

(19)



(11)

EP 3 302 097 B1

(12)

EUROPEAN PATENT SPECIFICATION

(45) Date of publication and mention
of the grant of the patent:

23.03.2022 Bulletin 2022/12

(21) Application number: **16804152.3**

(22) Date of filing: **27.05.2016**

(51) International Patent Classification (IPC):

A23L 2/60 ^(2006.01) **A61K 31/70** ^(2006.01)
C12P 19/56 ^(2006.01) **A61K 31/704** ^(2006.01)
C12N 15/81 ^(2006.01)

(52) Cooperative Patent Classification (CPC):

C12P 19/56; A23L 2/60; A61K 31/704

(86) International application number:

PCT/US2016/034781

(87) International publication number:

WO 2016/196345 (08.12.2016 Gazette 2016/49)

(54) **HEAT TREATMENT TO PRODUCE GLYCOSIDES**

WÄRMEBEHANDLUNG ZUR HERSTELLUNG VON GLYKOSIDEN

TRAITEMENT THERMIQUE POUR PRODUIRE DES GLYCOSIDES

(84) Designated Contracting States:

**AL AT BE BG CH CY CZ DE DK EE ES FI FR GB
GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO
PL PT RO RS SE SI SK SM TR**

(30) Priority: **29.05.2015 US 201562168142 P**

(43) Date of publication of application:

11.04.2018 Bulletin 2018/15

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EP 3 302 097 B1

Description

Background

[0001] Sugars, such as sucrose, fructose and glucose, are utilized to provide a pleasant taste to beverages, foods, pharmaceuticals, oral hygienic and cosmetic products. Sucrose, in particular, imparts a taste preferred by most consumers. Although sucrose provides superior sweetness characteristics, it is caloric. Non-caloric or lower caloric sweeteners have been introduced to satisfy consumer demand. Consumers also desire that these sweeteners have favorable taste characteristics.

[0002] Stevia is a genus of about 240 species of herbs and shrubs in the sunflower family (*Asteraceae*), native to subtropical and tropical regions and from western North America to South America. The species *Stevia rebaudiana*, commonly known as sweetleaf, sweet leaf, sugarleaf, or simply stevia, is widely grown for its sweet leaves. Stevia-based sweeteners may be obtained by extracting one or more sweet compounds from the leaves. Many of these compounds are glycosides of steviol, a diterpene compound. These diterpene glycosides are about 150 to 450 times sweeter than sugar. Steviol glycosides differ from each other by sweetness power as well as other sensory features contributing to taste quality such as bitterness, lingering aftertaste and the like. See Kinghorn, A. D., Stevia: The genus Stevia, Taylor & Francis, London (2002).

[0003] Examples of steviol glycosides are described in PCT International Patent Application Publication No. WO 2013/096420 (see, e.g., listing in Fig. 1); and in Ohta et. al., "Characterization of Novel Steviol Glycosides from Leaves of *Stevia rebaudiana* Morita," J. Appl. Glycosi., 57, 199-209 (2010) (see, e.g., Table 4 at p. 204). Structurally, the diterpene glycosides are characterized by a single base, steviol, and differ by the presence of carbohydrate residues at positions C13 and C19, as presented in Figs. 2a-2k of WO 2013/096420.

[0004] Typically, on a dry weight basis, the four major steviol glycosides found in the leaves of Stevia are dulcoside A (0.3%), rebaudioside C (0.6-1.0%), rebaudioside A (3.8%) and stevioside (9.1%). Other glycosides identified in Stevia extract include one or more of rebaudioside B, D, E, F, G, H, I, J, K, L, M, N, O, steviolbioside and rubusoside.

[0005] While the major steviol glycoside, rebaudioside A, is commonly used as sweetener in beverage applications it has off-taste issues. More recently, there has been focus on certain minor steviol glycosides which have better taste properties. For example, rebaudioside M has higher sweetness intensity and is more potent than other steviol glycosides (e.g., see Prakash, I., et al. (2013) Nat. Prod. Commun., 8: 1523-1526, and WO 2013/096420). Rebaudioside D tastes about 200-220 times sweeter than sucrose and in a sensory evaluation had a slow onset of sweetness and was very clean, namely sweeter overall than sucrose, less sweet lingering aftertaste compared to sucrose (e.g., see Prakash, I., et al. (2012) Int. J. Mol. Sci., 13:15126-15136).

[0006] Recombinant DNA technology has made it possible, and commercially viable, to produce desired steviol glycosides using a wide variety of host cells such as yeast. Recovery of intracellularly expressed products from within the host cells, however, is fraught with difficulties because of the necessity of rupturing or hydrolyzing the tough yeast cell wall. The choice of cell wall disruption can also affect the downstream processing of the recovered product and its subsequent purification from yeast cells or fermentation broths thereby affecting the efficiency and cost effectiveness of the process.

[0007] WO2013110673A describes a recombinant microorganism capable of producing diterpene and the preparation of the diterpene comprising fermenting the recombinant microorganism. WO2013110673A discloses that broth was incubated at 95° C for 5 minutes, acetonitrile was added, samples were mixed, spun down and diluted for analyzing the steviol or steviol glycoside by using LC/MS.

[0008] WO2014191581A describes recombinant microorganism for diterpene production. WO2014191581A discloses the cultures in the 96-well plate was incubated for 15 minutes at 95° C and cooled down, and to each well acetonitrile was added and homogenized for analyzing Reb A using LC/MS.

[0009] US2014329281A describes recombinant microorganisms that have been engineered to express recombinant genes encoding USTs and can produce steviol glycosides. US2014329281A discloses that DMSO was added to the culture samples which were vortexed, heated at 80° C for 15 minutes, and the steviol glycosides or 19-SMG content was analyzed by LC-MS.

Summary

[0010] Disclosed is a method to efficiently recover desired steviol glycosides produced by recombinant yeast technology in a cost effective manner.

[0011] The invention discloses a method of releasing steviol glycosides from a yeast host cell, comprising heating a fermentation medium comprising steviol glycoside producing engineered yeast host cells to a temperature in the range from 70° C to 75° C for 5 minutes to 48 hours to release one or more steviol glycosides from the engineered host cells into the fermentation medium, wherein the steviol glycoside is selected from rebaudioside M, rebaudioside D, rebaudi-

oside A, rebaudioside B or combinations thereof.

[0012] The invention discloses a method of releasing steviol glycosides from a yeast host cell, comprising the steps of:

- a) providing a fermentation medium comprising a yeast host cell engineered to produce one or more steviol glycosides; and
- b) releasing the one or more steviol glycosides from the engineered yeast host cell by heating the composition to a temperature in the range from 70° C to 75° C for 5 minutes to 48 hours, wherein the one or more steviol glycosides are released from an intracellular portion of the host cell into an extracellular portion of the fermentation medium, wherein the steviol glycoside is selected from rebaudioside M, rebaudioside D, rebaudioside A, rebaudioside B or combinations thereof.

[0013] A method of releasing steviol glycosides from a composition, the method comprising the step of heating the composition to a temperature in the range from 50° C to 95° C for 5 minutes to 48 hours to release the steviol glycosides from engineered host cells in the composition is described.

[0014] A method of releasing steviol glycosides, the method comprising the steps of:

- a) providing a composition comprising an engineered host cell to produce one or more steviol glycosides; and
- b) releasing the one or more steviol glycosides from the engineered host cell by heating the composition to a temperature in the range from 70° C to 75° C for 5 minutes to 48 hours is described.

[0015] A composition produced by the method according to the invention, comprising an engineered yeast host cell to produce one or more steviol glycosides, and one or more steviol glycosides that are released from the engineered host cell by heating the composition to a temperature in the range from 70° C to 75° C for 5 minutes to 48 hours is described.

[0016] Such methods result in releasing the glycosides into the composition while minimizing cell lysis.

Description of the Drawings

[0017]

- FIG. 1 shows a representative mevalonate pathway.
- FIG. 2 shows a representative non-mevalonate pathway.
- FIG. 3 shows a representative pathway for steviol production.
- FIG. 4 shows representative pathways for the biosynthesis of steviol glycosides from steviol.
- FIG. 5A is a digital image of a bright field microscopy image before heat treatment.
- FIG. 5B is a digital image of a bright field microscopy image after heat treatment.

Detailed Description

[0018] Embodiments of the disclosure described herein are not intended to be exhaustive or to limit the invention to the precise forms disclosed in the following detailed description. Rather a purpose of the embodiments chosen and described is the appreciation and understanding by others skilled in the art of the claims. The publications and patents disclosed here are provided solely for their disclosure.

[0019] Host cells such as yeast and fungus can be engineered for the synthesis of steviol glycosides. During fermentation only a fraction of the glycosides produced by the host cell may be released from within the cells into the fermentation media. To increase the glycoside amount released from the host cell such as yeast, mechanical and chemical disruption methods may be used. Mechanical disruption, namely the breaking of the cell, releases the intracellular components into the surrounding medium. The result is a complex mixture of not only the desired recombinant product but also other products such as protein, nucleic acids, cellular metabolites, cell fragments, complicating the recovery of the desired product. Chemical methods entail chemically altering or making permeable the cell membrane and/or wall structure to allow diffusion of the product from within the cell. Chemical treatments, however, have to be compatible with the organism and the release of the desired product.

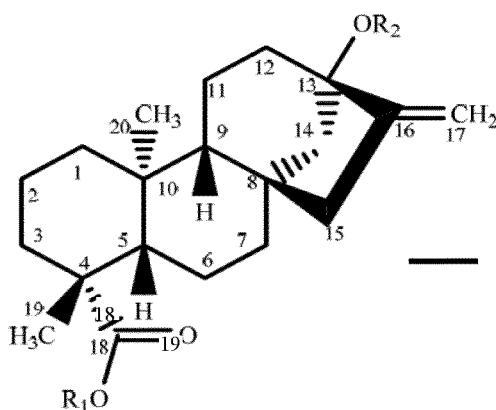
[0020] Described are methods of efficiently recovering steviol glycosides after fermentation using heat treatments. Heat applied for a given time period and temperature to a fermentation broth or fermentation medium that includes host cells (e.g. yeast and fungus). The host cells can produce the desired steviol glycosides. When the fermentation medium containing the host cells are heated, the desired glycosides can be released from the host cells with minimal lysis of the cells thereby enriching the fermentation media or composition for the desired glycoside.

[0021] The method of the invention uses engineered yeast capable of producing steviol glycosides. An engineered yeast capable of producing steviol glycosides can include one or more exogenous nucleic acids that encode enzyme(s)

that promote formation of one or more steviol glycosides in the cell.

[0022] As used herein, the term "steviol glycoside(s)" refers to glycosides of steviol. Exemplary steviol glycoside, include, but are not limited to, rebaudioside A, rebaudioside B, rebaudioside C, rebaudioside D, rebaudioside E, rebaudioside F, rebaudioside G, rebaudioside H, rebaudioside I, rebaudioside J, rebaudioside K, rebaudioside L, rebaudioside M, rebaudioside N, rebaudioside O, stevioside, steviolbioside, dulcoside A, and/or rubusoside. Engineered yeast can produce steviol glycosides that are naturally found ("naturally occurring") or ones that are not found in nature (non-naturally occurring). As used herein, the term "total steviol glycosides" (TSG) is calculated as the sum of the content of all steviol glycosides in a composition on a dry (anhydrous) basis.

[0023] Structurally, steviol glycosides have a central molecular moiety, which is a single steviol base, and glucopyranosyl residues, or other sugar residuals attached to the C₁₃ and C₁₉ atoms of the steviol base, according to the atom numbering on the base shown below. That is, glucopyranosyl residues represent groups R₁ and R₂ in the following formula:



[0024] Table A below shows the certain steviol glycosides and the corresponding R₁ and R₂ groups:

Table A

Compound name	R ₁ (C-19)	R ₂ (C-13)
Steviol	H	H
Stevioside	β-Glu	β-Glu-β-Glu (2->1)
Rebaudioside A	β-Glu	β-Glu-β-Glu (2->1) β-Glu (3->1)
Rebaudioside B	H	β-Glu-β-Glu (2->1) β-Glu (3->1)
Rebaudioside C	β-Glu	β-Glu-α-Rha (2->1) β-Glu (3->1)
Rebaudioside D	β-Glu-β-Glu (2->1)	β-Glu-β-Glu (2->1)
Rebaudioside E	β-Glu-β-Glu (2->1)	β-Glu-β-Glu (2->1)
Rebaudioside G	β-Glu	β-Glu-β-Glu (3->1)

(continued)

Compound name	R ₁ (C-19)	R ₂ (C-13)
Rebaudioside M	β -Glu- β -Glu (2->1) β -Glu (3->1)	β -Glu- β -Glu (2->1) β -Glu (3->1)
Rebaudioside N	β -Glu- α -Rha (2->1) β -Glu (3->1)	β -Glu- β -Glu (2->1) β -Glu (3->1)
Rebaudioside O	β -Glu- α -Rha (2->1)- β -Glu (3->1) β -Glu (3->1)	β -Glu- β -Glu (2->1) β -Glu (3->1)
Glu: glucose Rha: rhamnose		

[0025] The steviol glycosides may be produced in a fermentation process. The fermentation process can use a genetically modified organism that is engineered for the production of one or more steviol glycosides, such as rebaudioside A (Reb A), rebaudioside M (Reb M) and/or rebaudioside D (Reb D). Production of steviol glycosides can be carried out using an engineered microbial strain having a set of enzymes that provide a pathway for the synthesis of steviol glycosides.

[0026] Various yeast or fungal host cells can be engineered to provide a pathway to one or more steviol glycosides. Such cells can be transformed with one or more DNA construct encoding enzymes for steviol glycoside synthesis. Exemplary yeast and fungus that can be used for hosts for exogenous DNA constructs encoding steviol glycoside pathway enzymes, include, but are not limited to the genus of, *Candida*, *Pichia* (*Hansenula*), *Kloeckera* (*Hanseniaspora*), *Kluyveromyces*, *Rhodotorula*, *Torulopsis*, *Zygosaccharomyces*, *Saccharomycete*, *Yarrowia*, and *Saccharomyces*. Exemplary species include *Candida albicans*, *Pichia pastoris*, *Saccharomyces cerevisiae*, and *Schizosaccharomyces pombe*. Further, host cells can also include genetic modifications other than those of the steviol glycoside pathway that may provide improved performance during fermentation.

[0027] An "engineered host cell" refers to yeast and fungal cells having at least one exogenous DNA sequence that is introduced into the cell, either integrated into the cell's genome or present on an extrachromosomal construct, such as a plasmid in bacteria or episomes. While the specification may describe yeast cells in detail, it should be understood that the method may also be adapted to fungal cells.

[0028] The term "exogenous" refers to a molecule, such as a nucleic acid, or an activity, such as an enzyme activity. The exogenous molecule is introduced into the host yeast or fungus. An exogenous nucleic acid can be introduced into the host organism by well-known techniques and can be maintained external to the host's chromosomal material (e.g., maintained on a non-integrating vector), or can be integrated into the host's chromosome, such as by a recombination event. Generally, the genome of an engineered yeast or fungus is augmented through the stable introduction of one or more recombinant genes. An exogenous nucleic acid can encode an enzyme, or portion thereof, that is either homologous or heterologous to the host organism. An exogenous nucleic acid can be in the form of a "recombinant gene or DNA construct" referring to a nucleic acid that is in one or more ways manipulated through molecular techniques to be in a form that does not naturally exist. The term "non-natural" may be used to characterize a molecule, such as a nucleic acid or protein, or an organism that does not naturally exist in nature.

[0029] The term "heterologous" (e.g., "non-native") refers to a molecule or activity that is from a source that is different than the referenced molecule or organism. Accordingly, a gene or protein that is heterologous to a referenced organism is a gene or protein not found in that organism. In the context of the disclosure, a "heterologous glycosyltransferase" refers to a glycosyltransferase polypeptide that is different from any glycosyltransferase polypeptide that may be native to the host organism. For example, a specific glycosyltransferase gene found in a first species and exogenously introduced into a host yeast or fungal organism that is different than the first species is "heterologous" to the host yeast or fungal organism.

[0030] The engineered yeast or fungus can use an auxotrophic marker suitable for selecting for a transformant. The host cell can include modifications (e.g. deletions) in one or more genes that control auxotrophies, such as LYS2, LEU2, HIS3, URA3, URA5, and TRP1. Using a host cell having a desired genetic background for introduction of one or more exogenous genes, one or more gene construct(s) is introduced into a cell to integrate into the genome, or to be stably maintained and allow for expression. Methods for introducing a gene construct into a host cell include transformation,

transduction, transfection, co-transfection, electroporation. In particular, yeast transformation can be carried out using the lithium acetate method, the protoplast method, and the like. The gene construct to be introduced may be incorporated into a chromosome in the form of a plasmid, or by insertion into the gene of a host, or through homologous recombination with the gene of a host. The transformed yeast into which the gene construct has been introduced can be selected with a selectable marker (for example, an auxotrophic marker as mentioned above). Further confirmation can be made by measuring the activity of the expressed protein.

[0031] The transformation of exogenous nucleic acid sequences including the steviol pathway genes can be confirmed using methods well known in the art. Such methods include, for example, nucleic acid analysis such as Northern blots or polymerase chain reaction (PCR) amplification, or immunoblotting for expression of gene products, or other suitable analytical methods to test the expression of the introduced nucleic acid sequences or their corresponding gene product. It is understood by those skilled in the art that the exogenous nucleic acid is expressed in an amount sufficient to produce the desired product, and it is further understood that expression levels can be optimized to obtain sufficient expression using methods well known in the art and as disclosed herein.

[0032] The terpenoid compounds, isopentenyl diphosphate (IPP) and dimethylallyl diphosphate (DMAPP), can serve as chemical precursors to steviol glycosides in an engineered host cell (e.g. yeast). Some organisms, including plants, insect, and some microbial species, have a mevalonate (MVA) pathway that converts acetyl-CoA through a series of chemical intermediates to IPP and DMAPP. Some organisms produce IPP and DMAPP through the non-mevalonate pathway (also known as the methyl D-erythritol 4-phosphate or MEP pathway) starting with glyceraldehyde-3-phosphate (G3P) and pyruvate (PYR).

[0033] The yeast *Saccharomyces cerevisiae* naturally expresses genes of the mevalonate pathway. Mevalonate pathway genes include: (a1) acetoacetyl CoA thiolase (EC 2.3.1.9), (b1) 3-hydroxy-3-methylglutaryl-coenzyme A (HMG-CoA) synthase (EC 4.1.3.5); (c1) HMG-CoA reductase (EC 1.1.1.34); (d1) mevalonate kinase (EC 2.7.1.36); (e1) phosphomevalonate kinase (EC 2.7.4.2); and (f1) mevalonate diphosphate decarboxylase (EC 4.1.1.33). Mevalonate pathway enzymes convert acetyl-CoA to IPP as follows: acetyl-CoA → acetoacetyl-CoA → 3-hydroxy-3-methylglutaryl-CoA → mevalonate → mevalonate-5-phosphate → mevalonate-5-pyrophosphate → IPP. See also FIG. 1

[0034] The engineered yeast can include one or more modifications to increase the flux from acetyl-CoA to IPP and/or DMAPP, thereby providing an increased pool of IPP and/or DMAPP for use in a pathway to steviol. The modifications can include, for example, increasing expression or activity of one or more mevalonate pathway enzymes (a1)-(f1), such as by placing a nucleic acid encoding an enzyme that is homologous or heterologous to the yeast cell under the control of a promoter that provides increased expression, using multiple copies of the nucleic acid, and/or using a heterologous enzyme, a variant enzyme (e.g., including one or more amino acid substitutions), or a variant heterologous enzyme that provides a higher level of enzymatic activity as compared to the native enzyme.

[0035] Alternatively, the non-mevalonate (MEP) pathway can be used to provide IPP and DMAPP as precursors to steviol glycoside production. The yeast *Saccharomyces cerevisiae* does not naturally express genes of the MEP pathway, but can optionally be engineered to provide MEP pathway genes. Theoretically, the MEP pathway is more energetically efficient generally because it loses less carbon as CO₂ as compared to the MVA pathway (MEP pathway: 1 CO₂/IPP; MVA pathway: 4 CO₂/IPP; sugar as carbon source).

[0036] In particular, the non-mevalonate (MEP) pathway compounds, isopentenyl diphosphate (IPP), dimethylallyl diphosphate (DMAPP), are generated through a series of intermediates leading from glyceraldehydes-3-phosphate (G3P) and pyruvate (PYR), and a number of enzymes are responsible for this conversion. Enzymes involved in a biosynthetic pathway from G3P and PYR to IPP and DMAPP include (a2) 1-deoxy-D-xylulose-5-phosphate synthase (DXS), (b2) 1-Deoxy-D-xylulose-5-phosphate reductoisomerase (ispC), (c2) 4-diphosphocytidyl-2C-methyl-D-erythritol synthase (IspD), (d2) 4-diphosphocytidyl-2-C-methyl-D-erythritol kinase (IspE), (e2) 2C-Methyl-D-erythritol-2,4-cyclodiphosphate Synthase (IspF), (f2) 1-hydroxy-2-methyl-2-(E)-butenyl-4-diphosphate synthase (IspG), (g2) 4-hydroxy-3-methyl-2-(E)-butenyl-4-diphosphate reductase (IspH), and (h2) isopentenyl-diphosphate isomerase (IDI). See FIG. 2

[0037] The engineered yeast used to produce steviol glycosides by fermentation can have one or more genetic modifications to increase the flux from G3P and PYR to IPP and/or DMAPP, thereby providing an increased pool of IPP and/or DMAPP for use in a pathway to steviol. The modifications can include, for example, increasing expression or activity of one or more enzymes (a2) - (h2), such as by placing a nucleic acid encoding an enzyme that is heterologous to the yeast cell under the control of a promoter that provides increased expression, using multiple copies of the nucleic acid, and/or using a heterologous enzyme, a variant enzyme (e.g., one including one or more amino acid substitutions), or a variant heterologous enzyme that provides a high levels of enzymatic activity.

[0038] The engineered yeast used to produce steviol glycosides by fermentation can also include a pathway to convert IPP and/or DMAPP to steviol. For example, in some aspects the engineered yeast can include exogenous nucleic acids expressing the following enzymes: (a3) geranyl geranyldiphosphate synthase (GGPPS), (b3) copalyl diphosphate synthase (CPS), (c3) kaurene synthase (KS), (d3) kaurene oxidase (KO), and (e3) kaurenoic acid 13-hydroxylase (KAH). Enzymes of the mevalonate pathway converts IPP and/or DMAPP to steviol as follows: IPP/ DMAPP → geranyl geranyldiphosphate → copalyl diphosphate → kaurene → kaurenoic acid → steviol. (See FIG. 3) Exogenous nucleic acids

encoding enzymes (a3) - (e3) that are heterologous to the yeast cell can be placed under the control of a promoter that provides increased expression, using multiple copies of the nucleic acid, and/or using a variant enzyme (e.g., one including one or more amino acid substitutions), or a variant heterologous enzyme that provides a high levels of enzymatic activity.

[0039] The methods of the disclosure for producing steviol glycoside(s) by fermentation can use engineered yeast including one or more steviol biosynthesis enzymes selected from the group of geranylgeranyl diphosphate synthase, copalyl diphosphate synthase, kaurene synthase, kaurene oxidase, kaurenoic acid 13-hydroxylase (KAH), steviol synthetase, deoxyxylulose 5-phosphate synthase (DXS), D-1-deoxyxylulose 5-phosphate reductoisomerase (DXR), 4-diphosphocytidyl-2-C-methyl-D-erythritol synthase (CMS), 4-diphosphocytidyl-2-C-methyl-D-erythritol kinase (CMK), 4-diphosphocytidyl-2-C-methyl-D-erythritol 2,4-cyclodiphosphate synthase (MCS), 1-hydroxy-2-methyl-2(E)-butenyl 4-diphosphate synthase (HDS), 1-hydroxy-2-methyl-2(E)-butenyl 4-diphosphate reductase (HDR), acetoacetyl-CoA thiolase, truncated HMG-CoA reductase, mevalonate kinase, phosphomevalonate kinase, mevalonate pyrophosphate decarboxylase, cytochrome P450 reductase etc.

[0040] The engineered yeast used to produce steviol glycosides by fermentation can have any pathway to convert steviol to a steviol glycoside. If more than one steviol glycoside pathway enzymes are present in the engineered yeast, the yeast may be able to produce different steviol glycosides. For example, the yeast may be able to produce two, three, four, five, six, seven, eight, nine, ten, or more than 10 different steviol glycoside species. Reb A, Reb B, Reb D and/or Reb M are produced by the engineered yeast cells of the method of the invention.

[0041] The steviol glycoside pathway can include one or more uridine diphosphate (UDP) glycosyltransferases (UGTs) that mediate the transfer of glycosyl residues from activated nucleotide sugars to acceptor molecules. In the case of a steviol glycoside pathway, a monosaccharide unit can be transferred to a hydroxyl or carboxyl moiety on a steviol or steviol glycoside molecule, or to a hydroxyl group on a glucose group that is attached to the steviol base. See FIG. 4. UGTs have been classified into families and subfamilies based on sequence homology. See Li et al., 2001, J. Biol. Chem. 276:4338-4343. A superfamily of over 100 genes encoding UGTs, each containing a 42 amino acid consensus sequence, has been identified in the model plant *Arabidopsis thaliana*, and genes encoding UGTs have also been identified in several other higher plant species.

[0042] Exemplary UDP-glucosyltransferase can be any UDP-glucosyltransferase capable of adding at least one glucose unit to the steviol and or steviol glycoside substrate to provide the desired steviol glycoside. The engineered yeast can include one or more UDP-glucosyltransferase selected from group UGT74G1 (SEQ ID NO: 1), UGT85C2 (SEQ ID NO: 2), UGT76G1 (SEQ ID NO: 3), UGT91D2 (SEQ ID NO: 4), and also UGTs having substantial (e.g. >85%, >75%, >65%, >55%, >45% and >35%) identity to these polypeptides. An engineered yeast can include one or more exogenous nucleic acid molecule(s) that code for these UGTs.

[0043] The engineered yeast can also include one or more UGT and UDP-glucose recycling enzyme(s). An exemplary UDP-glucosyltransferase capable of adding at least one glucose unit to rubusoside to form stevioside is UGT91D2 (SEQ ID NO: 4). An exemplary UDP-glucosyltransferase capable of adding at least one glucose unit to stevioside to form rebaudioside A is UGT76G1 (SEQ ID NO: 3). An exemplary UDP-glucosyltransferase capable of adding at least one glucose unit to rebaudioside A to form rebaudioside D is UGT91D2 (SEQ ID NO: 4). An exemplary UDP-glucosyltransferase capable of adding at least one glucose unit to rebaudioside D to form rebaudioside M is UGT76G1 (SEQ ID NO: 3).

[0044] Exemplary publications that describe engineered microorganisms for steviol glycoside production include, for example, U.S. Patent Application Publication No. 2014/0357588, International Patent Application Publication Nos. WO 2014/193934, WO 2014/193888, and WO 2014/122227.

[0045] An engineered yeast useful for the production of steviol glycosides may express the following enzymes: geranylgeranyl diphosphate synthase (GGPPS), ent-copalyl diphosphate synthase (CDPS), kaurene oxidase (KO), kaurene synthase (KS); steviol synthase (KAH), cytochrome P450 reductase (CPR), UGT74G1, UGT76G1, UGT91D2, UGT85C2 and a EUGT11. WO2014/122227 describes an engineered yeast strain that express these enzymes. The UDP-glucosyltransferases can be a gene encoding a polypeptide for example, UGT74G1 (SEQ ID NO: 1), UGT85C2 (SEQ ID NO: 2), UGT76G1 (SEQ ID NO: 3), UGT91D2 (SEQ ID NO: 4), and a EUGT11 (SEQ ID NO: 13); these genes encode polypeptides capable of carrying out a number of reactions such as a) a gene encoding a polypeptide capable of beta 1,2 glucosylation of the C2' of the 19-O glucose of a steviol glycoside; (b) a gene encoding a polypeptide capable of beta 1,2 glucosylation of the C2' of the 13-O-glucose of a steviol glycoside; (c) a gene encoding a polypeptide capable of beta 1,3 glucosylation of the C3' of the 19-O-glucose of a steviol glycoside; (d) a gene encoding a polypeptide capable of beta 1,3 glucosylation of the C3' of the 13-O-glucose of a steviol glycoside; (i) a gene encoding a polypeptide capable of glucosylation of the 13-OH of steviol or a steviol glycoside; (j) a gene encoding a polypeptide capable of glucosylation of the C-19 carboxyl of steviol or a steviol glycoside. For example, UGT85C2 carries out reaction (i); UGT74G1 carries out reaction (j); UGT91D2 carries out reactions (a; weakly), (b); UGT76G1 carries out reactions (c) and (d) EUGT11 carries out reactions (a), (b; less well).

[0046] The engineered host cell may express one or more exogenous nucleic acid(s) encoding one or more of the following proteins heterologous to the host cell: GGPPS polypeptide, an enf-copalyl diphosphate synthase (CDPS)

polypeptide, a kaurene oxidase (KO) polypeptide, a kaurene synthase (KS) polypeptide; a steviol synthase (KAH) polypeptide, a cytochrome P450 reductase (CPR) polypeptide, a UGT74G1 polypeptide, a UGT76G1 polypeptide, a UGT91 d2 polypeptide, and a EUGT11 polypeptide.

[0047] In another embodiment, the engineered yeast expresses one or more exogenous nucleic acid(s) encoding one or more of the following proteins heterologous to the host cell: a GGPPS polypeptide, a truncated *Zea mays* CDPS polypeptide, an *Arabidopsis thaliana* KS polypeptide a *Stevia rebaudiana* KO polypeptide, an *Arabidopsis thaliana* ATR2 polypeptide, an *Oryza sativa* EUGT 11 polypeptide, a SrKAH1 polypeptide, a *Stevia rebaudiana* CPR8 polypeptide, an *Stevia rebaudiana* UGT85C2 polypeptide, an *Stevia rebaudiana* UGT74G1 polypeptide, a *Stevia rebaudiana* UGT76G1 polypeptide, a *Stevia rebaudiana* UGT91D2 variant or functional homolog, and a UGT91D2e-b polypeptide.

[0048] Steviol glycoside-producing *S. cerevisiae* strains were constructed using methods as described in WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328. The following sequences were used for construction of a parent strain (Strain A): a recombinant gene encoding a *Synechococcus* sp GGPPS polypeptide (SEQ ID NO:6), a recombinant gene encoding a truncated *Zea mays* CDPS polypeptide (SEQ ID NO:7), a recombinant gene encoding an *Arabidopsis thaliana* KS polypeptide (SEQ ID NO:8), a recombinant gene encoding a recombinant *Stevia rebaudiana* KO polypeptide (SEQ ID NO:9, SEQ ID NO:10), a recombinant gene encoding an *Arabidopsis thaliana* ATR2 polypeptide (SEQ ID NO:11, SEQ ID NO:12), a recombinant gene encoding an *Oryza sativa* EUGT 11 polypeptide (SEQ ID NO:13), a recombinant gene encoding an SrKAH1 polypeptide (SEQ ID NO:14, SEQ ID NO:15), a recombinant gene encoding an *Stevia rebaudiana* CPR8 polypeptide (SEQ ID NO:16, SEQ ID NO:17), a recombinant gene encoding an *Stevia rebaudiana* UGT85C2 polypeptide (SEQ ID NO:2), a recombinant gene encoding an *Stevia rebaudiana* UGT74G1 polypeptide (SEQ ID NO:1), a recombinant gene encoding an *Stevia rebaudiana* UGT76G1 polypeptide (SEQ ID NO:3), and a recombinant gene encoding an *Stevia rebaudiana* UGT91D2 variant (or functional homolog), UGT91D2e-b, (SEQ ID NO:4) polypeptide produced steviol glycosides.

[0049] The UGT91D2e-b variant of UGT91D2 (SEQ ID NO:5 from PCT/US2012/050021) includes a substitution of a methionine for leucine at position 211 and a substitution of an alanine for valine at position 286. (Additional variants, except T144S, M152L, L213F, S364P, and G384C variants, described in Table 12 and Example 11 of PCT/US2012/050021 could be used.) GeneArt codon-optimized sequence encoding a *Stevia rebaudiana* UGT91D2e-b with the amino acid modifications L211M and V286A (SEQ ID NO:4 for amino acid sequence; codon optimized nucleotide sequence is set forth in SEQ ID NO:5).

[0050] Strain B is derived from the parent strain described above and additionally includes a codon-optimized CPR1 from *Stevia rebaudiana* (SEQ ID NO: 18 corresponding to amino acid SEQ ID NO: 19).

[0051] The engineered yeast may express one or more exogenous nucleic acid(s) encoding one or more of the following proteins heterologous to the yeast: GGPPS polypeptide, an ent-copalyl diphosphate synthase (CDPS) polypeptide, a kaurene oxidase (KO) polypeptide, a kaurene synthase (KS) polypeptide; a steviol synthase (KAH) polypeptide, a cytochrome P450 reductase (CPR) polypeptide, a UGT74G1 polypeptide, a UGT76G1 polypeptide, a UGT91D2 polypeptide, and a EUGT11 polypeptide.

[0052] The engineered yeast may express one or more exogenous nucleic acid(s) encoding one or more of the following proteins heterologous to the yeast: a GGPPS polypeptide, a truncated *Zea mays* CDPS polypeptide, an *A. thaliana* KS polypeptide a *S. rebaudiana* KO polypeptide, an *A. thaliana* ATR2 polypeptide, an *O. sativa* EUGT 11 polypeptide, a SrKAH1 polypeptide, a *S. rebaudiana* CPR8 polypeptide, an *S. rebaudiana* UGT85C2 polypeptide, an *S. rebaudiana* UGT74G1 polypeptide, a *S. rebaudiana* UGT76G1 polypeptide, a *S. rebaudiana* UGT91D2 variant or functional homolog, and a UGT91D2e-b polypeptide

[0053] The engineered host cells (e.g., yeast cells) can be used to produce the desired steviol glycosides such as rebaudioside M, rebaudioside D, and rebaudioside A by fermentation. "Fermentation" as used here refers to the assimilation of the medium such as the carbohydrates to produce the desired steviol glycosides through aerobic fermentation. The term "medium" or grammatical equivalents such as "media" refers to a liquid composition in which the engineered yeast or fungus can be maintained, grown, or fermented, or combinations thereof. A "medium" may also be referred to as a "broth" or "cell culture," and terms such as "fermentation" that qualifies the term "medium" and its equivalents may be used to more specifically define the type of cellular activity that is occurring in the medium.

[0054] A medium can be defined with regards to the components present in the medium, and amounts thereof, including, but not limited to: (a) carbon sources, including carbohydrates such as glucose and starch products such as maltodextrin; (b) nitrogen sources, such as yeast nitrogen base, ammonium hydroxide, urea, ammonium sulfate, yeast extract or any combination thereof; (c) salts, such as potassium phosphate (monobasic, dibasic), magnesium sulfate, sodium chloride, and calcium chloride; (d) vitamins, such as biotin, calcium pantothenate, folic acid, (myo)-inositol, nicotinic acid, p-aminobenzoic acid, pyridoxine HCl, riboflavin, thiamine HCL, and citric acid; and/or (e) trace metals such as boric acid, copper sulfate, cobalt chloride, calcium chloride, potassium iodide, ferric chloride, magnesium sulfate, manganese chloride, sodium molybdate, and zinc sulfate. The medium can also be defined with regards to its pH, and biocompatible acids, bases, and buffers that are used to control the pH in the medium.

[0055] Fermentation of the engineered yeast can be performed using starch and/or sugar containing plant material

derivable from any plant and plant part, such as tubers, roots, stems, leaves and seeds. Starch and/or sugar comprising plant material can be obtained from cereal, such as barley, wheat, maize, rye, sorghum, millet, barley, potatoes, cassava, or rice, and any combination thereof. The starch-and/or sugar comprising plant material can be processed, such as by methods such as milling, malting, or partially malting. The fermentation media may include a treated starch. For example, the fermentation media can include a partially hydrolyzed starch. The partially hydrolyzed starch can include high molecular weight dextrins and high molecular weight maltodextrins. A partially hydrolyzed starch product can be used that have amounts of starch and starch degradation products within desired ranges.

[0056] Fermentation can be carried out under conditions and in medium suitable for production of the desired steviol glycosides such as rebaudioside M, rebaudioside D, and rebaudioside A. The fermentation conditions generally use oxygen (aerobic conditions), a carbon source, and a nutrient (nitrogen) base. Fermentation can be carried out using a fed batch or continuous process. The fermentation minimal medium includes glucose (5g/L), ammonium sulfate (5g/L), potassium dihydrogenphosphate (3g/L), magnesium sulphate (0.5 g/L), trace elements, and vitamins (e.g., see, Verduyn, C. et al. (1992) *Yeast* 8, 501-517). The pH of the fermentation media can be at about pH 4 to pH 5 and the fermentation can be carried out at a temperature at about 30° C. Fermentation can be carried out using a first growth phase in base medium, followed by a longer feeding phase using a glucose-containing defined feed medium (with trace metals, vitamins, and salts).

[0057] Fermentation may be carried out using a pH greater than 5.0. The pH may range from about 5.5 to 8 or 5.8 to 7.5. The engineered yeast may be grown initially at a lower pH such as less than 6.0 and fermentation carried out a higher pH such as greater than 5.0. Other methods of fermentation for producing the desired steviol glycosides are described in the application titled "Fermentation Methods for Producing Steviol Glycosides Using High pH and Compositions Obtained Therefrom," U.S. Pat. No. 10815513, and in International PCT application titled "Fermentation Methods for Producing Steviol Glycosides," PCT/US2016/034728 and filed concurrently with the present application.

[0058] Optionally, fermentation can be carried out in media containing other intermediates such as steviol, rebudioside A, stevioside, rubusoside, steviol monoside, steviolbioside, dulcoside A, rebaudioside C, rebaudioside E, and the like.

[0059] The fermentation step can also be referred to as a "steviol glycoside producing step." The fermentation medium may be maintained at a temperature less than 40° C during the fermentation step. The fermentation medium may be maintained at a temperature in the range of about 25 to 35° C during the fermentation step.

[0060] After fermentation, but before any treatment (e.g., heat treatment) to release or enrich for the desired steviol glycosides, the fermentation broth may contain only a fraction of the desired glycosides, with the majority of the desired glycosides within the engineered host cell. Disclosed are methods of releasing or enriching for the desired glycoside from the yeast host cell by using heat.

[0061] The heat treatment may be carried out on the whole cell fermentation broth, which contains the engineered yeast capable of producing the desired steviol glycosides and the desired steviol glycoside products. The engineered yeast after fermentation but before any treatment (e.g. heat treatment) may be separated from the fermentation broth and transferred into any suitable composition such as a liquid media.

[0062] The fermentation broth or composition containing the transferred yeast cells is heated at temperatures ranging from 70°C to 75°C for 5 minutes to 48 hours. Heating may be 70°C to 75°C for 5 minutes to 2 hours, or 75°C for 5 minutes to 2 hours. The heating may be at 70°C to 75°C for 5 minutes or from 70°C to 75°C for 2 hours.

[0063] The heat treatment releases the steviol glycosides into the medium or enriches the steviol glycoside content of the medium such that the extracellular glycoside content is greater than 90% of the total (i.e., both intracellular and extracellular) glycoside content in the fermentation broth or composition. The heat treatment step may release steviol glycosides into the fermentation medium such that the extracellular steviol glycoside content is greater than 80%, 85%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99%, of the total (i.e., both intracellular and extracellular) steviol glycoside content in the fermentation broth or The heat treatment step may release steviol glycosides into the fermentation medium such that the extracellular steviol glycoside content is in the range of about 80 to 100%, 85 to 100%, 90 to 100%, 90 to 99%, 92 to 98%, or 90 to 95% of the total (i.e., both intracellular and extracellular) steviol glycoside content in the fermentation broth or composition.

[0064] The heat treatment of the fermentation broth or composition releases Reb A, Reb B, Reb M, Reb D or combinations thereof. In some embodiments, the heat treatment enriches for Reb A such that the extracellular Reb A is greater than 50 % of the total (i.e., both intracellular and extracellular) Reb A present in the fermentation broth or composition. In some embodiments, the extracellular Reb A is greater than 40 %, 45%, 48%, 52%, 55%, 57%, 60%, 62%, 65%, 67%, 70%, 75%, 80%, 85%, 90%, 95%, or 99% of the total (i.e., both intracellular and extracellular) Reb A present in the fermentation broth or composition. In some embodiments, the Reb A amount, can be released extracellularly in the range of 30% to 100 %, 50 % to 100 %, 80 % to 90 % of the total amount of Reb A produced during the fermentation.

[0065] In some embodiments, the heat treatment enriches for Reb D and/or Reb M such that the Reb D and/or Reb M released extracellularly is greater than 33 % of the total (i.e. both intracellular and extracellular) Reb D and/or Reb M present in the fermentation broth or composition. In some embodiments, the extracellular Reb D, Reb M, and/or the total of Reb D and Reb M is greater than 40 %, 45%, 48%, 52%, 55%, 57%, 60%, 62%, 65%, 67%, 70%, 75%, 80%, 85%,

90%, 95%, or 99% of the total (i.e., both intracellular and extracellular) Reb D, Reb M, and/or the total of Reb D and Reb M present in the fermentation broth or composition. In some embodiments, the combined amount of Reb D and Reb M, released extracellularly can be present in the range of 30% to 100%, 50% to 100%, 80% to 90% of the total amount of Reb D and/or Reb M present in the fermentation broth or composition.

[0066] In other embodiments, the Reb D released, can be present extracellularly in the range of 30% to 100%, 50% to 100%, 80% to 90% of the total amount of Reb D present in the fermentation broth or composition.

[0067] In other embodiments, the Reb M released, can be present extracellularly in the range of 30% to 100%, 50% to 100%, 80% to 90% of the total amount of Reb M present in the fermentation broth or composition.

[0068] In some embodiments, the Reb B released, can be present extracellularly in the range of 25% to 100%, 30% to 95%, 40% to 80% of the total amount of Reb B present in the fermentation broth or composition. In some embodiments, the extracellular Reb B is greater than 30%. 33%, 35%, 40%, 45%, 48%, 52%, 55%, 57%, 60%, 62%, 65%, 67%, 70%, 75%, 80%, 85%, 90%, 95%, or 99% of the total (i.e., both intracellular and extracellular) Reb B present in the fermentation broth or composition. The disclosed heat treatment may not result in significant degradation from Reb DM and/or Reb A to Reb B.

[0069] The disclosed heat treatment may result in the release of the desired glycosides yet may not result in extensive lysis of the cell. Cell lysis may be determined, for example, by the amount of nitrogen or phosphorus or both released into the fermentation broth (e.g., cell free) or composition after the fermentation and heat treatment. The heating of the fermentation broth or composition may not increase nitrogen and/or phosphorus content in the cell free fermentation broth or composition. The extracellular nitrogen content in the cell free fermentation broth or composition after the heat treatment may not increase by more than 45% of the total (i.e. extracellular and intracellular) nitrogen present in the whole cell fermentation broth or composition. The extracellular phosphorous content in the fermentation broth or composition after the heat treatment may not increase by more than 70% of the total (i.e. extracellular and intracellular) phosphorous present in the cell-containing fermentation broth or composition.

[0070] The extracellular nitrogen content may not increase by more than 45% of the total nitrogen present in the cell-containing fermentation broth or composition and/or the extracellular phosphorous content in the fermentation broth or composition after the heat treatment may not increase by more than 70% of the total phosphorous present in the cell-containing fermentation broth or composition when heated at 70°C to 75°C for 5 minutes to 2 hours.

[0071] When the fermentation medium is heated at 70°C to 75°C for 5 minutes to 48 hours, the extracellular nitrogen content may not increase by more than 30%, 33%, 35%, 37%, 40%, 42%, 47%, or 50% of the total nitrogen present in the cell-containing fermentation broth or When the fermentation medium is heated at 70°C to 75°C for 5 minutes to 48 hours, the extracellular phosphorus content may not increase by more than 50%, 55%, 60%, 63%, 65%, 67%, 73%, 75%, 77%, or 80% of the total phosphorus present in the cell-containing fermentation broth.

[0072] The intracellular and/or extracellular content of various components of the fermentation broth (e.g., steviol glycosides, nitrogen, or phosphorus) can be determined as described in Example 1.

[0073] The heat treatment of the fermentation broth may also be evaluated based on the cell viability. While the desired glycosides may be released from the yeast cell into the fermentation broth, the yeast viability is decreased while limiting cell lysis. Cell lysis may be determined, for example, by plating the whole cell broth on Potato Dextrose Antibiotic Agar or by cell viability counter (e.g., Cellometer™).

[0074] Heating may be carried out by using a heat exchanger in which heat is transferred to hot water or steam. Heating may be carried out using a heat exchanger with a holding loop, direct steam injection, or bulk heating of the material utilizing steam applied to a jacketed vessel. The yeast may be concentrated for example, by centrifugation or filtration, before a heat treatment. The centrifuged or filtered yeast may be transferred into other suitable media or composition and then used for heating.

[0075] Following the release of the desired recombinant product (e.g. steviol glycosides) from the host cell into the medium, thereby enriching for the desired product, various procedures can be used for further isolation and purification of the desired recombinant product. Such procedures include centrifugation, ultrafiltration, precipitation and chromatographic procedures.

[0076] After the steviol glycosides are released or enriched, the steviol glycosides may be further purified by preparative HPLC as described, for example in International Patent Application Publication No. WO 2009/140394.

[0077] The fermentation broth or composition after the heat treatment can then be centrifuged or filtered to remove the engineered cells. The fermentation broth can optionally be treated to remove low molecular weight components (glucose, basic nutrients, and salts), such as by membrane dialysis.

[0078] If it is desired to provide a composition with steviol glycosides including Reb A, Reb B, Reb M, Reb D, and combinations thereof in enriched or purified form, or where Reb A, Reb B, Reb M, Reb D, and combinations thereof are separated from other steviol glycosides, or separated from one another, further purification can be carried out. Such enrichment or purification of steviol glycoside components can be carried out on liquid fermentation media, or the fermentation media can then be dried down prior to purification. For example, fermentation media can be dried down using lyophilization to form a dry composition (e.g., powder or flakes) including steviol glycosides with Reb A, Reb B,

Reb M, Reb D, and combinations thereof that can be subsequently processed.

[0079] Dried fermentation broth or composition enriched for steviol glycosides including Reb A, Reb B, Reb M, and/or Reb D, may be used as the starting material for purification. For example, a solvent or solvent combination can be added to the dried fermentation broth to dissolve or suspend materials that include the steviol glycosides. An exemplary combination for dissolving the steviol glycosides is a mixture of water and an alcohol (e.g., 50:50 ethanol:water). To facilitate dissolving or resuspending, the dried broth materials can be heated at a temperature above room temperature, such as in the range of 40°C - 60°C. Mechanical disruption of the dried fermentation broth materials can also be performed, such as by sonication. The dissolved or resuspended fermentation broth materials can be filtered using a micron or sub-micron filtration system prior to further purification, such as by preparative chromatography.

[0080] Dried fermentation broth or composition enriched for steviol glycoside compounds can be subjected to other purification processes, such as reverse phase liquid chromatography. A suitable resin can be used to retain steviol glycoside compounds in the column, with removal of hydrophilic compounds which get washed through the column with a liquid such as water. Elution of steviol glycosides including Reb A, Reb B, Reb M, Reb D, and combinations thereof from the column can be accomplished using a suitable solvent or solvent combination such as acetonitrile or methanol.

[0081] The steviol glycosides such as Reb A, Reb B, Reb M, Reb D, and combinations thereof can be purified using preparative liquid chromatography, such as high pressure liquid chromatography (HPLC) or ultra high pressure liquid chromatography (UHPLC). A steviol glycoside composition with Reb A, Reb B, Reb M, Reb D, and combinations thereof can be dissolved in a mobile phase, such as a mixture of water and an solvent (e.g., methanol, ethanol, acetonitrile) at a desired ratio (e.g., 60% water, 40% methanol, v/v). The composition can also be heated to enhance dissolution of the steviol glycoside material, such as heating at about 50°C. The solution can also be filtered prior to injection into the column, such as using a 0.2 µm filter. Phenomenex Kinetex XB-C18 5 µm, core-shell silica solid support, and stationary phase of C18 with iso-butyl side chains and TMS endcapping. The flow rate through the column can be based on column properties (such as about 20 mL/min), with a maximum pressure of 400 bar. Reb A, Reb B, Reb M, Reb D, and combinations thereof can be identified by their elution times from the column. In exemplary flow conditions Reb A, Reb B, Reb M, Reb D, and combinations thereof can elute from the column within 60 minutes. One of skill in the art will appreciate that the elution times for the Reb A, Reb B, Reb M, Reb D, and combinations thereof can vary with changes in solvent and/or equipment. One of skill in the art will also understand that although the process described below assumes certain order of the described steps, this order can be altered in some cases.

[0082] The fermentation broth or composition enriched for steviol glycosides including Reb A, Reb M, and/or Reb D, may be dried and used as the starting material for further purification. A steviol glycoside composition with Reb A, Reb M, Reb D, and combinations thereof can be dissolved in water and passed through adsorption chromatographic column to remove hydrophilic impurities in the composition. Adsorbed steviol glycosides can be desorbed from the column with ethanol/water mixture. Upon removal of the ethanol, the steviol glycoside liquid stream can be further purified with ion exchange chromatography to remove ions and color bodies. Additional treatment with activated carbon could be performed. To further separate specific steviol glycosides from others, a crystallization step could be used. The crystals may be enriched with Reb M, Reb D, Reb A, or other steviol glycosides. The crystals can be dried under heat and vacuum or can be re-slurried with water and spray dried.

Example 1

Production Of Reb D, Reb M And Reb A In Fed Batch Fermentation With Ammonium Hydroxide As The Primary Nitrogen Source

[0083] For inoculum preparation, a yeast strain designated strain B was cultured in 150 mLs of seed flask medium in 1 liter shake flasks at 250 rpm and 30°C for 20-24 hours.

Table 1 - Seed Flask Medium

Component	Formula	Concentration	Units
Biospringer 0251 yeast extract		7.5	g/L
Glucose monohydrate	$C_6H_{12}O_6 \cdot H_2O$	22.0	g/L

[0084] For the fermentation, 75 mL of seed culture was transferred into initial fermentation medium (Tables 2, 3 and 4) with a starting volume of 0.75 liters. Temperature was maintained at 30°C throughout. The air flow rate was 1.75 SLPM and the agitation rate was automatically controlled to increase in a stepwise manner from 400 to 900 rpm during the fermentation. Glucose concentration was kept limiting by controlling flow rates of feed medium (Table 5). A 2-phase

feeding strategy involved an initial exponential phase beginning at 10 hours with a growth rate of $\mu = 0.12$ 1/h while the second phase of feeding (or feed phase II) started at 33 hours with a constant flow rate of 0.180 mls/minute. Feeding was continued until a final volume of 1.95 liters was obtained by 120 hours.

[0085] pH was maintained at pH 5 with 12% NH_4OH throughout fermentation. Antifoam addition was controlled by utilization of foam control probes with 10 wt% antifoam solution (Ivanhoe 1163B). The medium was based on Verduyn et al (Verduyn C, Postma E, Scheffers WA, Van Dijken JP. Yeast. 1992 Jul; 8(7):501-17) with modifications as described in Tables 2 through 5.

Table 2 - Initial Fermentation Medium

Component	Formula	Concentration	Units
Glucose monohydrate	$\text{C}_6\text{H}_{12}\text{O}_6 \cdot \text{H}_2\text{O}$	22.0	g/L
Ammonium sulfate	$(\text{NH}_4)_2\text{SO}_4$	5.0	g/L
Monobasic potassium phosphate	KH_2PO_4	3.0	g/L
Magnesium sulfate heptahydrate	$\text{MgSO}_4 \cdot 7 \text{H}_2\text{O}$	0.5	g/L
Trace metals stock		10.0	ml/L
Vitamin stock		12.0	ml/L

Table 3 - Trace Metals Stock Solution

Component	Formula	Concentration	Units
Disodium edetate	$\text{C}_{10}\text{H}_{14}\text{N}_2\text{Na}_2\text{O}_8 \cdot 2\text{H}_2\text{O}$	15	g/L
Zinc sulfate heptahydrate	$\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$	4.5	g/L
Manganese (II) chloride tetrahydrate	$\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$	1.026	g/L
Cobalt (II) chloride hexahydrate	$\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$	0.32	g/L
Copper (II) sulfate heptahydrate	$\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$	0.3	g/L
Sodium molybdate dihydrate	$\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$	0.4	g/L
Calcium chloride dihydrate	$\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$	3	g/L
Iron (II) sulfate heptahydrate	$\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$	3	g/L
Boric acid	H_3BO_3	1	g/L
Potassium iodide	KI	0.1	g/L

Table 4 - Vitamin Stock Solution

Component	Formula	Concentration	Units
d-Biotin	$\text{C}_{10}\text{H}_{16}\text{N}_2\text{O}_3\text{S}$	50	mg/L
Calcium pantothenate	$\text{C}_{18}\text{H}_{32}\text{CaN}_2\text{O}_{10}$	1000	mg/L
Nicotinic acid	$\text{C}_6\text{H}_5\text{NO}_2$	1000	mg/L
Thiamine hydrochloride	$\text{C}_{12}\text{H}_{17}\text{ClN}_4\text{OS} \cdot \text{HCl}$	1000	mg/L
Pyridoxine hydrochloride	$\text{C}_8\text{H}_{11}\text{NO}_3 \cdot \text{HCl}$	1000	mg/L
p-aminobenzoic acid	$\text{C}_7\text{H}_7\text{NO}_2$	200	mg/L
myo-inositol	$\text{C}_6\text{H}_{12}\text{O}_5$	25000	mg/L

Table 5 - Fermentation Feed Medium

Component	Formula	Concentration	Units
Glucose monohydrate	$C_6H_{12}O_5 \cdot H_2O$	660	g/L
Urea (in urea treatments only)	NH_2CONH_2	33	g/L
Antifoam		1.3	g/L
Potassium sulfate	K_2SO_4	4.2	g/L
Sodium sulfate	Na_2SO_4	0.336	g/L
Magnesium sulfate heptahydrate	$MgSO_4 \cdot 7H_2O$	6.12	g/L
Monobasic potassium phosphate	KH_2PO_4	10.8	g/L
Trace metal stock		14.4	mL/L
Vitamin stock		14.4	mL/L

Example 2

Enrichment or Release and Analysis of Steviol Glycosides

[0086] The fermentation broth generated from a recombinant yeast fermentation as described in Example 1 was used. The measurement of total steviol glycosides including intracellular and extracellular concentrations as well as the total nitrogen content (intracellular and extracellular) and total phosphorous content (intracellular and extracellular) was carried out by agitating the fermentation samples including the yeast cells to ensure that yeast cells were mixed and did not settle to the bottom of the vial. 100 μ L of the mixed fermentation broth were pipetted into a 2 mL microcentrifuge tube. 900 μ L of 61% methanol (extraction solvent) was added into the 2 ml microcentrifuge tube and agitated by placing on a sample rotator for 10 min to extract the steviol glycosides. The samples were then centrifuged at 10K rpm in a microcentrifuge for 3 min and the clarified supernatant was pipetted into an autosampler vial for analysis.

[0087] The fermentation broth (including cells) was heated at various temperatures and for various times. Heating was accomplished using a hot water bath and the sample was prepared as follows: 1 ml of the fermentation broth (including cells) was placed into a headspace vial for each temperature and time point studied. The vials were sealed and placed into the water bath and allowed to equilibrate for 5 minutes at each temperature studied prior to maintaining the samples at the desired temperature for the desired amount of time. At the end of the designated time, the samples were removed from the water bath and were cooled at 4°C. 4 ml of sterile water was added to each vial after which the sample was transferred to a 14 ml sterile tube. Samples were vortexed and 1 ml was removed for determining cell viability by the plating method. The remainder of the sample was centrifuged at 4700 rpm for 30 minutes at room temperature to obtain clear, cell-free supernatant. The cell-free supernatant was removed for further analysis, for example the extracellular content of steviol glycosides, nitrogen, and phosphorus in the fermentation medium (Phosphorous via ICP, Nitrogen via Antek and glycoside via UHPLC).

[0088] The amount of glycosides released into the fermentation broth after the heat treatment was determined. Reb A, Reb D and Reb M were measured by ultra-performance liquid chromatography (UPLC). The cell free supernatant was also used to evaluate nitrogen and phosphorous release into the fermentation broth. Nitrogen levels were analyzed using the combustion method and an ANTEK™ nitrogen analyzer (Antek 9000 nitrogen detector utilizing nitrogen combustion and reaction with ozone to produce metastable nitrogen dioxide, which is detected via chemiluminescence and phosphorous levels were analyzed using inductively coupled plasma spectrometry ICP, Perkin Elmer Optima 3200DV, UV detection at 213.617nm, samples dissolved in 0.5% HNO_3). Yeast viability was determined using the Potato Dextrose Antibiotic Easy gel plates (Microbiology Labs test kits), incubated at room temperature for 5 days. UPLC Method for glycoside separation

[0089] The steviol glycosides were separated using an Eclipse C18 Plus RRHD column (3mm x 150mm x 1.8um) with Eclipse Plus C18 guard (column 3mm x 5mm x 1.8um). The mobile phase used was channel A: 10mM phosphate buffer, pH 2.6 and channel B acetonitrile. The flow rate was 0.6 ml/min, the column temperature was 40° C and the detection was performed at ultraviolet absorption of 210 nm. The gradient elution profile in which the ratio of phosphate buffer to acetonitrile changed is shown below:

- i. Initial: 80% A/20% B
- ii. 7 minutes: 70% A/30% B
- iii. 12 minutes: 70% A/30% B
- iv. 15 minutes: 45% A/55% B
- v. 18 minutes: 45% A/55% B

- vi. 20 minutes: 20% A/80% B
- vii. 25 minutes: 20% A/80% B
- viii. 27 minutes: 15% A/85% B
- ix. 30 minutes: 15% A/85% B
- x. 30.5 minutes: 80% A/20% B
- xi. 35 minutes: 80% A/20% B

[0090] The UPLC results, shown as the % of extracellular Reb A, Reb D & M released from the yeast cell after the heat treatment is shown in Table 6. The % of the extracellular nitrogen and phosphorus are also shown.

[0091] Total nitrogen in the fermentation broth including cells was determined using a Leco Nitrogen Analyzer (Leco TruMac N, St. Joseph, MI). 1 ml of homogeneously mixed fermentation broth including cells were applied to a nickel lined ceramic combustion container for nitrogen combustion analysis.

[0092] Total phosphorus in the fermentation broth including cells was determined by ashing 5 mL of homogeneously mixed fermentation broth with a propane torch in a ceramic crucible. The resultant ash was held at 525°C overnight prior to dissolving the ash with 10 mls of 0.5% HNO₃. Phosphorus content in the nitric acid solution was determined using ICP.

Table 6.

Heat treatment condition		% extracellular Reb D&M	% extracellular Reb A	% extracellular Reb B	Reb B (% extracellular)	% extracellular N	% extracellular P
Temperature, °C	Time hr						
24	1	34%	48%	20%	28%	20%	45%
	2	34%	48%	21%	28%	21%	45%
	24	43%	52%	25%	30%	25%	51%
	48	47%	58%	30%	34%	30%	56%
	70	48%	59%	31%	34%	31%	55%

EP 3 302 097 B1

	Temperature, °C	Time, hr	% extracellular D&M	% extracellular A	% extracellular B	% extracellular N	% extracellular P
5	50	0.08	44%	54%	30%	23%	46%
		0.5	56%	67%	36%	28%	55%
		1	72%	88%	41%	37%	65%
		1.5	76%	85%	44%	40%	67%
10		2	88%	91%	46%	50%	76%
		24	98%	99%	58%	88%	93%
		48	95%	97%	60%	91%	92%
		70	93%	96%	61%	91%	97%
15		72	92%	96%	60%	93%	95%
		96	91%	96%		91%	95%
		120	87%	92%		92%	95%

	Temperature, °C	Time, hr	% extracellular D&M	Reb B (% extracellular)	% extracellular N	% extracellular P
20	70	0.08	101%		36%	64%
		0.25	101%	45%	35%	62%
25		0.5	101%		36%	64%
		0.75	99%	45%	35%	66%
		1	101%		36%	66%
		1.5	101%	46%	37%	65%
30		2	99%		37%	64%

	Temperature, °C	Time, hr	% extracellular D&M	% extracellular A	Reb B (% extracellular)	% extracellular N	% extracellular P
35	75	0.08	96%	92%	44%	35%	65%
		0.5	98%	94%	45%	36%	68%
		1	96%	94%	46%	35%	69%
40		2	95%	93%	47%	36%	67%
		24	80%	82%	69%	38%	74%
		48	74%	78%	86%	40%	82%
45		70	68%	72%	98%	44%	88%

	Temperature, °C	Time, hr	% extracellular D&M	% extracellular A	Reb B (% extracellular)	% extracellular N	% extracellular P
50	95	0.08	98%	93%	47%	32%	65%
		0.25	100%	99%	49%	33%	65%
		0.5	101%	99%	53%	36%	66%
55		1	96%	91%	55%	35%	67%

[0093] As can be seen from Table 6, a heat treatment at a higher temperature and for a shorter time results in an increase in the release of the Reb A, Reb D and Reb M from within the yeast cell while affecting a heat kill but with

EP 3 302 097 B1

minimal cell lysis as evaluated by the amount of nitrogen and phosphorus in the medium after the heat treatment. The table also shows that at higher temperatures (50°C, 75°C and 95°C and longer times), Reb DM and RebA can degrade to Reb B resulting in a lower Reb DM content in the broth and an increase in Reb B content.

[0094] Viable yeast counts after heat treatment are shown in Table 7. It takes between 2 and 24 hours at 50°C to kill the yeast while no more than 5 minutes at 75°C or 95°C to kill the yeast.

Table 7

Temperature	Time	YM (CFU/ml)
24°C	24 h	1.02E+09
50°C	1h	2.03E+04
	2h	600
	24h	< 10
	48h	< 10
	72h	< 10
75°C	0.08 h	< 10
	0.5h	< 10
	1h	< 10
	2h	< 10
	24h	< 10
	48h	< 10
	70h	< 10
95°C	0.08h	< 10
	1h	< 10

Example 3

[0095] Fermentation broth from a strain designated A in a two-phase process was heat treated at 75°C for one hour. The two phase rates of feeding are shown below:

Strain	pH control	Phase I	Phase II
B	5.0	0.12u	0.166 mL/min

[0096] Yeast cells were stained by the fluorescent dyes based on the Nexcelom Cellometer method. Live cells before heat treatment appeared green under microscope and dead cells after heat treatment appeared red. Bright field microscopy images showed a lack of debris after heat treatment (compare FIG. 5A with FIG. 5B) indicating that the yeast cells were intact even after heat treatment.

SEQUENCE LISTING

[0097]

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<120> HEAT TREATMENT TO PRODUCE GLYOSIDES

<130> CAR0211/WO

<160> 22

EP 3 302 097 B1

<170> PatentIn version 3.5

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      Gly Glu Thr Val Ser Val Pro Gly Phe Pro Val Leu Gln Arg Trp Glu
      165          170          175

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EP 3 302 097 B1

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			195					200					205				
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					325					330					335		
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70	Asn	Gly	Ile	Val	Arg	Arg	Gly	Asn	Leu	Ala	Ser	Cys	Ile	Lys	Met	Ile	
					405					410					415		
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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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		210					215					220					
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25	Gln	Lys	Gly	Ser	Val	Val	Tyr	Val	Ala	Leu	Gly	Ser	Glu	Ala	Leu	Val	
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EP 3 302 097 B1

Lys Ala Asn Ala Arg Glu Leu Ser Lys Ile Tyr Asn Asp Thr Lys Val
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EP 3 302 097 B1

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50 <213> Synechococcus sp.

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EP 3 302 097 B1

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EP 3 302 097 B1

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	225					230					235					240	
15	Gly	Lys	Asp	Gln	Ala	Ala	Ala	Lys	Ala	Thr	Tyr	Pro	Ser	Leu	Leu	Gly	
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20	Leu	Glu	Ala	Ser	Arg	Gln	Lys	Ala	Glu	Glu	Leu	Ile	Gln	Ser	Ala	Lys	
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25	Glu	Ala	Leu	Arg	Pro	Tyr	Gly	Ser	Gln	Ala	Glu	Pro	Leu	Leu	Ala	Leu	
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EP 3 302 097 B1

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		210					215						220				
	Phe	Pro	Tyr	Asp	His	Gln	Ala	Leu	Gln	Gly	Ile	Tyr	Ser	Ser	Arg	Glu	
	225					230					235					240	
30	Ile	Lys	Met	Lys	Arg	Ile	Pro	Lys	Glu	Val	Met	His	Thr	Val	Pro	Thr	
					245					250					255		
35	Ser	Ile	Leu	His	Ser	Leu	Glu	Gly	Met	Pro	Gly	Leu	Asp	Trp	Ala	Lys	
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					325					330					335		
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EP 3 302 097 B1

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45	Val	Asn	Asn	Asp	Val	Tyr	Leu	Glu	Leu	Ala	Arg	Met	Asp	Phe	Asn	His	
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55	Thr	Glu	Asn	Arg	Leu	Met	Asp	Phe	Gly	Val	Ala	Gln	Glu	Asp	Ala	Leu	
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EP 3 302 097 B1

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10	Glu	Asp	Ile	Ile	His	Lys	Leu	Leu	Arg	Ser	Ala	Trp	Ala	Glu	Trp	Val		660	665	670
15	Arg	Glu	Lys	Ala	Asp	Ala	Ala	Asp	Ser	Val	Cys	Asn	Gly	Ser	Ser	Ala		675	680	685
	Val	Glu	Gln	Glu	Gly	Ser	Arg	Met	Val	His	Asp	Lys	Gln	Thr	Cys	Leu		690	695	700
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25	Ala	Ala	Ser	Glu	Asp	Gly	Asp	Arg	Arg	Ile	Ile	Gln	Leu	Thr	Gly	Ser		725	730	735
30	Ile	Cys	Asp	Ser	Leu	Lys	Gln	Lys	Met	Leu	Val	Ser	Gln	Asp	Pro	Glu		740	745	750
	Lys	Asn	Glu	Glu	Met	Met	Ser	His	Val	Asp	Asp	Glu	Leu	Lys	Leu	Arg		755	760	765
35	Ile	Arg	Glu	Phe	Val	Gln	Tyr	Leu	Leu	Arg	Leu	Gly	Glu	Lys	Lys	Thr		770	775	780
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EP 3 302 097 B1

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EP 3 302 097 B1

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45	Arg Asp Lys Tyr Leu Lys Lys Glu Val Glu Asp Ala Leu Ala Phe Pro 405 410 415		
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	Asn Phe Cys Gln Ser Ile His Arg Glu Glu Met Glu Arg Leu Asp Arg 465 470 475 480		
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	Leu Ala Tyr Cys Tyr Phe Ser Gly Ala Ala Thr Leu Phe Ser Pro Glu 500 505 510		

EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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Arg Asp Val His Arg Ser Leu His Thr Ile Ala Gln Glu Gln Gly Ser
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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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EP 3 302 097 B1

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5	Tyr	Leu	Phe	Thr	Thr	Gln	Leu	Arg	Arg	Lys	Ser	Ala	Asn	Leu	Pro	Pro
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10			35					40					45			
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	Ile	Leu	Gln	Leu	Gln	Leu	Gly	Tyr	Arg	Arg	Val	Leu	Val	Ile	Ser	Ser
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20	Pro	Ser	Ala	Ala	Glu	Glu	Cys	Phe	Thr	Asn	Asn	Asp	Val	Ile	Phe	Ala
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30																
35																
40																
45																
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EP 3 302 097 B1

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15	Arg Val Asp Glu Asn Arg Leu Leu Ile Arg Lys Leu Arg Ser Ser Ser 145 150 155 160		
20	Ser Pro Val Thr Leu Ile Thr Val Phe Tyr Ala Leu Thr Leu Asn Val 165 170 175		
25	Ile Met Arg Met Ile Ser Gly Lys Arg Tyr Phe Asp Ser Gly Asp Arg 180 185 190		
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35	Leu Leu Leu Ala Gly Ala Ser Asn Val Gly Asp Tyr Leu Pro Ile Leu 210 215 220		
40	Asn Trp Leu Gly Val Lys Ser Leu Glu Lys Lys Leu Ile Ala Leu Gln 225 230 235 240		
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	Ile Arg Ser Phe Val Leu Gly Leu Leu Ala Ala Gly Ser Asp Thr Ser 290 295 300		
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	Val Leu Lys Lys Ala Gln Ala Glu Ile Asp Arg Val Ile Gly Asn Asn 325 330 335		
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EP 3 302 097 B1

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10	Arg	Gly	Thr	Met	Leu	Ile	Val	Asn	Gln	Trp	Ala	Ile	His	His	Asp	Pro
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15	Lys	Val	Trp	Asp	Asp	Pro	Glu	Thr	Phe	Lys	Pro	Glu	Arg	Phe	Gln	Gly
				405						410					415	
20	Leu	Glu	Gly	Thr	Arg	Asp	Gly	Phe	Lys	Leu	Met	Pro	Phe	Gly	Ser	Gly
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25	Arg	Arg	Gly	Cys	Pro	Gly	Glu	Gly	Leu	Ala	Ile	Arg	Leu	Leu	Gly	Met
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40	Val	Pro	Leu	Val	Ala	Lys	Cys	Lys	Pro	Arg	Ser	Glu	Met	Thr	Asn	Leu
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<213> Stevia rebaudiana

<400> 16

EP 3 302 097 B1

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EP 3 302 097 B1

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<400> 17

EP 3 302 097 B1

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20	Val	Val	Leu	Val	Trp	Arg	Arg	Ser	Ser	Thr	Lys	Lys	Ser	Ala	Leu	Glu	
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25	Pro	Pro	Val	Ile	Val	Val	Pro	Lys	Arg	Val	Gln	Glu	Glu	Glu	Val	Asp	
					85					90					95		
30	Asp	Gly	Lys	Lys	Lys	Val	Thr	Val	Phe	Phe	Gly	Thr	Gln	Thr	Gly	Thr	
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40	Glu	Lys	Ala	Val	Phe	Lys	Val	Ile	Asp	Leu	Asp	Asp	Tyr	Ala	Ala	Asp	
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50	Phe	Leu	Ala	Thr	Tyr	Gly	Asp	Gly	Glu	Pro	Thr	Asp	Asn	Ala	Ala	Arg	
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55	Phe	Tyr	Lys	Trp	Phe	Thr	Glu	Gly	Asp	Ala	Lys	Gly	Glu	Trp	Leu	Asn	
				180				185						190			
60	Lys	Leu	Gln	Tyr	Gly	Val	Phe	Gly	Leu	Gly	Asn	Arg	Gln	Tyr	Glu	His	
			195					200					205				
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70	Ala	Lys	Arg	Leu	Val	Pro	Val	Gly	Leu	Gly	Asp	Asp	Asp	Gln	Cys	Ile	
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75	Glu	Asp	Asp	Phe	Thr	Ala	Trp	Lys	Glu	Leu	Val	Trp	Pro	Glu	Leu	Asp	
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EP 3 302 097 B1

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10	Leu	Ser	Glu	Asp	Tyr	Ser	Tyr	Thr	Asn	Gly	His	Ala	Val	His	Asp	Ala	
		290					295					300					
15	Gln	His	Pro	Cys	Arg	Ser	Asn	Val	Ala	Val	Lys	Lys	Glu	Leu	His	Ser	
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40	Leu	Gly	Gly	Ala	Ser	Leu	Pro	Pro	Pro	Phe	Pro	Pro	Cys	Thr	Leu	Arg	
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					405					410					415		
50	Ala	Leu	Leu	Ala	Leu	Ala	Ala	His	Ala	Thr	Asp	Pro	Ser	Glu	Ala	Asp	
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60	Trp	Ile	Val	Ala	Ser	Gln	Arg	Ser	Leu	Leu	Glu	Val	Met	Glu	Ala	Phe	
	450						455					460					
65	Pro	Ser	Ala	Lys	Pro	Ser	Leu	Gly	Val	Phe	Phe	Ala	Ser	Val	Ala	Pro	
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70	Arg	Leu	Gln	Pro	Arg	Tyr	Tyr	Ser	Ile	Ser	Ser	Ser	Pro	Lys	Met	Ala	
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75	Pro	Asp	Arg	Ile	His	Val	Thr	Cys	Ala	Leu	Val	Tyr	Glu	Lys	Thr	Pro	
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EP 3 302 097 B1

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10	Arg	Thr	Ser	Asn	Phe	Arg	Leu	Pro	Ser	Asp	Pro	Lys	Val	Pro	Val	Ile	
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					565					570					575		
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25	Leu	Phe	Phe	Gly	Cys	Arg	Asn	Arg	Lys	Val	Asp	Phe	Ile	Tyr	Glu	Asn	
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30	Glu	Leu	Asn	Asn	Phe	Val	Glu	Thr	Gly	Ala	Leu	Ser	Glu	Leu	Ile	Val	
		610					615					620					
35	Ala	Phe	Ser	Arg	Glu	Gly	Pro	Thr	Lys	Glu	Tyr	Val	Gln	His	Lys	Met	
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40	Ser	Glu	Lys	Ala	Ser	Asp	Ile	Trp	Asn	Leu	Leu	Ser	Glu	Gly	Ala	Tyr	
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EP 3 302 097 B1

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EP 3 302 097 B1

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Asp Pro Thr Thr Leu Pro Ala Leu Lys Met Leu Val Glu Asn Arg Glu
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Leu Leu Thr Leu Phe Thr Thr Ser Phe Ala Val Leu Ile Gly Cys Leu
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Val Phe Leu Met Trp Arg Arg Ser Ser Ser Lys Lys Leu Val Gln Asp
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35

Pro Val Pro Gln Val Ile Val Val Lys Lys Lys Glu Lys Glu Ser Glu
 85 90 95

40

Val Asp Asp Gly Lys Lys Lys Val Ser Ile Phe Tyr Gly Thr Gln Thr
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Gly Thr Ala Glu Gly Phe Ala Lys Ala Leu Val Glu Glu Ala Lys Val
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Arg Tyr Glu Lys Thr Ser Phe Lys Val Ile Asp Leu Asp Asp Tyr Ala
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Ala Asp Asp Asp Glu Tyr Glu Glu Lys Leu Lys Lys Glu Ser Leu Ala
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Ala Asn Phe Tyr Lys Trp Phe Thr Glu Gly Asp Asp Lys Gly Glu Trp
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EP 3 302 097 B1

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20	Leu	Asp	Gln	Leu	Leu	Arg	Asp	Glu	Asp	Asp	Thr	Ser	Val	Thr	Thr	Pro	
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25	Tyr	Thr	Ala	Ala	Val	Leu	Glu	Tyr	Arg	Val	Val	Tyr	His	Asp	Lys	Pro	
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30	Ala	Asp	Ser	Tyr	Ala	Glu	Asp	Gln	Thr	His	Thr	Asn	Gly	His	Val	Val	
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35	His	Asp	Ala	Gln	His	Pro	Ser	Arg	Ser	Asn	Val	Ala	Phe	Lys	Lys	Glu	
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40	Leu	His	Thr	Ser	Gln	Ser	Asp	Arg	Ser	Cys	Thr	His	Leu	Glu	Phe	Asp	
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50	Tyr	Ser	Glu	Asn	Leu	Ser	Glu	Val	Val	Asp	Glu	Ala	Leu	Lys	Leu	Leu	
			355					360					365				
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60	Gly	Thr	Pro	Ile	Gly	Gly	Ala	Ser	Leu	Pro	Pro	Pro	Phe	Pro	Pro	Cys	
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65	Thr	Leu	Arg	Asp	Ala	Leu	Thr	Arg	Tyr	Ala	Asp	Val	Leu	Ser	Ser	Pro	
					405					410					415		
70	Lys	Lys	Val	Ala	Leu	Leu	Ala	Leu	Ala	Ala	His	Ala	Ser	Asp	Pro	Ser	
				420					425					430			
75	Glu	Ala	Asp	Arg	Leu	Lys	Phe	Leu	Ala	Ser	Pro	Ala	Gly	Lys	Asp	Glu	

EP 3 302 097 B1

	435	440	445
5	Tyr Ala Gln Trp Ile Val 450	Ala Asn Gln Arg Ser 455	Leu Leu Glu Val Met 460
10	Gln Ser Phe Pro Ser 465	Ala Lys Pro Pro 470	Leu Gly Val Phe Phe Ala Ala 475 480
15	Val Ala Pro Arg 485	Leu Gln Pro Arg Tyr Tyr 490	Ser Ile Ser Ser Ser Pro 495
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25	Thr Thr Pro Ala Gly Arg Ile His Arg Gly Leu Cys Ser Thr Trp Met 515 520 525		
30	Lys Asn Ala Val Pro Leu Thr Glu Ser Pro Asp Cys Ser Gln Ala Ser 530 535 540		
35	Ile Phe Val Arg Thr Ser Asn Phe Arg Leu Pro Val Asp Pro Lys Val 545 550 555 560		
40	Pro Val Ile Met Ile Gly Pro Gly Thr Gly Leu Ala Pro Phe Arg Gly 565 570 575		
45	Phe Leu Gln Glu Arg Leu Ala Leu Lys Glu Ser Gly Thr Glu Leu Gly 580 585 590		
50	Ser Ser Ile Phe Phe Phe Gly Cys Arg Asn Arg Lys Val Asp Phe Ile 595 600 605		
55	Tyr Glu Asp Glu Leu Asn Asn Phe Val Glu Thr Gly Ala Leu Ser Glu 610 615 620		
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65	His Lys Met Ser Gln Lys Ala Ser Asp Ile Trp Lys Leu Leu Ser Glu 645 650 655		
70	Gly Ala Tyr Leu Tyr Val Cys Gly Asp Ala Lys Gly Met Ala Lys Asp 660 665 670		
75	Val His Arg Thr Leu His Thr Ile Val Gln Glu Gln Gly Ser Leu Asp 675 680 685		

EP 3 302 097 B1

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5 Tyr Leu Arg Asp Val Trp
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<211> 511

10 <212> PRT

<213> Rubus suavissimus

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1 5 10 15

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Ser Phe Phe Ser Asn Lys Ser Ala Gln Ala Lys Leu Pro Pro Val Pro
35 40 45

25 Val Val Pro Gly Leu Pro Val Ile Gly Asn Leu Leu Gln Leu Lys Glu
50 55 60

30 Lys Lys Pro Tyr Gln Thr Phe Thr Arg Trp Ala Glu Glu Tyr Gly Pro
65 70 75 80

Ile Tyr Ser Ile Arg Thr Gly Ala Ser Thr Met Val Val Leu Asn Thr
35 85 90 95

Thr Gln Val Ala Lys Glu Ala Met Val Thr Arg Tyr Leu Ser Ile Ser
100 105 110

40 Thr Arg Lys Leu Ser Asn Ala Leu Lys Ile Leu Thr Ala Asp Lys Cys
115 120 125

45 Met Val Ala Ile Ser Asp Tyr Asn Asp Phe His Lys Met Ile Lys Arg
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50 Tyr Ile Leu Ser Asn Val Leu Gly Pro Ser Ala Gln Lys Arg His Arg
145 150 155 160

Ser Asn Arg Asp Thr Leu Arg Ala Asn Val Cys Ser Arg Leu His Ser
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55 Gln Val Lys Asn Ser Pro Arg Glu Ala Val Asn Phe Arg Arg Val Phe
180 185 190

EP 3 302 097 B1

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			195					200					205				
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		210					215					220					
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10	225					230					235					240	
	Glu	Val	Asp	Trp	Arg	Asp	Phe	Phe	Pro	Tyr	Leu	Arg	Trp	Ile	Pro	Asn	
					245					250					255		
15	Thr	Arg	Met	Glu	Thr	Lys	Ile	Gln	Arg	Leu	Tyr	Phe	Arg	Arg	Lys	Ala	
				260					265					270			
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25		290					295					300					
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					325					330					335		
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35				340					345					350			
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			355					360					365				
40	Tyr	Leu	Asn	Ala	Val	Phe	His	Glu	Thr	Leu	Arg	Lys	His	Ser	Pro	Ala	
		370					375					380					
	Ala	Leu	Val	Pro	Leu	Arg	Tyr	Ala	His	Glu	Asp	Thr	Gln	Leu	Gly	Gly	
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				420					425					430			
55	Arg	Phe	Leu	Asp	Pro	Lys	Phe	Asp	Pro	Met	Asp	Leu	Tyr	Lys	Thr	Met	
			435					440					445				

EP 3 302 097 B1

	Ala	Phe	Gly	Ala	Gly	Lys	Arg	Val	Cys	Ala	Gly	Ser	Leu	Gln	Ala	Met	
	450						455					460					
5	Leu	Ile	Ala	Cys	Pro	Thr	Ile	Gly	Arg	Leu	Val	Gln	Glu	Phe	Glu	Trp	
	465					470					475					480	
10	Lys	Leu	Arg	Asp	Gly	Glu	Glu	Glu	Asn	Val	Asp	Thr	Val	Gly	Leu	Thr	
					485					490					495		
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30	Ser	Ile	Val	Val	Arg	Trp	Ala	Trp	Ser	Val	Val	Asn	Trp	Val	Trp	Phe	
				20					25					30			
35	Lys	Pro	Lys	Lys	Leu	Glu	Arg	Phe	Leu	Arg	Glu	Gln	Gly	Leu	Lys	Gly	
			35					40					45				
40	Asn	Ser	Tyr	Arg	Phe	Leu	Tyr	Gly	Asp	Met	Lys	Glu	Asn	Ser	Ile	Leu	
		50					55					60					
45	Leu	Lys	Gln	Ala	Arg	Ser	Lys	Pro	Met	Asn	Leu	Ser	Thr	Ser	His	Asp	
	65					70					75					80	
50	Ile	Ala	Pro	Gln	Val	Thr	Pro	Phe	Val	Asp	Gln	Thr	Val	Lys	Ala	Tyr	
					85					90					95		
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				100					105					110			
60	Met	Asn	Pro	Glu	Asp	Leu	Lys	Asp	Val	Leu	Thr	Lys	Asn	Val	Asp	Phe	
			115					120					125				
65	Val	Lys	Pro	Ile	Ser	Asn	Pro	Leu	Ile	Lys	Leu	Leu	Ala	Thr	Gly	Ile	
		130					135					140					
70	Ala	Ile	Tyr	Glu	Gly	Glu	Lys	Trp	Thr	Lys	His	Arg	Arg	Ile	Ile	Asn	
	145					150					155					160	

EP 3 302 097 B1

	Pro	Thr	Phe	His	Ser	Glu	Arg	Leu	Lys	Arg	Met	Leu	Pro	Ser	Phe	His	
					165					170					175		
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				180					185					190			
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			195					200					205				
15	Ser	Ala	Asp	Val	Ile	Ser	Arg	Thr	Ala	Phe	Gly	Thr	Ser	Tyr	Lys	Lys	
		210					215					220					
20	Gly	Gln	Lys	Ile	Phe	Glu	Leu	Leu	Arg	Glu	Gln	Val	Ile	Tyr	Val	Thr	
	225					230					235					240	
25	Lys	Gly	Phe	Gln	Ser	Phe	Tyr	Ile	Pro	Gly	Trp	Arg	Phe	Leu	Pro	Thr	
					245					250					255		
30	Lys	Met	Asn	Lys	Arg	Met	Asn	Glu	Ile	Asn	Glu	Glu	Ile	Lys	Gly	Leu	
				260					265					270			
35	Ile	Arg	Gly	Ile	Ile	Ile	Asp	Arg	Glu	Gln	Ile	Ile	Lys	Ala	Gly	Glu	
			275					280					285				
40	Glu	Thr	Asn	Asp	Asp	Leu	Leu	Gly	Ala	Leu	Met	Glu	Ser	Asn	Leu	Lys	
		290					295					300					
45	Asp	Ile	Arg	Glu	His	Gly	Lys	Asn	Asn	Lys	Asn	Val	Gly	Met	Ser	Ile	
	305					310					315					320	
50	Glu	Asp	Val	Ile	Gln	Glu	Cys	Lys	Leu	Phe	Tyr	Phe	Ala	Gly	Gln	Glu	
					325					330					335		
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				340					345					350			
60	Gln	Asn	Trp	Gln	Asp	Arg	Ala	Arg	Gln	Glu	Val	Leu	Gln	Val	Phe	Gly	
			355					360					365				
65	Ser	Ser	Lys	Pro	Asp	Phe	Asp	Gly	Leu	Ala	His	Leu	Lys	Val	Val	Thr	
		370					375					380					
70	Met	Ile	Leu	Leu	Glu	Val	Leu	Arg	Leu	Tyr	Pro	Pro	Val	Ile	Glu	Leu	
	385					390					395					400	
75	Ile	Arg	Thr	Ile	His	Lys	Lys	Thr	Gln	Leu	Gly	Lys	Leu	Ser	Leu	Pro	
				405						410					415		

EP 3 302 097 B1

	Glu	Gly	Val	Glu	Val	Arg	Leu	Pro	Thr	Leu	Leu	Ile	His	His	Asp	Lys
				420					425					430		
5	Glu	Leu	Trp	Gly	Asp	Asp	Ala	Asn	Gln	Phe	Asn	Pro	Glu	Arg	Phe	Ser
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		450					455					460				
15	Gly	Ala	Gly	Pro	Arg	Ile	Cys	Ile	Gly	Gln	Asn	Phe	Ser	Met	Met	Glu
	465					470					475					480
20	Ala	Lys	Leu	Ala	Leu	Ala	Leu	Ile	Leu	Gln	His	Phe	Thr	Phe	Glu	Leu
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25	Ser	Pro	Ser	His	Ala	His	Ala	Pro	Ser	His	Arg	Ile	Thr	Leu	Gln	Pro
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45	Val	Ile	Phe	Glu	Asn	Arg	Glu	Leu	Val	Ala	Ile	Leu	Thr	Thr	Ser	Ile
			35					40					45			
50	Ala	Val	Met	Ile	Gly	Cys	Phe	Val	Val	Leu	Met	Trp	Arg	Arg	Ala	Gly
		50					55					60				
55	Ser	Arg	Lys	Val	Lys	Asn	Val	Glu	Leu	Pro	Lys	Pro	Leu	Ile	Val	His
	65					70					75					80
	Glu	Pro	Glu	Pro	Glu	Val	Glu	Asp	Gly	Lys	Lys	Lys	Val	Ser	Ile	Phe
					85					90					95	
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				100					105					110		

EP 3 302 097 B1

	Asp	Glu	Ala	Lys	Ala	Arg	Tyr	Glu	Lys	Ala	Thr	Phe	Arg	Val	Val	Asp	
			115					120					125				
5	Leu	Asp	Asp	Tyr	Ala	Ala	Asp	Asp	Asp	Gln	Tyr	Glu	Glu	Lys	Leu	Lys	
		130					135					140					
10	Asn	Glu	Ser	Phe	Ala	Val	Phe	Leu	Leu	Ala	Thr	Tyr	Gly	Asp	Gly	Glu	
	145					150					155					160	
15	Pro	Thr	Asp	Asn	Ala	Ala	Arg	Phe	Tyr	Lys	Trp	Phe	Ala	Glu	Gly	Lys	
					165					170					175		
20	Glu	Arg	Gly	Glu	Trp	Leu	Gln	Asn	Leu	His	Tyr	Ala	Val	Phe	Gly	Leu	
				180					185					190			
25	Gly	Asn	Arg	Gln	Tyr	Glu	His	Phe	Asn	Lys	Ile	Ala	Lys	Val	Ala	Asp	
			195					200					205				
30	Glu	Leu	Leu	Glu	Ala	Gln	Gly	Gly	Asn	Arg	Leu	Val	Lys	Val	Gly	Leu	
	210						215					220					
35	Gly	Asp	Asp	Asp	Gln	Cys	Ile	Glu	Asp	Asp	Phe	Ser	Ala	Trp	Arg	Glu	
	225					230					235					240	
40	Ser	Leu	Trp	Pro	Glu	Leu	Asp	Met	Leu	Leu	Arg	Asp	Glu	Asp	Asp	Ala	
					245					250					255		
45	Thr	Thr	Val	Thr	Thr	Pro	Tyr	Thr	Ala	Ala	Val	Leu	Glu	Tyr	Arg	Val	
				260					265					270			
50	Val	Phe	His	Asp	Ser	Ala	Asp	Val	Ala	Ala	Glu	Asp	Lys	Ser	Trp	Ile	
			275					280					285				
55	Asn	Ala	Asn	Gly	His	Ala	Val	His	Asp	Ala	Gln	His	Pro	Phe	Arg	Ser	
	290						295					300					
60	Asn	Val	Val	Val	Arg	Lys	Glu	Leu	His	Thr	Ser	Ala	Ser	Asp	Arg	Ser	
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65	Cys	Ser	His	Leu	Glu	Phe	Asn	Ile	Ser	Gly	Ser	Ala	Leu	Asn	Tyr	Glu	
				325						330					335		
70	Thr	Gly	Asp	His	Val	Gly	Val	Tyr	Cys	Glu	Asn	Leu	Thr	Glu	Thr	Val	
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EP 3 302 097 B1

	355	360	365
5	Ile Tyr Thr Asp Asn Glu Asp Gly Thr Pro Leu Gly Gly Ser Ser Leu 370 375 380		
10	Pro Pro Pro Phe Pro Ser Cys Thr Leu Arg Thr Ala Leu Thr Arg Tyr 385 390 395 400		
15	Ala Asp Leu Leu Asn Ser Pro Lys Lys Ser Ala Leu Leu Ala Leu Ala 405 410 415		
20	Ala His Ala Ser Asn Pro Val Glu Ala Asp Arg Leu Arg Tyr Leu Ala 420 425 430		
25	Ser Pro Ala Gly Lys Asp Glu Tyr Ala Gln Ser Val Ile Gly Ser Gln 435 440 445		
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40	Tyr Ser Ile Ser Ser Ser Pro Arg Met Ala Pro Ser Arg Ile His Val 485 490 495		
45	Thr Cys Ala Leu Val Tyr Asp Lys Met Pro Thr Gly Arg Ile His Lys 500 505 510		
50	Gly Val Cys Ser Thr Trp Met Lys Asn Ser Val Pro Met Glu Lys Ser 515 520 525		
55	His Glu Cys Ser Trp Ala Pro Ile Phe Val Arg Gln Ser Asn Phe Lys 530 535 540		
	Leu Pro Ala Glu Ser Lys Val Pro Ile Ile Met Val Gly Pro Gly Thr 545 550 555 560		
	Gly Leu Ala Pro Phe Arg Gly Phe Leu Gln Glu Arg Leu Ala Leu Lys 565 570 575		
	Glu Ser Gly Val Glu Leu Gly Pro Ser Ile Leu Phe Phe Gly Cys Arg 580 585 590		
	Asn Arg Arg Met Asp Tyr Ile Tyr Glu Asp Glu Leu Asn Asn Phe Val 595 600 605		

Glu Thr Gly Ala Leu Ser Glu Leu Val Ile Ala Phe Ser Arg Glu Gly
610 615 620

5 Pro Thr Lys Glu Tyr Val Gln His Lys Met Ala Glu Lys Ala Ser Asp
625 630 635 640

10 Ile Trp Asn Leu Ile Ser Glu Gly Ala Tyr Leu Tyr Val Cys Gly Asp
645 650 655

15 Ala Lys Gly Met Ala Lys Asp Val His Arg Thr Leu His Thr Ile Met
660 665 670

Gln Glu Gln Gly Ser Leu Asp Ser Ser Lys Ala Glu Ser Met Val Lys
675 680 685

20 Asn Leu Gln Met Asn Gly Arg Tyr Leu Arg Asp Val Trp
690 695 700

Claims

1. A method of releasing steviol glycosides from a yeast host cell, comprising the steps of:

a) providing a fermentation medium comprising a yeast host cell engineered to produce one or more steviol glycosides; and

b) releasing the one or more steviol glycosides from the engineered yeast host cell by heating the fermentation medium to a temperature in the range from 70° C to 75° C for 5 minutes to 48 hours, wherein the one or more steviol glycosides are released from an intracellular portion of the yeast host cell into an extracellular portion of the fermentation medium;

wherein the steviol glycoside is selected from rebaudioside M, rebaudioside D, rebaudioside A, rebaudioside B or combinations thereof.

2. The method of claim 1, wherein the heating is 75°C for 5 minutes to 2 hours.

3. The method of claim 1 or 2, wherein the heating does not increase extracellular nitrogen content to more than 45% of the total nitrogen components of the fermentation medium.

4. The method of any one of claims 1-3, wherein the heating is for 5 minutes to 2 hours.

5. The method of any one of the preceding claims, wherein the engineered yeast host cell is selected from the group consisting of genus, *Candida*, *Pichia* (*Hansenula*), *Kloeckera* (*Hanseniaspora*), *Kluyveromyces*, *Rhodotorula*, *Torulopsis*, *Zygosaccharomyces*, *Saccharomycete*, *Yarrowia*, and *Saccharomyces*.

6. The method of any one of the preceding claims, wherein the rebaudioside M, rebaudioside D, rebaudioside A, rebaudioside B or combinations thereof released are greater than 90% of the rebaudioside M, rebaudioside D, rebaudioside A, rebaudioside B or combinations compared to the total glycosides produced and found both intracellularly and extracellularly.

7. The method of any one of the preceding claims, wherein the rebaudioside D and/or rebaudioside M released extracellularly is greater than 33 % of the total of both intracellular and extracellular rebaudioside D and/or rebaudioside M present in the fermentation medium.

8. The method of any one of the preceding claims, wherein rebaudioside D released, is present extracellularly in the range of 30% to 100% of the total amount of rebaudioside D present in the fermentation medium.

9. The method of any one of the preceding claims, wherein the rebaudioside M released, is present extracellularly in the range of 30% to 100%, of the total amount of rebaudioside M present in the fermentation medium.
10. The method of any one of the preceding claims, wherein the rebaudioside A amount, is released extracellularly in the range of 30% to 100 %, 50 % to 100 %, 80 % to 90 % of the total amount of rebaudioside A produced during the fermentation.
11. The method of any one of the preceding claims, wherein the rebaudioside B amount, is released extracellularly in the range of 25% to 100%, 30% to 95%, 40% to 80% of the total amount of rebaudioside B produced during the fermentation.

Patentansprüche

1. Verfahren zum Freisetzen von Steviolglycosiden aus einer Hefewirtszelle, das folgende Schritte umfasst:
 - a) Bereitstellen eines Fermentationsmediums, das eine Hefewirtszelle umfasst, die verändert wird, um ein oder mehrere Steviolglycoside zu erzeugen; und
 - b) Freisetzen des einen oder der mehreren Steviolglycoside aus der veränderten Hefewirtszelle durch Erwärmen des Fermentationsmediums auf eine Temperatur in dem Bereich von 70 °C bis 75 °C für 5 Minuten bis 48 Stunden lang, wobei das eine oder die mehreren Steviolglycoside aus einem intrazellulären Abschnitt der Hefewirtszelle in einem extrazellulären Abschnitt des Fermentationsmediums freigesetzt werden; wobei das Steviolglycosid aus Rebaudiosid M, Rebaudiosid D, Rebaudiosid A, Rebaudiosid B oder Kombinationen davon ausgewählt ist.
2. Verfahren nach Anspruch 1, wobei das Erwärmen 75 °C für 5 Minuten bis 2 Stunden lang beträgt.
3. Verfahren nach Anspruch 1 oder 2, wobei das Erwärmen ein Gehalt an extrazellulärem Stickstoff nicht auf mehr als 45 % der Gesamtsumme an Stickstoffkomponenten des Fermentationsmediums erhöht.
4. Verfahren nach einem der Ansprüche 1-3, wobei das Erwärmen für 5 Minuten bis 2 Stunden lang beträgt.
5. Verfahren nach einem der vorhergehenden Ansprüche, wobei die veränderte Hefewirtszelle aus der Gruppe ausgewählt ist, die aus der Gattung *Candida*, *Pichia* (*Hansenula*), *Kloeckera* (*Hanseniaspora*), *Kluyveromyces*, *Rhodotorula*, *Torulopsis*, *Zygosaccharomyces*, *Saccharomycete*, *Yarrowia* und *Saccharomyces* besteht.
6. Verfahren nach einem der vorhergehenden Ansprüche, wobei das freigesetzte Rebaudiosid M, Rebaudiosid D, Rebaudiosid A, Rebaudiosid B oder Kombinationen davon über 90 % des Rebaudiosids M, Rebaudiosids D, Rebaudiosids A, Rebaudiosids B oder Kombinationen im Vergleich zu der Gesamtsumme an Glycosiden, die sowohl intrazellulär als auch extrazellulär erzeugt und gefunden werden, beträgt.
7. Verfahren nach einem der vorhergehenden Ansprüche, wobei das extrazellulär freigesetzte Rebaudiosid D und/oder Rebaudiosid M über 33 % der Gesamtsumme an sowohl intrazellulärem als auch extrazellulärem Rebaudiosid D und/oder Rebaudiosid M, die in dem Fermentationsmedium vorhanden sind, beträgt.
8. Verfahren nach einem der vorhergehenden Ansprüche, wobei freigesetztes Rebaudiosid D in dem Bereich von 30 % bis 100 % der Gesamtmenge an Rebaudiosid D, das in dem Fermentationsmedium vorhanden ist, extrazellulär vorhanden ist.
9. Verfahren nach einem der vorhergehenden Ansprüche, wobei das freigesetzte Rebaudiosid M in dem Bereich von 30 % bis 100 % der Gesamtmenge an Rebaudiosid M, das in dem Fermentationsmedium vorhanden ist, extrazellulär vorhanden ist.
10. Verfahren nach einem der vorhergehenden Ansprüche, wobei die Menge an Rebaudiosid A in dem Bereich von 30 % bis 100 %, 50 % bis 100 %, 80 % bis 90 % der Gesamtmenge an Rebaudiosid A, die während der Fermentation erzeugt wird, extrazellulär freigesetzt wird.
11. Verfahren nach einem der vorhergehenden Ansprüche, wobei die Menge an Rebaudiosid B in dem Bereich von 25

% bis 100 %, 30 % bis 95 %, 40 % bis 80 % der Gesamtmenge an Rebaudiosid B, die während der Fermentation erzeugt wird, extrazellulär freigesetzt wird.

5 Revendications

1. Procédé de libération de glycosides de stéviol à partir d'une cellule hôte de levure, comprenant les étapes suivantes

a) la fourniture d'un milieu de fermentation comprenant une cellule hôte de levure modifiée pour produire un ou plusieurs glycosides de stéviol ; et

b) la libération du ou des glycosides de stéviol à partir de la cellule hôte de levure modifiée en chauffant le milieu de fermentation à une température comprise entre 70 °C et 75 °C pendant 5 minutes à 48 heures, le ou les glycosides de stéviol étant libérés d'une partie intracellulaire de la cellule hôte de levure dans une partie extracellulaire du milieu de fermentation ;

le glycoside de stéviol étant choisi parmi le rébaudioside M, le rébaudioside D, le rébaudioside A, le rébaudioside B ou leurs combinaisons.

2. Procédé selon la revendication 1, le chauffage étant à 75 °C pendant 5 minutes à 2 heures.

3. Procédé selon la revendication 1 ou 2, le chauffage n'augmentant pas la teneur en azote extracellulaire à plus de 45 % des composants d'azote totaux du milieu de fermentation.

4. Procédé selon l'une quelconque des revendications 1 à 3, le chauffage étant de 5 minutes à 2 heures.

5. Procédé selon l'une quelconque des revendications précédentes, la cellule hôte de levure modifiée étant choisie dans le groupe constitué par le genre, *Candida*, *Pichia* (*Hansenula*), *Kloeckera* (*Hanseniaspora*), *Kluyveromyces*, *Rhodotorula*, *Torulopsis*, *Zygosaccharomyces*, *Saccharomycete*, *Yarrowia* et *Saccharomyces*.

6. Procédé selon l'une quelconque des revendications précédentes, le rébaudioside M, le rébaudioside D, le rébaudioside A, le rébaudioside B ou leurs combinaisons libérés étant supérieurs à 90 % du rébaudioside M, du rébaudioside D, du rébaudioside A, du rébaudioside B ou des combinaisons par rapport aux glycosides totaux produits et trouvés à la fois au niveau intracellulaire et extracellulaire.

7. Procédé selon l'une quelconque des revendications précédentes, le rébaudioside D et/ou le rébaudioside M libéré de manière extracellulaire étant supérieur à 33 % du total à la fois du rébaudioside D et/ou du rébaudioside M intracellulaire et extracellulaire présents dans le milieu de fermentation.

8. Procédé selon l'une quelconque des revendications précédentes, le rébaudioside D libéré étant présent de manière extracellulaire dans la plage de 30 % à 100 % de la quantité totale de rébaudioside D présente dans le milieu de fermentation.

9. Procédé selon l'une quelconque des revendications précédentes, le rébaudioside M libéré étant présent de manière extracellulaire dans la plage de 30 % à 100 % de la quantité totale de rébaudioside M présente dans le milieu de fermentation.

10. Procédé selon l'une quelconque des revendications précédentes, la quantité de rébaudioside A étant libérée de manière extracellulaire dans la plage de 30 % à 100 %, de 50 % à 100 %, de 80 % à 90 % de la quantité totale de rébaudioside A produite pendant la fermentation.

11. Procédé selon l'une quelconque des revendications précédentes, la quantité de rébaudioside B étant libérée de manière extracellulaire dans la plage de 25 % à 100 %, de 30 % à 95 %, de 40 % à 80 % de la quantité totale de rébaudioside B produite pendant la fermentation.

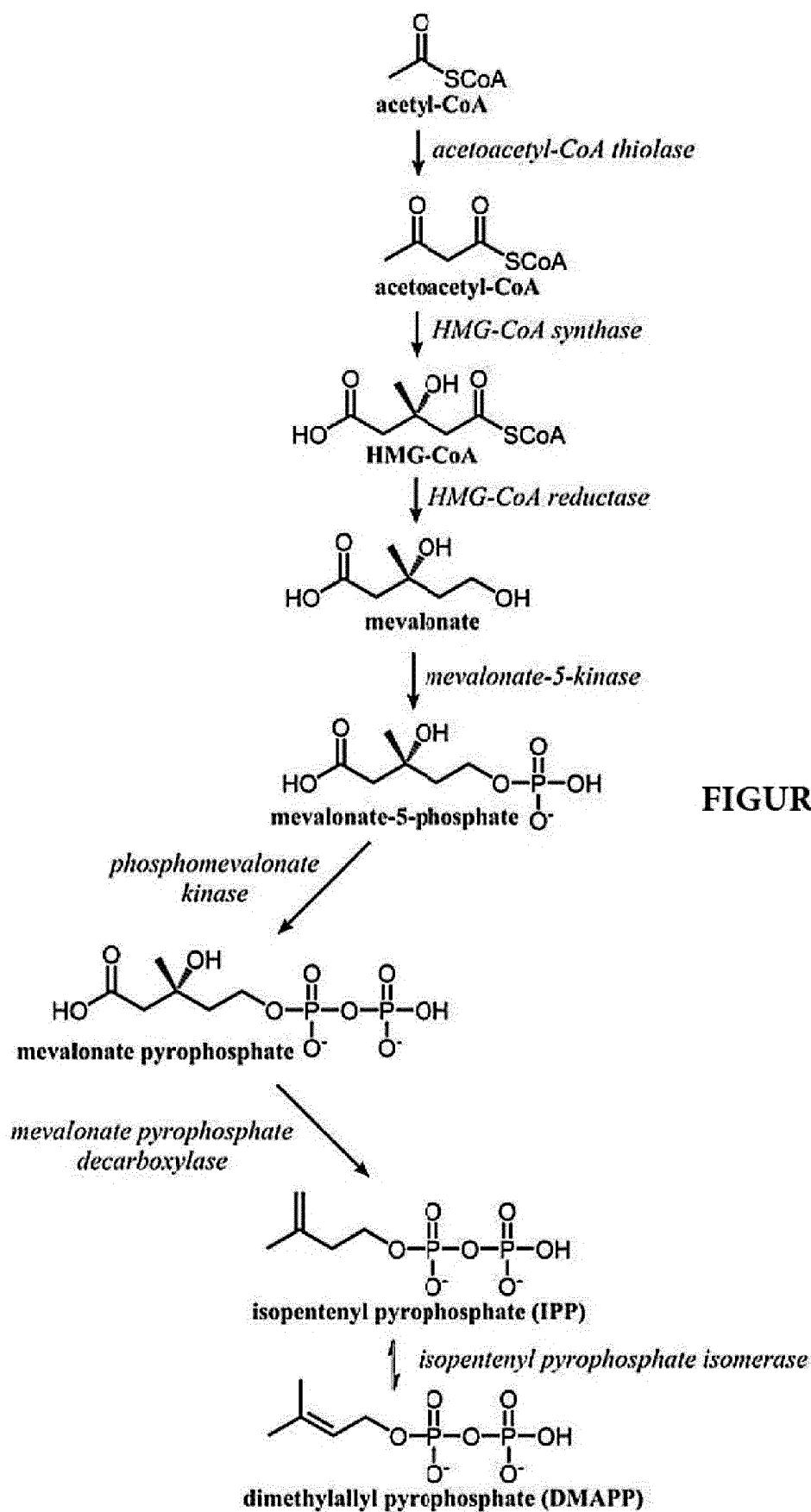


FIGURE 1

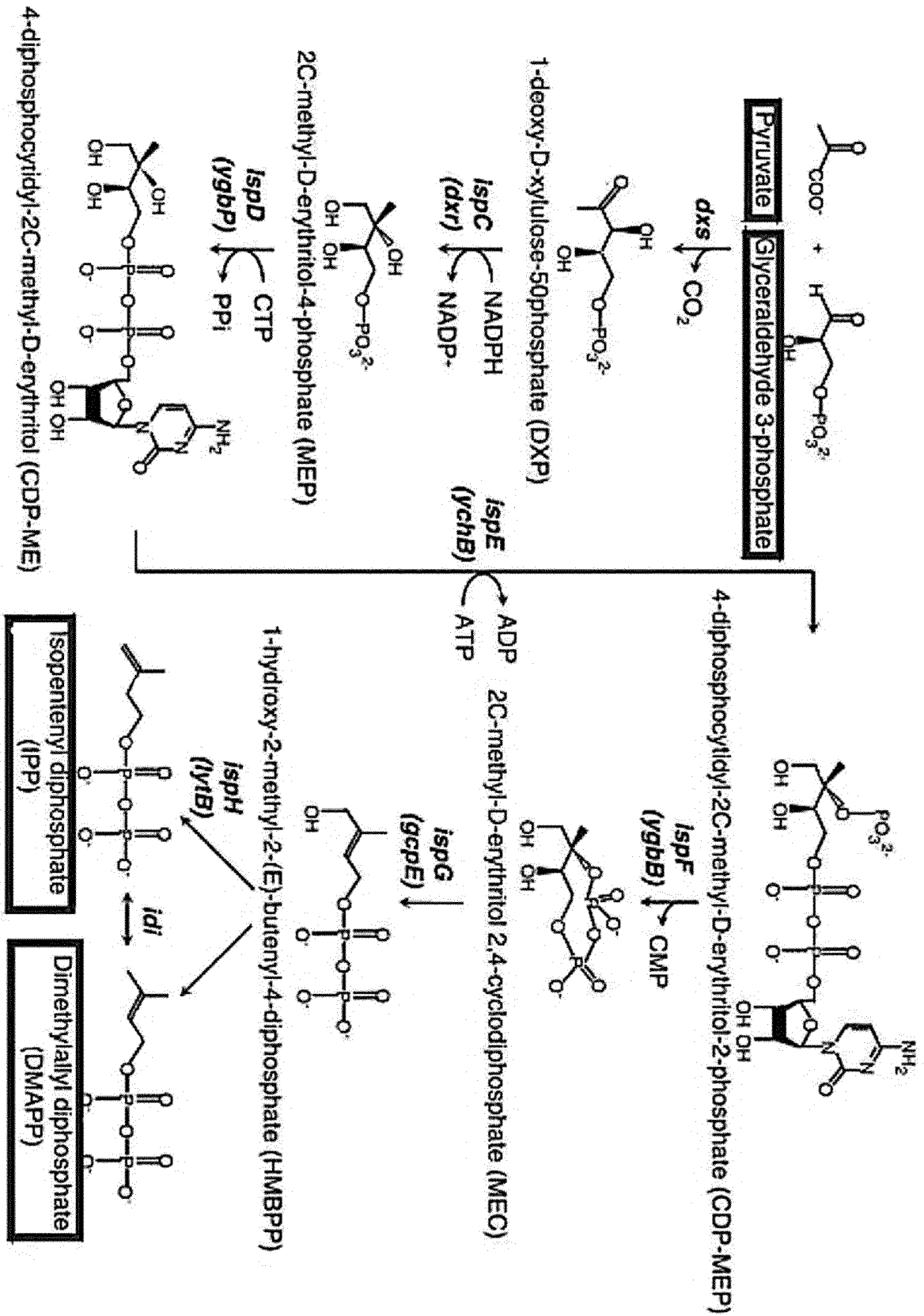


FIGURE 2

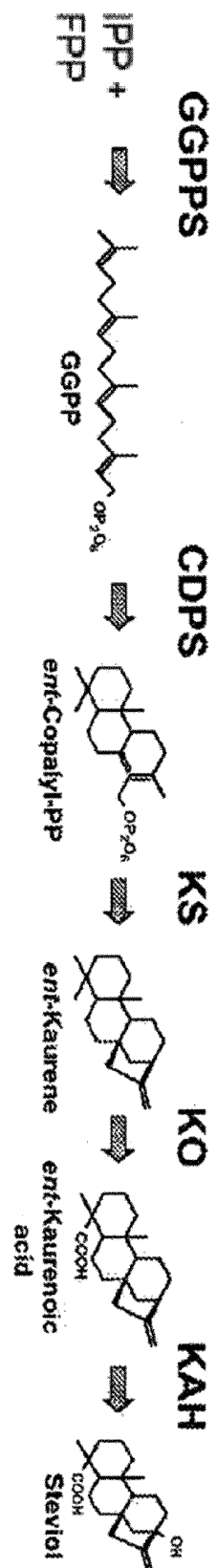


FIGURE 3

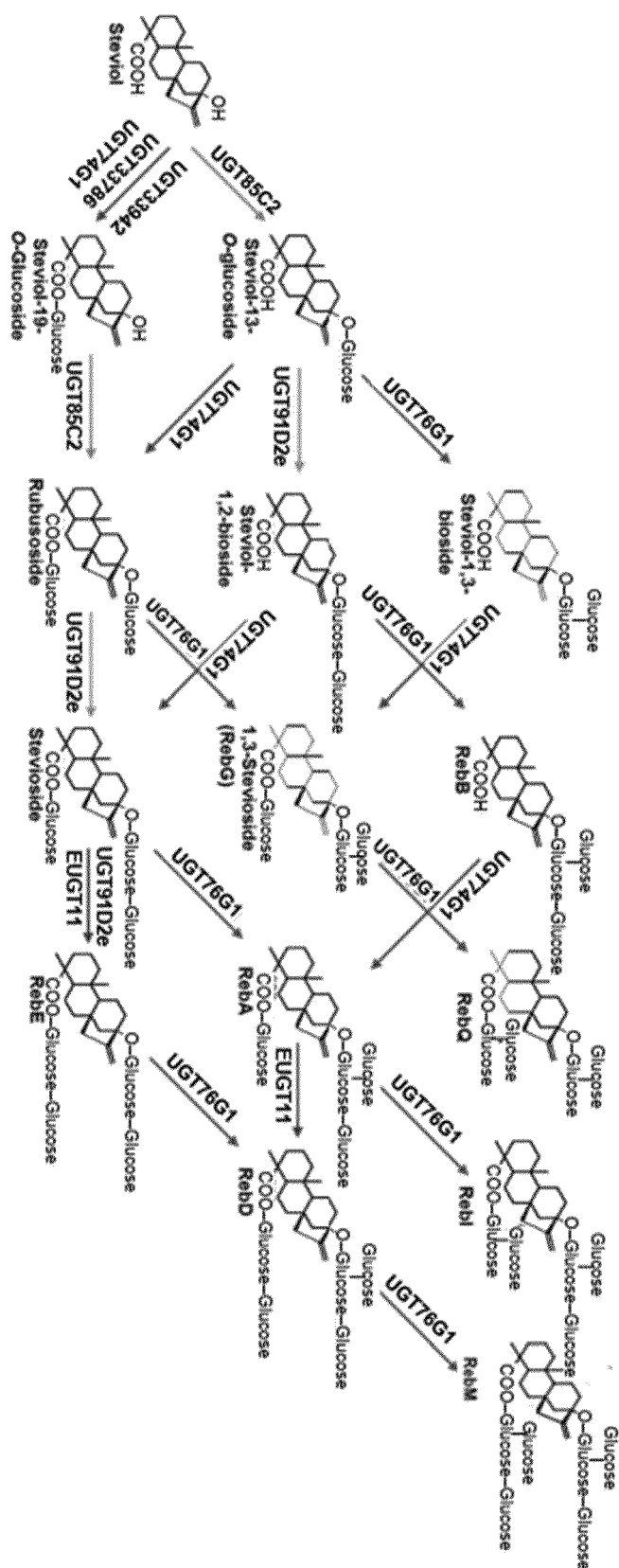


FIGURE 4

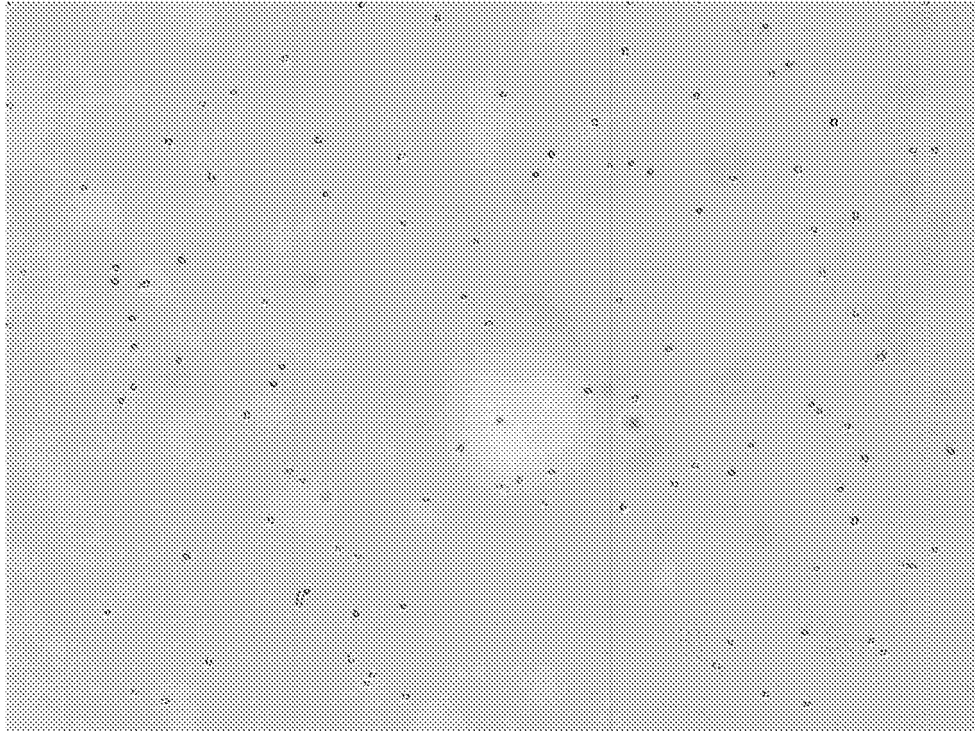


FIGURE 5 A

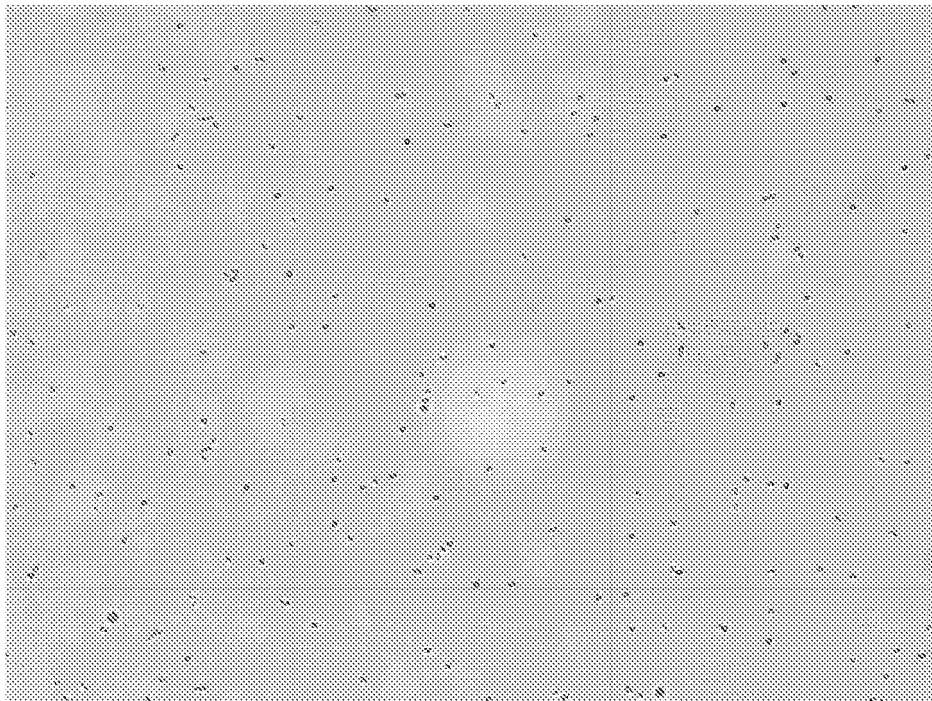


FIGURE 5 B

REFERENCES CITED IN THE DESCRIPTION

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