

(12) United States Patent

Yu et al.

(54) COLD-TOLERANT PLANTS EXPRESSING

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Kaohsiung (TW)

MYBS3 AND DREB1A PROTEINS

Assignee: Academia Sinica, Taipei (TW)

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patent is extended or adjusted under 35

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Related U.S. Application Data

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- (51) Int. Cl. C12N 15/67 (2006.01)C12N 15/82 (2006.01)C07K 14/415 (2006.01)
- (52) U.S. Cl.

USPC 800/289; 800/298; 800/320.2; 800/278

(10) Patent No.:

US 8,859,851 B2

(45) **Date of Patent:**

Oct. 14, 2014

(58) Field of Classification Search

None

See application file for complete search history.

(56)**References Cited**

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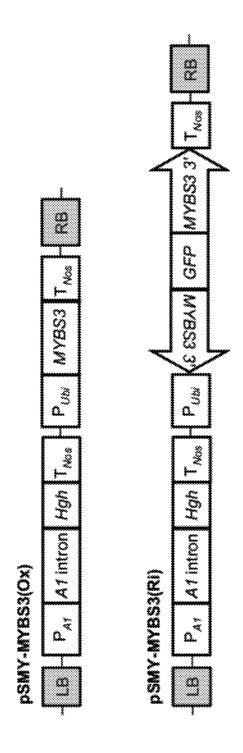
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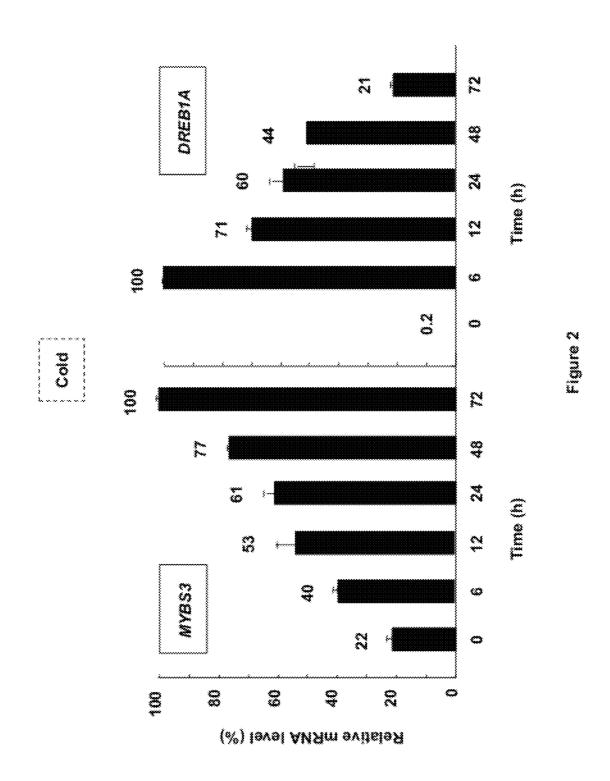
ABSTRACT (57)

Disclosed are chill- or cold-tolerant plants and methods of making the plants. Also disclosed are methods for identifying a plant that is tolerant to chill.

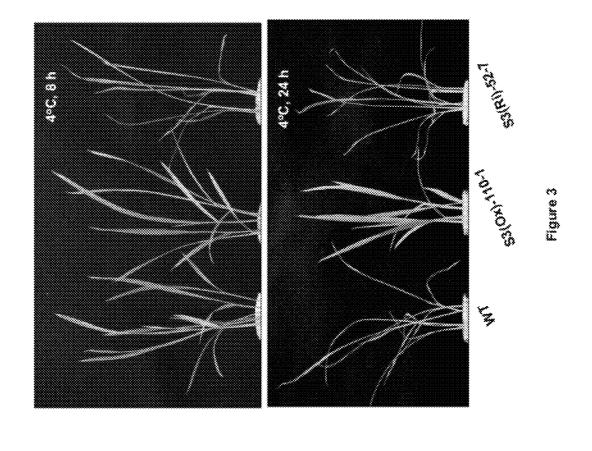
6 Claims, 6 Drawing Sheets



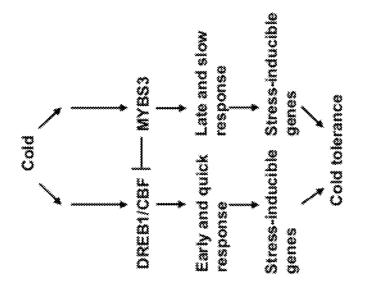
Figure



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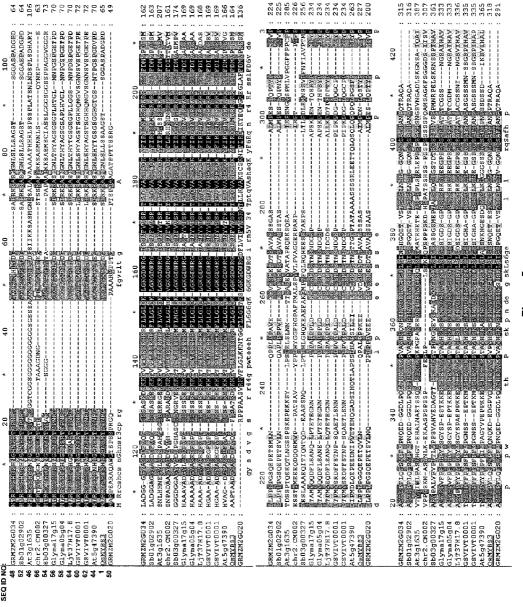


Figure 5 (conti.)

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*
CRMZMZG034 : **
Sb01g02902 : **
At3g1635 : **
Chr2.CM002 : A----- : 308
Sb03g00327 : **
Glyma17g15 : **
LjT37H17.8 : **
CSYLYT0001 : NGKILLVWRL : 345
At5g47390 : **
CSMYE83 : **
GRMZMZG020 : **
CSMYE83 : **
CSMY
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COLD-TOLERANT PLANTS EXPRESSING MYBS3 AND DREB1A PROTEINS

RELATED APPLICATION

This application claims priority of U.S. Provisional Application No. 61/370,351, filed on Aug. 3, 2010. The prior application is incorporated herein by reference in its entirety.

BACKGROUND

Various plants, including most important food crops, are not tolerant to cold. For example, rice seedlings are particularly sensitive to chilling in early spring in temperate and 2

subtropical zones and in high elevation areas. Improvement of chilling tolerance in rice or other food crops may significantly increase their production. There is a need for coldtolerant plants.

SUMMARY

This invention relates to using transgenic technology to generate cold-tolerant plants.

Accordingly, one aspect of this invention features a transgenic plant having a cell that has a MYBS3 gene expression level that is higher than a wild type MYBS3 protein level. Listed below is polypeptide sequence (SEQ ID NO: 1) and the nucleic acid sequence (SEQ ID NO: 2) of rice OsMybS3:

																				GGAG*	(SEQ	ID	NO:	1)
61	GG	GGG																		GTGC				
			М	Т	R	R	C	S	н	C	S	Н	N	G	н	N	S	R	Т	C				
121	CC	CAA	CCG	CGG	GGT	CAA	GAT	СТТ	CGG	GGT	'GCG	ССТ	CAC	CGA	TGG	СТС	CAT	CCG	CAA	GAGC				
			R																	S				
181																				CGCC				
	А	S	М	G	N	L	S	L	L	S	S	Α	Α	G	S	Т	S	G	G	A				
041	Шα	aaa	aaa	aar	aaa	aaa	aa*	aaa	aaa	aac	1070	aaa	aaa	aa 3	aaa	om v	aaa	ата	aa 3	aara				
241			A																	CGAC				
	٥		-	ט	G	-	ע		-	-	-	-	-	ט	G	-	-	5	ט	D				
301	тт	CGT	CCA	GGG	СТТ	CTC	CTC	CGC	CAC	CCG	CGA	.CCG	CAA	GAA	GGG	TGT	TCC'	TTG	GAC	TGAA				
	F	V	Q	G	F	S	S	Α	Т	R	D	R	K	K	G	V	P	W	Т	E				
361																				AGGA				
	Е	Е	Н	R	R	F	Ь	Ь	G	L	Q	K	Ь	G	K	G	D	W	R	G				
421	ΔТ	ሮሞሮ	TCG	таа	ጥጥጥ	ССТ	сст	מידמ	AAG	Δ Δ C	י ארר	тΔС	тсъ	ΔСТ	AGC	CAG	ጥሮል፡	ፐርሮ	тсъ	GAAA				
721			R																					
													~						~					
481	TA	TTT	TAT	ACG	CCA	ATC	CAA	TAT	GAC	CAG	AAG	GAA	AAG.	AAG	GTC	TAG	CCT	TTT	TGA	CATG				
	Y	F	I	R	Q	S	N	М	Т	R	R	K	R	R	S	S	L	F	D	M				
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541			AGA D																	CCAA				
	v	r	D	ш	٥	1-1	D	ш	-	r	п	-	G	G	Q	ш	F	15	1	Q				
601	GT.	ATT	AAA'	ГCА	ACC	AGC.	ACT.	ACC	TCC.	ACC	'GAA	.GGA	GGA	AGA	GGA	GGT	AGA'	TTC	TAT	GGAG				
	V	L	N	Q	P	Α	L	P	Ρ	Ρ	K	E	E	E	E	V	D	S	Μ	E				
661																				TTTG				
	S	D	Т	S	Α	V	А	E	S	S	S	Α	S	Α	Τ	M	Р	ע	N	ь				
721	CA	GTC	GAC	CTA	TCC	AGT	GAT	TGT	TCC.	AGC	TTA:	ттт	CTC	GCC	CTT	ттт	GCA.	ATT	CTC	GGTT				
			т																					
781																				TGTC				
	Р	F	W	Q	N	Q	K	D	Ε	D	G	Ρ	V	Q	Ε	Т	Н	Ε	Ι	V				
0/1	7.7.	מממ	ጥረታጥ	דירירי	지근만	ጥሮአ	ጥጥሮ	777	av a	ממר	ית איני	ראא	CCT	ሞር፡አ	ጥር፡አ	сст	ጥረታም	тсс	ር አ ጥ	GTCG				
041			V																					
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901	AA	GCT	CAG	CAT.	AGG	AGA	GTC	CAA	TCA	AGA	GAC	AGA	GTC	TAC	TTC	TCT	TTC.	ATT.	AAA	TCTG				
	K	L	S	Ι	G	E	S	N	Q	E	T	E	S	Т	S	L	S	L	И	L				
961																				GGCA				
	٧	G	G	Q	1/1	ĸ	Q	5	А	ь	н	А	1/1	Р	Р	1	ĸ	А	Q	А				
021	TG.	ATC	TGG	TTG	TGC	ACA	CAA	CTG	CAT	TTA	GAT	GAA	TCC	CAG	GCA	AAA	TAA	GCT	TTG	CCTC				
081	CT	TGT	TTT'	TTT	GTT	TTT	ATT	TTA	AGA	TTA	ACC	GTT	CTC	CGT	AGT	CTG	TAT	CAT	GTG	CTGT (SEQ I	D 1	VO: 2	2)
141	AA	GTT	ATG	CTA	TGT	ATG	AAT	GTA	TCT	GTT	'GTT	TGT	CTG	GCA	.CAC	ATG	ATA	AAT	CAC	TCTA				
201	тC	יים איים	አሮኦ	N N N	ጥሮኦ	СФУ	አ ጥር	CITE 76	OTP C	CTC	ነ አጥ C	ጥጥሮ	СТС	Cirrin	CITIN	CTTC	ጥጥረ።	ጥአኦ	አረጥ	CTTT				
.∠∪⊥	16	ıIА	ALA	наа	ı CA	GIA.	MIG	GIA	نی 1 ت	CIG	AIC	11C	GIG	GIT	GIA	ı C I G	116	ı AA	MC I	CIII				

encodes a polypeptide that is at least 40%, e.g., 50, 60, 70, 80, 85, 90, 95, 98, 99, or 100% identical to rice MYBS3 protein

1261 TATAAGAAAAAAAATATTAGTTAGTC

In the above-described transgenic plant, the MYBS3 gene 65 (SEQ ID NO: 1). For example, the MYBS3 gene can be a rice MYBS3 gene or its homologues, such as those from other plants (e.g., maize, wheat, barley, sorghum, sugarcane, turf

grass, Miscanthus, switchgrass, soybean, canola, potato, tomato, bean, pea, or jatropha). The exemplary homologues are listed below:

> At5g47390 (DNA)

(SEQ ID NO: 43) $\tt ATGACTCGTCGATGTTCTCACTGCAATCACAATGGCCACAACTCTCGGACTTGTCCCAATCGCGG$ $\tt CGTGAAGCTCTTTGGTGTTCGGCTCACCGAAGGTTCGATCCGGAAAAGTGCAAGTATGGGTAATC$ ${\tt TTAGCCATTACACGGGTTCTGGATCGGGTGGGCATGGAACCGGGTCCAACACTCCGGGTTCTCCG}$ $\tt GGTGATGTCCCTGACCATGTCGCTGGTGATGGTTACGCTTCTGAGGATTTCGTTGCTGGCTCTTC$ $\verb|CTCTAGCCGCGAGAGAAAGAAAGGAACTCCATGGACAGGAGGAAGAACACAGGATGTTCTTATTAG|$ $\tt GTTTACAGAAGCTGGGTAAAGGTGATTGGAGAGGTATCTCAAGAAACTATGTGACCACTAGGACA$ CCTACACAAGTTGCTAGCCATGCTCAGAAGTATTTCATCAGACAATCCAATGTCTCTCGTCGCAA ${\tt AAGACGTTCTAGTCTCTTTGATATGGTTCCTGATGAGGTTGGAGATATTCCCATGGATTTGCAAG}$ $\verb|AACCAGAGGAAGATAATATTCCTGTGGAAACTGAAATGCAAGGTGCTGACTCTATTCATCAGACA|$ $\tt CTTGCTCCTAGCTCACTTCACGCACCGTCAATCTTGGAAATCGAAGAATGTGAATCAATGGACTC$ $\tt CACAAACTCTACCACCGGGGAACCAACCGCAACTGCCGCTGCTGCTTCTTCTTCTTCCAGACTAG$ AAGAAACCACACAACTGCAATCACAACTGCAACCGCAGCCGCAACTACCTGGCTCATTCCCCATA CTATATCCGACCTACTTTTCACCATATTACCCGTTTCCATTCCCAATATGGCCTGCTGGTTATGT TCCTGAACCACCCAAGAAAGAGGAAACTCATGAAATTCTCAGACCAACTGCTGTGCACTCGAAAG GGAGAATCCGATCAGTCTCTTTCATTGAAGCTAGGTGGCGGGTCATCTTCAAGACAATCAGCATT ${\tt TCACCCGAATCCTAGCTCTGATAGTTCAGACATCAAAAGCGTGATACACGCTTTATAAAAGACCT}$ GAGGAAGTGATGGTCTAAAATGGG

>At5q47390 (Protein)

(SEQ ID NO: 44)

MTRRCSHCNHNGHNSRTCPNRGVKLFGVRLTEGSIRKSASMGNLSHYTGSGSGGHGTGSNTPGSP
GDVPDHVAGDGYASEDFVAGSSSSRERKKGTPWTEEEHRMFLLGLQKLGKGDWRGISRNYVTTRT
PTQVASHAQKYFIRQSNVSRRKRRSSLFDMVPDEVGDIPMDLQEPEEDNIPVETEMQGADSIHQT
LAPSSLHAPSILEIEECESMDSTNSTTGEPTATAAAASSSSRLEETTQLQSQLQPQPQLPGSFPI
LYPTYFSPYYPFPFPIWPAGYVPEPPKKEETHEILRPTAVHSKAPINVDELLGMSKLSLAESNKH
GESDQSLSLKLGGGSSSRQSAFHPNPSSDSSDIKSVIHAL

> At3g16350 (DNA)

(SEQ ID NO: 45)

-continued

TCTCACTCAATAATACCACAGAAGCTGAAGAGGTCGTAGCCACGGCGCCACGGCACAGGAAAAATCT
CAAGAAGCTATAGAACCATCAAATGGTGTTTCACCAATGCTAGTCCCGGGTGGCTTCTTTCCTCC
TTGTTTTCCAGTGACTTACACGATTTGGCTCCCTGCGTCACTTCACGGAACAGAACATGCCTTAA
ACGCTGAGACTTCTTCTCAGCAGCATCAGGTCCTAAAACCAAAACCTGGATTTGCTAAAGAACGT
GTGAACATGGACGAGTTGGTCGGTATGTCTCAGCTTAGCATAGGAATGGCGACAAGACACGAAAC
CGAAACTTCCCCTTCCCCGCTATCTTTGAGACTAGAGCCCTCAAGGCCATCAGCGTTTCACTCGA
ATGGCTCGGTTAATGGTGCAGATTTGAGTAAAGGCAACAGCGGGTTTCAGGCTATCTAA

>At3g1635 (Protein)

(SEQ ID NO: 46)

MTRRCSHCSNNGHNSRTCPTRGGGTCGGSGGGGGGGGGGGGGSGSSSAVKLFGVRLTDGSIIKKSAS
MGNLSALAVAAAAATHHRLSPSSPLATSNLNDSPLSDHARYSNLHHNEGYLSDDPAHGSGSSHRR
GERKRGVPWTEEEHRLFLVGLQKLGKGDWRGISRNYVTSRTPTQVASHAQKYFIRHTSSSRRKRR
SSLFDMVTDEMVTDSSPTQEEQTLNGSSPSKEPEKKSYLPSLELSLNNTTEAEEVVATAPRQEKS
QEAIEPSNGVSPMLVPGGFFPPCFPVTYTIWLPASLHGTEHALNAETSSQQHQVLKPKPGFAKER
VNMDELVGMSQLSIGMATRHETETSPSPLSLRLEPSRPSAFHSNGSVNGADLSKGNSAIQAI

> GRMZM2G034110_T01 (DNA)

(SEQ ID NO: 47)

>GRMZM2G034110 T01 (Protein)

(SEO ID NO: 48)

MTRRCSHCSHNGHNSRTCPNRGVKIFGVHLTDGSAIRKSASMGNLSLLSAGSTSGGASPADGPDL
ADGGGGYASDDFVQGSSSASRDRKKGVPWTEEEHRRFLLGLQKLGKGDWRGISRNFVVSRTPTQV
ASHAQKYFIRQSNMSRRKRRSSLFDMVPDESMDLPPLPGSQEPETSMLNQPPLPPAVEEEVESME
SDTSAVAESSGASALMPESLQPTYPMIVPAYFSPFLQFSVPFWPNQEDGGDLPQETHEIVKPVAV
HSONPINVDELVGMSKLSIWEHGOETVSTSLSLNLLGGONROSAFHANPOTRAQA

> GRMZM2G020934_T01 (DNA)

(SEO ID NO: 49)

>GRMZM2G020934_T01 (Protein)

(SEO ID NO: 50)

MARPSARAQAWGTSPSSPRGQPAAARRPPTGPTSPTAAGATPPTTSSRGRPPPÄASVRKVFLGLK
KNTGGFLLGITKSLGKGDWRGDFLVIFVVSKNTLLKVAKSCSKNIFIRQIKYEQKGREGLAFFDM
VPDESMDLPPLPGSQEPETSVLNQPPLPPPVEEEEEVESMESDTSAVAESSAASALMPESLQPTY
PMIVPAYFSPFLQFSVPFWPNQEDGGDLPQETHEIVKPVAVHSKNPINVDELVSMSKLSIGEPGQ
ETVSTSLSLNLLVGQNRQSAFHANPQTRAQA

> Sb01g029020.1 (DNA)

(SEQ ID NO: 51)

>Sb01g029020.1 (Protein)

(SEQ ID NO: 52)

MTRRCSHCSHNGHNSRTCPNRGVKIFGVRLTDGSAIRKSASMGNLSLLSAGSTSGGASPADGPDL
ADGGAGGYASDDFVQGSSSASRERKKGVPWTEEEHRRFLLGLQKLGKGDWRGISRNFVVSRTPTQ
VASHAQKYFIRQSNMSRRKRRSSLFDMVPDESMDLPPLPGSQEPETSVLNQAPLPPPPVEEEVESM
ESDTSAVAESSTASALMPESLQPNYPMIVPAYFSPFLQFSVPFWPNQEDGGDLPQETHEIVKPVA
VHSKNPINVDELVGMSKLSIGEPGQETVSTSLSLNLLGGQNRQSAFHANPQTRAQA

>Sb03g003270.1 (DNA)

(SEQ ID NO: 53) $\tt CGGGCACGCCTCCTGCTCGACGAATGGCCGCGTCGAGCGGAAGAAAGGTACACCTTGGACTGAAG$ ${\tt AAGAGCATAGAATGTTTCTGATGGGTCTTCAGAAGCTTGGTAAGGGAGATTGGCGCGGGATATCT}$ $\tt CGAAACTTTGTTGTTTCCAGGACCCCGACTCAAGTGGCAAGCCATGCTCAAAAGTACTTTATTAG$ AACTTTTCAGCTAAGGCAGCATGAGGAATCTGAATATGCAGAACCTTCATTGACATTACCAGATT TAGAGATGAACTCCAGTGTACCATTCAATACCATAGCTGTTCCGACGATGCCAGCATTCTACCCT GCGTTGGTCCCTGTTCCACTAACTCTTTGGCCTCCAAGTGTTGCCCATGTGGAGGACGCAGGCAC AACCCATGAAATCCTAAAACCAACTCCTTTGAATGGTAAGGAGGTGATTAAAGCAGATGATGTTG $\tt TTGGTATGTCTAAGCTCAGCATTGGTGAGGCCAGCTCTTGGCTCCATGGAACCCACAGCTCTTTCC$ $\tt CTTCAGCTTATTGGATCGACAGATACAAGGCAGTCAGCTTTTCATGTGAGTCCACCAATGAATAG$ ACCTGAACTAAGCAAGAGAAACAGCAGTCCAATTCATGCCGTTTGA

>Sb03g003270.1 (Protein)

(SEQ ID NO: 54)

MTRRCSHCSNNGHNSRTCPARSGGGVRLFGVRLTTAPAPAAMKKSASMSCIASSLGGGSGGGSSPP
AGGVGGGRGGGDGGAGYVSDDPGHASCSTNGRVERKKGTPWTEEEHRMFLMGLQKLGKGDWRGIS
RNFVVSRTPTQVASHAQKYFIRQTNSSRRKRRSSLFDMVAEMPVDESLAAAEQITIQNTQDEAAS
SNQLPTLHLGHQKEAEFAKQMPTFQLRQHEESEYAEPSLTLPDLEMNSSVPFNTIAVPTMPAFYP
ALVPVPLTLWPPSVAHVEDAGTTHEILKPTPLNGKEVIKADDVVGMSKLSIGEASSGSMEPTALS
LQLIGSTDTRQSAFHVSPPMNRPELSKRNSSPIHAV

> Glyma17g15330.1 (DNA)

(SEQ ID NO: 55)

>Glyma17g15330.1 (Protein)

(SEQ ID NO: 56)
MTRRCSHCSHNGHNSRTCPNRGVKLFGVRLTDGSIRKSASMGNLTHYAGSGSGPLHTGLNNPGSP
GETPDHAAAVADGYLSEDFVPGSSSSSRERKKGVPWTEEEHRMFLLGLQKLGKGDWRGIARTYVI
SRTPTQVASHAQKYFIRQSNVSRRKRRSSLFDIVADEAADTAMVQQDFLSANQLPTETEGNNPLP
APPPLDEECESMDSTNSNDGEPAPSKPENTQSSYPMLYPAYYSPVFPFPLPYWSGYSPESTKKEE
THEVLKPTAVHSKSPINVDELVGISKLSLGESIGDSGPSSLSRKLIEEGPSRQSAFHATPTCGSS
NGSAIHAV

> Glyma05g04950.1 (DNA)

(SEO ID NO: 57) GGTTAAGCTCTTCGGGGTCCGATTAACCGACGGGTCGATCCGGAAGAGCGCCAGCATGGGCAACC TAACCCACTACGCTGGTTCCGGGTCGGCCCCGCTCCATGTCGGGTTGAATAACCCGGGTTCACCC GGGGAGACGCCCGATCACGCCGCCGCCGCCGACGCTACGCCTCCGAGGACTTCGTTCCCGG GTCTTCTTCTAGCTCCCGTGAAAGAAGAAGGGTGTTCCATGGACTGAGGAGGAACATAGAATGT TTTTGCTCGGATTGCAGAAGCTGGCCAAAGGTGATTGCCGTGGAATTGCAAGGAACTATGTTATA ${\tt TCAAGGACGCCTACTCAAGTGGCCAGCCATGCTCAGAAATATTTCATCAGGCAAAGCAATGTGTC}$ CAGGCGAAAAAGACGGTCCAGCTTGTTTGATATTGTTGCAGATGAAGCAGCTGACACTGCAATGG ${\tt TACAGCAAGACTTCTTGTCTGCTAATGAGTTACCAACTGAAACAGAAGGCAATAACCCCTTGCCT}$ $\tt GCTCCTCCCCTCGATGAAGAGTGTGAATCAATGGATTCCACAAACTCAAATGATGGAGAGCC$ $\tt TGCCCCATCAAAGCCAGAAAACACACCATCCTTATCCTATGTTATATCCTGCGTATTATTCTC$ ${\tt CAGTGTTCCCGTTTCCTCTGCCCTATTGGTCAGGATACAGTCCAGAGCCCACCAAGAAGGAGGAA}$ ${\tt ACACATGAAGTGCTGAAACCAACTGCAGTACATTCTAAAAGCCCTATCAATGTTGATGAACTGGT}$ $\tt TGGCATATCAAAACTGAGTTTAGGGGAGTCTATTGGTGACTCGGGTCCCTCCACCCTGTCTCGAA$ AACTTATTGAAGAAGGACCCTCTAGACAATCAGCTTTTCATGCAACACCAACATGTGGTGATATG AATGGCAGTGCCATCCATGCAGTTTAA

>Glyma05g04950.1 (Protein)

(SEQ ID NO: 58)
MTRRCSHCSHNGHNSRTCPNRGVKLFGVRLTDGSIRKSASMGNLTHYAGSGSAPLHVGLNNPGSP
GETPDHAAAAADGYASEDFVPGSSSSSRERKKGVPWTEEEHRMFLLGLQKLGKGDWRGIARNYVI
SRTPTQVASHAQKYFIRQSNVSRRKRRSSLFDIVADEAADTAMVQQDFLSANELPTETEGNNPLP
APPPLDEECESMDSTNSNDGEPAPSKPENTHPSYPMLYPAYYSPVFPFPLPYWSGYSPEPTKKEE
THEVLKPTAVHSKSPINVDELVGISKLSLGESIGDSGPSTLSRKLIEEGPSRQSAFHATPTCGDM
NGSAIHAV

> GSVIVT00013475001 (DNA)

(SEQ ID NO: 59)
ATGACTCGCCGCTGCTCGCATTGCAGTCACAACGGGCACAATTCCAGGACATGCCCCAACCGCGG
GGTCAAGATCTTCGGGGTTCGATTGACTGATGGGTTGATCCGTAAGAGTGCTAGTATGGGCAATC
TCAGCCACTACGCCGGGTCGACCTCTGGTCATCATCAGAACGGCGTTTCCGGTAACAATTCGGTC
TCTCCCGGAGAGACTCCAGAGCACGGCGCCGCGGCCGATGGATACGCCTCCGAGGGTTTCGTTCC
CGGTTCATCATCCAGCCGGGAGCGCAAGAAAGGCACTCCATGGACTGAAGAGGAACACAGAATGT
TTCTACTTGGACTGCAGAAAGCTTGGAAAAAGGGGATTGCCGTGAATTTATGTTATA

>GSVIVT00013475001 (Protein)

(SEO ID NO: 60)

MTRRCSHCSHNGHNSRTCPNRGVKIFGVRLTDGLIRKSASMGNLSHYAGSTSGHHQNGVSGNNSV
SPGETPEHGAAADGYASEGFVPGSSSSRERKKGTPWTEEEHRMFLLGLQKLGKGDWRGISRNYVI
SRTPTQVASHAQKYFIRQTNVSRRKRRSSLFDIVADESVDTPMVSRDFFSTNPSQAETLSNNPLP
VPPALDEECESMDSTNSNDGEPPIPKPDGLQGCPPVIYPTYFSPFFPFSFPFWPGNSSEPTKMET
HEVLKPTAVHSKSPINVDELVGMSKLSLGESIGHAGPSSLTLKLLEGSSRQSAFHANPASGSSSM
NSSGSPIHAV

> GSVIVT00012218001 (DNA)

(SEQ ID NO: 61)

 $\tt ATGACTCGCCGCTGCTCGCATTGCAGTCACAACGGGCACAATTCCAGGACATGCCCCAACCGCGG$ GGTCAAGATCTTCGGGGTTCGATTGACTGATGGGTTGATCCGTAAGAGTGCTAGTATGGGCAATC ${\tt TCAGCCACTACGCCGGGTCGACCTCTGGTCATCATCAGAACGGCGTTTCCGGTAACAATTCGGTC}$ $\tt TCTCCCGGAGAGACTCCAGAGCACGGCGCCGCGGCCGATGGATACGCCTCCGAGGGTTTCGTTCC$ $\tt CGGTTCATCATCCAGCCGGGAGCGCAAGAAAGGCACTCCATGGACTGAAGAGAACACAGAATGT$ $\tt TTCTACTTGGACTGCAGAAGCTTGGAAAAGGGGATTGGCGTGGAATTTCACGTAATTATGTTATA$ ${\tt TCAAGGACACCTACTCAAGTCGCCAGCCATGCTCAGAAATATTTCATCAGGCAAACCAATGTGTC}$ ${\tt TAGGAGAAAAAGACGGTCCAGCTTGTTTGATATTGTAGCTGATGAATCTGTTGACACTCCAATGG}$ ${\tt TATCACGGGATTTCTTCTCCACCAACCCTTCGCAAGCTGAAACACTAAGCAATAACCCATTGCCT}$ $\tt GTTCCTCCGGCTCTGGATGAAGAATGTGAATCAATGGATTCTACCAACTCGAATGATGAGAACC$ ${\tt ACCCATTCCAAAGCCGGATGGCTTACAAGGCTGTCCCCCAGTAATATATCCTACTTATTTCTCGC}$ ${\tt CATTCTTCCCATTTTCTTTTCCATTCTGGCCGGGAAACAGTTCAGAGCCAACTAAAATGGAGACT}$ TGCTTGAAGGGTCAAGCAGCAATCTGCTTTCCATGCTAATCCAGCCTCTGGCAGTTCAAGCATG AACTCGAGCGGCAGTCCAATCCATGCACCCAATGGGAAGATTCTGCTGGTATGGAGATTGTAG

>GSVIVT00012218001 (Protein)

(SEQ ID NO: 62)

MTRRCSHCSHNGHNSRTCPNRGVKIFGVRLTDGLIRKSASMGNLSHYAGSTSGHHQNGVSGNNSV
SPGETPEHGAAADGYASEGFVPGSSSSRERKKGTPWTEEEHRMFLLGLQKLGKGDWRGISRNYVI
SRTPTQVASHAQKYFIRQTNVSRRKRRSSLFDIVADESVDTPMVSRDFFSTNPSQAETLSNNPLP
VPPALDEECESMDSTNSNDGEPPIPKPDGLQGCPPVIYPTYFSPFFPFSFPFWPGNSSEPTKMET
HEVLKPTAVHSKSPINVDELVGMSKLSLGESIGHAGPSSLTLKLLEGSSRQSAFHANPASGSSSM

NSSGSPIHAPNGKILLVWRL

> LjT37H17.80.nd (DNA)

(SEQ ID NO: 63)

ATGACCCGGCGATGCTCGCATTGCAGCCATGGTGGCCACAACGCCAGGACCTGCCCCAACCGCGG ${\tt AGTCAAGCTTTTCGGTGTCCGATTGACTGATGGCTCGATCCGGAAGAGTGCTAGTATGGGTAATC}$ ${\tt TCACCCACTACACTGGCTCCGGGTCTGGACCTCTTCTTGGTGGGTCCAATAACCCTGATTCTCCC}$ $\tt GGTGAAACCCCTGATCACGCCGCCGCTGCTGACGGTTACGCCTCTGAGGATTTTGTTCCTGGCTC$ $\tt TTCTTCTAGCTCCCGTGAAAGAAAAAAGGGCACTCCATGGACTGAGGAGGAACACAGAATGTTTT$ ${\tt TACTTGGATTGCAGAAACTGGGCAAAGGTGATTGGCGTGGAATTGCAAGGAACTATGTTATTTCA}$ ${\tt AGGACACCTACTCAAGTGGCCAGTCATGCTCAGAAATATTTCATCAGGCAAAGCAATGTGTCTAG}$ GAGAAAGAGACGGTCCAGCTTGTTTGATATTGTTGCAGATGATGCGTCCGACACTCCAATGGTAG AGCAAGACTTCTTGTCAGCTAATCAGCTACAGACTGAAACAGAAGGCCAATAACCCTTTGCCTGCT $\tt CCTCCTCCCATTGATGAAGAGTGTGAATCCATGGATTCCACAAACTCAATAGATGGAGACTCTGC$ CCTGTTAAAGCCCGACACTCCAATACCGCCAACCTACCCGGTGTTATATCCTGCATATTATCCTC CATTCTACCCGTATCCTCTGCCTTATTGGTCTGGATACAGTCCTGCAGAGCCCCCAAAGAAAAGAG GAAAACTCGTCGAAGAAGGACCTTCCAGACAATCAGCTTTTCATGCTACTCCAGCATGTGGCAGT TCAAATATAAATGGCAGTGTCATACATGCAGTTTAA

>LjT37H17.80.nd (Protein)

(SEQ ID NO: 64)

MTRRCSHCSHGGHNARTCPNRGVKLFGVRLTDGSIRKSASMGNLTHYTGSGSGPLLGGSNNPDSP
GETPDHAAAADGYASEDFVPGSSSSSRERKKGTPWTEEEHRMFLLGLQKLGKGDWRGIARNYVIS
RTPTQVASHAQKYFIRQSNVSRRKRRSSLFDIVADDASDTPMVEQDFLSANQLQTETEGNNPLPA
PPPIDEECESMDSTNSIDGDSALLKPDTPIPPTYPVLYPAYYPPFYPYPPLPYWSGYSPAEPPKKE
ETHEVVKPTAVLSKSPINVDELVGMSKLSLGDSIGDSGPSSLSRKLVEEGPSRQSAFHATPACGS
SNINGSVIHAV

>chr2.CM0028.230.nd (DNA)

TTATTAGTGTTGCTTGA

(SEQ ID NO: 65)

ATGTCTCGCACGTGCTCACAGTGCGGCAACAACGGCCACAACTCCCGCACATGCACCGACACCGC
CGCCGCTGGAGACAACGGCATCATGCTCTTCGGCGTGCGCCTCACCGAAGGCTCCACCTCCTC
CCGCCTTCATCAGGAAGAGCGCTAGCATGAACAACCTCTCCCAGTATAACGAACCCGAATCCAAC
CCCGCTGACGCAGCTGGCTACGCCTCCGACGACGTCGTTCATCCCTCCGCACGCCCCGCGACCG
CAAGCGAGGTGTGCCTTGGACGGAAGAAGAACACAAACTGTTTCTGTTGGGATTGCATAAAGTGG
GGAAGGGAGATTGGAGAGGAATTTCTAGAAACTTCGTCAAAACTCGCACACCCACTCAGGTTGCT
AGTCATGCTCAGAAGTATTTCCTCCGCCGTCACAACCATAACCGCCGGCGCCGGAGATCTAGCCT
TTTCGACATCACCACCGATACGGTGATGGAATCTTCAACAATAATGGAGGAAGAACAAGATCAGC
CAAGCGTTTCCAATGGCTACCTCCGCCGTGTATCCGCCGTTACATTACGGTGGCTTCCACGGC
CCAGCGTTTCCAATGGCTCTGTCTCCGGTGGTATTGCCGGTGGCCGGAGGGAAAACCGGCAAG
GCCGATTAGGCCAACGCCGATTTTCCCTGTGCCTCCGTCTTCTAAGATGGCTAGTTTGAACTTGA
AAGAGAAAGCAGCTTCTCCTTCCCCTTTCTTCCATTTGAGCCTCTACCGCTGTCGCCTGAAGCTG
CAGCCATCTCCGCCGCCGTCCAAGGATCATTCTCCGGCAACCAGTAGCCACTCGTCGCCATCATC
GCCGTCTTCTTCATCATCTTTTCAGGCTATGTCTGCAGGGAAGGTTCAGCGGTGGGAGAGATAGCA

>chr2.CM0028.230.nd (Protein)

(SEQ ID NO: 66)

15

20

MSRTCSQCGNNGHNSRTCTDTAAAGDNGIMLFGVRLTEGSTSSSAFIRKSASMNNLSQYNEPESN

PADAAGYASDDVVHPSARARDRKRGVPWTEEEHKLFLLGLHKVGKGDWRGISRNFVKTRTPTQVA

 ${\tt SHAQKYFLRRHNHNRRRRSSLFDITTDTVMESSTIMEEEQDQQEMVPPATSAVYPPLHYGGFHG}$

 ${\tt PAFPMALSPVVLPVAGGERPARPIRPTPIFPVPPSSKMASLNLKEKAASPSPSSPFEPLPLSLKL}$

QPSPPPSKDHSPATSSHSSPSSPSSSSSFQAMSAGKFSGGGDSIISVA

As shown below, SEQ ID NOs: 44, 46, 48, ..., and 66 share at least 44% homology with SEQ ID NO: 1. Also see FIG. 5 for a multiple protein sequence alignment.

No.	Species	Gene ID	Identities (%)	Positives (%)
1	Arabidopsis thaliana	AT5G47390	56	66
2	Arabidopsis thaliana	AT3G16350	45	55
3	Zea May	GRMZM2G034110_T01	86	90
4	Zea May	GRMZM2G020934_T01	62	69
5	Sorghum bicolor (Sorghum)	Sb01g029020.1	87	90
6	Sorghum bicolor (Sorghum)	Sb03g003270.1	44	56
7	Glycine max (Soybean)	Glyma17g15330.1	61	72
8	Glycine max (Soybean)	Glyma05g04950.1	58	71
9	Vitis vinifera (Wine Grape)	GSVIVT00013475001	62	71
10	Vitis vinifera (Wine Grape)	GSVIVT00012218001	62	71

-continued

; _	No.	Species	Gene ID	Identities (%)	Positives (%)
	11	Lotus japonicus (Lotus)	LjT37H17.80.nd	59	71
)	12	Lotus japonicus (Lotus)	chr2.CM0028.230.nd	57	68

As used herein, "percent homology" of two sequences is determined using the algorithm described in Karlin and Atts-25 chul, Proc, Natl. Acad. Sci. USA 87:2264-2268, 1990, modified as described in Karlin and Altschul, Proc, Natl. Acad. Sci. USA 90:5873-5877, 1993. Such an algorithm is incorporated into the NBLAST and XBLAST programs of Altschul et al., J Mol. Biol. 215:403-410, 1990. BLAST protein searches are 30 performed with the XBLAST program, score=50, wordlength=3, to obtain amino acid sequences homologous to a reference polypeptide. To obtain gapped alignments for comparison purposes, Gapped BLAST is utilized as described in Altschul et al., Nucleic Acids Res. 25:3389-3402, 1997. When utilizing the BLAST and Gapped BLAST programs, the default parameters of the respective programs (e.g., XBLAST and NBLAST) are used. See the NCBI website at ncbi.nlm,nih.gov.

In a preferred embodiment, the cell also has a DREB1 gene expression level that is higher than a wild type DREB1 protein level so that the plant over-expresses both MYBS3 and DREB1. Listed below are cDNA and protein sequences of DREB1A, 1B, 1C, and 1D.

OsDREB1A (dehydration-responsive element-binding protein 1A) LOC_Os09g35030 CDS >12009.m06503

-continued

Protein >12009.m06503

(SEQ ID NO: 4)

 $\verb|MCGIKQEMSGESSGSPCSSASAERQHQTVWTAPPKRPAGRTKFRETRHPVFRGVRRRGNA|$

 ${\tt GRWVCEVRVPGRRGCRLWLGTFDTAEGAARAHDAAMLAINAGGGGGGGGACCLNFADSAWL}$

 ${\tt LAVPRSYRTLADVRHAVAEAVEDFFRRRLADDALSATSSSSTTPSTPRTDDDEESAATDG}$

DESSSPASDLAFELDVLSDMGWDLYYASLAQGMLMEPPSAALGDDGDAILADVPLWSY*

OsDREB1B (dehydration-responsive element-binding protein 1B)LOC Os09g35010

CDS >12009.m06501

(SEQ ID NO: 5)

 $\tt ATGGAGGTGGAGGGGGGGGTACAGGACGGTGTGGTCGGAGCCGCCGAAGAGGCCGGCG$

GGAAGGACCAAGTTCAGGGAGACGAGGCACCCGGTGTACCGCGGCGTGCGGCGCGCGG

 $\tt GCCGCGCTGGCGCTCCGCGGCAGGCCGCCTGCCTCAACTTCGCCGACTCCGCGTGGCGG$

ATGCCGCCGTCCCCGCGTCCGCCGCGCTCGCCGGCGCGAGGGGGGTCAGGGACGCCGTC

 $\tt GCGTGGTGGGAGGACGGCGAGCTCGGCGGCTCCGACATGCCGCTCTGGAGCTACTAA$

Protein >12009.m06501

(SEQ ID NO: 6)

MEVEEAAYRTVWSEPPKRPAGRTKFRETRHPVYRGVRRRGGRPGAAGRWVCEVRVPGARG

 $\tt SRLWLGTFATAEAAARAHDAAALALRGRAACLNFADSAWRMPPVPASAALAGARGVRDAV$

 ${\tt AVAVEAFQRQSAAPSSPAETFANDGDEEEDNKDVLPVAAAEVFDAGAFELDDGFRFGGMD}$

 ${\tt AGSYYASLAQGLLVEPPAAGAWWEDGELAGSDMPLWSY*}$

OsDREB1C (dehydration-responsive element-binding protein 1C)LOC_0s06g03670

CDC -13106 m0030F

(SEQ ID NO: 7)

GCGGGGAGGACCAAGTTCAGGGAGACGAGGCACCCGGTGTACCGCGGCGTGCGGCGGCGG

 $\tt GGGCCCGCGGGGCGTGTGTGCGAGGTCAGGGAGCCCAACAAGAAGTCCCGCATCTGG$

GCCGCCGCCGCCCCCATGATGATGCAGTACCAGGACGACATGGCGGCGACGCCGTCC

GCCGGCGGCTACGGCGGCGGCGACGTCACACTCTGGAGCTACTGA

Protein >13106.m00305

(SEO ID NO: 8)

MEYYEQEEYATVTSAPPKRPAGRTKFRETRHPVYRGVRRRGPAGRWVCEVREPNKKSRIW

LGTFATAEAAARAHDVAALALRGRGACLNFADSARLLRVDPATLATPDDIRRAAIELAES

CPHDAAAAAASSSAAAVEASAAAAPAMMMOYODDMAATPSSYDYAYYGNMDFDOPSYYYD

 ${\tt GMGGGGEYQSWQMDGDDDGGAGGYGGGDVTLWSY*}$

-continued

OsDREB1D (dehydration-responsive element-binding protein 1D)LOC_Os06g06970 CDS >13106.m00721

(SEQ ID NO: 9) ATGGAGAAGAACACCGCCGCGAGGGGCAATTGATGACCTCCTCCGCGAGGGGAGCGACGCCG

TCGTCGCCGAAGCGGCCGGCGGGGCGAACCAAGTTCCAGGAGACGAGGCACCTAGTGTTC

CGTGGGGTGCGATGGCGTGGGTGCGCGGGGCGGTGGGTGTGCAAGGTGCGTGTCCCGGGC

AGCCGCGGTGACCGTTTCTGGATAGGCACGTCTGACACCGCCGAGGAGACCGCGCGCACG

CACGACGCCGCCATGCTCGCCTTGTGCGGGGCCTCCGCCAGCCTCAACTTCGCCGACTCT

GCCTGGCTGCTCCACGTCCCGCGCCCCCGTCGTCTCCCGGACTCCGGCCACCAGCTGCC

ATGTCAAAGCTTATCAGCAGTAGCAGAGCAAAAGGATCGTTGTGCCTGCGAAAAAATCCC

ATTTCATTTTGCATGGTTACAAATTCTTACACTGCTCTTTTTGCTCGAATACATTATATTG

CAGATGAATTCAATGATCGTTTTAATCCACGAATTATCAAAATATCAAGTCTTTCTGCTA

CTAACCATGATAACACACCACCTTTTTCAATGGAGGAGGTAG

Protein >13106.m00721

(SEQ ID NO: 10)

 ${\tt MEKNTAASGQLMTSSAEATPSSPKRPAGRTKFQETRHLVFRGVRWRGCAGRWVCKVRVPG}$

 $\tt SRGDRFWIGTSDTAEETARTHDAAMLALCGASASLNFADSAWLLHVPRAPVVSGLRPPAA$

 ${\tt RCATRCLQGHRRVPAPGRGSTATATATSGDAASTAPPSAPVLSAKQCEFIFLSSLDCWML}$

MSKLISSSRAKGSLCLRKNPISFCMVTNSYTALLLEYIILQMNSMIVLIHELSKYQVFLL

LTMITHHLFQWRR*

The above-mentioned level of wild type MYBS3 or DREB1 protein can be the MYBS3 or DREB1 gene expression level in a wild type cell of a wild type plant. To achieve this overexpression, the genes can be under the control of native, constitutive, tissue-specific, developmental stage-specific, or other inducible promoters.

The above-described transgenic plant can be rice, maize, wheat, barley, sorghum, sugarcane, turf grass, Miscanthus, switchgrass, soybean, canola, potato, tomato, bean, pea, or jatropha. The transgenic plant is more tolerant to chill than the wild type plant.

In a second aspect, the invention features a method of generating the above-described plant. The method includes steps of introducing into a cell of a plant a first nucleic acid that encodes a polypeptide containing the amino acid sequence of MYBS3 protein; and expressing the MYBS3 protein in the cell. The level of the MYBS3 protein in the cell is higher than a wild type MYBS3 protein level. In one 55 embodiment, the method further includes introducing into the cell of the plant a second nucleic acid that encodes a second polypeptide containing the amino acid sequence of DREB1 protein; and expressing the DREB1 protein in the cell. The level of the DREB1 protein in the cell is higher than a wild 60 type DREB1 protein level.

In a third aspect, the invention features a method of identifying a plant that is tolerant to chill. The method can be carried out by obtaining a sample from a candidate plant, and determining the MYBS3 gene expression level in the sample. 65 The candidate plant is determined to be tolerant to chill if the expression level is above a predetermined level. The prede-

termined level is the level in a wild type plant. With this method, one can use MYBS3 as a marker for molecular breeding plants and selecting chill tolerant plants.

The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims.

BRIEF DESCRIPTION OF DRAWINGS

FIG. 1 shows two constructs for respectively over-expressing and under-expressing MYBS3 in transgenic rice; namely, pSMY-MYBS3(Ox), an over-expression construct; and pSMY-MYBS3(Ri), an under-expression construct. P_{A1} , Hgh, T_{Nos} , P_{Ubi} , LB, and RB denote A1 promoter, hygromycin phosphotransferase, nopalin synthase terminator, Ubi promoter, left border of T-DNA, and right border of T-DNA, respectively.

FIG. 2 is a bar graph showing expression of MYBS3 and DREB1A in 10-day-old rice seedlings that were shifted from 28° C. to 4° C. and incubated for 72 h. Total RNAs were isolated from seedlings and subjected to quantitative real-time RT-PCR analysis. The highest mRNA level was assigned a value of 100, and the mRNA levels of other samples were calculated relative to this value. The error bars indicate the SE of three replicates.

FIG. 3 includes two photographs showing 10-day-old seedlings of wild type (WT) and lines S3(Ox)-110-1 and S3(Ri)-52-7 that were incubated at 4° C. for 8 h (A) or 24 h (B).

FIG. 4 is a schematic flow chart for a proposed role of MYBS3 in cold stress tolerance in rice.

FIG. 5 is a sequence alignment of thirteen MYB3 proteins, one from rice and two from each of *Arabidopsis thaliana*, *Zea May*, sorghum, soybean, twine grape, and lotus.

DETAILED DESCRIPTION

This invention is based, at least in part, on the unexpected discoveries of that over-expression of MYBS3 in a plant 10 increased the cold tolerance.

MYBS3 is a single DNA-binding repeat (1R) MYB transcription factor previously shown to mediate sugar signaling in rice. MYBS3 also plays a critical role in cold adaptation in rice. Gain- and loss-of-function analyses indicates that 15 MYBS3 is sufficient and necessary for enhancing cold tolerance in rice. Transgenic rice constitutively over-expressing MYBS3 tolerates 4° C. for at least 1 week, and exhibits no yield penalty in normal field conditions. Transcription profiling of transgenic rice over- or under-expressing MYBS3 lead 20 to identification of many genes in the MYBS3-mediated cold signaling pathway. Several genes activated by MYBS3 as well as inducible by cold have previously been implicated in various abiotic stress response and/or tolerance in rice and other plant species. Surprisingly, MYBS3 represses the well- 25 known DREB1/CBF-dependent cold signaling pathway in rice, and the repression appears to act at the transcriptional level. DREB1 responds quickly and transiently while MYBS3 responds slowly to cold stress, which suggests distinct pathways act sequentially and complementarily for 30 adapting short- and long-term cold stress in rice. This novel pathway, which controls cold adaptation in rice as well as other plants, forms the basis for this invention.

A transgenic plant described in the invention can be generated by introducing into the plant or a part thereof an 35 expression construct comprising a DNA sequence encoding a MYB3 protein, a DREB1 protein, or both. Expression constructs are provided by the present invention for the stable transformation of plants with a gene encoding a MYB3 protein, a DREB1 protein, or both. These constructs comprise a 40 DNA sequence encoding a MYB3 protein or a DREB1 protein which is operably linked to regulatory sequences which are capable of directing the expression of a MYB3 protein, a DREB1 protein, or both. These regulatory sequences may also include sequences capable of directing transcription in 45 plants, either constitutively, or stage and/or tissue specific, depending on the use of the plant or parts thereof. The expression constructs provided may be inserted into a vector, preferably a plasmid, used in bacteria-mediated transformation of the selected plant host. The expression construct is then preferably integrated into the genome of the plant host.

A transgene is a nucleic acid sequence (encoding, e.g., one or more subject polypeptides), which is partly or entirely heterologous to a plant cell into which it is introduced, or, is homologous to an endogenous gene of the plant or cell into 55 which it is introduced but is intended to be inserted into the plant genome in such a way as to alter the genome (e.g., it is inserted at a location which differs from that of the natural gene or its insertion results in a knockout). A transgene can include one or more operably linked transcriptional regulatory sequences (e.g., an enhancer sequence) and any other nucleic acid, such as an intron, that may be necessary for optimal expression of a nucleic acid of interest.

A "transformed" and "transgenic cell refer to a host cell or organism into which a heterologous nucleic acid molecule 65 has been introduced. The nucleic acid molecule can be stably integrated into the genome generally known in the art.

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Accordingly, a transgenic cell is a cell containing a transgene. A transgenic plant is any plant in which one or more, or all, of the cells of the plant include a transgene. The transgene can be introduced into the cell by introduction into a precursor cell by way of deliberate genetic manipulation, such as by T-DNA mediated transfer, electroporation, or protoplast transformation. The transgene may be integrated within a chromosome, or it may be an extrachromosomally replicating DNA.

The term "heterologous" refers to portions of a nucleic acid and indicates that the nucleic acid comprises two or more subsequences that are not found in the same relationship to each other in nature. For instance, a nucleic acid that is recombinantly produced typically has two or more sequences from unrelated genes synthetically arranged to make a new functional nucleic acid, e.g., a promoter from one source and a coding region from another source. The two nucleic acids are thus heterologous to each other in this context. When added to a cell, the recombinant nucleic acids would also be heterologous to the endogenous genes of the cell. Thus, in a chromosome, a heterologous nucleic acid would include a non-native (non-naturally occurring) nucleic acid that has integrated into the chromosome, or a non-native (non-naturally occurring) extra-chromosomal nucleic acid. In contrast, a naturally translocated piece of chromosome would not be considered heterologous in the context of this patent application, as it comprises an endogenous nucleic acid sequence that is native to the mutated cell. A heterologous nucleic acid, gene, or protein can be one that originates from a foreign species, or, if from the same species, is substantially modified from its original form. For example, a maize ubiquitin (Ubi) promoter operably linked to a nucleic acid sequence encoding a rice MYB3 protein is one form of a sequence heterologous to rice. If a promoter and a coding sequence are from the same species, one or both of them can be substantially modified from their original forms.

Rice is one of the most important food crops in the world, and increases in rice yield could significantly ease the pressure on world food production. Rice is also a powerful model for functional genomics study for dissecting genetic networks of stress responses in cereal crops. Low temperatures are one of the major environmental stresses that adversely affect rice productivity in temperate and subtropical zones and in high elevation areas. Rice seedlings are particularly sensitive to chilling in early spring in these areas, leading to slow seedling development, yellowing, withering, reduced tillering and stunted growth (Andaya, et al., 2003, J Exp Bot 54: 2579-2585). Rice cannot be grown in approximately 7,000,000 hectares of land in south and south-east Asia due to cold stress (Sthapit, et al., 1998, Crop Sci 38: 660-665); in temperate regions such as California (USA), cold is an important stress that results in delayed heading and yield reduction due to spikelet sterility (Peterson, et al., 1974, California Agriculture, 28(7), 12-14). Thus, improvement of chilling tolerance may significantly increase rice production.

Plants respond and adapt to cold stress at the molecular and cellular levels as well as induce an array of biochemical and physiological alterations that enable them to survive (Bohnert et al., 1995, Cell 7: 1099-1111; and Browse, et al., 2001, Curr Opin Plant Biol 4: 241-246). Under cold stress, the expression of many genes is induced in various plant species (Hughes, et al., 1996, J Exp Bot 47: 291; and Thomashow, 1999, Annu Rev Plant Physiol Plant Mol Biol 50: 571-59), and the products of these genes function not only in adaptations promoting stress tolerance, e.g., biosynthesis of osmotica (Chen, et al., 2002, Curr Opin Plant Biol 5: 250-257; and Taji, et al., 2002, Plant J 29: 417-426), generation of antioxidants (Prasad, et al., 1994, Plant Cell 6: 65-74), and increased

membrane fluidity (Murata and Los, 1997, Plant Physiol 115: 875-879; and Orvar, et al., 2000, Plant J 23: 785-794), but also in the regulation of gene expression and signaling transduction in stress responses, e.g., transcription factors and proteins involved in RNA processing and nuclear export 5 (Yamaguchi-Shinozaki, et al., 2006, Annu Rev Plant Biol 57: 781-803; and Chinnusamy, et al., 2007, Trends Plant Sci 12: 444-451). Deciphering the mechanisms by which plants perceive and transmit cold signals to cellular machinery to activate adaptive responses is of critical importance for developing breeding strategies to enhance cold stress tolerance in crops.

In Arabidopsis and rice, the CBF/DREB1-dependent cold response pathway has been shown to play a predominant role in freezing-tolerance through the process of cold acclimation 15 (Thomashow, 1999, Annu Rev Plant Physiol Plant Mol Biol 50: 571-59; Yamaguchi-Shinozaki, et al., 2006, Annu Rev Plant Biol 57: 781-803; and Chinnusamy, et al., 2007, Trends Plant Sci 12: 444-451). The DREB1/CBF family, including DREB1A/CBF3, DREB1B/CBF1, and DREB1C/CBF2, are 20 able to bind to and activate the cis-acting elements DRE (dehydration-responsive element) (Yamaguchi-Shinozaki, et al., 1994, Plant Cell 6: 251-264; and Stockinger, et al., 1997, Proc Natl Acad Sci USA 94: 1035-104) or CRT (C-repeat) (Baker, et al., 1994, Plant Mol Biol 24: 701-713) on promot- 25 ers of several cold-responsive genes (CORs) (Gilmour, et al., 1998, Plant J 16: 433-442; Jaglo-Ottosen, et al., 1998, Science 280: 104-106; Liu, et al., 1998, Plant Cell 10: 1391-1406; Medina, et al., 1999, Plant Physiol 119: 463-470).

Rice DREB1A and DREB1B are induced by cold stress, 30 and constitutive over-expression of these genes leads to induction of stress-responsive genes, increased tolerance to high-salt and cold, and growth retardation under normal conditions in transgenic Arabidopsis and rice (Dubouzet, et al., 2003, Plant J 33: 751-763; and Ito, et al., 2006, Plant Cell 35 Physiol 47: 141-153), indicating the evolutionary conservation of the DREB1/CBF cold-responsive pathway in monocots and dicots. However, in comparison to Arabidopsis and other cereals like wheat and barley that cold acclimate (Wen, et al., 2002, Plant Physiol 129: 1880-1891), rice does not 40 undergo acclimation process and is more sensitive to low temperature exposures. Microarray analysis demonstrated the existence of 22 cold-regulated genes in rice, which have not been reported in Arabidopsis (Rabbani, et al., 2003, Plant Physiol 133: 1755-1767). These studies also indicate that 45 plant species vary in their abilities to adapt to cold stress.

Other rice proteins have also been shown to be involved in cold tolerance. For example, a zinc-finger protein iSAP1 confers cold, dehydration, and salt tolerance in transgenic tobacco (Mukhopadhyay, et al., 2004, Proc Natl Acad Sci 50 USA 101: 6309-6314); the rice MYB4 transcription factor confers chilling and freezing tolerances by enhancing the COR gene expression and proline accumulation in Arabidopsis (Vannini, et al., 2004, Plant J 37: 115-127), and improves cold and drought tolerances by accumulating osmolyte in 55 transgenic apples (Pasquali, et al., 2008, Plant Cell Rep 27: 1677-1686). Overexpression of the rice cold-, drought, and salt-inducible MYB3R-2 (an R1R2R3 MYB) gene enhances cold, drought, and salt tolerance by regulating some stressresponsive genes involved in the CBF-dependent or CBF- 60 independent pathways in Arabidopsis (Dai, et al., 2007, Plant Physiol 143: 1739-1751; and Ma, et al., 2009, Plant Physiol 150: 244-256).

The expression of DREB1 is subjected to regulation by several factors. For example, it is affected by members in the 65 same DREB1 family. The *Arabidopsis* cbf2 mutant, in which CBF2/DREB 1 C is disrupted, shows higher freezing, dehy-

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dration and salt tolerance than the wild-type plant, indicating that DREB1C/CBF2 acts as a repressor of CBF1/DREB1B and CBF3/DREB1A expression (Novillo, et al., 2004, Proc Natl Acad Sci USA 101: 3985-3990). The expression of DREB1/CBF is activated by Inducer of CBF Expression 1, ICE1 (a MYC-like basic helix-loop-helix-type transcription factor) (Chinnusamy, et al., 2003, Genes Dev 17: 1043-1054), CAX1 (a Ca²⁺/H⁺ transporter) (Catala, et al., 2003, Plant Cell 15: 2940-2951), CBL1 (a Ca²⁺ sensor) (Albrecht, et al, 2003, Plant J 36: 457-470), and LOS4 (a DEAD-box RNA helicase) (Gong, et al., 2002, Proc Natl Acad Sci USA 99: 11507-11512), and repressed by FRY2 (a transcription factor) (Xiong, et al., 2002, Proc Natl Acad Sci USA 99: 10899-1090), HOS1 (a putative RING finger E3 ligase) (Lee, et al., 2001, Genes Dev 15: 912-924), and ZAT12 (a C₂H₂ zinc finger transcription factor) (Vogel, et al., 2005, Plant J 41: 195-211), during cold acclimation in Arabidopsis. The mechanism by which these factors affect the expression of CBF/DREB1 is not clear.

Previously, three MYB transcription factors, MYBS1, MYBS2, and MYBS3 each with a single DNA binding domain (1R MYB), were identified in rice and shown to bind specifically to the TA box (TATCCA) in the sugar response complex (SRC) of α-amylase gene (αAmy3) promoter (Lu, et al., 2002, Plant Cell 14: 1963-1980). MYBS1 and MYBS2 transactivate, while MYBS3 represses, the sugar starvationinducible a Amy3 SRC activity in rice (Lu, et al., 2002, Plant Cell 14: 1963-1980). The rice MYBS3 homologue in Arabidopsis (AGI code: At5g47390) is activated by ABA, CdCl₂ and NaCl (Yanhui, et al., 2006, Plant Mol Biol 60: 107-124). It is unexpected that the expression of MYBS3 is induced by cold. By both gain- and loss-of-function analyses, MYBS3 is shown essential for cold stress tolerance in rice. Transcription profiling of transgenic rice over- or under-expressing MYBS3 led to identification of genes that are activated or repressed by MYBS3 and play diverse functions. The DREB1-dependent cold response signaling pathway is among those repressed by MYBS3 in rice. This finding suggests that the DREB1- and MYBS3-dependent pathways may complement each other and act sequentially to adapt to immediate and persistent cold stress in rice.

The specific examples below are to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever. Without further elaboration, it is believed that one skilled in the art can, based on the description herein, utilize the present invention to its fullest extent. All publications cited herein are hereby incorporated by reference in their entirety. Further, any mechanism proposed below does not in any way restrict the scope of the claimed invention.

EXAMPLE

Materials and Methods

Plant Materials

The rice cultivar *Oryza sativa* L. cv. Tainung 67 was used in this study. Induction of rice calli was performed as described (Yu, et al., 1991, J Biol Chem 266: 21131-21137). For hydroponic culture of rice seedlings, seeds were sterilized with 3% NaOCl for 30 min, washed extensively with distilled water, and germinated in Petri dishes with wetted filter papers at 37° C. in the dark. After 48 h incubation, germinated seeds were cultivated in a half-strength Kimura B solution containing the following macronutrients: (NH4) $_2$ SO $_4$ (0.18 mM), KNO $_3$ (0.09 mM), MgSO $_4$ (0.27 mM), KH $_2$ PO $_4$ (0.09 mM), and Ca(NO $_3$) $_2$ (0.18 mM), and micronutrients: Fe-citrate (0.03 $_4$ M), H $_3$ BO $_3$ (2.5 $_4$ M), MnSO $_4$ H $_2$ O (0.2 $_4$ M), ZnSO $_4$.7H $_2$ O

Primers

 $(0.2~\mu M),~CuSO_4.5H_2O~(0.05~\mu M),~and~H_2MoO_4~(0.05~\mu M).$ The pH of the solution was adjusted to 4.7-4.8 using 0.5 N HCl. The culture solution was replaced with fresh solution every 2 days. Seedlings were grown under a 14-h light/10-h dark cycle for 10 days in a 28° C. chamber before treatments.

All primers used for the cloning of cDNAs or plasmid constructions and quantitative RT-PCR are listed in Table 1 below.

TABLE 1

-	Primers		
Target gene	Primer	Annealing temperature (° C.)	
MYBS3	S3F1: 5'-CCT TTCTGGCAA AATCAG AAAGA-3' (SEQ ID NO: 11) S3R1: 5'-ATG AACTGG AACAGGCTTGACA-3' (SEQ ID NO: 12)	60	78
MYBS3 promoter	S3PF: GCGGATCCCCTTTTGACTTGCAGGTTAATTACTTCAGG (SEQ ID NO: 13) S3PR: CATGCCATGGTTTAAACCCCCCCTCCGTCCCTCCACCTTCC (SEQ ID NO: 14)	68	2500
DREB1A	1AF: 5'-GGACCTGTACTACGCGAGCTT-3' (SEQ ID NO: 15) 1AR: 5'-GGCAAAATTGTACAGTTGATTGA-3' (SEQ ID NO: 16)	60	138
DREB1A promoter	1APro: 5'-TTGACCGGGATACCGAATTA-3' (SEQ ID NO: 17) 1APro: 5-GTAATGGCGATGGGAGAAGA-3' (SEQ ID NO: 18)	60	1054
DREB1B	1BF: 5'-AGCTCGCCGGCTCCGACA-3' (SEQ ID NO: 19) 1BR: 5'-GGGAGAAATCTGGCACATTCC-3' (SEQ ID NO: 20)	60	194
DREB1B promoter	1BPro: 5'-AGGTAAGCCATTAGCGCATG-3' (SEQ ID NO: 21) 1BPro: 5'-GGATGACTCTCTCTGGTTCA-3' (SEQ ID NO: 22)	60	747
DREB1C	1CF: 5'-GAGTTGGAGCTAGCAGTTTTGAG-3' (SEQ ID NO: 23) 1CR: 5'-TAGCTGTATAGGAGGAGCAAAGC-3' (SEQ ID NO: 24)	60	54
Amy3	Amy3F: 5'-GTAGGCAGGCTCTCTAGCCTCTAGG-3' (SEQ ID NO: 25) Amy3R: 5'-GTAGGCAGGCTCTCTAGCCTCTAGG-3' (SEQ ID NO: 26)	60	112
Cytochrome P450	CytF: 5'-GTCATCCAGGAGACGATGAGG-3' (SEQ ID NO: 27) CytR: 5'-GATGTTGCGGAACAGAGGTAG-3' (SEQ ID NO: 28)	60	129
18S rRNA	18SF: 5'-CCTATCAACTTTCGATGGTAGGATA-3' (SEQ ID NO: 29) 18SR: 5'-CGTTAAGGGATTTAGATTGTACTCATT-3' (SEQ ID NO: 30)	60	229
Trehalose-6- phosphate phosphatase 3 gene (Os02g44230)	T6PP1 F: 5'-GGAGTTCCTCAATTTCTTGGTG-3' (SEQ ID NO: 31) T6PP1 R: 5'-CGCCTCGGAAACTACAGTTATT-3' (SEQ ID NO: 32)	60	116
Trehalose-6- phosphate phosphatase 2 gene (Os10g40550)	T6PP F: 5'-AGGATGCATTCAAGGTTCTGA-3' (SEQ ID NO: 33) T6PP R: 5'-CAAGATGCCAGTTTCTTCAGG-3' (SEQ ID NO: 34) 2	60	139
GFP	GFP_F: 5'-CCTGTCCTTTTACCAGACAACC-3' (SEQ ID NO: 35) GFP_R: 5'-GGACCATGTGGTCTCTTTTC-3' (SEQ ID NO: 36)	60	85
Multidrug- resistantce (0s01g50100)	MRT_F: 5'-CAGGCAGAGGAACAGGTGAT-3' (SEQ ID NO: 37) MRT_R: 5'-CGTACCGGAACAAGCTGAAC-3' (SEQ ID NO: 38)	60	108
Glutamate decarboxylase (Os03g13300)	GD_F: 5' AAGACGCTGCTGATTGATATGAT-3' (SEQ ID NO: 39) eGD_R: 5'-TGGTAGCTCACACCATGAATGTA-3' (SEQ ID NO: 40)	60	50
WRKY77 (0s01g40260)	WRKY77_F: 5'-GGAATGGACAATTAGTTTGTCTCC-3' (SEQ ID NO: 41) WRKY77 R: 5'-ATATATCGATGGGCCGTAATTTT-3' (SEQ ID NO: 42)	60	73

Plasmid Construction

The GATEWAY gene cloning system (Invitrogen, Carlsbad, CA) was used to construct the MYBS3-GFP fusion gene. Briefly, the full-length cDNA of MYBS3 was inserted 65 between the attL1 and attL2 sites in pENTR/D-TOPO, generating the entry vector pENT-MYBS3. The CaMV35S pro-

moter upstream of GFP in pCAMBIA1302 (see the website at cambia.org.au/daisy/cambia/585.html) was replaced with the maize ubiquitin (Ubi) promoter, and the ccdB DNA fragment flanked by attR1 and attR2 sties was inserted between the Ubi promoter and GFP, generating the destination vector pDEST-GFP. MYBS3 in pENT-MYBS3 was then inserted upstream

Microarray Analysis

of GFP in pDEST-GFP through the GATEWAY lambda recombination system, generating p1302-MYBS3-GFP. The 2.5-kb MYBS3 promoter fragment (upstream of ATG) was PCR synthesized and used for replacement of the Ubi promoter in pDEST-GFP, generating the Ubi::MYBS3-GFP construct.

For generating constructs used for the embryo transient expression assay, the 1054-bp DREB1A and 747-bp DREB1B promoters (upstream of ATG) were PCR-synthesized and fused upstream of Luc cDNA in pLuc (Lu, et al., 1998, J Biol Chem 273: 10120-10131). Plasmid p3Luc.18 contains αAmy3 SRC (–186 to –82 upstream of the transcription start site) fused to the CaMV35S minimal promoter-Adh intron-Luc fusion gene (Lu, et al., 1998, J Biol Chem 273: 1510120-10131).

For generating constructs used for rice transformation, plasmid pBS-MYBS3 (Lu, et al., 2002, Plant Cell 14: 1963-1980) containing the Ubi promoter fused upstream of MYBS3 cDNA was linearized with EcoRI and inserted into 20 the binary vector pSMY1H (Ho, et al., 2000, Plant Physiol 122: 57-66), generating pSMY-MYBS3 (FIG. 1). To make the MYBS3 RNA interference construct, a 227-bp sequence derived from the 3'UTR of MYBS3 cDNA was synthesized by PCR, fused in antisense and sense orientations flanking the 25 750-bp GFP cDNA. This MYBS3 RNAi fragment was used to replace the MYBS3 cDNA in pUbi-MYBS3, generating pUbi-MYBS3(Ri). pUbi-MYBS3(Ri) was linearized with EcoRI and inserted into the binary vector pSMY1H, generating pSMY-MYBS3(Ri) (FIG. 1).

Rice Transformation

Plasmids p1302-MYBS3-GFP, pSMY-MYBS3, pSMY-MYBS3(Ri) as constructed above were introduced into *Agrobacterium tumefaciens* strain EHA101, and rice transformation was performed as described elsewhere (Ho, et al., 2000, 35 Plant Physiol 122: 57-66).

RNA Extraction and Real-Time Quantitative RT-PCR Analysis

Total RNA was extracted from leaves of rice seedlings with Trizol reagent (Invitrogen) and treated with RNase-free 40 DNase I (Promega, Madison, Wis.). Four µg of RNA was used for cDNA preparation with reverse transcriptase (Applied Biosystems, Foster City, Calif.), and cDNA was then diluted to 10 ng/µl. Five µl of cDNA was mixed with primers and the 2× Power SYBR Green PCR Master Mix reagent, and applied 45 to an ABI 7500 Fast Real-Time PCR System (Applied Biosystems). The quantitative variation between different samples was evaluated by the delta-delta CT method, and the amplification of 18S ribosomal RNA was used as an internal control to normalize all data.

Subcellular Localization of MYBS3-GFP Fusion Protein

Protoplasts were isolated from transformed calli as described (Lu, et al., 1998, J Biol Chem 273: 10120-10131). GFP expression was detected under a LSM510 confocal laser scanning microscope (Carl Zeiss) using a 40× objective lens 55 and the confocal microscopy software Release 2.8 (Carl Zeiss).

Stress Treatments

Ten-day-old seedlings cultured in the half-strength Kimura B solution at 28° C. and with 16-h light and 8-h dark cycle in 60 a growth chamber were used for all stress treatments. Stress treatments are as follows: ABA, seedlings were transferred to a culture solution containing $20\,\mu\text{M}$ ABA; drought, seedlings were air-dried until 10% or 30% of fresh weight was lost; cold, seedlings were transferred to 4° C.; salt, seedlings were transferred to a culture solution containing 200 mM NaCl; heat, seedlings were transferred to 45° C.

Total RNA was extracted from leaves of rice seedlings using the Qiagen RNeasy Plant Mini Kits (Qiagen, Valencia, Calif.) according to the Qiagen manual. RNA quality was examined by the Agilent 2100 bioanalyzer (Affymetrix, Palo Alto, Calif.), and biotinylated target RNA was prepared from total RNA. Samples were hybridized to the Affymetrix Rice GeneChip (Affymetrix, Santa Clara, Calif.) as described in the GeneChip Expression Analysis Technical Manual. Two biological replicates were performed for cold treated samples per time point.

The hybridization signals were scanned with an Affymetrix GeneChip scanner 3000 7G, and the cell intensity (CEL) files were obtained from software Affymetrix GCOS version 1.4 (Affymetrix). CEL files were loaded into GeneSpring GX 9.0 (Agilent Technologies, Palo Alto, Calif.). Filtering tools in the GeneSpring software were used to identify genes significantly up-regulated and down-regulated between different chips. All genes up-regulated or down-regulated by overexpression or under-expression of MYBS3 or by cold are listed in Tables S7-S11 of Su, et al, Plant Physiol. 2010; 153(1):145-58.

Accession Number

DREB1A (Os09g35030); DREB1B (Os09g35010); DREB1C (Os06g03670); αAmy3/RAmy3D (Os08g36910); Cytochrome P450 gene (Os02g47470); Glutamate decarboxylase gene (Os03g13300); WRKY77 (Os01g40260); Multidrug resistance protein 4 gene (Os01g50100); Trehalose-6-phosphate phosphatase 1 gene (Os02g44230); and Trehalose-6-phosphate phosphatase 2 gene (Os10g40550). Results

Expression of MYBS3 is Ubiquitous and Activated by Cold Stress

Expression of MYBS3 was found to be ubiquitous in all tissues in 7-day-old seedlings and 3-month-old mature plants (including spike, stem, sheath, leaf, senescent leaf, leaf in light, root in light, leaf in dark, and root in dark) and in suspension cells of rice cultured in media with or without sucrose for 2 days. The regulation of MYBS3 expression by various stresses was investigated by subjecting 10-day-old rice seedlings to ABA (20 μM), drought (air dry), cold (4° C.), salt (200 mM NaCl), and heat (45° C.) treatments. The accumulation of MYBS3 mRNA was induced by cold in roots and by cold and salt in shoots, but reduced by ABA in shoots. In 10-day-old rice seedlings, the expression pattern of MYBS3 and DREB1A under cold stress was further compared. The amount of MYBS3 mRNA was detectable at 28° C., and increased 5-fold at 4° C. after 72 h; in contrast, the accumulation of DREB1A mRNA was barely detectable at 28° C., increased drastically after shifting to 4° C. and peaked at 6 h, but declined to one fifth after 72 h (FIG. 2).

To determine whether MYBS3 is regulated by cold at the transcriptional level, the 2.5-kb MYBS3 promoter was fused to the reporter gene GFP encoding a green fluorescence protein and introduced into the rice genome. Ubi promoter fused to GFP was used as a control. Transgenic rice seedlings were grown at 4° C. Under the control of MYBS3 promoter, the accumulation of GFP mRNA was 2.5 times higher at 12 h and stayed high up to 24 h. In contrast, under the control of Ubi promoter, the accumulation of GFP mRNA decreased by nearly 50% at 6 h, and then stayed at similar levels up to 24 h. This result indicates that the MYBS3 promoter is activated by cold.

MYBS3 is a transcriptional repressor of αAmy3 SRC in rice suspension cells (Lu, et al., 2002, Plant Cell 14: 1963-1980). To determine whether MYBS3 is localized in nucleus, the Ubi promoter was fused to the MYBS3-GFP fusion DNA. The Ubi::MYBS3-GFP and Ubi::GFP constructs were introduced into the rice genome. Protoplasts were isolated from transformed calli, incubated at 4° C. or 28° C., and examined.

32 TABLE 3-continued

Accumulation of MYBS3-GFP was detected mainly in the nucleus, whereas GFP alone was distributed throughout the cell except the vacuole, at both 4° C. and 28° C., suggesting that MYBS3 is constitutively localized in the nucleus.

MYBS3 is Sufficient and Necessary for Cold Tolerance in 5

Since MYBS3 was induced by cold, its role in cold tolerance in rice was explored by gain- and loss-of-function approaches. Constructs Ubi::MYBS3 and Ubi::MYBS3 (RNAi) (RNA interference) (FIG. 1) were introduced into the 10 rice genome, and several transgenic lines were obtained. Compared to the untransformed wild type (WT) rice, the accumulation of MYBS3 mRNA was 62.5 and 19.4 times that of WT in MYBS3-overexpression [MYBS3(Ox)] lines S3(Ox)-110-1 and S3(Ox)-112-7, and 0.19 and 0.16 times 15 that of WT in MYBS3-underexpression [MYBS3(Ri)] lines S3(Ri)-42-10 and S3(Ri)-52-7. Each of these lines contained only one copy of inserted DNA.

To test the cold tolerance of transgenic rice, seedlings were shifted from 28° C. to 4° C. MYBS3(Ox) lines and WT 20 remained normal while MYBS3(Ri) lines started to show leaf rolling at 4° C. after 8 h (FIG. 3A), and both WT and MYBS3 (Ri) lines showed leaf rolling and wilting at 4° C. after 24 h in hydroponic culture (FIG. 3B) or 1 week in soil. Seedlings seemed to be more sensitive to cold in hydroponic culture, 25 probably due to weaker growth in hydroponic culture than in soil. Line S3(Ox)-110-1, which accumulated three times more MYBS3 mRNA than S3(Ox)-112-7, conferred higher cold tolerance than line S3(Ox)-112-7. Quantitative analysis also indicated that MYBS3(Ox) lines were more cold tolerant 30 than WT and MYBS3(Ri) lines, and WTs were more cold tolerant than MYBS3(Ri) lines (Table 2). These observations suggest that MYBS3 is sufficient and necessary for cold tolerance in rice, and the degree of cold tolerance correlates with the MYBS3 expression level.

TABLE 2

MYBS3(Ox) lines are more cold tolerant.						
Line	No. of plants survived*	Total no. of plants tested	Survival rate (%)			
Wild type	3	30	10.0 ± 0.0			
S3(Ox)-110-1	18	21	85.7 ± 10.5			
S3(Ox)-112-7	26	32	81.3 ± 11.1			
S3(Ri)-42-10	0	20	0.0 ± 0.0			
S3(Ri)-52-7	0	21	0.0 ± 0.0			

*Number of plants survived after exposure to 4° C. for 24 h. Experiments were repeated 4 times. Five to eight plants per line were tested in each experiment. The survival rate is a mean \pm SD.

The morphology of transgenic rice was similar to the WT, 50 except under greenhouse growth conditions, where plants of the MYBS3(Ox) lines were 20% shorter, had 30% lower tiller numbers, and headed 1 week later than the WT and MYBS3 (Ri) line. However, in field conditions, most agronomic traits and yield of MYBS3(Ox) lines were similar to those of the 55 WT (Table 3).

TABLE 3

Comparison of agronomic traits of a MYBS3(Ox) line with wild type grown in field.					
Trait	WT	S3(Ox)-110-1			
Plant height (cm)	101.6 ± 3.7*	95.5 ± 5.4			
Tiller number	12.3 ± 2.5	19.5 ± 6.8			
Panicle number/plant	12.9 ± 2.9	19.0 ± 6.1			
Panicle length (cm)	18.9 ± 1.3	18.4 ± 0.1			

Comparison of agronomic traits of a MYBS3(Ox) line with wild type grown in field Trait WT S3(Ox)-110-1 Grain number/panicle 118.0 ± 14.5 103.5 ± 8.8 Fertility (%) 95.7 ± 1.2 93.8 ± 1.7 Grain yield (g/plant) 41.7 ± 11.0 45.4 ± 15.9 1000 Grains weight (g) 26.3 ± 0.5 24.3 ± 0.4

Twenty plants each of WT and line S3(Ox)-110-1 per replicate, and with total of 3 replicates, were grown during February to July, 2008. The value is a mean±SD.

MYBS3 Regulates the Expression of Genes with Diverse

To identify downstream genes regulated by MYBS3 under cold stress, seedlings of S3(Ox)-110-1, S3(Ri)-52-7 and WT were grown at 4 and 28° C. for 24 h. Total RNAs were isolated for microarray analysis using the Affymetrix rice gene chip array containing 55,515 probe sets. Relative change was calculated by comparing the data for MYBS3(Ox) line or MYBS3(Ri) line against those for WT grown at 4° C. and 28° C., generating six comparisons. Only relative changes of 3-fold or more were taken to be significantly different. Based on a Venn diagram analysis, 89 genes were up-regulated in the MYBS3(Ox) line (compared with WT) at either 4 or 28° C., and 1466 genes were up-regulated in WT at 4° C. (compared with 28° C.). Among these genes, 17 genes were up-regulated by over-expression of MYBS3 as well as up-regulated by cold in WT (Table S1 of Su, et al, Plant Physiol. 2010; 153(1):145-58). On the other hand, 291 genes were down-regulated in the MYBS3(Ox) line (compared with WT) at either 4 or 28° C., and 871 genes were down-regulated in WT at 4° C. (com-35 pared with 28° C.). Among these genes, 53 genes were downregulated by over-expression of MYBS3 as well as downregulated by cold in WT (Table S1 of Su, et al, Plant Physiol. 2010; 153(1):145-58).

Another analysis revealed that 389 genes were up-regu-40 lated in the MYBS3(Ri) line (compared with WT) at either 4 or 28° C. Among these genes, 17 genes were up-regulated by under-expression of MYBS3 as well as up-regulated by cold in WT (Table S2 of Su, et al, Plant Physiol. 2010; 153(1):145-58). On the other hand, 124 genes were down-regulated in the MYBS3(Ri) line (compared with WT) at either 4 or 28° C. Among these genes, 37 genes were down-regulated by overexpression of MYBS3 as well as by cold in WT (Table S2 of Su, et al, Plant Physiol. 2010; 153(1):145-58).

The cold- and MYBS3-regulated genes seem to be involved in diverse functions, and many of them have also been shown to be regulated by drought and salt stresses (Tables S1 and S2 of Su, et al, Plant Physiol. 2010; 153(1): 145-58). Among the 17 genes up-regulated by over-expression of MYBS3 as well as up-regulated by cold in WT, five genes that have also been shown to be up-regulated by drought (Table Si of Su, et al, Plant Physiol. 2010; 153(1): 145-58) and cold (Jain, et al., 2007, Plant Physiol 143: 1467-1483), such as genes encoding glutamate decarboxylase, WRKY77, multidrug resistance protein 4, and trehalose-6-60 phosphate phosphatases (TPP1 and TPP2), were selected for further quantitative real-time RT-PCR analysis. The accumulation of mRNA of all five genes was significantly increased in WT and further increased in the MYBS3(Ox) line but reduced in the MYBS3(Ri) line at 4° C. (Table S3 of Su, et al, 65 Plant Physiol. 2010; 153(1):145-58), indicating that these genes are downstream of the MYBS3-mediated cold signaling pathway.

MYBS3 Suppresses the DREB1-Dependent Pathway Under Prolonged Cold Stress

We noticed that in the microarray analysis, the DREB1 family, including DREB1A, DREB1B and DREB1C, and another two DREB1-like genes (ERF#025 and ERF#104) 5 were up-regulated in WT at 4° C., but the induction was surprisingly reduced or abolished in the MYBS3(Ox) line at 4° C. To investigate how MYBS3 regulates DREB1 gene expression, the accumulation of mRNAs of three DREB1 genes was further analyzed with the quantitative real-time 10 RT-PCR analysis. Compared with WT, accumulation of MYBS3 mRNA increased significantly at 28° C. and was further induced 2-fold at 4° C. in the MYBS3(Ox) line. The accumulation of MYBS3 mRNA was reduced in the MYBS3 (Ri) line at both 4 and 28° C. In contrast, the cold-induced 15 DREB1A, DREB1B and DREB1C expression was significantly suppressed in the MYBS3(Ox) line at 4° C. Furthermore, the cold inducibility of αAmy3/RAmy3D and a cytochrome P450 gene, both members of the cold-inducible DREB1A regulon (Ito, et al., 2006, Plant Cell Physiol 47: 20 141-153), were also significantly reduced in the MYBS3(Ox) line at 4° C. The accumulation of DREB1, αAmy3/RAmy3D and cytochrome P450 mRNAs were significantly higher in the MYBS3(Ri) line than in the MYBS3(Ox) line at 4° C., although levels did not reach to that in WT at 4° C

MYBS3 has been shown to repress αAmy3 SRC through the TA box (Lu, et al., 2002, Plant Cell 14: 1963-1980). Examination of promoter regions within 1 kb upstream of the translation start codon (ATG) revealed the presence of TA box and/or its variants in DREB1 genes. To determine whether 30 MYBS3 represses DREB1 promoters, a rice embryo transient expression assay was performed. Rice embryos were cotransfected with the effector construct containing Ubi promoter fused to MYBS3 cDNA and the reporter construct containing DREB1A (1054 bp), DREB1B (747 bp) or αAmy3 SRC (105 35 bp) promoter sequence fused to luciferase cDNA (Luc). Both DREB1 promoters were significantly induced at 4° C., but only the DREB1B promoter was repressed by over-expression of MYBS3 at 4° C. The αAmy3 SRC was repressed by over-expression of MYBS3 at both 4 and 28° C., consistent 40 with the role of MYBS3 as a repressor of αAmy3 SRC (Lu, et al., 2002, Plant Cell 14: 1963-1980). These results indicate that MYBS3 could repress DREB1B promoter and αAmy3 SRC at 4° C.

Discussion

A Novel MYBS3-Mediated Cold Signaling Pathway

Both gain- and loss-of-function analyses demonstrated that the MYBS3-mediated pathway was essential for cold stress tolerance in rice. DREB1A responded early and transiently, which is consistent with previous reports in *Arabidopsis* and 50 rice (Liu, et al., 1998, Plant Cell 10: 1391-1406; Shinwari, et al., 1998, Biochem Biophys Res Commun 250: 161-170; Dubouzet, et al., 2003, Plant J 33: 751-763; and Vogel, et al., 2005, Plant J 41: 195-211), whereas MYBS3 responded relatively slowly, to cold stress in rice (FIG. 4). The DREB1-55 mediated process is most likely crucial in responding to short term cold stress (cold shock), and the MYBS3-mediated system is more important for long-term adaptation to persistent cold stress.

Transcriptome profiling analyses suggest that multiple 60 cold response pathways exist in *Arabidopsis* and rice (Fowler, et al., 2002, Plant Cell 14: 1675-1690; Vogel, et al., 2005, Plant J 41: 195-211; Cheng, et al., 2007, BMC Genomics 8: 175; and Chinnusamy, et al., 2007, Trends Plant Sci 12: 444-451). However, the MYBS3-mediated cold signaling 65 pathway has never been observed previously. MYBS3 acts as a transcriptional repressor of αAmy3 SRC in the sugar sig-

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naling pathway in rice (Lu, et al., 2002, Plant Cell 14: 1963-1980), and is constitutively localized in the nucleus in cultured rice suspension cells. These studies indicate that MYBS3 may play multiple regulatory roles in plant growth in addition to cold response in rice. Consequently, gene expression in MYBS3(Ox) or MYBS3(Ri) line altered at 28° C. may not, whereas that altered at 4° C. may, be involved in cold response.

The MYBS3-regulated genes encompass a wide range of functions. In the microarray analysis, among the 17 genes up-regulated for at least 3-fold by over-expression of MYBS3 as well as by cold in WT, several of them have previously been implicated in stress responses and/or tolerance in plants (Table S1 of Su, et al, Plant Physiol. 2010; 153(1):145-58), such as glutamate decarboxylase, which catalyzes the conversion of glutamate to γ-aminobutyrate (GABA) and is activated in response to heat in Arabidopsis roots (Bouche et al., 2004, Plant Mol Biol 55: 315-325) and to anoxia in rice roots (Aurisano, et al., 1995, Plant Cell Physiol. 36: 1525-1529); WRKY77, which activates the ABA-inducible HVA22 promoter in cereal grains (Xie, et al., 2005, Plant Physiol 137: 176-189) and several WRKYs have been shown to confer biotic and abiotic stress tolerance in *Arabidopsis* (Ross, et al., 2007, J Integr Plant Biol 49: 827-842; Lai et al., 2008, BMC Plant Biol 8: 68; and Zhou et al., 2008, Plant Biotechnol J 6: 486-503); multidrug resistance protein 4, whose expression is activated by arsenate and arsenite stresses in rice seedlings (Chakrabarty, et al., 2009, Chemosphere 74: 688-702) and its homologous genes confer salt tolerance (Lee, et al., 2004, Plant Physiol 134: 528-538) and oxidative stress tolerance against pathogens (Sun, et al., 2006, Plant Cell 18: 3686-3705) in various plant species.

TPPs are a group of genes worthwhile noting. Trehalose is a disaccharide sugar widely distributed in bacteria, fungi, plants and invertebrate animals, and is produced from glucose by trehalose-6-phosphate synthase (TPS) and TPP, and serves as sugar storage, metabolic regulator, and protectant again abiotic stresses (Strom, et al., 1993, Mol Microbiol 8: 205-210; and Elbein, et al., 2003, Glycobiology 13: 17R-27R). Trehalose has been shown to stabilize dehydrated enzymes, proteins, and lipid membranes, as well as to protect biological structures from damage during desiccation (Elbein, et al., 2003, Glycobiology 13: 17R-27R). TPP1 and TPP2 are two major TPP genes expressed in rice seedlings (Shima, et al., 2007, Febs J 274: 1192-1201). Their expression is induced by cold and other abiotic stresses (Pramanik, et al., 2005, Plant Mol Biol 58: 751-762; Shima, et al., 2007, Febs J 274: 1192-1201; Ge, et al., 2008, Planta 228: 191-201). Trehalose accumulates rapidly and transiently, which follows the transient induction of TPP activity, in rice tissues during chilling stress (Pramanik, et al., 2005, Plant Mol Biol 58: 751-762). Overexpression of TPS and TPP enhances accumulation of trehalose and tolerance to cold stress in transgenic tobacco and rice (Garg, et al., 2002, Proc Natl Acad Sci USA 99: 15898-15903; Jang, et al., 2003 Plant Physiol 131: 516-524; Ge, et al., 2008, Planta 228: 191-201; and Iordachescu, et al., 2008, J Integr Plant Biol 50: 1223-1229). However, the regulatory mechanism of TPPs by cold or other stresses is unclear.

The accumulation of these MYBS3-activated genes were significantly increased in the MYBS3(Ox) line and decreased in the MYBS3(Ri) line at 4° C. MYBS3 confers stress tolerance to transgenic rice through the activation of these genes whose products are involved either in the regulation of gene expression for cold adaptation or for protection of cells from chilling injury.

Complexity in Cold Regulation

The temporal expression patterns and magnitudes of activation of DREB1A and MYBS3 expression by cold are quite different (FIG. 2). Several factors have been found to regulate the expression of DREB1/CBF as mentioned in the introduc- 5 tion, but the detailed information about the cold signaling pathways upstream of DREB1/CBF is rather limited. Recently, a calmodulin binding transcription factor (CAMTA) was found to bind to the conserved motif 2 (CM2) present in promoters (within 200 by upstream of ATG), and function as a positive regulator, of the rapidly cold-inducible CBF2 and ZAT12 transcription factors in Arabidopsis (Doherty, et al., 2009, Plant Cell 21: 972-984). CM2 is present in one copy in the MYBS3 promoter (-117 to -112 upstream of ATG) (Table S4 of Su, et al, Plant Physiol. 2010; 15 153(1):145-58). For cold up-regulated but MYBS3 downregulated genes, CM2 is present in two copies each in DREB1B (-134 to -129 and -80 to -75) and α Amy3 (-158to -153 and -149 to -144) promoters; for cold up-regulated and MYBS3 up-regulated genes, CM2 is present in the 20 glutamate decarboxylase (-54 to -49) and WRKY77 promoters (-96 to -91) (Table S4 of Su, et al, Plant Physiol. 2010; 153(1):145-58). Some of other CMs shared by the Arabidopsis CBF2 and ZAT12 promoters (Doherty, et al., 2009, Plant Cell 21: 972-984) could also be found in DREB1B, 25 DREB1C, αAmy3 and cytochrome P450 promoters (Table S4 of Su, et al, Plant Physiol. 2010; 153(1):145-58), but the function of these cis-acting elements and the identify of their interacting transcription factors in cold signaling have not been determined (Doherty, et al., 2009, Plant Cell 21: 972-30 984).

CMs 1-7 have been found in the *Arabidopsis* CBF2 promoter (within 200 by upstream of ATG) (Doherty, et al., 2009, Plant Cell 21: 972-984), however, only CM4 is present in the 1-kb promoter region of DREB1C (the rice CBF2 homolog), suggesting that the mechanism of cold regulation on the DREB1/CBF family might have diverged throughout evolution. No CM is present in the 1-kb promoter region of DREB1A, indicating unidentified cis-acting element(s) could be responsible for cold induction of DREB1A. It appears that 40 combinations of various cis-acting elements and interacting transcription factors constitute the quantitative and temporal regulation of the DREB1- and MYBS3-dependent cold signaling cascades.

It is also noticed that the DREB1A target sequence DRE 45 (Ito, et al., 2006, Plant Cell Physiol 47: 141-153), is present in α Amy3 (-153 to -148) and cytochrome P450 (-605 to -600) promoters, and interestingly, it overlaps with the two CM2s in α Amy3 promoter. None of the 1-kb promoter regions of MYBS3-activated genes further characterized in this study 50 contains DRE.

How MYBS3 represses the expression of the DREB1 regulon is unclear. The TA box has been shown to function in both sense and antisense orientations (Lu, et al., 1998, J Biol Chem 273: 10120-10131). Promoters of the cold inducible but 55 MYBS3 repressible genes, except cytochrome P450, contain TA box or its variants (Yu, 1999, Regulation of alpha-amylase gene expression. In K Shimamoto, ed, Molecular Biology of Rice, Springer-Verlag, Tokyo, pp 161-178; and Wang, et al., 2007, Plant Mol Biol 63: 441-463) in sense or antisense 60 orientation (Table S5 of Su, et al, Plant Physiol. 2010; 153 (1):145-58), which could be the target of repression by MYBS3. However, in the transient expression assay, the 747bp DREB1B promoter and 105-bp αAmy3 SRC, but not the 1054-bp DREB1A promoter, were repressed by over-expression of MYBS3 at 4° C. One explanation is that the TA3 box (-625 to -620) in the 1054-bp DREB1A promoter did not

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function as well as the TA2 box (-85 to -80) in the 747-bp DREB1B promoter and the TA1 box (the canonical TA box) (2 copies between -116 to -105) in the 105-bp α Amy3 SRC in the rice embryo transient expression assay.

How MYBS3 activates the expression of downstream genes in the cold signaling pathway, by serving as a transcriptional activator or repressing a transcriptional repressor, is unclear. However, except TPP2, other MYBS3 up-regulated genes also contain TA box or its variants (Table S5 of Su, et al, Plant Physiol. 2010; 153(1):145-58). Both MYBS1 and MYBS3 bind specifically to the TA box, however, MYBS1 activates and MYBS3 represses aAmy3 SRC under sugar starvation (Lu, et al., 2002, Plant Cell 14: 1963-1980). MYBs with one single DNA binding domain (1R MYB) have been proposed to bind DNA as a dimer, and MYBS1 does whereas MYBS3 doesn't form a homodimer (Lu, et al., 2002, Plant Cell 14: 1963-1980). Whether MYBS3 could be converted into an activator, by interacting with other 1R MYB and forming a heterodimer or with other transcription factor(s), remains for further study.

Taken together, above studies suggest the complexity of cold regulation in plants, which involves multiple cis-acting elements and transcription factors. Additionally, the regulation of the MYBS3-dependent pathway differs from that of the DREB1- or ROS-mediated pathways in response to cold stress (Ito, et al., 2006, Plant Cell Physiol 47: 141-153; Cheng, et al., 2007, BMC Genomics 8: 175), which suggests that MYBS3 defines a new signaling pathway mediating cold adaptation in rice. It appears that distinct regulatory pathways function in fine tuning the qualitative and quantitative gene expressions for short- and long-term cold adaptation in rice (FIG. 4).

MYBS3 as a Tool for Improving Cold Stress Tolerance in Crops

Compared with microbial TPP, the rice TPP has been shown to be rather unstable, which leads to low level accumulation of trehalose in rice under normal growth conditions (Shima, et al., 2007, Febs J 274: 1192-1201). Although in WT plant, the expression of two rice TPPs is induced by cold, it peaks around 24 h and declined afterward at 4-6° C. (Pramanik, et al., 2005, Plant Mol Biol 58: 751-762; and Ge, et al., 2008, Planta 228: 191-201). The significant activation of TPP expression in the MYBS3(Ox) line may increase the accumulation of trehalose to levels high enough to confer cold tolerance in rice.

In the MYBS3(Ri) line, the expression of three DREB1 genes were 50-94% of the WT at 4° C., probably due to weaker growth and reduced cellular activities of plants under cold stress, as mentioned above that MYBS3 may play multiple regulatory roles in plant growth in addition to cold response in rice. However, it suggests that high-level DREB1 expression is insufficient to sustain cold tolerance if the level of MYBS3 expression is too low to efficiently activate the TPP-mediated cold response pathway. Consequently, the sequential expression of DREB1 and MYBS3 provides rice two complementary mechanisms for conferring cold tolerance in rice, with the DREB1-mediated process mediates the immediate cold shock response and the MYBS3-mediated system adjusts the long-term cold adaptation in rice. The antithetical regulation of aAmy3 in rice seedlings by two different pathways is physiologically meaningful: the transient activation of αAmy3 expression by DREB1 allows hydrolysis of reserved starch for providing immediate need of carbon source and energy to combat the cold shock, while the subsequent suppression of αAmy3 expression by MYBS3 allows rice to conserve carbohydrates until re-growth is allowed at elevated temperatures. It would be interesting to

test whether stacking of these two systems, by over-expression of both DREB1 and MYBS3, could further enhance the cold tolerance in rice.

Overexpression of proteins or enzymes associated with stress responses has been a common practice in improving stress tolerance of crop plants. However, constitutive overexpression of these proteins frequently leads to impaired plant growth or yield penalty. For example, though transgenic Arabidopsis and rice constitutively overexpressing CBF/DREB1 and a NAC6 transcription factor are highly tolerant to freezing, the growth rates of these transgenic plants, however, are severely retarded under normal growth conditions (Jaglo-Ottosen, et al., 1998, Science 280: 104-106; Liu, et al., 1998, Plant Cell 10: 1391-1406; Kasuga, et al., 1999, Nat Biotechnol 17: 287-291; Gilmour, et al., 2000, Plant Physiol 124: 1854-1865; Ito, et al., 2006, Plant Cell Physiol 47: 141-153; and Nakashima et al., 2007, Plant J 51: 617-630). Using stress-inducible promoters for the expression of these transcription factors minimize their negative effects on plant growth (Kasuga, et al., 1999, Nat Biotechnol 17: 287-291; 20 and Nakashima, et al., 2007, Plant J 51: 617-630). Transgenic seedlings were able to withstand 4° C. for at least 1 week after

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shifting from 28° C., which could significantly protect seedlings from chilling injury in rice fields in areas where are easily prone to transient temperature drops in early spring. Although the growth of MYBS3(Ox) lines was affected to certain extent in the greenhouse, the growth and yield of line S3(Ox)-110-1 was normal in field (Table 3). In conclusion, MYBS3 can be used for the improvement of cold tolerance in rice and possibly other crop plants.

Other Embodiments

All of the features disclosed in this specification may be combined in any combination. Each feature disclosed in this specification may be replaced by an alternative feature serving the same, equivalent, or similar purpose. Thus, unless expressly stated otherwise, each feature disclosed is only an example of a generic series of equivalent or similar features.

A number of embodiments of the invention have been described. Nevertheless, it will be understood that various modifications may be made without departing from the spirit and scope of the invention. Accordingly, other embodiments are within the scope of the following claims.

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	_

											-	con	tin	ued			
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Glu	Asp	Asn	Ile 180	Pro	Val	Glu	Thr	Glu 185	Met	Gln	Gly	Ala	Asp 190	Ser	Ile		
His	Gln	Thr 195	Leu	Ala	Pro	Ser	Ser 200	Leu	His	Ala	Pro	Ser 205	Ile	Leu	Glu		
Ile	Glu 210	Glu	Cys	Glu	Ser	Met 215	Asp	Ser	Thr	Asn	Ser 220	Thr	Thr	Gly	Glu		
Pro 225	Thr	Ala	Thr	Ala	Ala 230	Ala	Ala	Ser	Ser	Ser 235	Ser	Arg	Leu	Glu	Glu 240		
Thr	Thr	Gln	Leu	Gln 245	Ser	Gln	Leu	Gln	Pro 250	Gln	Pro	Gln	Leu	Pro 255	Gly		
Ser	Phe	Pro	Ile 260	Leu	Tyr	Pro	Thr	Tyr 265	Phe	Ser	Pro	Tyr	Tyr 270	Pro	Phe		
Pro	Phe	Pro 275	Ile	Trp	Pro	Ala	Gly 280	Tyr	Val	Pro	Glu	Pro 285	Pro	Lys	Lys		
Glu	Glu 290	Thr	His	Glu	Ile	Leu 295	Arg	Pro	Thr	Ala	Val 300	His	Ser	Lys	Ala		
Pro 305	Ile	Asn	Val	Asp	Glu 310	Leu	Leu	Gly	Met	Ser 315	Lys	Leu	Ser	Leu	Ala 320		
Glu	Ser	Asn	Lys	His 325	Gly	Glu	Ser	Asp	Gln 330	Ser	Leu	Ser	Leu	Lys 335	Leu		
Gly	Gly	Gly	Ser 340	Ser	Ser	Arg	Gln	Ser 345	Ala	Phe	His	Pro	Asn 350	Pro	Ser		
Ser	Asp	Ser 355	Ser	Asp	Ile	Lys	Ser 360	Val	Ile	His	Ala	Leu 365					
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															ggaggg	120	
															attatt	180	
	_			_		_									gcaacg	240	
aaaa	ayaç	J-9 (-yay	cacy	שש ני	aacci		9 90	, ccg	July	cugi	-990	aac ,	3263	gcaacy	240	′

20 180 caccaccgtt tatctccgtc gtctcctctg gcgacgtcaa atcttaatga ttcgccgtta teggateatg eccgatacte taatttgeat cataatgaag ggtatttate tgatgateet 360 gctcatggtt ctgggtctag tcaccgtcgt ggtgagagga agagaggtgt tccttggact 420 gaagaggaac atagactatt cttagtcggt cttcagaaac tcgggaaagg agattggcgc 480 ggtatttcga gaaactatgt aacgtcaaga actcctacac aagtggctag tcatgctcaa aagtatttta ttegacatae tagtteaage egeaggaaaa gaeggtetag eetettegae 600 atggttacag atgagatggt aaccgattca tcgccaacac aggaagagca gaccttaaac 660 ggttcctctc caagcaagga acctgaaaag aaaagctacc ttccttcact tgagctctca 720 780 ctcaataata ccacagaagc tgaagaggtc gtagccacgg cgccacgaca ggaaaaatct caagaagcta tagaaccatc aaatggtgtt tcaccaatgc tagtcccggg tggcttcttt 840

cct	cctt	gtt t	tcca	agtga	ac ti	cacac	cgatt	t tg	geted	cctg	cgt	cact	ca (eggaa	acagaa	900
cat	gcctt	taa a	acgct	gaga	ac ti	ctt	ctcaç	g caq	gcato	agg	tcct	caaa	acc a	aaaa	cctgga	960
ttt	gctaa	aag a	aacgt	gtga	aa ca	atgga	acgaç	g tto	ggtcg	ggta	tgt	ctca	gct 1	tagca	atagga	1020
atg	gcgad	caa ç	gacad	cgaaa	ac co	gaaa	ette	e eet	tccc	eege	tato	cttt	gag a	actaç	gagccc	1080
tca	aggco	cat o	cage	gttt	ca ct	cgaa	atggo	e teg	ggtta	aatg	gtg	caga	tt q	gagta	aaaggc	1140
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Gly	Gly	Gly 35	Gly	Gly	Gly	Gly	Gly 40	Ser	Gly	Ser	Ser	Ser 45	Ala	Val	Lys	
Leu	Phe 50	Gly	Val	Arg	Leu	Thr 55	Asp	Gly	Ser	Ile	Ile 60	Lys	Lys	Ser	Ala	
Ser 65	Met	Gly	Asn	Leu	Ser 70	Ala	Leu	Ala	Val	Ala 75	Ala	Ala	Ala	Ala	Thr 80	
His	His	Arg	Leu	Ser 85	Pro	Ser	Ser	Pro	Leu 90	Ala	Thr	Ser	Asn	Leu 95	Asn	
Asp	Ser	Pro	Leu 100	Ser	Asp	His	Ala	Arg 105	Tyr	Ser	Asn	Leu	His 110	His	Asn	
Glu	Gly	Tyr 115	Leu	Ser	Asp	Asp	Pro 120	Ala	His	Gly	Ser	Gly 125	Ser	Ser	His	
Arg	Arg 130	Gly	Glu	Arg	Lys	Arg 135	Gly	Val	Pro	Trp	Thr 140	Glu	Glu	Glu	His	
Arg 145	Leu	Phe	Leu	Val	Gly 150	Leu	Gln	Lys	Leu	Gly 155	rys	Gly	Asp	Trp	Arg 160	
Gly	Ile	Ser	Arg	Asn 165	Tyr	Val	Thr	Ser	Arg 170	Thr	Pro	Thr	Gln	Val 175	Ala	
Ser	His	Ala	Gln 180	Lys	Tyr	Phe	Ile	Arg 185	His	Thr	Ser	Ser	Ser 190	Arg	Arg	
Lys	Arg	Arg 195	Ser	Ser	Leu	Phe	Asp 200	Met	Val	Thr	Asp	Glu 205	Met	Val	Thr	
Asp	Ser 210	Ser	Pro	Thr	Gln	Glu 215	Glu	Gln	Thr	Leu	Asn 220	Gly	Ser	Ser	Pro	
Ser 225	Lys	Glu	Pro	Glu	Lys 230	Lys	Ser	Tyr	Leu	Pro 235	Ser	Leu	Glu	Leu	Ser 240	
Leu	Asn	Asn	Thr	Thr 245	Glu	Ala	Glu	Glu	Val 250	Val	Ala	Thr	Ala	Pro 255	Arg	
Gln	Glu	Lys	Ser 260	Gln	Glu	Ala	Ile	Glu 265	Pro	Ser	Asn	Gly	Val 270	Ser	Pro	
Met	Leu	Val 275	Pro	Gly	Gly	Phe	Phe 280	Pro	Pro	Сув	Phe	Pro 285	Val	Thr	Tyr	
Thr	Ile 290	Trp	Leu	Pro	Ala	Ser 295	Leu	His	Gly	Thr	Glu 300	His	Ala	Leu	Asn	

Ala Glu Thr Ser Ser Gln Gln His Gln Val Leu Lys Pro Lys Pro Gly

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305 310 315 320	
Phe Ala Lys Glu Arg Val Asn Met Asp Glu Leu Val Gly Met Ser Gln 325 330 335	
Leu Ser Ile Gly Met Ala Thr Arg His Glu Thr Glu Thr Ser Pro Ser 340 345 350	
Pro Leu Ser Leu Arg Leu Glu Pro Ser Arg Pro Ser Ala Phe His Ser 355 360 365	
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agcatgggga acctetecet ecteteegeg ggateeacea geggeggege gteeceegee	180
gacgggcccg acctcgccga cggcggcggg ggctacgcct ccgacgactt cgtccagggg	240
tegteeteeg eeageegega tegaaagaaa ggtgtteett ggaetgaaga agaacaeegg	300
aggtttttgc tgggattaca aaagctcggg aaaggggatt ggcgaggaat ttctcgtaat	360
tttgtggtct caagaacacc tactcaagta gcaagtcatg ctcaaaagta ttttatacgc	420
caatcaaata tgagcagaag gaagagaagg tctagccttt tcgacatggt tcctgatgag	480
tccatggacc ttccgcccct tcctggaagt caagaaccag agacctcaat gttaaatcaa	540
ccgccactgc ctcctgctgt ggaggaggag gtggaatcga tggagtcaga tacttctgct	600
gtcgcagaga gttctggagc ttctgctctc atgcccgaga gtttacagcc tacctatccg	660
atgattgttc cagettattt etegeegtte ttgcaattet cagtteettt etggeeaaat	720
caggaagatg gaggcgatct tccccaagaa acacacgaga ttgtcaagcc tgttgcagtt	780
catteecaga atecaattaa tgttgatgaa etegtgggea tgteaaaget aageatatgg	840
gagcatggtc aggagacagt gtctacttct ctgtcgctaa atctgctagg gggtcaaaat	900
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Thr Cys Pro Asn Arg Gly Val Lys Ile Phe Gly Val His Leu Thr Asp 20 25 30	
Gly Ser Ala Ile Arg Lys Ser Ala Ser Met Gly Asn Leu Ser Leu Leu 35 40 45	
Ser Ala Gly Ser Thr Ser Gly Gly Ala Ser Pro Ala Asp Gly Pro Asp 50 55 60	

Leu Ala Asp Gly Gly Gly Tyr Ala Ser Asp Asp Phe Val Gln Gly 65 70 75 80

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Ser	Ser	Ser	Ala	Ser 85	Arg	Asp	Arg	Lys	Lys	Gly	Val	Pro	Trp	Thr 95	Glu	
Glu	Glu	His	Arg 100	Arg	Phe	Leu	Leu	Gly 105	Leu	Gln	Lys	Leu	Gly 110	Lys	Gly	
Asp	Trp	Arg 115	Gly	Ile	Ser		Asn 120	Phe	Val	Val	Ser	Arg 125	Thr	Pro	Thr	
Gln	Val 130	Ala	Ser	His	Ala	Gln 135	Lys	Tyr	Phe	Ile	Arg 140	Gln	Ser	Asn	Met	
Ser 145	Arg	Arg	Lys	Arg	Arg 150	Ser	Ser	Leu	Phe	Asp 155	Met	Val	Pro	Asp	Glu 160	
Ser	Met	Asp	Leu	Pro 165	Pro	Leu	Pro	Gly	Ser 170	Gln	Glu	Pro	Glu	Thr 175	Ser	
Met	Leu	Asn	Gln 180	Pro	Pro	Leu	Pro	Pro 185	Ala	Val	Glu	Glu	Glu 190	Val	Glu	
Ser	Met	Glu 195	Ser	Asp	Thr	Ser	Ala 200	Val	Ala	Glu	Ser	Ser 205	Gly	Ala	Ser	
Ala	Leu 210	Met	Pro	Glu	Ser	Leu 215	Gln	Pro	Thr	Tyr	Pro 220	Met	Ile	Val	Pro	
Ala 225	Tyr	Phe	Ser	Pro	Phe 230	Leu	Gln	Phe	Ser	Val 235	Pro	Phe	Trp	Pro	Asn 240	
Gln	Glu	Asp	Gly	Gly 245	Asp	Leu	Pro	Gln	Glu 250	Thr	His	Glu	Ile	Val 255	Lys	
Pro	Val	Ala	Val 260	His	Ser	Gln	Asn	Pro 265	Ile	Asn	Val	Asp	Glu 270	Leu	Val	
Gly	Met	Ser 275	ГÀз	Leu	Ser	Ile	Trp 280	Glu	His	Gly	Gln	Glu 285	Thr	Val	Ser	
Thr	Ser 290	Leu	Ser	Leu	Asn	Leu 295	Leu	Gly	Gly	Gln	Asn 300	Arg	Gln	Ser	Ala	
Phe 305	His	Ala	Asn	Pro	Gln 310	Thr	Arg	Ala	Gln	Ala 315						
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															cagaag	360
															caccc	420
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876

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agcatgggga	acctctccct	cctctccgcg	ggatccacca	gcggcggcgc	gtcccccgcc	180
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Thr Cys Pro Asn Arg Gly Val Lys Ile Phe Gly Val Arg Leu Thr Asp 25

Gly Ser Ala Ile Arg Lys Ser Ala Ser Met Gly Asn Leu Ser Leu Leu

Ser Ala Gly Ser Thr Ser Gly Gly Ala Ser Pro Ala Asp Gly Pro Asp

Leu Ala Asp Gly Gly Ala Gly Gly Tyr Ala Ser Asp Asp Phe Val Gln 65 $$ 70 $$ 75 $$ 80 70

Gly Ser Ser Ser Ala Ser Arg Glu Arg Lys Lys Gly Val Pro Trp Thr $85 \hspace{0.5cm} 90 \hspace{0.5cm} 95$

Glu Glu Glu His Arg Arg Phe Leu Leu Gly Leu Gln Lys Leu Gly Lys $100 \hspace{1.5cm} 100 \hspace{1.5cm} 105 \hspace{1.5cm} 110 \hspace{1.5cm}$

Gly Asp Trp Arg Gly Ile Ser Arg Asn Phe Val Val Ser Arg Thr Pro

Thr Gln Val Ala Ser His Ala Gln Lys Tyr Phe Ile Arg Gln Ser Asn

Met Ser Arg Arg Lys Arg Arg Ser Ser Leu Phe Asp Met Val Pro Asp 150 155

Glu Ser Met Asp Leu Pro Pro Leu Pro Gly Ser Gln Glu Pro Glu Thr 170

Ser Val Leu Asn Gln Ala Pro Leu Pro Pro Pro Val Glu Glu Val 185

Glu Ser Met Glu Ser Asp Thr Ser Ala Val Ala Glu Ser Ser Thr Ala 200

Ser Ala Leu Met Pro Glu Ser Leu Gln Pro Asn Tyr Pro Met Ile Val

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gatgatgttg ttggtatgtc taagctcagc attggtgagg ccagctctgg ctccatggaa cccacagctc tttcccttca gcttattgga tcgacagata caaggcagtc agcttttcat

gtgagtccac caatgaatag acctgaacta agcaagagaa acagcagtcc aattcatgcc

ggcacaaccc atgaaatcct aaaaccaact cctttgaatg gtaaggaggt gattaaagca

gtttga 1086

1080

<210> SEQ ID NO 54

<211> LENGTH: 361

<212> TYPE: PRT

<213 > ORGANISM: Sorghum bicolor

<400> SEQUENCE: 54

Met Thr Arg Arg Cys Ser His Cys Ser Asn Asn Gly His Asn Ser Arg 10

Thr Cys Pro Ala Arg Ser Gly Gly Gly Val Arg Leu Phe Gly Val Arg 25

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Leu Thr Th		Pro	Ala	Pro	Ala 40	Ala	Met	Lys	Lys	Ser 45	Ala	Ser	Met	
Ser Cys Il	e Ala	Ser	Ser	Leu 55	Gly	Gly	Gly	Ser	Gly 60	Gly	Ser	Ser	Pro	
Pro Ala Gl 65	y Gly	Val	Gly 70	Gly	Gly	Arg	Gly	Gly 75	Gly	Asp	Gly	Gly	Ala 80	
Gly Tyr Va	ıl Ser	Asp 85	Asp	Pro	Gly	His	Ala 90	Ser	Сув	Ser	Thr	Asn 95	Gly	
Arg Val G	u Arg 100	Lys	ГÀа	Gly	Thr	Pro 105	Trp	Thr	Glu	Glu	Glu 110	His	Arg	
Met Phe Le		Gly	Leu	Gln	Lys 120	Leu	Gly	Lys	Gly	Asp 125	Trp	Arg	Gly	
Ile Ser An 130	g Asn	Phe	Val	Val 135	Ser	Arg	Thr	Pro	Thr 140	Gln	Val	Ala	Ser	
His Ala Gl 145	n Lys	Tyr	Phe 150	Ile	Arg	Gln	Thr	Asn 155	Ser	Ser	Arg	Arg	160 Lys	
Arg Arg Se	r Ser	Leu 165	Phe	Asp	Met	Val	Ala 170	Glu	Met	Pro	Val	Asp 175	Glu	
Ser Leu Al	a Ala 180	Ala	Glu	Gln	Ile	Thr 185	Ile	Gln	Asn	Thr	Gln 190	Asp	Glu	
Ala Ala Se		Asn	Gln	Leu	Pro 200	Thr	Leu	His	Leu	Gly 205	His	Gln	ГÀа	
Glu Ala Gl 210	u Phe	Ala	Lys	Gln 215	Met	Pro	Thr	Phe	Gln 220	Leu	Arg	Gln	His	
Glu Glu Se 225	er Glu	Tyr	Ala 230	Glu	Pro	Ser	Leu	Thr 235	Leu	Pro	Asp	Leu	Glu 240	
Met Asn Se	r Ser	Val 245	Pro	Phe	Asn	Thr	Ile 250	Ala	Val	Pro	Thr	Met 255	Pro	
Ala Phe Ty	r Pro 260	Ala	Leu	Val	Pro	Val 265	Pro	Leu	Thr	Leu	Trp 270	Pro	Pro	
Ser Val Al 27		Val	Glu	Asp	Ala 280	Gly	Thr	Thr	His	Glu 285	Ile	Leu	Lys	
Pro Thr Pr 290	o Leu	Asn	Gly	Lys 295	Glu	Val	Ile	ГÀа	Ala 300	Asp	Asp	Val	Val	
Gly Met Se 305	er Lys	Leu	Ser 310	Ile	Gly	Glu	Ala	Ser 315	Ser	Gly	Ser	Met	Glu 320	
Pro Thr Al	a Leu	Ser 325	Leu	Gln	Leu	Ile	Gly 330	Ser	Thr	Asp	Thr	Arg 335	Gln	
Ser Ala Ph	e His 340	Val	Ser	Pro	Pro	Met 345	Asn	Arg	Pro	Glu	Leu 350	Ser	ГÀа	
Arg Asn Se		Pro	Ile	His	Ala 360	Val								
<210> SEQ ID NO 55 <211> LENGTH: 1002 <212> TYPE: DNA <213> ORGANISM: Glycine max														
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atgggcaato														180
aaccccggtt	egce	cggg	ga aa	accc	ccgat	c cat	gaaq	gccg	cagt	cgc	cga (eggti	acttg	240

tcc	gagga	act t	cgtt	caaq	gg gt	tctt	ettet	ago	ctcc	gtg	aaaq	gaaaq	gaa q	gggt	gttcca	300
tgga	actga	agg a	aggaa	acata	ag aa	atgtt	ttta	a cto	cggat	ttgc	agaa	agct	ggg (caaa	ggtgat	360
tgg	gtgg	gaa t	tgca	aagga	ac ct	tatgt	tata	a tca	aagga	acac	cta	ctcaa	agt 9	ggcta	agccat	420
gcto	cagaa	aat a	attto	catca	ag go	cagaç	gcaat	gt	gtcca	aggc	gga	aaaga	acg (gtcca	agcttg	480
tttç	gatat	tg t	tgca	agat	ga aç	gcago	ctgad	c act	tgcaa	atgg	taca	agcaa	aga (cttct	tgtct	540
gcta	aatca	agt t	cacco	cact	ga aa	acaga	aaggo	c aat	taaco	ccct	tgc	cagct	tcc 1	teet	ccctc	600
gaco	gaaga	igt 9	gegaa	atcca	at g	gatto	ccaca	a aad	ctcaa	aatg	atg	gaga	gaa 1	tgcc	ccatca	660
aago	ccaga	aaa a	acaca	acagt	c at	tctta	atcca	a ato	gttat	tatc	ctg	catat	tta 1	ttct	ccggtg	720
ttco	ecgtt	tc o	etete	gecet	a ti	tggt	cagga	a tao	cagto	ccag	agt	ccaco	caa 🤉	gaag	gaggag	780
acad	catga	aag t	tacto	gaago	cc aa	actgo	cagtt	cat	ttcta	aaaa	gcc	ctato	caa 1	tgtt	gatgaa	840
ctg	gttgg	gca t	ttca	aaaat	t ga	agttt	aggg	g gaq	gteta	attg	gtga	actct	tgg 1	tacat	cctct	900
ctgt	ctc	gaa a	aactt	tatco	ga aq	gaag	gacco	c tct	tagad	cagt	cago	ctttt	tca 1	tgcaa	acaccg	960
acat	gtgg	gca (gttca	aaat	gg ca	agtgo	ccato	c cat	tgcag	gttt	aa					1002
<211 <212 <213	0 > SI L > LI 2 > T\ 3 > OF	ENGTI (PE : RGAN	H: 33 PRT ISM:	33 Gly	cine	max										
Met	Thr	Arg	Arg	Cys	Ser	His	Cys	Ser	His	Asn	Gly	His	Asn	Ser	Arg	
1				5					10					15		
Thr	Cys	Pro	Asn 20	Arg	Gly	Val	Lys	Leu 25	Phe	Gly	Val	Arg	Leu 30	Thr	Asp	
Gly	Ser	Ile 35	Arg	Lys	Ser	Ala	Ser 40	Met	Gly	Asn	Leu	Thr 45	His	Tyr	Ala	
Gly	Ser 50	Gly	Ser	Gly	Pro	Leu 55	His	Thr	Gly	Leu	Asn 60	Asn	Pro	Gly	Ser	
Pro 65	Gly	Glu	Thr	Pro	Asp 70	His	Ala	Ala	Ala	Val 75	Ala	Asp	Gly	Tyr	Leu 80	
Ser	Glu	Asp	Phe	Val 85	Pro	Gly	Ser	Ser	Ser 90	Ser	Ser	Arg	Glu	Arg 95	Lys	
Lys	Gly	Val	Pro	Trp	Thr	Glu	Glu	Glu 105	His	Arg	Met	Phe	Leu 110	Leu	Gly	
Leu	Gln	Lys 115	Leu	Gly	Lys	Gly	Asp 120	Trp	Arg	Gly	Ile	Ala 125	Arg	Thr	Tyr	
Val	Ile 130	Ser	Arg	Thr	Pro	Thr 135	Gln	Val	Ala	Ser	His	Ala	Gln	Lys	Tyr	
Phe		Arg	Gln	Ser	Asn 150		Ser	Arg	Arg	Lys 155		Arg	Ser	Ser	Leu 160	
	Asp	Ile	Val			Glu	Ala	Ala	_		Ala	Met	Val	Gln		
				165					170					175		
Asp	Phe	Leu	Ser 180	Ala	Asn	Gln	Leu	Pro 185	Thr	Glu	Thr	Glu	Gly 190	Asn	Asn	
Pro	Leu	Pro 195	Ala	Pro	Pro	Pro	Leu 200	Asp	Glu	Glu	CAa	Glu 205	Ser	Met	Asp	
Ser	Thr 210	Asn	Ser	Asn	Asp	Gly 215	Glu	Pro	Ala	Pro	Ser 220	Lys	Pro	Glu	Asn	

Thr Gln Ser Ser Tyr Pro Met Leu Tyr Pro Ala Tyr Tyr Ser Pro Val 225 230 240

79 80

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Phe Pro Phe Pro Leu Pro Tyr Trp Ser Gly Tyr Ser Pro Glu Ser Thr

Lys Lys Glu Glu Thr His Glu Val Leu Lys Pro Thr Ala Val His Ser 265

Lys Ser Pro Ile Asn Val Asp Glu Leu Val Gly Ile Ser Lys Leu Ser

Leu Gly Glu Ser Ile Gly Asp Ser Gly Pro Ser Ser Leu Ser Arg Lys 295

Leu Ile Glu Glu Gly Pro Ser Arg Gln Ser Ala Phe His Ala Thr Pro

Thr Cys Gly Ser Ser Asn Gly Ser Ala Ile His Ala Val

<210> SEQ ID NO 57

<211> LENGTH: 1002

<212> TYPE: DNA

<213> ORGANISM: Glycine max

<400> SEQUENCE: 57

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<210> SEQ ID NO 58

<211> LENGTH: 333

<212> TYPE: PRT

<213 > ORGANISM: Glycine max

<400> SEQUENCE: 58

Met Thr Arg Arg Cys Ser His Cys Ser His Asn Gly His Asn Ser Arg 1.0

Thr Cys Pro Asn Arg Gly Val Lys Leu Phe Gly Val Arg Leu Thr Asp

Gly Ser Ile Arg Lys Ser Ala Ser Met Gly Asn Leu Thr His Tyr Ala 35 40

Gly Ser Gly Ser Ala Pro Leu His Val Gly Leu Asn Asn Pro Gly Ser

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50 55 60	
Pro Gly Glu Thr Pro Asp His Ala Ala Ala Ala Ala Asp Gly Tyr Ala 65 70 75 80	
Ser Glu Asp Phe Val Pro Gly Ser Ser Ser Ser Ser Arg Glu Arg Lys 85 90 95	
Lys Gly Val Pro Trp Thr Glu Glu Glu His Arg Met Phe Leu Leu Gly 100 105 110	
Leu Gln Lys Leu Gly Lys Gly Asp Trp Arg Gly Ile Ala Arg Asn Tyr 115 120 125	
Val Ile Ser Arg Thr Pro Thr Gln Val Ala Ser His Ala Gln Lys Tyr 130 135 140	
Phe Ile Arg Gln Ser Asn Val Ser Arg Arg Lys Arg Arg Ser Ser Leu 145 150 155 160	
Phe Asp Ile Val Ala Asp Glu Ala Ala Asp Thr Ala Met Val Gln Gln 165 170 175	
Asp Phe Leu Ser Ala Asn Glu Leu Pro Thr Glu Thr Glu Gly Asn Asn 180 185 190	
Pro Leu Pro Ala Pro Pro Pro Leu Asp Glu Glu Cys Glu Ser Met Asp 195 200 205	
Ser Thr Asn Ser Asn Asp Gly Glu Pro Ala Pro Ser Lys Pro Glu Asn 210 215 220	
Thr His Pro Ser Tyr Pro Met Leu Tyr Pro Ala Tyr Tyr Ser Pro Val 225 230 235 240	
Phe Pro Phe Pro Leu Pro Tyr Trp Ser Gly Tyr Ser Pro Glu Pro Thr 245 250 255	
Lys Lys Glu Glu Thr His Glu Val Leu Lys Pro Thr Ala Val His Ser 260 265 270	
Lys Ser Pro Ile Asn Val Asp Glu Leu Val Gly Ile Ser Lys Leu Ser 275 280 285	
Leu Gly Glu Ser Ile Gly Asp Ser Gly Pro Ser Thr Leu Ser Arg Lys 290 295 300	
Leu Ile Glu Glu Gly Pro Ser Arg Gln Ser Ala Phe His Ala Thr Pro 305 310 315 320	
Thr Cys Gly Asp Met Asn Gly Ser Ala Ile His Ala Val 325 330	
<210> SEQ ID NO 59 <211> LENGTH: 1008 <212> TYPE: DNA <213> ORGANISM: Vitis vinifera <400> SEQUENCE: 59	
atgactogoo gotgotogoa ttgoagtoac aacgggoaca attocaggao atgococaac	60
cgcggggtca agatettcgg ggttcgattg actgatgggt tgatccgtaa gagtgctagt	
atgggcaatc tcagccacta cgccgggtcg acctctggtc atcatcagaa cggcgtttcc	180
ggtaacaatt cggtctctcc cggagagact ccagagcacg gcgccgcggc cgatggatac	240
gcctccgagg gtttcgttcc cggttcatca tccagccggg agcgcaagaa aggcactcca	300
tggactgaag aggaacacag aatgtttcta cttggactgc agaagcttgg aaaaggggat	360
tggcgtggaa tttcacgtaa ttatgttata tcaaggacac ctactcaagt cgccagccat	420
gctcagaaat atttcatcag gcaaactaat gtgtctagga gaaaaagacg gtccagcttg	480

tttgatattg tagctgatga atctgtcgac actccaatgg tatcacggga tttcttctcc

accaaccett egeaagetga aacactaage aataacceat tgeetgttee teeggetetg	600
gatgaagaat gtgaatcaat ggattctacc aactcgaatg atggagaacc gcccattcca	660
aageeggatg gettacaagg etgteeecca gtaatatate etaettattt eteaecatte	720
ttcccatttt cttttccatt ctggccggga aacagttcag agccaactaa aatggagact	780
catgaggtgc ttaagccaac agctgtacat tctaagagtc caatcaatgt tgatgagctg	840
gttggcatgt caaaactgag tttaggagaa tccatcggtc atgctggtcc ctcctctcc	900
acactgaaac tgcttgaagg gtcaagcagg caatctgctt tccatgctaa tccagcctct	960
ggcagttcaa gcatgaactc gagcggcagt ccaatccatg cagtttga	1008
<210> SEQ ID NO 60 <211> LENGTH: 335 <212> TYPE: PRT <213> ORGANISM: Vitis vinifera	
<400> SEQUENCE: 60	
Met Thr Arg Arg Cys Ser His Cys Ser His Asn Gly His Asn Ser Arg 1 10 15	
Thr Cys Pro Asn Arg Gly Val Lys Ile Phe Gly Val Arg Leu Thr Asp 20 25 30	
Gly Leu Ile Arg Lys Ser Ala Ser Met Gly Asn Leu Ser His Tyr Ala 35 40 45	
Gly Ser Thr Ser Gly His His Gln Asn Gly Val Ser Gly Asn Asn Ser 50 55 60	
Val Ser Pro Gly Glu Thr Pro Glu His Gly Ala Ala Ala Asp Gly Tyr 65 70 75 80	
Ala Ser Glu Gly Phe Val Pro Gly Ser Ser Ser Ser Arg Glu Arg Lys 85 90 95	
Lys Gly Thr Pro Trp Thr Glu Glu Glu His Arg Met Phe Leu Leu Gly 100 105 110	
Leu Gln Lys Leu Gly Lys Gly Asp Trp Arg Gly Ile Ser Arg Asn Tyr 115 120 125	
Val Ile Ser Arg Thr Pro Thr Gln Val Ala Ser His Ala Gln Lys Tyr 130 135 140	
Phe Ile Arg Gln Thr Asn Val Ser Arg Arg Lys Arg Arg Ser Ser Leu 145 150 155 160	
Phe Asp Ile Val Ala Asp Glu Ser Val Asp Thr Pro Met Val Ser Arg 165 170 175	
Asp Phe Phe Ser Thr Asn Pro Ser Gln Ala Glu Thr Leu Ser Asn Asn 180 185 190	
Pro Leu Pro Val Pro Pro Ala Leu Asp Glu Glu Cys Glu Ser Met Asp 195 200 205	
Ser Thr Asn Ser Asn Asp Gly Glu Pro Pro Ile Pro Lys Pro Asp Gly 210 215 220	
Leu Gln Gly Cys Pro Pro Val Ile Tyr Pro Thr Tyr Phe Ser Pro Phe 225 230 235 240	
Phe Pro Phe Ser Phe Pro Phe Trp Pro Gly Asn Ser Ser Glu Pro Thr	
Lys Met Glu Thr His Glu Val Leu Lys Pro Thr Ala Val His Ser Lys 260 265 270	
Ser Pro Ile Asn Val Asp Glu Leu Val Gly Met Ser Lys Leu Ser Leu 275 280 285	
	

Gly Glu Ser Ile Gly His Ala Gly Pro Ser Ser Leu Thr Leu Lys Leu

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290	295 30	00
Leu Glu Gly Ser 305	Ser Arg Gln Ser Ala Phe His Al	la Asn Pro Ala Ser 320
Gly Ser Ser Ser	Met Asn Ser Ser Gly Ser Pro II 325 330	le His Ala Val 335
<210> SEQ ID NO <211> LENGTH: 1 <212> TYPE: DNA <213> ORGANISM:	.038	
<400> SEQUENCE:	61	
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cgcggggtca agat	cttcgg ggttcgattg actgatgggt tg	gateegtaa gagtgetagt 120
atgggcaatc tcag	ccacta cgccgggtcg acctctggtc at	ccatcagaa cggcgtttcc 180
ggtaacaatt cggt	ctetee eggagagaet eeagageaeg ge	egeegegge egatggatae 240
gcctccgagg gttt	egttee eggtteatea teeageeggg ag	gegeaagaa aggeaeteea 300
tggactgaag agga	acacag aatgtttcta cttggactgc ag	gaagettgg aaaaggggat 360
tggcgtggaa tttc	acgtaa ttatgttata tcaaggacac ct	tactcaagt cgccagccat 420
gctcagaaat attt	catcag gcaaaccaat gtgtctagga ga	aaaaagacg gtccagcttg 480
tttgatattg tagc	tgatga atctgttgac actccaatgg ta	atcacggga tttcttctcc 540
accaaccett egca	lagetga aacaetaage aataaceeat te	geetgttee teeggetetg 600
gatgaagaat gtga	atcaat ggattctacc aactcgaatg at	eggagaacc acccattcca 660
aagccggatg gctt	acaagg ctgtccccca gtaatatatc ct	eacttattt ctcgccattc 720
ttcccatttt cttt	tccatt ctggccggga aacagttcag ag	gccaactaa aatggagact 780
catgaggtgc ttaa	gccaac agctgtacat tctaagagtc ca	aatcaatgt tgatgagctg 840
gttggcatgt caaa	actgag tttaggagaa tccatcggtc at	egetggtee etectetete 900
acactgaaac tgct	tgaagg gtcaagcagg caatctgctt to	ccatgctaa tccagcctct 960
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ctggtatgga gatt	gtag	1038
<210> SEQ ID NO <211> LENGTH: 3 <212> TYPE: PRT <213> ORGANISM:	45	
<400> SEQUENCE:	62	
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Thr Cys Pro Asn 20	a Arg Gly Val Lys Ile Phe Gly Va 25	al Arg Leu Thr Asp 30
Gly Leu Ile Arg 35	Lys Ser Ala Ser Met Gly Asn Le 40	eu Ser His Tyr Ala 45
Gly Ser Thr Ser	Gly His His Gln Asn Gly Val Se	
Val Ser Pro Gly	Glu Thr Pro Glu His Gly Ala Al	la Ala Asp Gly Tyr 80
Ala Ser Glu Gly	Phe Val Pro Gly Ser Ser Ser Se 85 90	er Arg Glu Arg Lys 95

Lys Gly Thr Pro Trp Thr Glu Glu Glu His Arg Met Phe Leu Leu Gly

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100 105 110	
Leu Gln Lys Leu Gly Lys Gly Asp Trp Arg Gly Ile Ser Arg Asn Ty 115 120 125	r
Val Ile Ser Arg Thr Pro Thr Gln Val Ala Ser His Ala Gln Lys Ty 130 135 140	r
Phe Ile Arg Gln Thr Asn Val Ser Arg Arg Lys Arg Arg Ser Ser Le 145 150 155 16	
Phe Asp Ile Val Ala Asp Glu Ser Val Asp Thr Pro Met Val Ser Ar- 165 170 175	g
Asp Phe Phe Ser Thr Asn Pro Ser Gln Ala Glu Thr Leu Ser Asn As 180 185 190	n
Pro Leu Pro Val Pro Pro Ala Leu Asp Glu Glu Cys Glu Ser Met As 195 200 205	р
Ser Thr Asn Ser Asn Asp Gly Glu Pro Pro Ile Pro Lys Pro Asp Gl 210 215 220	У
Leu Gln Gly Cys Pro Pro Val Ile Tyr Pro Thr Tyr Phe Ser Pro Ph 225 230 235 235 24	
Phe Pro Phe Ser Phe Pro Phe Trp Pro Gly Asn Ser Ser Glu Pro Th 245 250 255	.r
Lys Met Glu Thr His Glu Val Leu Lys Pro Thr Ala Val His Ser Ly 260 265 270	's
Ser Pro Ile Asn Val Asp Glu Leu Val Gly Met Ser Lys Leu Ser Le 275 280 285	u
Gly Glu Ser Ile Gly His Ala Gly Pro Ser Ser Leu Thr Leu Lys Le 290 295 300	u
Leu Glu Gly Ser Ser Arg Gln Ser Ala Phe His Ala Asn Pro Ala Se 305 310 315 32	
Gly Ser Ser Ser Met Asn Ser Ser Gly Ser Pro Ile His Ala Pro Ass 325 330 335	n
Gly Lys Ile Leu Leu Val Trp Arg Leu 340 345	
<210> SEQ ID NO 63 <211> LENGTH: 1011 <212> TYPE: DNA <213> ORGANISM: Lotus japonicus <400> SEQUENCE: 63	
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atgggtaate teaccacta cactggetee gggtetggae etettettgg tgggtee	
aaccetgatt eteeeggtga aacceetgat eaeggegeeg etgetgaegg ttaegee	
gaggattttg ttcctggctc ttcttctagc tcccgtgaaa gaaaaaaaggg cactcca	
actgaggagg aacacagaat gtttttactt ggattgcaga aactgggcaa aggtgat	
cgtggaattg caaggaacta tgttatttca aggacaccta ctcaagtggc cagtcat	
cagaaatatt tcatcaggca aagcaatgtg tctaggagaa agagacggtc cagcttg	
gatattgttg cagatgatgc gtccgacact ccaatggtag agcaagactt cttgtca	gct 540
aatcagctac agactgaaac agaaggcaat aaccetttge etgeteetee teecatt	gat 600

gaagagtgtg aatccatgga ttccacaaac tcaatagatg gagactctgc cctgttaaag

cccgacactc caataccgcc aacctacccg gtgttatatc ctgcatatta tcctccattc

660

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taccegtate etetge	cctta ti	tggtctg	ga ta	cagt	cctg	caga	agcc	ccc a	aaaga	aaagag	780
gagacacatg aagtg	gtgaa go	ccaactg	g gt	gctti	tcca	aaaq	gccca	aat o	caato	gtggat	840
gaacttgtcg gcatg	tcaaa a	ctgagtti	g gg	agact	tcca	ttg	gtgad	ctc 1	ggc	ccctcc	900
tctctgtctc gaaaa	ctcgt c	gaagaag	ga cc	ttcc	agac	aat	cagct	ttt 1	cato	gctact	960
ccagcatgtg gcagt	tcaaa ta	ataaatg	gc ag	tgtca	atac	atg	cagt	tta a	a		1011
<pre><210> SEQ ID NO 6 <211> LENGTH: 336 <212> TYPE: PRT <213> ORGANISM: 1</pre>	б	aponicus	3								
<400> SEQUENCE:	64										
Met Thr Arg Arg (Cys Ser 5	His Cys	s Ser	His 10	Gly	Gly	His	Asn	Ala 15	Arg	
Thr Cys Pro Asn 2	Arg Gly	Val Ly:	Leu 25	Phe	Gly	Val	Arg	Leu 30	Thr	Asp	
Gly Ser Ile Arg I	Lys Ser	Ala Sei	. Met	Gly	Asn	Leu	Thr 45	His	Tyr	Thr	
Gly Ser Gly Ser (Gly Pro		ı Gly	Gly	Ser	Asn 60		Pro	Asp	Ser	
Pro Gly Glu Thr 1	Pro Asp 70		a Ala	Ala	Ala 75		Gly	Tyr	Ala	Ser 80	
Glu Asp Phe Val 1	Pro Gly	Ser Sei	s Ser			Arg	Glu	Arg	_		
Gly Thr Pro Trp	85 Thr Glu	Glu Glu		90 Arg	Met	Phe	Leu		95 Gly	Leu	
100			105					110			
Gln Lys Leu Gly 1 115	Lya Gly	Asp Trp	_	Gly	Ile	Ala	Arg 125	Asn	Tyr	Val	
Ile Ser Arg Thr I 130	Pro Thr	Gln Val	l Ala	Ser	His	Ala 140	Gln	Lys	Tyr	Phe	
Ile Arg Gln Ser A	Asn Val 150	Ser Ar	g Arg	Lys	Arg 155	Arg	Ser	Ser	Leu	Phe 160	
Asp Ile Val Ala	Asp Asp 165	Ala Sei	Asp	Thr 170	Pro	Met	Val	Glu	Gln 175	Asp	
Phe Leu Ser Ala A	Asn Gln	Leu Gli	n Thr 185	Glu	Thr	Glu	Gly	Asn 190	Asn	Pro	
Leu Pro Ala Pro 1	Pro Pro	Ile Asp		Glu	Cys	Glu	Ser 205	Met	Asp	Ser	
Thr Asn Ser Ile 2	Asp Gly	Asp Ser	Ala	Leu	Leu	Lys 220	Pro	Asp	Thr	Pro	
Ile Pro Pro Thr	Tyr Pro 230		ı Tyr	Pro	Ala 235		Tyr	Pro	Pro	Phe 240	
Tyr Pro Tyr Pro 1	Leu Pro	Tyr Tr	Ser	-		Ser	Pro	Ala			
Pro Lys Lys Glu (245 Glu Thr	His Gl	ı Val	250 Val	Lys	Pro	Thr	Ala	255 Val	Leu	
260			265		_			270			
Ser Lys Ser Pro 1 275	ile Asn	Val Asp 280		ьeu	val	σтλ	Met 285	ser	гуз	ьеи	
Ser Leu Gly Asp 9 290	Ser Ile	Gly Asp 295) Ser	Gly	Pro	Ser 300	Ser	Leu	Ser	Arg	
Lys Leu Val Glu (305	Glu Gly 310	Pro Sei	Arg	Gln	Ser 315	Ala	Phe	His	Ala	Thr 320	
Pro Ala Cys Gly	Ser Ser	Asn Ile	e Asn	Gly	Ser	Val	Ile	His	Ala	Val	

91 92

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<210> SEQ ID NO 65 <211> LENGTH: 927

<212> TYPE: DNA

<213> ORGANISM: Lotus japonicus

325

<400> SEQUENCE: 65

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<210> SEQ ID NO 66

<211> LENGTH: 308 <212> TYPE: PRT

<213> ORGANISM: Lotus japonicus

<400> SEQUENCE: 66

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Thr Cys Thr Asp Thr Ala Ala Ala Gly Asp Asn Gly Ile Met Leu Phe

Gly Val Arg Leu Thr Glu Gly Ser Thr Ser Ser Ser Ala Phe Ile Arg

Lys Ser Ala Ser Met Asn Asn Leu Ser Gln Tyr Asn Glu Pro Glu Ser

Asn Pro Ala Asp Ala Ala Gly Tyr Ala Ser Asp Asp Val Val His Pro

Ser Ala Arg Ala Arg Asp Arg Lys Arg Gly Val Pro Trp Thr Glu Glu

Glu His Lys Leu Phe Leu Leu Gly Leu His Lys Val Gly Lys Gly Asp 105

 $\hbox{Trp Arg Gly Ile Ser Arg Asn Phe Val Lys Thr Arg Thr Pro Thr Gln } \\$ 120

Val Ala Ser His Ala Gln Lys Tyr Phe Leu Arg Arg His Asn His Asn 135 140

Arg Arg Arg Arg Ser Ser Leu Phe Asp Ile Thr Thr Asp Thr Val 155 150

-continued

Met Glu Ser Ser Thr 16e Met Glu Glu Glu Glu Asp Gln Gln Gln Glu Gln Asp Gln Gln Glu Met 175

Val Pro Pro Ala Thr Ser Ala Val Tyr Pro Pro Leu His Tyr Gly Gly 190

Phe His Gly Pro Ala Phe Pro Met Ala Pro Pro Met Ala Arg Pro Ile Arg Pro Thr Pro Ile 220

Val Ala Gly Gly Glu Arg Pro Ala Arg Pro Ile Arg Pro Ile Arg Pro Thr Pro Ile 225

Phe Pro Val Pro Pro Ser Ser Lys Met Ala Ser Leu Asn Leu Lys Glu 240

Lys Ala Ala Gly Gln Pro Ser Ser Ser Pro Pro Pro Pro Pro Ser Lys Met Ala Ser Leu Asn Leu Lys Glu 255

Ser Leu Lys Leu Gln Pro Ser Pro Pro Pro Pro Pro Ser Lys Asp His Ser Pro Ala Thr Ser Ser His Ser Pro 265

Phe Gln Ala Met Ser Ala Gly Lys Phe Ser Gly Gly Gly Asp Ser Ile Ser Val Ala Ser Val Ala Ser Val Ala Ser Lys Asp Ser Ile Ser Val Ala Ser Val Ala Ser Val Ala

What is claimed is:

- 1. A transgenic plant comprising:
- a first transgene that contains a first nucleic acid sequence operably linked to a promoter, the first nucleic acid sequence encoding a MYBS3 protein having the sequence of SEQ ID NO:1, and
- a second transgene that contains a second nucleic acid sequence operably linked to a promoter, the second nucleic acid sequence encoding a DREB1A protein having the sequence of SEQ ID NO:4,
- wherein the transgenic plant expresses both a higher level of the MYBS3 protein and a higher level of the DREB1A protein as compared to a host plant of the transgenic plant that lacks the first transgene and the second transgene, the transgenic plant being more tolerant to chill as compared to the host plant.
- 2. The transgenic plant of claim 1, wherein the plant is rice, maize, wheat, barley, sorghum, sugarcane, turf grass, Miscanthus, switchgrass, napier grass, soybean, canola, potato, tomato, bean, pea, or jatropha.
 - 3. The transgenic plant of claim 2, wherein the plant is rice.

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- 4. A method of generating the transgenic plant of claim 1, 35 comprising
 - introducing into a cell of a host plant the first transgene and the second transgene; and
 - identifying and selecting a transgenic plant that is more tolerant to chill as compared to the host plant.
 - 5. The method of claim 4, wherein the host plant is rice, maize, wheat, barley, sorghum, sugarcane, turf grass, Miscanthus, switchgrass, napier grass, soybean, canola, potato, tomato, bean, pea, or jatropha.
 - 6. The method of claim 5, wherein the host plant is rice.

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