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(72) Inventors; and

(71) Applicants: **KHAYAMIAN, Mohammad Ali** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR). **ABDOLAHAD, Mohammad** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR). **SALEMIZADEH PARIZI, Mohammad** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR). **GHADERINIA, Mohammad Reza** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR). **ABADIJOO, Hamed** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR). **VANAEI, Shohreh** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR). **SIMAEE, Hossein** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR). **SHALILEH, Shahriar** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR). **FARAMARZPOUR DARZINI, Mahsa** [IR/IR]; Nano Bio Electronic Devices Lab, College of Engineering, University of Tehran., Tehran, 14395-515 (IR).

(74) Agent: **IDESAZAN ASR AFTAB**; Unit 7, Pardis building, Behboudi St., Sattarkhan, Tehran (IR).

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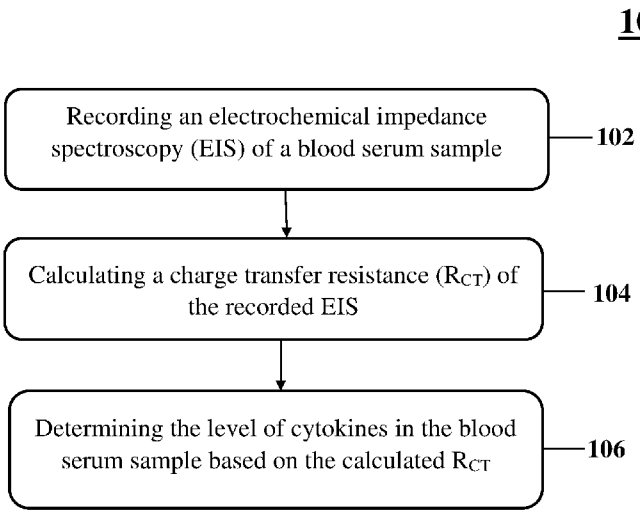


FIG. 1A

(57) Abstract: Disclosed herein is method for measuring a level of total cytokines in a blood serum sample. The method includes recording an electrochemical impedance spectroscopy (EIS) associated with the blood serum sample, calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS, and determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} by looking up the calculated R_{CT} in a calibration dataset including a set of total cytokine mass values respective to a set of R_{CT} values.

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REAL-TIME TRACING OF CYTOKINE STORM IN BLOOD SERUM OF COVID-19 PATIENTS

TECHNICAL FIELD

[0001] The present disclosure generally relates to detecting cytokine storm in a patient, and particularly, to a label-free electrochemical approach to detect COVID-19 infection status of a patient by recording and analyzing an electrochemical impedance spectroscopy (EIS) of a blood serum sample of the patient.

BACKGROUND ART

[0002] COVID-19 as a new member of fetal coronaviruses severely infects the lower respiratory tract and leads to several common symptoms including dry cough, fever and shortness of breath in many patients. Due to the highly contagious nature of this virus due to both person-to-person and airborne transmission, along with a daily upsurge of newly infected patients globally, COVID-19 was announced as a pandemic by the World Health Organization (WHO) in 2019.

[0003] There are various aspects about COVID-19 virus which are still quite unknowns in the scientific community. However, some insight has been gleaned over the last year based on case studies. For example, after entry of COVID-19 to a respiratory tract, three phases are presumed: viral replication, immune hyperactivity or cytokine storm, and finally a pulmonary dysfunction. In fact, it appears that when infected with COVID-19, the immune system itself attacks the infected patient. When the white blood cells are faced with the virus or virus-infected cells, they become hyperactivated and unleash (secrete) a storm of cytokines into the blood medium to call other immune cells. Furthermore, reports demonstrate that cytokine storm is the most relevant reason behind the mortality of the COVID-19 patients by inducing an acute respiratory distress syndrome (ARDS). On this basis, notable fraction of clinical trials are attempting to suppress the hyperactivation of the immune system in response to diagnosis of a COVID-19 infection.

[0004] Hence, monitoring the acute elevation of cytokines could be a reliable warning approach to check the severity of the disease in the infected patients. For example, S. M. Russell et al. in “Biosensors for Managing the COVID-19 Cytokine Storm: Challenges Ahead”, ACS

Sensors, 2020, studied cytokine storm in COVID-19 as a key parameter and reported that biosensors are needed for COVID-19 management and must be rapid, sufficiently sensitive to detect cytokines in whole blood, and independent of centralized equipment. Many methods were developed for rapid and precise tracing of cytokine storm in blood. However, many of these conventional approaches are time-consuming and have expensive protocols. Nonetheless, the conventional Enzyme-linked immunosorbent assay (ELISA) method even with its limitations such as cross-reactivity of the antibodies such as reported by M. S. Draz et al. in Nanoscale, 2018, vol. 10, pages 11841-11849, is still the most utilized technique among all proposed procedures.

[0005] Hence, there is still a need for a real-time, simple, cost-effective and precise method, system, and associated apparatus for fast reliable monitoring detection of cytokine storm in patients, and specifically, in patients with COVID-19 symptoms. Furthermore, there is a need for a fast-diagnosing and easy-to-use method to detect COVID-19 infection in a suspicious person to avoid further spreading the virus. Moreover, there is a need for a cost-effective and simple method to detect COVID-19 in suspected patients without any need for expensive and complicated techniques and devices.

SUMMARY OF THE DISCLOSURE

[0006] This summary is intended to provide an overview of the subject matter of this patent, and is not intended to identify essential elements or key elements of the subject matter, nor is it intended to be used to determine the scope of the claimed implementations. The proper scope of this patent may be ascertained from the claims set forth below in view of the detailed description below and the drawings.

[0007] In one general aspect, the present disclosure describes an exemplary method for measuring a level of total cytokines in a blood serum sample. The method may include recording an electrochemical impedance spectroscopy (EIS) associated with the blood serum sample, calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS, and determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} by looking up the calculated R_{CT} in a calibration dataset comprising a set of total cytokine mass values respective to a set of R_{CT} values.

[0008] In an exemplary implementation, the method may further include generating the calibration dataset. In an exemplary implementation, generating the calibration dataset may include generating the set of R_{CT} values and generating the set of total cytokine mass values corresponding to the set of R_{CT} values. In an exemplary implementation, generating the set of R_{CT} values may include recording a set of EIS plots from a respective plurality of blood serum samples drawn from a corresponding plurality of persons and calculating the set of R_{CT} values respective to the set of recorded EIS plots by measuring a diameter of a semicircular curved part of each of the recorded EIS plots. In an exemplary implementation, generating the set of total cytokine mass values corresponding to the set of R_{CT} values may include measuring total cytokines mass of each of the plurality of blood serum samples utilizing a cytokine measurement assay.

[0009] In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} may further include diagnosing an infection of a chronic inflammatory disease (CID). In an exemplary implementation, diagnosing the CID may include detecting the infection of a chronic inflammatory disease for a person associated with the blood serum sample if the calculated R_{CT} is more than a threshold value.

[0010] In an exemplary implementation, detecting the infection of the chronic inflammatory disease may include detecting a COVID-19 infection of the person if the calculated R_{CT} is equal to or more than 440 Ω. In an exemplary implementation, detecting the infection of the chronic inflammatory disease may include detecting a COVID-19 infection status of the person based on the calculated R_{CT}. In an exemplary implementation, detecting the COVID-19 infection status of the person based on the calculated R_{CT} may include detecting a healthy (normal) state for the person if the calculated R_{CT} is less than 340 Ω, detecting a moderate COVID-19 infection of the person if the calculated R_{CT} is between 440 Ω and 610 Ω, and detecting a severe COVID-19 infection of the person if the calculated R_{CT} is more than 715 Ω.

[0011] In an exemplary implementation, determining the level of total cytokines in a blood serum sample based on the calculated R_{CT} may further include grading a severity of the chronic inflammatory disease infection of the person. In an exemplary implementation, grading the severity of the chronic inflammatory disease infection of the person may include detecting the person is in a healthy (normal) state if the calculated R_{CT} is within a first range of the calibration dataset, detecting the person is in a moderately infected state if the calculated R_{CT} is within a second range of the calibration dataset, and detecting the person is in a severely infected state

if the calculated R_{CT} is within a third range of the calibration dataset. Where, the first range of the calibration dataset may include a first set of R_{CT} values less than the threshold value measured from a group of healthy people. The second range of the calibration dataset may include a second set of R_{CT} values between the threshold value and a severity borderline value measured from a group of patients with moderate levels of cytokines. The third range of the calibration dataset may include a third set of R_{CT} values more than the severity borderline value measured from a group of patients with high levels of cytokines.

[0012] In an exemplary implementation, recording the EIS associated with the blood serum sample may include drawing a blood sample from a person, separating the blood serum sample from the blood sample by centrifuging the blood sample, forming a mixture by mixing the blood serum sample with an electrolyte solution comprising at least one metal ion, putting the mixture in contact with three-integrated electrodes of a biosensor, and recording an EIS from the mixture in contact with the three-integrated electrodes using an electrochemical stimulator-analyzer.

[0013] In an exemplary implementation, recording the EIS from the mixture in contact with the three-integrated electrodes may include recording the EIS of the blood serum sample in contact with the three-integrated electrodes at an AC potential amplitude between 5 mV and 20 mV while sweeping a frequency range between 0.01 Hz and 100 kHz.

[0014] In an exemplary implementation, putting the mixture in contact with the three-integrated electrodes of the biosensor may include dropping the mixture on a surface of three-integrated electrodes of the biosensor. In an exemplary embodiment, the three-integrated electrodes may include a working electrode including a circular-shaped sensing part, a counter electrode including a partially annular part placed around the circular-shaped sensing part of the working electrode, and a reference electrode placed adjacent to both the circular-shaped sensing part of the working electrode and the partially annular part of the counter electrode.

[0015] In an exemplary embodiment, the circular-shaped sensing part of the working electrode may include a bilayer of graphene/copper adhered on a first bilayer of gold/titanium (Au/Ti) deposited on a substrate. The working electrode may be configured to be an attachment site for the mixture. In an exemplary embodiment, the counter electrode may include a second bilayer of Au/Ti deposited on the substrate. The counter electrode may be configured to acquire the electrical response from the working electrode. In an exemplary embodiment, the reference electrode may include a third bilayer of Au/Ti deposited on the substrate. The reference

electrode may be configured to adjust a specific voltage between the working and the counter electrodes.

[0016] In an exemplary embodiment, the bilayer of graphene/copper may include a layer of graphene sheets with a thickness between 0.1 nm and 1 nm deposited on a layer of copper with a thickness between 1 μm and 30 μm. In an exemplary embodiment, each of the bilayers of Au/Ti may include a layer of gold with a thickness between 10 nm and 50 nm deposited on a layer of titanium with a thickness between 3 nm and 10 nm. In an exemplary embodiment, the substrate may include at least one of a glass substrate, a silicon substrate, a ceramic substrate, and combinations thereof.

[0017] In another aspect of the present disclosure, an exemplary method for identifying a status of COVID-19 infection of a person is described. The method may include drawing a blood sample from a person suspected to be infected with COVID-19 virus, separating a blood serum sample from the blood sample by centrifuging the blood sample, recording an electrochemical impedance spectroscopy (EIS) associated with the blood serum sample, calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS, and determining a COVID-19 infection status of the person based on the calculated R_{CT} if the calculated R_{CT} is equal to or more than a threshold value.

[0018] In an exemplary implementation, the method may further include generating a calibration dataset. In an exemplary implementation, generating the calibration dataset may include generating a set of R_{CT} values and generating a set of total cytokine mass values corresponding to the set of R_{CT} values. In an exemplary implementation, generating the set of R_{CT} values may include recording a set of EIS plots from a respective plurality of blood serum samples drawn from a corresponding plurality of persons and calculating the set of R_{CT} values respective to the set of recorded EIS plots by measuring a diameter of a semicircular curved part of each of the recorded EIS plots. In an exemplary implementation, generating the set of total cytokine mass values corresponding to the set of R_{CT} values may include measuring total cytokines mass of each of the plurality of blood serum samples utilizing a cytokine measurement assay.

[0019] In an exemplary implementation, determining the COVID-19 infection status of the person based on the calculated R_{CT} may include comparing the calculated R_{CT} with the set of R_{CT} values of the calibration dataset, detecting the person is in a healthy (normal) state if the calculated R_{CT} is within a first range of the calibration dataset, detecting the person is in a

moderately COVID-19 infected state if the calculated R_{CT} is within a second range of the calibration dataset, and detecting the person is in a severely infected state if the calculated R_{CT} is within a third range of the calibration dataset. In an exemplary embodiment, the first range of the calibration dataset may include a first set of R_{CT} values less than the threshold value. The first set of R_{CT} values may be measured from a group of healthy people. In an exemplary embodiment, the second range of the calibration dataset may include a second set of R_{CT} values between the threshold value and a severity borderline value. The second set of R_{CT} values may be measured from a group of moderately infected patients with COVID-19 who may have moderate levels of cytokines. In an exemplary embodiment, the third range of the calibration dataset may include a third set of R_{CT} values more than the severity borderline value. The third set of R_{CT} values may be measured from a group of severely infected patients with COVID-19 who may have high levels of cytokines.

[0020] In an exemplary implementation, determining the COVID-19 infection status of the person may include detecting the Covid-19 infection of the person if the calculated R_{CT} is equal to or more than 440 Ω. In an exemplary implementation, determining the COVID-19 infection status of the person may include detecting a healthy (normal) state for the person if the calculated R_{CT} is less than 340 Ω, detecting a moderate COVID-19 infection of the person if the calculated R_{CT} is between 440 Ω and 610 Ω, and detecting a severe COVID-19 infection of the person if the calculated R_{CT} is more than 715 Ω.

[0021] In an exemplary implementation, recording the EIS associated with the blood serum sample may include forming a mixture by mixing the blood serum sample with an electrolyte solution comprising a solution of at least one metal ion, putting the mixture in contact with three-integrated electrodes of a biosensor, and recording the EIS of the mixture in contact with the three-integrated electrodes using an electrochemical stimulator-analyzer.

[0022] In an exemplary implementation, recording the EIS from the mixture in contact with the three-integrated electrodes may include recording the EIS of the mixture in contact with the three-integrated electrodes at an AC potential amplitude between 5 mV and 20 mV while sweeping a frequency range between 0.01 Hz and 100 kHz. In an exemplary embodiment, the electrochemical stimulator-analyzer may include a potentiostat.

[0023] In an exemplary implementation, putting the mixture in contact with the three-integrated electrodes of the biosensor may include dropping the mixture on a surface of three-integrated electrodes of the biosensor. In an exemplary embodiment, the three-integrated

electrodes may include a working electrode including a circular-shaped sensing part, a counter electrode including a partially annular part placed around the circular-shaped sensing part of the working electrode, and a reference electrode placed adjacent to both the circular-shaped sensing part of the working electrode and the partially annular part of the counter electrode.

[0024] In an exemplary embodiment, the circular-shaped sensing part of the working electrode may include a bilayer of graphene/copper adhered on a first bilayer of gold/titanium (Au/Ti) deposited on a substrate. The working electrode may be configured to be an attachment site for the mixture. In an exemplary embodiment, the counter electrode may include a second bilayer of Au/Ti deposited on the substrate. The counter electrode may be configured to acquire an electrical response from the working electrode. In an exemplary embodiment, the reference electrode may include a third bilayer of Au/Ti deposited on the substrate. The reference electrode may be configured to adjust a specific voltage between the working and the counter electrodes.

[0025] In an exemplary embodiment, the bilayer of graphene/copper may include a layer of graphene sheets with a thickness between 0.1 nm and 1 nm deposited on a layer of copper with a thickness between 1 μm and 30 μm. In an exemplary embodiment, each of the bilayers of Au/Ti may include a layer of gold with a thickness between 10 nm and 50 nm deposited on a layer of titanium with a thickness between 3 nm and 10 nm. In an exemplary embodiment, the substrate may include at least one of a glass substrate, a silicon substrate, a ceramic substrate, and combinations thereof.

BRIEF DESCRIPTION OF THE DRAWINGS

[0026] The drawing figures depict one or more implementations in accord with the present teachings, by way of example only, not by way of limitation. In the figures, like reference numerals refer to the same or similar elements.

[0027] FIG. 1A shows an exemplary method for measuring a level of cytokines in a blood serum sample, consistent with one or more exemplary embodiments of the present disclosure.

[0028] FIG. 1B shows another exemplary implementation of an exemplary method for measuring a level of cytokines in a blood serum sample, consistent with one or more exemplary embodiments of the present disclosure.

[0029] FIG. 1C shows an exemplary method for identifying a status of COVID-19 infection of a person, consistent with one or more exemplary embodiments of the present disclosure.

[0030] FIG. 2 shows an exemplary method for recording the electrochemical impedance spectroscopy (EIS) associated with the blood serum sample, consistent with one or more exemplary embodiments of the present disclosure.

[0031] FIG. 3A shows a schematic view of an exemplary biosensor for recording an EIS from a blood serum sample, consistent with one or more exemplary embodiments of the present disclosure.

[0032] FIG. 3B shows a schematic view of an exemplary upper surface of an exemplary biosensor for recording an EIS from a blood serum sample, consistent with one or more exemplary embodiments of the present disclosure.

[0033] FIG. 4 shows a schematic view of an exemplary EIS response recorded from a mixture of a blood serum sample and an electrolyte solution put in contact with exemplary three-integrated electrodes, consistent with one or more exemplary embodiments of the present disclosure.

[0034] FIG. 5A shows comparing secreted levels of IL-6, TNF-α, and IFN-γ in blood serum of normal donors and patients with moderate and severe hypercytokinemia, consistent with one or more exemplary embodiments of the present disclosure.

[0035] FIG. 5B shows comparing total cytokine mass in blood serum of normal donors and patients with moderate and severe hypercytokinemia, consistent with one or more exemplary embodiments of the present disclosure.

[0036] FIG. 6 shows comparing secreted levels of immunoglobulin G (IGG), immunoglobulin M (IGM), and immunoglobulin A (IGA) antibodies in blood serum of normal donors and patients with moderate and severe hypercytokinemia, consistent with one or more exemplary embodiments of the present disclosure.

[0037] FIG. 7 shows an exemplary mass spectrum of blood serum of the three groups of normal, moderate and severe cytokine strom and an exemplary graph representing mass to charge (m/z) peaks and their abundance in the mass spectrum of the blood serums for the normal and hypercytokinemia groups, consistent with one or more exemplary embodiments of the present disclosure.

[0038] FIG. 8 shows a Raman spectrum of graphene/Cu of an exemplary working electrode of an exemplary fabricated biosensor, consistent with one or more exemplary embodiments of the present disclosure.

[0039] FIG. 9 shows exemplary X-ray diffraction analysis (XRD) analysis of graphene/Cu of an exemplary working electrode of an exemplary fabricated biosensor, consistent with one or more exemplary embodiments of the present disclosure.

[0040] FIG. 10 shows contact angle comparison of a normal blood serum sample over graphene, gold, and carbon substrates, consistent with one or more exemplary embodiments of the present disclosure.

[0041] FIG. 11A shows an exemplary Nyquist plot for 9 samples of blood serum for each group of normal, moderate, and severely infected patients, consistent with one or more exemplary embodiments of the present disclosure.

[0042] FIG. 11B shows comparing average values of charge transfer resistance (R_{CT}) for three groups of group of normal, moderate, and severely infected patients, consistent with one or more exemplary embodiments of the present disclosure.

[0043] FIG. 12A shows R_{CT} values for 63 individual normal persons and infected patients, consistent with one or more exemplary embodiments of the present disclosure.

[0044] FIG. 12B shows increase percentage of R_{CT} for COVID-19 patients in different progression stages relative to an avergae value of normal group, consistent with one or more exemplary embodiments of the present disclosure.

[0045] FIG. 13A shows confusion matrix of diagnostic results obtained by EIS analysis and ELISA method for 63 persons, consistent with one or more exemplary embodiments of the present disclosure.

[0046] FIG. 13B shows exemplary receiver operating characteristic curve (ROC) curves for comparing EIS analysis with ELISA technique on detection of cytokine storm, consistent with one or more exemplary embodiments of the present disclosure.

[0047] FIG. 14 shows a calibration curve based on measured R_{CT} values and total cytokines mass for three groups of normal, moderate, and severe cases, consistent with one or more exemplary embodiments of the present disclosure.

[0048] FIG. 15 shows exemplary CT scan images of three exemplary persons from three groups of normal, moderate, and severe cases, consistent with one or more exemplary embodiments of the present disclosure.

[0049] FIG. 16A shows confusion matrix of diagnostic results obtained by EIS analysis and RT-PCR method for 63 persons, consistent with one or more exemplary embodiments of the present disclosure.

[0050] FIG. 16B shows confusion matrix of diagnostic results obtained by EIS analysis and CT scan method for 63 persons, consistent with one or more exemplary embodiments of the present disclosure.

[0051] FIG. 16C shows ROC curves for comparing EIS analysis with RT-PCR and CT scan techniques on detection of cytokine storm, consistent with one or more exemplary embodiments of the present disclosure.

DESCRIPTION OF EMBODIMENTS

[0052] In the following detailed description, numerous specific details are set forth by way of examples in order to provide a thorough understanding of the relevant teachings. However, it should be apparent that the present teachings may be practiced without such details. In other instances, well known methods, procedures, components, and/or circuitry have been described at a relatively high-level, without detail, in order to avoid unnecessarily obscuring aspects of the present teachings.

[0053] Blood serum consists of many proteins including antibodies, cytokines, hormones, etc. with different molecular weights and dielectric properties. In chronic inflammatory diseases with pathogens such as SARS-CoV-2 virus, many inflammatory signals are produced and cytokine release into the blood serum becomes out of control. Such a phenomenon is so-called cytokine storm and is the most prominent feature of the disease.

[0054] Concurrent with the pandemic announcement of SARS-CoV-2 infection by World Health Organization (WHO), a variety of reports were published affirming the cytokine storm having the most significant impact on mortality of infected patients with COVID-19 virus. Although cytokine storm arises in many other types of diseases such as sepsis, Flu, Graft versus host disease (GvHD), etc., currently the best available cohort with significant level of cytokine storm for developing new technologies is COVID-19 infected patients during this pandemic. Hence, cytokine storm as an evidenced consequence in most of the COVID-19 patients could offer a promising opportunity to use blood as a disease progression marker of COVID-19 infection.

[0055] Herein, a label-free electrochemical impedimetric approach for monitoring a level of cytokines and detecting cytokine storm in an unprocessed blood sample is disclosed. In an exemplary implementation, the disclosed method may be utilized for real-time monitoring amounts of cytokines in a person's bloodstream, and subsequently diagnosing a cytokine storm

as a reliable indicator of an inflammatory disease, such as COVID-19 infection. Herein, an electrochemical impedimetric biosensor, a system including the electrochemical impedimetric biosensor, and a method utilizing the electrochemical impedimetric biosensor and the system is disclosed for simply, real-time and accurately detecting cytokines level in a blood sample, and thereafter detecting an inflammatory disease infectious in the blood sample.

[0056] Exemplary biosensor, system and method disclosed here, utilizes electrochemical impedance spectroscopy (EIS) by measuring electrical impedance of cytokines of a blood sample in an electrolyte medium. Charge transfer resistance (R_{CT}) is a comparing parameter between studied groups and a reliable criterion, which may be affected by a presence of dielectric materials in a media. Hence, impact of a glycoprotein cytokine on R_{CT} elevation due to its weight may be expected especially for a blood serum drawn from a COVID-19 patient with a cytokine storm indication.

[0057] EIS may be utilized for quantifying overall immune activity of patients. Herein, a biosensing method for non-direct detection of cytokine storm in an inflammatory disease infected patients (e.g., COVID-19 infected patients) and their screening according to a severity of their immune activity based on a recorded EIS of cytokine storm in an unprocessed blood serum sample is disclosed. To improve electrochemical interface between biological macromolecules (such as cytokines) and electrodes, nanomaterials may be the best choice for an active surface of an electrode (e.g., a working electrode) of an electrochemical impedimetric biosensor. Herein, graphene sheets deposited on copper substrate due to its better charge mobility and electrical conductivity may be utilized as a working electrode of an electrochemical impedimetric biosensor. Moreover, graphene may apply much better site binding with polarizable bio-agents, such as cytokines in comparison with other possible usable materials, such as gold and carbon for coating on an exemplary working electrode. In an exemplary embodiment, a blood serum sample may have more tendency to a graphene layer on an electrode in comparison with a layer of gold or carbon. Exemplary biosensor, system and method may show a meaningful correlation between an immune hyperactivation in COVID-19 patients (induced by acute elevation of total mass of cytokines in blood serum) and an increased R_{CT}. In addition, EIS results obtained by exemplary method may be compared and approved with a cytokine mass measurement technique, such as enzyme-linked immunosorbent assay (ELISA) analysis. Moreover, exemplary biosensor may be utilized for indirect detection of patients suffering from the COVID-19 disease and results may be

validated by conventional Reverse transcription polymerase chain reaction (RT-PCR) and computed tomography (CT) scan techniques on normal and infected cases.

[0058] FIG. 1A shows exemplary method **100** for measuring a level of cytokines in a blood serum sample, consistent with one or more exemplary embodiments of the present disclosure. Exemplary method **100** may include recording an electrochemical impedance spectroscopy (EIS) associated with the blood serum sample (step **102**), calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS (step **104**), and determining the level of cytokines in the blood serum sample based on the calculated R_{CT} by looking up the calculated R_{CT} in a calibration dataset comprising a set of cytokine mass values versus a set of R_{CT} values (step **106**).

[0059] In detail, step **102** may include recording an electrochemical impedance spectroscopy (EIS) associated with the blood serum sample. In an exemplary implementation, recording the EIS associated with the blood serum sample may include recording an EIS of the blood serum sample. **FIG. 2** shows an exemplary method **200** for recording the EIS associated with the blood serum sample (step **102**), consistent with one or more exemplary embodiments of the present disclosure. In an exemplary implementation, recording the EIS associated with the blood serum sample (step **102**) may include drawing a blood sample from a person (step **202**), separating the blood serum sample from the blood sample by centrifuging the blood sample (step **204**), forming a mixture by mixing the blood serum sample with an electrolyte solution (step **206**), putting the mixture in contact with three-integrated electrodes of a biosensor (step **208**), and recording an EIS from the mixture in contact with the three-integrated electrodes using an electrochemical stimulator-analyzer (step **210**).

[0060] In detail, step **202** may include drawing a blood sample from a person. In an exemplary embodiment, exemplary method **100** may be utilized for measuring a level of cytokines in a blood serum sample extracted from a blood sample drawn from a person suspected to be infected with COVID-19 virus. In an exemplary implementation, drawing the blood sample from the person (step **202**) may include drawing a blood sample from a person suspected to be infected with COVID-19 virus. In an exemplary implementation, drawing the blood sample from the person may include drawing a blood sample between about 200 μL and about 1 cc blood sample from the person.

[0061] In an exemplary implementation, separating the blood serum sample from the blood sample (step **204**) may include forming the blood serum sample in a separated phase from the

blood sample by centrifuging the blood sample and extracting the blood serum sample from a top part of the centrifuged blood sample. In an exemplary implementation, centrifuging the blood sample may include centrifuging the blood sample at a speed of about 3000 rpm for about 10 minutes.

[0062] In an exemplary implementation, forming a mixture by mixing the blood serum sample with an electrolyte solution (step **206**) may include mixing the blood serum sample with a solution containing one or more metal ions. The one or more metal ions may be used as a charge carrier agent for EIS measurements and recordings. In an exemplary embodiment, the solution containing one or more metal ions may include a solution of potassium ferricyanide ($K_3Fe(CN)_6$ and/or $K_4Fe(CN)_6$) with a concentration of about 5 mM of the potassium ferricyanide in water.

[0063] In an exemplary implementation, putting the mixture in contact with three-integrated electrodes of a biosensor (step **208**) may include dropping the mixture on a surface of the three-integrated electrodes of the biosensor. **FIG. 3A** shows a schematic view of an exemplary biosensor **300** for recording an EIS from a blood serum sample, consistent with one or more exemplary embodiments of the present disclosure. In an exemplary embodiment, biosensor **300** may include three-integrated electrodes **302**, **304**, and **306** patterned and fabricated on exemplary substrate **308**.

[0064] In an exemplary embodiment, substrate **308** may include at least one of a glass substrate, a silicon substrate, a ceramic substrate, and combinations thereof. In an exemplary embodiment, substrate **308** may have a thickness between about 0.5 mm and about 5 mm.

[0065] In an exemplary embodiment, three-integrated electrodes **302**, **304**, and **306** may include working electrode **302**, counter electrode **304**, and reference electrode **306**. **FIG. 3B** shows a schematic view of upper surface **320** of biosensor **300** shown in **FIG. 3A**, consistent with one or more exemplary embodiments of the present disclosure.

[0066] Referring to **FIG. 3B**, working electrode **302** may include exemplary circular-shaped sensing part **322** and a first distal end **324**. In an exemplary embodiment, circular-shaped sensing part **322** of working electrode **302** may be configured to be an attachment site for a sample, such as the mixture of the blood serum sample and the electrolyte solution. In an exemplary embodiment, the first distal end **324** may be configured to be connected to an electrochemical stimulator-analyzer, for example, a potentiostat device.

[0067] In an exemplary embodiment, working electrode **302** may include a first bilayer of gold/titanium (Au/Ti) deposited on upper surface **320** of biosensor **300**. In an exemplary embodiment, the first bilayer of Au/Ti may include a layer of Ti with a thickness between about 3 nm and about 10 nm deposited on upper surface **320** of biosensor **300** and a layer of Au with a thickness between about 10 nm and about 50 nm deposited on the layer of Ti. Accordingly, the first distal end **324** may include the first bilayer of Au/Ti deposited on upper surface **320** of biosensor **300**. In an exemplary embodiment, circular-shaped sensing part **322** may further include a layer of an electrical conductive hydrophilic material adhered on the first bilayer of Au/Ti deposited on upper surface **320** of biosensor **300**. In an exemplary embodiment, circular-shaped sensing part **322** may include a bilayer of graphene/copper adhered on the first bilayer of Au/Ti. In an exemplary embodiment, the bilayer of graphene/copper may include a monolayer of graphene including a layer of graphene sheets with a thickness between about 0.1 nm and about 1 nm deposited on a layer of copper (Cu) with a thickness between about 1 μm and about 30 μm. In an exemplary embodiment, the first bilayer of Au/Ti may be deposited on upper surface **320** of biosensor **300** utilizing a sputtering technique.

[0068] Referring again to **FIG. 3B**, counter electrode **304** may include partially annular part **326** and a second distal end **328**. In an exemplary embodiment, partially annular part **326** of counter electrode **304** may be placed around circular-shaped sensing part **322** of working electrode **302**. In an exemplary embodiment, partially annular part **326** of counter electrode **304** may be configured to acquire an electrical response from working electrode **302**. In an exemplary embodiment, the second distal end **328** may be configured to be connected to the electrochemical stimulator-analyzer, for example, a potentiostat device.

[0069] In an exemplary embodiment, counter electrode **304** may include a second bilayer of Au/Ti deposited on upper surface **320** of biosensor **300**. In an exemplary embodiment, the second bilayer of Au/Ti may include a layer of Ti with a thickness between about 3 nm and about 10 nm deposited on upper surface **320** of biosensor **300** and a layer of Au with a thickness between about 10 nm and about 50 nm deposited on the layer of Ti. In an exemplary embodiment, the second bilayer of Au/Ti may be deposited on upper surface **320** of biosensor **300** utilizing the sputtering technique.

[0070] With more reference to **FIG. 3B**, reference electrode **306** may include active part **330** and a third distal end **332**. In an exemplary embodiment, active part **330** of reference electrode **306** may be placed adjacent to both working electrode **302** and counter electrode **304**. In an

exemplary embodiment, active part **330** of reference electrode **306** may be configured to adjust a specific voltage between working electrode **302** and counter electrode **304**. In an exemplary embodiment, the third distal end **332** may be configured to be connected to the electrochemical stimulator-analyzer, for example, a potentiostat device.

[0071] In an exemplary embodiment, reference electrode **306** may include a third bilayer of Au/Ti deposited on upper surface **320** of biosensor **300**. In an exemplary embodiment, the third bilayer of Au/Ti may include a layer of Ti with a thickness between about 3 nm and about 10 nm deposited on upper surface **320** of biosensor **300** and a layer of Au with a thickness between about 10 nm and about 50 nm deposited on the layer of Ti. In an exemplary embodiment, the third bilayer of Au/Ti may be deposited on upper surface **320** of biosensor **300** utilizing the sputtering technique.

[0072] In further detail with respect to **FIG. 3A**, in an exemplary embodiment, biosensor **300** may further include exemplary liquid holder **310**. In an exemplary embodiment, liquid holder **310** may be configured to hold a sample, such as the mixture of the blood serum sample and the electrolyte solution there inside on surface of three-integrated electrodes **302**, **304**, and **306**. In an exemplary embodiment, liquid holder **310** may be placed around circular-shaped sensing part **322** of working electrode **302**, partially annular part **326** of counter electrode **304**, and active part **330** of reference electrode **306**.

[0073] Referring back to **FIG. 2**, step **210** may include recording an EIS from the mixture in contact with three-integrated electrodes **302**, **304**, and **306** using an electrochemical stimulator-analyzer. In an exemplary implementation, recording the EIS from the mixture in contact with three-integrated electrodes **302**, **304**, and **306** may include recording the EIS of the mixture in contact with three-integrated electrodes **302**, **304**, and **306** at a AC potential amplitude between about 5 mV and 2 about 0 mV while sweeping a frequency range between about 0.01 Hz and about 100 kHz. In an exemplary embodiment, the electrochemical stimulator-analyzer may include a potentiostat device.

[0074] In an exemplary implementation, recording the EIS from the mixture in contact with three-integrated electrodes **302**, **304**, and **306** (step **210**) may include plotting a Nyquist response for the blood serum sample. **FIG. 4** shows a schematic view of exemplary EIS response **400** recorded from a mixture of a blood serum sample and an electrolyte solution put in contact with three-integrated electrodes **302**, **304**, and **306**, consistent with one or more exemplary embodiments of the present disclosure. Exemplary EIS response **400** may include

exemplary Nyquist plot **400** with a semicircular curve shape including a set of recorded imaginary part of impedance (Z^{II} (Ω)) versus a set of recorded real part of impedance (Z^{I} (Ω)).

[0075] Referring back to **FIG. 1A**, step **104** may include calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS obtained from step **102**. In an exemplary implementation, calculating the R_{CT} of the recorded EIS may include measuring a diameter of a semicircular curve of exemplary EIS **400** as shown in **FIG. 4**.

[0076] Moreover, step **106** of exemplary method **100** may include determining the level of total cytokines in the blood serum sample based on the calculated R_{CT}. In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} may include looking up the calculated R_{CT} in a calibration dataset comprising a set of total cytokine mass values respective to a set of R_{CT} values.

[0077] In an exemplary implementation, exemplary method **100** may further include generating the calibration dataset. **FIG. 1B** shows another exemplary implementation of exemplary method **110** for measuring a level of cytokines in a blood serum sample, consistent with one or more exemplary embodiments of the present disclosure. Exemplary method **110** may include generating a calibration dataset including a set of total cytokines mass values respective to a set of R_{CT} values (step **101**), recording an electrochemical impedance spectroscopy (EIS) associated with the blood serum sample (step **102**), calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS (step **104**), and determining the level of cytokines in the blood serum sample based on the calculated R_{CT} by looking up the calculated R_{CT} in the calibration dataset (step **106**).

[0078] In detail, step **101** may include generating a calibration dataset including a set of total cytokines mass values respective to a set of R_{CT} values. In an exemplary implementation, generating the calibration dataset (step **101**) may include generating the set of R_{CT} values and generating the set of total cytokines mass values corresponding to the set of R_{CT} values.

[0079] In an exemplary implementation, generating the set of R_{CT} values may include recording a set of electrochemical impedance spectroscopy (EIS) plots from a respective plurality of blood serum samples drawn from a corresponding plurality of persons and calculating the set of R_{CT} values respective to the set of recorded EIS plots by measuring a diameter of a semicircular curved part of each of the recorded EIS plots. In an exemplary

implementation, recording the set of EIS plots from the respective plurality of blood serum samples may be carried out similar to step **102** of exemplary method **100** described in detail hereinabove. In an exemplary implementation, calculating the set of R_{CT} values respective to the set of recorded EIS plots may be carried out similar to step **104** of exemplary method **100** described in detail hereinabove.

[0080] In an exemplary implementation, generating the set of total cytokines mass values corresponding to the set of R_{CT} values may include measuring total cytokines mass of each of the plurality of blood serum samples utilizing a cytokine measurement assay. In an exemplary embodiment, the cytokine measurement assay may include at least one of enzyme-linked immunosorbent assay (ELISA), Matrix-assisted laser desorption/ionization mass spectrometry (MALDI MS), and combinations thereof.

[0081] In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106**) may include looking up the calculated R_{CT} in the calibration dataset by comparing the calculated R_{CT} with R_{CT} values in the calibration dataset and obtaining a respective total cytokines mass value corresponding to the calculated R_{CT} in the calibration dataset.

[0082] In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106**) may further include detecting or diagnosing an infection of a chronic inflammatory disease (CID) such as COVID-19 infection. In an exemplary implementation, diagnosing the infection of the chronic inflammatory disease may include diagnosing the infection of the chronic inflammatory disease in a person associated with the blood serum sample whom the blood sample may be drawn from her. In an exemplary implementation, diagnosing the infection of the chronic inflammatory disease in the person associated with the blood serum sample may include detecting the infection of the chronic inflammatory disease for the person responsive to the calculated R_{CT} being more than a threshold value. In an exemplary embodiment, the chronic inflammatory disease (CID) may include at least one of COVID-19 disease, SARS-CoV-2 infection, flu, Graft versus host disease (GvHD), and combinations thereof. In an exemplary embodiment, the threshold value may include a R_{CT} value of about 340 Ω for COVID-19 infection. In an exemplary embodiment, the threshold value may include a R_{CT} value of about 440 Ω for COVID-19 infection.

[0083] In an exemplary implementation, generating the calibration dataset (step **101**) may include generating a calibration dataset from a plurality of persons including three groups of persons based on a grade of infection of the chronic inflammatory disease. The plurality of persons may be categorized in three groups, including a first group including healthy (normal) persons, a second group including moderately infected persons, and a third group including severely infected persons. The healthy (normal) persons may include a plurality of persons with no symptoms and no clinical diagnosis of an infection of the chronic inflammatory disease. The healthy (normal) persons may include a plurality of persons with total cytokines mass values of their blood serum samples less than a reference value. The second group may include a plurality of moderately infected persons with the chronic inflammatory disease that may be diagnosed and categorized by clinical diagnosis methods and assays. The third group may include a plurality of severely infected persons with the chronic inflammatory disease that may be diagnosed and categorized by clinical diagnosis methods and assays.

[0084] In an exemplary implementation, people may be categorized in three groups of persons including the first group including healthy (normal) persons, the second group including moderately infected persons, and the third group including severely infected persons based on clinical judgments and assays. The three groups of persons may be addressed with known status of severity of the chronic inflammatory disease based on clinical results and judgments. In an exemplary implementation, a plurality of persons may be clinically classified into a first group of healthy persons, a second group of moderately COVID-19 infected patients, and a third group of severely COVID-19 infected patients. The calibration dataset may be generated in step **101** for the first group of healthy persons, the second group of moderately COVID-19 infected patients, and the third group of severely COVID-19 infected patients.

[0085] In an exemplary implementation, generating the calibration dataset (step **101**) may further include setting a severity borderline value between a set of R_{CT} values associated with the second group and a set of R_{CT} values associated with the third group based on clinical diagnosis and symptoms. The severity borderline value may include a highest value of R_{CT} for a set of R_{CT} values associated with the second group and also a lowest value of R_{CT} for a set of R_{CT} values associated with the third group. In an exemplary embodiment, the severity borderline value may include a R_{CT} value of about 610 Ω for COVID-19 infection. In an exemplary embodiment, the severity borderline value may include a R_{CT} value of about 715 Ω for COVID-19 infection.

[0086] Accordingly, in an exemplary embodiment, the generated calibration dataset may include a first set of R_{CT} values versus a first set of total cytokines mass values, a second set of R_{CT} values versus a second set of total cytokines mass values, and a third set of R_{CT} values versus a third set of total cytokines mass values. The first set of R_{CT} values may include R_{CT} values being less than the threshold value representing no cytokine storm and no infection of the chronic inflammatory disease in the respective first group of persons. The second set of R_{CT} values may include R_{CT} values between the threshold value and the severity borderline value representing a moderate cytokine storm and moderate infection of the chronic inflammatory disease in the respective second group of persons. The third set of R_{CT} values may include R_{CT} values more than the severity borderline value representing a severe cytokine storm and severe infection of the chronic inflammatory disease in the respective third group of persons.

[0087] In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106** of methods **100** and **110**) may further include grading a severity of the chronic inflammatory disease infection of the person. In an exemplary implementation, grading the severity of the chronic inflammatory disease infection of the person may include one of detecting the person being in a healthy (normal) state if the calculated R_{CT} is within a first range of the calibration dataset including the first set of R_{CT} values less than the threshold value, detecting the person being in a moderately infected state if the calculated R_{CT} is within a second range of the calibration dataset including the second set of R_{CT} values between the threshold value and the severity borderline value, and detecting the blood serum sample being in a severely infected state if the calculated R_{CT} is within a third range of the calibration dataset including a third set of R_{CT} values more than the severity borderline value.

[0088] In an exemplary implementation, generating the calibration dataset (step **101**) may include generating the set of R_{CT} values and generating the set of total cytokines mass values corresponding to the set of R_{CT} values for a plurality of people with known status of a chronic inflammatory disease (CID) infection, for example, a plurality of people with known status of COVID-19 infection. In an exemplary embodiment, the plurality of people with known status of COVID-19 infection may include a first group of healthy persons, a second group of moderately COVID-19 infected patients, and a third group of severely COVID-19 infected patients.

[0089] In an exemplary embodiment, the calibration dataset may include three sets of data, including a first set, a second set, and a third set. The first set may include a first set of calculated R_{CT} values for the first group of healthy persons respective to a first set of total cytokines mass values measured for the first group of healthy persons. In an exemplary embodiment, the first set of calculated R_{CT} values may include R_{CT} values less than a COVID-19-threshold value, and the first set of total cytokines mass values may include total cytokines mass values less than a COVID-19-reference value. The COVID-19-reference value may include an amount of total cytokines mass representing a cytokine storm. In an exemplary embodiment, the second set may include a second set of calculated R_{CT} values for the second group of moderately COVID-19 infected patients respective to a second set of total cytokines mass values measured for the second group of moderately COVID-19 infected patients. In an exemplary embodiment, the second set of calculated R_{CT} values may include R_{CT} values more than the COVID-19-threshold value and less than a severity COVID-19-borderline R_{CT} value. Correspondingly, the second set of total cytokines mass values may include total cytokines mass values between the COVID-19-reference value and a severity COVID-19-borderline total cytokines value. In an exemplary embodiment, the third set may include a third set of calculated R_{CT} values for the third group of severely COVID-19 infected patients respective to a third set of total cytokines mass values measured for the third group of severely COVID-19 infected patients. In an exemplary embodiment, the third set of calculated R_{CT} values may include R_{CT} values more than the severity COVID-19-borderline R_{CT} value. Correspondingly, the third set of total cytokines mass values may include total cytokines mass values more than the severity COVID-19-borderline total cytokines value.

[0090] In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106**) may include detecting COVID-19 infection status of a person suspected to be infected with COVID-19. In an exemplary implementation, detecting the COVID-19 infection status of the person may include detecting the Covid-19 infection of the person if the calculated R_{CT} is equal to or more than about 440 Ω. In an exemplary implementation, detecting the COVID-19 infection status of the person may include detecting a healthy (normal) state for the person if the calculated R_{CT} is less than about 340 Ω, detecting a moderate COVID-19 infection of the person if the calculated R_{CT} is between about 440 Ω and about 610 Ω, and detecting a severe COVID-19 infection of the person if the calculated R_{CT} is more than about 715 Ω.

[0091] It should be noted that all COVID-19 infected patients may suffer from a cytokine storm with total cytokines mass values more than the COVID-19-reference value. Moreover, severely COVID-19 infected patients may suffer from an intensive cytokine storm with total cytokines mass values more than the severity COVID-19-borderline total cytokines value. In an exemplary embodiment, the calculated R_{CT} values may increase with increasing total cytokines mass values representing a COVID-19 infection and in some cases, a severity of COVID-19 infection. In an exemplary embodiment, the COVID-19-threshold value may include about 440 Ω corresponding to the COVID-19-reference of about 7400 kDa, representing a border between the first group of healthy persons, and the second and third groups of COVID-19 infected patients. In an exemplary embodiment, the severity COVID-19-borderline R_{CT} value may include about 715 Ω corresponding to the COVID-19-reference of about 11600 kDa, representing a border between the second group of moderately COVID-19 infected patients and the third group of severely COVID-19 infected patients.

[0092] In an exemplary implementation, generating the calibration dataset (step **101**) may further include generating (i.e., fitting or estimating) a mathematical relationship between the set of R_{CT} values and the set of respective total cytokines mass values. In such implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106**) may include calculating an amount of total cytokines in the blood serum sample utilizing the mathematical relationship between the set of R_{CT} values and the set of respective total cytokines mass values.

[0093] In an exemplary embodiment, the relationship between the set of R_{CT} values and the set of respective total cytokines mass values may be defined by:

[0094] $$TCM = (R_{CT} - 283.75)/0.0338 \quad \text{(Eq. 1)}$$

[0095] Where TCM is total cytokines mass of the blood serum sample (kDa), and R_{CT} is the calculated R_{CT} (Ω). In an exemplary embodiment, a cytokine storm for the person, and subsequently, a COVID-19 infection of the person may be detected in step **106** if the calculated TCM is equal to or more than a pre-determined value of total cytokines mass, for example, the COVID-19-reference of about 3000 kDa.

[0096] In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106** of methods **100** and **110**) may further include diagnosing COVID-19 infection of the person if the calculated R_{CT} is equal to or more than the COVID-19-threshold value.

[0097] Furthermore, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106** of methods **100** and **110**) may further include grading a severity of COVID-19 infection of the person based on the calculated R_{CT}. In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106** of methods **100** and **110**) may include comparing the calculated R_{CT} with the set of R_{CT} values of the calibration dataset, detecting the person is in a healthy (normal) state if the calculated R_{CT} is within the first set of the calibration dataset, detecting the person is in a moderately COVID-19 infected state if the calculated R_{CT} is within the second set of the calibration dataset, and detecting the person is in a severely infected state if the calculated R_{CT} is within the third set of the calibration dataset.

[0098] According to exemplary embodiments described hereinabove, methods **100** and **110** may be utilized for diagnosing and grading COVID-19 infection of a person. **FIG. 1C** shows exemplary method **120** for identifying a status of COVID-19 infection of a person, consistent with one or more exemplary embodiments of the present disclosure. Exemplary method **120** may include generating a calibration dataset including a set of total cytokines mass values respective to a set of R_{CT} values (step **122** similar to step **101**), recording an electrochemical impedance spectroscopy (EIS) of a blood serum sample of a person suspected to be infected with COVID-19 (step **124** similar to step **102**), calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS (step **126** similar to step **104**), and detecting a COVID-19 infection of the person responsive to the calculated R_{CT} being equal to or more than a threshold value (step **128** similar to step **106**).

[0099] In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} (step **106** of methods **100** and **110**) may further include detecting a cytokine storm in the blood serum sample if the calculated R_{CT} is more than the threshold value. In an exemplary implementation, determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} may include detecting the cytokine storm in the blood serum sample if the calculated R_{CT} is more than about 440 Ω corresponding to a total cytokines mass of more than about 7400 kDa. In an exemplary implementation, detecting the cytokine storm in the blood serum sample may include detecting an infection of a chronic inflammatory disease of a person associated with the blood serum sample whom the blood sample may be drawn from her. In an exemplary implementation, detecting the cytokine storm in the blood serum sample may include detecting a moderate infection of the chronic

inflammatory disease of the person if the calculated R_{CT} is between about 440 Ω and about 610 Ω. In an exemplary implementation, detecting the cytokine storm in the blood serum sample may include detecting a severe infection of the chronic inflammatory disease of the person if the calculated R_{CT} is more than about 715 Ω.

[00100] **EXAMPLE 1: Cytokine analysis of blood serum samples by ELISA method**

[00101] In this example, blood samples were collected from normal donators, moderately and severely infected patients (with signs of inflammatory diseases such as COVID-19) in three individual cohorts which were selected based on clinical judgments of the patients. An adequate volume of blood was sampled from normal donators and the patients and after serum isolation, cytokine measurement was performed by the ELISA method.

[00102] Although in hypercytokinemia many cytokines may be secreted into the blood, three key cytokines named Interleukin-6 (IL-6), Tumor Necrosis Factor-α (TNF-α) and Interferon-γ (IFN-γ) play more important role in cytokine storm relative to the others and show the most elevated levels in blood serum of the patients. On this basis, ELISA procedure was carried out to measure a number of produced cytokines by the immune system of each person.

[00103] In brief, known concentrations of recombinant human IL-6, TNF-α or INF-γ and experimental blood samples were added and incubated in polystyrene microtiter plates coated with an antibody against the appointed cytokine, followed by incubation with an enzyme-linked polyclonal antibody directed to the cytokine. Next, a substrate solution for the enzyme was added, and the color development was stopped by adding a solution of 2 N H_2SO_4. The absorbance was measured with a microtiter plate spectrophotometer. An amount of IL-6, TNF-α, or INF-γ present in each blood sample was determined from a standard curve generated in each assay and expressed as picograms per milliliter. The sensitivity of the enzyme-linked immunosorbent assay for IL-6, TNF-α and INF-γ was 0.70 pg/ml, 1.88 pg/mL, and 8 pg/mL, respectively. The reproducibility of all measurements was within about 10%.

[00104] **FIG. 5A** shows comparing secreted levels of IL-6 (graph **502**), TNF-α (graph **504**), and IFN-γ (graph **506**) in the blood serum of normal donors and patients with moderate and severe hypercytokinemia, consistent with one or more exemplary embodiments of the present disclosure. It may be observed that the expressed levels of IL-6, TNF-α and IFN-γ cytokines in the infected patients were markedly higher than the normal group. Furthermore, analyzing the cytokines for the two groups of moderately and severely infected patients demonstrates a

correlated secretion pattern of those three cytokines with the severity of the disease. Here, hypercytokinemia alone means sum of the moderate and severe cases.

[00105] For more clarification, total molecular weight for all three types of cytokines were calculated for all patients including both normal and infected cases. To do this, for each person the amount of the expressed cytokine was multiplied by its molecular mass (IL-6: 23 kDa, TNF-α: 25 kDa and IFN-γ: 19 kDa) and summed for the all three cytokines. **FIG. 5B** shows comparing total cytokine mass in the blood serum of normal donors and patients with moderate and severe hypercytokinemia, consistent with one or more exemplary embodiments of the present disclosure. As presented in this figure, the total molecular weight of the mentioned cytokines for the suspicious COVID-19 patients was meaningfully increased as the stage of the disease becomes inferior.

[00106] Aside from the cytokine analysis, immunoglobulins G, M and A were also investigated for the normal and infected groups. **FIG. 6** shows comparing secreted levels of immunoglobulin G (IGG) (graph **602**), immunoglobulin M (IGM) (graph **604**), and immunoglobulin A (IGA) (graph **606**) antibodies in the blood serum of normal donors and patients with moderate and severe hypercytokinemia, consistent with one or more exemplary embodiments of the present disclosure. As may be seen, no meaningful correlation in the infected COVID-19 patients compared to the normal donors could be detected in the case of the IGG, IGM and IGA antibodies, while significant difference existed in the case of cytokines.

[00107] Moreover, mass spectroscopy (MS) analysis of plasma blood samples were analyzed by Matrix-assisted laser desorption/ionization (MALDI)-TOF mass spectrometry. For MS analysis, after diluting each sample up to 5 times and then ZipTip performing, the sample was spotted on MALDI plate and was mixed with alphacyano-4-hydroxy-cinnamic acid (CHCA) in 50% CAN containing 0.1% TFA (with 1:2 ratio) as a matrix solution. Then, the sample was air dried and analyzed in linear positive mode. **FIG. 7** shows mass spectrum **702** of the blood serum of the three groups of normal, moderate and severe cytokine strom and graph **704** representing mass to charge (m/z) peaks and their abundance in the mass spectrum of the blood serums for the normal and hypercytokinemia groups, consistent with one or more exemplary embodiments of the present disclosure. The obtained mass spectrum revealed a total increase in intensity of mass peaks for the patients with severe symptoms and cytokine storm. Moreover, in some m/z such as 22474 which is close to the molecular weight of three mentioned cytokines, a meaningful increase may be observed in the mass peaks of normal samples (4) to severe

patients (39). This might be in a well correlation with cytokine storm based tracing of people suspicious to COVID-19.

[00108] **EXAMPLE 2: Fabrication and characterization of an exemplary biosensor**

[00109] In this example, an exemplary biosensor similar to biosensor **300** was fabricated. At first, a bilayer of Au/Ti (30/5 nm) was deposited on top of a glass slide as a substrate using a sputtering system. Then, a circular pattern of three-electrode array of the biosensor was transferred to the substrate using a standard lithography method. Thereafter, a layer of graphene/Cu was placed and adhered on top of a working electrode of the three-electrode array. A sensing part of the three-electrode array was sealed by a holder; allowing for dropping a sample onto the sensing part inside the holder. The biosensor included an integrated three-electrode system including a circular working electrode with a diameter of about 5 mm, a counter electrode surrounding the circular working electrode and a reference electrode near both the circular working electrode and the counter electrode. A distance between working, reference and counter electrodes was about 1 mm. Sensing material of reference and counter electrodes were gold, and sensing material of the working electrode was a graphene monolayer on top of a copper substrate.

[00110] **FIG. 8** shows a Raman spectrum of graphene/Cu of the working electrode of exemplary fabricated biosensor, consistent with one or more exemplary embodiments of the present disclosure. The Raman spectra shows two considerable peaks at 1562 cm^{-1} and 2712 cm^{-1} related to G-band (in-plane vibration mode) and 2D-band (double resonance scattering of inter-valley) of graphene layer, respectively. These peaks confirm the presence of graphene layer on Cu. As observed in Raman spectrum, there is not significant D-band peak in diagram, which illustrates approximately no-defects on graphene.

[00111] Formation of graphene/Cu was also investigated by X-ray diffraction (XRD) technique. **FIG. 9** shows XRD analysis of graphene/Cu of the working electrode of exemplary fabricated biosensor, consistent with one or more exemplary embodiments of the present disclosure. A sharp diffraction peak at 59.2° is related to Cu with plane of (200). Also, a tiny peak at 42.4° corresponds to the graphene layer.

[00112] It should be noted that a material that may be chosen for depositing on surface of electrodes to interact with a blood serum sample, may play a significant role in accuracy and validity of electrical responses acquired from exemplary fabricated biosensor. Accordingly, three types of electrode materials including gold (Au), carbon, and graphene were investigated

as candidates for EIS analysis. Adhesion and spread of a blood serum sample on the selected substrates were evaluated by contact angle method. As the water composes the most part of the blood serum sample, the blood serum sample may have more tendency to hydrophilic substrates. **FIG. 10** shows contact angle comparison of a normal blood serum sample over graphene, gold and carbon substrates, consistent with one or more exemplary embodiments of the present disclosure. It may be observed that the maximum contact angle was attributed to the carbon surface with about 118° and the minimum was for graphene electrode with angle of about 69°. The results may demonstrate that the blood serum is mostly attracted to the graphene-coated electrode rather than the Au and carbon.

[00113] **EXAMPLE 3: Derivation of R_{CT} in human blood serum samples**

[00114] Patients suspicious to be infected by SARS-CoV-2 virus was selected based on the severity of the disease. The presence and severity of the disease was reported based on the symptoms and clinical exams of the patients presented in latest guidelines for clinical management of COVID-19 patients published by WHO. The serum samples were prepared from the residues of patients' and candidates' blood who did serological test to check themselves. When the blood was sampled and coagulated, the serum was isolated from the whole blood by centrifugation. Human blood samples from normal and suspicious patients were collected and then divided into clot activator tubes. For serum isolation, the clot activator tubes were maintained at 37°C for 15 minutes and then centrifuged for about 10 min at about 3000 rpm. Serum sample as a top part of the centrifuged liquid samples was extracted for EIS measurements. For EIS analysis, the serum was mixed in a ratio of about 3:1 with an electrolyte solution (0.1 M KCl, 5 mM $K_3[Fe(CN)_6]$ and 5 mM $K_4[Fe(CN)_6]$). Then, about 200 µL of the produced mixture was added to an exemplary three-electrode biosensor similar to biosensor **300**. EIS was performed using a potentiostat for a potential amplitude of about 10 mV and in a frequency range of about 10 mHz to 100 KHz. Charge transfer resistance (R_{CT}) was calculated based on the Nyquist diagram for each sample. Finally, the R_{CT} was compared between the cases without sign of cytokine storm and those who have hypercytokinemia based on the obtained results utilizing the ELISA method according to EXAMPLE 1 hereinabove.

[00115] **FIG. 11A** shows Nyquist plot for 9 samples of blood serum for each group of normal, moderate, and severely infected patients, consistent with one or more exemplary embodiments of the present disclosure. This figure demonstrates the obtained Nyquist plot for the serums of normal people without any symptoms and patients suspicious to the COVID-19 disease. Three

levels of responses with distinct curves may be observed in this figure. Smallest curves were recorded for normal donators and largest curves were observed in the EIS responses of the patients with severe infection. **FIG. 11B** shows comparing average values of R_{CT} for three groups of group of normal, moderate, and severely infected patients, consistent with one or more exemplary embodiments of the present disclosure. As could be deduced from this figure, a semicircle diameter of Nyquist plots in **FIG. 11A** as an indicator of the R_{CT} for normal groups and different stages of the patients showed meaningful ranges.

[00116] **FIG. 12A** shows R_{CT} values for 63 individual normal persons and infected patients, consistent with one or more exemplary embodiments of the present disclosure. It may be observed that R_{CT} for all of normal serum samples is less than about 377 Ω while for the moderate cases stand in a range of about 441 Ω to about 620 Ω and severe ones are more than about 750 Ω. Ranges from about 377 Ω to about 441 Ω and about 620 Ω to about 750 Ω may be free bands between normal-moderate and moderate-severe groups, respectively. The results of the impedimetric analysis showed an increasing correlation between R_{CT} (dielectric properties of blood serum) and the disease from infection to progression in stage. Lowest R_{CT} was recorded from samples of normal people while highest R_{CT} was recorded from samples of severely infected patients. **FIG. 12B** shows increase percentage of R_{CT} for COVID-19 patients in different progression stages relative to an avergae value of normal group, consistent with one or more exemplary embodiments of the present disclosure. An average increase ($\%\Delta R_{CT}/R_{CT}$) of about 101% was observed for the infected patients compared to the normal donators. Such a value of $\%\Delta R_{CT}/R_{CT}$ for moderate and severe cases were about 65% and about 138%, respectively.

[00117] To assess a correlation of hypercytokinemia with the increased charge transfer resistance obtained by the EIS, Receiver-Operating Characteristic (ROC) analysis was performed. Gold standard for positively scoring a patient with cytokine storm was ELISA tests (according to EXAMPLE 1) from the three cytokines of IL-6, TNF-α and IFN-γ and evaluating the total cytokine mass. **FIG. 13A** shows confusion matrix of diagnostic results obtained by EIS analysis and ELISA method for 63 persons, consistent with one or more exemplary embodiments of the present disclosure. **FIG. 13B** shows ROC curves for comparing EIS analysis with ELISA technique on detection of cytokine storm, consistent with one or more exemplary embodiments of the present disclosure. The statistical analysis shows a sensitivity of 100% and specificity of 77% with AUC of 0.95 for the biosensor with respect to the ELISA

as the gold standard for cytokine measurement. It is worth to note that a sum of the moderate and severe cases was studied for the ROC analysis.

[00118] Moreover, a calibration curves for the obtained R_{CT} values versus total cytokine mass measured according to EXAMPLE 1, were plotted. **FIG. 14** shows a calibration curve based on measured R_{CT} values and total cytokines mass for all three groups of normal, moderate and severe cases, consistent with one or more exemplary embodiments of the present disclosure. In this figure, dots illustrates the charge transfer resistance data, whereas the line is corresponding data fitting results on the basis of simple linear regression method. Based on the curves, there may be a satisfactory linear relationship with r^2=0.80 between the R_{CT} and the total mass of the cytokines. Total cytokine mass for the patients was between 7395 to 20770 kDa while this value for the normal group was between 1206 to 2345 kDa. Also, this value in moderate patients was about half of that in severe cases (**FIG. 5B**).

[00119] EXAMPLE 4: Evaluating the capability of the biosensor for COVID-19 detection

[00120] In this example, capability of the fabricated biosensor similar to biosensor **300** and utilized by an exemplary method similar to method **120** as described in above examples was investigated for detecting patients with COVID-19 disease since a cytokine storm may be an evidenced consequence in most of the COVID-19 patients. Gold standard for positively scoring a COVID-19 infected patient was either RT-PCR or CT-Scan. **FIG. 15** shows exemplary CT images of three exemplary persons from three groups of normal, moderate, and severe cases, consistent with one or more exemplary embodiments of the present disclosure. Exemplary chest CT image **1502** of a normal lung (ID#4) shows no abnormal opacification while for a moderate case shown in image **1504**, glass-ground opacification (GGO) may be seen in both lobes of lung (ID#29). Such a bilateral GGO pattern in severe cases may be more intensive with more involved area of both lung lobes accompanied with consolidation as shown for patient ID#55 in image **1506**.

[00121] **Table 1** compares detailed data of 63 persons analyzed by EIS sensor, RT-PCR, CT-Scan and cytokine analyses. As presented, three of patients (ID#23, ID#59, and ID#60) with clinical symptoms of infection were also analyzed in a case of cytokine storm as well as their CT/PCR results. Interestingly, their R_{CT} value was placed in range of severely infected patients with CT involvement but the RT-PCR showed no signs of COVID-19 infection and diagnosis of acute Flu was more probable. On the other hand, R_{CT} for three cases (ID#6, ID#8, and ID#11) were in range of moderately infected patients, while CT/PCR was negative and no cytokine

storm was detected by the ELISA results. These false signals of EIS for such cases might be due to an unknown non-specific binding of other elevated reagents in the blood serum of tested patients. Moreover, four false negatives were recorded by EIS analysis with normal RT-PCR and chest CT results (ID#3, ID#7, ID#9, and ID#13). The blood analysis of these patients revealed non-elevated IL-6, TNF-α and IFN-γ cytokines which indicated that the immune system was not reacted to the virus yet and just the lungs were involved in the infection. Hence, no cytokine storm was happened and neither ELISA nor EIS diagnosed the COVID-19 disease. In case of COVID-19 detection, about 92% of sensitivity with respect to both CT/PCR, 67% and 50% of specificity as well as AUC of 0.91 and 0.73 was reached based on CT and RT-PCR methods, as shown respectively in **FIGs. 16A**, **16B**, and **16C**. **FIG. 16A** shows confusion matrix of diagnostic results obtained by EIS analysis and RT-PCR method for 63 persons, consistent with one or more exemplary embodiments of the present disclosure. **FIG. 16B** shows confusion matrix of diagnostic results obtained by EIS analysis and CT scan method for 63 persons, consistent with one or more exemplary embodiments of the present disclosure. **FIG. 16C** shows ROC curves for comparing EIS analysis with RT-PCR and CT scan techniques on detection of cytokine storm, consistent with one or more exemplary embodiments of the present disclosure.

[00122] Table 1. EIS, CT, RT-PCR and total cytokine mass results of normal and COVID-19 patients

Patient ID	R_{CT}	$\Delta R_{CT}/R_{CT}$ (Avg) %	CT scan	RT-PCR	Total cytokine mass (kDa)
1	250	-	☒	☒	1206.46
2	315	-	☒	☒	1710.1
3	328	-	☑	☑	1806.7
4	263	-	☒	☒	1367.5
5	278	-	☒	☒	1665.4
6	456	29.9	☒	☒	2345.4
7	347	-	☑	☑	2073.4
8	598	70.37	☒	☒	2010.7
9	301	-	☑	☑	1614
10	288	-	☒	☒	1556.8
11	510	45.3	☒	☒	2090.6
12	294	-	☒	☒	1753.6
13	336	-	☑	☑	1756.9
14	472	34.47	☑	☑	8326
15	495	41.02	☑	☑	8715
16	582	65.8	☑	☑	10046

17	542	54.41	☑	☑	9368
18	780	122.2	☑	☑	9852
19	481	37.03	☑	☑	8412
20	510	45.29	☑	☑	8760
21	559	59.25	☑	☑	9523
22	544	54.98	☑	☑	9437
23	620	76.63	☑	☒	9682
24	829	136.18	☑	☑	9036
25	571	62.67	☑	☑	9818
26	467	33.04	☑	☑	8113
27	452	28.77	☑	☑	7910
28	486	38.46	☑	☑	8798
29	610	73.79	☑	☑	10822
30	441	25.64	☑	☑	7395
31	535	52.42	☑	☑	9209
32	503	43.30	☑	☑	8899
33	526	49.85	☑	☑	9100
34	596	69.80	☑	☑	10178
35	800	127.92	☑	☑	8724
36	459	30.77	☑	☑	8743
37	758	115.95	☑	☑	7997
38	902	156.98	☑	☑	10200
39	570	62.4	☑	☑	9826
40	740	110.82	☑	☑	13062
41	784	123.36	☑	☑	15712
42	818	133.05	☑	☑	17462
43	728	107.4	☑	☑	12524
44	867	147	☑	☑	17866
45	911	159.54	☑	☑	18907
46	845	140.74	☑	☑	17689
47	799	127.63	☑	☑	16949
48	887	152.7	☑	☑	18251
49	778	121.65	☑	☑	15201
50	715	103.7	☑	☑	11620
51	750	113.67	☑	☑	12534
52	786	123.93	☑	☑	15833
53	765	117.95	☑	☑	14644
54	896	155.27	☑	☑	18901
55	932	165.52	☑	☑	19573
56	831	136.75	☑	☑	17321
57	751	113.96	☑	☑	14015
58	1003	185.75	☑	☑	20770
59	980	179.2	☑	☒	15493
60	900	156.41	☑	☒	13401
61	802	128.49	☑	☑	15840
62	890	153.56	☑	☑	17411

63	888	152.99	☑	☑	15768

[00123] **Table 2** shows descriptive comparison on performance of cytokine storm and COVID-19 detection between exemplary method based on EIS analysis and ELISA, CT, and RT-PCR methods. These outcomes may mean that exemplary method based on EIS analysis and exemplary biosensor strongly responses to the patients with cytokine storm activity in their blood serum not affected by type of disease. So, exemplary method based on EIS analysis may compete with the current ELISA method on detection of the cytokine storm in the serum samples due to remarkable features of exemplary method and biosensor. In addition, since most of the patients infected with COVID-19 disease suffer from the cytokine storm, hence the results of EIS-based method on diagnosis of the infected patients by SARS-CoV-2 virus seems acceptable and could be compared with current methods of diagnosis such as RT-PCR and CT scan.

[00124] **Table 2.** Descriptive comparison on performance of cytokine storm and COVID-19 detection between EIS analysis and ELISA, CT, and RT-PCR methods

Method	Sample	Time	Accuracy	Cost	Procedure
Comparing methods for cytokine storm detection					
ELISA	Serum	< 90 min	High	Low-Cost	Simple
Our Method	Serum	< 10 min	High	Low-Cost	Simple
Comparing methods for COVID-19 detection					
CT scan	Lung	< 15 s	High	Expensive	Simple
RT-PCR	Saliva	< 3 h	High	Expensive	Complex
Our Method	Serum	< 10 min	Medium	Low-Cost	Simple

Industrial Applicability

[00125] Exemplary biosensor and method utilizing the biosensor disclosed here may include effective tools for real-time detecting at least one of cytokine storm, an infection of a chronic inflammatory disease (CID), and an infection of COVID-19 of a person. The biosensor and method may be helpful as a complementary diagnostic tool to warn about presence of cytokine storm in patients who are suspicious to COVID-19 during COVID-19 pandemic. The biosensor and method may declare diagnostic responses from serum of blood in less than about

10 minutes with full match between total cytokine's levels and EIS responses recorded by the method utilizing the biosensor. The biosensor with a sensitivity of about 92% may offer this biosensor could be applied for precise COVID-19 diagnosis. The biosensor based on the advantages of easy fabrication, fast detection and high accuracy may also be used for evaluating new drugs and vaccines developed for treatment of cytokine storm in COVID-19 patients.

[00126] While the foregoing has described what are considered to be the best mode and/or other examples, it is understood that various modifications may be made therein and that the subject matter disclosed herein may be implemented in various forms and examples, and that the teachings may be applied in numerous applications, only some of which have been described herein. It is intended by the following claims to claim any and all applications, modifications and variations that fall within the true scope of the present teachings.

[00127] Unless otherwise stated, all measurements, values, ratings, positions, magnitudes, sizes, and other specifications that are set forth in this specification, including in the claims that follow, are approximate, not exact. They are intended to have a reasonable range that is consistent with the functions to which they relate and with what is customary in the art to which they pertain.

[00128] The scope of protection is limited solely by the claims that now follow. That scope is intended and should be interpreted to be as broad as is consistent with the ordinary meaning of the language that is used in the claims when interpreted in light of this specification and the prosecution history that follows and to encompass all structural and functional equivalents. Notwithstanding, none of the claims are intended to embrace subject matter that fails to satisfy the requirement of Sections 101, 102, or 103 of the Patent Act, nor should they be interpreted in such a way. Any unintended embracement of such subject matter is hereby disclaimed.

[00129] Except as stated immediately above, nothing that has been stated or illustrated is intended or should be interpreted to cause a dedication of any component, step, feature, object, benefit, advantage, or equivalent to the public, regardless of whether it is or is not recited in the claims.

[00130] It will be understood that the terms and expressions used herein have the ordinary meaning as is accorded to such terms and expressions with respect to their corresponding respective areas of inquiry and study except where specific meanings have otherwise been set forth herein. Relational terms such as first and second and the like may be

used solely to distinguish one entity or action from another without necessarily requiring or implying any actual such relationship or order between such entities or actions. The terms "comprises," "comprising," or any other variation thereof, are intended to cover a non-exclusive inclusion, such that a process, method, article, or apparatus that comprises a list of elements does not include only those elements but may include other elements not expressly listed or inherent to such process, method, article, or apparatus. An element proceeded by "a" or "an" does not, without further constraints, preclude the existence of additional identical elements in the process, method, article, or apparatus that comprises the element.

[00131] The Abstract of the Disclosure is provided to allow the reader to quickly ascertain the nature of the technical disclosure. It is submitted with the understanding that it will not be used to interpret or limit the scope or meaning of the claims. In addition, in the foregoing Detailed Description, it can be seen that various features are grouped together in various implementations. This is for purposes of streamlining the disclosure, and is not to be interpreted as reflecting an intention that the claimed implementations require more features than are expressly recited in each claim. Rather, as the following claims reflect, inventive subject matter lies in less than all features of a single disclosed implementation. Thus, the following claims are hereby incorporated into the Detailed Description, with each claim standing on its own as a separately claimed subject matter.

[00132] While various implementations have been described, the description is intended to be exemplary, rather than limiting and it will be apparent to those of ordinary skill in the art that many more implementations and implementations are possible that are within the scope of the implementations. Although many possible combinations of features are shown in the accompanying figures and discussed in this detailed description, many other combinations of the disclosed features are possible. Any feature of any implementation may be used in combination with or substituted for any other feature or element in any other implementation unless specifically restricted. Therefore, it will be understood that any of the features shown and/or discussed in the present disclosure may be implemented together in any suitable combination. Accordingly, the implementations are not to be restricted except in light of the attached claims and their equivalents. Also, various modifications and changes may be made within the scope of the attached claims.

What is claimed is:

1. A method for measuring a level of total cytokines in a blood serum sample, comprising:

recording an electrochemical impedance spectroscopy (EIS) associated with the blood serum sample;

calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS; and

determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} by looking up the calculated R_{CT} in a calibration dataset comprising a set of total cytokine mass values respective to a set of R_{CT} values.

2. The method of claim 1, further comprising generating the calibration dataset, comprising:

generating the set of R_{CT} values, comprising:

recording a set of EIS plots from a respective plurality of blood serum samples drawn from a corresponding plurality of persons; and

calculating the set of R_{CT} values respective to the set of recorded EIS plots by measuring a diameter of a semicircular curved part of each of the recorded EIS plots; and

generating the set of total cytokine mass values corresponding to the set of R_{CT} values by measuring total cytokines mass of each of the plurality of blood serum samples utilizing a cytokine measurement assay.

3. The method of claim 1, wherein determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} further comprises detecting an infection of a chronic inflammatory disease (CID), comprising:

detecting the infection of the chronic inflammatory disease for a person associated with the blood serum sample responsive to the calculated R_{CT} being more than a threshold value.

4. The method of claim 3, wherein determining the level of total cytokines in the blood serum sample based on the calculated R_{CT} further comprises grading a severity of the chronic inflammatory disease infection of the person, comprising:

detecting the person is in a healthy (normal) state if the calculated R_{CT} is within a first range of the calibration dataset, the first range of the calibration dataset comprising a first set of R_{CT} values less than the threshold value, the first set of R_{CT} values measured from a group of healthy people;

detecting the person is in a moderately infected state if the calculated R_{CT} is within a second range of the calibration dataset, the second range of the calibration dataset comprising a second set of R_{CT} values between the threshold value and a severity borderline value, the second set of R_{CT} values measured from a group of patients with moderate levels of cytokines; and

detecting the person is in a severely infected state if the calculated R_{CT} is within a third range of the calibration dataset, the third range of the calibration dataset comprising a third set of R_{CT} values more than the severity borderline value, the third set of R_{CT} values measured from a group of patients with high levels of cytokines.

5. The method of claim 4, wherein detecting the infection of the chronic inflammatory disease comprises detecting a COVID-19 infection of the person if the calculated R_{CT} is equal to or more than 440 Ω.

6. The method of claim 4, wherein detecting the infection of the chronic inflammatory disease comprises detecting a COVID-19 infection status of the person based on the calculated R_{CT}, comprising:

detecting a healthy (normal) state for the person if the calculated R_{CT} is less than 340 Ω;

detecting a moderate COVID-19 infection of the person if the calculated R_{CT} is between 440 Ω and 610 Ω; and

detecting a severe COVID-19 infection of the person if the calculated R_{CT} is more than 715 Ω.

7. The method of claim 1, wherein recording the EIS associated with the blood serum sample comprises:

drawing a blood sample from a person;

separating the blood serum sample from the blood sample by centrifuging the blood sample;

forming a mixture by mixing the blood serum sample with an electrolyte solution comprising at least one metal ion;

putting the mixture in contact with three-integrated electrodes of a biosensor; and

recording an EIS from the mixture in contact with the three-integrated electrodes using an electrochemical stimulator-analyzer.

8. The method of claim 7, wherein recording the EIS from the mixture in contact with the three-integrated electrodes comprises recording the EIS of the blood serum sample in contact

with the three-integrated electrodes at an AC potential amplitude between 5 mV and 20 mV while sweeping a frequency range between 0.01 Hz and 100 kHz.

9. The method of claim 7, wherein putting the mixture in contact with the three-integrated electrodes of the biosensor comprises dropping the mixture on a surface of three-integrated electrodes of the biosensor, the three-integrated electrodes comprising:

a working electrode comprising a circular-shaped sensing part, the circular-shaped sensing part comprising a bilayer of graphene/copper adhered on a first bilayer of gold/titanium (Au/Ti) deposited on a substrate, the working electrode configured to be an attachment site for the mixture;

a counter electrode comprising a partially annular part placed around the working electrode, the counter electrode comprising a second bilayer of Au/Ti deposited on the substrate, the counter electrode configured to acquire the electrical response from the working electrode; and

a reference electrode placed adjacent to both the working electrode and the counter electrode, the reference electrode comprising a third bilayer of Au/Ti deposited on the substrate, the reference electrode configured to adjust a specific voltage between the working and the counter electrodes.

10. The method of claim 9, wherein:

the bilayer of graphene/copper comprises a layer of graphene sheets with a thickness between 0.1 nm and 1 nm deposited on a layer of copper with a thickness between 1 μm and 30 μm, and

each of the bilayers of Au/Ti comprises a layer of gold with a thickness between 10 nm and 50 nm deposited on a layer of titanium with a thickness between 3 nm and 10 nm.

11. A method for identifying a status of COVID-19 infection of a person, comprising:

drawing a blood sample from a person suspected to be infected with COVID-19 virus;

separating a blood serum sample from the blood sample by centrifuging the blood sample;

recording an electrochemical impedance spectroscopy (EIS) associated with the blood serum sample;

calculating a charge transfer resistance (R_{CT}) of the recorded EIS by measuring a diameter of a semicircular curved part of the recorded EIS; and

determining a COVID-19 infection status of the person based on the calculated R_{CT} if the calculated R_{CT} is equal to or more than a threshold value.

12. The method of claim 11, wherein determining the COVID-19 infection status of the person comprises:

detecting a healthy (normal) state for the person if the calculated R_{CT} is less than 340 Ω;

detecting a moderate COVID-19 infection of the person if the calculated R_{CT} is between 440 Ω and 610 Ω; and

detecting a severe COVID-19 infection of the person if the calculated R_{CT} is more than 715 Ω.

13. The method of claim 11, further comprising generating a calibration dataset, comprising:

generating a set of R_{CT} values, comprising:

recording a set of EIS plots from a respective plurality of blood serum samples drawn from a corresponding plurality of persons; and

calculating the set of R_{CT} values respective to the set of recorded EIS plots by measuring a diameter of a semicircular curved part of each of the recorded EIS plots; and

generating a set of total cytokine mass values corresponding to the set of R_{CT} values by measuring total cytokines mass of each of the plurality of blood serum samples utilizing a cytokine measurement assay.

14. The method of claim 13, wherein determining the COVID-19 infection status of the person based on the calculated R_{CT} comprises:

comparing the calculated R_{CT} with the set of R_{CT} values of the calibration dataset;

detecting the person is in a healthy (normal) state if the calculated R_{CT} is within a first range of the calibration dataset, the first range of the calibration dataset comprising a first set of R_{CT} values less than the threshold value, the first set of R_{CT} values measured from a group of healthy people;

detecting the person is in a moderately COVID-19 infected state if the calculated R_{CT} is within a second range of the calibration dataset, the second range of the calibration dataset comprising a second set of R_{CT} values between the threshold value and a severity borderline value, the second set of R_{CT} values measured from a group of moderately infected patients with COVID-19; and

detecting the person is in a severely infected state if the calculated R_{CT} is within a third range of the calibration dataset, the third range of the calibration dataset comprising a third set of R_{CT} values more than the severity borderline value, the third set of R_{CT} values measured from a group of severely infected patients with COVID-19.

15. The method of claim 11, wherein determining the COVID-19 infection of the person comprises detecting a COVID-19 infection of the person if the calculated R_{CT} is equal to or more than 440 Ω.

16. The method of claim 11, wherein recording the EIS associated with the blood serum sample comprises:

forming a mixture by mixing the blood serum sample with an electrolyte solution comprising a solution of at least one metal ion;

putting the mixture in contact with three-integrated electrodes of a biosensor; and

recording the EIS of the mixture in contact with the three-integrated electrodes using an electrochemical stimulator-analyzer.

17. The method of claim 16, wherein recording the EIS from the mixture in contact with the three-integrated electrodes comprises recording the EIS of the mixture in contact with the three-integrated electrodes at an AC potential amplitude between 5 mV and 20 mV while sweeping a frequency range between 0.01 Hz and 100 kHz.

18. The method of claim 16, wherein the electrochemical stimulator-analyzer comprises a potentiostat.

19. The method of claim 16, wherein putting the mixture in contact with the three-integrated electrodes of the biosensor comprises dropping the mixture on a surface of three-integrated electrodes of the biosensor, the three-integrated electrodes comprising:

a working electrode comprising a circular-shaped sensing part, the circular-shaped sensing part comprising a bilayer of graphene/copper adhered on a first bilayer of gold/titanium (Au/Ti) deposited on a substrate, the working electrode configured to be an attachment site for the mixture;

a counter electrode comprising a partially annular part placed around the working electrode, the counter electrode comprising a second bilayer of Au/Ti deposited on the substrate, the counter electrode configured to acquire the electrical response from the working electrode; and

a reference electrode placed adjacent to both the working electrode and the counter electrode, the reference electrode comprising a third bilayer of Au/Ti deposited on the substrate, the reference electrode configured to adjust a specific voltage between the working and the counter electrodes.

20. The method of claim 19, wherein:

the bilayer of graphene/copper comprises a layer of graphene sheets with a thickness between 0.1 nm and 1 nm deposited on a layer of copper with a thickness between 1 μm and 30 μm;

each of the bilayers of Au/Ti comprises a layer of gold with a thickness between 10 nm and 50 nm deposited on a layer of titanium with a thickness between 3 nm and 10 nm; and

the substrate comprises at least one of a glass substrate, a silicon substrate, a ceramic substrate, and combinations thereof.

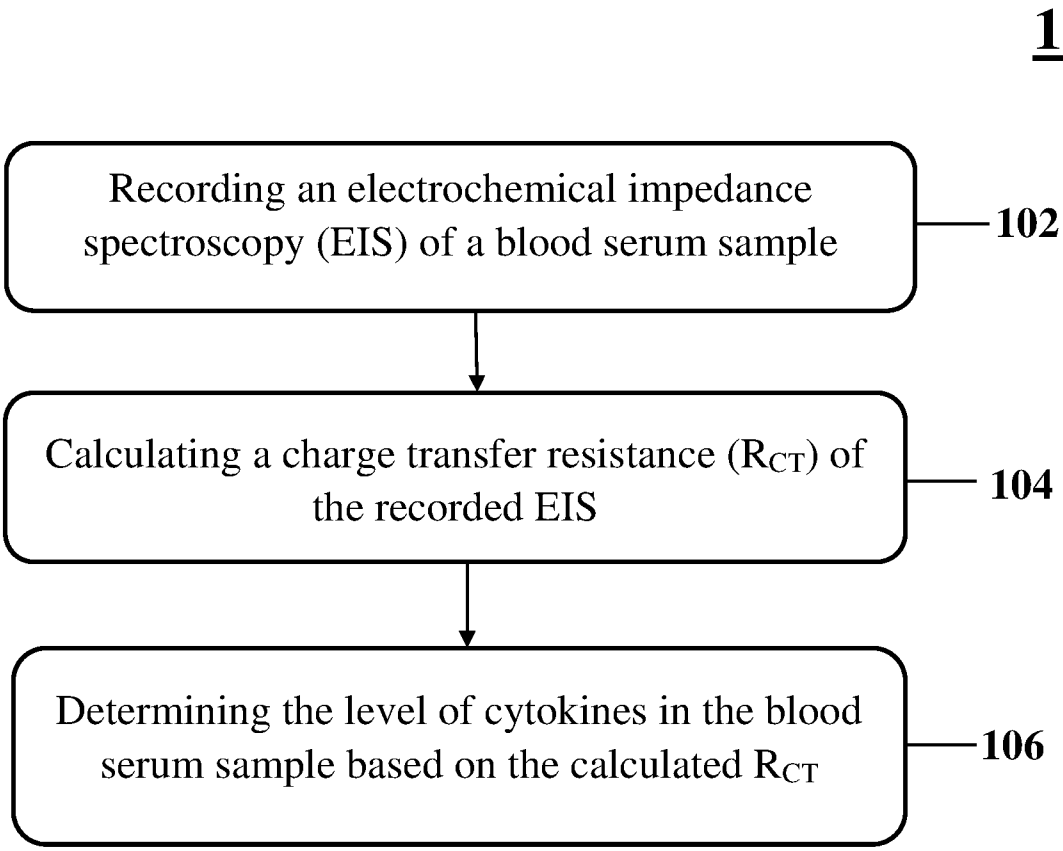

100
Recording an electrochemical impedance spectroscopy (EIS) of a blood serum sample
102
Calculating a charge transfer resistance (R_{CT}) of the recorded EIS
104
Determining the level of cytokines in the blood serum sample based on the calculated R_{CT}
106

FIG. 1A

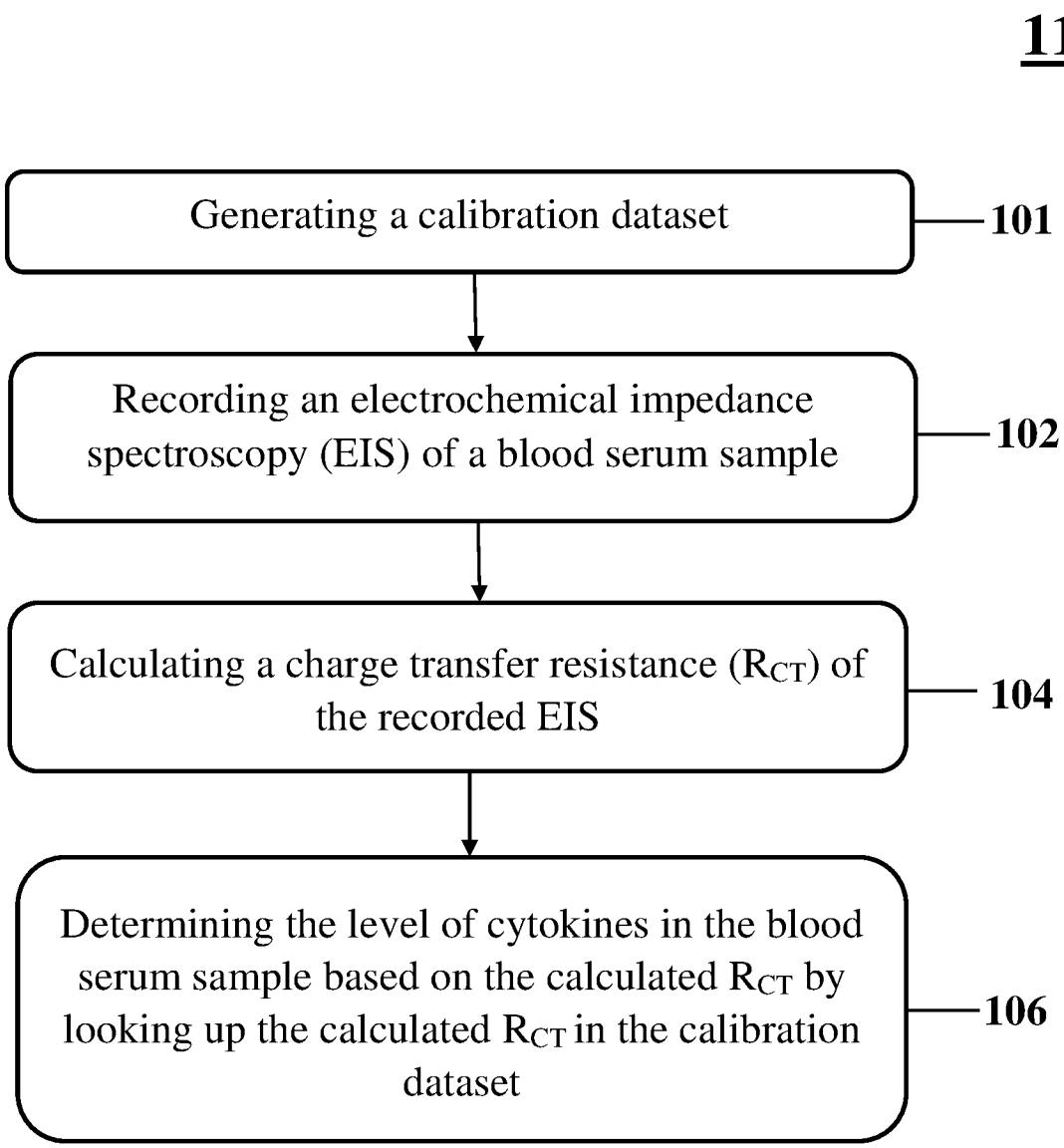

110
Generating a calibration dataset
101
Recording an electrochemical impedance spectroscopy (EIS) of a blood serum sample
102
Calculating a charge transfer resistance (R_{CT}) of the recorded EIS
104
Determining the level of cytokines in the blood serum sample based on the calculated R_{CT} by looking up the calculated R_{CT} in the calibration dataset
106

FIG. 1B

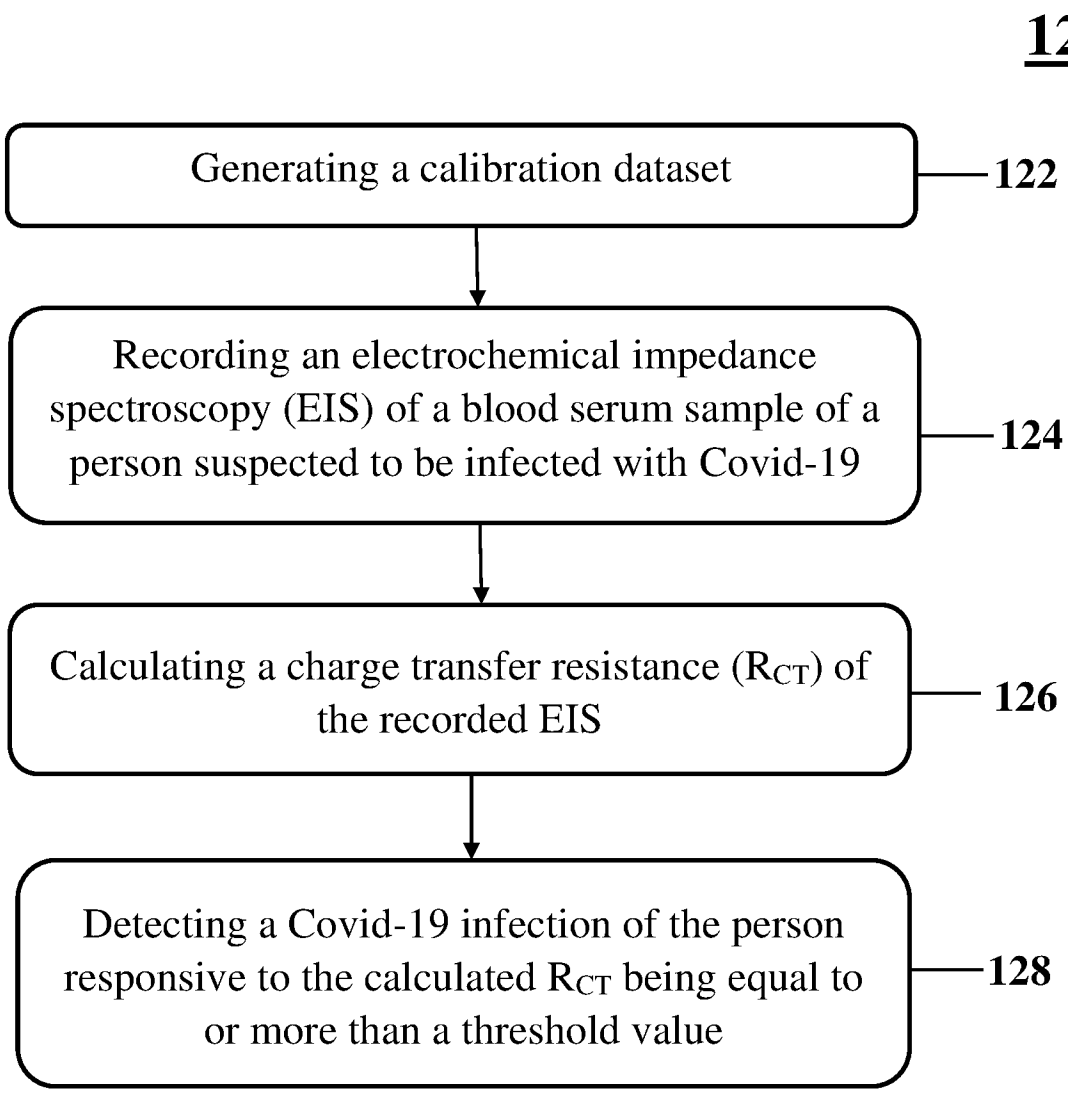

120
Generating a calibration dataset
122
Recording an electrochemical impedance spectroscopy (EIS) of a blood serum sample of a person suspected to be infected with Covid-19
124
Calculating a charge transfer resistance (R_{CT}) of the recorded EIS
126
Detecting a Covid-19 infection of the person responsive to the calculated R_{CT} being equal to or more than a threshold value
128

FIG. 1C

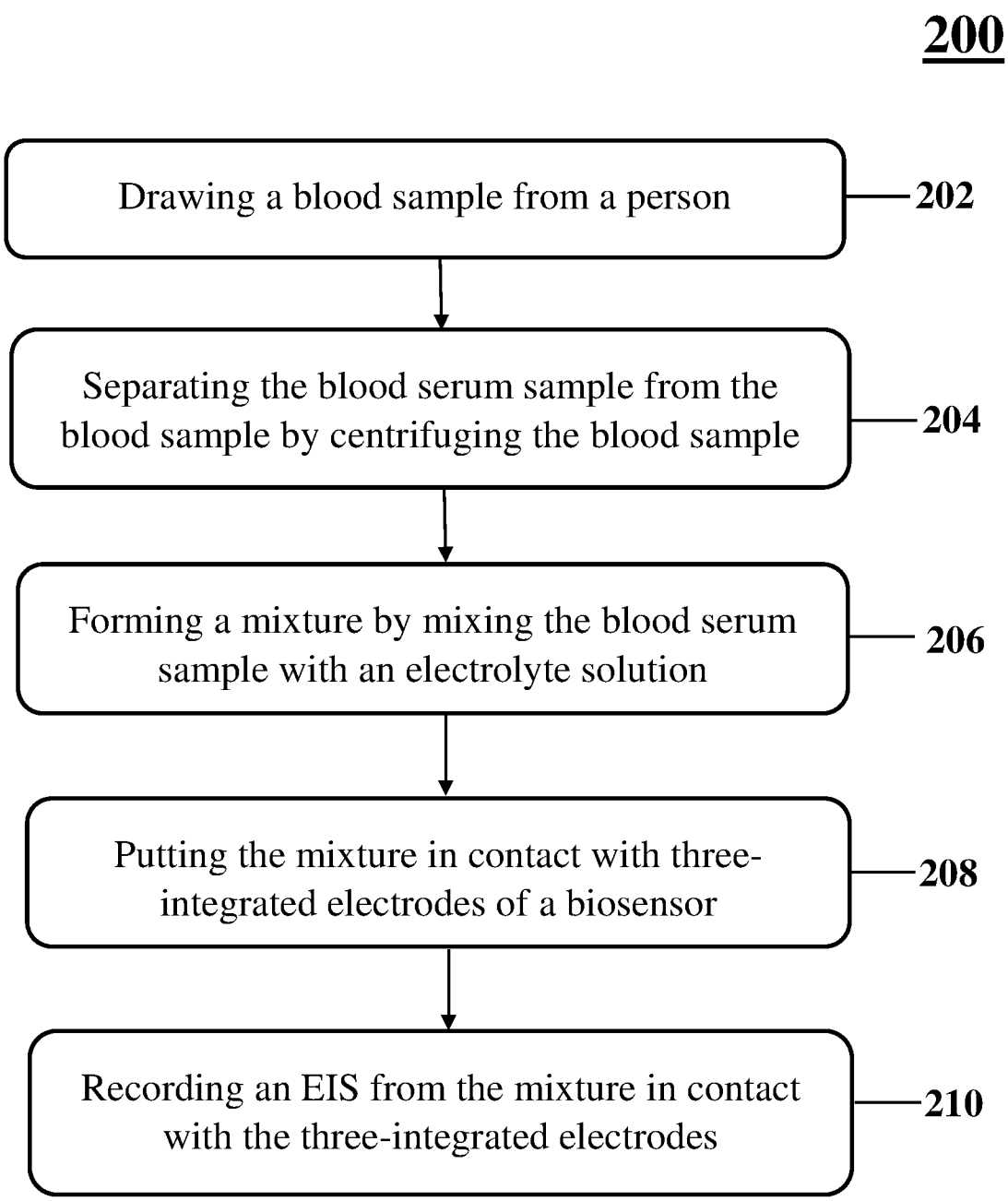

200
Drawing a blood sample from a person
202
Separating the blood serum sample from the blood sample by centrifuging the blood sample
204
Forming a mixture by mixing the blood serum sample with an electrolyte solution
206
Putting the mixture in contact with three-integrated electrodes of a biosensor
208
Recording an EIS from the mixture in contact with the three-integrated electrodes
210

FIG. 2

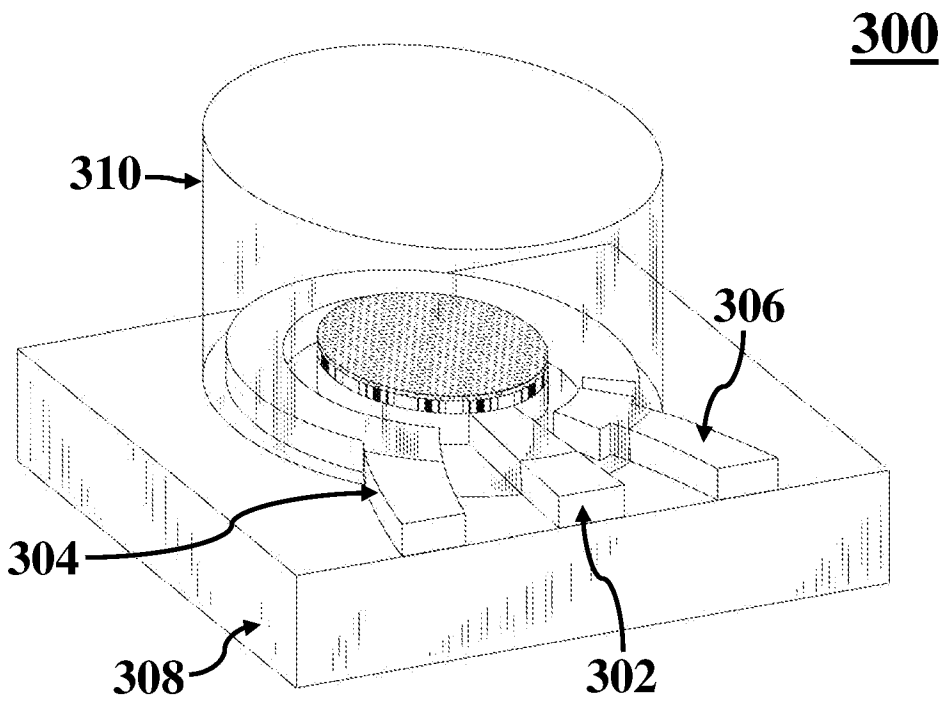


FIG. 3A

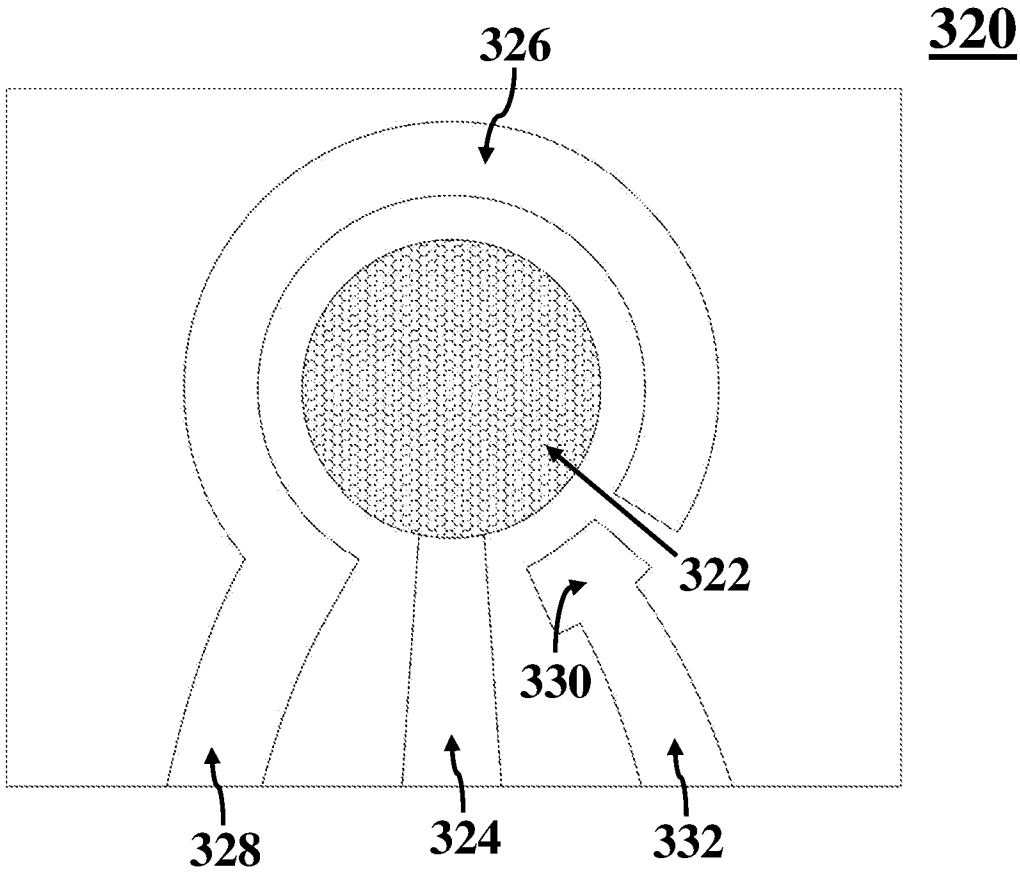


FIG. 3B

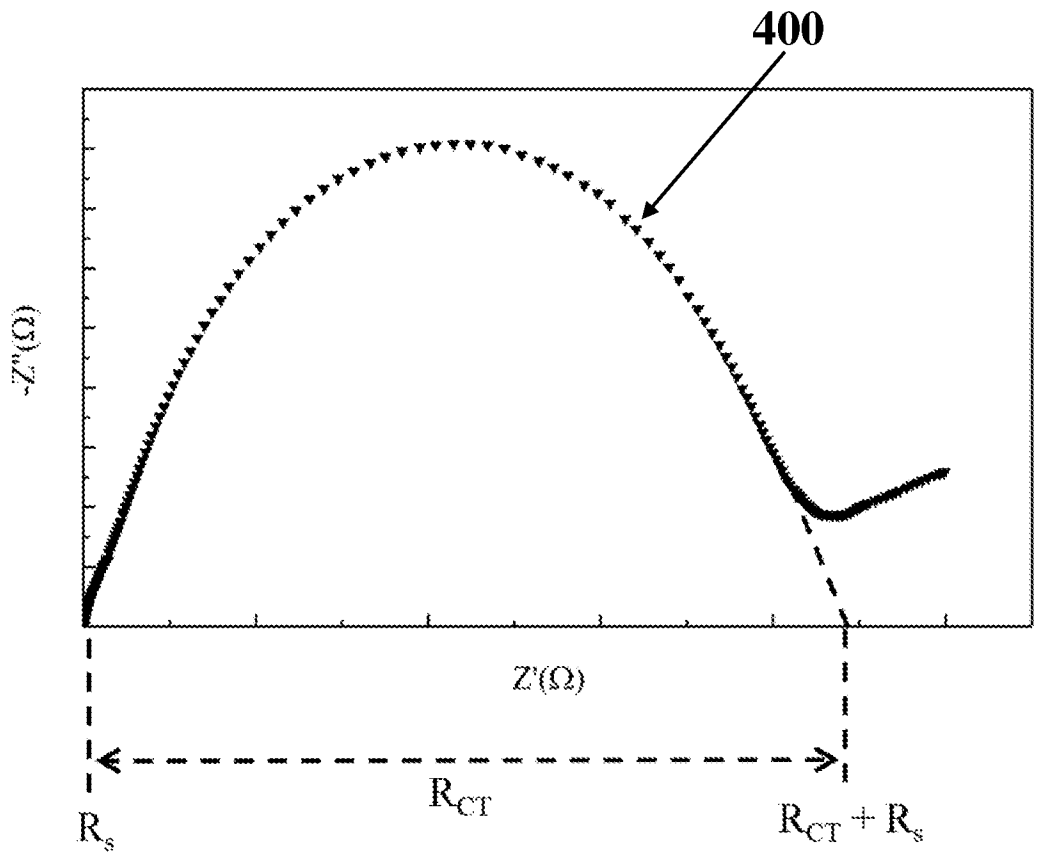


FIG. 4

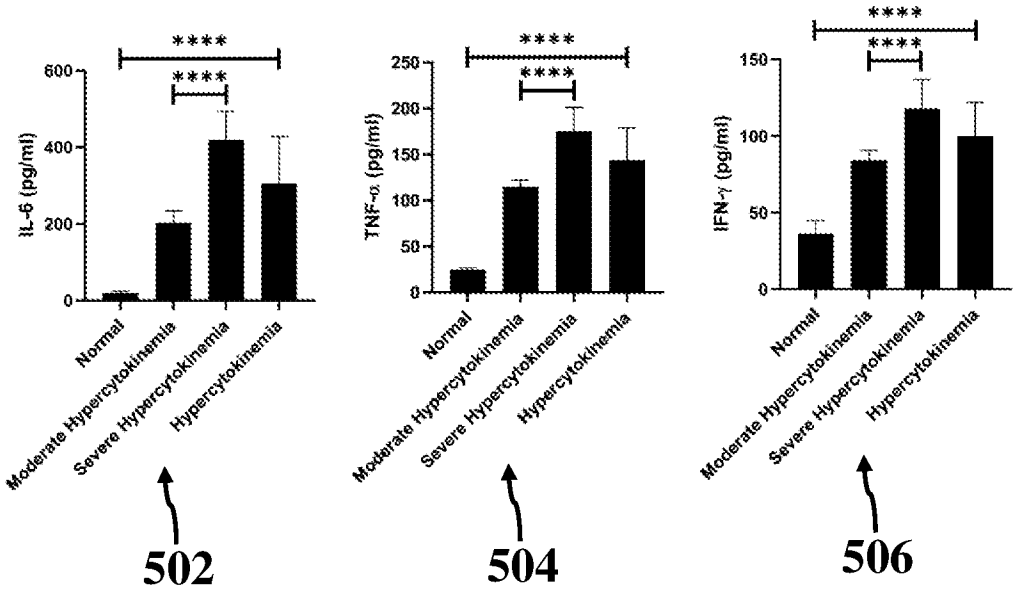

IL-6 (pg/ml)
600
400
200
0
****
****
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
Hypercytokinemia
502
TNF-α (pg/ml)
250
200
150
100
50
0
****
****
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
Hypercytokinemia
504
IFN-γ (pg/ml)
150
100
50
0
****
****
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
Hypercytokinemia
506

FIG. 5A

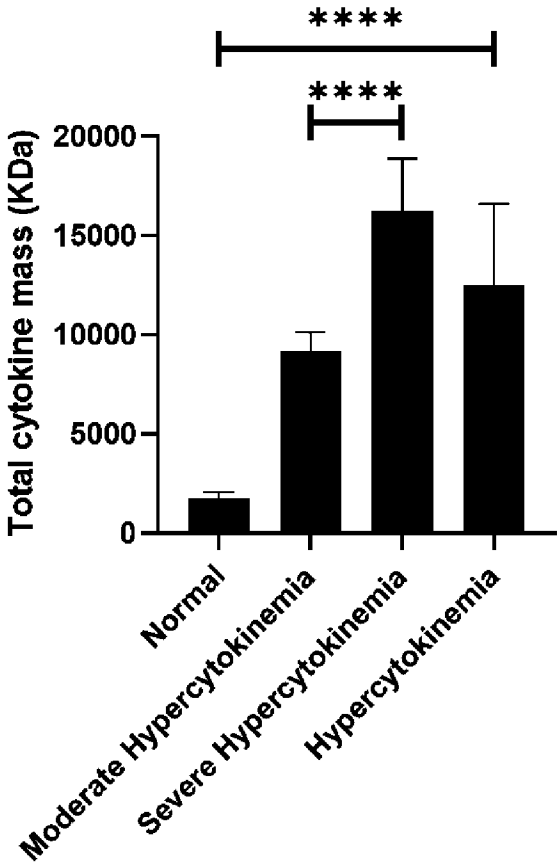

****
****
Total cytokine mass (KDa)
20000
15000
10000
5000
0
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
Hypercytokinemia

FIG. 5B

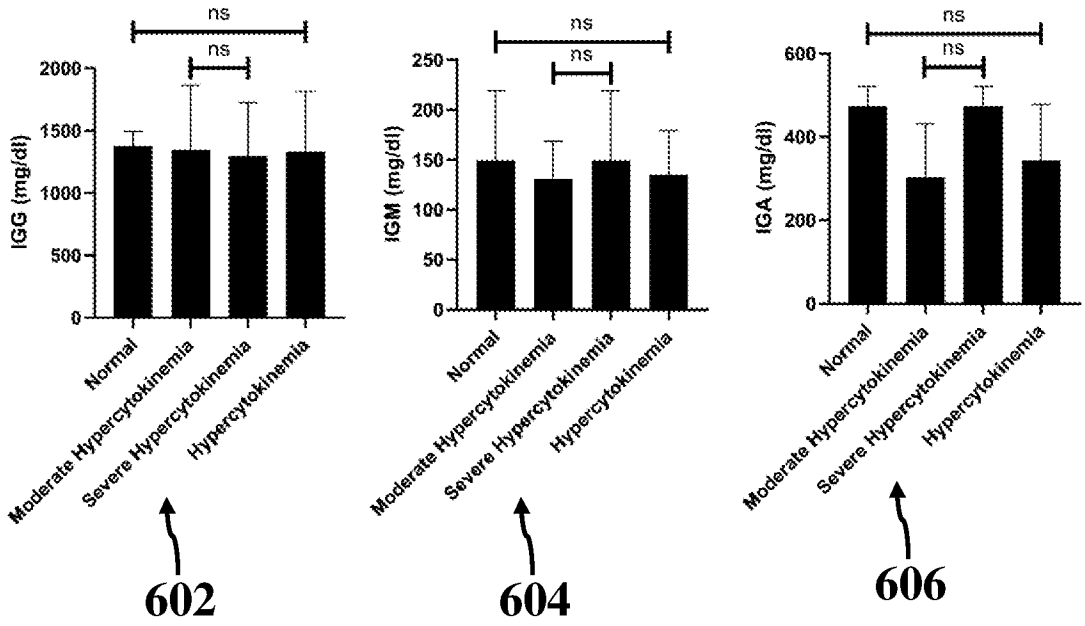

ns
ns
2000
1500
1000
500
0
IGG (mg/dl)
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
Hypercytokinemia
602
ns
ns
250
200
150
100
50
0
IGM (mg/dl)
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
Hypercytokinemia
604
ns
ns
600
400
200
0
IGA (mg/dl)
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
Hypercytokinemia
606

FIG. 6

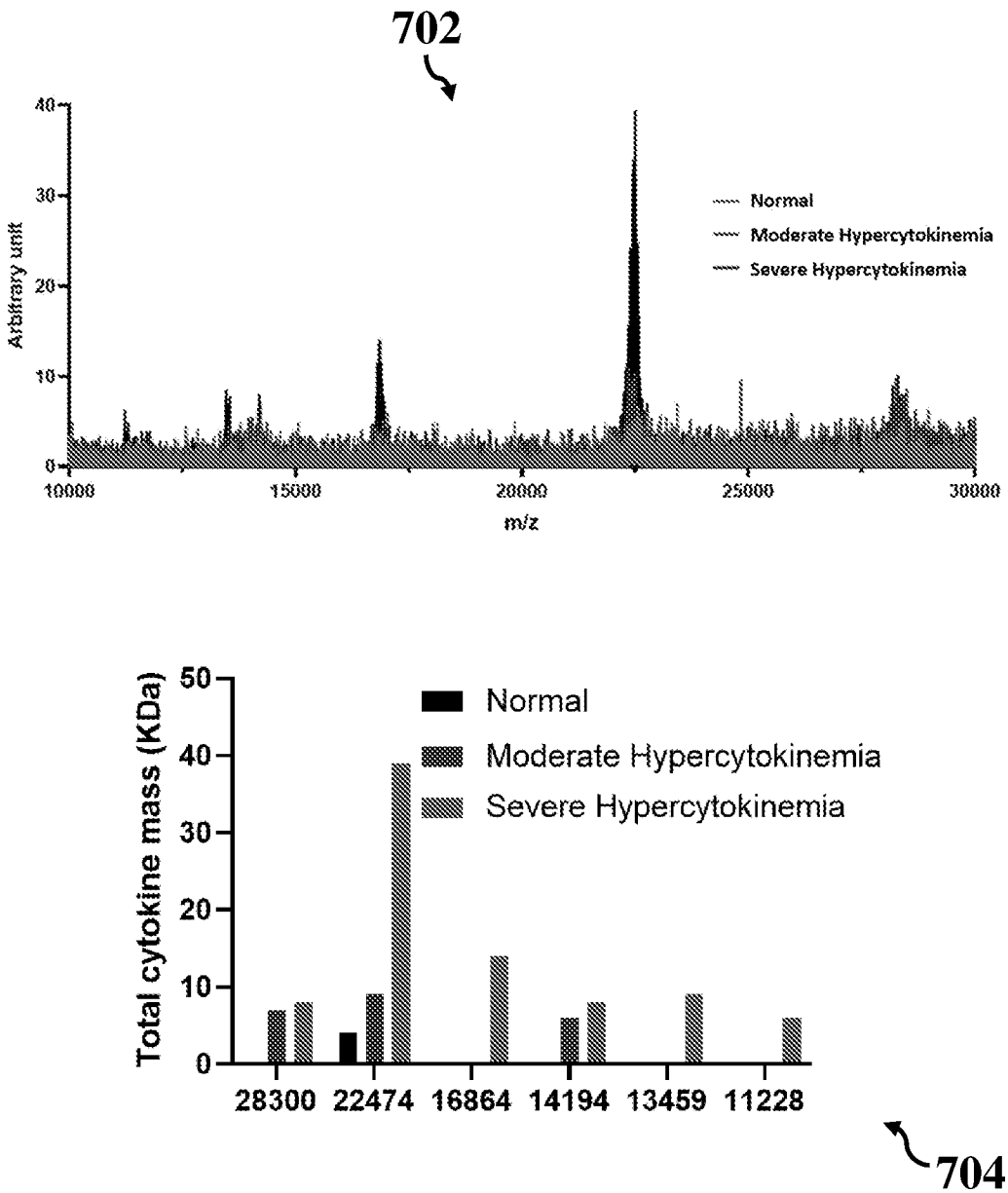

702
Arbitrary unit
40
30
20
10
0
10000
15000
20000
25000
30000
m/z
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
Total cytokine mass (KDa)
50
40
30
20
10
0
28300
22474
16864
14194
13459
11228
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
704

FIG. 7

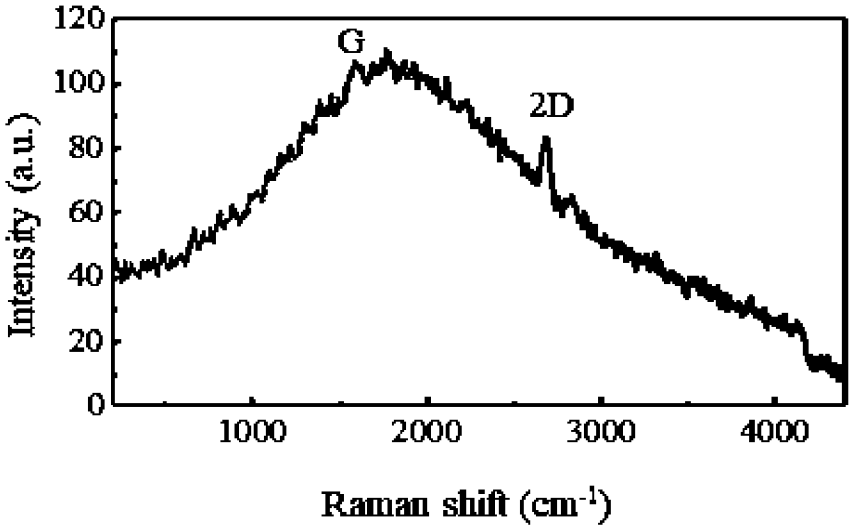

120
100
80
60
40
20
0
Intensity (a.u.)
G
2D
1000
2000
3000
4000
Raman shift (cm-1)

FIG. 8

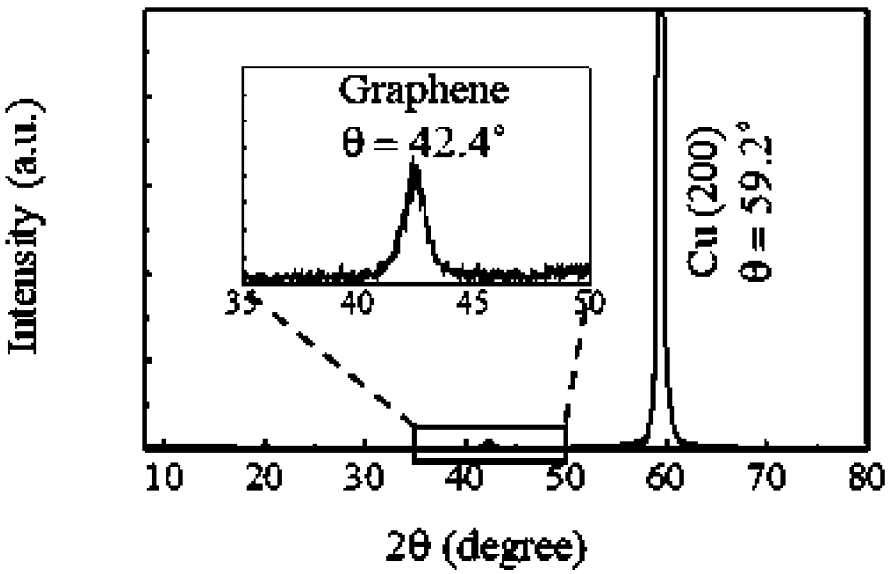

Graphene
θ = 42.4°
Cu (200)
θ = 59.2°
Intensity (a.u.)
35
40
45
50
10
20
30
40
50
60
70
80
2θ (degree)

FIG. 9

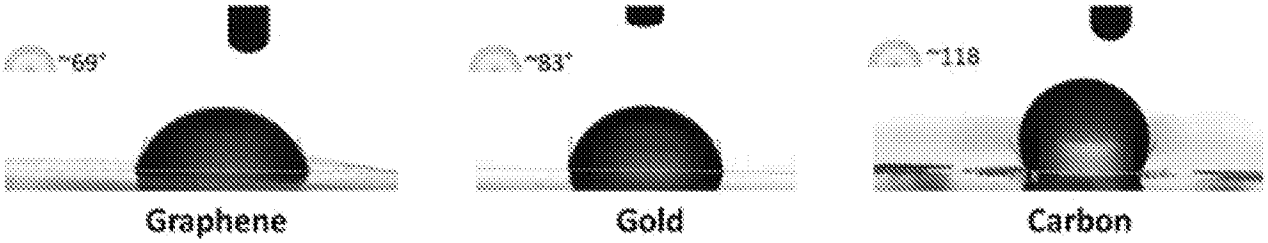


FIG. 10

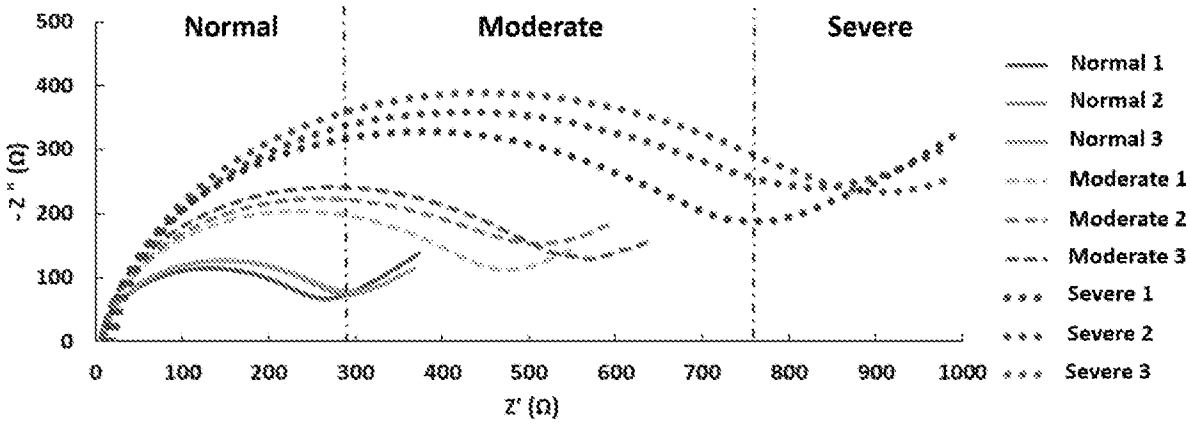

Normal
Moderate
Severe
500
400
300
200
100
0
-Z'' (Ω)
0
100
200
300
400
500
600
700
800
900
1000
Z' (Ω)
Normal 1
Normal 2
Normal 3
Moderate 1
Moderate 2
Moderate 3
Severe 1
Severe 2
Severe 3

FIG. 11A

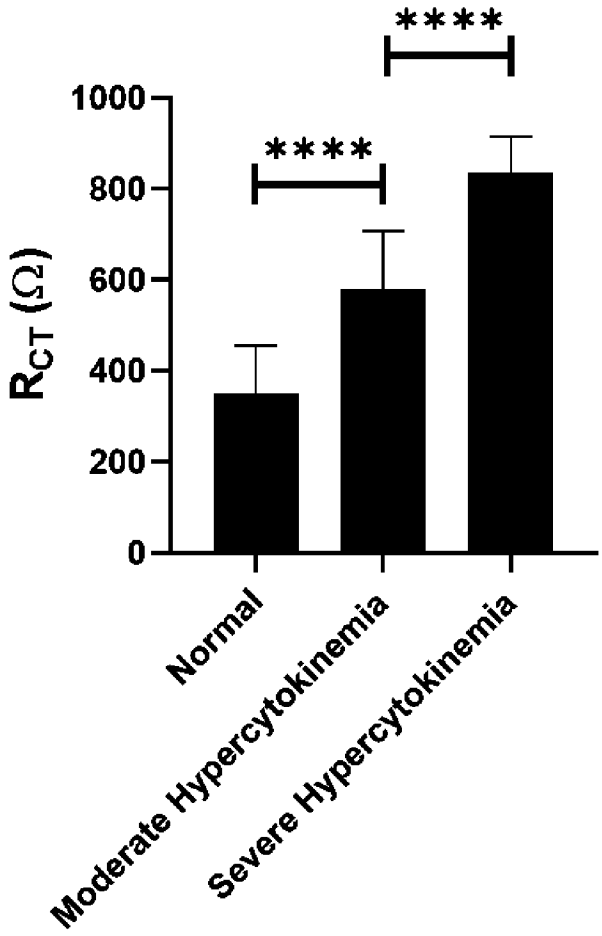

****
****
1000
800
600
400
200
0
R_{CT} (Ω)
Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia

FIG. 11B

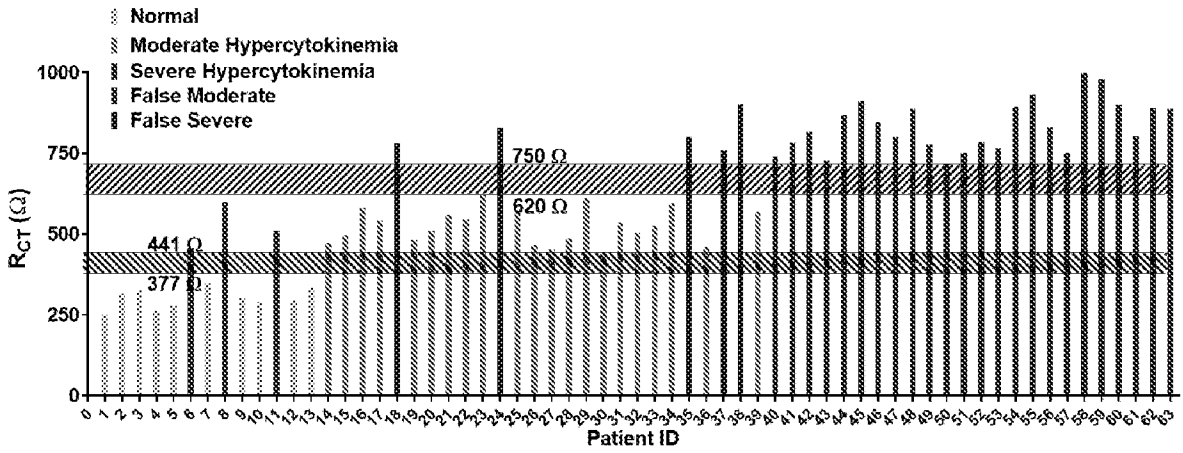

Normal
Moderate Hypercytokinemia
Severe Hypercytokinemia
False Moderate
False Severe
1000
750
500
250
0
R_{CT} (Ω)
750 Ω
620 Ω
441 Ω
377 Ω
Patient ID

FIG. 12A

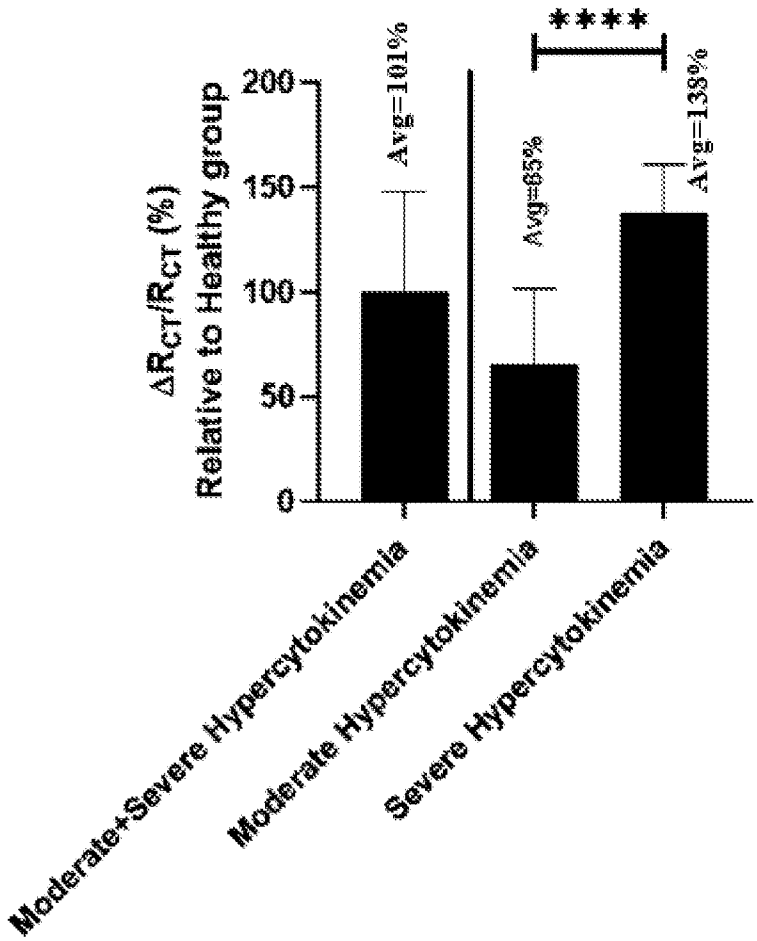

****
Avg=101%
Avg=65%
Avg=138%
200
150
100
50
0
ΔRCT/RCT (%)
Relative to Healthy group
Moderate+Severe Hypercytokinemia
Moderate Hypercytokinemia
Severe Hypercytokinemia

FIG. 12B

ELISA

EIS Analysis	Number of Cases: 63	Positive	Negative	Total
	Positive	True Positive: 50	False Positive: 3	53
	Negative	False Negative: 0	True Negative: 10	10
	Total	50	13	

FIG. 13A

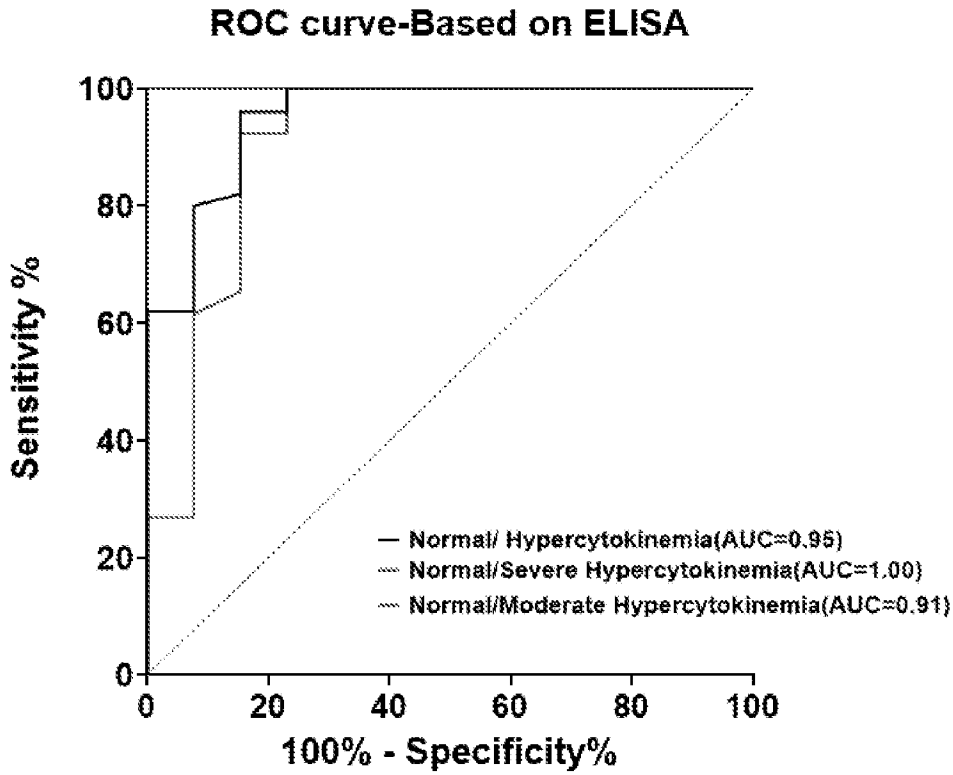

ROC curve-Based on ELISA
Sensitivity %
100
80
60
40
20
0
0
20
40
60
80
100
100% - Specificity%
Normal/ Hypercytokinemia(AUC=0.95)
Normal/Severe Hypercytokinemia(AUC=1.00)
Normal/Moderate Hypercytokinemia(AUC=0.91)

FIG. 13B

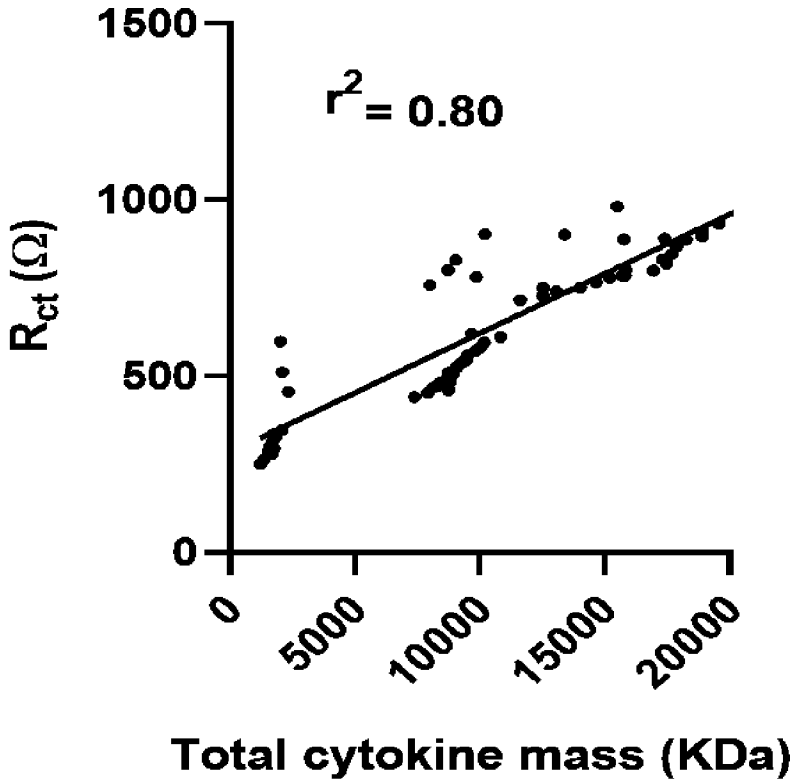

1500
1000
500
0
r^2 = 0.80
R_{ct} (Ω)
0
5000
10000
15000
20000
Total cytokine mass (KDa)

FIG. 14

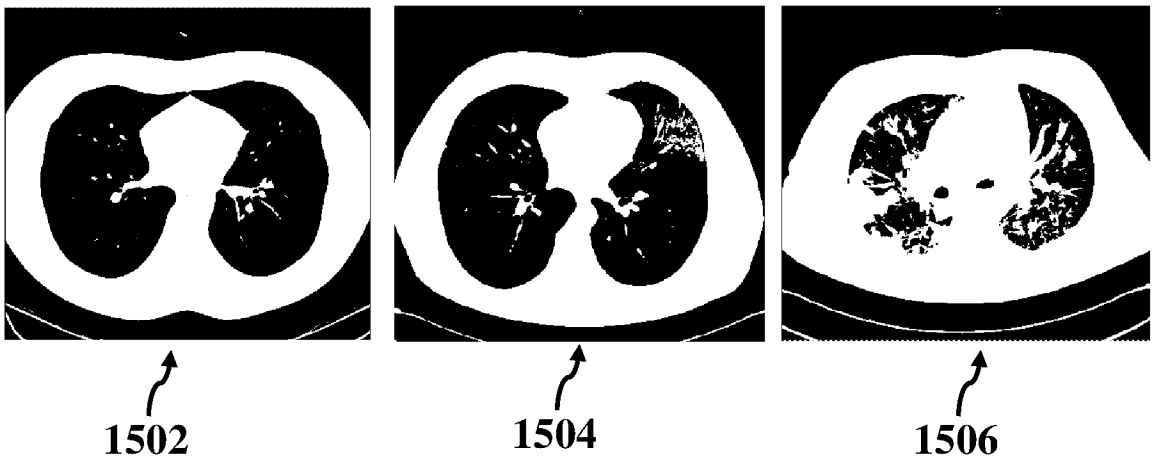


FIG. 15

	RT-PCR			
	Number of Cases: 63	Positive	Negative	Total
EIS Analysis	Positive	True Positive: 47	False Positive: 6	53
	Negative	False Negative: 1	True Negative: 9	10
	Total	48	15	Sensitivity ≈ 92% Specificity ≈ 50%

FIG. 16A

CT Scan

EIS Analysis	Number of Cases: 63	Positive	Negative	Total
	Positive	True Positive: 50	False Positive: 3	53
	Negative	False Negative: 1	True Negative: 9	10
	Total	51	12	Sensitivity ≈ 93% Specificity ≈ 67%

FIG. 16B

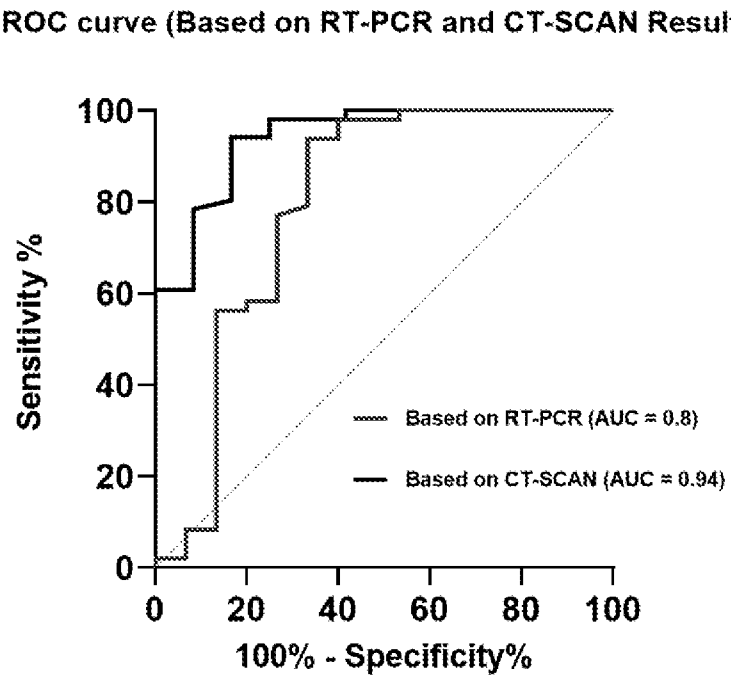

ROC curve (Based on RT-PCR and CT-SCAN Results)
Sensitivity %
100
80
60
40
20
0
0
20
40
60
80
100
100% - Specificity%
Based on RT-PCR (AUC = 0.8)
Based on CT-SCAN (AUC = 0.94)

FIG. 16C