Office de la Propriété Intellectuelle du Canada
Un organisme d'Industrie Canada

Canadian Intellectual Property Office
An agency of Industry Canada

CA 2473733 A1 2004/01/08

(21) 2 473 733

(12) DEMANDE DE BREVET CANADIEN
CANADIAN PATENT APPLICATION

(13) A1

(86) Date de dépôt PCT/PCT Filing Date: 2003/01/21
(87) Date publication PCT/PCT Publication Date: 2004/01/08
(85) Entrée phase nationale/National Entry: 2004/07/16
(86) N° demande PCT/PCT Application No.: US 2003/001983
(87) N° publication PCT/PCT Publication No.: 2004/003140
(30) Priorités/Priorities: 2002/01/18 (60/350,325) US;
2002/06/14 (60/389,108) US; 2002/12/19 (60/435,361) US

(51) Cl.Int.^{7}/Int.Cl.^{7} C12N 15/19, A61P 29/00, A61P 35/00, A61K 38/19, A61K 38/17, C07K 16/28, C12P 21/02, C07K 14/715, C12N 15/63, G01N 33/566

(71) Demandeur/Applicant:
ZYMOGENETICS, INC., US

(72) Inventeurs/Inventors:
SPRECHER, CINDY A., US;
GAO, ZEREN, US;
KUIJPER, JOSEPH L., US;
DASOVICH, MARIA M., US;
GRANT, FRANCIS J., US;
...

(74) Agent: MBM & CO.

(54) Titre : RECEPTEURS MULTIMERES DES CYTOKINES CONTENANT ZCYTOR17
(54) Title: CYTOKINE RECEPTOR ZCYTOR17 MULTIMERS

(57) Abrégé/Abstract:
Novel polypeptide combinations, polynucleotides encoding the polypeptides, and related compositions and methods are disclosed for zcytor17-containing multimeric or heterodimer cytokine receptors that may be used as novel cytokine antagonists, and within methods for detecting ligands that stimulate the proliferation and/or development of hematopoietic, lymphoid and myeloid cells in vitro and in vivo. The present invention also includes methods for producing the multimeric or heterodimeric cytokine receptor, uses therefor and antibodies thereto.

CA 2473733 A1 2004/01/08

(21) 2 473 733

(13) A1

(72) **Inventeurs(suite)/Inventors(continued)**: PRESNELL, SCOTT R., US; WHITMORE, THEODORE E., US; HAMMOND, ANGELA K., US; NOVAK, JULIA E., US; GROSS, JANE A., US; DILLON, STACEY R., US

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau

(43) International Publication Date
8 January 2004 (08.01.2004)

PCT

(10) International Publication Number
WO 2004/003140 A2

(51) International Patent Classification[7]: C12N

(21) International Application Number:
PCT/US2003/001983

(22) International Filing Date: 21 January 2003 (21.01.2003)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:

60/350,325	18 January 2002 (18.01.2002)	US
60/389,108	14 June 2002 (14.06.2002)	US
60/435,361	19 December 2002 (19.12.2002)	US

(71) Applicant: **ZYMOGENETICS, INC.** [US/US]; 1201 Eastlake Avenue East, Seattle, WA 98102 (US).

(72) Inventors: **SPRECHER, Cindy, A.**; 3549 Osprey Drive, Sierra Vista, AZ 85650 (US). **GAO, Zeren**; 9816 229th Lane NE, Redmond, WA 98053 (US). **KUIJPER, Joseph, L.**; 6524 NE 192nd Place, Kenmore, WA 98028 (US). **DASOVICH, Maria, M.**; 602 15th Ave East, Seattle, WA 98112 (US). **GRANT, Francis, J.**; 7714 37th Ave NE, Seattle, WA 98115 (US). **PRESNELL, Scott, R.**; 2902 N. Puget Sound Ave, Tacoma, WA 98407 (US). **WHITMORE, Theodore, E.**; 6916 152nd Avee NE, Redmond, WA 98052 (US). **HAMMOND, Angela, K.**; 26774 230th Place SE, Maple Valley, WA 98038 (US). **NOVAK, Julia, E.**; 10699 Battle Point Dr. NE, Bainbridge Island, WA 98110 (US). **GROSS, Jane, A.**; 4258 NE 74th Street, Seattle, WA 98115 (US). **DILLON, Stacey, R.**; 268 Northeast 42nd Street #2, Seattle, WA 98105 (US).

(74) Agent: **WALSH, Brian, J.**; ZymoGenetics, Inc., 1201 Eastlake Avenue East, Seattle, WA 98102 (US).

(81) Designated States *(national)*: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, OM, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, TJ, TM, TN, TR, TT, TZ, UA, UG, UZ, VC, VN, YU, ZA, ZM, ZW.

(84) Designated States *(regional)*: ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, SE, SI, SK, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:
— *without international search report and to be republished upon receipt of that report*

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

WO 2004/003140 A2

(54) Title: CYTOKINE RECEPTOR ZCYTOR17 MULTIMERS

(57) Abstract: Novel polypeptide combinations, polynucleotides encoding the polypeptides, and related compositions and methods are disclosed for zcytor17-containing multimeric or heterodimer cytokine receptors that may be used as novel cytokine antagonists, and within methods for detecting ligands that stimulate the proliferation and/or development of hematopoietic, lymphoid and myeloid cells in vitro and in vivo. The present invention also includes methods for producing the multimeric or heterodimeric cytokine receptor, uses therefor and antibodies thereto.

DEMANDE OU BREVET VOLUMINEUX

LA PRÉSENTE PARTIE DE CETTE DEMANDE OU CE BREVET COMPREND PLUS D'UN TOME.

CECI EST LE TOME 1 DE 2
CONTENANT LES PAGES 1 À 314

NOTE : Pour les tomes additionels, veuillez contacter le Bureau canadien des brevets

JUMBO APPLICATIONS/PATENTS

THIS SECTION OF THE APPLICATION/PATENT CONTAINS MORE THAN ONE VOLUME

THIS IS VOLUME 1 OF 2
CONTAINING PAGES 1 TO 314

NOTE: For additional volumes, please contact the Canadian Patent Office

NOM DU FICHIER / FILE NAME :

NOTE POUR LE TOME / VOLUME NOTE:

CYTOKINE RECEPTOR ZCYTOR17 MULTIMERS

BACKGROUND OF THE INVENTION

Proliferation and differentiation of cells of multicellular organisms are controlled by hormones and polypeptide growth factors. These diffusable molecules allow cells to communicate with each other and act in concert to form cells, tissues and organs, and to repair damaged tissue. Examples of hormones and growth factors include the steroid hormones (e.g., estrogen, testosterone), parathyroid hormone, follicle stimulating hormone, the interleukins, platelet derived growth factor (PDGF), epidermal growth factor (EGF), granulocyte-macrophage colony stimulating factor (GM-CSF), erythropoietin (EPO) and calcitonin.

Hormones and growth factors influence cellular metabolism by binding to receptors. Receptors may be integral membrane proteins that are linked to signaling pathways within the cell, such as second messenger systems. Other classes of receptors are soluble molecules, such as the transcription factors.

Cytokines generally stimulate proliferation or differentiation of cells of the hematopoietic lineage or participate in the immune and inflammatory response mechanisms of the body. Examples of cytokines which affect hematopoiesis are erythropoietin (EPO), which stimulates the development of red blood cells; thrombopoietin (TPO), which stimulates development of cells of the megakaryocyte lineage; and granulocyte-colony stimulating factor (G-CSF), which stimulates development of neutrophils. These cytokines are useful in restoring normal blood cell levels in patients suffering from anemia, thrombocytopenia, and neutropenia or receiving chemotherapy for cancer.

The interleukins are a family of cytokines that mediate immunological responses, including inflammation. The interleukins mediate a variety of inflammatory pathologies. Central to an immune response are T cells, which produce many cytokines and adaptive immunity to antigens. Cytokines produced by T cells have been classified as type 1 and type 2 (Kelso, A. *Immun. Cell Biol. 76*:300-317, 1998). Type 1 cytokines

include IL-2, IFN-γ, LT-α, and are involved in inflammatory responses, viral immunity, intracellular parasite immunity and allograft rejection. Type 2 cytokines include IL-4, IL-5, IL-6, IL-10 and IL-13, and are involved in humoral responses, helminth immunity and allergic response. Shared cytokines between Type 1 and 2 include IL-3, GM-CSF and TNF-α. There is some evidence to suggest that Type 1 and Type 2 producing T cell populations preferentially migrate into different types of inflamed tissue.

Mature T cells may be activated, i.e., by an antigen or other stimulus, to produce, for example, cytokines, biochemical signaling molecules, or receptors that further influence the fate of the T cell population.

B cells can be activated via receptors on their cell surface including B cell receptor and other accessory molecules to perform accessory cell functions, such as production of cytokines.

Monocytes/macrophages and T-cells can be activated by receptors on their cell surface and play a central role in the immune response by presenting antigen to lymphocytes and also act as accessory cells to lymphocytes by secreting numerous cytokines.

Natural killer (NK) cells have a common progenitor cell with T cells and B cells, and play a role in immune surveillance. NK cells, which comprise up to 15% of blood lymphocytes, do not express antigen receptors, and therefore do not use MHC recognition as requirement for binding to a target cell. NK cells are involved in the recognition and killing of certain tumor cells and virally infected cells. *In vivo*, NK cells are believed to require activation, however, *in vitro*, NK cells have been shown to kill some types of tumor cells without activation.

The demonstrated *in vivo* activities of these cytokines illustrate the enormous clinical potential of, and need for, other cytokines, cytokine agonists, and cytokine antagonists or binding partners. The present invention addresses these needs by providing a new hematopoietic multimeric cytokine receptor, as well as related compositions and methods.

The present invention provides such polypeptides for these and other uses that should be apparent to those skilled in the art from the teachings herein.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is an illustration of a multiple alignment of human zcytor17lig (SEQ ID NO:2) (zcytor17lig), and mouse zcytor17lig (SEQ ID NO:11) (mzcytor17lig), mouse IL-3 (mIL-3) (SEQ ID NO:100), and human IL-3 (hIL-3) (SEQ ID NO:102).

Figure 2 is an illustration of a multiple alignment of human zcytor17lig (SEQ ID NO:2) (zcytor17lig), and mouse zcytor17lig (SEQ ID NO:11) (mzcytor17lig).

Figure 3 is a Hopp/Woods hydrophilicity plot of human zcytor17lig (SEQ ID NO:2).

Figure 4 is a multiple alignment of zcytor17 polynucleotide sequences SEQ ID NO:109, SEQ ID NO:113, SEQ ID NO:5, SEQ ID NO:111, and SEQ ID NO:115.

Figure 5 is an alignment of human zcytor17 (ZCYTOR) (SEQ ID NO:5) and mouse zcytor17 (M17R-O) (SEQ ID NO:117). Between the two sequences, identical residues (:), Conserved residues (.) and gaps (-) are indicated.

SUMMARY OF THE INVENTION

The present invention provides an isolated multimeric or heterodimeric cytokine receptor comprising at least one polypeptide having at least 90 percent sequence identity with SEQ ID NO:111 or SEQ ID NO:109; and wherein the multimeric or heterodimeric cytokine receptor binds a ligand comprising SEQ ID NO:2. Optionally, the isolated multimeric or heterodimeric cytokine receptor may further comprise a cytokine-binding domain of a class I cytokine receptor. The cytokine-binding domain of the class I cytokine receptor may comprise amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 979 of SEQ ID NO:7, or amino acid residue 1 to amino acid residue 979 of SEQ ID NO:7. The isolated multimeric or heterodimeric cytokine receptor may antagonize an activity of SEQ ID NO:2. The isolated multimeric or heterodimeric cytokine receptor may inhibit

proliferation of hematopoietic cells, inhibit proliferation of immune cells, inhibit proliferation of inflammatory cells, inhibit an immune response, inhibit an inflammatory response, or inhibit proliferation of tumor cells of epithelial origin. The isolated multimeric or heterodimeric cytokine receptor may be soluble. The isolated multimeric or heterodimeric cytokine receptor may further comprises an affinity tag, such as, for instance, polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™ peptide, streptavidin binding peptide, and immunoglobulin F_c polypeptide, or cytotoxic molecule, such as, for instance, a toxin or radionuclide. The isolated multimeric or heterodimeric cytokine receptor wherein the polypeptide having at least 90 percent identity with SEQ ID NO:111 may comprise amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 227, amino acid residue 1 to amino acid residue 519, amino acid residue 1 to amino acid residue 543, or amino acid residue 1 to amino acid residue 732. The isolated multimeric or heterodimeric cytokine receptor wherein the polypeptide having at least 90 percent identity with SEQ ID NO:109 may comprise amino acid residue 1 to amino acid residue 649 of SEQ ID NO:109, or amino acid residue 20 to amino acid residue 649 of SEQ ID NO:109.

The present invention also provides an isolated multimeric or heterodimeric cytokine receptor comprising at least one polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111. The at least one polypeptide may comprise amino acid residue 1 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 732 of SEQ ID NO:111, or amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111. The isolated multimeric or heterodimeric cytokine receptor may further comprise a cytokine-binding domain of a class I cytokine receptor, for instance, amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 1 to amino acid

residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 979 of SEQ ID NO:7, or amino acid residue 1 to amino acid residue 979 of SEQ ID NO:7. The isolated multimeric or heterodimeric cytokine receptor may antagonize an activity of a ligand comprising SEQ ID NO:2. The isolated multimeric or heterodimeric cytokine receptor may inhibit proliferation of hematopoietic cells, inhibit proliferation of immune cells, inhibit proliferation of inflammatory cells, inhibit an immune response, inhibit an inflammatory response, or inhibit proliferation of tumor cells of epithelial origin. Optionally, the isolated multimeric or heterodimeric cytokine receptor may be is soluble. The isolated multimeric or heterodimeric cytokine receptor may further comprise an affinity tag, such as, for instance, polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™ peptide, streptavidin binding peptide, and immunoglobulin F_c polypeptide, or cytotoxic molecule, such as, for instance, a toxin or radionuclide.

The present invention also provides a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7.

The present invention also provides an isolated polynucleotide that encodes a cytokine receptor polypeptide comprising an amino acid sequence having at least 90 percent sequence identity with SEQ ID NO:111 or SEQ ID NO:109, wherein the cytokine receptor polypeptide forms a multimeric or heterodimeric cytokine receptor, and wherein the multimeric or heterodimeric cytokine receptor binds a ligand comprising SEQ ID NO:2. The multimeric or heterodimeric cytokine receptor may further comprise a cytokine-binding domain of a class I cytokine receptor, such as, for instance, amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 761 of SEQ ID NO:7, amino

acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 979 of SEQ ID NO:7, or amino acid residue 1 to amino acid residue 979 of SEQ ID NO:7. The multimeric or heterodimeric cytokine receptor may antagonize an activity of SEQ ID NO:2. The multimeric or heterodimeric cytokine receptor may inhibit proliferation of hematopoietic cells, inhibit proliferation of immune cells, inhibit proliferation of inflammatory cells, inhibit an immune response, inhibit an inflammatory response, or inhibit proliferation of tumor cells of epithelial origin. Optionally, the multimeric or heterodimeric cytokine receptor may be soluble. The multimeric or heterodimeric cytokine receptor may further comprise an affinity tag, such as, for instance, polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™ peptide, streptavidin binding peptide, and immunoglobulin F_c polypeptide, or cytotoxic molecule, such as, for instance, a toxin or radionuclide. The encoded cytokine receptor polypeptide having at least 90 percent identity with SEQ ID NO:111 may comprise amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 543 of SEQ ID NO:111, or amino acid residue 1 to amino acid residue 732 of SEQ ID NO:111. The encoded cytokine receptor polypeptide having at least 90 percent identity with SEQ ID NO:109 may comprise amino acid residue 1 to amino acid residue 649 of SEQ ID NO:109, or amino acid residue 20 to amino acid residue 649 of SEQ ID NO:109.

The present invention also provides an isolated polynucleotide that encodes a cytokine receptor polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111, wherein the cytokine receptor polypeptide forms a multimeric or heterodimeric cytokine receptor. The cytokine receptor polypeptide may comprise amino acid residue 1 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 543 of SEQ ID

NO:111, amino acid residue 1 to amino acid residue 732 of SEQ ID NO:111, or amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111. The multimeric or heterodimeric cytokine receptor may further comprise a cytokine-binding domain of a class I cytokine receptor, such as, for instance, amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 979 of SEQ ID NO:7, or amino acid residue 1 to amino acid residue 979 of SEQ ID NO:7. The multimeric or heterodimeric cytokine receptor may antagonize an activity of a ligand comprising SEQ ID NO:2. The multimeric or heterodimeric cytokine receptor may inhibit proliferation of hematopoietic cells, inhibit proliferation of immune cells, inhibit proliferation of inflammatory cells, inhibit an immune response, inhibit an inflammatory response, or inhibit proliferation of tumor cells of epithelial origin. Optionally, the multimeric or heterodimeric cytokine receptor may be soluble. The multimeric or heterodimeric cytokine receptor may further comprise an affinity tag or cytotoxic molecule as described herein.

The present invention also provides an expression vector that comprises the the following operably linked elements: a transcription promoter; a DNA segment encoding a cytokine receptor polypeptide having at least 90 percent sequence identity with SEQ ID NO:111; and a transcription terminator; wherein the cytokine receptor polypeptide forms a multimeric or heterodimeric cytokine receptor, and wherein the multimeric or heterodimeric cytokine receptor binds a ligand comprising SEQ ID NO:2.

Alternatively, the present invention also provides an expression vector that comprises the following operably linked elements: a) a first transcription promoter; a first DNA segment encoding a cytokine receptor polypeptide having at least 90 percent sequence identity with SEQ ID NO:111; and a first transcription terminator; and b) a second transcription promoter; a second DNA segment encoding a cytokine-binding domain of a class I cytokine receptor; and a second transcription terminator; wherein the cytokine receptor polypeptide and the class I cytokine receptor form a

multimeric or heterodimeric cytokine receptor; and wherein the multimeric or heterodimeric cytokine receptor binds to a ligand comprising SEQ ID NO:2.

Alternatively, the present invention also provides an expression vector that comprises the following operably linked elements: a) a first transcription promoter; a first DNA segment encoding a polypeptide having at least 90 percent sequence identity with SEQ ID NO:111; and a first transcription terminator; and b) a second transcription promoter; a second DNA segment encoding at least a portion of a class I cytokine receptor; and a second transcription terminator; wherein the polypeptide and the class I cytokine receptor form a multimeric cytokine receptor; and wherein the multimeric cytokine receptor binds to at least a portion of SEQ ID NO:2.

The expression vectors of the present invention may further include a secretory signal sequence linked to the first and second DNA segments. The multimeric or heterodimeric cytokine receptor may be soluble, membrane-bound, or attached to a solid support. The multimeric or heterodimeric cytokine receptor may antagonize an activity of a ligand comprising SEQ ID NO:2. The multimeric or heterodimeric cytokine receptor may inhibit proliferation of hematopoietic cells, inhibit proliferation of immune cells, inhibit proliferation of inflammatory cells, inhibit an immune response, inhibit an inflammatory response, or inhibit proliferation of tumor cells of epithelial origin. Optionally, the multimeric or heterodimeric cytokine receptor may be soluble. The multimeric or heterodimeric cytokine receptor may further comprise an affinity tag or cytotoxic molecule as described herein.

The present invention also provides a cultured cell including an expression vector as discribed herein, wherein the cell expresses the polypeptide or polypeptides encoded by the DNA segment or segments. The cell may secrete the multimeric or heterodimeric cytokine receptor. The multimeric cytokine receptor may bind and/or antagonize an activity of SEQ ID NO:2 as further described herein.

The present invention also provides a cultured cell which includes a first expression vector comprising: a) a transcription promoter; b) a DNA segment encoding a cytokine receptor polypeptide having at least 90 percent sequence identity with SEQ ID NO:111; and c) a transcription terminator; and a second expression vector comprising: a) a transcription promoter; b) a DNA segment encoding a cytokine-

binding domain of a class I cytokine receptor; and c) a transcription terminator; wherein the cytokine receptor polypeptide and the class I cytokine receptor form a multimeric or heterodimeric cytokine receptor, and wherein the multimeric or heterodimeric cytokine receptor binds to a ligand that comprises SEQ ID NO:2. The first and second expression vectors may include a secretory signal sequence operably linked to the first and second DNA segments. The cultured cell may further comprise a third expression vector which includes a) a transcription promoter; b) a DNA segment encoding a cytokine-binding domain of a second class I cytokine receptor; and c) a transcription terminator; wherein the cytokine receptor polypeptide, the first class I cytokine receptor, and the second class I cytokine receptor form a multimeric cytokine receptor. The cytokine-binding domain of a class I cytokine receptor may be of SEQ ID NO:7 and/or SEQ ID NO:9. Optionally, the multimeric or heterodimeric cytokine receptor may be soluble. The multimeric or heterodimeric cytokine receptor may further include an affinity tag as described herein. The multimeric or heterodimeric cytokine receptor may bind to at least a portion of SEQ ID NO:2 and/or antagonize an activity of SEQ ID NO:2 as described herein.

The present invention also provides a method of producing an antibody to a multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor. The method includes inoculating an animal with the multimeric or heterodimeric cytokine receptor, wherein the multimeric or heterodimeric cytokine receptor elicits an immune response in the animal to produce an antibody that specifically binds the multimeric or heterodimeric cytokine receptor; and isolating the antibody from the animal. The antibody may optionally be a monoclonal antibody. The antibody may optionally be a neutralizing antibody. The antibody may specifically bind to a multimeric or heterodimeric cytokine receptor as described herein.

The present invention also provides a composition which includes an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and a pharmaceutically acceptable vehicle. The binding domain of the class I cytokine receptor may include amino acid

residue 28 to amino acid residue 429 of SEQ ID NO:7. The soluble multimeric or heterodimeric cytokine receptor may bind to a ligand comprising SEQ ID NO:2. The soluble multimeric or heterodimeric cytokine receptor may further include an affinity tag or cytotoxic molecule as described herein. The composition may antagonize an activity of a ligand comprising SEQ ID NO:2. The composition may inhibit proliferation of hematopoietic cells, inhibit proliferation of immune cells, inhibit proliferation of inflammatory cells, inhibit an immune response, inhibit an inflammatory response, or inhibit proliferation of tumor cells of epithelial origin

The present invention also provides a method of producing a multimeric or heterodimeric cytokine receptor comprising culturing a cell as described herein, and isolating the multimeric or heterodimeric cytokine receptor produced by the cell.

The present invention also provides an immune cell inhibiting composition which includes an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and a pharmaceutically acceptable vehicle; wherein the soluble multimeric or heterodimeric cytokine receptor inhibits the proliferation of immune cells.

The present invention also provides an immune response inhibiting composition which includes an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and a pharmaceutically acceptable vehicle; wherein the soluble multimeric or heterodimeric cytokine receptor inhibits an immune response.

The present invention also provides an inflammatory cell inhibiting composition which includes an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and a pharmaceutically acceptable vehicle; wherein the soluble multimeric or heterodimeric cytokine receptor inhibits the proliferation of inflammatory cells.

The present invention also provides an inflammatory response inhibiting composition which includes an effective amount of a soluble multimeric or

heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and a pharmaceutically acceptable vehicle; wherein the soluble multimeric or heterodimeric cytokine receptor inhibits an inflammatory response.

The present invention also provides a method of inhibiting an immune response in a mammal exposed to an antigen or pathogen. The method includes (a) determining directly or indirectly the level of antigen or pathogen present in the mammal; (b) administering a composition comprising a soluble multimeric or heterodimeric cytokine receptor in a pharmaceutically acceptable vehicle; (c) determining directly or indirectly the level of antigen or pathogen in the mammal; and (d) comparing the level of the antigen or pathogen in step (a) to the antigen or pathogen level in step (c), wherein a change in the level is indicative of inhibiting an immune response. The method may further comprise (e) re-administering a composition comprising a multimeric cytokine receptor in a pharmaceutically acceptable vehicle; (f) determining directly or indirectly the level of antigen or pathogen in the mammal; and (g) comparing the number of the antigen or pathogen level in step (a) to the antigen level in step (f), wherein a change in the level is indicative of inhibiting an immune response.

The present invention also provides a method for reducing hematopoietic cells and/or hematopoietic progenitors cells in a mammal. The method includes culturing bone marrow or peripheral blood cells with a composition comprising an effective amount of a soluble multimeric or heterodimeric cytokine receptor to produce a decrease in the number of lymphoid cells in the bone marrow or peripheral blood cells as compared to bone marrow or peripheral blood cells cultured in the absence of the multimeric cytokine receptor. The hematopoietic cells and hematopoietic cell progenitors may be lymphoid, which can be monocytic cells, macrophages, or T cells.

The present invention also provides a method of detecting the presence of a multimeric or heterodimeric cytokine receptor in a biological sample. The method includes contacting the biological sample with an antibody, or an antibody fragment, as described herein, wherein the contacting is performed under conditions that allow the

binding of the antibody or antibody fragment to the biological sample; and detecting any of the bound antibody or bound antibody fragment.

The present invention also provides a method of a method of killing cancer cells. The method includes obtaining *ex vivo* a tissue or biological sample containing cancer cells from a patient, or identifying cancer cells *in vivo*; producing a multimeric or heterodimeric cytokine receptor by a method as described herein; formulating the multimeric or heterodimeric cytokine receptor in a pharmaceutically acceptable vehicle; and administering to the patient or exposing the cancer cells to the multimeric or heterodimeric cytokine receptor formulation; wherein the multimeric or heterodimeric cytokine receptor kills the cells. The multimeric or heterodimeric cytokine receptor may be further conjugated to a toxin.

The present invention also provides an antibody that specifically binds to a multimerc or heterodimeric cytokine receptor as described herein. The antibody may be a polyclonal antibody, a murine monoclonal antibody, a humanized antibody derived from a murine monoclonal antibody, an antibody fragment, a neutralizing antibody, or a human monoclonal antibody. The antibody or antibody fragment may specifically bind to a multimeric or heterodimeric cytokine receptor of the present invention which may comprise a cytokine receptor polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor. The antibody may further include a radionuclide, enzyme, substrate, cofactor, fluorescent marker, chemiluminescent marker, peptide tag, magnetic particle, drug, or toxin.

The present invention also provides a method for inhibiting zcytor17lig-induced proliferation or differentiation of hematopoietic cells and hematopoietic progenitor cells. The method includes culturing bone marrow or peripheral blood cells with a composition comprising an amount of a soluble multimeric or heterodimeric cytokine receptor comprising a cytokine receptor polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor sufficient to reduce proliferation or differentiation of the hematopoietic cells in the bone marrow or peripheral blood cells as compared to bone marrow or peripheral blood cells cultured in the absence of the soluble multimeric

or heterodimeric cytokine receptor. The hematopoietic cells and hematopoietic progenitor cells may be lymphoid cells, such as macrophages or T cells.

The present invention also provides a method of reducing zcytor17lig-induced induced inflammation. The method includes administering to a mammal with inflammation an amount of a composition comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor sufficient to reduce inflammation.

The present invention also provides a method of suppressing an inflammatory response in a mammal with inflammation. The method includes (1) determining a level of an inflammatory molecule; (2) administering a composition comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor in a pharmaceutically acceptable vehicle; (3) determining a post administration level of the inflammatory molecule; (4) comparing the level of the inflammatory molecule in step (1) to the level of the inflammatory molecule in step (3), wherein a lack of increase or a decrease the inflammatory molecule level is indicative of suppressing an inflammatory response.

The present invention also provides a method for inhibiting zcytor17lig-induced proliferation or differentiation of hematopoietic cells and hematopoietic progenitor cells. The method includes culturing bone marrow or peripheral blood cells with a composition comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor in a pharmaceutically acceptable vehicle sufficient to reduce proliferation or differentiation of the hematopoietic cells in the bone marrow or peripheral blood cells as compared to bone marrow or peripheral blood cells cultured in the absence of soluble multimeric or heterodimeric cytokine receptor. The hematopoietic cells and hematopoietic progenitor cells may be lymphoid cells, such as macrophages or T cells.

The present invention also provides a method of reducing zcytor17lig-induced induced inflammation. The method includes administering to a mammal with inflammation an amount of a composition comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor in a pharmaceutically acceptable vehicle sufficient to reduce

inflammation.

The present invention also provides a method of suppressing an inflammatory response in a mammal with inflammation. The method includes (1) determining a level of an inflammatory molecule; (2) administering a composition comprising a multimeric or heterodimeric cytokine receptor which comprises amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 in a pharmaceutically acceptable vehicle; (3) determining a post administration level of the inflammatory molecule; (4) comparing the level of the inflammatory molecule in step (1) to the level of the inflammatory molecule in step (3), wherein a lack of increase or a decrease in the inflammatory molecule level is indicative of suppressing an inflammatory response.

The present invention also provides a method of treating a mammal afflicted with an inflammatory disease in which zcytor17lig plays a role. The method includes administering an antagonist of zcytor17lig to the mammal such that the inflammation is reduced, wherein the antagonist is a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor in a pharmaceutically acceptable vehicle. The inflammatory disease may be a chronic inflammatory disease, such as, for instance, inflammatory bowel disease, ulcerative colitis, Crohn's disease, atopic dermatitis, eczema, or psoriasis. The inflammatory disease may be an acute inflammatory disease, such as, for instance, endotoxemia, septicemia, toxic shock syndrome, or infectious disease. Optinally, the soluble multimeric or heterodimeric cytokine receptor may further comprise a radionuclide, enzyme, substrate, cofactor, fluorescent marker, chemiluminescent marker, peptide tag, magnetic particle, drug, or toxin.

The present invention also provides a method for detecting inflammation in a patient. The method includes obtaining a tissue or biological sample from a patient; incubating the tissue or biological sample with a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor under conditions wherein the soluble multimeric or heterodimeric cytokine receptor binds to its complementary polypeptide in the tissue or biological sample;

visualizing the soluble multimeric or heterodimeric cytokine receptor bound in the tissue or biological sample; and comparing levels of soluble multimeric or heterodimeric cytokine receptor bound in the tissue or biological sample from the patient to a normal control tissue or biological sample, wherein an increase in the level of soluble multimeric or heterodimeric cytokine receptor bound to the patient tissue or biological sample relative to the normal control tissue or biological sample is indicative of inflammation in the patient.

The present invention also provides a method for detecting a multiple cytokine receptor ligand from a test sample. The method includes contacting the test sample with a multimeric or heterodimeric cytokine receptor comprising a cytokine receptor polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and detecting the binding of the multimeric or heterodimeric cytokine receptor to the ligand in the test sample.

DETAILED DESCRIPTION OF THE INVENTION

Definitions

Prior to setting forth the invention in detail, it may be helpful to the understanding thereof to define the following terms:

Unless otherwise specified, "a," "an," "the," and "at least one" are used interchangeably and mean one or more than one.

The term “affinity tag” is used herein to denote a polypeptide segment that can be attached to a second polypeptide to provide for purification or detection of the second polypeptide or provide sites for attachment of the second polypeptide to a substrate. In principal, any peptide or protein for which an antibody or other specific binding agent is available can be used as an affinity tag. Affinity tags include a poly-histidine tract, protein A (Nilsson et al., *EMBO J. 4*:1075, 1985; Nilsson et al., *Methods Enzymol. 198*:3, 1991), glutathione S transferase (Smith and Johnson, *Gene 67*:31, 1988), Glu-Glu affinity tag (Grussenmeyer et al., *Proc. Natl. Acad. Sci. USA 82*:7952-4, 1985), substance P, Flag™ peptide (Hopp et al., *Biotechnology 6*:1204-10, 1988), streptavidin binding peptide, or other antigenic epitope or binding domain. See, in

general, Ford et al., *Protein Expression and Purification 2*: 95-107, 1991. DNAs encoding affinity tags are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, NJ).

The term "allelic variant" is used herein to denote any of two or more alternative forms of a gene occupying the same chromosomal locus. Allelic variation arises naturally through mutation, and may result in phenotypic polymorphism within populations. Gene mutations can be silent (no change in the encoded polypeptide) or may encode polypeptides having altered amino acid sequence. The term allelic variant is also used herein to denote a protein encoded by an allelic variant of a gene.

The terms “amino-terminal” and “carboxyl-terminal” are used herein to denote positions within polypeptides. Where the context allows, these terms are used with reference to a particular sequence or portion of a polypeptide to denote proximity or relative position. For example, a certain sequence positioned carboxyl-terminal to a reference sequence within a polypeptide is located proximal to the carboxyl terminus of the reference sequence, but is not necessarily at the carboxyl terminus of the complete polypeptide.

The term “complement/anti-complement pair” denotes non-identical moieties that form a non-covalently associated, stable pair under appropriate conditions. For instance, biotin and avidin (or streptavidin) are prototypical members of a complement/anti-complement pair. Other exemplary complement/anti-complement pairs include receptor/ligand pairs, antibody/antigen (or hapten or epitope) pairs, sense/antisense polynucleotide pairs, and the like. Where subsequent dissociation of the complement/anti-complement pair is desirable, the complement/anti-complement pair preferably has a binding affinity of $<10^{9}\ M^{-1}$.

The term “complements of a polynucleotide molecule” denotes a polynucleotide molecule having a complementary base sequence and reverse orientation as compared to a reference sequence. For example, the sequence 5' ATGCACGGG 3' is complementary to 5' CCCGTGCAT 3'.

The term “contig” denotes a polynucleotide that has a contiguous stretch of identical or complementary sequence to another polynucleotide. Contiguous sequences are said to “overlap” a given stretch of polynucleotide sequence either in

their entirety or along a partial stretch of the polynucleotide. For example, representative contigs to the polynucleotide sequence 5'-ATGGCTTAGCTT-3' are 5'-TAGCTTgagtct-3' and 3'-gtcgacTACCGA-5'.

The term "degenerate nucleotide sequence" denotes a sequence of nucleotides that includes one or more degenerate codons (as compared to a reference polynucleotide molecule that encodes a polypeptide). Degenerate codons contain different triplets of nucleotides, but encode the same amino acid residue (i.e., GAU and GAC triplets each encode Asp).

The term "expression vector" is used to denote a DNA molecule, linear or circular, that comprises a segment encoding a polypeptide of interest operably linked to additional segments that provide for its transcription. Such additional segments include promoter and terminator sequences, and may also include one or more origins of replication, one or more selectable markers, an enhancer, a polyadenylation signal, etc. Expression vectors are generally derived from plasmid or viral DNA, or may contain elements of both.

The term "isolated", when applied to a polynucleotide, denotes that the polynucleotide has been removed from its natural genetic milieu and is thus free of other extraneous or unwanted coding sequences, and is in a form suitable for use within genetically engineered protein production systems. Such isolated molecules are those that are separated from their natural environment and include cDNA and genomic clones. Isolated DNA molecules of the present invention are free of other genes with which they are ordinarily associated, but may include naturally occurring 5' and 3' untranslated regions such as promoters and terminators. The identification of associated regions will be evident to one of ordinary skill in the art (see for example, Dynan and Tijan, *Nature 316*:774-78, 1985).

An "isolated" polypeptide or protein is a polypeptide or protein that is found in a condition other than its native environment, such as apart from blood and animal tissue. In a preferred form, the isolated polypeptide is substantially free of other polypeptides, particularly other polypeptides of animal origin. It is preferred to provide the polypeptides in a highly purified form, i.e. greater than 95% pure, more preferably greater than 99% pure. When used in this context, the term "isolated" does not exclude

the presence of the same polypeptide in alternative physical forms, such as dimers or alternatively glycosylated or derivatized forms.

The term "neoplastic", when referring to cells, indicates cells undergoing new and abnormal proliferation, particularly in a tissue where in the proliferation is uncontrolled and progressive, resulting in a neoplasm. The neoplastic cells can be either malignant, i.e., invasive and metastatic, or benign.

The term "operably linked", when referring to DNA segments, indicates that the segments are arranged so that they function in concert for their intended purposes, e.g., transcription initiates in the promoter and proceeds through the coding segment to the terminator.

The term "ortholog" denotes a polypeptide or protein obtained from one species that is the functional counterpart of a polypeptide or protein from a different species. Sequence differences among orthologs are the result of speciation.

"Paralogs" are distinct but structurally related proteins made by an organism. Paralogs are believed to arise through gene duplication. For example, α-globin, β-globin, and myoglobin are paralogs of each other.

A "polynucleotide" is a single- or double-stranded polymer of deoxyribonucleotide or ribonucleotide bases read from the 5' to the 3' end. Polynucleotides include RNA and DNA, and may be isolated from natural sources, synthesized *in vitro*, or prepared from a combination of natural and synthetic molecules. Sizes of polynucleotides are expressed as base pairs (abbreviated "bp"), nucleotides ("nt"), or kilobases ("kb"). Where the context allows, the latter two terms may describe polynucleotides that are single-stranded or double-stranded. When the term is applied to double-stranded molecules it is used to denote overall length and will be understood to be equivalent to the term "base pairs". It will be recognized by those skilled in the art that the two strands of a double-stranded polynucleotide may differ slightly in length and that the ends thereof may be staggered as a result of enzymatic cleavage; thus all nucleotides within a double-stranded polynucleotide molecule may not be paired.

A "polypeptide" is a polymer of amino acid residues joined by peptide bonds, whether produced naturally or synthetically. Polypeptides of less than about 10 amino acid residues are commonly referred to as "peptides".

The term "promoter" is used herein for its art-recognized meaning to denote a portion of a gene containing DNA sequences that provide for the binding of RNA polymerase and initiation of transcription. Promoter sequences are commonly, but not always, found in the 5' non-coding regions of genes.

A "protein" is a macromolecule comprising one or more polypeptide chains. A protein may also comprise non-peptidic components, such as carbohydrate groups. Carbohydrates and other non-peptidic substituents may be added to a protein by the cell in which the protein is produced, and will vary with the type of cell. Proteins are defined herein in terms of their amino acid backbone structures; substituents such as carbohydrate groups are generally not specified, but may be present nonetheless.

The term "receptor" denotes a cell-associated protein that binds to a bioactive molecule (i.e., a ligand) and mediates the effect of the ligand on the cell. Membrane-bound receptors are characterized by a multi-peptide structure comprising an extracellular ligand-binding domain and an intracellular effector domain that is typically involved in signal transduction. Binding of ligand to receptor results in a conformational change in the receptor that causes an interaction between the effector domain and other molecule(s) in the cell. This interaction in turn leads to an alteration in the metabolism of the cell. Metabolic events that are linked to receptor-ligand interactions include gene transcription, phosphorylation, dephosphorylation, increases in cyclic AMP production, mobilization of cellular calcium, mobilization of membrane lipids, cell adhesion, hydrolysis of inositol lipids and hydrolysis of phospholipids. In general, receptors can be membrane bound, cytosolic or nuclear; monomeric (e.g., thyroid stimulating hormone receptor, beta-adrenergic receptor) or multimeric (e.g., PDGF receptor, growth hormone receptor, IL-3 receptor, GM-CSF receptor, G-CSF receptor, erythropoietin receptor and IL-6 receptor).

The term "secretory signal sequence" denotes a DNA sequence that encodes a polypeptide (a "secretory peptide") that, as a component of a larger polypeptide, directs the larger polypeptide through a secretory pathway of a cell in which it is synthesized. The larger polypeptide is commonly cleaved to remove the secretory peptide during transit through the secretory pathway.

A "soluble receptor" is a receptor polypeptide that is not bound to a cell membrane. Soluble receptors are most commonly ligand-binding receptor polypeptides that lack transmembrane and cytoplasmic domains. Soluble receptors can comprise additional amino acid residues, such as affinity tags that provide for purification of the polypeptide or provide sites for attachment of the polypeptide to a substrate, or immunoglobulin constant region sequences. Many cell-surface receptors have naturally occurring, soluble counterparts that are produced by proteolysis. Soluble receptor polypeptides are said to be substantially free of transmembrane and intracellular polypeptide segments when they lack sufficient portions of these segments to provide membrane anchoring or signal transduction, respectively.

The term "splice variant" is used herein to denote alternative forms of RNA transcribed from a gene. Splice variation arises naturally through use of alternative splicing sites within a transcribed RNA molecule, or less commonly between separately transcribed RNA molecules, and may result in several mRNAs transcribed from the same gene. Splice variants may encode polypeptides having altered amino acid sequence. The term splice variant is also used herein to denote a protein encoded by a splice variant of an mRNA transcribed from a gene.

Molecular weights and lengths of polymers determined by imprecise analytical methods (e.g., gel electrophoresis) will be understood to be approximate values. When such a value is expressed as "about" X or "approximately" X, the stated value of X will be understood to be accurate to ±10%.

The present invention is based in part upon the discovery of a novel multimeric cytokine receptor protein having the structure of a class I cytokine receptor, referred to herein as "multimeric cytokine receptor," or "zcytor17 multimeric cytokine receptor." The multimeric cytokine receptor includes at least a portion of a zcytor17 receptor subunit, disclosed in the commonly owned U.S. Patent Application Serial No. 09/892,949. Another receptor subunit polypeptide that may be included in the multimeric cytokine receptor of the present invention includes at least a portion of at least one polypeptide of a class I cytokine receptor, such as OSMRbeta and/or WSX-1. For example, the deduced amino acid sequence indicated that zcytor17 belongs to the

receptor subfamily that includes gp130, LIF, IL-12, oncostatinM receptor beta (OSMRbeta) (SEQ ID NO:7), WSX-1 receptors (SEQ ID NO:9) (Sprecher , CA et al., *Biochem. Biophys. Res. Comm.*, *246*:81-90 (1998); and U.S. Patent No. 5,925,735), DCRS2 (WIPO Publication No. WO 00/73451), the IL-2 receptor β-subunit and the β-common receptor (i.e., IL-3, IL-5, and GM-CSF receptor subunits). A further example of class I cytokine receptor subunit polypeptides that may be included in the multimeric cytokine receptor are the receptors for IL-2, IL-4, IL-7, Lif, IL-12, IL-15, EPO, TPO, GM-CSF and G-CSF (Cosman, *Cytokine*, *5*(2):95-106 (1993)).

Cytokine receptor subunits are characterized by a multi-domain structure comprising an extracellular domain, a transmembrane domain that anchors the polypeptide in the cell membrane, and an intracellular domain. The extracellular domain may be a ligand-binding domain, and the intracellular domain may be an effector domain involved in signal transduction, although ligand-binding and effector functions may reside on separate subunits of a multimeric receptor. The ligand-binding domain may itself be a multi-domain structure. Multimeric receptors include homodimers (e.g., PDGF receptor αα and ββ isoforms, erythropoietin receptor, MPL, and G-CSF receptor), heterodimers whose subunits each have ligand-binding and effector domains (e.g., PDGF receptor αβ isoform), and multimers having component subunits with disparate functions (e.g., IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, and GM-CSF receptors). Some receptor subunits are common to a plurality of receptors. For example, the AIC2B subunit, which cannot bind ligand on its own but includes an intracellular signal transduction domain, is a component of IL-3 and GM-CSF receptors. Many cytokine receptors can be placed into one of four related families on the basis of the structure and function. Hematopoietic receptors, for example, are characterized by the presence of a domain containing conserved cysteine residues and the WSXWS motif (SEQ ID NO:3). Cytokine receptor structure has been reviewed by Urdal, *Ann. Reports Med. Chem. 26*:221-228, 1991 and Cosman, *Cytokine 5*:95-106, 1993. Under selective pressure for organisms to acquire new biological functions, new receptor family members likely arise from duplication of existing receptor genes leading to the existence of multi-gene families. Family members thus contain vestiges of the ancestral gene, and these characteristic features can be exploited in the isolation

and identification of additional family members. Thus, the cytokine receptor superfamily is subdivided into several families, for example, the immunoglobulin family (including CSF-1, MGF, IL-1, and PDGF receptors); the hematopoietin family (including IL-2 receptor β-subunit, GM-CSF receptor α-subunit, GM-CSF receptor β-subunit; and G-CSF, EPO, IL-3, IL-4, IL-5, IL-6, IL-7, and IL-9 receptors); TNF receptor family (including TNF (p80) TNF (p60) receptors, CD27, CD30, CD40, Fas, and NGF receptor).

Analysis of the zcytor17 sequence suggests that it is a member of the same receptor subfamily as the gp130, LIF, IL-12, WSX-1, IL-2 receptor β-subunit, IL-3, IL-4, and IL-6 receptors. Certain receptors in this subfamily (e.g., G-CSF) associate to form homodimers that transduce a signal. Other members of the subfamily (e.g., gp130, IL-6, IL-11, and LIF receptors) combine with a second subunit (termed a β-subunit) to bind ligand and transduce a signal. Specific β-subunits associate with a plurality of specific cytokine receptor subunits. For example, the β-subunit gp130 (Hibi et al., *Cell 63*:1149-1157, 1990) associates with receptor subunits specific for IL-6, IL-11, and LIF (Gearing et al., *EMBO J. 10*:2839-2848, 1991; Gearing et al., U.S. Patent No. 5,284,755). Oncostatin M binds to a heterodimer of LIF receptor and gp130. CNTF binds to trimeric receptors comprising CNTF receptor, LIF receptor, and gp130 subunits.

A multimeric cytokine receptor of the present invention can be a heterodimer, trimer, tetramer, pentamer, and the like, comprising at least a portion of zcytor17 and at least a portion of a class I cytokine receptor. In addition, a multimeric cytokine receptor can be soluble, membrane-bound, or attached to a solid support. Analysis of the tissue distribution of the mRNA of the zcytor17 receptor revealed expression in activated CD4+ and CD8+ T-cell subsets, CD14+ monocytes, and weaker expression in CD19+ B-cells. Moreover, the mRNA was present in both resting or activated monocytic cell lines THP-1 (ATCC No. TIB-202), U937 (ATCC No. CRL-1593.2) and HL60 (ATCC No. CCL-240).

Nucleotide sequences of representative zcytor17-encoding DNA are described in SEQ ID NO:110 (from nucleotide 171 to 2366), with its deduced 732 amino acid sequence described in SEQ ID NO:111; SEQ ID NO:108 (from nucleotide

162 to 2108), with its deduced 649 amino acid sequence described in SEQ ID NO:109; and in SEQ ID NO:4 (from nucleotide 497 to 2482), with its deduced 662 amino acid sequence described in SEQ ID NO:5. In its entirety, the zcytor17 polypeptide (SEQ ID NO:111, SEQ ID NO:109 or SEQ ID NO:5) represents a full-length polypeptide segment (residue 1 (Met) to residue 732 (Val) of SEQ ID NO:111; residue 1 (Met) to residue 649 (Ile) of SEQ ID NO:109; residue 1 (Met) to residue 662 (Ile) of SEQ ID NO:5). The domains and structural features of the zcytor17 polypeptides are further described below.

Analysis of the zcytor17 polypeptide encoded by the DNA sequence of SEQ ID NO:110 revealed an open reading frame encoding 732 amino acids (SEQ ID NO:111) comprising a predicted secretory signal peptide of 19 amino acid residues (residue 1 (Met) to residue 19 (Ala) of SEQ ID NO:111), and a mature polypeptide of 713 amino acids (residue 20 (Ala) to residue 732 (Val) of SEQ ID NO:111). Analysis of the zcytor17 polypeptide encoded by the DNA sequence of SEQ ID NO:108 revealed an open reading frame encoding 649 amino acids (SEQ ID NO:109) comprising a predicted secretory signal peptide of 19 amino acid residues (residue 1 (Met) to residue 19 (Ala) of SEQ ID NO:109), and a mature polypeptide of 630 amino acids (residue 20 (Ala) to residue 649 (Ile) of SEQ ID NO:109). Analysis of the zcytor17 polypeptide encoded by the DNA sequence of SEQ ID NO:4 revealed an open reading frame encoding 662 amino acids (SEQ ID NO:5) comprising a predicted secretory signal peptide of 32 amino acid residues (residue 1 (Met) to residue 32 (Ala) of SEQ ID NO:5), and a mature polypeptide of 630 amino acids (residue 33 (Ala) to residue 662 (Ile) of SEQ ID NO:5). In addition to the WSXWS motif (SEQ ID NO:3) (corresponding to residues 211 to 215 of SEQ ID NO:111 and SEQ ID NO:109; and residues 224 to 228 of SEQ ID NO:5), the receptor comprises an extracellular domain (residues 20 (Ala) to 519 (Glu) of SEQ ID NO:111 and SEQ ID NO:109; residues 33 (Ala) to 532 (Glu) of SEQ ID NO:5) which includes a cytokine-binding domain of approximately 200 amino acid residues (residues 20 (Ala) to 227 (Pro) of SEQ ID NO:111 and SEQ ID NO:109; residues 33 (Ala) to 240 (Pro) of SEQ ID NO:5); a domain linker (residues 122 (Thr) to 125 (Pro) of SEQ ID NO:111 and SEQ ID NO:109; residues 135 (Thr) to 138 (Pro) of SEQ ID NO:111); a penultimate strand

region (residues 194 (Phe) to 202 (Arg) of SEQ ID NO:111 and SEQ ID NO:109; residues 207 (Phe) to 215 (Arg) of SEQ ID NO:5); a fibronectin type III domain (residues 228 (Cys) to 519 (Glu) of SEQ ID NO:111 and SEQ ID NO:109; residues 241 (Cys) to 532 (Glu) of SEQ ID NO:5); a transmembrane domain (residues 520 (Ile) to 543 (Leu) of SEQ ID NO:111 and SEQ ID NO:109; residues 533 (Ile) to 556 (Leu) of SEQ ID NO:5); complete intracellular signaling domain (residues 544 (Lys) to 732 (Val) of SEQ ID NO:111; residues 544 (Lys) to 649 (Ile) of SEQ ID NO:109; and residues 557 (Lys) to 662 (Ile) of SEQ ID NO:5) which contains a "Box I" signaling site (residues 554 (Trp) to 560 (Pro) of SEQ ID NO:111 and SEQ ID NO:109; residues 567 (Trp) to 573 (Pro) of SEQ ID NO:5), and a "Box II" signaling site (residues 617 (Gln) to 620 (Phe) of SEQ ID NO:111 and SEQ ID NO:109; residues 630 (Gln) to 633 (Phe) of SEQ ID NO:5). Those skilled in the art will recognize that these domain boundaries are approximate, and are based on alignments with known proteins and predictions of protein folding. In addition to these domains, conserved receptor features in the encoded receptor include (as shown in SEQ ID NO:111 and SEQ ID NO:109) a conserved Cys residue at position 30 (position 43 as shown in SEQ ID NO:5), CXW motif (wherein X is any amino acid) at positions 40-42 (positions 53-55 as shown in SEQ ID NO:5), Trp residue at position 170 (position 183 as shown in SEQ ID NO:5), and a conserved Arg residue at position 202 (position 215 as shown in SEQ ID NO:5). The corresponding polynucleotides encoding the zcytor17 polypeptide regions, domains, motifs, residues and sequences described above are as shown in SEQ ID NO:110, SEQ ID NO:108, and SEQ ID NO:4.

Moreover, truncated forms of the zcytor17 polypeptide appear to be naturally expressed. Both forms encode soluble zcytor17 receptors. A polynucleotide encoding a "long-form" of the soluble zcytor17 receptor, truncated within the fibronectin type III domain, is shown in SEQ ID NO:112 and the corresponding polypeptide is shown in SEQ ID NO:113. This truncated form encodes residues 1 (Met) through 324 (Lys) of SEQ ID NO:111 and SEQ ID NO:109), and thus comprises an intact signal sequence, WSXWS (SEQ ID NO:3) motif, linker, cytokine binding domain, penultimate strand, and conserved, Cys, CXW motif, Trp and Arg residues as described above. A polynucleotide encoding a "short-form" of the soluble zcytor17

receptor, truncated at the end of the cytokine binding domain is shown in SEQ ID NO:114 and the corresponding polypeptide is shown in SEQ ID NO:115. This truncated form encodes a 239 residue polypeptide that is identical to residues 1 (Met) through 225 (Glu) of SEQ ID NO:111 and SEQ ID NO:109 and then diverges, and thus comprises an intact signal sequence, WSXWS (SEQ ID NO:3) motif, linker, cytokine binding domain, penultimate strand, and conserved, Cys, CXW motif, Trp and Arg residues as described above. A multiple alignment of the truncated forms compared to the full-length forms of zcytor17 is shown in Figure 1.

Moreover, the zcytor17 cDNA of SEQ ID NO:110, SEQ ID NO:108, SEQ ID NO:112, and SEQ ID NO:114 encode polypeptides that may use an alternative initiating methionine (at nucleotide 75 of SEQ ID NO:110, at nucleotide 66 of SEQ ID NO:108, at nucleotide 66 of SEQ ID NO:112, and at nucleotide 66 of SEQ ID NO:114) that would encode a polypeptide in the same open reading frame (ORF) as the zcytor17 polypeptides of SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:113, and SEQ ID NO:115. Use of the alternative initiating methionine would add 32 amino acids (shown in SEQ ID NO:48) in-frame to the N-terminus of SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:113, and SEQ ID NO:111. In addition, nucleotide 536 of SEQ ID NO:4 may serve as an alternative initiating methionine, thus generating the same N-terminus (starting at amino acid 14 (Met) of SEQ ID NO:5) and signal polypeptide sequence, as SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:113, and SEQ ID NO:115. Moreover, the second Met at amino acid number 2 in the SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:113, and SEQ ID NO:115 sequences (similarly at amino acid number 15 (Met) in SEQ ID NO:5) may also serve as an alternative starting methionine for the polypeptides.

Nucleotide sequences of representative OSMRbeta-encoding DNA are described in SEQ ID NO:6 (from nucleotide 368 to 3304), with its deduced 979 amino acid sequence described in SEQ ID NO:7. In its entirety, the OSMRbeta polypeptide (SEQ ID NO:7) represents a full-length polypeptide segment (residue 1 (Met) to residue 979 (Cys) of SEQ ID NO:7. The domains and structural features of the OSMRbeta polypeptides are further described below.

Analysis of the OSMRbeta polypeptide encoded by the DNA sequence

of SEQ ID NO:6 revealed an open reading frame encoding 979 amino acids (SEQ ID NO:7) comprising a predicted secretory signal peptide of 27 amino acid residues (residue 1 (Met) to residue 27 (Ala) of SEQ ID NO:7), and a mature polypeptide of 952 amino acids (residue 28 (Glu) to residue 979 (Cys) of SEQ ID NO:7. In addition to the two WSXWS motifs (SEQ ID NO:3) (corresponding to residues 129 to 133 and residues 415 to 419 of SEQ ID NO:7), the receptor comprises an extracellular domain (residues 28 (Glu) to 739 (Ser) of SEQ ID NO:7); which includes a cytokine-binding domain of approximately 400 amino acid residues (residues 28 (Glu) to 429 (Ala) of SEQ ID NO:7, which includes two linker domains (residues 31 (Pro) to 34 (Pro) and residues 343 (Asn) to 347 (Thr)), three regions of cytokine binding (residues 35 (Val) to 137 (Glu), residues 240 (Pro) to 342 (Glu), and residues 348 (Asn) to 429 (Ala), an immuglobulin domain (residues 138 (Val) to 239 (Glu), two penultimate strand regions (residues 106 (His) to 115 (Lys) and residues 398 (Thr) to 405 (Arg) of SEQ ID NO:7), and a fibronectin type III domain (residues 430 (Pro) to 739 (Ser) of SEQ ID NO:7); a transmembrane domain (residues 740 (Met) to 761 (Leu) of SEQ ID NO:7); complete intracellular signaling domain (residues 762 (Lys) to 979 (Cys) of SEQ ID NO:7) which contains a "Box I" signaling site (residues 771 (Tyr) to 777 (Pro) of SEQ ID NO:7), and a "Box II" signaling site (residues 829 (Glu) to 832 (Leu) of SEQ ID NO:7). Those skilled in the art will recognize that these domain boundaries are approximate, and are based on alignments with known proteins and predictions of protein folding. In addition to these domains, conserved receptor features in the encoded receptor include (as shown in SEQ ID NO:7) conserved Trp residues at positions 52 and 353, a conserved Cys residue at position 288, CXW motif (wherein X is any amino acid) at positions 294-296, and a conserved Arg residue at position 405. The corresponding polynucleotides encoding the OSMRbeta polypeptide regions, domains, motifs, residues and sequences described above are as shown in SEQ ID NO:6.

The presence of transmembrane regions, and conserved and low variance motifs generally correlates with or defines important structural regions in proteins. Regions of low variance (e.g., hydrophobic clusters) are generally present in regions of structural importance (Sheppard, P. et al., *supra.*). Such regions of low variance often contain rare or infrequent amino acids, such as Tryptophan. The regions

flanking and between such conserved and low variance motifs may be more variable, but are often functionally significant because they may relate to or define important structures and activities such as binding domains, biological and enzymatic activity, signal transduction, cell-cell interaction, tissue localization domains and the like.

The regions of conserved amino acid residues in zcytor17, described above, can be used as tools to identify new family members. For instance, reverse transcription-polymerase chain reaction (RT-PCR) can be used to amplify sequences encoding the conserved regions from RNA obtained from a variety of tissue sources or cell lines. In particular, highly degenerate primers designed from the zcytor17 sequences are useful for this purpose. Designing and using such degenerate primers may be readily performed by one of skill in the art.

The present invention also contemplates a multimeric zcytor17 receptor, as detailed herein, which is capable of intracellular signaling. Such receptors may include at least a portion of at least one extracellular domain of a zcytor17 receptor, and an intracellular domain from a zcytor17 receptor or another class I cytokine receptor. In addition to the extracellular domain of zcytor17, the multimeric cytokine receptor can also include the extracellular domain of at least a portion of class I cytokine receptor, for instance, the ligand binding domains of OSMRbeta receptor and/or WSX-1 receptor. Alternatively, the multimeric cytokine receptor may include the extracellular domain of another receptor, such as another class I cytokine receptor, and the intracellular domain of zcytor17 to effect intracellular signaling.

The present invention further contemplates a multimeric cytokine receptor that is soluble. For example, a multimeric cytokine receptor may be, for instance, a heterodimer which includes, for example, a portion of the extracellular domain of zcytor17 and a portion of the extracellular domain of a class I cytokine receptor, such as OSMRbeta (SEQ ID NO:7) and/or WSX-1 (SEQ ID NO:9). Additionally, a soluble multimeric cytokine receptor may also include an affinity tag, such as an immuglobulin F_c polypeptide. The soluble multimeric cytokine receptor can be expressed as a fusion with an immunoglobulin heavy chain constant region, such as an F_c fragment, which contains two constant region domains and lacks the variable region. Such fusions are typically secreted as multimeric molecules wherein the F_c

portions are disulfide bonded to each other and two non-Ig polypeptides are arrayed in closed proximity to each other. Fusions of this type can be used for example, for dimerization, increasing stability and *in vivo* half-life, to affinity purify ligand, as *in vitro* assay tool or antagonist.

Through processes of cloning, and proliferation assays described in detail herein, a multimeric cytokine receptor of the present invention has been shown to bind a novel ligand polypeptide (zcytor17lig) (SEQ ID NO:2), disclosed in commonly owned U.S. Patent Application Serial No. 60/350,325 and commonly owned U.S. Patent Application Serial No. 60/375,323, with high specificity. Zcytor17lig was isolated from a cDNA library generated from activated human peripheral blood cells (hPBCs), which were selected for CD3. CD3 is a cell surface marker unique to cells of lymphoid origin, particularly T cells.

A zcytor17lig positive clone was isolated, and sequence analysis revealed that the polynucleotide sequence contained within the plasmid DNA was novel. The secretory signal sequence is comprised of amino acid residues 1 (Met) to 23 (Ala), and the mature polypeptide is comprised of amino acid residues 24 (Ser) to 164 (Thr) (as shown in SEQ ID NO:2). Further, N-Terminal sequencing analysis of purified zcytor17lig from 293T cells showed an N-terminus at residue 27 (Leu) as shown in SEQ ID NO:2, with the mature polypeptide comprised of amino acid residues 27 (Leu) to 164 (Thr) (as shown in SEQ ID NO:2).

In general, cytokines are predicted to have a four-alpha helix structure, with helices A, C and D being most important in ligand-receptor interactions, and are more highly conserved among members of the family. Referring to the human zcytor17lig amino acid sequence shown in SEQ ID NO:2, alignment of human zcytor17lig, human IL-3, and human cytokine amino acid sequences it is predicted that zcytor17lig helix A is defined by amino acid residues 38-52; helix B by amino acid residues 83-98; helix C by amino acid residues 104-117; and helix D by amino acid residues 137-152; as shown in SEQ ID NO:2. Structural analysis suggests that the A/B loop is long, the B/C loop is short and the C/D loop is long. This loop structure results in an up-up-down-down helical organization. Based on 4-helix bundle structure, the cysteine residues within zcytor17lig that are conserved correspond to amino acid

residues 72, 133, and 147 of SEQ ID NO:2; and 74, 137, and 151 of SEQ ID NO:11 described herein. Consistent cysteine placement is further confirmation of the four-helical-bundle structure. Also highly conserved in the zcytor17lig is the Glu residue as shown in SEQ ID NO:2 at residue 43.

Moreover, the predicted amino acid sequence of murine zcytor17lig shows 31% identity to the predicted human protein over the entire length of the sequences (SEQ ID NO:2 and SEQ ID NO:11). Based on comparison between sequences of human and murine zcytor17lig conserved residues were found in the regions predicted to encode alpha helices C and D. The corresponding polynucleotides encoding the human zcytor17lig polypeptide regions, domains, motifs, residues and sequences described herein are as shown in SEQ ID NO:1.

While helix D is relatively conserved between human and murine zcytor17lig, helix C is the most conserved. While both species have predominant acidic amino acids in this region, the differences may account for species specificity in interaction between zcytor17lig and its receptor, zcytor17, comprising monomeric, heterodimeric (e.g., zcytor17/OSMRbeta, WSX-1/OSMRbeta, zcytor17/WSX-1) or multimeric (e.g., zcytor17/OSMRbeta/WSX-1) receptors. Loop A/B and helix B of zcytor17lig are marginally conserved, and helix C through Loop C/D into helix D is most conserved between species; conservation through this region suggests that it is functionally significant. The D helices of human and murine zcytor17lig are also conserved. Zcytor17 receptor antagonists may be designed through mutations within zcytor17lig helix D. These may include truncation of the protein from residue Thr156 (SEQ ID NO:2), or conservation of residues that confer binding of the ligand to the receptor, but diminish signaling activity.

Four-helical bundle cytokines are also grouped by the length of their component helices. "Long-helix" form cytokines generally consist of between 24-30 residue helices, and include IL-6, ciliary neutrotrophic factor (CNTF), leukemia inhibitory factor (LIF) and human growth hormone (hGH). "Short-helix" form cytokines generally consist of between 18-21 residue helices and include IL-2, IL-4 and GM-CSF. Zcytor17lig is believed to be a new member of the short-helix form cytokine group. Studies using CNTF and IL-6 demonstrated that a CNTF helix can be

exchanged for the equivalent helix in IL-6, conferring CTNF-binding properties to the chimera. Thus, it appears that functional domains of four-helical cytokines are determined on the basis of structural homology, irrespective of sequence identity, and can maintain functional integrity in a chimera (Kallen et al., *J. Biol. Chem. 274*:11859-11867, 1999). Therefore, the helical domains of zcytor17lig may be useful for preparing chimeric fusion molecules, particularly with other short-helix form cytokines to determine and modulate receptor binding specificity. The present invention also envisions fusion proteins engineered with helix A and/or helix D, and fusion proteins that combine helical and loop domains from other short-form cytokines such as IL-2, IL-4, IL-15, Lif, IL-12, IL-3 and GM-CSF.

The polynucleotide sequence for human IL-2 is shown in SEQ ID NO:176 and the corresponding amino acid sequence is shown in SEQ ID NO:177. The secretory signal sequence is comprised of amino acid residues 1 (Met) to 20 (Ser) of SEQ ID NO:177; nucleotides 48 to 107 of SEQ ID NO:176. The mature polypeptide is comprised of amino acid residues 21 (Ala) to 156 (Thr) of SEQ ID NO:177; nucleotides 108 to 515 of SEQ ID NO:176. Helix A of human IL-2 is comprised of amino acid residues 27 (Thr) to 48 (Leu) of SEQ ID NO:177; nucleotides 126 to 191 of SEQ ID NO:176. Helix B of human IL-2 comprises Helix B1 and Helix B2. Helix B1 of human IL-2 is comprised of amino acid residues 73 (Ala) to 80 (Gln) of SEQ ID NO:177; nucleotides 264 to 287 of SEQ ID NO:176. Helix B2 of human IL-2 is comprised of amino acid residues 83 (Glu) to 92 (Val) of SEQ ID NO:177; nucleotides 294 to 323 of SEQ ID NO:176. Thus, Helix B (comprising Helices B1 and B2) of IL-2 is represented by the amino acid sequence of SEQ ID NO:183 (nucleotide sequence of SEQ ID NO:182) wherein amino acid residues 9 and 10 can be any amino acid. SEQ ID NO:183 is identical to amino acids 73 (Ala) to 92 (Val) of SEQ ID NO:177 wherein amino acids 81 and 82 are any amino acid. In a preferred form, Helix B of IL-2 comprises amino acids 73 (Ala) to 92 (Val) of SEQ ID NO:177; nucleotides 264 to 323 of SEQ ID NO:176. Helix C of human IL-2 is comprised of amino acid residues 102 (His) to 116 (Val) of SEQ ID NO:177 nucleotides 351 to 395 of SEQ ID NO:176. Helix D of human IL-2 is comprised of amino acid residues 134 (Thr) to 149 (Gln) of SEQ ID NO:177; nucleotides 447 to 494 of SEQ ID NO:176.

The polynucleotide sequence for human IL-4 is shown in SEQ ID NO:178 and the corresponding amino acid sequence is shown in SEQ ID NO:179. The secretory signal sequence is comprised of amino acid residues 1 (Met) to 24 (Gly) of SEQ ID NO:179; nucleotides 64 to 135 of SEQ ID NO:178. The mature polypeptide is comprised of amino acid residues 25 (His) to 153 (Ser) of SEQ ID NO:179; nucleotides 136 to 522 of SEQ ID NO:178. Helix A of human IL-4 is comprised of amino acid residues 30 (Thr) to 42 (Thr) of SEQ ID NO:179; nucleotides 151 to 189 of SEQ ID NO:178. Helix B of human IL-4 is comprised of amino acid residues 65 (Glu) to 83 (His) of SEQ ID NO:179; nucleotides 256 to 312 of SEQ ID NO:178. Helix C of human IL-4 is comprised of amino acid residues 94 (Ala) to 118 (Ala) of SEQ ID NO:179; nucleotides 343 to 417 of SEQ ID NO:178. Helix D of human IL-4 is comprised of amino acid residues 133 (Leu) to 151 (Cys) of SEQ ID NO:179; nucleotides 460 to 516 of SEQ ID NO:178.

The polynucleotide sequence for human GM-CSF is shown in SEQ ID NO:180 and the corresponding amino acid sequence is shown in SEQ ID NO:181. The secretory signal sequence is comprised of amino acid residues 1 (Met) to 17 (Ser) of SEQ ID NO:181; nucleotides 9 to 59 of SEQ ID NO:180. The mature polypeptide is comprised of amino acid residues 18 (Ala) to 144 (Glu) of SEQ ID NO:181; nucleotides 60 to 440 of SEQ ID NO:180. Helix A of human GM-CSF is comprised of amino acid residues 30 (Trp) to 44 (Asn) of SEQ ID NO:181; nucleotides 96 to 140 of SEQ ID NO:180. Helix B of human GM-CSF is comprised of amino acid residues 72 (Leu) to 81 (Gln) of SEQ ID NO:181; nucleotides 222 to 251 of SEQ ID NO:180. Helix C of human GM-CSF is comprised of amino acid residues 85 (Gly) to 103 (Gln) of SEQ ID NO:181; nucleotides 261 to 317 of SEQ ID NO:180. Helix D of human GM-CSF is comprised of amino acid residues 120 (Phe) to 131 (Leu) of SEQ ID NO:181; nucleotides 366 to 401 of SEQ ID NO:180.

The amino acid residues comprising helices A, B, C, and D, for human zcytor17lig, IL-3, IL-2, IL-4, and GM-CSF are shown in Table 1.

Table 1

	Helix A	Helix B	Helix C	Helix D	
zcytor17lig	38-52	83-98	104-117	137-152	of SEQ ID NO:2

IL-3	35-45	73-86	91-103	123-141	of SEQ ID NO:102
IL-2	27-48	73-92	102-116	134-149	of SEQ ID NO:177 or Helix B as described in SEQ ID NO:183
IL-4	30-42	65-83	94-118	133-151	of SEQ ID NO:179
GM-CSF	30-44	72-81	85-103	120-131	of SEQ ID NO:181

The present invention provides polynucleotide molecules, including DNA and RNA molecules that encode the zcytor17 polypeptides disclosed herein that can be included in the multimeric cytokine receptor. Those skilled in the art will recognize that, in view of the degeneracy of the genetic code, considerable sequence variation is possible among these polynucleotide molecules. SEQ ID NO:120, SEQ ID NO:121 and SEQ ID NO:122 are degenerate DNA sequences that encompass all DNAs that encode the zcytor17 polypeptide of SEQ ID NO:111, SEQ ID NO:109 and SEQ ID NO:5, respectively, and fragments thereof. Those skilled in the art will recognize that the degenerate sequences of SEQ ID NO:120, SEQ ID NO:121 and SEQ ID NO:122 also provide all RNA sequences encoding SEQ ID NO:111, SEQ ID NO:109 and SEQ ID NO:5 by substituting U for T. Thus, zcytor17 polypeptide-encoding polynucleotides comprising nucleotide 1 to nucleotide 2196 of SEQ ID NO:120, nucleotide 1 to nucleotide 1947 of SEQ ID NO:121, and nucleotide 1 to nucleotide 1986 of SEQ ID NO:122 and their RNA equivalents are contemplated by the present invention. Table 2 sets forth the one-letter codes used within SEQ ID NO:120, SEQ ID NO:121 and SEQ ID NO:122 to denote degenerate nucleotide positions. "Resolutions" are the nucleotides denoted by a code letter. "Complement" indicates the code for the complementary nucleotide(s). For example, the code Y denotes either C or T, and its complement R denotes A or G, A being complementary to T, and G being complementary to C.

TABLE 2

Nucleotide	Resolution	Complement	Resolution
A	A	T	T
C	C	G	G
G	G	C	C
T	T	A	A
R	A|G	Y	C|T
Y	C|T	R	A|G
M	A|C	K	G|T
K	G|T	M	A|C
S	C|G	S	C|G
W	A|T	W	A|T
H	A|C|T	D	A|G|T
B	C|G|T	V	A|C|G
V	A|C|G	B	C|G|T
D	A|G|T	H	A|C|T
N	A|C|G|T	N	A|C|G|T

The degenerate codons used in SEQ ID NO:120, SEQ ID NO:121 and SEQ ID NO:122, encompassing all possible codons for a given amino acid, are set forth in Table 3.

TABLE 3

Amino Acid	One Letter Code	Codons	Degenerate Codon
Cys	C	TGC TGT	TGY
Ser	S	AGC AGT TCA TCC TCG TCT	WSN
Thr	T	ACA ACC ACG ACT	ACN
Pro	P	CCA CCC CCG CCT	CCN
Ala	A	GCA GCC GCG GCT	GCN
Gly	G	GGA GGC GGG GGT	GGN
Asn	N	AAC AAT	AAY
Asp	D	GAC GAT	GAY
Glu	E	GAA GAG	GAR
Gln	Q	CAA CAG	CAR
His	H	CAC CAT	CAY
Arg	R	AGA AGG CGA CGC CGG CGT	MGN
Lys	K	AAA AAG	AAR
Met	M	ATG	ATG
Ile	I	ATA ATC ATT	ATH
Leu	L	CTA CTC CTG CTT TTA TTG	YTN
Val	V	GTA GTC GTG GTT	GTN
Phe	F	TTC TTT	TTY
Tyr	Y	TAC TAT	TAY
Trp	W	TGG	TGG
Ter	.	TAA TAG TGA	TRR
Asn|Asp	B		RAY
Glu|Gln	Z		SAR
Any	X		NNN

One of ordinary skill in the art will appreciate that some ambiguity is introduced in determining a degenerate codon, representative of all possible codons encoding each amino acid. For example, the degenerate codon for serine (WSN) can, in some circumstances, encode arginine (AGR), and the degenerate codon for arginine (MGN) can, in some circumstances, encode serine (AGY). A similar relationship exists between codons encoding phenylalanine and leucine. Thus, some polynucleotides encompassed by the degenerate sequence may encode variant amino acid sequences, but one of ordinary skill in the art can easily identify such variant sequences by reference to the amino acid sequences of SEQ ID NO:111, SEQ ID NO:109 and SEQ ID NO:5; or SEQ ID NO:117 and SEQ ID NO:119. Variant sequences can be readily tested for functionality as described herein.

One of ordinary skill in the art will also appreciate that different species can exhibit "preferential codon usage." In general, see, Grantham, et al., *Nuc. Acids Res. 8*:1893-912, 1980; Haas, et al. *Curr. Biol. 6*:315-24, 1996; Wain-Hobson et al., *Gene 13*:355-64, 1981; Grosjean and Fiers, *Gene 18*:199-209, 1982; Holm, *Nuc. Acids Res. 14*:3075-87, 1986; Ikemura, *J. Mol. Biol. 158*:573-97, 1982. As used herein, the term "preferential codon usage" or "preferential codons" is a term of art referring to protein translation codons that are most frequently used in cells of a certain species, thus favoring one or a few representatives of the possible codons encoding each amino acid (See Table 3). For example, the amino acid Threonine (Thr) may be encoded by ACA, ACC, ACG, or ACT, but in mammalian cells ACC is the most commonly used codon; in other species, for example, insect cells, yeast, viruses or bacteria, different Thr codons may be preferential. Preferential codons for a particular species can be introduced into the polynucleotides of the present invention by a variety of methods known in the art. Introduction of preferential codon sequences into recombinant DNA can, for example, enhance production of the protein by making protein translation more efficient within a particular cell type or species. Therefore, the degenerate codon sequences disclosed in SEQ ID NO:120, SEQ ID NO:121 and SEQ ID NO:122 serve as templates for optimizing expression of zcytor17 polynucleotides in various cell types and species commonly used in the art and disclosed herein. Sequences containing preferential codons can be tested and optimized for expression in various species, and

tested for functionality as disclosed herein.

As previously noted, the isolated polynucleotides of the present invention include DNA and RNA. Methods for preparing DNA and RNA are well known in the art. In general, RNA is isolated from a tissue or cell that produces large amounts of zcytor17 RNA. Such tissues and cells are identified by Northern blotting (Thomas, *Proc. Natl. Acad. Sci. USA 77*:5201, 1980), and include PBLs, spleen, thymus, bone marrow, prostate, and lymph tissues, human erythroleukemia cell lines, acute monocytic leukemia cell lines, other lymphoid and hematopoietic cell lines, and the like. Total RNA can be prepared using guanidinium isothiocyanate extraction followed by isolation by centrifugation in a CsCl gradient (Chirgwin et al., *Biochemistry 18*:52-94, 1979). Poly $(A)^+$ RNA is prepared from total RNA using the method of Aviv and Leder (*Proc. Natl. Acad. Sci. USA 69*:1408-12, 1972). Complementary DNA (cDNA) is prepared from poly$(A)^+$ RNA using known methods. In the alternative, genomic DNA can be isolated. Polynucleotides encoding zcytor17 polypeptides are then identified and isolated by, for example, hybridization or polymerase chain reaction (PCR) (Mullis, U.S. Patent No. 4,683,202).

A full-length clone encoding zcytor17 can be obtained by conventional cloning procedures. Complementary DNA (cDNA) clones are preferred, although for some applications (e.g., expression in transgenic animals) it may be preferable to use a genomic clone, or to modify a cDNA clone to include at least one genomic intron. Methods for preparing cDNA and genomic clones are well known and within the level of ordinary skill in the art, and include the use of the sequence disclosed herein, or parts thereof, for probing or priming a library. Expression libraries can be probed with antibodies to zcytor17, receptor fragments, or other specific binding partners.

The polynucleotides of the present invention can also be synthesized using DNA synthesis machines. Currently the method of choice is the phosphoramidite method. If chemically synthesized double stranded DNA is required for an application such as the synthesis of a gene or a gene fragment, then each complementary strand is made separately. The production of short polynucleotides (60 to 80 bp) is technically straightforward and can be accomplished by synthesizing the complementary strands and then annealing them. However, for producing longer polynucleotides (>300 bp),

special strategies are usually employed, because the coupling efficiency of each cycle during chemical DNA synthesis is seldom 100%. To overcome this problem, synthetic genes (double-stranded) are assembled in modular form from single-stranded fragments that are from 20 to 100 nucleotides in length.

An alternative way to prepare a full-length gene is to synthesize a specified set of overlapping oligonucleotides (40 to 100 nucleotides). After the 3' and 5' short overlapping complementary regions (6 to 10 nucleotides) are annealed, large gaps still remain, but the short base-paired regions are both long enough and stable enough to hold the structure together. The gaps are filled and the DNA duplex is completed via enzymatic DNA synthesis by *E. coli* DNA polymerase I. After the enzymatic synthesis is completed, the nicks are sealed with T4 DNA ligase. Double-stranded constructs are sequentially linked to one another to form the entire gene sequence which is verified by DNA sequence analysis. See Glick and Pasternak, *Molecular Biotechnology, Principles & Applications of Recombinant DNA*, (ASM Press, Washington, D.C. 1994); Itakura et al., *Annu. Rev. Biochem. 53*: 323-56, 1984 and Climie et al., *Proc. Natl. Acad. Sci. USA 87*:633-7, 1990. Moreover, other sequences are generally added that contain signals for proper initiation and termination of transcription and translation.

The present invention also provides reagents which will find use in diagnostic applications. For example, the zcytor17lig gene, a probe comprising zcytor17lig DNA or RNA or a subsequence thereof, can be used to determine if the zcytor17lig gene is present on a human chromosome, such as chromosome 12, or if a gene mutation has occurred. Zcytor17lig is located at the 12q24.31 region of chromosome 12 (Example 13). Detectable chromosomal aberrations at the zcytor17lig gene locus include, but are not limited to, aneuploidy, gene copy number changes, loss of heterozygosity (LOH), translocations, insertions, deletions, restriction site changes and rearrangements. Such aberrations can be detected using polynucleotides of the present invention by employing molecular genetic techniques, such as restriction fragment length polymorphism (RFLP) analysis, short tandem repeat (STR) analysis employing PCR techniques, and other genetic linkage analysis techniques known in the art (Sambrook et al., *ibid.*; Ausubel et. al., *ibid.*; Marian, *Chest 108*:255-65, 1995).

The precise knowledge of a gene's position can be useful for a number of purposes, including: 1) determining if a sequence is part of an existing contig and obtaining additional surrounding genetic sequences in various forms, such as YACs, BACs or cDNA clones; 2) providing a possible candidate gene for an inheritable disease which shows linkage to the same chromosomal region; and 3) cross-referencing model organisms, such as mouse, which may aid in determining what function a particular gene might have.

One of skill in the art would recognize that the 12q24 region is frequently involved in gross genomic rearrangements, including translocations, deletions, inversions, and duplications, that are associated with various cancers. The Mitelman Database of Chromosomal Aberrations in Cancer, at the Cancer Genome Anatomy Project, National Insitutes of Health, Bethesda, Md located on the Internet lists 199 cases of cancers with genomic rearrangements involving 12q24. Of these, most are part of complex karyotypes with other rearrangements; however, in some cases the rearrangement involving 12q24 is the only genomic alteration. Given the expression of the receptor for zcytor17lig on cells of lymphoid and myeloid lineages, it is particularly significant to note that there are at least 4 cases of myeloid leukemia reported in the literature in which either translocation (2 cases: Yamagata et al, *Cancer Genet Cytogenet 97*:90-93, 1997; Dunphy and Batanian, *Cancer Genet Cytogenet 114*:51-57, 1999) or duplication (2 cases: Bonomi et al, *Cancer Genet Cytogenet 108*:75-78, 1999) are the sole genomic alteration. This suggests that a gene or genes residing within 12q24 could be directly involved in the malignant transformation of these patients' cells. Inappropriate over expression of zcytor17lig could contribute to malignant transformation by promoting aberrant proliferation of receptor-bearing cells, through either autocrine or paracrine mechanisms. Inhibition of zcytor17lig activity could thus inhibit growth of such cells. Alternatively, a genomic rearrangement resulting in inactivation of the zcytor17lig gene may promote malignant transformation and/or metastasis by removing zcytor17lig immunoregulatory functions. Indeed, a gene suppressing metastasis in prostate cancer has been mapped to 12q24-qter (Ichikawa et al, *Asian J Androl 2*:167-171, 2000). If zcytor17lig is the gene within this region responsible for the suppression of metastasis, then zcytor17lig itself may have

therapeutic value in the treatment of cancer.

A diagnostic could assist physicians in determining the type of disease and appropriate associated therapy, or assistance in genetic counseling. As such, the inventive anti-zcytor17lig antibodies, polynucleotides, and polypeptides can be used for the detection of zcytor17lig polypeptide, mRNA or anti-zcytor17lig antibodies, thus serving as markers and be directly used for detecting or genetic diseases or cancers, as described herein, using methods known in the art and described herein. Further, zcytor17lig polynucleotide probes can be used to detect abnormalities or genotypes associated with chromosome 12q24.3 deletions and translocations associated with human diseases, or other translocations involved with malignant progression of tumors or other 12q24.3 mutations, which are expected to be involved in chromosome rearrangements in malignancy; or in other cancers. Similarly, zcytor17lig polynucleotide probes can be used to detect abnormalities or genotypes associated with chromosome 12 trisomy and chromosome loss associated with human diseases or spontaneous abortion. Thus, zcytor17lig polynucleotide probes can be used to detect abnormalities or genotypes associated with these defects.

One of skill in the art would recognize that zcytor17lig polynucleotide probes are particularly useful for diagnosis of gross chromosomal abnormalities associated with loss of heterogeneity (LOH), chromosome gain (e.g., trisomy), translocation, DNA amplification, and the like. Translocations within chromosomal locus 12q24.3 wherein the zcytor17lig gene is located are known to be associated with human disease. For example, 12q24 deletions and translocations, duplications and trisomy are associated with cancers as discussed above. Thus, since the zcytor17lig gene maps to this critical region, zcytor17lig polynucleotide probes of the present invention can be used to detect abnormalities or genotypes associated with 12q24 translocation, deletion and trisomy, and the like, described above.

As discussed above, defects in the zcytor17lig gene itself may result in a heritable human disease state. Molecules of the present invention, such as the polypeptides, antagonists, agonists, polynucleotides and antibodies of the present invention would aid in the detection, diagnosis prevention, and treatment associated with a zcytor17lig genetic defect. In addition, zcytor17lig polynucleotide probes can be

used to detect allelic differences between diseased or non-diseased individuals at the zcytor17lig chromosomal locus. As such, the zcytor17lig sequences can be used as diagnostics in forensic DNA profiling.

In general, the diagnostic methods used in genetic linkage analysis, to detect a genetic abnormality or aberration in a patient, are known in the art. Analytical probes will be generally at least 20 nt in length, although somewhat shorter probes can be used (e.g., 14-17 nt). PCR primers are at least 5 nt in length, preferably 15 or more, more preferably 20-30 nt. For gross analysis of genes, or chromosomal DNA, a zcytor17lig polynucleotide probe may comprise an entire exon or more. Exons are readily determined by one of skill in the art by comparing zcytor17lig sequences (SEQ ID NO:1) with the genomic DNA for mouse zcytor17lig (SEQ ID NO:76). In general, the diagnostic methods used in genetic linkage analysis, to detect a genetic abnormality or aberration in a patient, are known in the art. Most diagnostic methods comprise the steps of (a) obtaining a genetic sample from a potentially diseased patient, diseased patient or potential non-diseased carrier of a recessive disease allele; (b) producing a first reaction product by incubating the genetic sample with a zcytor17lig polynucleotide probe wherein the polynucleotide will hybridize to complementary polynucleotide sequence, such as in RFLP analysis or by incubating the genetic sample with sense and antisense primers in a PCR reaction under appropriate PCR reaction conditions; (iii) visualizing the first reaction product by gel electrophoresis and/or other known methods such as visualizing the first reaction product with a zcytor17lig polynucleotide probe wherein the polynucleotide will hybridize to the complementary polynucleotide sequence of the first reaction; and (iv) comparing the visualized first reaction product to a second control reaction product of a genetic sample from wild type patient, or a normal or control individual. A difference between the first reaction product and the control reaction product is indicative of a genetic abnormality in the diseased or potentially diseased patient, or the presence of a heterozygous recessive carrier phenotype for a non-diseased patient, or the presence of a genetic defect in a tumor from a diseased patient, or the presence of a genetic abnormality in a fetus or pre-implantation embryo. For example, a difference in restriction fragment pattern, length of PCR products, length of repetitive sequences at the zcytor17lig genetic locus, and the

like, are indicative of a genetic abnormality, genetic aberration, or allelic difference in comparison to the normal wild type control. Controls can be from unaffected family members, or unrelated individuals, depending on the test and availability of samples. Genetic samples for use within the present invention include genomic DNA, mRNA, and cDNA isolated from any tissue or other biological sample from a patient, which includes, but is not limited to, blood, saliva, semen, embryonic cells, amniotic fluid, and the like. The polynucleotide probe or primer can be RNA or DNA, and will comprise a portion of SEQ ID NO:1, the complement of SEQ ID NO:1, or an RNA equivalent thereof. Such methods of showing genetic linkage analysis to human disease phenotypes are well known in the art. For reference to PCR based methods in diagnostics see generally, Mathew (ed.), *Protocols in Human Molecular Genetics* (Humana Press, Inc. 1991), White (ed.), *PCR Protocols: Current Methods and Applications* (Humana Press, Inc. 1993), Cotter (ed.), *Molecular Diagnosis of Cancer* (Humana Press, Inc. 1996), Hanausek and Walaszek (eds.), *Tumor Marker Protocols* (Humana Press, Inc. 1998), Lo (ed.), *Clinical Applications of PCR* (Humana Press, Inc. 1998), and Meltzer (ed.), *PCR in Bioanalysis* (Humana Press, Inc. 1998).

Mutations associated with the zcytor17lig locus can be detected using nucleic acid molecules of the present invention by employing standard methods for direct mutation analysis, such as restriction fragment length polymorphism analysis, short tandem repeat analysis employing PCR techniques, amplification-refractory mutation system analysis, single-strand conformation polymorphism detection, RNase cleavage methods, denaturing gradient gel electrophoresis, fluorescence-assisted mismatch analysis, and other genetic analysis techniques known in the art (see, for example, Mathew (ed.), *Protocols in Human Molecular Genetics* (Humana Press, Inc. 1991), Marian, *Chest 108*:255 (1995), Coleman and Tsongalis, *Molecular Diagnostics* (Human Press, Inc. 1996), Elles (ed.) *Molecular Diagnosis of Genetic Diseases* (Humana Press, Inc. 1996), Landegren (ed.), *Laboratory Protocols for Mutation Detection* (Oxford University Press 1996), Birren *et al.* (eds.), *Genome Analysis, Vol. 2: Detecting Genes* (Cold Spring Harbor Laboratory Press 1998), Dracopoli *et al.* (eds.), *Current Protocols in Human Genetics* (John Wiley & Sons 1998), and Richards and Ward, "Molecular Diagnostic Testing," in *Principles of Molecular Medicine*, pages 83-

88 (Humana Press, Inc. 1998). Direct analysis of an zcytor17lig gene for a mutation can be performed using a subject's genomic DNA. Methods for amplifying genomic DNA, obtained for example from peripheral blood lymphocytes, are well-known to those of skill in the art (see, for example, Dracopoli *et al.* (eds.), *Current Protocols in Human Genetics*, at pages 7.1.6 to 7.1.7 (John Wiley & Sons 1998)).

The present invention further provides counterpart polypeptides and polynucleotides from other species (orthologs). These species include, but are not limited to mammalian, avian, amphibian, reptile, fish, insect and other vertebrate and invertebrate species. Of particular interest are zcytor17 polypeptides from other mammalian species, including murine, porcine, ovine, bovine, canine, feline, equine, and other primate polypeptides. Orthologs of human zcytor17 can be cloned using information and compositions provided by the present invention in combination with conventional cloning techniques. For example, a cDNA can be cloned using mRNA obtained from a tissue or cell type that expresses zcytor17 as disclosed herein. Suitable sources of mRNA can be identified by probing Northern blots with probes designed from the sequences disclosed herein. A library is then prepared from mRNA of a positive tissue or cell line. A zcytor17-encoding cDNA can then be isolated by a variety of methods, such as by probing with a complete or partial human cDNA or with one or more sets of degenerate probes based on the disclosed sequences. A cDNA can also be cloned using PCR (Mullis, *supra.*), using primers designed from the representative human zcytor17 sequence disclosed herein. Within an additional method, the cDNA library can be used to transform or transfect host cells, and expression of the cDNA of interest can be detected with an antibody to zcytor17 polypeptide. Similar techniques can also be applied to the isolation of genomic clones.

A polynucleotide sequence for the mouse ortholog of human zcytor17 has been identified and is shown in SEQ ID NO:116 and the corresponding amino acid sequence shown in SEQ ID NO:117. Analysis of the mouse zcytor17 polypeptide encoded by the DNA sequence of SEQ ID NO:116 revealed an open reading frame encoding 662 amino acids (SEQ ID NO:117) comprising a predicted secretory signal

peptide of 45 amino acid residues (residue 1 (Met) to residue 45 (Ala) of SEQ ID NO:117), and a mature polypeptide of 617 amino acids (residue46 (Val) to residue 662 (Cys) of SEQ ID NO:117). Moreover, an additional Met residue, Met (28) can be used as a starting methionine; comprising a second predicted secretory signal peptide of 18 amino acid residues (residue 28 (Met) to residue 45 (Ala) of SEQ ID NO:117), and the same mature polypeptide of 617 amino acids (residue46 (Val) to residue 662 (Cys) of SEQ ID NO:117. In addition to the WSXWS motif (SEQ ID NO:3) corresponding to residues 224-228 of SEQ ID NO:117, the receptor comprises an extracellular domain from residues 46 (Val) to 533 (Glu) of SEQ ID NO:117) that includes a cytokine-binding domain of approximately 200 amino acid residues (residues 46 (Val) to 240 (Pro) of SEQ ID NO:117) and a fibronectin III domain (residues 241 (His) to 533 (Glu) of SEQ ID NO:117); a CXW motif (residues 66 (Cys) to 68 (Trp) of SEQ ID NO:117); a domain linker (residues 142 (Thr) to 145 (Pro) of SEQ ID NO:117); a penultimate strand region (residues 207 (Phe) to 215 (Arg) of SEQ ID NO:117); a transmembrane domain (residues 534 (Ile) to 550 (Ile) of SEQ ID NO:117); complete intracellular signaling domain (residues 551 (Lys) to 662 (Cys) of SEQ ID NO:117) which contains a "Box I" signaling site (residues 568 (Cys) to 574 (Pro) of SEQ ID NO:117), and a "Box II" signaling site (residues 628 (Glu) to 631 (leu) of SEQ ID NO:117). Conserved residues common to class I cytokine receptors, are at residues 56 (Cys), 187 (Trp), and 215 (Arg). A comparison of the human and mouse amino acid sequences reveals that both the human and orthologous polypeptides contain corresponding structural features described above (and, see, Figure 2). The mature sequence for the mouse zcytor17 begins at Val_{46} (as shown in SEQ ID NO:117), which corresponds to Ala_{33} (as shown in SEQ ID NO:5) in the human sequence. There is about 61% identity between the mouse and human sequences over the entire amino acid sequence corresponding to SEQ ID NO:5 and SEQ ID NO:117. The above percent identity was determined using a FASTA program with ktup=1, gap opening penalty=12, gap extension penalty=2, and substitution matrix=BLOSUM62, with other parameters set as default. The corresponding polynucleotides encoding the mouse zcytor17 polypeptide regions, domains, motifs, residues and sequences described above are as shown in SEQ ID NO:116.

Moreover, a truncated soluble form of the mouse zcytor17 receptor polypeptide appears to be naturally expressed. A polynucleotide sequence for a truncated soluble form of the mouse zcytor17 receptor has been identified and is shown in SEQ ID NO:118 and the corresponding amino acid sequence shown in SEQ ID NO:119. Analysis of the truncated soluble mouse zcytor17 polypeptide encoded by the DNA sequence of SEQ ID NO:118 revealed an open reading frame encoding 547 amino acids (SEQ ID NO:119) comprising a predicted secretory signal peptide of 45 amino acid residues (residue 1 (Met) to residue 45 (Ala) of SEQ ID NO:119), and a mature polypeptide of 502 amino acids (residue46 (Val) to residue 547 (Val) of SEQ ID NO:119). Moreover, an additional Met residue, Met (28) can be used as a starting methionine; comprising a second predicted secretory signal peptide of 18 amino acid residues (residue 28 (Met) to residue 45 (Ala) of SEQ ID NO:119), and the same mature polypeptide of 502 amino acids (residue46 (Val) to residue 547 (Val) of SEQ ID NO:119. In addition to the WSXWS motif (SEQ ID NO:3) corresponding to residues 224-228 of SEQ ID NO:119, the receptor comprises an extracellular domain from residues 46 (Val) to 533 (Trp) of SEQ ID NO:119) that includes a cytokine-binding domain of approximately 200 amino acid residues (residues 46 (Val) to 240 (Pro) of SEQ ID NO:119) and a fibronectin III domain (residues 241 (His) to 533 (Trp) of SEQ ID NO:119); a CXW motif (residues 66 (Cys) to 68 (Trp) of SEQ ID NO:119); a domain linker (residues 142 (Thr) to 145 (Pro) of SEQ ID NO:119); a penultimate strand region (residues 207 (Phe) to 215 (Arg) of SEQ ID NO:119); and a C-terminal tail region (residues 534 (Leu) to 547 (Val). Conserved residues common to class I cytokine receptors, are at residues 56 (Cys), 187 (Trp), and 215 (Arg). A comparison of the human and mouse amino acid sequences, including the truncated soluble mouse zcytor17, reveals that both the human and orthologous polypeptides contain corresponding structural features described above (and, see, Figure 2). The corresponding polynucleotides encoding the truncated soluble mouse zcytor17 polypeptide regions, domains, motifs, residues and sequences described above are as shown in SEQ ID NO:118.

Those skilled in the art will recognize that the sequences disclosed in SEQ ID NO:110, SEQ ID NO:108 and SEQ ID NO:4 represent alleles of human

zcytor17 and that allelic variation and alternative splicing are expected to occur. Allelic variants of this sequence can be cloned by probing cDNA or genomic libraries from different individuals according to standard procedures. Allelic variants of the DNA sequence shown in SEQ ID NO:110, SEQ ID NO:108 or SEQ ID NO:4, including those containing silent mutations and those in which mutations result in amino acid sequence changes, are within the scope of the present invention, as are proteins which are allelic variants of SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:5 SEQ ID NO:117 or SEQ ID NO:119. cDNAs generated from alternatively spliced mRNAs, which retain the properties of the zcytor17 polypeptide are included within the scope of the present invention, as are polypeptides encoded by such cDNAs and mRNAs. Allelic variants and splice variants of these sequences can be cloned by probing cDNA or genomic libraries from different individuals or tissues according to standard procedures known in the art. For example, the short-form and long-form soluble zcytor17 receptors described above, and in SEQ ID NO:112 and SEQ ID NO:113 or SEQ ID NO:114 and SEQ ID NO:115 can be considered allelic or splice variants of zcytor17.

The present invention also provides isolated zcytor17 polypeptides that are substantially similar to the polypeptides of SEQ ID NO:111, SEQ ID NO:109 or SEQ ID NO:5 and their orthologs, e.g., SEQ ID NO:117 and SEQ ID NO:119. The term "substantially similar" is used herein to denote polypeptides having at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or greater than 99% sequence identity to the sequences shown in SEQ ID NO:111, SEQ ID NO:109 or SEQ ID NO:5 or their orthologs, e.g., SEQ ID NO:117 and SEQ ID NO:119. Such polypeptides will more preferably be at least 90% identical, and most preferably 95% or more identical to SEQ ID NO:111, SEQ ID NO:109 and SEQ ID NO:5 or its orthologs.) Percent sequence identity is determined by conventional methods. See, for example, Altschul et al., *Bull. Math. Bio. 48*: 603-616, 1986 and Henikoff and Henikoff, *Proc. Natl. Acad. Sci. USA 89*:10915-10919, 1992. Briefly, two amino acid sequences are aligned to optimize the alignment scores using a gap opening penalty of 10, a gap extension penalty of 1, and the "blosum 62" scoring matrix of Henikoff and Henikoff (*ibid.*) as shown in Table 4 (amino acids are indicated by the

standard one-letter codes). The percent identity is then calculated as:

$$\frac{\text{Total number of identical matches}}{\text{[length of the longer sequence plus the number of gaps introduced into the longer sequence in order to align the two sequences]}} \times 100$$

Table 4

	A	R	N	D	C	Q	E	G	H	I	L	K	M	F	P	S	T	W	Y	V
A	4																			
R	-1	5																		
N	-2	0	6																	
D	-2	-2	1	6																
C	0	-3	-3	-3	9															
Q	-1	1	0	0	-3	5														
E	-1	0	0	2	-4	2	5													
G	0	-2	0	-1	-3	-2	-2	6												
H	-2	0	1	-1	-3	0	0	-2	8											
I	-1	-3	-3	-3	-1	-3	-3	-4	-3	4										
L	-1	-2	-3	-4	-1	-2	-3	-4	-3	2	4									
K	-1	2	0	-1	-3	1	1	-2	-1	-3	-2	5								
M	-1	-1	-2	-3	-1	0	-2	-3	-2	1	2	-1	5							
F	-2	-3	-3	-3	-2	-3	-3	-3	-1	0	0	-3	0	6						
P	-1	-2	-2	-1	-3	-1	-1	-2	-2	-3	-3	-1	-2	-4	7					
S	1	-1	1	0	-1	0	0	0	-1	-2	-2	0	-1	-2	-1	4				
T	0	-1	0	-1	-1	-1	-1	-2	-2	-1	-1	-1	-1	-2	-1	1	5			
W	-3	-3	-4	-4	-2	-2	-3	-2	-2	-3	-2	-3	-1	1	-4	-3	-2	11		
Y	-2	-2	-2	-3	-2	-1	-2	-3	2	-1	-1	-2	-1	3	-3	-2	-2	2	7	
V	0	-3	-3	-3	-1	-2	-2	-3	-3	3	1	-2	1	-1	-2	-2	0	-3	-1	4

Sequence identity of polynucleotide molecules is determined by similar methods using a ratio as disclosed above.

Those skilled in the art appreciate that there are many established algorithms available to align two amino acid sequences. The "FASTA" similarity search algorithm of Pearson and Lipman is a suitable protein alignment method for examining the level of identity shared by an amino acid sequence disclosed herein and the amino acid sequence of a putative variant zcytor17. The FASTA algorithm is described by Pearson and Lipman, *Proc. Nat'l Acad. Sci. USA 85:*2444 (1988), and by Pearson, *Meth. Enzymol. 183*:63 (1990).

Briefly, FASTA first characterizes sequence similarity by identifying regions shared by the query sequence (*e.g.,* SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:5, SEQ ID NO:117 and SEQ ID NO:119) and a test sequence that have either the highest density of identities (if the ktup variable is 1) or pairs of identities (if ktup=2), without considering conservative amino acid substitutions, insertions, or deletions. The ten regions with the highest density of identities are then rescored by comparing the similarity of all paired amino acids using an amino acid substitution matrix, and the ends of the regions are "trimmed" to include only those residues that contribute to the highest score. If there are several regions with scores greater than the "cutoff" value (calculated by a predetermined formula based upon the length of the sequence and the ktup value), then the trimmed initial regions are examined to determine whether the regions can be joined to form an approximate alignment with gaps. Finally, the highest scoring regions of the two amino acid sequences are aligned using a modification of the Needleman-Wunsch-Sellers algorithm (Needleman and Wunsch, *J. Mol. Biol. 48*:444 (1970); Sellers, *SIAM J. Appl. Math. 26*:787 (1974)), which allows for amino acid insertions and deletions. Preferred parameters for FASTA analysis are: ktup=1, gap opening penalty=10, gap extension penalty=1, and substitution matrix=BLOSUM62, with other parameters set as default. These parameters can be introduced into a FASTA program by modifying the scoring matrix file ("SMATRIX"), as explained in Appendix 2 of Pearson, *Meth. Enzymol. 183*:63 (1990).

FASTA can also be used to determine the sequence identity of nucleic acid molecules using a ratio as disclosed above. For nucleotide sequence comparisons,

the ktup value can range between one to six, preferably from three to six, most preferably three, with other FASTA program parameters set as default.

The BLOSUM62 table (Table 4) is an amino acid substitution matrix derived from about 2,000 local multiple alignments of protein sequence segments, representing highly conserved regions of more than 500 groups of related proteins (Henikoff and Henikoff, *Proc. Nat'l Acad. Sci. USA 89*:10915 (1992)). Accordingly, the BLOSUM62 substitution frequencies can be used to define conservative amino acid substitutions that may be introduced into the amino acid sequences of the present invention. Although it is possible to design amino acid substitutions based solely upon chemical properties (as discussed below), the language "conservative amino acid substitution" preferably refers to a substitution represented by a BLOSUM62 value of greater than -1. For example, an amino acid substitution is conservative if the substitution is characterized by a BLOSUM62 value of 0, 1, 2, or 3. According to this system, preferred conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 1 (e.*g.*, 1, 2 or 3), while more preferred conservative amino acid substitutions are characterized by a BLOSUM62 value of at least 2 (e.*g.*, 2 or 3).

Variant zcytor17 polypeptides or substantially homologous zcytor17 polypeptides are characterized as having one or more amino acid substitutions, deletions or additions. These changes are preferably of a minor nature, that is conservative amino acid substitutions (see Table 5) and other substitutions that do not significantly affect the folding or activity of the polypeptide; small deletions, typically of one to about 30 amino acids; and small amino- or carboxyl-terminal extensions, such as an amino-terminal methionine residue, a small linker peptide of up to about 20-25 residues, or an affinity tag. The present invention thus includes polypeptides that comprise a sequence that is at least 80%, preferably at least 90%, and more preferably 95% or more identical to the corresponding region of SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:5, SEQ ID NO:117 or SEQ ID NO:119 excluding the tags, extension, linker sequences and the like. Polypeptides comprising affinity tags can further comprise a proteolytic cleavage site between the zcytor17 polypeptide and the affinity tag. Suitable sites include thrombin cleavage sites and factor Xa cleavage sites.

Table 5

Conservative amino acid substitutions

Basic:	arginine
	lysine
	histidine
Acidic:	glutamic acid
	aspartic acid
Polar:	glutamine
	asparagine
Hydrophobic:	leucine
	isoleucine
	valine
Aromatic:	phenylalanine
	tryptophan
	tyrosine
Small:	glycine
	alanine
	serine
	threonine
	methionine

The present invention provides an isolated an isolated multimeric cytokine receptor comprising a polypeptide having at least 90 percent sequence identity with SEQ ID NO:111, SEQ ID NO:109, or SEQ ID NO:5; and at least a portion of at least one class I cytokine receptor, wherein the multimeric cytokine receptor binds to at least a portion of SEQ ID NO:2. The polypeptide having at least 90 percent sequence identity with SEQ ID NO:111 may include, for instance, amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 227 of SEQ ID NO:519, amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 544 to amino acid residue 732 of SEQ ID NO:111, amino acid residue 544 to amino acid residue 649 of SEQ ID NO:109, amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 649 of SEQ ID NO:109, and combinations thereof. The at least a portion of at least one class I cytokine receptor includes, for example, OSMRbeta (SEQ ID NO:7) and/or WSX-1 (SEQ ID NO:9). For example, the at least a portion of at least one class I cytokine receptor comprising at least a portion of SEQ ID NO:7 may comprise amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 35 to amino acid residue 137 of SEQ ID NO:7, amino acid residue 240 to amino acid residue 342 of SEQ ID NO:7, amino acid residue 348 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 762 to amino acid residue 979 of SEQ ID NO:7, and/or combinations thereof. The multimeric cytokine receptor may be a heterodimer, trimer, tetramer, pentamer, or the like. In addition, the multimeric cytokine receptor may be soluble, immobilized on a solid support, or membrane-bound. Optionally, the multimeric cytokine receptor may antagonize an activity of SEQ ID NO:2, such as inhibit or reduce the proliferation of hematopoietic, immune, and/or inflammatory cells, or inhibit or reduce the enhancement of the hematopoietic, immune, and/or inflammatory process, or inhibit or reduce the differentiation of hematopoietic cells, for instance lymphoid cells such as monocytic cells, macrophages and/or T cells. The multimeric cytokine receptor of the present invention may also comprise an affinity tag selected from the group of polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™

peptide, streptavidin binding peptide, and an immunoglobulin F_c polypeptide.

The present invention also provides for a ligand/receptor complex comprising a polypeptide which includes at least a portion of SEQ ID NO:2; and a soluble multimeric cytokine receptor comprising at least a portion of at least one polypeptide selected from the group of SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:7, and SEQ ID NO:9, wherein the polypeptide is attached to the soluble multimeric cytokine receptor. The soluble multimeric cytokine receptor may comprise the extracellular domain and/or transmembrane domain of zcytor17 (SEQ ID NO:111), OSMRbeta (SEQ ID NO:7), and/or WSX-1 (SEQ ID NO:9). For example, the soluble multimeric cytokine receptor may comprise amino acid residue 20 to 227 to SEQ ID NO:111, amino acid residue 20 to 519 to SEQ ID NO:111, amino acid residue 20 to 543 to SEQ ID NO:111, amino acid residue 28 to 739 to SEQ ID NO:7, amino acid residue 28 to 429 to SEQ ID NO:7, amino acid residue 35 to 137 to SEQ ID NO:7, amino acid residue 240 to 342 to SEQ ID NO:7, amino acid residue 348 to 429 to SEQ ID NO:7, or combinations thereof. The soluble multimeric cytokine receptor may be a heterodimer, trimer, tetramer, pentamer, or the like. The multimeric cytokine receptor of the present invention may also comprise an affinity tag as described herein. The polypeptide of the ligand/receptor complex may comprise amino acid residues of SEQ ID NO:2 selected from the group of 38 to 152, 27 to 164, 24 to 164, 1 to 164, 38 to 52, 83 to 98, 104 to 117, 137 to 152, and combinations thereof.

The ligand/receptor complex of the present may also comprise a fusion protein. The fusion protein may comprise at least four polypeptides, wherein the order of polypeptides from N-terminus to C-terminus are a first polypeptide comprising amino acid residues 38-52 of SEQ ID NO:2; a first spacer of 6-27 amino acid residues; a second polypeptide comprising amino acid residues selected from the group of (a) IL-2 helix B residues of SEQ ID NO:183; (b) IL-4 helix B residues 65-83 of SEQ ID NO:179; (c) IL-3 helix B residues 73-86 of SEQ ID NO:102; (d) GM-CSF helix B residues 72-81 of SEQ ID NO:181; and (e) amino acid residues 83-98 of SEQ ID NO:2; a second spacer of 5-11 amino acid residues; a third polypeptide comprising amino acid residues selected from the group of (a) IL-2 helix C residues 102-116 of SEQ ID NO:177; (b) IL-4 helix C residues 94-118 of SEQ ID NO:179; (c) IL-3 helix C residues

91-103 of SEQ ID NO:102; (d) GM-CSF helix C residues 85-103 of SEQ ID NO:181; and (e) amino acid residues 104-117 of SEQ ID NO:2; a third spacer of 3-29 amino acid residues; and a fourth polypeptide comprising amino acid residues selected from the group of (a) IL-2 helix D residues 134-149 of SEQ ID NO:177 ; (b) IL-3 helix D residues 123-141 of SEQ ID NO:102; (c) IL-4 helix D residues 133-151 of SEQ ID NO:179; (d) GM-CSF helix D residues 120-131 of SEQ ID NO:181; and (e) amino acid residues 137-152 of SEQ ID NO:2; and a multimeric cytokine receptor comprising at least a portion of at least one polypeptide selected from the group of SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:7, and SEQ ID NO:9; wherein the fusion protein is attached to the multimeric cytokine receptor.

Alternatively, the fusion protein may comprise at least four polypeptides, wherein the order of polypeptides from N-terminus to C-terminus are a first polypeptide comprising amino acid residues selected from a group of (a) IL-2 helix A residues 27-48 of SEQ ID NO:177; (b) IL-4 helix A residues 30-42 of SEQ ID NO:179; (c) IL-3 helix A residues 35-45 of SEQ ID NO:102; (d) GM-CSF helix A residues 30-44 of SEQ ID NO:181; and (e) amino acids residues 38-52 of SEQ ID NO:2; a first spacer of 6-27 amino acid residues; a second polypeptide comprising amino acid residues selected from the group of (a) IL-2 helix B residues of SEQ ID NO:183; (b); IL-4 helix B residues 65-83 of SEQ ID NO:179; (c) IL-3 helix B residues 73-86 of SEQ ID NO:102; (d) GM-CSF helix B residues 72-81 of SEQ ID NO:181; and (e) amino acid residues 83-98 of SEQ ID NO:2; a second spacer of 5-11 amino acid residues; a third polypeptide comprising amino acid residues selected from the group of (a) IL-2 helix C residues 102-116 of SEQ ID NO:177; (b) IL-4 helix C residues 94-118 of SEQ ID NO:179; (c) IL-3 helix C residues 91-103 of SEQ ID NO:102; (d) GM-CSF helix C residues 85-103 of SEQ ID NO:181; and (e) amino acid residues 104-117 of SEQ ID NO:2; a third spacer of 3-29 amino acid residues; and a fourth polypeptide comprising amino acid residues from 137-152 of SEQ ID NO:2; and a multimeric cytokine receptor comprising at least a portion of at least one polypeptide selected from the group of SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:7, and SEQ ID NO:9; wherein the fusion protein is attached to the multimeric cytokine receptor. A multimeric cytokine receptor may comprise at least one of the following polypeptides of SEQ ID NO:111, SEQ ID

NO:109, SEQ ID NO:7, SEQ ID NO:9, or the extracellular domains thereof. The ligand/receptor complex can be soluble and may additionally include an affinity tag as described herein.

The present invention also provides an isolated and purified polynucleotide that encodes a polypeptide comprising an amino acid sequence having at least 90 percent sequence identity with SEQ ID NO:111, SEQ ID NO:109, or SEQ ID NO:5, wherein the polypeptide and at least a portion of at least one class I cytokine receptor form a multimeric cytokine receptor, and wherein the multimeric cytokine receptor binds to at least a portion of SEQ ID NO:2. The present invention also provides an isolated and purified polynucleotide that encodes a polypeptide comprising at least a portion of at least one of SEQ ID NO:111, SEQ ID NO:109, or SEQ ID NO:5, wherein the polypeptide and at least a portion of a class I cytokine receptor form a multimeric cytokine receptor, and wherein the multimeric cytokine receptor binds to at least a portion of SEQ ID NO:2. The polynucleotide may encode a polypeptide that is included in a soluble multimeric cytokine receptor and that may also include an affinity tag as described herein. The polypeptide may comprise, for example, amino acid residue 20 to 227 to SEQ ID NO:111, amino acid residue 20 to 519 to SEQ ID NO:111, amino acid residue 20 to 543 to SEQ ID NO:111, and/or combinations thereof. In addition, the at least a portion of at least one class I cytokine receptor may comprise, for example, amino acid residue 28 to 739 to SEQ ID NO:7, amino acid residue 28 to 429 to SEQ ID NO:7, amino acid residue 35 to 137 to SEQ ID NO:7, amino acid residue 240 to 342 to SEQ ID NO:7, amino acid residue 348 to 429 to SEQ ID NO:7, and/or combinations thereof. The soluble multimeric cytokine receptor may be a heterodimer, trimer, tetramer, pentamer, or the like. The at least a portion of SEQ ID NO:2 may include, for instance amino acid residues of SEQ ID NO:2 selected from the group of 38 to 152, 27 to 164, 24 to 164, 1 to 164, 38 to 52, 83 to 98, 104 to 117, 137 to 152, and combinations thereof. Optionally, the multimeric cytokine receptor may antagonize an activity of SEQ ID NO:2 as described herein.

The present invention further provides a variety of other polypeptide fusions and related multimeric proteins comprising one or more polypeptide fusions. For example, a zcytor17 polypeptide can be prepared as a fusion to a dimerizing protein

as disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Preferred dimerizing proteins in this regard include immunoglobulin constant region domains. Immunoglobulin-zcytor17 polypeptide fusions can be expressed in genetically engineered cells to produce a variety of multimeric zcytor17 analogs. Auxiliary domains can be fused to zcytor17 polypeptides to target them to specific cells, tissues, or macromolecules (e.g., collagen). A zcytor17 polypeptide can be fused to two or more moieties, such as an affinity tag for purification and a targeting domain. Polypeptide fusions can also comprise one or more cleavage sites, particularly between domains. See, Tuan et al., *Connective Tissue Research 34*:1-9, 1996. For example, one or more domains from zcytor17 soluble receptor can be joined to a soluble cytokine receptor, such as OSMRbeta and/or WSX-1, which may enhance their biological properties or efficiency of production. Additionally, the soluble multimeric cytokine receptor may further include an affinity tag. An affinity tag can be, for example, a tag selected from the group of polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™ peptide, streptavidin binding peptide, and an immunoglobulin F_c polypeptide.

The proteins of the present invention can also comprise non-naturally occurring amino acid residues. Non-naturally occurring amino acids include, without limitation, *trans*-3-methylproline, 2,4-methanoproline, *cis*-4-hydroxyproline, *trans*-4-hydroxyproline, *N*-methylglycine, *allo*-threonine, methylthreonine, hydroxyethylcysteine, hydroxyethylhomocysteine, nitroglutamine, homoglutamine, pipecolic acid, thiazolidine carboxylic acid, dehydroproline, 3- and 4-methylproline, 3,3-dimethylproline, *tert*-leucine, norvaline, 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, and 4-fluorophenylalanine. Several methods are known in the art for incorporating non-naturally occurring amino acid residues into proteins. For example, an *in vitro* system can be employed wherein nonsense mutations are suppressed using chemically aminoacylated suppressor tRNAs. Methods for synthesizing amino acids and aminoacylating tRNA are known in the art. Transcription and translation of plasmids containing nonsense mutations is carried out in a cell-free system comprising an *E. coli* S30 extract and commercially available enzymes and other reagents. Proteins are purified by chromatography. See, for example, Robertson et al., J. *Am. Chem. Soc.*

113:2722, 1991; Ellman et al., *Methods Enzymol. 202*:301, 1991; Chung et al., *Science 259*:806-9, 1993; and Chung et al., *Proc. Natl. Acad. Sci. USA 90*:10145-9, 1993). In a second method, translation is carried out in *Xenopus* oocytes by microinjection of mutated mRNA and chemically aminoacylated suppressor tRNAs (Turcatti et al., *J. Biol. Chem. 271*:19991-8, 1996). Within a third method, *E. coli* cells are cultured in the absence of a natural amino acid that is to be replaced (e.g., phenylalanine) and in the presence of the desired non-naturally occurring amino acid(s) (e.g., 2-azaphenylalanine, 3-azaphenylalanine, 4-azaphenylalanine, or 4-fluorophenylalanine). The non-naturally occurring amino acid is incorporated into the protein in place of its natural counterpart. See, Koide et al., *Biochem. 33*:7470-7476, 1994. Naturally occurring amino acid residues can be converted to non-naturally occurring species by *in vitro* chemical modification. Chemical modification can be combined with site-directed mutagenesis to further expand the range of substitutions (Wynn and Richards, *Protein Sci. 2*:395-403, 1993).

A limited number of non-conservative amino acids, amino acids that are not encoded by the genetic code, non-naturally occurring amino acids, and unnatural amino acids may be substituted for zcytor17 amino acid residues.

Essential amino acids in the polypeptides of the present invention can be identified according to procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells, *Science 244*: 1081-5, 1989; Bass et al., *Proc. Natl. Acad. Sci. USA 88*:4498-502, 1991). In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity (e.g. ligand binding and signal transduction) as disclosed below to identify amino acid residues that are critical to the activity of the molecule. See also, Hilton et al., *J. Biol. Chem. 271*:4699-4708, 1996. Sites of ligand-receptor, protein-protein or other biological interaction can also be determined by physical analysis of structure, as determined by such techniques as nuclear magnetic resonance, crystallography, electron diffraction or photoaffinity labeling, in conjunction with mutation of putative contact site amino acids. See, for example, de Vos et al., Science 255:306-312, 1992; Smith et al., *J. Mol. Biol. 224*:899-904, 1992; Wlodaver et al., *FEBS Lett. 309*:59-64, 1992. The identities of essential

amino acids can also be inferred from analysis of homologies with related receptors.

Determination of amino acid residues that are within regions or domains that are critical to maintaining structural integrity can be determined. Within these regions one can determine specific residues that will be more or less tolerant of change and maintain the overall tertiary structure of the molecule. Methods for analyzing sequence structure include, but are not limited to, alignment of multiple sequences with high amino acid or nucleotide identity and computer analysis using available software (e.g., the Insight II® viewer and homology modeling tools; MSI, San Diego, CA), secondary structure propensities, binary patterns, complementary packing and buried polar interactions (Barton, *Current Opin. Struct. Biol. 5*:372-376, 1995 and Cordes et al., *Current Opin. Struct. Biol. 6*:3-10, 1996). In general, when designing modifications to molecules or identifying specific fragments determination of structure will be accompanied by evaluating activity of modified molecules.

Amino acid sequence changes are made in zcytor17 polypeptides so as to minimize disruption of higher order structure essential to biological activity. For example, when the zcytor17 polypeptide comprises one or more helices, changes in amino acid residues will be made so as not to disrupt the helix geometry and other components of the molecule where changes in conformation abate some critical function, for example, binding of the molecule to its binding partners. The effects of amino acid sequence changes can be predicted by, for example, computer modeling as disclosed above or determined by analysis of crystal structure (see, e.g., Lapthorn et al., *Nat. Struct. Biol. 2*:266-268, 1995). Other techniques that are well known in the art compare folding of a variant protein to a standard molecule (e.g., the native protein). For example, comparison of the cysteine pattern in a variant and standard molecules can be made. Mass spectrometry and chemical modification using reduction and alkylation provide methods for determining cysteine residues which are associated with disulfide bonds or are free of such associations (Bean et al., *Anal. Biochem. 201*:216-226, 1992; Gray, *Protein Sci. 2*:1732-1748, 1993; and Patterson et al., *Anal. Chem. 66*:3727-3732, 1994). It is generally believed that if a modified molecule does not have the same disulfide bonding pattern as the standard molecule folding would be affected. Another well known and accepted method for measuring folding is circular dichrosism

(CD). Measuring and comparing the CD spectra generated by a modified molecule and standard molecule is routine (Johnson, *Proteins 7*:205-214, 1990). Crystallography is another well known method for analyzing folding and structure. Nuclear magnetic resonance (NMR), digestive peptide mapping and epitope mapping are also known methods for analyzing folding and structural similarities between proteins and polypeptides (Schaanan et al., *Science 257*:961-964, 1992).

A Hopp/Woods hydrophilicity profile of the zcytor17 protein sequence as shown in SEQ ID NO:2, SEQ ID NO:46, SEQ ID NO:54, SEQ ID NO:57 and SEQ ID NO:93 can be generated (Hopp et al., *Proc. Natl. Acad. Sci. 78*:3824-3828, 1981; Hopp, *J. Immun. Meth. 88*:1-18, 1986 and Triquier et al., *Protein Engineering 11*:153-169, 1998). See, Figure 1. The profile is based on a sliding six-residue window. Buried G, S, and T residues and exposed H, Y, and W residues were ignored. For example, in zcytor17, hydrophilic regions include amino acid residues 43 through 48 of SEQ ID NO:2 and SEQ ID NO:46 (residues 56 through 61 of SEQ ID NO:54), amino acid residues 157 through 162 of SEQ ID NO:2 and SEQ ID NO:46 (residues 170 through 175 of SEQ ID NO:54), amino acid residues 158 through 163 of SEQ ID NO:2 and SEQ ID NO:46 (residues 171 through 176 of SEQ ID NO:54), amino acid residues 221 through 226 of SEQ ID NO:2 and SEQ ID NO:46 (residues 234 through 239 of SEQ ID NO:54), and amino acid residues 426 through 431 of SEQ ID NO:2 and SEQ ID NO:46 (residues 439 through 444 of SEQ ID NO:54).

Those skilled in the art will recognize that hydrophilicity or hydrophobicity will be taken into account when designing modifications in the amino acid sequence of a zcytor17 polypeptide, so as not to disrupt the overall structural and biological profile. Of particular interest for replacement are hydrophobic residues selected from the group consisting of Val, Leu and Ile or the group consisting of Met, Gly, Ser, Ala, Tyr and Trp. For example, residues tolerant of substitution could include such residues as shown in SEQ ID NO:2, SEQ ID NO:46, SEQ ID NO:54, SEQ ID NO:57 and SEQ ID NO:93. However, Cysteine residues would be relatively intolerant of substitution.

The identities of essential amino acids can also be inferred from analysis of sequence similarity between class I cytokine receptor family members with zcytor17.

Using methods such as "FASTA" analysis described previously, regions of high similarity are identified within a family of proteins and used to analyze amino acid sequence for conserved regions. An alternative approach to identifying a variant zcytor17 polynucleotide on the basis of structure is to determine whether a nucleic acid molecule encoding a potential variant zcytor17 polynucleotide can hybridize to a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:45 or SEQ ID NO:53, as discussed above.

Other methods of identifying essential amino acids in the polypeptides of the present invention are procedures known in the art, such as site-directed mutagenesis or alanine-scanning mutagenesis (Cunningham and Wells, *Science 244*:1081 (1989), Bass et al., *Proc. Natl Acad. Sci. USA 88*:4498 (1991), Coombs and Corey, "Site-Directed Mutagenesis and Protein Engineering," in *Proteins: Analysis and Design*, Angeletti (ed.), pages 259-311 (Academic Press, Inc. 1998)). In the latter technique, single alanine mutations are introduced at every residue in the molecule, and the resultant mutant molecules are tested for biological activity as disclosed below to identify amino acid residues that are critical to the activity of the molecule. See also, Hilton *et al., J. Biol. Chem. 271*:4699 (1996).

The present invention also includes a multimeric cytokine receptor which includes functional fragments of zcytor17 polypeptides and nucleic acid molecules encoding such functional fragments. A "functional" zcytor17 or fragment thereof defined herein is characterized by its ability to mediate proliferative or differentiating activity, by its ability to induce or inhibit specialized cell functions, or by its ability to bind specifically to an anti-zcytor17 antibody or zcytor17 ligand (either soluble or immobilized). Moreover, functional fragments also include the signal peptide, intracellular signaling domain, and the like. As previously described herein, zcytor17 is characterized by a class I cytokine receptor structure. Thus, the present invention further provides fusion proteins encompassing: (a) polypeptide molecules comprising an extracellular domain, cytokine-binding domain, or intracellular domain described herein; and (b) functional fragments comprising one or more of these domains. The other polypeptide portion of the fusion protein may comprise at least a portion of one or more of another class I cytokine receptor, for example, gp130, LIF,

IL-12, WSX-1, IL-2 receptor β-subunit and the β-common receptor (i.e., IL3, IL-5, and GM-CSF receptor β-subunits), or by a non-native and/or an unrelated secretory signal peptide that facilitates secretion of the fusion protein.

Routine deletion analyses of nucleic acid molecules can be performed to obtain functional fragments of a nucleic acid molecule that encodes a zcytor17 polypeptide. As an illustration, DNA molecules having the nucleotide sequence of SEQ ID NO:1, SEQ ID NO:45 or SEQ ID NO:53 or fragments thereof, can be digested with *Bal*31 nuclease to obtain a series of nested deletions. These DNA fragments are then inserted into expression vectors in proper reading frame, and the expressed polypeptides are isolated and tested for zcytor17 activity, or for the ability to bind anti-zcytor17 antibodies or zcytor17 ligand. One alternative to exonuclease digestion is to use oligonucleotide-directed mutagenesis to introduce deletions or stop codons to specify production of a desired zcytor17 fragment. Alternatively, particular fragments of a zcytor17 polynucleotide can be synthesized using the polymerase chain reaction.

Standard methods for identifying functional domains are well-known to those of skill in the art. For example, studies on the truncation at either or both termini of interferons have been summarized by Horisberger and Di Marco, *Pharmac. Ther.* *66*:507 (1995). Moreover, standard techniques for functional analysis of proteins are described by, for example, Treuter et al., *Molec. Gen. Genet. 240*:113 (1993); Content et al., "Expression and preliminary deletion analysis of the 42 kDa 2-5A synthetase induced by human interferon," in *Biological Interferon Systems, Proceedings of ISIR-TNO Meeting on Interferon Systems*, Cantell (ed.), pages 65-72 (Nijhoff 1987); Herschman, "The EGF Receptor," in *Control of Animal Cell Proliferation 1*, Boynton et al., (eds.) pages 169-199 (Academic Press 1985); Coumailleau et al., *J. Biol. Chem.* *270*:29270 (1995); Fukunaga et al., *J. Biol. Chem. 270*:25291 (1995); Yamaguchi et al., *Biochem. Pharmacol. 50*:1295 (1995); and Meisel et al., *Plant Molec. Biol. 30*:1 (1996).

Multiple amino acid substitutions can be made and tested using known methods of mutagenesis and screening, such as those disclosed by Reidhaar-Olson and Sauer (*Science 241*:53-57, 1988) or Bowie and Sauer (*Proc. Natl. Acad. Sci. USA* *86*:2152-2156, 1989). Briefly, these authors disclose methods for simultaneously

randomizing two or more positions in a polypeptide, selecting for functional polypeptide, and then sequencing the mutagenized polypeptides to determine the spectrum of allowable substitutions at each position. Other methods that can be used include phage display (e.g., Lowman et al., *Biochem. 30*:10832-10837, 1991; Ladner et al., U.S. Patent No. 5,223,409; Huse, WIPO Publication WO 92/062045) and region-directed mutagenesis (Derbyshire et al., *Gene 46*:145, 1986; Ner et al., *DNA 7*:127, 1988).

Variants of the disclosed zcytor17 DNA and polypeptide sequences can be generated through DNA shuffling as disclosed by Stemmer, *Nature 370*:389-91, 1994, Stemmer, *Proc. Natl. Acad. Sci. USA 91*:10747-51, 1994 and WIPO Publication WO 97/20078. Briefly, variant DNAs are generated by *in vitro* homologous recombination by random fragmentation of a parent DNA followed by reassembly using PCR, resulting in randomly introduced point mutations. This technique can be modified by using a family of parent DNAs, such as allelic variants or DNAs from different species, to introduce additional variability into the process. Selection or screening for the desired activity, followed by additional iterations of mutagenesis and assay provides for rapid "evolution" of sequences by selecting for desirable mutations while simultaneously selecting against detrimental changes.

Mutagenesis methods as disclosed herein can be combined with high-throughput, automated screening methods to detect activity of cloned, mutagenized zcytor17 receptor polypeptides in host cells. Preferred assays in this regard include cell proliferation assays and biosensor-based ligand-binding assays, which are described below. Mutagenized DNA molecules that encode active receptors or portions thereof (e.g., ligand-binding fragments, signaling domains, and the like) can be recovered from the host cells and rapidly sequenced using modern equipment. These methods allow the rapid determination of the importance of individual amino acid residues in a polypeptide of interest, and can be applied to polypeptides of unknown structure.

The present invention also provides a novel multimeric cytokine receptor in which a segment comprising at least a portion of one or more of the domains of zcytor17, for instance, secretory, extracellular, transmembrane, and intracellular, is fused to another polypeptide, for example, an extracellular domain of a class I cytokine

receptor, such as OSMRbeta and/or WSX-1. Fusion is preferably done by splicing at the DNA level to allow expression of chimeric molecules in recombinant production systems. The resultant molecules are then assayed for such properties as improved solubility, improved stability, prolonged clearance half-life, improved expression and secretion levels, and pharmacodynamics. Such a multimeric cytokine receptor may further comprise additional amino acid residues (e.g., a polypeptide linker) between the component proteins or polypeptides. A domain linker may comprise a sequence of amino acids from about 3 to about 20 amino acids long, from about 5 to 15 about amino acids long, from about 8 to about 12 amino acids long, and about 10 amino acids long. One function of a linker is to separate the active protein regions to promote their independent bioactivity and permit each region to assume its bioactive conformation independent of interference from its neighboring structure.

`Using the methods discussed herein, one of ordinary skill in the art can identify and/or prepare a variety of polypeptide fragments or variants of SEQ ID NO:111, SEQ ID NO:109, SEQ ID NO:5, SEQ ID NO:117 and SEQ ID NO:119 that retain the signal transduction or ligand binding activity. For example, one can make a zcytor17 "soluble receptor" by preparing a variety of polypeptides that are substantially homologous to the cytokine-binding domain (residues 20 (Ala) to 227 (Pro) of SEQ ID NO:111 and SEQ ID NO:109; residues 33 (Ala) to 240 (Pro) of SEQ ID NO:5), the extracellular domain (residues 20 (Ala) to 519 (Glu) of SEQ ID NO:111 and SEQ ID NO:109; residues 33 (Ala) to 532 (Glu) of SEQ ID NO:5), or allelic variants or species orthologs thereof (e.g., see SEQ ID NO:117 and SEQ ID NO:119 and functional fragments thereof as described herein)) and retain ligand-binding activity of the wild-type zcytor17 protein. Moreover, variant zcytor17 soluble receptors such as those shown in SEQ ID NO:113 and SEQ ID NO:115 can be isolated. Such polypeptides may include additional amino acids from, for example, part or all of the transmembrane and intracellular domains. Such polypeptides may also include additional polypeptide segments as generally disclosed herein such as labels, affinity tags, and the like.

For any zcytor17 polypeptide, including variants, soluble receptors, and fusion polypeptides or proteins, one of ordinary skill in the art can readily generate a fully degenerate polynucleotide sequence encoding that variant using the information

set forth in Tables 1 and 2 above.

The zcytor17 multimeric cytokine receptors of the present invention, including full-length polypeptides, biologically active fragments, and fusion polypeptides, can be produced in genetically engineered host cells according to conventional techniques. Suitable host cells are those cell types that can be transformed or transfected with exogenous DNA and grown in culture, and include bacteria, fungal cells, and cultured higher eukaryotic cells. Eukaryotic cells, particularly cultured cells of multicellular organisms, are preferred. Techniques for manipulating cloned DNA molecules and introducing exogenous DNA into a variety of host cells are disclosed by Sambrook et al., Molecular Cloning: *A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989, and Ausubel et al., eds., *Current Protocols in Molecular Biology*, John Wiley and Sons, Inc., NY, 1987.

The present invention also provides an expression vector comprising an isolated and purified DNA molecule including the following operably linked elements: a first transcription promoter, a first DNA segment encoding a polypeptide having at least 90 percent sequence identity with SEQ ID NO:111, and a first transcription terminator; and a second transcription promoter, a second DNA segment encoding at least a portion of a class I cytokine receptor, and a second transcription terminator; wherein the polypeptide and the class I cytokine receptor form a multimeric cytokine receptor; and wherein the multimeric cytokine receptor binds to at least a portion of SEQ ID NO:2. The DNA molecule may further comprise a secretory signal sequence operably linked to the first and second DNA segments. The multimeric cytokine receptor may be soluble and/or may further comprise an affinity tag as described herein. In addition, the multimeric cytokine receptor may antagonize an activity of SEQ ID NO:2 as described herein. The at least at portion of a class I cytokine receptor may comprise portions of SEQ ID NO:7 and/or SEQ ID NO:9, such as, for instance, amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 35 to amino acid residue 137 of SEQ ID NO:7, amino acid residue 240 to amino acid residue 342 of SEQ ID NO:7, amino acid residue 348 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 762 to

amino acid residue 979 of SEQ ID NO:7, or combinations thereof. The present invention also provides a cultured cell containing the above-described expression vector.

In general, a DNA sequence, for example, encoding a zcytor17 polypeptide is operably linked to other genetic elements required for its expression, generally including a transcription promoter and terminator, within an expression vector. The vector will also commonly contain one or more selectable markers and one or more origins of replication, although those skilled in the art will recognize that within certain systems selectable markers may be provided on separate vectors, and replication of the exogenous DNA may be provided by integration into the host cell genome. Selection of promoters, terminators, selectable markers, vectors and other elements is a matter of routine design within the level of ordinary skill in the art. Many such elements are described in the literature and are available through commercial suppliers.

To direct, for example, a zcytor17 polypeptide into the secretory pathway of a host cell, a secretory signal sequence (also known as a leader sequence, prepro sequence or pre sequence) is provided in the expression vector. The secretory signal sequence may be that of zcytor17, or may be derived from another secreted protein (e.g., t-PA) or synthesized *de novo*. The secretory signal sequence is operably linked to the zcytor17 DNA sequence, i.e., the two sequences are joined in the correct reading frame and positioned to direct the newly synthesized polypeptide into the secretory pathway of the host cell. Secretory signal sequences are commonly positioned 5' to the DNA sequence encoding the polypeptide of interest, although certain secretory signal sequences may be positioned elsewhere in the DNA sequence of interest (see, e.g., Welch et al., U.S. Patent No. 5,037,743; Holland et al., U.S. Patent No. 5,143,830).

Alternatively, the secretory signal sequence contained in the polypeptides of the present invention is used to direct other polypeptides into the secretory pathway. The present invention provides for such fusion polypeptides. A signal fusion polypeptide can be made wherein a secretory signal sequence derived from amino acid 1 (Met) to amino acid 19 (Ala) of SEQ ID NO:111 and SEQ ID

NO:109, or wherein a secretory signal sequence derived from amino acid 1 (Met) to amino acid 32 (Ala) of SEQ ID NO:5, or amino acid 1 (Met) to amino acid 45 (Ala) of SEQ ID NO:117 or SEQ ID NO:119), or amino acid 28 (Met) to residue 45 (Ala) of SEQ ID NO:117 or SEQ ID NO:119), is operably linked to another polypeptide using methods known in the art and disclosed herein. The secretory signal sequence contained in the fusion polypeptides of the present invention is preferably fused amino-terminally to an additional peptide to direct the additional peptide into the secretory pathway. Such constructs have numerous applications known in the art. For example, these novel secretory signal sequence fusion constructs can direct the secretion of an active component of a normally non-secreted protein. Such fusions may be used *in vivo* or *in vitro* to direct peptides through the secretory pathway.

The present invention also provides a cultured cell comprising a first expression vector comprising a DNA molecule containing the following operably linked elements: a transcription promoter, a DNA segment encoding a polypeptide having at least 90 percent sequence identity with SEQ ID NO:111, and a transcription terminator; and a second expression vector comprising a transcription promoter, a DNA segment encoding at least a portion of a class I cytokine receptor, and a transcription terminator; wherein the polypeptide and the class I cytokine receptor form a multimeric cytokine receptor. The first and second expression vectors may further comprise a secretory signal sequence operably linked to the first and second DNA segments. The multimeric cytokine receptor may be soluble, may be a heterodimer, and/or may further comprise an affinity tag as described herein. In addition, the multimeric cytokine receptor may antagonize an activity of SEQ ID NO:2 as described herein. The at least at portion of a class I cytokine receptor may comprise portions of SEQ ID NO:7 and/or SEQ ID NO:9, such as, for instance, amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 35 to amino acid residue 137 of SEQ ID NO:7, amino acid residue 240 to amino acid residue 342 of SEQ ID NO:7, amino acid residue 348 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 762 to amino acid residue 979 of SEQ ID NO:7, or combinations thereof.

Cultured mammalian cells are suitable hosts within the present invention. Methods for introducing exogenous DNA into mammalian host cells include calcium phosphate-mediated transfection (Wigler et al., *Cell 14*:725, 1978; Corsaro and Pearson, *Somatic Cell Genetics 7*:603, 1981: Graham and Van der Eb, *Virology 52*:456, 1973), electroporation (Neumann et al., *EMBO J. 1*:841-845, 1982), DEAE-dextran mediated transfection (Ausubel et al., *ibid.*), and liposome-mediated transfection (Hawley-Nelson et al., Focus 15:73, 1993; Ciccarone et al., *Focus 15*:80, 1993, and viral vectors (Miller and Rosman, *BioTechniques 7*:980-90, 1989; Wang and Finer, *Nature Med. 2*:714-716, 1996). The production of recombinant polypeptides in cultured mammalian cells is disclosed, for example, by Levinson et al., U.S. Patent No. 4,713,339; Hagen et al., U.S. Patent No. 4,784,950; Palmiter et al., U.S. Patent No. 4,579,821; and Ringold, U.S. Patent No. 4,656,134. Suitable cultured mammalian cells include the COS-1 (ATCC No. CRL 1650), COS-7 (ATCC No. CRL 1651), BHK (ATCC No. CRL 1632), BHK 570 (ATCC No. CRL 10314), 293 (ATCC No. CRL 1573; Graham et al., *J. Gen. Virol. 36*:59-72, 1977) and Chinese hamster ovary (e.g. CHO-K1; ATCC No. CCL 61) cell lines. Additional suitable cell lines are known in the art and available from public depositories such as the American Type Culture Collection, Rockville, Maryland. In general, strong transcription promoters are preferred, such as promoters from SV-40 or cytomegalovirus. See, e.g., U.S. Patent No. 4,956,288. Other suitable promoters include those from metallothionein genes (U.S. Patent Nos. 4,579,821 and 4,601,978) and the adenovirus major late promoter.

Drug selection is generally used to select for cultured mammalian cells into which foreign DNA has been inserted. Such cells are commonly referred to as "transfectants". Cells that have been cultured in the presence of the selective agent and are able to pass the gene of interest to their progeny are referred to as "stable transfectants." A preferred selectable marker is a gene encoding resistance to the antibiotic neomycin. Selection is carried out in the presence of a neomycin-type drug, such as G-418 or the like. Selection systems can also be used to increase the expression level of the gene of interest, a process referred to as "amplification." Amplification is carried out by culturing transfectants in the presence of a low level of the selective agent and then increasing the amount of selective agent to select for cells that produce

high levels of the products of the introduced genes. A preferred amplifiable selectable marker is dihydrofolate reductase, which confers resistance to methotrexate. Other drug resistance genes (e.g., hygromycin resistance, multi-drug resistance, puromycin acetyltransferase) can also be used. Alternative markers that introduce an altered phenotype, such as green fluorescent protein, or cell surface proteins such as CD4, CD8, Class I MHC, placental alkaline phosphatase may be used to sort transfected cells from untransfected cells by such means as FACS sorting or magnetic bead separation technology.

Other higher eukaryotic cells can also be used as hosts, including plant cells, insect cells and avian cells. The use of *Agrobacterium rhizogenes* as a vector for expressing genes in plant cells has been reviewed by Sinkar et al., *J. Biosci. (Bangalore) 11*:47-58, 1987. Transformation of insect cells and production of foreign polypeptides therein is disclosed by Guarino et al., U.S. Patent No. 5,162,222 and WIPO publication WO 94/06463. Insect cells can be infected with recombinant baculovirus, commonly derived from *Autographa californica nuclear polyhedrosis virus* (AcNPV). See, King, L.A. and Possee, R.D., *The Baculovirus Expression System: A Laboratory Guide*, London, Chapman & Hall; O'Reilly, D.R. et al., *Baculovirus Expression Vectors: A Laboratory Manual*, New York, Oxford University Press., 1994; and, Richardson, C. D., Ed., *Baculovirus Expression Protocols. Methods in Molecular Biology*, Totowa, NJ, Humana Press, 1995. A second method of making recombinant zcytor17 baculovirus utilizes a transposon-based system described by Luckow (Luckow, V.A, et al., *J Virol 67*:4566-79, 1993). This system, which utilizes transfer vectors, is sold in the Bac-to-Bac™ kit (Life Technologies, Rockville, MD). This system utilizes a transfer vector, pFastBac1™ (Life Technologies) containing a Tn7 transposon to move the DNA encoding the zcytor17 polypeptide into a baculovirus genome maintained in *E. coli* as a large plasmid called a "bacmid." See, Hill-Perkins, M.S. and Possee, R.D., *J Gen Virol 71*:971-6, 1990; Bonning, B.C. et al., *J Gen Virol 75*:1551-6, 1994; and, Chazenbalk, G.D., and Rapoport, B., *J Biol Chem 270*:1543-9, 1995. In addition, transfer vectors can include an in-frame fusion with DNA encoding an epitope tag at the C- or N-terminus of the expressed zcytor17 polypeptide, for example, a Glu-Glu epitope tag (Grussenmeyer, T. et al., *Proc. Natl. Acad. Sci.*

82:7952-4, 1985). Using a technique known in the art, a transfer vector containing zcytor17 is transformed into *E. coli*, and screened for bacmids which contain an interrupted lacZ gene indicative of recombinant baculovirus. The bacmid DNA containing the recombinant baculovirus genome is isolated, using common techniques, and used to transfect *Spodoptera frugiperda* cells, e.g., Sf9 cells. Recombinant virus that expresses zcytor17 is subsequently produced. Recombinant viral stocks are made by methods commonly used in the art.

The recombinant virus is used to infect host cells, typically a cell line derived from the fall armyworm, *Spodoptera frugiperda*. See, in general, Glick and Pasternak, *Molecular Biotechnology: Principles and Applications of Recombinant DNA*, ASM Press, Washington, D.C., 1994. Another suitable cell line is the High FiveO™ cell line (Invitrogen) derived from *Trichoplusia ni* (U.S. Patent No. 5,300,435). Commercially available serum-free media are used to grow and maintain the cells. Suitable media are Sf900 II™ (Life Technologies) or ESF 921™ (Expression Systems) for the Sf9 cells; and Ex-cellO405™ (JRH Biosciences, Lenexa, KS) or Express FiveO™ (Life Technologies) for the *T. ni* cells. Procedures used are generally described in available laboratory manuals (King, L. A. and Possee, R.D., *ibid.*; O'Reilly, D.R. et al., *ibid.*; Richardson, C. D., *ibid.*). Subsequent purification of the zcytor17 polypeptide from the supernatant can be achieved using methods described herein.

Fungal cells, including yeast cells, can also be used within the present invention. Yeast species of particular interest in this regard include *Saccharomyces cerevisiae*, *Pichia pastoris*, and *Pichia methanolica*. Methods for transforming *S. cerevisiae* cells with exogenous DNA and producing recombinant polypeptides therefrom are disclosed by, for example, Kawasaki, U.S. Patent No. 4,599,311; Kawasaki et al., U.S. Patent No. 4,931,373; Brake, U.S. Patent No. 4,870,008; Welch et al., U.S. Patent No. 5,037,743; and Murray et al., U.S. Patent No. 4,845,075. Transformed cells are selected by phenotype determined by the selectable marker, commonly drug resistance or the ability to grow in the absence of a particular nutrient (e.g., leucine). A preferred vector system for use in *Saccharomyces cerevisiae* is the *POT1* vector system disclosed by Kawasaki et al. (U.S. Patent No. 4,931,373), which allows transformed cells to be selected by growth in glucose-containing media.

Suitable promoters and terminators for use in yeast include those from glycolytic enzyme genes (see, e.g., Kawasaki, U.S. Patent No. 4,599,311; Kingsman et al., U.S. Patent No. 4,615,974; and Bitter, U.S. Patent No. 4,977,092) and alcohol dehydrogenase genes. See also U.S. Patents Nos. 4,990,446; 5,063,154; 5,139,936 and 4,661,454. Transformation systems for other yeasts, including *Hansenula polymorpha, Schizosaccharomyces pombe, Kluyveromyces lactis, Kluyveromyces fragilis, Ustilago maydis, Pichia pastoris, Pichia methanolica, Pichia guillermondii* and *Candida maltosa* are known in the art. See, for example, Gleeson et al., *J. Gen. Microbiol. 132*:3459-3465, 1986 and Cregg, U.S. Patent No. 4,882,279. *Aspergillus* cells may be utilized according to the methods of McKnight et al., U.S. Patent No. 4,935,349. Methods for transforming *Acremonium chrysogenum* are disclosed by Sumino et al., U.S. Patent No. 5,162,228. Methods for transforming *Neurospora* are disclosed by Lambowitz, U.S. Patent No. 4,486,533.

The use of *Pichia methanolica* as host for the production of recombinant proteins is disclosed in WIPO Publications WO 97/17450, WO 97/17451, WO 98/02536, and WO 98/02565. DNA molecules for use in transforming *P. methanolica* will commonly be prepared as double-stranded, circular plasmids, which are preferably linearized prior to transformation. For polypeptide production in *P. methanolica*, it is preferred that the promoter and terminator in the plasmid be that of a *P. methanolica* gene, such as a *P. methanolica* alcohol utilization gene (*AUG1* or *AUG2*). Other useful promoters include those of the dihydroxyacetone synthase (DHAS), formate dehydrogenase (FMD), and catalase (CAT) genes. To facilitate integration of the DNA into the host chromosome, it is preferred to have the entire expression segment of the plasmid flanked at both ends by host DNA sequences. A preferred selectable marker for use in *Pichia methanolica* is a *P. methanolica ADE2* gene, which encodes phosphoribosyl-5-aminoimidazole carboxylase (AIRC; EC 4.1.1.21), which allows *ade2* host cells to grow in the absence of adenine. For large-scale, industrial processes where it is desirable to minimize the use of methanol, it is preferred to use host cells in which both methanol utilization genes (*AUG1* and *AUG2*) are deleted. For production of secreted proteins, host cells deficient in vacuolar protease genes (*PEP4* and *PRB1*) are preferred. Electroporation is used to facilitate the introduction of a plasmid

containing DNA encoding a polypeptide of interest into *P. methanolica* cells. It is preferred to transform *P. methanolica* cells by electroporation using an exponentially decaying, pulsed electric field having a field strength of from 2.5 to 4.5 kV/cm, preferably about 3.75 kV/cm, and a time constant (t) of from 1 to 40 milliseconds, most preferably about 20 milliseconds.

Prokaryotic host cells, including strains of the bacteria *Escherichia coli*, *Bacillus* and other genera are also useful host cells within the present invention. Techniques for transforming these hosts and expressing foreign DNA sequences cloned therein are well known in the art (see, e.g., Sambrook et al., *ibid.*). When expressing a zcytor17 polypeptide in bacteria such as *E. coli*, the polypeptide may be retained in the cytoplasm, typically as insoluble granules, or may be directed to the periplasmic space by a bacterial secretion sequence. In the former case, the cells are lysed, and the granules are recovered and denatured using, for example, guanidine isothiocyanate or urea. The denatured polypeptide can then be refolded and dimerized by diluting the denaturant, such as by dialysis against a solution of urea and a combination of reduced and oxidized glutathione, followed by dialysis against a buffered saline solution. In the latter case, the polypeptide can be recovered from the periplasmic space in a soluble and functional form by disrupting the cells (by, for example, sonication or osmotic shock) to release the contents of the periplasmic space and recovering the protein, thereby obviating the need for denaturation and refolding.

Transformed or transfected host cells are cultured according to conventional procedures in a culture medium containing nutrients and other components required for the growth of the chosen host cells. A variety of suitable media, including defined media and complex media, are known in the art and generally include a carbon source, a nitrogen source, essential amino acids, vitamins and minerals. Media may also contain such components as growth factors or serum, as required. The growth medium will generally select for cells containing the exogenously added DNA by, for example, drug selection or deficiency in an essential nutrient which is complemented by the selectable marker carried on the expression vector or co-transfected into the host cell. *P. methanolica* cells are cultured in a medium comprising adequate sources of carbon, nitrogen and trace nutrients at a temperature of about 25°C

to 35°C. Liquid cultures are provided with sufficient aeration by conventional means, such as shaking of small flasks or sparging of fermentors. A preferred culture medium for *P. methanolica* is YEPD (2% D-glucose, 2% Bacto™ Peptone (Difco Laboratories, Detroit, MI), 1% Bacto™ yeast extract (Difco Laboratories), 0.004% adenine and 0.006% L-leucine).

Within one aspect of the present invention, a zcytor17 multimeric cytokine receptor (including transmembrane and intracellular domains) is produced by a cultured cell, and the cell is used to screen for ligands for the receptor, including the natural ligand (SEQ ID NO:2), as well as agonists and antagonists of the natural ligand. To summarize this approach, a cDNA or gene encoding the receptor is combined with other genetic elements required for its expression (e.g., a transcription promoter), and the resulting expression vector is inserted into a host cell. Cells that express the DNA and produce functional receptor are selected and used within a variety of screening systems.

Mammalian cells suitable for use in expressing the novel receptors of the present invention and transducing a receptor-mediated signal include cells that express a β-subunit, such as gp130, and cells that co-express gp130 and LIF receptor (Gearing et al., *EMBO J. 10*:2839-2848, 1991; Gearing et al., U.S. Patent No. 5,284,755). In this regard it is generally preferred to employ a cell that is responsive to other cytokines that bind to receptors in the same subfamily, such as IL-6 or LIF, because such cells will contain the requisite signal transduction pathway(s). Preferred cells of this type include BaF3 cells (Palacios and Steinmetz, *Cell 41*: 727-734, 1985; Mathey-Prevot et al., *Mol. Cell. Biol. 6*: 4133-4135, 1986), the human TF-1 cell line (ATCC number CRL-2003) and the DA-1 cell line (Branch et al., *Blood 69*:1782, 1987; Broudy et al., *Blood 75*:1622-1626, 1990). In the alternative, suitable host cells can be engineered to produce a β-subunit or other cellular component needed for the desired cellular response. For example, the murine cell line BaF3 (Palacios and Steinmetz, *Cell 41*:727-734, 1985; Mathey-Prevot et al., *Mol. Cell. Biol. 6*: 4133-4135, 1986), a baby hamster kidney (BHK) cell line, or the CTLL-2 cell line (ATCC TIB-214) can be transfected to express the mouse gp130 subunit, or mouse gp130 and LIF receptor, in addition to zcytor17. It is generally preferred to use a host cell and receptor(s) from the

same species, however this approach allows cell lines to be engineered to express multiple receptor subunits from any species, thereby overcoming potential limitations arising from species specificity. In the alternative, species homologs of the human receptor cDNA can be cloned and used within cell lines from the same species, such as a mouse cDNA in the BaF3 cell line. Cell lines that are dependent upon one hematopoietic growth factor, such as IL-3, can thus be engineered to become dependent upon a zcytor17 ligand or anti-zcytor17 antibody.

Cells expressing functional zcytor17 are used within screening assays. A variety of suitable assays are known in the art. These assays are based on the detection of a biological response in the target cell. One such assay is a cell proliferation assay. Cells are cultured in the presence or absence of a test compound, and cell proliferation is detected by, for example, measuring incorporation of tritiated thymidine or by colorimetric assay based on the reduction or metabolic breakdown of Alymar Blue™ (AccuMed, Chicago, IL) or 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) (Mosman, *J. Immunol. Meth. 65*:55-63, 1983). An alternative assay format uses cells that are further engineered to express a reporter gene. The reporter gene is linked to a promoter element that is responsive to the receptor-linked pathway, and the assay detects activation of transcription of the reporter gene. A preferred promoter element in this regard is a serum response element, STAT or SRE (see, for example, Shaw et al., *Cell 56*:563-572, 1989). A preferred such reporter gene is a luciferase gene (de Wet et al., *Mol. Cell. Biol. 7*:725, 1987). Expression of the luciferase gene is detected by luminescence using methods known in the art (e.g., Baumgartner et al., *J. Biol. Chem. 269*:19094-29101, 1994; Schenborn and Goiffin, *Promega Notes 41*:11, 1993). Luciferase assay kits are commercially available from, for example, Promega Corp., Madison, WI. Target cell lines of this type can be used to screen libraries of chemicals, cell-conditioned culture media, fungal broths, soil samples, water samples, and the like. For example, a bank of cell- or tissue-conditioned media samples can be assayed on a target cell to identify cells that produce ligand. Positive cells are then used to produce a cDNA library in a mammalian cell expression vector, which is divided into pools, transfected into host cells, and expressed. Media samples from the transfected cells are then assayed, with subsequent division of pools,

retransfection, subculturing, and re-assay of positive cells to isolate a clonal cell line expressing the ligand. Media samples conditioned by kidney, liver, spleen, thymus, other lymphoid tissues, or T-cells are preferred sources of ligand for use in screening procedures.

The present invention also provides a method of producing a multimeric cytokine receptor. The method includes culturing a cell as described herein, and isolating the multimeric cytokine receptor produced by the cell.

The present invention also provides a method for detecting a multiple cytokine receptor ligand in a test sample. The method includes contacting the test sample with a multimeric cytokine receptor comprising a polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 or SEQ ID NO:109 (amino acid residue 33 to amino acid residue 240 of SEQ ID NO:5), and at least a portion of at least one class I cytokine receptor; and detecting the binding of the multimer cytokine receptor to the ligand in the test sample. The at least a portion of at least one class I cytokine receptor can include, for example, a portion of SEQ ID NO:9 and/or a portion of SEQ ID NO:7, such as, for instance, amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 35 to amino acid residue 137 of SEQ ID NO:7, amino acid residue 240 to amino acid residue 342 of SEQ ID NO:7, amino acid residue 348 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, and/or combinations thereof.

A natural ligand for a zcytor17 multimeric cytokine receptor of the present invention can also be identified by mutagenizing a cytokine-dependent cell line expressing zcytor17 and culturing it under conditions that select for autocrine growth. See WIPO publication WO 95/21930. Within a typical procedure, cells expressing zcytor17 are mutagenized, such as with EMS. The cells are then allowed to recover in the presence of the required cytokine, then transferred to a culture medium lacking the cytokine. Surviving cells are screened for the production of a ligand for zcytor17 multimeric cytokine receptor, such as by adding soluble receptor polypeptide comprising the zcytor17 cytokine-binding domain and at least a portion of a class I cytokine receptor, such as the cytokine-binding domain of OSMRbeta (SEQ ID NO:7) and/or WSX-1 (SEQ ID NO:9), as described herein to the culture medium to compete

against the ligand or by assaying conditioned media on wild-type cells compared to transfected cells expressing the zcytor17 multimeric cytokine receptor. Preferred cell lines for use within this method include cells that are transfected to express gp130 or gp130 in combination with LIF receptor. Preferred such host cell lines include transfected CTLL-2 cells (Gillis and Smith, *Nature 268*:154-156, 1977) and transfected BaF3 cells.

Moreover, a secretion trap method employing zcytor17 soluble multimeric cytokine receptor can be used to isolate a zcytor17 ligand, such as SEQ ID NO:2 (Aldrich, et al, *Cell 87*: 1161-1169, 1996). A cDNA expression library prepared from a known or suspected ligand source is transfected into COS-7 cells. The cDNA library vector generally has an SV40 origin for amplification in COS-7 cells, and a CMV promoter for high expression. The transfected COS-7 cells are grown in a monolayer and then fixed and permeabilized. Tagged or biotin-labeled zcytor17 soluble multimeric cytokine receptor, described herein, is then placed in contact with the cell layer and allowed to bind cells in the monolayer that express an anti-complementary molecule, i.e., a zcytor17 ligand. A cell expressing a ligand will thus be bound with receptor molecules. An anti-tag antibody (anti-Ig for Ig fusions, M2 or anti-FLAG for FLAG-tagged fusions, streptavidin, anti-Glu-Glu tag, and the like) which is conjugated with horseradish peroxidase (HRP) is used to visualize these cells to which the tagged or biotin-labeled zcytor17 soluble multimeric cytokine receptor has bound. The HRP catalyzes deposition of a tyramide reagent, for example, tyramide-FITC. A commercially-available kit can be used for this detection (for example, Renaissance TSA-Direct™ Kit; NEN Life Science Products, Boston, MA). Cells which express zcytor17 multimeric cytokine receptor ligand will be identified under fluorescence microscopy as green cells and picked for subsequent cloning of the ligand using procedures for plasmid rescue as outlined in Aldrich, et al, *supra.*, followed by subsequent rounds of secretion trap assay, or conventional screening of cDNA library pools, until single clones are identified.

As a multimeric receptor complex, the activity of zcytor17 polypeptide can be measured by a silicon-based biosensor microphysiometer which measures the extracellular acidification rate or proton excretion associated with receptor binding and

subsequent physiologic cellular responses. An exemplary device is the Cytosensor™ Microphysiometer manufactured by Molecular Devices, Sunnyvale, CA. A variety of cellular responses, such as cell proliferation, ion transport, energy production, inflammatory response, regulatory and receptor activation, and the like, can be measured by this method. See, for example, McConnell, H.M. et al., *Science 257*:1906-1912, 1992; Pitchford, S. et al., *Meth. Enzymol. 228*:84-108, 1997; Arimilli, S. et al., J. *Immunol. Meth. 212*:49-59, 1998; Van Liefde, I. Et al., *Eur. J. Pharmacol. 346*:87-95, 1998. The microphysiometer can be used for assaying eukaryotic, prokaryotic, adherent or non-adherent cells. By measuring extracellular acidification changes in cell media over time, the microphysiometer directly measures cellular responses to various stimuli, including agonists, ligands, or antagonists of the zcytor17 polypeptide. Preferably, the microphysiometer is used to measure responses of a zcytor17-expressing eukaryotic cell, compared to a control eukaryotic cell that does not express zcytor17 polypeptide. Zcytor17-expressing eukaryotic cells comprise cells into which zcytor17 has been transfected or infected via adenovirus vector, and the like, as described herein, creating a cell that is responsive to zcytor17-modulating stimuli, or are cells naturally expressing zcytor17, such as zcytor17-expressing cells derived from lymphoid, spleen, thymus tissue or PBLs. Differences, measured by an increase or decrease in extracellular acidification, in the response of cells expressing zcytor17, relative to a control, are a direct measurement of zcytor17-modulated cellular responses. Moreover, such zcytor17-modulated responses can be assayed under a variety of stimuli. Also, using the microphysiometer, there is provided a method of identifying agonists and antagonists of zcytor17 multimeric cytokine receptor, comprising providing cells expressing a zcytor17 multimeric cytokine receptor, culturing a first portion of the cells in the absence of a test compound, culturing a second portion of the cells in the presence of a test compound, and detecting an increase or a decrease in a cellular response of the second portion of the cells as compared to the first portion of the cells. Antagonists and agonists, including the natural ligand for zcytor17 multimeric cytokine receptor, can be rapidly identified using this method.

A zcytor17 multimeric cytokine receptor can be expressed as a fusion with an immunoglobulin heavy chain constant region, typically an F_c fragment, which

contains two constant region domains and lacks the variable region. Methods for preparing such fusions are disclosed in U.S. Patents Nos. 5,155,027 and 5,567,584. Such fusions are typically secreted as multimeric molecules wherein the F_c portions are disulfide bonded to each other and two non-Ig polypeptides are arrayed in closed proximity to each other. Fusions of this type can be used for example, for dimerization, increasing stability and *in vivo* half-life, to affinity purify ligand, as *in vitro* assay tool or antagonist. For use in assays, the chimeras are bound to a support via the F_c region and used in an ELISA format.

Additional assays provided by the present invention include the use of hybrid receptor polypeptides. These hybrid polypeptides fall into two general classes. Within the first class, the intracellular domain of zcytor17, comprising approximately residues 544 (Lys) to 732 (Val) of SEQ ID NO:111, residues 544 (Lys) to 649 (Ile) of SEQ ID NO:109, or residues 557 (Lys) to 662 (Ile) of SEQ ID NO:5, or residues 551 (Lys) to 662 (Cys) of SEQ ID NO:117 is joined to the ligand-binding domain of a second receptor. It is preferred that the second receptor be a hematopoietic cytokine receptor, such as, for instance, mpl receptor (Souyri et al., *Cell 63*:1137-1147, 1990). The hybrid receptor will further comprise a transmembrane domain, which may be derived from either receptor. A DNA construct encoding the hybrid receptor is then inserted into a host cell. Cells expressing the hybrid receptor are cultured in the presence of a ligand for the binding domain and assayed for a response. This system provides a means for analyzing signal transduction mediated by zcytor17 while using readily available ligands. This system can also be used to determine if particular cell lines are capable of responding to signals transduced by zcytor17. A second class of hybrid receptor polypeptides comprise the extracellular (ligand-binding) domain (approximately residues 20 (Ala) to 519 (Glu) of SEQ ID NO:111 and SEQ ID NO:109; approximately residues 33 (Ala) to 532 (Glu) of SEQ ID NO:5) or cytokine-binding domain of zcytor17 (approximately residues 20 (Ala) to 227 (Pro) of SEQ ID NO:111 and SEQ ID NO:109; or approximately residues 33 (Ala) to 240 (Pro) of SEQ ID NO:5; approximately residues 46 (Val) to 533 (Glu) of SEQ ID NO:117; or approximately residues 46 (Val) to 533 (Trp) of SEQ ID NO:119) with a cytoplasmic domain of a second receptor, preferably a cytokine receptor, and a transmembrane domain. The

transmembrane domain may be derived from either receptor. Hybrid receptors of this second class are expressed in cells known to be capable of responding to signals transduced by the second receptor. Together, these two classes of hybrid receptors enable the use of a broad spectrum of cell types within receptor-based assay systems.

The expression of WSX-1 is strongest in thymus, spleen, PBL, and lymph node, as well as increased expression observed for activated T-cells. The tissue distribution for OSMRbeta is described as very broad. The tissue distribution of these three receptors suggests that a target for zcytor17lig is hematopoietic lineage cells, in particular T-cells, monocytes/macrophages and lymphoid progenitor cells and lymphoid cells. Other known four-helical-bundle cytokines that act on lymphoid cells include IL-2, IL-4, IL-7, and IL-15. For a review of four-helical-bundle cytokines, see, Nicola et al., *Advances in Protein Chemistry* 52:1-65, 1999 and Kelso, A., *Immunol. Cell Biol.* *76*:300-317, 1998.

Conditioned media (CM) from CD3+ selected, PMA/Ionomycin-stimulated human peripheral blood cells supported the growth of BaF3 cells that expressed the zcytor17 receptor, OSMRbeta and WSX-1 receptor and were otherwise dependent on IL-3. Conditioned medias from cells that were not: 1) PMA/Ionomycin-stimulated; or were not: 2) CD3 selected (with or without PMA/Ionomycin stimulation) did not support the growth of Baf3 cells expressing zcytor17, OSMRbeta and WSX-1 (BaF3/zcytor17/WSX-1/OSMRbeta) receptor-expressing cells. Control experiments demonstrated that this proliferative activity was not attributable to other known growth factors, and that the ability of such conditioned media to stimulate proliferation of zcytor17/WSX-1/OSMRbeta receptor-expressing cells could be neutralized by a soluble form of the zcytor17 receptor.

Conditioned-media from CD3+ selected cells activated with PMA/Ionomycin also supported growth of BaF3 cells that expressed the zcytor17 receptor and OSMRbeta receptor (zcytor17/OSMRbeta), while BaF3 cells expressing only zcytor17 receptor and WSX-1 receptor (zcytor17/WSX-1), or containing only the OSMRbeta receptor, were not stimulated by this conditioned-media.

Proliferation of zcytor17/WSX-1/OSMRbeta receptor-expressing BaF3 cells exposed to CM from CD3+ selected, PMA/Ionomycin-stimulated human

peripheral blood cells were identified by visual inspection of the cultures and/or by proliferation assay. Many suitable proliferation assays are known in the art, and include assays for reduction of a dye such as AlamarBlue™ (AccuMed International, Inc. Westlake, Ohio), 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (Mosman, *J. Immunol. Meth. 65*: 55-63, 1983); 3,(4,5 dimethyl thiazol-2yl)-5-3-carboxymethoxyphenyl-2H-tetrazolium; 2,3-bis(2-methoxy-4-nitro-5-sulfophenyl)-5-[(phenylamino)carbonyl]-2H-tetrazolium hydroxide; and cyanoditolyl-tetrazolium chloride (which are commercially available from Polysciences, Inc., Warrington, PA); mitogenesis assays, such as measurement of incorporation of ^{3}H-thymidine; dye exclusion assays using, for example, naphthalene black or trypan blue; dye uptake using diacetyl fluorescein; and chromium release. See, in general, Freshney, *Culture of Animal Cells: A Manual of Basic Technique*, 3rd ed., Wiley-Liss, 1994, which is incorporated herein by reference.

A cDNA library was prepared from CD3+ selected, PMA- and Ionomycin-stimulated primary human peripheral blood cells. The CD3+ selected, PMA- and Ionomycin-stimulated human peripheral blood cells cDNA library was divided into pools containing multiple cDNA molecules and was transfected into a host cell line, for example, BHK 570 cells (ATCC accession no. 10314). The transfected host cells were cultured in a medium that did not contain exogenous growth factors (e.g., 5% FBS) and conditioned medium was collected. The conditioned media were assayed for the ability to stimulate proliferation of BaF3 cells transfected with the zcytor17, WSX-1, and OSMRbeta receptors. cDNA pools producing conditioned medium that stimulated BaF3/zcytor17/WSX-1/OSMRbeta receptor cells were identified. This pooled plasmid cDNA was electroporated into *E. coli.* cDNA was isolated from single colonies and transfected individually into BHK 570 cells. Positive clones were identified by a positive result in the BaF3/zcytor17/WSX-1/OSMRbeta receptor proliferation assay, and the activity was confirmed by neutralization of proliferation using the soluble zcytor17 receptor.

In view of the tissue distribution observed for zcytor17 receptor agonists (including the natural zcytor17lig/ substrate/ cofactor/ etc.) and/or antagonists have enormous potential in both *in vitro* and *in vivo* applications. Compounds identified as

zcytor17lig agonists are useful for expansion, proliferation, activation, differentiation, and/or induction or inhibition of specialized cell functions of cells involved in homeostasis of hematopoiesis and immune function. For example, zcytor17lig and agonist compounds are useful as components of defined cell culture media, and may be used alone or in combination with other cytokines and hormones to replace serum that is commonly used in cell culture. Agonists are thus useful in specifically promoting the growth and/or development of T-cells, B-cells, monocytes/macrophages, NK cells, cytotoxic lymphocytes, and other cells of the lymphoid and myeloid lineages in culture.

Antagonists are also useful as research reagents for characterizing sites of ligand-receptor interaction. Antagonists are useful to inhibit expansion, proliferation, activation, and/or differentiation of cells involved in regulating hematopoiesis. Inhibitors of zcytor17lig activity (zcytor17lig antagonists) include anti-zcytor17lig antibodies and soluble multimeric cytokine receptors, as well as other peptidic and non-peptidic agents (including ribozymes).

A zcytor17lig-binding protein, such as a multimeric cytokine receptor of the present invention, can also be used for purification of ligand. The multimeric cytokine receptor is immobilized on a solid support, such as beads of agarose, cross-linked agarose, glass, cellulosic resins, silica-based resins, polystyrene, cross-linked polyacrylamide, or like materials that are stable under the conditions of use. Methods for linking polypeptides to solid supports are known in the art, and include amine chemistry, cyanogen bromide activation, N-hydroxysuccinimide activation, epoxide activation, sulfhydryl activation, and hydrazide activation. The resulting medium will generally be configured in the form of a column, and fluids containing ligand are passed through the column one or more times to allow ligand to bind to the receptor polypeptide. The ligand is then eluted using changes in salt concentration, chaotropic agents (guanidine HCl), or pH to disrupt ligand-receptor binding.

An assay system that uses a ligand-binding receptor (or an antibody, one member of a complement/anti-complement pair) or a binding fragment thereof, and a commercially available biosensor instrument (BIAcore, Pharmacia Biosensor, Piscataway, NJ) may be advantageously employed. Such receptor, antibody, member of a complement/anti-complement pair or fragment is immobilized onto the surface of a

receptor chip. Use of this instrument is disclosed by Karlsson, *J. Immunol. Methods 145*:229-40 (1991) and Cunningham and Wells, *J. Mol. Biol. 234*:554-63 (1993). A receptor, antibody, member or fragment is covalently attached, using amine or sulfhydryl chemistry, to dextran fibers that are attached to gold film within the flow cell. A test sample is passed through the cell. If a ligand, epitope, or opposite member of the complement/anti-complement pair is present in the sample, it will bind to the immobilized receptor, antibody or member, respectively, causing a change in the refractive index of the medium, which is detected as a change in surface plasmon resonance of the gold film. This system allows the determination of on- and off-rates, from which binding affinity can be calculated, and assessment of stoichiometry of binding. Alternatively, ligand/receptor binding can be analyzed using SELDI(TM) technology (Ciphergen, Inc., Palo Alto, CA).

Ligand-binding receptor polypeptides can also be used within other assay systems known in the art. Such systems include Scatchard analysis for determination of binding affinity (see Scatchard, *Ann. NY Acad. Sci. 51*: 660-72 (1949)) and calorimetric assays (Cunningham et al., *Science 253*:545-48 (1991); and Cunningham et al., *Science 245*:821-25 (1991)).

The present invention also provides an antibody that specifically binds to a polypeptide or at least at portion of a multimeric cytokine receptor as described herein.

Zcytor17 multimeric cytokine receptors can also be used to prepare antibodies that bind to epitopes, peptides or polypeptides thereof. The multimeric cytokine receptor or a fragment thereof serves as an antigen (immunogen) to inoculate an animal and elicit an immune response. One of skill in the art would recognize that antigenic, epitope-bearing polypeptides may contain a sequence of at least 6, preferably at least 9, and more preferably at least 15 to about 30 contiguous amino acid residues of a polypeptide(s) of the multimeric cytokine receptor, such as zcytor17 (SEQ ID NO:111), OSMRbeta (SEQ ID NO:7), and/or WSX-1 (SEQ ID NO:9). Polypeptides comprising a larger portion of a multimeric cytokine receptor, i.e., from 30 to 100 residues up to the entire length of the amino acid sequence are included. Antigens or

immunogenic epitopes can also include attached tags, adjuvants, carriers and vehicles, as described herein.

Antibodies from an immune response generated by inoculation of an animal with these antigens can be isolated and purified as described herein. Methods for preparing and isolating polyclonal and monoclonal antibodies are well known in the art. See, for example, *Current Protocols in Immunology*, Cooligan, et al. (eds.), National Institutes of Health, John Wiley and Sons, Inc., 1995; Sambrook et al., *Molecular Cloning: A Laboratory Manual, Second Edition*, Cold Spring Harbor, NY, 1989; and Hurrell, J. G. R., Ed., *Monoclonal Hybridoma Antibodies: Techniques and Applications*, CRC Press, Inc., Boca Raton, FL, 1982.

As would be evident to one of ordinary skill in the art, polyclonal antibodies can be generated from inoculating a variety of warm-blooded animals such as horses, cows, goats, sheep, dogs, chickens, rabbits, mice, and rats with a multimeric cytokine receptor or a fragment thereof. The immunogenicity of a multimeric cytokine receptor may be increased through the use of an adjuvant, such as alum (aluminum hydroxide) or Freund's complete or incomplete adjuvant. Multimeric cytokine receptors useful for immunization also include fusion polypeptides, such as fusions of zcytor17, OSMRbeta, and/or WSX-1, or a portion thereof with an immunoglobulin polypeptide or with maltose binding protein. The polypeptide immunogen may be a full-length molecule or a portion thereof. If the polypeptide portion is "hapten-like", such portion may be advantageously joined or linked to a macromolecular carrier (such as keyhole limpet hemocyanin (KLH), bovine serum albumin (BSA) or tetanus toxoid) for immunization.

As used herein, the term "antibodies" includes polyclonal antibodies, affinity-purified polyclonal antibodies, monoclonal antibodies, and antigen-binding fragments, such as $F(ab')_2$ and Fab proteolytic fragments. Genetically engineered intact antibodies or fragments, such as chimeric antibodies, Fv fragments, single chain antibodies and the like, as well as synthetic antigen-binding peptides and polypeptides, are also included. Non-human antibodies may be humanized by grafting non-human CDRs onto human framework and constant regions, or by incorporating the entire non-human variable domains (optionally “cloaking” them with a human-like surface by

replacement of exposed residues, wherein the result is a "veneered" antibody). In some instances, humanized antibodies may retain non-human residues within the human variable region framework domains to enhance proper binding characteristics. Through humanizing antibodies, biological half-life may be increased, and the potential for adverse immune reactions upon administration to humans is reduced. Moreover, human antibodies can be produced in transgenic, non-human animals that have been engineered to contain human immunoglobulin genes as disclosed in WIPO Publication WO 98/24893. It is preferred that the endogenous immunoglobulin genes in these animals be inactivated or eliminated, such as by homologous recombination.

Antibodies are considered to be specifically binding if: 1) they exhibit a threshold level of binding activity, and 2) they do not significantly cross-react with related polypeptide molecules. A threshold level of binding is determined if anti-multimeric cytokine receptor antibodies herein bind to a multimeric cytokine receptor, peptide or epitope with an affinity at least 10-fold greater than the binding affinity to control (non-multimeric cytokine receptor) protein. It is preferred that the antibodies exhibit a binding affinity (K_a) of 10^6 M^{-1} or greater, preferably 10^7 M^{-1} or greater, more preferably 10^8 M^{-1} or greater, and most preferably 10^9 M^{-1} or greater. The binding affinity of an antibody can be readily determined by one of ordinary skill in the art, for example, by Scatchard analysis (Scatchard, G., *Ann. NY Acad. Sci. 51*: 660-672 (1949)).

Whether anti-multimeric cytokine receptor antibodies do not significantly cross-react with related polypeptide molecules is shown, for example, by the antibody detecting zcytor17 multimeric cytokine receptor but not known related polypeptides using a standard Western blot analysis (Ausubel et al., *ibid.*). Examples of known related polypeptides are those disclosed in the prior art, such as known orthologs, and paralogs, and similar known members of a protein family. Screening can also be done using non-human multimeric cytokine receptor, and multimeric cytokine receptor mutant polypeptides. Moreover, antibodies can be "screened against" known related polypeptides, to isolate a population that specifically binds to the multimeric cytokine receptor. For example, antibodies raised to multimeric cytokine receptor are adsorbed to related polypeptides adhered to insoluble matrix; antibodies specific to multimeric cytokine receptor will flow through the matrix under the proper buffer

conditions. Screening allows isolation of polyclonal and monoclonal antibodies non-crossreactive to known closely related polypeptides (*Antibodies: A Laboratory Manual*, Harlow and Lane (eds.), Cold Spring Harbor Laboratory Press, 1988; *Current Protocols in Immunology*, Cooligan, et al. (eds.), National Institutes of Health, John Wiley and Sons, Inc., 1995). Screening and isolation of specific antibodies is well known in the art. See, *Fundamental Immunology*, Paul (eds.), Raven Press, 1993; Getzoff et al., *Adv. in Immunol. 43*: 1-98, 1988; *Monoclonal Antibodies: Principles and Practice*, Goding, J.W. (eds.), *Academic Press Ltd.*, 1996; Benjamin et al., *Ann. Rev. Immunol. 2*: 67-101, 1984. Specifically binding anti-multimeric cytokine receptor antibodies can be detected by a number of methods in the art, and disclosed below.

A variety of assays known to those skilled in the art can be utilized to detect antibodies which bind to multimeric cytokine receptor proteins or polypeptides. Exemplary assays are described in detail in *Antibodies: A Laboratory Manual*, Harlow and Lane (Eds.), Cold Spring Harbor Laboratory Press, 1988. Representative examples of such assays include: concurrent immunoelectrophoresis, radioimmunoassay, radioimmuno-precipitation, enzyme-linked immunosorbent assay (ELISA), dot blot or Western blot assay, inhibition or competition assay, and sandwich assay. In addition, antibodies can be screened for binding to wild-type versus mutant multimeric cytokine receptor protein or polypeptide.

Within another aspect the present invention provides an antibody produced by the method as disclosed above, wherein the antibody binds to at least a portion of a multimer cytokine receptor comprising at least a portion of SEQ ID NO:111, SEQ ID NO:109, or SEQ ID NO:5. In one embodiment, the antibody disclosed above specifically binds to a polypeptide shown in SEQ ID NO:111, SEQ ID NO:109, or SEQ ID NO:5. In another embodiment, the antibody can be a monoclonal antibody or a polyclonal antibody.

Antibodies to multimeric cytokine receptor may be used for tagging cells that express multimeric cytokine receptor; for isolating multimeric cytokine receptor by affinity purification; for diagnostic assays for determining circulating levels of multimeric cytokine receptor; for detecting or quantitating soluble multimeric cytokine receptor as a marker of underlying pathology or disease; in analytical methods

employing FACS; for screening expression libraries; for generating anti-idiotypic antibodies; and as neutralizing antibodies or as antagonists to block multimeric cytokine receptor activity *in vitro* and *in vivo*. Suitable direct tags or labels include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like; indirect tags or labels may feature use of biotin-avidin or other complement/anti-complement pairs as intermediates. Antibodies herein may also be directly or indirectly conjugated to drugs, toxins, radionuclides and the like, and these conjugates used for *in vivo* diagnostic or therapeutic applications. Moreover, antibodies to multimeric cytokine receptor or fragments thereof may be used *in vitro* to detect denatured multimeric cytokine receptor or fragments thereof in assays, for example, Western Blots or other assays known in the art.

Suitable detectable molecules may be directly or indirectly attached to the multimeric cytokine receptor or antibody, and include radionuclides, enzymes, substrates, cofactors, inhibitors, fluorescent markers, chemiluminescent markers, magnetic particles and the like. Suitable cytotoxic molecules may be directly or indirectly attached to the polypeptide or antibody, and include bacterial or plant toxins (for instance, diphtheria, toxin, saporin, *Pseudomonas* exotoxin, ricin, abrin and the like), as well as therapeutic radionuclides, such as iodine-131, rhenium-188 or yttrium-90 (either directly attached to the polypeptide or antibody, or indirectly attached through means of a chelating moiety, for instance). Multimeric cytokine receptors or antibodies may also be conjugated to cytotoxic drugs, such as adriamycin. For indirect attachment of a detectable or cytotoxic molecule, the detectable or cytotoxic molecule can be conjugated with a member of a complementary/anticomplementary pair, where the other member is bound to the polypeptide or antibody portion. For these purposes, biotin/streptavidin is an exemplary complementary/anticomplementary pair.

A soluble multimeric cytokine receptor can also act as zcytor17lig "antagonists" to block zcytor17lig binding and signal transduction *in vitro* and *in vivo*. These anti-zcytor17lig binding proteins would be useful for inhibiting zcytor17lig activity or protein-binding.

Polypeptide-toxin fusion proteins or antibody-toxin fusion proteins can be used for targeted cell or tissue inhibition or ablation (for instance, to treat cancer cells or tissues). Alternatively, if the polypeptide has multiple functional domains (i.e., an activation domain or a receptor binding domain, plus a targeting domain), a fusion protein including only the targeting domain may be suitable for directing a detectable molecule, a cytotoxic molecule or a complementary molecule to a cell or tissue type of interest. In instances where the domain only fusion protein includes a complementary molecule, the anti-complementary molecule can be conjugated to a detectable or cytotoxic molecule. Such domain-complementary molecule fusion proteins thus represent a generic targeting vehicle for cell/tissue-specific delivery of generic anti-complementary-detectable/ cytotoxic molecule conjugates.

Moreover, inflammation is a protective response by an organism to fend off an invading agent. Inflammation is a cascading event that involves many cellular and humoral mediators. On one hand, suppression of inflammatory responses can leave a host immunocompromised; however, if left unchecked, inflammation can lead to serious complications including chronic inflammatory diseases (e.g., rheumatoid arthritis, multiple sclerosis, inflammatory bowel disease and the like), septic shock and multiple organ failure. Importantly, these diverse disease states share common inflammatory mediators. The collective diseases that are characterized by inflammation have a large impact on human morbidity and mortality. Therefore it is clear that anti-inflammatory antibodies and binding polypeptides, such as anti-zcytor17lig antibodies and binding polypeptides described herein, could have crucial therapeutic potential for a vast number of human and animal diseases, from asthma and allergy to autoimmunity and septic shock. As such, use of anti-inflammatory anti zcytor17lig antibodies and binding polypeptides described herein can be used therapeutically as zcytor17lig antagonists described herein, particularly in diseases such as arthritis, endotoxemia, inflammatory bowel disease, psoriasis, related disease and the like.

1. Arthritis

Arthritis, including osteoarthritis, rheumatoid arthritis, arthritic joints as a result of injury, and the like, are common inflammatory conditions which would benefit from the therapeutic use of anti-inflammatory antibodies and binding

polypeptides, such as anti-zcytor17lig antibodies and binding polypeptides of the present invention. For example, rheumatoid arthritis (RA) is a systemic disease that affects the entire body and is one of the most common forms of arthritis. It is characterized by the inflammation of the membrane lining the joint, which causes pain, stiffness, warmth, redness and swelling. Inflammatory cells release enzymes that may digest bone and cartilage. As a result of rheumatoid arthritis, the inflamed joint lining, the synovium, can invade and damage bone and cartilage leading to joint deterioration and severe pain amongst other physiologic effects. The involved joint can lose its shape and alignment, resulting in pain and loss of movement.

Rheumatoid arthritis (RA) is an immune-mediated disease particularly characterized by inflammation and subsequent tissue damage leading to severe disability and increased mortality. A variety of cytokines are produced locally in the rheumatoid joints. Numerous studies have demonstrated that IL-1 and TNF-alpha, two prototypic pro-inflammatory cytokines, play an important role in the mechanisms involved in synovial inflammation and in progressive joint destruction. Indeed, the administration of TNF-alpha and IL-1 inhibitors in patients with RA has led to a dramatic improvement of clinical and biological signs of inflammation and a reduction of radiological signs of bone erosion and cartilage destruction. However, despite these encouraging results, a significant percentage of patients do not respond to these agents, suggesting that other mediators are also involved in the pathophysiology of arthritis (Gabay, *Expert. Opin. Biol. Ther.* 2(2):135-149, 2002). One of those mediators could be zcytor17lig, and as such a molecule that binds or inhibits zcytor17lig, such as anti zcytor17lig antibodies or binding partners, could serve as a valuable therapeutic to reduce inflammation in rheumatoid arthritis, and other arthritic diseases.

There are several animal models for rheumatoid arthritis known in the art. For example, in the collagen-induced arthritis (CIA) model, mice develop chronic inflammatory arthritis that closely resembles human rheumatoid arthritis. Since CIA shares similar immunological and pathological features with RA, this makes it an ideal model for screening potential human anti-inflammatory compounds. The CIA model is a well-known model in mice that depends on both an immune response, and an inflammatory response, in order to occur. The immune response comprises the

interaction of B-cells and CD4+ T-cells in response to collagen, which is given as antigen, and leads to the production of anti-collagen antibodies. The inflammatory phase is the result of tissue responses from mediators of inflammation, as a consequence of some of these antibodies cross-reacting to the mouse's native collagen and activating the complement cascade. An advantage in using the CIA model is that the basic mechanisms of pathogenesis are known. The relevant T-cell and B-cell epitopes on type II collagen have been identified, and various immunological (e.g., delayed-type hypersensitivity and anti-collagen antibody) and inflammatory (e.g., cytokines, chemokines, and matrix-degrading enzymes) parameters relating to immune-mediated arthritis have been determined, and can thus be used to assess test compound efficacy in the CIA model (Wooley, *Curr. Opin. Rheum. 3*:407-20, 1999; Williams et al., *Immunol. 89*:9784-788, 1992; Myers et al., *Life Sci. 61*:1861-78, 1997; and Wang et al., *Immunol. 92*:8955-959, 1995).

The administration of soluble zcytor17 comprising polypeptides (including heterodimeric and multimeric receptors described herein), such as zcytor17-Fc4 or other zcytor17 soluble and fusion proteins to these CIA model mice was used to evaluate the use of zcytor17 to ameliorate symptoms and alter the course of disease. As a molecule that modulates immune and inflammatory response, zcytor17lig, may induce production of SAA, which is implicated in the pathogenesis of rheumatoid arthritis, zcytor17lig antagonists may reduce SAA activity *in vitro* and *in vivo*, the systemic or local administration of zcytor17lig antagonists such as anti-zcytor17lig antibodies or binding partners, zcytor17 comprising polypeptides (including heterodimeric and multimeric receptors described herein), such as zcytor17-Fc4 or other zcytor17 soluble and fusion proteins can potentially suppress the inflammatory response in RA. Other potential therapeutics include zcytor17 polypeptides, soluble heterodimeric and multimeric receptor polypeptides, or anti zcytor17lig antibodies or binding partners of the present invention, and the like.

2. Endotoxemia

Endotoxemia is a severe condition commonly resulting from infectious agents such as bacteria and other infectious disease agents, sepsis, toxic shock syndrome, or in immunocompromised patients subjected to opportunistic infections,

and the like. Therapeutically useful of anti-inflammatory antibodies and binding polypeptides, such as anti-zcytor17lig antibodies and binding polypeptides of the present invention, could aid in preventing and treating endotoxemia in humans and animals. Other potential therapeutics include zcytor17 polypeptides, soluble heterodimeric and multimeric receptor polypeptides, or anti zcytor17lig antibodies or binding partners of the present invention, and the like, could serve as a valuable therapeutic to reduce inflammation and pathological effects in endotoxemia.

Lipopolysaccharide (LPS) induced endotoxemia engages many of the proinflammatory mediators that produce pathological effects in the infectious diseases and LPS induced endotoxemia in rodents is a widely used and acceptable model for studying the pharmacological effects of potential pro-inflammatory or immunomodulating agents. LPS, produced in gram-negative bacteria, is a major causative agent in the pathogenesis of septic shock (Glausner et al., Lancet 338:732, 1991). A shock-like state can indeed be induced experimentally by a single injection of LPS into animals. Molecules produced by cells responding to LPS can target pathogens directly or indirectly. Although these biological responses protect the host against invading pathogens, they may also cause harm. Thus, massive stimulation of innate immunity, occurring as a result of severe Gram-negative bacterial infection, leads to excess production of cytokines and other molecules, and the development of a fatal syndrome, septic shock syndrome, which is characterized by fever, hypotension, disseminated intravascular coagulation, and multiple organ failure (Dumitru et al. Cell 103:1071-1083, 2000).

These toxic effects of LPS are mostly related to macrophage activation leading to the release of multiple inflammatory mediators. Among these mediators, TNF appears to play a crucial role, as indicated by the prevention of LPS toxicity by the administration of neutralizing anti-TNF antibodies (Beutler et al., *Science 229*:869, 1985). It is well established that 1ug injection of *E. coli* LPS into a C57Bl/6 mouse will result in significant increases in circulating IL-6, TNF-alpha, IL-1, and acute phase proteins (for example, SAA) approximately 2 hours post injection. The toxicity of LPS appears to be mediated by these cytokines as passive immunization against these mediators can result in decreased mortality (Beutler et al., *Science 229*:869, 1985). The

potential immunointervention strategies for the prevention and/or treatment of septic shock include anti-TNF mAb, IL-1 receptor antagonist, LIF, IL-10, and G-CSF. Since LPS induces the production of pro- inflammatory factors possibly contributing to the pathology of endotoxemia, the neutralization of zcytor17lig activity, SAA or other pro-inflammatory factors by antagonizing zcytor17lig polypeptide can be used to reduce the symptoms of endotoxemia, such as seen in endotoxic shock. Other potential therapeutics include zcytor17 polypeptides, soluble heterodimeric and multimeric receptor polypeptides, or anti-zcytor17lig antibodies or binding partners of the present invention, and the like.

3 Inflammatory Bowel Disease. IBD

In the United States approximately 500,000 people suffer from Inflammatory Bowel Disease (IBD) which can affect either colon and rectum (Ulcerative colitis) or both, small and large intestine (Crohn's Disease). The pathogenesis of these diseases is unclear, but they involve chronic inflammation of the affected tissues. Potential therapeutics include zcytor17 polypeptides, soluble heterodimeric and multimeric receptor polypeptides, or anti-zcytor17lig antibodies or binding partners of the present invention, and the like, could serve as a valuable therapeutic to reduce inflammation and pathological effects in IBD and related diseases.

Ulcerative colitis (UC) is an inflammatory disease of the large intestine, commonly called the colon, characterized by inflammation and ulceration of the mucosa or innermost lining of the colon. This inflammation causes the colon to empty frequently, resulting in diarrhea. Symptoms include loosening of the stool and associated abdominal cramping, fever and weight loss. Although the exact cause of UC is unknown, recent research suggests that the body's natural defenses are operating against proteins in the body which the body thinks are foreign (an "autoimmune reaction"). Perhaps because they resemble bacterial proteins in the gut, these proteins may either instigate or stimulate the inflammatory process that begins to destroy the lining of the colon. As the lining of the colon is destroyed, ulcers form releasing mucus, pus and blood. The disease usually begins in the rectal area and may eventually extend through the entire large bowel. Repeated episodes of inflammation lead to thickening of the wall of the intestine and rectum with scar tissue. Death of colon

tissue or sepsis may occur with severe disease. The symptoms of ulcerative colitis vary in severity and their onset may be gradual or sudden. Attacks may be provoked by many factors, including respiratory infections or stress.

Although there is currently no cure for UC available, treatments are focused on suppressing the abnormal inflammatory process in the colon lining. Treatments including corticosteroids immunosuppressives (eg. azathioprine, mercaptopurine, and methotrexate) and aminosalicytates are available to treat the disease. However, the long-term use of immunosuppressives such as corticosteroids and azathioprine can result in serious side effects including thinning of bones, cataracts, infection, and liver and bone marrow effects. In the patients in whom current therapies are not successful, surgery is an option. The surgery involves the removal of the entire colon and the rectum.

There are several animal models that can partially mimic chronic ulcerative colitis. The most widely used model is the 2,4,6-trinitrobenesulfonic acid/ethanol (TNBS) induced colitis model, which induces chronic inflammation and ulceration in the colon. When TNBS is introduced into the colon of susceptible mice via intra-rectal instillation, it induces T-cell mediated immune response in the colonic mucosa, in this case leading to a massive mucosal inflammation characterized by the dense infiltration of T-cells and macrophages throughout the entire wall of the large bowel. Moreover, this histopathologic picture is accompanies by the clinical picture of progressive weight loss (wasting), bloody diarrhea, rectal prolapse, and large bowel wall thickening (Neurath et al. *Intern. Rev. Immunol. 19*:51-62, 2000).

Another colitis model uses dextran sulfate sodium (DSS), which induces an acute colitis manifested by bloody diarrhea, weight loss, shortening of the colon and mucosal ulceration with neutrophil infiltration. DSS-induced colitis is characterized histologically by infiltration of inflammatory cells into the lamina propria, with lymphoid hyperplasia, focal crypt damage, and epithelial ulceration. These changes are thought to develop due to a toxic effect of DSS on the epithelium and by phagocytosis of lamina propria cells and production of TNF-alpha and IFN-gamma. Despite its common use, several issues regarding the mechanisms of DSS about the relevance to the human disease remain unresolved. DSS is regarded as a T cell-independent model

because it is observed in T cell-deficient animals such as SCID mice.

The administration of anti-zcytor17lig antibodies or binding partners, soluble zcytor17 comprising polypeptides (including heterodimeric and multimeric receptors), such as zcytor17-Fc4 or other zcytor17 soluble and fusion proteins to these TNBS or DSS models can be used to evaluate the use of zcytor17lig antagonists to ameliorate symptoms and alter the course of gastrointestinal disease. Zcytor17lig may play a role in the inflammatory response in colitis, and the neutralization of zcytor17lig activity by administrating zcytor17lig antagonists is a potential therapeutic approach for IBD. Other potential therapeutics include zcytor17 polypeptides, soluble heterodimeric and multimeric receptor polypeptides, or anti-zcytor17lig antibodies or binding partners of the present invention, and the like.

4. Psoriasis

Psoriasis is a chronic skin condition that affects more than seven million Americans. Psoriasis occurs when new skin cells grow abnormally, resulting in inflamed, swollen, and scaly patches of skin where the old skin has not shed quickly enough. Plaque psoriasis, the most common form, is characterized by inflamed patches of skin ("lesions") topped with silvery white scales. Psoriasis may be limited to a few plaques or involve moderate to extensive areas of skin, appearing most commonly on the scalp, knees, elbows and trunk. Although it is highly visible, psoriasis is not a contagious disease. The pathogenesis of the diseases involves chronic inflammation of the affected tissues. Zcytor17 polypeptides, soluble heterodimeric and multimeric receptor polypeptides, or anti-zcytor17lig antibodies or binding partners of the present invention, and the like, could serve as a valuable therapeutic to reduce inflammation and pathological effects in psoriasis, other inflammatory skin diseases, skin and mucosal allergies, and related diseases.

Psoriasis is a T-cell mediated inflammatory disorder of the skin that can cause considerable discomfort. It is a disease for which there is no cure and affects people of all ages. Psoriasis affects approximately two percent of the populations of European and North America. Although individuals with mild psoriasis can often control their disease with topical agents, more than one million patients worldwide require ultraviolet or systemic immunosuppressive therapy. Unfortunately, the

inconvenience and risks of ultraviolet radiation and the toxicities of many therapies limit their long-term use. Moreover, patients usually have recurrence of psoriasis, and in some cases rebound, shortly after stopping immunosuppressive therapy.

Differentiation is a progressive and dynamic process, beginning with pluripotent stem cells and ending with terminally differentiated cells. Pluripotent stem cells that can regenerate without commitment to a lineage express a set of differentiation markers that are lost when commitment to a cell lineage is made. Progenitor cells express a set of differentiation markers that may or may not continue to be expressed as the cells progress down the cell lineage pathway toward maturation. Differentiation markers that are expressed exclusively by mature cells are usually functional properties such as cell products, enzymes to produce cell products, and receptors. The stage of a cell population's differentiation is monitored by identification of markers present in the cell population.

There is evidence to suggest that factors that stimulate specific cell types down a pathway towards terminal differentiation or dedifferentiation affect the entire cell population originating from a common precursor or stem cell.

A multimeric cytokine receptor of the present invention can be useful for stimulating proliferation, activation, differentiation and/or induction or inhibition of specialized cell function of cells of the involved homeostasis of the hematopoiesis and immune function. In particular, multimeric cytokine receptors as described herein are useful for stimulating proliferation, activation, differentiation, induction or inhibition of specialized cell functions of cells of the hematopoietic lineages, including, but not limited to, T cells, B cells, monocytes/macrophages, NK cells, neutrophils, endothelial cells, fibroblasts, eosinophils, chondrocytes, mast cells, langerhan cells, monocytes, and macrophages, as well as epithelial cells. Epithelial cells include, for example, ameloblasts, chief cells, chromatophores, enterochramaffin cells, enterochromaffin-like cells, goblet cells, granulosa cells, keratinocytes, dendritic cells, labyrinth supporting cells, melanocytes, merkel cells, paneth cells, parietal cells, sertoli cells, and the like.

The present invention also provides a method for reducing hematopoietic cells and hematopoietic cell progenitors of a mammal. The method includes culturing bone marrow or peripheral blood cells with a composition

comprising an effective amount of a soluble multimeric cytokine receptor to produce a decrease in the number of lymphoid cells in the bone marrow or peripheral blood cells as compared to bone marrow or peripheral blood cells cultured in the absence of the multimeric cytokine receptor. The hematopoietic cells and hematopoietic cell progenitors can be lymphoid cells, such as monocytic cells, macrophages, or T cells.

The present invention also provides a method of inhibiting an immune response in a mammal exposed to an antigen or pathogen. The method includes (a) determining directly or indirectly the level of antigen or pathogen present in the mammal; (b) administering a composition comprising a soluble multimeric cytokine receptor in an acceptable pharmaceutical vehicle; (c) determining directly or indirectly the level of antigen or pathogen in the mammal; and (d) comparing the level of the antigen or pathogen in step (a) to the antigen or pathogen level in step (c), wherein a change in the level is indicative of inhibiting an immune response. The method may further include (e) re-administering a composition comprising a multimeric cytokine receptor in an acceptable pharmaceutical vehicle; (f) determining directly or indirectly the level of antigen or pathogen in the mammal; and (g) comparing the number of the antigen or pathogen level in step (a) to the antigen level in step (f), wherein a change in the level is indicative of inhibiting an immune response.

Alternatively, the method can include (a) determining a level of an antigen- or pathogen-specific antibody; (b) administering a composition comprising a soluble multimeric cytokine receptor in an acceptable pharmaceutical vehicle; (c) determining a post administration level of antigen- or pathogen-specific antibody; (d) comparing the level of antibody in step (a) to the level of antibody in step (c), wherein a decrease in antibody level is indicative of inhibiting an immune response.

Zcytor17lig was isolated from tissue known to have important immunological function and which contain cells that play a role in the immune system. Zcytor17lig is expressed in CD3+ selected, activated peripheral blood cells, and it has been shown that zcytor17lig expression increases after T cell activation. Moreover, results of experiments described in the Examples section herein suggest that a multimeric cytokine receptor of the present invention can have an effect on the growth/expansion of monocytes/macrophages, T-cells, B-cells, NK cells and/or

differentiated state of monocytes/macrophages, T-cells, B-cells, NK cells or these cells' progenitors. Factors that both stimulate proliferation of hematopoietic progenitors and activate mature cells are generally known, however, proliferation and activation can also require additional growth factors. For example, it has been shown that IL-7 and Steel Factor (c-kit ligand) were required for colony formation of NK progenitors. IL-15 plus IL-2 in combination with IL-7 and Steel Factor was more effective (Mrózek et al., *Blood 87*:2632-2640, 1996). However, unidentified cytokines may be necessary for proliferation of specific subsets of NK cells and/or NK progenitors (Robertson et. al., *Blood 76*:2451-2438, 1990). Similarly, zcytor17lig may act alone or in concert or synergy with other cytokines to enhance growth, proliferation expansion and modification of differentiation of monocytes/macrophages, T-cells, B-cells or NK cells.

Assays measuring differentiation include, for example, measuring cell markers associated with stage-specific expression of a tissue, enzymatic activity, functional activity or morphological changes (Watt, *FASEB, 5*:281-284 (1991); Francis, *Differentiation 57*:63-75 (1994); and Raes, *Adv. Anim. Cell Biol. Technol. Bioprocesses*, 161-171 (1989)). Alternatively, zcytor17lig polypeptide itself can serve as an additional cell-surface or secreted marker associated with stage-specific expression of a tissue. As such, direct measurement of zcytor17lig polypeptide, or its loss of expression in a tissue as it differentiates, can serve as a marker for differentiation of tissues.

Similarly, direct measurement of zcytor17lig polypeptide, or its loss of expression in a tissue can be determined in a tissue or in cells as they undergo tumor progression. Increases in invasiveness and motility of cells, or the gain or loss of expression of zcytor17lig in a pre-cancerous or cancerous condition, in comparison to normal tissue, can serve as a diagnostic for transformation, invasion and metastasis in tumor progression. As such, knowledge of a tumor's stage of progression or metastasis will aid the physician in choosing the most proper therapy, or aggressiveness of treatment, for a given individual cancer patient. Methods of measuring gain and loss of expression (of either mRNA or protein) are well known in the art and described herein and can be applied to zcytor17lig expression. For example, appearance or disappearance of polypeptides that regulate cell motility can be used to aid diagnosis

and prognosis of prostate cancer (Banyard, J. and Zetter, B.R., *Cancer and Metast. Rev. 17*:449-458, 1999). As an effector of cell motility, zcytor17lig gain or loss of expression may serve as a diagnostic for lymphoid, B-cell, epithelial, hematopoietic and other cancers.

Moreover, the activity and effect of zcytor17lig on tumor progression and metastasis can be measured *in vivo*. Several syngeneic mouse models have been developed to study the influence of polypeptides, compounds or other treatments on tumor progression. In these models, tumor cells passaged in culture are implanted into mice of the same strain as the tumor donor. The cells will develop into tumors having similar characteristics in the recipient mice, and metastasis will also occur in some of the models. Appropriate tumor models for our studies include the Lewis lung carcinoma (ATCC No. CRL-1642) and B16 melanoma (ATCC No. CRL-6323), amongst others. These are both commonly used tumor lines, syngeneic to the C57BL6/J mouse, that are readily cultured and manipulated *in vitro*. Tumors resulting from implantation of either of these cell lines are capable of metastasis to the lung in C57BL6/J mice. The Lewis lung carcinoma model has recently been used in mice to identify an inhibitor of angiogenesis (O'Reilly MS, et al. *Cell 79*: 315-328,1994). C57BL6/J mice are treated with an experimental agent either through daily injection of recombinant protein, agonist or antagonist or a one time injection of recombinant adenovirus. Three days following this treatment, 10^5 to 10^6 cells are implanted under the dorsal skin. Alternatively, the cells themselves may be infected with recombinant adenovirus, such as one expressing zcytor17lig, before implantation so that the protein is synthesized at the tumor site or intracellularly, rather than systemically. The mice normally develop visible tumors within 5 days. The tumors are allowed to grow for a period of up to 3 weeks, during which time they may reach a size of 1500 - 1800 mm^3 in the control treated group. Tumor size and body weight are carefully monitored throughout the experiment. At the time of sacrifice, the tumor is removed and weighed along with the lungs and the liver. The lung weight has been shown to correlate well with metastatic tumor burden. As an additional measure, lung surface metastases are counted. The resected tumor, lungs and liver are prepared for histopathological examination, immunohistochemistry, and *in situ* hybridization, using methods known in

the art and described herein. The influence of the expressed polypeptide in question, e.g., zcytor17lig, on the ability of the tumor to recruit vasculature and undergo metastasis can thus be assessed. In addition, aside from using adenovirus, the implanted cells can be transiently transfected with zcytor17lig. Use of stable zcytor17lig transfectants as well as use of induceable promoters to activate zcytor17lig expression *in vivo* are known in the art and can be used in this system to assess zcytor17lig induction of metastasis. Moreover, purified zcytor17lig or zcytor17lig conditioned media can be directly injected in to this mouse model, and hence be used in this system. For general reference see, O'Reilly MS, et al. *Cell 79*:315-328, 1994; and Rusciano D, et al. Murine Models of Liver Metastasis. *Invasion Metastasis 14*:349-361, 1995.

A soluble multimeric cytokine receptor of the present invention or antibodies thereto may be useful in treating tumorgenesis, and therefore would be useful in the treatment of cancer. Zcytor17lig is expressed in activated T-cells, monocytes and macrophages, and is linked to a region of the human chromosome wherein translocations are common in leukemias. Moreover, the zcytor17lig is shown to act through a cytokine receptor, zcytor17 multimeric cytokine receptor, which is also expressed in activated T-cells, monocytes and macrophages. Over stimulation of activated T-cells, monocytes and macrophages by zcytor17lig could result in a human disease state such as an immune cell cancer. As such, identifying zcytor17lig expression, polypeptides (e.g., by anti-zcytor17lig antibodies, zcytor17 soluble multimeric cytokine receptors (e.g., zcytor17 receptor, heterodimers (e.g., zcytor17/OSMRbeta, zcytor17/WSX-1), multimers (e.g., zcytor17/OSMRbeta/WSX-1)), or other zcytor17lig binding partners) can serve as a diagnostic, and can serve as antagonists of zcytor17lig proliferative activity. The ligand could be administered in combination with other agents already in use including both conventional chemotherapeutic agents as well as immune modulators such as interferon alpha. Alpha/beta interferons have been shown to be effective in treating some leukemias and animal disease models, and the growth inhibitory effects of interferon-alpha and zcytor17lig may be additive.

NK cells are thought to play a major role in elimination of metastatic tumor cells and patients with both metastases and solid tumors have decreased levels of NK cell activity (Whiteside et. al., *Curr. Top. Microbiol. Immunol. 230*:221-244, 1998). An agent that stimulates NK cells would be useful in the elimination of tumors.

The present invention provides a method of reducing proliferation of a neoplastic monocytes/macrophages comprising administering to a mammal with a monocyte/macrophage neoplasm an amount of a composition including a soluble multimeric cytokine receptor or antibody thereto sufficient to reduce proliferation of the neoplastic monocytes/macrophages.

The present invention provides a method for inhibiting activation or differentiation of monocytes/macrophages. Monocytes are incompletely differentiated cells that migrate to various tissues where they mature and become macrophages. Macrophages play a central role in the immune response by presenting antigen to lymphocytes and play a supportive role as accessory cells to lymphocytes by secreting numerous cytokines. Macrophages can internalize extracellular molecules and upon activation have an increased ability to kill intracellular microorganisms and tumor cells. Activated macrophages are also involved in stimulating acute or local inflammation.

In another aspect, the present invention provides a method of reducing proliferation of a neoplastic B or T-cells comprising administering to a mammal with a B or T cell neoplasm an amount of a composition including a soluble multimeric cytokine receptor sufficient to reducing proliferation of the neoplastic monocytes/macrophages. Furthermore, the zcytor17lig antagonist can be a ligand/toxin fusion protein.

A zcytor17 multimeric cytokine receptor-saporin fusion toxin may be employed against a similar set of leukemias and lymphomas, extending the range of leukemias that can be treated with a cytokine antagonsist. For example, such leukemias can be those that over-express zcytor17 receptors (e.g., zcytor17 receptor, heterodimers (e.g., zcytor17/OSMRbeta, zcytor17/WSX-1), multimers (e.g., zcytor17/OSMRbeta/WSX)). Fusion toxin mediated activation of the zcytor17 receptor, zcytor17 receptor heterodimers or multimers (e.g., zcytor17/OSMRbeta, zcytor17/WSX-1 or zcytor17/WSX-1/OSMR) provides two independent means to

inhibit the growth of the target cells, the first being identical to the effects seen by the ligand alone, and the second due to delivery of the toxin through receptor internalization. The lymphoid and monocyte restricted expression pattern of the zcytor17 receptor suggests that the ligand-saporin conjugate can be tolerated by patients.

The tissue distribution of receptors for a given cytokine offers a strong indication of the potential sites of action of that cytokine. Expression of zcytor17 was seen in monocytes and B-cells, with a dramatic increase of expression upon activation for CD3+, CD4+, and CD8+ T-cells. In addition, two monocytic cell lines, THP-1 (Tsuchiya et al., *Int. J. Cancer 26*:171-176, 1980) and U937 (Sundstrom et al., *Int. J. Cancer 17*:565-577, 1976), were also positive for zcytor17 expression.

Northern analysis of WSX-1 receptor revealed transcripts in all tissues examined, with increased levels of expression in human spleen, thymus, lymph node, bone marrow, and peripheral blood leukocytes. Also, expression levels of WSX-1 increased upon activation of T-cells.

Expression of OSMR is reported to be very broad (Mosley et al, *JBC 271*:32635-32643, 1996). This distribution of zcytor17, WSX-1, and OSMRbeta receptors supports a role for zcytor17lig in immune responses, especially expansion of T-cells upon activation or a role in the monocyte/macrophage arm of the immune system.

Thus, particular embodiments of the present invention are directed toward use of a soluble multimeric cytokine receptor, for instance zcytor17/WSX-1/OSMR, and zcytor17/OSMR heterodimers, as antagonists in inflammatory and immune diseases or conditions such as pancreatitis, type I diabetes (IDDM), pancreatic cancer, pancreatitis, Graves Disease, inflammatory bowel disease (IBD), Crohn's Disease, colon and intestinal cancer, diverticulosis, autoimmune disease, sepsis, organ or bone marrow transplant; inflammation due to trauma, surgery or infection; amyloidosis; splenomegaly; graft versus host disease; and where inhibition of inflammation, immune suppression, reduction of proliferation of hematopoietic, immune, inflammatory or lymphoid cells, macrophages, T-cells (including Th1 and Th2 cells, CD4+ and CD8+ cells), suppression of immune response to a pathogen or

antigen. Moreover the presence of zcytor17 expression in activated immune cells such as activated CD4+ and CD19+ cells showed that zcytor17 receptor may be involved in the body's immune defensive reactions against foreign invaders: such as microorganisms and cell debris, and could play a role in immune responses during inflammation and cancer formation. As such, antibodies and binding partners of the present invention that are agonistic or antagonistic to zcytor17 receptor function, such as a soluble zcytor17 multimeric cytokine receptor, can be used to modify immune response and inflammation.

The zcytor17lig structure and tissue expression suggests a role in early hematopoietic or thymocyte development and immune response regulation or inflammation. These processes involve stimulation of cell proliferation and differentiation in response to the binding of one or more cytokines to their cognate receptors. In view of the tissue distribution observed for this zcytor17lig, agonists (including the natural receptor(s)) and antagonists have enormous potential in both *in vitro* and *in vivo* applications. Compounds identified as zcytor17lig agonists are useful for stimulating proliferation and development of target cells *in vitro* and *in vivo*. For example, agonist compounds, zcytor17lig, or anti-zcytor17lig antibodies, are useful as components of defined cell culture media, and may be used alone or in combination with other cytokines and hormones to replace serum that is commonly used in cell culture. Agonists are thus useful in specifically promoting the growth and/or development or activation of monocytes, T-cells, B-cells, and other cells of the lymphoid and myeloid lineages, and hematopoietic cells in culture.

The molecules of the present invention have particular use in the monocyte/macrophage arm of the immune system. Methods are known that can assess such activity. For example, interferon gamma (IFNγ) is a potent activator of mononuclear phagocytes. For example, an increase in expression of zcytor17 upon activation of THP-1 cells (ATCC No. TIB-202) with interferon gamma could suggest that this receptor is involved in monocyte activation. Monocytes are incompletely differentiated cells that migrate to various tissues where they mature and become macrophages. Macrophages play a central role in the immune response by presenting antigen to lymphocytes and play a supportive role as accessory cells to lymphocytes by

secreting numerous cytokines. Macrophages can internalize extracellular molecules and upon activation have an increased ability to kill intracellular microorganisms and tumor cells. Activated macrophages are also involved in stimulating acute or local inflammation. Moreover, monocyte-macrophage function has been shown to be abnormal in a variety of diseased states. For example see, Johnston, RB, *New Eng. J. Med. 318*:747-752, 1998.

One of skill in the art would recognize that agonists of zcytor17 multimeric cytokine receptor, such as zcytor17lig, are useful. For example, depressed migration of monocytes has been reported in populations with a predisposition to infection, such as newborn infants, patients receiving corticosteroid or other immunosuppressive therapy, and patients with diabetes mellitus, burns, or AIDS. Agonists for zcytor17 multimeric cytokine receptor, such as zcytor17lig, could result in an increase in the ability of monocytes to migrate and possibly prevent infection in these populations. There is also a profound defect of phagocytic killing by mononuclear phagocytes from patients with chronic granulomatous disease. This results in the formation of subcutaneous abscesses, as well as abscesses in the liver, lungs, spleen, and lymph nodes. An agonist of zcytor17 multimeric cytokine receptor, such as zcytor17lig, could correct or improve this phagocytic defect. In addition, defective monocyte cytotoxicity has been reported in patients with cancer and Wiskott-Aldrich syndrome (eczema, thrombocytopenia, and recurrent infections). Activation of monocytes by agonists of zcytor17 multimeric cytokine receptor, such as zcytor17lig, could aid in treatment of these conditions. The monocyte-macrophage system is prominently involved in several lipid-storage diseases (sphingolipidoses) such as Gaucher's disease. Resistance to infection can be impaired because of a defect in macrophage function, which could be treated by agonists to zcytor17 multimeric cytokine receptor such as zcytor17lig.

Moreover, one of skill in the art would recognize that antagonists of a zcytor17 multimeric cytokine receptor are useful. For example, in atherosclerotic lesions, one of the first abnormalities is localization of monocyte/macrophages to endothelial cells. These lesions could be prevented by use of antagonists to zcytor17lig. Zcytor17 soluble multimeric cytokine receptors, such as, for instance, heterodimers and

trimers, can also be used as antagonists to the zcytor17lig. Moreover, monoblastic leukemia is associated with a variety of clinical abnormalities that reflect the release of the biologic products of the macrophage, examples include high levels of lysozyme in the serum and urine and high fevers. Moreover, such leukemias exhibit an abnormal increase of monocytic cells. These effects could possibly be prevented by antagonists to zcytor17lig, such as described herein.

Using methods known in the art, and disclosed herein, one of skill could readily assess the activity of a zcytor17 multimeric cytokine receptor in the disease states disclosed herein, inflammation, cancer, or infection as well as other disease states involving monocytic cells. In addition, as zcytor17lig is expressed in a T-cell, macrophage and monocyte-specific manner, and these diseases involve abnormalities in monocytic cells, such as cell proliferation, function, localization, and activation, the polynucleotides, polypeptides, and antibodies of the present invention can be used to as diagnostics to detect such monocytic cell abnormalities, and indicate the presence of disease. Such methods involve taking a biological sample from a patient, such as blood, saliva, or biopsy, and comparing it to a normal control sample. Histological, cytological, flow cytometric, biochemical and other methods can be used to determine the relative levels or localization of zcytor17lig, or cells expressing zcytor17lig, i.e., monocytes, in the patient sample compared to the normal control. A change in the level (increase or decrease) of zcytor17lig expression, or a change in number or localization of monocytes (e.g., increase or infiltration of monocytic cells in tissues where they are not normally present) compared to a control would be indicative of disease. Such diagnostic methods can also include using radiometric, fluorescent, and colorimetric tags attached to polynucleotides, polypeptides or antibodies of the present invention. Such methods are well known in the art and disclosed herein.

Amino acid sequences having zcytor17lig activity can be used to modulate the immune system by binding soluble zcytor17 multimeric cytokine receptor, and thus, preventing the binding of zcytor17lig with endogenous zcytor17lig receptor. Zcytor17lig antagonists, such as a zcytor17 multimeric cytokine receptor, can also be used to modulate the immune system by inhibiting the binding of Zcytor17 ligand with the endogenous zcytor17lig receptor. Accordingly, the present invention includes the

use of a multimeric cytokine receptor that can be also used to treat a subject which produces an excess of either zcytor17lig or Zcytor17 comprising receptor(s). Suitable subjects include mammals, such as humans or veterinary animals.

Zcytor17lig has been shown to be expressed in activated mononuclear cells, and may be involved in regulating inflammation. As such, polypeptides of the present invention can be assayed and used for their ability to modify inflammation, or can be used as a marker for inflammation. Methods to determine proinflammatory and antiinflammatory qualities of zcytor17lig are known in the art and discussed herein. Moreover, it may be involved in up-regulating the production of acute phase reactants, such as serum amyloid A (SAA), α1-antichymotrypsin, and haptoglobin, and that expression of zcytor17 receptor ligand may be increased upon injection of lipopolysaccharide (LPS) *in vivo* that are involved in inflammatory response (Dumoutier, L. et al., *Proc. Nat'l. Acad. Sci. 97*:10144-10149 (2000)). Production of acute phase proteins, such as SAA, is considered a short-term survival mechanism where inflammation is beneficial; however, maintenance of acute phase proteins for longer periods contributes to chronic inflammation and can be harmful to human health. For review, see Uhlar, CM and Whitehead, AS, *Eur. J. Biochem. 265*:501-523 (1999); and Baumann H. and Gauldie, J. *Immunology Today 15*:74-80 (1994). Moreover, the acute phase protein SAA is implicated in the pathogenesis of several chronic inflammatory diseases, is implicated in atherosclerosis and rheumatoid arthritis, and is the precursor to the amyloid A protein deposited in amyloidosis (Uhlar, CM and Whitehead, *supra.*). Thus, where a ligand such as zcytor17lig that acts as a pro-inflammatory molecule and induces production of SAA, antagonists would be useful in treating inflammatory disease and other diseases associated with acute phase response proteins induced by the ligand. Such antagonists are provided by the present invention. For example, a method of reducing inflammation comprises administering to a mammal with or at risk of developing inflammation an amount of a composition of a soluble multimeric cytokine receptor that is sufficient to reduce inflammation. Moreover, a method of suppressing an inflammatory response in a mammal with inflammation can comprise: (1) determining a level of serum amyloid A protein; (2) administering a composition comprising a soluble multimeric cytokine receptor polypeptide as

described herein in an acceptable pharmaceutical vehicle; (3) determining a post administration level of serum amyloid A protein; (4) comparing the level of serum amyloid A protein in step (1) to the level of serum amyloid A protein in step (3), wherein a lack of increase or a decrease in serum amyloid A protein level is indicative of suppressing an inflammatory response.

Like zcytor17lig, analysis of the tissue distribution of the mRNA corresponding it's zcytor17 receptor cDNA showed that mRNA level was highest in monocytes and prostate cells, and is elevated in activated monocytes, and activated CD4+, activated CD8+, and activated CD3+ cells. Hence, zcytor17 receptor is also implicated in inducing inflammatory and immune response. Thus, particular embodiments of the present invention are directed toward use of zcytor17lig-antibodies, and zcytor17lig, as well as soluble zcytor17 receptor heterodimers as antagonists in inflammatory and immune diseases or conditions such as pancreatitis, type I diabetes (IDDM), pancreatic cancer, pancreatitis, Graves Disease, inflammatory bowel disease (IBD), Crohn's Disease, colon and intestinal cancer, diverticulosis, autoimmune disease, sepsis, organ or bone marrow transplant; inflammation due to trauma, sugery or infection; amyloidosis; splenomegaly; graft versus host disease; and where inhibition of inflammation, immune suppression, reduction of proliferation of hematopoietic, immune, inflammatory or lymphoid cells, macrophages, T-cells (including Th1 and Th2 cells, CD4+ and CD8+ cells), suppression of immune response to a pathogen or antigen. Moreover the presence of zcytor17 receptor and zcytor17lig expression in activated immune cells such as activated CD3+, monocytes, CD4+ and CD19+ cells showed that zcytor17 receptor may be involved in the body's immune defensive reactions against foreign invaders: such as microorganisms and cell debris, and could play a role in immune responses during inflammation and cancer formation. As such, zcytor17lig and zcytor17lig-antibodies of the present invention that are agonistic or antagonistic to zcytor17 receptor function, can be used to modify immune response and inflammation.

Moreover, zcytor17lig polypeptides that bind zcytor17 multimeric cytokine receptors and antibodies thereto are useful to:

1) Antagonize or block signaling via a zcytor17 multimeric cytokine

receptor in the treatment of acute inflammation, inflammation as a result of trauma, tissue injury, surgery, sepsis or infection, and chronic inflammatory diseases such as asthma, inflammatory bowel disease (IBD), chronic colitis, splenomegaly, rheumatoid arthritis, recurrent acute inflammatory episodes (e.g., tuberculosis), and treatment of amyloidosis, and atherosclerosis, Castleman's Disease, asthma, and other diseases associated with the induction of acute-phase response.

2) Antagonize or block signaling via the zcytor17 multimeric cytokine receptor in the treatment of autoimmune diseases such as IDDM, multiple sclerosis (MS), systemic Lupus erythematosus (SLE), myasthenia gravis, rheumatoid arthritis, and IBD to prevent or inhibit signaling in immune cells (e.g. lymphocytes, monocytes, leukocytes) via zcytor17 receptor (Hughes C et al., *J. Immunol 153*: 3319-3325 (1994)). Asthma, allergy and other atopic disease may be treated with an MAb against, for example, soluble zcytor17 multimeric cytokine receptors or zcytor17/CRF2-4 heterodimers, to inhibit the immune response or to deplete offending cells. Blocking or inhibiting signaling via zcytor17 multimeric cytokine receptor, using the polypeptides and antibodies of the present invention, may also benefit diseases of the pancreas, kidney, pituitary and neuronal cells. IDDM, NIDDM, pancreatitis, and pancreatic carcinoma may benefit. Zcytor17 multimeric cytokine receptor may serve as a target for MAb therapy of cancer where an antagonizing MAb inhibits cancer growth and targets immune-mediated killing. (Holliger P, and Hoogenboom, *H Nature Biotech. 16*: 1015-1016 (1998)). Mabs to soluble zcytor17 receptor monomers, homodimers, heterodimers and multimers may also be useful to treat nephropathies such as glomerulosclerosis, membranous neuropathy, amyloidosis (which also affects the kidney among other tissues), renal arteriosclerosis, glomerulonephritis of various origins, fibroproliferative diseases of the kidney, as well as kidney dysfunction associated with SLE, IDDM, type II diabetes (NIDDM), renal tumors and other diseases.

3) Agonize or initiate signaling via the zcytor17 multimeric cytokine receptor in the treatment of autoimmune diseases such as IDDM, MS, SLE, myasthenia gravis, rheumatoid arthritis, and IBD. Zcytor17lig may signal lymphocytes or other immune cells to differentiate, alter proliferation, or change production of cytokines or

cell surface proteins that ameliorate autoimmunity. Specifically, modulation of a T-helper cell response to an alternate pattern of cytokine secretion may deviate an autoimmune response to ameliorate disease (Smith JA et al., *J. Immunol. 160*:4841-4849 (1998)). Similarly, zcytor17lig may be used to signal, deplete and deviate immune cells involved in asthma, allergy and atopoic disease. Signaling via zcytor17 multimeric cytokine receptor may also benefit diseases of the pancreas, kidney, pituitary and neuronal cells. IDDM, NIDDM, pancreatitis, and pancreatic carcinoma may benefit. Zcytor17 multimeric cytokine receptor may serve as a target for MAb therapy of pancreatic cancer where a signaling MAb inhibits cancer growth and targets immune-mediated killing (Tutt, AL et al., *J Immunol. 161*: 3175-3185 (1998)). Similarly T-cell specific leukemias, lymphomas, plasma cell dyscrasia (e.g., multiple myeloma), and carcinoma may be treated with monoclonal antibodies (e.g., neutralizing antibody) to zcytor17-comprising soluble receptors of the present invention.

Soluble zcytor17 multimeric cytokine receptors as described herein can be used to neutralize/block zcytor17 receptor ligand activity in the treatment of autoimmune disease, atopic disease, NIDDM, pancreatitis and kidney dysfunction as described above. A soluble form of zcytor17 multimeric cytokine receptor may be used to promote an antibody response mediated by T cells and/or to promote the production of IL-4 or other cytokines by lymphocytes or other immune cells.

A soluble zcytor17 multimeric cytokine receptor may be useful as antagonists of zcytor17lig. Such antagonistic effects can be achieved by direct neutralization or binding of its natural ligand. In addition to antagonistic uses, the soluble receptors can bind zcytor17lig and act as carrier or vehicle proteins, in order to transport zcytor17lig to different tissues, organs, and cells within the body. As such, the soluble receptors can be fused or coupled to molecules, polypeptides or chemical moieties that direct the soluble-receptor-ligand complex to a specific site, such as a tissue, specific immune cell, monocytes, or tumor. For example, in acute infection or some cancers, benefit may result from induction of inflammation and local acute phase response proteins. Thus, the soluble receptors described herein or antibodies thereto can be used to specifically direct the action of a pro-inflammatory zcytor17lig ligand. See, Cosman, D. *Cytokine 5*: 95-106 (1993); and Fernandez-Botran, R. *Exp. Opin.*

Invest. Drugs 9:497-513 (2000).

Moreover, the soluble zcytor17 multimeric cytokine receptors can be used to stabilize the zcytor17lig, to increase the bioavailability, therapeutic longevity, and/or efficacy of the ligand by stabilizing the ligand from degradation or clearance, or by targeting the ligand to a site of action within the body. For example, the naturally occurring IL-6/soluble IL-6R complex stabilizes IL-6 and can signal through the gp130 receptor. See, Cosman, D. *supra.*, and Fernandez-Botran, R. *supra.* Moreover, Zcytor17 may be combined with a cognate ligand such as its ligand to comprise a ligand/soluble receptor complex. Such complexes may be used to stimulate responses from cells presenting a companion receptor subunit. The cell specificity of zcytor17 multimeric cytokine receptor/zcytor17lig complexes may differ from that seen for the ligand administered alone. Furthermore the complexes may have distinct pharmacokinetic properties such as affecting half-life, dose/response and organ or tissue specificity. Zcytor17 multimeric cytokine receptor/ligand complexes thus may have agonist activity to enhance an immune response or stimulate mesangial cells or to stimulate hepatic cells. Alternatively, only tissues expressing a signaling subunit the heterodimerizes with the complex may be affected analogous to the response to IL6/IL6R complexes (Hirota H. et al., *Proc. Nat'l. Acad. Sci. 92*:4862-4866 (1995); and Hirano, T. in Thomason, A. (Ed.) "The Cytokine Handbook", 3rd Ed., p. 208-209). Soluble receptor/cytokine complexes for IL12 and CNTF display similar activities.

Zcytor17lig may also be used within diagnostic systems for the detection of circulating levels of ligand, and in the detection of acute phase inflammatory response. Within a related embodiment, antibodies or other agents that specifically bind to zcytor17lig can be used to detect circulating zcytor17lig polypeptides; conversely, zcytor17lig itself can be used to detect circulating or locally-acting receptor polypeptides. Elevated or depressed levels of ligand or receptor polypeptides may be indicative of pathological conditions, including inflammation or cancer. Moreover, detection of acute phase proteins or molecules such as zcytor17lig can be indicative of a chronic inflammatory condition in certain disease states (e.g., rheumatoid arthritis). Detection of such conditions serves to aid in disease diagnosis as well as help a physician in choosing proper therapy.

Polynucleotides encoding a zcytor17 multimeric cytokine receptor are useful within gene therapy applications where it is desired to increase or inhibit zcytor17lig activity. If a mammal has a mutated or absent zcytor17 gene, the zcytor17 gene of the present invention can be introduced into the cells of the mammal. In one embodiment, a gene encoding a zcytor17 multimeric cytokine receptor is introduced *in vivo* in a viral vector. Such vectors include an attenuated or defective DNA virus, such as, but not limited to, herpes simplex virus (HSV), papillomavirus, Epstein Barr virus (EBV), adenovirus, adeno-associated virus (AAV), and the like. Defective viruses, which entirely or almost entirely lack viral genes, are preferred. A defective virus is not infective after introduction into a cell. Use of defective viral vectors allows for administration to cells in a specific, localized area, without concern that the vector can infect other cells. Examples of particular vectors include, but are not limited to, a defective herpes simplex virus 1 (HSV1) vector (Kaplitt et al., *Molec. Cell. Neurosci.* *2*:320-30 (1991)); an attenuated adenovirus vector, such as the vector described by Stratford-Perricaudet et al., *J. Clin. Invest. 90*:626-30 (1992); and a defective adeno-associated virus vector (Samulski et al., *J. Virol. 61*:3096-101 (1987); and Samulski et al., *J. Virol. 63*:3822-8 (1989)).

A zcytor17 gene of the present invention can be introduced in a retroviral vector, e.g., as described in Anderson et al., U.S. Patent No. 5,399,346; Mann et al. *Cell 33*:153 (1983); Temin et al., U.S. Patent No. 4,650,764; Temin et al., U.S. Patent No. 4,980,289; Markowitz et al., *J. Virol. 62*:1120 (1988); Temin et al., U.S. Patent No. 5,124,263; International Patent Publication No. WO 95/07358, published March 16, 1995 by Dougherty et al.; and Kuo et al., *Blood 82*:845 (1993). Alternatively, the vector can be introduced by lipofection *in vivo* using liposomes. Synthetic cationic lipids can be used to prepare liposomes for *in vivo* transfection of a gene encoding a marker (Felgner et al., *Proc. Natl. Acad. Sci. USA 84*:7413-7 (1987); Mackey et al., *Proc. Natl. Acad. Sci. USA 85*:8027-31 (1988)). The use of lipofection to introduce exogenous genes into specific organs *in vivo* has certain practical advantages. Molecular targeting of liposomes to specific cells represents one area of benefit. More particularly, directing transfection to particular cells represents one area of benefit. For instance, directing transfection to particular cell types would be particularly

advantageous in a tissue with cellular heterogeneity, such as the immune system, pancreas, liver, kidney, and brain. Lipids may be chemically coupled to other molecules for the purpose of targeting. Targeted peptides (e.g., hormones or neurotransmitters), proteins such as antibodies, or non-peptide molecules can be coupled to liposomes chemically.

It is possible to remove the target cells from the body; to introduce the vector as a naked DNA plasmid; and then to re-implant the transformed cells into the body. Naked DNA vectors for gene therapy can be introduced into the desired host cells by methods known in the art, e.g., transfection, electroporation, microinjection, transduction, cell fusion, DEAE dextran, calcium phosphate precipitation, use of a gene gun or use of a DNA vector transporter. See, e.g., Wu et al., *J. Biol. Chem. 267*:963-7 (1992); and Wu et al., *J. Biol. Chem. 263*:14621-4 (1988).

Antisense methodology can be used to inhibit zcytor17 multimeric cytokine receptor gene transcription, such as to inhibit cell proliferation *in vivo*. Polynucleotides that are complementary to a segment of a zcytor17-encoding polynucleotide (e.g., a polynucleotide as set forth in SEQ ID NO:110. SEQ ID NO:108, or SEQ ID NO:4) are designed to bind to zcytor17lig-encoding mRNA and to inhibit translation of such mRNA. Such antisense polynucleotides are used to inhibit expression of zcytor17lig polypeptide-encoding genes in cell culture or in a subject.

Mice engineered to express the zcytor17lig gene, referred to as "transgenic mice," and mice that exhibit a complete absence of zcytor17lig gene function, referred to as "knockout mice," may also be generated (Snouwaert et al., *Science 257*:1083 (1992); Lowell et al., *Nature 366*:740-42 (1993); Capecchi, M.R., *Science 244*: 1288-1292 (1989); Palmiter, R.D. et al. *Annu Rev Genet. 20*: 465-499 (1986)). For example, transgenic mice that over-express zcytor17lig, either ubiquitously or under a tissue-specific or tissue-restricted promoter can be used to ask whether over-expression causes a phenotype. For example, over-expression of a wild-type zcytor17lig polypeptide, polypeptide fragment or a mutant thereof may alter normal cellular processes, resulting in a phenotype that identifies a tissue in which zcytor17lig expression is functionally relevant and may indicate a therapeutic target for the zcytor17lig, its agonists or antagonists. For example, a preferred transgenic mouse

to engineer is one that over-expresses the zcytor17lig (amino acid residues 23-164 of SEQ ID NO:2; or 24-163 of SEQ ID NO:11). Moreover, such over-expression may result in a phenotype that shows similarity with human diseases. Similarly, knockout zcytor17lig mice can be used to determine where zcytor17lig is absolutely required *in vivo*. The phenotype of knockout mice is predictive of the *in vivo* effects of that a zcytor17lig antagonist, such as a soluble zcytor17 multimeric cytokine receptor, may have. The human or mouse zcytor17lig cDNA described herein can be used to generate knockout mice. These mice may be employed to study the zcytor17lig gene and the protein encoded thereby in an *in vivo* system, and can be used as *in vivo* models for corresponding human diseases. Moreover, transgenic mice expression of zcytor17lig antisense polynucleotides or ribozymes directed against zcytor17lig, described herein, can be used analogously to transgenic mice described above. Studies may be carried out by administration of purified zcytor17lig protein, as well.

The present invention also provides a composition which includes an effective amount of a soluble multimeric cytokine receptor comprising a polypeptide comprising amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111 and at least a portion of at least one class I cytokine receptor; and a pharmaceutically acceptable vehicle. The polypeptide may be comprised of various fragement or portions of the extracellular domain of SEQ ID NO:111, SEQ ID NO:109, and/or SEQ ID NO:5, such as for instance, amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111. The at least a portion of at least one class I cytokine receptor can include, for example, a portion of SEQ ID NO:9 and/or a portion of SEQ ID NO:7, such as, for instance, amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 35 to amino acid residue 137 of SEQ ID NO:7, amino acid residue 240 to amino acid residue 342 of SEQ ID NO:7, amino acid residue 348 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, and/or combinations thereof. The multimeric cytokine receptor may further include an affinity tag as described herein.

The present invention also provides an immune cell inhibiting composition which includes an effective amount of a soluble multimeric cytokine

receptor comprising a polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and at least a portion of at least one class I cytokine receptor; and a pharmaceutically acceptable vehicle, wherein the soluble multimeric cytokine receptor inhibits the proliferation of immune cells.

The present invention also provides an inflammatory cell inhibiting composition which includes an effective amount of a soluble multimeric cytokine receptor comprising a polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and at least a portion of at least one class I cytokine receptor; and a pharmaceutically acceptable vehicle, wherein the soluble multimeric cytokine receptor inhibits the proliferation of inflammatory cells.

Experimental evidence suggests a role for zcytor17lig in the progression of diseases that involve the skin or epithelium of internal surfaces, such as, for instance, large intestine, small intestine, pancrease, lung, prostate, uterus, and the like. First, as disclosed herein, zcytor17 receptors, including both OSM receptor beta and zcytor17, are expressed in several cell types located in epithelial surfaces including cell lines derived from lung epithelium, lung fibroblast, prostate, colon, breast, liver epithelium, bone and skin epithelium, bone fibroblast, and the like. Moreover, as disclosed herein, examples from each of these cell types also responded to zcytor17lig activation of a STAT reporter construct. In addition, several cell lines responded to zcytor17lig stimulation by producing increased levels of IL-6, IL-8, MCP-1 (a chemotactic factor) as described herein. In whole, these data suggest a role for zcytor17lig in diseases that involve the epithelium such as, for instance, atopic dermatitis; dermatitis; psoriasis; psoriatic arthritis; eczema; gingivitis; peridontal disease; inflammatory bowel diseases (IBD) (e.g., ulcerative colitis, Crohn's disease); reproductive disorders, such as, for instance, cervical dysplasia, cervical cancer; other skin diseases like cancers: sarcomas; canrcinomas; melanoma, etc. i.e, not just inflammatory diseases, since immune system is involved in activating/curing cancers; diseases involving barrier dysfunction, such as, for instance, graft-versus-host disease (GVHD) and irritable bowel syndrome (IBS); and diseases that involve lung epithelium, such as asthma, emphysema, and the like. In addition, the release of cytokines IL-6, IL-8, and MCP-1 by cells exposed to zcytor17lig suggests that zcytor17lig is involved in inflammation. Therefore, regulation of

zcytor17lig can be useful in the treatment of autoimmune, inflammatory, or cancerous diseases associated with the tissues that express receptor. These diseases include, for example, prostatitis, hepatitis, osteoarthritis, and the like. Zcytor17lig may positively or negatively directly or indirectly regulate these diseases. Therefore, the administration of zcytor17lig can be used to treat diseases as described herein directly or with molecules that inhibit zcytor17lig activity including, for example, both monoclonal antibodies to zcytor17lig or monoclonal antibodies to zcytor17, or monoclonal antibodies that recognize the zcytor17 and OSM receptor beta complex.

Data also suggests that zcytor17lig may be involved in the regulation of TH2 T cell mediated diseases. First, zcytor17lig is made by the TH2 subset of activated T cells. TH2 cells express more zcytor17lig as compared to TH1 cells. In addition, at least two lung epithelial cell lines (SK-LU-1, A549) were stimulated to increase IL13 receptor alpha-2 mRNA in response to zcyto17 ligand stimulation as described herein. There is an association of IL-13 receptor alpha2 chain and tumorigenicity of human breast and pancreatic tumors. This suggests that zcytor17lig may play a role in regulating tumorigenicity of these types of cancers, as well as other cancers. Therefore, the administration of a zcytor17lig antagonist or direct use of zcytor17lig may be useful in treatment of these types of cancers, benign or malignant and at various grades (grades I-IV) and stages (e.g., TNM or AJC staging methods) of tumor development, in mammals, preferably humans.

It is well-known in the art that IL13 is involved in the generation of activated TH2 cells and in TH2 mediated diseases, such as asthma, atopic dermatitis, and the like. Zcytor17lig or zcytor17lig antagonists may be useful in the treatment of diseases that involved TH2 T cells. This would include diseases such as, for instance, atopic dermatitis, asthma, as well as other diseases that are exacerbated by activated TH2 cells. The involvement of zcytor17lig in diseases, such as, for instance, atopic dermatitis, is also supported by the phenotype of the transgenic mice that overexpress zcytor17lig and develop symptoms of atopic dermatitis as described herein.

Despite the preferential expression of zcytor17lig by TH2 cells, there is still some expression of zcytor17lig in TH1 cells and in CD8+ T cells. Therefore, zcytor17lig or its antagonists may be useful in treating diseases that involve immune

modulation of activated T cells including, for example, viral infection, cancers, graft rejection, and the like.

Zcytor17lig may also be involved in the development of cancer. There is expression of the zcytor17 and OSM receptor beta receptors in human bone fibroblast osteosarcomas, human skin fibroblast melanoma, colon epithelial carcinoma, adenocarcinoma, breast epithelial adenocarcinoma, prostate epithelial adenosarcoma, and lung epithelial adenocarcinoma and carcinoma. Therefore, it may be useful to treat tumors of epithelial origin with either zcytor17lig, fragments thereof, or zcytor17lig antagonists which include, but are not limited to, carcinoma, adenocarcinoma, and melanoma. Notwithstanding, zcytor17lig or a zcytor17lig antagonist may be used to treat a cancer, or reduce one or more symptoms of a cancer, from a cancer including but not limited to, squamous cell or epidermoid carcinoma, basal cell carcinoma, adenocarcinoma, papillary carcinoma, cystadenocarcinoma, bronchogenic carcinoma, bronchial adenoma, melanoma, renal cell carcinoma, hepatocellular carcinoma, transitional cell carcinoma, choriocarcinoma, seminoma, embryonal carcinoma, malignant mixed tumor of salivary gland origin, Wilms' tumor, immature teratoma, teratocarcinoma, and other tumors comprising at least some cells of epithelial origin.

Generally, the dosage of administered zcytor17lig polypeptide (or Zcytor16 analog or fusion protein) will vary depending upon such factors as the patient's age, weight, height, sex, general medical condition and previous medical history. Typically, it is desirable to provide the recipient with a dosage of zcytor17lig polypeptide which is in the range of from about 1 pg/kg to 10 mg/kg (amount of agent/body weight of patient), although a lower or higher dosage also may be administered as circumstances dictate. One skilled in the art can readily determine such dosages, and adjustments thereto, using methods known in the art.

Administration of a zcytor17 multimeric receptor agonist or antagonist to a subject can be topical, inhalant, intravenous, intraarterial, intraperitoneal, intramuscular, subcutaneous, intrapleural, intrathecal, by perfusion through a regional catheter, or by direct intralesional injection. When administering therapeutic proteins by injection, the administration may be by continuous infusion or by single or multiple boluses.

Additional routes of administration include oral, mucosal-membrane, pulmonary, and transcutaneous. Oral delivery is suitable for polyester microspheres, zein microspheres, proteinoid microspheres, polycyanoacrylate microspheres, and lipid-based systems (see, for example, DiBase and Morrel, "Oral Delivery of Microencapsulated Proteins," in *Protein Delivery: Physical Systems*, Sanders and Hendren (eds.), pages 255-288 (Plenum Press 1997)). The feasibility of an intranasal delivery is exemplified by such a mode of insulin administration (see, for example, Hinchcliffe and Illum, *Adv. Drug Deliv. Rev. 35*:199 (1999)). Dry or liquid particles comprising Zcytor17 multimeric receptor agonist or antagonist can be prepared and inhaled with the aid of dry-powder dispersers, liquid aerosol generators, or nebulizers (*e.g.*, Pettit and Gombotz, *TIBTECH 16*:343 (1998); Patton *et al., Adv. Drug Deliv. Rev. 35*:235 (1999)). This approach is illustrated by the AERX diabetes management system, which is a hand-held electronic inhaler that delivers aerosolized insulin into the lungs. Studies have shown that proteins as large as 48,000 kDa have been delivered across skin at therapeutic concentrations with the aid of low-frequency ultrasound, which illustrates the feasibility of trascutaneous administration (Mitragotri *et al., Science 269*:850 (1995)). Transdermal delivery using electroporation provides another means to administer a molecule having Zcytor17 multimeric receptor binding activity (Potts *et al., Pharm. Biotechnol. 10*:213 (1997)).

A pharmaceutical composition comprising a protein, polypeptide, or peptide having Zcytor17 multimeric receptor binding activity can be formulated according to known methods to prepare pharmaceutically useful compositions, whereby the therapeutic proteins are combined in a mixture with a pharmaceutically acceptable vehicle. A composition is said to be in a "pharmaceutically acceptable vehicle" if its administration can be tolerated by a recipient patient. Sterile phosphate-buffered saline is one example of a pharmaceutically acceptable vehicle. Other suitable vehicles are well-known to those in the art. See, for example, Gennaro (ed.), *Remington's Pharmaceutical Sciences*, 19th Edition (Mack Publishing Company 1995).

For purposes of therapy, molecules having Zcytor17 multimeric receptor binding activity and a pharmaceutically acceptable vehicle are administered to a patient in a therapeutically effective amount. A combination of a protein, polypeptide, or

peptide having Zcytor17 multimeric receptor binding activity and a pharmaceutically acceptable vehicle is said to be administered in a "therapeutically effective amount" or "effective amount" if the amount administered is physiologically significant. An agent is physiologically significant if its presence results in a detectable change in the physiology of a recipient patient. For example, an agent used to treat inflammation is physiologically significant if its presence alleviates at least a portion of the inflammatory response.

A pharmaceutical composition comprising Zcytor17lig (or Zcytor17lig analog or fusion protein) can be furnished in liquid form, in an aerosol, or in solid form. Liquid forms, are illustrated by injectable solutions, aerosols, droplets, topological solutions and oral suspensions. Exemplary solid forms include capsules, tablets, and controlled-release forms. The latter form is illustrated by miniosmotic pumps and implants (Bremer *et al., Pharm. Biotechnol. 10*:239 (1997); Ranade, "Implants in Drug Delivery," in *Drug Delivery Systems*, Ranade and Hollinger (eds.), pages 95-123 (CRC Press 1995); Bremer *et al.*, "Protein Delivery with Infusion Pumps," in *Protein Delivery: Physical Systems*, Sanders and Hendren (eds.), pages 239-254 (Plenum Press 1997); Yewey *et al.*, "Delivery of Proteins from a Controlled Release Injectable Implant," in *Protein Delivery: Physical Systems*, Sanders and Hendren (eds.), pages 93-117 (Plenum Press 1997)). Other solid forms include creams, pastes, other topological applications, and the like.

Liposomes provide one means to deliver therapeutic polypeptides to a subject intravenously, intraperitoneally, intrathecally, intramuscularly, subcutaneously, or via oral administration, inhalation, or intranasal administration. Liposomes are microscopic vesicles that consist of one or more lipid bilayers surrounding aqueous compartments (see, generally, Bakker-Woudenberg *et al., Eur. J. Clin. Microbiol. Infect. Dis. 12 (Suppl. 1)*:S61 (1993), Kim, *Drugs 46*:618 (1993), and Ranade, "Site-Specific Drug Delivery Using Liposomes as Carriers," in *Drug Delivery Systems*, Ranade and Hollinger (eds.), pages 3-24 (CRC Press 1995)). Liposomes are similar in composition to cellular membranes and as a result, liposomes can be administered safely and are biodegradable. Depending on the method of preparation, liposomes may be unilamellar or multilamellar, and liposomes can vary in size with diameters ranging

from 0.02 μm to greater than 10 μm. A variety of agents can be encapsulated in liposomes: hydrophobic agents partition in the bilayers and hydrophilic agents partition within the inner aqueous space(s) (see, for example, Machy *et al., Liposomes In Cell Biology And Pharmacology* (John Libbey 1987), and Ostro *et al., American J. Hosp. Pharm. 46*:1576 (1989)). Moreover, it is possible to control the therapeutic availability of the encapsulated agent by varying liposome size, the number of bilayers, lipid composition, as well as the charge and surface characteristics of the liposomes.

Liposomes can adsorb to virtually any type of cell and then slowly release the encapsulated agent. Alternatively, an absorbed liposome may be endocytosed by cells that are phagocytic. Endocytosis is followed by intralysosomal degradation of liposomal lipids and release of the encapsulated agents (Scherphof *et al., Ann. N.Y. Acad. Sci. 446*:368 (1985)). After intravenous administration, small liposomes (0.1 to 1.0 μm) are typically taken up by cells of the reticuloendothelial system, located principally in the liver and spleen, whereas liposomes larger than 3.0 μm are deposited in the lung. This preferential uptake of smaller liposomes by the cells of the reticuloendothelial system has been used to deliver chemotherapeutic agents to macrophages and to tumors of the liver.

The reticuloendothelial system can be circumvented by several methods including saturation with large doses of liposome particles, or selective macrophage inactivation by pharmacological means (Claassen *et al., Biochim. Biophys. Acta 802*:428 (1984)). In addition, incorporation of glycolipid- or polyethelene glycol-derivatized phospholipids into liposome membranes has been shown to result in a significantly reduced uptake by the reticuloendothelial system (Allen *et al., Biochim. Biophys. Acta 1068*:133 (1991); Allen *et al., Biochim. Biophys. Acta 1150*:9 (1993)).

Liposomes can also be prepared to target particular cells or organs by varying phospholipid composition or by inserting receptors or ligands into the liposomes. For example, liposomes, prepared with a high content of a nonionic surfactant, have been used to target the liver (Hayakawa *et al.*, Japanese Patent 04-244,018; Kato *et al., Biol. Pharm. Bull. 16*:960 (1993)). These formulations were prepared by mixing soybean phospatidylcholine, α-tocopherol, and ethoxylated hydrogenated castor oil (HCO-60) in methanol, concentrating the mixture under

vacuum, and then reconstituting the mixture with water. A liposomal formulation of dipalmitoylphosphatidylcholine (DPPC) with a soybean-derived sterylglucoside mixture (SG) and cholesterol (Ch) has also been shown to target the liver (Shimizu *et al., Biol. Pharm. Bull. 20*:881 (1997)).

Alternatively, various targeting ligands can be bound to the surface of the liposome, such as antibodies, antibody fragments, carbohydrates, vitamins, and transport proteins. For example, liposomes can be modified with branched type galactosyllipid derivatives to target asialoglycoprotein (galactose) receptors, which are exclusively expressed on the surface of liver cells (Kato and Sugiyama, *Crit. Rev. Ther. Drug Carrier Syst. 14*:287 (1997); Murahashi *et al., Biol. Pharm. Bull.20*:259 (1997)). Similarly, Wu *et al., Hepatology 27*:772 (1998), have shown that labeling liposomes with asialofetuin led to a shortened liposome plasma half-life and greatly enhanced uptake of asialofetuin-labeled liposome by hepatocytes. On the other hand, hepatic accumulation of liposomes comprising branched type galactosyllipid derivatives can be inhibited by preinjection of asialofetuin (Murahashi *et al., Biol. Pharm. Bull.20*:259 (1997)). Polyaconitylated human serum albumin liposomes provide another approach for targeting liposomes to liver cells (Kamps *et al., Proc. Nat'l Acad. Sci. USA 94*:11681 (1997)). Moreover, Geho, *et al.* U.S. Patent No. 4,603,044, describe a hepatocyte-directed liposome vesicle delivery system, which has specificity for hepatobiliary receptors associated with the specialized metabolic cells of the liver.

In a more general approach to tissue targeting, target cells are prelabeled with biotinylated antibodies specific for a ligand expressed by the target cell (Harasym *et al., Adv. Drug Deliv. Rev. 32*:99 (1998)). After plasma elimination of free antibody, streptavidin-conjugated liposomes are administered. In another approach, targeting antibodies are directly attached to liposomes (Harasym *et al., Adv. Drug Deliv. Rev. 32*:99 (1998)).

Polypeptides having Zcytor17 multimeric receptor binding activity can be encapsulated within liposomes using standard techniques of protein microencapsulation (see, for example, Anderson *et al., Infect. Immun. 31*:1099 (1981), Anderson *et al., Cancer Res. 50*:1853 (1990), and Cohen *et al., Biochim. Biophys. Acta 1063*:95 (1991), Alving *et al.* "Preparation and Use of Liposomes in Immunological

Studies," in *Liposome Technology,* 2nd Edition, Vol. III, Gregoriadis (ed.), page 317 (CRC Press 1993), Wassef *et al., Meth. Enzymol. 149*:124 (1987)). As noted above, therapeutically useful liposomes may contain a variety of components. For example, liposomes may comprise lipid derivatives of poly(ethylene glycol) (Allen *et al., Biochim. Biophys. Acta 1150*:9 (1993)).

Degradable polymer microspheres have been designed to maintain high systemic levels of therapeutic proteins. Microspheres are prepared from degradable polymers such as poly(lactide-co-glycolide) (PLG), polyanhydrides, poly (ortho esters), nonbiodegradable ethylvinyl acetate polymers, in which proteins are entrapped in the polymer (Gombotz and Pettit, *Bioconjugate Chem. 6*:332 (1995); Ranade, "Role of Polymers in Drug Delivery," in *Drug Delivery Systems*, Ranade and Hollinger (eds.), pages 51-93 (CRC Press 1995); Roskos and Maskiewicz, "Degradable Controlled Release Systems Useful for Protein Delivery," in *Protein Delivery: Physical Systems*, Sanders and Hendren (eds.), pages 45-92 (Plenum Press 1997); Bartus *et al., Science 281*:1161 (1998); Putney and Burke, *Nature Biotechnology 16*:153 (1998); Putney, *Curr. Opin. Chem. Biol. 2*:548 (1998)). Polyethylene glycol (PEG)-coated nanospheres can also provide vehicles for intravenous administration of therapeutic proteins (see, for example, Gref *et al., Pharm. Biotechnol. 10*:167 (1997)).

The present invention also contemplates chemically modified polypeptides having binding Zcytor17 multimeric receptor activity such as zcytor17 multimeric receptor heterodimeric or multimeric soluble receptors, and Zcytor17 multimeric receptor antagonists, for example anti-zcytor17 multimeric receptor antibodies or binding polypeptides, which a polypeptide is linked with a polymer, as discussed above.

Other dosage forms can be devised by those skilled in the art, as shown, for example, by Ansel and Popovich, *Pharmaceutical Dosage Forms and Drug Delivery Systems,* 5th Edition (Lea & Febiger 1990), Gennaro (ed.), *Remington's Pharmaceutical Sciences*, 19th Edition (Mack Publishing Company 1995), and by Ranade and Hollinger, *Drug Delivery Systems* (CRC Press 1996).

As an illustration, pharmaceutical compositions may be supplied as a kit comprising a container that comprises a polypeptide with a Zcytor17 multimeric

receptor extracellular domain, e.g., zcytor17 multimeric receptor heterodimeric or multimeric soluble receptors, or a Zcytor17 multimeric receptor antagonist (*e.g.*, a neutralizing antibody or antibody fragment that binds a Zcytor17 multimeric receptor polypeptide). Therapeutic polypeptides can be provided in the form of an injectable solution for single or multiple doses, or as a sterile powder that will be reconstituted before injection. Alternatively, such a kit can include a dry-powder disperser, liquid aerosol generator, or nebulizer for administration of a therapeutic polypeptide. Such a kit may further comprise written information on indications and usage of the pharmaceutical composition. Moreover, such information may include a statement that the Zcytor17 multimeric receptor composition is contraindicated in patients with known hypersensitivity to Zcytor17 multimeric receptor.

The complete disclosure of all patents, patent applications, and publications, and electronically available material (e.g., GenBank amino acid and nucleotide sequence submissions) cited herein are incorporated by reference. The foregoing detailed description and examples have been given for clarity of understanding only. No unnecessary limitations are to be understood therefrom. The invention is not limited to the exact details shown and described, for variations obvious to one skilled in the art will be included within the invention defined by the claims.

EXAMPLES

Example 1

Construction of MPL-zcytor17 Polypeptide Chimera: MPL Extracellular and TM Domain Fused to the zcytor17 Intracellular Signaling Domain

The 5' extracellular domain of the murine MPL receptor was isolated from a plasmid containing the murine MPL receptor (PHZ1/MPL plasmid) by digestion with EcoRI and BamHI generating a 1164 bp fragment. The digestion was run on a 1% agarose gel and the fragment was isolated using the Qiaquick gel extraction kit (Qiagen) as per manufacturer's instructions. The rest of the MPL extracellular domain and transmembrane domain were generated using PCR with primers ZC6,673 (SEQ ID

NO:13) and ZC29,082 (SEQ ID NO:14). The reaction conditions were as follows: 15 cycles at 94°C for 1 min., 55°C for 1 min., 72°C for 2 min.; followed by 72°C for 7 min.; then a 4°C soak. The PCR product was run on a 1% agarose gel and the approximately 400bp MPL receptor fragment was isolated using Qiaquick™ gel extraction kit (Qiagen) as per manufacturer's instructions.

The intracellular domain of human zcytor17 was isolated from a plasmid containing zcytor17 receptor cDNA (#23/pCAP) using PCR with primers ZC29,083 (SEQ ID NO:15) and ZC29,145 (SEQ ID NO:16). The polynucleotide sequence that corresponds to the zcytor17 receptor coding sequence is shown in SEQ ID NO:5. The reaction conditions were as per above. The PCR product was run on a 1% agarose gel and the approximately 320 bp zcytor17 fragment isolated using Qiaquick gel extraction kit as per manufacturer's instructions.

Each of the isolated PCR fragments described above were mixed at a 1:1 volumetric ratio and used in a PCR reaction using ZC6673 (SEQ ID NO:13) and ZC29145 (SEQ ID NO:16) to create all but the 5' MPL portion of the MPL-zcytor17 chimera. The reaction conditions were as follows: 15 cycles at 94°C for 1 min., 55°C for 1 min., 72°C for 2 min.; followed by 72°C for 7 min.; then a 4°C soak. The entire PCR product was run on a 1% agarose gel and the approximately 700bp MPL-zcytor17 chimera fragment isolated using Qiaquick gel extraction kit (Qiagen) as per manufacturer's instructions. The MPL-zcytor17 chimera fragment was digested with BamHI (BRL) and XbaI (Boerhinger Mannheim) as per manufacturer's instructions. The entire digest was run on a 1% agarose gel and the cleaved MPL-zcytor17 chimera isolated using Qiaquick™ gel extraction kit (Qiagen) as per manufacturer's instructions. The resultant cleaved MPL-zvytor17 chimera plus 5' MPL EcoRI/BamHI fragment described above were inserted into an expression vector to generate the full MPL-zcytor17 chimeric receptor as described below.

Recipient expression vector pZP-7 was digested with EcoRI (BRL) and Xba1 (BRL) as per manufacturer's instructions, and gel purified as described above. This vector fragment was combined with the EcoRI and XbaI cleaved MPL-zcytor17 PCR chimera isolated above and the EcoRI and BamHI 5' MPL fragment isolated above in a ligation reaction. The ligation was run using T4 Ligase (Epicentre

Technologies), at room temperature for 1 hour as per manufacturer's instructions. A sample of the ligation was electroporated into DH10B ElectroMAX™ electrocompetent *E. coli* cells (25μF, 200Ω, 1.8V). Transformants were plated on LB+Ampicillin plates and single colonies screened by miniprep (Qiagen) and digestion with EcoRI to check for the MPL-zcytor17 chimera. EcoRI digestion of correct clones yield about a 2kb fragment. Confirmation of the MPL-zcytor17 chimera sequence was made by sequence analyses. The insert was approximately 3.1 kb, and was full-length.

Example 2

MPL-zcytor17 Chimera Based Proliferation in BAF3 Assay Using Alamar Blue

A. Construction of BaF3 Cells Expressing MPL-zcytor17 chimera

BaF3, an interleukin-3 (IL-3) dependent pre-lymphoid cell line derived from murine bone marrow (Palacios and Steinmetz, *Cell 41*: 727-734, 1985; Mathey-Prevot et al., *Mol. Cell. Biol. 6*: 4133-4135, 1986), was maintained in complete media (RPMI medium (JRH Bioscience Inc., Lenexa, KS) supplemented with 10% heat-inactivated fetal calf serum, 1-2 ng/ml murine IL-3 (mIL-3) (R & D, Minneapolis, MN), 2mM L-glutaMax-1™ (Gibco BRL), 1 mM Sodium Pyruvate (Gibco BRL), and PSN antibiotics (GIBCO BRL)). Prior to electroporation, pZP-7/MPL-zcytor17 plasmid DNA was prepared and purified using a Qiagen Maxi Prep kit (Qiagen) as per manufacturer's instructions. BaF3 cells for electroporation were washed twice in RPMI media and then resuspended in RPMI media at a cell density of 10^7 cells/ml. One ml of resuspended BaF3 cells was mixed with 30 μg of the pZP-7/MPL-zcytor17 plasmid DNA and transferred to separate disposable electroporation chambers (GIBCO BRL). At room temperature cells were given 5x.1msec shocks at 800 volts followed by 5x2ms shocks at 600 volts delivered by an electroporation apparatus (Cyto-Pulse). Alternatively, cells were electroporated with two serial pulses (800 μFAD/300 V; followed by 1180 μFAD/300 V) delivered by a Cell-Porator (GibcoBRL) electroporation apparatus. The electroporated cells were transferred to 50 ml of complete media and placed in an incubator for 15-24 hours (37°C, 5% CO2). Then Geneticin™ (Gibco) selection (1mg/ml G418) was added to the cells in a T-162 flask to isolate the G418-resistant pool. Pools of the transfected BaF3 cells, hereinafter called

BaF3/MPL-zcytor17 cells, were assayed for signaling capability as described below.

B. Testing the signaling capability of the BaF3/MPL-zcytor17 cells using an Alamar Blue Proliferation Assay

BaF3/MPL-zcytor17 cells were spun down and washed in the complete media, described above, but without mIL-3 (hereinafter referred to as "mIL-3 free media"). The cells were spun and washed 3 times to ensure the removal of the mIL-3. Cells were then counted in a hemacytometer. Cells were plated in a 96-well format at 5000 cells per well in a volume of 100 µl per well using the mIL-3 free media.

Proliferation of the BaF3/MPL-zcytor17 cells was assessed using murine thrombopoietin (mTPO) diluted with mIL-3 free media to 200 ng/ml, 100ng/ml, 50ng/ml, 25ng/ml, 12.5ng/ml, 6.25ng/ml, 3.1ng/ml, 1.5 ng/ml concentrations. One hundred microliters of the diluted mTPO was added to the BaF3/MPL-zcytor17 cells. The total assay volume was 200 µl. Negative controls were run in parallel using mIL-3 free media only, without the addition of mTPO. The assay plates were incubated at 37°C, 5% CO_2 for 3 days at which time Alamar Blue (Accumed, Chicago, IL) was added at 20µl/well. Alamar Blue gives a fluorometric readout based on the metabolic activity of cells, and is thus a direct measurement of cell proliferation in comparison to a negative control. Plates were again incubated at 37°C, 5% CO_2 for 24 hours. Plates were read on the Fmax™ plate reader (Molecular Devices Sunnyvale, CA) using the SoftMax™ Pro program, at wavelengths 544 (Excitation) and 590 (Emission), or a Wallac Victor 2 plate reader (PerkinElmer Life Sciences, Boston, MA).

Results confirmed the signaling capability of the intracellular portion of the zcytor17 receptor, as the thrombopoietin induced proliferation at approximately 9-13 fold over background at mTPO concentrations of 50ng/ml and greater.

Example 3

Construction of Expression Vector Expressing Full-length zcytor17: pZp7pX/zcytor17

A. Cloning of full length zcytor17 cDNA for expression:

To obtain a full-length zcytor17 cDNA, 5' and 3' PCR products were isolated and joined using an internal PstI site. The PCR primers were designed using

the nucleotide sequence SEQ ID NO:4 and include BamHI and Xho I restriction sites for cloning purposes.

A 5' PCR product was generated using a WI-38 cDNA library as a template and oligonucleotides ZC29,359 (SEQ ID NO:18) and ZC27,899 (SEQ ID NO:19) as primers. WI-38 is an in-house cDNA library generated from a human embryonic lung cell line (ATCC CRL-2221). This 5' PCR reaction was run as follows: 30 cycles at 94°C for 1 minute, 65°C for 1 minute, 72°C for 2 minutes, then 72°C for 7 minutes; 10°C soak. The PCR reaction used approximately 3 µg of plasmid prepared from the cDNA library, 20 pmoles of each oligonucleotide, and five units of PWO DNA polymerase (Roche). About 90% of the 5' PCR product was ethanol precipitated, digested with BamHI and PstI and gel purified on a 1.0% agarose gel. The approximately 600 bp band was excised and used for ligation to the cloning vector pUC18 digested with BamHI and PstI. The resulting transformants were sequenced to confirm the zcytor17 cDNA sequence. For one of these transformants, plasmid DNA was prepared and digested with BamHI and PstI. The resulting approximately 600 bp band was gel purified and used for a ligation below to form a full-length cDNA.

A 3' PCR product was generated using a human testes in-house cDNA library as a template and oligonucleotides ZC27,895 (SEQ ID NO:20) and ZC29,122 (SEQ ID NO:21) as primers. This 3' PCR reaction was run as follows: 30 cycles at 94° C for 45 seconds, 65°C for 45 seconds, 72°C for 2 minutes, then 72°C for 7 minutes; 10 °C soak. The entire 3' PCR reaction was gel purified on a 1.0% agarose gel and the major 1500 bp band excised. This band was cloned into the PCR Blunt II TOPO vector using the Zeroblunt TOPO kit (Invitrogen). The resulting transformants were sequenced to confirm the zcytor17 cDNA sequence. For one of these transformants, plasmid DNA was prepared and digested with PstI and XhoI. The resulting approximately 1500 bp band was gel purified. A three-part ligation was performed with the 5' BamHI to Pst I fragment above, the 3' PstI to XhoI fragment, and the expression vector pZp7pX digested with BamHI and XhoI. This generated a pZp7pX plasmid containing a full-length cDNA for zcytor17 (SEQ ID NO:4), designated pZp7p/zcytor17. The full length zcytor17 cDNA in pZp7p/zcytor17 has a silent mutation that changes the T to G at position 1888 of SEQ ID NO:4 (encoding a Gly

residue at residue 464 of SEQ ID NO:5). As this mutation is silent, the zcytor17 cDNA in pZp7p/zcytor17 encodes the polypeptide as shown in SEQ ID NO:5. Plasmid pZp7pX is a mammalian expression vector containing an expression cassette having the CMV promoter, intron A, multiple restriction sites for insertion of coding sequences, and a human growth hormone terminator. The plasmid also has an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a puromycin resistance gene and the SV40 terminator.

B. Construction of Expression Vector Expressing Full-Length WSX-1

The entire WSX-1 receptor (SEQ ID NO:9) was isolated from a plasmid containing the WSX-1 receptor cDNA (SEQ ID NO:8) (United States Patent No. 5,925,735). hWSX-1/pBluescript SK(+) plasmid DNA (Stratagene, La Jolla, CA) was digested with EcoRI and XhoI to generate a 1075 bp fragment, and also digested with XhoI and XbaI to generate a 900 bp fragment. Both digests were run on a 1% agarose gel and the cleaved WSX-1 fragments isolated.

Recipient expression vector pZp7Z was digested with EcoRI and XbaI and gel purified as described above. This vector fragment was combined with the two cleaved zcytor17 fragments isolated above in a ligation reaction using T4 Ligase (BRL). The ligation was incubated at room temperature overnight. A sample of the ligation was electroporated in to DH10B electroMAX™ electrocompetent *E. coli* cells (25μF, 200Ω, 2.3V). Six colonies were grown in culture and miniprepped DNA was prepared and digested to confirm the correct WSX-1 full-length insert of 2.0 kb. The resulting plasmid is pZPZ7Z/WSX-1.

Example 4

Zcytor17 based proliferation in BAF3 assay using Alamar Blue

A. Construction of BaF3 Cells Expressing zcytor17 receptor, WSX-1 receptor and OSMR

BaF3 cells expressing the full-length zcytor17 receptor were constructed as per Example 2A above, using 30μg of the zcytor17 expression vector, described in

Example 3A. One exception is that in place of Geneticin selection, 2µg/ml of Puromycin (ClonTech) was added to the transfected cells in a T-162 flask to isolate the puromycin-resistant pool. The BaF3 cells expressing the zcytor17 receptor mRNA were designated as BaF3/zcytor17. To obtain clones, Baf3/zcytor17 cells were counted in a hemocytometer and plated at 1 cell/well, 0.5 cell/well, 0.1 cell/well, and 0.01 cell/well in 96-well dishes. Fifteen clones were scaled up to T75 flasks, and five clones were assayed for zcytor17 expression. Total RNA was isolated from cell pellets using a S.N.A.P. ™ total RNA Isolation Kit (InVitrogen). First-strand cDNA was synthesized using the proSTAR™ First Strand RT-PCR kit, and then PCR with zcytor17 specific primers ZC29,180 (SEQ ID N0:22) and ZC29,122 (SEQ ID NO:23) was performed to screen the clones for expression of zcytor17. One clone, BaF3/zcytor17#15 was chosen to expand and transfect with the WSX-1 expression vector.

BaF3 cells expressing zcytor17 and full-length WSX-1 were constructed as per Example 2A above, using 30 µg of the WSX-1 expression vector WSX-1/pZp7Z (Example 3B) to electroporate the BaF3/zcytor17#15 cells. One exception is that in place of Geneticin selection, 200 µg/ml Zeocin (InVitrogen) was added to the transfected cells in a T-162 flask to isolate the zeocin-resistant pool. The BaF3 cells expressing zcytor17 and WSX-1 were designated BaF3/zcytor17/hWSX-1. To obtain clones, pools of Baf3/zcytor17/hWSX-1 cells were plated at limiting dilution in 96-well plates. The resulting clones were expanded and total RNA was isolated using a S.N.A.P. ™ total RNA Isolation Kit (InVitrogen). First-strand cDNA was synthesized using the proSTAR™ First Strand RT-PCR kit, and then PCR with WSX-1 specific primers ZC9791 (SEQ ID NO:24) and ZC9793 (SEQ ID NO:25) was used to screen the clones for expression of WSX-1. One clone, BaF3/zcytor17/hWSX-1#5 was chosen to expand further and transfect with the OSMRbeta expression vector.

BaF3 cells expressing zcytor17, WSX-1 and full-length OSMRbeta were constructed as per Example 2A above, using 30 µg of the OSMRbeta expression vector OSMR/pZp7NX described in Example 29 to electroporate the BaF3/zcytor17/hWSX-1#5 cells. The BaF3 cells expressing zcytor17, WSX-1, and OSMRbeta mRNA were designated BaF3/zcytor17/WSX-1/OSMR. To obtain clones, pools of BaF3/zcytor17/WSX-1/OSMRbeta cells were plated at limiting dilution in 96-well

plates. Individual clones were expanded and total RNA was isolated using a S.N.A.P. ™ total RNA Isolation Kit (InVitrogen). First-strand cDNA was synthesized using the proSTAR™ First Strand RT-PCR kit, and then PCR with OSMRbeta specific primers ZC40109 (SEQ ID NO:26) and ZC40112 (SEQ ID NO:27) was used to screen the clones for expression of zcytor17, WSX-1, and OSMR. One clone, BaF3/zcytor17/WSX-1/OSMR#5 was selected and these cells were used to screen for zcytor17lig as described below in Examples 5 and 6.

B. Construction of BaF3 Cells Expressing zcytor17 receptor and OSMR

BaF3 cells expressing the full-length zcytor17 receptor were constructed as per Example 2A above, using 30µg of the zcytor17 expression vector, described in Example 3A. One exception is that in place of Geneticin selection, 2µg/ml of Puromycin (ClonTech) was added to the transfected cells in a T-162 flask to isolate the puromycin-resistant pool. The BaF3 cells expressing the zcytor17 receptor mRNA were designated as BaF3/zcytor17. To obtain clones, pools of Baf3/zcytor17 cells were plated at limiting dilution in 96-well plates. These clones were expanded in culture and total RNA was isolated using a S.N.A.P.™ total RNA Isolation Kit (InVitrogen). First-strand cDNA was synthesized using the proSTAR™ First Strand RT-PCR kit, and then PCR was used to screen the clones for expression of zcytor17. One clone, BaF3/zcytor17 #15 was chosen to expand and transfect with the OSMRbeta expression vector.

BaF3 cells expressing zcytor17 and full-length OSMRbeta were constructed as per Example 2A above, using 30 µg of the OSMRbeta expression vector OSMR/pZp7NX (example 29) to electroporate the BaF3/zcytor17#15 cells. The BaF3 cells expressing zcytor17 and OSMRbeta mRNA were designated BaF3/zcytor17/OSMR. These cells were used to screen for zcytor17lig as described below in Example 5.

Example 5

Screening for zcytor17lig using BaF3/Zcytor17/WSX-1/OSMRbeta cells using an Alamar Blue Proliferation Assay

A. Activation of CCRF-CEM and CCRF-HSB2 cells to test for presence of zcytor17lig

CCRF-CEM and CCRF-HSB2 cells were obtained from ATCC and stimulated in culture to produce conditioned media to test for the presence of zcytor17lig activity as described below. The suspension cells were seeded at 2 x 10^5 cells/ml or 5 x 10^5 cells/ml in RPMI-1640 media supplemented with 10% FBS, 2 mM L-glutamine (GibcoBRL), 1X PSN (GibcoBRL), and activated with 10 ng/ml Phorbol-12-myristate-13-acetate (PMA) (Calbiochem, San Diego, CA) and 0.5µg/ml Ionomycin™ (Calbiochem) for 24 or 48hrs. The supernatant from the stimulated cells was used to assay proliferation of the BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells as described below.

B. Screening for zcytor17lig using BaF3/Zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells using an Alamar Blue Proliferation Assay

BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells were spun down and washed in mIL-3 free media. The cells were spun and washed 3 times to ensure the removal of the mIL-3. Cells were then counted in a hemacytometer. Cells were plated in a 96-well format at 5000 cells per well in a volume of 100 µl per well using the mIL-3 free media.

Proliferation of the BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells was assessed using conditioned media from activated CCRFCEM and CCRF-HSB2 cells (see Example 5A). Conditioned media was diluted with mIL-3 free media to 50%, 25%, 12.5%, 6.25%, 3.125%, 1.5%, 0.75%, and 0.375% concentrations. 100 µl of the diluted conditioned media was added to the BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells. The total assay volume is 200 µl. The assay plates were incubated at 37°C, 5% CO_2 for 3-5 days at which time Alamar Blue (Accumed, Chicago, IL) was added at 20 µl/well. Plates were again incubated at 37°C, 5% CO_2 for 24 hours. Plates were read on the Fmax™ plate reader (Molecular devices) as described above (Example 2).

Results confirmed the proliferative response of the BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells to a factor present in the activated CCRF-CEM and CCRF-HSB2 conditioned media. The

response, as measured, was approximately 10-fold over background at the 25% concentration. The untransfected BaF3 cells did not proliferate in response to this factor, nor did BaF3 cells transfected with zcytor17 and WSX-1 (BaF3/zcytor17/WXS-1 cells), showing that this factor was specific for Zcytor17/OSMRbeta or zcytor17/OSMRbeta/WSX-1 receptors. Moreover soluble zcytor17 receptor diminished this proliferative activity of zcytor17lig in the BaF3/zcytor17/WSX-1/OSMRbeta cells (see, Example 11). Similar results are expected in BaF3/zcytor17/OSMRbeta cells.

C. Human Primary Source used to isolate zcytor17lig

One hundred milliliters blood draws were taken from each of six donors. The blood was drawn using 10X 10 ml vacutainer tubes containing heparin. Blood was pooled from six donors (600ml), diluted 1:1 in PBS, and separated using a Ficoll-Paque® PLUS (Pharmacia Biotech). The isolated primary human cell yield after separation on the ficoll gradient was $1.2X10^9$ cells.

Cells were suspended in 9.6 ml MACS buffer (PBS, 0.5% EDTA, 2mM EDTA). 1.6 ml of cell suspension was removed and 0.4 ml CD3 microbeads (Miltenyi Biotec, Auburn, CA) added. The mixture was incubated for 15 min. at 4°C. These cells labeled with CD3 beads were washed with 30 ml MACS buffer, and then resuspended in 2 ml MACS buffer.

A VS+ column (Miltenyi) was prepared according to the manufacturer's instructions. The VS+ column was then placed in a VarioMACS™ magnetic field (Miltenyi). The column was equilibrated with 5 ml MACS buffer. The isolated primary human cells were then applied to the column. The CD3 negative cells were allowed to pass through. The column was rinsed with 9 ml (3 X 3 ml) MACS buffer. The column was then removed from the magnet and placed over a 15 ml falcon tube. CD3+ cells were eluted by adding 5 ml MACS buffer to the column and bound cells flushed out using the plunger provided by the manufacturer. The incubation of the cells with the CD3 magnetic beads, washes, and VS+ column steps (incubation through elution) above were repeated five more times. The resulting CD3+ fractions from the six column separations were pooled. The yield of CD3+ selected human cells were $3X10^8$

total cells.

A sample of the pooled CD3+ selected human cells was removed for staining and sorting on a fluorescent antibody cell sorter (FACS) to assess their purity. The human CD3+ selected cells were 91% CD3+ cells.

The human CD3+ selected cells were activated by incubating in RPMI + 5% FBS + PMA 10 ng/ml and Ionomycin 0.5 µg/ml (Calbiochem) for 13 hours 37°C. The supernatant from these activated CD3+ selected human cells was tested for zcytor17lig activity as described below. Moreover, the activated CD3+ selected human cells were used to prepare a cDNA library, as described in Example 6, below.

D. Testing supernatant from activated CD3+ selected human cells for zcytor17lig using BaF3/Zcytor17/WSX-1/OSMRbeta cells and an Alamar Blue Proliferation Assay

BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells were spun down and washed in mIL-3 free media. The cells were spun and washed 3 times to ensure the removal of the mIL-3. Cells were then counted in a hemacytometer. Cells were plated in a 96-well format at 5000 cells per well in a volume of 100 µl per well using the mIL-3 free media.

Proliferation of the BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells were assessed using conditioned media from activated CD3+ selected human cells (see Example 5C) diluted with mIL-3 free media to 25%, 12.5%, 6.25%, 3.125%, 1.5%, 0.75%, 0.375% and 0.187% concentrations. 100 µl of the diluted conditioned media was added to the BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells. The total assay volume was 200 µl. The assay plates were incubated and assayed as described in Example 5B.

Results confirmed the proliferative response of the BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells to a factor present in the activated CD3+ selected human Cell conditioned media. The response, as measured, was approximately 15-fold over background at the 25% concentration. The untransfected BaF3 cells did not proliferate in response to this factor, nor did BaF3 cells transfected with zcytor17 and WSX-1 (BaF3/zcytor17/WXS-1 cells), showing that this factor is specific for Zcytor17/OSMRbeta or zcytor17/OSMRbeta/WSX-1

receptors.

Example 6

Cloning of human zcytor17lig from a human CD3+ selected cell Library

Screening of a primary human activated CD3+ selected cell cDNA library revealed an isolated cDNA that is a novel member of the four-helix bundle cytokine family. This cDNA encoded the zcytor17lig. The cDNA was identified by screening for activity of the zcytor17lig using the zcytor17/WSX-1/OSM receptors.

A. The vector for CD3+ selected library construction

The vector for CD3+ selected library construction was pZP7NX. The pZP7NX vector was constructed as follows: The coding region for the DHFR selective marker in vector pZP7 was removed by DNA digestion with NcoI and PstI restriction enzymes (Boehringer Mannheim). The digested DNA was run on 1% agarose gel, cut out and gel purified using the Qiagen Gel Extraction Kit (Qiagen) as per manufacturer's instructions. A DNA fragment representing the coding region of Zeocin selective marker was amplified by PCR method with primers ZC13,946 (SEQ ID NO:28) and ZC13,945 (SEQ ID NO:29), and pZeoSV2(+) as a template. There are additional PstI and BclI restriction sites in primer ZC13,946 (SEQ ID NO:28), and additional NcoI and SfuI sites in primer ZC13,945 (SEQ ID NO:29). The PCR fragment was cut with PstI and NcoI restriction enzymes and cloned into pZP7 vector prepared by cleaving with the same two enzymes and subsequent gel purification. This vector was named pZP7Z. Then the Zeocin coding region was removed by DNA digestion of vector pZP7Z with BclI and SfuI restriction enzymes. The digested DNA was run on 1% agarose gel, cut out and gel purified, and then ligated with a DNA fragment of Neomycin coding region cut from pZem228 vector (deposited at the American Type Culture Collection (ATCC), Manassas, VA; ATCC Deposit No. 69446) with the same restriction enzymes (BclI and SfuI).

This new vector was named pZP7N, in which the coding region for DHFR selective marker was replaced by the coding region for a Neomycin selective marker from vector pZem228. A stuffer fragment including an Xho1 site was added to

pZP7N to create a vector suitable for high efficiency directional cloning of cDNA; this new vector was called pZP7NX. To prepare the vector for cDNA, 20µg of pZP7NX was digested with 20 units of EcoR1 (Life Technologies Gaithersberg, MD) and 20 units of Xho1 (Boehringer Mannheim Indianapolis,IN) for 5 hours at 37°C, then 68°C for 15 minutes. The digest was then run on a 0.8% low melt agarose 1XTAE gel to separate the stuffer from the vector. The vector band was excised and digested with "beta-Agarase" (New England Biolabs, Beverly, MA) following the manufacturer's recommendations. After ethanol precipitation the digested vector was resuspended in water to 45ng/ml in preparation for ligation of CD3+ selected cDNA library described below.

B. Preparation of primary human activated CD3+ selected cell cDNA library

Approximately $1.5X10^8$ primary human CD3+ selected cells stimulated in ionomycin/PMA were isolated by centrifugation after culturing at 37°C for 13 hours (Example 5C). Total RNA was isolated from the cell pellet using the "RNeasy Midi" kit from Qiagen, Inc. (Valencia, CA). mRNA was isolated from 225 micrograms of total RNA using the "MPG mRNA purification kit" from CPG Inc. (Lincoln Park, NJ). 3.4 micrograms of mRNA was isolated and converted to double stranded cDNA using the following procedure.

First strand cDNA from stimulated human CD3+ selected cells was synthesized as follows. Nine µl Oligo d(T)-selected poly(A) CD3+ RNA at a concentration of 0.34 µg/µl and 1.0 µl of 1 µg/µl first strand primer ZC18,698 (SEQ ID NO:30) containing an *Xho*I restriction site were mixed and heated at 65°C for 4 minutes and cooled by chilling on ice. First strand cDNA synthesis was initiated by the addition of 9 µl of first strand buffer (5x SUPERSCRIPT® buffer; (Life Technologies), 4 µl of 100 mM dithiothreitol and 2 µl of a deoxynucleotide triphosphate solution containing 10 mM each of dATP, dGTP, dTTP and 5-methyl-dCTP (Pharmacia Biotech Inc.) to the RNA-primer mixture. The reaction mixture was incubated at 45°C for 4 minutes followed by the addition of 8 µl of 200 U/µl SuperscriptII®, RNase H- reverse transcriptase (Life technologies). The reaction was incubated at 45°C for 45 minutes followed by an incubation ramp of 1°C every 2 minutes to 50°C where the reaction was

held for 10 minutes. To denature any secondary structure and allow for additional extension of the cDNA the reaction was then heated to 70°C for 2 minutes then dropped to 55°C for 4 minutes after which 2 μl of SuperscriptII® RT was added and incubated an additional 15 minutes followed by a ramp up to 70°C 1minute/1°C. Unincorporated nucleotides were removed from the cDNA by twice precipitating in the presence of 2 μg of glycogen carrier, 2.0 M ammonium acetate and 2.5 volume ethanol, followed by a 100 μl wash with 70% ethanol. The cDNA was resuspended in 98 μl water for use in second strand synthesis.

Second strand synthesis was performed on the first strand cDNA under conditions that promoted first strand priming of second strand synthesis resulting in DNA hairpin formation. The second strand reaction contained 98 μl of the first strand cDNA, 30 μl of 5x polymerase I buffer (100 mM Tris: HCl, pH 7.5, 500 mM KCl, 25 mM $MgCl_2$, 50 mM $(NH_4)_2SO_4$), 2 μl of 100 mM dithiothreitol, 6 μl of a solution containing 10 mM of each deoxynucleotide triphosphate, 5 μl of 5 mM b-NAD, 1 μl of 3 U/μl *E. coli* DNA ligase (New England Biolabs Inc.) and 4 μl of 10 U/μl *E. coli* DNA polymerase I (New England Biolabs Inc.). The reaction was assembled at room temperature and was incubated at room temperature for 2 minutes followed by the addition of 4 μl of 3.8 U/μl RNase H (Life Technologies). The reaction was incubated at 15°C for two hours followed by a 15 minute incubation at room temperature. 10 μl of 1M TRIS pH7.4 was added to the reaction and extracted twice with phenol/chloroform and once with chloroform, the organic phases were then back extracted with 50 μl of TE (10mM TRIS pH 7.4, 1mM EDTA), pooled with the other aqueous and ethanol precipitated in the presence of 0.3 M sodium acetate. The pellet was washed with 100 μl 70% ethanol air dried and resuspended in 40 μl water.

The single-stranded DNA of the hairpin structure was cleaved using mung bean nuclease. The reaction mixture contained 40 μl of second strand cDNA, 5 μl of 10x mung bean nuclease buffer (Life technologies), 5 μl of mung bean nuclease (Pharmacia Biotech Corp.) diluted to 1U/μl in 1X mung bean nuclease buffer. The reaction was incubated at 37°C for 45 minutes. The reaction was terminated by the addition of 10 μl of 1 M Tris: HCl, pH 7.4 followed by sequential phenol/chloroform and chloroform extractions as described above. Following the extractions, the cDNA

was ethanol precipitated in the presence of 0.3 M sodium acetate. The pellet was washed with 100 µl 70% ethanol air dried and resuspended in 38 µl water.

The resuspended cDNA was blunt-ended with T4 DNA polymerase. The cDNA, which was resuspended in 38 µl of water, was mixed with 12 µl 5x T4 DNA polymerase buffer (250 mM Tris:HCl, pH 8.0, 250 mM KCl, 25 mM $MgCl_2$), 2 µl 0.1 M dithiothreitol, 6 µl of a solution containing 10 mM of each deoxynucleotide triphosphate and 2 µl of 1 U/µl T4 DNA polymerase (Boehringer Mannheim Corp.). After an incubation of 45 minutes at 15°C, the reaction was terminated by the addition of 30 µl TE followed by sequential phenol/chloroform and chloroform extractions and back extracted with 20 µl TE as described above. The DNA was ethanol precipitated in the presence of 2 µl Pellet Paint™ (Novagen) carrier and 0.3 M sodium acetate and was resuspended 11 µl of water.

*Eco*RI adapters were ligated onto the 5' ends of the cDNA described above to enable cloning into an expression vector. 11 µl of cDNA and 4 µl of 65 pmole/µl of *Eco* RI hemiphophorylated adaptor (Pharmacia Biotech Corp) were mixed with 5 µl 5x ligase buffer (Life Technologies), 2 µl of 10 mM ATP and 3 µl of 1 U/µl T4 DNA ligase (Life Technologies), 1 µl 10X ligation buffer (Promega Corp), 9 µl water. The extra dilution with 1X buffer was to prevent the pellet paint from precipitating. The reaction was incubated 9 hours in a water bath temperature ramp from 10°C to 22°C over 9 hours, followed by 45 minutes at 25°C. The reaction was terminated by incubation at 68°C for 15 minutes.

To facilitate the directional cloning of the cDNA into an expression vector, the cDNA was digested with *Xho*I, resulting in a cDNA having a 5' *Eco* RI cohesive end and a 3' *Xho*I cohesive end. The *Xho*I restriction site at the 3' end of the cDNA had been previously introduced using the ZC18698 (SEQ ID NO:31) primer. Restriction enzyme digestion was carried out in a reaction mixture containing 35 µl of the ligation mix described above, 6 µl of 10x H buffer (Boehringer Mannheim Corp.), 1 µl of 2mg/ml BSA (Biolabs Corp.), 17 µl water and 1.0 µl of 40 U/µl *Xho*I (Boehringer Mannheim). Digestion was carried out at 37°C for 1 hour. The reaction was terminated by incubation at 68°C for 15 minutes followed by ethanol precipitation, washing drying as described above and resuspension in 30 µl water.

The resuspended cDNA was heated to 65°C for 5 minutes and cooled on ice, 4 μl of 5X gel loading dye (Research Genetics Corp.) was added, the cDNA was loaded onto a 0.8% low melt agarose 1X TAE gel (SEA PLAQUE GTG™ low melt agarose; FMC Corp.) and electrophoresed. The contaminating adapters and cDNA below 0.6 Kb in length were excised from the gel. The electrodes were reversed, molten agarose was added to fill in the wells, the buffer was changed and the cDNA was electrophoresed until concentrated near the lane origin. The area of the gel containing the concentrated cDNA was excised and placed in a microfuge tube, and the agarose was melted by heating to 65°C for 15 minutes. Following equilibration of the sample to 45°C, 2 μl of 1 U/μl Beta-agarase I (Biolabs, Inc.) was added, and the mixture was incubated for 90 min. at 45°C to digest the agarose. After incubation, 1 tenth volume of 3 M Na acetate was added to the sample, and the mixture was incubated on ice for 15 minutes. The sample was centrifuged at 14,000 x g for 15 minutes at room temperature to remove undigested agarose, the cDNA was ethanol precipitated, washed in 70% ethanol, air-dried and resuspended in 40 μl water.

To determine the optimum ratio of cDNA to vector several ligations were assembled and electroporated. Briefly, 2 μl of 5X T4 ligase buffer (Life Technologies), 1 μl of 10mM ATP, 1 μl pZP7NX digested with EcoR1-Xho1, 1 μl T4 DNA ligase diluted to 0.25u/μl (Life Technologies) water to 10 μl and 0.5, 1,2 or 3 μl of cDNA were mixed in 4 separate ligations, incubated at 22°C for 4 hours, 68°C for 20 minutes, sodium acetate-ethanol precipitated, washed, dried and resuspended in 10 μl. A single microliter of each ligation was electroporated into 40 μl DH10b ElectroMax™ electrocompetent bacteria (Life Technologies) using a 0.1cm cuvette (Biorad) and a Genepulser, pulse controller™ (Biorad) set to 2.5KV, 25lF, 200ohms. These cells were immediately resuspended in 1 ml SOC broth (Manniatis et al. *supra.*) followed by 500 μl of 50% glycerol-SOC as a preservative. These "glycerol stocks " were frozen in several aliquots at -70°C. An aliquot of each was thawed and plated serially on LB-agar plates supplemented with ampicillin at 100 μg/ml. Colony numbers indicated that the optimum ratio of CD3+ cDNA to pZP7NX vector was 1 μl to 45 ng; such a ligation yielded 4.5 million primary clones.

For the purpose of screening the library using a BaF3-based proliferation

assay (Example 5) glycerol stocks from above were diluted into liquid cultures of 100 or 250 clones per pool in deep well microtiter plates, grown 24 hours at 37°C with shaking and plasmid isolated using a Qiagen kit following the manufacturer's instructions. Such DNA was subsequently transfected into BHK cells, media conditioned 72 hours, harvested and stored at -80°C, and subsequently placed on 5K BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells for 72 hours after which proliferation was assessed using an "Alamar blue" fluorescence assay (Example 5B and Example 2B).

Example 7

Expression Cloning of human zcytor17lig

The glycerol stocks from the activated human CD3+ selected cell library (Example 6) were added to Super Broth II™ (Becton Dickinson, Cockeysville, MD) + 0.1 mg/ml ampicillin (amp) at a concentration of 250 cells per 800 microliters. The *E. coli* were allowed to equilibrate for 24 hours at room temperature. At the time of inoculation, 400 microliters was plated on LB + amp plates to determine the actual titer of the inoculation. After 24 hours the plates were counted and then the final concentration of the SuperBrothII™ + *E. coli* was adjusted so that the final concentration was 250 cells per 1.2 ml. Three times 2 liters were inoculated for a total of 6 liters. The media were then plated into 96-well deep well blocks (Qiagen). Plating was done on the 8-channel Q-Fill2™ dispenser (Genetix, Christchurch, Dorset, UK). The *E. coli* were grown overnight at 37°C shaking at 250 rotations/min. on a New Brunswick Scientific Innova 4900 multi-tier environment shaker. The *E. coli* were spun out of solution at 3000 rpm, using a Beckman GS-6KR centrifuge. These *E. coli* pellets were frozen at -20°C or used fresh before miniprepping the plasmid DNA. Each pellet contains approximately 250 cDNA clones from the human CD3+ selected cell library.

These pools of 250 cDNA clones were then mini-prepped using QIAprep™ 96 Turbo Miniprep kit (Qiagen). Plasmid DNA was eluted using 125 µl of TE (10 mM Tris pH 8, 1 mM EDTA). This plasmid DNA was then used to transfect BHK cells.

BHK transfection

BHK cells were plated in 96-well tissue culture plates at a density of 12,000 cells per well in a volume of 100 µl. per well. Culture media was DMEM (GibcoBRL), 5% heat-inactivated fetal bovine serum, 2 mM L-glutamine (GibcoBRL), 1X PSN (GibcoBRL), 1 mM NaPyruvate (GibcoBRL).

The following day, BHK cells were washed once with 100 µl SFA. SFA is serum-free media which is DMEM/F12 or DMEM (Gibco/BRL), 2 mM GlutaMax™ (Gibco/BRL), 1 mM NaPyruvate, 10 µg/ml transferrin, 5 µg/ml insulin, 10 µg/ml fetuin, 2 µg/ml selenium, 25mM HEPES (Gibco/BRL), 100 µM non-essential amino acids (Gibco/BRL).

A DNA/Lipofectamine™ mix was made as follows: 2.2 µl Lipofectamine™ reagent (Gibco/BRL) was combined with 102.8 µl SFA at room temperature; approximately 5 µl of the plasmid DNA (200 ng/µl) was then added to the Lipofectamine™/SFA to form the DNA/Lipofectamine™ mixture, which was incubated at room temperature for 30 minutes. The SFA was removed from the BHK cells and the cells were incubated with 50 µl of the DNA/lipofectamine™ mix for 5 hours at 37°C with 5% CO_2. Fifty µl of the DNA/Lipofectamine™ mixture was added to each of two wells of the BHK cells, so that transfections were done in duplicate.

After BHK cells were incubated with DNA/Lipofectamine™ mix for 5 hours, the DNA/Lipofectamine™ mix was removed and 100 µl culture media was added. Cells were incubated overnight, the media was removed and replaced with 100 µl. culture media. After culturing cells for 48-72 hours, conditioned media was removed, frozen at -80°C for a minimum of 20 minutes, thawed, and then 50 µl was assayed in the Baf3 proliferation assay, described in Example 5, to identify pools of 250 clones with ligand activity.

Twenty 96-well plates were screened in a single assay. This represented approximately 250 cDNAs/well or 480,000 cDNAs total. Of these, conditioned media from approximately 60 wells (representing 250 cDNAs per well) tested positive in the proliferation assay. One of these positive pools was chosen to break down and isolate a single cDNA that would encode the zcytor17lig. This was pool 62A12.

For pool 62A12, 1 µl. of DNA was used to transform ElectroMax™ DH10B cells (Gibco/BRL) by electroporation. The transformants were plated on LB + amp (100 µg/ml) plates to give single colonies. From the electroporated pool, 672 individual colonies were selected by toothpick into seven 96-well plates containing 1.2 ml of SuperBrothII™ per well. These plates were numbered #62.1 through #62.7. These were cultured overnight and the plasmid DNA miniprepped as above. For all seven plates, plasmid DNA from the breakdown plates was transfected into BHK cells and assayed by proliferation as above, except that transfections were not done in duplicate.

Two positive clones 62.6C7 and 62.6E9 were identified by activity from a total of 672 clones. Plasmid DNA miniprepped from clone 62.6E9 was sequenced and a tentative identification was obtained, but a mixed sequence was obtained from this positive clones. To further isolate the zcytor17lig cDNA to a single clone, 1 µl of DNA from pool 62.6E9 was used to electroporate DH10B cells and the transformants plated on LB + amp (100 µg/ml) plates to give single colonies. Plasmid DNA miniprepped from several colonies was sequenced to give the exact DNA sequence. The polynucleotide sequence of zcytor17lig was full-length (SEQ ID NO:1) and its corresponding amino acid sequence is shown (SEQ ID NO:2).

Example 8

Construction of Mammalian Expression Vectors That Express zcytor17 Soluble Receptors: zcytor17CEE, zcytor17CFLG, zcytor17CHIS and zcytor17-Fc4

A. Construction of zcytor17 Mammalian Expression Vector containing zcytor17CEE, zcytor17CFLG and zcytor17CHIS

An expression vector was prepared for the expression of the soluble, extracellular domain of the zcytor17 polypeptide, pZp9zcytor17CEE, where the construct is designed to express a zcytor17 polypeptide comprised of the predicted initiating methionine and truncated adjacent to the predicted transmembrane domain, and with a C-terminal Glu-Glu tag (SEQ ID NO:32).

An approximately 1500 bp PCR product was generated using ZC29,451 (SEQ ID NO:33) and ZC29,124 (SEQ ID NO:34) as PCR primers to add EcoRI and

BamHI restriction sites. A human HPVS in-house cDNA library was used as a template and PCR amplification was performed as follows: 30 cycles at 94°C for 1 minute, 65°C for 1 minute, 72°C for 1.5 minutes, then 72°C for 7 minutes; 10°C soak. The PCR reaction was ethanol precipitated and digested with EcoRI and BamHI restriction enzymes. The digested PCR product was gel purified on a 1.0% agarose gel and the approximately 1500 bp band excised. This band was then re-amplified using identical primers with the following cycling: 30 cycles at 94°C for 1 minute, 65°C for 1 minute, 72°C for 3 minutes, then 72°C for 7 minutes; 10°C soak. The PCR reaction was ethanol precipitated and digested with EcoRI and BamHI restriction enzymes. The digested PCR product was gel purified on a 1.0% agarose gel and the approximately 1500 bp band excised. The excised DNA was subcloned into plasmid CEEpZp9 that had been cut with EcoRI and BamHI, to generate plasmid with a GLU-GLU C-terminally tagged soluble receptor for zcytor17, zcytor17CEEpZp9. The extracellular domain in the zcytor17CEE cDNA in zcytor17CEEpZp9 has a silent mutation that changes the T to C at position 1705 of SEQ ID NO:4 (encoding a Pro residue at residue 403 of SEQ ID NO:5). As this mutation is silent, the zcytor17 cDNA in zcytor17CEEpZp9 encodes the polypeptide as shown in SEQ ID NO:5. Moreover, because of the construct used, a Gly-Ser residue pair is inserted C-terminal to the end of the soluble, extracellular domain of zcytor17 and prior to the C-terminal Glu-Glu Tag (SEQ ID NO:32). As such, the tag at the C-terminus of the zcytor17 extracellular domain, was a Glu-Glu tag as shown in (SEQ ID NO:17). Plasmid CEEpZp9 is a mammalian expression vector containing an expression cassette having the mouse metallothionein-1 promoter, multiple restriction sites for insertion of coding sequences, and a human growth hormone terminator. The plasmid also has an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a DHFR gene and the SV40 terminator. Using standard molecular biological techniques zcytor17CEEpZp9 was electroporated into DH10B competent cells (GIBCO BRL, Gaithersburg, MD) according to manufacturer's direction and plated onto LB plates containing 100 μg/ml ampicillin, and incubated overnight. Colonies were screened by restriction analysis, or PCR from DNA prepared from individual colonies. The insert sequence of positive clones was verified by

sequence analysis. A large scale plasmid preparation was done using a QIAGEN® Maxi prep kit (Qiagen) according to manufacturer's instructions.

The same process is used to prepare the zcytor17 soluble receptors with a C-terminal his tag, composed of 6 His residues in a row; and a C-terminal FLAG® tag (SEQ ID NO:36), zcytor17CFLAG. To construct these constructs, the aforementioned vector has either the HIS or the FLAG® tag in place of the glu-glu tag (e.g., SEQ ID NO:17; SEQ ID NO:32 or SEQ ID NO:35).

B. Mammalian Expression Construction of Soluble human zcytor17 receptor: zcytor17-Fc4

An expression vector, pEZE-2 hzcytor17/Fc4, was prepared to express a C-terminally Fc4 tagged soluble version of hzcytor17 (human zcytor17-Fc4) in PF CHO cells. PF CHO cells are an in house CHO cell line adapted for growth in protein-free medium (ExCell 325 PF medium; JRH Biosciences). The in house CHO cell line was originally derived from CHO DG44 cells (G. Urlaub, J. Mitchell, E. Kas, L.A. Chasin, V.L. Funanage, T.T. Myoda and J.L. Hamlin, "The Effect Of Gamma Rays at the Dihydrofolate Reductase Locus: Deletions and Inversions," Somatic Cell and *Molec. Genet., 12*: 555-566 (1986). A fragment of zcytor17 cDNA that includes the polynucleotide sequence from extracellular domain of the zcytor17 receptor was fused in frame to the Fc4 polynucleotide sequence (SEQ ID NO:37) to generate a zcytor17-Fc4 fusion (SEQ ID NO:38 and SEQ ID NO:39). The pEZE-2 vector is a mammalian expression vector that contains the Fc4 polynucleotide sequence and a cloning site that allows rapid construction of C-terminal Fc4 fusions using standard molecular biology techniques.

A 1566 base pair fragment was generated by PCR, containing the extracellular domain of human zcytor17 and the first two amino acids of Fc4 (Glu and Pro) with FseI and BglII sites coded on the 5' and 3' ends, respectively. This PCR fragment was generated using primers ZC29,157 (SEQ ID NO:40) and ZC29,150 (SEQ ID NO:41) by amplification from a plasmid containing the extracellular domain of human zcytor17 (pZp9zcytor17CEE) (Example 8). The PCR reaction conditions were as follows: 25 cycles of 94°C for 1 minute, 60°C for 1 minute, and 72°C for 2 minutes;

1 cycle at 72°C for 10 minutes; followed by a 4°C soak. The fragment was digested with FseI and BglII restriction endonucleases and subsequently purified by 1% gel electrophoresis and band purification using QiaQuick gel extraction kit (Qiagen). The resulting purified DNA was ligated for 5 hours at room temperature into a pEZE-2 vector previously digested with FseI and BglII containing Fc4 3' of the FseI and BglII sites.

Two μl of the ligation mix was electroporated in 37 μl DH10B electrocompetent *E. coli* (Gibco) according to the manufacturer's directions. The transformed cells were diluted in 400 μl of LB media and plated onto LB plates containing 100 μg/ml ampicillin. Clones were analyzed by restriction digests and positive clones were sent for DNA sequencing to confirm the sequence of the fusion construct. 1 μl of a positive clone was transformed into 37 μl of DH10B electrocompetent *E. coli* and streaked on a LB/amp plate. A single colony was picked from this streaked plate to start a 250 ml LB/amp culture that was then grown overnight at 37°C with shaking at 250 rpm. This culture was used to generate 750 μg of purified DNA using a Qiagen plasmid Maxi kit (Qiagen).

Example 9

Transfection And Expression Of Zcytor17 Soluble Receptor Polypeptides

BHK 570 cells (ATCC No. CRL-10314), DG-44 CHO, or other mammalian cells are plated at about $1.2X10^6$ cells/well (6-well plate) in 800 μl of appropriate serum free (SF) media (e.g., DMEM, Gibco/BRL High Glucose) (Gibco BRL, Gaithersburg, MD). The cells are transfected with expression plasmids containing zcytor17CEE, zcytor17CFLG, zcytor17CHIS or zcytor17-Fc4 (Example 8), using Lipofectin™ (Gibco BRL), in serum free (SF) media according to manufacturer's instruction. Single clones expressing the soluble receptors are isolated, screened and grown up in cell culture media, and purified using standard techniques.

A. Mammalian expression of soluble human zcytor17CEE receptor

BHK 570 cells (ATCC NO: CRL-10314) were plated in T-75 tissue culture flasks and allowed to grow to approximately 50 to 70% confluence at 37°C, 5%

CO_2, in DMEM/FBS media (DMEM, Gibco/BRL High Glucose, (Gibco BRL, Gaithersburg, MD), 5% fetal bovine serum, 1 mM L-glutamine (JRH Biosciences, Lenea, KS), 1 mM sodium pyruvate (Gibco BRL)). The cells were then transfected with the plasmid containing zcytor17CEE (Example 8) using Lipofectamine™ (Gibco BRL), in serum free (SF) media formulation (DMEM, 10 mg/ml transferrin, 5 mg/ml insulin, 2 mg/ml fetuin, 1% L-glutamine and 1% sodium pyruvate). Ten μg of the plasmid DNA pZp9zcytor17CEE (Example 8) was diluted into a 15ml tube to a total final volume of 500 μl with SF media. 50 μl of Lipofectamine was mixed with 450 μl of SF medium. The Lipofectamine mix was added to the DNA mix and allowed to incubate approximately 30 minutes at room temperature. Four ml of SF media was added to the DNA:Lipofectamine mixture. The cells were rinsed once with 5 ml of SF media, aspirated, and the DNA:Lipofectamine mixture was added. The cells were incubated at 37°C for five hours, and then 5 ml of DMEM/10%FBS media was added. The flask was incubated at 37°C overnight after which time the cells were split into the selection media (DMEM/FBS media from above with the addition of 1 μM methotrexate or 10 μM Methotrexate (Sigma Chemical Co., St. Louis, Mo.) in 150 mm plates at 1:2, 1:10, and 1:50. Approximately 10 days post-transfection, one 150 mm plate of 1 μM methotrexate resistant colonies was trypsinized, the cells were pooled, and one-half of the cells were replated in 10 μM methotrexate; to further amplify expression of the zcytor17CEE protein. A conditioned-media sample from this pool of amplified cells was tested for expression levels using SDS-PAGE and Western analysis.

B. Mammalian expression of soluble human zcytor17-Fc4 receptor

Five replicates of 200 μg of pEZE-2hzcytor17Fc4 plasmid DNA (Example 8) were linearized by restriction digestion with FspI, a restriction enzyme that cuts once within the vector and does not disturb genes necessary for expression. 200 μg of CHO cell genomic DNA was added to each replicate as carrier DNA and then the DNA was precipitated by addition of 0.1 volumes of 3M Sodium Acetate pH 5.2 and 2.2 volumes ethanol followed by a 15 minute ice incubation and microcentrifugation at 4°C. The resulting DNA pellets were washed in 70% ethanol and air dried before being resuspended in 100 μl protein free (PF) CHO non-selection growth media (21 g/L PF

CHO Ex Cell 325 /200 mM L-glutamine (Gibco)/100 mM sodium pyruvate (Gibco)/1x HT Supplement (Gibco). Ten million PF CHO passage 61 cells were added to the DNA in 600 µl of PF CHO non-selection growth media and then electroporated in a Gene Pulser II Electroporation system (BioRad) using 950 µF capacitance and 300 Kv using a 0.4 cm gap Gene Pulser (BioRad) electroporation cuvette. All 5 replicates of the electroporated cells were pooled and directly selected in –HT media (21 g/L PF CHO Ex Cell 325/ 200 mM L-glutamine (Gibco)/100 mM sodium pyruvate (Gibco). Cells were selected for 15 days in –HT media before being passaged at 4 x 10^5 ml into 50 nm MTX selection. Eight days later cells were seeded at 3.5X10^5 cells/ml into 200 mM MTX selection. After one week, cells were seeded at 4X10^5 cells/ml into 1 µM MTX selection. After two weeks at 1 µM MTX, cells were seeded at 1X10^6 cells/ml into 50 ml to generate conditioned medium. The resulting 72 hour conditioned media was analyzed by probing western blots with an antibody generated against human Ig. The cells produced hzcytor17/Fc4 protein at approximately 1 mg/L.

C. Larger-scale mammalian expression of soluble human zcytor17-Fc4 receptor

Two hundred micrograms of pEZE-2hzcytor17Fc4 plasmid DNA (Example 8) was linearized by restriction digestion with FspI, a restriction enzyme that cuts once within the pEZE-2 vector and does not disturb genes necessary for expression. 200 µg of CHO genomic DNA (prepared in-house) was added as carrier DNA and then the DNA was precipitated by addition of 0.1 volumes of 3M Sodium Acetate pH 5.2 and 2.5 volumes ethanol followed by microcentrifugation at Room temperature. Five replicate DNA pellets were made and transformed. The resulting DNA pellet was washed in 70% ethanol and air dried before being resuspended in 100 µl PF CHO non-selection growth media (21 g/L PF CHO Ex Cell 325 /200 mM L-glutamine (Gibco)/100 mM sodium pyruvate (Gibco)/1x HT Supplement (Gibco). Ten million PF CHO cells were added to the DNA in 600 µl of PF CHO non-selection growth media and then electroporated in a Gene Pulser II Electroporation system (BioRad) using 950 µF capacitance and 300 volts using a 0.4 cm gap Gene Pulser (BioRad) electroporation cuvette. The electroporated cells were pooled and put directly into selection in –HT media (21 g/L PF CHO Ex Cell 325/ 200 mM L-glutamine

(Gibco)/100 mM sodium pyruvate (Gibco). Cells were selected for 14 days in –HT media before being passaged at 4 x 10^5/ml into 50 nm MTX selection. Cells were amplified to 200nM MTX and then to 1uM MTX. The –HT, 50nM, and 1uM pools were seeded at 1 x 10^6 c/ml for 48 hours, and the resulting conditioned media was analyzed by probing western blots with an antibody generated against human Ig.

Example 10

Purification of zcytor17 soluble receptors from BHK 570 and CHO cells

A. Transient mammalian expression and purification of soluble human zcytor17-Fc4 receptor

pEZE-2hzcytor17Fc4 plasmid DNA (Example 11B) was introduced into 40 maxi plates of BHK cells using Lipofectamine (Gibco BRL) as described herein and in manufacturer's instructions. Cells were allowed to recover overnight, then were rinsed and refed with serum-free medium (SL7V4, made in-house). After 72 hours, the media was collected and filtered, and cells were refed with serum-free medium. After 72 hours, the media was again collected and filtered.

The serum-free conditioned media (2 x 1.5 L batches) from transiently transfected BHK cells was pumped over a 1.5 ml Protein A-agarose column in 20 mM Tris, pH 7.5, 0.5 M NaCl. The column was washed extensively with this buffer and then the bound protein was eluted with 1 ml of 0.2 M glycine, pH 2.5, 0.5 M NaCl. The eluted protein was collected into 0.1 ml of 2 M Tris, pH 8.5.

Aliquots were collected for SDS-polyacrylamide gel electrophoresis and the bulk zcytor17-Fc was dialyzed overnight against PBS. The soluble receptor was sterile filtered and placed in aliquots at –80°C.

B. Purification of zcytor17-Fc4

Recombinant carboxyl terminal Fc4 tagged zcytor17 (Example 8 and Example 9) was produced from transfected CHO cells. The CHO transfection was performed using methods known in the art. Approximately five-liters of conditioned media were harvested and sterile filtered using Nalgene 0.2μm filters.

Protein was purified from the filtered media by a combination of Poros

50 protein A affinity chromatography (PerSeptive Biosystems, 1-5559-01, Framingham, MA) and Superdex 200 gel exclusion chromatography column (Amersham Pharmacia Biotech, Piscataway, NJ). Culture medium was directly loaded onto a 10x70mm (5.5-ml bed volume) protein A affinity column at a flow of about 3-10 ml/minute. Following column washing for ten column volumes of PBS, bound protein was eluted by five column volumes of 0.1 M glycine, pH 3.0 at 10 ml/minute). Fractions of 2 ml each were collected into tubes containing 100μl of 2.0 M Tris, pH 8.0, in order to neutralize the eluted proteins. Samples from the affinity column were analyzed by SDS-PAGE with coomassie staining and Western blotting for the presence of zcytor17-Fc4 using human Ig-HRP. Zcytor17-Fc4-containing fractions were pooled and concentrated to 1-2 ml using Biomax-30 concentrator (Millipore), and loaded onto a 20x580 mm Superdex 200 gel filtration column. The fractions containing purified zcytor17-Fc4 were pooled, filtered through 0.2 μm filter, aliquoted into 100 μl each, and frozen at –80°C. The concentration of the final purified protein was determined by BCA assay (Pierce, Rockford, IL).

C. SDS-PAGE and Western blotting analysis of zcytor17/Fc4

Recombinant zcytor17-Fc4 was analyzed by SDS-PAGE (Nupage 4-12%, Invitrogen, Carlsbad, CA) with coomassie staining method and Western blotting using human Ig-HRP. Either the conditioned media or purified protein was electrophoresed using an Invitrogen Novex's Xcell II mini-cell, and transferred to nitrocellulose (0.2 mm; Invitrogen, Carlsbad, CA) at room temperature using Novex's Xcell II blot module with stirring according to directions provided in the instrument manual. The transfer was run at 500 mA for one hour in a buffer containing 25 mM Tris base, 200 mM glycine, and 20% methanol. The filters were then blocked with 10% non-fat dry milk in PBS for 10 minutes at room temperature. The nitrocellulose was quickly rinsed, then the human Ig-HRP antibody (1:2000) was added in PBS containing 2.5% non-fat dry milk. The blots were incubated for two hours at room temperature, or overnight at 4°C, with gentle shaking. Following the incubation, the blots were washed three times for 10 minutes each in PBS, then quickly rinsed in H_2O. The blots were developed using commercially available chemiluminescent substrate

reagents (SuperSignal® ULTRA reagents 1 and 2 mixed 1:1; reagents obtained from Pierce, Rockford, IL), and the signal was captured using Lumi-Imager's Lumi Analyst 3.0 software (Boehringer Mannheim GmbH, Germany) for exposure times ranging from 10 second to 5 minutes or as necessary.

The purified zcytor17-Fc4 appeared as a single band with either the coomassie or silver staining at about 220 kDa under non-reducing conditions, and at about 120 kDa under reducing conditions, suggesting the dimeric form of zcytor17-Fc4 under non-reducing conditions as expected.

Example 11

Assay using zcytor17 soluble receptor zcytor17-Fc4 Soluble receptor in competitive inhibition assay

BaF3/zcytor17/WSX-1/OSMRbeta cells or BaF3/zcytor17/OSMRbeta cells were spun down and washed in mIL-3 free media. The cells were spun and washed 3 times to ensure the removal of the mIL-3. Cells were then counted in a hemacytometer. Cells were plated in a 96-well format at 5000 cells per well in a volume of 100 µl per well using the mIL-3 free media.

Both conditioned media from the CCRF-CEM and CCRF-HSB2 cell activation and the human CD3+ selected cells, described in Example 5, were added in separate experiments at 25%, 12.5%, 6.25%, 3.125%, 1.5%, 0.75%, 0.375%, and 0.187 % concentrations, with or without zcytor17 soluble receptors (Zcytor17-Fc4; See, Example 9 and Example 10) at 1-10 µg/ml. The total assay volume was 200 µl.

The assay plates were incubated at 37°C, 5% CO_2 for 3-5 days at which time Alamar Blue (Accumed) was added at 20 µl/well. Plates were again incubated at 37°C, 5% CO_2 for 16-24 hours. Plates were read on the Fmax™ plate reader (Molecular Devices) as described in Example 2. Results demonstrated partial inhibition of cell growth with zcytor17-Fc4 soluble receptor at 10 µg/ml, confirming that the factor in each sample was specific for the zcytor17 receptor.

Titration curves, diluting out the soluble receptor, or soluble receptor heterodimers and trimers comprising zcytor17 receptor (e.g., zcytor17/OSMR, zcytor17/WSX-1, or zcytor17/OSMR/WSX-1, or other Class I cytokine receptor

subunits) are also run using the above stated assay to determine whether zcytor17 receptors are able to completely inhibit growth, for example, at low or physiologic concentrations.

Example 12

Secretion trap assay

A secretion trap assay is used to test the binding of the zcytor17lig to receptors comprising zcytor17 receptor, such as the zcytor17 receptor or receptor heterodimers and trimers comprising zcytor17 receptor (e.g., zcytor17/OSMR, zcytor17/WSX-1, or zcytor17/OSMR/WSX-1, or other Class I cytokine receptor subunits). Zcytor17lig plasmid DNA is transfected into COS cells, and used to assess binding of the zcytor17lig to receptors comprising zcytor17 receptor by secretion trap as described below.

A. COS Cell Transfections

The COS cell transfection is performed as follows: Mix about 800 ng of zcytor17lig cDNA and 5µl Lipofectamine™ in 92 µl serum free DMEM media (55mg sodium pyruvate, 146mg L-glutamine, 5mg transferrin, 2.5mg insulin, 1µg selenium and 5mg fetuin in 500ml DMEM), incubate at room temperature for 30 minutes and then add 400 µl serum free DMEM media. Add this 500µl mixture onto $1.5x10^5$ COS cells/well plated on 12-well tissue culture plate and incubate for 5 hours at 37°C. Add 500µl 20% FBS DMEM media (100 ml FBS, 55 mg sodium pyruvate and 146mg L-glutamine in 500ml DMEM) and incubate overnight.

B. Secretion Trap Assay

The secretion trap is performed as follows: Media is rinsed off cells with PBS and then fixed for 15 minutes with 1.8% Formaldehyde in PBS. Cells are then washed with TNT (0.1M Tris-HCL, 0.15M NaCl, and 0.05% Tween-20 in H_2O), and permeated with 0.1% Triton-X in PBS for 15 minutes, and again washed with TNT. Cells are blocked for 1 hour with TNB (0.1M Tris-HCL, 0.15M NaCl and 0.5% Blocking Reagent (NEN Renaissance TSA-Direct Kit) in H_2O), and washed again with

TNT. If using the biotinylated receptor protein, the cells are blocked for 15 minute incubations with Avidin and then Biotin (Vector Labs) washing in-between with TNT. Depending on which soluble receptor is used, the cells are incubated for 1 hour in TNB with: (A) 1-3 μg/ml zcytor17 soluble receptor zcytor17-Fc4 fusion protein (Example 10); (B) 1-3 μg/ml zcytor17/OSMRbeta soluble receptor protein; (C) 1-3 μg/ml zcytor17/WSX-1 soluble receptor protein; or (D) 1-3 μg/ml zcytor17/OSMR/WSX-1 soluble receptor protein. Cells are then washed with TNT. Depending on which soluble receptor is used (e.g., if labeled with an Fc4 tag (SEQ ID NO:37), C-terminal FLAG tag (SEQ ID NO:36), or CEE tag (SEQ ID NO:32; SEQ ID NO:35)), cells are incubated for another hour with: (A) 1:200 diluted goat-anti-human Ig-HRP (Fc specific); (B) 1:1000 diluted M2-HRP; (C) 1:1000 diluted anti-GluGlu antibody-HRP; or (D) 1:300 diluted streptavidin-HRP (NEN kit) in TNB, for example. Again cells are washed with TNT.

Positive binding is detected with fluorescein tyramide reagent diluted 1:50 in dilution buffer (NEN kit) and incubated for 4-6 minutes, and washed with TNT. Cells are preserved with Vectashield Mounting Media (Vector Labs Burlingame, CA) diluted 1:5 in TNT. Cells are visualized using a FITC filter on fluorescent microscope.

Example 13

Chromosomal Assignment and Placement of the gene sequence for the zcytor17lig

The zcytor17lig gene sequence was mapped to human chromosome 12 using the commercially available version of the "Stanford G3 Radiation Hybrid Mapping Panel" (Research Genetics, Inc., Huntsville, AL). The "Stanford G3 RH Panel" contains DNA from each of 83 radiation hybrid clones of the whole human genome, plus two control DNAs (the RM donor and the A3 recipient). A publicly available WWW server (e.g., Standford University) allows chromosomal localization of markers and genes.

For the mapping of the zcytor17lig gene sequence with the "Stanford G3 RH Panel", 20 μl reactions were set up in a 96-well microtiter plate compatible for PCR (Stratagene, La Jolla, CA) and used in a "RoboCycler Gradient 96" thermal cycler (Stratagene). Each of the 95 PCR reactions consisted of 2 μl 10X PCR reaction buffer (Qiagen, Inc., Valencia, CA), 1.6 μl dNTPs mix (2.5 mM each, PERKIN-ELMER,

Foster City, CA), 1 μl sense primer, ZC41,458 (SEQ ID NO:42), 1 μl antisense primer, ZC41,457 (SEQ ID NO:43), 2 μl "RediLoad" (Research Genetics, Inc., Huntsville, AL), 0.1 μl Qiagen HotStarTaq DNA Polymerase (5 units/μl), 25 ng of DNA from an individual hybrid clone or control and distilled water for a total volume of 20 μl. The reactions were overlaid with an equal amount of mineral oil and sealed. The PCR cycler conditions were as follows: an initial 1 cycle 15 minute denaturation at 95°C, 35 cycles of a 45 second denaturation at 95°C, 1 minute annealing at 53°C and 1 minute and 15 seconds extension at 72°C, followed by a final 1 cycle extension of 7 minutes at 72°C. The reactions were separated by electrophoresis on a 2% agarose gel (EM Science, Gibbstown, NJ) and visualized by staining with ethidium bromide.

The results showed linkage of the zcytor17lig gene sequence to the chromosome 12 marker SHGC-83339 with a LOD score of >11 and at a distance of 17 cR_10000 from the marker. This marker positions zcytor17lig gene in the 12q24.31 chromosomal region.

Example 14

Identification and cloning of murine zcytor17lig

A. Identification of full length murine zcytor17lig

Using the human zcytor17lig peptide sequence (SEQ ID NO:2) to query an in house DNA database, a murine cDNA, Genbank Accession No. AK005939, was identified as a potential partial sequence for the murine zcytor17lig. The AK005939 cDNA sequence was used to query a database containing murine genomic fragments. A genomic contig of the murine zcytor17lig was assembled (SEQ ID NO:76). Prediction of coding potential on this genomic fragment with the program Genscan revealed a likely cDNA sequence, with the same gene structure as the human zcytor17lig. A murine cDNA sequence is represented in SEQ ID NO:10, and corresponding polypeptide sequence is shown in SEQ ID NO:11.

B. Cloning of mouse zcytor17lig from a mouse testis cDNA library by PCR

Based on the genomic sequence (SEQ ID NO:76), two PCR primers were designed and used to identify a cDNA source of mouse zcytor17lig by PCR.

These Primers ZC41498 (SEQ ID NO:86) and ZC41496 (SEQ ID NO:87) were designed to the putative 5' and 3' untranslated regions of the mouse sequences (SEQ ID NO:76 and SEQ ID NO:10). Several cDNA sources were screened by PCR, including Marathon-ready cDNAs (Clontech) and aliquots of locally made cDNA libraries. Products were visualized on 1% agarose gels. Bands of the expected size were observed in reactions utilizing a mouse testis cDNA library template. These PCR reactions were successfully performed in approximately 50 μl volumes with or without 10% DMSO, using pfu turbo polymerase (Stratagene) according to the manufacturer's recommendations; with an additional application of a wax hot-start employing hot start 50s (Molecular Bioproducts, Inc. San Diego, CA). PCR thermocycling was performed with a single cycle of 94°C for 4 min; followed by 40 cycles of 94°C: 30 seconds, 48°C: 30 seconds, 72°C: 50 seconds; with additional final 72°C extension for 7 minutes. The two PCR reactions were pooled and purified using low melt agarose and Gelase agarose digesting enzyme (Epicenter, Inc. Madison, WI) according to the manufacturer's recommendations.

DNA sequence determination of these PCR products revealed a murine zcytor17 cDNA sequence (SEQ ID NO:90) which comprised an ORF identical to SEQ ID NO:10, confirming that SEQ ID NO:10 encoded the mouse zcytor17lig polypeptide. PCR primers, ZC41583 (SEQ ID NO:88) and ZC41584 (SEQ ID NO:89), were then used to add FseI and AscI restriction sites and a partial Kozak sequence to the mcytor17lig open reading frame and termination codon (SEQ ID NO:92). A Robocycler 40 thermocycler (Stratagene) was used to run a temperature gradient of annealing temperatures and cycling as follows. Pfu turbo polymerase (Stratagene) was applied as described above, but only in 10% DMSO. Cycling was performed with a single cycle of 94°C for 4 min; followed by 20 cycles of 94°C: 30 seconds, 65°C to 51°C gradient: 30 seconds, 72°C: 1 minute; and a single 72°C extension for 7 minutes. The template for this second thermocycling reaction was 1 μl of the initial gel-purified mcytor17lig PCR product, above. Resulting PCR product from the three lowest temperature reactions were pooled and gel purified using the Gelase (Epicenter) method described above. This purified mzcytor17lig was digested with FseI and AscI and ligated into a pZP7X vector modified to have FseI and AscI sites in its cloning site.

Plasmid pZP7X is a mammalian expression vector containing an expression cassette having the mouse metallothionein-1 (MT-1) promoter, multiple restriction sites for insertion of coding sequences, and a human growth hormone terminator. The plasmid also has an *E. coli* origin of replication, a mammalian selective marker expression unit having an SV40 promoter, enhancer and origin of replication, a DHFR gene, and the SV40 terminator. The cloned murine cDNA sequence is represented in SEQ ID NO:90, and corresponding polypeptide sequence is shown in SEQ ID NO:91 (which is identical to SEQ ID NO:11).

Example 15

Isolation of mouse zcytor17lig cDNA clone from an activated mouse spleen library

A. Murine Primary Source used to isolate mouse zcytor17lig

Mouse spleens from Balb/C mice, are collected and mashed between frosted-end slides to create a cell suspension. The isolated primary mouse cell yield is expected to be about $6.4X10^8$ cells prior to selection described below.

The spleen cells are suspended in 9.6 ml MACS buffer (PBS, 0.5% EDTA, 2mM EDTA). 1.6 ml of cell suspension is removed and 0.4 ml CD90 (Thy1.2) microbeads (Miltenyi Biotec) added. The mixture is incubated for 15 min. at 4°C. These cells labeled with CD90 beads are washed with 30 ml MACS buffer, and then resuspended in 2 ml MACS buffer.

A VS+ column (Miltenyi) is prepared according to the manufacturer's instructions. The VS+ column is then placed in a VarioMACS™ magnetic field (Miltenyi). The column is equilibrated with 5 ml MACS buffer. The isolated primary mouse cells are then applied to the column. The CD90 negative cells are allowed to pass through. The column is rinsed with 9 ml (3 X 3 ml) MACS buffer. The column is then removed from the magnet and placed over a 15 ml falcon tube. CD90+ cells are eluted by adding 5 ml MACS buffer to the column and bound cells flushed out using the plunger provided by the manufacturer. The incubation of the cells with the CD90 magnetic beads, washes, and VS+ column steps (incubation through elution) above are repeated once more. The resulting CD90+ fractions from the 2 column separations are pooled. The yield of CD90+ selected mouse spleen cells are expected to be about

$1X10^8$ total cells.

A sample of the pooled CD90+ selected mouse cells is removed for staining and sorting on a fluorescent antibody cell sorter (FACS) to assess their purity. A PE-conjugated hamster anti-mouse CD3ε antibody (PharMingen) is used for staining and sorting the CD90+ selected cells. The mouse CD90+ selected cells should be about 93% CD3+ cells, suggesting the cells are 93% T-cells.

The murine CD90+ selected cells are activated by incubating $3X10^6$ cells/ml in RPMI + 5% FBS + PMA 10 ng/ml and Ionomycin 0.5 μg/ml (Calbiochem) for overnight at 37°C. The supernatant from these activated CD90+ selected mouse cells is tested for zcytor17lig activity as described below. Moreover, the activated CD90+ selected mouse cells are used to prepare a cDNA library, as described in Example 16, below.

Example 16

Cloning of mouse zcytor17lig from a mouse CD90+ selected cell library

Screening of a primary mouse activated CD90+ selected cell cDNA library can reveal isolated cDNA that is a novel member of the four-helix bundle cytokine family that would encode the mouse ortholog of the human zcytor17lig. The cDNA is identified by hybridization screening.

A. The vector for CD90+ selected library construction

The vector, pZP7N is used for CD3+ selected library construction (See Example 6A)

B. Preparation of primary mouse activated CD90+ selected cell cDNA library

Approximately $1.5X10^8$ primary mouse CD90+ selected cells stimulated in ionomycin/PMA (Example 15) are isolated by centrifugation. Total RNA is isolated from the cell pellet, and converted to double stranded cDNA as described in Example 6B. This DNA is subsequently transfected into BHK cells, as described in Example 6B, and proliferation is assessed using an "Alamar blue" fluorescence assay (Example 2B).

For the purpose of screening the library by secretion trap cloning, a

complex, amplified form of the library is needed to transfect COS-7 cells. 4.8 million clones are plated on 110 15cm LB-agar plates supplemented with 100 μg/ml ampicillin, 10 μg/ml methicillin. After growing the plates overnight at 37°C the bacteria are harvested by scraping and pelleted. Plasmid DNA is extracted from the pelleted bacteria using a Nucleobond-giga™ (Clonetech) following the manufacturer's instructions. This plasmid is then used to transfect COS-7 cells on slides and screened using the secretion trap technique described below (Example 17).

C. Screening the activated mouse cDNA library

Approximately $5X10^5$ clones are plated on 10 LB/Amp Maxi plates. The colonies are lifted, denatured, neutralized, and cross-linked using the standard procedure (Sambrook, J. et al. *supra.*). Fifty nanograms of the 300 bp 5' RACE PCR fragment (Example 14) is labeled with ^{32}P using Prime-Itr RmT random primer labeling kit (Stratagene). The 10 filters are hybridized with this labeled probe at 65°C overnight using ExpressHyb™ Hybridization Solution (Clontech). The filters are then washed sequentially at 60°C for 1 hour three times with 0.2xSSC (30 mM NaCl, 3 mM sodium citrate, pH 7.0), 0.1% SDS; and then at 65°C for 1 hour. The filters are exposed at -80°C overnight, and the X-ray film are developed. Agar plugs containing the positive colonies are pulled, and the clones plated on 10-cm LB/Amp plates. The colonies are then filter-lifted and hybridized again following the same procedure described above. Single DNA clones are isolated and sequenced using standard methods, to identify the mouse cDNA.

Example 17

Mouse zcytor17lig does not bind to human zcytor17 soluble receptor in secretion trap assay

The DNA for mouse clone mzcytor17lig/pZP7 was transfected into COS cells, and the binding of zcytor17 comprising soluble receptors (human zcytor17 soluble receptor zcytor17-Fc4 (Example 10), or soluble receptor heterodimers (zcytor17/WSX-1 or BaF3/zcytor17/OSMRbeta), to the transfected COS cells were tested by a secretion trap assay (Example 12). The assay confirmed that the mouse

zcytor17lig does not bind to human zcytor17 soluble receptor.

The COS cell transfection was performed as per Example 12 using about 0.7 μg mouse zcytor17lig cDNA (Example 16) in 3 μl.

The secretion trap was performed as per example 12 using, for example, 1 μg/ml zcytor17 soluble receptor Fc4 fusion protein (Example 10) (or zcytor17 comprising soluble receptor heterodimers as described herein) in TNB, and 1:200 diluted goat-anti-human Ig-HRP (Fc specific) in TNB for the detectable antibody. Positive binding of the soluble human zcytor17 receptor to the prepared fixed cells was not detected with fluorescein tyramide reagent as per Example 12. Cells were preserved and visualized according to Example 12.

Results indicated that the mouse zcytor17lig does not bind to human zcytor17 soluble receptor (or zcytor17 comprising soluble receptor heterodimers as described herein).

Example 18

Expression of mouse zcytor17lig in mammalian cells

Mammalian expression of mouse zcytor17lig

BHK 570 cells (ATCC No: CRL-10314) were plated in 10 cm tissue culture dishes and allowed to grow to approximately 20% confluence overnight at 37°C, 5% CO_2, in DMEM/FBS media (DMEM, Gibco/BRL High Glucose media; Gibco BRL, Gaithersburg, MD), 5% fetal bovine serum (Hyclone, Logan, UT), 1 mM L-glutamine (JRH Biosciences, Lenexa, KS), 1 mM sodium pyruvate (Gibco BRL). The cells were then transfected with the plasmid mzcytor17lig/pZP7X (Example 14) using a mammalian stable Lipofectamine (GibcoBRL) transfection kit according to the manufacturer's instructions.

One day after transfection, the cells were split 1:10 and 1:20 into the selection media (DMEM/FBS media with the addition of 1 μM methotrexate (Sigma Chemical Co., St. Louis, MO)) in 150 mm plates. The media on the cells was replaced with fresh selection media at day 5 post-transfection. Approximately 10 days post-transfection, methotrexate resistant colonies were trypsinized and the cells pooled and plated into large-scale culture flasks. Once the cells were grown to approximately 90%

confluence, they were rinsed with PBS three times, and cultured with serum-free ESTEP2 media (DMEM (Gibco BRL), 0.11 g/l Na Pyruvate, 3.7 g/l $NaHCO_3$, 2.5 mg/l insulin, 5 mg/l transferrin, pH7.0) conditioned media. The conditioned media was collected three days later, and put into a BaF3 proliferation assay using Alamar Blue, described in Example 19 below.

Example 19

Mouse zcytor17lig does not activate human zcytor17 receptor in BaF3 assay using Alamar Blue

Proliferation of BaF3/zcytor17, BaF3/zcytor17/OSMRbeta and BaF3/zcytor17/WSX-1 cells (Example 4, and 5B) was assessed using serum-free conditioned media from BHK cells expressing mouse zcytor17lig (Example 18). BaF3/Zcytor17, BaF3/zcytor17/OSMRbeta and BaF3/zcytor17/WSX-1 cells were spun down, washed and plated in mIL-3 free media as described in Example 5B. Conditioned media from BHK cells expressing mouse zcytor17lig (Example 18) was diluted with mIL-3 free media to 50%, 25%, 12.5%, 6.25%, 3.125%, 1.5%, 0.75% and 0.375% concentrations. The proliferation assay was performed as per Example 5B. The results of this assay were negative, indicating that mouse zcytor17lig does not activate human zcytor17, zcytor17/OSMRbeta, or zcytor17/WSX-1 receptor complexes.

Example 20

Human zcytor17lig activates human, zcytor17/OSMRbeta receptor, in luciferase assay

A. Construction of BaF3/KZ134/zcytor17 cell line

The KZ134 plasmid was constructed with complementary oligonucleotides ZC12,749 (SEQ ID NO:44) and ZC12,748 (SEQ ID NO:45) that contain STAT transcription factor binding elements from 4 genes, which includes a modified c-fos Sis inducible element (m67SIE, or hSIE) (Sadowski, H. et al., *Science 261*:1739-1744, 1993), the p21 SIE1 from the p21 WAF1 gene (Chin, Y. et al., *Science 272*:719-722, 1996), the mammary gland response element of the β-casein gene (Schmitt-Ney, M. et al., *Mol. Cell. Biol. 11*:3745-3755, 1991), and a STAT inducible element of the Fcg RI gene, (Seidel, H. et al., *Proc. Natl. Acad. Sci. 92*:3041-3045,

1995). These oligonucleotides contain Asp718-XhoI compatible ends and were ligated, using standard methods, into a recipient firefly luciferase reporter vector with a c-fos promoter (Poulsen, L.K. et al., *J. Biol. Chem. 273*:6229-6232, 1998) digested with the same enzymes and containing a neomycin selectable marker. The KZ134 plasmid was used to stably transfect BaF3 cells, using standard transfection and selection methods, to make the BaF3/KZ134 cell line.

A stable BaF3/KZ134 indicator cell line, expressing the full-length zcytor17 receptor or zcytor17/OSMRbeta receptor was constructed as per Example 4. Clones were diluted, plated and selected using standard techniques. Clones were screened by luciferase assay (see Example 20B, below) using the human zcytor17lig conditioned media or purified zcytor17lig protein (see Example 35, below) as an inducer. Clones with the highest luciferase response (via STAT luciferase) and the lowest background were selected. Stable transfectant cell lines were selected. The cell lines were called BaF3/KZ134/zcytor17 or BaF3/KZ134/zcytor17/OSMRbeta depending on the receptors transfected into the cell line.

Similarly, BHK cell lines were also constructed using the method described herein, and were used in luciferase assays described herein. The cell lines were called BHK/KZ134/zcytor17 or BHK/KZ134/zcytor17/OSMRbeta depending on the receptors transfected into the cell line.

B. Human zcytor17lig activates human zcytor17 receptor in BaF3/KZ134/zcytor17/OSMRbeta or BHK/KZ134/zcytor17/OSMRbeta luciferase assay

BaF3/KZ134/zcytor17 and BaF3/KZ134/zcytor17/OSMRbeta cells were spun down and washed in mIL-3 free media. The cells were spun and washed 3 times to ensure removal of mIL-3. Cells were then counted in a hemacytometer. Cells were plated in a 96-well format at about 30,000 cells per well in a volume of 100 µl per well using the mIL-3 free media. The same procedure was used for untransfected BaF3/KZ134 cells for use as a control in the subsequent assay. BHK/KZ134/zcytor17 or BHK/KZ134/zcytor17/OSMRbeta cells were plated in a 96-well format at 15,000 cells per well in 100 µl media. Parental BHK/KZ134 cells were used as a control.

STAT activation of the BaF3/KZ134/Zcytor17,

BaF3/KZ134/zcytor17/OSMRbeta, BHK/KZ134/zcytor17, or BHK/KZ134/zcytor17/OSMRbeta cells was assessed using (1) conditioned media from BHK570 cells transfected with the human zcytor17lig (Example 7), (2) conditioned media from BHK570 cells transfected with the mouse zcytor17lig (Example 18), (3) purified human zcytor17lig (Example 35), or (4) mIL-3 free media to measure media-only control response. Conditioned media was diluted with RPMI mIL-3 free media to 50%, 25%, 12.5%, 6.25%, 3.125%, 1.5%, 0.75% and 0.375% concentrations. Purified human zcytor17lig was diluted to a concentration of 1200, 600, 300, 150, 75, 37.5, 18.75, or 9.4 pM. 100 µl of the diluted conditioned media or protein was added to the BaF3/KZ134/Zcytor17, BaF3/KZ134/zcytor17/OSMRbeta, BHK/KZ134/zcytor17, or BHK/KZ134/zcytor17/OSMRbeta cells. The assay using the conditioned media was done in parallel on untransfected BaF3/KZ134 or BHK/KZ134 cells as a control. The total assay volume was 200 µl. The assay plates were incubated at 37°C, 5% CO_2 for 24 hours at which time the BaF3 cells were pelleted by centrifugation at 2000 rpm for 10 min., and the media was aspirated and 25 µl of lysis buffer (Promega) was added. For the BHK cell lines, the centrifugation step was not necessary as the cells are adherant. After 10 minutes at room temperature, the plates were measured for activation of the STAT reporter construct by reading them on a luminometer (Labsystems Luminoskan, model RS) which added 40 µl of luciferase assay substrate (Promega) at a five second integration.

The results of this assay confirmed that the STAT reporter response of the BaF3/KZ134/zcytor17/OSMRbeta and BHK/KZ134/zcytor17/OSMRbeta cells to the human zcytor17lig when compared to either the BaF3/KZ134/zcytor17 cells, the BHK/KZ134/zcytor17 cells or the untransfected BaF3/KZ134 or BHK/KZ134 control cells, showed that the response was mediated through the zcytor17/OSMRbeta receptors. The results also showed that the mouse zcytor17lig does not activate the STAT reporter assay through the human receptor complex.

Example 21

Mouse zcytor17lig is active in mouse bone marrow assay

A. Isolation of Non-adherent Low Density Marrow Cells:

Fresh mouse femur aspirate (marrow) is obtained from 6-10 week old male Balb/C or C57BL/6 mice. The marrow is then washed with RPMI+10% FBS (JRH, Lenexa KS; Hyclone, Logan UT) and suspended in RPMI+10% FBS as a whole marrow cell suspension. The whole marrow cell suspension is then subjected to a density gradient (Nycoprep, 1.077, Animal; Gibco BRL) to enrich for low density, mostly mononuclear, cells as follows: The whole marrow cell suspension (About 8 ml) is carefully pipeted on top of about 5 ml Nycoprep gradient solution in a 15 ml conical tube, and then centrifuged at 600X g for 20 minutes. The interface layer, containing the low density mononuclear cells, is then removed, washed with excess RPMI+10% FBS, and pelleted by centrifugation at 400X g for 5-10 minutes. This pellet is resuspended in RPMI +10% FBS and plated in a T-75 flask at approximately 10^6 cells/ml, and incubated at 37°C 5% CO_2 for approximately 2 hours. The resulting cells in suspension are Non-Adherent Low Density (NA LD) Marrow Cells.

B. 96-Well Assay

NA LD Mouse Marrow Cells are plated at 25,000 to 45,000 cells/well in 96 well tissue culture plates in RPMI +10% FBS + 1ng/mL mouse Stem Cell Factor (mSCF) (R&D Systems, Minneapolis, MN), plus 5% conditioned medium from one of the following: (1) BHK 570 cells expressing mouse zcytor17lig (Example 18), (2) BHK 570 cells expressing human zcytor17lig (Example 7), or (3) control BHK 570 cells containing vector and not expressing either Ligand. These cells are then subjected to a variety of cytokine treatments to test for expansion or differentiation of hematopoietic cells from the marrow. For testing, the plated NA LD mouse marrow cells are subjected to human Interleukin-15 (hIL-15) (R&D Systems), or one of a panel of other cytokines (R&D Systems). Serial dilution of hIl-15, or the other cytokines, are tested, with 2-fold serial dilution from about 50 ng/ml down to about 0.5 ng/ml concentration. After 8 to 12 days the 96-well assays are scored for cell proliferation by Alamar blue assay as described in Example 5B.

C. Results from the 96-well NA LD Mouse Marrow assay

Conditioned media from the BHK cells expressing both mouse and

human zcytor17lig can promote the expansion of a population of hematopoietic cells either alone or in synergy with other cytokines in the NA LD mouse marrow in comparison to control BHK conditioned medium. The population hematopoietic cells expanded by the mouse zcytor17lig with or without other cytokines, and those hematopoietic cells expanded by the human zcytor17lig with or without other cytokines, are further propagated in cell culture. These hematopoietic cells are stained with a Phycoerythrin labeled anti-Pan NK cell antibody (PharMingen) and subjected to flow cytometry analysis, which demonstrated that the expanded cells stained positively for this natural killer (NK) cell marker. Similarly, other specific hematopoietic cell markers can be used to determine expansion of, for example, CD4+ or CD8+ T-cells, other T-cell populations, B-cells, and other immune cell markers.

The same 96-well assay is run, using fresh human marrow cells bought from Poietic Technologies, Gaithersburg, MD. Again, a positive result shows that zcytor17lig alone or in synergy with other cytokines, the mouse and human zcytor17lig can expand a hematopoietic cell population that is stained positively for specific cell markers as disclosed above.

Example 22

Constructs for generating zcytor17lig Transgenic Mice

A. Construct for expressing human zcytor17lig from the MT-1 promoter

Oligonucleotides are designed to generate a PCR fragment containing a consensus Kozak sequence and the human zcytor17lig coding region. These oligonucleotides are designed with an FseI site at the 5' end and an AscI site at the 3' end to facilitate cloning into (a) pMT12-8, our standard transgenic vector, or (b) pKFO51, a lymphoid-specific transgenic vector (Example 22B).

PCR reactions are carried out with about 200 ng human zcytor17lig template (SEQ ID NO:1) and oligonucleotides designed to amplify the full-length or active portion of the zcytor17lig. PCR reaction conditions are determined using methods known in the art. PCR products are separated by agarose gel electrophoresis and purified using a QiaQuick™ (Qiagen) gel extraction kit. The isolated, correct sized DNA fragment is digested with FseI and AscI (Boerhinger-Mannheim), ethanol

precipitated and ligated into pMT12-8 previously digested with FseI and AscI. The pMT12-8 plasmid, designed for expressing a gene of interest in liver and other tissues in transgenic mice, contains an expression cassette flanked by 10 kb of MT-1 5' DNA and 7 kb of MT-1 3' DNA. The expression cassette comprises the MT-1 promoter, the rat insulin II intron, a polylinker for the insertion of the desired clone, and the human growth hormone (hGH) poly A sequence.

About one microliter of each ligation reaction is electroporated into DH10B ElectroMax™ competent cells (GIBCO BRL, Gaithersburg, MD) according to manufacturer's direction and plated onto LB plates containing 100 μg/ml ampicillin, and incubated overnight. Colonies are picked and grown in LB media containing 100 μg/ml ampicillin. Miniprep DNA is prepared from the picked clones and screened for the human zcytor17lig insert by restriction digestion with EcoRI alone, or FseI and AscI combined, and subsequent agarose gel electrophoresis. Maxipreps of the correct pMT-human zcytor17lig are performed. A SalI fragment containing with 5' and 3' flanking sequences, the MT-1 promoter, the rat insulin II intron, human zcytor17lig cDNA and the hGH poly A sequence is prepared to be used for microinjection into fertilized murine oocytes. Microinjection and production of transgenic mice are produced as described in Hogan, B. et al. *Manipulating the Mouse Embryo*, 2nd ed., Cold Spring Harbor Laboratory Press, NY, 1994.

B. Construct for expressing human zcytor17lig from the lymphoid-specific EμLCK promoter

Oligonucleotides are designed to generate a PCR fragment containing a consensus Kozak sequence and the human zcytor17lig coding region. These oligonucleotides are designed with an FseI site at the 5' end and an AscI site at the 3' end to facilitate cloning into pKFO51, a lymphoid-specific transgenic vector.

PCR reactions are carried out with about 200 ng human zcytor17lig template (SEQ ID NO:1) and oligonucleotides designed to amplify the full-length or active portion of the zcytor17lig. A PCR reaction is performed using methods known in the art. The isolated, correct sized DNA fragment is digested with FseI and AscI (Boerhinger-Mannheim), ethanol precipitated and ligated into pKFO51 previously

digested with FseI and AscI. The pKFO51 transgenic vector is derived from p1026X (Iritani, B.M., et al., *EMBO J. 16*:7019-31, 1997) and contains the T cell-specific lck proximal promoter, the B/T cell-specific immunoglobulin μ heavy chain enhancer, a polylinker for the insertion of the desired clone, and a mutated hGH gene that encodes an inactive growth hormone protein (providing 3' introns and a polyadenylation signal).

About one microliter of each ligation reaction is electroporated, plated, clones picked and screened for the human zcytor17lig insert by restriction digestion as described above. A correct clone of pKFO51-zcytor17lig is verified by sequencing, and a maxiprep of this clone is performed. A NotI fragment, containing the lck proximal promoter and immunoglobulin μ enhancer (EμLCK), zcytor17lig cDNA, and the mutated hGH gene is prepared to be used for microinjection into fertilized murine oocytes.

C. Construct for expressing mouse zcytor17lig from the EF1alpha promoter

Primers ZC41,498 (SEQ ID NO:86) and ZC41,496 (SEQ ID NO:87) were used to PCR a mouse testis cDNA library template. These PCR reactions were successfully performed in approximately 50 μl volumes with or without 10% DMSO, using pfu turbo polymerase (Stratagene) according to the manufacturer's recommendations; with an additional application of a wax hot-start employing hot start 50s (Molecular Bioproducts, Inc. San Diego, CA). PCR thermocycling was performed with a single cycle of 94°C for 4 min; followed by 40 cycles of 94°C: 30 seconds, 48°C: 30 seconds, 72°C: 50 seconds; with additional final 72°C extension for 7 minutes. The two PCR reactions were pooled and purified using low melt agarose and Gelase agarose digesting enzyme (Epicenter, Inc. Madison, WI) according to the manufacturer's recommendations.

DNA sequenced PCR products revealed a murine zcytor17 cDNA sequence (SEQ ID NO:90) which comprised an ORF identical to SEQ ID NO:10, confirming that SEQ ID NO:10 encoded the mouse zcytor17lig polypeptide. PCR primers, ZC41583 (SEQ ID NO:88) and ZC41584 (SEQ ID NO:89), were then used to add FseI and AscI restriction sites and a partial Kozak sequence to the mcytor17lig open reading frame and termination codon (SEQ ID NO:92). A Robocycler 40

thermocycler (Stratagene) was used to run a temperature gradient of annealing temperatures and cycling as follows. Pfu turbo polymerase (Stratagene) was applied as described above, but only in 10% DMSO. Cycling was performed with a single cycle of 94°C for 4 min; followed by 20 cycles of 94°C: 30 seconds, 65°C to 51°C gradient: 30 seconds, 72°C: 1 minute; and a single 72°C extension for 7 minutes. The template for this second thermocycling reaction was 1 μl of the initial gel-purified mcytor17lig PCR product, above. Resulting PCR product from the three lowest temperature reactions were pooled and gel purified using the Gelase (Epicenter) method described above. This purified fragment was then digested with FseI and AscI and ligated into a pZP7X vector modified to have FseI and AscI sites in its cloning site. This was sent to sequencing to confirm the correct sequence. The cloned murine cDNA sequence is represented in SEQ ID NO:90, and corresponding polypeptide sequence is shown in SEQ ID NO:91 (which is identical to SEQ ID NO:11).

The isolated, correct sized DNA fragment digested with FseI and AscI (Boerhinger-Mannheim) was subcloned into a plasmid containing EF1alpha promoter previously digested with FseI and AscI. Maxipreps of the correct EF1alpha mouse zcytor17lig were performed. The expression cassette contains the EF1alpha promoter (with a deleted FseI site), the EF1alpha intron, SUR IRES like site to facilitate expression, a polylinker flanked with rat insulin II sites on the 5'end which adds FseI PmeI AscI sites for insertion of the desired clone, and the human growth hormone (hGH) poly A sequence. A 7.5kb NotI fragment containing the EF1alpha promoter expression cassette and mouse zcytor17lig was prepared to be used for microinjection into fertilized murine oocytes. The EF1alpha plsdmid was obtained from Louis-Marie of the Laboratoire de Differenciation Cellulaire, as described in Taboit-Dameron et al., 1999, *Transgenic Research 8*:223-235.

D. Construct for expressing mouse zcytor17lig from the lymphoid-specific EμLCK promoter

Oligonucleotides were designed to generate a PCR fragment containing a consensus Kozak sequence and the mouse zcytor17lig coding region. These oligonucleotides were designed with an FseI site at the 5' end and an AscI site at the 3'

end to facilitate cloning into pKFO51 (see Example 22B, above).

The isolated, correct sized zcytor17lig DNA fragment used in EF1alpha constructs, digested with FseI and AscI (Boerhinger-Mannheim), was subcloned into a plasmid containing pKFO51, a lymphoid-specific transgenic vector. The pKFO51 transgenic vector is derived from p1026X (Iritani, B.M., et al., *EMBO J. 16*:7019-31, 1997) and contains the T cell-specific lck proximal promoter, the B/T cell-specific immunoglobulin μ heavy chain enhancer, a polylinker for the insertion of the desired clone, and a mutated hGH gene that encodes an inactive growth hormone protein (providing 3' introns and a polyadenylation signal). A 6.5kb NotI fragment, containing the lck proximal promoter and immunoglobulin μ enhancer (EμLCK), mouse zcytor17lig cDNA, and the mutated hGH gene was prepared to be used for microinjection into fertilized murine oocytes (Example 41).

Example 23

Construction of mammalian expression vectors that express zcytor17lig-CEE

A. Construction of zCytor17Lig-CEE/pZMP21

An expression plasmid containing all or part of a polynucleotide encoding human zCytor17lig was constructed via homologous recombination. The plasmid was called zCytor17Lig-CEE/pZMP21.

The construction of zCytor17Lig-CEE/pZMP21 was accomplished by generating a zCytor17Lig-CEE fragment (SEQ ID NO:95) using PCR amplification. The DNA template used for the production of the zCytor17Lig-CEE fragment was zCytor17Lig/pZP7nx. The primers used for the production of the zCytor17Lig-CEE fragment were: (1) ZC41607 (SEQ ID NO:97) (sense sequence), which includes from the 5' to the 3' end: 28bp of the vector flanking sequence (5' of the insert) and 21 bp corresponding to the 5' sequence of zCytor17Lig; and (2) ZC41605 (SEQ ID NO:98) (anti-sense sequence), which includes from the 5' to the 3' end: 37 bp of the vector flanking sequence (3' of the insert), 3 bp of the stop codon, 21 bp encoding a C-terminal EE tag, and 21 bp corresponding to the 3' end of zCytor17Lig sequence. The fragment resulting from the above PCR amplification is a copy of the template zCytor17Lig with the addition of a C-terminal EE tag, yielding a final product

zCytor17Lig-CEE.

PCR reactions were run as follows: To a 100μl final volume was added: 10μl of 10x Taq Polymerase Reaction Buffer with 15mM MgCl (Gibco), 1μl of Taq DNA Polymerase (5 units/μl, Gibco), 3μl of 10mM dNTPs, 78μl dH2O, 3μl of a 20 pmol/μl stock of primer ZC41607 (SEQ ID NO:97) 3μl of a 20 pmol/μl stock of primer ZC41605 (SEQ ID NO:98), and 2μl of a 0.13μg/μl stock of zCytor17lig template DNA. A volume equal to 50μl of mineral oil was added to the mixture. The reaction was heated to 94°C for 5 minutes, followed by 35 cycles at 94°C for 1 minute; 55°C for 2 minutes; 72°C for 3 minutes; followed by a 10 minute extension at 72°C and held at 4°C until the reaction was collected.

The plasmid pZMP21 was restriction digested with BglII enzyme, cleaned with a QiaQuick PCR Purification Kit (Qiagen) using a microcentrifuge protocol, and used for recombination with the PCR fragment. Plasmid pZMP21 was constructed from pZMP20 which was constructed from pZP9 (deposited at the American Type Culture Collection, 10801 University Boulevard, Manassas, VA 20110-2209, and is designated No. 98668) with the yeast genetic elements taken from pRS316 (deposited at the American Type Culture Collection, 10801 University Boulevard, Manassas, VA 20110-2209, and designated No. 77145), an IRES element from poliovirus, and the extracellular domain of CD8, truncated at the carboxyl terminal end of the transmembrane domain. PZMP21 is a mammalian expression vector containing an expression cassette having the MPSV promoter, immunoglobulin signal peptide intron, multiple restriction sites for insertion of coding sequences, a stop codon and a human growth hormone terminator. The plasmid also has an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a DHFR gene, the SV40 terminator, as well as the URA3 and CEN-ARS sequences required for selection and replication in *S. cerevisiae*.

Fifty microliters of competent yeast cells (*S. cerevisiae*) were independently combined with 100 ng of cut plasmid, 5 μl of previously described PCR mixture, and transferred to a 0.2 cm electroporation cuvette. The yeast/DNA mixture was electropulsed at 0.75 kV (5 kV/cm), ∞ ohms, 25 μF. Each cuvette had 600 μl of 1.2 M sorbitol added, and the yeast was plated in one 100 μl aliquot and one 300 μl

aliquot onto two URA-D plates and incubated at 30°C. After about 72 hours, the Ura+ yeast transformants from a single plate were resuspended in 1 ml H_2O and spun briefly to pellet the yeast cells. The cell pellet was resuspended in 500 μl of lysis buffer (2% Triton X-100, 1% SDS, 100 mM NaCl, 10 mM Tris, pH 8.0, 1 mM EDTA). The 500 μl of the lysis mixture was added to an Eppendorf tube containing 300 μl acid washed 600μm glass beads and 300 μl phenol-chloroform, vortexed for 1 minute intervals two or three times, followed by a 5 minute spin in a Eppendorf centrifuge at maximum speed. Three hundred microliters of the aqueous phase was transferred to a fresh tube, and the DNA precipitated with 600 μl 100% ethanol (EtOH), followed by centrifugation for 10 minutes at 4°C. The DNA pellet was then washed with 500μl 70% EtOH, followed by centrifugation for 1 minute at 4°C. The DNA pellet was resuspended in 30 μl H_2O.

Transformation of electrocompetent *E. coli* cells (MC1061) was done with 5 μl of the yeast DNA prep and 50 μl of MC1061 cells. The cells were electropulsed at 2.0 kV, 25 μF and 400 ohms(Ω). Following electroporation, 600μl SOC (2% Bacto‘ Tryptone (Difco, Detroit, MI), 0.5% yeast extract (Difco), 10 mM NaCl, 2.5 mM KCl, 10 mM $MgCl_2$, 10 mM $MgSO_4$, 20 mM glucose) was added. The electroporated *E.coli* cells were plated in a 200μl and a 50μl aliquot on two LB AMP plates (LB broth (Lennox), 1.8% Bacto Agar (Difco), 100 mg/L Ampicillin). The plates were incubated upside down for about 24 hours at 37°C. Three Ampicillin-resistant colonies were selected at random and submitted for sequence analysis of the insert. Large scale plasmid DNA was isolated from a sequence-confirmed clone using the Qiagen Maxi kit (Qiagen) according to manufacturer's instructions.

B. Mammalian Expression of human zcytor17lig

Full-length zCytor17Lig protein was produced in BHK cells transfected with zCytor17Lig-CEE/pZMP21 (Example 23A). BHK 570 cells (ATCC CRL-10314) were plated in T75 tissue culture flasks and allowed to grow to approximately 50 to 70% confluence at 37°C, 5% CO_2, in growth media (SL7V4, 5%FBS, 1% pen/strep). The cells were then transfected with zCytor17Lig-CEE/pZMP21 by liposome-mediated transfection (using Lipofectamine™; Life Technologies), in serum free (SF) media

(SL7V4). The plasmid (16μg) was diluted into 1.5 ml tubes to a total final volume of 640 μl with SF media. Thirty-five microliters of the lipid mixture was mixed with 605 μl of SF medium, and the resulting mixture was allowed to incubate approximately 15 minutes at room temperature. Five milliliters of SF media was then added to the DNA:lipid mixture. The cells were rinsed once with 10 ml of PBS, the PBS was decanted, and the DNA:lipid mixture was added. The cells are incubated at 37°C for five hours, then 15 ml of media (SL7V4, 5% FBS, 1% pen/strep) was added to each plate. The plates were incubated at 37°C overnight, and the DNA:lipid media mixture was replaced with selection media (SL7V4, 5% FBS, 1% pen/strep, 1μM methotrexate) the next day. Approximately 10 days post-transfection, methotrexate-resistant colonies from the T75 transfection flask were trypsinized, and the cells were pooled and plated into a T-162 flask and transferred to large-scale culture.

Example 24

Expression of zcytor17 Soluble Receptor in E. coli

A. Construction of expression vector pCMH01 that expresses huzcytor17/MBP-6H fusion polypeptide

An expression plasmid containing a polynucleotide encoding a zcytor17 soluble receptor fused C-terminally to maltose binding protein (MBP) was constructed via homologous recombination. The fusion polypeptide contains an N-terminal approximately 388 amino acid MBP portion fused to any of the zcytor17 soluble receptors described herein. A fragment of zcytor17 cDNA (SEQ ID NO:4) was isolated using PCR as described herein. Two primers were used in the production of the zcytor17 fragment in a standard PCR reaction: (1) one containing about 40 bp of the vector flanking sequence and about 25 bp corresponding to the amino terminus of the zcytor17, and (2) another containing about 40 bp of the 3' end corresponding to the flanking vector sequence and about 25 bp corresponding to the carboxyl terminus of the zcytor17. Two μl of the 100 μl PCR reaction was run on a 1.0% agarose gel with 1 x TBE buffer for analysis, and the expected approximately fragment was seen. The remaining PCR reaction was combined with the second PCR tube and precipitated with 400 μl of absolute ethanol. The precipitated DNA used for recombining into the Sma1

cut recipient vector pTAP170 to produce the construct encoding the MBP-zcytor17 fusion, as described below.

Plasmid pTAP170 was derived from the plasmids pRS316 and pMAL-c2. The plasmid pRS316 is a *Saccharomyces cerevisiae* shuttle vector (Hieter P. and Sikorski, R., *Genetics 122*:19-27, 1989). pMAL-C2 (NEB) is an *E. coli* expression plasmid. It carries the *tac* promoter driving *MalE* (gene encoding MBP) followed by a His tag, a thrombin cleavage site, a cloning site, and the *rrnB* terminator. The vector pTAP170 was constructed using yeast homologous recombination. 100ng of EcoR1 cut pMAL-c2 was recombined with 1μg Pvu1 cut pRS316, 1μg linker, and 1μg Sca1/EcoR1 cut pRS316. The linker consisted of oligos zc19,372 (SEQ ID NO:172) (100pmole): zc19,351 (SEQ ID NO:173) (1pmole): zc19,352 (SEQ ID NO:174) (1pmole), and zc19,371 (SEQ ID NO:175) (100pmole) combined in a PCR reaction. Conditions were as follows: 10 cycles of 94°C for 30 seconds, 50°C for 30 seconds, and 72°C for 30 seconds; followed by 4°C soak. PCR products were concentrated via 100% ethanol precipitation.

One hundred microliters of competent yeast cells (*S. cerevisiae*) were combined with 10 μl of a mixture containing approximately 1 μg of the human zcytor17 insert, and 100 ng of SmaI digested pTAP170 vector, and transferred to a 0.2 cm electroporation cuvette. The yeast/DNA mixture was electropulsed at 0.75 kV (5 kV/cm), infinite ohms, 25 μF. To each cuvette was added 600 μl of 1.2 M sorbitol. The yeast was then plated in two 300 μl aliquots onto two -URA D plates and incubated at 30°C.

After about 48 hours, the Ura+ yeast transformants from a single plate were picked, DNA was isolated, and transformed into electrocompetent *E. coli* cells (e.g., MC1061, Casadaban et. al. *J. Mol. Biol. 138*, 179-207), and plated on MM/CA +KAN 25 μg/L plates (Pryor and Leiting, *Protein Expression and Purification 10*:309-319, 1997) using standard procedures. Cells were grown in MM/CA with 25 μg/ml Kanomyacin for two hours, shaking, at 37°C. One ml of the culture was induced with 1mM IPTG. Two to four hours later the 250 μl of each culture was mixed with 250 μl acid washed glass beads and 250 μl Thorner buffer with 5% βME and dye (8M urea,

100 mM Tris pH7.0, 10% glycerol, 2mM EDTA, 5% SDS). Samples were vortexed for one minute and heated to 65°C for 10 minutes. 20 µl were loaded per lane on a 4%-12% PAGE gel (NOVEX). Gels were run in 1XMES buffer. The positive clones were designated pCMH01 and subjected to sequence analysis.

One microliter of sequencing DNA was used to transform strain BL21. The cells were electropulsed at 2.0 kV, 25 µF and 400 ohms. Following electroporation, 0.6 ml MM/CA with 25 µg/L Kanomyacin. Cells were grown in MM/CA and induced with ITPG as described above. The positive clones were used to grow up for protein purification of the huzcytor17/MBP-6H fusion protein using standard techniques.

B. Purification of huzcytor17/MBP-6H soluble receptor from E.coli fermentation

Unless otherwise noted, all operations were carried out at 4°C. The following procedure was used for the purification of recombinant huzcytor17/MBP-6H soluble receptor polypeptide. *E. coli* cells containing the pCMH01 construct and expressing huzcytor17/MBP-6H soluble receptor polypeptide were constructed using standard molecular biology methods and cultured in SuperBroth II (12 g/L Casien, 24 g/L Yeast Extract, 11.4 g/L di-potassium phosphate, 1.7 g/L Mono-potassium phosphate; Becton Dickenson, Cockeysville, MD). The resulting cells were harvested and frozen in 0.5% glycerol. Twenty grams of the frozen cells were used for protein purification.

Thawed cells were resuspended in 500mL Amylose Equilibration buffer (20mM Tris, 100mM NaCl, pH 8.0). A French Press cell breaking system (Constant Systems Ltd., Warwick, UK) with a temperature setting of -7°C to -10°C and 30K PSI was used to lyse the cells. The resuspended cells were checked for breakage by A_{600} readings before and after cycling through the French Press. The lysed cell suspension was pelleted at 10,000G for 30 minutes. Supernatant was harvested from the debris pellet for protein purification.

Twenty-five milliliters of Amylose resin (New England Biolabs, Beverly, MA) was poured into a Bio-Rad, 2.5 cm D x 10 cm H glass column. The column was packed and equilibrated by gravity with 10 column volumes (CVs) of

Amylose Equilibration buffer. The harvested cell supernatant was batch loaded to the Amylose resin, overnight with rocking. The loaded resin was returned to the glass column, washed with 10 CVs Amylose Equilibration buffer, and eluted by gravity with ~2 CVs Amylose Elution buffer (Amylose Equilibration buffer, 10 mM Maltose, Fluka Biochemical, Switzerland). Ten 5 ml fractions were collected over the elution profile and assayed for absorbance at 280 and 320 nM. The Amylose resin was regenerated with 1 CV of distilled H_2O, 5 CVs of 0.1% (w/v) SDS (Sigma), 5 CVs of distilled H_2O, 5 CVs of Amylose Equilibration buffer, and finally 1 CV of Amylose Storage buffer (Amylose Equilibration buffer, 0.02% (w/v) Sodium Azide). The regenerated resin was stored at 4°C.

Elution profile fractions of interest were pooled and dialyzed in a 10K dialysis chamber (Slide-A-Lyzer, Pierce Immunochemical) against 4 changes of 4L PBS pH 7.4 (Sigma) over an 8 hour time period. Following dialysis, the material harvested represented the purified huzcytor17/MBP-6H polypeptide. The purified huzcytor17/MBP-6H polypeptide was filter sterilized and analyzed via SDS-PAGE Coomassie staining for an appropriate molecular weight product. The concentration of the huzcytor17/MBP-6H polypeptide was determined by BCA analysis to be 0.76 mg/ml.

Purified huzcytor17/MBP-6H polypeptide was appropriately formulated for the immunizaton of rabbits and sent to R & R Research and Development (Stanwood, WA) for polyclonal antibody production (Example 25, below).

Example 25

Human zcytor17 Receptor Polyclonal Antibody

A. Preparation and Purification

Polyclonal antibodies were prepared by immunizing 2 female New Zealand white rabbits with the purified recombinant protein huzcytor17/MBP-6H (Example 24). The rabbits were each given an initial intraperitoneal (IP) injection of 200 μg of purified protein in Complete Freund's Adjuvant followed by booster IP injections of 100 μg protein in Incomplete Freund's Adjuvant every three weeks. Seven to ten days after the administration of the second booster injection (3 total

injections), the animals were bled and the serum was collected. The animals were then boosted and bled every three weeks.

The huzcytor17/MBP-6H specific rabbit serum was pre-adsorbed of anti-MBP antibodies using a CNBr-SEPHAROSE 4B protein column (Pharmacia LKB) that was prepared using 10 mg of non-specific purified recombinant MBP-fusion protein per gram of CNBr-SEPHAROSE. The huzcytor17/MBP-6H-specific polyclonal antibodies were affinity purified from the pre-adsorbed rabbit serum using a CNBr-SEPHAROSE 4B protein column (Pharmacia LKB) that was prepared using 10 mg of the specific antigen purified recombinant protein huzcytor17/MBP-6H. Following purification, the polyclonal antibodies were dialyzed with 4 changes of 20 times the antibody volume of PBS over a time period of at least 8 hours. Human zcytor17-specific antibodies were characterized by ELISA using 500 ng/ml of the purified recombinant protein huzcytor17/MBP-6H as antibody target. The lower limit of detection (LLD) of the rabbit anti-huzcytor17/MBP-6H affinity purified antibody is 500 pg/ml on its specific purified recombinant antigen huzcytor17/MBP-6H.

B. SDS-PAGE and Western blotting analysis of Rabbit Anti-human ZcytoR17 MBP-6H antibody

Rabbit anti-human ZcytoR17 MBP-6H antibody was tested by SDS-PAGE (NuPage 4-12%, Invitrogen, Carlsbad, CA) with coomassie staining method and Western blotting using goat anti-rabbit IgG-HRP. Either purified protein (200-25 ng) or conditioned media containing zcytor17 was electrophoresed using an Invitrogen Novex's Xcell II mini-cell, and transferred to nitrocellulose (0.2 mm; Invitrogen, Carlsbad, CA) at room temperature using Novex's Xcell blot module with stirring according to directions provided in the instrument manual. The transfer was run at 300 mA for one hour in a buffer containing 25 mM Tris base, 200 mM glycine, and 20% methanol. The filter was then blocked with Western A buffer (in house, 50 mM Tris, pH 7.4, 5 mM EDTA, pH 8.0, 0.05% Igepal CA-630, 150 mM NaCl, and 0.25% gelatin) overnight with gentle rocking at 4°C. The nitrocellulose was quickly rinsed, then the rabbit anti-human zcytoR17 MBP-6H (1:1000) was added in Western A buffer. The blot was incubated for 1.5 hours at room temperature with gentle rocking. The blot

was rinsed 3 times for 5 minutes each in Western A, then goat anti-rabbit IgG HRP antibody (1:1000) was added in Western A buffer. The blot was incubated for 1.25 hours at room temperature with gentle rocking. The blot was rinsed 3 times for 5 minutes each in Western A, then quickly rinsed in H_20. The blot was developed using commercially available chemiluminescent substrate reagents (ECLWestern blotting detection reagents 1 and 2 mixed 1:1; reagents obtained from Amersham Pharmacia Biotech, Buckinghamshire, England) and the blot was exposed to X-ray film for up to15 minutes.

The rabbit anti-human zcytoR17 MBP-6H was able to detect human zcytor17 present in conditioned media as well as zcytoR17 purified protein as a band at 120 kDa under reducing conditions.

Example 26

Tissue Distribution of Mouse zcytor17 in Tissue Panels Using PCR

A panel of cDNAs from murine tissues was screened for mouse zcytor17 expression using PCR. The panel was made in-house and contained 94 marathon cDNA and cDNA samples from various normal and cancerous murine tissues and cell lines are shown in Table 6, below. The cDNAs came from in-house libraries or marathon cDNAs from in-house RNA preps, Clontech RNA, or Invitrogen RNA. The mouse marathon cDNAs were made using the marathon-Ready™ kit (Clontech, Palo Alto, CA) and QC tested with mouse transferrin receptor primers ZC10,651 (SEQ ID NO:46) and ZC10,565 (SEQ ID NO:47) and then diluted based on the intensity of the transferrin band. To assure quality of the amplified library samples in the panel, three tests for quality control (QC) were run: (1) To assess the RNA quality used for the libraries, the in-house cDNAs were tested for average insert size by PCR with vector oligos that were specific for the vector sequences for an individual cDNA library; (2) Standardization of the concentration of the cDNA in panel samples was achieved using standard PCR methods to amplify full length alpha tubulin or G3PDH cDNA using a 5' vector oligo: ZC14,063 (SEQ ID NO:48) and 3' alpha tubulin specific oligo primer ZC17,574 (SEQ ID NO:49) or 3' G3PDH specific oligo primer ZC17,600 (SEQ ID NO:50); and (3) a sample was sent to sequencing to check for possible ribosomal or

mitochondrial DNA contamination. The panel was set up in a 96-well format that included a mouse genomic DNA (Clontech, Palo Alto, CA) positive control sample. Each well contained approximately 0.2-100 pg/μl of cDNA. The PCR was set up using oligos ZC38,065 (SEQ ID NO:51) and ZC38,068 (SEQ ID NO:52), TaKaRa Ex Taq™ (TAKARA Shuzo Co LTD, Biomedicals Group, Japan), and Rediload dye (Research Genetics, Inc., Huntsville, AL). The amplification was carried out as follows: 1 cycle at 94°C for 5 minutes; 5 cycles of 94°C for 30 seconds, 68°C for 30 seconds; 35 cycles of 94°C for 30 seconds, 56°C for 30 seconds and 72°C for 30 seconds, followed by 1 cycle at 72°C for 5 minutes. About 10 μl of the PCR reaction product was subjected to standard Agarose gel electrophoresis using a 4% agarose gel. The correct predicted DNA fragment size was observed in brain, CD90+ cells, dendritic, embryo, MEWt#2, Tuvak-prostate cell line, salivary gland, skin and testis.

The DNA fragment for skin and testis were excised and purified using a Gel Extraction Kit (Qiagen, Chatsworth, CA) according to manufacturer's instructions. Fragments were confirmed by sequencing to show that they were indeed mouse zcytor17.

Table 6

Tissue/Cell line	#samples		Tissue/Cell line	#samples
229	1			
7F2	1			
Adipocytes-Amplified	1			
aTC1.9	1			
Brain	4			
CCC4	1			
CD90+ Amplified	1			
OC10B	1			
Dentritic	1			
Embyro	1			
Heart	2			
Kidney	3			
Liver	2			
Lung	2			
MEWt#2	1			
P388D1	1			
Pancreas	1			
Placenta	2			
Jakotay-Prostate Cell Line	1			
Nelix-Prostate Cell Line	1			
Paris-Prostate Cell Line	1			
Torres-Prostate Cell Line	1			
Tuvak-Prostate Cell Line	1			
Salivary Gland	2			
Skeletal Muscle	1			
Skin	2			
Small Intestine	1			
Smooth Muscle	2			
Spleen	2			
Stomach	1			
Testis	3			
Thymus	1			

Example 27: Human Zcytor17 Expression in Various Tissues Using Real-Time Quantitative RT/PCR

A. Primers and Probes for Human Zcytor17, OSMRbeta and Zcytor17lig for Conventional and Quantitative RT-PCR

Real-time quantitative RT-PCR using the ABI PRISM 7900 Sequence Detection System (PE Applied Biosystems, Inc., Foster City, CA) has been previously

described (See, Heid, C.A. et al., *Genome Research 6*:986-994, 1996; Gibson, U.E.M. et al., *Genome Research 6*:995-1001, 1996; Sundaresan, S. et al., *Endocrinology 139*:4756-4764, 1998). This method incorporates use of a gene specific probe containing both reporter and quencher fluorescent dyes. When the probe is intact the reporter dye emission is negated due to the close proximity of the quencher dye. During PCR extension using additional gene-specific forward and reverse primers, the probe is cleaved by 5' nuclease activity of Taq polymerase which releases the reporter dye from the probe resulting in an increase in fluorescent emission.

The primers and probes used for real-time quantitative RT-PCR analyses of human Zcytor17, OSMRbeta and Zcytor17ligand expression were designed using the primer design software Primer Express™ (PE Applied Biosystems, Foster City, CA). Primers for human Zcytor17 were designed spanning an intron-exon junction to eliminate possible amplification of genomic DNA. The forward primer, ZC37,877 (SEQ ID NO:53) and the reverse primer, ZC37,876 (SEQ ID NO:54) were used in a PCR reaction at a 200 nM concentration to synthesize a 73 bp product. The corresponding Zcytor17 TaqMan® probe, designated ZC37,776 (SEQ ID NO:55) was synthesized and labeled by PE Applied Biosystems and used in each PCR reaction at a concentration of 200nM. The ZC37,776 (SEQ ID NO:55) probe was labeled at the 5'end with a reporter fluorescent dye (6-carboxy-fluorescein) (FAM) (PE Applied Biosystems) and at the 3' end with a fluorescent quencher dye (6-carboxy-tetramethyl-rhodamine) (TAMRA) (Epoch Biosciences, Bothell, WA).

Primers for human OSMRbeta were designed spanning an intron-exon junction to eliminate possible amplification of genomic DNA. The forward primer, ZC43,891 (SEQ ID NO:137) and the reverse primer, ZC43,900 (SEQ ID NO:138) were used in a PCR reaction (below) at a 200 nM concentration. The corresponding OSMRbeta TaqMan® probe, designated ZC43,896 (SEQ ID NO:139) was synthesized and labeled by PE Applied Biosystems and used in each PCR reaction at a concentration of 200nM. The ZC43,896 (SEQ ID NO:139) probe was labeled at the 5'end with a reporter fluorescent dye (6-carboxy-fluorescein) (FAM) (PE Applied Biosystems) and at the 3' end with a non-fluorescent quencher dye (ECLIPSE) (Epoch Biosciences).

Primers for human Zcytor17ligand were designed spanning an intron-exon junction to eliminate possible amplification of genomic DNA. The forward primer, ZC43,280 (SEQ ID NO:140) and the reverse primer, ZC43,281 (SEQ ID NO:141) were used in a PCR reaction (below) at about 200 nM concentration. The corresponding Zcytor17ligand TaqMan® probe, designated ZC43,275 (SEQ ID NO:142) was synthesized and labeled by PE Applied Biosystems and used in each PCR reaction at a concentration of 200nM. The ZC43,275 (SEQ ID NO:142) probe was labeled at the 5'end with a reporter fluorescent dye (6-carboxy-fluorescein) (FAM) (PE Applied Biosystems) and at the 3' end with a non-fluorescent quencher dye (ECLIPSE) (Epoch Biosciences).

As a control to test the integrity and quality of RNA samples tested, all RNA samples were screened for either rRNA or GUS using primer and probe sets either ordered from PE Applied Biosystems (rRNA kit) or designed in-house (GUS). The rRNA kit contained the forward primer (SEQ ID NO:56), the rRNA reverse primer (SEQ ID NO:57), and the rRNA TaqMan® probe (SEQ ID NO:58). The rRNA probe was labeled at the 5'end with a reporter fluorescent dye VIC (PE Applied Biosystems) and at the 3' end with the quencher fluorescent dye TAMRA (PE Applied Biosystems). The GUS primers and probe were generated in-house and used in each PCR reaction at 200nM and 100nM, respectively. The forward primer was ZC40,574 (SEQ ID NO:143) and the reverse primer was ZC40,575 (SEQ ID NO:144). The GUS probe ZC43,017 (SEQ ID NO:145) was labeled at the 5'end with a reporter fluorescent dye (Yakima-Yellow) (Epoch Biosciences) and at the 3'end with a non-fluorescent quencher dye (ECLIPSE) (Epoch Biosciences). The rRNA and GUS results also serve as an endogenous control and allow for the normalization of the Zcytor17 mRNA expression results seen in the test samples.

For conventional non-quantitative RT-PCR, primers were designed using the primer design software Primer Express™ (PE Applied Biosystems, Foster City, CA). The human zcytor17 primers generate an approximately 1000 base pair product and are as follows: forward primer ZC28,917 (SEQ ID NO:83), and reverse primer ZC28,480 (SEQ ID NO:146). The human OSMRbeta primers generate a 202 base pair product and are as follows: forward primer ZC41,653(SEQ ID NO:147) and

reverse primer ZC41,655 (SEQ ID NO:148). The human Zcytor17ligand primers generate a 305 base pair product and are as follows: forward primer ZC41,703 (SEQ ID NO:149) and reverse primer ZC41,704 (SEQ ID NO:150).

B. Primers and Probes for Murine Zcytor17, OSMRbeta and Zcytor17ligand for Conventional and Quantitative RT-PCR

The primers and probes used for real-time quantitative RT-PCR analyses of murine Zcytor17, OSMRbeta and Zcytor17lig expression were designed using the primer design software Primer Express™ (PE Applied Biosystems, Foster City, CA). Primers for murine Zcytor17 were designed spanning an intron-exon junction to eliminate possible amplification of genomic DNA. The forward primer, ZC43,272 (SEQ ID NO:151) and the reverse primer, ZC43,273 (SEQ ID NO:152) were used in the PCR reactions (below) at 300 nM concentration. The corresponding Zcytor17 TaqMan® probe, designated ZC43,478 (SEQ ID NO:153) was synthesized and labeled by PE Applied Biosystems. The ZC43,478 (SEQ ID NO:153) probe was labeled at the 5'end with a reporter fluorescent dye (6-carboxy-fluorescein) (FAM) (PE Applied Biosystems) and at the 3' end with a quencher fluorescent dye (6-carboxy-tetramethyl-rhodamine) (TAMRA) (PE Applied Biosystems). The ZC43,478 (SEQ ID NO:153) probe was used in the PCR reactions at a concentration of 100nM.

Primers for murine Zcytor17ligand were designed spanning an intron-exon junction to eliminate possible amplification of genomic DNA. The forward primer, ZC43,278 (SEQ ID NO:154) and the reverse primer, ZC43,279 (SEQ ID NO:155) were used in the PCR reactions at 500 nM concentration. The corresponding Zcytor17ligand TaqMan® probe, designated ZC43,276 (SEQ ID NO:156) was synthesized and labeled by PE Applied Biosystems. The ZC43,478 (SEQ ID NO:153) probe was labeled at the 5'end with a reporter fluorescent dye (6-carboxy-fluorescein) (FAM) (PE Applied Biosystems) and at the 3' end with a non-fluorescent quencher dye (ECLIPSE) (Epoch Biosciences). The ZC43,276 (SEQ ID NO:156) probe was used in the PCR reactions (below) at a concentration of 200nM.

Primers for murine OSMRbeta were designed spanning an intron-exon junction to eliminate possible amplification of genomic DNA. The forward primer,

ZC43,045 (SEQ ID NO:157) and the reverse primer, ZC43,046 (SEQ ID NO:158) were used in the PCR reactions at a 300 nM concentration. The corresponding OSMRbeta TaqMan® probe, designated ZC43,141(SEQ ID NO:159) was synthesized and labeled by Epoch Biosciences. The ZC43,141 (SEQ ID NO:159) probe was labeled at the 5'end with a reporter fluorescent dye (6-carboxy-fluorescein) (FAM) (PE Applied Biosystems) and at the 3' end with a non-fluorescent quencher dye (ECLIPSE) (Epoch Biosciences). The ZC43,141 (SEQ ID NO:159) probe was used in the PCR reactions (below) at a concentration of 100nM.

As a control to test the integrity and quality of RNA samples tested, all RNA samples were screened for either murine GUS or transferrin receptor using primers and probes designed using the primer design program Primer Express™ (PE Applied Biosystems Inc., Foster City, CA). The murine GUS primers are as follows: forward primer, ZC43,004 (SEQ ID NO:160), reverse primer, ZC43,005 (SEQ ID NO:161), and TaqMan® probe ZC43,018 (SEQ ID NO:162). The murine GUS probe ZC43,018 (SEQ ID NO:162) was labeled at the 5'end with a reporter fluorescent dye Yakima-Yellow (Epoch Biosciences) and at the 3' end with the non-fluorescent quencher dye ECLIPSE (Epoch Biosciences). The murine GUS primers were used in the PCR reactions at 300 nM and the probe, ZC43,018 (SEQ ID NO:162), was used at 100nM. In some cases murine Transferrin Receptor was used instead of GUS as the endogenous control. The transferrin receptor forward primer, ZC40,269 (SEQ ID NO:163) and the reverse primer, ZC40,268 (SEQ ID NO:164) were used at 300nM. The transferrin receptor probe, ZC40,298 (SEQ ID NO:165) was used in PCR at 100nM and was labeled at the 5'end with a reporter fluorescent dye VIC (PE Applied Biosystems) and at the 3'end with a fluorescent quencher dye (TAMRA) (PE Applied Biosystems). The murine GUS and transferrin receptor results also serve as an endogenous control and allow for the normalization of the Zcytor17, OSMRbeta and Zcytor17ligand mRNA expression results seen in the test samples.

For conventional semi-quantitative RT-PCR, primers were designed using the primer design software Primer Express™ (PE Applied Biosystems). The murine Zcytor17 primers generate a 276 base pair product and are as follows: forward primer ZC43,140 (SEQ ID NO:166), and reverse primer ZC43,139 (SEQ ID NO:167).

The murine OSMRbeta primers generate a 575 base pair product and are as follows: forward primer ZC41,608 (SEQ ID NO:168) and reverse primer ZC41,609 (SEQ ID NO:169). The murine Zcytor17ligand primers generate a 657bp product and are as follows: forward primer ZC41,502 (SEQ ID NO:170) and reverse primer ZC41,500 (SEQ ID NO:171).

C. Protocols for Realtime Quantitative RT-PCR and Conventional Semi-quantitative RT-PCR

Relative levels of Zcytor17, OSMRbeta and Zcytor17ligand mRNA were determined by analyzing total RNA samples using the one-step RT-PCR method (PE Applied Biosystems). Total RNA from Zcytor17- and OSMRbeta-transfected BAF cells (human) or BHK cells (murine) was isolated by standard methods and used to generate a standard curve used for quantitation of Zcytor17 and OSMRbeta. The curve consisted of 10-fold serial dilutions ranging from 100-0.01ng/µl with each standard curve point analyzed in triplicate. Similarly, for Zcytor17ligand, activated CD4+ T cell RNA (previously shown to make Zcytor17ligand) was used to generate a standard curve in the same 100-0.01ng/µl range. Total RNA from human or murine cells was analyzed in triplicate for either human or murine Zcytor17, OSMRbeta and Zcytor17ligand transcript levels and for one of the following endogenous control genes: rRNA, GUS or transferrin receptor. In a total volume of 10 µl, each RNA sample was subjected to a One-Step RT-PCR reaction containing: approximately 50-100ng of total RNA in a preformulated 2X master mix containing an internal control dye (ROX)(carboxy-x-rhodamine) and Thermo-Start® DNA Polymerase (Abgene, Surrey, UK); appropriate primers for the gene of interest (see parts A and B of current example); the appropriate probe (see parts A and B for concentration); Superscript® reverse transcriptase (50 U/µl) (PE Applied Biosystems), and an appropriate volume of RNase-free water. PCR thermal cycling conditions were as follows: an initial reverse transcription (RT) step of one cycle at 48°C for 30 minutes; followed by a Thermo-Start® enzyme activation step of one cycle at 95°C for 10 minutes; followed by 40 cycles of amplification at 95°C for 15 seconds and 60°C for 1 minute. Relative Zcytor17, OSMRbeta and Zcytor17ligand RNA levels were determined by using the Standard Curve Method as described by the

manufacturer, PE Biosystems (User Bulletin #2: ABI Prism 7700 Sequence Detection System, Relative Quantitation of Gene Expression, December 11, 1997). The rRNA, GUS or Transferrin Receptor measurements were used to normalize the levels of the gene of interest.

The semi-quantitative RT-PCR reactions used the 'Superscript One-Step RT-PCR System with Platinum Taq' (Invitrogen, Carlsbad, CA). Each 25μl reaction consisted of the following: 12.5μl of 2X Reaction Buffer, 0.5μl (20pmol/μl) of forward primer, 0.5μl (20pmol/μl) of reverse primer, 0.4μl RT/Taq polymerase mix, 5.0μl of Rediload Gel Loading Buffer (Invitrogen), 5.1μl RNase-free water, and 1.0μl total RNA (100ng/μl). The amplification was carried out as follows: one cycle at 45°C for 30 minutes followed by 35-38 cycles of 94°C, 20 seconds; Variable annealing temp (See Table 7 below), 20 seconds; 72°C, 45 seconds; then ended with a final extension at 72°C for 5 minutes. Eight to ten microliters of the PCR reaction product was subjected to standard agarose gel electrophoresis using a 2% agarose gel.

Table 7

Murine Zcytor17	58°C anneal temp
Murine OSMRbeta	60°C anneal temp
Murine Zcytor17ligand	52°C anneal temp
Human Zcytor17	55°C anneal temp
Human OSMRbeta	59°C anneal temp
Human Zcytor17ligand	59°C anneal temp

D. Isolation of RNA from Human and Murine PBMC Subsets and Cell Lines

Blood was drawn from several anonymous donors and peripheral blood mononuclear cells (PBMC) isolated using standard Ficoll gradient methodology. Monocytes were then isolated using the Monocyte Isolation Kit and the Magnetic Cell Separation System (Miltenyi Biotec, Auburn, CA). The monocytes were then plated onto ultra-low adherence 24-well plates in endotoxin-free media. They were either unstimulated or treated with recombinant human IFNg (R&D Systems Inc.) at 10ng/ml. Cells were collected at 24 and 48 hours. In similar manner, CD4+ and CD8+ T cells were isolated from PBMC's using the anti-CD4 or anti-CD8 magnetic beads from

Miltenyi Biotec. Cells were then activated for 4 or 16 hours in tissue culture plates coated with 0.5μg/ml anti-CD3 antibodies in media containing 5μg/ml anti-CD28 antibodies. NK cells were also isolated from PBMC's using Miltenyi's anti-CD56 coated magnetic beads. Some of the NK cells were collected at time zero for RNA and the others were plated in media containing Phorbol Myristate Acetate (PMA) (5ng/ml) and ionomycin (0.5μg/ml) for 24 hours. Additionally, several human monocyte-like cell lines, U937, THP-1 and HL-60, were collected in either their resting or activated states. U937 cells were activated overnight with PMA (10ng/ml). HL-60's were activated overnight with PMA (10ng/ml) or for 72 and 96 hours with IFNg (10ng/ml) to drive them down a monocytic pathway. THP-1 cells were activated overnight with a combination of LPS (10ng/ml) and IFNg (10ng/ml). RNA was prepared from all primary cells using the RNeasy Midiprep™ Kit (Qiagen, Valencia, CA) as per manufacturer's instructions. Carryover DNA was removed using the DNA-*Free*™ kit (Ambion, Inc., Austin, TX). RNA concentration was determined using standard spectrophotometry and RNA quality determined using the Bioanalyzer 2100 (Agilent Technologies, Palo Alto, CA).

Murine T Cell RNA was collected using a variety of methods well-known in the art. Primary splenic CD4+ and CD8+ T cells were isolated from the spleens of C57Bl/6 mice using antibody-coated magnetic beads and the Magnetic Cell Separation System from Miltenyi Biotec. The CD4+ and CD8+ T cells were then activated by culturing the cells in 24-well plates coated with anti-CD3 antibodies (500ng/ml) in media containing anti-CD28 antibodies at 5μg/ml. Cells were harvested for RNA at 0, 4 and 16 hours. Similarly, CD4+ T cells were isolated and then skewed towards a Th1 or Th2 phenotype using the following protocol. Since C57Bl/6 T cells are already skewed in the Th1 direction, all that was required was to activate for 6 hours with 0.5μg/ml PMA and 10ng/ml ionomycin. 'Th2' skewing was obtained by plating naïve CD4+ T cells with 2.5μg/ml anti-CD28, 10ng/ml mIL-2 (R&D Systems Inc.) and 25ng/ml mIL-4 (R&D Systems) into plates coated with 0.5μg/ml anti-CD3. After 2 days in culture, cells were resuspended in media containing 10ng/ml mIL-2 (R&D Systems) and 25ng/ml mIL-4. Cells were cultured for an additional three days then activated with PMA and ionomycin for 6 hours.

One additional set of Th1 and Th2 skewed T cells was derived using the T Cell Receptor Transgenic DO11.10 T cell line. All cells were plated into anti-CD3 and anti-CD28 coated plates. The 'Th1' cells were plated in media containing mIL-12 (1ng/ml) and anti-IL-4 (10μg/ml). The 'Th2' cells were plated in media containing mIL-4 (10ng/ml) and anti-IFNg (10μg/ml). After 24 hours, all cultures were given mIL-2 (10ng/ml). After two more days, the media on the cells was changed and new media containing the aforementioned cytokines was added and cells were cultured an additional 4 days before being harvested.

All of the murine T cell RNA was prepared using the RNeasy Midiprep™ Kit (Qiagen) and contaminating DNA was removed using the DNA-*free*™ kit from Ambion.

E. Isolation of RNA from the Murine Models of Pancreatitis and Irritable Bowel Disease

To induce a condition similar to human Irritable Bowel Disease (IBD), the hybrid mouse strain C57Bl6/129S6F1 was used. Mice were divided into 4 groups with an average size of six mice per group. Group 1 was given no Dextran Sulfate Sodium (DSS) and was sacrificed on day 14. Group 2 received 2% DSS for two days prior to being sacrificed. Group 3 received 2% DSS for seven days prior to sacrifice. Group 4 received 2% DSS for seven days then allowed to recover for seven days and was sacrificed on day 14. On the day of sacrifice, the distal colon sections were removed and placed in RNA*later*™ (Ambion). The colon sections were homogenized using standard techniques and RNA was isolated using the RNeasy Midiprep™ Kit (Qiagen). Contaminating DNA was removed by DNA-*free*™ (Ambion) treatment as per manufacturer's instructions.

In a different study, acute pancreatitis was induced in male CD-1 mice by caerulein injection. Mice were divided into three groups (n= 8 mice/group). Group 1 animals were given seven i.p. injections (1 injection per hour) with Vehicle (saline), and then sacrificed at 12 and 24 hours following the first injection. Groups 2 and 3 were given seven i.p. injections of caerulein (Sigma) (Catalog#C-9026) at a dose of 50μg/kg/hr for six hours (1 injection per hour). Group 2 was sacrificed at 12 hrs after

the first injection and Group 3 was sacrificed at 24 hrs following the first injection. Pancreases were removed at the time of sacrifice and snap frozen for RNA isolation. Tissues were homogenized and RNA was isolated using the Qiagen RNeasy Midiprep™ Kit.

In yet another study, murine Zcytor17ligand transgenic mice were generated and observed for phenotypic changes (see Example 41). Piloerection and hair loss was observed in many of the transgenic mice. Four transgenic mice were sacrificed and skin samples from both normal and hairless areas were removed and snap frozen for later RNA isolation. Skin sections from two non-transgenic control mice were collected as well. Skin samples were homogenized and then digested with Proteinase K (Qiagen) (Catalog# 19133) for 20 minutes at 60°C. RNA was then isolated using the Qiagen RNeasy Midiprep™ Kit following manufacturer's instructions. Carryover DNA was removed using DNA-*free*™ kit from Ambion.

F. Results of Quantitative and Semi-Quantitative RT-PCR for Human Zcytor17, OSMRbeta and Zcytor17ligand

Zcytor17 and OSMRbeta expression was examined by quantitative RT-PCR in four sets of primary human monocytes that were either in their resting state or activated with IFNg for 24 or 48 hours. Zcytor17 expression was below detection in the unstimulated cells but increased dramatically after the 24-hour activation with IFNg, and was the highest after 48 hours of activation. In all cases OSMRbeta was below detection. Zcytor17ligand was not tested in these samples.

In the primary T cells, Zcytor17 was below detection in both the resting CD4+ and CD8+ subsets. After a four-hour activation, however, expression of Zcytor17 went up in both subsets and then decreased to a slightly lower level at the 16 hour time point. OSMRbeta was below detection in these samples. Zcytor17ligand expression was examined using semi-quantitative RT-PCR. No expression was detected in the unstimulated CD4+ and CD8+ T cells. However, after the four hour activation, high levels of Zcytor17ligand were detected. This level dropped somewhat at the 16 hour time point.

Expression of Zcytor17 was not examined in NK cells. OSMRb was

below detection in these samples. Zcytor17ligand expression was below detection in the resting NK cells, however there was a faint signal generated by the activated NK cells suggesting that these cells may make Zcytor17ligand under certain conditions.

In the human monocyte–like cell lines, U937, THP-1 and HL-60, OSMRbeta expression was below detection in all of the resting and activated samples except for activated THP-1 samples where a faint signal was detected. Zcytor17 expression was high in both the U937 and THP-1 resting cell lines and showed a strong upregulation following activation. Expression in U937's was the highest of any cell type. In the HL-60's, Zcytor17 was expressed at moderate levels in the unstimulated cells and decreased upon stimulation with PMA. However, the expression of Zcytor17 was dramatically upregulated in the HL-60's when stimulated with IFNg for 72 and 96 hours. All of the human expression data is summarized in Table 8 below.

Table 8

Primary Human Monocytes	Activation Status	Zcytor17	OSMRbeta	Zcytor17Ligand
Human Monocytes	Unstim	-	-	
Human Monocytes	Act. 24hr IFNg	+	-	
Human Monocytes	Act. 48hr IFNg	++	-	
Human CD4+	Unstim	-	-	-
Human CD4+	Act 4hr	++	-	++
Human CD4+	Act. 16hr	+	-	+
Human CD8+	Unstim	-	-	-
Human CD8+	Act 4hr	++	-	++
Human CD8+	Act. 16hr	+	-	+
Human NK Cells	Unstim		-	-
Human NK Cells	Act 24hr		-	+
U937	Unstim	++	-	-
U937	Act. 16hr	+++	-	-
THP-1	Unstim	++	-	-
THP-1	Act. 16hr	+++	+	-
HL-60	Unstim	++	-	-
HL-60	Act. 16hr PMA	+	-	-
HL-60	Act. 72hr IFNg	+++	-	-
HL-60	Act. 96hr IFNg	+++	-	-

G. Results of Quantitative and Semi-Quantitative RT-PCR for Murine Zcytor17, OSMRbeta and Zcytor17ligand

Murine Zcytor17, OSMRbeta and Zcytor17ligand expression levels were examined in several murine T cells populations and the results are summarized in Table 9 below. Murine Zcytor17 expression was tested by semi-quantitative RT-PCR and shown to be at low levels on both resting and activated primary CD4+ T cells. Expression of Zcytor17 was detected on resting CD8+ T cells and then seemed to drop upon activation with anti-CD3 and anti-CD28 antibodies at both the 4- and 16-hour time points. OSMRbeta expression was measured by quantitative RT-PCR and shown to be expressed in resting and activated CD4+ and CD8+ T cells. The expression of OSMRbeta went up after a 4-hour activation and then returned to the unstimulated levels by 16 hours in both the CD4+ and CD8+ T cells. Zcytor17ligand was detected by quantitative RT-PCR and shown to be expressed at very low levels in unstimulated CD4+ T cells. However, following a 4-hour activation, Zcytor17ligand expression was dramatically upregulated and then dropped slightly by the 16-hour time point. In CD8+ T cells, no Zcytor17ligand was detected in the unstimulated cells. There was some Zcytor17ligand expression at the 4-hour time point, but by 16 hours expression levels had dropped back below detection.

In the DO11.10 T cells, Zcytor17 expression was detected in the naïve and Th2 skewed cells, but not in the Th1 skewed cells. OSMRbeta expression was at low levels in the naïve DO11.10 cells. There was a dramatic increase in OSMRbeta expression levels in the Th1 skewed cells and a moderate increase of expression in the Th2-skewed cells. The Zcytor17ligand expression in these cells was shown to be predominantly by the Th2 skewed subset. Low levels were detected in the Th1 subset and no expression was detected in the naïve cells. These results are summarized in the Table 9 below.

In the primary CD4+ T cells that were skewed in either the Th1 or Th2 direction, Zcyto17 wasn't examined. OSMRbeta expression was detected in all three samples with the highest levels found in the Th2 sample. Similar to the DO11.10 results, Zcytor17ligand expression was detected at high levels in the Th2 skewed subset, with a small amount detected in the Th1 subset and levels were below detection in the unstimulated cells. These results are summarized in the Table 9 below.

Table 9

Murine T Cells	Zcytor17	OSMRbeta	Zcytor17ligand
CD4+ T Cells Unstimulated	+	+	+/-
CD4+ T Cells 4hr Activation	+	++	++
CD4+ T Cells 16hr Activation	+	+	+
CD8+ T Cells Unstimulated	+	+	-
CD8+ T Cells 4hr Activation	+/-	++	+
CD8+ T Cells 16hr Activation	-	+	-
DO11.10 Naïve	+	+	-
DO11.10 Th1	-	+++	+
DO11.10 Th2	+	++	++
CD4+ T Cells Unstimulated		++	-
CD4+ T Cells - Th1 Skewed		+++	+
CD4+ T Cells - Th2 Skewed		++	+++

In the Zcytor17ligand transgenic skin samples, Zcytor17, OSMRbeta and Zcytor17ligand expression levels were determined using quantitative RT-PCR. Zcytor17 was shown to be present in all samples at roughly equivalent levels. There were dramatically higher levels of OSMRbeta expression in the non-transgenic control animals than the transgenic samples. Zcytor17ligand expression was below detection in the non-transgenic control animals with moderate to high levels of expression in the transgenic animals. The results are summarized in Table 10 below.

Table 10

Murine Zcytor17ligand Transgenic Skin	*Skin* Phenotype	Zcytor17	OSMRbeta	Zcytor17ligand
Wild Type Mouse	Normal	+	+++	-
Wild Type Mouse	Normal	+	+++	-
Transgenic #1	Normal	+	+	+
Transgenic #1	Hair Loss	+	+	+
Transgenic #2	Normal	+	+	+
Transgenic #2	Hair Loss	+	+	+
Transgenic #3	Normal	+	+	+
Transgenic #3	Hair Loss	+	+	+
Transgenic #4	Normal	+	+	+++
Transgenic #4	Hair Loss	+	+	+++

In a different experiment, Zcytor17, OSMRbeta and Zcytor17ligand expression levels were measured by quantitative RT-PCR in the pancreases of mice

subjected to acute pancreatitis. Zcytor17 expression was below detection in all of the samples. OSMRbeta expression was seen at low levels in the normal control samples (Group 1), but showed a strong upregulation at the 12-hour time point (Group 2) and slightly lower levels at the 24-hour time point (Group 3). Zcytor17ligand expression was below detection in the control animals, but showed high levels in both of the caerulein injected groups. The data is summarized in Table 11 below.

Table 11

Pancreatitis Model	Description	Zcytor17	OSMRbeta	Zcytor17ligand
Group 1	Normal Control	-	+	-
Group 2	12hr Post Injection	-	+++	++
Group 3	24hr Post Injection	-	++	++

In another experiment, Zcytor17 and OSMRbeta expression levels were examined in the distal colons of mice subjected to DSS treatment. In this murine model of Inflammatory Bowel Disease, expression levels of both genes were determined by quantitative RT-PCR and are summarized in Table 12 below. Zcytor17 expression levels increased with the severity of the disease, with low levels of expression in the Group 1 normal animals and increasing amounts seen Groups 2 and 3. In the Group 4 animals, the Zcytor17 levels had returned to more normal levels. Unlike Zcytor17 expression, OSMRbeta levels were the highest in the control animals and levels actually decreased in all three DSS treated groups.

Table 12

IBD Model	Description	SAC Day		Zcytor17	OSMRbeta
Group 1	Normal Control	14		+	++
Group 2	DSS-Treated 2 days	2		++	+
Group 3	DSS-Treated 7 days	7		+++	+
Group 4	DSS-Treated 7 days	14		+	+

Example 28

Human Zcytor17lig Tissue Distribution Expression based on RT-PCR Analysis of Multiple Tissue First-Strand cDNAs

Gene expression of the zcytor17lig was examined using commercially

available normalized multiple tissue first-strand cDNA panels (OriGene Technologies, Inc. Rockville, MD; BD Biosciences Clontech, Palo Alto, CA). These included the OriGene "Human Tissue Rapid-Scan™ Panel" (Cat. #CHSCA-101, containing 22 different tissues, bone marrow, and plasma blood leucocytes) and the BD Biosciences Clontech "Human Blood Fractions MTC™ Panel" (Cat. #K1428-1, containing 9 different blood fractions).

PCR reactions were set up using the zcytor17lig specific oligo primers ZC41,458 (SEQ ID NO:60), and ZC41,457 (SEQ ID NO:61), which yield a 139 bp product, and ZC41,459 (SEQ ID NO:62), and ZC41,460 (SEQ ID NO:63), which yield a 92 bp product, Qiagen HotStarTaq DNA polymerase and buffer (Qiagen, Inc., Valencia, CA), dH_2O, and *Red*iLoad™ dye (Research Genetics, Inc., Huntville, AL). The PCR cycler conditions were as follows: an initial 1 cycle 15 minute denaturation at 95°C, 35 cycles of a 45 second denaturation at 95°C, 1 minute annealing at 53°C or 56°C and 1 minute and 15 seconds extension at 72°C, followed by a final 1 cycle extension of 7 minutes at 72°C. The reactions were separated by electrophoresis on a 2% agarose gel (EM Science, Gibbstown, NJ) and visualized by staining with ethidium bromide.

A DNA fragment of the correct size was observed in the following human adult tissues using the OriGene "Human Tissue Rapid-Scan™ Panel": testis, plasma blood leucocytes (PBL), and bone marrow.

A DNA fragment of the correct size was observed in the following human blood fractions using the BD Biosciences Clontech "Human Blood Fractions MTC™ Panel": activated mononuclear cells (B- & T-cells and monocytes), activated CD8+ cells (T-suppressor/cytotoxic), activated CD4+ cells (T-helper/inducer) and faintly in resting CD8+ cells.

Example 29

Cloning the human Oncostatin M receptor

The OncostatinM beta receptor (OSMRbeta) is a type I cytokine receptor with structural similarity to IL12R-B2. ZcytoR17 has structural similarity to IL12R - B1. The OSMRbeta and zcytor17 were tested to see whether they could interact as

subunits in a cytokine signaling complex, and whether together they could act as a signaling receptor, or soluble receptor antagonist, for zcytor17lig.

To isolate OSMRbeta, oligonucleotide PCR primers ZC39982 (SEQ ID NO:64) and ZC39983 (SEQ ID NO:65) were designed to amplify the full length coding region of the human OncostatinM beta cDNA sequence (SEQ ID NO:6) (Genbank Accession No. U60805; Mosley B, *JBC Volume 271*, Number 50, Issue of December 20, 1996 pp. 32635-32643).

PCR reactions were run on an array of cDNA library templates using a robust polymerase, Advantage II (Clonetech, PaloAlto, CA), in order to identify a source of the cDNA. The template DNA used was from amplified cDNA plasmid libraries each containing 5 million independent cDNA clones. Reactions were assembled as per manufacturer's instructions using 400 fmol/µl of each oligonucleotide and 2-20 ng/µl purified plasmid library DNA as template. The cDNA libraries were derived from the following human tissues and cell lines: fetal brain, prostate smooth muscle, bone marrow, RPMI1588, thyroid, WI-38, testis, stimulated peripheral blood mononuclear cells, stimulated CD3+ cells, THP-1, activated tonsil, HACAT and fetal liver. Reactions were performed on a thermocycler machine using the following conditions: 30 cycles of 95°C for 20 seconds, 68°C for 3 minutes. At the conclusion of 30 cycles an additional single extension cycle of 8 minutes at 68°C was run. PCR products were visualized by TAE agarose, gel electrophoresis in the presence of ethidium bromide followed by UV illumination. The most abundant product was found to be from a prostate smooth muscle cDNA library. The PCR reaction using prostate smooth muscle template and oligonucleotides ZC39982 (SEQ ID NO:64) and ZC39983 (SEQ ID NO:65) was repeated using a less robust but higher fidelity thermostable DNA polymerase "turboPFu", (Stratagene, La Jolla, CA). Thirty amplification cycles were run with the following conditions: denaturing at 94°C, 30 seconds, annealing at 63°C 45 seconds, extension at 72°C 3.5 minutes. A single band product was gel purified on a 0.8% TAE, agarose gel.

This DNA was then amplified again using primers ZC39980 (SEQ ID NO:66) and ZC39981 (SEQ ID NO:67) designed to include restriction enzyme recognition sequences to allow the cloning of this cDNA into a mammalian expression

vector.

The PCR reaction was performed using "TurboPfu" and the purified PCR product for 15 cycles of: 95°C 1 minute, 64°C 1 minute 20 seconds, 72°C 4.5 minutes. The PCR reaction was then digested with EcoR1 and Xho1 (Invitrogen, Carlsbad,CA) and gel purified as described above. A mammalian expression vector, pZ7NX, was prepared by digesting with EcoR1 and Xho1 and the PCR product was ligated to this vector and electroporated into *E. coli* DH10b cells. Several bacterial colonies were isolated and sequenced. One clone was correct with the exception of a single non-conservative mutation. In order to change this base to match the expected sequence, an oligonucleotide spanning mutation and a neighboring Pst1 restriction site was used in a PCR reaction with "TurboPfu" using the pZP7Nx-h. OncostatinM R plasmid previously sequenced as a template. The PCR amplified DNA was digested with Pst1 and Xho1 and cloned back into the pZP7Nx-h OncostatinM R plasmid in place of the Pst1/Xho1 fragment containing the offending mutation. This new plasmid was sequenced over the recently amplified Pst1 to Xho1 region to confirm the correction and make sure no other errors were created in the amplification process. This analysis confirmed sequence that matched the expected sequence over the coding region. The sequence is shown in SEQ ID NO:6, and corresponding amino acid sequence shown in SEQ ID NO:7.

Example 30

Constructs for Generating a Human Zcytor17 / OncostatinM receptor (OSMRbeta) Heterodimer

A system for construction, expression and purification of such soluble heterodimeric receptors is known in the art, and has been adapted to the receptor pair, human oncostatin M receptor (OSMRbeta) and human zcytor17. For this construct, the polynucleotide for the soluble receptor for OSMRbeta is shown in SEQ ID NO:68 and corresponding polypeptide is shown in SEQ ID NO:69; and the polynucleotide for the soluble receptor for human zcytor17 is shown in SEQ ID NO:70 and corresponding polypeptide is shown in SEQ ID NO:71.

To construct a cell line expressing a secreted soluble hzcytor17/human

OSMRbeta heterodimer, a construct was made so that the resulting heterodimeric soluble receptor comprises the extracellular domain of human OSMRbeta fused to the heavy chain of IgG gamma1 (Fc4) (SEQ ID NO:37) with a Glu-Glu tag (SEQ ID NO:35) at the C-terminus; while the extracellular domain of zcytoR17 is fused to Fc4 (SEQ ID NO:37) with a His tag (SEQ ID NO:72) at the C-terminus. For both of the hzcytor17 and human OSMRbeta arms of the heterodimer a Gly-Ser spacer of 12 amino acids (SEQ ID NO:73) was engineered between the extracellular portion of the receptor and the N-terminus of Fc4.

A. Construction of human soluble OSMRbeta/Fc4-CEE

For construction of the human soluble OSMRbeta/Fc4-CEE portion of the heterodimer the extracellular portion of human OSMRbeta was isolated using PCR with oligos ZC14063 (SEQ ID NO:48) and ZC41557 (SEQ ID NO:74) under PCR reaction conditions as follows: 30 cycles of 95°C for 60 sec., 57°C for 30 sec., and 72°C for 100 sec.; and 72°C for 7 min. PCR products were purified using QIAquick PCR Purification Kit (Qiagen), digested with EcoR1 and BglII (Boerhinger-Mannheim), separated by gel electrophoresis and purified using a QIAquick gel extraction kit (Qiagen).

The expression cassette, plasmid backbone and Fc4-GluGlu tag portion of the chimera were contained within a previously made in house plasmid vector. The plasmid vector was digested with EcoR1 and BamH1 (Boerhinger-Mannheim), separated by gel electrophoresis and purified using a QIAquick gel extraction kit (Qiagen). The digested and purified fragments of human OSMRbeta and Fc4-cEE containing plasmid were ligated together using T4 DNA Ligase (Life Technologies, Bethesda, MD) using standard ligation methods. Minipreps of the resulting ligation were screened for an EcoRI/Sma1 insert of the correct size (772bp) for the soluble OSMRbeta and positive minipreps were sequenced to confirm accuracy of the PCR reaction. This new plasmid construction is termed pZP9-ONCOMR-Fc4CEE.

B. Construction of human soluble Zcytor17 / Fc4-CHIS

For construction of the hzcytor17/Fc4-CHIS portion of the heterodimer,

the extracellular portion of human zcytor17 was isolated by digestion of a plasmid previously containing Zcytor17-Fc4 soluble receptor. The plasmid was first digested with Sal1 (New England Biolabs, Beverly, MA) after which the reaction was serially phenol chloroform extracted and ethanol precipitated. The digested DNA was then treated with T4 DNA Polymerase (Boerhinger-Mannheim), to fill in the 5' overhangs created by the Sal1 digestion, leaving the DNA ends blunt, after which the reaction was serially phenol chloroform extracted and ethanol precipitated. The blunted DNA was then further digested with BglII to cut at the 3' end.), separated by gel electrophoresis and purified using a QIAquick gel extraction kit (Qiagen) as per manufacturer's instruction. The resulting DNA fragment containing the sequence coding for the extracellular domain of zcytoR17 was ligated into an Fc4-CHIS tag containing mammalian expression vector prepared as follows.

The expression cassette, plasmid backbone and Fc4-CHIS tag portion of the chimera were contained within a previously made in house plasmid vector. This plasmid vector was digested with EcoR1(Boerhinger-Mannheim) after which the reaction was serially phenol chloroform extracted and ethanol precipitated. The digested DNA was then treated with T4 DNA Polymerase (Boerhinger-Mannheim), to fill in the 5' overhangs created by the EcoR1 digestion, leaving the DNA ends blunt, after which the reaction was serially phenol chloroform extracted and ethanol precipitated. The blunted DNA was then further digested with BamH1 (Boerhinger-Mannheim) to cut at the 3' end, separated by gel electrophoresis and purified using a QIAquick gel extraction kit (Qiagen). The digested and purified fragments of human zcytor17 and Fc4-CHIS containing plasmid were ligated together using T4 DNA Ligase (Life Technologies, Bethesda, MD) using standard ligation methods.

Minipreps of the resulting ligation were screened by PCR using the zcytor17 specific sense primer ZC29180 (SEQ ID NO:22) and the Fc4 specific antisense primer ZC29232 (SEQ ID NO:75) with the following PCR reaction conditions: 30 cycles of 94°C for 60 sec., 68°C for 150 sec; and 72°C for 7 min. An expected product size of 848 bp confirmed the correct assembly of the plasmid termed pZEM228 hzcytor17/Fc4HIS.

A second zcytor17-Fc4 construction was created for use in generating

homodimer protein from COS cells. Briefly the coding region for the full fusion protein was isolated by digestion of a plasmid previously containing Zcytor17-Fc4 soluble receptor with Sal1 (Boerhinger-Mannheim). The reaction was serially phenol chloroform extracted and ethanol precipitated. The digested DNA was then treated with T4 DNA Polymerase (Boerhinger-Mannheim), to fill in the 5' overhangs created by the EcoR1 digestion, leaving the DNA ends blunt, after which the reaction was serially phenol chloroform extracted and ethanol precipitated. The blunted DNA was then further digested with Not1 (Boerhinger-Mannheim) to cut at the 3' end, separated by gel electrophoresis and purified using a QIAquick gel extraction kit (Qiagen). A mammalian expression vector containing a CMV driven expression cassette was digested to generate compatible ends and the 2 fragments were ligated together. Minipreps of the resulting ligation were screened by PCR using the vector specific sense primer ZC14063 (SEQ ID NO:48) and the zcytor17 specific antisense primer ZC27899 (SEQ ID NO:19) with the following PCR reaction conditions: 30 cycles of 94°C for 30 sec., 64°C for 30 sec; 70°C for 90 sec; and 72°C for 7 min. An expected product size of approximately 1000 bp confirmed the correct assembly of the plasmid termed pZP7NX-hzcytor17-Fc4. This plasmid was subsequently transfected into COS cells using Lipofectamine (Gibco/BRL), as per manufacturer's instructions. The cells were conditioned for 60 hours in DMEM + 5%FBS (Gibco/BRL) after which the protein was purified over a protein G-sepharose 4B chromatography column and made available for *in vitro* bioassays, for example, such as those described herein.

C. Generating a Human Zcytor17 / OncostatinM receptor (OSMRbeta)

About 16 μg each of the pZP9-ONCOMR-Fc4CEE and pZEM228 hzcytor17/Fc4HIS were co-transfected into BHK-570 (ATCC No. CRL-10314) cells using lipofectamine (Gibco/BRL), as per manufacturer's instructions. The transfected cells were selected for 10 days in DMEM + 5%FBS (Gibco/BRL) containing 0.5 mg/ml G418 (Gibco/BRL) and 250 nM methyltrexate (MTX)(Sigma, St. Louis, MO) for 10 days.

The resulting pool of doubly-selected cells was used to generate the heterodimeric protein. Three cell Factories (Nunc, Denmark) of this pool were used to

generate 10 L of serum free conditioned medium. This conditioned media was passed over a 1 ml protein-A column and eluted in (10) 750 microliter fractions. Four of these fractions found to have the highest concentration were pooled and dialyzed (10 kD MW cutoff) against PBS. The desired heterodimeric soluble zcytor17/OSMRbeta protein complex was isolated from other media components by passing the pool over a Nickel column and washing the column with various concentrations of Imidazole. The soluble zcytor17/OSMRbeta protein eluted at intermediate concentrations of Imidazole, while hzcytor17/Fc4HIS homodimer eluted at higher concentrations of Imidazole.

Example 31

Tissue Distribution of Human zcytor17 in Tissue Panels Using Northern Blot and PCR

A. Human zcytor17 Tissue Distribution using Northern Blot

Human Multiple Tissue Northern Blots (Human 12-lane MTN Blot I and II, and Human Immune System MTN Blot II; Human Endocrine MTN, Human Fetal MTN Blot II, Human Multiple Tissue Array) (Clontech) as well as in house blots containing various tissues were probed to determine the tissue distribution of human zcytor17 expression. The in-house prepared blots included the following tissue and cell line mRNA: SK-Hep-1 cells, THP1 cells, Adrenal gland (Clontech); Kidney (Clontech), Liver (Clontech and Invitrogen); Spinal cord (Clontech), Testis (Clontech), Human CD4+ T-cells, Human CD8+ T-cells, Human CD19+ T-cells, human mixed lymphocyte reaction (MLR), THP1 cell line (ATCC No. TIB-202), U937 cell line, p388D1 mouse lymphoblast cell line (ATCC No. CCL-46) with or without stimulation by Ionomycin; and WI-38 human embryonic lung cell line (ATCC No. CRL-2221) with or without stimulation by Ionomycin.

An approximately 500 bp PCR derived probe for zcytor17 (SE ID NO:4) was amplified using oligonucleotides ZC28,575 (SEQ ID NO:77) and ZC27,899 (SEQ ID NO:19) as primers. The PCR amplification was carried out as follows: 30 cycles of 94°C for 1 minute, 65°C for 1 minute, and 72°C for 1 minute; followed by 1 cycle at 72°C for 7 minutes. The PCR product was visualized by agarose gel electrophoresis and the approximately 500 bp PCR product was gel purified as described herein. The probe was radioactively labeled using the PRIME IT II™ Random Primer Labeling Kit

(Stratagene) according to the manufacturer's instructions. The probe was purified using a NUCTRAP™ push column (Stratagene). EXPRESSHYB™ (Clontech) solution was used for the prehybridization and as a hybridizing solution for the Northern blots. Prehybridization was carried out at 68°C for 2 hours. Hybridization took place overnight at 68°C with about $1.5X10^6$ cpm/ml of labeled probe. The blots were washed three times at room temperature in 2X SSC, 0.05% SDS, followed by 1 wash for 10 minutes in 2X SSC, 0.1% SDS at 50°C. Several faint bands were seen after several days exposure. An approximately 9 kb transcript was seen in trachea, skeletal muscle and thymus; an approximately 2 kb transcript was seen in PBL, HPV, U937 and THP-1 cells; and about a 1.2 kb transcript was seen in placenta, bone marrow and thyroid, and HPV and U937 cells. In all the tissues listed above, the signal intensity was faint. There appeared to be little expression in most normal tissues, suggesting that zcytor17 expression may be dependent on activation of the cell or tissues in which it is expressed.

B. Tissue Distribution in tissue panels using PCR

A panel of cDNAs from human tissues was screened for zcytor17 expression using PCR. The panel was made in-house and contained 94 marathon cDNA and cDNA samples from various normal and cancerous human tissues and cell lines as shown below in Table 13. The cDNAs came from in-house libraries or marathon cDNAs from in-house RNA preps, Clontech RNA, or Invitrogen RNA. The marathon cDNAs were made using the marathon-Ready™ kit (Clontech, Palo Alto, CA) and QC tested with clathrin primers ZC21195 (SEQ ID NO:78) and ZC21196 (SEQ ID NO:79) and then diluted based on the intensity of the clathrin band. To assure quality of the panel samples, three tests for quality control (QC) were run: (1) To assess the RNA quality used for the libraries, the in-house cDNAs were tested for average insert size by PCR with vector oligos that were specific for the vector sequences for an individual cDNA library; (2) Standardization of the concentration of the cDNA in panel samples was achieved using standard PCR methods to amplify full length alpha tubulin or G3PDH cDNA using a 5' vector oligo ZC14,063 (SEQ ID NO:48) and 3' alpha tubulin specific oligo primer ZC17,574 (SEQ ID NO:49) or 3'

G3PDH specific oligo primer ZC17,600 (SEQ ID NO:50); and (3) a sample was sent to sequencing to check for possible ribosomal or mitochondrial DNA contamination. The panel was set up in a 96-well format that included a human genomic DNA (Clontech, Palo Alto, CA) positive control sample. Each well contained approximately 0.2-100 pg/µl of cDNA. The PCR reactions were set up using oligos ZC26,358 (SEQ ID NO:80) and ZC26,359 (SEQ ID NO:81), TaKaRa Ex Taq™ (TAKARA Shuzo Co LTD, Biomedicals Group, Japan), and Rediload dye (Research Genetics, Inc., Huntsville, AL). The amplification was carried out as follows: 1 cycle at 94°C for 2 minutes, 35 cycles of 94°C for 30 seconds, 66.3°C for 30 seconds and 72°C for 30 seconds, followed by 1 cycle at 72°C for 5 minutes. About 10 µl of the PCR reaction product was subjected to standard agarose gel electrophoresis using a 4% agarose gel. The correct predicted DNA fragment size was observed in lymph node, prostate, thyroid, HPV (prostate epithelia), HPVS (prostate epithelia, selected), lung tumor, uterus tumor reactions, along with the genomic DNA reaction.

The DNA fragment for prostate tissue (2 samples), HPV (prostate epithelia), HPVS (prostate epithelia, selected), and genomic were excised and purified using a Gel Extraction Kit (Qiagen, Chatsworth, CA) according to manufacturer's instructions. Fragments were confirmed by sequencing to show that they were indeed zcytor17.

Table 13

Tissue/Cell line	#samples		Tissue/Cell line	#samples
Adrenal gland	1		Bone marrow	3
Bladder	1		Fetal brain	3
Bone Marrow	1		Islet	2
Brain	1		Prostate	3
Cervix	1		RPMI #1788 (ATCC # CCL-156)	2
Colon	1		Testis	4
Fetal brain	1		Thyroid	2
Fetal heart	1		WI38 (ATCC # CCL-75	2
Fetal kidney	1		ARIP (ATCC # CRL-1674 - rat)	1
Fetal liver	1		HaCat - human keratinocytes	1
Fetal lung	1		HPV (ATCC # CRL-2221)	1
Fetal muscle	1		Adrenal gland	1
Fetal skin	1		Prostate SM	2
Heart	2		CD3+ selected PBMC's Ionomycin + PMA stimulated	1
K562 (ATCC # CCL-243)	1		HPVS (ATCC # CRL-2221) - selected	1
Kidney	1		Heart	1
Liver	1		Pituitary	1
Lung	1		Placenta	2
Lymph node	1		Salivary gland	1
Melanoma	1		HL60 (ATCC # CCL-240)	3
Pancreas	1		Platelet	1
Pituitary	1		HBL-100	1
Placenta	1		Renal mesangial	1
Prostate	1		T-cell	1
Rectum	1		Neutrophil	1
Salivary Gland	1		MPC	1

Skeletal muscle	1		Hut-102 (ATCC # TIB-162)	1
Small intestine	1		Endothelial	1
Spinal cord	1		HepG2 (ATCC # HB-8065)	1
Spleen	1		Fibroblast	1
Stomach	1		E. Histo	1
Testis	2			
Thymus	1			
Thyroid	1			
Trachea	1			
Uterus	1			
Esophagus tumor	1			
Gastric tumor	1			
Kidney tumor	1			
Liver tumor	1			
Lung tumor	1			
Ovarian tumor	1			
Rectal tumor	1			
Uterus tumor	1			

C. Expression analysis of zcytoR17 by PCR and Northern

Annotation of the cell types and growth conditions that affect expression of the receptor is a useful means of elucidating its function and predicting a source of ligand. To that end a wide variety of tissue and cell types were surveyed by PCR. The thermostable polymerase Advantage II™ (Clontech, La Jolla, CA) was used with the oligonucleotide primers ZC29,180 (SEQ ID NO:22) and ZC29,179 (SEQ ID NO:82) and 1-10ng of the various cDNA templates listed below for 30 amplification cycles of (94°C, 30 sec.; 66°C, 20 sec.; 68°C, 1 min. 30 sec.). Following this, 20% of each reaction was run out on 0.8% agarose, TAE /ethidium bromide gels and visualized with UV light. Samples were then scored on the basis of band intensity. See Table 14 below.

Table 14

Cells and Conditions	Score 0-5
Hel stimulated with PMA	0
U937	3
MCF-7	0
HuH7	1
Human follicle	0
HT-29	0
HEPG2	0
HepG2 stimulated with IL6	0
Human dermal endothelial	0
Human venous endothelial	0
Human CD4+	0
BEWO	0
Human CD19+	1
Human PBMC stimulated with PHA, PMA, Ionomycin, IL2, IL4, TNFα 24 hours	0
Human PBMC stimulated with LPS, PWM, IFNγ, TNFα, 24 hours	0
Human PBMC all of the above conditions for 48 hours	4
HUVEC p.2	4
RPMI1788	0
TF1	0
Monkey spleen T cells stimulated with PMA, Ionomycin	0
Human prostate epithelia HPV transformed	5
Human tonsils, inflamed	0
HACAT	0
Human chondrocyte	1
Human synoviacyte	1
THP1	5
REH	0

Of the strong positive PCR signals, two were from the human monocyte cell lines U937 and THP1.

These two cell lines along with a prostate epithelia line were selected for further analysis by Northern blot. Previous attempts at visualizing a transcript by northern analysis using mRNA from various tissues yielded weak and diffuse signals in the surprisingly large size range of 7-10kb making this data difficult to interpret. A denaturing formaldehyde/MOPS/0.8%agarose gel was prepared (RNA Methodologies, Farrell, RE Academic Press) and 2 μg of polyA+ mRNA was run for each sample along side an RNA ladder (Life Technologies, Bethesda, MD). The gel was then transferred to Hybond nylon (Amersham, Buckinghamshire, UK), UV crosslinked, and hybridized in ExpressHyb solution (Clontech, LaJolla, CA) at 68°C overnight using a probe to human zcytoR17 generated by PCR with the oligos ZC28,575 (SEQ ID NO:77), and ZC27,899 (SEQ ID NO:19) and labeled with a Megaprime ^{32}P kit (Amersham). The northern blot was subsequently washed with 0.2xSSC+0.1%SDS at 65C for 15 minutes and exposed to film for 7 days with intensifying screens. A prominent 8kb band was seen in both the prostate epithelia and U937 lanes while a fainter band was present in the THP1 lane.

To optimize the cDNA used as a hybridization probe, four different regions of the full-length human zcytoR17 sequence were amplified by PCR, labeled and hybridized as described above to southern blots containing genomic and amplified cDNA library DNA. The four probes, herein designated probes A-D, were amplified using the following primer pairs: (A) ZC28,575 (SEQ ID NO:77), ZC27,899 (SEQ ID NO:19); (B) ZC27,895 (SEQ ID NO:20), ZC28,917 (SEQ ID NO:83); (C) ZC28,916 (SEQ ID NO:84), ZC28,918 (SEQ ID NO:85); and (D) ZC28,916 (SEQ ID NO:84), ZC29,122 (SEQ ID NO:21). Human genomic DNA along with amplified cDNA libraries demonstrated to contain zcytor17 by PCR were digested with EcoR1 and Xho1 to liberate inserts and run out on duplicate TAE/0.8%agarose gels, denatured with 0.5M NaOH, 1.5 M NaCl, blotted to Hybond, UV crosslinked and each hybridized with a distinct probe. Probe B was found to have the least nonspecific binding and strongest signal. Thus, Probe B was used for all subsequent hybridizations.

Given that the THP1 cells are an excellent model of circulating monocytes and expressed zcytor17 at low levels we treated them with a variety of compounds in an effort to increase expression of zcytoR17. The cells were grown to a density of 2e5/ml, washed and resuspended in various stimulating media, grown for four or thirty hours, and harvested for RNA preparations. Each media was supplemented with one of the following drugs or pairs of cytokines: LPS 2ug/ml (Sigma Chemicals, St. Louis, MO), hTNFα 2 ng/ml (R&D Systems, Minneapolis, MN), hGM-CSF 2ng/ml (R&D Systems, Minneapolis, MN), hIFNγ 50 ng/ml (R&D Systems, Minneapolis, MN), hMCSF 1ng/ml (R&D Systems, Minneapolis, MN), hIL6 1ng/ml (R&D Systems, Minneapolis, MN), hIL1β 2ng/ml (R&D Systems, Minneapolis, MN), hIFNγ 50ng/ml+hIL4 0.5ng/ml (R&D Systems, Minneapolis, MN), hIFNγ 50ng/ml+hIL10 1ng/ml (R&D Systems, Minneapolis, MN), PMA 10ng/ml (Calbiochem, San Diego, CA) and an untreated control. At the end of the culture period Total RNA was prepared using an RNAeasy Midi-kit (Qiagen, Valencia, CA). Poly A+ RNA was selected from the total RNA using an MPG kit (CPG, Lincoln Park, NJ). Two micrograms of polyA+ RNA from each condition was run on formaldehyde/MOPS/agarose gels, transferred to nylon and UV crosslinked as described above. These northern blots were then hybridized, as above, to probe B at 68°C overnight, washed at high stringency with 0.2XSSC, 0.1%SDS at 65C, exposed to film overnight then exposed to phosphor screens for signal quantitation. A dominant 8 kb mRNA as well a relatively weaker 2.8 kb band were seen in all lanes. A 20-fold increase in zcytor17 mRNA was seen in RNA from cells treated with hIFNγ for 30 hours, this effect was slightly muted with simultaneous treatment with IL4. Minor 3 fold increases in mRNA were seen in RNA from cells treated with LPS, TNFα and GM-CSF while MCSF, IL6, and IL1β had no effect on zcytor17 mRNA levels. Taken together this data suggests a role for the zcytor17 receptor and its ligand in monocyte macrophage biology and by extension any number of disease processes in which these cells participate.

Example 32

Tissue Distribution of human zcytor17 in Tissue Panels Using Northern Blot and PCR

A human zcytor17lig cDNA fragment was obtained using PCR with gene specific primers: Sense primer ZC41438 (SEQ ID NO:93) and antisense primer ZC41437 (SEQ ID NO:94) and template human zcytor17lig cDNA (SEQ ID NO:90) This fragment was purified using standard methods and about 25ng labeled with ^{32}P alpha dCTP using the Prime-It RmT random primer labeling kit (Stratagene) and hybridized in Ultrahyb, (Ambion) and used to expose Biomax film/intensifying screens per the manufacturer's recommendations in each case. New, previously unused blots Including the Clontech Human 12 lane MTN, the human brain MTN II, and the human brain MTN blot IV, the human immune system MTN II, and the human MTE array II, from Clontech were hybridized overnight at 42°C per the Ambion ultrahyb method. Non-specific radioactive counts were washed off using .1SSC/.5% SDS at 55°C. The positive blots included the human 12 lane MTN (Clontech). Of the 12 tissues examined, only placenta was positive for an approximately 1.2 KB transcript.

Example 33

Construction of mammalian expression vectors that express human zcytor17lig-CEE

A. Construction of zCytor17Lig-CEE/pZMP21

An expression plasmid containing all or part of a polynucleotide encoding zCytor17Lig-CEE (SEQ ID NO:95) was constructed via homologous recombination. The plasmid was called zCytor17Lig-CEE/pZMP21.

The construction of zCytor17Lig-CEE/pZMP21 was accomplished by generating a zCytor17Lig-CEE fragment using PCR amplification. The DNA template used for the production of the zCytor17Lig-CEE fragment was zCytor17Lig/pZP7nx. The primers used for the production of the zCytor17Lig-CEE fragment were: (1) ZC41,607 (SEQ ID NO:97) (sense sequence), which includes from the 5' to the 3' end: 28bp of the vector flanking sequence (5' of the insert) and 21 bp corresponding to the 5' sequence of zCytor17Lig; and (2) ZC41,605 (SEQ ID NO:98) (anti-sense sequence), which includes from the 5' to the 3' end: 37 bp of the vector flanking sequence (3' of the insert), 3 bp of the stop codon, 21 bp encoding a C-terminal EE tag, and 21 bp corresponding to the 3' end of zCytor17Lig sequence. The fragment resulting from the above PCR amplification is a copy of the template zCytor17Lig with the addition of a

C-terminal EE tag, yielding a final product zCytor17Lig-CEE.

PCR reactions were run as follows: To a 100μl final volume was added: 10μl of 10x Taq Polymerase Reaction Buffer with 15mM MgCl (Gibco), 1μl of Taq DNA Polymerase (5 units/μl, Gibco), 3μl of 10mM dNTPs, 78μl dH_2O, 3μl of a 20 pmol/μl stock of primer ZC41,607 (SEQ ID NO:97) 3μl of a 20 pmol/μl stock of primer ZC41,605 (SEQ ID NO:98), and 2μl of a 0.13μg/μl stock of zCytor17lig template DNA. A volume equal to 50μl of mineral oil was added to the mixture. The reaction was heated to 94°C for 5 minutes, followed by 35 cycles at 94°C for 1 minute; 55°C for 2 minutes; 72°C for 3 minutes; followed by a 10 minute extension at 72°C and held at 4°C until the reaction was collected.

The plasmid pZMP21 was restriction digested with BglII enzyme, cleaned with a QiaQuick PCR Purification Kit (Qiagen) using a microcentrifuge protocol, and used for recombination with the PCR fragment. Plasmid pZMP21 was constructed from pZMP20 which was constructed from pZP9 (deposited at the American Type Culture Collection, 10801 University Boulevard, Manassas, VA 20110-2209, and is designated No. 98668) with the yeast genetic elements from pRS316 (deposited at the American Type Culture Collection, 10801 University Boulevard, Manassas, VA 20110-2209, and designated No. 77145), an IRES element from poliovirus, and the extracellular domain of CD8, truncated at the carboxyl terminal end of the transmembrane domain. PZMP21 is a mammalian expression vector containing an expression cassette having the MPSV promoter, immunoglobulin signal peptide intron, multiple restriction sites for insertion of coding sequences, a stop codon and a human growth hormone terminator. The plasmid also has an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a DHFR gene, the SV40 terminator, as well as the URA3 and CEN-ARS sequences required for selection and replication in *S. cerevisiae*.

Fifty microliters of competent yeast cells (*S. cerevisiae*) were independently combined with 100 ng of cut plasmid, 5 μl of previously described PCR mixture, and transferred to a 0.2 cm electroporation cuvette. The yeast/DNA mixture was electropulsed at 0.75 kV (5 kV/cm), ∞ ohms, 25 μF. Each cuvette had 600 μl of 1.2 M sorbitol added, and the yeast was plated in one 100 μl aliquot and one 300 μl

aliquot onto two URA-D plates and incubated at 30°C. After about 72 hours, the Ura+ yeast transformants from a single plate were resuspended in 1 ml H_2O and spun briefly to pellet the yeast cells. The cell pellet was resuspended in 500 μl of lysis buffer (2% Triton X-100, 1% SDS, 100 mM NaCl, 10 mM Tris, pH 8.0, 1 mM EDTA). The 500 μl of the lysis mixture was added to an Eppendorf tube containing 300 μl acid washed 600μm glass beads and 300 μl phenol-chloroform, vortexed for 1 minute intervals two or three times, followed by a 5 minute spin in a Eppendorf centrifuge at maximum speed. Three hundred microliters of the aqueous phase was transferred to a fresh tube, and the DNA precipitated with 600 μl 100% ethanol (EtOH), followed by centrifugation for 10 minutes at 4°C. The DNA pellet was then washed with 500μl 70% EtOH, followed by centrifugation for 1 minute at 4°C. The DNA pellet was resuspended in 30 μl H_2O.

Transformation of electrocompetent *E. coli* cells (MC1061) was done with 5 μl of the yeast DNA prep and 50 μl of MC1061 cells. The cells were electropulsed at 2.0 kV, 25 μF and 400 ohms(Ω). Following electroporation, 600μl SOC (2% Bacto‘ Tryptone (Difco, Detroit, MI), 0.5% yeast extract (Difco), 10 mM NaCl, 2.5 mM KCl, 10 mM $MgCl_2$, 10 mM $MgSO_4$, 20 mM glucose) was added. The electroporated *E. coli* cells were plated in a 200μl and a 50μl aliquot on two LB AMP plates (LB broth (Lennox), 1.8% Bacto Agar (Difco), 100 mg/L Ampicillin). The plates were incubated upside down for about 24 hours at 37°C. Three Ampicillin-resistant colonies were selected at random and submitted for sequence analysis of the insert. Large-scale plasmid DNA was isolated from a sequence-confirmed clone using the Qiagen Maxi kit (Qiagen) according to manufacturer's instructions.

B. Construction of mouse zCytor17Lig(m)-CEE/pZMP21

An expression plasmid containing the entire polynucleotide encoding murine zCytor17Lig-CEE (SEQ ID NO:104 and SEQ ID NO:105) was also constructed via homologous recombination using the method described in Example 33A above. The primers used were: (1) ZC41643 (SEQ ID NO:106) (forward, 5' to 3' sense) having a 28bp vector overlap 5' of the insertion point; 21 bp of the 5' end of zcytor17lig(m) and (2) ZC41641 (SEQ ID NO:107) (reverse, 5' to 3' anti-sense) having a 37bp vector

overlap 3' of the insertion point; 3 bp stop codon; 21 bp C-terminal EE tag; 24 bp of the 3' end of zCytor17Lig(m)-CEE. The plasmid was called zcytor17lig(m)-CEE/pZMP21. The polynucleotide sequence of zcytor17lig(m)-CEE is shown in SEQ ID NO:104, and corresponding polypeptide sequence is shown in SEQ ID NO:105.

Example 34

Transfection And Expression Of zcytor17lig-CEE Polypeptides

A. Expression of human zCytor17Lig-CEE/pZMP21 in 293T cells

ZCytor17Lig-CEE was expressed transiently in 293T cells (Stanford University School of Medicine, Stanford, CA; ATCC No. SD-3515) to generate initial purified protein. The day before the transfection, 293T cells were seeded at $6.5x10^4$ cells/cm^2 in 30 T162 culture flasks with a total volume of 30ml of culture media (SL7V4 +5%FBS +1% Pen/Strep) per flask. The cells were allowed to incubate for 24 hours at 37°C.

A DNA/Liposome mixture was prepared as follows: Two 50 ml conical tubes were filled with 25 mLs of transfection media (SL7V4 +1%Pen/Strep) and 1.13 mg of zCytor17Lig-CEE/pZMP21 (Example 33) was added to each. A separate set of two 50 ml conical tubes were filled with 22 ml of transfection media (above) and 3 ml of liposomes (Lipofectamine, Gibco) was added to each. For each set of tubes, one tube of DNA was added to one tube of liposomes and the DNA/liposome mix was incubated for 30 minutes. The two 50 ml conical tubes containing the DNA/liposome mixtures were pooled (about 100 ml) and 300 ml of transfection media was added.
The 30 flasks of the 293T cells were decanted, washed 1x with about 15 ml of PBS, and 12.5 ml of the diluted DNA/liposome mixture was added to each flask. The flasks were incubated for 3 hours at 37°C. After the incubation period, 25 ml of culture media (above) were added to each T162 flask. The transfection media was harvested after approximately 96 hours and was used for protein purification (Example 35).

B. Expression of mouse zCytor17Lig-CEE(m)/pZMP21 in 293T cells

Mouse zCytor17Lig(m)-CEE was expressed transiently in 293T cells as described in Example 34A and cultured media was used for protein purification (Example 35).

Example 35

Purification of Zcytor17lig-CEE from 293T cells

Unless otherwise noted, all operations were carried out at 4°C. The following procedure was used for purifying both mouse and human Zcytor17lig containing C-terminal Glu-Glu (EE) tags (SEQ ID NO:103). Conditioned media from 293T cells expressing Zcytor17lig-CEE (Example 34) was purified. Total target protein concentrations of the conditioned media were determined via SDS-PAGE and Western blot analysis with the anti-EE antibody.

A 5.5 ml column of anti-EE Poros 50 A (PE BioSystems, Framingham, MA) (prepared as described below) was poured in a Waters AP-1, 1 cm x 7cm glass column (Waters, Milford, MA). The column was flow packed and equilibrated on a BioCad Sprint (PE BioSystems, Framingham, MA) with phosphate buffered saline (PBS) pH 7.4. The conditioned media was adjusted with NaCl to 0.3 M and the pH adjusted to 7.2. The conditioned media was then loaded on the column overnight with about 3 ml/minute flow rate. The column was washed with 10 column volumes (CVs) of PBS pH 7.4, and again washed with 3CVs 5X Sigma PBS pH 7.4. It was step eluted with 0.5 M Acetate, 0.5 M NaCl, pH 2.5 at 3 ml/minute. The fraction tubes contained 1ml Tris base (no pH adjustment) to neutralize the elution immediately. The column was again washed for 2CVs with 5X Sigma PBS, pH 7.4 to neutralize the column and then equilibrated in PBS (pH 7.4). Two ml fractions were collected over the entire elution chromatography and absorbance at 280 and 215 nM were monitored; the pass through and wash pools were also saved and analyzed. The 5X PBS and the acid elution peak fractions were analyzed for the target protein via SDS-PAGE Silver staining and Western Blotting with the primary antibody anti-EE and secondary antibody, anti mouse-HRP conjugated. The acid elution fractions of interest were pooled and concentrated from 38 ml to 0.8ml using a 5000 Dalton molecular weight cutoff membrane spin concentrator (Millipore, Bedford, MA) according to the manufacturer's instructions.

To separate Zcytor17lig-CEE from aggregated material and any other contaminating co-purifying proteins, the pooled concentrated fractions were subjected

to size exclusion chromatography on a 1.6 x 60 cm (120 ml) Superdex 75 (Pharmacia, Piscataway, NJ) column equilibrated and loaded in PBS at a flow rate of 1.0 ml/min using a BioCad Sprint. Three milliliter fractions were collected across the entire chromatography and the absorbance at 280 and 215 nM were monitored. The peak fractions were characterized via SDS-PAGE Silver staining, and only the most pure fractions were pooled. This material represented purified Zcytor17lig -CEE protein.

On Western blotted, Coomassie Blue and Silver stained SDS-PAGE gels, the Zcytor17lig-CEE was one major band. The protein concentration of the purified material was performed by BCA analysis (Pierce, Rockford, IL) and the protein was aliquoted, and stored at -80°C according to standard procedures.

To prepare PorosA50 anti-EE, a 65 ml bed volume of Poros A50 (PE Biosystems) was washed with 100 ml of water and then 0.1 M triethanolamine, pH 8.2 (TEA, ICN, Aurora, Ohio), 1 M Na_2S0_4, pH 8.8 containing 0.02% sodium azide using a vacuum flask filter unit. The EE monoclonal antibody solution, at a concentration of 2 mg/ml in a volume of 300 ml, was mixed with the washed resin in a volume of 250 ml. After an overnight incubation at room temperature, the unbound antibody was removed by washing the resin with 5 volumes of 200 mM TEA, 1 M Na_2S0_4, pH 8.8 containing 0.02% sodium azide as described above. The resin was resuspended in 2 volumes of TEA, 1 M Na_2S0_4, pH 8.8 containing 0.02% sodium azide and transferred to a suitable container. Three ml of 25 mg/ml (68mM) Disuccinimidyl suberate (in DMSO supplied by Pierce, Rockford, IL) is added and the solution is incubated for three hours at room temperature. Nonspecific sites on the resin were then blocked by incubating for 10 min at room temperature with 5 volumes of 20 mM ethanolamine (Sigma, St. Louis, MO) in 200 mM TEA, pH 8.8 using the vacuum flask filter unit. The resin is washed with PBS, pH 7.4, followed by 0.1 M Glycine, pH 3 and then neutralized with 10X PBS. After washing with distilled water, the final coupled anti-EE Poros-A 50 resin was stored at 4°C in 20% Ethanol.

Example 36

N-terminal sequencing of human and mouse Zcytor17lig

A. N-terminal sequencing of human Zcytor17lig

Standard automated N-terminal polypeptide sequencing (Edman degradation) was performed using reagents from Applied Biosystems. N-terminal sequence analysis was performed on a Model 494 Protein Sequencer System (Applied Biosystems, Inc., Foster City, CA). Data analysis was performed with Model 610A Data Analysis System for Protein Sequencing, version 2.1a (Applied Biosystems).

A purified human zcytor17lig-CEE sample (Example 35) was supplied. The sample was loaded onto a prepared glass fiber filter for n-terminal sequencing. The glass fiber filter was prepared by precycling it with Biobrene ™.

N-terminal sequence analysis of the secreted human zcytor17lig polypeptide did not verify the predicted cleavage site of the signal sequence but resulted in a mature start at residue 27(Leu) in SEQ ID NO:2 of the human zcytor17lig precursor sequence.

B. N-terminal sequencing of human Zcytor17lig

Standard automated N-terminal polypeptide sequencing (Edman degradation) was performed using reagents from Applied Biosystems. N-terminal sequence analysis was performed on a Model 494 Protein Sequencer System (Applied Biosystems, Inc., Foster City, CA). Data analysis was performed with Model 610A Data Analysis System for Protein Sequencing, version 2.1a (Applied Biosystems).

A purified mouse zcytor17lig-CEE sample was supplied as captured on Protein G Sepharose/anti-EE beads (Example 35). The beads were placed in reducing SDS PAGE sample buffer and on a boiling water bath before running on SDS PAGE, using a Novex SDS PAGE system (4-12% Bis-Tris MES NuPAGE; Invitrogen) as per manufacturer's instructions. The gel was electrotransferred to a Novex PVDF membrane (Invitrogen), and Coomassie blue stained (Sigma, St. Louis, MO) using standard methods. Corresponding anti-EE Western blots were performed to identify the zcytor17lig band for N-terminal protein sequencing. The mouse anti-EE IgG HRP conjugated antibody used was produced in house.

N-terminal sequence analysis of the secreted mouse zcytor17lig polypeptide verified the predicted cleavage site of the signal sequence resulting in a mature start at 31 (Ala) in reference to SEQ ID NO:11 and SEQ ID NO:91 of the mouse

zcytor17lig precursor sequence.

Example 37

Cos cell binding assay

A binding assay was used to test the binding of the zcytor17lig to receptors comprising zcytor17 receptor, such as the zcytor17 receptor or receptor heterodimers and trimers comprising zcytor17 receptor (e.g., zcytor17/OSMR, zcytor17/WSX-1, or zcytor17/OSMR/WSX-1, or other Class I cytokine receptor subunits). Zcytor17 receptor plasmid DNA was transfected into COS cells and transfected COS cells were used to assess binding of the zcytor17lig to receptors comprising zcytor17 receptor as described below.

A. COS Cell Transfections

The COS cell transfection was performed as follows: Mix 800 ng receptor plasmid DNA in the following combinations: pZp7pX/zcytor17 alone; pZp7Z/WSX-1 alone; pZp7NX/OSMR alone; pZp7pX/zcytor17 + pZp7NX/OSMR; pZp7pX/zcytor17 + pZp7Z/WSX-1; pZp7NX/OSMR + pZp7Z/WSX-1; pZp7pX/zcytor17 + pZp7NX/OSMR + pZp7Z/WSX-1) and 4ul Lipofectamine™ in 80 μl serum free DMEM media (55mg sodium pyruvate, 146mg L-glutamine, 5mg transferrin, 2.5mg insulin, 1μg selenium and 5mg fetuin in 500ml DMEM), incubate at room temperature for 30 minutes and then add 320 μl serum free DMEM media. Add this 400ul mixture onto 2 x10^5 COS cells/well plated on 12-well tissue culture plate (fibronectin-coated) and incubate for 5 hours at 37°C. Add 500ul 20% FBS DMEM media (100 ml FBS, 55 mg sodium pyruvate and 146mg L-glutamine in 500ml DMEM) and incubate overnight.

B. Binding Assay

The binding assay was performed as follows: media was rinsed off cells with PBS + 0.1% BSA, and then cells were blocked for 60 minutes with the same solution. The cells were then incubated for 1 hour in PBS + 0.1% BSA with 1.0 μg/ml zcytor17ligCEE purified protein. Cells were then washed with PBS + 0.1% BSA and

incubated for another hour with 1:1000 diluted mouse anti-GluGlu antibody. Again cells were washed with PBS + 0.1% BSA, then incubated for 1 hour with 1:200 diluted goat anti-mouse-HRP conjugated antibody.

Positive binding was detected with fluorescein tyramide reagent diluted 1:50 in dilution buffer (NEN kit) and incubated for 4-6 minutes, and washed with PBS + 0.1% BSA. Cells were fixed for 15 minutes with 1.8% Formaldehyde in PBS, then washed with PBS + 0.1%BSA. Cells were preserved with Vectashield Mounting Media (Vector Labs Burlingame, CA) diluted 1:5 in PBS. Cells were visualized using a FITC filter on fluorescent microscope.

Positive binding was detected for cells transfected with zcytor17 only, zcytor17+OSMRbeta, zcytor17+WSX-1, and zcytor17+OSMRbeta+WSX-1. No binding was detected for cells transfected with WSX-1 + OSMRbeta, with OSMRbeta only, or with WSX-1 only.

Example 38

Mouse zcytor17lig activates mouse zcytor17/OSMRbeta receptor-in luciferase assay

A. Cloning of full-length mouse zcytor17 and mouse OSMRbeta for expression

A mouse testes cDNA library was screened for a full-length clone of mouse zcytoR17. The library was plated at 65,500 cfu/plate on 24 LB + Amp plates. Filter lifts were prepared using Hybond N (Amersham-Pharmacia Biotech, Inc., Piscataway, NJ) on a total of approximately 1.6 million colonies. The filters were marked with a hot needle for orientation and then denatured for 6 minutes in 0.5 M NaOH and 1.5 M Tris-HCl, pH 7.2. The filters were then neutralized in 1.5 M NaCl and 0.5 M Tris-HCl, pH 7.2 for 6 minutes. The DNA was affixed to the filters using a UV crosslinker (Stratalinker®, Stratagene, La Jolla, CA) at 1200 joules. The filters were then left to dry overnight at room temperature.

The next day, the filters were pre-washed at 65°C in pre-wash buffer consisting of 0.25X SSC, 0.25% SDS and 1mM EDTA. Cell debris was manually removed using Kimwipes® (Kimberly-Clark) and the solution was changed 3 times over a period of 1 hour. Filters were air dried and stored at room temperature until needed. The filters were then prehybridized for approximately 3 hours at 63°C in 20 ml

of ExpressHyb™Hybridization Solution (Clontech, Palo Alto, CA).

Probe B (Example 31) was generated by PCR from human zcytoR17 template using oligonucleotide primers ZC27,895 (SEQ ID NO:20) and ZC28,917 (SEQ ID NO:83) and was radioactively labeled with ^{32}P using a commercially available kit (Megaprime DNA Labeling System; Amersham Pharmacia Biotech, Piscataway, NJ) according to the manufacturer's instructions. The probe was purified using a Stratagene™ push column (NucTrap® column; Stratagene, La Jolla, CA). The probe was denatured at 100°C for 15 min and added to ExpressHyb™. Filters were hybridized in 15 ml hybridizing solution containing 1.6 x 10^6 cpm/ml of probe at 63°C overnight. Filters were washed at 55°C in 2X SSC, 0.1 % SDS and 1 mM EDTA and exposed to X-ray film at -80°C for 4 1/2 days. Thirteen positives were picked from the plates as plugs and placed in 1 ml LB +amp in 1.7 ml tubes. Tubes were placed at 4°C overnight. These 13 positives were subjected to two further rounds of purification. The tertiary plates were outgrown at 37°C after filter lifts were taken and single colonies were picked and sent to sequencing. Three of these were determined to contain sequence of the mouse ortholog of zcytoR17.

In addition, a PCR product was generated using CTLL-2 cDNA as a template and oligonucleotides ZC38,239 (SEQ ID NO:123) and ZC38,245 (SEQ ID NO:124) as primers. CTLL-2 is a mouse cytotoxic T lymphocyte cell line (ATCC No. TIB-214). This PCR reaction was run as follows: 1 cycle at 95°C for 1 minute, 30 cycles at 95°C for 15 seconds, 68°C for 3 minutes, then 68°C for 10 minutes; 4°C soak. The PCR reaction used approximately 0.5 ng of cDNA, 20 pmoles of each oligonucleotide, and 1 µl of Advantage II polymerase mix (ClonTech). About 6% of the PCR product was used as a template in a new PCR reaction, as above, except with oligonucleotides ZC38,239 (SEQ ID NO:123) and ZC38,238 (SEQ ID NO:125). This PCR reaction was run as follows: 30 cycles at 94°C for 45 seconds, 65°C for 45 seconds, 72°C for 1 minute, then 72°C for 7 minutes; 10°C soak. Most of the PCR reaction was loaded on a 1.0% agarose gel and the predominant band at approximately 360 bp was excised, the DNA fragment was eluted, and DNA sequencing was performed.

The sequence of the mouse zcytor17 polynucleotide is shown in SEQ ID

NO:126 and the corresponding amino acid sequence shown in SEQ ID NO:127. In addition, a truncated soluble form of the mouse zcytor17 polynucleotide is shown in SEQ ID NO:128 and the corresponding amino acid sequence shown in SEQ ID NO:129.

To obtain a full-length mouse OSMRbeta cDNA, 5' and 3' PCR products were isolated and joined using an internal BamHI site. The PCR primers were designed using the nucleotide sequence SEQ ID NO:134 and include EcoRI and XbaI restriction sites for cloning purposes. The genomic mouse OSMRbeta nucleic acid sequence is shown in SEQ ID NO:134, wherein the coding sequence encompasses residues 780 to 3692 encoding a mouse OSMRbeta 970 amino acid polypeptide, which is shown in SEQ ID NO:135. A degenerate nucleic acid sequence which encodes the polypeptide of SEQ ID NO:135 is shown in SEQ ID NO:136.

A 5' PCR product was generated using an in-house 3T3-L1 (differentiated mouse adipocyte) cDNA library as a template and oligonucleotides ZC41,764 (SEQ ID NO:130) and ZC41,598 (SEQ ID NO:131) as primers. This 5' PCR reaction was run as follows: 30 cycles at 95°C for 45 seconds, 55°C for 45 seconds, 72°C for 1 minute 30 seconds, then 72°C for 7 minutes; 4°C soak. The PCR reaction used approximately 3 μg of plasmid prepared from the cDNA library, 20 pmoles of each oligonucleotide, and five units of *Pwo* DNA polymerase (Roche). About 90% of the 5' PCR product was digested with EcoRI and BamHI and gel purified on a 1.0% agarose gel. The approximately 1446 bp band was excised and used for ligation (see below).

A 3' PCR product was generated using a mouse placenta in-house cDNA library as a template and oligonucleotides ZC41,948 (SEQ ID NO:132) and ZC41,766 (SEQ ID NO:133) as primers. This 3' PCR reaction was run as follows: 30 cycles at 95°C for 45 seconds, 55°C for 45 seconds, 72°C for 1 minute 30 seconds, then 72°C for 7 minutes; 4°C soak. The PCR reaction used approximately 3 μg of plasmid prepared from the cDNA library, 20 pmoles of each oligonucleotide, and five units of *Pwo* DNA polymerase (Roche). About 90% of the 3' PCR product was digested with BamHI and XbaI and gel purified on a 1.0% agarose gel. The approximately 2200 bp band was excised and used for ligation along with the 5' PCR product (described above) to the

expression vector pZP-5Z digested with EcoRI and XbaI. The three-part ligation was performed with the 5' EcoRI to BamHI fragment above, the 3' BamHI to XbaI fragment, and the expression vector pZP-5Z digested with EcoRI and XbaI. This generated a pZP-5Z plasmid containing a full-length cDNA for mouse OSMRbeta (nucleotides 780 to 3692 of SEQ ID NO:134), designated pZP-5Z/OSMRbeta. The full length mouse OSMRbeta cDNA in pZP5Z/OSMRbeta has two amino acid insertions from SEQ ID NO:135. There is a duplication of amino acid Glycine at position 370 and a duplication of amino acid Glutamic Acid at position 526. Plasmid pZP-5Z is a mammalian expression vector containing an expression cassette having the CMV promoter, multiple restriction sites for insertion of coding sequences, and a human growth hormone terminator. The plasmid also has an *E. coli* origin of replication, a mammalian selectable marker expression unit having an SV40 promoter, enhancer and origin of replication, a zeocin resistance gene and the SV40 terminator.

The resulting transformants were sequenced to confirm the mouse OSMRbeta cDNA sequence.

B. Construction of BaF3/KZ134/zcytor17m, BaF3/KZ134/zcytor17m/OSMRbetam, BHK/KZ134/zcytor17m, and BHK/KZ134/zcytor17m/OSMRbetam cell lines

Stable BaF3/KZ134 and BHK/KZ134 cell lines (Example 20) were transfected with an expression plasmid encoding full-length mouse zcytor17, pZP-7P/zcytor17m (Example 38A), to create BaF3/KZ134/zcytor17m and BHK/KZ134/zcytor17m cells, respectively. The mouse OSMRbeta expression plasmid, pZP-5Z/OSMRbetam (Example 38A), was then transfected into these cells to create BaF3/KZ134/zcytor17m/OSMRbetam and BHK/KZ134/zcytor17m/OSMRbetam cell lines, respectively. Methods were as described in Example 4 with the exception that Baf3/KZ134/zcytor17m and BHK/KZ134/zcytor17m were selected with, in addition to Geneticin, 2ug/ml puromycin while Baf3/KZ134/zcytor17m/OSMRbetam and BHK/KZ134/zcytor17m/OSMRbetam were selected with, in addition to Geneticin, 2ug/ml puromycin and 200 ug/ml zeocin.

Clones were diluted, plated and selected using standard techniques. Clones were screened by luciferase assay (see Example 20, above) using the mouse

zcytor17lig conditioned media or purified mouse zcytor17lig protein (Example 35) as an inducer. Clones with the highest luciferase response (via STAT luciferase) and the lowest background were selected. Stable transfectant cell lines were selected.

C. Mouse Zcytor17lig activates mouse zcytor17 receptor in BaF3/KZ134/zcytor17m/OSMRbetam or BHK/KZ134/zcytor17m/OSMRbetam luciferase assay

Cell lines were plated for luciferase assays as described in Example 20 above. STAT activation of the BaF3/KZ134/Zcytor17m, BaF3/KZ134/zcytor17m/OSMRbetam, BHK/KZ134/zcytor17m, or BHK/KZ134/zcytor17m/OSMRbetam cells was assessed using (1) conditioned media from BHK570 cells transfected with the human zcytor17lig (Example 7), (2) conditioned media from BHK570 cells transfected with the mouse zcytor17lig (Example 18), (3) purified mouse and human zcytor17lig (Example 35), and (4) mIL-3 free media to measure media-only control response. Luciferase assays were performed as described in Example 20.

The results of this assay confirm the STAT reporter response of the BaF3/KZ134/zcytor17m/OSMRbetam and BHK/KZ134/zcytor17m/OSMRbetam cells to the mouse zcytor17lig when compared to either the BaF3/KZ134/zcytor17m cells, the BHK/KZ134/zcytor17m cells or the untransfected BaF3/KZ134 or BHK/KZ134 control cells, and show that the response is mediated through the mouse zcytor17/OSMRbeta receptors. The results also show that the human zcytor17lig does not activate the STAT reporter assay through the mouse receptor complex.

Example 39

Human zcytor17 ligand binding to zcytor17 and zcytor17/OSMRbeta by Flow Cytometry

The biotinylation of human zcytor17L was done as follows: 100μL of zcytor17 at 5.26mg/mL was combined with 30μL of 10mg/mL EZ-link Sulfo-NHS-LC-biotin (Pierce, Rockford, IL) dissolved in ddH_2O. This solution was incubated on a rocker for 30 minutes at room temperature. After biotinylation the solution was

dialyzed in PBS using a Slide-A-Lyzer dialysis cassette.

To test the binding properties of human zcytor17 ligand to different receptor combinations both BHK and BAF3 cells were transfected with expression plasmids using standard techniques well-known in the art. These plasmids were transfected into both cell lines in the following combinations: zcytor17 alone, OSMRbeta alone, and both zcytor17 and OSMRbeta. Transfection was performed as detailed above. Untransfected BHK and BAF3 cells were used as controls. Cells were stained by FACS as follows: 2E5 cells were stained with either: 2.0μg/mL, 100ng/mL, 10ng/mL, 1.0ng/mL, 100pg/mL, 10pg/mL, 1.0pg/mL of biotinylated zcytor17L or left unstained for 30 minutes on ice in FACS buffer (PBS + 2%BSA + 2%NHS (Gemini) + 2%NGS). Cells were washed 1.5 times and then stained with SA-PE (Jackson Immuno Laboratories) at 1:250 for 30 minutes on ice. Cells were then washed 1.5 times with FACS buffer and resuspended in FACS buffer and analyzed by FACS on a BD FACSCaliber using CellQuest software (Becton Dickinson, Mountain View, CA).

Both BHK and BAF3 cells showed that zcytor17 ligand bound to both zcytor17 alone and in combination with OSMRbeta with the binding to the zcytor17/OSMRbeta heterodimer being slightly stronger. No binding was seen in either cell lines expressing OSMRbeta alone. The zcytor17 ligand bound in a concentration dependent manner. The mean fluorescent intensity (MFI) values for the BHK binding are shown below in Table 15.

Table 15

zcytor17 μg/mL	2.0	0.100	0.010	0.001	0.0001	0.00001	0.000001	0.0
BHK C17+OSMRbeta	3780	2126	328	53	17	15	14	13
BHK-C17	3032	1600	244	39	16	15	14	15
BHK-OSMRbeta	13	X	X	X	X	X	X	0
BHK-WT	15	14	13	X	X	X	X	13

zcytor17 μg/mL	10.0	3.33	1.11	0.37	0.12	0.04	0.00
BAF3-C17+OSMRbeta	531	508	489	441	364	247	7
BAF3-OSMRbeta	6	5	5	5	5	5	11
BAF3-WT	13	13	12	12	12	12	13

zcytor17 ng/mL	100.0	10.0	1.0	0.0
BAF3-C17	347	72	17	7

Example 40

Gene Expression Array Analysis of Human Zcytor17lig Treated Cells

RNA was isolated from human zcytor17lig treated A549 cells, zcytor17lig treated SK-LU-1 cells, and untreated control cells using a RNeasy Midi Kit (Qiagen, Valencia, CA) according to the manufactures instructions.

Gene expression profiling of the cells treated with zcytor17lig and the respective control cells was carried out using GEArray Q series cDNA expression arrays (SuperArray Inc., Bethesda, MD). The Q Series cDNA expression arrays contain up to 96 cDNA fragments associated with a specific biological pathway, or genes with similar functions or structural features. Comparison of arrays from treated and control cells allows for a determination of the up and down regulation of specific genes. Probe labeling, hybridization and detection were carried out according to the manufactures instructions. Chemiluminscent signal detection and data acquisition was carried out on a Lumi-Imager workstation (Roche, Indianapolis, IN). The resulting image data was analyzed using ImageQuant 5.2 (Amersham Biosciences, Inc., Piscataway, NJ) and GEArray Analyzer 1.2 (SuperArray Inc., Bethesda, MD) software.

Analysis of the results from the Human Interleukin and Receptor Q series HS-014N arrays, showed, after normalization, an approximate 4.7 fold increase of IL13RA2 signal in the zcytor17lig treated human SK-LU-1 cells and an approximate 2.2 fold increase of the IL13RA2 signal in the zcytor17lig treated human A549 cells.

These results indicate that zcytor17lig significantly up regulated IL13RA2 in the SK-LU-1 and A549 cells. Both of these are established cell lines derived from human lung carcinomas (Blobel et al., *Virchows Arch B Cell Pathol Incl Mol Pathol.*, 1984;*45*(4):407-29). More specifically, A549 is characterized as a human pulmonary epithelial cell line (Lin, et al., *J Pharm Pharmacol.*, 2002 Sep;*54*(9):1271-8; Martinez et al., *Toxicol Sci.*, 2002 Oct;*69*(2):409-23).

Interleukin-13 (IL13), a cytokine secreted by activated T lymphocytes, has been demonstrated to be both necessary and sufficient for the expression of allergic asthma and for use in experimental models of asthma, which include airway hyperresponsiveness, eosinophil recruitment, and mucus overproduction (Wills-Karp et

al., *Science*, 1998;*282*:2258-2261). It has been shown, that selective neutralization of IL13 will ameliorate the asthma phenotype (Grunig et al., *Science*, 1998; *282*:2261-2263). It has also been reported that IL13 is involved in the up regulation of mucin gene MUC8 expression in human nasal polyp epithelium and cultured nasal epithelium (Kimm et al., *Acta Otolaryngol.*, 2002; Sep;*122*(6):638-643; Seong et al., *Acta Otolaryngol.*, 2002; Jun;*122*(4):401-407). MUC8, a major airway mucin glycoprotein, is implicated as playing a role in the pathogenesis of mucus hypersecretion in chronic sinusitis with polps (Seong et al., *Acta Otolaryngol.*, 2002; Jun;*122*(4):401-407).

Functionally, IL13 signals through a receptor complex consisting of the interleukin-13 receptor alpha-1 chain (IL13RA1) and IL-4 receptor alpha (IL4RA) (Daines and Hershey, *J Biol Chem.*, 2002; *22*(12):10387-10393). It has also been shown, that the interleukin-13 receptor alpha-2 (IL13RA2) binds IL13 with high affinity, but by itself (Daines and Hershey, *J Biol Chem.*, 2002; *22*(12):10387-10393). This receptor lacks, however, the cytoplasmic domain necessary for signaling and, therefore, is considered to be a decoy receptor. It has been shown that IL13RA2 is predominately an intracellular molecule that can be quickly mobilized from intracellular stores and surface expressed following cellular treatment with interferon (IFN)-gamma. The surface expression of IL13RA2 after IFN-gamma treatment does not involve protein synthesis and results in diminished IL13 signaling (Daines and Hershey, *J Biol Chem.*, 2002; *22*(12):10387-10393).

The results of the gene expression array analysis for zcytor17lig indicate the action of zcytor17lig to be novel to that of IFN-gamma in that the zcytor17lig treatment of lung epithelial derived cell lines resulted in a significant increase of IL13RA2 gene expression. It is conceivable, therefore, that zcytor17lig treatment would be beneficial in cases where long-term up regulation of IL13RA2 expression and down regulation of IL13 is desired such as in asthma, airway hyperactivity (AHR), and mucin regulation, including chronic sinusitis with polps.

Example 41

Murine zcytor17lig Trangenic Mice

To evaluate the *in vivo* effects of zcytor17lig overexpression, multiple founders of transgenic mice expressing the murine form of the gene were generated, driven by two different promoters: the lymphocyte-specific promoter Eμ/lck, and the ubiquitous promoter, EF1α (Example 22). Serum protein levels range from approximately 20-300 ng/ml. The Eμ/lck promoter generated mice with higher levels of serum protein than those in the EF1α-zcytor17lig transgenic mice.

The zcytor17lig transgenic mice developed a skin phenotype around 4-8 weeks of age. The fur of the transgenic mice became "ruffled," with obvious piloerection and mild to severe hair loss, usually on their backs, sides of the torso, and around their eyes. This phenotype was consistently found in mice with detectable levels of zcytor17lig protein in their serum. Among the founders, 100% incidence rate among the mice expressing the Eμ/lck -driven gene, and a 50% incidence in the EF1α-zcytor17lig transgenic mice was noted, correlating well with the relative levels of zcytor17lig that was detected in their serum. The transgenic skin appeared to be pruritic, as evidenced by the scratching behavior of the mice, sometimes excessive enough to induce excoriation and lesions of the skin, which usually became infected (with at least *Staphylococcus aureus*). The mice were originally identified with metal ear tags, but in most cases, the ear tags were forcibly removed by the mice themselves. This often resulted in severe damage to the external ear. These wounded ears often did not heal properly, as reflected in the presence of long-lasting pustules and crusting, and a seeping, expanding wound would that developed in many of the animals, behind and between their ears. Some of the transgenic mice also developed scabby wounds on their shoulders and neck. Skin lesions were observed in a subset of the animals, generally evolving on areas of skin where hair loss had already been apparent, and were often exacerbated by the scratching behavior of the mice.

RealTime quantitative RT-PCR was used to detect zcytor17lig RNA transcripts in transgenic (but not non-transgenic) skin samples, with the Eμ/lck transgenic skin expressing more zcytor17lig RNA than skin from EF1α-zcytor17lig transgenic mice. The genes encoding the zcytor17 receptor subunits, zcytor17 and OSM-Rbeta, were expressed in the skin of both non-transgenic and zcytor17lig-transgenic mice.

An examination of the lymphoid tissues from a subset of the Eμ/lck-transgenic founders by flow cytometry revealed a significant increase in the proportion of activated T cells in the spleen and lymph nodes of these mice. Two of the four mice analyzed had severely enlarged cervical lymph nodes, possibly due to the presence of lesions on their necks. A subtle increase in spleen weight and a slight increase in monocytes and neutrophils circulating in the blood of the transgenic mice was observed. There was no increase in a variety of cytokines tested, nor were there changes in the circulating serum amyloid A levels in these mice. The effects on the immune cells in the transgenic mice may be a direct or an indirect result of zcytor17lig, or are secondary effects of the skin lesions.

Histopathology was performed on many tissues other than skin, including liver, thymus, spleen, kidney, and testes, and no significant abnormalities in these organs were noted. Analysis of the transgenic skin, however, did reveal a number of alterations, which varied greatly depending upon the source and location of skin (e.g., normal, hairless, or lesional). In many cases, the ears of the transgenic mice had a thickened epidermis as compared to the non-transgenic controls (e.g., approximately 4 layers versus 2 layers), and the underlying tissues contained low to moderate numbers of inflammatory cells, which were primarily mononuclear with occasional neutrophils. The epidermis over the abdomen appeared multifocally slightly thicker in the transgenic, but there was no apparent increase in inflammatory cells in the underlying dermis or subcutis. In the hairless portions of skin from these mice, there were dilated hair follicles that contained some debris but no hair shafts (e.g., the hairs fell out by the roots). In the lesioned areas, there was severe thickening of the epidermis (acanthosis), increased keratin on the surface of the skin (hyperkeratosis), scattered ulcers of varying size and significant numbers of inflammatory cells in the dermis (mainly neutrophils, with varying numbers of macrophages and lymphocytes). The dermis also contained numerous mast cells bordering the lesions. Some of the hair shafts in the lesioned areas of the transgenic skin were in the active stage (anagen), in contrast to many of the hair shafts in "normal" areas which were in the involuting (catagen) to inactive (telogen) stage.

The phenotype of the zcytor17lig transgenic mice strongly resembles that of atopic dermatitis (AD) patients, and mouse models of AD. AD is a common chronic inflammatory disease that is characterized by hyperactivated cytokines of the helper T cell subset 2 (Th2). Zcytor17lig is preferentially expressed by Th2 vs. Th1 cells, which lends further credence to this comparison. Although the exact etiology of AD is unknown, multiple factors have been implicated, including hyperactive Th2 immune responses, autoimmunity, infection, allergens, and genetic predisposition. Key features of the disease include xerosis (dryness of the skin), pruritus (itchiness of the skin), conjunctivitis, inflammatory skin lesions, *Staphylococcus aureus* infection, elevated blood eosinophilia, elevation of serum IgE and IgG1, and chronic dermatitis with T cell, mast cell, macrophage and eosinophil infiltration. Colonization or infection with *S. aureus* has been recognized to exacerbate AD and perpetuate chronicity of this skin disease.

AD is often found in patients with asthma and allergic rhinitis, and is frequently the initial manifestation of allergic disease. About 20% of the population in Western countries suffer from these allergic diseases, and the incidence of AD in developed countries is rising for unknown reasons. AD typically begins in childhood and can often persist through adolescence into adulthood. Current treatments for AD include topical corticosteroids, oral cyclosporin A, non-corticosteroid immunosuppressants such as tacrolimus (FK506 in ointment form), and interferon-gamma. Despite the variety of treatments for AD, many patients' symptoms do not improve, or they have adverse reactions to medications, requiring the search for other, more effective therapeutic agents.

Epithelial cells, which express the heterodimeric receptor for zcytor17lig (zcytoR17 and OSM-Rbeta), are located at the sites (e.g., skin, gut, lung, etc.) of allergen entry into the body and interact closely with dendritic cells (professional antigen presenting cells) *in situ*. Dendritic cells play an important role in the pathogenesis of allergic diseases, and it is possible that zcytor17lig can interact with its receptor on epithelial cells in the skin and lung and influence immune responses in these organs. Zcytor17lig and its receptor(s) may therefore contribute to the pathogenesis of allergic diseases such as AD and asthma. Furthermore, the phenotype

of the zcytor17lig transgenic mice suggests that this ligand may play a role in wound healing, since the mice seem unable to repair damage to their ears, and often bear long-lasting lesions on their backs and sides. An antagonist of zcytor17lig might therefore represent a viable therapeutic for these and other indications.

Example 42

Luciferase Assay on Human Transformed Epithelial Cell Lines via Transient Infection with an Adenoviral STAT/SRE Reporter Gene

A wide variety of human transformed epithelial cell lines (see Table 16 below) were seeded in 96-well flat-bottom plates at 10,000 cell/well in regular growth media as specified for each cell type. The following day, the cells were infected with an adenovirus reporter construct, KZ136, at a multiplicity of infection of 5000. The KZ136 reporter contains the STAT elements in addition to a serum response element. The total volume was 100 ul/well using DMEM supplemented with 2 mM L-glutamine (GibcoBRL), 1 mM Sodium Pyruvate (GibcoBRL) and 1x Insulin-Transferrin-Selenium supplement (GibcoBRL) (hereinafter referred to as serum-free media). Cells were cultured overnight.

The following day, the media was removed and replaced with 100 μl of induction media. The induction media was human zcytor17 ligand diluted in serum-free media at 100ng/ml, 50 ng/ml, 25 ng/ml, 12.5 ng/ml, 6.25 ng/ml, 3.125 ng/ml and 1.56 ng/ml. A positive control of 20% FBS was used to validate the assay and to ensure the infection by adenovirus was successful. The cells were induced for 5 hours at which time the media was aspirated. The cells were then washed in 50 μl/well of PBS, and subsequently lysed in 30 μl/well of 1X cell lysis buffer (Promega). After a 10-minute incubation at room temperature, 25 μl/well of lysate was transferred to opaque white 96-well plates. The plates were then read on the Luminometer using 5-second integration with 40 μl/well injection of luciferase substrate (Promega).

The results revealed the ability of multiple epithelial cell lines to respond to zcytor17 ligand as shown in Table 16 below.

Table 16

Cell Line	Species	Tissue	Morphology	Disease	Fold Induction
A549	Human	Lung	Epithelial	Carcinoma	2x
Sk-Lu-1	Human	Lung	Epithelial	Adenocarcinoma	6x
WI-38	Human	Embryonic Lung	Fibroblast		Negative
MRC-5	Human	Lung	Fibroblast		Negative
DU 145	Human	Prostate	Epithelial	Carcinoma	10x
PZ-HPV-7	Human	Prostate	Epithelial	Transformed with HPV	5x
PC-3	Human	Prostate	Epithelial	Adenocarcinoma	Negative
U2OS	Human	Bone	Epithelial	Osteosarcoma	15.5x
SaOS2	Human	Bone	Epithelial	Osteosarcoma	22x
MG-63	Human	Bone	Fibroblast	Osteosarcoma	Negative
143B	Human	Bone	Fibroblast	Osteosarcoma	3.5x
HOS	Human	Bone	Fibroblast and Epithelial		8x
TRBMeC	Human	Vascular Bone Marrow	Epithelial		2x
HT144	Human	Skin	Fibroblast	Melanoma	5x
C32	Human	Skin		Melanoma	Negative
Sk-Mel-2	Human	Skin	Polygonal	Melanoma	2.7x
WM-115	Human	Skin	Epithelial	Melanoma	2x
HCT-116	Human	Colon	Epithelial	Carcinoma	Negative
HT-29	Human	Colon	Epithelial	Carcinoma	Negative

CA 02473733 2004-07-16

CaCo2	Human	Colon	Epithelial	Adenocarcinoma	3x
HBL-100	Human	Breast	Epithelial		1.5x
ME-180	Human	Cervix	Epithelial	Carcinoma	Negative
HeLa 299	Human	Cervix	Epithelial	Adenocarcinoma	Negative
SK-N-SH	Human	Brain	Epithelial	Neuroblastoma	Negative
U138 MG	Human	Brain	Polygonal	Glioblastoma	Negative
HepG2	Human	Liver	Epithelial	Carcinoma	Negative
Chang liver	Human	Liver	Epithelial		Negative
Sk-Hep-1	Human	Liver	Epithelial	Adenocarcinoma	4x
Int 407	Human	Intestine	Epithelial		Negative
3a-Sub E	Human	Placenta			Negative

Example 43

Cytokine production by human epithelial cell lines cultured with human zcytor17 ligand

Human disease-state epithelial cell lines (A549, human lung epithelial carcinoma; SkLu1, human lung epithelial adenocarcinoma; DU145, human prostate epithelial carcinoma; PZ-HPV-7, human prostate epithelial HPV transformed; U20S, human bone epithelial osteosarcoma) were screened for cytokine production in response to zcytor17 ligand *in vitro*. These cell lines have both zcytor17 and OSMR-beta, identified by RT-PCR, and respond to human zcytor17 ligand when assayed with the adenoviral luciferase reporter construct, KZ136 (Example 42). Cytokine production by these cell lines was determined in response to human zcytor17 ligand in a series of three experiments.

A. Cytokine production by human disease-state epithelial cell lines cultured with human zcytor17lig

Cells were plated at a density of 4.5 x 10^5 cells per well in a 6 well plate (Costar) and cultured in respective growth media. The cells were cultured with test reagents; 100ng/mL zcytor17 ligand, 10ng/mL Interferon gamma (IFN gamma) (R&D Systems, Minneapolis, MN), 10ng/mL Tumor Necrosis Factor alpha (TNF alpha) (R&D Systems, Minneapolis, MN), 10ng/mL IL-1beta (R&D Systems, Minneapolis, MN) or 100ug/mL Lipopolysaccharide (LPS) (Sigma). Supernatants were harvested at 24 and 48 hours and assayed for cytokines; GM-CSF (Granulocyte-Macrophage Colony-Stimulating Factor), IL-1b, IL-6, IL-8, MCP-1 (Macrophage Chemoattractant Protein-1) and TNFa. Multiplex Antibody Bead kits from BioSource International (Camarillo, CA) were used to measure cytokines in samples. Assays were read on a Luminex-100 instrument (Luminex, Austin, TX) and data was analyzed using MasterPlex software (MiraiBio, Alameda, CA). Cytokine production (pg/mL) for each cell line in the 24-hour samples is shown below in Table 17.

Table 17

GM-CSF pg/mL

	A549	SkLu1	DU145	U2OS	PZ-HPV-7
zcytor17L	18.80	10.26	16.19	13.26	14.10
IFN-g	16.19	13.36	11.56	16.26	11.81
IL-1b	104.60	126.44	76.77	338.25	27.32
TNFa	106.67	33.20	58.50	107.09	33.79
LPS	17.64	10.62	11.81	25.47	18.34
control	14.81	8.56	13.26	21.67	13.96

IL-1b pg/mL

	A549	SkLu1	DU145	U2OS	PZ-HPV-7
zcytor17L	26.90	30.17	28.77	29.07	28.00
IFN-g	29.07	35.33	21.96	26.90	26.73
IL-1b	1332.88	1256.17	979.02	1107.35	998.60
TNFa	31.11	33.28	35.33	31.24	25.66
LPS	33.28	28.77	29.07	31.11	31.24
control	28.77	28.77	26.73	31.24	29.07

IL-6 pg/mL

	A549	SkLu1	DU145	U2OS	PZ-HPV-7
zcytor17L	20.09	26.89	193.05	19.37	17.30
IFN-g	17.52	33.64	217.58	27.02	17.63
IL-1b	175.44	5920.19	2375.29	304.08	18.44
TNFa	354.16	1002.51	1612.17	103.58	18.33
LPS	18.06	35.65	162.18	22.42	17.30
control	17.63	27.80	71.23	19.32	17.19

IL-8 pg/mL

	A549	SkLu1	DU145	U2OS	PZ-HPV-7
zcytor17L	86.33	150.81	150.61	45.92	6.81
IFN-g	24.07	72.82	163.31	81.78	1.35
IL-1b	1726.24	4083.12	4407.79	5308.83	124.17
TNFa	3068.68	3811.75	2539.39	3324.02	69.65
LPS	20.28	167.13	230.39	115.08	7.95
control	14.92	109.78	107.27	93.44	9.49

MCP-1 pg/mL

	A549	SkLu1	DU145	U2OS	PZ-HPV-7
zcytor17L	8.97	187.29	26.84	105.15	7.20
IFN-g	7.30	267.99	17.05	88.68	7.71
IL-1b	8.11	8039.84	88.78	3723.81	4.70
TNFa	8.50	7100.37	153.26	3826.80	2.80
LPS	9.40	185.83	22.65	61.62	5.61
control	8.16	167.93	13.68	47.78	5.61

TNFa pg/mL

	A549	SkLu1	DU145	U2OS	PZ-HPV-7
zcytor17L	16.23	17.52	16.67	15.80	17.09
IFN-g	15.80	17.09	15.80	16.65	15.80
IL-1b	16.66	17.09	15.80	17.95	16.23
TNFa	1639.92	1648.83	2975.07	1348.33	3554.82
LPS	16.87	15.80	15.37	17.09	17.52
control	16.23	15.80	15.80	17.09	16.66

All cell lines tested produced GM-CSF and IL-8 in response to stimulation with control cytokines IL-1b and TNFa. Most cell lines produced IL-6 and MCP-1in response to IL-1b and TNFa stimulation. Zcytor17 ligand stimulated IL-6 production in the DU145 cell line compared to control (193pg/mL vs. 71pg/mL). Zcytor17 ligand stimulated 3 of 5 cell lines to produce IL-8 with the greatest effect seen in A549 cells (5 fold), and reduced IL-8 production in U2OS cells by 2 fold. There was a slight effect on MCP-1 production by DU145 and U2OS cells when cultured with zcytor17 ligand.

B. Cytokine production by normal human epithelial cell lines cultured with human zcytor17lig

In addition to the human epithelial cell lines, normal human bronchial epithelial cells (NHBE, Clonetics) were also tested. Cells were plated at a density of 1 x 10^5 cells per well in a 24 well plate and cultured with test reagents; 1000ng/mL, 100ng/mL and 10ng/mL zcytor17 ligand (A760F), 10ng/mL TNFa, 10ng/mL OSM, 10ng/mL IFNa, 10ng/mL TGFb or 10ng/mL Lymphotactin. Supernatants were harvested at 24 and 48 hours and assayed for cytokines; IL-6, IL-8, MCP-1, MIP-1a, RANTES and Eotaxin. Cytokines were assayed as previously described. Cytokine production (pg/mL) for each cell line in the 48-hour samples is shown below in Table 18.

Table 18

IL-6 pg/ml

	A549	DU145	SkLu1	U2OS	NHBE
r17L 1000ng/ml	24.5	56.3	32.1	25.2	64.5
r17lL 100ng/ml	25.0	65.0	31.0	25.4	50.2
r17L 10ng/ml	24.8	51.8	30.2	25.3	54.3
TNFa	272.9	355.4	437.5	36.1	299.3
OSM	26.4	73.5	112.4	25.6	80.4
IFNa	24.6	109.3	33.7	26.4	52.4
TGFb	24.4	102.6	42.7	27.8	268.9
control	24.5	36.3	29.9	25.2	47.9

IL-8 pg/ml

	A549	DU145	SkLu1	U2OS	NHBE
r17L 1000ng/ml	35.0	243.3	45.6	18.6	402.0
r17lL 100ng/ml	31.0	290.7	40.1	21.3	296.0
r17L 10ng/ml	30.4	240.4	33.4	18.9	361.8
TNFa	2809.3	2520.9	1385.2	784.9	1486.3
OSM	37.8	60.6	68.0	22.5	494.6
IFNa	18.9	315.3	39.5	33.1	231.6
TGFb	9.9	77.5	19.6	88.9	246.9
control	10.9	238.0	38.0	39.7	315.8

MCP-1 pg/ml

	A549	DU145	SkLu1	U2OS	NHBE
r17L 1000ng/ml	nd	nd	149.1	81.0	nd
r17lL 100ng/ml	nd	nd	130.6	81.9	nd
r17L 10ng/ml	nd	nd	111.7	49.1	nd
TNFa	nd	22.1	2862.6	1104.7	nd
OSM	nd	17.2	448.2	85.8	nd
IFNa	nd	nd	131.7	10.5	nd
TGFb	nd	1.7	54.5	27.6	nd
control	nd	nd	113.0	1.7	nd

nd = not detected

DU145 cells produced IL-6 in response to zcytor17 ligand, repeating the previous results in Example 43A. However, only A549 and U2OS had similar IL-8 responses as seen Example 43A. SkLu1 and U2OS cells both produced MCP-1 in response to zcytor17 ligand. Cytokine production by NHBE cells was marginal compared to controls.

C. Cytokine production by human disease-state epithelial cell lines co-cultured with human zcytor17lig and IFN gamma

Cells were plated at a density of 2 x 10^5 cells per well in 24 well plate and co-cultured with 10ng/mL IFN gamma +/- zcytor17 ligand at 100ng/mL, 10ng/mL or 1ng/mL. Supernatants were collected at 24 and 48 hours and assayed for IL-8 and MCP-1 as described above. Cytokine production (pg/mL) for each cell line in the 24-hour samples is shown below in Table 19.

Table 19

		IL-8 pg/ml	MCP-1 pg/ml
A549	10ng/mL IFNg+100ng/mL r17L	86.7	nd
	10ng/mL IFNg+10ng/mL r17L	75.1	nd
	10ng/mL IFNg+1ng/mL r17L	63.6	nd
	10ng/ml IFNg	35.4	nd
	control	36.6	nd
DU145	10ng/mL IFNg+100ng/mL r17L	102.3	nd
	10ng/mL IFNg+10ng/mL r17L	92.9	nd
	10ng/mL IFNg+1ng/mL r17L	79.9	nd
	10ng/ml IFNg	70.7	nd
	control	79.4	nd
SkLu1	10ng/mL IFNg+100ng/mL r17L	152.2	604.9
	10ng/mL IFNg+10ng/mL r17L	194.4	870.7
	10ng/mL IFNg+1ng/mL r17L	138.7	585.4
	10ng/ml IFNg	170.8	652.6
	control	203.0	292.3
U2OS	10ng/mL IFNg+100ng/mL r17L	106.8	357.0
	10ng/mL IFNg+10ng/mL r17L	108.2	347.7
	10ng/mL IFNg+1ng/mL r17L	109.9	293.3
	10ng/ml IFNg	118.8	159.8
	control	146.8	7.0

A549 cells produced IL-8 in response to zcytor17 ligand, however there was no effect of co-culturing cells with the addition of IFN gamma. U2OS cells made 20 fold more MCP-1 when cultured with IFNg and 50 fold more MCP-1 when cultured with IFN gamma + zcytor17 ligand.

Example 44

Zcytor17lig Effects on ^{3}H-TdR Incorporation in DU145 Prostate Epithelial Carcinoma Cells

Cells were seeded in 96-well tissue clusters (Falcon) at a density of 25,000/well in MEM (Life Technologies) growth medium supplemented with glutamine, pyruvate, non-essential amino acids (Life Technologies) and 10% fetal bovine serum (Hyclone). At confluence (24 hours later), cells were switched to growth arrest media by substituting 0.1% BSA (Life Technologies) for serum. After 48 hours to achieve cell synchronization, the growth-arrest medium was replaced with fresh medium. Then, human recombinant zcytor17lig (test reagent) was added at various

concentrations (from 0.24 to 60 ng/mL) (see Table 16 below), to test for the effect of the protein on basal DNA replication. Some wells received 2.5% FBS (Hyclone) in addition to zcytor17Ligand, in order to test effect of the protein on elevated levels of TdR incorporation. FBS 10% and 20ng/ml Platelet Derived Growth Factor-BB (PDGF-BB) (R&D) were used as positive control.

Eighteen hours following addition of zcytor17Ligand and the rest of the test reagents, cells were pulsed with 250 nCi/mL [^{3}H]-thymidine (NEN) for 4 hours. Following the 4-hour pulse, media were discarded and 100 μL trypsin solution (Life Technologies) was added in each well to dislodge the cells. The radioactivity incorporated by DU145 was determined by harvesting the cells with a Packard Filtermate 196 cell harvester and by counting the incorporated label using a Packard TopCount NXT microplate scintillation counter.

As can be seen in Table 20 below, zcytor17lig induced thymidine incorporation in quiescent cells (in 0.1% BSA) in a concentration-dependent manner. This effect reached 2.5-fold of the BSA control at the highest concentration used, 60 ng/mL. In addition, this effect of zcytor17lig was also detectable when the baseline incorporation was elevated by the addition of 2.5% FBS (in this series as potent a mitogen as 10% FBS). These results therefore indicate that under both basal and stimulated conditions zcytor17lig can act as a mitogenic factor for the DU145 carcinoma cells.

Table 16 shows the effects of zcytor17lig on thymidine incorporation by DU145 cells. Results are expressed in cpm/well and numbers are the mean±st.dev of triplicate wells.

Table 20

	0.1%BSA	2.5%FBS
BSA Control	1139±336	4228±600
Zcytor17lig (0.24ng/mL)	1430±136	4894±1037
Zcytor17lig (0.74ng/mL)	1657±32	5038±810
Zcytor17lig (2.22ng/mL)	1646±57	5162±808
Zcytor17lig (6.67ng/mL)	2226±189	6385±1613
Zcytor17lig (20ng/mL)	2168±108	5880±1085
Zcytor17lig (60ng/mL)	2512±111	6165±417
PDGF-BB (20ng/mL)	4094±202	5927±360

Example 45

Expression of huzcytor17Ligand in E. coli

A. Construction of expression vector pRPS01 that expresses huzcytor17Lig/MBP-6H fusion polypeptide

An expression plasmid containing a polynucleotide encoding a huzcytor17lig fused C-terminally to Maltose Binding Protein (MBP) was constructed via homologous recombination. The fusion polypeptide contains an N-terminal approximately 388 amino acid MBP portion fused to the huzcytor17Lig described herein. A fragment of huzcytor17lig cDNA was isolated using the PCR method as described herein. Two primers were used in the production of the zcytor17lig fragment in a standard PCR reaction: (1) one containing 40 bp of the vector flanking sequence and 20 bp corresponding to the amino terminus of the huzcytor17lig, and (2) another containing 40 bp of the 3' end corresponding to the flanking vector sequence and 20 bp corresponding to the carboxyl terminus of the huzcytor17lig. Two microliters of the 100 μl PCR reaction was run on a 1.0% agarose gel with 1 x TBE buffer for analysis, and the expected molecular weight fragment was observed. The remaining PCR

reaction was combined with the second PCR tube and precipitated with 400 μl of absolute ethanol. The precipitated DNA was used for recombination into the Sma1 cut recipient vector pTAP98 to produce the construct encoding the MBP-huzcytor17lig fusion, as described below.

The vector pTAP98 was constructed using yeast homologous recombination. One hundred nanograms of EcoR1 cut pMAL-c2 was recombined with 1μg Pvu1 cut pRS316, 1μg linker, and 1μg Sca1/EcoR1 cut pRS316 were combined in a PCR reaction. PCR products were concentrated via 100% ethanol precipitation. The competent yeast cell (*S. cerevisiae*) strain, SF838-9Dα, was combined with10 μl of a mixture containing approximately 1 μg of the huzcytor17lig PCR product (above) and 100 ng of SmaI digested pTAP98 vector, and electroporated at 0.75 kV, 25 μF and ∞ ohms. The resulting reaction mixture was plated onto URA-D plates and incubated at 30°C.

After 48 hours, the Ura+ yeast transformants from a single plate were selected. DNA was isolated and transformed into electrocompetent *E. coli* cells (e.g., MC1061, Casadaban et. al. *J. Mol. Biol. 138*, 179-207). The resulting *E. coli* cells were plated on MM/CA +AMP 100 mg/L plates (Pryor and Leiting, *Protein Expression and Purification 10*:309-319, 1997) using standard procedures. Four individual clones were harvested from the plates and inoculated into MM/CA with 100 μg/ml Ampicillin for two hours at 37°C. One milliliter of each of the culture was induced with 1mM IPTG. Approximately 2-4 hours later, 250 μl of each induced culture was mixed with 250 μl acid washed glass beads and 250 μl Thorner buffer with 5% βME and dye (8M urea, 100 mM Tris pH7.0, 10% glycerol, 2mM EDTA, 5% SDS). Samples were vortexed for one minute and heated to 65°C for 10 minutes. Twenty microliters of each sample was loaded per lane on a 4%-12% PAGE gel (NOVEX). Gels were run in 1XMES buffer. The positive clones were designated pRPS01 and subjected to sequence analysis.

One microliter of sequencing DNA was used to transform electrocompetent *E. coli* cell strain MC1061. The cells were electropulsed at 2.0 kV, 25 μF and 400 ohms. Following electroporation, cells were rescued 0.6 ml SOC and grown on LB+Amp plates at 37 °C overnight, with 100 mg/L Ampicillin. Four cultures were induced with ITPG and screened for positives as described above. The positive

clones were expanded for protein purification of the huzcytor17lig/MBP-6H fusion protein using standard techniques.

B. Purification of huzcytor17Lig/MBP-6H from E. coli fermentation

Unless otherwise noted, all operations were carried out at 4°C. The following procedure was used to purify recombinant huzcytor17Lig/MBP-6H polypeptide. *E. coli* cells containing the pRPS01 construct and expressing huzcytor17Lig/MBP-6H, were constructed using standard molecular biology methods and cultured in 50.0 g/L SuperBroth II (12 g/L Casien, 24 g/L Yeast Extract, 11.4 g/L di-potassium phosphate, 1.7 g/L Mono-potassium phosphate; Becton Dickenson, Cockeysville, MD), 5 g/L glycerol and 5 mL/L 1M Magnesium Sulfate. Twenty grams of cells were harvested and frozen for protein purification.

The thawed cells were resuspended in 500 mL Amylose Equilibration buffer (20mM Tris, 100mM NaCl, pH 8.0). A French Press cell breaking system (Constant Systems Ltd., Warwick, UK) with a temperature setting of -7°C to -10°C and 30K PSI was used to lyse the cells. The resuspended cells were assayed for breakage by A_{600} readings before and after cycling through the French Press. The processed cell suspension was pelleted at 10,000G for 30 minutes to remove the cellular debris and the supernatant was harvested for protein purification.

A 25 ml column of Amylose resin (New England Biolabs, Beverly, MA) (prepared as described below) was poured into a Bio-Rad, 2.5 cm D x 10 cm H glass column. The column was packed and equilibrated by gravity with 10 column volumes (CVs) of Amylose Equilibration buffer. The processed cell supernatant was batch loaded to the Amylose resin overnight, with rocking. The resin was returned to the Bio-Rad column and washed with 10 CV's of Amylose Equilibration buffer by gravity. The column was eluted with ~2 CV of Amylose Elution buffer (Amylose Equilibration buffer + 10 mM Maltose, Fluka Biochemical, Switzerland) by gravity. Ten 5 mL fractions were collected over the elution profile and assayed for Absorbance at 280 and 320 nM. The Amylose resin was regenerated with 1 CV of distilled H_2O, 5 CVs of 0.1% (w/v) SDS (Sigma), 5 CVs of distilled H_2O, 5 CVs of Amylose Equilibration buffer and finally 1 CV of Amylose Storage buffer (Amylose Equilibration buffer +

0.02% Sodium Azide). The regenerated column was stored at 4°C.

Elution profile fractions of interest were pooled and dialyzed in a 10K dialysis chamber (Slide-A-Lyzer, Pierce Immunochemicals) against 4 x 4L PBS pH 7.4 (Sigma) over an 8 hour time period to remove low molecular weight contaminants, buffer exchange and desalt. Following dialysis, the material harvested represented the purified huzcytor17Lig/MBP-6H polypeptide. The purified huzcytor17Lig/MBP-6H polypeptide was filter sterilized and analyzed via SDS-PAGE Coomassie staining for an appropriate molecular weight product. The concentration of the huzcytor17Lig/MBP-6H polypeptide was determined by BCA analysis to be 1.28 mg/mL.

Example 46

Human zcytor17lig Polyclonal Antibody

A. Preparation and Purification

Polyclonal antibodies were prepared by immunizing 2 female New Zealand white rabbits with the purified recombinant protein hzcytor17L/MBP-6H (Example 45). The rabbits were each given an initial intraperitoneal (IP) injection of 200 μg of purified protein in Complete Freund's Adjuvant followed by booster IP injections of 100 μg protein in Incomplete Freund's Adjuvant every three weeks. Seven to ten days after the administration of the second booster injection (3 total injections), the animals were bled and the serum was collected. The animals were then boosted and bled every three weeks.

The hzcytor17L/MBP-6H specific rabbit serum was pre-adsorbed of anti-MBP antibodies using a CNBr-SEPHAROSE 4B protein column (Pharmacia LKB) that was prepared using 10 mg of non-specific purified recombinant MBP-fusion protein per gram of CNBr-SEPHAROSE. The hzcytor17L/MBP-6H-specific polyclonal antibodies were affinity purified from the pre-adsorbed rabbit serum using a CNBr-SEPHAROSE 4B protein column (Pharmacia LKB) that was prepared using 10 mg of the specific antigen purified recombinant protein hzcytor17L/MBP-6H. Following purification, the polyclonal antibodies were dialyzed with 4 changes of 20 times the antibody volume of PBS over a time period of at least 8 hours. Hzcytor17-Ligand-specific antibodies were characterized by ELISA using 500 ng/ml of the

purified recombinant proteins hzcytor17L/MBP-6H or hzcytor17L-CEE produced in a baculovirus expression system as antibody targets. The lower limit of detection (LLD) of the rabbit anti-hzcytor17L/MBP-6H affinity purified antibody is 100 pg/ml on its specific purified recombinant antigen hzcytor17L/MBP-6H and 500 pg/ml on purified recombinant hzcytor17L-CEE produced in a baculovirus expression system.

B. SDS-PAGE and Western blotting analysis of Rabbit Anti-human ZcytoR17lig MBP-6H antibody

Rabbit Anti-human ZcytoR17lig MBP-6H antibody was tested by SDS-PAGE (NuPage 4-12%, Invitrogen, Carlsbad, CA) with coomassie staining method and Western blotting using goat anti-rabbit IgG-HRP. Human and mouse zcytor17lig purified protein (200-25 ng) was electrophoresed using an Invitrogen Novex's Xcell II mini-cell, and transferred to nitrocellulose (0.2 mm; Invitrogen, Carlsbad, CA) at room temperature using Novex's Xcell blot module with stirring according to directions provided in the instrument manual. The transfer was run at 300 mA for one hour in a buffer containing 25 mM Tris base, 200 mM glycine, and 20% methanol. The filter was then blocked with Western A buffer (in house, 50 mM Tris, pH 7.4, 5 mM EDTA, pH 8.0, 0.05% Igepal CA-630, 150 mM NaCl, and 0.25% gelatin) overnight with gentle rocking at 4°C. The nitrocellulose was quickly rinsed, then the rabbit anti-human zcytoR17lig MBP-6H (1:1000) was added in Western A buffer. The blot was incubated for 1.5 hours at room temperature with gentle rocking. The blot was rinsed 3 times for 5 minutes each in Western A, then goat anti-rabbit IgG HRP antibody (1:5000) was added in Western A buffer. The blot was incubated for 1 hour at room temperature with gentle rocking. The blot was rinsed 3 times for 5 minutes each in Western A, then quickly rinsed in H_20. The blot was developed using commercially available chemiluminescent substrate reagents (ECLWestern blotting detection reagents 1 and 2 mixed 1:1; reagents obtained from Amersham Pharmacia Biotech, Buckinghamshire, England) and the blot was exposed to x-ray film for up to 5 minutes.

The purified human zcytor17lig appeared as a large band at about 30 kDa and a smaller band at about 20 kDa under reduced conditions. The mouse zcytor17lig was not detected by the rabbit anti-human zcytor17lig antibody.

Example 47

Zcytor17lig Effects on U937 Monocyte Adhesion

to Transformed Bone Marrow Endothelial Cell (TRBMEC) Monolayer

Transformed Bone Marrow Endothelial Cells (TRBMEC) were seeded in 96-well tissue clusters (Falcon) at a density of 25,000/well in medium M131 (Cascade Biologics) supplemented with Microvascular Growth Supplement (MVGS) (Cascade Biologics). At confluence (24 hours later), cells were switched to M199 (Gibco-Life Technologies) supplemented with 1% Fetal Bovine Serum (Hyclone). Human recombinant zcytor17lig (test reagent) was added at various concentrations (from 0.4 to 10 ng/mL) (see Table 21 below), to test for the effect of the protein on immune cell-endothelial cell interactions resulting in adhesion. Some wells received 0.3ng/ml Tumor Necrosis Factor (TNFalpha R&D Systems), a known pro-inflammatory cytokine, in addition to zcytor17lig, to test an effect of the protein on endothelial cells under inflammatory conditions. TNFalpha at 0.3ng/ml alone was used as positive control and the concentration used represents approximately 70% of the maximal TNFalpha effect in this system, i.e., it does not induce maximal adherence of U937 cells (a human monocyte-like cell line) to the endothelium. For this reason, this setup can detect both upregulation and downregulation of the TNFalpha effects. Basal levels of adhesion both with and without TNFalpha were used as baseline to assess effect of test reagents.

After overnight incubation of the endothelial cells with the test reagents (zcytor17ligand ± TNFalpha), U937 cells, stained with 5μM Calcein-AM fluorescent marker (Molecular Probes), the cells were suspended in RPMI 1640 (no phenol-red) supplemented with 1% FBS and plated at 100,000 cells/well on the rinsed TRBMEC monolayer. Fluorescence levels at excitation/emission wavelengths of 485/538nm (Molecular Devices micro-plate reader, CytoFluor application) were measured 30 minutes later, before and after rinsing the well three times with warm RPMI 1640 (no phenol-red), to remove non-adherent U937. Pre-rinse (total) and post-rinse (adherence-specific) fluorescence levels were used to determine percent adherence (net

adherent/net total x 100 = % adherence).

As can be seen in Table 21 below, zcytor17lig when added alone affected the basal adherence of U937 cells to the endothelial monolayers at the concentration range used (less than 2-fold increases, $p<0.01$ by ANOVA test). By itself, the positive control, 0.3ng/mL TNFalpha, increased the adherence of U937 cells from a basal 5.8% to 35% (6-fold). In the presence of TNFalpha, zcytor17lig synergized with TNFalpha and further enhanced U937 adhesion in a concentration-dependent manner between 0.4 and 10ng/mL ($p<0.01$ by ANOVA test). At 10ng/mL, zcytor17lig enhanced the effect of TNFalpha by 62%. These results indicate that zcytor17lig may by itself be a pro-inflammatory agent. Zcytor17lig was able to synergize with sub-maximal concentrations of TNFalpha to increase monocyte adherence to endothelial cells. These results also show that endothelial cells, especially when exposed to pro-inflammatory cytokines such as TNFalpha, are a likely target tissue of zcytor17lig action. The consequence of zcytor17ligand on endothelial cells may be to heighten monocyte or macrophage adhesion to a site of proinflammatory activity. Activated monocytes and macrophages are important in many inflammatory diseases. Therefore inhibition of monocyte/macrophage adhesions may provide a therapeutic rationale for zcytor17ligand antagonists. This data would support the use of zcytor17 ligand antagonists for treatment lung diseases, vascular diseases, autoimmunity, tumor metastasis, disease involving allergic reactions, wound healing and diseases of the skin including contact, allergic or non-allergic dermatistic or psoriasis and inflammatory bowel disease. Table 21 shows the effects of zcytor17lig on U937 monocyte adhesion to TRBMEC endothelial monolayers. Results are expressed in percent adhesion and numbers are the mean±st.dev of triplicate wells.

Table 21

	Basal	*0.3ng/mL TNFalpha*
Basal	5.8±1.2	35±5.5
zcytor17lig 0.4 ng/mL	9±0.7	44.7±2.5
zcytor17lig 1.1 ng/mL	10.4±0.8	45.2±0.6
zcytor17lig 3.3 ng/mL	7.9±1.7	51.1±4
zcytor17lig 10 ng/mL	9.5±0.5	56.6±3.9

From the foregoing, it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

SEQUENCE LISTING

<110> Zymogenetics

<120> CYTOKINE RECEPTOR ZCYTOR17 MULTIMERS

<130> 02-02PC

<150> US 60/435,361
<151> 2002-12-19

<150> US 60/389,108
<151> 2002-06-14

<150> US 60/350,325
<151> 2002-01-18

<160> 183

<170> FastSEQ for Windows Version 4.0

<210> 1
<211> 904
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (28)...(519)

<400> 1
ctgaagctgg ccttgctctc tctcgcc atg gcc tct cac tca ggc ccc tcg acg 54
Met Ala Ser His Ser Gly Pro Ser Thr
1 5

tct gtg ctc ttt ctg ttc tgc tgc ctg gga ggc tgg ctg gcc tcc cac 102
Ser Val Leu Phe Leu Phe Cys Cys Leu Gly Gly Trp Leu Ala Ser His
10 15 20 25

acg ttg ccc gtc cgt tta cta cga cca agt gat gat gta cag aaa ata 150
Thr Leu Pro Val Arg Leu Leu Arg Pro Ser Asp Asp Val Gln Lys Ile
30 35 40

gtc gag gaa tta cag tcc ctc tcg aag atg ctt ttg aaa gat gtg gag 198
Val Glu Glu Leu Gln Ser Leu Ser Lys Met Leu Leu Lys Asp Val Glu
45 50 55

gaa gag aag ggc gtg ctc gtg tcc cag aat tac acg ctg ccg tgt ctc 246
Glu Glu Lys Gly Val Leu Val Ser Gln Asn Tyr Thr Leu Pro Cys Leu
60 65 70

agc cct gac gcc cag ccg cca aac aac atc cac agc cca gcc atc cgg 294

```
Ser Pro Asp Ala Gln Pro Pro Asn Asn Ile His Ser Pro Ala Ile Arg
     75                  80                  85

gca tat ctc aag aca atc aga cag cta gac aac aaa tct gtt att gat   342
Ala Tyr Leu Lys Thr Ile Arg Gln Leu Asp Asn Lys Ser Val Ile Asp
 90                  95                 100                 105

gag atc ata gag cac ctc gac aaa ctc ata ttt caa gat gca cca gaa   390
Glu Ile Ile Glu His Leu Asp Lys Leu Ile Phe Gln Asp Ala Pro Glu
                110                 115                 120

aca aac att tct gtg cca aca gac acc cat gaa tgt aaa cgc ttc atc   438
Thr Asn Ile Ser Val Pro Thr Asp Thr His Glu Cys Lys Arg Phe Ile
            125                 130                 135

ctg act att tct caa cag ttt tca gag tgc atg gac ctc gca cta aaa   486
Leu Thr Ile Ser Gln Gln Phe Ser Glu Cys Met Asp Leu Ala Leu Lys
        140                 145                 150

tca ttg acc tct gga gcc caa cag gcc acc act taaggccatc tcttcctttc 539
Ser Leu Thr Ser Gly Ala Gln Gln Ala Thr Thr
    155                 160

ggattggcag gaacttaagg agccttaaaa agatgaccga cagctaagtg tgggaactct 599
gccgtgattc cttaagtaca tttttccaat gaataatctc agggacccct catatgggct 659
agtcccggga gggctgagat gtgaatttgt gaattacctt gaaaaacatt aggttattgt 719
tattagtctt ggtatttatg gaatgctttt cttctgcagg cttaagtctt acttattata 779
ccctcgtgag ggtgggaggt ggcagctatg ttaatttatt gatatttatt gtactaagag 839
ttgtcaatgc tccctggggg agccctcgga atctatttaa taaattatat tgaatttttc 899
tcata                                                             904
```

<210> 2
<211> 164
<212> PRT
<213> Homo sapiens

<400> 2

```
Met Ala Ser His Ser Gly Pro Ser Thr Ser Val Leu Phe Leu Phe Cys
 1               5                  10                  15
Cys Leu Gly Gly Trp Leu Ala Ser His Thr Leu Pro Val Arg Leu Leu
            20                  25                  30
Arg Pro Ser Asp Asp Val Gln Lys Ile Val Glu Glu Leu Gln Ser Leu
        35                  40                  45
Ser Lys Met Leu Leu Lys Asp Val Glu Glu Glu Lys Gly Val Leu Val
    50                  55                  60
Ser Gln Asn Tyr Thr Leu Pro Cys Leu Ser Pro Asp Ala Gln Pro Pro
65                  70                  75                  80
Asn Asn Ile His Ser Pro Ala Ile Arg Ala Tyr Leu Lys Thr Ile Arg
                85                  90                  95
Gln Leu Asp Asn Lys Ser Val Ile Asp Glu Ile Ile Glu His Leu Asp
            100                 105                 110
Lys Leu Ile Phe Gln Asp Ala Pro Glu Thr Asn Ile Ser Val Pro Thr
        115                 120                 125
Asp Thr His Glu Cys Lys Arg Phe Ile Leu Thr Ile Ser Gln Gln Phe
    130                 135                 140
Ser Glu Cys Met Asp Leu Ala Leu Lys Ser Leu Thr Ser Gly Ala Gln
145                 150                 155                 160
Gln Ala Thr Thr
```

<210> 3
<211> 492

<212> DNA
<213> Artificial Sequence

<220>
<223> human zcytor17lig degenerate polynucleotide of SEQ ID NO:2

<221> misc_feature
<222> (1)...(492)
<223> n = A,T,C or G

<400> 3
atggcnwsnc aywsnggncc nwsnacnwsn gtnytnttyy tnttytgytg yytnggnggn 60
tggytngcnw sncayacnyt nccngtnmgn ytnytnmgnc cnwsngayga ygtncaraar 120
athgtngarg arytncarws nytnwsnaar atgytnytna argaygtnga rgargaraar 180
ggngtnytng tnwsncaraa ytayacnytn ccntgyytnw snccngaygc ncarccnccn 240
aayaayathc aywsnccngc nathmgngcn tayytnaara cnathmgnca rytngayaay 300
aarwsngtna thgaygarat hathgarcay ytngayaary tnathttyca rgaygcnccn 360
garacnaaya thwsngtncc nacngayacn caygartgya armgnttyat hytnacnath 420
wsncarcart tywsngartg yatggayytn gcnytnaarw snytnacnws nggngcncar 480
cargcnacna cn 492

<210> 4
<211> 2903
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (497)...(2482)

<400> 4
tgaaaagaca tgtgtgtgca gtatgaaaat tgagacagga aggcagagtg tcagcttgtt 60
ccacctcagc tgggaatgtg catcaggcaa ctcaagtttt tcaccacggc atgtgtctgt 120
gaatgtccgc aaaacattag tttcactctt gtcgccaggt tggagtacaa tggcacgatc 180
ttggctcact gcaacctctg cctcccgggt tcaagcgatt ctcctgcctc agcctcccga 240
gtagctggga ttacagttaa caataatgca atccatttcc cagcataagt gggtaagtgc 300
cactttgact tgggctgggc ttaaaagcac aagaaaagct cgcagacaat cagagtggaa 360
acactcccac atcttagtgt ggataaatta aagtccagat tgttcttcct gtcctgactt 420
gtgctgtggg aggtggagtt gcctttgatg caaatccttt gagccagcag aacatctgtg 480
gaacatcccc tgatac atg aag ctc tct ccc cag cct tca tgt gtt aac ctg 532
Met Lys Leu Ser Pro Gln Pro Ser Cys Val Asn Leu
1 5 10

ggg atg atg tgg acc tgg gca ctg tgg atg ctc cct tca ctc tgc aaa 580
Gly Met Met Trp Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys
15 20 25

ttc agc ctg gca gct ctg cca gct aag cct gag aac att tcc tgt gtc 628
Phe Ser Leu Ala Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val
30 35 40

tac tac tat agg aaa aat tta acc tgc act tgg agt cca gga aag gaa 676
Tyr Tyr Tyr Arg Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu
45 50 55 60

acc agt tat acc cag tac aca gtt aag aga act tac gct ttt gga gaa 724
Thr Ser Tyr Thr Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu
65 70 75

aaa cat gat aat tgt aca acc aat agt tct aca agt gaa aat cgt gct 772
Lys His Asp Asn Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala
80 85 90

```
tcg tgc tct ttt ttc ctt cca aga ata acg atc cca gat aat tat acc   820
Ser Cys Ser Phe Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr
         95                  100                 105

att gag gtg gaa gct gaa aat gga gat ggt gta att aaa tct cat atg   868
Ile Glu Val Glu Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met
    110                 115                 120

aca tac tgg aga tta gag aac ata gcg aaa act gaa cca cct aag att   916
Thr Tyr Trp Arg Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile
125                 130                 135                 140

ttc cgt gtg aaa cca gtt ttg ggc atc aaa cga atg att caa att gaa   964
Phe Arg Val Lys Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu
                145                 150                 155

tgg ata aag cct gag ttg gcg cct gtt tca tct gat tta aaa tac aca   1012
Trp Ile Lys Pro Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr
            160                 165                 170

ctt cga ttc agg aca gtc aac agt acc agc tgg atg gaa gtc aac ttc   1060
Leu Arg Phe Arg Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe
        175                 180                 185

gct aag aac cgt aag gat aaa aac caa acg tac aac ctc acg ggg ctg   1108
Ala Lys Asn Arg Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu
    190                 195                 200

cag cct ttt aca gaa tat gtc ata gct ctg cga tgt gcg gtc aag gag   1156
Gln Pro Phe Thr Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu
205                 210                 215                 220

tca aag ttc tgg agt gac tgg agc caa gaa aaa atg gga atg act gag   1204
Ser Lys Phe Trp Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu
                225                 230                 235

gaa gaa gct cca tgt ggc ctg gaa ctg tgg aga gtc ctg aaa cca gct   1252
Glu Glu Ala Pro Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala
            240                 245                 250

gag gcg gat gga aga agg cca gtg cgg ttg tta tgg aag aag gca aga   1300
Glu Ala Asp Gly Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg
        255                 260                 265

gga gcc cca gtc cta gag aaa aca ctt ggc tac aac ata tgg tac tat   1348
Gly Ala Pro Val Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr
    270                 275                 280

cca gaa agc aac act aac ctc aca gaa aca atg aac act act aac cag   1396
Pro Glu Ser Asn Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln
285                 290                 295                 300

cag ctt gaa ctg cat ctg gga ggc gag agc ttt tgg gtg tct atg att   1444
Gln Leu Glu Leu His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile
                305                 310                 315

tct tat aat tct ctt ggg aag tct cca gtg gcc acc ctg agg att cca   1492
Ser Tyr Asn Ser Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro
            320                 325                 330

gct att caa gaa aaa tca ttt cag tgc att gag gtc atg cag gcc tgc   1540
Ala Ile Gln Glu Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys
```

```
        335                 340                 345

gtt gct gag gac cag cta gtg gtg aag tgg caa agc tct gct cta gac   1588
Val Ala Glu Asp Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp
    350                 355                 360

gtg aac act tgg atg att gaa tgg ttt ccg gat gtg gac tca gag ccc   1636
Val Asn Thr Trp Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro
365                 370                 375                 380

acc acc ctt tcc tgg gaa tct gtg tct cag gcc acg aac tgg acg atc   1684
Thr Thr Leu Ser Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile
                385                 390                 395

cag caa gat aaa tta aaa cct ttc tgg tgc tat aac atc tct gtg tat   1732
Gln Gln Asp Lys Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr
            400                 405                 410

cca atg ttg cat gac aaa gtt ggc gag cca tat tcc atc cag gct tat   1780
Pro Met Leu His Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr
        415                 420                 425

gcc aaa gaa ggc gtt cca tca gaa ggt cct gag acc aag gtg gag aac   1828
Ala Lys Glu Gly Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn
    430                 435                 440

att ggc gtg aag acg gtc acg atc aca tgg aaa gag att ccc aag agt   1876
Ile Gly Val Lys Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser
445                 450                 455                 460

gag aga aag ggt atc atc tgc aac tac acc atc ttt tac caa gct gaa   1924
Glu Arg Lys Gly Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu
                465                 470                 475

ggt gga aaa gga ttc tcc aag aca gtc aat tcc agc atc ttg cag tac   1972
Gly Gly Lys Gly Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr
            480                 485                 490

ggc ctg gag tcc ctg aaa cga aag acc tct tac att gtt cag gtc atg   2020
Gly Leu Glu Ser Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met
        495                 500                 505

gcc agc acc agt gct ggg gga acc aac ggg acc agc ata aat ttc aag   2068
Ala Ser Thr Ser Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys
    510                 515                 520

aca ttg tca ttc agt gtc ttt gag att atc ctc ata act tct ctg att   2116
Thr Leu Ser Phe Ser Val Phe Glu Ile Ile Leu Ile Thr Ser Leu Ile
525                 530                 535                 540

ggt gga ggc ctt ctt att ctc att atc ctg aca gtg gca tat ggt ctc   2164
Gly Gly Gly Leu Leu Ile Leu Ile Ile Leu Thr Val Ala Tyr Gly Leu
                545                 550                 555

aaa aaa ccc aac aaa ttg act cat ctg tgt tgg ccc acc gtt ccc aac   2212
Lys Lys Pro Asn Lys Leu Thr His Leu Cys Trp Pro Thr Val Pro Asn
            560                 565                 570

cct gct gaa agt agt ata gcc aca tgg cat gga gat gat ttc aag gat   2260
Pro Ala Glu Ser Ser Ile Ala Thr Trp His Gly Asp Asp Phe Lys Asp
        575                 580                 585

aag cta aac ctg aag gag tct gat gac tct gtg aac aca gaa gac agg   2308
```

```
Lys Leu Asn Leu Lys Glu Ser Asp Asp Ser Val Asn Thr Glu Asp Arg
    590                 595                 600

atc tta aaa cca tgt tcc acc ccc agt gac aag ttg gtg att gac aag   2356
Ile Leu Lys Pro Cys Ser Thr Pro Ser Asp Lys Leu Val Ile Asp Lys
605                 610                 615                 620

ttg gtg gtg aac ttt ggg aat gtt ctg caa gaa att ttc aca gat gaa   2404
Leu Val Val Asn Phe Gly Asn Val Leu Gln Glu Ile Phe Thr Asp Glu
                625                 630                 635

gcc aga acg ggt cag gaa aac aat tta gga ggg gaa aag aat ggg act   2452
Ala Arg Thr Gly Gln Glu Asn Asn Leu Gly Gly Glu Lys Asn Gly Thr
            640                 645                 650

aga att ctg tct tcc tgc cca act tca ata taagtgtgga ctaaaatgcg     2502
Arg Ile Leu Ser Ser Cys Pro Thr Ser Ile
        655                 660

agaaaggtgt cctgtggtct atgcaaatta gaaaggacat gcagagtttt ccaactagga 2562
agactgaatc tgtggcccca agagaaccat ctctgaagac tgggtatgtg gtcttttcca 2622
cacatggacc acctacggat gcaatctgta atgcatgtgc atgagaagtc tgttattaag 2682
tagagtgtga aaacatggtt atggtaatag gaacagcttt taaaatgctt ttgtatttgg 2742
gcctttcata caaaaaagcc ataataccat tttcatgtaa tgctatactt ctatactatt 2802
ttcatgtaat actatacttc tatactattt tcatgtaata ctatacttct atactatttt 2862
catgtaatac tatacttcta tattaaagtt ttacccactc a                     2903

<210> 5
<211> 662
<212> PRT
<213> Homo sapiens

<400> 5
Met Lys Leu Ser Pro Gln Pro Ser Cys Val Asn Leu Gly Met Met Trp
 1               5                  10                  15
Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe Ser Leu Ala
            20                  25                  30
Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr Tyr Tyr Arg
        35                  40                  45
Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr Ser Tyr Thr
    50                  55                  60
Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys His Asp Asn
65                  70                  75                  80
Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser Cys Ser Phe
                85                  90                  95
Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile Glu Val Glu
            100                 105                 110
Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr Tyr Trp Arg
        115                 120                 125
Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe Arg Val Lys
    130                 135                 140
Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp Ile Lys Pro
145                 150                 155                 160
Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu Arg Phe Arg
                165                 170                 175
Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala Lys Asn Arg
            180                 185                 190
Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln Pro Phe Thr
        195                 200                 205
Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser Lys Phe Trp
    210                 215                 220
Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu Glu Ala Pro
225                 230                 235                 240
```

Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu Ala Asp Gly
245 250 255
Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val
260 265 270
Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro Glu Ser Asn
275 280 285
Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln Leu Glu Leu
290 295 300
His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser Tyr Asn Ser
305 310 315 320
Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala Ile Gln Glu
325 330 335
Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val Ala Glu Asp
340 345 350
Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val Asn Thr Trp
355 360 365
Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr Thr Leu Ser
370 375 380
Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln Gln Asp Lys
385 390 395 400
Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro Met Leu His
405 410 415
Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu Gly
420 425 430
Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile Gly Val Lys
435 440 445
Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu Arg Lys Gly
450 455 460
Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly Gly Lys Gly
465 470 475 480
Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly Leu Glu Ser
485 490 495
Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala Ser Thr Ser
500 505 510
Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr Leu Ser Phe
515 520 525
Ser Val Phe Glu Ile Ile Leu Ile Thr Ser Leu Ile Gly Gly Gly Leu
530 535 540
Leu Ile Leu Ile Ile Leu Thr Val Ala Tyr Gly Leu Lys Lys Pro Asn
545 550 555 560
Lys Leu Thr His Leu Cys Trp Pro Thr Val Pro Asn Pro Ala Glu Ser
565 570 575
Ser Ile Ala Thr Trp His Gly Asp Asp Phe Lys Asp Lys Leu Asn Leu
580 585 590
Lys Glu Ser Asp Asp Ser Val Asn Thr Glu Asp Arg Ile Leu Lys Pro
595 600 605
Cys Ser Thr Pro Ser Asp Lys Leu Val Ile Asp Lys Leu Val Val Asn
610 615 620
Phe Gly Asn Val Leu Gln Glu Ile Phe Thr Asp Glu Ala Arg Thr Gly
625 630 635 640
Gln Glu Asn Asn Leu Gly Gly Glu Lys Asn Gly Thr Arg Ile Leu Ser
645 650 655
Ser Cys Pro Thr Ser Ile
660

<210> 6
<211> 4171
<212> DNA
<213> Homo sapiens

<220>
<221> CDS

<222> (368)...(3304)

<400> 6

```
gggccgcctc tgcacgtccg ccccggagcc cgcacccgcg ccccacgcgc cgccgaggac 60
tcggcccggc tcgtggagcc cttcgcccgc ggcgtgagta cccccgaccc gcccgtcccc 120
gctctgctcg cgccctgccg ctgcgccgcc ctcggtggct tttccgacgg gcgagccccg 180
tgctgtgcgg gaaagaatcc gacaacttcg cagcccatcc cggctggacg cgaccgggag 240
tgcagcagcc cgttcccctc ctcggtgccg cctctgccca gcgtttgctt ggctgggcta 300
ccacctgcgc tcggacggcg ctcggagggt cctcgccccc ggcctgccta cctgaaaacc 360
agaactg atg gct cta ttt gca gtc ttt cag aca aca ttc ttc tta aca   409
       Met Ala Leu Phe Ala Val Phe Gln Thr Thr Phe Phe Leu Thr
        1               5                  10

ttg ctg tcc ttg agg act tac cag agt gaa gtc ttg gct gaa cgt tta   457
Leu Leu Ser Leu Arg Thr Tyr Gln Ser Glu Val Leu Ala Glu Arg Leu
 15                  20                  25                  30

cca ttg act cct gta tca ctt aaa gtt tcc acc aat tct acg cgt cag   505
Pro Leu Thr Pro Val Ser Leu Lys Val Ser Thr Asn Ser Thr Arg Gln
                 35                  40                  45

agt ttg cac tta caa tgg act gtc cac aac ctt cct tat cat cag gaa   553
Ser Leu His Leu Gln Trp Thr Val His Asn Leu Pro Tyr His Gln Glu
             50                  55                  60

ttg aaa atg gta ttt cag atc cag atc agt agg att gaa aca tcc aat   601
Leu Lys Met Val Phe Gln Ile Gln Ile Ser Arg Ile Glu Thr Ser Asn
         65                  70                  75

gtc atc tgg gtg ggg aat tac agc acc act gtg aag tgg aac cag gtt   649
Val Ile Trp Val Gly Asn Tyr Ser Thr Thr Val Lys Trp Asn Gln Val
     80                  85                  90

ctg cat tgg agc tgg gaa tct gag ctc cct ttg gaa tgt gcc aca cac   697
Leu His Trp Ser Trp Glu Ser Glu Leu Pro Leu Glu Cys Ala Thr His
 95                 100                 105                 110

ttt gta aga ata aag agt ttg gtg gac gat gcc aag ttc cct gag cca   745
Phe Val Arg Ile Lys Ser Leu Val Asp Asp Ala Lys Phe Pro Glu Pro
                115                 120                 125

aat ttc tgg agc aac tgg agt tcc tgg gag gaa gtc agt gta caa gat   793
Asn Phe Trp Ser Asn Trp Ser Ser Trp Glu Glu Val Ser Val Gln Asp
            130                 135                 140

tct act gga cag gat ata ttg ttc gtt ttc cct aaa gat aag ctg gtg   841
Ser Thr Gly Gln Asp Ile Leu Phe Val Phe Pro Lys Asp Lys Leu Val
        145                 150                 155

gaa gaa ggc acc aat gtt acc att tgt tac gtt tct agg aac att caa   889
Glu Glu Gly Thr Asn Val Thr Ile Cys Tyr Val Ser Arg Asn Ile Gln
    160                 165                 170

aat aat gta tcc tgt tat ttg gaa ggg aaa cag att cat gga gaa caa   937
Asn Asn Val Ser Cys Tyr Leu Glu Gly Lys Gln Ile His Gly Glu Gln
175                 180                 185                 190

ctt gat cca cat gta act gca ttc aac ttg aat agt gtg cct ttc att   985
Leu Asp Pro His Val Thr Ala Phe Asn Leu Asn Ser Val Pro Phe Ile
                195                 200                 205

agg aat aaa ggg aca aat atc tat tgt gag gca agt caa gga aat gtc   1033
Arg Asn Lys Gly Thr Asn Ile Tyr Cys Glu Ala Ser Gln Gly Asn Val
```

```
                210                 215                 220

agt gaa ggc atg aaa ggc atc gtt ctt ttt gtc tca aaa gta ctt gag   1081
Ser Glu Gly Met Lys Gly Ile Val Leu Phe Val Ser Lys Val Leu Glu
        225                 230                 235

gag ccc aag gac ttt tct tgt gaa acc gag gac ttc aag act ttg cac   1129
Glu Pro Lys Asp Phe Ser Cys Glu Thr Glu Asp Phe Lys Thr Leu His
    240                 245                 250

tgt act tgg gat cct ggg acg gac act gcc ttg ggg tgg tct aaa caa   1177
Cys Thr Trp Asp Pro Gly Thr Asp Thr Ala Leu Gly Trp Ser Lys Gln
255                 260                 265                 270

cct tcc caa agc tac act tta ttt gaa tca ttt tct ggg gaa aag aaa   1225
Pro Ser Gln Ser Tyr Thr Leu Phe Glu Ser Phe Ser Gly Glu Lys Lys
                275                 280                 285

ctt tgt aca cac aaa aac tgg tgt aat tgg caa ata act caa gac tca   1273
Leu Cys Thr His Lys Asn Trp Cys Asn Trp Gln Ile Thr Gln Asp Ser
            290                 295                 300

caa gaa acc tat aac ttc aca ctc ata gct gaa aat tac tta agg aag   1321
Gln Glu Thr Tyr Asn Phe Thr Leu Ile Ala Glu Asn Tyr Leu Arg Lys
        305                 310                 315

aga agt gtc aat atc ctt ttt aac ctg act cat cga gtt tat tta atg   1369
Arg Ser Val Asn Ile Leu Phe Asn Leu Thr His Arg Val Tyr Leu Met
    320                 325                 330

aat cct ttt agt gtc aac ttt gaa aat gta aat gcc aca aat gcc atc   1417
Asn Pro Phe Ser Val Asn Phe Glu Asn Val Asn Ala Thr Asn Ala Ile
335                 340                 345                 350

atg acc tgg aag gtg cac tcc ata agg aat aat ttc aca tat ttg tgt   1465
Met Thr Trp Lys Val His Ser Ile Arg Asn Asn Phe Thr Tyr Leu Cys
                355                 360                 365

cag att gaa ctc cat ggt gaa gga aaa atg atg caa tac aat gtt tcc   1513
Gln Ile Glu Leu His Gly Glu Gly Lys Met Met Gln Tyr Asn Val Ser
            370                 375                 380

atc aag gtg aac ggt gag tac ttc tta agt gaa ctg gaa cct gcc aca   1561
Ile Lys Val Asn Gly Glu Tyr Phe Leu Ser Glu Leu Glu Pro Ala Thr
        385                 390                 395

gag tac atg gcg cga gta cgg tgt gct gat gcc agc cac ttc tgg aaa   1609
Glu Tyr Met Ala Arg Val Arg Cys Ala Asp Ala Ser His Phe Trp Lys
    400                 405                 410

tgg agt gaa tgg agt ggt cag aac ttc acc aca ctt gaa gct gct ccc   1657
Trp Ser Glu Trp Ser Gly Gln Asn Phe Thr Thr Leu Glu Ala Ala Pro
415                 420                 425                 430

tca gag gcc cct gat gtc tgg aga att gtg agc ttg gag cca gga aat   1705
Ser Glu Ala Pro Asp Val Trp Arg Ile Val Ser Leu Glu Pro Gly Asn
                435                 440                 445

cat act gtg acc tta ttc tgg aag cca tta tca aaa ctg cat gcc aat   1753
His Thr Val Thr Leu Phe Trp Lys Pro Leu Ser Lys Leu His Ala Asn
            450                 455                 460

gga aag atc ctg ttc tat aat gta gtt gta gaa aac cta gac aaa cca   1801
```

```
Gly Lys Ile Leu Phe Tyr Asn Val Val Val Glu Asn Leu Asp Lys Pro
        465                 470                 475

tcc agt tca gag ctc cat tcc att cca gca cca gcc aac agc aca aaa   1849
Ser Ser Ser Glu Leu His Ser Ile Pro Ala Pro Ala Asn Ser Thr Lys
    480                 485                 490

cta atc ctt gac agg tgt tcc tac caa atc tgc gtc ata gcc aac aac   1897
Leu Ile Leu Asp Arg Cys Ser Tyr Gln Ile Cys Val Ile Ala Asn Asn
495                 500                 505                 510

agt gtg ggt gct tct cct gct tct gta ata gtc atc tct gca gac ccc   1945
Ser Val Gly Ala Ser Pro Ala Ser Val Ile Val Ile Ser Ala Asp Pro
                515                 520                 525

gaa aac aaa gag gtt gag gaa gaa aga att gca ggc aca gag ggt gga   1993
Glu Asn Lys Glu Val Glu Glu Glu Arg Ile Ala Gly Thr Glu Gly Gly
            530                 535                 540

ttc tct ctg tct tgg aaa ccc caa cct gga gat gtt ata ggc tat gtt   2041
Phe Ser Leu Ser Trp Lys Pro Gln Pro Gly Asp Val Ile Gly Tyr Val
        545                 550                 555

gtg gac tgg tgt gac cat acc cag gat gtg ctc ggt gat ttc cag tgg   2089
Val Asp Trp Cys Asp His Thr Gln Asp Val Leu Gly Asp Phe Gln Trp
    560                 565                 570

aag aat gta ggt ccc aat acc aca agc aca gtc att agc aca gat gct   2137
Lys Asn Val Gly Pro Asn Thr Thr Ser Thr Val Ile Ser Thr Asp Ala
575                 580                 585                 590

ttt agg cca gga gtt cga tat gac ttc aga att tat ggg tta tct aca   2185
Phe Arg Pro Gly Val Arg Tyr Asp Phe Arg Ile Tyr Gly Leu Ser Thr
                595                 600                 605

aaa agg att gct tgt tta tta gag aaa aaa aca gga tac tct cag gaa   2233
Lys Arg Ile Ala Cys Leu Leu Glu Lys Lys Thr Gly Tyr Ser Gln Glu
            610                 615                 620

ctt gct cct tca gac aac cct cac gtg ctg gtg gat aca ttg aca tcc   2281
Leu Ala Pro Ser Asp Asn Pro His Val Leu Val Asp Thr Leu Thr Ser
        625                 630                 635

cac tcc ttc act ctg agt tgg aaa gat tac tct act gaa tct caa cct   2329
His Ser Phe Thr Leu Ser Trp Lys Asp Tyr Ser Thr Glu Ser Gln Pro
    640                 645                 650

ggt ttt ata caa ggg tac cat gtc tat ctg aaa tcc aag gcg agg cag   2377
Gly Phe Ile Gln Gly Tyr His Val Tyr Leu Lys Ser Lys Ala Arg Gln
655                 660                 665                 670

tgc cac cca cga ttt gaa aag gca gtt ctt tca gat ggt tca gaa tgt   2425
Cys His Pro Arg Phe Glu Lys Ala Val Leu Ser Asp Gly Ser Glu Cys
                675                 680                 685

tgc aaa tac aaa att gac aac ccg gaa gaa aag gca ttg att gtg gac   2473
Cys Lys Tyr Lys Ile Asp Asn Pro Glu Glu Lys Ala Leu Ile Val Asp
            690                 695                 700

aac cta aag cca gaa tcc ttc tat gag ttt ttc atc act cca ttc act   2521
Asn Leu Lys Pro Glu Ser Phe Tyr Glu Phe Phe Ile Thr Pro Phe Thr
        705                 710                 715
```

```
agt gct ggt gaa ggc ccc agt gct acg ttc acg aag gtc acg act ccg   2569
Ser Ala Gly Glu Gly Pro Ser Ala Thr Phe Thr Lys Val Thr Thr Pro
    720                 725                 730

gat gaa cac tcc tcg atg ctg att cat atc cta ctg ccc atg gtt ttc   2617
Asp Glu His Ser Ser Met Leu Ile His Ile Leu Leu Pro Met Val Phe
735                 740                 745                 750

tgc gtc ttg ctc atc atg gtc atg tgc tac ttg aaa agt cag tgg atc   2665
Cys Val Leu Leu Ile Met Val Met Cys Tyr Leu Lys Ser Gln Trp Ile
                755                 760                 765

aag gag acc tgt tat cct gac atc cct gac cct tac aag agc agc atc   2713
Lys Glu Thr Cys Tyr Pro Asp Ile Pro Asp Pro Tyr Lys Ser Ser Ile
            770                 775                 780

ctg tca tta ata aaa ttc aag gag aac cct cac cta ata ata atg aat   2761
Leu Ser Leu Ile Lys Phe Lys Glu Asn Pro His Leu Ile Ile Met Asn
        785                 790                 795

gtc agt gac tgt atc cca gat gct att gaa gtt gta agc aag cca gaa   2809
Val Ser Asp Cys Ile Pro Asp Ala Ile Glu Val Val Ser Lys Pro Glu
    800                 805                 810

ggg aca aag ata cag ttc cta ggc act agg aag tca ctc aca gaa acc   2857
Gly Thr Lys Ile Gln Phe Leu Gly Thr Arg Lys Ser Leu Thr Glu Thr
815                 820                 825                 830

gag ttg act aag cct aac tac ctt tat ctc ctt cca aca gaa aag aat   2905
Glu Leu Thr Lys Pro Asn Tyr Leu Tyr Leu Leu Pro Thr Glu Lys Asn
                835                 840                 845

cac tct ggc cct ggc ccc tgc atc tgt ttt gag aac ttg acc tat aac   2953
His Ser Gly Pro Gly Pro Cys Ile Cys Phe Glu Asn Leu Thr Tyr Asn
            850                 855                 860

cag gca gct tct gac tct ggc tct tgt ggc cat gtt cca gta tcc cca   3001
Gln Ala Ala Ser Asp Ser Gly Ser Cys Gly His Val Pro Val Ser Pro
        865                 870                 875

aaa gcc cca agt atg ctg gga cta atg acc tca cct gaa aat gta cta   3049
Lys Ala Pro Ser Met Leu Gly Leu Met Thr Ser Pro Glu Asn Val Leu
    880                 885                 890

aag gca cta gaa aaa aac tac atg aac tcc ctg gga gaa atc cca gct   3097
Lys Ala Leu Glu Lys Asn Tyr Met Asn Ser Leu Gly Glu Ile Pro Ala
895                 900                 905                 910

gga gaa aca agt ttg aat tat gtg tcc cag ttg gct tca ccc atg ttt   3145
Gly Glu Thr Ser Leu Asn Tyr Val Ser Gln Leu Ala Ser Pro Met Phe
                915                 920                 925

gga gac aag gac agt ctc cca aca aac cca gta gag gca cca cac tgt   3193
Gly Asp Lys Asp Ser Leu Pro Thr Asn Pro Val Glu Ala Pro His Cys
            930                 935                 940

tca gag tat aaa atg caa atg gca gtc tcc ctg cgt ctt gcc ttg cct   3241
Ser Glu Tyr Lys Met Gln Met Ala Val Ser Leu Arg Leu Ala Leu Pro
        945                 950                 955

ccc ccg acc gag aat agc agc ctc tcc tca att acc ctt tta gat cca   3289
Pro Pro Thr Glu Asn Ser Ser Leu Ser Ser Ile Thr Leu Leu Asp Pro
    960                 965                 970
```

ggt gaa cac tac tgc taaccagcat gccgatttca taccttatgc tacacagaca 3344
Gly Glu His Tyr Cys
975

ttaagaagag cagagctggc accctgtcat caccagtggc cttggtcctt aatcccagta 3404
caatttgcag gtctggttta tataagacca ctacagtctg gctaggttaa aggccagagg 3464
ctatggaact taacactccc cattggagca agcttgccct agagacggca ggatcatggg 3524
agcatgctta ccttctgctg tttgttccag gctcaccttt agaacaggag acttgagctt 3584
gacctaagga tatgcattaa ccactctaca gactcccact cagtactgta cagggtggct 3644
gtggtcctag aagttcagtt tttactgagg aaatatttcc attaacagca attattatat 3704
tgaaggcttt aataaaggcc acaggagaca ttactatagc atagattgtc aaatgtaaat 3764
ttactgagcg tgttttataa aaaactcaca ggtgtttgag gccaaaacag attttagact 3824
taccttgaac ggataagaat ctatagttca ctgacacagt aaaattaact ctgtgggtgg 3884
gggcgggggg catagctcta atctaatata taaaatgtgt gatgaatcaa caagatttcc 3944
acaattcttc tgtcaagctt actacagtga aagaatggga ttggcaagta acttctgact 4004
tactgtcagt tgtacttctg ctccatagac atcagtattc tgccatcatt tttgatgact 4064
acctcagaac ataaaaagga acgtatatca cataattcca gtcacagttt ttggttcctc 4124
ttttctttca agaactatat ataaatgacc tgttttcacg cggccgc 4171

<210> 7
<211> 979
<212> PRT
<213> Homo sapiens

<400> 7
Met Ala Leu Phe Ala Val Phe Gln Thr Thr Phe Phe Leu Thr Leu Leu
1 5 10 15
Ser Leu Arg Thr Tyr Gln Ser Glu Val Leu Ala Glu Arg Leu Pro Leu
20 25 30
Thr Pro Val Ser Leu Lys Val Ser Thr Asn Ser Thr Arg Gln Ser Leu
35 40 45
His Leu Gln Trp Thr Val His Asn Leu Pro Tyr His Gln Glu Leu Lys
50 55 60
Met Val Phe Gln Ile Gln Ile Ser Arg Ile Glu Thr Ser Asn Val Ile
65 70 75 80
Trp Val Gly Asn Tyr Ser Thr Thr Val Lys Trp Asn Gln Val Leu His
85 90 95
Trp Ser Trp Glu Ser Glu Leu Pro Leu Glu Cys Ala Thr His Phe Val
100 105 110
Arg Ile Lys Ser Leu Val Asp Asp Ala Lys Phe Pro Glu Pro Asn Phe
115 120 125
Trp Ser Asn Trp Ser Ser Trp Glu Glu Val Ser Val Gln Asp Ser Thr
130 135 140
Gly Gln Asp Ile Leu Phe Val Phe Pro Lys Asp Lys Leu Val Glu Glu
145 150 155 160
Gly Thr Asn Val Thr Ile Cys Tyr Val Ser Arg Asn Ile Gln Asn Asn
165 170 175
Val Ser Cys Tyr Leu Glu Gly Lys Gln Ile His Gly Glu Gln Leu Asp
180 185 190
Pro His Val Thr Ala Phe Asn Leu Asn Ser Val Pro Phe Ile Arg Asn
195 200 205
Lys Gly Thr Asn Ile Tyr Cys Glu Ala Ser Gln Gly Asn Val Ser Glu
210 215 220
Gly Met Lys Gly Ile Val Leu Phe Val Ser Lys Val Leu Glu Glu Pro
225 230 235 240
Lys Asp Phe Ser Cys Glu Thr Glu Asp Phe Lys Thr Leu His Cys Thr
245 250 255
Trp Asp Pro Gly Thr Asp Thr Ala Leu Gly Trp Ser Lys Gln Pro Ser
260 265 270
Gln Ser Tyr Thr Leu Phe Glu Ser Phe Ser Gly Glu Lys Lys Leu Cys
275 280 285
Thr His Lys Asn Trp Cys Asn Trp Gln Ile Thr Gln Asp Ser Gln Glu

```
    290                 295                 300
Thr Tyr Asn Phe Thr Leu Ile Ala Glu Asn Tyr Leu Arg Lys Arg Ser
305                 310                 315                 320
Val Asn Ile Leu Phe Asn Leu Thr His Arg Val Tyr Leu Met Asn Pro
                325                 330                 335
Phe Ser Val Asn Phe Glu Asn Val Asn Ala Thr Asn Ala Ile Met Thr
            340                 345                 350
Trp Lys Val His Ser Ile Arg Asn Asn Phe Thr Tyr Leu Cys Gln Ile
        355                 360                 365
Glu Leu His Gly Glu Gly Lys Met Met Gln Tyr Asn Val Ser Ile Lys
    370                 375                 380
Val Asn Gly Glu Tyr Phe Leu Ser Glu Leu Glu Pro Ala Thr Glu Tyr
385                 390                 395                 400
Met Ala Arg Val Arg Cys Ala Asp Ala Ser His Phe Trp Lys Trp Ser
                405                 410                 415
Glu Trp Ser Gly Gln Asn Phe Thr Thr Leu Glu Ala Ala Pro Ser Glu
            420                 425                 430
Ala Pro Asp Val Trp Arg Ile Val Ser Leu Glu Pro Gly Asn His Thr
        435                 440                 445
Val Thr Leu Phe Trp Lys Pro Leu Ser Lys Leu His Ala Asn Gly Lys
    450                 455                 460
Ile Leu Phe Tyr Asn Val Val Val Glu Asn Leu Asp Lys Pro Ser Ser
465                 470                 475                 480
Ser Glu Leu His Ser Ile Pro Ala Pro Ala Asn Ser Thr Lys Leu Ile
                485                 490                 495
Leu Asp Arg Cys Ser Tyr Gln Ile Cys Val Ile Ala Asn Asn Ser Val
            500                 505                 510
Gly Ala Ser Pro Ala Ser Val Ile Val Ile Ser Ala Asp Pro Glu Asn
        515                 520                 525
Lys Glu Val Glu Glu Glu Arg Ile Ala Gly Thr Glu Gly Gly Phe Ser
    530                 535                 540
Leu Ser Trp Lys Pro Gln Pro Gly Asp Val Ile Gly Tyr Val Val Asp
545                 550                 555                 560
Trp Cys Asp His Thr Gln Asp Val Leu Gly Asp Phe Gln Trp Lys Asn
                565                 570                 575
Val Gly Pro Asn Thr Thr Ser Thr Val Ile Ser Thr Asp Ala Phe Arg
            580                 585                 590
Pro Gly Val Arg Tyr Asp Phe Arg Ile Tyr Gly Leu Ser Thr Lys Arg
        595                 600                 605
Ile Ala Cys Leu Leu Glu Lys Lys Thr Gly Tyr Ser Gln Glu Leu Ala
    610                 615                 620
Pro Ser Asp Asn Pro His Val Leu Val Asp Thr Leu Thr Ser His Ser
625                 630                 635                 640
Phe Thr Leu Ser Trp Lys Asp Tyr Ser Thr Glu Ser Gln Pro Gly Phe
                645                 650                 655
Ile Gln Gly Tyr His Val Tyr Leu Lys Ser Lys Ala Arg Gln Cys His
            660                 665                 670
Pro Arg Phe Glu Lys Ala Val Leu Ser Asp Gly Ser Glu Cys Cys Lys
        675                 680                 685
Tyr Lys Ile Asp Asn Pro Glu Glu Lys Ala Leu Ile Val Asp Asn Leu
    690                 695                 700
Lys Pro Glu Ser Phe Tyr Glu Phe Phe Ile Thr Pro Phe Thr Ser Ala
705                 710                 715                 720
Gly Glu Gly Pro Ser Ala Thr Phe Thr Lys Val Thr Thr Pro Asp Glu
                725                 730                 735
His Ser Ser Met Leu Ile His Ile Leu Leu Pro Met Val Phe Cys Val
            740                 745                 750
Leu Leu Ile Met Val Met Cys Tyr Leu Lys Ser Gln Trp Ile Lys Glu
        755                 760                 765
Thr Cys Tyr Pro Asp Ile Pro Asp Pro Tyr Lys Ser Ser Ile Leu Ser
    770                 775                 780
Leu Ile Lys Phe Lys Glu Asn Pro His Leu Ile Ile Met Asn Val Ser
785                 790                 795                 800
```

```
Asp Cys Ile Pro Asp Ala Ile Glu Val Val Ser Lys Pro Glu Gly Thr
                805                 810                 815
Lys Ile Gln Phe Leu Gly Thr Arg Lys Ser Leu Thr Glu Thr Glu Leu
            820                 825                 830
Thr Lys Pro Asn Tyr Leu Tyr Leu Leu Pro Thr Glu Lys Asn His Ser
        835                 840                 845
Gly Pro Gly Pro Cys Ile Cys Phe Glu Asn Leu Thr Tyr Asn Gln Ala
    850                 855                 860
Ala Ser Asp Ser Gly Ser Cys Gly His Val Pro Val Ser Pro Lys Ala
865                 870                 875                 880
Pro Ser Met Leu Gly Leu Met Thr Ser Pro Glu Asn Val Leu Lys Ala
                885                 890                 895
Leu Glu Lys Asn Tyr Met Asn Ser Leu Gly Glu Ile Pro Ala Gly Glu
            900                 905                 910
Thr Ser Leu Asn Tyr Val Ser Gln Leu Ala Ser Pro Met Phe Gly Asp
        915                 920                 925
Lys Asp Ser Leu Pro Thr Asn Pro Val Glu Ala Pro His Cys Ser Glu
    930                 935                 940
Tyr Lys Met Gln Met Ala Val Ser Leu Arg Leu Ala Leu Pro Pro Pro
945                 950                 955                 960
Thr Glu Asn Ser Ser Leu Ser Ser Ile Thr Leu Leu Asp Pro Gly Glu
                965                 970                 975
His Tyr Cys
```

<210> 8
<211> 2657
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (133)...(2040)

<400> 8

```
cggaggcggc ctgccggggt ggttcggctt cccgttgccg cctcgggcgc tgtacccaga 60
gctcgaagag gagcagcgcg gccgcgcgga cccggcaagg ctgggccgga ctcggggctc 120
ccgagggacg cc atg cgg gga ggc agg ggc gcc cct ttc tgg ctg tgg ccg 171
              Met Arg Gly Gly Arg Gly Ala Pro Phe Trp Leu Trp Pro
               1               5                  10

ctg ccc aag ctg gcg ctg ctg cct ctg ttg tgg gtg ctt ttc cag cgg   219
Leu Pro Lys Leu Ala Leu Leu Pro Leu Leu Trp Val Leu Phe Gln Arg
     15                  20                  25

acg cgt ccc cag ggc agc gcc ggg cca ctg cag tgc tac gga gtt gga   267
Thr Arg Pro Gln Gly Ser Ala Gly Pro Leu Gln Cys Tyr Gly Val Gly
 30                  35                  40                  45

ccc ttg ggc gac ttg aac tgc tcg tgg gag cct ctt ggg gac ctg gga   315
Pro Leu Gly Asp Leu Asn Cys Ser Trp Glu Pro Leu Gly Asp Leu Gly
                 50                  55                  60

gcc ccc tcc gag tta cac ctc cag agc caa aag tac cgt tcc aac aaa   363
Ala Pro Ser Glu Leu His Leu Gln Ser Gln Lys Tyr Arg Ser Asn Lys
             65                  70                  75

acc cag act gtg gca gtg gca gcc gga cgg agc tgg gtg gcc att cct   411
Thr Gln Thr Val Ala Val Ala Ala Gly Arg Ser Trp Val Ala Ile Pro
         80                  85                  90

cgg gaa cag ctc acc atg tct gac aaa ctc ctt gtc tgg ggc act aag   459
```

```
Arg Glu Gln Leu Thr Met Ser Asp Lys Leu Leu Val Trp Gly Thr Lys
     95                  100                 105

gca ggc cag cct ctc tgg ccc ccc gtc ttc gtg aac cta gaa acc caa   507
Ala Gly Gln Pro Leu Trp Pro Pro Val Phe Val Asn Leu Glu Thr Gln
110                 115                 120                 125

atg aag cca aac gcc ccc cgg ctg ggc cct gac gtg gac ttt tcc gag   555
Met Lys Pro Asn Ala Pro Arg Leu Gly Pro Asp Val Asp Phe Ser Glu
                130                 135                 140

gat gac ccc ctg gag gcc act gtc cat tgg gcc cca cct aca tgg cca   603
Asp Asp Pro Leu Glu Ala Thr Val His Trp Ala Pro Pro Thr Trp Pro
            145                 150                 155

tct cat aaa gtt ctg atc tgc cag ttc cac tac cga aga tgt cag gag   651
Ser His Lys Val Leu Ile Cys Gln Phe His Tyr Arg Arg Cys Gln Glu
        160                 165                 170

gcg gcc tgg acc ctg ctg gaa ccg gag ctg aag acc ata ccc ctg acc   699
Ala Ala Trp Thr Leu Leu Glu Pro Glu Leu Lys Thr Ile Pro Leu Thr
    175                 180                 185

cct gtt gag atc caa gat ttg gag cta gcc act ggc tac aaa gtg tat   747
Pro Val Glu Ile Gln Asp Leu Glu Leu Ala Thr Gly Tyr Lys Val Tyr
190                 195                 200                 205

ggc cgc tgc cgg atg gag aaa gaa gag gat ttg tgg ggc gag tgg agc   795
Gly Arg Cys Arg Met Glu Lys Glu Glu Asp Leu Trp Gly Glu Trp Ser
                210                 215                 220

ccc att ttg tcc ttc cag aca ccg cct tct gct cca aaa gat gtg tgg   843
Pro Ile Leu Ser Phe Gln Thr Pro Pro Ser Ala Pro Lys Asp Val Trp
            225                 230                 235

gta tca ggg aac ctc tgt ggg acg cct gga gga gag gaa cct ttg ctt   891
Val Ser Gly Asn Leu Cys Gly Thr Pro Gly Gly Glu Glu Pro Leu Leu
        240                 245                 250

cta tgg aag gcc cca ggg ccc tgt gtg cag gtg agc tac aaa gtc tgg   939
Leu Trp Lys Ala Pro Gly Pro Cys Val Gln Val Ser Tyr Lys Val Trp
    255                 260                 265

ttc tgg gtt gga ggt cgt gag ctg agt cca gaa gga att acc tgc tgc   987
Phe Trp Val Gly Gly Arg Glu Leu Ser Pro Glu Gly Ile Thr Cys Cys
270                 275                 280                 285

tgc tcc cta att ccc agt ggg gcg gag tgg gcc agg gtg tcc gct gtc  1035
Cys Ser Leu Ile Pro Ser Gly Ala Glu Trp Ala Arg Val Ser Ala Val
                290                 295                 300

aac gcc aca agc tgg gag cct ctc acc aac ctc tct ttg gtc tgc ttg  1083
Asn Ala Thr Ser Trp Glu Pro Leu Thr Asn Leu Ser Leu Val Cys Leu
            305                 310                 315

gat tca gcc tct gcc ccc cgt agc gtg gca gtc agc agc atc gct ggg  1131
Asp Ser Ala Ser Ala Pro Arg Ser Val Ala Val Ser Ser Ile Ala Gly
        320                 325                 330

agc acg gag cta ctg gtg acc tgg caa ccg ggg cct ggg gaa cca ctg  1179
Ser Thr Glu Leu Leu Val Thr Trp Gln Pro Gly Pro Gly Glu Pro Leu
    335                 340                 345
```

```
gag cat gta gtg gac tgg gct cga gat ggg gac ccc ctg gag aaa ctc   1227
Glu His Val Val Asp Trp Ala Arg Asp Gly Asp Pro Leu Glu Lys Leu
350                 355                 360                 365

aac tgg gtc cgg ctt ccc cct ggg aac ctc agt gct ctg tta cca ggg   1275
Asn Trp Val Arg Leu Pro Pro Gly Asn Leu Ser Ala Leu Leu Pro Gly
                370                 375                 380

aat ttc act gtc ggg gtc ccc tat cga atc act gtg acc gca gtc tct   1323
Asn Phe Thr Val Gly Val Pro Tyr Arg Ile Thr Val Thr Ala Val Ser
            385                 390                 395

gct tca ggc ttg gcc tct gca tcc tcc gtc tgg ggg ttc agg gag gaa   1371
Ala Ser Gly Leu Ala Ser Ala Ser Ser Val Trp Gly Phe Arg Glu Glu
        400                 405                 410

tta gca ccc cta gtg ggg cca acg ctt tgg cga ctc caa gat gcc cct   1419
Leu Ala Pro Leu Val Gly Pro Thr Leu Trp Arg Leu Gln Asp Ala Pro
    415                 420                 425

cca ggg acc ccc gcc ata gcg tgg gga gag gtc cca agg cac cag ctt   1467
Pro Gly Thr Pro Ala Ile Ala Trp Gly Glu Val Pro Arg His Gln Leu
430                 435                 440                 445

cga ggc cac ctc acc cac tac acc ttg tgt gca cag agt gga acc agc   1515
Arg Gly His Leu Thr His Tyr Thr Leu Cys Ala Gln Ser Gly Thr Ser
                450                 455                 460

ccc tcc gtc tgc atg aat gtg agt ggc aac aca cag agt gtc acc ctg   1563
Pro Ser Val Cys Met Asn Val Ser Gly Asn Thr Gln Ser Val Thr Leu
            465                 470                 475

cct gac ctt cct tgg ggt ccc tgt gag ctg tgg gtg aca gca tct acc   1611
Pro Asp Leu Pro Trp Gly Pro Cys Glu Leu Trp Val Thr Ala Ser Thr
        480                 485                 490

atc gct gga cag ggc cct cct ggt ccc atc ctc cgg ctt cat cta cca   1659
Ile Ala Gly Gln Gly Pro Pro Gly Pro Ile Leu Arg Leu His Leu Pro
    495                 500                 505

gat aac acc ctg agg tgg aaa gtt ctg cca ggc atc cta ttc ttg tgg   1707
Asp Asn Thr Leu Arg Trp Lys Val Leu Pro Gly Ile Leu Phe Leu Trp
510                 515                 520                 525

ggc ttg ttc ctg ttg ggg tgt ggc ctg agc ctg gcc acc tct gga agg   1755
Gly Leu Phe Leu Leu Gly Cys Gly Leu Ser Leu Ala Thr Ser Gly Arg
                530                 535                 540

tgc tac cac cta agg cac aaa gtg ctg ccc cgc tgg gtc tgg gag aaa   1803
Cys Tyr His Leu Arg His Lys Val Leu Pro Arg Trp Val Trp Glu Lys
            545                 550                 555

gtt cct gat cct gcc aac agc agt tca ggc cag ccc cac atg gag caa   1851
Val Pro Asp Pro Ala Asn Ser Ser Ser Gly Gln Pro His Met Glu Gln
        560                 565                 570

gta cct gag gcc cag ccc ctt ggg gac ttg ccc atc ctg gaa gtg gag   1899
Val Pro Glu Ala Gln Pro Leu Gly Asp Leu Pro Ile Leu Glu Val Glu
    575                 580                 585

gag atg gag ccc ccg ccg gtt atg gag tcc tcc cag ccc gcc cag gcc   1947
Glu Met Glu Pro Pro Pro Val Met Glu Ser Ser Gln Pro Ala Gln Ala
590                 595                 600                 605
```

```
acc gcc ccg ctt gac tct ggg tat gag aag cac ttc ctg ccc aca cct   1995
Thr Ala Pro Leu Asp Ser Gly Tyr Glu Lys His Phe Leu Pro Thr Pro
                610                 615                 620

gag gag ctg ggc ctt ctg ggg ccc ccc agg cca cag gtt ctg gcc       2040
Glu Glu Leu Gly Leu Leu Gly Pro Pro Arg Pro Gln Val Leu Ala
            625                 630                 635

tgaaccacac gtctggctgg gggctgccag ccaggctaga gggatgctca tgcaggttgc 2100
accccagtcc tggattagcc ctcttgatgg atgaagacac tgaggactca gagaggctga 2160
gtcacttacc tgaggacacc cagccaggca gagctgggat tgaaggaccc ctatagagaa 2220
gggcttggcc cccatgggga agacacggat ggaaggtgga gcaaaggaaa atacatgaaa 2280
ttgagagtgg cagctgcctg ccaaaatctg ttccgctgta acagaactga atttggaccc 2340
cagcacagtg gctcacgcct gtaatcccag cactttggca ggccaaggtg gaaggatcac 2400
ttagagctag gagtttgaga ccagcctggg caatatagca agacccctca ctacaaaaat 2460
aaaacatcaa aaacaaaaac aattagctgg gcatgatggc acacacctgt agtccgagcc 2520
acttgggagg ctgaggtggg aggatcggtt gagcccagga gttcgaagct gcagggacct 2580
ctgattgcac cactgcactc caggctgggt aacagaatga gaccttatct caaaaataaa 2640
caaactaata aaaagca                                              2657

<210> 9
<211> 636
<212> PRT
<213> Homo sapiens

<400> 9
Met Arg Gly Gly Arg Gly Ala Pro Phe Trp Leu Trp Pro Leu Pro Lys
 1               5                  10                  15
Leu Ala Leu Leu Pro Leu Leu Trp Val Leu Phe Gln Arg Thr Arg Pro
            20                  25                  30
Gln Gly Ser Ala Gly Pro Leu Gln Cys Tyr Gly Val Gly Pro Leu Gly
        35                  40                  45
Asp Leu Asn Cys Ser Trp Glu Pro Leu Gly Asp Leu Gly Ala Pro Ser
    50                  55                  60
Glu Leu His Leu Gln Ser Gln Lys Tyr Arg Ser Asn Lys Thr Gln Thr
65                  70                  75                  80
Val Ala Val Ala Ala Gly Arg Ser Trp Val Ala Ile Pro Arg Glu Gln
                85                  90                  95
Leu Thr Met Ser Asp Lys Leu Leu Val Trp Gly Thr Lys Ala Gly Gln
            100                 105                 110
Pro Leu Trp Pro Pro Val Phe Val Asn Leu Glu Thr Gln Met Lys Pro
        115                 120                 125
Asn Ala Pro Arg Leu Gly Pro Asp Val Asp Phe Ser Glu Asp Asp Pro
    130                 135                 140
Leu Glu Ala Thr Val His Trp Ala Pro Pro Thr Trp Pro Ser His Lys
145                 150                 155                 160
Val Leu Ile Cys Gln Phe His Tyr Arg Arg Cys Gln Glu Ala Ala Trp
                165                 170                 175
Thr Leu Leu Glu Pro Glu Leu Lys Thr Ile Pro Leu Thr Pro Val Glu
            180                 185                 190
Ile Gln Asp Leu Glu Leu Ala Thr Gly Tyr Lys Val Tyr Gly Arg Cys
        195                 200                 205
Arg Met Glu Lys Glu Glu Asp Leu Trp Gly Glu Trp Ser Pro Ile Leu
    210                 215                 220
Ser Phe Gln Thr Pro Pro Ser Ala Pro Lys Asp Val Trp Val Ser Gly
225                 230                 235                 240
Asn Leu Cys Gly Thr Pro Gly Gly Glu Glu Pro Leu Leu Leu Trp Lys
                245                 250                 255
Ala Pro Gly Pro Cys Val Gln Val Ser Tyr Lys Val Trp Phe Trp Val
            260                 265                 270
Gly Gly Arg Glu Leu Ser Pro Glu Gly Ile Thr Cys Cys Cys Ser Leu
        275                 280                 285
```

```
Ile Pro Ser Gly Ala Glu Trp Ala Arg Val Ser Ala Val Asn Ala Thr
    290                 295                 300
Ser Trp Glu Pro Leu Thr Asn Leu Ser Leu Val Cys Leu Asp Ser Ala
305                 310                 315                 320
Ser Ala Pro Arg Ser Val Ala Val Ser Ser Ile Ala Gly Ser Thr Glu
                325                 330                 335
Leu Leu Val Thr Trp Gln Pro Gly Pro Gly Glu Pro Leu Glu His Val
            340                 345                 350
Val Asp Trp Ala Arg Asp Gly Asp Pro Leu Glu Lys Leu Asn Trp Val
        355                 360                 365
Arg Leu Pro Pro Gly Asn Leu Ser Ala Leu Leu Pro Gly Asn Phe Thr
    370                 375                 380
Val Gly Val Pro Tyr Arg Ile Thr Val Thr Ala Val Ser Ala Ser Gly
385                 390                 395                 400
Leu Ala Ser Ala Ser Ser Val Trp Gly Phe Arg Glu Glu Leu Ala Pro
                405                 410                 415
Leu Val Gly Pro Thr Leu Trp Arg Leu Gln Asp Ala Pro Pro Gly Thr
            420                 425                 430
Pro Ala Ile Ala Trp Gly Glu Val Pro Arg His Gln Leu Arg Gly His
        435                 440                 445
Leu Thr His Tyr Thr Leu Cys Ala Gln Ser Gly Thr Ser Pro Ser Val
    450                 455                 460
Cys Met Asn Val Ser Gly Asn Thr Gln Ser Val Thr Leu Pro Asp Leu
465                 470                 475                 480
Pro Trp Gly Pro Cys Glu Leu Trp Val Thr Ala Ser Thr Ile Ala Gly
                485                 490                 495
Gln Gly Pro Pro Gly Pro Ile Leu Arg Leu His Leu Pro Asp Asn Thr
            500                 505                 510
Leu Arg Trp Lys Val Leu Pro Gly Ile Leu Phe Leu Trp Gly Leu Phe
        515                 520                 525
Leu Leu Gly Cys Gly Leu Ser Leu Ala Thr Ser Gly Arg Cys Tyr His
    530                 535                 540
Leu Arg His Lys Val Leu Pro Arg Trp Val Trp Glu Lys Val Pro Asp
545                 550                 555                 560
Pro Ala Asn Ser Ser Ser Gly Gln Pro His Met Glu Gln Val Pro Glu
                565                 570                 575
Ala Gln Pro Leu Gly Asp Leu Pro Ile Leu Glu Val Glu Glu Met Glu
            580                 585                 590
Pro Pro Pro Val Met Glu Ser Ser Gln Pro Ala Gln Ala Thr Ala Pro
        595                 600                 605
Leu Asp Ser Gly Tyr Glu Lys His Phe Leu Pro Thr Pro Glu Glu Leu
    610                 615                 620
Gly Leu Leu Gly Pro Pro Arg Pro Gln Val Leu Ala
625                 630                 635
```

<210> 10
<211> 755
<212> DNA
<213> Mus musculus

<220>
<221> CDS
<222> (1)...(489)

<400> 10

```
atg atc ttc cac aca gga aca acg aag cct acc ctg gtg ctg ctt tgc   48
Met Ile Phe His Thr Gly Thr Thr Lys Pro Thr Leu Val Leu Leu Cys
 1               5                  10                  15

tgt ata gga acc tgg ctg gcc acc tgc agc ttg tcc ttc ggt gcc cca   96
Cys Ile Gly Thr Trp Leu Ala Thr Cys Ser Leu Ser Phe Gly Ala Pro
            20                  25                  30
```

```
ata tcg aag gaa gac tta aga act aca att gac ctc ttg aaa caa gag   144
Ile Ser Lys Glu Asp Leu Arg Thr Thr Ile Asp Leu Leu Lys Gln Glu
         35                  40                  45

tct cag gat ctt tat aac aac tat agc ata aag cag gca tct ggg atg   192
Ser Gln Asp Leu Tyr Asn Asn Tyr Ser Ile Lys Gln Ala Ser Gly Met
     50                  55                  60

tca gca gac gaa tca ata cag ctg ccg tgt ttc agc ctg gac cgg gaa   240
Ser Ala Asp Glu Ser Ile Gln Leu Pro Cys Phe Ser Leu Asp Arg Glu
 65                  70                  75                  80

gca tta acc aac atc tcg gtc atc ata gca cat ctg gag aaa gtc aaa   288
Ala Leu Thr Asn Ile Ser Val Ile Ile Ala His Leu Glu Lys Val Lys
                 85                  90                  95

gtg ttg agc gag aac aca gta gat act tct tgg gtg ata aga tgg cta   336
Val Leu Ser Glu Asn Thr Val Asp Thr Ser Trp Val Ile Arg Trp Leu
            100                 105                 110

aca aac atc agc tgt ttc aac cca ctg aat tta aac att tct gtg cct   384
Thr Asn Ile Ser Cys Phe Asn Pro Leu Asn Leu Asn Ile Ser Val Pro
        115                 120                 125

gga aat act gat gaa tcc tat gat tgt aaa gtg ttc gtg ctt acg gtt   432
Gly Asn Thr Asp Glu Ser Tyr Asp Cys Lys Val Phe Val Leu Thr Val
    130                 135                 140

tta aag cag ttc tca aac tgc atg gca gaa ctg cag gct aag gac aat   480
Leu Lys Gln Phe Ser Asn Cys Met Ala Glu Leu Gln Ala Lys Asp Asn
145                 150                 155                 160

act aca tgc tgagtgatgg gggggggggg ggtgcagtgt cctcagcagt           529
Thr Thr Cys

gcctgtcctt cgagggctga gcttgcaacc caggacttaa ctccaaaggg actgtgcggt 589
cattactagt catgttattt atgtttttat tttgtccact gaaatcttgt tctgctaccc 649
tgtagggact ggaagtggca gctatattta tttatttatg tactgagttt gttaacgctc 709
catggaggag ccttcagagt ctatttaata aattatattg acatga                755

<210> 11
<211> 163
<212> PRT
<213> Mus musculus

<400> 11
Met Ile Phe His Thr Gly Thr Thr Lys Pro Thr Leu Val Leu Leu Cys
 1               5                  10                  15
Cys Ile Gly Thr Trp Leu Ala Thr Cys Ser Leu Ser Phe Gly Ala Pro
            20                  25                  30
Ile Ser Lys Glu Asp Leu Arg Thr Thr Ile Asp Leu Leu Lys Gln Glu
        35                  40                  45
Ser Gln Asp Leu Tyr Asn Asn Tyr Ser Ile Lys Gln Ala Ser Gly Met
    50                  55                  60
Ser Ala Asp Glu Ser Ile Gln Leu Pro Cys Phe Ser Leu Asp Arg Glu
65                  70                  75                  80
Ala Leu Thr Asn Ile Ser Val Ile Ile Ala His Leu Glu Lys Val Lys
                85                  90                  95
Val Leu Ser Glu Asn Thr Val Asp Thr Ser Trp Val Ile Arg Trp Leu
            100                 105                 110
Thr Asn Ile Ser Cys Phe Asn Pro Leu Asn Leu Asn Ile Ser Val Pro
```

```
        115                 120                 125
Gly Asn Thr Asp Glu Ser Tyr Asp Cys Lys Val Phe Val Leu Thr Val
    130                 135                 140
Leu Lys Gln Phe Ser Asn Cys Met Ala Glu Leu Gln Ala Lys Asp Asn
145                 150                 155                 160
Thr Thr Cys
```

<210> 12
<211> 489
<212> DNA
<213> Artificial Sequence

<220>
<223> mouse zcytor17lig degenerate polynucleotide of SEQ ID NO:11

<221> misc_feature
<222> (1)...(489)
<223> n = A,T,C or G

<400> 12
atgathttyc ayacnggnac nacnaarccn acnytngtny tnytntgytg yathggnacn 60
tggytngcna cntgywsnyt nwsnttyggn gcnccnathw snaargarga yytnmgnacn 120
acnathgayy tnytnaarca rgarwsncar gayytntaya ayaaytayws nathaarcar 180
gcnwsnggna tgwsngcnga ygarwsnath carytnccnt gyttywsnyt ngaymgngar 240
gcnytnacna ayathwsngt nathathgcn cayytngara argtnaargt nytnwsngar 300
aayacngtng ayacnwsntg ggtnathmgn tggytnacna ayathwsntg yttyaayccn 360
ytnaayytna ayathwsngt nccnggnaay acngaygarw sntaygaytg yaargtntty 420
gtnytnacng tnytnaarca rttywsnaay tgyatggcng arytncargc naargayaay 480
acnacntgy 489

<210> 13
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC6673

<400> 13
gcgcaaggtg ccgttcacag c 21

<210> 14
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29082

<400> 14
caatttgttg ggtttttta gcagcagtag gcccag 36

<210> 15
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29083

<400> 15
ctgggcctac tgctgctaaa aaaacccaac aaattg 36

<210> 16
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29145

<400> 16
gcgtctagag ggttatattg aagttgggca ggaaga 36

<210> 17
<211> 8
<212> PRT
<213> Artificial Sequence

<220>
<223> Gly-Ser Modified Glu-Glu (CEE) peptide tag

<400> 17
Gly Ser Glu Tyr Met Pro Met Glu
1 5

<210> 18
<211> 33
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29359

<400> 18
gcgggatcca tgaagctctc tccccagcct tca 33

<210> 19
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC27899

<400> 19
ccagaacttt gactccttga ccg 23

<210> 20
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC27895

<400> 20
gaagtcaact tcgctaagaa ccg 23

<210> 21
<211> 34
<212> DNA

<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29122

<400> 21
ccgctcgagt tatattgaag ttgggcagga agac 34

<210> 22
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29180

<400> 22
cctggagtcc ctgaaacgaa ag 22

<210> 23
<211> 34
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29122

<400> 23
ccgctcgagt tatattgaag ttgggcagga agac 34

<210> 24
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC9791

<400> 24
cgttccaaca aaacccagac 20

<210> 25
<211> 19
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC9793

<400> 25
tggcgttgac agcggacac 19

<210> 26
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC40109

<400> 26
ccattccagc accagccaac 20

<210> 27
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC40112

<400> 27
tacaacttca atagcatctg gg 22

<210> 28
<211> 40
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC13946

<400> 28
ccctgcagtg atcaacatgg ccaagttgac cagtgccgtt 40

<210> 29
<211> 45
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC13945

<400> 29
gcccatggac tagtttcgaa aggtcgagtg tcagtcctgc tcctc 45

<210> 30
<211> 34
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC18698

<400> 30
ttttttttctc gagacttttt ttttttttttt tttt 34

<210> 31

<220>
<223> Skipped Sequence

<400> 31
000

<210> 32
<211> 10
<212> PRT
<213> Artificial Sequence

<220>
<223> Glu-Glu (CEE) peptide tag with Gly-Ser residue pair

<400> 32
Gly Ser Gly Gly Glu Tyr Met Pro Met Glu

1 5 10

<210> 33
<211> 33
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29451

<400> 33
ccggaattcc cctgatacat gaagctctct ccc 33

<210> 34
<211> 33
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29124

<400> 34
cgcggatccc tcaaagacac tgaatgacaa tgt 33

<210> 35
<211> 6
<212> PRT
<213> Artificial Sequence

<220>
<223> Glu-Glu (CEE) peptide tag without Gly-Ser residue pair

<400> 35
Glu Tyr Met Pro Met Glu
1 5

<210> 36
<211> 8
<212> PRT
<213> Artificial Sequence

<220>
<223> C-terminal FLAG peptide sequence

<400> 36
Asp Tyr Lys Asp Asp Asp Asp Lys
1 5

<210> 37
<211> 684
<212> DNA
<213> Homo sapiens

<400> 37
tcagacaaaa ctcacacatg cccaccgtgc ccagcacctg aagccgaggg ggcaccgtca 60
gtcttcctct tccccccaaa acccaaggac accctcatga tctcccggac ccctgaggtc 120
acatgcgtgg tggtggacgt gagccacgaa gaccctgagg tcaagttcaa ctggtacgtg 180
gacggcgtgg aggtgcataa tgccaagaca aagccgcggg aggagcagta caacagcacg 240
taccgtgtgg tcagcgtcct caccgtcctg caccaggact ggctgaatgg caaggagtac 300

aagtgcaagg tctccaacaa agccctccca tcctccatcg agaaaaccat ctccaaagcc 360
aaagggcagc cccgagaacc acaggtgtac accctgcccc catcccggga tgagctgacc 420
aagaaccagg tcagcctgac ctgcctggtc aaaggcttct atcccagcga catcgccgtg 480
gagtgggaga gcaatgggca gccggagaac aactacaaga ccacgcctcc cgtgctggac 540
tccgacggct ccttcttcct ctacagcaag ctcaccgtgg acaagagcag gtggcagcag 600
gggaacgtct tctcatgctc cgtgatgcat gaggctctgc acaaccacta cacgcagaag 660
agcctctccc tgtctccggg taaa 684

<210> 38
<211> 2295
<212> DNA
<213> Artificial Sequence

<220>
<223> human zcytor17-Fc4 fusion polynucleotide

<221> CDS
<222> (1)...(2295)

<400> 38
atg aag ctc tct ccc cag cct tca tgt gtt aac ctg ggg atg atg tgg 48
Met Lys Leu Ser Pro Gln Pro Ser Cys Val Asn Leu Gly Met Met Trp
1 5 10 15

acc tgg gca ctg tgg atg ctc cct tca ctc tgc aaa ttc agc ctg gca 96
Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe Ser Leu Ala
20 25 30

gct ctg cca gct aag cct gag aac att tcc tgt gtc tac tac tat agg 144
Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr Tyr Tyr Arg
35 40 45

aaa aat tta acc tgc act tgg agt cca gga aag gaa acc agt tat acc 192
Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr Ser Tyr Thr
50 55 60

cag tac aca gtt aag aga act tac gct ttt gga gaa aaa cat gat aat 240
Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys His Asp Asn
65 70 75 80

tgt aca acc aat agt tct aca agt gaa aat cgt gct tcg tgc tct ttt 288
Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser Cys Ser Phe
85 90 95

ttc ctt cca aga ata acg atc cca gat aat tat acc att gag gtg gaa 336
Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile Glu Val Glu
100 105 110

gct gaa aat gga gat ggt gta att aaa tct cat atg aca tac tgg aga 384
Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr Tyr Trp Arg
115 120 125

tta gag aac ata gcg aaa act gaa cca cct aag att ttc cgt gtg aaa 432
Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe Arg Val Lys
130 135 140

cca gtt ttg ggc atc aaa cga atg att caa att gaa tgg ata aag cct 480
Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp Ile Lys Pro
145 150 155 160

gag ttg gcg cct gtt tca tct gat tta aaa tac aca ctt cga ttc agg 528
Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu Arg Phe Arg
165 170 175

```
aca gtc aac agt acc agc tgg atg gaa gtc aac ttc gct aag aac cgt   576
Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala Lys Asn Arg
            180                 185                 190

aag gat aaa aac caa acg tac aac ctc acg ggg ctg cag cct ttt aca   624
Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln Pro Phe Thr
        195                 200                 205

gaa tat gtc ata gct ctg cga tgt gcg gtc aag gag tca aag ttc tgg   672
Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser Lys Phe Trp
    210                 215                 220

agt gac tgg agc caa gaa aaa atg gga atg act gag gaa gaa gct cca   720
Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu Glu Ala Pro
225                 230                 235                 240

tgt ggc ctg gaa ctg tgg aga gtc ctg aaa cca gct gag gcg gat gga   768
Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu Ala Asp Gly
                245                 250                 255

aga agg cca gtg cgg ttg tta tgg aag aag gca aga gga gcc cca gtc   816
Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val
            260                 265                 270

cta gag aaa aca ctt ggc tac aac ata tgg tac tat cca gaa agc aac   864
Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro Glu Ser Asn
        275                 280                 285

act aac ctc aca gaa aca atg aac act act aac cag cag ctt gaa ctg   912
Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln Leu Glu Leu
    290                 295                 300

cat ctg gga ggc gag agc ttt tgg gtg tct atg att tct tat aat tct   960
His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser Tyr Asn Ser
305                 310                 315                 320

ctt ggg aag tct cca gtg gcc acc ctg agg att cca gct att caa gaa   1008
Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala Ile Gln Glu
                325                 330                 335

aaa tca ttt cag tgc att gag gtc atg cag gcc tgc gtt gct gag gac   1056
Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val Ala Glu Asp
            340                 345                 350

cag cta gtg gtg aag tgg caa agc tct gct cta gac gtg aac act tgg   1104
Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val Asn Thr Trp
        355                 360                 365

atg att gaa tgg ttt ccg gat gtg gac tca gag ccc acc acc ctt tcc   1152
Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr Thr Leu Ser
    370                 375                 380

tgg gaa tct gtg tct cag gcc acg aac tgg acg atc cag caa gat aaa   1200
Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln Gln Asp Lys
385                 390                 395                 400

tta aaa ccc ttc tgg tgc tat aac atc tct gtg tat cca atg ttg cat   1248
Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro Met Leu His
                405                 410                 415

gac aaa gtt ggc gag cca tat tcc atc cag gct tat gcc aaa gaa ggc   1296
Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu Gly
```

```
                420                 425                 430

gtt cca tca gaa ggt cct gag acc aag gtg gag aac att ggc gtg aag   1344
Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile Gly Val Lys
        435                 440                 445

acg gtc acg atc aca tgg aaa gag att ccc aag agt gag aga aag ggt   1392
Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu Arg Lys Gly
    450                 455                 460

atc atc tgc aac tac acc atc ttt tac caa gct gaa ggt gga aaa gga   1440
Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly Gly Lys Gly
465                 470                 475                 480

ttc tcc aag aca gtc aat tcc agc atc ttg cag tac ggc ctg gag tcc   1488
Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly Leu Glu Ser
                485                 490                 495

ctg aaa cga aag acc tct tac att gtt cag gtc atg gcc agc acc agt   1536
Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala Ser Thr Ser
            500                 505                 510

gct ggg gga acc aac ggg acc agc ata aat ttc aag aca ttg tca ttc   1584
Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr Leu Ser Phe
        515                 520                 525

agt gtc ttt gag gag ccc aga tct tca gac aaa act cac aca tgc cca   1632
Ser Val Phe Glu Glu Pro Arg Ser Ser Asp Lys Thr His Thr Cys Pro
    530                 535                 540

ccg tgc cca gca cct gaa gcc gag ggg gca ccg tca gtc ttc ctc ttc   1680
Pro Cys Pro Ala Pro Glu Ala Glu Gly Ala Pro Ser Val Phe Leu Phe
545                 550                 555                 560

ccc cca aaa ccc aag gac acc ctc atg atc tcc cgg acc cct gag gtc   1728
Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val
                565                 570                 575

aca tgc gtg gtg gtg gac gtg agc cac gaa gac cct gag gtc aag ttc   1776
Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe
            580                 585                 590

aac tgg tac gtg gac ggc gtg gag gtg cat aat gcc aag aca aag ccg   1824
Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro
        595                 600                 605

cgg gag gag cag tac aac agc acg tac cgt gtg gtc agc gtc ctc acc   1872
Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr
    610                 615                 620

gtc ctg cac cag gac tgg ctg aat ggc aag gag tac aag tgc aag gtc   1920
Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val
625                 630                 635                 640

tcc aac aaa gcc ctc cca tcc tcc atc gag aaa acc atc tcc aaa gcc   1968
Ser Asn Lys Ala Leu Pro Ser Ser Ile Glu Lys Thr Ile Ser Lys Ala
                645                 650                 655

aaa ggg cag ccc cga gaa cca cag gtg tac acc ctg ccc cca tcc cgg   2016
Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg
            660                 665                 670

gat gag ctg acc aag aac cag gtc agc ctg acc tgc ctg gtc aaa ggc   2064
```

```
Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly
        675                 680                 685

ttc tat ccc agc gac atc gcc gtg gag tgg gag agc aat ggg cag ccg   2112
Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro
    690                 695                 700

gag aac aac tac aag acc acg cct ccc gtg ctg gac tcc gac ggc tcc   2160
Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser
705                 710                 715                 720

ttc ttc ctc tac agc aag ctc acc gtg gac aag agc agg tgg cag cag   2208
Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln
                725                 730                 735

ggg aac gtc ttc tca tgc tcc gtg atg cat gag gct ctg cac aac cac   2256
Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His
            740                 745                 750

tac acg cag aag agc ctc tcc ctg tct ccg ggt aaa taa               2295
Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys  *
        755                 760
```

<210> 39
<211> 764
<212> PRT
<213> Artificial Sequence

<220>
<223> human zcytor17-Fc4 fusion polypeptide

<400> 39

```
Met Lys Leu Ser Pro Gln Pro Ser Cys Val Asn Leu Gly Met Met Trp
 1               5                  10                  15
Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe Ser Leu Ala
            20                  25                  30
Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr Tyr Tyr Arg
        35                  40                  45
Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr Ser Tyr Thr
    50                  55                  60
Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys His Asp Asn
65                  70                  75                  80
Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser Cys Ser Phe
                85                  90                  95
Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile Glu Val Glu
            100                 105                 110
Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr Tyr Trp Arg
        115                 120                 125
Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe Arg Val Lys
    130                 135                 140
Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp Ile Lys Pro
145                 150                 155                 160
Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu Arg Phe Arg
                165                 170                 175
Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala Lys Asn Arg
            180                 185                 190
Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln Pro Phe Thr
        195                 200                 205
Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser Lys Phe Trp
    210                 215                 220
Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu Glu Ala Pro
225                 230                 235                 240
```

Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu Ala Asp Gly
245 250 255
Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val
260 265 270
Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro Glu Ser Asn
275 280 285
Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln Leu Glu Leu
290 295 300
His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser Tyr Asn Ser
305 310 315 320
Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala Ile Gln Glu
325 330 335
Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val Ala Glu Asp
340 345 350
Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val Asn Thr Trp
355 360 365
Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr Thr Leu Ser
370 375 380
Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln Gln Asp Lys
385 390 395 400
Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro Met Leu His
405 410 415
Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu Gly
420 425 430
Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile Gly Val Lys
435 440 445
Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu Arg Lys Gly
450 455 460
Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly Gly Lys Gly
465 470 475 480
Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly Leu Glu Ser
485 490 495
Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala Ser Thr Ser
500 505 510
Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr Leu Ser Phe
515 520 525
Ser Val Phe Glu Glu Pro Arg Ser Ser Asp Lys Thr His Thr Cys Pro
530 535 540
Pro Cys Pro Ala Pro Glu Ala Glu Gly Ala Pro Ser Val Phe Leu Phe
545 550 555 560
Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val
565 570 575
Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe
580 585 590
Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro
595 600 605
Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr
610 615 620
Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val
625 630 635 640
Ser Asn Lys Ala Leu Pro Ser Ser Ile Glu Lys Thr Ile Ser Lys Ala
645 650 655
Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg
660 665 670
Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly
675 680 685
Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro
690 695 700
Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser
705 710 715 720
Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln
725 730 735
Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His

740 745 750
Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys
755 760

<210> 40
<211> 34
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29157

<400> 40
ctagtatggc cggccatgaa gctctctccc cagc 34

<210> 41
<211> 41
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29150

<400> 41
gtctgaagat ctgggctcct caaagacact gaatgacaat g 41

<210> 42
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41458

<400> 42
tggacctcgc actaaaatca t 21

<210> 43
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41457

<400> 43
aatcacggca gagttcccac ac 22

<210> 44
<211> 100
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC12749

<400> 44
gtaccttccc gtaaatccct ccccttcccg gaattacacc cgcgtatttc ccagaaaagg 60
aactgtagat ttctaggaat tcaatccttg gccacgcgtc 100

<210> 45
<211> 100

<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC12748

<400> 45
tcgagacgcg tggccaagga ttgaattcct agaaatctac agttcctttt ctgggaaata 60
cgcgggtgta attccgggaa ggggagggat ttacgggaag 100

<210> 46
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC10651

<400> 46
agcttttctg cagcagctct 20

<210> 47
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC10565

<400> 47
tttgcagaaa aggttgcaaa tgc 23

<210> 48
<211> 25
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC14063

<400> 48
caccagacat aatagctgac agact 25

<210> 49
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC17574

<400> 49
ggtrttgctc agcatgcaca c 21

<210> 50
<211> 24
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC17600

<400> 50

catgtaggcc atgaggtcca ccac 24

<210> 51
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC38065

<400> 51
ctttcctggg aatctgtgtc t 21

<210> 52
<211> 18
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC38068

<400> 52
cctccagctc tggtgctg 18

<210> 53
<211> 24
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC37877

<400> 53
caaaaaaccc aacaaattga ctca 24

<210> 54
<211> 25
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC37876

<400> 54
catgtggcta tactactttc agcag 25

<210> 55
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> zcytor17 receptor TaqMan® probe

<400> 55
ctgtgttggc ccaccgttcc ca 22

<210> 56
<211> 20
<212> DNA
<213> Artificial Sequence

<220>

<223> rRNA forward primer

<400> 56
cggctaccac atccaaggaa 20

<210> 57
<211> 18
<212> DNA
<213> Artificial Sequence

<220>
<223> the rRNA reverse primer

<400> 57
gctggaatta ccgcggct 18

<210> 58
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> rRNA TaqMan® probe

<400> 58
tgctggcacc agacttgccc tc 22

<210> 59

<220>
<223> Skipped Sequence

<400> 59
000

<210> 60
<211> 21
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41458

<400> 60
tggacctcgc actaaaatca t 21

<210> 61
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41457

<400> 61
aatcacggca gagttcccac ac 22

<210> 62
<211> 19
<212> DNA
<213> Artificial Sequence

<220>

<223> Oligonucleotide primer ZC41459

<400> 62
agaagggcgt gctcgtgtc 19

<210> 63
<211> 18
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41460

<400> 63
ccggatggct gggctgtg 18

<210> 64
<211> 25
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC39982

<400> 64
aatgtctgtg tagcataagg tatga 25

<210> 65
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC39983

<400> 65
cctgcctacc tgaaaaccag aa 22

<210> 66
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC39980

<400> 66
tttgaattcg ccaccatggc tctatttgca gtcttt 36

<210> 67
<211> 28
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC39981

<400> 67
ctgtctcgag tgctggttag cagtgttc 28

<210> 68
<211> 2217
<212> DNA

<213> Homo sapiens

<220>
<221> CDS
<222> (1)...(2217)

<400> 68

```
atg gct cta ttt gca gtc ttt cag aca aca ttc ttc tta aca ttg ctg   48
Met Ala Leu Phe Ala Val Phe Gln Thr Thr Phe Phe Leu Thr Leu Leu
 1               5                  10                  15

tcc ttg agg act tac cag agt gaa gtc ttg gct gaa cgt tta cca ttg   96
Ser Leu Arg Thr Tyr Gln Ser Glu Val Leu Ala Glu Arg Leu Pro Leu
            20                  25                  30

act cct gta tca ctt aaa gtt tcc acc aat tct acg cgt cag agt ttg   144
Thr Pro Val Ser Leu Lys Val Ser Thr Asn Ser Thr Arg Gln Ser Leu
        35                  40                  45

cac tta caa tgg act gtc cac aac ctt cct tat cat cag gaa ttg aaa   192
His Leu Gln Trp Thr Val His Asn Leu Pro Tyr His Gln Glu Leu Lys
    50                  55                  60

atg gta ttt cag atc cag atc agt agg att gaa aca tcc aat gtc atc   240
Met Val Phe Gln Ile Gln Ile Ser Arg Ile Glu Thr Ser Asn Val Ile
65                  70                  75                  80

tgg gtg ggg aat tac agc acc act gtg aag tgg aac cag gtt ctg cat   288
Trp Val Gly Asn Tyr Ser Thr Thr Val Lys Trp Asn Gln Val Leu His
                85                  90                  95

tgg agc tgg gaa tct gag ctc cct ttg gaa tgt gcc aca cac ttt gta   336
Trp Ser Trp Glu Ser Glu Leu Pro Leu Glu Cys Ala Thr His Phe Val
            100                 105                 110

aga ata aag agt ttg gtg gac gat gcc aag ttc cct gag cca aat ttc   384
Arg Ile Lys Ser Leu Val Asp Asp Ala Lys Phe Pro Glu Pro Asn Phe
        115                 120                 125

tgg agc aac tgg agt tcc tgg gag gaa gtc agt gta caa gat tct act   432
Trp Ser Asn Trp Ser Ser Trp Glu Glu Val Ser Val Gln Asp Ser Thr
    130                 135                 140

gga cag gat ata ttg ttc gtt ttc cct aaa gat aag ctg gtg gaa gaa   480
Gly Gln Asp Ile Leu Phe Val Phe Pro Lys Asp Lys Leu Val Glu Glu
145                 150                 155                 160

ggc acc aat gtt acc att tgt tac gtt tct agg aac att caa aat aat   528
Gly Thr Asn Val Thr Ile Cys Tyr Val Ser Arg Asn Ile Gln Asn Asn
                165                 170                 175

gta tcc tgt tat ttg gaa ggg aaa cag att cat gga gaa caa ctt gat   576
Val Ser Cys Tyr Leu Glu Gly Lys Gln Ile His Gly Glu Gln Leu Asp
            180                 185                 190

cca cat gta act gca ttc aac ttg aat agt gtg cct ttc att agg aat   624
Pro His Val Thr Ala Phe Asn Leu Asn Ser Val Pro Phe Ile Arg Asn
        195                 200                 205

aaa ggg aca aat atc tat tgt gag gca agt caa gga aat gtc agt gaa   672
Lys Gly Thr Asn Ile Tyr Cys Glu Ala Ser Gln Gly Asn Val Ser Glu
    210                 215                 220
```

```
ggc atg aaa ggc atc gtt ctt ttt gtc tca aaa gta ctt gag gag ccc   720
Gly Met Lys Gly Ile Val Leu Phe Val Ser Lys Val Leu Glu Glu Pro
225                 230                 235                 240

aag gac ttt tct tgt gaa acc gag gac ttc aag act ttg cac tgt act   768
Lys Asp Phe Ser Cys Glu Thr Glu Asp Phe Lys Thr Leu His Cys Thr
                245                 250                 255

tgg gat cct ggg acg gac act gcc ttg ggg tgg tct aaa caa cct tcc   816
Trp Asp Pro Gly Thr Asp Thr Ala Leu Gly Trp Ser Lys Gln Pro Ser
            260                 265                 270

caa agc tac act tta ttt gaa tca ttt tct ggg gaa aag aaa ctt tgt   864
Gln Ser Tyr Thr Leu Phe Glu Ser Phe Ser Gly Glu Lys Lys Leu Cys
        275                 280                 285

aca cac aaa aac tgg tgt aat tgg caa ata act caa gac tca caa gaa   912
Thr His Lys Asn Trp Cys Asn Trp Gln Ile Thr Gln Asp Ser Gln Glu
    290                 295                 300

acc tat aac ttc aca ctc ata gct gaa aat tac tta agg aag aga agt   960
Thr Tyr Asn Phe Thr Leu Ile Ala Glu Asn Tyr Leu Arg Lys Arg Ser
305                 310                 315                 320

gtc aat atc ctt ttt aac ctg act cat cga gtt tat tta atg aat cct  1008
Val Asn Ile Leu Phe Asn Leu Thr His Arg Val Tyr Leu Met Asn Pro
                325                 330                 335

ttt agt gtc aac ttt gaa aat gta aat gcc aca aat gcc atc atg acc  1056
Phe Ser Val Asn Phe Glu Asn Val Asn Ala Thr Asn Ala Ile Met Thr
            340                 345                 350

tgg aag gtg cac tcc ata agg aat aat ttc aca tat ttg tgt cag att  1104
Trp Lys Val His Ser Ile Arg Asn Asn Phe Thr Tyr Leu Cys Gln Ile
        355                 360                 365

gaa ctc cat ggt gaa gga aaa atg atg caa tac aat gtt tcc atc aag  1152
Glu Leu His Gly Glu Gly Lys Met Met Gln Tyr Asn Val Ser Ile Lys
    370                 375                 380

gtg aac ggt gag tac ttc tta agt gaa ctg gaa cct gcc aca gag tac  1200
Val Asn Gly Glu Tyr Phe Leu Ser Glu Leu Glu Pro Ala Thr Glu Tyr
385                 390                 395                 400

atg gcg cga gta cgg tgt gct gat gcc agc cac ttc tgg aaa tgg agt  1248
Met Ala Arg Val Arg Cys Ala Asp Ala Ser His Phe Trp Lys Trp Ser
                405                 410                 415

gaa tgg agt ggt cag aac ttc acc aca ctt gaa gct gct ccc tca gag  1296
Glu Trp Ser Gly Gln Asn Phe Thr Thr Leu Glu Ala Ala Pro Ser Glu
            420                 425                 430

gcc cct gat gtc tgg aga att gtg agc ttg gag cca gga aat cat act  1344
Ala Pro Asp Val Trp Arg Ile Val Ser Leu Glu Pro Gly Asn His Thr
        435                 440                 445

gtg acc tta ttc tgg aag cca tta tca aaa ctg cat gcc aat gga aag  1392
Val Thr Leu Phe Trp Lys Pro Leu Ser Lys Leu His Ala Asn Gly Lys
    450                 455                 460

atc ctg ttc tat aat gta gtt gta gaa aac cta gac aaa cca tcc agt  1440
Ile Leu Phe Tyr Asn Val Val Val Glu Asn Leu Asp Lys Pro Ser Ser
465                 470                 475                 480
```

```
tca gag ctc cat tcc att cca gca cca gcc aac agc aca aaa cta atc   1488
Ser Glu Leu His Ser Ile Pro Ala Pro Ala Asn Ser Thr Lys Leu Ile
                485                 490                 495

ctt gac agg tgt tcc tac caa atc tgc gtc ata gcc aac aac agt gtg   1536
Leu Asp Arg Cys Ser Tyr Gln Ile Cys Val Ile Ala Asn Asn Ser Val
            500                 505                 510

ggt gct tct cct gct tct gta ata gtc atc tct gca gac ccc gaa aac   1584
Gly Ala Ser Pro Ala Ser Val Ile Val Ile Ser Ala Asp Pro Glu Asn
        515                 520                 525

aaa gag gtt gag gaa gaa aga att gca ggc aca gag ggt gga ttc tct   1632
Lys Glu Val Glu Glu Glu Arg Ile Ala Gly Thr Glu Gly Gly Phe Ser
    530                 535                 540

ctg tct tgg aaa ccc caa cct gga gat gtt ata ggc tat gtt gtg gac   1680
Leu Ser Trp Lys Pro Gln Pro Gly Asp Val Ile Gly Tyr Val Val Asp
545                 550                 555                 560

tgg tgt gac cat acc cag gat gtg ctc ggt gat ttc cag tgg aag aat   1728
Trp Cys Asp His Thr Gln Asp Val Leu Gly Asp Phe Gln Trp Lys Asn
                565                 570                 575

gta ggt ccc aat acc aca agc aca gtc att agc aca gat gct ttt agg   1776
Val Gly Pro Asn Thr Thr Ser Thr Val Ile Ser Thr Asp Ala Phe Arg
            580                 585                 590

cca gga gtt cga tat gac ttc aga att tat ggg tta tct aca aaa agg   1824
Pro Gly Val Arg Tyr Asp Phe Arg Ile Tyr Gly Leu Ser Thr Lys Arg
        595                 600                 605

att gct tgt tta tta gag aaa aaa aca gga tac tct cag gaa ctt gct   1872
Ile Ala Cys Leu Leu Glu Lys Lys Thr Gly Tyr Ser Gln Glu Leu Ala
    610                 615                 620

cct tca gac aac cct cac gtg ctg gtg gat aca ttg aca tcc cac tcc   1920
Pro Ser Asp Asn Pro His Val Leu Val Asp Thr Leu Thr Ser His Ser
625                 630                 635                 640

ttc act ctg agt tgg aaa gat tac tct act gaa tct caa cct ggt ttt   1968
Phe Thr Leu Ser Trp Lys Asp Tyr Ser Thr Glu Ser Gln Pro Gly Phe
                645                 650                 655

ata caa ggg tac cat gtc tat ctg aaa tcc aag gcg agg cag tgc cac   2016
Ile Gln Gly Tyr His Val Tyr Leu Lys Ser Lys Ala Arg Gln Cys His
            660                 665                 670

cca cga ttt gaa aag gca gtt ctt tca gat ggt tca gaa tgt tgc aaa   2064
Pro Arg Phe Glu Lys Ala Val Leu Ser Asp Gly Ser Glu Cys Cys Lys
        675                 680                 685

tac aaa att gac aac ccg gaa gaa aag gca ttg att gtg gac aac cta   2112
Tyr Lys Ile Asp Asn Pro Glu Glu Lys Ala Leu Ile Val Asp Asn Leu
    690                 695                 700

aag cca gaa tcc ttc tat gag ttt ttc atc act cca ttc act agt gct   2160
Lys Pro Glu Ser Phe Tyr Glu Phe Phe Ile Thr Pro Phe Thr Ser Ala
705                 710                 715                 720

ggt gaa ggc ccc agt gct acg ttc acg aag gtc acg act ccg gat gaa   2208
Gly Glu Gly Pro Ser Ala Thr Phe Thr Lys Val Thr Thr Pro Asp Glu
```

725 730 735

cac tcc tcg 2217
His Ser Ser

<210> 69
<211> 739
<212> PRT
<213> Homo sapiens

<400> 69
Met Ala Leu Phe Ala Val Phe Gln Thr Thr Phe Phe Leu Thr Leu Leu
1 5 10 15
Ser Leu Arg Thr Tyr Gln Ser Glu Val Leu Ala Glu Arg Leu Pro Leu
20 25 30
Thr Pro Val Ser Leu Lys Val Ser Thr Asn Ser Thr Arg Gln Ser Leu
35 40 45
His Leu Gln Trp Thr Val His Asn Leu Pro Tyr His Gln Glu Leu Lys
50 55 60
Met Val Phe Gln Ile Gln Ile Ser Arg Ile Glu Thr Ser Asn Val Ile
65 70 75 80
Trp Val Gly Asn Tyr Ser Thr Thr Val Lys Trp Asn Gln Val Leu His
85 90 95
Trp Ser Trp Glu Ser Glu Leu Pro Leu Glu Cys Ala Thr His Phe Val
100 105 110
Arg Ile Lys Ser Leu Val Asp Asp Ala Lys Phe Pro Glu Pro Asn Phe
115 120 125
Trp Ser Asn Trp Ser Ser Trp Glu Glu Val Ser Val Gln Asp Ser Thr
130 135 140
Gly Gln Asp Ile Leu Phe Val Phe Pro Lys Asp Lys Leu Val Glu Glu
145 150 155 160
Gly Thr Asn Val Thr Ile Cys Tyr Val Ser Arg Asn Ile Gln Asn Asn
165 170 175
Val Ser Cys Tyr Leu Glu Gly Lys Gln Ile His Gly Glu Gln Leu Asp
180 185 190
Pro His Val Thr Ala Phe Asn Leu Asn Ser Val Pro Phe Ile Arg Asn
195 200 205
Lys Gly Thr Asn Ile Tyr Cys Glu Ala Ser Gln Gly Asn Val Ser Glu
210 215 220
Gly Met Lys Gly Ile Val Leu Phe Val Ser Lys Val Leu Glu Glu Pro
225 230 235 240
Lys Asp Phe Ser Cys Glu Thr Glu Asp Phe Lys Thr Leu His Cys Thr
245 250 255
Trp Asp Pro Gly Thr Asp Thr Ala Leu Gly Trp Ser Lys Gln Pro Ser
260 265 270
Gln Ser Tyr Thr Leu Phe Glu Ser Phe Ser Gly Glu Lys Lys Leu Cys
275 280 285
Thr His Lys Asn Trp Cys Asn Trp Gln Ile Thr Gln Asp Ser Gln Glu
290 295 300
Thr Tyr Asn Phe Thr Leu Ile Ala Glu Asn Tyr Leu Arg Lys Arg Ser
305 310 315 320
Val Asn Ile Leu Phe Asn Leu Thr His Arg Val Tyr Leu Met Asn Pro
325 330 335
Phe Ser Val Asn Phe Glu Asn Val Asn Ala Thr Asn Ala Ile Met Thr
340 345 350
Trp Lys Val His Ser Ile Arg Asn Asn Phe Thr Tyr Leu Cys Gln Ile
355 360 365
Glu Leu His Gly Glu Gly Lys Met Met Gln Tyr Asn Val Ser Ile Lys
370 375 380
Val Asn Gly Glu Tyr Phe Leu Ser Glu Leu Glu Pro Ala Thr Glu Tyr
385 390 395 400

```
Met Ala Arg Val Arg Cys Ala Asp Ala Ser His Phe Trp Lys Trp Ser
                405                 410                 415
Glu Trp Ser Gly Gln Asn Phe Thr Thr Leu Glu Ala Ala Pro Ser Glu
            420                 425                 430
Ala Pro Asp Val Trp Arg Ile Val Ser Leu Glu Pro Gly Asn His Thr
        435                 440                 445
Val Thr Leu Phe Trp Lys Pro Leu Ser Lys Leu His Ala Asn Gly Lys
    450                 455                 460
Ile Leu Phe Tyr Asn Val Val Val Glu Asn Leu Asp Lys Pro Ser Ser
465                 470                 475                 480
Ser Glu Leu His Ser Ile Pro Ala Pro Ala Asn Ser Thr Lys Leu Ile
                485                 490                 495
Leu Asp Arg Cys Ser Tyr Gln Ile Cys Val Ile Ala Asn Asn Ser Val
            500                 505                 510
Gly Ala Ser Pro Ala Ser Val Ile Val Ile Ser Ala Asp Pro Glu Asn
        515                 520                 525
Lys Glu Val Glu Glu Glu Arg Ile Ala Gly Thr Glu Gly Gly Phe Ser
    530                 535                 540
Leu Ser Trp Lys Pro Gln Pro Gly Asp Val Ile Gly Tyr Val Val Asp
545                 550                 555                 560
Trp Cys Asp His Thr Gln Asp Val Leu Gly Asp Phe Gln Trp Lys Asn
                565                 570                 575
Val Gly Pro Asn Thr Thr Ser Thr Val Ile Ser Thr Asp Ala Phe Arg
            580                 585                 590
Pro Gly Val Arg Tyr Asp Phe Arg Ile Tyr Gly Leu Ser Thr Lys Arg
        595                 600                 605
Ile Ala Cys Leu Leu Glu Lys Lys Thr Gly Tyr Ser Gln Glu Leu Ala
    610                 615                 620
Pro Ser Asp Asn Pro His Val Leu Val Asp Thr Leu Thr Ser His Ser
625                 630                 635                 640
Phe Thr Leu Ser Trp Lys Asp Tyr Ser Thr Glu Ser Gln Pro Gly Phe
                645                 650                 655
Ile Gln Gly Tyr His Val Tyr Leu Lys Ser Lys Ala Arg Gln Cys His
            660                 665                 670
Pro Arg Phe Glu Lys Ala Val Leu Ser Asp Gly Ser Glu Cys Cys Lys
        675                 680                 685
Tyr Lys Ile Asp Asn Pro Glu Glu Lys Ala Leu Ile Val Asp Asn Leu
    690                 695                 700
Lys Pro Glu Ser Phe Tyr Glu Phe Phe Ile Thr Pro Phe Thr Ser Ala
705                 710                 715                 720
Gly Glu Gly Pro Ser Ala Thr Phe Thr Lys Val Thr Thr Pro Asp Glu
                725                 730                 735
His Ser Ser
```

<210> 70
<211> 1557
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (1)...(1557)

<400> 70

```
atg atg tgg acc tgg gca ctg tgg atg ctc ccc tca ctc tgc aaa ttc   48
Met Met Trp Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe
 1               5                  10                  15

agc ctg gca gct ctg cca gct aag cct gag aac att tcc tgt gtc tac   96
Ser Leu Ala Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr
            20                  25                  30
```

```
tac tat agg aaa aat tta acc tgc act tgg agt cca gga aag gaa acc   144
Tyr Tyr Arg Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr
          35                  40                  45

agt tat acc cag tac aca gtt aag aga act tac gct ttt gga gaa aaa   192
Ser Tyr Thr Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys
     50                  55                  60

cat gat aat tgt aca acc aat agt tct aca agt gaa aat cgt gct tcg   240
His Asp Asn Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser
 65                  70                  75                  80

tgc tct ttt ttc ctt cca aga ata acg atc cca gat aat tat acc att   288
Cys Ser Phe Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile
                 85                  90                  95

gag gtg gaa gct gaa aat gga gat ggt gta att aaa tct cat atg aca   336
Glu Val Glu Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr
            100                 105                 110

tac tgg aga tta gag aac ata gcg aaa act gaa cca cct aag att ttc   384
Tyr Trp Arg Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe
        115                 120                 125

cgt gtg aaa cca gtt ttg ggc atc aaa cga atg att caa att gaa tgg   432
Arg Val Lys Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp
    130                 135                 140

ata aag cct gag ttg gcg cct gtt tca tct gat tta aaa tac aca ctt   480
Ile Lys Pro Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu
145                 150                 155                 160

cga ttc agg aca gtc aac agt acc agc tgg atg gaa gtc aac ttc gct   528
Arg Phe Arg Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala
                165                 170                 175

aag aac cgt aag gat aaa aac caa acg tac aac ctc acg ggg ctg cag   576
Lys Asn Arg Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln
            180                 185                 190

cct ttt aca gaa tat gtc ata gct ctg cga tgt gcg gtc aag gag tca   624
Pro Phe Thr Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser
        195                 200                 205

aag ttc tgg agt gac tgg agc caa gaa aaa atg gga atg act gag gaa   672
Lys Phe Trp Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu
    210                 215                 220

gaa gct cca tgt ggc ctg gaa ctg tgg aga gtc ctg aaa cca gct gag   720
Glu Ala Pro Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu
225                 230                 235                 240

gcg gat gga aga agg cca gtg cgg ttg tta tgg aag aag gca aga gga   768
Ala Asp Gly Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly
                245                 250                 255

gcc cca gtc cta gag aaa aca ctt ggc tac aac ata tgg tac tat cca   816
Ala Pro Val Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro
            260                 265                 270

gaa agc aac act aac ctc aca gaa aca atg aac act act aac cag cag   864
Glu Ser Asn Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln
```

```
        275                 280                 285

ctt gaa ctg cat ctg gga ggc gag agc ttt tgg gtg tct atg att tct   912
Leu Glu Leu His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser
    290                 295                 300

tat aat tct ctt ggg aag tct cca gtg gcc acc ctg agg att cca gct   960
Tyr Asn Ser Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala
305                 310                 315                 320

att caa gaa aaa tca ttt cag tgc att gag gtc atg cag gcc tgc gtt  1008
Ile Gln Glu Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val
                325                 330                 335

gct gag gac cag cta gtg gtg aag tgg caa agc tct gct cta gac gtg  1056
Ala Glu Asp Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val
            340                 345                 350

aac act tgg atg att gaa tgg ttt ccg gat gtg gac tca gag ccc acc  1104
Asn Thr Trp Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr
        355                 360                 365

acc ctt tcc tgg gaa tct gtg tct cag gcc acg aac tgg acg atc cag  1152
Thr Leu Ser Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln
    370                 375                 380

caa gat aaa tta aaa cct ttc tgg tgc tat aac atc tct gtg tat cca  1200
Gln Asp Lys Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro
385                 390                 395                 400

atg ttg cat gac aaa gtt ggc gag cca tat tcc atc cag gct tat gcc  1248
Met Leu His Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala
                405                 410                 415

aaa gaa ggc gtt cca tca gaa ggt cct gag acc aag gtg gag aac att  1296
Lys Glu Gly Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile
            420                 425                 430

ggc gtg aag acg gtc acg atc aca tgg aaa gag att ccc aag agt gag  1344
Gly Val Lys Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu
        435                 440                 445

aga aag ggt atc atc tgc aac tac acc atc ttt tac caa gct gaa ggt  1392
Arg Lys Gly Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly
    450                 455                 460

gga aaa gga ttc tcc aag aca gtc aat tcc agc atc ttg cag tac ggc  1440
Gly Lys Gly Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly
465                 470                 475                 480

ctg gag tcc ctg aaa cga aag acc tct tac att gtt cag gtc atg gcc  1488
Leu Glu Ser Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala
                485                 490                 495

agc acc agt gct ggg gga acc aac ggg acc agc ata aat ttc aag aca  1536
Ser Thr Ser Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr
            500                 505                 510

ttg tca ttc agt gtc ttt gag                                     1557
Leu Ser Phe Ser Val Phe Glu
        515
```

<210> 71
<211> 519
<212> PRT
<213> Homo sapiens

<400> 71
Met Met Trp Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe
1 5 10 15
Ser Leu Ala Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr
20 25 30
Tyr Tyr Arg Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr
35 40 45
Ser Tyr Thr Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys
50 55 60
His Asp Asn Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser
65 70 75 80
Cys Ser Phe Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile
85 90 95
Glu Val Glu Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr
100 105 110
Tyr Trp Arg Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe
115 120 125
Arg Val Lys Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp
130 135 140
Ile Lys Pro Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu
145 150 155 160
Arg Phe Arg Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala
165 170 175
Lys Asn Arg Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln
180 185 190
Pro Phe Thr Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser
195 200 205
Lys Phe Trp Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu
210 215 220
Glu Ala Pro Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu
225 230 235 240
Ala Asp Gly Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly
245 250 255
Ala Pro Val Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro
260 265 270
Glu Ser Asn Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln
275 280 285
Leu Glu Leu His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser
290 295 300
Tyr Asn Ser Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala
305 310 315 320
Ile Gln Glu Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val
325 330 335
Ala Glu Asp Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val
340 345 350
Asn Thr Trp Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr
355 360 365
Thr Leu Ser Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln
370 375 380
Gln Asp Lys Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro
385 390 395 400
Met Leu His Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala
405 410 415
Lys Glu Gly Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile
420 425 430
Gly Val Lys Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu
435 440 445
Arg Lys Gly Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly

```
    450                 455                 460
Gly Lys Gly Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly
465                 470                 475                 480
Leu Glu Ser Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala
                485                 490                 495
Ser Thr Ser Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr
            500                 505                 510
Leu Ser Phe Ser Val Phe Glu
        515
```

<210> 72
<211> 10
<212> PRT
<213> Artificial Sequence

<220>
<223> C-Terminal His peptide tag

<400> 72
Gly Ser Gly Gly His His His His His His
1 5 10

<210> 73
<211> 12
<212> PRT
<213> Artificial Sequence

<220>
<223> Gly-Ser spacer of 12 amino acids

<400> 73
Gly Ser Gly Ser Gly Ser Gly Ser Glu Pro Arg Ser
1 5 10

<210> 74
<211> 31
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41557

<400> 74
ttatagatct cgaggagtgt tcatccggag t 31

<210> 75
<211> 65
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29232

<400> 75
cgactgactc gagtcagtga tggtgatggt gatggccacc tgatccttta cccggagaca 60
gggag 65

<210> 76
<211> 3196
<212> DNA

<213> Mus musculus

<400> 76
tgtatgtctg cacagtttgt gtatgcctgg tgcccacaga ggctcgagag tgtcagattc 60
cccccaaaac tggagttaca gttttgagcc gccccatgct tgatagcaat caaacctggg 120
tcctctgaaa gagcatccag tgcatgtaac cactgaacca tctctccaaa ccatgaacat 180
cactttaatt ttttttaata gttaaaggat attttgattc taaagatgta aaagaacgtc 240
tcacctattt tgaaatttgg taataaatgt ttcttcaaag cttaaaaaaa ttagttcagg 300
ttttttttttt ttttcagtca gtgatttgct aagctgccca aactggctta gaatttgtga 360
ccctcttgtc tcagcatact gagtgttaag attacaagtg cacccccctac ccagttccca 420
taattaactg atccaccccc acccccatcc caccccactc ccattgcctg ggcaagtaac 480
tcttgagccc cattctggtt ctagagtctg aagtcacaaa ggtgcaggtg agaacgcaag 540
gacaagggca ggccctggag cacagatgcc ttctccttat gccttccctg tgttcactag 600
agccatcccc ctgcctccgg aattcccaca gatggatcgc tctgtggctt cttaaaactt 660
ccctgcaggg cactgaccct cagcccctct aagtcacttc ttccccagtg attgtacttt 720
tcaatcgggc ttcaaacttt cctctcatta aatcagcaag cactttccaa gaaaagagag 780
atgctcaaga tgccttcctg tgtggtatgt gtatgcgttt gtgtgtgtgc acgcatgtgt 840
gtgcatgtga ctcaatcttc tgccttgcct tgagggtaac ctcagcattt ccttccagcc 900
ctgctttccc caggccgagc cgaggctggc aacctttga aaatgttttc tggagaaaag 960
ctgagcaatg gttttgccat gggcgggcct ttgatctgct tcctcatgac aacccttat 1020
atattgcctg gtggccatgg cgaacacacc aggctccaga gaccacaggc aaagcgggcc 1080
ttcctcactc tcttaccgtc gccatgatct tccacacagg taccgctggc tccacacgca 1140
gctcagcatg gcttcagctc catggctctt atcatgttag gggaaggagc cgggaatggc 1200
tgcctcaggt ggttgctgga cagaggctgt tgtaactgaa gctgggatgg gcaggggcat 1260
ctgacctatc agctccatgg tatccttctt tttctccagg aacaacgaag cctaccctgg 1320
tgctgctttg ctgtatagga acctggctgg ccacctgcag cttgtccttc ggtgccccaa 1380
tatcgaagga agacttaaga actacaattg acctcttgaa acaagagtct caggatcttt 1440
ataacaacta tgtaagtgcc cttgagattg ttttttctta accatttctt taaaatgtct 1500
tattttgcta tctaagcaca gctatccttt ctcgatataa agccagctat ggaagccaga 1560
gaggcatggg gaaacattgg aattcggttg gggtgaaatg tttccaaggg ggtaaatgca 1620
ctagcagaag aggcagaggc agactggtcc agggactgaa accttggcag cttacgaaac 1680
actacaggat gtatgctccc tgaattcttt atctcaaatc cacccggctc acagtcccta 1740
ctaaacgagc attcttgctg aaagggcatc cttagagaag ggccagcttg attcaggaat 1800
cccccaagag caatgagagc cagtttcagc agccaaagat gtcctagtgg aagcagggtg 1860
tgaggatctt cctttgggtc tccgttgact aactaggcaa ctgtctgtgt gttcttggag 1920
catcctggag ggccctctgc ctggccagag cctggcacag gtacagcaca ggacccagaa 1980
agtgtgaata cttcatttcc ttgggaccgc ttagataact tcagttgaag caagtaacag 2040
ggaaactgat ggagacacag ataacctccc tgcccctctc acttcagtca ctgagcctcc 2100
gagaacaggt tgcagatggc taggggcagc ctcagccaga taggcggagg cagactgggt 2160
agaagcatcc ttaggaacca cggccaacct gggtgggtat gccatgtctt ctagctcata 2220
agccaactag accttcgatt cctgtagaca cagagttagt gatggcccaa gcttcagaag 2280
gttgttgtac caattagata aggtctgagg caggctagac acagaggaag ccctggaaat 2340
gagctgttct gagctgtagg gttgttacaa atgtcttcct tacaatattt caaacctcct 2400
ctttctacag agcataaagc aggcatctgg gatgtcagca gacgaatcaa tacagctgcc 2460
gtgtttcagc ctggaccggg aagcattaac caacatctcg gtcatcatag cacatctgga 2520
gaaagtcaaa gtgttgagcg agaacacagt agatacttct tgggtgataa gatggctaac 2580
aaacatcagc tgtttcaacc cactgaattt aaacatttct gtgcctggaa atactgatga 2640
atcctatgat tgtaaagtgt tcgtgcttac ggttttaaag cagttctcaa actgcatggc 2700
agaactgcag gctaaggaca atactacatg ctgagtgatg gggggggggg ggtgcagtgt 2760
cctcagcagt gcctgtcctt cgagggctga gcttgcaacc caggacttaa ctccaaaggg 2820
actgtgcggt cattactagt catgttattt atgttttttat tttgtccact gaaatcttgt 2880
tctgctaccc tgtagggact ggaagtggca gctatattta tttatttatg tactgagttt 2940
gttaacgctc catggaggag ccttcagagt ctatttaata aattatattg acatgatcac 3000
ataatcagtt ttggaatttg tgatggggtt gaaatcaaag attaggaatg ttctggaaat 3060
agtttatgct accctctccc tccattagac agactcatga gcaaataatc ccagcagcat 3120
cacgtgcatg ataaacatct ttgttccagg tcataagtac aatcactgtc cttttggtat 3180
gtaggctgga aactaa 3196

<210> 77
<211> 24
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC28575

<400> 77
ccaggaaagg aaaccagtta tacc 24

<210> 78
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC21195

<400> 78
gaggagacca taacccccga cag 23

<210> 79
<211> 23
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC21196

<400> 79
catagctccc accacacgat ttt 23

<210> 80
<211> 25
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC26358

<400> 80
aaaaccaaac gtacaacctc acggg 25

<210> 81
<211> 25
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC26359

<400> 81
gagcagccat acaccagagc agaca 25

<210> 82
<211> 19
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC29179

<400> 82
gcagggttgg gaacggtgg 19

<210> 83
<211> 20

<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC28917

<400> 83
tgcaagatgc tggaattgac 20

<210> 84
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC28916

<400> 84
agtcaattcc agcatcttgc 20

<210> 85
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC28918

<400> 85
tcacagagtc atcagactcc 20

<210> 86
<211> 18
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41498

<400> 86
ggctccagag accacagg 18

<210> 87
<211> 22
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41496

<400> 87
atgactagta atgaccgcac ag 22

<210> 88
<211> 39
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41583

<400> 88
cgtacgggcc ggccaccatg atcttccaca caggaacaa 39

<210> 89
<211> 36
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41584

<400> 89
tgacgaggcg cgcctcagca tgtagtattg tcctta 36

<210> 90
<211> 650
<212> DNA
<213> Mus musculus

<220>
<221> CDS
<222> (53)...(542)

<400> 90
ggctccagag accacaggca aagcgggcct tcctcactct cttaccgtcg cc atg atc 58
Met Ile
1

ttc cac aca gga aca acg aag cct acc ctg gtg ctg ctt tgc tgt ata 106
Phe His Thr Gly Thr Thr Lys Pro Thr Leu Val Leu Leu Cys Cys Ile
5 10 15

gga acc tgg ctg gcc acc tgc agc ttg tcc ttc ggt gcc cca ata tcg 154
Gly Thr Trp Leu Ala Thr Cys Ser Leu Ser Phe Gly Ala Pro Ile Ser
20 25 30

aag gaa gac tta aga act aca att gac ctc ttg aaa caa gag tct cag 202
Lys Glu Asp Leu Arg Thr Thr Ile Asp Leu Leu Lys Gln Glu Ser Gln
35 40 45 50

gat ctt tat aac aac tat agc ata aag cag gca tct ggg atg tca gca 250
Asp Leu Tyr Asn Asn Tyr Ser Ile Lys Gln Ala Ser Gly Met Ser Ala
55 60 65

gac gaa tca ata cag ctg ccg tgt ttc agc ctg gac cgg gaa gca tta 298
Asp Glu Ser Ile Gln Leu Pro Cys Phe Ser Leu Asp Arg Glu Ala Leu
70 75 80

acc aac atc tcg gtc atc ata gca cat ctg gag aaa gtc aaa gtg ttg 346
Thr Asn Ile Ser Val Ile Ile Ala His Leu Glu Lys Val Lys Val Leu
85 90 95

agc gag aac aca gta gat act tct tgg gtg ata aga tgg cta aca aac 394
Ser Glu Asn Thr Val Asp Thr Ser Trp Val Ile Arg Trp Leu Thr Asn
100 105 110

atc agc tgt ttc aac cca ctg aat tta aac att tct gtg cct gga aat 442
Ile Ser Cys Phe Asn Pro Leu Asn Leu Asn Ile Ser Val Pro Gly Asn
115 120 125 130

act gat gaa tcc tat gat tgt aaa gtg ttc gtg ctt acg gtt tta aag 490
Thr Asp Glu Ser Tyr Asp Cys Lys Val Phe Val Leu Thr Val Leu Lys
135 140 145

cag ttc tca aac tgc atg gca gaa ctg cag gct aag gac aat act aca 538

Gln Phe Ser Asn Cys Met Ala Glu Leu Gln Ala Lys Asp Asn Thr Thr
150 155 160

tgc t gagtgatggg gggggggtgc agtgtcctca gcagtgcctg tccttcgagg 592
Cys

gctgagcttg caacccagga cttaactcca aagggactgt gcggtcatta ctagtcat 650

<210> 91
<211> 163
<212> PRT
<213> Mus musculus

<400> 91
Met Ile Phe His Thr Gly Thr Thr Lys Pro Thr Leu Val Leu Leu Cys
1 5 10 15
Cys Ile Gly Thr Trp Leu Ala Thr Cys Ser Leu Ser Phe Gly Ala Pro
20 25 30
Ile Ser Lys Glu Asp Leu Arg Thr Thr Ile Asp Leu Leu Lys Gln Glu
35 40 45
Ser Gln Asp Leu Tyr Asn Asn Tyr Ser Ile Lys Gln Ala Ser Gly Met
50 55 60
Ser Ala Asp Glu Ser Ile Gln Leu Pro Cys Phe Ser Leu Asp Arg Glu
65 70 75 80
Ala Leu Thr Asn Ile Ser Val Ile Ile Ala His Leu Glu Lys Val Lys
85 90 95
Val Leu Ser Glu Asn Thr Val Asp Thr Ser Trp Val Ile Arg Trp Leu
100 105 110
Thr Asn Ile Ser Cys Phe Asn Pro Leu Asn Leu Asn Ile Ser Val Pro
115 120 125
Gly Asn Thr Asp Glu Ser Tyr Asp Cys Lys Val Phe Val Leu Thr Val
130 135 140
Leu Lys Gln Phe Ser Asn Cys Met Ala Glu Leu Gln Ala Lys Asp Asn
145 150 155 160
Thr Thr Cys

<210> 92
<211> 511
<212> DNA
<213> Artificial Sequence

<220>
<223> Cloned murine cDNA sequence (SEQ ID NO:90) w/FseI and AscI restriction sites and a partial Kozak sequence to the mcytor17lig open reading frame and termination codon

<400> 92
ggccggccac catgatcttc cacacaggaa caacgaagcc taccctggtg ctgctttgct 60
gtataggaac ctggctggcc acctgcagct tgtccttcgg tgccccaata tcgaaggaag 120
acttaagaac tacaattgac ctcttgaaac aagagtctca ggatctttat aacaactata 180
gcataaagca ggcatctggg atgtcagcag acgaatcaat acagctgccg tgtttcagcc 240
tggaccggga agcattaacc aacatctcgg tcatcatagc acatctggag aaagtcaaag 300
tgttgagcga gaacacagta gatacttctt gggtgataag atggctaaca aacatcagct 360
gtttcaaccc actgaattta aacatttctg tgcctggaaa tactgatgaa tcctatgatt 420
gtaaagtgtt cgtgcttacg gttttaaagc agttctcaaa ctgcatggca gaactgcagg 480
ctaaggacaa tactacatgc tgaggcgcgc c 511

<210> 93
<211> 21

<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41438

<400> 93
gccatggcct ctcactcagg c 21

<210> 94
<211> 24
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41437

<400> 94
ccagggagca ttgacaactc ttag 24

<210> 95
<211> 516
<212> DNA
<213> Artificial Sequence

<220>
<223> human zCytor17Lig-CEE polynucleotide

<221> CDS
<222> (1)...(516)

<400> 95
atg gcc tct cac tca ggc ccc tcg acg tct gtg ctc ttt ctg ttc tgc 48
Met Ala Ser His Ser Gly Pro Ser Thr Ser Val Leu Phe Leu Phe Cys
1 5 10 15

tgc ctg gga ggc tgg ctg gcc tcc cac acg ttg ccc gtc cgt tta cta 96
Cys Leu Gly Gly Trp Leu Ala Ser His Thr Leu Pro Val Arg Leu Leu
20 25 30

cga cca agt gat gat gta cag aaa ata gtc gag gaa tta cag tcc ctc 144
Arg Pro Ser Asp Asp Val Gln Lys Ile Val Glu Glu Leu Gln Ser Leu
35 40 45

tcg aag atg ctt ttg aaa gat gtg gag gaa gag aag ggc gtg ctc gtg 192
Ser Lys Met Leu Leu Lys Asp Val Glu Glu Glu Lys Gly Val Leu Val
50 55 60

tcc cag aat tac acg ctg ccg tgt ctc agc cct gac gcc cag ccg cca 240
Ser Gln Asn Tyr Thr Leu Pro Cys Leu Ser Pro Asp Ala Gln Pro Pro
65 70 75 80

aac aac atc cac agc cca gcc atc cgg gca tat ctc aag aca atc aga 288
Asn Asn Ile His Ser Pro Ala Ile Arg Ala Tyr Leu Lys Thr Ile Arg
85 90 95

cag cta gac aac aaa tct gtt att gat gag atc ata gag cac ctc gac 336
Gln Leu Asp Asn Lys Ser Val Ile Asp Glu Ile Ile Glu His Leu Asp
100 105 110

aaa ctc ata ttt caa gat gca cca gaa aca aac att tct gtg cca aca 384
Lys Leu Ile Phe Gln Asp Ala Pro Glu Thr Asn Ile Ser Val Pro Thr
115 120 125

```
gac acc cat gaa tgt aaa cgc ttc atc ctg act att tct caa cag ttt   432
Asp Thr His Glu Cys Lys Arg Phe Ile Leu Thr Ile Ser Gln Gln Phe
    130                 135                 140

tca gag tgc atg gac ctc gca cta aaa tca ttg acc tct gga gcc caa   480
Ser Glu Cys Met Asp Leu Ala Leu Lys Ser Leu Thr Ser Gly Ala Gln
145                 150                 155                 160

cag gcc acc act gaa gaa tac atg ccg atg gaa taa                   516
Gln Ala Thr Thr Glu Glu Tyr Met Pro Met Glu  *
                165                 170
```

<210> 96
<211> 171
<212> PRT
<213> Artificial Sequence

<220>
<223> human zCytor17Lig-CEE polypeptide

<400> 96

```
Met Ala Ser His Ser Gly Pro Ser Thr Ser Val Leu Phe Leu Phe Cys
 1               5                  10                  15
Cys Leu Gly Gly Trp Leu Ala Ser His Thr Leu Pro Val Arg Leu Leu
            20                  25                  30
Arg Pro Ser Asp Asp Val Gln Lys Ile Val Glu Glu Leu Gln Ser Leu
        35                  40                  45
Ser Lys Met Leu Leu Lys Asp Val Glu Glu Glu Lys Gly Val Leu Val
    50                  55                  60
Ser Gln Asn Tyr Thr Leu Pro Cys Leu Ser Pro Asp Ala Gln Pro Pro
65                  70                  75                  80
Asn Asn Ile His Ser Pro Ala Ile Arg Ala Tyr Leu Lys Thr Ile Arg
                85                  90                  95
Gln Leu Asp Asn Lys Ser Val Ile Asp Glu Ile Ile Glu His Leu Asp
            100                 105                 110
Lys Leu Ile Phe Gln Asp Ala Pro Glu Thr Asn Ile Ser Val Pro Thr
        115                 120                 125
Asp Thr His Glu Cys Lys Arg Phe Ile Leu Thr Ile Ser Gln Gln Phe
    130                 135                 140
Ser Glu Cys Met Asp Leu Ala Leu Lys Ser Leu Thr Ser Gly Ala Gln
145                 150                 155                 160
Gln Ala Thr Thr Glu Glu Tyr Met Pro Met Glu
                165                 170
```

<210> 97
<211> 49
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41607

<400> 97
tccagggaat tcatataggc cggccaccat ggcctctcac tcaggcccc 49

<210> 98
<211> 82
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41605

<400> 98
caaccccaga gctgttttaa ggcgcgcctc tagattatta ttccatcggc atgtattctt 60
cagtggtggc ctgttgggct cc 82

<210> 99
<211> 629
<212> DNA
<213> Mus musculus

<220>
<221> CDS
<222> (28)...(528)

<400> 99
aacccttggg aggaccagaa cgagaca atg gtt ctt gcc agc tct acc acc agc 54
Met Val Leu Ala Ser Ser Thr Thr Ser
1 5

atc cac acc atg ctg ctc ctg ctc ctg atg ctc ttc cac ctg gga ctc 102
Ile His Thr Met Leu Leu Leu Leu Leu Met Leu Phe His Leu Gly Leu
10 15 20 25

caa gct tca atc agt ggc cgg gat acc cac cgt tta acc aga acg ttg 150
Gln Ala Ser Ile Ser Gly Arg Asp Thr His Arg Leu Thr Arg Thr Leu
30 35 40

aat tgc agc tct att gtc aag gag att ata ggg aag ctc cca gaa cct 198
Asn Cys Ser Ser Ile Val Lys Glu Ile Ile Gly Lys Leu Pro Glu Pro
45 50 55

gaa ctc aaa act gat gat gaa gga ccc tct ctg agg aat aag agc ttt 246
Glu Leu Lys Thr Asp Asp Glu Gly Pro Ser Leu Arg Asn Lys Ser Phe
60 65 70

cgg aga gta aac ctg tcc aaa ttc gtg gaa agc caa gga gaa gtg gat 294
Arg Arg Val Asn Leu Ser Lys Phe Val Glu Ser Gln Gly Glu Val Asp
75 80 85

cct gag gac aga tac gtt atc aag tcc aat ctt cag aaa ctt aac tgt 342
Pro Glu Asp Arg Tyr Val Ile Lys Ser Asn Leu Gln Lys Leu Asn Cys
90 95 100 105

tgc ctg cct aca tct gcg aat gac tct gcg ctg cca ggg gtc ttc att 390
Cys Leu Pro Thr Ser Ala Asn Asp Ser Ala Leu Pro Gly Val Phe Ile
110 115 120

cga gat ctg gat gac ttt cgg aag aaa ctg aga ttc tac atg gtc cac 438
Arg Asp Leu Asp Asp Phe Arg Lys Lys Leu Arg Phe Tyr Met Val His
125 130 135

ctt aac gat ctg gag aca gtg cta acc tct aga cca cct cag ccc gca 486
Leu Asn Asp Leu Glu Thr Val Leu Thr Ser Arg Pro Pro Gln Pro Ala
140 145 150

tct ggc tcc gtc tct cct aac cgt gga acc gtg gaa tgt taa 528
Ser Gly Ser Val Ser Pro Asn Arg Gly Thr Val Glu Cys *
155 160 165

aacagcaggc agagcaccta aagtctgaat gttcctcatg gcccatggtc aaaaggattt 588
tacattcctt tatgccatca aatgtcttat caatttatct a 629

<210> 100
<211> 166
<212> PRT
<213> Mus musculus

<400> 100
Met Val Leu Ala Ser Ser Thr Thr Ser Ile His Thr Met Leu Leu Leu
1 5 10 15
Leu Leu Met Leu Phe His Leu Gly Leu Gln Ala Ser Ile Ser Gly Arg
20 25 30
Asp Thr His Arg Leu Thr Arg Thr Leu Asn Cys Ser Ser Ile Val Lys
35 40 45
Glu Ile Ile Gly Lys Leu Pro Glu Pro Glu Leu Lys Thr Asp Asp Glu
50 55 60
Gly Pro Ser Leu Arg Asn Lys Ser Phe Arg Arg Val Asn Leu Ser Lys
65 70 75 80
Phe Val Glu Ser Gln Gly Glu Val Asp Pro Glu Asp Arg Tyr Val Ile
85 90 95
Lys Ser Asn Leu Gln Lys Leu Asn Cys Cys Leu Pro Thr Ser Ala Asn
100 105 110
Asp Ser Ala Leu Pro Gly Val Phe Ile Arg Asp Leu Asp Asp Phe Arg
115 120 125
Lys Lys Leu Arg Phe Tyr Met Val His Leu Asn Asp Leu Glu Thr Val
130 135 140
Leu Thr Ser Arg Pro Pro Gln Pro Ala Ser Gly Ser Val Ser Pro Asn
145 150 155 160
Arg Gly Thr Val Glu Cys
165

<210> 101
<211> 674
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (10)...(464)

<400> 101
gatccaaac atg agc cgc ctg ccc gtc ctg ctc ctg ctc caa ctc ctg gtc 51
Met Ser Arg Leu Pro Val Leu Leu Leu Leu Gln Leu Leu Val
1 5 10

cgc ccc gga ctc caa gct ccc atg acc cag aca acg tcc ttg aag aca 99
Arg Pro Gly Leu Gln Ala Pro Met Thr Gln Thr Thr Ser Leu Lys Thr
15 20 25 30

agc tgg gtt aac tgc tct aac atg atc gat gaa att ata aca cac tta 147
Ser Trp Val Asn Cys Ser Asn Met Ile Asp Glu Ile Ile Thr His Leu
35 40 45

aag cag cca cct ttg cct ttg ctg gac ttc aac aac ctc aat ggg gaa 195
Lys Gln Pro Pro Leu Pro Leu Leu Asp Phe Asn Asn Leu Asn Gly Glu
50 55 60

gac caa gac att ctg atg gaa aat aac ctt cga agg cca aac ctg gag 243
Asp Gln Asp Ile Leu Met Glu Asn Asn Leu Arg Arg Pro Asn Leu Glu
65 70 75

gca ttc aac agg gct gtc aag agt tta cag aac gca tca gca att gag 291
Ala Phe Asn Arg Ala Val Lys Ser Leu Gln Asn Ala Ser Ala Ile Glu

80 85 90

agc att ctt aaa aat ctc ctg cca tgt ctg ccc ctg gcc acg gcc gca 339
Ser Ile Leu Lys Asn Leu Leu Pro Cys Leu Pro Leu Ala Thr Ala Ala
95 100 105 110

ccc acg cga cat cca atc cat atc aag gac ggt gac tgg aat gaa ttc 387
Pro Thr Arg His Pro Ile His Ile Lys Asp Gly Asp Trp Asn Glu Phe
115 120 125

cgg agg aaa ctg acg ttc tat ctg aaa acc ctt gag aat gcg cag gct 435
Arg Arg Lys Leu Thr Phe Tyr Leu Lys Thr Leu Glu Asn Ala Gln Ala
130 135 140

caa cag acg act ttg agc ctc gcg atc tt ttagtccaac gtccagctcg 484
Gln Gln Thr Thr Leu Ser Leu Ala Ile
145 150

ttctctgggc cttctcacca cagcgcctcg ggacatcaaa aacagcagaa cttctgaaac 544
ctctgggtca tctctcacac attccaggac cagaagcatt tcacctttc ctgcggcatc 604
agatgaattg ttaattatct aatttctgaa atgtgcagct cccatttggc cttgtgcggt 664
tgtgttctca 674

<210> 102
<211> 151
<212> PRT
<213> Homo sapiens

<400> 102
Met Ser Arg Leu Pro Val Leu Leu Leu Leu Gln Leu Leu Val Arg Pro
1 5 10 15
Gly Leu Gln Ala Pro Met Thr Gln Thr Thr Ser Leu Lys Thr Ser Trp
20 25 30
Val Asn Cys Ser Asn Met Ile Asp Glu Ile Ile Thr His Leu Lys Gln
35 40 45
Pro Pro Leu Pro Leu Leu Asp Phe Asn Asn Leu Asn Gly Glu Asp Gln
50 55 60
Asp Ile Leu Met Glu Asn Asn Leu Arg Arg Pro Asn Leu Glu Ala Phe
65 70 75 80
Asn Arg Ala Val Lys Ser Leu Gln Asn Ala Ser Ala Ile Glu Ser Ile
85 90 95
Leu Lys Asn Leu Leu Pro Cys Leu Pro Leu Ala Thr Ala Ala Pro Thr
100 105 110
Arg His Pro Ile His Ile Lys Asp Gly Asp Trp Asn Glu Phe Arg Arg
115 120 125
Lys Leu Thr Phe Tyr Leu Lys Thr Leu Glu Asn Ala Gln Ala Gln Gln
130 135 140
Thr Thr Leu Ser Leu Ala Ile
145 150

<210> 103
<211> 7
<212> PRT
<213> Artificial Sequence

<220>
<223> Alternative Glu-Glu (CEE) peptide tag without
Gly-Ser residue pair

<400> 103
Glu Glu Tyr Met Pro Met Glu
1 5

<210> 104
<211> 513
<212> DNA
<213> Artificial Sequence

<220>
<223> Mouse zCytor17Lig(m)-CEE polynucleotide

<221> CDS
<222> (1)...(513)

<400> 104

```
atg atc ttc cac aca gga aca acg aag cct acc ctg gtg ctg ctt tgc   48
Met Ile Phe His Thr Gly Thr Thr Lys Pro Thr Leu Val Leu Leu Cys
 1               5                  10                  15

tgt ata gga acc tgg ctg gcc acc tgc agc ttg tcc ttc ggt gcc cca   96
Cys Ile Gly Thr Trp Leu Ala Thr Cys Ser Leu Ser Phe Gly Ala Pro
            20                  25                  30

ata tcg aag gaa gac tta aga act aca att gac ctc ttg aaa caa gag   144
Ile Ser Lys Glu Asp Leu Arg Thr Thr Ile Asp Leu Leu Lys Gln Glu
        35                  40                  45

tct cag gat ctt tat aac aac tat agc ata aag cag gca tct ggg atg   192
Ser Gln Asp Leu Tyr Asn Asn Tyr Ser Ile Lys Gln Ala Ser Gly Met
    50                  55                  60

tca gca gac gaa tca ata cag ctg ccg tgt ttc agc ctg gac cgg gaa   240
Ser Ala Asp Glu Ser Ile Gln Leu Pro Cys Phe Ser Leu Asp Arg Glu
65                  70                  75                  80

gca tta acc aac atc tcg gtc atc ata gca cat ctg gag aaa gtc aaa   288
Ala Leu Thr Asn Ile Ser Val Ile Ile Ala His Leu Glu Lys Val Lys
                85                  90                  95

gtg ttg agc gag aac aca gta gat act tct tgg gtg ata aga tgg cta   336
Val Leu Ser Glu Asn Thr Val Asp Thr Ser Trp Val Ile Arg Trp Leu
            100                 105                 110

aca aac atc agc tgt ttc aac cca ctg aat tta aac att tct gtg cct   384
Thr Asn Ile Ser Cys Phe Asn Pro Leu Asn Leu Asn Ile Ser Val Pro
        115                 120                 125

gga aat act gat gaa tcc tat gat tgt aaa gtg ttc gtg ctt acg gtt   432
Gly Asn Thr Asp Glu Ser Tyr Asp Cys Lys Val Phe Val Leu Thr Val
    130                 135                 140

tta aag cag ttc tca aac tgc atg gca gaa ctg cag gct aag gac aat   480
Leu Lys Gln Phe Ser Asn Cys Met Ala Glu Leu Gln Ala Lys Asp Asn
145                 150                 155                 160

act aca tgc gaa gaa tac atg ccg atg gaa tga                      513
Thr Thr Cys Glu Glu Tyr Met Pro Met Glu  *
                165                 170
```

<210> 105
<211> 170
<212> PRT
<213> Artificial Sequence

<220>
<223> Mouse zCytor17Lig(m)-CEE polypeptide

<400> 105
Met Ile Phe His Thr Gly Thr Thr Lys Pro Thr Leu Val Leu Leu Cys
1 5 10 15
Cys Ile Gly Thr Trp Leu Ala Thr Cys Ser Leu Ser Phe Gly Ala Pro
20 25 30
Ile Ser Lys Glu Asp Leu Arg Thr Thr Ile Asp Leu Leu Lys Gln Glu
35 40 45
Ser Gln Asp Leu Tyr Asn Asn Tyr Ser Ile Lys Gln Ala Ser Gly Met
50 55 60
Ser Ala Asp Glu Ser Ile Gln Leu Pro Cys Phe Ser Leu Asp Arg Glu
65 70 75 80
Ala Leu Thr Asn Ile Ser Val Ile Ile Ala His Leu Glu Lys Val Lys
85 90 95
Val Leu Ser Glu Asn Thr Val Asp Thr Ser Trp Val Ile Arg Trp Leu
100 105 110
Thr Asn Ile Ser Cys Phe Asn Pro Leu Asn Leu Asn Ile Ser Val Pro
115 120 125
Gly Asn Thr Asp Glu Ser Tyr Asp Cys Lys Val Phe Val Leu Thr Val
130 135 140
Leu Lys Gln Phe Ser Asn Cys Met Ala Glu Leu Gln Ala Lys Asp Asn
145 150 155 160
Thr Thr Cys Glu Glu Tyr Met Pro Met Glu
165 170

<210> 106
<211> 49
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41643

<400> 106
tccagggaat tcatataggc cggccaccat gatcttccac acaggaaca 49

<210> 107
<211> 85
<212> DNA
<213> Artificial Sequence

<220>
<223> Oligonucleotide primer ZC41641

<400> 107
caaccccaga gctgttttaa ggcgcgcctc tagattatca ttccatcggc atgtattctt 60
cgcatgtagt attgtcctta gcctg 85

<210> 108
<211> 2529
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (162)...(2108)

<400> 108
tgtgtgtgca gtatgaaaat tgagacagga aggcagagtg tcagcttgtt ccacctcagc 60

```
tgggaatgtg catcaggcaa ctcaagtttt tcaccacggc atgtgtctgt gaatgtccgc 120
aaaacattct ctctccccag ccttcatgtg ttaacctggg g atg atg tgg acc tgg 176
                                              Met Met Trp Thr Trp
                                               1               5

gca ctg tgg atg ctc ccc tca ctc tgc aaa ttc agc ctg gca gct ctg   224
Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe Ser Leu Ala Ala Leu
                10                  15                  20

cca gct aag cct gag aac att tcc tgt gtc tac tac tat agg aaa aat   272
Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr Tyr Tyr Arg Lys Asn
            25                  30                  35

tta acc tgc act tgg agt cca gga aag gaa acc agt tat acc cag tac   320
Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr Ser Tyr Thr Gln Tyr
        40                  45                  50

aca gtt aag aga act tac gct ttt gga gaa aaa cat gat aat tgt aca   368
Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys His Asp Asn Cys Thr
    55                  60                  65

acc aat agt tct aca agt gaa aat cgt gct tcg tgc tct ttt ttc ctt   416
Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser Cys Ser Phe Phe Leu
70                  75                  80                  85

cca aga ata acg atc cca gat aat tat acc att gag gtg gaa gct gaa   464
Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile Glu Val Glu Ala Glu
                90                  95                  100

aat gga gat ggt gta att aaa tct cat atg aca tac tgg aga tta gag   512
Asn Gly Asp Gly Val Ile Lys Ser His Met Thr Tyr Trp Arg Leu Glu
            105                 110                 115

aac ata gcg aaa act gaa cca cct aag att ttc cgt gtg aaa cca gtt   560
Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe Arg Val Lys Pro Val
        120                 125                 130

ttg ggc atc aaa cga atg att caa att gaa tgg ata aag cct gag ttg   608
Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp Ile Lys Pro Glu Leu
    135                 140                 145

gcg cct gtt tca tct gat tta aaa tac aca ctt cga ttc agg aca gtc   656
Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu Arg Phe Arg Thr Val
150                 155                 160                 165

aac agt acc agc tgg atg gaa gtc aac ttc gct aag aac cgt aag gat   704
Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala Lys Asn Arg Lys Asp
                170                 175                 180

aaa aac caa acg tac aac ctc acg ggg ctg cag cct ttt aca gaa tat   752
Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln Pro Phe Thr Glu Tyr
            185                 190                 195

gtc ata gct ctg cga tgt gcg gtc aag gag tca aag ttc tgg agt gac   800
Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser Lys Phe Trp Ser Asp
        200                 205                 210

tgg agc caa gaa aaa atg gga atg act gag gaa gaa gct cca tgt ggc   848
Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu Glu Ala Pro Cys Gly
    215                 220                 225

ctg gaa ctg tgg aga gtc ctg aaa cca gct gag gcg gat gga aga agg   896
Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu Ala Asp Gly Arg Arg
```

```
230                 235                 240                 245

cca gtg cgg ttg tta tgg aag aag gca aga gga gcc cca gtc cta gag   944
Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val Leu Glu
                250                 255                 260

aaa aca ctt ggc tac aac ata tgg tac tat cca gaa agc aac act aac   992
Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro Glu Ser Asn Thr Asn
            265                 270                 275

ctc aca gaa aca atg aac act act aac cag cag ctt gaa ctg cat ctg   1040
Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln Leu Glu Leu His Leu
        280                 285                 290

gga ggc gag agc ttt tgg gtg tct atg att tct tat aat tct ctt ggg   1088
Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser Tyr Asn Ser Leu Gly
    295                 300                 305

aag tct cca gtg gcc acc ctg agg att cca gct att caa gaa aaa tca   1136
Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala Ile Gln Glu Lys Ser
310                 315                 320                 325

ttt cag tgc att gag gtc atg cag gcc tgc gtt gct gag gac cag cta   1184
Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val Ala Glu Asp Gln Leu
                330                 335                 340

gtg gtg aag tgg caa agc tct gct cta gac gtg aac act tgg atg att   1232
Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val Asn Thr Trp Met Ile
            345                 350                 355

gaa tgg ttt ccg gat gtg gac tca gag ccc acc acc ctt tcc tgg gaa   1280
Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr Thr Leu Ser Trp Glu
        360                 365                 370

tct gtg tct cag gcc acg aac tgg acg atc cag caa gat aaa tta aaa   1328
Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln Gln Asp Lys Leu Lys
    375                 380                 385

cct ttc tgg tgc tat aac atc tct gtg tat cca atg ttg cat gac aaa   1376
Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro Met Leu His Asp Lys
390                 395                 400                 405

gtt ggc gag cca tat tcc atc cag gct tat gcc aaa gaa ggc gtt cca   1424
Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu Gly Val Pro
                410                 415                 420

tca gaa ggt cct gag acc aag gtg gag aac att ggc gtg aag acg gtc   1472
Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile Gly Val Lys Thr Val
            425                 430                 435

acg atc aca tgg aaa gag att ccc aag agt gag aga aag ggt atc atc   1520
Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu Arg Lys Gly Ile Ile
        440                 445                 450

tgc aac tac acc atc ttt tac caa gct gaa ggt gga aaa gga ttc tcc   1568
Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly Gly Lys Gly Phe Ser
    455                 460                 465

aag aca gtc aat tcc agc atc ttg cag tac ggc ctg gag tcc ctg aaa   1616
Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly Leu Glu Ser Leu Lys
470                 475                 480                 485

cga aag acc tct tac att gtt cag gtc atg gcc agc acc agt gct ggg   1664
```

```
Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala Ser Thr Ser Ala Gly
                490                 495                 500

gga acc aac ggg acc agc ata aat ttc aag aca ttg tca ttc agt gtc   1712
Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr Leu Ser Phe Ser Val
            505                 510                 515

ttt gag att atc ctc ata act tct ctg att ggt gga ggc ctt ctt att   1760
Phe Glu Ile Ile Leu Ile Thr Ser Leu Ile Gly Gly Gly Leu Leu Ile
        520                 525                 530

ctc att atc ctg aca gtg gca tat ggt ctc aaa aaa ccc aac aaa ttg   1808
Leu Ile Ile Leu Thr Val Ala Tyr Gly Leu Lys Lys Pro Asn Lys Leu
    535                 540                 545

act cat ctg tgt tgg ccc acc gtt ccc aac cct gct gaa agt agt ata   1856
Thr His Leu Cys Trp Pro Thr Val Pro Asn Pro Ala Glu Ser Ser Ile
550                 555                 560                 565

gcc aca tgg cat gga gat gat ttc aag gat aag cta aac ctg aag gag   1904
Ala Thr Trp His Gly Asp Asp Phe Lys Asp Lys Leu Asn Leu Lys Glu
                570                 575                 580

tct gat gac tct gtg aac aca gaa gac agg atc tta aaa cca tgt tcc   1952
Ser Asp Asp Ser Val Asn Thr Glu Asp Arg Ile Leu Lys Pro Cys Ser
            585                 590                 595

acc ccc agt gac aag ttg gtg att gac aag ttg gtg gtg aac ttt ggg   2000
Thr Pro Ser Asp Lys Leu Val Ile Asp Lys Leu Val Val Asn Phe Gly
        600                 605                 610

aat gtt ctg caa gaa att ttc aca gat gaa gcc aga acg ggt cag gaa   2048
Asn Val Leu Gln Glu Ile Phe Thr Asp Glu Ala Arg Thr Gly Gln Glu
    615                 620                 625

aac aat tta gga ggg gaa aag aat ggg act aga att ctg tct tcc tgc   2096
Asn Asn Leu Gly Gly Glu Lys Asn Gly Thr Arg Ile Leu Ser Ser Cys
630                 635                 640                 645

cca act tca ata taagtgtgga ctaaaatgcg agaaaggtgt cctgtggtct       2148
Pro Thr Ser Ile

atgcaaatta gaaaggacat gcagagtttt ccaactagga agactgaatc tgtggcccca 2208
agagaaccat ctctgaagac tgggtatgtg gtcttttcca cacatggacc acctacggat 2268
gcaatctgta atgcatgtgc atgagaagtc tgttattaag tagagtgtga aaacatggtt 2328
atggtaatag gaacagcttt taaaatgctt ttgtatttgg gcctttcata caaaaaagcc 2388
ataataccat tttcatgtaa tgctatactt ctatactatt ttcatgtaat actatacttc 2448
tatactattt tcatgtaata ctatacttct atactatttt catgtaatac tatacttcta 2508
tattaaagtt ttacccactc a                                           2529

<210> 109
<211> 649
<212> PRT
<213> Homo sapiens

<400> 109
Met Met Trp Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe
 1               5                  10                  15
Ser Leu Ala Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr
            20                  25                  30
Tyr Tyr Arg Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr
        35                  40                  45
```

Ser Tyr Thr Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys
50 55 60
His Asp Asn Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser
65 70 75 80
Cys Ser Phe Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile
85 90 95
Glu Val Glu Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr
100 105 110
Tyr Trp Arg Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe
115 120 125
Arg Val Lys Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp
130 135 140
Ile Lys Pro Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu
145 150 155 160
Arg Phe Arg Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala
165 170 175
Lys Asn Arg Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln
180 185 190
Pro Phe Thr Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser
195 200 205
Lys Phe Trp Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu
210 215 220
Glu Ala Pro Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu
225 230 235 240
Ala Asp Gly Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly
245 250 255
Ala Pro Val Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro
260 265 270
Glu Ser Asn Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln
275 280 285
Leu Glu Leu His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser
290 295 300
Tyr Asn Ser Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala
305 310 315 320
Ile Gln Glu Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val
325 330 335
Ala Glu Asp Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val
340 345 350
Asn Thr Trp Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr
355 360 365
Thr Leu Ser Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln
370 375 380
Gln Asp Lys Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro
385 390 395 400
Met Leu His Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala
405 410 415
Lys Glu Gly Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile
420 425 430
Gly Val Lys Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu
435 440 445
Arg Lys Gly Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly
450 455 460
Gly Lys Gly Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly
465 470 475 480
Leu Glu Ser Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala
485 490 495
Ser Thr Ser Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr
500 505 510
Leu Ser Phe Ser Val Phe Glu Ile Ile Leu Ile Thr Ser Leu Ile Gly
515 520 525
Gly Gly Leu Leu Ile Leu Ile Ile Leu Thr Val Ala Tyr Gly Leu Lys
530 535 540
Lys Pro Asn Lys Leu Thr His Leu Cys Trp Pro Thr Val Pro Asn Pro

```
545                 550                 555                 560
Ala Glu Ser Ser Ile Ala Thr Trp His Gly Asp Asp Phe Lys Asp Lys
                565                 570                 575
Leu Asn Leu Lys Glu Ser Asp Asp Ser Val Asn Thr Glu Asp Arg Ile
            580                 585                 590
Leu Lys Pro Cys Ser Thr Pro Ser Asp Lys Leu Val Ile Asp Lys Leu
        595                 600                 605
Val Val Asn Phe Gly Asn Val Leu Gln Glu Ile Phe Thr Asp Glu Ala
    610                 615                 620
Arg Thr Gly Gln Glu Asn Asn Leu Gly Gly Glu Lys Asn Gly Thr Arg
625                 630                 635                 640
Ile Leu Ser Ser Cys Pro Thr Ser Ile
                645
```

<210> 110
<211> 2402
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (171)...(2366)

<400> 110
ggcacgaggt gtgtgtgcag tatgaaaatt gagacaggaa ggcagagtgt cagcttgttc 60
cacctcagct gggaatgtgc atcaggcaac tcaagttttt caccacggca tgtgtctgtg 120
aatgtccgca aaacattctc tctccccagc cttcatgtgt taacctgggg atg atg 176
                                                    Met Met
                                                     1

```
tgg acc tgg gca ctg tgg atg ctc ccc tca ctc tgc aaa ttc agc ctg   224
Trp Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe Ser Leu
          5                  10                  15

gca gct ctg cca gct aag cct gag aac att tcc tgt gtc tac tac tat   272
Ala Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr Tyr Tyr
      20                  25                  30

agg aaa aat tta acc tgc act tgg agt cca gga aag gaa acc agt tat   320
Arg Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr Ser Tyr
  35                  40                  45                  50

acc cag tac aca gtt aag aga act tac gct ttt gga gaa aaa cat gat   368
Thr Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys His Asp
                  55                  60                  65

aat tgt aca acc aat agt tct aca agt gaa aat cgt gct tcg tgc tct   416
Asn Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser Cys Ser
              70                  75                  80

ttt ttc ctt cca aga ata acg atc cca gat aat tat acc att gag gtg   464
Phe Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile Glu Val
          85                  90                  95

gaa gct gaa aat gga gat ggt gta att aaa tct cat atg aca tac tgg   512
Glu Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr Tyr Trp
      100                 105                 110

aga tta gag aac ata gcg aaa act gaa cca cct aag att ttc cgt gtg   560
Arg Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe Arg Val
  115                 120                 125                 130
```

aaa cca gtt ttg ggc atc aaa cga atg att caa att gaa tgg ata aag 608
Lys Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp Ile Lys
135 140 145

cct gag ttg gcg cct gtt tca tct gat tta aaa tac aca ctt cga ttc 656
Pro Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu Arg Phe
150 155 160

agg aca gtc aac agt acc agc tgg atg gaa gtc aac ttc gct aag aac 704
Arg Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala Lys Asn
165 170 175

cgt aag gat aaa aac caa acg tac aac ctc acg ggg ctg cag cct ttt 752
Arg Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln Pro Phe
180 185 190

aca gaa tat gtc ata gct ctg cga tgt gcg gtc aag gag tca aag ttc 800
Thr Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser Lys Phe
195 200 205 210

tgg agt gac tgg agc caa gaa aaa atg gga atg act gag gaa gaa gct 848
Trp Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu Glu Ala
215 220 225

cca tgt ggc ctg gaa ctg tgg aga gtc ctg aaa cca gct gag gcg gat 896
Pro Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu Ala Asp
230 235 240

gga aga agg cca gtg cgg ttg tta tgg aag aag gca aga gga gcc cca 944
Gly Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro
245 250 255

gtc cta gag aaa aca ctt ggc tac aac ata tgg tac tat cca gaa agc 992
Val Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro Glu Ser
260 265 270

aac act aac ctc aca gaa aca atg aac act act aac cag cag ctt gaa 1040
Asn Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln Leu Glu
275 280 285 290

ctg cat ctg gga ggc gag agc ttt tgg gtg tct atg att tct tat aat 1088
Leu His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser Tyr Asn
295 300 305

tct ctt ggg aag tct cca gtg gcc acc ctg agg att cca gct att caa 1136
Ser Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala Ile Gln
310 315 320

gaa aaa tca ttt cag tgc att gag gtc atg cag gcc tgc gtt gct gag 1184
Glu Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val Ala Glu
325 330 335

gac cag cta gtg gtg aag tgg caa agc tct gct cta gac gtg aac act 1232
Asp Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val Asn Thr
340 345 350

tgg atg att gaa tgg ttt ccg gat gtg gac tca gag ccc acc acc ctt 1280
Trp Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr Thr Leu
355 360 365 370

tcc tgg gaa tct gtg tct cag gcc acg aac tgg acg atc cag caa gat 1328
Ser Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln Gln Asp
375 380 385

aaa tta aaa cct ttc tgg tgc tat aac atc tct gtg tat cca atg ttg 1376
Lys Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro Met Leu
390 395 400

cat gac aaa gtt ggc gag cca tat tcc atc cag gct tat gcc aaa gaa 1424
His Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu
405 410 415

ggc gtt cca tca gaa ggt cct gag acc aag gtg gag aac att ggc gtg 1472
Gly Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile Gly Val
420 425 430

aag acg gtc acg atc aca tgg aaa gag att ccc aag agt gag aga aag 1520
Lys Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu Arg Lys
435 440 445 450

ggt atc atc tgc aac tac acc atc ttt tac caa gct gaa ggt gga aaa 1568
Gly Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly Gly Lys
455 460 465

gga ttc tcc aag aca gtc aat tcc agc atc ttg cag tac ggc ctg gag 1616
Gly Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly Leu Glu
470 475 480

tcc ctg aaa cga aag acc tct tac att gtt cag gtc atg gcc agc acc 1664
Ser Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala Ser Thr
485 490 495

agt gct ggg gga acc aac ggg acc agc ata aat ttc aag aca ttg tca 1712
Ser Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr Leu Ser
500 505 510

ttc agt gtc ttt gag att atc ctc ata act tct ctg att ggt gga ggc 1760
Phe Ser Val Phe Glu Ile Ile Leu Ile Thr Ser Leu Ile Gly Gly Gly
515 520 525 530

ctt ctt att ctc att atc ctg aca gtg gca tat ggt ctc aaa aaa ccc 1808
Leu Leu Ile Leu Ile Ile Leu Thr Val Ala Tyr Gly Leu Lys Lys Pro
535 540 545

aac aaa ttg act cat ctg tgt tgg ccc acc gtt ccc aac cct gct gaa 1856
Asn Lys Leu Thr His Leu Cys Trp Pro Thr Val Pro Asn Pro Ala Glu
550 555 560

agt agt ata gcc aca tgg cat gga gat gat ttc aag gat aag cta aac 1904
Ser Ser Ile Ala Thr Trp His Gly Asp Asp Phe Lys Asp Lys Leu Asn
565 570 575

ctg aag gag tct gat gac tct gtg aac aca gaa gac agg atc tta aaa 1952
Leu Lys Glu Ser Asp Asp Ser Val Asn Thr Glu Asp Arg Ile Leu Lys
580 585 590

cca tgt tcc acc ccc agt gac aag ttg gtg att gac aag ttg gtg gtg 2000
Pro Cys Ser Thr Pro Ser Asp Lys Leu Val Ile Asp Lys Leu Val Val
595 600 605 610

aac ttt ggg aat gtt ctg caa gaa att ttc aca gat gaa gcc aga acg 2048
Asn Phe Gly Asn Val Leu Gln Glu Ile Phe Thr Asp Glu Ala Arg Thr
615 620 625

ggt cag gaa aac aat tta gga ggg gaa aag aat ggg tat gtg acc tgc 2096
Gly Gln Glu Asn Asn Leu Gly Gly Glu Lys Asn Gly Tyr Val Thr Cys

630 635 640

ccc ttc agg cct gat tgt ccc ctg ggg aaa agt ttt gag gag ctc cca 2144
Pro Phe Arg Pro Asp Cys Pro Leu Gly Lys Ser Phe Glu Glu Leu Pro
645 650 655

gtt tca cct gag att ccg ccc aga aaa tcc caa tac cta cgt tcg agg 2192
Val Ser Pro Glu Ile Pro Pro Arg Lys Ser Gln Tyr Leu Arg Ser Arg
660 665 670

atg cca gag ggg acc cgc cca gaa gcc aaa gag cag ctt ctc ttt tct 2240
Met Pro Glu Gly Thr Arg Pro Glu Ala Lys Glu Gln Leu Leu Phe Ser
675 680 685 690

ggt caa agt tta gta cca gat cat ctg tgt gag gaa gga gcc cca aat 2288
Gly Gln Ser Leu Val Pro Asp His Leu Cys Glu Glu Gly Ala Pro Asn
695 700 705

cca tat ttg aaa aat tca gtg aca gcc agg gaa ttt ctt gtg tct gaa 2336
Pro Tyr Leu Lys Asn Ser Val Thr Ala Arg Glu Phe Leu Val Ser Glu
710 715 720

aaa ctt cca gag cac acc aag gga gaa gtc taaatgcgac catagcatga 2386
Lys Leu Pro Glu His Thr Lys Gly Glu Val
725 730

gaccctcggg gcctca 2402

<210> 111
<211> 732
<212> PRT
<213> Homo sapiens

<400> 111
Met Met Trp Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe
1 5 10 15
Ser Leu Ala Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr
20 25 30
Tyr Tyr Arg Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr
35 40 45
Ser Tyr Thr Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys
50 55 60
His Asp Asn Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser
65 70 75 80
Cys Ser Phe Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile
85 90 95
Glu Val Glu Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr
100 105 110
Tyr Trp Arg Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe
115 120 125
Arg Val Lys Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp
130 135 140
Ile Lys Pro Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu
145 150 155 160
Arg Phe Arg Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala
165 170 175
Lys Asn Arg Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln
180 185 190
Pro Phe Thr Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser
195 200 205
Lys Phe Trp Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu
210 215 220
Glu Ala Pro Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu

225 230 235 240
Ala Asp Gly Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly
245 250 255
Ala Pro Val Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro
260 265 270
Glu Ser Asn Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln
275 280 285
Leu Glu Leu His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser
290 295 300
Tyr Asn Ser Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala
305 310 315 320
Ile Gln Glu Lys Ser Phe Gln Cys Ile Glu Val Met Gln Ala Cys Val
325 330 335
Ala Glu Asp Gln Leu Val Val Lys Trp Gln Ser Ser Ala Leu Asp Val
340 345 350
Asn Thr Trp Met Ile Glu Trp Phe Pro Asp Val Asp Ser Glu Pro Thr
355 360 365
Thr Leu Ser Trp Glu Ser Val Ser Gln Ala Thr Asn Trp Thr Ile Gln
370 375 380
Gln Asp Lys Leu Lys Pro Phe Trp Cys Tyr Asn Ile Ser Val Tyr Pro
385 390 395 400
Met Leu His Asp Lys Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala
405 410 415
Lys Glu Gly Val Pro Ser Glu Gly Pro Glu Thr Lys Val Glu Asn Ile
420 425 430
Gly Val Lys Thr Val Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Glu
435 440 445
Arg Lys Gly Ile Ile Cys Asn Tyr Thr Ile Phe Tyr Gln Ala Glu Gly
450 455 460
Gly Lys Gly Phe Ser Lys Thr Val Asn Ser Ser Ile Leu Gln Tyr Gly
465 470 475 480
Leu Glu Ser Leu Lys Arg Lys Thr Ser Tyr Ile Val Gln Val Met Ala
485 490 495
Ser Thr Ser Ala Gly Gly Thr Asn Gly Thr Ser Ile Asn Phe Lys Thr
500 505 510
Leu Ser Phe Ser Val Phe Glu Ile Ile Leu Ile Thr Ser Leu Ile Gly
515 520 525
Gly Gly Leu Leu Ile Leu Ile Ile Leu Thr Val Ala Tyr Gly Leu Lys
530 535 540
Lys Pro Asn Lys Leu Thr His Leu Cys Trp Pro Thr Val Pro Asn Pro
545 550 555 560
Ala Glu Ser Ser Ile Ala Thr Trp His Gly Asp Asp Phe Lys Asp Lys
565 570 575
Leu Asn Leu Lys Glu Ser Asp Asp Ser Val Asn Thr Glu Asp Arg Ile
580 585 590
Leu Lys Pro Cys Ser Thr Pro Ser Asp Lys Leu Val Ile Asp Lys Leu
595 600 605
Val Val Asn Phe Gly Asn Val Leu Gln Glu Ile Phe Thr Asp Glu Ala
610 615 620
Arg Thr Gly Gln Glu Asn Asn Leu Gly Gly Glu Lys Asn Gly Tyr Val
625 630 635 640
Thr Cys Pro Phe Arg Pro Asp Cys Pro Leu Gly Lys Ser Phe Glu Glu
645 650 655
Leu Pro Val Ser Pro Glu Ile Pro Pro Arg Lys Ser Gln Tyr Leu Arg
660 665 670
Ser Arg Met Pro Glu Gly Thr Arg Pro Glu Ala Lys Glu Gln Leu Leu
675 680 685
Phe Ser Gly Gln Ser Leu Val Pro Asp His Leu Cys Glu Glu Gly Ala
690 695 700
Pro Asn Pro Tyr Leu Lys Asn Ser Val Thr Ala Arg Glu Phe Leu Val
705 710 715 720
Ser Glu Lys Leu Pro Glu His Thr Lys Gly Glu Val
725 730

<210> 112
<211> 1299
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (162)...(1133)

<400> 112
tgtgtgtgca gtatgaaaat tgagacagga aggcagagtg tcagcttgtt ccacctcagc 60
tgggaatgtg catcaggcaa ctcaagtttt tcaccacggc atgtgtctgt gaatgtccgc 120
aaaacattct ctctccccag ccttcatgtg ttaacctggg g atg atg tgg acc tgg 176
Met Met Trp Thr Trp
1 5

gca ctg tgg atg ctc ccc tca ctc tgc aaa ttc agc ctg gca gct ctg 224
Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe Ser Leu Ala Ala Leu
10 15 20

cca gct aag cct gag aac att tcc tgt gtc tac tac tat agg aaa aat 272
Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr Tyr Tyr Arg Lys Asn
25 30 35

tta acc tgc act tgg agt cca gga aag gaa acc agt tat acc cag tac 320
Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr Ser Tyr Thr Gln Tyr
40 45 50

aca gtt aag aga act tac gct ttt gga gaa aaa cat gat aat tgt aca 368
Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys His Asp Asn Cys Thr
55 60 65

acc aat agt tct aca agt gaa aat cgt gct tcg tgc tct ttt ttc ctt 416
Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser Cys Ser Phe Phe Leu
70 75 80 85

cca aga ata acg atc cca gat aat tat acc att gag gtg gaa gct gaa 464
Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile Glu Val Glu Ala Glu
90 95 100

aat gga gat ggt gta att aaa tct cat atg aca tac tgg aga tta gag 512
Asn Gly Asp Gly Val Ile Lys Ser His Met Thr Tyr Trp Arg Leu Glu
105 110 115

aac ata gcg aaa act gaa cca cct aag att ttc cgt gtg aaa cca gtt 560
Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe Arg Val Lys Pro Val
120 125 130

ttg ggc atc aaa cga atg att caa att gaa tgg ata aag cct gag ttg 608
Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp Ile Lys Pro Glu Leu
135 140 145

gcg cct gtt tca tct gat tta aaa tac aca ctt cga ttc agg aca gtc 656
Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu Arg Phe Arg Thr Val
150 155 160 165

aac agt acc agc tgg atg gaa gtc aac ttc gct aag aac cgt aag gat 704
Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala Lys Asn Arg Lys Asp
170 175 180

aaa aac caa acg tac aac ctc acg ggg ctg cag cct ttt aca gaa tat 752

```
Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln Pro Phe Thr Glu Tyr
            185                 190                 195

gtc ata gct ctg cga tgt gcg gtc aag gag tca aag ttc tgg agt gac   800
Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser Lys Phe Trp Ser Asp
        200                 205                 210

tgg agc caa gaa aaa atg gga atg act gag gaa gaa gct cca tgt ggc   848
Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu Glu Ala Pro Cys Gly
    215                 220                 225

ctg gaa ctg tgg aga gtc ctg aaa cca gct gag gcg gat gga aga agg   896
Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu Ala Asp Gly Arg Arg
230                 235                 240                 245

cca gtg cgg ttg tta tgg aag aag gca aga gga gcc cca gtc cta gag   944
Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val Leu Glu
                250                 255                 260

aaa aca ctt ggc tac aac ata tgg tac tat cca gaa agc aac act aac   992
Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro Glu Ser Asn Thr Asn
            265                 270                 275

ctc aca gaa aca atg aac act act aac cag cag ctt gaa ctg cat ctg  1040
Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln Leu Glu Leu His Leu
        280                 285                 290

gga ggc gag agc ttt tgg gtg tct atg att tct tat aat tct ctt ggg  1088
Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser Tyr Asn Ser Leu Gly
    295                 300                 305

aag tct cca gtg gcc acc ctg agg att cca gct att caa gaa aaa      1133
Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala Ile Gln Glu Lys
310                 315                 320

tagaaacttt acagatgcta gtcccagaca taaaagaaaa taatgttctg gatgtgcacg 1193
atggctcacg cctgtaatcc cagcactttg aggccaagac gggtggatcg ctgagttcag 1253
gagttcaaga caagtccagg caacatagtg aaaccttgtt tctaca               1299

<210> 113
<211> 324
<212> PRT
<213> Homo sapiens

<400> 113
Met Met Trp Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe
 1               5                  10                  15
Ser Leu Ala Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr
            20                  25                  30
Tyr Tyr Arg Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr
        35                  40                  45
Ser Tyr Thr Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys
    50                  55                  60
His Asp Asn Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser
65                  70                  75                  80
Cys Ser Phe Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile
                85                  90                  95
Glu Val Glu Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr
            100                 105                 110
Tyr Trp Arg Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe
        115                 120                 125
Arg Val Lys Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp
    130                 135                 140
```

```
Ile Lys Pro Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu
145                 150                 155                 160
Arg Phe Arg Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala
                165                 170                 175
Lys Asn Arg Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln
            180                 185                 190
Pro Phe Thr Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser
        195                 200                 205
Lys Phe Trp Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu
    210                 215                 220
Glu Ala Pro Cys Gly Leu Glu Leu Trp Arg Val Leu Lys Pro Ala Glu
225                 230                 235                 240
Ala Asp Gly Arg Arg Pro Val Arg Leu Leu Trp Lys Lys Ala Arg Gly
                245                 250                 255
Ala Pro Val Leu Glu Lys Thr Leu Gly Tyr Asn Ile Trp Tyr Tyr Pro
            260                 265                 270
Glu Ser Asn Thr Asn Leu Thr Glu Thr Met Asn Thr Thr Asn Gln Gln
        275                 280                 285
Leu Glu Leu His Leu Gly Gly Glu Ser Phe Trp Val Ser Met Ile Ser
    290                 295                 300
Tyr Asn Ser Leu Gly Lys Ser Pro Val Ala Thr Leu Arg Ile Pro Ala
305                 310                 315                 320
Ile Gln Glu Lys
```

<210> 114
<211> 1476
<212> DNA
<213> Homo sapiens

<220>
<221> CDS
<222> (162)...(878)

<400> 114

```
tgtgtgtgca gtatgaaaat tgagacagga aggcagagtg tcagcttgtt ccacctcagc 60
tgggaatgtg catcaggcaa ctcaagtttt tcaccacggc atgtgtctgt gaatgtccgc 120
aaaacattct ctctccccag ccttcatgtg ttaacctggg g atg atg tgg acc tgg 176
                                                 Met Met Trp Thr Trp
                                                  1               5

gca ctg tgg atg ctc ccc tca ctc tgc aaa ttc agc ctg gca gct ctg   224
Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe Ser Leu Ala Ala Leu
                10                  15                  20

cca gct aag cct gag aac att tcc tgt gtc tac tac tat agg aaa aat   272
Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr Tyr Tyr Arg Lys Asn
            25                  30                  35

tta acc tgc act tgg agt cca gga aag gaa acc agt tat acc cag tac   320
Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr Ser Tyr Thr Gln Tyr
        40                  45                  50

aca gtt aag aga act tac gct ttt gga gaa aaa cat gat aat tgt aca   368
Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys His Asp Asn Cys Thr
    55                  60                  65

acc aat agt tct aca agt gaa aat cgt gct tcg tgc tct ttt ttc ctt   416
Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser Cys Ser Phe Phe Leu
70                  75                  80                  85

cca aga ata acg atc cca gat aat tat acc att gag gtg gaa gct gaa   464
```

```
Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile Glu Val Glu Ala Glu
                 90                  95                 100

aat gga gat ggt gta att aaa tct cat atg aca tac tgg aga tta gag   512
Asn Gly Asp Gly Val Ile Lys Ser His Met Thr Tyr Trp Arg Leu Glu
            105                 110                 115

aac ata gcg aaa act gaa cca cct aag att ttc cgt gtg aaa cca gtt   560
Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe Arg Val Lys Pro Val
        120                 125                 130

ttg ggc atc aaa cga atg att caa att gaa tgg ata aag cct gag ttg   608
Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp Ile Lys Pro Glu Leu
    135                 140                 145

gcg cct gtt tca tct gat tta aaa tac aca ctt cga ttc agg aca gtc   656
Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu Arg Phe Arg Thr Val
150                 155                 160                 165

aac agt acc agc tgg atg gaa gtc aac ttc gct aag aac cgt aag gat   704
Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala Lys Asn Arg Lys Asp
                170                 175                 180

aaa aac caa acg tac aac ctc acg ggg ctg cag cct ttt aca gaa tat   752
Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln Pro Phe Thr Glu Tyr
            185                 190                 195

gtc ata gct ctg cga tgt gcg gtc aag gag tca aag ttc tgg agt gac   800
Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser Lys Phe Trp Ser Asp
        200                 205                 210

tgg agc caa gaa aaa atg gga atg act gag gaa gaa ggc aag cta ctc   848
Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu Glu Gly Lys Leu Leu
    215                 220                 225

cct gcg att ccc gtc ctg tct gct ctg gtg tagggctgct ttgggctaga     898
Pro Ala Ile Pro Val Leu Ser Ala Leu Val
230                 235

cttggtgggg tttgtcacca cctggttggg aatcatggaa tctcatgacc ccaggggccc 958
cctgtaccat cgagagtgag cctgcacaac tttgtgcccc aaaggcaaag gatcacattt 1018
taatactcat gaggttctta tactatacat gaaagggtat catatcattt gttttgtttt 1078
gttttgtttt tgagatggag tcttactctg tcacccagga tggagtgcag tgatgtgatc 1138
tcggctcact gccaccacca cctcccgagt tcaagcaatt cttgtgcctc agcctcccaa 1198
gtagctggga ttacaggggc ccacgaccat gcccggttga tttttgtatt tttagtagag 1258
aagggatatc accatgttgg ctaggctagt cttgaactcc tgacctcagg taatctgccc 1318
accttgacct cccaaagtgt tgggattaca ggcgtgagcc actgtgcccc gccagtatca 1378
tatcatctga aggtatcctg tgataaatta aagatacata ttgtgaatcc tggagctact 1438
actcaaaaaa taaataaagg tgtaactaat acaattta                         1476

<210> 115
<211> 239
<212> PRT
<213> Homo sapiens

<400> 115
Met Met Trp Thr Trp Ala Leu Trp Met Leu Pro Ser Leu Cys Lys Phe
 1               5                  10                  15
Ser Leu Ala Ala Leu Pro Ala Lys Pro Glu Asn Ile Ser Cys Val Tyr
            20                  25                  30
Tyr Tyr Arg Lys Asn Leu Thr Cys Thr Trp Ser Pro Gly Lys Glu Thr
        35                  40                  45
Ser Tyr Thr Gln Tyr Thr Val Lys Arg Thr Tyr Ala Phe Gly Glu Lys
```

```
    50                  55                  60
His Asp Asn Cys Thr Thr Asn Ser Ser Thr Ser Glu Asn Arg Ala Ser
65                  70                  75                  80
Cys Ser Phe Phe Leu Pro Arg Ile Thr Ile Pro Asp Asn Tyr Thr Ile
                85                  90                  95
Glu Val Glu Ala Glu Asn Gly Asp Gly Val Ile Lys Ser His Met Thr
            100                 105                 110
Tyr Trp Arg Leu Glu Asn Ile Ala Lys Thr Glu Pro Pro Lys Ile Phe
        115                 120                 125
Arg Val Lys Pro Val Leu Gly Ile Lys Arg Met Ile Gln Ile Glu Trp
    130                 135                 140
Ile Lys Pro Glu Leu Ala Pro Val Ser Ser Asp Leu Lys Tyr Thr Leu
145                 150                 155                 160
Arg Phe Arg Thr Val Asn Ser Thr Ser Trp Met Glu Val Asn Phe Ala
                165                 170                 175
Lys Asn Arg Lys Asp Lys Asn Gln Thr Tyr Asn Leu Thr Gly Leu Gln
            180                 185                 190
Pro Phe Thr Glu Tyr Val Ile Ala Leu Arg Cys Ala Val Lys Glu Ser
        195                 200                 205
Lys Phe Trp Ser Asp Trp Ser Gln Glu Lys Met Gly Met Thr Glu Glu
    210                 215                 220
Glu Gly Lys Leu Leu Pro Ala Ile Pro Val Leu Ser Ala Leu Val
225                 230                 235
```

<210> 116
<211> 2748
<212> DNA
<213> Mus musculus

<220>
<221> CDS
<222> (237)...(2222)

<400> 116

```
gatggggccc tgaatgttga tctgacagaa ttccagacca acctggtggt tattgtcctt 60
ttcatctggt catgctgaat atactctcaa gatgtgctgg agaaggtgct gctgtccggg 120
ctctcagaga aggcagtgct ggaggcgttc ctggcccggg tctcctccta ctgttcctgg 180
tagcccagcc ttctcggggt ggaaggagaa gctggccagg tgagctctga ggaagc atg 239
                                                              Met
                                                               1

ctg agc agc cag aag gga tcc tgc agc cag gaa cca ggg gca gcc cac  287
Leu Ser Ser Gln Lys Gly Ser Cys Ser Gln Glu Pro Gly Ala Ala His
             5                  10                  15

gtc cag cct ctg ggt gtg aac gct gga ata atg tgg acc ttg gca ctg  335
Val Gln Pro Leu Gly Val Asn Ala Gly Ile Met Trp Thr Leu Ala Leu
         20                  25                  30

tgg gca ttc tct ttc ctc tgc aaa ttc agc ctg gca gtc ctg ccg act  383
Trp Ala Phe Ser Phe Leu Cys Lys Phe Ser Leu Ala Val Leu Pro Thr
     35                  40                  45

aag cca gag aac att tcc tgc gtc ttt tac ttc gac aga aat ctg act  431
Lys Pro Glu Asn Ile Ser Cys Val Phe Tyr Phe Asp Arg Asn Leu Thr
 50                  55                  60                  65

tgc act tgg aga cca gag aag gaa acc aat gat acc agc tac att gtg  479
Cys Thr Trp Arg Pro Glu Lys Glu Thr Asn Asp Thr Ser Tyr Ile Val
                 70                  75                  80

act ttg act tac tcc tat gga aaa agc aat tat agt gac aat gct aca  527
```

```
Thr Leu Thr Tyr Ser Tyr Gly Lys Ser Asn Tyr Ser Asp Asn Ala Thr
                85                  90                  95

gag gct tca tat tct ttt ccc cgt tcc tgt gca atg ccc cca gac atc   575
Glu Ala Ser Tyr Ser Phe Pro Arg Ser Cys Ala Met Pro Pro Asp Ile
        100                 105                 110

tgc agt gtt gaa gta caa gct caa aat gga gat ggt aaa gtt aaa tct   623
Cys Ser Val Glu Val Gln Ala Gln Asn Gly Asp Gly Lys Val Lys Ser
    115                 120                 125

gac atc aca tat tgg cat tta atc tcc ata gca aaa acc gaa cca cct   671
Asp Ile Thr Tyr Trp His Leu Ile Ser Ile Ala Lys Thr Glu Pro Pro
130                 135                 140                 145

ata att tta agt gtg aat cca att tgt aat aga atg ttc cag ata caa   719
Ile Ile Leu Ser Val Asn Pro Ile Cys Asn Arg Met Phe Gln Ile Gln
                150                 155                 160

tgg aaa ccg cgt gaa aag act cgt ggg ttt cct tta gta tgc atg ctt   767
Trp Lys Pro Arg Glu Lys Thr Arg Gly Phe Pro Leu Val Cys Met Leu
            165                 170                 175

cgg ttc aga act gtc aac agt agc cgc tgg acg gaa gtc aat ttt gaa   815
Arg Phe Arg Thr Val Asn Ser Ser Arg Trp Thr Glu Val Asn Phe Glu
        180                 185                 190

aac tgt aaa cag gtc tgc aac ctc aca gga ctt cag gct ttc aca gaa   863
Asn Cys Lys Gln Val Cys Asn Leu Thr Gly Leu Gln Ala Phe Thr Glu
    195                 200                 205

tat gtc ctg gct cta cga ttc agg ttc aat gac tca aga tat tgg agc   911
Tyr Val Leu Ala Leu Arg Phe Arg Phe Asn Asp Ser Arg Tyr Trp Ser
210                 215                 220                 225

aag tgg agc aaa gaa gaa acc aga gtg act atg gag gaa gtt cca cat   959
Lys Trp Ser Lys Glu Glu Thr Arg Val Thr Met Glu Glu Val Pro His
                230                 235                 240

gtc ctg gac ctg tgg aga att ctg gaa cca gca gac atg aac gga gac   1007
Val Leu Asp Leu Trp Arg Ile Leu Glu Pro Ala Asp Met Asn Gly Asp
            245                 250                 255

agg aag gtg cga ttg ctg tgg aag aag gca aga gga gcc ccc gtc ttg   1055
Arg Lys Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val Leu
        260                 265                 270

gag aaa aca ttt ggc tac cac ata cag tac ttt gca gag aac agc act   1103
Glu Lys Thr Phe Gly Tyr His Ile Gln Tyr Phe Ala Glu Asn Ser Thr
    275                 280                 285

aac ctc aca gag ata aac aac atc acc acc cag cag tat gaa ctg ctt   1151
Asn Leu Thr Glu Ile Asn Asn Ile Thr Thr Gln Gln Tyr Glu Leu Leu
290                 295                 300                 305

ctg atg agc cag gca cac tct gtg tcc gtg act tct ttt aat tct ctt   1199
Leu Met Ser Gln Ala His Ser Val Ser Val Thr Ser Phe Asn Ser Leu
                310                 315                 320

ggc aag tcc caa gag acc atc ctg agg atc cca gat gtc cat gag aag   1247
Gly Lys Ser Gln Glu Thr Ile Leu Arg Ile Pro Asp Val His Glu Lys
            325                 330                 335
```

```
acc ttc cag tac att aag agc atg cag gcc tac ata gcc gag ccc ctg   1295
Thr Phe Gln Tyr Ile Lys Ser Met Gln Ala Tyr Ile Ala Glu Pro Leu
        340                 345                 350

ttg gtg gtg aac tgg caa agc tcc att cct gcg gtg gac act tgg ata   1343
Leu Val Val Asn Trp Gln Ser Ser Ile Pro Ala Val Asp Thr Trp Ile
    355                 360                 365

gtg gag tgg ctc cca gaa gct gcc atg tcg aag ttc cct gcc ctt tcc   1391
Val Glu Trp Leu Pro Glu Ala Ala Met Ser Lys Phe Pro Ala Leu Ser
370                 375                 380                 385

tgg gaa tct gtg tct cag gtc acg aac tgg acc atc gag caa gat aaa   1439
Trp Glu Ser Val Ser Gln Val Thr Asn Trp Thr Ile Glu Gln Asp Lys
                390                 395                 400

cta aaa cct ttc aca tgc tat aat ata tca gtg tat cca gtg ttg gga   1487
Leu Lys Pro Phe Thr Cys Tyr Asn Ile Ser Val Tyr Pro Val Leu Gly
            405                 410                 415

cac cga gtt gga gag ccg tat tca atc caa gct tat gcc aaa gaa gga   1535
His Arg Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu Gly
        420                 425                 430

act cca tta aaa ggt cct gag acc agg gtg gag aac atc ggt ctg agg   1583
Thr Pro Leu Lys Gly Pro Glu Thr Arg Val Glu Asn Ile Gly Leu Arg
    435                 440                 445

aca gcc acg atc aca tgg aag gag att cct aag agt gct agg aat gga   1631
Thr Ala Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Ala Arg Asn Gly
450                 455                 460                 465

ttt atc aac aat tac act gta ttt tac caa gct gaa ggt gga aaa gaa   1679
Phe Ile Asn Asn Tyr Thr Val Phe Tyr Gln Ala Glu Gly Gly Lys Glu
                470                 475                 480

ctc tcc aag act gtt aac tct cat gcc ctg cag tgt gac ctg gag tct   1727
Leu Ser Lys Thr Val Asn Ser His Ala Leu Gln Cys Asp Leu Glu Ser
            485                 490                 495

ctg aca cga agg acc tct tat act gtt tgg gtc atg gcc agc acc aga   1775
Leu Thr Arg Arg Thr Ser Tyr Thr Val Trp Val Met Ala Ser Thr Arg
        500                 505                 510

gct gga ggt acc aac ggg gtg aga ata aac ttc aag aca ttg tca atc   1823
Ala Gly Gly Thr Asn Gly Val Arg Ile Asn Phe Lys Thr Leu Ser Ile
    515                 520                 525

agt gtg ttt gaa att gtc ctt cta aca tct cta gtt gga gga ggc ctt   1871
Ser Val Phe Glu Ile Val Leu Leu Thr Ser Leu Val Gly Gly Gly Leu
530                 535                 540                 545

ctt cta ctt agc atc aaa aca gtg act ttt ggc ctc aga aag cca aac   1919
Leu Leu Leu Ser Ile Lys Thr Val Thr Phe Gly Leu Arg Lys Pro Asn
                550                 555                 560

cgg ttg act ccc ctg tgt tgt cct gat gtt ccc aac cct gct gaa agt   1967
Arg Leu Thr Pro Leu Cys Cys Pro Asp Val Pro Asn Pro Ala Glu Ser
            565                 570                 575

agt tta gcc aca tgg ctc gga gat ggt ttc aag aag tca aat atg aag   2015
Ser Leu Ala Thr Trp Leu Gly Asp Gly Phe Lys Lys Ser Asn Met Lys
        580                 585                 590
```

```
gag act gga aac tct ggg aac aca gaa gac gtg gtc cta aaa cca tgt   2063
Glu Thr Gly Asn Ser Gly Asn Thr Glu Asp Val Val Leu Lys Pro Cys
    595                 600                 605

ccc gtc ccc gcg gat ctc att gac aag ctg gta gtg aac ttt gag aat   2111
Pro Val Pro Ala Asp Leu Ile Asp Lys Leu Val Val Asn Phe Glu Asn
610                 615                 620                 625

ttt ctg gaa gta gtt ttg aca gag gaa gct gga aag ggt cag gcg agc   2159
Phe Leu Glu Val Val Leu Thr Glu Glu Ala Gly Lys Gly Gln Ala Ser
                630                 635                 640

att ttg gga gga gaa gcg aat gag tat atc tta tcc cag gaa cca agc   2207
Ile Leu Gly Gly Glu Ala Asn Glu Tyr Ile Leu Ser Gln Glu Pro Ser
            645                 650                 655

tgt cct ggc cat tgc tgaagctacc ctcagggtcc aggacagctg tcttgttggc   2262
Cys Pro Gly His Cys
        660

acttgactct ggcaggaacc tgatctctac ttttcttctc cctgtctccg gacactttct 2322
ctccttcatg cagagaccag gactagagcg gattcctcat ggtttgccag gctcctcagt 2382
ccttgctcgg gctcaggatc ttcaacaatg ccctttctgg gacactccat catccactta 2442
tatttatttt ttgcaacatt gtggattgaa cccagggact tgtttatgcg cgcaacttca 2502
gtaactgtgg cagagactta ggaatggaga tctgaccctt tgcagaaggt ttctggacat 2562
ccgtccctgt gtgagcctca gacagcattg tctttacttt gaatcagctt ccaagttaat 2622
aaaagaaaaa cagagaggtg gcataacagc tcctgcttcc tgacctgctt gagttccagt 2682
tctgacttcc tttggtgatg aacagcaatg tgggaagtgt aagctgaata aacccttcc 2742
tcccca                                                           2748
```

<210> 117
<211> 662
<212> PRT
<213> Mus musculus

<400> 117

```
Met Leu Ser Ser Gln Lys Gly Ser Cys Ser Gln Glu Pro Gly Ala Ala
 1               5                  10                  15
His Val Gln Pro Leu Gly Val Asn Ala Gly Ile Met Trp Thr Leu Ala
            20                  25                  30
Leu Trp Ala Phe Ser Phe Leu Cys Lys Phe Ser Leu Ala Val Leu Pro
        35                  40                  45
Thr Lys Pro Glu Asn Ile Ser Cys Val Phe Tyr Phe Asp Arg Asn Leu
    50                  55                  60
Thr Cys Thr Trp Arg Pro Glu Lys Glu Thr Asn Asp Thr Ser Tyr Ile
65                  70                  75                  80
Val Thr Leu Thr Tyr Ser Tyr Gly Lys Ser Asn Tyr Ser Asp Asn Ala
                85                  90                  95
Thr Glu Ala Ser Tyr Ser Phe Pro Arg Ser Cys Ala Met Pro Pro Asp
            100                 105                 110
Ile Cys Ser Val Glu Val Gln Ala Gln Asn Gly Asp Gly Lys Val Lys
        115                 120                 125
Ser Asp Ile Thr Tyr Trp His Leu Ile Ser Ile Ala Lys Thr Glu Pro
    130                 135                 140
Pro Ile Ile Leu Ser Val Asn Pro Ile Cys Asn Arg Met Phe Gln Ile
145                 150                 155                 160
Gln Trp Lys Pro Arg Glu Lys Thr Arg Gly Phe Pro Leu Val Cys Met
                165                 170                 175
Leu Arg Phe Arg Thr Val Asn Ser Ser Arg Trp Thr Glu Val Asn Phe
            180                 185                 190
Glu Asn Cys Lys Gln Val Cys Asn Leu Thr Gly Leu Gln Ala Phe Thr
        195                 200                 205
```

Glu Tyr Val Leu Ala Leu Arg Phe Arg Phe Asn Asp Ser Arg Tyr Trp
210 215 220
Ser Lys Trp Ser Lys Glu Glu Thr Arg Val Thr Met Glu Glu Val Pro
225 230 235 240
His Val Leu Asp Leu Trp Arg Ile Leu Glu Pro Ala Asp Met Asn Gly
245 250 255
Asp Arg Lys Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val
260 265 270
Leu Glu Lys Thr Phe Gly Tyr His Ile Gln Tyr Phe Ala Glu Asn Ser
275 280 285
Thr Asn Leu Thr Glu Ile Asn Asn Ile Thr Thr Gln Gln Tyr Glu Leu
290 295 300
Leu Leu Met Ser Gln Ala His Ser Val Ser Val Thr Ser Phe Asn Ser
305 310 315 320
Leu Gly Lys Ser Gln Glu Thr Ile Leu Arg Ile Pro Asp Val His Glu
325 330 335
Lys Thr Phe Gln Tyr Ile Lys Ser Met Gln Ala Tyr Ile Ala Glu Pro
340 345 350
Leu Leu Val Val Asn Trp Gln Ser Ser Ile Pro Ala Val Asp Thr Trp
355 360 365
Ile Val Glu Trp Leu Pro Glu Ala Ala Met Ser Lys Phe Pro Ala Leu
370 375 380
Ser Trp Glu Ser Val Ser Gln Val Thr Asn Trp Thr Ile Glu Gln Asp
385 390 395 400
Lys Leu Lys Pro Phe Thr Cys Tyr Asn Ile Ser Val Tyr Pro Val Leu
405 410 415
Gly His Arg Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu
420 425 430
Gly Thr Pro Leu Lys Gly Pro Glu Thr Arg Val Glu Asn Ile Gly Leu
435 440 445
Arg Thr Ala Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Ala Arg Asn
450 455 460
Gly Phe Ile Asn Asn Tyr Thr Val Phe Tyr Gln Ala Glu Gly Gly Lys
465 470 475 480
Glu Leu Ser Lys Thr Val Asn Ser His Ala Leu Gln Cys Asp Leu Glu
485 490 495
Ser Leu Thr Arg Arg Thr Ser Tyr Thr Val Trp Val Met Ala Ser Thr
500 505 510
Arg Ala Gly Gly Thr Asn Gly Val Arg Ile Asn Phe Lys Thr Leu Ser
515 520 525
Ile Ser Val Phe Glu Ile Val Leu Leu Thr Ser Leu Val Gly Gly Gly
530 535 540
Leu Leu Leu Leu Ser Ile Lys Thr Val Thr Phe Gly Leu Arg Lys Pro
545 550 555 560
Asn Arg Leu Thr Pro Leu Cys Cys Pro Asp Val Pro Asn Pro Ala Glu
565 570 575
Ser Ser Leu Ala Thr Trp Leu Gly Asp Gly Phe Lys Lys Ser Asn Met
580 585 590
Lys Glu Thr Gly Asn Ser Gly Asn Thr Glu Asp Val Val Leu Lys Pro
595 600 605
Cys Pro Val Pro Ala Asp Leu Ile Asp Lys Leu Val Val Asn Phe Glu
610 615 620
Asn Phe Leu Glu Val Val Leu Thr Glu Glu Ala Gly Lys Gly Gln Ala
625 630 635 640
Ser Ile Leu Gly Gly Glu Ala Asn Glu Tyr Ile Leu Ser Gln Glu Pro
645 650 655
Ser Cys Pro Gly His Cys
660

<210> 118
<211> 2728
<212> DNA

<213> Mus musculus

<220>
<221> CDS
<222> (237)...(1877)

<400> 118
gatggggccc tgaatgttga tctgacagaa ttccagacca acctggtggt tattgtcctt 60
ttcatctggt catgctgaat atactctcaa gatgtgctgg agaaggtgct gctgtccggg 120
ctctcagaga aggcagtgct ggaggcgttc ctggcccggg tctcctccta ctgttcctgg 180
tagcccagcc ttctcggggt ggaaggagaa gctggccagg tgagctctga ggaagc atg 239
Met
1

ctg agc agc cag aag gga tcc tgc agc cag gaa cca ggg gca gcc cac 287
Leu Ser Ser Gln Lys Gly Ser Cys Ser Gln Glu Pro Gly Ala Ala His
5 10 15

gtc cag cct ctg ggt gtg aac gct gga ata atg tgg acc ttg gca ctg 335
Val Gln Pro Leu Gly Val Asn Ala Gly Ile Met Trp Thr Leu Ala Leu
20 25 30

tgg gca ttc tct ttc ctc tgc aaa ttc agc ctg gca gtc ctg ccg act 383
Trp Ala Phe Ser Phe Leu Cys Lys Phe Ser Leu Ala Val Leu Pro Thr
35 40 45

aag cca gag aac att tcc tgc gtc ttt tac ttc gac aga aat ctg act 431
Lys Pro Glu Asn Ile Ser Cys Val Phe Tyr Phe Asp Arg Asn Leu Thr
50 55 60 65

tgc act tgg aga cca gag aag gaa acc aat gat acc agc tac att gtg 479
Cys Thr Trp Arg Pro Glu Lys Glu Thr Asn Asp Thr Ser Tyr Ile Val
70 75 80

act ttg act tac tcc tat gga aaa agc aat tat agt gac aat gct aca 527
Thr Leu Thr Tyr Ser Tyr Gly Lys Ser Asn Tyr Ser Asp Asn Ala Thr
85 90 95

gag gct tca tat tct ttt ccc cgt tcc tgt gca atg ccc cca gac atc 575
Glu Ala Ser Tyr Ser Phe Pro Arg Ser Cys Ala Met Pro Pro Asp Ile
100 105 110

tgc agt gtt gaa gta caa gct caa aat gga gat ggt aaa gtt aaa tct 623
Cys Ser Val Glu Val Gln Ala Gln Asn Gly Asp Gly Lys Val Lys Ser
115 120 125

gac atc aca tat tgg cat tta atc tcc ata gca aaa acc gaa cca cct 671
Asp Ile Thr Tyr Trp His Leu Ile Ser Ile Ala Lys Thr Glu Pro Pro
130 135 140 145

ata att tta agt gtg aat cca att tgt aat aga atg ttc cag ata caa 719
Ile Ile Leu Ser Val Asn Pro Ile Cys Asn Arg Met Phe Gln Ile Gln
150 155 160

tgg aaa ccg cgt gaa aag act cgt ggg ttt cct tta gta tgc atg ctt 767
Trp Lys Pro Arg Glu Lys Thr Arg Gly Phe Pro Leu Val Cys Met Leu
165 170 175

cgg ttc aga act gtc aac agt agc cgc tgg acg gaa gtc aat ttt gaa 815
Arg Phe Arg Thr Val Asn Ser Ser Arg Trp Thr Glu Val Asn Phe Glu
180 185 190

aac tgt aaa cag gtc tgc aac ctc aca gga ctt cag gct ttc aca gaa 863

```
Asn Cys Lys Gln Val Cys Asn Leu Thr Gly Leu Gln Ala Phe Thr Glu
    195                 200                 205

tat gtc ctg gct cta cga ttc agg ttc aat gac tca aga tat tgg agc   911
Tyr Val Leu Ala Leu Arg Phe Arg Phe Asn Asp Ser Arg Tyr Trp Ser
210                 215                 220                 225

aag tgg agc aaa gaa gaa acc aga gtg act atg gag gaa gtt cca cat   959
Lys Trp Ser Lys Glu Glu Thr Arg Val Thr Met Glu Glu Val Pro His
                230                 235                 240

gtc ctg gac ctg tgg aga att ctg gaa cca gca gac atg aac gga gac   1007
Val Leu Asp Leu Trp Arg Ile Leu Glu Pro Ala Asp Met Asn Gly Asp
            245                 250                 255

agg aag gtg cga ttg ctg tgg aag aag gca aga gga gcc ccc gtc ttg   1055
Arg Lys Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val Leu
        260                 265                 270

gag aaa aca ttt ggc tac cac ata cag tac ttt gca gag aac agc act   1103
Glu Lys Thr Phe Gly Tyr His Ile Gln Tyr Phe Ala Glu Asn Ser Thr
    275                 280                 285

aac ctc aca gag ata aac aac atc acc acc cag cag tat gaa ctg ctt   1151
Asn Leu Thr Glu Ile Asn Asn Ile Thr Thr Gln Gln Tyr Glu Leu Leu
290                 295                 300                 305

ctg atg agc cag gca cac tct gtg tcc gtg act tct ttt aat tct ctt   1199
Leu Met Ser Gln Ala His Ser Val Ser Val Thr Ser Phe Asn Ser Leu
                310                 315                 320

ggc aag tcc caa gag acc atc ctg agg atc cca gat gtc cat gag aag   1247
Gly Lys Ser Gln Glu Thr Ile Leu Arg Ile Pro Asp Val His Glu Lys
            325                 330                 335

acc ttc cag tac att aag agc atg cag gcc tac ata gcc gag ccc ctg   1295
Thr Phe Gln Tyr Ile Lys Ser Met Gln Ala Tyr Ile Ala Glu Pro Leu
        340                 345                 350

ttg gtg gtg aac tgg caa agc tcc att cct gcg gtg gac act tgg ata   1343
Leu Val Val Asn Trp Gln Ser Ser Ile Pro Ala Val Asp Thr Trp Ile
    355                 360                 365

gtg gag tgg ctc cca gaa gct gcc atg tcg aag ttc cct gcc ctt tcc   1391
Val Glu Trp Leu Pro Glu Ala Ala Met Ser Lys Phe Pro Ala Leu Ser
370                 375                 380                 385

tgg gaa tct gtg tct cag gtc acg aac tgg acc atc gag caa gat aaa   1439
Trp Glu Ser Val Ser Gln Val Thr Asn Trp Thr Ile Glu Gln Asp Lys
                390                 395                 400

cta aaa cct ttc aca tgc tat aat ata tca gtg tat cca gtg ttg gga   1487
Leu Lys Pro Phe Thr Cys Tyr Asn Ile Ser Val Tyr Pro Val Leu Gly
            405                 410                 415

cac cga gtt gga gag ccg tat tca atc caa gct tat gcc aaa gaa gga   1535
His Arg Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu Gly
        420                 425                 430

act cca tta aaa ggt cct gag acc agg gtg gag aac atc ggt ctg agg   1583
Thr Pro Leu Lys Gly Pro Glu Thr Arg Val Glu Asn Ile Gly Leu Arg
    435                 440                 445
```

```
aca gcc acg atc aca tgg aag gag att cct aag agt gct agg aat gga   1631
Thr Ala Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Ala Arg Asn Gly
450                 455                 460                 465

ttt atc aac aat tac act gta ttt tac caa gct gaa ggt gga aaa gaa   1679
Phe Ile Asn Asn Tyr Thr Val Phe Tyr Gln Ala Glu Gly Gly Lys Glu
                470                 475                 480

ctc tcc aag act gtt aac tct cat gcc ctg cag tgt gac ctg gag tct   1727
Leu Ser Lys Thr Val Asn Ser His Ala Leu Gln Cys Asp Leu Glu Ser
            485                 490                 495

ctg aca cga agg acc tct tat act gtt tgg gtc atg gcc agc acc aga   1775
Leu Thr Arg Arg Thr Ser Tyr Thr Val Trp Val Met Ala Ser Thr Arg
        500                 505                 510

gct gga ggt acc aac ggg gtg aga ata aac ttc aag aca ttg tca atc   1823
Ala Gly Gly Thr Asn Gly Val Arg Ile Asn Phe Lys Thr Leu Ser Ile
    515                 520                 525

agt gag tac tgg ctt cag gcc tca ttc tgg agt tta ctt cgg gtt gga   1871
Ser Glu Tyr Trp Leu Gln Ala Ser Phe Trp Ser Leu Leu Arg Val Gly
530                 535                 540                 545

aat gtt tgacaggagc aaggagagcc agcagagggc agcagagcat ggcttctcct   1927
Asn Val

gctctctctg gctcactcac ctcccaggag ttactgagga gctggcaaag ggagggctga 1987
gttagaccaa caggccattt tgatccttgc tggtaagcag ccacaaataa tcttaagatg 2047
aagcaagcaa catccacttc agcctcagcc acgtcaaagg ctgttgcctg agctcacact 2107
ggccagttcc taaatgtcag gagttgtgca atagaacctg ggaaggaaca actggttgat 2167
cagaggtcac tgacaaggga cttaatgtta ccatctgcgg tggggctttt gtttcgtttt 2227
gtttgtttgt tatgtgtatt caacttatca gcttttacgt tgaaaacatg aaaagcaaga 2287
caaatttgtt agatatcaca tataatgtga aatataatag tttaataatt gagtaggaaa 2347
gctgagggca tgtaatagac agagggaaaa gaagaggaaa gccagtctgg tctacaaagt 2407
gagttccagg acagccaggg ctacatggag aaaccctgtc tcaatcaatc aatcaatcaa 2467
tcaatcagtc aatcaatcaa aattcaagca gcattgacaa gttttgcaat aactactata 2527
aaccaaaaaa gtcatcttga tgtatctcag aagccccttg ttatttatgt tcctgaagac 2587
taaagtagac cgtggctctg agaaccatga gcaagataac acgttctgtc ctgcagccta 2647
acaatgcctt cttggtattc tttttgatac aacttctaaa ataacttttt tttaaaaaaa 2707
ataaaaatca tgttacagct a                                          2728
```

<210> 119
<211> 547
<212> PRT
<213> Mus musculus

<400> 119

```
Met Leu Ser Ser Gln Lys Gly Ser Cys Ser Gln Glu Pro Gly Ala Ala
 1               5                  10                  15
His Val Gln Pro Leu Gly Val Asn Ala Gly Ile Met Trp Thr Leu Ala
            20                  25                  30
Leu Trp Ala Phe Ser Phe Leu Cys Lys Phe Ser Leu Ala Val Leu Pro
        35                  40                  45
Thr Lys Pro Glu Asn Ile Ser Cys Val Phe Tyr Phe Asp Arg Asn Leu
    50                  55                  60
Thr Cys Thr Trp Arg Pro Glu Lys Glu Thr Asn Asp Thr Ser Tyr Ile
65                  70                  75                  80
Val Thr Leu Thr Tyr Ser Tyr Gly Lys Ser Asn Tyr Ser Asp Asn Ala
                85                  90                  95
Thr Glu Ala Ser Tyr Ser Phe Pro Arg Ser Cys Ala Met Pro Pro Asp
            100                 105                 110
```

Ile Cys Ser Val Glu Val Gln Ala Gln Asn Gly Asp Gly Lys Val Lys
115 120 125
Ser Asp Ile Thr Tyr Trp His Leu Ile Ser Ile Ala Lys Thr Glu Pro
130 135 140
Pro Ile Ile Leu Ser Val Asn Pro Ile Cys Asn Arg Met Phe Gln Ile
145 150 155 160
Gln Trp Lys Pro Arg Glu Lys Thr Arg Gly Phe Pro Leu Val Cys Met
165 170 175
Leu Arg Phe Arg Thr Val Asn Ser Ser Arg Trp Thr Glu Val Asn Phe
180 185 190
Glu Asn Cys Lys Gln Val Cys Asn Leu Thr Gly Leu Gln Ala Phe Thr
195 200 205
Glu Tyr Val Leu Ala Leu Arg Phe Arg Phe Asn Asp Ser Arg Tyr Trp
210 215 220
Ser Lys Trp Ser Lys Glu Glu Thr Arg Val Thr Met Glu Glu Val Pro
225 230 235 240
His Val Leu Asp Leu Trp Arg Ile Leu Glu Pro Ala Asp Met Asn Gly
245 250 255
Asp Arg Lys Val Arg Leu Leu Trp Lys Lys Ala Arg Gly Ala Pro Val
260 265 270
Leu Glu Lys Thr Phe Gly Tyr His Ile Gln Tyr Phe Ala Glu Asn Ser
275 280 285
Thr Asn Leu Thr Glu Ile Asn Asn Ile Thr Thr Gln Gln Tyr Glu Leu
290 295 300
Leu Leu Met Ser Gln Ala His Ser Val Ser Val Thr Ser Phe Asn Ser
305 310 315 320
Leu Gly Lys Ser Gln Glu Thr Ile Leu Arg Ile Pro Asp Val His Glu
325 330 335
Lys Thr Phe Gln Tyr Ile Lys Ser Met Gln Ala Tyr Ile Ala Glu Pro
340 345 350
Leu Leu Val Val Asn Trp Gln Ser Ser Ile Pro Ala Val Asp Thr Trp
355 360 365
Ile Val Glu Trp Leu Pro Glu Ala Ala Met Ser Lys Phe Pro Ala Leu
370 375 380
Ser Trp Glu Ser Val Ser Gln Val Thr Asn Trp Thr Ile Glu Gln Asp
385 390 395 400
Lys Leu Lys Pro Phe Thr Cys Tyr Asn Ile Ser Val Tyr Pro Val Leu
405 410 415
Gly His Arg Val Gly Glu Pro Tyr Ser Ile Gln Ala Tyr Ala Lys Glu
420 425 430
Gly Thr Pro Leu Lys Gly Pro Glu Thr Arg Val Glu Asn Ile Gly Leu
435 440 445
Arg Thr Ala Thr Ile Thr Trp Lys Glu Ile Pro Lys Ser Ala Arg Asn
450 455 460
Gly Phe Ile Asn Asn Tyr Thr Val Phe Tyr Gln Ala Glu Gly Gly Lys
465 470 475 480
Glu Leu Ser Lys Thr Val Asn Ser His Ala Leu Gln Cys Asp Leu Glu
485 490 495
Ser Leu Thr Arg Arg Thr Ser Tyr Thr Val Trp Val Met Ala Ser Thr
500 505 510
Arg Ala Gly Gly Thr Asn Gly Val Arg Ile Asn Phe Lys Thr Leu Ser
515 520 525
Ile Ser Glu Tyr Trp Leu Gln Ala Ser Phe Trp Ser Leu Leu Arg Val
530 535 540
Gly Asn Val
545

<210> 120
<211> 2196
<212> DNA
<213> Artificial Sequence

<220>
<223> Degenerate polynucleotide of SEQ ID NO:111

<221> misc_feature
<222> (1)...(2196)
<223> n = A, T, C or G

<400> 120
atgatgtgga cntgggcnyt ntggatgytn ccnwsnytnt gyaarttyws nytngcngcn 60
ytnccngcna arccngaraa yathwsntgy gtntaytayt aymgnaaraa yytnacntgy 120
acntggwsnc cnggnaarga racnwsntay acncartaya cngtnaarmg nacntaygcn 180
ttyggngara arcaygayaa ytgyacnacn aaywsnwsna cnwsngaraa ymgngcnwsn 240
tgywsnttyt tyytnccnmg nathacnath ccngayaayt ayacnathga rgtngargcn 300
garaayggng ayggngtnat haarwsncay atgacntayt ggmgnytnga raayathgcn 360
aaracngarc cnccnaarat httymgngtn aarccngtny tnggnathaa rmgnatgath 420
carathgart ggathaarcc ngarytngcn ccngtnwsnw sngayytnaa rtayacnytn 480
mgnttymgna cngtnaayws nacnwsntgg atggargtna ayttygcnaa raaymgnaar 540
gayaaraayc aracntayaa yytnacnggn ytncarccnt tyacngarta ygtnathgcn 600
ytnmgntgyg cngtnaarga rwsnaartty tggwsngayt ggwsncarga raaratgggn 660
atgacngarg argargcncc ntgyggnytn garytntggm gngtnytnaa rccngcngar 720
gcngayggnm gnmgnccngt nmgnytnytn tggaaraarg cnmgnggngc nccngtnytn 780
garaaracny tnggntayaa yathtggtay tayccngarw snaayacnaa yytnacngar 840
acnatgaaya cnacnaayca rcarytngar ytncayytng gnggngarws nttytgggtn 900
wsnatgathw sntayaayws nytnggnaar wsnccngtng cnacnytnmg nathccngcn 960
athcargara arwsnttyca rtgyathgar gtnatgcarg cntgygtngc ngargaycar 1020
ytngtngtna artggcarws nwsngcnytn gaygtnaaya cntggatgat hgartggtty 1080
ccngaygtng aywsngarcc nacnacnytn wsntgggarw sngtnwsnca rgcnacnaay 1140
tggacnathc arcargayaa rytnaarccn ttytggtgyt ayaayathws ngtntayccn 1200
atgytncayg ayaargtngg ngarccntay wsnathcarg cntaygcnaa rgarggngtn 1260
ccnwsngarg gnccngarac naargtngar aayathggng tnaaracngt nacnathacn 1320
tggaargara thccnaarws ngarmgnaar ggnathatht gyaaytayac naththytay 1380
cargcngarg gnggnaargg nttywsnaar acngtnaayw snwsnathyt ncartayggn 1440
ytngarwsny tnaarmgnaa racnwsntay athgtncarg tnatggcnws nacnwsngcn 1500
ggnggnacna ayggnacnws nathaaytty aaracnytnw snttywsngt nttygarath 1560
athytnatha cnwsnytnat hggnggnggn ytnytnathy tnathathyt nacngtngcn 1620
tayggnytna araarccnaa yaarytnacn cayytntgyt ggccnacngt nccnaayccn 1680
gcngarwsnw snathgcnac ntggcayggn gaygayttya argayaaryt naayytnaar 1740
garwsngayg aywsngtnaa yacngargay mgnathytna arccntgyws nacnccnwsn 1800
gayaarytng tnathgayaa rytngtngtn aayttyggna aygtnytnca rgarathtty 1860
acngaygarg cnmgnacngg ncargaraay aayytnggng gngaraaraa yggntaygtn 1920
acntgyccnt tymgnccnga ytgyccnytn ggnaarwsnt tygargaryt nccngtnwsn 1980
ccngarathc cnccnmgnaa rwsncartay ytnmgnwsnm gnatgccnga rggnacnmgn 2040
ccngargcna argarcaryt nytnttywsn ggncarwsny tngtnccnga ycayytntgy 2100
gargarggng cnccnaaycc ntayytnaar aaywsngtna cngcnmgnga rttyytngtn 2160
wsngaraary tnccngarca yacnaarggn gargtn 2196

<210> 121
<211> 1947
<212> DNA
<213> Artificial Sequence

<220>
<223> Degenerate polynucleotide of SEQ ID NO:109

<221> misc_feature
<222> (1)...(1947)
<223> n = A, G, C or G

<400> 121
atgatgtgga cntgggcnyt ntggatgytn ccnwsnytnt gyaarttyws nytngcngcn 60
ytnccngcna arccngaraa yathwsntgy gtntaytayt aymgnaaraa yytnacntgy 120
acntggwsnc cnggnaarga racnwsntay acncartaya cngtnaarmg nacntaygcn 180
ttyggngara arcaygayaa ytgyacnacn aaywsnwsna cnwsngaraa ymgngcnwsn 240

tgywsnttyt tyytnccnmg nathacnath ccngayaayt ayacnathga rgtngargcn 300
garaayggng ayggngtnat haarwsncay atgacntayt ggmgnytnga raayathgcn 360
aaracngarc cnccnaarat httymgngtn aarccngtny tnggnathaa rmgnatgath 420
carathgart ggathaarcc ngarytngcn ccngtnwsnw sngayytnaa rtayacnytn 480
mgnttymgna cngtnaayws nacnwsntgg atggargtna ayttygcnaa raaymgnaar 540
gayaaraayc aracntayaa yytnacnggn ytncarccnt tyacngarta ygtnathgcn 600
ytnmgntgyg cngtnaarga rwsnaartty tggwsngayt ggwsncarga raaratgggn 660
atgacngarg argargcncc ntgyggnytn garytntggm gngtnytnaa rccngcngar 720
gcngayggnm gnmgnccngt nmgnytnytn tggaaraarg cnmgnggngc nccngtnytn 780
garaaracny tnggntayaa yathtggtay tayccngarw snaayacnaa yytnacngar 840
acnatgaaya cnacnaayca rcarytngar ytncayytng gnggngarws nttytgggtn 900
wsnatgathw sntayaayws nytnggnaar wsnccngtng cnacnytnmg nathccngcn 960
athcargara arwsnttyca rtgyathgar gtnatgcarg cntgygtngc ngargaycar 1020
ytngtngtna artggcarws nwsngcnytn gaygtnaaya cntggatgat hgartggtty 1080
ccngaygtng aywsngarcc nacnacnytn wsntgggarw sngtnwsnca rgcnacnaay 1140
tggacnathc arcargayaa rytnaarccn ttytggtgyt ayaayathws ngtntayccn 1200
atgytncayg ayaargtngg ngarccntay wsnathcarg cntaygcnaa rgarggngtn 1260
ccnwsngarg gnccngarac naargtngar aayathggng tnaaracngt nacnathacn 1320
tggaargara thccnaarws ngarmgnaar ggnathatht gyaaytayac naththytay 1380
cargcngarg gnggnaargg nttywsnaar acngtnaayw snwsnathyt ncartayggn 1440
ytngarwsny tnaarmgnaa racnwsntay athgtncarg tnatggcnws nacnwsngcn 1500
ggnggnacna ayggnacnws nathaaytty aaracnytnw snttywsngt nttygarath 1560
athytnatha cnwsnytnat hggnggnggn ytnytnathy tnathathyt nacngtngcn 1620
tayggnytna araarccnaa yaarytnacn cayytntgyt ggccnacngt nccnaayccn 1680
gcngarwsnw snathgcnac ntggcayggn gaygayttya argayaaryt naayytnaar 1740
garwsngayg aywsngtnaa yacngargay mgnathytna arccntgyws nacnccnwsn 1800
gayaarytng tnathgayaa rytngtngtn aayttyggna aygtnytnca rgarathtty 1860
acngaygarg cnmgnacngg ncargaraay aayytnggng gngaraaraa yggnacnmgn 1920
athytnwsnw sntgyccnac nwsnath 1947

<210> 122
<211> 1986
<212> DNA
<213> Artificial Sequence

<220>
<223> Degenerate polynucleotide of SEQ ID NO:5

<221> misc_feature
<222> (1)...(1986)
<223> n = A, T, C or G

<400> 122
atgaarytnw snccncarcc nwsntgygtn aayytnggna tgatgtggac ntgggcnytn 60
tggatgytnc cnwsnytntg yaarttywsn ytngcngcny tnccngcnaa rccngaraay 120
athwsntgyg tntaytayta ymgnaaraay ytnacntgya cntggwsncc nggnaargar 180
acnwsntaya cncartayac ngtnaarmgn acntaygcnt tyggngaraa rcaygayaay 240
tgyacnacna aywsnwsnac nwsngaraay mgngcnwsnt gywsnttytt yytnccnmgn 300
athacnathc cngayaayta yacnathgar gtngargcng araayggnga yggngtnath 360
aarwsncaya tgacntaytg gmgnytngar aayathgcna aracngarcc nccnaarath 420
ttymgngtna arccngtnyt nggnathaar mgnatgathc arathgartg gathaarccn 480
garytngcnc cngtnwsnws ngayytnaar tayacnytnm gnttymgnac ngtnaaywsn 540
acnwsntgga tggargtnaa yttygcnaar aaymgnaarg ayaaraayca racntayaay 600
ytnacnggny tncarccntt yacngartay gtnathgcny tnmgntgygc ngtnaargar 660
wsnaarttyt ggwsngaytg gwsncargar aaratgggna tgacngarga rgargcnccn 720
tgyggnytng arytntggmg ngtnytnaar ccngcngarg cngayggnmg nmgnccngtn 780
mgnytnytnt ggaaraargc nmgnggngcn ccngtnytng araaracnyt nggntayaay 840
athtggtayt ayccngarws naayacnaay ytnacngara cnatgaayac nacnaaycar 900
carytngary tncayytngg nggngarwsn ttytgggtnw snatgathws ntayaaywsn 960
ytnggnaarw snccngtngc nacnytnmgn athccngcna thcargaraa rwsnttycar 1020
tgyathgarg tnatgcargc ntgygtngcn gargaycary tngtngtnaa rtggcarwsn 1080
wsngcnytng aygtnaayac ntggatgath gartggttyc cngaygtnga ywsngarccn 1140
acnacnytnw sntgggarws ngtnwsncar gcnacnaayt ggacnathca rcargayaar 1200

DEMANDE OU BREVET VOLUMINEUX

LA PRÉSENTE PARTIE DE CETTE DEMANDE OU CE BREVET COMPREND PLUS D'UN TOME.

CECI EST LE TOME 1 DE 2
CONTENANT LES PAGES 1 À 314

NOTE : Pour les tomes additionels, veuillez contacter le Bureau canadien des brevets

JUMBO APPLICATIONS/PATENTS

THIS SECTION OF THE APPLICATION/PATENT CONTAINS MORE THAN ONE VOLUME

THIS IS VOLUME 1 OF 2
CONTAINING PAGES 1 TO 314

NOTE: For additional volumes, please contact the Canadian Patent Office

NOM DU FICHIER / FILE NAME :

NOTE POUR LE TOME / VOLUME NOTE:

CLAIMS

What is claimed is:

1. An isolated multimeric or heterodimeric cytokine receptor comprising:

at least one polypeptide having at least 90 percent sequence identity with SEQ ID NO:111 or SEQ ID NO:109; and

wherein the multimeric or heterodimeric cytokine receptor binds a ligand comprising SEQ ID NO:2.

2. The isolated multimeric or heterodimeric cytokine receptor of claim 1 wherein the multimeric or heterodimeric cytokine receptor further comprises a cytokine-binding domain of a class I cytokine receptor.

3. The isolated multimeric or heterodimeric cytokine receptor of claim 2 wherein the cytokine-binding domain of the class I cytokine receptor comprises amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 979 of SEQ ID NO:7, or amino acid residue 1 to amino acid residue 979 of SEQ ID NO:7.

4. The isolated multimeric or heterodimeric cytokine receptor of claim 1 wherein the multimeric or heterodimeric cytokine receptor antagonizes an activity of SEQ ID NO:2.

5. The isolated multimeric or heterodimeric cytokine receptor of claim 1 wherein the multimeric or heterodimeric cytokine receptor inhibits proliferation of hematopoietic cells, inhibits proliferation of immune cells, inhibits proliferation of inflammatory cells, inhibits an immune response, inhibits an inflammatory response, or inhibits proliferation of tumor cells of epithelial origin.

6. The isolated multimeric or heterodimeric cytokine receptor of claim 1 wherein the multimeric or heterodimeric cytokine receptor is soluble.

7. The isolated multimeric or heterodimeric cytokine receptor of claim 1 further

comprising an affinity tag or cytotoxic molecule.

8. The isolated multimeric or heterodimeric cytokine receptor of claim 7 wherein the affinity tag is polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™ peptide, streptavidin binding peptide, or immunoglobulin F_c polypeptide.

9. The isolated multimeric or heterodimeric cytokine receptor of claim 7 wherein the cytotoxic molecule is a toxin or radionuclide.

10. The isolated multimeric or heterodimeric cytokine receptor of claim 1 wherein the polypeptide having at least 90 percent identity with SEQ ID NO:111 comprises amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 227, amino acid residue 1 to amino acid residue 519, amino acid residue 1 to amino acid residue 543, or amino acid residue 1 to amino acid residue 732.

11. The isolated multimeric or heterodimeric cytokine receptor of claim 1 wherein the polypeptide having at least 90 percent identity with SEQ ID NO:109 comprises amino acid residue 1 to amino acid residue 649 of SEQ ID NO:109, or amino acid residue 20 to amino acid residue 649 of SEQ ID NO:109.

12. An isolated multimeric or heterodimeric cytokine receptor comprising at least one polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111.

13. The isolated multimeric or heterodimeric cytokine receptor of claim 12 wherein the polypeptide comprises amino acid residue 1 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 732 of SEQ ID NO:111, or amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111.

14. The isolated multimeric or heterodimeric cytokine receptor of claim 12 further comprising a cytokine-binding domain of a class I cytokine receptor.

15. The isolated multimeric or heterodimeric cytokine receptor of claim 14 wherein the cytokine-binding domain of the class I cytokine receptor comprises amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 979 of SEQ ID NO:7, or amino acid residue 1 to amino acid residue 979 of SEQ ID NO:7.

16. The isolated multimeric or heterodimeric cytokine receptor of claim 12 wherein the multimeric or heterodimeric cytokine receptor antagonizes an activity of a ligand comprising SEQ ID NO:2.

17. The isolated multimeric or heterodimeric cytokine receptor of claim 12 wherein the multimeric or heterodimeric cytokine receptor inhibits proliferation of hematopoietic cells, inhibits proliferation of immune cells, inhibits proliferation of inflammatory cells, inhibits an immune response, inhibits an inflammatory response, or inhibits proliferation of tumor cells of epithelial origin.

18. The isolated multimeric or heterodimeric cytokine receptor of claim 12 wherein the multimeric or heterodimeric cytokine receptor is soluble.

19. The isolated multimeric or heterodimeric cytokine receptor of claim 12 further comprising an affinity tag or cytotoxic molecule.

20. The isolated multimeric or heterodimeric cytokine receptor of claim 19 wherein the affinity tag is polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™ peptide, streptavidin binding peptide, or immunoglobulin F_c polypeptide.

21. The isolated multimeric or heterodimeric cytokine receptor of claim 19 wherein the cytotoxic molecule is a toxin or radionuclide.

22. An isolated polynucleotide that encodes a cytokine receptor polypeptide comprising an amino acid sequence having at least 90 percent sequence identity with SEQ ID NO:111 or SEQ ID NO:109, wherein the cytokine receptor polypeptide forms a

multimeric or heterodimeric cytokine receptor, and wherein the multimeric or heterodimeric cytokine receptor binds a ligand comprising SEQ ID NO:2.

23. The isolated polynucleotide of claim 22 wherein the multimeric or heterodimeric cytokine receptor further comprises a cytokine-binding domain of a class I cytokine receptor.

24. The isolated polynucleotide of claim of 23 wherein the cytokine-binding domain of the class I cytokine receptor comprises amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 979 of SEQ ID NO:7, or amino acid residue 1 to amino acid residue 979 of SEQ ID NO:7.

25. The isolated polynucleotide of claim of 22 wherein the multimeric or heterodimeric cytokine receptor antagonizes an activity of SEQ ID NO:2.

26. The isolated polynucleotide of claim of 22 wherein the multimeric or heterodimeric cytokine receptor inhibits proliferation of hematopoietic cells, inhibits proliferation of immune cells, inhibits proliferation of inflammatory cells, inhibits an immune response, inhibits an inflammatory response, or inhibits proliferation of tumor cells of epithelial origin.

27. The isolated polynucleotide of claim of 22 wherein the multimeric or heterodimeric cytokine receptor is soluble.

28. The isolated polynucleotide of claim of 22 further comprising an affinity tag or cytotoxic molecule.

29. The isolated polynucleotide of claim of 28 wherein the affinity tag is polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™ peptide, streptavidin binding peptide, or immunoglobulin F_c polypeptide.

30. The isolated polynucleotide of claim 28 wherein the cytotoxic molecule is a toxin or radionuclide.

31. The isolated polynucleotide of claim 22 wherein the cytokine receptor polypeptide having at least 90 percent identity with SEQ ID NO:111 comprises amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 543 of SEQ ID NO:111, or amino acid residue 1 to amino acid residue 732 of SEQ ID NO:111.

32. The isolated polynucleotide of claim 22 wherein the cytokine receptor polypeptide having at least 90 percent identity with SEQ ID NO:109 comprises amino acid residue 1 to amino acid residue 649 of SEQ ID NO:109, or amino acid residue 20 to amino acid residue 649 of SEQ ID NO:109.

33. An isolated polynucleotide that encodes a cytokine receptor polypeptide comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111, wherein the cytokine receptor polypeptide forms a multimeric or heterodimeric cytokine receptor.

34. The isolated polynucleotide of claim 33 wherein the cytokine receptor polypeptide comprises amino acid residue 1 to amino acid residue 227 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 519 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 20 to amino acid residue 543 of SEQ ID NO:111, amino acid residue 1 to amino acid residue 732 of SEQ ID NO:111, or amino acid residue 20 to amino acid residue 732 of SEQ ID NO:111.

35. The isolated polynucleotide of claim 33 wherein the multimeric or heterodimeric cytokine receptor further comprises a cytokine-binding domain of a class I cytokine receptor.

36. The isolated polynucleotide of claim 35 wherein the cytokine-binding domain of the class I cytokine receptor comprises amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 429 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 739 of SEQ ID NO:7, amino acid residue 1

to amino acid residue 739 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 1 to amino acid residue 761 of SEQ ID NO:7, amino acid residue 28 to amino acid residue 979 of SEQ ID NO:7, or amino acid residue 1 to amino acid residue 979 of SEQ ID NO:7.

37. The isolated polynucleotide of claim 33 wherein the multimeric or heterodimeric cytokine receptor antagonizes an activity of a ligand comprising SEQ ID NO:2.

38. The isolated polynucleotide of claim 33 wherein the multimeric or heterodimeric cytokine receptor inhibits proliferation of hematopoietic cells, inhibits proliferation of immune cells, inhibits proliferation of inflammatory cells, inhibits an immune response, inhibits an inflammatory response, or inhibits proliferation of tumor cells of epithelial origin.

39. The isolated polynucleotide of claim 33 wherein the multimeric or heterodimeric cytokine receptor is soluble.

40. The isolated polynucleotide of claim 33 wherein the multimeric or heterodimeric cytokine receptor further comprises an affinity tag or cytotoxic molecule.

41. The isolated polynucleotide of claim 40 wherein the affinity tag is polyhistidine, protein A, glutathione S transferase, Glu-Glu, substance P, Flag™ peptide, streptavidin binding peptide, or immunoglobulin F_c polypeptide.

42. The isolated polynucleotide of claim 40 wherein the cytotoxic molecule is a toxin or radionuclide.

43. An expression vector comprising the following operably linked elements:

a transcription promoter;

a DNA segment encoding a cytokine receptor polypeptide having at least 90 percent sequence identity with SEQ ID NO:111; and

a transcription terminator;

wherein the cytokine receptor polypeptide forms a multimeric or heterodimeric cytokine receptor, and wherein the multimeric or heterodimeric cytokine receptor binds a ligand comprising SEQ ID NO:2.

44. An expression vector comprising the following operably linked elements:

a) a first transcription promoter; a first DNA segment encoding a cytokine receptor polypeptide having at least 90 percent sequence identity with SEQ ID NO:111; and a first transcription terminator; and

b) a second transcription promoter; a second DNA segment encoding a cytokine-binding domain of a class I cytokine receptor; and a second transcription terminator;

wherein the cytokine receptor polypeptide and the class I cytokine receptor form a multimeric or heterodimeric cytokine receptor; and

wherein the multimeric or heterodimeric cytokine receptor binds to a ligand comprising SEQ ID NO:2.

45. A cultured cell comprising an expression vector of claim 43, wherein the cell expresses the cytokine receptor polypeptide encoded by the DNA segment.

46. A cultured cell comprising an expression vector of claim 44, wherein the cell expresses the cytokine receptor polypeptide and the class I cytokine receptor encoded by the DNA segments.

47. A cultured cell comprising:

a first expression vector comprising:

a) a transcription promoter;

b) a DNA segment encoding a cytokine receptor polypeptide having at least 90 percent sequence identity with SEQ ID NO:111; and

c) a transcription terminator; and

a second expression vector comprising:

a) a transcription promoter;

b) a DNA segment encoding a cytokine-binding domain of a class I cytokine receptor; and

c) a transcription terminator;

wherein the cytokine receptor polypeptide and the class I cytokine receptor form a multimeric or heterodimeric cytokine receptor; and

wherein the multimeric or heterodimeric cytokine receptor binds to a ligand comprising SEQ ID NO:2.

48. A method of producing an antibody to a multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor, the method comprising:

inoculating an animal with the multimeric or heterodimeric cytokine receptor, wherein the multimeric or heterodimeric cytokine receptor elicits an immune response in the animal to produce an antibody that specifically binds the multimeric or heterodimeric cytokine receptor; and

isolating the antibody from the animal.

49. An antibody that specifically binds to the multimeric or heterodimeric cytokine receptor of claim 1.

50. An antibody that specifically binds to the multimeric or heterodimeric cytokine receptor of claim 12.

51. A composition comprising:

an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and

a pharmaceutically acceptable vehicle.

52. A method of producing a multimeric or heterodimeric cytokine receptor comprising:

culturing a cell according to claim 45; and

isolating the multimeric or heterodimeric cytokine receptor produced by the cell.

53. A method of producing a multimeric or heterodimeric cytokine receptor comprising:

culturing a cell according to claim 46; and

isolating the multimeric or heterodimeric cytokine receptor produced by the cell.

54. A method of producing a multimeric or heterodimeric cytokine receptor comprising:

culturing a cell according to claim 47; and

isolating the multimeric or heterodimeric cytokine receptor produced by the cell.

55. An immune cell inhibiting composition comprising:

an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and

a pharmaceutically acceptable vehicle;

wherein the soluble multimeric or heterodimeric cytokine receptor inhibits proliferation of immune cells.

56. An immune response inhibiting composition comprising:

an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and

a pharmaceutically acceptable vehicle;

wherein the soluble multimeric or heterodimeric cytokine receptor inhibits an immune response.

57. An inflammatory cell inhibiting composition comprising:

an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and

a pharmaceutically acceptable vehicle;

wherein the soluble multimeric or heterodimeric cytokine receptor

inhibits proliferation of inflammatory cells.

58. An inflammatory response inhibiting composition comprising:

an effective amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor; and

a pharmaceutically acceptable vehicle;

wherein the soluble multimeric or heterodimeric cytokine receptor inhibits an inflammatory response.

59. A method of inhibiting an immune response in a mammal exposed to an antigen or pathogen, the method comprising:

(a) determining directly or indirectly the level of antigen or pathogen present in the mammal;

(b) administering a composition comprising a soluble multimeric or heterodimeric cytokine receptor in a pharmaceutically acceptable vehicle;

(c) determining directly or indirectly the level of antigen or pathogen in the mammal; and

(d) comparing the level of the antigen or pathogen in step (a) to the antigen or pathogen level in step (c), wherein a change in the level is indicative of inhibiting an immune response.

60. The method of claim 59 further comprising:

(e) re-administering a composition comprising a multimeric or heterodimeric cytokine receptor in a pharmaceutically acceptable vehicle;

(f) determining directly or indirectly the level of antigen or pathogen in the mammal; and

(g) comparing the number of the antigen or pathogen level in step (a) to the antigen level in step (f), wherein a change in the level is indicative of inhibiting an immune response.

61. A method for reducing hematopoietic cells and hematopoietic progenitor cells in a mammal, the method comprising:

culturing bone marrow or peripheral blood cells with a composition comprising an effective amount of a soluble multimeric or heterodimeric cytokine receptor to produce a decrease in the number of lymphoid cells in the bone marrow or peripheral blood cells as compared to bone marrow or peripheral blood cells cultured in the absence of the multimeric or heterodimeric cytokine receptor.

62. The method of claim 61 wherein the hematopoietic cells and hematopoietic progenitor cells are lymphoid cells.

63. The method of claim 62 wherein the lymphoid cells are monocytic cells, macrophages or T cells.

64. A method of detecting the presence of a multimeric or heterodimeric cytokine receptor in a biological sample, comprising the steps of:

(a) contacting the biological sample with an antibody, or an antibody fragment, of claim 50, wherein the contacting is performed under conditions that allow the binding of the antibody or antibody fragment to the biological sample; and

(b) detecting any of the bound antibody or bound antibody fragment.

65. A method of killing cancer cells comprising,

obtaining *ex vivo* a tissue or biological sample containing cancer cells from a patient, or identifying cancer cells *in vivo*;

producing a multimeric or heterodimeric cytokine receptor by the method of claim 52;

formulating the multimeric or heterodimeric cytokine receptor in a pharmaceutically acceptable vehicle; and

administering to the patient or exposing the cancer cells to the multimeric or heterodimeric cytokine receptor formulation;

wherein the multimeric or heterodimeric cytokine receptor kills the cells.

66. A method of killing cancer cells of claim 65, wherein the multimeric or heterodimeric cytokine receptor is further conjugated to a toxin.

67. The antibody of claim 50, wherein the antibody is from the group of: (a) polyclonal antibody, (b) murine monoclonal antibody, (c) humanized antibody derived from (b), (d) an antibody fragment, and (e) human monoclonal antibody.

68. An antibody or antibody fragment that specifically binds to a multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor.

69. The antibody of claim 50, wherein the antibody further comprises a radionuclide, enzyme, substrate, cofactor, fluorescent marker, chemiluminescent marker, peptide tag, magnetic particle, drug, or toxin.

70. A method for inhibiting zcytor17lig-induced proliferation or differentiation of hematopoietic cells and hematopoietic progenitor cells comprising culturing bone marrow or peripheral blood cells with a composition comprising an amount of a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor sufficient to reduce proliferation or differentiation of the hematopoietic cells in the bone marrow or peripheral blood cells as compared to bone marrow or peripheral blood cells cultured in the absence of the soluble multimeric or heterodimeric cytokine receptor.

71. The method of claim 70, wherein the hematopoietic cells and hematopoietic progenitor cells are lymphoid cells.

72. The method of claim 71, wherein the lymphoid cells are macrophages or T cells.

73. A method of reducing zcytor17lig-induced induced inflammation comprising administering to a mammal with inflammation an amount of a composition comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor sufficient to reduce inflammation.

74. A method of suppressing an inflammatory response in a mammal with inflammation comprising:

(1) determining a level of an inflammatory molecule;

(2) administering a composition comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor in a pharmaceutically acceptable vehicle;

(3) determining a post administration level of the inflammatory molecule;

(4) comparing the level of the inflammatory molecule in step (1) to the level of the inflammatory molecule in step (3), wherein a lack of increase or a decrease the inflammatory molecule level is indicative of suppressing an inflammatory response.

75. The antibody of claim 68, wherein the antibody further comprises a radionuclide, enzyme, substrate, cofactor, fluorescent marker, chemiluminescent marker, peptide tag, magnetic particle, drug, or toxin.

76. A method for inhibiting zcytor17lig-induced proliferation or differentiation of hematopoietic cells and hematopoietic progenitor cells comprising culturing bone marrow or peripheral blood cells with a composition comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor in a pharmaceutically acceptable vehicle sufficient to reduce proliferation or differentiation of the hematopoietic cells in the bone marrow or peripheral blood cells as compared to bone marrow or peripheral blood cells cultured in the absence of soluble multimeric or heterodimeric cytokine receptor.

77. The method of claim 76, wherein the hematopoietic cells and hematopoietic progenitor cells are lymphoid cells.

78. The method of claim 77, wherein the lymphoid cells are macrophages or T cells.

79. A method of reducing zcytor17lig-induced induced inflammation comprising administering to a mammal with inflammation an amount of a composition comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor in a pharmaceutically acceptable vehicle sufficient to reduce inflammation.

80. A method of suppressing an inflammatory response in a mammal with inflammation comprising:

(1) determining a level of an inflammatory molecule;

(2) administering a composition comprising a multimeric or heterodimeric cytokine receptor which comprises amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 in a pharmaceutically acceptable vehicle;

(3) determining a post administration level of the inflammatory molecule;

(4) comparing the level of the inflammatory molecule in step (1) to the level of the inflammatory molecule in step (3), wherein a lack of increase or a decrease in the inflammatory molecule level is indicative of suppressing an inflammatory response.

81. A method of treating a mammal afflicted with an inflammatory disease in which zcytor17lig plays a role, comprising:

administering an antagonist of zcytor17lig to the mammal such that the inflammation is reduced, wherein the antagonist is a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine receptor in a pharmaceutically acceptable vehicle.

82. A method of claim 81, wherein the disease is a chronic inflammatory disease.

83. A method of claim 82, wherein the disease is a chronic inflammatory disease from the group of:

(a) inflammatory bowel disease;

(b) ulcerative colitis;

(c) Crohn's disease;

(d) atopic dermatitis;

(e) eczema; and

(f) psoriasis.

84. A method of claim 81, wherein the disease is an acute inflammatory disease.

85. A method of claim 84, wherein the disease is an acute inflammatory disease from the group of:

(a) endotoxemia;

(b) septicemia;

(c) toxic shock syndrome; and

(d) infectious disease.

86. A method of claim 81, wherein the soluble multimeric or heterodimeric cytokine receptor further comprises a radionuclide, enzyme, substrate, cofactor, fluorescent marker, chemiluminescent marker, peptide tag, magnetic particle, drug, or toxin.

87. A method for detecting inflammation in a patient, comprising:

obtaining a tissue or biological sample from a patient;

incubating the tissue or biological sample with a soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and a cytokine-binding domain of a class I cytokine

receptor under conditions wherein the soluble multimeric or heterodimeric cytokine receptor binds to its complementary polypeptide in the tissue or biological sample;

visualizing the soluble multimeric or heterodimeric cytokine receptor bound in the tissue or biological sample; and

comparing levels of soluble multimeric or heterodimeric cytokine receptor bound in the tissue or biological sample from the patient to a normal control tissue or biological sample,

wherein an increase in the level of soluble multimeric or heterodimeric cytokine receptor bound to the patient tissue or biological sample relative to the normal control tissue or biological sample is indicative of inflammation in the patient.

88. A soluble multimeric or heterodimeric cytokine receptor comprising amino acid residue 20 to amino acid residue 227 of SEQ ID NO:111 and amino acid residue 28 to amino acid residue 429 of SEQ ID NO:7.

```
                    1               15 16           30 31           45
1 zcytor17lig       ---ASHTLPVRLLRP SDDVQKIVEELQSLS KMLLKD--VEEEKGV     40
2 mzcytor17lig      ---ATCSLSFGAPIS KEDLRTTIDLLKQES QDLYNNYSIKQASGM     42
3 mIL-3             ASISGRDTHRLTRTL NCS-SIVKEIIGKL- --PEP----ELKT--     35
4 hIL-3             APMTQTTPLKTSW-V NCS-NMIDEIITHLK QPPLP--LLDFNNLN     41

                    46              60 61           75 76           90
1 zcytor17lig       LVSQNYTLPCLSPDA QPPNNIHSPAIRAYL KTIRQLDNKSVIDEI     85
2 mzcytor17lig      SADESIQLPCFSLDR EALTNISVIIAHLEK VKVLSE-NTVDTSWV     86
3 mIL-3             DDEGPSLRNKS---- -----FRRVNLSKFV ESQGEVDPEDRYVIK     71
4 hIL-3             GEDQDILMENN---- -----LRRPNLEAFN RAVKSL--QNASAIE     75

                    91             105 106          120 121          135
1 zcytor17lig       IEHLDKLIFQDAPET NISVPTDTHE---CK RFILTISQQFSECMD     127
2 mzcytor17lig      IRWLTNISCFNPLNL NISVPGNTDESYDCK VFVLTVLKQFSNCMA     131
3 mIL-3             SNLQKLNCCLPTSAN DSALPGVFIRDLD-- DFRKKLRFYMVH-LN     113
4 hIL-3             SILKNLLPCLPLATA APTRHPIHIKDGDWN EFRRKLTFY----LK     116

                    136            150 151          165 166          180
1 zcytor17lig       LALKSLTSGAQQATT                                     142
2 mzcytor17lig      ELQAKDNTTC                                          141
3 mIL-3             DLETVLTSRPPQPAS GSVSPNRGTVEC                        140
4 hIL-3             TLENA----QAQQTT LSLAIF                              133
```

FIG. 1

```
                10        20        30        40        50
ZCYTOR MASHSGPSTSVLFLFCCLGGWLASHTLPVRLLRPSDDVQKIVEELQSLSKMLLKD--VEE
       . . . ..... ..... ... ..... .. .. . .. ...
M17RL- MIFHTGTTKPTLVLLCCIGTWLATCSLSFGAPISKEDLRTTIDLLKQESQDLYNNYSIKQ
               10        20        30        40        50        60

       60        70        80        90       100       110
ZCYTOR EKGVLVSQNYTLPCLSPDAQPPNNIHSPAIRAYLKTIRQLDNKSVIDE-IIEHLDKLIFQ
       ..:. ....  :::.: : .. .::  ..: :.:.... :....: .. .:  :...
M17RL- ASGMSADESIQLPCFSLDREALTNI--SVIIAHLEKVKVLSENTVDTSWVIRWLTNISCF
               70        80        90       100       110

       120       130          140       150       160
ZCYTOR DAPETNISVPTDTHE---CKRFILTISQQFSECM-DLALKSLTSGAQQATT
       .. . :::::.:.:   X: :.::. .::::X .:. :. :.
M17RL- NPLNLNISVPGNTDESYDCKVFVLTVLKQFSNCMAELQAKDNTTC
      120       130       140       150       160
```

FIG. 2

```
;          Hydrophobic                                                Hydrophilic
;          -3        -2        -1          0         1         2         3
;           |---------|---------|---------  | ---------|---------|---------|
   1   0.00                                 M
   2    0.00                                A
   3    0.38                                S ====
   4    0.49                                H =====
   5    0.51                                S =====
   6    0.28                                G ===
   7   -0.31                             === P
   8   -0.72                          ======= S
   9   -1.13                       ========== T
  10   -1.55                    ============== s
  11   -1.90                 ================= V
  12   -2.25              ==================== L
  13   -1.85                 ================= F
  14   -1.77                  ================ L
  15   -1.77              ================== F
  16   -1.35                 ============= C
  17   -1.05                   ========== C
  18   -1.20                  =========== L
  19   -1.33                 ============ G
  20   -1.25                 ============ G
  21   -0.90                    ========= W
  22   -0.98                   ========== L
  23   -1.05                   ========== A
  24   -0.78                     ======== S
  25   -0.48                       ===== H
  26   -0.65                     ======= T
  27   -0.20                         == L
  28   -0.42                       ==== P
  29   -0.65                  ======= V
  30    0.15                         R ==
  31    0.15                         L ==
  32    0.45                         L =====
  33    0.45                         R =====
  34    1.25                         P ============
  35    1.30                         S ============
```

FIG. 3A

```
36    0.83                                    D ========
37    1.33                                    D ============
38    0.98                                    V =========
39    0.23                                    Q ==
40    0.23                                    K ==
41    0.98                                    I =========
42    0.65                                    V =======
43    0.18                                    E ==
44    0.53                                    E =====
45    0.48                                    L =====
46    0.03                                    Q
47    0.03                                    S
48    0.12                                    L =
49   -0.22                                 == S
50   -0.57                                 ====== K
51    0.23                                    M ==
52    0.68                                    L =======
53   -0.07                                  = L
54    0.65                                    K =======
55    1.45                                    D ============
56    2.25                                    V ======================
57    2.25                                    E ======================
58    1.75                                    E ================
59    1.75                                    E ================
60    0.95                                    K =========
61    0.20                                    G ==
62   -0.25                                  === V
63   -0.72                                ======= L
64   -0.68                                ======= V
65   -0.42                                 ==== S
66   -0.18                                   == Q
67   -0.23                                   == N
68   -0.28                                   === y
69   -0.48                                  ===== T
70   -0.82                                ======== L
71   -0.78                                ======== P
72   -0.72                                 ======= C
73    0.08                                    L =
74    0.00                                    S
```

FIG. 3B

```
75    0.20                                    P ==
76    0.50                                    D =====
77    0.45                                    A =====
78    0.48                                    Q =====
79    0.02                                    P
80   -0.20                                 == P
81   -0.32                                 === N
82   -0.27                                 === N
83   -0.27                                 === I
84   -0.38                                 ==== H
85   -0.72                                 ======= S
86    0.08                                    P =
87    0.08                                    A =
88   -0.35                                 ==== I
89   -0.65                                 ======= R
90   -0.07                                   = A
91    0.17                                    Y ==
92   -0.63                                 ====== L
93   -0.05                                   K
94    0.37                                    T ====
95    0.37                                    I ====
96    0.37                                    R ====
97    0.47                                    Q =====
98    1.27                                    L ============
99    0.82                                    D ========
100   0.53                                    N =====
101   0.53                                    K =====
102   0.53                                    S =====
103   1.00                                    V ==========
104   0.20                                    I ==
105  -0.15                                 = D
106   0.60                                    E ======
107   0.82                                    I ========
108   0.02                                    I
109   0.02                                    E
110   0.82                                    H ========
111   0.82                                    L ========
112   0.02                                    D
```

FIG. 3C

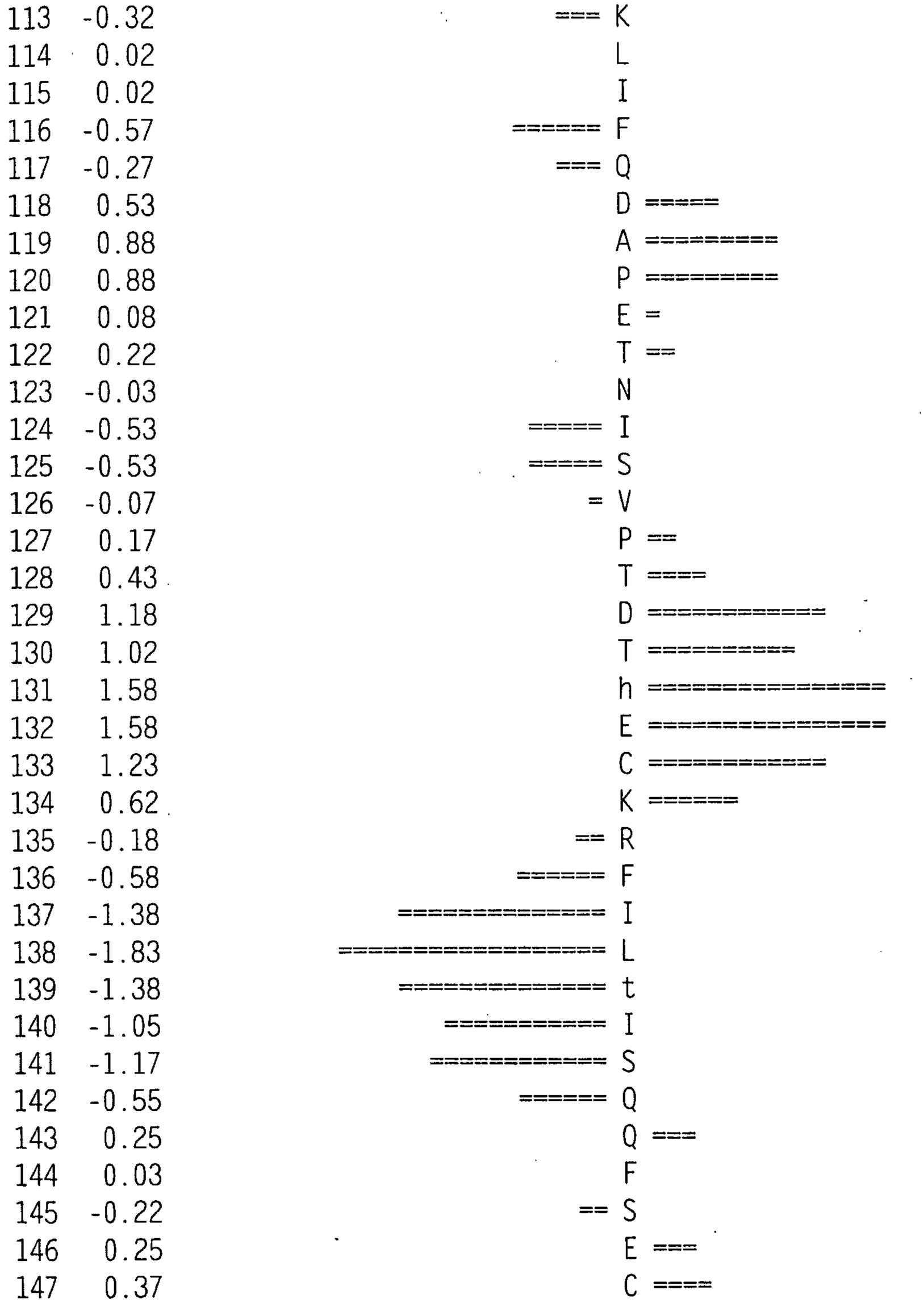

113 -0.32 === K
114 0.02 L
115 0.02 I
116 -0.57 ====== F
117 -0.27 === Q
118 0.53 D =====
119 0.88 A =========
120 0.88 P =========
121 0.08 E =
122 0.22 T ==
123 -0.03 N
124 -0.53 ===== I
125 -0.53 ===== S
126 -0.07 = V
127 0.17 P ==
128 0.43 T ====
129 1.18 D ============
130 1.02 T =========
131 1.58 h ================
132 1.58 E ================
133 1.23 C ============
134 0.62 K ======
135 -0.18 == R
136 -0.58 ====== F
137 -1.38 ============ I
138 -1.83 ================== L
139 -1.38 ============ t
140 -1.05 ========== I
141 -1.17 ============ S
142 -0.55 ====== Q
143 0.25 Q ===
144 0.03 F
145 -0.22 == S
146 0.25 E ===
147 0.37 C ====
148 0.23 M ==
149 -0.57 ====== D
150 0.10 L =

FIG. 3D

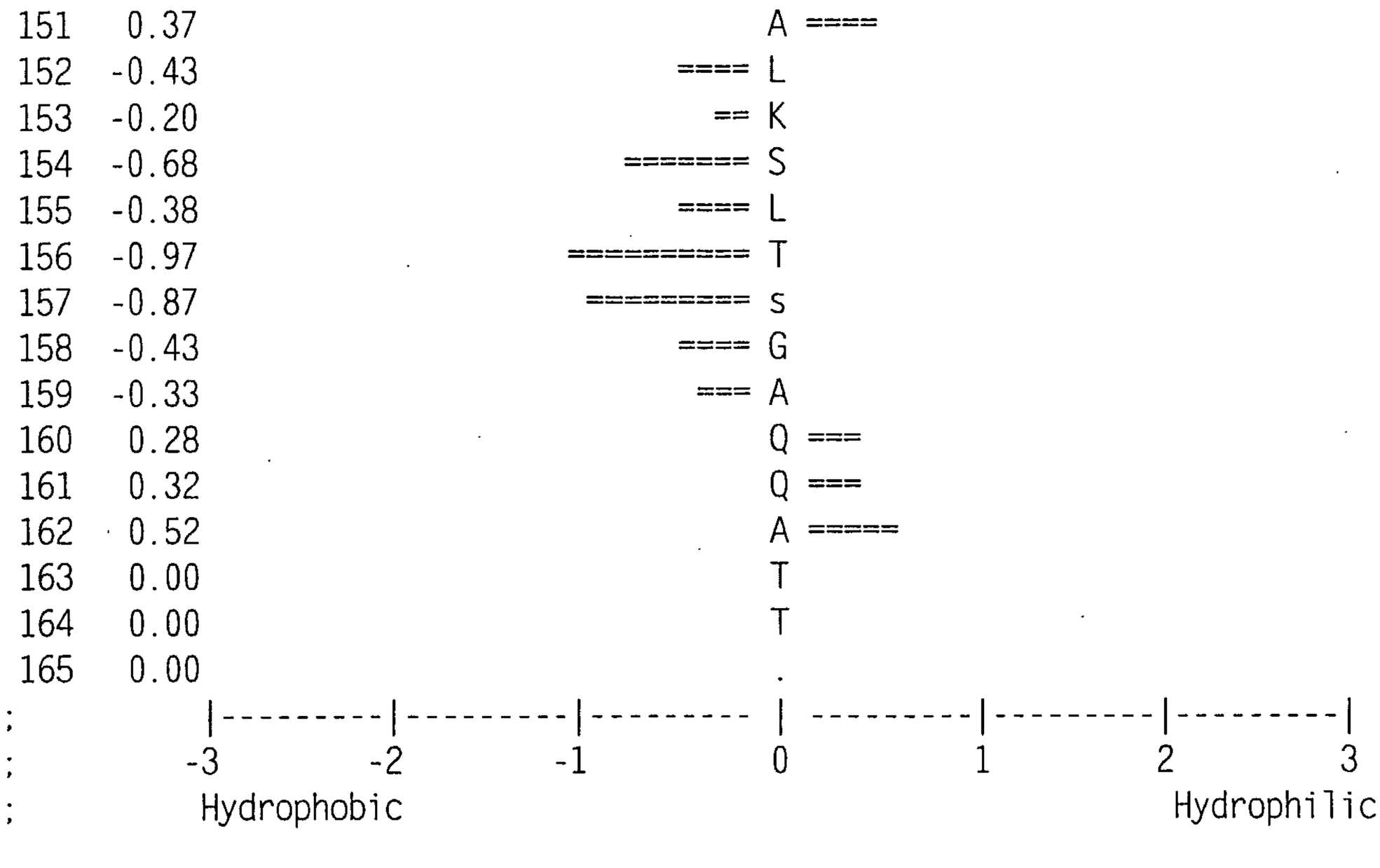

151 0.37 A ====
152 -0.43 ==== L
153 -0.20 == K
154 -0.68 ======= S
155 -0.38 ==== L
156 -0.97 ========= T
157 -0.87 ======== s
158 -0.43 ==== G
159 -0.33 === A
160 0.28 Q ===
161 0.32 Q ===
162 0.52 A =====
163 0.00 T
164 0.00 T
165 0.00 .
-3 -2 -1 0 1 2 3
Hydrophobic
Hydrophilic

FIG. 3E

```
                  1                                                   50
SEQ ID NO:109     MMWTWALWML PSLCKFSLAA LPAKPENISC VYYYRKNLTC TWSPGKETSY
SEQ ID NO:113     MMWTWALWML PSLCKFSLAA LPAKPENISC VYYYRKNLTC TWSPGKETSY
SEQ ID NO:111     MMWTWALWML PSLCKFSLAA LPAKPENISC VYYYRKNLTC TWSPGKETSY
SEQ ID NO:115     MMWTWALWML PSLCKFSLAA LPAKPENISC VYYYRKNLTC TWSPGKETSY

                  51                                                  100
SEQ ID NO:109     TQYTVKRTYA FGEKHDNCTT NSSTSENRAS CSFFLPRITI PDNYTIEVEA
SEQ ID NO:113     TQYTVKRTYA FGEKHDNCTT NSSTSENRAS CSFFLPRITI PDNYTIEVEA
SEQ ID NO:111     TQYTVKRTYA FGEKHDNCTT NSSTSENRAS CSFFLPRITI PDNYTIEVEA
SEQ ID NO:115     TQYTVKRTYA FGEKHDNCTT NSSTSENRAS CSFFLPRITI PDNYTIEVEA

                  101                                                 150
SEQ ID NO:109     ENGDGVIKSH MTYWRLENIA KTEPPKIFRV KPVLGIKRMI QIEWIKPELA
SEQ ID NO:113     ENGDGVIKSH MTYWRLENIA KTEPPKIFRV KPVLGIKRMI QIEWIKPELA
SEQ ID NO:111     ENGDGVIKSH MTYWRLENIA KTEPPKIFRV KPVLGIKRMI QIEWIKPELA
SEQ ID NO:115     ENGDGVIKSH MTYWRLENIA KTEPPKIFRV KPVLGIKRMI QIEWIKPELA

                  151                                                 200
SEQ ID NO:109     PVSSDLKYTL RFRTVNSTSW MEVNFAKNRK DKNQTYNLTG LQPFTEYVIA
SEQ ID NO:113     PVSSDLKYTL RFRTVNSTSW MEVNFAKNRK DKNQTYNLTG LQPFTEYVIA
SEQ ID NO:111     PVSSDLKYTL RFRTVNSTSW MEVNFAKNRK DKNQTYNLTG LQPFTEYVIA
SEQ ID NO:115     PVSSDLKYTL RFRTVNSTSW MEVNFAKNRK DKNQTYNLTG LQPFTEYVIA

                  201                                                 250
SEQ ID NO:109     LRCAVKESKF WSDWSQEKMG MTEEEAPCGL ELWRVLKPAE ADGRRPVRLL
SEQ ID NO:113     LRCAVKESKF WSDWSQEKMG MTEEEAPCGL ELWRVLKPAE ADGRRPVRLL
SEQ ID NO:111     LRCAVKESKF WSDWSQEKMG MTEEEAPCGL ELWRVLKPAE ADGRRPVRLL
SEQ ID NO:115     LRCAVKESKF WSDWSQEKMG MTEEEGKL.L PAIPVLSALV ~~~~~~~~~~

                  251                                                 300
SEQ ID NO:109     WKKARGAPVL EKTLGYNIWY YPESNTNLTE TMNTTNQQLE LHLGGESFWV
SEQ ID NO:113     WKKARGAPVL EKTLGYNIWY YPESNTNLTE TMNTTNQQLE LHLGGESFWV
SEQ ID NO:111     WKKARGAPVL EKTLGYNIWY YPESNTNLTE TMNTTNQQLE LHLGGESFWV
SEQ ID NO:115     ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~

                  301                                                 350
SEQ ID NO:109     SMISYNSLGK SPVATLRIPA IQEKSFQCIE VMQACVAEDQ LVVKWQSSAL
SEQ ID NO:113     SMISYNSLGK SPVATLRIPA IQEK~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:111     SMISYNSLGK SPVATLRIPA IQEKSFQCIE VMQACVAEDQ LVVKWQSSAL
SEQ ID NO:115     ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
```

FIG. 4A

```
                  351                                                 400
SEQ ID NO:109  DVNTWMIEWF PDVDSEPTTL SWESVSQATN WTIQQDKLKP FWCYNISVYP
SEQ ID NO:113  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:111  DVNTWMIEWF PDVDSEPTTL SWESVSQATN WTIQQDKLKP FWCYNISVYP
SEQ ID NO:115  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~

                  401                                                 450
SEQ ID NO:109  MLHDKVGEPY SIQAYAKEGV PSEGPETKVE NIGVKTVTIT WKEIPKSERK
SEQ ID NO:113  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:111  MLHDKVGEPY SIQAYAKEGV PSEGPETKVE NIGVKTVTIT WKEIPKSERK
SEQ ID NO:115  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~

                  451                                                 500
SEQ ID NO:109  GIICNYTIFY QAEGGKGFSK TVNSSILQYG LESLKRKTSY IVQVMASTSA
SEQ ID NO:113  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:111  GIICNYTIFY QAEGGKGFSK TVNSSILQYG LESLKRKTSY IVQVMASTSA
SEQ ID NO:115  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~

                  501                                                 550
SEQ ID NO:109  GGTNGTSINF KTLSFSVFEI ILITSLIGGG LLILIILTVA YGLKKPNKLT
SEQ ID NO:113  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:111  GGTNGTSINF KTLSFSVFEI ILITSLIGGG LLILIILTVA YGLKKPNKLT
SEQ ID NO:115  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~

                  551                                                 600
SEQ ID NO:109  HLCWPTVPNP AESSIATWHG DDFKDKLNLK ESDDSVNTED RILKPCSTPS
SEQ ID NO:113  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:111  HLCWPTVPNP AESSIATWHG DDFKDKLNLK ESDDSVNTED RILKPCSTPS
SEQ ID NO:115  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~

                  601                                                 650
SEQ ID NO:109  DKLVIDKLVV NFGNVLQEIF TDEARTGQEN NLGGEKNGTR ILSSCPTSI~
SEQ ID NO:113  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:111  DKLVIDKLVV NFGNVLQEIF TDEARTGQEN NLGGEKNGYV TCPFRPDCPL
SEQ ID NO:115  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~

                  651                                                 700
SEQ ID NO:109  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:113  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
SEQ ID NO:111  GKSFEELPVS PEIPPRKSQY LRSRMPEGTR PEAKEQLLFS GQSLVPDHLC
SEQ ID NO:115  ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~
```

FIG. 4B

```
                 701                                        733
SEQ ID NO:109    ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~
SEQ ID NO:113    ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~
SEQ ID NO:111    EEGAPNPYLK NSVTAREFLV SEKLPEHTKG EV~
SEQ ID NO:115    ~~~~~~~~~~ ~~~~~~~~~~ ~~~~~~~~~~ ~~~
```

FIG. 4C

```
                                    10        20        30        40
ZCYTOR                      MKLSPQPSCVNLGMMWTWALWMLPSLCKFSLAALPAKPENISCVYYY
                                    ::  :: :.::: ::: .  :::::::::::::::::.:.
M17R-O MLSSQKGSCSQEPGAAHVQPLGVNAGIMWTLALWAFSFLCKFSLAVLPTKPENISCVFYF
            10        20        30        40        50        60

            50        60        70        80        90       100
ZCYTOR RKNLTCTWSPGKETSYTQYTVKRTYAFGEKHDNCTTNSSTSENRASCSFFLPRITIPDNY
        .:::::: : :::. :.: :  ::.:.:.      .: : . ..:: ::        ::
M17R-O DRNLTCTWRPEKETNDTSYIVTLTYSYGK------SNYSDNATEASYSFPRSCAMPPDIC
                70        80                90       100       110

       110       120       130       140       150       160
ZCYTOR TIEVEAENGDGVIKSHMTYWRLENIAKTEPPKIFRVKPVLGIKRMIQIEWIKPELAPVSS
       ..:.:.:::: .:: .::::: :::::::: :. :::. .  ::.::.: ::.     .
M17R-O SVEVQAQNGDGKVKSDITYWHLISIAKTEPPIILSVNPICN--RMFQIQW-KPREKTRGF
          120       130       140       150       160       170

       170       180       190       200       210       220
ZCYTOR DLKYTLRFRTVNSTSWMEVNFAKNRKDKNQTYNLTGLQPFTEYVIALRCAVKESKFWSDW
        :   :::::::.. : :::: .: :   :. ::::: :::::.:::   .:.:.:: :
M17R-O PLVCMLRFRTVNSSRWTEVNF-ENCK---QVCNLTGLQAFTEYVLALRFRFNDSRYWSKW
               180       190           200       210       220

       230       240       250       260       270       280
ZCYTOR SQEKMGMTEEEAPCGLELWRVLKPAEADGRRPVRLLWKKARGAPVLEKTLGYNIWYYPES
       :.:.  .: :::.:  :.:::.:.::. .: : :::::::::::::::::.::.: :. :.
M17R-O SKEETRVTMEEVPHVLDLWRILEPADMNGDRKVRLLWKKARGAPVLEKTFGYHIQYFAEN
       230       240       250       260       270       280

       290       300       310       320       330       340
ZCYTOR NTNLTETMNTTNQQLELHLGGESFWVSMISYNSLGKSPVATLRIPAIQEKSFQCIEVMQA
       .:::::  : :.:: :: : ...  ::. :.::::::  . :::: .:.:.:: :. :::
M17R-O STNLTEINNITTQQYELLLMSQAHSVSVTSFNSLGKSQETILRIPDVHEKTFQYIKSMQA
       290       300       310       320       330       340

       350       360       370       380       390       400
ZCYTOR CVAEDQLVVKWQSSALDVNTWMIEWFPDVD-SEPTTLSWESVSQATNWTIQQDKLKPFWC
        .::  :::.:::: :.:::.:::.. :. .:::::::::::::::::::: :
M17R-O YIAEPLLVVNWQSSIPAVDTWIVEWLPEAAMSKFPALSWESVSQVTNWTIEQDKLKPFTC
       350       360       370       380       390       400
```

Fig. 5A

```
          410        420        430        440        450        460
ZCYTOR YNISVYPMLHDKVGEPYSIQAYAKEGVPSEGPETKVENIGVKTVTITWKEIPKSERKGII
       :::::::::  ::::::::::::::::: ::::.:::::.:::::::::: :.:.:
M17R-O YNISVYPVLGHRVGEPYSIQAYAKEGTPLKGPETRVENIGLRTATITWKEIPKSARNGFI
       410        420        430        440        450        460

          470        480        490        500        510        520
ZCYTOR CNYTIFYQAEGGKGFSKTVNSSILQYGLESLKRKTSYIVQVMASTSAGGTNGTSINFKTL
       ::::::::::: :::::::  ::  :::: :.::: : ::::: :::::: ::::::
M17R-O NNYTVFYQAEGGKELSKTVNSHALQCDLESLTRRTSYTVWVMASTRAGGTNGVRINFKTL
       470        480        490        500        510        520

          530        540        550        560        570        580
ZCYTOR SFSVFEIILITSLIGGGLLILIILTVAYGLKKPNKLTHLCWPTVPNPAESSIATWHGDDF
       :.::::::::::::::::: : ::.::::::.:: :: : :::::::::::: :: :
M17R-O SISVFEIVLLTSLVGGGLLLLSIKTVTFGLRKPNRLTPLCCPDVPNPAESSLATWLGDGF
       530        540        550        560        570        580

          590        600        610        620        630        640
ZCYTOR KDKLNLKESDDSVNTEDRILKPCSTPSDKLVIDKLVVNFGNVLQEIFTDEARTGQENNLG
       : : :.::. .: :::: .:::: .:.:  .::::::::: : :. .:.::  :: . ::
M17R-O K-KSNMKETGNSGNTEDVVLKPCPVPAD--LIDKLVVNFENFLEVVLTEEAGKGQASILG
        590        600        610         620        630        640

          650        660
ZCYTOR GEKNGTRILSSCPTSI
       :: :   :::. :.
M17R-O GEAN-EYILSQEPSCPGHC
           650        660
```

Fig 5B