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(71) Applicant(s)
BASF Plant Science GmbH

(72) Inventor(s)
Datla, Nagamani;Bauer, Jorg;Qiu, Xiao;Wu, Guohai;Cirpus, Petra

(74) Agent / Attorney
**Watermark Patent and Trade Marks Attorneys, Level 2 302 Burwood Road,
HAWTHORN, VIC, 3122**

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(54) Title: METHOD FOR PRODUCING POLYUNSATURATED FATTY ACIDS IN TRANSGENIC PLANTS

(54) Bezeichnung: VERFAHREN ZUR HERSTELLUNG MEHRFACH UNGESÄTTIGTER FETTSÄUREN IN TRANSGENEN
PFLANZEN

(57) Abstract: The invention relates to a method for producing polyunsaturated fatty acids in seeds of transgenic plants. According to said method, nucleic acids, coding for polypeptides with a ω -3-desaturase, Δ -12-desaturase, Δ -6-desaturase, Δ -6-elongase, Δ -5-desaturase, Δ -5-elongase and/or Δ -4-desaturase activity, preferably for polypeptides with a Δ -6-desaturase, Δ -6-elongase and Δ -5-desaturase activity, are introduced into the organism. The nucleic acid sequences are represented by SEQ ID NO: 11, SEQ ID NO: 27, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 and SEQ ID NO: 201. Advantageously, said nucleic acid sequences can be expressed in the organism optionally together with other nucleic acid sequences that code for polypeptides of the biosynthesis of the fatty acid or lipid metabolism. Nucleic acid sequences coding for a Δ -6-desaturase, Δ -5-desaturase, Δ -4-desaturase, Δ -12-desaturase and/or Δ -6-elongase activity are especially advantageous. Advantageously, said desaturases and elongases originate from thalassiosira, euglena or ostreococcus. The invention also relates to a method for producing oils and/or triacylglycerides with an increased content of long-chain polyunsaturated fatty acids. In a preferred embodiment, the invention also relates to a method for producing arachidonic acid, eicosapentaenoic acid or docosahexaenoic acid, and to a method for producing triglycerides with an increased content of unsaturated fatty acids, especially arachidonic acid, eicosapentaenoic acid and/or docosahexaenoic acid, in transgenic plants, preferably in seeds of the transgenic plants. The invention further relates to the production of a transgenic plant with an increased content of polyunsaturated fatty acids, especially arachidonic acid, eicosapentaenoic acid and/or docosahexaenoic acid, based on the expression of the elongases and desaturases used in the inventive method. The invention also relates to recombinant nucleic acid molecules containing, together or individually, nucleic acid sequences coding for the polypeptides with a Δ -6-desaturase, Δ -6-elongase, Δ -5-desaturase and Δ -5-elongase activity, and transgenic plants containing said recombinant nucleic acid molecules. Another part of the invention relates to oils, lipids and/or fatty acids produced according to the inventive method, and to the use thereof. Furthermore, the invention relates to unsaturated fatty acids and triglycerides with an increased content of unsaturated fatty acids, and to the use of the same.

(57) Zusammenfassung: Die vorliegende Erfindung betrifft ein Verfahren zur Herstellung von mehrfach ungesättigten Fettsäuren im Samen transgener Pflanzen, indem Nukleinsäuren in den Organismus eingebracht werden, die für Polypeptide mit ω -3-Desaturase-, Δ -12-Desaturase-, Δ -6-Desaturase-, Δ -6-Elongase-, Δ -5-Desaturase-, Δ -5-Elongase- und/oder Δ -4-Desaturaseaktivität bevorzugt für Polypeptide mit Δ -6-Desaturase-, Δ -6-Elongase- und Δ -5-Desaturaseaktivität codieren. Bei den Nukleinsäuresequenzen handelt es sich um die in SEQ ID NO: 11, SEQ ID NO: 27, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 und SEQ ID NO: 201 dargestellten Sequenzen. Vorteilhaft können diese Nukleinsäuresequenzen gegebenenfalls zusammen mit weiteren Nukleinsäuresequenzen, die für Polypeptide der Biosynthese des Fettsäure- oder Lipidstoffwechsels codieren, in dem Organismus exprimiert werden. Besonders vorteilhaft sind Nukleinsäuresequenzen, die für eine Δ -6-Desaturase-, eine Δ -5-Desaturase-, Δ -4-Desaturase-, Δ -12-Desaturase- und/oder Δ -6-Elongaseaktivität codieren. Vorteilhaft stammen diese Desaturasen und Elongasen aus Thalassiosira, Euglena oder Ostreococcus. Weiterhin betrifft die Erfindung ein Verfahren zur Herstellung von Ölen und/oder Triacylglyceriden mit einem erhöhten Gehalt an langkettigen mehrfach ungesättigten Fettsäuren. Die Erfindung betrifft in einer bevorzugten Ausführungsform ausserdem ein Verfahren zur Herstellung

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[CA/CA]; 2103 Kenderdine Road, Saskatoon Sk. S7N 4A9 (CA). **DATI.A, Nagamani** [CA/CA]; 527 Bayview Terrace, Saskatoon Sk. S7V 1B6 (CA).

(74) **Anwalt: PRESSLER, Uwe;** c/o BASF Aktiengesellschaft, 67056 Ludwigshafen (DE).

(81) **Bestimmungsstaaten** (*soweit nicht anders angegeben, für jede verfügbare nationale Schutzrechtsart*): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SM, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

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Zur Erklärung der Zweibuchstaben-Codes und der anderen Abkürzungen wird auf die Erklärungen ("Guidance Notes on Codes and Abbreviations") am Anfang jeder regulären Ausgabe der PCT-Gazette verwiesen.

von Arachidonsäure, Eicosapentaensäure oder Docosahexaensäure sowie ein Verfahren zur Herstellung von Triglyceriden mit einem erhöhten Gehalt an ungesättigten Fettsäuren, insbesondere Arachidonsäure, Eicosapentaensäure und/oder Docosahexaensäure, in transgenen Pflanzen vorteilhaft im Samen der transgenen Pflanze. Die Erfindung betrifft die Herstellung einer transgenen Pflanze mit erhöhtem Gehalt an mehrfach ungesättigten Fettsäuren, insbesondere Arachidonsäure, Eicosapentaensäure und/oder Docosahexaensäure, aufgrund der Expression der im erfindungsgemässen Verfahren verwendeten Elongasen und Desaturasen. Die Erfindung betrifft weiterhin rekombinante Nukleinsäuremoleküle, die die Nukleinsäuresequenzen, die für die Polypeptide mit Δ -6-Desaturase-, Δ -6-Elongase-, Δ -5-Desaturase- und Δ -5-Elongaseaktivität kodieren, gemeinsam oder einzeln enthalten, sowie transgene Pflanzen, die die vorgenannten rekombinanten Nukleinsäuremoleküle enthalten. Ein weiterer Teil der Erfindung betrifft Öle, Lipide und/oder Fettsäuren hergestellt nach dem erfindungsgemässen Verfahren und deren Verwendung. Ausserdem betrifft die Erfindung ungesättigte Fettsäuren sowie Triglyceride mit einem erhöhten Gehalt an ungesättigten Fettsäuren und deren Verwendung.

Process for the production of polyunsaturated fatty acids in transgenic plants

The present invention relates to a process for the production of polyunsaturated fatty acids in the seed of transgenic plants by introducing, into the organism, nucleic acids which encode polypeptides with ω 3-desaturase, Δ 12-desaturase, Δ 6-desaturase, Δ 6-elongase, Δ 5-desaturase, Δ 5-elongase and/or Δ 4-desaturase activity, preferably polypeptides with Δ 6-desaturase, Δ 6-elongase and Δ 5-desaturase activity.

The nucleic acid sequences are the sequences shown in SEQ ID NO: 11, SEQ ID NO: 27, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 and SEQ ID NO: 201. Preferably, a further nucleic acid sequence which encodes a polypeptide with a Δ 12-desaturase activity is additionally introduced into the plant, in addition to these nucleic acid sequences, and also expressed simultaneously. Especially preferably, this is the nucleic acid sequence shown in SEQ ID NO: 195.

These nucleic acid sequences can advantageously be expressed in the organism, if appropriate together with further nucleic acid sequences which encode polypeptides of the biosynthesis of the fatty acid or lipid metabolism. Especially advantageous are nucleic acid sequences which encode a Δ 6-desaturase, a Δ 5-desaturase, Δ 4-desaturase, Δ 12-desaturase and/or Δ 6-elongase activity. These desaturases and elongases originate advantageously from *Thalassiosira*, *Euglena* or *Ostreococcus*. Furthermore, the invention relates to a process for the production of oils and/or triacylglycerides with an elevated content of long-chain polyunsaturated fatty acids.

In a preferred embodiment, the invention furthermore relates to a process for the production of arachidonic acid, eicosapentaenoic acid or docosahexaenoic acid and to a process for the production of triglycerides with an elevated content of unsaturated fatty acids, in particular arachidonic acid, eicosapentaenoic acid and/or docosahexaenoic acid, in transgenic plants, advantageously in the seed of the transgenic plant. The invention relates to the generation of a transgenic plant with an elevated content of polyunsaturated fatty acids, in particular arachidonic acid, eicosapentaenoic acid and/or docosahexaenoic acid, as the result of the expression of the elongases and desaturases used in the process according to the invention.

The invention furthermore relates to recombinant nucleic acid molecules comprising the nucleic acid sequences which encode the polypeptides with Δ 6-desaturase, Δ 6-elongase, Δ 5-desaturase and Δ 5-elongase activity, either jointly or individually, and

transgenic plants which comprise the abovementioned recombinant nucleic acid molecules.

- 5 A further part of the invention relates to oils, lipids and/or fatty acids which have been produced by the process according to the invention, and to their use. Moreover, the invention relates to unsaturated fatty acids and to triglycerides with an elevated content of unsaturated fatty acids and to their use.

- 10 Lipid synthesis can be divided into two sections: the synthesis of fatty acids and their binding to sn-glycerol-3-phosphate, and the addition or modification of a polar head group. Usual lipids which are used in membranes comprise phospholipids, glycolipids, sphingolipids and phosphoglycerides. Fatty acid synthesis starts with the conversion of acetyl-CoA into malonyl-CoA by acetyl-CoA carboxylase or into acetyl-ACP by acetyl transacylase. After condensation reaction, these two product molecules together form
- 15 acetoacetyl-ACP, which is converted via a series of condensation, reduction and dehydration reactions so that a saturated fatty acid molecule with the desired chain length is obtained. The production of the unsaturated fatty acids from these molecules is catalyzed by specific desaturases, either aerobically by means of molecular oxygen or anaerobically (regarding the fatty acid synthesis in microorganisms, see F.C. Neidhardt et al. (1996) *E. coli* and *Salmonella*. ASM Press: Washington, D.C., p. 612-636 and references cited therein; Lengeler et al. (Ed.) (1999) *Biology of Prokaryotes*. Thieme: Stuttgart, New York, and the references therein, and Magnuson, K., et al. (1993) *Microbiological Reviews* 57:522-542 and the references therein). To undergo
- 20 the further elongation steps, the resulting phospholipid-bound fatty acids must be returned to the fatty acid CoA ester pool. This is made possibly by acyl-CoA:lysophospholipid acyltransferases. Moreover, these enzymes are capable of transferring the elongated fatty acids from the CoA esters back to the phospholipids. If appropriate, this reaction sequence can be followed repeatedly.
- 30 Furthermore, fatty acids must subsequently be transported to various modification sites and incorporated into the triacylglycerol storage lipid. A further important step during lipid synthesis is the transfer of fatty acids to the polar head groups, for example by glycerol fatty acid acyltransferase (see Frentzen, 1998, *Lipid*, 100(4-5):161-166).
- 35 With regard to publications on the biosynthesis of fatty acids in plants, desaturation, the lipid metabolism and the membrane transport of lipidic compounds, beta-oxidation, the modification of fatty acids and cofactors and the storage and assembly of triacylglycerol, including the references cited therein, see the following papers: Kinney,

- 1997, Genetic Engineering, Ed.: JK Setlow, 19:149-166; Ohlrogge and Browse, 1995, Plant Cell 7:957-970; Shanklin and Cahoon, 1998, Annu. Rev. Plant Physiol. Plant Mol. Biol. 49:611-641; Voelker, 1996, Genetic Engineering, Ed.: JK Setlow, 18:111-13; Gerhardt, 1992, Prog. Lipid R. 31:397-417; Gühnemann-Schäfer & Kindl, 1995, Biochim. Biophys Acta 1256:181-186; Kunau et al., 1995, Prog. Lipid Res. 34:267-342; Stymne et al., 1993, in: Biochemistry and Molecular Biology of Membrane and Storage Lipids of Plants, Eds.: Murata and Somerville, Rockville, American Society of Plant Physiologists, 150-158, Murphy & Ross 1998, Plant Journal. 13(1):1-16.
- 10 In the text which follows, polyunsaturated fatty acids are referred to as PUFA, PUFAs, LCPUFA or LCPUFAs (poly unsaturated fatty acids, PUFA, long chain poly unsaturated fatty acids, LCPUFA).

Fatty acids and triacylglycerides have a multiplicity of applications in the food industry, in animal nutrition, in cosmetics and the pharmacological sector. Depending on whether they are free saturated or unsaturated fatty acids or else triacylglycerides with an elevated content of saturated or unsaturated fatty acids, they are suitable for very different applications. Polyunsaturated fatty acids such as linoleic and linolenic acid are essential for mammals since they cannot be produced by the latter. This is why polyunsaturated ω 3-fatty acids and ω 6-fatty acids are an important constituent of human and animal food. Thus, for example, lipids with unsaturated fatty acids, specifically with polyunsaturated fatty acids, are preferred in human nutrition. The polyunsaturated ω 3-fatty acids are supposed to have a positive effect on the cholesterol level in the blood and thus on the prevention of heart disease. The risk of heart disease, strokes or hypertension can be reduced markedly by adding these ω 3-fatty acids to the food (Shimikawa 2001, World Rev. Nutr. Diet. 88, 100-108).

ω 3-fatty acids also have a positive effect on inflammatory, specifically on chronically inflammatory, processes in association with immunological diseases such as rheumatoid arthritis (Calder 2002, Proc. Nutr. Soc. 61, 345-358; Cleland and James 2000, J. Rheumatol. 27, 2305-2307). They are therefore added to foodstuffs, specifically to dietetic foodstuffs, or are employed in medicaments. ω 6-fatty acids such as arachidonic acid tend to have a negative effect in connection with these rheumatological diseases.

ω 3- and ω 6-fatty acids are precursors of tissue hormones, known as eicosanoids, such as the prostaglandins, which are derived from dihomo- γ -linolenic acid, arachidonic acid and eicosapentaenoic acid, and of the thromboxanes and leukotrienes, which are

derived from arachidonic acid and eicosapentaenoic acid. Eicosanoids (known as the PG₂ series) which are formed from the ω 6-fatty acids, generally promote inflammatory reactions, while eicosanoids (known as the PG₃ series) from ω 3-fatty acids have little or no proinflammatory effect.

- 5 Polyunsaturated long-chain ω 3-fatty acids such as eicosapentaenoic acid (= EPA, C20:5 ^{Δ 5,8,11,14,17}) or docosahexaenoic acid (= DHA, C22:6 ^{Δ 4,7,10,13,16,19}) are important components of human nutrition owing to their various roles in health aspects, including the development of the child brain, the functionality of the eyes, the synthesis of hormones and other signal substances, and the prevention of cardiovascular disorders, cancer and diabetes (Poulos, A Lipids 30:1-14, 1995; Horrocks, LA and Yeo YK Pharmacol Res 40:211-225, 1999). There is therefore a demand for the production of polyunsaturated long-chain fatty acids.

- Owing to the present-day composition of human food, an addition of polyunsaturated ω 3-fatty acids, which are preferentially found in fish oils, to the food is particularly important. Thus, for example, polyunsaturated fatty acids such as docosahexaenoic acid (= DHA, C22:6 ^{Δ 4,7,10,13,16,19}) or eicosapentaenoic acid (= EPA, C20:5 ^{Δ 5,8,11,14,17}) are added to infant formula to improve the nutritional value. The unsaturated fatty acid DHA is supposed to have a positive effect on the development and maintenance of brain function. There is therefore a demand for the production of polyunsaturated long-chain fatty acids.

- The various fatty acids and triglycerides are mainly obtained from microorganisms such as Mortierella or Schizochytrium or from oil-producing plants such as soybeans, oilseed rape, algae such as Crypthecodinium or Phaeodactylum and others, being obtained, as a rule, in the form of their triacylglycerides (= triglycerides = triglycerols). However, they can also be obtained from animals, for example, fish. The free fatty acids are advantageously prepared by hydrolysis. Very long-chain polyunsaturated fatty acids such as DHA, EPA, arachidonic acid (ARA, C20:4 ^{Δ 5,8,11,14}), dihomo- γ -linolenic acid (C20:3 ^{Δ 8,11,14}) or docosapentaenoic acid (DPA, C22:5 ^{Δ 7,10,13,16,19}) are, however, not synthesized in oil crops such as oilseed rape, soybeans, sunflowers and safflower. Conventional natural sources of these fatty acids are fish such as herring, salmon, sardine, redfish, eel, carp, trout, halibut, mackerel, zander or tuna, or algae.

- Depending on the intended use, oils with saturated or unsaturated fatty acids are preferred. In human nutrition, for example, lipids with unsaturated fatty acids, specifically polyunsaturated fatty acids, are preferred. The polyunsaturated ω 3-fatty acids are said to have a positive effect on the cholesterol level in the blood and thus on

- the possibility of preventing heart disease. The risk of heart disease, stroke or hypertension can be reduced markedly by adding these ω 3-fatty acids to the food. Also, ω 3-fatty acids have a positive effect on inflammatory, specifically on chronically inflammatory, processes in association with immunological diseases such as
- 5 rheumatoid arthritis. They are therefore added to foodstuffs, specifically to dietetic foodstuffs, or are employed in medicaments. ω 3-fatty acids such as arachidonic acid tend to have an adverse effect on these disorders in connection with these rheumatic diseases on account of our usual dietary intake.
- 10 Owing to their positive characteristics, there has been no lack of attempts in the past to make available genes which are involved in the synthesis of these fatty acids or triglycerides for the production of oils in various organisms with a modified content of unsaturated fatty acids. Thus, WO 91/13972 and its US equivalent describe a Δ 9-desaturase. WO 93/11245 claims a Δ 15-desaturase and WO 94/11516 a Δ 12-
- 15 desaturase. Further desaturases are described, for example, in EP-A-0 550 162, WO 94/18337, WO 97/30582, WO 97/21340, WO 95/18222, EP-A-0 794 250, Stucky et al., J. Biol. Chem., 265, 1990: 20144–20149, Wada et al., Nature 347, 1990: 200–203 or Huang et al., Lipids 34, 1999: 649–659. However, the biochemical
- 20 characterization of the various desaturases has been insufficient to date since the enzymes, being membrane-bound proteins, present great difficulty in their isolation and characterization (McKeon et al., Methods in Enzymol. 71, 1981: 12141–12147, Wang et al., Plant Physiol. Biochem., 26, 1988: 777–792). As a rule, membrane-bound desaturases are characterized by being introduced into a suitable organism which is subsequently analyzed for enzyme activity by analyzing the starting materials and the
- 25 products. Δ 6-Desaturases are described in WO 93/06712, US 5,614,393, US5614393 WO 96/21022, WO 00/21557 and WO 99/27111. The application of this enzyme for the production of fatty acids in transgenic organisms is described in WO 98/46763, WO 98/46764 and WO 98/46765. The expression of various desaturases is described and claimed in WO 99/64616 or WO 98/46776. As regards the expression efficacy of
- 30 desaturases and its effect on the formation of polyunsaturated fatty acids, it must be noted that the expression of a single desaturase as described to date has only resulted in low contents of unsaturated fatty acids/lipids such as, for example, γ -linolenic acid and stearidonic acid.
- There have been a number of attempts in the past to obtain elongase genes. Millar and
- 35 Kunst, 1997 (Plant Journal 12:121-131) and Millar et al., 1999 (Plant Cell 11:825-838) describe the characterization of plant elongases for the synthesis of monounsaturated long-chain fatty acids (C22:1) and for the synthesis of very long-chain fatty acids for the

formation of waxes in plants (C_{28} - C_{32}). The synthesis of arachidonic acid and EPA is described, for example, in WO 01/59128, WO 00/12720, WO 02/077213 and WO 02/08401. The synthesis of polyunsaturated C24-fatty acids is described, for example, in Tvrdik et al. 2000, J. Cell Biol. 149:707-718 or WO 02/44320.

- 5 Especially suitable microorganisms for the production of PUFAs are microorganisms such as microalgae such as *Phaeodactylum tricornutum*, *Porphyridium* species, *Thraustochytrium* species, *Schizochytrium* species or *Cryptocodinium* species, ciliates such as *Stylonychia* or *Colpidium*, fungi such as *Mortierella*, *Entomophthora* or *Mucor* and/or mosses such as *Physcomitrella*, *Ceratodon* and *Marchantia* (R. Vazhappilly & F. Chen (1998) *Botanica Marina* 41: 553-558; K. Totani & K. Oba (1987) *Lipids* 22: 1060-1062; M. Akimoto et al. (1998) *Appl. Biochemistry and Biotechnology* 73: 269-278). Strain selection has resulted in the development of a number of mutant strains of the microorganisms in question which produce a series of desirable compounds including PUFAs. However, the mutation and selection of strains with an improved production of a particular molecule such as the polyunsaturated fatty acids is a time-consuming and difficult process, which is why as described above recombinant methods are preferred. However, only limited amounts of the desired polyunsaturated fatty acids such as DPA, EPA or ARA can be produced with the aid of the abovementioned microorganisms; where, as a rule, they are generally obtained as fatty acid mixtures, depending on the microorganisms used.

- Higher plants comprise polyunsaturated fatty acids such as linoleic acid ($C_{18:2}$) and linolenic acid ($C_{18:3}$). ARA, EPA and DHA are found not at all in the seed oil of higher plants, or only in miniscule amounts (E. Ucciani: *Nouveau Dictionnaire des Huiles Végétales* [New Dictionary of the Vegetable Oils]. Technique & Documentation – Lavoisier, 1995. ISBN: 2-7430-0009-0). However, the production of LCPUFAs in higher plants, preferably in oilseed crops such as oilseed rape, linseed, sunflowers and soybeans, would be advantageous since large amounts of high-quality LCPUFAs for the food industry, animal nutrition and pharmaceutical purposes might be obtained economically. To this end, it is advantageous to introduce, into oilseed crops, genes which encode enzymes of the LCPUFA biosynthesis via recombinant methods and to express them therein. These genes encode for example $\Delta 6$ -desaturases, $\Delta 6$ -elongases, $\Delta 5$ -desaturases, $\Delta 5$ -elongases or $\Delta 4$ -desaturases. These genes can advantageously be isolated from microorganisms and lower plants which produce LCPUFAs and incorporate them in the membranes or triacylglycerides. Thus, it has already been possible to isolate $\Delta 6$ -desaturase genes from the moss *Physcomitrella patens* and $\Delta 6$ -elongase genes from *P. patens* and from the nematode *C. elegans*.

A variety of synthetic pathways is being discussed for the synthesis of arachidonic acid, eicosapentaenoic acid (EPA) and docosahexaenoic acid (DHA) (Figure 1). Thus, EPA or DHA are produced in marine bacteria such as *Vibrio* sp. or *Shewanella* sp. via the polyketide pathway (Yu, R. et al. *Lipids* 35:1061-1064, 2000; Takeyama, H. et al. *Microbiology* 143:2725-2731, 1997).

An alternative strategy is the alternating activity of desaturases and elongases (Zank, T.K. et al. *Plant Journal* 31:255-268, 2002; Sakuradani, E. et al. *Gene* 238:445-453, 1999). A modification of the above-described pathway by $\Delta 6$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and $\Delta 4$ -desaturase is the Sprecher pathway (Sprecher 2000, *Biochim. Biophys. Acta* 1486:219-231) in mammals. Instead of the $\Delta 4$ -desaturation, a further elongation step is effected here to give C_{24} , followed by a further $\Delta 6$ -desaturation and finally β -oxidation to give the C_{22} chain length. Thus what is known as Sprecher pathway (see Figure 1) is, however, not suitable for the production in plants and microorganisms since the regulatory mechanisms are not known.

Depending on their desaturation pattern, the polyunsaturated fatty acids can be divided into two large classes, viz. $\omega 6$ - or $\omega 3$ -fatty acids, which differ with regard to their metabolic and functional activities (Fig. 1).

The starting material for the $\omega 6$ -metabolic pathway is the fatty acid linoleic acid ($18:2^{\Delta 9,12}$) while the $\omega 3$ -pathway proceeds via linolenic acid ($18:3^{\Delta 9,12,15}$). Linolenic acid is formed by the activity of an $\omega 3$ -desaturase (Tocher et al. 1998, *Prog. Lipid Res.* 37, 73-117 ; Domergue et al. 2002, *Eur. J. Biochem.* 269, 4105-4113).

Mammals, and thus also humans, have no corresponding desaturase activity ($\Delta 12$ - and $\omega 3$ -desaturase) and must take up these fatty acids (essential fatty acids) via the food. Starting with these precursors, the physiologically important polyunsaturated fatty acids arachidonic acid (= ARA, $20:4^{\Delta 5,8,11,14}$), an $\omega 6$ -fatty acid and the two $\omega 3$ -fatty acids eicosapentaenoic acid (= EPA, $20:5^{\Delta 5,8,11,14,17}$) and docosahexaenoic acid (DHA, $22:6^{\Delta 4,7,10,13,17,19}$) are synthesized via the sequence of desaturase and elongase reactions. The application of $\omega 3$ -fatty acids shows the therapeutic activity described above in the treatment of cardiovascular diseases (Shimikawa 2001, *World Rev. Nutr. Diet.* 88, 100-108), inflammations (Calder 2002, *Proc. Nutr. Soc.* 61, 345-358) and arthritis (Cleland and James 2000, *J. Rheumatol.* 27, 2305-2307).

From the angle of nutritional physiology, it is therefore advantageous to achieve a shift

between the ω 6-synthetic pathway and the ω 3-synthetic pathway (see Figure 1) so that more ω 3-fatty acids are produced. The enzymatic activities of various ω 3-desaturases which desaturate C_{18:2}-, C_{22:4}- or C_{22:5}-fatty acids have been described in the literature (see Figure 1). However, none of the desaturases whose biochemistry has been

5 described converts a broad range of substrates of the ω 6-synthetic pathway into the corresponding fatty acids of the ω 3-synthetic pathway.

The elongation of fatty acids, by elongases, by 2 or 4 C atoms is of crucial importance for the production of C₂₀- and C₂₂-PUFAs, respectively. This process proceeds via 4

10 steps. The first step is the condensation of malonyl-CoA onto the fatty-acid-acyl-CoA by ketoacyl-CoA synthase (KCS, hereinbelow referred to as elongase). This is followed by a reduction step (ketoacyl-CoA reductase, KCR), a dehydration step (dehydratase) and a final reduction step (enoyl-CoA reductase). It has been postulated that the elongase activity affects the specificity and rate of the entire process (Millar

15 and Kunst, 1997 Plant Journal 12:121-131).

No specific elongase has been described to date for the production of DHA (C_{22:6} n-3) in organisms which do not naturally produce this fatty acid. Only elongases which provide C₂₀- or C₂₄-fatty acids have been described to date. A Δ 5-elongase activity has

20 not been described to date.

The first transgenic plants which comprise and express genes encoding LCPUFA biosynthesis enzymes and which, as a consequence, produce LCPUFAs were described for the first time, for example, in DE-A-102 19 203 (Process for the

25 production of polyunsaturated fatty acids in plants) or in WO 2004/071467. However, these plants produce LCPUFAs in amounts which require further optimization for processing the oils which are present in the plants. Thus, ARA content in the plants described in DE-A-102 19 203 only amounts to 0.4 to 2% and the EPA content only to 0.5 to 1%, in each case based on the total lipid content of the plants. WO 2004/071467

30 discloses higher contents of polyunsaturated C₂₀- and C₂₂-fatty acids such as ARA, EPA or DHA. However, the process disclosed has a series of grave disadvantages. It seems that DHA cannot be detected at all in the seeds in the process disclosed. To produce PUFAs, soybean is less suitable, owing to its low oil content of approximately only 20% by weight. Soybean is an advantageous protein source and is therefore

35 grown on a large scale. However, the oil content of soybeans is rather low. Moreover, the dihomog- γ -linolenic acid (=DGHL or HGLA) content obtained in the production process is much too high. HGLA is hardly detectable in fish oils or algal oils or microbial oils. A further disadvantage is that the plants disclosed in WO 2004/071467

were generated by cotransformation, which leads to the segregation of the characteristics in the subsequent generations, and thus to an increased selection effort.

To make possible the fortification of food and/or of feed with these polyunsaturated fatty acids, there is therefore a great need for a simple, inexpensive process for the production of these polyunsaturated fatty acids in plant systems, especially in the seed of transgenic plants.

The object of the invention was therefore to develop a process for the production of large amounts of polyunsaturated fatty acids, specifically ARA, EPA and DHA, in the seed of a transgenic plant. This object was achieved by the process according to the invention for the production of C₁₈-, C₂₀- and C₂₂- polyunsaturated fatty acids in the seed of a transgenic oil crop plant with a content of at least 20% by weight based on the total lipid content, which comprises the following process steps:

a) introducing, into the oil crop plant, at least one nucleic acid encoding a $\Delta 6$ -desaturase and at least one nucleic acid encoding a $\Delta 6$ -elongase, wherein the $\Delta 6$ -desaturase has the amino acid sequence shown in SEQ ID NO: 202 or an amino acid sequence having at least 60% sequence identity thereto, or

introducing, into the oil crop plant, at least one nucleic acid encoding a $\Delta 9$ -elongase, at least one nucleic acid encoding a $\Delta 8$ -desaturase, at least one nucleic acid encoding a $\Delta 6$ -desaturase and at least one nucleic acid encoding a $\Delta 6$ -elongase, wherein the $\Delta 6$ -desaturase has the amino acid sequence shown in SEQ ID NO: 202 or an amino acid sequence having at least 60% sequence identity thereto, and

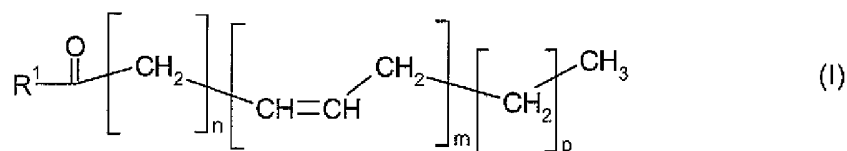
b) introducing, into the oil crop plant, at least one nucleic acid encoding a $\Delta 5$ -desaturase, and

c) introducing, into the oil crop plant, at least one nucleic acid encoding a $\Delta 5$ -elongase, and

d) introducing, into the oil crop plant, at least one nucleic acid encoding a $\Delta 4$ -desaturase.

9a

In one embodiment of the invention there is provided a process for the production of compounds of the general formula I



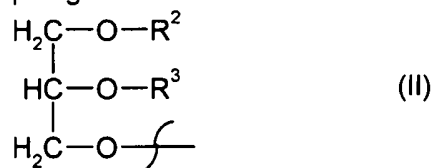
in the seeds of transgenic plants with a content of at least 20% by weight based on the total lipid content, which comprises the following process steps:

- a) introducing, into the organism, at least one nucleic acid sequence which encodes a $\Delta 9$ -elongase and $\Delta 6$ -desaturase activity, and
- b) introducing, into the organism, at least one nucleic acid sequence which encodes a $\Delta 8$ -desaturase and $\Delta 6$ -elongase activity, and
- c) introducing, into the organism, at least one nucleic acid sequence which encodes a $\Delta 5$ -desaturase activity, and
- d) introducing, into the organism, at least one nucleic acid sequence which encodes a $\Delta 5$ -elongase activity, and
- e) introducing, into the organism, at least one nucleic acid sequence which encodes a $\Delta 4$ -desaturase activity, and

where the variables and substituents in formula I have the following meanings:

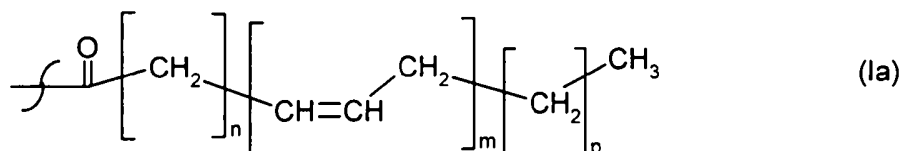
$\text{R}^1 =$ hydroxyl, coenzyme A (thioester), lysophosphatidylcholine,

lysophosphatidylethanolamine, lysophosphatidylglycerol, lysodiphosphatidylglycerol, lysophosphatidylserine, lysophosphatidylinositol, sphingo base or a radical of the formula II



5 $\text{R}^2 =$ hydrogen, lysophosphatidylcholine, lysophosphatidylethanolamine, lysophosphatidylglycerol, lysodiphosphatidylglycerol, lysophosphatidylserine, lysophosphatidylinositol or saturated or unsaturated C_2 - C_{24} -alkylcarbonyl,

10 $\text{R}^3 =$ hydrogen, saturated or unsaturated C_2 - C_{24} -alkylcarbonyl, or R^2 and R^3 independently of one another are a radical of the formula Ia:



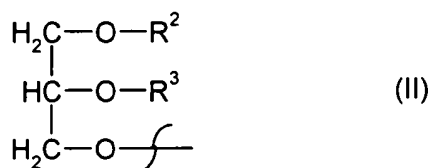
$n = 2, 3, 4, 5, 6, 7$ or 9 , $m = 2, 3, 4, 5$ or 6 and $p = 0$ or 3 . Advantageously, the variables n , m and p in the abovementioned formula I and Ia denote the following: $n = 2, 3$ or 5 , $m = 4, 5$ or 6 and $p = 0$ or 3 . In an especially advantageous embodiment of

15 the process, the variables n , m and p in the formulae I and Ia denote the following: $m = 4$, $n = 3$, $p = 3$ and the compounds of the general formula I and Ia thus denote arachidonic acid, and/or $m = 5$, $n = 3$, $p = 0$ and the compounds of the general formula I and Ia thus denote eicosapentaenoic acid, and/or $m = 5$, $n = 5$, $p = 0$ and the compounds of the general formula I and Ia thus denote docosapentaenoic acid is

20 and/or $m = 6$, $n = 3$, $p = 0$ and the compounds of the general formula I and Ia thus denote docosahexaenoic acid is.

R^1 in the general formula I is hydroxyl, coenzyme A (thioester), lysophosphatidylcholine, lysophosphatidylethanolamine, lysophosphatidylglycerol, lysodiphosphatidylglycerol, lysophosphatidylserine, lysophosphatidylinositol, sphingo base

25 or a radical of the formula II



The abovementioned radicals of R^1 are always bonded to the compounds of the general formula I in the form of their thioesters.

R^2 in the general formula II is hydrogen, lysophosphatidylcholine, lysophosphatidylethanolamine, lysophosphatidylglycerol, lysodiphosphatidylglycerol, lysophosphatidylserine, lysophosphatidylinositol or saturated or unsaturated C_2 - C_{24} -alkylcarbonyl.

Alkyl radicals which may be mentioned are substituted or unsubstituted, saturated or unsaturated C_2 - C_{24} -alkylcarbonyl chains such as ethylcarbonyl, n-propylcarbonyl, n-butylcarbonyl, n-pentylcarbonyl, n-hexylcarbonyl, n-heptylcarbonyl, n-octylcarbonyl, n-nonylcarbonyl, n-decylcarbonyl, n-undecylcarbonyl, n-dodecylcarbonyl, n-tridecylcarbonyl, n-tetradecylcarbonyl, n-pentadecylcarbonyl, n-hexadecylcarbonyl, n-heptadecylcarbonyl, n-octadecylcarbonyl, n-nonadecylcarbonyl, n-eicosylcarbonyl, n-docosanylcarbonyl- or n-tetracosanylcarbonyl, which comprise one or more double bonds. Saturated or unsaturated C_{10} - C_{22} -alkylcarbonyl radicals such as n-decylcarbonyl, n-undecylcarbonyl, n-dodecylcarbonyl, n-tridecylcarbonyl, n-tetradecylcarbonyl, n-pentadecylcarbonyl, n-hexadecylcarbonyl, n-heptadecylcarbonyl, n-octadecylcarbonyl, n-nonadecylcarbonyl, n-eicosylcarbonyl, n-docosanylcarbonyl or n-tetracosanylcarbonyl, which comprise one or more double bonds are preferred. Especially preferred are saturated and/or unsaturated C_{10} - C_{22} -alkylcarbonyl radicals such as C_{10} -alkylcarbonyl, C_{11} -alkylcarbonyl, C_{12} -alkylcarbonyl, C_{13} -alkylcarbonyl, C_{14} -alkylcarbonyl, C_{16} -alkylcarbonyl, C_{18} -alkylcarbonyl, C_{20} -alkylcarbonyl or C_{22} -alkylcarbonyl radicals which comprise one or more double bonds. Very especially preferred are saturated or unsaturated C_{16} - C_{22} -alkylcarbonyl radicals such as C_{16} -alkylcarbonyl, C_{18} -alkylcarbonyl, C_{20} -alkylcarbonyl or C_{22} -alkylcarbonyl radicals which comprise one or more double bonds. These advantageous radicals can comprise two, three, four, five or six double bonds. The especially preferred radicals with 20 or 22 carbon atoms in the fatty acid chain comprise up to six double bonds, advantageously three, four, five or six double bonds, especially preferably four, five or six double bonds, very especially preferably five or six. All the abovementioned radicals are derived from the corresponding fatty acids.

R^3 in the formula II is hydrogen, saturated or unsaturated C_2 - C_{24} -alkylcarbonyl.

Alkyl radicals which may be mentioned are substituted or unsubstituted, saturated or unsaturated C_2 - C_{24} -alkylcarbonyl chains such as ethylcarbonyl, n-propylcarbonyl, n-butylcarbonyl, n-pentylcarbonyl, n-hexylcarbonyl, n-heptylcarbonyl, n-octylcarbonyl,

n-nonylcarbonyl, n-decylcarbonyl, n-undecylcarbonyl, n-dodecylcarbonyl, n-tridecylcarbonyl, n-tetradecylcarbonyl, n-pentadecylcarbonyl, n-hexadecylcarbonyl, n-heptadecylcarbonyl, n-octadecylcarbonyl-, n-nonadecylcarbonyl, n-eicosylcarbonyl, n-docosanylcarbonyl- or n-tetracosanylcarbonyl, which comprise one or more double bonds. Saturated or unsaturated C₁₀-C₂₂-alkylcarbonyl radicals such as n-decylcarbonyl, n-undecylcarbonyl, n-dodecylcarbonyl, n-tridecylcarbonyl, n-tetradecylcarbonyl, n-pentadecylcarbonyl, n-hexadecylcarbonyl, n-heptadecylcarbonyl, n-octadecylcarbonyl, n-nonadecylcarbonyl, n-eicosylcarbonyl, n-docosanylcarbonyl or n-tetracosanylcarbonyl, which comprise one or more double bonds are preferred. Especially preferred are saturated and/or unsaturated C₁₀-C₂₂-alkylcarbonyl radicals such as C₁₀-alkylcarbonyl, C₁₁-alkylcarbonyl, C₁₂-alkylcarbonyl, C₁₃-alkylcarbonyl, C₁₄-alkylcarbonyl, C₁₆-alkylcarbonyl, C₁₈-alkylcarbonyl, C₂₀-alkylcarbonyl or C₂₂-alkylcarbonyl radicals which comprise one or more double bonds. Very especially preferred are saturated or unsaturated C₁₆-C₂₂-alkylcarbonyl radicals such as C₁₆-alkylcarbonyl, C₁₈-alkylcarbonyl, C₂₀-alkylcarbonyl or C₂₂-alkylcarbonyl radicals which comprise one or more double bonds. These advantageous radicals can comprise two, three, four, five or six double bonds. The especially preferred radicals with 20 or 22 carbon atoms in the fatty acid chain comprise up to six double bonds, advantageously three, four, five or six double bonds, especially preferably four, five or six double bonds, very especially preferably five or six. All the abovementioned radicals are derived from the corresponding fatty acids.

The abovementioned radicals of R¹, R² and R³ can be substituted by hydroxyl and/or epoxy groups and/or can comprise triple bonds.

The polyunsaturated fatty acids produced in the process according to the invention advantageously comprise at least two, advantageously three, four, five or six, double bonds. The fatty acids especially advantageously comprise four, five or six double bonds. Fatty acids produced in the process advantageously have 18, 20 or 22 C atoms in the fatty acid chain; the fatty acids preferably comprise 20 or 22 carbon atoms in the fatty acid chain. Saturated fatty acids are advantageously reacted to a minor degree, or not at all, by the nucleic acids used in the process. To a minor degree is to be understood as meaning that the saturated fatty acids are reacted with less than 5% of the activity, advantageously less than 3%, especially advantageously with less than 2%, very especially preferably with less than 1, 0.5, 0.25 or 0.125% of the activity in comparison with polyunsaturated fatty acids. These fatty acids which have been produced can be produced in the process as a single product or be present in a fatty acid mixture.

The nucleic acid sequences used in the process according to the invention take the form of isolated nucleic acid sequences which encode polypeptides with $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 4$ -desaturase activity.

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Nucleic acid sequences which are advantageously used in the process according to the invention are nucleic acid sequences which encode polypeptides with $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase activity selected from the group consisting of:

10

a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201, or

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b) nucleic acid sequences which, as the result of the degeneracy of the genetic code, can be derived from the amino acid sequences shown in SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 70, SEQ ID NO: 72, SEQ ID NO: 74, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID

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NO: 104, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138, SEQ ID NO: 184, SEQ ID NO: 194, SEQ ID NO: 198, SEQ ID NO: 200 or SEQ ID NO: 202, or

5

- c) derivatives of the nucleic acid sequence shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201, which encode polypeptides with at least 40% identity at the amino acid level with SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 70, SEQ ID NO: 72, SEQ ID NO: 74, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138, SEQ ID NO: 184, SEQ ID NO: 194, SEQ ID NO: 198, SEQ ID NO: 200 or SEQ ID NO: 202 and which have a $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase activity.

35

Advantageously, the substituents R^2 or R^3 in the general formulae I and II

independently of one another are saturated or unsaturated C₁₈-C₂₂-alkylcarbonyl; especially advantageously, are independently of one another C₁₈-, C₂₀- or C₂₂-alkylcarbonyl with at least two double bonds, advantageously with at least three, four, five or six double bonds, especially advantageously with at least four, five or six double bonds.

In a preferred embodiment of the process, a nucleic acid sequence which encodes polypeptides with ω 3-desaturase activity, selected from the group consisting of:

- 10 a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 87 or SEQ ID NO: 105, or
- b) nucleic acid sequences which can be derived from the amino acid sequence shown in SEQ ID NO: 88 or SEQ ID NO: 106 as the result of the degeneracy of the genetic code, or
- 15 c) derivatives of the nucleic acid sequence shown in SEQ ID NO: 87 or SEQ ID NO: 105, which encode polypeptides with at least 60% identity at the amino acid level with SEQ ID NO: 88 or SEQ ID NO: 106 and which have ω 3-desaturase activity

is additionally introduced into the transgenic plant.

In a further preferred embodiment of the process, that a nucleic acid sequence which encodes polypeptides with Δ 12-desaturase activity, selected from the group consisting of:

- 25 a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 107, SEQ ID NO: 109 or SEQ ID NO: 195, or
- 30 b) nucleic acid sequences which, as the result of the degeneracy of the genetic code, can be derived from the amino acid sequence shown in SEQ ID NO: 108, SEQ ID NO: 110 or SEQ ID NO: 196, or
- 35 c) derivatives of the nucleic acid sequence shown in SEQ ID NO: 107, SEQ ID NO: 109 or SEQ ID NO: 195, which encode polypeptides with at least 60% at the amino acid level with SEQ ID NO: 108, SEQ ID NO: 110 or SEQ ID NO: 196 and which have Δ 12-desaturase activity

is additionally introduced into the transgenic plant.

- These abovementioned $\Delta 12$ -desaturase sequences can be used alone or in
 5 combination with $\omega 3$ -desaturase sequences together with the nucleic acid sequences
 used in the process which encode $\Delta 9$ -elongases, $\Delta 6$ -desaturases, $\Delta 8$ -desaturases,
 $\Delta 6$ -elongases, $\Delta 5$ -desaturases, $\Delta 5$ -elongases or $\Delta 4$ -desaturases.

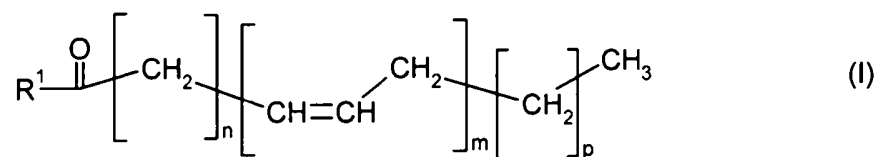
10 Table 1 shows the nucleic acid sequences, the organism of origin and the sequence ID
 number.

No.	Organism	Activity	Sequence number
1.	<i>Euglena gracilis</i>	$\Delta 8$ -Desaturase	SEQ ID NO: 1
2.	<i>Isochrysis galbana</i>	$\Delta 9$ -Elongase	SEQ ID NO: 3
3.	<i>Phaeodactylum tricornutum</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 5
4.	<i>Ceratodon purpureus</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 7
5.	<i>Physcomitrella patens</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 9
6.	<i>Thraustochytrium</i> sp.	$\Delta 5$ -Desaturase	SEQ ID NO: 11
7.	<i>Mortierella alpina</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 13
8.	<i>Caenorhabditis elegans</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 15
9.	<i>Borago officinalis</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 17
10.	<i>Ceratodon purpureus</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 19
11.	<i>Phaeodactylum tricornutum</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 21
12.	<i>Physcomitrella patens</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 23
13.	<i>Caenorhabditis elegans</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 25
14.	<i>Physcomitrella patens</i>	$\Delta 6$ -Elongase	SEQ ID NO: 27
15.	<i>Thraustochytrium</i> sp.	$\Delta 6$ -Elongase	SEQ ID NO: 29
16.	<i>Phytophthora infestans</i>	$\Delta 6$ -Elongase	SEQ ID NO: 31
17.	<i>Mortierella alpina</i>	$\Delta 6$ -Elongase	SEQ ID NO: 33
18.	<i>Mortierella alpina</i>	$\Delta 6$ -Elongase	SEQ ID NO: 35
19.	<i>Caenorhabditis elegans</i>	$\Delta 6$ -Elongase	SEQ ID NO: 37
20.	<i>Euglena gracilis</i>	$\Delta 4$ -Desaturase	SEQ ID NO: 39
21.	<i>Thraustochytrium</i> sp.	$\Delta 4$ -Desaturase	SEQ ID NO: 41

No.	Organism	Activity	Sequence number
22.	<i>Thalassiosira pseudonana</i>	$\Delta 5$ -Elongase	SEQ ID NO: 43
23.	<i>Thalassiosira pseudonana</i>	$\Delta 6$ -Elongase	SEQ ID NO: 45
24.	<i>Cryptocodinium cohnii</i>	$\Delta 5$ -Elongase	SEQ ID NO: 47
25.	<i>Cryptocodinium cohnii</i>	$\Delta 5$ -Elongase	SEQ ID NO: 49
26.	<i>Oncorhynchus mykiss</i>	$\Delta 5$ -Elongase	SEQ ID NO: 51
27.	<i>Oncorhynchus mykiss</i>	$\Delta 5$ -Elongase	SEQ ID NO: 53
28.	<i>Thalassiosira pseudonana</i>	$\Delta 5$ -Elongase	SEQ ID NO: 59
29.	<i>Thalassiosira pseudonana</i>	$\Delta 5$ -Elongase	SEQ ID NO: 61
30.	<i>Thalassiosira pseudonana</i>	$\Delta 5$ -Elongase	SEQ ID NO: 63
31.	<i>Thraustochytrium aureum</i>	$\Delta 5$ -Elongase	SEQ ID NO: 65
32.	<i>Ostreococcus tauri</i>	$\Delta 5$ -Elongase	SEQ ID NO: 67
33.	<i>Ostreococcus tauri</i>	$\Delta 6$ -Elongase	SEQ ID NO: 69
34.	<i>Primula farinosa</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 71
35.	<i>Primula vialii</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 73
36.	<i>Ostreococcus tauri</i>	$\Delta 5$ -Elongase	SEQ ID NO: 75
37.	<i>Ostreococcus tauri</i>	$\Delta 5$ -Elongase	SEQ ID NO: 77
38.	<i>Ostreococcus tauri</i>	$\Delta 5$ -Elongase	SEQ ID NO: 79
39.	<i>Ostreococcus tauri</i>	$\Delta 6$ -Elongase	SEQ ID NO: 81
40.	<i>Thraustochytrium</i> sp.	$\Delta 5$ -Elongase	SEQ ID NO: 83
41.	<i>Thalassiosira pseudonana</i>	$\Delta 5$ -Elongase	SEQ ID NO: 85
42.	<i>Phytophthora infestans</i>	$\omega 3$ -Desaturase	SEQ ID NO: 87
43.	<i>Ostreococcus tauri</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 89
44.	<i>Ostreococcus tauri</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 91
45.	<i>Ostreococcus tauri</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 93
46.	<i>Ostreococcus tauri</i>	$\Delta 4$ -Desaturase	SEQ ID NO: 95
47.	<i>Thalassiosira pseudonana</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 97
48.	<i>Thalassiosira pseudonana</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 99
49.	<i>Thalassiosira pseudonana</i>	$\Delta 5$ -Desaturase	SEQ ID NO: 101
50.	<i>Thalassiosira pseudonana</i>	$\Delta 4$ -Desaturase	SEQ ID NO: 103
51.	<i>Thalassiosira pseudonana</i>	$\omega 3$ -Desaturase	SEQ ID NO: 105
52.	<i>Ostreococcus tauri</i>	$\Delta 12$ -Desaturase	SEQ ID NO: 107

No.	Organism	Activity	Sequence number
53.	<i>Thalassiosira pseudonana</i>	$\Delta 12$ -Desaturase	SEQ ID NO: 109
54.	<i>Ostreococcus tauri</i>	$\Delta 6$ -Elongase	SEQ ID NO: 111
55.	<i>Ostreococcus tauri</i>	$\Delta 5$ -Elongase	SEQ ID NO: 113
56.	<i>Xenopus laevis</i> (BC044967)	$\Delta 5$ -Elongase	SEQ ID NO: 117
57.	<i>Ciona intestinalis</i> (AK112719)	$\Delta 5$ -Elongase	SEQ ID NO: 119
58.	<i>Euglena gracilis</i>	$\Delta 5$ -Elongase	SEQ ID NO: 131
59.	<i>Euglena gracilis</i>	$\Delta 5$ -Elongase	SEQ ID NO: 133
60.	<i>Arabidopsis thaliana</i>	$\Delta 5$ -Elongase	SEQ ID NO: 135
61.	<i>Arabidopsis thaliana</i>	$\Delta 5$ -Elongase	SEQ ID NO: 137
62.	<i>Phaeodactylum tricornutum</i>	$\Delta 6$ -Elongase	SEQ ID NO: 183
63.	<i>Phytium irregulare</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 193
64.	<i>Calendula officinalis</i>	$\Delta 12$ -Desaturase	SEQ ID NO: 195
65.	<i>Ostreococcus tauri</i>	$\Delta 5$ -Elongase	SEQ ID NO: 197
66.	<i>Ostreococcus tauri</i>	$\Delta 6$ -Elongase	SEQ ID NO: 199
67.	<i>Ostreococcus tauri</i>	$\Delta 6$ -Desaturase	SEQ ID NO: 201

In a further embodiment of the invention, a process to be developed for the production of large amounts of polyunsaturated fatty acids, specifically ARA and EPA, in a transgenic plant. This process is also suitable for the production of DHA. Thus, ARA, EPA, DHA or their mixtures can be produced in the process. A further embodiment of the invention is thus a process for the compounds of the general formula I



in transgenic plants, the process comprising:

- 10 a) introducing, into a plant, at least one nucleic acid sequence which encodes a polypeptide with a $\Delta 6$ -desaturase activity and is selected from the group consisting of:

- 5 i) a nucleic acid with the sequence shown in SEQ ID NO: 193 or SEQ ID NO: 201,
 ii) nucleic acid sequences which encode the amino acid sequence shown in
 SEQ ID NO: 194 or SEQ ID NO: 202,
 iii) nucleic acid sequences which hybridize under stringent conditions with the
 complementary strand of the nucleic acid sequence shown in SEQ ID NO:
 193 or SEQ ID NO: 201, and
 iv) nucleic acid sequences which have at least 60% identity with the sequence
 shown in SEQ ID NO: 193 or SEQ ID NO: 201,
- 10 b) introducing, into a plant, at least one nucleic acid sequence which encodes a
 polypeptide with a $\Delta 6$ -elongase activity and is selected from the group
 consisting of:
- 15 i) a nucleic acid with the sequence shown in SEQ ID NO: 27 or SEQ ID NO:
 199,
 ii) nucleic acid sequences which encode the amino acid sequence shown in
 SEQ ID NO: 28 or SEQ ID NO: 200,
 iii) nucleic acid sequences which hybridize under stringent conditions with the
20 complementary strand of the nucleic acid sequence shown in SEQ ID NO:
 27 or SEQ ID NO: 199, and
 iv) nucleic acid sequences which have at least 60% identity with the sequence
 shown in SEQ ID NO: 27 or SEQ ID NO: 199,
- 25 c) introducing, into a plant, at least one nucleic acid sequence which encodes a
 polypeptide with a $\Delta 5$ -desaturase activity and is selected from the group
 consisting of:
- 30 i) a nucleic acid with the sequence shown in SEQ ID NO: 11,
 ii) nucleic acid sequences which encode the amino acid sequence shown in
 SEQ ID NO: 12,
 iii) nucleic acid sequences which hybridize under stringent conditions with the
 complementary strand of the nucleic acid sequence shown in SEQ ID NO:
 11, and
35 iv) nucleic acid sequences which have at least 60% identity with the sequence
 shown in SEQ ID NO: 11,

where the variables and substituents in the formula I have the meaning given above.

The nucleic acid sequences which can be used in the process are described in WO 02/26946 ($\Delta 5$ -desaturase from *Thraustochytrium* ssp., SEQ ID NO: 11 and $\Delta 6$ -desaturase from *Phytium irregulare*, SEQ ID NO: 193) and in WO 01/59128 ($\Delta 6$ -elongase from *Physcomitrella patens*, SEQ ID NO: 27), which is expressly referred to here. However, in these cases, the formation of ARA and EPA was studied either not in transgenic plants, but only in microorganisms, or else no increase ARA and EPA synthesis was detected in the transgenic plants. Moreover, the nucleic acids according to the invention were not combined, in these applications, with nucleic acids which encode other enzymes of the fatty acid biosynthetic pathway.

Surprisingly, it has now been found that the coexpression of the nucleic acids with the sequences shown in SEQ ID NO: 11, 27, 193, 199 and 201 leads, in transgenic plants, to a greatly increased ARA content to up to more than 8%, advantageously up to more than 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19% or 20%, especially advantageously to more than 21%, 22%, 23%, 24% or 25%, based on the total lipid content of the plant (cf. Table 2, Table 3, Table 4 and Figure 31). The abovementioned percentages are percent by weight.

To further increase the yields in the process described for the production of oils and/or triglycerides with a content of polyunsaturated fatty acids, especially ARA, EPA or DHA or their mixtures, which is advantageously increased in comparison with oils and/or triglycerides from wild-type plants, it may be advantageous to increase the amount of the starting material for the fatty acid biosynthesis. This can be achieved for example by introducing a nucleic acid which encodes a polypeptide with the activity of a $\Delta 12$ -desaturase, and coexpressing it in the organism.

This is especially advantageously in oil-producing organisms such as the family Brassicaceae, such as the genus *Brassica*, for example oilseed rape, turnip rape or Indian mustard; the family Elaeagnaceae, such as the genus *Elaeagnus*, for the example the genus and species *Olea europaea* or the family Fabaceae, such as the genus *Glycine*, for example the genus and species *Glycine max*, which has a high oleic acid content, but only a low linoleic acid content (Mikoklajczak et al., Journal of the American Oil Chemical Society, 38, 1961, 678-681).

This is why, in a preferred embodiment of the present invention, a nucleic acid sequence which encodes a polypeptide with $\Delta 12$ -desaturase activity is additionally introduced into the transgenic plant.

Especially preferably, this nucleic acid sequence is selected from the group consisting of:

- 5 a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 195,
- b) nucleic acid sequences which encode the amino acid sequence shown in SEQ ID NO: 196,
- 10 c) nucleic acid sequences which hybridize under stringent conditions with the complementary strand of the nucleic acid sequence shown in SEQ ID NO: 195, and
- d) nucleic acid sequences which have at least 60% identity with the sequence shown in SEQ ID NO: 195.

15

The nucleic acid sequence with the SEQ ID NO: 195 is derived from *Calendula officinalis* and described in WO 01/85968, the disclosure of which is likewise incorporated in the present application by reference.

- 20 The $\Delta 12$ -desaturases used in the process according to the invention advantageously convert oleic acid (C18:1 ^{$\Delta 9$}) into linoleic acid (C18:2 ^{$\Delta 9,12$}) or C18:2 ^{$\Delta 6,9$} into C18:3 ^{$\Delta 6,9,12$} (gamma-linolenic acid = GLA), the starting materials for the synthesis of ARA, EPA and DHA. The $\Delta 12$ -desaturases advantageously convert fatty acids bound to phospholipids or CoA-fatty acid esters, advantageously bound to CoA-fatty acid esters. If an
- 25 elongation step has taken place beforehand, this advantageously leads to higher yields of synthetic products since, as a rule, elongation takes place at CoA-fatty acid esters, while desaturation predominantly takes place at the phospholipid or at the triglycerides. An exchange between the CoA-fatty acid esters and the phospholipids or triglycerides, which would require a further, potentially limiting, enzyme reaction, is thus not required.

30

The additional expression of the $\Delta 12$ -desaturase in the transgenic plants leads to a further increase in the ARA content up to more than 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19% and 20%, especially advantageously to more than 21%, 22%, 23%, 24% or 25%, based on the total lipid content of the plant (cf. Tables 3 and 4 and

35 Figure 32). The abovementioned percentages are percent by weight.

Further nucleic acid sequences which encode a polypeptide with a $\Delta 5$ -elongase activity can advantageously be introduced into the plants in the process according to the

invention.

Preference is given to those nucleic acid sequences which encode a $\Delta 5$ -elongase activity is chosen from the group consisting of:

5

a) a nucleic acid sequence was the sequence shown in SEQ ID NO: 43, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137 or SEQ ID NO: 197,

10

b) nucleic acid sequences which encode the amino acid sequence shown in SEQ ID NO: 44, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 86, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138 or SEQ ID NO: 198,

15

c) nucleic acid sequences which hybridize under stringent conditions with the complementary strand of the nucleic acid sequence shown in SEQ ID NO: 43, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137 or SEQ ID NO: 197, and

20

d) nucleic acid sequences which have at least 60% identity with the sequence shown in SEQ ID NO: 43, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137 or SEQ ID NO: 197.

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In a preferred embodiment of the process, the $\Delta 5$ -elongase genes are expressed under the control of a seed-specific promoter.

In a further advantageous embodiment of the process, all nucleic acid sequences are introduced into the plants on a shared recombinant nucleic acid molecule, it being possible for each nucleic acid sequence to be under the control of its own promoter and it being possible for this own promoter to take the form of a seed-specific promoter.

5

However, it is not only the nucleic acids detailed in the sequence listing which can successfully be employed in the invention to carry out the conversion; rather, even sequences which deviate to a certain degree from these sequences and which encode proteins with the essentially identical enzymatic activity can be employed. These take the form of nucleic acids which have a certain degree of identity or homology with the sequences specified in the sequence listing. An essentially identical enzymatic activity denotes proteins which have at least 20%, 30%, 40%, 50% or 60%, advantageously at least 70%, 80%, 90% or 95%, especially advantageously at least 96%, 97%, 98% or 99% of the enzymatic activity of the wild-type enzymes.

15

In order to determine the percentage of homology (= identity) of two amino acid sequences or of two nucleic acids, the sequences are written one under the other (for example, gaps may be introduced into the sequence of a protein or of a nucleic acid in order to generate optimal alignment with the other protein or the other nucleic acid). Then, the amino acid radicals or nucleotides at the corresponding amino acid positions or nucleotide positions are compared. If a position in a sequence is occupied by the same amino acid radical or the same nucleotide as the corresponding position in the other sequence, then the molecules are homologous at this position (i.e. amino acid or nucleic acid "homology" as used in the present context corresponds to amino acid or nucleic acid "identity"). The percentage of homology between the two sequences is a function of the number of positions which the sequences share (i.e. % homology = number of identical positions/total number of positions x 100). The terms homology and identity are therefore to be considered as synonymous.

30

The homology was calculated over the entire amino acid or nucleic acid sequence region. To compare various sequences, the skilled worker has available a series of programs which are based on various algorithms. The algorithms of Needleman and Wunsch or Smith and Waterman give particularly reliable results. The program PileUp (J. Mol. Evolution., 25, 351-360, 1987, Higgins et al., CABIOS, 5 1989: 151-153) or the programs Gap and BestFit [Needleman and Wunsch (J. Mol. Biol. 48; 443-453 (1970) and Smith and Waterman (Adv. Appl. Math. 2; 482-489 (1981))], which are part of the GCG software packet [Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA 53711 (1991)], were used to carry out the sequence comparisons. The

35

sequence homology data given above in percent were determined over the entire sequence region using the program GAP with the following settings: Gap Weight: 50, Length Weight: 3, Average Match: 10.000 and Average Mismatch: 0.000. Unless otherwise specified, these settings were always used as standard settings for sequence comparisons.

The skilled worker will recognize that DNA sequence polymorphisms which lead to modifications of the amino acid sequence of SEQ ID NO: 12, 28, 194, 196, 198, 200 and/or 202 may occur within a population. These natural variants usually cause a variance of from 1 to 5% in the nucleotide sequence of the $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 6$ -elongase gene. The scope of the invention is to comprise each and all of these nucleotide variation(s) and resulting amino acid polymorphisms in the $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 6$ -elongase which are the result of natural variation and which do not essentially modify the enzymatic activity.

Essential enzymatic activity of the $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -elongase or $\Delta 5$ -desaturase used in the process according to the invention is understood as meaning that they retain an enzymatic activity of at least 10%, preferably of at least 20%, especially preferably of at least 30%, 40%, 50% or at least 60% and most preferably at least 70%, 80%, 90%, 95%, 96%, 97%, 98% or 99% in comparison with the proteins/enzymes encoded by the sequence and its derivatives and that they are thus capable of participating in the metabolism of compounds which are required for the synthesis of fatty acids, fatty acid esters such as diacylglycerides and/or triacylglycerides in a plant or plant cell or in the transport of molecules across membranes, meaning C_{18} -, C_{20} - or C_{22} -carbon chains in the fatty acid molecule with double bonds at at least two, advantageously three, four or five, positions.

Likewise, the scope of the invention comprises nucleic acid molecules which hybridize under stringent conditions with the complementary strand of the $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 6$ -elongase nucleic acids used. The term "hybridizes under stringent conditions" as used in the present context is to describe hybridization and washing conditions under which nucleotide sequences with at least 60% homology to one another usually remain hybridized with one another. Conditions are preferably such that sequences with at least approximately 65%, 70%, 80% or 90%, preferably at least approximately 91%, 92%, 93%, 94% or 95%, and especially preferably at least approximately 96%, 97%, 98%, 99% or more homology to one another usually remain hybridized to one another. These stringent conditions are

known to the skilled worker and described, for example, in Current Protocols in Molecular Biology, John Wiley & Sons, N. Y. (1989), 6.3.1-6.3.6.

5 A preferred, nonlimiting, example of stringent hybridization conditions is hybridizations in 6 x sodium chloride/sodium citrate (= SSC) at approximately 45°C, followed by one or more washing steps in 0.2 x SSC, 0.1% SDS at 50 to 65°C. The skilled worker knows that these hybridization conditions differ depending on the type of nucleic acid and, for example when organic solvents are present, regarding temperature and buffer concentration. Under "standard hybridization conditions", for example, the hybridization
10 temperature is, depending on the type of nucleic acid, between 42°C and 58°C in aqueous buffer with a concentration of 0.1 to 5 x SSC (pH 7.2). If organic solvents, for example 50% formamide, are present in the abovementioned buffer, the temperature under standard conditions is approximately 42°C. Preferably the hybridization conditions for DNA:DNA hybrids, for example, are 0.1 x SSC and 20°C to 45°C,
15 preferably 30°C to 45°C. Preferably the hybridization conditions for DNA:RNA hybrids are, for example, 0.1 x SSC and 30°C to 55°C, preferably 45°C to 55°C. The abovementioned hybridization temperatures are determined for a nucleic acid with approximately 100 bp (= base pairs) in length and with a G + C content of 50% in the absence of formamide. The skilled worker knows how to determine the required
20 hybridization conditions on the basis of textbooks such as Sambrook et al., "Molecular Cloning", Cold Spring Harbor Laboratory, 1989; Hames and Higgins (Eds.) 1985, "Nucleic Acids Hybridization: A Practical Approach", IRL Press at Oxford University Press, Oxford; Brown (Ed.) 1991, "Essential Molecular Biology: A Practical Approach", IRL Press at Oxford University Press, Oxford.

25

By introducing one or more nucleotide substitutions, additions or deletions into a nucleotide sequence, it is possible to generate an isolated nucleic acid molecule which encodes a $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 6$ -elongase with one or more amino acid substitutions, additions or deletions. Mutations
30 can be introduced into one of the sequences by means of standard techniques, such as site-specific mutagenesis and PCR-mediated mutagenesis. It is preferred to generate conservative amino acid substitutions in one or more of the above nonessential amino acid radicals. In a "conservative amino acid substitution", the amino acid radical is replaced by an amino acid radical with a similar side chain. Families of
35 amino acid radicals with similar side chains have been defined in the art. These families comprise amino acids with basic side chains (for example lysine, arginine, histidine), acidic side chains (for example aspartic acid, glutamic acid), uncharged polar side chains (for example glycine, asparagine, glutamine, serine, threonine, tyrosine,

- cysteine), unpolar side chains (for example alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (for example threonine, valine, isoleucine) and aromatic side chains (for example tyrosine, phenylalanine, tryptophan, histidine). A predicted nonessential amino acid radical in a
- 5 $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 6$ -elongase is thus preferably replaced by another amino acid radical from the same family of side chains. In another embodiment, the mutations can, alternatively, be introduced randomly over all or part of the sequence encoding the $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 6$ -elongase, for example by saturation mutagenesis, and
- 10 the resulting mutants can be screened by recombinant expression for the hereindescribed $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 6$ -elongase activity in order to identify mutants which have retained the $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 6$ -elongase activity.
- 15 The polyunsaturated fatty acids produced in the process according to the invention advantageously comprise at least two, preferably three, four, five or six, double bonds. The fatty acids especially preferably comprise four, five or six double bonds. Fatty acids produced in the process preferably have a length of 20 C or 22 C atoms.
- 20 Saturated fatty acids are preferably reacted to a minor degree with the nucleic acids used in the process, or not at all. "A minor degree" is understood as meaning that, in comparison with polyunsaturated fatty acids, the saturated fatty acids are reacted with less than 5%, preferably with less than 3%, especially preferably with less than 2%, most preferably with less than 1, 0.5, 0.25 or 0.125% of the activity. The fatty acids
- 25 produced may constitute the only product of the process or else may be present in a fatty acid mixture.
- The polyunsaturated fatty acids produced in the process are advantageously bound in membrane lipids and/or triacylglycerides, but may also occur in the organisms as free
- 30 fatty acids or else bound in the form of other fatty acid esters. In this context, they may be present as "pure products" or else advantageously in the form of mixtures of various fatty acids or mixtures of different glycerides. The various fatty acids which are bound in the triacylglycerides can be derived from short-chain fatty acids with 4 to 6 C atoms, medium-chain fatty acids with 8 to 12 C atoms or long-chain fatty acids with 14 to 24 C
- 35 atoms, preferred are the long-chain fatty acids, especially preferred are the long-chain fatty acids LCPUFAs of C_{18} -, C_{20} - and/or C_{22} -fatty acids, very especially preferred are the long-chain fatty acids LCPUFAs of C_{20} - and/or C_{22} -fatty acids such as ARA, EPA, DHA or their combination.

The process according to the invention advantageously yields fatty acid esters with polyunsaturated C₁₈-, C₂₀- and/or C₂₂-fatty acid molecules with at least two double bonds in the fatty acid ester, advantageously with at least three, four, five or six double bonds in the fatty acid ester, especially advantageously four, five or six double bonds in the fatty acid ester, very especially advantageously at least five or six double bonds in the fatty acid ester. This advantageously leads to the synthesis of linoleic acid (=LA, C₁₈:2^{Δ^{9,12}}), γ-linolenic acid (= GLA, C₁₈:3^{Δ^{6,9,12}}), stearidonic acid (= SDA, C₁₈:4^{Δ^{6,9,12,15}}), dihomo-γ-linolenic acid (= DGLA, C₂₀:3^{Δ^{8,11,14}}), ω3-eicosatetraenoic acid (= ETA, C₂₀:4^{Δ^{5,8,11,14}}), arachidonic acid (ARA, C₂₀:4^{Δ^{5,8,11,14}}), eicosapentaenoic acid (EPA, C₂₀:4^{Δ^{5,8,11,14}}) or mixtures of these, ω3-eicosapentaenoic acid (= ETA, C₂₀:4^{Δ^{5,8,11,14,17}}), arachidonic acid (ARA, C₂₀:4^{Δ^{5,8,11,14}}), eicosapentaenoic acid (EPA, C₂₀:5^{Δ^{5,8,11,14,17}}), ω6-docosapentaenoic acid (C₂₂:5^{Δ^{4,7,10,13,16}}), ω6-docosapentaenoic acid (C₂₂:4^{Δ^{7,10,13,16}}), ω3-docosapentaenoic acid (=DPA, C₂₂:5^{Δ^{7,10,13,16,19}}), docosahexaenoic acid (= DHA, C₂₂:6^{Δ^{4,7,10,13,16,19}}) or their mixtures are preferably produced, and ARA, EPA and/or DHA are very especially produced. ω3-Fatty acids such as EPA and/or DHA, preferably DHA, are advantageously produced.

The fatty acid esters with polyunsaturated C₁₈-, C₂₀- and/or C₂₂-fatty acid molecules, advantageously with polyunsaturated C₂₀- and/or C₂₂-fatty acid molecules, can be isolated in the form of an oil or lipid, for example in the form of compounds such as sphingolipids, phosphoglycerides, lipids, glycolipids such as glycosphingolipids, phospholipids such as phosphatidylethanolamine, phosphatidylcholine, phosphatidylserine, phosphatidylglycerol, phosphatidylinositol or diphosphatidylglycerol, monoacylglycerides, diacylglycerides, triacylglycerides or other fatty acid esters such as the acetyl-coenzyme A esters which comprise the polyunsaturated fatty acids with at least two, three, four, five or six, preferably four, five or six, especially preferably five or six, double bonds, from the plants which were used for the preparation of the fatty acid esters. Preferably, they are isolated in the form of their diacylglycerides, triacylglycerides and/or in the form of phosphatidylcholine, especially preferably in the form of the triacylglycerides. In addition to these esters, the polyunsaturated fatty acids are also present in the plants as free fatty acids or bound in other compounds. As a rule, the various abovementioned compounds (fatty acid esters and free fatty acids) are present in the organisms with an approximate distribution of 80 to 90% by weight of triglycerides, 2 to 5% by weight of diglycerides, 5 to 10% by weight of monoglycerides, 1 to 5% by weight of free fatty acids, 2 to 8% by weight of phospholipids, the total of the various compounds amounting to 100% by weight.

In the method(s) according to the invention (for the purposes of the invention and the disclosure shown herein, the singular is to comprise the plural and vice versa), the LCPUFAs produced are produced in a content of at least 3, 5, 6, 7 or 8% by weight, advantageously at least 9, 10, 11, 12, 13, 14 or 15% by weight, preferably at least 16, 17, 18, 19 or 20% by weight, especially preferably at least 21, 22, 23, 24 or 25% by weight, very especially preferably at least 26, 27, 28, 29 or 30% by weight based on the total fatty acids in the transgenic organisms, advantageously in the seeds of the transgenic plants. Here, C₁₈- and/or C₂₀-fatty acids which are present in the host organisms are advantageously converted into the corresponding products such as ARA, EPA, DPA or DHA, to mention but a few by way of example, at the rate of at least 10%, advantageously at least 20%, especially advantageously at least 30%, very especially advantageously at least 40%. The fatty acids are advantageously produced in bound form.

Polyunsaturated C₂₀-fatty acids with four or five double bonds in the molecule are advantageously produced in the process in a content of all such fatty acids together of at least 15, 16, 17, 18, 19, or 20% by weight, advantageously at least 21, 22, 23, 24 or 25% by weight, especially advantageously at least 26, 27, 28, 29 or 30% by weight based on the total fatty acids in the seeds of the transgenic plants.

Polyunsaturated C₂₀- and/or C₂₂-fatty acids with four, five or six double bonds in the molecule are advantageously produced in the process in a content of all such fatty acids together of at least 15, 16, 17, 18, 19, or 20% by weight, advantageously at least 21, 22, 23, 24 or 25% by weight, especially advantageously at least 26, 27, 28, 29 or 30% by weight, very especially advantageously at least 31, 32, 33, 34 or 35% by weight based on the total fatty acids in the seeds of the transgenic plants.

ARA is produced in the process according to the invention in a content of at least 3, 5, 6, 7, 8, 9 or 10% by weight, advantageously at least 11, 12, 13, 14 or 15% by weight, preferably at least 16, 17, 18, 19 or 20% by weight, especially preferably at least 21, 22, 23, 24 or 25% by weight, most preferably at least 26% by weight, based on the total lipid content in the seeds of the transgenic plants.

EPA is produced in the process according to the invention in a content of at least 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9 or 1% by weight, advantageously at least 2, 3, 4 or 5% by weight, preferably at least 6, 7, 8, 9 or 10% by weight, especially preferably at least 11, 12, 13, 14 or 15% by weight and most preferably at least 16% by weight, based on the total lipid content in the seeds of transgenic plants.

DHA is produced in the process according to the invention in a content of at least 0.01 or 0.02% by weight, advantageously at least 0.03 or 0.05% by weight, advantageously at least 0.09 or 0.1% by weight, especially preferably at least 0.2 or 0.3% by weight and most preferably at least 0.35% by weight, based on the total lipid content in the seeds of the transgenic plants.

It is possible, with the aid of the nucleic acids used in the process according to the invention, for these unsaturated fatty acids to be positioned at the sn1, sn2 and/or sn3 position of the triglycerides which have advantageously been produced. Since in the process according to the invention the starting compounds linoleic acid (C18:2) and linolenic acid (C18:3) pass through a plurality of reaction steps, the end product of the process, such as, for example, arachidonic acid (ARA), eicosapentaenoic acid (EPA), ω 6-docosapentaenoic acid or DHA, are not obtained as absolutely pure products, small traces of the precursors are also always present in the end product. If, for example, both linoleic acid and linolenic acid are present in the starting organism, or the starting plants, the end product, such as ARA, EPA or DHA, are present as mixtures. It is advantageous that, in the end product ARA or DHA, only minor amounts of the in each case other end product should be present. This is why, in a DHA-comprising lipid and/or oil, less than 15, 14, 13, 12 or 11% by weight, advantageously less than 10, 9, 8, 7, 6 or 5% by weight, especially advantageously less than 4, 3, 2 or 1% by weight, of EPA and/or ARA should be present. This is why, in a EPA-comprising lipid and/or oil, less than 15, 14, 13, 12 or 11% by weight, advantageously less than 10, 9, 8, 7, 6 or 5% by weight, especially advantageously less than 4, 3, 2 or 1% by weight, of ARA should be present. This is also why less than 15, 14, 13, 12 or 11% by weight, advantageously less than 10, 9, 8, 7, 6 or 5% by weight, especially advantageously less than 4, 3, 2 or 1% by weight of EPA and/or DHA should be present in an ARA-comprising lipid and/or oil.

However, mixtures of different polyunsaturated C₂₀- and/or C₂₂-fatty acids in one product may also be desirable. In such cases, DHA-comprising lipids and/or oils may comprise at least 1, 2, 3, 4 or 5% by weight of ARA and/or EPA, advantageously at least 6, 7 or 8% by weight, especially advantageously at least 9, 10, 11, 12, 13, 14 or 15% by weight, very especially advantageously at least 16, 17, 18, 19, 20, 21, 22, 23, 24 or 25% by weight, based on the total lipid content in the seeds of the transgenic plants.

The precursors should advantageously not amount to more than 20% by weight, preferably not to more than 15% by weight, especially preferably not to more than 10% by weight, very especially preferably not to more than 5% by weight, based on the amount of the end product in question. Advantageously, only ARA, EPA or only DHA, bound or as free acids, are produced as end products in the process of the invention in a transgenic plant. If the compounds ARA, EPA and DHA are produced simultaneously, they are advantageously produced in a ratio of at least 1:1:2 (EPA:ARA:DHA), advantageously at least 1:1:3, preferably 1:1:4, especially preferably 1:1:5. If the compounds ARA and EPA are produced simultaneously, they are advantageously produced, in the plant, in a ratio of at least 1:6 (EPA:ARA), advantageously of at least 1:8, preferably of at least 1:10, especially preferably of at least 1:12.

Fatty acid esters or fatty acid mixtures produced by the process according to the invention advantageously comprise 6 to 15% of palmitic acid, 1 to 6% of stearic acid, 7-85% of oleic acid, 0.5 to 8% of vaccenic acid, 0.1 to 1% of arachic acid, 7 to 25% of saturated fatty acids, 8 to 85% of monounsaturated fatty acids and 60 to 85% of polyunsaturated fatty acids, in each case based on 100% and on the total fatty acid content of the organisms.

Moreover, the fatty acid esters or fatty acid mixtures which have been produced by the process of the invention advantageously comprise fatty acids selected from the group of the fatty acids erucic acid (13-docosaenoic acid), sterculic acid (9,10-methyleneoctadec-9-enoic acid), malvalic acid (8,9-methyleneheptadec-8-enoic acid), chaulmoogric acid (cyclopentenododecanoic acid), furan fatty acid (9,12-epoxyoctadeca-9,11-dienoic acid), vernolic acid (9,10-epoxyoctadec-12-enoic acid), tariric acid (6-octadecynoic acid), 6-nonadecynoic acid, santalbic acid (11-octadecen-9-ynoic acid), 6,9-octadecenynoic acid, pyrulic acid (10-heptadecen-8-ynoic acid), crepenyninic acid (9-octadecen-12-ynoic acid), 13,14-dihydrooropoeic acid, octadecen-13-ene-9,11-diynoic acid, petroselenic acid (cis-6-octadecenoic acid), 9c,12t-octadecadienoic acid, calendulic acid (8t10t12c-octadecatrienoic acid), catalpic acid (9t11t13c-octadecatrienoic acid), eleostearic acid (9c11t13t-octadecatrienoic acid), jacaric acid (8c10t12c-octadecatrienoic acid), punიცic acid (9c11t13c-octadecatrienoic acid), parinaric acid (9c11t13t15c-octadecatetraenoic acid), pinolenic acid (all-cis-5,9,12-octadecatrienoic acid), laballenic acid (5,6-octadecadienallenic acid), ricinoleic acid (12-hydroxyoleic acid) and/or coriolic acid (13-hydroxy-9c,11t-octadecadienoic acid). The abovementioned fatty acids are, as a rule, advantageously only found in traces in the fatty acid esters or fatty acid mixtures produced by the process according

to the invention, that is to say that, based on the total fatty acids, they occur to less than 30%, preferably to less than 25%, 24%, 23%, 22% or 21%, especially preferably to less than 20%, 15%, 10%, 9%, 8%, 7%, 6% or 5%, very especially preferably to less than 4%, 3%, 2% or 1%. In a further preferred form of the invention, these

5 abovementioned fatty acids occur to less than 0.9%, 0.8%, 0.7%, 0.6% or 0.5%, especially preferably to less than 0.4%, 0.3%, 0.2%, 0.1%, based on the total fatty acids. The fatty acid esters or fatty acid mixtures produced by the process according to the invention advantageously comprise less than 0.1%, based on the total fatty acids, or no butyric acid, no cholesterol, no clupanodonic acid (= docosapentaenoic acid,

10 C22:5^{Δ4,8,12,15,21}) and no nisinic acid (tetracosahexaenoic acid, C23:6^{Δ3,8,12,15,18,21}).

Owing to the nucleic acid sequences according to the invention or nucleic acid sequences used in the process according to the invention, an increase in the yield of polyunsaturated fatty acids, mainly ARA and EPA, but also DHA, of at least 50, 80 or

15 100%, advantageously at least 150, 200 or 250%, especially advantageously at least 300, 400, 500, 600, 700, 800 or 900%, very especially advantageously at least 1000, 1100, 1200, 1300, 1400 or 1500% in comparison with the nontransgenic starting plant, for example a plant such as *Brassica juncea*, *Brassica napus*, *Camelina sativa*, *Arabidopsis thaliana* or *Linum usitatissimum* when compared by means of GC

20 analysis; see Examples.

Advantageously, as described above, the polyunsaturated C₂₀- and/or C₂₂-fatty acids with four, five or six double bonds in the molecule, which are produced in the process, will comprise in the seeds of plants which comprise only very small amounts of C12:0-

25 or C14:0-fatty acids, or none at all. Even shorter saturated fatty acids, such as the fatty acids C4:0, C6:0, C8:0 or C10:0 should not be present in the lipid and/or oil or only in very small amounts. Only very small amounts are advantageously understood as amounts which, in GC analysis, are advantageously under 5, 4, 3, 2 or 1%, advantageously under 0.9, 0.8, 0.7, 0.6 or 0.5%, especially advantageously under 0.4,

30 0.3, 0.2 or 0.1%, very especially preferably under 0.09, 0.08, 0.07, 0.06, 0.05, 0.04, 0.03, 0.02 or 0.01 units area in the GC. The fatty acid C16:0 should advantageously be in a range of from 1 to 28% GC units area. The fatty acid C16:0 should advantageously be present in GC units area in amounts of less than 25%, 20%, 15% or 10%, advantageously less than 9%, 8%, 7%, 6% or 5%, especially advantageously less than

35 4%, 3%, 2% or 1% or not at all, in the lipids, oils and/or free fatty acids. The fatty acid C16:1 should advantageously amount to less than 1, 0.5, 0.4, 0.3, 0.2 or 0.1%, especially advantageously 0.09, 0.08, 0.07, 0.06, 0.05, 0.04, 0.03, 0.02 or 0.01 units area in the GC. Very especially preferably, the fatty acid C16:1 should not be present in

the oils and/or lipids produced by the process. The same applies to the fatty acids C15:0, C17:0, C16:1^{Δ3}trans, C16:4^{Δ4,7,10,13} and C18:5^{Δ3,6,9,12,15}. Besides oleic acid (C18:1^{Δ9}), the isomers (C18:1^{Δ7}, 18:1^{Δ11}) may also be present in the lipids, oils or free fatty acids. Advantageously in amounts of less than 5%, 4%, 3%, 2% or 1%, measured as units GC area. The fatty acids C20:0, C20:1, C24:0 and C24:1 should in each case be in the range of from 0 to 1%, 0 to 3% and 0 to 5%, respectively, units GC area. Furthermore, little dihomo-γ-linolenic acid (= DGLA) should be detectable in the GC analysis in units GC area in the seed oil and/or seed lipid. Little is understood as meaning less than 2, 1.9, 1.8, 1.7, 1.6 or 1.5%, advantageously less than 1.4, 1.3, 1.2, 1.1 or 1%, especially advantageously less than 0.9, 0.8, 0.7, 0.6, 0.5 or 0.4% in units GC area.

In a preferred embodiment of the process, DGLA and ARA should be produced in a ratio of from 1:1 up to 1:100, advantageously from 1:2 up to 1:80, especially advantageously from 1:3 up to 1:70, very especially from 1:5 up to 1:60.

In a further preferred embodiment, DGLA and EPA should be produced in a ratio of from 1:1 up to 1:100, advantageously from 1:2 up to 1:80, especially advantageously from 1:3 up to 1:70, very especially from 1:5 up to 1:60.

The lipids and/or oils produced in the process according to the invention should advantageously have a high unsaturated, advantageously polyunsaturated, fatty acid content of at least 30, 40 or 50% by weight, advantageously at least 60, 70 or 80% by weight, based on the total fatty acid content in the seeds of the transgenic plants.

All saturated fatty acids together should advantageously only amount to a small quantity in the plants preferably used in the process according to the invention. In this context, a small amount is understood as meaning an amount of less than 15%, 14%, 13%, 12%, 11% or 10%, preferably less than 9%, 8%, 7% or 6%, in units GC area.

Furthermore, the genes for the synthesis of the polyunsaturated fatty acids, which are used in the process and which have been introduced, in the process, via different processes, advantageously as host plant, should advantageously have a higher oil content than protein content in the seed, advantageous plants have an oil/protein content ratio of from 5:1, 4:1, 3:1, 2:1 or 1:1. In this context, the oil content based on the total weight of the seed should be in a range of 15-55%, advantageously between 25-50%, especially advantageously between 35-50%. Advantageous host plants used in the process should have a distribution of the unsaturated fatty acids such as oleic

acid, linoleic acid and linolenic acid, which are the starting compounds in the process according to the invention for the synthesis of polyunsaturated fatty acids, in the sn1, sn2 and sn3 position of the triglyceride, as shown in Table 5 hereinbelow, where rows No. 1-7 represent different advantageous alternatives of such distributions. n.p. means not present.

Table 5: Plants with advantageous fatty acid distribution in the sn1, sn2 and sn3 position on the triglyceride

No.	Oleic acid			Linoleic acid			α -Linolenic acid		
	sn1	sn2	sn3	sn1	sn2	sn3	sn1	sn2	sn3
1.	1	1	1	2	4	1	n.p.	n.p.	n.p.
2.	1.4	2.2	1	2.8	9	1	2	6.7	1
3.	0.8	0.8	1	1.1	1.6	1	1	0.8	1
4.	0.9	0.9	1	1.2	1.6	1	0.9	1	1
5.	0.9	0.9	1	1	1.3	1	1	1	1
6.	1	1.1	1	2	2.8	1	1	1	n.p.
7.	1.3	9.7	1	1	9	traces	1	n.p.	n.p.

The rows show the ratios of the following plants: row 1 = *Arachis hypogaea*, row 2 = *Brassica napus*, row 3 = *Glycine max*, row 4 = *Linum usitatissimum*, row 5 = *Zea mays*, row 6 = *Olea europaea* and row 7 = *Theobroma cacao*.

- Host plants which are advantageous for the process are those which have a high oleic acid content, that means at least 40, 50, 60 or 70% by weight based on the total fatty acid content of the plant, in comparison with linoleic acid and/or linolenic acid in the lipids and/or oils, especially in the triglyceride, such as, for example, *Anacardium occidentale*, *Argania spinosa*, *Bombax malabaricum*, *Brassica napus*, *Butyrospermum parkii*, high-oleic safflower (*Carthamus tinctorius*), *Citrullus colocythis*, *Corylus avellana*, *Curcubita foetidissima*, *Curcubita pepo*, *Guizotia abyssinica*, high-oleic sunflower (*Helianthus annuus*), *Macadamia integrifolia*, *Nigella sativa*, *Olea europaea*, *Papaver somniferum*, *Passiflora edulis*, *Persea americana*, *Prunus amygdalis*, *Prunus armeniaca*, *Prunus dulcis*, *Prunus communis*, *Sesamum indicum*, *Simarouba glauca*, *Thea sasungua*, or *Theobroma cacao*. Further advantageous plants have a higher content of the unsaturated fatty acids oleic acid, linoleic acid and α -linolenic acid in the sn2 position in comparison with the other positions sn1 and sn3. A higher content is understood as meaning ratios of (sn1:sn2:sn3) 1:1.1:1, 1:1.5:1 to 1:3:1. Advantageous

plants such as *Actinidia chinensis*, *Aleurites moluccana*, *Arnebia griffithii*, *Brassica alba*, *Brassica hirta*, *Brassica nigra*, *Brassica juncea*, *Brassica carinata*, *Camelina sativa*, *Cannabis sativa*, *Echium rubrum*, *Echium vulgare*, *Humulus lupulus*, *Juglans regia*, *Linum usitatissimum*, *Ocimum* spp., *Perilla frutescens*, *Portulaca oleracea*,
5 *Prunus cerasus*, *Salicornia bigelovii*, *Salvia hispanica* are also those which have a high α -linolenic acid content in the lipid and/or oil of the plant, that is to say an α -linolenic acid content of at least 10, 15 or 20% by weight, advantageously at least 25, 30, 35, 40, 45 or 50% by weight, based on the total fatty acid content of the plant. Very especially advantageous plants likewise show an advantageous preference for the sn2
10 position over the positions sn1 and sn3 in the triglyceride of from 1:1.1:1, 1:1.5:1 to 1:3:1 for the arachidonic acid, eicosapentaenoic acid or docosahexaenoic acid produced in the process.

Plants used for the process should advantageously have an erucic acid content of less
15 than 2% by weight based on the total fatty acid content of the plant. Also, the content of saturated fatty acids C16:0 and/or C18:0 should advantageously be less than 19, 18, 17, 16, 15, 14, 13, 12, 11 or 10% by weight, advantageously less than 9, 8, 7, 6 or 5% by weight, based on the total fatty acid content of the plant. Also, longer fatty acids such as C20:0 or C22:1 should advantageously not be present, or only in small
20 amounts, advantageously in amounts of less than 4, 3, 2 or 1% by weight, advantageously less than 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2 or 0.1% by weight based on the total fatty acid content of the plant in the plants used in the process. Typically, C16:1 is not present as fatty acid, or only present in small amounts, in the plants used for the process according to the invention. Small amounts are advantageously
25 understood as meaning fatty acid contents which are less than 4, 3, 2 or 1% by weight, advantageously less than 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2 or 0.1% by weight based on the total fatty acid content of the plant.

For economic reasons, that is to say because of the area under cultivation and the oil
30 yield, plants which are grown on a large scale, such as soybean, oilseed rape, mustard, *Camelina*, linseed, sunflower, oil palm, cotton, sesame, maize, olive, are preferred, preferably oilseed rape, *Camelina*, linseed, sunflower are used frequently as host plant in the process.

35 Chemically pure polyunsaturated fatty acids or fatty acid compositions can also be synthesized by the processes described above. To this end, the fatty acids or the fatty acid compositions are isolated from the plants, advantageously the seeds of the plants, in the known manner, for example via crushing the seeds, such as grinding, followed

by extraction, distillation, crystallization, chromatography or a combination of these methods. These chemically pure fatty acids or fatty acid compositions are advantageous for applications in the food industry sector, the cosmetic sector and especially the pharmacological industry sector.

5

Plants which are suitable for the process according to the invention are, in principle, all those plants which are capable of synthesizing fatty acids, such as all dicotyledonous or monocotyledonous plants, algae or mosses. Advantageous plants are selected from the group of the plant families Adelotheceaceae, Anacardiaceae, Asteraceae, Apiaceae,

- 10 Betulaceae, Boraginaceae, Brassicaceae, Bromeliaceae, Caricaceae, Cannabaceae, Compositae, Convolvulaceae, Cruciferae, Cucurbitaceae, Elaeagnaceae, Ericaceae, Euphorbiaceae, Fabaceae, Geraniaceae, Gramineae, Juglandaceae, Lauraceae, Leguminosae, Linaceae, Malvaceae, Moringaceae, Marchantiaceae, Onagraceae, Olacaceae, Oleaceae, Papaveraceae, Piperaceae, Pedaliaceae, Poaceae, Rosaceae
15 or Solanaceae, vorteilhaft Anacardiaceae, Asteraceae, Boraginaceae, Brassicaceae, Cannabaceae, Compositae, Cruciferae, Cucurbitaceae, Elaeagnaceae, Euphorbiaceae, Fabaceae, Geraniaceae, Gramineae, Leguminosae, Linaceae, Malvaceae, Moringaceae, Marchantiaceae, Onagraceae, Olacaceae, Oleaceae, Papaveraceae, Piperaceae, Pedaliaceae, Poaceae or Solaneae, but other plants which
20 are suitable for the process are vegetable plants or ornamentals such as Tagetes.

Examples which may be mentioned are the following plants selected from the group consisting of: Anacardiaceae such as the genera Pistacia, Mangifera, Anacardium, for example the genus and species *Pistacia vera* [pistachio], *Mangifer indica* [mango] or

- 25 *Anacardium occidentale* [cashew], Asteraceae, such as the genera Calendula, Carthamus, Centaurea, Cichorium, Cynara, Helianthus, Lactuca, Locusta, Tagetes, Valeriana, for example the genus and species *Artemisia sphaerocephala*, *Calendula officinalis* [common marigold], *Carthamus tinctorius* [safflower], *Centaurea cyanus* [cornflower], *Cichorium intybus* [chicory], *Cynara scolymus* [artichoke], *Helianthus annuus* [sunflower], *Lactuca sativa*, *Lactuca crispera*, *Lactuca esculenta*, *Lactuca scariola*
30 *L. ssp. sativa*, *Lactuca scariola* L. var. *integrata*, *Lactuca scariola* L. var. *integrifolia*, *Lactuca sativa* subsp. *romana*, *Locusta communis*, *Valeriana locusta* [salad vegetables], *Tagetes lucida*, *Tagetes erecta* or *Tagetes tenuifolia* [african or french marigold], Apiaceae, such as the genus Daucus, for example the genus and species
35 *Daucus carota* [carrot], Betulaceae, such as the genus Corylus, for example the genera and species *Corylus avellana* or *Corylus colurna* [hazelnut], Boraginaceae, such as the genus Adelocaryum, Alkanna, Anchusa, Borago, Brunnera, Cerinthe, Cynoglossum, Echium, Gastrocatyle, Lithospermum, Moltkia, Nonea, Onosma, Onosmodium,

- Paracaryum, Pectocarya, Symphytum for example the genus and species *Adelocarym coelestinum*, *Alkanna orientalis*, *Anchusa anzurea*, *Anchusa capensis*, *Anchusa hybrida*, *Borago officinalis* [borage], *Brunnera orientalis*, *Cerithe minor*, *Cynoglossum amabile*, *Cynoglossum lanceolatum*, *Echium rubrum*, *Echium vulgare*, *Gastrocatyle hispida*, *Lithospermum arvense*, *Lithospermum purpureocaeruleum*, *Moltkia aurea*,
5 *Moltkia coerules*, *Nonea macrosperma*, *Onosma sericeum*, *Onosmodium molle*, *Onosmodium occidentale*, *Paracaryum caelestinum*, *Pectocarya platycarpa*, *Symphytum officinale*, Brassicaceae, such as the genera *Brassica*, *Camelina*, *Melanosinapis*, *Sinapis*, *Arabadopsis*, for example the genera and species *Brassica alba*, *Brassica carinata*, *Brassica hirta*, *Brassica napus*, *Brassica rapa* ssp. [oilseed rape], *Sinapis arvensis* *Brassica juncea*, *Brassica juncea* var. *juncea*, *Brassica juncea* var. *crispifolia*, *Brassica juncea* var. *foliosa*, *Brassica nigra*, *Brassica sinapioides*, *Camelina sativa*, *Melanosinapis communis* [mustard], *Brassica oleracea* [fodder beet] or *Arabidopsis thaliana*, Bromeliaceae, such as the genera *Anana*, *Bromelia*
15 (pineapple), for example the genera and species *Anana comosus*, *Ananas ananas* or *Bromelia comosa* [pineapple], Caricaceae, such as the genus *Carica*, such as the genus and species *Carica papaya* [pawpaw], Cannabaceae, such as the genus *Cannabis*, such as the genus and species *Cannabis sativa* [hemp], Convolvulaceae, such as the genera *Ipomea*, *Convolvulus*, for example the genera and species
20 *Ipomoea batatas*, *Ipomoea pandurata*, *Convolvulus batatas*, *Convolvulus tiliaceus*, *Ipomoea fastigiata*, *Ipomoea tiliacea*, *Ipomoea triloba* or *Convolvulus panduratus* [sweet potato, batate], Chenopodiaceae, such as the genus *Beta*, such as the genera and species *Beta vulgaris*, *Beta vulgaris* var. *altissima*, *Beta vulgaris* var. *vulgaris*, *Beta maritima*, *Beta vulgaris* var. *perennis*, *Beta vulgaris* var. *conditiva* or *Beta vulgaris* var.
25 *esculenta* [sugarbeet], Cryptecodiniaceae, such as the genus *Cryptecodinium*, for example the genus and species *Cryptecodinium cohnii*, Cucurbitaceae, such as the genus *Cucurbita*, for example the genera and species *Cucurbita maxima*, *Cucurbita mixta*, *Cucurbita pepo* or *Cucurbita moschata* [pumpkin/squash], Elaeagnaceae, such as the genus *Elaeagnus*, for example the genus and species *Olea europaea* [olive],
30 Ericaceae, such as the genus *Kalmia*, for example the genera and species *Kalmia latifolia*, *Kalmia angustifolia*, *Kalmia microphylla*, *Kalmia polifolia*, *Kalmia occidentalis*, *Cistus chamaerhodendros* or *Kalmia lucida* [mountain laurel], Euphorbiaceae, such as the genera *Manihot*, *Janipha*, *Jatropha*, *Ricinus*, for example the genera and species *Manihot utilisissima*, *Janipha manihot*, *Jatropha manihot*, *Manihot aipil*, *Manihot dulcis*,
35 *Manihot manihot*, *Manihot melanobasis*, *Manihot esculenta* [cassava] or *Ricinus communis* [castor-oil plant], Fabaceae, such as the genera *Pisum*, *Albizia*, *Cathormion*, *Feuillea*, *Inga*, *Pithecolobium*, *Acacia*, *Mimosa*, *Medicajo*, *Glycine*, *Dolichos*, *Phaseolus*, soybean, for example the genera and species *Pisum sativum*, *Pisum*

- arvense*, *Pisum humile* [pea], *Albizia berteriana*, *Albizia julibrissin*, *Albizia lebbeck*,
Acacia berteriana, *Acacia littoralis*, *Albizia berteriana*, *Albizia berteriana*, *Cathormion*
berteriana, *Feuillea berteriana*, *Inga fragrans*, *Pithecellobium berterianum*,
Pithecellobium fragrans, *Pithecolobium berterianum*, *Pseudalbizzia berteriana*, *Acacia*
5 *julibrissin*, *Acacia nemu*, *Albizia nemu*, *Feuillea julibrissin*, *Mimosa julibrissin*, *Mimosa*
speciosa, *Sericanrda julibrissin*, *Acacia lebbeck*, *Acacia macrophylla*, *Albizia lebbeck*,
Feuillea lebbeck, *Mimosa lebbeck*, *Mimosa speciosa* [silk tree], *Medicago sativa*,
Medicago falcata, *Medicago varia* [alfalfa] *Glycine max*, *Dolichos soja*, *Glycine gracilis*,
Glycine hispida, *Phaseolus max*, *Soja hispida* or *Soja max* [soybean], Geraniaceae,
10 such as the genera *Pelargonium*, *Cocos*, *Oleum*, for example the genera and species
Cocos nucifera, *Pelargonium grossularioides* or *Oleum cocois* [coconut], Gramineae,
such as the genus *Saccharum*, for example the genus and species *Saccharum*
officinatum, Juglandaceae, such as the genera *Juglans*, *Wallia*, for example the genera
and species *Juglans regia*, *Juglans ailanthifolia*, *Juglans sieboldiana*, *Juglans cinerea*,
15 *Wallia cinerea*, *Juglans bixbyi*, *Juglans californica*, *Juglans hindsii*, *Juglans intermedia*,
Juglans jamaicensis, *Juglans major*, *Juglans microcarpa*, *Juglans nigra* or *Wallia nigra*
[walnut], Lauraceae, such as the genera *Persea*, *Laurus*, for example the genera and
species *Laurus nobilis* [bay], *Persea americana*, *Persea gratissima* or *Persea persea*
[avocado], Leguminosae, such as the genus *Arachis*, for example the genus and
20 species *Arachis hypogaea* [peanut], Linaceae, such as the genera *Adenolinum*, for
example the genera and species *Linum usitatissimum*, *Linum humile*, *Linum*
austriacum, *Linum bienne*, *Linum angustifolium*, *Linum catharticum*, *Linum flavum*,
Linum grandiflorum, *Adenolinum grandiflorum*, *Linum lewisii*, *Linum narbonense*, *Linum*
perenne, *Linum perenne* var. *lewisii*, *Linum pratense* or *Linum trigynum* [linseed],
25 Lythraeae, such as the genus *Punica*, for example the genus and species *Punica*
granatum [pomegranate], Malvaceae, such as the genus *Gossypium*, for example the
genera and species *Gossypium hirsutum*, *Gossypium arboreum*, *Gossypium*
barbadense, *Gossypium herbaceum* or *Gossypium thurberi* [cotton], Marchantiaceae,
such as the genus *Marchantia*, for example the genera and species *Marchantia*
30 *berteroana*, *Marchantia foliacea*, *Marchantia macropora*, Musaceae, such as the genus
Musa, for example the genera and species *Musa nana*, *Musa acuminata*, *Musa*
paradisiaca, *Musa* spp. [banana], Onagraceae, such as the genera *Camissonia*,
Oenothera, for example the genera and species *Oenothera biennis* or *Camissonia*
brevipes [evening primrose], Palmae, such as the genus *Elaeis*, for example the genus
and species *Elaeis guineensis* [oil palm], Papaveraceae, such as, for example, the
35 genus *Papaver*, for example the genera and species *Papaver orientale*, *Papaver*
rhoeas, *Papaver dubium* [poppy], Pedaliaceae, such as the genus *Sesamum*, for
example the genus and species *Sesamum indicum* [sesame], Piperaceae, such as the

- genera *Piper*, *Artanthe*, *Peperomia*, *Steffensia*, for example the genera and species *Piper aduncum*, *Piper amalago*, *Piper angustifolium*, *Piper auritum*, *Piper betel*, *Piper cubeba*, *Piper longum*, *Piper nigrum*, *Piper retrofractum*, *Artanthe adunca*, *Artanthe elongata*, *Peperomia elongata*, *Piper elongatum*, *Steffensia elongata* [cayenne pepper],
- 5 Poaceae, such as the genera *Hordeum*, *Secale*, *Avena*, *Sorghum*, *Andropogon*, *Holcus*, *Panicum*, *Oryza*, *Zea* (maize), *Triticum*, for example the genera and species *Hordeum vulgare*, *Hordeum jubatum*, *Hordeum murinum*, *Hordeum secalinum*, *Hordeum distichon*, *Hordeum aegiceras*, *Hordeum hexastichon*, *Hordeum hexastichum*, *Hordeum irregulare*, *Hordeum sativum*, *Hordeum secalinum* [barley], *Secale cereale*
- 10 [rye], *Avena sativa*, *Avena fatua*, *Avena byzantina*, *Avena fatua* var. *sativa*, *Avena hybrida* [oats], *Sorghum bicolor*, *Sorghum halepense*, *Sorghum saccharatum*, *Sorghum vulgare*, *Andropogon drummondii*, *Holcus bicolor*, *Holcus sorghum*, *Sorghum aethiopicum*, *Sorghum arundinaceum*, *Sorghum caffrorum*, *Sorghum cernuum*, *Sorghum dochna*, *Sorghum drummondii*, *Sorghum durra*, *Sorghum guineense*,
- 15 *Sorghum lanceolatum*, *Sorghum nervosum*, *Sorghum saccharatum*, *Sorghum subglabrescens*, *Sorghum verticilliflorum*, *Sorghum vulgare*, *Holcus halepensis*, *Sorghum miliaceum*, *Panicum militaceum* [millet], *Oryza sativa*, *Oryza latifolia* [rice], *Zea mays* [maize] *Triticum aestivum*, *Triticum durum*, *Triticum turgidum*, *Triticum hybernum*, *Triticum macha*, *Triticum sativum* or *Triticum vulgare* [wheat],
- 20 Porphyridiaceae, such as the genera *Chrootheca*, *Flintiella*, *Petrovanella*, *Porphyridium*, *Rhodella*, *Rhodorus*, *Vanhoeffenia*, for example the genus and species *Porphyridium cruentum*, Proteaceae, such as the genus *Macadamia*, for example the genus and species *Macadamia intergrifolia* [macadamia], Rosaceae, such as the genus *Prunus*, for example the genus and species *Prunus armeniaca*, *Prunus amygdalus*, *Prunus avilum*, Rubiaceae, such as the genus *Coffea*, for example the
- 25 genera and species *Coffea* spp., *Coffea arabica*, *Coffea canephora* or *Coffea liberica* [coffee], Scrophulariaceae, such as the genus *Scrophularia*, *Verbascum*, for example the genera and species *Scrophularia marilandica*, *Verbascum blattaria*, *Verbascum chaixii*, *Verbascum densiflorum*, *Verbascum lagurus*, *Verbascum longifolium*,
- 30 *Verbascum lychnitis*, *Verbascum nigrum*, *Verbascum olympicum*, *Verbascum phlomoides*, *Verbascum phoenicum*, *Verbascum pulverulentum* or *Verbascum thapsus* [mullein], Solanaceae, such as the genera *Capsicum*, *Nicotiana*, *Solanum*, *Lycopersicon*, for example the genera and species *Capsicum annuum*, *Capsicum annuum* var. *glabriusculum*, *Capsicum frutescens* [pepper], *Capsicum annuum*
- 35 [paprika], *Nicotiana tabacum*, *Nicotiana alata*, *Nicotiana attenuata*, *Nicotiana glauca*, *Nicotiana langsdorffii*, *Nicotiana obtusifolia*, *Nicotiana quadrivalvis*, *Nicotiana repanda*, *Nicotiana rustica*, *Nicotiana sylvestris* [tobacco], *Solanum tuberosum* [potato], *Solanum melongena* [eggplant] *Lycopersicon esculentum*, *Lycopersicon lycopersicum*,

- Lycopersicon pyriforme*, *Solanum integrifolium* or *Solanum lycopersicum* [tomato], Sterculiaceae, such as the genus *Theobroma*, for example the genus and species *Theobroma cacao* [cacao] or Theaceae, such as the genus *Camellia*, for example the genus and species *Camellia sinensis* [tea]. Further plants which may be mentioned are
- 5 the genus and species *Argania spinosa*, *Arnebia griffithii*, *Adansonia digitata*, *Orbignya martiana*, *Carum carvi*, *Bertholletia excelsa*, *Aleurites moluccana*, *Hydnocarpus kursii*, *Salvia hispanica*, *Vitis vinifera*, *Corvlus avellana*, *Humulus lupulus*, *Hyptis spicigera* and *Shorea stenoptera*.
- 10 Plants which are advantageously used in the process according to the invention are transgenic plants such as dicotyledonous or monocotyledonous plants. Plants which are especially advantageously used in the process according to the invention are transgenic plants which belong to the oil-producing plants, that is to say which are used
- 15 for the production of oils, such as, preferably, oil fruit crops which comprise large amounts of lipid compounds, such as peanut, oilseed rape, canola, sunflower, safflower (*Carthamus tinctoria*), poppy, mustard, hemp, castor-oil plant, olive, sesame, *Calendula*, *Punica*, evening primrose, mullein, thistle, wild roses, hazelnut, almond, macadamia, avocado, bay, pumpkin/squash, linseed, soybean, pistachios, borage,
- 20 trees (oil palm, coconut, walnut) or crops such as maize, wheat, rye, oats, triticale, rice, barley, cotton, cassava, pepper, *Tagetes*, Solanaceae plants such as potato, tobacco, eggplant and tomato, *Vicia* species, pea, alfalfa or bushy plants (coffee, cacao, tea), *Salix* species, and perennial grasses and fodder crops.

- Preferred plants according to the invention are oilseed and oil crop plants such as
- 25 peanut, oilseed rape, canola, sunflower, safflower, poppy, Indian mustard, mustard, hemp, castor-oil plant, olive, *Calendula*, *Punica*, evening primrose, pumpkin/squash, linseed, soybean, borage, trees (oil palm, coconut). Especially preferred are plants which are high in C18:2- and/or C18:3-fatty acids, such as sunflower, safflower, tobacco, mullein, sesame, cotton, pumpkin/squash, poppy, evening primrose, walnut,
- 30 linseed, hemp, thistle or safflower. Very especially preferred plants are plants such as safflower, sunflower, poppy, evening primrose, walnut, linseed, Indian mustard, *Camelina* or hemp.

- It is advantageous for the above-described processes according to the invention to
- 35 additionally introduce, into the plant, further nucleic acids which encode enzymes of the fatty acid or lipid metabolism, in addition to the nucleic acids introduced in steps (a) to (e) or (a) to (c) of the process, and the optionally introduced nucleic acid sequences which encode the ω 3-desaturases and/or the Δ 12-desaturases.

In principle, all genes of the fatty acid or lipid metabolism can be used in the process for the production of polyunsaturated fatty acids, advantageously in combination with the $\Delta 5$ -elongase(s), $\Delta 6$ -elongase(s) and/or $\omega 3$ -desaturases [for the purposes of the present invention, the plural is understood as encompassing the singular and vice versa]. Genes of the fatty acid or lipid metabolism selected from the group consisting of acyl-CoA dehydrogenase(s), acyl-ACP [= acyl carrier protein] desaturase(s), acyl-ACP thioesterase(s), fatty acid acyl transferase(s), acyl-CoA:lysophospholipid acyltransferases, fatty acid synthase(s), fatty acid hydroxylase(s), acetyl-coenzyme A carboxylase(s), acyl-coenzyme A oxidase(s), fatty acid desaturase(s), fatty acid acetylenases, lipoxygenases, triacylglycerol lipases, allenoxide synthases, hydroperoxide lyases or fatty acid elongase(s) are advantageously used in combination with the $\Delta 5$ -elongase, $\Delta 6$ -elongase and/or $\omega 3$ -desaturase. Genes selected from the group of the $\Delta 4$ -desaturases, $\Delta 5$ -desaturases, $\Delta 6$ -desaturases, $\Delta 8$ -desaturases, $\Delta 9$ -desaturases, $\Delta 12$ -desaturases, $\Delta 6$ -elongases or $\Delta 9$ -elongases are especially preferably used in combination with the above genes for the $\Delta 5$ -elongase, $\Delta 6$ -elongase and/or $\omega 3$ -desaturase, it being possible to use individual genes or a plurality of genes in combination. The abovementioned genes are advantageously used in combination with the $\Delta 6$ -elongase, $\Delta 5$ -elongase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase and/or $\Delta 12$ -desaturase used in accordance with the invention.

Genes selected from the group of the $\Delta 8$ -desaturases, $\Delta 9$ -desaturases, $\Delta 5$ -elongase or $\Delta 9$ -elongases are especially preferably used in combination with the abovementioned genes.

Owing to the enzymatic activity of the nucleic acids used in the process according to the invention which encode polypeptides with $\Delta 6$ -elongase, $\Delta 6$ -desaturase, $\Delta 5$ -desaturase and/or $\Delta 12$ -desaturase activity, advantageously in combination with nucleic acid sequences which encode polypeptides of the fatty acid or lipid metabolism, such as polypeptides with $\Delta 8$ -desaturase, or $\Delta 5$ - or $\Delta 9$ -elongase activity, a wide range of polyunsaturated fatty acids can be produced in the process according to the invention. Depending on the choice of plants used for the process according to the invention, mixtures of the various polyunsaturated fatty acids or individual polyunsaturated fatty acids, such as EPA or ARA, can be produced in free or bound form. Depending on the prevailing fatty acid composition in the starting plant (C18:2- or C18:3-fatty acids), fatty acids which are derived from C18:2-fatty acids, such as GLA, DGLA or ARA, or fatty acids which are derived from C18:3-fatty acids, such as SDA, ETA or EPA, are thus obtained. If only linoleic acid (= LA, C18:2 ^{$\Delta 9,12$}) is present as unsaturated fatty acid in

the plant used for the process, the process can only afford GLA, DGLA and ARA as products, all of which can be present as free fatty acids or in bound form. If only α -linolenic acid (= ALA, C18:3 ^{Δ 9,12,15}) is present as unsaturated fatty acid in the plant used for the process, as is the case, for example, in linseed, the process can only
5 afford SDA, ETA or EPA as products, all of which can be present as free fatty acids or in bound form, as described above.

Owing to the activity of Δ 6-desaturase and Δ 6-elongase, products formed are, for example, GLA and DGLA, or SDA and ETA, respectively, depending on the starting
10 plant and the unsaturated fatty acid present therein. DGLA or ETA or mixtures of these are preferentially formed. If Δ 5-desaturase is additionally introduced into the plant, ARA and/or EPA are also formed. If, moreover, genes which encode a Δ 5-elongase and/or Δ 4-desaturase activity are additionally introduced, the fatty acids DPA and/or DHA can be produced in the process according to the invention. Advantageously, only ARA,
15 EPA and/or DHA or mixtures of these are synthesized, depending on the fatty acid present in the plant, which acts as starting substance for the synthesis. Since biosynthetic cascades are involved, the end products in question are not present in pure form in the organisms. Small amounts of the precursor compounds are always additionally present in the end product. These small amounts amount to less than 20%
20 by weight, advantageously less than 15% by weight, especially advantageously less than 10% by weight, most advantageously less than 5, 4, 3, 2 or 1% by weight, based on the end products DGLA, ETA or their mixtures, or ARA, EPA or their mixtures, or ARA, EPA, DHA or their mixtures.

In addition to the production directly in the plant, of the starting fatty acids for the
25 enzymes used in the process of the invention, the fatty acids can also be fed externally. The production in the plant is preferred for reasons of economy. Substrates which are preferred for the production of ARA are linoleic acid (C18:2 ^{Δ 9,12}), γ -linolenic acid (C18:3 ^{Δ 6,9,12}) and dihomo- γ -linolenic acid (C20:3 ^{Δ 8,11,14}). Substrates which are preferred for the production of EPA are linolenic acid (C18:3 ^{Δ 9,12,15}), stearidonic acid
30 (C18:4 ^{Δ 6,9,12,15}) and eicosatetraenoic acid (C20:4 ^{Δ 8,11,14,17}). Substrates which are preferred for the production of DHA are linolenic acid (C18:3 ^{Δ 9,12,15}), stearidonic acid (C18:4 ^{Δ 6,9,12,15}), eicosatetraenoic acid (C20:4 ^{Δ 8,11,14,17}), EPA and DPA.

In comparison with the human elongases or elongases from non-human animals, such
35 as those from *Oncorhynchus*, *Xenopus* or *Ciona*, the Δ 5-elongases according to the invention have the advantageous characteristic that they do not elongate C₂₂-fatty acids to the corresponding C₂₄-fatty acids. Furthermore, they advantageously do not convert

fatty acids with a double bond in the $\Delta 6$ -position, as is the case with the human elongases or the elongases from non-human animals. Especially advantageously $\Delta 5$ -elongases preferentially only convert unsaturated C_{20} -fatty acids. These advantageous $\Delta 5$ -elongases contain some putative transmembrane helices (5 – 7). Advantageously, only C_{20} -fatty acids with one double bond in the $\Delta 5$ -position are converted, with $\omega 3$ - C_{20} -fatty acids being preferred (EPA). Moreover, in a preferred embodiment of the invention, they have the characteristic that, besides the $\Delta 5$ -elongase activity, they advantageously have no, or only relatively low, $\Delta 6$ -elongase activity. In contrast, the human elongases or non-human animal elongases have approximately the same activity towards fatty acids with a $\Delta 6$ - or $\Delta 5$ -double bond. These advantageous elongases are referred to what are known as monofunctional elongases. In contrast, the human elongases or the non-human animal elongases are referred to as multifunctional elongases, which, besides the abovementioned substrates, also convert monounsaturated C_{16} - and C_{18} -fatty acids, for example with $\Delta 9$ - or $\Delta 11$ -double bonds. In a yeast feeding test, in which EPA was added to the yeast as the substrate, the monofunctional elongases convert at least 15% by weight of the added EPA into docosapentaenoic acid (DPA, $C_{22:5}^{\Delta 7,10,13,16,19}$), advantageously at least 20% by weight, especially advantageously at least 25% by weight. If γ -linolenic acid (= GLA, $C_{18:3}^{\Delta 6,9,12}$) is added as the substrate, this acid is advantageously not elongated at all. Likewise, $C_{18:3}^{\Delta 6,9,12}$ is not elongated. In another advantageous embodiment, less than 60% by weight of the added GLA is converted into dihomogamma-linolenic acid (= $C_{20:3}^{\Delta 8,11,14}$), advantageously less than 55% by weight, preferably less than 50% by weight, especially advantageously less than 45% by weight, very especially advantageously less than 40% by weight. In a further, very preferred embodiment of the $\Delta 5$ -elongase activity according to the invention, GLA is not converted.

Figures 27 and 28 show the measured substrate specificities of the various elongases. Figure 27 shows the specificities of the multifunctional elongases from *Xenopus laevis* (Fig. 27 A), *Ciona intestinalis* (Fig. 27 B) and *Oncorhynchus mykiss* (Fig. 27 C). All these elongases convert a broad substrate spectrum. In the process according to the invention, this can lead to by-products, which must be converted by further enzymatic activities. This is why these enzymes are less preferred in the process according to the invention. The preferred monofunctional elongases and their substrate specificity are shown in Figure 28. Figure 28 A shows the specificity of the *Ostreococcus tauri* $\Delta 5$ -elongase. This enzyme only converts fatty acids with a double bond in the $\Delta 5$ -position. Advantageously, only C_{20} -fatty acids are converted. A similarly high substrate specificity is shown by the *Thalassiosira pseudonana* $\Delta 5$ -elongase (Fig. 28. C). Both the *Ostreococcus tauri* $\Delta 6$ -elongase (Fig. 28 B) as that of *Thalassiosira pseudonana* (Fig.

28 D) advantageously only convert fatty acids with a double bond in the $\Delta 6$ -position. Advantageously, only C^{18} -fatty acids are converted. The $\Delta 5$ -elongases from *Arabidopsis thaliana* and *Euglena gracilis* are also distinguished by their specificities.

- 5 Likewise, advantageous $\Delta 6$ -elongases according to the invention are distinguished by a high specificity, that is to say that C_{18} -fatty acids are preferentially elongated. They advantageously convert fatty acids with a double bond in the $\Delta 6$ -position. Especially advantageous $\Delta 6$ -elongases advantageously convert C_{18} -fatty acids with three or four double bonds in the molecule, which fatty acids must comprise a double bond in the
- 10 $\Delta 6$ -position. Moreover, in a preferred embodiment of the invention, they have the characteristic that, besides the $\Delta 6$ -elongase activity, they advantageously have no, or only relatively low, $\Delta 5$ -elongase activity. In contrast, the human elongases or non-human animal elongases have approximately the same activity towards fatty acids with a $\Delta 6$ - or $\Delta 5$ -double bond. These advantageous elongases are referred to as what are
- 15 known as monofunctional elongases. In contrast, the human elongases or the non-human animal elongases are referred to as multifunctional elongases, which, besides the abovementioned substrates, also convert monounsaturated C_{16} - and C_{18} -fatty acids, for example with $\Delta 9$ - or $\Delta 11$ -double bonds. In a yeast feeding test, in which EPA has been added to the yeasts as the substrate, the monofunctional elongases convert
- 20 at least 10% by weight of the added α -linolenic acid (= ALA, $C_{18:3}^{\Delta 9,12,15}$) or at least 40% by weight of added γ -linolenic acid (= GLA, $C_{18:3}^{\Delta 6,9,12}$), advantageously at least 20% by weight and 50% by weight, respectively, especially advantageously at least 25% by weight and 60% by weight, respectively. It is especially advantageous that $C_{18:4}^{\Delta 6,9,12,15}$ (stearidonic acid) is also elongated. Here, SDA is converted to at least
- 25 40% by weight, advantageously to at least 50% by weight, especially advantageously to at least 60% by weight, very especially advantageously to at least 70% by weight. Especially advantageous $\Delta 6$ -elongases show no, or only very low activity (less than 0.1% by weight conversion rate) toward the following substrates: $C_{18:1}^{\Delta 6}$, $C_{18:1}^{\Delta 9}$, $C_{18:1}^{\Delta 11}$, $C_{20:2}^{\Delta 11,14}$, $C_{20:3}^{\Delta 11,14,17}$, $C_{20:3}^{\Delta 8,11,14}$, $C_{20:4}^{\Delta 5,8,11,14}$, $C_{20:5}^{\Delta 5,8,11,14,17}$ or
- 30 $C_{22:4}^{\Delta 7,10,13,16}$.

Figures 29 and 30 and Table 21 show the measured substrate specificities of the various elongases.

- 35 In comparison with the known $\omega 3$ -desaturase, the $\omega 3$ -desaturase used in the process according to the invention has the advantageous characteristic that it is capable of desaturating a broad spectrum of $\omega 6$ -fatty acids, with C_{20} - and C_{22} -fatty acids such as $C_{20:2}$, $C_{20:3}$, $C_{20:4}$, $C_{22:4}$ or $C_{22:5}$ -fatty acids being preferentially desaturated. However,

the shorter C_{18} -fatty acids such as $C_{18:2}$ - or $C_{18:3}$ -fatty acids are also advantageously desaturated. Owing to these characteristics of $\omega 3$ -desaturase, it is advantageously possible to shift the fatty acid spectrum within an organism, advantageously within a plant or a fungus, from the $\omega 6$ -fatty acids towards the $\omega 3$ -fatty acids. The $\omega 3$ -desaturase according to the invention preferentially desaturates C_{20} -fatty acids. Within the organism, these fatty acids are converted to at least 10%, 15%, 20%, 25% or 30% from the existing fatty acid pool to give the corresponding $\omega 3$ -fatty acids. In comparison with the C_{18} -fatty acids, the activity of $\omega 3$ -desaturase is lower by a factor of 10, that is to say only approximately 1.5 to 3% of the fatty acids present in the fatty acid pool are converted into the corresponding $\omega 3$ -fatty acids. Preferred substrates of the $\omega 3$ -desaturase according to the invention are the $\omega 6$ -fatty acids bound in phospholipids. With reference to the desaturation of dihomo- γ -linolenic acid [$C_{20:4}^{\Delta 8,11,14}$], Figure 19 shows clearly that $\omega 3$ -desaturase advantageously does not differentiate between fatty acids bound at the sn1 or sn2 position when desaturation takes place. Both fatty acids bound at the sn1 position and fatty acids bound in the sn2 position in the phospholipids are desaturated. Another advantage is that $\omega 3$ -desaturase converts a broad range of phospholipids such as phosphatidylcholine (= PC), phosphatidylinositol (= PIS) or phosphatidylethanolamine (= PE). Finally, desaturation products are also found in the neutral lipids (= NL), i.e. in the triglycerides.

In comparison with the known $\Delta 4$ -desaturases, $\Delta 5$ -desaturases and $\Delta 6$ -desaturases, the advantage of the $\Delta 4$ -desaturases, $\Delta 5$ -desaturases and $\Delta 6$ -desaturases used in the process according to the invention is that they can convert fatty acids which are bound to phospholipids or CoA-fatty acid esters, advantageously CoA-fatty acid esters.

The $\Delta 12$ -desaturases used in the process according to the invention advantageously convert oleic acid ($C_{18:1}^{\Delta 9}$) into linoleic acid ($C_{18:2}^{\Delta 9,12}$) or $C_{18:2}^{\Delta 6,9}$ into $C_{18:3}^{\Delta 6,9,12}$ (= GLA). The $\Delta 12$ -desaturases used advantageously convert fatty acids which are bound to phospholipids or CoA-fatty acid esters, advantageously those which are bound to CoA-fatty acid esters.

Owing to the enzymatic activity of the nucleic acids used in the process according to the invention which encode polypeptides with $\Delta 5$ -elongase, $\Delta 6$ -elongase and/or $\omega 3$ -desaturase activity, advantageously in combination with nucleic acid sequences which encode polypeptides of the fatty acid or lipid metabolism, such as additionally polypeptides with $\Delta 4$ -, $\Delta 5$ -, $\Delta 6$ -, $\Delta 8$ -, $\Delta 12$ -desaturase or $\Delta 5$ -, $\Delta 6$ - or $\Delta 9$ -elongase activity, a very wide range of polyunsaturated fatty acids can be produced in the process according to the invention. Depending on the choice of the advantageous

plants used for the process according to the invention, mixtures of the various polyunsaturated fatty acids or individual polyunsaturated fatty acids such as EPA, ARA or DHA, can be produced in free or bound form. Depending on the prevailing fatty acid composition in the starting plant (C18:2- or C18:3-fatty acids), fatty acids which are derived from C18:2-fatty acids, such as GLA, DGLA or ARA, or which are derived from C18:3-fatty acids, such as SDA, ETA, EPA or DHA, are thus obtained. If only linoleic acid (= LA, C18:2^{Δ^{9,12}}) is present as unsaturated fatty acid in the plant used for the process, the process can only afford GLA, DGLA and ARA as products, all of which can be present as free fatty acids or in bound form. By expressing the additional ω3-desaturase in plants, the fatty acid spectrum can be shifted towards α-linolenic acid, DPA and DHA. However, this shift in the fatty acid spectrum is only relatively limited. More advantageous is such a shift in plants which, as described hereinbelow, already have a high α-linolenic acid content. If only α-linolenic acid (= ALA, C18:3^{Δ^{9,12,15}}) is present as unsaturated fatty acid in the plant, as is the case, for example, in linseed, the process can only afford SDA, ETA, EPA and/or DHA, which, as described above, may be present as free fatty acids or in bound form. Owing to the modification of the activity of the enzyme Δ5-elongase which plays a role in the synthesis, advantageously in combination with Δ4-, Δ5-, Δ6-, Δ12-desaturase and/or Δ6-elongase, or Δ4-, Δ5-, Δ8-, Δ12-desaturase, and/or Δ9-elongase, it is possible to produce, in a targeted fashion, only individual products in the abovementioned plants. Owing to the activity of Δ6-desaturase and Δ6-elongase, for example, GLA and DGLA, or SDA and ETA, are formed, depending on the starting plant and unsaturated fatty acids. DGLA or ETA or mixtures of these are preferentially formed. If Δ5-desaturase, Δ5-elongase and Δ4-desaturase are additionally introduced into the organisms, advantageously into the plant, ARA, EPA and/or DHA are additionally formed. This also applies to organisms into which Δ8-desaturase and Δ9-elongase have previously been introduced. Advantageously, only ARA, EPA or DHA or their mixtures are synthesized, depending on the fatty acid present in the plant, which acts as starting substance for the synthesis. Since biosynthetic cascades are involved, the end products in question are not present in pure form in the organisms. Small amounts of the precursor compounds are always additionally present in the end product. These small amounts amount to less than 20% by weight, advantageously less than 15% by weight, especially advantageously less than 10% by weight, very especially advantageously less than 5, 4, 3, 2, or 1% by weight, based on the end product DGLA, ETA or their mixtures, or ARA, EPA, DHA or their mixtures, advantageously EPA or DHA or their mixtures.

The nucleic acid with the SEQ ID NO: 53, which is derived from trout and which can be used in the process according to the invention, encodes a protein with high specificity

for the two C18:4^{Δ6,9,12,15}- and C20:5^{Δ5,8,11,14,17}-fatty acids, which are precursors for the synthesis of DHA (precursors and synthesis of DHA, see Figure 1). However, other fatty acids too are elongated by the enzyme. The protein encoded by SEQ ID NO: 53 thus has specificity for Δ6- and Δ5-fatty acids with additionally one ω3-double bond (Figure 2). Δ5-elongase has a keto-acyl-CoA synthase activity which advantageously elongates fatty acid residues of acyl-CoA esters by 2 carbon atoms.

The synthesis of DHA in yeast (*Saccharomyces cerevisiae*) was detected by the gene product of the abovementioned fish Δ5-elongase gene and further Δ5-elongases, the Δ5-desaturase from *Phaeodactylum* and the Δ4-desaturase from *Euglena* (Figure 3).

In addition to the production directly in the transgenic organism, advantageously in the transgenic plant, of the starting fatty acids for the Δ5-elongases, Δ6-elongases, Δ9-elongases, Δ4-desaturases, Δ5-desaturases, Δ6-desaturases, Δ12-desaturases and/or ω3-desaturases advantageously used in the process according to the invention, the fatty acids can also be shed externally. The production in the organism is preferred for reasons of economy. Preferred substrates of ω3-desaturase are linoleic acid (C18:2^{Δ9,12}), γ-linolenic acid (C18:3^{Δ6,9,12}), eicosadienoic acid (C20:2^{Δ11,14}), dihomo-γ-linolenic acid (C20:3^{Δ8,11,14}), arachidonic acid (C20:4^{Δ5,8,11,14}), docosatetraenoic acid (C22:4^{Δ7,10,13,16}) and docosapentaenoic acid (C22:5^{Δ4,7,10,13,15}).

To increase the yield in the above-described process for the production of oils and/or triglycerides with an advantageously elevated content of polyunsaturated fatty acids, it is advantageous to increase the amount of starting product for the synthesis of fatty acids; this can be achieved for example by introducing, into the organism, a nucleic acid which encodes a polypeptide with Δ12-desaturase activity. This is particularly advantageous in oil-producing organisms such as those from the family of the Brassicaceae, such as the genus *Brassica*, for example oilseed rape; the family of the Elaeagnaceae, such as the genus *Elaeagnus*, for example the genus and species *Olea europaea*, or the family Fabaceae, such as the genus *Glycine*, for example the genus and species *Glycine max*, which are high in oleic acid. Since these organisms are only low in linoleic acid (Mikoklajczak et al., Journal of the American Oil Chemical Society, 38, 1961, 678 - 681), the use of the abovementioned Δ12-desaturases for producing the starting material linoleic acid is advantageous.

Nucleic acids used in the process according to the invention are advantageously derived from plants such as algae, for example algae of the family of the Prasinophyceae such as the genera *Heteromastix*, *Mammella*, *Mantoniella*, *Micromonas*, *Nephroselmis*, *Ostreococcus*, *Prasinocladus*, *Prasinococcus*, *Pseudoscurfieldia*, *Pycnococcus*, *Pyramimonas*, *Scherffelia* or *Tetraselmis* such as the genera and species *Heteromastix longifillis*, *Mamiella gilva*, *Mantoniella squamata*, *Micromonas pusilla*, *Nephroselmis olivacea*, *Nephroselmis pyriformis*, *Nephroselmis*

- rotunda, *Ostreococcus tauri*, *Ostreococcus* sp. *Prasinocladus ascus*, *Prasinocladus lubricus*, *Pycnococcus provasolii*, *Pyramimonas amyliifera*, *Pyramimonas disomata*, *Pyramimonas obovata*, *Pyramimonas orientalis*, *Pyramimonas parkeae*, *Pyramimonas spinifera*, *Pyramimonas* sp., *Tetraselmis apiculata*, *Tetraselmis carteriaformis*,
5 *Tetraselmis chui*, *Tetraselmis convolutae*, *Tetraselmis desikacharyi*, *Tetraselmis gracilis*, *Tetraselmis hazeni*, *Tetraselmis impellucida*, *Tetraselmis inconspicua*, *Tetraselmis levis*, *Tetraselmis maculata*, *Tetraselmis marina*, *Tetraselmis striata*, *Tetraselmis subcordiformis*, *Tetraselmis suecica*, *Tetraselmis tetrabrachia*, *Tetraselmis tetrathele*, *Tetraselmis verrucosa*, *Tetraselmis verrucosa* fo. *rubens* or *Tetraselmis* sp.
10 or from algae of the family *Euglenaceae* such as from the genera *Ascoglena*, *Astasia*, *Colacium*, *Cyclidiopsis*, *Euglena*, *Euglenopsis*, *Hyalophacus*, *Khawkinea*, *Lepocinclis*, *Phacus*, *Strombomonas* or *Trachelomonas* such as the genera and species *Euglena acus*, *Euglena geniculata*, *Euglena gracilis*, *Euglena mixocylindrica*, *Euglena rostrifera*, *Euglena viridis*, *Colacium stentorium*, *Trachelomonas cylindrica* or *Trachelomonas*
15 *volvocina*. The nucleic acid sequences used in the process can also advantageously be derived from algae, such as the alga *Porphyridium cruentum*, *Isochrysis galbana* or *Chlorella minutissima*, *Chlorella vulgaris*, *Thraustochytrium aureum* or *Nannochloropsis oculata*. The nucleic acids used are advantageously derived from algae of the genera *Euglena*, *Mantoniella* or *Ostreococcus*.
- 20 Further advantageous plants as sources for the nucleic acid sequences used in the process according to the invention are algae such as *Isochrysis* or *Cryptothecodinium*, algae/diatoms such as *Thalassiosira* or *Phaeodactylum*, mosses such as *Physcomitrella* or *Ceratodon*, or higher plants such as the *Primulaceae* such as *Aleuritia*, *Calendula stellata*, *Osteospermum spinescens* or *Osteospermum*
25 *hyoseroides*, microorganisms such as fungi, such as *Aspergillus*, *Thraustochytrium*, *Phytophthora*, *Entomophthora*, *Mucor* or *Mortierella*, bacteria such as *Shewanella*, yeasts or animals such as nematodes such as *Caenorhabditis*, insects, frogs, sea cucumber or fish. The isolated nucleic acid sequences according to the invention are advantageously derived from an animal of the order of the vertebrates. Preferably, the
30 nucleic acid sequences are derived from the classes of the Vertebrata; Euteleostomi, Actinopterygii; Neopterygii; Teleostei; Euteleostei, Protacanthopterygii, Salmoniformes; Salmonidae or *Oncorhynchus* or Vertebrata, Amphibia, Anura, Pipidae, *Xenopus* or Evertebrata such as Protochordata, Tunicata, Holothuroidea, Cionidae such as *Amaroucium constellatum*, *Botryllus schlosseri*, *Ciona intestinalis*, *Molgula citrina*,
35 *Molgula manhattensis*, *Perophora viridis* or *Styela partita*. The nucleic acids are especially advantageously derived from fungi, animals, or from plants such as algae or mosses, preferably from the order of the Salmoniformes, such as the family of the Salmonidae, such as the genus *Salmo*, for example from the genera and species *Oncorhynchus mykiss*, *Trutta trutta* or *Salmo trutta fario*, from algae, such as the

genera *Mantoniella* or *Ostreococcus*, or from the diatoms such as the genera *Thalassiosira* or *Phaeodactylum* or from algae such as *Cryptocodinium*.

5 Advantageous nucleic acid used in the process according to the invention can also be derived from microorganisms such as fungi such as the genus *Mortierella*, *Phytium*, for example the genus and species *Mortierella alpiina*, *Mortierella elongata*, *Phytium irregulare*, *Phytium ultimum* or bacteria such as the genus *Shewanella*, for example the genus and species *Shewanella hanedai*.

10 The process according to the invention advantageously employs the abovementioned nucleic acid sequences or their derivatives or homologs which encode polypeptides which retain the enzymatic activity of the proteins encoded by nucleic acid sequences. These sequences, individually or in combination with the nucleic acid sequences which encode $\Delta 12$ -desaturase, $\Delta 4$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase,
15 $\Delta 6$ -elongase and/or $\omega 3$ -desaturase, are cloned into expression constructs and used for the introduction into, and expression in, organisms. Owing to their construction, these expression constructs make possible an advantageous optimal synthesis of the polyunsaturated fatty acids produced in the process according to the invention.

20 In a preferred embodiment, the process furthermore comprises the step of obtaining a transgenic plant which comprises the nucleic acid sequences used in the process, where the plant is transformed with a nucleic acid sequence according to the invention which encodes the $\Delta 12$ -desaturase, $\Delta 4$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase and/or $\omega 3$ -desaturase, a gene construct or a vector as
25 described below, alone or in combination with further nucleic acid sequences which encode proteins of the fatty acid or lipid metabolism. In a further preferred embodiment, this process furthermore comprises the step of obtaining the oils, lipids or free fatty acids from the seed of the plant, such as, for example, the seed of an oil crop, such as, for example, peanut, oilseed rap, canola, linseed, hemp, peanut, soybean, safflower,
30 hemp, sunflowers or borage.

In the case of plant cells, plant tissue or plant organs, "growing" is understood as meaning, for example, the cultivation on or in a nutrient medium, or of the intact plant on or in a substrate, for example in a hydroponic culture, potting compost or on arable
35 land.

The invention furthermore relates to gene constructs which comprise the nucleic acid sequences according to the invention which encode a $\Delta 5$ -desaturase, $\Delta 6$ -desaturase,

$\Delta 5$ -elongase or $\Delta 6$ -elongase, the nucleic acid being linked functionally with one or more regulatory signals. In addition, the gene construct may comprise further biosynthesis genes of the fatty acid or lipid metabolism selected from the group consisting of acyl-CoA dehydrogenase(s), acyl-ACP [= acyl carrier protein] desaturase(s), acyl-ACP thioesterase(s), fatty acid acyl transferase(s), acyl-CoA:lysophospholipid acyltransferases, fatty acid synthase(s), fatty acid hydroxylase(s), acetyl-coenzyme A carboxylase(s), acyl-coenzyme A oxidase(s), fatty acid desaturase(s), fatty acid acetylenases, lipoxygenases, triacylglycerol lipases, allenoxide synthases, hydroperoxide lyases or fatty acid elongase(s). Biosynthesis genes of the fatty acid or lipid metabolism selected from the group $\Delta 8$ -desaturase, $\Delta 9$ -desaturase, $\Delta 9$ -elongase or $\omega 3$ -desaturase are advantageously additionally present.

The nucleic acid sequences used in the process which encode proteins with $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 12$ -desaturase, $\Delta 5$ -elongase or $\Delta 6$ -elongase activity are advantageously introduced into the plant alone or, preferably, in combination with an expression cassette (= nucleic acid construct) which makes possible the expression of the nucleic acids in the plant. The nucleic acid construct can comprise more than one nucleic acid sequence with an enzymatic activity, for example, of a $\Delta 12$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 6$ -elongase.

To introduce the nucleic acids into the gene constructs, the nucleic acids used in the process are advantageously amplified and ligated in the known manner. Preferably, a procedure following the protocol for Pfu DNA polymerase or a Pfu/Taq DNA polymerase mixture is followed. The primers are selected taking into consideration the sequence to be amplified. The primers should expediently be chosen in such a way that the amplificate comprises the entire codogenic sequence from the start codon to the stop codon. After the amplification, the amplificate is expediently analyzed. For example, a gel-electrophoretic separation can be carried out, which is followed by a quantitative and a qualitative analysis. Thereafter, the amplificate can be purified following a standard protol (for example Qiagen). An aliquot of the purified amplificate is then available for the subsequent cloning step.

Suitable cloning vectors are generally known to the skilled worker. These include, in particular, vectors which are capable of replication in microbial systems, that is to say mainly vectors which ensure efficient cloning in yeasts or fungi and which make possible the stable transformation of plants. Those which must be mentioned in particular are various binary and cointegrated vector systems which are suitable for the T-DNA-mediated transformation. Such vector systems are, as a rule, characterized in

that they comprise at least the vir genes required for the Agrobacterium-mediated transformation and the T-DNA-delimiting sequences (T-DNA border). These vector systems preferably also comprise further cis-regulatory regions such as promoters and terminator sequences and/or selection markers, by means of which suitably

5 transformed organisms can be identified. While in the case of cointegrated vector systems vir genes and T-DNA sequences are arranged on the same vector, binary systems are based on at least two vectors, one of which bears vir genes, but no T-DNA, while a second one bears T-DNA, but no vir genes. Owing to this fact, the last-mentioned vectors are relatively small, easy to manipulate and capable of replication
10 both in *E. coli* and in Agrobacterium. These binary vectors include vectors from the series pBIB-HYG, pPZP, pBecks, pGreen. In accordance with the invention, Bin19, pBI101, pBinAR, pGPTV and pCAMBIA are used by preference. An overview of the binary vectors and their use is found in Hellens et al, Trends in Plant Science (2000) 5, 446–451.

15 In order to prepare the vectors, the vectors can first be linearized with restriction endonuclease(s) and then modified enzymatically in a suitable manner. Thereafter, the vector is purified, and an aliquot is employed for the cloning step. In the cloning step, the enzymatically cleaved and, if appropriate, purified amplificate is ligated with vector
20 fragments which have been prepared in a similar manner, using ligase. In this context, a particular nucleic acid construct, or vector or plasmid construct, can have one or more than one codogenic gene segments. The codogenic gene segments in these constructs are preferably linked functionally with regulatory sequences. The regulatory sequences include, in particular, plant sequences such as promoters and terminator
25 sequences. The constructs can advantageously be stably propagated in microorganisms, in particular in *E. coli* and *Agrobacterium tumefaciens*, under selection conditions and make possible a transfer of heterologous DNA into plants or microorganisms.

30 The nucleic acids used in the process can be introduced into plants, advantageously using cloning vectors, and thus be used in the transformation of plants such as those which are published and cited therein: Plant Molecular Biology and Biotechnology (CRC Press, Boca Raton, Florida), Chapter 6/7, p. 71-119 (1993); F.F. White, Vectors for Gene Transfer in Higher Plants; in: Transgenic Plants, Vol. 1, Engineering and
35 Utilization, Eds.: Kung and R. Wu, Academic Press, 1993, 15-38; B. Jenes et al., Techniques for Gene Transfer, in: Transgenic Plants, Vol. 1, Engineering and Utilization, Eds.: Kung and R. Wu, Academic Press (1993), 128-143; Potrykus, Annu. Rev. Plant Physiol. Plant Molec. Biol. 42 (1991), 205-225. Thus, the nucleic acids

and/or vectors used in the process can be used for the recombinant modification of a broad spectrum of plants so that the latter become better and/or more efficient PUFA producers.

- 5 A series of mechanisms by which a modification of the $\Delta 12$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase and/or $\Delta 6$ -desaturase protein is possible exists, so that the yield, production and/or production efficiency of the polyunsaturated fatty acids in a plant, preferably in an oilseed plant or oil crop, can be influenced directly owing to this modified protein. The number or activity of the $\Delta 12$ -Desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -
10 elongase, $\Delta 6$ -elongase or $\Delta 5$ -desaturase proteins or genes can be increased, so that greater amounts of the gene products and, ultimately, greater amounts of the compounds of the general formula I are produced. A *de novo* synthesis in a plant which has lacked the activity and ability to biosynthesize the compounds prior to introduction of the corresponding gene(s) is also possible. This applies analogously to the
15 combination with further desaturases or elongases or further enzymes of the fatty acid and lipid metabolism. The use of various divergent sequences, i.e. sequences which differ at the DNA sequence level, may also be advantageous in this context, or else the use of promoters which make possible a different gene expression in the course of time, for example as a function of the degree of maturity of a seed or an oil-storing
20 tissue.

- Owing to the introduction of a combination of $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase and/or $\Delta 5$ -desaturase genes into the plant, alone or in combination with other genes, it is not only possible to increase biosynthesis flux
25 towards the end product, but also to increase, or to create *de novo* the corresponding triacylglycerol composition. Likewise, the number or activity of other genes which are involved in the import of nutrients which are required for the biosynthesis of one or more fatty acids, oils, polar and/or neutral lipids, can be increased, so that the concentration of these precursors, cofactors or intermediates within the cells or within
30 the storage compartment is increased, whereby the ability of the cells to produce PUFAs is enhanced further. By optimizing the activity or increasing the number of one or more $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase or $\Delta 5$ -desaturase genes which are involved in the biosynthesis of these compounds, or by destroying the activity of one or more genes which are involved in the degradation of these
35 compounds, an enhanced yield, production and/or production efficiency of fatty acid and lipid molecules in plants is made possible.

The nucleic acid sequences used in the process are advantageously introduced into an expression cassette which makes possible the expression of the nucleic acids in plants.

5 In doing so, the nucleic acid sequences which encode $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase or $\Delta 5$ -desaturase are linked functionally with one or more regulatory signals, advantageously for enhancing gene expression. These regulatory sequences are intended to make possible the specific expression of the genes and proteins. Depending on the host organism, this may mean, for example, that the gene
10 is expressed and/or overexpressed only after induction has taken place, or else that it is expressed and/or overexpressed immediately. For example, these regulatory sequences take the form of sequences to which inductors or repressors bind, thus controlling the expression of the nucleic acid. In addition to these novel regulatory sequences, or instead of these sequences, the natural regulatory elements of these
15 sequences may still be present before the actual structural genes and, if appropriate, may have been genetically modified in such a way that their natural regulation is eliminated and the expression of the genes is enhanced. These modified promoters can also be positioned on their own before the natural gene in the form of part-sequences (= promotor with parts of the nucleic acid sequences used in accordance
20 with the invention) in order to enhance the activity. Moreover, the gene construct may advantageously also comprise one or more what are known as enhancer sequences in operable linkage with the promoter, which make possible an enhanced expression of the nucleic acid sequence. Additional advantageous sequences, such as further regulatory elements or terminator sequences, may also be inserted at the 3' end of the
25 DNA sequences.

The $\Delta 12$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 6$ -elongase genes may be present in one or more copies of the expression cassette (= gene construct). Preferably, only one copy of the genes is present in each expression
30 cassette. This gene construct, or the gene constructs, can be expressed together in the host plant. In this context, the gene construct(s) can be inserted in one or more vectors and be present in the cell in free form, or else be inserted in the genome. It is advantageous for the insertion of further genes in the host genome when the genes to be expressed are present together in one gene construct.

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In this context, the regulatory sequences or factors can, as described above, preferably have a positive effect on the gene expression of the genes introduced, thus enhancing it. Thus, an enhancement of the regulatory elements, advantageously at the

transcriptional level, may take place by using strong transcription signals such as promoters and/or enhancers. In addition, however, enhanced translation is also possible, for example by improving the stability of the mRNA.

- 5 In a further embodiment of the invention, one or more gene constructs comprising one or more sequences which are defined by SEQ ID NO: 11, SEQ ID NO: 27, SEQ ID NO: 193, SEQ ID NO: 195, SEQ ID NO: 197, SEQ ID NO: 199, SEQ ID NO: 201 or their derivatives and which encode polypeptides as shown in SEQ ID NO: 12, SEQ ID NO: 28, SEQ ID NO: 194, SEQ ID NO: 196, SEQ ID NO: 198, SEQ ID NO: 200, SEQ ID NO: 202 are present. The abovementioned $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase or $\Delta 5$ -desaturase proteins advantageously lead to a desaturation or elongation of fatty acids, the substrate advantageously having one, two, three or four double bonds and advantageously 18, 20 or 22 carbon atoms in the fatty acid molecule. The same applies to their homologs, derivatives or analogs which are linked functionally with one or more regulatory signals, preferably for enhancing gene expression.

- In principle, it is possible to use all natural promoters together with their regulatory sequences, such as those mentioned above, for the novel process. It is also possible and advantageous to use synthetic promoters, either in addition or alone, in particular when they mediate seed-specific expression, such as those described in WO 99/16890.

- In order to achieve a particularly high PUFA content, especially in transgenic plants, the PUFA biosynthesis genes should advantageously be expressed in oilseeds in a seed-specific manner. To this end, seed-specific promoters can be used, or those promoters which are active in the embryo and/or in the endosperm. In principle, seed-specific promoters can be isolated both from dicotyledonous and from monocotyledonous plants. Preferred promoters are listed hereinbelow: USP (= unknown seed protein) and vicilin (*Vicia faba*) [Bäumlein et al., Mol. Gen Genet., 1991, 225(3)], napin (oilseed rape) [US 5,608,152], conlinin (linseed) [WO 02/102970], acyl carrier protein (oilseed rape) [US 5,315,001 and WO 92/18634], oleosin (*Arabidopsis thaliana*) [WO 98/45461 and WO 93/20216], phaseolin (*Phaseolus vulgaris*) [US 5,504,200], Bce4 [WO 91/13980], legumes B4 (LegB4 promoter) [Bäumlein et al., Plant J., 2,2, 1992], Lpt2 and Lpt1 (barley) [WO 95/15389 and WO95/23230], seed-specific promoters from rice, maize and wheat [WO 99/16890], Amy32b, Amy 6-6 and aleurain [US 5,677,474], Bce4 (oilseed rape) [US 5,530,149], glycinin (soybean) [EP 571 741], phosphoenol pyruvate carboxylase (soybean) [JP 06/62870], ADR12-2 (soybean)

[WO 98/08962], isocitrate lyase (oilseed rape) [US 5,689,040] or α -amylase (barley) [EP 781 849].

5 Plant gene expression can also be facilitated via a chemically inducible promoter (see a review in Gatz 1997, *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 48:89-108).

Chemically inducible promoters are particularly suitable when it is desired that gene expression should take place in a time-specific manner. Examples of such promoters are a salicylic-acid-inducible promoter (WO 95/19443), a tetracyclin-inducible promoter (Gatz et al. (1992) *Plant J.* 2, 397-404) and an ethanol-inducible promoter.

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To ensure the stable integration of the biosynthesis genes into the transgenic plant over a plurality of generations, each of the nucleic acids which encode $\Delta 12$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase and/or $\Delta 5$ -desaturase and which are used in the process should be expressed under the control of a separate promoter,

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preferably a promoter which differs from the other promoters, since repeating sequence motifs can lead to instability of the T-DNA, or to recombination events. In this context, the expression cassette is advantageously constructed in such a way that a promoter is followed by a suitable cleavage site, advantageously in a polylinker, for insertion of the nucleic acid to be expressed and, if appropriate, a terminator sequence is positioned

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behind the polylinker. This sequence is repeated several times, preferably three, four, five, six or seven times, so that up to seven genes can be combined in one construct and introduced into the transgenic plant in order to be expressed. Advantageously, the sequence is repeated up to four times. To express the nucleic acid sequences, the

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latter are inserted behind the promoter via a suitable cleavage site, for example in the polylinker. Advantageously, each nucleic acid sequence has its own promoter and, if appropriate, its own terminator sequence. Such advantageous constructs are

disclosed, for example, in DE 101 02 337 or DE 101 02 338. However, it is also

possible to insert a plurality of nucleic acid sequences behind a shared promoter and, if appropriate, before a shared terminator sequence. Here, the insertion site, or the

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sequence, of the inserted nucleic acids in the expression cassette is not of critical importance, that is to say a nucleic acid sequence can be inserted at the first or last position in the cassette without its expression being substantially influenced thereby.

Advantageously, different promoters such as, for example, the USP, LegB4 or DC3 promoter, and different terminator sequences can be used in the expression cassette.

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However, it is also possible to use only one type of promoter in the cassette, which, however, may lead to undesired recombination events.

As described above, the transcription of the genes which have been introduced should advantageously be terminated by suitable terminator sequences at the 3' end of the biosynthesis genes which have been introduced (behind the stop codon). An example of a sequence which can be used in this context is the OCS1 terminator sequence. As
5 is the case with the promoters, different terminator sequences should be used for each gene.

As described above, the gene construct can also comprise further genes to be introduced into the plants. It is possible and advantageous to introduce into the host
10 plants, and to express, regulatory genes such as genes for inducers, repressors or enzymes which, owing to their enzyme activity, engage in the regulation of one or more genes of a biosynthesis pathway. These genes can be of heterologous or of homologous origin.

15 Moreover, further biosynthesis genes of the fatty acid or lipid metabolism can advantageously be present in the nucleic acid construct, or gene construct; however, these genes can also be present on one or more further nucleic acid constructs. A biosynthesis gene of the fatty acid or lipid metabolism which is preferably chosen is a gene from the group consisting of acyl-CoA dehydrogenase(s), acyl-ACP [= acyl carrier
20 protein] desaturase(s), acyl-ACP thioesterase(s), fatty acid acyl transferase(s), acyl-CoA:lysophospholipid acyltransferases, fatty acid synthase(s), fatty acid hydroxylase(s), acetyl-coenzyme A carboxylase(s), acyl-coenzyme A oxidase(s), fatty acid desaturase(s), fatty acid acetylenases, lipoxygenases, triacylglycerol lipases, allenoxide synthases, hydroperoxide lyases or fatty acid elongase(s) or combinations
25 thereof.

Especially advantageous nucleic acid sequences are biosynthesis genes of the fatty acid or lipid metabolism selected from the group of the acyl-CoA:lysophospholipid acyltransferase, ω 3-desaturase, Δ 8-desaturase, Δ 4-desaturase, Δ 9-desaturase, Δ 5-
30 elongase and/or Δ 9-elongase.

In this context, the abovementioned nucleic acids or genes can be cloned into expression cassettes, like those mentioned above, in combination with other elongases and desaturases and used for transforming plants with the aid of *Agrobacterium*.

35 Here, the regulatory sequences or factors can, as described above, preferably have a positive effect on, and thus enhance, the gene expression of the genes which have been introduced. Thus, enhancement of the regulatory elements can advantageously

take place at the transcriptional level by using strong transcription signals such as promoters and/or enhancers. However, an enhanced translation is also possible, for example by improving the stability of the mRNA. In principle, the expression cassettes can be used directly for introduction into the plants or else be introduced into a vector.

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These advantageous vectors, preferably expression vectors, comprise the nucleic acids which encode the $\Delta 12$ -desaturases, $\Delta 6$ -desaturases, $\Delta 5$ -elongases, $\Delta 6$ -elongases or $\Delta 5$ -desaturases and which are used in the process, or else a nucleic acid construct which comprises the nucleic acid used either alone or in combination with
10 further biosynthesis genes of the fatty acid or lipid metabolism such as the acyl-CoA:lysophospholipid acyltransferases, $\omega 3$ -desaturases, $\Delta 8$ -desaturases, $\Delta 9$ -desaturases, $\omega 3$ -desaturases, $\Delta 4$ -desaturases, $\Delta 5$ -elongases and/or $\Delta 9$ -elongases.

As used in the present context, the term "vector" refers to a nucleic acid molecule
15 which is capable of transporting another nucleic acid to which it is bound. One type of vector is a "plasmid", a circular double-stranded DNA loop into which additional DNA segments can be ligated. A further type of vector is a viral vector, it being possible for additional DNA segments to be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they have been introduced
20 (for example bacterial vectors with bacterial replication origin). Other vectors are advantageously integrated into the genome of a host cell when they are introduced into the host cell, and thus replicate together with the host genome. Moreover, certain vectors can govern the expression of genes with which they are in operable linkage. These vectors are referred to in the present context as "expression vectors". Usually,
25 expression vectors which are suitable for DNA recombination techniques take the form of plasmids. In the present description, "plasmid" and "vector" can be used exchangeably since the plasmid is the form of vector which is most frequently used. However, the invention is also intended to cover other forms of expression vectors, such as viral vectors, which exert similar functions. Furthermore, the term "vector" is
30 also intended to encompass other vectors with which the skilled worker is familiar, such as phages, viruses such as SV40, CMV, TMV, transposons, IS elements, phasmids, phagemids, cosmids, linear or circular DNA.

The recombinant expression vectors advantageously used in the process comprise the
35 nucleic acids or the described gene construct used in accordance with the invention in a form which is suitable for expressing the nucleic acids used in a host cell, which means that the recombinant expression vectors comprise one or more regulatory sequences, selected on the basis of the host cells used for the expression, which

regulatory sequence(s) is/are linked functionally with the nucleic acid sequence to be expressed. In a recombinant expression vector, "linked functionally" or "in operable linkage" means that the nucleotide sequence of interest is bound to the regulatory sequence(s) in such a way that the expression of the nucleotide sequence is possible and they are bound to each other in such a way that both sequences carry out the predicted function which is ascribed to the sequence (for example in an in-vitro transcription/translation system, or in a host cell if the vector is introduced into the host cell).

- 10 The term "regulatory sequence" is intended to comprise promoters, enhancers and other expression control elements (for example polyadenylation signals). These regulatory sequences are described, for example, in Goeddel: Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, CA (1990), or see: Gruber and Crosby, in: Methods in Plant Molecular Biology and Biotechnology, 15 CRC Press, Boca Raton, Florida, Eds.: Glick and Thompson, Chapter 7, 89-108, including the references cited therein. Regulatory sequences comprise those which govern the constitutive expression of a nucleotide sequence in many types of host cell and those which govern the direct expression of the nucleotide sequence only in specific host cells under specific conditions. The skilled worker knows that the design 20 of the expression vector can depend on factors such as the choice of host cell to be transformed, the desired expression level of the protein and the like.

- In a further embodiment of the process, the $\Delta 12$ -desaturases, $\Delta 6$ -desaturases, $\Delta 5$ -elongases, $\Delta 6$ -elongases and/or $\Delta 5$ -desaturases can be expressed in single-celled 25 plant cells (such as algae), see Falciatore et al., 1999, Marine Biotechnology 1 (3):239-251 and references cited therein, and in plant cells from higher plants (for example spermatophytes such as arable crops). Examples of plant expression vectors comprise those which are described in detail in: Becker, D., Kemper, E., Schell, J., and Masterson, R. (1992) "New plant binary vectors with selectable markers located 30 proximal to the left border", Plant Mol. Biol. 20:1195-1197; and Bevan, M.W. (1984) "Binary Agrobacterium vectors for plant transformation", Nucl. Acids Res. 12:8711-8721; Vectors for Gene Transfer in Higher Plants; in: Transgenic Plants, Vol. 1, Engineering and Utilization, Eds.: Kung and R. Wu, Academic Press, 1993, p. 15-38.

- 35 A plant expression cassette preferably comprises regulatory sequences which are capable of governing the expression of genes in plant cells and which are linked functionally so that each sequence can fulfill its function, such as transcriptional termination, for example polyadenylation signals. Preferred polyadenylation signals are

those which are derived from *Agrobacterium tumefaciens* T-DNA, such as gene 3 of the Ti plasmid pTiACH5 (Gielen et al., EMBO J. 3 (1984) 835 et seq.), which is known as octopine synthase, or functional equivalents thereof, but all other terminator sequences which are functionally active in plants are also suitable.

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Since the regulation of plant gene expression is very often not limited to the transcriptional level, a plant expression cassette preferably comprises other sequences which are linked functionally, such as translation enhancers, for example the overdrive sequence, which enhances the tobacco mosaic virus 5'-untranslated leader sequence, which increases the protein/RNA ratio (Gallie et al., 1987, Nucl. Acids Research 15:8693-8711).

As described above, the gene to be expressed must be linked functionally with a suitable promoter which triggers gene expression with the correct planning or in a cell- or tissue-specific manner. Utilizable promoters are constitutive promoters (Benfey et al., EMBO J. 8 (1989) 2195-2202), such as those which are derived from plant viruses, such as 35S CaMV (Franck et al., Cell 21 (1980) 285-294), 19S CaMV (see also US 5352605 and WO 84/02913), or constitutive plant promoters, such as the promoter of the Rubisco small subunit, which is described in US 4,962,028.

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As described above, plant gene expression can also be achieved via a chemically inducible promoter (see a review in Gatz 1997, Annu. Rev. Plant Physiol. Plant Mol. Biol., 48:89-108). Chemically inducible promoters are particularly suitable when it is desired that the gene expression takes place in a time-specific manner. Examples of such promoters are a salicylic-acid-inducible promoter (WO 95/19443), a tetracyclin-inducible promoter (Gatz et al. (1992) Plant J. 2, 397-404) and an ethanol-inducible promoter.

Promoters which respond to biotic or abiotic stress conditions are also suitable, for example the pathogen-induced PRP1 gene promoter (Ward et al., Plant. Mol. Biol. 22 (1993) 361-366), the heat-inducible tomato hsp80 promoter (US 5,187,267), the chill-inducible potato alpha-amylase promoter (WO 96/12814) or the wound-inducible pinII promoter (EP-A-0 375 091).

Especially preferred are those promoters which bring about the gene expression in tissues and organs in which the biosynthesis of fatty acids, lipids and oils takes place, in seed cells, such as cells of the endosperm and of the developing embryo. Suitable promoters are the oilseed rape napin promoter (US 5,608,152), the linseed Conlinin

promoter (WO 02/102970), the *Vicia faba* USP promoter (Baeumlein et al., Mol Gen Genet, 1991, 225 (3):459-67), the *Arabidopsis oleosin* promoter (WO 98/45461), the *Phaseolus vulgaris* phaseolin promoter (US 5,504,200), the *Brassica Bce4* promoter (WO 91/13980) or the legume B4 promoter (LeB4; Baeumlein et al., 1992, Plant
5 Journal, 2 (2):233-9), and promoters which bring about the seed-specific expression in monocotyledonous plants such as maize, barley, wheat, rye, rice and the like. Suitable noteworthy promoters are the barley lpt2 or lpt1 gene promoter (WO 95/15389 and WO 95/23230) or the promoters from the barley hordein gene, the rice glutelin gene, the rice oryzin gene, the rice prolamine gene, the wheat gliadine gene, the wheat glutelin
10 gene, the maize zeine gene, the oat glutelin gene, the sorghum kasirin gene or the rye secalin gene, which are described in WO 99/16890.

Other promoters which are also particularly suitable are those which bring about the plastid-specific expression, since plastids constitute the compartment in which
15 precursors and some end products of lipid biosynthesis are synthesized. Suitable promoters are the viral RNA polymerase promoter, described in WO 95/16783 and WO 97/06250, and the *Arabidopsis clpP* promoter, described in WO 99/46394.

In particular, it may be desired to bring about the multiparallel expression of the $\Delta 12$ -
20 desaturases, $\Delta 6$ -desaturases, $\Delta 5$ -elongases, $\Delta 6$ -elongases and/or $\Delta 5$ -desaturases used in the process. Such expression cassettes can be introduced via the simultaneous transformation of a plurality of individual expression constructs or, preferably, by combining a plurality of expression cassettes on one construct. Also, a plurality of vectors can be transformed with in each case a plurality of expression
25 cassettes and then transferred into the host cell.

Other preferred sequences for the use in operable linkage in plant gene expression cassettes are targeting sequences which are required for targeting the gene product into its corresponding cell compartment, for example into the vacuole, the nucleus, all
30 types of plastids, such as amyloplasts, chloroplasts, chromoplasts, the extracellular space, the mitochondria, the endoplasmic reticulum, elaioplasts, peroxisomes and other compartments of plant cells (see a review in Kermode, Crit. Rev. Plant Sci. 15, 4 (1996) 285-423 and references cited therein).

35 The process according to the invention employs the nucleic acid sequences with the SEQ ID NO: 11, SEQ ID NO: 27, SEQ ID NO: 193, SEQ ID NO: 195, SEQ ID NO: 197, SEQ ID NO: 199, SEQ ID NO: 201 or their derivatives or homologs which encode polypeptides which retain the enzymatic activity of the proteins encoded by nucleic acid

sequences. These sequences, individually or in combination with the nucleic acid sequences which encode the other enzymes used, are cloned into expression constructs and used for the transformation into, and expression in, plants. Owing to their construction, these expression constructs make possible an advantageous
5 optimal synthesis of the polyunsaturated fatty acids produced in the process according to the invention.

In a preferred embodiment, the process furthermore comprises the step of obtaining a cell or an intact plant which comprises the nucleic acid sequences used in the process,
10 where the cell and/or the plant is transformed with a nucleic acid sequence encoding a polypeptide with a $\Delta 12$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 6$ -elongase activity, a gene construct or a vector as described above, alone or in combination with further nucleic acid sequences which encode proteins of the fatty acid or lipid metabolism. The resulting cell is advantageously a cell of an oil-producing
15 organism such as an oil crop, such as, for example, peanut, oilseed rape, canola, linseed, hemp, peanut, soybean, safflower, hemp, mustard, sunflowers or borage.

For the purposes of the invention, "transgenic" or "recombinant" means with regard to, for example, a nucleic acid sequence, an expression cassette (= gene construct) or a
20 vector comprising the nucleic acid sequence according to the invention or an organism transformed with the nucleic acid sequences, expression cassettes or vectors according to the invention, all those constructions brought about by recombinant methods in which either

- 25 a) the nucleic acid sequence according to the invention, or
- b) a genetic control sequence which is operably linked with the nucleic acid sequence according to the invention, for example a promoter, or
- 30 c) a) and b)

are not located in their natural genetic environment or have been modified by recombinant methods, it being possible for the modification to take the form of, for example, a substitution, addition, deletion, inversion or insertion of one or more
35 nucleotide residues. The natural genetic environment is understood as meaning the natural genomic or chromosomal locus in the original organism or the presence in a genomic library. In the case of a genomic library, the natural genetic environment of the nucleic acid sequence is preferably retained, at least in part. The environment flanks

the nucleic acid sequence at least on one side and has a sequence length of at least 50 bp, preferably at least 500 bp, especially preferably at least 1000 bp, most preferably at least 5000 bp. A naturally occurring expression cassette – for example the naturally occurring combination of the natural promoter of the nucleic acid sequences used in the process according to the invention with the corresponding $\Delta 12$ -desaturase, $\Delta 4$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -elongase and/or $\Delta 5$ -elongase genes – becomes a transgenic expression cassette when this expression cassette is modified by non-natural, synthetic ("artificial") methods such as, for example, mutagenic treatment. Suitable methods are described, for example, in US 5,565,350 or WO 00/15815.

Transgenic plants for the purposes of the invention is therefore understood as meaning that the nucleic acids used in the process are not at their natural locus in the genome of the plant, it being possible for the nucleic acids to be expressed homologously or heterologously. However, transgenic also means that, while the nucleic acids according to the invention are at their natural position in the genome of the plant, however, the sequence having been modified with regard to the natural sequence, and/or that the regulatory sequences of the natural sequences have been modified. Transgenic is preferably understood as meaning the expression of the nucleic acids according to the invention or the nucleic acid sequences used in the process according to the invention at an unnatural locus in the genome, i.e. homologous or, preferably, heterologous expression of the nucleic acids takes place. Preferred transgenic plants are oilseed or oil fruit crops.

Plants which are suitable for use in the process according to the invention are, in principle, advantageously all plants which are capable of synthesizing fatty acids, specifically unsaturated fatty acids such as ARA, EPA and/or DHA, and which are suitable for the expression of recombinant genes. Examples are plants such as Arabidopsis, Asteraceae such as Calendula or crop plants such as soybean, peanut, castor-oil plant, sunflower, maize, cotton, flax, oilseed rape, coconut, oil palm, safflower (*Carthamus tinctorius*) or cacao bean. Plants which are naturally capable of synthesizing large amounts of oils are preferred, such as soybean, oilseed rape, Camelina, Indian mustard, coconut, oil palm, safflower (*Carthamus tinctorius*), flax, hemp, castor-oil plant, Calendula, peanut, cacao bean or sunflower or yeast such as *Saccharomyces cerevisiae*, with soybean, flax, oilseed rape, safflower, sunflower, Camelina, indian mustard or Calendula being especially preferred.

Further host cells which can be used for cloning the nucleic acid sequences used in the process according to the invention are detailed in: Goeddel, Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, CA (1990).

- 5 Expression strains which can be used, for example those with a lower protease activity, are described in: Gottesman, S., Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, California (1990) 119-128.

10 These include plant cells and certain tissues, organs and parts of plants in all their phenotypic forms such as anthers, fibers, root hairs, stalks, embryos, calli, cotyledons, petioles, harvested material, plant tissue, reproductive tissue and cell cultures which is derived from the actual transgenic plant and/or can be used for bringing about the transgenic plant.

- 15 Transgenic plants or advantageously the seeds thereof which comprise the polyunsaturated fatty acids in particular ARA, EPA and/or DHA, synthesized in the process according to the invention can advantageously be marketed directly without there being any need for the oils, lipids or fatty acids synthesized to be isolated. Plants for the process according to the invention are as meaning intact plants and all plant
20 parts, plant organs or plant parts such as leaf, stem, seeds, root, tubers, anthers, fibers, root hairs, stalks, embryos, calli, cotyledons, petioles, harvested material, plant tissue, reproductive tissue and cell cultures which are derived from the actual transgenic plant and/or can be used for bringing about the transgenic plant. In this context, the seed comprises all parts of the seed such as the seed coats, epidermal
25 cells, seed cells, endosperm or embryonic tissue.

In principle, the process according to the invention is also suitable for the production of polyunsaturated fatty acids, in particular ARA, EPA and/or DHA, in plant cell cultures, followed by obtaining the fatty acids from the cultures. In particular, they may take the
30 form of suspension or callus cultures.

However, the compound produced in the process according to the invention can also be isolated from the plants, advantageously the plant seeds, in the form of their oils, fat, lipids and/or free fatty acids. Polyunsaturated fatty acids produced by this process,
35 in particular ARA, EPA and/or DHA, can be harvested by harvesting the plants or plant seeds either from the culture in which they grow, or from the field.

In a further preferred embodiment, this process furthermore comprises the step of obtaining the oils, lipids or free fatty acids from the plant or from the crop. The crop may, for example, take the form of a greenhouse- or field-grown plant crop.

- 5 The oils, lipids or free fatty acids can be isolated via pressing or extraction of the plant parts, preferably the plant seeds. In this context, the oils, fats, lipids and/or free fatty acids can be obtained by what is known as cold-beating or cold-pressing without applying heat. To allow for greater ease of disruption of the plant parts, specifically the seeds, they are previously comminuted, steamed or roasted. The seeds which have
10 been pretreated in this manner can subsequently be pressed or extracted with solvents such as warm hexane. The solvent is subsequently removed.

- Thereafter, the resulting products which comprise the polyunsaturated fatty acids are processed further, i.e. refined. In this process, substances such as the plant mucilages
15 and suspended matter are first removed. What is known as desliming can be effected enzymatically or, for example, chemico-physically by addition of acid such as phosphoric acid. Thereafter, the free fatty acids are removed by treatment with a base, for example sodium hydroxide solution. The resulting product is washed thoroughly with water to remove the alkali remaining in the product and then dried. To remove the
20 pigment remaining in the product, the products are subjected to bleaching, for example using fuller's earth or active charcoal. At the end, the product is deodorized, for example using steam.

- The PUFAs or LCPUFAs produced by this process are preferably C₁₈-, C₂₀- or C₂₂-fatty acid molecules, advantageously C₂₀- or C₂₂-fatty acid molecules, with at least two
25 double bonds in the fatty acid molecule, preferably with three, four, five or six double bonds, especially preferably with four, five or six double bonds. These C₁₈-, C₂₀- or C₂₂-fatty acid molecules can be isolated from the plant in the form of an oil, a lipid or a free fatty acid. Examples of suitable plants are those mentioned above. Suitable organisms
30 are transgenic plants.

- One embodiment of the invention are therefore oils, lipids or fatty acids or fractions thereof which have been prepared by the above-described process, especially preferably oils, lipids or a fatty acid composition which comprise PUFAs and originate
35 from transgenic plants.

The fatty acids obtained in the process are also suitable as starting material for the chemical synthesis of products of value. For example, they can be used together or

alone for the production of pharmaceuticals, foodstuffs, feedstuffs or cosmetics.

As described above, these oils, lipids or fatty acids advantageously comprise 6 to 15% of palmitic acid, 1 to 6% of stearic acid, 7-85% of oleic acid, 0.5 to 8% of vaccenic acid, 0.1 to 1% of arachic acid, 7 to 25% of saturated fatty acids, 8 to 85% of monounsaturated fatty acids and 60 to 85% of polyunsaturated fatty acids, in each case based on 100% and on the total fatty acid content of the organisms.

Advantageous polyunsaturated fatty acids which are present in the fatty acid esters or fatty acid mixtures are preferably at least 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 0.7, 0.8, 0.9 or 1% of arachidonic acid, based on the total fatty acid content. Moreover, the fatty acid esters or fatty acid mixtures which have been produced by the process of the invention advantageously comprise fatty acids selected from the group of the fatty acids erucic acid (13-docosaenoic acid), sterculic acid (9,10-methyleneoctadec-9-enoic acid), malvalic acid (8,9-methyleneheptadec-8-enoic acid), chaulmoogric acid

(cyclopentenododecanoic acid), furan fatty acid (9,12-epoxyoctadeca-9,11-dienoic acid), vernolic acid (9,10-epoxyoctadec-12-enoic acid), tariric acid (6-octadecynoic acid), 6-nonadecynoic acid, santalbic acid (11-octadecen-9-ynoic acid), 6,9-octadecenynoic acid, pyruvic acid (10-heptadecen-8-ynoic acid), crepenyninic acid (9-octadecen-12-ynoic acid), 13,14-dihydrooropheic acid, octadecen-13-ene-9,11-diynoic acid, petroselenic acid (cis-6-octadecenoic acid), 9c,12t-octadecadienoic acid, calendulic acid (8t10t12c-octadecatrienoic acid), catalpic acid (9t11t13c-octadecatrienoic acid), eleostearic acid (9c11t13t-octadecatrienoic acid), jacaric acid (8c10t12c-octadecatrienoic acid), punitic acid (9c11t13c-octadecatrienoic acid), parinaric acid (9c11t13t15c-octadecatetraenoic acid), pinolenic acid (all-cis-5,9,12-octadecatrienoic acid), laballenic acid (5,6-octadecadienallenic acid), ricinoleic acid (12-hydroxyoleic acid) and/or coriolic acid (13-hydroxy-9c,11t-octadecadienoic acid).

The abovementioned fatty acids are, as a rule, advantageously only found in traces in the fatty acid esters or fatty acid mixtures produced by the process according to the invention, that is to say that, based on the total fatty acids, they occur to less than 30%, preferably to less than 25%, 24%, 23%, 22% or 21%, especially preferably to less than 20%, 15%, 10%, 9%, 8%, 7%, 6% or 5%, very especially preferably to less than 4%, 3%, 2% or 1%. In a further preferred form of the invention, these abovementioned fatty acids occur in amounts of less than 0.9%, 0.8%, 0.7%, 0.6% or 0.5%, especially preferably less than 0.4%, 0.3%, 0.2%, 0.1%, based on the total fatty acids. The fatty acid esters or fatty acid mixtures produced by the process according to the invention advantageously comprise less than 0.1%, based on the total fatty acids, and/or no butyric acid, no cholesterol, no clupanodonic acid (= docosapentaenoic acid, C22:5^{Δ4,8,12,15,21}) and no nisinic acid (tetracosahexaenoic acid, C23:6^{Δ3,8,12,15,18,21}).

As a rule, the abovementioned fatty acids are advantageously only found in traces in the fatty acid esters or fatty acid mixtures produced by the process according to the invention, that is to say that, based on the total fatty acids, they are found in amounts of less than 30%, preferably less than 25%, 24%, 23%, 22% or 21%, especially preferably less than 20%, 15%, 10%, 9%, 8%, 7%, 6% or 5%, very especially preferably less than 4%, 3%, 2% or 1%. In a further preferred embodiment of the invention, these abovementioned fatty acids are found relative to the total fatty acids in amounts of less than 0.9%, 0.8%, 0.7%, 0.6% or 0.5%, especially preferably less than 0.4%, 0.3%, 0.2%, 0.1%. The fatty acid esters or fatty acid mixtures produced by the process according to the invention advantageously comprise less than 0.1% based on the total fatty acids and/or no butyric acid, no cholesterol, no clupanodonic acid (= docosapentaenoic acid, C22:5^{Δ4,8,12,15,21}) and no nisinic acid (tetracosahexaenoic acid, C23:6^{Δ3,8,12,15,18,21}).

The oils, lipids or fatty acids according to the invention advantageously comprise at least 0.5%, 1%, 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9% or 10%, advantageously at least 11%, 12%, 13%, 14%, 15%, 16% or 17%, especially advantageously at least 18%, 19%, 20%, 21%, 22%, 23%, 24% or 25% of ARA or at least 0.5%, 1%, 2%, 3%, 4%, 5% or 6%, advantageously at least 7%, 8%, 9%, 10% or 11%, especially advantageously at least 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19% or 20% of EPA or at least 0.01%, 0.02%, 0.03%, 0.04% or 0.05% or 0.06%, advantageously at least 0.07%, 0.08%, 0.09% or 0.1%, especially advantageously at least 0.2%, 0.3% or 0.4% of DHA, based on the total fatty acid content of the production organism, advantageously of a plant, especially advantageously of an oil crop such as soybean, oilseed rape, coconut, oil palm, safflower, flax, hemp, castor-oil plant, Calendula, peanut, cacao bean, sunflower or the abovementioned other monocotyledonous or dicotyledonous oil crops. All percentages are by weight.

Owing to the nucleic acid sequences according to the invention, or the nucleic acid sequences used in the process according to the invention, it is possible to obtain an increase in the yield of polyunsaturated fatty acids, mainly ARA and EPA, but also DHA, of at least 50, 80 or 100%, advantageously at least 150, 200 or 250%, especially advantageously at least 300, 400, 500, 600, 700, 800 or 900%, very advantageously at least 1000, 1100, 1200, 1300, 1400 or 1500% in comparison with the non-transgenic starting plant, for example a plant such as *Brassica juncea*, *Brassica napus*, *Camelina sativa*, *Arabidopsis thaliana* or *Linum usitatissimum* when using a GC analysis for comparison purposes, see Examples.

The lipids and/or oils produced in the process according to the invention have a higher content of the unsaturated fatty acids oleic acid, linoleic acid and α -linolenic acid in the sn2-position in comparison with the other positions sn1 and sn3. A higher content is understood as meaning ratios of (sn1:sn2:sn3) 1:1.1:1, 1:1.5:1 to 1:3:1. Also, the arachidonic acid, eicosapentaenoic acid or docosahexaenoic acid produced in the process likewise show, in the lipids and/or oils, a preference for the sn2-position in the triglyceride in comparison with the positions sn1 and sn3 of advantageously 1:1.1:1, 1:1.5:1 to 1:3:1.

As described above, the polyunsaturated C₂₀- and/or C₂₂-fatty acids, produced in the process, with four, five or six double bonds in the molecule will in the seed of plants which comprise no, or only very small amounts, of C_{12:0}- or C_{14:0}-fatty acids. Even shorter saturated fatty acids such as the fatty acids C_{4:0}, C_{6:0}, C_{8:0} or C_{10:0}, too, should not be present in the lipid and/or oil, or only in small amounts. Only small amounts are understood as meaning, advantageously, amounts which, when analyzed by GC, advantageously amount to less than 5, 4, 3, 2 or 1%, advantageously less than 0.9, 0.8, 0.7, 0.6 or 0.5%, especially advantageously less than 0.4, 0.3, 0.2 or 0.1%, very especially preferably less than 0.09, 0.08, 0.07, 0.06, 0.05, 0.04, 0.03, 0.02 or 0.01 units GC peak area. The fatty acid C_{16:0} should advantageously be in the range of from 1 to 28% units GC peak area. Advantageously, the fatty acid C_{16:0} should be present in amounts of less than 25%, 20%, 15% or 10%, advantageously less than 9%, 8%, 7%, 6% or 5%, especially advantageously of less than 4%, 3%, 2% or 1% units GC peak area or not at all in the lipids, oils and/or free fatty acids. The fatty acid C_{16:1} should advantageously amount to less than 1, 0.5, 0.4, 0.3, 0.2 or 0.1%, especially advantageously 0.09, 0.08, 0.07, 0.06, 0.05, 0.04, 0.03, 0.02 or 0.01 units GC peak area. Very especially preferably, the fatty acid C_{16:1} should not be present in the oils and/or lipids produced in the process. The same applies to the fatty acids C_{15:0}, C_{17:0}, C_{16:1}^{Δ3}, trans, C_{16:4}^{Δ4,7,10,13} and C_{18:5}^{Δ3,6,9,12,15}. Besides oleic acid (C_{18:1}^{Δ9}), the isomers (C_{18:1}^{Δ7}, C_{18:1}^{Δ11}) may also be present in the lipids, oils or free fatty acids. Advantageously in amounts of less than 5%, 4%, 3%, 2% or 1%, measured as units GC peak area. Each of the fatty acids C_{20:0}, C_{20:1}, C_{24:0} and C_{24:1} should be present in a range of from 0 to 1%, 0 to 3% and 0 to 5% units GC peak area, respectively. Moreover, little dihomog- γ -linolenic acid (= DGLA) in terms of units GC peak area should be detectable in the seed oil and/or seed lipid in the GC analysis. Little is understood as meaning less than 2, 1.9, 1.8, 1.7, 1.6 and 1.5%, advantageously less than 1.4, 1.3, 1.2, 1.1 or 1%, especially advantageously less than 0.9, 0.8, 0.7, 0.6, 0.5 or 0.4% in terms of units GC peak area.

In a preferred embodiment of the process, DGLA and ARA should be produced in a ratio of from 1:1 up to 1:100, advantageously 1:2 up to 1:80, especially advantageously 1:3 up to 1:70, very especially preferably 1:5 up to 1:60.

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In a further preferred embodiment of the process, DGLA and EPA should be produced in a ratio of from 1:1 up to 1:100, advantageously 1:2 up to 1:80, especially advantageously 1:3 up to 1:70, very especially preferably 1:5 up to 1:60.

- 10 The lipids, oils and/or free fatty acids produced in the process according to the invention should advantageously have a high content of unsaturated fatty acids, advantageously of polyunsaturated acids, of at least 30, 40 or 50% by weight, advantageously of at least 60, 70 or 80% by weight, based on the total fatty acid content in the seeds of the transgenic plants.

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All saturated fatty acids together should advantageously only account for a small amount in the lipids, oils and/or free fatty acids, preferably used plants. In this context, a small amount is understood as meaning an amount of less than 15%, 14%, 13%, 12%, 11% or 10%, preferably less than 9%, 8%, 7% or 6% in units GC peak area.

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Lipids, oils and/or free fatty acids produced in the process should advantageously have an erucic acid content of less than 2% by weight based on the total fatty acid content of the plant. Advantageously, no erucic acid should be present in the lipids and/or oils.

- Also, the content of saturated fatty acids C16:0 and/or C18:0 should advantageously
25 be less than 19, 18, 17, 16, 15, 14, 13, 12, 11 or 10% by weight, advantageously less than 9, 8, 7, 6 or 5% by weight, based on the total fatty acid content of the lipids and/or oils. Also, longer fatty acids such as C20:0 or C22:1 should not be present at all or only in small amounts of advantageously less than 4, 3, 2 or 1% by weight, advantageously less than 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2 or 0.1% by weight, based on the total fatty acid content of the lipids and/or oils. Typically, no, or only small amounts, of C16:1 are
30 present as fatty acid in the lipids and/or oils produced in the process according to the invention. Small amounts are advantageously understood as meaning fatty acid contents of less than 4, 3, 2 or 1% by weight, advantageously less than 0.9, 0.8, 0.7, 0.6, 0.5, 0.4, 0.3, 0.2 or 0.1% by weight, based on the total fatty acid content of the
35 lipids and/or oils.

The oils, lipids, fatty acids or fatty acid mixtures according to the invention which are obtained after pressing are referred to as what is known as crude oils. They still

comprise all of the oil and/or lipid contents and also compounds which are soluble in these. Such compounds are the various tocopherols such as α -tocopherol, β -tocopherol, γ -tocopherol and/or δ -tocopherol or phytosterols such as brassicasterol, campesterol, stigmasterol, β -sitosterol, sitostanol, Δ^5 -avenasterol, Δ^5 ,24-stigmastadienol, Δ^7 -stigmasternol or Δ^7 -avenasterol. These compounds are present in a range of from 1 to 1000 mg/100 g, advantageously 10 to 800 mg/100 g of lipid or oil. Triterpenes such as germaniol, amyrin, cycloartenol and others may also be present in these lipids and oils. These lipids and/or oils comprise the polyunsaturated fatty acids produced in the process, such as ARA, EPA and/or DHA, bound in polar and unpolar lipids such as phospholipids, for example phosphatidylcholine, phosphatidylethanolamine, phosphatidylinositol, phosphatidylserine, phosphatidylglycerol, galactolipids, monoglycerides, diglycerides or triglycerides, to mention but a few. Lysophospholipids may also be present in the lipids and/or oils. These components of the lipids and/or oils can be separated from one another by suitable processes. Cholesterol is not present in these crude oils.

A further embodiment according to the invention is the use of the oil, lipid, fatty acids and/or the fatty acid composition in feedstuffs, foodstuffs, cosmetics or pharmaceuticals. The oils, lipids, fatty acids or fatty acid mixtures according to the invention can be used in the manner with which the skilled worker is familiar for mixing with other oils, lipids, fatty acids or fatty acid mixtures of animal origin such as, for example, fish oils. Typical of such fish oils short-chain fatty acids such as C12:0, C14:0, C14:1, branched C15:0, C15:0, C16:0 or C16:1. Polyunsaturated C16-fatty acids such as C16:2, C16:3 or C16:4, branched C17:0, C17:1, branched C18:0 and C19:0 and also C19:0 and C19:1 are also found in fish oil. Such fatty acids are typical of fish oils and are only found rarely, or not at all, in vegetable oils. Economically relevant fish oils are, for example, anchovy oil, menhaden oil, tuna oil, sardine oil, herring oil, mackerel oil, whale oil and salmon oil. These lipids and/or oils of animal origin can be used for mixing with the oils according to the invention in the form of crude oils, i.e. in the form of lipids and/or oils which have not yet been purified, or else various purified fractions may be used for mixing.

A further embodiment according to the invention is the use of the oil, lipid, fatty acids and/or fatty acid compositions in feedstuffs, foodstuffs, cosmetics or pharmaceuticals.

The oils, lipids, fatty acids or fatty acid mixtures according to the invention can be used in the manner with which the skilled worker is familiar for mixing with other oils, lipids, fatty acids or fatty acid mixtures of animal origin such as, for example, fish oils. Again,

these oils, lipids, fatty acids or fatty acid mixtures, which are composed of vegetable and animal constituents, may be used for the preparation of foodstuffs, feedstuffs, cosmetics or pharmaceuticals.

- 5 The term "oil", "lipid" or "fat" is understood as meaning a fatty acid mixture comprising unsaturated or saturated, preferably esterified, fatty acid(s). The oil, lipid or fat is preferably high in polyunsaturated free or, advantageously, esterified fatty acid(s), in particular linoleic acid, γ -linolenic acid, dihomo- γ -linolenic acid, arachidonic acid, α -linolenic acid, stearidonic acid, eicosatetraenoic acid, eicosapentaenoic acid,
- 10 docosapentaenoic acid or docosahexaenoic acid. The amount of unsaturated esterified fatty acids preferably amounts to approximately 30%, a content of 50% is more preferred, a content of 60%, 70%, 80%, 85% or more is even more preferred. For the analysis, the fatty acid content can, for example, be determined by gas chromatography after converting the fatty acids into the methyl esters by
- 15 transesterification. The oil, lipid or fat can comprise various other saturated or unsaturated fatty acids, for example calendulic acid, palmitic acid, palmitoleic acid, stearic acid, oleic acid and the like. The content of the various fatty acids in the oil or fat can vary, in particular depending on the starting organism.
- 20 The polyunsaturated fatty acids with advantageously at least two double bonds which are produced in the process are, as described above, for example sphingolipids, phosphoglycerides, lipids, glycolipids, phospholipids, monoacylglycerol, diacylglycerol, triacylglycerol or other fatty acid esters.
- 25 Starting from the polyunsaturated fatty acids with advantageously at least five or six double bonds, which acids have been prepared in the process according to the invention, the polyunsaturated fatty acids which are present can be liberated for example via treatment with alkali, for example aqueous KOH or NaOH, or acid hydrolysis, advantageously in the presence of an alcohol such as methanol or ethanol,
- 30 or via enzymatic cleavage, and isolated via, for example, phase separation and subsequent acidification via, for example, H_2SO_4 . The fatty acids can also be liberated directly without the above-described processing step.

- Mosses and algae are the only known plant systems which produce substantial
- 35 amounts of polyunsaturated fatty acids such as arachidonic acid (ARA) and/or eicosapentaenoic acid (EPA) and/or docosahexaenoic acid (DHA). Mosses comprise PUFAs in membrane lipids, while algae, organisms which are related to algae and a few fungi also accumulate substantial amounts of PUFAs in the triacylglycerol fraction.

This is why nucleic acid molecules which are isolated from such strains which also accumulate PUFAs in the triacylglycerol fraction are particularly advantageous for the process according to the invention and thus for the modification of the lipid and PUFA production system in a host, in particular plants such as oil crops, for example oilseed rape, canola, linseed, hemp, soybeans, sunflowers and borage. They can therefore be used advantageously in the process according to the invention.

After their introduction into a plant cell or plant, the nucleic acids used in the process can either be present on a separate plasmid or, advantageously, integrated into the genome of the host cell. In the case of integration into the genome, integration can be random or else be effected by recombination such that the native gene is replaced by the copy introduced, whereby the production of the desired compound by the cell is modulated, or by the use of a gene in trans, so that the gene is linked operably with a functional expression unit which comprises at least one sequence which ensures the expression of a gene and at least one sequence which ensures the polyadenylation of a functionally transcribed gene. The nucleic acids are advantageously introduced into the organisms via multiexpression cassettes or constructs for multiparallel expression, advantageously into the plants for the multiparallel seed-specific expression of genes.

Naturally, the coexpression of a plurality of genes can be effected not only by introducing the genes on a shared recombinant nucleic acid construct. Rather, individual genes can also be introduced separately – simultaneously or in succession, on a variety of constructs. In this case, the simultaneous presence in the plant which coexpresses all of the genes is ensured by using different selection markers. This plant can be the product of one or more transformation procedures, or else be a hybridization product of plants comprising one or more of the genes.

Substrates which are advantageously suitable for the nucleic acids which are used in the process according to the invention and which encode polypeptides with ω 3-desaturase, Δ 4-desaturase, Δ 5-desaturase, Δ 6-desaturase, Δ 8-desaturase, Δ 12-desaturase, Δ 5-elongase, Δ 6-elongase and/or Δ 9-elongase activity and/or the further nucleic acids used, such as the nucleic acids which encode polypeptides of the fatty acid or lipid metabolism selected from the group acyl-CoA dehydrogenase(s), acyl-ACP [= acyl carrier protein] desaturase(s), acyl-ACP thioesterase(s), fatty acid acyltransferase(s), acyl-CoA:lysophospholipid acyltransferase(s), fatty acid synthase(s), fatty acid hydroxylase(s), acetyl-coenzyme A carboxylase(s), acyl-coenzyme A oxidase(s), fatty acid desaturase(s), fatty acid acetylenases, lipxygenases, triacylglycerol lipases, allenoxide synthases, hydroperoxide lyases or fatty acid elongase(s) are advantageously C₁₆-, C₁₈-, C₂₀- or C₂₂-fatty acids. The fatty

acids converted as substrates in the process are preferably converted in the form of their acyl-CoA esters and/or their phospholipid esters. It is advantageous to use, in the process, desaturases with specificity for the acyl-CoA esters. The advantage here is that a substitution between the phospholipid esters, which are generally the substrate of the desaturation, and the acyl-CoA esters, can be dispensed with. Thus, a further enzyme step which, as has been shown, is limiting in some cases, can be dispensed with.

To produce the long-chain PUFAs according to the invention, the polyunsaturated C₁₆- or C₁₈-fatty acids must first be desaturated by the enzymatic activity of a desaturase and subsequently be elongated by at least two carbon atoms via an elongase. After one elongation cycle, this enzyme activity gives C₁₈- or C₂₀-fatty acids and after two elongation cycles C₂₀- or C₂₂-fatty acids. The activity of the desaturases and elongases used in the process according to the invention preferably leads to C₁₈-, C₂₀- and/or C₂₂-fatty acids, advantageously with at least two double bonds in the fatty acid molecule, preferably with three, four, five or six double bonds, especially preferably to give C₂₀- and/or C₂₂-fatty acids with at least three double bonds in the fatty acid molecule, preferably with three, four, five or six double bonds, very specially preferably with four, five or six double bonds in the molecule. Products of the process according to the invention which are especially preferred are arachidonic acid, eicosapentaenoic acid and/or docosahexaenoic acid. The C₁₈-fatty acids with at least two double bonds in the fatty acid can be elongated by the enzymatic activity according to the invention in the form of the free fatty acid or in the form of the esters, such as phospholipids, glycolipids, sphingolipids, phosphoglycerides, monoacylglycerol, diacylglycerol or triacylglycerol.

The preferred biosynthesis site of the fatty acids, oils, lipids or fats in the plants which are advantageously used is, for example, in general the seed or cell strata of the seed, so that seed-specific expression of the nucleic acids used in the process makes sense. However, it is obvious that the biosynthesis of fatty acids, oils or lipids need not be limited to the seed tissue, but can also take place in a tissue-specific manner in all the other parts of the plant, for example in epidermal cells or in the tubers.

Owing to the use of the nucleic acids according to the invention which encode a $\Delta 5$ -elongase, the polyunsaturated fatty acids produced in the process can be increased by at least 5%, preferably by at least 10%, especially preferably by at least 20%, very especially preferably by at least 50% in comparison with the wild type of the organisms which do not comprise the nucleic acids recombinantly.

In principle, the polyunsaturated fatty acids produced by the process according to the invention in the plants used in the process can be increased in two different ways.

5 Either the pool of free polyunsaturated fatty acids and/or the content of the esterified polyunsaturated fatty acids produced via the process can be enlarged.

Advantageously, the pool of esterified polyunsaturated fatty acids in the transgenic organisms is enlarged by the process according to the invention.

10 A further subject matter according to the invention are isolated nucleic acid sequences which encode polypeptides with $\Delta 5$ -elongase, the $\Delta 5$ -elongases encoded by the nucleic acid sequences converting C_{20} -fatty acids having at least four double bonds in the fatty acid molecule; which are advantageously ultimately incorporated into diacylglycerides and/or triacylglycerides.

15 A further subject matter of the invention is thus an isolated nucleic acid sequence which encodes polypeptides with $\Delta 5$ -elongase and which has the sequence shown in SEQ ID NO: 197.

20 A further subject matter of the invention is an isolated nucleic acid sequence which encodes polypeptides with $\Delta 6$ -elongase activity and which has the sequence shown in SEQ ID NO: 199.

25 Yet a further subject matter of the invention is an isolated nucleic acid sequence which encodes polypeptides with $\Delta 6$ -desaturase activity and which has the sequence shown in SEQ ID NO: 201.

The subject matters of the invention likewise extend to a recombinant nucleic acid molecule comprising:

- 30 a) one or more copies of a promoter which is active in plant cells, preferably in seed cells,
- b) at least one nucleic acid sequence with the sequence shown in SEQ ID NO: 193 or SEQ ID NO: 201 which encodes a $\Delta 6$ -desaturase activity,
- 35 c) at least one nucleic acid sequence with the sequence shown in SEQ ID NO: 11 which encodes a $\Delta 5$ -desaturase activity,
- d) at least one nucleic acid sequence with the sequence shown in SEQ ID NO: 27 or SEQ ID NO: 199 which encodes a $\Delta 6$ -elongase activity, and
- e) one or more copies of a terminator sequence.

Advantageously, an additional nucleic acid sequence with the sequence shown in SEQ ID NO: 195 and which encodes a $\Delta 12$ -desaturase may also advantageously be present in the recombinant abovementioned nucleic acid molecule.

5

In a further advantageous embodiment, an additional nucleic acid sequence with the sequence shown in SEQ ID NO: 197 and which encodes a $\Delta 5$ -elongase may also be present in the recombinant nucleic acid molecule.

- 10 Besides these abovementioned sequences, further biosynthetic genes of the fatty acid or lipid metabolism selected from the group consisting of acyl-CoA dehydrogenase(s), acyl-ACP [= acyl carrier protein] desaturase(s), acyl-ACP thioesterase(s), fatty acid acyltransferase(s), acyl-CoA:lysophospholipid acyltransferase(s), fatty acid synthase(s), fatty acid hydroxylase(s), acetyl-coenzyme A carboxylase(s), acyl-
- 15 coenzyme A oxidase(s), fatty acid desaturase(s), fatty acid acetylenases, lipoxygenases, triacylglycerol lipases, allenoxide synthases, hydroperoxide lyases or fatty acid elongase(s) may also be introduced into the recombinant nucleic acid molecule.

- 20 These genes are by preference genes of the fatty acid or lipid metabolism selected from the group consisting of $\Delta 4$ -desaturase, $\Delta 8$ -desaturase, $\Delta 9$ -desaturase or $\Delta 9$ -elongase.

- Yet a further subject matter of the invention are gene constructs which comprise the
- 25 nucleic acid sequences SEQ ID NO: 11, SEQ ID NO: 27, SEQ ID NO: 193, SEQ ID NO: 195, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201 according to the invention, the nucleic acid being functionally linked to one or more regulatory signals.

- All of the nucleic acid sequences used in the process according to the invention are
- 30 advantageously derived from a eukaryotic organism such as a plant, a microorganism such as an alga or an animal. By preference, the nucleic acid sequences are derived from the order Salmoniformes, *Xenopus* or *Ciona*, algae such as *Mantoniella*, *Cryptothecodinium*, *Euglena* or *Ostreococcus*, fungi such as the genus *Phytophthora* or from diatoms such as the genera *Thalassiosira* or *Phaeodactylum*.

35

The nucleic acid sequences used in the process which encode proteins with $\omega 3$ -desaturase, $\Delta 4$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 9$ -desaturase, $\Delta 12$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase or $\Delta 9$ -elongase activity are

advantageously introduced by themselves or by preference in combination with an expression cassette (= nucleic acid construct) which the expression of the nucleic acids in a plant. More than one nucleic acid sequence of an enzymatic activity such as, for example, a $\Delta 12$ -desaturase, $\Delta 4$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase and/or $\omega 3$ -desaturase may be present in the nucleic acid construct.

For introduction into the plant, the nucleic acids used in the process are advantageously subjected to amplification and ligation in the known manner as described above.

A series of mechanisms exist which enable a modification of the $\Delta 12$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 4$ -desaturase, $\Delta 6$ -desaturase and/or $\omega 3$ -desaturase protein according to the invention and of the further proteins used in the process, such as the $\Delta 12$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase or $\Delta 4$ -desaturase proteins, so that the yield, production and/or production efficiency of the advantageously polyunsaturated fatty acids in a plant, preferably in an oil crop plant, can be influenced directly as the result of this modified protein. The number or activity of the $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase proteins or genes can be increased so that larger amounts of the gene products and thus ultimately larger amounts of the compounds of the general formula I are produced. A *de-novo* synthesis in a plant which had lacked the activity and ability to biosynthesize the compounds prior to the introduction of the gene(s) in question is also possible. The same also applies analogously to the combination with further desaturases or elongases or further enzymes from the fatty acid and lipid metabolism. Also, the use of different, divergent sequences, i.e. sequences which differ at the DNA sequence level, may be advantageous, or the use of promoters for gene expression which makes possible a different temporal gene expression, for example depending on the degree of maturity of a seed or oil-storing tissue.

By introducing a $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 4$ -desaturase gene into a plant alone or in combination with other genes into a cell may not only increase the biosynthetic flux towards the end product, but also increase the corresponding triacylglycerol composition or create it *de novo*. Likewise, the number or activity of other genes in the import of nutrients required for the biosynthesis of one or more fatty acids, oils, polar and/or neutral lipids may be increased, so that the concentration of

these precursors, cofactors or intermediates within the cells or within the storage compartment is increased, whereby the ability of the cells to produce PUFAs is increased further, as described hereinbelow. By optimizing the activity or increasing the number of one or more $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase genes which are involved in the biosynthesis of these compounds, or by destroying the activity of one or more genes which are involving in breaking down these compounds, it may be possible to increase the yield, production and/or production efficiency of fatty acid and lipid molecules from organisms and advantageously from plants.

The isolated nucleic acid molecules used in the process according to the invention encode proteins or parts of these, the proteins or the individual protein or parts thereof comprising an amino acid sequence with sufficient homology with an amino acid sequence which is shown in the sequences SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 70, SEQ ID NO: 72, SEQ ID NO: 74, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138, SEQ ID NO: 184, SEQ ID NO: 194, SEQ ID NO: 198, SEQ ID NO: 200 or SEQ ID NO: 202 so that the proteins or parts thereof retain a $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase activity. The proteins or parts thereof, which is/are encoded by the nucleic acid molecule(s), preferably still retain(s) its/their essential enzymatic activity and the ability of participating in the metabolism of compounds required in the formation of cell membranes or lipid bodies in organisms, advantageously in plants, or in the transport of molecules across these membranes. Advantageously, the proteins encoded by the nucleic acid molecules have at least approximately 50%, preferably at least approximately 60% and more preferably at least approximately 70%, 80% or 90% and most preferably at least approximately 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identity with the amino sequences shown in SEQ ID NO: 2, SEQ ID NO: 4, SEQ

ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 70, SEQ ID NO: 72, SEQ ID NO: 74, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138, SEQ ID NO: 184, SEQ ID NO: 194, SEQ ID NO: 198, SEQ ID NO: 200 or SEQ ID NO: 202. For the purposes of the invention, homology or homologous is understood as meaning identity or identical.

The homology was calculated over the entire amino acid or nucleic acid sequence region. A series of programs which are based on the various algorithms are available for comparing different sequences. In this context, the algorithms of Needleman and Wunsch or Smith and Waterman give especially reliable results. To carry out the sequence alignments, the program PileUp (J. Mol. Evolution., 25, 351-360, 1987, Higgins et al., CABIOS, 5 1989: 151-153) or the programs Gap and BestFit [Needleman and Wunsch (J. Mol. Biol. 48; 443-453 (1970) and Smith and Waterman (Adv. Appl. Math. 2; 482-489 (1981))], which are part of the GCG software packet [Genetics Computer Group, 575 Science Drive, Madison Wisconsin, USA 53711 (1991)], were used. The sequence homology values stated above as percentages were determined over the entire sequence region using the program GAP, with the following settings: Gap Weight: 50, Length Weight: 3, Average Match: 10.000 and Average Mismatch: 0.000. Unless otherwise specified, these settings were always used as standard settings for sequence alignments.

Essential enzymatic activity of the Δ^{12} -desaturase, ω^3 -desaturase, Δ^9 -elongase, Δ^6 -desaturase, Δ^8 -desaturase, Δ^6 -elongase, Δ^5 -desaturase, Δ^5 -elongase or Δ^4 -desaturase used in the process according to the invention is understood as meaning that, in comparison with the proteins/enzymes encoded by the sequence SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41,

SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201 and their derivatives retain at least an enzymatic activity of at least 10%, preferably 20%, especially preferably 30% and very especially 40% and can thus participate in the metabolism of compounds required in the synthesis of fatty acids, fatty acid esters such as diacylglycerides and/or triacylglycerides in an organism, advantageously a plant or plant cell, or in the transport of molecules across membranes, meaning C₁₈-, C₂₀- or C₂₂-carbon chains in the fatty acid molecule with double bonds at at least two, advantageously three, four, five or six positions.

The nucleic acids which can be used advantageously in the process are derived from bacteria, fungi, diatoms, animals such as *Caenorhabditis* or *Oncorhynchus* or plants such as algae or mosses, such as the genera *Shewanella*, *Physcomitrella*, *Thraustochytrium*, *Fusarium*, *Phytophthora*, *Ceratodon*, *Mantoniella*, *Ostreococcus*, *Isochrysis*, *Aleurita*, *Muscarioides*, *Mortierella*, *Borago*, *Phaeodactylum*, *Cryptothecodinium*, specifically from the genera and species *Oncorhynchus mykiss*, *Xenopus laevis*, *Ciona intestinalis*, *Thalassiosira pseudonona*, *Mantoniella squamata*, *Ostreococcus* sp., *Ostreococcus tauri*, *Euglena gracilis*, *Physcomitrella patens*, *Phytophthora infestans*, *Fusarium gramineum*, *Cryptocodinium cohnii*, *Ceratodon purpureus*, *Isochrysis galbana*, *Aleurita farinosa*, *Thraustochytrium* sp., *Muscarioides vialii*, *Mortierella alpina*, *Borago officinalis*, *Phaeodactylum tricornutum*, *Caenorhabditis elegans* or especially advantageously from *Oncorhynchus mykiss*, *Euglena gracilis*, *Thalassiosira pseudonona* or *Cryptothecodinium cohnii*.

As an alternative, it is possible to use, in the process according to the invention, nucleotide sequences which encode a Δ 12-desaturase, ω 3-desaturase, Δ 9-elongase, Δ 6-desaturase, Δ 8-desaturase, Δ 6-elongase, Δ 5-desaturase, Δ 5-elongase or Δ 4-desaturase and which hybridize, advantageously under stringent conditions, with a nucleotide sequence as shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID

NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID
 NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID
 NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID
 NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID
 5 NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID
 NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID
 NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID
 NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ
 ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183,
 10 SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201.

The nucleic acid sequences used in the process are advantageously introduced in an
 expression cassette which enables the expression of the nucleic acids in organisms
 such as microorganisms or plants.

15

In this context, the nucleic acid sequences which encode the $\Delta 12$ -desaturase,
 $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase,
 $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase are advantageously linked functionally
 with one or more regulatory signals to increase gene expression. These regulatory
 20 sequences should enable the targeted expression of the genes and protein expression.
 For example, this may mean, depending on the host plant, that the gene is expressed
 and/or overexpressed only after induction has taken place, or else that it is expressed
 and/or overexpressed immediately. For example, these regulatory sequences take the
 form of sequences to which inducers or repressors bind and thus regulate the
 25 expression of the nucleic acid. In addition to these new regulatory sequences, or
 instead of these sequences, the natural regulation of these sequences may still be
 present before the actual structural genes and, if appropriate, may have been
 genetically modified in such a way that the natural regulation has been switched off and
 the expression of the genes enhanced. The expression cassette (= expression
 30 construct = gene construct) may, however, also be simpler in construction, that is to
 say no additional regulatory signals were inserted before the nucleic acid sequence or
 its derivatives, and the natural promoter together with its regulation was not removed.
 Instead, the natural regulatory sequence was mutated in such a way that regulation no
 longer takes place and/or gene expression is enhanced. These modified promoters can
 35 be placed before the natural gene in order to increase the activity either in the form of
 part-sequences (= promoter with parts of the nucleic acid sequences according to the
 invention) or else alone. Moreover, the gene construct can advantageously also
 comprise one or more what are known as "enhancer sequences" in functional linkage

with the promoter, and these enable an increased expression of the nucleic acid sequence. Also, it is possible to insert additional advantageous sequences at the 3' end of the DNA sequences, such as further regulatory elements or terminators. The $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 4$ -desaturase, $\Delta 5$ -desaturase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 5$ -elongase, $\Delta 6$ -elongase and/or $\Delta 9$ -elongase genes can be present in the expression cassette (= gene construct) as one or more copies. Advantageously, only in each case one copy of the genes is present in the expression cassette. This gene construct, or the gene constructs, can be expressed together in the host organism. In this context, the gene construct(s) can be inserted in one or more vectors and be present in the cell in free form or else inserted in the genome. It is advantageous for the insertion of further genes in the host genome when the genes to be expressed are present together in one gene construct.

In this context, the regulatory sequences or factors can, as described above, preferably have a positive effect on the gene expression of the genes which have been introduced, thus increasing it. Thus, enhancement of the regulatory elements can advantageously take place at the transcription level by using strong transcription signals such as promoters and/or enhancers. Besides, however, an enhancement of the translation is also possible, for example by improving the stability of the mRNA.

Advantageous regulatory sequences for the new process are present for example in promoters such as the plant promoters CaMV/35S [Franck et al., Cell 21 (1980) 285-294], PRP 1 [Ward et al., Plant Mol. Biol. 22 (1993)], SSU, OCS, lib4, usp, STLS1, B33, nos or in the ubiquitin or phaseolin promoter. Also advantageous in this context are inducible promoters, such as the promoters described in EP-A-0 388 186 (benzylsulfonamide-inducible), Plant J. 2, 1992:397-404 (Gatz et al., tetracyclin-inducible), EP-A-0 335 528 (abscisic-acid-inducible) or WO 93/21334 (ethanol- or cyclohexenol-inducible). Further suitable plant promoters are the promoter of cytosolic FBPase or the ST-LSI promoter from potato (Stockhaus et al., EMBO J. 8, 1989, 2445), the phosphoribosyl-pyrophosphate amidotransferase promoter from Glycine max (Genbank accession No. U87999) or the node-specific promoter described in EP-A-0 249 676. Especially advantageous promoters are promoters which enable the expression in tissues which are involved in the biosynthesis of fatty acids. Very especially advantageous are seed-specific promoters such as the USP promoter in accordance with the practice, but also other promoters such as the LeB4, DC3, phaseolin or napin promoters. Further especially advantageous promoters are seed-specific promoters which can be used for monocotyledonous or dicotyledonous plants and which are described in US 5,608,152 (napin promoter from oilseed rape), WO

98/45461 (oleosin promoter from *Arabidopsis*), US 5,504,200 (phaseolin promoter from *Phaseolus vulgaris*), WO 91/13980 (Bce4 promoter from *Brassica*), by Baeumlein et al., *Plant J.*, 2, 2, 1992:233-239 (LeB4 promoter from a legume), these promoters being suitable for dicots. The following promoters are suitable for example for monocots: lpt-2
5 or lpt-1 promoter from barley (WO 95/15389) and WO 95/23230), hordein promoter from barley and other promoters which are suitable and which are described in WO 99/16890.

10 In principle, it is possible to use all natural promoters together with their regulatory sequences, such as those mentioned above, for the novel process. Likewise, it is possible and advantageous to use synthetic promoters, either additionally or alone, especially when they mediate a seed-specific expression, such as, for example, as described in WO 99/16890.

15 To obtain a particularly high PUFA content especially in transgenic plants, the PUFA biosynthesis genes should advantageously be expressed in a seed-specific manner in oilseed crops. To this end, it is possible to use seed-specific promoters or those promoters which are active in the embryo and/or in the endosperm. In principle, seed-specific promoters can be isolated both from dicotyledonous and from
20 monocotyledonous plants. Such advantageous promoters are detailed further above, for example the USP, Vicilin, Napin, Oleosin, Phaseolin, Bce4, LegB4, Lpt2, lpt1, Amy32b, Amy 6-6, Aleurain or Bce4 promoter.

Moreover, chemically inducible promoters are also advantageously useful in the
25 process according to the invention.

Further advantageous promoters which are advantageously suitable for expression in soybean are the promoters of the β -conglycinin α -subunit, of the β -conglycinin β -subunit, of the Kunitz trypsin inhibitor, of annexin, of glysinin, of albumin 2S, of
30 legumin A1, of legumin A2 and that of BD30.

Especially advantageous promoters are the USP, LegB4, Fad3, SBP, DC-3 or cruciferin820 promoter.

35 Advantageous regulatory sequences which are used for the expression of the nucleic acid sequences used in the process according to the invention are terminators for the expression advantageously in soybean are Leg2A3', Kti3', Phas3', BD30 3' or AIS3'.

Especially advantageous terminators are the A7T, OCS, LeB3T or cat terminator.

To ensure a stable integration of the biosynthetic genes in the transgenic plant over several generations, each of the nucleic acids used in the process and which encodes
5 $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 4$ -desaturase should, as described above, be under the control of its own promoter, preferably of a different promoter, since repeating sequence motifs can lead to instability of the T-DNA, or to recombination events. As described above, the gene construct can also comprise
10 further genes which are to be introduced into the plant.

In this context, the regulatory sequences or factors used advantageously for the expression of the nucleic acids used in the process according to the invention can, as described above, preferably have a positive effect on the gene expression of the genes
15 introduced.

These advantageous vectors, preferably expression vectors, comprise the nucleic acids used in the process which encode the $\Delta 12$ -desaturases, $\omega 3$ -desaturases, $\Delta 9$ -elongases, $\Delta 6$ -desaturases, $\Delta 8$ -desaturases, $\Delta 6$ -elongases, $\Delta 5$ -desaturases,
20 $\Delta 5$ -elongases or $\Delta 4$ -desaturases, or a nucleic acid construct which the used nucleic acid alone or in combination with further biosynthesis genes of the fatty acid or lipid metabolism such as the acyl-CoA:lysophospholipid acyltransferases, $\omega 3$ -desaturases, $\Delta 4$ -desaturases, $\Delta 5$ -desaturases, $\Delta 6$ -desaturases, $\Delta 8$ -desaturases, $\Delta 9$ -desaturases, $\Delta 12$ -desaturases, $\omega 3$ -desaturases, $\Delta 5$ -elongases, $\Delta 6$ -elongases and/or $\Delta 9$ -elongases.
25

As described and used in the present context, the term "vector" refers to a nucleic acid molecule which is capable of transporting another nucleic acid to which it is bound.

The recombinant expression vectors used can be designed for expressing
30 $\Delta 12$ -desaturases, $\omega 3$ -desaturases, $\Delta 9$ -elongases, $\Delta 6$ -desaturases, $\Delta 8$ -desaturases, $\Delta 6$ -elongases, $\Delta 5$ -desaturases, $\Delta 5$ -elongases and/or $\Delta 4$ -desaturases in prokaryotic or eukaryotic cells. This is advantageous since, for the sake of simplicity, intermediate steps of the vector construction are frequently carried out in microorganisms. For example, the $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase,
35 $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 4$ -desaturase genes can be expressed in bacterial cells, insect cells (using baculovirus expression vectors), yeast cells and other fungal cells (see Romanos, M.A., et al. (1992) "Foreign gene expression in yeast: a review", Yeast 8:423-488; van den Hondel, C.A.M.J.J., et

al. (1991) "Heterologous gene expression in filamentous fungi", in: More Gene Manipulations in Fungi, J.W. Bennet & L.L. Lasure, Ed., pp. 396-428: Academic Press: San Diego; and van den Hondel, C.A.M.J.J., & Punt, P.J. (1991) "Gene transfer systems and vector development for filamentous fungi, in: Applied Molecular Genetics of Fungi, Peberdy, J.F., et al., Ed., pp. 1-28, Cambridge University Press: Cambridge), algae (Falciatore et al., 1999, Marine Biotechnology. 1, 3:239-251), ciliates of the types: Holotrichia, Peritrichia, Spirotrichia, Suctoria, Tetrahymena, Paramecium, Colpidium, Glaucoma, Platyophrya, Potomacus, Desaturaseudocohnilembus, Euplotes, Engelmaniella and Stylonychia, in particular the genus Stylonychia lemnae, using vectors following a transformation process as described in WO 98/01572, and preferably in cells of multi-celled plants (see Schmidt, R. and Willmitzer, L. (1988) "High efficiency Agrobacterium tumefaciens-mediated transformation of Arabidopsis thaliana leaf and cotyledon explants" Plant Cell Rep.: 583-586; Plant Molecular Biology and Biotechnology, C Press, Boca Raton, Florida, chapter 6/7, pp. 71-119 (1993); F.F. White, B. Jenes et al., Techniques for Gene Transfer, in: Transgenic Plants, Vol. 1, Engineering and Utilization, Ed.: Kung and R. Wu, Academic Press (1993), 128-43; Potrykus, Annu. Rev. Plant Physiol. Plant Molec. Biol. 42 (1991), 205-225 (and references cited therein)). Suitable host cells are furthermore discussed in Goeddel, Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, CA (1990). As an alternative, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7-promoter regulatory sequences and T7-polymerase.

In most cases, the expression of proteins in prokaryotes, advantageously for the simple detection of the enzyme activity for example for detecting the desaturase or elongase activity, is performed using vectors comprising constitutive or inducible promoters which control the expression of fusion or nonfusion proteins. Examples of typical fusion expression vectors are pGEX (Pharmacia Biotech Inc; Smith, D.B., and Johnson, K.S. (1988) Gene 67:31-40), pMAL (New England Labs, Beverly, MA) and pRIT5 (Pharmacia, Piscataway, NJ), where glutathione S-transferase (GST), maltose-E-binding protein and protein A, respectively, are fused with the recombinant target protein.

Examples of suitable inducible nonfusion E. coli expression vectors are, inter alia, pTrc (Amann et al. (1988) Gene 69:301-315) and pET 11d (Studier et al., Gene Expression Technology: Methods in Enzymology 185, Academic Press, San Diego, California (1990) 60-89). The target gene expression of the pTrc vector is based on the transcription from a hybrid trp-lac fusion promoter by host RNA polymerase. The target

gene expression from the pET 11d vector is based on the transcription of a T7-gn10-lac fusion promoter, which is mediated by a coexpressed viral RNA polymerase (T7 gn1). This viral polymerase is provided by the host strains BL21 (DE3) or HMS174 (DE3) from a resident λ -prophage which harbors a T7 gn1 gene under the
5 transcriptional control of the lacUV 5 promoter.

The skilled worker is familiar with other vectors which are suitable in prokaryotic organisms, these vectors are, for example E. coli, pLG338, pACYC184, the pBR series such as pBR322, the pUC series such as pUC18 or pUC19, the M113mp series,
10 pKC30, pRep4, pHS1, pHS2, pPLc236, pMBL24, pLG200, pUR290, pIN-III113-B1, λ gt11 or pBdCl, in Streptomyces pIJ101, pIJ364, pIJ702 or pIJ361, in Bacillus pUB110, pC194 or pBD214, in Corynebacterium pSA77 or pAJ667.

In a further embodiment, the expression vector is a yeast expression vector. Examples
15 of vectors for expression in the yeast *S. cerevisiae* comprise pYeDesaturase1 (Baldari et al. (1987) Embo J. 6:229-234), pMFa (Kurjan and Herskowitz (1982) Cell 30:933-943), pJRY88 (Schultz et al. (1987) Gene 54:113-123) and pYES2 (Invitrogen Corporation, San Diego, CA). Vectors and processes for the construction of vectors which are suitable for use in other fungi, such as the filamentous fungi, comprise those
20 which are described in detail in: van den Hondel, C.A.M.J.J., & Punt, P.J. (1991) "Gene transfer systems and vector development for filamentous fungi, in: Applied Molecular Genetics of fungi, J.F. Peberdy et al., Ed. pp. 1-28, Cambridge University Press: Cambridge, or in: More Gene Manipulations in Fungi [J.W. Bennett & L.L. Lasure, Ed., pp. 396-428: Academic Press: San Diego]. Further suitable yeast vectors are, for
25 example, pAG-1, YEp6, YEp13 or pEMBLYe23.

As an alternative, the Δ 12-desaturases, ω 3-desaturases, Δ 9-elongases, Δ 6-desaturases, Δ 8-desaturases, Δ 6-elongases, Δ 5-desaturases, Δ 5-elongases and/or Δ 4-desaturases can be expressed in insect cells using baculovirus expression vectors.
30 Baculovirus vectors which are available for the expression of proteins in cultured insect cells (for example Sf9 cells) comprise the pAc series (Smith et al. (1983) Mol. Cell Biol. 3:2156-2165) and the pVL series (Lucklow and Summers (1989) Virology 170:31-39).

The abovementioned vectors are only a small overview of possible suitable vectors.
35 Further plasmids are known to the skilled worker and are described, for example, in: Cloning Vectors (Ed., Pouwels, P.H., et al., Elsevier, Amsterdam-New York-Oxford, 1985, ISBN 0 444 904018). Further suitable expression systems for prokaryotic and eukaryotic cells, see the chapters 16 and 17 of Sambrook, J., Fritsch, E.F., and

Maniatis, T., *Molecular Cloning: A Laboratory Manual*, 2nd edition, Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989.

To detect the enzyme activity, $\Delta 12$ -desaturases, $\omega 3$ -desaturases, $\Delta 9$ -elongases, $\Delta 6$ -desaturases, $\Delta 8$ -desaturases, $\Delta 6$ -elongases, $\Delta 5$ -desaturases, $\Delta 5$ -elongases and/or $\Delta 4$ -desaturases can be expressed in single-cell plant cells (such as algae), see Falciatore et al., 1999, *Marine Biotechnology* 1 (3):239-251 and the references cited therein, and plant cells from higher plants (for example Spermatophytes, such as arable crops). Examples of plant expression vectors comprise those which are described in detail in: Becker, D., Kemper, E., Schell, J., and Masterson, R. (1992) "New plant binary vectors with selectable markers located proximal to the left border", *Plant Mol. Biol.* 20:1195-1197; and Bevan, M.W. (1984) "Binary Agrobacterium vectors for plant transformation", *Nucl. Acids Res.* 12:8711-8721; *Vectors for Gene Transfer in Higher Plants*; in: *Transgenic Plants*, Vol. 1, Engineering and Utilization, Ed.: Kung and R. Wu, Academic Press, 1993, p. 15-38.

A plant expression cassette preferably comprises regulatory sequences which are capable of controlling the gene expression in plant cells and which are functionally linked so that each sequence can fulfill its function, such as transcriptional termination, for example polyadenylation signals. Preferred polyadenylation signals are those which are derived from *Agrobacterium tumefaciens* T-DNA, such as the gene 3 of the Ti plasmid pTiACH5, which is known as octopine synthase (Gielen et al., *EMBO J.* 3 (1984) 835 et seq.) or functional equivalents of these, but all other terminators which are functionally active in plants are also suitable.

Since plant gene expression is very often not limited to transcriptional levels, a plant expression cassette preferably comprises other functionally linked sequences such as translation enhancers, for example the overdrive sequence, which comprises the 5'-untranslated tobacco mosaic virus leader sequence, which increases the protein/RNA ratio (Gallie et al., 1987, *Nucl. Acids Research* 15:8693-8711).

As described above, plant gene expression must be functionally linked to a suitable promoter which performs the expression of the gene in a timely, cell-specific or tissue-specific manner. Promoters which can be used are constitutive promoters (Benfey et al., *EMBO J.* 8 (1989) 2195-2202) such as those which are derived from plant viruses such as 35S CAMV (Franck et al., *Cell* 21 (1980) 285-294), 19S CaMV (see also US 5352605 and WO 84/02913) or plant promoters such as the promoter of the Rubisco small subunit, which is described in US 4,962,028.

Other preferred sequences for the use in functional linkage in plant gene expression cassettes are targeting sequences which are required for targeting the gene product into its relevant cell compartment (for a review, see Kermode, Crit. Rev. Plant Sci. 15, 4
5 (1996) 285-423 and references cited therein), for example into the vacuole, the nucleus, all types of plastids, such as amyloplasts, chloroplasts, chromoplasts, the extracellular space, the mitochondria, the endoplasmic reticulum, oil bodies, peroxisomes and other compartments of plant cells.

10 As described above, plant gene expression can also be facilitated via a chemically inducible promoter (for a review, see Gatz 1997, Annu. Rev. Plant Physiol. Plant Mol. Biol., 48:89-108). Chemically inducible promoters are particularly suitable if it is desired that genes are expressed in a time-specific manner. Examples of such promoters are a salicylic-acid-inducible promoter (WO 95/19443), a tetracyclin-inducible promoter (Gatz
15 et al. (1992) Plant J. 2, 397-404) and an ethanol-inducible promoter.

Promoters which respond to biotic or abiotic stress conditions are also suitable promoters, for example the pathogen-inducible PRP1-gene promoter (Ward et al., Plant Mol. Biol. 22 (1993) 361-366), the heat-inducible hsp80 promoter from tomato
20 (US 5,187,267), the cold-inducible alpha-amylase promoter from potato (WO 96/12814) or the wound-inducible pinII promoter (EP-A-0 375 091).

The promoters which are especially preferred are those which bring about the expression of genes in tissues and organs in which fatty acid, lipid and oil biosynthesis
25 takes place, in seed cells such as the cells of endosperm and of the developing embryo. Suitable promoters are the napin gene promoters from oilseed rape (US 5,608,152), the USP promoter from Vicia faba (Baeumlein et al., Mol. Gen. Genet, 1991, 225 (3):459-67), the oleosin promoter from Arabidopsis (WO 98/45461), the phaseolin promoter from Phaseolus vulgaris (US 5,504,200), the Bce4 promoter from
30 Brassica (WO 91/13980) or the legumin B4 promoter (LeB4; Baeumlein et al., 1992, Plant Journal, 2 (2):233-9), and promoters which bring about the seed-specific expression in monocotyledonous plants such as maize, barley, wheat, rye, rice and the like. Suitable promoters to be taken into consideration are the lpt2 or lpt1 gene
35 promoter from barley (WO 95/15389 and WO 95/23230) or those which are described in WO 99/16890 (promoters from the barley hordein gene, the rice glutelin gene, the rice oryzin gene, the rice prolamin gene, the wheat gliadin gene, wheat glutelin gene, the maize zein gene, the oat glutelin gene, the sorghum kasirin gene, the rye secalin gene).

- In particular, the multiparallel expression of the $\Delta 12$ -desaturases, $\omega 3$ -desaturases, $\Delta 9$ -elongases, $\Delta 6$ -desaturases, $\Delta 8$ -desaturases, $\Delta 6$ -elongases, $\Delta 5$ -desaturases, $\Delta 5$ -elongases and/or $\Delta 4$ -desaturases may be desired. Such expression cassettes can be introduced via a simultaneous transformation of a plurality of individual expression constructs or, preferably, by combining a plurality of expression cassettes on one construct. Also, it is possible to transform a plurality of vectors with in each case a plurality of expression cassettes and to transfer them to the host cell.
- Likewise especially suitable are promoters which bring about the plastid-specific expression since plastids are the compartment in which the precursors and some end products of lipid biosynthesis are synthesized. Suitable promoters such as the viral RNA-polymerase promoter, are described in WO 95/16783 and WO 97/06250, and the *clpP* promoter from *Arabidopsis*, described in WO 99/46394.
- Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. The terms "transformation" and "transfection", conjugation and transduction, as used in the present context, are intended to comprise a multiplicity of prior-art processes for introducing foreign nucleic acid (for example DNA) into a host cell, including calcium phosphate or calcium chloride coprecipitation, DEAE-dextran-mediated transfection, lipofection, natural competence, chemically mediated transfer, electroporation or particle bombardment. Suitable methods for the transformation or transfection of host cells, including plant cells, can be found in Sambrook et al. (Molecular Cloning: A Laboratory Manual, 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989) and other laboratory manuals, such as Methods in Molecular Biology, 1995, Vol. 44, Agrobacterium protocols, Ed.: Gartland and Davey, Humana Press, Totowa, New Jersey.
- The host organisms which are advantageously used are plant cells, preferably plants or parts thereof. Especially preferred plants are plants such as oilseed plants or oil crops, which comprise large amounts of lipid compounds, such as oilseed rape, evening primrose, hemp, thistle, peanut, canola, linseed, soybean, safflower, Indian mustard, sunflower, borage or plants such as maize, wheat, rye, oats, triticale, rice, barley, cotton, cassava, pepper, Tagetes, Solanaceae plants such as potato, tobacco, eggplant and tomato, *Vicia* species, pea, alfalfa, bushy plants (coffee, cacao, tea), *Salix* species, trees (oil palm, coconut) and perennial grasses and fodder crops. Especially preferred plants according to the invention are oil crops such as soybean,

peanut, oilseed rape, canola, linseed, hemp, evening primrose, sunflower, safflower, trees (oil palm, coconut).

As described above, a further subject matter according to the invention is an isolated
5 nucleic acid sequence which encodes polypeptides with $\Delta 5$ -elongase activity and
which has the sequence shown in SEQ ID NO: 197, where the elongase encoded by
the nucleic acid sequence does not elongate C_{16} - and C_{18} -fatty acids with one double
bond. Polyunsaturated C_{18} -fatty acids with one $\Delta 6$ -double bond, or C_{22} -fatty acids, are
not converted either. Advantageously, only polyunsaturated C_{20} -fatty acids with one $\Delta 5$ -
10 double bond are elongated by the enzymatic activity. Further subject matters of the
invention are, as described above, a $\Delta 6$ -elongase, $\Delta 6$ -desaturase and a $\Delta 12$ -
desaturase.

In an advantageous embodiment, the term "nucleic acid (molecule)" as used in the
15 present text additionally comprises the untranslated sequence at the 3' and at the 5'
terminus of the coding gene region: at least 500, preferably 200, especially preferably
100 nucleotides of the sequence upstream of the 5' terminus of the coding region and
at least 100, preferably 50, especially preferably 20 nucleotides of the sequence
downstream of the 3' terminus of the coding gene region. An "isolated" nucleic acid
20 molecule is separated from other nucleic acid molecules which are present in the
natural source of the nucleic acid. An "isolated" nucleic acid preferably has no
sequences which naturally flank the nucleic acid in the genomic DNA of the organism
from which the nucleic acid is derived (for example sequences which are located at the
5' and 3' termini of the nucleic acid). In various embodiments, the isolated
25 $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -
elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase molecule can, for example,
comprise less than approximately 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of
nucleotide sequences which naturally flank the nucleic acid molecule in the genomic
DNA of the cell from which the nucleic acid is derived.

30 The nucleic acid molecules used in the process, for example a nucleic acid molecule
with a nucleotide sequence of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID
NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO:
17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27,
35 SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37,
SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47,
SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61,
SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71,

SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201 or part thereof, can be isolated using standard techniques of molecular biology and the sequence information provided herein. Also, for example a homologous sequence or homologous, conserved sequence regions at the DNA or amino acid level can be identified with the aid of comparative algorithms. These sequence regions can be used as hybridization probe and standard hybridization techniques (such as, for example, described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989) for isolating further nucleic acid sequences which are useful in the process. Moreover, a nucleic acid molecule comprising a complete sequence of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201 or part thereof can be isolated by polymerase chain reaction, where oligonucleotide primers which on the basis of this sequence or parts thereof are used (for example, a nucleic acid molecule comprising the complete sequence or part thereof can be isolated by polymerase chain reaction using oligonucleotide primers which have been generated on the basis of this very sequence). For example, mRNA can be isolated from cells (for example by the guanidinium thiocyanate extraction process by Chirgwin et al. (1979) *Biochemistry* 18:5294-5299) and cDNA can be generated by means of reverse transcriptase (for example Moloney-MLV reverse transcriptase, from Gibco/BRL, Bethesda, MD, or AMV reverse transcriptase, from Seikagaku America, Inc., St. Petersburg, FL). Synthetic oligonucleotide primers for the amplification by means of polymerase chain reaction can be generated on the basis of one of the

- sequences shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201 or with the aid of the amino acid sequences shown in SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 70, SEQ ID NO: 72, SEQ ID NO: 74, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138, SEQ ID NO: 184, SEQ ID NO: 194, SEQ ID NO: 198, SEQ ID NO: 200 or SEQ ID NO: 202. One of the abovementioned nucleic acids can be amplified in accordance with standard PCR amplification techniques using cDNA or, alternatively, genomic DNA as template and suitable oligonucleotide primers. The nucleic acid amplified thus can be cloned into a suitable vector and characterized by means of DNA sequence analysis.
- Oligonucleotides which correspond to a desaturase nucleotide sequence can be generated by synthetic standard methods, for example using an automatic DNA synthesizer.
- Homologs of the $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase nucleic acid sequences used, with the sequence SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ

ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201, mean for example allelic variants with at least approximately 50 or 60%, preferably at least approximately 60 or 70%, more preferably at least approximately 70 or 80%, 90% or 95% and even more preferably at least approximately 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more identity or homology with one of the nucleotide sequences shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201 or their homologs, derivatives or analogs or parts thereof. Furthermore, isolated nucleic acid molecules of a nucleotide sequence which hybridize, for example under stringent conditions, with one of the nucleotide sequences shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID

NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201 or a part thereof. A part in accordance with the invention is understood as meaning, in this context, that at least 25 base pairs (= bp), 50 bp, 75 bp, 100 bp, 125 bp or 150 bp, preferably at least 175 bp, 200 bp, 225 bp, 250 bp, 275 bp or 300 bp, especially preferably 350 bp, 400 bp, 450 bp, 500 bp or more base pairs are used for the hybridization. Advantageously, the entire sequence may also be used. Allelic variants comprise in particular functional variants which can be obtained by deletion, insertion or substitution of nucleotides from/into the sequence shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201, the intention being, however, that the enzyme activity of the resulting protein synthesized advantageously being retained for the insertion of one or more genes. Proteins which still retain the enzymatic activity of Δ^{12} -desaturase, ω^3 -desaturase, Δ^9 -elongase, Δ^6 -desaturase, Δ^8 -desaturase, Δ^6 -elongase, Δ^5 -desaturase, Δ^5 -elongase or Δ^4 -desaturase, i.e. whose activity is essentially not reduced, mean proteins with at least 10%, preferably 20%, especially preferably 30%, very especially preferably 40% of the original enzyme activity in comparison with the protein encoded by SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID

NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201. The homology was calculated over the entire amino acid or nucleic acid sequence region. A series of programs based on a variety of algorithms is available to the skilled worker for comparing different sequences. In this context, the algorithms of Needleman and Wunsch or Smith and Waterman give particularly reliable results. To carry out the sequence alignments, the program PileUp (J. Mol. Evolution., 25, 351-360, 1987, Higgins et al., CABIOS, 5 (1989: 151-153) or the programs Gap and BestFit [Needleman and Wunsch (J. Mol. Biol. 48; 443-453 (1970) and Smith and Waterman (Adv. Appl. Math. 2; 482-489 (1981)), which are part of the GCG software packet [Genetics Computer Group, 575 Science Drive, Madison Wisconsin, USA 53711 (1991)], were used. The sequence homology values detailed above in percent were determined using the program GAP over the entire sequence region with the following settings: Gap Weight: 50, Length Weight: 3, Average Match: 10.000 and Average Mismatch: 0.000, which, unless otherwise specified, were always used as standard settings for sequence alignments.

Homologs of the abovementioned nucleic acid sequences also mean for example bacterial, fungal and plant homologs, truncated sequences, single-stranded DNA or RNA of the coding and noncoding DNA sequence or else derivatives such as, for example, promoter variants. The promoters upstream of the nucleotide sequences stated can be modified by one or more nucleotide substitutions, by insertion(s) and/or deletion(s), without, however, the functionality or activity of the promoters being adversely affected. Furthermore, it is possible that the activity of the promoters is increased by modifying their sequence, or that they are replaced completely by more active promoters, including those from heterologous organisms.

The abovementioned nucleic acids and protein molecules with $\Delta 12$ -desaturase, $\omega 3$ -desaturase, $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase and/or $\Delta 4$ -desaturase activity which are involved in the metabolism of lipids and fatty acids, PUFA cofactors and enzymes or in the transport of lipophilic compounds across membranes are used in the process according to the invention for modulating the production of PUFAs in transgenic plants such as maize, wheat, rye, oats, triticale, rice, barley, soybean, peanut, cotton, Linum species such as linseed or flax, Brassica species such as oilseed rape, canola, Indian mustard and

turnip rape, pepper, sunflower, borage, evening primrose and Tagetes, Solanaceae plants such as potato, tobacco, eggplant or tomato, Vicia species, pea, cassava, alfalfa, bushy plants (coffee, cacao, tea), Salix species, trees (oil palm, coconut) and perennial grasses and fodder crops either directly (for example when the
5 overexpression or optimization of a fatty acid biosynthetic protein has a direct effect on the yield, production and/or production efficiency of the fatty acid from modified organisms) and/or can have an indirect effect which nevertheless entails an increase in the yield, production and/or production efficiency of the PUFAs or a decrease of undesired compounds (for example when the modulation of the metabolism of lipids
10 and fatty acids, cofactors and enzymes results in changes in the yield, production and/or production efficiency or the composition of the desired compounds within the cells which, in turn, can have an effect on the production of one or more fatty acids).

Brassicaceae, Boraginaceae, Primulaceae or Linaceae are especially suitable for the
15 production of PUFAs, preferably of arachidonic acid, eicosapentaenoic acid or docosahexaenoic acid. Especially suitable for the production of PUFAs with the nucleic acid sequences according to the invention, advantageously, as described, in combination with further desaturases and elongases are Indian mustard (*Brassica juncea*), oilseed rape and *Camelina sativa*.

20 The combination of a variety of precursor molecules and biosynthetic enzymes leads to the production of different fatty acid molecules, which has a major effect on the composition of the lipids since polyunsaturated fatty acids (= PUFAs) are incorporated not only into triacylglycerol but also into membrane lipids.

25 Brassicaceae, Boraginaceae, Primulaceae or Linaceae are especially suitable for the production of PUFAs, for example stearidonic acid, eicosapentaenoic acid or docosahexaenoic acid. Linseed (*Linum usitatissimum*) and *Brassica juncea* and *Camelina sativa* are especially advantageously suitable for the production of PUFAs
30 with the nucleic acid sequences according to the invention, advantageously, as described, in combination with further desaturates and elongases.

Lipid synthesis can be divided into two sections: the synthesis of fatty acids and their
35 binding to sn-glycerol-3-phosphate, and the addition or modification of a polar head group. Usual lipids which are used in membranes comprise phospholipids, glycolipids, sphingolipids and phosphoglycerides. Fatty acid synthesis starts with the conversion of acetyl-CoA into malonyl-CoA by acetyl-CoA carboxylase or into acetyl-ACP by acetyl transacylase. After condensation reaction, these two product molecules together form

acetoacetyl-ACP, which is converted via a series of condensation, reduction and dehydration reactions so that a saturated fatty acid molecule with the desired chain length is obtained. The production of the unsaturated fatty acids from these molecules is catalyzed by specific desaturases, either aerobically by means of molecular oxygen or anaerobically (regarding the fatty acid synthesis in microorganisms, see F.C. Neidhardt et al. (1996) *E. coli* and *Salmonella*. ASM Press: Washington, D.C., p. 612-636 and references cited therein; Lengeler et al. (Ed.) (1999) *Biology of Prokaryotes*. Thieme: Stuttgart, New York, and the references therein, and Magnuson, K., et al. (1993) *Microbiological Reviews* 57:522-542 and the references therein). To undergo the further elongation steps, the resulting phospholipid-bound fatty acids must be returned from the phospholipids to the fatty acid CoA ester pool. This is made possible by acyl-CoA:lysophospholipid acyltransferases. Moreover, these enzymes are capable of transferring the elongated fatty acids from the CoA esters back to the phospholipids. If appropriate, this reaction sequence can be followed repeatedly.

Examples of precursors for PUFA biosynthesis are oleic acid, linoleic acid and linolenic acid. These C₁₈-carbon fatty acids must be elongated to C₂₀ and C₂₂ to obtain fatty acids of the eicosa and docosa chain type. It is possible, with the aid of the desaturases used in the process, such as the Δ 12-, ω 3-, Δ 4-, Δ 5-, Δ 6- and Δ 8-desaturases and/or the Δ 5-, Δ 6-, Δ 9-elongases to produce arachidonic acid, eicosapentaenoic acid, docosapentaenoic acid or docosahexaenoic acid, advantageously eicosapentaenoic acid and/or docosahexaenoic acid, and subsequently to use them for a variety of purposes in applications in the fields of foodstuffs, feedstuffs, cosmetics or pharmaceuticals. Using the abovementioned enzymes, C₂₀- and/or C₂₂-fatty acids with at least two, advantageously at least three, four, five or six double bonds in the fatty acid molecule, preferably C₂₀- or C₂₂-fatty acids with advantageously four, five or six double bonds in the fatty acid molecule can be produced. The desaturation can take place before or after elongation of the fatty acid in question. This is why the products of the desaturase activities and the further possible desaturation and elongation lead to preferred PUFAs with a higher degree of desaturation, including a further elongation of C₂₀- to C₂₂-fatty acids, to fatty acids such as γ -linolenic acid, dihomo- γ -linolenic acid, arachidonic acid, stearidonic acid, eicosatetraenoic acid or eicosapentaenoic acid. Substrates of the desaturases and elongases used in the process according to the invention are C₁₆-, C₁₈- or C₂₀-fatty acids such as, for example, linoleic acid, γ -linolenic acid, α -linolenic acid, dihomo- γ -linolenic acid, eicosatetraenoic acid or stearidonic acid. Preferred substrates are linoleic acid, γ -linolenic acid and/or α -linolenic acid, dihomo- γ -linolenic acid or arachidonic acid, eicosatetraenoic acid or eicosapentaenoic acid. The synthesized C₂₀-

to C₂₂-fatty acids with at least two, three, four, five or six, advantageously at least four, five or six double bonds in the fatty acid are obtained in the process according to the invention in the form of the free fatty acid or in the form of its esters, for example in the form of its glycerides.

5

The term "glyceride" is understood as meaning glycerol esterified with one, two or three carboxyl radicals (mono-, di- or triglyceride). "Glyceride" is also understood as meaning a mixture of various glycerides. The glyceride or glyceride mixture can comprise further additions, for example free fatty acids, antioxidants, proteins, carbohydrates, vitamins

10

and/or other substances.

A "glyceride" for the purposes of the process according to the invention is furthermore understood as meaning derivatives which are derived from glycerol. In addition to the above-described fatty acid glycerides, these also include glycerophospholipids and glyceroglycolipids. Preferred examples which may be mentioned here are the glycerophospholipids such as lecithin (phosphatidylcholine), cardiolipin, phosphatidylglycerol, phosphatidylserine and alkylacylglycerophospholipids.

15

Furthermore, fatty acids must subsequently be transported to various sites of modification and incorporated into the triacylglycerol storage lipid. A further important step in lipid synthesis is the transfer of fatty acids onto the polar head groups, for example by glycerol-fatty-acid acyltransferase (see Frentzen, 1998, Lipid, 100(4-5):161-166).

20

Publications on plant fatty acid biosynthesis, desaturation, the lipid metabolism and the transmembrane transport of fatty compounds, beta-oxidation, fatty acid modification and cofactors, triacylglycerol storage and assembly, including the references therein, see the following articles: Kinney, 1997, Genetic Engineering, Ed., JK Setlow, 19:149-166; Ohlrogge and Browse, 1995, Plant Cell 7:957-970; Shanklin and Cahoon, 1998, Annu. Rev. Plant Physiol. Plant Mol. Biol. 49:611-641; Voelker, 1996, Genetic Engineering, Ed.: JK Setlow, 18:111-13; Gerhardt, 1992, Prog. Lipid R. 31:397-417; Gühnemann-Schäfer & Kindl, 1995, Biochim. Biophys Acta 1256:181-186; Kunau et al., 1995, Prog. Lipid Res. 34:267-342; Stymme et al., 1993, in: Biochemistry and Molecular Biology of Membrane and Storage Lipids of Plants, Ed.: Murata and Somerville, Rockville, American Society of Plant Physiologists, 150-158, Murphy & Ross 1998, Plant Journal. 13(1):1-16.

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The PUFAs produced in the process comprise a group of molecules which higher

animals are no longer capable of synthesizing and must therefore take up, or which higher animals are no longer capable of synthesizing themselves in sufficient quantity and must therefore take up additionally, although they can be readily synthesized by other organisms such as bacteria; for example, cats are no longer capable of synthesizing arachidonic acid.

Phospholipids are to be understood as meaning, for the purposes of the invention, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylglycerol and/or phosphatidylinositol, advantageously phosphatidylcholine.

10

The terms "production" or "productivity" are known in the art and refer to the concentration of the fermentation product (compounds of the formula I) formed within a certain period of time and a certain fermentation volume (for example kg of product per hour per liter). They also encompass the productivity within a plant cell or a plant, i.e. the content of the desired fatty acids produced in the process based on the content of all fatty acids in this cell or plant. The term production efficiency encompasses the time required for obtaining a certain amount of product (for example the time required by the cell for establishing a certain throughput rate of a fine chemical). The term "yield" or "product/carbon yield" is known in the art and comprises the efficiency of the conversion of the carbon source into the product (i.e. the fine chemical). This is usually expressed for example as kg of product per kg of carbon source. By increasing the yield or production of the compound, the amount of the obtained molecules or of the suitable obtained molecules of this compound in a certain amount of culture is increased over a specified period.

25

The terms "biosynthesis" or "biosynthetic pathway" are known in the art and comprise the synthesis of a compound, preferably of an organic compound, by a cell starting from intermediates, for example in a multistep process which is highly regulated. The terms "catabolism" or "catabolic pathway" are known in the art and comprise the cleavage of a compound, preferably of an organic compound, by a cell to give catabolytes (in more general terms, smaller or less complex molecules), for example in a multistep process which is highly regulated.

30

The term "metabolism" is known in the art and encompasses the totality of the biochemical reactions which take place in an organism. Thus, the metabolism of a certain compound (for example the metabolism of a fatty acid) comprises the totality of the biosynthetic, modification and catabolic pathways of this compound in the cell.

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This invention is illustrated in greater detail by the examples which follow, which are not to be construed as limiting. The content of all of the references, patent applications, patents and published patent applications cited in the present patent application is herewith incorporated by reference.

5

Examples

Example 1: General cloning methods:

- 10 The cloning methods such as, for example, restriction cleavages, agarose gel electrophoresis, purification of DNA fragments, transfer of nucleic acids to nitrocellulose and nylon membranes, linkage of DNA fragments, transformation of *E. coli* cells, bacterial cultures and the sequence analysis of recombinant DNA were carried out as described by Sambrook et al. (1989) (Cold Spring Harbor Laboratory Press: ISBN 0-87969-309-6).
- 15

Example 2: Sequence analysis of recombinant DNA:

- Recombinant DNA molecules were sequenced with an ABI laser fluorescence DNA sequencer by the process of Sanger (Sanger et al. (1977) Proc. Natl. Acad. Sci. USA74, 5463-5467). Fragments resulting from a polymerase chain reaction were sequenced and verified to avoid polymerase errors in constructs to be expressed.
- 20

Example 3: Cloning genes from *Oncorhynchus mykiss*

25

As the result of a search for conserved regions in the protein sequences corresponding to the elongase genes detailed in the application, two sequences with suitable motifs were identified in the Genbank sequence database.

Name of gene	Genbank No.	Amino acids
OmELO2	CA385234, CA364848, CA366480	264
OmELO3	CA360014, CA350786	295

30

Total RNA from *Oncorhynchus mykiss* was isolated with the aid of the RNAeasy Kit from Qiagen (Valencia, CA, US). Poly-A+ RNA (mRNA) was isolated from the total RNA with the aid of oligo-dT cellulose (Sambrook et al., 1989). The RNA was subjected to reverse transcription using the reverse transcription system kit from Promega, and

the cDNA synthesized was cloned into the lambda ZAP vector (lambda ZAP Gold, Stratagene). The cDNA was depackaged in accordance with the manufacturer's instructions to give the plasmid DNA. The cDNA plasmid library was then used for the PCR for cloning expression plasmids.

5

Example 4: Cloning expression plasmids for the heterologous expression in yeasts

To clone the two sequences for heterologous expression in yeasts, the following oligonucleotides were used for the PCR reaction:

Primer	Nucleotide sequence
5' f* OmELO2	5' aagcttacataatggcttcaacatggcaa (SEQ ID NO: 179)
3' r* OmELO2	5' ggatccttatgtcttcttgccttctgtt (SEQ ID NO: 180)
5' f OmELO3	5' aagcttacataatggagactttta (SEQ ID NO: 181)
3' r OmELO3	5' ggatccttcagtcctccctcactttcc (SEQ ID NO: 182)

10 *f: forward, r: reverse

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

15 5.00 µl of 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

0.50 µl of Advantage polymerase (Clontech)

PCR reaction conditions:

20 Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

25 The PCR product was first incubated for 2 hours at 37°C with the restriction enzymes HindIII and BamHI. The yeast expression vector pYES3 (Invitrogen) was incubated in the same manner. Thereafter, the 812 bp PCR product and the 905 bp PCR product and the vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel

30 Purification Kit following the manufacturer's instructions. Thereafter, the vector and the elongase cDNA were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmids pYES3-OmELO2 and pYES3-OmELO3 were verified by sequencing and transformed into the *Saccharomyces* strain INVSc1 (Invitrogen) by

means of electroporation (1500 V). As a control, pYES3 was transformed in parallel. Thereafter, the yeasts were plated onto complete tryptophan dropout minimal medium supplement with 2% glucose. Cells which are capable of growing on without tryptophan in the medium thus comprise the corresponding plasmids pYES3, pYES3-OmELO2 (SEQ ID NO: 51) and pYES3-OmELO3 (SEQ ID NO: 53). After the selection, in each case two transformants were selected for the further functional expression.

Example 5: Cloning expression plasmids for the seed-specific expression in plants

To transform plants, a further transformation vector based on pSUN-USP was generated. To this end, NotI cleavage sites were introduced at the 5' and 3' termini of the coding sequence using the following primer pair:

PSUN-OmELO2

Forward: 5'-GCGGCCGCATAATGGCTTCAACATGGCAA (SEQ ID NO: 175)

Reverse: 3'-GCGGCCGCTTATGTCTTCTTGCTCTTCCTGTT (SEQ ID NO: 176)

PSUN-OmELO3

Forward: 5'-GCGGCCGCataatggagactttta (SEQ ID NO: 177)

Reverse: 3'-GCGGCCGctcagtcctccctcacttcc (SEQ ID NO: 178)

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

5.00 µl of 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

0.50 µl of Advantage polymerase (Clontech)

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

The PCR products were incubated with the restriction enzyme NotI for 16 hours at 37°C. The plant expression vector pSUN300-USP was incubated in the same manner.

Thereafter, the PCR products and the 7624 bp vector were separated by agarose gel electrophoresis, and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR products were ligated. The Rapid Ligation Kit

from Roche was used for this purpose. The resulting plasmids pSUN-OmELO2 and pSUN-OmELO3 were verified by sequencing.

pSUN300 is a derivative of the plasmid pPZP (Hajdukiewicz P., Svab, Z, Maliga P.,
5 (1994) The small versatile pPZP family of *Agrobacterium* binary vectors for plant transformation. *Plant Mol. Biol.* 25:989-994). pSUN-USP originated from pSUN300 by inserting a USP promoter as *EcoRI* fragment into pSUN 300. The polyadenylation signal is that of the octopine synthase gene from the *A. tumefaciens* Ti plasmid (ocs terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P., Seurinck, J.,
10 Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine synthase gene *J. Mol. Appl. Genet.* 1 (6), 499-511 (1982). The USP promoter corresponds to the nucleotides 1-684 (Genbank Accession X56240), part of the noncoding region of the USP gene being present in the promoter. The promoter fragment, which is 684 base pairs in size,
15 was amplified via a PCR reaction by standard methods, by means of commercially available T7 standard primer (Stratagene) and with the aid of a synthesized primer (primer sequence:
5'-GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGGGGGATCC
GGATCTGCTTGGCTATGAA-3', SEQ ID NO: 174). The PCR fragment was recut with
20 *EcoRI*/*Sall* and inserted into the vector pSUN300 with OCS terminator. This gave rise to the plasmid named pSUN-USP. The construct was used for transforming *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

Example 6: Lipid extraction from yeasts and seeds:

25 The effect of the genetic modification in plants, fungi, algae, ciliates or on the production of a desired compound (such as a fatty acid) can be determined by growing the modified microorganisms or the modified plant under suitable conditions (such as those described above) and analyzing the medium and/or the cellular components for
30 the elevated production of the desired product (i.e. of the lipids or a fatty acid). These analytical techniques are known to the skilled worker and comprise spectroscopy, thin-layer chromatography, various types of staining methods, enzymatic and microbiological methods and analytical chromatography such as high-performance liquid chromatography (see, for example, Ullman, Encyclopedia of Industrial Chemistry,
35 Vol. A2, p. 89-90 and p. 443-613, VCH: Weinheim (1985); Fallon, A., et al., (1987) "Applications of HPLC in Biochemistry" in: Laboratory Techniques in Biochemistry and Molecular Biology, Vol. 17; Rehm et al. (1993) Biotechnology, Vol. 3, Chapter III: "Product recovery and purification", p. 469-714, VCH: Weinheim; Belter, P.A., et al.

- (1988) Bioseparations: downstream processing for Biotechnology, John Wiley and Sons; Kennedy, J.F., and Cabral, J.M.S. (1992) Recovery processes for biological Materials, John Wiley and Sons; Shaeiwitz, J.A., and Henry, J.D. (1988) Biochemical Separations, in: Ullmann's Encyclopedia of Industrial Chemistry, Vol. B3; Chapter 11, p. 1-27, VCH: Weinheim; and Dechow, F.J. (1989) Separation and purification techniques in biotechnology, Noyes Publications).

- In addition to the abovementioned methods, plant lipids are extracted from plant material as described by Cahoon et al. (1999) Proc. Natl. Acad. Sci. USA 96 (22):12935-12940 and Browse et al. (1986) Analytic Biochemistry 152:141-145. The qualitative and quantitative analysis of lipids or fatty acids is described by Christie, William W., Advances in Lipid Methodology, Ayr/Scotland: Oily Press (Oily Press Lipid Library; 2); Christie, William W., Gas Chromatography and Lipids. A Practical Guide - Ayr, Scotland: Oily Press, 1989, Repr. 1992, IX, 307 pp. (Oily Press Lipid Library; 1); "Progress in Lipid Research, Oxford: Pergamon Press, 1 (1952) - 16 (1977) under the title: Progress in the Chemistry of Fats and Other Lipids CODEN.

- In addition to measuring the end product of the fermentation, it is also possible to analyze other components of the metabolic pathways which are used for the production of the desired compound, such as intermediates and by-products, in order to determine the overall production efficiency of the compound. The analytical methods comprise measuring the amount of nutrients in the medium (for example sugars, hydrocarbons, nitrogen sources, phosphate and other ions), measuring the biomass composition and the growth, analyzing the production of conventional metabolites of biosynthetic pathways and measuring gases which are generated during the fermentation. Standard methods for these measurements are described in Applied Microbial Physiology; A Practical Approach, P.M. Rhodes and P.F. Stanbury, Ed., IRL Press, p. 103-129; 131-163 and 165-192 (ISBN: 0199635773) and references cited therein.

- One example is the analysis of fatty acids (abbreviations: FAME, fatty acid methyl ester; GC-MS, gas liquid chromatography/mass spectrometry; TAG, triacylglycerol; TLC, thin-layer chromatography).

- The unambiguous detection for the presence of fatty acid products can be obtained by analyzing recombinant organisms using analytical standard methods: GC, GC-MS or TLC, as described on several occasions by Christie and the references therein (1997, in: Advances on Lipid Methodology, Fourth Edition: Christie, Oily Press, Dundee, 119-

169; 1998, Gaschromatographie-Massenspektrometrie-Verfahren [Gas chromatography/mass spectrometric methods], Lipide 33:343-353).

5 The material to be analyzed can be disrupted by sonication, grinding in a glass mill, liquid nitrogen and grinding or via other applicable methods. After disruption, the material must be centrifuged. The sediment is resuspended in distilled water, heated for 10 minutes at 100°C, cooled on ice and recentrifuged, followed by extraction for one hour at 90°C in 0.5 M sulfuric acid in methanol with 2% dimethoxypropane, which leads to hydrolyzed oil and lipid compounds, which give transmethyated lipids. These fatty
10 acid methyl esters are extracted in petroleum ether and finally subjected to a GC analysis using a capillary column (Chrompack, WCOT Fused Silica, CP-Wax-52 CB, 25 m, 0.32 mm) at a temperature gradient of between 170°C and 240°C for 20 minutes and 5 minutes at 240°C. The identity of the resulting fatty acid methyl esters must be defined using standards which are available from commercial sources (i.e. Sigma).

15 Plant material is initially homogenized mechanically by comminuting in a pestle and mortar to make it more amenable to extraction.

This is followed by heating at 100°C for 10 minutes and, after cooling on ice, by
20 resedimentation. The cell sediment is hydrolyzed for one hour at 90°C with 1 M methanolic sulfuric acid and 2% dimethoxypropane, and the lipids are transmethyated. The resulting fatty acid methyl esters (FAMES) are extracted in petroleum ether. The extracted FAMES are analyzed by gas liquid chromatography using a capillary column (Chrompack, WCOT Fused Silica, CP-Wax-52 CB, 25 m, 0.32 mm) and a temperature
25 gradient of from 170°C to 240°C in 20 minutes and 5 minutes at 240°C. The identity of the fatty acid methyl esters is confirmed by comparison with corresponding FAME standards (Sigma). The identity and position of the double bond can be analyzed further by suitable chemical derivatization of the FAME mixtures, for example to give 4,4-dimethoxyoxazolin derivatives (Christie, 1998) by means of GC-MS.

30 Yeasts which had been transformed with the plasmids pYES3, pYES3-OmELO2 and pYES3-OmELO3 as described in Example 4 were analyzed as follows:

The yeast cells from the main cultures were harvested by centrifugation (100 × g, 10
35 min, 20°C) and washed with 100 mM NaHCO₃, pH 8.0 in order to remove residual medium and fatty acids. Fatty acid methyl esters (FAMES) were prepared with the yeast cell sediments by acid methanolysis. To this end, the cell sediments were incubated for 1 hour at 80°C with 2 ml of 1N methanolic sulfuric acid and 2% (v/v)

dimethoxypropane. The FAMEs were extracted by twice extracting with petroleum ether (PE). To remove non-derivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO₃, pH 8.0, and 2 ml of distilled water. Thereafter, the PE phases were dried with Na₂SO₄, evaporated under argon and taken up in 100 µl of PE. The samples were separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 µm, Agilent) in a Hewlett-Packard 6850 gas chromatograph with flame ionization detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of 5°C/min and finally 10 minutes at 250°C (holding).

The signals were identified by comparing the retention times with corresponding fatty acid standards (Sigma).

The methodology is described for example in Napier and Michaelson, 2001, *Lipids* 36(8):761-766; Sayanova et al., 2001, *Journal of Experimental Botany*, 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

Example 7: Functional characterization of OmELO2 and OmELO3:

OmELO2 shows no elongase activity, while a pronounced activity was detected for OmELO3, using different substrates. The substrate specificity of OmELO3 was determined after expression and feeding with various fatty acids (Figure 2). The fed substrates can be detected in large amounts in all transgenic yeasts. All transgenic yeasts show that new fatty acids have been synthesized, to the products of the OmELO3 reaction. This means that the gene OmELO3 was expressed functionally.

Figure 2 demonstrates that OmELO3 has a substrate specificity which leads to the elongation of Δ5- and Δ6-fatty acids with one ω-double bond with high specificity.

Moreover, ω6-fatty acids (C18 and C20) were also elongated, with less specificity. The best substrates for OmELO3 were stearidonic acid (C18:4 ω3) and eicosapentaenoic acid (C20:5 ω3) (up to 66% elongation).

Example 8: Reconstitution of the synthesis of DHA in yeast

The reconstitution of the biosynthesis of DHA (22:6 ω3) was carried out starting from EPA (20:5 ω3) or stearidonic acid (18:4 ω3) by coexpressing OmELO3 together with the *Euglena gracilis* Δ4-desaturase or the *Phaeodactylum tricornutum* Δ5-desaturase and

the *Euglena gracilis* $\Delta 4$ -desaturase. To this end, the expression vectors pYes2-EgD4 and pESCLEu-PtD5 were additionally constructed. The abovementioned yeast strain which is already transformed with pYes3-OmElo3 (SEQ ID NO: 55), was then transformed further with pYes2-EgD4, or simultaneously with pYes2-EgD4 and pESCLEu-PtD5. The transformed yeasts were selected on complete minimal dropout tryptophan and uracil medium agar plates supplemented with 2% glucose in the case of the pYes3-pYes3-OmEIO/pYes2-EgD4 strain and complete minimal dropout tryptophan, uracil and leucine medium in the case of the pYes3-OmEIO/pYes2-EgD4+pESCLEu-PtD5 strain. Expression was then induced by addition of 2% (w/v) galactose. The cultures were subsequently incubated for a further 120 hours at 15°C.

Figure 3 shows the fatty acid profiles of transgenic yeasts which have been fed 20:5 $\omega 3$. In the control yeast (A), which had been transformed with the vector pYes3-OmElo3 and the blank vector pYes2, 20:5 $\omega 3$ was elongated highly efficiently to give 22:5 $\omega 3$ (65% elongation). The additional introduction of the EEg $\Delta 4$ -desaturase led to the conversion of 22:5 $\omega 3$ into 22:6 $\omega 3$ DHA. The fatty acid composition of the transgenic yeasts is shown in Figure 5. After coexpression of OmElo3 and EgD4, up to 3% DHA was detected in yeasts.

In a further coexpression experiment, OmElo3, EgD4 and a $\Delta 5$ -desaturase from *P. tricomutum* (PtD5) were expressed together. The transgenic yeasts were fed stearidonic acid (18:4 $\omega 3$) and analyzed (Figure 4). The fatty acid composition of these yeasts is shown in Figure 5. OmElo3 elongated the fed fatty acid 18:4 $\omega 3$ to give 20:4 $\omega 3$ (60% elongation). The latter was desaturated by PtD5 to give 20:5 $\omega 3$. The PtD5 activity amounted to 15%. Furthermore, 20:5 $\omega 3$ was elongated by EmElo3 to give 22:5 $\omega 3$. Thereafter, the newly synthesized 22:5 $\omega 3$ was desaturated to give 22:6 $\omega 3$ (DHA). Up to 0.7% of DHA was obtained in these experiments.

These experiments demonstrate that the sequences OmElo3, EgD4 and PtD5 which are used in the present invention are suitable for the production of DHA in eukaryotic cells.

Example 9: Generation of transgenic plants

- a) Generation of transgenic oilseed rape plants (modified process of Moloney et al., 1992, Plant Cell Reports, 8:238-242)

The binary vectors in *Agrobacterium tumefaciens* C58C1:pGV2260 or *Escherichia coli* (Deblaere et al, 1984, Nucl. Acids. Res. 13, 4777-4788) can be used for generating transgenic oilseed rape plants. To transform oilseed rape plants (Var. Drakkar, NPZ Nordeutsche Pflanzenzucht, Hohenlieth, Germany), a 1:50 dilution of an overnight
5 culture of a positively transformed agrobacterial colony in Murashige-Skoog medium (Murashige and Skoog 1962 Physiol. Plant. 15, 473) supplemented with 3% sucrose (3MS medium) is used. Petioles or hypocotyls of freshly germinated sterile oilseed rape plants (in each case approx. 1 cm²) are incubated with a 1:50 agrobacterial dilution for 5-10 minutes in a petri dish. This is followed by 3 days of coincubation in the dark at
10 25°C on 3MS medium supplemented with 0.8% Bacto agar. The cultures are then grown for 3 days at 16 hours light/8 hours dark. The cultivation is then continued in a weekly rhythm on MS medium supplemented with 500 mg/l Claforan (cefotaxim sodium), 50 mg/l kanamycin, 20 µM benzylaminopurine (BAP), now supplemented with 1.6 g/l of glucose. Growing shoots are transferred to MS medium supplemented with
15 2% sucrose, 250 mg/l Claforan and 0.8% Bacto agar. If no roots have developed after three weeks, 2-indolebutyric acid is added to the medium as growth hormone for rooting.

Regenerated shoots were obtained on 2MS medium supplemented with kanamycin
20 and Claforan; after rooting, they were transferred to compost and, after growing on for two weeks in a controlled-environment cabinet or in the greenhouse, allowed to flower, and mature seeds were harvested and analyzed by lipid analysis for elongase expression, such as $\Delta 5$ -elongase or $\Delta 6$ -elongase activity. In this manner, lines with elevated contents of polyunsaturated C₂₀- and C₂₂-fatty acids can be identified.

25

b) Generation of transgenic linseed plants

Transgenic linseed plants can be generated for example by the process of Bell et al., 1999, In Vitro Cell. Dev. Biol.-Plant. 35(6):456-465 by means of particle bombardment.
30 Usually, an agrobacteria-mediated transformations was used for the transformation of linseed, for example by the process of Mlynarova et al. (1994), Plant Cell Report 13: 282-285.

Example 10: Cloning $\Delta 5$ -elongase genes from *Thraustochytrium aureum* ATCC34304
35 and *Thraustochytrium* ssp.

Comparisons of the various elongase protein sequences found in the present application enabled the definition of conserved nucleic acid regions (histidin box: His-

Val-X-His-His, tyrosin box: Met-Tyr-X-Tyr-Tyr). An EST database of *T. aureum* ATCC34304 and *Thraustochytrium* ssp. was screened for further $\Delta 5$ -elongases with the aid of these sequences. The following new sequences were found:

Name of gene	Nucleotides	Amino acids
BioTaurELO1	828 bp	275
TL16y2	831	276

5

Total RNA from *T. aureum* ATCC34304 and *Thraustochytrium* ssp. was isolated with the aid of the RNeasy Kits from Qiagen (Valencia, CA, US). mRNA was isolated from the total RNA with the aid of the polyAtract isolation system (Promega). The mRNA was subjected to reverse transcription using the Marathon cDNA Amplification Kit (BD Biosciences) and adaptors were ligated in accordance with the manufacturer's instructions. The cDNA library was then employed for the PCR for cloning expression plasmids by means of 5'- and 3'-RACE (rapid amplification of cDNA ends).

10

Example 11: Cloning expression plasmids for the heterologous expression in yeasts

15

To clone the sequence for heterologous expression in yeasts, the following oligonucleotides were used for the PCR reaction:

Primer	Nucleotide sequence
5' f* BioTaurELO1	5' gacataatgacgagcaacatgag (SEQ ID NO: 170)
3' r* BioTaurELO1	5' cggcttaggccgacttggccttggg (SEQ ID NO: 171)
5' f*TL16y2	5' agacataatggacgtcgtcgagcagcaatg (SEQ ID NO: 172)
3' r*TL16y2	5' ttataggttcttctgcttcttggcgcc (SEQ ID NO: 173)

*f: forward, r: reverse

20

Composition of the PCR mix (50 μ l):

5.00 μ l template cDNA

5.00 μ l 10x buffer (Advantage polymerase) + 25mM MgCl₂

5.00 μ l of 2mM dNTP

1.25 μ l of each primer (10 pmol/ μ l)

25

0.50 μ l of Advantage polymerase (Clontech)

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

The PCR products BioTaurELO1 (see (SEQ ID NO: 65) and TL16y2 (see SEQ ID NO: 83) were incubated for 30 minutes at 21°C with the yeast expression vector pYES2.1-TOPO (Invitrogen) following the manufacturer's instructions. The PCR product is ligated into the vector by means of a T overhang and activity of a topoisomerase (Invitrogen). After incubation, *E. coli* DH5α cells were transformed. Suitable clones were identified by PCR, the plasmid DNA was isolated by means of Qiagen DNAeasy Kit and verified by sequencing. The correct sequence was then transformed into the *Saccharomyces* strain INVSc1 (Invitrogen) by electroporation (1500 V). As a control, the blank vector pYES2.1 was transformed in parallel. The yeasts were subsequently plated onto complete uracil dropout minimal medium supplemented with 2% glucose. Cells which were capable of growing in the medium without uracil thus comprise the corresponding plasmids pYES2.1, pYES2.1-BioTaurELO1 and pYES2.1-TL16y2. After the selection, in each case two transformants were selected for further functional expression.

Example 12: Cloning expression plasmids for the seed specific expression in plants

A further transformation vector based on pSUN-USP was generated for the transformation of plants. To this end, NotI cleavage sites were introduced at the 5' and 3' termini of the coding sequence, using the following primer pair:

PSUN-BioTaurELO1

Forward: 5'-GCGGCCGCATAATGACGAGCAACATGAGC (SEQ ID NO: 166)

Reverse: 3'-GCGGCCGCTTAGGCCGACTTGGCCTTGGG (SEQ ID NO: 167)

PSUN-TL16y2

Forward: 5'-GCGGCCGCACCATGGACGTCGTCGAGCAGCAATG (SEQ ID NO: 168)

Reverse: 5'-GCGGCCGCTTAGATGGTCTTCTGCTTCTTGGGCGCC (SEQ ID NO: 169)

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase)+ 25mM MgCl₂

5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

PCR reaction conditions:

5

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

10

The PCR products were incubated with the restriction enzyme NotI for 16 hours at 37°C. The plant expression vector pSUN300-USP was incubated in the same manner. Thereafter, the PCR products and the 7624 bp vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR products were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmids pSUN-BioTaurELO1 and pSUN-TL16y2 were verified by sequencing.

20

pSUN300 is a derivative of plasmid pPZP (Hajdukiewicz, P., Svab, Z., Maliga, P., (1994)

The small versatile pPZP family of *Agrobacterium* binary vectors for plant transformation. *Plant Mol Biol* 25:989-994). pSUN-USP originated from pSUN300, by inserting a USP promoter into pSUN300 in the form of an EcoRI fragment. The polyadenylation signal is that of the octopine synthase gene from the *A. tumefaciens* Ti plasmid (ocs-Terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P., Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine synthase gene *J. Mol. Appl. Genet.* 1 (6), 499-511 (1982)). The USP promoter corresponds to nucleotides 1 to 684 (Genbank Accession X56240), where part of the noncoding region of the USP gene is present in the promoter. The promoter fragment which is 684 base pairs in size was amplified by a PCR reaction and standard methods with the aid of a synthesized primer and by means of a commercially available T7 standard primer (Stratagene). (Primer sequence:

30

5'-GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGGGGGATCC

35

GGATCTGCTGGCTATGAA-3', SEQ ID NO: 165). The PCR fragment was recut with EcoRI/Sall and inserted into the vector pSUN300 with OCS terminator. This gave rise

to the plasmid with the name pSUN-USP. The construct was used for the transformation of *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

Lipids were extracted from yeasts and seeds as described for Example 6.

5

Example 13: Functional characterization of BioTaurELO1 and TL16y2:

The substrate specificity of BioTaurELO1 was determined after expression and feeding of various fatty acids (Figure 6). Figure 6 shows the feeding experiments for
10 determining the functionality and substrate specificity with yeast strains comprising either the vector pYes2.1 (control) or the vector pYes2.1-BioTaurELO1 (= BioTaur) with the $\Delta 5$ -elongase. In both approaches, 200 μ m of γ -linolenic acid and eicosapentaenoic acid were added to the yeast incubation medium and incubated for 24 hours. After the fatty acids had been extracted from the yeasts, they were transmethyated and
15 separated by gas chromatography. The elongation products originating from the two fatty acids which had been fed are identified by arrows.

The substrates which had been fed can be detected in large amounts in all transgenic yeasts. All transgenic yeasts show that new fatty acids have been synthesized, the
20 products of the BioTaurELO1 reaction. This means that the gene BioTaurELO1 has been expressed functionally.

Figure 6 shows that BioTaurELO1 has a substrate specificity which leads with high specificity to the elongation of $\Delta 5$ - and $\Delta 6$ -fatty acids with one $\omega 3$ -double bond.
25 Moreover, $\omega 6$ -fatty acids (C18 and C20) were also elongated. γ -Linolenic acid (C18:3 $\omega 6$) is converted with a conversion rate of 65.28%, stearidonic acid (C18:4 $\omega 3$) with a conversion rate of 65.66% and eicosapentaenoic acid (C20:5 $\omega 3$) with a conversion rate of 22.01%. The substrate specificities of the various feeding experiments are shown in Table 6 (see end of the description).

30

The conversion rate of GLA when feeding GLA and EPA was 65.28%. The conversion rate of EPA, again when feeding GLA and EPA, was 9.99%. When only EPA was fed, the EPA conversion rate was 22.01%. Arachidonic acid (= ARA) was also converted when fed. The conversion rate was 14.47%. Stearidonic acid (= SDA) was also
35 converted. In this case, the conversion rate was 65.66%.

The functionality and substrate specificity of TL16y2 were determined after expression and feeding of various fatty acids. Table 7 shows the feeding experiments. The feeding

experiments were carried out in the same manner as described for BioTaurELO1. The substrates which have been fed can be detected in large amounts in all transgenic yeasts. The transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the TL16y2 reaction (Fig. 11). This means that the gene TL16y2 has been expressed functionally.

Table 7: Expression of TL16y2 in yeast.

% areas in the gas-chromatographic analysis									
Plasmid	Fatty acid	C18:3 (n-6)	C18:4 (n-3)	C20:3 (n-6)	C20:4 (n-6)	C20:4 (n-3)	C20:5 (n-3)	C22:4 (n-6)	C22:5 (n-3)
pYES	250 μ m EPA						13.79		
TL16y2	250 μ m EPA						25.81		2.25
pYES	50 μ m EPA						5.07		
TL16y2	50 μ m EPA						2.48		1.73
pYES	250 μ m GLA	8.31							
TL16y2	250 μ m GLA	3.59		10.71					
pYES	250 μ m ARA				16.03				
TL16y2	250 μ m ARA				15.2		3.87		
pYES	250 μ m SDA		26.79			0.35			
TL16y2	250 μ m SDA		7.74			29.17			

10 The results with TL16y2, which are shown in Table 7, show the following conversion rates in % of the control: a) conversion rate of EPA in % (250 μ m): 8%, b) conversion rate of EPA in % (50 μ m): 41%; c) conversion rate of ARA in %: 20.3%, d) conversion rate of SDA in %: 79.4%, and e) conversion rate of GLA in %: 74.9%.

Thus, TL16y2 shows $\Delta 5$ -, $\Delta 6$ - and $\Delta 8$ -elongase activity. The activity is highest for C18-fatty acids with $\Delta 6$ -double bond. Then, C20-fatty acids with a $\Delta 5$ - or $\Delta 8$ -double bond are elongated, depending on the concentration of fatty acids which are fed.

5 Example 14: Cloning genes from *Ostreococcus tauri*

The search for conserved regions in the protein sequences with the aid of the elongase genes with $\Delta 5$ -elongase activity or $\Delta 6$ -elongase activity which are shown in the application allowed the identification of sequences with suitable motifs in an

10 *Ostreococcus tauri* sequence database (genomic sequences).

The sequences were the following:

Name of gene	SEQ ID	Amino acids
OtELO1, ($\Delta 5$ -elongase)	SEQ ID NO: 67	300
OtELO2, ($\Delta 6$ -elongase)	SEQ ID NO: 69	292

15 OtElo1 shows the highest similarity with an elongase from *Danio rerio* (GenBank AAN77156; identity approx. 26%), while OtElo2 shows the highest similarity with the *Physcomitrella Elo* (PSE) [approx. 36% identity] (alignments were carried out using the tBLASTn algorithm (Altschul et al., J. Mol. Biol. 1990, 215: 403-410).

20 The cloning procedure was as follows:

40 ml of an *Ostreococcus tauri* culture in the stationary phase were spun down, resuspended in 100 μ l of double-distilled water and stored at -20°C. The respective genomic DNAs were amplified on the basis of the PCR process. The relevant primer

25 pairs were selected in such a way that they bore the yeast consensus sequence for highly efficient translation (Kozak, Cell 1986, 44:283-292) next to the start codon. The amplification of the OtElo DNAs was carried out in each case using 1 μ l of defrosted cells, 200 μ m of dNTPs, 2.5 U *Taq* polymerase and 100 pmol of each primer in a total volume of 50 μ l. The PCR conditions were as follows: first denaturation for 5 minutes at

30 95°C, followed by 30 cycles of 30 seconds at 94°C, 1 minute at 55°C and 2 minutes at 72°C, and a last elongation step of 10 minutes at 72°C.

Example 15: Cloning expression plasmids for the heterologous expression in yeasts:

35 To characterize the function of the *Ostreococcus tauri* elongases, the open reading

frames of the DNAs in question were cloned downstream of the galactose-inducible GAL1 promoter of pYES2.1/V5-His-TOPO (Invitrogen), giving rise to pOTE1 and pOTE2.

- 5 The *Saccharomyces cerevisiae* strain 334 was transformed by electroporation (1500 v) with the vector pOTE1 or pOTE2. A yeast which was transformed with the blank vector pYES2 was used as the control. The transformed yeasts were selected on complete minimal dropout uracil medium (CMdum) agar plates supplemented with 2% glucose. After the selection, in each case three transformants were selected for the further
10 functional expression.

To express the Ot elongases, precultures of in each case 5 ml of dropout uracil CMdum liquid medium supplemented with 2% (w/v) raffinose were inoculated with the selected transformants and incubated for 2 days at 30°C, 200 rpm.

- 15 5 ml of CMdum liquid medium (without uracil) supplemented with 2% raffinose and 300 µm of various fatty acids were then inoculated with the precultures to an OD₆₀₀ of 0.05. The expression was induced by addition of 2% (w/v) galactose. The cultures were incubated for a further 96 hours at 20°C.

20

Example 16: Cloning of expression plasmids for the seed-specific expression in plants

- A further transformation vector based on pSUN-USP was generated for the transformation of plants. To this end, NotI cleavage sites were introduced at the 5' and
25 3' termini of the coding sequences, using PCR. The corresponding primer sequences are derived from the 5' and 3' regions of OtElo1 and OtElo2.

Composition of the PCR mix (50 µl):

- 30 5.00 µl template cDNA
5.00 µl 10x buffer (Advantage polymerase)+ 25mM MgCl₂
5.00 µl 2mM dNTP
1.25 µl of each primer (10 pmol/µl)
0.50 µl Advantage polymerase

35

The Advantage polymerase from Clontech was employed.

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

5

The PCR products were incubated with the restriction enzyme NotI for 16 hours at 37°C. The plant expression vector pSUN300-USP was incubated in the same manner. Thereafter, the PCR products and the vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was
10 purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR products were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmids pSUN-OtELO1 and pSUN-OtELO2 were verified by sequencing.

- 15 pSUN300 is a derivative of plasmid pPZP (Hajdukiewicz, P., Svab, Z., Maliga, P., (1994) The small versatile pPZP family of *Agrobacterium* binary vectors for plant transformation. *Plant Mol Biol* 25:989-994). pSUN-USP originated from pSUN300, by inserting a USP promoter into pSUN300 in the form of an EcoRI fragment. The polyadenylation signal is that of the *Ostreococcus* gene from the *A. tumefaciens* Ti
20 plasmid (ocs-Terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P., Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine synthase gene *J. Mol. Appl. Genet.* 1 (6), 499-511 (1982)). The USP promoter corresponds to nucleotides 1 to 684 (Genbank Accession X56240), where part of the
25 noncoding region of the USP gene is present in the promoter. The promoter fragment which is 684 base pairs in size was amplified by a PCR reaction and standard methods with the aid of a synthesized primer and by means of a commercially available T7 standard primer (Stratagene). (Primer sequence:
5'-GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGGGGGATCC
30 GGATCTGCTGGCTATGAA-3', SEQ ID NO: 164).
The PCR fragment was recut with EcoRI/Sall and inserted into the vector pSUN300 with OCS terminator. This gave rise to the plasmid with the name pSUN-USP. The construct was used for the transformation of *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

35

Example 17: Expression of OtELO1 and OtELO2 in yeasts

Yeasts which had been transformed with the plasmids pYES3, pYES3-OtELO1 and pYES3-OtELO2 as described in Example 15 were analyzed as follows:

The yeast cells from the main cultures were harvested by centrifugation (100 x g, 5 min, 20°C) and washed with 100 mM NaHCO₃, pH 8.0 to remove residual medium and fatty acids. Starting with the yeast cell sediments, fatty acid methyl esters (FAMES) were prepared by acid methanolysis. To this end, the cell sediments were incubated for one hour at 80°C together with 2 ml of 1 N methanolic sulfuric acid and 2% (v/v) of dimethoxypropane. The FAMES were extracted twice with petroleum ether (PE). To remove nonderivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO₃, pH 8.0 and 2 ml of distilled water. Thereafter, the PE phases were dried with Na₂SO₄, evaporated under argon and taken up in 100 µl of PE. The samples were separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 µm, Agilent) in a Hewlett-Packard 6850 gas chromatograph equipped with flame ionization detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of 5°C/min and finally 10 min at 250°C (holding).

The signals were identified by comparing the retention times with corresponding fatty acid standards (Sigma). The methodology is described for example in Napier and Michaelson, 2001, *Lipids*. 36(8):761-766; Sayanova et al., 2001, *Journal of Experimental Botany*. 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

Example 18: Functional characterization of OtELO1 and OtELO2:

The substrate specificity of OtELO1 could be determined after expression and the feeding of different fatty acids (Tab. 8). The substrates fed can be detected in large amounts in all of the transgenic yeasts. The transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the OtELO1 reaction. This means that the gene OtELO1 has been expressed functionally.

It can be seen from Table 7 that OtELO1 has a narrow substrate specificity. OtELO1 was only capable of elongating the C20-fatty acids eicosapentaenoic acid (Figure 7) and arachidonic acid (Figure 8), but preferred the ω3-desaturated eicosapentaenoic acid.

Table 8:

Fatty acid substrate	Conversion rate (in %)
16:0	-
16:1 ^{Δ9}	-
18:0	-
18:1 ^{Δ9}	-
18:1 ^{Δ11}	-
18:2 ^{Δ9,12}	-
18:3 ^{Δ6,9,12}	-
18:3 ^{Δ5,9,12}	-
20:3 ^{Δ8,11,14}	-
20:4 ^{Δ5,8,11,14}	10.8 ± 0.6
20:5 ^{Δ5,8,11,14,17}	46.8 ± 3,6
22:4 ^{Δ7,10,13,16}	-
22:6 ^{Δ4,7,10,13,16,19}	-

Table 8 shows the substrate specificity of the elongase OtElo1 for C20-polyunsaturated fatty acids with one double bond in Δ5-position in comparison with various fatty acids.

The yeasts which had been transformed with the vector pOTE1 were cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC. Each value represents the mean (n=3) ± standard deviation.

The substrate specificity of OtElo2 (SEQ ID NO: 81) could be determined after expression and the feeding of different fatty acids (Tab. 9). The substrates fed could be detected in large amounts in all of the transgenic yeasts. The transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the OtElo2 reaction. This means that the gene OtElo2 has been expressed functionally.

Table 9:

Fatty acid substrate	Conversion rate (in %)
16:0	-
16:1 ^{Δ9}	-

16:3 $\Delta^{7,10,13}$	
18:0	-
18:1 Δ^0	-
18:1 Δ^9	-
18:1 Δ^{11}	-
18:2 $\Delta^{9,12}$	-
18:3 $\Delta^{6,9,12}$	15.3$\pm$
18:3 $\Delta^{5,9,12}$	-
18:4 $\Delta^{6,9,12,15}$	21.1$\pm$
20:2 $\Delta^{11,14}$	-
20:3 $\Delta^{8,11,14}$	-
20:4 $\Delta^{5,8,11,14}$	-
20:5 $\Delta^{5,8,11,14,17}$	-
22:4 $\Delta^{7,10,13,16}$	-
22:5 $\Delta^{7,10,13,16,19}$	-
22:6 $\Delta^{4,7,10,13,16,19}$	-

Table 9 shows the substrate specificity of the elongase OtElo2 for various fatty acids.

5 The yeasts which had been transformed with the vector pOTE2 were cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC. Each value represents the mean (n=3) $\pm$ standard deviation.

10 The enzymatic activity shown in Table 9 clearly demonstrates that OTELO2 is a Δ^6 -elongase.

Example 19: Cloning genes from *Thalassiosira pseudonana*

15 The search for conserved regions in the protein sequences with the aid of the elongase genes with Δ^5 -elongase activity or Δ^6 -elongase activity which are shown in the application allowed the identification of two sequences with suitable motifs in a *Thalassiosira pseudonana* sequence database (genomic sequences). The sequences were the following:

Name of gene	SEQ ID	Amino acids
TpELO1 ($\Delta 5$ -elongase)	43	358
TpELO2 ($\Delta 5$ -elongase)	59	358
TpELO3 ($\Delta 6$ -elongase)	45	272

A 2 l culture of *T. pseudonana* was grown in f/2 medium (Guillard, R.R.L. 1975. Culture of phytoplankton for feeding marine invertebrates. In *Culture of Marine Invertebrate Animals* (Eds. Smith, W.L. and Chanley, M.H.), Plenum Press, New York, pp 29-60) for 14 d (= days) at a light intensity of 80 E/cm². After the cells had been spun down, RNA was isolated with the aid of the RNAeasy Kit from Quiagen (Valencia, CA, US) following the manufacturer's instructions. The mRNA was subjected to reverse transcription using the Marathon cDNA Amplification Kit (BD Biosciences) and adaptors were ligated in accordance with the manufacturer's instructions. Then, the cDNA library was used for the PCR for cloning expression plasmids by means of 5'- and 3'-RACE (rapid amplification of cDNA ends).

Example 20: Cloning expression plasmids for the heterologous expression in yeasts

The relevant primer pairs were selected in such a way that they bore the yeast consensus sequence for highly efficient translation (Kozak, Cell 1986, 44:283-292) next to the start codon. The amplification of the TpElo DNAs was carried out in each case using 1 μ l of cDNA, 200 μ M of dNTPs, 2.5 U of *Advantage* polymerase and 100 pmol of each primer in a total volume of 50 μ l. The PCR conditions were as follows: first denaturation for 5 minutes at 95°C, followed by 30 cycles of 30 seconds at 94°C, 1 minute at 55°C and 2 minutes at 72°C, and a last elongation step of 10 minutes at 72°C.

To clone the sequence for the heterologous expression in yeasts, the following oligonucleotides were used for the PCR reaction:

Name of gene and SEQ ID NO:	Primer sequence
TpELO1 ($\Delta 5$ -elongase), SEQ ID NO: 59	F:5'-accatgtgctcaccaccgccgctc (SEQ ID NO: 158) R:5'-ctacatggcaccagtaac (SEQ ID NO: 159)

TpELO2 ($\Delta 5$ -elongase), SEQ ID NO: 85	F:5'-accatgtgctcatcaccgccgctc (SEQ ID NO: 160) R:5'-ctacatggcaccagtaac (SEQ ID NO: 161)
TpELO3 ($\Delta 6$ -elongase), SEQ ID NO: 45	F:5'-accatggacgcctacaacgctgc (SEQ ID NO: 162) R:5'-ctaagcactcttctcttt (SEQ ID NO: 163)

*F=forward primer, R=reverse primer

The PCR products were incubated for 30 minutes at 21°C with the yeast expression
 5 vector pYES2.1-TOPO (Invitrogen) following the manufacturer's instructions. The PCR
 product is ligated into the vector by means of a T overhang and activity of a
 topoisomerase (Invitrogen). After incubation, *E. coli* DH5 α cells were transformed.
 Suitable clones were identified by PCR, the plasmid DNA was isolated by means of
 Qiagen DNAeasy Kit and verified by sequencing. The correct sequence was then
 10 transformed into the *Saccharomyces* strain INVSc1 (Invitrogen) by electroporation
 (1500 V). As a control, the blank vector pYES2.1 was transformed in parallel. The
 yeasts were subsequently plated onto complete uracil dropout minimal medium
 supplemented with 2% glucose. Cells which were capable of growing in the medium
 without uracil thus comprise the corresponding plasmids pYES2.1, pYES2.1-TpELO1,
 15 pYES2.1-TpELO2 and pYES2.1-TpELO3. After the selection, in each case two
 transformants were selected for further functional expression.

Example 21: Cloning expression plasmids for the seed specific expression in plants

20 A further transformation vector based on pSUN-USP is generated for the
 transformation of plants. To this end, NotI cleavage sites are introduced at the 5' and 3'
 termini of the coding sequences, using the following primer pair:

PSUN-TPELO1

25 Forward: 5'-GCGGCCGCACCATGTGCTCACCACCGCCGTC (SEQ ID NO: 152)
 Reverse: 3'-GCGGCCGCCTACATGGCACCAGTAAC (SEQ ID NO: 153)

PSUN-TPELO2

Forward: 5'-GCGGCCGCACCATGTGCTCATCACCACCGCCGTC (SEQ ID NO: 154)
 30 Reverse: 3'-GCGGCCGCCTACATGGCACCAGTAAC (SEQ ID NO: 155)

PSUN-TPELO3

Forward: 5'-GCGGCCGCaccatggacgcctacaacgctgc (SEQ ID NO: 156)

Reverse: 3'-GCGGCCGCCTAAGCACTCTTCTTCTTT (SEQ ID NO: 157)

5

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase)+ 25mM MgCl₂

5.00 µl 2mM dNTP

10

1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

PCR reaction conditions:

15

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

20

The PCR products are incubated with the restriction enzyme NotI for 16 hours at 37°C.

The plant expression vector pSUN300-USP is incubated in the same manner.

Thereafter, the PCR products and the 7624 bp vector are separated by agarose gel electrophoresis and the corresponding DNA fragments are excised. The DNA is

25

purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR products are ligated. The Rapid Ligation Kit from Roche is used for this purpose. The resulting plasmids pSUN-TPELO1, pSUN-TPELO2 and pSUN-TPELO3 are verified by sequencing.

30

pSUN300 is a derivative of plasmid pPZP (Hajdukiewicz, P, Svab, Z, Maliga, P., (1994)

The small versatile pPZP family of Agrobacterium binary vectors for plant

transformation. Plant Mol Biol 25:989-994). pSUN-USP originated from pSUN300, by inserting a USP promoter into pSUN300 in the form of an EcoRI fragment. The

polyadenylation signal is that of the octopine synthase gene from the A. tumefaciens Ti

35

plasmid (ocs-Terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P., Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and transcript map of the Agrobacterium tumefaciens Ti plasmid-encoded octopine synthase gene J. Mol. Appl. Genet. 1 (6), 499-511 (1982)). The USP promoter

corresponds to nucleotides 1 to 684 (Genbank Accession X56240), where part of the noncoding region of the USP gene is present in the promoter. The promoter fragment which is 684 base pairs in size was amplified by a PCR reaction and standard methods with the aid of a synthesized primer and by means of a commercially available T7 standard primer (Stratagene).

(Primer sequence:

5'-GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGGGGGATCC
GGATCTGCTGGCTATGAA-3'); SEQ ID NO: 151).

10

The PCR fragment was recut with EcoRI/Sall and inserted into the vector pSUN300 with OCS terminator. This gave rise to the plasmid with the name pSUN-USP. The construct was used for the transformation of *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

15

Lipids were extracted from yeasts and seeds as described for Example 6.

Example 22: Expression of TpELO1, TpELO2 and TpELO3 in yeasts

20 Yeasts which had been transformed with the plasmids pYES2, pYES2-TpELO1, pYES2-TpELO2 and pYES2-TpELO3 as in Example 4 were analyzed as follows:

The yeast cells from the main cultures were harvested by centrifugation ($100 \times g$, 5 min, 20°C) and washed with 100 mM NaHCO_3 , pH 8.0 in order to remove residual medium and fatty acids. Fatty acid methyl esters (FAMES) were prepared from the yeast cell sediments by acid methanolysis. To this end, the cell sediments were incubated for 1 hour at 80°C with 2 ml of 1 N methanolic sulfuric acid and 2% (v/v) dimethoxypropane. The FAMES were extracted by twice extracting with petroleum ether (PE). To remove nonderivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO_3 , pH 8.0, and 2 ml of distilled water. Thereafter, the PE phases were dried with Na_2SO_4 , evaporated under argon and taken up in 100 μl of PE. The samples were separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 μm , Agilent) in a Hewlett-Packard 6850 gas chromatograph with flame ionization detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of $5^{\circ}\text{C}/\text{min}$ and finally 10 minutes at 250°C (holding).

35

The signals were identified by comparing the retention times with corresponding fatty

acid standards (Sigma). The methodology is described for example in Napier and Michaelson, 2001, *Lipids* 36(8):761-766; Sayanova et al., 2001, *Journal of Experimental Botany*, 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

5

Example 23: Functional characterization of TpELO1 and TpELO3:

The substrate specificity of TpELO1 could be determined after expression and the feeding of different fatty acids (Figure 9). The substrates fed can be detected in large amounts in all of the transgenic yeasts. The transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the TpElo1 reaction. This means that the gene TpElo1 has been expressed functionally.

It can be seen from Table 10 that TpElo1 shows a narrow substrate specificity. TpElo1 was only capable of elongating the C20-fatty acids eicosapentaenoic acid and arachidonic acid, but preferred the ω 3-desaturated eicosapentaenoic acid.

The yeasts which had been transformed with the vector pYES2-TpELO1 were cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC.

Table 10: Expression of TpELO1 in yeast. Columns 1 and 3 show the control reactions for columns 2 (fed: 250 μ m 20:4 Δ 5,8,11,14) and 4 (fed: 250 μ m 20:4 Δ 5,8,11,14,17).

25

	Expression	Expression	Expression	Expression
Fatty acids	1	2	3	4
16:0	18.8	17.8	25.4	25.2
16:1 Δ^9	28.0	29.8	36.6	36.6
18:0	5.2	5.0	6.8	6.9
18:1 Δ^9	25.5	23.6	24.6	23.9
20:4 $\Delta^{5,8,11,14}$	22.5	23.4	-	-
22:4 $\Delta^{7,10,13,16}$	-	0.4	-	-
20:5 $\Delta^{5,8,11,14,17}$	-	-	6.6	6.5
22:5 $\Delta^{7,10,13,16,19}$	-	-	-	0.9
% conversion	0	1.7	0	12.2

The substrate specificity of TpElo3 could be determined after expression and the feeding of different fatty acids (Figure 10). The substrates fed can be detected in large amounts in all of the transgenic yeasts. The transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the TpElo3 reaction. This means that the gene TpElo3 has been expressed functionally.

It can be seen from Table 11 that TpElo3 shows a narrow substrate specificity. TpElo3 was only capable of elongating the C18-fatty acid γ -linolenic acid and stearidonic acid, but preferred the ω 3-desaturated stearidonic acid.

The yeasts which had been transformed with the vector pYES2-TpELO3 were cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC.

Table 11: Expression of TpELO3 in yeast. Column 1 shows the fatty acid profile of yeast without feeding. Column 2 shows the control reaction. In columns 3 to 6, the following were fed: γ -linolenic acid, stearidonic acid, arachidonic acid and eicosapentaenoic acid (250 μ m of each fatty acid).

Fatty acids	1	2	3	4	5	6
16:0	17.9	20.6	17.8	16.7	18.8	18.8
16:1 Δ^9	41.7	18.7	27.0	33.2	24.0	31.3
18:0	7.0	7.7	6.4	6.6	5.2	6.0
18:1 Δ^9	33.3	16.8	24.2	31.8	25.5	26.4
18:2 $\Delta^{9,12}$	-	36.1	-	-	-	-
18:3 $\Delta^{6,9,12}$	-	-	6.1	-	-	-
18:4 $\Delta^{6,9,12,15}$	-	-	-	1.7	-	-
20:2 $\Delta^{11,14}$	-	0	-	-	-	-
20:3 $\Delta^{8,11,14}$	-	-	18.5	-	-	-
20:4 $\Delta^{8,11,14,17}$	-	-	-	10.0	-	-
20:4 $\Delta^{5,8,11,14}$	-	-	-	-	22.5	-
22:4 $\Delta^{7,10,13,16}$	-	-	-	-	0	-
20:5 $\Delta^{5,8,11,14,17}$	-	-	-	-	-	17.4
22:5 $\Delta^{7,10,13,16,19}$	-	-	-	-	-	0
% conversion	0	0	75	85	0	0

Example 24: Cloning and expression plasmid for the heterologous expression of the Pi-omega3Des in yeasts

For the heterologous expression in yeasts, the Pi-omega3Des clone was cloned into the yeast expression vector pYES3 via PCR, using suitable Pi-omega3Des-specific primers. Here, exclusively the open reading frame, of the gene, which encodes the Pi-omega3Des protein was amplified and provided with two cleavage sites for cloning into the pYES3 expression vector:

- Forward Primer: 5'-TAAGCTTACATGGCGACGAAGGAGG (SEQ ID NO: 149)
Reverse Primer: 5'-TGGATCCACTTACGTGGACTTGGT (SEQ ID NO: 150)

Composition of the PCR mix (50 µl):

- 5.00 µl template cDNA
5.00 µl 10x buffer (Advantage polymerase)+ 25mM MgCl₂
5.00 µl 2mM dNTP
1.25 µl of each primer (10 pmol/µl of the 5'ATG primer and the 3' Stopp primer)
0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

20

PCR reaction conditions:

- Annealing temperature: 1 min 55°C
Denaturation temperature: 1 min 94°C
Elongation temperature: 2 min 72°C
Number of cycles: 35

- The PCR product was incubated with the restriction enzymes HindIII and BamHI for 2 hours at 37°C. The yeast expression vector pYES3 (Invitrogen) was incubated in the same manner. Thereafter, the 1104 bp PCR product and the vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and desaturase cDNA were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pYES3-Pi-omega3Des was verified by sequencing and transformed into the Saccharomyces strain INVSc1 (Invitrogen) by means of electroporation (1500 V). pYES3 was transformed in parallel to act as a control. Thereafter, the yeasts were plated onto complete minimal dropout tryptophan medium supplemented with 2%

glucose. Cells which were capable of growing in the medium without tryptophan thus comprise the relevant plasmids pYES3, pYES3-Pi-omega3Des. Following selection, in each case two transformants were selected for the further functional expression.

5 Example 25: Cloning expression plasmids for the seed specific expression in plants

A further transformation vector based on pSUN-USP was generated for the transformation of plants. To this end, NotI cleavage sites were introduced at the 5' and 3' termini of the coding sequence, using the following primer pair

10

PSUN-Pi-omega3Des

Reverse: 3'-GCGGCCGCTTACGTGGACTTGGTC (SEQ ID NO: 147)

Forward: 5'-GCGGCCGCatGGCGACGAAGGAGG (SEQ ID NO: 148)

15 Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase)+ 25mM MgCl₂

5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

20 0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

PCR reaction conditions:

25 Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

30 The PCR products were incubated with the restriction enzyme NotI for 4 hours at 37°C. The plant expression vector pSUN300-USP was incubated in the same manner.

Thereafter, the PCR products and the 7624 bp vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's

35 instructions. Thereafter, vector and PCR products were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pSUN-Piomega3Des was verified by sequencing.

Example 26: Expression of Pi-omega3Des in yeasts

Yeasts which had been transformed with the plasmid pYES3 or pYES3-Pi-omega3Des, as described in Example 24, were analyzed as follows:

- 5 The yeast cells from the main cultures were harvested by centrifugation ($100 \times g$, 5 min, 20°C) and washed with 100 mM NaHCO_3 , pH 8.0 in order to remove residual medium and fatty acids. Fatty acid methyl esters (FAMES) were prepared from the yeast cell sediments by acid methanolysis. To this end, the cell sediments were incubated for 1 hour at 80°C with 2 ml of 1 N methanolic sulfuric acid and 2% (v/v)
- 10 dimethoxypropane. The FAMES were extracted by twice extracting with petroleum ether (PE). To remove nonderivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO_3 , pH 8.0, and 2 ml of distilled water. Thereafter, the PE phases were dried with Na_2SO_4 , evaporated under argon and taken up in 100 μl of PE. The samples were separated on a DB-23 capillary column (30 m,
- 15 0.25 mm, 0.25 μm , Agilent) in a Hewlett-Packard 6850 gas chromatograph with flame ionization detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of $5^{\circ}\text{C}/\text{min}$ and finally 10 minutes at 250°C (holding).

- The signals were identified by comparing the retention times with corresponding fatty acid standards (Sigma). The methodology is described for example in Napier and
- 20 Michaelson, 2001, *Lipids* 36(8):761-766; Sayanova et al., 2001, *Journal of Experimental Botany*, 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

25 **Example 23: Functional characterization of Pi-omega3Des:**

- The substrate specificity of Pi-omega3Des could be determined after expression and the feeding of different fatty acids (Figures 12 to 18). The substrates fed are present in large amounts in all of the transgenic yeasts, which proves that these fatty acids have
- 30 been taken up into the yeasts. The transgenic yeasts demonstrate the synthesis of novel fatty acids, the products of the Pi-omega3Des reaction. This means that the gene Pi-omega3Des has been expressed functionally.

- Figure 12 represents the desaturation of linoleic acid ($18:2 \omega_6$ -fatty acid) to give
- 35 α -linolenic acid ($18:3 \omega_3$ -fatty acid) by Pi-omega3Des. The fatty acid methyl esters were synthesized by subjecting intact cells which had been transformed with the blank vector pYES2 (Figure 12 A) or the vector pYES3- Pi-omega3Des (Figure 12 B) to acid methanolysis. The yeasts were cultured in minimal medium in the presence of $18:2^{\Delta 9,12}$.

fatty acid (300 μ m). Thereafter, the FAMES were analyzed via GLC.

Figure 13 represents the desaturation of γ -linolenic acid (18:3 ω 6-fatty acid) to give stearidonic acid (18:4 ω 3-fatty acid) by Pi-omega3Des. The fatty acid methyl esters were synthesized by subjecting intact cells which had been transformed with the blank vector pYES2 (Figure 13 A) or the vector pYes3-Pi-omega3Des (Figure 13 B) to acid methanolysis. The yeasts were cultured in minimal medium in the presence of γ -C18:3 $^{\Delta 6,9,12}$ -fatty acid (300 μ m). Thereafter, the FAMES were analyzed via GLC.

Figure 14 represents the desaturation of C20:2- ω 6-fatty acid to give C20:3- ω 3-fatty acid by Pi-omega3Des. The fatty acid methyl esters were synthesized by subjecting intact cells which had been transformed with the blank vector pYES2 (Figure 14 A) or the vector pYes3-Pi-omega3Des (Figure 14 B) to acid methanolysis. The yeasts were cultured in minimal medium in the presence of C20:2 $^{\Delta 11,14}$ -fatty acid (300 μ m). Thereafter, the FAMES were analyzed via GLC.

Figure 15 represents the desaturation of C20:3- ω 6-fatty acid to give C20:4- ω 3-fatty acid by Pi-omega3Des. The fatty acid methyl esters were synthesized by subjecting intact cells which had been transformed with the blank vector pYES2 (Figure 15 A) or the vector pYes3-Pi-omega3Des (Figure 15 B) to acid methanolysis. The yeasts were cultured in minimal medium in the presence of C20:3 $^{\Delta 8,11,14}$ -fatty acid (300 μ m). Thereafter, the FAMES were analyzed via GLC.

Figure 16 shows the desaturation of arachidonic acid (C20:4- ω 6-fatty acid) to give eicosapentaenoic acid (C20:5- ω 3-fatty acid) by Pi-omega3Des.

The fatty acid methyl esters were synthesized by subjecting intact cells which had been transformed with the blank vector pYES2 (Figure 16 A) or the vector pYes3-Pi-omega3Des (Figure 16 B) to acid methanolysis. The yeasts were cultured in minimal medium in the presence of C20:4 $^{\Delta 5,8,11,14}$ -fatty acid (300 μ m). Thereafter, the FAMES were analyzed via GLC.

Figure 17 represents the desaturation of docosatetraenoic acid (C22:4- ω 6-fatty acid) to give docosapentaenoic acid (C22:5- ω 3-fatty acid) by Pi-omega3Des. The fatty acid methyl esters were synthesized by subjecting intact cells which had been transformed with the blank vector pYES2 (Figure 17 A) or the vector pYes3-Pi-omega3Des (Figure 17 B) to acid methanolysis. The yeasts were cultured in minimal medium in the presence of C22:4 $^{\Delta 7,10,13,16}$ -fatty acid (300 μ m). Thereafter, the FAMES were analyzed

via GLC.

The substrate specificity of Pi-omega3Des with regard to different fatty acids can be seen from Figure 18. The yeasts which had been transformed with the vector pYes3-
 5 Pi-omega3Des were cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC. Each value represents a mean of three measurements. The conversion rates (% desaturation) were calculated using the formula:

10
$$[\text{product}]/([\text{product}]+[\text{substrate}])*100.$$

As described in Example 9, Pi-omega3Des can also be used for generating transgenic plants. Then, the lipids can be extracted from the seeds of these plants as described under Example 6.

15

Example 28: Cloning desaturase genes from *Ostreococcus tauri*

The search for conserved regions in the protein sequences with the aid of conserved motifs (His boxes, Domergue et al. 2002, Eur. J. Biochem. 269, 4105-4113) allowed
 20 the identification of five sequences with corresponding motifs in an *Ostreococcus tauri* sequence database (genomic sequences). The sequences were the following:

Name of gene	SEQ ID	Amino acids	Homology
OtD4	SEQ ID NO: 95	536	Δ 4-desaturase
OtD5.1	SEQ ID NO: 91	201	Δ 5-desaturase
OtD5.2	SEQ ID NO: 93	237	Δ 5-desaturase
OtD6.1	SEQ ID NO: 89	456	Δ 6-desaturase
OtFad2	SEQ ID NO: 107	361	Δ 12-desaturase

The alignments for finding homologies of the individual genes were carried out using
 25 the tBLASTn algorithm (Altschul et al., J. Mol. Biol. 1990, 215: 403-410).

The cloning procedure was as follows:

40 ml of an *Ostreococcus tauri* culture in the stationary phase were spun down,
 30 resuspended in 100 μ l of double-distilled water and stored at -20°C. The respective genomic DNAs were amplified on the basis of the PCR process. The relevant primer

pairs were selected in such a way that they bore the yeast consensus sequence for highly efficient translation (Kozak, Cell 1986, 44:283-292) next to the start codon. The amplification of the OtDes DNAs was carried out in each case using 1 µl of defrosted cells, 200 µm of dNTPs, 2.5 U *Taq* polymerase and 100 pmol of each primer in a total
5 volume of 50 µl. The PCR conditions were as follows: first denaturation for 5 minutes at 95°C, followed by 30 cycles of 30 seconds at 94°C, 1 minute at 55°C and 2 minutes at 72°C, and a last elongation step of 10 minutes at 72°C.

The following primers were employed in the PCR:

10

OtDes6.1 Forward: 5'ggtaccacataatgtcgtggagacggaaaataacg3' (SEQ ID NO: 145)

OtDes6.1 Reverse: 5'ctcgagttacgccgtcttccggagtgttgcc3' (SEQ ID NO: 146)

Example 29: Cloning expression plasmids for the heterologous expression in yeasts:

15

To characterize the function of the desaturase OtDes6.1 (=Δ6-desaturase) from *Ostreococcus tauri*, the open reading frame of the DNA was cloned downstream of the galactose-inducible GAL1 promoter of pYES2.1/V5-His-TOPO (Invitrogen), giving rise to the corresponding clone pYES2.1-OtDes6.1. Further desaturase genes from

20

Ostreococcus can be cloned analogously.

The *Saccharomyces cerevisiae* strain 334 was transformed by electroporation (1500 v) with the vector pYES2.1-OtDes6.1. A yeast which was transformed with the blank vector pYES2 was used as the control. The transformed yeasts were selected on
25 complete minimal dropout uracil medium (CMdum) agar plates supplemented with 2% glucose. After the selection, in each case three transformants were selected for the further functional expression.

To express the OtDes6.1 desaturase, precultures of in each case 5 ml of dropout uracil
30 CMdum liquid medium supplemented with 2% (w/v) raffinose were inoculated with the selected transformants and incubated for 2 days at 30°C, 200 rpm. 5 ml of CMdum liquid medium (without uracil) supplemented with 2% raffinose and 300 µm of various fatty acids were then inoculated with the precultures to an OD₆₀₀ of 0.05. Expression was induced by addition of 2% (w/v) galactose. The cultures were incubated for a
35 further 96 hours at 20°C.

Example 30: Cloning of expression plasmids for the seed-specific expression in plants

A further transformation vector based on pSUN-USP is generated for the transformation of plants. To this end, NotI cleavage sites are introduced at the 5' and 3' termini of the coding sequences, using PCR. The corresponding primer sequences are derived from the 5' and 3' regions of the desaturases.

5

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase)+ 25mM MgCl₂

10 5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

15

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

20 Number of cycles: 35

The PCR products were incubated with the restriction enzyme NotI for 16 hours at 37°C. The plant expression vector pSUN300-USP was incubated in the same manner.

Thereafter, the PCR products and the vector were separated by agarose gel

25 electrophoresis and the corresponding DNA fragments were excised. The DNA was

purified by means of the Qiagen Gel Purification Kit following the manufacturer's

instructions. Thereafter, vector and PCR products were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmids were verified by

sequencing.

30

pSUN300 is a derivative of plasmid pPZP (Hajdukiewicz, P., Svab, Z., Maliga, P., (1994)

The small versatile pPZP family of *Agrobacterium* binary vectors for plant

transformation. *Plant Mol Biol* 25:989-994). pSUN-USP originated from pSUN300, by

inserting a USP promoter into pSUN300 in the form of an EcoRI fragment. The

35 polyadenylation signal is that of the *Ostreococcus* gene from the *A. tumefaciens* Ti

plasmid (ocs-Terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P.,

Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and

transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine

synthase gene J. Mol. Appl. Genet. 1 (6), 499-511 (1982)). The USP promoter corresponds to nucleotides 1 to 684 (Genbank Accession X56240), where part of the noncoding region of the USP gene is present in the promoter. The promoter fragment which is 684 base pairs in size was amplified by a PCR reaction and standard methods with the aid of a synthesized primer and by means of a commercially available T7 standard primer (Stratagene). (Primer sequence: 5'–
GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGGGGGATCC
GGATCTGCTGGCTATGAA–3', SEQ ID NO: 144).

The PCR fragment was recut with EcoRI/Sall and inserted into the vector pSUN300 with OCS terminator. This gave rise to the plasmid with the name pSUN-USP. The construct was used for the transformation of *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

Example 31: Expression of OtDes6.1 in yeasts

Yeasts which had been transformed with the plasmids pYES2, pYES2- OtDes6.2 as described in Example 4 were analyzed as follows:

The yeast cells from the main cultures were harvested by centrifugation (100 x g, 5 min, 20°C) and washed with 100 mM NaHCO₃, pH 8.0 to remove residual medium and fatty acids. Starting with the yeast cell sediments, fatty acid methyl esters (FAMES) were prepared by acid methanolysis. To this end, the cell sediments were incubated for one hour at 80°C together with 2 ml of 1 N methanolic sulfuric acid and 2% (v/v) of dimethoxypropane. The FAMES were extracted twice with petroleum ether (PE). To remove nonderivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO₃, pH 8.0 and 2 ml of distilled water. Thereafter, the PE phases were dried with Na₂SO₄, evaporated under argon and taken up in 100 µl of PE. The samples were separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 µm, Agilent) in a Hewlett-Packard 6850 gas chromatograph equipped with flame ionization detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of 5°C/min and finally 10 min at 250°C (holding).

The signals were identified by comparing the retention times with corresponding fatty acid standards (Sigma). The methodology is described for example in Napier and Michaelson, 2001, Lipids. 36(8):761-766; Sayanova et al., 2001, Journal of Experimental Botany. 52(360):1581-1585, Sperling et al., 2001, Arch. Biochem. Biophys. 388(2):293-298 and Michaelson et al., 1998, FEBS Letters. 439(3):215-218.

Example 32: Functional characterization of desaturases from *Ostreococcus*:

The substrate specificity of desaturases can be determined after expression in yeast
 5 (see examples Cloning desaturase genes, Yeast expression) by feeding by means of
 different yeasts. Descriptions for determining the individual activities are found in WO
 93/11245 for $\Delta 15$ -desaturases, WO 94/11516 for $\Delta 12$ -desaturases, WO 93/06712, US
 5,614,393, US 5614393, WO 96/21022, WO 0021557 and WO 99/27111 for $\Delta 6$ -
 desaturases, Qiu et al. 2001, J. Biol. Chem. 276, 31561-31566 for $\Delta 4$ -desaturases,
 10 Hong et al. 2002, Lipids 37, 863-868 for $\Delta 5$ -desaturases.

Table 12 represents the substrate specificity of the desaturase OtDes6.1 with regard to
 different fatty acids. The substrate specificity of OtDes6.1 was determined after
 expression and feeding of various fatty acids. The substrates which have been fed can
 15 be detected in large amounts in all of the transgenic yeasts. The transgenic yeasts
 demonstrated the synthesis of novel fatty acids, the products of the OtDes6.2 reaction
 (Fig. 20). This means that the gene OtDes6.1 has been expressed functionally.

The yeasts which had been transformed with the vector pYES2-OtDes6.1 were
 20 cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid
 methyl esters were synthesized by subjecting intact cells to acid methanolysis.
 Thereafter, the FAMES were analyzed via GLC. Each value represents the mean (n=3)
 $\pm$ standard deviation. The activity corresponds to the conversion rate calculated using
 the formula [substrate/(substrate+product)*100].

25 It can be seen from Table 12 that OtDes6.1 shows substrate specificity for linoleic and
 linolenic acid (18:2 and 18:3) since the highest activities are obtained with these fatty
 acids. In contrast, the activity for oleic acid (18:1) and palmitoleic acid (16:1) is
 markedly lower. The preferred conversion of linoleic and linolenic acid demonstrates
 30 that this desaturase is suitable for the production of polyunsaturated fatty acids.

Substrates	Activity in %
16:1 ^{$\Delta 9$}	5.6
18:1 ^{$\Delta 9$}	13.1
18:2 ^{$\Delta 9,12$}	68.7
18:3 ^{$\Delta 9,12,15$}	64.6

Figure 20 shows the conversion of linoleic acid by OtDes6.1. The FAMES were analyzed via gas chromatography. The substrate which has been fed (C18:2) is converted into γ -C18:3. Both the starting material and the resulting product are indicated by arrows.

5

Figure 21 represents the conversion of linoleic acid (= LA) and α -linolenic acid (= ALA) in the presence of OtDes6.1 to give γ -linolenic acid (= GLA) and stearidonic acid (= STA), respectively (Figure 21 A and C). Moreover, Figure 21 shows the conversion of linoleic acid (= LA) and α -linolenic acid (= ALA) in the presence of the Δ 6-desaturase OtDes6.1 together with the *Physcomitrella patens* Δ 6-elongase PSE1 (Zank et al. 2002, Plant J. 31:255-268) and the *Phaeodactylum tricornutum* Δ 5-desaturase PtD5 (Domergue et al. 2002, Eur. J. Biochem. 269, 4105-4113) to give dihomogamma-linolenic acid (= DHGLA) and arachidonic acid (= ARA, Figure 21 B) and dihomostearidonic acid (= DHSTA) and eicosapentaenoic acid (= EPA, Figure 21 D), respectively. Figure 21 shows clearly that the reaction products GLA and STA of the Δ 6-desaturase OtDes6.1 in the presence of the Δ 6-elongase PSE1 is elongated virtually quantitatively to give DHGLA and DHSTA, respectively. The subsequent desaturation by the Δ 5-desaturase PtD5 to give ARA and EPA, respectively, also proceeds smoothly. Approximately 25-30% of the elongase product is desaturated (Figure 21 B and D).

20

Table 13 which follows gives an overview of the *Ostreococcus* desaturases which have been cloned:

<i>Ostreococcus tauri</i> desaturases							
Name	bp	aa	Homology	Cyt. B5	His box1	His box2	His box3
OtD4	1611	536	Δ 4-desaturase	HPGG	HCANH	WRYHHQVSHH	QVEHHLFP
OtD5.1	606	201	Δ 5-desaturase	-	-	-	QVVHHLFP
OtD5.2	714	237	Δ 5-desaturase	-	-	WRYHHMVSHH	QIEHHLFP
OtD6.1	1443	480	Δ 6-desaturase	HPGG	HEGGH	WNSMHNKHH	QVIHHLFP
OtFAD2	1086	361	Δ 12-desaturase	-	HECGH	WQRSHAVHH	HVAHH

Example 33: Cloning desaturase genes from *Thalassiosira pseudonana*

- 5 The search for conserved regions in the protein sequences with the aid of conserved motifs (His boxes, see motifs) allowed the identification of six sequences with corresponding motifs in an *Thalassiosira pseudonana* sequence database (genomic sequences). The sequences were the following:

Name of gene	SEQ ID	Amino acids	Homology
TpD4	SEQ ID NO: 103	503	Δ 4-desaturase
TpD5-1	SEQ ID NO: 99	476	Δ 5-desaturase
TpD5-2	SEQ ID NO: 101	482	Δ 5-desaturase
TpD6	SEQ ID NO: 97	484	Δ 6-desaturase
TpFAD2	SEQ ID NO: 109	434	Δ 12-desaturase
TpO3	SEQ ID NO: 105	418	ω 3-desaturase

- 10 The cloning procedure was as follows:

- 40 ml of an *Thalassiosira pseudonana* culture in the stationary phase were spun down, resuspended in 100 μ l of double-distilled water and stored at -20°C. The respective genomic DNAs were amplified on the basis of the PCR method. The relevant primer
 15 pairs were selected in such a way that they bore the yeast consensus sequence for highly efficient translation (Kozak, Cell 1986, 44:283-292) next to the start codon. The amplification of the TpDes DNAs was carried out in each case using 1 μ l of defrosted cells, 200 μ M of dNTPs, 2.5 U *Taq* polymerase and 100 pmol of each primer in a total volume of 50 μ l. The PCR conditions were as follows: first denaturation for 5 minutes at
 20 95°C, followed by 30 cycles of 30 seconds at 94°C, 1 minute at 55°C and 2 minutes at 72°C, and a last elongation step of 10 minutes at 72°C.

Example 34: Cloning expression plasmids for the heterologous expression in yeasts:

- 25 To characterize the function of the desaturases from *Thalassiosira pseudonana*, the open reading frame of the respective DNA was cloned downstream of the galactose-inducible GAL1 promoter of pYES2.1/V5-His-TOPO (Invitrogen), giving rise to the corresponding pYES2.1 clone.
- 30 The *Saccharomyces cerevisiae* strain 334 is transformed by electroporation (1500 v)

with the vectors pYES2.1-TpDesaturasen. A yeast which is transformed with the blank vector pYES2 is used as the control. The transformed yeasts are selected on complete minimal dropout uracil medium (CMdum) agar plates supplemented with 2% glucose. After the selection, in each case three transformants are selected for the further functional expression.

To express the Tp desaturases, initially precultures of in each case 5 ml of dropout uracil CMdum liquid medium supplemented with 2% (w/v) raffinose are inoculated with the selected transformants and incubated for 2 days at 30°C, 200 rpm. 5 ml of liquid CMdum medium (without uracil) supplemented with 2% raffinose and 300 µm of various fatty acids are then inoculated with the precultures to an OD₆₀₀ of 0.05. The expression is induced by addition of 2% (w/v) galactose. The cultures are incubated for a further 96 hours at 20°C.

Example 35: Cloning of expression plasmids for the seed-specific expression in plants

A further transformation vector based on pSUN-USP is generated for the transformation of plants. To this end, NotI cleavage sites are introduced at the 5' and 3' termini of the coding sequences, using PCR. The corresponding primer sequences are derived from the 5' and 3' regions of the desaturases.

Composition of the PCR mix (50 µl):

- 5.00 µl template cDNA
- 5.00 µl 10x buffer (Advantage polymerase)+ 25mM MgCl₂
- 5.00 µl 2mM dNTP
- 1.25 µl of each primer (10 pmol/µl)
- 0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

PCR reaction conditions:

- Annealing temperature: 1 min 55°C
- Denaturation temperature: 1 min 94°C
- Elongation temperature: 2 min 72°C
- Number of cycles: 35

The PCR products are incubated with the restriction enzyme NotI for 16 hours at 37°C. The plant expression vector pSUN300-USP is incubated in the same manner.

Thereafter, the PCR products and the vector are separated by agarose gel electrophoresis and the corresponding DNA fragments are excised. The DNA was

5 purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR products are ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmids are verified by sequencing.

10 pSUN300 is a derivative of plasmid pPZP (Hajdukiewicz, P., Svab, Z., Maliga, P., (1994)

The small versatile pPZP family of *Agrobacterium* binary vectors for plant transformation. *Plant Mol Biol* 25:989-994). pSUN-USP originated from pSUN300, by inserting a USP promoter into pSUN300 in the form of an EcoRI fragment. The polyadenylation signal is the OCS gene from the *A. tumefaciens* Ti plasmid (ocs-

15 Terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P., Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine synthase gene *J. Mol. Appl. Genet.* 1 (6), 499-511 (1982)). The USP promoter corresponds to nucleotides 1 to 684 (Genbank Accession X56240), where part of the noncoding region of the USP

20 gene is present in the promoter. The promoter fragment which is 684 base pairs in size was amplified by a PCR reaction and standard methods with the aid of a synthesized primer and by means of a commercially available T7 standard primer (Stratagene). (Primer sequence: 5'-

GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGCGGGGATCC

25 GGATCTGCTGGCTATGAA-3', SEQ ID NO: 143).

The PCR fragment was recut with EcoRI/SalI and inserted into the vector pSUN300 with OCS terminator. This gave rise to the plasmid with the name pSUN-USP. The construct was used for the transformation of *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

30

Example 36: Expression of Tp desaturases in yeasts

Yeasts which have been transformed with the plasmids pYES2 and pYES2-TpDesaturase as described in Example 4 were analyzed as follows:

35

The yeast cells from the main cultures are harvested by centrifugation (100 x g, 5 min, 20°C) and washed with 100 mM NaHCO₃, pH 8.0 to remove residual medium and fatty acids. Starting with the yeast cell sediments, fatty acid methyl esters (FAMES) are

prepared by acid methanolysis. To this end, the cell sediments are incubated for one hour at 80°C together with 2 ml of 1 N methanolic sulfuric acid and 2% (v/v) of dimethoxypropane. The FAMEs were extracted twice with petroleum ether (PE). To remove nonderivatized fatty acids, the organic phases are washed in each case once
5 with 2 ml of 100 mM NaHCO₃, pH 8.0 and 2 ml of distilled water. Thereafter, the PE phases are dried with Na₂SO₄, evaporated under argon and taken up in 100 µl of PE. The samples are separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 µm, Agilent) in a Hewlett-Packard 6850 gas chromatograph equipped with flame ionization
10 detector. The conditions for the GLC analysis are as follows: the oven temperature is programmed from 50°C to 250°C with an increment of 5°C/min and finally 10 min at 250°C (holding).

The signals are identified by comparing the retention times with corresponding fatty acid standards (Sigma). The methodology is described for example in Napier and
15 Michaelson, 2001, *Lipids*. 36(8):761-766; Sayanova et al., 2001, *Journal of Experimental Botany*. 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

Example 37: Functional characterization of desaturases from *Thalassiosira*
20 *pseudonana*:

The substrate specificity of desaturases can be determined after expression in yeast (see examples Cloning desaturase genes, Yeast expression) by feeding by means of different yeasts. Descriptions for determining the individual activities are found in WO
25 93/11245 for Δ^{15} -desaturases, WO 94/11516 for Δ^{12} -desaturases, WO 93/06712, US 5,614,393, US 5614393, WO 96/21022, WO 0021557 and WO 99/27111 for Δ^6 -desaturases, Qiu et al. 2001, *J. Biol. Chem.* 276, 31561-31566 for Δ^4 -desaturases, Hong et al. 2002, *Lipids* 37, 863-868 for Δ^5 -desaturases.

30 The activity of the individual desaturases is calculated from the conversion rate using the formula [substrate/(substrate+product)*100]

Tables 11 and 12 which follow give an overview of the cloned *Thalassiosira pseudonana* desaturases.

Table 14: Length and characteristic features of the cloned *Thalassiosira pseudonana* desaturases

Desaturase	cDNA (bp)	Protein (aa)	Cyt. B5	His box1	His box2	His box3
TpD4	1512	503	HPGG	HDGNH	WELQHMLGHH	QIEHHLFP
TpD5-1	1431	476	HPGG	HDANH	WMAQHWTHH	QVEHHLFP
TpD5-2	1443	482	HPGG	HDANH	WLAQHWTHH	QVEHHLFP
TpD6	1449	484	HPGG	HDFLH	WKNKHNGHH	QVDHHLFP
TpFAD2 (d12)	1305	434	-	HECGH	HAKHH	HVAHHLFH
TpO3	1257	419	-	HDAGH	WLFMVTYLQHH	HVVHHLF

5 Table 15: Length, exons, homology and identities of the cloned desaturases.

Des.	GDN A (bp)	Exon 1	Exon 2	First Blast Hit	Hom./Iden.
TpD4	2633	496-1314	1571-2260	Thrautochitrium D4-des	56%/43%
TpD5-1	2630	490-800	900-2019	Phaeodactylum D5-des	74%/62%
TpD5-2	2643	532-765	854-2068	Phaeodactylum D5-des	72%/61%
TpD6	2371	379-480	630-1982	Phaeodactylum D6-des	83%/69%
TpFAD2	2667	728-2032	-	Phaeodactylum FAD2	76%/61%
TpO3	2402	403-988	1073-1743	Chaenorhabdids Fad2	49%/28%

The $\Delta 12$ -desaturase genes from *Ostreococcus* and *Thalassiosira* can also be cloned analogously to the above examples.

10

Example 38: Cloning elongase genes from *Xenopus laevis* and *Ciona intestinalis*

- The search for conserved regions (see consensus sequences, SEQ ID NO: 115 and SEQ ID NO: 116) in the protein sequences in gene databases (Genbank) with the aid of the elongase genes with $\Delta 5$ -elongase activity or $\Delta 6$ -elongase activity, which are detailed in the application, allowed the identification and isolation of further elongase sequences from other organisms. Further sequences were identified in each case from
- 5 X. laevis and from C. intestinalis, using suitable motifs. The sequences were the following:

Name of gene	Organism	Genbank No.	SEQ ID NO:	Amino acids
ELO(XI)	Xenopus laevis	BC044967	117	303
ELO(Ci)	Ciona intestinalis	AK112719	119	290

- 10 The cDNA clone of X. laevis was obtained from the NIH (National Institute of Health) [Genetic and genomic tools for Xenopus research: The NIH Xenopus initiative, Dev. Dyn. 225 (4), 384-391 (2002)].

- The cDNA clone of C. intestinalis was obtained from the University of Kyoto [Satou, Y., Yamada, L., Mochizuki, Y., Takatori, N., Kawashima, T., Sasaki, A., Hamagu-chi, M., Awazu, S., Yagi, K., Sasakura, Y., Nakayama, A., Ishikawa, H., Inaba, K. and Satoh, N. "A cDNA resource from the basal chordate Ciona intestinalis" JOURNAL Genesis 33 (4), 153-154 (2002)].
- 15

- 20 Example 39: Cloning expression plasmids for the heterologous expression in yeasts

- The elongase DNAs were amplified in each case using 1 μ l of cDNA, 200 μ M dNTPs, 2.5 U of *Advantage* polymerase and 100 pmol of each primer in a total volume of 50 μ l. The PCR conditions were as follows: first denaturation for 5 minutes at 95°C, followed
- 25 by 30 cycles of 30 seconds at 94°C, 1 minute at 55°C and 2 minutes at 72°C, and a final elongation step of 10 minutes at 72°C.

To clone the sequence for heterologous expression in yeasts, the following oligonucleotides were used for the PCR reaction:

Name of gene and SEQ ID NO:	Primer sequence
ELO(XI) SEQ ID NO: 121 SEQ ID NO: 122	F:5'-AGGATCC <u>ATGGC</u> CTTCAAGGAGCTCACATC R:5'- CCTCGAGTCAATGGTTTTTGCTTTTCAATGCACCG
ELO(Ci) SEQ ID NO: 123 SEQ ID NO: 124	F:5'-TAAGCTT <u>ATGG</u> ACGTACTTCATCGT R:5'-TCAGATCTTTAATCGGTTTTACCATT

*F=forward primer, R=reverse primer

The PCR products were incubated for 30 minutes at 21°C with the yeast expression vector pYES2.1-TOPO (Invitrogen) following the manufacturer's instructions. The PCR product is ligated into the vector by means of a T overhang and activity of a topoisomerase (Invitrogen). After incubation, *E. coli* DH5α cells were transformed. Suitable clones were identified by PCR, the plasmid DNA was isolated by means of Qiagen DNAeasy Kit and verified by sequencing. The correct sequence was then transformed into the *Saccharomyces* strain INVSc1 (Invitrogen) by electroporation (1500 V). As a control, the blank vector pYES2.1 was transformed in parallel. The yeasts were subsequently plated onto complete uracil dropout minimal medium supplemented with 2% glucose. Cells which were capable of growing in the medium without uracil thus comprise the corresponding plasmids pYES2.1, pYES2.1-ELO(XI) and pYES2.1-ELO(Ci). After the selection, in each case two transformants were selected for further functional expression.

Example 40: Cloning expression plasmids for the seed-specific expression in plants

A further transformation vector based on pSUN-USP is generated for the transformation of plants. To this end, NotI cleavage sites are introduced at the 5' and 3' ends of the coding sequence, using the following primer pair:

pSUN-ELO(XI)

25

Forward: 5'-GCGGCCGCACCATATGGCCTTCAAGGAGCTCACATC

(SEQ ID NO: 125)

Reverse: 3'-GCGGCCGCCTTCAATGGTTTTTGCTTTTCAATGCACCG

(SEQ ID NO: 126)

30

pSUN-ELO(Ci)

Forward: 5'-GCGGCCGCACCATGGACGTACTTCATCGT

(SEQ ID NO: 127)

Reverse: 3'-GCGGCCGCTTTAATCGGTTTTACCATT

(SEQ ID NO: 128)

5

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

5.00 µl 2mM dNTP

10 1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

PCR reaction conditions:

15

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

20

The PCR products were incubated with the restriction enzyme NotI for 16 hours at 37°C. The plant expression vector pSUN300-USP was incubated in the same manner.

Thereafter, the PCR products and the 7624 bp vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was

25

purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR products were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmids pSUN-ELO(XI) and pSUN-ELO(Ci) were verified by sequencing.

30

pSUN300 is a derivative of plasmid pPZP (Hajdukiewicz, P., Svab, Z., Maliga, P., (1994)

The small versatile pPZP family of *Agrobacterium* binary vectors for plant

transformation. *Plant Mol Biol* 25:989-994). pSUN-USP originated from pSUN300, by

inserting a USP promoter into pSUN300 in the form of an EcoRI fragment. The

polyadenylation signal is that of the Octopine synthase gene from the *A. tumefaciens* Ti

35

plasmid (ocs-Terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P.,

Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and

transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine

synthase gene *J. Mol. Appl. Genet.* 1 (6), 499-511 (1982)). The USP promoter

corresponds to nucleotides 1 to 684 (Genbank Accession X56240), where part of the noncoding region of the USP gene is present in the promoter. The promoter fragment which is 684 base pairs in size was amplified by a PCR reaction and standard methods with the aid of a synthesized primer and by means of a commercially available T7 standard primer (Stratagene).

Primer sequence:

5'-GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGGGGGATCC
GGATCTGCTGGCTATGAA-3' (SEQ ID NO: 129).

10

The PCR fragment was recut with EcoRI/Sall and inserted into the vector pSUN300 with OCS terminator. This gave rise to the plasmid with the name pSUN-USP. The construct was used for the transformation of *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

15

Lipids were extracted from yeasts and seeds as described for Example 6.

Example 41: Expression of ELO(XI) and ELO(Ci) in yeasts

20 Yeasts which had been transformed with the plasmids pYES2, pYES2-ELO(XI) and pYES2-ELO(Ci) as in Example 4 were analyzed as follows:

The yeast cells from the main cultures were harvested by centrifugation ($100 \times g$, 5 min, 20°C) and washed with 100 mM NaHCO_3 , pH 8.0 in order to remove residual medium and fatty acids. Fatty acid methyl esters (FAMES) were prepared from the yeast cell sediments by acid methanolysis. To this end, the cell sediments were incubated for 1 hour at 80°C with 2 ml of 1N methanolic sulfuric acid and 2% (v/v) dimethoxypropane. The FAMES were extracted by twice extracting with petroleum ether (PE). To remove non-derivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO_3 , pH 8.0, and 2 ml of distilled water. Thereafter, the PE phases were dried with Na_2SO_4 , evaporated under argon and taken up in 100 μl of PE. The samples were separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 μm , Agilent) in a Hewlett-Packard 6850 gas chromatograph with flame ionization detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of $5^{\circ}\text{C}/\text{min}$ and finally 10 minutes at 250°C (holding).

35

The signals were identified by comparing the retention times with corresponding fatty

acid standards (Sigma). The methodology is described for example in Napier and Michaelson, 2001, *Lipids* 36(8):761-766; Sayanova et al., 2001, *Journal of Experimental Botany*, 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

5

Example 42: Functional characterization of ELO(XI) and ELO(Ci):

The substrate specificity of ELO(XI) can be determined after expression and the feeding of different fatty acids (Figure 22). The substrates fed can be detected in large amounts in all of the transgenic yeasts. The transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the ELO(XI) reaction. This means that the gene ELO(XI) has been expressed functionally.

It can be seen from Table 16 that ELO(XI) shows a broad substrate specificity. Both C18- and C20-fatty acids are elongated, but a preference for $\Delta 5$ - and $\Delta 6$ -desaturated fatty acids can be observed.

The yeasts which had been transformed with the vector pYES2-ELO(XI) were cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC.

Table 16: Expression of ELO(XI) in yeast. The conversion rate of different starting materials (amounts fed: in each case 250 μ M) is described.

25

Starting materials	Conversion of the starting materials by ELO(XI) in %
16:0	3
16:1 $\Delta 9$	0
18:0	2
18:1 $\Delta 9$	0
18:2 $\Delta 9,12$	3
18:3 $\Delta 6,9,12$	12
18:3 $\Delta 5,9,12$	13
18:3 $\Delta 9,12,15$	3
18:4 $\Delta 6,9,12,15$	20
20:3 $\Delta 8,11,14$	5

20:3 ^{Δ11,14,17}	13
20:4 ^{Δ5,8,11,14}	15
20:5 ^{Δ5,8,11,14,17}	10
22:4 ^{Δ7,10,13,16}	0
22:6 ^{Δ4,7,10,13,16,19}	0

The substrate specificity of ELO(Ci) can be determined after expression and the feeding of different fatty acids (Fig. 23). The substrates fed can be detected in large amounts in all of the transgenic yeasts. The transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the ELO(Ci) reaction. This means that the gene ELO(Ci) has been expressed functionally.

Table 17: Expression of ELO(Ci) in yeast. The conversion rate of different starting materials (amounts fed: in each case 250 μM) is described.

Starting materials	Conversion of the starting materials by ELO(Ci) in %
16:0	0
16:1 ^{Δ9}	0
18:0	0
18:1 ^{Δ9}	0
18:2 ^{Δ9,12}	23
18:3 ^{Δ6,9,12}	10
18:3 ^{Δ5,9,12}	38
18:3 ^{Δ9,12,15}	25
18:4 ^{Δ6,9,12,15}	3
20:3 ^{Δ8,11,14}	10
20:3 ^{Δ11,14,17}	8
20:4 ^{Δ5,8,11,14}	10
20:5 ^{Δ5,8,11,14,17}	15
22:4 ^{Δ7,10,13,16}	0
22:6 ^{Δ4,7,10,13,16,19}	0

It can be seen from Table 17 that ELO(Ci) shows a broad substrate specificity. Both C18- and C20-fatty acids are elongated, but a preference for Δ5- and Δ6-desaturated fatty acids can be observed.

The yeasts which had been transformed with the vector pYES2-ELO(Ci) were cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC.

Example 43: Cloning genes from *Ostreococcus tauri*

The search for conserved regions in the protein sequences with the aid of the elongase genes with $\Delta 5$ -elongase activity or $\Delta 6$ -elongase activity, which have been described herein, allowed the identification of in each case two sequences with corresponding motifs in an *Ostreococcus tauri* sequence database (genomic sequences). The sequences were the following:

Name of gene	SEQ ID	Amino acids
OtELO1, ($\Delta 5$ -elongase)	SEQ ID NO: 67	300
OtELO1.2, ($\Delta 5$ -elongase)	SEQ ID NO: 113	300
OtELO2, ($\Delta 6$ -elongase)	SEQ ID NO: 69	292
OtELO2.1, ($\Delta 6$ -elongase)	SEQ ID NO: 111	292

OtElo1 and OtElo1.2 show the highest similarity with an elongase from *Danio rerio* (GenBank AAN77156; approximately 26% identity), while OtElo2 and OtElo2.1 show the highest similarity with *Physcomitrella Elo* (PSE) [approx. 36% identity] (alignments were carried out using the tBLASTn algorithm (Altschul et al., J. Mol. Biol. 1990, 215: 403-410)).

The elongases were cloned as follows:

40 ml of an *Ostreococcus tauri* culture in the stationary phase were spun down, resuspended in 100 μ l of double-distilled water and stored at -20°C . The respective genomic DNAs were amplified on the basis of the PCR method. The relevant primer pairs were selected in such a way that they bore the yeast consensus sequence for highly efficient translation (Kozak, Cell 1986, 44:283-292) next to the start codon. The amplification of the OtElo DNAs was carried out in each case using 1 μ l of defrosted cells, 200 μ M of dNTPs, 2.5 U *Taq* polymerase and 100 pmol of each primer in a total volume of 50 μ l. The PCR conditions were as follows: first denaturation for 5 minutes at 95°C , followed by 30 cycles of 30 seconds at 94°C , 1 minute at 55°C and 2 minutes at

72°C, and a last elongation step of 10 minutes at 72°C.

Example 44: Cloning expression plasmids for the heterologous expression in yeasts:

- 5 To characterize the function of the elongases from *Ostreococcus tauri*, the open reading frames of the respective DNAs were cloned downstream of the galactose-inducible GAL1 promoter of pYES2.1/V5-His-TOPO (Invitrogen), giving rise to pOTE1, pOTE1.2, pOTE2 and pOTE2.1.
- 10 The *Saccharomyces cerevisiae* strain 334 was transformed by electroporation (1500 V) with the vector pOTE1, pOTE1.2, pOTE2 and pOTE2.1, respectively. A yeast which was transformed with the blank vector pYES2 was used as the control. The transformed yeasts were selected on complete minimal dropout uracil medium (CMdum) agar plates supplemented with 2% glucose. After the selection, in each case
- 15 three transformants were selected for the further functional expression.

To express the Ot elongases, precultures of in each case 5 ml of liquid CMdum medium supplemented with 2% (w/v) raffinose, but without uracil, were inoculated with the selected transformants and incubated for 2 days at 30°C, 200 rpm. 5 ml of liquid

20 CMdum medium (without uracil) supplemented with 2% raffinose and 300 µm of various fatty acids were then inoculated with the precultures to an OD₆₀₀ of 0.05. The expression was induced by addition of 2% (w/v) galactose. The cultures were incubated for a further 96 hours at 20°C.

- 25 Example 45: Cloning of expression plasmids for the seed-specific expression in plants

A further transformation vector based on pSUN-USP was generated for the transformation of plants. To this end, NotI cleavage sites were introduced at the 5' and

30 3' ends of the coding sequences, using PCR. The corresponding primer sequences were derived from the 5' and 3' regions of OtElo1, OtElo1.2, OtElo2 and OtElo2.1.

Composition of the PCR mix (50 µl):

- 35 5.00 µl template cDNA
5.00 µl 10x buffer (Advantage polymerase)+ 25mM MgCl₂
5.00 µl 2mM dNTP
1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

PCR reaction conditions:

5 Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

- 10 The PCR products are incubated with the restriction enzyme NotI for 16 hours at 37°C. The plant expression vector pSUN300-USP is incubated in the same manner. Thereafter, the PCR products and the vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's
- 15 instructions. Thereafter, vector and PCR products were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmids pSUN-OtELO1, pSUN-OtELO1.2, pSUN-OtELO2 and pSUN-OtELO2.2 were verified by sequencing.

- pSUN300 is a derivative of plasmid pPZP (Hajdukiewicz, P., Svab, Z., Maliga, P., (1994)
- 20 The small versatile pPZP family of *Agrobacterium* binary vectors for plant transformation. *Plant Mol Biol* 25:989-994). pSUN-USP originated from pSUN300, by inserting a USP promoter into pSUN300 in the form of an EcoRI fragment. The polyadenylation signal is that of the *Ostreococcus* gene from the *A. tumefaciens* Ti plasmid (ocs-Terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P.,
- 25 Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine synthase gene *J. Mol. Appl. Genet.* 1 (6), 499-511 (1982)). The USP promoter corresponds to nucleotides 1 to 684 (Genbank Accession X56240), where part of the noncoding region of the USP gene is present in the promoter. The promoter fragment
- 30 which is 684 base pairs in size was amplified by a PCR reaction and standard methods with the aid of a synthesized primer and by means of a commercially available T7 standard primer (Stratagene).

Primer sequence:

35

5'-GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGGGGGATCC
GGATCTGCTGGCTATGAA-3', SEQ ID NO: 130.

The PCR fragment was recut with EcoRI/Sall and inserted into the vector pSUN300 with OCS terminator. This gave rise to the plasmid with the name pSUN-USP. The construct was used for the transformation of *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

5

Example 46: Expression of OtElo1, OtElo1.2, OtElo2 and OtELO2.2 in yeasts

Yeasts which had been transformed with the plasmids pYES3, pYES3-OtEIO1, pYES3-OtEIO1.2, pYES3- OtELO2 and pYES3-OtELO2.2 as described in Example 15 were
10 analyzed as follows:

The yeast cells from the main cultures were harvested by centrifugation (100 x g, 5 min, 20°C) and washed with 100 mM NaHCO₃, pH 8.0 to remove residual medium and fatty acids. Starting with the yeast cell sediments, fatty acid methyl esters (FAMES) were prepared by acid methanolysis. To this end, the cell sediments were incubated for
15 1 hour at 80°C together with 2 ml of 1 N methanolic sulfuric acid and 2% (v/v) of dimethoxypropane. The FAMES were extracted twice with petroleum ether (PE). To remove nonderivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO₃, pH 8.0 and 2 ml of distilled water. Thereafter, the PE
20 phases were dried with Na₂SO₄, evaporated under argon and taken up in 100 µl of PE. The samples were separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 µm, Agilent) in a Hewlett-Packard 6850 gas chromatograph equipped with flame ionization detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of 5°C/min and finally 10 min
25 at 250°C (holding).

The signals were identified by comparing the retention times with corresponding fatty acid standards (Sigma). The methodology is described for example in Napier and Michaelson, 2001, *Lipids*. 36(8):761-766; Sayanova et al., 2001, *Journal of*
30 *Experimental Botany*. 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

Example 47: Functional characterization of OtElo1, OtElo1.2, OtElo2 and OtELO2.1:

35 The substrate specificity of OtElo1 was determined after expression and feeding of different fatty acids (Table 18). The substrates which have been fed can be detected in large amounts in all transgenic yeasts. The transgenic yeasts showed the synthesis of novel fatty acids, the products of the OtElo1 reaction. This means that the gene OtElo1

was expressed functionally.

It can be seen from Table 18 that OtElo1 and OtElo1.2 have a narrow substrate specificity. OtElo1 and OtElo1.2 were only capable of elongating the C20-fatty acids
 5 eicosapentaenoic acid (Figure 24A, 24B) and arachidonic acid (Figure 25A, 25B), but preference was given to the ω 3-desaturated eicosapentaenoic acid.

Table 18 shows the substrate specificity of the elongase OtElo1 and OtElo1.2 for C20-
 10 poly unsaturated fatty acids with a double bond in the Δ 5-position in comparison with different fatty acids.

The yeasts which had been transformed with the vector pOTE1 or pOTE1.2 were cultured in minimal medium in the presence of the fatty acids stated. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis.
 15 Thereafter, the FAMES were analyzed via GLC.

The substrate specificity of OtElo2 (SEQ ID NO: 81) OtElo2.1 (SEQ ID NO: 111) can be determined after expression and the feeding of different fatty acids (Table 19). The substrates fed can be detected in large amounts in all of the transgenic yeasts. The
 20 transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the OtElo2 reaction. This means that the genes OtElo2 and OtElo2.1 have been expressed functionally.

Table 18:

25

Fatty acid substrate	Conversion rate of OtElo1 (in %)	Conversion rate of OtElo1.2 (in %)
16:0	-	-
16:1 ^{Δ9}	-	-
18:0	-	-
18:1 ^{Δ9}	-	-
18:1 ^{Δ11}	-	-
18:2 ^{Δ9,12}	-	-
18:3 ^{Δ6,9,12}	-	-
18:3 ^{Δ5,9,12}	-	-
20:3 ^{Δ8,11,14}	-	-
20:4 ^{Δ5,8,11,14}	10.8 $\pm$ 0.6	38.0

20:5 ^{Δ5,8,11,14,17}	46.8 ± 3.6	68.6
22:4 ^{Δ7,10,13,16}	-	-
22:6 ^{Δ4,7,10,13,16,19}	-	-

Table 19 shows the substrate specificity of the elongase OtElo2 and OtElo2.1 with regard to various fatty acids. OtElo2.1 shows a markedly higher activity.

- 5 The yeasts which had been transformed with the vector pOTE2 or pOTE2.1 were cultured in minimal medium in the presence of the fatty acids stated. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC.
- 10 The enzymatic activity shown in Table 19 clearly demonstrates that OtElo2 and OtElo2.1, respectively, are a $\Delta 6$ -elongase.

Table 19:

Fatty acid substrate	Conversion rate of OtElo2 (in %)	Conversion rate of OtElo2.2 (in %)
16:0	-	-
16:1 ^{Δ9}	-	-
16:3 ^{Δ7,10,13}	-	-
18:0	-	-
18:1 ^{Δ6}	-	-
18:1 ^{Δ9}	-	-
18:1 ^{Δ11}	-	-
18:2 ^{Δ9,12}	-	-
18:3 ^{Δ6,9,12}	15.3	55.7
18:3 ^{Δ5,9,12}	-	-
18:4 ^{Δ6,9,12,15}	21.1	70.4
20:2 ^{Δ11,14}	-	-
20:3 ^{Δ8,11,14}	-	-
20:4 ^{Δ5,8,11,14}	-	-
20:5 ^{Δ5,8,11,14,17}	-	-
22:4 ^{Δ7,10,13,16}	-	-
22:5 ^{Δ7,10,13,16,19}	-	-
22:6 ^{Δ4,7,10,13,16,19}	-	-

Figure 24 A – D shows the elongation of eicosapentaenoic acid by OtElo1 (B) and OtElo1.2 (D), respectively. The controls (A, C) do not show the elongation product (22:5 ω 3).

- 5 Figure 25 A – D shows the elongation of arachidonic acid by OtElo1 (B) and OtElo1.2 (D), respectively. The controls (A, C) do not show the elongation product (22:4 ω 6).

Example 48: Cloning elongase genes from *Euglena gracilis* and *Arabidopsis thaliana*

- 10 The search for conserved regions in the protein sequences with the aid of the elongase genes with Δ 5-elongase activity or Δ 6-elongase activity, which are detailed in the application, allowed the identification of sequences from *Arabidopsis thaliana* and *Euglena gracilis*, respectively, with corresponding motifs in sequence databases (Genbank, *Euglena* EST Bank). The sequences were the following:

15

Name of gene	SEQ ID	Amino acids
EGY1019 (<i>E. gracilis</i>)	SEQ ID NO: 131	262
EGY2019 (<i>E. gracilis</i>)	SEQ ID NO: 133	262
At3g06460 (<i>A. thaliana</i>)	SEQ ID NO: 135	298
At3g06470 (<i>A. thaliana</i>)	SEQ ID NO: 137	278

The *Euglena gracilis* elongases were cloned as follows:

- 20 The *Euglena gracilis* strain 1224-5/25 was obtained from the Sammlung für Algenkulturen Göttingen [Göttingen collection of algal cultures] (SAG). For the isolation, the strain was grown for 4 days at 23°C in medium II (Calvayrac R and Douce R, FEBS Letters 7:259-262, 1970) with a photoperiod of 8 h/16 h (light intensity 35 mol s⁻¹ m⁻²).

- 25 Total RNA of a four-day-old *Euglena* culture was isolated with the aid of the RNeasy Kit from Qiagen (Valencia, CA, US). poly-A⁺ RNA (mRNA) was isolated from the total RNA with the aid of oligo-dT-cellulose (Sambrook et al., 1989). The RNA was subjected to reverse transcription with the Reverse Transcription System Kit from Promega, and the cDNA synthesized was cloned into the lambda ZAP vector (lambda ZAP Gold, Stratagene). The cDNA was depackaged in accordance with the
- 30 manufacturer's instructions to give the plasmid DNA, and clones were partially sequenced for random sequencing. mRNA was isolated from the total RNA with the aid of the PolyAtract isolation system (Promega). The mRNA was subjected to reverse

transcription with the Marathon cDNA Amplification Kit (BD Biosciences) and the adaptors were ligated in accordance with the manufacturer's instructions. The cDNA library was then used for the PCR for cloning expression plasmids by means of 5'- and 3'-RACE (rapid amplification of cDNA ends).

5

The *Arabidopsis thaliana* elongases were cloned as follows:

Starting from the genomic DNA, primers for the two genes were derived at the 5' and the 3' end of the open reading frame.

10

The method of Chirgwin *et al.*, (1979) was used for isolating total RNA from *A. thaliana*. Leaves from 21-day-old plants were crushed in liquid nitrogen, treated with disruption buffer and incubated for 15 minutes at 37°C. After centrifugation (10 min, 4°C, 12 000 x g), the RNA in the supernatant was precipitated at -20°C for 5 hours using 0.02 volume of 3 M sodium acetate pH 5.0 and 0.75 volume ethanol. After a further centrifugation step, the RNA was taken up in 1 ml of TES per g of starting material, extracted once with one volume of phenol/chloroform and once with one volume of chloroform, and the RNA was precipitated with 2.5 M LiCl. Following subsequent centrifugation and washing with 80% ethanol, the RNA was resuspended in water. The cDNA was synthesized in accordance with the method of Sambrook *et al.* 1989, and an RT-PCR was carried out using the derived primers. The PCR products were cloned into the vector pYES2.1-TOPO (Invitrogen) in accordance with the manufacturer's instructions.

15

20

25 Example 49: Cloning expression plasmids for heterologous expression in yeasts:

To characterize the function of the *A. thaliana* elongases, the open reading frames of the DNAs in question were cloned downstream of the galactose-inducible GAL1 promoter of pYES2.1/V5-His-TOPO (Invitrogen), giving rise to pAt60 and pAt70.

30

The *Saccharomyces cerevisiae* strain 334 was transformed by electroporation (1500 V) with the vector pAt60 and pAt70, respectively. A yeast which was transformed with the blank vector pYES2.1 was used as the control. The transformed yeasts were selected on complete minimal dropout uracil medium (CMdum) agar plates supplemented with 2% glucose. After the selection, in each case three transformants were selected for the further functional expression.

35

To express the At elongases, precultures of in each case 5 ml of dropout uracil CMdum

liquid medium supplemented with 2% (w/v) raffinose were inoculated with the selected transformants and incubated for 2 days at 30°C, 200 rpm.

- 5 5 ml of liquid CMdum medium (without uracil) supplemented with 2% raffinose and 300 μ M of various fatty acids were then inoculated with the precultures to an OD₆₀₀ of 0.05. The expression was induced by addition of 2% (w/v) galactose. The cultures were incubated for a further 96 hours at 20°C.

Example 50: Expression of pAt60 and pAt70 in yeasts

10

Yeasts which had been transformed with the plasmids pYES2.1, pAt60 and pAt70 as described in Example 5 were analyzed as follows:

- 15 The yeast cells from the main cultures were harvested by centrifugation (100 x g, 5 min, 20°C) and washed with 100 mM NaHCO₃, pH 8.0 to remove residual medium and fatty acids. Starting with the yeast cell sediments, fatty acid methyl esters (FAMES) were prepared by acid methanolysis. To this end, the cell sediments were incubated for 1 hour at 80°C together with 2 ml of 1 N methanolic sulfuric acid and 2% (v/v) of dimethoxypropane. The FAMES were extracted twice with petroleum ether (PE). To
20 remove nonderivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO₃, pH 8.0 and 2 ml of distilled water. Thereafter, the PE phases were dried with Na₂SO₄, evaporated under argon and taken up in 100 μ l of PE. The samples were separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 μ m, Agilent) in a Hewlett-Packard 6850 gas chromatograph equipped with flame ionization
25 detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of 5°C/min and finally 10 min at 250°C (holding).

- 30 The signals were identified by comparing the retention times with corresponding fatty acid standards (Sigma). The methodology is described for example in Napier and Michaelson, 2001, *Lipids*. 36(8):761-766; Sayanova et al., 2001, *Journal of Experimental Botany*. 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

- 35 Example 51: Functional characterization of pAt60 and pAt70

The substrate specificity of the elongases At3g06460 and At3g06470 was determined after expression and feeding of various fatty acids (Table 20, Fig. 26). The substrates

which have been fed can be detected in all transgenic yeasts. The transgenic yeasts showed the synthesis of novel fatty acids, the products of the genes At3g06460 and At3g06470, respectively. This means that these genes have been expressed functionally.

5

Table 20: Elongation of EPA by the elongases At3g06460 and At3g06470, respectively. Measurement of the yeast extracts after feeding of 250 μ M EPA

Gene	Fatty acid fed	C20:5n-3 content	C22:5n-3 content
At3g06460	EPA (C20:5n-3)	20.8	0.6
At3g06460	EPA (C20:5n-3)	25.4	1.1
Conversion rate of EPA		At3g06460: 3.0%	At3g06470: 4.1%

10

Figure 26 represents the elongation of 20:5n-3 by the elongases At3g06470.

Example 52: Cloning an elongase from *Phaeodactylum tricornutum*

15 Starting from conserved regions in the protein sequences, degenerate primers were constructed with the aid of the elongase genes with $\Delta 6$ -elongase activity detailed in the application, and these primers were used for searching a *Phaeodactylum* cDNA library by means of PCR. The following primer sequences were employed:

Name of primer	Sequence 5'-3' orientation	Corresponding amino acids
Phaelo forward1	AA(C/T)CTUCTUTGGCTUTT(C/T)TA (SEQ ID NO: 185)	NLLWLFY
Phaelo reverse1	GA(C/T)TGUAC(A/G)AA(A/G)AA(C/T)TGUG C(A/G)AA (SEQ ID NO: 186)	FAQFFVQS

20

Nucleotide bases in brackets mean that a mixture of oligonucleotides with in each case one or the other nucleotide base are present.

Construction of the *Phaeodactylum* cDNA library:

25

A 2 l culture of *P. tricornutum* UTEX 646 was grown in f/2 medium (Guillard, R.R.L. 1975. Culture of phytoplankton for feeding marine invertebrates. In *Culture of Marine*

Invertebrate Animals (Eds. Smith, W.L. and Chanley, M.H.), Plenum Press, New York, pp 29-60) for 14 d (= days) at a light intensity of 35 E/cm². After centrifugation, frozen cells were ground to a fine powder in the presence of liquid nitrogen and resuspended in 2 ml of homogenization buffer (0.33 M sorbitol, 0.3 M NaCl, 10 mM EDTA, 10 mM EGTA, 2% SDS, 2% mercaptoethanol in 0.2 M Tris-Cl pH 8.5). After 4 ml of phenol and 2 ml of chloroform had been added, the mixture was shaken vigorously for 15 minutes at 40-50°C. Thereafter, the mixture was centrifuged (10 min × 10 000 g) and the aqueous phase was extracted stepwise with chloroform. Nucleic acids were then precipitated by addition of 1/20 volume 4 M sodium hydrogencarbonate solution and centrifuged. The pellet was taken up in 80 mM Tris-borate pH 7.0 and 1 mM EDTA, and the RNA was precipitated with 8 M lithium chloride. After centrifugation and washing with 70% strength ethanol, the RNA pellet was taken up in RNase-free water. Poly(A)-RNA was isolated using Dynabeads (Dynal, Oslo, Norway) following the manufacturer's instructions, and the first-strand cDNA synthesis was carried out using MLV-Rtase from Roche (Mannheim). Then, the second-strand synthesis was carried out using DNA polymerase I and Klenow fragment, followed by a digestion with RNaseH. The cDNA was then treated with T4 DNA polymerase, and EcoRI/XhoI adaptors (Pharmacia, Freiburg) were subsequently attached by means of T4 ligase. After digestion with XhoI, phosphorylation and gel separation, fragments greater than 300 bp were ligated into the phage lambda ZAP Express following the manufacturer's instructions (Stratagene, Amsterdam, the Netherlands). Following bulk excision of the cDNA library and plasmid recovery, the plasmid library was transformed into *E. coli* DH10B cells and employed for the PCR screening.

Using the abovementioned degenerate primers, it was possible to generate the PCR fragment with the sequence number SEQ ID NO: 187.

This fragment was labeled with digoxigenin (Roche, Mannheim) and used as probe for screening the phage library.

With the aid of the sequence SEQ ID NO: 187, it was possible to obtain the gene sequence SEQ ID NO: 183, which constitutes the full-RNA molecule of the *Phaeodactylum* Δ6-elongase:

Example 53: Cloning expression plasmids for the heterologous expression in yeasts

The relevant primer pairs were selected in such a way that they bore the yeast consensus sequence for highly efficient translation (Kozak, Cell 1986, 44:283-292) next

to the start codon. The amplification of the PtELO6 DNAs was carried out in each case using 1 μ l of cDNA, 200 μ M of dNTPs, 2.5 U Advantage polymerase and 100 pmol of each primer in a total volume of 50 μ l. The PCR conditions were as follows: first denaturation for 5 minutes at 95°C, followed by 30 cycles of 30 seconds at 94°C, 1 minute at 55°C and 2 minutes at 72°C, and a last elongation step of 10 minutes at 72°C.

To clone the sequence for the heterologous expression in yeasts, the following oligonucleotides were used for the PCR reaction:

10

Name of gene and SEQ ID NO:	Primer sequence
PtELO6 (SEQ ID NO: 183)	F:5'-GCGGCCGCACATAATGATGGTACCTTCAAG (SEQ ID NO: 188) R:3'-GAAGACAGCTTAATAGACTAGT (SEQ ID NO: 189)

*F=forward primer, R=reverse primer

The PCR products were incubated for 30 minutes at 21°C with the yeast expression vector pYES2.1-TOPO (Invitrogen) following the manufacturer's instructions. The PCR product (see SEQ ID NO: 192) was ligated into the vector by means of a T overhang and activity of a topoisomerase (Invitrogen). After incubation, *E. coli* DH5 α cells were transformed. Suitable clones were identified by PCR, the plasmid DNA was isolated by means of Qiagen DNAeasy Kit and verified by sequencing. The correct sequence was then transformed into the *Saccharomyces* strain INVSc1 (Invitrogen) by electroporation (1500 V). As a control, the blank vector pYES2.1 was transformed in parallel. The yeasts were subsequently plated onto complete uracil dropout minimal medium supplemented with 2% glucose. Cells which were capable of growing in the medium without uracil thus comprise the corresponding plasmids pYES2.1 and pYES2.1-PtELO6. After the selection, in each case two transformants were selected for further functional expression.

Example 54: Cloning expression plasmids for the seed-specific expression in plants

A further transformation vector based on pSUN-USP is generated for the transformation of plants. To this end, NotI cleavage sites are introduced at the 5' and 3' ends of the coding sequence, using the following primer pair:

PSUN-PtELO6

Forward: 5'-GCGGCCGCACCATGATGGTACCTTCAAGTTA (SEQ ID NO: 190)

5 Reverse: 3'-GAAGACAGCTTAATAGGCGGCCGC (SEQ ID NO: 191)

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

10 5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase

The Advantage polymerase from Clontech was employed.

15 PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

20 Number of cycles: 35

The PCR products are incubated with the restriction enzyme NotI for 16 hours at 37°C.

The plant expression vector pSUN300-USP is incubated in the same manner.

Thereafter, the PCR products and the 7624 bp vector are separated by agarose gel

25 electrophoresis and the corresponding DNA fragments are excised. The DNA is purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR products are ligated. The Rapid Ligation Kit from Roche is used for this purpose. The resulting plasmids pSUN-PtELO is verified by sequencing.

30

pSUN300 is a derivative of plasmid pPZP (Hajdukiewicz, P., Svab, Z., Maliga, P., (1994)

The small versatile pPZP family of *Agrobacterium* binary vectors for plant

transformation. *Plant Mol Biol* 25:989-994). pSUN-USP originated from pSUN300, by

inserting a USP promoter into pSUN300 in the form of an EcoRI fragment. The

35 polyadenylation signal is that of the Octopine synthase gene from the *A. tumefaciens* Ti plasmid (ocs-Terminator, Genbank Accession V00088) (De Greve, H., Dhaese, P., Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine

synthase gene J. Mol. Appl. Genet. 1 (6), 499-511 (1982)). The USP promoter corresponds to nucleotides 1 to 684 (Genbank Accession X56240), where part of the noncoding region of the USP gene is present in the promoter. The promoter fragment which is 684 base pairs in size was amplified by a PCR reaction and standard methods with the aid of a synthesized primer and by means of a commercially available T7 standard primer (Stratagene).

(Primer sequence: 5'-
GTCGACCCGCGGACTAGTGGGCCCTCTAGACCCGGGGGATCC
GGATCTGCTGGCTATGAA-3'; SEQ ID NO: 151).

The PCR fragment was recut with EcoRI/Sall and inserted into the vector pSUN300 with OCS terminator. This gave rise to the plasmid with the name pSUN-USP. The construct was used for the transformation of *Arabidopsis thaliana*, oilseed rape, tobacco and linseed.

Lipids were extracted from yeasts and seeds as described for Example 6.

Example 55: Expression of PtElo in yeasts

Yeasts which had been transformed with the plasmids pYES2 and pYES2-PtELO6 as in Example 4 were analyzed as follows:

The yeast cells from the main cultures were harvested by centrifugation ($100 \times g$, 5 min, 20°C) and washed with 100 mM NaHCO_3 , pH 8.0 in order to remove residual medium and fatty acids. Fatty acid methyl esters (FAMES) were prepared from the yeast cell sediments by acid methanolysis. To this end, the cell sediments were incubated for 1 hour at 80°C with 2 ml of 1N methanolic sulfuric acid and 2% (v/v) dimethoxypropane. The FAMES were extracted by twice extracting with petroleum ether (PE). To remove nonderivatized fatty acids, the organic phases were washed in each case once with 2 ml of 100 mM NaHCO_3 , pH 8.0, and 2 ml of distilled water. Thereafter, the PE phases were dried with Na_2SO_4 , evaporated under argon and taken up in 100 μl of PE. The samples were separated on a DB-23 capillary column (30 m, 0.25 mm, 0.25 μm , Agilent) in a Hewlett-Packard 6850 gas chromatograph with flame ionization detector. The conditions for the GLC analysis were as follows: the oven temperature was programmed from 50°C to 250°C with an increment of $5^{\circ}\text{C}/\text{min}$ and finally 10 minutes at 250°C (holding).

The signals were identified by comparing the retention times with corresponding fatty acid standards (Sigma). The methodology is described for example in Napier and Michaelson, 2001, *Lipids*. 36(8):761-766; Sayanova et al., 2001, *Journal of Experimental Botany*. 52(360):1581-1585, Sperling et al., 2001, *Arch. Biochem. Biophys.* 388(2):293-298 and Michaelson et al., 1998, *FEBS Letters*. 439(3):215-218.

Example 56: Functional characterization of PtELO6:

Figure 29 represents the conversion of C18:3^{Δ6,9,12} and C18:4^{Δ6,9,12,15}. The substrates are elongated by in each case two carbon atoms; this results in the fatty acids C20:3^{Δ8,11,14} and C20:4^{Δ8,11,14,17}, respectively. The substrate specificity of PtELO6 can be determined after expression and the feeding of different fatty acids (Figure 30). The substrates fed can be detected in large amounts in all of the transgenic yeasts. The transgenic yeasts demonstrated the synthesis of novel fatty acids, the products of the PtELO6 reaction. This means that the gene PtELO6 has been expressed functionally.

It can be seen from Table 21 that PtELO6 shows a narrow substrate specificity. PtELO6 was only capable of elongating the C18-fatty acids linoleic acid, linolenic acid, γ-linolenic acid and stearidonic acid, but preferred the ω3-desaturated stearidonic acid (see also Figure 30).

Feeding experiment: fatty acids (in bold) were added in each case in amounts of 250 μM. The underlined fatty acids were formed *de novo*.

Table 21: Substrate specificity of PtELO6

Fatty acid fed:		+18:2	+18:3	+18:3	+18:4
16:0	16.2	18.2	15.2	20	04:48
16:1	50.6	20.5	22.8	33.5	34.2
18:0	5.4	6.3	6.2	5.2	12.4
18:1	27.7	14.6	19.6	19.3	16.7
18:2		40			
18:3			32.9		
18:3				12.3	
18:4					4.5
20:2		<u>0.4</u>			
20:3			<u>3.4</u>		

20:3				<u>9.7</u>	
20:4					<u>14.5</u>
% elongation	0.0	0.99	9.37	44.09	76.32

The following fatty acids were fed, but not converted:

- 18:1^{Δ6}, 18:1^{Δ9}, 18:1^{Δ11}
- 5 • 20:2^{Δ11,14}, 20:3^{Δ11,14,17}, 20:3^{Δ8,11,14}, 20:4^{Δ5,8,11,14}, 20:5^{Δ5,8,11,14,17}
- 22:4^{Δ7,10,13,16}

The yeasts which had been transformed with the vector pYES2-PtELO6 were cultured in minimal medium in the presence of the fatty acids detailed. The fatty acid methyl esters were synthesized by subjecting intact cells to acid methanolysis. Thereafter, the FAMES were analyzed via GLC. The results shown in Figures 29 and 30 and in Table 19 were thus determined.

Example 57: Cloning expression plasmids for the seed-specific expression in plants

The general conditions described hereinbelow apply to all of the subsequent experiments, unless otherwise specified.

The following are preferably used in accordance with the invention for the examples which follow: Bin19, pBI101, pBinAR, pGPTV and pCAMBIA. An overview of binary vectors and their use is found in Hellens et al., Trends in Plant Science (2000) 5, 446-451. A pGPTV derivative as described in DE10205607 was used. This vector differs from pGPTV by an additionally inserted Ascl restriction cleavage site.

Starting point of the cloning procedure was the cloning vector pUC19 (Maniatis et al.). In the first step, the Conlinin promoter fragment was amplified using the following primers:

Cnl1 C 5': gaattcggcgcgccgagctcctcgagcaacggttccggcggtatagagttgggtaattcga
Cnl1 C 3': cccgggatcgatgccggcagatctccaccatttttggtggtgat

Composition of the PCR mix (50 µl):

5.00 µl template cDNA
5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂
5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)
0.50 µl Advantage polymerase (Clontech)

PCR reaction conditions:

5

Annealing temperature: 1 min 55°C
Denaturation temperature: 1 min 94°C
Elongation temperature: 2 min 72°C
Number of cycles: 35

10

The PCR product was first incubated with the restriction enzyme *EcoRI* for 2 hours at 37°C and then for 12 hours at 25°C with the restriction enzyme *SmaI*. The cloning vector pUC19 was incubated in the same manner. Thereafter, the PCR product and the cut, 2668 bp vector were separated by agarose gel electrophoresis and the
15 corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1-C was verified by sequencing.

20

In the next step, the OCS terminator (Genbank Accession V00088; De Greve, H., Dhaese, P., Seurinck, J., Lemmers, M., Van Montagu, M. and Schell, J. Nucleotide sequence and transcript map of the *Agrobacterium tumefaciens* Ti plasmid-encoded octopine synthase gene J. Mol. Appl. Genet. 1 (6), 499-511 (1982)) from the vector pGPVT-USP/OCS

25

(DE 102 05 607) was amplified using the following primers:

OCS_C 5': aggcctccatggcctgcttaatgagatatgagagacgcc
OCS_C 3': cccgggccggacaatcagtaaattgaacggag

30

Composition of the PCR mix (50 µl):

5.00 µl template cDNA
5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂
5.00 µl 2mM dNTP
1.25 µl of each primer (10 pmol/µl)
35 0.50 µl Advantage polymerase (Clontech)

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

5 Number of cycles: 35

The PCR product was first incubated with the restriction enzyme *Stu*I for 2 hours at 37°C and then for 12 hours at 25°C with the restriction enzyme *Sma*I. The vector pUC19-Cnl1-C was incubated for 12 hours at 25°C with the restriction enzyme *Sma*I.

10 Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1-C_OCS was
15 verified by sequencing.

In the next step, the Cnl1-B promoter was amplified by PCR using the following primers:

20 Cnl1-B 5': aggcctcaacggtccggcggtatag

Cnl1-B 3': cccgggggtaacgctagcgggcccgatatcggatcccatttttggtggtgattggtct

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

25 5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase (Clontech)

30 PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

35 Number of cycles: 35

The PCR product was first incubated with the restriction enzyme *Stu*I for 2 hours at 37°C and then for 12 hours at 25°C with the restriction enzyme *Sma*I. The vector

pUC19-Cnl1-C was incubated for 12 hours at 25°C with the restriction enzyme *Sma*I. Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1C_Cnl1B_OCS was verified by sequencing.

In a further step, the OCS terminator for Cnl1B was inserted. To this end, the PCR was carried out with the following primers:

OCS2 5': aggcctcctgctttaatgagatatgcgagac

OCS2 3': cccgggacgacaatcagtaaatgaacggag

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase (Clontech)

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

The PCR product was first incubated with the restriction enzyme *Stu*I for 2 hours at 37°C and then for 12 hours at 25°C with the restriction enzyme *Sma*I. The vector pUC19-Cnl1C_Cnl1B_OCS was incubated for 12 hours at 25°C with the restriction enzyme *Sma*I. Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1C_Cnl1B_OCS2 was verified by sequencing.

In the next step, the Cnl1-A promoter was amplified by PCR using the following primers:

Cnl1-B 5': aggcctcaacgggtccggcggtatagag

5 Cnl1-B 3': aggccttctagactgcaggcggccgcccgcatttttggtggtgattggt

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

10 5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase (Clontech)

PCR reaction conditions:

15

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

20

The PCR product was incubated for 2 hours at 37°C with the restriction enzyme *Stu*I.

The vector pUC19-Cnl1-C was incubated for 12 hours at 25°C with the restriction enzyme *Sma*I. Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The

25 DNA was purified by means of the Qiagen Gel Purification Kit in accordance with the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1C_Cnl1B_Cnl1A_OCS2 was verified by sequencing.

30 In a further step, the OCS terminator for Cnl1A was inserted. To this end, the PCR was carried out with the following primers:

OCS2 5': ggcctcctgcttaatgagatatgcga

OCS2 3': aagcttggcgcgccgagctcgtcgacggacaatcagtaaattgaacggaga

35

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂
5.00 µl 2mM dNTP
1.25 µl of each primer (10 pmol/µl)
0.50 µl Advantage polymerase (Clontech)

5

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

10 Elongation temperature: 2 min 72°C

Number of cycles: 35

The PCR product was first incubated with the restriction enzyme *Stu*I for 2 hours at 37°C and then for 2 hours at 37°C with the restriction enzyme *Hind*III. The vector
15 pUC19-Cnl1C_Cnl1B_Cnl1A_OCS2 was incubated for 2 hours at 37°C with the restriction enzyme *Stu*I and for 2 hours at 37°C with the restriction enzyme *Hind*III. Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's
20 instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1C_Cnl1B_Cnl1A_OCS3 was verified by sequencing.

In the next step, the plasmid pUC19-Cnl1C_Cnl1B_Cnl1A_OCS3 was used for cloning
25 the Δ6-, Δ5-desaturase and Δ6-elongase. To this end, the Δ6-desaturase from *Phytium irregulare* (WO02/26946) was amplified using the following PCR primers:

D6Des(Pir) 5': agatctatggtggacctaagcctggagtg

D6Des(Pir) 3': ccatggcccgggttacatcgctgggaactcggtgat

30

Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

5.00 µl 2mM dNTP

35 1.25 µl of each primer (10 pmol/µl)

0.50 µl Advantage polymerase (Clontech)

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

5 Number of cycles: 35

The PCR product was first incubated with the restriction enzyme *Bgl*II for 2 hours at 37°C and then for 2 hours at 37°C with the restriction enzyme *Nco*I. The vector pUC19-Cnl1C_Cnl1B_Cnl1A_OCS3 was incubated for 2 hours at 37°C with the restriction enzyme *Bgl*II and for 2 hours at 37°C with the restriction enzyme *Nco*I. Thereafter, the 10 PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this 15 purpose. The resulting plasmid pUC19-Cnl1_d6Des(Pir) was verified by sequencing.

In the next step, the plasmid pUC19-Cnl1_d6Des(Pir) was used for cloning the $\Delta 5$ -desaturase from *Thraustochytrium* ssp. (WO02/26946). To this end, the $\Delta 5$ -desaturase from *Thraustochytrium* ssp. was amplified using the following PCR primers:

20

D5Des(Tc) 5': gggatccatgggcaagggcagcgagggccg

D5Des(Tc) 3': ggcgccgacaccaagaagcaggactgagatatc

Composition of the PCR mix (50 μ l):

25

5.00 μ l template cDNA

5.00 μ l 10x buffer (Advantage polymerase) + 25mM MgCl₂

5.00 μ l 2mM dNTP

1.25 μ l of each primer (10 pmol/ μ l)

0.50 μ l Advantage polymerase (Clontech)

30

PCR reaction conditions:

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

35 Elongation temperature: 2 min 72°C

Number of cycles: 35

The PCR product was first incubated with the restriction enzyme *Bam*HI for 2 hours at 37°C and then for 2 hours at 37°C with the restriction enzyme *Eco*RV. The vector pUC19-Cnl1_d6Des(Pir) was incubated for 2 hours at 37°C with the restriction enzyme *Bam*HI and for 2 hours at 37°C with the restriction enzyme *Eco*RV. Thereafter, the
5 PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1_d6Des(Pir)_d5Des(Tc) was verified by
10 sequencing.

In the next step, the plasmid pUC19-Cnl1_d6Des(Pir)_d5Des(Tc) was used for cloning the $\Delta 6$ -elongase from *Physcomitrella patens* (WO01/59128), to which end an amplification with the following PCR primers was carried out:

15
D6Elo(Pp) 5': gcggccgcatggaggctcgtggagagattctacggtg
D6Elo(Pp) 3': gcaaaagggagctaaaactgagtgatctaga

Composition of the PCR mix (50 μ l):

20 5.00 μ l template cDNA
5.00 μ l 10x buffer (Advantage polymerase) + 25mM MgCl₂
5.00 μ l 2mM dNTP
1.25 μ l of each primer (10 pmol/ μ l)
0.50 μ l Advantage polymerase (Clontech)

25
PCR reaction conditions:

Annealing temperature: 1 min 55°C
Denaturation temperature: 1 min 94°C
30 Elongation temperature: 2 min 72°C
Number of cycles: 35

The PCR product was first incubated with the restriction enzyme *Not*I for 2 hours at 37°C and then for 2 hours at 37°C with the restriction enzyme *Xba*I. The vector pUC19-
35 Cnl1_d6Des(Pir)_d5Des(Tc) was incubated for 2 hours at 37°C with the restriction enzyme *Not*I and for 2 hours at 37°C with the restriction enzyme *Xba*I. Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was purified by means of the

Qiagen Gel Purification Kit following the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp) was verified by sequencing.

5

The binary vector for the plant transformation was generated starting from pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp). To this end, pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp) was incubated for 2 hours at 37°C with the restriction enzyme *Ascl*. The vector pGPTV was treated in the same manner.

- 10 Thereafter, the fragment from pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp) and the cut pGPTV vector were separated by agarose gel electrophoresis and the relevant DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit in accordance with the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pGPTV-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp) was verified by sequencing.

- 20 A further construct, pGPTV-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co), was used. To this end, an amplification was performed starting from pUC19-Cnl1C_OCS, using the following primers:

Cnl1_OCS 5': gtcgatcaacggttccggcggtagagttg

Cnl1_OCS 3': gtcgatcggacaatcagtaaattgaacggaga

- 25 Composition of the PCR mix (50 µl):

5.00 µl template cDNA

5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂

5.00 µl 2mM dNTP

1.25 µl of each primer (10 pmol/µl)

- 30 0.50 µl Advantage polymerase (Clontech)

PCR reaction conditions:

Annealing temperature: 1 min 55°C

- 35 Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

The PCR product was incubated for 2 hours at 37°C with the restriction enzyme *Sall*. The vector pUC19 was incubated for 2 hours at 37°C with the restriction enzyme *Sall*. Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA fragments were excised. The DNA was
5 purified by means of the Qiagen Gel Purification Kit in accordance with the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1_OCS was verified by sequencing.

- 10 In a further step, the $\Delta 12$ -desaturase gene from *Calendula officinalis* (WO01/85968) was cloned into pUC19-Cnl1_OCS. To this end, d12Des(Co) was amplified using the following primers:

D12Des(Co) 5': agatctatgggtgcaggcggtcgaatgc

- 15 D12Des(Co) 3': ccatggttaaattctattacgatacc

Composition of the PCR mix (50 μ l):

5.00 μ l template cDNA

5.00 μ l 10x buffer (Advantage polymerase) + 25mM MgCl₂

- 20 5.00 μ l 2mM dNTP

1.25 μ l of each primer (10 pmol/ μ l)

0.50 μ l Advantage polymerase (Clontech)

PCR reaction conditions:

- 25

Annealing temperature: 1 min 55°C

Denaturation temperature: 1 min 94°C

Elongation temperature: 2 min 72°C

Number of cycles: 35

- 30

The PCR product was incubated for 2 hours at 37°C with the restriction enzyme *Bgl*II and subsequently for 2 hours at the same temperature with *Nco*I. The vector pUC19-Cnl1_OCS was incubated in the same manner. Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the corresponding DNA
35 fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit in accordance with the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1_D12Des(Co) was verified by sequencing.

The plasmid pUC19-Cnl1_D12Des(Co) and the plasmid pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp) were incubated for 2 hours at 37°C with the restriction enzyme *Sall*. Thereafter, the vector fragment and the vector were separated
5 by agarose gel electrophoresis and the relevant DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit in accordance with the manufacturer's instructions. Thereafter, vector and vector fragment were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co) was verified by
10 sequencing.

The binary vector for the plant transformation was generated starting from pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co). To this end, pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co) was incubated for 2 hours at
15 37°C with the restriction enzyme *Ascl*. The vector pGPTV was treated in the same manner. Thereafter, the fragment from pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co) and the cut pGPTV vector were separated by agarose gel electrophoresis and the relevant DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit in
20 accordance with the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pGPTV-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co) was verified by sequencing.

25 A further vector which is suitable for the transformation of plants is pSUN2. To increase the number of expression cassettes present in the vector to more than four, this vector was used in combination with the Gateway System (Invitrogen, Karlsruhe). To this end, the Gateway cassette A was inserted into the vector pSUN2 in accordance with the manufacturer's instructions as described hereinbelow:

30 The pSUN2 vector (1 µg) was incubated for 1 hour with the restriction enzyme *EcoRV* at 37°C. Thereafter, the Gateway cassette A (Invitrogen, Karlsruhe) was ligated into the cut vector by means of the Rapid Ligation Kit from Roche, Mannheim. The resulting plasmid was transformed into *E. coli* DB3.1 cells (Invitrogen). The isolated plasmid
35 pSUN-GW was subsequently verified by sequencing.

In the second step, the expression cassette was excised from pUC19-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co) by means of *Ascl* and ligated

into the vector pSUN-GW, which had been treated in the same manner. The resulting plasmid pSUN-4G was used for further gene constructs.

To this end, a pENTR clone was first modified in accordance with the manufacturer's instructions (Invitrogen). The plasmid pENTR1A (Invitrogen) was incubated for 1 hour at 37°C with the restriction enzyme EcorI, subsequently treated for 30 minutes with Klenow enzyme and with one 1 µM dNTP mix, and the Ascl adaptor (5'-ggcgcgcc; phosphorylated at the 5' terminus, double-stranded) was then ligated into the vector pENTR1A. Into this modified, genes were stepwise inserted into the Cnl cassette as described above and transferred into the pENTR vector via Ascl.

The gene TL16y2 from *Thraustochytrium* ssp. (SEQ ID NO: 83) was transferred into the pSUN-4G vector in the abovedescribed manner:

In the next step, the plasmid pUC19-Cnl1C_Cnl1B_Cnl1A_OCS3 was used for cloning the Δ5-elongase TL16y2. To this end, the Δ5-elongase from *Thraustochytrium* ssp. was amplified using the following PCR primers:

TL16y2 5': agatct atggacgtcgtcgagcagca
TL16y2 3': ccatggcccggg agaagcagaagaccatctaa

Composition of the PCR mix (50 µl):

5.00 µl template cDNA
5.00 µl 10x buffer (Advantage polymerase) + 25mM MgCl₂
5.00 µl 2mM dNTP
1.25 µl of each primer (10 pmol/µl)
0.50 µl Advantage polymerase (Clontech)

PCR reaction conditions:

Annealing temperature: 1 min 55°C
Denaturation temperature: 1 min 94°C
Elongation temperature: 2 min 72°C
Number of cycles: 35

The PCR product was first incubated for 2 hours at 37°C with the restriction enzyme *Bgl*II and then for 2 hours at 37°C with the restriction enzyme *Nco*I. The vector pUC19-Cnl1C_Cnl1B_Cnl1A_OCS3 was incubated for 2 hours at 37°C with the restriction

enzyme *Bgl*II and for 2 hours at 37°C with the restriction enzyme *Nco*I. Thereafter, the PCR product and the cut vector were separated by agarose gel electrophoresis and the relevant DNA fragments were excised. The DNA was purified by means of the Qiagen Gel Purification Kit in accordance with the manufacturer's instructions. Thereafter, vector and PCR product were ligated. The Rapid Ligation Kit from Roche was used for this purpose. The resulting plasmid pUC19-Cnl1_TL16y2 was verified by sequencing. Thereafter, the cassette was excised using *Asc*I and ligated into an *Asc*I-pretreated pENTR vector. The resulting plasmid pENTR-Cnl1_TL16y2 was then incubated with the vector pSUN-4G in a recombination reaction in accordance with the manufacturer's instructions (Invitrogen). The product gave the vector pSUN-5G, which was used for the transformation of plants.

In a further step, the construct pSUN-8G was generated using the above-described methodology. To this end, 5' and 3' primers for the genes SEQ ID 41, 53, 87 and 113 with the above-described restriction cleavage sites and the first and in each case last 20 nucleotides of the open reading frame were generated, amplified under the standard conditions (see above) and ligated into the vector pENTR-Cnl.

A recombination reaction with the vector pSUN-4G gave rise to the construct pSUN-8G. This vector too was employed for the transformation of plants.

Example 58: Generation of transgenic plants

a) Generation of transgenic Indian mustard plants. The protocol for the transformation of oilseed rape plants was used (modification of the method of Moloney et al., 1992, Plant Cell Reports, 8:238-242)

To generate transgenic plants, the binary vectors pGPTV-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co), pSUN-5G and pSUN-8G which had been generated were transformed into *Agrobacterium tumefaciens* C58C1:pGV2260 (Deblaere et al., 1984, Nucl. Acids Res. 13, 4777-4788). To transform Indian mustard plants, a 1:50 dilution of an overnight culture of a positively transformed agrobacterial colony in Murashige-Skoog medium (Murashige and Skoog 1962 Physiol. Plant. 15, 473) supplemented with 3% sucrose (3MS medium) was used. Petioles or hypocotyls of freshly germinated sterile plants (in each case approx. 1 cm²) were incubated for 5-10 minutes with a 1:50 agrobacterial dilution in a Petri dish. This is followed by 3 days of coincubation in the dark at 25°C on 3MS medium supplemented with 0.8% Bacto agar. Cultivation was subsequently continued at 16

hours light/8 hours dark and in a weekly rhythm on MS medium supplemented with 500 mg/l of Claforan (cefotaxime-sodium), 50 mg/l kanamycin, 20 μ M benzylaminopurine (BAP) and 1.6 g/l glucose. Growing shoots were transferred to MS medium supplemented with 2% sucrose, 250 mg/l Claforan and 0.8% Bacto agar. If no roots
5 had formed after three weeks, 2-indolebutyric acid was added to the medium for rooting, to act as growth hormone.

Regenerated shoots were maintained on 2MS medium supplemented with kanamycin and Claforan, after rooting, transferred into soil and, after cultivation, grown for two
10 weeks in a controlled-environment cabinet or in a greenhouse, allowed to flower, mature seeds were harvested and studied for elongase expression such as $\Delta 6$ -elongase activity or $\Delta 5$ - or $\Delta 6$ -desaturase activity by means of lipid analyses. In this manner, lines with elevated contents of C20- and C22-polyunsaturated fatty acids were identified.

15 Transgenic oilseed rape plants were also generated successfully using this protocol.

b) Generation of transgenic linseed plants

20 The transgenic linseed plants can be generated for example by the method of Bell et al., 1999, *In Vitro Cell. Dev. Biol.-Plant.* 35(6): 456-465 by means of particle bombardment. Agrobacteria-mediated transformations can be carried out for example by the method of Mlynarova et al. (1994), *Plant Cell Report* 13: 282-285.

25 Example 59: Lipid extraction from seeds:

The effect of the genetic modification in plants on the production of a desired compound (such as a fatty acid) can be determined by growing the modified plant under suitable conditions (such as those described above) and analyzing the medium
30 and/or the cellular components for the elevated production of the desired product (i.e. of the lipids or a fatty acid). These analytical techniques are known to the skilled worker and comprise spectroscopy, thin-layer chromatography, various types of staining methods, enzymatic and microbiological methods and analytical chromatography such as high-performance liquid chromatography (see, for example, Ullman, *Encyclopedia of*
35 *Industrial Chemistry*, Vol. A2, pp 89-90 and pp 443-613, VCH: Weinheim (1985); Fallon, A., et al., (1987) "Applications of HPLC in Biochemistry" in: *Laboratory Techniques in Biochemistry and Molecular Biology*, Vol. 17; Rehm et al. (1993) *Biotechnology*, Vol. 3, Chapter III: "Product recovery and purification", pp 469-714,

- VCH: Weinheim; Belter, P.A., et al. (1988) *Bioseparations: downstream processing for Biotechnology*, John Wiley and Sons; Kennedy, J.F., and Cabral, J.M.S. (1992) *Recovery processes for biological Materials*, John Wiley and Sons; Shaeiwitz, J.A., and Henry, J.D. (1988) *Biochemical Separations*, in: *Ullmann's Encyclopedia of Industrial Chemistry*, Vol. B3; Chapter 11, pp 1-27, VCH: Weinheim; and Dechow, F.J. (1989) *Separation and purification techniques in biotechnology*, Noyes Publications).

- In addition to the abovementioned methods, plant lipids are extracted from plant material as described by Cahoon et al. (1999) *Proc. Natl. Acad. Sci. USA* 96 (22):12935-12940 and Browse et al. (1986) *Analytic Biochemistry* 152:141-145. The qualitative and quantitative analysis of lipids or fatty acids is described by Christie, William W., *Advances in Lipid Methodology*, Ayr/Scotland: Oily Press (Oily Press Lipid Library; 2); Christie, William W., *Gas Chromatography and Lipids. A Practical Guide* - Ayr, Scotland: Oily Press, 1989, Repr. 1992, IX, 307 pp (Oily Press Lipid Library; 1); "Progress in Lipid Research, Oxford: Pergamon Press, 1 (1952) - 16 (1977) under the title: Progress in the Chemistry of Fats and Other Lipids CODEN.

- In addition to measuring the end product of the fermentation, it is also possible to analyze other components of the metabolic pathways which are used for the production of the desired compound, such as intermediates and by-products, in order to determine the overall production efficiency of the compound. The analytical methods comprise measuring the amount of nutrients in the medium (for example sugars, hydrocarbons, nitrogen sources, phosphate and other ions), measuring the biomass composition and the growth, analyzing the production of conventional metabolites of biosynthetic pathways and measuring gases which are generated during the fermentation. Standard methods for these measurements are described in *Applied Microbial Physiology; A Practical Approach*, P.M. Rhodes and P.F. Stanbury, Ed., IRL Press, pp 103-129; 131-163 and 165-192 (ISBN: 0199635773) and references cited therein.

- One example is the analysis of fatty acids (abbreviations: FAME, fatty acid methyl ester; GC-MS, gas liquid chromatography/mass spectrometry; TAG, triacylglycerol; TLC, thin-layer chromatography).

- Unambiguous proof of the presence of fatty acid products can be obtained by analyzing recombinant organisms using standard analytical methods: GC, GC-MS or TLC, as described on several occasions by Christie and the references therein (1997, in: *Advances on Lipid Methodology*, Fourth Edition: Christie, Oily Press, Dundee, 119-169;

1998, Gaschromatographie-Massenspektrometrie-Verfahren [Gas chromatography/mass spectrometry methods], Lipide 33:343-353).

5 The material to be analyzed can be disrupted by sonication, grinding in a glass mill, liquid nitrogen and grinding or via other applicable methods. After disruption, the material must be centrifuged. The sediment is resuspended in distilled water, heated for 10 minutes at 100°C, cooled on ice and recentrifuged, followed by extraction for 1 hour at 90°C in 0.5 M sulfuric acid in methanol with 2% dimethoxypropane, which leads to hydrolyzed oil and lipid compounds, which give transmethylated lipids. These fatty acid methyl esters are extracted in petroleum ether and finally subjected to a GC analysis using a capillary column (Chrompack, WCOT Fused Silica, CP-Wax-52 CB, 25 µm, 0.32 mm) at a temperature gradient of between 170°C and 240°C for 20 minutes and 5 minutes at 240°C. The identity of the resulting fatty acid methyl esters must be defined using standards which are available from commercial sources (i.e. Sigma).

Plant material is initially homogenized mechanically by crushing in a pestle and mortar to make it more amenable to extraction.

20 This is followed by heating at 100°C for 10 minutes and, after cooling on ice, by resedimentation. The cell sediment is hydrolyzed for 1 hour at 90°C with 1 M methanolic sulfuric acid and 2% dimethoxypropane, and the lipids are transmethylated. The resulting fatty acid methyl esters (FAMES) are extracted in petroleum ether. The extracted FAMES are analyzed by gas liquid chromatography using a capillary column (Chrompack, WCOT Fused Silica, CP-Wax-52 CB, 25 m, 0.32 mm) and a temperature gradient of from 170°C to 240°C in 20 minutes and 5 minutes at 240°C. The identity of the fatty acid methyl esters is confirmed by comparison with corresponding FAME standards (Sigma). The identity and position of the double bond can be analyzed further by suitable chemical derivatization of the FAME mixtures, for example to give 4,4-dimethoxyoxazolin derivatives (Christie, 1998) by means of GC-MS.

Example 60: Analysis of the seeds from the transgenic plants which have been generated

35 Analogously to Example 59, the seeds of the plants which had been transformed with the constructs pGPTV-CnI1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co), pSUN-5G and pSUN-8G were analyzed. Figure XX shows the fatty acid spectrum of seeds with the construct pGPTV-CnI1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co). In

comparison with control plants which were not transformed (wild-type control, WT), a pronounced change in the fatty acid spectrum was observed. It was thus possible to demonstrate that the transformed genes are functional. Table 22 compiles the results of Figure 32.

5

Table 22:

Lines	Fatty acids								
	16:0	18:0	18:1	18:2	GLA	18:3	SDA	ARA	EPA
WT control	5.6	6.5	31.7	41.7	nd	12.1	nd	nd	nd
1424_Ko82_4	6.6	1.5	8.9	10.5	42.2	3.1	2.8	17.2	0.2
1424_Ko82_5	6.1	1.5	11.0	9.0	40.6	2.9	4.0	15.0	1.5
1424_Ko82_6	5.7	1.6	15.5	10.6	37.1	3.0	3.2	14.6	0.2
1424_Ko82_7	5.4	2.0	20.4	10.7	32.6	3.5	3.2	12.1	1.0
1424_Ko82_8	5.4	1.4	15.1	12.5	39.9	2.6	2.4	12.2	0.7
1424_Ko82_9	6.0	1.8	25.0	9.9	29.7	2.2	2.5	10.2	0.8
1424_Ko82_10	5.7	1.3	10.1	10.3	42.5	2.6	3.5	13.9	1.1
1424_Ko82_11	5.4	1.4	15.7	11.3	38.2	2.6	2.8	14.1	1.0

Here, the analysis of the seeds with the construct pSUN-5G reveals lines with a pronounced increase in the arachidonic acid content in comparison with the construct pGPTV-Cnl1_d6Des(Pir)_d5Des(Tc)_D6Elo(Pp)_D12Des(Co). In this context, lines with up to 25% ARA were obtained. The additional elongase (TL16y2) must be responsible for this effect (Figure 31, pSUN-5G). The results from this line are compiled in Table 23.

10

Table 23: Fatty acid analysis of transgenic seeds which have been transformed with the construct pSUN-5G.

Lines	Fatty acids									
	16:0	18:0	18:1	18:2 LA	18:3 GLA	18:3 ALA	18:4 SDA	20:3 HGLA	ARA	EPA
WT	5.2	2.3	34.2	37.9	0.0	11.6	0.0	0.0	0.0	0.0
16-1-2	4.2	1.6	20.1	21.5	25.9	4.1	1.8	1.7	8.9	0.8
16-1-3	5.8	2.3	9.9	14.6	33.6	3.1	2.2	2.2	16.0	1.4
16-1-8	5.0	2.8	11.1	12.6	34.9	2.2	1.8	2.6	16.3	1.2
16-2-1	4.9	1.6	14.5	17.4	32.9	3.5	2.0	1.6	12.3	1.0
16-2-5	5.5	3.3	12.9	13.8	32.9	2.9	2.2	1.4	15.4	1.4
16-4-2	5.8	2.5	18.8	14.7	32.0	3.5	2.3	1.2	12.0	1.2
16-4-3	5.9	2.0	19.7	15.0	32.0	3.8	2.4	1.1	11.4	1.2
16-7-2	6.2	4.4	14.3	10.2	30.7	2.0	2.1	1.7	19.4	1.9
16-7-3	5.0	2.5	21.6	13.6	30.7	2.1	1.8	1.5	12.6	1.1
16-7-4	5.3	4.1	18.8	19.5	23.1	4.2	2.2	2.9	11.3	1.4
16-7-5	7.4	1.8	4.2	6.8	33.7	1.8	2.7	2.6	25.8	2.6

5 Example 61: Detection of DHA in seeds of transgenic Indian mustard plants.

Seeds of plants which had been generated with the construct pSUN-8G as described in Example 58 were analyzed as described in Example 59. Besides the LCPUFAs arachidonic acid and eicosapentaenoic acid, docosahexaenoic acid, the product after conversion by the $\Delta 4$ -desaturase from *Thraustochytrium* and $\Delta 5$ -elongases from *Onchorynchis mykiss* and *Ostreococcus tauri*, was also detected in these seeds. Figure 33 shows the chromatogram with the modified fatty acid spectrum in comparison with an untransformed control plant. The results of several measurements are compiled in Table 24.

Table 24 shows the fatty acid analysis of transgenic seeds which have been transformed with the construct pSUN-8G.

- 5 In this experiment, the synthesis of docosahexaenoic acid in seeds was demonstrated for the first time. While the synthesis of DHA in higher plants has been described, for example in WO 2004/071467, the synthesis has not been demonstrated for seeds, only for an embryogenic cell culture.

Equivalents:

10

Many equivalents of the specific embodiments according to the invention described herein can be seen or found by the skilled worker by simple routine experiments. These equivalents are intended to be included in the patent claims.

Table 2: Fatty acid distribution in the seeds of the three different transgenic B. juncea lines

B. juncea lines	No.	18:1	18:2 (LA)	γ 18:3 (GLA)	α 18:3 (ALA)	18:4 (SDA)	20:3 (HGLA)	20:4 (ARA)
WT	1	33.2	38.2	0	12.2	0	0	0
	2	31.3	41.2	0	11.7	0	0	0
8-1424-5	1	25.1	12.8	26.4	3.5	2.4	0.6	8.3
	2	26	12.7	26.3	3.8	2.6	0.6	8.2
	3	25	12.5	25.9	3.4	2.4	0.8	8.5
8-1424-8	1	28.1	13.1	25	5.8	3.7	0.2	6.2
	2	24.7	14.8	26.4	5.2	3	0.3	6.8
8-1424-10	1	25.2	14.2	29.8	5.2	3.4	0.5	5
	2	27.2	12.7	27.9	4.2	2.9	0.3	6.3

The amounts of fatty acids were stated in % by weight.

LA = linoleic acid, GLA = γ -linolenic acid, ALA = α -linolenic acid, SDA = stearidonic acid, HGLA = dihomogamma-linolenic acid, ARA = arachidonic acid, ETA = eicosatetraenoic acid, EPA = eicosapentaenoic acid

Table 3: Fatty acid distribution in the seeds of the three different transgenic B. juncea lines

Sample	No.	18:1 $\Delta 9$	18:2 $\Delta 6,9$	18:2 $\Delta 9,12$ (LA)	18:3 $\Delta 6,9,12$ (GLA)	18:3 $\Delta 9,12,15$ (ALA)	18:4 $\Delta 6,9,12,15$ (SDA)	20:3 $\Delta 8,11,14$ (HGLA)	20:4 $\Delta 5,8,11,14$ (ARA)	20:4 $\Delta 8,11,14,17$ (ETA)	20:5 $\Delta 5,8,11,14,17$ (EPA)
WT	1	35.10	0.00	35.71	0.00	10.80	0.00	0.00	0.00	0.00	0.00
	2	27.79	0.00	32.83	0.00	8.94	0.71	0.00	0.00	0.00	0.00
9-1424-1	1	17.62	1.07	12.32	29.92	2.84	2.17	0.97	13.05	<0.01	1.21
	2	23.68	2.17	10.57	23.70	2.39	1.80	0.98	11.60	<0.01	1.16
	3	17.15	0.94	12.86	31.16	3.19	2.40	1.01	12.09	<0.01	1.16
9-1424-5	1	16.48	1.47	11.09	30.49	3.06	2.56	0.75	11.84	<0.01	1.24
	2	17.70	1.23	11.42	27.94	2.35	1.88	0.64	12.30	0.03	1.12
	3	19.29	1.05	10.95	26.11	2.85	2.11	1.07	12.09	<0.01	1.21
9-1424-6	1	24.71	0.00	41.87	0.00	12.32	0.00	0.00	0.00	0.00	0.00
	2	28.84	0.00	40.65	0.00	10.94	0.00	0.00	0.00	0.00	0.00
	3	29.28	0.00	41.34	0.00	10.76	0.00	0.00	0.00	0.00	0.00

Sample	No.	18:1 $\Delta 9$	18:2 $\Delta 6,9$	18:2 $\Delta 9,12$ (LA)	18:3 $\Delta 6,9,12$ (GLA)	18:3 $\Delta 9,12,15$ (ALA)	18:4 $\Delta 6,9,12,15$ (SDA)	20:3 $\Delta 8,11,14$ (HGLA)	20:4 $\Delta 5,8,11,14$ (ARA)	20:4 $\Delta 8,11,14,17$ (ETA)	20:5 $\Delta 5,8,11,14,17$ (EPA)
9-1424-7	1	32.41	0.00	37.26	0.00	10.05	0.00	0.00	0.00	0.00	0.00
	2	27.76	0.00	36.66	0.00	11.43	0.00	0.00	0.00	0.00	0.00
	3	32.03	0.00	36.27	0.00	9.27	0.00	0.00	0.00	0.00	0.00
9-1424-8	1	19.08	0.61	11.26	23.31	3.73	2.14	1.11	10.93	0.08	1.11
	2	20.34	3.78	10.07	19.59	2.36	1.72	0.68	8.21	<0.01	1.00
	3	28.27	0.00	37.19	0.00	9.32	0.00	0.00	0.00	0.00	0.00
9-1424-9	1	25.95	0.00	37.87	0.00	9.15	0.00	0.00	0.00	0.00	0.00
	2	22.94	0.00	42.69	0.00	9.14	0.00	0.00	0.00	0.00	0.00
	3	18.96	0.61	14.09	23.76	3.17	1.86	0.97	10.46	<0.01	0.94

The amounts of fatty acids were stated in % by weight.

LA = linoleic acid, GLA = γ -linolenic acid, ALA = α -linolenic acid, SDA = stearidonic acid, HGLA = dihomono- γ -linolenic acid, ARA = arachidonic acid, ETA = eicosatetraenoic acid, EPA = eicosapentaenoic acid

Table 4: Fatty acid analysis in seeds of *Brassica juncea*

[illegible]

The amounts of fatty acids were stated in % by weight.

LA = linoleic acid, GLA = γ -linolenic acid, ALA = α -linolenic acid, SDA = stearidonic acid, HGLA = dihomogamma-linolenic acid, ARA = arachidonic acid, ETA = eicosatetraenoic acid, EPA = eicosapentaenoic acid

Table 6: Conversion rates of the fatty acids which have been fed. The conversion rates were calculated using the formula

$$[\text{conversion rate}] = \frac{[\text{product}]}{[\text{substrate}] + [\text{product}]} * 100$$

BioTaur clones area in % of the GC analysis														
Clone	fatty acid	C16:0	C16:1 (n-7)	C18:0	C18:1 (n-9)	C18:3 (n-6)	C18:4 (n-3)	C20:3 (n-6)	C20:4 (n-6)	C20:4 (n-3)	C20:5 (n-3)	C22:4 (n-6)	C22:4 (n-3)	C22:5 (n-3)
Vector	none	21.261	41.576	4.670	25.330									
BioTaur	none	20.831	37.374	4.215	26.475									
Vector	GLA + EPA	22.053	23.632	5.487	17.289	11.574					13.792			
BioTaur	GLA + EPA	20.439	25.554	6.129	19.587	3.521		6.620			10.149			1.127
Vector	EPA	20.669	28.985	6.292	21.712						16.225			
BioTaur	EPA	20.472	26.913	6.570	23.131						11.519			3.251
Vector	ARA	23.169	23.332	6.587	12.735				27.069					
BioTaur	ARA	20.969	31.281	5.367	21.351				9.648			1.632		
Vector	SDA	18.519	12.626	6.642	6.344		47.911							
BioTaur	SDA	19.683	15.878	7.246	8.403		13.569			25.946			0.876	

Table 24: Fatty acid analysis of transgenic seeds which have been transformed with the construct pSUN-8G

I	16:0	18:0	18:1 Δ9	LA 18:2 Δ9,12	GLA 18:3 Δ6,9,12	ALA 18:3 Δ9,12,15	SDA 18:4 Δ6,9,12,15	HGLA 20:3 Δ8,11,14	ARA 20:4 Δ5,8,11,14	EPA 20:5 Δ5,8,11,14 ,17	22:5 Δ7,10,13,16, 19	DHA 22:6 Δ4,7,10,13,16, 19
WT	5.26	1.80	30.78	43.93	nd	12.47	nd	nd	nd	nd	nd	nd
Bj-17-1-3	4.73	2.28	19.30	14.04	31.48	3.09	2.40	1.70	3.37	8.65	0.19	0.25
Bj-17-2-1	4.34	2.17	17.60	15.56	29.97	3.37	2.44	2.14	4.05	9.14	0.23	0.40
Bj-17-4-3	4.31	1.70	14.45	16.94	35.54	3.43	2.39	0.10	5.09	9.43	0.24	0.23

II	% saturated fatty acids			% mono-unsaturated fatty acids		% poly-unsaturated fatty acids		% LCFAs	% VLCFAs
WT	7.96			35.43		56.62		97.71	2.29
Bj-17-1-3	9.18			24.95		65.87		79.64	20.36
Bj-17-2-1	9.83			25.44		64.73		80.44	19.56
Bj-17-4-3	14.05			20.36		65.60		75.27	24.73

LCFAs = all fatty acids up to a length of 18 carbon atoms in the fatty acid chain

VLCFAs = all fatty acids with a length of 20 or more carbon atoms in the fatty acid chain

Comprises/comprising and grammatical variations thereof when used in this specification are to be taken to specify the presence of stated features, integers, steps or components or groups thereof, but do not preclude the presence or addition of one or more other features, integers, steps, components or groups thereof.

THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. A process for the production of C₁₈-, C₂₀- and C₂₂- polyunsaturated fatty acids in the seed of a transgenic oil crop plant with a content of at least 20% by weight based on the total lipid content, which comprises the following process steps:

a) introducing, into the oil crop plant, at least one nucleic acid encoding a Δ 6-desaturase and at least one nucleic acid encoding a Δ 6-elongase, wherein the Δ 6-desaturase has the amino acid sequence shown in SEQ ID NO: 202 or an amino acid sequence having at least 60% sequence identity thereto, or

introducing, into the oil crop plant, at least one nucleic acid encoding a Δ 9-elongase, at least one nucleic acid encoding a Δ 8-desaturase, at least one nucleic acid encoding a Δ 6-desaturase and at least one nucleic acid encoding a Δ 6-elongase, wherein the Δ 6-desaturase has the amino acid sequence shown in SEQ ID NO: 202 or an amino acid sequence having at least 60% sequence identity thereto, and

b) introducing, into the oil crop plant, at least one nucleic acid encoding a Δ 5-desaturase, and

c) introducing, into the oil crop plant, at least one nucleic acid encoding a Δ 5-elongase, and

d) introducing, into the oil crop plant, at least one nucleic acid encoding a Δ 4-desaturase.

2. The process according to claim 1 wherein the C₂₀- and C₂₂- polyunsaturated fatty acids are selected from the group comprising arachidonic acid, eicosapentaenoic acid, docosapentaenoic acid and docosahexaenoic acid.
3. The process according to claim 1 or 2, wherein, in the seed of the transgenic oil crop plant, the content of all C₁₈-, C₂₀- and C₂₂- polyunsaturated fatty acids together amounts to at least 27% by weight based on the total lipid content.
4. The process according to claim 2, wherein, in the seed of the transgenic oil crop plant, the docosahexaenoic acid content amounts to at least 0.23% by weight based on the total lipid content.
5. The process according to any one of claims 1 to 4, wherein the nucleic acid sequences which encode polypeptides with Δ 9-elongase, Δ 6-desaturase, Δ 8-desaturase, Δ 6-elongase, Δ 5-desaturase, Δ 5-elongase or Δ 4-desaturase activity are selected from the group consisting of
 - a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201, or

b) nucleic acid sequences which, as the result of the degeneracy of the genetic code, can be derived from the amino acid sequences shown in SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 70, SEQ ID NO: 72, SEQ ID NO: 74, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138, SEQ ID NO: 184, SEQ ID NO: 194, SEQ ID NO: 198, SEQ ID NO: 200 or SEQ ID NO: 202, or

c) derivatives of the nucleic acid sequence shown in SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, SEQ ID NO: 71, SEQ ID NO: 73, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 111, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137, SEQ ID NO: 183, SEQ ID NO: 193, SEQ ID NO: 197, SEQ ID NO: 199 or SEQ ID NO: 201, which encode polypeptides with at least 40% identity at the amino acid level with SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10,

SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 70, SEQ ID NO: 72, SEQ ID NO: 74, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 112, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138, SEQ ID NO: 184, SEQ ID NO: 198, or SEQ ID NO: 200 or, in respect of $\Delta 6$ -desaturase, which encode polypeptides with at least 60% identity at the amino acid level with SEQ ID NO: 194 or SEQ ID NO: 202, and which have $\Delta 9$ -elongase, $\Delta 6$ -desaturase, $\Delta 8$ -desaturase, $\Delta 6$ -elongase, $\Delta 5$ -desaturase, $\Delta 5$ -elongase or $\Delta 4$ -desaturase activity.

6. The process according to any one of claims 1 to 5, wherein a nucleic acid sequence which encodes a polypeptide with $\omega 3$ -desaturase activity, selected from the group consisting of:

a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 87 or SEQ ID NO: 105, or

b) nucleic acid sequences which, as the result of the degeneracy of the genetic code, can be derived from the amino acid sequence shown in SEQ ID NO: 88 or SEQ ID NO: 106, or

c) derivatives of the nucleic acid sequence shown in SEQ ID NO: 87 or SEQ ID NO: 105, which encode polypeptides with at least 60% identity at the amino acid level with SEQ ID NO: 88 or SEQ ID NO: 106 and which have $\omega 3$ -desaturase activity

is additionally introduced into the transgenic oil crop plant.

7. The process according to any one of claims 1 to 6, wherein a nucleic acid sequence which encodes a polypeptide with $\Delta 12$ -desaturase activity, selected from the group consisting of:

a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 107, SEQ ID NO: 109 or SEQ ID NO: 195, or

b) nucleic acid sequences which, as the result of the degeneracy of the genetic code, can be derived from the amino acid sequence shown in SEQ ID NO: 108, SEQ ID NO: 110 or SEQ ID NO: 196, or

c) derivatives of the nucleic acid sequence shown in SEQ ID NO: 107, SEQ ID NO: 109 or SEQ ID NO: 195, which encode polypeptides with at least 60% identity at the amino acid level with SEQ ID NO: 108, SEQ ID NO: 110 or SEQ ID NO: 196 and which have $\Delta 12$ -desaturase activity

is additionally introduced into the transgenic oil crop plant.

8. The process according to any one of claims 1 to 7, wherein one or more nucleic acid sequences which encode a polypeptide or polypeptides involved in fatty acid biosynthesis or lipid metabolism selected from the group consisting of acyl-CoA dehydrogenase(s), acyl-ACP [= acyl carrier protein] desaturase(s), acyl-ACP thioesterase(s), fatty acid acyltransferase(s), acyl-CoA:lysophospholipid acyltransferase(s), fatty acid synthase(s), fatty acid hydroxylase(s), acetyl-coenzyme A carboxylase(s), acyl-coenzyme A oxidase(s), fatty acid desaturase(s), fatty acid acetylenases, lipoxygenases, triacylglycerol lipases, allenoxide synthases, hydroperoxide lyases and fatty acid elongase(s) is/are additionally introduced into the transgenic oil crop plant.

9. The process according to any one of claims 1 to 8, wherein the C₁₈-, C₂₀- and C₂₂- polyunsaturated fatty acids are isolated from the seed of the transgenic oil crop plant in the form of their oils, lipids or free fatty acids.

10. The process according to any one of the previous claims comprising:

a) introducing, into the oil crop plant, at least one nucleic acid sequence which encodes a polypeptide with a $\Delta 6$ -desaturase activity and is selected from the group consisting of:

i) a nucleic acid sequence which encodes the amino acid sequence shown in SEQ ID NO: 202, and

ii) a nucleic acid sequence which encodes an amino acid sequence which has at least 60% identity with the sequence shown in SEQ ID NO: 202, and

b) introducing, into the oil crop plant, at least one nucleic acid sequence which encodes a polypeptide with a $\Delta 6$ -elongase activity and is selected from the group consisting of:

i) a nucleic acid sequence with the sequence shown in SEQ ID NO: 27 or SEQ ID NO: 199,

ii) a nucleic acid sequence which encodes the amino acid sequence shown in SEQ ID NO: 28 or SEQ ID NO: 200,

iii) a nucleic acid sequence which hybridizes under stringent conditions with the complementary strand of the nucleic acid sequence shown in SEQ ID NO: 27 or SEQ ID NO: 199, and

iv) a nucleic acid sequence which has at least 60% identity with the sequence shown in SEQ ID NO: 27 or at least 80% identity with the sequence shown in SEQ ID NO: 199, and

c) introducing, into the oil crop plant, at least one nucleic acid sequence which encodes a polypeptide with a $\Delta 5$ -desaturase activity and is selected from the group consisting of:

i) a nucleic acid sequence with the sequence shown in SEQ ID NO: 11,

ii) a nucleic acid sequence which encodes the amino acid sequence shown in SEQ ID NO: 12,

iii) a nucleic acid sequence which hybridizes under stringent conditions with the complementary strand of the nucleic acid sequence shown in SEQ ID NO: 11, and

iv) a nucleic acid sequence which has at least 60% identity with the sequence shown in SEQ ID NO: 11, and

d) introducing, into the oil crop plant, a nucleic acid sequence which encodes a polypeptide with a $\Delta 5$ -elongase activity.

11. The process according to claim 10, wherein a nucleic acid sequence which encodes a polypeptide with a $\Delta 12$ -desaturase activity selected from the group consisting of:

a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 195,

b) a nucleic acid sequence which encodes the amino acid sequence shown in SEQ ID NO: 196,

c) a nucleic acid sequence which hybridizes under stringent conditions with the complementary strand of the nucleic acid sequence shown in SEQ ID NO: 195, and

d) a nucleic acid sequence which has at least 60% identity with the sequence shown in SEQ ID NO: 195,

is additionally introduced into the transgenic oil crop plant.

12. The process according to claim 11, wherein the $\Delta 12$ -desaturase and the $\Delta 5$ -elongase are expressed under the control of a seed-specific promoter.

13. The process according to claim 10, wherein the $\Delta 5$ -elongase nucleic acid sequence is selected from the group consisting of:

a) a nucleic acid sequence with the sequence shown in SEQ ID NO: 43, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137 or SEQ ID NO: 197,

b) a nucleic acid sequence which encodes the amino acid sequence shown in SEQ ID NO: 44, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, SEQ ID NO: 76, SEQ ID NO: 78, SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 86, SEQ ID NO: 114, SEQ ID NO: 118, SEQ ID NO: 120, SEQ ID NO: 132, SEQ ID NO: 134, SEQ ID NO: 136, SEQ ID NO: 138 or SEQ ID NO: 198,

c) a nucleic acid sequence which hybridizes under stringent conditions with the complementary strand of the nucleic acid sequence shown in SEQ ID NO: 43, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137 or SEQ ID NO: 197, and

d) a nucleic acid sequence which has at least 60% identity with the sequence shown in SEQ ID NO: 43, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 75, SEQ ID NO: 77, SEQ ID NO: 79, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 113, SEQ ID NO: 117, SEQ ID NO: 119, SEQ ID NO: 131, SEQ ID NO: 133, SEQ ID NO: 135, SEQ ID NO: 137 or SEQ ID NO: 197.

14. The process according to any one of claims 9 to 13, wherein all nucleic acid sequences are introduced into the oil crop plant on a shared recombinant nucleic acid molecule.

15. The process according to claim 14, wherein each nucleic acid sequence is under the control of its own promoter.

16. The process according to any one of claims 10 to 15, wherein the arachidonic acid concentration amounts to at least 25% based on the total lipid content, or wherein the eicosapentaenoic acid concentration amounts to at least 2.6% based on the total lipid content.

17. An oil, lipid or fatty acid or a fraction thereof, obtained by a process according to any one of the preceding claims.

18. The use of a $\Delta 12$ -desaturase, a $\Delta 6$ -desaturase, a $\Delta 5$ -desaturase, a $\Delta 6$ -elongase and a $\Delta 5$ -elongase as defined in any one of claims 10, 11 and 13, for the production of C_{18} -, C_{20} - and C_{22} - polyunsaturated fatty acids.

19. A recombinant nucleic acid molecule comprising:

a) one or more copies of a promoter which is active in plant cells, preferably a seed-specific promoter, and

b) at least one nucleic acid sequence as defined in claim 10 which encodes a polypeptide with $\Delta 6$ -desaturase activity, and

c) at least one nucleic acid sequence as defined in claim 10 which encodes a polypeptide with $\Delta 5$ -desaturase activity, and

d) at least one nucleic acid sequence as defined in claim 10 which encodes a polypeptide with $\Delta 6$ -elongase activity, and

e) at least one nucleic acid sequence as defined in claim 14 which encodes a polypeptide with $\Delta 5$ -elongase activity, and

f) one or more copies of a terminator sequence.

20. The recombinant nucleic acid molecule according to claim 19, additionally comprising a nucleic acid sequence as defined in claim 11 which encodes a $\Delta 12$ -desaturase.

21. The recombinant nucleic acid molecule according to claim 19 or 20, additionally comprising one or more nucleic acid sequences encoding a polypeptide or polypeptides involved in fatty acid biosynthesis or lipid metabolism selected from the group consisting of acyl-CoA dehydrogenase(s), acyl-ACP [= acyl carrier protein] desaturase(s), acyl-ACP thioesterase(s), fatty acid acyltransferase(s), acyl-CoA:lysophospholipid acyltransferase(s), fatty acid synthase(s), fatty acid hydroxylase(s), acetyl-coenzyme A carboxylase(s), acyl-coenzyme A oxidase(s), fatty acid desaturase(s), fatty acid acetylenases, lipoxygenases, triacylglycerol lipases, allenoxide synthases, hydroperoxide lyases and fatty acid elongase(s).

22. The recombinant nucleic acid molecule according to any one of claims 19 to 21 additionally comprising one or more nucleic acid sequences encoding a polypeptide or polypeptides selected from the group consisting of $\Delta 4$ -desaturase, $\Delta 8$ -desaturase, $\Delta 9$ -desaturase and $\Delta 9$ -elongase.

23. A transgenic oil crop plant comprising a recombinant nucleic acid molecule according to any one of claims 19 to 22.

24. A process of producing oils, lipids or fatty acid compositions by mixing oils, lipids or fatty acids produced by a process according to any one of claims 1 to 16 with animal or microbial oils, lipids or fatty acids.
25. The use of oils, lipids or fatty acids produced by a process according to any one of claims 1 to 16 in feed, foodstuffs, cosmetics or pharmaceuticals.
26. An isolated nucleic acid sequence which encodes a polypeptide with $\Delta 5$ -elongase activity and which has the sequence shown in SEQ ID NO: 197.
27. A gene construct comprising an isolated nucleic acid according to claim 26, wherein the nucleic acid is linked functionally to one or more regulatory signals.
28. The gene construct according to claim 27, wherein the nucleic acid construct comprises one or more nucleic acids encoding a polypeptide or polypeptides involved in fatty acid biosynthesis or lipid metabolism selected from the group consisting of acyl-CoA dehydrogenase(s), acyl-ACP [= acyl carrier protein] desaturase(s), acyl-ACP thioesterase(s), fatty acid acyltransferase(s), acyl-CoA:lysophospholipid acyltransferase(s), fatty acid synthase(s), fatty acid hydroxylase(s), acetyl-coenzyme A carboxylase(s), acyl-coenzyme A oxidase(s), fatty acid desaturase(s), fatty acid acetylenases, lipoxygenases, triacylglycerol lipases, allenoxide synthases, hydroperoxide lyases and fatty acid elongase(s).

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Figure 1: Various synthetic pathways for the biosynthesis of DHA (docosahexaenoic acid)

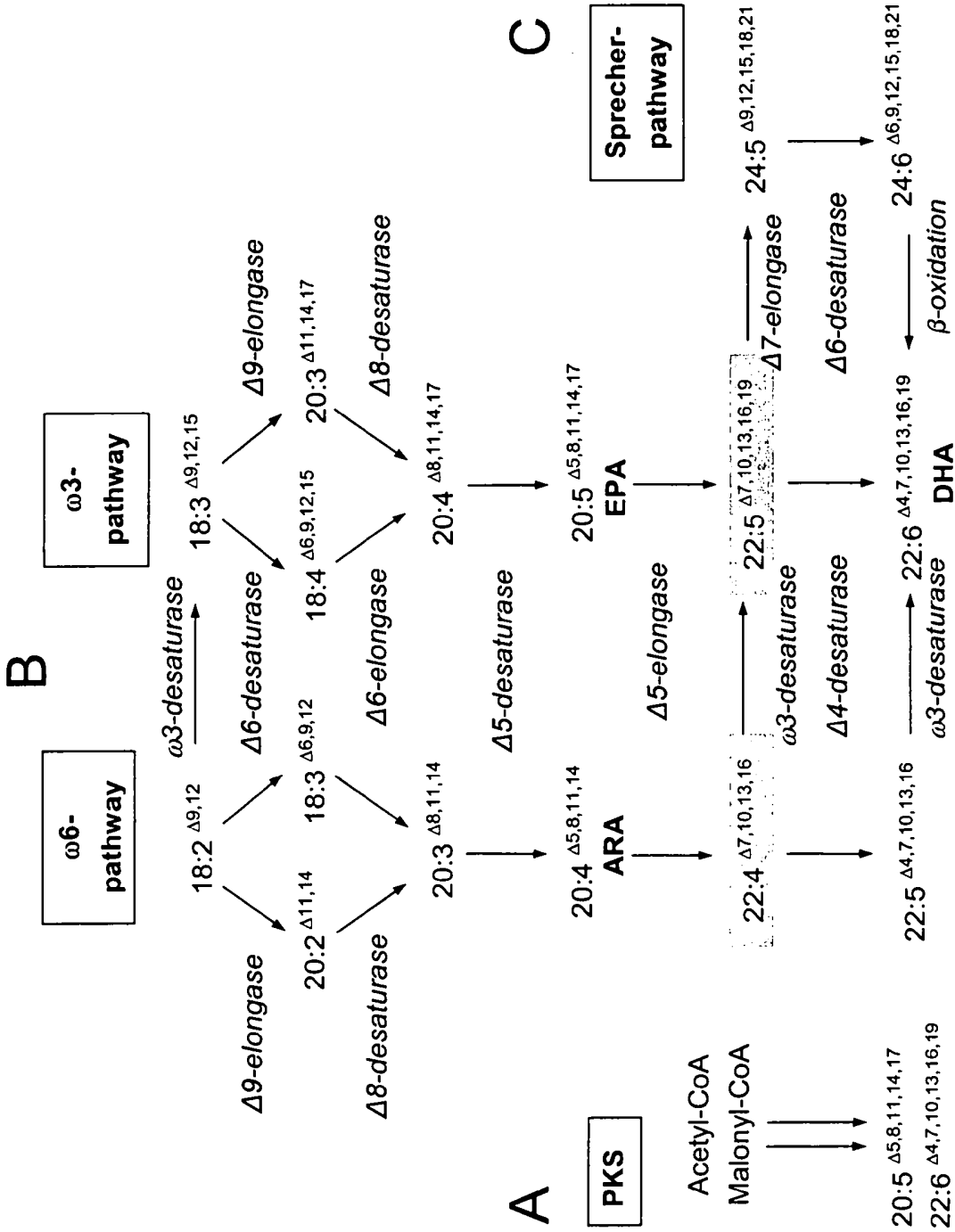


Figure 2: Substrate specificity of the $\Delta 5$ -elongase (SEQ ID NO: 53) with regard to different fatty acids

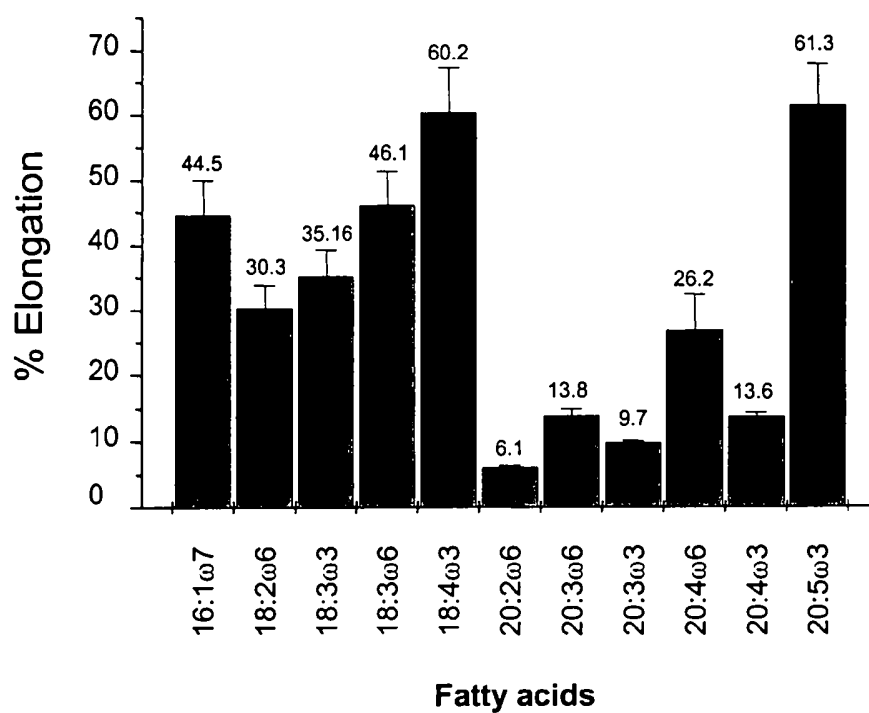


Figure 3: Reconstitution of DHA biosynthesis in yeast starting from 20:5 ω 3.

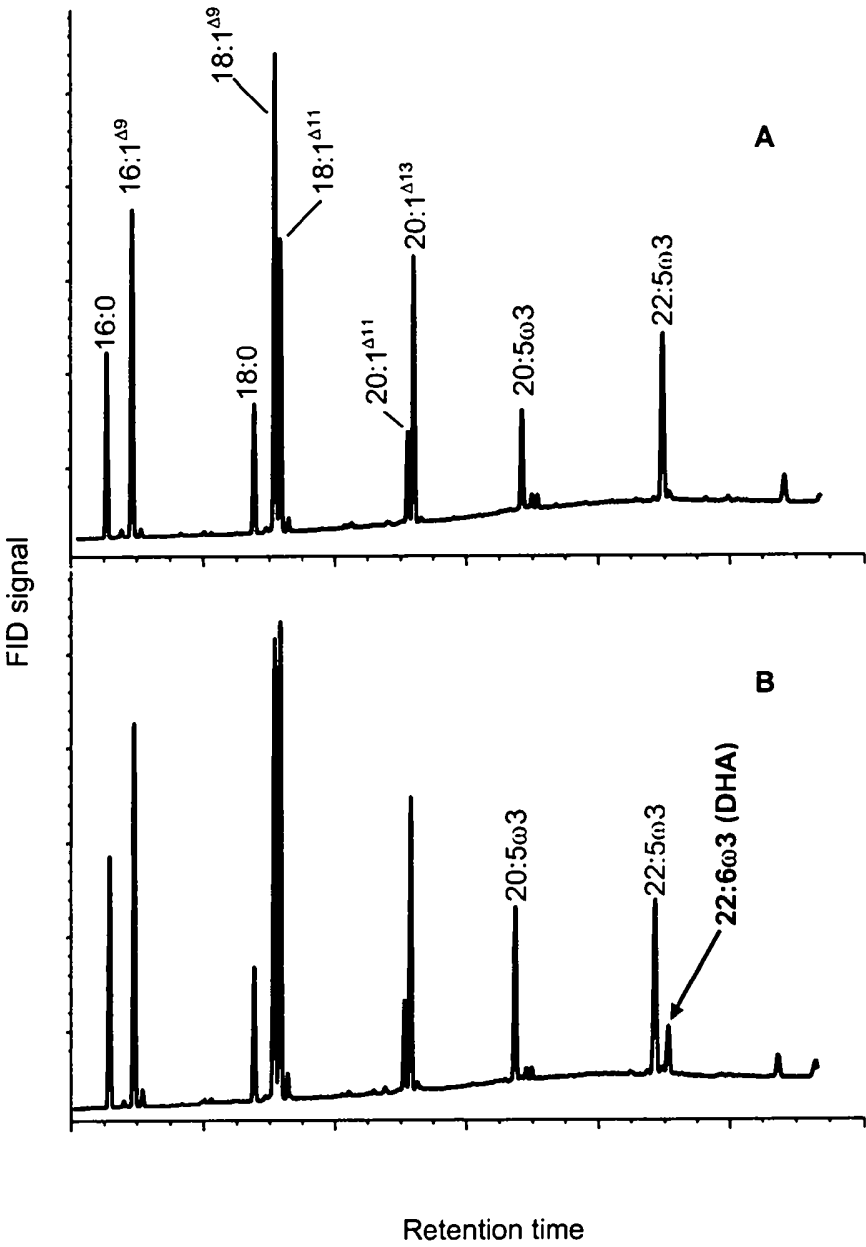


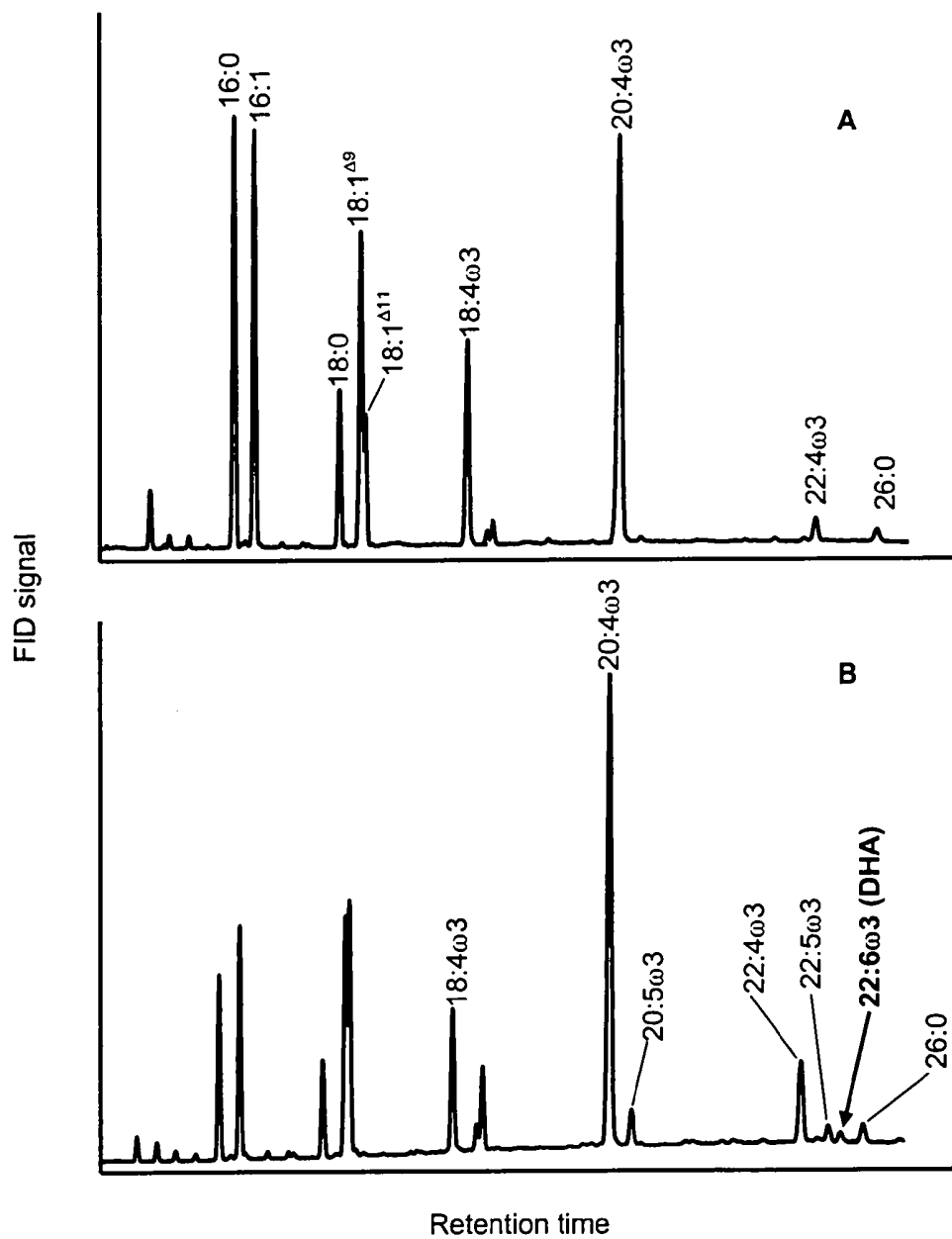
Figure 4: Reconstitution of DHA biosynthesis in yeast starting from 18:4 ω 3.

Figure 5: Fatty acid composition (in mol%) of transgenic yeasts which had been transformed with the vectors pYes3-OmELO3/pYes2-EgD4 or pYes3-OmELO3/pYes2-EgD4+pESCLEu-PtD5. The yeast cells were cultured in minimal medium without tryptophan and uracil/ and leucin in the presence of 250 μ M 20:5 $^{\Delta 5,8,11,14,17}$ and 18:4 $^{\Delta 6,9,12,15}$, respectively. The fatty acid methyl esters were obtained from cell sediments by acid methanolysis and analyzed via GLC. Each value represents the mean (n=4) $\pm$ standard deviation.

Fatty acids	pYes3-OmELO/pYes2-EgD4	pYes3-OmELO/pYes2-EgD4 EgD4 + pESCLEu-PtD5
	Feeding of 20:5 $^{\Delta 5,8,11,14,17}$	Feeding of 18:4 $^{\Delta 6,9,12,15}$
16:0	9.35 $\pm$ 1.61	7.35 $\pm$ 1.37
16:1 $^{\Delta 9}$	14.70 $\pm$ 2.72	10.02 $\pm$ 1.81
18:0	5.11 $\pm$ 1.09	4.27 $\pm$ 1.21
18:1 $^{\Delta 9}$	19.49 $\pm$ 3.01	10.81 $\pm$ 1.95
18:1 $^{\Delta 11}$	18.93 $\pm$ 2.71	11.61 $\pm$ 1.48
18:4 $^{\Delta 6,9,12,15}$	-	7.79 $\pm$ 1.29
20:1 $^{\Delta 11}$	3.24 $\pm$ 0.41	1.56 $\pm$ 0.23
20:1 $^{\Delta 13}$	11.13 $\pm$ 2.07	4.40 $\pm$ 0.78
20:4 $^{\Delta 8,11,14,17}$	-	30.05 $\pm$ 3.16
20:5 $^{\Delta 5,8,11,14,17}$	6.91 $\pm$ 1.10	3.72 $\pm$ 0.59
22:4 $^{\Delta 10,13,16,17}$	-	5.71 $\pm$ 1.30
22:5 $^{\Delta 7,10,13,16,19}$	8.77 $\pm$ 1.32	1.10 $\pm$ 0.27
22:6 $^{\Delta 4,7,10,13,16,19}$	2.73 $\pm$ 0.39	0.58 $\pm$ 0.10

Figure 6: Feeding experiment for determining the functionality and substrate specificity with yeast strains

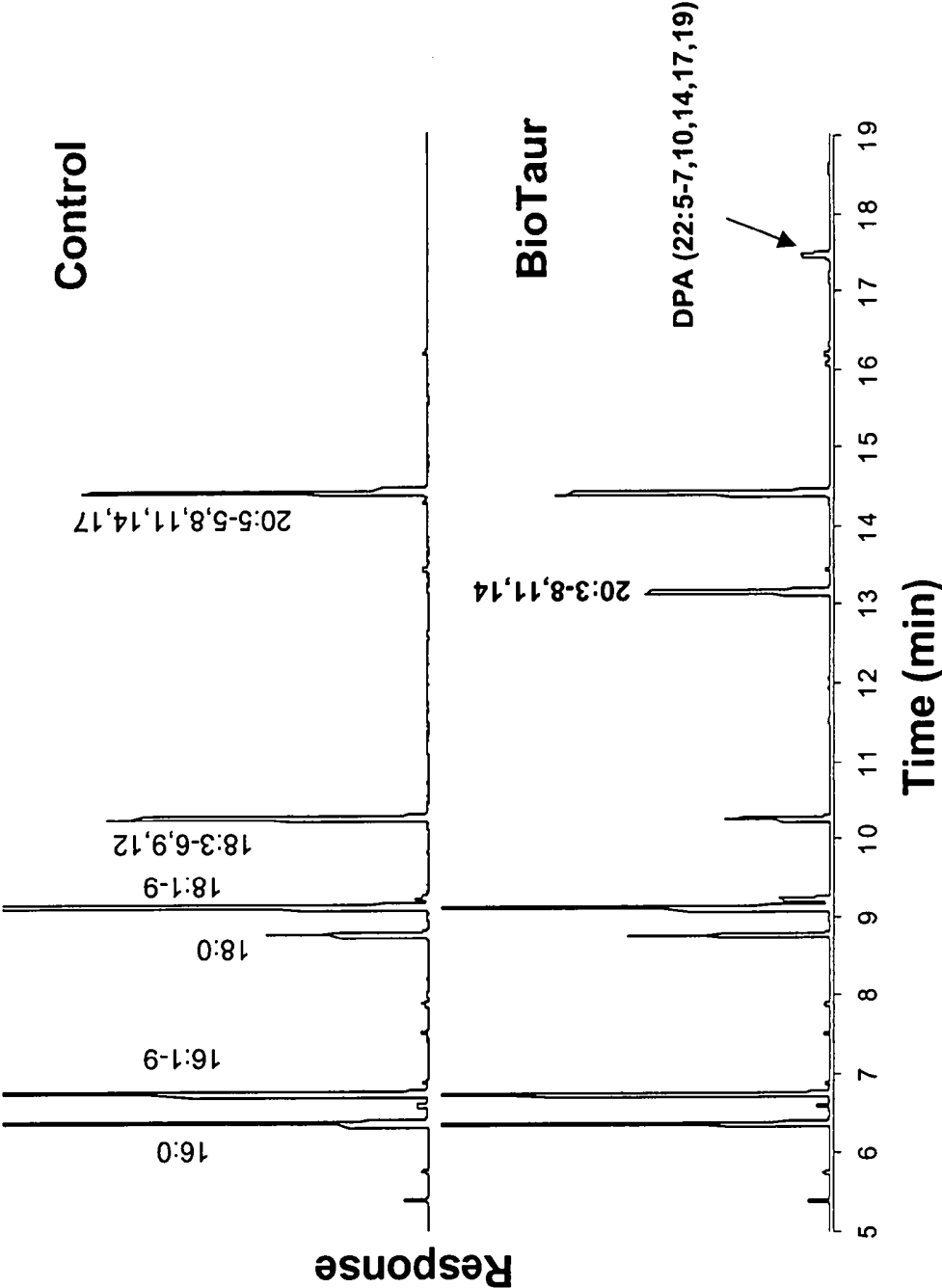


Figure 7: Elongation of eicosapentaenoic acid by OtElo1

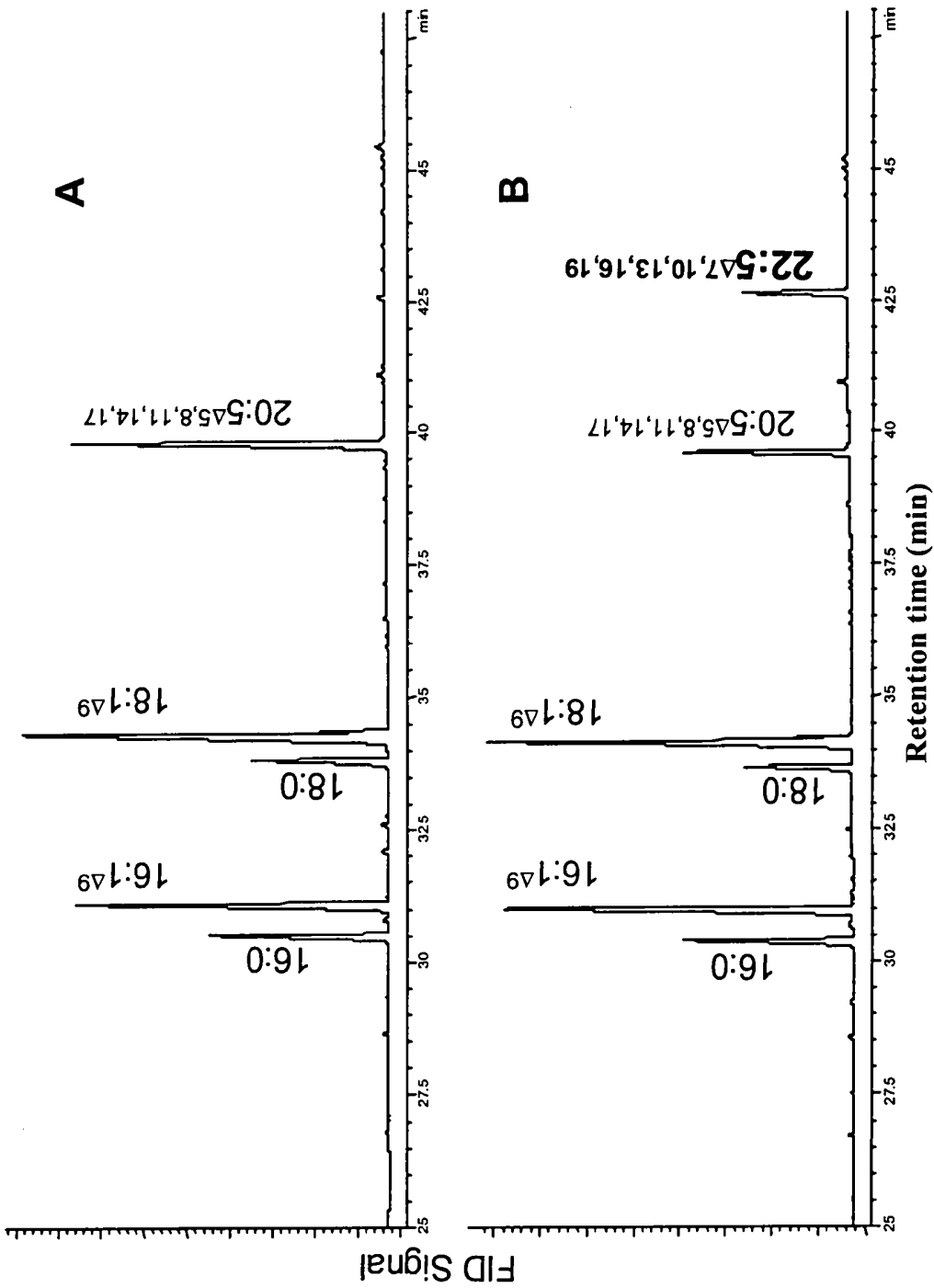


Figure 8: Elongation of arachidonic acid by Otlol1

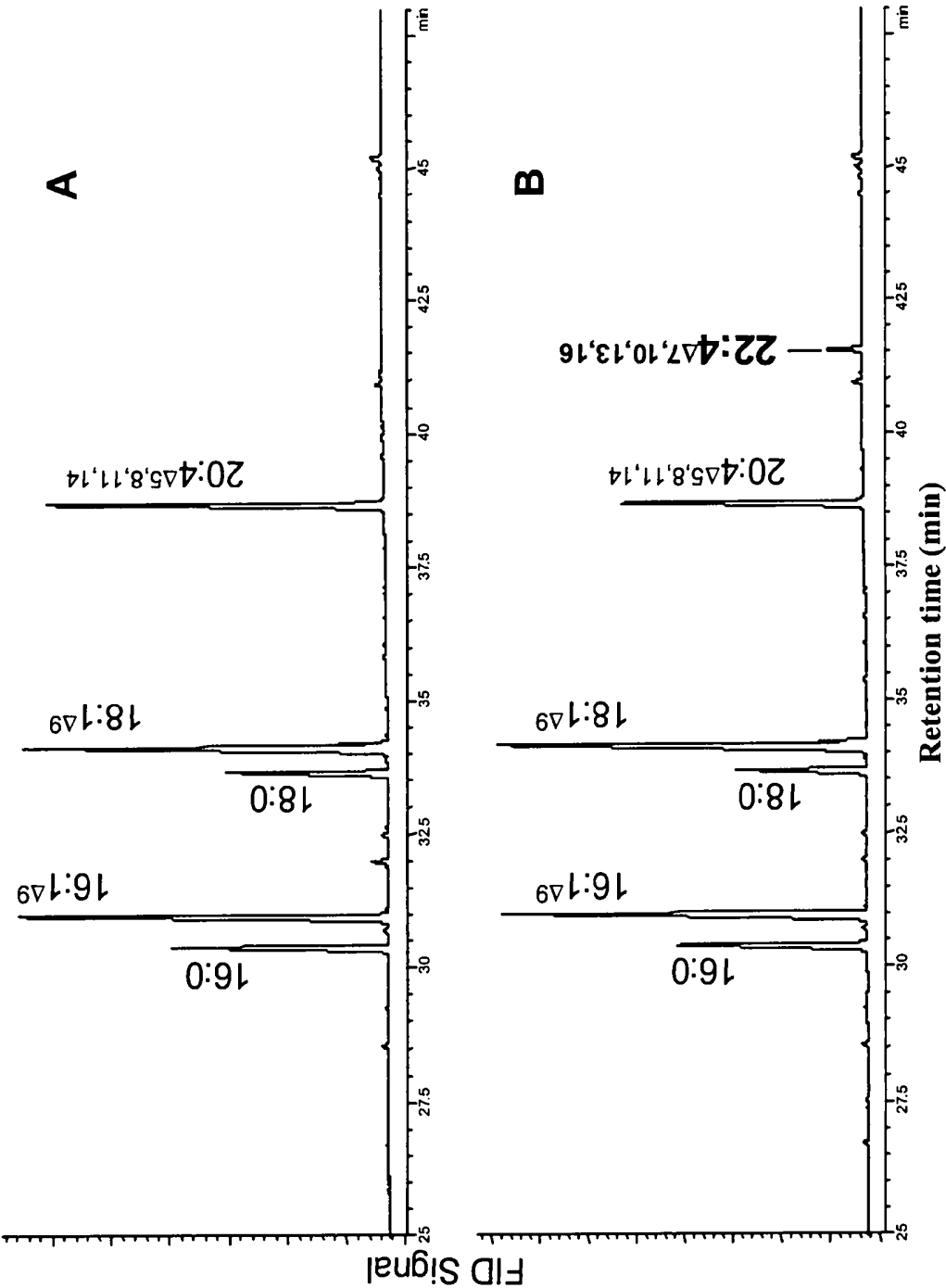


Figure 9: Expression of TpELO1 in yeast

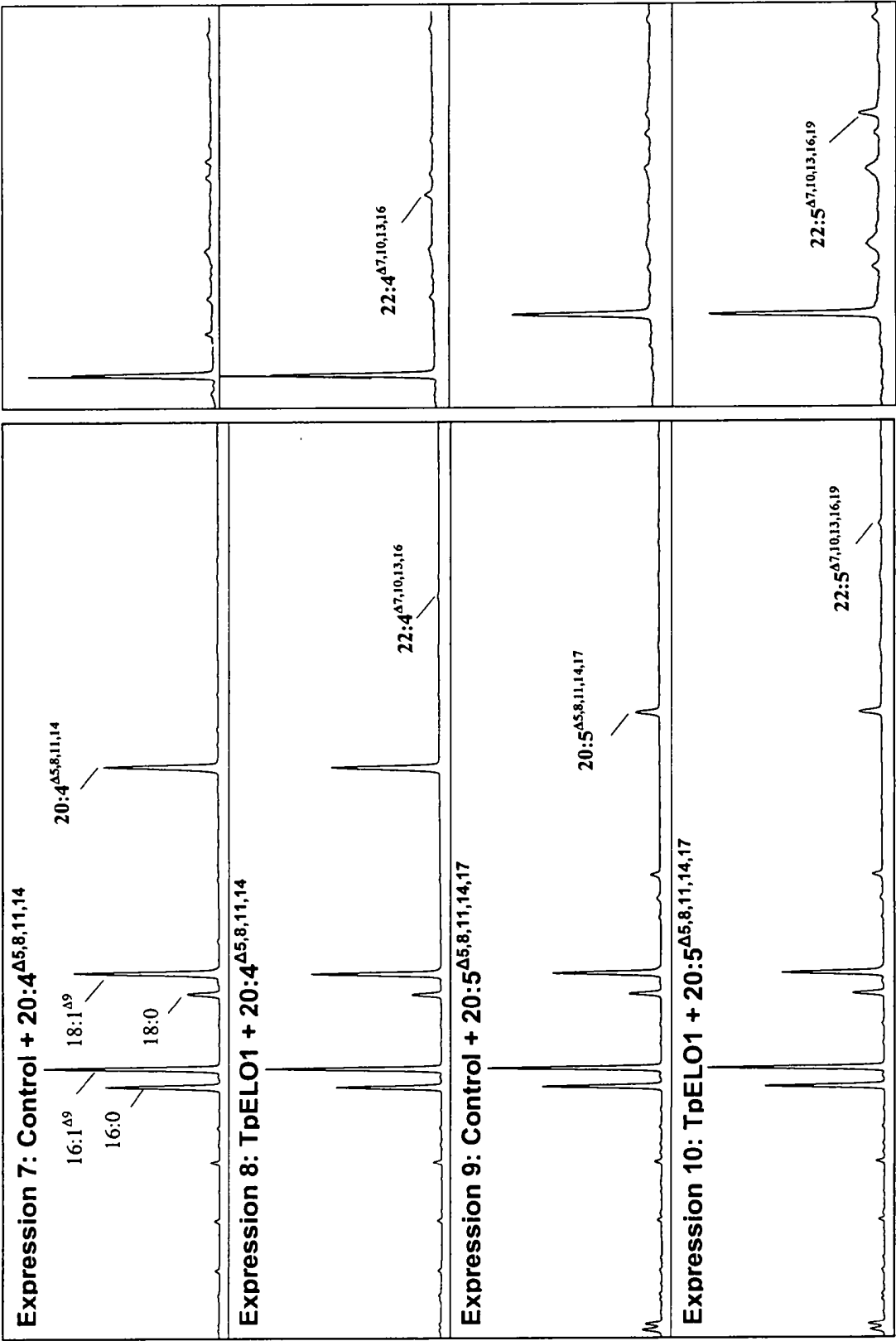


Figure 10: Expression of TpELO3 in yeast

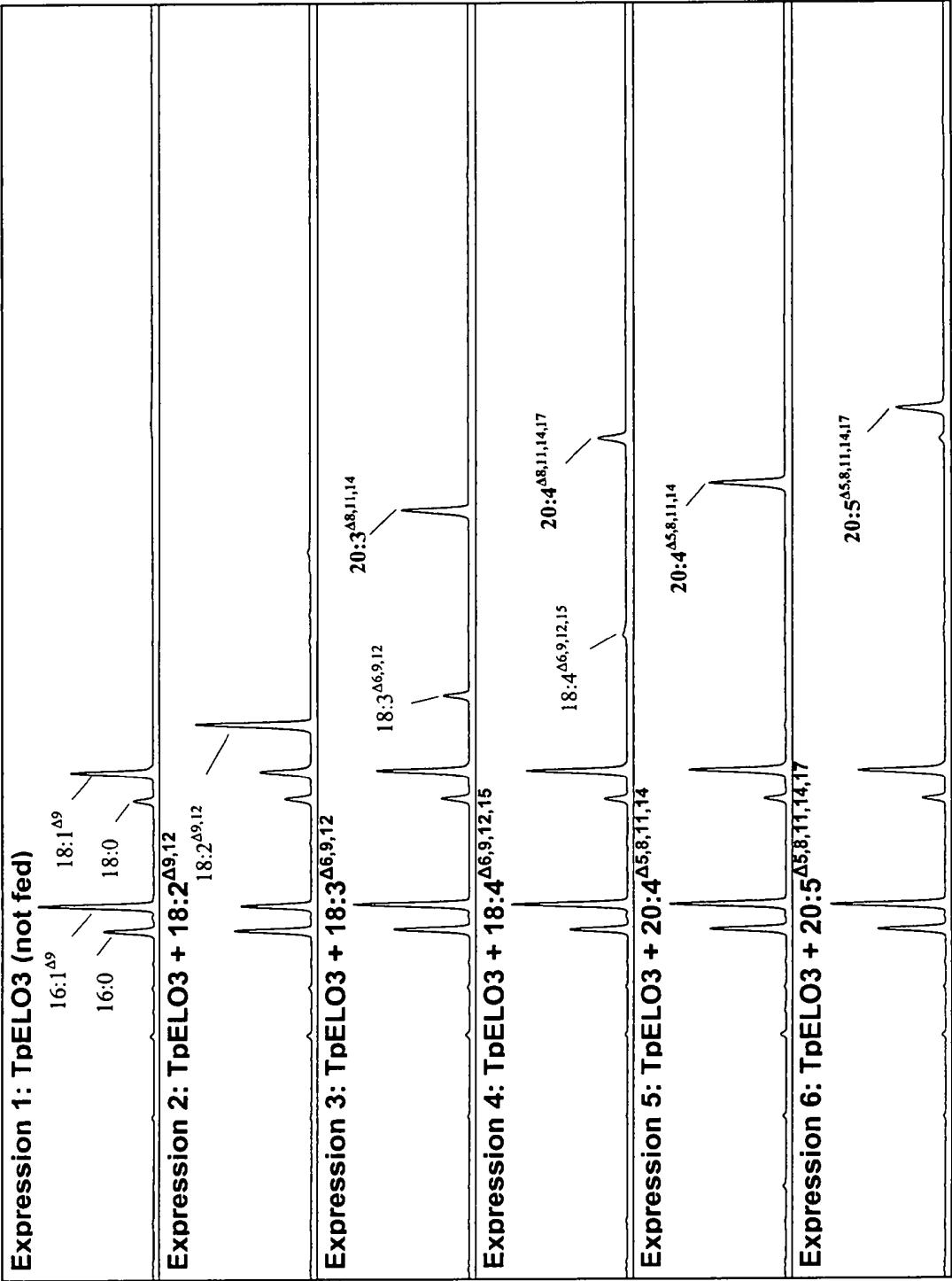


Figure 11: Expression of Thraustochytrium $\Delta 5$ -elongase TL16/pYES2.1 in yeast

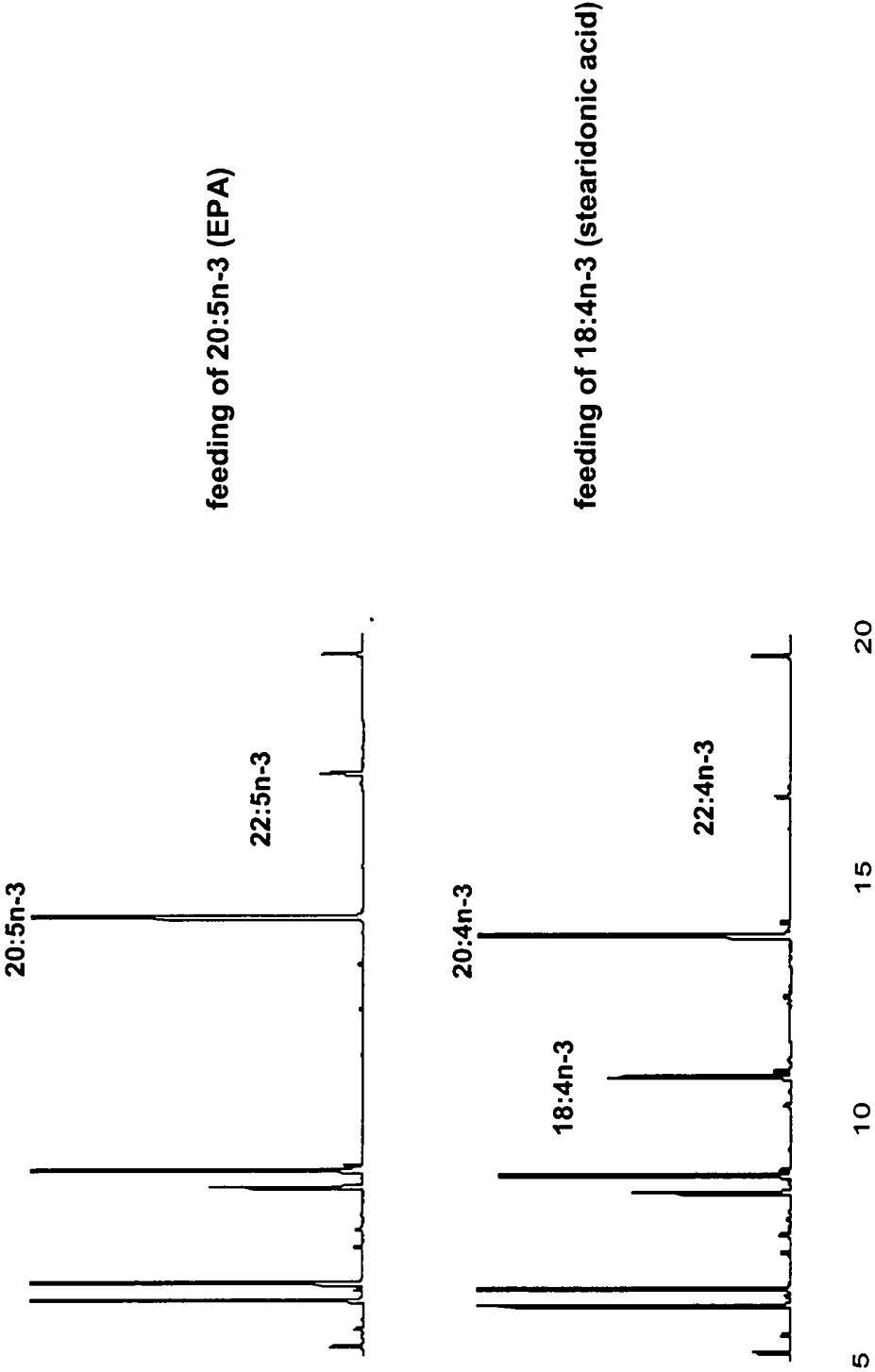


Figure 12: Desaturation of γ -linolenic acid (18:2 ω 6-fatty acid) to give α -linolenic acid (18:3 ω 3-fatty acid) by Pi-omega3Des.

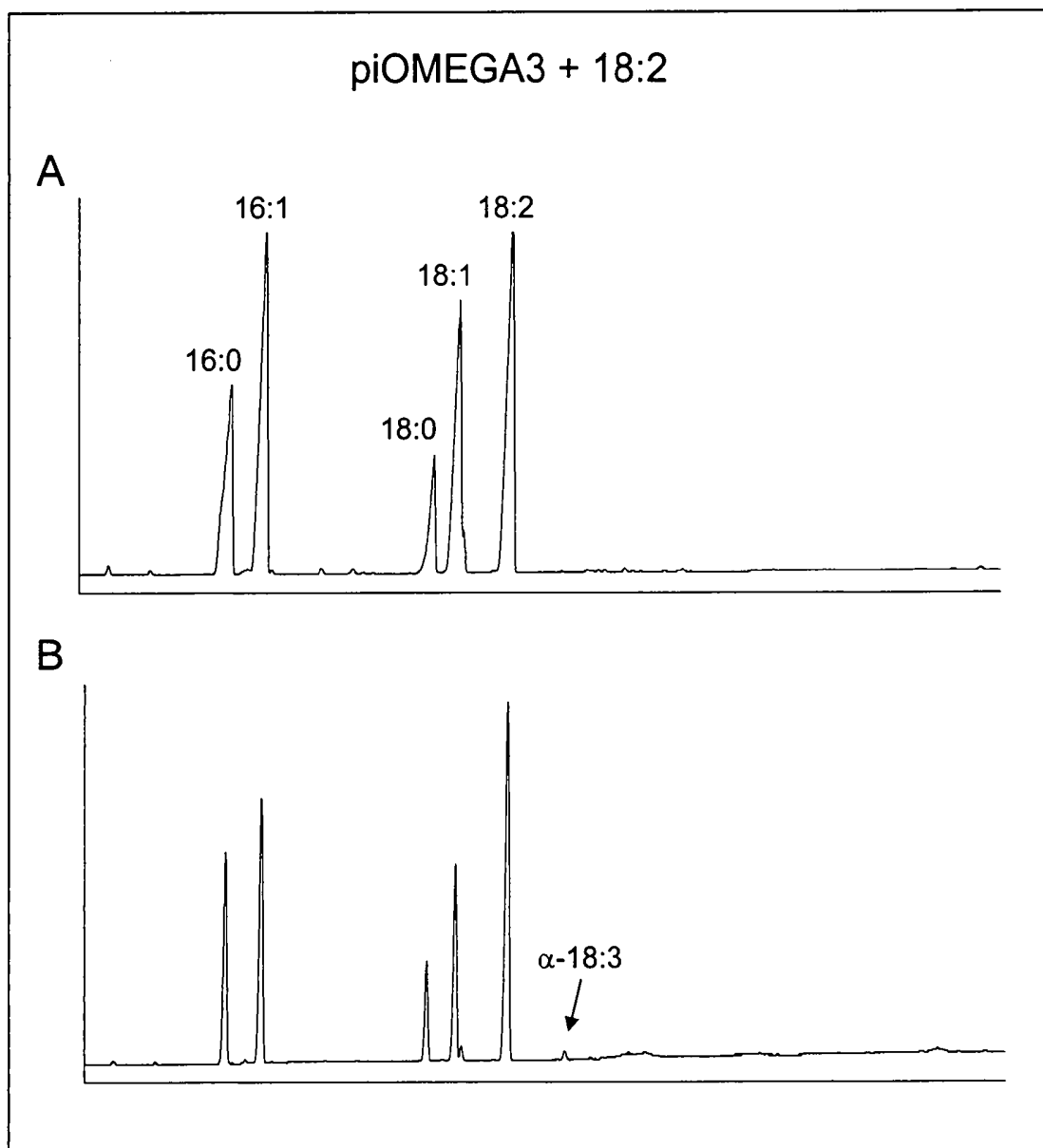


Figure 13: Desaturation of γ -linolenic acid (18:2 ω 6-fatty acid) to give stearidonic acid (18:4 ω 3-fatty acid) by Pi-omega3Des.

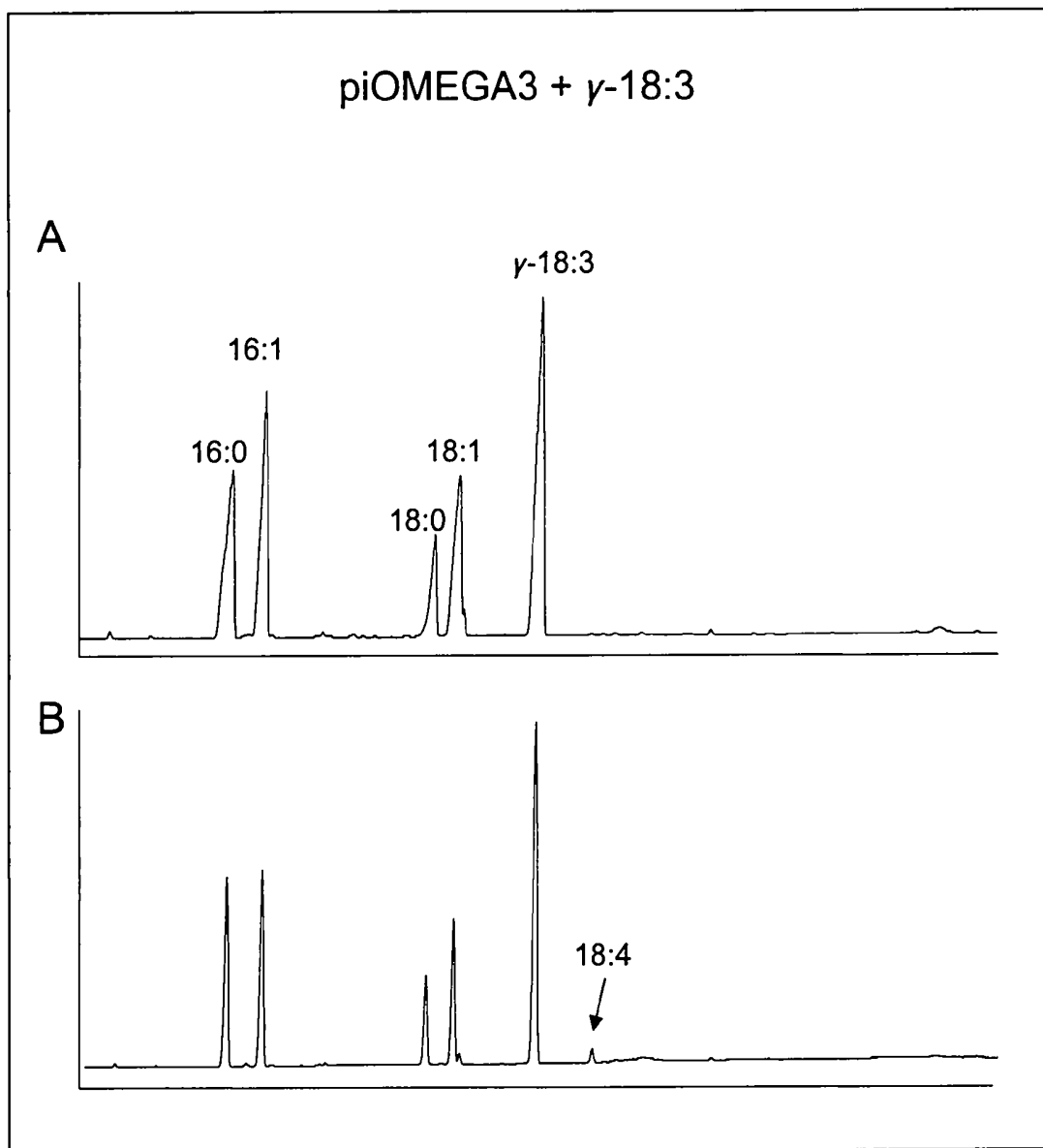


Figure 14: Desaturation of C20:2 ω 6-fatty acid to give C20:3 ω 3-fatty acid by Pi-omega3Des.

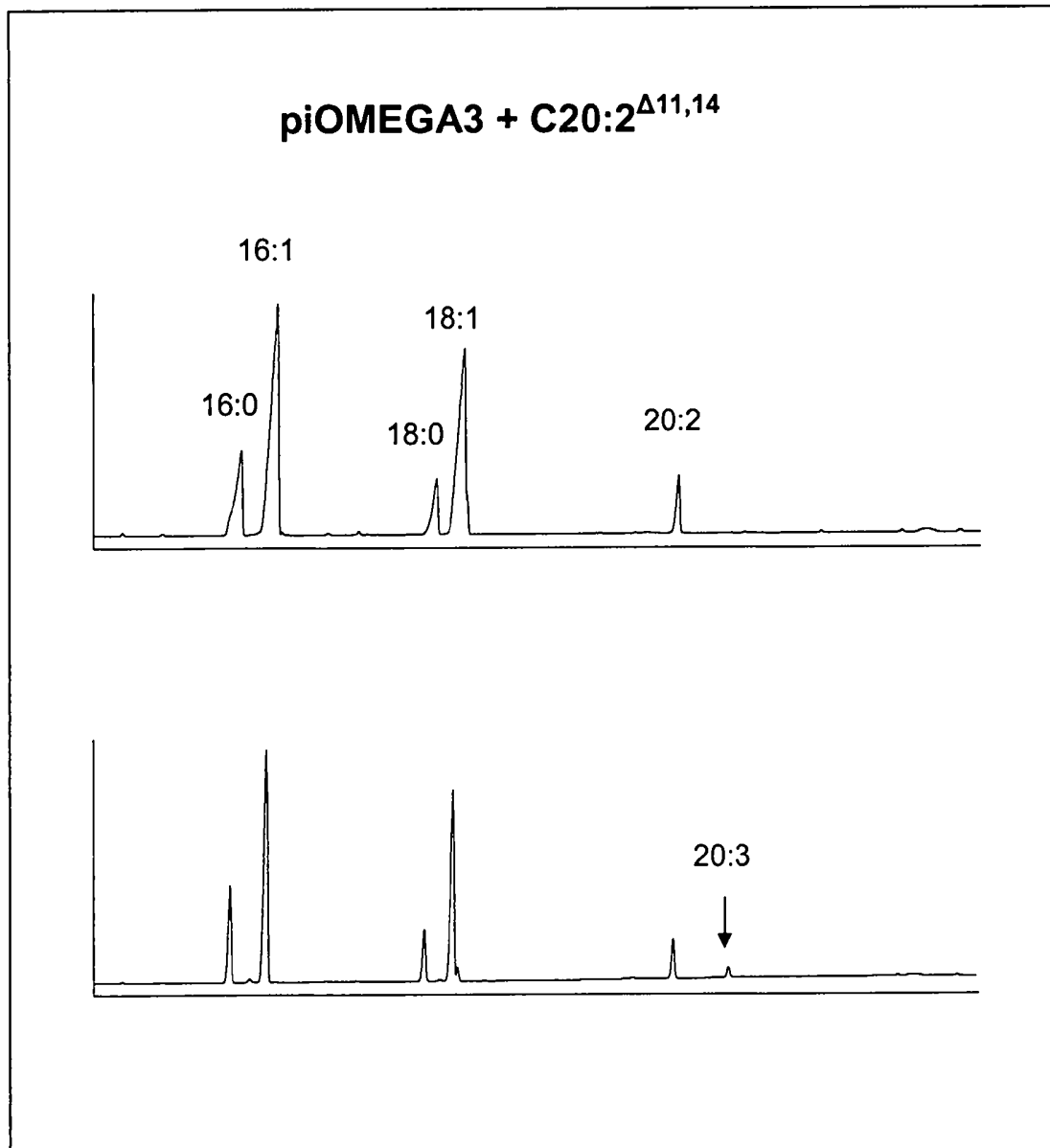


Figure 15: Desaturation of C20:3 ω 6-fatty acid to give C20:4 ω 3-fatty acid by Pi-
omega3Des.

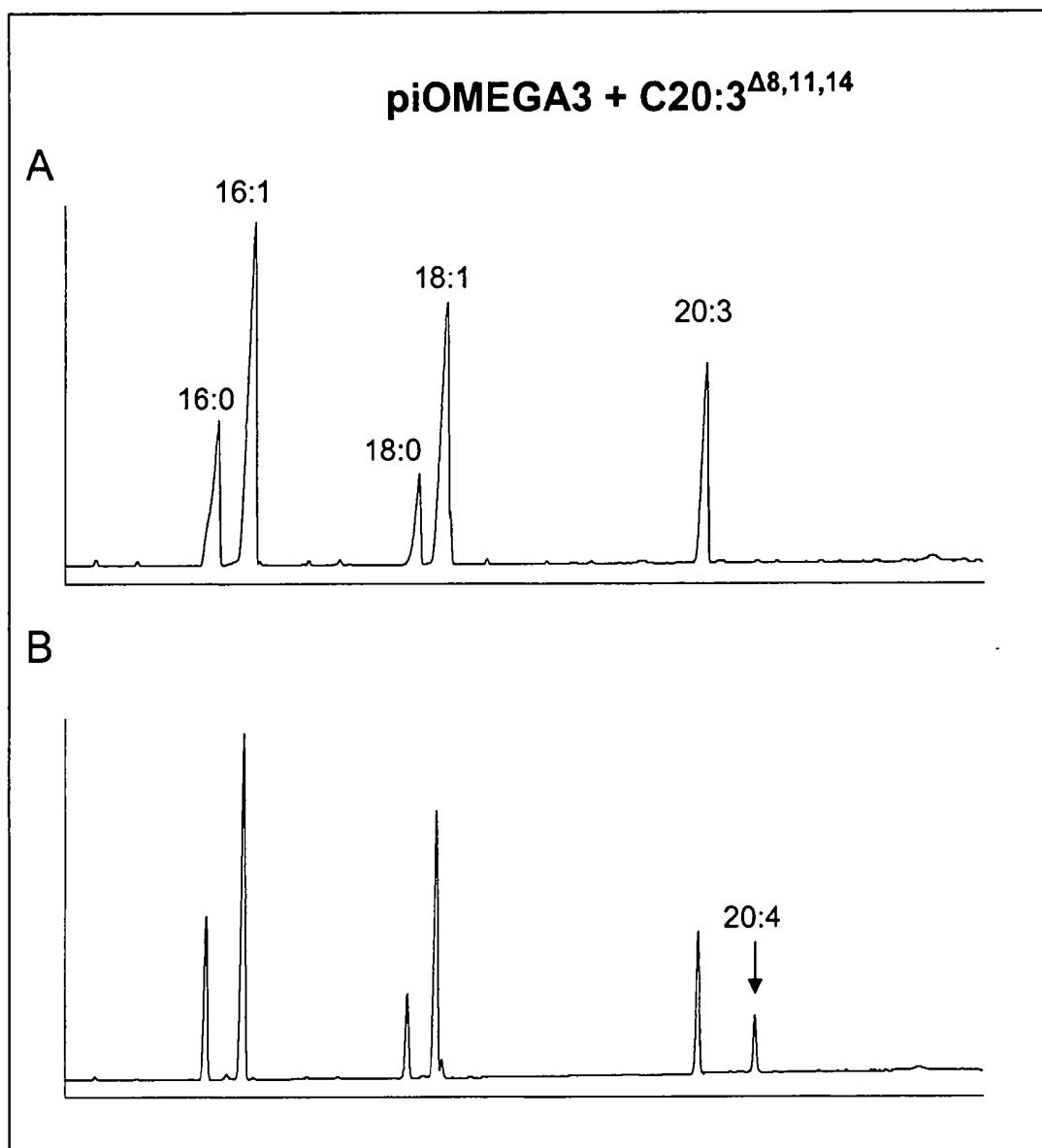


Figure 16: Desaturation of arachidonic acid (C20:4 ω 6-fatty acid) to give eicosapentaenoic acid (C20:5 ω 3-fatty acid) by Pi-omega3Des.

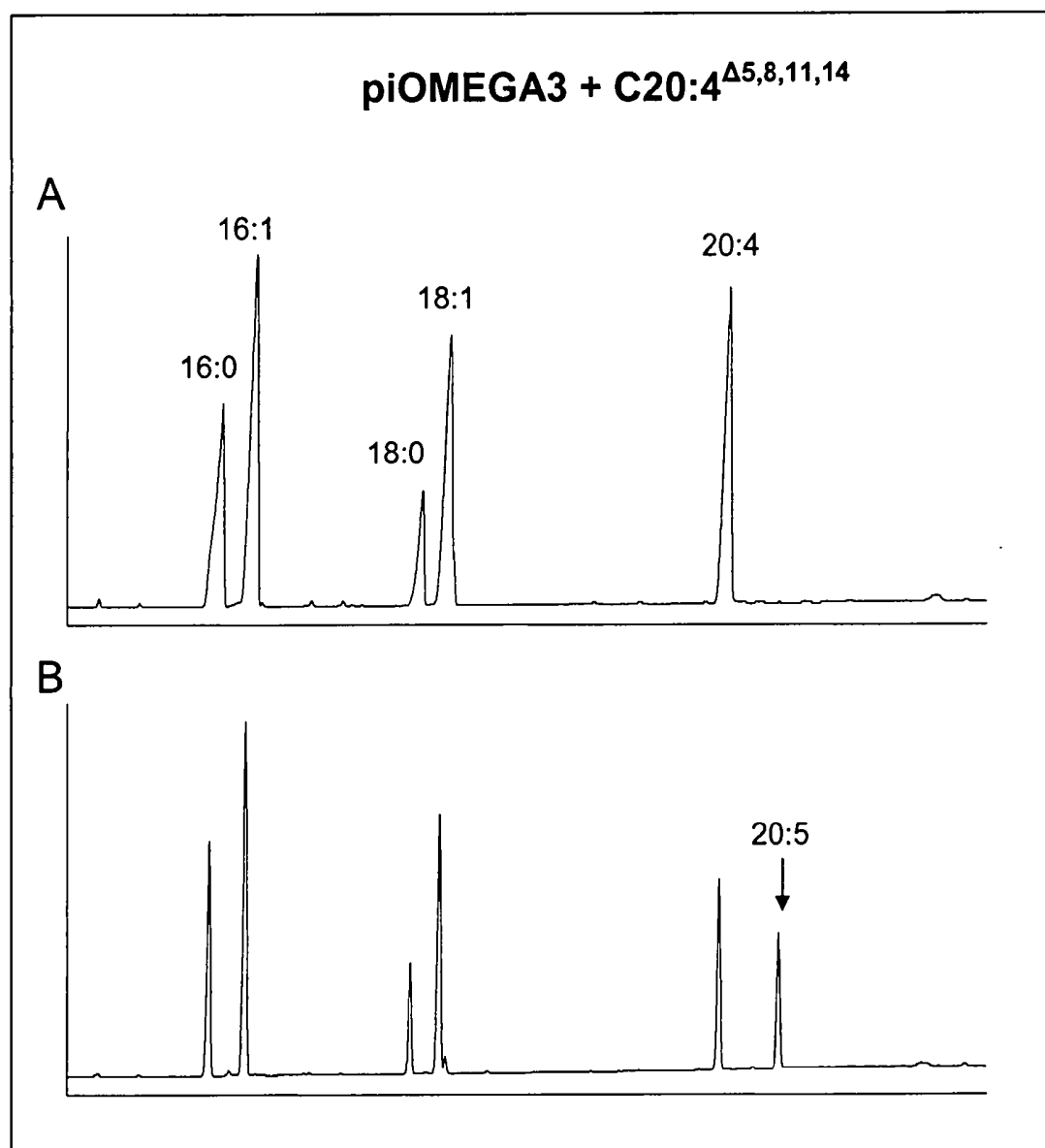


Figure 17: Desaturation of docosatetraenoic acid (C22:4 ω 6-fatty acid) to give docosapentaenoic acid (C22:5 ω 3-fatty acid) by Pi-omega3Des.

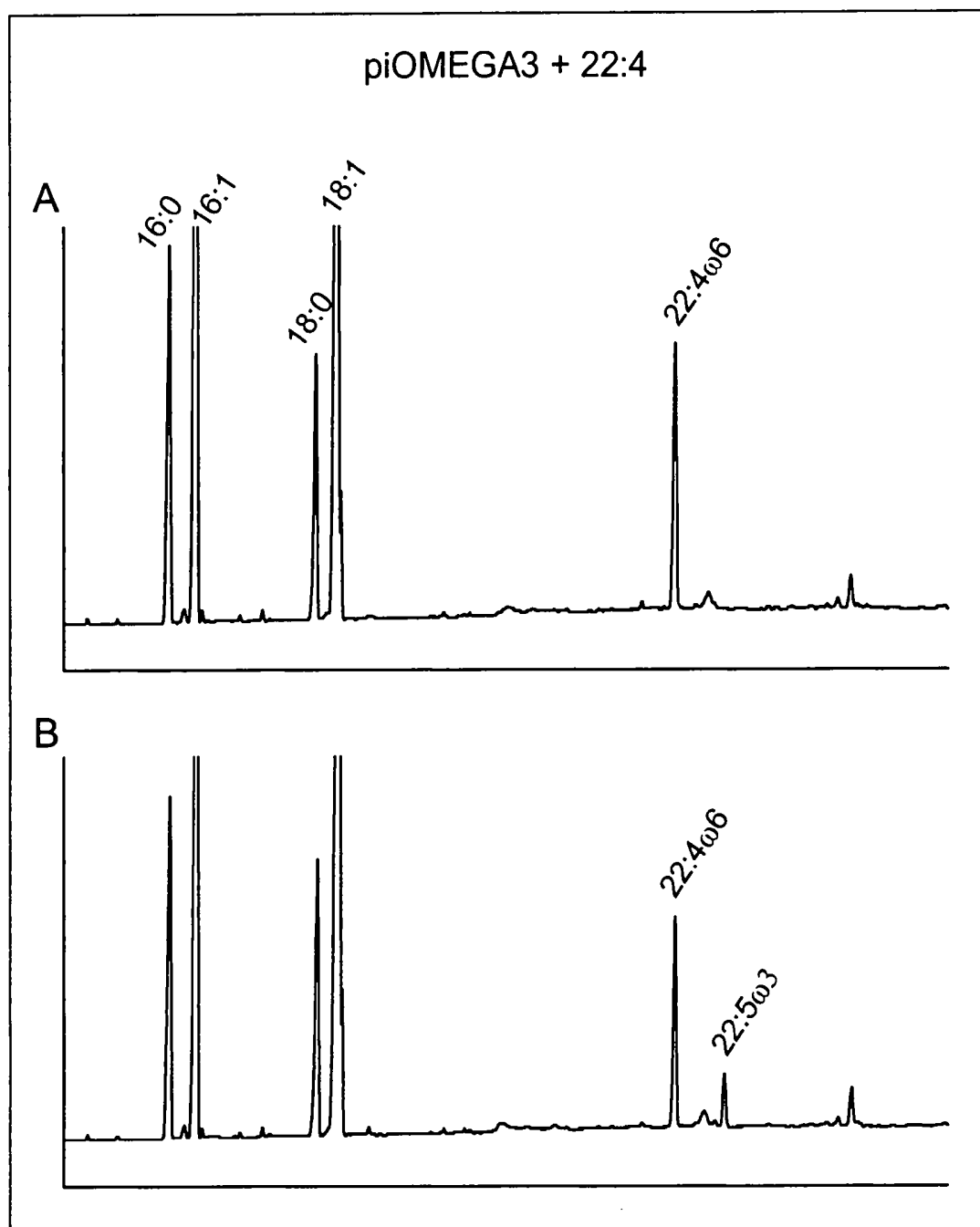


Figure 18: Substrate specificity of Pi-omega3Des with regard to different fatty acids

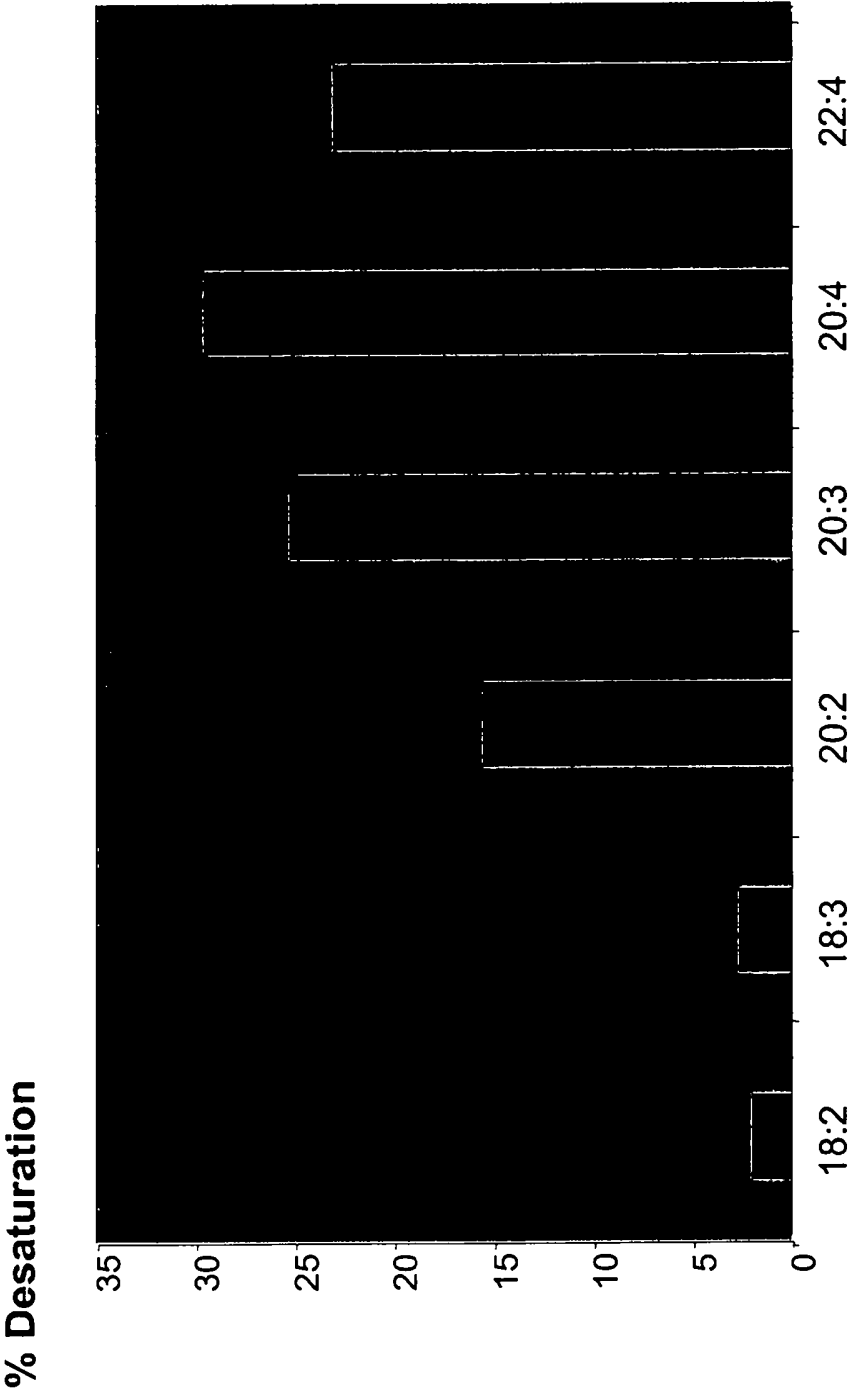


Figure 19: Desaturation of phospholipid-bound arachidonic acid to give EPA by Pi-Omega3Des

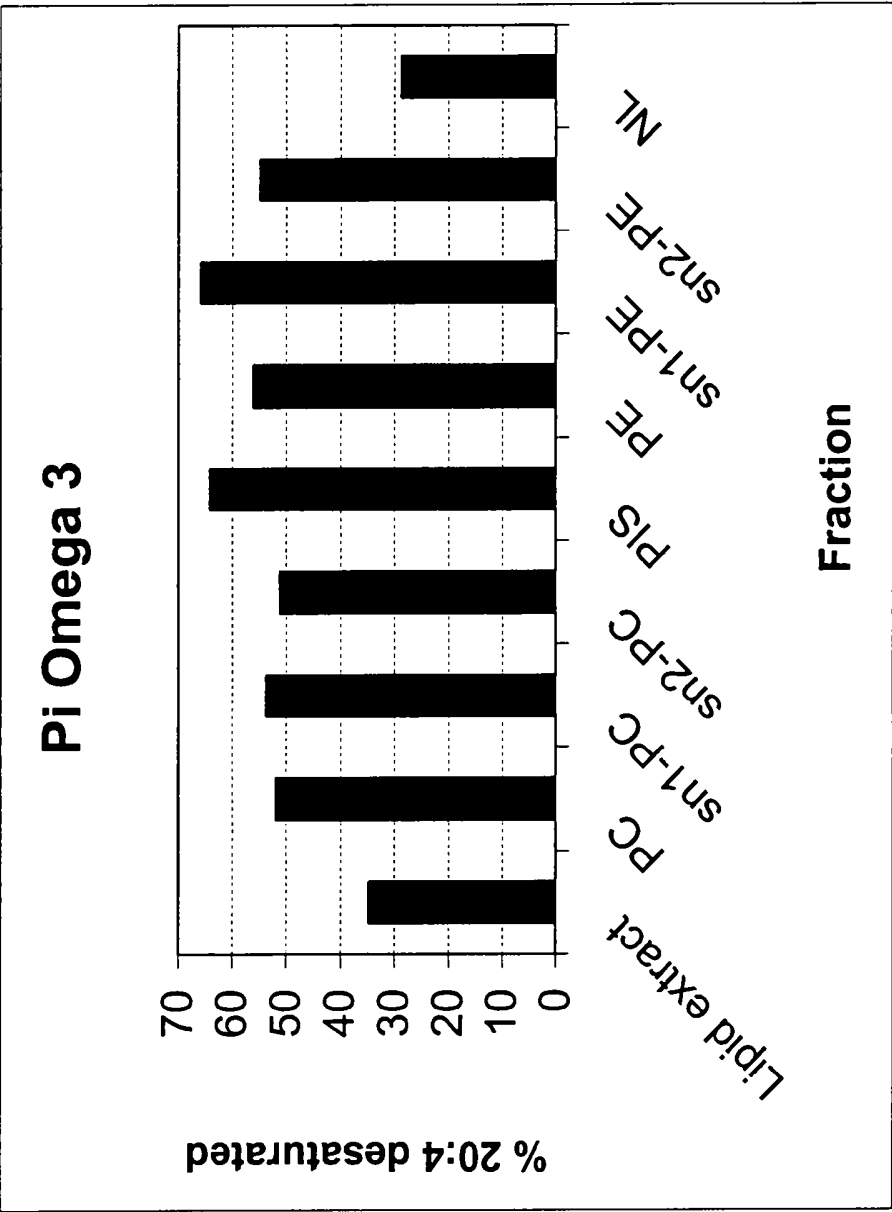
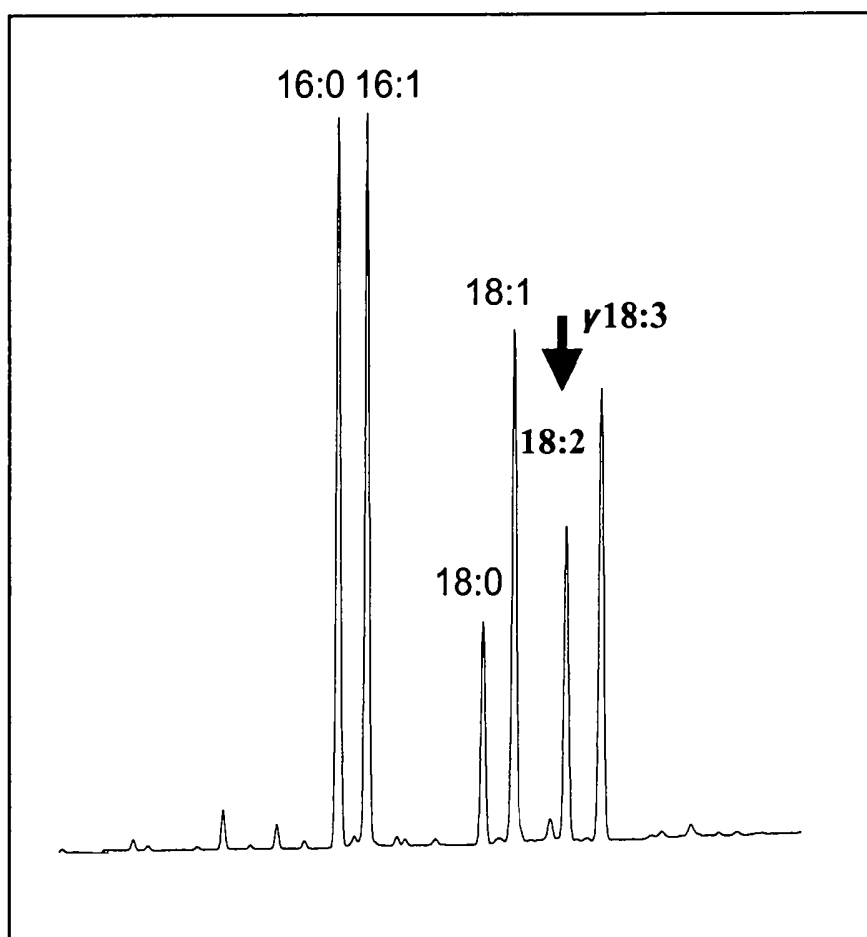


Figure 20: Conversion of linoleic acid (arrow) to give γ -linolenic acid (γ -18:3) by Ot-Des6.1.

Absorption mAU



Retention time

Figure 21: Conversion of linoleic acid and α -linolenic acid (A and C), and reconstitution of the ARA and EPA synthetic pathways, respectively, in yeast (B and D) in the presence of OtD6.1.

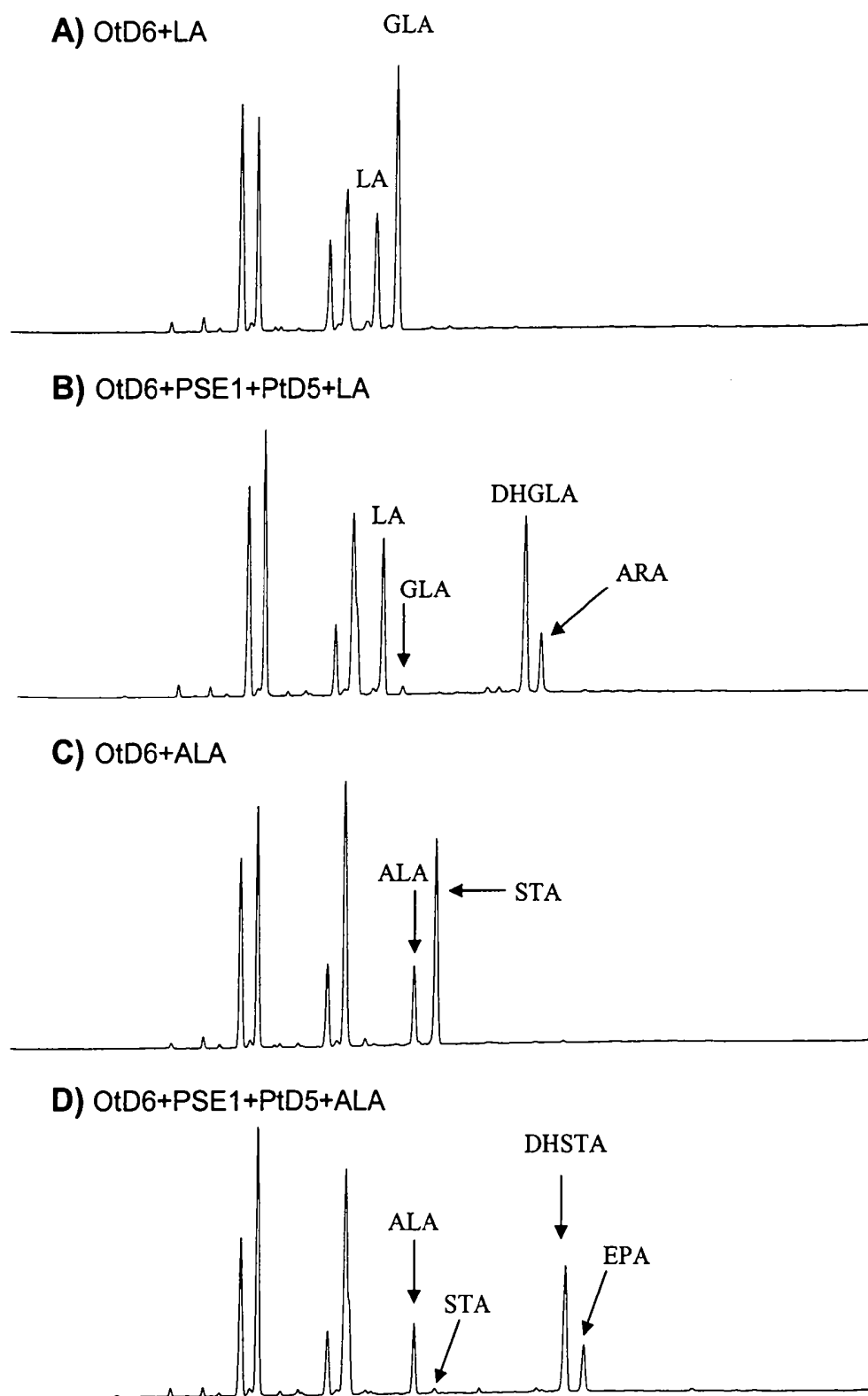


Figure 22: Expression of ELO(XI) in yeast

Absorption in mA

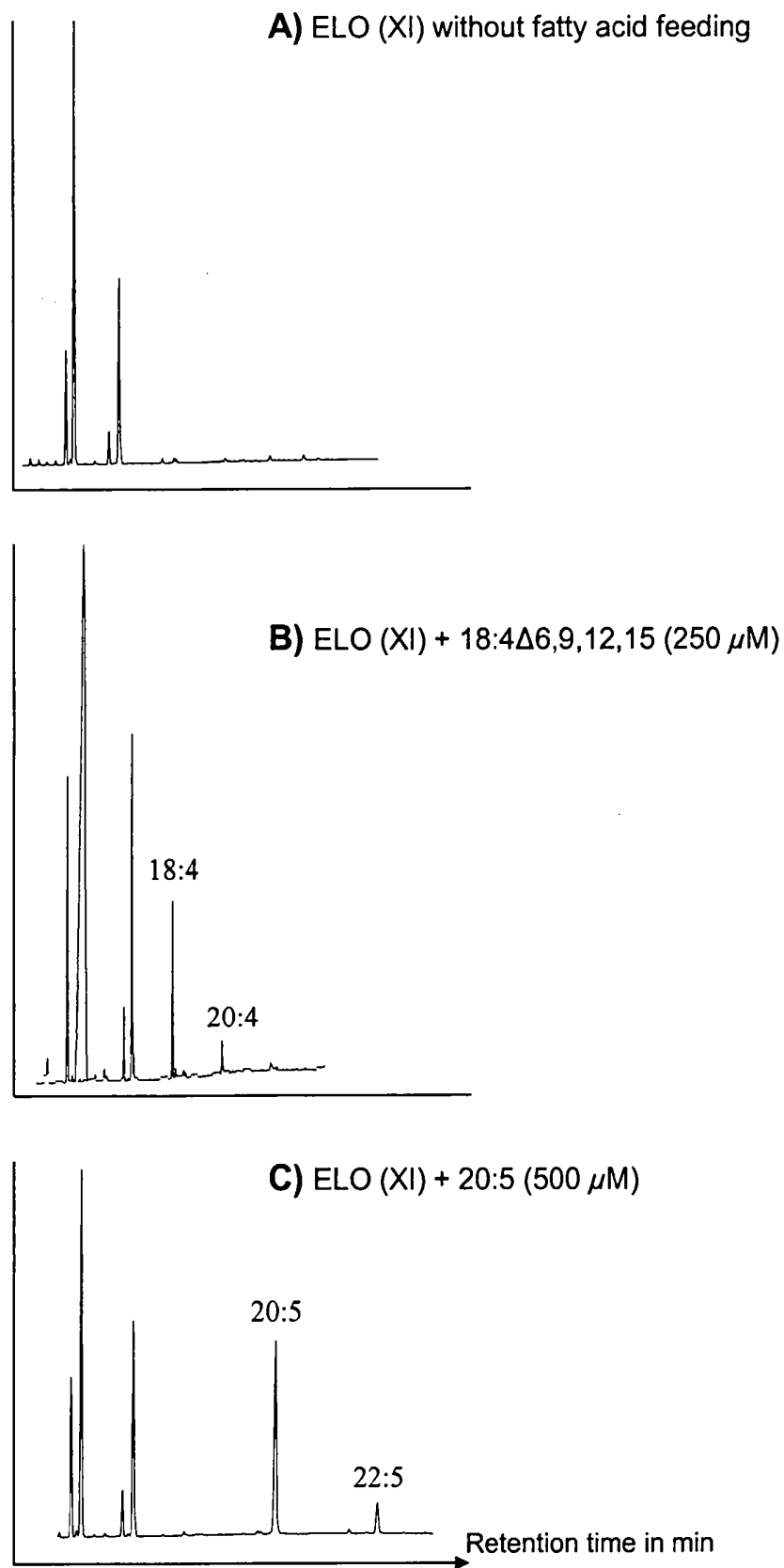


Figure 23:

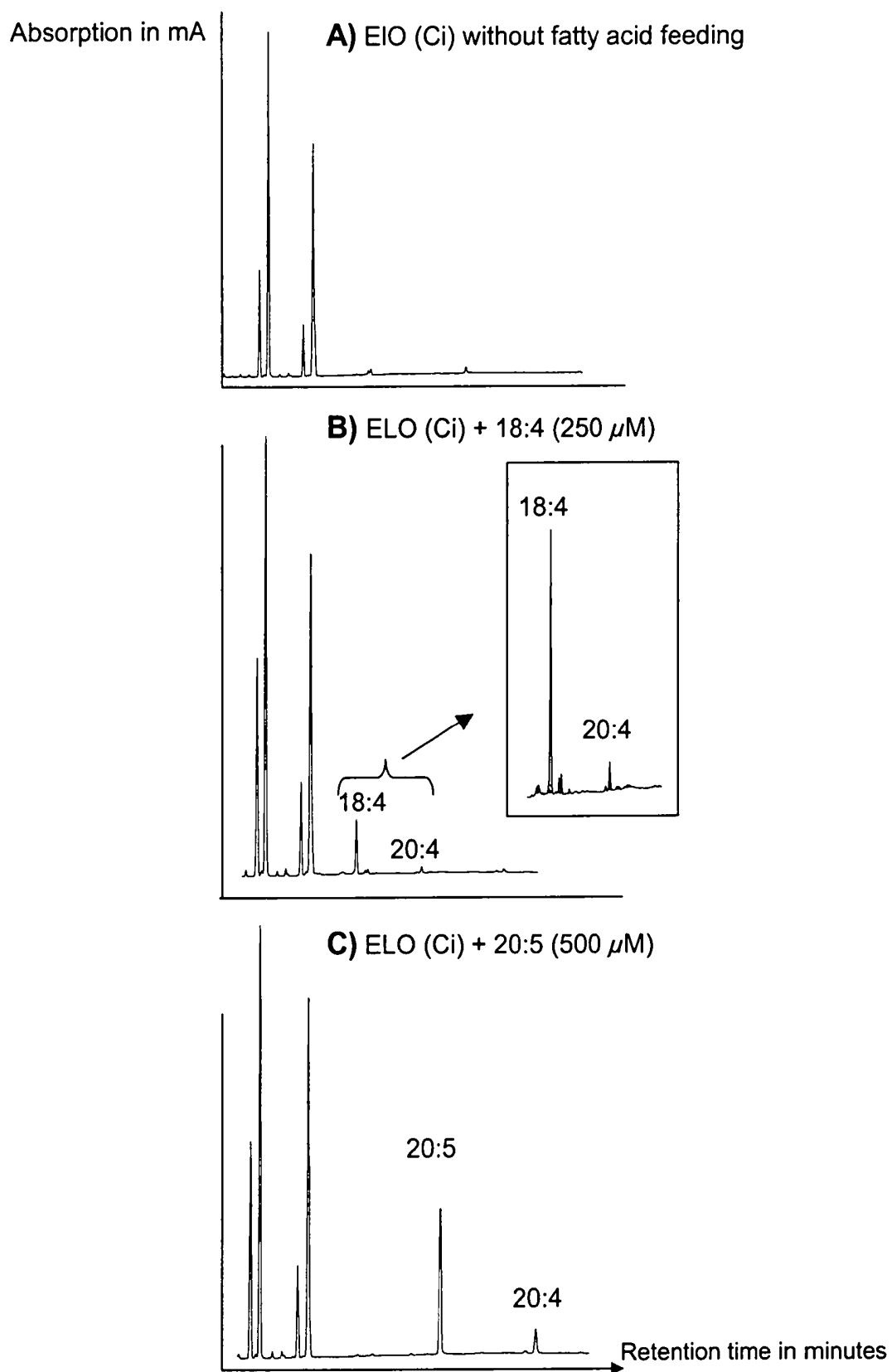


Figure 24: Elongation of eicosapentaenoic acid by OtElo1 (B) and OtElo1.2 (D), respectively. The controls (A, C) do not show the elongation product (22:5 ω 3).

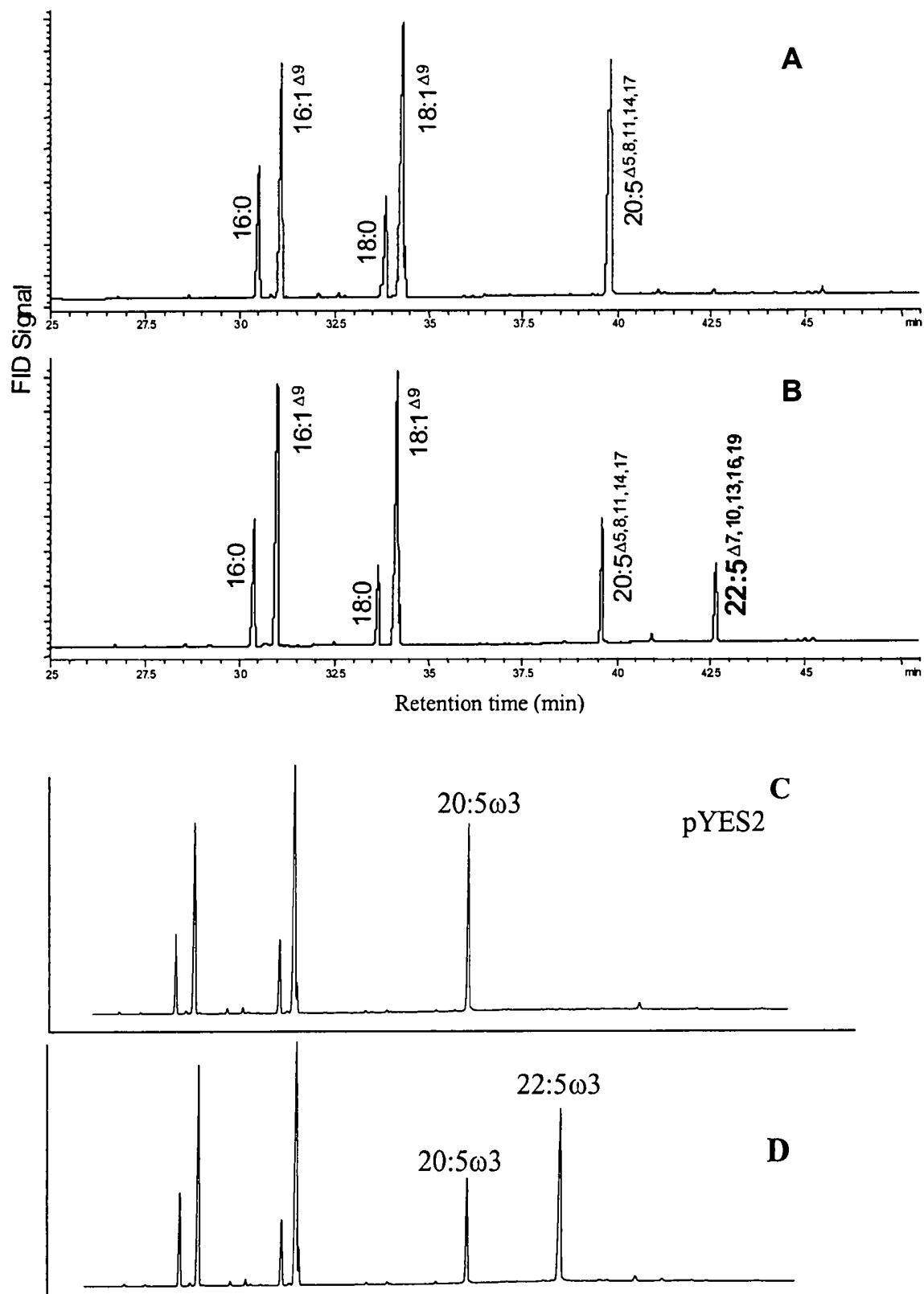


Figure 25: Elongation of arachidonic acid by OtElo1 (B) and OtElo1.2 (D), respectively. The controls (A, C) do not show the elongation product (22:4 ω 6).

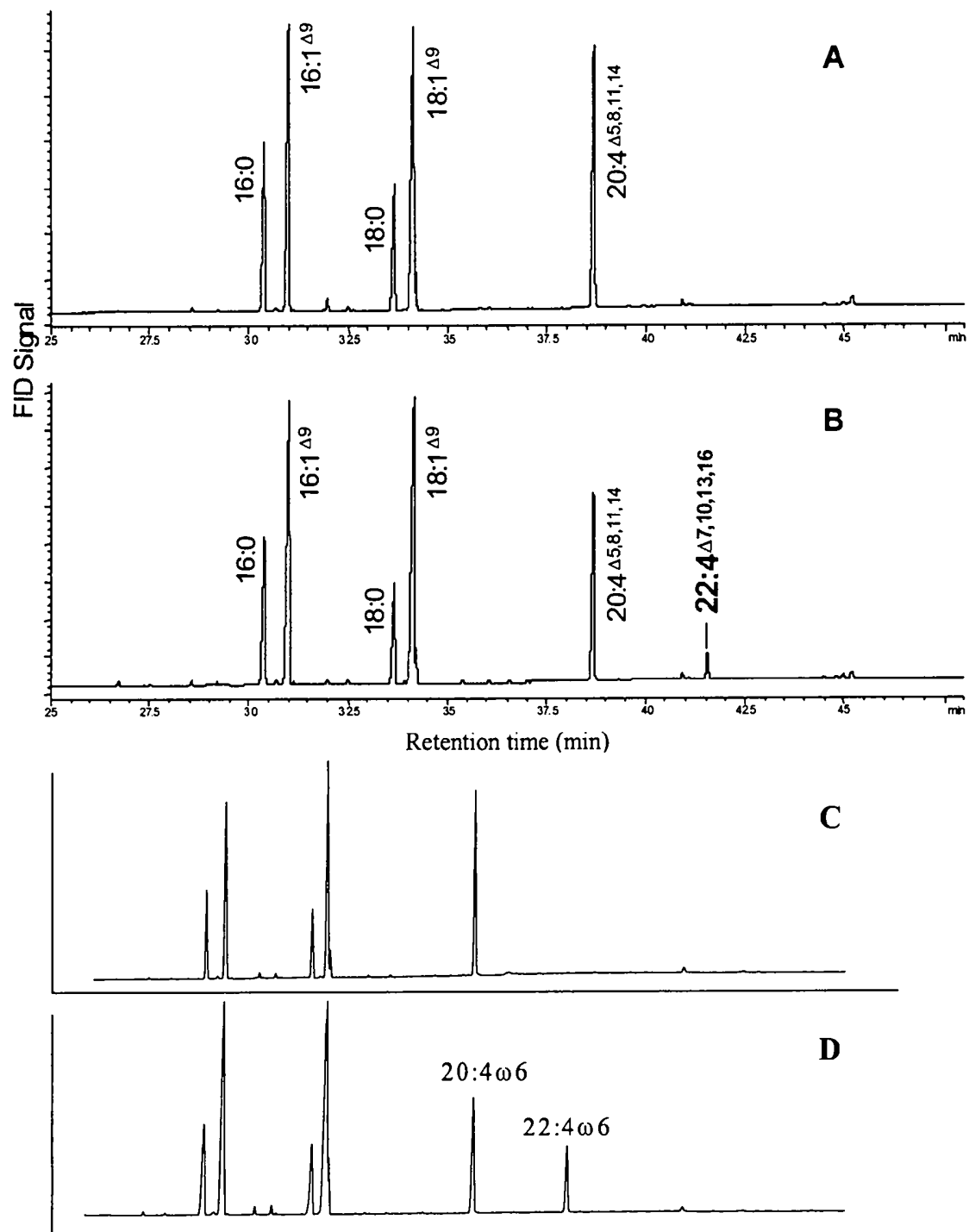


Figure 26: Elongation of 20:5n-3 by the elongases At3g06470.

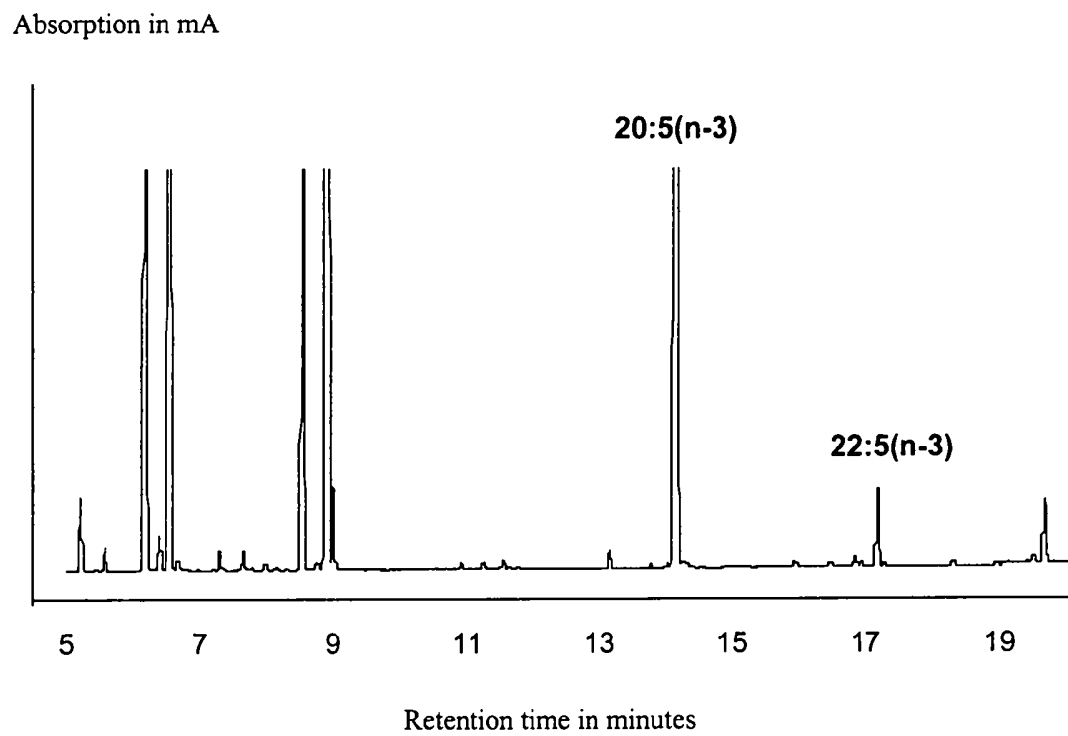


Figure 27: Substrate specificity of the *Xenopus* Elongase (A), *Ciona* Elongase (B) und *Oncorhynchus* Elongase (C)

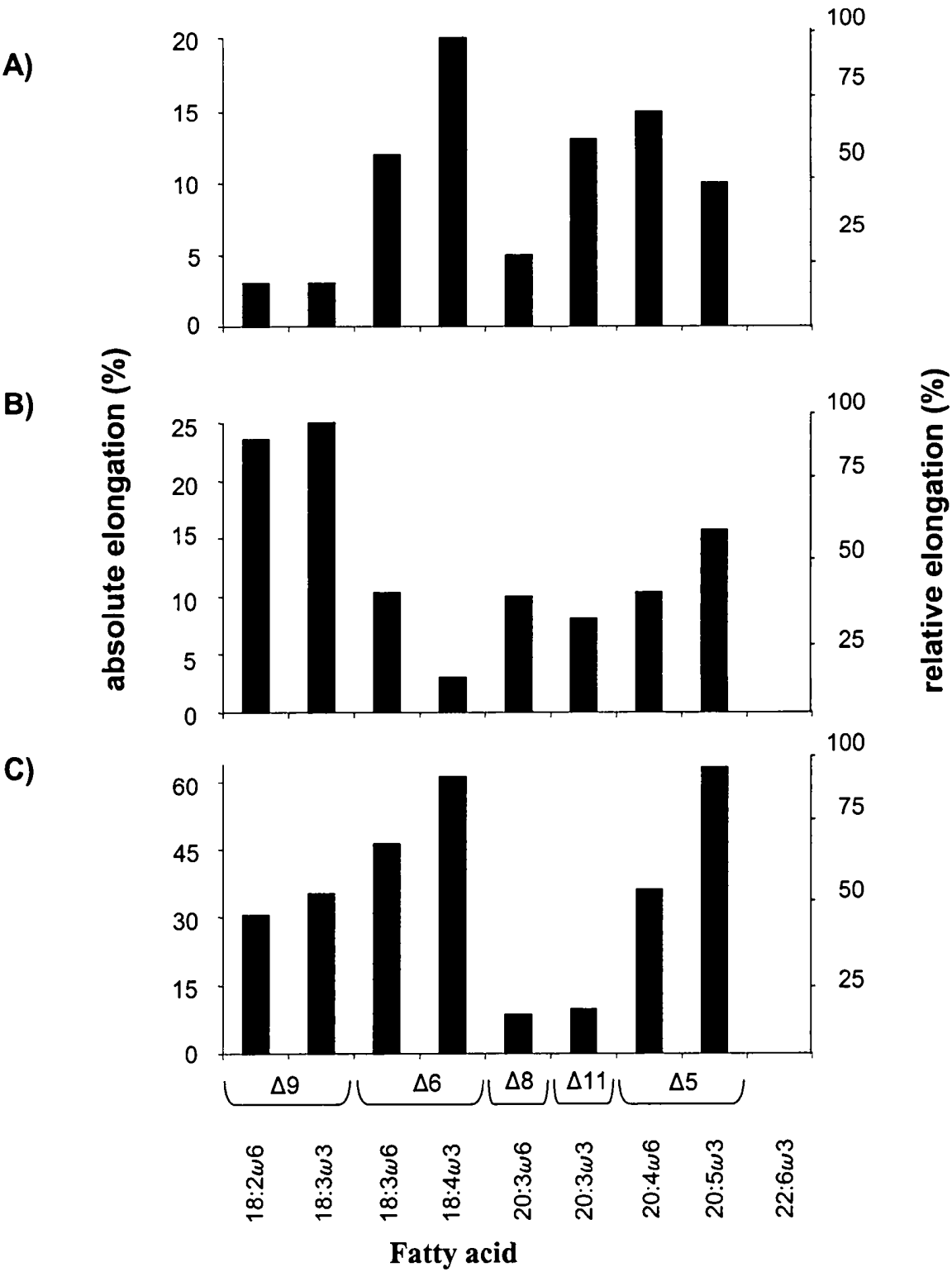


Figure 28: Substrate specificity of the *Ostreococcus* $\Delta 5$ -elongase (A), the *Ostreococcus* $\Delta 6$ -elongase (B), the *Thalassiosira* $\Delta 5$ -elongase (C) and the *Thalassiosira* *Ostreococcus* $\Delta 6$ -elongase (D)

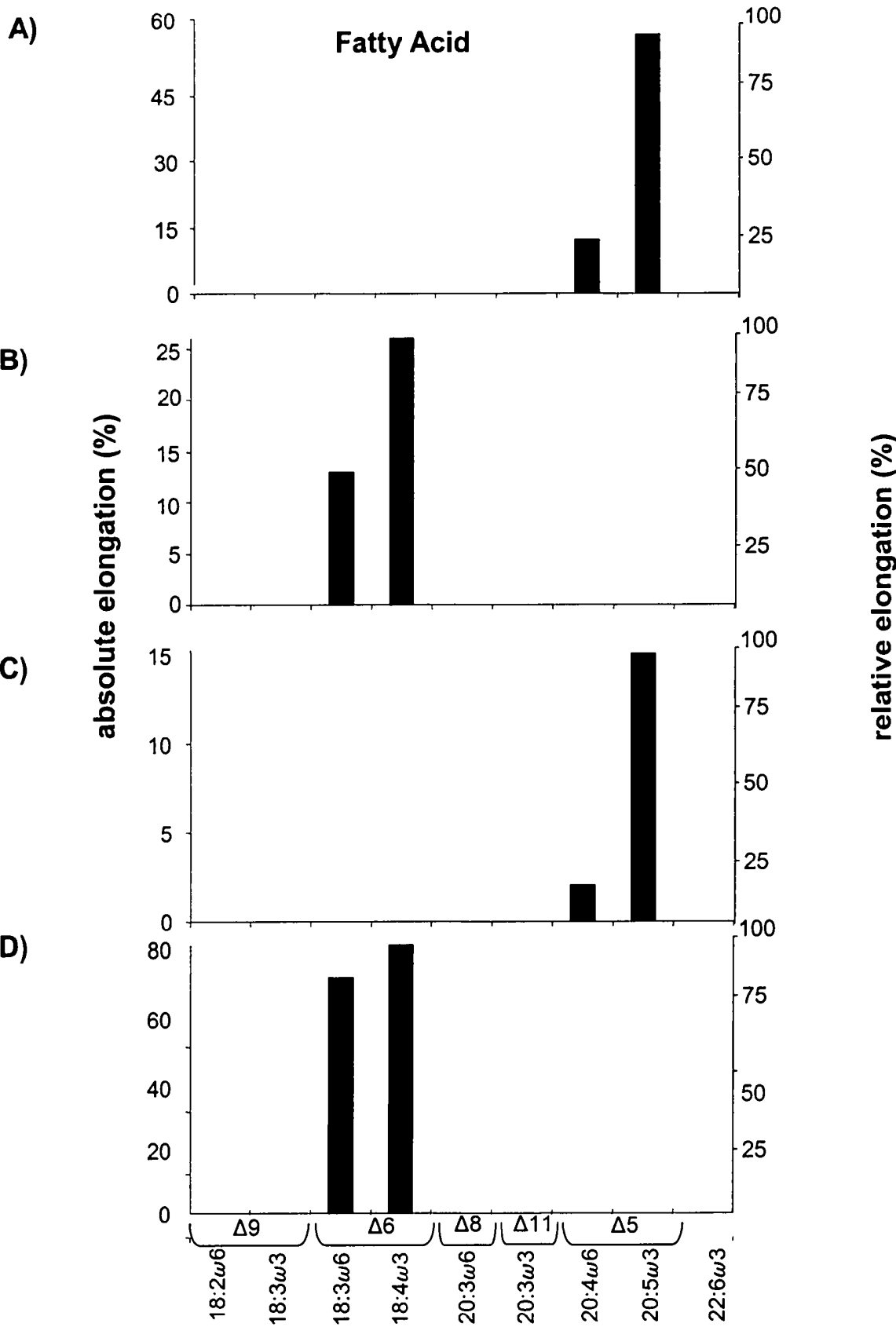
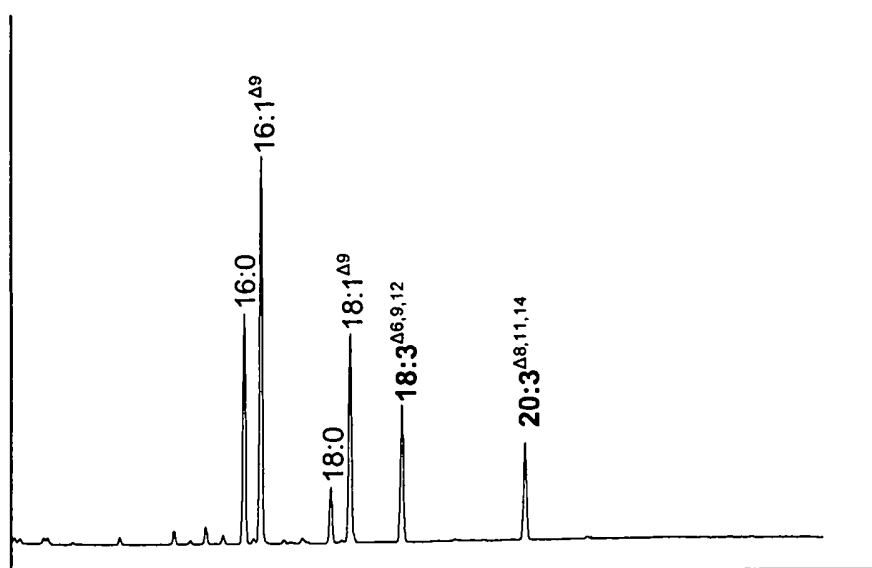


Figure 29: Expression of the *Phaeodactylum tricornutum* $\Delta 6$ -elongase (PtELO6) in yeast. A) shows the elongation of the C18:3 $^{\Delta 6,9,12}$ fatty acid and B) the elongation of the C18:3 $^{\Delta 6,9,12,15}$ fatty acid

A)



B)

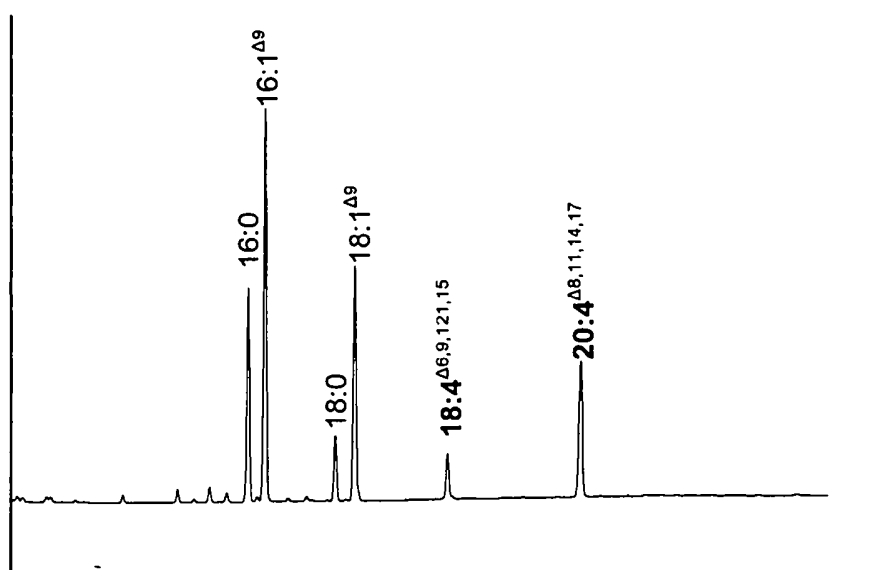


Figure 30: Figure 30 shows the substrate specificity of PtELO6 with regard to the substrates fed.

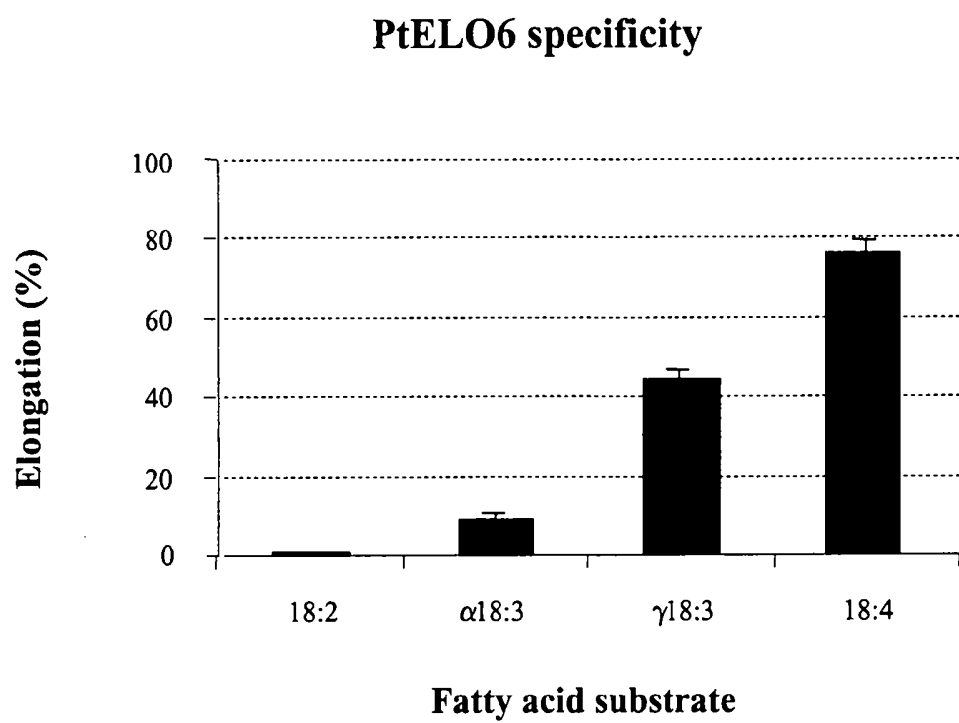


Figure 31: Gas-chromatographic analysis of the seed of a transgenic plant, transformed with pSUN-5G.

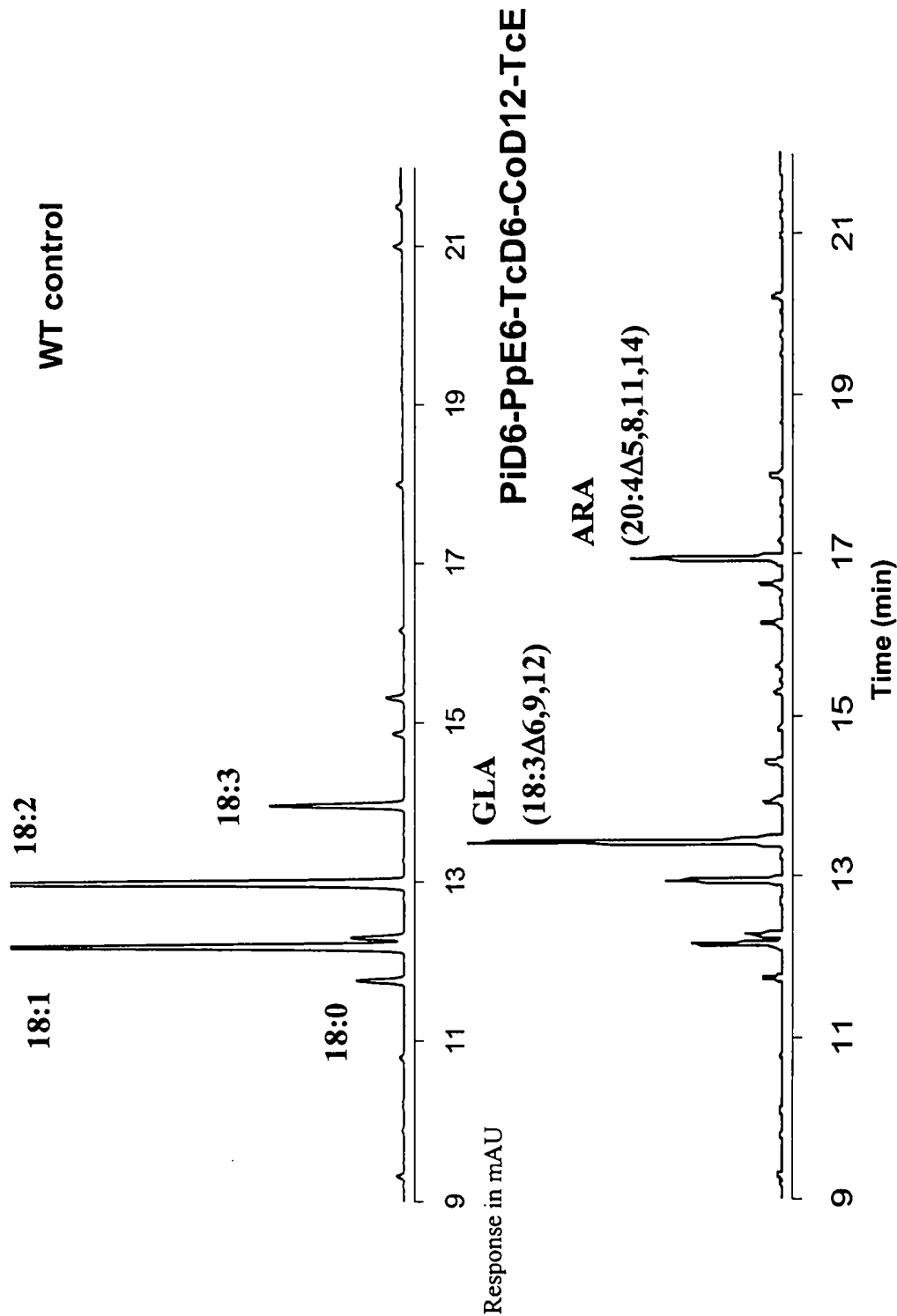


Figure 32: Gas-chromatographic analysis of the seed of a transgenic plant, transformed with pGPTV-D6Des(Pir)_D5Des(Tc)_D6Elo(PP)_12Des(Co)

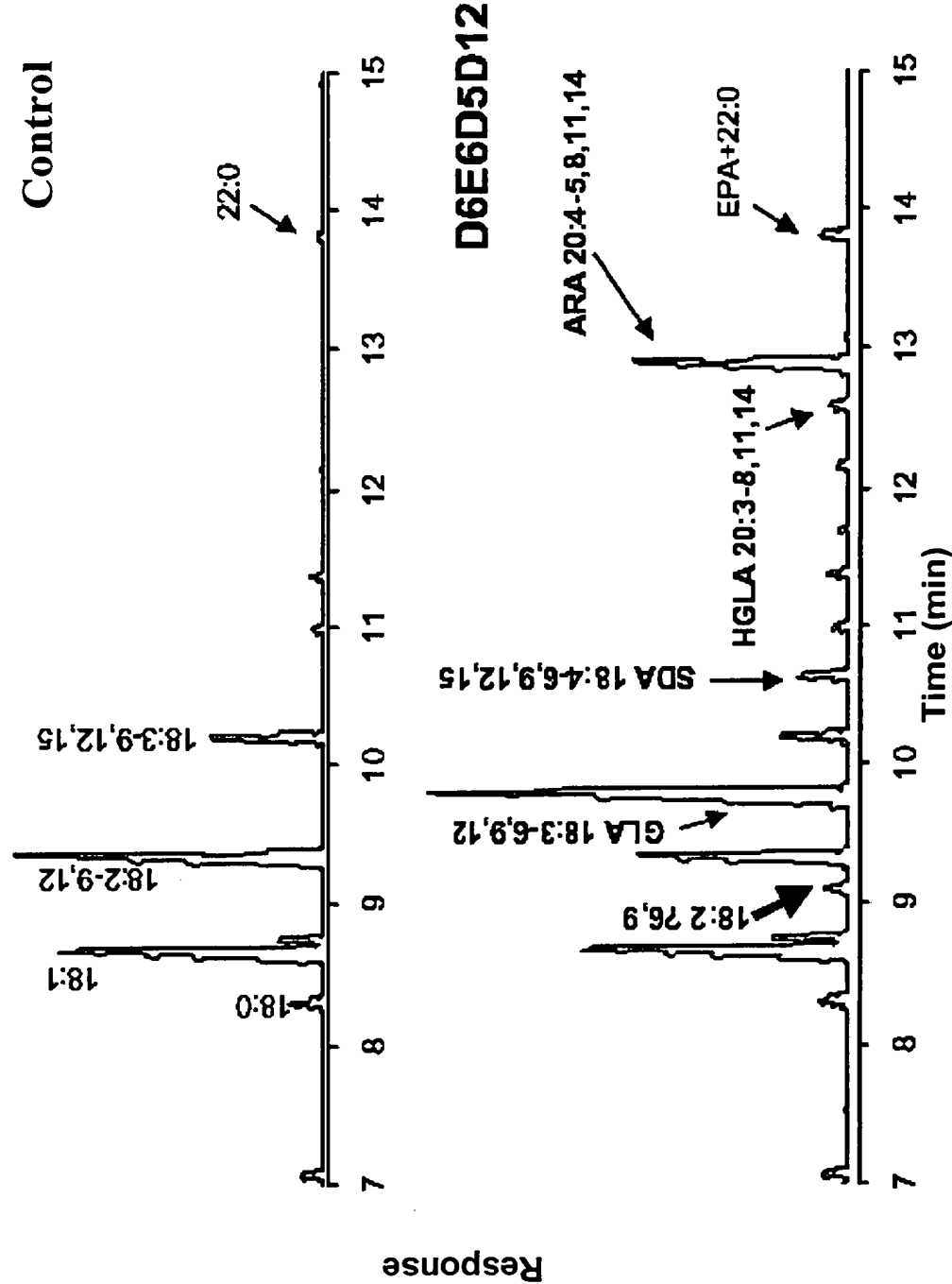
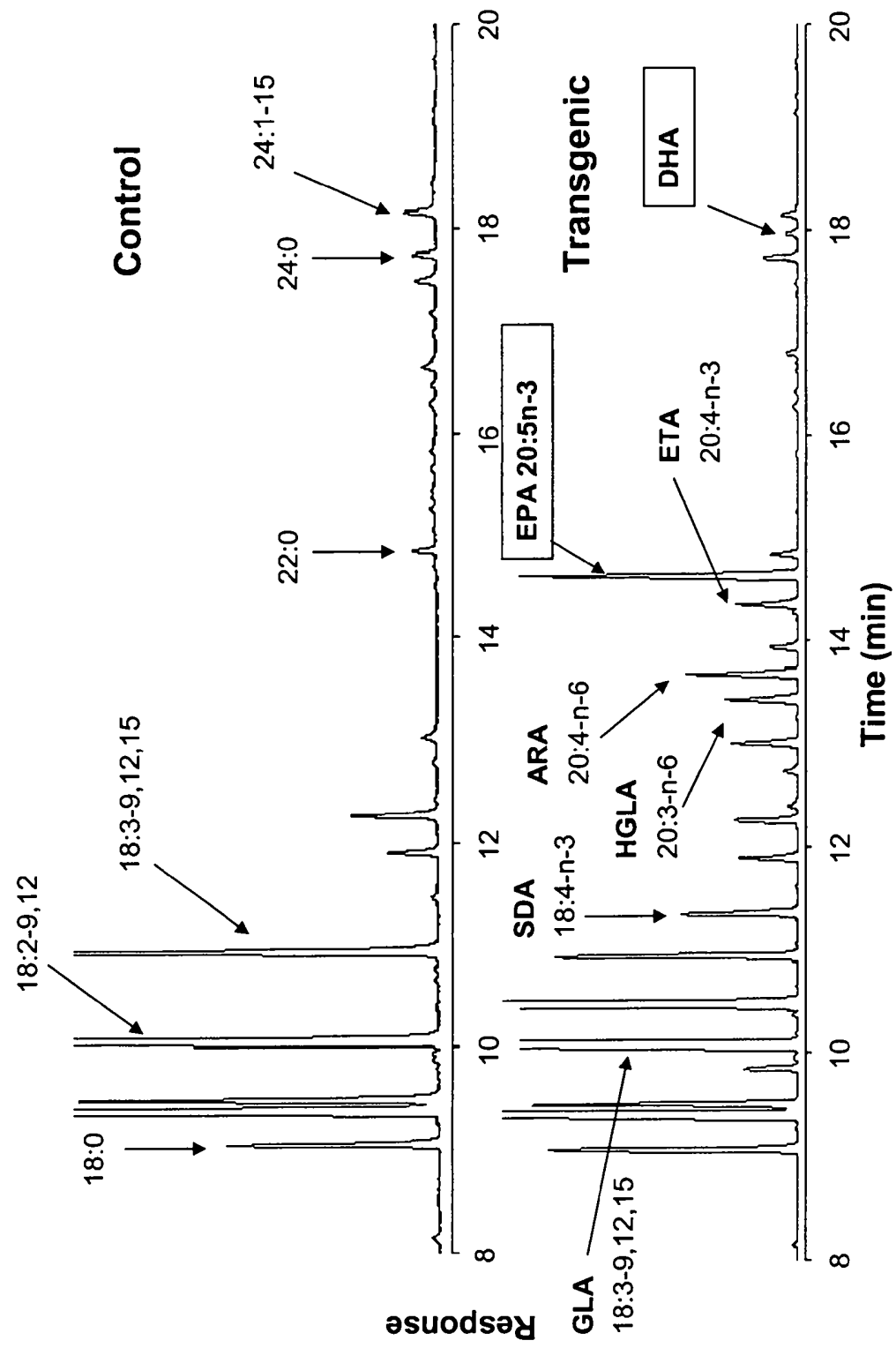


Figure 33: DHA in transgenic seeds of Brassica juncea. The plants were transformed with the construct pSUN-8G.



SEQUENCE LISTING

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<130> PF56186

<140> 20041035

<141> 2004-12-22

<160> 202

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<213> Isochrysis galbana

<400> 4

Met Ala Leu Ala Asn Asp Ala Gly Glu Arg Ile Trp Ala Ala Val Thr
 1 5 10 15

Asp Pro Glu Ile Leu Ile Gly Thr Phe Ser Tyr Leu Leu Leu Lys Pro
 20 25 30

Leu Leu Arg Asn Ser Gly Leu Val Asp Glu Lys Lys Gly Ala Tyr Arg

35

40

45

Thr Ser Met Ile Trp Tyr Asn Val Leu Leu Ala Leu Phe Ser Ala Leu
 50 55 60

Ser Phe Tyr Val Thr Ala Thr Ala Leu Gly Trp Asp Tyr Gly Thr Gly
 65 70 75 80

Ala Trp Leu Arg Arg Gln Thr Gly Asp Thr Pro Gln Pro Leu Phe Gln
 85 90 95

Cys Pro Ser Pro Val Trp Asp Ser Lys Leu Phe Thr Trp Thr Ala Lys
 100 105 110

Ala Phe Tyr Tyr Ser Lys Tyr Val Glu Tyr Leu Asp Thr Ala Trp Leu
 115 120 125

Arg Val Ser Phe Leu Gln Ala Phe His His Phe Gly Ala Pro Trp Asp
 130 135 140

Val Tyr Leu Gly Ile Arg Leu His Asn Glu Gly Val Trp Ile Phe Met
 145 150 155 160

Phe Phe Asn Ser Phe Ile His Thr Ile Met Tyr Thr Tyr Tyr Gly Leu
 165 170 175

Thr Ala Ala Gly Tyr Lys Phe Lys Ala Lys Pro Leu Ile Thr Ala Met
 180 185 190

Gln Ile Cys Gln Phe Val Gly Gly Phe Leu Leu Val Trp Asp Tyr Ile
 195 200 205

Asn Val Pro Cys Phe Asn Ser Asp Lys Gly Lys Leu Phe Ser Trp Ala
 210 215 220

Phe Asn Tyr Ala Tyr Val Gly Ser Val Phe Leu Leu Phe Cys His Phe
 225 230 235 240

Phe Tyr Gln Asp Asn Leu Ala Thr Lys Lys Ser Ala Lys Ala Gly Lys
 245 250 255

Gln Leu

<210> 5

<211> 1410

<212> DNA

<213> *Phaeodactylum tricornutum*

<220>

<221> CDS

<222> (1)..(1410)

<223> Delta-5 desaturase

<400> 5

atg gct ccg gat gcg gat aag ctt cga caa cgc cag acg act gcg gta	48
Met Ala Pro Asp Ala Asp Lys Leu Arg Gln Arg Gln Thr Thr Ala Val	
1 5 10 15	

gcg aag cac aat gct gct acc ata tcg acg cag gaa cgc ctt tgc agt	96
Ala Lys His Asn Ala Ala Thr Ile Ser Thr Gln Glu Arg Leu Cys Ser	
20 25 30	

ctg tct tcg ctc aaa ggc gaa gaa gtc tgc atc gac gga atc atc tat	144
Leu Ser Ser Leu Lys Gly Glu Glu Val Cys Ile Asp Gly Ile Ile Tyr	
35 40 45	

gac ctc caa tca ttc gat cat ccc ggg ggt gaa acg atc aaa atg ttt	192
Asp Leu Gln Ser Phe Asp His Pro Gly Gly Glu Thr Ile Lys Met Phe	
50 55 60	

ggg ggc aac gat gtc act gta cag tac aag atg att cac ccg tac cat	240
Gly Gly Asn Asp Val Thr Val Gln Tyr Lys Met Ile His Pro Tyr His	
65 70 75 80	

acc gag aag cat ttg gaa aag atg aag cgt gtc ggc aag gtg acg gat	288
Thr Glu Lys His Leu Glu Lys Met Lys Arg Val Gly Lys Val Thr Asp	
85 90 95	

ttc gtc tgc gag tac aag ttc gat acc gaa ttt gaa cgc gaa atc aaa	336
Phe Val Cys Glu Tyr Lys Phe Asp Thr Glu Phe Glu Arg Glu Ile Lys	
100 105 110	

cga gaa gtc ttc aag att gtg cga cga ggc aag gat ttc ggt act ttg	384
Arg Glu Val Phe Lys Ile Val Arg Arg Gly Lys Asp Phe Gly Thr Leu	
115 120 125	

gga tgg ttc ttc cgt gcg ttt tgc tac att gcc att ttc ttc tac ctg	432
Gly Trp Phe Phe Arg Ala Phe Cys Tyr Ile Ala Ile Phe Phe Tyr Leu	
130 135 140	

cag tac cat tgg gtc acc acg gga acc tct tgg ctg ctg gcc gtg gcc	480
Gln Tyr His Trp Val Thr Thr Gly Thr Ser Trp Leu Leu Ala Val Ala	
145 150 155 160	

tac gga atc tcc caa gcg atg att ggc atg aat gtc cag cac gat gcc	528
Tyr Gly Ile Ser Gln Ala Met Ile Gly Met Asn Val Gln His Asp Ala	
165 170 175	

aac cac ggg gcc acc tcc aag cgt ccc tgg gtc aac gac atg cta ggc	576
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Asn	His	Gly	Ala	Thr	Ser	Lys	Arg	Pro	Trp	Val	Asn	Asp	Met	Leu	Gly	
			180					185					190			
ctc	ggt	gcg	gat	ttt	att	ggt	ggt	tcc	aag	tgg	ctc	tgg	cag	gaa	caa	624
Leu	Gly	Ala	Asp	Phe	Ile	Gly	Gly	Ser	Lys	Trp	Leu	Trp	Gln	Glu	Gln	
		195					200					205				
cac	tgg	acc	cac	cac	gct	tac	acc	aat	cac	gcc	gag	atg	gat	ccc	gat	672
His	Trp	Thr	His	His	Ala	Tyr	Thr	Asn	His	Ala	Glu	Met	Asp	Pro	Asp	
	210					215					220					
agc	ttt	ggt	gcc	gaa	cca	atg	ctc	cta	ttc	aac	gac	tat	ccc	ttg	gat	720
Ser	Phe	Gly	Ala	Glu	Pro	Met	Leu	Leu	Phe	Asn	Asp	Tyr	Pro	Leu	Asp	
225					230					235					240	
cat	ccc	gct	cgt	acc	tgg	cta	cat	cgc	ttt	caa	gca	ttc	ttt	tac	atg	768
His	Pro	Ala	Arg	Thr	Trp	Leu	His	Arg	Phe	Gln	Ala	Phe	Phe	Tyr	Met	
				245					250					255		
ccc	gtc	ttg	gct	gga	tac	tgg	ttg	tcc	gct	gtc	ttc	aat	cca	caa	att	816
Pro	Val	Leu	Ala	Gly	Tyr	Trp	Leu	Ser	Ala	Val	Phe	Asn	Pro	Gln	Ile	
			260					265					270			
ctt	gac	ctc	cag	caa	cgc	ggc	gca	ctt	tcc	gtc	ggt	atc	cgt	ctc	gac	864
Leu	Asp	Leu	Gln	Gln	Arg	Gly	Ala	Leu	Ser	Val	Gly	Ile	Arg	Leu	Asp	
		275					280					285				
aac	gct	ttc	att	cac	tcg	cga	cgc	aag	tat	gcg	gtt	ttc	tgg	cgg	gct	912
Asn	Ala	Phe	Ile	His	Ser	Arg	Arg	Lys	Tyr	Ala	Val	Phe	Trp	Arg	Ala	
	290						295						300			
gtg	tac	att	gcg	gtg	aac	gtg	att	gct	ccg	ttt	tac	aca	aac	tcc	ggc	960
Val	Tyr	Ile	Ala	Val	Asn	Val	Ile	Ala	Pro	Phe	Tyr	Thr	Asn	Ser	Gly	
305					310					315					320	
ctc	gaa	tgg	tcc	tgg	cgt	gtc	ttt	gga	aac	atc	atg	ctc	atg	ggt	gtg	1008
Leu	Glu	Trp	Ser	Trp	Arg	Val	Phe	Gly	Asn	Ile	Met	Leu	Met	Gly	Val	
				325				330						335		
gcg	gaa	tcg	ctc	gcg	ctg	gcg	gtc	ctg	ttt	tcg	ttg	tcg	cac	aat	ttc	1056
Ala	Glu	Ser	Leu	Ala	Leu	Ala	Val	Leu	Phe	Ser	Leu	Ser	His	Asn	Phe	
			340					345					350			
gaa	tcc	gcg	gat	cgc	gat	ccg	acc	gcc	cca	ctg	aaa	aag	acg	gga	gaa	1104
Glu	Ser	Ala	Asp	Arg	Asp	Pro	Thr	Ala	Pro	Leu	Lys	Lys	Thr	Gly	Glu	
		355					360					365				
cca	gtc	gac	tgg	ttc	aag	aca	cag	gtc	gaa	act	tcc	tgc	act	tac	ggt	1152
Pro	Val	Asp	Trp	Phe	Lys	Thr	Gln	Val	Glu	Thr	Ser	Cys	Thr	Tyr	Gly	
	370					375					380					
gga	ttc	ctt	tcc	ggt	tgc	ttc	acg	gga	ggt	ctc	aac	ttt	cag	gtt	gaa	1200
Gly	Phe	Leu	Ser	Gly	Cys	Phe	Thr	Gly	Gly	Leu	Asn	Phe	Gln	Val	Glu	
385					390					395					400	
cac	cac	ttg	ttc	cca	cgc	atg	agc	agc	gct	tgg	tat	ccc	tac	att	gcc	1248
His	His	Leu	Phe	Pro	Arg	Met	Ser	Ser	Ala	Trp	Tyr	Pro	Tyr	Ile	Ala	
				405					410					415		
ccc	aag	gtc	cgc	gaa	att	tgc	gcc	aaa	cac	ggc	gtc	cac	tac	gcc	tac	1296
Pro	Lys	Val	Arg	Glu	Ile	Cys	Ala	Lys	His	Gly	Val	His	Tyr	Ala	Tyr	
			420					425					430			

tac ccg tgg atc cac caa aac ttt ctc tcc acc gtc cgc tac atg cac 1344
 Tyr Pro Trp Ile His Gln Asn Phe Leu Ser Thr Val Arg Tyr Met His
 435 440 445

gcg gcc ggg acc ggt gcc aac tgg cgc cag atg gcc aga gaa aat ccc 1392
Ala Ala Gly Thr Gly Ala Asn Trp Arg Gln Met Ala Arg Glu Asn Pro
450 455 460

ttg acc gga cgg gcg taa 1410
Leu Thr Gly Arg Ala
465

<210> 6

<211> 469

<212> PRT

<213> Phaeodactylum tricornutum

<400> 6

Met Ala Pro Asp Ala Asp Lys Leu Arg Gln Arg Gln Thr Thr Ala Val
1 5 10 15

Ala Lys His Asn Ala Ala Thr Ile Ser Thr Gln Glu Arg Leu Cys Ser
20 25 30

Leu Ser Ser Leu Lys Gly Glu Glu Val Cys Ile Asp Gly Ile Ile Tyr
35 40 45

Asp Leu Gln Ser Phe Asp His Pro Gly Gly Glu Thr Ile Lys Met Phe
50 55 60

Gly Gly Asn Asp Val Thr Val Gln Tyr Lys Met Ile His Pro Tyr His
65 70 75 80

Thr Glu Lys His Leu Glu Lys Met Lys Arg Val Gly Lys Val Thr Asp
85 90 95

Phe Val Cys Glu Tyr Lys Phe Asp Thr Glu Phe Glu Arg Glu Ile Lys
100 105 110

Arg Glu Val Phe Lys Ile Val Arg Arg Gly Lys Asp Phe Gly Thr Leu
115 120 125

Gly Trp Phe Phe Arg Ala Phe Cys Tyr Ile Ala Ile Phe Phe Tyr Leu
130 135 140

Gln Tyr His Trp Val Thr Thr Gly Thr Ser Trp Leu Leu Ala Val Ala
145 150 155 160

Tyr Gly Ile Ser Gln Ala Met Ile Gly Met Asn Val Gln His Asp Ala
 165 170 175

Asn His Gly Ala Thr Ser Lys Arg Pro Trp Val Asn Asp Met Leu Gly
 180 185 190

Leu Gly Ala Asp Phe Ile Gly Gly Ser Lys Trp Leu Trp Gln Glu Gln
 195 200 205

His Trp Thr His His Ala Tyr Thr Asn His Ala Glu Met Asp Pro Asp
 210 215 220

Ser Phe Gly Ala Glu Pro Met Leu Leu Phe Asn Asp Tyr Pro Leu Asp
 225 230 235 240

His Pro Ala Arg Thr Trp Leu His Arg Phe Gln Ala Phe Phe Tyr Met
 245 250 255

Pro Val Leu Ala Gly Tyr Trp Leu Ser Ala Val Phe Asn Pro Gln Ile
 260 265 270

Leu Asp Leu Gln Gln Arg Gly Ala Leu Ser Val Gly Ile Arg Leu Asp
 275 280 285

Asn Ala Phe Ile His Ser Arg Arg Lys Tyr Ala Val Phe Trp Arg Ala
 290 295 300

Val Tyr Ile Ala Val Asn Val Ile Ala Pro Phe Tyr Thr Asn Ser Gly
 305 310 315 320

Leu Glu Trp Ser Trp Arg Val Phe Gly Asn Ile Met Leu Met Gly Val
 325 330 335

Ala Glu Ser Leu Ala Leu Ala Val Leu Phe Ser Leu Ser His Asn Phe
 340 345 350

Glu Ser Ala Asp Arg Asp Pro Thr Ala Pro Leu Lys Lys Thr Gly Glu
 355 360 365

Pro Val Asp Trp Phe Lys Thr Gln Val Glu Thr Ser Cys Thr Tyr Gly
 370 375 380

Gly Phe Leu Ser Gly Cys Phe Thr Gly Gly Leu Asn Phe Gln Val Glu
 385 390 395 400

His His Leu Phe Pro Arg Met Ser Ser Ala Trp Tyr Pro Tyr Ile Ala
 405 410 415

Pro Lys Val Arg Glu Ile Cys Ala Lys His Gly Val His Tyr Ala Tyr
 420 425 430

Tyr Pro Trp Ile His Gln Asn Phe Leu Ser Thr Val Arg Tyr Met His
 435 440 445

Ala Ala Gly Thr Gly Ala Asn Trp Arg Gln Met Ala Arg Glu Asn Pro
 450 455 460

Leu Thr Gly Arg Ala
 465

<210> 7

<211> 1344

<212> DNA

<213> Ceratodon purpureus

<220>

<221> CDS

<222> (1)..(1344)

<223> Delta-5 desaturase

<400> 7

atg gta tta cga gag caa gag cat gag cca ttc ttc att aaa att gat 48
 Met Val Leu Arg Glu Gln Glu His Glu Pro Phe Phe Ile Lys Ile Asp
 1 5 10 15

gga aaa tgg tgt caa att gac gat gct gtc ctg aga tca cat cca ggt 96
 Gly Lys Trp Cys Gln Ile Asp Asp Ala Val Leu Arg Ser His Pro Gly
 20 25 30

ggg agt gca att act acc tat aaa aat atg gat gcc act acc gta ttc 144
 Gly Ser Ala Ile Thr Thr Tyr Lys Asn Met Asp Ala Thr Thr Val Phe
 35 40 45

cac aca ttc cat act ggt tct aaa gaa gcg tat caa tgg ctg aca gaa 192
 His Thr Phe His Thr Gly Ser Lys Glu Ala Tyr Gln Trp Leu Thr Glu
 50 55 60

ttg aaa aaa gag tgc cct aca caa gaa cca gag atc cca gat att aag 240
 Leu Lys Lys Glu Cys Pro Thr Gln Glu Pro Glu Ile Pro Asp Ile Lys
 65 70 75 80

gat gac cca atc aaa gga att gat gat gtg aac atg gga act ttc aat 288
 Asp Asp Pro Ile Lys Gly Ile Asp Asp Val Asn Met Gly Thr Phe Asn
 85 90 95

att tct gag aaa cga tct gcc caa ata aat aaa agt ttc act gat cta 336

13

Ile	Ser	Glu	Lys	Arg	Ser	Ala	Gln	Ile	Asn	Lys	Ser	Phe	Thr	Asp	Leu	
			100					105					110			
cgt	atg	cga	gtt	cgt	gca	gaa	gga	ctt	atg	gat	gga	tct	cct	ttg	ttc	384
Arg	Met	Arg	Val	Arg	Ala	Glu	Gly	Leu	Met	Asp	Gly	Ser	Pro	Leu	Phe	
			115				120					125				
tac	att	aga	aaa	att	ctt	gaa	aca	atc	ttc	aca	att	ctt	ttt	gca	ttc	432
Tyr	Ile	Arg	Lys	Ile	Leu	Glu	Thr	Ile	Phe	Thr	Ile	Leu	Phe	Ala	Phe	
	130					135					140					
tac	ctt	caa	tac	cac	aca	tat	tat	ctt	cca	tca	gct	att	cta	atg	gga	480
Tyr	Leu	Gln	Tyr	His	Thr	Tyr	Tyr	Leu	Pro	Ser	Ala	Ile	Leu	Met	Gly	
	145				150					155					160	
gtt	gcg	tgg	caa	caa	ttg	gga	tgg	tta	atc	cat	gaa	ttc	gca	cat	cat	528
Val	Ala	Trp	Gln	Gln	Leu	Gly	Trp	Leu	Ile	His	Glu	Phe	Ala	His	His	
				165				170						175		
cag	ttg	ttc	aaa	aac	aga	tac	tac	aat	gat	ttg	gcc	agc	tat	ttc	gtt	576
Gln	Leu	Phe	Lys	Asn	Arg	Tyr	Tyr	Asn	Asp	Leu	Ala	Ser	Tyr	Phe	Val	
			180					185					190			
gga	aac	ttt	tta	caa	gga	ttc	tca	tct	ggt	ggt	tgg	aaa	gag	cag	cac	624
Gly	Asn	Phe	Leu	Gln	Gly	Phe	Ser	Ser	Gly	Gly	Trp	Lys	Glu	Gln	His	
		195				200						205				
aat	gtg	cat	cac	gca	gcc	aca	aat	gtt	gtt	gga	cga	gac	gga	gat	ctt	672
Asn	Val	His	His	Ala	Ala	Thr	Asn	Val	Val	Gly	Arg	Asp	Gly	Asp	Leu	
	210					215					220					
gat	tta	gtc	cca	ttc	tat	gct	aca	gtg	gca	gaa	cat	ctc	aac	aat	tat	720
Asp	Leu	Val	Pro	Phe	Tyr	Ala	Thr	Val	Ala	Glu	His	Leu	Asn	Asn	Tyr	
	225				230					235					240	
tct	cag	gat	tca	tgg	gtt	atg	act	cta	ttc	aga	tgg	caa	cat	gtt	cat	768
Ser	Gln	Asp	Ser	Trp	Val	Met	Thr	Leu	Phe	Arg	Trp	Gln	His	Val	His	
				245					250					255		
tgg	aca	ttc	atg	tta	cca	ttc	ctc	cgt	ctc	tcg	tgg	ctt	ctt	cag	tca	816
Trp	Thr	Phe	Met	Leu	Pro	Phe	Leu	Arg	Leu	Ser	Trp	Leu	Leu	Gln	Ser	
			260					265						270		
atc	att	ttt	gtt	agt	cag	atg	cca	act	cat	tat	tat	gac	tat	tac	aga	864
Ile	Ile	Phe	Val	Ser	Gln	Met	Pro	Thr	His	Tyr	Tyr	Asp	Tyr	Tyr	Arg	
		275				280						285				
aat	act	gcg	att	tat	gaa	cag	gtt	ggt	ctc	tct	ttg	cac	tgg	gct	tgg	912
Asn	Thr	Ala	Ile	Tyr	Glu	Gln	Val	Gly	Leu	Ser	Leu	His	Trp	Ala	Trp	
		290				295					300					
tca	ttg	ggt	caa	ttg	tat	ttc	cta	ccc	gat	tgg	tca	act	aga	ata	atg	960
Ser	Leu	Gly	Gln	Leu	Tyr	Phe	Leu	Pro	Asp	Trp	Ser	Thr	Arg	Ile	Met	
	305				310					315					320	
ttc	ttc	ctt	gtt	tct	cat	ctt	gtt	gga	ggt	ttc	ctg	ctc	tct	cat	gta	1008
Phe	Phe	Leu	Val	Ser	His	Leu	Val	Gly	Gly	Phe	Leu	Leu	Ser	His	Val	
				325					330					335		
gtt	act	ttc	aat	cat	tat	tca	gtg	gag	aag	ttt	gca	ttg	agc	tcg	aac	1056
Val	Thr	Phe	Asn	His	Tyr	Ser	Val	Glu	Lys	Phe	Ala	Leu	Ser	Ser	Asn	
			340					345					350			
atc	atg	tca	aat	tac	gct	tgt	ctt	caa	atc	atg	acc	aca	aga	aat	atg	1104

14

Ile Met Ser Asn Tyr Ala Cys Leu Gln Ile Met Thr Thr Arg Asn Met
 355 360 365

aga cct gga aga ttc att gac tgg ctt tgg gga ggt ctt aac tat cag 1152
 Arg Pro Gly Arg Phe Ile Asp Trp Leu Trp Gly Gly Leu Asn Tyr Gln
 370 375 380

att gag cac cat ctt ttc cca acg atg cca cga cac aac ttg aac act 1200
 Ile Glu His His Leu Phe Pro Thr Met Pro Arg His Asn Leu Asn Thr
 385 390 395 400

gtt atg cca ctt gtt aag gag ttt gca gca gca aat ggt tta cca tac 1248
 Val Met Pro Leu Val Lys Glu Phe Ala Ala Ala Asn Gly Leu Pro Tyr
 405 410 415

atg gtc gac gat tat ttc aca gga ttc tgg ctt gaa att gag caa ttc 1296
 Met Val Asp Asp Tyr Phe Thr Gly Phe Trp Leu Glu Ile Glu Gln Phe
 420 425 430

cga aat att gca aat gtt gct gct aaa ttg act aaa aag att gcc tag 1344
 Arg Asn Ile Ala Asn Val Ala Ala Lys Leu Thr Lys Lys Ile Ala
 435 440 445

<210> 8

<211> 447

<212> PRT

<213> Ceratodon purpureus

<400> 8

Met Val Leu Arg Glu Gln Glu His Glu Pro Phe Phe Ile Lys Ile Asp
 1 5 10 15

Gly Lys Trp Cys Gln Ile Asp Asp Ala Val Leu Arg Ser His Pro Gly
 20 25 30

Gly Ser Ala Ile Thr Thr Tyr Lys Asn Met Asp Ala Thr Thr Val Phe
 35 40 45

His Thr Phe His Thr Gly Ser Lys Glu Ala Tyr Gln Trp Leu Thr Glu
 50 55 60

Leu Lys Lys Glu Cys Pro Thr Gln Glu Pro Glu Ile Pro Asp Ile Lys
 65 70 75 80

Asp Asp Pro Ile Lys Gly Ile Asp Asp Val Asn Met Gly Thr Phe Asn
 85 90 95

Ile Ser Glu Lys Arg Ser Ala Gln Ile Asn Lys Ser Phe Thr Asp Leu
 100 105 110

Arg Met Arg Val Arg Ala Glu Gly Leu Met Asp Gly Ser Pro Leu Phe
 115 120 125

Tyr Ile Arg Lys Ile Leu Glu Thr Ile Phe Thr Ile Leu Phe Ala Phe
 130 135 140

Tyr Leu Gln Tyr His Thr Tyr Tyr Leu Pro Ser Ala Ile Leu Met Gly
 145 150 155 160

Val Ala Trp Gln Gln Leu Gly Trp Leu Ile His Glu Phe Ala His His
 165 170 175

Gln Leu Phe Lys Asn Arg Tyr Tyr Asn Asp Leu Ala Ser Tyr Phe Val
 180 185 190

Gly Asn Phe Leu Gln Gly Phe Ser Ser Gly Gly Trp Lys Glu Gln His
 195 200 205

Asn Val His His Ala Ala Thr Asn Val Val Gly Arg Asp Gly Asp Leu
 210 215 220

Asp Leu Val Pro Phe Tyr Ala Thr Val Ala Glu His Leu Asn Asn Tyr
 225 230 235 240

Ser Gln Asp Ser Trp Val Met Thr Leu Phe Arg Trp Gln His Val His
 245 250 255

Trp Thr Phe Met Leu Pro Phe Leu Arg Leu Ser Trp Leu Leu Gln Ser
 260 265 270

Ile Ile Phe Val Ser Gln Met Pro Thr His Tyr Tyr Asp Tyr Tyr Arg
 275 280 285

Asn Thr Ala Ile Tyr Glu Gln Val Gly Leu Ser Leu His Trp Ala Trp
 290 295 300

Ser Leu Gly Gln Leu Tyr Phe Leu Pro Asp Trp Ser Thr Arg Ile Met
 305 310 315 320

Phe Phe Leu Val Ser His Leu Val Gly Gly Phe Leu Leu Ser His Val
 325 330 335

Val Thr Phe Asn His Tyr Ser Val Glu Lys Phe Ala Leu Ser Ser Asn
 340 345 350

Ile Met Ser Asn Tyr Ala Cys Leu Gln Ile Met Thr Thr Arg Asn Met
 355 360 365

Arg Pro Gly Arg Phe Ile Asp Trp Leu Trp Gly Gly Leu Asn Tyr Gln
370 375 380

Ile Glu His His Leu Phe Pro Thr Met Pro Arg His Asn Leu Asn Thr
385 390 395 400

Val Met Pro Leu Val Lys Glu Phe Ala Ala Ala Asn Gly Leu Pro Tyr
405 410 415

Met Val Asp Asp Tyr Phe Thr Gly Phe Trp Leu Glu Ile Glu Gln Phe
420 425 430

Arg Asn Ile Ala Asn Val Ala Ala Lys Leu Thr Lys Lys Ile Ala
435 440 445

<210> 9

<211> 1443

<212> DNA

<213> Physcomitrella patens

<220>

<221> CDS

<222> (1)..(1443)

<223> Delta-5 desaturase

<400> 9

atg gcg ccc cac tct gcg gat act gct ggg ctc gtg cct tct gac gaa 48
Met Ala Pro His Ser Ala Asp Thr Ala Gly Leu Val Pro Ser Asp Glu
1 5 10 15

ttg agg cta cga acg tcg aat tca aag ggt ccc gaa caa gag caa act 96
Leu Arg Leu Arg Thr Ser Asn Ser Lys Gly Pro Glu Gln Glu Gln Thr
20 25 30

ttg aag aag tac acc ctt gaa gat gtc agc cgc cac aac acc cca gca 144
Leu Lys Lys Tyr Thr Leu Glu Asp Val Ser Arg His Asn Thr Pro Ala
35 40 45

gat tgt tgg ttg gtg ata tgg ggc aaa gtc tac gat gtc aca agc tgg 192
Asp Cys Trp Leu Val Ile Trp Gly Lys Val Tyr Asp Val Thr Ser Trp
50 55 60

att ccc aat cat ccg ggg ggc agt ctc atc cac gta aaa gca ggg cag 240
Ile Pro Asn His Pro Gly Gly Ser Leu Ile His Val Lys Ala Gly Gln
65 70 75 80

gat tcc act cag ctt ttc gat tcc tat cac ccc ctt tat gtc agg aaa 288
Asp Ser Thr Gln Leu Phe Asp Ser Tyr His Pro Leu Tyr Val Arg Lys

				85				90				95							
atg	ctc	gcg	aag	tac	tgt	att	ggg	gaa	tta	gta	ccg	tct	gct	ggg	gat	336			
Met	Leu	Ala	Lys	Tyr	Cys	Ile	Gly	Glu	Leu	Val	Pro	Ser	Ala	Gly	Asp				
				100				105				110							
gac	aag	ttt	aag	aaa	gca	act	ctg	gag	tat	gca	gat	gcc	gaa	aat	gaa	384			
Asp	Lys	Phe	Lys	Lys	Ala	Thr	Leu	Glu	Tyr	Ala	Asp	Ala	Glu	Asn	Glu				
				115				120				125							
gat	ttc	tat	ttg	gtt	gtg	aag	caa	cga	gtt	gaa	tct	tat	ttc	aag	agt	432			
Asp	Phe	Tyr	Leu	Val	Val	Lys	Gln	Arg	Val	Glu	Ser	Tyr	Phe	Lys	Ser				
				130				135				140							
aac	aag	ata	aac	ccc	caa	att	cat	cca	cat	atg	atc	ctg	aag	tca	ttg	480			
Asn	Lys	Ile	Asn	Pro	Gln	Ile	His	Pro	His	Met	Ile	Leu	Lys	Ser	Leu				
				145				150				155				160			
ttc	att	ctt	ggg	gga	tat	ttc	gcc	agt	tac	tat	tta	gcg	ttc	ttc	tg	528			
Phe	Ile	Leu	Gly	Gly	Tyr	Phe	Ala	Ser	Tyr	Tyr	Leu	Ala	Phe	Phe	Trp				
				165				170				175							
tct	tca	agt	gtc	ctt	gtt	tct	ttg	ttt	ttc	gca	ttg	tg	atg	ggg	ttc	576			
Ser	Ser	Ser	Val	Leu	Val	Ser	Leu	Phe	Phe	Ala	Leu	Trp	Met	Gly	Phe				
				180				185				190							
ttc	gca	gcg	gaa	gtc	ggc	gtg	tcg	att	caa	cat	gat	gga	aat	cat	ggg	624			
Phe	Ala	Ala	Glu	Val	Gly	Val	Ser	Ile	Gln	His	Asp	Gly	Asn	His	Gly				
				195				200				205							
tca	tac	act	aaa	tg	cgt	ggc	ttt	gga	tat	atc	atg	gga	gcc	tcc	cta	672			
Ser	Tyr	Thr	Lys	Trp	Arg	Gly	Phe	Gly	Tyr	Ile	Met	Gly	Ala	Ser	Leu				
				210				215				220							
gat	cta	gtc	gga	gcc	agt	agc	ttc	atg	tg	aga	cag	caa	cac	gtt	gtg	720			
Asp	Leu	Val	Gly	Ala	Ser	Ser	Phe	Met	Trp	Arg	Gln	Gln	His	Val	Val				
				225				230				235				240			
gga	cat	cac	tcg	ttt	aca	aat	gtg	gac	aac	tac	gat	cct	gat	att	cgt	768			
Gly	His	His	Ser	Phe	Thr	Asn	Val	Asp	Asn	Tyr	Asp	Pro	Asp	Ile	Arg				
				245				250				255							
gtg	aaa	gat	cca	gat	gtc	agg	agg	gtt	gcg	acc	aca	caa	cca	aga	caa	816			
Val	Lys	Asp	Pro	Asp	Val	Arg	Arg	Val	Ala	Thr	Thr	Gln	Pro	Arg	Gln				
				260				265				270							
tg	tat	cat	gcg	tat	cag	cat	atc	tac	ctg	gca	gta	tta	tat	gga	act	864			
Trp	Tyr	His	Ala	Tyr	Gln	His	Ile	Tyr	Leu	Ala	Val	Leu	Tyr	Gly	Thr				
				275				280				285							
cta	gct	ctt	aag	agt	att	ttt	cta	gat	gat	ttc	ctt	gcg	tac	ttc	aca	912			
Leu	Ala	Leu	Lys	Ser	Ile	Phe	Leu	Asp	Asp	Phe	Leu	Ala	Tyr	Phe	Thr				
				290				295				300							
gga	tca	att	ggc	cct	gtc	aag	gtg	gcg	aaa	atg	acc	ccc	ctg	gag	ttc	960			
Gly	Ser	Ile	Gly	Pro	Val	Lys	Val	Ala	Lys	Met	Thr	Pro	Leu	Glu	Phe				
				305				310				315				320			
aac	atc	ttc	ttt	cag	gga	aag	ctg	cta	tat	gcg	ttc	tac	atg	ttc	gtg	1008			
Asn	Ile	Phe	Phe	Gln	Gly	Lys	Leu	Leu	Tyr	Ala</									

340	345	350	
tat gtg gct tct cag ctc att aca ggt tgg atg tta gct ttt ctt ttt			1104
Tyr Val Ala Ser Gln Leu Ile Thr Gly Trp Met Leu Ala Phe Leu Phe			
355	360	365	
caa gta gca cat gtc gtg gat gat gtt gca ttt cct aca cca gaa ggt			1152
Gln Val Ala His Val Val Asp Asp Val Ala Phe Pro Thr Pro Glu Gly			
370	375	380	
ggg aag gtg aag gga gga tgg gct gca atg cag gtt gca aca act acg			1200
Gly Lys Val Lys Gly Gly Trp Ala Ala Met Gln Val Ala Thr Thr Thr			
385	390	395	400
gat ttc agt cca cgc tca tgg ttc tgg ggt cat gtc tct gga gga tta			1248
Asp Phe Ser Pro Arg Ser Trp Phe Trp Gly His Val Ser Gly Gly Leu			
405	410	415	
aac aac caa att gag cat cat ctg ttt cca gga gtg tgc cat gtt cat			1296
Asn Asn Gln Ile Glu His His Leu Phe Pro Gly Val Cys His Val His			
420	425	430	
tat cca gcc att cag cct att gtc gag aag acg tgc aag gaa ttc gat			1344
Tyr Pro Ala Ile Gln Pro Ile Val Glu Lys Thr Cys Lys Glu Phe Asp			
435	440	445	
gtg cct tat gta gcc tac cca act ttt tgg act gcg ttg aga gcc cac			1392
Val Pro Tyr Val Ala Tyr Pro Thr Phe Trp Thr Ala Leu Arg Ala His			
450	455	460	
ttt gcg cat ttg aaa aag gtt gga ttg aca gag ttt cgg ctc gat ggc			1440
Phe Ala His Leu Lys Lys Val Gly Leu Thr Glu Phe Arg Leu Asp Gly			
465	470	475	480

tga	1443
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<210> 10

<211> 480

<212> PRT

<213> Physcomitrella patens

<400> 10

Met Ala Pro His Ser Ala Asp Thr Ala Gly Leu Val Pro Ser Asp Glu
1 5 10 15

Leu Arg Leu Arg Thr Ser Asn Ser Lys Gly Pro Glu Gln Glu Gln Thr
20 25 30

Leu Lys Lys Tyr Thr Leu Glu Asp Val Ser Arg His Asn Thr Pro Ala
35 40 45

Asp Cys Trp Leu Val Ile Trp Gly Lys Val Tyr Asp Val Thr Ser Trp
50 55 60

Ile Pro Asn His Pro Gly Gly Ser Leu Ile His Val Lys Ala Gly Gln
65 70 75 80

Asp Ser Thr Gln Leu Phe Asp Ser Tyr His Pro Leu Tyr Val Arg Lys
85 90 95

Met Leu Ala Lys Tyr Cys Ile Gly Glu Leu Val Pro Ser Ala Gly Asp
100 105 110

Asp Lys Phe Lys Lys Ala Thr Leu Glu Tyr Ala Asp Ala Glu Asn Glu
115 120 125

Asp Phe Tyr Leu Val Val Lys Gln Arg Val Glu Ser Tyr Phe Lys Ser
130 135 140

Asn Lys Ile Asn Pro Gln Ile His Pro His Met Ile Leu Lys Ser Leu
145 150 155 160

Phe Ile Leu Gly Gly Tyr Phe Ala Ser Tyr Tyr Leu Ala Phe Phe Trp
165 170 175

Ser Ser Ser Val Leu Val Ser Leu Phe Phe Ala Leu Trp Met Gly Phe
180 185 190

Phe Ala Ala Glu Val Gly Val Ser Ile Gln His Asp Gly Asn His Gly
195 200 205

Ser Tyr Thr Lys Trp Arg Gly Phe Gly Tyr Ile Met Gly Ala Ser Leu
210 215 220

Asp Leu Val Gly Ala Ser Ser Phe Met Trp Arg Gln Gln His Val Val
225 230 235 240

Gly His His Ser Phe Thr Asn Val Asp Asn Tyr Asp Pro Asp Ile Arg
245 250 255

Val Lys Asp Pro Asp Val Arg Arg Val Ala Thr Thr Gln Pro Arg Gln
260 265 270

Trp Tyr His Ala Tyr Gln His Ile Tyr Leu Ala Val Leu Tyr Gly Thr
275 280 285

Leu Ala Leu Lys Ser Ile Phe Leu Asp Asp Phe Leu Ala Tyr Phe Thr
290 295 300

Gly Ser Ile Gly Pro Val Lys Val Ala Lys Met Thr Pro Leu Glu Phe
305 310 315 320

20

Asn Ile Phe Phe Gln Gly Lys Leu Leu Tyr Ala Phe Tyr Met Phe Val
325 330 335

Leu Pro Ser Val Tyr Gly Val His Ser Gly Gly Thr Phe Leu Ala Leu
340 345 350

Tyr Val Ala Ser Gln Leu Ile Thr Gly Trp Met Leu Ala Phe Leu Phe
355 360 365

Gln Val Ala His Val Val Asp Asp Val Ala Phe Pro Thr Pro Glu Gly
370 375 380

Gly Lys Val Lys Gly Gly Trp Ala Ala Met Gln Val Ala Thr Thr Thr
385 390 395 400

Asp Phe Ser Pro Arg Ser Trp Phe Trp Gly His Val Ser Gly Gly Leu
405 410 415

Asn Asn Gln Ile Glu His His Leu Phe Pro Gly Val Cys His Val His
420 425 430

Tyr Pro Ala Ile Gln Pro Ile Val Glu Lys Thr Cys Lys Glu Phe Asp
435 440 445

Val Pro Tyr Val Ala Tyr Pro Thr Phe Trp Thr Ala Leu Arg Ala His
450 455 460

Phe Ala His Leu Lys Lys Val Gly Leu Thr Glu Phe Arg Leu Asp Gly
465 470 475 480

<210> 11

<211> 1320

<212> DNA

<213> Thraustrochytrium

$\langle 220 \rangle$

<221> CDS

<222> (1) .. (1320)

<223>

<400> 11

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Met Gly Lys Gly Ser Glu Gly Arg Ser Ala Ala Arg Glu Met Thr Ala

21

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gag gcg aac ggc gac aag cgg aaa acg att ctg atc gag ggc gtc ctg Glu Ala Asn Gly Asp Lys Arg Lys Thr Ile Leu Ile Glu Gly Val Leu 20 25 30				96
tac gac gcg acg aac ttt aag cac ccg ggc ggt tcg atc atc aac ttc Tyr Asp Ala Thr Asn Phe Lys His Pro Gly Gly Ser Ile Ile Asn Phe 35 40 45				144
ttg acc gag ggc gag gcc ggc gtg gac gcg acg cag gcg tac cgc gag Leu Thr Glu Gly Glu Ala Gly Val Asp Ala Thr Gln Ala Tyr Arg Glu 50 55 60				192
ttt cat cag cgg tcc ggc aag gcc gac aag tac ctc aag tcg ctg ccg Phe His Gln Arg Ser Gly Lys Ala Asp Lys Tyr Leu Lys Ser Leu Pro 65 70 75 80				240
aag ctg gat gcg tcc aag gtg gag tcg cgg ttc tcg gcc aaa gag cag Lys Leu Asp Ala Ser Lys Val Glu Ser Arg Phe Ser Ala Lys Glu Gln 85 90 95				288
gcg cgg cgc gac gcc atg acg cgc gac tac gcg gcc ttt cgc gag gag Ala Arg Arg Asp Ala Met Thr Arg Asp Tyr Ala Ala Phe Arg Glu Glu 100 105 110				336
ctc gtc gcc gag ggg tac ttt gac ccg tcg atc ccg cac atg att tac Leu Val Ala Glu Gly Tyr Phe Asp Pro Ser Ile Pro His Met Ile Tyr 115 120 125				384
cgc gtc gtg gag atc gtg gcg ctc ttc gcg ctc tcg ttc tgg ctc atg Arg Val Val Glu Ile Val Ala Leu Phe Ala Leu Ser Phe Trp Leu Met 130 135 140				432
tcc aag gcc tcg ccc acc tcg ctc gtg ctg ggc gtg gtg atg aac ggc Ser Lys Ala Ser Pro Thr Ser Leu Val Leu Gly Val Val Met Asn Gly 145 150 155 160				480
att gcg cag ggc cgc tgc ggc tgg gtc atg cac gag atg ggc cac ggg Ile Ala Gln Gly Arg Cys Gly Trp Val Met His Glu Met Gly His Gly 165 170 175				528
tcg ttc acg ggc gtc atc tgg ctc gac gac cgg atg tgc gag ttc ttc Ser Phe Thr Gly Val Ile Trp Leu Asp Asp Arg Met Cys Glu Phe Phe 180 185 190				576
tac ggc gtc ggc tgc ggc atg agc ggg cac tac tgg aag aac cag cac Tyr Gly Val Gly Cys Gly Met Ser Gly His Tyr Trp Lys Asn Gln His 195 200 205				624
agc aag cac cac gcc gcg ccc aac cgc ctc gag cac gat gtc gat ctc Ser Lys His His Ala Ala Pro Asn Arg Leu Glu His Asp Val Asp Leu 210 215 220				672
aac acg ctg ccc ctg gtc gcc ttt aac gag cgc gtc gtg cgc aag gtc Asn Thr Leu Pro Leu Val Ala Phe Asn Glu Arg Val Val Arg Lys Val 225 230 235 240				720
aag ccg gga tcg ctg ctg gcg ctc tgg ctg cgc gtg cag gcg tac ctc Lys Pro Gly Ser Leu Leu Ala Leu Trp Leu Arg Val Gln Ala Tyr Leu 245 250 255				768
ttt gcg ccc gtc tcg tgc ctg ctc atc ggc ctt ggc tgg acg ctc tac Phe Ala Pro Val Ser Cys Leu Leu Ile Gly Leu Gly Trp Thr Leu Tyr				816

260	265	270	
ctg cac ccg cgc tac atg ctg cgc acc aag cgg cac atg gag ttc gtc Leu His Pro Arg Tyr Met Leu Arg Thr Lys Arg His Met Glu Phe Val 275 280 285			864
tgg atc ttc gcg cgc tac att ggc tgg ttc tcg ctc atg ggc gct ctc Trp Ile Phe Ala Arg Tyr Ile Gly Trp Phe Ser Leu Met Gly Ala Leu 290 295 300			912
ggc tac tcg ccg ggc acc tcg gtc ggg atg tac ctg tgc tcg ttc ggc Gly Tyr Ser Pro Gly Thr Ser Val Gly Met Tyr Leu Cys Ser Phe Gly 305 310 315 320			960
ctc ggc tgc att tac att ttc ctg cag ttc gcc gtc agc cac acg cac Leu Gly Cys Ile Tyr Ile Phe Leu Gln Phe Ala Val Ser His Thr His 325 330 335			1008
ctg ccg gtg acc aac ccg gag gac cag ctg cac tgg ctc gag tac gcg Leu Pro Val Thr Asn Pro Glu Asp Gln Leu His Trp Leu Glu Tyr Ala 340 345 350			1056
gcc gac cac acg gtg aac att agc acc aag tcc tgg ctc gtc acg tgg Ala Asp His Thr Val Asn Ile Ser Thr Lys Ser Trp Leu Val Thr Trp 355 360 365			1104
tgg atg tcg aac ctg aac ttt cag atc gag cac cac ctc ttc ccc acg Trp Met Ser Asn Leu Asn Phe Gln Ile Glu His His Leu Phe Pro Thr 370 375 380			1152
gcg ccg cag ttc cgc ttc aag gaa atc agt cct cgc gtc gag gcc ctc Ala Pro Gln Phe Arg Phe Lys Glu Ile Ser Pro Arg Val Glu Ala Leu 385 390 395 400			1200
ttc aag cgc cac aac ctc ccg tac tac gac ctg ccc tac acg agc gcg Phe Lys Arg His Asn Leu Pro Tyr Tyr Asp Leu Pro Tyr Thr Ser Ala 405 410 415			1248
gtc tcg acc acc ttt gcc aat ctt tat tcc gtc ggc cac tcg gtc ggc Val Ser Thr Thr Phe Ala Asn Leu Tyr Ser Val Gly His Ser Val Gly 420 425 430			1296
gcc gac acc aag aag cag gac tga Ala Asp Thr Lys Lys Gln Asp 435			1320

<210> 12

<211> 439

<212> PRT

<213> Thraustrochytrium

<400> 12

Met Gly Lys Gly Ser Glu Gly Arg Ser Ala Ala Arg Glu Met Thr Ala
1 5 10 15

Glu Ala Asn Gly Asp Lys Arg Lys Thr Ile Leu Ile Glu Gly Val Leu

20					25					30					
Tyr	Asp	Ala	Thr	Asn	Phe	Lys	His	Pro	Gly	Gly	Ser	Ile	Ile	Asn	Phe
		35					40					45			
Leu	Thr	Glu	Gly	Glu	Ala	Gly	Val	Asp	Ala	Thr	Gln	Ala	Tyr	Arg	Glu
	50					55					60				
Phe	His	Gln	Arg	Ser	Gly	Lys	Ala	Asp	Lys	Tyr	Leu	Lys	Ser	Leu	Pro
65					70					75					80
Lys	Leu	Asp	Ala	Ser	Lys	Val	Glu	Ser	Arg	Phe	Ser	Ala	Lys	Glu	Gln
				85					90					95	
Ala	Arg	Arg	Asp	Ala	Met	Thr	Arg	Asp	Tyr	Ala	Ala	Phe	Arg	Glu	Glu
			100					105					110		
Leu	Val	Ala	Glu	Gly	Tyr	Phe	Asp	Pro	Ser	Ile	Pro	His	Met	Ile	Tyr
		115					120					125			
Arg	Val	Val	Glu	Ile	Val	Ala	Leu	Phe	Ala	Leu	Ser	Phe	Trp	Leu	Met
	130					135					140				
Ser	Lys	Ala	Ser	Pro	Thr	Ser	Leu	Val	Leu	Gly	Val	Val	Met	Asn	Gly
145					150					155					160
Ile	Ala	Gln	Gly	Arg	Cys	Gly	Trp	Val	Met	His	Glu	Met	Gly	His	Gly
				165					170					175	
Ser	Phe	Thr	Gly	Val	Ile	Trp	Leu	Asp	Asp	Arg	Met	Cys	Glu	Phe	Phe
			180					185					190		
Tyr	Gly	Val	Gly	Cys	Gly	Met	Ser	Gly	His	Tyr	Trp	Lys	Asn	Gln	His
	195						200					205			
Ser	Lys	His	His	Ala	Ala	Pro	Asn	Arg	Leu	Glu	His	Asp	Val	Asp	Leu
	210					215					220				
Asn	Thr	Leu	Pro	Leu	Val	Ala	Phe	Asn	Glu	Arg	Val	Val	Arg	Lys	Val
225					230					235					240
Lys	Pro	Gly	Ser	Leu	Leu	Ala	Leu	Trp	Leu	Arg	Val	Gln	Ala	Tyr	Leu
				245					250					255	
Phe	Ala	Pro	Val	Ser	Cys	Leu	Leu	Ile	Gly	Leu	Gly	Trp	Thr	Leu	Tyr
		260						265					270		
Leu	His	Pro	Arg	Tyr	Met	Leu	Arg	Thr	Lys	Arg	His	Met	Glu	Phe	Val

275

280

285

Trp Ile Phe Ala Arg Tyr Ile Gly Trp Phe Ser Leu Met Gly Ala Leu
 290 295 300

Gly Tyr Ser Pro Gly Thr Ser Val Gly Met Tyr Leu Cys Ser Phe Gly
 305 310 315 320

Leu Gly Cys Ile Tyr Ile Phe Leu Gln Phe Ala Val Ser His Thr His
 325 330 335

Leu Pro Val Thr Asn Pro Glu Asp Gln Leu His Trp Leu Glu Tyr Ala
 340 345 350

Ala Asp His Thr Val Asn Ile Ser Thr Lys Ser Trp Leu Val Thr Trp
 355 360 365

Trp Met Ser Asn Leu Asn Phe Gln Ile Glu His His Leu Phe Pro Thr
 370 375 380

Ala Pro Gln Phe Arg Phe Lys Glu Ile Ser Pro Arg Val Glu Ala Leu
 385 390 395 400

Phe Lys Arg His Asn Leu Pro Tyr Tyr Asp Leu Pro Tyr Thr Ser Ala
 405 410 415

Val Ser Thr Thr Phe Ala Asn Leu Tyr Ser Val Gly His Ser Val Gly
 420 425 430

Ala Asp Thr Lys Lys Gln Asp
 435

<210> 13

<211> 1341

<212> DNA

<213> *Mortierella alpina*

<220>

<221> CDS

<222> (1)..(1341)

<223> Delta-5 desaturase

<400> 13

25

atg gga acg gac caa gga aaa acc ttc acc tgg gaa gag ctg gcg gcc Met Gly Thr Asp Gln Gly Lys Thr Phe Thr Trp Glu Glu Leu Ala Ala 1 5 10 15	48
cat aac acc aag gac gac cta ctc ttg gcc atc cgc ggc agg gtg tac His Asn Thr Lys Asp Asp Leu Leu Leu Ala Ile Arg Gly Arg Val Tyr 20 25 30	96
gat gtc aca aag ttc ttg agc cgc cat cct ggt gga gtg gac act ctc Asp Val Thr Lys Phe Leu Ser Arg His Pro Gly Gly Val Asp Thr Leu 35 40 45	144
ctg ctc gga gct ggc cga gat gtt act ccg gtc ttt gag atg tat cac Leu Leu Gly Ala Gly Arg Asp Val Thr Pro Val Phe Glu Met Tyr His 50 55 60	192
gcg ttt ggg gct gca gat gcc att atg aag aag tac tat gtc ggt aca Ala Phe Gly Ala Ala Asp Ala Ile Met Lys Lys Tyr Tyr Val Gly Thr 65 70 75 80	240
ctg gtc tcg aat gag ctg ccc atc ttc ccg gag cca acg gtg ttc cac Leu Val Ser Asn Glu Leu Pro Ile Phe Pro Glu Pro Thr Val Phe His 85 90 95	288
aaa acc atc aag acg aga gtc gag ggc tac ttt acg gat cgg aac att Lys Thr Ile Lys Thr Arg Val Glu Gly Tyr Phe Thr Asp Arg Asn Ile 100 105 110	336
gat ccc aag aat aga cca gag atc tgg gga cga tac gct ctt atc ttt Asp Pro Lys Asn Arg Pro Glu Ile Trp Gly Arg Tyr Ala Leu Ile Phe 115 120 125	384
gga tcc ttg atc gct tcc tac tac gcg cag ctc ttt gtg cct ttc gtt Gly Ser Leu Ile Ala Ser Tyr Tyr Ala Gln Leu Phe Val Pro Phe Val 130 135 140	432
gtc gaa cgc aca tgg ctt cag gtg gtg ttt gca atc atc atg gga ttt Val Glu Arg Thr Trp Leu Gln Val Val Phe Ala Ile Ile Met Gly Phe 145 150 155 160	480
gcg tgc gca caa gtc gga ctc aac cct ctt cat gat gcg tct cac ttt Ala Cys Ala Gln Val Gly Leu Asn Pro Leu His Asp Ala Ser His Phe 165 170 175	528
tca gtg acc cac aac ccc act gtc tgg aag att ctg gga gcc acg cac Ser Val Thr His Asn Pro Thr Val Trp Lys Ile Leu Gly Ala Thr His 180 185 190	576
gac ttt ttc aac gga gca tcg tac ctg gtg tgg atg tac caa cat atg Asp Phe Phe Asn Gly Ala Ser Tyr Leu Val Trp Met Tyr Gln His Met 195 200 205	624
ctc ggc cat cac ccc tac acc aac att gct gga gca gat ccc gac gtg Leu Gly His His Pro Tyr Thr Asn Ile Ala Gly Ala Asp Pro Asp Val 210 215 220	672
tcg acg tct gag ccc gat gtt cgt cgt atc aag ccc aac caa aag tgg Ser Thr Ser Glu Pro Asp Val Arg Arg Ile Lys Pro Asn Gln Lys Trp 225 230 235 240	720
ttt gtc aac cac atc aac cag cac atg ttt gtt cct ttc ctg tac gga Phe Val Asn His Ile Asn Gln His Met Phe Val Pro Phe Leu Tyr Gly 245 250 255	768

26

ctg ctg gcg ttc aag gtg cgc att cag gac atc aac att ttg tac ttt 816
 Leu Leu Ala Phe Lys Val Arg Ile Gln Asp Ile Asn Ile Leu Tyr Phe
 260 265 270

gtc aag acc aat gac gct att cgt gtc aat ccc atc tcg aca tgg cac 864
 Val Lys Thr Asn Asp Ala Ile Arg Val Asn Pro Ile Ser Thr Trp His
 275 280 285

act gtg atg ttc tgg ggc ggc aag gct ttc ttt gtc tgg tat cgc ctg 912
 Thr Val Met Phe Trp Gly Gly Lys Ala Phe Phe Val Trp Tyr Arg Leu
 290 295 300

att gtt ccc ctg cag tat ctg ccc ctg ggc aag gtg ctg ctc ttg ttc 960
 Ile Val Pro Leu Gln Tyr Leu Pro Leu Gly Lys Val Leu Leu Leu Phe
 305 310 315 320

acg gtc gcg gac atg gtg tcg tct tac tgg ctg gcg ctg acc ttc cag 1008
 Thr Val Ala Asp Met Val Ser Ser Tyr Trp Leu Ala Leu Thr Phe Gln
 325 330 335

gcg aac cac gtt gtt gag gaa gtt cag tgg ccg ttg cct gac gag aac 1056
 Ala Asn His Val Val Glu Glu Val Gln Trp Pro Leu Pro Asp Glu Asn
 340 345 350

ggg atc atc caa aag gac tgg gca gct atg cag gtc gag act acg cag 1104
 Gly Ile Ile Gln Lys Asp Trp Ala Ala Met Gln Val Glu Thr Thr Gln
 355 360 365

gat tac gca cac gat tcg cac ctc tgg acc agc atc act ggc agc ttg 1152
 Asp Tyr Ala His Asp Ser His Leu Trp Thr Ser Ile Thr Gly Ser Leu
 370 375 380

aac tac cag gct gtg cac cat ctg ttc ccc aac gtg tcg cag cac cat 1200
 Asn Tyr Gln Ala Val His His Leu Phe Pro Asn Val Ser Gln His His
 385 390 395 400

tat ccc gat att ctg gcc atc atc aag aac acc tgc agc gag tac aag 1248
 Tyr Pro Asp Ile Leu Ala Ile Ile Lys Asn Thr Cys Ser Glu Tyr Lys
 405 410 415

gtt cca tac ctt gtc aag gat acg ttt tgg caa gca ttt gct tca cat 1296
 Val Pro Tyr Leu Val Lys Asp Thr Phe Trp Gln Ala Phe Ala Ser His
 420 425 430

ttg gag cac ttg cgt gtt ctt gga ctc cgt ccc aag gaa gag tag 1341
 Leu Glu His Leu Arg Val Leu Gly Leu Arg Pro Lys Glu Glu
 435 440 445

<210> 14

<211> 446

<212> PRT

<213> Mortierella alpina

<400> 14

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His Asn Thr Lys Asp Asp Leu Leu Leu Ala Ile Arg Gly Arg Val Tyr
 20 25 30

Asp Val Thr Lys Phe Leu Ser Arg His Pro Gly Gly Val Asp Thr Leu
 35 40 45

Leu Leu Gly Ala Gly Arg Asp Val Thr Pro Val Phe Glu Met Tyr His
 50 55 60

Ala Phe Gly Ala Ala Asp Ala Ile Met Lys Lys Tyr Tyr Val Gly Thr
 65 70 75 80

Leu Val Ser Asn Glu Leu Pro Ile Phe Pro Glu Pro Thr Val Phe His
 85 90 95

Lys Thr Ile Lys Thr Arg Val Glu Gly Tyr Phe Thr Asp Arg Asn Ile
 100 105 110

Asp Pro Lys Asn Arg Pro Glu Ile Trp Gly Arg Tyr Ala Leu Ile Phe
 115 120 125

Gly Ser Leu Ile Ala Ser Tyr Tyr Ala Gln Leu Phe Val Pro Phe Val
 130 135 140

Val Glu Arg Thr Trp Leu Gln Val Val Phe Ala Ile Ile Met Gly Phe
 145 150 155 160

Ala Cys Ala Gln Val Gly Leu Asn Pro Leu His Asp Ala Ser His Phe
 165 170 175

Ser Val Thr His Asn Pro Thr Val Trp Lys Ile Leu Gly Ala Thr His
 180 185 190

Asp Phe Phe Asn Gly Ala Ser Tyr Leu Val Trp Met Tyr Gln His Met
 195 200 205

Leu Gly His His Pro Tyr Thr Asn Ile Ala Gly Ala Asp Pro Asp Val
 210 215 220

Ser Thr Ser Glu Pro Asp Val Arg Arg Ile Lys Pro Asn Gln Lys Trp
 225 230 235 240

Phe Val Asn His Ile Asn Gln His Met Phe Val Pro Phe Leu Tyr Gly
 245 250 255

Leu Leu Ala Phe Lys Val Arg Ile Gln Asp Ile Asn Ile Leu Tyr Phe
 260 265 270

Val Lys Thr Asn Asp Ala Ile Arg Val Asn Pro Ile Ser Thr Trp His
 275 280 285

Thr Val Met Phe Trp Gly Gly Lys Ala Phe Phe Val Trp Tyr Arg Leu
 290 295 300

Ile Val Pro Leu Gln Tyr Leu Pro Leu Gly Lys Val Leu Leu Leu Phe
 305 310 315 320

Thr Val Ala Asp Met Val Ser Ser Tyr Trp Leu Ala Leu Thr Phe Gln
 325 330 335

Ala Asn His Val Val Glu Glu Val Gln Trp Pro Leu Pro Asp Glu Asn
 340 345 350

Gly Ile Ile Gln Lys Asp Trp Ala Ala Met Gln Val Glu Thr Thr Gln
 355 360 365

Asp Tyr Ala His Asp Ser His Leu Trp Thr Ser Ile Thr Gly Ser Leu
 370 375 380

Asn Tyr Gln Ala Val His His Leu Phe Pro Asn Val Ser Gln His His
 385 390 395 400

Tyr Pro Asp Ile Leu Ala Ile Ile Lys Asn Thr Cys Ser Glu Tyr Lys
 405 410 415

Val Pro Tyr Leu Val Lys Asp Thr Phe Trp Gln Ala Phe Ala Ser His
 420 425 430

Leu Glu His Leu Arg Val Leu Gly Leu Arg Pro Lys Glu Glu
 435 440 445

<210> 15

<211> 1344

<212> DNA

<213> *Caenorhabditis elegans*

<220>

<221> CDS

<222> (1)..(1344)

<223> Delta-5 desaturase

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1 5 10 15	
gga aaa tgg tgt caa att gac gat gct gtc ctg aga tca cat cca ggt	96
Gly Lys Trp Cys Gln Ile Asp Asp Ala Val Leu Arg Ser His Pro Gly	
20 25 30	
ggt agt gca att act acc tat aaa aat atg gat gcc act acc gta ttc	144
Gly Ser Ala Ile Thr Thr Tyr Lys Asn Met Asp Ala Thr Thr Val Phe	
35 40 45	
cac aca ttc cat act ggt tct aaa gaa gcg tat caa tgg ctg aca gaa	192
His Thr Phe His Thr Gly Ser Lys Glu Ala Tyr Gln Trp Leu Thr Glu	
50 55 60	
ttg aaa aaa gag tgc cct aca caa gaa cca gag atc cca gat att aag	240
Leu Lys Lys Glu Cys Pro Thr Gln Glu Pro Glu Ile Pro Asp Ile Lys	
65 70 75 80	
gat gac cca atc aaa gga att gat gat gtg aac atg gga act ttc aat	288
Asp Asp Pro Ile Lys Gly Ile Asp Asp Val Asn Met Gly Thr Phe Asn	
85 90 95	
att tct gag aaa cga tct gcc caa ata aat aaa agt ttc act gat cta	336
Ile Ser Glu Lys Arg Ser Ala Gln Ile Asn Lys Ser Phe Thr Asp Leu	
100 105 110	
cgt atg cga gtt cgt gca gaa gga ctt atg gat gga tct cct ttg ttc	384
Arg Met Arg Val Arg Ala Glu Gly Leu Met Asp Gly Ser Pro Leu Phe	
115 120 125	
tac att aga aaa att ctt gaa aca atc ttc aca att ctt ttt gca ttc	432
Tyr Ile Arg Lys Ile Leu Glu Thr Ile Phe Thr Ile Leu Phe Ala Phe	
130 135 140	
tac ctt caa tac cac aca tat tat ctt cca tca gct att cta atg gga	480
Tyr Leu Gln Tyr His Thr Tyr Tyr Leu Pro Ser Ala Ile Leu Met Gly	
145 150 155 160	
ggt gca tgg caa caa ttg gga tgg tta atc cat gaa ttc gca cat cat	528
Val Ala Trp Gln Gln Leu Gly Trp Leu Ile His Glu Phe Ala His His	
165 170 175	
cag ttg ttc aaa aac aga tac tac aat gat ttg gcc agc tat ttc gtt	576
Gln Leu Phe Lys Asn Arg Tyr Tyr Asn Asp Leu Ala Ser Tyr Phe Val	
180 185 190	
gga aac ttt tta caa gga ttc tca tct ggt ggt tgg aaa gag cag cac	624
Gly Asn Phe Leu Gln Gly Phe Ser Ser Gly Gly Trp Lys Glu Gln His	
195 200 205	
aat gtg cat cac gca gcc aca aat gtt gtt gga cga gac gga gat ctt	672
Asn Val His His Ala Ala Thr Asn Val Val Gly Arg Asp Gly Asp Leu	
210 215 220	
gat tta gtc cca ttc tat gct aca gtg gca gaa cat ctc aac aat tat	720
Asp Leu Val Pro Phe Tyr Ala Thr Val Ala Glu His Leu Asn Asn Tyr	
225 230 235 240	
tct cag gat tca tgg gtt atg act cta ttc aga tgg caa cat gtt cat	768

30

Ser	Gln	Asp	Ser	Trp	Val	Met	Thr	Leu	Phe	Arg	Trp	Gln	His	Val	His		
				245					250					255			
tgg	aca	ttc	atg	tta	cca	ttc	ctc	cgt	ctc	tcg	tgg	ctt	ctt	cag	tca		816
Trp	Thr	Phe	Met	Leu	Pro	Phe	Leu	Arg	Leu	Ser	Trp	Leu	Leu	Gln	Ser		
			260					265					270				
atc	att	ttt	gtt	agt	cag	atg	cca	act	cat	tat	tat	gac	tat	tac	aga		864
Ile	Ile	Phe	Val	Ser	Gln	Met	Pro	Thr	His	Tyr	Tyr	Asp	Tyr	Tyr	Arg		
		275					280					285					
aat	act	gcg	att	tat	gaa	cag	gtt	ggt	ctc	tct	ttg	cac	tgg	gct	tgg		912
Asn	Thr	Ala	Ile	Tyr	Glu	Gln	Val	Gly	Leu	Ser	Leu	His	Trp	Ala	Trp		
	290					295					300						
tca	ttg	ggt	caa	ttg	tat	ttc	cta	ccc	gat	tgg	tca	act	aga	ata	atg		960
Ser	Leu	Gly	Gln	Leu	Tyr	Phe	Leu	Pro	Asp	Trp	Ser	Thr	Arg	Ile	Met		
305				310					315						320		
ttc	ttc	ctt	gtt	tct	cat	ctt	gtt	gga	ggt	ttc	ctg	ctc	tct	cat	gta		1008
Phe	Phe	Leu	Val	Ser	His	Leu	Val	Gly	Gly	Phe	Leu	Leu	Ser	His	Val		
			325					330						335			
gtt	act	ttc	aat	cat	tat	tca	gtg	gag	aag	ttt	gca	ttg	agc	tcg	aac		1056
Val	Thr	Phe	Asn	His	Tyr	Ser	Val	Glu	Lys	Phe	Ala	Leu	Ser	Ser	Asn		
		340					345					350					
atc	atg	tca	aat	tac	gct	tgt	ctt	caa	atc	atg	acc	aca	aga	aat	atg		1104
Ile	Met	Ser	Asn	Tyr	Ala	Cys	Leu	Gln	Ile	Met	Thr	Thr	Arg	Asn	Met		
		355				360						365					
aga	cct	gga	aga	ttc	att	gac	tgg	ctt	tgg	gga	ggt	ctt	aac	tat	cag		1152
Arg	Pro	Gly	Arg	Phe	Ile	Asp	Trp	Leu	Trp	Gly	Gly	Leu	Asn	Tyr	Gln		
	370					375				380							
att	gag	cac	cat	ctt	ttc	cca	acg	atg	cca	cga	cac	aac	ttg	aac	act		1200
Ile	Glu	His	His	Leu	Phe	Pro	Thr	Met	Pro	Arg	His	Asn	Leu	Asn	Thr		
385				390					395						400		
gtt	atg	cca	ctt	gtt	aag	gag	ttt	gca	gca	gca	aat	ggt	tta	cca	tac		1248
Val	Met	Pro	Leu	Val	Lys	Glu	Phe	Ala	Ala	Ala	Asn	Gly	Leu	Pro	Tyr		
			405					410					415				
atg	gtc	gac	gat	tat	ttc	aca	gga	ttc	tgg	ctt	gaa	att	gag	caa	ttc		1296
Met	Val	Asp	Asp	Tyr	Phe	Thr	Gly	Phe	Trp	Leu	Glu	Ile	Glu	Gln	Phe		
		420					425					430					
cga	aat	att	gca	aat	gtt	gct	gct	aaa	ttg	act	aaa	aag	att	gcc	tag		1344
Arg	Asn	Ile	Ala	Asn	Val	Ala	Ala	Lys	Leu	Thr	Lys	Lys	Ile	Ala			
		435				440						445					

<210> 16

<211> 447

<212> PRT

<213> Caenorhabditis elegans

<400> 16

31

Met Val Leu Arg Glu Gln Glu His Glu Pro Phe Phe Ile Lys Ile Asp
 1 5 10 15
 Gly Lys Trp Cys Gln Ile Asp Asp Ala Val Leu Arg Ser His Pro Gly
 20 25 30
 Gly Ser Ala Ile Thr Thr Tyr Lys Asn Met Asp Ala Thr Thr Val Phe
 35 40 45
 His Thr Phe His Thr Gly Ser Lys Glu Ala Tyr Gln Trp Leu Thr Glu
 50 55 60
 Leu Lys Lys Glu Cys Pro Thr Gln Glu Pro Glu Ile Pro Asp Ile Lys
 65 70 75 80
 Asp Asp Pro Ile Lys Gly Ile Asp Asp Val Asn Met Gly Thr Phe Asn
 85 90 95
 Ile Ser Glu Lys Arg Ser Ala Gln Ile Asn Lys Ser Phe Thr Asp Leu
 100 105 110
 Arg Met Arg Val Arg Ala Glu Gly Leu Met Asp Gly Ser Pro Leu Phe
 115 120 125
 Tyr Ile Arg Lys Ile Leu Glu Thr Ile Phe Thr Ile Leu Phe Ala Phe
 130 135 140
 Tyr Leu Gln Tyr His Thr Tyr Tyr Leu Pro Ser Ala Ile Leu Met Gly
 145 150 155 160
 Val Ala Trp Gln Gln Leu Gly Trp Leu Ile His Glu Phe Ala His His
 165 170 175
 Gln Leu Phe Lys Asn Arg Tyr Tyr Asn Asp Leu Ala Ser Tyr Phe Val
 180 185 190
 Gly Asn Phe Leu Gln Gly Phe Ser Ser Gly Gly Trp Lys Glu Gln His
 195 200 205
 Asn Val His His Ala Ala Thr Asn Val Val Gly Arg Asp Gly Asp Leu
 210 215 220
 Asp Leu Val Pro Phe Tyr Ala Thr Val Ala Glu His Leu Asn Asn Tyr
 225 230 235 240
 Ser Gln Asp Ser Trp Val Met Thr Leu Phe Arg Trp Gln His Val His
 245 250 255

32

Trp Thr Phe Met Leu Pro Phe Leu Arg Leu Ser Trp Leu Leu Gln Ser
 260 265 270

Ile Ile Phe Val Ser Gln Met Pro Thr His Tyr Tyr Asp Tyr Tyr Arg
 275 280 285

Asn Thr Ala Ile Tyr Glu Gln Val Gly Leu Ser Leu His Trp Ala Trp
 290 295 300

Ser Leu Gly Gln Leu Tyr Phe Leu Pro Asp Trp Ser Thr Arg Ile Met
 305 310 315 320

Phe Phe Leu Val Ser His Leu Val Gly Gly Phe Leu Leu Ser His Val
 325 330 335

Val Thr Phe Asn His Tyr Ser Val Glu Lys Phe Ala Leu Ser Ser Asn
 340 345 350

Ile Met Ser Asn Tyr Ala Cys Leu Gln Ile Met Thr Thr Arg Asn Met
 355 360 365

Arg Pro Gly Arg Phe Ile Asp Trp Leu Trp Gly Gly Leu Asn Tyr Gln
 370 375 380

Ile Glu His His Leu Phe Pro Thr Met Pro Arg His Asn Leu Asn Thr
 385 390 395 400

Val Met Pro Leu Val Lys Glu Phe Ala Ala Ala Asn Gly Leu Pro Tyr
 405 410 415

Met Val Asp Asp Tyr Phe Thr Gly Phe Trp Leu Glu Ile Glu Gln Phe
 420 425 430

Arg Asn Ile Ala Asn Val Ala Ala Lys Leu Thr Lys Lys Ile Ala
 435 440 445

<210> 17

<211> 1683

<212> DNA

<213> Borago officinalis

<220>

<221> CDS

<222> (42)..(1388)

<223> Delta-6 desaturase

<400> 17

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Met Ala Ala Gln Ile	
1 5	
aag aaa tac att acc tca gat gaa ctc aag aac cac gat aaa ccc gga	104
Lys Lys Tyr Ile Thr Ser Asp Glu Leu Lys Asn His Asp Lys Pro Gly	
10 15 20	
gat cta tgg atc tcg att caa ggg aaa gcc tat gat gtt tcg gat tgg	152
Asp Leu Trp Ile Ser Ile Gln Gly Lys Ala Tyr Asp Val Ser Asp Trp	
25 30 35	
gtg aaa gac cat cca ggt ggc agc ttt ccc ttg aag agt ctt gct ggt	200
Val Lys Asp His Pro Gly Gly Ser Phe Pro Leu Lys Ser Leu Ala Gly	
40 45 50	
caa gag gta act gat gca ttt gtt gca ttc cat cct gcc tct aca tgg	248
Gln Glu Val Thr Asp Ala Phe Val Ala Phe His Pro Ala Ser Thr Trp	
55 60 65	
aag aat ctt gat aag ttt ttc act ggg tat tat ctt aaa gat tac tct	296
Lys Asn Leu Asp Lys Phe Phe Thr Gly Tyr Tyr Leu Lys Asp Tyr Ser	
70 75 80 85	
gtt tct gag gtt tct aaa gat tat agg aag ctt gtg ttt gag ttt tct	344
Val Ser Glu Val Ser Lys Asp Tyr Arg Lys Leu Val Phe Glu Phe Ser	
90 95 100	
aaa atg ggt ttg tat gac aaa aaa ggt cat att atg ttt gca act ttg	392
Lys Met Gly Leu Tyr Asp Lys Lys Gly His Ile Met Phe Ala Thr Leu	
105 110 115	
tgc ttt ata gca atg ctg ttt gct atg agt gtt tat ggg gtt ttg ttt	440
Cys Phe Ile Ala Met Leu Phe Ala Met Ser Val Tyr Gly Val Leu Phe	
120 125 130	
tgt gag ggt gtt ttg gta cat ttg ttt tct ggg tgt ttg atg ggg ttt	488
Cys Glu Gly Val Leu Val His Leu Phe Ser Gly Cys Leu Met Gly Phe	
135 140 145	
ctt tgg att cag agt ggt tgg att gga cat gat gct ggg cat tat atg	536
Leu Trp Ile Gln Ser Gly Trp Ile Gly His Asp Ala Gly His Tyr Met	
150 155 160 165	
gta gtg tct gat tca agg ctt aat aag ttt atg ggt att ttt gct gca	584
Val Val Ser Asp Ser Arg Leu Asn Lys Phe Met Gly Ile Phe Ala Ala	
170 175 180	
aat tgt ctt tca gga ata agt att ggt tgg tgg aaa tgg aac cat aat	632
Asn Cys Leu Ser Gly Ile Ser Ile Gly Trp Trp Lys Trp Asn His Asn	
185 190 195	
gca cat cac att gcc tgt aat agc ctt gaa tat gac cct gat tta caa	680
Ala His His Ile Ala Cys Asn Ser Leu Glu Tyr Asp Pro Asp Leu Gln	
200 205 210	
tat ata cca ttc ctt gtt gtg tct tcc aag ttt ttt ggt tca ctc acc	728
Tyr Ile Pro Phe Leu Val Val Ser Ser Lys Phe Phe Gly Ser Leu Thr	
215 220 225	

tct cat ttc tat gag aaa agg ttg act ttt gac tct tta tca aga ttc Ser His Phe Tyr Glu Lys Arg Leu Thr Phe Asp Ser Leu Ser Arg Phe 230 235 240 245	776
ttt gta agt tat caa cat tgg aca ttt tac cct att atg tgt gct gct Phe Val Ser Tyr Gln His Trp Thr Phe Tyr Pro Ile Met Cys Ala Ala 250 255 260	824
agg ctc aat atg tat gta caa tct ctc ata atg ttg ttg acc aag aga Arg Leu Asn Met Tyr Val Gln Ser Leu Ile Met Leu Leu Thr Lys Arg 265 270 275	872
aat gtg tcc tat cga gct cag gaa ctc ttg gga tgc cta gtg ttc tcg Asn Val Ser Tyr Arg Ala Gln Glu Leu Leu Gly Cys Leu Val Phe Ser 280 285 290	920
att tgg tac ccg ttg ctt gtt tct tgt ttg cct aat tgg ggt gaa aga Ile Trp Tyr Pro Leu Leu Val Ser Cys Leu Pro Asn Trp Gly Glu Arg 295 300 305	968
att atg ttt gtt att gca agt tta tca gtg act gga atg caa caa gtt Ile Met Phe Val Ile Ala Ser Leu Ser Val Thr Gly Met Gln Gln Val 310 315 320 325	1016
cag ttc tcc ttg aac cac ttc tct tca agt gtt tat gtt gga aag cct Gln Phe Ser Leu Asn His Phe Ser Ser Ser Val Tyr Val Gly Lys Pro 330 335 340	1064
aaa ggg aat aat tgg ttt gag aaa caa acg gat ggg aca ctt gac att Lys Gly Asn Asn Trp Phe Glu Lys Gln Thr Asp Gly Thr Leu Asp Ile 345 350 355	1112
tct tgt cct cct tgg atg gat tgg ttt cat ggt gga ttg caa ttc caa Ser Cys Pro Pro Trp Met Asp Trp Phe His Gly Gly Leu Gln Phe Gln 360 365 370	1160
att gag cat cat ttg ttt ccc aag atg cct aga tgc aac ctt agg aaa Ile Glu His His Leu Phe Pro Lys Met Pro Arg Cys Asn Leu Arg Lys 375 380 385	1208
atc tcg ccc tac gtg atc gag tta tgc aag aaa cat aat ttg cct tac Ile Ser Pro Tyr Val Ile Glu Leu Cys Lys Lys His Asn Leu Pro Tyr 390 395 400 405	1256
aat tat gca tct ttc tcc aag gcc aat gaa atg aca ctc aga aca ttg Asn Tyr Ala Ser Phe Ser Lys Ala Asn Glu Met Thr Leu Arg Thr Leu 410 415 420	1304
agg aac aca gca ttg cag gct agg gat ata acc aag ccg ctc ccg aag Arg Asn Thr Ala Leu Gln Ala Arg Asp Ile Thr Lys Pro Leu Pro Lys 425 430 435	1352
aat ttg gta tgg gaa gct ctt cac act cat ggt taa aattaccctt Asn Leu Val Trp Glu Ala Leu His Thr His Gly 440 445	1398
agttcatgta ataatttgag attatgtatc tcctatgttt gtgtcttgctc ttggttctac	1458
ttgttgaggt cattgcaact tgtcttttat gggtttattag atgtttttta atatatttta	1518
gaggttttgc tttcatctcc attattgatg aataaggagt tgcataattgt caattgttgt	1578
gctcaatatc tgatatatttg gaatgtactt tgtaccactg tgttttcagt tgaagctcat	1638

gtgtacttct atagactttg tttaaattgg tatgtcatgt tattt

1683

<210> 18

<211> 448

<212> PRT

<213> Borago officinalis

<400> 18

Met Ala Ala Gln Ile Lys Lys Tyr Ile Thr Ser Asp Glu Leu Lys Asn
1 5 10 15

His Asp Lys Pro Gly Asp Leu Trp Ile Ser Ile Gln Gly Lys Ala Tyr
20 25 30

Asp Val Ser Asp Trp Val Lys Asp His Pro Gly Gly Ser Phe Pro Leu
35 40 45

Lys Ser Leu Ala Gly Gln Glu Val Thr Asp Ala Phe Val Ala Phe His
50 55 60

Pro Ala Ser Thr Trp Lys Asn Leu Asp Lys Phe Phe Thr Gly Tyr Tyr
65 70 75 80

Leu Lys Asp Tyr Ser Val Ser Glu Val Ser Lys Asp Tyr Arg Lys Leu
85 90 95

Val Phe Glu Phe Ser Lys Met Gly Leu Tyr Asp Lys Lys Gly His Ile
100 105 110

Met Phe Ala Thr Leu Cys Phe Ile Ala Met Leu Phe Ala Met Ser Val
115 120 125

Tyr Gly Val Leu Phe Cys Glu Gly Val Leu Val His Leu Phe Ser Gly
130 135 140

Cys Leu Met Gly Phe Leu Trp Ile Gln Ser Gly Trp Ile Gly His Asp
145 150 155 160

Ala Gly His Tyr Met Val Val Ser Asp Ser Arg Leu Asn Lys Phe Met
165 170 175

Gly Ile Phe Ala Ala Asn Cys Leu Ser Gly Ile Ser Ile Gly Trp Trp
180 185 190

36

Lys Trp Asn His Asn Ala His His Ile Ala Cys Asn Ser Leu Glu Tyr
 195 200 205

Asp Pro Asp Leu Gln Tyr Ile Pro Phe Leu Val Val Ser Ser Lys Phe
 210 215 220

Phe Gly Ser Leu Thr Ser His Phe Tyr Glu Lys Arg Leu Thr Phe Asp
 225 230 235 240

Ser Leu Ser Arg Phe Phe Val Ser Tyr Gln His Trp Thr Phe Tyr Pro
 245 250 255

Ile Met Cys Ala Ala Arg Leu Asn Met Tyr Val Gln Ser Leu Ile Met
 260 265 270

Leu Leu Thr Lys Arg Asn Val Ser Tyr Arg Ala Gln Glu Leu Leu Gly
 275 280 285

Cys Leu Val Phe Ser Ile Trp Tyr Pro Leu Leu Val Ser Cys Leu Pro
 290 295 300

Asn Trp Gly Glu Arg Ile Met Phe Val Ile Ala Ser Leu Ser Val Thr
 305 310 315 320

Gly Met Gln Gln Val Gln Phe Ser Leu Asn His Phe Ser Ser Ser Val
 325 330 335

Tyr Val Gly Lys Pro Lys Gly Asn Asn Trp Phe Glu Lys Gln Thr Asp
 340 345 350

Gly Thr Leu Asp Ile Ser Cys Pro Pro Trp Met Asp Trp Phe His Gly
 355 360 365

Gly Leu Gln Phe Gln Ile Glu His His Leu Phe Pro Lys Met Pro Arg
 370 375 380

Cys Asn Leu Arg Lys Ile Ser Pro Tyr Val Ile Glu Leu Cys Lys Lys
 385 390 395 400

His Asn Leu Pro Tyr Asn Tyr Ala Ser Phe Ser Lys Ala Asn Glu Met
 405 410 415

Thr Leu Arg Thr Leu Arg Asn Thr Ala Leu Gln Ala Arg Asp Ile Thr
 420 425 430

Lys Pro Leu Pro Lys Asn Leu Val Trp Glu Ala Leu His Thr His Gly
 435 440 445

<210> 19

<211> 1563

<212> DNA

<213> *Ceratodon purpureus*

<220>

<221> CDS

<222> (1)..(1563)

<223> Delta-6 desaturase

<400> 19

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Met	Val	Ser	Gln	Gly	Gly	Gly	Leu	Ser	Gln	Gly	Ser	Ile	Glu	Glu	Asn	
1				5					10					15		

att	gac	gtt	gag	cac	ttg	gca	acg	atg	ccc	ctc	gtc	agt	gac	ttc	cta	96
Ile	Asp	Val	Glu	His	Leu	Ala	Thr	Met	Pro	Leu	Val	Ser	Asp	Phe	Leu	
			20					25					30			

aat	gtc	ctg	gga	acg	act	ttg	ggc	cag	tgg	agt	ctt	tcc	act	aca	ttc	144
Asn	Val	Leu	Gly	Thr	Thr	Leu	Gly	Gln	Trp	Ser	Leu	Ser	Thr	Thr	Phe	
		35					40					45				

gct	ttc	aag	agg	ctc	acg	act	aag	aaa	cac	agt	tcg	gac	atc	tcg	gtg	192
Ala	Phe	Lys	Arg	Leu	Thr	Thr	Lys	Lys	His	Ser	Ser	Asp	Ile	Ser	Val	
	50						55				60					

gag	gca	caa	aaa	gaa	tcg	gtt	gcg	cgg	ggg	cca	gtt	gag	aat	att	tct	240
Glu	Ala	Gln	Lys	Glu	Ser	Val	Ala	Arg	Gly	Pro	Val	Glu	Asn	Ile	Ser	
65						70				75					80	

caa	tcg	gtt	gcg	cag	ccc	atc	agg	cgg	agg	tgg	gtg	cag	gat	aaa	aag	288
Gln	Ser	Val	Ala	Gln	Pro	Ile	Arg	Arg	Arg	Trp	Val	Gln	Asp	Lys	Lys	
			85						90					95		

ccg	gtt	act	tac	agc	ctg	aag	gat	gta	gct	tcg	cac	gat	atg	ccc	cag	336
Pro	Val	Thr	Tyr	Ser	Leu	Lys	Asp	Val	Ala	Ser	His	Asp	Met	Pro	Gln	
			100					105					110			

gac	tgc	tgg	att	ata	atc	aaa	gag	aag	gtg	tat	gat	gtg	agc	acc	ttc	384
Asp	Cys	Trp	Ile	Ile	Ile	Lys	Glu	Lys	Val	Tyr	Asp	Val	Ser	Thr	Phe	
		115					120					125				

gct	gag	cag	cac	cct	gga	ggc	acg	gtt	atc	aac	acc	tac	ttc	gga	cga	432
Ala	Glu	Gln	His	Pro	Gly	Gly	Thr	Val	Ile	Asn	Thr	Tyr	Phe	Gly	Arg	
	130					135					140					

gac	gcc	aca	gat	gtt	ttc	tct	act	ttc	cac	gca	tcc	acc	tca	tgg	aag	480
Asp	Ala	Thr	Asp	Val	Phe	Ser	Thr	Phe	His	Ala	Ser	Thr	Ser	Trp	Lys	
145				150						155					160	

att	ctt	cag	aat	ttc	tac	atc	ggg	aac	ctt	gtt	agg	gag	gag	ccg	act	528
Ile	Leu	Gln	Asn	Phe	Tyr	Ile	Gly	Asn	Leu	Val	Arg	Glu	Glu	Pro	Thr	
				165					170						175	

ttg gag ctg ctg aag gag tac aga gag ttg aga gcc ctt ttc ttg aga	576
Leu Glu Leu Leu Lys Glu Tyr Arg Glu Leu Arg Ala Leu Phe Leu Arg	
180 185 190	
gaa cag ctt ttc aag agt tcc aaa tcc tac tac ctt ttc aag act ctc	624
Glu Gln Leu Phe Lys Ser Ser Lys Ser Tyr Tyr Leu Phe Lys Thr Leu	
195 200 205	
ata aat gtt tcc att gtt gcc aca agc att gcg ata atc agt ctg tac	672
Ile Asn Val Ser Ile Val Ala Thr Ser Ile Ala Ile Ile Ser Leu Tyr	
210 215 220	
aag tct tac cgg gcg gtt ctg tta tca gcc agt ttg atg ggc ttg ttt	720
Lys Ser Tyr Arg Ala Val Leu Leu Ser Ala Ser Leu Met Gly Leu Phe	
225 230 235 240	
att caa cag tgc gga tgg ttg tct cac gat ttt cta cac cat cag gta	768
Ile Gln Gln Cys Gly Trp Leu Ser His Asp Phe Leu His His Gln Val	
245 250 255	
ttt gag aca cgc tgg ctc aat gac gtt gtt ggc tat gtg gtc ggc aac	816
Phe Glu Thr Arg Trp Leu Asn Asp Val Val Gly Tyr Val Val Gly Asn	
260 265 270	
gtt gtt ctg gga ttc agt gtc tcg tgg tgg aag acc aag cac aac ctg	864
Val Val Leu Gly Phe Ser Val Ser Trp Trp Lys Thr Lys His Asn Leu	
275 280 285	
cat cat gct gct ccg aat gaa tgc gac caa aag tac aca ccg att gat	912
His His Ala Ala Pro Asn Glu Cys Asp Gln Lys Tyr Thr Pro Ile Asp	
290 295 300	
gag gat att gat act ctc ccc atc att gct tgg agt aaa gat ctc ttg	960
Glu Asp Ile Asp Thr Leu Pro Ile Ile Ala Trp Ser Lys Asp Leu Leu	
305 310 315 320	
gcc act gtt gag agc aag acc atg ttg cga gtt ctt cag tac cag cac	1008
Ala Thr Val Glu Ser Lys Thr Met Leu Arg Val Leu Gln Tyr Gln His	
325 330 335	
cta ttc ttt ttg gtt ctt ttg acg ttt gcc cgg gcg agt tgg cta ttt	1056
Leu Phe Phe Leu Val Leu Leu Thr Phe Ala Arg Ala Ser Trp Leu Phe	
340 345 350	
tgg agc gcg gcc ttc act ctc agg ccc gag ttg acc ctt ggc gag aag	1104
Trp Ser Ala Ala Phe Thr Leu Arg Pro Glu Leu Thr Leu Gly Glu Lys	
355 360 365	
ctt ttg gag agg gga acg atg gct ttg cac tac att tgg ttt aat agt	1152
Leu Leu Glu Arg Gly Thr Met Ala Leu His Tyr Ile Trp Phe Asn Ser	
370 375 380	
gtt gcg ttt tat ctg ctc ccc gga tgg aaa cca gtt gta tgg atg gtg	1200
Val Ala Phe Tyr Leu Leu Pro Gly Trp Lys Pro Val Val Trp Met Val	
385 390 395 400	
gtc agc gag ctc atg tct ggt ttc ctg ctg gga tac gta ttt gta ctc	1248
Val Ser Glu Leu Met Ser Gly Phe Leu Leu Gly Tyr Val Phe Val Leu	
405 410 415	
agt cac aat gga atg gag gtg tac aat acg tca aag gac ttc gtg aat	1296
Ser His Asn Gly Met Glu Val Tyr Asn Thr Ser Lys Asp Phe Val Asn	
420 425 430	

gcc cag att gca tcg act cgc gac atc aaa gca ggg gtg ttt aat gat 1344
Ala Gln Ile Ala Ser Thr Arg Asp Ile Lys Ala Gly Val Phe Asn Asp
435 440 445

tgg ttc acc gga ggt ctc aac aga cag att gag cat cat cta ttt cca 1392
 Trp Phe Thr Gly Gly Leu Asn Arg Gln Ile Glu His His Leu Phe Pro
 450 455 460

acg atg ccc agg cac aac ctt aat aaa att tct cct cac gtg gag act 1440
Thr Met Pro Arg His Asn Leu Asn Lys Ile Ser Pro His Val Glu Thr
465 470 475 480

ttg tgc aag aag cat gga ctg gtc tac gaa gac gtg agc atg gct tcg	1488
Leu Cys Lys Lys His Gly Leu Val Tyr Glu Asp Val Ser Met Ala Ser	
485	490
	495

ggc act tac cgg gtt ttg aaa aca ctt aag gac gtt gcc gat gct gct 1536
Gly Thr Tyr Arg Val Leu Lys Thr Leu Lys Asp Val Ala Asp Ala Ala
500 505 510

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tca cac cag cag ctt gct gcg agt tga      1563
Ser His Gln Gln Leu Ala Ala Ser
      515                      520
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<210> 20

<211> 520

<212> PRT

<213> Ceratodon purpureus

<400> 20

Met Val Ser Gln Gly Gly Gly Leu Ser Gln Gly Ser Ile Glu Glu Asn
1 5 10 15

Ile Asp Val Glu His Leu Ala Thr Met Pro Leu Val Ser Asp Phe Leu
20 25 30

Asn Val Leu Gly Thr Thr Leu Gly Gln Trp Ser Leu Ser Thr Thr Phe
35 40 45

Ala Phe Lys Arg Leu Thr Thr Lys Lys His Ser Ser Asp Ile Ser Val
50 55 60

Glu Ala Gln Lys Glu Ser Val Ala Arg Gly Pro Val Glu Asn Ile Ser
65 70 75 80

Gln Ser Val Ala Gln Pro Ile Arg Arg Arg Trp Val Gln Asp Lys Lys
85 90 95

Pro Val Thr Tyr Ser Leu Lys Asp Val Ala Ser His Asp Met Pro Gln
100 105 110

Asp Cys Trp Ile Ile Ile Lys Glu Lys Val Tyr Asp Val Ser Thr Phe
 115 120 125

Ala Glu Gln His Pro Gly Gly Thr Val Ile Asn Thr Tyr Phe Gly Arg
 130 135 140

Asp Ala Thr Asp Val Phe Ser Thr Phe His Ala Ser Thr Ser Trp Lys
 145 150 155 160

Ile Leu Gln Asn Phe Tyr Ile Gly Asn Leu Val Arg Glu Glu Pro Thr
 165 170 175

Leu Glu Leu Leu Lys Glu Tyr Arg Glu Leu Arg Ala Leu Phe Leu Arg
 180 185 190

Glu Gln Leu Phe Lys Ser Ser Lys Ser Tyr Tyr Leu Phe Lys Thr Leu
 195 200 205

Ile Asn Val Ser Ile Val Ala Thr Ser Ile Ala Ile Ile Ser Leu Tyr
 210 215 220

Lys Ser Tyr Arg Ala Val Leu Leu Ser Ala Ser Leu Met Gly Leu Phe
 225 230 235 240

Ile Gln Gln Cys Gly Trp Leu Ser His Asp Phe Leu His His Gln Val
 245 250 255

Phe Glu Thr Arg Trp Leu Asn Asp Val Val Gly Tyr Val Val Gly Asn
 260 265 270

Val Val Leu Gly Phe Ser Val Ser Trp Trp Lys Thr Lys His Asn Leu
 275 280 285

His His Ala Ala Pro Asn Glu Cys Asp Gln Lys Tyr Thr Pro Ile Asp
 290 295 300

Glu Asp Ile Asp Thr Leu Pro Ile Ile Ala Trp Ser Lys Asp Leu Leu
 305 310 315 320

Ala Thr Val Glu Ser Lys Thr Met Leu Arg Val Leu Gln Tyr Gln His
 325 330 335

Leu Phe Phe Leu Val Leu Leu Thr Phe Ala Arg Ala Ser Trp Leu Phe
 340 345 350

Trp Ser Ala Ala Phe Thr Leu Arg Pro Glu Leu Thr Leu Gly Glu Lys
 355 360 365

Leu Leu Glu Arg Gly Thr Met Ala Leu His Tyr Ile Trp Phe Asn Ser
 370 375 380

Val Ala Phe Tyr Leu Leu Pro Gly Trp Lys Pro Val Val Trp Met Val
 385 390 395 400

Val Ser Glu Leu Met Ser Gly Phe Leu Leu Gly Tyr Val Phe Val Leu
 405 410 415

Ser His Asn Gly Met Glu Val Tyr Asn Thr Ser Lys Asp Phe Val Asn
 420 425 430

Ala Gln Ile Ala Ser Thr Arg Asp Ile Lys Ala Gly Val Phe Asn Asp
 435 440 445

Trp Phe Thr Gly Gly Leu Asn Arg Gln Ile Glu His His Leu Phe Pro
 450 455 460

Thr Met Pro Arg His Asn Leu Asn Lys Ile Ser Pro His Val Glu Thr
 465 470 475 480

Leu Cys Lys Lys His Gly Leu Val Tyr Glu Asp Val Ser Met Ala Ser
 485 490 495

Gly Thr Tyr Arg Val Leu Lys Thr Leu Lys Asp Val Ala Asp Ala Ala
 500 505 510

Ser His Gln Gln Leu Ala Ala Ser
 515 520

<210> 21

<211> 1434

<212> DNA

<213> Phaeodactylum tricornutum

<220>

<221> CDS

<222> (1)..(1434)

<223> Delta-6 desaturase

<400> 21

atg ggc aaa gga ggg gac gct cgg gcc tcg aag ggc tca acg gcg gct

42

Met	Gly	Lys	Gly	Gly	Asp	Ala	Arg	Ala	Ser	Lys	Gly	Ser	Thr	Ala	Ala	
1				5					10					15		
cgc	aag	atc	agt	tgg	cag	gaa	gtc	aag	acc	cac	gcg	tct	ccg	gag	gac	96
Arg	Lys	Ile	Ser	Trp	Gln	Glu	Val	Lys	Thr	His	Ala	Ser	Pro	Glu	Asp	
			20					25					30			
gcc	tgg	atc	att	cac	tcc	aat	aag	gtc	tac	gac	gtg	tcc	aac	tgg	cac	144
Ala	Trp	Ile	Ile	His	Ser	Asn	Lys	Val	Tyr	Asp	Val	Ser	Asn	Trp	His	
			35				40					45				
gaa	cat	ccc	gga	ggc	gcc	gtc	att	ttc	acg	cac	gcc	ggg	gac	gac	atg	192
Glu	His	Pro	Gly	Gly	Ala	Val	Ile	Phe	Thr	His	Ala	Gly	Asp	Asp	Met	
		50				55					60					
acg	gac	att	ttc	gct	gcc	ttt	cac	gca	ccc	gga	tcg	cag	tcg	ctc	atg	240
Thr	Asp	Ile	Phe	Ala	Phe	His	Ala	Pro	Gly	Ser	Gln	Ser	Leu	Met		
65					70				75					80		
aag	aag	ttc	tac	att	ggc	gaa	ttg	ctc	ccg	gaa	acc	acc	ggc	aag	gag	288
Lys	Lys	Phe	Tyr	Ile	Gly	Glu	Leu	Leu	Pro	Glu	Thr	Thr	Gly	Lys	Glu	
				85					90					95		
ccg	cag	caa	atc	gcc	ttt	gaa	aag	ggc	tac	cgc	gat	ctg	cgc	tcc	aaa	336
Pro	Gln	Gln	Ile	Ala	Phe	Glu	Lys	Gly	Tyr	Arg	Asp	Leu	Arg	Ser	Lys	
			100					105					110			
ctc	atc	atg	atg	ggc	atg	ttc	aag	tcc	aac	aag	tgg	ttc	tac	gtc	tac	384
Leu	Ile	Met	Met	Gly	Met	Phe	Lys	Ser	Asn	Lys	Trp	Phe	Tyr	Val	Tyr	
		115					120					125				
aag	tgc	ctc	agc	aac	atg	gcc	att	tgg	gcc	gcc	gcc	tgt	gct	ctc	gtc	432
Lys	Cys	Leu	Ser	Asn	Met	Ala	Ile	Trp	Ala	Ala	Ala	Cys	Ala	Leu	Val	
		130				135					140					
ttt	tac	tcg	gac	cgc	ttc	tgg	gta	cac	ctg	gcc	agc	gcc	gtc	atg	ctg	480
Phe	Tyr	Ser	Asp	Arg	Phe	Trp	Val	His	Leu	Ala	Ser	Ala	Val	Met	Leu	
145					150				155					160		
gga	aca	ttc	ttt	cag	cag	tcg	gga	tgg	ttg	gca	cac	gac	ttt	ctg	cac	528
Gly	Thr	Phe	Phe	Gln	Gln	Ser	Gly	Trp	Leu	Ala	His	Asp	Phe	Leu	His	
				165				170					175			
cac	cag	gtc	ttc	acc	aag	cgc	aag	cac	ggg	gat	ctc	gga	gga	ctc	ttt	576
His	Gln	Val	Phe	Thr	Lys	Arg	Lys	His	Gly	Asp	Leu	Gly	Gly	Leu	Phe	
			180					185					190			
tgg	ggg	aac	ctc	atg	cag	ggt	tac	tcc	gta	cag	tgg	tgg	aaa	aac	aag	624
Trp	Gly	Asn	Leu	Met	Gln	Gly	Tyr	Ser	Val	Gln	Trp	Trp	Lys	Asn	Lys	
		195				200						205				
cac	aac	gga	cac	cac	gcc	gtc	ccc	aac	ctc	cac	tgc	tcc	tcc	gca	gtc	672
His	Asn	Gly	His	His	Ala	Val	Pro	Asn	Leu	His	Cys	Ser	Ser	Ala	Val	
		210				215					220					
gcg	caa	gat	ggg	gac	ccg	gac	atc	gat	acc	atg	ccc	ctt	ctc	gcc	tgg	720
Ala	Gln	Asp	Gly	Asp	Pro	Asp	Ile	Asp	Thr	Met	Pro	Leu	Leu	Ala	Trp	
225					230				235					240		
tcc	gtc	cag	caa	gcc	cag	tct	tac	cgg	gaa	ctc	caa	gcc	gac	gga	aag	768
Ser	Val	Gln	Gln	Ala	Gln	Ser	Tyr	Arg	Glu	Leu	Gln	Ala	Asp	Gly	Lys	
				245					250					255		
gat	tcg	ggg	ttg	gtc	aag	ttc	atg	atc	cgt	aac	caa	tcc	tac	ttt	tac	816

43

Asp	Ser	Gly	Leu	Val	Lys	Phe	Met	Ile	Arg	Asn	Gln	Ser	Tyr	Phe	Tyr		
			260					265					270				
ttt	ccc	atc	ttg	ttg	ctc	gcc	cgc	ctg	tcg	tgg	ttg	aac	gag	tcc	ttc	864	
Phe	Pro	Ile	Leu	Leu	Leu	Ala	Arg	Leu	Ser	Trp	Leu	Asn	Glu	Ser	Phe		
		275					280					285					
aag	tgc	gcc	ttt	ggg	ctt	gga	gct	gcg	tcg	gag	aac	gct	gct	ctc	gaa	912	
Lys	Cys	Ala	Phe	Gly	Leu	Gly	Ala	Ala	Ser	Glu	Asn	Ala	Ala	Leu	Glu		
	290					295					300						
ctc	aag	gcc	aag	ggt	ctt	cag	tac	ccc	ctt	ttg	gaa	aag	gct	ggc	atc	960	
Leu	Lys	Ala	Lys	Gly	Leu	Gln	Tyr	Pro	Leu	Leu	Glu	Lys	Ala	Gly	Ile		
305					310					315					320		
ctg	ctg	cac	tac	gct	tgg	atg	ctt	aca	gtt	tcg	tcc	ggc	ttt	gga	cgc	1008	
Leu	Leu	His	Tyr	Ala	Trp	Met	Leu	Thr	Val	Ser	Ser	Gly	Phe	Gly	Arg		
				325					330					335			
ttc	tcg	ttc	gcg	tac	acc	gca	ttt	tac	ttt	cta	acc	gcg	acc	gcg	tcc	1056	
Phe	Ser	Phe	Ala	Tyr	Thr	Ala	Phe	Tyr	Phe	Leu	Thr	Ala	Thr	Ala	Ser		
			340					345					350				
tgt	gga	ttc	ttg	ctc	gcc	att	gtc	ttt	ggc	ctc	ggc	cac	aac	ggc	atg	1104	
Cys	Gly	Phe	Leu	Leu	Ala	Ile	Val	Phe	Gly	Leu	Gly	His	Asn	Gly	Met		
		355				360						365					
gcc	acc	tac	aat	gcc	gac	gcc	cgt	ccg	gac	ttc	tgg	aag	ctc	caa	gtc	1152	
Ala	Thr	Tyr	Asn	Ala	Asp	Ala	Arg	Pro	Asp	Phe	Trp	Lys	Leu	Gln	Val		
	370					375					380						
acc	acg	act	cgc	aac	gtc	acg	ggc	gga	cac	ggt	ttc	ccc	caa	gcc	ttt	1200	
Thr	Thr	Thr	Arg	Asn	Val	Thr	Gly	Gly	His	Gly	Phe	Pro	Gln	Ala	Phe		
385					390					395					400		
gtc	gac	tgg	ttc	tgt	ggt	ggc	ctc	cag	tac	caa	gtc	gac	cac	cac	tta	1248	
Val	Asp	Trp	Phe	Cys	Gly	Gly	Leu	Gln	Tyr	Gln	Val	Asp	His	His	Leu		
				405				410						415			
ttc	ccc	agc	ctg	ccc	cga	cac	aat	ctg	gcc	aag	aca	cac	gca	ctg	gtc	1296	
Phe	Pro	Ser	Leu	Pro	Arg	His	Asn	Leu	Ala	Lys	Thr	His	Ala	Leu	Val		
			420					425					430				
gaa	tcg	ttc	tgc	aag	gag	tgg	ggt	gtc	cag	tac	cac	gaa	gcc	gac	ctt	1344	
Glu	Ser	Phe	Cys	Lys	Glu	Trp	Gly	Val	Gln	Tyr	His	Glu	Ala	Asp	Leu		
		435					440					445					
gtg	gac	ggg	acc	atg	gaa	gtc	ttg	cac	cat	ttg	ggc	agc	gtg	gcc	ggc	1392	
Val	Asp	Gly	Thr	Met	Glu	Val	Leu	His	His	Leu	Gly	Ser	Val	Ala	Gly		
	450					455					460						
gaa	ttc	gtc	gtg	gat	ttt	gta	cgc	gat	gga	ccc	gcc	atg	taa			1434	
Glu	Phe	Val	Val	Asp	Phe	Val	Arg	Asp	Gly	Pro	Ala	Met					
465					470					475							

<210> 22

<211> 477

<212> PRT

<213> Phaeodactylum tricornutum

<400> 22

Met Gly Lys Gly Gly Asp Ala Arg Ala Ser Lys Gly Ser Thr Ala Ala
 1 5 10 15
 Arg Lys Ile Ser Trp Gln Glu Val Lys Thr His Ala Ser Pro Glu Asp
 20 25 30
 Ala Trp Ile Ile His Ser Asn Lys Val Tyr Asp Val Ser Asn Trp His
 35 40 45
 Glu His Pro Gly Gly Ala Val Ile Phe Thr His Ala Gly Asp Asp Met
 50 55 60
 Thr Asp Ile Phe Ala Ala Phe His Ala Pro Gly Ser Gln Ser Leu Met
 65 70 75 80
 Lys Lys Phe Tyr Ile Gly Glu Leu Leu Pro Glu Thr Thr Gly Lys Glu
 85 90 95
 Pro Gln Gln Ile Ala Phe Glu Lys Gly Tyr Arg Asp Leu Arg Ser Lys
 100 105 110
 Leu Ile Met Met Gly Met Phe Lys Ser Asn Lys Trp Phe Tyr Val Tyr
 115 120 125
 Lys Cys Leu Ser Asn Met Ala Ile Trp Ala Ala Ala Cys Ala Leu Val
 130 135 140
 Phe Tyr Ser Asp Arg Phe Trp Val His Leu Ala Ser Ala Val Met Leu
 145 150 155 160
 Gly Thr Phe Phe Gln Gln Ser Gly Trp Leu Ala His Asp Phe Leu His
 165 170 175
 His Gln Val Phe Thr Lys Arg Lys His Gly Asp Leu Gly Gly Leu Phe
 180 185 190
 Trp Gly Asn Leu Met Gln Gly Tyr Ser Val Gln Trp Trp Lys Asn Lys
 195 200 205
 His Asn Gly His His Ala Val Pro Asn Leu His Cys Ser Ser Ala Val
 210 215 220
 Ala Gln Asp Gly Asp Pro Asp Ile Asp Thr Met Pro Leu Leu Ala Trp
 225 230 235 240

45

Ser Val Gln Gln Ala Gln Ser Tyr Arg Glu Leu Gln Ala Asp Gly Lys
245 250 255

Asp Ser Gly Leu Val Lys Phe Met Ile Arg Asn Gln Ser Tyr Phe Tyr
260 265 270

Phe Pro Ile Leu Leu Leu Ala Arg Leu Ser Trp Leu Asn Glu Ser Phe
275 280 285

Lys Cys Ala Phe Gly Leu Gly Ala Ala Ser Glu Asn Ala Ala Leu Glu
290 295 300

Leu Lys Ala Lys Gly Leu Gln Tyr Pro Leu Leu Glu Lys Ala Gly Ile
305 310 315 320

Leu Leu His Tyr Ala Trp Met Leu Thr Val Ser Ser Gly Phe Gly Arg
325 330 335

Phe Ser Phe Ala Tyr Thr Ala Phe Tyr Phe Leu Thr Ala Thr Ala Ser
340 345 350

Cys Gly Phe Leu Leu Ala Ile Val Phe Gly Leu Gly His Asn Gly Met
355 360 365

Ala Thr Tyr Asn Ala Asp Ala Arg Pro Asp Phe Trp Lys Leu Gln Val
370 375 380

Thr Thr Thr Arg Asn Val Thr Gly Gly His Gly Phe Pro Gln Ala Phe
385 390 395 400

Val Asp Trp Phe Cys Gly Gly Leu Gln Tyr Gln Val Asp His His Leu
405 410 415

Phe Pro Ser Leu Pro Arg His Asn Leu Ala Lys Thr His Ala Leu Val
420 425 430

Glu Ser Phe Cys Lys Glu Trp Gly Val Gln Tyr His Glu Ala Asp Leu
435 440 445

Val Asp Gly Thr Met Glu Val Leu His His Leu Gly Ser Val Ala Gly
450 455 460

Glu Phe Val Val Asp Phe Val Arg Asp Gly Pro Ala Met
465 470 475

<210> 23

<211> 1578

<212> DNA

<213> *Physcomitrella patens*

<220>

<221> CDS

<222> (1)..(1578)

<223> Delta-6 desaturase

<400> 23

atg gta ttc gcg ggc ggt gga ctt cag cag ggc tct ctc gaa gaa aac	48
Met Val Phe Ala Gly Gly Gly Leu Gln Gln Gly Ser Leu Glu Glu Asn	
1 5 10 15	

atc gac gtc gag cac att gcc agt atg tct ctc ttc agc gac ttc ttc	96
Ile Asp Val Glu His Ile Ala Ser Met Ser Leu Phe Ser Asp Phe Phe	
20 25 30	

agt tat gtg tct tca act gtt ggt tcg tgg agc gta cac agt ata caa	144
Ser Tyr Val Ser Ser Thr Val Gly Ser Trp Ser Val His Ser Ile Gln	
35 40 45	

cct ttg aag cgc ctg acg agt aag aag cgt gtt tcg gaa agc gct gcc	192
Pro Leu Lys Arg Leu Thr Ser Lys Lys Arg Val Ser Glu Ser Ala Ala	
50 55 60	

gtg caa tgt ata tca gct gaa gtt cag aga aat tcg agt acc cag gga	240
Val Gln Cys Ile Ser Ala Glu Val Gln Arg Asn Ser Ser Thr Gln Gly	
65 70 75 80	

act gcg gag gca ctc gca gaa tca gtc gtg aag ccc acg aga cga agg	288
Thr Ala Glu Ala Leu Ala Glu Ser Val Val Lys Pro Thr Arg Arg Arg	
85 90 95	

tca tct cag tgg aag aag tcg aca cac ccc cta tca gaa gta gca gta	336
Ser Ser Gln Trp Lys Lys Ser Thr His Pro Leu Ser Glu Val Ala Val	
100 105 110	

cac aac aag cca agc gat tgc tgg att gtt gta aaa aac aag gtg tat	384
His Asn Lys Pro Ser Asp Cys Trp Ile Val Val Lys Asn Lys Val Tyr	
115 120 125	

gat gtt tcc aat ttt gcg gac gag cat ccc gga gga tca gtt att agt	432
Asp Val Ser Asn Phe Ala Asp Glu His Pro Gly Gly Ser Val Ile Ser	
130 135 140	

act tat ttt gga cga gac ggc aca gat gtt ttc tct agt ttt cat gca	480
Thr Tyr Phe Gly Arg Asp Gly Thr Asp Val Phe Ser Ser Phe His Ala	
145 150 155 160	

gct tct aca tgg aaa att ctt caa gac ttt tac att ggt gac gtg gag	528
Ala Ser Thr Trp Lys Ile Leu Gln Asp Phe Tyr Ile Gly Asp Val Glu	
165 170 175	

agg gtg gag ccg act cca gag ctg ctg aaa gat ttc cga gaa atg aga	576
Arg Val Glu Pro Thr Pro Glu Leu Leu Lys Asp Phe Arg Glu Met Arg	
180 185 190	

gct ctt ttc ctg agg gag caa ctt ttc aaa agt tgc aaa ttg tac tat	624
Ala Leu Phe Leu Arg Glu Gln Leu Phe Lys Ser Ser Lys Leu Tyr Tyr	
195 200 205	
ggt atg aag ctg ctc acg aat gtt gct att ttt gct gcg agc att gca	672
Val Met Lys Leu Leu Thr Asn Val Ala Ile Phe Ala Ala Ser Ile Ala	
210 215 220	
ata ata tgt tgg agc aag act att tca gcg gtt ttg gct tca gct tgt	720
Ile Ile Cys Trp Ser Lys Thr Ile Ser Ala Val Leu Ala Ser Ala Cys	
225 230 235 240	
atg atg gct ctg tgt ttc caa cag tgc gga tgg cta tcc cat gat ttt	768
Met Met Ala Leu Cys Phe Gln Gln Cys Gly Trp Leu Ser His Asp Phe	
245 250 255	
ctc cac aat cag gtg ttt gag aca cgc tgg ctt aat gaa gtt gtc ggg	816
Leu His Asn Gln Val Phe Glu Thr Arg Trp Leu Asn Glu Val Val Gly	
260 265 270	
tat gtg atc ggc aac gcc gtt ctg ggg ttt agt aca ggg tgg tgg aag	864
Tyr Val Ile Gly Asn Ala Val Leu Gly Phe Ser Thr Gly Trp Trp Lys	
275 280 285	
gag aag cat aac ctt cat cat gct gct cca aat gaa tgc gat cag act	912
Glu Lys His Asn Leu His His Ala Ala Pro Asn Glu Cys Asp Gln Thr	
290 295 300	
tac caa cca att gat gaa gat att gat act ctc ccc ctc att gcc tgg	960
Tyr Gln Pro Ile Asp Glu Asp Ile Asp Thr Leu Pro Leu Ile Ala Trp	
305 310 315 320	
agc aag gac ata ctg gcc aca gtt gag aat aag aca ttc ttg cga atc	1008
Ser Lys Asp Ile Leu Ala Thr Val Glu Asn Lys Thr Phe Leu Arg Ile	
325 330 335	
ctc caa tac cag cat ctg ttc ttc atg ggt ctg tta ttt ttc gcc cgt	1056
Leu Gln Tyr Gln His Leu Phe Phe Met Gly Leu Leu Phe Phe Ala Arg	
340 345 350	
ggt agt tgg ctc ttt tgg agc tgg aga tat acc tct aca gca gtg ctc	1104
Gly Ser Trp Leu Phe Trp Ser Trp Arg Tyr Thr Ser Thr Ala Val Leu	
355 360 365	
tca cct gtc gac agg ttg ttg gag aag gga act gtt ctg ttt cac tac	1152
Ser Pro Val Asp Arg Leu Leu Glu Lys Gly Thr Val Leu Phe His Tyr	
370 375 380	
ttt tgg ttc gtc ggg aca gcg tgc tat ctt ctc cct ggt tgg aag cca	1200
Phe Trp Phe Val Gly Thr Ala Cys Tyr Leu Leu Pro Gly Trp Lys Pro	
385 390 395 400	
tta gta tgg atg gcg gtg act gag ctc atg tcc ggc atg ctg ctg ggc	1248
Leu Val Trp Met Ala Val Thr Glu Leu Met Ser Gly Met Leu Leu Gly	
405 410 415	
ttt gta ttt gta ctt agc cac aat ggg atg gag gtt tat aat tgc tct	1296
Phe Val Phe Val Leu Ser His Asn Gly Met Glu Val Tyr Asn Ser Ser	
420 425 430	
aaa gaa ttc gtg agt gca cag atc gta tcc aca cgg gat atc aaa gga	1344
Lys Glu Phe Val Ser Ala Gln Ile Val Ser Thr Arg Asp Ile Lys Gly	
435 440 445	

aac ata ttc aac gac tgg ttc act ggt ggc ctt aac agg caa ata gag 1392
 Asn Ile Phe Asn Asp Trp Phe Thr Gly Gly Leu Asn Arg Gln Ile Glu
 450 455 460

cat cat ctt ttc cca aca atg ccc agg cat aat tta aac aaa ata gca 1440
 His His Leu Phe Pro Thr Met Pro Arg His Asn Leu Asn Lys Ile Ala
 465 470 475 480

cct aga gtg gag gtg ttc tgt aag aaa cac ggt ctg gtg tac gaa gac 1488
 Pro Arg Val Glu Val Phe Cys Lys Lys His Gly Leu Val Tyr Glu Asp
 485 490 495

gta tct att gct acc ggc act tgc aag gtt ttg aaa gca ttg aag gaa 1536
 Val Ser Ile Ala Thr Gly Thr Cys Lys Val Leu Lys Ala Leu Lys Glu
 500 505 510

gtc gcg gag gct gcg gca gag cag cat gct acc acc agt taa 1578
 Val Ala Glu Ala Ala Ala Glu Gln His Ala Thr Thr Ser
 515 520 525

<210> 24

<211> 525

<212> PRT

<213> Physcomitrella patens

<400> 24

Met Val Phe Ala Gly Gly Gly Leu Gln Gln Gly Ser Leu Glu Glu Asn
 1 5 10 15

Ile Asp Val Glu His Ile Ala Ser Met Ser Leu Phe Ser Asp Phe Phe
 20 25 30

Ser Tyr Val Ser Ser Thr Val Gly Ser Trp Ser Val His Ser Ile Gln
 35 40 45

Pro Leu Lys Arg Leu Thr Ser Lys Lys Arg Val Ser Glu Ser Ala Ala
 50 55 60

Val Gln Cys Ile Ser Ala Glu Val Gln Arg Asn Ser Ser Thr Gln Gly
 65 70 75 80

Thr Ala Glu Ala Leu Ala Glu Ser Val Val Lys Pro Thr Arg Arg Arg
 85 90 95

Ser Ser Gln Trp Lys Lys Ser Thr His Pro Leu Ser Glu Val Ala Val
 100 105 110

His Asn Lys Pro Ser Asp Cys Trp Ile Val Val Lys Asn Lys Val Tyr
 115 120 125

Asp Val Ser Asn Phe Ala Asp Glu His Pro Gly Gly Ser Val Ile Ser
 130 135 140

Thr Tyr Phe Gly Arg Asp Gly Thr Asp Val Phe Ser Ser Phe His Ala
 145 150 155 160

Ala Ser Thr Trp Lys Ile Leu Gln Asp Phe Tyr Ile Gly Asp Val Glu
 165 170 175

Arg Val Glu Pro Thr Pro Glu Leu Leu Lys Asp Phe Arg Glu Met Arg
 180 185 190

Ala Leu Phe Leu Arg Glu Gln Leu Phe Lys Ser Ser Lys Leu Tyr Tyr
 195 200 205

Val Met Lys Leu Leu Thr Asn Val Ala Ile Phe Ala Ala Ser Ile Ala
 210 215 220

Ile Ile Cys Trp Ser Lys Thr Ile Ser Ala Val Leu Ala Ser Ala Cys
 225 230 235 240

Met Met Ala Leu Cys Phe Gln Gln Cys Gly Trp Leu Ser His Asp Phe
 245 250 255

Leu His Asn Gln Val Phe Glu Thr Arg Trp Leu Asn Glu Val Val Gly
 260 265 270

Tyr Val Ile Gly Asn Ala Val Leu Gly Phe Ser Thr Gly Trp Trp Lys
 275 280 285

Glu Lys His Asn Leu His His Ala Ala Pro Asn Glu Cys Asp Gln Thr
 290 295 300

Tyr Gln Pro Ile Asp Glu Asp Ile Asp Thr Leu Pro Leu Ile Ala Trp
 305 310 315 320

Ser Lys Asp Ile Leu Ala Thr Val Glu Asn Lys Thr Phe Leu Arg Ile
 325 330 335

Leu Gln Tyr Gln His Leu Phe Phe Met Gly Leu Leu Phe Phe Ala Arg
 340 345 350

Gly Ser Trp Leu Phe Trp Ser Trp Arg Tyr Thr Ser Thr Ala Val Leu
 355 360 365

Ser Pro Val Asp Arg Leu Leu Glu Lys Gly Thr Val Leu Phe His Tyr
 370 375 380

Phe Trp Phe Val Gly Thr Ala Cys Tyr Leu Leu Pro Gly Trp Lys Pro
385 390 395 400

Leu Val Trp Met Ala Val Thr Glu Leu Met Ser Gly Met Leu Leu Gly
405 410 415

Phe Val Phe Val Leu Ser His Asn Gly Met Glu Val Tyr Asn Ser Ser
420 425 430

Lys Glu Phe Val Ser Ala Gln Ile Val Ser Thr Arg Asp Ile Lys Gly
435 440 445

Asn Ile Phe Asn Asp Trp Phe Thr Gly Gly Leu Asn Arg Gln Ile Glu
450 455 460

His His Leu Phe Pro Thr Met Pro Arg His Asn Leu Asn Lys Ile Ala
465 470 475 480

Pro Arg Val Glu Val Phe Cys Lys Lys His Gly Leu Val Tyr Glu Asp
485 490 495

Val Ser Ile Ala Thr Gly Thr Cys Lys Val Leu Lys Ala Leu Lys Glu
500 505 510

Val Ala Glu Ala Ala Ala Glu Gln His Ala Thr Thr Ser
515 520 525

<210> 25

<211> 1332

<212> DNA

<213> *Caenorhabditis elegans*

<220>

<221> CDS

<222> (1)..(1332)

<223> Delta-6 desaturase

<400> 25

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Met Val Val Asp Lys Asn Ala Ser Gly Leu Arg Met Lys Val Asp Gly
1 5 10 15

aaa tgg ctc tac ctt agc gag gaa ttg gtg aag aaa cat cca gga gga 96

51

Lys	Trp	Leu	Tyr	Leu	Ser	Glu	Glu	Leu	Val	Lys	Lys	His	Pro	Gly	Gly	
		20						25					30			
gct	gtt	att	gaa	caa	tat	aga	aat	tcg	gat	gct	act	cat	att	ttc	cac	144
Ala	Val	Ile	Glu	Gln	Tyr	Arg	Asn	Ser	Asp	Ala	Thr	His	Ile	Phe	His	
		35					40					45				
gct	ttc	cac	gaa	gga	tct	tct	cag	gct	tat	aag	caa	ctt	gac	ctt	ctg	192
Ala	Phe	His	Glu	Gly	Ser	Ser	Gln	Ala	Tyr	Lys	Gln	Leu	Asp	Leu	Leu	
	50					55					60					
aaa	aag	cac	gga	gag	cac	gat	gaa	ttc	ctt	gag	aaa	caa	ttg	gaa	aag	240
Lys	Lys	His	Gly	Glu	His	Asp	Glu	Phe	Leu	Glu	Lys	Gln	Leu	Glu	Lys	
65					70					75					80	
aga	ctt	gac	aaa	gtt	gat	atc	aat	gta	tca	gca	tat	gat	gtc	agt	gtt	288
Arg	Leu	Asp	Lys	Val	Asp	Ile	Asn	Val	Ser	Ala	Tyr	Asp	Val	Ser	Val	
			85					90					95			
gca	caa	gaa	aag	aaa	atg	gtt	gaa	tca	ttc	gaa	aaa	cta	cga	cag	aag	336
Ala	Gln	Glu	Lys	Lys	Met	Val	Glu	Ser	Phe	Glu	Lys	Leu	Arg	Gln	Lys	
			100					105					110			
ctt	cat	gat	gat	gga	tta	atg	aaa	gca	aat	gaa	aca	tat	ttc	ctg	ttt	384
Leu	His	Asp	Asp	Gly	Leu	Met	Lys	Ala	Asn	Glu	Thr	Tyr	Phe	Leu	Phe	
		115					120					125				
aaa	gcg	att	tca	aca	ctt	tca	att	atg	gca	ttt	gca	ttt	tat	ctt	cag	432
Lys	Ala	Ile	Ser	Thr	Leu	Ser	Ile	Met	Ala	Phe	Ala	Phe	Tyr	Leu	Gln	
	130					135					140					
tat	ctt	gga	tgg	tat	att	act	tct	gca	tgt	tta	tta	gca	ctt	gca	tgg	480
Tyr	Leu	Gly	Trp	Tyr	Ile	Thr	Ser	Ala	Cys	Leu	Leu	Ala	Leu	Ala	Trp	
145					150					155					160	
caa	caa	ttc	gga	tgg	tta	aca	cat	gag	ttc	tgc	cat	caa	cag	cca	aca	528
Gln	Gln	Phe	Gly	Trp	Leu	Thr	His	Glu	Phe	Cys	His	Gln	Gln	Pro	Thr	
			165					170					175			
aag	aac	aga	cct	ttg	aat	gat	act	att	tct	ttg	ttc	ttt	ggt	aat	ttc	576
Lys	Asn	Arg	Pro	Leu	Asn	Asp	Thr	Ile	Ser	Leu	Phe	Phe	Gly	Asn	Phe	
			180					185					190			
tta	caa	gga	ttt	tca	aga	gat	tgg	tgg	aag	gac	aag	cat	aac	act	cat	624
Leu	Gln	Gly	Phe	Ser	Arg	Asp	Trp	Trp	Lys	Asp	Lys	His	Asn	Thr	His	
		195					200					205				
cac	gct	gcc	aca	aat	gta	att	gat	cat	gac	ggt	gat	atc	gac	ttg	gca	672
His	Ala	Ala	Thr	Asn	Val	Ile	Asp	His	Asp	Gly	Asp	Ile	Asp	Leu	Ala	
		210				215					220					
cca	ctt	ttc	gca	ttt	att	cca	gga	gat	ttg	tgc	aag	tat	aag	gcc	agc	720
Pro	Leu	Phe	Ala	Phe	Ile	Pro	Gly	Asp	Leu	Cys	Lys	Tyr	Lys	Ala	Ser	
225					230					235					240	
ttt	gaa	aaa	gca	att	ctc	aag	att	gta	cca	tat	caa	cat	ctc	tat	ttc	768
Phe	Glu	Lys	Ala	Ile	Leu	Lys	Ile	Val	Pro	Tyr	Gln	His	Leu	Tyr	Phe	
				245				250						255		
acc	gca	atg	ctt	cca	atg	ctc	cgt	ttc	tca	tgg	act	ggt	cag	tca	gtt	816
Thr	Ala	Met	Leu	Pro	Met	Leu	Arg	Phe	Ser	Trp	Thr	Gly	Gln	Ser	Val	
			260					265					270			
caa	tgg	gta	ttc	aaa	gag	aat	caa	atg	gag	tac	aag	gtc	tat	caa	aga	864

52

Gln Trp Val Phe Lys Glu Asn Gln Met Glu Tyr Lys Val Tyr Gln Arg
 275 280 285
 aat gca ttc tgg gag caa gca aca att gtt gga cat tgg gct tgg gta 912
 Asn Ala Phe Trp Glu Gln Ala Thr Ile Val Gly His Trp Ala Trp Val
 290 295 300
 ttc tat caa ttg ttc tta tta cca aca tgg cca ctt cgg gtt gct tat 960
 Phe Tyr Gln Leu Phe Leu Leu Pro Thr Trp Pro Leu Arg Val Ala Tyr
 305 310 315 320
 ttc att att tca caa atg gga gga ggc ctt ttg att gct cac gta gtc 1008
 Phe Ile Ile Ser Gln Met Gly Gly Gly Leu Leu Ile Ala His Val Val
 325 330 335
 act ttc aac cat aac tct gtt gat aag tat cca gcc aat tct cga att 1056
 Thr Phe Asn His Asn Ser Val Asp Lys Tyr Pro Ala Asn Ser Arg Ile
 340 345 350
 tta aac aac ttc gcc gct ctt caa att ttg acc aca cgc aac atg act 1104
 Leu Asn Asn Phe Ala Ala Leu Gln Ile Leu Thr Thr Arg Asn Met Thr
 355 360 365
 cca tct cca ttc att gat tgg ctt tgg ggt gga ctc aat tat cag atc 1152
 Pro Ser Pro Phe Ile Asp Trp Leu Trp Gly Gly Leu Asn Tyr Gln Ile
 370 375 380
 gag cac cac ttg ttc cca aca atg cca cgt tgc aat ctg aat gct tgc 1200
 Glu His His Leu Phe Pro Thr Met Pro Arg Cys Asn Leu Asn Ala Cys
 385 390 395 400
 gtg aaa tat gtg aaa gaa tgg tgc aaa gag aat aat ctt cct tac ctc 1248
 Val Lys Tyr Val Lys Glu Trp Cys Lys Glu Asn Asn Leu Pro Tyr Leu
 405 410 415
 gtc gat gac tac ttt gac gga tat gca atg aat ttg caa caa ttg aaa 1296
 Val Asp Asp Tyr Phe Asp Gly Tyr Ala Met Asn Leu Gln Gln Leu Lys
 420 425 430
 aat atg gct gag cac att caa gct aaa gct gcc taa 1332
 Asn Met Ala Glu His Ile Gln Ala Lys Ala Ala
 435 440

<210> 26

<211> 443

<212> PRT

<213> Caenorhabditis elegans

<400> 26

Met Val Val Asp Lys Asn Ala Ser Gly Leu Arg Met Lys Val Asp Gly
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 Lys Trp Leu Tyr Leu Ser Glu Glu Leu Val Lys Lys His Pro Gly Gly
 20 25 30

53

Ala Val Ile Glu Gln Tyr Arg Asn Ser Asp Ala Thr His Ile Phe His
 35 40 45

Ala Phe His Glu Gly Ser Ser Gln Ala Tyr Lys Gln Leu Asp Leu Leu
 50 55 60

Lys Lys His Gly Glu His Asp Glu Phe Leu Glu Lys Gln Leu Glu Lys
 65 70 75 80

Arg Leu Asp Lys Val Asp Ile Asn Val Ser Ala Tyr Asp Val Ser Val
 85 90 95

Ala Gln Glu Lys Lys Met Val Glu Ser Phe Glu Lys Leu Arg Gln Lys
 100 105 110

Leu His Asp Asp Gly Leu Met Lys Ala Asn Glu Thr Tyr Phe Leu Phe
 115 120 125

Lys Ala Ile Ser Thr Leu Ser Ile Met Ala Phe Ala Phe Tyr Leu Gln
 130 135 140

Tyr Leu Gly Trp Tyr Ile Thr Ser Ala Cys Leu Leu Ala Leu Ala Trp
 145 150 155 160

Gln Gln Phe Gly Trp Leu Thr His Glu Phe Cys His Gln Gln Pro Thr
 165 170 175

Lys Asn Arg Pro Leu Asn Asp Thr Ile Ser Leu Phe Phe Gly Asn Phe
 180 185 190

Leu Gln Gly Phe Ser Arg Asp Trp Trp Lys Asp Lys His Asn Thr His
 195 200 205

His Ala Ala Thr Asn Val Ile Asp His Asp Gly Asp Ile Asp Leu Ala
 210 215 220

Pro Leu Phe Ala Phe Ile Pro Gly Asp Leu Cys Lys Tyr Lys Ala Ser
 225 230 235 240

Phe Glu Lys Ala Ile Leu Lys Ile Val Pro Tyr Gln His Leu Tyr Phe
 245 250 255

Thr Ala Met Leu Pro Met Leu Arg Phe Ser Trp Thr Gly Gln Ser Val
 260 265 270

Gln Trp Val Phe Lys Glu Asn Gln Met Glu Tyr Lys Val Tyr Gln Arg
 275 280 285

54

Asn Ala Phe Trp Glu Gln Ala Thr Ile Val Gly His Trp Ala Trp Val
290 295 300

Phe Tyr Gln Leu Phe Leu Leu Pro Thr Trp Pro Leu Arg Val Ala Tyr
305 310 315 320

Phe Ile Ile Ser Gln Met Gly Gly Gly Leu Leu Ile Ala His Val Val
325 330 335

Thr Phe Asn His Asn Ser Val Asp Lys Tyr Pro Ala Asn Ser Arg Ile
340 345 350

Leu Asn Asn Phe Ala Ala Leu Gln Ile Leu Thr Thr Arg Asn Met Thr
355 360 365

Pro Ser Pro Phe Ile Asp Trp Leu Trp Gly Gly Leu Asn Tyr Gln Ile
370 375 380

Glu His His Leu Phe Pro Thr Met Pro Arg Cys Asn Leu Asn Ala Cys
385 390 395 400

Val Lys Tyr Val Lys Glu Trp Cys Lys Glu Asn Asn Leu Pro Tyr Leu
405 410 415

Val Asp Asp Tyr Phe Asp Gly Tyr Ala Met Asn Leu Gln Gln Leu Lys
420 425 430

Asn Met Ala Glu His Ile Gln Ala Lys Ala Ala
435 440

<210> 27

<211> 873

<212> DNA

<213> Physcomitrella patens

<220>

<221> CDS

<222> (1)..(873)

<223> Delta-6 elongase

<400> 27

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Met Glu Val Val Glu Arg Phe Tyr Gly Glu Leu Asp Gly Lys Val Ser
1 5 10 15

cag ggc gtg aat gca ttg ctg ggt agt ttt ggg gtg gag ttg acg gat	96
Gln Gly Val Asn Ala Leu Leu Gly Ser Phe Gly Val Glu Leu Thr Asp	
20 25 30	
acg ccc act acc aaa ggc ttg ccc ctc gtt gac agt ccc aca ccc atc	144
Thr Pro Thr Thr Lys Gly Leu Pro Leu Val Asp Ser Pro Thr Pro Ile	
35 40 45	
gtc ctc ggt gtt tct gta tac ttg act att gtc att gga ggg ctt ttg	192
Val Leu Gly Val Ser Val Tyr Leu Thr Ile Val Ile Gly Gly Leu Leu	
50 55 60	
tgg ata aag gcc agg gat ctg aaa ccg cgc gcc tcg gag cca ttt ttg	240
Trp Ile Lys Ala Arg Asp Leu Lys Pro Arg Ala Ser Glu Pro Phe Leu	
65 70 75 80	
ctc caa gct ttg gtg ctt gtg cac aac ctg ttc tgt ttt gcg ctc agt	288
Leu Gln Ala Leu Val Leu Val His Asn Leu Phe Cys Phe Ala Leu Ser	
85 90 95	
ctg tat atg tgc gtg ggc atc gct tat cag gct att acc tgg cgg tac	336
Leu Tyr Met Cys Val Gly Ile Ala Tyr Gln Ala Ile Thr Trp Arg Tyr	
100 105 110	
tct ctc tgg ggc aat gca tac aat cct aaa cat aaa gag atg gcg att	384
Ser Leu Trp Gly Asn Ala Tyr Asn Pro Lys His Lys Glu Met Ala Ile	
115 120 125	
ctg gta tac ttg ttc tac atg tct aag tac gtg gaa ttc atg gat acc	432
Leu Val Tyr Leu Phe Tyr Met Ser Lys Tyr Val Glu Phe Met Asp Thr	
130 135 140	
gtt atc atg ata ctg aag cgc agc acc agg caa ata agc ttc ctc cac	480
Val Ile Met Ile Leu Lys Arg Ser Thr Arg Gln Ile Ser Phe Leu His	
145 150 155 160	
gtt tat cat cat tct tca att tcc ctc att tgg tgg gct att gct cat	528
Val Tyr His His Ser Ser Ile Ser Leu Ile Trp Trp Ala Ile Ala His	
165 170 175	
cac gct cct ggc ggt gaa gca tat tgg tct gcg gct ctg aac tca gga	576
His Ala Pro Gly Gly Glu Ala Tyr Trp Ser Ala Ala Leu Asn Ser Gly	
180 185 190	
gtg cat gtt ctc atg tat gcg tat tac ttc ttg gct gcc tgc ctt cga	624
Val His Val Leu Met Tyr Ala Tyr Tyr Phe Leu Ala Ala Cys Leu Arg	
195 200 205	
agt agc cca aag tta aaa aat aag tac ctt ttt tgg ggc agg tac ttg	672
Ser Ser Pro Lys Leu Lys Asn Lys Tyr Leu Phe Trp Gly Arg Tyr Leu	
210 215 220	
aca caa ttc caa atg ttc cag ttt atg ctg aac tta gtg cag gct tac	720
Thr Gln Phe Gln Met Phe Gln Phe Met Leu Asn Leu Val Gln Ala Tyr	
225 230 235 240	
tac gac atg aaa acg aat gcg cca tat cca caa tgg ctg atc aag att	768
Tyr Asp Met Lys Thr Asn Ala Pro Tyr Pro Gln Trp Leu Ile Lys Ile	
245 250 255	
ttg ttc tac tac atg atc tcg ttg ctg ttt ctt ttc ggc aat ttt tac	816
Leu Phe Tyr Tyr Met Ile Ser Leu Leu Phe Leu Phe Gly Asn Phe Tyr	
260 265 270	

gta caa aaa tac atc aaa ccc tct gac gga aag caa aag gga gct aaa 864
Val Gln Lys Tyr Ile Lys Pro Ser Asp Gly Lys Gln Lys Gly Ala Lys
275 280 285

act gag tga 873
Thr Glu
290

<210> 28

<211> 290

<212> PRT

<213> Physcomitrella patens

<400> 28

Met Glu Val Val Glu Arg Phe Tyr Gly Glu Leu Asp Gly Lys Val Ser
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Gln Gly Val Asn Ala Leu Leu Gly Ser Phe Gly Val Glu Leu Thr Asp
20 25 30

Thr Pro Thr Thr Lys Gly Leu Pro Leu Val Asp Ser Pro Thr Pro Ile
35 40 45

Val Leu Gly Val Ser Val Tyr Leu Thr Ile Val Ile Gly Gly Leu Leu
50 55 60

Trp Ile Lys Ala Arg Asp Leu Lys Pro Arg Ala Ser Glu Pro Phe Leu
65 70 75 80

Leu Gln Ala Leu Val Leu Val His Asn Leu Phe Cys Phe Ala Leu Ser
85 90 95

Leu Tyr Met Cys Val Gly Ile Ala Tyr Gln Ala Ile Thr Trp Arg Tyr
100 105 110

Ser Leu Trp Gly Asn Ala Tyr Asn Pro Lys His Lys Glu Met Ala Ile
115 120 125

Leu Val Tyr Leu Phe Tyr Met Ser Lys Tyr Val Glu Phe Met Asp Thr
130 135 140

Val Ile Met Ile Leu Lys Arg Ser Thr Arg Gln Ile Ser Phe Leu His
145 150 155 160

Val Tyr His His Ser Ser Ile Ser Leu Ile Trp Trp Ala Ile Ala His
165 170 175

His Ala Pro Gly Gly Glu Ala Tyr Trp Ser Ala Ala Leu Asn Ser Gly
180 185 190

Val His Val Leu Met Tyr Ala Tyr Tyr Phe Leu Ala Ala Cys Leu Arg
195 200 205

Ser Ser Pro Lys Leu Lys Asn Lys Tyr Leu Phe Trp Gly Arg Tyr Leu
210 215 220

Thr Gln Phe Gln Met Phe Gln Phe Met Leu Asn Leu Val Gln Ala Tyr
225 230 235 240

Tyr Asp Met Lys Thr Asn Ala Pro Tyr Pro Gln Trp Leu Ile Lys Ile
245 250 255

Leu Phe Tyr Tyr Met Ile Ser Leu Leu Phe Leu Phe Gly Asn Phe Tyr
260 265 270

Val Gln Lys Tyr Ile Lys Pro Ser Asp Gly Lys Gln Lys Gly Ala Lys
275 280 285

Thr Glu
290

<210> 29

<211> 1049

<212> DNA

<213> Thraustochytrium

<220>

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<222> (43) . . (858)

<223> Delta-6 elongase

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              Met Met Glu Pro
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ctc gac agg tac agg gcg ctg gcg gag ctc gcc gcg agg tac gcc agc 102
Leu Asp Arg Tyr Arg Ala Leu Ala Glu Leu Ala Ala Arg Tyr Ala Ser
5 10 15 20

tcg gcg gcc ttc aag tgg caa gtc acg tac gac gcc aag gac agc ttc 150

58

Ser	Ala	Ala	Phe	Lys	Trp	Gln	Val	Thr	Tyr	Asp	Ala	Lys	Asp	Ser	Phe		
				25					30					35			
gtc	ggg	ccc	ctg	gga	atc	cgg	gag	ccg	ctc	ggg	ctc	ctg	gtg	ggc	tcc	198	
Val	Gly	Pro	Leu	Gly	Ile	Arg	Glu	Pro	Leu	Gly	Leu	Leu	Val	Gly	Ser		
				40					45					50			
gtg	gtc	ctc	tac	ctg	agc	ctg	ctg	gcc	gtg	gtc	tac	gcg	ctg	cgg	aac	246	
Val	Val	Leu	Tyr	Leu	Ser	Leu	Leu	Ala	Val	Val	Tyr	Ala	Leu	Arg	Asn		
				55					60					65			
tac	ctt	ggc	ggc	ctc	atg	gcg	ctc	cgc	agc	gtg	cat	aac	ctc	ggg	ctc	294	
Tyr	Leu	Gly	Gly	Leu	Met	Ala	Leu	Arg	Ser	Val	His	Asn	Leu	Gly	Leu		
				70					75					80			
tgc	ctc	ttc	tcg	ggc	gcc	gtg	tgg	atc	tac	acg	agc	tac	ctc	atg	atc	342	
Cys	Leu	Phe	Ser	Gly	Ala	Val	Trp	Ile	Tyr	Thr	Ser	Tyr	Leu	Met	Ile		
				85					90					95			100
cag	gat	ggg	cac	ttt	cgc	agc	ctc	gag	gcg	gca	acg	tgc	gag	ccg	ctc	390	
Gln	Asp	Gly	His	Phe	Arg	Ser	Leu	Glu	Ala	Ala	Thr	Cys	Glu	Pro	Leu		
				105					110					115			
aag	cat	ccg	cac	ttc	cag	ctc	atc	agc	ttg	ctc	ttt	gcg	ctg	tcc	aag	438	
Lys	His	Pro	His	Phe	Gln	Leu	Ile	Ser	Leu	Leu	Phe	Ala	Leu	Ser	Lys		
				120					125					130			
atc	tgg	gag	tgg	ttc	gac	acg	gtg	ctc	ctc	atc	gtc	aag	ggc	aac	aag	486	
Ile	Trp	Glu	Trp	Phe	Asp	Thr	Val	Leu	Leu	Ile	Val	Lys	Gly	Asn	Lys		
				135					140					145			
ctc	cgc	ttc	ctg	cac	gtc	ttg	cac	cac	gcc	acg	acc	ttt	tgg	ctc	tac	534	
Leu	Arg	Phe	Leu	His	Val	Leu	His	His	Ala	Thr	Thr	Phe	Trp	Leu	Tyr		
				150					155					160			
gcc	atc	gac	cac	atc	ttt	ctc	tcg	tcc	atc	aag	tac	ggc	gtc	gcg	gtc	582	
Ala	Ile	Asp	His	Ile	Phe	Leu	Ser	Ser	Ile	Lys	Tyr	Gly	Val	Ala	Val		
				165					170					175			180
aat	gct	ttc	atc	cac	acc	gtc	atg	tac	gcg	cac	tac	ttc	cgc	cca	ttc	630	
Asn	Ala	Phe	Ile	His	Thr	Val	Met	Tyr	Ala	His	Tyr	Phe	Arg	Pro	Phe		
				185					190					195			
ccg	aag	ggc	ttg	cgc	ccg	ctt	att	acg	cag	ttg	cag	atc	gtc	cag	ttc	678	
Pro	Lys	Gly	Leu	Arg	Pro	Leu	Ile	Thr	Gln	Leu	Gln	Ile	Val	Gln	Phe		
				200					205					210			
att	ttc	agc	atc	ggc	atc	cat	acc	gcc	att	tac	tgg	cac	tac	gac	tgc	726	
Ile	Phe	Ser	Ile	Gly	Ile	His	Thr	Ala	Ile	Tyr	Trp	His	Tyr	Asp	Cys		
				215					220					225			
gag	ccg	ctc	gtg	cat	acc	cac	ttt	tgg	gaa	tac	gtc	acg	ccc	tac	ctt	774	
Glu	Pro	Leu	Val	His	Thr	His	Phe	Trp	Glu	Tyr	Val	Thr	Pro	Tyr	Leu		
				230					235					240			
ttc	gtc	gtg	ccc	ttc	ctc	atc	ctc	ttt	ttc	aat	ttt	tac	ctg	cag	cag	822	
Phe	Val	Val	Pro	Phe	Leu	Ile	Leu	Phe	Phe	Asn	Phe	Tyr					

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<210> 30

<211> 271

<212> PRT

<213> Thraustochytrium

<400> 30

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Arg Tyr Ala Ser Ser Ala Ala Phe Lys Trp Gln Val Thr Tyr Asp Ala
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Lys Asp Ser Phe Val Gly Pro Leu Gly Ile Arg Glu Pro Leu Gly Leu
 35 40 45

Leu Val Gly Ser Val Val Leu Tyr Leu Ser Leu Leu Ala Val Val Tyr
 50 55 60

Ala Leu Arg Asn Tyr Leu Gly Gly Leu Met Ala Leu Arg Ser Val His
 65 70 75 80

Asn Leu Gly Leu Cys Leu Phe Ser Gly Ala Val Trp Ile Tyr Thr Ser
 85 90 95

Tyr Leu Met Ile Gln Asp Gly His Phe Arg Ser Leu Glu Ala Ala Thr
 100 105 110

Cys Glu Pro Leu Lys His Pro His Phe Gln Leu Ile Ser Leu Leu Phe
 115 120 125

Ala Leu Ser Lys Ile Trp Glu Trp Phe Asp Thr Val Leu Leu Ile Val
 130 135 140

Lys Gly Asn Lys Leu Arg Phe Leu His Val Leu His His Ala Thr Thr
 145 150 155 160

Phe Trp Leu Tyr Ala Ile Asp His Ile Phe Leu Ser Ser Ile Lys Tyr
 165 170 175

60

Gly Val Ala Val Asn Ala Phe Ile His Thr Val Met Tyr Ala His Tyr
 180 185 190

Phe Arg Pro Phe Pro Lys Gly Leu Arg Pro Leu Ile Thr Gln Leu Gln
 195 200 205

Ile Val Gln Phe Ile Phe Ser Ile Gly Ile His Thr Ala Ile Tyr Trp
 210 215 220

His Tyr Asp Cys Glu Pro Leu Val His Thr His Phe Trp Glu Tyr Val
 225 230 235 240

Thr Pro Tyr Leu Phe Val Val Pro Phe Leu Ile Leu Phe Phe Asn Phe
 245 250 255

Tyr Leu Gln Gln Tyr Val Leu Ala Pro Ala Lys Thr Lys Lys Ala
 260 265 270

<210> 31

<211> 837

<212> DNA

<213> *Phytophthora infestans*

<220>

<221> CDS

<222> (1)..(837)

<223> Delta-6 elongase

<400> 31

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 1 5 10 15

gag gcc aag ctg ctg gac tgg gtc gac cct gag ggc ggc tgg aag gtg 96
 Glu Ala Lys Leu Leu Asp Trp Val Asp Pro Glu Gly Gly Trp Lys Val
 20 25 30

cat cct atg gca gac tac ccc cta gcc aac ttc tcc agc gtc tac gcc 144
 His Pro Met Ala Asp Tyr Pro Leu Ala Asn Phe Ser Ser Val Tyr Ala
 35 40 45

atc tgc gtc gga tac ttg ctc ttc gta atc ttc ggc acg gcc ctg atg 192
 Ile Cys Val Gly Tyr Leu Leu Phe Val Ile Phe Gly Thr Ala Leu Met
 50 55 60

aaa atg gga gtc ccc gcc atc aag acc agt cca tta cag ttt gtg tac 240
 Lys Met Gly Val Pro Ala Ile Lys Thr Ser Pro Leu Gln Phe Val Tyr
 65 70 75 80


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aac ccc atc caa gtc att gcc tgc tct tat atg tgc gtg gag gcc gcc      288
Asn Pro Ile Gln Val Ile Ala Cys Ser Tyr Met Cys Val Glu Ala Ala
                        85                      90                      95

atc cag gcc tac cgc aac ggc tac acc gcc gcc ccg tgc aac gcc ttt      336
Ile Gln Ala Tyr Arg Asn Gly Tyr Thr Ala Ala Pro Cys Asn Ala Phe
                        100                      105                      110

aag tcc gac gac ccc gtc atg ggc aac gtt ctg tac ctc ttc tat ctc      384
Lys Ser Asp Asp Pro Val Met Gly Asn Val Leu Tyr Leu Phe Tyr Leu
                        115                      120                      125

tcc aag atg ctc gac ctg tgc gac aca gtc ttc att atc cta gga aag      432
Ser Lys Met Leu Asp Leu Cys Asp Thr Val Phe Ile Ile Leu Gly Lys
                        130                      135                      140

aag tgg aaa cag ctt tcc atc ttg cac gtg tac cac cac ctt acc gtg      480
Lys Trp Lys Gln Leu Ser Ile Leu His Val Tyr His His Leu Thr Val
                        145                      150                      155                      160

ctt ttc gtc tac tat gtg acg ttc cgc gcc gct cag gac ggg gac tca      528
Leu Phe Val Tyr Tyr Val Thr Phe Arg Ala Ala Gln Asp Gly Asp Ser
                        165                      170                      175

tat gct acc atc gtg ctc aac ggc ttc gtg cac acc atc atg tac act      576
Tyr Ala Thr Ile Val Leu Asn Gly Phe Val His Thr Ile Met Tyr Thr
                        180                      185                      190

tac tac ttc gtc agc gcc cac acg cgc aac att tgg tgg aag aag tac      624
Tyr Tyr Phe Val Ser Ala His Thr Arg Asn Ile Trp Trp Lys Lys Tyr
                        195                      200                      205

ctc acg cgc att cag ctt atc cag ttc gtg acc atg aac gtg cag ggc      672
Leu Thr Arg Ile Gln Leu Ile Gln Phe Val Thr Met Asn Val Gln Gly
                        210                      215                      220

tac ctg acc tac tct cga cag tgc cca ggc atg cct cct aag gtg ccg      720
Tyr Leu Thr Tyr Ser Arg Gln Cys Pro Gly Met Pro Pro Lys Val Pro
                        225                      230                      235                      240

ctc atg tac ctt gtg tac gtg cag tca ctc ttc tgg ctc ttc atg aat      768
Leu Met Tyr Leu Val Tyr Val Gln Ser Leu Phe Trp Leu Phe Met Asn
                        245                      250                      255

ttc tac att cgc gcg tac gtg ttc ggc ccc aag aaa ccg gcc gtg gag      816
Phe Tyr Ile Arg Ala Tyr Val Phe Gly Pro Lys Lys Pro Ala Val Glu
                        260                      265                      270

gaa tcg aag aag aag ttg taa      837
Glu Ser Lys Lys Lys Leu
                        275

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<210> 32

<211> 278

<212> PRT

<213> Phytophthora infestans

<400> 32

Met Ser Thr Glu Leu Leu Gln Ser Tyr Tyr Ala Trp Ala Asn Ala Thr
 1 5 10 15

Glu Ala Lys Leu Leu Asp Trp Val Asp Pro Glu Gly Gly Trp Lys Val
 20 25 30

His Pro Met Ala Asp Tyr Pro Leu Ala Asn Phe Ser Ser Val Tyr Ala
 35 40 45

Ile Cys Val Gly Tyr Leu Leu Phe Val Ile Phe Gly Thr Ala Leu Met
 50 55 60

Lys Met Gly Val Pro Ala Ile Lys Thr Ser Pro Leu Gln Phe Val Tyr
 65 70 75 80

Asn Pro Ile Gln Val Ile Ala Cys Ser Tyr Met Cys Val Glu Ala Ala
 85 90 95

Ile Gln Ala Tyr Arg Asn Gly Tyr Thr Ala Ala Pro Cys Asn Ala Phe
 100 105 110

Lys Ser Asp Asp Pro Val Met Gly Asn Val Leu Tyr Leu Phe Tyr Leu
 115 120 125

Ser Lys Met Leu Asp Leu Cys Asp Thr Val Phe Ile Ile Leu Gly Lys
 130 135 140

Lys Trp Lys Gln Leu Ser Ile Leu His Val Tyr His His Leu Thr Val
 145 150 155 160

Leu Phe Val Tyr Tyr Val Thr Phe Arg Ala Ala Gln Asp Gly Asp Ser
 165 170 175

Tyr Ala Thr Ile Val Leu Asn Gly Phe Val His Thr Ile Met Tyr Thr
 180 185 190

Tyr Tyr Phe Val Ser Ala His Thr Arg Asn Ile Trp Trp Lys Lys Tyr
 195 200 205

Leu Thr Arg Ile Gln Leu Ile Gln Phe Val Thr Met Asn Val Gln Gly
 210 215 220

Tyr Leu Thr Tyr Ser Arg Gln Cys Pro Gly Met Pro Pro Lys Val Pro
 225 230 235 240

Leu Met Tyr Leu Val Tyr Val Gln Ser Leu Phe Trp Leu Phe Met Asn
 245 250 255

Phe Tyr Ile Arg Ala Tyr Val Phe Gly Pro Lys Lys Pro Ala Val Glu
 260 265 270

Glu Ser Lys Lys Lys Leu
 275

<210> 33

<211> 954

<212> DNA

<213> Mortierella alpina

<220>

<221> CDS

<222> (1)..(954)

<223> Delta-6 elongase

<400> 33

atg gcc gcc gca atc ttg gac aag gtc aac ttc ggc att gat cag ccc 48
 Met Ala Ala Ala Ile Leu Asp Lys Val Asn Phe Gly Ile Asp Gln Pro
 1 5 10 15

ttc gga atc aag ctc gac acc tac ttt gct cag gcc tat gaa ctc gtc 96
 Phe Gly Ile Lys Leu Asp Thr Tyr Phe Ala Gln Ala Tyr Glu Leu Val
 20 25 30

acc gga aag tcc atc gac tcc ttc gtc ttc cag gag ggc gtc acg cct 144
 Thr Gly Lys Ser Ile Asp Ser Phe Val Phe Gln Glu Gly Val Thr Pro
 35 40 45

ctc tcg acc cag aga gag gtc gcc atg tgg act atc act tac ttc gtc 192
 Leu Ser Thr Gln Arg Glu Val Ala Met Trp Thr Ile Thr Tyr Phe Val
 50 55 60

gtc atc ttt ggt ggt cgc cag atc atg aag agc cag gac gcc ttc aag 240
 Val Ile Phe Gly Gly Arg Gln Ile Met Lys Ser Gln Asp Ala Phe Lys
 65 70 75 80

ctc aag ccc ctc ttc atc ctc cac aac ttc ctc ctg acg atc gcg tcc 288
 Leu Lys Pro Leu Phe Ile Leu His Asn Phe Leu Leu Thr Ile Ala Ser
 85 90 95

gga tcg ctg ttg ctc ctg ttc atc gag aac ctg gtc ccc atc ctc gcc 336
 Gly Ser Leu Leu Leu Leu Phe Ile Glu Asn Leu Val Pro Ile Leu Ala
 100 105 110

aga aac gga ctt ttc tac gcc atc tgc gac gac ggt gcc tgg acc cag 384
 Arg Asn Gly Leu Phe Tyr Ala Ile Cys Asp Asp Gly Ala Trp Thr Gln
 115 120 125

cgc ctc gag ctc ctc tac tac ctc aac tac ctg gtc aag tac tgg gag 432

64

Arg	Leu	Glu	Leu	Leu	Tyr	Tyr	Leu	Asn	Tyr	Leu	Val	Lys	Tyr	Trp	Glu		
130						135					140						
ttg	gcc	gac	acc	gtc	ttt	ttg	gtc	ctc	aag	aag	aag	cct	ctt	gag	ttc	480	
Leu	Ala	Asp	Thr	Val	Phe	Leu	Val	Leu	Lys	Lys	Lys	Pro	Leu	Glu	Phe		
145					150				155					160			
ctg	cac	tac	ttc	cac	cac	tcg	atg	acc	atg	gtt	ctc	tgc	ttt	gtc	cag	528	
Leu	His	Tyr	Phe	His	His	Ser	Met	Thr	Met	Val	Leu	Cys	Phe	Val	Gln		
				165					170					175			
ctt	gga	gga	tac	act	tca	gtg	tcc	tgg	gtc	cct	att	acc	ctc	aac	ttg	576	
Leu	Gly	Gly	Tyr	Thr	Ser	Val	Ser	Trp	Val	Pro	Ile	Thr	Leu	Asn	Leu		
			180					185					190				
act	gtc	cac	gtc	ttc	atg	tac	tac	tac	tac	atg	cgc	tcc	gct	gcc	ggc	624	
Thr	Val	His	Val	Phe	Met	Tyr	Tyr	Tyr	Tyr	Met	Arg	Ser	Ala	Ala	Gly		
		195				200						205					
gtt	cgc	atc	tgg	tgg	aag	cag	tac	ttg	acc	act	ctc	cag	atc	gtc	cag	672	
Val	Arg	Ile	Trp	Trp	Lys	Gln	Tyr	Leu	Thr	Thr	Leu	Gln	Ile	Val	Gln		
	210					215					220						
ttc	gtt	ctt	gac	ctc	gga	ttc	atc	tac	ttc	tgc	gcc	tac	acc	tac	ttc	720	
Phe	Val	Leu	Asp	Leu	Gly	Phe	Ile	Tyr	Phe	Cys	Ala	Tyr	Thr	Tyr	Phe		
225					230					235				240			
gcc	ttc	acc	tac	ttc	ccc	tgg	gct	ccc	aac	gtc	ggc	aag	tgc	gcc	ggc	768	
Ala	Phe	Thr	Tyr	Phe	Pro	Trp	Ala	Pro	Asn	Val	Gly	Lys	Cys	Ala	Gly		
				245					250					255			
acc	gag	ggt	gct	gct	ctc	ttt	ggc	tgc	gga	ctc	ctc	tcc	agc	tat	ctc	816	
Thr	Glu	Gly	Ala	Ala	Leu	Phe	Gly	Cys	Gly	Leu	Leu	Ser	Ser	Tyr	Leu		
			260					265					270				
ttg	ctc	ttt	atc	aac	ttc	tac	cgc	att	acc	tac	aat	gcc	aag	gcc	aag	864	
Leu	Leu	Phe	Ile	Asn	Phe	Tyr	Arg	Ile	Thr	Tyr	Asn	Ala	Lys	Ala	Lys		
		275					280					285					
gca	gcc	aag	gag	cgt	gga	agc	aac	ttt	acc	ccc	aag	act	gtc	aag	tcc	912	
Ala	Ala	Lys	Glu	Arg	Gly	Ser	Asn	Phe	Thr	Pro	Lys	Thr	Val	Lys	Ser		
		290				295					300						
ggc	gga	tcg	ccc	aag	aag	ccc	tcc	aag	agc	aag	cac	atc	taa			954	
Gly	Gly	Ser	Pro	Lys	Lys	Pro	Ser	Lys	Ser	Lys	His	Ile					
305					310					315							

<210> 34

<211> 317

<212> PRT

<213> Mortierella alpina

<400> 34

Met	Ala	Ala	Ala	Ile	Leu	Asp	Lys	Val	Asn	Phe	Gly	Ile	Asp	Gln	Pro
1				5					10					15	

65

Phe Gly Ile Lys Leu Asp Thr Tyr Phe Ala Gln Ala Tyr Glu Leu Val
 20 25 30

Thr Gly Lys Ser Ile Asp Ser Phe Val Phe Gln Glu Gly Val Thr Pro
 35 40 45

Leu Ser Thr Gln Arg Glu Val Ala Met Trp Thr Ile Thr Tyr Phe Val
 50 55 60

Val Ile Phe Gly Gly Arg Gln Ile Met Lys Ser Gln Asp Ala Phe Lys
 65 70 75 80

Leu Lys Pro Leu Phe Ile Leu His Asn Phe Leu Leu Thr Ile Ala Ser
 85 90 95

Gly Ser Leu Leu Leu Leu Phe Ile Glu Asn Leu Val Pro Ile Leu Ala
 100 105 110

Arg Asn Gly Leu Phe Tyr Ala Ile Cys Asp Asp Gly Ala Trp Thr Gln
 115 120 125

Arg Leu Glu Leu Leu Tyr Tyr Leu Asn Tyr Leu Val Lys Tyr Trp Glu
 130 135 140

Leu Ala Asp Thr Val Phe Leu Val Leu Lys Lys Lys Pro Leu Glu Phe
 145 150 155 160

Leu His Tyr Phe His His Ser Met Thr Met Val Leu Cys Phe Val Gln
 165 170 175

Leu Gly Gly Tyr Thr Ser Val Ser Trp Val Pro Ile Thr Leu Asn Leu
 180 185 190

Thr Val His Val Phe Met Tyr Tyr Tyr Tyr Met Arg Ser Ala Ala Gly
 195 200 205

Val Arg Ile Trp Trp Lys Gln Tyr Leu Thr Thr Leu Gln Ile Val Gln
 210 215 220

Phe Val Leu Asp Leu Gly Phe Ile Tyr Phe Cys Ala Tyr Thr Tyr Phe
 225 230 235 240

Ala Phe Thr Tyr Phe Pro Trp Ala Pro Asn Val Gly Lys Cys Ala Gly
 245 250 255

Thr Glu Gly Ala Ala Leu Phe Gly Cys Gly Leu Leu Ser Ser Tyr Leu
 260 265 270

66

Leu Leu Phe Ile Asn Phe Tyr Arg Ile Thr Tyr Asn Ala Lys Ala Lys
 275 280 285

Ala Ala Lys Glu Arg Gly Ser Asn Phe Thr Pro Lys Thr Val Lys Ser
 290 295 300

Gly Gly Ser Pro Lys Lys Pro Ser Lys Ser Lys His Ile
 305 310 315

<210> 35

<211> 957

<212> DNA

<213> Mortierella alpina

<220>

<221> CDS

<222> (1)..(957)

<223> Delta-6 elongase

<400> 35

atg gag tcg att gcg cca ttc ctc cca tca aag atg ccg caa gat ctg 48
 Met Glu Ser Ile Ala Pro Phe Leu Pro Ser Lys Met Pro Gln Asp Leu
 1 5 10 15

ttt atg gac ctt gcc acc gct atc ggt gtc cgg gcc gcg ccc tat gtc 96
 Phe Met Asp Leu Ala Thr Ala Ile Gly Val Arg Ala Ala Pro Tyr Val
 20 25 30

gat cct ctc gag gcc gcg ctg gtg gcc cag gcc gag aag tac atc ccc 144
 Asp Pro Leu Glu Ala Ala Leu Val Ala Gln Ala Glu Lys Tyr Ile Pro
 35 40 45

acg att gtc cat cac acg cgt ggg ttc ctg gtc gcg gtg gag tcg cct 192
 Thr Ile Val His His Thr Arg Gly Phe Leu Val Ala Val Glu Ser Pro
 50 55 60

ttg gcc cgt gag ctg ccg ttg atg aac ccg ttc cac gtg ctg ttg atc 240
 Leu Ala Arg Glu Leu Pro Leu Met Asn Pro Phe His Val Leu Leu Ile
 65 70 75 80

gtg ctc gct tat ttg gtc acg gtc ttt gtg ggc atg cag atc atg aag 288
 Val Leu Ala Tyr Leu Val Thr Val Phe Val Gly Met Gln Ile Met Lys
 85 90 95

aac ttt gag cgg ttc gag gtc aag acg ttt tcg ctc ctg cac aac ttt 336
 Asn Phe Glu Arg Phe Glu Val Lys Thr Phe Ser Leu Leu His Asn Phe
 100 105 110

tgt ctg gtc tcg atc agc gcc tac atg tgc ggt ggg atc ctg tac gag 384
 Cys Leu Val Ser Ile Ser Ala Tyr Met Cys Gly Gly Ile Leu Tyr Glu
 115 120 125

gct tat cag gcc aac tat gga ctg ttt gag aac gct gct gat cat acc 432
 Ala Tyr Gln Ala Asn Tyr Gly Leu Phe Glu Asn Ala Ala Asp His Thr
 130 135 140

ttc aag ggt ctt cct atg gcc aag atg atc tgg ctc ttc tac ttc tcc 480
 Phe Lys Gly Leu Pro Met Ala Lys Met Ile Trp Leu Phe Tyr Phe Ser
 145 150 155 160

aag atc atg gag ttt gtc gac acc atg atc atg gtc ctc aag aag aac 528
 Lys Ile Met Glu Phe Val Asp Thr Met Ile Met Val Leu Lys Lys Asn
 165 170 175

aac cgc cag atc tcc ttc ttg cac gtt tac cac cac agc tcc atc ttc 576
 Asn Arg Gln Ile Ser Phe Leu His Val Tyr His His Ser Ser Ile Phe
 180 185 190

acc atc tgg tgg ttg gtc acc ttt gtt gca ccc aac ggt gaa gcc tac 624
 Thr Ile Trp Trp Leu Val Thr Phe Val Ala Pro Asn Gly Glu Ala Tyr
 195 200 205

ttc tct gct gcg ttg aac tcg ttc atc cat gtg atc atg tac ggc tac 672
 Phe Ser Ala Ala Leu Asn Ser Phe Ile His Val Ile Met Tyr Gly Tyr
 210 215 220

tac ttc ttg tcg gcc ttg ggc ttc aag cag gtg tcg ttc atc aag ttc 720
 Tyr Phe Leu Ser Ala Leu Gly Phe Lys Gln Val Ser Phe Ile Lys Phe
 225 230 235 240

tac atc acg cgc tcg cag atg aca cag ttc tgc atg atg tcg gtc cag 768
 Tyr Ile Thr Arg Ser Gln Met Thr Gln Phe Cys Met Met Ser Val Gln
 245 250 255

tct tcc tgg gac atg tac gcc atg aag gtc ctt ggc cgc ccc gga tac 816
 Ser Ser Trp Asp Met Tyr Ala Met Lys Val Leu Gly Arg Pro Gly Tyr
 260 265 270

ccc ttc ttc atc acg gct ctg ctt tgg ttc tac atg tgg acc atg ctc 864
 Pro Phe Phe Ile Thr Ala Leu Leu Trp Phe Tyr Met Trp Thr Met Leu
 275 280 285

ggt ctc ttc tac aac ttt tac aga aag aac gcc aag ttg gcc aag cag 912
 Gly Leu Phe Tyr Asn Phe Tyr Arg Lys Asn Ala Lys Leu Ala Lys Gln
 290 295 300

gcc aag gcc gac gct gcc aag gag aag gca agg aag ttg cag taa 957
 Ala Lys Ala Asp Ala Ala Lys Glu Lys Ala Arg Lys Leu Gln
 305 310 315

<210> 36

<211> 318

<212> PRT

<213> Mortierella alpina

<400> 36

Met Glu Ser Ile Ala Pro Phe Leu Pro Ser Lys Met Pro Gln Asp Leu
 1 5 10 15

Phe Met Asp Leu Ala Thr Ala Ile Gly Val Arg Ala Ala Pro Tyr Val
 20 25 30

Asp Pro Leu Glu Ala Ala Leu Val Ala Gln Ala Glu Lys Tyr Ile Pro
 35 40 45

Thr Ile Val His His Thr Arg Gly Phe Leu Val Ala Val Glu Ser Pro
 50 55 60

Leu Ala Arg Glu Leu Pro Leu Met Asn Pro Phe His Val Leu Leu Ile
 65 70 75 80

Val Leu Ala Tyr Leu Val Thr Val Phe Val Gly Met Gln Ile Met Lys
 85 90 95

Asn Phe Glu Arg Phe Glu Val Lys Thr Phe Ser Leu Leu His Asn Phe
 100 105 110

Cys Leu Val Ser Ile Ser Ala Tyr Met Cys Gly Gly Ile Leu Tyr Glu
 115 120 125

Ala Tyr Gln Ala Asn Tyr Gly Leu Phe Glu Asn Ala Ala Asp His Thr
 130 135 140

Phe Lys Gly Leu Pro Met Ala Lys Met Ile Trp Leu Phe Tyr Phe Ser
 145 150 155 160

Lys Ile Met Glu Phe Val Asp Thr Met Ile Met Val Leu Lys Lys Asn
 165 170 175

Asn Arg Gln Ile Ser Phe Leu His Val Tyr His His Ser Ser Ile Phe
 180 185 190

Thr Ile Trp Trp Leu Val Thr Phe Val Ala Pro Asn Gly Glu Ala Tyr
 195 200 205

Phe Ser Ala Ala Leu Asn Ser Phe Ile His Val Ile Met Tyr Gly Tyr
 210 215 220

Tyr Phe Leu Ser Ala Leu Gly Phe Lys Gln Val Ser Phe Ile Lys Phe
 225 230 235 240

Tyr Ile Thr Arg Ser Gln Met Thr Gln Phe Cys Met Met Ser Val Gln
 245 250 255

Ser Ser Trp Asp Met Tyr Ala Met Lys Val Leu Gly Arg Pro Gly Tyr
 260 265 270

Pro Phe Phe Ile Thr Ala Leu Leu Trp Phe Tyr Met Trp Thr Met Leu
 275 280 285

Gly Leu Phe Tyr Asn Phe Tyr Arg Lys Asn Ala Lys Leu Ala Lys Gln
 290 295 300

Ala Lys Ala Asp Ala Ala Lys Glu Lys Ala Arg Lys Leu Gln
 305 310 315

<210> 37

<211> 867

<212> DNA

<213> *Caenorhabditis elegans*

<220>

<221> CDS

<222> (1)..(867)

<223> Delta-6 elongase

<400> 37

atg gct cag cat ccg ctc gtt caa cgg ctt ctc gat gtc aaa ttc gac 48
 Met Ala Gln His Pro Leu Val Gln Arg Leu Leu Asp Val Lys Phe Asp
 1 5 10 15

acg aaa cga ttt gtg gct att gct act cat ggg cca aag aat ttc cct 96
 Thr Lys Arg Phe Val Ala Ile Ala Thr His Gly Pro Lys Asn Phe Pro
 20 25 30

gac gca gaa ggt cgc aag ttc ttt gct gat cac ttt gat gtt act att 144
 Asp Ala Glu Gly Arg Lys Phe Phe Ala Asp His Phe Asp Val Thr Ile
 35 40 45

cag gct tca atc ctg tac atg gtc gtt gtg ttc gga aca aaa tgg ttc 192
 Gln Ala Ser Ile Leu Tyr Met Val Val Val Phe Gly Thr Lys Trp Phe
 50 55 60

atg cgt aat cgt caa cca ttc caa ttg act att cca ctc aac atc tgg 240
 Met Arg Asn Arg Gln Pro Phe Gln Leu Thr Ile Pro Leu Asn Ile Trp
 65 70 75 80

aat ttc atc ctc gcc gca ttt tcc atc gca gga gct gtc aaa atg acc 288
 Asn Phe Ile Leu Ala Ala Phe Ser Ile Ala Gly Ala Val Lys Met Thr
 85 90 95

cca gag ttc ttt gga acc att gcc aac aaa gga att gtc gca tcc tac 336
 Pro Glu Phe Phe Gly Thr Ile Ala Asn Lys Gly Ile Val Ala Ser Tyr
 100 105 110

tgc aaa gtg ttt gat ttc acg aaa gga gag aat gga tac tgg gtg tgg 384

70

Cys Lys Val Phe Asp Phe Thr Lys Gly Glu Asn Gly Tyr Trp Val Trp
 115 120 125
 ctc ttc atg gct tcc aaa ctt ttc gaa ctt gtt gac acc atc ttc ttg 432
 Leu Phe Met Ala Ser Lys Leu Phe Glu Leu Val Asp Thr Ile Phe Leu
 130 135 140
 gtt ctc cgt aaa cgt cca ctc atg ttc ctt cac tgg tat cac cat att 480
 Val Leu Arg Lys Arg Pro Leu Met Phe Leu His Trp Tyr His His Ile
 145 150 155 160
 ctc acc atg atc tac gcc tgg tac tct cat cca ttg acc cca gga ttc 528
 Leu Thr Met Ile Tyr Ala Trp Tyr Ser His Pro Leu Thr Pro Gly Phe
 165 170 175
 aac aga tac gga att tat ctt aac ttt gtc gtc cac gcc ttc atg tac 576
 Asn Arg Tyr Gly Ile Tyr Leu Asn Phe Val Val His Ala Phe Met Tyr
 180 185 190
 tct tac tac ttc ctt cgc tcg atg aag att cgc gtg cca gga ttc atc 624
 Ser Tyr Tyr Phe Leu Arg Ser Met Lys Ile Arg Val Pro Gly Phe Ile
 195 200 205
 gcc caa gct atc aca tct ctt caa atc gtt caa ttc atc atc tct tgc 672
 Ala Gln Ala Ile Thr Ser Leu Gln Ile Val Gln Phe Ile Ile Ser Cys
 210 215 220
 gcc gtt ctt gct cat ctt ggt tat ctc atg cac ttc acc aat gcc aac 720
 Ala Val Leu Ala His Leu Gly Tyr Leu Met His Phe Thr Asn Ala Asn
 225 230 235 240
 tgt gat ttc gag cca tca gta ttc aag ctc gca gtt ttc atg gac aca 768
 Cys Asp Phe Glu Pro Ser Val Phe Lys Leu Ala Val Phe Met Asp Thr
 245 250 255
 aca tac ttg gct ctt ttc gtc aac ttc ttc ctc caa tca tat gtt ctc 816
 Thr Tyr Leu Ala Leu Phe Val Asn Phe Phe Leu Gln Ser Tyr Val Leu
 260 265 270
 cgc gga gga aaa gac aag tac aag gca gtg cca aag aag aag aac aac 864
 Arg Gly Gly Lys Asp Lys Tyr Lys Ala Val Pro Lys Lys Lys Asn Asn
 275 280 285
 taa 867

<210> 38

<211> 288

<212> PRT

<213> Caenorhabditis elegans

<400> 38

Met Ala Gln His Pro Leu Val Gln Arg Leu Leu Asp Val Lys Phe Asp
 1 5 10 15

Thr Lys Arg Phe Val Ala Ile Ala Thr His Gly Pro Lys Asn Phe Pro
 20 25 30

Asp Ala Glu Gly Arg Lys Phe Phe Ala Asp His Phe Asp Val Thr Ile
 35 40 45

Gln Ala Ser Ile Leu Tyr Met Val Val Val Phe Gly Thr Lys Trp Phe
 50 55 60

Met Arg Asn Arg Gln Pro Phe Gln Leu Thr Ile Pro Leu Asn Ile Trp
 65 70 75 80

Asn Phe Ile Leu Ala Ala Phe Ser Ile Ala Gly Ala Val Lys Met Thr
 85 90 95

Pro Glu Phe Phe Gly Thr Ile Ala Asn Lys Gly Ile Val Ala Ser Tyr
 100 105 110

Cys Lys Val Phe Asp Phe Thr Lys Gly Glu Asn Gly Tyr Trp Val Trp
 115 120 125

Leu Phe Met Ala Ser Lys Leu Phe Glu Leu Val Asp Thr Ile Phe Leu
 130 135 140

Val Leu Arg Lys Arg Pro Leu Met Phe Leu His Trp Tyr His His Ile
 145 150 155 160

Leu Thr Met Ile Tyr Ala Trp Tyr Ser His Pro Leu Thr Pro Gly Phe
 165 170 175

Asn Arg Tyr Gly Ile Tyr Leu Asn Phe Val Val His Ala Phe Met Tyr
 180 185 190

Ser Tyr Tyr Phe Leu Arg Ser Met Lys Ile Arg Val Pro Gly Phe Ile
 195 200 205

Ala Gln Ala Ile Thr Ser Leu Gln Ile Val Gln Phe Ile Ile Ser Cys
 210 215 220

Ala Val Leu Ala His Leu Gly Tyr Leu Met His Phe Thr Asn Ala Asn
 225 230 235 240

Cys Asp Phe Glu Pro Ser Val Phe Lys Leu Ala Val Phe Met Asp Thr
 245 250 255

Thr Tyr Leu Ala Leu Phe Val Asn Phe Phe Leu Gln Ser Tyr Val Leu
 260 265 270

Arg Gly Gly Lys Asp Lys Tyr Lys Ala Val Pro Lys Lys Lys Asn Asn
 275 280 285

<210> 39

<211> 1626

<212> DNA

<213> *Euglena gracilis*

<220>

<221> CDS

<222> (1)..(1626)

<223> Delta-4 desaturase

<400> 39

atg ttg gtg ctg ttt ggc aat ttc tat gtc aag caa tac tcc caa aag	48
Met Leu Val Leu Phe Gly Asn Phe Tyr Val Lys Gln Tyr Ser Gln Lys	
1 5 10 15	

aac ggc aag ccg gag aac gga gcc acc cct gag aac gga gcg aag ccg	96
Asn Gly Lys Pro Glu Asn Gly Ala Thr Pro Glu Asn Gly Ala Lys Pro	
20 25 30	

caa cct tgc gag aac ggc acg gtg gaa aag cga gag aat gac acc gcc	144
Gln Pro Cys Glu Asn Gly Thr Val Glu Lys Arg Glu Asn Asp Thr Ala	
35 40 45	

aac gtt cgg ccc acc cgt cca gct gga ccc ccg ccg gcc acg tac tac	192
Asn Val Arg Pro Thr Arg Pro Ala Gly Pro Pro Pro Ala Thr Tyr Tyr	
50 55 60	

gac tcc ctg gca gtg tgc ggg cag ggc aag gag cgg ctg ttc acc acc	240
Asp Ser Leu Ala Val Ser Gly Gln Gly Lys Glu Arg Leu Phe Thr Thr	
65 70 75 80	

gat gag gtg agg cgg cac atc ctc ccc acc gat ggc tgg ctg acg tgc	288
Asp Glu Val Arg Arg His Ile Leu Pro Thr Asp Gly Trp Leu Thr Cys	
85 90 95	

cac gaa gga gtc tac gat gtc act gat ttc ctt gcc aag cac cct ggt	336
His Glu Gly Val Tyr Asp Val Thr Asp Phe Leu Ala Lys His Pro Gly	
100 105 110	

ggc ggt gtc atc acg ctg ggc ctt gga agg gac tgc aca atc ctc atc	384
Gly Gly Val Ile Thr Leu Gly Leu Gly Arg Asp Cys Thr Ile Leu Ile	
115 120 125	

gag tca tac cac cct gct ggg cgc ccg gac aag gtg atg gag aag tac	432
Glu Ser Tyr His Pro Ala Gly Arg Pro Asp Lys Val Met Glu Lys Tyr	
130 135 140	

cgc att ggt acg ctg cag gac ccc aag acg ttc tat gct tgg gga gag	480
Arg Ile Gly Thr Leu Gln Asp Pro Lys Thr Phe Tyr Ala Trp Gly Glu	
145 150 155 160	

tcc gat ttc tac cct gag ttg aag cgc cgg gcc ctt gca agg ctg aag	528
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73

Ser	Asp	Phe	Tyr	Pro	Glu	Leu	Lys	Arg	Arg	Ala	Leu	Ala	Arg	Leu	Lys	
				165					170						175	
gag	gct	ggt	cag	gcg	cgg	cgc	ggc	ggc	ctt	ggg	gtg	aag	gcc	ctc	ctg	576
Glu	Ala	Gly	Gln	Ala	Arg	Arg	Gly	Gly	Leu	Gly	Val	Lys	Ala	Leu	Leu	
			180					185					190			
gtg	ctc	acc	ctc	ttc	ttc	gtg	tcg	tgg	tac	atg	tgg	gtg	gcc	cac	aag	624
Val	Leu	Thr	Leu	Phe	Phe	Val	Ser	Trp	Tyr	Met	Trp	Val	Ala	His	Lys	
		195					200					205				
tcc	ttc	ctc	tgg	gcc	gcc	gtc	tgg	ggc	ttc	gcc	ggc	tcc	cac	gtc	ggg	672
Ser	Phe	Leu	Trp	Ala	Ala	Val	Trp	Gly	Phe	Ala	Gly	Ser	His	Val	Gly	
	210					215					220					
ctg	agc	atc	cag	cac	gat	ggc	aac	cac	ggc	gcg	ttc	agc	cgc	aac	aca	720
Leu	Ser	Ile	Gln	His	Asp	Gly	Asn	His	Gly	Ala	Phe	Ser	Arg	Asn	Thr	
225					230					235					240	
ctg	gtg	aac	cgc	ctg	gcg	ggg	tgg	ggc	atg	gac	ttg	atc	ggc	gcg	tcg	768
Leu	Val	Asn	Arg	Leu	Ala	Gly	Trp	Gly	Met	Asp	Leu	Ile	Gly	Ala	Ser	
				245					250					255		
tcc	acg	gtg	tgg	gag	tac	cag	cac	gtc	atc	ggc	cac	cac	cag	tac	acc	816
Ser	Thr	Val	Trp	Glu	Tyr	Gln	His	Val	Ile	Gly	His	His	Gln	Tyr	Thr	
			260					265					270			
aac	ctc	gtg	tcg	gac	acg	cta	ttc	agt	ctg	cct	gag	aac	gat	ccg	gac	864
Asn	Leu	Val	Ser	Asp	Thr	Leu	Phe	Ser	Leu	Pro	Glu	Asn	Asp	Pro	Asp	
		275					280					285				
gtc	ttc	tcc	agc	tac	ccg	ctg	atg	cgc	atg	cac	ccg	gat	acg	gcg	tgg	912
Val	Phe	Ser	Ser	Tyr	Pro	Leu	Met	Arg	Met	His	Pro	Asp	Thr	Ala	Trp	
	290					295					300					
cag	ccg	cac	cac	cgc	ttc	cag	cac	ctg	ttc	gcg	ttc	cca	ctg	ttc	gcc	960
Gln	Pro	His	His	Arg	Phe	Gln	His	Leu	Phe	Ala	Phe	Pro	Leu	Phe	Ala	
305					310					315					320	
ctg	atg	aca	atc	agc	aag	gtg	ctg	acc	agc	gat	ttc	gct	gtc	tgc	ctc	1008
Leu	Met	Thr	Ile	Ser	Lys	Val	Leu	Thr	Ser	Asp	Phe	Ala	Val	Cys	Leu	
				325					330					335		
agc	atg	aag	aag	ggg	tcc	atc	gac	tgc	tcc	tcc	agg	ctc	gtc	cca	ctg	1056
Ser	Met	Lys	Lys	Gly	Ser	Ile	Asp	Cys	Ser	Ser	Arg	Leu	Val	Pro	Leu	
			340					345					350			
gag	ggg	cag	ctg	ctg	ttc	tgg	ggg	gcc	aag	ctg	gcg	aac	ttc	ctg	ttg	1104
Glu	Gly	Gln	Leu	Leu	Phe	Trp	Gly	Ala	Lys	Leu	Ala	Asn	Phe	Leu	Leu	
		355					360					365				
cag	att	gtg	ttg	cca	tgc	tac	ctc	cac	ggg	aca	gct	atg	ggc	ctg	gcc	1152
Gln	Ile	Val	Leu	Pro	Cys	Tyr	Leu	His	Gly	Thr	Ala	Met	Gly	Leu	Ala	
	370					375					380					
ctc	ttc	tct	gtt	gct	cac	ctt	gtg	tcg	ggg	gag	tac	ctc	gcg	atc	tgc	1200
Leu	Phe	Ser	Val	Ala	His	Leu	Val	Ser	Gly	Glu	Tyr	Leu	Ala	Ile	Cys	
	385				390					395					400	
ttc	atc	atc	aac	cac	atc	agc	gag	tct	tgt	gag	ttt	atg	aat	aca	agc	1248
Phe	Ile	Ile	Asn	His	Ile	Ser	Glu	Ser	Cys	Glu	Phe	Met	Asn	Thr	Ser	
			405						410					415		
ttt	caa	acc	gcc	gcc	cgg	agg	aca	gag	atg	ctt	cag	gca	gca	cat	cag	1296

74

Phe Gln Thr Ala Ala Arg Arg Thr Glu Met Leu Gln Ala Ala His Gln
 420 425 430
 gca gcg gag gcc aag aag gtg aag ccc acc cct cca ccg aac gat tgg 1344
 Ala Ala Glu Ala Lys Lys Val Lys Pro Thr Pro Pro Asn Asp Trp
 435 440 445
 gct gtg aca cag gtc caa tgc tgc gtg aat tgg aga tca ggt ggc gtg 1392
 Ala Val Thr Gln Val Gln Cys Val Asn Trp Arg Ser Gly Gly Val
 450 455 460
 ttg gcc aat cac ctc tct gga ggc ttg aac cac cag atc gag cat cat 1440
 Leu Ala Asn His Leu Ser Gly Gly Leu Asn His Gln Ile Glu His His
 465 470 475 480
 ctg ttc ccc agc atc tgc cat gcc aac tac ccc acc atc gcc cct gtt 1488
 Leu Phe Pro Ser Ile Ser His Ala Asn Tyr Pro Thr Ile Ala Pro Val
 485 490 495
 gtg aag gag gtg tgc gag gag tac ggg ttg ccg tac aag aat tac gtc 1536
 Val Lys Glu Val Cys Glu Glu Tyr Gly Leu Pro Tyr Lys Asn Tyr Val
 500 505 510
 acg ttc tgg gat gca gtc tgt ggc atg gtt cag cac ctc cgg ttg atg 1584
 Thr Phe Trp Asp Ala Val Cys Gly Met Val Gln His Leu Arg Leu Met
 515 520 525
 ggt gct cca ccg gtg cca acg aac ggg gac aaa aag tca taa 1626
 Gly Ala Pro Pro Val Pro Thr Asn Gly Asp Lys Lys Ser
 530 535 540

<210> 40

<211> 541

<212> PRT

<213> *Euglena gracilis*

<400> 40

Met Leu Val Leu Phe Gly Asn Phe Tyr Val Lys Gln Tyr Ser Gln Lys
 1 5 10 15
 Asn Gly Lys Pro Glu Asn Gly Ala Thr Pro Glu Asn Gly Ala Lys Pro
 20 25 30
 Gln Pro Cys Glu Asn Gly Thr Val Glu Lys Arg Glu Asn Asp Thr Ala
 35 40 45
 Asn Val Arg Pro Thr Arg Pro Ala Gly Pro Pro Pro Ala Thr Tyr Tyr
 50 55 60
 Asp Ser Leu Ala Val Ser Gly Gln Gly Lys Glu Arg Leu Phe Thr Thr
 65 70 75 80

75

Asp Glu Val Arg Arg His Ile Leu Pro Thr Asp Gly Trp Leu Thr Cys
 85 90 95

His Glu Gly Val Tyr Asp Val Thr Asp Phe Leu Ala Lys His Pro Gly
 100 105 110

Gly Gly Val Ile Thr Leu Gly Leu Gly Arg Asp Cys Thr Ile Leu Ile
 115 120 125

Glu Ser Tyr His Pro Ala Gly Arg Pro Asp Lys Val Met Glu Lys Tyr
 130 135 140

Arg Ile Gly Thr Leu Gln Asp Pro Lys Thr Phe Tyr Ala Trp Gly Glu
 145 150 155 160

Ser Asp Phe Tyr Pro Glu Leu Lys Arg Arg Ala Leu Ala Arg Leu Lys
 165 170 175

Glu Ala Gly Gln Ala Arg Arg Gly Gly Leu Gly Val Lys Ala Leu Leu
 180 185 190

Val Leu Thr Leu Phe Phe Val Ser Trp Tyr Met Trp Val Ala His Lys
 195 200 205

Ser Phe Leu Trp Ala Ala Val Trp Gly Phe Ala Gly Ser His Val Gly
 210 215 220

Leu Ser Ile Gln His Asp Gly Asn His Gly Ala Phe Ser Arg Asn Thr
 225 230 235 240

Leu Val Asn Arg Leu Ala Gly Trp Gly Met Asp Leu Ile Gly Ala Ser
 245 250 255

Ser Thr Val Trp Glu Tyr Gln His Val Ile Gly His His Gln Tyr Thr
 260 265 270

Asn Leu Val Ser Asp Thr Leu Phe Ser Leu Pro Glu Asn Asp Pro Asp
 275 280 285

Val Phe Ser Ser Tyr Pro Leu Met Arg Met His Pro Asp Thr Ala Trp
 290 295 300

Gln Pro His His Arg Phe Gln His Leu Phe Ala Phe Pro Leu Phe Ala
 305 310 315 320

Leu Met Thr Ile Ser Lys Val Leu Thr Ser Asp Phe Ala Val Cys Leu
 325 330 335

76

Ser Met Lys Lys Gly Ser Ile Asp Cys Ser Ser Arg Leu Val Pro Leu
 340 345 350

Glu Gly Gln Leu Leu Phe Trp Gly Ala Lys Leu Ala Asn Phe Leu Leu
 355 360 365

Gln Ile Val Leu Pro Cys Tyr Leu His Gly Thr Ala Met Gly Leu Ala
 370 375 380

Leu Phe Ser Val Ala His Leu Val Ser Gly Glu Tyr Leu Ala Ile Cys
 385 390 395 400

Phe Ile Ile Asn His Ile Ser Glu Ser Cys Glu Phe Met Asn Thr Ser
 405 410 415

Phe Gln Thr Ala Ala Arg Arg Thr Glu Met Leu Gln Ala Ala His Gln
 420 425 430

Ala Ala Glu Ala Lys Lys Val Lys Pro Thr Pro Pro Pro Asn Asp Trp
 435 440 445

Ala Val Thr Gln Val Gln Cys Cys Val Asn Trp Arg Ser Gly Gly Val
 450 455 460

Leu Ala Asn His Leu Ser Gly Gly Leu Asn His Gln Ile Glu His His
 465 470 475 480

Leu Phe Pro Ser Ile Ser His Ala Asn Tyr Pro Thr Ile Ala Pro Val
 485 490 495

Val Lys Glu Val Cys Glu Glu Tyr Gly Leu Pro Tyr Lys Asn Tyr Val
 500 505 510

Thr Phe Trp Asp Ala Val Cys Gly Met Val Gln His Leu Arg Leu Met
 515 520 525

Gly Ala Pro Pro Val Pro Thr Asn Gly Asp Lys Lys Ser
 530 535 540

<210> 41

<211> 1548

<212> DNA

<213> Thraustochytrium

<220>

<221> CDS

<222> (1)..(1548)

<223> Delta-4 desaturase

<400> 41

atg	acg	gtc	ggg	ttt	gac	gaa	acg	gtg	act	atg	gac	acg	gtc	cgc	aac	48
Met	Thr	Val	Gly	Phe	Asp	Glu	Thr	Val	Thr	Met	Asp	Thr	Val	Arg	Asn	
1			5						10					15		

cac	aac	atg	ccg	gac	gac	gcc	tgg	tgc	gcg	atc	cac	ggc	acc	gtg	tac	96
His	Asn	Met	Pro	Asp	Asp	Ala	Trp	Cys	Ala	Ile	His	Gly	Thr	Val	Tyr	
			20					25					30			

gac	atc	acc	aag	ttc	agc	aag	gtg	cac	ccc	ggc	ggg	gac	atc	atc	atg	144
Asp	Ile	Thr	Lys	Phe	Ser	Lys	Val	His	Pro	Gly	Gly	Asp	Ile	Ile	Met	
		35					40					45				

ctg	gcc	gct	ggc	aag	gag	gcc	acc	atc	ctg	ttc	gag	acc	tac	cac	atc	192
Leu	Ala	Ala	Gly	Lys	Glu	Ala	Thr	Ile	Leu	Phe	Glu	Thr	Tyr	His	Ile	
	50					55					60					

aag	ggc	gtc	ccg	gac	gcg	gtg	ctg	cgc	aag	tac	aag	gtc	ggc	aag	ctc	240
Lys	Gly	Val	Pro	Asp	Ala	Val	Leu	Arg	Lys	Tyr	Lys	Val	Gly	Lys	Leu	
65					70					75					80	

ccc	cag	ggc	aag	aag	ggc	gaa	acg	agc	cac	atg	ccc	acc	ggg	ctc	gac	288
Pro	Gln	Gly	Lys	Lys	Gly	Glu	Thr	Ser	His	Met	Pro	Thr	Gly	Leu	Asp	
			85						90					95		

tcg	gcc	tcc	tac	tac	tcg	tgg	gac	agc	gag	ttt	tac	agg	gtg	ctc	cgc	336
Ser	Ala	Ser	Tyr	Tyr	Ser	Trp	Asp	Ser	Glu	Phe	Tyr	Arg	Val	Leu	Arg	
			100					105					110			

gag	cgc	gtc	gcc	aag	aag	ctg	gcc	gag	ccc	ggc	ctc	atg	cag	cgc	gcg	384
Glu	Arg	Val	Ala	Lys	Lys	Leu	Ala	Glu	Pro	Gly	Leu	Met	Gln	Arg	Ala	
		115				120					125					

cgc	atg	gag	ctc	tgg	gcc	aag	gcg	atc	ttc	ctc	ctg	gca	ggt	ttc	tgg	432
Arg	Met	Glu	Leu	Trp	Ala	Lys	Ala	Ile	Phe	Leu	Leu	Ala	Gly	Phe	Trp	
	130					135					140					

ggc	tcc	ctt	tac	gcc	atg	tgc	gtg	cta	gac	ccg	cac	ggc	ggt	gcc	atg	480
Gly	Ser	Leu	Tyr	Ala	Met	Cys	Val	Leu	Asp	Pro	His	Gly	Gly	Ala	Met	
145					150					155					160	

gta	gcc	gcc	gtt	acg	ctc	ggc	gtg	ttc	gct	gcc	ttt	gtc	gga	act	tgc	528
Val	Ala	Ala	Val	Thr	Leu	Gly	Val	Phe	Ala	Ala	Phe	Val	Gly	Thr	Cys	
			165						170					175		

atc	cag	cac	gac	ggc	agc	cac	ggc	gcc	ttc	tcc	aag	tcg	cga	ttc	atg	576
Ile	Gln	His	Asp	Gly	Ser	His	Gly	Ala	Phe	Ser	Lys	Ser	Arg	Phe	Met	
			180					185					190			

aac	aag	gcg	gcg	ggc	tgg	acc	ctc	gac	atg	atc	ggc	gcg	agt	gcg	atg	624
Asn	Lys	Ala	Ala	Gly	Trp	Thr	Leu	Asp	Met	Ile	Gly	Ala	Ser	Ala	Met	
		195				200					205					

acc	tgg	gag	atg	cag	cac	gtt	ctt	ggc	cac	cac	ccg	tac	acc	aac	ctc	672
Thr	Trp	Glu	Met	Gln	His	Val	Leu	Gly	His	His	Pro	Tyr	Thr	Asn	Leu	
	210					215					220					

atc gag atg gag aac ggt ttg gcc aag gtc aag ggc gcc gac gtc gac Ile Glu Met Glu Asn Gly Leu Ala Lys Val Lys Gly Ala Asp Val Asp 225 230 235 240	720
ccg aag aag gtc gac cag gag agc gac ccg gac gtc ttc agt acg tac Pro Lys Lys Val Asp Gln Glu Ser Asp Pro Asp Val Phe Ser Thr Tyr 245 250 255	768
ccg atg ctt cgc ctg cac ccg tgg cac cgc cag ccg ttt tac cac aag Pro Met Leu Arg Leu His Pro Trp His Arg Gln Arg Phe Tyr His Lys 260 265 270	816
ttc cag cac ctg tac gcc ccg ttt atc ttt ggg tct atg acg att aac Phe Gln His Leu Tyr Ala Pro Phe Ile Phe Gly Ser Met Thr Ile Asn 275 280 285	864
aag gtg att tcc cag gat gtc ggg gtt gtg ctg cgc aag cgc ctg ttc Lys Val Ile Ser Gln Asp Val Gly Val Val Leu Arg Lys Arg Leu Phe 290 295 300	912
cag atc gac gcc aac tgc ccg tat ggc agc ccc tgg tac gtg gcc cgc Gln Ile Asp Ala Asn Cys Arg Tyr Gly Ser Pro Trp Tyr Val Ala Arg 305 310 315 320	960
ttc tgg atc atg aag ctc ctc acc acg ctc tac atg gtg gcg ctt ccc Phe Trp Ile Met Lys Leu Leu Thr Thr Leu Tyr Met Val Ala Leu Pro 325 330 335	1008
atg tac atg cag ggg cct gct cag ggc ttg aag ctt ttc ttc atg gcc Met Tyr Met Gln Gly Pro Ala Gln Gly Leu Lys Leu Phe Phe Met Ala 340 345 350	1056
cac ttc acc tgc gga gag gtc ctc gcc acc atg ttt att gtc aac cac His Phe Thr Cys Gly Glu Val Leu Ala Thr Met Phe Ile Val Asn His 355 360 365	1104
atc atc gag ggc gtc agc tac gct tcc aag gac gcg gtc aag ggc gtc Ile Ile Glu Gly Val Ser Tyr Ala Ser Lys Asp Ala Val Lys Gly Val 370 375 380	1152
atg gct ccg ccg cgc act gtg cac ggt gtc acc ccg atg cag gtg acg Met Ala Pro Pro Arg Thr Val His Gly Val Thr Pro Met Gln Val Thr 385 390 395 400	1200
caa aag gcg ctc agt gcg gcc gag tcg gcc aag tcg gac gcc gac aag Gln Lys Ala Leu Ser Ala Ala Glu Ser Ala Lys Ser Asp Ala Asp Lys 405 410 415	1248
acg acc atg atc ccc ctc aac gac tgg gcc gct gtg cag tgc cag acc Thr Thr Met Ile Pro Leu Asn Asp Trp Ala Ala Val Gln Cys Gln Thr 420 425 430	1296
tct gtg aac tgg gct gtc ggg tcg tgg ttt tgg aac cac ttt tcg ggc Ser Val Asn Trp Ala Val Gly Ser Trp Phe Trp Asn His Phe Ser Gly 435 440 445	1344
ggc ctc aac cac cag att gag cac cac tgc ttc ccc caa aac ccc cac Gly Leu Asn His Gln Ile Glu His His Cys Phe Pro Gln Asn Pro His 450 455 460	1392
acg gtc aac gtc tac atc tcg ggc atc gtc aag gag acc tgc gaa gaa Thr Val Asn Val Tyr Ile Ser Gly Ile Val Lys Glu Thr Cys Glu Glu 465 470 475 480	1440

tac ggc gtg ccg tac cag gct gag atc agc ctc ttc tct gcc tat ttc 1488
 Tyr Gly Val Pro Tyr Gln Ala Glu Ile Ser Leu Phe Ser Ala Tyr Phe
 485 490 495

aag atg ctg tcg cac ctc cgc acg ctc ggc aac gag gac ctc acg gcc 1536
 Lys Met Leu Ser His Leu Arg Thr Leu Gly Asn Glu Asp Leu Thr Ala
 500 505 510

tgg tcc acg tga 1548
 Trp Ser Thr
 515

<210> 42

<211> 515

<212> PRT

<213> Thraustochytrium

<400> 42

Met Thr Val Gly Phe Asp Glu Thr Val Thr Met Asp Thr Val Arg Asn
 1 5 10 15

His Asn Met Pro Asp Asp Ala Trp Cys Ala Ile His Gly Thr Val Tyr
 20 25 30

Asp Ile Thr Lys Phe Ser Lys Val His Pro Gly Gly Asp Ile Ile Met
 35 40 45

Leu Ala Ala Gly Lys Glu Ala Thr Ile Leu Phe Glu Thr Tyr His Ile
 50 55 60

Lys Gly Val Pro Asp Ala Val Leu Arg Lys Tyr Lys Val Gly Lys Leu
 65 70 75 80

Pro Gln Gly Lys Lys Gly Glu Thr Ser His Met Pro Thr Gly Leu Asp
 85 90 95

Ser Ala Ser Tyr Tyr Ser Trp Asp Ser Glu Phe Tyr Arg Val Leu Arg
 100 105 110

Glu Arg Val Ala Lys Lys Leu Ala Glu Pro Gly Leu Met Gln Arg Ala
 115 120 125

Arg Met Glu Leu Trp Ala Lys Ala Ile Phe Leu Leu Ala Gly Phe Trp
 130 135 140

Gly Ser Leu Tyr Ala Met Cys Val Leu Asp Pro His Gly Gly Ala Met
 145 150 155 160

Val Ala Ala Val Thr Leu Gly Val Phe Ala Ala Phe Val Gly Thr Cys
 165 170 175

Ile Gln His Asp Gly Ser His Gly Ala Phe Ser Lys Ser Arg Phe Met
 180 185 190

Asn Lys Ala Ala Gly Trp Thr Leu Asp Met Ile Gly Ala Ser Ala Met
 195 200 205

Thr Trp Glu Met Gln His Val Leu Gly His His Pro Tyr Thr Asn Leu
 210 215 220

Ile Glu Met Glu Asn Gly Leu Ala Lys Val Lys Gly Ala Asp Val Asp
 225 230 235 240

Pro Lys Lys Val Asp Gln Glu Ser Asp Pro Asp Val Phe Ser Thr Tyr
 245 250 255

Pro Met Leu Arg Leu His Pro Trp His Arg Gln Arg Phe Tyr His Lys
 260 265 270

Phe Gln His Leu Tyr Ala Pro Phe Ile Phe Gly Ser Met Thr Ile Asn
 275 280 285

Lys Val Ile Ser Gln Asp Val Gly Val Val Leu Arg Lys Arg Leu Phe
 290 295 300

Gln Ile Asp Ala Asn Cys Arg Tyr Gly Ser Pro Trp Tyr Val Ala Arg
 305 310 315 320

Phe Trp Ile Met Lys Leu Leu Thr Thr Leu Tyr Met Val Ala Leu Pro
 325 330 335

Met Tyr Met Gln Gly Pro Ala Gln Gly Leu Lys Leu Phe Phe Met Ala
 340 345 350

His Phe Thr Cys Gly Glu Val Leu Ala Thr Met Phe Ile Val Asn His
 355 360 365

Ile Ile Glu Gly Val Ser Tyr Ala Ser Lys Asp Ala Val Lys Gly Val
 370 375 380

Met Ala Pro Pro Arg Thr Val His Gly Val Thr Pro Met Gln Val Thr
 385 390 395 400

Gln Lys Ala Leu Ser Ala Ala Glu Ser Ala Lys Ser Asp Ala Asp Lys
 405 410 415

Thr Thr Met Ile Pro Leu Asn Asp Trp Ala Ala Val Gln Cys Gln Thr
 420 425 430

Ser Val Asn Trp Ala Val Gly Ser Trp Phe Trp Asn His Phe Ser Gly
 435 440 445

Gly Leu Asn His Gln Ile Glu His His Cys Phe Pro Gln Asn Pro His
 450 455 460

Thr Val Asn Val Tyr Ile Ser Gly Ile Val Lys Glu Thr Cys Glu Glu
 465 470 475 480

Tyr Gly Val Pro Tyr Gln Ala Glu Ile Ser Leu Phe Ser Ala Tyr Phe
 485 490 495

Lys Met Leu Ser His Leu Arg Thr Leu Gly Asn Glu Asp Leu Thr Ala
 500 505 510

Trp Ser Thr
 515

<210> 43

<211> 960

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(960)

<223> Delta-5 elongase

<400> 43

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 Met Val Leu Tyr Asn Val Ala Gln Val Leu Leu Asn Gly Trp Thr Val
 1 5 10 15

tat gcg att gtg gat gcg gtg atg aat aga gac cat ccg ttt att gga 96
 Tyr Ala Ile Val Asp Ala Val Met Asn Arg Asp His Pro Phe Ile Gly
 20 25 30

agt aga agt ttg gtt ggg gcg gcg ttg cat agt ggg agc tcg tat gcg 144
 Ser Arg Ser Leu Val Gly Ala Ala Leu His Ser Gly Ser Ser Tyr Ala
 35 40 45

gtg tgg gtt cat tat tgt gat aag tat ttg gag ttc ttt gat acg tat 192

82

Val	Trp	Val	His	Tyr	Cys	Asp	Lys	Tyr	Leu	Glu	Phe	Phe	Asp	Thr	Tyr												
50																55	60										
ttt	atg	gtg	ttg	agg	ggg	aaa	atg	gac	cag	atg	gta	ctt	ggt	gaa	gtt	240											
Phe	Met	Val	Leu	Arg	Gly	Lys	Met	Asp	Gln	Met	Val	Leu	Gly	Glu	Val												
65																70	75										80
ggt	ggc	agt	gtg	tgg	tgt	ggc	gtt	gga	tat	atg	gat	atg	gag	aag	atg	288											
Gly	Gly	Ser	Val	Trp	Cys	Gly	Val	Gly	Tyr	Met	Asp	Met	Glu	Lys	Met												
85																90	95										
ata	cta	ctc	agc	ttt	gga	gtg	cat	cgg	tct	gct	cag	gga	acg	ggg	aag	336											
Ile	Leu	Leu	Ser	Phe	Gly	Val	His	Arg	Ser	Ala	Gln	Gly	Thr	Gly	Lys												
100																105	110										
gct	ttc	acc	aac	aac	gtt	acc	aat	cca	cat	ctc	acg	ctt	cca	cct	cat	384											
Ala	Phe	Thr	Asn	Asn	Val	Thr	Asn	Pro	His	Leu	Thr	Leu	Pro	Pro	His												
115																120	125										
tct	aca	aaa	aca	aaa	aaa	cag	gtc	tcc	ttc	ctc	cac	atc	tac	cac	cac	432											
Ser	Thr	Lys	Thr	Lys	Lys	Gln	Val	Ser	Phe	Leu	His	Ile	Tyr	His	His												
130																135	140										
acg	acc	ata	gcg	tgg	gca	tgg	tgg	atc	gcc	ctc	cgc	ttc	tcc	ccc	ggt	480											
Thr	Thr	Ile	Ala	Trp	Ala	Trp	Trp	Ile	Ala	Leu	Arg	Phe	Ser	Pro	Gly												
145																150	155										160
gga	gac	att	tac	ttc	ggg	gca	ctc	ctc	aac	tcc	atc	atc	cac	gtc	ctc	528											
Gly	Asp	Ile	Tyr	Phe	Gly	Ala	Leu	Leu	Asn	Ser	Ile	Ile	His	Val	Leu												
165																170	175										
atg	tat	tcc	tac	tac	gcc	ctt	gcc	cta	ctc	aag	gtc	agt	tgt	cca	tgg	576											
Met	Tyr	Ser	Tyr	Tyr	Ala	Leu	Ala	Leu	Leu	Lys	Val	Ser	Cys	Pro	Trp												
180																185	190										
aaa	cga	tac	ctg	act	caa	gct	caa	tta	ttg	caa	ttc	aca	agt	gtg	gtg	624											
Lys	Arg	Tyr	Leu	Thr	Gln	Ala	Gln	Leu	Leu	Gln	Phe	Thr	Ser	Val	Val												
195																200	205										
gtt	tat	acg	ggg	tgt	acg	ggt	tat	act	cat	tac	tat	cat	acg	aag	cat	672											
Val	Tyr	Thr	Gly	Cys	Thr	Gly	Tyr	Thr	His	Tyr	Tyr	His	Thr	Lys	His												
210																215	220										
gga	gcg	gat	gag	aca	cag	cct	agt	tta	gga	acg	tat	tat	ttc	tgt	tgt	720											
Gly	Ala	Asp	Glu	Thr	Gln	Pro	Ser	Leu	Gly	Thr	Tyr	Tyr	Phe	Cys	Cys												
225																230	235										240
gga	gtg	cag	gtg	ttt	gag	atg	gtt	agt	ttg	ttt	gta	ctc	ttt	tcc	atc	768											
Gly	Val	Gln	Val	Phe	Glu	Met	Val	Ser	Leu	Phe	Val	Leu	Phe	Ser	Ile												
245																250	255										
ttt	tat	aaa	cga	tcc	tat	tcg	aag	aag	aac	aag	tca	gga	gga	aag	gat	816											
Phe	Tyr	Lys	Arg	Ser	Tyr	Ser	Lys	Lys	Asn	Lys	Ser	Gly	Gly	Lys	Asp												
260																265	270										
agc	aag	aag	aat	gat	gat	ggg	aat	aat	gag	gat	caa	tgt	cac	aag	gct	864											
Ser	Lys	Lys	Asn	Asp	Asp	Gly	Asn	Asn	Glu	Asp	Gln	Cys	His	Lys	Ala												
275																280	285										
atg	aag	gat	ata	tcg	gag	ggt	gcg	aag	gag	ggt	gtg	ggg	cat	gca	gcg	912											
Met	Lys	Asp	Ile	Ser	Glu	Gly	Ala	Lys	Glu	Val	Val	Gly	His	Ala	Ala												
290																295	300										
aaq	gat	gct	gga	aaq	ttg	gtg	gct	acg	aga	gta	agg	tgt	aaq	gtg	taa	960											

83

Lys Asp Ala Gly Lys Leu Val Ala Thr Arg Val Arg Cys Lys Val
 305 310 315

<210> 44

<211> 319

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 44

Met Val Leu Tyr Asn Val Ala Gln Val Leu Leu Asn Gly Trp Thr Val
 1 5 10 15

Tyr Ala Ile Val Asp Ala Val Met Asn Arg Asp His Pro Phe Ile Gly
 20 25 30

Ser Arg Ser Leu Val Gly Ala Ala Leu His Ser Gly Ser Ser Tyr Ala
 35 40 45

Val Trp Val His Tyr Cys Asp Lys Tyr Leu Glu Phe Phe Asp Thr Tyr
 50 55 60

Phe Met Val Leu Arg Gly Lys Met Asp Gln Met Val Leu Gly Glu Val
 65 70 75 80

Gly Gly Ser Val Trp Cys Gly Val Gly Tyr Met Asp Met Glu Lys Met
 85 90 95

Ile Leu Leu Ser Phe Gly Val His Arg Ser Ala Gln Gly Thr Gly Lys
 100 105 110

Ala Phe Thr Asn Asn Val Thr Asn Pro His Leu Thr Leu Pro Pro His
 115 120 125

Ser Thr Lys Thr Lys Lys Gln Val Ser Phe Leu His Ile Tyr His His
 130 135 140

Thr Thr Ile Ala Trp Ala Trp Trp Ile Ala Leu Arg Phe Ser Pro Gly
 145 150 155 160

Gly Asp Ile Tyr Phe Gly Ala Leu Leu Asn Ser Ile Ile His Val Leu
 165 170 175

Met Tyr Ser Tyr Tyr Ala Leu Ala Leu Leu Lys Val Ser Cys Pro Trp
 180 185 190

Lys Arg Tyr Leu Thr Gln Ala Gln Leu Leu Gln Phe Thr Ser Val Val
 195 200 205

Val Tyr Thr Gly Cys Thr Gly Tyr Thr His Tyr Tyr His Thr Lys His
 210 215 220

Gly Ala Asp Glu Thr Gln Pro Ser Leu Gly Thr Tyr Tyr Phe Cys Cys
 225 230 235 240

Gly Val Gln Val Phe Glu Met Val Ser Leu Phe Val Leu Phe Ser Ile
 245 250 255

Phe Tyr Lys Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp
 260 265 270

Ser Lys Lys Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala
 275 280 285

Met Lys Asp Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala
 290 295 300

Lys Asp Ala Gly Lys Leu Val Ala Thr Arg Val Arg Cys Lys Val
 305 310 315

<210> 45

<211> 819

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(819)

<223> Delta-5 elongase

<400> 45

atg gac gcc tac aac gct gca atg gat aag atc ggt gcc gcc atc atc 48
 Met Asp Ala Tyr Asn Ala Ala Met Asp Lys Ile Gly Ala Ala Ile Ile
 1 5 10 15

gat tgg tct gat ccc gat gga aag ttc cgt gcc gat aga gag gac tgg 96
 Asp Trp Ser Asp Pro Asp Gly Lys Phe Arg Ala Asp Arg Glu Asp Trp
 20 25 30

tgg ctc tgc gac ttc cgt agc gcc atc acc atc gcc ctc atc tac atc 144
 Trp Leu Cys Asp Phe Arg Ser Ala Ile Thr Ile Ala Leu Ile Tyr Ile
 35 40 45

gcc ttc gtc atc ctc ggt tcc gcc gtc atg caa tcc ctc ccc gca atg 192
 Ala Phe Val Ile Leu Gly Ser Ala Val Met Gln Ser Leu Pro Ala Met
 50 55 60

gat ccc tac ccc atc aaa ttc ctc tac aac gtc tcc caa atc ttc ctt 240
 Asp Pro Tyr Pro Ile Lys Phe Leu Tyr Asn Val Ser Gln Ile Phe Leu
 65 70 75 80

tgt gcc tac atg act gtc gag gcg gga ttt ttg gcc tac cgc aat gga 288
 Cys Ala Tyr Met Thr Val Glu Ala Gly Phe Leu Ala Tyr Arg Asn Gly
 85 90 95

tat acc gtc atg cct tgc aat cat ttc aat gtg aat gat cct ccc gtg 336
 Tyr Thr Val Met Pro Cys Asn His Phe Asn Val Asn Asp Pro Pro Val
 100 105 110

gcg aat ctt ctt tgg ttg ttt tat att tcc aag gtg tgg gac ttt tgg 384
 Ala Asn Leu Leu Trp Leu Phe Tyr Ile Ser Lys Val Trp Asp Phe Trp
 115 120 125

gat acc att ttc att gtg ttg ggg aag aag tgg cgt caa tta tct ttc 432
 Asp Thr Ile Phe Ile Val Leu Gly Lys Lys Trp Arg Gln Leu Ser Phe
 130 135 140

ttg cat gta tac cat cac acc acc atc ttt cta ttc tat tgg ctg aat 480
 Leu His Val Tyr His His Thr Thr Ile Phe Leu Phe Tyr Trp Leu Asn
 145 150 155 160

gcc aat gtc ttg tac gat ggt gac atc ttc ctt acc atc ttg ctc aat 528
 Ala Asn Val Leu Tyr Asp Gly Asp Ile Phe Leu Thr Ile Leu Leu Asn
 165 170 175

gga ttc atc cac acg gtg atg tac acg tat tac ttc atc tgt atg cat 576
 Gly Phe Ile His Thr Val Met Tyr Thr Tyr Tyr Phe Ile Cys Met His
 180 185 190

acc aaa gat tcc aag acg ggc aag agt ctt cct ata tgg tgg aag tcg 624
 Thr Lys Asp Ser Lys Thr Gly Lys Ser Leu Pro Ile Trp Trp Lys Ser
 195 200 205

agt ttg acg gcg ttt cag ttg ttg caa ttc act atc atg atg agt cag 672
 Ser Leu Thr Ala Phe Gln Leu Leu Gln Phe Thr Ile Met Met Ser Gln
 210 215 220

gct acc tac ctt gtc ttc cac ggg tgt gat aag gtg tcg ctt cgt atc 720
 Ala Thr Tyr Leu Val Phe His Gly Cys Asp Lys Val Ser Leu Arg Ile
 225 230 235 240

acg att gtg tac ttt gtg tcc ctt ttg agt ttg ttc ttc ctt ttt gct 768
 Thr Ile Val Tyr Phe Val Ser Leu Leu Ser Leu Phe Phe Leu Phe Ala
 245 250 255

cag ttc ttt gtg caa tca tac atg gca ccc aaa aag aag aag agt gct 816
 Gln Phe Phe Val Gln Ser Tyr Met Ala Pro Lys Lys Lys Lys Ser Ala
 260 265 270

tag 819

<210> 46
 <211> 272

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 46

Met Asp Ala Tyr Asn Ala Ala Met Asp Lys Ile Gly Ala Ala Ile Ile
 1 5 10 15

Asp Trp Ser Asp Pro Asp Gly Lys Phe Arg Ala Asp Arg Glu Asp Trp
 20 25 30

Trp Leu Cys Asp Phe Arg Ser Ala Ile Thr Ile Ala Leu Ile Tyr Ile
 35 40 45

Ala Phe Val Ile Leu Gly Ser Ala Val Met Gln Ser Leu Pro Ala Met
 50 55 60

Asp Pro Tyr Pro Ile Lys Phe Leu Tyr Asn Val Ser Gln Ile Phe Leu
 65 70 75 80

Cys Ala Tyr Met Thr Val Glu Ala Gly Phe Leu Ala Tyr Arg Asn Gly
 85 90 95

Tyr Thr Val Met Pro Cys Asn His Phe Asn Val Asn Asp Pro Pro Val
 100 105 110

Ala Asn Leu Leu Trp Leu Phe Tyr Ile Ser Lys Val Trp Asp Phe Trp
 115 120 125

Asp Thr Ile Phe Ile Val Leu Gly Lys Lys Trp Arg Gln Leu Ser Phe
 130 135 140

Leu His Val Tyr His His Thr Thr Ile Phe Leu Phe Tyr Trp Leu Asn
 145 150 155 160

Ala Asn Val Leu Tyr Asp Gly Asp Ile Phe Leu Thr Ile Leu Leu Asn
 165 170 175

Gly Phe Ile His Thr Val Met Tyr Thr Tyr Tyr Phe Ile Cys Met His
 180 185 190

Thr Lys Asp Ser Lys Thr Gly Lys Ser Leu Pro Ile Trp Trp Lys Ser
 195 200 205

Ser Leu Thr Ala Phe Gln Leu Leu Gln Phe Thr Ile Met Met Ser Gln
 210 215 220

87

Ala Thr Tyr Leu Val Phe His Gly Cys Asp Lys Val Ser Leu Arg Ile
225 230 235 240

Thr Ile Val Tyr Phe Val Ser Leu Leu Ser Leu Phe Phe Leu Phe Ala
245 250 255

Gln Phe Phe Val Gln Ser Tyr Met Ala Pro Lys Lys Lys Lys Ser Ala
260 265 270

<210> 47

<211> 936

<212> DNA

<213> Crypthecodinium cohnii

<220>

<221> CDS

<222> (1)..(936)

<223> Delta-5 elongase

<400> 47

atg tct gcc ttc atg act ctc cca cag gct ctc tcc gat gtg acc tcg 48
Met Ser Ala Phe Met Thr Leu Pro Gln Ala Leu Ser Asp Val Thr Ser
1 5 10 15

gcc ttg gtc acg ctg gga aag gat gtc tcc agc cct tca gct ttt caa 96
Ala Leu Val Thr Leu Gly Lys Asp Val Ser Ser Pro Ser Ala Phe Gln
20 25 30

gct gtc act ggc ttc tgc agg gag cag tgg ggg att ccg aca gta ttc 144
Ala Val Thr Gly Phe Cys Arg Glu Gln Trp Gly Ile Pro Thr Val Phe
35 40 45

tgc ctg ggc tac ttg gcc atg gtc tac gcg gcc aga aga ccc ctc ccg 192
Cys Leu Gly Tyr Leu Ala Met Val Tyr Ala Ala Arg Arg Pro Leu Pro
50 55 60

cag cac ggc tac atg gtt gcg gtg gac cgt tgc ttc gct gct tgg aac 240
Gln His Gly Tyr Met Val Ala Val Asp Arg Cys Phe Ala Ala Trp Asn
65 70 75 80

ttg gct ctc tct gtc ttc agc act tgg ggc ttc tac cac atg gct gtc 288
Leu Ala Leu Ser Val Phe Ser Thr Trp Gly Phe Tyr His Met Ala Val
85 90 95

ggg ctc tac aac atg aca gag acg agg ggc ttg caa ttc acc atc tgc 336
Gly Leu Tyr Asn Met Thr Glu Thr Arg Gly Leu Gln Phe Thr Ile Cys
100 105 110

ggc tgc act ggg gag ctc gtg cag aac ctt cag act ggc cca acc gct 384
Gly Ser Thr Gly Glu Leu Val Gln Asn Leu Gln Thr Gly Pro Thr Ala
115 120 125

ctg gcg ctc tgc ctc ttc tgc ttc agc aag atc ccc gag ttg atg gac 432
 Leu Ala Leu Cys Leu Phe Cys Phe Ser Lys Ile Pro Glu Leu Met Asp
 130 135 140

acg gtg ttt ctc atc ctg aag gcc aag aag gtc cgc ttc ttg cag tgg 480
 Thr Val Phe Leu Ile Leu Lys Ala Lys Lys Val Arg Phe Leu Gln Trp
 145 150 155 160

tac cac cat gcc aca gtc atg ctc ttc tgt tgg ctc gcc ctc gcg acg 528
 Tyr His His Ala Thr Val Met Leu Phe Cys Trp Leu Ala Leu Ala Thr
 165 170 175

gag tac act cct ggc ttg tgg ttt gcg gcg acg aac tac ttc gtg cac 576
 Glu Tyr Thr Pro Gly Leu Trp Phe Ala Ala Thr Asn Tyr Phe Val His
 180 185 190

tcc atc atg tac atg tac ttc ttc ctc atg acc ttc aag tcg gcc gcg 624
 Ser Ile Met Tyr Met Tyr Phe Phe Leu Met Thr Phe Lys Ser Ala Ala
 195 200 205

aag gtg gtg aag ccc atc gcc cct ctc atc aca gtt atc cag att gct 672
 Lys Val Val Lys Pro Ile Ala Pro Leu Ile Thr Val Ile Gln Ile Ala
 210 215 220

cag atg gtc tgg ggc ctc atc gtc aac ggc atc gcc atc acc acc ttc 720
 Gln Met Val Trp Gly Leu Ile Val Asn Gly Ile Ala Ile Thr Thr Phe
 225 230 235 240

ttc acg act ggt gcc tgc cag atc cag tct gtg act gtg tat tcg gcc 768
 Phe Thr Thr Gly Ala Cys Gln Ile Gln Ser Val Thr Val Tyr Ser Ala
 245 250 255

atc atc atg tac gct tcg tac ttc tac ctg ttc tcc cag ctc ttc ttc 816
 Ile Ile Met Tyr Ala Ser Tyr Phe Tyr Leu Phe Ser Gln Leu Phe Phe
 260 265 270

gag gcc cat ggt gcc gct ggc aag aac aag aag aag ttg acc cgc gag 864
 Glu Ala His Gly Ala Ala Gly Lys Asn Lys Lys Lys Leu Thr Arg Glu
 275 280 285

ctc tct cga aaa atc tcg gag gct ctc ctg aac acc ggt gac gag gtt 912
 Leu Ser Arg Lys Ile Ser Glu Ala Leu Leu Asn Thr Gly Asp Glu Val
 290 295 300

tcc aag cac ctg aag gtg aat tga 936
 Ser Lys His Leu Lys Val Asn
 305 310

<210> 48

<211> 311

<212> PRT

<213> *Crypthecodinium cohnii*

<400> 48

Met Ser Ala Phe Met Thr Leu Pro Gln Ala Leu Ser Asp Val Thr Ser
 1 5 10 15

Ala Leu Val Thr Leu Gly Lys Asp Val Ser Ser Pro Ser Ala Phe Gln
 20 25 30

Ala Val Thr Gly Phe Cys Arg Glu Gln Trp Gly Ile Pro Thr Val Phe
 35 40 45

Cys Leu Gly Tyr Leu Ala Met Val Tyr Ala Ala Arg Arg Pro Leu Pro
 50 55 60

Gln His Gly Tyr Met Val Ala Val Asp Arg Cys Phe Ala Ala Trp Asn
 65 70 75 80

Leu Ala Leu Ser Val Phe Ser Thr Trp Gly Phe Tyr His Met Ala Val
 85 90 95

Gly Leu Tyr Asn Met Thr Glu Thr Arg Gly Leu Gln Phe Thr Ile Cys
 100 105 110

Gly Ser Thr Gly Glu Leu Val Gln Asn Leu Gln Thr Gly Pro Thr Ala
 115 120 125

Leu Ala Leu Cys Leu Phe Cys Phe Ser Lys Ile Pro Glu Leu Met Asp
 130 135 140

Thr Val Phe Leu Ile Leu Lys Ala Lys Lys Val Arg Phe Leu Gln Trp
 145 150 155 160

Tyr His His Ala Thr Val Met Leu Phe Cys Trp Leu Ala Leu Ala Thr
 165 170 175

Glu Tyr Thr Pro Gly Leu Trp Phe Ala Ala Thr Asn Tyr Phe Val His
 180 185 190

Ser Ile Met Tyr Met Tyr Phe Phe Leu Met Thr Phe Lys Ser Ala Ala
 195 200 205

Lys Val Val Lys Pro Ile Ala Pro Leu Ile Thr Val Ile Gln Ile Ala
 210 215 220

Gln Met Val Trp Gly Leu Ile Val Asn Gly Ile Ala Ile Thr Thr Phe
 225 230 235 240

Phe Thr Thr Gly Ala Cys Gln Ile Gln Ser Val Thr Val Tyr Ser Ala
 245 250 255

Ile Ile Met Tyr Ala Ser Tyr Phe Tyr Leu Phe Ser Gln Leu Phe Phe
 260 265 270

Glu Ala His Gly Ala Ala Gly Lys Asn Lys Lys Lys Leu Thr Arg Glu
 275 280 285

Leu Ser Arg Lys Ile Ser Glu Ala Leu Leu Asn Thr Gly Asp Glu Val
 290 295 300

Ser Lys His Leu Lys Val Asn
 305 310

<210> 49

<211> 927

<212> DNA

<213> *Crypthecodinium cohnii*

<220>

<221> CDS

<222> (1)..(927)

<223> Delta-5 elongase

<400> 49

atg gct tcc tac caa caa gca ttc tcc gaa ttg gct aga gct ttg tcc 48
 Met Ala Ser Tyr Gln Gln Ala Phe Ser Glu Leu Ala Arg Ala Leu Ser
 1 5 10 15

act ttg aac cac gac ttc tcc agc gtc gag cca ttc aaa gtc gtg acg 96
 Thr Leu Asn His Asp Phe Ser Ser Val Glu Pro Phe Lys Val Val Thr
 20 25 30

cag ttc tgc agg gac cag tgg gcg atc ccg aca gtc ttt tgc atc ggt 144
 Gln Phe Cys Arg Asp Gln Trp Ala Ile Pro Thr Val Phe Cys Ile Gly
 35 40 45

tac ttg gca atg gtc tac gcc acg cga aga cct atc gcg aag cac ccc 192
 Tyr Leu Ala Met Val Tyr Ala Thr Arg Arg Pro Ile Ala Lys His Pro
 50 55 60

tac atg tct ctc gtg gat cgc tgc ttt gcg gcc tgg aac ttg ggc ctc 240
 Tyr Met Ser Leu Val Asp Arg Cys Phe Ala Ala Trp Asn Leu Gly Leu
 65 70 75 80

tcg ctc ttc agt tgc tgg ggc ttc tac cac atg gca gtg gga ctc tcc 288
 Ser Leu Phe Ser Cys Trp Gly Phe Tyr His Met Ala Val Gly Leu Ser
 85 90 95

cac acc act tgg aat ttc ggg ctc cag ttc acc atc tgc ggc agc acc 336
 His Thr Thr Trp Asn Phe Gly Leu Gln Phe Thr Ile Cys Gly Ser Thr
 100 105 110

acg gag ctt gtg aat ggc ttc cag aag ggc ccg gcg gcc ctc gcc ctc 384

91

Thr Glu Leu Val Asn Gly Phe Gln Lys Gly Pro Ala Ala Leu Ala Leu
 115 120 125
 atc ctg ttc tgc ttc tcc aag atc ccg gag ttg ggc gac acc gtc ttc 432
 Ile Leu Phe Cys Phe Ser Lys Ile Pro Glu Leu Gly Asp Thr Val Phe
 130 135 140
 ttg atc ttg aag gga aag aag gtc cgc ttc ttg cag tgg tac cac cac 480
 Leu Ile Leu Lys Gly Lys Lys Val Arg Phe Leu Gln Trp Tyr His His
 145 150 155 160
 acg acc gtg atg ctc ttc tgt tgg atg gcc ttg gcg act gag tac act 528
 Thr Thr Val Met Leu Phe Cys Trp Met Ala Leu Ala Thr Glu Tyr Thr
 165 170 175
 cct gga ttg tgg ttc gcg gcc acg aac tac ttc gtg cac tcc atc atg 576
 Pro Gly Leu Trp Phe Ala Ala Thr Asn Tyr Phe Val His Ser Ile Met
 180 185 190
 tac atg tac ttc ttc ctc atg acc ttc aag acg gcc gcc ggc atc atc 624
 Tyr Met Tyr Phe Phe Leu Met Thr Phe Lys Thr Ala Ala Gly Ile Ile
 195 200 205
 aag ccc atc gcg cct ctc atc acc atc atc cag atc tcc cag atg gtc 672
 Lys Pro Ile Ala Pro Leu Ile Thr Ile Ile Gln Ile Ser Gln Met Val
 210 215 220
 tgg ggc ttg gtc gtg aac gcc atc gcc gtc ggc acc ttc ttc acc aca 720
 Trp Gly Leu Val Val Asn Ala Ile Ala Val Gly Thr Phe Phe Thr Thr
 225 230 235 240
 ggc aac tgc cag atc cag gca gtg aca gtc tac tcc gcc atc gtg atg 768
 Gly Asn Cys Gln Ile Gln Ala Val Thr Val Tyr Ser Ala Ile Val Met
 245 250 255
 tac gcc tcc tac ttc tac ctc ttc ggc cag ctc ttc ttc gag gcc cag 816
 Tyr Ala Ser Tyr Phe Tyr Leu Phe Gly Gln Leu Phe Phe Glu Ala Gln
 260 265 270
 ggt tcg gct gga aag gac aag aag aag ttg gcc cga gag ctg agc cga 864
 Gly Ser Ala Gly Lys Asp Lys Lys Lys Leu Ala Arg Glu Leu Ser Arg
 275 280 285
 aag gtc tcg cgg gct ctc aca gca acg ggc gaa gag gtg tcg aag cac 912
 Lys Val Ser Arg Ala Leu Thr Ala Thr Gly Glu Glu Val Ser Lys His
 290 295 300
 atg aag gtg aat tga 927
 Met Lys Val Asn
 305

<210> 50

<211> 308

<212> PRT

<213> Crypthecodinium cohnii

<400> 50

92

Met Ala Ser Tyr Gln Gln Ala Phe Ser Glu Leu Ala Arg Ala Leu Ser
1 5 10 15

Thr Leu Asn His Asp Phe Ser Ser Val Glu Pro Phe Lys Val Val Thr
20 25 30

Gln Phe Cys Arg Asp Gln Trp Ala Ile Pro Thr Val Phe Cys Ile Gly
35 40 45

Tyr Leu Ala Met Val Tyr Ala Thr Arg Arg Pro Ile Ala Lys His Pro
50 55 60

Tyr Met Ser Leu Val Asp Arg Cys Phe Ala Ala Trp Asn Leu Gly Leu
65 70 75 80

Ser Leu Phe Ser Cys Trp Gly Phe Tyr His Met Ala Val Gly Leu Ser
85 90 95

His Thr Thr Trp Asn Phe Gly Leu Gln Phe Thr Ile Cys Gly Ser Thr
100 105 110

Thr Glu Leu Val Asn Gly Phe Gln Lys Gly Pro Ala Ala Leu Ala Leu
115 120 125

Ile Leu Phe Cys Phe Ser Lys Ile Pro Glu Leu Gly Asp Thr Val Phe
130 135 140

Leu Ile Leu Lys Gly Lys Lys Val Arg Phe Leu Gln Trp Tyr His His
145 150 155 160

Thr Thr Val Met Leu Phe Cys Trp Met Ala Leu Ala Thr Glu Tyr Thr
165 170 175

Pro Gly Leu Trp Phe Ala Ala Thr Asn Tyr Phe Val His Ser Ile Met
180 185 190

Tyr Met Tyr Phe Phe Leu Met Thr Phe Lys Thr Ala Ala Gly Ile Ile
195 200 205

Lys Pro Ile Ala Pro Leu Ile Thr Ile Ile Gln Ile Ser Gln Met Val
210 215 220

Trp Gly Leu Val Val Asn Ala Ile Ala Val Gly Thr Phe Phe Thr Thr
225 230 235 240

Gly Asn Cys Gln Ile Gln Ala Val Thr Val Tyr Ser Ala Ile Val Met
245 250 255

93

Tyr Ala Ser Tyr Phe Tyr Leu Phe Gly Gln Leu Phe Phe Glu Ala Gln
 260 265 270

Gly Ser Ala Gly Lys Asp Lys Lys Lys Leu Ala Arg Glu Leu Ser Arg
 275 280 285

Lys Val Ser Arg Ala Leu Thr Ala Thr Gly Glu Glu Val Ser Lys His
 290 295 300

Met Lys Val Asn
 305

<210> 51

<211> 795

<212> DNA

<213> Oncorhynchus mykiss

<220>

<221> CDS

<222> (1)..(795)

<223> Delta-5 elongase

<400> 51

atg gct tca aca tgg caa agc gtt cag tcc atg cgc cag tgg att tta 48
 Met Ala Ser Thr Trp Gln Ser Val Gln Ser Met Arg Gln Trp Ile Leu
 1 5 10 15

gag aat gga gat aaa agg aca gac cca tgg cta ctg gtc tac tcc cct 96
 Glu Asn Gly Asp Lys Arg Thr Asp Pro Trp Leu Leu Val Tyr Ser Pro
 20 25 30

atg cca gtg gcc att ata ttc ctc ctc tat ctt ggt gtg gtc tgg gct 144
 Met Pro Val Ala Ile Ile Phe Leu Leu Tyr Leu Gly Val Val Trp Ala
 35 40 45

ggg ccc aag ctg atg aaa cgc agg gaa cca gtt gat ctc aag gct gta 192
 Gly Pro Lys Leu Met Lys Arg Arg Glu Pro Val Asp Leu Lys Ala Val
 50 55 60

ctc att gtc tac aac ttc gcc atg gtc tgc ctg tct gtc tac atg ttc 240
 Leu Ile Val Tyr Asn Phe Ala Met Val Cys Leu Ser Val Tyr Met Phe
 65 70 75 80

cat gag ttc ttg gtc acg tcc ttg ctg tct aac tac agt tac ctg tgt 288
 His Glu Phe Leu Val Thr Ser Leu Leu Ser Asn Tyr Ser Tyr Leu Cys
 85 90 95

caa cct gtg gat tac agc act agt cca ctg gcg atg agg atg gcc aaa 336
 Gln Pro Val Asp Tyr Ser Thr Ser Pro Leu Ala Met Arg Met Ala Lys
 100 105 110

gta tgc tgg tgg ttt ttc ttc tcc aag gtc ata gaa ttg gct gac acg 384
 Val Cys Trp Trp Phe Phe Phe Ser Lys Val Ile Glu Leu Ala Asp Thr
 115 120 125

gtg ttc ttc atc ctg agg aag aag aac agt cag ctg act ttc ctg cat 432
 Val Phe Phe Ile Leu Arg Lys Lys Asn Ser Gln Leu Thr Phe Leu His
 130 135 140

gtc tat cac cat ggc acc atg atc ttc aac tgg tgg gca ggg gtc aag 480
 Val Tyr His His Gly Thr Met Ile Phe Asn Trp Trp Ala Gly Val Lys
 145 150 155 160

tat ctg gct gga ggc caa tgc ttc ttc atc ggc ctg ctc aat acc ttt 528
 Tyr Leu Ala Gly Gly Gln Ser Phe Phe Ile Gly Leu Leu Asn Thr Phe
 165 170 175

gtg cac atc gtg atg tac tct tac tac gga ctg gct gcc ctg ggg cct 576
 Val His Ile Val Met Tyr Ser Tyr Tyr Gly Leu Ala Ala Leu Gly Pro
 180 185 190

cac acg cag aag tac tta tgg tgg aag cgc tat ctg acc tca ctg cag 624
 His Thr Gln Lys Tyr Leu Trp Trp Lys Arg Tyr Leu Thr Ser Leu Gln
 195 200 205

ctg ctc cag ttt gtc ctg ttg acc act cac act ggc tac aac ctc ttc 672
 Leu Leu Gln Phe Val Leu Leu Thr Thr His Thr Gly Tyr Asn Leu Phe
 210 215 220

act gag tgt gac ttc ccg gac tcc atg aac gct gtg gtg ttt gcc tac 720
 Thr Glu Cys Asp Phe Pro Asp Ser Met Asn Ala Val Val Phe Ala Tyr
 225 230 235 240

tgt gtc agt ctc att gct ctc ttc agc aac ttc tac tat cag agc tac 768
 Cys Val Ser Leu Ile Ala Leu Phe Ser Asn Phe Tyr Tyr Gln Ser Tyr
 245 250 255

ctc aac agg aag agc aag aag aca taa 795
 Leu Asn Arg Lys Ser Lys Lys Thr
 260

<210> 52

<211> 264

<212> PRT

<213> Oncorhynchus mykiss

<400> 52

Met Ala Ser Thr Trp Gln Ser Val Gln Ser Met Arg Gln Trp Ile Leu
 1 5 10 15

Glu Asn Gly Asp Lys Arg Thr Asp Pro Trp Leu Leu Val Tyr Ser Pro
 20 25 30

Met Pro Val Ala Ile Ile Phe Leu Leu Tyr Leu Gly Val Val Trp Ala
 35 40 45

Gly Pro Lys Leu Met Lys Arg Arg Glu Pro Val Asp Leu Lys Ala Val
 50 55 60

Leu Ile Val Tyr Asn Phe Ala Met Val Cys Leu Ser Val Tyr Met Phe
 65 70 75 80

His Glu Phe Leu Val Thr Ser Leu Leu Ser Asn Tyr Ser Tyr Leu Cys
 85 90 95

Gln Pro Val Asp Tyr Ser Thr Ser Pro Leu Ala Met Arg Met Ala Lys
 100 105 110

Val Cys Trp Trp Phe Phe Phe Ser Lys Val Ile Glu Leu Ala Asp Thr
 115 120 125

Val Phe Phe Ile Leu Arg Lys Lys Asn Ser Gln Leu Thr Phe Leu His
 130 135 140

Val Tyr His His Gly Thr Met Ile Phe Asn Trp Trp Ala Gly Val Lys
 145 150 155 160

Tyr Leu Ala Gly Gly Gln Ser Phe Phe Ile Gly Leu Leu Asn Thr Phe
 165 170 175

Val His Ile Val Met Tyr Ser Tyr Tyr Gly Leu Ala Ala Leu Gly Pro
 180 185 190

His Thr Gln Lys Tyr Leu Trp Trp Lys Arg Tyr Leu Thr Ser Leu Gln
 195 200 205

Leu Leu Gln Phe Val Leu Leu Thr Thr His Thr Gly Tyr Asn Leu Phe
 210 215 220

Thr Glu Cys Asp Phe Pro Asp Ser Met Asn Ala Val Val Phe Ala Tyr
 225 230 235 240

Cys Val Ser Leu Ile Ala Leu Phe Ser Asn Phe Tyr Tyr Gln Ser Tyr
 245 250 255

Leu Asn Arg Lys Ser Lys Lys Thr
 260

<210> 53

<211> 885

<212> DNA

<213> Oncorhynchus mykiss

<220>

<221> CDS

<222> (1)..(885)

<223> Delta-5 elongase

<400> 53

atg	gag	act	ttt	aat	tat	aaa	cta	aac	atg	tac	ata	gac	tca	tgg	atg	48
Met	Glu	Thr	Phe	Asn	Tyr	Lys	Leu	Asn	Met	Tyr	Ile	Asp	Ser	Trp	Met	
1			5						10					15		

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Gly	Pro	Arg	Asp	Glu	Arg	Val	Gln	Gly	Trp	Leu	Leu	Leu	Asp	Asn	Tyr	
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Pro	Pro	Thr	Phe	Ala	Leu	Thr	Val	Met	Tyr	Leu	Leu	Ile	Val	Trp	Met	
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Gly	Pro	Lys	Tyr	Met	Arg	His	Arg	Gln	Pro	Val	Ser	Cys	Arg	Gly	Leu	
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Leu	Leu	Val	Tyr	Asn	Leu	Gly	Leu	Thr	Ile	Leu	Ser	Phe	Tyr	Met	Phe	
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Tyr	Glu	Met	Val	Ser	Ala	Val	Trp	His	Gly	Asp	Tyr	Asn	Phe	Phe	Cys	
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caa	gac	aca	cac	agt	gca	gga	gaa	acc	gat	acc	aag	atc	ata	aat	gtg	336
Gln	Asp	Thr	His	Ser	Ala	Gly	Glu	Thr	Asp	Thr	Lys	Ile	Ile	Asn	Val	
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ctg	tgg	tgg	tac	tac	ttc	tcc	aag	ctc	ata	gag	ttt	atg	gat	acc	ttc	384
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Phe	Phe	Ile	Leu	Arg	Lys	Asn	Asn	His	Gln	Ile	Thr	Phe	Leu	His	Ile	
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Tyr	His	His	Ala	Ser	Met	Leu	Asn	Ile	Trp	Trp	Phe	Val	Met	Asn	Trp	
145					150					155					160	

gtg	ccc	tgt	ggt	cac	tcc	tac	ttt	ggt	gcc	tcc	ctg	aac	agc	ttc	atc	528
Val	Pro	Cys	Gly	His	Ser	Tyr	Phe	Gly	Ala	Ser	Leu	Asn	Ser	Phe	Ile	
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cat	gtc	ctg	atg	tac	tct	tac	tat	ggg	ctc	tct	gct	gtc	ccg	gcc	ttg	576
His	Val	Leu	Met	Tyr	Ser	Tyr	Tyr	Gly	Leu	Ser	Ala	Val	Pro	Ala	Leu	
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cgg	ccc	tat	cta	tgg	tgg	aag	aaa	tac	atc	aca	caa	gta	cag	ctg	att	624
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97

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 Cys Asp Phe Pro Arg Gly Trp Leu Tyr Phe Gln Ile Phe Tyr Val Ile
 225 230 235 240
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 245 250 255
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 His Leu Val Ser Gln Lys Lys Glu Tyr His Gln Asn Gly Ser Val Ala
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 tca ttg aat ggc cat gtg aat ggg gtg aca ccc acg gaa acc att aca 864
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 Gly Pro Lys Tyr Met Arg His Arg Gln Pro Val Ser Cys Arg Gly Leu
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 Tyr Glu Met Val Ser Ala Val Trp His Gly Asp Tyr Asn Phe Phe Cys
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98

Gln Asp Thr His Ser Ala Gly Glu Thr Asp Thr Lys Ile Ile Asn Val
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 115 120 125

Phe Phe Ile Leu Arg Lys Asn Asn His Gln Ile Thr Phe Leu His Ile
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Tyr His His Ala Ser Met Leu Asn Ile Trp Trp Phe Val Met Asn Trp
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Val Pro Cys Gly His Ser Tyr Phe Gly Ala Ser Leu Asn Ser Phe Ile
 165 170 175

His Val Leu Met Tyr Ser Tyr Tyr Gly Leu Ser Ala Val Pro Ala Leu
 180 185 190

Arg Pro Tyr Leu Trp Trp Lys Lys Tyr Ile Thr Gln Val Gln Leu Ile
 195 200 205

Gln Phe Phe Leu Thr Met Ser Gln Thr Ile Cys Ala Val Ile Trp Pro
 210 215 220

Cys Asp Phe Pro Arg Gly Trp Leu Tyr Phe Gln Ile Phe Tyr Val Ile
 225 230 235 240

Thr Leu Ile Ala Leu Phe Ser Asn Phe Tyr Ile Gln Thr Tyr Lys Lys
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His Leu Val Ser Gln Lys Lys Glu Tyr His Gln Asn Gly Ser Val Ala
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His Arg Lys Val Arg Gly Asp
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acaataaaga ttctacaata ctagctttta tggttatgaa gaggaaaaat tggcagtaac 180

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cggtttgat tacttcttat tcaaatgtaa taaaagtatc aacaaaaaat tgttaataata 420

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cgactcacta tagggaatat taagcttaca ta atg gag act ttt aat tat aaa 533

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Leu Asn Met Tyr Ile Asp Ser Trp Met Gly Pro Arg Asp Glu Arg Val
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Gln Gly Trp Leu Leu Leu Asp Asn Tyr Pro Pro Thr Phe Ala Leu Thr
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gtc atg tac ctg ctg atc gta tgg atg ggg ccc aag tac atg aga cac 677

Val Met Tyr Leu Leu Ile Val Trp Met Gly Pro Lys Tyr Met Arg His
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aga cag ccg gtg tct tgc cgg ggt ctc ctc ttg gtc tac aat ctg ggc 725

Arg Gln Pro Val Ser Cys Arg Gly Leu Leu Leu Val Tyr Asn Leu Gly
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ctc acg atc ttg tcc ttc tat atg ttc tat gag atg gtg tct gct gtg 773

Leu Thr Ile Leu Ser Phe Tyr Met Phe Tyr Glu Met Val Ser Ala Val
75 80 85

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Trp His Gly Asp Tyr Asn Phe Phe Cys Gln Asp Thr His Ser Ala Gly
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Glu Thr Asp Thr Lys Ile Ile Asn Val Leu Trp Trp Tyr Tyr Phe Ser
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Lys Leu Ile Glu Phe Met Asp Thr Phe Phe Phe Ile Leu Arg Lys Asn
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Asn His Gln Ile Thr Phe Leu His Ile Tyr His His Ala Ser Met Leu
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Phe Gly Ala Ser Leu Asn Ser Phe Ile His Val Leu Met Tyr Ser Tyr	
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Asn Phe Tyr Ile Gln Thr Tyr Lys Lys His Leu Val Ser Gln Lys Lys	
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Gly Pro Lys Tyr Met Arg His Arg Gln Pro Val Ser Cys Arg Gly Leu
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Leu Leu Val Tyr Asn Leu Gly Leu Thr Ile Leu Ser Phe Tyr Met Phe
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Tyr Glu Met Val Ser Ala Val Trp His Gly Asp Tyr Asn Phe Phe Cys
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Gln Asp Thr His Ser Ala Gly Glu Thr Asp Thr Lys Ile Ile Asn Val
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Phe Phe Ile Leu Arg Lys Asn Asn His Gln Ile Thr Phe Leu His Ile
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Tyr His His Ala Ser Met Leu Asn Ile Trp Trp Phe Val Met Asn Trp
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Val Pro Cys Gly His Ser Tyr Phe Gly Ala Ser Leu Asn Ser Phe Ile
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Thr Leu Ile Ala Leu Phe Ser Asn Phe Tyr Ile Gln Thr Tyr Lys Lys
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Asp Pro Trp Leu Leu Val Tyr Ser Pro Met Pro Val Ala Ile Ile Phe
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Leu Leu Tyr Leu Gly Val Val Trp Ala Gly Pro Lys Leu Met Lys Arg
40                               45                               50                               55

agg gaa cca gtt gat ctc aag gct gta ctc att gtc tac aac ttc gcc      725
Arg Glu Pro Val Asp Leu Lys Ala Val Leu Ile Val Tyr Asn Phe Ala
                               60                               65                               70

atg gtc tgc ctg tct gtc tac atg ttc cat gag ttc ttg gtc acg tcc      773
Met Val Cys Leu Ser Val Tyr Met Phe His Glu Phe Leu Val Thr Ser
                               75                               80                               85

ttg ctg tct aac tac agt tac ctg tgt caa cct gtg gat tac agc act      821
Leu Leu Ser Asn Tyr Ser Tyr Leu Cys Gln Pro Val Asp Tyr Ser Thr
                               90                               95                               100

agt cca ctg gcg atg agg atg gcc aaa gta tgc tgg tgg ttt ttc ttc      869
Ser Pro Leu Ala Met Arg Met Ala Lys Val Cys Trp Trp Phe Phe Phe
105                               110                               115

tcc aag gtc ata gaa ttg gct gac acg gtg ttc ttc atc ctg agg aag      917
Ser Lys Val Ile Glu Leu Ala Asp Thr Val Phe Phe Ile Leu Arg Lys
120                               125                               130                               135

aag aac agt cag ctg act ttc ctg cat gtc tat cac cat ggc acc atg      965
Lys Asn Ser Gln Leu Thr Phe Leu His Val Tyr His His Gly Thr Met
                               140                               145                               150

atc ttc aac tgg tgg gca ggg gtc aag tat ctg gct gga ggc caa tcg      1013

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106

Ile Phe Asn Trp Trp Ala Gly Val Lys Tyr Leu Ala Gly Gly Gln Ser	
155 160 165	
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Phe Phe Ile Gly Leu Leu Asn Thr Phe Val His Ile Val Met Tyr Ser	
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tac tac gga ctg gct gcc ctg ggg cct cac acg cag aag tac tta tgg	1109
Tyr Tyr Gly Leu Ala Ala Leu Gly Pro His Thr Gln Lys Tyr Leu Trp	
185 190 195	
tgg aag cgc tat ctg acc tca ctg cag ctg ctc cag ttt gtc ctg ttg	1157
Trp Lys Arg Tyr Leu Thr Ser Leu Gln Leu Leu Gln Phe Val Leu Leu	
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acc act cac act ggc tac aac ctc ttc act gag tgt gac ttc ccg gac	1205
Thr Thr His Thr Gly Tyr Asn Leu Phe Thr Glu Cys Asp Phe Pro Asp	
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tcc atg aac gct gtg gtg ttt gcc tac tgt gtc agt ctc att gct ctc	1253
Ser Met Asn Ala Val Val Phe Ala Tyr Cys Val Ser Leu Ile Ala Leu	
235 240 245	
ttc agc aac ttc tac tat cag agc tac ctc aac agg aag agc aag aag	1301
Phe Ser Asn Phe Tyr Tyr Gln Ser Tyr Leu Asn Arg Lys Ser Lys Lys	
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<210> 58

<211> 264

<212> PRT

<213> *Oncorhynchus mykiss*

<400> 58

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Met Ala Ser Thr Trp Gln Ser Val Gln Ser Met Arg Gln Trp Ile Leu
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Glu Asn Gly Asp Lys Arg Thr Asp Pro Trp Leu Leu Val Tyr Ser Pro
20           25           30

```

```

Met Pro Val Ala Ile Ile Phe Leu Leu Tyr Leu Gly Val Val Trp Ala
35           40           45

```

```

Gly Pro Lys Leu Met Lys Arg Arg Glu Pro Val Asp Leu Lys Ala Val
50           55           60

```

```

Leu Ile Val Tyr Asn Phe Ala Met Val Cys Leu Ser Val Tyr Met Phe
65           70           75           80

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```

His Glu Phe Leu Val Thr Ser Leu Leu Ser Asn Tyr Ser Tyr Leu Cys
85           90           95

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Gln Pro Val Asp Tyr Ser Thr Ser Pro Leu Ala Met Arg Met Ala Lys
100          105          110

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Val Cys Trp Trp Phe Phe Phe Ser Lys Val Ile Glu Leu Ala Asp Thr
115          120          125

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Val Phe Phe Ile Leu Arg Lys Lys Asn Ser Gln Leu Thr Phe Leu His
130          135          140

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Val Tyr His His Gly Thr Met Ile Phe Asn Trp Trp Ala Gly Val Lys
145 150 155 160

Tyr Leu Ala Gly Gly Gln Ser Phe Phe Ile Gly Leu Leu Asn Thr Phe
165 170 175

Val His Ile Val Met Tyr Ser Tyr Tyr Gly Leu Ala Ala Leu Gly Pro
180 185 190

His Thr Gln Lys Tyr Leu Trp Trp Lys Arg Tyr Leu Thr Ser Leu Gln
195 200 205

Leu Leu Gln Phe Val Leu Leu Thr Thr His Thr Gly Tyr Asn Leu Phe
210 215 220

Thr Glu Cys Asp Phe Pro Asp Ser Met Asn Ala Val Val Phe Ala Tyr
225 230 235 240

Cys Val Ser Leu Ile Ala Leu Phe Ser Asn Phe Tyr Tyr Gln Ser Tyr
245 250 255

Leu Asn Arg Lys Ser Lys Lys Thr
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<210> 59

<211> 1077

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(1077)

<223> Delta-5 elongase

<400> 59

atg tgc tca tca ccg ccg tca caa tcc aaa aca aca tcc ctc cta gca 48
Met Cys Ser Ser Pro Pro Ser Gln Ser Lys Thr Thr Ser Leu Leu Ala
1 5 10 15

cgg tac acc acc gcc gcc ctc ctc ctc ctc acc ctc aca aca tgg tgc 96
Arg Tyr Thr Thr Ala Ala Leu Leu Leu Leu Thr Leu Thr Thr Trp Cys
20 25 30

cac ttc gcc ttc cca gcc gcc acc gcc aca ccc ggc ctc acc gcc gaa 144

111

His	Phe	Ala	Phe	Pro	Ala	Ala	Thr	Ala	Thr	Pro	Gly	Leu	Thr	Ala	Glu	
		35					40					45				
atg	cac	tcc	tac	aaa	gtc	cca	ctc	ggc	ctc	acc	gta	ttc	tac	ctg	ctg	192
Met	His	Ser	Tyr	Lys	Val	Pro	Leu	Gly	Leu	Thr	Val	Phe	Tyr	Leu	Leu	
	50					55					60					
agt	cta	ccg	tca	cta	aag	tac	gtt	acg	gac	aac	tac	ctt	gcc	aaa	aag	240
Ser	Leu	Pro	Ser	Leu	Lys	Tyr	Val	Thr	Asp	Asn	Tyr	Leu	Ala	Lys	Lys	
65					70					75					80	
tat	gat	atg	aag	tca	ctc	cta	acg	gaa	tca	atg	gtg	ttg	tac	aat	gtg	288
Tyr	Asp	Met	Lys	Ser	Leu	Leu	Thr	Glu	Ser	Met	Val	Leu	Tyr	Asn	Val	
			85						90					95		
gcg	caa	gtg	ctg	ctc	aat	ggg	tgg	acg	gtg	tat	gcg	att	gtg	gat	gcg	336
Ala	Gln	Val	Leu	Asn	Gly	Trp	Thr	Val	Tyr	Ala	Ile	Val	Asp	Ala		
		100					105					110				
gtg	atg	aat	aga	gac	cat	ccg	ttt	att	gga	agt	aga	agt	ttg	gtt	ggg	384
Val	Met	Asn	Arg	Asp	His	Pro	Phe	Ile	Gly	Ser	Arg	Ser	Leu	Val	Gly	
		115					120					125				
gcg	gcg	ttg	cat	agt	ggg	agc	tcg	tat	gcg	gtg	tgg	gtt	cat	tat	tgt	432
Ala	Ala	Leu	His	Ser	Gly	Ser	Ser	Tyr	Ala	Val	Trp	Val	His	Tyr	Cys	
		130				135					140					
gat	aag	tat	ttg	gag	ttc	ttt	gat	acg	tat	ttt	atg	gtg	ttg	agg	ggg	480
Asp	Lys	Tyr	Leu	Glu	Phe	Phe	Asp	Thr	Tyr	Phe	Met	Val	Leu	Arg	Gly	
145					150					155					160	
aaa	atg	gac	cag	gtc	tcc	ttc	ctc	cac	atc	tac	cac	cac	acg	acc	ata	528
Lys	Met	Asp	Gln	Val	Ser	Phe	Leu	His	Ile	Tyr	His	His	Thr	Thr	Ile	
			165						170					175		
gcg	tgg	gca	tgg	tgg	atc	gcc	ctc	cgc	ttc	tcc	ccc	ggc	gga	gac	att	576
Ala	Trp	Ala	Trp	Trp	Ile	Ala	Leu	Arg	Phe	Ser	Pro	Gly	Gly	Asp	Ile	
			180					185					190			
tac	ttc	ggg	gca	ctc	ctc	aac	tcc	atc	atc	cac	gtc	ctc	atg	tat	tcc	624
Tyr	Phe	Gly	Ala	Leu	Leu	Asn	Ser	Ile	Ile	His	Val	Leu	Met	Tyr	Ser	
		195					200					205				
tac	tac	gcc	ctt	gcc	cta	ctc	aag	gtc	agt	tgt	cca	tgg	aaa	cga	tac	672
Tyr	Tyr	Ala	Leu	Ala	Leu	Leu	Lys	Val	Ser	Cys	Pro	Trp	Lys	Arg	Tyr	
		210				215					220					
ctg	act	caa	gct	caa	tta	ttg	caa	ttc	aca	agt	gtg	gtg	gtt	tat	acg	720
Leu	Thr	Gln	Ala	Gln	Leu	Leu	Gln	Phe	Thr	Ser	Val	Val	Val	Tyr	Thr	
225					230					235					240	
ggg	tgt	acg	ggc	tat	act	cat	tac	tat	cat	acg	aag	cat	gga	gcg	gat	768
Gly	Cys	Thr	Gly	Tyr	Thr	His	Tyr	Tyr	His	Thr	Lys	His	Gly	Ala	Asp	
				245					250					255		
gag	aca	cag	cct	agt	tta	gga	acg	tat	tat	ttc	tgt	tgt	gga	gtg	cag	816
Glu	Thr	Gln	Pro	Ser	Leu	Gly	Thr	Tyr	Tyr	Phe	Cys	Cys	Gly	Val	Gln	
			260					265					270			
gtg	ttt	gag	atg	gtt	agt	ttg	ttt	gta	ctc	ttt	tcc	atc	ttt	tat	aaa	864
Val	Phe	Glu	Met	Val	Ser	Leu	Phe	Val	Leu	Phe	Ser	Ile	Phe	Tyr	Lys	
		275					280					285				
cga	tcc	tat	tcg	aag	aag	aac	aag	tca	gga	gga	aag	gat	agc	aag	aag	912

112

Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp Ser Lys Lys
 290 295 300
 aat gat gat ggg aat aat gag gat caa tgt cac aag gct atg aag gat 960
 Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala Met Lys Asp
 305 310 315 320
 ata tcg gag ggt gcg aag gag gtt gtg ggg cat gca gcg aag gat gct 1008
 Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala Lys Asp Ala
 325 330 335
 gga aag ttg gtg gct acg gcg agt aag gct gta aag agg aag gga act 1056
 Gly Lys Leu Val Ala Thr Ala Ser Lys Ala Val Lys Arg Lys Gly Thr
 340 345 350
 cgt gtt act ggt gcc atg tag 1077
 Arg Val Thr Gly Ala Met
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<210> 60

<211> 358

<212> PRT

<213> Thalassiosira pseudonana

<400> 60

Met Cys Ser Ser Pro Pro Ser Gln Ser Lys Thr Thr Ser Leu Leu Ala
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Arg Tyr Thr Thr Ala Ala Leu Leu Leu Leu Thr Leu Thr Thr Trp Cys
 20 25 30

His Phe Ala Phe Pro Ala Ala Thr Ala Thr Pro Gly Leu Thr Ala Glu
 35 40 45

Met His Ser Tyr Lys Val Pro Leu Gly Leu Thr Val Phe Tyr Leu Leu
 50 55 60

Ser Leu Pro Ser Leu Lys Tyr Val Thr Asp Asn Tyr Leu Ala Lys Lys
 65 70 75 80

Tyr Asp Met Lys Ser Leu Leu Thr Glu Ser Met Val Leu Tyr Asn Val
 85 90 95

Ala Gln Val Leu Leu Asn Gly Trp Thr Val Tyr Ala Ile Val Asp Ala
 100 105 110

Val Met Asn Arg Asp His Pro Phe Ile Gly Ser Arg Ser Leu Val Gly
 115 120 125

113

Ala Ala Leu His Ser Gly Ser Ser Tyr Ala Val Trp Val His Tyr Cys
 130 135 140

Asp Lys Tyr Leu Glu Phe Phe Asp Thr Tyr Phe Met Val Leu Arg Gly
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Lys Met Asp Gln Val Ser Phe Leu His Ile Tyr His His Thr Thr Ile
 165 170 175

Ala Trp Ala Trp Trp Ile Ala Leu Arg Phe Ser Pro Gly Gly Asp Ile
 180 185 190

Tyr Phe Gly Ala Leu Leu Asn Ser Ile Ile His Val Leu Met Tyr Ser
 195 200 205

Tyr Tyr Ala Leu Ala Leu Leu Lys Val Ser Cys Pro Trp Lys Arg Tyr
 210 215 220

Leu Thr Gln Ala Gln Leu Leu Gln Phe Thr Ser Val Val Val Tyr Thr
 225 230 235 240

Gly Cys Thr Gly Tyr Thr His Tyr Tyr His Thr Lys His Gly Ala Asp
 245 250 255

Glu Thr Gln Pro Ser Leu Gly Thr Tyr Tyr Phe Cys Cys Gly Val Gln
 260 265 270

Val Phe Glu Met Val Ser Leu Phe Val Leu Phe Ser Ile Phe Tyr Lys
 275 280 285

Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp Ser Lys Lys
 290 295 300

Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala Met Lys Asp
 305 310 315 320

Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala Lys Asp Ala
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Gly Lys Leu Val Ala Thr Ala Ser Lys Ala Val Lys Arg Lys Gly Thr
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Arg Val Thr Gly Ala Met
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<210> 61

<211> 933

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(933)

<223> Delta-5 elongase

<400> 61

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Ser	Leu	Pro	Ser	Leu	Lys	Tyr	Val	Thr	Asp	Asn	Tyr	Leu	Ala	Lys	Lys	
			20					25					30			

tat	gat	atg	aag	tca	ctc	cta	acg	gaa	tca	atg	gtg	ttg	tac	aat	gtg	144
Tyr	Asp	Met	Lys	Ser	Leu	Leu	Thr	Glu	Ser	Met	Val	Leu	Tyr	Asn	Val	
		35					40					45				

gcg	caa	gtg	ctg	ctc	aat	ggg	tgg	acg	gtg	tat	gcg	att	gtg	gat	gcg	192
Ala	Gln	Val	Leu	Leu	Asn	Gly	Trp	Thr	Val	Tyr	Ala	Ile	Val	Asp	Ala	
	50					55					60					

gtg	atg	aat	aga	gac	cat	ccg	ttt	att	gga	agt	aga	agt	ttg	gtt	ggg	240
Val	Met	Asn	Arg	Asp	His	Pro	Phe	Ile	Gly	Ser	Arg	Ser	Leu	Val	Gly	
65					70					75				80		

gcg	gcg	ttg	cat	agt	ggg	agc	tcg	tat	gcg	gtg	tgg	gtt	cat	tat	tgt	288
Ala	Ala	Leu	His	Ser	Gly	Ser	Ser	Tyr	Ala	Val	Trp	Val	His	Tyr	Cys	
			85						90				95			

gat	aag	tat	ttg	gag	ttc	ttt	gat	acg	tat	ttt	atg	gtg	ttg	agg	ggg	336
Asp	Lys	Tyr	Leu	Glu	Phe	Phe	Asp	Thr	Tyr	Phe	Met	Val	Leu	Arg	Gly	
			100					105					110			

aaa	atg	gac	cag	gtc	tcc	ttc	ctc	cac	atc	tac	cac	cac	acg	acc	ata	384
Lys	Met	Asp	Gln	Val	Ser	Phe	Leu	His	Ile	Tyr	His	His	Thr	Thr	Ile	
		115					120					125				

gcg	tgg	gca	tgg	tgg	atc	gcc	ctc	cgc	ttc	tcc	ccc	ggg	gga	gac	att	432
Ala	Trp	Ala	Trp	Trp	Ile	Ala	Leu	Arg	Phe	Ser	Pro	Gly	Gly	Asp	Ile	
	130					135					140					

tac	ttc	ggg	gca	ctc	ctc	aac	tcc	atc	atc	cac	gtc	ctc	atg	tat	tcc	480
Tyr	Phe	Gly	Ala	Leu	Leu	Asn	Ser	Ile	Ile	His	Val	Leu	Met	Tyr	Ser	
145					150					155					160	

tac	tac	gcc	ctt	gcc	cta	ctc	aag	gtc	agt	tgt	cca	tgg	aaa	cga	tac	528
Tyr	Tyr	Ala	Leu	Ala	Leu	Leu	Lys	Val	Ser	Cys	Pro	Trp	Lys	Arg	Tyr	
			165					170					175			

ctg	act	caa	gct	caa	tta	ttg	caa	ttc	aca	agt	gtg	gtg	gtt	tat	acg	576
Leu	Thr	Gln	Ala	Gln	Leu	Leu	Gln	Phe	Thr	Ser	Val	Val	Val	Tyr	Thr	
			180					185							190	

ggg tgt acg ggt tat act cat tac tat cat acg aag cat gga gcg gat 624
 Gly Cys Thr Gly Tyr Thr His Tyr Tyr His Thr Lys His Gly Ala Asp
 195 200 205
 gag aca cag cct agt tta gga acg tat tat ttc tgt tgt gga gtg cag 672
 Glu Thr Gln Pro Ser Leu Gly Thr Tyr Tyr Phe Cys Cys Gly Val Gln
 210 215 220
 gtg ttt gag atg gtt agt ttg ttt gta ctc ttt tcc atc ttt tat aaa 720
 Val Phe Glu Met Val Ser Leu Phe Val Leu Phe Ser Ile Phe Tyr Lys
 225 230 235 240
 cga tcc tat tcg aag aag aac aag tca gga gga aag gat agc aag aag 768
 Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp Ser Lys Lys
 245 250 255
 aat gat gat ggg aat aat gag gat caa tgt cac aag gct atg aag gat 816
 Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala Met Lys Asp
 260 265 270
 ata tcg gag ggt gcg aag gag gtt gtg ggg cat gca gcg aag gat gct 864
 Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala Lys Asp Ala
 275 280 285
 gga aag ttg gtg gct acg gcg agt aag gct gta aag agg aag gga act 912
 Gly Lys Leu Val Ala Thr Ala Ser Lys Ala Val Lys Arg Lys Gly Thr
 290 295 300
 cgt gtt act ggt gcc atg tag 933
 Arg Val Thr Gly Ala Met
 305 310

<210> 62

<211> 310

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 62

Met His Ser Tyr Lys Val Pro Leu Gly Leu Thr Val Phe Tyr Leu Leu
 1 5 10 15
 Ser Leu Pro Ser Leu Lys Tyr Val Thr Asp Asn Tyr Leu Ala Lys Lys
 20 25 30
 Tyr Asp Met Lys Ser Leu Leu Thr Glu Ser Met Val Leu Tyr Asn Val
 35 40 45
 Ala Gln Val Leu Leu Asn Gly Trp Thr Val Tyr Ala Ile Val Asp Ala
 50 55 60
 Val Met Asn Arg Asp His Pro Phe Ile Gly Ser Arg Ser Leu Val Gly
 65 70 75 80

Ala Ala Leu His Ser Gly Ser Ser Tyr Ala Val Trp Val His Tyr Cys
85 90 95

Asp Lys Tyr Leu Glu Phe Phe Asp Thr Tyr Phe Met Val Leu Arg Gly
100 105 110

Lys Met Asp Gln Val Ser Phe Leu His Ile Tyr His His Thr Thr Ile
115 120 125

Ala Trp Ala Trp Trp Ile Ala Leu Arg Phe Ser Pro Gly Gly Asp Ile
130 135 140

Tyr Phe Gly Ala Leu Leu Asn Ser Ile Ile His Val Leu Met Tyr Ser
145 150 155 160

Tyr Tyr Ala Leu Ala Leu Leu Lys Val Ser Cys Pro Trp Lys Arg Tyr
165 170 175

Leu Thr Gln Ala Gln Leu Leu Gln Phe Thr Ser Val Val Val Tyr Thr
180 185 190

Gly Cys Thr Gly Tyr Thr His Tyr Tyr His Thr Lys His Gly Ala Asp
195 200 205

Glu Thr Gln Pro Ser Leu Gly Thr Tyr Tyr Phe Cys Cys Gly Val Gln
210 215 220

Val Phe Glu Met Val Ser Leu Phe Val Leu Phe Ser Ile Phe Tyr Lys
225 230 235 240

Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp Ser Lys Lys
245 250 255

Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala Met Lys Asp
260 265 270

Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala Lys Asp Ala
275 280 285

Gly Lys Leu Val Ala Thr Ala Ser Lys Ala Val Lys Arg Lys Gly Thr
290 295 300

Arg Val Thr Gly Ala Met
305 310

<210> 63

<211> 933

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(933)

<223> Delta-5 elongase

<400> 63

atg	cac	tcc	tac	aaa	gtc	cca	ctc	ggt	ctc	acc	gta	ttc	tac	ctg	ctg	48
Met	His	Ser	Tyr	Lys	Val	Pro	Leu	Gly	Leu	Thr	Val	Phe	Tyr	Leu	Leu	
1				5					10					15		

agt	cta	ccg	tca	cta	aag	tac	gtt	acg	gac	aac	tac	ctt	gcc	aaa	aag	96
Ser	Leu	Pro	Ser	Leu	Lys	Tyr	Val	Thr	Asp	Asn	Tyr	Leu	Ala	Lys	Lys	
			20					25					30			

tat	gat	atg	aag	tca	ctc	cta	acg	gaa	tca	atg	gtg	ttg	tac	aat	gtg	144
Tyr	Asp	Met	Lys	Ser	Leu	Leu	Thr	Glu	Ser	Met	Val	Leu	Tyr	Asn	Val	
		35					40					45				

gcg	caa	gtg	ctg	ctc	aat	ggg	tgg	acg	gtg	tat	gcg	att	gtg	gat	gcg	192
Ala	Gln	Val	Leu	Leu	Asn	Gly	Trp	Thr	Val	Tyr	Ala	Ile	Val	Asp	Ala	
	50					55					60					

gtg	atg	aat	aga	gac	cat	ccg	ttt	att	gga	agt	aga	agt	ttg	gtt	ggg	240
Val	Met	Asn	Arg	Asp	His	Pro	Phe	Ile	Gly	Ser	Arg	Ser	Leu	Val	Gly	
65					70					75					80	

gcg	gcg	ttg	cat	agt	ggg	agc	tgc	tat	gcg	gtg	tgg	gtt	cat	tat	tgt	288
Ala	Ala	Leu	His	Ser	Gly	Ser	Ser	Tyr	Ala	Val	Trp	Val	His	Tyr	Cys	
			85						90					95		

gat	aag	tat	ttg	gag	ttc	ttt	gat	acg	tat	ttt	atg	gtg	ttg	agg	ggg	336
Asp	Lys	Tyr	Leu	Glu	Phe	Phe	Asp	Thr	Tyr	Phe	Met	Val	Leu	Arg	Gly	
			100					105					110			

aaa	atg	gac	cag	gtc	tcc	ttc	ctc	cac	atc	tac	cac	cac	acg	acc	ata	384
Lys	Met	Asp	Gln	Val	Ser	Phe	Leu	His	Ile	Tyr	His	His	Thr	Thr	Ile	
		115					120					125				

gcg	tgg	gca	tgg	tgg	atc	gcc	ctc	cgc	ttc	tcc	ccc	ggt	gga	gac	att	432
Ala	Trp	Ala	Trp	Trp	Ile	Ala	Leu	Arg	Phe	Ser	Pro	Gly	Gly	Asp	Ile	
	130					135					140					

tac	ttc	ggg	gca	ctc	ctc	aac	tcc	atc	atc	cac	gtc	ctc	atg	tat	tcc	480
Tyr	Phe	Gly	Ala	Leu	Leu	Asn	Ser	Ile	Ile	His	Val	Leu	Met	Tyr	Ser	
145					150					155					160	

tac	tac	gcc	ctt	gcc	cta	ctc	aag	gtc	agt	tgt	cca	tgg	aaa	cga	tac	528
Tyr	Tyr	Ala	Leu	Ala	Leu	Leu	Lys	Val	Ser	Cys	Pro	Trp	Lys	Arg	Tyr	
			165						170					175		

ctg	act	caa	gct	caa	tta	ttg	caa	ttc	aca	agt	gtg	gtg	gtt	tat	acg	576
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

118

Leu Thr Gln Ala Gln Leu Leu Gln Phe Thr Ser Val Val Val Tyr Thr
 180 185 190
 ggg tgt acg ggt tat act cat tac tat cat acg aag cat gga gcg gat 624
 Gly Cys Thr Gly Tyr Thr His Tyr Tyr His Thr Lys His Gly Ala Asp
 195 200 205
 gag aca cag cct agt tta gga acg tat tat ttc tgt tgt gga gtg cag 672
 Glu Thr Gln Pro Ser Leu Gly Thr Tyr Tyr Phe Cys Cys Gly Val Gln
 210 215 220
 gtg ttt gag atg gtt agt ttg ttt gta ctc ttt tcc atc ttt tat aaa 720
 Val Phe Glu Met Val Ser Leu Phe Val Leu Phe Ser Ile Phe Tyr Lys
 225 230 235 240
 cga tcc tat tcg aag aag aac aag tca gga gga aag gat agc aag aag 768
 Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp Ser Lys Lys
 245 250 255
 aat gat gat ggg aat aat gag gat caa tgt cac aag gct atg aag gat 816
 Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala Met Lys Asp
 260 265 270
 ata tcg gag ggt gcg aag gag gtt gtg ggg cat gca gcg aag gat gct 864
 Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala Lys Asp Ala
 275 280 285
 gga aag ttg gtg gct acg gcg agt aag gct gta aag agg aag gga act 912
 Gly Lys Leu Val Ala Thr Ala Ser Lys Ala Val Lys Arg Lys Gly Thr
 290 295 300
 cgt gtt act ggt gcc atg tag 933
 Arg Val Thr Gly Ala Met
 305 310

<210> 64

<211> 310

<212> PRT

<213> Thalassiosira pseudonana

<400> 64

Met His Ser Tyr Lys Val Pro Leu Gly Leu Thr Val Phe Tyr Leu Leu
 1 5 10 15
 Ser Leu Pro Ser Leu Lys Tyr Val Thr Asp Asn Tyr Leu Ala Lys Lys
 20 25 30
 Tyr Asp Met Lys Ser Leu Leu Thr Glu Ser Met Val Leu Tyr Asn Val
 35 40 45
 Ala Gln Val Leu Leu Asn Gly Trp Thr Val Tyr Ala Ile Val Asp Ala
 50 55 60

119

Val Met Asn Arg Asp His Pro Phe Ile Gly Ser Arg Ser Leu Val Gly
65 70 75 80

Ala Ala Leu His Ser Gly Ser Ser Tyr Ala Val Trp Val His Tyr Cys
85 90 95

Asp Lys Tyr Leu Glu Phe Phe Asp Thr Tyr Phe Met Val Leu Arg Gly
100 105 110

Lys Met Asp Gln Val Ser Phe Leu His Ile Tyr His His Thr Thr Ile
115 120 125

Ala Trp Ala Trp Trp Ile Ala Leu Arg Phe Ser Pro Gly Gly Asp Ile
130 135 140

Tyr Phe Gly Ala Leu Leu Asn Ser Ile Ile His Val Leu Met Tyr Ser
145 150 155 160

Tyr Tyr Ala Leu Ala Leu Leu Lys Val Ser Cys Pro Trp Lys Arg Tyr
165 170 175

Leu Thr Gln Ala Gln Leu Leu Gln Phe Thr Ser Val Val Val Tyr Thr
180 185 190

Gly Cys Thr Gly Tyr Thr His Tyr Tyr His Thr Lys His Gly Ala Asp
195 200 205

Glu Thr Gln Pro Ser Leu Gly Thr Tyr Tyr Phe Cys Cys Gly Val Gln
210 215 220

Val Phe Glu Met Val Ser Leu Phe Val Leu Phe Ser Ile Phe Tyr Lys
225 230 235 240

Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp Ser Lys Lys
245 250 255

Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala Met Lys Asp
260 265 270

Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala Lys Asp Ala
275 280 285

Gly Lys Leu Val Ala Thr Ala Ser Lys Ala Val Lys Arg Lys Gly Thr
290 295 300

Arg Val Thr Gly Ala Met
305 310

<210> 65

<211> 825

<212> DNA

<213> *Thraustochytrium aureum*

<220>

<221> CDS

<222> (1)..(825)

<223> Delta-5 elongase

<400> 65

atg	acg	agc	aac	atg	agc	gcg	tgg	ggc	gtc	gcc	gtc	gac	cag	acg	cag	48
Met	Thr	Ser	Asn	Met	Ser	Ala	Trp	Gly	Val	Ala	Val	Asp	Gln	Thr	Gln	
1				5					10				15			

cag	gtc	gtc	gac	cag	atc	atg	ggc	ggc	gcc	gag	ccg	tac	aag	ctg	aca	96
Gln	Val	Val	Asp	Gln	Ile	Met	Gly	Gly	Ala	Glu	Pro	Tyr	Lys	Leu	Thr	
			20				25						30			

gaa	ggg	cgc	atg	acg	aac	gtc	gag	acg	atg	ctg	gcg	atc	gag	tgc	ggc	144
Glu	Gly	Arg	Met	Thr	Asn	Val	Glu	Thr	Met	Leu	Ala	Ile	Glu	Cys	Gly	
		35				40						45				

tac	gcc	gcc	atg	ctg	ctg	ttc	ctg	acc	ccg	atc	atg	aag	cag	gcc	gag	192
Tyr	Ala	Ala	Met	Leu	Leu	Phe	Leu	Thr	Pro	Ile	Met	Lys	Gln	Ala	Glu	
	50					55					60					

aag	ccc	ttc	gag	ctc	aag	tcc	ttc	aag	ctc	gcc	cac	aac	ctg	ttc	ctg	240
Lys	Pro	Phe	Glu	Leu	Lys	Ser	Phe	Lys	Leu	Ala	His	Asn	Leu	Phe	Leu	
65					70					75				80		

ttc	gtc	ctg	tcc	gcc	tac	atg	tgc	ctc	gag	acc	gtc	cgc	cag	gcc	tac	288
Phe	Val	Leu	Ser	Ala	Tyr	Met	Cys	Leu	Glu	Thr	Val	Arg	Gln	Ala	Tyr	
			85					90					95			

ctt	gcg	ggc	tac	tcg	gtg	ttc	ggc	aac	gac	atg	gag	aag	ggc	agc	gag	336
Leu	Ala	Gly	Tyr	Ser	Val	Phe	Gly	Asn	Asp	Met	Glu	Lys	Gly	Ser	Glu	
			100				105						110			

ccg	cac	gcg	cac	ggc	atg	gcc	caa	atc	gtg	tgg	atc	ttt	tac	gtg	tcc	384
Pro	His	Ala	His	Gly	Met	Ala	Gln	Ile	Val	Trp	Ile	Phe	Tyr	Val	Ser	
		115				120						125				

aag	gcg	tac	gag	ttc	gtg	gac	acg	ctg	atc	atg	atc	ctg	tgc	aaa	aag	432
Lys	Ala	Tyr	Glu	Phe	Val	Asp	Thr	Leu	Ile	Met	Ile	Leu	Cys	Lys	Lys	
	130					135					140					

ttc	aac	cag	gtc	tcc	gtc	ctg	cac	gtg	tac	cac	cac	gcc	acc	atc	ttt	480
Phe	Asn	Gln	Val	Ser	Val	Leu	His	Val	Tyr	His	His	Ala	Thr	Ile	Phe	
145				150					155					160		

gct	atc	tgg	ttt	atg	atc	gcc	aag	tac	gcc	ccg	ggc	ggc	gac	gca	tac	528
Ala	Ile	Trp	Phe	Met	Ile	Ala	Lys	Tyr	Ala	Pro	Gly	Gly	Asp	Ala	Tyr	
				165					170					175		

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ttt agc gtc atc ctg aac tcg ttc gtg cac acc gtc atg tac gcg tac      576
Phe Ser Val Ile Leu Asn Ser Phe Val His Thr Val Met Tyr Ala Tyr
      180                      185                      190

tac ttc ttc tcg tcg cag ggc ttc ggg ttc gtc aag ccg atc aag ccg      624
Tyr Phe Phe Ser Ser Gln Gly Phe Gly Phe Val Lys Pro Ile Lys Pro
      195                      200                      205

tac atc acc tcg ctg cag atg acg cag ttc atg gcg atg ctc gtg cag      672
Tyr Ile Thr Ser Leu Gln Met Thr Gln Phe Met Ala Met Leu Val Gln
      210                      215                      220

tcg ctg tac gac tac ctt tac ccg tgc gac tac ccg cag ggg ctc gtc      720
Ser Leu Tyr Asp Tyr Leu Tyr Pro Cys Asp Tyr Pro Gln Gly Leu Val
      225                      230                      235                      240

aag ctc ctc ggc gtg tac atg ctc acc ctg ctt gcg ctc ttc ggc aac      768
Lys Leu Leu Gly Val Tyr Met Leu Thr Leu Leu Ala Leu Phe Gly Asn
      245                      250                      255

ttt ttc gtg cag agc tac ctc aag aag tcg aac aag ccc aag gcc aag      816
Phe Phe Val Gln Ser Tyr Leu Lys Lys Ser Asn Lys Pro Lys Ala Lys
      260                      265                      270

tcg gcc taa      825
Ser Ala

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<210> 66

<211> 274

<212> PRT

<213> Thraustochytrium aureum

<400> 66

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Met Thr Ser Asn Met Ser Ala Trp Gly Val Ala Val Asp Gln Thr Gln
1                      5                      10                      15

Gln Val Val Asp Gln Ile Met Gly Gly Ala Glu Pro Tyr Lys Leu Thr
      20                      25                      30

Glu Gly Arg Met Thr Asn Val Glu Thr Met Leu Ala Ile Glu Cys Gly
      35                      40                      45

Tyr Ala Ala Met Leu Leu Phe Leu Thr Pro Ile Met Lys Gln Ala Glu
      50                      55                      60

Lys Pro Phe Glu Leu Lys Ser Phe Lys Leu Ala His Asn Leu Phe Leu
      65                      70                      75                      80

Phe Val Leu Ser Ala Tyr Met Cys Leu Glu Thr Val Arg Gln Ala Tyr
      85                      90                      95

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Leu Ala Gly Tyr Ser Val Phe Gly Asn Asp Met Glu Lys Gly Ser Glu
 100 105 110

Pro His Ala His Gly Met Ala Gln Ile Val Trp Ile Phe Tyr Val Ser
 115 120 125

Lys Ala Tyr Glu Phe Val Asp Thr Leu Ile Met Ile Leu Cys Lys Lys
 130 135 140

Phe Asn Gln Val Ser Val Leu His Val Tyr His His Ala Thr Ile Phe
 145 150 155 160

Ala Ile Trp Phe Met Ile Ala Lys Tyr Ala Pro Gly Gly Asp Ala Tyr
 165 170 175

Phe Ser Val Ile Leu Asn Ser Phe Val His Thr Val Met Tyr Ala Tyr
 180 185 190

Tyr Phe Phe Ser Ser Gln Gly Phe Gly Phe Val Lys Pro Ile Lys Pro
 195 200 205

Tyr Ile Thr Ser Leu Gln Met Thr Gln Phe Met Ala Met Leu Val Gln
 210 215 220

Ser Leu Tyr Asp Tyr Leu Tyr Pro Cys Asp Tyr Pro Gln Gly Leu Val
 225 230 235 240

Lys Leu Leu Gly Val Tyr Met Leu Thr Leu Leu Ala Leu Phe Gly Asn
 245 250 255

Phe Phe Val Gln Ser Tyr Leu Lys Lys Ser Asn Lys Pro Lys Ala Lys
 260 265 270

Ser Ala

<210> 67

<211> 903

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(903)

<223> Delta-5 elongase

<400> 67

atg	agc	gcc	tcc	ggt	gcg	ctg	ctg	ccc	gcg	atc	gcg	ttc	gcc	gcg	tac	48
Met	Ser	Ala	Ser	Gly	Ala	Leu	Leu	Pro	Ala	Ile	Ala	Phe	Ala	Ala	Tyr	
1				5					10					15		

gcg	tac	gcg	acg	tac	gcc	tac	gcc	ttt	gag	tgg	tcg	cac	gcg	aat	ggc	96
Ala	Tyr	Ala	Thr	Tyr	Ala	Tyr	Ala	Phe	Glu	Trp	Ser	His	Ala	Asn	Gly	
			20					25					30			

atc	gac	aac	gtc	gac	gcg	cgc	gag	tgg	atc	ggt	gcg	ctg	tcg	ttg	agg	144
Ile	Asp	Asn	Val	Asp	Ala	Arg	Glu	Trp	Ile	Gly	Ala	Leu	Ser	Leu	Arg	
		35				40						45				

ctc	ccg	gcg	atc	gcg	acg	acg	atg	tac	ctg	ttg	ttc	tgc	ctg	gtc	gga	192
Leu	Pro	Ala	Ile	Ala	Thr	Thr	Met	Tyr	Leu	Leu	Phe	Cys	Leu	Val	Gly	
	50					55					60					

ccg	agg	ttg	atg	gcg	aag	cgc	gag	gcg	ttc	gac	ccg	aag	ggg	ttc	atg	240
Pro	Arg	Leu	Met	Ala	Lys	Arg	Glu	Ala	Phe	Asp	Pro	Lys	Gly	Phe	Met	
65					70				75					80		

ctg	gcg	tac	aat	gcg	tat	cag	acg	gcg	ttc	aac	gtc	gtc	gtg	ctc	ggg	288
Leu	Ala	Tyr	Asn	Ala	Tyr	Gln	Thr	Ala	Phe	Asn	Val	Val	Val	Leu	Gly	
			85						90					95		

atg	ttc	gcg	cga	gag	atc	tcg	ggg	ctg	ggg	cag	ccc	gtg	tgg	ggg	tca	336
Met	Phe	Ala	Arg	Glu	Ile	Ser	Gly	Leu	Gly	Gln	Pro	Val	Trp	Gly	Ser	
			100					105					110			

acc	atg	ccg	tgg	agc	gat	aga	aaa	tcg	ttt	aag	atc	ctc	ctc	ggg	gtg	384
Thr	Met	Pro	Trp	Ser	Asp	Arg	Lys	Ser	Phe	Lys	Ile	Leu	Leu	Gly	Val	
		115					120					125				

tgg	ttg	cac	tac	aac	aac	caa	tat	ttg	gag	cta	ttg	gac	act	gtg	ttc	432
Trp	Leu	His	Tyr	Asn	Asn	Gln	Tyr	Leu	Glu	Leu	Leu	Asp	Thr	Val	Phe	
	130					135					140					

atg	gtt	gcg	cgc	aag	aag	acg	aag	cag	ttg	agc	ttc	ttg	cac	gtt	tat	480
Met	Val	Ala	Arg	Lys	Lys	Thr	Lys	Gln	Leu	Ser	Phe	Leu	His	Val	Tyr	
145					150					155					160	

cat	cac	gcc	ctg	ttg	atc	tgg	gcg	tgg	tgg	ttg	gtg	tgt	cac	ttg	atg	528
His	His	Ala	Leu	Leu	Ile	Trp	Ala	Trp	Trp	Leu	Val	Cys	His	Leu	Met	
			165					170						175		

gcc	acg	aac	gat	tgt	atc	gat	gcc	tac	ttc	ggc	gcg	gcg	tgc	aac	tcg	576
Ala	Thr	Asn	Asp	Cys	Ile	Asp	Ala	Tyr	Phe	Gly	Ala	Ala	Cys	Asn	Ser	
			180					185					190			

ttc	att	cac	atc	gtg	atg	tac	tcg	tat	tat	ctc	atg	tcg	gcg	ctc	ggc	624
Phe	Ile	His	Ile	Val	Met	Tyr	Ser	Tyr	Tyr	Leu	Met	Ser	Ala	Leu	Gly	
		195					200					205				

att	cga	tgc	ccg	tgg	aag	cga	tac	atc	acc	cag	gct	caa	atg	ctc	caa	672
Ile	Arg	Cys	Pro	Trp	Lys	Arg	Tyr	Ile	Thr	Gln	Ala	Gln	Met	Leu	Gln	
	210					215					220					

ttc	gtc	att	gtc	ttc	gcg	cac	gcc	gtg	ttc	gtg	ctg	cgt	cag	aag	cac	720
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124

Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240
 tgc ccg gtc acc ctt cct tgg gcg caa atg ttc gtc atg acg aac atg 768
 Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255
 ctc gtg ctc ttc ggg aac ttc tac ctc aag gcg tac tcg aac aag tcg 816
 Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270
 cgc ggc gac ggc gcg agt tcc gtg aaa cca gcc gag acc acg cgc gcg 864
 Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285
 ccc agc gtg cga cgc acg cga tct cga aaa att gac taa 903
 Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 68

<211> 300

<212> PRT

<213> *Ostreococcus tauri*

<400> 68

Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Phe Ala Ala Tyr
 1 5 10 15
 Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30
 Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45
 Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly
 50 55 60
 Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met
 65 70 75 80
 Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly
 85 90 95
 Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser
 100 105 110
 Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val
 115 120 125

125

Trp Leu His Tyr Asn Asn Gln Tyr Leu Glu Leu Leu Asp Thr Val Phe
 130 135 140

Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr
 145 150 155 160

His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met
 165 170 175

Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser
 180 185 190

Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
 195 200 205

Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
 210 215 220

Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240

Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255

Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270

Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285

Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 69

<211> 879

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(879)

<223> Delta-6 elongase

<400> 69
 atg agt ggc tta cgt gca ccc aac ttt tta cac aga ttc tgg aca aag 48
 Met Ser Gly Leu Arg Ala Pro Asn Phe Leu His Arg Phe Trp Thr Lys
 1 5 10 15

tgg gac tac gcg att tcc aaa gtc gtc ttc acg tgt gcc gac agt ttt 96
 Trp Asp Tyr Ala Ile Ser Lys Val Val Phe Thr Cys Ala Asp Ser Phe
 20 25 30

cag tgg gac atc ggg cca gtg agt tcg agt acg gcg cat tta ccc gcc 144
 Gln Trp Asp Ile Gly Pro Val Ser Ser Ser Thr Ala His Leu Pro Ala
 35 40 45

att gaa tcc cct acc cca ctg gtg act agc ctc ttg ttc tac tta gtc 192
 Ile Glu Ser Pro Thr Pro Leu Val Thr Ser Leu Leu Phe Tyr Leu Val
 50 55 60

aca gtt ttc ttg tgg tat ggt cgt tta acc agg agt tca gac aag aaa 240
 Thr Val Phe Leu Trp Tyr Gly Arg Leu Thr Arg Ser Ser Asp Lys Lys
 65 70 75 80

att aga gag cct acg tgg tta aga aga ttc ata ata tgt cat aat gcg 288
 Ile Arg Glu Pro Thr Trp Leu Arg Arg Phe Ile Ile Cys His Asn Ala
 85 90 95

ttc ttg ata gtc ctc agt ctt tac atg tgc ctt ggt tgt gtg gcc caa 336
 Phe Leu Ile Val Leu Ser Leu Tyr Met Cys Leu Gly Cys Val Ala Gln
 100 105 110

gcg tat cag aat gga tat act tta tgg ggt aat gaa ttc aag gcc acg 384
 Ala Tyr Gln Asn Gly Tyr Thr Leu Trp Gly Asn Glu Phe Lys Ala Thr
 115 120 125

gaa act cag ctt gct ctc tac att tac att ttt tac gta agt aaa ata 432
 Glu Thr Gln Leu Ala Leu Tyr Ile Tyr Ile Phe Tyr Val Ser Lys Ile
 130 135 140

tac gag ttt gta gat act tac att atg ctt ctc aag aat aac ttg cgg 480
 Tyr Glu Phe Val Asp Thr Tyr Ile Met Leu Leu Lys Asn Asn Leu Arg
 145 150 155 160

caa gta agt ttc cta cac att tat cac cac agc acg att tcc ttt att 528
 Gln Val Ser Phe Leu His Ile Tyr His His Ser Thr Ile Ser Phe Ile
 165 170 175

tgg tgg atc att gct cgg agg gct ccg ggt ggt gat gct tac ttc agc 576
 Trp Trp Ile Ile Ala Arg Arg Ala Pro Gly Gly Asp Ala Tyr Phe Ser
 180 185 190

gcg gcc ttg aac tca tgg gta cac gtg tgc atg tac acc tat tat cta 624
 Ala Ala Leu Asn Ser Trp Val His Val Cys Met Tyr Thr Tyr Tyr Leu
 195 200 205

tta tca acc ctt att gga aaa gaa gat cct aag cgt tcc aac tac ctt 672
 Leu Ser Thr Leu Ile Gly Lys Glu Asp Pro Lys Arg Ser Asn Tyr Leu
 210 215 220

tgg tgg ggt cgc cac cta acg caa atg cag atg ctt cag ttt ttc ttc 720
 Trp Trp Gly Arg His Leu Thr Gln Met Gln Met Leu Gln Phe Phe Phe
 225 230 235 240

aac gta ctt caa gcg ttg tac tgc gct tcg ttc tct acg tat ccc aag 768
 Asn Val Leu Gln Ala Leu Tyr Cys Ala Ser Phe Ser Thr Tyr Pro Lys
 245 250 255

ttt ttg tcc aaa att ctg ctc gtc tat atg atg agc ctt ctc ggc ttg 816
Phe Leu Ser Lys Ile Leu Leu Val Tyr Met Met Ser Leu Leu Gly Leu
260 265 270

t t t g g g c a t t t c t a c t a t t c c a a g c a c a t a g c a g c a g c t a a g c t c c a g 864
Phe Gly His Phe Tyr Tyr Ser Lys His Ile Ala Ala Ala Lys Leu Gln
275 280 285

aaa aaa cag cag tga 879
Lys Lys Gln Gln
290

<210> 70

<211> 292

<212> PRT

<213> *Ostreococcus tauri*

<400> 70

Met Ser Gly Leu Arg Ala Pro Asn Phe Leu His Arg Phe Trp Thr Lys
1 5 10 15

Trp Asp Tyr Ala Ile Ser Lys Val Val Phe Thr Cys Ala Asp Ser Phe
20 25 30

Gln Trp Asp Ile Gly Pro Val Ser Ser Ser Thr Ala His Leu Pro Ala
35 40 45

Ile Glu Ser Pro Thr Pro Leu Val Thr Ser Leu Leu Phe Tyr Leu Val
50 55 60

Thr Val Phe Leu Trp Tyr Gly Arg Leu Thr Arg Ser Ser Asp Lys Lys
65 70 75 80

Ile Arg Glu Pro Thr Trp Leu Arg Arg Phe Ile Ile Cys His Asn Ala
85 90 95

Phe Leu Ile Val Leu Ser Leu Tyr Met Cys Leu Gly Cys Val Ala Gln
100 105 110

Ala Tyr Gln Asn Gly Tyr Thr Leu Trp Gly Asn Glu Phe Lys Ala Thr
115 120 125

Glu Thr Gln Leu Ala Leu Tyr Ile Tyr Ile Phe Tyr Val Ser Lys Ile
130 135 140

Tyr Glu Phe Val Asp Thr Tyr Ile Met Leu Leu Lys Asn Asn Leu Arg
145 150 155 160

Gln Val Ser Phe Leu His Ile Tyr His His Ser Thr Ile Ser Phe Ile
 165 170 175

Trp Trp Ile Ile Ala Arg Arg Ala Pro Gly Gly Asp Ala Tyr Phe Ser
 180 185 190

Ala Ala Leu Asn Ser Trp Val His Val Cys Met Tyr Thr Tyr Tyr Leu
 195 200 205

Leu Ser Thr Leu Ile Gly Lys Glu Asp Pro Lys Arg Ser Asn Tyr Leu
 210 215 220

Trp Trp Gly Arg His Leu Thr Gln Met Gln Met Leu Gln Phe Phe Phe
 225 230 235 240

Asn Val Leu Gln Ala Leu Tyr Cys Ala Ser Phe Ser Thr Tyr Pro Lys
 245 250 255

Phe Leu Ser Lys Ile Leu Leu Val Tyr Met Met Ser Leu Leu Gly Leu
 260 265 270

Phe Gly His Phe Tyr Tyr Ser Lys His Ile Ala Ala Ala Lys Leu Gln
 275 280 285

Lys Lys Gln Gln
 290

<210> 71

<211> 1362

<212> DNA

<213> *Primula farinosa*

<220>

<221> CDS

<222> (1)..(1362)

<223> Delta-6 desaturase

<400> 71

atg gct aac aaa tct cca cca aac ccc aaa aca ggt tac ata acc agc 48
 Met Ala Asn Lys Ser Pro Pro Asn Pro Lys Thr Gly Tyr Ile Thr Ser
 1 5 10 15

tca gac ctg aaa tcc cac aac aag gca ggt gac cta tgg ata tca atc 96

Ser	Asp	Leu	Lys	Ser	His	Asn	Lys	Ala	Gly	Asp	Leu	Trp	Ile	Ser	Ile	
		20						25					30			
cac	ggc	caa	gtc	tac	gac	gtg	tcc	tct	tgg	gcc	gcc	ctt	cat	ccg	ggg	144
His	Gly	Gln	Val	Tyr	Asp	Val	Ser	Ser	Trp	Ala	Ala	Leu	His	Pro	Gly	
		35					40					45				
ggc	act	gcc	cct	ctc	atg	gcc	ctt	gca	gga	cac	gac	gtg	acc	gat	gct	192
Gly	Thr	Ala	Pro	Leu	Met	Ala	Leu	Ala	Gly	His	Asp	Val	Thr	Asp	Ala	
	50					55					60					
ttc	ctc	gcg	tac	cat	ccc	cct	tcc	act	gcc	cgt	ctc	ctc	cct	cct	ctc	240
Phe	Leu	Ala	Tyr	His	Pro	Pro	Ser	Thr	Ala	Arg	Leu	Leu	Pro	Pro	Leu	
65					70					75					80	
tct	acc	aac	ctc	ctt	ctt	caa	aac	cac	tcc	gtc	tcc	ccc	acc	tcc	tca	288
Ser	Thr	Asn	Leu	Leu	Leu	Gln	Asn	His	Ser	Val	Ser	Pro	Thr	Ser	Ser	
			85						90					95		
gac	tac	cgc	aaa	ctc	ctc	gac	aac	ttc	cat	aaa	cat	ggc	ctt	ttc	cgc	336
Asp	Tyr	Arg	Lys	Leu	Leu	Asp	Asn	Phe	His	Lys	His	Gly	Leu	Phe	Arg	
			100					105					110			
gcc	agg	ggc	cac	act	gct	tac	gcc	acc	ttc	gtc	ttc	atg	ata	gcg	atg	384
Ala	Arg	Gly	His	Thr	Ala	Tyr	Ala	Thr	Phe	Val	Phe	Met	Ile	Ala	Met	
		115					120					125				
ttt	cta	atg	agc	gtg	act	gga	gtc	ctt	tgc	agc	gac	agt	gcg	tgg	gtc	432
Phe	Leu	Met	Ser	Val	Thr	Gly	Val	Leu	Cys	Ser	Asp	Ser	Ala	Trp	Val	
	130					135					140					
cat	ttg	gct	agc	ggc	gga	gca	atg	ggg	ttc	gcc	tgg	atc	caa	tgc	gga	480
His	Leu	Ala	Ser	Gly	Gly	Ala	Met	Gly	Phe	Ala	Trp	Ile	Gln	Cys	Gly	
145					150					155					160	
tgg	ata	ggt	cac	gac	tct	ggg	cat	tac	cgg	att	atg	tct	gac	agg	aaa	528
Trp	Ile	Gly	His	Asp	Ser	Gly	His	Tyr	Arg	Ile	Met	Ser	Asp	Arg	Lys	
			165						170					175		
tgg	aac	tgg	ttc	gcg	caa	atc	cta	agc	aca	aac	tgc	ctc	cag	ggg	att	576
Trp	Asn	Trp	Phe	Ala	Gln	Ile	Leu	Ser	Thr	Asn	Cys	Leu	Gln	Gly	Ile	
			180					185					190			
agt	atc	ggg	tgg	tgg	aag	tgg	aac	cat	aat	gcg	cac	cac	atc	gct	tgc	624
Ser	Ile	Gly	Trp	Trp	Lys	Trp	Asn	His	Asn	Ala	His	His	Ile	Ala	Cys	
		195					200					205				
aat	agc	ctg	gat	tac	gac	ccc	gac	ctc	cag	tat	atc	cct	ttg	ctc	gtc	672
Asn	Ser	Leu	Asp	Tyr	Asp	Pro	Asp	Leu	Gln	Tyr	Ile	Pro	Leu	Leu	Val	
	210					215					220					
gtc	tcc	ccc	aag	ttc	ttc	aac	tcc	ctt	act	tct	cgt	ttc	tac	gac	aag	720
Val	Ser	Pro	Lys	Phe	Phe	Asn	Ser	Leu	Thr	Ser	Arg	Phe	Tyr	Asp	Lys	
225					230					235					240	
aag	ctg	aac	ttc	gac	ggc	gtg	tcg	agg	ttt	ctg	ggt	tgc	tac	cag	cac	768
Lys	Leu	Asn	Phe	Asp	Gly	Val	Ser	Arg	Phe	Leu	Val	Cys	Tyr	Gln	His	
			245						250					255		
tgg	acg	ttt	tat	ccg	gtc	atg	tgt	gtc	gct	agg	ctg	aac	atg	ctc	gcg	816
Trp	Thr	Phe	Tyr	Pro	Val	Met	Cys	Val	Ala	Arg	Leu	Asn	Met	Leu	Ala	
			260					265					270			
cag	tca	ttt	ata	acg	ctt	ttc	tcg	agt	agg	gag	gtg	tgc	cat	agg	gcg	864

130

Gln Ser Phe Ile Thr Leu Phe Ser Ser Arg Glu Val Cys His Arg Ala	
275 280 285	
caa gag gtt ttc gga ctt gcc gtg ttt tgg gtt tgg ttt ccg ctt tta	912
Gln Glu Val Phe Gly Leu Ala Val Phe Trp Val Trp Phe Pro Leu Leu	
290 295 300	
ctt tct tgt tta cct aat tgg ggc gag agg att atg ttt ttg ctt gcg	960
Leu Ser Cys Leu Pro Asn Trp Gly Glu Arg Ile Met Phe Leu Leu Ala	
305 310 315 320	
agc tat tcc gtt acg ggg ata caa cac gtg cag ttc agc ttg aac cat	1008
Ser Tyr Ser Val Thr Gly Ile Gln His Val Gln Phe Ser Leu Asn His	
325 330 335	
ttt tct tcg gac gtc tat gtg ggc ccg cca gta ggt aat gac tgg ttc	1056
Phe Ser Ser Asp Val Tyr Val Gly Pro Pro Val Gly Asn Asp Trp Phe	
340 345 350	
aag aaa cag act gcc ggg aca ctt aac ata tcg tgc ccg gcg tgg atg	1104
Lys Lys Gln Thr Ala Gly Thr Leu Asn Ile Ser Cys Pro Ala Trp Met	
355 360 365	
gat tgg ttc cat ggc ggg tta cag ttt cag gtc gag cac cac ttg ttt	1152
Asp Trp Phe His Gly Gly Leu Gln Phe Gln Val Glu His His Leu Phe	
370 375 380	
ccg cgg atg cct agg ggt cag ttt agg aag att tct cct ttt gtg agg	1200
Pro Arg Met Pro Arg Gly Gln Phe Arg Lys Ile Ser Pro Phe Val Arg	
385 390 395 400	
gat ttg tgt aag aaa cac aac ttg cct tac aat atc gcg tct ttt act	1248
Asp Leu Cys Lys Lys His Asn Leu Pro Tyr Asn Ile Ala Ser Phe Thr	
405 410 415	
aaa gcg aat gtg ttt acg ctt aag acg ctg aga aat acg gcc att gag	1296
Lys Ala Asn Val Phe Thr Leu Lys Thr Leu Arg Asn Thr Ala Ile Glu	
420 425 430	
gct cgg gac ctc tct aat ccg ctc cca aag aat atg gtg tgg gaa gct	1344
Ala Arg Asp Leu Ser Asn Pro Leu Pro Lys Asn Met Val Trp Glu Ala	
435 440 445	
ctt aaa act ctc ggg tga	1362
Leu Lys Thr Leu Gly	
450	

<210> 72

<211> 453

<212> PRT

<213> Primula farinosa

<400> 72

Met Ala Asn Lys Ser Pro Pro Asn Pro Lys Thr Gly Tyr Ile Thr Ser
1 5 10 15

131

Ser Asp Leu Lys Ser His Asn Lys Ala Gly Asp Leu Trp Ile Ser Ile
 20 25 30
 His Gly Gln Val Tyr Asp Val Ser Ser Trp Ala Ala Leu His Pro Gly
 35 40 45
 Gly Thr Ala Pro Leu Met Ala Leu Ala Gly His Asp Val Thr Asp Ala
 50 55 60
 Phe Leu Ala Tyr His Pro Pro Ser Thr Ala Arg Leu Leu Pro Pro Leu
 65 70 75 80
 Ser Thr Asn Leu Leu Leu Gln Asn His Ser Val Ser Pro Thr Ser Ser
 85 90 95
 Asp Tyr Arg Lys Leu Leu Asp Asn Phe His Lys His Gly Leu Phe Arg
 100 105 110
 Ala Arg Gly His Thr Ala Tyr Ala Thr Phe Val Phe Met Ile Ala Met
 115 120 125
 Phe Leu Met Ser Val Thr Gly Val Leu Cys Ser Asp Ser Ala Trp Val
 130 135 140
 His Leu Ala Ser Gly Gly Ala Met Gly Phe Ala Trp Ile Gln Cys Gly
 145 150 155 160
 Trp Ile Gly His Asp Ser Gly His Tyr Arg Ile Met Ser Asp Arg Lys
 165 170 175
 Trp Asn Trp Phe Ala Gln Ile Leu Ser Thr Asn Cys Leu Gln Gly Ile
 180 185 190
 Ser Ile Gly Trp Trp Lys Trp Asn His Asn Ala His His Ile Ala Cys
 195 200 205
 Asn Ser Leu Asp Tyr Asp Pro Asp Leu Gln Tyr Ile Pro Leu Leu Val
 210 215 220
 Val Ser Pro Lys Phe Phe Asn Ser Leu Thr Ser Arg Phe Tyr Asp Lys
 225 230 235 240
 Lys Leu Asn Phe Asp Gly Val Ser Arg Phe Leu Val Cys Tyr Gln His
 245 250 255
 Trp Thr Phe Tyr Pro Val Met Cys Val Ala Arg Leu Asn Met Leu Ala
 260 265 270

132

Gln Ser Phe Ile Thr Leu Phe Ser Ser Arg Glu Val Cys His Arg Ala
 275 280 285

Gln Glu Val Phe Gly Leu Ala Val Phe Trp Val Trp Phe Pro Leu Leu
 290 295 300

Leu Ser Cys Leu Pro Asn Trp Gly Glu Arg Ile Met Phe Leu Leu Ala
 305 310 315 320

Ser Tyr Ser Val Thr Gly Ile Gln His Val Gln Phe Ser Leu Asn His
 325 330 335

Phe Ser Ser Asp Val Tyr Val Gly Pro Pro Val Gly Asn Asp Trp Phe
 340 345 350

Lys Lys Gln Thr Ala Gly Thr Leu Asn Ile Ser Cys Pro Ala Trp Met
 355 360 365

Asp Trp Phe His Gly Gly Leu Gln Phe Gln Val Glu His His Leu Phe
 370 375 380

Pro Arg Met Pro Arg Gly Gln Phe Arg Lys Ile Ser Pro Phe Val Arg
 385 390 395 400

Asp Leu Cys Lys Lys His Asn Leu Pro Tyr Asn Ile Ala Ser Phe Thr
 405 410 415

Lys Ala Asn Val Phe Thr Leu Lys Thr Leu Arg Asn Thr Ala Ile Glu
 420 425 430

Ala Arg Asp Leu Ser Asn Pro Leu Pro Lys Asn Met Val Trp Glu Ala
 435 440 445

Leu Lys Thr Leu Gly
 450

<210> 73

<211> 1362

<212> DNA

<213> *Primula vialii*

<220>

<221> CDS

<222> (1)..(1362)

<223> Delta-6 desaturase

<400> 73

atg gct aac aaa tct cca cca aac ccc aaa aca ggt tac att acc agc	48
Met Ala Asn Lys Ser Pro Pro Asn Pro Lys Thr Gly Tyr Ile Thr Ser	
1 5 10 15	
tca gac ctg aaa ggg cac aac aaa gca gga gac cta tgg ata tca atc	96
Ser Asp Leu Lys Gly His Asn Lys Ala Gly Asp Leu Trp Ile Ser Ile	
20 25 30	
cac ggg gag gta tac gac gtg tcc tcg tgg gcc ggc ctt cac ccg ggg	144
His Gly Glu Val Tyr Asp Val Ser Ser Trp Ala Gly Leu His Pro Gly	
35 40 45	
ggc agt gcc ccc ctc atg gcc ctc gca gga cac gac gta acc gac gct	192
Gly Ser Ala Pro Leu Met Ala Leu Ala Gly His Asp Val Thr Asp Ala	
50 55 60	
ttt cta gcg tat cat cct cct tct acc gcc cgc ctc ctc cct ccc ctc	240
Phe Leu Ala Tyr His Pro Pro Ser Thr Ala Arg Leu Leu Pro Pro Leu	
65 70 75 80	
tcc acc aac ctc ctc ctt caa aac cac tcc gtc tcc ccc acc tcc tct	288
Ser Thr Asn Leu Leu Leu Gln Asn His Ser Val Ser Pro Thr Ser Ser	
85 90 95	
gac tac cgc aaa ctc ctc cac aac ttc cat aaa att ggt atg ttc cgc	336
Asp Tyr Arg Lys Leu Leu His Asn Phe His Lys Ile Gly Met Phe Arg	
100 105 110	
gcc agg ggc cac act gct tac gcc acc ttc gtc atc atg ata gtg atg	384
Ala Arg Gly His Thr Ala Tyr Ala Thr Phe Val Ile Met Ile Val Met	
115 120 125	
ttt cta acg agc gtg acc gga gtc ctt tgc agc gac agt gcg tgg gtc	432
Phe Leu Thr Ser Val Thr Gly Val Leu Cys Ser Asp Ser Ala Trp Val	
130 135 140	
cat ctg gct agc ggc gca gca atg ggg ttc gcc tgg atc cag tgc gga	480
His Leu Ala Ser Gly Ala Ala Met Gly Phe Ala Trp Ile Gln Cys Gly	
145 150 155 160	
tgg ata ggt cac gac tct ggg cat tac cgg att atg tct gac agg aaa	528
Trp Ile Gly His Asp Ser Gly His Tyr Arg Ile Met Ser Asp Arg Lys	
165 170 175	
tgg aac tgg ttc gcg cag gtc ctg agc aca aac tgc ctc cag ggg atc	576
Trp Asn Trp Phe Ala Gln Val Leu Ser Thr Asn Cys Leu Gln Gly Ile	
180 185 190	
agt atc ggg tgg tgg aag tgg aac cat aac gcc cac cac att gct tgc	624
Ser Ile Gly Trp Trp Lys Trp Asn His Asn Ala His His Ile Ala Cys	
195 200 205	
aat agc ctg gac tac gac ccc gac ctc cag tat atc cct ttg ctc gtg	672
Asn Ser Leu Asp Tyr Asp Pro Asp Leu Gln Tyr Ile Pro Leu Leu Val	
210 215 220	
gtc tcc ccc aag ttc ttc aac tcc ctt act tct cgt ttc tac gac aag	720
Val Ser Pro Lys Phe Phe Asn Ser Leu Thr Ser Arg Phe Tyr Asp Lys	
225 230 235 240	

aag ctg aat ttc gac ggc gtg tca agg ttt ctg gtt tgc tac cag cac Lys Leu Asn Phe Asp Gly Val Ser Arg Phe Leu Val Cys Tyr Gln His 245 250 255	768
tgg acg ttt tat cca gtc atg tgt gtc gct agg cta aac atg atc gca Trp Thr Phe Tyr Pro Val Met Cys Val Ala Arg Leu Asn Met Ile Ala 260 265 270	816
cag tcg ttt ata acg ctt ttc tcg agc agg gag gtg ggt cat agg gcg Gln Ser Phe Ile Thr Leu Phe Ser Ser Arg Glu Val Gly His Arg Ala 275 280 285	864
caa gag att ttc gga ctt gct gtg ttt tgg gtt tgg ttt ccg ctc ctg Gln Glu Ile Phe Gly Leu Ala Val Phe Trp Val Trp Phe Pro Leu Leu 290 295 300	912
ctc tct tgc tta cct aat tgg agc gag agg att atg ttt ctg cta gcg Leu Ser Cys Leu Pro Asn Trp Ser Glu Arg Ile Met Phe Leu Leu Ala 305 310 315 320	960
agc tat tcc gtt acg ggg ata cag cac gtg cag ttc agc ttg aac cat Ser Tyr Ser Val Thr Gly Ile Gln His Val Gln Phe Ser Leu Asn His 325 330 335	1008
ttt tct tcg gac gtc tac gtg ggc ccg cca gta gct aac gac tgg ttc Phe Ser Ser Asp Val Tyr Val Gly Pro Pro Val Ala Asn Asp Trp Phe 340 345 350	1056
aag aaa cag act gct ggg aca ctt aac ata tcg tgc ccg gcg tgg atg Lys Lys Gln Thr Ala Gly Thr Leu Asn Ile Ser Cys Pro Ala Trp Met 355 360 365	1104
gac tgg ttc cat ggc ggg ttg cag ttt cag gtc gag cac cac ttg ttt Asp Trp Phe His Gly Gly Leu Gln Phe Gln Val Glu His His Leu Phe 370 375 380	1152
ccg cgg atg cct agg ggt cag ttt agg aag att tct cct ttt gtg agg Pro Arg Met Pro Arg Gly Gln Phe Arg Lys Ile Ser Pro Phe Val Arg 385 390 395 400	1200
gat ttg tgt aag aaa cac aac ttg cct tac aat atc gcg tct ttt act Asp Leu Cys Lys Lys His Asn Leu Pro Tyr Asn Ile Ala Ser Phe Thr 405 410 415	1248
aaa gca aac gtg ttg acg ctt aag acg ctg aga aat acg gcc att gag Lys Ala Asn Val Leu Thr Leu Lys Thr Leu Arg Asn Thr Ala Ile Glu 420 425 430	1296
gct cgg gac ctc tct aat ccg acc cca aag aat atg gtg tgg gaa gcc Ala Arg Asp Leu Ser Asn Pro Thr Pro Lys Asn Met Val Trp Glu Ala 435 440 445	1344
gtc cac aca cac ggc tag Val His Thr His Gly 450	1362

<210> 74

<211> 453

<212> PRT

<213> Primula vialii

<400> 74

Met Ala Asn Lys Ser Pro Pro Asn Pro Lys Thr Gly Tyr Ile Thr Ser
1 5 10 15

Ser Asp Leu Lys Gly His Asn Lys Ala Gly Asp Leu Trp Ile Ser Ile
20 25 30

His Gly Glu Val Tyr Asp Val Ser Ser Trp Ala Gly Leu His Pro Gly
35 40 45

Gly Ser Ala Pro Leu Met Ala Leu Ala Gly His Asp Val Thr Asp Ala
50 55 60

Phe Leu Ala Tyr His Pro Pro Ser Thr Ala Arg Leu Leu Pro Pro Leu
65 70 75 80

Ser Thr Asn Leu Leu Leu Gln Asn His Ser Val Ser Pro Thr Ser Ser
85 90 95

Asp Tyr Arg Lys Leu Leu His Asn Phe His Lys Ile Gly Met Phe Arg
100 105 110

Ala Arg Gly His Thr Ala Tyr Ala Thr Phe Val Ile Met Ile Val Met
115 120 125

Phe Leu Thr Ser Val Thr Gly Val Leu Cys Ser Asp Ser Ala Trp Val
130 135 140

His Leu Ala Ser Gly Ala Ala Met Gly Phe Ala Trp Ile Gln Cys Gly
145 150 155 160

Trp Ile Gly His Asp Ser Gly His Tyr Arg Ile Met Ser Asp Arg Lys
165 170 175

Trp Asn Trp Phe Ala Gln Val Leu Ser Thr Asn Cys Leu Gln Gly Ile
180 185 190

Ser Ile Gly Trp Trp Lys Trp Asn His Asn Ala His His Ile Ala Cys
195 200 205

Asn Ser Leu Asp Tyr Asp Pro Asp Leu Gln Tyr Ile Pro Leu Leu Val
210 215 220

Val Ser Pro Lys Phe Phe Asn Ser Leu Thr Ser Arg Phe Tyr Asp Lys
225 230 235 240

Lys Leu Asn Phe Asp Gly Val Ser Arg Phe Leu Val Cys Tyr Gln His
 245 250 255

Trp Thr Phe Tyr Pro Val Met Cys Val Ala Arg Leu Asn Met Ile Ala
 260 265 270

Gln Ser Phe Ile Thr Leu Phe Ser Ser Arg Glu Val Gly His Arg Ala
 275 280 285

Gln Glu Ile Phe Gly Leu Ala Val Phe Trp Val Trp Phe Pro Leu Leu
 290 295 300

Leu Ser Cys Leu Pro Asn Trp Ser Glu Arg Ile Met Phe Leu Leu Ala
 305 310 315 320

Ser Tyr Ser Val Thr Gly Ile Gln His Val Gln Phe Ser Leu Asn His
 325 330 335

Phe Ser Ser Asp Val Tyr Val Gly Pro Pro Val Ala Asn Asp Trp Phe
 340 345 350

Lys Lys Gln Thr Ala Gly Thr Leu Asn Ile Ser Cys Pro Ala Trp Met
 355 360 365

Asp Trp Phe His Gly Gly Leu Gln Phe Gln Val Glu His His Leu Phe
 370 375 380

Pro Arg Met Pro Arg Gly Gln Phe Arg Lys Ile Ser Pro Phe Val Arg
 385 390 395 400

Asp Leu Cys Lys Lys His Asn Leu Pro Tyr Asn Ile Ala Ser Phe Thr
 405 410 415

Lys Ala Asn Val Leu Thr Leu Lys Thr Leu Arg Asn Thr Ala Ile Glu
 420 425 430

Ala Arg Asp Leu Ser Asn Pro Thr Pro Lys Asn Met Val Trp Glu Ala
 435 440 445

Val His Thr His Gly
 450

<210> 75

<211> 903

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(903)

<223> Delta-5 elongase

<400> 75

atg	agc	gcc	tcc	ggt	gcg	ctg	ctg	ccc	gcg	atc	gcg	tcc	gcc	gcg	tac	48
Met	Ser	Ala	Ser	Gly	Ala	Leu	Leu	Pro	Ala	Ile	Ala	Ser	Ala	Ala	Tyr	
1				5					10					15		

gcg	tac	gcg	acg	tac	gcc	tac	gcc	ttt	gag	tgg	tcg	cac	gcg	aat	ggc	96
Ala	Tyr	Ala	Thr	Tyr	Ala	Tyr	Ala	Phe	Glu	Trp	Ser	His	Ala	Asn	Gly	
			20					25					30			

atc	gac	aac	gtc	gac	gcg	cgc	gag	tgg	atc	ggt	gcg	ctg	tcg	ttg	agg	144
Ile	Asp	Asn	Val	Asp	Ala	Arg	Glu	Trp	Ile	Gly	Ala	Leu	Ser	Leu	Arg	
		35				40						45				

ctc	ccg	gcg	atc	gcg	acg	acg	atg	tac	ctg	ttg	ttc	tgc	ctg	gtc	gga	192
Leu	Pro	Ala	Ile	Ala	Thr	Thr	Met	Tyr	Leu	Leu	Phe	Cys	Leu	Val	Gly	
	50					55					60					

ccg	agg	ttg	atg	gcg	aag	cgc	gag	gcg	ttc	gac	ccg	aag	ggg	ttc	atg	240
Pro	Arg	Leu	Met	Ala	Lys	Arg	Glu	Ala	Phe	Asp	Pro	Lys	Gly	Phe	Met	
65				70					75					80		

ctg	gcg	tac	aat	gcg	tat	cag	acg	gcg	ttc	aac	gtc	gtc	gtg	ctc	ggg	288
Leu	Ala	Tyr	Asn	Ala	Tyr	Gln	Thr	Ala	Phe	Asn	Val	Val	Val	Leu	Gly	
			85					90						95		

atg	ttc	gcg	cga	gag	atc	tcg	ggg	ctg	ggg	cag	ccc	gtg	tgg	ggg	tca	336
Met	Phe	Ala	Arg	Glu	Ile	Ser	Gly	Leu	Gly	Gln	Pro	Val	Trp	Gly	Ser	
			100					105					110			

acc	atg	ccg	tgg	agc	gat	aga	aaa	tcg	ttt	aag	atc	ctc	ctc	ggg	gtg	384
Thr	Met	Pro	Trp	Ser	Asp	Arg	Lys	Ser	Phe	Lys	Ile	Leu	Leu	Gly	Val	
		115					120					125				

tgg	ttg	cac	tac	aac	aac	aaa	tat	ttg	gag	cta	ttg	gac	act	gtg	ttc	432
Trp	Leu	His	Tyr	Asn	Asn	Lys	Tyr	Leu	Glu	Leu	Leu	Asp	Thr	Val	Phe	
	130					135					140					

atg	gtt	gcg	cgc	aag	aag	acg	aag	cag	ttg	agc	ttc	ttg	cac	gtt	tat	480
Met	Val	Ala	Arg	Lys	Lys	Thr	Lys	Gln	Leu	Ser	Phe	Leu	His	Val	Tyr	
145					150					155					160	

cat	cac	gcc	ctg	ttg	atc	tgg	gcg	tgg	tgg	ttg	gtg	tgt	cac	ttg	atg	528
His	His	Ala	Leu	Leu	Ile	Trp	Ala	Trp	Trp	Leu	Val	Cys	His	Leu	Met	
			165					170						175		

gcc	acg	aac	gat	tgt	atc	gat	gcc	tac	ttc	ggc	gcg	gcg	tgc	aac	tcg	576
Ala	Thr	Asn	Asp	Cys	Ile	Asp	Ala	Tyr	Phe	Gly	Ala	Ala	Cys	Asn	Ser	
			180					185					190			

ttc	att	cac	atc	gtg	atg	tac	tcg	tat	tat	ctc	atg	tcg	gcg	ctc	ggc	624
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

138

Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
 195 200 205
 att cga tgc ccg tgg aag cga tac atc acc cag gct caa atg ctc caa 672
 Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
 210 215 220
 ttc gtc att gtc ttc gcg cac gcc gtg ttc gtg ctg cgt cag aag cac 720
 Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240
 tgc ccg gtc acc ctt cct tgg gcg caa atg ttc gtc atg acg aac atg 768
 Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255
 ctc gtg ctc ttc ggg aac ttc tac ctc aag gcg tac tcg aac aag tcg 816
 Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270
 cgc ggc gac ggc gcg agt tcc gtg aaa cca gcc gag acc acg cgc gcg 864
 Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285
 ccc agc gtg cga cgc acg cga tct cga aaa att gac taa 903
 Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 76

<211> 300

<212> PRT

<213> *Ostreococcus tauri*

<400> 76

Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Ser Ala Ala Tyr
 1 5 10 15
 Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30
 Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45
 Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly
 50 55 60
 Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met
 65 70 75 80
 Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly
 85 90 95

139

Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser
100 105 110

Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val
115 120 125

Trp Leu His Tyr Asn Asn Lys Tyr Leu Glu Leu Leu Asp Thr Val Phe
130 135 140

Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr
145 150 155 160

His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met
165 170 175

Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser
180 185 190

Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
195 200 205

Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
210 215 220

Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
225 230 235 240

Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
245 250 255

Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
260 265 270

Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
275 280 285

Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
290 295 300

<210> 77

<211> 903

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(903)

<223> Delta-5 elongase

<400> 77

atg agc gcc tcc ggt gcg ctg ctg ccc gcg atc gcg ttc gcc gcg tac	48
Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Phe Ala Ala Tyr	
1 5 10 15	
gcg tac gcg acg tac gcc tac gcc ttt gag tgg tcg cac gcg aat ggc	96
Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly	
20 25 30	
atc gac aac gtc gac gcg cgc gag tgg atc ggt gcg ctg tcg ttg agg	144
Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg	
35 40 45	
ctc ccg gcg atc gcg acg acg atg tac ctg ttg ttc tgc ctg gtc gga	192
Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly	
50 55 60	
ccg agg ttg atg gcg aag cgc gag gcg ttc gac ccg aag ggg ttc atg	240
Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met	
65 70 75 80	
ctg gcg tac aat gcg tat cag acg gcg ttc aac gtc gtc gtg ctc ggg	288
Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly	
85 90 95	
atg ttc gcg cga gag atc tcg ggg ctg ggg cag ccc gtg tgg ggg tca	336
Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser	
100 105 110	
acc atg ccg tgg agc gat aga aaa tcg ttt aag atc ctc ctc ggg gtg	384
Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val	
115 120 125	
tgg ttg cac tac aac aac aaa tat ttg gag cta ttg gac act gtg ttc	432
Trp Leu His Tyr Asn Asn Lys Tyr Leu Glu Leu Leu Asp Thr Val Phe	
130 135 140	
atg gtt gcg cgc aag aag acg aag cag ttg agc ttc ttg cac gtt tat	480
Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr	
145 150 155 160	
cat cac gcc ctg ttg atc tgg gcg tgg tgg ttg gtg tgt cac ttg atg	528
His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met	
165 170 175	
gcc acg aac gat tgt atc gat gcc tac ttc ggc gcg gcg tgc aac tcg	576
Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser	
180 185 190	
ttc att cac atc gtg atg tac tcg tat tat ctc atg tcg gcg ctc ggc	624
Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly	
195 200 205	
att cga tgc ccg tgg aag cga tac atc acc cag gct caa atg ctc caa	672
Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln	
210 215 220	

ttc gtc att gtc ttc gcg cac gcc gtg ttc gtg ctg cgt cag aag cac 720
 Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240

tgc ccg gtc acc ctt cct tgg gcg caa atg ttc gtc atg acg aac atg 768
 Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255

ctc gtg ctc ttc ggg aac ttc tac ctc aag gcg tac tcg aac aag tcg 816
 Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270

cgc ggc gac ggc gcg agt tcc gtg aaa cca gcc gag acc acg cgc gcg 864
 Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285

ccc agc gtg cga cgc acg cga tct cga aaa att gac taa 903
 Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 78

<211> 300

<212> PRT

<213> *Ostreococcus tauri*

<400> 78

Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Phe Ala Ala Tyr
 1 5 10 15

Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30

Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45

Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly
 50 55 60

Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met
 65 70 75 80

Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly
 85 90 95

Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser
 100 105 110

Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val
 115 120 125

Trp Leu His Tyr Asn Asn Lys Tyr Leu Glu Leu Leu Asp Thr Val Phe
 130 135 140

Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr
 145 150 155 160

His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met
 165 170 175

Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser
 180 185 190

Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
 195 200 205

Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
 210 215 220

Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240

Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255

Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270

Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285

Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 79

<211> 903

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(903)

<223> Delta-5 elongase

<400> 79
 atg agc gcc tcc ggt gcg ctg ctg ccc gcg atc gcg tcc gcc gcg tac 48
 Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Ser Ala Ala Tyr
 1 5 10 15
 gcg tac gcg acg tac gcc tac gcc ttt gag tgg tcg cac gcg aat ggc 96
 Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30
 atc gac aac gtc gac gcg cgc gag tgg atc ggt gcg ctg tcg ttg agg 144
 Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45
 ctc ccg gcg atc gcg acg acg atg tac ctg ttg ttc tgc ctg gtc gga 192
 Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly
 50 55 60
 ccg agg ttg atg gcg aag cgc gag gcg ttc gac ccg aag ggg ttc atg 240
 Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met
 65 70 75 80
 ctg gcg tac aat gcg tat cag acg gcg ttc aac gtc gtc gtg ctc ggg 288
 Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly
 85 90 95
 atg ttc gcg cga gag atc tcg ggg ctg ggg cag ccc gtg tgg ggg tca 336
 Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser
 100 105 110
 acc atg ccg tgg agc gat aga aaa tcg ttt aag atc ctc ctc ggg gtg 384
 Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val
 115 120 125
 tgg ttg cac tac aac aac caa tat ttg gag cta ttg gac act gtg ttc 432
 Trp Leu His Tyr Asn Asn Gln Tyr Leu Glu Leu Leu Asp Thr Val Phe
 130 135 140
 atg gtt gcg cgc aag aag acg aag cag ttg agc ttc ttg cac gtt tat 480
 Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr
 145 150 155 160
 cat cac gcc ctg ttg atc tgg gcg tgg tgg ttg gtg tgt cac ttg atg 528
 His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met
 165 170 175
 gcc acg aac gat tgt atc gat gcc tac ttc ggc gcg gcg tgc aac tcg 576
 Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser
 180 185 190
 ttc att cac atc gtg atg tac tcg tat tat ctc atg tcg gcg ctc ggc 624
 Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
 195 200 205
 att cga tgc ccg tgg aag cga tac atc acc cag gct caa atg ctc caa 672
 Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
 210 215 220
 ttc gtc att gtc ttc gcg cac gcc gtg ttc gtg ctg cgt cag aag cac 720
 Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240
 tgc ccg gtc acc ctt cct tgg gcg caa atg ttc gtc atg acg aac atg 768

144

Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255
 ctc gtg ctc ttc ggg aac ttc tac ctc aag gcg tac tcg aac aag tcg 816
 Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270
 cgc ggc gac ggc gcg agt tcc gtg aaa cca gcc gag acc acg cgc gcg 864
 Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285
 ccc agc gtg cga cgc acg cga tct cga aaa att gac taa 903
 Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 80

<211> 300

<212> PRT

<213> *Ostreococcus tauri*

<400> 80

Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Ser Ala Ala Tyr
 1 5 10 15
 Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30
 Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45
 Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly
 50 55 60
 Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met
 65 70 75 80
 Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly
 85 90 95
 Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser
 100 105 110
 Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val
 115 120 125
 Trp Leu His Tyr Asn Asn Gln Tyr Leu Glu Leu Leu Asp Thr Val Phe
 130 135 140

145

Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr
 145 150 155 160

His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met
 165 170 175

Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser
 180 185 190

Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
 195 200 205

Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
 210 215 220

Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240

Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255

Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270

Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285

Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 81

<211> 879

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(879)

<223> Delta-6 elongase

<400> 81

atg agt ggc tta cgt gca ccc aac ttt tta cac aga ttc tgg aca aag
 Met Ser Gly Leu Arg Ala Pro Asn Phe Leu His Arg Phe Trp Thr Lys
 1 5 10 15

tgg gac tac gcg att tcc aaa gtc gtc ttc acg tgt gcc gac agt ttt	96
Trp Asp Tyr Ala Ile Ser Lys Val Val Phe Thr Cys Ala Asp Ser Phe	
20 25 30	
cag tgg gac atc ggg cca gtg agt tcg agt acg gcg cat tta ccc gcc	144
Gln Trp Asp Ile Gly Pro Val Ser Ser Ser Thr Ala His Leu Pro Ala	
35 40 45	
att gaa tcc cct acc cca ctg gtg act agc ctc ttg ttc tac tta gtc	192
Ile Glu Ser Pro Thr Pro Leu Val Thr Ser Leu Leu Phe Tyr Leu Val	
50 55 60	
aca gtt ttc ttg tgg tat ggt cgt tta acc agg agt tca gac aag aaa	240
Thr Val Phe Leu Trp Tyr Gly Arg Leu Thr Arg Ser Ser Asp Lys Lys	
65 70 75 80	
att aga gag cct acg tgg tta aga aga ttc ata ata tgt cat aat gcg	288
Ile Arg Glu Pro Thr Trp Leu Arg Arg Phe Ile Ile Cys His Asn Ala	
85 90 95	
ttc ttg ata gtc ctc agt ctt tac atg tgc ctt ggt tgt gtg gcc caa	336
Phe Leu Ile Val Leu Ser Leu Tyr Met Cys Leu Gly Cys Val Ala Gln	
100 105 110	
gcg tat cag aat gga tat act tta tgg ggt aat gaa ttc aag gcc acg	384
Ala Tyr Gln Asn Gly Tyr Thr Leu Trp Gly Asn Glu Phe Lys Ala Thr	
115 120 125	
gaa act cag ctt gct ctc tac att tac att ttt tac gta agt aaa ata	432
Glu Thr Gln Leu Ala Leu Tyr Ile Tyr Ile Phe Tyr Val Ser Lys Ile	
130 135 140	
tac gag ttt gta gat act tac att atg ctt ctc aag aat aac ttg cgg	480
Tyr Glu Phe Val Asp Thr Tyr Ile Met Leu Leu Lys Asn Asn Leu Arg	
145 150 155 160	
caa gta aga ttc cta cac act tat cac cac agc acg att tcc ttt att	528
Gln Val Arg Phe Leu His Thr Tyr His His Ser Thr Ile Ser Phe Ile	
165 170 175	
tgg tgg atc att gct cgg agg gct ccg ggt ggt gat gct tac ttc agc	576
Trp Trp Ile Ile Ala Arg Arg Ala Pro Gly Gly Asp Ala Tyr Phe Ser	
180 185 190	
gcg gcc ttg aac tca tgg gta cac gtg tgc atg tac acc tat tat cta	624
Ala Ala Leu Asn Ser Trp Val His Val Cys Met Tyr Thr Tyr Tyr Leu	
195 200 205	
tta tca acc ctt att gga aaa gaa gat cct aag cgt tcc aac tac ctt	672
Leu Ser Thr Leu Ile Gly Lys Glu Asp Pro Lys Arg Ser Asn Tyr Leu	
210 215 220	
tgg tgg ggt cgc cac cta acg caa atg cag atg ctt cag ttt ttc ttc	720
Trp Trp Gly Arg His Leu Thr Gln Met Gln Met Leu Gln Phe Phe Phe	
225 230 235 240	
aac gta ctt caa gcg ttg tac tgc gct tcg ttc tct acg tat ccc aag	768
Asn Val Leu Gln Ala Leu Tyr Cys Ala Ser Phe Ser Thr Tyr Pro Lys	
245 250 255	
ttt ttg tcc aaa att ctg ctc gtc tat atg atg agc ctt ctc ggc ttg	816
Phe Leu Ser Lys Ile Leu Leu Val Tyr Met Met Ser Leu Leu Gly Leu	
260 265 270	

aaa aaa cag cag tga 879
Lys Lys Gln Gln
290

<213> *Ostreococcus tauri*

Gln Val Arg Phe Leu His Thr Tyr His His Ser Thr Ile Ser Phe Ile
165 170 175

Trp Trp Ile Ile Ala Arg Arg Ala Pro Gly Gly Asp Ala Tyr Phe Ser
 180 185 190

Ala Ala Leu Asn Ser Trp Val His Val Cys Met Tyr Thr Tyr Tyr Leu
 195 200 205

Leu Ser Thr Leu Ile Gly Lys Glu Asp Pro Lys Arg Ser Asn Tyr Leu
 210 215 220

Trp Trp Gly Arg His Leu Thr Gln Met Gln Met Leu Gln Phe Phe Phe
 225 230 235 240

Asn Val Leu Gln Ala Leu Tyr Cys Ala Ser Phe Ser Thr Tyr Pro Lys
 245 250 255

Phe Leu Ser Lys Ile Leu Leu Val Tyr Met Met Ser Leu Leu Gly Leu
 260 265 270

Phe Gly His Phe Tyr Tyr Ser Lys His Ile Ala Ala Ala Lys Leu Gln
 275 280 285

Lys Lys Gln Gln
 290

<210> 83

<211> 831

<212> DNA

<213> Thraustochytrium sp.

<220>

<221> CDS

<222> (1)..(831)

<223> Delta-5 elongase

<400> 83

atg gac gtc gtc gag cag caa tgg cgc cgc ttc gtg gac gcc gtg gac 48
 Met Asp Val Val Glu Gln Gln Trp Arg Arg Phe Val Asp Ala Val Asp
 1 5 10 15

aac gga atc gtg gag ttc atg gag cat gag aag ccc aac aag ctg aac 96
 Asn Gly Ile Val Glu Phe Met Glu His Glu Lys Pro Asn Lys Leu Asn
 20 25 30

gag ggc aag ctc ttc acc tcg acc gag gag atg atg gcg ctt atc gtc 144

Glu Gly Lys Leu Phe Thr Ser Thr Glu Glu Met Met Ala Leu Ile Val	
35 40 45	
ggc tac ctg gcg ttc gtg gtc ctc ggg tcc gcc ttc atg aag gcc ttt	192
Gly Tyr Leu Ala Phe Val Val Leu Gly Ser Ala Phe Met Lys Ala Phe	
50 55 60	
gtc gat aag cct ttc gag ctc aag ttc ctc aag ctc gtg cac aac atc	240
Val Asp Lys Pro Phe Glu Leu Lys Phe Leu Lys Leu Val His Asn Ile	
65 70 75 80	
ttc ctc acc ggt ctg tcc atg tac atg gcc acc gag tgc gcg cgc cag	288
Phe Leu Thr Gly Leu Ser Met Tyr Met Ala Thr Glu Cys Ala Arg Gln	
85 90 95	
gca tac ctc ggc ggc tac aag ctc ttt ggc aac ccg atg gag aag ggc	336
Ala Tyr Leu Gly Gly Tyr Lys Leu Phe Gly Asn Pro Met Glu Lys Gly	
100 105 110	
acc gag tcg cac gcc ccg ggc atg gcc aac atc atc tac atc ttc tac	384
Thr Glu Ser His Ala Pro Gly Met Ala Asn Ile Ile Tyr Ile Phe Tyr	
115 120 125	
gtg agc aag ttc ctc gaa ttc ctc gac acc gtc ttc atg atc ctc ggc	432
Val Ser Lys Phe Leu Glu Phe Leu Asp Thr Val Phe Met Ile Leu Gly	
130 135 140	
aag aag tgg aag cag ctc agc ttt ctc cac gtc tac cac cac gcg agc	480
Lys Lys Trp Lys Gln Leu Ser Phe Leu His Val Tyr His His Ala Ser	
145 150 155 160	
atc agc ttc atc tgg ggc atc atc gcc cgc ttc gcg ccc ggt ggc gac	528
Ile Ser Phe Ile Trp Gly Ile Ile Ala Arg Phe Ala Pro Gly Gly Asp	
165 170 175	
gcc tac ttc tct acc atc ctc aac agc agc gtg cat gtc gtg ctc tac	576
Ala Tyr Phe Ser Thr Ile Leu Asn Ser Ser Val His Val Val Leu Tyr	
180 185 190	
ggc tac tac gcc tcg acc acc ctc ggc tac acc ttc atg cgc ccg ctg	624
Gly Tyr Tyr Ala Ser Thr Thr Leu Gly Tyr Thr Phe Met Arg Pro Leu	
195 200 205	
cgc ccg tac att acc acc att cag ctc acg cag ttc atg gcc atg gtc	672
Arg Pro Tyr Ile Thr Thr Ile Gln Leu Thr Gln Phe Met Ala Met Val	
210 215 220	
gtc cag tcc gtc tat gac tac tac aac ccc tgc gac tac ccg cag ccc	720
Val Gln Ser Val Tyr Asp Tyr Tyr Asn Pro Cys Asp Tyr Pro Gln Pro	
225 230 235 240	
ctc gtc aag ctg ctc ttc tgg tac atg ctc acc atg ctc ggc ctc ttc	768
Leu Val Lys Leu Leu Phe Trp Tyr Met Leu Thr Met Leu Gly Leu Phe	
245 250 255	
ggc aac ttc ttc gtg cag cag tac ctc aag ccc aag gcg ccc aag aag	816
Gly Asn Phe Phe Val Gln Gln Tyr Leu Lys Pro Lys Ala Pro Lys Lys	
260 265 270	
cag aag acc atc taa	831
Gln Lys Thr Ile	
275	

<210> 84

<211> 276

<212> PRT

<213> Thraustochytrium sp.

<400> 84

Met Asp Val Val Glu Gln Gln Trp Arg Arg Phe Val Asp Ala Val Asp
 1 5 10 15

Asn Gly Ile Val Glu Phe Met Glu His Glu Lys Pro Asn Lys Leu Asn
 20 25 30

Glu Gly Lys Leu Phe Thr Ser Thr Glu Glu Met Met Ala Leu Ile Val
 35 40 45

Gly Tyr Leu Ala Phe Val Val Leu Gly Ser Ala Phe Met Lys Ala Phe
 50 55 60

Val Asp Lys Pro Phe Glu Leu Lys Phe Leu Lys Leu Val His Asn Ile
 65 70 75 80

Phe Leu Thr Gly Leu Ser Met Tyr Met Ala Thr Glu Cys Ala Arg Gln
 85 90 95

Ala Tyr Leu Gly Gly Tyr Lys Leu Phe Gly Asn Pro Met Glu Lys Gly
 100 105 110

Thr Glu Ser His Ala Pro Gly Met Ala Asn Ile Ile Tyr Ile Phe Tyr
 115 120 125

Val Ser Lys Phe Leu Glu Phe Leu Asp Thr Val Phe Met Ile Leu Gly
 130 135 140

Lys Lys Trp Lys Gln Leu Ser Phe Leu His Val Tyr His His Ala Ser
 145 150 155 160

Ile Ser Phe Ile Trp Gly Ile Ile Ala Arg Phe Ala Pro Gly Gly Asp
 165 170 175

Ala Tyr Phe Ser Thr Ile Leu Asn Ser Ser Val His Val Val Leu Tyr
 180 185 190

Gly Tyr Tyr Ala Ser Thr Thr Leu Gly Tyr Thr Phe Met Arg Pro Leu
 195 200 205

151

Arg Pro Tyr Ile Thr Thr Ile Gln Leu Thr Gln Phe Met Ala Met Val
 210 215 220

Val Gln Ser Val Tyr Asp Tyr Tyr Asn Pro Cys Asp Tyr Pro Gln Pro
 225 230 235 240

Leu Val Lys Leu Leu Phe Trp Tyr Met Leu Thr Met Leu Gly Leu Phe
 245 250 255

Gly Asn Phe Phe Val Gln Gln Tyr Leu Lys Pro Lys Ala Pro Lys Lys
 260 265 270

Gln Lys Thr Ile
 275

<210> 85

<211> 1077

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(1077)

<223> Delta-5 elongase

<400> 85

atg tgc tca cca ccg ccg tca caa tcc aaa aca aca tcc ctc cta gca 48
 Met Cys Ser Pro Pro Pro Ser Gln Ser Lys Thr Thr Ser Leu Leu Ala
 1 5 10 15

cgg tac acc acc gcc gcc ctc ctc ctc ctc acc ctc aca acg tgg tgc 96
 Arg Tyr Thr Thr Ala Ala Leu Leu Leu Leu Thr Leu Thr Thr Trp Cys
 20 25 30

cac ttc gcc ttc cca gcc gcc acc gcc aca ccc ggc ctc acc gcc gaa 144
 His Phe Ala Phe Pro Ala Ala Thr Ala Thr Pro Gly Leu Thr Ala Glu
 35 40 45

atg cac tcc tac aaa gtc cca ctc ggt ctc acc gta ttc tac ctg ctg 192
 Met His Ser Tyr Lys Val Pro Leu Gly Leu Thr Val Phe Tyr Leu Leu
 50 55 60

agt cta ccg tca cta aag tac gtt acg gac aac tac ctt gcc aaa aag 240
 Ser Leu Pro Ser Leu Lys Tyr Val Thr Asp Asn Tyr Leu Ala Lys Lys
 65 70 75 80

tat gat atg aag tca ctc ctg acg gaa tca atg gtg ttg tac aat gtg 288
 Tyr Asp Met Lys Ser Leu Leu Thr Glu Ser Met Val Leu Tyr Asn Val
 85 90 95

gcg caa gtg ctg ctc aat ggg tgg acg gtg tat gcg att gtg gat gcg Ala Gln Val Leu Leu Asn Gly Trp Thr Val Tyr Ala Ile Val Asp Ala 100 105 110	336
gtg atg aat aga gac cat cct ttt att gga agt aga agt ttg gtt ggg Val Met Asn Arg Asp His Pro Phe Ile Gly Ser Arg Ser Leu Val Gly 115 120 125	384
gcg gcg ttg cat agt ggg agc tcg tat gcg gtg tgg gtt cat tat tgt Ala Ala Leu His Ser Gly Ser Ser Tyr Ala Val Trp Val His Tyr Cys 130 135 140	432
gat aag tat ttg gag ttc ttt gat acg tat ttt atg gtg ttg agg ggg Asp Lys Tyr Leu Glu Phe Phe Asp Thr Tyr Phe Met Val Leu Arg Gly 145 150 155 160	480
aaa atg gac cag gtc tcc ttc ctc cac atc tac cac cac acg acc ata Lys Met Asp Gln Val Ser Phe Leu His Ile Tyr His His Thr Thr Ile 165 170 175	528
gcg tgg gca tgg tgg atc gcc ctc cgc ttc tcc ccc ggc gga gac att Ala Trp Ala Trp Trp Ile Ala Leu Arg Phe Ser Pro Gly Gly Asp Ile 180 185 190	576
tac ttc ggg gca ctc ctc aac tcc atc atc cac gtc ctc atg tat tcc Tyr Phe Gly Ala Leu Leu Asn Ser Ile Ile His Val Leu Met Tyr Ser 195 200 205	624
tac tac gcc ctt gcc cta ctc aag gtc agt tgt cca tgg aaa cga tac Tyr Tyr Ala Leu Ala Leu Leu Lys Val Ser Cys Pro Trp Lys Arg Tyr 210 215 220	672
ttg act caa gct caa tta ttg caa ttc aca agt gtg gtg gtt tat acg Leu Thr Gln Ala Gln Leu Leu Gln Phe Thr Ser Val Val Val Tyr Thr 225 230 235 240	720
ggg tgt acg ggt tat act cat tac tat cat acg aag cat gga gcg gat Gly Cys Thr Gly Tyr Thr His Tyr Tyr His Thr Lys His Gly Ala Asp 245 250 255	768
gag aca cag cct agt tta gga acg tat tat ttc tgt tgt gga gtg cag Glu Thr Gln Pro Ser Leu Gly Thr Tyr Tyr Phe Cys Cys Gly Val Gln 260 265 270	816
gtg ttt gag atg gtt agt ttg ttt gta ctc ttt tcc atc ttt tat aaa Val Phe Glu Met Val Ser Leu Phe Val Leu Phe Ser Ile Phe Tyr Lys 275 280 285	864
cga tcc tat tcg aag aag aac aag tca gga gga aag gat agc aag aag Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp Ser Lys Lys 290 295 300	912
aat gat gat ggg aat aat gag gat caa tgt cac aag gct atg aag gat Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala Met Lys Asp 305 310 315 320	960
ata tcg gag ggt gcg aag gag gtt gtg ggg cat gca gcg aag gat gct Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala Lys Asp Ala 325 330 335	1008
gga aag ttg gtg gct acg gcg agt aag gct gta aag agg aag gga act Gly Lys Leu Val Ala Thr Ala Ser Lys Ala Val Lys Arg Lys Gly Thr 340 345 350	1056

cgt gtt act ggt gcc atg tag
 Arg Val Thr Gly Ala Met
 355

1077

<210> 86

<211> 358

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 86

Met Cys Ser Pro Pro Pro Ser Gln Ser Lys Thr Thr Ser Leu Leu Ala
 1 5 10 15

Arg Tyr Thr Thr Ala Ala Leu Leu Leu Leu Thr Leu Thr Thr Trp Cys
 20 25 30

His Phe Ala Phe Pro Ala Ala Thr Ala Thr Pro Gly Leu Thr Ala Glu
 35 40 45

Met His Ser Tyr Lys Val Pro Leu Gly Leu Thr Val Phe Tyr Leu Leu
 50 55 60

Ser Leu Pro Ser Leu Lys Tyr Val Thr Asp Asn Tyr Leu Ala Lys Lys
 65 70 75 80

Tyr Asp Met Lys Ser Leu Leu Thr Glu Ser Met Val Leu Tyr Asn Val
 85 90 95

Ala Gln Val Leu Leu Asn Gly Trp Thr Val Tyr Ala Ile Val Asp Ala
 100 105 110

Val Met Asn Arg Asp His Pro Phe Ile Gly Ser Arg Ser Leu Val Gly
 115 120 125

Ala Ala Leu His Ser Gly Ser Ser Tyr Ala Val Trp Val His Tyr Cys
 130 135 140

Asp Lys Tyr Leu Glu Phe Phe Asp Thr Tyr Phe Met Val Leu Arg Gly
 145 150 155 160

Lys Met Asp Gln Val Ser Phe Leu His Ile Tyr His His Thr Thr Ile
 165 170 175

Ala Trp Ala Trp Trp Ile Ala Leu Arg Phe Ser Pro Gly Gly Asp Ile
 180 185 190

Tyr Phe Gly Ala Leu Leu Asn Ser Ile Ile His Val Leu Met Tyr Ser
 195 200 205

Tyr Tyr Ala Leu Ala Leu Leu Lys Val Ser Cys Pro Trp Lys Arg Tyr
 210 215 220

Leu Thr Gln Ala Gln Leu Leu Gln Phe Thr Ser Val Val Val Tyr Thr
 225 230 235 240

Gly Cys Thr Gly Tyr Thr His Tyr Tyr His Thr Lys His Gly Ala Asp
 245 250 255

Glu Thr Gln Pro Ser Leu Gly Thr Tyr Tyr Phe Cys Cys Gly Val Gln
 260 265 270

Val Phe Glu Met Val Ser Leu Phe Val Leu Phe Ser Ile Phe Tyr Lys
 275 280 285

Arg Ser Tyr Ser Lys Lys Asn Lys Ser Gly Gly Lys Asp Ser Lys Lys
 290 295 300

Asn Asp Asp Gly Asn Asn Glu Asp Gln Cys His Lys Ala Met Lys Asp
 305 310 315 320

Ile Ser Glu Gly Ala Lys Glu Val Val Gly His Ala Ala Lys Asp Ala
 325 330 335

Gly Lys Leu Val Ala Thr Ala Ser Lys Ala Val Lys Arg Lys Gly Thr
 340 345 350

Arg Val Thr Gly Ala Met
 355

<210> 87

<211> 1086

<212> DNA

<213> Phytophthora infestans

<220>

<221> CDS

<222> (1)..(1086)

<223> Omega-3 desaturase

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<400> 87
atg gcg acg aag gag gcg tat gtg ttc ccc act ctg acg gag atc aag      48
Met Ala Thr Lys Glu Ala Tyr Val Phe Pro Thr Leu Thr Glu Ile Lys
1          5          10          15

cgg tcg cta cct aaa gac tgt ttc gag gct tcg gtg cct ctg tcg ctc      96
Arg Ser Leu Pro Lys Asp Cys Phe Glu Ala Ser Val Pro Leu Ser Leu
          20          25          30

tac tac acc gtg cgt tgt ctg gtg atc gcg gtg gct cta acc ttc ggt      144
Tyr Tyr Thr Val Arg Cys Leu Val Ile Ala Val Ala Leu Thr Phe Gly
          35          40          45

ctc aac tac gct cgc gct ctg ccc gag gtc gag agc ttc tgg gct ctg      192
Leu Asn Tyr Ala Arg Ala Leu Pro Glu Val Glu Ser Phe Trp Ala Leu
          50          55          60

gac gcc gca ctc tgc acg ggc tac atc ttg ctg cag ggc atc gtg ttc      240
Asp Ala Ala Leu Cys Thr Gly Tyr Ile Leu Leu Gln Gly Ile Val Phe
65          70          75          80

tgg ggc ttc ttc acg gtg ggc cac gat gcc ggc cac ggc gcc ttc tcg      288
Trp Gly Phe Phe Thr Val Gly His Asp Ala Gly His Gly Ala Phe Ser
          85          90          95

cgc tac cac ctg ctt aac ttc gtg gtg ggc act ttc atg cac tcg ctc      336
Arg Tyr His Leu Leu Asn Phe Val Val Gly Thr Phe Met His Ser Leu
          100          105          110

atc ctc acg ccc ttc gag tcg tgg aag ctc acg cac cgt cac cac cac      384
Ile Leu Thr Pro Phe Glu Ser Trp Lys Leu Thr His Arg His His His
          115          120          125

aag aac acg ggc aac att gac cgt gac gag gtc ttc tac ccg caa cgc      432
Lys Asn Thr Gly Asn Ile Asp Arg Asp Glu Val Phe Tyr Pro Gln Arg
          130          135          140

aag gcc gac gac cac ccg ctg tct cgc aac ctg att ctg gcg ctc ggg      480
Lys Ala Asp Asp His Pro Leu Ser Arg Asn Leu Ile Leu Ala Leu Gly
145          150          155          160

gca gcg tgg ctc gcc tat ttg gtc gag ggc ttc cct cct cgt aag gtc      528
Ala Ala Trp Leu Ala Tyr Leu Val Glu Gly Phe Pro Pro Arg Lys Val
          165          170          175

aac cac ttc aac ccg ttc gag cct ctg ttc gtg cgt cag gtg tca gct      576
Asn His Phe Asn Pro Phe Glu Pro Leu Phe Val Arg Gln Val Ser Ala
          180          185          190

gtg gta atc tct ctt ctc gcc cac ttc ttc gtg gcc gga ctc tcc atc      624
Val Val Ile Ser Leu Leu Ala His Phe Phe Val Ala Gly Leu Ser Ile
          195          200          205

tat ctg agc ctc cag ctg ggc ctt aag acg atg gca atc tac tac tat      672
Tyr Leu Ser Leu Gln Leu Gly Leu Lys Thr Met Ala Ile Tyr Tyr Tyr
          210          215          220

gga cct gtt ttt gtg ttc ggc agc atg ctg gtc att acc acc ttc cta      720
Gly Pro Val Phe Val Phe Gly Ser Met Leu Val Ile Thr Thr Phe Leu
225          230          235          240

cac cac aat gat gag gag acc cca tgg tac gcc gac tcg gag tgg acg      768

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156

His His Asn Asp Glu Glu Thr Pro Trp Tyr Ala Asp Ser Glu Trp Thr
 245 250 255
 tac gtc aag ggc aac ctc tcg tcc gtg gac cga tcg tac ggc gcg ctc 816
 Tyr Val Lys Gly Asn Leu Ser Ser Val Asp Arg Ser Tyr Gly Ala Leu
 260 265 270
 att gac aac ctg agc cac aac atc ggc acg cac cag atc cac cac ctt 864
 Ile Asp Asn Leu Ser His Asn Ile Gly Thr His Gln Ile His His Leu
 275 280 285
 ttc cct atc att ccg cac tac aaa ctc aag aaa gcc act gcg gcc ttc 912
 Phe Pro Ile Ile Pro His Tyr Lys Leu Lys Lys Ala Thr Ala Ala Phe
 290 295 300
 cac cag gct ttc cct gag ctc gtg cgc aag agc gac gag cca att atc 960
 His Gln Ala Phe Pro Glu Leu Val Arg Lys Ser Asp Glu Pro Ile Ile
 305 310 315 320
 aag gct ttc ttc cgg gtt gga cgt ctc tac gca aac tac ggc gtt gtg 1008
 Lys Ala Phe Phe Arg Val Gly Arg Leu Tyr Ala Asn Tyr Gly Val Val
 325 330 335
 gac cag gag gcg aag ctc ttc acg cta aag gaa gcc aag gcg gcg acc 1056
 Asp Gln Glu Ala Lys Leu Phe Thr Leu Lys Glu Ala Lys Ala Ala Thr
 340 345 350
 gag gcg gcg gcc aag acc aag tcc acg taa 1086
 Glu Ala Ala Ala Lys Thr Lys Ser Thr
 355 360

<210> 88

<211> 361

<212> PRT

<213> *Phytophthora infestans*

<400> 88

Met Ala Thr Lys Glu Ala Tyr Val Phe Pro Thr Leu Thr Glu Ile Lys
 1 5 10 15
 Arg Ser Leu Pro Lys Asp Cys Phe Glu Ala Ser Val Pro Leu Ser Leu
 20 25 30
 Tyr Tyr Thr Val Arg Cys Leu Val Ile Ala Val Ala Leu Thr Phe Gly
 35 40 45
 Leu Asn Tyr Ala Arg Ala Leu Pro Glu Val Glu Ser Phe Trp Ala Leu
 50 55 60
 Asp Ala Ala Leu Cys Thr Gly Tyr Ile Leu Leu Gln Gly Ile Val Phe
 65 70 75 80

157

Trp Gly Phe Phe Thr Val Gly His Asp Ala Gly His Gly Ala Phe Ser
85 90 95

Arg Tyr His Leu Leu Asn Phe Val Val Gly Thr Phe Met His Ser Leu
100 105 110

Ile Leu Thr Pro Phe Glu Ser Trp Lys Leu Thr His Arg His His His
115 120 125

Lys Asn Thr Gly Asn Ile Asp Arg Asp Glu Val Phe Tyr Pro Gln Arg
130 135 140

Lys Ala Asp Asp His Pro Leu Ser Arg Asn Leu Ile Leu Ala Leu Gly
145 150 155 160

Ala Ala Trp Leu Ala Tyr Leu Val Glu Gly Phe Pro Pro Arg Lys Val
165 170 175

Asn His Phe Asn Pro Phe Glu Pro Leu Phe Val Arg Gln Val Ser Ala
180 185 190

Val Val Ile Ser Leu Leu Ala His Phe Phe Val Ala Gly Leu Ser Ile
195 200 205

Tyr Leu Ser Leu Gln Leu Gly Leu Lys Thr Met Ala Ile Tyr Tyr Tyr
210 215 220

Gly Pro Val Phe Val Phe Gly Ser Met Leu Val Ile Thr Thr Phe Leu
225 230 235 240

His His Asn Asp Glu Glu Thr Pro Trp Tyr Ala Asp Ser Glu Trp Thr
245 250 255

Tyr Val Lys Gly Asn Leu Ser Ser Val Asp Arg Ser Tyr Gly Ala Leu
260 265 270

Ile Asp Asn Leu Ser His Asn Ile Gly Thr His Gln Ile His His Leu
275 280 285

Phe Pro Ile Ile Pro His Tyr Lys Leu Lys Lys Ala Thr Ala Ala Phe
290 295 300

His Gln Ala Phe Pro Glu Leu Val Arg Lys Ser Asp Glu Pro Ile Ile
305 310 315 320

Lys Ala Phe Phe Arg Val Gly Arg Leu Tyr Ala Asn Tyr Gly Val Val
325 330 335

Asp Gln Glu Ala Lys Leu Phe Thr Leu Lys Glu Ala Lys Ala Ala Thr
 340 345 350

Glu Ala Ala Ala Lys Thr Lys Ser Thr
 355 360

<210> 89

<211> 1371

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(1371)

<223> Delta-6 desaturase

<400> 89

atg tgc gtg gag acg gaa aat aac gat ggg atc ccc acg gtg gag atc 48
 Met Cys Val Glu Thr Glu Asn Asn Asp Gly Ile Pro Thr Val Glu Ile
 1 5 10 15

gcg ttc gac ggt gag cgc gag cgg gcg gag gca aac gtg aag ctg tcc 96
 Ala Phe Asp Gly Glu Arg Glu Arg Ala Glu Ala Asn Val Lys Leu Ser
 20 25 30

gcg gag aag atg gag ccg gcg gcg ctg gcg aag acg ttc gcg agg cgg 144
 Ala Glu Lys Met Glu Pro Ala Ala Leu Ala Lys Thr Phe Ala Arg Arg
 35 40 45

tac gtc gtg atc gag ggg gtg gag tac gat gtg acg gat ttt aag cac 192
 Tyr Val Val Ile Glu Gly Val Glu Tyr Asp Val Thr Asp Phe Lys His
 50 55 60

ccg gga gga acg gtt att ttc tat gcg ttg tca aac acc ggg gcg gac 240
 Pro Gly Gly Thr Val Ile Phe Tyr Ala Leu Ser Asn Thr Gly Ala Asp
 65 70 75 80

gcg acg gaa gcg ttc aag gag ttt cat cat cgg tcg aga aag gcg agg 288
 Ala Thr Glu Ala Phe Lys Glu Phe His His Arg Ser Arg Lys Ala Arg
 85 90 95

aaa gcc ttg gcg gcg ctc ccg tct cga ccg gcc aag acg gcc aag gtg 336
 Lys Ala Leu Ala Ala Leu Pro Ser Arg Pro Ala Lys Thr Ala Lys Val
 100 105 110

gac gac gcg gag atg ctc caa gat ttc gcc aag tgg cgg aaa gaa ttg 384
 Asp Asp Ala Glu Met Leu Gln Asp Phe Ala Lys Trp Arg Lys Glu Leu
 115 120 125

gag aga gat gga ttc ttc aag ccc tct ccg gcg cac gtg gcg tat cgc 432
 Glu Arg Asp Gly Phe Phe Lys Pro Ser Pro Ala His Val Ala Tyr Arg
 130 135 140

ttc gcc gag ctc gcg gcg atg tac gct ctc ggg acg tac ctg atg tac	480
Phe Ala Glu Leu Ala Ala Met Tyr Ala Leu Gly Thr Tyr Leu Met Tyr	
145 150 155 160	
gct cga tac gtc gtc tcc tcg gtg ctc gtg tac gct tgc ttt ttc ggc	528
Ala Arg Tyr Val Val Ser Ser Val Leu Val Tyr Ala Cys Phe Phe Gly	
165 170 175	
gcc cga tgc ggt tgg gtg cag cac gag ggc gga cac agc tcg ctg acg	576
Ala Arg Cys Gly Trp Val Gln His Glu Gly Gly His Ser Ser Leu Thr	
180 185 190	
ggc aac att tgg tgg gac aag cgc atc cag gcc ttc aca gcc ggg ttc	624
Gly Asn Ile Trp Trp Asp Lys Arg Ile Gln Ala Phe Thr Ala Gly Phe	
195 200 205	
ggt ctc gcc ggt agc ggc gac atg tgg aac tcg atg cac aac aag cat	672
Gly Leu Ala Gly Ser Gly Asp Met Trp Asn Ser Met His Asn Lys His	
210 215 220	
cac gcg acg cct caa aag gtt cgt cac gac atg gat ctg gac acc acc	720
His Ala Thr Pro Gln Lys Val Arg His Asp Met Asp Leu Asp Thr Thr	
225 230 235 240	
ccc gcg gtg gcg ttc ttc aac acc gcg gtg gaa gac aat cgt ccc cgt	768
Pro Ala Val Ala Phe Phe Asn Thr Ala Val Glu Asp Asn Arg Pro Arg	
245 250 255	
ggc ttt agc aag tac tgg ttg cgc ctt cag gcg tgg acc ttc atc ccc	816
Gly Phe Ser Lys Tyr Trp Leu Arg Leu Gln Ala Trp Thr Phe Ile Pro	
260 265 270	
gtg acg tcc ggc ttg gtg ctc ctt ttc tgg atg ttt ttc ctc cac ccc	864
Val Thr Ser Gly Leu Val Leu Leu Phe Trp Met Phe Phe Leu His Pro	
275 280 285	
tcc aag gct ttg aag ggt ggc aag tac gaa gag ttg gtg tgg atg ctc	912
Ser Lys Ala Leu Lys Gly Gly Lys Tyr Glu Glu Leu Val Trp Met Leu	
290 295 300	
gcc gcg cac gtc atc cgc acg tgg acg atc aag gcg gtg acc gga ttc	960
Ala Ala His Val Ile Arg Thr Trp Thr Ile Lys Ala Val Thr Gly Phe	
305 310 315 320	
acc gcg atg cag tcc tac ggc tta ttt ttg gcg acg agc tgg gtg agc	1008
Thr Ala Met Gln Ser Tyr Gly Leu Phe Leu Ala Thr Ser Trp Val Ser	
325 330 335	
ggc tgc tat ctg ttt gca cac ttc tcc acg tcg cac acg cac ctg gat	1056
Gly Cys Tyr Leu Phe Ala His Phe Ser Thr Ser His Thr His Leu Asp	
340 345 350	
gtg gtg ccc gcg gac gag cat ctc tcc tgg gtt cga tac gcc gtc gat	1104
Val Val Pro Ala Asp Glu His Leu Ser Trp Val Arg Tyr Ala Val Asp	
355 360 365	
cac acg atc gac atc gat ccg agt caa ggt tgg gtg aac tgg ttg atg	1152
His Thr Ile Asp Ile Asp Pro Ser Gln Gly Trp Val Asn Trp Leu Met	
370 375 380	
ggc tac ctc aac tgc caa gtc atc cac cac ctc ttt ccg agc atg ccg	1200
Gly Tyr Leu Asn Cys Gln Val Ile His His Leu Phe Pro Ser Met Pro	
385 390 395 400	

cag ttc cgc cag ccc gag gta tct cgc cgc ttc gtc gcc ttt gcg aaa 1248
 Gln Phe Arg Gln Pro Glu Val Ser Arg Arg Phe Val Ala Phe Ala Lys
 405 410 415

aag tgg aac ctc aac tac aag gtc atg acc tac gcc ggt gcg tgg aag 1296
 Lys Trp Asn Leu Asn Tyr Lys Val Met Thr Tyr Ala Gly Ala Trp Lys
 420 425 430

gca acg ctc gga aac ctc gac aac gtg ggt aag cac tac tac gtg cac 1344
 Ala Thr Leu Gly Asn Leu Asp Asn Val Gly Lys His Tyr Tyr Val His
 435 440 445

ggc caa cac tcc gga aag acg gcg taa 1371
 Gly Gln His Ser Gly Lys Thr Ala
 450 455

<210> 90

<211> 456

<212> PRT

<213> *Ostreococcus tauri*

<400> 90

Met Cys Val Glu Thr Glu Asn Asn Asp Gly Ile Pro Thr Val Glu Ile
 1 5 10 15

Ala Phe Asp Gly Glu Arg Glu Arg Ala Glu Ala Asn Val Lys Leu Ser
 20 25 30

Ala Glu Lys Met Glu Pro Ala Ala Leu Ala Lys Thr Phe Ala Arg Arg
 35 40 45

Tyr Val Val Ile Glu Gly Val Glu Tyr Asp Val Thr Asp Phe Lys His
 50 55 60

Pro Gly Gly Thr Val Ile Phe Tyr Ala Leu Ser Asn Thr Gly Ala Asp
 65 70 75 80

Ala Thr Glu Ala Phe Lys Glu Phe His His Arg Ser Arg Lys Ala Arg
 85 90 95

Lys Ala Leu Ala Ala Leu Pro Ser Arg Pro Ala Lys Thr Ala Lys Val
 100 105 110

Asp Asp Ala Glu Met Leu Gln Asp Phe Ala Lys Trp Arg Lys Glu Leu
 115 120 125

Glu Arg Asp Gly Phe Phe Lys Pro Ser Pro Ala His Val Ala Tyr Arg
 130 135 140

Phe Ala Glu Leu Ala Ala Met Tyr Ala Leu Gly Thr Tyr Leu Met Tyr
 145 150 155 160

Ala Arg Tyr Val Val Ser Ser Val Leu Val Tyr Ala Cys Phe Phe Gly
 165 170 175

Ala Arg Cys Gly Trp Val Gln His Glu Gly Gly His Ser Ser Leu Thr
 180 185 190

Gly Asn Ile Trp Trp Asp Lys Arg Ile Gln Ala Phe Thr Ala Gly Phe
 195 200 205

Gly Leu Ala Gly Ser Gly Asp Met Trp Asn Ser Met His Asn Lys His
 210 215 220

His Ala Thr Pro Gln Lys Val Arg His Asp Met Asp Leu Asp Thr Thr
 225 230 235 240

Pro Ala Val Ala Phe Phe Asn Thr Ala Val Glu Asp Asn Arg Pro Arg
 245 250 255

Gly Phe Ser Lys Tyr Trp Leu Arg Leu Gln Ala Trp Thr Phe Ile Pro
 260 265 270

Val Thr Ser Gly Leu Val Leu Leu Phe Trp Met Phe Phe Leu His Pro
 275 280 285

Ser Lys Ala Leu Lys Gly Gly Lys Tyr Glu Glu Leu Val Trp Met Leu
 290 295 300

Ala Ala His Val Ile Arg Thr Trp Thr Ile Lys Ala Val Thr Gly Phe
 305 310 315 320

Thr Ala Met Gln Ser Tyr Gly Leu Phe Leu Ala Thr Ser Trp Val Ser
 325 330 335

Gly Cys Tyr Leu Phe Ala His Phe Ser Thr Ser His Thr His Leu Asp
 340 345 350

Val Val Pro Ala Asp Glu His Leu Ser Trp Val Arg Tyr Ala Val Asp
 355 360 365

His Thr Ile Asp Ile Asp Pro Ser Gln Gly Trp Val Asn Trp Leu Met
 370 375 380

Gly Tyr Leu Asn Cys Gln Val Ile His His Leu Phe Pro Ser Met Pro
 385 390 395 400

Gln Phe Arg Gln Pro Glu Val Ser Arg Arg Phe Val Ala Phe Ala Lys
 405 410 415

Lys Trp Asn Leu Asn Tyr Lys Val Met Thr Tyr Ala Gly Ala Trp Lys
 420 425 430

Ala Thr Leu Gly Asn Leu Asp Asn Val Gly Lys His Tyr Tyr Val His
 435 440 445

Gly Gln His Ser Gly Lys Thr Ala
 450 455

<210> 91

<211> 606

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(606)

<223> Delta-5 desaturase

<400> 91

atg tac ggt ttg cta tcg ctc aag tcg tgc ttc gtc gac gat ttc aac 48
 Met Tyr Gly Leu Leu Ser Leu Lys Ser Cys Phe Val Asp Asp Phe Asn
 1 5 10 15

gcc tac ttc tcc gga cgc atc ggc tgg gtc aag gtg atg aag ttc acc 96
 Ala Tyr Phe Ser Gly Arg Ile Gly Trp Val Lys Val Met Lys Phe Thr
 20 25 30

cgc ggc gag gcg atc gca ttt tgg ggc acc aag ctc ttg tgg gcc gcg 144
 Arg Gly Glu Ala Ile Ala Phe Trp Gly Thr Lys Leu Leu Trp Ala Ala
 35 40 45

tat tac ctc gcg ttg ccg cta aag atg tcg cat cgg ccg ctc gga gaa 192
 Tyr Tyr Leu Ala Leu Pro Leu Lys Met Ser His Arg Pro Leu Gly Glu
 50 55 60

ctc ctc gca ctc tgg gcc gtc acc gag ttc gtc acc gga tgg ctg ttg 240
 Leu Leu Ala Leu Trp Ala Val Thr Glu Phe Val Thr Gly Trp Leu Leu
 65 70 75 80

gcg ttc atg ttc caa gtc gcc cac gtc gtc ggc gag gtt cac ttc ttc 288
 Ala Phe Met Phe Gln Val Ala His Val Val Gly Glu Val His Phe Phe
 85 90 95

acc ctc gac gcg aag aac cgc gtg aac ttg gga tgg gga gag gca cag 336

163

Thr Leu Asp Ala Lys Asn Arg Val Asn Leu Gly Trp Gly Glu Ala Gln
 100 105 110

ctc atg tcg agc gcg gat ttc gcc cac gga tcc aag ttt tgg acg cac 384
 Leu Met Ser Ser Ala Asp Phe Ala His Gly Ser Lys Phe Trp Thr His
 115 120 125

ttc tcc gga ggc tta aac tac caa gtc gtc cac cat ctc ttc ccg ggc 432
 Phe Ser Gly Gly Leu Asn Tyr Gln Val Val His His Leu Phe Pro Gly
 130 135 140

gtc tgc cac gtg cac tat ccc gcg ctc gcg cca att att aag gcg gca 480
 Val Cys His Val His Tyr Pro Ala Leu Ala Pro Ile Ile Lys Ala Ala
 145 150 155 160

gct gag aag cac ggc ctc cac tac cag att tac ccc acg ttt tgg tcc 528
 Ala Glu Lys His Gly Leu His Tyr Gln Ile Tyr Pro Thr Phe Trp Ser
 165 170 175

gcc ctg cgc gcg cac ttc cgg cac ctc gcc aac gtc ggc cgc gcc gcg 576
 Ala Leu Arg Ala His Phe Arg His Leu Ala Asn Val Gly Arg Ala Ala
 180 185 190

tac gta ccg tcc ctc caa acc gtc gga tga 606
 Tyr Val Pro Ser Leu Gln Thr Val Gly
 195 200

<210> 92

<211> 201

<212> PRT

<213> *Ostreococcus tauri*

<400> 92

Met Tyr Gly Leu Leu Ser Leu Lys Ser Cys Phe Val Asp Asp Phe Asn
 1 5 10 15

Ala Tyr Phe Ser Gly Arg Ile Gly Trp Val Lys Val Met Lys Phe Thr
 20 25 30

Arg Gly Glu Ala Ile Ala Phe Trp Gly Thr Lys Leu Leu Trp Ala Ala
 35 40 45

Tyr Tyr Leu Ala Leu Pro Leu Lys Met Ser His Arg Pro Leu Gly Glu
 50 55 60

Leu Leu Ala Leu Trp Ala Val Thr Glu Phe Val Thr Gly Trp Leu Leu
 65 70 75 80

Ala Phe Met Phe Gln Val Ala His Val Val Gly Glu Val His Phe Phe
 85 90 95

164

Thr Leu Asp Ala Lys Asn Arg Val Asn Leu Gly Trp Gly Glu Ala Gln
 100 105 110

Leu Met Ser Ser Ala Asp Phe Ala His Gly Ser Lys Phe Trp Thr His
 115 120 125

Phe Ser Gly Gly Leu Asn Tyr Gln Val Val His His Leu Phe Pro Gly
 130 135 140

Val Cys His Val His Tyr Pro Ala Leu Ala Pro Ile Ile Lys Ala Ala
 145 150 155 160

Ala Glu Lys His Gly Leu His Tyr Gln Ile Tyr Pro Thr Phe Trp Ser
 165 170 175

Ala Leu Arg Ala His Phe Arg His Leu Ala Asn Val Gly Arg Ala Ala
 180 185 190

Tyr Val Pro Ser Leu Gln Thr Val Gly
 195 200

<210> 93

<211> 714

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(714)

<223> Delta-5 desaturase

<400> 93

atg gtg agc cat cac tcg tac tgt aac gac gcg gat ttg gat cag gat 48
 Met Val Ser His His Ser Tyr Cys Asn Asp Ala Asp Leu Asp Gln Asp
 1 5 10 15

gtg tac acc gca ctg ccg ctc ctg cgc ctg gac ccg tct cag gag ttg 96
 Val Tyr Thr Ala Leu Pro Leu Leu Arg Leu Asp Pro Ser Gln Glu Leu
 20 25 30

aag tgg ttt cat cga tac cag gcg ttt tac gcc ccg ctc atg tgg ccg 144
 Lys Trp Phe His Arg Tyr Gln Ala Phe Tyr Ala Pro Leu Met Trp Pro
 35 40 45

ttt ttg tgg ctc gcg gcg cag ttt ggc gac gcg cag aac atc ctg atc 192
 Phe Leu Trp Leu Ala Ala Gln Phe Gly Asp Ala Gln Asn Ile Leu Ile
 50 55 60

165

gac cga gcg tcg ccg ggc gtc gcg tac aag gga ttg atg gcg aac gag 240
 Asp Arg Ala Ser Pro Gly Val Ala Tyr Lys Gly Leu Met Ala Asn Glu
 65 70 75 80
 gtc gcg ctg tac gtt ctc ggt aag gtt tta cac ttt ggt ctt ctc ctc 288
 Val Ala Leu Tyr Val Leu Gly Lys Val Leu His Phe Gly Leu Leu Leu
 85 90 95
 ggc gtt cct gcg tac ttg cac gga ttg tcc aac gcg atc gtt cca ttc 336
 Gly Val Pro Ala Tyr Leu His Gly Leu Ser Asn Ala Ile Val Pro Phe
 100 105 110
 ttg gcg tac ggc gca ttc ggc tcc ttc gtc ctg tgc tgg ttc ttc atc 384
 Leu Ala Tyr Gly Ala Phe Gly Ser Phe Val Leu Cys Trp Phe Phe Ile
 115 120 125
 gtc agc cat aac ctc gaa gcg ctg aca ccc gtt aac ctt aac aag tcc 432
 Val Ser His Asn Leu Glu Ala Leu Thr Pro Val Asn Leu Asn Lys Ser
 130 135 140
 acg aag aac gac tgg ggg gcg tgg cag atc gag aca tcg gcg tct tgg 480
 Thr Lys Asn Asp Trp Gly Ala Trp Gln Ile Glu Thr Ser Ala Ser Trp
 145 150 155 160
 ggc aac gcg ttc tgg agc ttc ttc tct gga ggt ctg aac ctg caa atc 528
 Gly Asn Ala Phe Trp Ser Phe Phe Ser Gly Gly Leu Asn Leu Gln Ile
 165 170 175
 gag cac cac ctc ttc ccg ggc atg gcg cac aac ctg tac ccg aag atg 576
 Glu His His Leu Phe Pro Gly Met Ala His Asn Leu Tyr Pro Lys Met
 180 185 190
 gtg ccg atc atc aag gac gag tgt gcg aaa gcg ggc gtt cgc tac acc 624
 Val Pro Ile Ile Lys Asp Glu Cys Ala Lys Ala Gly Val Arg Tyr Thr
 195 200 205
 ggt tac ggt ggc tac acc ggc ctg ctc ccg atc acc cgc gac atg ttc 672
 Gly Tyr Gly Gly Tyr Thr Gly Leu Leu Pro Ile Thr Arg Asp Met Phe
 210 215 220
 tcc tac ctc cat aag tgt ggc cga acg gcg aaa cta gcc taa 714
 Ser Tyr Leu His Lys Cys Gly Arg Thr Ala Lys Leu Ala
 225 230 235

<210> 94

<211> 237

<212> PRT

<213> *Ostreococcus tauri*

<400> 94

Met Val Ser His His Ser Tyr Cys Asn Asp Ala Asp Leu Asp Gln Asp
 1 5 10 15

Val Tyr Thr Ala Leu Pro Leu Leu Arg Leu Asp Pro Ser Gln Glu Leu
 20 25 30

Lys Trp Phe His Arg Tyr Gln Ala Phe Tyr Ala Pro Leu Met Trp Pro
 35 40 45

Phe Leu Trp Leu Ala Ala Gln Phe Gly Asp Ala Gln Asn Ile Leu Ile
 50 55 60

Asp Arg Ala Ser Pro Gly Val Ala Tyr Lys Gly Leu Met Ala Asn Glu
 65 70 75 80

Val Ala Leu Tyr Val Leu Gly Lys Val Leu His Phe Gly Leu Leu Leu
 85 90 95

Gly Val Pro Ala Tyr Leu His Gly Leu Ser Asn Ala Ile Val Pro Phe
 100 105 110

Leu Ala Tyr Gly Ala Phe Gly Ser Phe Val Leu Cys Trp Phe Phe Ile
 115 120 125

Val Ser His Asn Leu Glu Ala Leu Thr Pro Val Asn Leu Asn Lys Ser
 130 135 140

Thr Lys Asn Asp Trp Gly Ala Trp Gln Ile Glu Thr Ser Ala Ser Trp
 145 150 155 160

Gly Asn Ala Phe Trp Ser Phe Phe Ser Gly Gly Leu Asn Leu Gln Ile
 165 170 175

Glu His His Leu Phe Pro Gly Met Ala His Asn Leu Tyr Pro Lys Met
 180 185 190

Val Pro Ile Ile Lys Asp Glu Cys Ala Lys Ala Gly Val Arg Tyr Thr
 195 200 205

Gly Tyr Gly Gly Tyr Thr Gly Leu Leu Pro Ile Thr Arg Asp Met Phe
 210 215 220

Ser Tyr Leu His Lys Cys Gly Arg Thr Ala Lys Leu Ala
 225 230 235

<210> 95

<211> 1611

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(1611)

<223> Delta-4 desaturase

<400> 95

atg tac ctc gga cgc ggc cgt ctc gag agc ggg acg acg cga ggg atg	48
Met Tyr Leu Gly Arg Gly Arg Leu Glu Ser Gly Thr Thr Arg Gly Met	
1 5 10 15	

atg cgg acg cac gcg cgg cga ccg tcg acg acg tcg aat ccg tgc gcg	96
Met Arg Thr His Ala Arg Arg Pro Ser Thr Thr Ser Asn Pro Cys Ala	
20 25 30	

cgg tca cgc gtg cgt aag acg acg gag cga tcg ctc gcg cga gtg cga	144
Arg Ser Arg Val Arg Lys Thr Thr Glu Arg Ser Leu Ala Arg Val Arg	
35 40 45	

cga tcg acg agt gag aag gga agc gcg ctc gtg ctc gag cga gag agc	192
Arg Ser Thr Ser Glu Lys Gly Ser Ala Leu Val Leu Glu Arg Glu Ser	
50 55 60	

gaa cgg gag aag gag gag gga ggg aaa gcg cga gcg gag gga ttg cga	240
Glu Arg Glu Lys Glu Glu Gly Gly Lys Ala Arg Ala Glu Gly Leu Arg	
65 70 75 80	

ttc caa cgc ccg gac gtc gcc gcg ccg ggg gga gcg gat cct tgg aac	288
Phe Gln Arg Pro Asp Val Ala Ala Pro Gly Gly Ala Asp Pro Trp Asn	
85 90 95	

gac gag aag tgg aca aag acc aag tgg acg gta ttc aga gac gtc gcg	336
Asp Glu Lys Trp Thr Lys Thr Lys Trp Thr Val Phe Arg Asp Val Ala	
100 105 110	

tac gat ctc gat cct ttc ttc gct cga cac ccc gga gga gac tgg ctc	384
Tyr Asp Leu Asp Pro Phe Phe Ala Arg His Pro Gly Gly Asp Trp Leu	
115 120 125	

ctg aac ttg gcc gtg gga cga gac tgc acc gcg ctc atc gaa tcc tat	432
Leu Asn Leu Ala Val Gly Arg Asp Cys Thr Ala Leu Ile Glu Ser Tyr	
130 135 140	

cac ttg cga cca gag gtg gcg acg gct cgt ttc aga atg ctg ccc aaa	480
His Leu Arg Pro Glu Val Ala Thr Ala Arg Phe Arg Met Leu Pro Lys	
145 150 155 160	

ctc gag gat ttt ccc gtc gag gcc gtg ccc aag tcc ccg aga ccg aac	528
Leu Glu Asp Phe Pro Val Glu Ala Val Pro Lys Ser Pro Arg Pro Asn	
165 170 175	

gat tcg ccg tta tac aac aac att cgc aac cga gtc gcg gaa gag ctc	576
Asp Ser Pro Leu Tyr Asn Asn Ile Arg Asn Arg Val Arg Glu Glu Leu	
180 185 190	

ttc cca gag gag gga aag aat atg cac aga cag ggc ggc gac cac ggc	624
Phe Pro Glu Glu Gly Lys Asn Met His Arg Gln Gly Gly Asp His Gly	
195 200 205	

gac ggt gac gat tct ggg ttt cgc cgc ctt ttg ctt atg ccg tgt acc	672
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Asp	Gly	Asp	Asp	Ser	Gly	Phe	Arg	Arg	Leu	Leu	Leu	Met	Pro	Cys	Thr	
210						215					220					
tat	tcc	ctt	ccg	ggg	gtt	cct	ttc	cgg	ctg	cct	cct	cgg	gtc	tcg	cgg	720
Tyr	Ser	Leu	Pro	Gly	Val	Pro	Phe	Arg	Leu	Pro	Pro	Arg	Val	Ser	Arg	
225					230					235					240	
ggg	cgt	gga	ttg	gtc	tca	cga	ttc	agg	cac	tgc	gcc	aac	cac	ggc	gcg	768
Gly	Arg	Gly	Leu	Val	Ser	Arg	Phe	Arg	His	Cys	Ala	Asn	His	Gly	Ala	
				245					250					255		
atg	tct	cct	tcg	ccg	gcc	gtt	aac	ggc	gtc	ctc	ggg	ttg	acg	aac	gat	816
Met	Ser	Pro	Ser	Pro	Ala	Val	Asn	Gly	Val	Leu	Gly	Leu	Thr	Asn	Asp	
			260					265					270			
ctc	atc	ggc	ggc	tcg	tcc	ttg	atg	tgg	aga	tat	cac	cac	caa	gtc	agc	864
Leu	Ile	Gly	Gly	Ser	Ser	Leu	Met	Trp	Arg	Tyr	His	His	Gln	Val	Ser	
		275					280					285				
cac	cac	att	cat	tgc	aac	gac	aac	gcc	atg	gat	caa	gac	gtg	tac	acg	912
His	His	Ile	His	Cys	Asn	Asp	Asn	Ala	Met	Asp	Gln	Asp	Val	Tyr	Thr	
		290				295					300					
gcg	atg	cca	tta	ttg	cgt	ttc	gac	gct	cgc	cgg	ccc	aag	tcc	tgg	tac	960
Ala	Met	Pro	Leu	Leu	Arg	Phe	Asp	Ala	Arg	Pro	Lys	Ser	Trp	Tyr		
305				310					315					320		
cat	cgc	ttc	cag	cag	tgg	tac	atg	ttt	tta	gcg	ttc	ccg	ttg	ttg	cag	1008
His	Arg	Phe	Gln	Gln	Trp	Tyr	Met	Phe	Leu	Ala	Phe	Pro	Leu	Leu	Gln	
			325						330					335		
gtt	gcc	ttc	caa	gtc	gga	gac	att	gcc	gca	ctg	ttc	acg	cgt	gat	acc	1056
Val	Ala	Phe	Gln	Val	Gly	Asp	Ile	Ala	Ala	Leu	Phe	Thr	Arg	Asp	Thr	
			340					345					350			
gaa	ggc	gct	aag	ctt	cac	ggg	gcg	acg	acg	tgg	gag	ctt	acc	acg	gtt	1104
Glu	Gly	Ala	Lys	Leu	His	Gly	Ala	Thr	Thr	Trp	Glu	Leu	Thr	Thr	Val	
		355					360					365				
gtc	ctc	ggg	aag	att	gtg	cac	ttc	ggg	ctt	ttg	ttg	ggg	ccg	ttg	atg	1152
Val	Leu	Gly	Lys	Ile	Val	His	Phe	Gly	Leu	Leu	Leu	Gly	Pro	Leu	Met	
		370				375						380				
aac	cac	gcg	gtg	agt	tct	gtt	ttg	ctg	ggg	atc	gtc	ggg	ttc	atg	gcg	1200
Asn	His	Ala	Val	Ser	Ser	Val	Leu	Leu	Gly	Ile	Val	Gly	Phe	Met	Ala	
385					390					395					400	
tgc	caa	ggg	ata	gtt	ctg	gcg	tgc	acg	ttt	gct	gtg	agt	cac	aat	gtc	1248
Cys	Gln	Gly	Ile	Val	Leu	Ala	Cys	Thr	Phe	Ala	Val	Ser	His	Asn	Val	
			405						410					415		
gcg	gag	gcg	aag	ata	cct	gag	gac	acc	gga	gga	gaa	gcc	tgg	gag	aga	1296
Ala	Glu	Ala	Lys	Ile	Pro	Glu	Asp	Thr	Gly	Gly	Glu	Ala	Trp	Glu	Arg	
			420					425					430			
gat	tgg	ggg	gtc	cag	cag	ttg	gtg	act	agc	gcc	gac	tgg	ggg	gga	aag	1344
Asp	Trp	Gly	Val	Gln	Gln	Leu	Val	Thr	Ser	Ala	Asp	Trp	Gly	Gly	Lys	
		435					440					445				
ata	ggg	aac	ttc	ttc	acg	ggg	ggc	ctc	aac	ttg	caa	gtt	gag	cac	cac	1392
Ile	Gly	Asn	Phe	Phe	Thr	Gly	Gly	Leu	Asn	Leu	Gln	Val	Glu	His	His	
		450				455					460					
ttg	ttt	ccg	gcg	att	tgc	ttc	gtc	cac	tac	ccg	gac	atc	gcg	aag	atc	1440

169

Leu Phe Pro Ala Ile Cys Phe Val His Tyr Pro Asp Ile Ala Lys Ile
 465 470 475 480
 gtg aag gaa gaa gcg gcc aag ctc aac atc cct tac gcg tct tac agg 1488
 Val Lys Glu Glu Ala Ala Lys Leu Asn Ile Pro Tyr Ala Ser Tyr Arg
 485 490 495
 act ctt cct ggt att ttc gtc caa ttc tgg aga ttt atg aag gac atg 1536
 Thr Leu Pro Gly Ile Phe Val Gln Phe Trp Arg Phe Met Lys Asp Met
 500 505 510
 ggc acg gct gag caa att ggt gaa gtt cca ttg ccg aag att ccc aac 1584
 Gly Thr Ala Glu Gln Ile Gly Glu Val Pro Leu Pro Lys Ile Pro Asn
 515 520 525
 ccg cag ctc gcg ccg aag ctc gct tag 1611
 Pro Gln Leu Ala Pro Lys Leu Ala
 530 535

<210> 96

<211> 536

<212> PRT

<213> *Ostreococcus tauri*

<400> 96

Met Tyr Leu Gly Arg Gly Arg Leu Glu Ser Gly Thr Thr Arg Gly Met
 1 5 10 15
 Met Arg Thr His Ala Arg Arg Pro Ser Thr Thr Ser Asn Pro Cys Ala
 20 25 30
 Arg Ser Arg Val Arg Lys Thr Thr Glu Arg Ser Leu Ala Arg Val Arg
 35 40 45
 Arg Ser Thr Ser Glu Lys Gly Ser Ala Leu Val Leu Glu Arg Glu Ser
 50 55 60
 Glu Arg Glu Lys Glu Glu Gly Gly Lys Ala Arg Ala Glu Gly Leu Arg
 65 70 75 80
 Phe Gln Arg Pro Asp Val Ala Ala Pro Gly Gly Ala Asp Pro Trp Asn
 85 90 95
 Asp Glu Lys Trp Thr Lys Thr Lys Trp Thr Val Phe Arg Asp Val Ala
 100 105 110
 Tyr Asp Leu Asp Pro Phe Phe Ala Arg His Pro Gly Gly Asp Trp Leu
 115 120 125

170

Leu Asn Leu Ala Val Gly Arg Asp Cys Thr Ala Leu Ile Glu Ser Tyr
 130 135 140

His Leu Arg Pro Glu Val Ala Thr Ala Arg Phe Arg Met Leu Pro Lys
 145 150 155 160

Leu Glu Asp Phe Pro Val Glu Ala Val Pro Lys Ser Pro Arg Pro Asn
 165 170 175

Asp Ser Pro Leu Tyr Asn Asn Ile Arg Asn Arg Val Arg Glu Glu Leu
 180 185 190

Phe Pro Glu Glu Gly Lys Asn Met His Arg Gln Gly Gly Asp His Gly
 195 200 205

Asp Gly Asp Asp Ser Gly Phe Arg Arg Leu Leu Leu Met Pro Cys Thr
 210 215 220

Tyr Ser Leu Pro Gly Val Pro Phe Arg Leu Pro Pro Arg Val Ser Arg
 225 230 235 240

Gly Arg Gly Leu Val Ser Arg Phe Arg His Cys Ala Asn His Gly Ala
 245 250 255

Met Ser Pro Ser Pro Ala Val Asn Gly Val Leu Gly Leu Thr Asn Asp
 260 265 270

Leu Ile Gly Gly Ser Ser Leu Met Trp Arg Tyr His His Gln Val Ser
 275 280 285

His His Ile His Cys Asn Asp Asn Ala Met Asp Gln Asp Val Tyr Thr
 290 295 300

Ala Met Pro Leu Leu Arg Phe Asp Ala Arg Arg Pro Lys Ser Trp Tyr
 305 310 315 320

His Arg Phe Gln Gln Trp Tyr Met Phe Leu Ala Phe Pro Leu Leu Gln
 325 330 335

Val Ala Phe Gln Val Gly Asp Ile Ala Ala Leu Phe Thr Arg Asp Thr
 340 345 350

Glu Gly Ala Lys Leu His Gly Ala Thr Thr Trp Glu Leu Thr Thr Val
 355 360 365

Val Leu Gly Lys Ile Val His Phe Gly Leu Leu Leu Gly Pro Leu Met
 370 375 380

171

Asn His Ala Val Ser Ser Val Leu Leu Gly Ile Val Gly Phe Met Ala
 385 390 395 400

Cys Gln Gly Ile Val Leu Ala Cys Thr Phe Ala Val Ser His Asn Val
 405 410 415

Ala Glu Ala Lys Ile Pro Glu Asp Thr Gly Gly Glu Ala Trp Glu Arg
 420 425 430

Asp Trp Gly Val Gln Gln Leu Val Thr Ser Ala Asp Trp Gly Gly Lys
 435 440 445

Ile Gly Asn Phe Phe Thr Gly Gly Leu Asn Leu Gln Val Glu His His
 450 455 460

Leu Phe Pro Ala Ile Cys Phe Val His Tyr Pro Asp Ile Ala Lys Ile
 465 470 475 480

Val Lys Glu Glu Ala Ala Lys Leu Asn Ile Pro Tyr Ala Ser Tyr Arg
 485 490 495

Thr Leu Pro Gly Ile Phe Val Gln Phe Trp Arg Phe Met Lys Asp Met
 500 505 510

Gly Thr Ala Glu Gln Ile Gly Glu Val Pro Leu Pro Lys Ile Pro Asn
 515 520 525

Pro Gln Leu Ala Pro Lys Leu Ala
 530 535

<210> 97

<211> 1455

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(1455)

<223> Delta-6 desaturase

<400> 97

atg gga aaa gga gga gac gca gcc gca gct acc aag cgt agt gga gca
 Met Gly Lys Gly Gly Asp Ala Ala Ala Ala Thr Lys Arg Ser Gly Ala
 1 5 10 15

ttg aaa ttg gcg gag aag ccg cag aag tac act tgg cag gag gtg aag	96
Leu Lys Leu Ala Glu Lys Pro Gln Lys Tyr Thr Trp Gln Glu Val Lys	
20 25 30	
aag cac atc acc ccc gac gat gcc tgg gta gtc cac caa aac aaa gtc	144
Lys His Ile Thr Pro Asp Asp Ala Trp Val Val His Gln Asn Lys Val	
35 40 45	
tac gac gtc tcc aac tgg tac gac cac ccc ggt gga gcc gtg gtg ttc	192
Tyr Asp Val Ser Asn Trp Tyr Asp His Pro Gly Gly Ala Val Val Phe	
50 55 60	
acc cac gcc gga gac gac atg acg gac atc ttc gcc gcc ttc cac gcc	240
Thr His Ala Gly Asp Asp Met Thr Asp Ile Phe Ala Ala Phe His Ala	
65 70 75 80	
caa ggc tct cag gcc atg atg aag aag ttt tac att gga gat ttg att	288
Gln Gly Ser Gln Ala Met Met Lys Lys Phe Tyr Ile Gly Asp Leu Ile	
85 90 95	
ccg gag agt gtg gag cat aag gat caa aga cag ttg gat ttc gag aag	336
Pro Glu Ser Val Glu His Lys Asp Gln Arg Gln Leu Asp Phe Glu Lys	
100 105 110	
gga tat cgt gat tta cgg gcc aag ctt gtc atg atg ggg atg ttc aag	384
Gly Tyr Arg Asp Leu Arg Ala Lys Leu Val Met Met Gly Met Phe Lys	
115 120 125	
tcg agt aag atg tat tat gca tac aag tgc tcg ttc aat atg tgc atg	432
Ser Ser Lys Met Tyr Tyr Ala Tyr Lys Cys Ser Phe Asn Met Cys Met	
130 135 140	
tgg ttg gtg gcg gtg gcc atg gtg tac tac tcg gac agt ttg gca atg	480
Trp Leu Val Ala Val Ala Met Val Tyr Tyr Ser Asp Ser Leu Ala Met	
145 150 155 160	
cac att gga tcg gct ctc ttg ttg gga ttg ttc tgg cag cag tgt gga	528
His Ile Gly Ser Ala Leu Leu Leu Gly Leu Phe Trp Gln Gln Cys Gly	
165 170 175	
tgg ctt gcg cac gac ttt ctt cac cac caa gtc ttt aag caa cga aag	576
Trp Leu Ala His Asp Phe Leu His His Gln Val Phe Lys Gln Arg Lys	
180 185 190	
tac gga gat ctc gtt ggc atc ttt tgg gga gat ctc atg cag ggg ttc	624
Tyr Gly Asp Leu Val Gly Ile Phe Trp Gly Asp Leu Met Gln Gly Phe	
195 200 205	
tcg atg cag tgg tgg aag aac aag cac aat ggc cac cat gct gtt ccc	672
Ser Met Gln Trp Trp Lys Asn Lys His Asn Gly His His Ala Val Pro	
210 215 220	
aac ttg cac aac tct tcc ttg gac agt cag gat ggt gat ccc gat att	720
Asn Leu His Asn Ser Ser Leu Asp Ser Gln Asp Gly Asp Pro Asp Ile	
225 230 235 240	
gat acc atg cca ctc ctt gct tgg agt ctc aag cag gct cag agt ttc	768
Asp Thr Met Pro Leu Leu Ala Trp Ser Leu Lys Gln Ala Gln Ser Phe	
245 250 255	
aga gag atc aat aag gga aag gac agt acc ttc gtc aag tac gct atc	816
Arg Glu Ile Asn Lys Gly Lys Asp Ser Thr Phe Val Lys Tyr Ala Ile	
260 265 270	

aaa ttc cag gca ttc aca tac ttc ccc atc ctc ctc ttg gct cgc atc	864
Lys Phe Gln Ala Phe Thr Tyr Phe Pro Ile Leu Leu Ala Arg Ile	
275 280 285	
tct tgg ttg aat gaa tcc ttc aaa act gca ttc gga ctc gga gct gcc	912
Ser Trp Leu Asn Glu Ser Phe Lys Thr Ala Phe Gly Leu Gly Ala Ala	
290 295 300	
tcg gag aat gcc aag ttg gag ttg gag aag cgt gga ctt cag tac cca	960
Ser Glu Asn Ala Lys Leu Glu Leu Glu Lys Arg Gly Leu Gln Tyr Pro	
305 310 315 320	
ctt ttg gag aag ctt gga atc acc ctt cat tac act ttg atg ttc gtc	1008
Leu Leu Glu Lys Leu Gly Ile Thr Leu His Tyr Thr Trp Met Phe Val	
325 330 335	
ctc tct tcc gga ttt gga agg tgg tct ctt cca tat tcc atc atg tat	1056
Leu Ser Ser Gly Phe Gly Arg Trp Ser Leu Pro Tyr Ser Ile Met Tyr	
340 345 350	
ttc ttc act gcc aca tgc tcc tcg gga ctt ttc ctc gca ttg gtc ttt	1104
Phe Phe Thr Ala Thr Cys Ser Ser Gly Leu Phe Leu Ala Leu Val Phe	
355 360 365	
gga ttg gga cac aac ggt atg tca gtg tac gat gcc acc acc cga cct	1152
Gly Leu Gly His Asn Gly Met Ser Val Tyr Asp Ala Thr Thr Arg Pro	
370 375 380	
gac ttc tgg caa ctc caa gtc acc act aca cgt aac atc att ggt gga	1200
Asp Phe Trp Gln Leu Gln Val Thr Thr Thr Arg Asn Ile Ile Gly Gly	
385 390 395 400	
cac ggc att ccc caa ttc ttt gtg gat tgg ttc tgc ggt gga ttg caa	1248
His Gly Ile Pro Gln Phe Phe Val Asp Trp Phe Cys Gly Gly Leu Gln	
405 410 415	
tac caa gtg gat cac cac ctc ttc ccc atg atg cct aga aac aat atc	1296
Tyr Gln Val Asp His His Leu Phe Pro Met Met Pro Arg Asn Asn Ile	
420 425 430	
gcg aaa tgc cac aag ctt gtg gag tca ttc tgt aag gag tgg ggt gtg	1344
Ala Lys Cys His Lys Leu Val Glu Ser Phe Cys Lys Glu Trp Gly Val	
435 440 445	
aag tac cat gag gcc gat atg tgg gat ggt acc gtg gaa gtg ttg caa	1392
Lys Tyr His Glu Ala Asp Met Trp Asp Gly Thr Val Glu Val Leu Gln	
450 455 460	
cat ctc tcc aag gtg tcg gat gat ttc ctt gtg gag atg gtg aag gat	1440
His Leu Ser Lys Val Ser Asp Asp Phe Leu Val Glu Met Val Lys Asp	
465 470 475 480	
ttc cct gcc atg taa	1455
Phe Pro Ala Met	

<210> 98

<211> 484

<212> PRT

<213> Thalassiosira pseudonana

<400> 98

Met Gly Lys Gly Gly Asp Ala Ala Ala Ala Thr Lys Arg Ser Gly Ala
 1 5 10 15

Leu Lys Leu Ala Glu Lys Pro Gln Lys Tyr Thr Trp Gln Glu Val Lys
 20 25 30

Lys His Ile Thr Pro Asp Asp Ala Trp Val Val His Gln Asn Lys Val
 35 40 45

Tyr Asp Val Ser Asn Trp Tyr Asp His Pro Gly Gly Ala Val Val Phe
 50 55 60

Thr His Ala Gly Asp Asp Met Thr Asp Ile Phe Ala Ala Phe His Ala
 65 70 75 80

Gln Gly Ser Gln Ala Met Met Lys Lys Phe Tyr Ile Gly Asp Leu Ile
 85 90 95

Pro Glu Ser Val Glu His Lys Asp Gln Arg Gln Leu Asp Phe Glu Lys
 100 105 110

Gly Tyr Arg Asp Leu Arg Ala Lys Leu Val Met Met Gly Met Phe Lys
 115 120 125

Ser Ser Lys Met Tyr Tyr Ala Tyr Lys Cys Ser Phe Asn Met Cys Met
 130 135 140

Trp Leu Val Ala Val Ala Met Val Tyr Tyr Ser Asp Ser Leu Ala Met
 145 150 155 160

His Ile Gly Ser Ala Leu Leu Leu Gly Leu Phe Trp Gln Gln Cys Gly
 165 170 175

Trp Leu Ala His Asp Phe Leu His His Gln Val Phe Lys Gln Arg Lys
 180 185 190

Tyr Gly Asp Leu Val Gly Ile Phe Trp Gly Asp Leu Met Gln Gly Phe
 195 200 205

Ser Met Gln Trp Trp Lys Asn Lys His Asn Gly His His Ala Val Pro
 210 215 220

Asn Leu His Asn Ser Ser Leu Asp Ser Gln Asp Gly Asp Pro Asp Ile
 225 230 235 240

Asp Thr Met Pro Leu Leu Ala Trp Ser Leu Lys Gln Ala Gln Ser Phe
245 250 255

Arg Glu Ile Asn Lys Gly Lys Asp Ser Thr Phe Val Lys Tyr Ala Ile
260 265 270

Lys Phe Gln Ala Phe Thr Tyr Phe Pro Ile Leu Leu Leu Ala Arg Ile
275 280 285

Ser Trp Leu Asn Glu Ser Phe Lys Thr Ala Phe Gly Leu Gly Ala Ala
290 295 300

Ser Glu Asn Ala Lys Leu Glu Leu Glu Lys Arg Gly Leu Gln Tyr Pro
305 310 315 320

Leu Leu Glu Lys Leu Gly Ile Thr Leu His Tyr Thr Trp Met Phe Val
325 330 335

Leu Ser Ser Gly Phe Gly Arg Trp Ser Leu Pro Tyr Ser Ile Met Tyr
340 345 350

Phe Phe Thr Ala Thr Cys Ser Ser Gly Leu Phe Leu Ala Leu Val Phe
355 360 365

Gly Leu Gly His Asn Gly Met Ser Val Tyr Asp Ala Thr Thr Arg Pro
370 375 380

Asp Phe Trp Gln Leu Gln Val Thr Thr Thr Arg Asn Ile Ile Gly Gly
385 390 395 400

His Gly Ile Pro Gln Phe Phe Val Asp Trp Phe Cys Gly Gly Leu Gln
405 410 415

Tyr Gln Val Asp His His Leu Phe Pro Met Met Pro Arg Asn Asn Ile
420 425 430

Ala Lys Cys His Lys Leu Val Glu Ser Phe Cys Lys Glu Trp Gly Val
435 440 445

Lys Tyr His Glu Ala Asp Met Trp Asp Gly Thr Val Glu Val Leu Gln
450 455 460

His Leu Ser Lys Val Ser Asp Asp Phe Leu Val Glu Met Val Lys Asp
465 470 475 480

Phe Pro Ala Met

<210> 99

<211> 1431

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(1431)

<223> Delta-5 desaturase

<400> 99

atg ccc ccc aac gcc gat atc tcc cgc atc cgc aac cgc atc ccc acc	48
Met Pro Pro Asn Ala Asp Ile Ser Arg Ile Arg Asn Arg Ile Pro Thr	
1 5 10 15	

aaa aca ggt acc gtt gcc tct gcc gac aac aac gac ccc gcc acc caa	96
Lys Thr Gly Thr Val Ala Ser Ala Asp Asn Asn Asp Pro Ala Thr Gln	
20 25 30	

tcc gtc cga acc ctc aaa tct ctc aag ggc aac gag gtc gtc atc aac	144
Ser Val Arg Thr Leu Lys Ser Leu Lys Gly Asn Glu Val Val Ile Asn	
35 40 45	

ggc aca att tat gac att gct gac ttt gtc cat cct gga gga gag gtt	192
Gly Thr Ile Tyr Asp Ile Ala Asp Phe Val His Pro Gly Gly Glu Val	
50 55 60	

gtc aag ttc ttt ggt ggg aat gat gtt act att cag tat aat atg att	240
Val Lys Phe Phe Gly Gly Asn Asp Val Thr Ile Gln Tyr Asn Met Ile	
65 70 75 80	

cat ccg tat cat acg ggg aaa cat ctg gag aag atg aag gct gtt gga	288
His Pro Tyr His Thr Gly Lys His Leu Glu Lys Met Lys Ala Val Gly	
85 90 95	

aag gtt gta gat tgg cag tcg gac tac aag ttc gac acc ccc ttt gaa	336
Lys Val Val Asp Trp Gln Ser Asp Tyr Lys Phe Asp Thr Pro Phe Glu	
100 105 110	

cga gag atc aaa tca gaa gtg ttc aag atc gta cgt cgc ggg cgt gag	384
Arg Glu Ile Lys Ser Glu Val Phe Lys Ile Val Arg Arg Gly Arg Glu	
115 120 125	

ttc ggc aca aca ggc tac ttc ctc cgt gcc ttt ttc tac atc gct ctc	432
Phe Gly Thr Thr Gly Tyr Phe Leu Arg Ala Phe Phe Tyr Ile Ala Leu	
130 135 140	

ttc ttc acc atg caa tac act ttc gcc aca tgc acc acc ttc acc acc	480
Phe Phe Thr Met Gln Tyr Thr Phe Ala Thr Cys Thr Thr Phe Thr Thr	
145 150 155 160	

tac gat cac tgg tat cag agt ggt gta ttc atc gca att gtg ttt ggt	528
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177

Tyr	Asp	His	Trp	Tyr	Gln	Ser	Gly	Val	Phe	Ile	Ala	Ile	Val	Phe	Gly	
				165					170					175		
att	tca	cag	gca	ttc	att	ggg	ttg	aat	gtc	cag	cac	gat	gcc	aat	cac	576
Ile	Ser	Gln	Ala	Phe	Ile	Gly	Leu	Asn	Val	Gln	His	Asp	Ala	Asn	His	
			180					185					190			
gga	gct	gcc	agt	aag	cgt	ccc	tgg	gtg	aat	gac	ttg	ttg	gga	ttt	gga	624
Gly	Ala	Ala	Ser	Lys	Arg	Pro	Trp	Val	Asn	Asp	Leu	Leu	Gly	Phe	Gly	
			195				200					205				
acg	gat	ttg	att	gga	tct	aac	aaa	tgg	aat	tgg	atg	gca	cag	cat	tgg	672
Thr	Asp	Leu	Ile	Gly	Ser	Asn	Lys	Trp	Asn	Trp	Met	Ala	Gln	His	Trp	
	210					215					220					
act	cat	cac	gct	tac	act	aac	cat	agt	gag	aag	gat	ccc	gat	agc	ttc	720
Thr	His	His	Ala	Tyr	Thr	Asn	His	Ser	Glu	Lys	Asp	Pro	Asp	Ser	Phe	
	225					230				235					240	
agc	tcg	gaa	cct	atg	ttt	gca	ttc	aat	gac	tat	ccc	att	gga	cac	ccg	768
Ser	Ser	Glu	Pro	Met	Phe	Ala	Phe	Asn	Asp	Tyr	Pro	Ile	Gly	His	Pro	
				245					250					255		
aag	aga	aag	tgg	tgg	cat	agg	ttc	cag	gga	ggg	tac	ttc	ctc	ttc	atg	816
Lys	Arg	Lys	Trp	Trp	His	Arg	Phe	Gln	Gly	Gly	Tyr	Phe	Leu	Phe	Met	
			260					265					270			
ctt	gga	ctt	tac	tgg	ctc	tcg	act	gta	ttc	aat	ccg	caa	ttc	att	gat	864
Leu	Gly	Leu	Tyr	Trp	Leu	Ser	Thr	Val	Phe	Asn	Pro	Gln	Phe	Ile	Asp	
		275					280					285				
ctt	cgt	caa	cgt	ggg	gct	cag	tac	gtc	gga	att	caa	atg	gag	aat	gat	912
Leu	Arg	Gln	Arg	Gly	Ala	Gln	Tyr	Val	Gly	Ile	Gln	Met	Glu	Asn	Asp	
	290					295					300					
ttc	att	gtc	aag	agg	agg	aag	tac	gcc	gtt	gca	ttg	agg	atg	atg	tac	960
Phe	Ile	Val	Lys	Arg	Arg	Lys	Tyr	Ala	Val	Ala	Leu	Arg	Met	Met	Tyr	
	305					310				315					320	
att	tac	ttg	aac	att	gtc	agc	ccc	ttc	atg	aac	aat	ggg	ttg	agc	tgg	1008
Ile	Tyr	Leu	Asn	Ile	Val	Ser	Pro	Phe	Met	Asn	Asn	Gly	Leu	Ser	Trp	
			325						330					335		
tct	acc	ttt	gga	atc	atc	atg	ttg	atg	gga	atc	agc	gag	agt	ctc	act	1056
Ser	Thr	Phe	Gly	Ile	Ile	Met	Leu	Met	Gly	Ile	Ser	Glu	Ser	Leu	Thr	
			340					345					350			
ctc	agt	gtg	ctc	ttc	tcg	ttg	tct	cac	aac	ttc	atc	aat	tcg	gat	cgt	1104
Leu	Ser	Val	Leu	Phe	Ser	Leu	Ser	His	Asn	Phe	Ile	Asn	Ser	Asp	Arg	
		355					360					365				
gat	cct	acg	gct	gac	ttc	aaa	aag	acc	gga	gaa	caa	gtg	tgc	tgg	ttc	1152
Asp	Pro	Thr	Ala	Asp	Phe	Lys	Lys	Thr	Gly	Glu	Gln	Val	Cys	Trp	Phe	
	370					375					380					
aag	tcg	cag	gtg	gag	act	tcg	tct	acc	tat	ggg	ggg	ttt	att	tcc	gga	1200
Lys	Ser	Gln	Val	Glu	Thr	Ser	Ser	Thr	Tyr	Gly	Gly	Phe	Ile	Ser	Gly	
	385				390					395					400	
tgt	ctt	acg	gga	gga	ctc	aac	ttt	cag	gtg	gaa	cat	cat	ctc	ttt	ccc	1248
Cys	Leu	Thr	Gly	Gly	Leu	Asn	Phe	Gln	Val	Glu	His	His	Leu	Phe	Pro	
			405						410					415		
cgt	atg	agc	agt	gct	tgg	tat	cct	tac	att	gca	cct	acg	gtt	cgt	gag	1296

178

Arg Met Ser Ser Ala Trp Tyr Pro Tyr Ile Ala Pro Thr Val Arg Glu
 420 425 430

gtt tgc aag aag cac ggg gtg aac tac gct tat tat cct tgg att ggg 1344
 Val Cys Lys Lys His Gly Val Asn Tyr Ala Tyr Tyr Pro Trp Ile Gly
 435 440 445

cag aat ttg gta tca aca ttc aaa tac atg cat cgc gct ggt agt gga 1392
 Gln Asn Leu Val Ser Thr Phe Lys Tyr Met His Arg Ala Gly Ser Gly
 450 455 460

gcc aac tgg gag ctc aag ccg ttg tct gga agt gcc taa 1431
 Ala Asn Trp Glu Leu Lys Pro Leu Ser Gly Ser Ala
 465 470 475

<210> 100

<211> 476

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 100

Met Pro Pro Asn Ala Asp Ile Ser Arg Ile Arg Asn Arg Ile Pro Thr
 1 5 10 15

Lys Thr Gly Thr Val Ala Ser Ala Asp Asn Asn Asp Pro Ala Thr Gln
 20 25 30

Ser Val Arg Thr Leu Lys Ser Leu Lys Gly Asn Glu Val Val Ile Asn
 35 40 45

Gly Thr Ile Tyr Asp Ile Ala Asp Phe Val His Pro Gly Gly Glu Val
 50 55 60

Val Lys Phe Phe Gly Gly Asn Asp Val Thr Ile Gln Tyr Asn Met Ile
 65 70 75 80

His Pro Tyr His Thr Gly Lys His Leu Glu Lys Met Lys Ala Val Gly
 85 90 95

Lys Val Val Asp Trp Gln Ser Asp Tyr Lys Phe Asp Thr Pro Phe Glu
 100 105 110

Arg Glu Ile Lys Ser Glu Val Phe Lys Ile Val Arg Arg Gly Arg Glu
 115 120 125

Phe Gly Thr Thr Gly Tyr Phe Leu Arg Ala Phe Phe Tyr Ile Ala Leu
 130 135 140

179

Phe Phe Thr Met Gln Tyr Thr Phe Ala Thr Cys Thr Thr Phe Thr Thr
 145 150 155 160

Tyr Asp His Trp Tyr Gln Ser Gly Val Phe Ile Ala Ile Val Phe Gly
 165 170 175

Ile Ser Gln Ala Phe Ile Gly Leu Asn Val Gln His Asp Ala Asn His
 180 185 190

Gly Ala Ala Ser Lys Arg Pro Trp Val Asn Asp Leu Leu Gly Phe Gly
 195 200 205

Thr Asp Leu Ile Gly Ser Asn Lys Trp Asn Trp Met Ala Gln His Trp
 210 215 220

Thr His His Ala Tyr Thr Asn His Ser Glu Lys Asp Pro Asp Ser Phe
 225 230 235 240

Ser Ser Glu Pro Met Phe Ala Phe Asn Asp Tyr Pro Ile Gly His Pro
 245 250 255

Lys Arg Lys Trp Trp His Arg Phe Gln Gly Gly Tyr Phe Leu Phe Met
 260 265 270

Leu Gly Leu Tyr Trp Leu Ser Thr Val Phe Asn Pro Gln Phe Ile Asp
 275 280 285

Leu Arg Gln Arg Gly Ala Gln Tyr Val Gly Ile Gln Met Glu Asn Asp
 290 295 300

Phe Ile Val Lys Arg Arg Lys Tyr Ala Val Ala Leu Arg Met Met Tyr
 305 310 315 320

Ile Tyr Leu Asn Ile Val Ser Pro Phe Met Asn Asn Gly Leu Ser Trp
 325 330 335

Ser Thr Phe Gly Ile Ile Met Leu Met Gly Ile Ser Glu Ser Leu Thr
 340 345 350

Leu Ser Val Leu Phe Ser Leu Ser His Asn Phe Ile Asn Ser Asp Arg
 355 360 365

Asp Pro Thr Ala Asp Phe Lys Lys Thr Gly Glu Gln Val Cys Trp Phe
 370 375 380

Lys Ser Gln Val Glu Thr Ser Ser Thr Tyr Gly Gly Phe Ile Ser Gly
 385 390 395 400

180

Cys Leu Thr Gly Gly Leu Asn Phe Gln Val Glu His His Leu Phe Pro
405 410 415

Arg Met Ser Ser Ala Trp Tyr Pro Tyr Ile Ala Pro Thr Val Arg Glu
420 425 430

Val Cys Lys Lys His Gly Val Asn Tyr Ala Tyr Tyr Pro Trp Ile Gly
435 440 445

Gln Asn Leu Val Ser Thr Phe Lys Tyr Met His Arg Ala Gly Ser Gly
450 455 460

Ala Asn Trp Glu Leu Lys Pro Leu Ser Gly Ser Ala
465 470 475

<210> 101

<211> 1449

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(1449)

<223> Delta-5 desaturase

<400> 101

atg cca ccc aac gcc gag gtc aaa aac ctc cgt tca cgt tcc atc cca 48
Met Pro Pro Asn Ala Glu Val Lys Asn Leu Arg Ser Arg Ser Ile Pro
1 5 10 15

acg aag aag tcc agt tca tcg tca tcc acc gcg aac gac gat ccg gct 96
Thr Lys Lys Ser Ser Ser Ser Ser Thr Ala Asn Asp Asp Pro Ala
20 25 30

acc caa tcc acc tca cct gtg aac cga acc ctc aag tct ttg aat gga 144
Thr Gln Ser Thr Ser Pro Val Asn Arg Thr Leu Lys Ser Leu Asn Gly
35 40 45

aac gaa ata gct att gac ggt gtc atc tat gat att gat ggc ttt gtc 192
Asn Glu Ile Ala Ile Asp Gly Val Ile Tyr Asp Ile Asp Gly Phe Val
50 55 60

cat cct gga gga gag gtt att agc ttc ttt gga ggc aac gat gtg act 240
His Pro Gly Gly Glu Val Ile Ser Phe Phe Gly Gly Asn Asp Val Thr
65 70 75 80

gta cag tac aaa atg att cat ccg tat cat aat agt aag cat ctc gag 288
Val Gln Tyr Lys Met Ile His Pro Tyr His Asn Ser Lys His Leu Glu
85 90 95

aag atg aga gcc gtt gga aag att gca gac tac tcc aca gag tac aag	336
Lys Met Arg Ala Val Gly Lys Ile Ala Asp Tyr Ser Thr Glu Tyr Lys	
100 105 110	
ttc gac aca ccc ttt gaa cga gag atc aaa tcc gaa gtg ttc aaa atc	384
Phe Asp Thr Pro Phe Glu Arg Glu Ile Lys Ser Glu Val Phe Lys Ile	
115 120 125	
gtc cgt cga gga cgt gaa ttc ggt aca aca gga tat ttc ctc cgt gcc	432
Val Arg Arg Gly Arg Glu Phe Gly Thr Thr Gly Tyr Phe Leu Arg Ala	
130 135 140	
ttc ttc tac att gct ctc ttc ttc acc atg caa tac acc ttc gcc aca	480
Phe Phe Tyr Ile Ala Leu Phe Phe Thr Met Gln Tyr Thr Phe Ala Thr	
145 150 155 160	
tgc act acc ttc acc acc tac gat cat tgg tat caa agt ggt gta ttc	528
Cys Thr Thr Phe Thr Thr Tyr Asp His Trp Tyr Gln Ser Gly Val Phe	
165 170 175	
atc gcc att gtg ttt ggt atc tca caa gct ttc att ggg ttg aat gta	576
Ile Ala Ile Val Phe Gly Ile Ser Gln Ala Phe Ile Gly Leu Asn Val	
180 185 190	
caa cat gat gcc aat cac gga gct gct agc aaa cga cct tgg gtg aat	624
Gln His Asp Ala Asn His Gly Ala Ala Ser Lys Arg Pro Trp Val Asn	
195 200 205	
gat ctc ctt gga tct gga gct gat ctc atc ggt gga tgc aaa tgg aac	672
Asp Leu Leu Gly Ser Gly Ala Asp Leu Ile Gly Gly Cys Lys Trp Asn	
210 215 220	
tgg ttg gct cag cat tgg act cat cat gcg tat acc aat cac gct gat	720
Trp Leu Ala Gln His Trp Thr His His Ala Tyr Thr Asn His Ala Asp	
225 230 235 240	
aaa gat cct gat agc ttt agt tcc gag ccg gtc ttc aac ttt aac gat	768
Lys Asp Pro Asp Ser Phe Ser Ser Glu Pro Val Phe Asn Phe Asn Asp	
245 250 255	
tat ccc att ggt cac ccc aaa aga aag tgg tgg cat agg ttc caa ggg	816
Tyr Pro Ile Gly His Pro Lys Arg Lys Trp Trp His Arg Phe Gln Gly	
260 265 270	
ctc tac ttc cta atc atg ctg agt ttc tat tgg gta tcg atg gta ttc	864
Leu Tyr Phe Leu Ile Met Leu Ser Phe Tyr Trp Val Ser Met Val Phe	
275 280 285	
aac cca caa gtt atc gac ctc cgt cat gct gga gct gcc tac gtt gga	912
Asn Pro Gln Val Ile Asp Leu Arg His Ala Gly Ala Ala Tyr Val Gly	
290 295 300	
ttt cag atg gag aac gac ttt atc gtc aaa cgg aga aag tat gca atg	960
Phe Gln Met Glu Asn Asp Phe Ile Val Lys Arg Arg Lys Tyr Ala Met	
305 310 315 320	
gca ctt cgt gca atg tac ttc tat ttc aac atc tat tgt ccg att gtc	1008
Ala Leu Arg Ala Met Tyr Phe Tyr Phe Asn Ile Tyr Cys Pro Ile Val	
325 330 335	
aac aat gga ttg act tgg tcg aca gtt gga atc atc ctc tta atg gga	1056
Asn Asn Gly Leu Thr Trp Ser Thr Val Gly Ile Ile Leu Leu Met Gly	
340 345 350	

gtt agc gaa agc ttc atg ctc tcc ggt cta ttc gta ctc tca cac aac 1104
 Val Ser Glu Ser Phe Met Leu Ser Gly Leu Phe Val Leu Ser His Asn
 355 360 365
 ttt gaa aat tcc gaa cgt gat cct acc tct gag tat cgc aag act ggt 1152
 Phe Glu Asn Ser Glu Arg Asp Pro Thr Ser Glu Tyr Arg Lys Thr Gly
 370 375 380
 gag caa gta tgt tgg ttc aag tct caa gtg gag act tct tct acc tac 1200
 Glu Gln Val Cys Trp Phe Lys Ser Gln Val Glu Thr Ser Ser Thr Tyr
 385 390 395 400
 gga ggt atc gtt gct ggg tgt ctc act ggt gga ctc aac ttt caa gtg 1248
 Gly Gly Ile Val Ala Gly Cys Leu Thr Gly Gly Leu Asn Phe Gln Val
 405 410 415
 gag cat cat ttg ttc ccg agg atg agc agt gct tgg tat cct ttc atc 1296
 Glu His His Leu Phe Pro Arg Met Ser Ser Ala Trp Tyr Pro Phe Ile
 420 425 430
 gcg ccg aag gtt aga gag att tgt aag aag cat gga gtt aga tac gct 1344
 Ala Pro Lys Val Arg Glu Ile Cys Lys Lys His Gly Val Arg Tyr Ala
 435 440 445
 tac tat ccg tac atc tgg cag aac ttg cat tct acc gtg agt tac atg 1392
 Tyr Tyr Pro Tyr Ile Trp Gln Asn Leu His Ser Thr Val Ser Tyr Met
 450 455 460
 cat ggg acg gga acg gga gct aga tgg gag ctt cag ccg ttg tct gga 1440
 His Gly Thr Gly Thr Gly Ala Arg Trp Glu Leu Gln Pro Leu Ser Gly
 465 470 475 480
 agg gcg tag 1449
 Arg Ala

<210> 102

<211> 482

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 102

Met Pro Pro Asn Ala Glu Val Lys Asn Leu Arg Ser Arg Ser Ile Pro
 1 5 10 15
 Thr Lys Lys Ser Ser Ser Ser Ser Ser Thr Ala Asn Asp Asp Pro Ala
 20 25 30
 Thr Gln Ser Thr Ser Pro Val Asn Arg Thr Leu Lys Ser Leu Asn Gly
 35 40 45
 Asn Glu Ile Ala Ile Asp Gly Val Ile Tyr Asp Ile Asp Gly Phe Val
 50 55 60

His Pro Gly Gly Glu Val Ile Ser Phe Phe Gly Gly Asn Asp Val Thr
65 70 75 80

Val Gln Tyr Lys Met Ile His Pro Tyr His Asn Ser Lys His Leu Glu
85 90 95

Lys Met Arg Ala Val Gly Lys Ile Ala Asp Tyr Ser Thr Glu Tyr Lys
100 105 110

Phe Asp Thr Pro Phe Glu Arg Glu Ile Lys Ser Glu Val Phe Lys Ile
115 120 125

Val Arg Arg Gly Arg Glu Phe Gly Thr Thr Gly Tyr Phe Leu Arg Ala
130 135 140

Phe Phe Tyr Ile Ala Leu Phe Phe Thr Met Gln Tyr Thr Phe Ala Thr
145 150 155 160

Cys Thr Thr Phe Thr Thr Tyr Asp His Trp Tyr Gln Ser Gly Val Phe
165 170 175

Ile Ala Ile Val Phe Gly Ile Ser Gln Ala Phe Ile Gly Leu Asn Val
180 185 190

Gln His Asp Ala Asn His Gly Ala Ala Ser Lys Arg Pro Trp Val Asn
195 200 205

Asp Leu Leu Gly Ser Gly Ala Asp Leu Ile Gly Gly Cys Lys Trp Asn
210 215 220

Trp Leu Ala Gln His Trp Thr His His Ala Tyr Thr Asn His Ala Asp
225 230 235 240

Lys Asp Pro Asp Ser Phe Ser Ser Glu Pro Val Phe Asn Phe Asn Asp
245 250 255

Tyr Pro Ile Gly His Pro Lys Arg Lys Trp Trp His Arg Phe Gln Gly
260 265 270

Leu Tyr Phe Leu Ile Met Leu Ser Phe Tyr Trp Val Ser Met Val Phe
275 280 285

Asn Pro Gln Val Ile Asp Leu Arg His Ala Gly Ala Ala Tyr Val Gly
290 295 300

Phe Gln Met Glu Asn Asp Phe Ile Val Lys Arg Arg Lys Tyr Ala Met
305 310 315 320

Ala Leu Arg Ala Met Tyr Phe Tyr Phe Asn Ile Tyr Cys Pro Ile Val
 325 330 335

Asn Asn Gly Leu Thr Trp Ser Thr Val Gly Ile Ile Leu Leu Met Gly
 340 345 350

Val Ser Glu Ser Phe Met Leu Ser Gly Leu Phe Val Leu Ser His Asn
 355 360 365

Phe Glu Asn Ser Glu Arg Asp Pro Thr Ser Glu Tyr Arg Lys Thr Gly
 370 375 380

Glu Gln Val Cys Trp Phe Lys Ser Gln Val Glu Thr Ser Ser Thr Tyr
 385 390 395 400

Gly Gly Ile Val Ala Gly Cys Leu Thr Gly Gly Leu Asn Phe Gln Val
 405 410 415

Glu His His Leu Phe Pro Arg Met Ser Ser Ala Trp Tyr Pro Phe Ile
 420 425 430

Ala Pro Lys Val Arg Glu Ile Cys Lys Lys His Gly Val Arg Tyr Ala
 435 440 445

Tyr Tyr Pro Tyr Ile Trp Gln Asn Leu His Ser Thr Val Ser Tyr Met
 450 455 460

His Gly Thr Gly Thr Gly Ala Arg Trp Glu Leu Gln Pro Leu Ser Gly
 465 470 475 480

Arg Ala

<210> 103

<211> 1512

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(1512)

<223> Delta-4 desaturase

<400> 103
 atg tgc aac ggc aac ctc cca gca tcc acc gca cag ctc aag tcc acc 48
 Met Cys Asn Gly Asn Leu Pro Ala Ser Thr Ala Gln Leu Lys Ser Thr
 1 5 10 15

 tcg aag ccc cag cag caa cat gag cat cgc acc atc tcc aag tcc gag 96
 Ser Lys Pro Gln Gln Gln His Glu His Arg Thr Ile Ser Lys Ser Glu
 20 25 30

 ctc gcc caa cac aac acg ccc aaa tca gca tgg tgt gcc gtc cac tcc 144
 Leu Ala Gln His Asn Thr Pro Lys Ser Ala Trp Cys Ala Val His Ser
 35 40 45

 act ccc gcc acc gac cca tcc cac tcc aac aac aaa caa cac gca cac 192
 Thr Pro Ala Thr Asp Pro Ser His Ser Asn Asn Lys Gln His Ala His
 50 55 60

 cta gtc ctc gac att acc gac ttt gcg tcc cgc cat cca ggg gga gag 240
 Leu Val Leu Asp Ile Thr Asp Phe Ala Ser Arg His Pro Gly Gly Asp
 65 70 75 80

 ctc atc ctc ctc gct tcc ggc aaa gac gcc tcg gtg ctg ttt gaa aca 288
 Leu Ile Leu Leu Ala Ser Gly Lys Asp Ala Ser Val Leu Phe Glu Thr
 85 90 95

 tac cat cca cgt gga gtt ccg acg tct ctc att caa aag ctg cag att 336
 Tyr His Pro Arg Gly Val Pro Thr Ser Leu Ile Gln Lys Leu Gln Ile
 100 105 110

 gga gtg atg gag gag gag gcg ttt cgg gat tcg ttt tac agt tgg act 384
 Gly Val Met Glu Glu Glu Ala Phe Arg Asp Ser Phe Tyr Ser Trp Thr
 115 120 125

 gat tct gac ttt tat act gtg ttg aag agg agg gtt gtg gag cgg ttg 432
 Asp Ser Asp Phe Tyr Thr Val Leu Lys Arg Arg Val Val Glu Arg Leu
 130 135 140

 gag gag agg ggg ttg gac agg agg gga tcg aaa gag att tgg atc aag 480
 Glu Glu Arg Gly Leu Asp Arg Arg Gly Ser Lys Glu Ile Trp Ile Lys
 145 150 155 160

 gct ttg ttc ttg ttg gtt gga ttt tgg tac tgt ttg tac aag atg tat 528
 Ala Leu Phe Leu Leu Val Gly Phe Trp Tyr Cys Leu Tyr Lys Met Tyr
 165 170 175

 act acg tcg gat atc gat cag tac ggt att gcc att gcc tat tct att 576
 Thr Thr Ser Asp Ile Asp Gln Tyr Gly Ile Ala Ile Ala Tyr Ser Ile
 180 185 190

 gga atg gga acc ttt gcg gca ttc atc ggc acg tgt att caa cac gat 624
 Gly Met Gly Thr Phe Ala Ala Phe Ile Gly Thr Cys Ile Gln His Asp
 195 200 205

 gga aat cac ggt gca ttc gct cag aac aag tta ctc aac aag ttg gct 672
 Gly Asn His Gly Ala Phe Ala Gln Asn Lys Leu Leu Asn Lys Leu Ala
 210 215 220

 ggg tgg acg ttg gat atg att ggt gcg agt gcg ttt acg tgg gag ctt 720
 Gly Trp Thr Leu Asp Met Ile Gly Ala Ser Ala Phe Thr Trp Glu Leu
 225 230 235 240

 cag cac atg ctg ggg cat cat cca tat acg aat gtg ttg gat ggg gtg 768

186

Gln	His	Met	Leu	Gly 245	His	His	Pro	Tyr	Thr 250	Asn	Val	Leu	Asp	Gly 255	Val	
gag Glu	gag Glu	gag Glu	agg Arg 260	aag Lys	gag Glu	agg Arg	ggg Gly	gag Glu 265	gat Asp	gtt Val	gct Ala	ttg Leu	gaa Glu 270	gaa Glu	aag Lys	816
gat Asp	cag Gln	gat Asp 275	ttt Phe	gaa Glu	gtt Val	gcc Ala	aca Thr 280	tcc Ser	gga Gly	cga Arg	tta Leu	tat Tyr 285	cat His	att Ile	gat Asp	864
gcc Ala	aat Asn 290	gta Val	cgt Arg	tat Tyr	ggt Gly	tcg Ser 295	gta Val	tgg Trp	aat Asn	gtc Val	atg Met 300	agg Arg	ttt Phe	tgg Trp	gct Ala	912
atg Met 305	aag Lys	gtc Val	att Ile	acg Thr	atg Met 310	gga Gly	tat Tyr	atg Met	atg Met	gga Gly 315	tta Leu	cca Pro	atc Ile	tac Tyr	ttt Phe 320	960
cat His	gga Gly	gta Val	ctg Leu	agg Arg 325	gga Gly	gtt Val	gga Gly	ttg Leu	ttt Phe 330	gtt Val	att Ile	ggg Gly	cat His	ttg Leu 335	gcg Ala	1008
tgt Cys	gga Gly	gag Glu	ttg Leu 340	ttg Leu	gcg Ala	acg Thr	atg Met	ttt Phe 345	att Ile	gtg Val	aat Asn	cac His	gtc Val 350	att Ile	gag Glu	1056
ggt Gly	gtg Val	agt Ser 355	tat Tyr	gga Gly	acg Thr	aag Lys	gat Asp 360	ttg Leu	gtt Val	ggt Gly	ggt Gly	gcg Ala 365	agt Ser	cat His	gta Val	1104
gat Asp	gag Glu 370	aag Lys	aag Lys	att Ile	gtc Val	aag Lys 375	cca Pro	acg Thr	act Thr	gta Val	ttg Leu 380	gga Gly	gat Asp	aca Thr	cca Pro	1152
atg Met 385	gta Val	aag Lys	act Thr	cgc Arg	gag Glu 390	gag Glu	gca Ala	ttg Leu	aaa Lys	agc Ser 395	aac Asn	agc Ser	aat Asn	aac Asn	aac Asn 400	1200
aag Lys	aag Lys	aag Lys	gga Gly 405	gag Glu	aag Lys	aac Asn	tcg Ser	gta Val 410	cca Pro	tcc Ser	gtt Val	cca Pro	ttc Phe	aac Asn 415	gac Asp	1248
tgg Trp	gca Ala	gca Ala	gtc Val 420	caa Gln	tgc Cys	cag Gln	acc Thr	tcc Ser 425	gtg Val	aat Asn	tgg Trp	tct Ser	cca Pro 430	ggc Gly	tca Ser	1296
tgg Trp	ttc Phe	tgg Trp 435	aat Asn	cac His	ttt Phe	tct Ser	ggg Gly 440	gga Gly	ctc Leu	tct Ser	cat His	cag Gln 445	att Ile	gag Glu	cat His	1344
cac His	ttg Leu 450	ttc Phe	ccc Pro	agc Ser	att Ile	tgt Cys 455	cat His	aca Thr	aac Asn	tac Tyr	tgt Cys 460	cat His	atc Ile	cag Gln	gat Asp	1392
gtt Val 465	gtg Val	gag Glu	agt Ser	acg Thr	tgt Cys 470	gct Ala	gag Glu	tac Tyr	gga Gly	gtt Val 475	ccg Pro	tat Tyr	cag Gln	agt Ser	gag Glu 480	1440
agt Ser	aat Asn	ttg Leu	ttt Phe	gtt Val 485	gct Ala	tat Tyr	gga Gly	aag Lys	atg Met 490	att Ile	agt Ser	cat His	ttg Leu	aag Lys 495	ttt Phe	1488
ttg	ggt	aaa	gcc	aag	tgt	gag	tag									1512

Leu Gly Lys Ala Lys Cys Glu
500

<210> 104

<211> 503

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 104

Met Cys Asn Gly Asn Leu Pro Ala Ser Thr Ala Gln Leu Lys Ser Thr
1 5 10 15

Ser Lys Pro Gln Gln Gln His Glu His Arg Thr Ile Ser Lys Ser Glu
20 25 30

Leu Ala Gln His Asn Thr Pro Lys Ser Ala Trp Cys Ala Val His Ser
35 40 45

Thr Pro Ala Thr Asp Pro Ser His Ser Asn Asn Lys Gln His Ala His
50 55 60

Leu Val Leu Asp Ile Thr Asp Phe Ala Ser Arg His Pro Gly Gly Asp
65 70 75 80

Leu Ile Leu Leu Ala Ser Gly Lys Asp Ala Ser Val Leu Phe Glu Thr
85 90 95

Tyr His Pro Arg Gly Val Pro Thr Ser Leu Ile Gln Lys Leu Gln Ile
100 105 110

Gly Val Met Glu Glu Glu Ala Phe Arg Asp Ser Phe Tyr Ser Trp Thr
115 120 125

Asp Ser Asp Phe Tyr Thr Val Leu Lys Arg Arg Val Val Glu Arg Leu
130 135 140

Glu Glu Arg Gly Leu Asp Arg Arg Gly Ser Lys Glu Ile Trp Ile Lys
145 150 155 160

Ala Leu Phe Leu Leu Val Gly Phe Trp Tyr Cys Leu Tyr Lys Met Tyr
165 170 175

Thr Thr Ser Asp Ile Asp Gln Tyr Gly Ile Ala Ile Ala Tyr Ser Ile
180 185 190

Gly Met Gly Thr Phe Ala Ala Phe Ile Gly Thr Cys Ile Gln His Asp
 195 200 205
 Gly Asn His Gly Ala Phe Ala Gln Asn Lys Leu Leu Asn Lys Leu Ala
 210 215 220
 Gly Trp Thr Leu Asp Met Ile Gly Ala Ser Ala Phe Thr Trp Glu Leu
 225 230 235 240
 Gln His Met Leu Gly His His Pro Tyr Thr Asn Val Leu Asp Gly Val
 245 250 255
 Glu Glu Glu Arg Lys Glu Arg Gly Glu Asp Val Ala Leu Glu Glu Lys
 260 265 270
 Asp Gln Asp Phe Glu Val Ala Thr Ser Gly Arg Leu Tyr His Ile Asp
 275 280 285
 Ala Asn Val Arg Tyr Gly Ser Val Trp Asn Val Met Arg Phe Trp Ala
 290 295 300
 Met Lys Val Ile Thr Met Gly Tyr Met Met Gly Leu Pro Ile Tyr Phe
 305 310 315 320
 His Gly Val Leu Arg Gly Val Gly Leu Phe Val Ile Gly His Leu Ala
 325 330 335
 Cys Gly Glu Leu Leu Ala Thr Met Phe Ile Val Asn His Val Ile Glu
 340 345 350
 Gly Val Ser Tyr Gly Thr Lys Asp Leu Val Gly Gly Ala Ser His Val
 355 360 365
 Asp Glu Lys Lys Ile Val Lys Pro Thr Thr Val Leu Gly Asp Thr Pro
 370 375 380
 Met Val Lys Thr Arg Glu Glu Ala Leu Lys Ser Asn Ser Asn Asn Asn
 385 390 395 400
 Lys Lys Lys Gly Glu Lys Asn Ser Val Pro Ser Val Pro Phe Asn Asp
 405 410 415
 Trp Ala Ala Val Gln Cys Gln Thr Ser Val Asn Trp Ser Pro Gly Ser
 420 425 430
 Trp Phe Trp Asn His Phe Ser Gly Gly Leu Ser His Gln Ile Glu His
 435 440 445

His Leu Phe Pro Ser Ile Cys His Thr Asn Tyr Cys His Ile Gln Asp
 450 455 460

Val Val Glu Ser Thr Cys Ala Glu Tyr Gly Val Pro Tyr Gln Ser Glu
 465 470 475 480

Ser Asn Leu Phe Val Ala Tyr Gly Lys Met Ile Ser His Leu Lys Phe
 485 490 495

Leu Gly Lys Ala Lys Cys Glu
 500

<210> 105

<211> 1257

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(1257)

<223> Omega-3 desaturase

<400> 105

atg tac aga tta aca tcc acc ttc ctc atc gca ttg gca ttc tcc tcc 48
 Met Tyr Arg Leu Thr Ser Thr Phe Leu Ile Ala Leu Ala Phe Ser Ser
 1 5 10 15

tcc atc aat gcc ttc tct cca caa cgg cca cca cgt act atc acc aaa 96
 Ser Ile Asn Ala Phe Ser Pro Gln Arg Pro Pro Arg Thr Ile Thr Lys
 20 25 30

agt aaa gtc caa agc acc gtg cta ccc ata ccg acc aag gat gat ctg 144
 Ser Lys Val Gln Ser Thr Val Leu Pro Ile Pro Thr Lys Asp Asp Leu
 35 40 45

aac ttt ctc caa cca caa ctc gat gag aat gat ctc tac ctc gac gat 192
 Asn Phe Leu Gln Pro Gln Leu Asp Glu Asn Asp Leu Tyr Leu Asp Asp
 50 55 60

gtc aac act cca cca aga gca ggt acc atc atg aag atg ttg ccg aag 240
 Val Asn Thr Pro Pro Arg Ala Gly Thr Ile Met Lys Met Leu Pro Lys
 65 70 75 80

gaa acg ttc aac att gat aca gca act tca ttg ggt tac ttt ggt atg 288
 Glu Thr Phe Asn Ile Asp Thr Ala Thr Ser Leu Gly Tyr Phe Gly Met
 85 90 95

gat atg gca gcg gtt gta tcg tcc atg acg ttg cta aat gct att gta 336
 Asp Met Ala Ala Val Val Ser Ser Met Thr Leu Leu Asn Ala Ile Val
 100 105 110

act tcg gat cag tac cat gct ctt cca ctt cct ctc caa gca gca aca	384
Thr Ser Asp Gln Tyr His Ala Leu Pro Leu Pro Leu Gln Ala Ala Thr	
115 120 125	
gtg att ccc ttt cag cta ttg gct ggg ttc gcc atg tgg tgt atg tgg	432
Val Ile Pro Phe Gln Leu Leu Ala Gly Phe Ala Met Trp Cys Met Trp	
130 135 140	
tgc att gga cac gat gct gga cat tct act gtt tcg aag aca aag tgg	480
Cys Ile Gly His Asp Ala Gly His Ser Thr Val Ser Lys Thr Lys Trp	
145 150 155 160	
atc aac cga gtc gtt ggt gaa gtg gct cat tct gtt gtt tgt ctc acg	528
Ile Asn Arg Val Val Gly Glu Val Ala His Ser Val Val Cys Leu Thr	
165 170 175	
cog ttc gtg cct tgg cag atg tcg cat agg aaa cac cat ttg aat cac	576
Pro Phe Val Pro Trp Gln Met Ser His Arg Lys His His Leu Asn His	
180 185 190	
aat cat att gaa aag gac tac tct cat aag tgg tac agt cgc gac gag	624
Asn His Ile Glu Lys Asp Tyr Ser His Lys Trp Tyr Ser Arg Asp Glu	
195 200 205	
ttt gat gat atc cca caa ctc tat aag aca ttt ggc tac aac cca aga	672
Phe Asp Asp Ile Pro Gln Leu Tyr Lys Thr Phe Gly Tyr Asn Pro Arg	
210 215 220	
atg atg caa ctt cca ttc ctc tac ttc atg tat ctt gca ttg gga att	720
Met Met Gln Leu Pro Phe Leu Tyr Phe Met Tyr Leu Ala Leu Gly Ile	
225 230 235 240	
cca gat ggt ggg cat gtt gtg ttc tac gga aga atg tgg gaa gga gtg	768
Pro Asp Gly Gly His Val Val Phe Tyr Gly Arg Met Trp Glu Gly Val	
245 250 255	
tca ttg cag aag aag ttt gat gct gct att tct gtg gcc gta tca tgt	816
Ser Leu Gln Lys Lys Phe Asp Ala Ala Ile Ser Val Ala Val Ser Cys	
260 265 270	
gca act gct gga tcg ctt tgg atg aat atg ggt aca gca gac ttc acg	864
Ala Thr Ala Gly Ser Leu Trp Met Asn Met Gly Thr Ala Asp Phe Thr	
275 280 285	
gtg gta tgc atg gtt cct tgg cta gtt cta tcg tgg tgg ctc ttc atg	912
Val Val Cys Met Val Pro Trp Leu Val Leu Ser Trp Trp Leu Phe Met	
290 295 300	
gta aca tac ctt cag cat cat tca gaa gac gga aag cta tac act gat	960
Val Thr Tyr Leu Gln His His Ser Glu Asp Gly Lys Leu Tyr Thr Asp	
305 310 315 320	
gaa acg ttt aca ttt gaa aag gga gcc ttc gag acc gtg gat cgt tcg	1008
Glu Thr Phe Thr Phe Glu Lys Gly Ala Phe Glu Thr Val Asp Arg Ser	
325 330 335	
tac ggc aag ttg atc aac cga atg tcg cat cac atg atg gac ggt cac	1056
Tyr Gly Lys Leu Ile Asn Arg Met Ser His His Met Met Asp Gly His	
340 345 350	
gtg gtg cac cac ttg ttc ttt gaa cgt gta cct cac tac aga tta gag	1104
Val Val His His Leu Phe Phe Glu Arg Val Pro His Tyr Arg Leu Glu	
355 360 365	

gca gct acc gaa gct ctt gtg aaa gga atg gat gaa acg gga cag aaa 1152
 Ala Ala Thr Glu Ala Leu Val Lys Gly Met Asp Glu Thr Gly Gln Lys
 370 375 380

cat ttg tac aaa tac att gat act cct gat ttc aat gcc gag att gtc 1200
 His Leu Tyr Lys Tyr Ile Asp Thr Pro Asp Phe Asn Ala Glu Ile Val
 385 390 395 400

aac gga ttt cgc gac aat tgg ttc ctt gtt gaa gag gag aac atc aaa 1248
 Asn Gly Phe Arg Asp Asn Trp Phe Leu Val Glu Glu Glu Asn Ile Lys
 405 410 415

agg gag tag 1257
 Arg Glu

<210> 106

<211> 418

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 106

Met Tyr Arg Leu Thr Ser Thr Phe Leu Ile Ala Leu Ala Phe Ser Ser
 1 5 10 15

Ser Ile Asn Ala Phe Ser Pro Gln Arg Pro Pro Arg Thr Ile Thr Lys
 20 25 30

Ser Lys Val Gln Ser Thr Val Leu Pro Ile Pro Thr Lys Asp Asp Leu
 35 40 45

Asn Phe Leu Gln Pro Gln Leu Asp Glu Asn Asp Leu Tyr Leu Asp Asp
 50 55 60

Val Asn Thr Pro Pro Arg Ala Gly Thr Ile Met Lys Met Leu Pro Lys
 65 70 75 80

Glu Thr Phe Asn Ile Asp Thr Ala Thr Ser Leu Gly Tyr Phe Gly Met
 85 90 95

Asp Met Ala Ala Val Val Ser Ser Met Thr Leu Leu Asn Ala Ile Val
 100 105 110

Thr Ser Asp Gln Tyr His Ala Leu Pro Leu Pro Leu Gln Ala Ala Thr
 115 120 125

Val Ile Pro Phe Gln Leu Leu Ala Gly Phe Ala Met Trp Cys Met Trp
 130 135 140

Cys Ile Gly His Asp Ala Gly His Ser Thr Val Ser Lys Thr Lys Trp
 145 150 155 160

Ile Asn Arg Val Val Gly Glu Val Ala His Ser Val Val Cys Leu Thr
 165 170 175

Pro Phe Val Pro Trp Gln Met Ser His Arg Lys His His Leu Asn His
 180 185 190

Asn His Ile Glu Lys Asp Tyr Ser His Lys Trp Tyr Ser Arg Asp Glu
 195 200 205

Phe Asp Asp Ile Pro Gln Leu Tyr Lys Thr Phe Gly Tyr Asn Pro Arg
 210 215 220

Met Met Gln Leu Pro Phe Leu Tyr Phe Met Tyr Leu Ala Leu Gly Ile
 225 230 235 240

Pro Asp Gly Gly His Val Val Phe Tyr Gly Arg Met Trp Glu Gly Val
 245 250 255

Ser Leu Gln Lys Lys Phe Asp Ala Ala Ile Ser Val Ala Val Ser Cys
 260 265 270

Ala Thr Ala Gly Ser Leu Trp Met Asn Met Gly Thr Ala Asp Phe Thr
 275 280 285

Val Val Cys Met Val Pro Trp Leu Val Leu Ser Trp Trp Leu Phe Met
 290 295 300

Val Thr Tyr Leu Gln His His Ser Glu Asp Gly Lys Leu Tyr Thr Asp
 305 310 315 320

Glu Thr Phe Thr Phe Glu Lys Gly Ala Phe Glu Thr Val Asp Arg Ser
 325 330 335

Tyr Gly Lys Leu Ile Asn Arg Met Ser His His Met Met Asp Gly His
 340 345 350

Val Val His His Leu Phe Phe Glu Arg Val Pro His Tyr Arg Leu Glu
 355 360 365

Ala Ala Thr Glu Ala Leu Val Lys Gly Met Asp Glu Thr Gly Gln Lys
 370 375 380

His Leu Tyr Lys Tyr Ile Asp Thr Pro Asp Phe Asn Ala Glu Ile Val
 385 390 395 400

Asn Gly Phe Arg Asp Asn Trp Phe Leu Val Glu Glu Glu Asn Ile Lys
 405 410 415

Arg Glu

<210> 107

<211> 1086

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(1086)

<223> Delta-12 desaturase

<400> 107

atg cag gag ggg gtg cga aac att ccg aac gag tgc ttt gag acg gga 48
 Met Gln Glu Gly Val Arg Asn Ile Pro Asn Glu Cys Phe Glu Thr Gly
 1 5 10 15

cat ctt gaa aga ccc tgg cgt tcc ggc cgg tgt ggg cgc gat ccc ggt 96
 His Leu Glu Arg Pro Trp Arg Ser Gly Arg Cys Gly Arg Asp Pro Gly
 20 25 30

tcg aat tgg ggc gct ggc ttc cgc ttt ttt tcg ctc aag ggg ttt tgg 144
 Ser Asn Trp Gly Ala Gly Phe Arg Phe Phe Ser Leu Lys Gly Phe Trp
 35 40 45

tgg ccg gcg tgg tgg gcg tac gcg ttc gtg acg ggg acg gcg gcc act 192
 Trp Pro Ala Trp Trp Ala Tyr Ala Phe Val Thr Gly Thr Ala Ala Thr
 50 55 60

ggg tgt tgg gtc gcc gcg cac gag tgc ggg cac ggc gcg ttc agc gat 240
 Gly Cys Trp Val Ala Ala His Glu Cys Gly His Gly Ala Phe Ser Asp
 65 70 75 80

aac aag acg ttg caa gat gcg gtt gga tac gtg ttg cac tcg ttg ctc 288
 Asn Lys Thr Leu Gln Asp Ala Val Gly Tyr Val Leu His Ser Leu Leu
 85 90 95

ttg gtg ccg tac ttt tct tgg cag cga tca cac gcg gtg cat cac tcg 336
 Leu Val Pro Tyr Phe Ser Trp Gln Arg Ser His Ala Val His His Ser
 100 105 110

agg acg aat cac gtt ctt gag ggc gag acg cac gtg ccg gcg cgc ttg 384
 Arg Thr Asn His Val Leu Glu Gly Glu Thr His Val Pro Ala Arg Leu
 115 120 125

ggg acg gaa gac gcc aac gtc gtg ttc aag ctt cgc gaa ttg atc ggt 432

194

Gly Thr Glu Asp Ala Asn Val Val Phe Lys Leu Arg Glu Leu Ile Gly	
130 135 140	
gaa ggg ccg ttc acg ttt ttc aac ctc gtc ggc gtc ttc gcg ctc gga	480
Glu Gly Pro Phe Thr Phe Phe Asn Leu Val Gly Val Phe Ala Leu Gly	
145 150 155 160	
tgg ccg att tac ttg ctc acc ggc gcg agc ggc gga ccg gtg cgc ggt	528
Trp Pro Ile Tyr Leu Leu Thr Gly Ala Ser Gly Gly Pro Val Arg Gly	
165 170 175	
aac acg aac cac ttc tta ccc ttc atg ggc gag aaa ggt aag cac gcg	576
Asn Thr Asn His Phe Leu Pro Phe Met Gly Glu Lys Gly Lys His Ala	
180 185 190	
ctg ttc ccg ggt aag tgg gcg aag aag gtg tgg cag tct gac atc ggc	624
Leu Phe Pro Gly Lys Trp Ala Lys Lys Val Trp Gln Ser Asp Ile Gly	
195 200 205	
gtt gtt gcc gtc ctg ggc gcg ctc gcg gct tgg gcg gcg cac agc ggg	672
Val Val Ala Val Leu Gly Ala Leu Ala Ala Trp Ala Ala His Ser Gly	
210 215 220	
att gcc aca gtg atg gca ctc tac gtc ggc ccg tac atg gtg acc aac	720
Ile Ala Thr Val Met Ala Leu Tyr Val Gly Pro Tyr Met Val Thr Asn	
225 230 235 240	
ttt tgg ctc gtc ttg tac acg tgg tta cag cac acc gac gtt gac gtg	768
Phe Trp Leu Val Leu Tyr Thr Trp Leu Gln His Thr Asp Val Asp Val	
245 250 255	
ccg cac ttc gag ggc gac gat tgg aac ttg gtc aag ggg gca ttc atg	816
Pro His Phe Glu Gly Asp Asp Trp Asn Leu Val Lys Gly Ala Phe Met	
260 265 270	
acg atc gat cgc ccg tac ggc cca gtt ttt gat ttc ttg cac cac cgc	864
Thr Ile Asp Arg Pro Tyr Gly Pro Val Phe Asp Phe Leu His His Arg	
275 280 285	
atc ggc agc acg cac gtc gcg cac cac atc aac aca cca ttc ccg cat	912
Ile Gly Ser Thr His Val Ala His His Ile Asn Thr Pro Phe Pro His	
290 295 300	
tac aag gct caa atg gcg acg gat gcg cta aag gag gcg tat ccc gac	960
Tyr Lys Ala Gln Met Ala Thr Asp Ala Leu Lys Glu Ala Tyr Pro Asp	
305 310 315 320	
ctc tac ctt tac gat cca act ccg atc gcg acc gct acg tgg cgc gtg	1008
Leu Tyr Leu Tyr Asp Pro Thr Pro Ile Ala Thr Ala Thr Trp Arg Val	
325 330 335	
ggg agc aag tgc atc gcc gtc gtg aag aag gga gac gaa tgg gtg ttc	1056
Gly Ser Lys Cys Ile Ala Val Val Lys Lys Gly Asp Glu Trp Val Phe	
340 345 350	
acg gat aag caa ctc ccg gtc gcg gcg tga	1086
Thr Asp Lys Gln Leu Pro Val Ala Ala	
355 360	
<210> 108	
<211> 361	

<212> PRT

<213> *Ostreococcus tauri*

<400> 108

Met Gln Glu Gly Val Arg Asn Ile Pro Asn Glu Cys Phe Glu Thr Gly
 1 5 10 15

His Leu Glu Arg Pro Trp Arg Ser Gly Arg Cys Gly Arg Asp Pro Gly
 20 25 30

Ser Asn Trp Gly Ala Gly Phe Arg Phe Phe Ser Leu Lys Gly Phe Trp
 35 40 45

Trp Pro Ala Trp Trp Ala Tyr Ala Phe Val Thr Gly Thr Ala Ala Thr
 50 55 60

Gly Cys Trp Val Ala Ala His Glu Cys Gly His Gly Ala Phe Ser Asp
 65 70 75 80

Asn Lys Thr Leu Gln Asp Ala Val Gly Tyr Val Leu His Ser Leu Leu
 85 90 95

Leu Val Pro Tyr Phe Ser Trp Gln Arg Ser His Ala Val His His Ser
 100 105 110

Arg Thr Asn His Val Leu Glu Gly Glu Thr His Val Pro Ala Arg Leu
 115 120 125

Gly Thr Glu Asp Ala Asn Val Val Phe Lys Leu Arg Glu Leu Ile Gly
 130 135 140

Glu Gly Pro Phe Thr Phe Phe Asn Leu Val Gly Val Phe Ala Leu Gly
 145 150 155 160

Trp Pro Ile Tyr Leu Leu Thr Gly Ala Ser Gly Gly Pro Val Arg Gly
 165 170 175

Asn Thr Asn His Phe Leu Pro Phe Met Gly Glu Lys Gly Lys His Ala
 180 185 190

Leu Phe Pro Gly Lys Trp Ala Lys Lys Val Trp Gln Ser Asp Ile Gly
 195 200 205

Val Val Ala Val Leu Gly Ala Leu Ala Ala Trp Ala Ala His Ser Gly
 210 215 220

196

Ile Ala Thr Val Met Ala Leu Tyr Val Gly Pro Tyr Met Val Thr Asn
225 230 235 240

Phe Trp Leu Val Leu Tyr Thr Trp Leu Gln His Thr Asp Val Asp Val
245 250 255

Pro His Phe Glu Gly Asp Asp Trp Asn Leu Val Lys Gly Ala Phe Met
260 265 270

Thr Ile Asp Arg Pro Tyr Gly Pro Val Phe Asp Phe Leu His His Arg
275 280 285

Ile Gly Ser Thr His Val Ala His His Ile Asn Thr Pro Phe Pro His
290 295 300

Tyr Lys Ala Gln Met Ala Thr Asp Ala Leu Lys Glu Ala Tyr Pro Asp
305 310 315 320

Leu Tyr Leu Tyr Asp Pro Thr Pro Ile Ala Thr Ala Thr Trp Arg Val
325 330 335

Gly Ser Lys Cys Ile Ala Val Val Lys Lys Gly Asp Glu Trp Val Phe
340 345 350

Thr Asp Lys Gln Leu Pro Val Ala Ala
355 360

<210> 109

<211> 1305

<212> DNA

<213> *Thalassiosira pseudonana*

<220>

<221> CDS

<222> (1)..(1305)

<223> Delta-12 desaturase

<400> 109

atg gga aag gga gga aga tca gta acc cgc gct caa aca gca gaa aag 48
Met Gly Lys Gly Arg Ser Val Thr Arg Ala Gln Thr Ala Glu Lys
1 5 10 15

tca gca cac acc atc caa acc ttc acc gac ggc cga tgg gtc tcc ccc 96
Ser Ala His Thr Ile Gln Thr Phe Thr Asp Gly Arg Trp Val Ser Pro
20 25 30

tac aac ccc ctc gca aaa gat gca cct gaa ctc ccc tcc aag ggt gaa	144
Tyr Asn Pro Leu Ala Lys Asp Ala Pro Glu Leu Pro Ser Lys Gly Glu	
35 40 45	
atc aag gcg gtc atc ccc aaa gag tgc ttc gaa cga agc tac ctc cac	192
Ile Lys Ala Val Ile Pro Lys Glu Cys Phe Glu Arg Ser Tyr Leu His	
50 55 60	
tcc atg tac ttc gtc ctc cgt gac acc gtc atg gcc gtg gcc tgc gcc	240
Ser Met Tyr Phe Val Leu Arg Asp Thr Val Met Ala Val Ala Cys Ala	
65 70 75 80	
tac atc gcc cac tca acg ctc tcc acc gat att ccc tcc gag tta ctg	288
Tyr Ile Ala His Ser Thr Leu Ser Thr Asp Ile Pro Ser Glu Leu Leu	
85 90 95	
agc gtg gac gca ctc aaa tgg ttc ctc gga tgg aac acc tac gcc ttt	336
Ser Val Asp Ala Leu Lys Trp Phe Leu Gly Trp Asn Thr Tyr Ala Phe	
100 105 110	
tgg atg ggg tgc att ctc acc gga cac tgg gtc cta gcc cat gaa tgt	384
Trp Met Gly Cys Ile Leu Thr Gly His Trp Val Leu Ala His Glu Cys	
115 120 125	
gga cat ggt gca ttc tct ccc tct cag acg ttt aat gac ttt tgg ggg	432
Gly His Gly Ala Phe Ser Pro Ser Gln Thr Phe Asn Asp Phe Trp Gly	
130 135 140	
ttc att atg cat cag gcg gtg ttg gtt ccg tat ttc gcc tgg cag tac	480
Phe Ile Met His Gln Ala Val Leu Val Pro Tyr Phe Ala Trp Gln Tyr	
145 150 155 160	
tct cat gcg aag cat cat cga cgt acc aac aac att atg gat ggg gag	528
Ser His Ala Lys His His Arg Arg Thr Asn Asn Ile Met Asp Gly Glu	
165 170 175	
agc cat gtg ccc aat atc gcc aag gaa atg gga ttg aac gag aag aat	576
Ser His Val Pro Asn Ile Ala Lys Glu Met Gly Leu Asn Glu Lys Asn	
180 185 190	
gag cgc agt gga gga tat gcc gcc att cat gag gct att gga gat gga	624
Glu Arg Ser Gly Gly Tyr Ala Ala Ile His Glu Ala Ile Gly Asp Gly	
195 200 205	
ccc ttt gcg atg ttt caa atc ttt gct cac ttg gtg atc ggg tgg cct	672
Pro Phe Ala Met Phe Gln Ile Phe Ala His Leu Val Ile Gly Trp Pro	
210 215 220	
att tac ttg atg gga ttt gct tcc act gga cgt ctc ggt cag gat ggg	720
Ile Tyr Leu Met Gly Phe Ala Ser Thr Gly Arg Leu Gly Gln Asp Gly	
225 230 235 240	
aag gaa ctt cag gct gga gag atc atc gac cat tac cgt cct tgg agt	768
Lys Glu Leu Gln Ala Gly Glu Ile Ile Asp His Tyr Arg Pro Trp Ser	
245 250 255	
aag atg ttc ccc acc aag ttg cga ttc aaa att gct ctt tcg aca ctt	816
Lys Met Phe Pro Thr Lys Leu Arg Phe Lys Ile Ala Leu Ser Thr Leu	
260 265 270	
gga gtg att gcc gcc tgg gtt ggg ttg tac ttt gct gca caa gag tat	864
Gly Val Ile Ala Ala Trp Val Gly Leu Tyr Phe Ala Ala Gln Glu Tyr	
275 280 285	

gga gtc ttg ccc gtg gtt ctt tgg tac att ggc cca ctc atg tgg aat 912
 Gly Val Leu Pro Val Val Leu Trp Tyr Ile Gly Pro Leu Met Trp Asn
 290 295 300

cag gcg tgg ctt gtg ctc tac act tgg ctt cag cac aat gat ccc tcc 960
 Gln Ala Trp Leu Val Leu Tyr Thr Trp Leu Gln His Asn Asp Pro Ser
 305 310 315 320

gtg cct caa tat gga agt gac gaa tgg aca tgg gtc aag gga gct ttg 1008
 Val Pro Gln Tyr Gly Ser Asp Glu Trp Thr Trp Val Lys Gly Ala Leu
 325 330 335

tcg acg att gat cgc ccg tat ggt atc ttt gac ttc ttc cat cac aag 1056
 Ser Thr Ile Asp Arg Pro Tyr Gly Ile Phe Asp Phe Phe His His Lys
 340 345 350

att gga agc act cac gta gct cat cat ttg ttc cac gag atg cca ttt 1104
 Ile Gly Ser Thr His Val Ala His His Leu Phe His Glu Met Pro Phe
 355 360 365

tac aag gcg gat gtg gct act gcg tcg atc aag ggt ttc ttg gag ccg 1152
 Tyr Lys Ala Asp Val Ala Thr Ala Ser Ile Lys Gly Phe Leu Glu Pro
 370 375 380

aag gga ctt tac aac tat gat cca acg cct tgg tat gtg gcc atg tgg 1200
 Lys Gly Leu Tyr Asn Tyr Asp Pro Thr Pro Trp Tyr Val Ala Met Trp
 385 390 395 400

agg gtg gcc aag act tgt cat tat att gag gat gtg gat gga gtt cag 1248
 Arg Val Ala Lys Thr Cys His Tyr Ile Glu Asp Val Asp Gly Val Gln
 405 410 415

tat tat aag agt ttg gag gat gtg cct ttg aag aag gat gcc aag aag 1296
 Tyr Tyr Lys Ser Leu Glu Asp Val Pro Leu Lys Lys Asp Ala Lys Lys
 420 425 430

tct gat tag 1305
 Ser Asp

<210> 110

<211> 434

<212> PRT

<213> *Thalassiosira pseudonana*

<400> 110

Met Gly Lys Gly Gly Arg Ser Val Thr Arg Ala Gln Thr Ala Glu Lys
1 5 10 15

Ser Ala His Thr Ile Gln Thr Phe Thr Asp Gly Arg Trp Val Ser Pro
20 25 30

Tyr Asn Pro Leu Ala Lys Asp Ala Pro Glu Leu Pro Ser Lys Gly Glu
35 40 45

Ile Lys Ala Val Ile Pro Lys Glu Cys Phe Glu Arg Ser Tyr Leu His
 50 55 60

Ser Met Tyr Phe Val Leu Arg Asp Thr Val Met Ala Val Ala Cys Ala
 65 70 75 80

Tyr Ile Ala His Ser Thr Leu Ser Thr Asp Ile Pro Ser Glu Leu Leu
 85 90 95

Ser Val Asp Ala Leu Lys Trp Phe Leu Gly Trp Asn Thr Tyr Ala Phe
 100 105 110

Trp Met Gly Cys Ile Leu Thr Gly His Trp Val Leu Ala His Glu Cys
 115 120 125

Gly His Gly Ala Phe Ser Pro Ser Gln Thr Phe Asn Asp Phe Trp Gly
 130 135 140

Phe Ile Met His Gln Ala Val Leu Val Pro Tyr Phe Ala Trp Gln Tyr
 145 150 155 160

Ser His Ala Lys His His Arg Arg Thr Asn Asn Ile Met Asp Gly Glu
 165 170 175

Ser His Val Pro Asn Ile Ala Lys Glu Met Gly Leu Asn Glu Lys Asn
 180 185 190

Glu Arg Ser Gly Gly Tyr Ala Ala Ile His Glu Ala Ile Gly Asp Gly
 195 200 205

Pro Phe Ala Met Phe Gln Ile Phe Ala His Leu Val Ile Gly Trp Pro
 210 215 220

Ile Tyr Leu Met Gly Phe Ala Ser Thr Gly Arg Leu Gly Gln Asp Gly
 225 230 235 240

Lys Glu Leu Gln Ala Gly Glu Ile Ile Asp His Tyr Arg Pro Trp Ser
 245 250 255

Lys Met Phe Pro Thr Lys Leu Arg Phe Lys Ile Ala Leu Ser Thr Leu
 260 265 270

Gly Val Ile Ala Ala Trp Val Gly Leu Tyr Phe Ala Ala Gln Glu Tyr
 275 280 285

Gly Val Leu Pro Val Val Leu Trp Tyr Ile Gly Pro Leu Met Trp Asn
 290 295 300

Gln Ala Trp Leu Val Leu Tyr Thr Trp Leu Gln His Asn Asp Pro Ser
305 310 315 320

Val Pro Gln Tyr Gly Ser Asp Glu Trp Thr Trp Val Lys Gly Ala Leu
325 330 335

Ser Thr Ile Asp Arg Pro Tyr Gly Ile Phe Asp Phe Phe His His Lys
340 345 350

Ile Gly Ser Thr His Val Ala His His Leu Phe His Glu Met Pro Phe
355 360 365

Tyr Lys Ala Asp Val Ala Thr Ala Ser Ile Lys Gly Phe Leu Glu Pro
370 375 380

Lys Gly Leu Tyr Asn Tyr Asp Pro Thr Pro Trp Tyr Val Ala Met Trp
385 390 395 400

Arg Val Ala Lys Thr Cys His Tyr Ile Glu Asp Val Asp Gly Val Gln
405 410 415

Tyr Tyr Lys Ser Leu Glu Asp Val Pro Leu Lys Lys Asp Ala Lys Lys
420 425 430

Ser Asp

<210> 111

<211> 879

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(879)

<223> Delta-6 elongase

<400> 111

atg agt ggc tta cgt gca ccc aac ttt tta cac aga ttc tgg aca aag 48
Met Ser Gly Leu Arg Ala Pro Asn Phe Leu His Arg Phe Trp Thr Lys
1 5 10 15

tgg gac tac gcg att tcc aaa gtc gtc ttc acg tgt gcc gac agt ttt 96

201

Trp	Asp	Tyr	Ala	Ile	Ser	Lys	Val	Val	Phe	Thr	Cys	Ala	Asp	Ser	Phe	
			20					25					30			
cag	tgg	gac	atc	ggg	cca	gtg	agt	tcg	agt	acg	gcg	cat	tta	ccc	gcc	144
Gln	Trp	Asp	Ile	Gly	Pro	Val	Ser	Ser	Ser	Thr	Ala	His	Leu	Pro	Ala	
		35				40					45					
att	gaa	tcc	cct	acc	cca	ctg	gtg	act	agc	ctc	ttg	ttc	tac	tta	gtc	192
Ile	Glu	Ser	Pro	Thr	Pro	Leu	Val	Thr	Ser	Leu	Leu	Phe	Tyr	Leu	Val	
	50					55				60						
aca	gtt	ttc	ttg	tgg	tat	ggg	cgt	tta	acc	agg	agt	tca	gac	aag	aaa	240
Thr	Val	Phe	Leu	Trp	Tyr	Gly	Arg	Leu	Thr	Arg	Ser	Ser	Asp	Lys	Lys	
65					70				75						80	
att	aga	gag	cct	acg	tgg	tta	aga	aga	ttc	ata	ata	tgt	cat	aat	gcg	288
Ile	Arg	Glu	Pro	Thr	Trp	Leu	Arg	Arg	Phe	Ile	Ile	Cys	His	Asn	Ala	
			85						90					95		
ttc	ttg	ata	gtc	ctc	agt	ctt	tac	atg	tgc	ctt	ggg	tgt	gtg	gcc	caa	336
Phe	Leu	Ile	Val	Leu	Ser	Leu	Tyr	Met	Cys	Leu	Gly	Cys	Val	Ala	Gln	
			100					105					110			
gcg	tat	cag	aat	gga	tat	act	tta	tgg	ggg	aat	gaa	ttc	aag	gcc	acg	384
Ala	Tyr	Gln	Asn	Gly	Tyr	Thr	Leu	Trp	Gly	Asn	Glu	Phe	Lys	Ala	Thr	
		115					120					125				
gaa	act	cag	ctt	gct	ctc	tac	att	tac	att	ttt	tac	gta	agt	aaa	ata	432
Glu	Thr	Gln	Leu	Ala	Leu	Tyr	Ile	Tyr	Ile	Phe	Tyr	Val	Ser	Lys	Ile	
	130					135					140					
tac	gag	ttt	gta	gat	act	tac	att	atg	ctt	ctc	aag	aat	aac	ttg	cgg	480
Tyr	Glu	Phe	Val	Asp	Thr	Tyr	Ile	Met	Leu	Lys	Asn	Asn	Leu	Arg		
145					150				155					160		
caa	gta	agt	ttc	cta	cac	att	tat	cac	cac	agc	acg	att	tcc	ttt	att	528
Gln	Val	Ser	Phe	Leu	His	Ile	Tyr	His	His	Ser	Thr	Ile	Ser	Phe	Ile	
				165				170						175		
tgg	tgg	atc	att	gct	cgg	agg	gct	ccg	ggg	ggg	gat	gct	tac	ttc	agc	576
Trp	Trp	Ile	Ile	Ala	Arg	Arg	Ala	Pro	Gly	Gly	Asp	Ala	Tyr	Phe	Ser	
		180						185					190			
gcg	gcc	ttg	aac	tca	tgg	gta	cac	gtg	tgc	atg	tac	acc	tat	tat	cta	624
Ala	Ala	Leu	Asn	Ser	Trp	Val	His	Val	Cys	Met	Tyr	Thr	Tyr	Tyr	Leu	
		195					200					205				
tta	tca	acc	ctt	att	gga	aaa	gaa	gat	cct	aag	cgt	tcc	aac	tac	ctt	672
Leu	Ser	Thr	Leu	Ile	Gly	Lys	Glu	Asp	Pro	Lys	Arg	Ser	Asn	Tyr	Leu	
	210					215					220					
tgg	tgg	ggg	cgc	cac	cta	acg	caa	atg	cag	atg	ctt	cag	ttt	ttc	ttc	720
Trp	Trp	Gly	Arg	His	Leu	Thr	Gln	Met	Gln	Met	Leu	Gln	Phe	Phe	Phe	
225					230				235					240		
aac	gta	ctt	caa	gcg	ttg	tac	tgc	gct	tcg	ttc	tct	acg	tat	ccc	aag	768
Asn	Val	Leu	Gln	Ala	Leu	Tyr	Cys	Ala	Ser	Phe	Ser	Thr	Tyr	Pro	Lys	
				245				250						255		
ttt	ttg	tcc	aaa	att	ctg	ctc	gtc	tat	atg	atg	agc	ctt	ctc	ggc	ttg	816
Phe	Leu	Ser	Lys	Ile	Leu	Leu	Val	Tyr	Met	Met	Ser	Leu	Leu	Gly	Leu	
			260					265					270			
ttt	ggg	cat	ttc	tac	tat	tcc	aag	cac	ata	gca	gca	gct	aag	ctc	cag	864

202

Phe Gly His Phe Tyr Tyr Ser Lys His Ile Ala Ala Ala Lys Leu Gln
 275 280 285

aaa aaa cag cag tga
 Lys Lys Gln Gln
 290

879

<210> 112

<211> 292

<212> PRT

<213> *Ostreococcus tauri*

<400> 112

Met Ser Gly Leu Arg Ala Pro Asn Phe Leu His Arg Phe Trp Thr Lys
 1 5 10 15

Trp Asp Tyr Ala Ile Ser Lys Val Val Phe Thr Cys Ala Asp Ser Phe
 20 25 30

Gln Trp Asp Ile Gly Pro Val Ser Ser Ser Thr Ala His Leu Pro Ala
 35 40 45

Ile Glu Ser Pro Thr Pro Leu Val Thr Ser Leu Leu Phe Tyr Leu Val
 50 55 60

Thr Val Phe Leu Trp Tyr Gly Arg Leu Thr Arg Ser Ser Asp Lys Lys
 65 70 75 80

Ile Arg Glu Pro Thr Trp Leu Arg Arg Phe Ile Ile Cys His Asn Ala
 85 90 95

Phe Leu Ile Val Leu Ser Leu Tyr Met Cys Leu Gly Cys Val Ala Gln
 100 105 110

Ala Tyr Gln Asn Gly Tyr Thr Leu Trp Gly Asn Glu Phe Lys Ala Thr
 115 120 125

Glu Thr Gln Leu Ala Leu Tyr Ile Tyr Ile Phe Tyr Val Ser Lys Ile
 130 135 140

Tyr Glu Phe Val Asp Thr Tyr Ile Met Leu Leu Lys Asn Asn Leu Arg
 145 150 155 160

Gln Val Ser Phe Leu His Ile Tyr His His Ser Thr Ile Ser Phe Ile
 165 170 175

203

Trp Trp Ile Ile Ala Arg Arg Ala Pro Gly Gly Asp Ala Tyr Phe Ser
 180 185 190

Ala Ala Leu Asn Ser Trp Val His Val Cys Met Tyr Thr Tyr Tyr Leu
 195 200 205

Leu Ser Thr Leu Ile Gly Lys Glu Asp Pro Lys Arg Ser Asn Tyr Leu
 210 215 220

Trp Trp Gly Arg His Leu Thr Gln Met Gln Met Leu Gln Phe Phe Phe
 225 230 235 240

Asn Val Leu Gln Ala Leu Tyr Cys Ala Ser Phe Ser Thr Tyr Pro Lys
 245 250 255

Phe Leu Ser Lys Ile Leu Leu Val Tyr Met Met Ser Leu Leu Gly Leu
 260 265 270

Phe Gly His Phe Tyr Tyr Ser Lys His Ile Ala Ala Ala Lys Leu Gln
 275 280 285

Lys Lys Gln Gln
 290

<210> 113

<211> 903

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(903)

<223> Delta-5 elongase

<400> 113

atg agc gcc tcc ggt gcg ctg ctg ccc gcg atc gcg ttc gcc gcg tac 48
 Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Phe Ala Ala Tyr
 1 5 10 15

gcg tac gcg acg tac gcc tac gcc ttt gag tgg tcg cac gcg aat ggc 96
 Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30

atc gac aac gtc gac gcg cgc gag tgg atc ggt gcg ctg tcg ttg agg 144
 Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45

ctc	ccg	gcg	atc	gcg	acg	acg	atg	tac	ctg	ttg	ttc	tgc	ctg	gtc	gga	192
Leu	Pro	Ala	Ile	Ala	Thr	Thr	Met	Tyr	Leu	Leu	Phe	Cys	Leu	Val	Gly	
	50						55					60				
ccg	agg	ttg	atg	gcg	aag	cgc	gag	gcg	ttc	gac	ccg	aag	ggg	ttc	atg	240
Pro	Arg	Leu	Met	Ala	Lys	Arg	Glu	Ala	Phe	Asp	Pro	Lys	Gly	Phe	Met	
65					70					75					80	
ctg	gcg	tac	aat	gcg	tat	cag	acg	gcg	ttc	aac	gtc	gtc	gtg	ctc	ggg	288
Leu	Ala	Tyr	Asn	Ala	Tyr	Gln	Thr	Ala	Phe	Asn	Val	Val	Val	Leu	Gly	
			85						90					95		
atg	ttc	gcg	cga	gag	atc	tcg	ggg	ctg	ggg	cag	ccc	gtg	tgg	ggg	tca	336
Met	Phe	Ala	Arg	Glu	Ile	Ser	Gly	Leu	Gly	Gln	Pro	Val	Trp	Gly	Ser	
			100					105					110			
acc	atg	ccg	tgg	agc	gat	aga	aaa	tcg	ttt	aag	atc	ctc	ctc	ggg	gtg	384
Thr	Met	Pro	Trp	Ser	Asp	Arg	Lys	Ser	Phe	Lys	Ile	Leu	Leu	Gly	Val	
		115					120					125				
tgg	ttg	cac	tac	aac	aac	aaa	tat	ttg	gag	cta	ttg	gac	act	gtg	ttc	432
Trp	Leu	His	Tyr	Asn	Asn	Lys	Tyr	Leu	Glu	Leu	Leu	Asp	Thr	Val	Phe	
	130					135					140					
atg	gtt	gcg	cgc	aag	aag	acg	aag	cag	ttg	agc	ttc	ttg	cac	gtt	tat	480
Met	Val	Ala	Arg	Lys	Lys	Thr	Lys	Gln	Leu	Ser	Phe	Leu	His	Val	Tyr	
145					150					155					160	
cat	cac	gcc	ctg	ttg	atc	tgg	gcg	tgg	tgg	ttg	gtg	tgt	cac	ttg	atg	528
His	His	Ala	Leu	Leu	Ile	Trp	Ala	Trp	Trp	Leu	Val	Cys	His	Leu	Met	
			165					170						175		
gcc	acg	aac	gat	tgt	atc	gat	gcc	tac	ttc	ggc	gcg	gcg	tgc	aac	tcg	576
Ala	Thr	Asn	Asp	Cys	Ile	Asp	Ala	Tyr	Phe	Gly	Ala	Ala	Cys	Asn	Ser	
			180					185					190			
ttc	att	cac	atc	gtg	atg	tac	tcg	tat	tat	ctc	atg	tcg	gcg	ctc	ggc	624
Phe	Ile	His	Ile	Val	Met	Tyr	Ser	Tyr	Tyr	Leu	Met	Ser	Ala	Leu	Gly	
		195					200					205				
att	cga	tgc	ccg	tgg	aag	cga	tac	atc	acc	cag	gct	caa	atg	ctc	caa	672
Ile	Arg	Cys	Pro	Trp	Lys	Arg	Tyr	Ile	Thr	Gln	Ala	Gln	Met	Leu	Gln	
	210					215					220					
ttc	gtc	att	gtc	ttc	gcg	cac	gcc	gtg	ttc	gtg	ctg	cgt	cag	aag	cac	720
Phe	Val	Ile	Val	Phe	Ala	His	Ala	Val	Phe	Val	Leu	Arg	Gln	Lys	His	
225					230					235					240	
tgc	ccg	gtc	acc	ctt	cct	tgg	gcg	caa	atg	ttc	gtc	atg	acg	aac	atg	768
Cys	Pro	Val	Thr	Leu	Pro	Trp	Ala	Gln	Met	Phe	Val	Met	Thr	Asn	Met	
				245					250					255		
ctc	gtg	ctc	ttc	ggg	aac	ttc	tac	ctc	aag	gcg	tac	tcg	aac	aag	tcg	816
Leu	Val	Leu	Phe	Gly	Asn	Phe	Tyr	Leu	Lys	Ala	Tyr	Ser	Asn	Lys	Ser	
			260					265					270			
cgc	ggc	gac	ggc	gcg	agt	tcc	gtg	aaa	cca	gcc	gag	acc	acg	cgc	gcg	864
Arg	Gly	Asp	Gly	Ala	Ser	Ser	Val	Lys	Pro	Ala	Glu	Thr	Thr	Arg	Ala	
		275					280					285				
ccc	agc	gtg	cga	cgc	acg	cga	tct	cga	aaa	att	gac	taa				903
Pro	Ser	Val	Arg	Arg	Thr	Arg	Ser	Arg	Lys	Ile	Asp					
	290					295					300					

<210> 114

<211> 300

<212> PRT

<213> *Ostreococcus tauri*

<400> 114

Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Phe Ala Ala Tyr
 1 5 10 15

Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30

Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45

Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly
 50 55 60

Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met
 65 70 75 80

Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly
 85 90 95

Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser
 100 105 110

Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val
 115 120 125

Trp Leu His Tyr Asn Asn Lys Tyr Leu Glu Leu Leu Asp Thr Val Phe
 130 135 140

Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr
 145 150 155 160

His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met
 165 170 175

Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser
 180 185 190

Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
 195 200 205

Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
 210 215 220

Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240

Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255

Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270

Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285

Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 115

<211> 13

<212> PRT

<213> Consensus

<220>

<221> MISC_FEATURE

<222> (1)..(13)

<223> Xaa in the sequence at positions 2, 3, 4, 6, 7, 8 and 9 has the
 meaning given in Table A.

<400> 115

Asn Xaa Xaa Xaa His Xaa Xaa Met Tyr Xaa Tyr Tyr Xaa
 1 5 10

<210> 116

<211> 10

<212> PRT

<213> Consensus

<220>

<221> MISC_FEATURE

<222> (1)..(10)

<223> Xaa at positions 3, 4, 5 and 6 the sequence has the meaning given in Table A.

<400> 116

His	His	Xaa	Xaa	Xaa	Xaa	Trp	Ala	Trp	Trp
1				5					10

<210> 117

<211> 909

<212> DNA

<213> *Xenopus laevis*

<220>

<221> CDS

<222> (1)..(909)

<223> Delta-5 elongase

<400> 117

atg	gcc	ttc	aag	gag	ctc	aca	tca	agg	gca	gtg	ctc	ctg	tat	gat	gaa	48
Met	Ala	Phe	Lys	Glu	Leu	Thr	Ser	Arg	Ala	Val	Leu	Leu	Tyr	Asp	Glu	
1				5					10					15		

tgg	att	aaa	gat	gct	gat	cct	agg	gtt	gaa	gac	tgg	cca	ctc	atg	tcc	96
Trp	Ile	Lys	Asp	Ala	Asp	Pro	Arg	Val	Glu	Asp	Trp	Pro	Leu	Met	Ser	
			20					25					30			

tct	cct	atc	cta	caa	acc	atc	atc	atc	ggc	gct	tac	atc	tac	ttt	gtc	144
Ser	Pro	Ile	Leu	Gln	Thr	Ile	Ile	Ile	Gly	Ala	Tyr	Ile	Tyr	Phe	Val	
			35					40					45			

aca	tca	ttg	ggc	cca	agg	atc	atg	gag	aac	agg	aag	ccg	ttt	gct	ctg	192
Thr	Ser	Leu	Gly	Pro	Arg	Ile	Met	Glu	Asn	Arg	Lys	Pro	Phe	Ala	Leu	
		50					55					60				

aag	gag	atc	atg	gca	tgt	tac	aac	tta	ttc	atg	ggt	ctg	ttt	tct	gtg	240
Lys	Glu	Ile	Met	Ala	Cys	Tyr	Asn	Leu	Phe	Met	Val	Leu	Phe	Ser	Val	
65					70					75				80		

tac	atg	tgc	tat	gag	ttt	ctc	atg	tcg	ggc	tgg	gct	act	gga	tat	tcc	288
Tyr	Met	Cys	Tyr	Glu	Phe	Leu	Met	Ser	Gly	Trp	Ala	Thr	Gly	Tyr	Ser	
				85					90					95		

ttt	aga	tgt	gac	att	gtt	gac	tac	tct	cag	tca	cct	cag	gcg	tta	cgg	336
Phe	Arg	Cys	Asp	Ile	Val	Asp	Tyr	Ser	Gln	Ser	Pro	Gln	Ala	Leu	Arg	

100	105	110	
atg gcc tgg acc tgc tgg ctc ttc tat ttt tca aag ttc att gaa tta			384
Met Ala Trp Thr Cys Trp Leu Phe Tyr Phe Ser Lys Phe Ile Glu Leu			
115	120	125	
tta gac act gtt ttc ttt gtg ctg cgt aag aag aac agc cag att aca			432
Leu Asp Thr Val Phe Phe Val Leu Arg Lys Lys Asn Ser Gln Ile Thr			
130	135	140	
ttc ctg cac gtc tat cac cac tcc att atg cct tgg acg tgg tgg ttt			480
Phe Leu His Val Tyr His His Ser Ile Met Pro Trp Thr Trp Trp Phe			
145	150	155	160
gga gtc aaa ttt gct cca ggt ggt ttg ggc aca ttc cat gca ctg gtg			528
Gly Val Lys Phe Ala Pro Gly Gly Leu Gly Thr Phe His Ala Leu Val			
165	170	175	
aac tgt gtg gtc cat gtt atc atg tac agc tac tac ggc ctg tca gcc			576
Asn Cys Val Val His Val Ile Met Tyr Ser Tyr Tyr Gly Leu Ser Ala			
180	185	190	
ttg ggg cct gcc tac cag aag tac ctg tgg tgg aaa aag tac atg acg			624
Leu Gly Pro Ala Tyr Gln Lys Tyr Leu Trp Trp Lys Lys Tyr Met Thr			
195	200	205	
tct atc caa ctg acc cag ttc ttg atg gtt act ttt cac atc ggc cag			672
Ser Ile Gln Leu Thr Gln Phe Leu Met Val Thr Phe His Ile Gly Gln			
210	215	220	
ttc ttc ttc atg gag aat tgc ccg tac cag tat ccc gtc ttc ttg tat			720
Phe Phe Phe Met Glu Asn Cys Pro Tyr Gln Tyr Pro Val Phe Leu Tyr			
225	230	235	240
gtc att tgg ctg tac ggg ttc gtt ttc tta atc ttg ttc ctc aac ttc			768
Val Ile Trp Leu Tyr Gly Phe Val Phe Leu Ile Leu Phe Leu Asn Phe			
245	250	255	
tgg ttc cac gct tac atc aaa gga cag agg ctg ccg aaa gcc gtc caa			816
Trp Phe His Ala Tyr Ile Lys Gly Gln Arg Leu Pro Lys Ala Val Gln			
260	265	270	
aat ggc cac tgc aag aac aac aac aac caa gaa aac act tgg tgc aag			864
Asn Gly His Cys Lys Asn Asn Asn Asn Gln Glu Asn Thr Trp Cys Lys			
275	280	285	
aac aaa aac cag aaa aac ggt gca ttg aaa agc aaa aac cat tga			909
Asn Lys Asn Gln Lys Asn Gly Ala Leu Lys Ser Lys Asn His			
290	295	300	

<210> 118

<211> 302

<212> PRT

<213> *Xenopus laevis*

<400> 118

Met Ala Phe Lys Glu Leu Thr Ser Arg Ala Val Leu Leu Tyr Asp Glu

1	5	10	15
Trp Ile Lys Asp Ala Asp Pro Arg Val Glu Asp Trp Pro Leu Met Ser	20	25	30
Ser Pro Ile Leu Gln Thr Ile Ile Ile Gly Ala Tyr Ile Tyr Phe Val	35	40	45
Thr Ser Leu Gly Pro Arg Ile Met Glu Asn Arg Lys Pro Phe Ala Leu	50	55	60
Lys Glu Ile Met Ala Cys Tyr Asn Leu Phe Met Val Leu Phe Ser Val	65	70	75
Tyr Met Cys Tyr Glu Phe Leu Met Ser Gly Trp Ala Thr Gly Tyr Ser	85	90	95
Phe Arg Cys Asp Ile Val Asp Tyr Ser Gln Ser Pro Gln Ala Leu Arg	100	105	110
Met Ala Trp Thr Cys Trp Leu Phe Tyr Phe Ser Lys Phe Ile Glu Leu	115	120	125
Leu Asp Thr Val Phe Phe Val Leu Arg Lys Lys Asn Ser Gln Ile Thr	130	135	140
Phe Leu His Val Tyr His His Ser Ile Met Pro Trp Thr Trp Trp Phe	145	150	155
Gly Val Lys Phe Ala Pro Gly Gly Leu Gly Thr Phe His Ala Leu Val	165	170	175
Asn Cys Val Val His Val Ile Met Tyr Ser Tyr Tyr Gly Leu Ser Ala	180	185	190
Leu Gly Pro Ala Tyr Gln Lys Tyr Leu Trp Trp Lys Lys Tyr Met Thr	195	200	205
Ser Ile Gln Leu Thr Gln Phe Leu Met Val Thr Phe His Ile Gly Gln	210	215	220
Phe Phe Phe Met Glu Asn Cys Pro Tyr Gln Tyr Pro Val Phe Leu Tyr	225	230	235
Val Ile Trp Leu Tyr Gly Phe Val Phe Leu Ile Leu Phe Leu Asn Phe	245	250	255
Trp Phe His Ala Tyr Ile Lys Gly Gln Arg Leu Pro Lys Ala Val Gln			

260

265

270

Asn Gly His Cys Lys Asn Asn Asn Asn Gln Glu Asn Thr Trp Cys Lys
 275 280 285

Asn Lys Asn Gln Lys Asn Gly Ala Leu Lys Ser Lys Asn His
 290 295 300

<210> 119

<211> 870

<212> DNA

<213> Ciona intestinalis

<220>

<221> CDS

<222> (1)..(870)

<223> Delta-5 elongase

<400> 119

atg gac gta ctt cat cgt ttc tta gga ttc tac gaa tgg acg ctg act 48
 Met Asp Val Leu His Arg Phe Leu Gly Phe Tyr Glu Trp Thr Leu Thr
 1 5 10 15

ttc gcg gac ccc cga gtg gca aaa tgg cct tta ata gaa aac ccc ctt 96
 Phe Ala Asp Pro Arg Val Ala Lys Trp Pro Leu Ile Glu Asn Pro Leu
 20 25 30

cct aca att gct att gtg ttg ctg tac ctg gcg ttt gtt ctg tat att 144
 Pro Thr Ile Ala Ile Val Leu Tyr Leu Ala Phe Val Leu Tyr Ile
 35 40 45

ggg ccg cgt ttt atg cga aaa aga gca cca gtt gac ttt ggt tta ttc 192
 Gly Pro Arg Phe Met Arg Lys Arg Ala Pro Val Asp Phe Gly Leu Phe
 50 55 60

ctc cct gga tat aac ttt gct ttg gtt gca tta aat tat tat atc ctg 240
 Leu Pro Gly Tyr Asn Phe Ala Leu Val Ala Leu Asn Tyr Tyr Ile Leu
 65 70 75 80

caa gaa gtg gtc act ggg agt tat ggg gct ggg tat gat ttg gtt tgc 288
 Gln Glu Val Val Thr Gly Ser Tyr Gly Ala Gly Tyr Asp Leu Val Cys
 85 90 95

aca cca ctt cga agt gat tcc tac gat ccc aat gaa atg aag gtt gca 336
 Thr Pro Leu Arg Ser Asp Ser Tyr Asp Pro Asn Glu Met Lys Val Ala
 100 105 110

aac gct gta tgg tgg tat tat gta tcc aag ata ata gag ttg ttt gat 384
 Asn Ala Val Trp Trp Tyr Tyr Val Ser Lys Ile Ile Glu Leu Phe Asp
 115 120 125

211

act gtg ttg ttc act cta cgc aaa cga gac cga caa gta act ttc ctt 432
 Thr Val Leu Phe Thr Leu Arg Lys Arg Asp Arg Gln Val Thr Phe Leu
 130 135 140

cat gtt tat cac cat tct acc atg ccc ctg ttg tgg tgg att ggg gca 480
 His Val Tyr His His Ser Thr Met Pro Leu Leu Trp Trp Ile Gly Ala
 145 150 155 160

aag tgg gtg cct ggt ggg caa tca ttt gtt ggc atc ata ctg aac tcc 528
 Lys Trp Val Pro Gly Gly Gln Ser Phe Val Gly Ile Ile Leu Asn Ser
 165 170 175

agt gtt cat gtt atc atg tat acg tac tat gga ttg tca gcc ttg ggg 576
 Ser Val His Val Ile Met Tyr Thr Tyr Tyr Gly Leu Ser Ala Leu Gly
 180 185 190

cct cac atg cag aag ttt cta tgg tgg aag aaa tat atc aca atg ttg 624
 Pro His Met Gln Lys Phe Leu Trp Trp Lys Lys Tyr Ile Thr Met Leu
 195 200 205

caa ctg gtt caa ttt gtt ctt gcc atc tac cat act gct cga tca ttg 672
 Gln Leu Val Gln Phe Val Leu Ala Ile Tyr His Thr Ala Arg Ser Leu
 210 215 220

tac gtt aaa tgt ccc tcg cct gtt tgg atg cac tgg gca ctt atc ttg 720
 Tyr Val Lys Cys Pro Ser Pro Val Trp Met His Trp Ala Leu Ile Leu
 225 230 235 240

tac gct ttc tca ttc att ttg ctt ttc tca aac ttc tac atg cat gcc 768
 Tyr Ala Phe Ser Phe Ile Leu Leu Phe Ser Asn Phe Tyr Met His Ala
 245 250 255

tat atc aag aaa tca aga aaa ggg aaa gag aat ggc agt cga gga aaa 816
 Tyr Ile Lys Lys Ser Arg Lys Gly Lys Glu Asn Gly Ser Arg Gly Lys
 260 265 270

ggt ggt gta agt aat gga aag gaa aag ctg cac gct aat ggt aaa acc 864
 Gly Gly Val Ser Asn Gly Lys Glu Lys Leu His Ala Asn Gly Lys Thr
 275 280 285

gat taa 870
 Asp

<210> 120

<211> 289

<212> PRT

<213> Ciona intestinalis

<400> 120

Met Asp Val Leu His Arg Phe Leu Gly Phe Tyr Glu Trp Thr Leu Thr
 1 5 10 15

Phe Ala Asp Pro Arg Val Ala Lys Trp Pro Leu Ile Glu Asn Pro Leu
 20 25 30

Pro Thr Ile Ala Ile Val Leu Leu Tyr Leu Ala Phe Val Leu Tyr Ile
 35 40 45

Gly Pro Arg Phe Met Arg Lys Arg Ala Pro Val Asp Phe Gly Leu Phe
 50 55 60

Leu Pro Gly Tyr Asn Phe Ala Leu Val Ala Leu Asn Tyr Tyr Ile Leu
 65 70 75 80

Gln Glu Val Val Thr Gly Ser Tyr Gly Ala Gly Tyr Asp Leu Val Cys
 85 90 95

Thr Pro Leu Arg Ser Asp Ser Tyr Asp Pro Asn Glu Met Lys Val Ala
 100 105 110

Asn Ala Val Trp Trp Tyr Tyr Val Ser Lys Ile Ile Glu Leu Phe Asp
 115 120 125

Thr Val Leu Phe Thr Leu Arg Lys Arg Asp Arg Gln Val Thr Phe Leu
 130 135 140

His Val Tyr His His Ser Thr Met Pro Leu Leu Trp Trp Ile Gly Ala
 145 150 155 160

Lys Trp Val Pro Gly Gly Gln Ser Phe Val Gly Ile Ile Leu Asn Ser
 165 170 175

Ser Val His Val Ile Met Tyr Thr Tyr Tyr Gly Leu Ser Ala Leu Gly
 180 185 190

Pro His Met Gln Lys Phe Leu Trp Trp Lys Lys Tyr Ile Thr Met Leu
 195 200 205

Gln Leu Val Gln Phe Val Leu Ala Ile Tyr His Thr Ala Arg Ser Leu
 210 215 220

Tyr Val Lys Cys Pro Ser Pro Val Trp Met His Trp Ala Leu Ile Leu
 225 230 235 240

Tyr Ala Phe Ser Phe Ile Leu Leu Phe Ser Asn Phe Tyr Met His Ala
 245 250 255

Tyr Ile Lys Lys Ser Arg Lys Gly Lys Glu Asn Gly Ser Arg Gly Lys
 260 265 270

Gly Gly Val Ser Asn Gly Lys Glu Lys Leu His Ala Asn Gly Lys Thr
 275 280 285

Asp

<210> 121

<211> 30

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(30)

<223>

<400> 121

aggatccatg gccttcaagg agctcacatc

30

<210> 122

<211> 35

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(35)

<223>

<400> 122

cctcgagtca atggtttttg cttttcaatg caccg

35

<210> 123

<211> 25

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(25)

<223>

<400> 123

taagcttatg gacgtacttc atcgt

25

<210> 124

<211> 26

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(26)

<223>

<400> 124

tcagatcttt aatcggtttt accatt

26

<210> 125

<211> 34

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(34)

<223>

<400> 125

gcggccgcac catggccttc aaggagctca catc

34

<210> 126

<211> 38

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(38)

<223>

<400> 126
gcggccgcct tcaatggttt ttgcttttca atgcaccg

38

<210> 127

<211> 29

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(29)

<223>

<400> 127
gcggccgcac catggacgta cttcatcgt

29

<210> 128

<211> 27

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(27)

<223>

<400> 128
gcggccgctt taatcggttt taccatt

27

<210> 129

<211> 60

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(60)

<223>

<400> 129
gtcgaccgc ggactagtgg gccctctaga cccgggggat ccgatctgc tggctatgaa

60

<210> 130

<211> 60

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(60)

<223>

<400> 130
gtcgaccgc ggactagtgg gccctctaga cccgggggat ccgatctgc tggctatgaa

60

<210> 131

<211> 789

<212> DNA

<213> *Euglena gracilis*

<220>

<221> CDS

<222> (1)..(789)

<223> Delta-5 elongase

<400> 131

atg	ctg	ggg	gcc	atc	gcg	gac	gtc	gtg	ctc	cgg	ggg	ccc	gcc	gca	ttc	48
Met	Leu	Gly	Ala	Ile	Ala	Asp	Val	Val	Leu	Arg	Gly	Pro	Ala	Ala	Phe	
1			5						10					15		

cac	tgg	gac	cct	gcc	acc	acc	ccg	ctc	gca	tcg	atc	gtc	agc	ccc	tgt	96
His	Trp	Asp	Pro	Ala	Thr	Thr	Pro	Leu	Ala	Ser	Ile	Val	Ser	Pro	Cys	
			20					25					30			

gtg	gcc	tcc	gtg	gcg	tac	ctg	ggg	gcc	atc	ggg	ctg	ctg	aag	cgc	cgc	144
Val	Ala	Ser	Val	Ala	Tyr	Leu	Gly	Ala	Ile	Gly	Leu	Leu	Lys	Arg	Arg	
		35					40					45				

act	gga	ccg	gag	gtc	cgc	tcc	aag	ccc	ttc	gag	ctg	cta	cac	aac	ggg	192
Thr	Gly	Pro	Glu	Val	Arg	Ser	Lys	Pro	Phe	Glu	Leu	Leu	His	Asn	Gly	
	50					55					60					

ctg	ctg	gtg	ggc	tgg	tcc	ctc	gtg	gtg	ctg	ctc	ggg	acg	ctg	tac	ggc	240
Leu	Leu	Val	Gly	Trp	Ser	Leu	Val	Val	Leu	Leu	Gly	Thr	Leu	Tyr	Gly	
65					70					75					80	

gcg	ttc	cag	cgc	gtg	cag	gag	gac	ggc	cgg	ggg	gtg	cag	gcc	ctc	ctg	288
Ala	Phe	Gln	Arg	Val	Gln	Glu	Asp	Gly	Arg	Gly	Val	Gln	Ala	Leu	Leu	
				85					90					95		

tgc	acc	cag	cgg	cca	cca	tct	cag	atc	tgg	gac	ggc	ccg	gtg	ggg	tac	336
Cys	Thr	Gln	Arg	Pro	Pro	Ser	Gln	Ile	Trp	Asp	Gly	Pro	Val	Gly	Tyr	
			100					105						110		

ttc	acg	tac	ctc	ttc	tac	ctc	gcg	aag	tac	tgg	gag	ctg	gcg	gac	act	384
Phe	Thr	Tyr	Leu	Phe	Tyr	Leu	Ala	Lys	Tyr	Trp	Glu	Leu	Ala	Asp	Thr	
		115					120					125				

gtc	atc	ctc	gcc	ctc	cgc	cag	aag	ccc	acc	atc	ccc	ctc	cac	gtc	tac	432
Val	Ile	Leu	Ala	Leu	Arg	Gln	Lys	Pro	Thr	Ile	Pro	Leu	His	Val	Tyr	
	130					135					140					

cat	cac	gcc	gtc	atg	ctg	ttc	atc	gtg	tgg	tcg	tgg	ttc	gcg	cac	ccc	480
His	His	Ala	Val	Met	Leu	Phe	Ile	Val	Trp	Ser	Trp	Phe	Ala	His	Pro	
145					150					155					160	

tgg	ctc	gag	ggg	agc	tgg	tgg	tgc	tcc	ctg	gtc	aac	tct	ttc	atc	cac	528
Trp	Leu	Glu	Gly	Ser	Trp	Trp	Cys	Ser	Leu	Val	Asn	Ser	Phe	Ile	His	
			165						170					175		

acg	gtg	atg	tac	tcg	tac	tac	acc	ctg	acg	gtg	gtt	ggc	atc	aac	cct	576
Thr	Val	Met	Tyr	Ser	Tyr	Tyr	Thr	Leu	Thr	Val	Val	Gly	Ile	Asn	Pro	
			180					185					190			

tgg	tgg	aag	aag	tgg	atg	acc	acc	atg	cag	atc	atc	cag	ttc	atc	acg	624
Trp	Trp	Lys	Lys	Trp	Met	Thr	Thr	Met	Gln	Ile	Ile	Gln	Phe	Ile	Thr	
		195					200					205				

ggc	tgc	gtg	tac	gtc	atg	gcg	ttc	ttc	ggc	cta	tat	tat	gcc	ggg	gcg	672
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----

218

Gly Cys Val Tyr Val Met Ala Phe Phe Gly Leu Tyr Tyr Ala Gly Ala
 210 215 220
 ggc tgc acc tcc aac gtg tac act gcc tgg ttc tcg atg ggg gtc aac 720
 Gly Cys Thr Ser Asn Val Tyr Thr Ala Trp Phe Ser Met Gly Val Asn
 225 230 235 240
 ctc agc ttt ctg tgg ctc ttc gct ctt ttc ttc cgc cgg tca tac agc 768
 Leu Ser Phe Leu Trp Leu Phe Ala Leu Phe Phe Arg Arg Ser Tyr Ser
 245 250 255
 aaa cct agc cgg aag gag tag 789
 Lys Pro Ser Arg Lys Glu
 260

<210> 132

<211> 262

<212> PRT

<213> Euglena gracilis

<400> 132

Met Leu Gly Ala Ile Ala Asp Val Val Leu Arg Gly Pro Ala Ala Phe
 1 5 10 15
 His Trp Asp Pro Ala Thr Thr Pro Leu Ala Ser Ile Val Ser Pro Cys
 20 25 30
 Val Ala Ser Val Ala Tyr Leu Gly Ala Ile Gly Leu Leu Lys Arg Arg
 35 40 45
 Thr Gly Pro Glu Val Arg Ser Lys Pro Phe Glu Leu Leu His Asn Gly
 50 55 60
 Leu Leu Val Gly Trp Ser Leu Val Val Leu Leu Gly Thr Leu Tyr Gly
 65 70 75 80
 Ala Phe Gln Arg Val Gln Glu Asp Gly Arg Gly Val Gln Ala Leu Leu
 85 90 95
 Cys Thr Gln Arg Pro Pro Ser Gln Ile Trp Asp Gly Pro Val Gly Tyr
 100 105 110
 Phe Thr Tyr Leu Phe Tyr Leu Ala Lys Tyr Trp Glu Leu Ala Asp Thr
 115 120 125
 Val Ile Leu Ala Leu Arg Gln Lys Pro Thr Ile Pro Leu His Val Tyr
 130 135 140

219

His His Ala Val Met Leu Phe Ile Val Trp Ser Trp Phe Ala His Pro
 145 150 155 160

Trp Leu Glu Gly Ser Trp Trp Cys Ser Leu Val Asn Ser Phe Ile His
 165 170 175

Thr Val Met Tyr Ser Tyr Tyr Thr Leu Thr Val Val Gly Ile Asn Pro
 180 185 190

Trp Trp Lys Lys Trp Met Thr Thr Met Gln Ile Ile Gln Phe Ile Thr
 195 200 205

Gly Cys Val Tyr Val Met Ala Phe Phe Gly Leu Tyr Tyr Ala Gly Ala
 210 215 220

Gly Cys Thr Ser Asn Val Tyr Thr Ala Trp Phe Ser Met Gly Val Asn
 225 230 235 240

Leu Ser Phe Leu Trp Leu Phe Ala Leu Phe Phe Arg Arg Ser Tyr Ser
 245 250 255

Lys Pro Ser Arg Lys Glu
 260

<210> 133

<211> 789

<212> DNA

<213> *Euglena gracilis*

<220>

<221> CDS

<222> (1)..(789)

<223> Delta-5 elongase

<400> 133

atg ctg ggg gcc atc gcg gac gtc gtg ctc cgg ggg ccc gcc gca ttc 48
 Met Leu Gly Ala Ile Ala Asp Val Val Leu Arg Gly Pro Ala Ala Phe
 1 5 10 15

cac tgg gac cct gcc acc acc ccg ctc gca tcg atc gtc agc ccc tgt 96
 His Trp Asp Pro Ala Thr Thr Pro Leu Ala Ser Ile Val Ser Pro Cys
 20 25 30

gtg gcc tcc gtg gcg tac ctg ggg gcc atc ggg ctg ctg aag cgc cgc 144
 Val Ala Ser Val Ala Tyr Leu Gly Ala Ile Gly Leu Leu Lys Arg Arg
 35 40 45

act gga ccg gag gtc cgc tcc aag ccc ttc gag ctg cta cac aac ggg	192
Thr Gly Pro Glu Val Arg Ser Lys Pro Phe Glu Leu Leu His Asn Gly	
50 55 60	
ctg ctg gtg ggc tgg tcc ctc gtg gtg ctg ctc ggg acg ctg tac ggc	240
Leu Leu Val Gly Trp Ser Leu Val Val Leu Leu Gly Thr Leu Tyr Gly	
65 70 75 80	
gcg tac cag cgc gtg cag gag gac ggc cgg ggg gtg cag gcc ctg ctg	288
Ala Tyr Gln Arg Val Gln Glu Asp Gly Arg Gly Val Gln Ala Leu Leu	
85 90 95	
tgc acc cag cgg cca cca tct cag atc tgg gac ggc cgg gtg ggg tac	336
Cys Thr Gln Arg Pro Pro Ser Gln Ile Trp Asp Gly Pro Val Gly Tyr	
100 105 110	
ttc acg tac ctt ttc tac ctc gcg aag tac tgg gag ctg gtg gac act	384
Phe Thr Tyr Leu Phe Tyr Leu Ala Lys Tyr Trp Glu Leu Val Asp Thr	
115 120 125	
gtc atc ctc gcc ctc cgc cag aag ccc acc atc ccc ctc cac gtc tac	432
Val Ile Leu Ala Leu Arg Gln Lys Pro Thr Ile Pro Leu His Val Tyr	
130 135 140	
cat cac gcc gtc atg ctg ttc att gtg tgg tcg tgg ttc gcg cac ccc	480
His His Ala Val Met Leu Phe Ile Val Trp Ser Trp Phe Ala His Pro	
145 150 155 160	
tgg ctc gag ggg agc tgg tgg tgc tcc ctg gtc aac tct ttc atc cac	528
Trp Leu Glu Gly Ser Trp Trp Cys Ser Leu Val Asn Ser Phe Ile His	
165 170 175	
acg gtg atg tac tcg tat tac acc ctg acg gtg gtt ggc atc aac cct	576
Thr Val Met Tyr Ser Tyr Tyr Thr Leu Thr Val Val Gly Ile Asn Pro	
180 185 190	
tgg tgg aag aag tgg atg acc acc atg cag atc atc cag ttc atc acg	624
Trp Trp Lys Lys Trp Met Thr Thr Met Gln Ile Ile Gln Phe Ile Thr	
195 200 205	
ggc tgc gtg tac gtc acg gcg ttc ttc ggc cta tac tat gcc ggg gcg	672
Gly Cys Val Tyr Val Thr Ala Phe Phe Gly Leu Tyr Tyr Ala Gly Ala	
210 215 220	
ggc tgc acc tcc aac gtg tac act gcc tgg ttc tcg atg ggg gtc aac	720
Gly Cys Thr Ser Asn Val Tyr Thr Ala Trp Phe Ser Met Gly Val Asn	
225 230 235 240	
ctc agc ttt ctg tgg ctc ttc gct ctt ttc ttc cgc cgg tcg tac agc	768
Leu Ser Phe Leu Trp Leu Phe Ala Leu Phe Phe Arg Arg Ser Tyr Ser	
245 250 255	
aaa cct agc cgg aag gag tag	789
Lys Pro Ser Arg Lys Glu	
260	

<210> 134

<211> 262

<212> PRT

<213> *Euglena gracilis*

<400> 134

Met Leu Gly Ala Ile Ala Asp Val Val Leu Arg Gly Pro Ala Ala Phe
 1 5 10 15

His Trp Asp Pro Ala Thr Thr Pro Leu Ala Ser Ile Val Ser Pro Cys
 20 25 30

Val Ala Ser Val Ala Tyr Leu Gly Ala Ile Gly Leu Leu Lys Arg Arg
 35 40 45

Thr Gly Pro Glu Val Arg Ser Lys Pro Phe Glu Leu Leu His Asn Gly
 50 55 60

Leu Leu Val Gly Trp Ser Leu Val Val Leu Leu Gly Thr Leu Tyr Gly
 65 70 75 80

Ala Tyr Gln Arg Val Gln Glu Asp Gly Arg Gly Val Gln Ala Leu Leu
 85 90 95

Cys Thr Gln Arg Pro Pro Ser Gln Ile Trp Asp Gly Pro Val Gly Tyr
 100 105 110

Phe Thr Tyr Leu Phe Tyr Leu Ala Lys Tyr Trp Glu Leu Val Asp Thr
 115 120 125

Val Ile Leu Ala Leu Arg Gln Lys Pro Thr Ile Pro Leu His Val Tyr
 130 135 140

His His Ala Val Met Leu Phe Ile Val Trp Ser Trp Phe Ala His Pro
 145 150 155 160

Trp Leu Glu Gly Ser Trp Trp Cys Ser Leu Val Asn Ser Phe Ile His
 165 170 175

Thr Val Met Tyr Ser Tyr Tyr Thr Leu Thr Val Val Gly Ile Asn Pro
 180 185 190

Trp Trp Lys Lys Trp Met Thr Thr Met Gln Ile Ile Gln Phe Ile Thr
 195 200 205

Gly Cys Val Tyr Val Thr Ala Phe Phe Gly Leu Tyr Tyr Ala Gly Ala
 210 215 220

Gly Cys Thr Ser Asn Val Tyr Thr Ala Trp Phe Ser Met Gly Val Asn
 225 230 235 240

Leu Ser Phe Leu Trp Leu Phe Ala Leu Phe Phe Arg Arg Ser Tyr Ser
 245 250 255

Lys Pro Ser Arg Lys Glu
 260

<210> 135

<211> 897

<212> DNA

<213> Arabidopsis thaliana

<220>

<221> CDS

<222> (1)..(897)

<223> Delta-5 elongase

<400> 135

atg gca tct gtt tac tcc acc cta acc tac tgg ctc gtc cac cac ccc 48
 Met Ala Ser Val Tyr Ser Thr Leu Thr Tyr Trp Leu Val His His Pro
 1 5 10 15

tac att gcc aac ttc acg tgg acc gaa ggt gaa aca cta ggc tcc acc 96
 Tyr Ile Ala Asn Phe Thr Trp Thr Glu Gly Glu Thr Leu Gly Ser Thr
 20 25 30

gtt ttc ttt gtc ttt gtc gtc gtc tcc ctt tac ctc tcc gcc aca ttc 144
 Val Phe Phe Val Phe Val Val Ser Leu Tyr Leu Ser Ala Thr Phe
 35 40 45

ctc ctc cga tac acc gtc gat tca ctc ccc aca ctc ggt ccc cgc att 192
 Leu Leu Arg Tyr Thr Val Asp Ser Leu Pro Thr Leu Gly Pro Arg Ile
 50 55 60

ctc aaa cca atc aca gcc gtt cac agc ctc att ctc ttc ctc ctc tcc 240
 Leu Lys Pro Ile Thr Ala Val His Ser Leu Ile Leu Phe Leu Leu Ser
 65 70 75 80

tta acc atg gcc gtt ggt tgc act ctc tcc cta atc tct tcc tcg gac 288
 Leu Thr Met Ala Val Gly Cys Thr Leu Ser Leu Ile Ser Ser Ser Asp
 85 90 95

ccg aag gcg cgt ctc ttc gac gcc gtt tgt ttc ccc ctc gac gtg aaa 336
 Pro Lys Ala Arg Leu Phe Asp Ala Val Cys Phe Pro Leu Asp Val Lys
 100 105 110

cct aag gga ccg ctt ttc ttt tgg gct caa gtc ttt tac ctc tcg aag 384
 Pro Lys Gly Pro Leu Phe Phe Trp Ala Gln Val Phe Tyr Leu Ser Lys
 115 120 125

atc ctt gag ttc gta gac aca ctt ctc atc ata ctc aac aaa tca atc 432

223

Ile Leu Glu Phe Val Asp Thr Leu Leu Ile Ile Leu Asn Lys Ser Ile
 130 135 140

caa cgg ctc tcg ttc ctc cac gtc tac cac cac gca acg gtt gtg att 480
 Gln Arg Leu Ser Phe Leu His Val Tyr His His Ala Thr Val Val Ile
 145 150 155 160

ttg tgc tac ctc tgg tta cga aca cgt caa tcg atg ttt cct gtt ggg 528
 Leu Cys Tyr Leu Trp Leu Arg Thr Arg Gln Ser Met Phe Pro Val Gly
 165 170 175

ctc gtg ttg aac tcg acg gtc cat gtg att atg tac ggg tac tat ttc 576
 Leu Val Leu Asn Ser Thr Val His Val Ile Met Tyr Gly Tyr Tyr Phe
 180 185 190

ctc tgc gct atc gga tcg agg ccc aag tgg aag aag ttg gtg acg aat 624
 Leu Cys Ala Ile Gly Ser Arg Pro Lys Trp Lys Lys Leu Val Thr Asn
 195 200 205

ttt caa atg gtt cag ttt gct ttc ggc atg ggg tta gga gcc gct tgg 672
 Phe Gln Met Val Gln Phe Ala Phe Gly Met Gly Leu Gly Ala Ala Trp
 210 215 220

atg ctc cca gag cat tat ttc ggg tcg ggt tgc gcc ggg att tgg aca 720
 Met Leu Pro Glu His Tyr Phe Gly Ser Gly Cys Ala Gly Ile Trp Thr
 225 230 235 240

gtt tat ttc aat ggt gtg ttt act gct tct cta ttg gct ctc ttc tac 768
 Val Tyr Phe Asn Gly Val Phe Thr Ala Ser Leu Leu Ala Leu Phe Tyr
 245 250 255

aac ttc cac tcc aag aac tat gag aag act aca acg tcg cct ttg tat 816
 Asn Phe His Ser Lys Asn Tyr Glu Lys Thr Thr Thr Ser Pro Leu Tyr
 260 265 270

aag atc gaa tcc ttt ata ttt att cac gga gag agg tgg gca aat aaa 864
 Lys Ile Glu Ser Phe Ile Phe Ile His Gly Glu Arg Trp Ala Asn Lys
 275 280 285

gcg att aca tta ttt tcc aag aaa aac gat taa 897
 Ala Ile Thr Leu Phe Ser Lys Lys Asn Asp
 290 295

<210> 136

<211> 298

<212> PRT

<213> Arabidopsis thaliana

<400> 136

Met Ala Ser Val Tyr Ser Thr Leu Thr Tyr Trp Leu Val His His Pro
 1 5 10 15

Tyr Ile Ala Asn Phe Thr Trp Thr Glu Gly Glu Thr Leu Gly Ser Thr
 20 25 30

224

Val Phe Phe Val Phe Val Val Val Ser Leu Tyr Leu Ser Ala Thr Phe
 35 40 45

Leu Leu Arg Tyr Thr Val Asp Ser Leu Pro Thr Leu Gly Pro Arg Ile
 50 55 60

Leu Lys Pro Ile Thr Ala Val His Ser Leu Ile Leu Phe Leu Leu Ser
 65 70 75 80

Leu Thr Met Ala Val Gly Cys Thr Leu Ser Leu Ile Ser Ser Ser Asp
 85 90 95

Pro Lys Ala Arg Leu Phe Asp Ala Val Cys Phe Pro Leu Asp Val Lys
 100 105 110

Pro Lys Gly Pro Leu Phe Phe Trp Ala Gln Val Phe Tyr Leu Ser Lys
 115 120 125

Ile Leu Glu Phe Val Asp Thr Leu Leu Ile Ile Leu Asn Lys Ser Ile
 130 135 140

Gln Arg Leu Ser Phe Leu His Val Tyr His His Ala Thr Val Val Ile
 145 150 155 160

Leu Cys Tyr Leu Trp Leu Arg Thr Arg Gln Ser Met Phe Pro Val Gly
 165 170 175

Leu Val Leu Asn Ser Thr Val His Val Ile Met Tyr Gly Tyr Tyr Phe
 180 185 190

Leu Cys Ala Ile Gly Ser Arg Pro Lys Trp Lys Lys Leu Val Thr Asn
 195 200 205

Phe Gln Met Val Gln Phe Ala Phe Gly Met Gly Leu Gly Ala Ala Trp
 210 215 220

Met Leu Pro Glu His Tyr Phe Gly Ser Gly Cys Ala Gly Ile Trp Thr
 225 230 235 240

Val Tyr Phe Asn Gly Val Phe Thr Ala Ser Leu Leu Ala Leu Phe Tyr
 245 250 255

Asn Phe His Ser Lys Asn Tyr Glu Lys Thr Thr Thr Ser Pro Leu Tyr
 260 265 270

Lys Ile Glu Ser Phe Ile Phe Ile His Gly Glu Arg Trp Ala Asn Lys
 275 280 285

Ala Ile Thr Leu Phe Ser Lys Lys Asn Asp
290 295

<210> 137

<211> 837

<212> DNA

<213> Arabidopsis thaliana

<220>

<221> CDS

<222> (1)..(837)

<223> Delta-5 elongase

<400> 137

atg gca tca att tac tcc tct tta acc tac tgg ctc gtt aac cac ccc 48
Met Ala Ser Ile Tyr Ser Ser Leu Thr Tyr Trp Leu Val Asn His Pro
1 5 10 15

tac atc tcc aat ttt act tgg atc gaa ggt gaa acc cta ggc tcc acc 96
Tyr Ile Ser Asn Phe Thr Trp Ile Glu Gly Glu Thr Leu Gly Ser Thr
20 25 30

gtc ttt ttc gta tcc gtc gta gtc tcc gtt tac ctc tcc gcc acg ttc 144
Val Phe Phe Val Ser Val Val Val Ser Val Tyr Leu Ser Ala Thr Phe
35 40 45

ctc ctc cga tcc gcc atc gat tca ctc cca tca ctc agt cca cgt atc 192
Leu Leu Arg Ser Ala Ile Asp Ser Leu Pro Ser Leu Ser Pro Arg Ile
50 55 60

ctc aaa ccg atc aca gcc gtc cac agc cta atc ctc tgt ctc ctc tcc 240
Leu Lys Pro Ile Thr Ala Val His Ser Leu Ile Leu Cys Leu Leu Ser
65 70 75 80

tta gtc atg gcc gtc ggt tgc act ctc tca ata acc tca tct cac gcg 288
Leu Val Met Ala Val Gly Cys Thr Leu Ser Ile Thr Ser Ser His Ala
85 90 95

tct tca gat ccg atg gcg cgt ttc ctt cac gcg att tgc ttt ccc gtc 336
Ser Ser Asp Pro Met Ala Arg Phe Leu His Ala Ile Cys Phe Pro Val
100 105 110

gac gtt aaa cct aac gga ccg ctt ttc ttc tgg gct caa gtc ttc tac 384
Asp Val Lys Pro Asn Gly Pro Leu Phe Phe Trp Ala Gln Val Phe Tyr
115 120 125

ctc tcg aag atc ctc gag ttc gga gac acg atc ctc atc ata ctc ggc 432
Leu Ser Lys Ile Leu Glu Phe Gly Asp Thr Ile Leu Ile Ile Leu Gly
130 135 140

aaa tca atc caa cgg cta tcc ttc ctc cac gtg tac cac cac gcg acg 480
Lys Ser Ile Gln Arg Leu Ser Phe Leu His Val Tyr His His Ala Thr
145 150 155 160

gtt gtg gtc atg tgt tat ctc tgg ctc cga act cgc caa tcg atg ttt 528
 Val Val Val Met Cys Tyr Leu Trp Leu Arg Thr Arg Gln Ser Met Phe
 165 170 175
 ccg att gcg ctc gtg acg aat tcg acg gta cac gtc atc atg tac ggt 576
 Pro Ile Ala Leu Val Thr Asn Ser Thr Val His Val Ile Met Tyr Gly
 180 185 190
 tac tac ttc ctc tgc gcc gtt gga tcg agg ccc aag tgg aag aga ttg 624
 Tyr Tyr Phe Leu Cys Ala Val Gly Ser Arg Pro Lys Trp Lys Arg Leu
 195 200 205
 gtg acg gat tgt cag att gtt cag ttt gtt ttc agt ttc ggg tta tcc 672
 Val Thr Asp Cys Gln Ile Val Gln Phe Val Phe Ser Phe Gly Leu Ser
 210 215 220
 ggt tgg atg ctc cga gag cac tta ttc ggg tcg ggt tgc acc ggg att 720
 Gly Trp Met Leu Arg Glu His Leu Phe Gly Ser Gly Cys Thr Gly Ile
 225 230 235 240
 tgg gga tgg tgt ttc aac gct gca ttt aat gct tct ctt ttg gct ctc 768
 Trp Gly Trp Cys Phe Asn Ala Ala Phe Asn Ala Ser Leu Leu Ala Leu
 245 250 255
 ttt tcc aac ttc cat tca aag aat tat gtc aag aag cca acg aga gag 816
 Phe Ser Asn Phe His Ser Lys Asn Tyr Val Lys Lys Pro Thr Arg Glu
 260 265 270
 gat ggc aaa aaa agc gat tag 837
 Asp Gly Lys Lys Ser Asp
 275

<210> 138

<211> 278

<212> PRT

<213> Arabidopsis thaliana

<400> 138

Met Ala Ser Ile Tyr Ser Ser Leu Thr Tyr Trp Leu Val Asn His Pro
 1 5 10 15
 Tyr Ile Ser Asn Phe Thr Trp Ile Glu Gly Glu Thr Leu Gly Ser Thr
 20 25 30
 Val Phe Phe Val Ser Val Val Val Ser Val Tyr Leu Ser Ala Thr Phe
 35 40 45
 Leu Leu Arg Ser Ala Ile Asp Ser Leu Pro Ser Leu Ser Pro Arg Ile
 50 55 60
 Leu Lys Pro Ile Thr Ala Val His Ser Leu Ile Leu Cys Leu Leu Ser
 65 70 75 80

Leu Val Met Ala Val Gly Cys Thr Leu Ser Ile Thr Ser Ser His Ala
85 90 95

Ser Ser Asp Pro Met Ala Arg Phe Leu His Ala Ile Cys Phe Pro Val
100 105 110

Asp Val Lys Pro Asn Gly Pro Leu Phe Phe Trp Ala Gln Val Phe Tyr
115 120 125

Leu Ser Lys Ile Leu Glu Phe Gly Asp Thr Ile Leu Ile Ile Leu Gly
130 135 140

Lys Ser Ile Gln Arg Leu Ser Phe Leu His Val Tyr His His Ala Thr
145 150 155 160

Val Val Val Met Cys Tyr Leu Trp Leu Arg Thr Arg Gln Ser Met Phe
165 170 175

Pro Ile Ala Leu Val Thr Asn Ser Thr Val His Val Ile Met Tyr Gly
180 185 190

Tyr Tyr Phe Leu Cys Ala Val Gly Ser Arg Pro Lys Trp Lys Arg Leu
195 200 205

Val Thr Asp Cys Gln Ile Val Gln Phe Val Phe Ser Phe Gly Leu Ser
210 215 220

Gly Trp Met Leu Arg Glu His Leu Phe Gly Ser Gly Cys Thr Gly Ile
225 230 235 240

Trp Gly Trp Cys Phe Asn Ala Ala Phe Asn Ala Ser Leu Leu Ala Leu
245 250 255

Phe Ser Asn Phe His Ser Lys Asn Tyr Val Lys Lys Pro Thr Arg Glu
260 265 270

Asp Gly Lys Lys Ser Asp
275

<210> 139

<211> 6

<212> PRT

<213> Consensus

<220>

<221> MISC_FEATURE

<222> (1)..(6)

<223> Xaa in positions 3 and 4 the sequence has the meaning
given in Table A.

<400> 139

Leu His Xaa Xaa His His
1 5

<210> 140

<211> 8

<212> PRT

<213> Consensus

<220>

<221> MISC_FEATURE

<222> (1)..(8)

<223> Xaa at positions 2, 3, 5 and 6 the sequence has the
meaning given in Table A.

<400> 140

Thr Xaa Xaa Gln Xaa Xaa Gln Phe
1 5

<210> 141

<211> 6

<212> PRT

<213> Consensus

<220>

<221> MISC_FEATURE

<222> (1)..(6)

<223> Xaa at position 3 the sequence has the meaning given
in Table A.

<400> 141

Asp Thr Xaa Phe Met Val
1 5

<210> 142

<211> 8

<212> PRT

<213> Consensus

<220>

<221> MISC_FEATURE

<222> (1)..(8)

<223> Xaa at positions 5 and 6 in the sequence has the meaning
given in Table A.

<400> 142

Thr Gln Ala Gln Xaa Xaa Gln Phe
1 5

<210> 143

<211> 60

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(60)

<223>

<400> 143
gtcgacccgc ggactagtgg gccctctaga cccgggggat ccggatctgc tggctatgaa 60

<210> 144

<211> 60

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(60)

<223>

<400> 144
gtcgacccgc ggactagtgg gccctctaga cccgggggat ccggatctgc tggctatgaa 60

<210> 145

<211> 36

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(36)

<223>

<400> 145
ggtaccacat aatgtgcgtg gagacggaaa ataacg 36

<210> 146

<211> 33

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(33)

<223>

<400> 146
ctcgagttac gccgtctttc cggagtgttg gcc 33

<210> 147

<211> 24

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(24)

<223>

<400> 147
gcggccgctt acgtggactt ggtc 24

<210> 148

<211> 24

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(24)

<223>

<400> 148
gcggccgcat ggcgacgaag gagg 24

<210> 149

<211> 25

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(25)

<223>

<400> 149

taagcttaca tggcgacgaa ggagg

25

<210> 150

<211> 24

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(24)

<223>

<400> 150

tggatccact tacgtggact tgg

24

<210> 151

<211> 60

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(60)

<223>

<400> 151

gtcgacccgc ggactagtgg gccctctaga cccgggggat ccggatctgc tggctatgaa

60

<210> 152

<211> 31

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(31)

<223>

<400> 152
gcggccgcac catgtgctca ccaccgccgt c

31

<210> 153

<211> 26

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(26)

<223>

<400> 153
gcggccgcct acatggcacc agtaac

26

<210> 154

<211> 31

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(31)

<223>

<400> 154
gcggccgcac catgtgctca tcaccgccgt c 31

<210> 155

<211> 26

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(26)

<223>

<400> 155
gcggccgcct acatggcacc agtaac 26

<210> 156

<211> 31

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(31)

<223>

<400> 156
gcggccgcac catggacgcc tacaacgctg c 31

<210> 157

<211> 27

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(27)

<223>

<400> 157

gcggccgcct aagcactctt cttcttt

27

<210> 158

<211> 23

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(23)

<223>

<400> 158

accatgtgct caccaccgcc gtc

23

<210> 159

<211> 18

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(18)

<223>

<400> 159

ctacatggca ccagtaac

18

<210> 160

<211> 23

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(23)

<223>

<400> 160
accatgtgct catcaccgcc gtc

23

<210> 161

<211> 18

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(18)

<223>

<400> 161
ctacatggca ccagtaac

18

<210> 162

<211> 23

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(23)

<223>

<400> 162
accatggacg cctacaacgc tgc 23

<210> 163

<211> 19

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(19)

<223>

<400> 163
ctaagcactc ttcttcttt 19

<210> 164

<211> 60

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(60)

<223>

<400> 164
gtcgaccgc ggactagtgg gccctctaga cccgggggat ccggatctgc tggctatgaa 60

<210> 165

<211> 60

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(60)

<223>

<400> 165

gtcgacccgc ggactagtgg gccctctaga cccgggggat ccggatctgc tggctatgaa 60

<210> 166

<211> 29

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(29)

<223>

<400> 166

gcggccgcat aatgacgagc aacatgagc 29

<210> 167

<211> 29

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(29)

<223>

<400> 167

gcggccgctt aggcgactt ggccttggg 29

<210> 168

<211> 34

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(34)

<223>

<400> 168

gcggccgcac catggacgtc gtcgagcagc aatg

34

<210> 169

<211> 36

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(36)

<223>

<400> 169

gcggccgctt agatggtctt ctgcttcttg ggcgcc

36

<210> 170

<211> 23

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(23)

<223>

<400> 170
gacataatga cgagcaacat gag 23

<210> 171

<211> 25

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(25)

<223>

<400> 171
cggcttaggc cgacttgGCC ttggg 25

<210> 172

<211> 30

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(30)

<223>

<400> 172
agacataatg gacgtcgtcg agcagcaatg 30

<210> 173

<211> 28

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(28)

<223>

<400> 173

ttagatgggtc ttctgcttct tgggcgcc

28

<210> 174

<211> 60

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(60)

<223>

<400> 174

gtcgacccgc ggactagtgg gccctctaga cccgggggat ccggatctgc tggctatgaa

60

<210> 175

<211> 29

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(29)

<223>

<400> 175

gcggccgcat aatggcttca acatggcaa

29

<210> 176

<211> 32

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(32)

<223>

<400> 176

gcggccgctt atgtcttctt gctcttctg tt

32

<210> 177

<211> 26

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(26)

<223>

<400> 177

gcggccgcat aatggagact tttaat

26

<210> 178

<211> 28

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(28)

<223>

<400> 178
gcggccgctc agtccccct cactttcc 28

<210> 179

<211> 29

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(29)

<223>

<400> 179
aagcttacat aatggcttca acatggcaa 29

<210> 180

<211> 30

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(30)

<223>

<400> 180
ggatccttat gtcttcttgc tcttctgtt 30

<210> 181

<211> 26

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(26)

<223>

<400> 181

aagcttacat aatggagact tttaat

26

<210> 182

<211> 27

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(27)

<223>

<400> 182

ggatccttca gtccccctc actttcc

27

<210> 183

<211> 993

<212> DNA

<213> *Phaeodactylum tricornutum*

<220>

<221> CDS

<222> (103)..(939)

<223> Delta-6 elongase

<400> 183

ggctctttgt ggtagctatc gtcacacac gcaggtcgtt gtcactatc gtgatccgta

60

tattgaccgt gcacttgtgt aaaacagaga tatttcaaga gt atg atg gta cct
Met Met Val Pro

114

1

tca agt tat gac gag tat atc gtc atg gtc aac gac ctt ggc gac tct

162

245

Ser 5	Ser	Tyr	Asp	Glu	Tyr 10	Ile	Val	Met	Val	Asn 15	Asp	Leu	Gly	Asp	Ser 20	
att	ctg	agc	tgg	gcc	gac	cct	gat	cac	tat	cgt	gga	cat	acc	gag	gga	210
Ile	Leu	Ser	Trp	Ala	Asp	Pro	Asp	His	Tyr 30	Arg	Gly	His	Thr	Glu	Gly	
				25										35		
tgg	gag	ttc	act	gac	ttt	tct	gct	gct	ttt	agc	att	gcc	gtc	gcg	tac	258
Trp	Glu	Phe	Thr	Asp	Phe	Ser	Ala	Ala	Phe	Ser	Ile	Ala	Val	Ala	Tyr	
			40					45					50			
ctc	ctg	ttt	gtc	ttt	gtt	gga	tct	ctc	att	atg	agt	atg	gga	gtc	ccc	306
Leu	Leu	Phe	Val	Phe	Val	Gly	Ser	Leu	Ile	Met	Ser	Met	Gly	Val	Pro	
		55					60					65				
gca	att	gac	cct	tat	ccg	ctc	aag	ttt	gtc	tac	aat	gtt	tca	cag	att	354
Ala	Ile	Asp	Pro	Tyr	Pro	Leu	Lys	Phe	Val	Tyr	Asn	Val	Ser	Gln	Ile	
	70					75					80					
atg	ctt	tgt	gct	tac	atg	acc	att	gaa	gcc	agt	ctt	cta	gct	tat	cgt	402
Met	Leu	Cys	Ala	Tyr	Met	Thr	Ile	Glu	Ala	Ser	Leu	Leu	Ala	Tyr	Arg	
85					90					95					100	
aac	ggc	tac	aca	ttc	tgg	cct	tgc	aac	gat	tgg	gac	ttt	gaa	aag	ccg	450
Asn	Gly	Tyr	Thr	Phe	Trp	Pro	Cys	Asn	Asp	Trp	Asp	Phe	Glu	Lys	Pro	
				105					110					115		
cct	atc	gct	aag	ctc	ctc	tgg	ctc	ttt	tac	gtt	tcc	aaa	att	tgg	gat	498
Pro	Ile	Ala	Lys	Leu	Leu	Trp	Leu	Phe	Tyr	Val	Ser	Lys	Ile	Trp	Asp	
			120					125					130			
ttt	tgg	gac	acc	atc	ttt	att	gtt	ctc	ggg	aag	aag	tgg	cgt	caa	ctt	546
Phe	Trp	Asp	Thr	Ile	Phe	Ile	Val	Leu	Gly	Lys	Lys	Trp	Arg	Gln	Leu	
		135					140					145				
tcc	ttc	ctg	cac	gtc	tac	cat	cac	acc	acc	atc	ttt	ctc	ttc	tac	tgg	594
Ser	Phe	Leu	His	Val	Tyr	His	His	Thr	Thr	Ile	Phe	Leu	Phe	Tyr	Trp	
	150					155					160					
ttg	aat	gca	cat	gta	aac	ttt	gat	ggt	gat	att	ttc	ctc	acc	atc	gtc	642
Leu	Asn	Ala	His	Val	Asn	Phe	Asp	Gly	Asp	Ile	Phe	Leu	Thr	Ile	Val	
165					170					175					180	
ttg	aac	ggt	ttc	atc	cac	acc	gtc	atg	tac	acg	tac	tac	ttc	att	tgc	690
Leu	Asn	Gly	Phe	Ile	His	Thr	Val	Met	Tyr	Thr	Tyr	Tyr	Phe	Ile	Cys	
				185					190					195		
atg	cac	acc	aag	gtc	cca	gag	acc	ggc	aaa	tcc	ttg	ccc	att	tgg	tgg	738
Met	His	Thr	Lys	Val	Pro	Glu	Thr	Gly	Lys	Ser	Leu	Pro	Ile	Trp	Trp	
			200					205					210			
aaa	tct	agt	ttg	aca	agc	atg	cag	ctg	gtg	cag	ttc	atc	acg	atg	atg	786
Lys	Ser	Ser	Leu	Thr	Ser	Met	Gln	Leu	Val	Gln	Phe	Ile	Thr	Met	Met	
		215					220					225				
acg	cag	gct	atc	atg	atc	ttg	tac	aag	ggc	tgt	gct	gct	ccc	cat	agc	834
Thr	Gln	Ala	Ile	Met	Ile	Leu	Tyr	Lys	Gly	Cys	Ala	Ala	Pro	His	Ser	
		230				235					240					
cgg	gtg	gtg	aca	tcg	tac	ttg	gtt	tac	att	ttg	tcg	ctc	ttt	att	ttg	882
Arg	Val	Val	Thr	Ser	Tyr	Leu	Val	Tyr	Ile	Leu	Ser	Leu	Phe	Ile	Leu	
245					250					255					260	
ttc	gcc	cag	ttc	ttt	gtc	agc	tca	tac	ctc	aag	ccg	aag	aag	aag	aag	930

246

Phe Ala Gln Phe Phe Val Ser Ser Tyr Leu Lys Pro Lys Lys Lys Lys
 265 270 275

aca gct taa gcgaaatttg ggtctacgtt aaaacaatta cgttacaaaa 979
 Thr Ala

aaaaaaaaaa aaaa 993

<210> 184

<211> 278

<212> PRT

<213> Phaeodactylum tricornutum

<400> 184

Met Met Val Pro Ser Ser Tyr Asp Glu Tyr Ile Val Met Val Asn Asp
 1 5 10 15

Leu Gly Asp Ser Ile Leu Ser Trp Ala Asp Pro Asp His Tyr Arg Gly
 20 25 30

His Thr Glu Gly Trp Glu Phe Thr Asp Phe Ser Ala Ala Phe Ser Ile
 35 40 45

Ala Val Ala Tyr Leu Leu Phe Val Phe Val Gly Ser Leu Ile Met Ser
 50 55 60

Met Gly Val Pro Ala Ile Asp Pro Tyr Pro Leu Lys Phe Val Tyr Asn
 65 70 75 80

Val Ser Gln Ile Met Leu Cys Ala Tyr Met Thr Ile Glu Ala Ser Leu
 85 90 95

Leu Ala Tyr Arg Asn Gly Tyr Thr Phe Trp Pro Cys Asn Asp Trp Asp
 100 105 110

Phe Glu Lys Pro Pro Ile Ala Lys Leu Leu Trp Leu Phe Tyr Val Ser
 115 120 125

Lys Ile Trp Asp Phe Trp Asp Thr Ile Phe Ile Val Leu Gly Lys Lys
 130 135 140

Trp Arg Gln Leu Ser Phe Leu His Val Tyr His His Thr Thr Ile Phe
 145 150 155 160

Leu Phe Tyr Trp Leu Asn Ala His Val Asn Phe Asp Gly Asp Ile Phe
 165 170 175

Leu Thr Ile Val Leu Asn Gly Phe Ile His Thr Val Met Tyr Thr Tyr
180 185 190

Tyr Phe Ile Cys Met His Thr Lys Val Pro Glu Thr Gly Lys Ser Leu
195 200 205

Pro Ile Trp Trp Lys Ser Ser Leu Thr Ser Met Gln Leu Val Gln Phe
210 215 220

Ile Thr Met Met Thr Gln Ala Ile Met Ile Leu Tyr Lys Gly Cys Ala
225 230 235 240

Ala Pro His Ser Arg Val Val Thr Ser Tyr Leu Val Tyr Ile Leu Ser
245 250 255

Leu Phe Ile Leu Phe Ala Gln Phe Phe Val Ser Ser Tyr Leu Lys Pro
260 265 270

Lys Lys Lys Lys Thr Ala
275

<210> 185

<211> 20

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(20)

<223> N in positions 3 and 18 is C or T.

<400> 185
aanctuctut ggctuttnta

20

<210> 186

<211> 23

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(23)

<223> N in positions 3 and 15 is C or T. N in positions
9, 12 and 21 is A or G.

<400> 186

gantguacna anaantgugc naa

23

<210> 187

<211> 446

<212> DNA

<213> PCR fragment

<220>

<221> misc_feature

<222> (1)..(446)

<223> PCR fragment

<400> 187

aagctcctct ggctctttta cgtttccaaa atttgggatt tttgggacac catctttatt	60
gttctcggga agaagtggcg tcaactttcc ttctgcacg tctaccatca caccaccatc	120
tttctcttct actggttgaa tgcacatgta aactttgatg gtgatatttt cctcaccatc	180
gtcttgaacg gtttcatcca caccgtcatg tacacgtact acttcatttg catgcacacc	240
aagggtcccag agaccggcaa atccttgccc atttggtgga aatctagttt gacaagcatg	300
cagctgggtgc agttcatcac gatgatgacg caggctatca tgatcttgta caagggctgt	360
gctgctcccc atagccgggt ggtgacatcg tacttggttt acattttgtc gctctttatt	420
ttgttcgccc agttctttgt cagctc	446

<210> 188

<211> 30

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(30)

<223>

<400> 188

gcggccgcac ataatgatgg taccttcaag

30

<210> 189

<211> 22

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(22)

<223>

<400> 189

gaagacagct taatagacta gt

22

<210> 190

<211> 31

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(31)

<223>

<400> 190

gcggccgcac catgatggta ccttcaagtt a

31

<210> 191

<211> 24

<212> DNA

<213> Primer

<220>

<221> misc_feature

<222> (1)..(24)

<223>

<400> 191

gaagacagct taataggcgg ccgc

24

<210> 192

<211> 859

<212> DNA

<213> PCR product

<400> 192

gcgggccgcac ataatgatgg taccttcaag ttatgacgag tataatcgta tgggtcaacga	60
ccttgggcgac tctattctga gctggggcga ccctgatcac tatcgtggac ataccgaggg	120
atgggagttc actgactttt ctgctgcttt tagcattgcc gtcgcgtacc tcctgtttgt	180
ctttgttga tctctcatta tgagtatggg agtccccgca attgaccctt atccgctcaa	240
gtttgtctac aatgtttcac agattatgct ttgtgcttac atgaccattg aagccagtct	300
tctagcttat cgtaacggct acacattctg gccttgcaac gattgggact ttgaaaagcc	360
gcctatcgct aagctcctct ggctctttta cgtttccaaa atttgggatt tttgggacac	420
catctttatt gttctcggga agaagtggcg tcaactttcc ttctgcacg tctaccatca	480
caccaccatc tttctcttct actggttgaa tgcacatgta aactttgatg gtgatatttt	540
cctcaccatc gtcttgaacg gtttcatcca caccgtcatg tacacgtact acttcatttg	600
catgcacacc aaggtccag agaccggcaa atccttgccc atttggtgga aatctagttt	660
gacaagcatg cagctggtgc agttcatcac gatgatgacg caggctatca tgatcttgta	720
caagggctgt gctgctcccc atagccgggt ggtgacatcg tacttggttt acattttgtc	780
gctctttatt ttgttcgccc agttctttgt cagctcatat ctcaagccga agaagaagaa	840
gacagcttaa tagactagt	859

<210> 193

<211> 1380

<212> DNA

<213> Phytium irregulare

<220>

<221> CDS

<222> (1)..(1380)

<223> Delta-6 desaturase

<400> 193

atg	gtg	gac	ctc	aag	cct	gga	gtg	aag	cgc	ctg	gtg	agc	tgg	aag	gag	48
Met	Val	Asp	Leu	Lys	Pro	Gly	Val	Lys	Arg	Leu	Val	Ser	Trp	Lys	Glu	
1				5					10					15		

atc	cgc	gag	cac	gcg	acg	ccc	gcg	acc	gcg	tgg	atc	gtg	att	cac	cac	96
Ile	Arg	Glu	His	Ala	Thr	Pro	Ala	Thr	Ala	Trp	Ile	Val	Ile	His	His	
			20					25					30			

aag	gtc	tac	gac	atc	tcc	aag	tgg	gac	tcg	cac	ccg	ggt	ggc	tcc	gtg	144
Lys	Val	Tyr	Asp	Ile	Ser	Lys	Trp	Asp	Ser	His	Pro	Gly	Gly	Ser	Val	
		35					40					45				

atg	ctc	acg	cag	gcc	ggc	gag	gac	gcc	acg	gac	gcc	ttc	gcg	gtc	ttc	192
Met	Leu	Thr	Gln	Ala	Gly	Glu	Asp	Ala	Thr	Asp	Ala	Phe	Ala	Val	Phe	
	50					55					60					

cac	ccg	tcc	tcg	gcg	ctc	aag	ctg	ctc	gag	cag	ttc	tac	gtc	ggc	gac	240
His	Pro	Ser	Ser	Ala	Leu	Lys	Leu	Leu	Glu	Gln	Phe	Tyr	Val	Gly	Asp	
65					70					75					80	

gtg	gac	gaa	acc	tcc	aag	gcc	gag	atc	gag	ggg	gag	ccg	gcg	agc	gac	288
Val	Asp	Glu	Thr	Ser	Lys	Ala	Glu	Ile	Glu	Gly	Glu	Pro	Ala	Ser	Asp	
				85					90					95		

gag	gag	cgc	gcg	cgc	cgc	gag	cgc	atc	aac	gag	ttc	atc	gcg	tcc	tac	336
Glu	Glu	Arg	Ala	Arg	Arg	Glu	Arg	Ile	Asn	Glu	Phe	Ile	Ala	Ser	Tyr	
			100					105					110			

cgc	cgt	ctg	cgc	gtc	aag	gtc	aag	ggc	atg	ggg	ctc	tac	gac	gcc	agc	384
Arg	Arg	Leu	Arg	Val	Lys	Val	Lys	Gly	Met	Gly	Leu	Tyr	Asp	Ala	Ser	
		115					120					125				

gcg	ctc	tac	tac	gcg	tgg	aag	ctc	gtg	agc	acg	ttc	ggc	atc	gcg	gtg	432
Ala	Leu	Tyr	Tyr	Ala	Trp	Lys	Leu	Val	Ser	Thr	Phe	Gly	Ile	Ala	Val	
	130					135						140				

ctc	tcg	atg	gcg	atc	tgc	ttc	ttc	ttc	aac	agt	ttc	gcc	atg	tac	atg	480
Leu	Ser	Met	Ala	Ile	Cys	Phe	Phe	Phe	Asn	Ser	Phe	Ala	Met	Tyr	Met	
145					150					155					160	

gtc	gcc	ggc	gtg	att	atg	ggg	ctc	ttc	tac	cag	cag	tcc	gga	tgg	ctg	528
Val	Ala	Gly	Val	Ile	Met	Gly	Leu	Phe	Tyr	Gln	Gln	Ser	Gly	Trp	Leu	
				165					170						175	

gcg cac gac ttc ttg cac aac cag gtg tgc gag aac cgc acg ctc ggc Ala His Asp Phe Leu His Asn Gln Val Cys Glu Asn Arg Thr Leu Gly 180 185 190	576
aac ctt atc ggc tgc ctc gtg ggc aac gcc tgg cag ggc ttc agc atg Asn Leu Ile Gly Cys Leu Val Gly Asn Ala Trp Gln Gly Phe Ser Met 195 200 205	624
cag tgg tgg aag aac aag cac aac ctg cac cac gcg gtg ccg aac ctg Gln Trp Trp Lys Asn Lys His Asn Leu His His Ala Val Pro Asn Leu 210 215 220	672
cac agc gcc aag gac gag ggc ttc atc ggc gac ccg gac atc gac acc His Ser Ala Lys Asp Glu Gly Phe Ile Gly Asp Pro Asp Ile Asp Thr 225 230 235 240	720
atg ccg ctg ctg gcg tgg tct aag gag atg gcg cgc aag gcg ttc gag Met Pro Leu Leu Ala Trp Ser Lys Glu Met Ala Arg Lys Ala Phe Glu 245 250 255	768
tcg gcg cac ggc ccg ttc ttc atc cgc aac cag gcg ttc cta tac ttc Ser Ala His Gly Pro Phe Phe Ile Arg Asn Gln Ala Phe Leu Tyr Phe 260 265 270	816
ccg ctg ctg ctg ctc gcg cgc ctg agc tgg ctc gcg cag tcg ttc ttc Pro Leu Leu Leu Leu Ala Arg Leu Ser Trp Leu Ala Gln Ser Phe Phe 275 280 285	864
tac gtg ttc acc gag ttc tcg ttc ggc atc ttc gac aag gtc gag ttc Tyr Val Phe Thr Glu Phe Ser Phe Gly Ile Phe Asp Lys Val Glu Phe 290 295 300	912
gac gga ccg gag aag gcg ggt ctg atc gtg cac tac atc tgg cag ctc Asp Gly Pro Glu Lys Ala Gly Leu Ile Val His Tyr Ile Trp Gln Leu 305 310 315 320	960
gcg atc ccg tac ttc tgc aac atg agc ctg ttt gag ggc gtg gca tac Ala Ile Pro Tyr Phe Cys Asn Met Ser Leu Phe Glu Gly Val Ala Tyr 325 330 335	1008
ttc ctc atg ggc cag gcg tcc tgc ggc ttg ctc ctg gcg ctg gtg ttc Phe Leu Met Gly Gln Ala Ser Cys Gly Leu Leu Leu Ala Leu Val Phe 340 345 350	1056
agt att ggc cac aac ggc atg tcg gtg tac gag cgc gaa acc aag ccg Ser Ile Gly His Asn Gly Met Ser Val Tyr Glu Arg Glu Thr Lys Pro 355 360 365	1104
gac ttc tgg cag ctg cag gtg acc acg acg cgc aac atc cgc gcg tcg Asp Phe Trp Gln Leu Gln Val Thr Thr Thr Arg Asn Ile Arg Ala Ser 370 375 380	1152
gta ttc atg gac tgg ttc acc ggt ggc ttg aac tac cag atc gac cat Val Phe Met Asp Trp Phe Thr Gly Gly Leu Asn Tyr Gln Ile Asp His 385 390 395 400	1200
cac ctg ttc ccg ctc gtg ccg cgc cac aac ttg cca aag gtc aac gtg His Leu Phe Pro Leu Val Pro Arg His Asn Leu Pro Lys Val Asn Val 405 410 415	1248
ctc atc aag tcg cta tgc aag gag ttc gac atc ccg ttc cac gag acc Leu Ile Lys Ser Leu Cys Lys Glu Phe Asp Ile Pro Phe His Glu Thr 420 425 430	1296

agc aag gaa ttt atc acc gag ttc cca gcg atg taa 1380
 Ser Lys Glu Phe Ile Thr Glu Phe Pro Ala Met
 450 455

<213> Phytium irregulare

Val Ala Gly Val Ile Met Gly Leu Phe Tyr Gln Gln Ser Gly Trp Leu
165 170 175

Ala His Asp Phe Leu His Asn Gln Val Cys Glu Asn Arg Thr Leu Gly
 180 185 190

Asn Leu Ile Gly Cys Leu Val Gly Asn Ala Trp Gln Gly Phe Ser Met
 195 200 205

Gln Trp Trp Lys Asn Lys His Asn Leu His His Ala Val Pro Asn Leu
 210 215 220

His Ser Ala Lys Asp Glu Gly Phe Ile Gly Asp Pro Asp Ile Asp Thr
 225 230 235 240

Met Pro Leu Leu Ala Trp Ser Lys Glu Met Ala Arg Lys Ala Phe Glu
 245 250 255

Ser Ala His Gly Pro Phe Phe Ile Arg Asn Gln Ala Phe Leu Tyr Phe
 260 265 270

Pro Leu Leu Leu Leu Ala Arg Leu Ser Trp Leu Ala Gln Ser Phe Phe
 275 280 285

Tyr Val Phe Thr Glu Phe Ser Phe Gly Ile Phe Asp Lys Val Glu Phe
 290 295 300

Asp Gly Pro Glu Lys Ala Gly Leu Ile Val His Tyr Ile Trp Gln Leu
 305 310 315 320

Ala Ile Pro Tyr Phe Cys Asn Met Ser Leu Phe Glu Gly Val Ala Tyr
 325 330 335

Phe Leu Met Gly Gln Ala Ser Cys Gly Leu Leu Leu Ala Leu Val Phe
 340 345 350

Ser Ile Gly His Asn Gly Met Ser Val Tyr Glu Arg Glu Thr Lys Pro
 355 360 365

Asp Phe Trp Gln Leu Gln Val Thr Thr Thr Arg Asn Ile Arg Ala Ser
 370 375 380

Val Phe Met Asp Trp Phe Thr Gly Gly Leu Asn Tyr Gln Ile Asp His
 385 390 395 400

His Leu Phe Pro Leu Val Pro Arg His Asn Leu Pro Lys Val Asn Val
 405 410 415

Leu Ile Lys Ser Leu Cys Lys Glu Phe Asp Ile Pro Phe His Glu Thr
 420 425 430

Gly Phe Trp Glu Gly Ile Tyr Glu Val Val Asp His Leu Ala Asp Ile
 435 440 445

Ser Lys Glu Phe Ile Thr Glu Phe Pro Ala Met
 450 455

<210> 195

<211> 1152

<212> DNA

<213> Calendula officinalis

<220>

<221> CDS

<222> (1)..(1152)

<223> Delta-12 desaturase

<400> 195

atg ggt gca ggc ggt cga atg caa gat ccc acc aac ggt ggc aac aaa 48
 Met Gly Ala Gly Gly Arg Met Gln Asp Pro Thr Asn Gly Gly Asn Lys
 1 5 10 15

acc gag ccc gaa cca atc caa cgg gtc cca cat gaa aaa ccc cca ttc 96
 Thr Glu Pro Glu Pro Ile Gln Arg Val Pro His Glu Lys Pro Pro Phe
 20 25 30

aca gtt gga gac atc aag aaa gcg atc cca cct cat tgt ttc aac cga 144
 Thr Val Gly Asp Ile Lys Lys Ala Ile Pro Pro His Cys Phe Asn Arg
 35 40 45

tcg gta att cgt tca ttt tca tac gtc ttt tac gac ctc aca atc gcg 192
 Ser Val Ile Arg Ser Phe Ser Tyr Val Phe Tyr Asp Leu Thr Ile Ala
 50 55 60

tca atc ttg tac tac att gcc aac aat tac atc tct acc ctc cct agc 240
 Ser Ile Leu Tyr Tyr Ile Ala Asn Asn Tyr Ile Ser Thr Leu Pro Ser
 65 70 75 80

cgg ctc gcc tac gtg gca tgg ccc gtt tac tgg gcc gtc caa ggg tgc 288
 Pro Leu Ala Tyr Val Ala Trp Pro Val Tyr Trp Ala Val Gln Gly Cys
 85 90 95

gtc tta acc ggg gtg tgg gtc ata gcc cac gaa tgt ggc cat cat gct 336
 Val Leu Thr Gly Val Trp Val Ile Ala His Glu Cys Gly His His Ala
 100 105 110

ttt agc gac cac caa tgg ctc gat gac acc gtg ggt ctc gtc ttg cac 384
 Phe Ser Asp His Gln Trp Leu Asp Asp Thr Val Gly Leu Val Leu His
 115 120 125

tcg ttc cta ctc gtg ccc tac ttt tcg tgg aaa tat agc cac cgt agg 432

256

Ser	Phe	Leu	Leu	Val	Pro	Tyr	Phe	Ser	Trp	Lys	Tyr	Ser	His	Arg	Arg	
130						135					140					
cac	cac	tcg	aac	acg	ggc	tcg	atc	gag	cac	gat	gag	gtt	ttc	gtc	ccg	480
His	His	Ser	Asn	Thr	Gly	Ser	Ile	Glu	His	Asp	Glu	Val	Phe	Val	Pro	
145					150					155					160	
aag	ttg	aaa	tcg	ggc	gtc	cgg	tca	acc	gcc	cgg	tac	cta	aac	aac	cca	528
Lys	Leu	Lys	Ser	Gly	Val	Arg	Ser	Thr	Ala	Arg	Tyr	Leu	Asn	Asn	Pro	
				165					170					175		
ccg	ggc	cga	atc	ttg	acc	cta	ctc	gta	acc	cta	acc	ctc	ggg	ttg	cct	576
Pro	Gly	Arg	Ile	Leu	Thr	Leu	Leu	Val	Thr	Leu	Thr	Leu	Gly	Trp	Pro	
			180					185					190			
cta	tac	ctc	acg	ttc	aac	gtt	tcg	ggc	cgt	tac	tac	gac	cgg	ttc	gcg	624
Leu	Tyr	Leu	Thr	Phe	Asn	Val	Ser	Gly	Arg	Tyr	Tyr	Asp	Arg	Phe	Ala	
	195					200						205				
tgc	cat	ttc	gac	ccg	aat	agc	ccg	atc	tac	tcg	aag	cgc	gaa	cgg	gct	672
Cys	His	Phe	Asp	Pro	Asn	Ser	Pro	Ile	Tyr	Ser	Lys	Arg	Glu	Arg	Ala	
	210					215					220					
caa	atc	ttc	ata	tcc	gac	gcc	ggg	atc	tta	gcc	gta	gtc	ttc	gta	ctc	720
Gln	Ile	Phe	Ile	Ser	Asp	Ala	Gly	Ile	Leu	Ala	Val	Val	Phe	Val	Leu	
225				230						235					240	
ttc	cga	ctc	gca	atg	acc	aaa	ggg	ctc	acg	tgg	gtc	cta	acc	atg	tac	768
Phe	Arg	Leu	Ala	Met	Thr	Lys	Gly	Leu	Thr	Trp	Val	Leu	Thr	Met	Tyr	
			245					250						255		
ggg	ggc	ccg	tta	ctc	gtg	gtc	aac	ggg	ttc	cta	gtc	ttg	atc	aca	ttc	816
Gly	Gly	Pro	Leu	Leu	Val	Val	Asn	Gly	Phe	Leu	Val	Leu	Ile	Thr	Phe	
		260					265					270				
cta	caa	cac	act	cac	cct	tcg	ctc	ccg	cac	tat	gac	tca	acc	gaa	tgg	864
Leu	Gln	His	Thr	His	Pro	Ser	Leu	Pro	His	Tyr	Asp	Ser	Thr	Glu	Trp	
	275					280					285					
gat	tgg	tta	cgt	ggg	gcc	ctc	acc	aca	atc	gac	cgt	gat	tac	ggg	atc	912
Asp	Trp	Leu	Arg	Gly	Ala	Leu	Thr	Thr	Ile	Asp	Arg	Asp	Tyr	Gly	Ile	
	290				295					300						
cta	aac	aaa	gtg	ttc	cat	aac	ata	acc	gac	act	cac	gtg	gcc	cac	cat	960
Leu	Asn	Lys	Val	Phe	His	Asn	Ile	Thr	Asp	Thr	His	Val	Ala	His	His	
305				310					315						320	
ttg	ttc	tct	aca	atg	cct	cat	tac	cat	gca	atg	gaa	gcc	acg	aag	gtg	1008
Leu	Phe	Ser	Thr	Met	Pro	His	Tyr	His	Ala	Met	Glu	Ala	Thr	Lys	Val	
			325					330					335			
atc	aaa	ccg	att	ttg	ggc	gat	tat	tat	cag	ttt	gac	ggg	acc	tcg	att	1056
Ile	Lys	Pro	Ile	Leu	Gly	Asp	Tyr	Tyr	Gln	Phe	Asp	Gly	Thr	Ser	Ile	
		340					345					350				
ttt	aag	gcg	atg	tat	cgg	gaa	aca	aag	gag	tgc	att	tat	gtt	gat	aag	1104
Phe	Lys	Ala	Met	Tyr	Arg	Glu	Thr	Lys	Glu	Cys	Ile	Tyr	Val	Asp	Lys	
	355					360					365					
gat	gag	gag	gtg	aaa	gat	ggg	gtt	tat	tgg	tat	cgt	aat	aag	att	taa	1152
Asp	Glu	Glu	Val	Lys	Asp	Gly	Val	Tyr	Trp	Tyr	Arg	Asn	Lys	Ile		
	370				375						380					

<210> 196

<211> 383

<212> PRT

<213> Calendula officinalis

<400> 196

Met Gly Ala Gly Gly Arg Met Gln Asp Pro Thr Asn Gly Gly Asn Lys
 1 5 10 15

Thr Glu Pro Glu Pro Ile Gln Arg Val Pro His Glu Lys Pro Pro Phe
 20 25 30

Thr Val Gly Asp Ile Lys Lys Ala Ile Pro Pro His Cys Phe Asn Arg
 35 40 45

Ser Val Ile Arg Ser Phe Ser Tyr Val Phe Tyr Asp Leu Thr Ile Ala
 50 55 60

Ser Ile Leu Tyr Tyr Ile Ala Asn Asn Tyr Ile Ser Thr Leu Pro Ser
 65 70 75 80

Pro Leu Ala Tyr Val Ala Trp Pro Val Tyr Trp Ala Val Gln Gly Cys
 85 90 95

Val Leu Thr Gly Val Trp Val Ile Ala His Glu Cys Gly His His Ala
 100 105 110

Phe Ser Asp His Gln Trp Leu Asp Asp Thr Val Gly Leu Val Leu His
 115 120 125

Ser Phe Leu Leu Val Pro Tyr Phe Ser Trp Lys Tyr Ser His Arg Arg
 130 135 140

His His Ser Asn Thr Gly Ser Ile Glu His Asp Glu Val Phe Val Pro
 145 150 155 160

Lys Leu Lys Ser Gly Val Arg Ser Thr Ala Arg Tyr Leu Asn Asn Pro
 165 170 175

Pro Gly Arg Ile Leu Thr Leu Leu Val Thr Leu Thr Leu Gly Trp Pro
 180 185 190

Leu Tyr Leu Thr Phe Asn Val Ser Gly Arg Tyr Tyr Asp Arg Phe Ala
 195 200 205

258

Cys His Phe Asp Pro Asn Ser Pro Ile Tyr Ser Lys Arg Glu Arg Ala
 210 215 220

Gln Ile Phe Ile Ser Asp Ala Gly Ile Leu Ala Val Val Phe Val Leu
 225 230 235 240

Phe Arg Leu Ala Met Thr Lys Gly Leu Thr Trp Val Leu Thr Met Tyr
 245 250 255

Gly Gly Pro Leu Leu Val Val Asn Gly Phe Leu Val Leu Ile Thr Phe
 260 265 270

Leu Gln His Thr His Pro Ser Leu Pro His Tyr Asp Ser Thr Glu Trp
 275 280 285

Asp Trp Leu Arg Gly Ala Leu Thr Thr Ile Asp Arg Asp Tyr Gly Ile
 290 295 300

Leu Asn Lys Val Phe His Asn Ile Thr Asp Thr His Val Ala His His
 305 310 315 320

Leu Phe Ser Thr Met Pro His Tyr His Ala Met Glu Ala Thr Lys Val
 325 330 335

Ile Lys Pro Ile Leu Gly Asp Tyr Tyr Gln Phe Asp Gly Thr Ser Ile
 340 345 350

Phe Lys Ala Met Tyr Arg Glu Thr Lys Glu Cys Ile Tyr Val Asp Lys
 355 360 365

Asp Glu Glu Val Lys Asp Gly Val Tyr Trp Tyr Arg Asn Lys Ile
 370 375 380

<210> 197

<211> 903

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(903)

<223> Delta-5 elongase

<400> 197
 atg tct gct tct gga gct ttg ttg cct gct att gct ttc gct gct tac 48
 Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Phe Ala Ala Tyr
 1 5 10 15

gct tac gct acc tac gct tat gct ttc gag tgg tct cat gct aac gga 96
 Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30

atc gat aac gtg gat gct aga gag tgg att gga gct ttg tct ttg aga 144
 Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45

ctc cct gca att gct acc acc atg tac ctc ttg ttc tgc ctt gtg gga 192
 Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly
 50 55 60

cct aga ttg atg gct aag agg gag gct ttt gat cct aag gga ttc atg 240
 Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met
 65 70 75 80

ctc gct tac aac gct tac caa acc gct ttc aac gtt gtg gtg ctc gga 288
 Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly
 85 90 95

atg ttc gct aga gag atc tct gga ttg gga caa cct gtt tgg gga tct 336
 Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser
 100 105 110

act atg cct tgg agc gat agg aag tcc ttc aag att ttg ttg gga gtg 384
 Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val
 115 120 125

tgg ctc cat tac aac aat aag tac ctc gag ttg ttg gat act gtg ttc 432
 Trp Leu His Tyr Asn Asn Lys Tyr Leu Glu Leu Leu Asp Thr Val Phe
 130 135 140

atg gtg gct agg aaa aag acc aag cag ctc tct ttc ttg cat gtg tac 480
 Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr
 145 150 155 160

cat cat gct ttg ttg att tgg gct tgg tgg ctt gtt tgt cat ctc atg 528
 His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met
 165 170 175

gct acc aac gat tgc atc gat gct tat ttc gga gct gct tgc aac tct 576
 Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser
 180 185 190

ttc atc cac atc gtg atg tac tcc tac tac ctc atg tct gct ttg gga 624
 Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
 195 200 205

att aga tgc cct tgg aag aga tat atc acc cag gct cag atg ttg caa 672
 Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
 210 215 220

ttc gtg atc gtg ttc gct cat gct gtt ttc gtg ctc aga caa aag cac 720
 Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
 225 230 235 240

tgc cct gtt act ttg cct tgg gca caa atg ttc gtg atg aca aat atg 768
 Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
 245 250 255

ttg gtg ctc ttc gga aac ttc tac ctc aag gct tac tct aac aag tct 816
 Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
 260 265 270

agg gga gat gga gct tct tct gtt aag cct gct gag act act aga gca 864
 Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
 275 280 285

cct tct gtg aga aga acc agg tcc agg aag atc gat tga 903
 Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
 290 295 300

<210> 198

<211> 300

<212> PRT

<213> *Ostreococcus tauri*

<400> 198

Met Ser Ala Ser Gly Ala Leu Leu Pro Ala Ile Ala Phe Ala Ala Tyr
 1 5 10 15

Ala Tyr Ala Thr Tyr Ala Tyr Ala Phe Glu Trp Ser His Ala Asn Gly
 20 25 30

Ile Asp Asn Val Asp Ala Arg Glu Trp Ile Gly Ala Leu Ser Leu Arg
 35 40 45

Leu Pro Ala Ile Ala Thr Thr Met Tyr Leu Leu Phe Cys Leu Val Gly
 50 55 60

Pro Arg Leu Met Ala Lys Arg Glu Ala Phe Asp Pro Lys Gly Phe Met
 65 70 75 80

Leu Ala Tyr Asn Ala Tyr Gln Thr Ala Phe Asn Val Val Val Leu Gly
 85 90 95

Met Phe Ala Arg Glu Ile Ser Gly Leu Gly Gln Pro Val Trp Gly Ser
 100 105 110

Thr Met Pro Trp Ser Asp Arg Lys Ser Phe Lys Ile Leu Leu Gly Val
 115 120 125

Trp Leu His Tyr Asn Asn Lys Tyr Leu Glu Leu Leu Asp Thr Val Phe
 130 135 140

Met Val Ala Arg Lys Lys Thr Lys Gln Leu Ser Phe Leu His Val Tyr
 145 150 155 160

His His Ala Leu Leu Ile Trp Ala Trp Trp Leu Val Cys His Leu Met
165 170 175

Ala Thr Asn Asp Cys Ile Asp Ala Tyr Phe Gly Ala Ala Cys Asn Ser
180 185 190

Phe Ile His Ile Val Met Tyr Ser Tyr Tyr Leu Met Ser Ala Leu Gly
195 200 205

Ile Arg Cys Pro Trp Lys Arg Tyr Ile Thr Gln Ala Gln Met Leu Gln
210 215 220

Phe Val Ile Val Phe Ala His Ala Val Phe Val Leu Arg Gln Lys His
225 230 235 240

Cys Pro Val Thr Leu Pro Trp Ala Gln Met Phe Val Met Thr Asn Met
245 250 255

Leu Val Leu Phe Gly Asn Phe Tyr Leu Lys Ala Tyr Ser Asn Lys Ser
260 265 270

Arg Gly Asp Gly Ala Ser Ser Val Lys Pro Ala Glu Thr Thr Arg Ala
275 280 285

Pro Ser Val Arg Arg Thr Arg Ser Arg Lys Ile Asp
290 295 300

<210> 199

<211> 879

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (1)..(879)

<223> Delta-6 elongase

<400> 199

atg tct gga ttg agg gct cct aac ttc ttg cat agg ttc tgg acc aag 48
Met Ser Gly Leu Arg Ala Pro Asn Phe Leu His Arg Phe Trp Thr Lys
1 5 10 15

tgg gat tac gct atc tct aag gtg gtg ttc act tgc gct gat tct ttc 96

262

Trp	Asp	Tyr	Ala	Ile	Ser	Lys	Val	Val	Phe	Thr	Cys	Ala	Asp	Ser	Phe	
			20					25					30			
cag	tgg	gat	atc	gga	cct	gtt	tct	tct	tct	acc	gct	cat	ttg	cct	gct	144
Gln	Trp	Asp	Ile	Gly	Pro	Val	Ser	Ser	Ser	Thr	Ala	His	Leu	Pro	Ala	
		35				40						45				
att	gag	tct	cct	act	cct	ttg	gtg	acc	tct	ttg	ctc	ttc	tac	ttg	gtg	192
Ile	Glu	Ser	Pro	Thr	Pro	Leu	Val	Thr	Ser	Leu	Leu	Phe	Tyr	Leu	Val	
	50					55					60					
act	gtg	ttc	ttg	tgg	tac	gga	aga	ttg	acc	aga	tcc	tcc	gat	aag	aag	240
Thr	Val	Phe	Leu	Trp	Tyr	Gly	Arg	Leu	Thr	Arg	Ser	Ser	Asp	Lys	Lys	
65					70				75						80	
atc	aga	gag	cct	acc	tgg	ttg	agg	aga	ttc	atc	atc	tgc	cac	aac	gct	288
Ile	Arg	Glu	Pro	Thr	Trp	Leu	Arg	Arg	Phe	Ile	Ile	Cys	His	Asn	Ala	
			85						90					95		
ttc	ttg	att	gtg	ctc	tcc	ttg	tac	atg	tgt	ttg	gga	tgc	gtt	gct	caa	336
Phe	Leu	Ile	Val	Leu	Ser	Leu	Tyr	Met	Cys	Leu	Gly	Cys	Val	Ala	Gln	
			100					105					110			
gct	tac	caa	aac	gga	tac	acc	ttg	tgg	gga	aac	gag	ttc	aag	gct	act	384
Ala	Tyr	Gln	Asn	Gly	Tyr	Thr	Leu	Trp	Gly	Asn	Glu	Phe	Lys	Ala	Thr	
		115					120					125				
gag	acc	caa	ttg	gct	ctc	tac	atc	tac	atc	ttc	tac	gtg	tcc	aag	atc	432
Glu	Thr	Gln	Leu	Ala	Leu	Tyr	Ile	Tyr	Ile	Phe	Tyr	Val	Ser	Lys	Ile	
	130					135						140				
tac	gag	ttc	gtg	gat	acc	tac	atc	atg	ctc	ctc	aag	aac	aac	ctc	agg	480
Tyr	Glu	Phe	Val	Asp	Thr	Tyr	Ile	Met	Leu	Leu	Lys	Asn	Asn	Leu	Arg	
145					150						155				160	
caa	gtg	tct	ttc	ttg	cac	atc	tac	cac	cac	tct	acc	atc	tct	ttc	atc	528
Gln	Val	Ser	Phe	Leu	His	Ile	Tyr	His	His	Ser	Thr	Ile	Ser	Phe	Ile	
				165					170					175		
tgg	tgg	atc	atc	gct	aga	aga	gca	cct	gga	gga	gat	gct	tat	ttc	tcc	576
Trp	Trp	Ile	Ile	Ala	Arg	Arg	Ala	Pro	Gly	Gly	Asp	Ala	Tyr	Phe	Ser	
			180					185					190			
gct	gct	ctc	aac	tct	tgg	gtt	cat	gtg	tgc	atg	tac	act	tac	tac	ctc	624
Ala	Ala	Leu	Asn	Ser	Trp	Val	His	Val	Cys	Met	Tyr	Thr	Tyr	Tyr	Leu	
		195					200					205				
ctc	tct	acc	ttg	att	gga	aag	gaa	gat	cct	aag	agg	tct	aac	tac	ctc	672
Leu	Ser	Thr	Leu	Ile	Gly	Lys	Glu	Asp	Pro	Lys	Arg	Ser	Asn	Tyr	Leu	
	210					215					220					
tgg	tgg	gga	agg	cat	ttg	acc	caa	atg	caa	atg	ctc	cag	ttc	ttc	ttc	720
Trp	Trp	Gly	Arg	His	Leu	Thr	Gln	Met	Gln	Met	Leu	Gln	Phe	Phe	Phe	
225					230					235					240	
aac	gtg	ctc	caa	gct	ctt	tat	tgc	gct	tcc	ttc	tcc	act	tac	cct	aag	768
Asn	Val	Leu	Gln	Ala	Leu	Tyr	Cys	Ala	Ser	Phe	Ser	Thr	Tyr	Pro	Lys	
				245					250					255		
ttc	ctc	tcc	aag	atc	ttg	ctc	gtg	tac	atg	atg	tct	ttg	ctc	gga	ctt	816
Phe	Leu	Ser	Lys	Ile	Leu	Leu	Val	Tyr	Met	Met	Ser	Leu	Leu	Gly	Leu	
			260					265					270			
ttc	gga	cac	ttc	tac	tac	tct	aag	cac	atc	gct	gct	gct	aag	ttg	caa	864

263

Phe Gly His Phe Tyr Tyr Ser Lys His Ile Ala Ala Ala Lys Leu Gln
 275 280 285

aag aag cag cag tga
 Lys Lys Gln Gln
 290

879

<210> 200

<211> 292

<212> PRT

<213> *Ostreococcus tauri*

<400> 200

Met Ser Gly Leu Arg Ala Pro Asn Phe Leu His Arg Phe Trp Thr Lys
 1 5 10 15

Trp Asp Tyr Ala Ile Ser Lys Val Val Phe Thr Cys Ala Asp Ser Phe
 20 25 30

Gln Trp Asp Ile Gly Pro Val Ser Ser Ser Thr Ala His Leu Pro Ala
 35 40 45

Ile Glu Ser Pro Thr Pro Leu Val Thr Ser Leu Leu Phe Tyr Leu Val
 50 55 60

Thr Val Phe Leu Trp Tyr Gly Arg Leu Thr Arg Ser Ser Asp Lys Lys
 65 70 75 80

Ile Arg Glu Pro Thr Trp Leu Arg Arg Phe Ile Ile Cys His Asn Ala
 85 90 95

Phe Leu Ile Val Leu Ser Leu Tyr Met Cys Leu Gly Cys Val Ala Gln
 100 105 110

Ala Tyr Gln Asn Gly Tyr Thr Leu Trp Gly Asn Glu Phe Lys Ala Thr
 115 120 125

Glu Thr Gln Leu Ala Leu Tyr Ile Tyr Ile Phe Tyr Val Ser Lys Ile
 130 135 140

Tyr Glu Phe Val Asp Thr Tyr Ile Met Leu Leu Lys Asn Asn Leu Arg
 145 150 155 160

Gln Val Ser Phe Leu His Ile Tyr His His Ser Thr Ile Ser Phe Ile
 165 170 175

264

Trp Trp Ile Ile Ala Arg Arg Ala Pro Gly Gly Asp Ala Tyr Phe Ser
 180 185 190

Ala Ala Leu Asn Ser Trp Val His Val Cys Met Tyr Thr Tyr Tyr Leu
 195 200 205

Leu Ser Thr Leu Ile Gly Lys Glu Asp Pro Lys Arg Ser Asn Tyr Leu
 210 215 220

Trp Trp Gly Arg His Leu Thr Gln Met Gln Met Leu Gln Phe Phe Phe
 225 230 235 240

Asn Val Leu Gln Ala Leu Tyr Cys Ala Ser Phe Ser Thr Tyr Pro Lys
 245 250 255

Phe Leu Ser Lys Ile Leu Leu Val Tyr Met Met Ser Leu Leu Gly Leu
 260 265 270

Phe Gly His Phe Tyr Tyr Ser Lys His Ile Ala Ala Ala Lys Leu Gln
 275 280 285

Lys Lys Gln Gln
 290

<210> 201

<211> 1421

<212> DNA

<213> *Ostreococcus tauri*

<220>

<221> CDS

<222> (26)..(1399)

<223> Delta-6 desaturase

<400> 201

ggatccttaa ttaaggcgcg ccaaa atg tgt gtt gag acc gag aac aac gat 52
 Met Cys Val Glu Thr Glu Asn Asn Asp
 1 5

gga atc cct act gtg gag atc gct ttc gat gga gag aga gaa aga gct 100
 Gly Ile Pro Thr Val Glu Ile Ala Phe Asp Gly Glu Arg Glu Arg Ala
 10 15 20 25

gag gct aac gtg aag ttg tct gct gag aag atg gaa cct gct gct ttg 148
 Glu Ala Asn Val Lys Leu Ser Ala Glu Lys Met Glu Pro Ala Ala Leu
 30 35 40

gct aag acc ttc gct aga aga tac gtg gtt atc gag gga gtt gag tac	196
Ala Lys Thr Phe Ala Arg Arg Tyr Val Val Ile Glu Gly Val Glu Tyr	
45 50 55	
gat gtg acc gat ttc aaa cat cct gga gga acc gtg att ttc tac gct	244
Asp Val Thr Asp Phe Lys His Pro Gly Gly Thr Val Ile Phe Tyr Ala	
60 65 70	
ctc tct aac act gga gct gat gct act gag gct ttc aag gag ttc cac	292
Leu Ser Asn Thr Gly Ala Asp Ala Thr Glu Ala Phe Lys Glu Phe His	
75 80 85	
cac aga tct aga aag gct agg aag gct ttg gct gct ttg cct tct aga	340
His Arg Ser Arg Lys Ala Arg Lys Ala Leu Ala Ala Leu Pro Ser Arg	
90 95 100 105	
cct gct aag acc gct aaa gtg gat gat gct gag atg ctc cag gat ttc	388
Pro Ala Lys Thr Ala Lys Val Asp Asp Ala Glu Met Leu Gln Asp Phe	
110 115 120	
gct aag tgg aga aag gag ttg gag agg gac gga ttc ttc aag cct tct	436
Ala Lys Trp Arg Lys Glu Leu Glu Arg Asp Gly Phe Phe Lys Pro Ser	
125 130 135	
cct gct cat gtt gct tac aga ttc gct gag ttg gct gct atg tac gct	484
Pro Ala His Val Ala Tyr Arg Phe Ala Glu Leu Ala Ala Met Tyr Ala	
140 145 150	
ttg gga acc tac ttg atg tac gct aga tac gtt gtg tcc tct gtg ttg	532
Leu Gly Thr Tyr Leu Met Tyr Ala Arg Tyr Val Val Ser Ser Val Leu	
155 160 165	
gtt tac gct tgc ttc ttc gga gct aga tgt gga tgg gtt caa cat gag	580
Val Tyr Ala Cys Phe Phe Gly Ala Arg Cys Gly Trp Val Gln His Glu	
170 175 180 185	
gga gga cat tct tct ttg acc gga aac atc tgg tgg gat aag aga atc	628
Gly Gly His Ser Ser Leu Thr Gly Asn Ile Trp Trp Asp Lys Arg Ile	
190 195 200	
caa gct ttc act gct gga ttc gga ttg gct gga tct gga gat atg tgg	676
Gln Ala Phe Thr Ala Gly Phe Gly Leu Ala Gly Ser Gly Asp Met Trp	
205 210 215	
aac tcc atg cac aac aag cac cat gct act cct caa aaa gtg agg cac	724
Asn Ser Met His Asn Lys His His Ala Thr Pro Gln Lys Val Arg His	
220 225 230	
gat atg gat ttg gat acc act cct gct gtt gct ttc ttc aac acc gct	772
Asp Met Asp Leu Asp Thr Thr Pro Ala Val Ala Phe Phe Asn Thr Ala	
235 240 245	
gtg gag gat aat aga cct agg gga ttc tct aag tac tgg ctc aga ttg	820
Val Glu Asp Asn Arg Pro Arg Gly Phe Ser Lys Tyr Trp Leu Arg Leu	
250 255 260 265	
caa gct tgg acc ttc att cct gtg act tct gga ttg gtg ttg ctc ttc	868
Gln Ala Trp Thr Phe Ile Pro Val Thr Ser Gly Leu Val Leu Leu Phe	
270 275 280	
tgg atg ttc ttc ctc cat cct tct aag gct ttg aag gga gga aag tac	916
Trp Met Phe Phe Leu His Pro Ser Lys Ala Leu Lys Gly Gly Lys Tyr	
285 290 295	

gag gag ctt gtg tgg atg ttg gct gct cat gtg att aga acc tgg acc 964
 Glu Glu Leu Val Trp Met Leu Ala Ala His Val Ile Arg Thr Trp Thr
 300 305 310
 att aag gct gtt act gga ttc acc gct atg caa tcc tac gga ctc ttc 1012
 Ile Lys Ala Val Thr Gly Phe Thr Ala Met Gln Ser Tyr Gly Leu Phe
 315 320 325
 ttg gct act tct tgg gtt tcc gga tgc tac ttg ttc gct cac ttc tct 1060
 Leu Ala Thr Ser Trp Val Ser Gly Cys Tyr Leu Phe Ala His Phe Ser
 330 335 340 345
 act tct cac acc cat ttg gat gtt gtt cct gct gat gag cat ttg tct 1108
 Thr Ser His Thr His Leu Asp Val Val Pro Ala Asp Glu His Leu Ser
 350 355 360
 tgg gtt agg tac gct gtg gat cac acc att gat atc gat cct tct cag 1156
 Trp Val Arg Tyr Ala Val Asp His Thr Ile Asp Ile Asp Pro Ser Gln
 365 370 375
 gga tgg gtt aac tgg ttg atg gga tac ttg aac tgc caa gtg att cat 1204
 Gly Trp Val Asn Trp Leu Met Gly Tyr Leu Asn Cys Gln Val Ile His
 380 385 390
 cac ctc ttc cct tct atg cct caa ttc aga caa cct gag gtg tcc aga 1252
 His Leu Phe Pro Ser Met Pro Gln Phe Arg Gln Pro Glu Val Ser Arg
 395 400 405
 aga ttc gtt gct ttc gct aag aag tgg aac ctc aac tac aag gtg atg 1300
 Arg Phe Val Ala Phe Ala Lys Lys Trp Asn Leu Asn Tyr Lys Val Met
 410 415 420 425
 act tat gct gga gct tgg aag gct act ttg gga aac ctc gat aat gtg 1348
 Thr Tyr Ala Gly Ala Trp Lys Ala Thr Leu Gly Asn Leu Asp Asn Val
 430 435 440
 gga aag cac tac tac gtg cac gga caa cat tct gga aag acc gct tga 1396
 Gly Lys His Tyr Tyr Val His Gly Gln His Ser Gly Lys Thr Ala
 445 450 455
 taa ttaattaagg cgcgccgaat tc 1421

<210> 202

<211> 456

<212> PRT

<213> *Ostreococcus tauri*

<400> 202

Met Cys Val Glu Thr Glu Asn Asn Asp Gly Ile Pro Thr Val Glu Ile
 1 5 10 15

Ala Phe Asp Gly Glu Arg Glu Arg Ala Glu Ala Asn Val Lys Leu Ser
 20 25 30

267

Ala Glu Lys Met Glu Pro Ala Ala Leu Ala Lys Thr Phe Ala Arg Arg
 35 40 45
 Tyr Val Val Ile Glu Gly Val Glu Tyr Asp Val Thr Asp Phe Lys His
 50 55 60
 Pro Gly Gly Thr Val Ile Phe Tyr Ala Leu Ser Asn Thr Gly Ala Asp
 65 70 75 80
 Ala Thr Glu Ala Phe Lys Glu Phe His His Arg Ser Arg Lys Ala Arg
 85 90 95
 Lys Ala Leu Ala Ala Leu Pro Ser Arg Pro Ala Lys Thr Ala Lys Val
 100 105 110
 Asp Asp Ala Glu Met Leu Gln Asp Phe Ala Lys Trp Arg Lys Glu Leu
 115 120 125
 Glu Arg Asp Gly Phe Phe Lys Pro Ser Pro Ala His Val Ala Tyr Arg
 130 135 140
 Phe Ala Glu Leu Ala Ala Met Tyr Ala Leu Gly Thr Tyr Leu Met Tyr
 145 150 155 160
 Ala Arg Tyr Val Val Ser Ser Val Leu Val Tyr Ala Cys Phe Phe Gly
 165 170 175
 Ala Arg Cys Gly Trp Val Gln His Glu Gly Gly His Ser Ser Leu Thr
 180 185 190
 Gly Asn Ile Trp Trp Asp Lys Arg Ile Gln Ala Phe Thr Ala Gly Phe
 195 200 205
 Gly Leu Ala Gly Ser Gly Asp Met Trp Asn Ser Met His Asn Lys His
 210 215 220
 His Ala Thr Pro Gln Lys Val Arg His Asp Met Asp Leu Asp Thr Thr
 225 230 235 240
 Pro Ala Val Ala Phe Phe Asn Thr Ala Val Glu Asp Asn Arg Pro Arg
 245 250 255
 Gly Phe Ser Lys Tyr Trp Leu Arg Leu Gln Ala Trp Thr Phe Ile Pro
 260 265 270
 Val Thr Ser Gly Leu Val Leu Leu Phe Trp Met Phe Phe Leu His Pro
 275 280 285

268

Ser Lys Ala Leu Lys Gly Gly Lys Tyr Glu Glu Leu Val Trp Met Leu
290 295 300

Ala Ala His Val Ile Arg Thr Trp Thr Ile Lys Ala Val Thr Gly Phe
305 310 315 320

Thr Ala Met Gln Ser Tyr Gly Leu Phe Leu Ala Thr Ser Trp Val Ser
325 330 335

Gly Cys Tyr Leu Phe Ala His Phe Ser Thr Ser His Thr His Leu Asp
340 345 350

Val Val Pro Ala Asp Glu His Leu Ser Trp Val Arg Tyr Ala Val Asp
355 360 365

His Thr Ile Asp Ile Asp Pro Ser Gln Gly Trp Val Asn Trp Leu Met
370 375 380

Gly Tyr Leu Asn Cys Gln Val Ile His His Leu Phe Pro Ser Met Pro
385 390 395 400

Gln Phe Arg Gln Pro Glu Val Ser Arg Arg Phe Val Ala Phe Ala Lys
405 410 415

Lys Trp Asn Leu Asn Tyr Lys Val Met Thr Tyr Ala Gly Ala Trp Lys
420 425 430

Ala Thr Leu Gly Asn Leu Asp Asn Val Gly Lys His Tyr Tyr Val His
435 440 445

Gly Gln His Ser Gly Lys Thr Ala
450 455