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Mammalian progenitor or stem cells are expanded in vitro by increasing the levels of β -catenin in the cell. The expanded cells substantially maintain their original phenotype including the ability to give rise to multiple types of differentiated cells. The intracellular levels of β -catenin may be manipulated by providing exogenous β -catenin protein to the cell, or by introduction into the cell of a genetic construct encoding β -catenin. The β -catenin may be a wild-type or stabilized mutant form of the protein. Preferably the long term cell culture medium substantially lacks stromal cells and cytokines.



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(57) Abstract: Mammalian progenitor or stem cells are expanded *in vitro* by increasing the levels of β -catenin in the cell. The expanded cells substantially maintain their original phenotype including the ability to give rise to multiple types of differentiated cells. The intracellular levels of β -catenin may be manipulated by providing exogenous β -catenin protein to the cell, or by introduction into the cell of a genetic construct encoding β -catenin. The β -catenin may be a wild-type or stabilized mutant form of the protein. Preferably the long term cell culture medium substantially lacks stromal cells and cytokines.

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EXPANSION OF STEM AND PROGENITOR CELLS BY BETA-CATENIN

INTRODUCTION

Beta-catenin is a pivotal player in the signaling pathway initiated by Wnt proteins, which are mediators of several developmental processes. Beta-catenin activity is controlled by a large number of binding partners that affect the stability and the localization of beta-catenin, and it is thereby able to participate in such varying processes as gene expression and cell adhesion. Activating mutations in beta-catenin and in components regulating its stability have been found to contribute to upregulation of cell proliferation.

The β -catenin protein becomes stabilized in response to Wnt/Wg, moves to the nucleus and forms complexes with the LEF1/TCF transcription factors to regulate gene expression. The level of cytosolic β -catenin is determined by its interaction with a number of proteins including those in a multiprotein complex of Axin, GSK-3 β , APC and other proteins. The mechanism by which the Wnt signal is transmitted to this complex is unclear but it involves interaction of Wnt with its receptors, which are members of Frizzled family of seven transmembrane proteins. The stabilization of β -catenin stimulates the expression of genes including c-myc, c-jun, fra-1, and cyclin D1. This pathway is negatively regulated by Axin.

Beta-catenin is also an adherens junction protein. Adherens junctions are critical for the establishment and maintenance of epithelial layers, such as those lining organ surfaces. AJs mediate adhesion between cells, communicate a signal that neighboring cells are present, and anchor the actin cytoskeleton. In serving these roles, AJs regulate normal cell growth and behavior. At several stages of embryogenesis, wound healing, and tumor cell metastasis, cells form and leave epithelia. This process, which involves the disruption and reestablishment of epithelial cell-cell contacts, may be regulated by the disassembly and assembly of AJs. AJs may also function in the transmission of the 'contact inhibition' signal, which instructs cells to stop dividing once an epithelial sheet is complete.

For many purposes, there is an interest in being able to expand stem and progenitor cells in culture. However, it is not simply a matter of maintaining cell viability for the stem cells, but also of ensuring that the stem cells increase in numbers without losing their distinctive phenotype. Current protocols for the *in vitro* culture of hematopoietic stem cells generally require one or a cocktail of cytokines, such as c-kit ligand (stem cell growth factor), flt-3, thrombopoietin, IL-6, etc. While a substantial increase in cell number can be obtained with such cultures, they do not provide for expanded number of cells that retain a capacity for long term repopulation of all hematopoietic lineages. See Domen and Weissman (1999) Mol Med Today 5(5):201-8; or Ziegler and Kanz (1998) Curr Opin Hematol 5(6):434-40.

Stem cells have also been grown in co-culture with stromal cells. However, it is particularly desirable to expand stem cells in a culture of known composition, rather than relying upon the presence of other cells for their maintenance.

There continues to be a strong demand for improvements in the *in vitro* culture of stem cells and progenitor cells. The present invention addresses this need.

SUMMARY OF THE INVENTION

Methods are provided for the expansion of progenitor or stem cells *in vitro*, whereby the cells retain their pluripotential phenotype after expansion. The intracellular level of β -catenin is increased in the cells in culture, either by providing exogenous β -catenin protein to the cell, or by introduction into the cell of a genetic construct encoding β -catenin. The β -catenin may be a wild type protein appropriate for the species from which the cells are derived, or preferably, a stabilized mutant form of the protein. The alteration in cellular levels of β -catenin provide for increased number of cells in cycle, and leads to cultures that containing proliferating cells that maintain an undifferentiated phenotype *in vitro*. The expanded cell populations are useful as a source of stem cells, e.g. to reconstitute function in a host that is deficient in a particular cell lineage or lineages. In one embodiment of the invention, the target cells are hematopoietic stem cells, which may be used in transplantation to restore hematopoietic function to autologous or allogeneic recipients.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. Activated beta-catenin retrovirus induces increased growth of stem cells. Stem cells infected with control or beta-catenin-GFP retrovirus were sorted and cultured on 96 well plates for two days in the presence or absence of steel factor, and cell numbers were counted at the end of the culture period.

Figure 2. Stem cells infected with beta-catenin retain many stem cell markers in long term culture. Beta-catenin infected stem cell spheres were harvested from long term cultures at 5 weeks, trypsinized and allowed to express their surface proteins for 12 hours. Subsequently they were harvested and stained with antibodies to Thy1.1, Sca1, c-kit, and lineage antigens (B220, Mac-1, Gr-1, Ter119, CD5, CD3, CD8/4).

Figure 3. Stem cells infected with beta-catenin have the ability to give rise to multiple lineages when transplanted. 100,000 beta-catenin infected stem cells were harvested from long term cultures at 7 weeks, trypsinized and injected into lethally irradiated (950 Rads) allotype marked recipients along with 300,000 rescuing bone marrow cells from the host. Analysis of reconstitution along various lineages was carried out at 4 weeks after transplantation.

DESCRIPTION OF THE SPECIFIC EMBODIMENTS

Mammalian progenitor or stem cells are expanded *in vitro* by increasing the levels of β -catenin in the cell. The intracellular levels of β -catenin may be manipulated by providing exogenous β -catenin protein to the cell, or by introduction into the cell of a genetic construct encoding β -catenin. The β -catenin may be a wild-type or stabilized mutant form of the protein. Preferably the long term cell culture medium substantially lacks stromal cells and cytokines. Cultures that provide stem cell activity can be obtained for at least three weeks, frequently six weeks and can be eight weeks or more. The culture media that are employed are conventional media for the growth of mammalian cells, optionally in the absence of serum using only defined protein factors. In the absence of the β -catenin, the medium is inefficient at maintaining growth of the undifferentiated cells.

In the first few days of culture, the expansion of stem/progenitor cells is limited, usually the number of phenotypic stem/progenitor cells is maintained, or slightly increased. After 2 to 3 weeks in the subject culture conditions, there is a substantial proliferation of cells having the desired phenotype, where the number of cells having a functional stem/progenitor cell phenotype is expanded.

DEFINITIONS

It is to be understood that this invention is not limited to the particular methodology, protocols, cell lines, animal species or genera, and reagents described, as such may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

As used herein the singular forms "a", "and", and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a cell" includes a plurality of such cells and reference to "the culture" includes reference to one or more cultures and equivalents thereof known to those skilled in the art, and so forth. All technical and scientific terms used herein have the same meaning as commonly understood to one of ordinary skill in the art to which this invention belongs unless clearly indicated otherwise.

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β -catenin: The term β -catenin, as used herein, is intended to refer to both wild-type and stabilized forms of the β -catenin protein, and to fusion proteins and derivatives thereof. Usually the protein will be of mammalian origin, although the protein from other species may find use. The protein is conserved between species, for example the human sequence is active in mouse cells. The sequences of many β -catenin proteins are publicly known. For convenience, the sequences of the human and mouse homologs of this protein are provided

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in the sequence listing, as SEQ ID NO:1; and SEQ ID NO:2, respectively. In one embodiment of the invention, a stabilized form of beta-catenin is used.

The ubiquitin-dependent proteolysis system is involved in the regulation of beta-catenin turnover. Beta-catenin becomes stabilized when proteasome-mediated proteolysis is inhibited and this leads to the accumulation of multi-ubiquitinated forms of beta-catenin (Aberle et al. (1997) EMBO J 16(13):3797-804). Substitution of the serine residues in the glycogen synthase kinase 3 β (GSK3 β) phosphorylation consensus motif of beta-catenin inhibits ubiquitination and results in stabilization of the protein. Examples of stabilized β -catenins include those with the amino acid changes D32Y; D32G; S33F; S33Y; G34E; S37C; S37F; T41I; S45Y; and deletion of AA 1-173. A number of publications describe stabilized β -catenin mutations. For example, see Morin et al. (1997) Science 275(5307):1787-90; Palacios et al. (1998) Cancer Res 58(7):1344-7; Muller et al. (1998) Genes Chromosomes Cancer 22(1):37-41; Miyoshi et al. (1998) Cancer Res 58(12):2524-7; Zurawel et al. (1998) Cancer Res. 58, 896-899; Voeller et al. (1998) Cancer Res. 58, 2520-2526; etc.

The sequence of the beta-catenin polypeptide may be altered in various ways known in the art to generate targeted changes in sequence. The polypeptide will usually be substantially similar to the sequences provided herein, i.e. will differ by at least one amino acid, and may differ by at least two but not more than about ten amino acids. Deletions may further include larger changes, such as deletions of a domain or exon, providing for active peptide fragments of the protein. Other modifications of interest include tagging, e.g. with the FLAG system, HA, green fluorescent protein, etc. Such alterations may be used to alter properties of the protein, by affecting the stability, specificity, etc. The protein may be joined to a wide variety of other oligopeptides or proteins for a variety of purposes, particular for facilitating transport across membranes.

Techniques for *in vitro* mutagenesis of cloned genes are known. Examples of protocols for scanning mutations may be found in Gustin et al., Biotechniques 14:22 (1993); Barany, Gene 37:111-23 (1985); Colicelli et al., Mol Gen Genet 199:537-9 (1985); and Prentki et al., Gene 29:303-13 (1984). Methods for site specific mutagenesis can be found in Sambrook et al., Molecular Cloning: A Laboratory Manual, CSH Press 1989, pp. 15.3-15.108; Weiner et al., Gene 126:35-41 (1993); Sayers et al., Biotechniques 13:592-6 (1992); Jones and Winistorfer, Biotechniques 12:528-30 (1992); Barton et al., Nucleic Acids Res 18:7349-55 (1990); Marotti and Tomich, Gene Anal Tech 6:67-70 (1989); and Zhu Anal Biochem 177:120-4 (1989).

Expression construct: In one embodiment of the invention, the beta-catenin is delivered to the targeted stem or progenitor cells by introduction of an exogenous nucleic acid expression vector into the cells. Many vectors useful for transferring exogenous genes

into target mammalian cells are available. The vectors may be episomal, e.g. plasmids, virus derived vectors such cytomegalovirus, adenovirus, etc., or may be integrated into the target cell genome, through homologous recombination or random integration, e.g. retrovirus derived vectors such MMLV, HIV-1, ALV, etc.

5 Retrovirus based vectors have been shown to be particularly useful when the target cells are hematopoietic stem cells. For example, see Baum *et al.* (1996) J Hematother 5(4):323-9; Schwarzenberger *et al.* (1996) Blood 87:472-478; Nolte *et al.* (1996) P.N.A.S. 93:2414-2419; and Maze *et al.* (1996) P.N.A.S. 93:206-210. Lentivirus vectors have also been described for use with hematopoietic stem cells, for example see Mochizuki *et al.* 10 (1998) J Virol 72(11):8873-83. The use of adenovirus based vectors with hematopoietic cells has also been published, see Ogniben and Haas (1998) Recent Results Cancer Res 144:86-92.

Various techniques known in the art may be used to transfect the target cells, e.g. electroporation, calcium precipitated DNA, fusion, transfection, lipofection and the like. The 15 particular manner in which the DNA is introduced is not critical to the practice of the invention.

Combinations of retroviruses and an appropriate packaging line may be used, where the capsid proteins will be functional for infecting the target cells. Usually, the cells and virus will be incubated for at least about 24 hours in the culture medium. Commonly used 20 retroviral vectors are "defective", i.e. unable to produce viral proteins required for productive infection. Replication of the vector requires growth in the packaging cell line.

The host cell specificity of the retrovirus is determined by the envelope protein, env (p120). The envelope protein is provided by the packaging cell line. Envelope proteins are of at least three types, ecotropic, amphotropic and xenotropic. Retroviruses packaged with 25 ecotropic envelope protein, e.g. MMLV, are capable of infecting most murine and rat cell types. Ecotropic packaging cell lines include BOSC23 (Pear *et al.* (1993) P.N.A.S. 90:8392-8396). Retroviruses bearing amphotropic envelope protein, e.g. 4070A (Danos *et al.*, *supra.*), are capable of infecting most mammalian cell types, including human, dog and mouse. Amphotropic packaging cell lines include PA12 (Miller *et al.* (1985) Mol. Cell. Biol. 5:431- 30 437); PA317 (Miller *et al.* (1986) Mol. Cell. Biol. 6:2895-2902) GRIP (Danos *et al.* (1988) PNAS 85:6460-6464). Retroviruses packaged with xenotropic envelope protein, e.g. AKR env, are capable of infecting most mammalian cell types, except murine cells.

The sequences at the 5' and 3' termini of the retrovirus are long terminal repeats (LTR). A number of LTR sequences are known in the art and may be used, including the 35 MMLV-LTR; HIV-LTR; AKR-LTR; FIV-LTR; ALV-LTR; etc. Specific sequences may be accessed through public databases. Various modifications of the native LTR sequences are also known. The 5' LTR acts as a strong promoter, driving transcription of the β -catenin

gene after integration into a target cell genome. For some uses, however, it is desirable to have a regulatable promoter driving expression. Where such a promoter is included, the promoter function of the LTR will be inactivated. This is accomplished by a deletion of the U3 region in the 3' LTR, including the enhancer repeats and promoter, that is sufficient to
5 inactivate the promoter function. After integration into a target cell genome, there is a rearrangement of the 5' and 3' LTR, resulting in a transcriptionally defective provirus, termed a "self-inactivating vector".

Suitable inducible promoters are activated in a desired target cell type, either the transfected cell, or progeny thereof. By transcriptional activation, it is intended that
10 transcription will be increased above basal levels in the target cell by at least about 100 fold, more usually by at least about 1000 fold. Various promoters are known that are induced in hematopoietic cell types, e.g. IL-2 promoter in T cells, immunoglobulin promoter in B cells, etc.

Preferred genetic constructs are those that can be removed from the target cells after
15 expansion. This can be accomplished by the use of a transient vector system, or by including a heterologous recombination site that flanks the beta-catenin coding sequence. In this manner, after expansion the construct can be removed prior to use of the expanded cell population. Preferably a detectable marker, e.g. green fluorescent protein, luciferase, cell surface proteins suitable for antibody selection methods, etc. is included in the expression
20 vector, such that after deletion of the construct the cells can be readily isolated that lack the exogenous beta-catenin.

The term "heterologous recombination site" is meant to encompass any introduced genetic sequence that facilitates site-specific recombination. In general, such sites facilitate recombination by interaction of a specific enzyme with two such sites. Exemplary
25 heterologous recombination sites include, but are not necessarily limited to, *lox* sequences with recombination mediated by Cre enzyme; *frt* sequences (Golic et al. (1989) *Cell* 59:499-509; O'Gorman et al. (1991) *Science* 251:1351-5; recombination mediated by the FLP recombinase), the recognition sequences for the pSR1 recombinase of *Zygosaccharomyces rouxii* (Matsuzaki et al. (1990) *J. Bacteriol.* 172:610-8), and the like.

Sequences encoding *lox* sites are of particular interest for use in the present
30 invention. A *lox* site is a nucleotide sequence at which the gene product of the *cre* gene, referred to herein as "Cre," catalyzes site-specific recombination. A particularly preferred *lox* site is a *loxP* site. The sequence of *loxP*, which is 34 bp in length, is known and can be produced synthetically or can be isolated from bacteriophage P1 by methods known in the art (see, e.g. Hoess et al. (1982) *Proc. Natl. Acad. Sci. USA* 79:3398). The *loxP* site is
35 composed of two 13 bp inverted repeats separated by an 8 bp spacer region. Other suitable

lox sites include *loxB*, *loxL*, and *loxR*, which can be isolated from *E. coli* (Hoess et al. (1982) *Proc. Natl. Acad. Sci. USA* 22:3398).

In an alternative method, expression vectors that provide for the transient expression in mammalian cells may be used. In general, transient expression involves the use of an expression vector that is able to replicate efficiently in a host cell, such that the host cell accumulates many copies of the expression vector and, in turn, synthesizes high levels of a desired polypeptide encoded by the expression vector. Transient expression systems, comprising a suitable expression vector and a host cell, allow for the convenient short term expansion of cells, but do not affect the long term genotype of the cell.

Translocation modified β -catenin: In some cases it is desirable to provide exogenous β -catenin protein, rather than transducing the cells with an expression construct. The beta-catenin may be added to the culture medium at high levels. Preferably the beta-catenin is modified so as to increase its transport into the cells.

In one embodiment of the invention, tat protein is used to deliver beta-catenin. The preferred transport polypeptides are characterized by the presence of the tat basic region amino acid sequence (amino acids 49-57 of naturally-occurring tat protein); the absence of the tat cysteine-rich region amino acid sequence (amino acids 22-36 of naturally-occurring tat protein) and the absence of the tat exon 2-encoded carboxy-terminal domain (amino acids 73-86 of naturally-occurring tat protein). Transport polypeptides are attached to beta-catenin by chemical cross-linking or by genetic fusion, where the beta-catenin moiety may be a wild-type or stabilized form. A unique terminal cysteine residue is a preferred means of chemical cross-linking.

Stem cell: The term stem cell is used herein to refer to a mammalian cell that has the ability both to self-renew, and to generate differentiated progeny (see Morrison et al. (1997) *Cell* 88:287-298). Generally, stem cells also have one or more of the following properties: an ability to undergo asynchronous, or symmetric replication, that is where the two daughter cells after division can have different phenotypes; extensive self-renewal capacity; capacity for existence in a mitotically quiescent form; and clonal regeneration of all the tissue in which they exist, for example the ability of hematopoietic stem cells to reconstitute all hematopoietic lineages. "Progenitor cells" differ from stem cells in that they typically do not have the extensive self-renewal capacity, and often can only regenerate a subset of the lineages in the tissue from which they derive, for example only lymphoid, or erythroid lineages in a hematopoietic setting.

Stem cells may be characterized by both the presence of markers associated with specific epitopes identified by antibodies and the absence of certain markers as identified by

the lack of binding of specific antibodies. Stem cells may also be identified by functional assays both *in vitro* and *in vivo*, particularly assays relating to the ability of stem cells to give rise to multiple differentiated progeny.

Stem cells of interest include hematopoietic stem cells and progenitor cells derived therefrom (U.S. Pat. No. 5,061,620); neural crest stem cells (see Morrison *et al.* (1999) Cell 96:737-749); embryonic stem cells; mesenchymal stem cells; mesodermal stem cells; *etc.*

Other hematopoietic "progenitor" cells of interest include cells dedicated to lymphoid lineages, e.g. immature T cell and B cell populations. The methods of the present invention are useful in expanding selected populations of these cells.

Purified populations of stem or progenitor cells may be used to initiate the cultures. For example, human hematopoietic stem cells may be positively selected using antibodies specific for CD34, thy-1; or negatively selected using lineage specific markers which may include glycophorin A, CD3, CD24, CD16, CD14, CD38, CD45RA, CD36, CD2, CD19, CD56, CD66a, and CD66b; T cell specific markers, tumor specific markers, *etc.* Markers useful for the separation of mesodermal stem cells include FcγRII, FcγRIII, Thy-1, CD44, VLA-4α, LFA-1β, HSA, ICAM-1, CD45, Aa4.1, Sca-1, *etc.* Neural crest stem cells may be positively selected with antibodies specific for low-affinity nerve growth factor receptor (LNGFR), and negatively selected for the markers sulfatide, glial fibrillary acidic protein (GFAP), myelin protein P₀, peripherin and neurofilament. Human mesenchymal stem cells may be positively separated using the markers SH2, SH3 and SH4.

The cells of interest are typically mammalian, where the term refers to any animal classified as a mammal, including humans, domestic and farm animals, and zoo, laboratory, sports, or pet animals, such as dogs, horses, cats, cows, mice, rats, rabbits, *etc.* Preferably, the mammal is human.

The cells which are employed may be fresh, frozen, or have been subject to prior culture. They may be fetal, neonate, adult. Hematopoietic cells may be obtained from fetal liver, bone marrow, blood, particularly G-CSF or GM-CSF mobilized peripheral blood, or any other conventional source. The manner in which the stem cells are separated from other cells of the hematopoietic or other lineage is not critical to this invention. As described above, a substantially homogeneous population of stem or progenitor cells may be obtained by selective isolation of cells free of markers associated with differentiated cells, while displaying epitopic characteristics associated with the stem cells.

Culture medium: The stem or progenitor cells are grown *in vitro* in an appropriate liquid nutrient medium. Generally, the seeding level will be at least about 10 cells/ml, more usually at least about 100 cells/ml and generally not more than about 10⁵ cells/ml, usually not more than about 10⁴ cells/ml.

Various media are commercially available and may be used, including Ex vivo serum free medium; Dulbecco's Modified Eagle Medium (DMEM), RPMI, Iscove's medium, etc. The medium may be supplemented with serum or with defined additives. Appropriate antibiotics to prevent bacterial growth and other additives, such as pyruvate (0.1-5 mM), glutamine (0.5-5 mM), 2-mercaptoethanol ($1-10 \times 10^{-5}$ M) may also be included.

Culture in serum-free medium is of particular interest. The medium may be any conventional culture medium, generally supplemented with additives such as iron-saturated transferrin, human serum albumin, soy bean lipids, linoleic acid, cholesterol, alpha thioglycerol, crystalline bovine hemin, etc., that allow for the growth of hematopoietic cells.

Preferably the expansion medium is free of cytokines, particularly cytokines that induce cellular differentiation. The term cytokine may include lymphokines, monokines and growth factors. Included among the cytokines are thrombopoietin (TPO); nerve growth factors such as NGF- β ; platelet-growth factor; transforming growth factors (TGFs) such as TGF- α and TGF- β ; erythropoietin (EPO); interferons such as interferon- α , - β , and - γ ; colony stimulating factors (CSFs) such as macrophage-CSF (M-CSF); granulocyte-macrophage-CSF (GM-CSF); and granulocyte-CSF (G-CSF); interleukins (ILs) such as IL-1, IL-1 α , IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-11, IL-12; etc. In some circumstances, proliferative factors that do not induce cellular differentiation may be included in the cultures, e.g. c-kit ligand, LIF, and the like.

EXPANSION OF STEM/PROGENITOR CELLS

A population of cells comprising progenitor and/or stem cells is cultured *in vitro* in the presence of enhanced levels of β -catenin, either by genetically altering the cells, or by providing exogenous β -catenin, as described above. The upregulation in β -catenin is sufficient to maintain or increase the number of assayable progenitor cells in the culture. The number of assayable progenitor cells may be demonstrated by a number of assays. After one week the progenitor cell cloning efficiency will usually be at least about 75% that of the starting cell population, more usually 100% that of the starting cell population, and may be as high as 200% that of the starting cell population.

Following the initial period, there is an increased expansion, where the number of assayable cells having the functional phenotype of the initial cell population can increase from about 5 to about 100 fold or more. After this time, the cells can remain in cycle, and expansion is limited primarily by considerations of space. The cells can be frozen using conventional methods at any time, usually after the first week of culture.

Frequently stem cells are isolated from biological sources in a quiescent state. Certain expression vectors, particularly retroviral vectors, do not effectively infect non-cycling cells. Cultures established with these vectors as a source of beta-catenin sequences are

induced to enter the cell cycle by a short period of time in culture with growth factors. For example, hematopoietic stem cells are induced to divide by culture with c-kit ligand, which may be combined with LIF, IL-11 and thrombopoietin. After 24 to 72 hours in culture with cytokines, the medium is changed, and the cells are contacted with the retroviral culture,
5 using culture conditions as described above.

After seeding the culture medium, the culture medium is maintained under conventional conditions for growth of mammalian cells, generally about 37° C and 5% CO₂ in 100% humidified atmosphere. Fresh media may be conveniently replaced, in part, by removing a portion of the media and replacing it with fresh media. Various commercially
10 available systems have been developed for the growth of mammalian cells to provide for removal of adverse metabolic products, replenishment of nutrients, and maintenance of oxygen. By employing these systems, the medium may be maintained as a continuous medium, so that the concentrations of the various ingredients are maintained relatively constant or within a predescribed range. Such systems can provide for enhanced
15 maintenance and growth of the subject cells using the designated media and additives.

These cells may find various applications for a wide variety of purposes. The cell populations may be used for screening various additives for their effect on growth and the mature differentiation of the cells. In this manner, compounds which are complementary, agonistic, antagonistic or inactive may be screened, determining the effect of the compound
20 in relationship with one or more of the different cytokines.

The populations may be employed as grafts for transplantation. For example, hematopoietic cells are used to treat malignancies, bone marrow failure states and congenital metabolic, immunologic and hematologic disorders. Marrow samples may be taken from patients with cancer, and enriched populations of hematopoietic stem cells
25 isolated by means of density centrifugation, counterflow centrifugal elutriation, monoclonal antibody labeling and fluorescence activated cell sorting. The stem cells in this cell population are then expanded *in vitro* and can serve as a graft for autologous marrow transplantation. The graft will be infused after the patient has received curative chemo-radiotherapy.

The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the present invention, and are not intended to limit the scope of what the inventors regard as their invention nor are they intended to represent that the experiments below are all or the only experiments
35 performed. Efforts have been made to ensure accuracy with respect to numbers used (e.g. amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is

weight average molecular weight, temperature is in degrees Centigrade, and pressure is at or near atmospheric.

All publications and patent applications cited in this specification are herein incorporated by reference as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference.

The present invention has been described in terms of particular embodiments found or proposed by the present inventor to comprise preferred modes for the practice of the invention. It will be appreciated by those of skill in the art that, in light of the present disclosure, numerous modifications and changes can be made in the particular embodiments exemplified without departing from the intended scope of the invention. For example, due to codon redundancy, changes can be made in the underlying DNA sequence without affecting the protein sequence. Moreover, due to biological functional equivalency considerations, changes can be made in protein structure without affecting the biological action in kind or amount. All such modifications are intended to be included within the scope of the appended claims.

EXPERIMENTAL

Bone marrow cells from BCL2 transgenic mice were isolated, enriched for c-kit over a magnetic column, and then stained with antibodies to sort the Sca1⁺ Thy1.1^{lo} c-kit⁺ lin^{-lo} population on a cell sorter. The cells were double sorted to ensure a high level of purity.

The cells were cultured to initiate cell cycle with Steel factor 100ng/ml with 5% serum in X-Vivo 15 containing the retrovirus. At 3 days 50% of media was replaced with only X-vivo 15, and this dilution was repeated every 2 days. The cells were then cultured with supernatant containing retrovirus encoding activated beta-catenin and recombinant steel factor. The increased growth of the stem cells is shown in Figure 1.

The retroviral supernatant had been generated in commercially available X-vivo 15 media using phoenix cells and a MSCV retroviral construct containing beta-catenin driven by the LTR. The retroviral construct is called MSCV and contains an IRES-GFP, in order to label infected cells. The activating beta catenin mutation is a mutation at the amino terminus that prevents phosphorylation and subsequent degradation by proteosomes. The accumulation of beta catenin in the cytosol allows it to translocate to the nucleus where it associates with the LEF/TCF family of transcription factors to turn on gene expression.

50% of the culture supernatant was replaced every day for 3 days. At the end of this culture period the media was replaced with X-vivo 15. Clusters of cells grew out of this culture, and were analyzed at 5 weeks. By May-Gruenwald-Geimsa staining, these cells appeared to have an immature phenotype with large nuclei and small cytoplasm. By FACS staining a majority of cells are Thy1^{lo} Sca-1⁺Lin^{lo}/-kit^{lo}, a phenotype resembling that of stem

cells. About 50% of the cells are Lin⁻ (LT-HSC phenotype), and 50% Lin^{lo} (ST-HSC phenotype). The analysis is shown in Figure 2.

5 These cells give rise to lineage positive cells at 4 weeks when transplanted into lethally irradiated mice suggesting that they are able to differentiate to various lineages *in vivo*, while remaining immature *in vitro*.

Lethally irradiated mice were injected with 300, 000 host bone marrow and 100,000 cultured cells. Peripheral blood was take at a later time, at 2 weeks, 3 weeks and 4 weeks so far. Donor type was marked with Ly5.1+ cells. Level of differentiation was determined by using antibodies to mature lineage markers. The results are shown in Figure 3,
10 demonstrating that stem cells over-expressing β -catenin have the ability to give rise to multiple lineages when transplanted.

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SEQUENCE LISTING

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Roeland Nusse
Irving L. Weissman

<120> Use of beta-catenin in the expansion of
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Val Val Asn	Lys Ala Ala	Val Met Val	His Gln Leu	Ser Lys Lys	Glu	
	170	175	180			
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Ala Ser Arg	His Ala Ile	Met Arg Ser	Pro Gln Met	Val Ser Ala	Ile	
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Val Arg Thr	Met Gln Asn	Thr Asn Asp	Val Glu Thr	Ala Arg Cys	Thr	
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gct ggg acc	ctt cac aac	ctt tct cac	cac cgc gag	ggc ttg ctg	gcc	787
Ala Gly Thr	Leu His Asn	Leu Ser His	His Arg Glu	Gly Leu Leu	Ala	
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cca gtg gat tct gta ctg ttc tac gcc atc acg aca ctg cat aat ctc Pro Val Asp Ser Val Leu Phe Tyr Ala Ile Thr Thr Leu His Asn Leu 250 255 260	883
ctg ctc cat cag gaa gga gct aaa atg gca gtg cgc cta gct ggt gga Leu Leu His Gln Glu Gly Ala Lys Met Ala Val Arg Leu Ala Gly Gly 265 270 275	931
ctg cag aaa atg gtt gct ttg ctc aac aaa aca aac gtg aaa ttc ttg Leu Gln Lys Met Val Ala Leu Leu Asn Lys Thr Asn Val Lys Phe Leu 280 285 290	979
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His Trp Pro Leu Ile Lys Ala Thr Val Gly Leu Ile Arg Asn Leu Ala	
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 35 40 45
 Lys Gly Asn Pro Glu Glu Glu Asp Val Asp Thr Ser Gln Val Leu Tyr
 50 55 60
 Glu Trp Glu Gln Gly Phe Ser Gln Ser Phe Thr Gln Glu Gln Val Ala
 65 70 75 80
 Asp Ile Asp Gly Gln Tyr Ala Met Thr Arg Ala Gln Arg Val Arg Ala
 85 90 95
 Ala Met Phe Pro Glu Thr Leu Asp Glu Gly Met Gln Ile Pro Ser Thr
 100 105 110
 Gln Phe Asp Ala Ala His Pro Thr Asn Val Gln Arg Leu Ala Glu Pro
 115 120 125
 Ser Gln Met Leu Lys His Ala Val Val Asn Leu Ile Asn Tyr Gln Asp
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 Asp Ala Glu Leu Ala Thr Arg Ala Ile Pro Glu Leu Thr Lys Leu Leu
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 Asn Asp Glu Asp Gln Val Val Val Asn Lys Ala Ala Val Met Val His
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 Gln Leu Ser Lys Lys Glu Ala Ser Arg His Ala Ile Met Arg Ser Pro
 180 185 190
 Gln Met Val Ser Ala Ile Val Arg Thr Met Gln Asn Thr Asn Asp Val
 195 200 205
 Glu Thr Ala Arg Cys Thr Ala Gly Thr Leu His Asn Leu Ser His His
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Val	Lys	Met	Leu	Gly	Ser	Pro	Val	Asp	Ser	Val	Leu	Phe	Tyr	Ala	Ile
				245					250					255	
Thr	Thr	Leu	His	Asn	Leu	Leu	Leu	His	Gln	Glu	Gly	Ala	Lys	Met	Ala
			260					265					270		
Val	Arg	Leu	Ala	Gly	Gly	Leu	Gln	Lys	Met	Val	Ala	Leu	Leu	Asn	Lys
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Pro	Gln	Ala	Leu	Val	Asn	Ile	Met	Arg	Thr	Tyr	Thr	Tyr	Glu	Lys	Leu
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Leu	Trp	Thr	Thr	Ser	Arg	Val	Leu	Lys	Val	Leu	Ser	Val	Cys	Ser	Ser
			340					345					350		
Asn	Lys	Pro	Ala	Ile	Val	Glu	Ala	Gly	Gly	Met	Gln	Ala	Leu	Gly	Leu
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Val	Thr	Cys	Ala	Ala	Gly	Ile	Leu	Ser	Asn	Leu	Thr	Cys	Asn	Asn	Tyr
			420					425					430		
Lys	Asn	Lys	Met	Met	Val	Cys	Gln	Val	Gly	Gly	Ile	Glu	Ala	Leu	Val
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Ile	Cys	Ala	Leu	Arg	His	Leu	Thr	Ser	Arg	His	Gln	Glu	Ala	Glu	Met
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Ala	Gln	Asn	Ala	Val	Arg	Leu	His	Tyr	Gly	Leu	Pro	Val	Val	Val	Lys
				485					490					495	
Leu	Leu	His	Pro	Pro	Ser	His	Trp	Pro	Leu	Ile	Lys	Ala	Thr	Val	Gly
			500					505					510		
Leu	Ile	Arg	Asn	Leu	Ala	Leu	Cys	Pro	Ala	Asn	His	Ala	Pro	Leu	Arg
		515					520					525			
Glu	Gln	Gly	Ala	Ile	Pro	Arg	Leu	Val	Gln	Leu	Leu	Val	Arg	Ala	His
	530					535					540				
Gln	Asp	Thr	Gln	Arg	Arg	Thr	Ser	Met	Gly	Gly	Thr	Gln	Gln	Gln	Phe
545					550					555					560
Val	Glu	Gly	Val	Arg	Met	Glu	Glu	Ile	Val	Glu	Gly	Cys	Thr	Gly	Ala
				565					570					575	
Leu	His	Ile	Leu	Ala	Arg	Asp	Val	His	Asn	Arg	Ile	Val	Ile	Arg	Gly
			580					585					590		
Leu	Asn	Thr	Ile	Pro	Leu	Phe	Val	Gln	Leu	Leu	Tyr	Ser	Pro	Ile	Glu
		595					600					605			
Asn	Ile	Gln	Arg	Val	Ala	Ala	Gly	Val	Leu	Cys	Glu	Leu	Ala	Gln	Asp
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Lys	Glu	Ala	Ala	Glu	Ala	Ile	Glu	Ala	Glu	Gly	Ala	Thr	Ala	Pro	Leu
625					630					635					640
Thr	Glu	Leu	Leu	His	Ser	Arg	Asn	Glu	Gly	Val	Ala	Thr	Tyr	Ala	Ala
				645					650					655	
Ala	Val	Leu	Phe	Arg	Met	Ser	Glu	Asp	Lys	Pro	Gln	Asp	Tyr	Lys	Lys
			660					665					670		
Arg	Leu	Ser	Val	Glu	Leu	Thr	Ser	Ser	Leu	Phe	Arg	Thr	Glu	Pro	Met
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Ala	Trp	Asn	Glu	Thr	Ala	Asp	Leu	Gly	Leu	Asp	Ile	Gly	Ala	Gln	Gly
	690					695					700				

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Glu	Ala	Leu	Gly	Tyr	Arg	Gln	Asp	Asp	Pro	Ser	Tyr	Arg	Ser	Phe	His
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Ser	Gly	Gly	Tyr	Gly	Gln	Asp	Ala	Leu	Gly	Met	Asp	Pro	Met	Met	Glu
				725					730						735
His	Glu	Met	Gly	Gly	His	His	Pro	Gly	Ala	Asp	Tyr	Pro	Val	Asp	Gly
			740					745					750		
Leu	Pro	Asp	Leu	Gly	His	Ala	Gln	Asp	Leu	Met	Asp	Gly	Leu	Pro	Pro
		755					760					765			
Gly	Asp	Ser	Asn	Gln	Leu	Ala	Trp	Phe	Asp	Thr	Asp	Leu			
770						775					780				

WHAT IS CLAIMED IS:

1. A method for *in vitro* expansion of mammalian stem or progenitor cells, the method comprising:

increasing the intracellular concentration of β -catenin in a progenitor or stem cell in an *in vitro* culture medium for a period of time sufficient for said progenitor or stem cell to divide;

wherein the number of cells having the functional phenotype of said stem or progenitor cells is expanded.

2. The method of Claim 1, wherein said stem or progenitor cell is a stem cell.

3. The method of Claim 2, wherein said stem cell is a hematopoietic stem cell.

4. The method of Claim 2, wherein said stem cell is a neural crest stem cell.

5. The method of Claim 2, wherein said stem cell is a mesenchymal stem cell.

6. The method of Claim 2, wherein said stem cell is an embryonic stem cell.

7. The method of Claim 3, wherein said hematopoietic stem cell is a human cell.

8. The method of Claim 1, wherein said step of increasing the intracellular concentration of β -catenin comprises:

introduction of an exogenous nucleic acid comprising beta-catenin coding sequences operably linked to a promoter.

9. The method of Claim 8, wherein said beta-catenin is a wild-type beta-catenin.

10. The method of Claim 8, wherein said beta-catenin is a stabilized mutant beta-catenin.

11. The method of Claim 8, wherein said exogenous nucleic acid is a retroviral vector.

12. The method of Claim 11, wherein said retroviral vector comprises sites for recombination, flanking said beta-catenin coding sequences.

13. The method of Claim 8, wherein said exogenous nucleic acid is an episomal vector.

14. The method of Claim 1, wherein said step of increasing the intracellular
5 concentration of β -catenin comprises:
addition of exogenous beta-catenin to said culture medium.

15. The method of Claim 14, wherein said beta-catenin is a wild-type beta-catenin.

10

16. The method of Claim 14, wherein said beta-catenin is a stabilized mutant beta-catenin.

15

17. The method of Claim 14, wherein said beta-catenin is genetically fused to a transport moiety.

18. The method of Claim 17, wherein said transport moiety is a fragment of HIV tat protein.

20

19. The method of Claim 1, wherein said stem or progenitor cell is a progenitor cell.

20. The method of Claim 19, wherein said progenitor cell is a hematopoietic progenitor cell.

25

21. The method of Claim 20, wherein said hematopoietic progenitor cell is a lymphoid cell.

22. The method of Claim 21, wherein said lymphoid cell is a B cell.

30

23. The method of Claim 21, wherein said lymphoid cell is a T cell.

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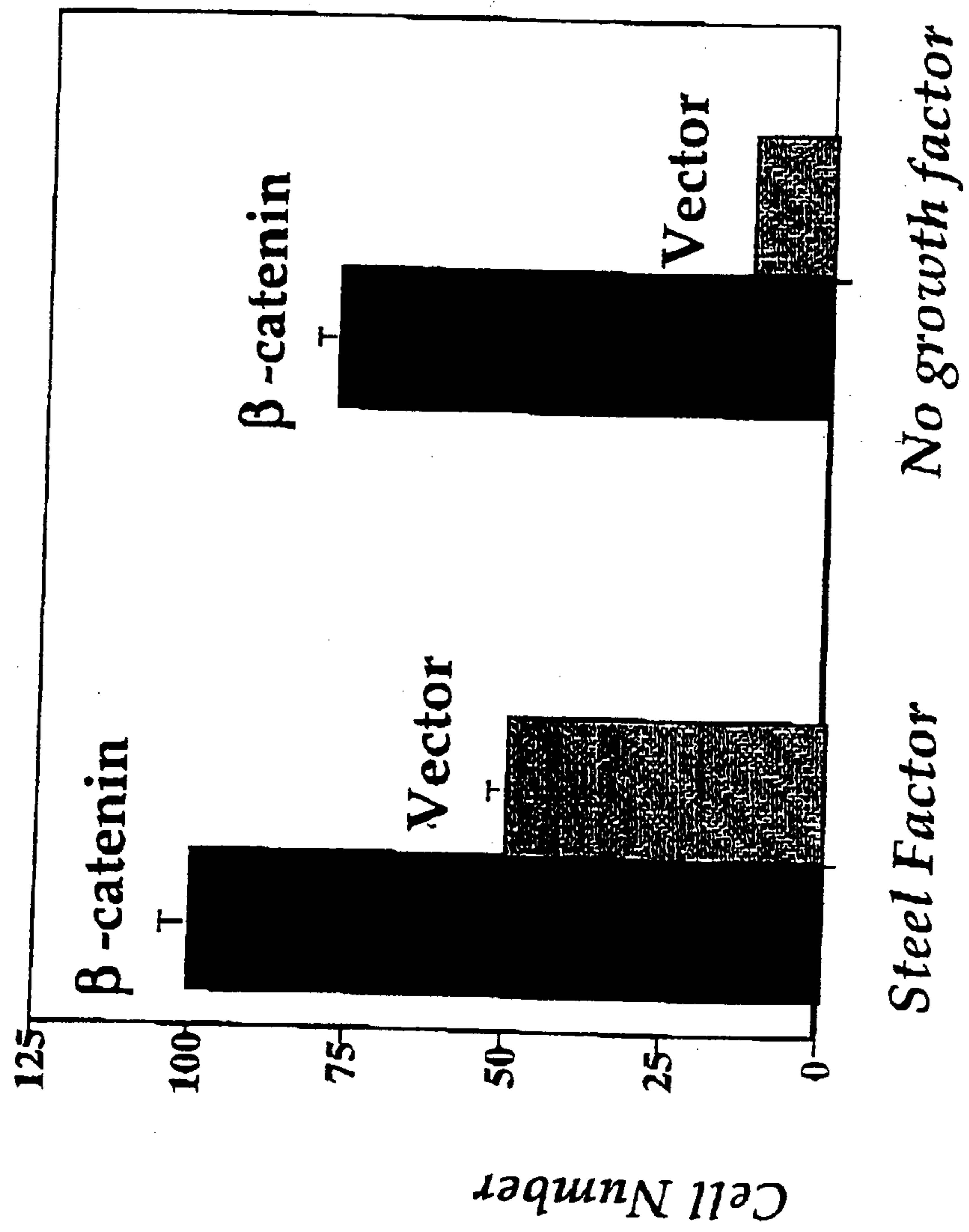


Figure 1

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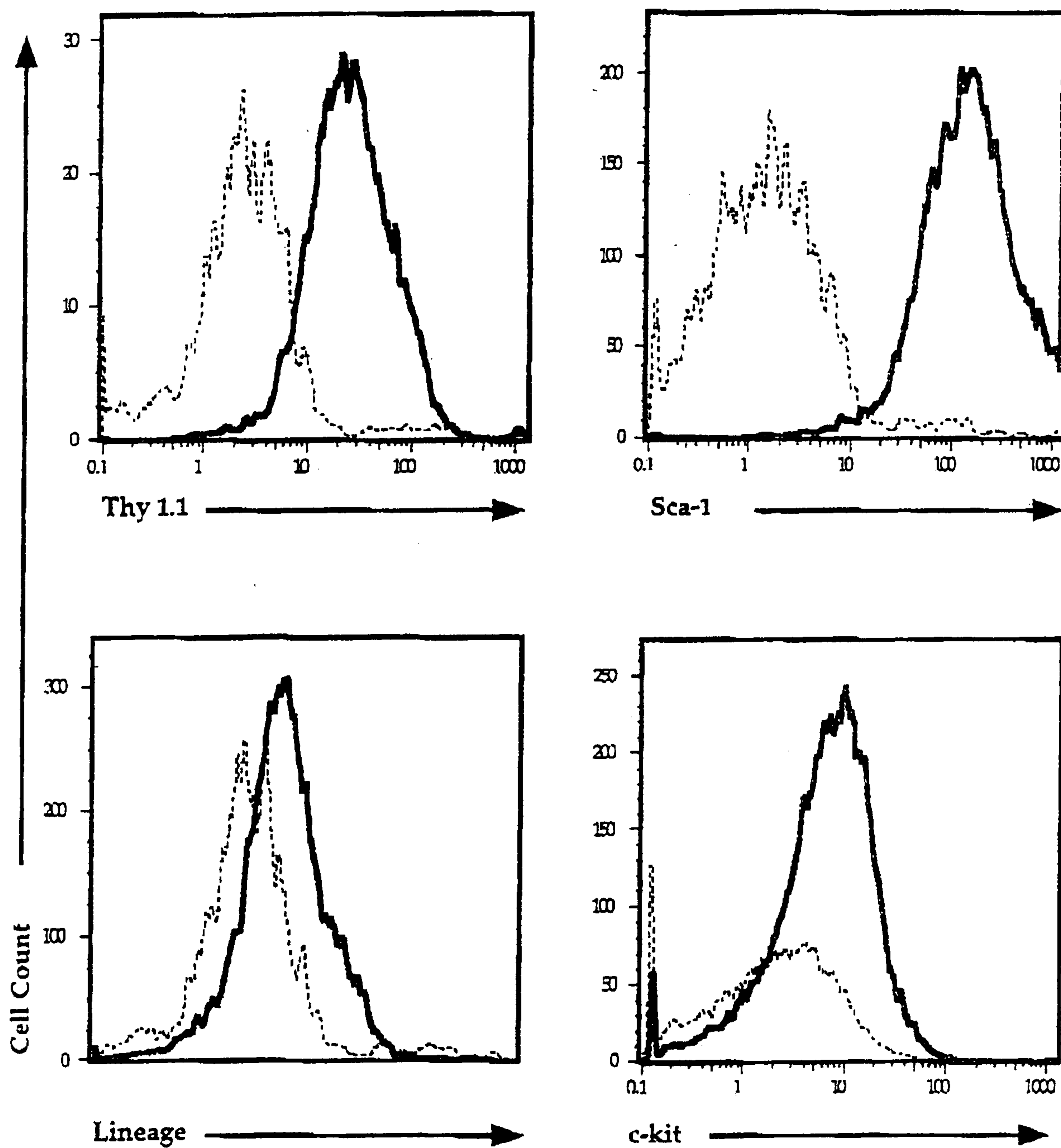


Figure 2

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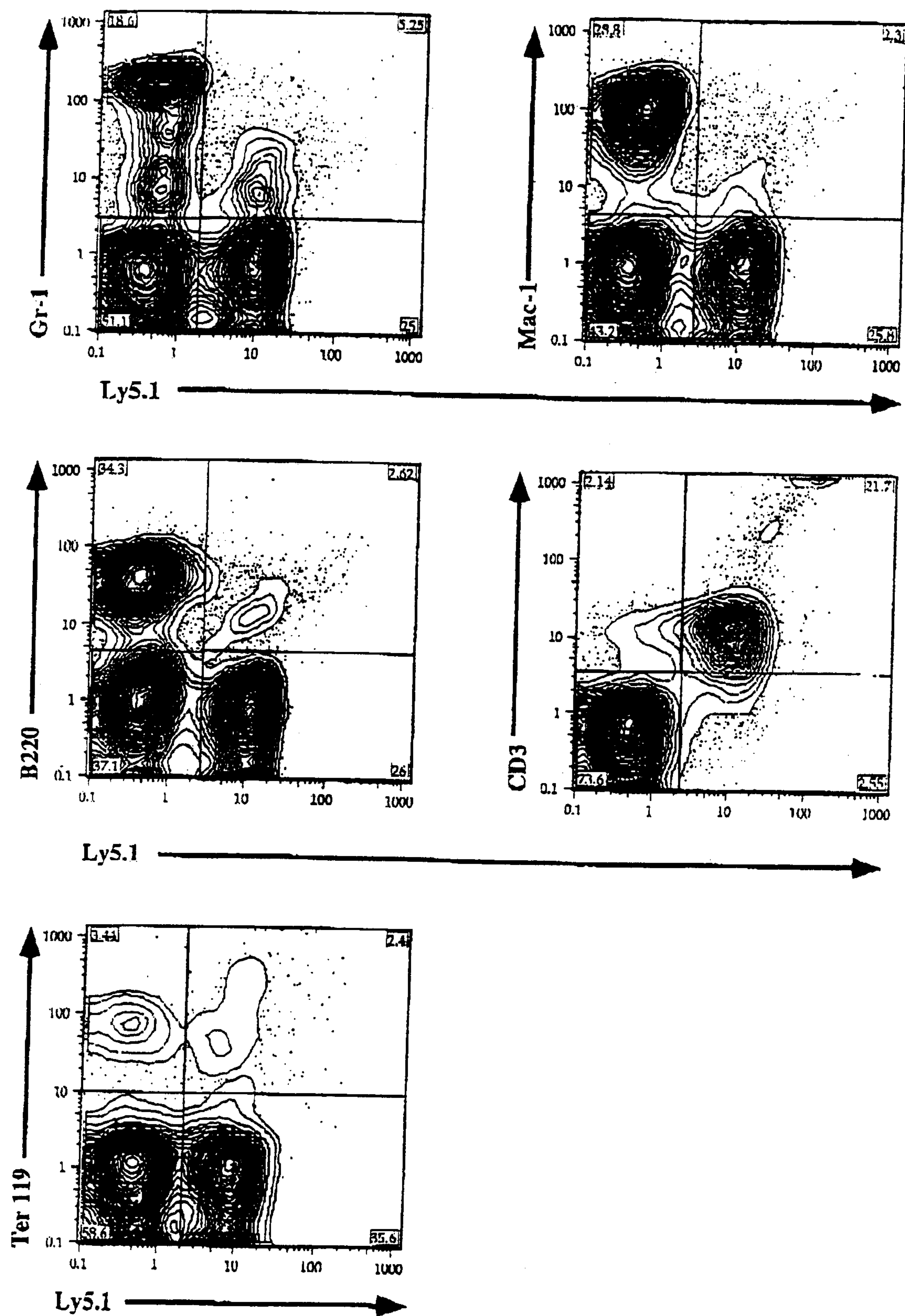


Figure 3