième cluster de gènes Vλ-Jλ-Cλ, de sorte que la souris exprime une chaîne légère dérivée d'un gène hVλ, d'un gène hJλ et d'un gène Cλ de souris, les amplificateurs endogènes Enh2.4, Enh λ3' de souris et Enh 3.1 étant maintenus intacts dans ledit allèle de chaîne légère λ.

2. Souris conforme à la revendication 1, dans laquelle l'allèle de chaîne légère λ (lambda) endogène comprend plusieurs segments de gènes Vλ humains.

3. Souris conforme à l'une des revendications précédentes, dans laquelle l'allèle de chaîne légère λ (lambda) endogène comprend

- a) au moins 12 segments de gènes Vλ humains non-réarrangés,
- b) ou de 13 à 28 segments de gènes Vλ humains non-réarrangés.

4. Souris conforme à la revendication 3, dans laquelle les segments de gènes Vλ humains incluent

- a) hVλ3-1, hVλ4-3, hVλ2-8, hVλ3-8, hVλ3-10, hVλ2-11 et hVλ3-12,

b) ou hVλ2-14, hVλ3-16, hVλ2-18, hVλ3-19, hVλ3-21, hVλ3-22, hVλ2-23, hVλ3-25 et hVλ3-27.

5. Souris conforme à la revendication 1, dans laquelle l'allèle de chaîne légère λ (lambda) endogène comprend un et un seul segment de gène Jλ non-réarrangé, qui est hJλ1.

6. Souris conforme à la revendication 1, dans laquelle l'allèle de chaîne légère λ (lambda) endogène comprend quatre segments de gènes Jλ humains non-réarrangés.

7. Souris conforme à la revendication 6, dans laquelle les quatre segments de gènes Jλ humains non-réarrangés sont Jλ1, Jλ2, Jλ3 et Jλ7.

8. Souris conforme à la revendication 1, dans laquelle ledit allèle de chaîne légère λ (lambda) endogène ne comprend pas de segment de gène Vλ endogène.

9. Souris conforme à l'une des revendications 1 à 3, 5 et 8, dans laquelle l'allèle de chaîne légère κ (kappa) endogène est délété.

10. Souris conforme à la revendication 1, dans laquelle l'allèle de chaîne légère λ endogène de souris comprend :

- a) une séquence contiguë du locus de chaîne légère λ humaine qui va de Vλ3-12 à Vλ3-1,
- b) une séquence contiguë du locus de chaîne légère λ humaine qui va de Vλ3-29 à Vλ3-1,
- c) ou une séquence contiguë du locus de chaîne légère λ humaine qui va de Vλ3-29 à Vλ3-1 et une séquence contiguë du locus de chaîne légère λ humaine qui va de Vλ5-52 à Vλ1-40.

11. Cellule isolée qui exprime une chaîne légère dérivée d'un gène hVλ, d'un gène hJλ, et d'un gène Cλ de souris, laquelle cellule provient ou peut être obtenue à partir d'une souris conforme à l'une des revendications précédentes, et laquelle cellule comporte un segment de gène Vλ humain réarrangé et un segment de gène Jλ humain réarrangé, et une délétion d'un premier cluster de gènes Vλ-Jλ-Cλ dudit allèle de chaîne légère λ endogène de souris et le remplacement, total ou partiel, des segments de gènes Vλ-1λ du deuxième cluster de gènes Vλ-Jλ-Cλ dudit allèle par lesdits segments de gènes Vλ et Jλ humains qui sont opérationnellement reliés à un gène de région constante Cλ de souris intact dudit deuxième cluster de gènes Vλ-Jλ-Cλ, les amplificateurs endogènes Enh2.4, Enh λ3' de souris et Enh 3.1 étant maintenus intacts dans ledit allèle de chaîne légère λ.

12. Cellule isolée conforme à la revendication 11, laquelle cellule est un lymphocyte B.

13. Cellule souche embryonnaire (CSE) de souris isolée, comportant un allèle de chaîne légère λ (lambda) endogène comprenant un segment de gène Vλ humain non-réarrangé et un segment de gène Jλ humain non-réarrangé, laquelle cellule comporte une délétion d'un premier cluster de gènes Vλ-Jλ-Cλ dudit allèle de chaîne légère λ endogène de souris et le remplacement, total ou partiel, des segments de gènes Vλ-Jλ du deuxième cluster de gènes Vλ-Jλ-Cλ dudit allèle par lesdits segments de gènes Vλ et Jλ humains qui sont opérationnellement reliés à un gène de région constante Cλ de souris intact dudit deuxième cluster de gènes Vλ-Jλ-Cλ, les amplificateurs endogènes Enh2.4, Enh λ3' de souris et Enh 3.1 étant maintenus intacts dans ledit allèle de chaîne légère λ.

14. Embryon de souris qui comprend une cellule CES de souris conforme à la revendication 13, ou qui est produit ou peut être obtenu à partir d'une telle cellule, lequel embryon de souris comporte un allèle de chaîne légère λ (lambda) endogène comprenant un segment de gène Vλ humain non-réarrangé et un segment de gène Jλ humain non-réarrangé, et duquel embryon une cellule comporte une délétion d'un premier cluster de gènes Vλ-Jλ-Cλ dudit allèle de chaîne légère λ endogène de souris et le remplacement, total ou partiel, des segments de gènes Vλ-Jλ du deuxième cluster de gènes Vλ-Jλ-Cλ dudit allèle par lesdits segments de gènes Vλ et Jλ humains qui sont opérationnellement reliés à un gène de région constante Cλ de souris intact dudit deuxième cluster de gènes Vλ-Jλ-Cλ, les amplificateurs endogènes Enh2.4, Enh λ3' de souris et Enh 3.1 étant maintenus intacts dans ledit allèle de chaîne légère λ.

15. Utilisation d'un lymphocyte B conforme à la revendication 12 pour produire un hybridome qui exprime une chaîne légère dérivée d'un gène hVλ, d'un gène hJλ et d'un gène Cλ de souris, lequel hybridome provient ou peut être obtenu à partir d'une souris conforme à l'une des revendications 1 à 10, laquelle cellule comporte un segment de gène Vλ humain réarrangé et un segment de gène Jλ humain réarrangé, et une délétion d'un premier cluster de gènes Vλ-Jλ-Cλ dudit allèle de chaîne légère λ endogène de souris et le remplacement, total ou partiel, des segments

de gènes V λ -J λ du deuxième cluster de gènes V λ -J λ -C λ dudit allèle par lesdits segments de gènes V λ et J λ humains qui sont opérationnellement reliés à un gène de région constante C λ de souris intact dudit deuxième cluster de gènes V λ -J λ -C λ , les amplificateurs endogènes Enh2.4, Enh λ 3' de souris et Enh 3.1 étant maintenus intacts dans ledit allèle de chaîne légère λ .

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16. Hybridome qui exprime une chaîne légère dérivée d'un gène hV λ , d'un gène hJ λ et d'un gène C λ de souris, lequel hybridome provient ou peut être obtenu à partir d'une souris conforme à l'une des revendications 1 à 10, et laquelle cellule comporte un segment de gène V λ humain réarrangé et un segment de gène J λ humain réarrangé, et une délétion d'un premier cluster de gènes V λ -J λ -C λ dudit allèle de chaîne légère λ endogène de souris et le remplacement, total ou partiel, des segments de gènes V λ -J λ du deuxième cluster de gènes V λ -J λ -C λ dudit allèle par lesdits segments de gènes V λ et J λ humains qui sont opérationnellement reliés à un gène de région constante C λ de souris intact dudit deuxième cluster de gènes V λ -J λ -C λ , les amplificateurs endogènes Enh2.4, Enh λ 3' de souris et Enh 3.1 étant maintenus intacts dans ledit allèle de chaîne légère λ .

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17. Procédé de production d'un anticorps dans une souris, comportant les étapes suivantes :

- a) exposer à un antigène une souris conforme à la revendication 1,
- b) laisser la souris développer une réponse immunitaire à l'antigène,
- c) et isoler, à partir de la souris issue de l'étape (b), un anticorps qui reconnaît spécifiquement l'antigène, ou isoler, à partir de la souris issue de l'étape (b), une cellule comprenant un domaine d'immunoglobuline qui reconnaît spécifiquement l'antigène, ou identifier, chez la souris issue de l'étape (b), une séquence d'acide nucléique codant un domaine variable de chaîne légère et/ou de chaîne lourde qui se lie à l'antigène,

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ledit anticorps comportant une chaîne légère dérivée d'un gène hV λ , d'un gène hJ λ et d'un gène C λ de souris.

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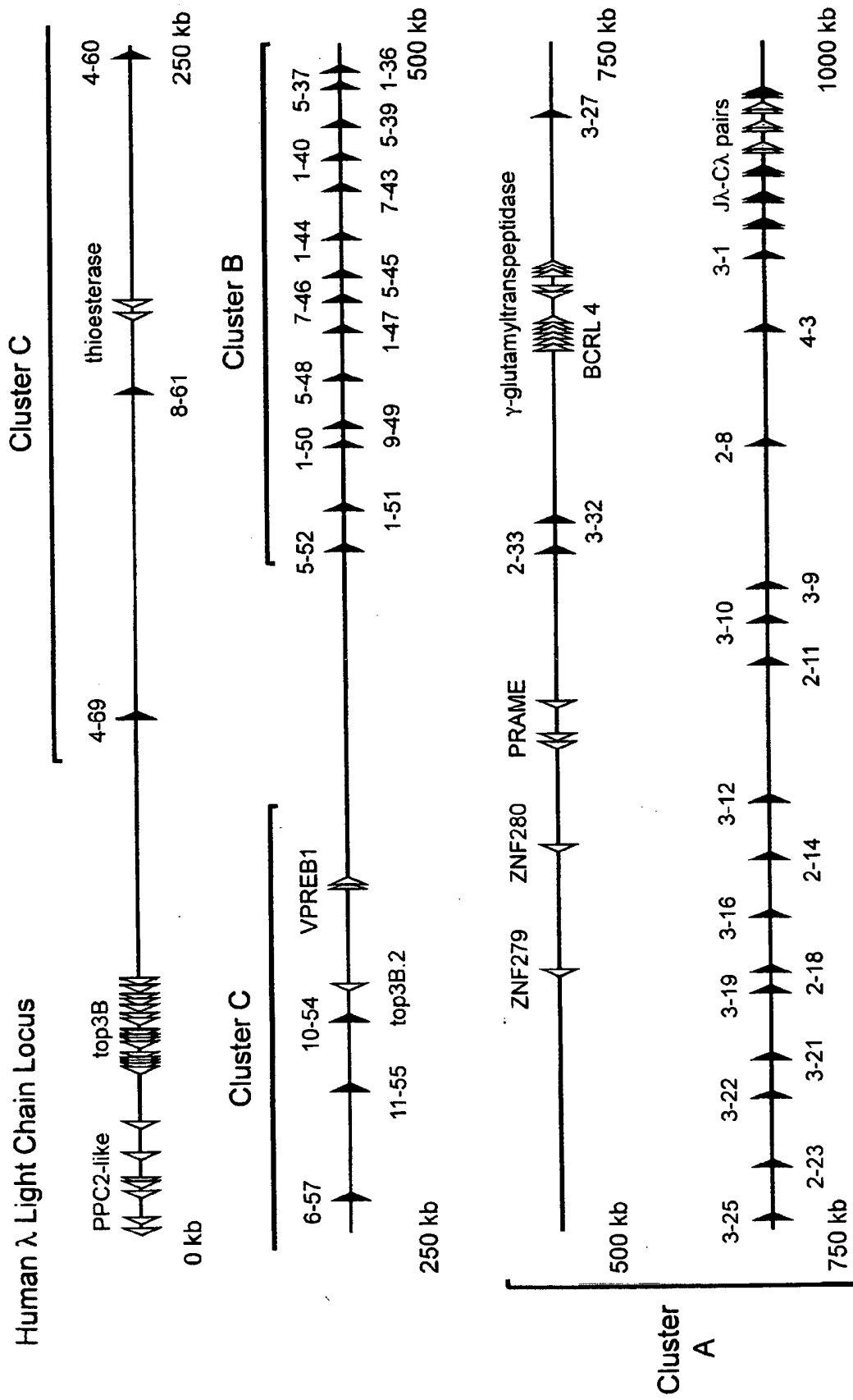


FIG. 1

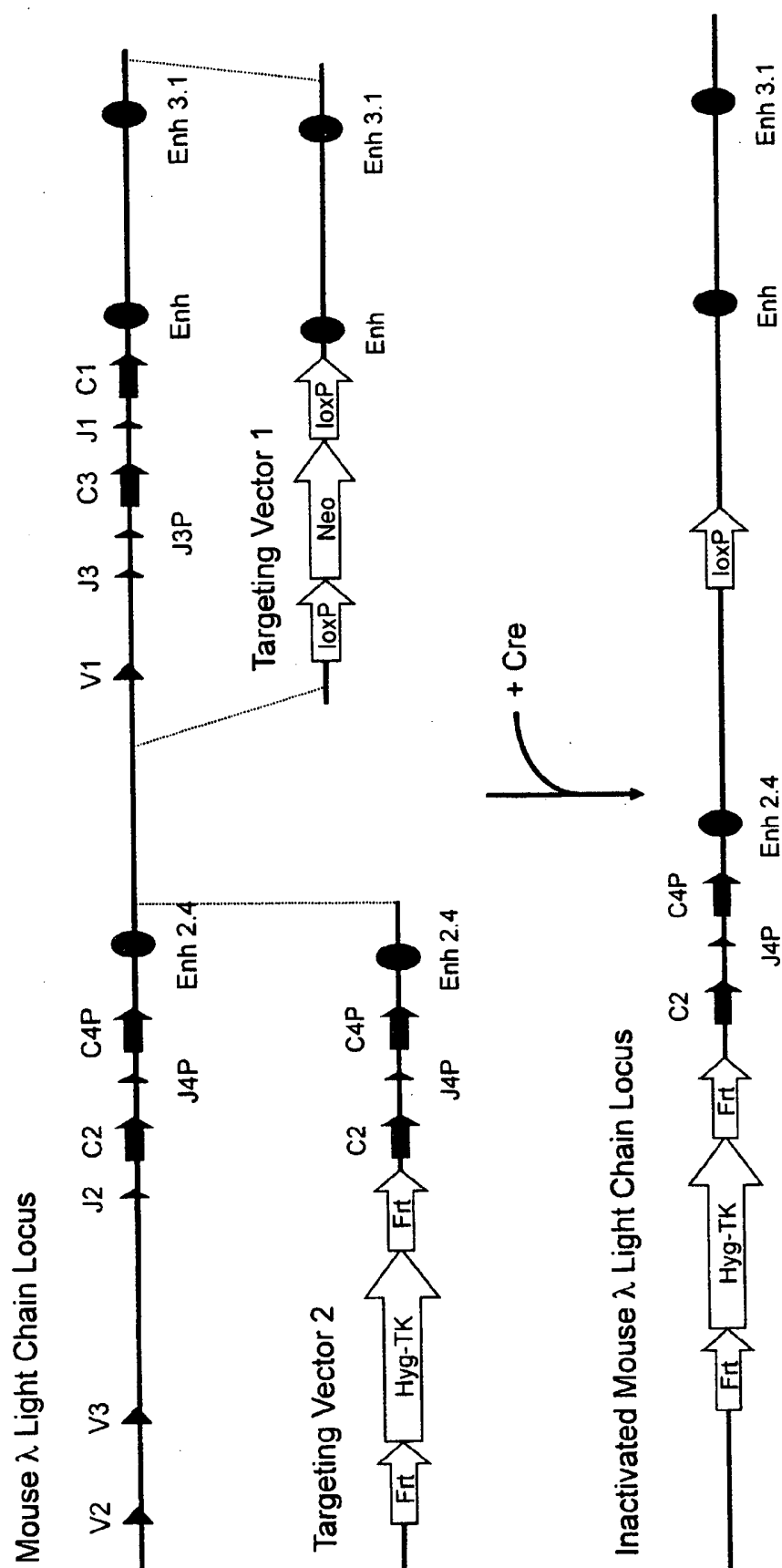


FIG. 2

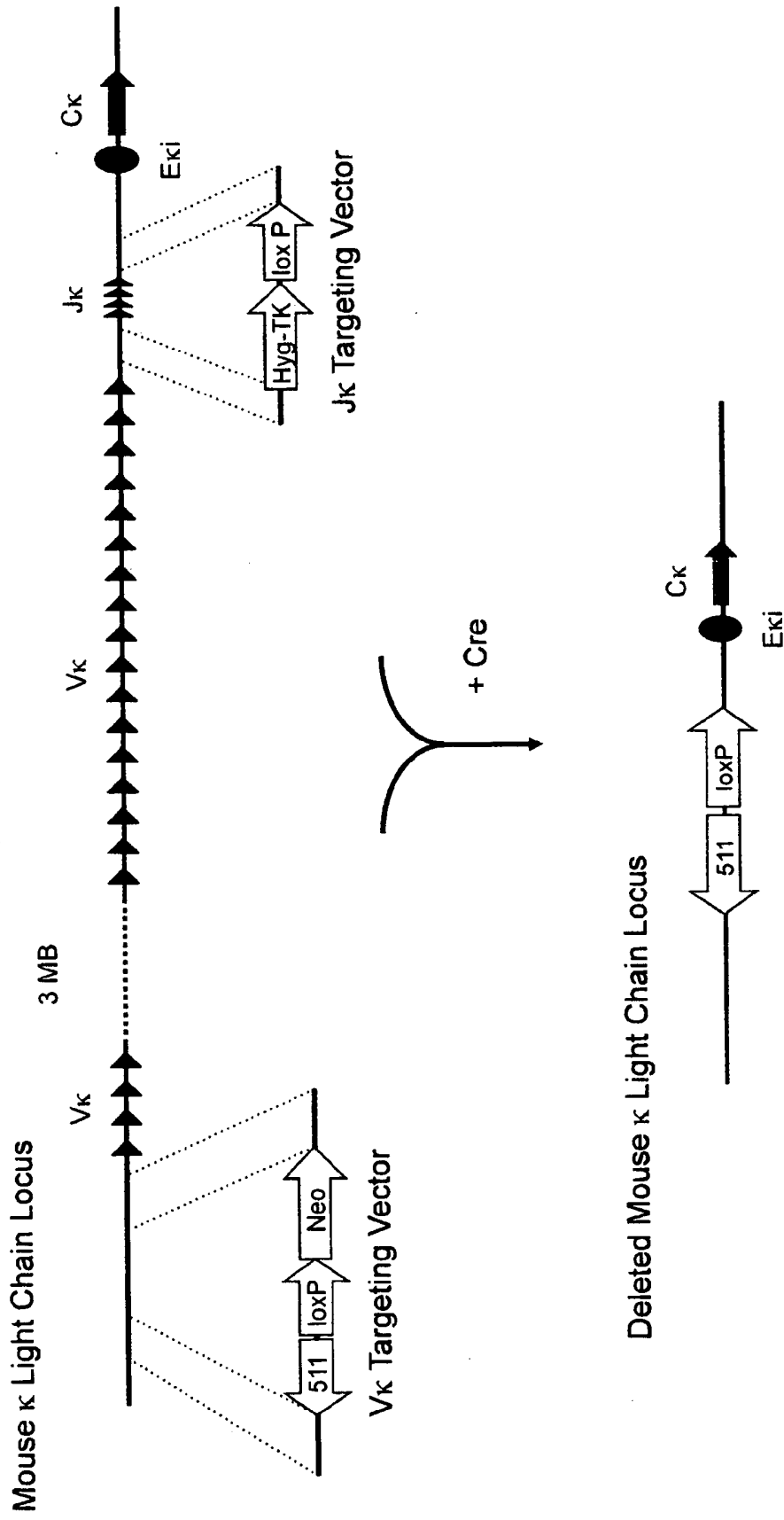


FIG. 3

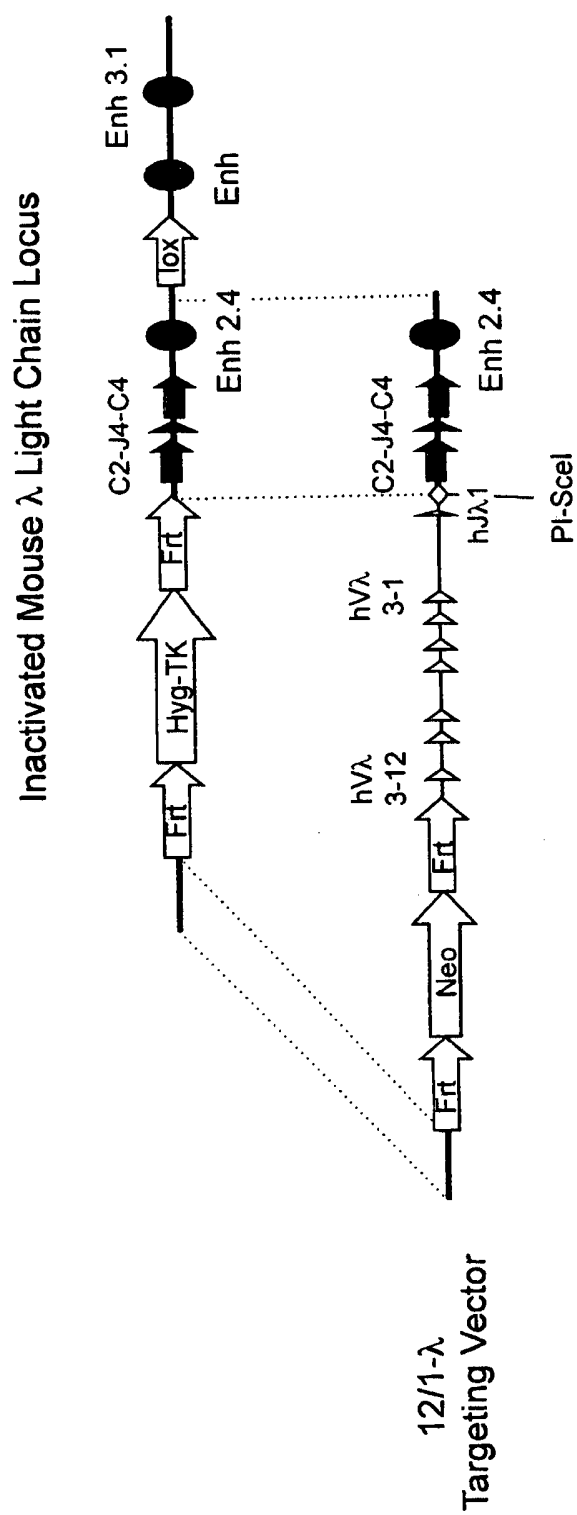


FIG. 4A

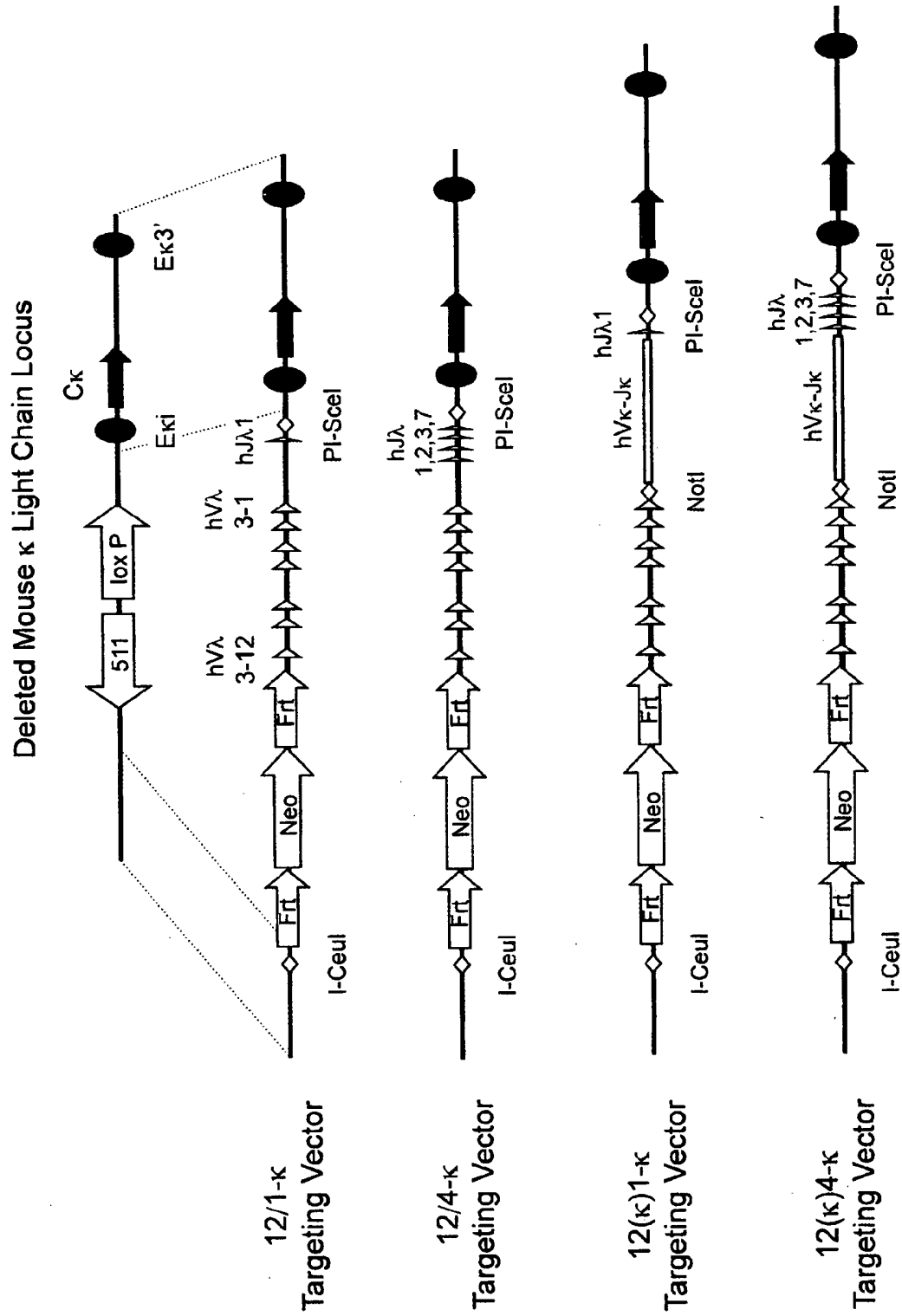


FIG. 4B

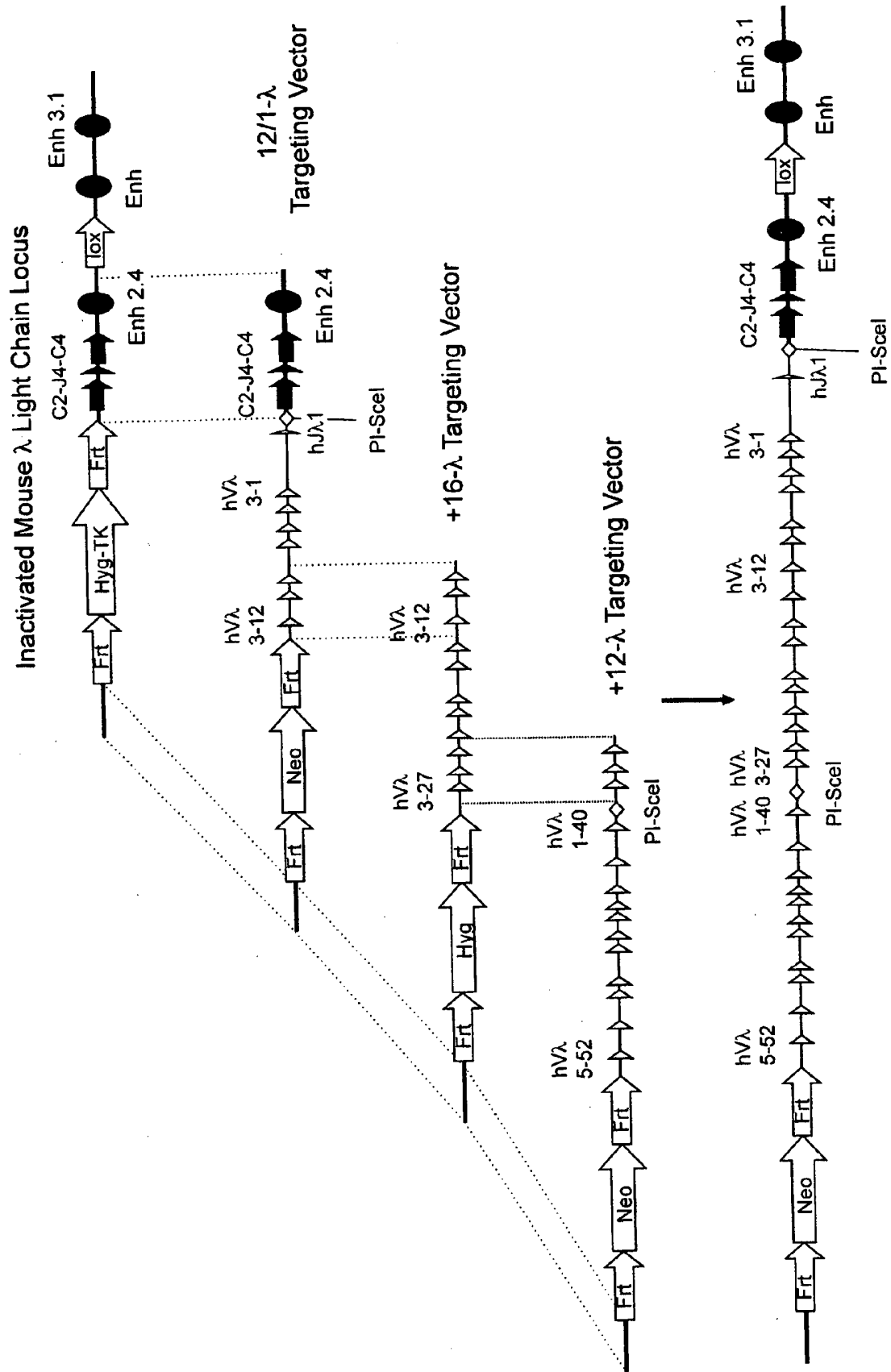


FIG. 5A

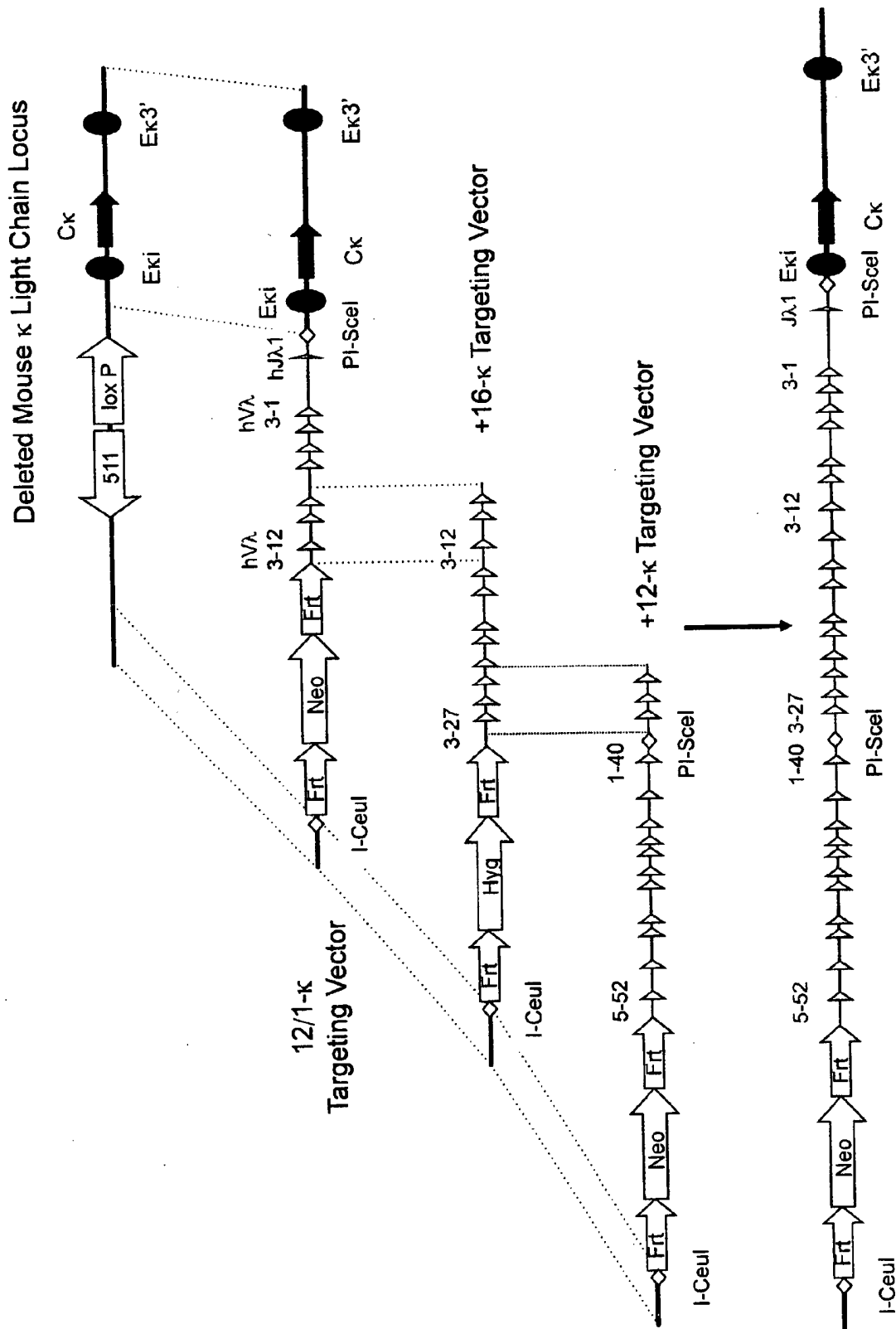


FIG. 5B

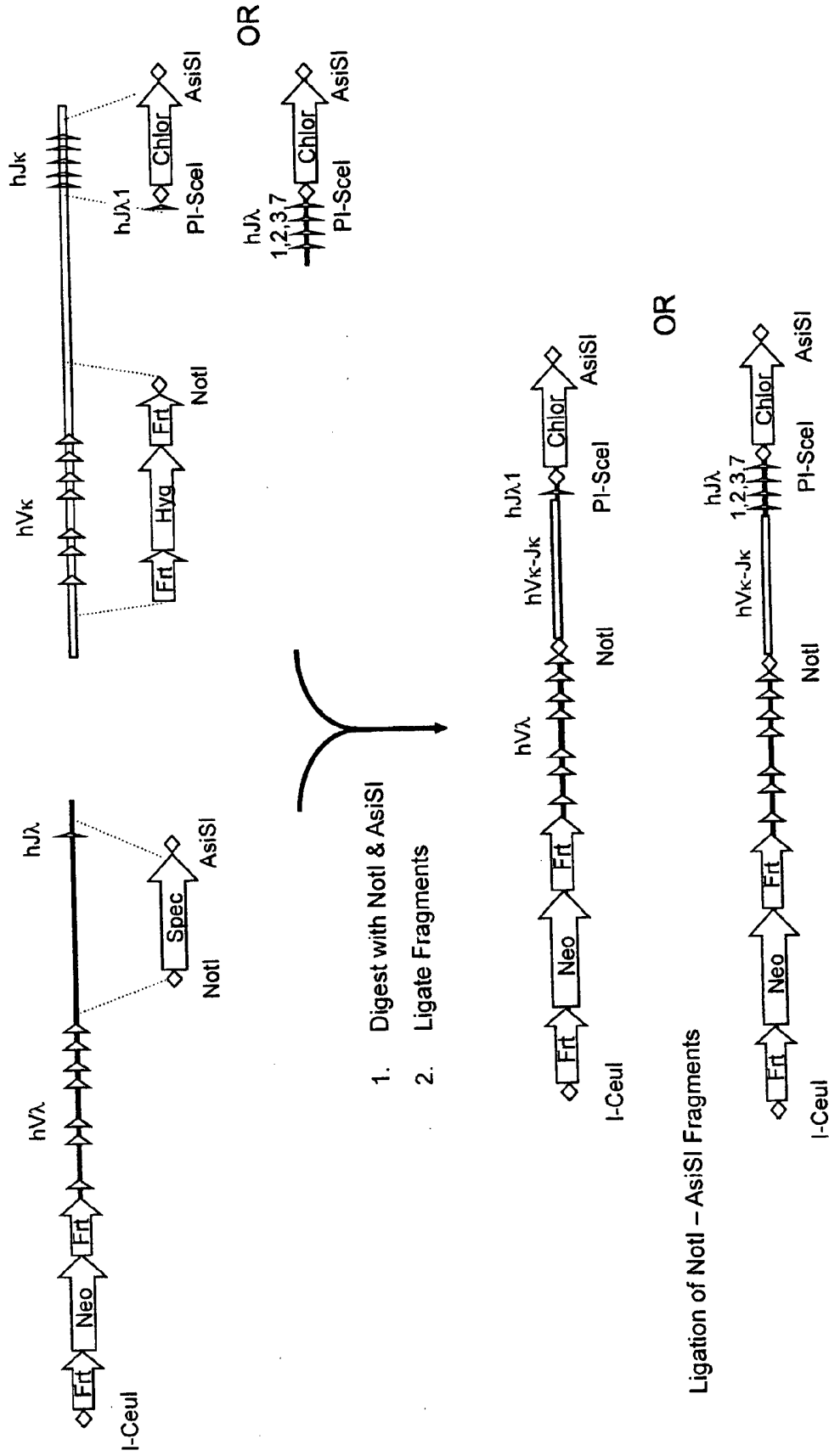


FIG. 6

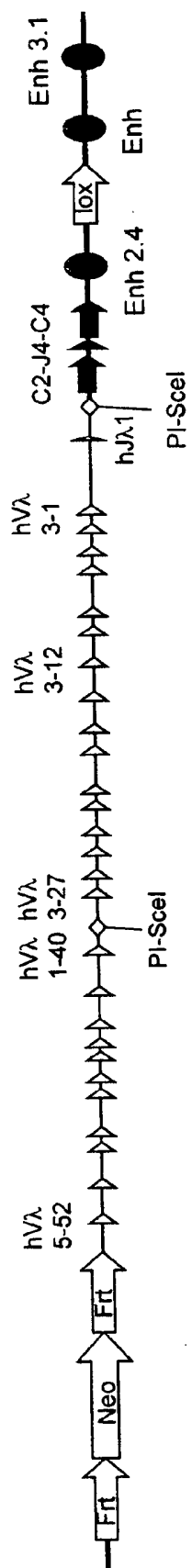
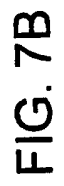


FIG. 7A



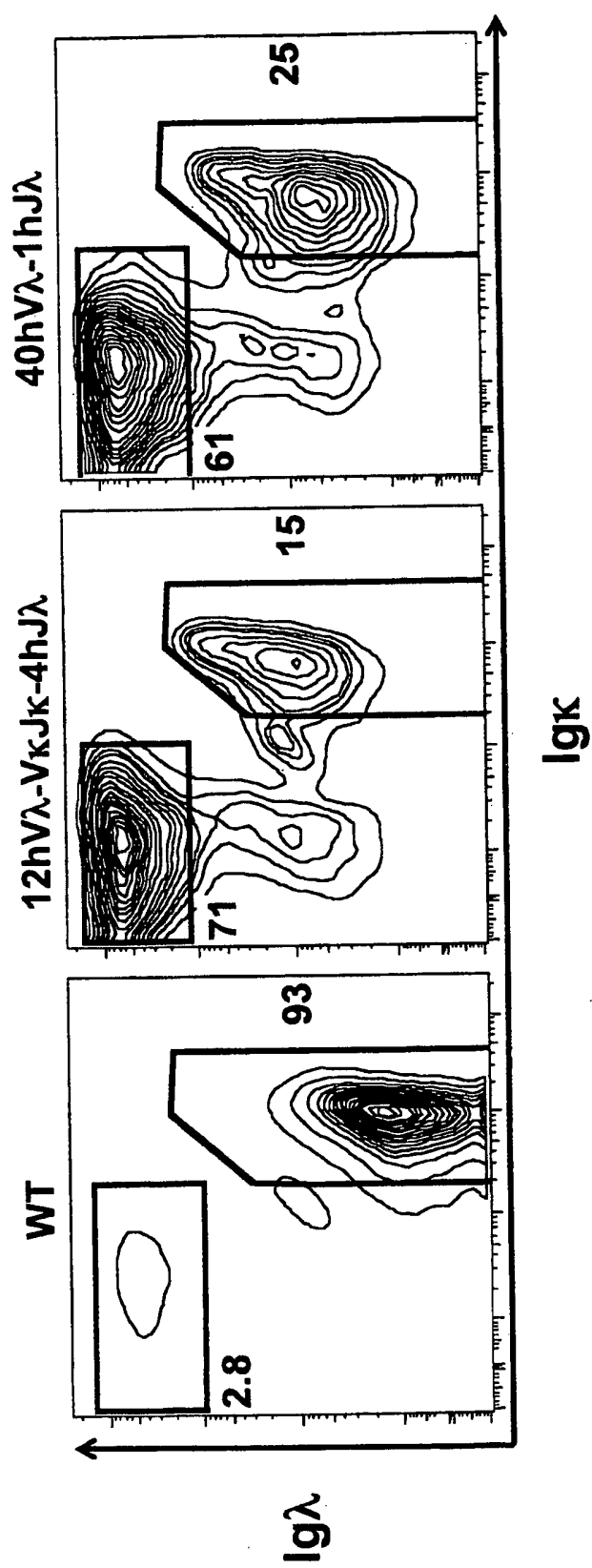


FIG. 8A

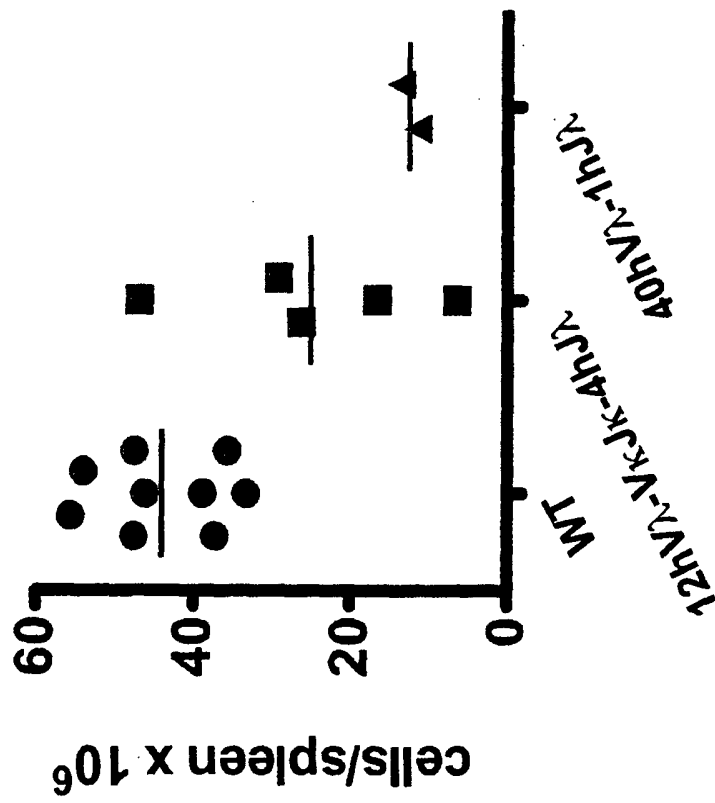


FIG. 8B

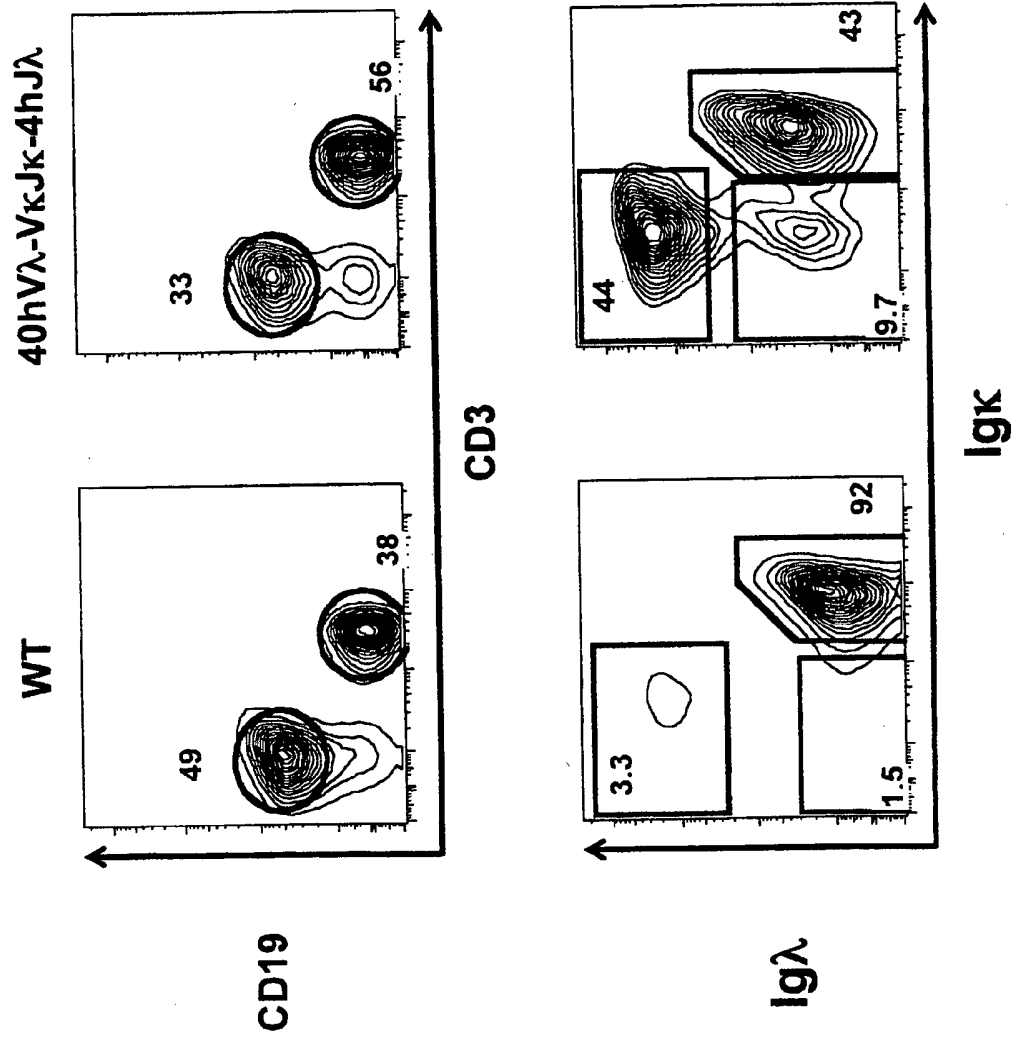


FIG. 9A

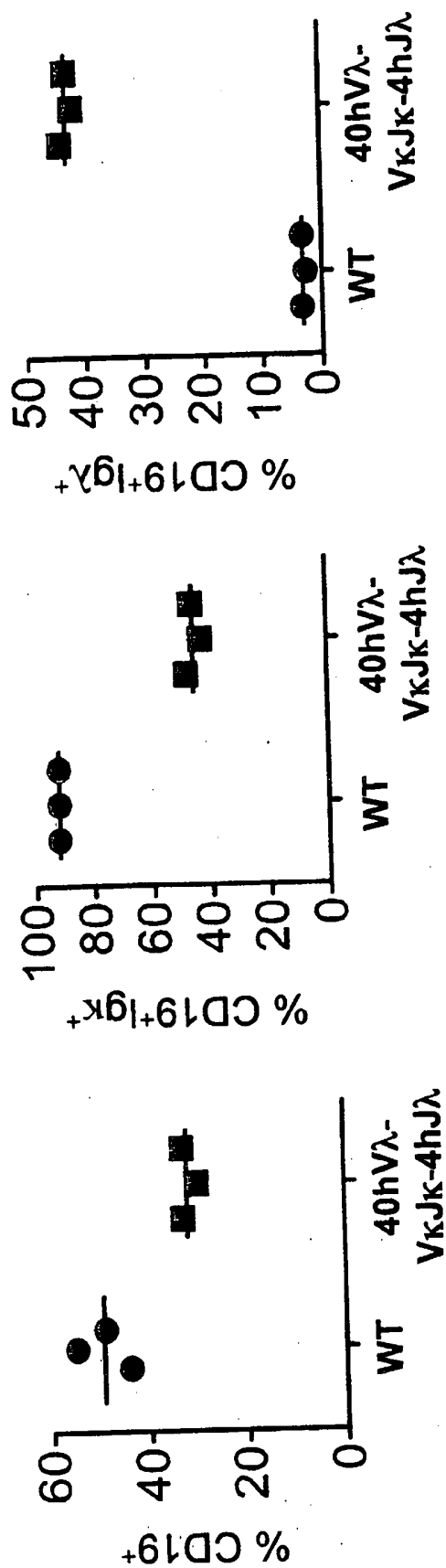


FIG. 9B

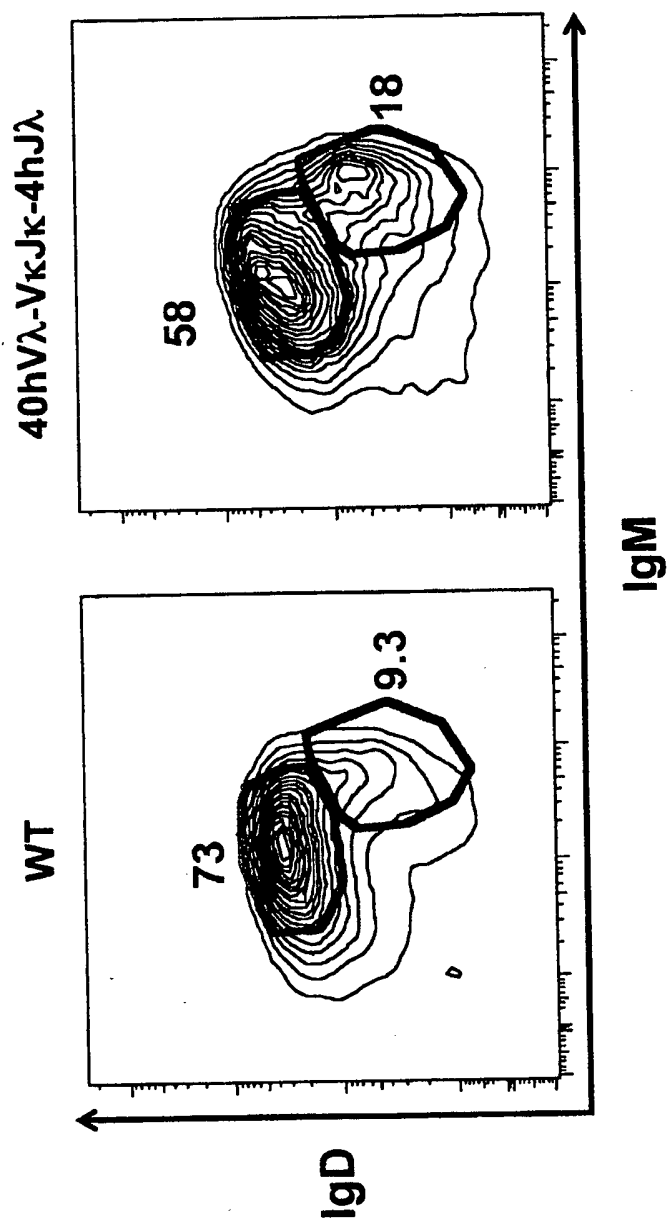


FIG. 9C

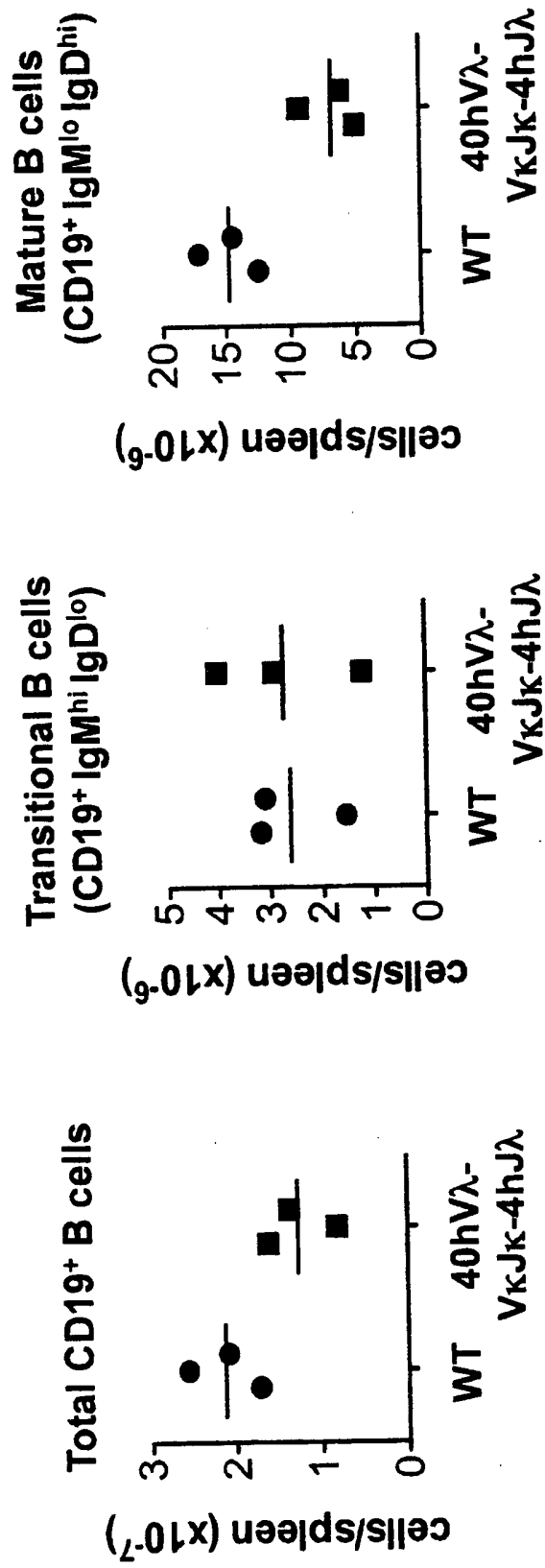


FIG. 9D

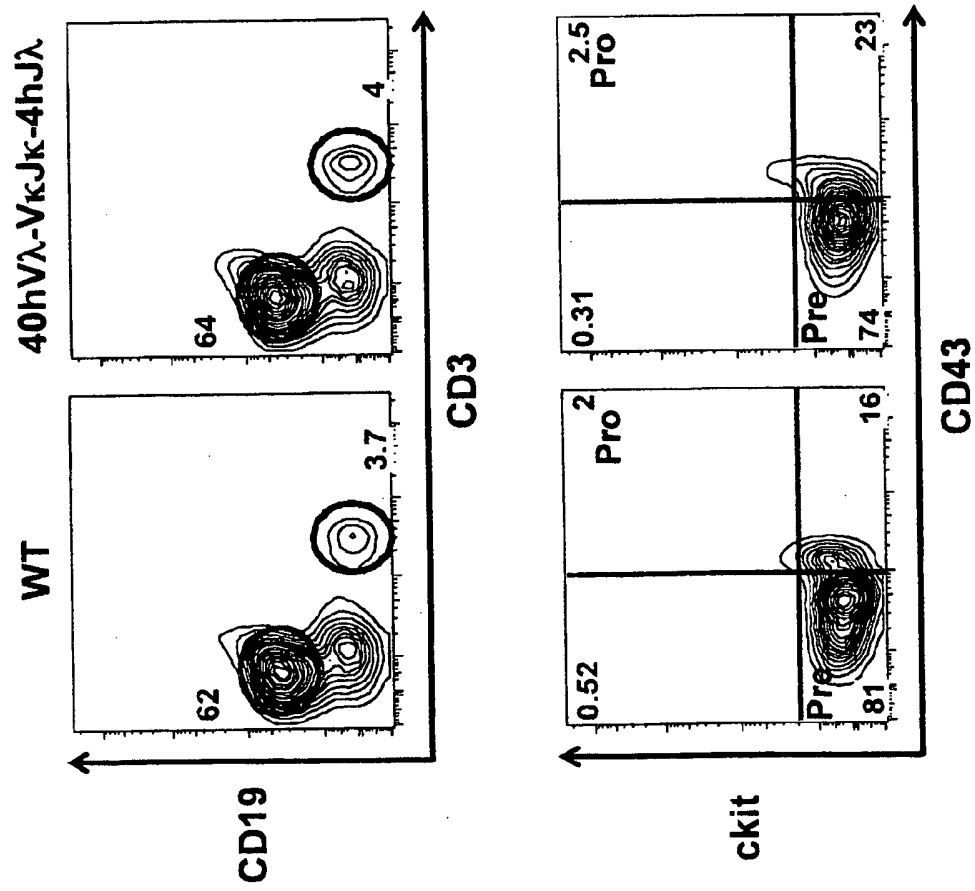


FIG. 10A

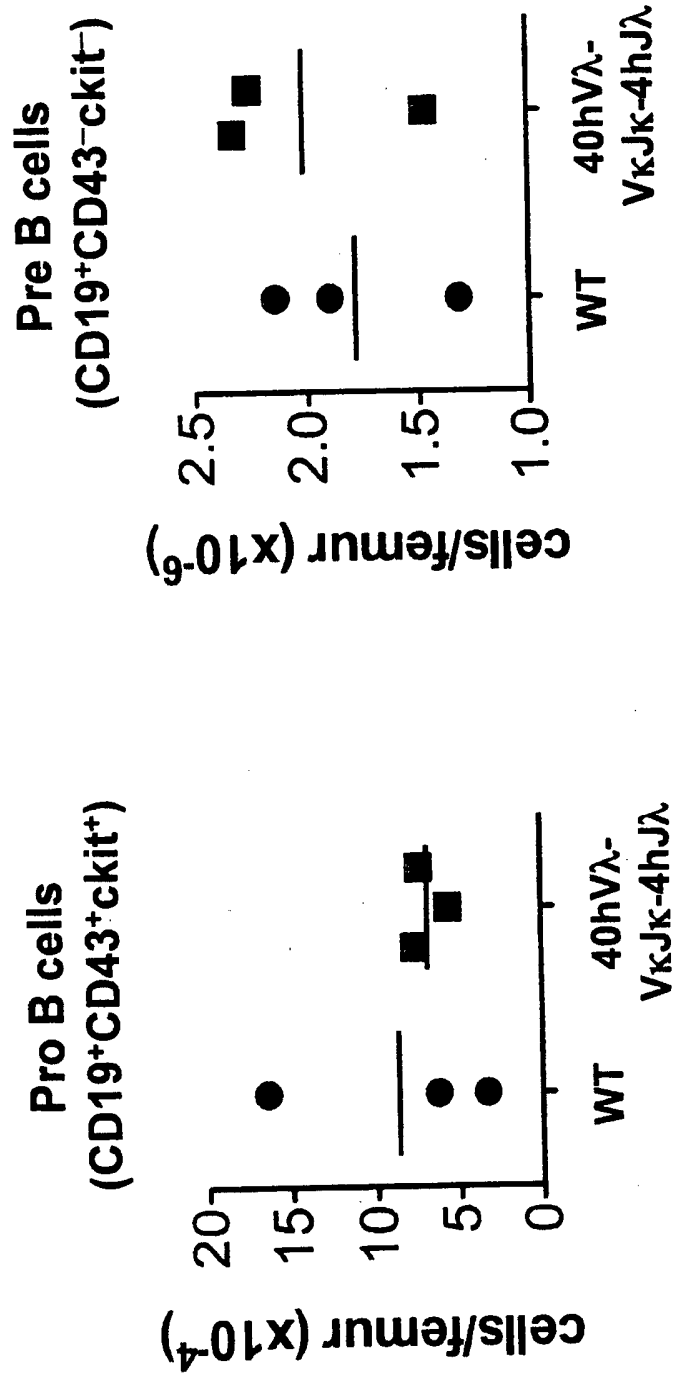


FIG. 10B

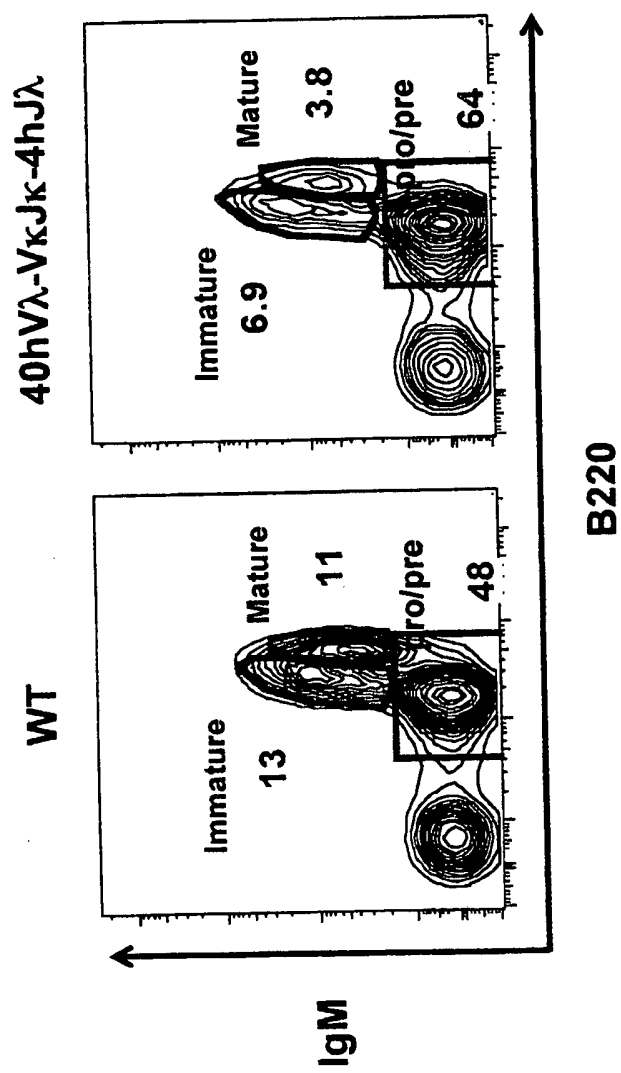


FIG. 10C

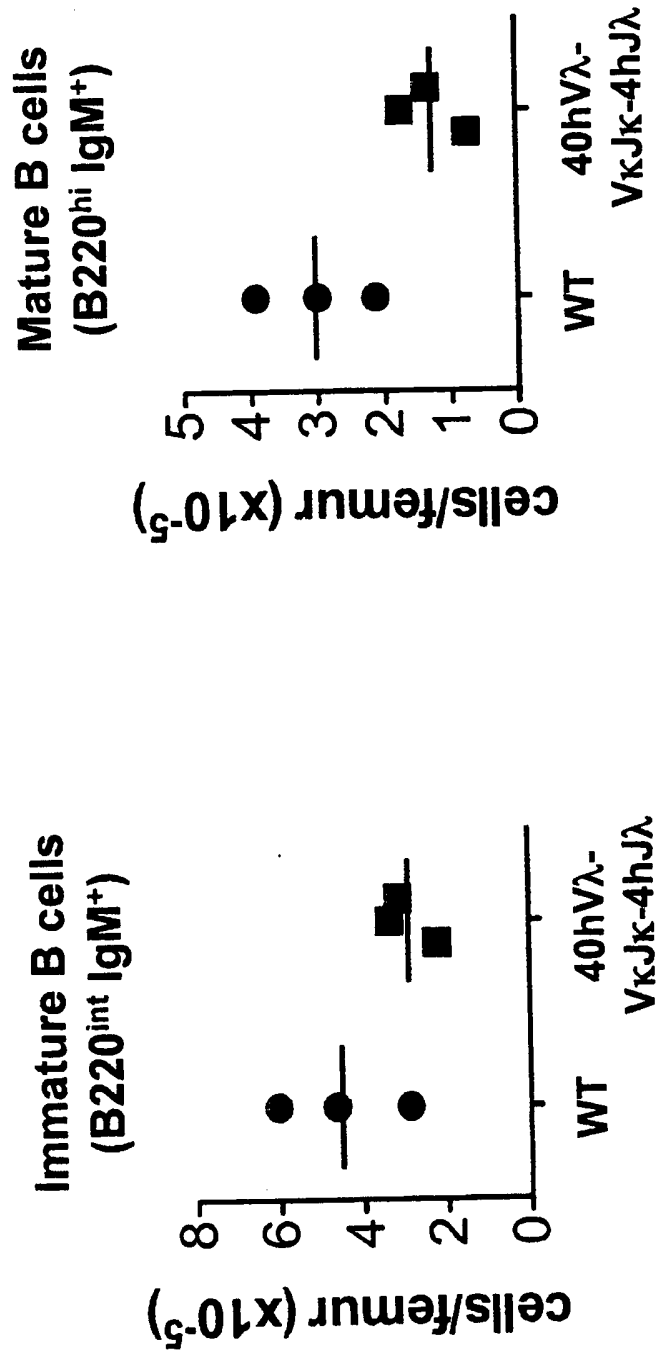


FIG. 10D

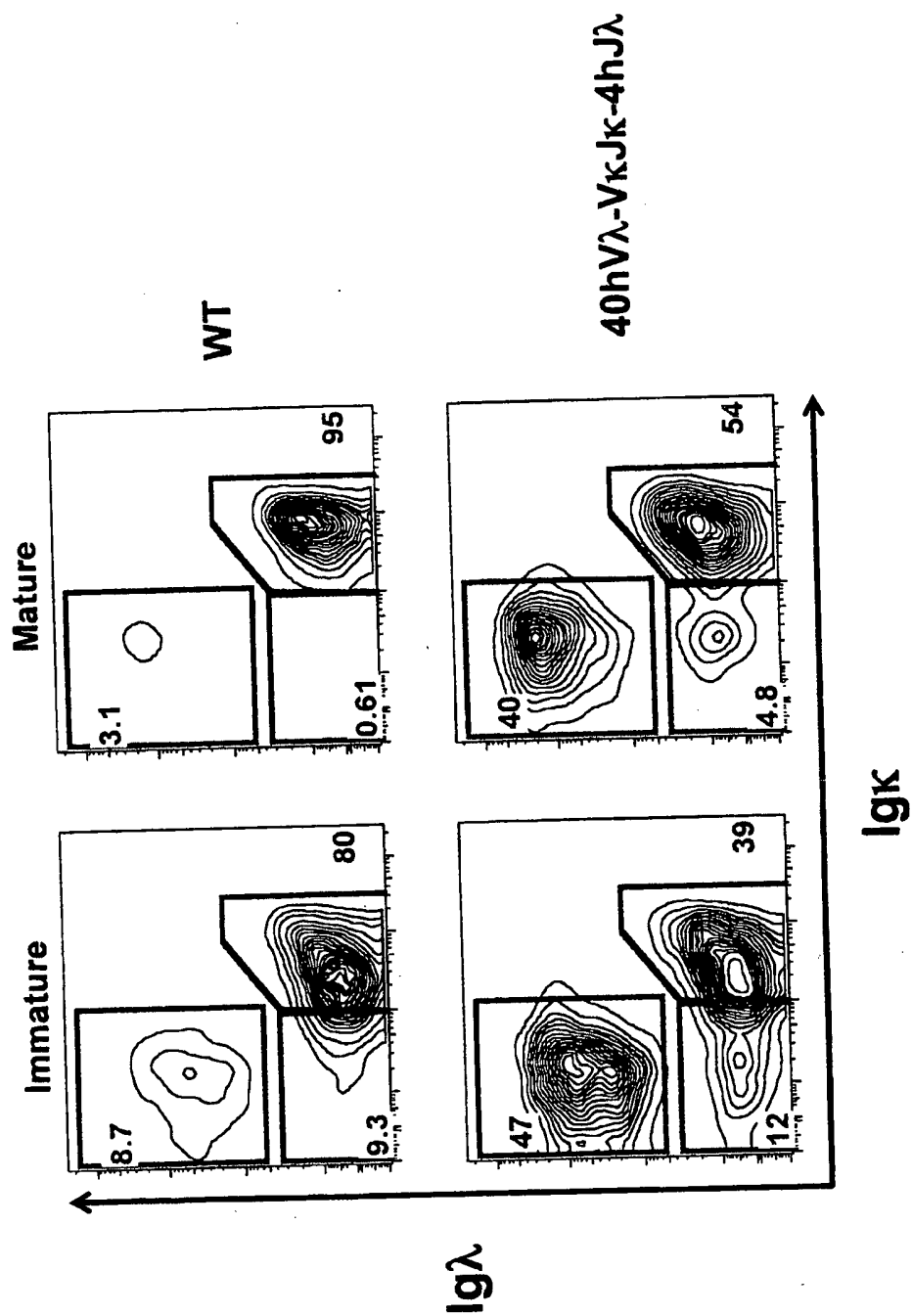


FIG. 10E

	3' Human V λ	Human J λ 1	5' Mouse C κ
A6	GCAACAATT	<u>tcgtcttcggaaactgggaccaaaggtcaccgctcctag</u>	<u>gggctgatgctgcaccaaactgtatccatcttc</u>
B6	GCAACAATT	atgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
F6	GCAACAATT	atgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
B7	GCAACAATT	atgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
E7	GCAACAAT	gtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
F7	GCAACAATT	atgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
C8	GCAACAATT	atgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
E12	CAAGTCGGTT	gtgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
1-4	TGAGTGCT	tatgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
1-20	TGAGTGCg	gcttttTTtGGAACTGGGACCAAGGTCACCGTCCCTAG	gggctgatgctgcaccaaactgtatccatcttc
3B43	CTGAATGGT	tatgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
5-8	AGTGATAAT	catgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
5-19	AGTGGTGCT	tatgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
1010	AGCAGCACT	tatgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
11A1	AGCAGCGCT	tatgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
7A8	GGTGGTGCT	tatgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
3A3	AGTAGCACT	tatgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
2-7	AGCAGCACT	tatgtcttcggaaactgggaccaaaggtcaccgctcctag	gggctgatgctgcaccaaactgtatccatcttc
FWR4		F G T G T K V T V L G A D A A P T V S I F	

FIG. 11

	3' Human V λ	Human J λ	5' Mouse C κ
5-2	CAGCCTGAGTGGTTC	TGTGTTCCGAGGAGGCACCCGGCTGACCGCCCTCG	GGGCTGATGCTGCACCAACTGTATCCATC
2-5	CAGCCTGAGTGGTT	ATGTCCTTCGGAACCTGGGACCAAGGTCACCGTCCTAG	GGGCTGATGCTGCACCAACTGTATCCATC
1-3	CAGCCTGAATGGT	GCTGTGTTCCGAGGAGGCACCCAGCTGACCGCCCTCG	GGGCTGATGCTGCACCAACTGTATCCATC
4B-1	CAGCCTGAGTGGTC	GGGTGTTCCGGCGGAGGGACCAAGCTGACCGTCCTAG	GGGCTGATGCTGCACCAACTGTATCCATC
3B-5	CAGCAGCACTGC	TGTGTTCCGAGGAGGCACCCAGCTGACCGCCCTCG	GGGCTGATGCTGCACCAACTGTATCCATC
7A-1	CAGCAGTGGTAAT	GCTGTGTTCCGAGGAGGCACCCAGCTGACCGCCCTCG	GGGCTGATGCTGCACCAACTGTATCCATC
5-1	CAGCAGTGGTAATCATAG	GGTGTTCCGGCGGAGGGACCAAGCTGACCGTCCTAG	GGGCTGATGCTGCACCAACTGTATCCATC
4A-1	CAGCCTGAGTGGTT	ATGTCCTTCGGAACCTGGGACCAAGGTCACCGTCCTAG	GGGCTGATGCTGCACCAACTGTATCCATC
11A-1	CAGCAGCGCT	GTGGTATTCCGGCGGAGGGACCAAGCTGACCGTCCTAG	GGGCTGATGCTGCACCAACTGTATCCATC
5-7	CTACTATGGTGGTGCTC	GGGTGTTCCGGCGGAGGGACCAAGCTGACCGTCCTAG	GGGCTGATGCTGCACCAACTGTATCCATC
5-4	CTCCTATAGTGGTGCTCGa	GTATTCCGGCGGAGGGACCAAGCTGACCGTCCTAG	GGGCTGATGCTGCACCAACTGTATCCATC
2-3	GAGCAACTTCGTGT	CTGTGTTCCGAGGAGGCACCCAGCTGACCGCCCTCG	GGGCTGATGCTGCACCAACTGTATCCATC
FWR4		F G G G T K L T V L G A D A A P T V S I	

FIG. 12

	3' Human V λ	Human J λ 1	5' Mouse C λ 2
2D1	<u>GCAGGCAGCAACAATTa</u>	<u>aGCTTTCGGAAC</u> TGGACCAAGGTCACCGTCCTAG	<u>GTCAGCCC</u> AAGTCCACTCCCCACTCTC
2D9	GACAGCAGTGGTAATCAT	TATGTCTTTCGGAACTGGACCAAGGTCACCGTCCTAG	GTCAGCCCAAGTCCACTCCCCACTCTC
3E15	GACAGCAGCACTGCC	GTCTTTCGGAACTGGACCAAGGTCACCGTCCTAG	GTCAGCCCAAGTCCACTCCCCACTCTC
FWR4		F G T G T K V T V L G Q P K S T P T L	

FIG. 13

REFERENCES CITED IN THE DESCRIPTION

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**EMBERI LAMBDA VARIÁBILIS RÉGIÓKAT ÉS EGÉR KONSTANS RÉGIÓKAT TARTALMAZÓ KÖNNYŰLÁNCOT
EXPRIMÁLÓ EGEREK
SZABADALMI IGÉNYPONTOK**

1. Egér, amelyben megtalálható egy endogén λ (lambda) könnyűlánc allél, amely tartalmaz egy újrapendezetlen emberi lambda variábilis (hVA) és egy újrapendezetlen emberi lambda kapcsoló (hJA) génszegmenst, ahol az egérben megtalálható az említett endogén egér λ könnyűlánc allél első VA-JA-CA génklaszterének deléciója és az említett emberi VA és JA génszegmenseket tartalmazó allél második VA-JA-CA génklaszterében lévő VA-JA génszegmensének - részben, vagy egészben történő - helyettesítésével, amelyek műtéti úton kapcsoltak az említett második VA-JA-CA génklaszter egy érintetlen egér lambda konstans (CA) régiójához, így az egér egy hVA, hJA és egy egér CA génből származtatott könnyűláncot exprimál, ahol az Enh 2.4, egér lambda 3' enhancer (Enh) és Enh 3.1 endogén enhancerek érintetlenek maradnak az említett λ könnyűlánc allélben.
2. Az egér az 1-es igénypont szerint, ahol az endogén λ (lambda) könnyűlánc allél több emberi VA génszegmenst tartalmaz.
3. Az egér az eddigi igénypontok bármelyike szerint, ahol az endogén λ (lambda) könnyűlánc allél rendelkezik:
 - a) legalább 12 újrapendezetlen emberi VA génszegmennel; vagy
 - b) 13-28 újrapendezetlen emberi VA génszegmennel.
4. Az egér a 3-as igénypont szerint, ahol az emberi VA génszegmensek közé értendő:
 - a) a hVA3-1, hVA4-3, hVA2-8, hVA3-8, hVA3-10, hVA2-11 és hVA3-12 szegmensek; vagy
 - b) a hVA2-14, hVA3-16, hVA2-18, hVA3-19, hVA3-21, hVA3-22, hVA2-23, hVA3-25 és hVA3-27 szegmensek.
5. Az egér az 1-es igénypont szerint, ahol az endogén λ (lambda) könnyűlánc allél egy újrapendezetlen JA génszegmenst tartalmaz, amely a hJA1.
6. Az egér az 1-es igénypont szerint, ahol az endogén λ (lambda) könnyűlánc allél négy újrapendezetlen emberi JA génszegmenst tartalmaz.
7. Az egér a 6-os igénypont szerint, ahol a négy újrapendezetlen emberi JA génszegmens a JA1, JA2, JA3 és JA7.
8. Az egér az 1-es igénypont szerint, ahol az endogén λ (lambda) könnyűlánc allélból hiányzik egy endogén VA génszegmens.
9. Az egér az 1-3, az 5-ös és 8-as igénypontok bármelyike szerint, ahol az endogén κ (kappa) könnyűlánc allél törölt.
10. Az egér az 1-es igénypont szerint, ahol az endogén egér λ könnyűlánc allél tartalmazza:
 - a) az emberi λ könnyűlánc lokusz egy folyamatos szekvenciáját, amely VA3-12-től a VA3-1-ig terjed;
 - b) az emberi λ könnyűlánc lokusz egy folyamatos szekvenciáját, amely VA3-29-től a VA3-1-ig terjed; vagy
 - c) egy az emberi λ könnyűlánc lokusz folyamatos szekvenciáját, amely VA3-29-től VA3-1-ig terjed és egy az emberi λ könnyűlánc lokusz folyamatos szekvenciáját, amely VA5-52-től a VA1-40-ig terjed.
11. Izolált sejt, amely egy hVA, hJA és egy CA egér génből származtatott könnyűláncot exprimál, ahol a sejt az eddigi igénypontok bármelyike szerint meghatározott egérből származik vagy abból kinyerhető, és ahol a sejtben megtalálható egy újrapendezett emberi VA génszegmens és egy újrapendezett emberi JA génszegmens, és rendelkezik az említett endogén egér λ könnyűlánc allél első VA-JA-CA génklaszterének deléciójával, és az említett emberi VA és JA génszegmenseket tartalmazó allél második VA-JA-CA génklaszterében lévő VA-JA génszegmensének - részben, vagy egészben történő - helyettesítésével, amelyek műtéti úton kapcsoltak az említett második VA-JA-CA génklaszter egy érintetlen egér CA konstans régióhoz, és ahol az Enh 2.4, egér lambda 3' enhancer (Enh) és Enh 3.1 endogén enhancerek érintetlenek maradnak az említett λ könnyűlánc allélben.
12. Az izolált sejt a 11-es igénypont szerint, ahol a sejt egy B-limfocita.
13. Izolált egér embrionális őssejt (ES), amelyben megtalálható egy endogén λ (lambda) könnyűlánc allél, amely tartalmaz egy újrapendezetlen emberi VA és egy újrapendezetlen emberi JA génszegmenst, ahol a sejtben megtalálható az említett endogén egér λ könnyűlánc allél első VA-JA-CA génklaszterének deléciója és az említett emberi VA és JA génszegmenseket tartalmazó

allél második V λ -J λ -C λ génklaszterében lévő V λ -J λ génszegmensének - részben, vagy egészben történő - helyettesítése, amelyek műtéti úton kapcsolak az említett második V λ -J λ -C λ génklaszter egy érintetlen egér C λ konstans régióhoz, ahol az Enh 2.4, egér lambda 3' enhancer (Enh) és Enh 3.1 endogén enhancerek érintetlenek maradnak az említett λ könnyűlánc allélban.

14. Egér embrió, amely a 13-as igénypont szerint meghatározott összejet tartalmazza, vagy abból fejlődött ki, vagy abból kinyerhető, ahol az egér embrióban megtalálható egy endogén λ (lambda) könnyűlánc allél, amely tartalmaz egy újraprendezett emberi V λ és egy újraprendezett emberi J λ génszegmenst, ahol a sejtből megtalálható az említett endogén egér λ könnyűlánc allél első V λ -J λ -C λ génklaszterének deléciója és az említett emberi V λ és J λ génszegmenseket tartalmazó allél második V λ -J λ -C λ génklaszterében lévő V λ -J λ génszegmensének - részben, vagy egészben történő - helyettesítése, amelyek műtéti úton kapcsolak az említett második V λ -J λ -C λ génklaszter egy érintetlen egér C λ konstans régióhoz, és ahol az Enh 2.4, egér lambda 3' enhancer (Enh) és Enh 3.1 endogén enhancerek érintetlenek maradnak az említett λ könnyűlánc allélban.

15. A 12-es igénypont szerint meghatározott B-limfocita olyan hibridóma sejt létrehozásához, amely egy hV λ , hJ λ és egy egér C λ génből származtatott könnyűláncot exprimál, ahol a hibridóma sejt az 1-10 igénypontok bármelyike szerint meghatározott egérből származik vagy abból kinyerhető, ahol a sejtből megtalálható egy újraprendezett emberi V λ génszegmens és egy újraprendezett emberi J λ génszegmens, és az említett endogén egér λ könnyűlánc allél első V λ -J λ -C λ génklaszterének deléciója és az említett emberi V λ és J λ génszegmenseket tartalmazó allél második V λ -J λ -C λ génklaszterében lévő V λ -J λ génszegmensének - részben, vagy egészben történő - helyettesítésével, amelyek műtéti úton kapcsolak az említett második V λ -J λ -C λ génklaszter egy érintetlen egér C λ konstans régióhoz, és ahol az Enh 2.4, egér lambda 3' enhancer (Enh) és Enh 3.1 endogén enhancerek érintetlenek maradnak az említett λ könnyűlánc allélban.

16. Hibridóma sejt, amely egy hV λ , hJ λ és egy C λ egér génből származtatott könnyűláncot exprimál, ahol a hibridóma sejt az 1-10 igénypontok bármelyike szerint meghatározott egérből származik vagy abból kinyerhető, ahol a sejtből megtalálható egy újraprendezett emberi V λ génszegmens és egy újraprendezett emberi J λ génszegmens, és az említett endogén egér λ könnyűlánc allél első V λ -J λ -C λ génklaszterének deléciója és az emberi V λ és J λ génszegmenseket tartalmazó allél második V λ -J λ -C λ génklaszterében lévő V λ -J λ génszegmensének - részben, vagy egészben történő - helyettesítése, amelyek műtéti úton kapcsolak az említett második V λ -J λ -C λ génklaszter egy érintetlen egér C λ konstans régióhoz, és ahol az Enh 2.4, egér lambda 3' enhancer (Enh) és Enh 3.1 endogén enhancerek érintetlenek maradnak az említett λ könnyűlánc allélban.

17. Módszer antitest létrehozására egérben, amely tartalmazza:

- a) egér kitételét egy antigénnek az 1-es igénypont szerint;
- b) az egér általi, az antigénre adott immunválasz kifejlődésének lehetővé tételét; és
- c) egy antitest leválasztását a (b) szerinti egérből, amely kifejezetten az antigént ismeri fel, vagy egy antitest leválasztását a (b) szerinti egérből, amely tartalmaz egy immunoglobulin domént, amely kifejezetten az antigént ismeri fel, vagy egy olyan nehézlánc és/vagy könnyűlánc variábilis domént kódoló nukleinsav-szekvencia azonosítását a (b) szerinti egérben, amely kötődik az antigénhez, ahol az antitest egy hV λ , hJ λ és egy C λ egér génből származtatott könnyűláncot tartalmaz.