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(54) Titre : METHODE D'AJOUT D'ACYLE AROMATIQUE AUX SUCRES AUX POSITIONS 3' ET 5 D'ANTHOCYANE

(54) Title: A METHOD OF ADDING AROMATIC ACYL GROUPS TO SUGARS AT BOTH THE 3' AND 5 POSITIONS OF ANTHOCYANINS

(57) Abrégé/Abstract:

A method of acylating the 3' position of anthocyanin using an enzyme that transfers an aromatic acyl group to a sugar at the 3' position of anthocyanin or a gene encoding the enzyme.

ABSTRACT

A method of acylating the 3' position of anthocyanin using an enzyme that transfers an aromatic acyl group to a sugar at the 3' position of anthocyanin or a gene encoding the enzyme.

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CECI EST LE TOME 1 DE 2
CONTENANT LES PAGES 1 À 31

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THIS IS VOLUME 1 OF 2
CONTAINING PAGES 1 TO 31

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- 1 -

DESCRIPTION

A METHOD OF ADDING AROMATIC ACYL GROUPS TO SUGARS AT BOTH
THE 3' AND 5 POSITIONS OF ANTHOCYANINS

5 FIELD OF THE INVENTION

The present invention relates to a method of altering anthocyanin bluer and more stable using an enzyme that transfers an aromatic acyl group to the 3'-position of anthocyanin or a gene encoding said enzyme,
10 and can be applied to the alteration and stabilization of anthocyanin pigments and to the alteration and stabilization of flower color. More specifically, it relates to a method of making the color of flowers blue and stabilizing it using an aromatic acyltransferase that
15 transfers an aromatic acyl group to the 3'-position of anthocyanin derived from plants including *Gentiana triflora* var. *japonica* or a cDNA encoding said enzyme.

The present invention also relates to a method of altering anthocyanin bluer and more stable using a single
20 enzyme that transfers aromatic acyl groups to sugars at multiple positions of anthocyanin or gene encoding said enzyme, and can be applied to the alteration and stabilization of anthocyanin pigments and to the alteration and stabilization of flower color.

25 BACKGROUND OF THE INVENTION

The flower industry strives to develop new and different varieties of flowers. An effective way to create such novel varieties is the manipulation of flower color where classical breeding techniques have been used
30 to produce a wide range of colors for most of the commercial varieties. This approach has been limited, however, by the constraints of a particular species' gene pool and for this reason it is rare for a single species to have the full spectrum of colored varieties.

35 Flower color is predominantly due to two types of

- 2 -

pigments: flavonoids and carotenoids. Flavonoids mainly contribute a wide range of color from yellow to red to blue, whereas carotenoids mainly contribute color tones such as orange or yellow. The flavonoids which are a major contribution to flower color are a class of compounds called anthocyanins. The chromophoric group of anthocyanins is anthocyanidins, and as major anthocyanidins, there are known pelargonidin, cyanidin and delphinidin. Plants are known to have a wide variety of anthocyanins, and the diversity thereof is one of the causes of the diversity of flower colors. Structures of hundreds of anthocyanins have already been determined, and the hydroxyl group at the 3 position of most anthocyanins has been modified with sugars (Harbone, in The Flavonoids: 565, 1986).

The biosynthetic pathway for anthocyanins is common among flowering plants up to the biosynthesis of the 3-glucosides (Holton et al., Plant Cell 7: 1071, 1995), and subsequently they undergo various modifications such as glycosylation, acylation and methylation in species- and variety-specific manners. Such differences in modification patterns in varieties are one of the reason for diversities in anthocyanins, i.e. diversities in flower colors. Generally the more aromatic acyl groups modify anthocyanins, the more stabilized and bluer anthocyanins become (Harbone, in The Flavonoids: 565, 1986; Norio Saito, TANPAKUSITU KAKUSAN KOUSO (Proteins, Nucleic Acids, Enzymes) 47: 202, 2002). Furthermore, flower color may be affected by the formation of metal complex of anthocyanins, the copigment effect by flavonoid compounds such as flavonol and flavone, and pH of vacuoles in which anthocyanins are localized (Forkmann, Plant Breeding 106: 1, 1991).

Biosynthesis of flavonoids including anthocyanidin has been extensively studied. All the genes for enzymes involved in anthocyanin biosynthesis have been cloned, and genes for the transcription factors therefor have

also been obtained. Therefore, the artificially
modification of the expression of these genes can alter
the structure and the amount of flavonoids accumulated in
flowers, and thereby can change flower color. There are
5 some reports on the modification of anthocyanin
structures and flower color by a molecular biological
technique and gene transformation into plants (Forkmann
G. & Martens S. (2001), Curr. Opin. Biotechnology, 12:
155-160; Tanaka Y. & Mason J. (2003), In: Singh RP &
10 Jaiwal PK (ed.) Plant genetic engineering, pp. 361-385,
SCI tech publishing, Houston).

One possible method for making flower color blue is
to increase the number of hydroxyl groups of B ring of
anthocyanin. An enzyme that catalyzes a reaction of
15 hydroxylating the 3' position of anthocyanin (flavonoid
3'-hydroxylase: F3'H) and an enzyme that catalyzes a
reaction of hydroxylating the 3' and the 5' position of
anthocyanin (flavonoid 3',5'-hydroxylase: F3'5'H) are
important in altering flower color. In general,
20 pelargonidin (one hydroxyl group in B ring) is contained
in orange- to red-colored flowers, cyanidin (two hydroxyl
groups in B ring) is contained in red- to magenta-colored
flowers, and delphinidin (three hydroxyl groups in B
ring) is contained in purple- to blue-colored flowers.
25 In most cases, plant species that do not have purple- to
blue-colored varieties often lack the ability to produce
delphinidin, and are represented by roses, chrysanthemums
and carnations.

For these plants, the creation of purple- to blue-
30 colored varieties by biotechnology has long attracted
attention. In fact, by expressing the F3'5'H gene
essential for the production of delphinidin, carnations
whose flower color is blue purple were produced (Tanaka Y.
& Mason J. (2003), In: Singh RP & Jaiwal PK (ed.) Plant
35 genetic engineering, pp. 361-385, SCI tech publishing,
Houston), and it became possible to produce delphinidin
in flower petals, but the flower color has not been fully

blue yet. Thus, in order to make flower color sheer blue, the introduction of the F3'5'H gene alone is not sufficient, and further contrivances may be required.

5 Actually anthocyanins contained in blue flowers are often modified with aromatic acyl groups via sugars (Honda & Saito, Heterocycles 56: 633 (2002)). Thus, one possible method of making flower color blue is to modify anthocyanins with aromatic acyl groups such as caffeoyl groups, coumaroyl groups and sinapoyl groups (Tanaka Y. & Mason J. (2003), In: Singh RP & Jaiwal PK (ed.) Plant genetic engineering, pp. 361-385, SCI tech publishing, Houston).

15 Generally, anthocyanin is slightly reddened by glycosylation, and the addition of aromatic acyl groups via sugars makes the color of anthocyanin blue (Forkmann, Plant Breeding 106: 1, 1991). Also, anthocyanin is a compound unstable in neutral solutions, and the stability is enhanced by modification with sugars or acyl groups (Forkmann, Plant Breeding 106: 1, 1991). An experiment 20 using anthocyanins from morning glories (*Pharbitis nil*) revealed that acylated anthocyanins to which an aromatic acyl group such as, for example, coumaric or caffeic acid was bound showed a hypsochromic shift (Dangle et al., Phytochemistry 34: 1119, 1993).

25 As for anthocyanins acylated with aromatic acyl groups, many isolation examples from nature have been reported including awobanin (Goto and Kondo, Angew. Chem. Int. Ed. Engl. 30: 17, 1991) derived from *Commelina communis* (Honda & Saito, Heterocycles 56: 633 (2002)). 30 For example, anthocyanins from blue flowers have multiple aromatic acyl groups as represented by cinerarin (derived from *cineraria*), gentiodelphin (derived from *Gentiana triflora*), heavenly blue anthocyanin (derived from *Pharbitis nil*), ternatin (derived from *Clitoria ternatea*) and lobelinin (derived from *Lobelia*). 35

Cinerarin (Goto et al., Tetrahedron 25: 6021, 1984) derived from *cineraria* (*Senecio cruentus*) has one

aliphatic acyl group and three aromatic acyl groups, and these aromatic acyl groups are reported to contribute to the stabilization of pigments in neutral aqueous solutions (Goto et al., Tetrahedron 25: 6021, 1984).

5 Gentiodelphin (DEL 3G-5CafG-3' CafG) which is a major pigment of *Gentiana triflora* petals has a delphinidin 3-glycoside as the basic backbone, and two side chains comprising one glucose molecule and one caffeic acid molecule on the hydroxyl groups at the 5 position and the

10 3' position. It is reported that the side chains at the 5 and 3' position comprised of sugar-acyl group contributed to a sandwich-type of intra-molecular stacking, resulting in the stabilization of pigments in aqueous solutions (Yoshida et al., Tetrahedron 48: 4313,

15 1992). Furthermore, it has been confirmed that among the two side chains of sugar-acyl group, the glucosylacyl group at the 3' position rather than the 5 position contributes more strongly to the stabilization and blueing of pigments (Yoshida et al., Phytochemistry 54:

20 85, 2000).

The aromatic acyl transfer reaction was first demonstrated in *Silene* (Kamsteeg et al., Biochem. Physiol. Pflanzen 175: 403, 1980), a plant of the family Caryophyllaceae, in 1980, and a similar aromatic acyl

25 transferase activity was also found in the solubilized enzyme fraction of *Matthiola* as well (Teusch et al., Phytochemistry 26: 991, 1986). Subsequently, an anthocyanin 5-aromatic acyltransferase (hereinafter 5AT) that transfers aromatic acyl groups such as caffeic acid

30 and coumaric acid to sugars at the 5 position of anthocyanins was isolated from *Gentiana triflora* (Fujiwara et al., Eur. J. Biochem. 249, 45, 1997), and based on the internal amino acid sequences of the purified enzyme, cDNA that codes for 5AT of *Gentiana*

35 *triflora* was isolated (Fujiwara et al., Plant J., 16, 421, 1998).

Based on this gene, a homolog was isolated from

Torenia (WO 2005/017147), and furthermore based on the amino acid sequence conserved between these enzymes, a Perilla cDNA coding for the enzyme (3AT) that transfers aromatic acyl groups to the sugar at the 3 position of anthocyanins was isolated (Yonekura-Sakibara et al., Plant Cell Physiol 41: 495, 2000). Using the Perilla 3AT gene, the 3AT gene was cloned from lavender of the same family Labiatae (WO 1996/25500).

An enzyme gene that transfers an acyl group to anthocyanidin-3-rutinoside has been obtained from petunia (National Publication of Translated Version (Kohyo) No. 2003-528603). When the Perilla 3AT gene or the torenia 5AT gene was introduced into roses, anthocyanin in which aromatic acyl groups were added to the 3 position or the 5 position was formed in petals, but it failed to significantly alter flower color blue, and the maximum absorption spectra just shifted to the long wavelength by about 1-2 nm.

The reason for this, as reported by Yoshida et al. (Yoshida et al., Tetrahedron 48: 4313, 1992), it was thought that acylation of A ring or C ring such as the 3 or 5 position is not fully effective, and that acylation at the 3' position is necessary for blueing and stabilization of an anthocyanin, and more preferably acylation at multiple positions including the 3' position is necessary. Since there is in fact anthocyanins containing an aromatic acyl group attached to a sugar at the 3' position, the presence of an enzyme (3' AT) that catalyzes a reaction of transferring an aromatic acyl group to a sugar at the 3' position may be postulated. However, there is no report on a measurement for 3' AT reaction and no 3' AT enzyme or a gene encoding for a 3' AT has been isolated so far.

All acyltransferases reported so far act on the 3 position or the 5 position of anthocyanin, and the site specificity of the reaction has been reported to be high (Fujiwara et al., Plant J., 16, 421, 1998; Yonekura-

Sakibara et al., Plant Cell Physiol 41: 495, 2000).

Therefore, the acylation at the 3' position using a known aromatic acyltransferase was thought to be impossible.

There have been no report for an aromatic acyltransferase

5 that have an activity of transferring aromatic acyl groups to multiple positions of anthocyanins. Thus, with the level of conventional technology, it was impossible, for example, to create a recombinant plant and transfer aromatic acyl groups to sugars at the 3' position or
10 multiple positions including the 3' position of anthocyanin. That is, it was impossible to add aromatic acyl groups to sugars at the 3' position or multiple positions including the 3' position of an anthocyanin in order to make a bluer and more stable anthocyanin, and to
15 make bluer and more stable flower color.

Patent document 1: WO 1996/25500

Patent document 2: WO 2005/017147

Patent document 3: National Publication of Translated Version (Kohyo) No. 2003-528603

20 Non-patent document 1: Harbone, in The Flavonoids: 565, 1986

Non-patent document 2: Holton et al., Plant Cell 7: 1071, 1995

Non-patent document 3: Harbone, in The Flavonoids: 25 565, 1986

Non-patent document 4: Norio Saito, TANPAKUSITU KAKUSAN KOUSO (Proteins, Nucleic Acids, Enzymes) 47: 202, 2002

Non-patent document 5: Forkmann, Plant Breeding 106: 30 1, 1991

Non-patent document 6: Forkmann G. & Martens S. (2001), Curr. Opin. Biotechnology, 12: 155-160

Non-patent document 7: Tanaka Y. & Mason J. (2003), In: Singh RP & Jaiwal PK (ed.) Plant genetic engineering, pp. 361-385, SCI tech publishing, Houston
35

Non-patent document 8: Honda & Saito, Heterocycles 56: 633 (2002)

Non-patent document 9: Forkmann, Plant Breeding 106: 1, 1991

Non-patent document 10: Dangle et al.,
Phytochemistry 34: 1119, 1993

5 Non-patent document 11: Goto et al., Tetrahedron 25: 6021, 1984

Non-patent document 12: Yoshida et al., Tetrahedron 48: 4313, 1992

10 Non-patent document 13: Yoshida et al.,
Phytochemistry 54: 85, 2000

Non-patent document 14: Goto and Kondo, Angew. Chem. Int. Ed. Engl. 30: 17, 1991

Non-patent document 15: Kamsteeg et al., Biochem. Physiol. Pflanzen 175: 403, 1980

15 Non-patent document 16: Teusch et al.,
Phytochemistry 26: 991, 1986

Non-patent document 17: Fujiwara et al., Eur. J. Biochem. 249, 45, 1997

20 Non-patent document 18: Fujiwara et al., Plant J., 16, 421, 1998

Non-patent document 19: Yonekura-Sakakibara et al., Plant Cell Physiol 41: 495, 2000

DISCLOSURE OF THE INVENTION

25 As described in the above report by Yoshida et al.,
the aromatic acyl groups of anthocyanin contribute to the
stabilization and blueing of anthocyanin, and
specifically the sugar-acyl group side chain at the 3'
position contributes more strongly than that at the 5
position. It is also believed that the sugar-acyl group
30 side chain at multiple positions including the 3'
position make anthocyanins bluer and more stable. Thus,
by using an enzyme that transfers an aromatic acyl group
to the 3' position of anthocyanin or multiple positions
including the 3' position, or a gene encoding said
35 enzyme, it seems to be possible to artificially modify
anthocyanins and alter anthocyanins to more stable
compounds, or to increase bluish hue of anthocyanins.

As described above, the transfer of aromatic acyl groups to the 3' position is very effective for the stabilization and blueing of anthocyanins. For that purpose, an aromatic acyltransferase that transfers aromatic acyl groups to the 3' position of anthocyanin or a gene encoding the enzyme is essential. The present inventors have investigated in detail the enzymatic properties of the anthocyanin 5-aromatic acyltransferase isolated from *Gentiana triflora*, and demonstrated that the 5-aromatic acyltransferase of *Gentiana triflora* also have an activity of 3'-acyl transfer. Thus, we clarified that, in spite of a single enzyme, the enzyme catalyzes the aromatic acyltransferring reactions to sugars at both of the 5 and the 3' positions of anthocyanins.

Thus, the present invention provides a method of making a bluer and more stable anthocyanin by adding an aromatic acyl group to the 3' position of anthocyanin. It also provides a method of making flower color bluer and more stable by introducing and expressing a gene encoding an aromatic acyltransferase into plants.

Thus, the present invention (1) provides a method of acylating the 3' position of anthocyanins using an enzyme that transfers an aromatic acyl group to a sugar at the 3' position of anthocyanin or a gene encoding the enzyme.

(2) The present invention also provides a method of stabilizing anthocyanins by using an enzyme that transfers an aromatic acyl group to a sugar at the 3' position of anthocyanin or a gene encoding the enzyme.

(3) The present invention also provides a method of blueing anthocyanins by using an enzyme that transfers an aromatic acyl group to a sugar at the 3' position of anthocyanin or a gene encoding the enzyme.

(4) The present invention further provides a method of acylating a pigment of interest by expressing a gene encoding an aromatic acyltransferase that transfers an aromatic acyl group to the 3' position of anthocyanin in plants.

5 (5) The present invention further provides a method of stabilizing a pigment of interest by introducing a gene encoding an aromatic acyltransferase that transfers an aromatic acyl group to the 3' position of anthocyanin, and acylating the pigment of interest in plants.

10 (6) The present invention also provides a method of blueing a pigment of interest by introducing a gene encoding an aromatic acyltransferase that transfers an aromatic acyl group to the 3' position of anthocyanin, and acylating the pigment of interest in plants.

15 (7) The present invention further provides a plant obtained by a method described in any of the above (4) - (6), a vegetative propagation product or a seed of a plant, or a progeny plant of a plant, a vegetative propagation product or a seed of a plant having properties identical to those of a plant.

20 (8) The present invention also provides a method of adding aromatic acyl groups to sugars at multiple positions of an anthocyanin, which comprises using a single enzyme that transfers aromatic acyl groups to sugars at multiple positions of an anthocyanin or a gene encoding the enzyme.

25 (9) The present invention further provides a method of stabilizing anthocyanin, which comprises using a single enzyme that transfers aromatic acyl groups to sugars at multiple positions of an anthocyanin or a gene encoding the enzyme.

30 (10) The present invention further provides a method of blueing anthocyanin, which comprises using a single enzyme that transfers aromatic acyl groups to sugars at multiple positions of an anthocyanin or a gene encoding the enzyme.

35 (11) The present invention further provides a method according to any of the above (8) - (10) wherein one of the above multiple positions is the 3' position of an anthocyanin.

(12) The present invention also provides a method of

- 11 -

acylating a pigment of interest by expressing a single enzyme that has activities of transferring aromatic acyl groups to sugars at multiple positions of an anthocyanin or a gene encoding the enzyme in plants.

5 (13) The present invention also provides a method of stabilizing a pigment of interest by introducing a single enzyme that has activities of transferring aromatic acyl groups to sugars at multiple positions of an anthocyanin or a gene encoding the enzyme, and acylating the pigment
10 of interest in plants.

(14) The present invention also provides a method of blueing a pigment of interest by introducing a single enzyme that has activities of transferring aromatic acyl groups to sugars at multiple positions of anthocyanin or
15 a gene encoding the enzyme, and acylating the pigment of interest in plants.

(15) The present invention further provides a method according to any of the above (12) - (14) wherein one of the above multiple positions is a sugar at the 3'
20 position of an anthocyanin.

(16) The present invention also provides a plant obtained by an method described in any of the above (12) - (15), a vegetative propagation product or a seed of a plant, or a progeny plant of a plant, a vegetative
25 propagation product or a seed of a plant having properties identical to those of the plant.

(17) The present invention provides a gene encoding a protein having an amino acid sequence as set forth in SEQ ID NO: 4 or 6 and having an activity of transferring
30 an aromatic acyl group to a sugar at the 3' position of anthocyanin, or a gene encoding a protein having a sequence identity of 70% or greater to an amino acid sequence and having an activity of transferring an aromatic acyl group to a sugar at the 3' position of
35 anthocyanin, or a gene encoding a protein having a sequence identity of 70% or greater to the nucleotide sequence as set forth in SEQ ID NO: 3 or 5 and having an

- 12 -

activity of transferring an aromatic acyl group to a sugar at the 3' position of anthocyanin.

5 (18) The present invention provides a gene encoding a protein having an amino acid sequence as set forth in SEQ ID NO: 4 or 6 and having an activity of transferring aromatic acyl groups to sugars at multiple positions of an anthocyanin, or a gene encoding a protein having a sequence identity of 70% or greater to an amino acid sequence and having an activity of transferring aromatic
10 acyl groups to sugars at multiple positions of an anthocyanin, or a gene encoding a protein having a sequence identity of 70% or greater to the nucleotide sequence as set forth in SEQ ID NO: 3 or 5 and having an activity of transferring aromatic acyl groups to sugars
15 at multiple positions of an anthocyanin.

(19) The present invention also provides the gene according to the above (18) wherein one of the multiple positions is a sugar at the 3' position of an anthocyanin.

20 (20) The present invention also provides a vector comprising the gene according to any of the above (17) - (19).

(21) The present invention also provides a host transformed with the vector according to the above (20).

25 (22) The present invention also provides a protein encoded by the gene according to any of the above (17) - (19).

(23) The present invention also provides a method of producing a protein having an activity of transferring a
30 sugar to the 3' position of a flavonoid, which method comprises culturing or growing the host according to the above (21), and harvesting the protein from the host.

(24) The present invention also provides a plant in which the gene according to any of the above (17) - (19)
35 has been introduced, or a progeny having properties identical thereto, or a tissue thereof.

(25) The present invention also provides a cut

- 13 -

flower of the plant according to the above (24) or a cut flower of a progeny having properties identical thereto.

5 (26) The present invention also provides a method of acylating the 3' position of anthocyanin, which method comprises using the gene according to any of the above (17) - (19).

(27) The present invention also provides a method of stabilizing anthocyanin, which method comprises using the gene according to any of the above (17) - (19).

10 (28) The present invention also provides a method of blueing anthocyanin, which method comprises using the gene according to any of the above (17) - (19).

15 (29) The present invention further provides a method of expressing the gene according to any of the above (17) - (19) in a plant and acylating the pigment of interest in the plant.

(30) The present invention also provides a method of stabilizing a pigment of interest which comprises introducing the gene according to any of the above (17) - 20 (19) to a plant and acylating the pigment of interest in the plant.

25 (31) The present invention also provides a method of blueing a pigment of interest which comprises introducing the gene according to any of the above (17) - (19) to a plant and acylating the pigment of interest in the plant.

(32) The present invention further provides a method of adding aromatic acyl groups to sugars at multiple positions of an anthocyanin, which comprises using the gene according to any of the above (17) - (19).

30 (33) The present invention further provides the method according to the above 32 wherein one of the multiple positions is the 3' position of anthocyanin.

BRIEF EXPLANATION OF THE DRAWINGS

35 Fig. 1 shows the structural formulas, names and abbreviations of anthocyanin compounds related to the present invention.

- 14 -

Fig. 2 shows the structural formulas, names and abbreviations of anthocyanin compounds related to the present invention.

5 Fig. 3 is a graph showing time course changes of reaction products when 50 μM of DEL 3G-5G-3'G was used as the substrate. In the figure, triG represents DEL 3G-5G-3'G, 5Caf represents DEL 3G-5CafG-3'G, 3'Caf represents DEL 3G-5G-3'CafG, and 5,3'Caf represents gentiodelphin (DEL 3G-5CafG-3'CafG).

10 Fig. 4 is a graph showing time course changes of reaction products when 100 μM of DEL 3G-5G-3'G was used as the substrate. In the figure, triG represents DEL 3G-5G-3'G, 5Caf represents DEL 3G-5CafG-3'G, 3'Caf represents DEL 3G-5G-3'CafG, and 5,3'Caf represents Gentiodelphin (DEL 3G-5CafG-3'CafG).

15 Fig. 5 is a graph showing time course changes of reaction products when 200 μM of DEL 3G-5G-3'G was used as the substrate. In the figure, triG represents DEL 3G-5G-3'G, 5Caf represents DEL 3G-5CafG-3'G, 3'Caf represents DEL 3G-5G-3'CafG, and 5,3'Caf represents Gentiodelphin (DEL 3G-5CafG-3'CafG).

20 Fig. 6 shows the result of SDS-PAGE and Western blot of a protein partially purified from the petal of *Gentiana triflora*. The left figure shows the result of SDS-PAGE and the right figure shows the result of Western blot against the GAT4 antibody. In the figure, M represents a molecular marker, lane 1 represents the result of 40 - 70% ammonium sulfate-saturated precipitate, and lane 2 represents the result of the active fraction after the Dyematrix column.

30 Fig. 7 shows a result of flower color simulation using the Medio squeeze liquid.

BEST MODE FOR CARRYING OUT THE INVENTION

35 Though the present invention describes a method that employs an aromatic acyltransferase that transfers an

- 15 -

aromatic acyl group to the 3'-position of anthocyanin or a gene encoding enzyme, genes used and proteins encoded by the gene are not limited to them. Proteins having an amino acid sequence that has been modified by the
5 addition or deletion of a plurality of amino acids, or by replacement with other amino acids are known to maintain the enzyme activity similar to that of the original protein. Thus, as long as the activity of transferring an aromatic acyl group to a sugar at the 3' position has
10 been maintained, protein having an amino acid sequence that has been modified by the addition or deletion of one or a plurality of amino acids, or by replacement with other amino acids, and a gene encoding the protein are also encompassed by the present invention.

15 The present invention also encompasses a gene encoding a protein that has an amino acid sequence identity of 70% or greater, preferably 90% or greater to the amino acid sequence of the aromatic acyltransferase that transfers an aromatic acyl group to the 3' position
20 of anthocyanin derived from *Gentiana triflora*, amino acid sequence as set forth in SEQ ID NO: 4 or 6, and that has an activity of transferring an aromatic acyl group to a sugar at the 3' position of anthocyanin.

The present invention also encompasses cases which
25 use a gene hybridizing to a gentian DNA encoding an aromatic acyltransferase for a sugar at the 3' position of anthocyanin under a relatively mild condition of 5 x SSC and 50°C, and encoding a protein with an activity of transferring an aromatic acyl group to the 3' position.
30 Furthermore, the present invention also encompasses cases which use a gene hybridizing to a gentian DNA encoding an 3'-aromatic acyltransferase under a stringent condition, and encoding a protein with an activity of transferring an aromatic acyl group to the 3' position.

35 Although the stringent condition as used herein is, for example, 2 x SSC and 65°C, it is not limited to this condition since the hybridization condition varies

- 16 -

depending on the length and the base composition of DNA used. Genes selected by such hybridization include naturally occurring ones, for example genes derived from plants containing anthocyanin to which an aromatic acyl group has been added at the 3' position, for example genes derived from *Clitoria ternatea*, *lobelia* or *cineraria*, but not limited to those derived from plants. Thus, any genes that encode enzymes having an activity of transferring an aromatic acyl group to the 3' position of anthocyanin may be used. The gene selected by hybridization may be cDNA or genomic DNA.

It is also possible to purify aromatic acyltransferases that transfer aromatic acyl groups to the 3' position from plants such as *lobelia* and *Clitoria ternatea* that contain anthocyanin to which an aromatic acyl group has been added to the 3' position by the purification method per se of the enzyme from the *Gentiana triflora* or modifying the method. Furthermore, by determining the amino acid sequence of the purified enzyme, a gene encoding said enzyme can be cloned.

DNA encoding a protein having the altered amino acid sequence can be synthesized using a known site-directed mutagenesis or a PCR method. For example, a DNA fragment of which an amino acid sequence is desired to be altered may be obtained by obtaining cDNA or genomic DNA by restriction enzyme treatment, and, with this as a template, using primers corresponding to the alteration of the desired amino acid sequence, and performing site-directed mutagenesis or a PCR method to obtain a DNA fragment corresponding to the alteration of the desired amino acid sequence. Then, the alteration-introduced DNA fragment may be ligated to a DNA fragment encoding another portion of the enzyme of interest.

Alternatively, in order to obtain a DNA that encodes an enzyme comprising a shortened amino acid sequence, for example, an amino acid sequence longer than the amino acid sequence of interest, for example a full-length

- 17 -

amino acid sequence, may be cleaved with the desired restriction enzyme, and if the resulting DNA fragment does not encode the entire amino acid sequence of interest, a DNA fragment corresponding to the amino acid sequence of the lacking part may be synthesized and ligated. By expressing the gene thus obtained in the *Escherichia coli* (*E. coli*) or a yeast expression system, and measuring the activity of transferring an aromatic acyl group to the 3' position in said *E. coli* or yeast extract, the gene obtained may be confirmed to encode an aromatic acyltransferase. A DNA encoding the amino acid sequence of interest may also be synthesized.

The present invention also encompasses cases in which aromatic acyltransferase extracted from recombinant vectors, specifically expression vectors, and host cells transformed with said vectors. As host cells, prokaryotes or eukaryotes may be used. As prokaryotes, there can be used conventional known host cells including, for example, bacteria belonging to genus *Escherichia* such as *Escherichia coli*, microorganisms belonging to genus *Bacillus* such as *Bacillus subtilis*, and the like. As eukaryotes, there can be used, for example, eukaryotic microorganisms, preferably yeast or filamentous fungi.

As the yeast, there can be mentioned yeast of the genus *Saccharomyces* such as *Saccharomyces cereviceae*, and as the filamentous fungi, there can be mentioned microorganisms of the genus *Aspergillus* such as *Aspergillus oryzae* and *Aspergillus niger*, and of the genus *Penicillium*. Furthermore, animal cells or plant cells may be used, and as the animal cells, cell systems such as mice, hamsters, and human cells may be used. Furthermore, insect cells such as silkworm cells or silkworm larvae per se may be used as the host.

Expression vectors may contain expression control regions such as promoters and terminators and replication origins depending on the species of the host into which

they are to be introduced. As the promoters of expression vectors for bacteria such as *E. coli*, there can be used conventionally known promoters such as a *trc* promoter, *tac* promoter, and *lac* promoter. As the
5 promoters for yeast, there can be used, for example, the glyceraldehyde-3-phosphate dehydrogenase promoter and the PH05 promoter, and the promoters for the filamentous fungi include, but not limited to, promoters such as amylase and *trpC*. Also as the promoters for animal
10 cells, there can be used viral promoters such as the SV40 early promoter and the SV40 late promoter.

Expression vectors may be prepared using restriction enzymes, ligases and the like according to standard methods. Transformation of host cells with expression
15 vectors may also be conducted according to conventionally known methods. As the expression vectors for plants, there can be used binary vectors such as pBI121 when *Agrobacterium* is used, and *E. coli* vectors such as pUC19 when particle guns are used. Furthermore, plant cells
20 that were transformed with said expression vector may be selected with a marker gene such as an antibiotics-resistant gene, and redifferentiated using a condition of a suitable plant hormone, etc. to obtain transformed plants.

25 The use of aromatic acyltransferase capable of transferring an aromatic acyl group to the 3' position of anthocyanin obtained by culturing or cultivating the thus transformed host cells or transformed plants and recovering and/or purifying from the culture by standard
30 methods such as filtration, centrifugation, cell disruption, gel filtration chromatography and ion exchange chromatography is also encompassed by the present invention.

35 With the present state of technology in the art, it is possible to introduce a gene into plants and express the gene in a constitutive or tissue-specific manner, and the expression of the gene of interest can also be

suppressed by the antisense method or the cosuppression method. Examples of plants that can be transformed include, but not limited to, roses, chrysanthemums, carnations, calceolarias, cyclamens, orchids, prairies
5 gentians, freesias, gerberas, gladioluses, gypsophyllas, kalanchoes, lilies, pelargoniums, geraniums, petunias, torenias, tulips, rice, barley, wheat, rapeseed, potatoes, tomatoes, poplar, bananas, eucalyptus, sweet potatoes, soy beans, alfalfa, lupin, corn, cauliflower
10 and the like.

Thus, those used in the method of aromatic acylation to the 3' position of anthocyanin are not limited to acyltransferases derived from *Gentiana triflora*, or cDNA or gene encoding the enzyme from *Gentiana triflora*, or a
15 recombinant enzyme obtained by expressing the cDNA or gene in a host such as *E. coli*, and aromatic acyltransferases can also be used to transfer aromatic acyl groups to the 3'-position of anthocyanin obtained from a wide variety of other organisms, cDNA or gene
20 encoding the enzymes, or recombinant enzymes obtained by expressing the cDNA or gene in a host such as *E. coli*.

Furthermore, though the present invention used CoA esters such as p-coumaroyl CoA and caffeoyl-CoA as the donor for the acyl group, p-coumaroyl or hydroxy
25 cinnamoyl-1-O-glucose such as cinnapoyl-1-O-glucose may also be used as the donor for the acyl group (Glassgen and Seitz, *Planta* 186: 582, 1992), and thus these may be used as the enzyme claimed in the present invention.

EXAMPLES

30 The present invention will now be explained in more detail with reference to specific embodiments. Unless otherwise specified, experimental procedures were as described in Sambrook et al., *Molecular Cloning* (Cold Spring Harbor Laboratory Press, 1989), PCT-JP96-00348,
35 and reports by Fujisawa et al. (1997, 1998).

Example 1. Expression of cDNA of *Gentiana triflora*-

- 20 -

derive acyl transferase in Escherichia coli and
purification of recombinant protein

By cleaving a construct pGeAT102 (Fujiwara et al.,
Plant J., 16, 421, 1998) for E. coli expression of cDNA
of Gentiana triflora-derived acyltransferase with
NcoI/HindIII, a fragment containing the coding region of
the acyltransferase and the 3'-untranslated region was
subcloned to the NcoI/HindIII site of E. coli expression
vector pQE60 (QIAGEN) to obtain a construct pQE8 for E.
coli expression.

An E. coli strain JM109, in which pQE8 was
introduced was cultured in a SB medium at 37°C to OD600nm
= 0.8, and then further cultured at a reduced temperature
of 15°C for 1 hour, to which IPTG was added to a final
concentration of 0.1M to induce the expression of the
gene of Gentiana triflora acyltransferase. After
culturing at 15°C for 1 hour, cells were collected and
sonication-disrupted, and used in the following
purification. The disrupted cells were subjected to DE52
(Whatman), and the flow-through fraction with 25 mM Tris-
HCl (pH 7.5) containing 150 mM NaCl was collected.
Salting out with ammonium sulfate was further performed
to collect a 40-60% ammonium sulfate-saturated fraction,
which was dissolved in a small amount of 20 mM Tris-HCl
(pH 7.5) and dialyzed using Sephadex G-25 (Pharmacia)
equilibrated with 20 mM Tris-HCl (pH 7.5).

After this was loaded to DEAE-TOYOPEARL (TOSOH
Corporation), and eluted with a linear gradient of 0 to
0.5 M NaCl-containing in 20 mM Tris-HCl (pH 7.5), an
activity of 5-acyltransferase to delphinidin 3,5-
diglucoside (DEL 3G-5G) was present in the eluted
fraction of 120 - 240 mM NaCl. The active fraction was
dialyzed against a 20 mM Tris-HCl (pH 7.5), and then
loaded to Blue Sepharose (Pharmacia). No activity was
found in the flow-through fraction of Blue Sepharose, and
the adsorbed fraction was eluted with 20 mM Tris-HCl (pH

- 21 -

7.5) containing 1 M NaCl.

Subsequently, this fraction was adsorbed to Phenyl Sepharose (Pharmacia), and eluted with a linear gradient of 40 to 0% ammonium sulfate in 20 mM Tris-HCl (pH 7.5). The active fraction was eluted with 0% ammonium sulfate. This was dialyzed against 20 mM Tris-HCl (pH 7.5), then allowed to adsorb to Dyematrix column Orange A (Amicon), and eluted with 10 mM Tris-HCl (pH 7.5) containing 0.3 M NaCl and 0.5 mM DTT. No activity was found in the flow-through fraction, and the activity was only found in the adsorbed fraction, which was concentrated with Centricon-10 (Amicon).

Example 2. Measurement of activity of recombinant acyltransferase

The enzymatic activity of the recombinant enzyme obtained in Example 1 was measured with 3 types of delphinidin derivatives (delphinidin 3,5,3'-triglucoside (DEL 3G-5G-3'G), delphinidin 3-glucosyl-5-caffeoylglucosyl-3'-glucoside (DEL 3G-5CafG-3'G), delphinidin 3-glucosyl-5-glucosyl-3'-caffeoylglucoside (DEL 3G-5G-3'CafG)) shown in Fig. 1 and Fig. 2 as substrates. The reaction mixture contained 100 mM potassium phosphate buffer (pH 8.5), 0.2 mM caffeoyl-CoA, 250 mM each of the above substrates and 5 μ l of the enzyme solution in a total volume of 50 μ l, and the reaction was carried out at 30°C for 15 minutes. An equal volume of 90% acetonitrile solution containing 0.1% trifluoroacetic acid (TFA) was added to the reaction mixture to stop the reaction.

In order to analyze time course changes of reaction products, 5 μ l of a 25-fold diluted enzyme solution and each of 50 μ M, 100 μ M or 200 μ M of DEL 3G-5G-3'G were used as a substrate, and the reaction was stopped at 2.5, 5, 10, and 20 minutes later to analyze the products. The reaction products were analyzed with a reverse phase high performance liquid chromatography (HPLC) using a DE-41

column (4.6 × 250 mm, Shodex). Samples were eluted with an liner gradient of 20-50% of acetonitrile containing 0.5% TFA at 0.6 ml/min for 15 minutes, and then isocratic elution at 0.6 ml/min for 10 minutes, and detected with SPD-M10A (SHIMAZU Corporation) at a wavelength of 250 - 600 nm. The HPLC elution time and absorption spectra of products were compared with those of authentic samples to identify the structure of products.

As a result of reaction using the above three types of compounds as the substrate, the recombinant acyltransferase reacted with all of the substrates. With 250 mM DEL 3G-5CafG-3'G as the substrate, 94.1% was converted to DEL 3G-5CafG-3'CafG. On the other hand, with 250 mM DEL 3G-5G-3'CafG as the substrate, 95.2% was converted to DEL 3G-5CafG-3'CafG. When 250 mM DEL 3G-5G-3'G was used as a substrate, 7.2% was converted to DEL 3G-5CafG-3'G to which an aromatic acyl group was added at the 5 position alone, and 58.7% was converted to DEL 3G-5CafG-3'CafG, while only the trace amount of DEL 3G-5G-3'CafG was produced, to which an aromatic acyl group was added at the 3' position alone (1% or less).

When the time course changes of the reaction was measured using a diluted enzyme solution with DEL 3G-5G-3'G as a substrate, the amount produced of DEL 3G-5CafG-3'G and DEL 3G-5CafG-3'CafG increased according to the reaction time. The amount of DEL 3G-5G-3'CafG was much smaller than the other two substrates, and barely detected when the amount of the substrate and the reaction time were increased (Fig. 3, Fig. 4, and Fig. 5). This result consistent with those obtained when the undiluted enzyme solution was reacted to DEL 3G-5G-3'G.

From the above result, it was revealed that a recombinant protein obtained by expressing the gene of an acyltransferase derived from *Gentiana triflora* in *E. coli* has an activity of transferring acyl groups to sugars on both the 5 position and the 3' position of anthocyanin. Thus, although a previous report (Fujiwara et al., Plant

J., 16, 421, 1998) showed that this enzyme was thought to transfer an aromatic acyl group only to the 5 position of anthocyanin, the present invention revealed that this enzyme transfers aromatic acyl groups to sugars at both of the 5 and the 3' position of anthocyanin.

Furthermore, considering the reaction products with DEL 3G-5G-3'G as a substrate, it is likely that the addition of an aromatic acyl group to the 5-glucose precedes the addition to the 3'-glucose.

Example 3. Purification of aromatic acyl group transferase derived from Gentiana triflora

The recombinant protein obtained by expressing a gentian cDNA of an aromatic acyltransferase in *E. coli* turned out to have an activity of transferring aromatic acyl groups to glucoses at the 5 and the 3' position of a anthocyanin (Example 2). In order to confirm that an enzyme naturally occurring in gentian petals also has both activities of the 5-aromatic acyltransferase and the 3'-aromatic acyl transferase, this enzyme was purified from gentian petals. In a series of purification, as described in Example 2, each eluted fraction of column chromatography was measured for 5-aromatic acyltransferase activity with DEL 3G-5G as a substrate and for 3'-aromatic acyltransferase activity with DEL 3G-5G-3'G as a substrate.

According to a report by Fujiwara et al. (Fujiwara et al., *Eur. J. Biochem.* 249: 45, 1997) in which a gentian 5-aromatic acyltransferase was purified, 40 - 70% ammonium sulfate-saturated fraction was obtained from extract of approximately 100 g of *Gentiana* petals. This fraction was dialyzed with Sephadex G-25 (Pharmacia) equilibrated with 20 mM Tris-HCl (pH 7.0) containing 10 μ M p-aminophenylmethanesulfonyl fluoride (APMSF) and 1 mM DTT(hereinafter referred to as the Tris buffer), then it was loaded to MONO Q (Pharmacia) equilibrated with the Tris buffer. The unadsorbed fraction was washed off with the Tris buffer, and then eluted with a liner gradient of

- 24 -

0 - 0.5 M NaCl in the Tris buffer at a flow rate of 5 ml/min for 20 minutes. The activity of transferring an aromatic acyl group was present in fractions eluted at 0.2 - 0.42 M NaCl.

5 The active fractions were loaded to HiTrap Blue (Pharmacia) and extensively washed in the Tris buffer, and then the adsorbed fraction was eluted with a Tris buffer containing 0.9 M NaCl. The activity was present in the adsorbed fraction. Then, the active fraction was
10 loaded to a DEAE-Sepharose (Pharmacia). After extensively washing with the Tris buffer, the adsorbed fraction was eluted with a linear gradient of 0-0.5M NaCl in the Tris buffer at 0.5 ml/min for 60 minutes. The activity was found in the fractions eluted with 0.22 -
15 0.3 mM NaCl. The active fraction concentrated with Centricon 30 (Amicon) was loaded to a Dyematrix column Red (Amicon). After washing off the unabsorbed fraction with the Tris buffer, the adsorbed fraction was eluted with the Tris buffer containing 1.5 M KCl. When the
20 eluted protein was subject to SDS-PAGE, only a single band was detected at molecular weight of 52 kDa which is an estimated molecular weight for an anthocyanin 5-acyltransferase.

25 In a series of purifications, each eluted fraction of column chromatography was measured for the 5-aromatic acyltransferase activity with DEL 3G-5G as a substrate and for the 5, 3'-aromatic acyltransferase activity with DEL 3G-5G-3'G as a substrate, and the fractions with the highest activity in both acyltransferase reactions were
30 completely identical. When activities for three types of substrates (DEL 3G-5G-3'G, DEL 3G-5CafG-3'G, DEL 3G-5G-3'CafG) were measured using the active fraction that adsorbed to the Dyematrix column Red as in Example 2, all of the substrates reacted and exhibited characteristics
35 similar to those of Example 2.

 After the active fraction that adsorbed to the Dyematrix column Red was separated with SDS

polyacrylamide gel electrophoresis, the separated protein was transferred to a nitrocellulose membrane Hybond-ECL (Amersham) according to the method described before (Towbin et al., Proc. Natl. Acad. Sci. 76: 4350, 1979), and crossed against an antibody specific to a gentian 5-anthocyanin acyltransferase (Fujiwara et al., Eur. J. Biochem. 249: 45, 1997) in a Western blotting, which detected one clear band (Fig. 6).

These results revealed that the anthocyanin 3'-acyltransferase activity and the anthocyanin 5-acyltransferase activity present in gentian petals were derived from a single protein.

Example 4. Stabilization and bluing of anthocyanin with an acyltransferase derived from *Gentiana triflora*

Relative stability of DEL 3G-5G-3'G, DEL 3G-5CafG-3'G, DEL 3G-5G-3'CafG and DEL 3G-5CafG-3'CafG has already been reported; in an aqueous solution of pH 6.5, DEL 3G-5CafG-3'CafG is most stable and refractory to fading, followed by DEL 3G-5G-3'CafG, DEL 3G-5CafG-3'G, and DEL 3G-5G-3'G in this order (Yoshida et al., Phytochemistry 54: 85, 2000). As for the absorbance maximum, that for DEL 3G-5CafG-3'CafG is the longest wavelength followed by DEL 3G-5G-3'CafG, DEL 3G-5CafG-3'G and DEL 3G-5G-3'G in this order. Thus, this report shows that DEL 3G-5CafG-3'CafG exhibit the most blueish in surface followed by DEL 3G-5G-3'CafG (Yoshida et al., Phytochemistry 54: 85, 2000).

In order to simulate the flower color when the gene for gentian anthocyanin 5,3'-acyltransferase was introduced into roses and DEL 3G-5CafG-3'CafG was accumulated in rose petals, the color development of a purified pigment suspended in juice squeezed from rose petals (cv. Medeo) was measured. As for purified pigments, DEL 3G-5CafG-3'CafG, and as a comparative control, DEL 3G-5G, cyanidin 3,5-diglucoside (CYA 3G-5G), pelargonidin 3,5-diglucoside (PEL 3G-5G) and malvidin 3,5-diglucoside (MAL 3G-5G) were used. About 20 g of

Medeo petals frozen at -80°C for more than one hour were squeezed by a garlic squeezer for household use, and centrifuged at 1000 rpm for 1 minute to remove the debris of the petals, and the supernatant was prepared as the squeezed juice.

Twenty μl of 50 mM DMSO solution of purified pigment was added to 1 ml of the squeezed juice, and kept for 10 minutes, and the absorption and transmittance spectra at 380 - 780 nm were measured with a spectrophotometer UV-2500PC (SHIMADZU Corporation). The transmittance spectrum values were converted into CIE $L^*a^*b^*$ color system (JISZ8729). The Royal Horticultural Society color chart (RHSCC) number was referenced based on the color value (CIE $L^*a^*b^*$ color system) to check the approximate colors using the color classification system Version 2.1.1 (The Japan Research Institute, Co. Ltd., Japan; Japanese Unexamined Patent Publication No. 2002-016935). By using this system, an approximate RHSCC number can be objectively selected. The final concentration of the pigment added to the squeezed juice approximately identical to the average anthocyanin concentration in the vacuoles of rose petals. However, since the absorbance for 3G-5CafG-3'CafG was too high, it was diluted 4-fold prior to measurement. Medeo is a variety that shows an average petal pH (pH 4.38) among the garden species of roses.

As shown in Fig. 7, DEL 3G-5CafG-3'CafG exhibited the largest maximum absorption spectrum among five purified pigments. The approximate color was 89A in the Royal Horticultural Society color chart (RHSCC), which was obtained based on the $L^*a^*b^*$ value converted from the transmission spectrum, and the DEL 3G-5CafG-3'CafG exhibited the strongest blue color among the five purified pigments as shown in Fig. 7.

From this result, it seems to be possible to produce DEL 3G-5CafG-3'CafG in rose petals and as a result to produce rose flowers with blue color by coexpressing the

- 27 -

genes for anthocyanin 5,3'-aromatic acyltransferase, the F3'5'H gene (WO 2004/020637) and the gene of 3'-glycosyltransferase (WO 2001/92509) in rose petals,. Also it seems to be possible to create blue flower varieties in carnations, chrysanthemums, petunias, verbenas, nierenbergias, lilies and so on.

Example 5. Isolation of 5,3'-position aromatic acyltransferase homolog of anthocyanin from Gentiana triflora-related species

In the genus *Gentiana*, there are various related species such as *Gentiana rubicunda* and *Gentiana yakushimensis* in addition to *Gentiana triflora*. Isolation of 5,3'- aromatic acyltransferase homolog from these species was attempted by PCR.

Since the gene for gentian 5,3'-aromatic acyltransferase was known to contain no introns, genomic DNAs extracted from the leaves of the related species were used as templates in PCR with primers specific for the gene of 5,3'-aromatic acyltransferase. An QIAGEN's DNeasy kit was used for the extraction of genomic DNA following a method recommended by the manufacturer. Primers, GAT4-Met-F and GAT4-B, specific for the gene of 5,3'-acyltransferase had the following sequences, and the full-length cDNA containing the entire coding region can be amplified by PCR using these primers. The reaction condition for PCR is as described below.

Sequences of primers:

GAT4-Met-F: TCA TTA TGG AGC AAA TCC AAA (SEQ ID NO: 1)

GAT4-B: CAT GTC AGG TGT GAG GTT CAA C (SEQ ID NO: 2)

PCR condition:

Denaturation reaction: 94°C for 5 minutes, 1 cycle

Amplification reaction: 94°C for 1 minute, 55°C for 1 minute 30 seconds, 72°C for 3 minutes, 30 cycles

Extension reaction: 72°C for 7 minutes, 1 cycle

- 28 -

In this PCR, a band at expected size, 1.5kb, was amplified in three related species, *Gentiana yakushimensis*, *ochroleuca* and *wutaiensis*. These fragments were collected and cloned into a pCRII-TOPO (Invitrogen) and their nucleotide sequences were determined. The nucleotide sequences and the corresponding amino acid sequences of the amplified fragments obtained from each species are shown in Sequence Listing.

10 *Gentiana yakushimensis*: SEQ ID NO: 3 and 4
 Gentiana ochroleuca and *wutaiensis*: SEQ ID NO: 5 and 6

 Fragments obtained from *ochroleuca* and *wutaiensis* turned out to encode the identical amino acid sequence. The identity with 5,3'-aromatic acyltransferase first obtained from *Gentiana triflora* was 95% for that from *Gentiana yakushimensis* and 90% for those obtained from *ochroleuca* and *wutaiensis*. From this high identity, the proteins encoded in these DNAs seem to be homologs to 5,3'-aromatic acyltransferase.

20 Example 6. Expression of 5,3'-aromatic acyltransferase of anthocyanin from *Gentiana triflora* in *nierembergia*

 The gene for 5,3'- aromatic acyltransferase from *Gentiana triflora* was introduced into *neirembergia* together with the genes for gentian 3'-glucosyltransferase and for pansy F3'5'H gene. In transformants, it is expected that the 3' position of DEL 3G-5G was first glucosylated by gentian 3'-glucosyltransferase to form DEL 3G-5G-3'G, on which gentian 5,3'-aromatic acyltransferase may act to form a final product gentiodelphin (DEL 3G-5CafG-3'CafG).

30 A expression construct pSPB1536 was prepared by introducing an expression cassette of 3'-glucosyltransferase into the HindIII and the EcoRI sites of a binary vector for expression in plants (van Engelen FA et al. (1995) Transgenic Res. 4: 288-290), an expression cassette of pansy F3'5'H into the PacI site

and an expression cassette of 5,3'-aromatic acyltransferase into the AscI site. Any of the expression cassettes is regulated by the 35S promoter derived from a cauliflower mosaic virus, and has the Agrobacterium-derived nopaline synthase terminator sequence downstream to each structural gene.

Transformation of *nierembergia* was conducted as described in a report by Tanaka et al. (Tanaka et al. (2005) Plant Cell Tiss. Org. Cult. 80: 1-24).

The expression of three genes (gentian 5,3'-aromatic acyltransferase gene, gentian 3'-glycosyl transferase gene, and pansy F3'5'H gene) in *nierembergia* transformants was confirmed by RT-PCR. For the lines where transcription of all three genes were confirmed, the petal color was analyzed in a similar manner to that described in a report by Mizutani et al. (Fukuchi-Mizutani et al. (2003) Plant Physiol. 132: 1652-1663), but the expected final product gentiodelphin was not detected in any of the lines.

On the other hand, a crude enzyme was extracted from the petals of the transformants where transcription of all three genes were confirmed, in a manner as described in the report by Fujiwara et al. (Fujiwara et al., (1997) Eur. J. Biochem. 249: 45-51). Using this crude extract as a enzyme solution, in vitro activity of 5,3'-aromatic acyltransferase was measured with DEL 3G-5G-3'G as the substrate in a manner similar to Example 2; then, formation of gentiodelphin was confirmed. On the other hand, when the crude enzyme from a non-recombinant *nierembergia* was used as the control, gentiodelphin was not detected. This result revealed that the transgenic *nierembergia* has 5,3'-aromatic acyltransferase activity, i.e., an activity of transferring aromatic acyl groups to both 5 and 3' position of DEL 3G-5G-3'G used as a substrate.

However, when the activity of the 3'-glucosyltransferase was measured in vitro as described in

- 30 -

a report by Mizutani et al. (Fukuchi-Mizutani et al. (2003) Plant Physiol. 132: 1652-1663) with crude enzyme solution from nierembergia transformants, no 3'-glucosyltransferase activity was detected. These results confirmed that a protein with an activity of 5,3'-aromatic acyltransferase is indeed present in the cell of the nierembergia transformants. However, the reason why the expected gentiodelphin was not detected in the petals of the transformants was because the protein of the 3'-glucosyltransferase that should work prior to the 5,3'-aromatic acyltransferase was not synthesized in the cell of nierembergia, or did not function even if it was synthesized.

Effect of the Invention

As described above, the present invention has demonstrated that a recombinant aromatic acyltransferase obtained by expressing a gentian aromatic acyltransferase gene in E. coli has an activity of transferring aromatic acyl groups not only to a sugar at the 5 position of delphinidin glucosides, but also to a sugar at the 3' position thereof. It further revealed that the naturally occurring anthocyanin 5-aromatic acyltransferase purified from gentian petals also has an activity of transferring an aromatic acyl group to a sugar at the 3' position, i.e., unlike the conventional anthocyanin aromatic acyltransferase, a single enzyme transfers aromatic acyl groups to sugars at both 5 position and 3' position of an anthocyanin. Furthermore, it was possible to express this gene in different species of plants, thus to obtain the 5,3'-aromatic acyltransferase activity.

It is generally believed that the aromatic acyl group at the 3' position contributes to the stabilization and blueing of anthocyanin more strongly than the aromatic acyl group at the 5 position, and the presence of sugar-acyl side chains at multiple positions including 3' position is more preferred. Thus, as described in the present invention, it is possible to create an

- 31 -

anthocyanin that has a more stable and bluish hue by
conducting aromatic acylation at both 5 and 3' position
of an anthocyanin glucosides using a 5,3'-aromatic
acyltransferase that transfers aromatic acyl groups to
5 both 5 and 3' position. Furthermore, by expressing the
gene for the said enzyme in plants together with other
genes essential for anthocyanin biosynthesis or
anthocyanin modification, it is possible to make flower
color, mainly comprising of anthocyanins, more stable and
10 bluer.

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-32-

CLAIMS

1. A method of adding aromatic acyl groups at both the 3' and 5 positions of an anthocyanin, said method comprising introducing a nucleic acid molecule into a host cell expressing the anthocyanin, wherein the nucleic acid molecule encodes a protein having (i) at least 90% identity to the amino acid sequence of SEQ ID NO:4 or 6 and (ii) aromatic acyl transferase activity.

2. The method of claim 1, wherein the protein has the amino acid sequence of SEQ ID NO:4 or 6.

3. The method of claim 1 or 2, wherein the host cell is a plant cell.

4. A method of stabilizing an anthocyanin by adding aromatic acyl groups at both the 3' and 5 positions of the anthocyanin, said method comprising introducing a nucleic acid molecule into a host cell expressing the anthocyanin, wherein the nucleic acid molecule encodes a protein having (i) at least 90% identity to the amino acid sequence of SEQ ID NO:4 or 6 and (ii) aromatic acyl transferase activity.

5. The method of claim 4, wherein the protein has the amino acid sequence of SEQ ID NO:4 or 6.

6. The method of claim 4 or 5, wherein the host cell is a plant cell.

7. A method of blueing an anthocyanin by adding aromatic acyl groups at both the 3' and 5 positions of the anthocyanin, said method comprising introducing a nucleic acid molecule into a host cell expressing the anthocyanin, wherein the nucleic acid molecule encodes a protein having (i) at least 90% identity to the amino acid sequence of SEQ ID NO:4 or 6 and (ii) aromatic acyl transferase activity.

-33-

8. The method of claim 7, wherein the protein has the amino acid sequence of SEQ ID NO:4 or 6.

9. The method of claim 7 or 8, wherein the host cell is a plant cell.

Application number / numéro de demande: JP05 20257

Figures: 6-7

Pages: _____

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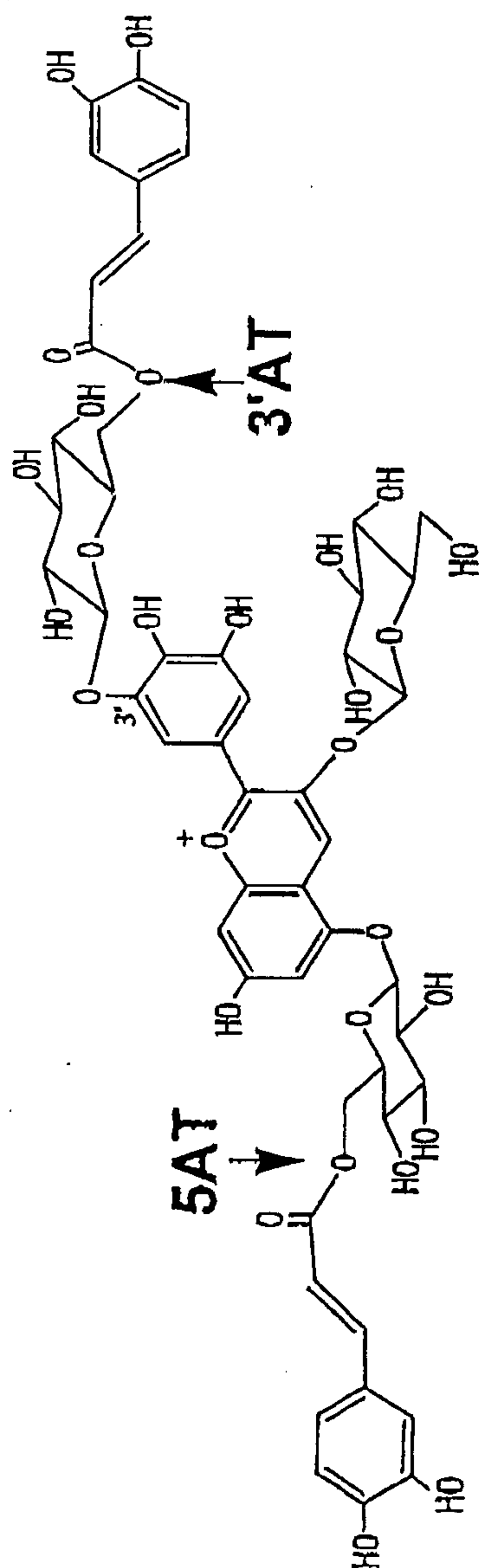
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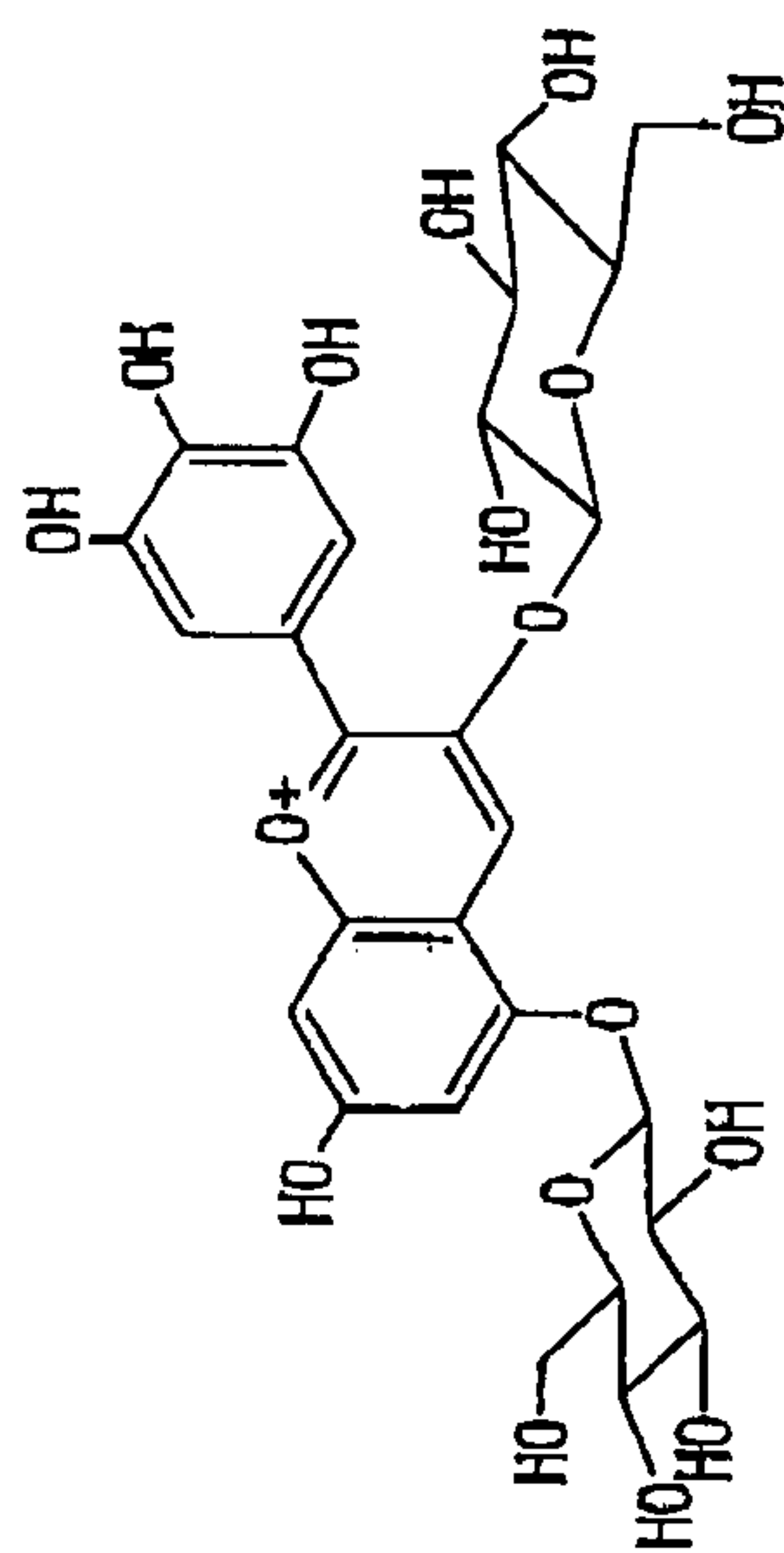
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Pour vous renseigner si vous pouvez commander une copie des items ne
pouvant être balayés, veuillez visiter le site web de l'OPIC au [HTTP://CIPO.GC.CA](http://CIPO.GC.CA)

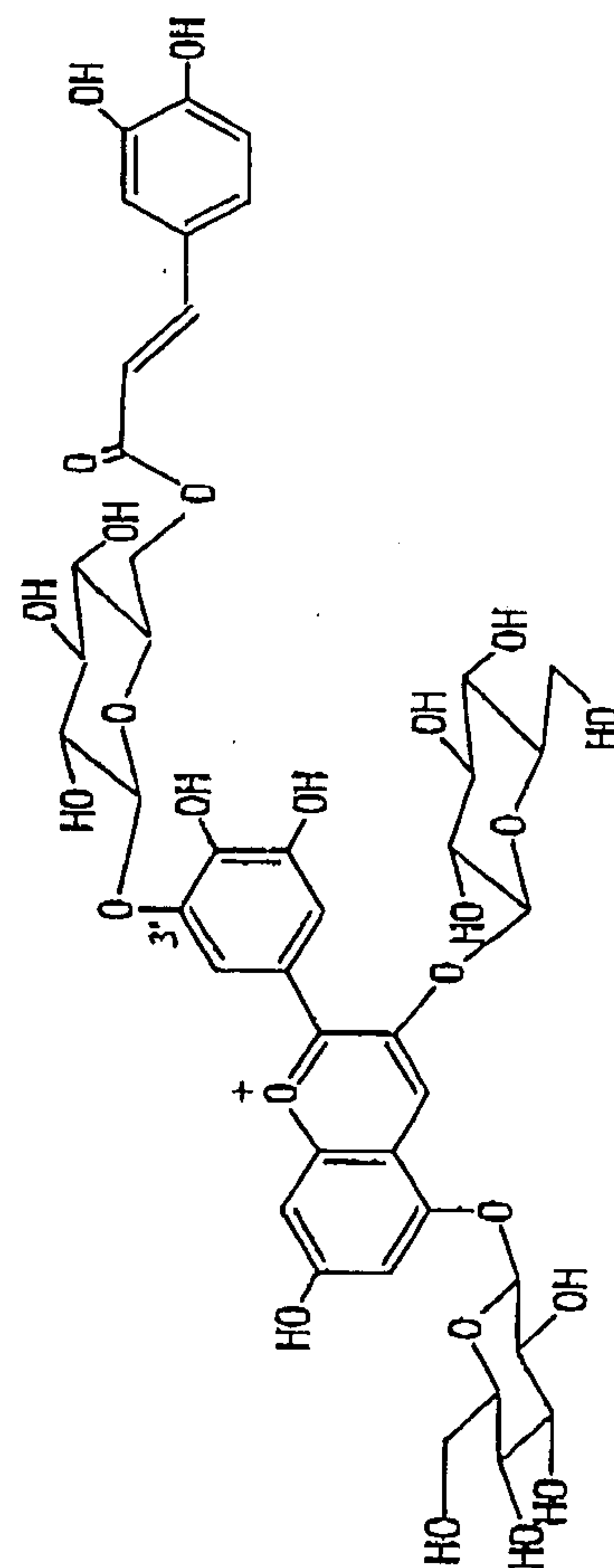
Fig.1



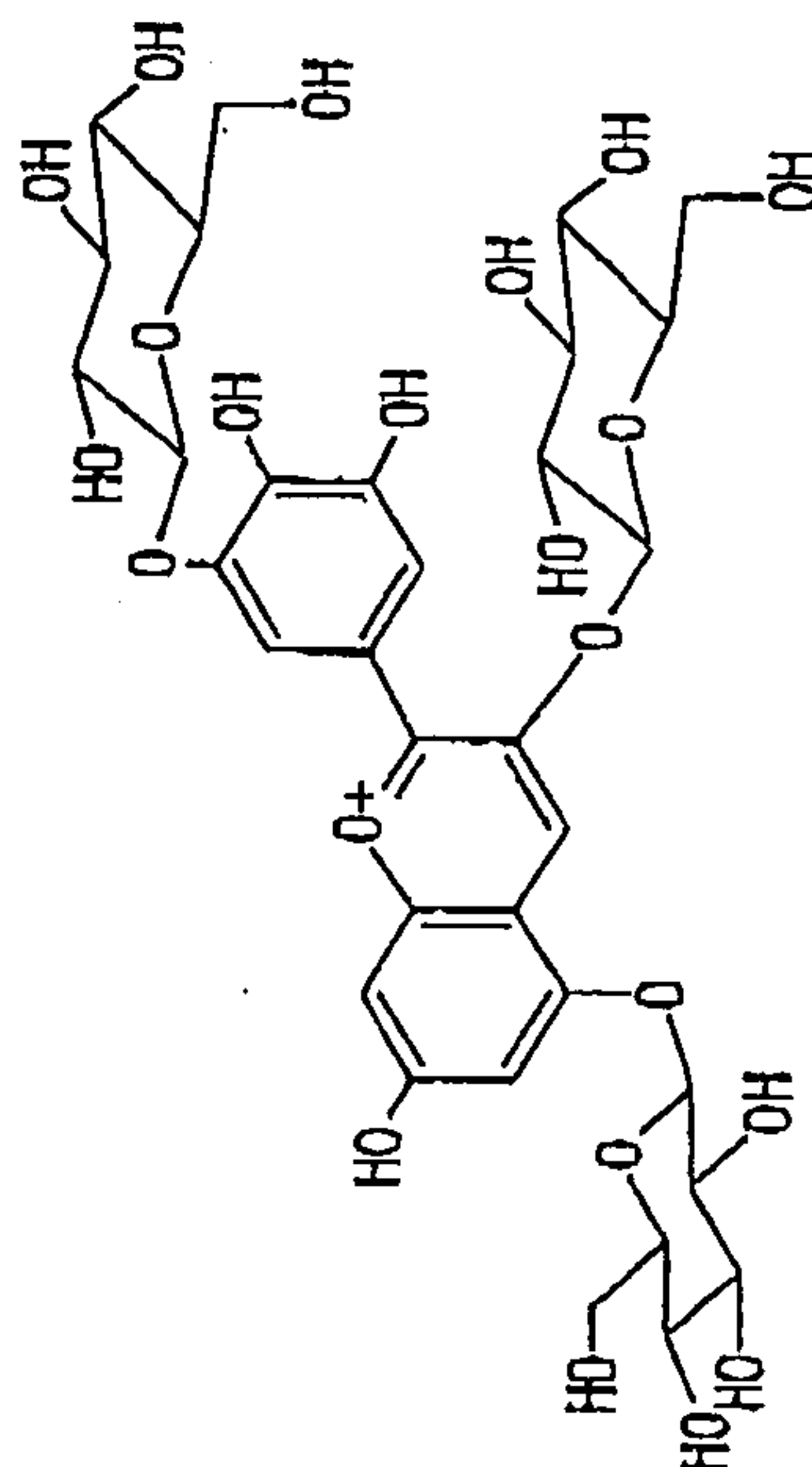
DELPHINIDIN 3-GLUCOSYL-5-CAFFEYL-GLUCOSYL-3'-CAFFEYL-GLUCOSIDE
(GENTIODELPHIN, DEL 3G-5CafG-3'CafG)



DELPHINIDIN 3,5-DIGLUCOSIDE
(DEL 3G-5G)

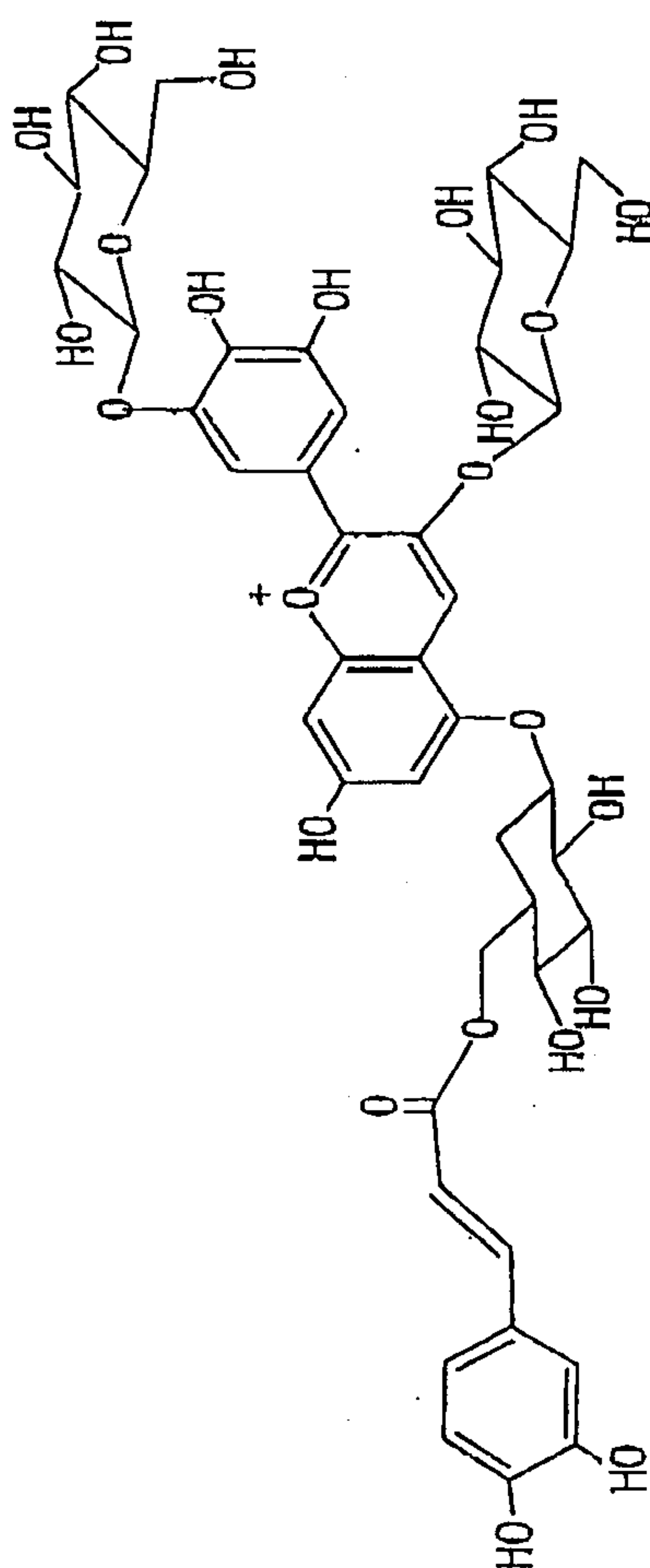


DELPHINIDIN 3,5-DIGLUCOSYL-3'-CAFFEYLGLUCOSIDE
(DEL 3G-5G-3'CafG)



DELPHINIDIN 3,5,3'-TRIGLUCOSIDE
(DEL 3G-5G-3'G)

Fig. 2



2/5

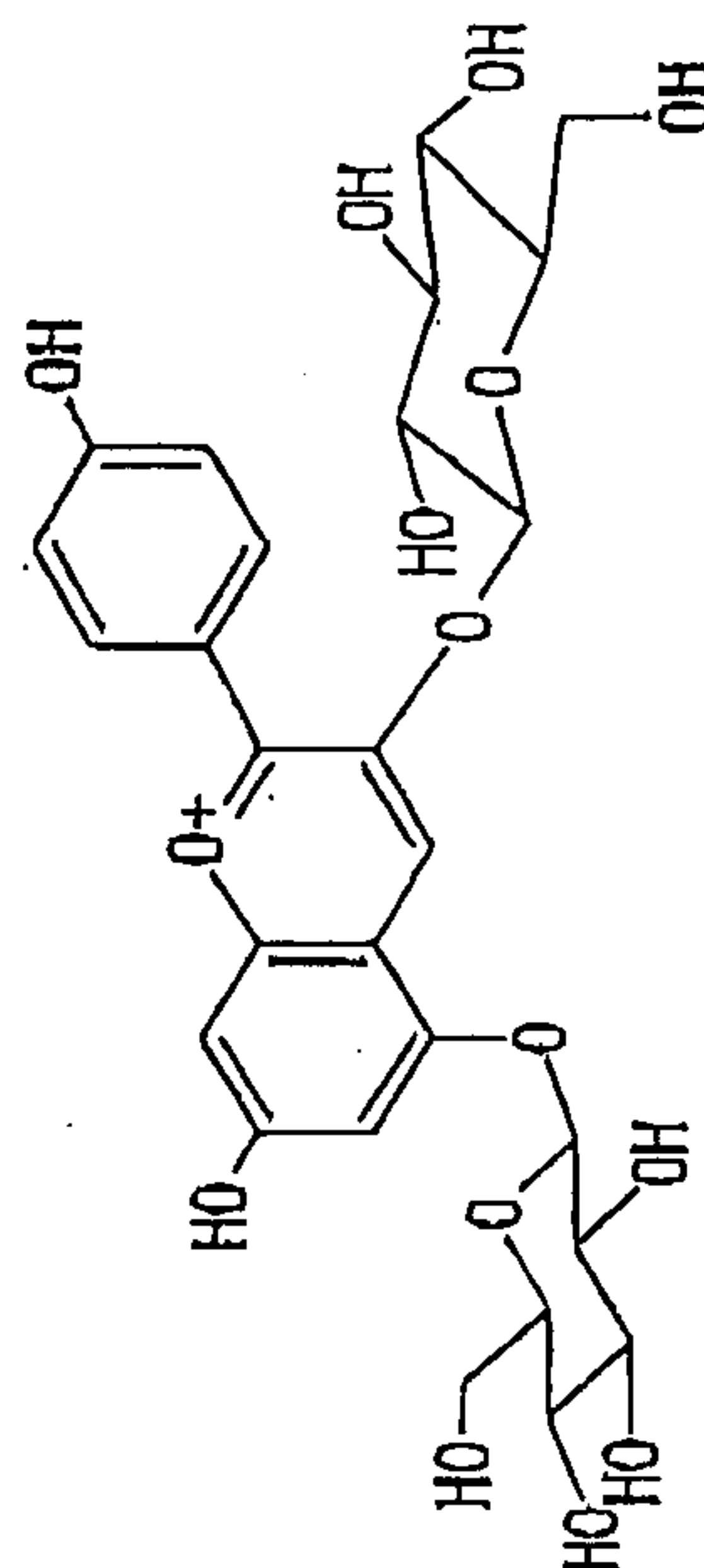
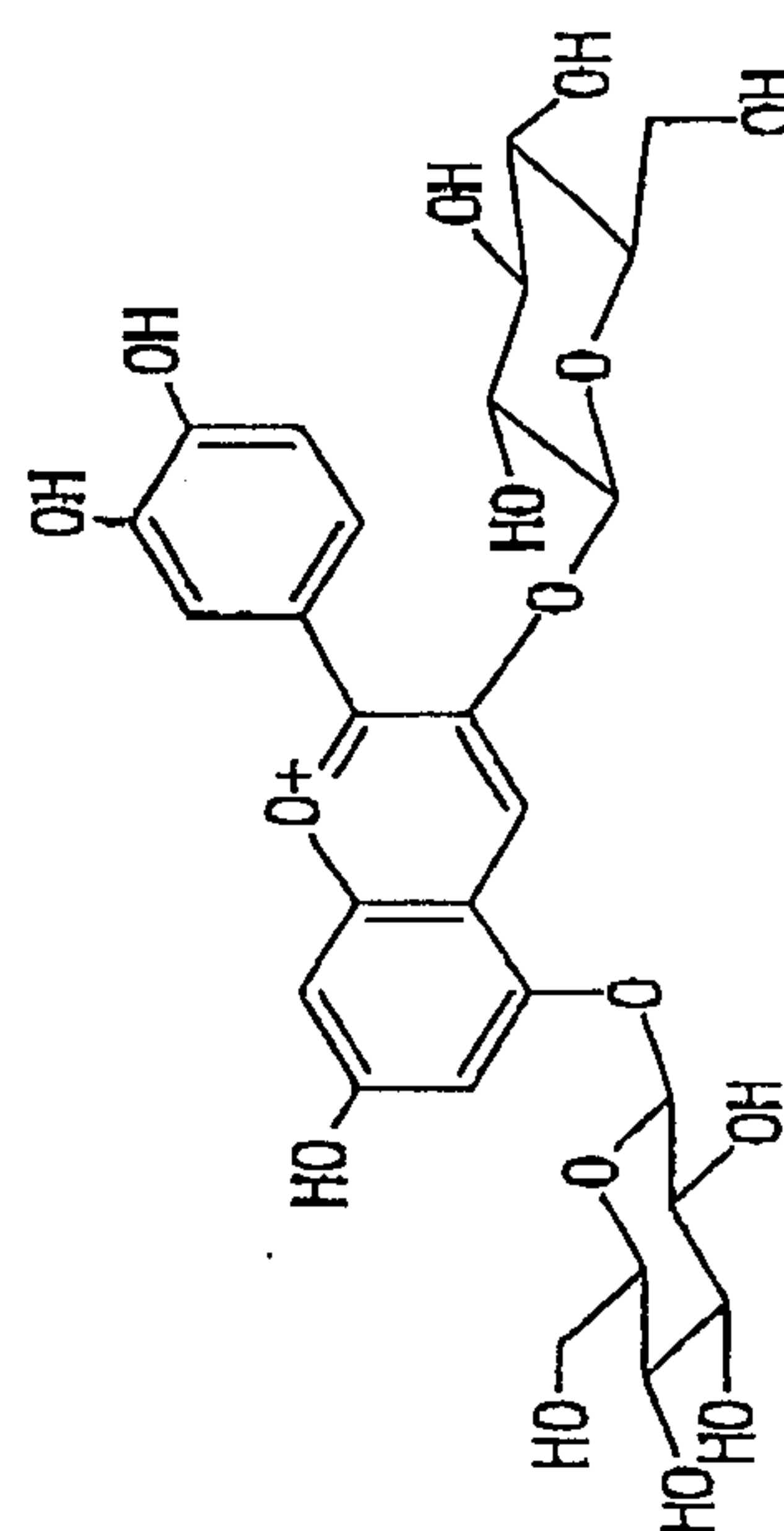
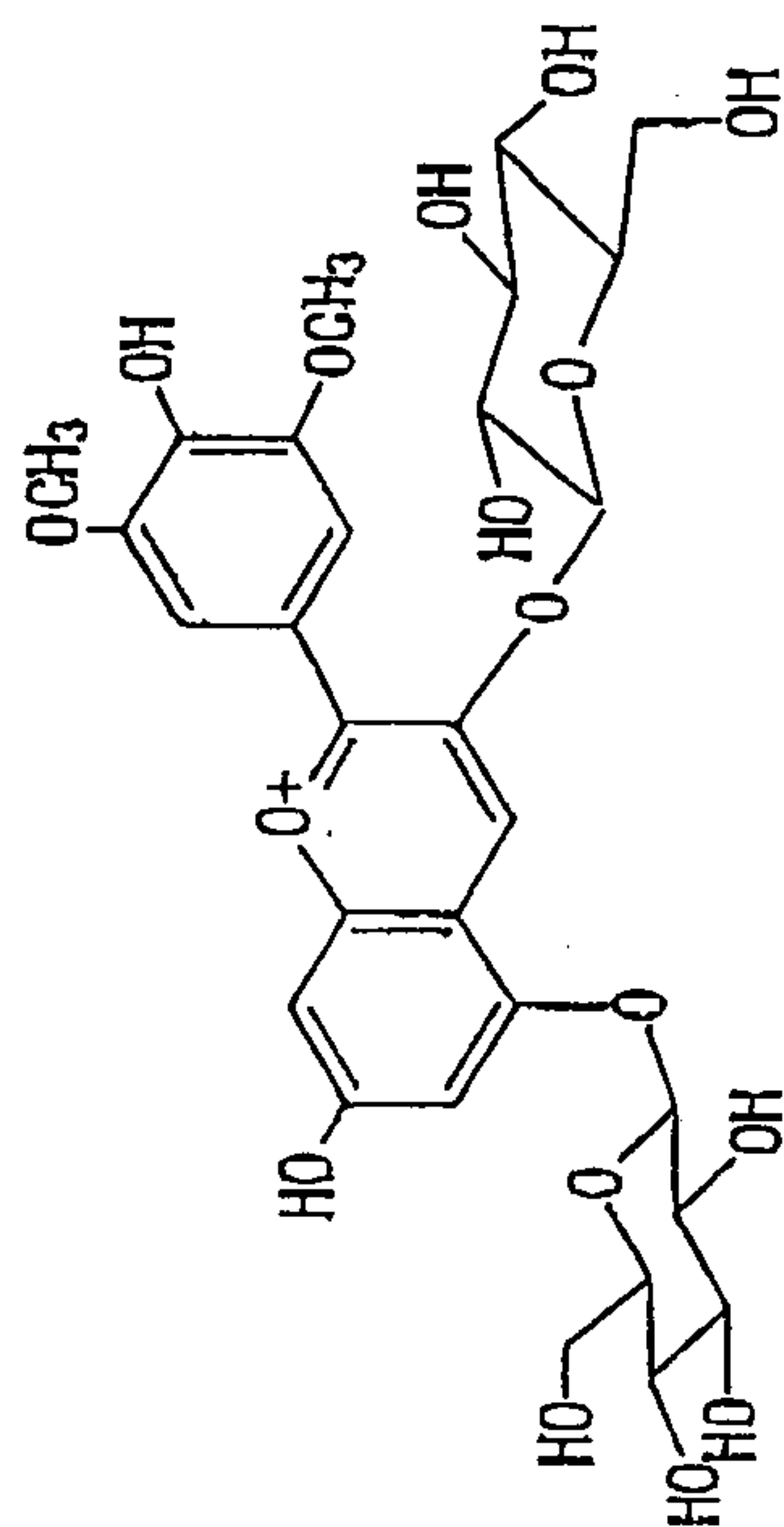
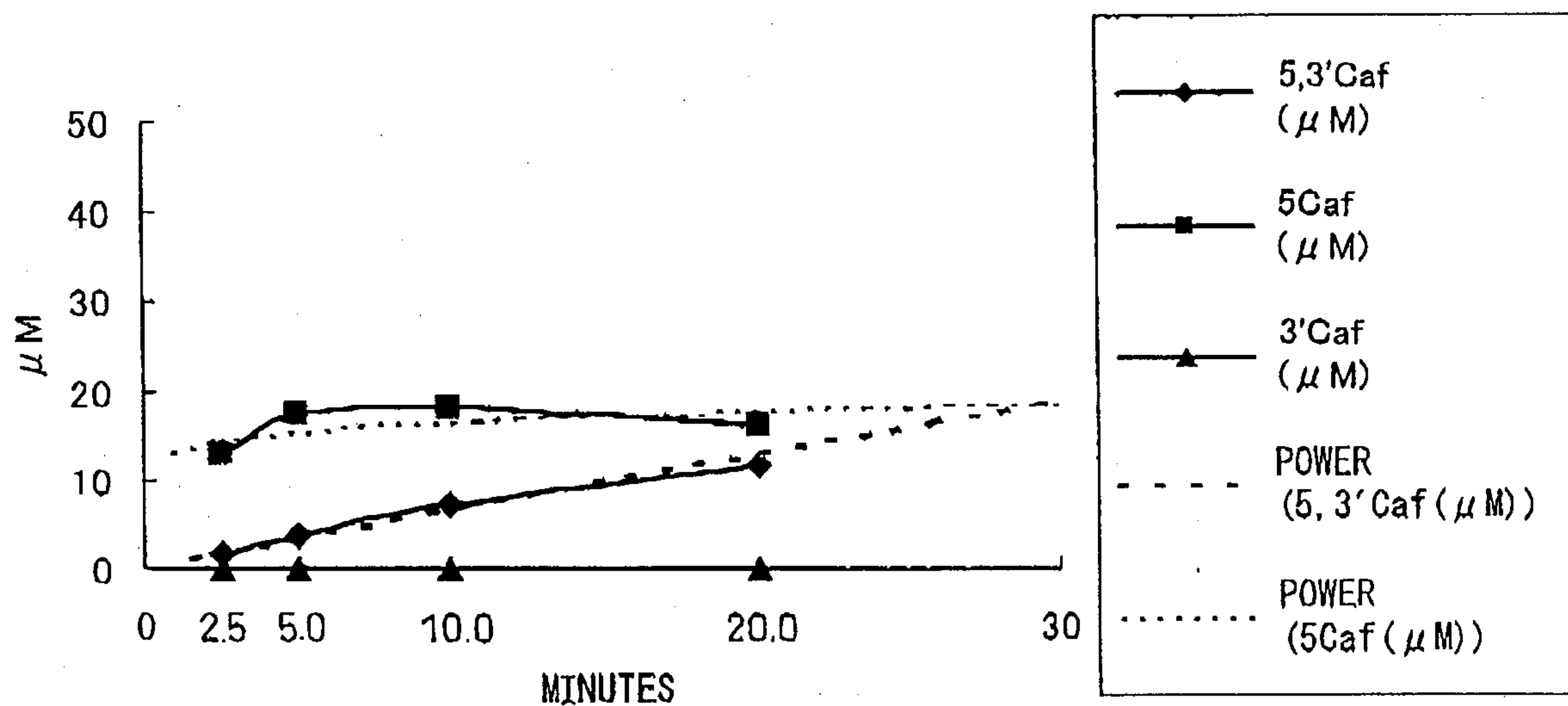


Fig.3

(A)



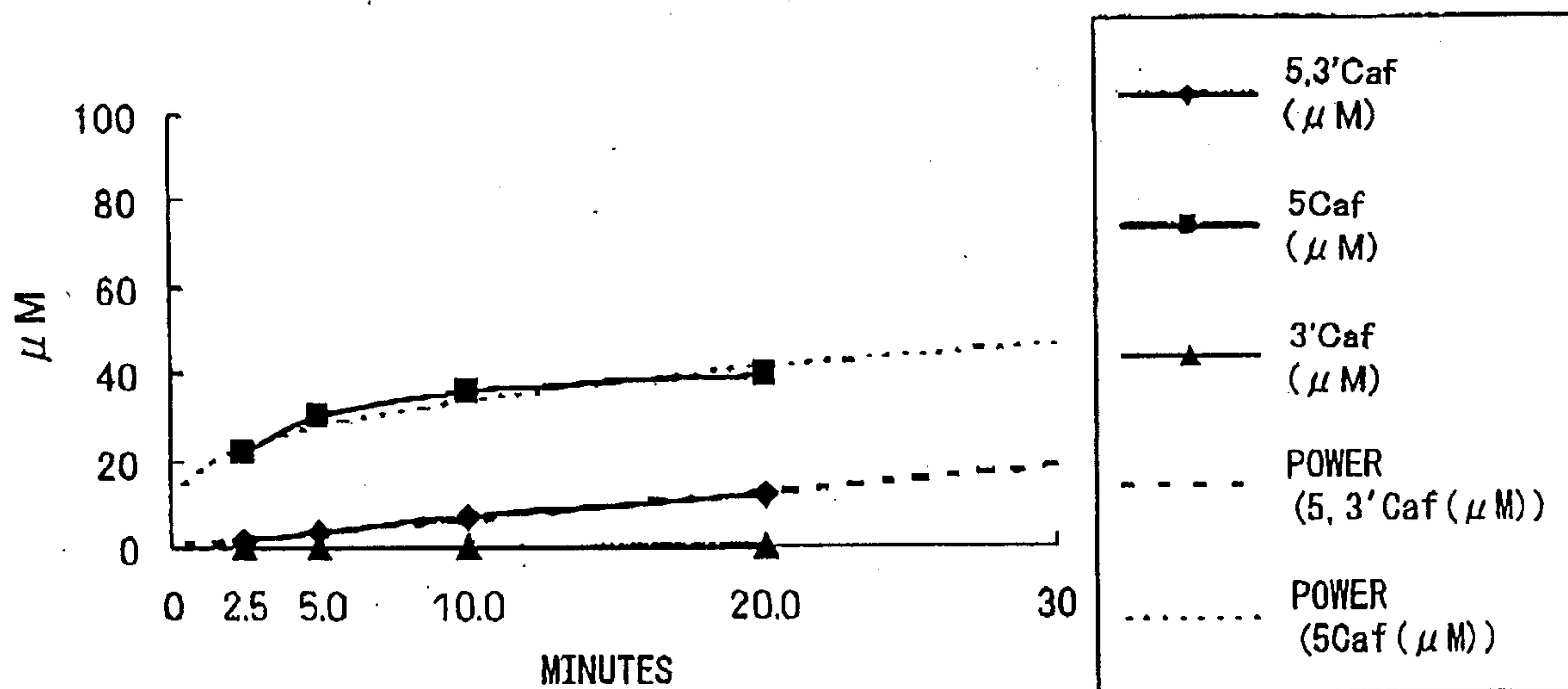
(B)

50 μM DEL 3G-5G-3'G

TIME (MINUTES)	5,3'Caf (μM)	5Caf (μM)	3'Caf (μM)	triG (μM)
2.5	1.65	13.05	0.00	35.3
5.0	3.90	17.40	0.00	28.7
10.0	7.30	18.15	0.00	24.6
20.0	11.70	16.20	0.00	22.1

Fig.4

(A)



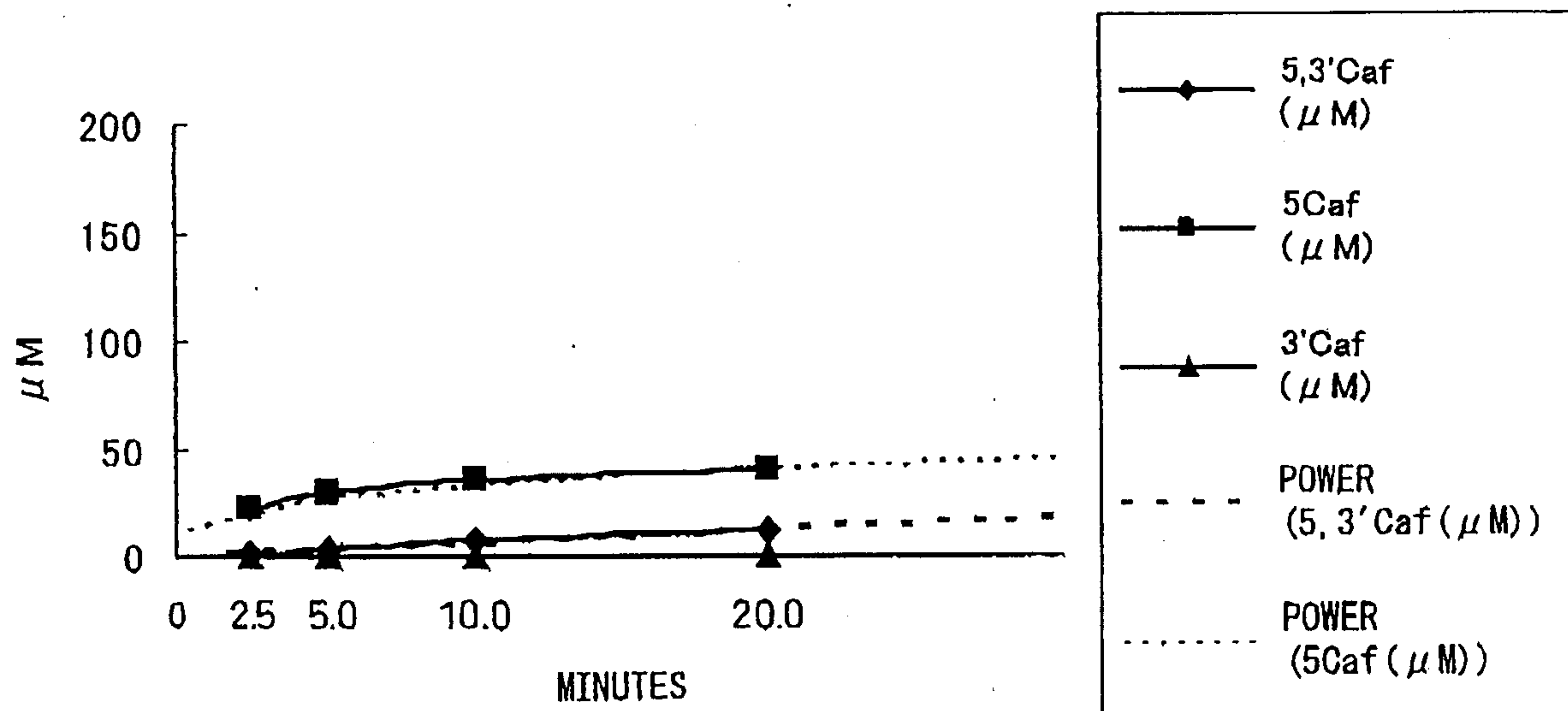
(B)

100 μM DEL 3G-5G-3'G

TIME (MINUTES)	5,3'Caf (μM)	5Caf (μM)	3'Caf (μM)	triG (μM)
2.5	1.70	21.80	0.00	76.5
5.0	3.50	30.10	0.00	66.4
10.0	6.80	35.60	0.00	57.6
20.0	11.80	39.60	0.00	48.6

Fig.5

(A)



(B)

200 μM DEL 3G-5G-3'G

TIME (MINUTES)	5,3'Caf (μM)	5Caf (μM)	3'Caf (μM)	triG (μM)
2.5	1.60	31.20	0.00	167.2
5.0	3.00	44.40	0.00	152.6
10.0	7.00	60.60	0.20	132.2
20.0	11.60	70.00	0.60	117.8