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(54) Title: MEDICAL COMPOSITIONS FOR INTRAVESICAL TREATMENT OF BLADDER CANCER

(57) Abstract: Anti-cancer coating compositions comprising 3-hydroxymethyl-5-aziridinyl-1-1-methyl-2-[1H-indole-4,7-dione]propenol (E09) are disclosed. More specifically, the coating compositions comprise E09 and a formulation vehicle. The formulation vehicle improves the solubility and stability of E09. Additionally, the coating compositions can include coating agents that provide better adhesion of the coating composition to the bladder wall during intravesical delivery of the coating composition.



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MEDICAL COMPOSITIONS FOR INTRAVESICAL TREATMENT OF BLADDER CANCER

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Application No. 60/344,446, filed November 1, 2001, and whose entire contents are hereby incorporated by reference.

BACKGROUND OF THE INVENTION

Bladder cancer accounts for approximately 2% of all malignant cancers and is the fifth and tenth most common cancer in men and women, respectively. The American Cancer Society estimated that 54,500 new cases and 11,700 deaths would have occurred in 1997. Superficial bladder cancers (pTa, pT1 and CIS) account for 70-80% of cancers at first presentation. Management of superficial bladder cancer may be achieved by endoscopic surgical resection often followed by a course of adjuvant intravesical chemotherapy or immunotherapy with the aim of both eradicating remaining tumor cells and preventing tumor recurrence (Herr HW (1987) Intravesical therapy—a critical review. *Urol Clin N Am* 14:399-404). Both anti-neoplastics (Mitomycin C [MMC], epirubicin and thioTEPA) and immunotherapy (BCG) administered intravesically are effective at reducing tumor recurrence rates although it is unclear whether disease progression to muscle invasive tumors is prevented (Newling D (1990) Intravesical therapy in the management of superficial transitional cell carcinoma of the bladder: the experience of the EORTC GU group, *Br J Cancer* 61:497-499; Oosterlink et al. (1993) A prospective European Organization for Research and Treatment of Cancer Genitourinary Group randomized trial comparing transurethral resection followed by a single instillation of epirubicin or water in single stage Ta, T1 papillary carcinoma of the bladder. *J Urol* 149:749-752). This observation in conjunction with the fact that mortality from bladder cancer is still high underscores the need to develop more effective therapeutic agents (Oosterlink et al. 1993).

One such therapeutic agent is MMC which belongs to a class of compounds known as bioreductive drugs (Workman 1994). MMC represents one of the antineoplastic agents used to treat superficial bladder cancers (Maffezzini et al, 1996, Tolley et al, 1996). MMC is activated to a cytotoxic species by cellular reductases

although the role of specific reductase enzymes involved in bioreductive activation remains poorly defined and controversial (Cummings et al, 1998a). This is particularly true for the enzyme NQO1 (NAD(P)H:Quinone oxidoreductase, EC 1.6.99.2) which is a cytosolic flavoprotein which catalyses the two electron reduction of various quinone based compounds using either NADH or NADPH as electron donors (Schlager and Powis, 1988, Siegel et al, 1990). The structurally related compound EO9 (5-aziridinyl-3-hydroxymethyl-1-methyl-2-[1H-indole-4,7-dione]prop-(3-en-a-ol), is however a much better substrate for NQO1 than MMC (Walton et al, 1991) and a good correlation exists between NQO1 activity and chemosensitivity in vitro under aerobic conditions (Robertson et al, 1994, Fitzsimmons et al, 1996, Smitkamp-Wilms et al, 1994). Under hypoxic conditions however, EO9's properties are markedly different with little or no potentiation of EO9 toxicity observed in NQO1 rich cells (Plumb and Workman, 1994). In NQO1 deficient cell lines however, large hypoxic cytotoxicity ratios have been reported (Workman, 1994). Therefore, EO9 has the potential to exploit the aerobic fraction of NQO1 rich tumors or the hypoxic fraction of NQO1 deficient tumors (Workman, 1994).

EO9 has been clinically evaluated but despite reports of three partial remissions in phase I clinical trials, no activity was seen against NSCLC, gastric, breast, pancreatic and colon cancers in subsequent phase II trials (Schellens et al, 1994, Dirix et al, 1996). These findings are particularly disappointing in view of the preclinical studies (Hendriks et al, 1993) together with reports that several tumor types have elevated NQO1 levels (Malkinson et al, 1992, Smitkamp-Wilms et al, 1995, Siegel et al, 1998). Several possible explanations have been proposed to explain EO9's lack of clinical efficacy (Connors, 1996, Phillips et al, 1998). Recent studies have demonstrated that the failure of EO9 in the clinic may not be due to poor pharmacodynamic interactions but may be the result of poor drug delivery to tumors (Phillips et al, 1998). The rapid plasma elimination of EO9 ($t_{1/2} = 10$ min in humans) in conjunction with poor penetration through multicell layers suggests that EO9 will not penetrate more than a few microns from a blood vessel within its pharmacokinetic lifespan (Schellens et al, 1994, Phillips et al, 1998). Intratumoural administration of EO9 to NQO1 rich and deficient tumors produced significant growth delays (although a distinction between damage to the aerobic or hypoxic fraction was not determined) suggesting that if EO9 can be delivered to tumors, therapeutic effects may be achieved (Cummings et al, 1998b). While these undesirable characteristics are a serious setback for the treatment of systemic disease, paradoxically they may be

advantageous for treating cancers which arise in a third compartment such as superficial bladder cancer. In this scenario, drug delivery is not problematical via the intravesical route and the penetration of EO9 into avascular tissue can be increased by maintenance of therapeutically relevant drug concentrations within the bladder (using a one hour instillation period for example). While this method of instilling EO9 within the bladder may be useful, there still remains a need for drug delivery vehicles that are capable of delivering an effective amount of EO9 within the bladder.

BRIEF SUMMARY OF THE INVENTION

In a broad aspect, the present invention is directed to compositions for treating cancer. More specifically, the compositions of the present invention comprise pharmaceutical products formulated for intravesical instillation to treat bladder cancer. The pharmaceutical products comprise bioreductive alkylating indoloquinone with anti-tumor effects such as, but not limited to, 3-hydroxymethyl-5-aziridinyl-1-1-methyl-2-[1H-indole-4,7-dione]propenol (EO9) and a formulation vehicle. The formulation vehicles of the present invention improves the physical characteristics of the solution such as solubility, lyophilization, and ease of reconstitution of the lyophilized solution.

According to one embodiment of the present invention, the composition of the present invention comprises 3-hydroxymethyl-5-aziridinyl-1-1-methyl-2-[1H-indole-4,7-dione]propenol (EO9) and a formulation vehicle. According to one embodiment, the formulation vehicle is a mixture of tert-butanol and water. In another embodiment, the formulation vehicle is a mixture of ethanol and water. In yet another embodiment, the formulation vehicle is 2-hydroxypropyl- β -cyclodextrin. These composition embodiments of the present invention can be lyophilized by techniques known or developed in the art. The lyophilized compositions of the present invention are

According to another embodiment of the present invention, the composition of the present invention comprises EO9 and a coating agent. The coating agent allows for better adhesion of the composition to the bladder wall. Consequently, the composition and, in particular, the EO9 contacts and may be able to penetrate the avascular tissue that comprises for a time sufficient to treat the bladder cancer. In one embodiment of the present invention, the coating agent is propylene glycol. In other exemplary embodiments of the present invention, the coating agent can be selected from the group consisting of hydroxypropylcellulose, carboxymethylcellulose, chitosan hydrochloride, lectin, or polycarbophil. In yet another embodiment of the present invention, the compositions of the present invention can be delivered to the bladder

wall by a liposome. In another embodiment, the compositions of the present invention can be delivered to the bladder wall by a microsphere. In another embodiment, the compositions of the present invention can be delivered to a patient intravenously.

BRIEF DESCRIPTION OF DRAWINGS

Figure 1 . Validation of the polyclonal anti-rat NQO1 antibody for use in immunohistochemical analysis of human NQO1. Panel A: Western blot analysis of cell extracts (12.5 μ g protein loaded per lane) for NQO1. Lanes 1-5 represent extracts from DLD-1 (794 ± 121 nmol/min/mg), HT-29 (688 ± 52 nmol/min/mg), H460 (1652 ± 142 nmol/min/mg), MT1 (287 ± 53 nmol/min/mg), and RT112 (30 ± 3 nmol/min/mg) respectively where the values in parenthesis represent NQO1 activity. Lane 6 represents molecular weight markers (ECL protein molecular weight markers, Amersham Pharmacia Biotech, UK). Panel B: Western blot analysis using purified human recombinant NQO1. Lanes 1-5 represent protein amounts of 0.25, 0.125, 0.0625, 0.0312 and 0.0156 pmol respectively. Panel C: Western blot analysis of cell extracts (25 μ g protein loaded per lane) derived from H460 cells (lanes 1-2) and BE cells (lanes 3-4).

Figure 2. Immunohistochemical localization of NQO1 in human bladder tumors, normal bladder, urethra and ureter. Tumors (panels A,B and C) were classified as G2 pTa (panel A, [x 200]) and G3 pT2 (panels B [x 100]) and G3 pT4 (panel C [x 200]) which had high to intermediate levels of NQO1 activity as determined by biochemical methods. Panel D (x100) represents a histological section through a macroscopically normal looking section of bladder from a patient who underwent cystectomy for a G3 pT4 tumor ; no tumor was identified in these sections but some inflammatory change was evident. Panels E and F (x200) represent urethra and ureter with no evidence of invasive or in situ carcinoma in these sections. All sections have been stained with NQO1 antibody. Negative staining (without primary antibody) were clear (data not shown).

Figure 3. The relationship between NQO1 activity and the response of a panel of cell lines to E09 (panel A) or MMC (panel B) under normal physiological pH of 7.4 (o) or acidic pH values of 6.0 (•). Regression analysis data (as determined by Sigma Plot graphics) for E09 at pH 7.4 were $r = 0.886$, slope = -0.52 and at pH 6.0, regression analysis data for E09 was $r = 0.804$ and slope = -0.51. For MMC, regression analysis at pH 7.4 was $r = 0.849$, slope = -0.19 and at pH 6.0, $r = 0.609$, slope = -0.23.

Figure 4. Response of HT-29 multicell spheroids following a one hour exposure to EO9 under acidic (pHe = 6.0, 0) and physiological (pHe = 7.4, 0) extracellular pH conditions. Values presented are the means of 3 independent experiments \pm standard deviation.

DETAILED DESCRIPTION OF THE INVENTION

The embodiments of the present invention are directed to compositions for treating bladder cancer via intravesical instillation. According to one embodiment, the composition of the present invention comprises 3-hydroxymethyl-5-aziridinyl-1-1-methyl-2-[1H-indole-4,7-dione]propenol (EO9) and a formulation vehicle. The formulation vehicles of the present invention are solvents that improves the solubility and stability of EO9. In a broad aspect of the present invention, the formulation vehicles of the present invention can be a mixture of an alcohol and water. According to the various embodiments of the present invention, EO9 dissolves in the formulation vehicles without physical manipulation such as grinding. Because the compositions of the present invention are capable of dissolving greater amounts of EO9, additional flexibility with respect to dosage units is achieved. According to one embodiment, a content of 8.0 mg of EO9 per dosage unit is contemplated. In other embodiments, instillation doses range from approximately 0.5 mg to approximately 16 mg in a total volume of 40 mL.

In addition to improving the solubility of EO9, the formulation vehicles of the present invention are good lyophilization vehicles. For example, the formulation vehicles of the present invention minimizes the time to lyophilize the compositions of the present invention. Accordingly, in one embodiment of the present invention, it is possible to lyophilize the compositions of the present invention in less than approximately 4.5 days. Furthermore, the compositions of the present invention are stable after undergoing lyophilization (see table 4). It is believed that the formulation vehicles of the present invention minimize the crystallization of EO9 during the lyophilization process. Consequently, by reducing the amount of crystallization of EO9, a smaller volume of fluid is required to reconstitute the compositions of the present invention. As a result, a larger batch size can be achieved due to the reduced reconstitution volumes for the lyophilized composition.

According to one embodiment, the composition of the present invention comprises EO9 and a formulation vehicle comprising tert-butanol. According to another embodiment of the present invention, the formulation vehicle comprises

mixture of ethanol and water. In yet another embodiment, the formulation vehicle is 2-hydroxypropyl- β -cyclodextrin. In one exemplary embodiment, the formulation vehicle comprises 40% tert-butanol in water. As those skilled in the art will appreciate, the amount of tert-butanol may be varied. The tert-butanol solution better dissolves EO9 as compared to water. By utilizing a tert-butanol formulation vehicle, solubility of EO9 is at least 9.5 mg/ml whereas the solubility of EO9 is approximately 0.2 mg/ml in water. Consequently, a smaller volume of the tert-butanol is required to dissolve a given amount of EO9. Additionally, a greater amount of EO9 may be dissolved in a given solution. That is, the compositions of the present invention will have a higher concentration of EO9 as compared to a solution where EO9 is dissolved in water.

According to another embodiment of the present invention, the composition comprises, EO9, a formulation vehicle, and a bulking agent. In one exemplary embodiment, lactose can be utilized as the bulking agent. As those skilled in the art will appreciate, it is contemplated that other bulking agents known or developed in the art may be utilized. According to another exemplary embodiment, the composition of the present invention can be buffered. In one embodiment, the composition is buffered to a pH ranging from approximately 9 to approximately 9.5. The composition can be buffered with any known or developed buffering agents. The compositions of the present invention can either be compounded for intravesical delivery or lyophilized. As those skilled in the art will appreciate, the compositions of the present invention can be lyophilized by those methods known or developed in the art. The lyophilized compositions can be reconstituted by a reconstitution vehicle. According to one exemplary embodiment, the reconstitution vehicle comprises 2% sodium bicarbonate, 0.02% disodium edetate and propylene glycol: water (60:40 V/V). This reconstitution vehicle dissolves the lyophilized composition of the present invention and produces a stable solution for administration for up to 24 hours. Additionally, the reconstitution vehicle of the present invention provides an ampoule having an extractable volume of 5 mL of reconstituted EO9 comprising propylene glycol/water/sodium bicarbonate/sodium edetate 60/40/2/0.02% v/v/w/w.

In another aspect of the present invention, the compositions of the present invention also comprises coating agents. The coating agents of the present invention provide better adhesion of the composition to the bladder wall. Consequently, the composition and, in particular, the EO9 contacts and may be able to penetrate the avascular tissue that comprises for a time sufficient to treat the bladder cancer. In one embodiment of the present invention, the coating agent is propylene glycol. In other

exemplary embodiments of the present invention, the coating agent can be selected from the group consisting of hydroxypropylcellulose, carboxymethylcellulose, chitosan hydrochloride, lectin, or polycarbophil.

In yet another embodiment of the present invention, the compositions of the present invention can be delivered to the bladder wall by a liposome. According to one embodiment of the present invention, the liposomes used are unilamellar or multilamellar and contain at least one cationic phospholipid such as stearylamine, 1,2-diacyl-3-trimethylammonium-propane (TAP) or 1,2-triacyl-3-dimethylammonium-propane (DAP). In another embodiment of the present invention, the surface liposomes may be coated with polyethylene glycol to prolong the circulating half-life of the liposomes. In yet another embodiment of the present invention, neutrally charged liposomes such as, but not limited to, phosphatidylcholine and cholesterol can also be used for liposomal entrapment of the compositions of the present invention. In another embodiment, the compositions of the present invention can be delivered to the bladder wall by a microsphere such as those known or developed in the art.

In yet another embodiment, the compositions of the present invention can be delivered to a patient intravenously. The lyophilized composition of the present invention can be reconstituted using the formulation vehicles of the present invention. The reconstituted composition can then be diluted to a desired concentration and delivered to a patient intravenously.

The following experiments were conducted to determine the activity of NQ01 in a series of human bladder tumors and normal bladder tissue by both enzymatic and immunohistochemical techniques. Furthermore, the following experiments evaluate strategies for reducing possible system toxicity arising from intravesical therapy based upon the fact that the aerobic activity of EO9 against cell lines is enhanced under mild acidic conditions (Phillips et al., 1992). Administration of EO9 in an acidic vehicle would result in greater activity within the bladder and any drug absorbed into the blood stream would become relatively inactive due to the rise in extracellular pH. The following experiments also determine the role of NQ01 in the activation of EO9 under acidic conditions.

Collection of tumor and normal bladder specimens. Ethical approval for tissue collection was obtained from the Local Research Ethical Committee (Bradford NHS Trust) and samples taken from patients following informed consent. A total of 17 paired cold pinch biopsies were taken from bladder tumors and macroscopically normal looking bladder mucosa at cystoscopy, immediately prior to formal

transurethral resection of the tumor. Three specimens were taken from patients undergoing cystectomy and tumor and normal samples dissected by pathologists within one hour of surgical removal. Specimens were flash frozen in liquid nitrogen and transported for NQO1 enzyme analysis. Further biopsies were taken of the normal bladder mucosa immediately adjacent to the previous biopsy site and sent at the end of the procedure, along with the resected tumor, in formalin for routine histological analysis. In this way bladder tumor and normal bladder urothelium enzymology could be directly correlated with the appropriate tissue histology in each patient. Immunohistochemistry was performed from the subsequently archived wax blocks prepared for histology.

Biochemical determination of NQO1 activity. Cell cultures in exponential growth were trypsinised, washed twice with Hanks balanced salt solution (HBSS) and sonicated on ice (3 x 30sec bursts at 40% duty cycle and output setting 4 on a Semat 250 cell sonicator). NQO1 activity and protein concentration was determined as described below. Tissues were homogenised (10% w/v homogenate) in sucrose (0.25M) using a 1ml tissue homogeniser (Fisher Scientific). Cytosolic fractions were prepared by centrifugation of the homogenate at 18,000 g for 4 min followed by further centrifugation of the supernatant at 110,000 g for 1 h at 4°C in a Beckman Optima TL ultracentrifuge. Activity of NQO1 in the supernatant was determined spectrophotometrically (Beckman DU650 spectrophotometer) by measuring the dicumarol sensitive reduction of dichlorophenolindophenol (DCPIP, Sigma Aldrich, UK) at 600 nm (Traver et al, 1992). This assay has been extensively validated for use in measuring NQO1 activity in both tissue and cell homogenates and has been shown to be preferable to other assays for NQO1 activity (Hodnick and Sartorelli, 1997). Each reaction contained NADH (200 μ M), DCPIP (40 μ M, Sigma Aldrich, UK), Dicumarol (20 μ M, when required, Sigma Aldrich, UK), cytosolic fraction of tissues (50 μ l per assay) in a final volume of 1 ml Tris HCl buffer (50 mM, pH 7.4) containing bovine serum albumin (0.7 mg ml⁻¹, Sigma Aldrich, UK). Rates of DCPIP reduction were calculated from the initial linear part of the reaction curve (30s) and results were expressed in terms of nmol DCPIP reduced /min/mg protein using a molar extinction coefficient of 21mNT⁻¹ cm⁻¹ for DCPIP. Protein concentration was determined using the Bradford assay (Bradford, 1976).

Immunohistochemistry. Polyclonal antibodies (raised in rabbits) to purified rat NQO1 were a gift from Professor Richard Knox (Enact Pharma Plc). Validation of the antibody for use in immunohistochemistry studies was performed by Western blot

analysis using both purified human recombinant NQO1 and cell extracts derived from a panel of cell lines of human origin. These cell lines included H460 (human NSCLC), RT112 (human bladder carcinoma), HT-29 (human colon carcinoma), BE (human colon carcinoma), MT1 (human breast) and DLD-1 (human colon carcinoma). The BE cell line has been genotyped for the C609T polymorphic variant of NQO1 and is a homozygous mutant (and therefore devoid of NQO1 enzyme activity) with respect to this polymorphism (Traver et al, 1992). Cells were washed in ice cold phosphate buffered saline and lysed by sonication (30 seconds on ice) in Tris HCl (50 mM, pH 7.5) containing 2mM EGTA, 2mM PMSF and 25 $\mu\text{g ml}^{-1}$ leupeptin. Protein concentration was estimated using the Bradford assay (Bradford, 1976) and a total of 12.5 μg of protein (in Lamelli sample loading buffer) applied to a 12% SDS-PAGE gel. Following electrophoretic transfer to nitrocellulose paper, membranes were blocked in TBS/Tween 20 (0.1%) containing 5% non-fat dry milk for 1 h at room temperature. Membranes were washed in TBS/Tween 20 (0.1 %) prior to the addition of rabbit anti-rat NQO1 antibody (1:100 dilution) and incubated at room temperature for 1 h. Membranes were extensively washed in TBS/Tween 20 (0.1%) followed by the addition of anti-rabbit IgG horseradish peroxidase conjugated secondary antibody (1:5000 dilution in TBS/Tween 20). Proteins were visualised by ECL based chemiluminescence as described by the manufacturer (Amersham Pharmacia Biotech, Bucks, UK).

For immunohistochemical studies, all tissues (both tumor and normal bladder mucosa) were fixed in 10% formalin, processed routinely and embedded in paraffin wax. Two sections of each tissue block were placed on one slide, one section served as the test and the other as a negative control (no primary antibody). A total of 5 sections from each sample were stained for NQO1 (plus negative controls) and tumor and normal samples from a total of 17 patients were analysed. Sections (5 μm) were dewaxed, rehydrated and incubated with primary antibody (1:400 dilution) for 4 hours. Sections were then washed and incubated with biotinylated mouse anti rabbit IgG for 30 min prior to immunoperoxidase staining using VECTASTAIN ABC reagents and DAB (Vector Laboratories Ltd, Peterborough,UK). Sections were counterstained with haematoxylin according to standard procedures.

Cell culture and chemosensitivity studies. E09 was a gift from NDDO Oncology, Amsterdam and MMC was obtained from the Department of Pharmacy, St Lukes Hospital, Bradford. H460 (human NSCLC) cell line was obtained from the American Type Culture Collection (ATCC). HT-29 (human colon carcinoma), RT112/83 (human bladder carcinoma epithelial), EJ138 (human bladder carcinoma)

and T24/83 (human bladder transitional cell carcinoma) cell lines were obtained from the European Collection of Animal Cell Cultures (ECACC). A2780 (human ovarian carcinoma) and BE (human colon carcinoma) cells were gifts from Dr T Ward (Paterson Institute, Manchester, UK). All cell lines were maintained as monolayer cultures in RPMI 1640 culture medium supplemented with fetal calf serum (10%), sodium pyruvate (2 mM), L-glutamine (2mM), penicillin/streptomycin (50 IU/ml / 50 µg/ml) and buffered with HEPES (25 mM). All cell culture materials were purchased from Gibco BRL (Paisley, UK). Cells were exposed to MMC or E09 at a range of doses for one hour and chemosensitivity was assessed following a five day recovery period using the MTT assay, details of which have been described elsewhere (Phillips et al, 1992). The pH of the medium used during drug exposure was adjusted using small aliquots of concentrated HCl (40 µM conc HCl [10.5M] to 20 ml medium gives a pH of 6.0). Calibration curves were conducted over a broad range of pH values in culture medium (pH 3.5 to 11) and the stability of the pH conditions monitored over a one hour incubation period at 37°C. At all pH values, no significant changes in the pH of the medium was observed over the one hour drug exposure period (data not presented).

HT-29 multicell spheroids were prepared by seeding 5×10^5 cells into T25 flasks which had been based coated with agar (1% w/v) and incubated for 24h at 37°C. Immature spheroids were then transferred to a spinner flask (Techne) containing 250ml of RPMI 1640 growth medium and spheroids were kept in suspension by stirring at 50 rpm. When spheroids reached a diameter of approximately 500 µm, they were harvested for chemosensitivity studies. Multicell spheroids were exposed to a range of E09 concentrations at pH 6.0 and 7.4 for one hour at 37°C. Following drug incubation, spheroids were washed twice in HBSS prior to disaggregation into single cells using trypsin EDTA. Disaggregated spheroids were then washed in HBSS and then plated into 96 well plates (1×10^3 cells per well), and incubated at 37°C for four days. Chemosensitivity was assessed using the NM assay as described elsewhere (Phillips et al, 1992).

The role of NQO1 in the activation of E09 at pH values of 7.4 and 6.0 was evaluated using the NQO1 inhibitor Flavone Acetic Acid (FAA), details of which are described elsewhere (Phillips, 1999). FAA is a competitive inhibitor of NQO1 with respect to NADH and at a final concentration of 2 mM, inhibition of NQO1 is > 95% whereas the activity of cytochrome P450 reductase and cytochrome b5 reductase is not substantially altered (<5% inhibition). Briefly, H460 cells (NQO1 rich) were plated

into 96 well plates at a density of 2×10^3 cells per well. Following an overnight incubation at 37°C , medium was replaced with fresh medium (pH 7.4) containing a non-toxic concentration of FAA (2mM) and incubated for one hour at 37°C . Medium was then replaced with fresh medium containing E09 (range of drug concentrations) and FAA (2mM) at either pH 7.4 or 6.0. Following a further one hour incubation at 37°C , cells were washed twice with HBSS and incubated at 37°C in growth medium for five days. Chemosensitivity was determined by the NM assay as described above and results were expressed in terms of IC_{50} values, selectivity ratios (IC_{50} at pH 7.4 / IC_{50} at pH 6.0) and protection ratios (IC_{50} FAA/E09 combinations / IC_{50} for E09 alone).

Substrate specificity. The influence of acidic pH on substrate specificity for purified human NQO1 was determined as described previously (Phillips 1996, Walton et al, 1991). NQO1 mediated reduction of the quinone to the hydroquinone species is difficult to detect by conventional assays thereby necessitating the use of a reporter signal generating step. In this assay, the hydroquinone acts as an intermediate electron acceptor which subsequently reduces cytochrome c which can readily be detected spectrophotometrically. Recombinant human NQO1 was derived from E.coli transformed with the pKK233-2 expression plasmid containing the full length cDNA sequence for human NQO1 isolated from the (Beall et al, 1994). Following IPTG induction, NQO1 was purified by cybacron blue affinity chromatography, details of which are described elsewhere (Phillips, 1996). The purified protein had a molecular weight of approximately 31 kDa and a specific activity of 139 $\mu\text{mol DCPIP reduced} / \text{min} / \text{mg protein}$ (Phillips, 1996). Reduction of E09 by recombinant human NQO1 was determined at pH 6.0 and 7.4 by measuring the rate of reduction of cytochrome c was measured at 550 nm on a Beckman DU 650 spectrophotometer according to previously published methods (Phillips, 1996). Results were expressed in terms of $\mu\text{mol cytochrome c reduced} / \text{min} / \text{mg protein}$ using a molar extinction coefficient of $21.1 \text{ mM}^{-1} \text{ cm}^{-1}$ for cytochrome c.

Measurement of intracellular pH. Intracellular pH was determined using the fluorescent pH indicator BCECF (2,7-bis-(2-carboxy-ethyl)-5-(and-6) carboxyfluorescein (Molecular Probes, Eugene, USA) according to manufacturers instructions. Confluent flasks of cells were washed with HBSS to remove any traces of serum containing RPMI medium and then incubated with the esterified form of BCECF (BCECF-AM) at a concentration of 2 μM in HBSS for one hour at 37°C . The non-denaturing detergent Pluronic was added to the probe to aid dispersion. Cells were

then washed to remove all traces of BCECF-AM and then trypsinized before being suspended in serum-free / phenol red-free RPM1 medium (Gibco BRL, Paisley, UK) at a concentration of 10^6 cells per ml at pH 6 for one hour. Fluorescence measurement was determined in a Perkin-Elmer fluorescence spectrophotometer in UV grade disposable 4ml cuvettes (Fischer Scientific) with excitation wavelengths 500nm and 450nm (excitation bandpass slit of 10nm) and emission wavelength fixed at 530nm (emission bandpass slit of 2.5nm). These were determined to be optimal settings for the machine and system under study. An in-situ calibration was performed for every pH_i determination with a range of six pH's from 4 to 9 using the ionophore nigericin at a concentration of 22.8 p,M to equilibrate pHe with pH_i. Calculation of the ratio of fluorescence at 500nm / 450nm was calculated after subtraction of background fluorescence from blanks at each pH (serum free, phenol red free RPMI without cells).

Activity of NQO1 in tumor and normal bladder specimens. The biochemical activity of NQO1 in paired samples of tumor (grade/stage ranging from G2 pTa to G2/G3 T4) and normal bladder mucosa (with three cystectomy specimens) taken from a series of 20 patients is presented in table 1. Within the tumor specimens, a broad range of NQO1 activity existed ranging from 571.4 nmoUmin/mg to undetectable (< 0.1 nmol/min/mg). In histologically nonnal bladder mucosa specimens, NQO1 activity ranged from 190.9 to < 0.1 nmoUmin/mg. In the majority of patients NQO1 activity in the tumor was greater than in the normal bladder mucosa. Tumor grade and stage did not correlate with NQO1 activity (table 1).

Validation of NQO1 antibody and inununohistochemical localization of NQO1. Western blot analysis demonstrates that polyclonal anti rat NQO1 antibody cross reacts with human NQO1 (figure 1) with a single band at approximately 31 kDa observed for both cell extracts and purified human NQO1. Titration of purified NQO 1 results in a decrease in band intensity (figure 1B) and in cell extracts, band intensity was qualitatively consistent with NQO1 enzyme activity (figure 1A). In addition, the antibody does not detect NQO1 in the BE cell line which is devoid of NQO1 activity as a result of the C609T polymorphism (figure 1C). No non-specific bands were observed on Western blots. Immunoperoxidase staining of NQO1 protein in tumor tissue, bladder wall, ureter and urethra are presented in figure 2. Superficial and invasive tumors (pTa- panel A, G3 pT2 - panel B and G3pT4 - panel C) with high to intermediate levels of NQO1 as determined by biochemical assays (patient numbers 1, 4 and 5 in table 1) clearly stained positive for NQO1. Staining was confined to the cytoplasm of tumor cells with little or no staining of stromal cells (panels B and C).

In other tumors with intermediate or low levels of NQO1 activity, staining was heterogeneous with pockets of cells containing high levels of NQO1 protein (data not shown). Normal bladder wall sections were obtained from a patient who underwent cystectomy (G3pT4 bladder tumor), ureter and urethra were obtained from another patient who underwent cystectomy (G3 pT3a bladder tumor). In the bladder wall, no NQO1 staining was observed in the urothelium (panel D) although slight staining was present in smooth muscle layers. The urethra (panel E) was negative although cells on the luminal surface of the ureter were positively stained (panel F). The basal layers of the ureter lining were however negatively stained (panel F). No evidence of invasive malignancy or in situ carcinoma were observed in the ureter and urethra or in the section of bladder wall presented (panel D). In 16 other normal bladder biopsy and cystectomy specimens, no positive staining of the urothelium was observed (data not shown).

Influence of pH on substrate specificity and chemosensitivity. The ability of E09 to serve as a substrate for NQO1 was not influenced by pH with specific activities of 21.10 ± 2.3 and 21.30 ± 1.5 pmol cytochrome c reduced/min/mg protein at pH 7.4 and 6.0 respectively. The response of a panel of cell lines with a range of NQO1 activity (<1.0 to $1,898 \pm 276$ nmol/min/mg) to E09 and MMC at pHe values of 7.4 and 6.0 is presented in table 2 and figure 2. At pHe = 7.4, a good correlation existed between NQO1 activity and chemosensitivity to E09 (figure 3). In the case of MMC (table 2, figure 3), a relationship between NQO1 and chemosensitivity was apparent (at pHe 7.4) although this relationship was not as prominent as shown by E09 with a narrow range of IC_{50} values (range 0.9 to 7.0 μ M) observed in cell lines which cover a broad range of NQO1 activity (ranging from <1.0 to 1,898 nmol/min/mg). Both MMC and E09 are preferentially more toxic to cells at pHe values of 6.0 although much greater potentiation of E09 activity is seen with SR values (SR = selectivity ratio defined as IC_{50} pHe 7.4 / IC_{50} pHe 6.0) ranging from 3.92 to 17.21 for E09 compared with 1.02 to 4.50 for MMC (table 2). The activity of E09 was enhanced in both NQO1 rich and deficient cell lines when pHe was reduced to 6.0 and the relationship between NQO1 and chemosensitivity remained good when cells were exposed to E09 under acidic conditions (figure 3). No cell kill was observed in control cultures when the pHe was decreased to 6.0 (in the absence of drug) as determined by the MTT assay. The response of H460 cells to E09 at pHe values of 7.4 and 6.0 in the presence and absence of FAA (2mM) is presented in table 3. At both pHe values, the response of H460 cells to E09 was reduced in the presence of FAA. Protection ratios defined as

the IC50 for E09 plus FAA divided by the IC50 value for E09 alone were similar for cells under acidic and physiological pHe values (14.63 and 13.95 respectively, table 3). Selectivity ratios defined as the IC50 at pHe 7.4 divided by the IC50 at pHe 6.0 in the presence and absence of FAA were also similar with SR values of 6.31 and 6.02 for E09 alone and E09 plus FAA respectively (table 3). The response of HT-29 multicell spheroids to E09 is presented in figure 4. Spheroids exposed to E09 at pHe 6.0 were significantly more responsive than at pHe 7.4 with IC50 values of 9.89 ± 0.89 and 24.24 ± 3.29 AM respectively. Spheroids were significantly less responsive to E09 than the same cells exposed to E09 as monolayers at both pHe values with ratios of IC50 values for spheroids to monolayers of 202 and 341 at pHe values of 7.4 and 6.0 respectively.

Influence of acidic pHe conditions on pHi. PM values following a one hour incubation at pHe 6.0 were 6.44 ± 0.04 , 6.51 ± 0.02 and 6.42 ± 0.05 in A549, RT112/83 and A2780 cells respectively. Addition of the ionophore nigericin (after a one hour incubation at pHe 6.0) resulted in the equilibration of pHe and p11i.

In terms of bioreductive drug development, two of the critical factors which will ultimately determine selectivity are the enzymology of tumors and the presence of hypoxia (Workman, 1994). As outlined in the introduction, the presence or absence of NQO1 is central to the design of appropriate E09 based therapeutic strategies aimed at targeting either the aerobic (NQO1 rich cells) or hypoxic fraction (NQO1 deficient tumors) of tumors. Workman (1994) has outlined a proposed mechanism for the different properties of E09 under aerobic and hypoxic conditions based on the hypothesis that it is the semiquinone (product of one electron reduction) rather than the hydroquinone which is responsible for toxicity. In NQO1 deficient cells, the semiquinone produced as a result of one electron reductases would be relatively non-toxic as it would rapidly redox cycle back to the parent compound. Free radical species generated as a result of redox cycling would be detoxified by superoxide dismutase or catalase but under hypoxic conditions, the semiquinone would be relatively stable. If this were the major toxic species, then the activity of E09 against cells with low NQO1 would be potentiated. In NQO1 rich cells however, the major product formed would be the hydroquinone. Aerobic toxicity could be generated as a result of the back oxidation of the hydroquinone to the semiquinone species or the parent quinone (Butler et al, 1996) resulting in free radical generation. Under hypoxic conditions however the hydroquinone will be more stable and if this is relatively non-toxic, then the activity of E09 against NQO1 cells under hypoxia would not be

potentiated. Whilst the mechanism of action of E09 under aerobic and hypoxic conditions is complex, the biological data suggest that E09 should target the aerobic fraction of NQO1 rich tumors or the hypoxic fraction of NQO1 deficient tumors (Workman, 1994).

Analysis of NQO1 activity in tumor and normal bladder tissues has clearly identified patients whose tumors are either NQO1 rich or NQO1 deficient (table 1). Within the subset of NQO1 rich tumors, enzyme activity is elevated relative to the normal bladder urothelium. Immunohistochemical studies confirm these biochemical measurements with staining confined to tumor cells as opposed to normal stromal cells (fig 2, panels A, B and C). Within normal bladder tissues, NQO1 staining was absent from the urothelial lining of the bladder (fig 2, panel D) and the urethra (fig 2, panel E). Faint staining of the superficial layers of the ureter (fig 2, panel F) was observed although the underlying basal layers of the ureter were negatively stained. Similarly, faint staining of the smooth muscle layers of the bladder, ureter and urethra were also observed (data not shown). These studies suggest that a proportion of patients with bladder tumors (at various grades and stages of the disease) exhibit a significant differential in terms of NQO1 activity which could potentially be exploited by E09 based therapies directed against the aerobic fraction of tumor cells. With regards to the ability of E09 to selectively kill hypoxic NQO1 deficient cells, a subset of patients also exist whose tumors are devoid of NQO1 activity (table 1). It is not known whether or not bladder tumors contain regions of low oxygen tension and further studies are required using hypoxia markers such as pimonidazole (Kennedy et al, 1997) to address this issue and to establish the relationship between NQO1 activity and hypoxia in tumors.

Whilst biochemical and immunohistochemical studies demonstrate that a subset of patients exist which have the appropriate tumor enzymology to activate E09 (under aerobic conditions), intravesical chemotherapy can result in systemic toxicity due to the drug entering the blood supply. This study has also evaluated a potential strategy for minimizing any risk of systemic toxicity based upon the hypothesis that administration of E09 in an acidic vehicle would enhance the potency of E09 (Phillips et al, 1992) within the bladder and that any drug reaching the blood stream would become relatively inactive due to a rise in pHe. Selectivity for aerobic cells would still be determined by NQO1 activity and therefore it is essential to determine the role that NQO1 plays in the activation of E09 under acidic pHe conditions. In a panel of cell lines with a broad spectrum of NQO1 activity, reducing the pHe to 6.0 enhances the

potency of E09 under aerobic conditions in all cases (with SR values ranging from 3.92 to 17.21, table 2). In the case of MMC, potency is also enhanced at low pHe values although the magnitude of the pH dependent increase in toxicity is reduced (SR values ranging from 1.02 to 4.50, table 2) compared with E09. With respect to MMC, one explanation for increased activity under acidic conditions has been attributed to the fact that MMC becomes a substrate for NQO1 under acidic conditions (Pan et al, 1993, Siegel et al, 1993). This is not the case with E09 as rates of reduction of E09 by purified human NQO1 are not influenced by pH (21.10 ± 2.30 and 21.30 ± 1.50 nmol cytochrome c reduced / min / mg protein at pH 7.4 and 6.0 respectively). Recent studies have demonstrated that the activity of E09 is enhanced under acidic conditions (pHe = 6.5) but only when the intracellular pH is reduced (pHi = 6.5) by co-incubation with nigericin (Kuin et al, 1999). The results of this study are in agreement with this finding as pHi becomes acidic (pHi values range from 6.42 ± 0.05 to 6.51 ± 0.02 depending on the cell line) when cells are cultured under pHe 6.0 conditions.

In the panel of cell lines used in this study, a good correlation exists between NQO1 activity and chemosensitivity at both pHe values of 7.4 and 6.0 (figure 3). A strong relationship between NQO1 activity and response under aerobic conditions (at pHe 7.4) has been established previously by several groups (Robertson et al, 1994, Fitzsimmons et al, 1996, Smitkamp-Wilms et al, 1994) and there is clear evidence that NQO1 plays a central role in the mechanism of action of E09 under aerobic conditions (Workman, 1994). The good correlation between NQO1 activity and response at pHe 6.0, in conjunction with the fact that E09 is still a good substrate for NQO1 at pH 6.0, suggests that NQO1 plays a significant role in E09's mechanism of action at acidic pHe values under aerobic conditions. It is of interest to note however that the activity of E09 against BE cells (which are devoid of NQO1 activity as a result of the C609T polymorphism, Traver et al, 1992) is also enhanced under acidic pHe conditions (table 2). This suggests that there is a NQO1 independent mechanism for the increased activity of E09 under acidic conditions. This is confirmed by the use of the NQO1 inhibitor FAA where the 'protection ratios' (defined as the ratio of IC₅₀ values for E09 plus FAA divided by the IC₅₀ values for E09) are similar at both pHe 7.4 and 6.0 (13.95 and 14.63 respectively, table 3). If NQO1 played a central role in the activation of E09 at pHe 6.0, then the protection ratio at pHe 6.0 would be significantly greater than the protection ratio at pHe 7.4. The mechanism behind the NQO1 independent activation of E09 is unclear although it is a well known fact that the reactivity of

aziridine ring structures is enhanced by protonation resulting in ring opening to the aziridinium ion which is a potent alkylating species (Mossoba et al, 1985, Gutierrez, 1989). Alternatively, E09 is a substrate for other one electron reductases (Maliepaard et al, 1995, Saunders et al, 2000) and further studies designed to evaluate whether E09's metabolism by these enzymes is pH dependent needs to be determined. The potency of E09 can be enhanced further by reducing pHe below 6.0 (Phillips et al, 1992) but these conditions are unlikely to provide significant clinical benefits as E09 becomes progressively more unstable when pH is reduced to 5.5 ($t_{1/2} = 37$ min). From a pharmacological standpoint, administration of E09 in a vehicle at pH 6.0 would appear desirable. Not only would this result in significant enhancement of E09 activity but also the stability of E09 would be sufficient ($t_{1/2} = 2.5$ h) to maintain drug exposure parameters at a therapeutic level.

With regards to the activity of E09 against three dimensional culture models in vitro, this study has demonstrated that reducing the pHe to 6.0 enhances the potency of E09 against multicell spheroids although the magnitude of this effect is reduced compared with monolayer cultures (figure 4). It is not known whether or not reduction in pHe results in greater cell kill throughout the spheroid or if it is confined to the surface of the spheroid exposed to medium. In comparison with MMC, previous studies using histocultures exposed to MMC demonstrated that no difference in toxicity exists between physiological and acidic pHe conditions (Yen et al, 1996). The pH dependent increase in E09 toxicity against spheroids suggests that manipulation of pHe may not only be of use in treating a multilayered solid bladder tumor but may offer an advantage over MMC. It should however be stated that multicell spheroids are significantly less responsive to E09 than mono] ayers, presumably because of the poor penetration properties of E09 through avascular tissue (Phillips et al, 1998). E09 can nevertheless kill >90% of cells in spheroids (figure 4) suggesting that a higher doses at least, the penetration of E09 is sufficient to eradicate cells which reside some distance away from the surface of the spheroid.

In conclusion, the results of this study have demonstrated that within a population of patients with bladder tumors at various stages and grades of the disease, there exists a great heterogeneity regarding the expression of NQO1. The majority of patients have tumors possessing elevated levels of NQO 1 while a small subset of patients appear to be devoid of NQO1 activity. The heterogeneous nature of NQO1 activity described here is consistent with several other studies in various tumor types (Malkinson et al, 1992, Smitkamp-Wilms et al, 1995, Siegel et al, 1998). These

findings reinforce the view that 'enzyme profiling' of individual patients could be valuable prior to therapeutic intervention with bioreductive drugs (Workman, 1994). This is to our knowledge the first study to characterize NQO1 activity and cellular localization in bladder tumors and provide strong evidence to support the evaluation of E09 against superficial and locally invasive bladder tumors. This study has clearly demonstrated that under aerobic conditions, E09 is much more potent under acid conditions (pH6.0) than at physiological pH (pH7.4). The mechanism for this increased E09 potency appears to be NQO1 independent and whilst this will not improve (or reduce) selectivity, it may prove beneficial in terms of reducing the therapeutically effective dose of E09. Dose reduction in conjunction with the fact that a reduction in the potency of E09 due to the increased pHe in the blood stream suggests that systemic toxicity arising from the intravesical administration of E09 would be low. In addition, this study shows that under physiological conditions the activity of E09 is much lower in tissues with "normal" expression of NQO1 compared to "high" NQO1 expressing tissues (i.e. the tumors). The results of this study provide strong evidence in support of the proposal that intravesical administration of E09 may have activity against bladder tumors.

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Table 1.
Tumor histology reports and NQO1 activity in paired samples of bladder tumor and normal bladder mucosa.

Patient No.	Tumor histology	NQO1 Activity Tumor (nmol/min/mg)	NQO1 Activity Normal (nmol/min/mg)	Ratio of NQO1 levels in tumor to normal tissue.
1 f,s,i,p	G2 pTa	571.4	< 0.1	571.
2 m,s,r	G3 pT2	273.3	<0.1	273.
3 f,s,i	G1pTa	107.80	<0.1	107.
4 m,e,i	G3 pT2/3	73.36	<0.1	73.3
5 m,s,i	G3pT4 (0'	81.30	4.10	19.8
6 ^h	G2PT1	309.50	25.20	12.1
7 m,n,r,o	G3 pT2	10.00	<0.1	10.0
8 f n,i	G3pT2	9.80	<0.1	9.80
9 m,n,i	G2 pT2	4.40	<0.1	4.40
10 m,s,c	G3 pT2	34.01	8.50	4.00
11 ^{ms}	G 1 pTa	69.76	22.20	3.14
12 ^{,,n}	G1pTa	42.16	15.30	2.73
13 m,n,i	G3 pT2	179.6	72.12	2.49
14 m,e,i	G2/G3 T4 (C)	89.70	63.30	1.41
15 m,n,r	G3 pT2	0.40	<0.1	0.40
16 m,e,c,o	G3 PT3 (C)	21.60	61.70	0.35
17 f n,i	G2 PTI	58.40	190.90	0.30
18 m,e,o	G2 PTI	<0.1	<0.1	0
19 f n,i	G2 PT I.	<0.1	<0.1	0
20 m,e,c,r	G2 pT0	<0.1	<0.1	0

^mMale, ^fFemale, ^s Smoker, [']Non-smoker, ^e Ex-smoker, [°] Intravesical chemotherapy prior to specimen collection, ^r Radiotherapy prior to specimen collection, ['] First presentation, ^P Previous malignancy other than bladder, ^h No medical history available, ^o Possible occupational carcinogen exposure (i.e., dye industry worker).

(C) denotes cystectomy specimens. In all cases, protein levels following preparation of the cytosolic fraction were greater than 0.1 mg/ml.

Table 2.

The relationship between NQO1 activity and chemosensitivity to E09 and MMC under physiological and acidic pHe conditions.

Cell line	Drug		NQO1 (nmol/min/mg)	IC50 pHe 7.4 (nM)	IC50 pHe 6.0 (nM)	SR*
H460	E09		1652 ± 142	60 ± 10	9.5 ± 2	6.31
HT-29	E09	-	688 ± 52	120 ± 53	29 ± 10	4.13
T24/83	E09		285 ± 28	290 ± 65	60 ± 18	4.83
A2780	E09		159 ± 33	200 ± 50	51 ± 14	3.92
EJ138	E09		83 ± 14	310 ± 95	39 ± 7	7.94
RT112	E09		30 ± 3	1050 ± 75	61 ± 13	17.21
BE	E09		<0.1	5300 ± 169	1300 ± 75-	4.07
H460	MMC		1652 ± 142	900 ± 200	220 ± 130	4.50
HT-29	MMC		688 ± 52	1050 ± 210	500 ± 240	2.10
T24/83	MMC		285 ± 28	2150 ± 93	2100 ± 800	1.02
A2780	MMC		159 ± 33	2400 ± 340	1400 ± 130	1.71
EJ138	MMC		83 ± 14	1600 ± 200	1400 ± 250	1.14
RT112	MMC		30 ± 3	3350 ± 250	2000 ± 500	1.67
BE	MMC		<0.1	7000 ± 192	4400 ± 215	1.59

All results presented are the mean of 3 independent experiments (SD values omitted in the interests of presentation).

* SR (selectivity ratio) = IC50 at pH 7.4 / IC50 at pH 6.0

Table 3.

Response of H460 cells to E09 in the presence or absence of FAA (2mm) at pHe values of 7.4 and 6.0.

Drug	pHe	IC50 (nM)	SR*	PR**
E09	7.4	60.0 ± 8.1	'	-
E09	6.0	9.5 ± 2.6	6.31	-
E09/FAA	7.4	837 ± 45	-	13.95
E09/FAA	6.0-	139 ± 27	6.02	14.63

* SR = Selectivity Ratio defined as the ratio of IC50 values at pHe = 7.4 divided by the IC50 at pHe = 6.0.

** Plf = Protection ratio defined as the ratio of IC50 values for E09 plus FAA divided by the IC50 values for E09 alone.

All values represent the mean ± standard deviation for three independent experiments.

Table 4
Neoquin 8mg/vial lyophilised product

Storage	test item	time (months)				
		0	1	2	3	6
5°C	content*	102,7 ± 1,2	na	na	103,8 ± 0,8	100,6 ± 0,6
	purity**	99,9 ± 0,008	na	na	99,5 ± 0,03	99,6 ± 0,03
	residual moisture***	6.0%	na	na	7.0%	6.3%
	pH after reconstitution****	9.5	na	na	na	9.4
25°C/60 %RH	content	102,7 ± 1,2	103,4 ± 0,7	102,1 ± 0,2	102,6 ± 1,3	97,4 ± 1,0
	purity	99,9 ± 0,008	99,9 ± 0,05	99,9 ± 0,01	99,2 ± 0,07	98,7 ± 0,2
	residual moisture	6.0%	na	na	5.9%	5.9%
	pH after reconstitution****	9.5	na	na	na	9.4
40°C/75 %RH	content	102,7 ± 1,2	102,3 ± 1,1	100,4 ± 1,3	101,3 ± 0,2	86,4 ± 2,0
	purity	99,9 ± 0,008	99,8 ± 0,01	99,7 ± 0,04	98,4 ± 0,07	97,5 ± 0,2
	residual moisture	6.0%	na	na	6.2%	6.3%
	pH after reconstitution****	9.5	na	na	na	9.5

*content as % of labelled content n=3

**purity as chromatographic purity n=3

What is claimed is:

1. An anti-cancer formulation comprising:
a buffered solution comprising 3-hydroxymethyl-5-aziridinyl-1-1-methyl-2-[1H-indole-4,7-dione]propenol and a formulation vehicle.
2. The anti-cancer formulation of claim 1 wherein the anti-cancer formulation is lyophilized.
3. The anti-cancer formulation of claim 1 wherein the buffered solution has a pH ranging from approximately 9 to approximately 9.5.
4. The anti-cancer formulation of claim 1 wherein the formulation vehicle is selected from the group consisting of tert-butanol/water, ethanol/water, and 2-hydroxypropyl- β -cyclodextrin solution.
5. The anti-cancer formulation of claim 1 further comprising a bulking agent.
6. The anti-cancer formulation of claim 5 wherein the bulking agent is lactose.
7. A anti-cancer formulation for treating bladder cancer, the formulation comprising:
a buffered, lyophilized solution comprising 3-hydroxymethyl-5-aziridinyl-1-1-methyl-2-[1H-indole-4,7-dione]propenol and a formulation vehicle.
8. The anti-cancer formulation of claim 7 wherein the buffered, lyophilized solution has a pH ranging from approximately 9 to approximately 9.5.
9. The anti-cancer formulation of claim 7 wherein the formulation vehicle is selected from the group consisting of tert-butanol/water, ethanol/water, and 2-hydroxypropyl- β -cyclodextrin solution.
10. The anti-cancer formulation of claim 7 further comprising a bulking agent.
11. The anti-cancer formulation of claim 10 wherein the bulking agent is lactose.
12. An anti-cancer formulation comprising:
3-hydroxymethyl-5-aziridinyl-1-1-methyl-2-[1H-indole-4,7-dione]propenol dissolved in a buffered formulation vehicle, the buffered formulation vehicle having a pH ranging from approximately 9 to approximately 9.5, and wherein the formulation vehicle is selected from the group consisting of tert-butanol/water, ethanol/water, and 2-hydroxypropyl- β -cyclodextrin solution.

13. The anti-cancer formulation of claim 12 further comprising a bulking agent.

14. The anti-cancer formulation of claim 13 wherein the bulking agent is lactose.

15. An anti-cancer formulation comprising:
an indolequinone having a pH within a range of approximately 9 and approximately 9.5, a refractive index within a range of approximately 1.393 to approximately 1.406, and a relative density of within a range of approximately 0.94 to approximately 0.95.

16. The anti-cancer formulation of claim 15 wherein the indolequinone is 3-hydroxymethyl-5-aziridinyl-1-methyl-2-[1H-indole-4,7-dione]propenol.

17. The anti-cancer formulation of claim 16 further comprising a coating composition.

18. The anti-cancer formulation of claim 17 wherein the coating composition is selected from the group consisting of propylene glycol, hydroxypropylcellulose, carboxymethylcellulose, chitosan hydrochloride, lectin, and polycarbophil.

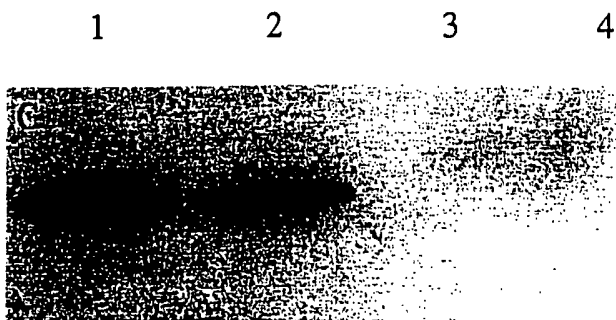
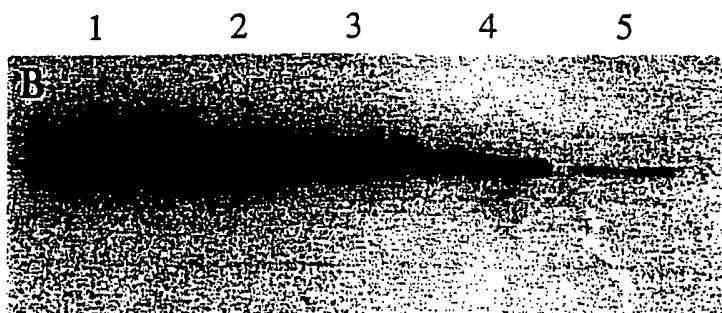
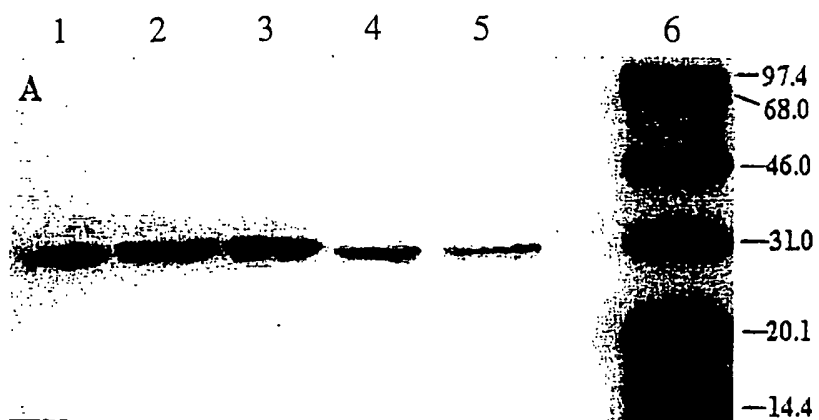


FIG 1

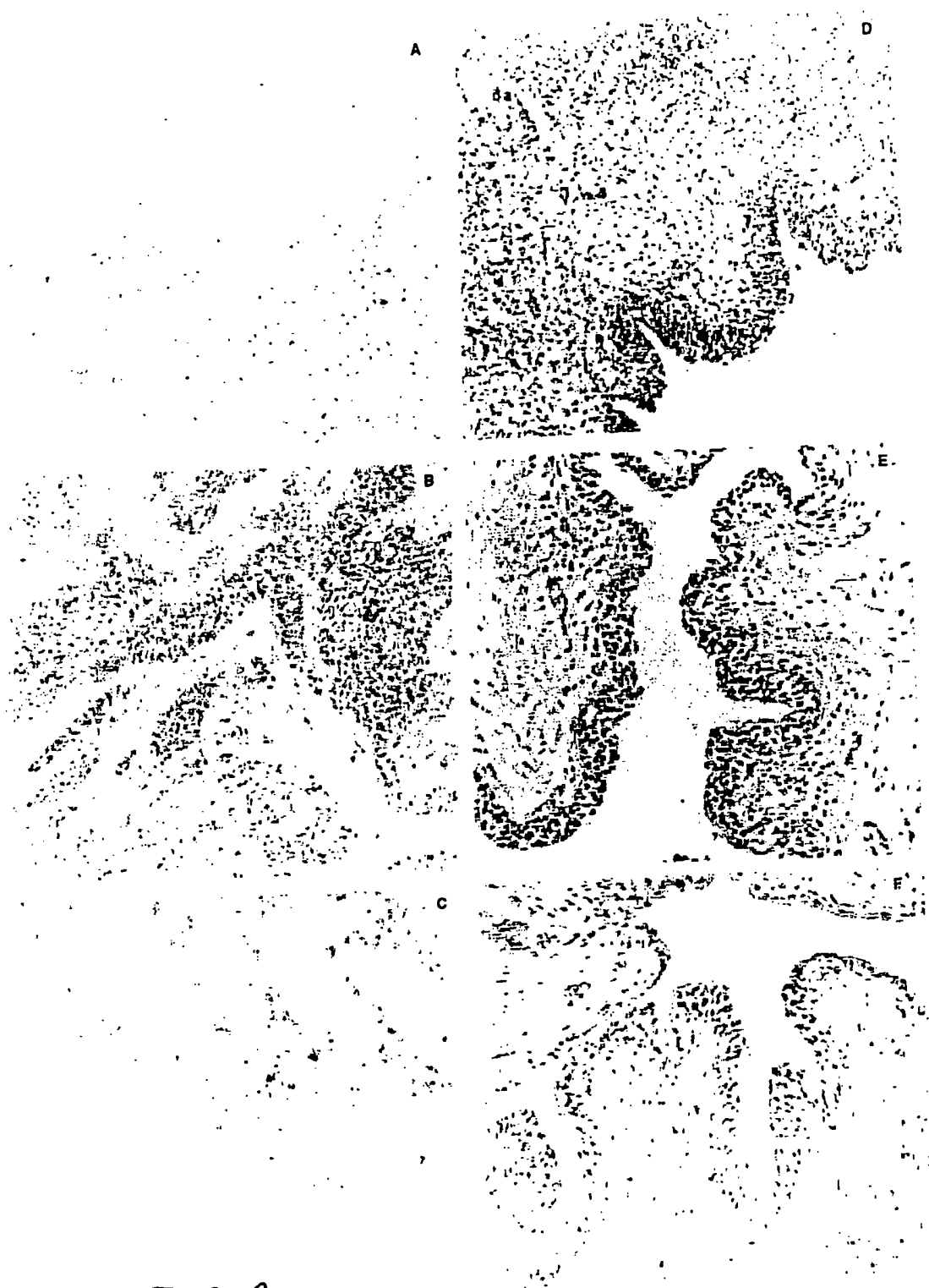


FIG. 2

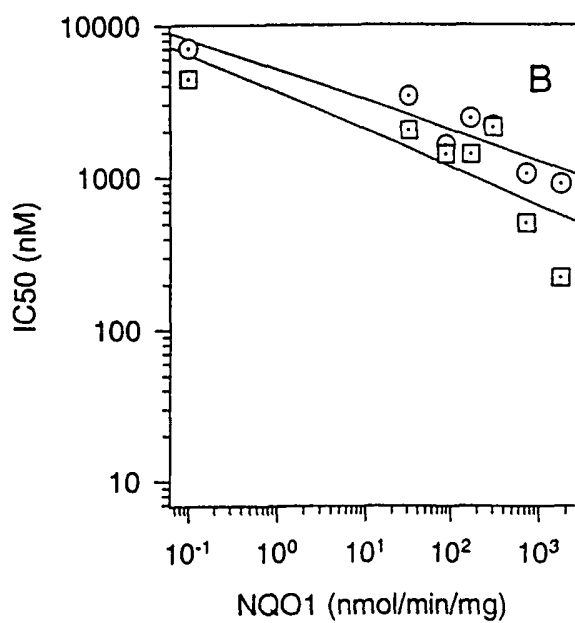
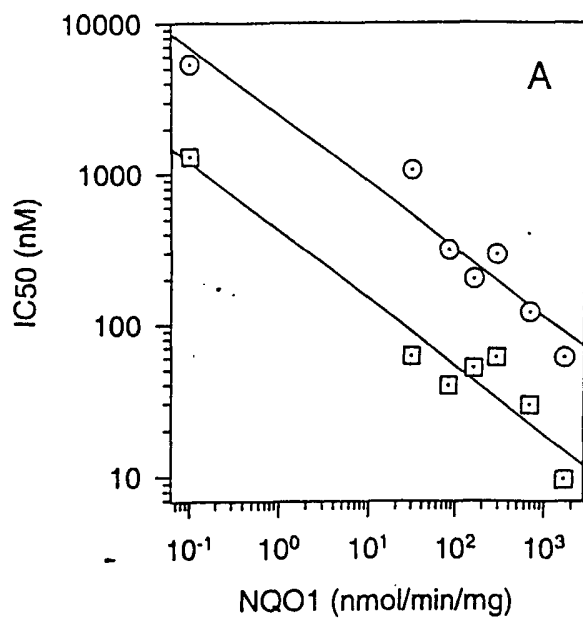


FIG. 3

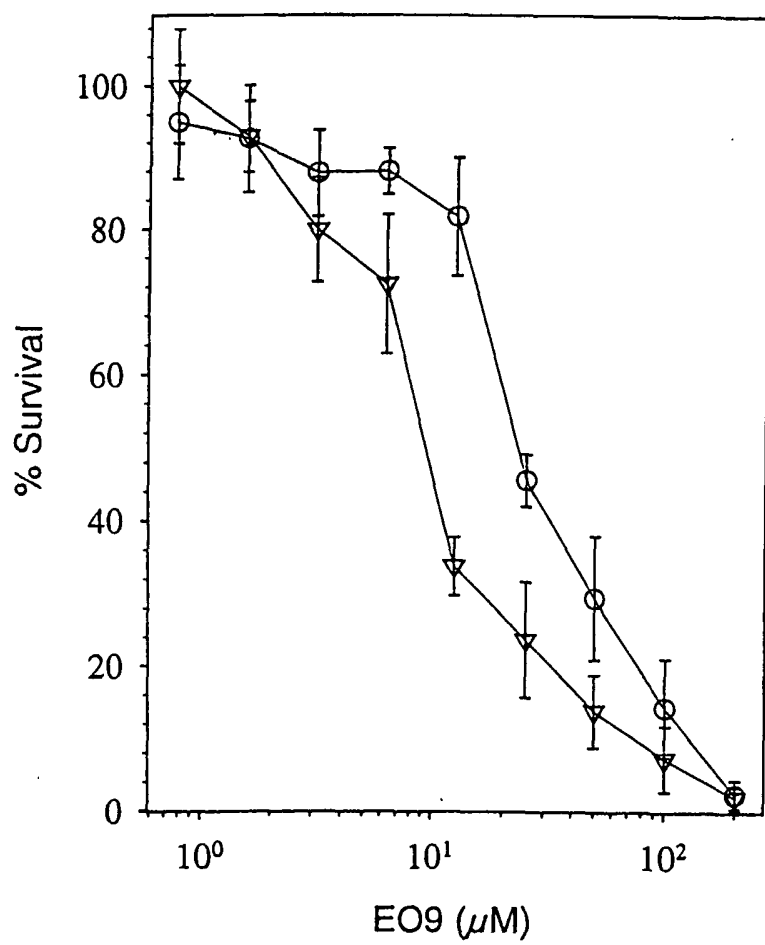


FIG. 4