



US 20040002109A9

(19) **United States**

(12) **Patent Application Publication**

Kostenis

(10) **Pub. No.: US 2004/0002109 A9**

(48) **Pub. Date: Jan. 1, 2004**

CORRECTED PUBLICATION

(54) **PROCESS FOR IDENTIFYING
MODULATORS OF G-PROTEIN-COUPLED
RECEPTORS**

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(21) Appl. No.: **09/899,295**

(22) Filed: **Jul. 6, 2001**

Prior Publication Data

(15) Correction of US 2002/0127601 A1 Sep. 12, 2002
See Filing Date INID (22).

(65) US 2002/0127601 A1 Sep. 12, 2002

(30) **Foreign Application Priority Data**
Jul. 8, 2000 (DE)..... 10033353.2

Publication Classification

(51) **Int. Cl.⁷** **G01N 33/53**; G01N 33/567

(52) **U.S. Cl.** **435/7.1**; 435/7.2

(57) **ABSTRACT**

The invention relates to a widely applicable process for identifying chemical compounds, which modulate G-protein-coupled receptors, by means of novel hybrid G-proteins with broad receptor specificity and very high expression and also to chemical compounds which can be identified by such a process.

Fig. 1

		1	7	9	10	<u>αN Helix</u>	
α_q (WT α)	MTLESIMAC					EEAK	-
-6α	MAC					EEAK	-
α_{11}	MTLESMMAC					DEVK	-
$\alpha_{i1,3}$	MGC					AEDK	-
$\alpha_{o1,2}$	MGC					AEER	-
α_{t1}	MGA					AEEK	-
α_s	MGCLGNSKTEDQ					RNEEK	-

Fig. 2

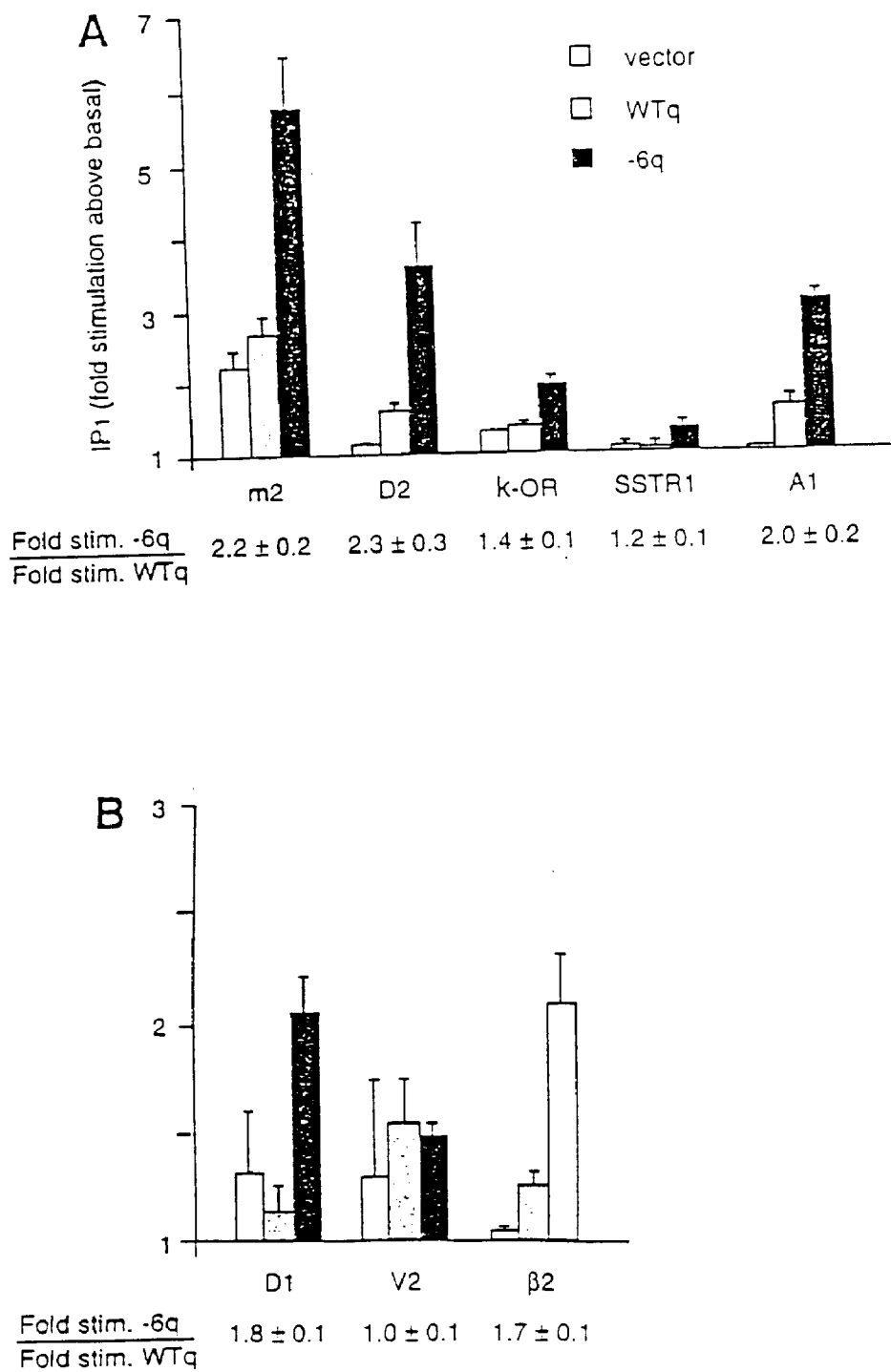


Fig. 3

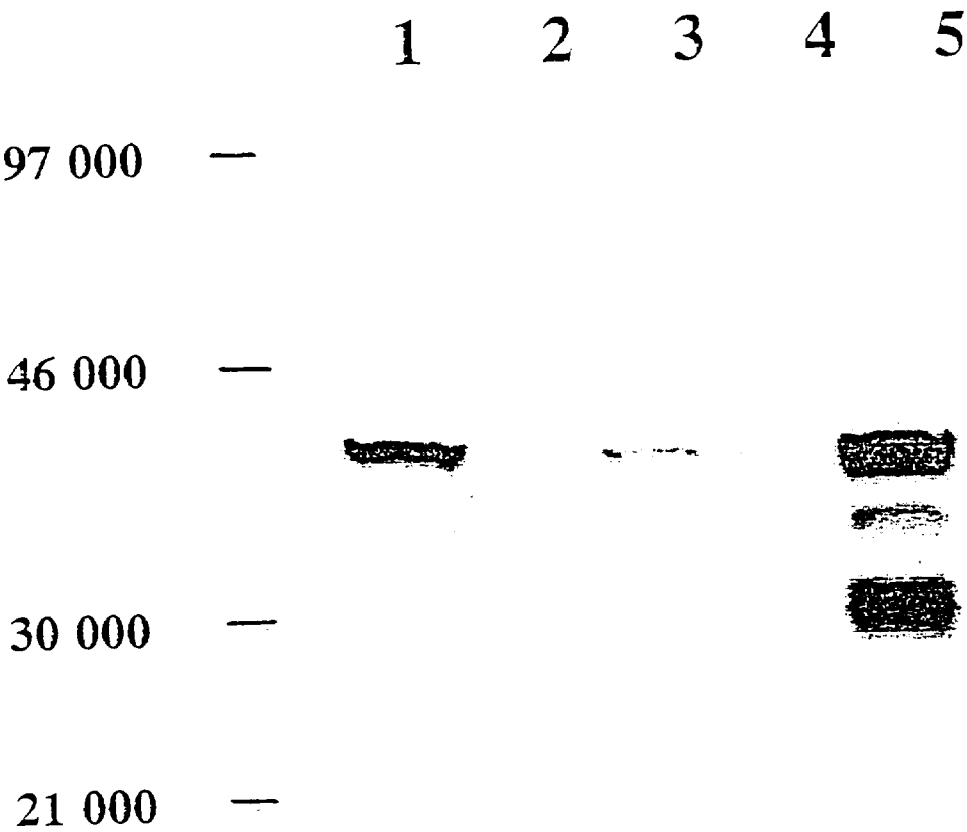


Fig. 4

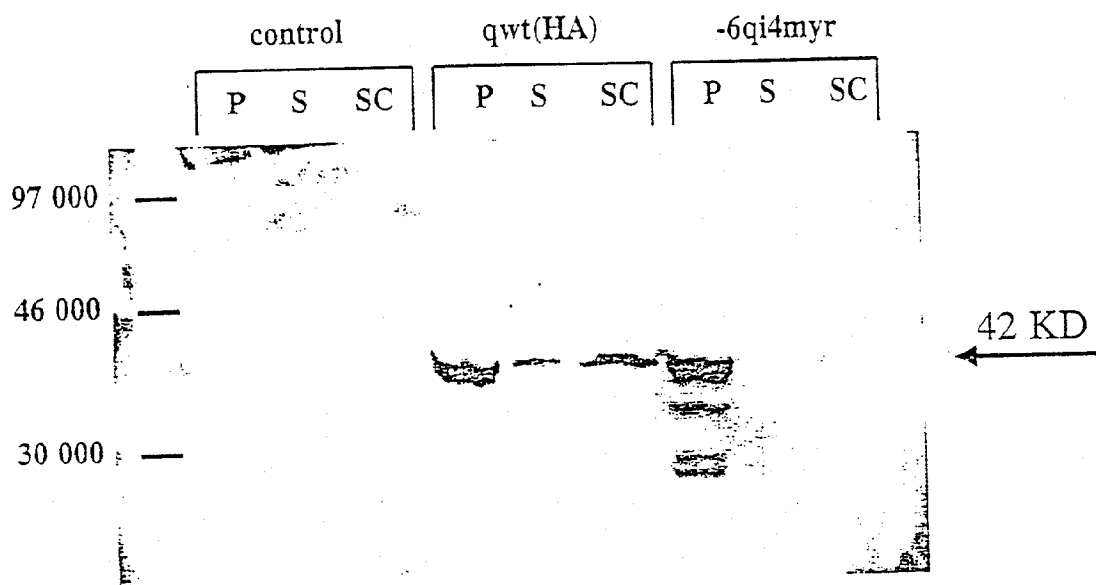


Fig. 5

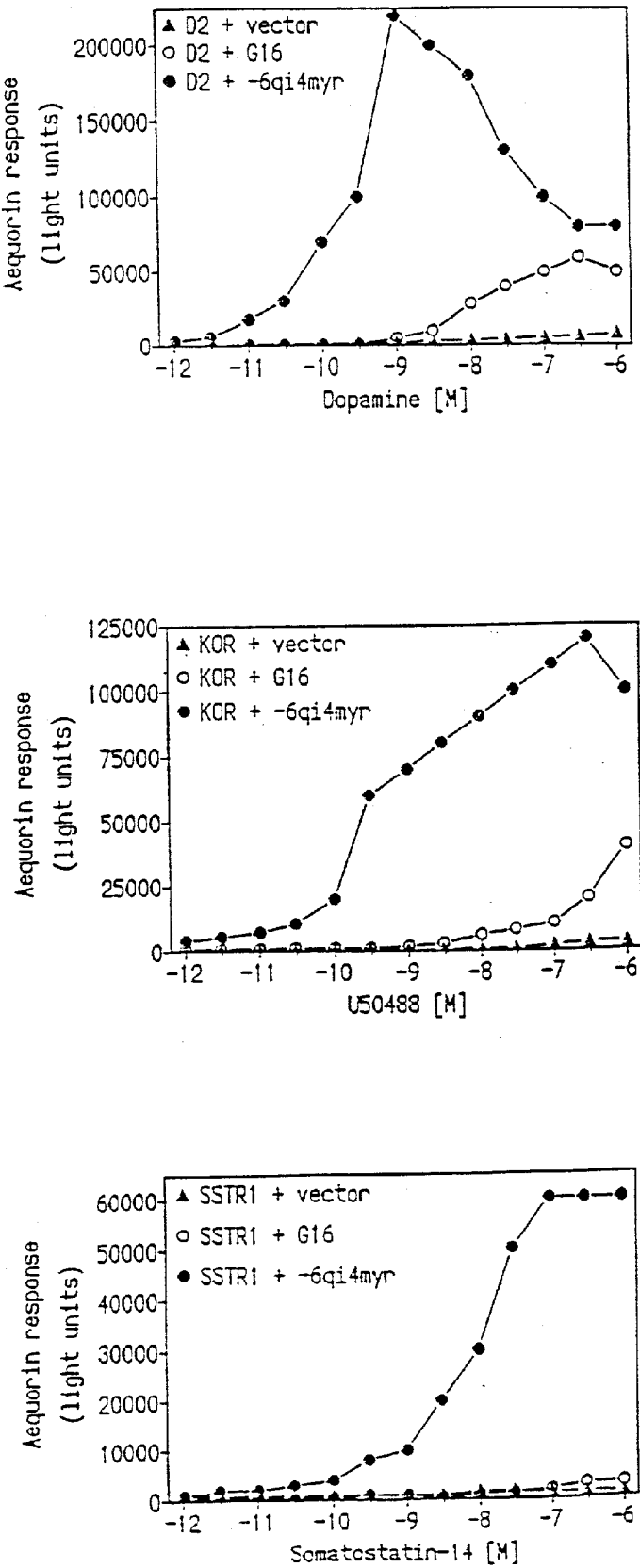


Fig. 6

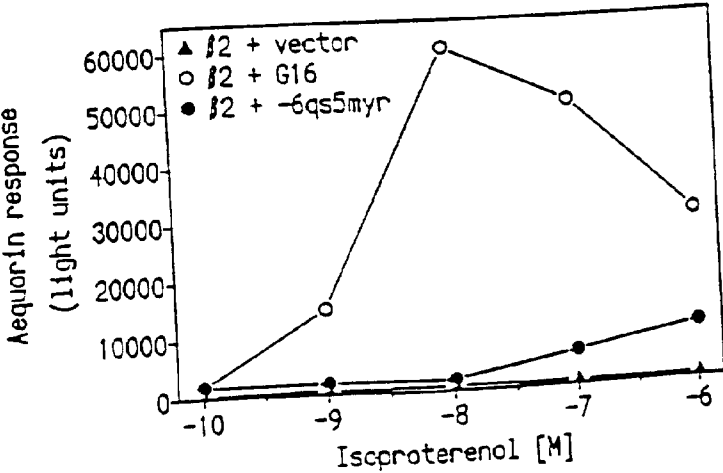
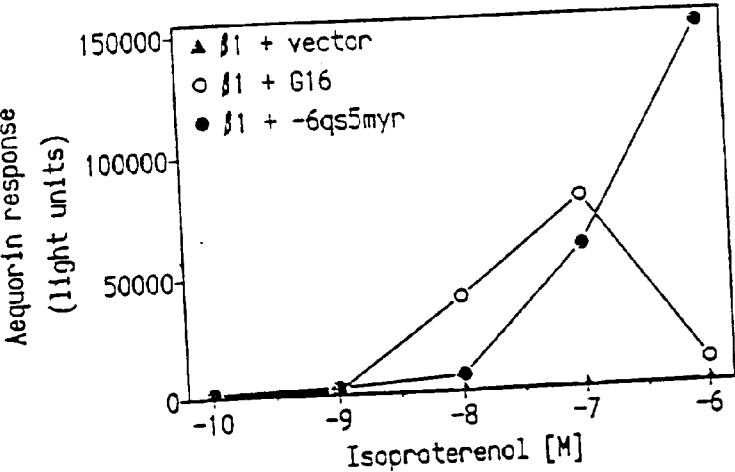
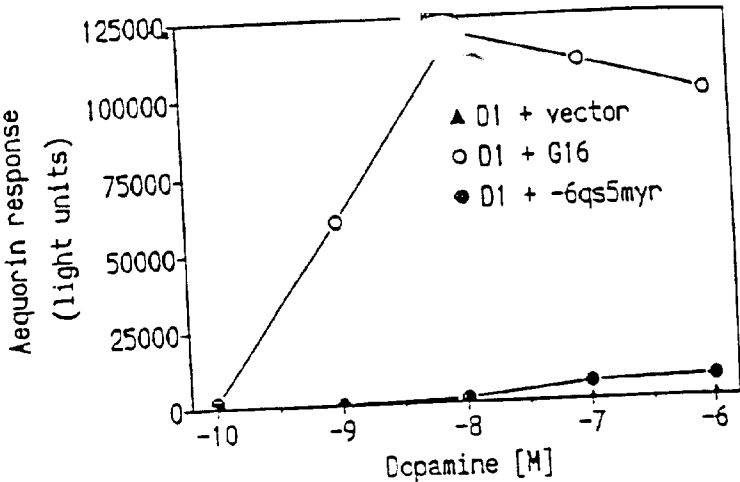
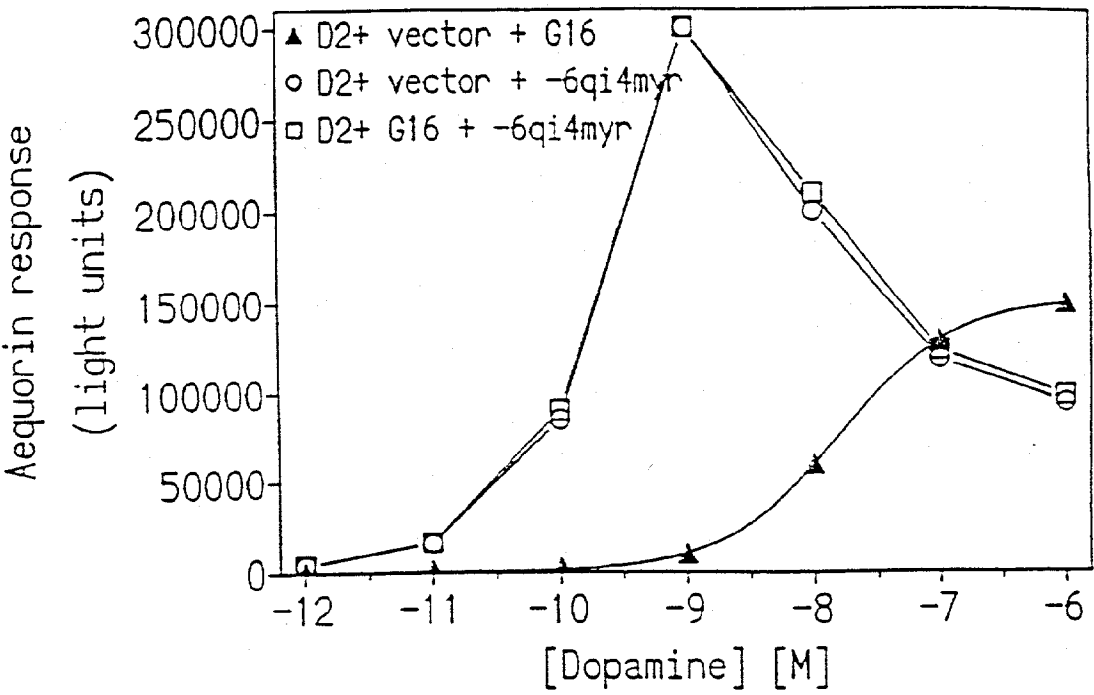


Fig. 7



PROCESS FOR IDENTIFYING MODULATORS OF G-PROTEIN-COUPLED RECEPTORS

TECHNICAL FIELD

[0001] The invention relates to a process for identifying chemical compounds which modulate G-protein-coupled receptors, by means of novel hybrid G-proteins with broad receptor specificity, and also to chemical compounds which can be identified by such a process

BACKGROUND OF THE INVENTION

[0002] G-protein-coupled receptors (GPCRs) play an important role in a multiplicity of physiological processes. They are one of the most important protein families known to date, and it is assumed that in the human genome about 1000 genes code for members of this receptor class. GPCRs have a characteristic structure: they are peptide threads which meander in the form of α -helices seven times through the phospholipid bilayer of the cell membrane, arranging themselves in a circle. It is estimated that about 60% of the pharmaceuticals presently available through prescription bind to GPCRs. This underlines the importance of this receptor class to the pharmaceutical research industry.

[0003] G-protein-coupled receptors share a common mechanism of action. Binding of an extracellular ligand leads to a conformational change in the receptor protein that allows it to make contact with a guanine-nucleotide binding protein (G-protein). G-proteins are located on the cytoplasmic side of the plasma membrane and mediate the extracellular signal in the cell interior to trigger various intracellular reactions.

[0004] GPCRs are the most important therapeutic target proteins to date. An estimated 40% of the pharmaceuticals prescribed by doctors act as agonists or antagonists of GPCRs. Owing to the size and importance of this protein family and in view of the fact that chemical binding partners for many GPCRs are unknown (orphan GPCRs), it can be assumed that this receptor class will be one of the most important reservoirs for suitable target proteins in the search for novel medicinal substances in the future.

[0005] GPCRs are integral membrane proteins that transfer a signal mediated via a mostly hydrophilic signal substance bound to the outer side of the cell into the cell interior via a family of G-proteins. Depending on the receptor specificity and the G-proteins activated thereby, activated GPCRs trigger various signal transduction pathways. Depending on the receptor type, various actions are evoked, all of which lead to the formation of second messengers. Second messengers are intracellular messenger molecules, such as, for example, cAMP, cGMP, and Ca^{2+} , formed in or released into the cytosol in response to an extracellular signal and which trigger reactions in the cell through the activation or deactivation of intracellular proteins. Thus, activation of a membrane-bound adenylate cyclase may lead to an increase in the intracellular cAMP level, and inhibition may lead to a decrease. Stimulation of a cGMP-specific phosphodiesterase may lead to a reduction in the cGMP level. The activated G-protein can also lead, for example, to an increase of Ca^{2+} or K^{+} ions by binding to an ion channel. Furthermore, an activated G-protein can affect activation of a phospholipase and thus formation of inositol 1,4,5-tris-

phosphate and diacylglycerol. This, in turn, leads either to a Ca^{2+} increase or to activation of a protein kinase C, with further effects in both cases.

[0006] The heterotrimeric G-proteins are located on the inside of the plasma membrane. They comprise the three subunits α , β and γ . When an activated receptor makes contact with the G-protein heterotrimer, it dissociates into an α subunit and a $\beta\gamma$ complex. Both the activated α subunit and the $\beta\gamma$ complex can influence intracellular effector proteins. The G-protein α subunit family is presently divided into four different classes ($\text{G}\alpha\text{s}$, $\text{G}\alpha\text{i}$, $\text{G}\alpha\text{q}$ and $\text{G}\alpha\text{12}$ classes).

[0007] GPCRs are classified according to the G-proteins that they contact. GPCRs of the $\text{G}\alpha\text{s}$ class mediate adenylate cyclase stimulation via activation of $\text{G}\alpha\text{s}$ and increase the intracellular cAMP concentration. GPCRs of the $\text{G}\alpha\text{i}$ class mediate adenylate cyclase inhibition via activation of $\text{G}\alpha\text{i}$ and decrease intracellular cAMP. GPCRs of the $\text{G}\alpha\text{q}$ class mediate stimulation of various phospholipase C beta ($\text{PLC}\beta$) isoforms via activation of $\text{G}\alpha\text{q}$ and lead to hydrolysis of membrane-bound phosphatidylinositol 4,5-bisphosphate to give diacylglycerol and inositol 1,4,5-trisphosphate (IP3). IP3 releases Ca^{2+} from intracellular depots.

[0008] Most GPCRs can make contact only with one G-protein α subunit family, and, therefore, are selective for a particular signal transduction pathway. This narrow specificity is a great hindrance to the identification of chemical compounds capable of modulating GPCR-dependent signal transduction pathways.

[0009] Moreover, a suitable signal which can be utilized in a screening assay with high sample throughput is obtained only from those signal transduction pathways in which, for example, G-protein activation leads to an increase in the intracellular Ca^{2+} level.

[0010] Hybrid G-proteins with altered receptor specificity and signal transduction pathway linkage may be constructed by joining together parts of various G-proteins using known molecular biology and biochemistry methods.

[0011] Hybrid G-proteins are fusion constructs which combine sequences of various $\text{G}\alpha$ subunits within one protein. Thus it is possible, for example by fusion of the $\text{G}\alpha\text{i}$ receptor recognition region to the $\text{G}\alpha\text{q}$ effector activation region, to prepare a $\text{G}\alpha\text{q/i}$ hybrid which receives signals from $\text{G}\alpha\text{i}$ -coupled receptors but switches on the $\text{G}\alpha\text{q-PLC}\beta$ signal transduction pathway. Such a hybrid, in which the C-terminal 5 amino acids of $\text{G}\alpha\text{q}$ is replaced by the corresponding $\text{G}\alpha\text{i}$ sequence ($\text{G}\alpha\text{qi5}$), was first described by Conklin et al., Nature 363, 274-276 (1993).

[0012] This "recoupling" of receptors has the advantage that the assay endpoint (increase in intracellular Ca^{2+} concentration in comparison with adenylate cyclase inhibition) is more readily accessible through measurement methods and can be used in high throughput screening.

[0013] However, the disadvantage of the $\text{G}\alpha\text{q/G}\alpha\text{i}$ fusion constructs is that they are unable to activate some GPCRs, such as, for example, the SSTR1 receptor qi5 (Conklin et al., Mol. Pharmacol. 50, 885-890 (1996)).

[0014] Similarly, fusion constructs between $\text{G}\alpha\text{q}$ and $\text{G}\alpha\text{s}$ have been described. These too have the disadvantage that they cannot link all $\text{G}\alpha\text{s}$ -coupled receptors to the $\text{PLC}\beta$ signal

transduction pathway, such as the β 2-adrenergic receptor and the dopamine D1 receptor, for example.

[0015] Besides C-terminal modifications for altering the linking of receptors to particular signal transduction pathways, an N-terminal modification of G α q has been described which allows the G-protein to receive and pass on signals from several different receptors. In this G α q protein, the 6 highly conserved N-terminal amino acids were deleted (Kostenis et al., J. Biol. Chem. 272, 19107-19110 (1997)). This deletion allows the resulting Gq (also called -6q) to receive signals not only from Gq- but also from Gs- or Gi/o-coupled receptors and to pass them on to PLC β .

[0016] This mutant G α subunit also recognizes receptors such as the SSTR1 somatostatin receptor, the dopamine D1 receptor and the adrenergic β 2 receptor. However, even this mutant is unable to recognize the receptor edg5. Moreover, the signal intensity of this mutant is so weak that it is unusable in practice (Kostenis et al., J. Biol. Chem. 272, 19107-19110 (1997)).

[0017] Another known G α subunit is G α 16 which links GPCRs from various functional classes to the PLC β -Ca²⁺ signal transduction pathway. G α 16 is a G-protein with broad receptor specificity and has been disclosed in WO 97/48820 (title: Promiscuous G-protein compositions and their use). G α 16 is practically nonselective by nature. But even this subunit is not universally applicable, because receptors such as the edg5 receptor or the SSTR1 somatostatin receptor couple to it only weakly, if at all.

[0018] Thus, it would be very useful if a G-protein were available that could be activated by other functional GPCR classes, could also give a sufficiently strong signal in the cell. Such a G-protein could be utilized in a screening assay, such as a high throughput screening assay, to identify compounds modulating GPCRs and/or the appropriate dependent signal transduction pathways, for example a signal such as the increase or decrease in the intracellular Ca²⁺ concentration.

[0019] The object of the present invention is therefore to provide further hybrid G-proteins characterized by having recognizable broad specificity with respect to GPCRs. These G-proteins can be used in screening processes to identify chemical compounds by the coupling of the G-proteins to a signal pathway leading to an increase in the intracellular Ca²⁺ concentration. In addition, these proteins can be expressed at such a high level that signal intensity is improved.

SUMMARY OF THE INVENTION

[0020] The invention relates to a process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism, wherein said process comprises:

[0021] a) providing at least one cell which contains at least one GPCR-dependent signal transduction pathway and which produces one or more than one G-protein;

[0022] b) providing at least one chemical compound to be studied;

[0023] c) contacting the cell or cells of a) with one or more chemical compounds of b);

[0024] d) determining the quantitative or qualitative effect of the chemical compounds of b) on the signal transduction pathway of the cells of a) by means of a signal transduction pathway-dependent measurable signal.

BRIEF DESCRIPTION OF THE DRAWINGS

[0025] FIG. 1 represents an alignment of the amino-terminal regions of various G α proteins.

[0026] FIG. 2 shows a stimulation of the PLC β signal transduction pathway by means of the -6q-G α protein variation by Gi/o-coupled (A) and Gs-coupled (B) receptors using the maximum concentration of the relevant agonist.

[0027] FIG. 3 shows an SDS-PAGE Western blot with increased expression of -6qi4myr in comparison with -6qi4. The expression of other G α proteins is also shown.

[0028] FIG. 4 depicts an SDS-PAGE Western blot showing fractionation of qWT and -6qi4myr into a membrane-containing particle fraction (P) and a soluble fraction (S; SC). The G-protein α subunits were detected by the 12CA5 monoclonal antibody resulting in protein bands of ~42 KD.

[0029] FIG. 5 shows the linking of various Gi/o-coupled receptors to the PLC β signal transduction pathway by -6qi4myr. D2, KOR and SSTR1 are Gi/o-coupled receptors. The controls used were a vector construct and the G α 16 protein (G16).

[0030] FIG. 6 shows that Gs-coupled receptors are linked to the PLC β signal transduction pathway by -6qs5myr. β 1, β 2 and D1 are Gs-coupled receptors. A vector construct and the G-protein G α 16 (G16) serve as references.

[0031] FIG. 7 shows the linking of the Gi/o-coupled dopamine D2 receptor to the PLC β -Ca²⁺ signal transduction pathway in the presence of the low-sensitivity α subunit G α 16 (G16), in the presence of the very sensitive G α subunit -6qi4myr and in the presence of a combination of G α 16 and -6qi4myr. It is evident that the potential activation of calcium release by -6qi4myr is not adversely affected by the presence of G α 16.

DETAILED DESCRIPTION OF THE INVENTION

[0032] The action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism can be modified in an inhibiting or stimulating manner by a chemical compound. A chemical compound presents an inhibiting effect if the signal transduction pathway-dependent measurable signal is weaker in the presence of the chemical compound than in its absence. Compounds evoking such an effect are called antagonists. A chemical compound presents a stimulating effect if the signal transduction pathway-dependent measurable signal is stronger in the presence of the chemical compound than in its absence. Such compounds are called agonists.

[0033] In one embodiment of the invention, the process makes use of a cell which produces at least two G-proteins. Said G-proteins may depend on one or on different GPCRs. In principle, all G-proteins are suitable for carrying out the

process according to the invention, regardless of their receptor specificity, their sequence, their structure, the species for which they are specific, or the cell, tissue or organ from which they originate.

[0034] In one embodiment, cells producing at least one G-protein from among -6qi4myr, -6qs5myr, -6qi4, -6qs5 are used. The G-proteins -6qi4myr, -6qs5myr, -6qi4, -6qs5 are hybrid G-proteins assembled from portions of different mouse G-proteins, in some cases, containing additional modifications. The G-proteins may be produced by the cell individually or in combination. Apart from the hybrid G-proteins already mentioned, a cell may produce Gα16. Further, each of the G-proteins may be present in a cell individually or in combination with one or more other G-proteins. Gα16 should always be produced in a cell in combination with another of the G-proteins mentioned above.

[0035] The names and amino acid sequences of some example G-proteins according to the invention are as follows: -6qi4myr, SEQ ID NO:2; -6qi5myr, SEQ ID NO:4; -6qi4, SEQ ID NO:6; -6qs5, SEQ ID NO:8, and Gα16, SEQ ID NO:10.

[0036] The chemical compound is commonly provided in soluble form, for example dissolved in water. Besides the solvent, the solution may contain buffer substances, salts, or auxiliaries such as solubilizers, detergents, preservatives, or other substances.

[0037] Provision of a cell includes its production, cultivation, and processing. Cells are provided, for example, by preparing suitable cell material from organs or tissues, or by propagating suitable cell lines or microorganisms. Various suitable culture media can be used for cultivation. The cells are maintained at the optimum temperature for the organism from which they are provided. Where appropriate, preservatives, antibiotics, pH indicators, blood serum components, blood serum, auxiliaries, or other substances are added to the growth medium. Processes for production, cultivation and further processing are described in standard textbooks (One example: Basic Cell Culture; Ed. J. M. Davis; IRL Press; 1994).

[0038] In some embodiments of the process described above, the cell of a vertebrate, insect, or yeast species, or of *Caenorhabditis elegans* (*C. elegans*) is provided. In some embodiments, a HeLa, 293, COS, or CHO cell, or a *Saccharomyces cerevisiae* cell is provided.

[0039] In one embodiment of the invention, the intracellular Ca²⁺ concentration is used as a signal transduction pathway-dependent measurable signal for determining the quantitative or qualitative effect of a chemical compound to be studied on a cell signal transduction pathway. The change in intracellular Ca²⁺ concentration can be detected, for example, by using aequorin, a dye, or by the FLIPR™ technique from Molecular Devices Corp. (1311 Orleans Ave., Sunnyvale, Calif. 94089; 800-635-5577).

[0040] In another embodiment, the processes as described above may be used for identifying a pharmaceutical.

[0041] The invention also relates to at least one chemical compound which modifies the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism, with said chemical compound

being identified by at least one process of this invention. Such chemical compounds could include, for example, hormones, scents, or pharmaceuticals that alter the chemical structure of hydrophilic signal substances which induce GPCRs.

[0042] The invention further relates to a polynucleotide sequence coding for a polypeptide having the property of a G-protein, which comprises a polypeptide selected from:

[0043] a) a polypeptide having an amino acid sequence according to SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6 or SEQ ID NO:8;

[0044] b) a polypeptide according to a) lacking one or more amino acids;

[0045] c) a polypeptide according to a) having an additional one or more amino acids;

[0046] d) an allelic variant of the polypeptide according to a).

[0047] The allelic variants include polypeptides comprising a polynucleotide sequence of an allelic variant of the corresponding gene. An allelic variant of a gene is an alternate form occupying the same locus in a particular chromosome or linkage structure and differing from other alleles of the locus at one or more mutational sites.

[0048] In addition, the invention relates to a polynucleotide comprising a polynucleotide sequence selected from:

[0049] a) a polynucleotide sequence according to SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5 or SEQ ID NO:7 or the corresponding sequence complementary thereto;

[0050] b) a polynucleotide sequence hybridizing with a polynucleotide sequence according to a) under stringent conditions.

[0051] The stringency is determined by the temperature and salt content. By varying the stringency, it is possible to adjust the extent of base pairing of two homologous nucleotide sequences. The extent of base pairing also depends on the length and base composition of a polynucleotide. Stringent conditions in accordance with this invention are present if 95% or more of the polynucleotide sequence and the hybridizing sequence are complementary.

[0052] In one embodiment of a polynucleotide sequence or a polynucleotide as described above, the polynucleotide is part of a recombinant vector construct. Recombinant vector constructs may be prepared with the help of knowledge in the art as illustrated, for example, in F. M. Ausubel et al., Current Protocols in Molecular Biology, Wiley & Sons, New York. The preparation entails inserting a polynucleotide coding for an amino acid sequence according to the sequence information described above (SEQ ID NO:2, 4, 6, or 8) or a polynucleotide sequence according to the sequence information described above (SEQ ID NO:1, 3, 5, or 7) into a base vector. A base vector is a vector into which a polynucleotide sequence can be inserted using molecular biology methods, and which can be cloned in a microorganism, for example, a bacterial, fungal, or cell culture cell. The base vector may comprise, for example, a phage, phagemid, plasmid, cosmid, viral, yeast artificial chromosome (YAC) or other type of vector. Non-limiting examples of base vectors are pUC18, pUC19, pBluescript, pKS, and pSK. The

base vector may comprise, for example, a plasmid having an antibiotic resistance marker, an origin of replication suitable for propagating the plasmid in bacteria or cell cultures, and a promoter suitable for expressing the genes comprised in the inserted polynucleotide sequence. The polynucleotide sequence is inserted via suitable restriction cleavage sites using appropriate restriction enzymes commercially available from companies such as New England BioLabs, Roche Diagnostics, Stratagene, and others. Such restriction cleavage sites may be those of the restriction enzymes BamHI, EcoRI, SalI, and EcoRV, for example.

[0053] In another embodiment, the recombinant vector construct comprises an expression vector usable in eukaryotes and/or prokaryotes. An expression vector contains a promoter which can be linked functionally to a polynucleotide sequence so that a protein encoded by said polynucleotide sequence is synthesized in a microorganism, for example, such as a bacterium or a fungus, or in the cell of a eukaryotic cell line. The promoter may be inducible, by means of tryptophan for example, or may be constitutive. Some non-limiting examples of expression vectors are pUC18, pUC19, pBluescript, and pCDNA3.1.

[0054] The invention further relates to a host cell which may comprise a polynucleotide or a recombinant vector construct as described above. In one embodiment, the host cell comprises a human cell. In other embodiments, the host cell comprises the cell of a vertebrate, insect, bacterium, or yeast species, or *C. elegans*. In yet other embodiments, the cell comprises a HeLa, 293, COS or CHO cell, or an *Escherichia coli* or *Saccharomyces cerevisiae* cell. Other eukaryotic cells or cell lines, or other bacteria, such as *Bacillus* or *Streptomyces* species, and fungi, such as *Penicillium* or *Aspergillus* species, may also be used.

[0055] The invention also relates to the production of a host cell as described above by introducing a polynucleotide according to one or more of the polynucleotide sequences as disclosed in SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, and 8 or a recombinant vector construct as characterized above into a eukaryotic or prokaryotic cell. The polynucleotide sequences may be introduced for example, by electroporation, by Ca^{2+} phosphate precipitation of the eukaryotic or prokaryotic cells together with the polynucleotide sequence, or by other transformation methods.

[0056] A host cell of this kind may be used for carrying out an above-described process of this invention.

[0057] The invention also relates to a protein having an amino acid sequence selected from: SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8.

[0058] Moreover, the invention relates to a process for preparing a protein comprising an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8, wherein the process comprises the following steps:

[0059] a) producing a host cell containing an appropriate polynucleotide sequence and prepared as described above;

[0060] b) cultivating said host cell in a growth medium suitable for the host cell and inducing expression of the protein encoded by the polynucleotide sequence;

[0061] c) obtaining the cell material and disrupting the cells;

[0062] d) removing the protein by means of biochemical methods for protein purification.

[0063] For preparing and purifying the proteins denoted, known methods, as described in F. M. Ausubel et al., *Current Protocols in Molecular Biology*, Wiley & Sons, New York, may be used accordingly.

[0064] A protein having an amino acid sequence according to SEQ ID NO:2, 4, 6, or 8 or prepared according to the process described may be used for producing antibodies.

EXAMPLES

Example 1

[0065] Activation of a Signal Transduction Pathway via the $\text{G}\alpha$ -protein Mutant -6q by Various Receptors

[0066] COS7 cells were cultured in DMEM (Dulbecco's modified Eagle's medium) with 10% FCS (fetal calf serum) at 37° C. (5% CO_2). For transfections, 1×10^6 cells were seeded in 100-mm plates. About 24 hours later, the cells were cotransformed with the expression plasmids αq or -6q (1 μg DNA/100 mm plate) and, in each case, one of the following receptor constructs (4 μg DNA/100 mm plate): M2 (muscarinic receptor in pCD), D2 (dopamine receptor in pCDNAI), kappa (opioid receptor in pCDNA3), SSTR1 (somatostatin receptor in pCMV), A1 (adenosine receptor in CDM7), D1 (dopamine receptor in pCDNAI), V2 (vasopressin receptor in pCD-ps), $\beta 2$ (adrenergic receptor in pSVL).

[0067] About 24 hours after transfection, the cells were divided into equal portions in 6-well plates and 3 $\mu\text{Ci/ml}$ ^3H -myo-inositol (20 Ci/mmol) in DMEM was added. After incubation for 24 hours, the cells were incubated with HBSS (Hank's Balanced Salt Solution; +10 mM LiCl) at room temperature for 20 minutes. The cells were then stimulated with the appropriate agonists for one hour, and the increase in intracellular inositol monophosphates (IP1) was determined by anion exchange chromatography. IP1 is a signal molecule, that is generated in the PLC- β -signal transduction pathway and leads in the further course of the signal transduction to an increase in intracellular Ca^{2+} concentration.

[0068] The results which follow were obtained with the $\text{G}\alpha$ -protein construct -6q. Compared with the wild-type sequence (WTq), denoted αq in FIG. 1, this mutant lacks the six highly conserved amino acid residues at the amino-terminal end, as depicted in FIG. 1. Moreover, FIG. 1 presents additional sequence examples. Mutants of this kind and receptor constructs used were prepared with the aid of standard molecular biological methods, as described in detail, for example in F. M. Ausubel et al., *Current Protocols in Molecular Biology*, Wiley & Sons, New York.

[0069] COS7 cells expressing WTq or -6q various Gi/o-coupled receptors (A) or Gs-coupled receptors (B) were incubated (37° C.) in the presence and absence of the appropriate agonists (see below) for 1 hour. The increase in intracellular IP1 concentration was determined as described above. The data represent averages $\pm$ S.E. of 3-7 independent

experiments, with each determination performed in triplicate. The following ligands were used:

[0070] FIG. 2A: m2 (muscarinic receptor): carbachol (100 μ M); D2 (dopamine receptor): (-)-quinpirole (10 μ M); K-OR (kappa (opioid receptor)): (-)-U50488 (10 μ M); SSTR1 (somatostatin receptor): somatostatin 14 (1 μ M); B, A1 (adenosine receptor): R(-)-PIA (10 mM);

[0071] FIG. 2B: D1 (dopamine receptor): dopamine (1 mM); V2 (vasopressin receptor): AVP (1 nM); β 2 (adrenergic receptor): (-)-isoproterenol (200 μ M). The numbers below the figures indicate the extent of the particular PLC stimulation as relative increase in PLC stimulation from -6q to WTq.

[0072] FIG. 2 shows that the G α -protein mutant -6q stimulates IP1 formation depending on different receptor classes. The experimental results for -6q in **FIG. 2** are compared with stimulation of IP1 by means of the wild-type construct (WTq) and with the vector construct without any G α insert (vector). IP1 release by means of the -6q construct succeeds both with Gi/o-coupled (**FIG. 2A:** m2, D2, k-OR, SSTR1, A1) and with Gs-coupled (**FIG. 2B:** D1, V2, β 2) receptors.

Example 2

[0073] Preparation of Highly Expressed Mutants of G α Proteins with Broad Receptor Specificity

[0074] Hybrid G-protein α subunits, that lack the six highly conserved amino acids of the amino terminus and that simultaneously have either an α i or α s sequence at the C terminus were constructed. They are denoted -6qi4 or -6qs5, corresponding to the α i sequence or α s sequence they contain. The construct -6qi4 links the Gs-coupled receptors and also some of the Gi/o-coupled receptors, such as the SSTR1 and edg5 receptors, to the PLC β signal transduction pathway. G α 16 cannot link the edg5 receptor to the PLC β signal transduction pathway. G α 16 is a G-protein with broad receptor specificity and has been disclosed in WO 97/48820 (title: Promiscuous G-protein compositions and their use).

[0075] The construct -6qs5 links the Gi/o-coupled receptors and the Gs-coupled receptors to the PLC β signal transduction pathway and also recognizes receptors such as the dopamine D1 receptor or the adrenergic β 2 receptor.

[0076] A combination of the two G-protein α subunits -6qi4 and -6qs5 in one cell line thus recognizes a wider range of GPCRs than each subunit separately or than G α 16.

[0077] The applicability of -6qi4 and -6qs5 G α subunits in technical screening procedures could be further improved if their expression were increased, because this would result in a stronger signal.

[0078] For this reason, additional myristoylation/palmitoylation recognition sequences were inserted into the amino-terminal region of the G α subunits to produce -6qi4myr and -6qs5myr from -6qi4 and -6qs5, respectively. The protein sequence of -6qi4myr and -6qs5myr at the amino terminus is MGCC, in contrast to MACC in the original sequence of the -6q variants. Therefore, the novel constructs, -6qi4myr and -6qs5myr, contain a consensus sequence for myristoylation/palmitoylation. It is known that removing myristyl or palmityl residues from G-proteins leads to a redistribution in the cell. Loss of palmitate or

myristate residues influences the expression pattern of the G-proteins in such a way that G-protein α subunits are found both in the cell membrane and in the cytosol, but are mainly cytosol-localized. However, only the membrane-bound G-proteins can pass the signals from GPCRs on to intracellular effectors. Only the consequences of removing a consensus sequence for palmitoylation/myristoylation by mutation were known. It was not known if introducing an additional consensus site for myristoylation/palmitoylation into the G α deletion mutants would affect expression. However, it was possible to show that introducing additional palmitoylation/myristoylation sites increases the amount of G α subunits expressed in the cell membrane (**FIG. 3, FIG. 4**). The SDS-PAGE Western blot (sodium dodecyl sulfate polyacrylamide gel electrophoresis Western blot) in **FIG. 3** shows distinctly increased expression of -6qi4myr compared to -6qi4. **FIG. 4** depicts an SDS-PAGE Western blot of a fractionation of qwt and -6qi4myr into a membrane-containing particle fraction (P) and a soluble fraction (S; SC). The variant with a higher degree of myristoylation/palmitoylation, -6qi4myr, is present only in the particle fraction.

[0079] For the SDS-PAGE Western blot, all G-protein α subunits were detected by the 12CA5 monoclonal antibody (coupled to horseradish peroxidase; Roche Biosciences), which is directed against the HA epitope tag contained in all of the G-protein constructs (generally the peptide sequence YPYDVPDYA). In qWT the HA tag replaces amino acids 125-130, while in the N-terminally deleted G-proteins (-6q, -6qi4, -6qi4myr) it replaces amino acids 119-124. 20 μ g of membrane protein, prepared from transfected COS7 cells, were in each case fractionated by means of SDS PAGE gel electrophoresis (for example, at 10% polyacrylamide) and blotted onto nitrocellulose, and the G-protein α subunits were detected by the 12CA5 antibody. Immunoreactive G-proteins were visualized using a chemiluminescence system (Amersham).

Example 3

[0080] Stimulation of Various Highly Expressed G α -proteins with Broad Receptor Specificity by Different Receptors

[0081] Stimulation of the highly expressed G α variants, -6qs5myr and -6qi4myr, by different receptors is depicted in **FIG. 5** and **FIG. 6**. **FIG. 5** shows that -6qi4myr is connected by Gi/o-coupled receptors (for example, dopamine D2, edg5, CCR5, SSTR1, and KOR) to the PLC β signal transduction pathway and leads to a strong signal which is proportional to Ca²⁺ release. The controls used were a vector construct and the G α 16 protein (G16). **FIG. 6** shows that Gs-coupled receptors are linked to the PLC β signal transduction pathway by -6qs5myr. The G-protein G α 16 (G16) acted as a control.

[0082] To experimentally determine the released Ca²⁺ concentration with the aequorin system, CHO cells were cotransfected with the apo-aequorin expression plasmid cytAEQ/pCDNAI, the receptor DNA mentioned above (for example, SSTR1, KOR, D2, D1, or β 2) and the G-protein α subunits G α 16 and -6qi4myr with the use of lipofectamine. After incubation in OPTIMEM medium for 10 hours, the cells were washed once with RPMI 1640 medium and incubated with 5 μ M coelenterazine f in RPMI 1640 at 37° C. for 2 hours. The cells were then washed twice with PBS and stimulated using the appropriate receptor agonists: somatostatin 14 for the SSTR1 receptor, U50488 for the

kappa opioid receptor, (-)-quinpirole for the dopamine D2 receptor, dopamine for the dopamine D1 receptor and isoproterenol for the β 2 receptor. Agonist stimulation of Gi/o-coupled receptors (SSTR1, KOR, and D2) and Gs-coupled receptors (D1 and β 2) leads to activation of the G-proteins $G\alpha_{16}$ and -6qi4myr followed by stimulation of PLC β and intracellular Ca^{2+} release. Ca^{2+} binding to the apo-aequorin-coelenterazine complex leads to light emission which was measured using a luminometer (TOPCOUNT®, Hewlett Packard).

SEQUENCE LISTING

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	130	135	140
Tyr Asp Arg Arg Arg	Glu Tyr Gln Leu Ser	Asp Ser Thr Lys Tyr	Tyr
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	165	170	175
Gln Asp Val Leu Arg	Val Arg Val Pro Thr	Thr Gly Ile Ile Glu	Tyr
	180	185	190
Pro Phe Asp Leu Gln	Ser Val Ile Phe Arg	Met Val Asp Val Gly	Gly
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Gln Arg Ser Glu Arg	Arg Lys Trp Ile His	Cys Phe Glu Asn Val	Thr
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Ser Ile Met Phe Leu	Val Ala Leu Ser Glu	Tyr Asp Gln Val Leu	Val
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Glu Ser Asp Asn Glu	Asn Arg Met Glu Glu	Ser Lys Ala Leu Phe	Arg
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Thr Ile Ile Thr Tyr	Pro Trp Phe Gln Asn	Ser Ser Val Ile Leu	Phe
	260	265	270
Leu Asn Lys Lys Asp	Leu Leu Glu Glu Lys	Ile Met Tyr Ser His	Leu
	275	280	285
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Ala Arg Glu Phe Ile	Leu Lys Met Phe Val	Asp Leu Asn Pro Asp	Ser
	305	310	315
Asp Lys Ile Ile Tyr	Ser His Phe Thr Cys	Ala Thr Asp Thr Glu	Asn
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<213> ORGANISM: Mus musculus

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Phe Ile Lys Gln Met Arg Ile Ile His Gly Ser Gly Tyr Ser Asp Glu
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Met Gln Ala Met Ile Arg Ala Met Asp Thr Leu Lys Ile Pro Tyr Lys
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agacgacgag aatatcaatt atctgactct accaaatact atcttaatga ctgggaccgc 480

gtagctgacc ctgcctacct gcctacgcaa caagatgtgc ttagagttcg agtccccacc 540

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His	Phe	Thr	Cys	Ala	Thr	Asp	Thr	Glu	Asn	Ile	Arg	Phe	Val	Phe	Ala	325	330	335	
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Tyr Glu His Asn Lys Ala His Ala Gln Leu Val Arg Glu Val Asp Val
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  1             5             10             15
Asp Glu Lys Ala Ala Ala Arg Val Asp Gln Glu Ile Asn Arg Ile Leu
          20             25             30
Leu Glu Gln Lys Lys Gln Asp Arg Gly Glu Leu Lys Leu Leu Leu Leu
      35             40             45
Gly Pro Gly Glu Ser Gly Lys Ser Thr Phe Ile Lys Gln Met Arg Ile
  50             55             60
Ile His Gly Ala Gly Tyr Ser Glu Glu Glu Arg Lys Gly Phe Arg Pro
  65             70             75             80
Leu Val Tyr Gln Asn Ile Phe Val Ser Met Arg Ala Met Ile Glu Ala
          85             90             95
Met Glu Arg Leu Gln Ile Pro Phe Ser Arg Pro Glu Ser Lys His His
      100             105             110
Ala Ser Leu Val Met Ser Gln Asp Pro Tyr Lys Val Thr Thr Phe Glu
      115             120             125
Lys Arg Tyr Ala Ala Ala Met Gln Trp Leu Trp Arg Asp Ala Gly Ile
      130             135             140
Arg Ala Cys Tyr Glu Arg Arg Arg Glu Phe His Leu Leu Asp Ser Ala
      145             150             155             160
Val Tyr Tyr Leu Ser His Leu Glu Arg Ile Thr Glu Glu Gly Tyr Val
      165             170             175
Pro Thr Ala Gln Asp Val Leu Arg Ser Arg Met Pro Thr Thr Gly Ile
      180             185             190
Asn Glu Tyr Cys Phe Ser Val Gln Lys Thr Asn Leu Arg Ile Val Asp
      195             200             205
Val Gly Gly Gln Lys Ser Glu Arg Lys Lys Trp Ile His Cys Phe Glu
      210             215             220
Asn Val Ile Ala Leu Ile Tyr Leu Ala Ser Leu Ser Glu Tyr Asp Gln
      225             230             235             240
Cys Leu Glu Glu Asn Asn Gln Glu Asn Arg Met Lys Glu Ser Leu Ala
      245             250             255
Leu Phe Gly Thr Ile Leu Glu Leu Pro Trp Phe Lys Ser Thr Ser Val
      260             265             270
Ile Leu Phe Leu Asn Lys Thr Asp Ile Leu Glu Glu Lys Ile Pro Thr
      275             280             285
Ser His Leu Ala Thr Tyr Phe Pro Ser Phe Gln Gly Pro Lys Gln Asp
      290             295             300
Ala Glu Ala Ala Lys Arg Phe Ile Leu Asp Met Tyr Thr Arg Met Tyr
      305             310             315             320
Thr Gly Cys Val Asp Gly Pro Glu Gly Ser Lys Lys Gly Ala Arg Ser
      325             330             335
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-continued

Arg	Arg	Leu	Phe	Ser	His	Tyr	Thr	Cys	Ala	Thr	Asp	Thr	Gln	Asn	Ile
			340					345					350		
Arg	Lys	Val	Phe	Lys	Asp	Val	Arg	Asp	Ser	Val	Leu	Ala	Arg	Tyr	Leu
		355					360				365				
Asp	Glu	Ile	Asn	Leu	Leu										
	370														

What is claimed is:

1. A process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism, wherein said process comprises the following steps:

- a) providing at least one cell which contains at least one GPCR-dependent signal transduction pathway and which produces one or more than one G-protein;
- b) providing at least one chemical compound to be studied;
- c) contacting the cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the cell of a) by means of a signal transduction pathway-dependent measurable signal.

2. The process as claimed in claim 1, wherein the cell provided according to a) produces at least two G-proteins.

3. The process as claimed in claim 1, wherein the cell provided according to a) produces at least two G-proteins selected from -6qi4myr, -6qs5myr, -6qi4, -6qs5, and Gα16.

4. The process as claimed in claim 2, wherein the cell provided according to a) produces at least two G-proteins selected from -6qi4myr, -6qs5myr, -6qi4, -6qs5, and Gα16.

5. The process as claimed in claim 1, wherein the cell provided according to a) produces at least one G-protein selected from -6qi4myr, -6qs5myr, -6qi4, and -6qs5.

6. The process as claimed in claim 2, wherein the cell provided according to a) produces at least one G-protein selected from -6qi4myr, -6qs5myr, -6qi4, and -6qs5.

7. The process as claimed in claim 1, wherein the cell provided according to a) produces at least one protein having an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8.

8. The process as claimed in claim 2, wherein the cell provided according to a) produces at least one protein having an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8.

9. The process as claimed in claim 3, wherein the cell provided according to a) produces at least one protein having an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8.

10. The process as claimed in claim 4, wherein the cell provided according to a) produces at least one protein having an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8.

11. The process as claimed in claim 5, wherein the cell provided according to a) produces at least one protein

having an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8.

12. The process as claimed in claim 6, wherein the cell provided according to a) produces at least one protein having an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:8.

13. The process as claimed in claim 1, wherein the cell provided according to a) is the cell of a vertebrate species, an insect species, a yeast species, or a *C. elegans*.

14. The process as claimed in claim 13, wherein the cell provided is a HeLa, 293, COS or CHO cell, or a cell of *Saccharomyces cerevisiae*.

15. The process as claimed in claim 1, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

16. The process as claimed in claim 2, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

17. The process as claimed in claim 3, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

18. The process as claimed in claim 4, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

19. The process as claimed in claim 5, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

20. The process as claimed in claim 6, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

21. The process as claimed in claim 7, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

22. The process as claimed in claim 8, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

23. The process as claimed in claim 9, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

24. The process as claimed in claim 10, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

25. The process as claimed in claim 11, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

26. The process as claimed in claim 12, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

27. The process as claimed in claim 13, wherein the intracellular Ca²⁺ concentration is the signal transduction pathway-dependent measurable signal.

106. A compound which modifies the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism and which is identified by the process as claimed in claim 50.

107. A compound which modifies the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism and which is identified by the process as claimed in claim 51.

108. A compound which modifies the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism and which is identified by the process as claimed in claim 52.

109. A compound which modifies the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism and which is identified by the process as claimed in claim 53.

110. A compound which modifies the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism and which is identified by the process as claimed in claim 54.

111. A compound which modifies the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism and which is identified by the process as claimed in claim 55.

112. A compound which modifies the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism and which is identified by the process as claimed in claim 56.

113. A polynucleotide sequence coding for a polypeptide having the property of a G-protein, wherein the polypeptide sequence is selected from:

- a) SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8;
- b) a sequence according to a) lacking one or more amino acids;
- c) a sequence according to a) having an additional one or more amino acids; and
- d) an allelic variant of a sequence according to a).

114. A polynucleotide comprising a polynucleotide sequence selected from:

- a) SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, the corresponding sequence complementary thereto; and
- b) a polynucleotide sequence hybridizing with a polynucleotide sequence according to a) under stringent conditions.

115. The polynucleotide as claimed in claim 113, wherein the polynucleotide is part of a recombinant vector construct.

116. The polynucleotide as claimed in claim 114, wherein the polynucleotide is part of a recombinant vector construct.

117. The polynucleotide as claimed in claim 115, wherein the recombinant vector construct is an expression vector usable in eukaryotes and/or prokaryotes.

118. The polynucleotide as claimed in claim 116, wherein the recombinant vector construct is an expression vector usable in eukaryotes and/or prokaryotes.

119. The polynucleotide as claimed in claim 117, wherein the expression vector contains a constitutive and/or inducible promoter.

120. The polynucleotide as claimed in claim 118, wherein the expression vector contains a constitutive and/or inducible promoter.

121. A host cell comprising a polynucleotide as claimed in claim 113.

122. A host cell comprising a polynucleotide as claimed in claim 114.

123. A host cell comprising a polynucleotide as claimed in claim 115.

124. A host cell comprising a polynucleotide as claimed in claim 116.

125. A host cell comprising a polynucleotide as claimed in claim 117.

126. A host cell comprising a polynucleotide as claimed in claim 118.

127. A host cell comprising a polynucleotide as claimed in claim 119.

128. A host cell comprising a polynucleotide as claimed in claim 120.

129. The host cell as claimed in claim 121, wherein the host cell is a human cell.

130. The host cell as claimed in claim 122, wherein the host cell is a human cell.

131. The host cell as claimed in claim 123, wherein the host cell is a human cell.

132. The host cell as claimed in claim 124, wherein the host cell is a human cell.

133. The host cell as claimed in claim 125, wherein the host cell is a human cell.

134. The host cell as claimed in claim 126, wherein the host cell is a human cell.

135. The host cell as claimed in claim 127, wherein the host cell is a human cell.

136. The host cell as claimed in claim 128, wherein the host cell is a human cell.

137. The host cell as claimed in claim 121, wherein the host cell is the cell of a vertebrate species, an insect species, a bacterial species, a yeast species, or *C. elegans*.

138. The host cell as claimed in claim 122, wherein the host cell is the cell of a vertebrate species, an insect species, a bacterial species, a yeast species, or *C. elegans*.

139. The host cell as claimed in claim 123, wherein the host cell is the cell of a vertebrate species, an insect species, a bacterial species, a yeast species, or *C. elegans*.

140. The host cell as claimed in claim 124, wherein the host cell is the cell of a vertebrate species, an insect species, a bacterial species, a yeast species, or *C. elegans*.

141. The host cell as claimed in claim 125, wherein the host cell is the cell of a vertebrate species, an insect species, a bacterial species, a yeast species, or *C. elegans*.

142. The host cell as claimed in claim 126, wherein the host cell is the cell of a vertebrate species, an insect species, a bacterial species, a yeast species, or *C. elegans*.

143. The host cell as claimed in claim 127, wherein the host cell is the cell of a vertebrate species, an insect species, a bacterial species, a yeast species, or *C. elegans*.

144. The host cell as claimed in claim 128, wherein the host cell is the cell of a vertebrate species, an insect species, a bacterial species, a yeast species, or *C. elegans*.

145. The host cell as claimed in claim 137, wherein the cell is a HeLa, 293, COS or CHO cell, an *Escherichia coli* cell or *Saccharomyces cerevisiae* cell.

146. The host cell as claimed in claim 138, wherein the cell is a HeLa, 293, COS or CHO cell, an *Escherichia coli* cell or *Saccharomyces cerevisiae* cell.

147. The host cell as claimed in claim 139, wherein the cell is a HeLa, 293, COS or CHO cell, an *Escherichia coli* cell or *Saccharomyces cerevisiae* cell.

148. The host cell as claimed in claim 140, wherein the cell is a HeLa, 293, COS or CHO cell, an *Escherichia coli* cell or *Saccharomyces cerevisiae* cell.

149. The host cell as claimed in claim 141, wherein the cell is a HeLa, 293, COS or CHO cell, an *Escherichia coli* cell or *Saccharomyces cerevisiae* cell.

150. The host cell as claimed in claim 142, wherein the cell is a HeLa, 293, COS or CHO cell, an *Escherichia coli* cell or *Saccharomyces cerevisiae* cell.

151. The host cell as claimed in claim 143, wherein the cell is a HeLa, 293, COS or CHO cell, an *Escherichia coli* cell or *Saccharomyces cerevisiae* cell.

153. The host cell as claimed in claim 144, wherein the cell is a HeLa, 293, COS or CHO cell, an *Escherichia coli* cell or *Saccharomyces cerevisiae* cell.

153. A method of producing a host cell, wherein a polynucleotide as claimed in claim 115 is introduced into a eukaryotic or prokaryotic cell.

154. A method of producing a host cell comprising a polynucleotide sequence selected from:

- a) SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, or the corresponding complementary sequence thereto; and
- b) a polynucleotide hybridizing with a polynucleotide sequence according to a) under stringent conditions,
- c) wherein a polynucleotide as claimed in claim 116 is introduced into a eukaryotic or prokaryotic cell.

155. A method of producing a host cell comprising a polynucleotide sequence selected from:

- a) SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, or the corresponding complementary sequence thereto; and
- b) a polynucleotide hybridizing with a polynucleotide sequence according to a) under stringent conditions,
- c) wherein a polynucleotide as claimed in claim 117 is introduced into a eukaryotic or prokaryotic cell.

156. A method of producing a host cell comprising a polynucleotide sequence selected from:

- a) SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, or the corresponding complementary sequence thereto; and
- b) a polynucleotide hybridizing with a polynucleotide sequence according to a) under stringent conditions,
- c) wherein a polynucleotide as claimed in claim 118 is introduced into a eukaryotic or prokaryotic cell.

157. A method of producing a host cell, comprising a polynucleotide sequence selected from:

- a) SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, or the corresponding complementary sequence thereto; and
- b) a polynucleotide hybridizing with a polynucleotide sequence according to a) under stringent conditions,

c) wherein a polynucleotide as claimed in claim 119 is introduced into a eukaryotic or prokaryotic cell.

158. A method of producing a host cell, comprising a polynucleotide sequence selected from:

- a) SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:7, or the corresponding complementary sequence thereto; and
- b) a polynucleotide hybridizing with a polynucleotide sequence according to a) under stringent conditions,
- c) wherein a polynucleotide as claimed in claim 120 is introduced into a eukaryotic or prokaryotic cell.

159. A method of using the host cell as claimed in claim 121 in a process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism comprising:

- a) providing said host cell;
- b) providing at least one chemical compound to be studied;
- c) contacting the host cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the host cell of a) by means of a signal transduction pathway-dependent measurable signal.

160. A method of using the host cell as claimed in claim 122 in a process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism comprising:

- a) providing said host cell;
- b) providing at least one chemical compound to be studied;
- c) contacting the host cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the host cell of a) by means of a signal transduction pathway-dependent measurable signal.

161. A method of using the host cell as claimed in claim 123 in a process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism comprising:

- a) providing said host cell;
- b) providing at least one chemical compound to be studied;
- c) contacting the host cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the host cell of a) by means of a signal transduction pathway-dependent measurable signal.

162. A method of using the host cell as claimed in claim 124 in a process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism comprising:

- a) providing said host cell;
- b) providing at least one chemical compound to be studied;
- c) contacting the host cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the host cell of a) by means of a signal transduction pathway-dependent measurable signal.

163. A method of using the host cell as claimed in claim 125 in a process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism comprising:

- a) providing said host cell;
- b) providing at least one chemical compound to be studied;
- c) contacting the host cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the host cell of a) by means of a signal transduction pathway-dependent measurable signal.

164. A method of using the host cell as claimed in claim 126 in a process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism comprising:

- a) providing said host cell;
- b) providing at least one chemical compound to be studied;
- c) contacting the host cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the host cell of a) by means of a signal transduction pathway-dependent measurable signal.

165. A method of using the host cell as claimed in claim 127 in a process for identifying a chemical compound

modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism comprising:

- a) providing said host cell;
- b) providing at least one chemical compound to be studied;
- c) contacting the host cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the host cell of a) by means of a signal transduction pathway-dependent measurable signal.

166. A method of using the host cell as claimed in claim 128 in a process for identifying a chemical compound modifying the action of at least one G-protein-coupled receptor (GPCR)-dependent signal transduction pathway of an organism comprising:

- a) providing said host cell;
- b) providing at least one chemical compound to be studied;
- c) contacting the host cell of a) with one or more of the chemical compounds of b);
- d) determining the quantitative or qualitative effect of the chemical compound or compounds of b) on the signal transduction pathway of the host cell of a) by means of a signal transduction pathway-dependent measurable signal.

167. A protein having an amino acid sequence selected from SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, and SEQ ID NO:10.

168. A process for preparing a protein as claimed in claim 167 comprising:

- a) providing a host cell;
- b) cultivating the host cell of a) in a growth medium suitable for the host cell and inducing expression of the protein;
- c) disrupting the cells and obtaining the cell material;
- d) removing the protein from other proteins of the disrupted cells of c).

169. A method of using the protein as claimed in claim 167 for producing antibodies.

170. A method of using the protein as claimed in claim 168 for producing antibodies.

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