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(54) **SCATTERED BRANCHED-CHAIN FATTY ACIDS AND BIOLOGICAL PRODUCTION THEREOF**

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(57) **ABSTRACT**

Methods and cells for producing scattered branched-chain fatty acids are provided. For example, the invention provides a method for producing branched-chain fatty acid comprising a methyl on one or more even number carbons. The method comprises culturing a cell comprising an exogenous or over-expressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of propionyl-CoA to methylmalonyl-CoA and/or an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of succinyl-CoA to methylmalonyl-CoA, under conditions allowing expression of the polynucleotide(s) and production of branched-chain fatty acid. The cell produces more branched-chain fatty acid comprising a methyl on one or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s). A cell that produces branched-chain fatty acid and the branched-chain fatty acid also are provided.

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C07C 53/126 (2006.01)
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C12N 1/21 (2006.01)

Figure 1: *mutA* nucleotide sequence (SEQ.ID NO: 1)

ATGGCAAGCACGGACCAGGGTACCAACCCGGCAGACACCGACGACCTGACGCCAACCACT
CTGAGTCTGGCGGGCGATTTTCCGAAAGCAACCGAAGAACAGTGGGAGCGCGAAGTGGAG
AAAGTTCTGAACCGTGGCCGTCCGCCGGAGAAACAGCTGACGTTTGCGGAATGTCTGAAA
CGCCTGACGGTCCACACAGTAGACGGCATTGACATTGTGCCAATGTATCGCCCCGAAAGAT
GCGCCGAAGAACTGGGTACCCAGGCGTTGCCCATTTACACGTGGGACCACGGTTCGT
AATGGCGATATGGACGCATGGGATGTCCGTGCACTGCATGAAGATCCGGATGAGAAATTT
ACGCGCAAAGCGATTCTGGAAGGGCTGGAACGCGGGGTACATCTCTGCTGCTGCGTGTG
GACCCGGACGCTATTGCTCCAGAACACCTGGATGAAGTGCTGTCTGACGTGCTGCTGGAG
ATGACCAAAGTAGAAGTCTTTAGTCGTTACGATCAAGGCGCCGCTGCCGAGGCGCTGGTA
TCTGTGTACGAGCGCAGCGATAAACCGGCTAAGGACCTGGCTCTGAATCTGGGTCTGGAC
CCGATCGCCTTCGCGGCACTGCAGGGGACGGAACCTGATCTGACTGTCCTGGGTGATTGG
GTGCGTCGCCTGGCAAAAATTTAGCCCAGATTCTCGTGCAGTGACCATCGATGCGAACATT
TATCATAATGCGGGTGCGGGCGATGTAGCAGAGCTGGCTTGGGCCCTGGCTACCGGTGCG
GAATATGTTTCGTGCACTGGTAGAACAAGGTTTTACGGCGACCGAGGCGTTCGATACGATT
AACTTTTCGTGTGACCGCAACCCATGATCAGTTTTCTGACAATCGCGCGTCTGCGCGCACTG
CGTGAGGCGTGGGCGCGCATTTGGGGAGGTATTTGGGGTTGATGAGGATAAACGTGGCGCC
CGTCAAAATGCGATCACGAGTTGGCGCGATGTGACACGCGAGGACCCGTATGTGAATATC
CTGCGCGGGAGCATCGCTACATTTTCTGCAAGCGTGGGTGGGGCCGAAAGTATTACAAC
CTGCCTTTTACCCAGGCACTGGGTCTGCCAGAAGACGATTTTTCCGCTGCGTATCGCTCGT
AATACCGGTATCGTTCTGGCCGAAGAAGTGAACATCGGTCGTGTTAATGATCCGGCCGGC
GGTAGCTATTACGTGGAAAGTCTGACTCGTAGTCTGGCCGATGCAGCGTGGAAAGAGTTC
CAAGAAGTGGAGAACTGGGCGGCATGAGCAAGGCGGTGATGACGGAACATGTAACGAAA
GTGCTGGATGCCTGCAATGCAGAACGCGGAAACGCCTGGCCAATCGCAAACAGCCGATT
ACCGCAGTAAGCGAATTTCTTATGATTGGGGCGCGCTCTATCGAAACGAAACCTTTTCCT
GCCGCACCGGCCCGTAAAGGTCTGGCATGGCATCGCGACAGTGAAGTATTCGAACAACCTG
ATGGATCGCAGCACCAGTGTGAGTGAACGTCCAAAGGTTTTCTGGCGTGCCTGGGCACA
CGTCGTGACTTCGGTGGTTCGTGAGGGTTTTAGCAGCCCAGTGTGGCATATCGCAGGCATT
GACACCCACAGGTTGAGGGTGGCACAACCGCAGAAATCGTAGAAGCATTCAAGAAATCT
GGGGCACAAGTTGCGGATCTGTGCTCTAGCGCCAAAGTGTACGCTCAGCAGGGTCTGGAG
GTGGCCAAAGCTCTGAAAGCAGCTGGCGCCAAAGCCCTGTATCTGAGCGGTGCCTTTAAG
GAGTTCGGCGATGATGCGGCTGAGGCGGAGAACTGATCGATGGTGCCTGTTTATGGGT
ATGGATGTGGTTGACACTCTGTCTAGTACGCTGGACATTCTGGGTGTAGCAAAGTAA

Figure 2: *mutB* nucleotide sequence (SEQ.ID NO: 2)

ATCACT
 ACACTGCCTCGTTTTGACTCTGTTGACCTGGGGAACGCGCCTGTTCCGGCGGATGCGGCC
 CGTCGCTTCGAGGAAC TGGCGGCAAAGCGGGCACGGGTGAGGCGTGGGAGACCGCGGAG
 CAGATTCGGTTGGTACACTGTTCAATGAAGACGTTTACAAAGATATGGACTGGCTGGAC
 ACGTACGCCGGGATTCCGCCATTCGTTTACGGCCCGTACGCGACGATGTACGCTTCCGT
 CCGTGGACAATTCGTCAATACGCCGGTTTAGCACGGCGAAAAGAAAGTAATGCTTCTAC
 CGCCGTAACCTGGCGGGGGCAAAGGGTCTGTCTGTGGCATTGACCTGCCGACCCAC
 CGCGTTACGATAGCGATAATCCGCGCGTGGCAGGGGACGTGGGTATGGCCGGGTGGCC
 ATCGACAGTATTTACGACATGCGTGAAC TTTGACGGCATTCCGCTGGACCAGATGAGC
 GTGAGTATGACGATGAATGGTGCCGCTCCTGCCGATTCTGGCACTGTATGTGGTTACAGCC
 GAAGAACAAGGTGTGAAGCCGGAACAGCTGGCTGGCACCATCCAGAACGATATTTCTGAAG
 GAGTTCATGGTGCCTAACACCTATATCTATCCGCGCAACCGTCTATGCGCATCATCAGT
 GAGATCTTTGCGTATACTAGTGCAAATATGCCGAAGTGGAACTCTATCAGTATTAGTGGC
 TATCACATGCAGGAGGCGGGCGCCACTGCCGATATCGAAATGGCCTATACGCTGGCCGAT
 GCGTGTGATTAATTCGTGCAGGCGAAAGCGTCCGTCTGAACGTGGACCAGTTCGCCCCG
 CGTCTGAGCTTCTTTGGGGTATTGGCATGAATTTCTTTATGGAAGTCGAAAAC TGGCGT
 GCCGCCCGCATGCTGTGGGCCAAACTGGTGCACCAATTCGGCCCGAAGAACCCGAAGAGC
 ATGAGCCTGCCACGCACAGTCAAACCAGCGGCTGGAGCCTGACCGCGCAGGACGTATAT
 AACAAAGTAGTTGCGACCTGTATTGAGGCGATGGCAGCCACCCAGGGTACACCCAGAGC
 CTGCATACAAACTCTCTGGACGAGGCCATCGCACTGCCGACAGACTTCAGCGCCCCGATC
 GCGCGTAATACTCAACTGTTTCTGCAACAGGAAAGCGGTACTACCCGTGTGATCGATCCG
 TGGTCTGGCAGTGCATATGTGAGGAAC TGAACCTGGGATCTGGCCCGTAAAGCGTGGGGT
 CATATCCAGGAAGTCGAGAAAAGTGGGTGATGGCTAAAGCAATTGAGAAAAGGCATCCCG
 AAAATGCGCATGAAGAAGCGGCAGCGCCACCCAAGCACGCATCGACAGCGGTCCGCCAG
 CCGCTGATTGGCGTGAACAAATATCGCCTGGAACATGAACCGCCACTGGATGTTCTGAAA
 GTAGATAAECTTACCGTCTTGGCGGAGCAGAAAGCGAAACTGGTTAAGCTGCGTGGCGAA
 CCGGATCCTGAGAAAAGTTAAAGCGGCGCTGGATAAAATCACTTGGGCCCGGGGCAACCCG
 GATGATAAAGACCAGACCGTAATCTGCTGAAGCTGTGTATTGACGCGGGTCTGTGCTATG
 GCGACTGTCCGGCAAATGAGCGATGCGCTGGAGAAAAGTATTTGGTCTTATAACCGCGCAA
 ATTCGTAATACTGTTGCTATAGCAAGGAAGTTAAGAATACTCCAGAAGTAGAAGAA
 GCGCGTGAAC TGGTAGAAGAA TTTGAGCAGGCTGAAGGTCCCGTCCACGCATTTCTGCTG
 GCCAAAATGGGCCAGGATGGCCATGATCGCGGTGAGAAAGTTATTGCTACTGCTTATGCT
 GATCTGGGCTTCGATGTTGATGTGGCCCC TCTGTTCCAGACTCCAGAGGAAACTGCCCGC
 CAGGCTGTTGAAGCTGACGTCCATGTCTGTTGGCGTTAGCTCTCTGGCTGGCAGCCATCTG
 ACCCTGGTCCCTGCTCTGCGCAAGGAAC TGGATAAGCTGGGCCGCCCTGATATTCTGATT
 ACTGTGGCGGCGTCAATCCTGAACAGGATTTGATGAACTGCGCAAGGATGGCGCTGTC
 GAAATTTATACCCTGGCACCGTCATTCCTGAATCTGCTATTTCTCTGGTCAAGAAGCTG
 CGCCCTAGCCTCGATGCCTAACTCGAG

Figure 3: MutA protein sequence (SEQ.ID NO: 3)

MARTYAGHSSAAASNALYRRNLAKGQTGLSVAFDLPTQTGYDPDHVLRGGEVGVKGVPI SHIGDMRALFDQ
 TPLGQMNTSMTNATAMWILAMYQVAAFDQATAADEFDPASVVKATGGTTQNDITKEYLSRGTYYFAPAPS
 LRLITDMVSYTVSDIPKWNPINICSYHLQEAGATPVQEIAYAMSTAI AVLDAVRDAGQVPQERFGEVVAR
 ISFFVNAGVRFVEEMCKMRAFVELWDELTRERYGVTDKQRRFRYGVQVNSLGLTEAQ PENNVQRIVLEM
 LAVTLKSGARARAVQLPAWNEALGLPRPWDQQWLSLRMQQVLAYESDLLEYEDLFEGSAVVEAKVAELVAG
 AKAEIARVAELGGAVAAVESGYMKSALVASHALRRQRIEAGEDIVGVNKFETTEPNPLTADLDTAIQSV
 DAGVEAAA AKAVREWRETRDADPVKRERAVAALARLKAAQT DENLMEASIECARAEVTTGEWAQALREV
 FGEFRAPTGVTVGLTGGAGAELS AVRERVAGLRDELGETLRVLVKGKPLDGH SNGAEQIAVRARDAG
 FEVIYQGIRLTPAQIVAAVSEDEVHLVGISILSGSHMELIPEVLDRLEAGAGDIPVIVGGIIPESDAAK
 LKATGVAEVFTPKDFGLNDIMGRFVDVIRDSRLTTAAPT V

Figure 4: MutB protein sequence (SEQ.ID NO: 4)

MTVAPKRPAAMTLAAHFPERTQEQRDLVACVVNKRPEQHLSCDDAVATMRSHLECGLDIEPLYMKSSDPVPLGVP
 GAMFFTRGRALRDADVDPVDRQVHDDPDAAATRQLVLADLENGVTSVWLHVHGADGLAPNDVAEALAEVRLLELAPVVVS
 SWDDQTAADALYAVLSCSRASSGNLGHDP LGAAARTGSAPDLAPLADAVRRLADHGEIRAITVDTRVHGDAGVTVD
 EVAFALATGVAYLRHLESEGVDVAEAFRNIIEFRVSATADQFLTAALRALRRAWARIGESVGVPESTRGATTHAVTSG
 RIFTRDDAWTNILRSTLATFGASLGGADAITVLPFDTVSGLPTPFRRRIARNTQIILAEESNVARVTDPAAGGSWYVET
 LTDDVAKAAWETFQIEI ESAGGMVAALANGLVAQRILAAVAERDAALATRSTPITGVSTFPLAGEKPLERVVRAELPVQ
 PNALAPHRDSAIFALRDRSAAYATEHGHAPRVSVPTLDPRAADRRIDAVNLLTVAGIDAVDGDTESSAAALTGTDKG
 YEGVAKDMDVVAFLSDLLDTTGAPA

Figure 5: Methylmalonyl-CoA epimerase nucleotide sequence (SEQ.ID NO: 5)

GAATTCAGGAGTCCCTCCATTATGCTGACCCGCATCGATCACATTGGCATCGCATGCTTTGAT
 CTGGATAAAAACCGTAGAGTTCTATCGCGCCACCTACGGCTTTTGGAGTGTTCATAGCGAA
 GTAACCGAAGAACAGGGCGTGCCTGAAGCCATGCTGAAAAATCAACGAAACTAGTGATGGT
 GGGGGCAGCTATCTGCAACTGCTGGAACCGACACCGCCGACTCTACAGTTCGTAAGTGG
 CTGGACAAGAATGGCGAAGGCGTTCATCACATTGCGTTCGGTACGGCTGATGIGGATCAA
 GACCGCGCAGATATTAAGATAAGGGTGTGCGTGTCTGTACGAGGAGCCACGCGGTGGT
 AGCATGGGTAGCCGTATTACGTTCCCTGCACCCATAAGACTGTGATGGTGTGCTGACTGAG
 CTGGTCACCTCTGCCCGGTGCAAAAGTCCGGAACATTAAGGCTT

Figure 6: Methylmalonyl-CoA epimerase protein sequence (SEQ.ID NO: 6)

MLTRIDHIGIACFDLTKTVEFYRATYGFVVFHSEVNEEQVREAMLKINETSDGGASYLQLLEPTRPDSTVAKWLDKN
 GEGVHHIAFGTADVDQDAADIKDKGVRVLYEEPRRSGSMGSRITFLHPKDCHGVLTELVTSA PVESPEH

Figure 7: DNA sequence for *accA1* (AF113603.1) (SEQ.ID NO: 7)

GTCCGCAAGGTGCTCATCGCCAATCGTGGCGAAATCGCTGTCGCGGTGGCCCGGGCTGCCGGGACGCGG
 GGATCCGCAAGCCTGGCCGCTACCGGGATCCGGACCGGGAACGCGTTGCAAGTCCGTCCCGCTGATGAGGC
 GTTCGCCCTGGGTGGTGACACCCCGGCAACGAGCTATCGGACATCGCCAAAGTCCCAAAGCCCGCGCG
 GATCGCGCCGCGACCCATCCACCCGGCTACGGATTCTCTCGGAGAACCCGAGTTCGCGCAGGCGG
 TCCGGAACCGCCGCTGATCGGATCGGCCCGCCCGCCGACGCCATCCGCGACCGTGGCGAAAAGTCCG
 CGCCCGCCACATCGCCACAGCGGGCCGGCCCGCCCTGGTCGCGCGCACCCCGACCCCGCTCCCGCGCG
 GACGAGTTCGTCGCTTCGCCAAGSAGCACGGCCGCCCCATCGCCATCAAGSCCGCTTCGCGCGCGCG
 GCGCGCCCTCAAGGTTCGCGCGCCTTCGAAGAGGTGCGGAGCTGTACGACTCCCGCGCCCGCGAGGC
 CGTGGCCSCTTCGCGCGCGGAGTGTCTGTCGAGCCCTACCTCGACAAACCCCGCCAGTGGAGACC
 CAGTGCCTGGCCGACACCCACGGCAACGTGGTGTGCTCCACCCCGGACTGCTCCCTCCAGCGCCGCC
 ACCAAAACCTCCCTCGAGGAGCCCGCCCGCCCTTCTCTCCGAGCCCGACAGCCAGCCCTACTCATC
 CTCGAAGCCATCTCGAAGGAGGCGGCTACGGCGCGCGCGGACCCGTGGASTCCCTCGTGGCATGGAC
 GGCAGCACTCTCTCGGAGTCAACACCGCCCTCCAGGTGAGGACCCCGGTCACCGAGGAAATCGCCG
 GCATCGACTGGTCCCGGAGATGTCCGCATCGCCGACCGGAGGAACTCGGTTACGACGACCCCGCCCT
 GCGCGCCACTCTTCGAGTTCGGATCGCGATCAACCGCGAGGACCCCGCGCGCGCTTCCTGCCC GCCCGCG
 ACCGTCAACCTCTTCGACGCGCCCAACCGGCCCGCGCGTCCGCTGGACCGCGCGCTCGAGTCCGGCTCCG
 TCAATCGCCCGCCCTGGGACCTCCCTTCGCAAACTGATCGTCAACCGCGCACCCCGCGCGGAGGAC
 CCAGCGCGCGCCCGCGCCCTGGACGAGTTCACCGTCGAGGGCATGGCCAGCGCCATCCCTTCCACCGC
 ACCGTCTCCCGGACCCCGCCCTTCGCCCCGAACTCACCGGCTCCACCGACCCCTTCCCTCCACACCC
 GGTGGATCGAGACGGAGTTCSTCAACGAGATCAAGCCCTCACCCACCGCCCGCACACCGAGACGGAGCA
 GGATTCGCGCGCGGAGACGGTCTGCTCGAGGTCGCGCGCAAGCGCCGGAAGTCTCCCTCCCTCCAGC
 CTGGGCACTGCTCCCTGGCCCGGACCGGCTGCGCGCGCGCGCGCCCGCCAAAGCGCGCGCGCAAGAA
 CCGCGCGCGCGCCCTCGCGCGCACCCCTCGGCTCCCGATGCAGGGCAGGATCGTCAAGATCGCGCTCGA
 GGAAGGCAGGAAGTCCAGGAGGCGACCTCATCGTCTGACTCGAGGCGATCAAGATGGAAACAGCCCTC
 AACGCCACAGGTCGCGGACCAATCAAGGGCTCACCGCGAGGTCGGCGCTCCCTCACCTCCGCGCGCG
 CCATCTCGGAGATCAAGGACTGA

Figure 8: DNA sequence for *pccB* (AF113605.1) (SEQ.ID NO: 8)

ATGTCCGAGCCGGAAGAGCAGCAGCCCGACATCCACACGACCCGCGGCAAGCTCGCGGATCTCAGGCGCC
GTATCGAGGAAGCGACGACGCGCGGTTCCGCACGCGCCGTCGAGAAGCAGCAGCCCAAGGGCAAGCTGAC
GGCTCGTGAACGCATCGACTCCTCCTCGACGAGGGTTCCTTCGTGAGCTGGACGAGTTCGCCCCGGCAC
CGCTCCACCAACTTCGGCCTCGACGCCAACCCGCCCTACGGCGACGGCGTTCGTCACCCGGCTACGGCACCG
TCGACGGCCCGCCCGTGGCCGTCTTCTCCCAGGACTTCACCGTCTTCGGCGGCGCGCTGGGCGAGGTCTA
CGGCCAGAAGATCGTCAAGGTGATGGACTTCGCCCTCAAGACCCGGCTGCCCGGTCGTTCGGCATCAACGAC
TCCGCGGCGCGCCGCATCCAGGAGGGCGTGGCCCTCCCTCGGCGCCTACGGCGAGATCTTCCGCCGCAACA
CCCAGCCCTCCGGCGTATCCCGCAGATCAGCCTGGTTCGTTCGGCCCGTGTGCGGGCGGCGCGGTGTACTC
CCCCGGATCACCAGACTTCACGGTGTGGTGGACCAGACCAGCCACATGTTTCAACCCGGTCCCGACGTC
ATCAAGACGGTACCAGGCGAGGACGTCGGCTTCGAGGAGCTGGGCGGCGCCCGCACCCACAACCTCCACCT
CGGGCGTGGCCACCACATGGCCCGGCGACGAGAAGGACGCGGTTCGAGTACGTCGAAGCAGCTCCTGTGTA
CCTGCCGTCCAACAACCTTCTCCGAGCCCCCGCCTTCCCGGAGGAGGCGGACCTCGCGGTACCGGACGAG
GACGCGGAGCTGGACACGATCGTCCGGACTCGGCGAACAGCCCTACGACATGCACTCCGTTCATCGAGC
ACGTCTGGACGACCGCGAGTTCCTTCGAGACGCAACCCCTTTCGCGCCGAACATCCTCACCGGCTTCGG
CCCGGTGGAGGGCCCGCCGGTCCGCATCGTCGCCAACAGCCCATGCAGTTCGCCGGCTGCCTGGACATC
ACGGCTCCGAGAAGGGCGCCCGCTTCGTGCGCACCTGCGACGCCCTCAACGTCGCCCGCTCACCTTCG
TGGACGTCCCCGGCTTCTGCGCCGGCTCGACCAGGAGCAGCAGCGCATCATCCGCCGCGGCGCCAAGCT
GATCTTCGCCCTACGCCGAGGCCACGGTGCCTCATCACGGTTCATACCCGCAAGGCCTTCGGCGGCGCC
TACGACGTTCAGGGCTCCAAGCACCTGGGCGCGGACCTCAACCTGGCCTGGCCCAACCCGAGATCGCCG
TCATGGGCGCCCAAGGCGCGGTCAACATCCTGCACCCGCGCACCATCGCCGACGCGCGGTGACGACGCGGA
GGCCACCCCGGCGCCCGCTGATCCAGGAGTACGAGGACGCCCTCCTCAACCCCTACACGCGGCGCGAACGC
GGCTACGTGACGCGGTGATCATGCCCTCCGACACTCGCCGCCACATCGTCCGCGGCTGCGCCAGCTGC
GCACCAAGCGGAGTCCCTGCCCGAAGAAGCACGGCAACATCCCCCTGTAA

Figure 9: Protein sequence for AccA1 (SEQ.ID NO: 9)

MRKVLIANRGEI AVRVARACRDAGIASVAVYADPDRDALHVRAADEAFALGGDTPATSYL
DIAKVLKAARESGADAIHPGYGFLSENAEFAQAVLDAGLIWIGPPPHAIRDRGEKVAARH
IAQRAGAPLVAGTDPDVS GADEVVAVFAKEHGLPIA IKAAFGGGGRGLKVARTLEEVPELY
DSAVREAVA AFGRGECFVERYLDKPRHVETQCLADTHGNVVVVSTRDCSLQRRHQKLVEE
APAPFLSEAQTEQLYSSSKAILKEAGYGGAGTVEFLVGM DGTIFFLEVNTRLQVEHPVTE
EVAGIDL VREMFR IADGEE LGYDDPALRGHSFEFRINGEDPGRGFLPAPGTVTLFDAPTG
PGVRLDAGVESGSVIGPAWDSLLAKLIVTGRTRAEALQRAARALDEFTVEGMATAIPFHR
TVVRDPAFAPELTGSTDPFTVHTRWIETEFVNEIKPFTTPADTETDEESGRETVVVEVGG
KRLEVSLPSSLGMSLARTGLAAGARPKRRAAKKSGPAASGDTLASPMQGTIVKIAVEEGQ
EVQEGDLIVVLEAMKMEQPLNAHRSGTIKGLTAEVGASLTSGAAICEIKD*

Figure 10: Protein sequence for PccB (SEQ.ID NO: 10)

MSEPEEQQPDIIHTTAGKLADLRRRIEEATHAGSARAVEKQHAKGKLTARERIDLLLDEGS
FVELDEFARHRSTNFGLDANRPYGDGVVTGYGTV DGRPVAVFSQDFTVFGGALGEVYGQK
IVKVMDFALKTGCPVVGINDSGGARIQEGVASLGAYGEIFRRNTHASGVIPQISLVVGP C
AGGAVYSPAITDFTVMVDQTS HMFITGPDVIKTVTGEDVGFEEELGGARTHNSTSGVAHMM
AGDEKDAVEYVKQLLSYLP SNNLSEPPAFPEEADLAVTDEDAELDTIVPDSANQPYDMHS
VIEHVLDDAEFFETQPLFAPN ILTGFRVEGRPVGIVANQPMQFAGCLDITASEKAARFV
RTCDAFNVPLTFVDVPGFLPGVDQEH DGIIRRGAKLI FAYAEATVPLITVITRKAFGGA
YDVMGSKHLGADLNLAWPTAQI AVMGAQGAVNILHRRTIADAGDDAEATRARLIQEYEDA
LLNPYTAERGYVDAVIMPSDTRRHIVRGLRQLRTKRESLPPKKHGNIP L*

Figure 11: Element 1: PlacO1 sequence + phage T7 gene10 ribosome binding site (SEQ.ID NO: 11)

aattgtgagcggataacaattgacattgtgagcggataacaagatactgagcacatcagcaggacgcactgaccgaattcaataat
tttgtttaactttaagaaggagatatacat

Figure 12: Element 2: Optimized accA1 gene sequence (SEQ.ID NO: 12)

atgcgcaaaagtgcctgatctggcaaccgtgggtaaatcgccgtctcgtgtggcaecgcgcgtcgtgatgcaggtattgcaagtgttggcgggtatgccc
atccggatcgcgatgcgctgcatgttcgtgcggccgatgaagcctttgcaactggcggcgggtgataccccggcaacagcgtatctggatattgcaaaagt
gctgaaagcagcgcgcgaaagcgggtgcggatgccatccatccgggctacgggtttctctgtctgaaaatgcagaaatttgcacaggcgggtctcgatgca
ggctctgatttgatcggtcocgcocgcgatgcaattcgtgatctggggcgaataagtgccgcacgccacatcgccagcgtgcaggcgcgcgcgtgg
ttgcgggcaaccgggaccgggtttctggtcagatgaagcggttgccgtttgcccagaacatggcctgcgattgogatcaaaagcagcattcggcgg
lggcggllcggcglcgaaglggccccglaaccgggaagagllccggaaclglalgalagcgcagllcggcgaagcggllggcagcglggcggllggll
qaatgcttctgtgaaacgctacctgqataaaaccgctcatgttgaacccaagtgtctgcccggatagcgaacggcaacgctgcttctggttagcaccocg
attgctctctgcaacgtcgccaccagaaaacgggtggaagaagcaccggcgcgcgtttctgagcgaagcccagaccgaacagcgtgtagctctagtaa
agcgtattctgaaagaagcgggttacgtggcgcgcggtaacgggttgaattctcgttggcgtatggatggcaccatlagctttctggaagttaacaccggt
ctgcaagttagacatccgggtgaccgaaqaaagttgcggcattgcatctggtgocgcaaatgttctgtatccagatggcgaagaaactgggttacgatg
atccggcgtcgcgggtcacagctttgaaattctgattaatggcgaagatccggcgcgcgttttctgcggcgcgggcaccgctgacgctgttoga
tgcaccgaccggtccggcgttctgtctggaatgcgggtgtgaaagtggtagcgttattggcgcggcatgggatagcctgctggcgaactgatcgtt
accggctgtaacgcgcgcogaagcgtgcaacgtgcagcaacgtccctggatgaattaccgtggaaaggcatggcgaacggcattcgtttctatcgca
ccgtggttctgtatccggcattcgcgcggcaactgacccgctctaccgatccgttccaccgtgcacacgcctggatcgaaccgaatttctgtaacga
aatcaaacggttaccacgcggcgggataccgaaacggaaagaagaagtggctgcgaaacgggtggttctggaagtggcgggttaaagctctggaagt
tctctgcgagcagcctgggtatgctctctgcgcgtaccggctctgcgcggcggcgcgcctccgaaacgtcgcgcagcgaaaaaatctggtccggcgc
caagcgggtgataccctggcagctccgatgcagggcacgaattgtgaaatcgcagtggaagaaggtcaggaagtgcaggaaggcagctctgattgtgt
gctggaagcagatgaaaatggaaacagcgtgaaatgcccacgtagcggcaaccatcaaaggcctgacggcogaagtggtgcatctctgaccagtggc
cggccatttgcgaaatcaaaagattaa

Figure 13: Element3: Spacer sequence (Restriction sites and phage T7 gene10 ribosome binding site) (SEQ.ID NO. 13)

agatctgcccgcgatctagaaaataattttgtttaactttaagaaggagatataatc

Figure 14: Element4: Optimized pccB (SEQ.ID NO: 14)

atgagtgaaaccggaagaacagcagcggatattcataaccacggcaggcaaacctggcggatccgcgtcgcggtatcgaagaagcaaccatgcaagta
ggcgcagctgcagtggaaaaaacagcaocgcgaaggtaaacagcggcccgccgaacgtatcgatctgctgctggatgaagcagttttgtgaaactgga
tgaaatttgcaacgcaccgtgacccaactttggcttggaatgcgaatcgcccgatggcgatgggtggtttaccggttacggtaacgggtggatggctgt
ccgggtggcagtttttagccaggtttttaccggttgcggcgggtgcaatggcgaagtttaacggctcagaaaaatcgtgaaagttaggattccgcgctga
aaacgggctgcgggtgggttggatattaaccatacggcgggtgcccgcacccggaaggtgttgcctctctggggcgcgtatggcgaatctttcgcg
taatacccatgagcagtgggctgattccgcagatcagcctgggtgggtggctcgtgcggggcgggtgcccgtttactctccggcattaccgattttacg
gtgatgggtgatcagaccagtcacatgttcaatacgggcccggatgtgatcaaaacggttacgggcgaagatgtgggtttgaaagaacggcgggtg
cagctaccacaacagcagcagctctggcgttgcgcatacacaaggcgggtgatgaaagatgacgtggaaatagtttaaacagctgctgagttacctgca
gagcaacaatctgctgaaacgcggcgttcccgaagaagcagacotggcgggtgacccgatgaagatgcccgaactggatcagatcgttccggattct
gcaaatcagccgtacgatatgcaacagtggtattgaaacagttctggatgatgcggaattttcgaaaacccagccgctgtttgcccgaacattctga
cgggtttcggctggtggaaggtcgtccggcgggtatcgctgcaaatcagccgatgcagtttggcgggttgcctggatataccgcctccgaaaaagc
ggccgcctttgtgctgtaacctgtagtgcgttcaacgtgcccgggtctgacgtttgtggatgttccgggcttccgcccgggtgttatcaggaacatgat
ggcattatccgcggtgggtgcgaactgattttgctgatgcccgaagcaaccgtgcccgtgattaccggtatcaccgcgcaaaagcattccggcgtgct
acgatgtgatggcagcaaacatctgggtgcccgatctgaaacctggcattggcggacccgcagatcgcagtgatgggycgcaggggtgcccgttaatat
ctgcaccgcgtaaccatccagatgcagctgatgatgcagagcgaacggcgcagcctgctgattcaggaatatgaaatgcgctgctgaaaccgctat
accgcagcggaaacgtgggtacgtggatgcccgttattatgcccagcagatacccccgtcatabctgctgctgctgctgctgctgctgctgctgctgctg
aatctctgcccgcgaaaaaacaggtaataatccgctgtaa

Figure 15: Entire synthetic sequence for propionyl-CoA carboxylase gene expression. (SEQ.ID NO: 15)

aattgtgagcggataaacaattgacattgtgagcggataaacaagatactgagcacatcagcaggacgcactgacccaattcaataaatttggttfaact
ttaaagaaggagataacacataatgagcaaaagtgtgatgtgcaaacgggtgaaatogcoogtctcgtgtggcaogcogcgtgtcgtgatgcaggtattgca
agtgttgcgggtgatgcogcatccggatcogcogtgcgctgcatgttcgtgcccgcgatgaagcctttgcaactggggcgtgatccccggcaacgagct
atctggalattgcaaaaagtgctgaaagcagcgcgcgaaagcggctgcggatgcaatccatccggggctacggllttctgctgcaaaaagcagaalltg
acaagcgggttctgqatgcaggtotgatttctgcatcggctccgcccgcogcatgcaattcgtgatctgggcgataaaagtggccgcacgccaatccccaq
cgtgcagggcgcgcogcgtggttgcggggcaccocggaccocgggtttctggtgcagatgaagtggttgcggtttgcocaaagaaacatgcccctgcogattgcga
tcaaaagcagcatccggcggltggcggltcgcggctgaaagggcccgtaacctggaagaagtccggaaactgatagatagccagttccgcgaagcgggt
ggcagcgtttggccgctggtgaatgcttcgtggaacgctacctggataaacccgctcatgttgaaccccagctgtctggcggatagcgaacggcaacgtg
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gaagaactgggttacgatgatccggcctcgcggctcacagctttgaaattctgatataatggcgaagatccggcgcgtggtttctcgcggcgcogcgg
gcaacgctgacgctgttcgatgcaaccgctcggggcgtcgtctggaatgcggctggaagaagtggtagcgttatggcccggcatgggatagcct
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gccattccgtttcatcgcacccgtggttcgtgatccggcaatcgcgcggcaactgcacggctctaccgatccgttccaccgcaacacgcgctggatccg
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cggtaaacgtctggaagtctctcgcgcagcagcctgggtatagcttggcgcgtaccggtctggcggcggcgcogcctccgaaacgtcgcgcagc
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aaggcagctcgtattggtgctggaagcgaagaaatggaacagcgcgctgaaatgcccatcgttagcggcaccatcaaaagcctgacggccgaagtggg
tgcactctgaccagtgccgcggccatttgcgaaatcaaaagataaagatctgcggcgcogctctagaaataaatttggtttaactttaagaaggagat
alalcaatgagtgaaaccggaagaacagcagccggalalalcaatccacggcaggaacatggcggalclggcglcgcglaagaaagcaacccaatg
caggtagcgcacgtgcagtggaaaaacagcaacgcgaaaggttaaacctgacggccgcgaaacgtatcgatctgctgctggatgaaggcagcttctgtta
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tcgcogtaataaccatagcagtggtgctgatccgcagatcagcctgggtgggttggctcgtgtagcggcgggtgcogcttaccctccggccattaccgat
tttacggtgaggttgaacagaccgtcacatgttcaatcagcggccggatgtagtaaaaacogttagcggcgaagatgggttttgaagaactgg
gcccgtgacgttcccccaacagcagctctcgcgttcgcgcatcaatggccggtagtaaaaaagatgcogtggaaatagttaaaacagctgctcagttta
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gatctcgaactcagccgtagcagatgcaacagtgatgaaacagcttctggatgagcggaaatttccgaaaccagccogctgttggccogaaca
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aatgatgcatattaccgocgtggtgcgaaactgattttgcgtatgcccgaagcaacocgtgocgctgattaccggtatcagcgcgcaagcattcggcg
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ccgtataccgcagcggaaacgtggttaccgtggatcgggttattatgcgcagcagataaccgcogctcatatcgtcgtggctcggctcagcgcgtaaga
aacgtgaatctctgcgcgcaaaaaaacacgtaataattccogctgtaa

Figure 16: Forward primer for PrpE (SEQ.ID NO: 16)

AAACTGCAGAGGAGGACAGCTATGTCTTTTAGCGAATTTTATCAG

Figure 17: Reverse primer for PrpE (SEQ.ID NO: 17)

AAAGGATCCCTATTCTTCGATCGCCTGGCGAATTTG

Figure 18: MMAT domain sequence from Mycobacterium bovis BCG (SEQ.ID NO: 18)

LVECLREVADGDALYDAAVGHGDRGPFVWVFSGQSSQWAMGTQLLASEPVEAATIAKLEP
VIAAESGFSVTEAITAQQTVTGIDKVPQAVFAVQVALAATMEQTYGVRPGAVVGHSMGES
AAAUVAGALSLEDAARVICRRSKLMTRITAGAGAMGVELPAKQVNSELMARGIDDVVSV
VASPQSTVIGGTSITRDLIARWEQRDVMAREVAVDVASHSPQVDPILDDLAAALADIAPI
MTPKVPYYSATLFDPRQPVCDGAYWVDNLRNTVQFAAAVQAAMEDGYRVFAELSPHLL
THAVEQTGRSLDKSVAAALAGMRREQLPHGRGLLELHRAGAALDYSAIYPAGRLVDAP
LPWATHARLFIDDDGQZQRAQGA

Figure 19: Protein sequence for the *Mycobacterium bovis* BCG MAS (YP_979046) (SEQ.ID NO: 19)

ME SRVT PVAVIGMGCRLEGGINS PDKLWESLLRGDDLVEIIPPDRWDADDYYDPEPGVPGRSVSRWGGFL
DDVAGFDAAEFFGISEREATSIDPQQRILLETSWEAIEHAGLDPASLAGSSTAVFTGLTHEDYLVLTITTAG
GLASPVVVTGLNNSVASGRHAHTLGLHGPA MPTFTACSSGLMAVHLACRS LHDGEADLALAGCCAVLLEP
HACVAASAQGMSSSTGRCHSFDADADG FVRSEGCAMVLLKRLPDALRDGNRIFAVVRGTATNQDGR TETL
TMPSEDAQVAVYRAALAAAGVQPETVGVVVAHGTGTPIGDPIEYRSLARVYGAGTPCALGSAKSNMGHST
ASAGTVGLIKAILSLRHGVVPLLHFNRLPDELS DVE TGLFVPOAVTFWPNGNDHTPKRVAVSSFGMSGT
NVHAI VEEAPAEASAPESSPGDAEVGPRLEFMLSSTSSDALRQTARQLATWVEEHQDCVAASDLAYTLARG
RAHRPVRTAVVAANLPELVEGLREVADGDALYDAAVGHGDRGPVWVFSGQGSQWAAAMGTQLLASEPVFAA
TIAKLEPVIAAESGFSVIEAITAQQTVTGIDKVQPAVFAVQVALAATMEQTYGVRPGAVVGHSMGESAAA
VVAGALSLEDAARVICRRSKLMTRIAGAGAMGSVELPAKQVNSELMARGIDDVVVSVVASPQSTVIGGTS
DTVRDLIARWEQRDVMAREVAVDVASHIS PQVDPILDDLAAAALADIAPMTPKVPYYSATLFPDFREQPVCDG
AYWVDNLRNTVQFAAAVQAAMEDGYRVFAELSPHPLLTHAVEQTGRSLDMSVAALAGMRREQPLPHGLRG
LLTELHRAGAALDYSALYPAGRLVDAPLP AWTHARLFI DDDGQEQRAQGACTITVHPLLGSHVRLTEEPE
RHVWQGDVGTSVLSWLSDHQVHNVAALPGAAYCEMALAAA AAEVFG EAAEVRDITFEQMLLLEDEQTFIDAV
ASIDAPGVVNFVETNTRDGETTRHATAALRAAEDDCPPPGYDITALLQAHPHAVNGTAMRESFAERGVTL
GAAFFGLTTAHTAEGAATVLAEVALPASIRFQQGAYRIHPALLDACFQSVGAGVQAGTATGGLLLPLGV
RSLRAYGPTNRNARYCYTRLTKAFNDGTRGGEADLDVLDDEHGTVLLAVRGLRMGTGTSE RDERDRLV SERL
LTLGWQQRALPEVGDGEAGSWLLIDTSNAVDTPEMLASTLTDALKSHGPQGT ECASLSWSVQDTPPNDQA
GLEKLSQQLRGRDGVVIVYGFVRVGDPEHSLLAGREQVRHLVRI TRFLAEFEGELPRLFVVTRQAQIVKP
HDSGERANLEQAGLRGLLRVISEHPMLRITLLIDVDEHTDVERVAQQILSGSSEDETAWRNCDWYVARLT
PSPGLGHEERRTAVLDPDHDGMRVQVRRGDLQTELFVVASDRVPPGPGQIEVAVSMS SINFDVLI AFGRF
PIIDDRPQLGMDFVGVVAVGEGVTGHQVGDVRVGGFSEGGCWRTFLTC DANLAVTLPPGLTDEQAITAA
TAHATAWYGLNDLAQIKAGDKVLLHSA TGGVCCAAISLARAKGAEI FATAGNPAKRAMLRLDMCV EHVYDS
RSVEFAEQIRRD TDGYVDIVLNSLTGAAQRAGLELLAFGGREVEICKADVYGNTRLGLFPFRRLTFYY
LDLALMSVTPQDRVRELLATVFKLTADGVLTAPQCTHYPLAEAAADIRAMSNAEHTGKLVLDVPRSGRRS
VAVTPEQAPLYRRDGSYIITGGLGGLG LFFASKLAAAAGCGRIVLTARSQPNPKARQTI EGLRAAGADIV
ECGNIAEPDTRADRLVSAATATGLPLRGVLESAAVVEDATLTNI TDELDLDRDWSPKVFGS WNLHRA TLGQP
LDWFC LFPSSGAALLGSPGQ GAYAAANSWVDVFAHWRR AQGLPVS AIAWGA WGEVGRATFLAEGGEIMITP
EEGAYAFETLVRH DRAYSYIPI LGAPWLADLVRRS PWGEMFASTGQRSRGP SKFRMELLSLPQDEWAGR
LRRLLVEQASVILRRITIDADRSFIEYGLDSLGMLEMRTHVETETGIRLTPKV IATNNTARALAQYLADTL
AEEQAAA PAAS

Figure 20: Codon-optimized MMAT domain DNA sequence from *Mycobacterium bovis* BCG (SEQ.ID NO: 20)

CTGTTGGAAGGCTCGCTGAAETTGCCGATGGTGTATGCAC TGTATGATGCAGCAGTGGGTCATGGCGAT
CGTGGTCCGGTTGGGTTGTTAGCGGCCAGGGTCTCAGTGGGCAGCGATGGGCACCCAGCTGCTGGCAAGCGAA
CCGGTTTTTGCCGCAACGATTCGCAAACTGGAACCGGTGATCGCGGCCGAAAGTGGCTTCAGCGTTACCGAAGCA
ATTACGGCGCAGCAGACCGTGACGGGTATCGATAAAGTGCAGCCGGCCGTTTTCCGAGTTCAGGTGGCGCTGGCA
CGCAGCATGGAACAGACGTACCGCGTTCTGTCGGGTGCGATGGTTGTCACAGTATGGGTGAAAGCGCCGACGG
GTGGTTGCAGCCGCCCTGAGTCTGGAAGATGCCGCACGTGTGATTTGCCCTCGCAGCAAAC TGTATGACCCGATC
GCAGGTGCAGGTGGGATGGG CAGCGTGGAACTGCCCGCAAACAGGTAACTCTGAAC TGTATGGCCGCGGTATT
GATGATGIGGTTGTCTGTCTGTGGCGTCTCCGCAGAGTACCGTGTATGGCCGCAC CAGTGATACCGTTCGTGAT
CTGATCCGCGCTTGGGAACAGCCGATGTGATGGCCCGCGAAAGTTGCCGPGGATGTTGCAAGCCATCTCCGCAG
GTTGATCCGATTCGTGGATGATCTGGCGCGGCACTGGCAGATATTGCACCGATGACCCGAAAGTGCCTATTAC
AGCCGACGCTGTTGATCCGCGTGAACAGCCGGTGTGTGATGGCCCTATTGGGTGATAACCTGGCGCAATACC
GTGCAAGTITGGCGCGGCGAGTTACGGCGGCGATGGAAGATGGTTACCGTGTGTTGCGCGAACTGCTCCGCATCCG
CTGCTGACCCACGCACTGGAACAGACGGGTCCCTCTCTGGATATGAGTGTTCAGCAGCACGGCCGATATGCCTCGC
GAAACAGCTGCGCGCATGGCTGCGTGGTCTGCTGACCGAAGTGCACCGTGCAGGTGCAGCACTGGATTATAGC
GCAC TGTACCCGCGCAGGTGCTGTGGTGGATGACCCGCTGCCGGCATGGACGCACGCACGCTGTTCATCGATGAT
GATGCGCCAGGAACAGCGCGCACAGGGTCCG

Figure 21: Alignment of a codon-optimized MMAT domain from *Mycobacterium bovis* BCG with the original sequence:

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Optimized 1 CTSGTGGAAGGCCTGCGTGAAGTTGCCGATGGTGATGCACGTATGATGCAGCAGTGGGT
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Original 1 CTCGTGAGGGTTTGCGCAGGTTGGCCGACGGTGACGCCCTCTATGACGCCGCGGTGGGA

Optimized 61 CATGCCGATCGTGGTCCSGTTTGGGIGTATTAGCGGCCAGGGTCTCAGTGGGCAGCGATG
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Original 61 CACGGTGTATCGAGGACCGGTCTGGGTCCTTCGCCGCAAGGGTCCGACTGGCCGCGGATG

Optimized 121 GGCACCCAGCTGCTGGCAAGCGAACCGGTTTTTGCCGCAACGATTGCAAACTGGAACCG
|||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Original 121 GGCACGCAATGCTCGCCAGCGAACAGTGTTCGGCGCCACCATCGCCAAGCTGGAGCCG

Optimized 181 GTGATCGCGGCCGAAAAGTGGCTTCAGCGTTACCGAAGCAATTACGGCGCAGCAGACCGTG
|||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Original 181 GTGATCGCCGAGAAATCGGGATTCGCGGTGACCGAGGCGATAACGGCGCAGCAGACCGTG

Optimized 241 ACGGGTATCGATAAAGTGCAGCCGGCCGTTTTTCGCAGTTCAGGTGGCGCTGGCAGCGACG
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Original 241 ACCGGAATCGACAAGTGCAGCCGGCAGTGTTCGCCGTTCAGGTTCGGCTGGCCGCCACC

Optimized 301 ATGGAACAGACGTACGGCGTTCGTCCGGGTGCAGTGGTGGTCCACAGTATGGGTGAAAAGC
|||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Original 301 ATGGAGCAAACCTACGGAGTGCGGCCGGGCGCGGTTCGTGGACACTCGATGGGTGAGTCCG

Optimized 361 GCCGCAGCGGTGGTTGCAGCGCCCTGAGTCTGGAAGATGCCGCACGTGTGATTTGCCGCT
|||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Original 361 GCCCGCGCGTTCGTCCGGGGGCACTGTGCTCGAGGACCGCGCGCGCTCATTTGCCCGC

Optimized 421 CGCAGCAAAGTATGACCCGATCGCAGGTGCAGGTGCGATGGGCAGCGTGGAACTGCCG
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Original 421 CGCTCGAAGTATGATGACCCGCATAGCCGGTGTGGTGCATGGGCTCGGTGGAAATGCCC

Optimized 481 GCAAAACAGGTTAACTCTGAAGTATGAGCGCGCGGTTATTGATGATGTGGTTGTGTCTGT
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Original 481 GCCAAGCAAGTGAATTCGGAGCTGATGGCACGCCGAATCGACAGATGTTGTCTGTCTCGGTG

Optimized 541 GTGGCTCTCCGACAGTACCGTGTATTGGCGGCACCAGTATACGGTTCGTGATCTGATC
|||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Original 541 GTGGCTCCCGCAATCCACGGTGTATCGGGGTACGAGCGACACCGTTCGTGACCTCATC

Optimized 601 GCGCCTTGGGAACAGCGGATGTGATGGCGCGCAAGTTCGGTGGATGTTGCAAGCCAT
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Original 601 GCCCGTTGGGAGCAGCGGACGTAAGGCGCGGAGGTTGGCCGTGACGTTGGCTTCGCAC

Optimized 661 TCTCCGAGGTTGATCCGATTCGGATGATCTGGCGCGGCACTGGCAGATATGACCCG
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Original 661 TCGCTCAAGTCGATCCGATACTCGACGATTTGGCCGCGGCGCTGGCGGACATTGCTCCG

Optimized 721 ATGACCCCGAAAGTCCCGTATTACAGCCGACCCCTGTTTGATCCCGTCAACAGCCGGTG
|||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Original 721 ATGACGCCAAGGTGCCGTACTACTCGGCGACCTGTTGACCCCGCGGACAGCCGGTG

Optimized 781 TGTGATGCGCCCTATTGGGTTGATAACCTGCGCAATACCGTGCAGTTTGGCGCGGCAGTT
||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| ||| |||
Original 781 TCGGATGCGCCTACTGGGTTGACAATCTGCGCAACACGGTGCAGTTCGCCGCGGCGGTG

Optimized 841 CAGCGCGGATGGAAGATGGTTACCGTGTGTTCCGCGAACTGTCTCCGATCCGCTGCTG
|||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Original 841 CAGGCTGCGATGGAGGACGGCTACCGGCTCTTCGCGAGCTGTGCCCCACCCGCTGCTT

Optimized 901 ACCACGCGCTGGAACAGACGGGTCCGCTCTCTCGATATGAGTGTTCAGCACTGCCCGTG
|||| | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | | |
Original 901 ACCACGCCGTCGAACAGACGGCCSAAGCCTCGACATGTCGGTCCGCCCCCTGGCCGCG

```

Optimized 961 ATGCGTCGCGAACAGCCGCTGCCGCATGGCCTGCGTGGTCTGCTGACCGAACTGCACCGT
||||| || || ||||| ||||||||| ||||| || ||||| || |||||
Original 961 ATGCGGCGAGAGCAGCCTCTGCCGCATGGTCTGCGCGGCTTGCTGACGGAGCTGCACCGC

Optimized 1021 GCAGGTGCAGCACTGGATTATAGCGCACTGTACCCGGCAGGTCGTCTGGTGGATGCACCG
|| || || || |||| ||| || ||||| || || || || ||||| ||||| |||
Original 1021 GCGGGCGCCGCTTTGGACTATTCCGCGCTGTATCCCGCTGGGCGGCTGGTGGATGCGCCG

Optimized 1081 CTGCCGGCATGGACGCACGCACGTCTGTTTCATCGATGATGATGGCCAGGAACAGCGCGCA
||||||| ||||| ||||| || || ||||| ||||| || ||||| |||
Original 1081 CTGCCGGCGTGGACCCACGCCCGCCTATTTCATCGACGATGATGGGCAAGAACAGCGGGCA

Optimized 1141 CAGGGTGCG
|| |||||
Original 1141 CAAGGTGCC

Figure 22: Protein sequence of *Salmonella enterica* propionyl CoA synthase PrpE (AAC44817) (SEQ.ID NO. 21)

MSFSEFYQRSINEPEAFWAEQARRIDWRQPFPTQTLDHSPFPFARWFCGGTTNLCHNAVDRWRDKQPEALA
LIAVSSSETDEERTFTFSQLHDEVNIVAAMLLSLGVQRGDRVLVYMPMIAEAQITLLACARIGAIHSVVFG
GFASHSVAARIDARPALIVSADAGARGGKILPYKKLLDDAIAQHQPKHVLLVDRGLAKMAWVDGRDL
DFATLRQQHLGASVPVAVLESNETSCLYTSGTGKPKGVQRDVGGYAVVALATSMDTIFGGKAGGVFFCA
SDIGWVVGHSYIVYAPLLAGMAFIVYEGLPTYPDGCVVWKIVEKYQVNRMFSAPTAIRVLLKKFPTAQIRN
HDLSSLEALYLACEPLDEPTASWVTEFLCVPVIDNYWQTESOWPIMALARALDDRPSRLGSPCVPMYCYN
VQLLEVTGEPGGINCKMLVLEGPLPPGCIQTWGGDDARFVKTYWSLFRQVYATFDWGIIRDAEGYFFI
LGRITDVINIAGHRLGTREIEBESSYPNVAEVAVVGIKDALKGQVAVAFVIPKQSDTLADREAARDEEN
AIMALVDNQIGHFRPAHVWFVSQLPKTRSGKMLRRTIQAIICEGRDPGDLTTIDDPASLQQIRQAIEE

Figure 23: DNA sequence of *Salmonella enterica* propionyl CoA synthase PrpE (SEQ.ID NO. 22)

ATGTCCTTTTAGCGAATTTATCAGCGTTCATTAAACGAACCGGAGGCGTCTGGGCGGAG
CAGGCCCGCGTATCGACTGGCGCAGACCGCTTACGCAGACCGTGGATCATAGCCGTCCA
CCGTTTGCCCGCTGGTTTGGCGCGGCACCACCTAAGTATGTCATAACCGCGTCGACCGC
TGGCGGGATAAACAGCCGAGCGCGCTGGCGCTGATGGCGTCTCATCAGAGACCGATGAA
GAGCCGACATTTACCTTCAGCCAGTTGCATGATGAAGTCAACATTTGGCGCCGCAATGTTG
CTGTCCGCTGGCGCTGCAGCCCTCGCGATCCCGCTATGGCTATATGCCGATGATTGCCGAA
GCGCAGATAACCTCTGCGCTGCGCGCGCATGGCGCGATCCATTCGGTGGTCTTTGGCG
GGTTTTGCTTCGCACAGCGTGGCGCGCGCATGACGATGCCAGACCGCGCTGATTGTG
TCGGCGGATGCCGAGCGCGGGCGGTAATACTGCGGTATAAAAAGCTGCTCGATGAC
GCTATTGCGCAGGCGCAGCATCAGCCGAAACCGTTCTGCTGGTGGACAGAGGGCTGGCG
AAAAATGGCATGGTGGATGGCGCGATCTGGATTTTGGCACGTTGGCCAGCAGCAGCATCTC
GGCGCGAGCGTCCCGTGGCTGGCTGGAATCCAACGAAACCTCGTGCAATCTTTTACACC
TCCCGCACTACCGCAAACCGAAAGCGTCCAGCGCGACGTCGGCGGTTATGCCGTGGCG
CTGGCAACCTCGATGGACACCAATTTTGGCGGCAAGGCGGGCGCGTATTCTTTTGCACA
TCGGATATCGGCTGGGTCGTCGGCCACTCCTATATCGTTTACCGCGCGTGGCTGGCAGGC
ATGGCGACTATTGTTTACGAAGGACTGCCGACGTACCGGACTGCGGGGTCTGGTGGAAA
ATTTGTCGAGAAATACCAGSTTAACCGGATGTTTTCCGCCCGACCGGATTCGGCTGCTG
AAAAAATCCCGACGCGCAAAATCCGCAATCAGGATCTCTCCTCGCTGGAGGCGCTTTAT
CTGGCCGGTTCAGCCGCTGGACGAGCCGACCGCCAGTTGGGTAACGGAGACGCTGGGCGTA
CCGGTCATCGACAATTTATGGCAGACGGAGTCCGGCTGGCCGATCATGGCGCTGGCCCGC
GCGCTGGACGACAGGCGCTCGCGTCTGGGAAGTCCCGCGTGGCGATGTACGGTTATAAC
GTCCAGCTACTCAATGAAGTCAACCGGCAACCTGCGCGCATAAATGAAAAGGGGATGCTG
GTGATCGAAGGGCGCTGCCGCGGGCTGTATTTCAGACTATTTGGGGCGACGATGCGCGT
TTTTGTAAGACTTACTGGTCGCTGTTTAAACCGTCAGGTTTATGCCACTTTCGACTGGGGA
ATCCGCGACGCGAGGGGTATTACTTTATTCTGGGCCGTACCGATGATGTGATTAATATT
CGGGTTCATCGGCTGGGACCGGAGAAATAGAAGAAAGTATCTCCAGCTACCCGAAACGTA
GCGGAAGTGGCGGTAGTGGGGATAAAGACGCTCTGAAAGGGCAGGTAGCGGTGGCGTTT
GTCATTCGAAGCAGAGCGATACGCTGGCGGATCGCGAGGCGGGCGCGCAGGAAAAC
GCGATTTATGGCGCTGGTGGACAACCGATCGGTCACTTTGGTTCGTCGGCGCATGTCGG
TTTTTTTCGAGCTCCCAAAACCGCTTCCGAAAAGATGCTTCGCGCACGATCCAGGCG
ATCTCGAAGGCCGCGATCCGGCGATCTGACAACCATGACGATCCCGCGTCTGTCAG
CAAAATTCGCCAGCGCATCGAACA

Figure 24

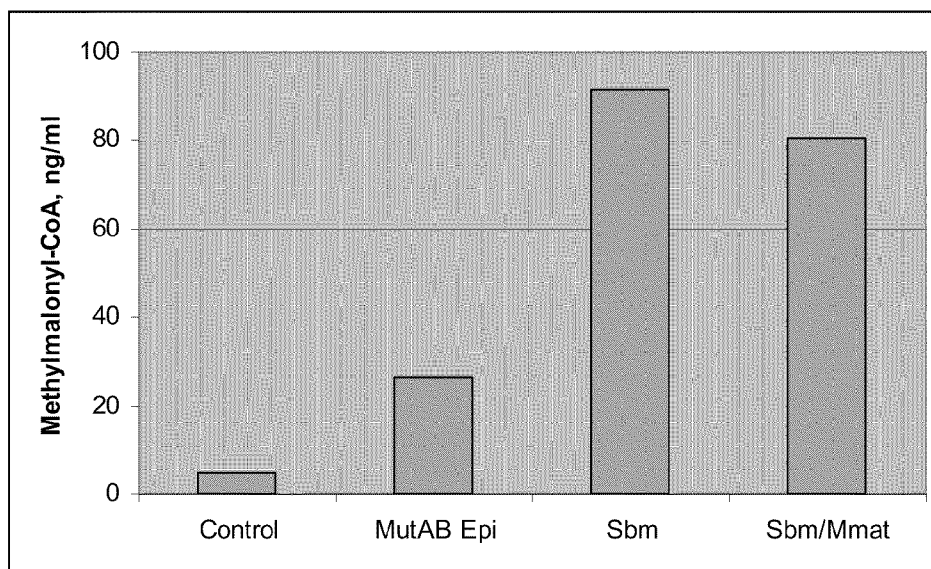


Figure 25

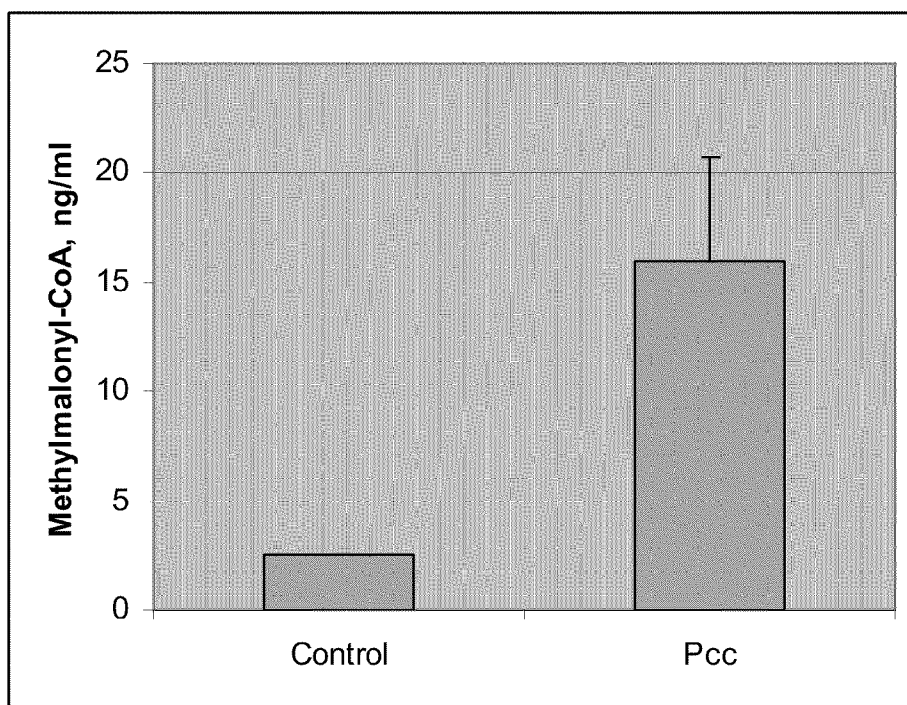


Figure 26

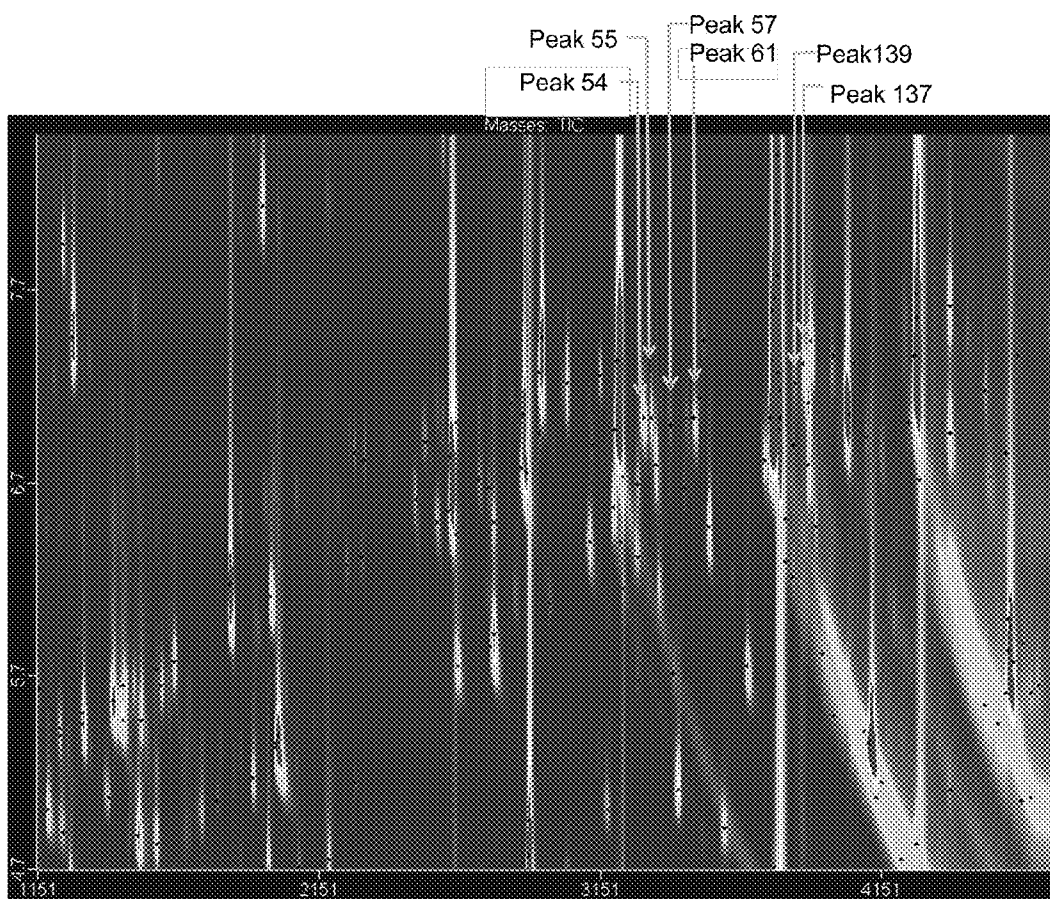


Figure 27

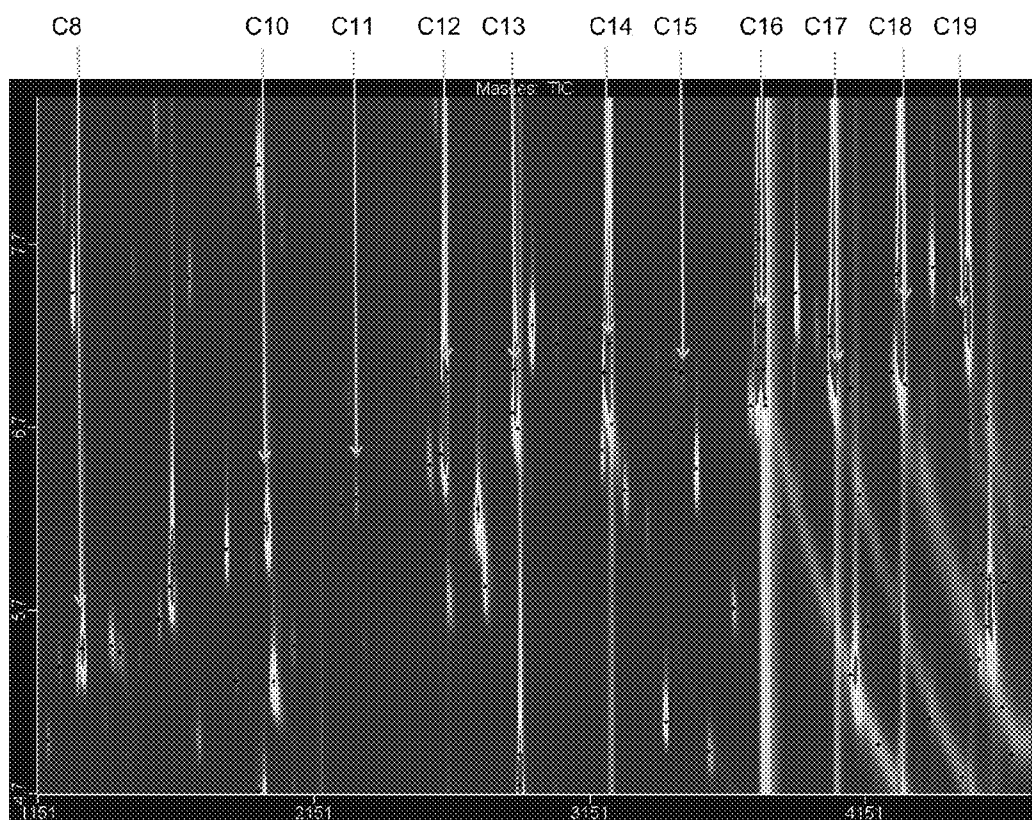
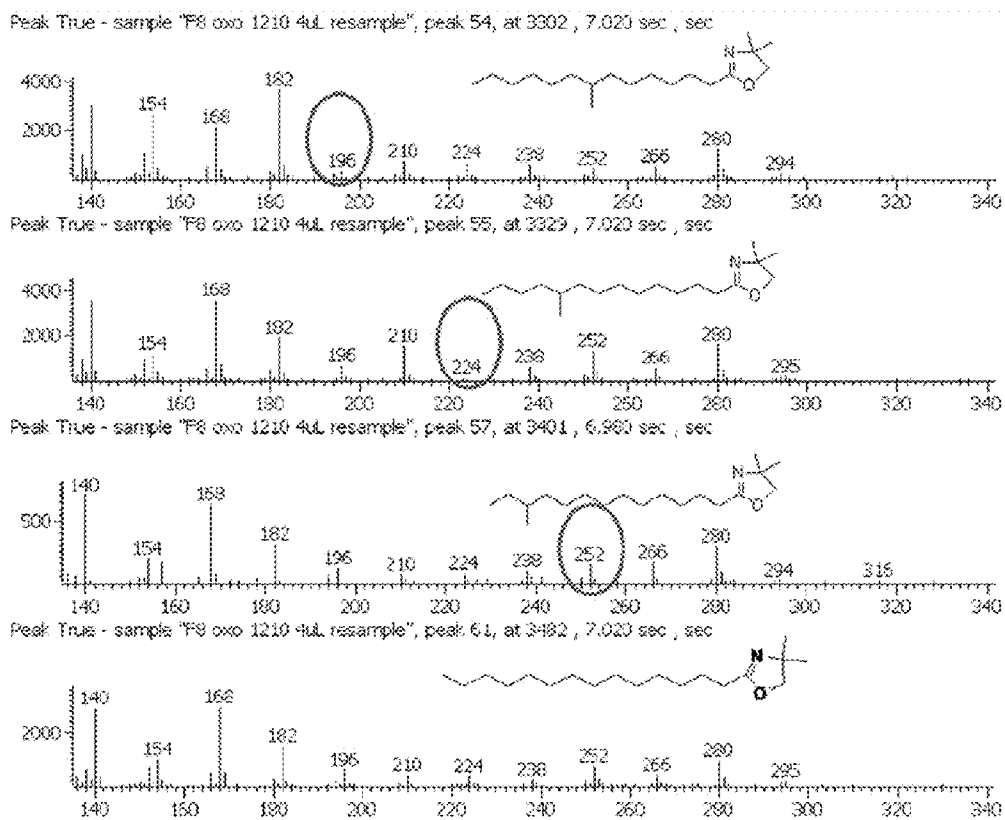


Figure 28



SCATTERED BRANCHED-CHAIN FATTY ACIDS AND BIOLOGICAL PRODUCTION THEREOF

CROSS-REFERENCE TO RELATED APPLICATIONS AND INCORPORATION BY REFERENCE

[0001] This application claims priority to U.S. Provisional Patent Application No. 61/294,274, filed Jan. 12, 2010, which is hereby incorporated by reference in its entirety.

FIELD OF THE INVENTION

[0002] The invention relates to cells and methods for producing fatty acids, and more particularly relates to cells and methods for producing scattered branched-chain fatty acids.

BACKGROUND OF THE INVENTION

[0003] Branched-chain fatty acids are carboxylic acids with a methyl or ethyl branch on one or more carbons that can be either chemically synthesized or isolated from certain animals and bacteria. While certain bacteria, such as *Escherichia coli*, do not naturally produce branched-chain fatty acids, some bacteria, such as members of the genera *Bacillus* and *Streptomyces*, can naturally produce these fatty acids. For example, *Streptomyces avermitilis* and *Bacillus subtilis* both produce branched-chain fatty acids with from 14 to 17 total carbons, with the branches in the iso and anteiso positions (Cropp et al., *Can. J. Microbiology* 46: 506-14 (2000); De Mendoza et al., *Biosynthesis and Function of Membrane Lipids, in Bacillus subtilis and Other Gram-Positive Bacteria*, Sonenshein and Losick, eds., American Society for Microbiology (1993)). However, these organisms do not produce branched-chain fatty acids in amounts that are commercially useful. Another limitation of these natural organisms is that they apparently do not produce medium-chain branched-chain fatty acids, such as those with 11 or 13 carbons. In addition, if fatty acids having particular chain lengths, branches on particular carbons, or branches at positions other than the iso and anteiso positions are desired, these fatty acids may not be available or easily isolated from a natural organism in meaningful quantities.

[0004] As such, there remains a need for commercially useful, bacterially-produced, branched-chain fatty acids. In addition, there remains a need for a method of producing such branched-chain fatty acids.

SUMMARY OF THE INVENTION

[0005] Methods and cells for producing scattered branched-chain fatty acids are provided. In certain embodiments, the method for producing branched-chain fatty acids in a cell includes expressing in the cell one or more recombinant polypeptides that catalyze the conversion of methylmalonyl-CoA to methylmalonyl-ACP; and culturing the cell under conditions suitable for producing the polypeptide, such that branched-chain fatty acids are produced.

[0006] Also provided is a method for producing branched-chain fatty acids in a cell, the method including expressing in the cell one or more recombinant polypeptides that increase the production of methylmalonyl-CoA in the cell; and culturing the cell under conditions suitable for producing the recombinant polypeptide, such that branched-chain fatty acids are produced.

[0007] In certain embodiments, a method for producing branched-chain fatty acids in a cell is provided, the method including expressing in the cell a polypeptide that has propionyl-CoA synthetase activity; inhibiting propionylation of the propionyl-CoA synthetase; and culturing the cell under conditions suitable for producing the polypeptide, such that branched-chain fatty acids are produced.

[0008] Further provided is a method for producing branched-chain fatty acids in a cell, the method including expressing in the cell a polypeptide that has methylmalonyl-CoA mutase activity; expressing in a cell a polypeptide that has methylmalonyl-CoA epimerase activity; and culturing the cell under conditions suitable for producing the polypeptides, such that branched-chain fatty acids are produced.

[0009] A composition comprising a mixture of biologically-produced branched-chain fatty acids is also provided. The composition can include branched-chain fatty acids having a chain length of C12 to C16 and from about 1 to about 3 methyl branches positioned on one or more even-numbered carbons.

[0010] In certain embodiments, a method for producing branched-chain fatty acids in a cell is provided, the method including expressing in the cell one or more recombinant polypeptides that increase the production of methylmalonyl-CoA in the cell; expressing in the cell a recombinant polypeptide that catalyzes the conversion of methylmalonyl-CoA to methylmalonyl-ACP; and culturing the cell under conditions suitable for producing the recombinant polypeptide, such that branched-chain fatty acids are produced.

[0011] In addition, in certain embodiments, a method for producing branched-chain fatty acids in a cell is provided, the method including expressing in the cell one or more recombinant polypeptides that increase the production of methylmalonyl-CoA in the cell; expressing in the cell a recombinant polypeptide that catalyzes the conversion of methylmalonyl-CoA to methylmalonyl-ACP; expressing in the cell a recombinant thioesterase; and culturing the cell under conditions suitable for producing the recombinant polypeptide, such that branched-chain fatty acids are produced.

[0012] Also provided is a method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 10 to 18 carbons and branching at the second carbon. The method includes modifying the cell to increase carbon flow to methylmalonyl-CoA; and culturing the cell under conditions suitable for carbon flow to methylmalonyl-CoA to be increased, such that branched-chain fatty acids having a chain length from about 10 to about 18 carbons and branching at the second carbon are produced. In certain embodiments, the branching can be on the fourth, sixth, eighth, tenth, or twelfth carbon.

[0013] In certain embodiments, a method for producing branched-chain fatty acids in a cell is provided, the branched-chain fatty acids having a chain length from about 10 to 18 carbons and branching at the second carbon. The method includes modifying the cell to generate methylmalonyl-ACP from methylmalonyl-CoA; and culturing the cell under conditions suitable for generation of methylmalonyl-ACP from methylmalonyl-CoA, such that branched-chain fatty acids having a chain length from about 10 to about 18 carbons and branching at the second carbon are produced. In certain embodiments, the branching can be on the fourth, sixth, eighth, tenth, or twelfth carbon.

[0014] A method for producing modified fatty acids in a cell is also provided, the method including providing a cell

having type II fatty acid synthase activity; expressing in the cell one or more recombinant polypeptides that catalyze formation of at least one intermediate metabolite, wherein the at least one intermediate metabolite is incorporated by the type II fatty acid synthase; and culturing the cell under conditions suitable for producing the recombinant polypeptide, such that modified fatty acids are produced.

[0015] Further provided is an *Escherichia* cell that produces branched-chain fatty acids having a chain length from about 10 to about 18 carbons and comprising one or more methyl branches on one or more even-numbered carbons.

[0016] The invention further provides a method for producing branched-chain fatty acid comprising a methyl on one or more even number carbons. The method comprises culturing a cell comprising (aa) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of propionyl-CoA to methylmalonyl-CoA and/or (bb) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of succinyl-CoA to methylmalonyl-CoA. The cell is cultured under conditions allowing expression of the polynucleotide(s) and production of branched-chain fatty acid. Optionally, the method further comprises extracting from the culture the branched-chain fatty acid or a product of the branched-chain fatty acid. Also provided is a cell comprising (i) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding an acyl transferase lacking polyketide synthesis activity, and (ii) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a propionyl-CoA carboxylase and/or an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA mutase, which are expressed in the cell. The cell produces more branched-chain fatty acid comprising a methyl on one or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s).

[0017] The following numbered paragraphs each succinctly define one or more exemplary variations of the invention:

[0018] 1. A method for producing branched-chain fatty acid comprising a methyl on one or more even number carbons, the method comprising culturing a cell comprising

[0019] (aa) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of propionyl-CoA to methylmalonyl-CoA and/or (bb) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of succinyl-CoA to methylmalonyl-CoA, under conditions allowing expression of the polynucleotide(s) and production of branched-chain fatty acid, wherein the cell produces more fatty acid comprising a methyl on one or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s).

[0020] 2. The method of paragraph 1 further comprising extracting from culture the branched-chain fatty acid or a product of the branched-chain fatty acid.

[0021] 3. The method of paragraph 1 or paragraph 2, wherein the polypeptide that catalyzes the conversion of propionyl-CoA to methylmalonyl-CoA is a propionyl-CoA carboxylase and/or the polypeptide that catalyzes the conversion of succinyl-CoA to methylmalonyl-CoA is a methylmalonyl-CoA mutase.

[0022] 4. The method of paragraph 3, wherein (i) the propionyl-CoA carboxylase is *Streptomyces coelicolor* PccB and AccA1 or PccB and AccA2 and/or (ii) the methylmalonyl-CoA mutase is *Janibacter* sp. HTCC2649 methylmalonyl-CoA mutase, *S. cinnamomensis* MutA and MutB, or *E. coli* Sbm.

[0023] 5. The method of paragraph 3, wherein (i) the methylmalonyl-CoA mutase comprises an amino acid sequence having at least about 80% sequence identity to the amino acid sequence set forth in SEQ ID NOs: 3, 4, or 28 and/or (ii) the propionyl-CoA carboxylase comprises an amino acid sequence having at least about 80% sequence identity to the amino acid sequence set forth in SEQ ID NOs: 9 and 10.

[0024] 6. The method of any one of paragraphs 3-5, wherein the cell comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA mutase and further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA epimerase.

[0025] 7. The method of any one of paragraphs 1-6, wherein the cell further comprises an exogenous or overexpressed polynucleotide encoding an acyl transferase lacking polyketide synthesis activity and/or an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a thioesterase.

[0026] 8. The method of paragraph 7, wherein the acyl transferase is FabD, an acyl transferase domain of a polyketide synthase, or an acyl transferase domain of *Mycobacterium* mycocerosic acid synthase.

[0027] 9. The method of any one of paragraphs 1-8, wherein the cell has been modified to attenuate endogenous methylmalonyl-CoA mutase activity, endogenous methylmalonyl-CoA decarboxylase activity, and/or endogenous acyl transferase activity.

[0028] 10. The method of any one of paragraphs 1-9, wherein the cell produces a Type II fatty acid synthase.

[0029] 11. The method of any one of paragraphs 1-10, wherein the cell is *Escherichia coli*.

[0030] 12. A branched-chain fatty acid produced by the method of any one of paragraphs 1-11.

[0031] 13. A cell comprising: (i) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding an acyl transferase lacking polyketide synthesis activity, and (ii) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a propionyl-CoA carboxylase and/or an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA mutase, wherein the polynucleotide(s) are expressed and the cell produces more branched-chain fatty acid comprising a methyl on one or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s).

[0032] 14. The cell of paragraph 13, wherein (i) the propionyl-CoA carboxylase is *Streptomyces coelicolor* PccB and AccA1 or PccB and AccA2 and/or (ii) the methylmalonyl-CoA mutase is *Janibacter* sp. HTCC2649 methylmalonyl-CoA mutase, *S. cinnamomensis* MutA and MutB, or *E. coli* Sbm.

[0033] 15. The cell of paragraph 13, wherein (i) the methylmalonyl-CoA mutase comprises an amino acid sequence having at least about 80% sequence identity to the amino acid sequence set forth in SEQ ID NOs: 3, 4, or 28 and/or (ii) the propionyl-CoA carboxylase comprises an amino acid

sequence having at least about 80% sequence identity to the amino acid sequence set forth in SEQ ID NOs: 9 and 10.

[0034] 16. The cell of any one of paragraphs 13-15, wherein the cell comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA mutase and further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA epimerase.

[0035] 17. The cell of any one of paragraphs 13-16, wherein the acyl transferase is FabD, an acyl transferase domain of a polyketide synthase, or an acyl transferase domain of *Mycobacterium mycocerosic acid synthase*.

[0036] 18. The cell of any one of paragraphs 13-17, wherein the cell further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a thioesterase.

[0037] 19. The cell of any one of paragraphs 13-18, wherein the cell has been modified to attenuate endogenous methylmalonyl-CoA mutase activity, endogenous methylmalonyl-CoA decarboxylase activity, and/or endogenous acyl transferase activity.

[0038] 20. The cell of any one of paragraphs 13-19, wherein the cell is *Escherichia coli*.

[0039] 21. A method for producing branched-chain fatty acids in a cell comprising: a. expressing in the cell one or more recombinant polypeptides that catalyze the conversion of methylmalonyl-CoA to methylmalonyl-ACP; and b. culturing the cell under conditions suitable for producing the polypeptide, such that branched-chain fatty acids are produced.

[0040] 22. The method of paragraph 21, wherein the polypeptide is an acyl transferase.

[0041] 23. The method of paragraph 21, wherein the polypeptide is encoded by fabD.

[0042] 24. The method of paragraph 22, wherein the polypeptide is a polyketide synthase or a portion thereof.

[0043] 25. The method of paragraph 21, wherein the polypeptide is a *Mycobacterium mycocerosic acid synthase* or a portion thereof

[0044] 26. The method of paragraph 21, wherein the polypeptide has at least about 60% sequence identity to a sequence set forth in SEQ ID NO: 19.

[0045] 27. The method of paragraph 21, wherein the method further includes expressing in the cell a polypeptide that encodes an exogenous thioesterase.

[0046] 28. The method of paragraph 21, wherein the cell is an *Escherichia coli*.

[0047] 29. The method of paragraph 21, wherein the cell produces higher levels of branched-chain fatty acids after expression of the polypeptide than it did prior to expression of the polypeptide.

[0048] 30. The method of paragraph 21, wherein the branched-chain fatty acids comprise one or more methyl branches.

[0049] 31. The method of paragraph 30, wherein the one or more methyl branches are on even numbered carbons.

[0050] 32. The method of paragraph 21, wherein the branched-chain fatty acids are not naturally produced in the cell.

[0051] 33. Branched-chain fatty acids produced by the method of paragraph 21.

[0052] 34. A cell comprising at least one recombinant polypeptide that catalyzes the conversion of methylmalonyl-CoA to methylmalonyl-ACP, wherein the cell comprising the

recombinant polypeptide produces more branched-chain fatty acids than an otherwise similar cell that does not comprise the recombinant polypeptide.

[0053] 35. A method for producing branched-chain fatty acids in a cell comprising: a. expressing in the cell one or more recombinant polypeptides that increase the production of methylmalonyl-CoA in the cell; and b. culturing the cell under conditions suitable for producing the recombinant polypeptide, such that branched-chain fatty acids are produced.

[0054] 36. The method of paragraph 35, wherein expression of the polypeptide results in increased propionyl-CoA synthetase activity in the cell.

[0055] 37. The method of paragraph 35, wherein the polypeptide has propionyl-CoA carboxylase activity.

[0056] 38. The method of paragraph 35, wherein the polypeptide has at least about 60% sequence identity to a sequence set forth in SEQ ID NO: 9 or SEQ ID NO: 10.

[0057] 39. The method of paragraph 35, wherein the method further includes expressing in the cell a polypeptide that encodes an exogenous thioesterase.

[0058] 40. The method of paragraph 35, wherein the cell is an *Escherichia coli* cell.

[0059] 41. The method of paragraph 35, wherein the cell produces higher levels of branched-chain fatty acids after expression of the polypeptide than it did prior to expression of the polypeptide.

[0060] 42. The method of paragraph 35, wherein the branched-chain fatty acids comprise one or more methyl branches.

[0061] 43. The method of paragraph 42, wherein the one or more methyl branches are on even numbered carbons.

[0062] 44. The method of paragraph 35, wherein the branched-chain fatty acids are not naturally produced in the cell.

[0063] 45. Branched-chain fatty acids produced by the method of paragraph 35.

[0064] 46. A cell comprising at least one recombinant polypeptide that increases the production of methylmalonyl-CoA in the cell, wherein the cell comprising the recombinant polypeptide produces more branched-chain fatty acids than an otherwise similar cell that does not comprise the recombinant polypeptide.

[0065] 47. A method for producing branched-chain fatty acids in a cell comprising: a. expressing in the cell a polypeptide that has propionyl-CoA synthetase activity; b. inhibiting propionylation of the propionyl-CoA synthetase; and c. culturing the cell under conditions suitable for producing the polypeptide, such that branched-chain fatty acids are produced.

[0066] 48. The method of paragraph 47, wherein the polypeptide does not include a lysine that is subject to propionylation.

[0067] 49. The method of paragraph 47, wherein step c) includes providing a source of resveratrol into a culture medium used to culture the cell.

[0068] 50. The method of paragraph 47, wherein the cell does not include an N-acetyltransferase enzyme responsible for propionylation of the propionyl-CoA synthetase.

[0069] 51. The method of paragraph 47, wherein the polypeptide has at least about 60% sequence identity to the protein encoded by SEQ ID NO: 22.

- [0070] 52. The method of paragraph 47, wherein the cell contains increased enzymatic activity for removal of propionyl groups from one or more lysine residues of propionyl-CoA synthetase.
- [0071] 53. The method of paragraph 47, wherein the method further includes expressing in the cell a polypeptide that encodes an exogenous thioesterase.
- [0072] 54. The method of paragraph 47, wherein the cell is an *Escherichia* cell.
- [0073] 55. The method of paragraph 47, wherein the cell produces higher levels of branched-chain fatty acids after expression of the polypeptide than it did prior to expression of the polypeptide.
- [0074] 56. The method of paragraph 47, wherein the branched-chain fatty acids comprise one or more methyl branches.
- [0075] 57. The method of paragraph 56, wherein the one or more methyl branches are on even numbered carbons.
- [0076] 58. The method of paragraph 47, wherein the branched-chain fatty acids are not naturally produced in the cell.
- [0077] 59. Branched-chain fatty acids produced by the method of paragraph 47.
- [0078] 60. A method for producing branched-chain fatty acids in a cell comprising: a. expressing in the cell a polypeptide that has methylmalonyl-CoA mutase activity; b. expressing in a cell a polypeptide that has methylmalonyl-CoA epimerase activity; and c. culturing the cell under conditions suitable for producing the polypeptides, such that branched-chain fatty acids are produced.
- [0079] 61. The method of paragraph 60, wherein the methylmalonyl-CoA mutase polypeptide has at least about 60% sequence identity to a sequence set forth in SEQ ID NO: 3 or SEQ ID NO: 4.
- [0080] 62. The method of paragraph 60, wherein the methylmalonyl-CoA epimerase polypeptide has at least about 60% sequence identity to a sequence set forth in SEQ ID NO: 6.
- [0081] 63. The method of paragraph 60, wherein the method further includes expressing in the cell a polypeptide that encodes an exogenous thioesterase.
- [0082] 64. The method of paragraph 60, wherein the cell is an *Escherichia* cell.
- [0083] 65. The method of paragraph 60, wherein the cell produces higher levels of branched-chain fatty acids after expression of the polypeptide than it did prior to expression of the polypeptide.
- [0084] 66. The method of paragraph 60, wherein the branched-chain fatty acids comprise one or more methyl branches.
- [0085] 67. The method of paragraph 66, wherein the one or more methyl branches are on even numbered carbons.
- [0086] 68. The method of paragraph 60, wherein the branched-chain fatty acids are not naturally produced in the cell.
- [0087] 69. Branched-chain fatty acids produced by the method of paragraph 60.
- [0088] 70. A cell comprising recombinant polypeptides having methylmalonyl-CoA mutase activity and methylmalonyl-CoA epimerase activity, wherein the cell comprising the recombinant polypeptides produces more branched-chain fatty acids than an otherwise similar cell that does not comprise the recombinant polypeptide.
- [0089] 71. A composition comprising a mixture of biologically-produced branched-chain fatty acids, the branched-chain fatty acids having a chain length of C12 to C16 and from about 1 to about 3 methyl branches positioned on one or more even-numbered carbons.
- [0090] 72. A method for producing branched-chain fatty acids in a cell comprising: a. expressing in the cell one or more recombinant polypeptides that increase the production of methylmalonyl-CoA in the cell; b. expressing in the cell a recombinant polypeptide that catalyzes the conversion of methylmalonyl-CoA to methylmalonyl-ACP; and c. culturing the cell under conditions suitable for producing the recombinant polypeptide, such that branched-chain fatty acids are produced.
- [0091] 73. The method of paragraph 72, wherein the cell has a deletion in a gene for a methylmalonyl-CoA decarboxylase.
- [0092] 74. The method of paragraph 72, wherein the cell additionally produces a recombinant polypeptide with a 3-ke-toacyl-ACP synthase activity that recognizes methylmalonyl-ACP as a substrate.
- [0093] 75. A method for producing branched-chain fatty acids in a cell comprising: a. expressing in the cell one or more recombinant polypeptides that increase the production of methylmalonyl-CoA in the cell; b. expressing in the cell a recombinant polypeptide that catalyzes the conversion of methylmalonyl-CoA to methylmalonyl-ACP; c. expressing in the cell a recombinant thioesterase; and d. culturing the cell under conditions suitable for producing the recombinant polypeptide, such that branched-chain fatty acids are produced.
- [0094] 76. The method of paragraph 75, wherein the cell has a deletion in a gene for a methylmalonyl-CoA decarboxylase.
- [0095] 77. The method of paragraph 75, wherein the cell additionally produces a recombinant polypeptide with a 3-ke-toacyl-ACP synthase activity that recognizes methylmalonyl-ACP as a substrate.
- [0096] 78. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 10 to 18 carbons and branching at the second carbon, the method comprising: a. modifying the cell to increase carbon flow to methylmalonyl-CoA; and b. culturing the cell under conditions suitable for carbon flow to methylmalonyl-CoA to be increased, such that branched-chain fatty acids having a chain length from about 10 to about 18 carbons and branching at the second carbon are produced.
- [0097] 79. The method of paragraph 78, wherein the branching at the second carbon is a methyl branch.
- [0098] 80. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 10 to 18 carbons and branching at the fourth carbon, the method comprising: a. modifying the cell to increase carbon flow to methylmalonyl-CoA; and b. culturing the cell under conditions suitable for carbon flow to methylmalonyl-CoA to be increased, such that branched-chain fatty acids having a chain length from about 10 to about 18 carbons and branching at the fourth carbon are produced.
- [0099] 81. The method of paragraph 80, wherein the branching at the fourth carbon is a methyl branch.
- [0100] 82. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 10 to 18 carbons and branching at the sixth carbon, the method comprising: a. modifying the cell to

increase carbon flow to methylmalonyl-CoA; and b. culturing the cell under conditions suitable for carbon flow to methylmalonyl-CoA to be increased, such that branched-chain fatty acids having a chain length from about 10 to about 18 carbons and branching at the sixth carbon are produced.

[0101] 83. The method of paragraph 82, wherein the branching at the sixth carbon is a methyl branch.

[0102] 84. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 12 to 18 carbons and branching at the eighth carbon, the method comprising: a. modifying the cell to increase carbon flow to methylmalonyl-CoA; and b. culturing the cell under conditions suitable for carbon flow to methylmalonyl-CoA to be increased, such that branched-chain fatty acids having a chain length from about 12 to about 18 carbons and branching at the eighth carbon are produced.

[0103] 85. The method of paragraph 84, wherein the branching at the eighth carbon is a methyl branch.

[0104] 86. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 14 to 18 carbons and branching at the tenth carbon, the method comprising: a. modifying the cell to increase carbon flow to methylmalonyl-CoA; and b. culturing the cell under conditions suitable for carbon flow to methylmalonyl-CoA to be increased, such that branched-chain fatty acids having a chain length from about 14 to about 18 carbons and branching at the tenth carbon are produced.

[0105] 87. The method of paragraph 86, wherein the branching at the tenth carbon is a methyl branch.

[0106] 88. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 16 to 18 carbons and branching at the twelfth carbon, the method comprising: a. modifying the cell to increase carbon flow to methylmalonyl-CoA; and b. culturing the cell under conditions suitable for carbon flow to methylmalonyl-CoA to be increased, such that branched-chain fatty acids having a chain length from about 16 to about 18 carbons and branching at the twelfth carbon are produced.

[0107] 89. The method of paragraph 88, wherein the branching at the twelfth carbon is a methyl branch.

[0108] 90. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 10 to 18 carbons and branching at the second carbon, the method comprising: a. modifying the cell to generate methylmalonyl-ACP from methylmalonyl-CoA; and b. culturing the cell under conditions suitable for generation of methylmalonyl-ACP from methylmalonyl-CoA, such that branched-chain fatty acids having a chain length from about 10 to about 18 carbons and branching at the second carbon are produced.

[0109] 91. The method of paragraph 90, wherein the branching at the second carbon is a methyl branch.

[0110] 92. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 10 to 18 carbons and branching at the fourth carbon, the method comprising: a. modifying the cell to generate methylmalonyl-ACP from methylmalonyl-CoA; and b. culturing the cell under conditions suitable for generation of methylmalonyl-ACP from methylmalonyl-CoA, such that branched-chain fatty acids having a chain length from about 10 to about 18 carbons and branching at the fourth carbon are produced.

[0111] 93. The method of paragraph 92, wherein the branching at the fourth carbon is a methyl branch.

[0112] 94. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 10 to 18 carbons and branching at the sixth carbon, the method comprising: a. modifying the cell to generate methylmalonyl-ACP from methylmalonyl-CoA; and b. culturing the cell under conditions suitable for generation of methylmalonyl-ACP from methylmalonyl-CoA, such that branched-chain fatty acids having a chain length from about 10 to about 18 carbons and branching at the sixth carbon are produced.

[0113] 95. The method of paragraph 94, wherein the branching at the sixth carbon is a methyl branch.

[0114] 96. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 12 to 18 carbons and branching at the eighth carbon, the method comprising: a. modifying the cell to generate methylmalonyl-ACP from methylmalonyl-CoA; and b. culturing the cell under conditions suitable for generation of methylmalonyl-ACP from methylmalonyl-CoA, such that branched-chain fatty acids having a chain length from about 12 to about 18 carbons and branching at the eighth carbon are produced.

[0115] 97. The method of paragraph 96, wherein the branching at the eighth carbon is a methyl branch.

[0116] 98. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 14 to 18 carbons and branching at the tenth carbon, the method comprising: a. modifying the cell to generate methylmalonyl-ACP from methylmalonyl-CoA; and b. culturing the cell under conditions suitable for generation of methylmalonyl-ACP from methylmalonyl-CoA, such that branched-chain fatty acids having a chain length from about 14 to about 18 carbons and branching at the tenth carbon are produced.

[0117] 99. The method of paragraph 98, wherein the branching at the tenth carbon is a methyl branch.

[0118] 100. A method for producing branched-chain fatty acids in a cell, the branched-chain fatty acids having a chain length from about 16 to 18 carbons and branching at the twelfth carbon, the method comprising: a. modifying the cell to generate methylmalonyl-ACP from methylmalonyl-CoA; and b. culturing the cell under conditions suitable for generation of methylmalonyl-ACP from methylmalonyl-CoA, such that branched-chain fatty acids having a chain length from about 16 to about 18 carbons and branching at the twelfth carbon are produced.

[0119] 101. The method of paragraph 100, wherein the branching at the twelfth carbon is a methyl branch.

[0120] 102. A method for producing modified fatty acids in a cell comprising: a. providing a cell having type II fatty acid synthase activity; b. expressing in the cell one or more recombinant polypeptides that catalyze formation of at least one intermediate metabolite, wherein the at least one intermediate metabolite is incorporated by the type II fatty acid synthase; and c. culturing the cell under conditions suitable for producing the recombinant polypeptide, such that modified fatty acids are produced.

[0121] 103. The method of paragraph 102, wherein the cell is an *Escherichia* cell.

[0122] 104. The method of paragraph 102, wherein the intermediate metabolite is methylmalonyl-ACP.

[0123] 105. The method of paragraph 102, wherein the polypeptide(s) catalyze the conversion of methylmalonyl-CoA to methylmalonyl-ACP.

[0124] 106. The method of paragraph 102, wherein the cell produces higher levels of modified fatty acids after expression of the polypeptide than it did prior to expression of the polypeptide.

[0125] 107. The method of paragraph 102, wherein the modified fatty acids comprise one or more methyl branches on even-numbered carbons.

[0126] 108. The method of paragraph 102, wherein the polypeptide is an acyl transferase.

[0127] 109. The method of paragraph 102, wherein the polypeptide is encoded by *fabD*.

[0128] 110. The method of paragraph 102, wherein the polypeptide is a polyketide synthase or a portion thereof.

[0129] 111. The method of paragraph 102, wherein the polypeptide is a *Mycobacterium* mycocerosic acid synthase or a portion thereof.

[0130] 112. An *Escherichia* cell that produces branched-chain fatty acids having a chain length from about 10 to about 18 carbons and comprising one or more methyl branches on one or more even-numbered carbons.

BRIEF DESCRIPTION OF THE DRAWINGS

[0131] FIG. 1 is a *mutA* nucleotide sequence (SEQ ID NO: 1).

[0132] FIG. 2 is a *mutB* nucleotide sequence (SEQ ID NO: 2).

[0133] FIG. 3 is a *MutA* protein sequence (SEQ ID NO: 3).

[0134] FIG. 4 is a *MutB* protein sequence (SEQ ID NO: 4).

[0135] FIG. 5 is a methylmalonyl-CoA epimerase nucleotide sequence (SEQ ID NO: 5).

[0136] FIG. 6 is a methylmalonyl-CoA epimerase protein sequence (SEQ ID NO: 6).

[0137] FIG. 7 is a DNA sequence for *accA1* (GenBank Accession No. AF113603.1) (SEQ ID NO: 7).

[0138] FIG. 8 is a DNA sequence for *pccB* (GenBank Accession No. AF113605.1) (SEQ ID NO: 8).

[0139] FIG. 9 is a protein sequence for *AccA1* (SEQ ID NO: 9).

[0140] FIG. 10 is a protein sequence for *PccB* (SEQ ID NO: 10).

[0141] FIG. 11 shows element 1 including the *P_{Llac0-1}* sequence and the phage T7 gene10 ribosome binding site (SEQ ID NO: 11).

[0142] FIG. 12 shows element 2 including the optimized *accA1* gene sequence (SEQ ID NO: 12).

[0143] FIG. 13 shows element 3 including the spacer sequence (SEQ ID NO: 13).

[0144] FIG. 14 shows element 4 including the optimized *pccB* sequence (SEQ ID NO: 14).

[0145] FIG. 15 is a synthetic sequence for propionyl-CoA carboxylase gene expression (SEQ ID NO: 15).

[0146] FIG. 16 is the forward primer sequence for *PrpE* (SEQ ID NO: 16).

[0147] FIG. 17 is the reverse primer sequence for *PrpE* (SEQ ID NO: 17).

[0148] FIG. 18 is the MMAT domain sequence from *Mycobacterium bovis* BCG (SEQ ID NO: 18).

[0149] FIG. 19 is a protein sequence for the *Mycobacterium bovis* BCG MAS (GenBank Accession No. YP_979046) (SEQ ID NO: 19).

[0150] FIG. 20 is a codon-optimized MMAT domain DNA sequence from *Mycobacterium bovis* BCG (SEQ ID NO: 20).

[0151] FIG. 21 is an alignment of a codon-optimized MMAT domain from *Mycobacterium bovis* BCG with the original sequence (SEQ ID NOs: 20 and 21).

[0152] FIG. 22 is the protein sequence of *Salmonella enterica* propionyl CoA synthase *PrpE* (GenBank Accession No. AAC44817) (SEQ ID NO: 22).

[0153] FIG. 23 is the DNA sequence of *Salmonella enterica* propionyl CoA synthase *PrpE* (SEQ ID NO: 23).

[0154] FIG. 24 is a bar graph illustrating methylmalonyl-CoA production (ng/ml) in *E. coli* strain K27-Z1 harboring *pTrcHisA pZA31* (control), *pZA31 mutAB Ss epi* (*MutAB Epi*), *pTrcHisA Ec sbm* (*Sbm*), or *pTrcHisA Ec sbm pZA31 Mb mmat* (*Sbm/Mmat*). No methylmalonyl-CoA was identified in the control sample; the figure indicates the background level of detection.

[0155] FIG. 25 is a bar graph illustrating methylmalonyl-CoA production (ng/ml) in *E. coli* BW25113 (control) and BW25113 harboring *pZA31-accA1-pccB* (*Pcc*). No methylmalonyl-CoA was identified in the control sample; the figure indicates the background level of detection. Two biological replicates are represented.

[0156] FIG. 26 is a two-dimensional (2D) representation of the 2D Total Ion Chromatogram resulting from a sample of fatty acid produced by BL21 Star (DE3) *E. coli* harboring *pTrcHisA Ec sbm So ce epi pZA31 mmat*. Light areas on the figure indicate the presence of sample material. Peak names and arrows indicate samples that were further characterized by mass spectrometry.

[0157] FIG. 27 is a two-dimensional (2D) representation of the 2D Total Ion Chromatogram resulting from a sample produced by a control strain, BL21 Star (DE3) *E. coli* harboring *pTrcHisA pZA31*. No branched-chain fatty acid was detected. Arrows indicate the presence of straight-chain fatty acid derivatives of the indicated chain length.

[0158] FIG. 28 is a representation of the mass spectra of peaks 54, 55, and 57 identified in FIG. 26. Eight- and ten-carbon branched-chain fatty acids are depicted in the top two profiles and were identified by the almost complete absence of the circled fragment. A twelve-branched fatty acid was tentatively identified and is depicted in the third profile.

DETAILED DESCRIPTION OF THE INVENTION

[0159] The invention relates to improved biological production of scattered branched-chain fatty acids. In addition, in certain embodiments, the invention provides improved compositions of biologically produced scattered branched-chain fatty acids having defined chain lengths with methyl branches at one or more even-numbered carbons within the fatty acid. In addition, in certain embodiments, the fatty acid length can be tailored to a predetermined length, such as, for example, to produce fatty acids with a backbone of C12 to C16. In certain embodiments, the methods and/or cells can produce a mixture of fatty acids having varied numbers of methyl branches, varied positions of the methyl branches, and varied length of the fatty acids, such as, for example, a mixture of fatty acids having a chain length of C12 to C16 and from about 0 to about 3 methyl branches positioned on one or more even-numbered carbons.

[0160] As used herein, “amplify,” “amplified,” or “amplification” refers to any process or protocol for copying a polynucleotide sequence into a larger number of polynucleotide molecules, e.g., by reverse transcription, polymerase chain reaction, and ligase chain reaction.

[0161] As used herein, an “antisense sequence” refers to a sequence that specifically hybridizes with a second polynucleotide sequence. For instance, an antisense sequence is a DNA sequence that is inverted relative to its normal orientation for transcription. Antisense sequences can express an RNA transcript that is complementary to a target mRNA molecule expressed within the host cell (e.g., it can hybridize to target mRNA molecule through Watson-Crick base pairing).

[0162] As used herein, “cDNA” refers to a DNA that is complementary or identical to an mRNA, in either single stranded or double stranded form.

[0163] As used herein, the carbons in fatty acids are numbered with the first carbon as part of the carboxylic acid group, and the second carbon (C2) adjacent to the first. The numbers continue so that the highest number carbon is farthest from the carboxylic acid group. “Even number” carbons include C2, C4, C6, C8, C10, C12, C14, and so on.

[0164] As used herein, “complementary” refers to a polynucleotide that can base pair with a second polynucleotide. Put another way, “complementary” describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, a polynucleotide having the sequence 5'-GTCCGA-3' is complementary to a polynucleotide with the sequence 5'-TCGGAC-3'.

[0165] As used herein, a “conservative substitution” refers to the substitution in a polypeptide of an amino acid with a functionally similar amino acid. Put another way, a conservative substitution involves replacement of an amino acid residue with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined within the art, and include amino acids with basic side chains (e.g., lysine, arginine, and histidine), acidic side chains (e.g., aspartic acid and glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, and cysteine), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, and tryptophan), beta-branched side chains (e.g., threonine, valine, and isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, and histidine).

[0166] As used herein, “encoding” refers to the inherent property of nucleotides to serve as templates for synthesis of other polymers and macromolecules. Unless otherwise specified, a “nucleotide sequence encoding an amino acid sequence” includes all nucleotide sequences that are degenerate versions of each other and that encode the same amino acid sequence.

[0167] As used herein, “endogenous” refers to polynucleotides, polypeptides, or other compounds that are expressed naturally or originate within an organism or cell. That is, endogenous polynucleotides, polypeptides, or other compounds are not exogenous. For instance, an “endogenous” polynucleotide or peptide is present in the cell when the cell was originally isolated from nature.

[0168] As used herein, “expression vector” refers to a vector comprising a recombinant polynucleotide comprising expression control sequences operatively linked to a nucleotide sequence to be expressed. For example, suitable expression vectors include, without limitation, autonomously replicating vectors or vectors integrated into the chromosome. In some instances, an expression vector is a viral-based vector.

[0169] As used herein, “exogenous” refers to any polynucleotide or polypeptide that is not naturally expressed or

produced in the particular cell or organism where expression is desired. Exogenous polynucleotides, polypeptides, or other compounds are not endogenous.

[0170] As used herein, “hybridization” includes any process by which a strand of a nucleic acid joins with a complementary nucleic acid strand through base-pairing. Thus, the term refers to the ability of the complement of the target sequence to bind to a test (i.e., target) sequence, or vice-versa.

[0171] As used herein, “hybridization conditions” are typically classified by degree of “stringency” of the conditions under which hybridization is measured. The degree of stringency can be based, for example, on the melting temperature (T_m) of the nucleic acid binding complex or probe. For example, “maximum stringency” typically occurs at about $T_m - 5^\circ \text{C}$. (5° below the T_m of the probe); “high stringency” at about $5-10^\circ \text{C}$. below the T_m ; “intermediate stringency” at about $10-20^\circ$ below the T_m of the probe; and “low stringency” at about $20-25^\circ \text{C}$. below the T_m . Alternatively, or in addition, hybridization conditions can be based upon the salt or ionic strength conditions of hybridization and/or one or more stringency washes. For example, $6\times \text{SSC}$ =very low stringency; $3\times \text{SSC}$ =low to medium stringency; $1\times \text{SSC}$ =medium stringency; and $0.5\times \text{SSC}$ =high stringency. Functionally, maximum stringency conditions may be used to identify nucleic acid sequences having strict (i.e., about 100%) identity or near-strict identity with the hybridization probe; while high stringency conditions are used to identify nucleic acid sequences having about 80% or more sequence identity with the probe.

[0172] As used herein, “identical” or percent “identity” in the context of two or more polynucleotide or polypeptide sequences refers to two or more sequences that are the same or have a specified percentage of nucleotides or amino acid residues that are the same, when compared and aligned for maximum correspondence, as measured using sequence comparison algorithms or by visual inspection.

[0173] As used herein, “long-chain fatty acids” refers to fatty acids with aliphatic tails longer than 14 carbons. In some embodiments of the invention, long-chain fatty acids are provided that comprise 15, 16, 17, 18, 19, 20, 21, or 22 carbons in the carbon backbone.

[0174] As used herein, “medium-chain fatty acids” refers to fatty acids with aliphatic tails between 6 and 14 carbons. In certain embodiments, the medium-chain fatty acids can have from 11 to 13 carbons.

[0175] As used herein, “naturally-occurring” refers to an object that can be found in nature. For example, a polypeptide or polynucleotide sequence that is present in an organism (including viruses) that can be isolated from a source in nature and which has not been intentionally modified by man in the laboratory is naturally-occurring.

[0176] As used herein, “operably linked,” when describing the relationship between two DNA regions or two polypeptide regions, means that the regions are functionally related to each other. For example, a promoter is operably linked to a coding sequence if it controls the transcription of the sequence; a ribosome binding site is operably linked to a coding sequence if it is positioned so as to permit translation; and a signal sequence is operably linked to a peptide if it functions as a signal sequence, such as by participating in the secretion of the mature form of the protein.

[0177] As used herein, “overexpression” refers to expression of a polynucleotide to produce a product (e.g., a polypeptide or RNA) at a higher level than the polynucleotide is

normally expressed in the host cell. An overexpressed polynucleotide is generally a polynucleotide native to the host cell, the product of which is generated in a greater amount than that normally found in the host cell. Overexpression is achieved by, for instance and without limitation, operably linking the polynucleotide to a different promoter than the polynucleotide's native promoter or introducing additional copies of the polynucleotide into the host cell.

[0178] As used herein, "polynucleotide" refers to a polymer composed of nucleotides. The polynucleotide may be in the form of a separate fragment or as a component of a larger nucleotide sequence construct, which has been derived from a nucleotide sequence isolated at least once in a quantity or concentration enabling identification, manipulation, and recovery of the sequence and its component nucleotide sequences by standard molecular biology methods, for example, using a cloning vector. When a nucleotide sequence is represented by a DNA sequence (i.e., A, T, G, C), this also includes an RNA sequence (i.e., A, U, G, C) in which "U" replaces "T." Put another way, "polynucleotide" refers to a polymer of nucleotides removed from other nucleotides (a separate fragment or entity) or can be a component or element of a larger nucleotide construct, such as an expression vector or a polycistronic sequence. Polynucleotides include DNA, RNA and cDNA sequences.

[0179] As used herein, "polypeptide" refers to a polymer composed of amino acid residues which may or may not contain modifications such as phosphates and formyl groups.

[0180] As used herein, "recombinant expression vector" refers to a DNA construct used to express a polynucleotide that encodes a desired polypeptide. A recombinant expression vector can include, for example, a transcriptional subunit comprising (i) an assembly of genetic elements having a regulatory role in gene expression, for example, promoters and enhancers, (ii) a structural or coding sequence which is transcribed into mRNA and translated into protein, and (iii) appropriate transcription and translation initiation and termination sequences. Recombinant expression vectors are constructed in any suitable manner. The nature of the vector is not critical, and any vector may be used, including plasmid, virus, bacteriophage, and transposon. Possible vectors for use in the invention include, but are not limited to, chromosomal, non-chromosomal and synthetic DNA sequences, e.g., bacterial plasmids; phage DNA; yeast plasmids; and vectors derived from combinations of plasmids and phage DNA, DNA from viruses such as vaccinia, adenovirus, fowl pox, baculovirus, SV40, and pseudorabies.

[0181] As used herein, "primer" refers to a polynucleotide that is capable of specifically hybridizing to a designated polynucleotide template and providing a point of initiation for synthesis of a complementary polynucleotide when the polynucleotide primer is placed under conditions in which synthesis is induced.

[0182] As used herein, "recombinant polynucleotide" refers to a polynucleotide having sequences that are not naturally joined together. A recombinant polynucleotide may be included in a suitable vector, and the vector can be used to transform a suitable host cell. A host cell that comprises the recombinant polynucleotide is referred to as a "recombinant host cell." The polynucleotide is then expressed in the recombinant host cell to produce, e.g., a "recombinant polypeptide."

[0183] As used herein, "specific hybridization" refers to the binding, duplexing, or hybridizing of a polynucleotide preferentially to a particular nucleotide sequence under stringent conditions.

[0184] As used herein, "stringent conditions" refers to conditions under which a probe will hybridize preferentially to its target subsequence, and to a lesser extent to, or not at all to, other sequences.

[0185] As used herein, "short-chain fatty acids" refers to fatty acids having aliphatic tails with fewer than 6 carbons.

[0186] As used herein, "substantially homologous" or "substantially identical" in the context of two nucleic acids or polypeptides, generally refers to two or more sequences or subsequences that have at least 40%, 60%, 80%, 90%, 95%, 96%, 97%, 98% or 99% nucleotide or amino acid residue identity, when compared and aligned for maximum correspondence, as measured using sequence comparison algorithms or by visual inspection. The substantial identity can exist over any suitable region of the sequences, such as, for example, a region that is at least about 50 residues in length, a region that is at least about 100 residues, or a region that is at least about 150 residues. In certain embodiments, the sequences are substantially identical over the entire length of either or both comparison biopolymers.

[0187] In one embodiment, the invention relates to a novel method of producing scattered branched-chain fatty acids (or products derived from scattered branched-chain fatty acid) using bacteria. In general, the method includes increasing the supply of methylmalonyl-CoA and/or the conversion of methylmalonyl-CoA to methylmalonyl-ACP within the cell, incorporating the branch from the methylmalonyl-CoA into the fatty acid, and, optionally, using a thioesterase to specify the range of size of the fatty acids. In certain embodiments, the method provides branched-chain fatty acids having a chain length of C12 to C16. In addition, in certain embodiments, the branched-chain fatty acids have from about 0 to about 3 methyl branches, such as from about 1 to about 3 methyl branches, such as, for example, from about 1 to about 2 methyl branches, or 1, 2, or 3 methyl branches positioned on one or more carbons. In certain embodiments, the methyl branches are positioned on even-numbered carbons.

[0188] In one embodiment, scattered branched-chain fatty acid production is increased by increasing the production of methylmalonyl-CoA within the cell via, e.g., propionyl-CoA and/or succinyl-CoA intermediates. Thus, in one aspect, the invention provides a method for producing branched-chain fatty acid comprising a methyl on one or more even number carbons. The method comprises culturing a cell comprising an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of propionyl-CoA to methylmalonyl-CoA and/or an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of succinyl-CoA to methylmalonyl-CoA. The cell is cultured under conditions allowing expression of the polynucleotide(s) and production of the branched-chain fatty acid. The cell produces more branched-chain fatty acid comprising a methyl branch on one or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s) (e.g., a cell of the same cell type or derived from the same organism that does not comprise the polynucleotide(s)). Propionyl-CoA is converted to methylmalonyl-CoA by, e.g., the action of a propionyl-CoA carboxylase. Any propionyl-CoA carboxylase that catalyzes the

conversion of propionyl-CoA to methylmalonyl-CoA is suitable for use in the inventive method. An exemplary propionyl-CoA carboxylase is a carboxylase from *Streptomyces coelicolor*, which comprises two heterologous subunits encoded by *pccB* and by either *accA1* or *accA2*. In certain embodiments, the cell of the inventive method is engineered to produce *PccB* and *AccA1* or *PccB* and *AccA2*. In one aspect, the cell comprises one or more polynucleotides encoding polypeptide(s) comprising an amino acid sequence at least about 80% identical (e.g., 85%, 90%, 95%, or 100% identical) to the amino acid sequences set forth in SEQ ID NO: 9 and/or 10. Additional, non-limiting examples of polypeptides that catalyze the conversion of propionyl-CoA to methylmalonyl-CoA are propionyl-CoA carboxylases from *Mycobacterium smegmatis*, *Homo sapiens*, *Acinetobacter baumannii*, *Brucella suis*, *Saccharopolyspora erythraea*, *Burkholderia glumae*, and *Aedes aegypti*, as well as the propionyl-CoA carboxylases set forth in Table A.

nucleotide encoding lactate dehydrogenase, lactate CoA transferase, lactyl-CoA dehydratase, and/or acrylyl-CoA reductase.

[0191] In addition, in any aspect of the invention, carbon flow to branch pathways not contributing to formation of the desired branched-chain fatty acid is minimized by attenuation of endogenous enzyme activity responsible for the diversion of carbon. Complete abolishment of endogenous activity is not required; any reduction in activity is suitable in the context of the invention. Enzyme activity is attenuated (i.e., reduced or abolished) by, for example, mutating the coding sequence for the enzyme to create a non-functional or reduced-function polypeptide, by removing all or part of the coding sequence for the enzyme from the cellular genome, by interfering with translation of an RNA transcript encoding the enzyme (e.g., using antisense oligonucleotides), or by manipulating the expression control sequences influencing expression of the enzyme. For example, in one aspect, the cell

TABLE A

Organism	GenBank Accession	Description	SEQ ID NO:
<i>Ehrlichia chaffeensis</i>	YP_507303	Propionyl-CoA carboxylase alpha subunit (PCCA)	51
<i>Ehrlichia chaffeensis</i>	YP_507410	Propionyl-CoA carboxylase beta subunit (PCCB)	52
<i>Agrobacterium vitis</i>	YP_002547482	Propionyl-CoA carboxylase alpha subunit (PCCA)	53
<i>Agrobacterium vitis</i>	YP_002547479	Propionyl-CoA carboxylase beta subunit (PCCB)	54
<i>Methylobacterium extorquens</i>	YP_003069256	Propionyl-CoA carboxylase alpha subunit (PCCA)	55
<i>Methylobacterium extorquens</i>	YP_003065890	Propionyl-CoA carboxylase beta subunit (PCCB)	56
<i>Sinorhizobium meliloti</i>	NP_437988	Propionyl-CoA carboxylase alpha subunit (PCCA)	57
<i>Sinorhizobium meliloti</i>	NP_437987	Propionyl-CoA carboxylase beta subunit (PCCB)	58
<i>Ruegeria pomeroyi</i>	YP_166352	Propionyl-CoA carboxylase alpha subunit (PCCA)	59
<i>Ruegeria pomeroyi</i>	YP_166345	Propionyl-CoA carboxylase beta subunit (PCCB)	60

[0189] Optionally, the cell is modified to increase carbon flow to propionyl-CoA (and then onward to methylmalonyl-CoA) by, for example, increasing expression of (i.e., overexpressing) *prpE* or other propionyl-CoA synthetase genes. Alternatively or in addition, an exogenous polynucleotide comprising a nucleic acid sequence encoding a propionyl-CoA synthetase is introduced into the host cell to upregulate propionyl-CoA production. Additionally, feeding host cells (e.g., microbes) large amounts of methionine, isoleucine, valine, threonine, propionic acid, and/or odd-chain length fatty acids (such as valeric acid) increases production of the propionyl-CoA precursor of methylmalonyl-CoA.

[0190] Methylmalonyl-CoA production via propionyl-CoA also is increased utilizing the metabolic pathway that converts pyruvate to propionyl-CoA, with lactate, lactoyl-CoA, and acrylyl-CoA as intermediates. Carbon flow to propionyl-CoA is upregulated by overproducing the enzymes of the pathway, producing exogenous enzymes catalyzing one or more conversions of the pathway, and/or by providing pyruvate or lactate in larger amounts than normally found in the host cell. For example, in any embodiment of the invention, the cell comprises an exogenous or overexpressed poly-

is modified to prevent methylmalonyl-CoA degradation, thereby increasing the amount of methylmalonyl-CoA available for conversion to methylmalonyl-ACP. Methylmalonyl-CoA degradation is reduced by, for example, deleting or inactivating methylmalonyl-CoA decarboxylase from the host. Put another way, the cell is modified to attenuate endogenous methylmalonyl-CoA decarboxylase activity. In *E. coli*, for example, methylmalonyl-CoA decarboxylase activity is attenuated by, for example, deleting or mutating *ygfG*. Optionally, endogenous acyl transferase activity is attenuated. Alternatively or in addition, methylmalonyl-CoA production within the cell is increased by preventing alternative metabolism of propionyl-CoA to succinyl-CoA, such as, for example, by deleting or otherwise reducing (attenuating) the activity of an endogenous methylmalonyl-CoA mutase gene. Optionally, methylmalonyl-CoA levels are increased by increasing the degradation of valine directly to methylmalonyl-CoA. Valine degradation comprises the following intermediates: α -ketoisovalerate, isobutyryl-CoA, methacrylyl-CoA, β -hydroxyisobutyryl-CoA, β -hydroxyisobutyrate, and methylmalonate semialdehyde. Optionally, methylmalonate semialdehyde is converted directly to methylmalonyl-CoA or

indirectly through a propionyl-CoA intermediate. In an exemplary embodiment, the cell of the invention comprises an overexpressed or exogenous polynucleotide comprising a nucleic acid sequence encoding one or more of the following enzymes: L-valine:2-oxoglutarate aminotransferase, 2-oxoisovalerate dehydrogenase, isobutyryl-CoA:FAD oxidoreductase, 3-hydroxy-isobutyryl-CoA hydro-lyase, 3-hydroxyisobutyryl-CoA hydrolase, 3-hydroxyisobutyrate dehydrogenase, and/or methylmalonate-semialdehyde dehydrogenase. Methylmalonate-semialdehyde dehydrogenase catalyzes the production of propanoyl-CoA, which can be converted to methylmalonyl-CoA by propanoyl-CoA carboxylase.

[0192] In one aspect, the cell comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of succinyl-CoA to methylmalonyl-CoA. An exemplary polypeptide that catalyzes the reaction is methylmalonyl-CoA mutase. In any embodiment of the invention, the cell is engineered to overexpress a methylmalonyl-CoA mutase gene, such as, for example, sbm (encoding Sleeping Beauty mutase) in *E. coli*. Alternatively or in addition, an exogenous polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA mutase is expressed in the cell. Exemplary methylmalonyl-CoA mutases include, but are not limited to, Sbm from *E. coli*, MutA and/or MutB from *Streptomyces cinnamonensis*, and methylmalonyl-CoA mutases from *Janibacter* sp. HTCC2649, *Corynebacterium glutamicum*, *Euglena gracilis*, *Homo sapiens*, *Propionibacterium shermanii*, *Bacillus megaterium*, and *Mycobacterium smegmatis*. Additional, non-limiting examples of polypeptides that catalyze the conversion of succinyl-CoA to methylmalonyl-CoA are provided in Table B.

[0194] Depending on the substrate specificity of the fatty acid synthase produced by the cell, a methylmalonyl-CoA epimerase also may be desired to facilitate use of methylmalonyl-CoA as a precursor in fatty acid synthesis. Thus, in one aspect, the cell further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA epimerase. Methylmalonyl-CoA epimerases suitable for use in the invention include, but are not limited to, *Sorangium cellulosum* So ce 56 methylmalonyl-CoA epimerase, *Streptomyces sviveus* ATCC 29083 methylmalonyl-CoA epimerase, *Kribbella flavida* DSM 17836 methylmalonyl-CoA epimerase, and methylmalonyl-CoA epimerases from *Homo sapiens*, *Bacillus megaterium*, and *Mycobacterium smegmatis*.

[0195] Production of branched-chain fatty acid comprising a methyl branch on one or more even number carbons also is enhanced by upregulating conversion of methylmalonyl-CoA to methylmalonyl-ACP. In one or more embodiments, conversion of methylmalonyl-CoA to methylmalonyl-ACP is increased in the cell by engineering the cell to produce an acyl transferase (such as the acyl transferase encoded by fabD in *E. coli*) to catalyze the formation of methylmalonyl-ACP from methylmalonyl-CoA. Put another way, in one aspect, the cell further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding an acyl transferase. Any suitable acyl transferase can be used, such as, for example and without limitation, an acyl transferase domain from a polyketide synthase, such as those involved in the synthesis of monensin, epothilone, amphotericin, candidin, nystatin, pimaricin, ascomycin, rapamycin, avermectin, spinosad, mycinamicin, niddamycin, oleandomycin, megalomicin, nanchangmycin, picromycin, rifamycin, oligomycin erythromycin, polyenes, and macrolides, and an acyl

TABLE B

Organism	GenBank Accession	Description	SEQ ID NO.
<i>Bacillus megaterium</i>	YP_003564880	methylmalonyl-CoA mutase small subunit (mutA)	61
<i>Bacillus megaterium</i>	YP_003564879	methylmalonyl-CoA mutase large subunit (mutB)	62
<i>Mycobacterium tuberculosis</i>	YP_001282809	methylmalonyl-CoA mutase small subunit (mutA)	63
<i>Mycobacterium tuberculosis</i>	YP_001282810	methylmalonyl-CoA mutase large subunit (mutB)	64
<i>Corynebacterium glutamicum</i>	YP_225814	methylmalonyl-CoA mutase small subunit (mutA)	65
<i>Corynebacterium glutamicum</i>	YP_225813	methylmalonyl-CoA mutase large subunit (mutB)	66
<i>Rhodococcus erythropolis</i>	YP_002766535	methylmalonyl-CoA mutase small subunit (mutA)	67
<i>Rhodococcus erythropolis</i>	YP_002766536	methylmalonyl-CoA mutase large subunit (mutB)	68
<i>Porphyromonas gingivalis</i>	NP_905776	methylmalonyl-CoA mutase small subunit (mutA)	69
<i>Porphyromonas gingivalis</i>	NP_905777	methylmalonyl-CoA mutase large subunit (mutB)	70

[0193] In one aspect, the cell comprises one or more polynucleotides encoding polypeptide(s) comprising an amino acid sequence at least about 80% identical (e.g., 85%, 90%, 95%, or 100% identical) to the amino acid sequences set forth in SEQ ID NO: 3, 4, and/or 28. The cell can comprise polynucleotides encoding a methylmalonyl-CoA mutase, a propionyl-CoA carboxylase, or both.

transferase domain from *Mycobacterium mycrocerosic* acid synthase. Acyl transferase domains from larger fatty acid synthase enzymes, such as *Mycobacterium mycrocerosic* acid synthase, act upon methylmalonyl-CoA in the absence of other enzymatic domains of the larger synthase. Optionally, the acyl transferase lacks polyketide synthesis activity. By "polyketide synthesis activity" is meant enzymatic activity,

other than acyl transferase activity, that catalyzes the production of polyketides in a host cell, such as, for example and without limitation, acyltransferase activity, ketoacyl synthase activity, ketoacyl reductase activity, dehydratase activity, enoyl reductase activity, acyl carrier protein activity, and thioesterase activity.

[0196] Alternatively, or in addition, in certain embodiments, a 3-ketoacyl-ACP synthase domain, such as, for example, a domain from a polyketide synthase or a mycoerotic acid synthase, is added to the fatty acid synthase of the host cell. In certain embodiments, the host cell (e.g., microbe) is engineered to include both acyl transferase and 3-ketoacyl-ACP synthase domains that can recognize methylmalonyl-CoA. In addition, in certain embodiments, genes for the endogenous acyl transferase and/or 3-ketoacyl-ACP synthase activities can be attenuated (e.g., deleted) to minimize the amount of malonyl-CoA incorporation in fatty acid synthesis.

[0197] In certain embodiments, the invention includes use of a thioesterase to specify the chain length of the fatty acid, such as, for example, to produce medium-chain fatty acids. In certain embodiments, the host cell further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a thioesterase. In one aspect, the host cell (e.g., bacteria) is engineered to produce a thioesterase that assists in the production of medium-chain branched-chain fatty acids. Alternatively, the host cell is engineered to produce (or overproduce) a thioesterase that assists in the production of long-chain branched-chain fatty acids. Exemplary thioesterases include, for example, the mallard uropygial gland thioesterase, the California bay thioesterase, the rat mammary gland thioesterase II, *E. coli* TesA, the *Cuphea wrightii* thioesterase, and other thioesterases suitable for production of the desired chain-length fatty acids.

[0198] Optionally, the cell is modified to produce (or increase the production of) branched acyl-CoA, which is a substrate for elongase in the production of long chain fatty acid. In this regard, in an exemplary embodiment of the invention, the cell comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid encoding a coenzyme-A synthetase, which converts branched-chain fatty acid to branched acyl-CoA. Examples of coenzyme-A synthetases include, but are not limited to, the coenzyme-A synthetase from *Leishmania braziliensis* (GenBank Accession No. XP_001561614), and the coenzyme-A synthetase from *Escherichia coli* (GenBank Accession No. YP_541006). Optionally, the cell comprises exogenous or overexpressed polynucleotide(s) comprising a nucleic acid sequence encoding an elongase to increase the length of the carbon backbone. Elongases are enzyme complexes that exhibit 3-ketoacyl-CoA synthase, 3-ketoacyl-CoA reductase, 3-hydroxyacyl-CoA dehydratase, and enoyl-CoA reductase activities, and generally utilize malonyl-CoA as an extension unit for extending the carbon chain. When a methyl-malonyl CoA is used as an extension unit by the enzyme complex, additional methyl branches are introduced at even carbon positions. Exemplary elongases include, but are not limited to, elongases comprising the one or more of the following subunits: *Saccharomyces cerevisiae* 3-ketoacyl-CoA synthase (GenBank Accession No. NP_013476), 3-ketoacyl-CoA reductase (GenBank Accession No. NP_009717), 3-hydroxyacyl-CoA dehydratase (GenBank Accession No. NP_012438) and enoyl-CoA reductase (GenBank Accession No. NP_010269); and *Arabidopsis thaliana* col 3-ketoacyl-CoA synthase (GenBank Accession No. NP_849861), 3-ketoacyl-

CoA reductase (GenBank Accession No. NP_564905), 3-hydroxyacyl-CoA dehydratase (GenBank Accession No. NP_193180), and enoyl-CoA reductase (GenBank Accession No. NP_191096).

[0199] Any suitable cell or organism, such as, for example, bacterial cells and other prokaryotic cells, and yeast cells, can be used in the context of the invention. In one aspect, the invention relates to cells, such as *Escherichia* cells (e.g., *E. coli*), which naturally produce Type II fatty acid synthase and/or do not naturally produce scattered branched-chain fatty acid (i.e., branched-chain fatty acid comprising a methyl branch on one or more even numbered carbons). These cells are engineered to produce the branched-chain fatty acids as described herein. Alternatively, the cell naturally produces branched-chain fatty acid and is modified as described herein to produce higher levels of branched-chain fatty acid (or different proportions of different types of branched-chain fatty acid) compared to an unmodified cell. In certain embodiments, fatty acid is manufactured using bacteria known to make the methylmalonyl-CoA precursor, such as *Streptomyces*, *Mycobacterium* or *Corynebacterium*. These bacteria are, in one aspect, engineered to produce (i) an acyl transferase to increase carbon flux to methylmalonyl-ACP that is incorporated in the fatty acid synthesis pathway and/or (ii) a thioesterase to control the chain length.

[0200] Exemplary bacteria that are suitable for use in the invention include, but are not limited to, *Spirochaeta aurantia*, *Spirochaeta littoralis*, *Pseudomonas maltophilia*, *Pseudomonas putrefaciens*, *Xanthomonas campestris*, *Legionella anisa*, *Moraxella catarrhalis*, *Thermus aquaticus*, *Flavobacterium aquatile*, *Bacteroides asaccharolyticus*, *Bacteroides fragilis*, *Succinimonas amylolytica*, *Desulfovibrio africanus*, *Micrococcus agilis*, *Stomatococcus mucilaginosus*, *Planococcus citreus*, *Marinococcus albus*, *Staphylococcus aureus*, *Peptostreptococcus anaerobius*, *Ruminococcus albus*, *Sarcina lutea*, *Sporolactobacillus inulinus*, *Clostridium thermocellum*, *Sporosarcina ureae*, *Desulfotomaculum nigrificans*, *Listeria monocytogenes*, *Brochothrix thermosphacta*, *Renibacterium salmoninarum*, *Kurthia zopfii*, *Corynebacterium aquaticum*, *Arthrobacter radiotolerans*, *Brevibacterium fermentans*, *Propionibacterium acidipropionici*, *Eubacterium lentum*, *Cytophaga aquatilis*, *Sphingobacterium multivorumb*, *Capnocytophaga gingivalis*, *Sporocytophaga myxococcoides*, *Flexibacter elegans*, *Myxococcus coralloides*, *Archangium gephyra*, *Stigmatella aurantia*, *Oerskovia turbata*, *Escherichia coli*, *Bacillus subtilis*, *Salmonella typhimurium*, *Corynebacterium glutamicum*, *Streptomyces coelicolor*, *Streptomyces lividans*, *Clostridium thermocellum* and *Saccharomonospora viridis*.

[0201] In one aspect, the fatty acid produced by the inventive cell comprises about 80% to about 100% (wt.) (e.g., about 85%, about 90%, or about 95%) linear and branched-chain fatty acid. Of the linear and branched-chain fatty acids produced by the cell, approximately 1% to approximately 95% or more (e.g., 5%, 10%, 15%, 20%, 30%, 50%, 60%, 75%, 85%, or 100%) is branched-chain fatty acid comprising a methyl group on one or more even carbons. In some embodiments, the cell does not produce, or produces only trace amounts of, fatty acid comprising methyl branching on odd numbered carbons. By "trace amount" is meant less than 1% of the total fatty acid content produced by the cell. Alternatively or in addition, in one aspect, the mixture of fatty acids produced by the cell comprises no more than 50% end-terminal-branched fatty acid (i.e., fatty acids that contain branch-

ing on a carbon atom that is within 40% of the non-functionalized terminus of the longest carbon chain). Optionally, the inventive cell is modified to preferentially produce branched-chain fatty acid with desired chain lengths, e.g., about six to about 18 carbons or more in the carbon backbone (not including the methyl branch(es)). In some embodiments, the host cell preferentially generates long chain fatty acid, medium-length chain fatty acid, short chain fatty acid, or a desired combination fatty acids (e.g., 60%, 70%, 80%, 85%, 90%, 95% or more of the branched-chain fatty acid produced by the cell comprises the desired number of carbons). In addition, in certain embodiments, the engineered cells tolerate large amounts of branched-chain fatty acid in the growth medium, plasma membrane, or lipid droplets, and/or produce branched-chain fatty acid more economically than an unmodified cell by, e.g., using a less expensive feedstock, requiring less fermentation time, and the like.

[0202] The polynucleotide(s) encoding one or more polypeptides that catalyze the reaction(s) for producing branched-chain fatty acid may be derived from any source. Depending on the embodiment of the invention, the polynucleotide is isolated from a natural source such as bacteria, algae, fungi, plants, or animals; produced via a semi-synthetic route (e.g., the nucleic acid sequence of a polynucleotide is codon-optimized for expression in a particular host cell, such as *E. coli*); or synthesized de novo. In certain embodiments, it is advantageous to select an enzyme from a particular source based on, e.g., the substrate specificity of the enzyme, the type of branched-chain fatty acid produced by the source, or the level of enzyme activity in a given host cell. In one aspect of the invention, the enzyme and corresponding polynucleotide are naturally found in the host cell and overexpression of the polynucleotide is desired. In this regard, in some instances, additional copies of the polynucleotide are introduced in the host cell to increase the amount of enzyme available for fatty acid production. Overexpression of a native polynucleotide also is achieved by upregulating endogenous promoter activity, or operably linking the polynucleotide to a more robust promoter. Exogenous enzymes and their corresponding polynucleotides also are suitable for use in the context of the invention, and the features of the biosynthesis pathway or end product can be tailored depending on the particular enzyme used. If desired, the polynucleotide(s) is isolated or derived from the branched-chain fatty acid-producing organisms described herein.

[0203] In certain embodiments, the cell produces an analog or variant of a polypeptide described herein. Amino acid sequence variants of the polypeptide include substitution, insertion, or deletion variants, and variants may be substantially homologous or substantially identical to the unmodified polypeptides as set out above. In certain embodiments, the variants retain at least some of the biological activity, e.g., catalytic activity, of the polypeptide. Other variants include variants of the polypeptide that retain at least about 50%, preferably at least about 75%, more preferably at least about 90%, of the biological activity.

[0204] Substitution variants typically exchange one amino acid for another at one or more sites within the protein. Substitutions of this kind can be conservative, that is, one amino acid is replaced with one of similar shape and charge. Conservative substitutions include, for example, the changes of: alanine to serine; arginine to lysine; asparagine to glutamine; aspartate to glutamate; cysteine to serine; glutamine to asparagine; glutamate to aspartate; isoleucine to

leucine or valine; leucine to valine or isoleucine; lysine to arginine; methionine to leucine or isoleucine; phenylalanine to tyrosine, leucine or methionine; serine to threonine; threonine to serine; tryptophan to tyrosine; tyrosine to tryptophan or phenylalanine; and valine to isoleucine or leucine.

[0205] In some instances, the recombinant cell comprises an analog or variant of the exogenous or overexpressed polynucleotide(s) described herein. Nucleic acid sequence variants include one or more substitutions, insertions, or deletions, and variants may be substantially homologous or substantially identical to the unmodified polynucleotide. Polynucleotide variants or analogs encode mutant enzymes having at least partial activity of the unmodified enzyme. Alternatively, polynucleotide variants or analogs encode the same amino acid sequence as the unmodified polynucleotide. Codon-optimized sequences, for example, generally encode the same amino acid sequence as the parent/native sequence but contain codons that are preferentially expressed in a particular host organism.

[0206] A polypeptide or polynucleotide “derived from” an organism contains one or more modifications to the native amino acid sequence or nucleotide sequence and exhibits similar, if not better, activity compared to the native enzyme (e.g., at least 70%, at least 80%, at least 90%, at least 95%, at least 100%, or at least 110% the level of activity of the native enzyme). For example, enzyme activity is improved in some contexts by directed evolution of a parent/native sequence. Additionally or alternatively, an enzyme coding sequence is mutated to achieve feedback resistance. Thus, in one or more embodiments of the invention, the polypeptide encoded by the exogenous polynucleotide is feedback resistant and/or is modified to alter the activity of the native enzyme. A polynucleotide “derived from” a reference polynucleotide encompasses, but is not limited to, a polynucleotide comprising a nucleic acid sequence that has been codon-optimized for expression in a desired host cell.

[0207] The cell of the invention may comprise any combination of polynucleotides described herein to produce branched-chain fatty acid comprising a methyl branch on one or more even number carbons. For example, the invention provides a cell comprising (i) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding an acyl transferase lacking polyketide synthesis activity, and (ii) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a propionyl-CoA carboxylase and/or an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA mutase, wherein the polynucleotide(s) are expressed and the cell produces more branched-chain fatty acid comprising a methyl on one or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s). Recombinant cells can be produced in any suitable manner to establish an expression vector within the cell. The expression vector can include the exogenous polynucleotide operably linked to expression elements, such as, for example, promoters, enhancers, ribosome binding sites, operators and activating sequences. Such expression elements may be regulatable, for example, inducible (via the addition of an inducer). Alternatively or in addition, the expression vector can include additional copies of a polynucleotide encoding a native gene product operably linked to expression elements. Representative examples of useful promoters include, but are not limited to: the LTR (long terminal 35 repeat from a retrovirus) or SV40 promoter, the *E. coli* lac,

tet, or trp promoter, the phage Lambda P_L promoter, and other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses. In one aspect, the expression vector also includes appropriate sequences for amplifying expression. The expression vector can comprise elements to facilitate incorporation of polynucleotides into the cellular genome. Introduction of the expression vector or other polynucleotides into cells can be performed using any suitable method, such as, for example, transformation, electroporation, microinjection, microprojectile bombardment, calcium phosphate precipitation, modified calcium phosphate precipitation, cationic lipid treatment, photoporation, fusion methodologies, receptor mediated transfer, or polybrene precipitation. Alternatively, the expression vector or other polynucleotides can be introduced by infection with a viral vector, by conjugation, by transduction, or by other any other suitable method.

[0208] Cells, such as bacterial cells, containing the polynucleotides encoding the proteins described herein can be cultured under conditions appropriate for growth of the cells and expression of the polynucleotides. Cells expressing the protein can be identified by any suitable methods, such as, for example, by PCR screening, screening by Southern blot analysis, or screening for the expression of the protein. In certain embodiments, cells that contain the polynucleotide(s) can be selected by including a selectable marker in the DNA construct, with subsequent culturing of cells containing a selectable marker gene, under conditions appropriate for survival of only those cells that express the selectable marker gene. The introduced DNA construct can be further amplified by culturing genetically modified cells under appropriate conditions (e.g., culturing genetically modified cells containing an amplifiable marker gene in the presence of a concentration of a drug at which only cells containing multiple copies of the amplifiable marker gene can survive). Cells that contain and express polynucleotides encoding the exogenous proteins can be referred to herein as genetically modified cells. Bacterial cells that contain and express polynucleotides encoding the exogenous protein can be referred to as genetically modified bacterial cells.

[0209] Exemplary cells of the invention include *E. coli* BW25113 comprising pTrcHisA mmat and pZA31-accA1-pccB, which was deposited with American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, Va., on Dec. 14, 2010, under the provisions of the Budapest Treaty for the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure ("Budapest Treaty"), and assigned Deposit Accession No. [XXX] on [DATE], and *E. coli* BL21 Star (DE3) comprising pTrcHisA Ec sbm So ce epi and pZA31 mmat which was deposited with American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, Va., on Dec. 14, 2010, under the provisions of the Budapest Treaty for the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure ("Budapest Treaty"), and assigned Deposit Accession No. [XXX] on [DATE]. The invention also includes variants or progeny of the cells described herein that retain the phenotypic characteristics of the recombinant microbe. A substantially pure monoculture of the cell described herein (i.e., a culture comprising at least 80% or at least 90% of a desired cell) also is provided.

[0210] Any cell culture conditions appropriate for growing a host cell and synthesizing branched-chain fatty acid is suitable for use in the inventive method. Addition of fatty acid

synthesis intermediates, precursors, and/or co-factors for the enzymes associated with branched-chain fatty acid synthesis to the culture is contemplated herein. In certain embodiments, the genetically modified cells (such as genetically modified bacterial cells) have an optimal temperature for growth, such as, for example, a lower temperature than normally encountered for growth and/or fermentation. For example, in certain embodiments, incorporation of branched-chain fatty acids into the membrane may increase membrane fluidity, a property normally associated with low growth temperatures. In addition, in certain embodiments, cells of the invention may exhibit a decline in growth at higher temperatures as compared to normal growth and/or fermentation temperatures as typically found in cells of the type.

[0211] The inventive method optionally comprises extracting branched-chain fatty acid from the culture. Fatty acids can be extracted from the culture medium and measured using any suitable manner. Suitable extraction methods include, for example, methods as described in: Bligh et al., A rapid method for total lipid extraction and purification, *Can. J. Biochem. Physiol.* 37:911-917 (1959). In certain embodiments, production of fatty acids in the culture supernatant or in the membrane fraction of recombinant cells can be measured. In this embodiment, cultures are prepared in the standard manner, although nutrients (e.g., 2-methylbutyrate, isoleucine) that may provide a boost in substrate supply can be added to the culture. Cells are harvested by centrifugation, acidified with hydrochloric or perchloric acid, and extracted with chloroform and methanol, with the fatty acids entering the organic layer. The fatty acids are converted to methyl esters, using methanol at 100° C. The methyl esters are separated by gas chromatography (GC) and compared with known standards of fatty acids (purchased from Larodan or Sigma). Confirmation of chemical identity is carried out by combined GC/mass spec, with further mass spec analysis of fragmented material carried out if necessary.

[0212] In one embodiment, the cell utilizes the branched-chain fatty acid as a precursor to make one or more other products. Products biosynthesized (i.e., derived) from branched-chain fatty acid include, but are not limited to, phospholipids, triglycerides, alkanes, olefins, wax esters, fatty alcohols, and fatty aldehydes. Some host cells naturally generate one or more products derived from branched-chain fatty acid; other host cells are genetically engineered to convert branched-chain fatty acid to, e.g., an alkane, olefin, wax ester, fatty alcohol, phospholipid, triglyceride, and/or fatty aldehyde. Organisms and genetic modifications thereof to synthesize products derived from branched-chain fatty acids are further described in, e.g., International Patent Publication Nos. WO 2007/136762, WO 2008/151149, and WO 2010/062480, and U.S. Patent Application Publication US 2010/0298612, all of which are hereby incorporated by reference in their entirety. In one aspect, the inventive method comprises extracting a product derived from branched-chain fatty acid (phospholipid, triglyceride, alkane, olefin, wax ester, fatty alcohol, and/or fatty aldehyde synthesized in the cell from branched-chain fatty acid) from the culture. Any extraction method is appropriate, including the extraction methods described in International Patent Publication Nos. WO 2007/136762, WO 2008/151149, and WO 2010/062480, and U.S. Patent Application Publication Nos. US 2010/0251601, US 20100242345, US 20100105963, and US 2010/0298612.

[0213] The inventive cell preferably produces more branched-chain fatty acid comprising a methyl branch on one

or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s). Methods of measuring fatty acid released into the fermentation broth or culture media or liberated from cellular fractions are described herein. Branched-chain fatty acid production is not limited to fatty acid accumulated in the culture, however, but also includes fatty acid used as a precursor for downstream reactions yielding products derived from branched-chain fatty acid. Thus, products derived from branched-chain fatty acid (e.g., phospholipids, triglycerides, fatty alcohols, olefins, wax esters, fatty aldehydes, and alkanes) are, in some embodiments, surrogates for measuring branched-chain fatty acid production in a host cell. Methods of measuring fatty acid content in phospholipid in the cell membrane are described herein. Similarly, measurement of degradation products of branched-chain fatty acids also is instructive as to the amount of branched-chain fatty acid is produced in a host cell. Depending on the particular embodiment of the invention, the inventive cell produces at least 3%, at least 5%, at least 10%, at least 20%, at least 25%, or at least 50% more branched-chain fatty acid than an otherwise similar cell that does not comprise the polynucleotide(s).

[0214] The invention further provides a composition comprising the branched-chain fatty acids described herein. For example, the invention provides a composition comprising a branched-chain fatty acid comprising between 10-18 carbons in the carbon backbone, such as fatty acids comprising between 10 and 16 carbons (e.g., fatty acids comprising 10, 11, 12, 13, 14, 15, or 16 carbons), with branching on one or more even numbered carbons (e.g., C2, C4, C6, C8, C10, C12, C14, and/or C16). A composition comprising longer-chain fatty acid also is provided, such as a composition comprising between 19 and 22 carbons in the longest carbon chain. A composition comprising a combination of any of the fatty acids described herein also is provided (e.g., a composition comprising fatty acids of varying lengths and/or branch locations along the carbon backbone).

[0215] The following examples further describe and demonstrate embodiments within the scope of the invention. The examples are given solely for the purpose of illustration and are not to be construed as limitations of the invention, as many variations thereof are possible without departing from the spirit and scope of the invention.

Example 1

Construction of Methylmalonyl-CoA Mutase Expression Vector

[0216] There are numerous genes annotated to encode the two subunits of methylmalonyl-CoA mutase. *Janibacter* sp. HTCC2649 encodes two such genes. Synthetic versions of these genes were prepared, with the codon usage altered to match that used by many *E. coli* genes (i.e., the coding sequence was codon-optimized for expression in *E. coli*). By analogy to other methylmalonyl-CoA mutase genes, these synthetic genes were named mutA (SEQ ID NO: 1) and mutB (SEQ ID NO: 2), corresponding to the MutA (SEQ ID NO: 3) and MutB (SEQ ID NO: 4) protein subunits. In the synthetic DNA, an extra three base pairs were added (encoding an alanine residue immediately after the initiation methionine) in mutA to facilitate introduction of an NcoI site. An XhoI restriction site was also placed after the coding sequence of mutB for insertion into the pBAD vector (Invitrogen). The NcoI/XhoI fragment was cloned into pBAD.

Example 2

Construction of Methylmalonyl-CoA Epimerase Expression Vector

[0217] There are numerous genes annotated to encode methylmalonyl-CoA mutase. One such gene is from *Streptomyces sviveus*. A synthetic gene can be constructed (SEQ ID NO: 5) using codon usage similar to *E. coli* genes and with EcoRI and Hind III sites flanking the coding region. An *E. coli* Shine-Dalgarno sequence can be added between the EcoRI site and the initiation codon for the epimerase gene. The predicted protein product is the same as the predicted protein product from the *S. sviveus* gene (SEQ ID NO: 6). The epimerase gene can be cloned into the pBAD-mutAB construct using the EcoRI and Hind III restriction sites (downstream of mutB) to form the pBAD-mutAB-epimerase gene plasmid. *E. coli* cultures can be grown at 27° C. after induction with arabinose and supplemented with hydroxycobalamin to achieve expression of functional methylmalonyl-CoA mutase and branched-chain fatty acid production.

Example 3

Construction of Propionyl-CoA Carboxylase Expression Vector

[0218] Nucleotide sequences (SEQ ID NO: 7 and SEQ ID NO: 8) encoding the two propionyl-CoA carboxylase subunits AccA1 (GenBank Accession NO. AF113603.1; SEQ ID NO: 9) and PccB (GenBank Accession No. AF113605.1; SEQ ID NO: 10)), respectively, from the *Streptomyces coelicolor* A3(2) propionyl-CoA carboxylase (Rodriguez E., Gramajo H., *Microbiology*. 1999 November; 145:3109-19), were codon-optimized for *E. coli* expression. A gene construct for expressing propionyl-CoA carboxylase was constructed with the following elements sequentially 1) P_{Llac0-1} promoter and operator plus T7 gene10 ribosomal binding site (SEQ ID NO: 11); 2) optimized accA1 (SEQ ID NO: 12); 3) three restriction site sequences including BglII, NotI and XbaI and a T7 gene10 ribosome binding site (SEQ ID NO: 13); and 4) codon-optimized pccB (SEQ ID NO: 14). The synthesized DNA fragments were cloned into the XhoI and PstI sites of expression vector pZA31-MCS (Expressys, Ruelzheim, Germany), resulting in plasmid pZA31-accA1-pccB (SEQ ID NO: 15).

Example 4

Construction of Propionyl-CoA Synthetase Expression Vector

[0219] The *Salmonella enterica* propionyl-CoA synthetase gene, prpE, was amplified using PCR and the primers set forth in SEQ ID NO: 16 and SEQ ID NO: 17, and placed behind a Shine-Dalgarno sequence in the plasmid pZA31-accA1-pccB (SEQ ID NO: 15) using the restriction enzymes PstI and BamHI. Enhanced propionyl-CoA synthetase production is expected to increase synthetic flux to propionyl-CoA.

Example 5

Reduction of Propionylation of Propionyl-CoA Synthetase

[0220] In *S. enterica*, propionyl-CoA synthetase is subject to inhibition by propionylation at lysine 592 when propionyl-CoA levels accumulate. (Garrity et al, *J. Biol. Chem.*, Vol.

282, Issue 41, 30239-30245, Oct. 12, 2007). Similar enzyme modulation may occur in other species, although the position of the modified lysine may be different. Several strategies to overcome this inhibition will be tested and compared. First, the propionyl-CoA synthetase gene will be mutated to change the coding capacity from lysine (at the site of propionylation) to arginine or other amino acids to prevent propionylation. Second, a source of resveratrol or other sirtuin activators will be introduced into the culture medium to activate sirtuin to depropionylate PrpE. Third, the endogenous N-acetyltransferase enzyme responsible for the propionylation reaction will be knocked out. For example, if working with *S. enterica*, pat could be deleted. As another example, if working with *B. subtilis*, acuA could be deleted. Fourth, the flux of propionyl-CoA into fatty acid synthesis will be increased by increasing propionyl-CoA carboxylase activity to keep free propionyl-CoA levels down. Fifth, the sirtuin activity will be increased, thus increasing deacetylation of propionyl-CoA carboxylase. For example, the *S. enterica* cobB expression could be increased.

Example 6

Creation of an Expression Vector Comprising the Coding Sequence of the MMAT (Methylmalonyl-CoA Acyl Transferase) Domain from *Mycobacterium Mycocerosic Acid Synthase (MAS)*.

[0221] *Mycobacterium* MAS is a multifunctional protein that catalyzes the synthesis of mycocerosic acid and that contains a domain with MMAT activity. The MMAT domain (amino acids 508-890) (SEQ ID NO: 18) of MAS from *Mycobacterium bovis* BCG (YP_979046) (SEQ ID NO: 19) was codon optimized for *E. coli* expression (SEQ ID NO: 20). The optimized sequence was synthesized and cloned into vector pTrcHisA (Invitrogen) between the BamHI and HindIII sites. The resulting construct fused the MMAT domain with the His tag leader peptide encoded by the vector. The expression vector was introduced into a recombinant *E. coli* host that produces methylmalonyl-CoA. MMAT activity catalyzes the formation of methylmalonyl-ACP, which subsequently can be incorporated into the type II fatty acid synthesis pathway to form methyl branches at even positions of the fatty acid chain.

Example 7

Method for Detecting Acyl-CoA

[0222] This example describes an exemplary method for detecting and quantifying an acyl-CoA (e.g., methylmalonyl-CoA) in a sample, such as a sample of recombinant host cells producing branched-chain fatty acid.

[0223] A stable, labeled (deuterium) internal standard-containing master mix was prepared comprising d₃-3-hydroxymethylglutaryl-CoA (200 µl of 50 µg/ml stock in 10 ml of 15% trichloroacetic acid). An aliquot (500 µl) of the master mix was added to a 2 ml tube. Silicone oil (AR200; Sigma catalog number 85419; 800 µl) was layered onto the master mix. An *E. coli* culture (800 µl) was layered gently on top of the silicone oil, and the resulting sample was subjected to centrifugation at 20,000×g for five minutes at 4° C. in an Eppendorf 5417 C centrifuge. A portion (300 µl) of the master mix-containing layer was transferred to an empty tube and frozen on dry ice for 30 minutes.

[0224] The acyl-CoA content of samples was determined using HPLC/MS/MS. Individual coenzyme-A standards

(propionyl-CoA, methylmalonyl-CoA, succinyl-CoA, malonyl-CoA, isobutyryl-CoA, isovaleryl-CoA, and acetyl-CoA) were purchased from Sigma Chemical Company (St. Louis, Mo.) and prepared as 500 µg/ml stocks in methanol. The analytes were pooled, and standards with all of the analytes were prepared by dilution with 15% trichloroacetic acid. Standards for regression were prepared by transferring 500 µl of the working standards to an autosampler vial containing 10 µl of the 50 µg/ml internal standard. Sample peak areas (or heights) were normalized to the stable-labeled internal standard (d₃-3-hydroxymethylglutaryl-CoA, Cayman Chemical Co.). Samples were assayed by HPLC/MS/MS on a Sciex API5000 mass spectrometer in positive ion Turbo Ion Spray. Separation was carried out by reversed-phase high performance liquid chromatography using a Phenomenex Onyx Monolithic C18 column (2×50 mm) and mobile phases of (1) 5 mM ammonium acetate, 5 mM dimethylbutylamine, 6.5 mM acetic acid and (2) acetonitrile with 0.1% formic acid, with the gradient set forth in Table C.

TABLE C

Time	Mobile Phase A (%)	Mobile Phase B (%)
0 min	97.5	2.5
1.0 min	97.5	2.5
2.5 min	91.0	9.0
5.5 min	45	55
6.0 min	45	55
6.1 min	97.5	2.5
7.5 min	—	—
9.5 min	End Run	—

[0225] The conditions on the mass spectrometer were: DP 160, CUR 30, GS1 65, GS2 65, IS 4500, CAD 7, TEMP 650 C. The transitions set forth in Table D were used for the multiple reaction monitoring (MRM).

TABLE D

Compound	Precursor Ion*	Product Ion*	Collision Energy	CXP
n-Propionyl-CoA	824.3	317.2	41	32
Methylmalonyl-CoA	868.1	317.1	42	31
Succinyl-CoA	868.2	361.1	49	38
Malonyl-CoA	854.2	347.2	41	36
Isobutyryl-CoA	838.3	345.2	45	34
Isovaleryl-CoA	852.2	345.2	45	34
Acetyl-CoA	810.3	303.2	43	30
d ₃ -3-Hydroxymethylglutaryl-CoA	915.2	408.2	49	13

*Energy (Volts) for MS/MS analysis

Example 8

Analysis of Fatty Acids Produced by Host Cells

[0226] This example illustrates a method of analyzing branched-chain fatty acids produced by cells (e.g., recombinant microbes).

[0227] Cell cultures (approximately 1.5 ml) were frozen in 2.0 ml glass vials and stored at -20° C. until ready for processing. Samples were chilled on dry ice for 30 minutes and lyophilized overnight (-16 hours) until dry. A 10 µl aliquot of internal standard (glyceryl trionadecanoate (Sigma catalog number T4632-1G)) was added to each vial, followed by 400

μL of 0.5 N NaOH (in methanol). The vial was capped and vortexed for 10 seconds. Samples were incubated at 65°C. for 30-50 minutes. Samples were then removed from the incubator, and 500 μL of boron trifluoride reagent (Aldrich catalog number B1252) was added. The samples were vortexed again for 10 seconds, incubated at 65°C. for 10-15 minutes, and cooled to room temperature (approximately 20 minutes). Hexane (350 μL) was added, and the samples were again vortexed for 10 seconds. If the phases did not separate, 50-100 μL of saturated salt solution (5 g NaCl to 5 ml water) was added, and the sample was vortexed for 10 seconds. At least 100 μL of the top hexane layer was placed into the gas chromatography vial. The vial was capped and stored at 4°C. until analyzed by gas chromatography.

[0228] Gas chromatography was performed as described in Table E below. A bacterial acid methyl ester standard (Sigma catalog number 47080-U) and a fatty acid methyl ester standard (Sigma catalog number 47885-U) were used to identify peaks in samples. A sample check standard using glyceryl tripalmitate (Sigma catalog number T5888-1G) was used to confirm esterification of samples. A blank standard (internal standard only) was used to assess background noise.

TABLE E

Gas Chromatograph	HP 5890 GC Series II				
Detector	FID 360° C. 40 ml/min Hydrogen, 400 ml/min Air				
Carrier Gas	Helium				
Quantitative Program	GC Chemstation A.09.03. (Agilent)				
Column	VF-5 ms 15 M \times 0.150 mm \times 0.15 μm Varian catalog number CP9035				
Injection Liner	Gooseneck (with glass wool packing)				
Injector	HP 7673				
Injection Syringe	10 μL				
Injection Mode	Split 25:1				
Injection volume	4 μL (Plunger Speed = fast; 5 sample pumps)				
Pre Injection Solvent Washes	2 samples				
Post Injection Solvent Washes	3 for both acetone and hexane				
Injector Temperature	325° C.				
Total Program Time	16 minutes				
	Initial Temp. (° C.)	Initial Time (min)	Rate (° C./min)	Final Temp (° C.)	Final Time (min)
Thermal Program	90	0.75	20.0 25.0	325 350	1.0 2.5

Example 9

Construction of Expression Vectors Comprising *S. Cinnamomensis* mutA and mutB and *S. sviveus* epi.

[0229] A synthetic DNA construct was generated comprising *Streptomyces cinnamomensis* mutA (SEQ ID NO: 24) (GenBank Accession No. AAA03040.1), *S. cinnamomensis* mutB (SEQ ID NO: 25) (GenBank Accession No. AAA03041.1), and a *Streptomyces sviveus* ATCC 29083 methylmalonyl-CoA epimerase gene (SEQ ID NO: 26) (GenBank Accession No. ZP_06919825.1). The genes were codon-optimized for expression in *E. coli*. An EcoRI restriction site was placed on the 5' end, and a BamHI site was placed on the 3' end of the synthesized gene construct. These sites were subsequently used for cloning into a pZA31 vector (Expressys, Ruelzheim, Germany). A ribosome binding

sequence and spacer was placed before the mutA and epimerase gene start codons (SEQ ID NO: 27). The plasmid was designated pZA31 mutAB Ss epi.

Example 10

Construction of Expression Vectors Comprising Sbm and malE/sbm Polynucleotides

[0230] Sleeping beauty mutase (Sbm) (also known as methylmalonyl-CoA mutase (MCM)) is an enzyme that catalyzes the rearrangement of succinyl-CoA to L-methylmalonyl-CoA. The enzyme is vitamin B12 (cobalamin) dependent. Methylmalonyl-CoA is a building block for scattered branch-chain fatty acids (SBCFA) (i.e., branched-chain fatty acid comprising a methyl branch on one or more even number carbons of the fatty acid backbone). Plasmids comprising a polynucleotide encoding Sbm were generated to introduce multiple copies of the Sbm coding sequence, downstream of a regulatable promoter, into *E. coli* host cells.

[0231] A polynucleotide was synthesized based on the sequence of *E. coli* sbm (SEQ ID NO: 28) (GenBank Accession No. NP_417392.1) from *E. coli* strain MG1655. The nucleic acid sequence was codon-optimized to match the pattern of highly expressed *E. coli* genes while maintaining the native amino acid sequence of the enzyme. The generated nucleic acid sequence is set forth in SEQ ID NO: 29. A BamHI and an XbaI site were added at the 5' end of the synthetic Sbm coding sequence with the sequence GGATC-CATGTCCTAGA (SEQ ID NO: 49) adjacent to the ATG translation initiation sequence. A SacI restriction site sequence was added to the 3' end of the synthetic Sbm coding sequence. The gene was synthesized, cloned into a pUC57 vector, and sequenced (GenScript, Piscataway, N.J.). The synthetic sbm was then released from pUC57 by restriction enzymes BamHI and Sad, and sub-cloned into plasmid pTrcHisA (Invitrogen, Carlsbad, Calif.) in frame with the poly-histidine sequence (GenScript, Piscataway, N.J.). The plasmid was designated pTrcHisA Ec sbm. The sequence was confirmed by sequencing (GenScript, Piscataway, N.J.). The recombinant protein encoded by the sequence contained a poly-histidine sequence (Met-Gly-Gly-Ser-His-His-His-His-His-His-Gly-Met-Ala-Ser-Met-Thr-Gly-Gly-Gln-Gln-Met-Gly-Arg-Thr-Asp-Asp-Asp-Asp-Lys-Asp-Arg-Trp-Gly-Ser (SEQ ID NO: 50)) and a full-length native Sbm amino acid sequence.

[0232] A recombinant methylmalonyl-CoA mutase has been reported to be insoluble in *E. coli* (Korotkova, N., and M. E. Lidstrom. *J. Biological Chemistry* 279: 13652-8 (2004)). Translation fusion with maltose-binding protein (MBP, encoded by malE) prevents aggregation of recombinant proteins (Kapust, R. B., and D. S. Waugh. *Protein Science* 8: 1668-74 (1999)). A recombinant construct was generated by inserting malE upstream of sbm. The malE polynucleotide was synthesized based on the sequence of maltose binding protein (*E. coli* MG1655 GenBank NC_000913.2 (GenScript, Piscataway, N.J.)). A BamHI site was placed adjacent to the translation initiation codon of malE, and an XbaI site was placed immediately 5' to the stop codon of the malE sequence (SEQ ID NO: 30). Also, one nucleotide was changed (T438 to C438) to remove a restriction site recognition sequence for BglII.

[0233] The MalE coding sequence (SEQ ID NO: 30) was first synthesized and cloned into a pUC57 plasmid. After confirming its sequence, the malE polynucleotide was released using restriction enzymes BamHI and XbaI. The

released male was then re-cloned into plasmid pTrcHisA Ec sbm at BamHI and XbaI sites (GenScript, Piscataway, N.J.). The resulting plasmid was designated pTrcHisA Ec male Ec sbm. The recombinant protein encoded by pTrcHisA Ec male Ec sbm contains three peptides: the poly-histidine tag, full-length MBP, and full-length Sbm.

Example 11

Construction of a Recombinant Expression Vector Comprising a Polynucleotide Encoding the Methylmalonyl-CoA Acyl Transferase (MMAT) Domain from *Mycobacterium Mycrocercosic Acid Synthase* (MAS).

[0234] *Mycobacterium MAS* is a multifunctional protein containing MMAT activity that catalyzes the synthesis of mycrocercosic acid. The nucleic acid sequence encoding the MMAT domain (amino acids 508-890) (SEQ ID NO: 18) of MAS from *Mycobacterium bovis* BCG (GenBank Accession No. YP_979046) (SEQ ID NO: 19) was codon-optimized for *E. coli* expression (SEQ ID NO: 20). The optimized sequence, designated "mmat," was synthesized and cloned into vector pTrcHisA (Invitrogen) between the BamHI and HindIII sites. The resulting construct fused the MMAT domain with the poly-histidine tag encoded by the vector. The expression vector (pTrcHisA mmat) was introduced into a recombinant *E. coli* host that produces methylmalonyl-CoA. MMAT activity catalyzes the formation of methylmalonyl-ACP, which is incorporated by Type II fatty acid synthase into fatty acid, forming methyl branches at even positions of the fatty acid chain.

[0235] An expression vector encoding *Mycobacterium bovis* BCG fused to a poly-histidine tag also was generated. The pTrcHisA mmat plasmid DNA described above was amplified by PCR using oligonucleotides synthesized to include 5'-KpnI (SEQ ID NO: 31) and 3'-HindIII restriction sites (SEQ ID NO: 32) (Integrated DNA Technologies, Inc., Coralville, Iowa). PCR was run on samples having 1 μ l (2 ng) pTrcHisA mmat DNA, 1.5 μ l of a 10 μ M stock of each primer, 5 μ l of 10 \times Pfx reaction mix (Invitrogen Carlsbad, Calif.), 0.5 μ l of Pfx DNA polymerase (1.25 units), and 41 μ l of water. PCR conditions were as follows: the samples were initially incubated at 95 $^{\circ}$ C. for three minutes, followed by 30 cycles at 95 $^{\circ}$ C. for 30 seconds (strand separation), 58 $^{\circ}$ C. for 30 seconds (primer annealing), and 68 $^{\circ}$ C. primer extension for 1.5 minutes. Following the cycles, the samples were incubated for 10 minutes at 68 $^{\circ}$ C., and the samples were then held at 4 $^{\circ}$ C.

[0236] The PCR products were purified using a QIAquick $^{\circledR}$ PCR Purification Kit (Qiagen), digested with restriction enzymes KpnI and HindIII and ligated (Fast-Link Epicentre Biotechnologies, Madison, Wis.) with KpnI/HindIII-digested pZA31MCS (Expressys, Ruelzheim, Germany). The ligation mix was used to transform *E. coli* DH5 α TM (Invitrogen Carlsbad, Calif.). Isolated colonies were screened by PCR using a sterile pipette tip stab as an inoculum into a reaction tube containing only water, followed by addition of the remaining PCR reaction cocktail (AccuPrimeTM SuperMixII, Invitrogen Carlsbad, Calif.) and primers as described above.

[0237] Recombinant plasmids were isolated and purified using the QIAprep $^{\circledR}$ Spin Miniprep Kit (Qiagen) and characterized by restriction enzyme digestion (DraI, KpnI and HindIII from New England Biolabs, Beverly, Mass.). The

plasmids were subsequently used to transform BW25113 (*E. coli* Genetics Stock Center, New Haven, Conn.) made competent using the calcium chloride method. Transformants were selected on Luria agar plates containing 34 μ g/ml chloramphenicol. Plasmid DNA was isolated and purified using the QIAfilterTM Plasmid Midi Kit (Qiagen). DNA sequencing confirmed that the insert was mmat (SEQ ID NO: 34). The resulting plasmid incorporating a poly-histidine tag was designated pZA31 mmat.

Example 12

Method of Generating a Recombinant Host Cell Comprising an Exogenous Polynucleotide Encoding a Propionyl-CoA Carboxylase and an Exogenous Polynucleotide Encoding a Methylmalonyl-CoA Acyl Transferase (MMAT) Domain from *Mycobacterium Mycrocercosic Acid Synthase* (MAS).

[0238] This example describes an exemplary method for making a cell comprising an exogenous polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of propionyl-CoA to methylmalonyl-CoA and an exogenous polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of methylmalonyl-CoA to methylmalonyl-ACP. The method entails co-transduction of *E. coli* with plasmids containing a propionyl-CoA carboxylase gene from *Streptomyces coelicolor* and a gene encoding a MMAT domain from *Mycobacterium MAS*.

[0239] *E. coli* BW25113 cells (*E. coli* Genetic Stock Center, New Haven, Conn.) were made chemically competent for plasmid DNA transformation by a calcium chloride method. Actively growing 50 ml *E. coli* cultures were grown to an optical density (at 600 nm) of ~0.4. Cultures were quickly chilled on ice, and the bacteria were recovered by centrifugation at 2700 \times g for 10 minutes. The supernatant was discarded and pellets were gently suspended in 30 ml of an ice-cold 80 mM MgCl₂, 20 mM CaCl₂ solution. Cells were again recovered by centrifugation at 2700 \times g for 10 minutes. The supernatant was discarded and pellets were gently resuspended in 2 ml of an ice-cold 0.1 M CaCl₂ solution.

[0240] Cells were transformed on ice in pre-chilled 14 ml round-bottom centrifuge tubes. Approximately 25 ng of each of pTrcHisA mmat and pZA31-accA1-pccB (described above) was incubated on ice with 100 μ l of competent cells for 30 minutes. The cells were heat shocked at 42 $^{\circ}$ C. for 90 seconds and immediately placed on ice for two minutes. Pre-warmed SOC medium (500 μ l; Invitrogen, Carlsbad, Calif.) was added and the cells allowed to recover at 37 $^{\circ}$ C. with 225 rpm shaking. A portion (50 μ l) of the transformed cell mix was spread onto selective LB agar 100 mg/ml ampicillin and 34 mg/ml chloramphenicol plates to select for cells carrying the pTrcHisA mmat and pZA31/32-accA1-pccB plasmids. Individual colonies were picked from each plate and streaked onto LB agar (with ampicillin and chloramphenicol) to confirm the antibiotic resistance phenotype. Restriction endonuclease digestion analysis of isolated plasmid DNA with HaeII verified the plasmid DNA pool for each strain. A sample of *E. coli* BW25113 comprising pTrcHisA mmat and pZA31-accA1-pccB was deposited with American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, Va., on Dec. 14, 2010, under the provisions of the Budapest Treaty for the International Recognition of the

Deposit of Microorganisms for the Purpose of Patent Procedure (“Budapest Treaty”), and assigned Deposit Accession No. [XXX] on [DATE].

Example 13

Construction of an Expression Vector Encoding *Sorangium Cellulosum* So ce 56 Methylmalonyl-CoA Epimerase

[0241] A *S. cellulosum* methylmalonyl-CoA epimerase synthetic gene (So ce epi) was designed and synthesized (SEQ ID NO: 37). The coding sequence was codon-optimization for expression in *E. coli* and modified to remove restriction sites (GenScript, Piscataway, N.J.). The nucleic acid sequence was flanked with a SacI site and a synthetic ribosome binding site from the pBAD vector (Invitrogen, Carlsbad, Calif.) adjacent to the translation initiation sequence (SEQ ID NO: 39). The synthetic gene was cloned as a SacI/PstI fragment into pTrcHisA Ec sbm and pTrcHisA Ec malE Ec sbm, with the resulting plasmids designated as pTrcHisA Ec sbm So ce epi and pTrcHisA Ec malE Ec sbm So ce epi, respectively.

Example 14

Construction of an Expression Vector Encoding *Kribbella Flavida* DSM 17836 Methylmalonyl-CoA Epimerase

[0242] A *K. flavida* methylmalonyl-CoA epimerase gene (Kf epi) was designed and synthesized (SEQ ID NO: 35). The coding sequence was optimized for expression in *E. coli* and restriction sites were removed (GenScript, Piscataway, N.J.). The gene was flanked with a Sad site and a synthetic ribosome binding site from the pBAD vector adjacent to the translation initiation sequence (SEQ ID NO: 39). The synthetic gene was cloned as a SacI/PstI fragment into pTrcHisA Ec sbm and pTrcHisA Ec malE Ec sbm. The resulting plasmids were designated pTrcHisA Ec sbm Kf epi and pTrcHisA Ec malE Ec sbm Kf epi, respectively.

Example 15

Production of Host Cells Producing Branched-Chain Fatty Acid

[0243] This example describes the production of branched-chain fatty acid using a recombinant host cell (e.g., *E. coli*) expressing polynucleotides encoding a propionyl-CoA carboxylase or a methylmalonyl-CoA mutase and a methylmalonyl-CoA epimerase, in some instances in conjunction with a polynucleotide encoding an acyl transferase and/or thioesterase.

[0244] It is useful to have the capacity to tailor the fatty acid chain length. Branched fatty acids of different lengths have different physical properties suitable for different commercial applications. To demonstrate the capacity to tailor the chain length of branched fatty acids, *E. coli* 'tesA (Cho, H., and J. E. Cronan, Jr. *J. Biological Chemistry* 270: 4216-9 (1995)) was incorporated into expression vectors described above and inserted into host cells. To create a pTrc Ec 'tesA expression vector, a truncated *E. coli* tesA ('tesA) cDNA (SEQ ID NO: 40) was created by PCR amplification of the *E. coli* tesA gene (GenBank Accession No. L06182). A 5' primer (SEQ ID NO: 41) was designed to anneal after the 26th codon of tesA, modifying the 27th codon from an alanine to a

methionine and creating a NcoI restriction site. A 3' primer (SEQ ID NO: 43) incorporating a BamHI restriction site was designed. PCR was performed with 50 µl of Pfu Ultra II Hotstart 2× master mix (Agilent Technologies, Santa Clara, Calif.), 1 µl of a mix of the two primers (10 µmoles of each), 1 µl of *E. coli* BW25113 genomic DNA, and 48 µl of water. PCR began with a two minute incubation at 95° C., followed by 30 cycles of 20 seconds at 95° C. for denaturation, 20 seconds for annealing at 58° C., and 15 seconds at 72° C. for extension. The sample was incubated at 72° C. for three minutes and then held at 4° C. The PCR product (Ec 'tesA) was purified using a QIAquick® PCR Purification Kit (Qiagen, Valencia, Calif.). The bacterial expression vector pTrcHisA and the 'tesA PCR product were digested with NcoI and BamHI. The digested vector and insert were ligated using Fast-Link (Epicentre Biotechnologies, Madison, Wis.). The ligation mix was then used to transform *E. coli* TOP 10 cells (Invitrogen, Carlsbad, Calif.). Recombinant plasmids were isolated using a QIAprep0 Spin Miniprep Kit (Qiagen) and characterized by gel electrophoresis of restriction digests with HaeII. DNA sequencing confirmed that the 'tesA insert had been cloned and that the insert encoded the expected amino acid sequence (SEQ ID NO: 45). The resulting plasmid was designated pTrc Ec 'tesA.

[0245] To limit gene expression, the truncated *E. coli* 'tesA gene was subcloned into the low-copy bacterial expression vector pZS21-MCS (Expressys, Ruelzheim, Germany). The expression vector pTrc Ec 'tesA was a template in a PCR reaction using a 5' primer designed to create a flanking XhoI restriction site and include the pTrcHisA lac promoter (to replace the pZS21-MCS vector tet promoter) (SEQ ID NO: 46) and a 3' primer incorporating a HindIII restriction site (SEQ ID NO: 47). PCR was performed with 50 µl of Pfu Ultra II Hotstart 2× master mix (Agilent Technologies, Santa Clara, Calif.), 1 µl of a mix of the two primers (10 µmoles of each), 1 µl of pTrc Ec 'tesA plasmid DNA (6 ng), and 48 µl of water. PCR began with a two minute incubation at 95° C., followed by 30 cycles of 20 seconds at 95° C. for denaturation, 20 seconds for annealing at 57° C., and 20 seconds at 72° C. for extension. The sample was incubated at 72° C. for three minutes and then held at 4° C. The PCR product was purified using a QIAquick® PCR Purification Kit (Qiagen, Valencia, Calif.). The bacterial expression vector pZS21-MCS and the Ec 'tesA PCR product were digested with XhoI and HindIII. The digested vector and insert were ligated using Fast-Link (Epicentre Biotechnologies, Madison, Wis.). The ligation mix was then used to transform *E. coli* TOP10 cells (Invitrogen, Carlsbad, Calif.). Recombinant plasmids were isolated using a QIAprep® Spin Miniprep Kit (Qiagen) and characterized by gel electrophoresis of restriction digests with HaeII. DNA sequencing confirmed that the 'tesA insert had been cloned and that the insert encoded the expected amino acid sequence (SEQ ID NO: 45). The resulting plasmid was designated pZS22 Ec 'tesA.

[0246] An *E. coli* strain deficient in fatty acid degradation (Voelker, T. A., and H. M. Davies. *J. Bacteriology* 176: 7320-7 (1994)) and able to regulate transcription of recombinant genes was generated as follows. An *E. coli* K-12 strain (K27) defective in fadD lacks the fatty acyl-CoA synthetase responsible for an initial step in fatty acid degradation. The strain K27 (F⁻, tyrT58(AS), fadD88, mel-1; CGSC Strain #5478) was obtained from the *E. coli* Genetic Stock Center (New Haven, Conn.). A genomic regulation cassette from strain DH5αZ1 [lac^q, PN25-tetR, Sp^R, deoR, supE44, Δ(lac-

ZYA-argFV169), $\phi 80$ lacZ Δ M15 (Expressys, Ruelzheim, Germany)] was introduced into the host strain. The transducing phage P1 vir was charged with DH5 α Z1 DNA as follows. A logarithmically growing culture (5 ml LB broth containing 0.2% glucose and 5 mM CaCl₂) of donor strain, DH5 α Z1, was infected with a 100 μ l of a lysate stock of P1 vir phage. The culture was further incubated three hours for the infected cells to lyse. The debris was pelleted, and the supernatant was further cleared through a 0.45 μ m syringe filter unit. The fresh lysate was titered by spotting 10 μ l of serial 1:10 dilutions of lysate in TM buffer (10 mM MgSO₄/10 mM Tris.Cl, pH 7.4) onto a 100 mm LB (with 2.5 mM CaCl₂) plate overlaid with a cultured lawn of *E. coli* in LB top agar (with 2.5 mM CaCl₂). The process was repeated using the newly created phage stock until the phage titer surpassed 10⁹ pfu/ml.

[0247] The higher titer phage stock was used to transduce fragments of the DH5 α Z1 genome into a recipient K27 strain. An overnight culture (1.5 ml) of K27 was pelleted and resuspended in 750 μ l of a P1 salts solution (10 mM CaCl₂/5 mM MgSO₄). 100 μ l of the suspended cells was inoculated with varying amounts of DH5 α Z1 donor P1 vir lysate (1, 10, and 100 μ l) in sterile test tubes. The phage was allowed to adsorb to the cells for 30 minutes at 37° C. Absorption was terminated by addition of 1 ml LB broth plus 200 μ l of 1 M sodium citrate, and the cultures were further incubated for 1 hour at 37° C. with aeration. The cultures were pelleted, and the cells suspended in 100 μ l of LB broth (plus 0.2 M sodium citrate) and spread onto LB agar plates with 50 μ g/mL spectinomycin. Spectinomycin-resistant strains were isolated, and genomic DNAs were screened by PCR for the presence of tetR, lacI^q and fadD88. One such transductant was named K27-Z1 and used in further studies.

[0248] To transform K27-Z1, competent cells were placed on ice in pre-chilled 14 ml round bottom centrifuge tubes. Each plasmid was incubated with 50 μ l of chemically competent K27-Z1 cells (Cohen, S. N., Change, A. C. Y., and L. Hsu. *Proceedings National Academy Sciences U.S.A.* 69: 2110-4 (1972)) for 30 minutes. The cells were heat shocked at 42° C. for 90 seconds and immediately placed on ice for two minutes. Pre-warmed SOC medium (250 μ l) (Invitrogen, Carlsbad, Calif.) was added, and the cells were allowed to recover at 37° C. with 125 rpm shaking for one hour. Transformed cell mix (20 μ l) was spread onto selective LB agar with 100 μ g/ml ampicillin to select for cells carrying the pTrcHisA-based plasmids. Transformed cell mix (50 μ l) was spread onto LB agar with 34 μ g/ml chloramphenicol to select for cells carrying the pZA31-based plasmids. Transformed cell mix (150 μ l) was spread onto LB agar with 100 μ g/ml ampicillin and 34 μ g/ml chloramphenicol to select for cells carrying both the pTrcHisA-based and pZA31-based plasmids. In some cases, the creation of triple transformants required two transformations: a double transformant was originally created, made competent, and transformed by a third plasmid.

[0249] Using the methods described above, *E. coli* strain K27-Z1 was transduced with pTrcHisA pZA31 (control), pZA31 mutAB Ss epi, pTrcHisA Ec sbm, and pTrcHisA Ec sbm/pZA31 Mb mmat. The bacteria were cultured in M9 with glycerol (0.2%) at 22° C. in flasks that were coated with black Scotch duct tape. After the bacteria reached an optical density (600 nm) of 0.4, a mix of IPTG, anhydrotetracycline, arabinose and hydroxocobalamin hydrochloride was added to the culture, giving final concentrations of 1 mM, 100 ng/ml, 0.2%, and 20 μ M, respectively. Twenty-four hours later, the

bacteria were harvested for coenzyme A analysis. Methylmalonyl-CoA production is illustrated in FIG. 24. Host cells producing exogenous methylmalonyl-CoA mutase and methylmalonyl-CoA epimerase (encoded by pZA31 mutAB Ss epi) produced over 25 ng methylmalonyl-CoA per ml culture. Host cells comprising additional copies of the Sbm (methylmalonyl-CoA mutase) coding sequence produced over three times the amount of methylmalonyl-CoA per ml of culture, and co-expression of an methylmalonyl-CoA acyl transferase reduced the amount of methylmalonyl-CoA present in the culture medium.

[0250] Production of methylmalonyl-CoA in host cells expressing exogenous propionyl-CoA carboxylase also was studied and is illustrated in FIG. 25. BW25113 (control) and BW25113 containing pZA31-accA1-pccB (labeled as Pcc in the figure) were cultured in LB, and the coenzyme-A thioesters were isolated and characterized as described above. Host cells comprising a polynucleotide encoding an exogenous propionyl-CoA carboxylase produced over about 15 ng methylmalonyl-CoA per ml of culture.

[0251] When Ec *tesA* was present, less longer-chain (fifteen and seventeen carbons) and more mid-chain (thirteen carbons) branched fatty acids were produced by the host cell, indicating that production of thioesterase increases the proportion of medium chain-length branched fatty acids produced by the inventive method.

Example 16

Analysis of Scattered Branched Fatty Acid by Two-Dimensional (2D) Gas Chromatography

[0252] To identify branched fatty acids produced by recombinant *E. coli* produced as described herein, fatty acids were isolated from bacterial cultures and derivatives were generated to facilitate identification. The fatty acid derivatives were separated by 2D gas chromatography and mass spectrometry was used to characterize fragmented samples. Derivatization of fatty acids to their 4,4' dimethylloxazoline derivatives prior to analysis via mass spectrometry has been described (Zhang, J. Y., QT. Yu, B. N. Liu and Z. H. Huang, *Biomed Env. Mass Spectrom.* 15:33 (1988)). By careful examination of minor spectral differences, it possible to determine the location of branch points on the backbones of fatty acid derivatives.

[0253] One liter of bacterial samples in LB (modified to contain only 0.5 mg/ml sodium chloride, unless otherwise indicated) with cyanocobalamin (20 μ M) were cultured at 22° C. for 25 hours following induction with IPTG, anhydrotetracycline, and arabinose. A cell pellet was collected by centrifugation at 3500 rpm, and the supernatant was discarded. The cell pellet was suspended in the remaining liquid, and the slurry was transferred into Pyrex tubes (#9826, Corning Inc., Lowell, Mass.). An equal volume of chloroform was added, and the sample was dried at room temperature overnight.

[0254] To produce samples for analysis, cell pellets (0.5 grams) were placed in a round bottom flask, and 0.5 grams of KOH pellets and 25 ml of water were added. The *E. coli* pellets and KOH solution were refluxed for three hours, and the sample was allowed to cool. Concentrated HCl was added drop-wise, using a methyl orange endpoint to ensure fatty carboxylic acids were in the acid form. The acidified aqueous solution was then extracted three times with 25 ml aliquots of hexane to extract the fatty acids into the organic layer.

[0255] To convert fatty acid to oxazoline derivatives, the hexane extract was evaporated to dryness and reconstituted

into 5 ml of hexane to which sodium sulfate was added as a drying agent. After evaporating the sample to a 1 ml volume, a portion (0.6 ml) was decanted into a Reactitherm™ vial. The hexane in the Reactitherm™ vial was again evaporated to dryness, and 2 ml of 2-methyl-2-aminopropanol was added. The vial was capped and heated for 4 hours at 200° C. The cooled 2-methyl-2-aminopropanol solution was transferred to a scintillation vial, to which 5 ml of methylene chloride was added. The sample was washed with three 5 ml volumes of water. Sodium sulfate was added to the methylene chloride to remove any residual water, and an aliquot was transferred to a GC vial for analysis.

[0256] The derivatized samples were analyzed on a Leco Pegasus 4D Comprehensive 2D gas chromatograph time-of-flight mass spectrometer equipped with a 30M Supelco GammaDex 120 (Supelco 24307) column in the first dimension and a 2M Varian VF5-MS (Varian CP9034) column in the second dimension. Retention times of key chain-length fatty acids (in both first and second dimensions) in test samples were confirmed by identical preparation and analysis of a Supelco (47080-U) BAME (bacterial acid methyl ester) standard mixture. Using these columns, 4,4'-dimethylloxazoline-derivatized branched-chain fatty acids were expected to elute prior to their linear chain-length homologs in the first dimension, and this was confirmed by the iso and anteiso structural isomers of C15 methyl esters (derivatized to their 4,4'-dimethylloxazoline derivatives) in the BAME standard reference above.

[0257] The profile of fatty acids produced by two strains was compared. The first strain was engineered to produce branched fatty acids [BL21 Star (DE3) (pTrcHisA Ec sbm So ce epi pZA31 mmat)] and the second was a control strain [BL21 Star (DE3) (pTrcHisA pZA31)]. A sample of *E. coli* BL21 Star (DE3) comprising pTrcHisA Ec sbm So ce epi and pZA31 mmat was deposited with American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, Va., on Dec. 14, 2010, under the provisions of the Budapest Treaty for the International Recognition of the Deposit of Microorganisms for the Purpose of Patent Procedure ("Budapest Treaty"), and assigned Deposit Accession No. [XXX] on [DATE]. The sample from the first strain revealed several peaks in the region where branched fatty acids were expected (FIG. 26), whereas the sample from the control strain revealed no such peaks (FIG. 27). For example, several peaks (labeled 54, 55, and 57) were in a position consistent with branched C15 acids, and peaks 137 and 139 were in a position expected for branched C17 acids. Mass spectrometry established that these peaks comprise branched fatty acids.

[0258] The mass spectral fragmentation pattern of oxazoline derivatives was used to confirm that the fatty acids identified using 2D GC contained branches. Oxazoline derivatives fragment along the length of the carbon chain starting from the functional end of the molecule. If a branch point occurs along the backbone, there is a gap in the mass spectrum pattern; which peak is missing (or reduced) depends on the location of the branch. FIG. 28 depicts the mass spectra of the peaks labeled 54, 55, and 57 in FIG. 26 as oxazoline derivatives of methyl-branched tetradecanoic fatty acids. The ions circled exhibit reduced or no intensity relative to the reference spectrum of linear pentadecanoic fatty acid (bottom spectrum), and were assigned as 8-methyl, 10-methyl, and 12-methyl (anteiso) tetradecanoic fatty acid (all as oxazoline derivatives). Peak 57 was tentatively identified as the anteiso C15 oxazoline derivative despite the similarity to the mass spec

data for the linear sample because 1) peak 61 migrated at the position of an anteiso C15 standard on 2D gas chromatography, 2) the 252 molecular weight ion is present in slightly lower amounts relative to the nearby 238 and 266 molecular weight ions, and 3) anteiso compounds can be difficult to identify by this technique. The 8- and 10-branched fatty acids are shown in the top two profiles of FIG. 28, readily identified by the almost complete absence of the fragment circled. Peaks 137 and 139 in FIG. 26 were assigned as 8-methylhexadecanoic acid and 12-methylhexadecanoic acids (as oxazoline derivatives). Thus, B132 Star (DE3) (pTrcHisA Ec sbm So ce epi pZA31 mmat) (i.e., a recombinant microbe comprising overexpressed or recombinant polynucleotides encoding a methylmalonyl-CoA mutase, a methylmalonyl-CoA epimerase, and an acyl transferase) generated branched-chain C15 and C17 fatty acids comprising methyl branches on even-number carbons.

[0259] Branched fatty acid production also was observed in host cells producing exogenous propionyl-CoA carboxylase and *Streptomyces coelicolor* methylmalonyl-CoA mutase. The propionyl-CoA carboxylase gene-containing strain produced the branched fatty acids shown in Table F.

TABLE F

Peak #	Proposed Compound ID	Formula	Molecular Weight	
			DMOX	as fatty acid
38	6-methyl, dodecanoic acid (DMOX)	C ₁₃ H ₃₃ (C ₄ H ₈ NO)	267	214
40	8-methyl, dodecanoic acid (DMOX)	C ₁₃ H ₃₃ (C ₄ H ₈ NO)	267	214
61	6-methyl, tridecanoic acid (DMOX)	C ₁₄ H ₃₅ (C ₄ H ₈ NO)	281	228
62	8-methyl, tridecanoic acid (DMOX)	C ₁₄ H ₃₅ (C ₄ H ₈ NO)	281	228
101	6-methyl, tetradecanoic acid (DMOX)	C ₁₅ H ₃₇ (C ₄ H ₈ NO)	295	242
103	10-methyl, tetradecanoic acid (DMOX)	C ₁₅ H ₃₇ (C ₄ H ₈ NO)	295	242
140	10-methyl, pentadecanoic acid (DMOX)	C ₁₆ H ₃₉ (C ₄ H ₈ NO)	309	256
182	8-methyl, hexadecanoic acid (DMOX)	C ₁₇ H ₄₁ (C ₄ H ₈ NO)	323	270
189	12-methyl, hexadecanoic acid (DMOX)	C ₁₇ H ₄₁ (C ₄ H ₈ NO)	323	270

[0260] The *S. coelicolor* methylmalonyl-CoA mutase gene-containing microbe (BL21 Star (DE3) harboring pZA31 mutAB Ss epi pTrcHisA mmat) produced four branched fatty acids: 6-methyltetradecanoic acid, 10-methyltetradecanoic acid, 6-methylhexadecanoic acid, and 12-methylhexadecanoic acid.

[0261] Using 2D gas chromatography and mass spectrometry, fatty acid profiles were compared for two recombinant strains comprising Ec sbm, So ce epi, Mb mmat and containing or lacking a thioesterase coding sequence (*tesA). The amount of branched C15 fatty acids relative to branched C17 fatty acids was greater in the *tesA-containing strain. The area percent ratio of branched C15 fatty acid to branched C17 fatty acids in K27-Z1 (pTrcHisA Ec sbm So ce epi pZA31 mmat) was 1.4, while the ratio produced by K27-Z1 (pTrcHisA Ec sbm So ce epi pZA31 mmat pZS22 Ec *tesA) was 7.0. Expression of a thioesterase shortened the chain length of branched fatty acids.

[0262] These results demonstrate that a cell of the invention producing propionyl-CoA carboxylase or producing methyl-

malonyl-CoA mutase, methylmalonyl-CoA epimerase, and acyl transferase generates branched-chain fatty acids comprising methyl branches on even-number carbons. Recombinant host cells further comprising a polynucleotide encoding a thioesterase preferentially produce fatty acid comprising shorter chain length.

[0263] The dimensions and values disclosed herein are not to be understood as being strictly limited to the exact numerical values recited. Instead, unless otherwise specified, each such dimension is intended to mean both the recited value and a functionally equivalent range surrounding that value. For example, a dimension disclosed as “40 mm” is intended to mean “about 40 mm.”

[0264] Every document cited herein, including any cross referenced or related patent or application, is hereby incorporated herein by reference in its entirety unless expressly

excluded or otherwise limited. The citation of any document is not an admission that it is prior art with respect to any invention disclosed or claimed herein or that it alone, or in any combination with any other reference or references, teaches, suggests or discloses any such invention. Further, to the extent that any meaning or definition of a term in this document conflicts with any meaning or definition of the same term in a document incorporated by reference, the meaning or definition assigned to that term in this document shall govern.

[0265] While particular embodiments of the invention have been illustrated and described, it would be obvious to those skilled in the art that various other changes and modifications can be made without departing from the spirit and scope of the invention. It is therefore intended to cover in the appended claims all such changes and modifications that are within the scope of this invention.

SEQUENCE LISTING

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cgtgaggcgt gggcgcgcat tggggaggta tttggggttg atgaggataa acgtggcgcc      960
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<212> TYPE: DNA

<213> ORGANISM: Escherichia coli

<400> SEQUENCE: 2

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<210> SEQ ID NO 3

<211> LENGTH: 671

<212> TYPE: PRT

<213> ORGANISM: Janibacter sp. HTCC2649

<400> SEQUENCE: 3

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35          40          45
Arg Gly Glu Val Gly Lys Val Gly Val Pro Ile Ser His Ile Gly Asp
50          55          60
Met Arg Ala Leu Phe Asp Gln Ile Pro Leu Gly Gln Met Asn Thr Ser
65          70          75          80
Met Thr Ile Asn Ala Thr Ala Met Trp Leu Leu Ala Met Tyr Gln Val
85          90          95
Ala Ala Glu Asp Gln Ala Thr Ala Ala Asp Glu Asp Pro Ala Ser Val
100         105         110
Val Lys Ala Leu Gly Gly Thr Thr Gln Asn Asp Ile Ile Lys Glu Tyr
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Leu Ser Arg Gly Thr Tyr Val Phe Ala Pro Ala Pro Ser Leu Arg Leu
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Ile Thr Asp Met Val Ser Tyr Thr Val Ser Asp Ile Pro Lys Trp Asn
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Pro Ile Asn Ile Cys Ser Tyr His Leu Gln Glu Ala Gly Ala Thr Pro
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Val Gln Glu Ile Ala Tyr Ala Met Ser Thr Ala Ile Ala Val Leu Asp
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 Gly Leu Pro Arg Pro Trp Asp Gln Gln Trp Ser Leu Arg Met Gln Gln
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 Val Leu Ala Tyr Glu Ser Asp Leu Leu Glu Tyr Glu Asp Leu Phe Glu
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<211> LENGTH: 146

<212> TYPE: PRT

<213> ORGANISM: Escherichia coli

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 35 40 45
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 <213> ORGANISM: Streptomyces sviveus

<400> SEQUENCE: 6

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			20					25					30		
Ser	Glu	Val	Asn	Glu	Glu	Gln	Gly	Val	Arg	Glu	Ala	Met	Leu	Lys	Ile
		35					40					45			
Asn	Glu	Thr	Ser	Asp	Gly	Gly	Ala	Ser	Tyr	Leu	Gln	Leu	Leu	Glu	Pro
	50					55					60				
Thr	Arg	Pro	Asp	Ser	Thr	Val	Ala	Lys	Trp	Leu	Asp	Lys	Asn	Gly	Glu
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Gly	Val	His	His	Ile	Ala	Phe	Gly	Thr	Ala	Asp	Val	Asp	Gln	Asp	Ala
				85					90					95	
Ala	Asp	Ile	Lys	Asp	Lys	Gly	Val	Arg	Val	Leu	Tyr	Glu	Glu	Pro	Arg
			100					105					110		
Arg	Gly	Ser	Met	Gly	Ser	Arg	Ile	Thr	Phe	Leu	His	Pro	Lys	Asp	Cys
		115					120					125			
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 <212> TYPE: DNA
 <213> ORGANISM: Streptomyces coelicolor
 <300> PUBLICATION INFORMATION:
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 <309> DATABASE ENTRY DATE: 1999-12-08
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aacgcccaca ggtccggcac catcaagggc ctaccgcgc aggtcggcgc ctccctcacc 1740
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<210> SEQ ID NO 8
<211> LENGTH: 1593
<212> TYPE: DNA
<213> ORGANISM: Streptomyces coelicolor
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: GenBank / AF113605.1
<309> DATABASE ENTRY DATE: 1999-12-08
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(1593)

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<400> SEQUENCE: 8

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ctcaggcgcg gtatcgagga agcgaacgc gccggttccg cacgcgcctg cgagaagcag 120
cacgccaaag gaaagctgac ggctcgtgaa cgcacgcacc tcctcctcga cgagggttcc 180
ttcgtcgcgc tggacagatt cgcctccgac cgtccacca acttcggcct cgaagccaac 240
cgcctctacg gcgacggcgt cgtcaccggc tacggcaccg tcgacggcgc ccccggtggc 300
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atcgtcaagg tgatggactt cgcctcaag accggctgcc cggctcgtcg catcaacgac 420
tccggcggcg cccgcatcca ggaggcgtg gcctccctcg gcgcctacgg cgagatcttc 480
cgcgcgaaca cccacgcctc cggcgtgatc ccgcagatca gcctggctgt cggcccgtgt 540
gcggcggcgc cgggtgactc ccccgcatc accgacttca cggtgatggt ggaccagacc 600

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agccacatgt tcatcaccgg tcccgaagtc atcaagacgg tcaccggcga ggacgtcggc 660
ttcaggagagc tggggcggcgc ccgcaaccac aactccacct cgggcgtggc ccaccacatg 720
gccggcgacg agaaggacgc ggtcgagtac gtcaagcagc tcctgtcgta cctgccgtcc 780
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gagcggcagc tggacacgat cgtcccggac tggcgaacc agccctacga catgactcc 900
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<210> SEQ ID NO 9
<211> LENGTH: 590
<212> TYPE: PRT
<213> ORGANISM: Streptomyces coelicolor
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: GenBank / AF113603.1
<309> DATABASE ENTRY DATE: 1999-12-08
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(590)

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<400> SEQUENCE: 9

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Ala Arg Ala Cys Arg Asp Ala Gly Ile Ala Ser Val Ala Val Tyr Ala
20           25           30
Asp Pro Asp Arg Asp Ala Leu His Val Arg Ala Ala Asp Glu Ala Phe
35           40           45
Ala Leu Gly Gly Asp Thr Pro Ala Thr Ser Tyr Leu Asp Ile Ala Lys
50           55           60
Val Leu Lys Ala Ala Arg Glu Ser Gly Ala Asp Ala Ile His Pro Gly
65           70           75           80
Tyr Gly Phe Leu Ser Glu Asn Ala Glu Phe Ala Gln Ala Val Leu Asp
85           90           95
Ala Gly Leu Ile Trp Ile Gly Pro Pro Pro His Ala Ile Arg Asp Arg
100          105          110
Gly Glu Lys Val Ala Ala Arg His Ile Ala Gln Arg Ala Gly Ala Pro
115          120          125
Leu Val Ala Gly Thr Pro Asp Pro Val Ser Gly Ala Asp Glu Val Val
130          135          140
Ala Phe Ala Lys Glu His Gly Leu Pro Ile Ala Ile Lys Ala Ala Phe
145          150          155          160

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Gly Gly Gly Gly Arg Gly Leu Lys Val Ala Arg Thr Leu Glu Glu Val
 165 170 175
 Pro Glu Leu Tyr Asp Ser Ala Val Arg Glu Ala Val Ala Ala Phe Gly
 180 185 190
 Arg Gly Glu Cys Phe Val Glu Arg Tyr Leu Asp Lys Pro Arg His Val
 195 200 205
 Glu Thr Gln Cys Leu Ala Asp Thr His Gly Asn Val Val Val Val Ser
 210 215 220
 Thr Arg Asp Cys Ser Leu Gln Arg Arg His Gln Lys Leu Val Glu Glu
 225 230 235 240
 Ala Pro Ala Pro Phe Leu Ser Glu Ala Gln Thr Glu Gln Leu Tyr Ser
 245 250 255
 Ser Ser Lys Ala Ile Leu Lys Glu Ala Gly Tyr Gly Gly Ala Gly Thr
 260 265 270
 Val Glu Phe Leu Val Gly Met Asp Gly Thr Ile Phe Phe Leu Glu Val
 275 280 285
 Asn Thr Arg Leu Gln Val Glu His Pro Val Thr Glu Glu Val Ala Gly
 290 295 300
 Ile Asp Leu Val Arg Glu Met Phe Arg Ile Ala Asp Gly Glu Glu Leu
 305 310 315 320
 Gly Tyr Asp Asp Pro Ala Leu Arg Gly His Ser Phe Glu Phe Arg Ile
 325 330 335
 Asn Gly Glu Asp Pro Gly Arg Gly Phe Leu Pro Ala Pro Gly Thr Val
 340 345 350
 Thr Leu Phe Ser Asp Ala Pro Thr Gly Pro Gly Val Arg Leu Asp Ala Gly
 355 360 365
 Val Glu Ser Gly Ser Val Ile Gly Pro Ala Trp Asp Ser Leu Leu Ala
 370 375 380
 Lys Leu Ile Val Thr Gly Arg Thr Arg Ala Glu Ala Leu Gln Arg Ala
 385 390 395 400
 Ala Arg Ala Leu Asp Glu Phe Thr Val Glu Gly Met Ala Thr Ala Ile
 405 410 415
 Pro Phe His Arg Thr Val Val Arg Asp Pro Ala Phe Ala Pro Glu Leu
 420 425 430
 Thr Gly Ser Thr Asp Pro Phe Thr Val His Thr Arg Trp Ile Glu Thr
 435 440 445
 Glu Phe Val Asn Glu Ile Lys Pro Phe Thr Thr Pro Ala Asp Thr Glu
 450 455 460
 Thr Asp Glu Glu Ser Gly Arg Glu Thr Val Val Val Glu Val Gly Gly
 465 470 475 480
 Lys Arg Leu Glu Val Ser Leu Pro Ser Ser Leu Gly Met Ser Leu Ala
 485 490 495
 Arg Thr Gly Leu Ala Ala Gly Ala Arg Pro Lys Arg Arg Ala Ala Lys
 500 505 510
 Lys Ser Gly Pro Ala Ala Ser Gly Asp Thr Leu Ala Ser Pro Met Gln
 515 520 525
 Gly Thr Ile Val Lys Ile Ala Val Glu Glu Gly Gln Glu Val Gln Glu
 530 535 540
 Gly Asp Leu Ile Val Val Leu Glu Ala Met Lys Met Glu Gln Pro Leu
 545 550 555 560

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Asn Ala His Arg Ser Gly Thr Ile Lys Gly Leu Thr Ala Glu Val Gly
565 570 575

Ala Ser Leu Thr Ser Gly Ala Ala Ile Cys Glu Ile Lys Asp
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<210> SEQ ID NO 10
 <211> LENGTH: 530
 <212> TYPE: PRT
 <213> ORGANISM: Streptomyces coelicolor
 <300> PUBLICATION INFORMATION:
 <308> DATABASE ACCESSION NUMBER: GenBank / AF113605.1
 <309> DATABASE ENTRY DATE: 1999-12-08
 <313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(530)

<400> SEQUENCE: 10

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Lys Leu Ala Asp Leu Arg Arg Arg Ile Glu Glu Ala Thr His Ala Gly
20 25 30

Ser Ala Arg Ala Val Glu Lys Gln His Ala Lys Gly Lys Leu Thr Ala
35 40 45

Arg Glu Arg Ile Asp Leu Leu Leu Asp Glu Gly Ser Phe Val Glu Leu
50 55 60

Asp Glu Phe Ala Arg His Arg Ser Thr Asn Phe Gly Leu Asp Ala Asn
65 70 75 80

Arg Pro Tyr Gly Asp Gly Val Val Thr Gly Tyr Gly Thr Val Asp Gly
85 90 95

Arg Pro Val Ala Val Phe Ser Gln Asp Phe Thr Val Phe Gly Gly Ala
100 105 110

Leu Gly Glu Val Tyr Gly Gln Lys Ile Val Lys Val Met Asp Phe Ala
115 120 125

Leu Lys Thr Gly Cys Pro Val Val Gly Ile Asn Asp Ser Gly Gly Ala
130 135 140

Arg Ile Gln Glu Gly Val Ala Ser Leu Gly Ala Tyr Gly Glu Ile Phe
145 150 155 160

Arg Arg Asn Thr His Ala Ser Gly Val Ile Pro Gln Ile Ser Leu Val
165 170 175

Val Gly Pro Cys Ala Gly Gly Ala Val Tyr Ser Pro Ala Ile Thr Asp
180 185 190

Phe Thr Val Met Val Asp Gln Thr Ser His Met Phe Ile Thr Gly Pro
195 200 205

Asp Val Ile Lys Thr Val Thr Gly Glu Asp Val Gly Phe Glu Glu Leu
210 215 220

Gly Gly Ala Arg Thr His Asn Ser Thr Ser Gly Val Ala His His Met
225 230 235 240

Ala Gly Asp Glu Lys Asp Ala Val Glu Tyr Val Lys Gln Leu Leu Ser
245 250 255

Tyr Leu Pro Ser Asn Asn Leu Ser Glu Pro Pro Ala Phe Pro Glu Glu
260 265 270

Ala Asp Leu Ala Val Thr Asp Glu Asp Ala Glu Leu Asp Thr Ile Val
275 280 285

Pro Asp Ser Ala Asn Gln Pro Tyr Asp Met His Ser Val Ile Glu His
290 295 300

Val Leu Asp Asp Ala Glu Phe Phe Glu Thr Gln Pro Leu Phe Ala Pro

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305		310		315		320
Asn Ile Leu Thr	Gly Phe Gly Arg Val	Glu Gly Arg Pro Val	Gly Ile			
	325	330	335			
Val Ala Asn Gln	Pro Met Gln Phe Ala	Gly Cys Leu Asp	Ile Thr Ala			
	340	345	350			
Ser Glu Lys Ala	Ala Arg Phe Val	Arg Thr Cys Asp	Ala Phe Asn Val			
	355	360	365			
Pro Val Leu Thr	Phe Val Asp Val	Pro Gly Phe Leu	Pro Gly Val Asp			
	370	375	380			
Gln Glu His Asp	Gly Ile Ile Arg	Arg Gly Ala Lys	Leu Ile Phe Ala			
	385	390	395			400
Tyr Ala Glu Ala	Thr Val Pro Leu	Ile Thr Val Ile	Thr Arg Lys Ala			
	405	410	415			
Phe Gly Gly Ala	Tyr Asp Val Met	Gly Ser Lys His	Leu Gly Ala Asp			
	420	425	430			
Leu Asn Leu Ala	Trp Pro Thr Ala	Gln Ile Ala Val	Met Gly Ala Gln			
	435	440	445			
Gly Ala Val Asn	Ile Leu His Arg	Arg Thr Ile Ala	Asp Ala Gly Asp			
	450	455	460			
Asp Ala Glu Ala	Thr Arg Ala Arg	Leu Ile Gln Glu	Tyr Glu Asp Ala			
	465	470	475			480
Leu Leu Asn Pro	Tyr Thr Ala Ala	Glu Arg Gly Tyr	Val Asp Ala Val			
	485	490	495			
Ile Met Pro Ser	Asp Thr Arg Arg	His Ile Val Arg	Gly Leu Arg Gln			
	500	505	510			
Leu Arg Thr Lys	Arg Glu Ser Leu	Pro Pro Lys Lys	His Gly Asn Ile			
	515	520	525			
Pro Leu						
	530					

<210> SEQ ID NO 11
 <211> LENGTH: 116
 <212> TYPE: DNA
 <213> ORGANISM: Streptomyces coelicolor

<400> SEQUENCE: 11

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<210> SEQ ID NO 12
 <211> LENGTH: 1773
 <212> TYPE: DNA
 <213> ORGANISM: Streptomyces coelicolor

<400> SEQUENCE: 12

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cgtgatgcag gtattgcaag tgttgcgggt tatgccgatc cggatcgcca tgcgctgcat    120
gttcgtgctg ccgatgaagc ctttgcaactg ggcggtgata ccccggaac gagctatctg    180
gatattgcaa aagtgcgtgaa agcagcgcgc gaaagcgggt cggatgccaat ccatccgggc    240
tacggttttc tgtctgaaaa tgcagaattt gcacagccgg tcttgatgc aggtctgatt    300
tggatcggtc cgccgccgca tgcaattcgt gatctgggcy ataaagtggc cgcacgccac    360
```

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atcgcccage gtgcaggcgc gccgctgggt gcgggcaccc cggaccgggt ttctggtgca 420
gatgaagtgg ttgcttttgc caaagaacat ggccctgccga ttgcatcaa agcagcattc 480
ggcgggtggcg gtccgctgtc gaaagtggcc cgtaccctgg aagaagtcc ggaactgtat 540
gatagcgcag ttcgcgaagc ggtggcagcg tttggcctg gtgaatgctt cgtggaacgc 600
tacctggata aaccgcgtca tgttgaacc cagtgtctgg cggatagca cggcaactg 660
gttgtgggta gcaccgcga ttgctctctg caacgtgcc accagaaact ggtggaagaa 720
gcaccggcgc cgtttctgag cgaagcccag accgaacagc tgtatagctc tagtaaagcg 780
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aatgcccacg ttagcggcac catcaaagc ctgacggcgc aagtgggtgc atctctgacc 1740
agtggcgcgg ccatttgca aatcaaagat taa 1773

```

```

<210> SEQ ID NO 13
<211> LENGTH: 57
<212> TYPE: DNA
<213> ORGANISM: Streptomyces coelicolor

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```

<400> SEQUENCE: 13

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```

agatctgcgg ccgatctag aaataatgtt gtttaacttt aagaaggaga tatattc 57

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<210> SEQ ID NO 14
<211> LENGTH: 1593
<212> TYPE: DNA
<213> ORGANISM: Streptomyces coelicolor

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<400> SEQUENCE: 14

```

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cacgcgaaag gtaaactgac ggcccgcgaa cgtatcgatc tgetgctgga tgaaggcagt 180
tttgttgaac tggatgaatt tgcacgccac cgtagcacca actttggtct ggatgcgaat 240
cgcccgtatg gcgatgggtg ggttaccggt tacggtaagg tggatggctg tccgggtgca 300

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gcgggcgggtg ccgtttactc tccggccatt accgatttta cggtgatggt tgatcagacc 600
agtcacatgt tcattacggg cccgatgtg atcaaaaccg ttacgggcga agatgtgggt 660
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aacaatctgt ctgaaccgcc ggcgttcccg gaagaagcag acctggcggg gaccgatgaa 840
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gataccgcc gtcatatcgt gcgtggtctg cgtcagctgc gtacgaaacg tgaatctctg 1560
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```

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<210> SEQ ID NO 15
<211> LENGTH: 3539
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic nucleotide

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<400> SEQUENCE: 15

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aattgtgagc ggataacaat tgacattgtg agcggataac aagatactga gcacatcagc 60
aggacgcact gaccgaattc aataattttg ttaacttta agaaggagat atacatatgc 120
gcaaagtget gattgcgaac cgtggtgaaa tcgccgttcg tgtggcacgc gcgtgtcgtg 180
atgcaggtat tgcaagtgtt gcggtgatg ccgatccgga tcgcgatgcg ctgcatgttc 240
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cccagcgtgc aggcgcgcgc ctggttgcgg gcaccccgga cccggttctt ggtgcagatg 540
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gtggcggtcg cggctctgaaa gtggcccgtg ccctggaaga agttccgga ctgtatgata 660

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atctgaacct ggcatggccc accgcacaga tcgcagtgat gggcgcgcag ggtgccgtta 3300
atattctgca ccgccgtacc atcgcagatg caggtgatga tgcagaagcg acgcgcgcac 3360
gtctgattca ggaatatgaa gatgcgctgc tgaaccgta taccgcagcg gaacgtggtt 3420
acgtggatgc ggttattatg ccgagcgata cccgccgta ttcgtgctg ggtctgctc 3480
agctgcgtac gaaactgtaa tctctgccgc cgaaaaaaca cggtaatatt ccgctgtaa 3539

```

```

<210> SEQ ID NO 16
<211> LENGTH: 45
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic primer

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<400> SEQUENCE: 16

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```

aaactgcaga ggaggacagc tatgtctttt agcgaatttt atcag 45

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```

<210> SEQ ID NO 17
<211> LENGTH: 36
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic primer

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```

<400> SEQUENCE: 17

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```

aaaggatccc tattcttctga tcgcctggcg aatttg 36

```

```

<210> SEQ ID NO 18
<211> LENGTH: 383
<212> TYPE: PRT
<213> ORGANISM: Mycobacterium bovis

```

```

<400> SEQUENCE: 18

```

```

Leu Val Glu Gly Leu Arg Glu Val Ala Asp Gly Asp Ala Leu Tyr Asp
1           5           10          15
Ala Ala Val Gly His Gly Asp Arg Gly Pro Val Trp Val Phe Ser Gly
20          25          30
Gln Gly Ser Gln Trp Ala Ala Met Gly Thr Gln Leu Leu Ala Ser Glu
35          40          45
Pro Val Phe Ala Ala Thr Ile Ala Lys Leu Glu Pro Val Ile Ala Ala
50          55          60
Glu Ser Gly Phe Ser Val Thr Glu Ala Ile Thr Ala Gln Gln Thr Val
65          70          75          80
Thr Gly Ile Asp Lys Val Gln Pro Ala Val Phe Ala Val Gln Val Ala
85          90          95
Leu Ala Ala Thr Met Glu Gln Thr Tyr Gly Val Arg Pro Gly Ala Val
100         105         110
Val Gly His Ser Met Gly Glu Ser Ala Ala Ala Val Val Ala Gly Ala
115         120         125

```

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Leu Ser Leu Glu Asp Ala Ala Arg Val Ile Cys Arg Arg Ser Lys Leu
  130                               135                       140

Met Thr Arg Ile Ala Gly Ala Gly Ala Met Gly Ser Val Glu Leu Pro
  145                               150                       155                       160

Ala Lys Gln Val Asn Ser Glu Leu Met Ala Arg Gly Ile Asp Asp Val
                               165                               170                       175

Val Val Ser Val Val Ala Ser Pro Gln Ser Thr Val Ile Gly Gly Thr
                               180                               185                       190

Ser Asp Thr Val Arg Asp Leu Ile Ala Arg Trp Glu Gln Arg Asp Val
  195                               200                       205

Met Ala Arg Glu Val Ala Val Asp Val Ala Ser His Ser Pro Gln Val
  210                               215                       220

Asp Pro Ile Leu Asp Asp Leu Ala Ala Ala Leu Ala Asp Ile Ala Pro
  225                               230                       235                       240

Met Thr Pro Lys Val Pro Tyr Tyr Ser Ala Thr Leu Phe Asp Pro Arg
                               245                               250                       255

Glu Gln Pro Val Cys Asp Gly Ala Tyr Trp Val Asp Asn Leu Arg Asn
                               260                               265                       270

Thr Val Gln Phe Ala Ala Ala Val Gln Ala Ala Met Glu Asp Gly Tyr
  275                               280                       285

Arg Val Phe Ala Glu Leu Ser Pro His Pro Leu Leu Thr His Ala Val
  290                               295                       300

Glu Gln Thr Gly Arg Ser Leu Asp Met Ser Val Ala Ala Leu Ala Gly
  305                               310                       315                       320

Met Arg Arg Glu Gln Pro Leu Pro His Gly Leu Arg Gly Leu Leu Thr
                               325                               330                       335

Glu Leu His Arg Ala Gly Ala Ala Leu Asp Tyr Ser Ala Leu Tyr Pro
  340                               345                       350

Ala Gly Arg Leu Val Asp Ala Pro Leu Pro Ala Trp Thr His Ala Arg
  355                               360                       365

Leu Phe Ile Asp Asp Asp Gly Gln Glu Gln Arg Ala Gln Gly Ala
  370                               375                       380

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<210> SEQ ID NO 19
<211> LENGTH: 2111
<212> TYPE: PRT
<213> ORGANISM: Mycobacterium bovis
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: GenBank / YP_97046
<309> DATABASE ENTRY DATE: 2010-12-14
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(2111)

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<400> SEQUENCE: 19

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Met Glu Ser Arg Val Thr Pro Val Ala Val Ile Gly Met Gly Cys Arg
  1           5           10

Leu Pro Gly Gly Ile Asn Ser Pro Asp Lys Leu Trp Glu Ser Leu Leu
  20           25           30

Arg Gly Asp Asp Leu Val Thr Glu Ile Pro Pro Asp Arg Trp Asp Ala
  35           40           45

Asp Asp Tyr Tyr Asp Pro Glu Pro Gly Val Pro Gly Arg Ser Val Ser
  50           55           60

Arg Trp Gly Gly Phe Leu Asp Asp Val Ala Gly Phe Asp Ala Glu Phe
  65           70           75           80

Phe Gly Ile Ser Glu Arg Glu Ala Thr Ser Ile Asp Pro Gln Gln Arg

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			85			90			95						
Leu	Leu	Leu	Glu	Thr	Ser	Trp	Glu	Ala	Ile	Glu	His	Ala	Gly	Leu	Asp
			100					105					110		
Pro	Ala	Ser	Leu	Ala	Gly	Ser	Ser	Thr	Ala	Val	Phe	Thr	Gly	Leu	Thr
		115						120					125		
His	Glu	Asp	Tyr	Leu	Val	Leu	Thr	Thr	Thr	Ala	Gly	Gly	Leu	Ala	Ser
	130						135					140			
Pro	Tyr	Val	Val	Thr	Gly	Leu	Asn	Asn	Ser	Val	Ala	Ser	Gly	Arg	Ile
145					150					155					160
Ala	His	Thr	Leu	Gly	Leu	His	Gly	Pro	Ala	Met	Thr	Phe	Asp	Thr	Ala
				165					170						175
Cys	Ser	Ser	Gly	Leu	Met	Ala	Val	His	Leu	Ala	Cys	Arg	Ser	Leu	His
			180						185					190	
Asp	Gly	Glu	Ala	Asp	Leu	Ala	Leu	Ala	Gly	Gly	Cys	Ala	Val	Leu	Leu
		195					200					205			
Glu	Pro	His	Ala	Cys	Val	Ala	Ala	Ser	Ala	Gln	Gly	Met	Leu	Ser	Ser
	210						215					220			
Thr	Gly	Arg	Cys	His	Ser	Phe	Asp	Ala	Asp	Ala	Asp	Gly	Phe	Val	Arg
225						230				235					240
Ser	Glu	Gly	Cys	Ala	Met	Val	Leu	Leu	Lys	Arg	Leu	Pro	Asp	Ala	Leu
				245						250					255
Arg	Asp	Gly	Asn	Arg	Ile	Phe	Ala	Val	Val	Arg	Gly	Thr	Ala	Thr	Asn
			260					265						270	
Gln	Asp	Gly	Arg	Thr	Glu	Thr	Leu	Thr	Met	Pro	Ser	Glu	Asp	Ala	Gln
		275					280						285		
Val	Ala	Val	Tyr	Arg	Ala	Ala	Leu	Ala	Ala	Ala	Gly	Val	Gln	Pro	Glu
	290						295					300			
Thr	Val	Gly	Val	Val	Glu	Ala	His	Gly	Thr	Gly	Thr	Pro	Ile	Gly	Asp
305					310					315					320
Pro	Ile	Glu	Tyr	Arg	Ser	Leu	Ala	Arg	Val	Tyr	Gly	Ala	Gly	Thr	Pro
				325					330						335
Cys	Ala	Leu	Gly	Ser	Ala	Lys	Ser	Asn	Met	Gly	His	Ser	Thr	Ala	Ser
			340					345						350	
Ala	Gly	Thr	Val	Gly	Leu	Ile	Lys	Ala	Ile	Leu	Ser	Leu	Arg	His	Gly
		355					360						365		
Val	Val	Pro	Pro	Leu	Leu	His	Phe	Asn	Arg	Leu	Pro	Asp	Glu	Leu	Ser
	370					375						380			
Asp	Val	Glu	Thr	Gly	Leu	Phe	Val	Pro	Gln	Ala	Val	Thr	Pro	Trp	Pro
385					390					395					400
Asn	Gly	Asn	Asp	His	Thr	Pro	Lys	Arg	Val	Ala	Val	Ser	Ser	Phe	Gly
				405					410					415	
Met	Ser	Gly	Thr	Asn	Val	His	Ala	Ile	Val	Glu	Glu	Ala	Pro	Ala	Glu
			420					425					430		
Ala	Ser	Ala	Pro	Glu	Ser	Ser	Pro	Gly	Asp	Ala	Glu	Val	Gly	Pro	Arg
		435					440					445			
Leu	Phe	Met	Leu	Ser	Ser	Thr	Ser	Ser	Asp	Ala	Leu	Arg	Gln	Thr	Ala
	450					455						460			
Arg	Gln	Leu	Ala	Thr	Trp	Val	Glu	Glu	His	Gln	Asp	Cys	Val	Ala	Ala
465					470					475					480
Ser	Asp	Leu	Ala	Tyr	Thr	Leu	Ala	Arg	Gly	Arg	Ala	His	Arg	Pro	Val
				485					490					495	

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Arg Thr Ala Val Val Ala Ala Asn Leu Pro Glu Leu Val Glu Gly Leu
 500 505 510

Arg Glu Val Ala Asp Gly Asp Ala Leu Tyr Asp Ala Ala Val Gly His
 515 520 525

Gly Asp Arg Gly Pro Val Trp Val Phe Ser Gly Gln Gly Ser Gln Trp
 530 535 540

Ala Ala Met Gly Thr Gln Leu Leu Ala Ser Glu Pro Val Phe Ala Ala
 545 550 555 560

Thr Ile Ala Lys Leu Glu Pro Val Ile Ala Ala Glu Ser Gly Phe Ser
 565 570 575

Val Thr Glu Ala Ile Thr Ala Gln Gln Thr Val Thr Gly Ile Asp Lys
 580 585 590

Val Gln Pro Ala Val Phe Ala Val Gln Val Ala Leu Ala Ala Thr Met
 595 600 605

Glu Gln Thr Tyr Gly Val Arg Pro Gly Ala Val Val Gly His Ser Met
 610 615 620

Gly Glu Ser Ala Ala Ala Val Val Ala Gly Ala Leu Ser Leu Glu Asp
 625 630 635 640

Ala Ala Arg Val Ile Cys Arg Arg Ser Lys Leu Met Thr Arg Ile Ala
 645 650 655

Gly Ala Gly Ala Met Gly Ser Val Glu Leu Pro Ala Lys Gln Val Asn
 660 665 670

Ser Glu Leu Met Ala Arg Gly Ile Asp Asp Val Val Val Ser Val Val
 675 680 685

Ala Ser Pro Gln Ser Thr Val Ile Gly Gly Thr Ser Asp Thr Val Arg
 690 695 700

Asp Leu Ile Ala Arg Trp Glu Gln Arg Asp Val Met Ala Arg Glu Val
 705 710 715 720

Ala Val Asp Val Ala Ser His Ser Pro Gln Val Asp Pro Ile Leu Asp
 725 730 735

Asp Leu Ala Ala Ala Leu Ala Asp Ile Ala Pro Met Thr Pro Lys Val
 740 745 750

Pro Tyr Tyr Ser Ala Thr Leu Phe Asp Pro Arg Glu Gln Pro Val Cys
 755 760 765

Asp Gly Ala Tyr Trp Val Asp Asn Leu Arg Asn Thr Val Gln Phe Ala
 770 775 780

Ala Ala Val Gln Ala Ala Met Glu Asp Gly Tyr Arg Val Phe Ala Glu
 785 790 795 800

Leu Ser Pro His Pro Leu Leu Thr His Ala Val Glu Gln Thr Gly Arg
 805 810 815

Ser Leu Asp Met Ser Val Ala Ala Leu Ala Gly Met Arg Arg Glu Gln
 820 825 830

Pro Leu Pro His Gly Leu Arg Gly Leu Leu Thr Glu Leu His Arg Ala
 835 840 845

Gly Ala Ala Leu Asp Tyr Ser Ala Leu Tyr Pro Ala Gly Arg Leu Val
 850 855 860

Asp Ala Pro Leu Pro Ala Trp Thr His Ala Arg Leu Phe Ile Asp Asp
 865 870 875 880

Asp Gly Gln Glu Gln Arg Ala Gln Gly Ala Cys Thr Ile Thr Val His
 885 890 895

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Pro	Leu	Leu	Gly	Ser	His	Val	Arg	Leu	Thr	Glu	Glu	Pro	Glu	Arg	His
			900					905					910		
Val	Trp	Gln	Gly	Asp	Val	Gly	Thr	Ser	Val	Leu	Ser	Trp	Leu	Ser	Asp
		915					920					925			
His	Gln	Val	His	Asn	Val	Ala	Ala	Leu	Pro	Gly	Ala	Ala	Tyr	Cys	Glu
	930					935					940				
Met	Ala	Leu	Ala	Ala	Ala	Ala	Glu	Val	Phe	Gly	Glu	Ala	Ala	Glu	Val
945					950					955					960
Arg	Asp	Ile	Thr	Phe	Glu	Gln	Met	Leu	Leu	Leu	Asp	Glu	Gln	Thr	Pro
				965					970					975	
Ile	Asp	Ala	Val	Ala	Ser	Ile	Asp	Ala	Pro	Gly	Val	Val	Asn	Phe	Thr
		980						985					990		
Val	Glu	Thr	Asn	Arg	Asp	Gly	Glu	Thr	Thr	Arg	His	Ala	Thr	Ala	Ala
		995					1000					1005			
Leu	Arg	Ala	Ala	Glu	Asp	Asp	Cys	Pro	Pro	Pro	Gly	Tyr	Asp	Ile	
	1010					1015					1020				
Thr	Ala	Leu	Leu	Gln	Ala	His	Pro	His	Ala	Val	Asn	Gly	Thr	Ala	
	1025					1030						1035			
Met	Arg	Glu	Ser	Phe	Ala	Glu	Arg	Gly	Val	Thr	Leu	Gly	Ala	Ala	
	1040					1045						1050			
Phe	Gly	Gly	Leu	Thr	Thr	Ala	His	Thr	Ala	Glu	Ala	Gly	Ala	Ala	
	1055					1060						1065			
Thr	Val	Leu	Ala	Glu	Val	Ala	Leu	Pro	Ala	Ser	Ile	Arg	Phe	Gln	
	1070					1075						1080			
Gln	Gly	Ala	Tyr	Arg	Ile	His	Pro	Ala	Leu	Leu	Asp	Ala	Cys	Phe	
	1085					1090						1095			
Gln	Ser	Val	Gly	Ala	Gly	Val	Gln	Ala	Gly	Thr	Ala	Thr	Gly	Gly	
	1100					1105						1110			
Leu	Leu	Leu	Pro	Leu	Gly	Val	Arg	Ser	Leu	Arg	Ala	Tyr	Gly	Pro	
	1115					1120						1125			
Thr	Arg	Asn	Ala	Arg	Tyr	Cys	Tyr	Thr	Arg	Leu	Thr	Lys	Ala	Phe	
	1130					1135						1140			
Asn	Asp	Gly	Thr	Arg	Gly	Gly	Glu	Ala	Asp	Leu	Asp	Val	Leu	Asp	
	1145					1150						1155			
Glu	His	Gly	Thr	Val	Leu	Leu	Ala	Val	Arg	Gly	Leu	Arg	Met	Gly	
	1160					1165						1170			
Thr	Gly	Thr	Ser	Glu	Arg	Asp	Glu	Arg	Asp	Arg	Leu	Val	Ser	Glu	
	1175					1180						1185			
Arg	Leu	Leu	Thr	Leu	Gly	Trp	Gln	Gln	Arg	Ala	Leu	Pro	Glu	Val	
	1190					1195						1200			
Gly	Asp	Gly	Glu	Ala	Gly	Ser	Trp	Leu	Leu	Ile	Asp	Thr	Ser	Asn	
	1205					1210						1215			
Ala	Val	Asp	Thr	Pro	Asp	Met	Leu	Ala	Ser	Thr	Leu	Thr	Asp	Ala	
	1220					1225						1230			
Leu	Lys	Ser	His	Gly	Pro	Gln	Gly	Thr	Glu	Cys	Ala	Ser	Leu	Ser	
	1235					1240						1245			
Trp	Ser	Val	Gln	Asp	Thr	Pro	Pro	Asn	Asp	Gln	Ala	Gly	Leu	Glu	
	1250					1255						1260			
Lys	Leu	Gly	Ser	Gln	Leu	Arg	Gly	Arg	Asp	Gly	Val	Val	Ile	Val	
	1265					1270						1275			
Tyr	Gly	Pro	Arg	Val	Gly	Asp	Pro	Asp	Glu	His	Ser	Leu	Leu	Ala	

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1280	1285	1290
Gly Arg 1295	Glu Gln Val Arg His 1300	Leu Val Arg Ile Thr Arg Glu Leu 1305
Ala Glu 1310	Phe Glu Gly Glu Leu 1315	Pro Arg Leu Phe Val Val Thr Arg 1320
Gln Ala 1325	Gln Ile Val Lys Pro 1330	His Asp Ser Gly Glu Arg Ala Asn 1335
Leu Glu 1340	Gln Ala Gly Leu Arg 1345	Gly Leu Leu Arg Val Ile Ser Ser 1350
Glu His 1355	Pro Met Leu Arg Thr 1360	Thr Leu Ile Asp Val Asp Glu His 1365
Thr Asp 1370	Val Glu Arg Val Ala 1375	Gln Gln Leu Leu Ser Gly Ser Glu 1380
Glu Asp 1385	Glu Thr Ala Trp Arg 1390	Asn Gly Asp Trp Tyr Val Ala Arg 1395
Leu Thr 1400	Pro Ser Pro Leu Gly 1405	His Glu Glu Arg Arg Thr Ala Val 1410
Leu Asp 1415	Pro Asp His Asp Gly 1420	Met Arg Val Gln Val Arg Arg Pro 1425
Gly Asp 1430	Leu Gln Thr Leu Glu 1435	Phe Val Ala Ser Asp Arg Val Pro 1440
Pro Gly 1445	Pro Gly Gln Ile Glu 1450	Val Ala Val Ser Met Ser Ser Ile 1455
Asn Phe 1460	Ala Asp Val Leu Ile 1465	Ala Phe Gly Arg Phe Pro Ile Ile 1470
Asp Asp 1475	Arg Glu Pro Gln Leu 1480	Gly Met Asp Phe Val Gly Val Val 1485
Thr Ala 1490	Val Gly Glu Gly Val 1495	Thr Gly His Gln Val Gly Asp Arg 1500
Val Gly 1505	Gly Phe Ser Glu Gly 1510	Gly Cys Trp Arg Thr Phe Leu Thr 1515
Cys Asp 1520	Ala Asn Leu Ala Val 1525	Thr Leu Pro Pro Gly Leu Thr Asp 1530
Glu Gln 1535	Ala Ile Thr Ala Ala 1540	Thr Ala His Ala Thr Ala Trp Tyr 1545
Gly Leu 1550	Asn Asp Leu Ala Gln 1555	Ile Lys Ala Gly Asp Lys Val Leu 1560
Ile His 1565	Ser Ala Thr Gly Gly 1570	Val Gly Gln Ala Ala Ile Ser Ile 1575
Ala Arg 1580	Ala Lys Gly Ala Glu 1585	Ile Phe Ala Thr Ala Gly Asn Pro 1590
Ala Lys 1595	Arg Ala Met Leu Arg 1600	Asp Met Gly Val Glu His Val Tyr 1605
Asp Ser 1610	Arg Ser Val Glu Phe 1615	Ala Glu Gln Ile Arg Arg Asp Thr 1620
Asp Gly 1625	Tyr Gly Val Asp Ile 1630	Val Leu Asn Ser Leu Thr Gly Ala 1635
Ala Gln 1640	Arg Ala Gly Leu Glu 1645	Leu Leu Ala Phe Gly Gly Arg Phe 1650
Val Glu 1655	Ile Gly Lys Ala Asp 1660	Val Tyr Gly Asn Thr Arg Leu Gly 1665

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Leu	Phe	Pro	Phe	Arg	Arg	Gly	Leu	Thr	Phe	Tyr	Tyr	Leu	Asp	Leu
1670						1675						1680		
Ala	Leu	Met	Ser	Val	Thr	Gln	Pro	Asp	Arg	Val	Arg	Glu	Leu	Leu
1685						1690					1695			
Ala	Thr	Val	Phe	Lys	Leu	Thr	Ala	Asp	Gly	Val	Leu	Thr	Ala	Pro
1700						1705					1710			
Gln	Cys	Thr	His	Tyr	Pro	Leu	Ala	Glu	Ala	Ala	Asp	Ala	Ile	Arg
1715						1720					1725			
Ala	Met	Ser	Asn	Ala	Glu	His	Thr	Gly	Lys	Leu	Val	Leu	Asp	Val
1730						1735					1740			
Pro	Arg	Ser	Gly	Arg	Arg	Ser	Val	Ala	Val	Thr	Pro	Glu	Gln	Ala
1745						1750					1755			
Pro	Leu	Tyr	Arg	Arg	Asp	Gly	Ser	Tyr	Ile	Ile	Thr	Gly	Gly	Leu
1760						1765					1770			
Gly	Gly	Leu	Gly	Leu	Phe	Phe	Ala	Ser	Lys	Leu	Ala	Ala	Ala	Gly
1775						1780					1785			
Cys	Gly	Arg	Ile	Val	Leu	Thr	Ala	Arg	Ser	Gln	Pro	Asn	Pro	Lys
1790						1795					1800			
Ala	Arg	Gln	Thr	Ile	Glu	Gly	Leu	Arg	Ala	Ala	Gly	Ala	Asp	Ile
1805						1810					1815			
Val	Val	Glu	Cys	Gly	Asn	Ile	Ala	Glu	Pro	Asp	Thr	Ala	Asp	Arg
1820						1825					1830			
Leu	Val	Ser	Ala	Ala	Thr	Ala	Thr	Gly	Leu	Pro	Leu	Arg	Gly	Val
1835						1840					1845			
Leu	His	Ser	Ala	Ala	Val	Val	Glu	Asp	Ala	Thr	Leu	Thr	Asn	Ile
1850						1855					1860			
Thr	Asp	Glu	Leu	Ile	Asp	Arg	Asp	Trp	Ser	Pro	Lys	Val	Phe	Gly
1865						1870					1875			
Ser	Trp	Asn	Leu	His	Arg	Ala	Thr	Leu	Gly	Gln	Pro	Leu	Asp	Trp
1880						1885					1890			
Phe	Cys	Leu	Phe	Ser	Ser	Gly	Ala	Ala	Leu	Leu	Gly	Ser	Pro	Gly
1895						1900					1905			
Gln	Gly	Ala	Tyr	Ala	Ala	Ala	Asn	Ser	Trp	Val	Asp	Val	Phe	Ala
1910						1915					1920			
His	Trp	Arg	Arg	Ala	Gln	Gly	Leu	Pro	Val	Ser	Ala	Ile	Ala	Trp
1925						1930					1935			
Gly	Ala	Trp	Gly	Glu	Val	Gly	Arg	Ala	Thr	Phe	Leu	Ala	Glu	Gly
1940						1945					1950			
Gly	Glu	Ile	Met	Ile	Thr	Pro	Glu	Glu	Gly	Ala	Tyr	Ala	Phe	Glu
1955						1960					1965			
Thr	Leu	Val	Arg	His	Asp	Arg	Ala	Tyr	Ser	Gly	Tyr	Ile	Pro	Ile
1970						1975					1980			
Leu	Gly	Ala	Pro	Trp	Leu	Ala	Asp	Leu	Val	Arg	Arg	Ser	Pro	Trp
1985						1990					1995			
Gly	Glu	Met	Phe	Ala	Ser	Thr	Gly	Gln	Arg	Ser	Arg	Gly	Pro	Ser
2000						2005					2010			
Lys	Phe	Arg	Met	Glu	Leu	Leu	Ser	Leu	Pro	Gln	Asp	Glu	Trp	Ala
2015						2020					2025			
Gly	Arg	Leu	Arg	Arg	Leu	Leu	Val	Glu	Gln	Ala	Ser	Val	Ile	Leu
2030						2035					2040			

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Arg	Arg	Thr	Ile	Asp	Ala	Asp	Arg	Ser	Phe	Ile	Glu	Tyr	Gly	Leu
	2045					2050					2055			
Asp	Ser	Leu	Gly	Met	Leu	Glu	Met	Arg	Thr	His	Val	Glu	Thr	Glu
	2060					2065					2070			
Thr	Gly	Ile	Arg	Leu	Thr	Pro	Lys	Val	Ile	Ala	Thr	Asn	Asn	Thr
	2075					2080					2085			
Ala	Arg	Ala	Leu	Ala	Gln	Tyr	Leu	Ala	Asp	Thr	Leu	Ala	Glu	Glu
	2090					2095					2100			
Gln	Ala	Ala	Ala	Pro	Ala	Ala	Ser							
	2105					2110								

<210> SEQ ID NO 20

<211> LENGTH: 1149

<212> TYPE: DNA

<213> ORGANISM: Mycobacterium bovis

<400> SEQUENCE: 20

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ctggtggaag gctcgcgtga agttgccgat ggtgatgcac tgtatgatgc agcagtgagg      60
catggcgatc gtggtccggt ttgggtgttt agcggccagg gttctcagtg ggcagcgatg      120
ggcaccacgc tgctggcaag cgaaccgggt tttgccgcaa cgattgcaaa actggaaccg      180
gtgatcgcgg ccgaaagtgg cttcagcgtt accgaagcaa ttacggcgca gcagaccgtg      240
acgggtatcg ataaagtgca gccggccggt ttcgcagttc aggtggcgct ggcagcgacg      300
atggaacaga cgtacggcgt tcgctccggg gcagtggttg gtcacagtat gggtgaaagc      360
gccgcagcgg tggttgcagg cgcctgagc ctggaagatg ccgcacgtgt gatttgccgt      420
cgcagcaaac tgatgacccg tatcgcaggt gcaggtgcga tgggcagcgt ggaactgccg      480
gcaaaacagg ttaactctga actgatggcg cgcggtattg atgatgtggt tgtgtctggt      540
gtggcgcttc cgcagagtac cgtgattggc ggcaccagtg atacggttcg tgatctgatc      600
gcgcggtggg aacagcgcga tgtgatggcg cgcaagtgg ccgtggatgt tgcaagccat      660
tctccgcagg ttgatccgat tctggatgat ctggcggcgg cactggcaga tattgcaccg      720
atgaccccca aagtgcctga ttacagcgcg acgctgtttg atccgcgtga acagccggtg      780
tgtgatggcg cctattgggt tgataacctg cgcaataccg tgcagtttgc ggcggcagtt      840
caggcggcga tggaagtggg ttaccgtgtg ttcgcggaac tgtctccgca tccgctgctg      900
acccacgcag tggaacagac gggtcgctct ctggatatga gtgttcagc actggccggt      960
atgctgcgcg aacagccgct gccgcattgg ctgctgtggtc tgctgacga actgcaccgt     1020
gcaggtgcag cactggatta tagcgcactg tacccggcag gtcgtctggt ggatgcaccg     1080
ctgcccggcat ggacgcacgc acgtctgttc atcgatgatg atggccagga acagcgcgca     1140
cagggtgctg                                     1149

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<210> SEQ ID NO 21

<211> LENGTH: 1149

<212> TYPE: DNA

<213> ORGANISM: Mycobacterium bovis

<400> SEQUENCE: 21

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ctcgtcgagg gtttgcgcga ggtggccgac ggtgacgccc tctatgacgc ggcggtgagg      60
caccgtgate gaggaccggt ctgggtcttc tccgggcaag ggtcgcagtg ggcggcgatg     120
ggcagcgaat tgctcgcag cgaaccagtg ttcgcggcca ccatcgccaa gctggagccg     180

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gtgatcgccg cagaatcggg attctcgggt accgaggcga taacggcgca gcagaccgtg 240
accggaatcg acaaagtga gccggcagtg ttcgccgttc aggtcgcggt gcccgccacc 300
atggagcaaa cctacggagt gcggccgggc gcggtcgtcg gacactcgat gggtagtgctg 360
gcccgggccg tcgtcggggg ggcactgtcg ctcgaggacg cggcgcgcggt catttgccgc 420
cgctcgaagc tgatgacctg catagccggt gctggtgcca tgggctcggg ggaattgccc 480
gccaaagcaag tgaattcggg gctgatggca cgcggaatcg acgatgttgt ggtctcggtg 540
gtggcgctcc cgcaatccac ggtgatcggc ggtacgagcg acaccgttcg tgacctcacc 600
gcccgttggg agcagcggga cgtgatggcg cgcgaggtgg ccgtcgacgt ggcgtcgcac 660
tcgcctcaag tcgatccgat actcgacgat ttggcccgcg cgctggcgga cattgtcccg 720
atgacgcccc aggtgcccga ctactcggcg accctgttcg acccgcgcca gcagccggtg 780
tgcgatggcg cttactgggt ggacaatctg cgcaacacgg tgcagttcgc cgcggcggtg 840
caggctgcga tggaggacgg ctaccgggtc ttcgcgagc tgctgcccc cccgctgctt 900
accacgcccg tcgaacagac gggccgaagc ctcgacatgt cggtcgccc cctggccggc 960
atgcccggcg agcagcctct gccgatgggt ctgcgcggtt tgctgacgga gctgcaccgc 1020
gcgggcccgc ctttgacta ttcgcgctg tatcccctg ggcggctggt ggatgcgccg 1080
ctgcccggct ggaccacgc ccgcctatc atcgacgatg atgggcaaga acagcgggca 1140
caaggtgcc 1149

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<210> SEQ ID NO 22
<211> LENGTH: 628
<212> TYPE: PRT
<213> ORGANISM: Salmonella enterica
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: GenBank / AAC44817
<309> DATABASE ENTRY DATE: 1999-08-05
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(628)

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<400> SEQUENCE: 22

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Met Ser Phe Ser Glu Phe Tyr Gln Arg Ser Ile Asn Glu Pro Glu Ala
1           5           10           15
Phe Trp Ala Glu Gln Ala Arg Arg Ile Asp Trp Arg Gln Pro Phe Thr
20           25           30
Gln Thr Leu Asp His Ser Arg Pro Pro Phe Ala Arg Trp Phe Cys Gly
35           40           45
Gly Thr Thr Asn Leu Cys His Asn Ala Val Asp Arg Trp Arg Asp Lys
50           55           60
Gln Pro Glu Ala Leu Ala Leu Ile Ala Val Ser Ser Glu Thr Asp Glu
65           70           75           80
Glu Arg Thr Phe Thr Phe Ser Gln Leu His Asp Glu Val Asn Ile Val
85           90           95
Ala Ala Met Leu Leu Ser Leu Gly Val Gln Arg Gly Asp Arg Val Leu
100          105          110
Val Tyr Met Pro Met Ile Ala Glu Ala Gln Ile Thr Leu Leu Ala Cys
115          120          125
Ala Arg Ile Gly Ala Ile His Ser Val Val Phe Gly Gly Phe Ala Ser
130          135          140
His Ser Val Ala Ala Arg Ile Asp Asp Ala Arg Pro Ala Leu Ile Val
145          150          155          160

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-continued

Ser Ala Asp Ala Gly Ala Arg Gly Gly Lys Ile Leu Pro Tyr Lys Lys
 165 170 175
 Leu Leu Asp Asp Ala Ile Ala Gln Ala Gln His Gln Pro Lys His Val
 180 185 190
 Leu Leu Val Asp Arg Gly Leu Ala Lys Met Ala Trp Val Asp Gly Arg
 195 200 205
 Asp Leu Asp Phe Ala Thr Leu Arg Gln Gln His Leu Gly Ala Ser Val
 210 215 220
 Pro Val Ala Trp Leu Glu Ser Asn Glu Thr Ser Cys Ile Leu Tyr Thr
 225 230 235 240
 Ser Gly Thr Thr Gly Lys Pro Lys Gly Val Gln Arg Asp Val Gly Gly
 245 250 255
 Tyr Ala Val Ala Leu Ala Thr Ser Met Asp Thr Ile Phe Gly Gly Lys
 260 265 270
 Ala Gly Gly Val Phe Phe Cys Ala Ser Asp Ile Gly Trp Val Val Gly
 275 280 285
 His Ser Tyr Ile Val Tyr Ala Pro Leu Leu Ala Gly Met Ala Thr Ile
 290 295 300
 Val Tyr Glu Gly Leu Pro Thr Tyr Pro Asp Cys Gly Val Trp Trp Lys
 305 310 315 320
 Ile Val Glu Lys Tyr Gln Val Asn Arg Met Phe Ser Ala Pro Thr Ala
 325 330 335
 Ile Arg Val Leu Lys Lys Phe Pro Thr Ala Gln Ile Arg Asn His Asp
 340 345 350
 Leu Ser Ser Leu Glu Ala Leu Tyr Leu Ala Gly Glu Pro Leu Asp Glu
 355 360 365
 Pro Thr Ala Ser Trp Val Thr Glu Thr Leu Gly Val Pro Val Ile Asp
 370 375 380
 Asn Tyr Trp Gln Thr Glu Ser Gly Trp Pro Ile Met Ala Leu Ala Arg
 385 390 395 400
 Ala Leu Asp Asp Arg Pro Ser Arg Leu Gly Ser Pro Gly Val Pro Met
 405 410 415
 Tyr Gly Tyr Asn Val Gln Leu Leu Asn Glu Val Thr Gly Glu Pro Cys
 420 425 430
 Gly Ile Asn Glu Lys Gly Met Leu Val Ile Glu Gly Pro Leu Pro Pro
 435 440 445
 Gly Cys Ile Gln Thr Ile Trp Gly Asp Asp Ala Arg Phe Val Lys Thr
 450 455 460
 Tyr Trp Ser Leu Phe Asn Arg Gln Val Tyr Ala Thr Phe Asp Trp Gly
 465 470 475 480
 Ile Arg Asp Ala Glu Gly Tyr Tyr Phe Ile Leu Gly Arg Thr Asp Asp
 485 490 495
 Val Ile Asn Ile Ala Gly His Arg Leu Gly Thr Arg Glu Ile Glu Glu
 500 505 510
 Ser Ile Ser Ser Tyr Pro Asn Val Ala Glu Val Ala Val Val Gly Ile
 515 520 525
 Lys Asp Ala Leu Lys Gly Gln Val Ala Val Ala Phe Val Ile Pro Lys
 530 535 540
 Gln Ser Asp Thr Leu Ala Asp Arg Glu Ala Ala Arg Asp Glu Glu Asn
 545 550 555 560

-continued

Ala Ile Met Ala Leu Val Asp Asn Gln Ile Gly His Phe Gly Arg Pro
 565 570 575

Ala His Val Trp Phe Val Ser Gln Leu Pro Lys Thr Arg Ser Gly Lys
 580 585 590

Met Leu Arg Arg Thr Ile Gln Ala Ile Cys Glu Gly Arg Asp Pro Gly
 595 600 605

Asp Leu Thr Thr Ile Asp Asp Pro Ala Ser Leu Gln Gln Ile Arg Gln
 610 615 620

Ala Ile Glu Glu
 625

<210> SEQ ID NO 23

<211> LENGTH: 1884

<212> TYPE: DNA

<213> ORGANISM: *Salmonella enterica*

<400> SEQUENCE: 23

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atgtctttaa gcaaatTTTA tcagcgttcc attaacgaac cggaggcggt ctgggccgag    60
caggcccggc gtatcgactg gcgacagccg tttacgcaga cgctggatca tagccgtcca    120
ccgtttgccc gctggttttg cggcggcacc actaacttat gtcataacgc cgtcgaccgc    180
tggcgggata aacagccgga ggcgctggcg ctgattgccg tctcatcaga gaccgatgaa    240
gagcgcacat ttaccttcag ccagttgcat gatgaagtca acattgtggc cgccatggtg    300
ctgtcgctgg gcgtgcagcg tggcgatcgc gtattggtct atatgccgat gattgccgaa    360
gcgcagataa ccctgctggc ctgcgcgcgc attggcgcga tccattcggg ggtctttggc    420
ggttttgccct cgcacagcgt ggcggcgcgc attgacgatg ccagaccggc gctgattgtg    480
tcggcggatg ccggagcgcg gggcggtaaa atcctgccgt ataaaaagct gctcgatgac    540
gctattgcgc aggcgcagca tcagccgaaa cacgttctgc tgggtggacag agggctggcg    600
aaaaatggcat ggggtgatgg gcgcgatctg gattttgcca cgttgcgcca gcagcatctc    660
ggcgcgagcg tgcgggtggc gtggctggaa tccaacgaaa cctcgtgcat tctttacacc    720
tccggcacta ccggcaaaccc gaaaggcgtc cagcgcgacg tcggcgggta tgcgggtggcg    780
ctggcaacct cgatggacac catttttggc ggcaaggcgg gcggcgtatt cttttgcgca    840
tcggatatcg gctgggtcgt cggccactcc tataatcgtt acgcgccggt gctggcaggc    900
atggcgacta ttgtttacga aggactgccg acgtaccggg actgcggggt ctggtggaaa    960
attgtcgaga aataccagggt taaccggatg ttttccgccc cgaccgcgat tcgctgctg    1020
aaaaaattcc cgacggcgca aatccgcaat cacgatctct cctcgtgga ggcgctttat    1080
ctggccggty agccgctgga cgagccgacg gccagttggg taacggagac gctgggcgta    1140
ccggtcatcg acaattattg gcagaccggag tccggctggc cgatcatggc gctggcccgc    1200
gcgctggacg acaggccgct gcgtctggga agtcccggcg tgccgatgta cggttataac    1260
gtccagctac tcaatgaagt caccggcgaa ccttgcggca taaatgaaaa ggggatgctg    1320
gtgatcgaag gcccgctgcc gccgggctgt attcagacta tttggggcga cgatgcgcgt    1380
tttgtgaaga cttactggtc gctgtttaac cgtcaggttt atgccacttt cgaactgggga    1440
atccgcgacg ccgaggggta ttactttatt ctgggccgta ccgatgatgt gattaatatt    1500
gcgggtcacc ggctggggac gcgagaaaata gaagaaaagta tctccagcta cccgaacgta    1560
gcggaagtgg cggtagtggg gataaaagac gctctgaaag ggcaggtagc ggtggcgttt    1620

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gtcattccga agcagagcga tacgctggcg gatcgcgagg cggcgcgcga cgaggaaaac 1680
gcgattatgg cgctggtgga caaccagatc ggtcactttg gtcgtccggc gcatgtctgg 1740
tttgtttcgc agctcccca aacgcgttcc ggaaagatgc ttcgccgcac gatccaggcg 1800
atctgcgaag gccgcgatcc gggcgatctg acaaccattg acgatcccgc gtcggttcag 1860
caaattcgcc aggcgatcga agaa 1884

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<210> SEQ ID NO 24
<211> LENGTH: 616
<212> TYPE: PRT
<213> ORGANISM: Streptomyces cinnamomensis

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<400> SEQUENCE: 24

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Met Thr Val Leu Pro Asp Asp Gly Leu Ser Leu Ala Ala Glu Phe Pro
1      5      10     15
Asp Ala Thr His Glu Gln Trp His Arg Leu Val Glu Gly Val Val Arg
20     25     30
Lys Ser Gly Lys Asp Val Ser Gly Thr Ala Ala Glu Glu Ala Leu Ser
35     40     45
Thr Thr Leu Glu Asp Gly Leu Thr Thr Arg Pro Leu Tyr Thr Ala Arg
50     55     60
Asp Ala Ala Pro Asp Ala Gly Phe Pro Gly Phe Ala Pro Phe Val Arg
65     70     75     80
Gly Ser Val Pro Glu Gly Asn Thr Pro Gly Gly Trp Asp Val Arg Gln
85     90     95
Arg Tyr Ala Ser Ala Asp Pro Ala Arg Thr Asn Glu Ala Val Leu Thr
100    105    110
Asp Leu Glu Asn Gly Val Thr Ser Leu Trp Leu Thr Leu Gly Ser Ala
115    120    125
Gly Leu Pro Val Thr Gly Leu Glu Arg Ala Leu Asp Gly Val Tyr Leu
130    135    140
Asp Leu Val Pro Val Ala Leu Asp Ala Gly Ser Glu Ala Ala Thr Ala
145    150    155    160
Ala Arg Glu Leu Leu Arg Leu Tyr Glu Ala Ala Gly Val Ala Asp Asp
165    170    175
Ala Val Arg Gly Thr Leu Gly Ala Asp Pro Leu Gly His Glu Ala Arg
180    185    190
Thr Gly Glu Lys Ser Thr Ser Phe Ala Ala Val Ala Glu Leu Ala Arg
195    200    205
Leu Cys Gly Glu Arg Tyr Pro Gly Leu Arg Ala Leu Thr Val Asp Ala
210    215    220
Leu Pro Tyr His Glu Ala Gly Ala Ser Ala Ala Gln Glu Leu Gly Ala
225    230    235    240
Ser Leu Ala Thr Gly Val Glu Tyr Leu Arg Ala Leu His Asp Lys Gly
245    250    255
Leu Gly Val Glu Lys Ala Phe Ala Gln Leu Glu Phe Arg Phe Ala Ala
260    265    270
Thr Ala Asp Gln Phe Leu Thr Ile Ala Lys Leu Arg Ala Ala Arg Arg
275    280    285
Leu Trp Ala Arg Val Ala Glu Val Ser Gly Val Pro Ala Ala Gly Ala
290    295    300

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Gln Arg Gln His Ala Val Thr Ser Pro Val Met Met Thr Arg Arg Asp
 305 310 315 320
 Pro Trp Val Asn Met Leu Arg Thr Thr Val Ala Cys Leu Gly Ala Gly
 325 330 335
 Val Gly Gly Ala Asp Ala Val Thr Val Leu Pro Phe Asp His Glu Leu
 340 345 350
 Gly Leu Pro Asp Ala Phe Ala Arg Arg Ile Ala Arg Asn Thr Ser Thr
 355 360 365
 Ile Leu Leu Glu Glu Ser His Leu Ala Arg Val Ile Asp Pro Ala Gly
 370 375 380
 Gly Ser Trp Tyr Val Glu Arg Leu Thr Asp Glu Leu Ala His Ala Ala
 385 390 395 400
 Trp Asp Phe Phe Lys Glu Ile Glu Arg Ala Asp Gly Gln Val Ala Ala
 405 410 415
 Leu Arg Ser Gly Leu Val Gly Asp Arg Ile Ala Ala Thr Trp Ala Glu
 420 425 430
 Arg Arg Lys Lys Leu Ala Arg Arg Arg Glu Pro Ile Thr Gly Val Ser
 435 440 445
 Glu Phe Pro Leu Leu Thr Glu Arg Pro Val Glu Arg Glu Pro Ala Pro
 450 455 460
 Ala Ala Pro Pro Gly Gly Leu Pro Arg Val Arg Arg Asp Glu Ala Tyr
 465 470 475 480
 Glu Glu Leu Arg Gly Arg Ser Asp Ala His Leu Glu Ala Thr Gly Ala
 485 490 495
 Arg Pro Lys Val Phe Ile Ala Ala Leu Gly Pro Ala Ala Ala His Thr
 500 505 510
 Ala Arg Ala Thr Phe Ala Ala Asn Leu Phe Met Ala Gly Gly Val Glu
 515 520 525
 Pro Val His Asp Pro Val Ser Val Asp Ala Glu Thr Ala Ala Glu Ala
 530 535 540
 Phe Ala Ala Ser Gly Ala Thr Val Ala Cys Leu Cys Ser Ser Asp Val
 545 550 555 560
 Leu Tyr Ala Glu Gln Ala Glu Ala Val Ala Arg Ala Leu Lys Ser Ala
 565 570 575
 Gly Ala Leu Arg Val Phe Leu Ala Gly Arg Gly Glu Phe Ala Asp Ile
 580 585 590
 Asp Glu Tyr Val Phe Ala Gly Cys Asp Ala Val Ala Val Leu Thr Ser
 595 600 605
 Thr Leu Asp Arg Met Gly Val Ala
 610 615

<210> SEQ ID NO 25

<211> LENGTH: 733

<212> TYPE: PRT

<213> ORGANISM: Streptomyces cinnamomensis

<400> SEQUENCE: 25

Met Arg Ile Pro Glu Phe Asp Asp Ile Glu Leu Gly Ala Gly Gly Gly
 1 5 10 15
 Pro Ser Gly Ser Ala Glu Gln Trp Arg Ala Ala Val Lys Glu Ser Val
 20 25 30
 Gly Lys Ser Glu Ser Asp Leu Leu Trp Glu Thr Pro Glu Gly Ile Ala
 35 40 45

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Val Lys Pro Leu Tyr Thr Gly Ala Asp Val Glu Gly Leu Asp Phe Leu
 50 55 60

Glu Thr Tyr Pro Gly Val Ala Pro Tyr Leu Arg Gly Pro Tyr Pro Thr
 65 70 75 80

Met Tyr Val Asn Gln Pro Trp Thr Ile Arg Gln Tyr Ala Gly Phe Ser
 85 90 95

Thr Ala Glu Glu Ser Asn Ala Phe Tyr Arg Arg Asn Leu Ala Ala Gly
 100 105 110

Gln Lys Gly Leu Ser Val Ala Phe Asp Leu Pro Thr His Arg Gly Tyr
 115 120 125

Asp Ser Asp His Pro Arg Val Thr Gly Asp Val Gly Met Ala Gly Val
 130 135 140

Ala Ile Asp Ser Ile Tyr Asp Met Arg Gln Leu Phe Asp Gly Ile Pro
 145 150 155 160

Leu Asp Lys Met Thr Val Ser Met Thr Met Asn Gly Ala Val Leu Pro
 165 170 175

Val Leu Ala Leu Tyr Ile Val Ala Ala Glu Glu Gln Gly Val Pro Pro
 180 185 190

Glu Lys Leu Ala Gly Thr Ile Gln Asn Asp Ile Leu Lys Glu Phe Met
 195 200 205

Val Arg Asn Thr Tyr Ile Tyr Pro Pro Lys Pro Ser Met Arg Ile Ile
 210 215 220

Ser Asp Ile Phe Ala Tyr Thr Ser Gln Lys Met Pro Arg Tyr Asn Ser
 225 230 235 240

Ile Ser Ile Ser Gly Tyr His Ile Gln Glu Ala Gly Ala Thr Ala Asp
 245 250 255

Leu Glu Leu Ala Tyr Thr Leu Ala Asp Gly Val Glu Tyr Leu Arg Ala
 260 265 270

Gly Gln Glu Ala Gly Leu Asp Val Asp Ala Phe Ala Pro Arg Leu Ser
 275 280 285

Phe Phe Trp Ala Ile Gly Met Asn Phe Phe Met Glu Val Ala Lys Leu
 290 295 300

Arg Ala Ala Arg Leu Leu Trp Ala Lys Leu Val Lys Gln Phe Asp Pro
 305 310 315

Lys Asn Ala Lys Ser Leu Ser Leu Arg Thr His Ser Gln Thr Ser Gly
 325 330 335

Trp Ser Leu Thr Ala Gln Asp Val Phe Asn Asn Val Thr Arg Thr Cys
 340 345 350

Val Glu Ala Met Ala Ala Thr Gln Gly His Thr Gln Ser Leu His Thr
 355 360 365

Asn Ala Leu Asp Glu Ala Leu Ala Leu Pro Thr Asp Phe Ser Ala Arg
 370 375 380

Ile Ala Arg Asn Thr Gln Leu Leu Ile Gln Gln Glu Ser Gly Thr Thr
 385 390 395 400

Arg Thr Ile Asp Pro Trp Gly Gly Ser Ala Tyr Val Glu Lys Leu Thr
 405 410 415

Tyr Asp Leu Ala Arg Arg Ala Trp Gln His Ile Glu Glu Val Glu Ala
 420 425 430

Ala Gly Gly Met Ala Gln Ala Ile Asp Ala Gly Ile Pro Lys Leu Arg
 435 440 445

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Val Glu Glu Ala Ala Ala Arg Thr Gln Ala Arg Ile Asp Ser Gly Arg
 450 455 460
 Gln Pro Val Ile Gly Val Asn Lys Tyr Arg Val Asp Thr Asp Glu Gln
 465 470 475 480
 Ile Asp Val Leu Lys Val Asp Asn Ser Ser Val Arg Ala Gln Gln Ile
 485 490 495
 Glu Lys Leu Arg Arg Leu Arg Glu Glu Arg Asp Asp Ala Ala Cys Gln
 500 505 510
 Asp Ala Leu Arg Ala Leu Thr Ala Ala Ala Glu Arg Gly Pro Gly Gln
 515 520 525
 Gly Leu Glu Gly Asn Leu Leu Ala Leu Ala Val Asp Ala Ala Arg Ala
 530 535 540
 Lys Ala Thr Val Gly Glu Ile Ser Asp Ala Leu Glu Ser Val Tyr Gly
 545 550 555 560
 Arg His Ala Gly Gln Ile Arg Thr Ile Ser Gly Val Tyr Arg Thr Glu
 565 570 575
 Ala Gly Gln Ser Pro Ser Val Glu Arg Thr Arg Ala Leu Val Asp Ala
 580 585 590
 Phe Asp Glu Ala Glu Gly Arg Arg Pro Arg Ile Leu Val Ala Lys Met
 595 600 605
 Gly Gln Asp Gly His Asp Arg Gly Gln Lys Val Ile Ala Ser Ala Phe
 610 615 620
 Ala Asp Leu Gly Phe Asp Val Asp Val Gly Pro Leu Phe Gln Thr Pro
 625 630 635 640
 Ala Glu Val Ala Arg Gln Ala Val Glu Ala Asp Val His Ile Val Gly
 645 650 655
 Val Ser Ser Leu Ala Ala Gly His Leu Thr Leu Val Pro Ala Leu Arg
 660 665 670
 Glu Glu Leu Ala Ala Glu Gly Arg Asp Asp Ile Met Ile Val Val Gly
 675 680 685
 Gly Val Ile Pro Pro Gln Asp Val Glu Ala Leu His Glu Ala Gly Ala
 690 695 700
 Thr Ala Val Phe Pro Pro Gly Thr Val Ile Pro Asp Ala Ala His Asp
 705 710 715 720
 Leu Val Lys Arg Leu Ala Ala Asp Leu Gly His Glu Leu
 725 730

<210> SEQ ID NO 26

<211> LENGTH: 146

<212> TYPE: PRT

<213> ORGANISM: *Streptomyces sviveus*

<400> SEQUENCE: 26

Met Leu Thr Arg Ile Asp His Ile Gly Ile Ala Cys Phe Asp Leu Asp
 1 5 10 15
 Lys Thr Val Glu Phe Tyr Arg Ala Thr Tyr Gly Phe Glu Val Phe His
 20 25 30
 Ser Glu Val Asn Glu Glu Gln Gly Val Arg Glu Ala Met Leu Lys Ile
 35 40 45
 Asn Glu Thr Ser Asp Gly Gly Ala Ser Tyr Leu Gln Leu Leu Glu Pro
 50 55 60
 Thr Arg Pro Asp Ser Thr Val Ala Lys Trp Leu Asp Lys Asn Gly Glu
 65 70 75 80

-continued

Gly Val His His Ile Ala Phe Gly Thr Ala Asp Val Asp Gln Asp Ala
85 90 95

Ala Asp Ile Lys Asp Lys Gly Val Arg Val Leu Tyr Glu Glu Pro Arg
100 105 110

Arg Gly Ser Met Gly Ser Arg Ile Thr Phe Leu His Pro Lys Asp Cys
115 120 125

His Gly Val Leu Thr Glu Leu Val Thr Ser Ala Pro Val Glu Ser Pro
130 135 140

Glu His
145

<210> SEQ ID NO 27
<211> LENGTH: 4553
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic nucleotide

<400> SEQUENCE: 27

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gaattcaaaa ttaagaggta tatattaatg accgtgctgc cggatgacgg tctgagtctg    60
gcagccgaat ttccggatgc gacgcatgaa cagtggcacc gtctggttga aggcgtgggt    120
cgcaaatcag gcaaagatgt ctccggcacc gcagctgaag aagccctgag caccacgctg    180
gaagacggtc tgaccacgcg tccgctgtat acggcacgtg atgcagcacc ggacgctggt    240
tttcggggtt tcgcgccgtt tgtgctggc tcagttccgg agggtaaacac cccgggcgggt    300
tgggatgtgc gtcaacgtta cgcacgtgca gaccggcac gtaccaacga agcagtgtctg    360
acggatctgg aaaatggtgt taccagcctg tggctgacgc tgggttctgc aggtctgccc    420
gtgaccggtc tggaacgtgc actggatggt gtttatctgg acctggccc ggtggcactg    480
gatgcaggta gcgaagcagc taccgacgca cgtgaactgc tgcgtctgta cgaagcagct    540
ggtggttctg atgacgcagt ccgtggcagc ctgggtgcag atccgctggg ccatgaagca    600
cgaccgggtg aaaaaagtac gtcctttgca gcagtggcag aactggcagc tctgtgcccgt    660
gaacgttata cgggtctgcg cgctctgacc gttgatgcgc tgcctacca tgaagctggc    720
gcgtcagcag ctcaagaaact gggcgcttcg ctggcgaccg gtgtggaata tctgctgctg    780
ctgcacgata aaggcctggg tgttgaaaaa gccttcgcac agctggaatt tcgcttcgcg    840
gccaccgceg accaatttct gacgattgcc aaactgcgtg cagctcgtcg cctgtgggca    900
cgtggttcag aagtcagtg cgtgcccgc gcaggtgcac agcgtcaaca tgcagtcacc    960
tcccgggtga tgatgacgcg tcgcatccg tgggtgaaca tgctgcttac caccggttget    1020
tgtctgggtg caggtgtcgg cgggtctgat gcagttaccg tcttccggtt cgatcacgaa    1080
ctgggtctgc cggacgcctt tgcacgtcgc attgcgctga ataccagtac gatcctgctg    1140
gaagaatccc atctgcccgg tgtcattgat ccggcaggcg gtagctggtg tgtggaacgc    1200
ctgaccgatg aactggccca cgcagcttgg gactttttca aagaatcga acgtgcagat    1260
ggtcaggctc cagcactgcg tagcggcctg gtgggtgacc gcattgcagc tacctgggca    1320
gaacgtcgca aaaaactggc gcgtcgccgt gaaccgatca ccggtgtgtc tgaatttccg    1380
ctgtgacggc aacgcccggg tgaacgtgaa ccggcaccgg cagcaccgcc gggcgggtctg    1440
ccgcccgtgc gccgtgatga agcctacgaa gaactgcgtg gtcgttctga cgcacacctg    1500

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gaagctaccg	gtgcacgtcc	gaaagtgttc	attgcagctc	tgggtccggc	agcagcacat	1560
accgctcgtg	cgacgttcgc	tgcgaacctg	tttatggcgg	gcggtgttga	accggtccac	1620
gatcctgtga	gctttgacgc	ggaaaaccgc	gcagaagcct	ttgctgcgtc	tggcgccacg	1680
gttgcatgcc	tgtgtagctc	tgatgtcctg	tatgcggaac	aagccgaagc	agtcgctcgt	1740
gcgctgaaaa	gtgccgggtc	actgcgtgtt	ttcctggcag	gccgcggtga	atttgcggat	1800
atcgacgaat	acgtgtttgc	aggttgcgat	gctgtcgcag	tgctgaacctc	cacgctggac	1860
cgatagggtg	ttgcgtaatg	cgtattccgg	aatttgatga	catcgaactg	ggtgccggcg	1920
gtggcccgtc	aggttcggca	gaacagtggc	gtgcagcagt	gaaagaaagc	gttggtaaaa	1980
gcgaatctga	tctgctgtgg	gaaaccccg	aaggcattgc	tgttaaaccg	ctgtacacgg	2040
gtgccgatgt	cgaaggcctg	gacttctcgg	aaacctatcc	gggtgtcgca	cgtacctgc	2100
gtggtccgta	tccgaccatg	tacgtgaacc	agccgtggac	gatccgcaa	tacgcgggtt	2160
ttagcaccgc	cgaagaatct	aacgcattct	atcgtcgcaa	tctggcagct	ggccagaaa	2220
gtctgagtgt	ggcgtttgat	ctgccgaccc	atcgtggcta	cgattccgac	cacccgcgtg	2280
tcacgggtga	cgtgggtatg	gccggcgtgg	caattgatag	catctatgac	atgcgtcagc	2340
tgttcgatgg	tattccgctg	gacaaaatga	ccgtttctat	gacgatgaac	ggcgctgtgc	2400
tgccggttct	ggcgtgtgat	atcgtggcgg	ccgaagaaca	gggtgttccg	ccgaaaaaac	2460
tggcgggcac	catccaaaac	gatatcctga	aagaatttat	ggttcgtaac	acgtacatct	2520
accgcgcgaa	accgagtatg	cgcattatct	ccgatatctt	cgctataacc	tcacagaaaa	2580
tgccgcgcta	caacagtatc	tccatctcag	gttatcatat	ccaagaagca	ggcgctaccg	2640
cggatctgga	actggcctac	acgctggcag	acggtgttga	atatctgcgt	gctggtcagg	2700
aagcgggcct	ggatgtgcac	gcctttgcac	cgccctgag	ctttttctgg	gccattggca	2760
tgaacttttt	catggaagtg	gcaaaaactgc	gtgcagctcg	cctgctgtgg	gcgaaaactgg	2820
ttaaacagtt	tgatccgaaa	aatgcgaaat	cgctgagcct	gcgtaccac	tcccagacgt	2880
caggttggtc	gctgaccgcc	caagatgttt	tcaacaatgt	cacccgcacg	tgcgtggaag	2940
caatggcagc	aaccagggtg	catacgcaat	caactgcacac	caacgcctg	gatgaagctc	3000
tggcgctgcc	gaccgacttt	tccgctcgtg	ttgcgcgcaa	tacgcagctg	ctgatccagc	3060
aagaaagcgg	caccacgcgt	accattgatc	cgtggggtgg	ctctgcgtat	gtggaaaaac	3120
tgaagtacga	cctggcacgt	cgcgcattggc	agcatatcga	agaagttgaa	gcagcgggtg	3180
gcatggccca	agcaattgat	gcgggcatcc	cgaaactgcg	tgtggaagaa	gcggcagcac	3240
gtaccaggc	acgcattgat	tctggtcgtc	aaccggctcat	cgccgtgaa	aaatatcgcg	3300
tggatacggg	cgaacagatt	gatgttctga	aagtgcgcaa	tagctctgtt	cgcgcgcagc	3360
aaatcgaaaa	actgcgtcgc	ctgcgtgaag	aacgcgatga	cgctgcgtgt	caggatgctc	3420
tgcgtgcact	gaccgcagca	gctgaacgtg	gtccgggtca	gggtctggaa	ggtaactctgc	3480
tggctctggc	agtggatgca	gcacgtgcc	aagcaaccgt	tggcgaaatt	tcagacgcac	3540
tggaatcgg	ctacggtcgt	catgcgggcc	agattcgac	catcagtggt	gtgtatcgca	3600
cggagcggg	ccaatctccg	agtgtogaac	gtaccgcgc	cctgggtggat	gcatttgacg	3660
aagctgaagg	tcgtcgcggc	cgtattctgg	ttgcaaaaat	gggtcaggat	ggccacgacc	3720
gcggccaaaa	agtcacgcgt	tccgcgtttg	ccgatctggg	tttcgatgct	gacgtgggtc	3780

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cgctgttcca gaccccgccc gaagtggcac gtcaagctgt ggaagcggat gttcatattg 3840
ttggtgtcag ttccctggca gctggtcacc tgacgctggt tccggcactg cgtgaagaac 3900
tggcggccga aggtcgcgat gacattatga tcgtggttgg tggcgtcatt ccgcccgagg 3960
atgtggaage cctgcatgaa gcaggtgcta ccgcggtttt tccgccgggc acggtcatcc 4020
cggatgcagc tcatgacctg gtgaaacgtc tggcagcaga tctgggtcac gaactgtaaa 4080
agcttaaaat taagaggat atattaatgc tgaccgccat cgatcacatt ggcatcgcat 4140
gctttgatct ggataaaacc gtagagttct atcgcgccac ctacggcttt gaggtgtttc 4200
atagcgaagt aaacgaagaa cagggcgtgc gtgaagccat gctgaaaatc aacgaaacta 4260
gtgatggtgg ggcgactat ctgcaactgc tggaaaccgac acgcccgac tctacagttg 4320
ctaagtggct ggacaagaat ggccaaggcg ttcacacat tgcgttcggt acggctgatg 4380
tggatcaaga cgcggcagat attaaagata aggggtgtgcg tgttctgtac gaggagccac 4440
gccgtggtag catgggtagc cgtattacgt tcctgcaccc taaagactgt catggtgtgc 4500
tgactgagct ggtcacctct gccccggtcg aaagtccgga acattaaggt acc 4553

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<210> SEQ ID NO 28

<211> LENGTH: 717

<212> TYPE: PRT

<213> ORGANISM: Escherichia coli

<400> SEQUENCE: 28

```

Met Ser Arg Met Ser Asn Val Gln Glu Trp Gln Gln Leu Ala Asn Lys
1          5          10          15
Glu Leu Ser Arg Arg Glu Lys Thr Val Asp Ser Leu Val His Gln Thr
20          25          30
Ala Glu Gly Ile Ala Ile Lys Pro Leu Tyr Thr Glu Ala Asp Leu Asp
35          40          45
Asn Leu Glu Val Thr Gly Thr Leu Pro Gly Leu Pro Pro Tyr Val Arg
50          55          60
Gly Pro Arg Ala Thr Met Tyr Thr Ala Gln Pro Trp Thr Ile Arg Gln
65          70          75          80
Tyr Ala Gly Phe Ser Thr Ala Lys Glu Ser Asn Ala Phe Tyr Arg Arg
85          90          95
Asn Leu Ala Ala Gly Gln Lys Gly Leu Ser Val Ala Phe Asp Leu Ala
100         105         110
Thr His Arg Gly Tyr Asp Ser Asp Asn Pro Arg Val Ala Gly Asp Val
115        120        125
Gly Lys Ala Gly Val Ala Ile Asp Thr Val Glu Asp Met Lys Val Leu
130        135        140
Phe Asp Gln Ile Pro Leu Asp Lys Met Ser Val Ser Met Thr Met Asn
145        150        155        160
Gly Ala Val Leu Pro Val Leu Ala Phe Tyr Ile Val Ala Ala Glu Glu
165        170        175
Gln Gly Val Thr Pro Asp Lys Leu Thr Gly Thr Ile Gln Asn Asp Ile
180        185        190
Leu Lys Glu Tyr Leu Cys Arg Asn Thr Tyr Ile Tyr Pro Pro Lys Pro
195        200        205
Ser Met Arg Ile Ile Ala Asp Ile Ile Ala Trp Cys Ser Gly Asn Met
210        215        220

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Pro Arg Phe Asn Thr Ile Ser Ile Ser Gly Tyr His Met Gly Glu Ala
 225 230 235 240
 Gly Ala Asn Cys Val Gln Gln Val Ala Phe Thr Leu Ala Asp Gly Ile
 245 250 255
 Glu Tyr Ile Lys Ala Ala Ile Ser Ala Gly Leu Lys Ile Asp Asp Phe
 260 265 270
 Ala Pro Arg Leu Ser Phe Phe Phe Gly Ile Gly Met Asp Leu Phe Met
 275 280 285
 Asn Val Ala Met Leu Arg Ala Ala Arg Tyr Leu Trp Ser Glu Ala Val
 290 295 300
 Ser Gly Phe Gly Ala Gln Asp Pro Lys Ser Leu Ala Leu Arg Thr His
 305 310 315 320
 Cys Gln Thr Ser Gly Trp Ser Leu Thr Glu Gln Asp Pro Tyr Asn Asn
 325 330 335
 Val Ile Arg Thr Thr Ile Glu Ala Leu Ala Ala Thr Leu Gly Gly Thr
 340 345 350
 Gln Ser Leu His Thr Asn Ala Phe Asp Glu Ala Leu Gly Leu Pro Thr
 355 360 365
 Asp Phe Ser Ala Arg Ile Ala Arg Asn Thr Gln Ile Ile Ile Gln Glu
 370 375 380
 Glu Ser Glu Leu Cys Arg Thr Val Asp Pro Leu Ala Gly Ser Tyr Tyr
 385 390 395 400
 Ile Glu Ser Leu Thr Asp Gln Ile Val Lys Gln Ala Arg Ala Ile Ile
 405 410 415
 Gln Gln Ile Asp Glu Ala Gly Gly Met Ala Lys Ala Ile Glu Ala Gly
 420 425 430
 Leu Pro Lys Arg Met Ile Glu Glu Ala Ser Ala Arg Glu Gln Ser Leu
 435 440 445
 Ile Asp Gln Gly Lys Arg Val Ile Val Gly Val Asn Lys Tyr Lys Leu
 450 455 460
 Asp His Glu Asp Glu Thr Asp Val Leu Glu Ile Asp Asn Val Met Val
 465 470 475 480
 Arg Asn Glu Gln Ile Ala Ser Leu Glu Arg Ile Arg Ala Thr Arg Asp
 485 490 495
 Asp Ala Ala Val Thr Ala Ala Leu Asn Ala Leu Thr His Ala Ala Gln
 500 505 510
 His Asn Glu Asn Leu Leu Ala Ala Val Asn Ala Ala Arg Val Arg
 515 520 525
 Ala Thr Leu Gly Glu Ile Ser Asp Ala Leu Glu Val Ala Phe Asp Arg
 530 535 540
 Tyr Leu Val Pro Ser Gln Cys Val Thr Gly Val Ile Ala Gln Ser Tyr
 545 550 555 560
 His Gln Ser Glu Lys Ser Ala Ser Glu Phe Asp Ala Ile Val Ala Gln
 565 570 575
 Thr Glu Gln Phe Leu Ala Asp Asn Gly Arg Arg Pro Arg Ile Leu Ile
 580 585 590
 Ala Lys Met Gly Gln Asp Gly His Asp Arg Gly Ala Lys Val Ile Ala
 595 600 605
 Ser Ala Tyr Ser Asp Leu Gly Phe Asp Val Asp Leu Ser Pro Met Phe
 610 615 620
 Ser Thr Pro Glu Glu Ile Ala Arg Leu Ala Val Glu Asn Asp Val His

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625	630	635	640
Val Val Gly Ala Ser Ser Leu Ala Ala Gly His Lys Thr Leu Ile Pro	645	650	655
Glu Leu Val Glu Ala Leu Lys Lys Trp Gly Arg Glu Asp Ile Cys Val	660	665	670
Val Ala Gly Gly Val Ile Pro Pro Gln Asp Tyr Ala Phe Leu Gln Glu	675	680	685
Arg Gly Val Ala Ala Ile Tyr Gly Pro Gly Thr Pro Met Leu Asp Ser	690	695	700
Val Arg Asp Val Leu Asn Leu Ile Ser Gln His His Asp	705	710	715

<210> SEQ ID NO 29

<211> LENGTH: 2166

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic nucleotide

<400> SEQUENCE: 29

```

ggatccatgt ctagaatgag caacgtgcag gaatggcagc agctggcga taaagaactg      60
agccgtcgcg aaaaaacggt tgattctctg gtgcatcaga ccgccgaagg tategcaatt    120
aaaccgctgt ataccgaagc ggatctggat aacctggaag tgaccggtac gctgcccgggt    180
ctgccgccgt atgttctgtg tccgcgtgcg accatgtaca cggcacagcc gtggacgatt    240
cgtcagtatg cgggcttcag caccgcaaaa gaatctaacg cattttaccg tcgcaatctg    300
gcgccggggtc agaaaaggct gagcgtggcg tttgatctgg cccccaccg tggttacgat    360
tctgataaac cgcgcgttgc gggcgatgtg ggtaaagcag gcgttgcgat cgatacggtg    420
gaagatatga aagttctgtt cgatcagatt cgcctggata aatgagtgtagcatgacc      480
atgaatggcg cggttctgcc ggtgctggcc ttttatatcg tggcagcggga agaacagggt    540
gttacgccgg ataaactgac cggcaogatc cagaacgata ttctgaaaga atacctgtgc    600
cgtaataacct atatttaccg gccgaaaccg tctatgcgca ttatcgcaga tattatcgcg    660
tggtgtagtg gtaacatgcc gcgtttcaat acgatctcta ttagtggcta tcatatgggt    720
gaagccggcg caaactgcgt tcagcagggt gcctttaccg tggcagatgg tategaatac    780
attaaagccg caatcagtg cggcctgaaa attgatgatt tcgccccgcg cctgagcttt    840
ttctttggca ttggtatgga tctgtttatg aatgtggcca tgctgcgtgc ggcccgtat     900
ctgtggagcg aagcagtttc tggctttggc ggcagagacc cgaaaagcct ggcactgcgt    960
accatttgcc agacgagtgg ttggagcctg accgaacagg acccgtacaa caatgtgatc   1020
cgcaccacga ttgaagcgct ggcagcaacc ctgggtggtg cgcagagcct gcacaccaac   1080
gcgttcgatg aagccctggg tctgcogacg gatttttagcg cccgtatcgc acgcaatacc   1140
cagattatca ttcaggaaga atctgaactg tgcgtacagg ttgatccgct ggcgggcagt   1200
tattacatcg aaagcctgac cgatcagatt gttaaacagg cgcgtgcgat cattcagcag   1260
attgatgaag caggcgggat ggcaaaagcg atcgaagcgg gcctgccgaa acgtatgatt   1320
gaagaagcct ctgcacgcga acagagtctg atcgatcagg gtaaacgtgt gattgttggc   1380
gtgaacaaat aaaaactgga tcatgaagat gaaaccgatg tgctggaaat cgataacggt   1440
atggtgcgta atgaacagat cgccagcctg gaacgtattc gcgcaaccgg cgatgatgcc   1500

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gcagttacgg cggccctgaa cgcactgacc catgcagcgc agcacaacga aaatctgctg 1560
gccgcagcgg tgaatgccgc acgtgttcgc gcgacgctgg gtgaaatttc tgatgcactg 1620
gaagtggcgt tcgatcgcta tctggttccg agtcagtgcg ttaccggcgt gatcgcccag 1680
agttaccatc agagcgaaaa aagcgcatct gaatttgatg cgattgtggc ccagaccgaa 1740
cagtttctgg cagataacgg ccgtcgcccg cgtatcctga ttgccaaaat gggtcaggat 1800
ggccacgatc gcggtgcgaa agtgatcgcg tctgcctata gtgatctggg cttcgatggt 1860
gatctgtctc cgatgtttag tacgcccgaa gaaattgcac gtctggcggg tgaanaatgat 1920
gtgcatgtgg ttggtgccag ctctctggcg gcgggtcaca aaaccctgat tccggaactg 1980
gtggaagcgc tgaanaaatg gggtcgcgaa gatatctgtg tggttgcggg cggtgtgatt 2040
ccgcccagc attatcgctt tctgcaagaa cgtgggtgtg cagcaatcta cggtcggggc 2100
accccgatgc tggatagtgt tcgcatgtg ctgaatctga ttagccagca tcacgattaa 2160
gagctc 2166

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<210> SEQ ID NO 30
<211> LENGTH: 1206
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Synthetic nucleotide

```

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<400> SEQUENCE: 30

```

```

ggatccatga aaataaaaac aggtgcacgc atcctcgcac tatccgcatt aacgacgatg 60
atgttttccg cctcggctct cgccaaaatc gaagaaggta aactggtaat ctggattaaac 120
ggcgataaag gctataacgg tctcgtgaa gtcggtaaga aattcgagaa agataccgga 180
attaagtca ccggttagca tccggataaa ctggaagaga aattcccaca ggttgcggca 240
actggcgatg gccctgacat tatcttctgg gcacacgacc gctttggtgg ctacgctcaa 300
tctggcctgt tgggtgaaat cccccggac aaagcgttcc aggacaagct gtatccgttt 360
acctgggatg ccgtacgta caacggcaag ctgattgctt acccgatcgc tgttgaagcg 420
ttatcgtga ttataaaca agacgtgctg ccgaaccgc caaaaactg ggaagagatc 480
ccggcgctgg ataaagaact gaaagcgaaa ggtaagagcg cgctgatgtt caacctgcaa 540
gaaccgtact tcacctggcc gctgattgct gctgacgggg gttatgcggt caagatgaa 600
aacggcaagt acgacattaa agacgtgggc gtggataacg ctggcgcgaa agcgggtctg 660
accttctcgg ttgacctgat taaaaaaca cacatgaatg cagacaccga ttaactccatc 720
gcagaagctg ctttaataa aggcgaaaca gcgatgacca tcaacggccc gtgggcatgg 780
tccaacatcg acaccagcaa agtgaattat ggtgtaacgg tactgccgac cttcaagggt 840
caaccatcca aaccgttcgt tggcgtgctg agcgcaggta ttaacggcgc cagtccgaac 900
aaagagctgg cgaagaggtt cctcgaaaac tatctgctga ctgatgaagg tctggaagcg 960
gttaataaag acaaaccgct ggggtgccga gcgctgaagt cttacgagga agagttggcg 1020
aaagatccac gtattgccgc caccatggaa aacgcccaga aagtgaaat catgccgaac 1080
atcccgcaga tgtccgcttt ctggtatgcc gtgcgtactg cggtgatcaa cgccgccagc 1140
ggtcgtcaga ctgctgatga agccctgaaa gacgcgcaga ctcgtatcac caagtctaga 1200
gagctc 1206

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<210> SEQ ID NO 31
 <211> LENGTH: 38
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic primer

 <400> SEQUENCE: 31

 gagaggtacc atgggggggtt ctcatcatca tcatcatc 38

<210> SEQ ID NO 32
 <211> LENGTH: 38
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic primer

 <400> SEQUENCE: 32

 cagccaagct tttattacgc accctgtgcg cgctgttc 38

<210> SEQ ID NO 33
 <211> LENGTH: 1263
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic nucleotide

 <400> SEQUENCE: 33

 atgggggggtt ctcatcatca tcatcatcat ggtatggcta gcatgactgg tggacagcaa 60
 atgggtcggg atctgtacga cgatgacgat aaggatcgat ggggatccct ggtggaaggc 120
 ctgcgtagaag ttgccgatgg tgatgcactg tatgatgcag cagtgggtca tggcgatcgt 180
 ggtccggttt ggggtgttag cggccagggt tctcagtggg cagcgatggg caccagctg 240
 ctggcaagcg aaccggtttt tgccgcaacg attgcaaac tggaaaccgg gatcgcggcc 300
 gaaagtggct tcagcgttac cgaagcaatt acggcgcagc agaccgtgac gggtatcgat 360
 aaagtgcagc cggccgtttt cgcagttcag gtggcgctgg cagcgacgat ggaacagacg 420
 tacggcggtc gtccgggtgc agtggttggc cacagtatgg gtgaaagcgc cgcagcggtg 480
 gttgcaggcg ccctgagtct ggaagatgcc gcacgtgtga tttgccgtcg cagcaaaactg 540
 atgaccgta tcgcaggtgc aggtgcgatg ggcagcgtgg aactgccggc aaaacaggtt 600
 aactctgaac tgatggcgcg cggatattgat gatgtggttg tgtctgttgt ggcgtctccg 660
 cagagtaccg tgattggcgg caccagtgat acggttcgtg atctgatcgc gcgttgggaa 720
 cagcgcgatg tgatggcgcg cgaagttgcc gtggatgttg caagccattc tccgcaggtt 780
 gatccgatte tggatgatct ggccggcgca ctggcagata ttgcaccgat gaccccgaaa 840
 gtgccgtatt acagcgcgac gctgtttgat ccgcgtgaac agccggtgtg tgatggcgcc 900
 tattgggttg ataacctgcg caataccgtg cagtttgcgg cggcagttca ggcggcgatg 960
 gaagatgggt accgtgtggt cgccgaaactg tctccgcatc cgctgctgac ccacgcagtg 1020
 gaacagacgg gtcgctctct ggatatgagt gttgcagcac tggccggtat gcgtcgcgaa 1080
 cagccgctgc cgcattggcct gcgtggtctg ctgaccgaac tgcaccgtgc aggtgcagca 1140
 ctggattata gcgcaactgta cccggcaggt cgtctggttg atgcaccgct gccggcatgg 1200
 acgcacgcac gtctgttcat cgatgatgat ggccaggaac agcgcgcaca ggtgtcgtaa 1260

-continued

taa

1263

<210> SEQ ID NO 34

<211> LENGTH: 419

<212> TYPE: PRT

<213> ORGANISM: Mycobacterium bovis

<400> SEQUENCE: 34

Met Gly Gly Ser His His His His His His Gly Met Ala Ser Met Thr
1 5 10 15

Gly Gly Gln Gln Met Gly Arg Asp Leu Tyr Asp Asp Asp Asp Lys Asp
20 25 30

Arg Trp Gly Ser Leu Val Glu Gly Leu Arg Glu Val Ala Asp Gly Asp
35 40 45

Ala Leu Tyr Asp Ala Ala Val Gly His Gly Asp Arg Gly Pro Val Trp
50 55 60

Val Phe Ser Gly Gln Gly Ser Gln Trp Ala Ala Met Gly Thr Gln Leu
65 70 75 80

Leu Ala Ser Glu Pro Val Phe Ala Ala Thr Ile Ala Lys Leu Glu Pro
85 90 95

Val Ile Ala Ala Glu Ser Gly Phe Ser Val Thr Glu Ala Ile Thr Ala
100 105 110

Gln Gln Thr Val Thr Gly Ile Asp Lys Val Gln Pro Ala Val Phe Ala
115 120 125

Val Gln Val Ala Leu Ala Ala Thr Met Glu Gln Thr Tyr Gly Val Arg
130 135 140

Pro Gly Ala Val Val Gly His Ser Met Gly Glu Ser Ala Ala Ala Val
145 150 155 160

Val Ala Gly Ala Leu Ser Leu Glu Asp Ala Ala Arg Val Ile Cys Arg
165 170 175

Arg Ser Lys Leu Met Thr Arg Ile Ala Gly Ala Gly Ala Met Gly Ser
180 185 190

Val Glu Leu Pro Ala Lys Gln Val Asn Ser Glu Leu Met Ala Arg Gly
195 200 205

Ile Asp Asp Val Val Val Ser Val Val Ala Ser Pro Gln Ser Thr Val
210 215 220

Ile Gly Gly Thr Ser Asp Thr Val Arg Asp Leu Ile Ala Arg Trp Glu
225 230 235 240

Gln Arg Asp Val Met Ala Arg Glu Val Ala Val Asp Val Ala Ser His
245 250 255

Ser Pro Gln Val Asp Pro Ile Leu Asp Asp Leu Ala Ala Ala Leu Ala
260 265 270

Asp Ile Ala Pro Met Thr Pro Lys Val Pro Tyr Tyr Ser Ala Thr Leu
275 280 285

Phe Asp Pro Arg Glu Gln Pro Val Cys Asp Gly Ala Tyr Trp Val Asp
290 295 300

Asn Leu Arg Asn Thr Val Gln Phe Ala Ala Ala Val Gln Ala Ala Met
305 310 315 320

Glu Asp Gly Tyr Arg Val Phe Ala Glu Leu Ser Pro His Pro Leu Leu
325 330 335

Thr His Ala Val Glu Gln Thr Gly Arg Ser Leu Asp Met Ser Val Ala
340 345 350

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Ala Leu Ala Gly Met Arg Arg Glu Gln Pro Leu Pro His Gly Leu Arg
 355 360 365

Gly Leu Leu Thr Glu Leu His Arg Ala Gly Ala Ala Leu Asp Tyr Ser
 370 375 380

Ala Leu Tyr Pro Ala Gly Arg Leu Val Asp Ala Pro Leu Pro Ala Trp
 385 390 395 400

Thr His Ala Arg Leu Phe Ile Asp Asp Asp Gly Gln Glu Gln Arg Ala
 405 410 415

Gln Gly Ala

<210> SEQ ID NO 35
 <211> LENGTH: 464
 <212> TYPE: DNA
 <213> ORGANISM: Kribbella flavida DSM

<400> SEQUENCE: 35

gagctcagga ggaattaacc atggaacacc tgacggcgac ccagaccctg tttgaagcga 60
 ttgaccacgt tggcgttgca gttgeggatt ttgatgaagc agtgcgtttt tatgcagaaa 120
 ccttcggcat gacggtgctt catgaagaag ttaacgaaga acagggtggt cgtgaagcaa 180
 tgctgtcaat tggcgattcg ggtagctcta tccaactgct ggcgcccgtg tccgatagtt 240
 ccccgattgc caaatttctg gaccgcaatg gcccggtat ccagcaactg gcctatcgtg 300
 tccgcatct ggacgcagt ggcgcaacc tgcgtgaacg tggcgcgcaa ctgctgtacg 360
 acgaaccgcg tcgcgcaacg gctggttctc gtattaactt cattcatccg aaatcggcgg 420
 gcggcgtcct ggtggaactg gtggaaccgg ctcgctaact gcag 464

<210> SEQ ID NO 36
 <211> LENGTH: 145
 <212> TYPE: PRT
 <213> ORGANISM: Kribbella flavida DSM

<400> SEQUENCE: 36

Met Glu His Leu Thr Ala Thr Gln Thr Leu Phe Glu Ala Ile Asp His
 1 5 10 15

Val Gly Val Ala Val Ala Asp Phe Asp Glu Ala Val Arg Phe Tyr Ala
 20 25 30

Glu Thr Phe Gly Met Thr Val Ala His Glu Glu Val Asn Glu Glu Gln
 35 40 45

Gly Val Arg Glu Ala Met Leu Ser Ile Gly Asp Ser Gly Ser Ser Ile
 50 55 60

Gln Leu Leu Ala Pro Leu Ser Asp Ser Ser Pro Ile Ala Lys Phe Leu
 65 70 75 80

Asp Arg Asn Gly Pro Gly Ile Gln Gln Leu Ala Tyr Arg Val Arg Asp
 85 90 95

Leu Asp Ala Val Ser Ala Thr Leu Arg Glu Arg Gly Ala Gln Leu Leu
 100 105 110

Tyr Asp Glu Pro Arg Arg Gly Thr Ala Gly Ser Arg Ile Asn Phe Ile
 115 120 125

His Pro Lys Ser Ala Gly Gly Val Leu Val Glu Leu Val Glu Pro Ala
 130 135 140

Arg
 145

-continued

<210> SEQ ID NO 37
 <211> LENGTH: 545
 <212> TYPE: DNA
 <213> ORGANISM: Sorangium cellulosum

<400> SEQUENCE: 37

```

gagctcagga ggaattaacc atggctccgc cggcaacgcg tccggctccg gctgcaccga    60
cgggcctgcc gacccaacgt gaaccgatga aagaccagat tccgggcttt ctgttcattg    120
atcatatcgc gatggccgtg cgggcaggcc aactggacgc acaagttaa gcctatgaaa    180
tgctgggctt tcgtgaagtt catcgcaag aagtccgtgg tgcggatcag gtgcgcaag    240
ttatgctgcg tattggtgat agcgacaacc acgtccaact gctggaaccg ctgagcccgg    300
aatctccggt tcaaaaactg atcgagaaaa acggcggtcg cggcggttgc gcacatgtgg    360
cttaccgtgt cagtgatgtg caagcggcct ttgacgaact gaaagcgcgt ggcttccgca    420
ttatcgatgc agctccgctg ccgggcagcc gtggcaccac gattttcttt gttcaccgcg    480
gctcacgcga cgatgcccg ttcggtcacc tgattgaagt tgtccagtca catggctaac    540
tgcag                                                                    545
  
```

<210> SEQ ID NO 38
 <211> LENGTH: 172
 <212> TYPE: PRT
 <213> ORGANISM: Sorangium cellulosum

<400> SEQUENCE: 38

```

Met Ala Pro Pro Ala Thr Arg Pro Ala Pro Ala Ala Pro Thr Gly Leu
1          5          10          15
Pro Thr Gln Arg Glu Pro Met Lys Asp Gln Ile Pro Gly Phe Leu Phe
          20          25          30
Ile Asp His Ile Ala Met Ala Val Pro Ala Gly Gln Leu Asp Ala Gln
          35          40          45
Val Lys Ala Tyr Glu Met Leu Gly Phe Arg Glu Val His Arg Glu Glu
          50          55          60
Val Arg Gly Ala Asp Gln Val Arg Glu Val Met Leu Arg Ile Gly Asp
65          70          75          80
Ser Asp Asn His Val Gln Leu Leu Glu Pro Leu Ser Pro Glu Ser Pro
          85          90          95
Val Gln Lys Leu Ile Glu Lys Asn Gly Gly Arg Gly Gly Phe Ala His
          100          105          110
Val Ala Tyr Arg Val Ser Asp Val Gln Ala Ala Phe Asp Glu Leu Lys
          115          120          125
Ala Arg Gly Phe Arg Ile Ile Asp Ala Ala Pro Arg Pro Gly Ser Arg
          130          135          140
Gly Thr Thr Ile Phe Phe Val His Pro Arg Ser Arg Asp Asp Ala Pro
145          150          155          160
Phe Gly His Leu Ile Glu Val Val Gln Ser His Gly
          165          170
  
```

<210> SEQ ID NO 39
 <211> LENGTH: 26
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:

-continued

<223> OTHER INFORMATION: Synthetic primer

<400> SEQUENCE: 39

taagagctca ggaggaatta accatg 26

<210> SEQ ID NO 40

<211> LENGTH: 566

<212> TYPE: DNA

<213> ORGANISM: Escherichia coli

<400> SEQUENCE: 40

catgccatgg cggacacggt attgattctg ggtgatagcc tgagcgcgg gtatcgaatg 60

tctgccagcg cggcctggcc tgccttgttg aatgataagt ggcagagtaa aacgtcggta 120

gttaatgcc gcatcagcgg cgacacctcg caacaaggac tggcgcgcct tccggctctg 180

ctgaaacagc atcagcccg tgggtgctg gttgaactgg gcggcaatga cggtttgcgt 240

ggttttcagc cacagcaaac cgagcaaac ctgcgccaga ttttcagga tgtcaaagcc 300

gccaacgctg aaccattggt aatgcaata cgtctgcctg caaactatgg tccgcttat 360

aatgaagcct ttagcgcct ttaccccaaa ctcgccaag agtttgatgt tccgctgctg 420

cccttttta tggagaggt ctacctcaag ccacaatgga tgcaggatga cggattcat 480

cccaccgcg acgccagcc gtttattgcc gactggatgg cgaagcagtt gcagcctta 540

gtaaatcatg actcataagg atccgc 566

<210> SEQ ID NO 41

<211> LENGTH: 32

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic primer

<400> SEQUENCE: 41

catgccatgg cggacacggt attgattctg gg 32

<210> SEQ ID NO 42

<211> LENGTH: 9

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic peptide

<400> SEQUENCE: 42

Met Ala Asp Thr Leu Leu Ile Leu Gly

1 5

<210> SEQ ID NO 43

<211> LENGTH: 36

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Synthetic primer

<400> SEQUENCE: 43

gcggatcctt atgagtcag atttactaaa ggctgc 36

<210> SEQ ID NO 44

<211> LENGTH: 9

<212> TYPE: PRT

<213> ORGANISM: Artificial Sequence

-continued

<220> FEATURE:
 <223> OTHER INFORMATION: Synthetic peptide

<400> SEQUENCE: 44

Ser Asp His Asn Val Leu Pro Gln Leu
 1 5

<210> SEQ ID NO 45
 <211> LENGTH: 183
 <212> TYPE: PRT
 <213> ORGANISM: Escherichia coli

<400> SEQUENCE: 45

Met Ala Asp Thr Leu Leu Ile Leu Gly Asp Ser Leu Ser Ala Gly Tyr
 1 5 10 15

Arg Met Ser Ala Ser Ala Ala Trp Pro Ala Leu Leu Asn Asp Lys Trp
 20 25 30

Gln Ser Lys Thr Ser Val Val Asn Ala Ser Ile Ser Gly Asp Thr Ser
 35 40 45

Gln Gln Gly Leu Ala Arg Leu Pro Ala Leu Leu Lys Gln His Gln Pro
 50 55 60

Arg Trp Val Leu Val Glu Leu Gly Gly Asn Asp Gly Leu Arg Gly Phe
 65 70 75 80

Gln Pro Gln Gln Thr Glu Gln Thr Leu Arg Gln Ile Leu Gln Asp Val
 85 90 95

Lys Ala Ala Asn Ala Glu Pro Leu Leu Met Gln Ile Arg Leu Pro Ala
 100 105 110

Asn Tyr Gly Arg Arg Tyr Asn Glu Ala Phe Ser Ala Ile Tyr Pro Lys
 115 120 125

Leu Ala Lys Glu Phe Asp Val Pro Leu Leu Pro Phe Phe Met Glu Glu
 130 135 140

Val Tyr Leu Lys Pro Gln Trp Met Gln Asp Asp Gly Ile His Pro Asn
 145 150 155 160

Arg Asp Ala Gln Pro Phe Ile Ala Asp Trp Met Ala Lys Gln Leu Gln
 165 170 175

Pro Leu Val Asn His Asp Ser
 180

<210> SEQ ID NO 46
 <211> LENGTH: 33
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic primer

<400> SEQUENCE: 46

cattactcga ggcactccc gttctggata atg

33

<210> SEQ ID NO 47
 <211> LENGTH: 35
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic primer

<400> SEQUENCE: 47

gggaagctta tgagtcatga ttactaaag gctgc

35

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<210> SEQ ID NO 48
 <211> LENGTH: 9
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic peptide

<400> SEQUENCE: 48

Ser Asp His Asn Val Leu Pro Gln Leu
 1 5

<210> SEQ ID NO 49
 <211> LENGTH: 15
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic nucleotide

<400> SEQUENCE: 49

ggatccatgt ctaga

15

<210> SEQ ID NO 50
 <211> LENGTH: 34
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Synthetic peptide

<400> SEQUENCE: 50

Met Gly Gly Ser His His His His His His Gly Met Ala Ser Met Thr
 1 5 10 15

Gly Gly Gln Gln Met Gly Arg Thr Asp Asp Asp Asp Lys Asp Arg Trp
 20 25 30

Gly Ser

<210> SEQ ID NO 51
 <211> LENGTH: 655
 <212> TYPE: PRT
 <213> ORGANISM: Ehrlichia chaffeensis
 <300> PUBLICATION INFORMATION:
 <308> DATABASE ACCESSION NUMBER: NCBI / YP_507303
 <309> DATABASE ENTRY DATE: 2010-05-14
 <313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(655)

<400> SEQUENCE: 51

Met Ile Lys Lys Ile Leu Ile Ala Asn Arg Gly Glu Ile Ala Cys Arg
 1 5 10 15

Val Met Arg Thr Ala Arg Lys Met Gly Ile Ser Cys Val Ala Val Tyr
 20 25 30

Ser Asn Ala Asp Val Tyr Ser Leu His Val Leu Ser Ala Glu Glu Ala
 35 40 45

Val Asn Ile Gly Pro Ala Pro Val Asn Gln Ser Tyr Leu Asn Met Glu
 50 55 60

Lys Ile Cys Glu Val Ala Cys Asn Thr Gly Val Asp Ala Val His Pro
 65 70 75 80

Gly Tyr Gly Phe Leu Ser Glu Asn Ala Asp Phe Pro Glu Lys Leu Glu
 85 90 95

Gln Tyr Asn Ile Lys Phe Ile Gly Pro Ser Ser Thr Ser Ile Arg Met
 100 105 110

-continued

Met	Ala	Asp	Lys	Ile	Thr	Ser	Lys	Lys	Ile	Ala	Glu	Ser	Ala	Lys	Val
		115					120					125			
Asn	Ile	Ile	Pro	Gly	Tyr	Met	Gly	Ile	Val	Asp	Ser	Val	His	Glu	Ala
	130					135					140				
Lys	Glu	Ile	Ala	Lys	Ser	Ile	Gly	Phe	Pro	Val	Met	Ile	Lys	Ala	Thr
145					150					155					160
Ala	Gly	Gly	Gly	Gly	Lys	Gly	Met	Arg	Ile	Val	Lys	Ser	Ser	Glu	Glu
				165					170						175
Ile	Glu	Gln	Ala	Phe	Thr	Ser	Ala	Thr	Asn	Glu	Ala	Ala	Lys	Asn	Phe
			180					185						190	
Arg	Asp	Gly	Arg	Ile	Phe	Ile	Glu	Lys	Tyr	Val	Glu	Leu	Pro	Arg	His
		195					200					205			
Ile	Glu	Ile	Gln	Ile	Ile	Ala	Asp	Lys	His	Gly	Asn	Ile	Val	Cys	Leu
	210					215					220				
Gly	Glu	Arg	Glu	Cys	Ser	Ile	Gln	Arg	His	Asn	Gln	Lys	Val	Ile	Glu
225					230					235					240
Glu	Thr	Pro	Ser	Pro	Phe	Leu	Asp	Glu	Glu	Thr	Arg	Gln	Lys	Met	Tyr
				245						250					255
Gln	Gln	Cys	Val	Asn	Leu	Ala	Lys	Lys	Val	Gly	Tyr	Tyr	Ser	Ala	Gly
			260					265						270	
Thr	Ile	Glu	Phe	Ile	Val	Asp	Gln	Asp	Lys	Gln	Phe	Tyr	Phe	Leu	Glu
		275					280					285			
Met	Asn	Thr	Arg	Leu	Gln	Val	Glu	His	Pro	Val	Thr	Glu	Leu	Val	Thr
		290				295					300				
Gly	Ile	Asp	Ile	Val	Glu	Glu	Met	Ile	Arg	Ile	Ala	Asp	Gly	Glu	Glu
305					310					315					320
Leu	Arg	Phe	Thr	Gln	Gln	Asp	Val	Lys	Phe	Thr	Gly	Ser	Ala	Ile	Glu
				325					330						335
Ala	Arg	Val	Tyr	Ala	Glu	Asn	Pro	Thr	Lys	Asn	Phe	Leu	Pro	Ser	Ser
			340					345						350	
Gly	Arg	Ile	Ala	Tyr	Tyr	Ser	Ala	Pro	Met	Pro	Asn	Asp	Asn	Leu	Arg
		355					360					365			
Ile	Asp	Ser	Gly	Val	Phe	Glu	Gly	Ala	Glu	Val	Ser	Met	Phe	Tyr	Asp
	370					375						380			
Pro	Met	Ile	Ala	Lys	Val	Cys	Thr	Tyr	Gly	Lys	Asn	Arg	Asp	Glu	Ala
385					390					395					400
Val	Ser	Phe	Met	Gln	Arg	Tyr	Leu	Asn	Glu	Phe	Tyr	Ile	Gly	Gly	Ile
				405					410						415
Ala	Asn	Asn	Ile	Asp	Phe	Leu	Leu	Ser	Val	Phe	His	His	Pro	Val	Phe
			420					425						430	
Ile	Ser	Gly	Asn	Ile	Asn	Thr	Lys	Phe	Ile	Glu	Gln	Phe	Tyr	Phe	Asp
		435					440						445		
Gly	Phe	Gln	Gly	Asn	Pro	Leu	Thr	Lys	Ala	Cys	Ile	Lys	Leu	Phe	Ile
	450					455					460				
Leu	Thr	Ser	Leu	Cys	Ile	Phe	Phe	Gln	Asp	Glu	Tyr	Gly	Ile	His	Gly
465					470					475					480
Val	Glu	Leu	Cys	Glu	Asn	Arg	Glu	Leu	Ala	Val	Tyr	Val	Asp	Gly	Gln
				485					490					495	
Lys	Tyr	Leu	Ile	Ser	Ala	Lys	Tyr	Glu	Asn	Gly	Arg	Val	Leu	Ala	Ile
		500						505						510	
Tyr	Asp	Gln	Cys	Glu	Tyr	Leu	Val	Val	Ser	Thr	Trp	Asn	Val	Asn	Phe

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Gln Glu Asp Leu Gly Gly Ala Lys Val His Ala Ser Lys Thr Gly Ile
 210 215 220
 Ala Asp Leu Val Phe His Asn Glu Ile Glu Ala Leu Leu Gln Val Arg
 225 230 235 240
 Arg Phe Met Asn Phe Ile Pro Ser Asn Asn Met Glu Ser Ile Gly Ser
 245 250 255
 Gln Ser Ala Ser Asn Phe Ile Asn Met Glu Asp Leu Ser Leu Asn Thr
 260 265 270
 Leu Val Pro Lys Asn Ser Thr Thr Pro Tyr Asn Met Tyr Glu Leu Leu
 275 280 285
 Glu Lys Val Cys Asp Glu Arg Leu Phe Tyr Glu Ile Lys Pro Asp Phe
 290 295 300
 Ala Arg Asn Ile Ile Ile Gly Phe Gly Lys Ile Gly Gly Tyr Asn Val
 305 310 315 320
 Gly Leu Val Ala Asn Gln Pro Leu His Leu Ala Gly Cys Leu Asp Ile
 325 330 335
 Asp Ala Ser Arg Lys Gly Ala Arg Phe Ile Arg Phe Cys Asp Ala Phe
 340 345 350
 Asn Ile Pro Val Ile Thr Phe Ile Asp Val Pro Gly Phe Met Pro Gly
 355 360 365
 Val Asn Gln Glu His Ser Gly Ile Ile Ala His Gly Ala Lys Leu Leu
 370 375 380
 Tyr Ala Tyr Ala Glu Ala Thr Val Pro Lys Ile Ser Val Ile Val Arg
 385 390 395 400
 Lys Ala Tyr Gly Gly Ala Tyr Ile Val Met Asn Ser Lys His Leu Cys
 405 410 415
 Gly Asp Val Asn Tyr Ala Trp Gln Asp Ala Glu Ile Ala Val Met Gly
 420 425 430
 Ala Glu Gly Ala Val Glu Ile Ile Phe Arg Asn Glu Lys Asp Lys Asp
 435 440 445
 Lys Ile Gln His Ile Ile Asp Glu Tyr Arg Thr Thr Ile Val Asn Pro
 450 455 460
 Tyr Val Ala Ala Ser Arg Gly Tyr Ile Asp Asp Ile Ile Val Pro Ser
 465 470 475 480
 Arg Thr Arg Glu His Leu Phe Lys Ser Leu Gln Phe Leu Glu Lys Lys
 485 490 495
 Lys Val His Lys Ile Met Arg Lys His Asp Asn Leu Pro Leu
 500 505 510

<210> SEQ ID NO 53
 <211> LENGTH: 666
 <212> TYPE: PRT
 <213> ORGANISM: Agrobacterium vitis
 <300> PUBLICATION INFORMATION:
 <308> DATABASE ACCESSION NUMBER: NCBI / YP_002547482
 <309> DATABASE ENTRY DATE: 2010-04-01
 <313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(666)

<400> SEQUENCE: 53

Met Ala Ile Ser Lys Ile Leu Ile Ala Asn Arg Gly Glu Ile Ala Cys
 1 5 10 15
 Arg Val Ile Lys Thr Ala Lys Arg Met Gly Ile Ala Thr Val Ala Val
 20 25 30

-continued

Tyr Ser Asp Ala Asp Ala Asn Ala Leu His Val Lys Leu Ala Asp Glu
35 40 45
Ala Val His Ile Gly Pro Ser Pro Ser Asn Gln Ser Tyr Ile Val Ile
50 55 60
Asp Lys Ile Leu Glu Ala Ile Arg Gln Thr Gly Ala Asp Ala Val His
65 70 75 80
Pro Gly Tyr Gly Phe Leu Ser Glu Asn Ala Ala Phe Ala Glu Ala Leu
85 90 95
Asp Lys Ala Gly Val Ala Phe Ile Gly Pro Pro Val Gly Ala Ile Lys
100 105 110
Ala Met Gly Asp Lys Ile Thr Ser Lys Lys Leu Ala Ala Glu Ala Gly
115 120 125
Val Ser Thr Val Pro Gly His Met Gly Leu Ile Ala Asp Ala Asp Glu
130 135 140
Ala Val Lys Ile Ala Ala Gln Ile Gly Tyr Pro Val Met Ile Lys Ala
145 150 155 160
Ser Ala Gly Gly Gly Gly Lys Gly Met Arg Ile Ala Trp Asn Asp Ala
165 170 175
Glu Ala Arg Glu Gly Phe Gln Ser Ser Lys Asn Glu Ala Met Asn Ser
180 185 190
Phe Gly Asp Asp Arg Ile Phe Ile Glu Lys Phe Val Asp Gln Pro Arg
195 200 205
His Ile Glu Ile Gln Val Leu Gly Asp Lys His Gly Asn Val Leu Tyr
210 215 220
Leu Gly Glu Arg Glu Cys Ser Ile Gln Arg Arg Asn Gln Lys Val Ile
225 230 235 240
Glu Glu Ala Pro Ser Pro Phe Leu Asp Ala Asp Thr Arg Lys Ala Met
245 250 255
Gly Glu Gln Ala Val Ala Leu Ala Lys Ala Val Gly Tyr Tyr Ser Ala
260 265 270
Gly Thr Val Glu Phe Ile Val Asp Gly Asn Arg Asn Phe Tyr Phe Leu
275 280 285
Glu Met Asn Thr Arg Leu Gln Val Glu His Pro Val Thr Glu Leu Ile
290 295 300
Thr Gly Leu Asp Leu Val Glu Gln Met Ile Arg Val Ala Ser Gly Glu
305 310 315 320
Thr Leu Ala Leu Ala Gln Gly Asp Val Thr Leu Thr Gly Trp Ala Val
325 330 335
Glu Ser Arg Leu Tyr Ala Glu Asp Pro Tyr Arg Asn Phe Leu Pro Ser
340 345 350
Ile Gly Arg Leu Ser Arg Tyr Arg Pro Pro Ser Glu Gly Gln Gln Ala
355 360 365
Asp Gly Thr Val Val Arg Asn Asp Thr Gly Val Phe Glu Gly Gly Glu
370 375 380
Ile Ser Met Tyr Tyr Asp Pro Met Val Ala Lys Leu Cys Thr Trp Gly
385 390 395 400
Pro Asp Arg Ile Thr Ala Ile Asp Ala Met Ser Ala Ala Leu Asp Arg
405 410 415
Phe Glu Val Glu Gly Ile Gly His Asn Leu Pro Phe Leu Ser Ala Val
420 425 430
Met Gln His Pro Arg Phe Arg Ser Gly Lys Ile Thr Thr Ala Phe Ile

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435					440					445					
Ala	Glu	Glu	Phe	Pro	Glu	Gly	Phe	Ser	Gly	Val	Glu	Pro	Asp	Glu	Met
450					455					460					
Ala	Gly	Lys	Thr	Leu	Ala	Ala	Ile	Ala	Ala	Leu	Val	His	Gln	Arg	Arg
465					470					475					480
Glu	Ala	Arg	Ala	Ala	Gln	Val	Ser	Gly	Thr	Met	Gly	Asn	His	Ala	Arg
			485						490					495	
Thr	Ile	Gly	Arg	Asp	Trp	Val	Val	Gly	Leu	Ala	Glu	Gln	Asn	Tyr	Pro
			500					505					510		
Leu	Thr	Leu	Ser	Thr	Asp	Pro	Gly	Ser	Met	Met	Phe	Ala	Asp	Gly	Asn
		515					520					525			
Val	Leu	Ser	Val	Asp	Gly	Val	Trp	Gln	Pro	Gly	Gln	Thr	Leu	Ala	Ile
	530					535					540				
Phe	Thr	Val	Asn	Gly	Gln	Ser	Ile	Gly	Leu	Lys	Ile	Asp	Leu	Lys	Gly
545				550						555					560
Pro	Ala	Ile	Arg	Leu	Arg	Trp	Arg	Gly	Met	Asp	Val	Val	Ala	His	Val
				565					570					575	
Arg	Asn	Pro	Arg	Val	Ala	Glu	Leu	Ala	Arg	Leu	Met	Pro	Arg	Lys	Leu
			580					585					590		
Pro	Pro	Asp	Thr	Ser	Lys	Met	Leu	Leu	Cys	Pro	Met	Pro	Gly	Val	Val
		595					600					605			
Thr	Gly	Ile	Ala	Val	Ala	Glu	Gly	Asp	Ala	Val	Glu	Ala	Gly	Gln	Ala
	610					615					620				
Leu	Ala	Thr	Val	Glu	Ala	Met	Lys	Met	Glu	Asn	Ile	Leu	Lys	Ala	Glu
625					630					635					640
Arg	Arg	Gly	Val	Val	Lys	Arg	Leu	Val	Ala	Lys	Ala	Gly	Gln	Ser	Leu
				645					650					655	
Ala	Val	Asp	Glu	Leu	Ile	Met	Glu	Phe	Glu						
		660					665								

<210> SEQ ID NO 54
 <211> LENGTH: 510
 <212> TYPE: PRT
 <213> ORGANISM: Agrobacterium vitis
 <300> PUBLICATION INFORMATION:
 <308> DATABASE ACCESSION NUMBER: NCBI / YP_002547479
 <309> DATABASE ENTRY DATE: 2010-04-01
 <313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(510)

<400> SEQUENCE: 54

Met	Pro	Thr	Ile	Leu	Asp	Gln	Leu	Glu	Ser	Arg	Arg	Ala	Glu	Ala	Arg
1				5					10					15	
Leu	Gly	Gly	Gly	Glu	Lys	Arg	Ile	Asp	Ala	Gln	His	Ala	Lys	Gly	Lys
			20					25					30		
Leu	Thr	Ala	Arg	Glu	Arg	Ile	Glu	Ile	Leu	Leu	Asp	Glu	Gly	Ser	Phe
		35					40					45			
Glu	Glu	Tyr	Asp	Met	Tyr	Val	Thr	His	Arg	Cys	Ala	Asp	Phe	Gly	Met
	50					55					60				
Asp	Gly	Gln	Lys	Val	Ala	Gly	Asp	Gly	Val	Val	Thr	Gly	Trp	Gly	Thr
65					70					75					80
Ile	Asn	Gly	Arg	Gln	Val	Tyr	Val	Phe	Ser	Gln	Asp	Phe	Thr	Val	Leu
				85					90					95	
Gly	Gly	Ser	Leu	Ser	Glu	Thr	His	Ala	Gln	Lys	Ile	Cys	Lys	Ile	Met
			100					105						110	

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Asp Met Ala Val Arg Val Gly Ala Pro Val Ile Gly Ile Asn Asp Ser
 115 120 125

Gly Gly Ala Arg Ile Gln Glu Gly Val Ala Ser Leu Ala Gly Tyr Ala
 130 135 140

Glu Val Phe Arg Arg Asn Ala Glu Val Ser Gly Val Ile Pro Gln Ile
 145 150 155 160

Ser Val Ile Met Gly Pro Cys Ala Gly Gly Ala Val Tyr Ser Pro Ala
 165 170 175

Met Thr Asp Phe Ile Phe Met Val Arg Asp Thr Ser Tyr Met Phe Val
 180 185 190

Thr Gly Pro Asp Val Val Lys Thr Val Thr Asn Glu Ile Val Thr Ala
 195 200 205

Glu Glu Leu Gly Gly Ala Gly Thr His Thr Lys Lys Ser Ser Val Ala
 210 215 220

Asp Gly Ala Phe Glu Asn Asp Val Glu Ala Leu Glu Gln Val Arg Leu
 225 230 235 240

Leu Phe Asp Phe Leu Pro Leu Asn Asn Arg Glu Lys Pro Pro Lys Arg
 245 250 255

Pro Phe Tyr Asp Asp Pro Ala Arg Leu Glu Met Arg Leu Asp Thr Leu
 260 265 270

Ile Pro Asp Ser Ser Thr Lys Pro Tyr Asp Met Lys Glu Leu Ile His
 275 280 285

Ala Leu Ala Asp Glu Gly Asp Phe Phe Glu Leu Gln Glu Ala Phe Ala
 290 295 300

Lys Asn Ile Ile Thr Gly Phe Ile Arg Leu Glu Gly Gln Thr Val Gly
 305 310 315 320

Val Val Ala Asn Gln Pro Met Val Leu Ala Gly Cys Leu Asp Ile Asp
 325 330 335

Ser Ser Arg Lys Ala Ala Arg Phe Val Arg Phe Cys Asp Ala Phe Ser
 340 345 350

Ile Pro Ile Leu Thr Leu Val Asp Val Pro Gly Phe Leu Pro Gly Val
 355 360 365

Ala Gln Glu Tyr Gly Gly Val Ile Lys His Gly Ala Lys Leu Leu Phe
 370 375 380

Ala Tyr Ser Glu Ala Thr Val Pro Met Val Thr Leu Ile Thr Arg Lys
 385 390 395 400

Ala Tyr Gly Gly Ala Tyr Asp Val Met Ala Ser Lys His Ile Gly Ala
 405 410 415

Asp Val Asn Tyr Ala Trp Pro Thr Ala Glu Ile Ala Val Met Gly Ala
 420 425 430

Lys Gly Ala Thr Glu Ile Leu Tyr Arg Ser Glu Leu Ala Asp Pro Glu
 435 440 445

Lys Ile Ala Ala Arg Thr Arg Glu Tyr Glu Glu Arg Phe Ala Asn Pro
 450 455 460

Phe Val Ala Ala Glu Arg Gly Phe Ile Asp Glu Val Ile Met Pro His
 465 470 475 480

Ser Ser Arg Lys Arg Ile Ala Arg Ala Phe Ala Ser Leu Arg Gly Lys
 485 490 495

Gln Val Ala Thr His Trp Lys Lys His Asp Thr Ile Pro Leu
 500 505 510

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<210> SEQ ID NO 55
<211> LENGTH: 667
<212> TYPE: PRT
<213> ORGANISM: Methylobacterium extorquens
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / YP_003069256
<309> DATABASE ENTRY DATE: 2010-04-16
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(667)

<400> SEQUENCE: 55

Met Phe Asp Lys Ile Leu Ile Ala Asn Arg Gly Glu Ile Ala Cys Arg
 1           5           10           15

Ile Ile Lys Thr Ala Gln Lys Met Gly Ile Lys Thr Val Ala Val Tyr
 20           25           30

Ser Asp Ala Asp Arg Asp Ala Val His Val Ala Met Ala Asp Glu Ala
 35           40           45

Val Asn Ile Gly Pro Ala Pro Ala Ala Gln Ser Tyr Leu Leu Ile Glu
 50           55           60

Lys Ile Ile Asp Ala Cys Lys Gln Thr Gly Ala Gln Ala Val His Pro
 65           70           75           80

Gly Tyr Gly Phe Leu Ser Glu Arg Glu Ser Phe Pro Lys Ala Leu Ala
 85           90           95

Glu Ala Gly Ile Val Phe Ile Gly Pro Asn Pro Gly Ala Ile Ala Ala
 100          105          110

Met Gly Asp Lys Ile Glu Ser Lys Lys Ala Ala Ala Ala Glu Val
 115          120          125

Ser Thr Val Pro Gly Phe Leu Gly Val Ile Glu Ser Pro Glu His Ala
 130          135          140

Val Thr Ile Ala Asp Glu Ile Gly Tyr Pro Val Met Ile Lys Ala Ser
 145          150          155          160

Ala Gly Gly Gly Gly Lys Gly Met Arg Ile Ala Glu Ser Ala Asp Glu
 165          170          175

Val Ala Glu Gly Phe Ala Arg Ala Lys Ser Glu Ala Ser Ser Ser Phe
 180          185          190

Gly Asp Asp Arg Val Phe Val Glu Lys Phe Ile Thr Asp Pro Arg His
 195          200          205

Ile Glu Ile Gln Val Ile Gly Asp Lys His Gly Asn Val Ile Tyr Leu
 210          215          220

Gly Glu Arg Glu Cys Ser Ile Gln Arg Arg Asn Gln Lys Val Ile Glu
 225          230          235          240

Glu Ala Pro Ser Pro Leu Leu Asp Glu Glu Thr Arg Arg Lys Met Gly
 245          250          255

Glu Gln Ala Val Ala Leu Ala Lys Ala Val Asn Tyr Asp Ser Ala Gly
 260          265          270

Thr Val Glu Phe Val Ala Gly Gln Asp Lys Ser Phe Tyr Phe Leu Glu
 275          280          285

Met Asn Thr Arg Leu Gln Val Glu His Pro Val Thr Glu Met Ile Thr
 290          295          300

Gly Leu Asp Leu Val Glu Leu Met Ile Arg Val Ala Ala Gly Glu Thr
 305          310          315          320

Leu Pro Leu Thr Gln Asp Gln Val Lys Leu Asp Gly Trp Ala Val Glu
 325          330          335

Ser Arg Val Tyr Ala Glu Asp Pro Thr Arg Asn Phe Leu Pro Ser Ile

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340					345					350					
Gly	Arg	Leu	Thr	Thr	Tyr	Gln	Pro	Pro	Glu	Glu	Gly	Pro	Leu	Gly	Gly
	355						360					365			
Ala	Ile	Val	Arg	Asn	Asp	Thr	Gly	Val	Glu	Glu	Gly	Gly	Glu	Ile	Ala
	370					375					380				
Ile	His	Tyr	Asp	Pro	Met	Ile	Ala	Lys	Leu	Val	Thr	Trp	Ala	Pro	Thr
385					390					395					400
Arg	Leu	Glu	Ala	Ile	Asp	Ala	Gln	Ala	Thr	Ala	Leu	Asp	Ala	Phe	Ala
				405					410					415	
Ile	Glu	Gly	Ile	Arg	His	Asn	Ile	Pro	Phe	Leu	Ala	Thr	Leu	Met	Ala
			420					425					430		
His	Pro	Arg	Trp	Arg	Asp	Gly	Arg	Leu	Ser	Thr	Gly	Phe	Ile	Lys	Glu
		435					440					445			
Glu	Phe	Pro	Glu	Gly	Phe	Ile	Ala	Pro	Glu	Pro	Glu	Gly	Pro	Val	Ala
	450					455					460				
His	Arg	Leu	Ala	Ala	Val	Ala	Ala	Ala	Ile	Asp	His	Lys	Leu	Asn	Ile
465					470					475					480
Arg	Lys	Arg	Gly	Ile	Ser	Gly	Gln	Met	Arg	Asp	Pro	Ser	Leu	Leu	Thr
				485					490					495	
Phe	Gln	Arg	Glu	Arg	Val	Val	Val	Leu	Ser	Gly	Gln	Arg	Phe	Asn	Val
			500					505					510		
Thr	Val	Asp	Pro	Asp	Gly	Asp	Asp	Leu	Leu	Val	Thr	Phe	Asp	Asp	Gly
		515					520					525			
Thr	Thr	Ala	Pro	Val	Arg	Ser	Ala	Trp	Arg	Pro	Gly	Ala	Pro	Val	Trp
	530						535				540				
Ser	Gly	Thr	Val	Gly	Asp	Gln	Ser	Ile	Ala	Ile	Gln	Val	Arg	Pro	Leu
545					550					555					560
Leu	Asn	Gly	Val	Phe	Leu	Gln	His	Ala	Gly	Ala	Ala	Ala	Glu	Ala	Arg
				565					570					575	
Val	Phe	Thr	Arg	Arg	Glu	Ala	Glu	Leu	Ala	Asp	Leu	Met	Pro	Val	Lys
			580					585					590		
Glu	Asn	Ala	Gly	Ser	Gly	Lys	Gln	Leu	Leu	Cys	Pro	Met	Pro	Gly	Leu
	595						600					605			
Val	Lys	Gln	Ile	Met	Val	Ser	Glu	Gly	Gln	Glu	Val	Lys	Asn	Gly	Glu
	610						615					620			
Pro	Leu	Ala	Ile	Val	Glu	Ala	Met	Lys	Met	Glu	Asn	Val	Leu	Arg	Ala
625					630					635					640
Glu	Arg	Asp	Gly	Thr	Ile	Ser	Lys	Ile	Ala	Ala	Lys	Glu	Gly	Asp	Ser
				645					650					655	
Leu	Ala	Val	Asp	Ala	Val	Ile	Leu	Glu	Phe	Ala					
		660						665							

<210> SEQ ID NO 56

<211> LENGTH: 510

<212> TYPE: PRT

<213> ORGANISM: Methylobacterium extorquens

<300> PUBLICATION INFORMATION:

<308> DATABASE ACCESSION NUMBER: NCBI / YP_003065890

<309> DATABASE ENTRY DATE: 2010-04-16

<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(510)

<400> SEQUENCE: 56

Met	Lys	Asp	Ile	Leu	Glu	Lys	Leu	Glu	Glu	Arg	Arg	Ala	Gln	Ala	Arg
1			5						10					15	

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Leu Gly Gly Gly Glu Lys Arg Leu Glu Ala Gln His Thr Arg Gly Lys
20 25 30

Leu Thr Ala Arg Glu Arg Ile Glu Leu Leu Leu Asp His Gly Ser Phe
35 40 45

Glu Glu Phe Asp Met Phe Val Gln His Arg Ser Thr Asp Phe Gly Met
50 55 60

Glu Lys Gln Lys Ile Pro Gly Asp Gly Val Val Thr Gly Trp Gly Thr
65 70 75 80

Val Asn Gly Arg Thr Val Phe Leu Phe Ser Lys Asp Phe Thr Val Phe
85 90 95

Gly Gly Ser Leu Ser Glu Ala His Ala Ala Lys Ile Val Lys Val Gln
100 105 110

Asp Met Ala Leu Lys Met Arg Ala Pro Ile Ile Gly Ile Phe Asp Ala
115 120 125

Gly Gly Ala Arg Ile Gln Glu Gly Val Ala Ala Leu Gly Gly Tyr Gly
130 135 140

Glu Val Phe Arg Arg Asn Val Ala Ala Ser Gly Val Ile Pro Gln Ile
145 150 155 160

Ser Val Ile Met Gly Pro Cys Ala Gly Gly Asp Val Tyr Ser Pro Ala
165 170 175

Met Thr Asp Phe Ile Phe Met Val Arg Asp Thr Ser Tyr Met Phe Val
180 185 190

Thr Gly Pro Asp Val Val Lys Thr Val Thr Asn Glu Val Val Thr Ala
195 200 205

Glu Glu Leu Gly Gly Ala Lys Val His Thr Ser Lys Ser Ser Ile Ala
210 215 220

Asp Gly Ser Phe Glu Asn Asp Val Glu Ala Ile Leu Gln Ile Arg Arg
225 230 235 240

Leu Leu Asp Phe Leu Pro Ala Asn Asn Ile Glu Gly Val Pro Glu Ile
245 250 255

Glu Ser Phe Asp Asp Val Asn Arg Leu Asp Lys Ser Leu Asp Thr Leu
260 265 270

Ile Pro Asp Asn Pro Asn Lys Pro Tyr Asp Met Gly Glu Leu Ile Arg
275 280 285

Arg Val Val Asp Glu Gly Asp Phe Phe Glu Ile Gln Ala Ala Tyr Ala
290 295 300

Arg Asn Ile Ile Thr Gly Phe Gly Arg Val Glu Gly Arg Thr Val Gly
305 310 315 320

Phe Val Ala Asn Gln Pro Leu Val Leu Ala Gly Val Leu Asp Ser Asp
325 330 335

Ala Ser Arg Lys Ala Ala Arg Phe Val Arg Phe Cys Asn Ala Phe Ser
340 345 350

Ile Pro Ile Val Thr Phe Val Asp Val Pro Gly Phe Leu Pro Gly Thr
355 360 365

Ala Gln Glu Tyr Gly Gly Leu Ile Lys His Gly Ala Lys Leu Leu Phe
370 375 380

Ala Tyr Ser Gln Ala Thr Val Pro Leu Val Thr Ile Ile Thr Arg Lys
385 390 395 400

Ala Phe Gly Gly Ala Tyr Asp Val Met Ala Ser Lys His Val Gly Ala
405 410 415

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Asp Leu Asn Tyr Ala Trp Pro Thr Ala Gln Ile Ala Val Met Gly Ala
      420                               425           430

Lys Gly Ala Val Glu Ile Ile Phe Arg Ala Glu Ile Gly Asp Ala Asp
      435                               440           445

Lys Ile Ala Glu Arg Thr Lys Glu Tyr Glu Asp Arg Phe Leu Ser Pro
      450                               455           460

Phe Val Ala Ala Glu Arg Gly Tyr Ile Asp Glu Val Ile Met Pro His
      465                               470           475           480

Ser Thr Arg Lys Arg Ile Ala Arg Ala Leu Gly Met Leu Arg Thr Lys
      485                               490           495

Glu Met Glu Gln Pro Trp Lys Lys His Asp Asn Ile Pro Leu
      500                               505           510

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<210> SEQ ID NO 57
<211> LENGTH: 670
<212> TYPE: PRT
<213> ORGANISM: Sinorhizobium meliloti
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / NP_437988
<309> DATABASE ENTRY DATE: 2010-04-01
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(670)

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<400> SEQUENCE: 57

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Met Gly His Met Phe Lys Lys Ile Leu Ile Ala Asn Arg Gly Glu Ile
 1      5      10      15

Ala Cys Arg Val Ile Arg Thr Thr Lys Ala Leu Gly Ile Pro Thr Val
 20      25      30

Ala Val Tyr Ser Asp Ala Asp Arg Asp Ala Met His Val Arg Met Ala
 35      40      45

Asp Glu Ala Val His Ile Gly Pro Ser Pro Ser Ser Gln Ser Tyr Ile
 50      55      60

Val Ile Glu Asn Ile Leu Ala Ala Ile Arg Arg Thr Gly Ala Asp Ala
 65      70      75      80

Val His Pro Gly Tyr Gly Phe Leu Ser Glu Asn Ala Ala Phe Ala Glu
 85      90      95

Ala Leu Glu Lys Asp Gly Val Thr Phe Ile Gly Pro Pro Val Arg Ala
100      105      110

Ile Glu Ala Met Gly Asp Lys Ile Thr Ser Lys Lys Leu Ala Ala Glu
115      120      125

Ala Gly Val Phe Thr Val Pro Gly His Met Gly Leu Ile Glu Asp Ala
130      135      140

Asp Glu Ala Ala Arg Ile Ala Ala Glu Ile Gly Phe Pro Val Met Ile
145      150      155      160

Lys Ala Ser Ala Gly Gly Gly Gly Lys Gly Met Arg Ile Ala Trp Asn
165      170      175

Glu Arg Glu Ala Arg Glu Gly Phe Gln Ser Ser Arg Asn Glu Ala Lys
180      185      190

Ser Ser Phe Gly Asp Asp Arg Ile Phe Ile Glu Lys Phe Val Thr Glu
195      200      205

Pro Arg His Ile Glu Ile Gln Val Leu Gly Asp Lys His Gly Asn Ile
210      215      220

Leu Tyr Leu Gly Glu Arg Glu Cys Ser Ile Gln Arg Arg Asn Gln Lys
225      230      235      240

Val Ile Glu Glu Ala Pro Ser Pro Phe Leu Asp Glu Lys Thr Arg Arg

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245					250					255					
Ala	Met	Gly	Glu	Gln	Ala	Val	Ala	Leu	Ala	Lys	Ala	Val	Gly	Tyr	His
			260					265					270		
Ser	Ala	Gly	Thr	Val	Glu	Phe	Ile	Val	Asp	Ala	Gly	Arg	Asn	Phe	Tyr
		275					280					285			
Phe	Leu	Glu	Met	Asn	Thr	Arg	Leu	Gln	Val	Glu	His	Pro	Val	Thr	Glu
	290					295					300				
Leu	Val	Thr	Gly	Leu	Asp	Leu	Val	Glu	Gln	Met	Ile	Arg	Val	Ala	Ala
305						310					315				320
Gly	Ala	Lys	Leu	Ala	Phe	Ala	Gln	Lys	Asp	Val	Lys	Leu	Asp	Gly	Trp
				325					330					335	
Ala	Ile	Glu	Ser	Arg	Leu	Tyr	Ala	Glu	Asp	Pro	Tyr	Arg	Thr	Phe	Leu
			340					345					350		
Pro	Ser	Ile	Gly	Arg	Leu	Thr	Arg	Tyr	Arg	Pro	Pro	Glu	Glu	Gly	Thr
		355					360					365			
Gln	Ala	Asp	Gly	Thr	Val	Ile	Arg	Asn	Asp	Thr	Gly	Val	Phe	Glu	Gly
	370					375					380				
Gly	Glu	Ile	Ser	Met	Tyr	Tyr	Asp	Pro	Met	Ile	Ala	Lys	Leu	Cys	Thr
385						390					395				400
Trp	Gly	Pro	Asp	Arg	Leu	Thr	Ala	Val	Arg	Ala	Met	Ala	Asp	Ala	Leu
				405					410					415	
Asp	Ala	Phe	Glu	Val	Glu	Gly	Ile	Gly	His	Asn	Leu	Pro	Phe	Leu	Ala
			420					425					430		
Ala	Val	Met	Gln	Gln	Glu	Arg	Phe	His	Glu	Gly	Arg	Leu	Thr	Thr	Ala
		435					440					445			
Tyr	Ile	Ala	Glu	Glu	Phe	Ala	Gly	Gly	Phe	His	Gly	Val	Ala	Leu	Asp
	450					455					460				
Asp	Ala	Ser	Ala	Arg	Lys	Leu	Ala	Ala	Val	Ala	Ala	Thr	Val	Asn	Gln
465						470					475				480
Thr	Leu	Gln	Glu	Arg	Ala	Ser	Arg	Ile	Ser	Gly	Thr	Ile	Gly	Asn	His
				485					490					495	
Arg	Arg	Val	Val	Gly	His	Glu	Trp	Val	Thr	Ser	Leu	Asp	Gly	His	Glu
			500					505					510		
Ile	Gln	Val	Thr	Cys	Glu	Val	Ser	Ala	Asp	Gly	Thr	Tyr	Val	Arg	Phe
		515					520					525			
Ala	Asp	Gly	Thr	Ser	Val	Ser	Val	Ala	Thr	Asp	Trp	Ala	Pro	Gly	Arg
	530					535					540				
Thr	Arg	Ala	Ala	Phe	Asn	Ile	Asp	Asn	Gln	Pro	Met	Ser	Val	Lys	Val
545						550					555				560
Glu	Leu	Ala	Gly	Pro	Gly	Ile	Arg	Leu	Arg	Trp	Arg	Gly	Ile	Asp	Val
				565					570					575	
Val	Ala	Arg	Val	Arg	Ser	Pro	Arg	Ile	Ala	Glu	Leu	Ala	Arg	Leu	Met
			580					585					590		
Pro	Lys	Lys	Leu	Pro	Pro	Asp	Thr	Ser	Lys	Met	Leu	Leu	Cys	Pro	Met
		595					600					605			
Pro	Gly	Val	Val	Thr	Ser	Ile	Thr	Val	Lys	Ala	Gly	Glu	Thr	Val	Glu
	610					615					620				
Ala	Gly	Gln	Ala	Ile	Ala	Val	Val	Glu	Ala	Met	Lys	Met	Glu	Asn	Ile
625						630					635				640
Leu	Arg	Ala	Glu	Lys	Arg	Ala	Ile	Val	Lys	Arg	Val	Ala	Ile	Glu	Ala
				645					650					655	

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Gly Ala Ser Leu Ala Val Asp Glu Leu Ile Met Glu Phe Glu
 660 665 670

<210> SEQ ID NO 58
 <211> LENGTH: 510
 <212> TYPE: PRT
 <213> ORGANISM: Sinorhizobium meliloti
 <300> PUBLICATION INFORMATION:
 <308> DATABASE ACCESSION NUMBER: NCBI / NP_437987
 <309> DATABASE ENTRY DATE: 2010-04-01
 <313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(510)

<400> SEQUENCE: 58

Met Arg Ala Val Leu Glu Gln Val Glu Ala Arg Arg Ala Glu Ala Arg
 1 5 10 15
 Ala Gly Gly Gly Glu Arg Arg Ile Ala Ala Gln His Gly Lys Gly Lys
 20 25 30
 Leu Thr Ala Arg Glu Arg Ile Asp Val Leu Leu Asp Glu Gly Ser Phe
 35 40 45
 Glu Glu Tyr Asp Met Tyr Val Thr His Arg Ser Val Asp Phe Gly Met
 50 55 60
 Ala Gly Gln Lys Ile Pro Gly Asp Gly Val Val Thr Gly Trp Gly Thr
 65 70 75 80
 Ile Asn Gly Arg Gln Val Tyr Val Phe Ser Gln Asp Phe Thr Val Leu
 85 90 95
 Gly Gly Ser Leu Ser Glu Thr His Ala Gln Lys Ile Cys Lys Ile Met
 100 105 110
 Asp Met Ala Ala Arg Asn Gly Ala Pro Val Ile Gly Leu Asn Asp Ser
 115 120 125
 Gly Gly Ala Arg Ile Gln Glu Gly Val Ala Ser Leu Ala Gly Tyr Ala
 130 135 140
 Glu Val Phe Arg Arg Asn Ala Glu Val Ser Gly Val Ile Pro Gln Ile
 145 150 155 160
 Ser Val Ile Met Gly Pro Cys Ala Gly Gly Ala Val Tyr Ser Pro Ala
 165 170 175
 Met Thr Asp Phe Ile Phe Met Val Arg Asp Ser Ser Tyr Met Phe Val
 180 185 190
 Thr Gly Pro Asp Val Val Lys Thr Val Thr Asn Glu Ile Val Thr Ala
 195 200 205
 Glu Glu Leu Gly Gly Ala Arg Thr His Thr Thr Lys Ser Ser Val Ala
 210 215 220
 Asp Gly Ala Tyr Glu Asn Asp Ile Glu Ala Leu Glu His Val Arg Leu
 225 230 235 240
 Leu Phe Asp Phe Leu Pro Leu Asn Asn Arg Glu Lys Pro Pro Val Arg
 245 250 255
 Pro Phe His Asp Asp Pro Gly Arg Leu Glu Met Arg Leu Asp Ser Leu
 260 265 270
 Ile Pro Asp Ser Ala Ala Lys Pro Tyr Asp Met Lys Glu Leu Ile Leu
 275 280 285
 Ala Ile Ala Asp Glu Ala Asp Phe Phe Glu Leu Gln Ala Ser Phe Ala
 290 295 300
 Arg Asn Ile Ile Thr Gly Phe Ile Arg Ile Glu Gly Gln Thr Val Gly
 305 310 315 320

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Val Ile Ala Asn Gln Pro Met Val Leu Ala Gly Cys Leu Asp Ile Asp
          325          330          335
Ser Ser Arg Lys Ala Ala Arg Phe Val Arg Phe Cys Asp Ala Phe Ser
          340          345          350
Ile Pro Ile Leu Thr Leu Val Asp Val Pro Gly Phe Leu Pro Gly Thr
          355          360          365
Ala Gln Glu Tyr Gly Gly Val Ile Lys His Gly Ala Lys Leu Leu Phe
          370          375          380
Ala Tyr Ser Gln Ala Thr Val Pro Met Val Thr Leu Ile Thr Arg Lys
          385          390          395          400
Ala Tyr Gly Gly Ala Tyr Asp Val Met Ala Ser Lys His Ile Gly Ala
          405          410          415
Asp Val Asn Tyr Ala Trp Pro Thr Ala Glu Ile Ala Val Met Gly Ala
          420          425          430
Lys Gly Ala Thr Glu Ile Leu Tyr Arg Ser Glu Leu Gly Asp Pro Ala
          435          440          445
Lys Ile Ala Ala Arg Thr Lys Glu Tyr Glu Glu Arg Phe Ala Asn Pro
          450          455          460
Phe Val Ala Ala Glu Arg Gly Phe Ile Asp Glu Val Ile Met Pro His
          465          470          475          480
Ser Ser Arg Arg Arg Ile Ala Arg Ala Phe Ala Ser Leu Arg Asn Lys
          485          490          495
Gln Val Glu Thr Arg Trp Arg Lys His Asp Thr Ile Pro Leu
          500          505          510

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<210> SEQ ID NO 59
<211> LENGTH: 681
<212> TYPE: PRT
<213> ORGANISM: Ruegeria pomeroyi
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / YP_166352
<309> DATABASE ENTRY DATE: 2010-06-29
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(681)

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<400> SEQUENCE: 59

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```

Met Phe Asn Lys Ile Leu Ile Ala Asn Arg Gly Glu Ile Ala Cys Arg
 1          5          10          15
Val Ile Lys Thr Ala Arg Lys Met Gly Ile Ser Thr Val Ala Ile Tyr
          20          25          30
Ser Asp Ala Asp Lys Gln Ala Leu His Val Gln Met Ala Asp Glu Ala
          35          40          45
Val His Ile Gly Pro Pro Pro Ala Asn Gln Ser Tyr Ile Val Ile Asp
          50          55          60
Lys Val Met Ala Ala Ile Arg Ala Thr Gly Ala Gln Ala Val His Pro
          65          70          75          80
Gly Tyr Gly Phe Leu Ser Glu Asn Ser Lys Phe Ala Glu Ala Leu Glu
          85          90          95
Ala Glu Gly Val Ile Phe Val Gly Pro Pro Lys Gly Ala Ile Glu Ala
          100          105          110
Met Gly Asp Lys Ile Thr Ser Lys Lys Ile Ala Gln Glu Ala Asn Val
          115          120          125
Ser Thr Val Pro Gly Tyr Met Gly Leu Ile Glu Asp Ala Asp Glu Ala
          130          135          140
Val Lys Ile Ser Asn Gln Ile Gly Tyr Pro Val Met Ile Lys Ala Ser

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145					150						155				160
Ala	Gly	Gly	Gly	Gly	Lys	Gly	Met	Arg	Ile	Ala	Trp	Asn	Asp	Gln	Glu
				165					170					175	
Ala	Arg	Glu	Gly	Phe	Gln	Ser	Ser	Lys	Asn	Glu	Ala	Ala	Asn	Ser	Phe
			180					185					190		
Gly	Asp	Asp	Arg	Ile	Phe	Ile	Glu	Lys	Phe	Val	Thr	Gln	Pro	Arg	His
	195						200					205			
Ile	Glu	Ile	Gln	Val	Leu	Cys	Asp	Ser	His	Gly	Asn	Gly	Ile	Tyr	Leu
	210					215					220				
Gly	Glu	Arg	Glu	Cys	Ser	Ile	Gln	Arg	Arg	Asn	Gln	Lys	Val	Val	Glu
225				230						235					240
Glu	Ala	Pro	Ser	Pro	Phe	Leu	Asp	Glu	Ala	Thr	Arg	Arg	Ala	Met	Gly
				245					250					255	
Glu	Gln	Ala	Val	Ala	Leu	Ala	Lys	Ala	Val	Gly	Tyr	Ala	Ser	Ala	Gly
			260					265					270		
Thr	Val	Glu	Phe	Ile	Val	Asp	Gly	Gln	Lys	Asn	Phe	Tyr	Phe	Leu	Glu
		275					280					285			
Met	Asn	Thr	Arg	Leu	Gln	Val	Glu	His	Pro	Val	Thr	Glu	Leu	Ile	Thr
	290					295					300				
Gly	Val	Asp	Leu	Val	Glu	Gln	Met	Ile	Arg	Val	Ala	Ala	Gly	Glu	Pro
305					310					315					320
Leu	Ser	Ile	Thr	Gln	Gly	Asp	Val	Lys	Leu	Thr	Gly	Trp	Ala	Ile	Glu
				325					330					335	
Asn	Arg	Leu	Tyr	Ala	Glu	Asp	Pro	Tyr	Arg	Gly	Phe	Leu	Pro	Ser	Ile
			340					345					350		
Gly	Arg	Leu	Thr	Arg	Tyr	Arg	Pro	Pro	Ala	Glu	Thr	Ala	Ala	Gly	Pro
		355					360					365			
Leu	Leu	Val	Asn	Gly	Lys	Trp	Gln	Gly	Asp	Ala	Pro	Ser	Gly	Glu	Ala
	370					375					380				
Ala	Val	Arg	Asn	Asp	Thr	Gly	Val	Tyr	Glu	Gly	Gly	Glu	Ile	Ser	Met
385					390					395					400
Tyr	Tyr	Asp	Pro	Met	Ile	Ala	Lys	Leu	Cys	Thr	Trp	Ala	Pro	Thr	Arg
				405					410					415	
Ala	Ala	Ala	Ile	Glu	Ala	Met	Arg	Ile	Ala	Leu	Asp	Ser	Phe	Glu	Val
			420					425					430		
Glu	Gly	Ile	Gly	His	Asn	Leu	Pro	Phe	Leu	Ser	Ala	Val	Met	Asp	His
		435					440					445			
Pro	Lys	Phe	Ile	Ser	Gly	Asp	Met	Thr	Thr	Ala	Phe	Ile	Ala	Glu	Glu
	450					455					460				
Tyr	Pro	Glu	Gly	Phe	Glu	Gly	Val	Asn	Leu	Pro	Glu	Thr	Asp	Leu	Arg
465					470					475					480
Arg	Val	Ala	Ala	Ala	Ala	Ala	Ala	Met	His	Arg	Val	Ala	Glu	Ile	Arg
				485					490					495	
Arg	Thr	Arg	Val	Ser	Gly	Arg	Met	Asp	Asn	His	Glu	Arg	Arg	Val	Gly
			500					505					510		
Thr	Glu	Trp	Val	Val	Thr	Leu	Gln	Gly	Ala	Asp	Phe	Pro	Val	Thr	Ile
		515					520					525			
Ala	Ala	Asp	His	Asp	Gly	Ser	Thr	Val	Ser	Phe	Asp	Asp	Gly	Ser	Ser
	530					535					540				
Met	Arg	Val	Thr	Ser	Asp	Trp	Thr	Pro	Gly	Asp	Gln	Leu	Ala	Asn	Leu
545					550					555					560

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Met Val Asp Gly Ala Pro Leu Val Leu Lys Val Gly Lys Ile Ser Gly
      565                               570                               575

Gly Phe Arg Ile Arg Thr Arg Gly Ala Asp Leu Lys Val His Val Arg
      580                               585                               590

Thr Pro Arg Gln Ala Glu Leu Ala Arg Leu Met Pro Glu Lys Leu Pro
      595                               600                               605

Pro Asp Thr Ser Lys Met Leu Leu Cys Pro Met Pro Gly Leu Ile Val
      610                               615                               620

Lys Val Asp Val Glu Val Gly Gln Glu Val Gln Glu Gly Gln Ala Leu
      625                               630                               635

Cys Thr Ile Glu Ala Met Lys Met Glu Asn Ile Leu Arg Ala Glu Lys
      645                               650                               655

Lys Gly Val Val Ala Lys Ile Asn Ala Ser Ala Gly Asn Ser Leu Ala
      660                               665                               670

Val Asp Asp Val Ile Met Glu Phe Glu
      675                               680

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<210> SEQ ID NO 60
<211> LENGTH: 510
<212> TYPE: PRT
<213> ORGANISM: Ruegeria pomeroyi
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / YP_166345
<309> DATABASE ENTRY DATE: 2010-06-29
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(510)

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<400> SEQUENCE: 60

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Met Lys Asp Ile Leu Ser Glu Leu Glu Thr Arg Arg Glu Ala Ala Arg
 1      5      10      15

Leu Gly Gly Gly Gln Lys Arg Ile Asp Ala Gln His Ala Arg Gly Lys
 20     25     30

Leu Thr Ala Arg Glu Arg Ile Glu Leu Leu Leu Asp Glu Asp Ser Phe
 35     40     45

Glu Glu Phe Asp Met Phe Val Ser His Arg Cys Thr Asp Phe Gly Met
 50     55     60

Glu Lys Gln Arg Pro Ala Gly Asp Gly Val Val Thr Gly Trp Gly Thr
 65     70     75     80

Ile Asn Gly Arg Met Val Tyr Val Phe Ser Gln Asp Phe Thr Val Phe
 85     90     95

Gly Gly Ser Leu Ser Glu Thr His Ala Gln Lys Ile Cys Lys Ile Met
100    105    110

Asp Met Ala Val Gln Asn Gly Ala Pro Val Ile Gly Ile Asn Asp Ser
115    120    125

Gly Gly Ala Arg Ile Gln Glu Gly Val Ala Ser Leu Ala Gly Tyr Ala
130    135    140

Glu Val Phe Gln Arg Asn Ile Met Ala Ser Gly Val Val Pro Gln Ile
145    150    155    160

Ser Val Ile Met Gly Pro Cys Ala Gly Gly Ala Val Tyr Ser Pro Ala
165    170    175

Met Thr Asp Phe Ile Phe Met Val Lys Asp Thr Ser Tyr Met Phe Val
180    185    190

Thr Gly Pro Asp Val Val Lys Thr Val Thr Asn Glu Val Val Thr Ala
195    200    205

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Glu Glu Leu Gly Gly Ala Ser Thr His Thr Arg Lys Ser Ser Val Ala
 210 215 220
 Asp Gly Ala Phe Glu Asn Asp Val Glu Ala Leu Ala Glu Val Arg Arg
 225 230 235 240
 Leu Val Asp Phe Leu Pro Leu Asn Asn Arg Glu Lys Pro Pro Val Arg
 245 250 255
 Pro Phe Phe Asp Glu Pro Gly Arg Ile Glu Ala Ser Leu Asp Thr Leu
 260 265 270
 Val Pro Glu Asn Ala Asn Thr Pro Tyr Asp Met Lys Glu Leu Ile Asn
 275 280 285
 Lys Ile Ala Asp Glu Gly Asp Phe Tyr Glu Ile Gln Glu Asp Phe Ala
 290 295 300
 Lys Asn Ile Ile Thr Gly Phe Ile Arg Leu Glu Gly Gln Thr Val Gly
 305 310 315 320
 Val Val Ala Asn Gln Pro Met Ile Leu Ala Gly Cys Leu Asp Ile Asp
 325 330 335
 Ser Ser Arg Lys Ala Ala Arg Phe Val Arg Phe Cys Asp Cys Phe Glu
 340 345 350
 Ile Pro Ile Leu Thr Leu Val Asp Val Pro Gly Phe Leu Pro Gly Thr
 355 360 365
 Ser Gln Glu Tyr Gly Gly Val Ile Lys His Gly Ala Lys Leu Leu Phe
 370 375 380
 Ala Tyr Gly Glu Ala Thr Val Pro Lys Val Thr Val Ile Thr Arg Lys
 385 390 395 400
 Ala Tyr Gly Gly Ala Tyr Asp Val Met Ala Ser Lys His Leu Arg Gly
 405 410 415
 Asp Phe Asn Tyr Ala Trp Pro Thr Ala Glu Ile Ala Val Met Gly Ala
 420 425 430
 Lys Gly Ala Thr Glu Ile Ile His Arg Ala Asp Leu Gly Asp Ala Asp
 435 440 445
 Lys Ile Ala Ala His Thr Lys Asp Tyr Glu Gly Arg Phe Ala Asn Pro
 450 455 460
 Phe Val Ala Ala Glu Arg Gly Phe Ile Asp Glu Val Ile Gln Pro Arg
 465 470 475 480
 Ser Thr Arg Lys Arg Val Ser Arg Ala Phe Ala Ser Leu Arg Gly Lys
 485 490 495
 Ser Leu Lys Asn Pro Trp Lys Lys His Asp Asn Ile Pro Leu
 500 505 510

<210> SEQ ID NO 61
 <211> LENGTH: 678
 <212> TYPE: PRT
 <213> ORGANISM: Bacillus megaterium
 <300> PUBLICATION INFORMATION:
 <308> DATABASE ACCESSION NUMBER: NCBI / YP_003564880
 <309> DATABASE ENTRY DATE: 2010-12-17
 <313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(678)
 <400> SEQUENCE: 61

Met Lys Thr Asn Thr Leu Ser Phe His Glu Phe Thr Arg Thr Pro Lys
 1 5 10 15
 Glu Asp Trp Ala Gln Glu Val Ser Lys Asn Thr Ala Ile Ser Ser Lys
 20 25 30
 Glu Thr Leu Glu Asn Ile Phe Leu Lys Pro Leu Tyr Phe Glu Ser Asp

-continued

35					40					45					
Thr	Ala	His	Leu	Asp	Tyr	Leu	Gln	Gln	Ser	Pro	Ala	Gly	Ile	Asp	Tyr
50					55					60					
Leu	Arg	Gly	Ala	Gly	Lys	Glu	Ser	Tyr	Ile	Leu	Gly	Glu	Trp	Glu	Ile
65					70					75					80
Thr	Gln	Lys	Ile	Asp	Leu	Pro	Ser	Ile	Lys	Glu	Ser	Asn	Lys	Leu	Leu
				85					90					95	
Leu	His	Ser	Leu	Arg	Asn	Gly	Gln	Asn	Thr	Ala	Ala	Phe	Thr	Cys	Ser
			100					105					110		
Glu	Ala	Met	Arg	Gln	Gly	Lys	Asp	Ile	Asp	Glu	Ala	Thr	Glu	Ala	Glu
		115					120					125			
Val	Ala	Ser	Gly	Ala	Thr	Ile	Ser	Thr	Leu	Glu	Asp	Val	Ala	His	Leu
		130					135					140			
Phe	Gln	His	Val	Ala	Leu	Glu	Ala	Val	Pro	Leu	Phe	Leu	Asn	Thr	Gly
145					150					155					160
Cys	Thr	Ser	Val	Pro	Leu	Leu	Ser	Phe	Leu	Lys	Ala	Tyr	Cys	Val	Asp
				165					170					175	
His	Asn	Phe	Asn	Met	Arg	Gln	Leu	Lys	Gly	Thr	Val	Gly	Met	Asp	Pro
			180					185					190		
Leu	Gly	Thr	Leu	Ala	Glu	Tyr	Gly	Arg	Val	Pro	Leu	Ser	Thr	Arg	Asp
		195					200						205		
Leu	Tyr	Asp	His	Leu	Ala	Tyr	Ala	Thr	Arg	Leu	Ala	His	Ser	Asn	Val
		210					215					220			
Pro	Glu	Leu	Lys	Thr	Ile	Ile	Val	Ser	Ser	Ile	Pro	Tyr	His	Asn	Ser
225					230					235				240	
Gly	Ala	Asn	Ala	Val	Gln	Glu	Leu	Ala	Tyr	Met	Leu	Ala	Thr	Gly	Val
				245					250					255	
Gln	Tyr	Ile	Asp	Glu	Cys	Ile	Lys	Arg	Gly	Leu	Ser	Leu	His	Gln	Val
			260					265						270	
Leu	Pro	His	Met	Thr	Phe	Ser	Phe	Ser	Val	Ser	Ser	His	Leu	Phe	Met
			275				280					285			
Glu	Ile	Ser	Lys	Leu	Arg	Ala	Phe	Arg	Met	Leu	Trp	Ala	Asn	Val	Val
			290				295					300			
Arg	Ala	Phe	Asp	Asp	Thr	Ala	Val	Ser	Val	Pro	Phe	Ile	His	Thr	Glu
305					310					315				320	
Thr	Ser	His	Leu	Thr	Gln	Ser	Lys	Glu	Asp	Met	Tyr	Thr	Asn	Ala	Leu
				325					330					335	
Arg	Ser	Thr	Val	Gln	Ala	Phe	Ala	Ser	Ile	Val	Gly	Gly	Ala	Asp	Ser
			340					345					350		
Leu	His	Ile	Glu	Pro	Tyr	Asp	Ser	Val	Thr	Ser	Ser	Ser	Ser	Gln	Phe
			355				360					365			
Ala	His	Arg	Leu	Ala	Arg	Asn	Thr	His	Leu	Ile	Leu	Gln	His	Glu	Thr
			370				375					380			
His	Ile	Ser	Lys	Val	Met	Asp	Pro	Ala	Gly	Gly	Ser	Trp	Tyr	Val	Glu
385					390					395				400	
Ala	Tyr	Thr	His	Glu	Leu	Met	Thr	Lys	Ala	Trp	Glu	Leu	Phe	Gly	Asn
				405					410					415	
Ile	Glu	Asp	His	Gly	Gly	Met	Glu	Glu	Ala	Leu	Lys	Gln	Gly	Arg	Ile
			420				425						430		
Gln	Asp	Glu	Val	Glu	Gln	Met	Lys	Val	Lys	Arg	Gln	Glu	Asp	Ile	Glu
			435				440					445			

-continued

Cys Arg Ile Glu Arg Leu Ile Gly Val Thr His Tyr Ala Pro Lys Gln
 450 455 460
 Gln Asp Ala Ser Gln Glu Ile Lys Ser Thr Pro Phe Lys Lys Glu Glu
 465 470 475 480
 Ile Lys Met Asp Lys Tyr Ser Asp Gln Asn Ala Ser Glu Phe Ser Ser
 485 490 495
 Asn Leu Ser Leu Glu Asp Tyr Thr Lys Leu Ala Ser Lys Gly Val Thr
 500 505 510
 Ala Gly Trp Met Leu Lys Gln Met Ala Lys Gln Thr Gln Pro Asp Ser
 515 520 525
 Val Val Pro Leu Thr Lys Trp Arg Ala Ala Glu Lys Phe Glu Lys Ile
 530 535 540
 Arg Val Tyr Thr Lys Gly Met Ser Ile Gly Ile Met Glu Leu Thr Asp
 545 550 555 560
 Pro Ser Ser Arg Lys Lys Ala Glu Ile Ala Arg Ser Leu Phe Glu Ser
 565 570 575
 Ala Gly Phe Ala Cys Glu Thr Ile Lys Asn Ile Asp Ser Tyr Val Glu
 580 585 590
 Ile Ala Asp Trp Met Asn Glu Gln Lys His Glu Ala Tyr Val Ile Cys
 595 600 605
 Gly Ser Asp Glu Leu Val Glu Lys Leu Leu Thr Lys Ala Met Thr Tyr
 610 615 620
 Phe Glu Glu Asp Ser Val Tyr Val Tyr Val Val Gly Glu Glu His Val
 625 630 635 640
 Ser Arg Lys Thr Gln Trp Gln Gln Lys Gly Val Met Ser Val Ile His
 645 650 655
 Pro Lys Thr Asn Val Ile Gln Cys Val Lys Lys Leu Leu Cys Ala Leu
 660 665 670
 Glu Val Glu Val His Val
 675

<210> SEQ ID NO 62
 <211> LENGTH: 716
 <212> TYPE: PRT
 <213> ORGANISM: Bacillus megaterium
 <300> PUBLICATION INFORMATION:
 <308> DATABASE ACCESSION NUMBER: NCBI - YP_003564879
 <309> DATABASE ENTRY DATE: 2010-12-17
 <313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(716)

<400> SEQUENCE: 62

Met Tyr Lys Lys Pro Ser Phe Ser Asn Ile Pro Leu Ser Phe Ser Lys
 1 5 10 15
 Gln Gln Arg Glu Asp Asp Val Thr Gln Ser Ser Tyr Thr Ala Phe Gln
 20 25 30
 Thr Asn Glu Gln Ile Glu Leu Lys Ser Val Tyr Thr Lys Lys Asp Arg
 35 40 45
 Asp Asn Leu Asp Phe Ile His Phe Ala Pro Gly Val Pro Pro Phe Val
 50 55 60
 Arg Gly Pro Tyr Ala Thr Met Tyr Val Asn Arg Pro Trp Thr Ile Arg
 65 70 75 80
 Gln Tyr Ala Gly Tyr Ser Thr Ala Glu Glu Ser Asn Ala Phe Tyr Arg
 85 90 95

-continued

Arg	Asn	Leu	Ala	Ala	Gly	Gln	Lys	Gly	Leu	Ser	Val	Ala	Phe	Asp	Leu
			100					105						110	
Ala	Thr	His	Arg	Gly	Tyr	Asp	Ser	Asp	His	Pro	Arg	Val	Val	Gly	Asp
		115					120					125			
Val	Gly	Lys	Ala	Gly	Val	Ala	Ile	Asp	Ser	Met	Met	Asp	Met	Lys	Gln
		130				135					140				
Leu	Phe	Glu	Gly	Ile	Pro	Leu	Asp	Gln	Met	Ser	Val	Ser	Met	Thr	Met
145					150					155					160
Asn	Gly	Ala	Val	Leu	Pro	Ile	Leu	Ala	Phe	Tyr	Ile	Val	Thr	Ala	Glu
				165					170					175	
Glu	Gln	Gly	Val	Lys	Lys	Glu	Lys	Leu	Ala	Gly	Thr	Ile	Gln	Asn	Asp
			180					185					190		
Ile	Leu	Lys	Glu	Tyr	Met	Val	Arg	Asn	Thr	Tyr	Ile	Tyr	Pro	Pro	Glu
		195					200					205			
Met	Ser	Met	Arg	Ile	Ile	Ala	Asp	Ile	Phe	Lys	Tyr	Thr	Ala	Glu	Tyr
		210				215					220				
Met	Pro	Lys	Phe	Asn	Ser	Ile	Ser	Ile	Ser	Gly	Tyr	His	Met	Gln	Glu
225					230					235					240
Ala	Gly	Ala	Pro	Ala	Asp	Leu	Glu	Leu	Ala	Tyr	Thr	Leu	Ala	Asp	Gly
				245					250					255	
Leu	Glu	Tyr	Val	Arg	Thr	Gly	Leu	Lys	Ala	Gly	Ile	Thr	Ile	Asp	Ala
			260					265					270		
Phe	Ala	Pro	Arg	Leu	Ser	Phe	Phe	Trp	Ala	Ile	Gly	Met	Asn	Tyr	Phe
		275					280					285			
Met	Glu	Val	Ala	Lys	Met	Arg	Ala	Gly	Arg	Leu	Leu	Trp	Ala	Lys	Leu
		290				295					300				
Met	Lys	Gln	Phe	Glu	Pro	Asp	Asn	Pro	Lys	Ser	Leu	Ala	Leu	Arg	Thr
305					310					315					320
His	Ser	Gln	Thr	Ser	Gly	Trp	Ser	Leu	Thr	Glu	Gln	Asp	Pro	Phe	Asn
				325					330					335	
Asn	Val	Ile	Arg	Thr	Cys	Val	Glu	Ala	Leu	Ala	Ala	Val	Ser	Gly	His
			340					345					350		
Thr	Gln	Ser	Leu	His	Thr	Asn	Ala	Leu	Asp	Glu	Ala	Ile	Ala	Leu	Pro
		355					360					365			
Thr	Asp	Phe	Ser	Ala	Arg	Ile	Ala	Arg	Asn	Thr	Gln	Leu	Tyr	Leu	Gln
		370				375					380				
Asn	Glu	Thr	Glu	Ile	Cys	Ser	Val	Ile	Asp	Pro	Trp	Gly	Gly	Ser	Tyr
385					390				395						400
Tyr	Val	Glu	Ser	Leu	Thr	Asn	Glu	Leu	Met	Ile	Lys	Ala	Trp	Lys	His
				405					410					415	
Leu	Glu	Glu	Ile	Glu	Gln	Leu	Gly	Gly	Met	Thr	Lys	Ala	Ile	Glu	Ala
			420					425					430		
Gly	Val	Pro	Lys	Met	Lys	Ile	Glu	Glu	Ala	Ala	Ala	Arg	Arg	Gln	Ala
		435					440					445			
Arg	Ile	Asp	Ser	Gln	Ala	Glu	Ile	Ile	Val	Gly	Val	Asn	Gln	Phe	Gln
		450				455						460			
Pro	Glu	Gln	Glu	Glu	Pro	Leu	Asp	Ile	Leu	Asp	Ile	Asp	Asn	Thr	Ala
465					470					475					480
Val	Arg	Met	Lys	Gln	Leu	Glu	Lys	Leu	Lys	Lys	Ile	Arg	Ser	Glu	Arg
				485					490					495	
Asn	Glu	Gln	Ala	Val	Ile	Glu	Ala	Leu	Asn	Arg	Leu	Thr	Asn	Cys	Ala

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                    500                505                510
Lys Thr Gly Glu Gly Asn Leu Leu Ala Phe Ala Val Glu Ala Ala Arg
      515                      520                      525

Ala Arg Ala Thr Leu Gly Glu Ile Ser Glu Ala Ile Glu Lys Val Ala
      530                      535                      540

Gly Arg His Gln Ala Thr Ser Lys Ser Val Ser Gly Val Tyr Ser Ala
545                      550                      555                      560

Glu Phe Val His Arg Asp Gln Ile Glu Glu Val Arg Lys Leu Thr Ala
      565                      570                      575

Glu Phe Leu Glu Gly Glu Gly Arg Arg Pro Arg Ile Leu Val Ala Lys
      580                      585                      590

Met Gly Gln Asp Gly His Asp Arg Gly Ser Lys Val Ile Ser Thr Ala
      595                      600                      605

Phe Ala Asp Leu Gly Phe Asp Val Asp Ile Gly Pro Leu Phe Gln Thr
      610                      615                      620

Pro Gln Glu Thr Ala Arg Gln Ala Val Glu Asn Asp Val His Val Ile
625                      630                      635                      640

Gly Ile Ser Ser Leu Ala Ala Gly His Lys Thr Leu Leu Pro Gln Leu
      645                      650                      655

Val Asp Glu Leu Lys Lys Leu Glu Arg Asp Asp Ile Val Val Ile Val
      660                      665                      670

Gly Gly Val Ile Pro Lys Gln Asp Tyr Ser Phe Leu Leu Glu His Gly
      675                      680                      685

Ala Ser Ala Ile Phe Gly Pro Gly Thr Val Ile Pro Lys Ala Ala Val
      690                      695                      700

Ser Val Leu His Glu Ile Lys Lys Arg Leu Glu Glu
705                      710                      715

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<210> SEQ ID NO 63
<211> LENGTH: 615
<212> TYPE: PRT
<213> ORGANISM: Mycobacterium tuberculosis
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / YP_001282809
<309> DATABASE ENTRY DATE: 2010-05-13
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(615)

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<400> SEQUENCE: 63

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Met Ser Ile Asp Val Pro Glu Arg Ala Asp Leu Glu Gln Val Arg Gly
1                      5                      10                      15

Arg Trp Arg Asn Ala Val Ala Gly Val Leu Ser Lys Ser Asn Arg Thr
      20                      25                      30

Asp Ser Ala Gln Leu Gly Asp His Pro Glu Arg Leu Leu Asp Thr Gln
      35                      40                      45

Thr Ala Asp Gly Phe Ala Ile Arg Ala Leu Tyr Thr Ala Phe Asp Glu
      50                      55                      60

Leu Pro Glu Pro Pro Leu Pro Gly Gln Trp Pro Phe Val Arg Gly Gly
65                      70                      75                      80

Asp Pro Leu Arg Asp Val His Ser Gly Trp Lys Val Ala Glu Ala Phe
      85                      90                      95

Pro Ala Asn Gly Ala Thr Ala Asp Thr Asn Ala Ala Val Leu Ala Ala
      100                     105                     110

Leu Gly Glu Gly Val Ser Ala Leu Leu Ile Arg Val Gly Glu Ser Gly
115                     120                     125

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Val Ala Pro Asp Arg Leu Thr Ala Leu Leu Ser Gly Val Tyr Leu Asn
 130 135 140
 Leu Ala Pro Val Ile Leu Asp Ala Gly Ala Asp Tyr Arg Pro Ala Cys
 145 150 155 160
 Asp Val Met Leu Ala Leu Val Ala Gln Leu Asp Pro Gly Gln Arg Asp
 165 170 175
 Thr Leu Ser Ile Asp Leu Gly Ala Asp Pro Leu Thr Ala Ser Leu Arg
 180 185 190
 Asp Arg Pro Ala Pro Pro Ile Glu Glu Val Val Ala Val Ala Ser Arg
 195 200 205
 Ala Ala Gly Glu Arg Gly Leu Arg Ala Ile Thr Val Asp Gly Pro Ala
 210 215 220
 Phe His Asn Leu Gly Ala Thr Ala Ala Thr Glu Leu Ala Ala Thr Val
 225 230 235 240
 Ala Ala Ala Val Ala Tyr Leu Arg Val Leu Thr Glu Ser Gly Leu Val
 245 250 255
 Val Ser Asp Ala Leu Arg Gln Ile Ser Phe Arg Leu Ala Ala Asp Asp
 260 265 270
 Asp Gln Phe Met Thr Leu Ala Lys Met Arg Ala Leu Arg Gln Leu Trp
 275 280 285
 Ala Arg Val Ala Glu Val Val Gly Asp Pro Gly Gly Gly Ala Ala Val
 290 295 300
 Val His Ala Glu Thr Ser Leu Pro Met Met Thr Gln Arg Asp Pro Trp
 305 310 315
 Val Asn Met Leu Arg Cys Thr Leu Ala Ala Phe Gly Ala Gly Val Gly
 325 330 335
 Gly Ala Asp Thr Val Leu Val His Pro Phe Asp Val Ala Ile Pro Gly
 340 345 350
 Gly Phe Pro Gly Thr Ala Ala Gly Phe Ala Arg Arg Ile Ala Arg Asn
 355 360 365
 Thr Gln Leu Leu Leu Leu Glu Ser His Val Gly Arg Val Leu Asp
 370 375 380
 Pro Ala Gly Gly Ser Trp Phe Val Glu Glu Leu Thr Asp Arg Leu Ala
 385 390 395 400
 Arg Arg Ala Trp Gln Arg Phe Gln Ala Ile Glu Ala Arg Gly Gly Phe
 405 410 415
 Val Glu Ala His Asp Phe Leu Ala Gly Gln Ile Ala Glu Cys Ala Ala
 420 425 430
 Arg Arg Ala Asp Asp Ile Ala His Arg Arg Leu Ala Ile Thr Gly Val
 435 440 445
 Asn Glu Tyr Pro Asn Leu Gly Glu Pro Ala Leu Pro Pro Gly Asp Pro
 450 455 460
 Thr Ser Pro Val Arg Arg Tyr Ala Ala Gly Phe Glu Ala Leu Arg Asp
 465 470 475 480
 Arg Ser Asp His His Leu Ala Arg Thr Gly Ala Arg Pro Arg Val Leu
 485 490 495
 Leu Leu Pro Leu Gly Pro Leu Ala Glu His Asn Ile Arg Thr Thr Phe
 500 505 510
 Ala Thr Asn Leu Leu Ala Ser Gly Gly Ile Glu Ala Ile Asp Pro Gly
 515 520 525

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Thr Val Asp Ala Gly Thr Val Gly Asn Ala Val Ala Asp Ala Gly Ser
  530                               535                               540

Pro Ser Val Ala Val Ile Cys Gly Thr Asp Ala Arg Tyr Arg Asp Glu
545                               550                               555                               560

Val Ala Asp Ile Val Gln Ala Ala Arg Ala Ala Gly Val Ser Arg Val
                               565                               570                               575

Tyr Leu Ala Gly Pro Glu Lys Ala Leu Gly Asp Ala Ala His Arg Pro
  580                               585                               590

Asp Glu Phe Leu Thr Ala Lys Ile Asn Val Val Gln Ala Leu Ser Asn
  595                               600                               605

Leu Leu Thr Arg Leu Gly Ala
  610                               615

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<210> SEQ ID NO 64
<211> LENGTH: 750
<212> TYPE: PRT
<213> ORGANISM: Mycobacterium tuberculosis
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / YP_001282810
<309> DATABASE ENTRY DATE: 2010-05-13
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(750)

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<400> SEQUENCE: 64

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Met Thr Thr Lys Thr Pro Val Ile Gly Ser Phe Ala Gly Val Pro Leu
  1                               5                               10                               15

His Ser Glu Arg Ala Ala Gln Ser Pro Thr Glu Ala Ala Val His Thr
                               20                               25                               30

His Val Ala Ala Ala Ala Ala Ala His Gly Tyr Thr Pro Glu Gln Leu
  35                               40                               45

Val Trp His Thr Pro Glu Gly Ile Asp Val Thr Pro Val Tyr Ile Ala
  50                               55                               60

Ala Asp Arg Ala Ala Ala Glu Ala Glu Gly Tyr Pro Leu His Ser Phe
  65                               70                               75                               80

Pro Gly Glu Pro Pro Phe Val Arg Gly Pro Tyr Pro Thr Met Tyr Val
                               85                               90                               95

Asn Gln Pro Trp Thr Ile Arg Gln Tyr Ala Gly Phe Ser Thr Ala Ala
  100                              105                              110

Asp Ser Asn Ala Phe Tyr Arg Arg Asn Leu Ala Ala Gly Gln Lys Gly
  115                              120                              125

Leu Ser Val Ala Phe Asp Leu Ala Thr His Arg Gly Tyr Asp Ser Asp
  130                              135                              140

His Pro Arg Val Gln Gly Asp Val Gly Met Ala Gly Val Ala Ile Asp
  145                              150                              155                              160

Ser Ile Leu Asp Met Arg Gln Leu Phe Asp Gly Ile Asp Leu Ser Thr
  165                              170                              175

Val Ser Val Ser Met Thr Met Asn Gly Ala Val Leu Pro Ile Leu Ala
  180                              185                              190

Leu Tyr Val Val Ala Ala Glu Glu Gln Gly Val Ala Pro Glu Gln Leu
  195                              200                              205

Ala Gly Thr Ile Gln Asn Asp Ile Leu Lys Glu Phe Met Val Arg Asn
  210                              215                              220

Thr Tyr Ile Tyr Pro Pro Lys Pro Ser Met Arg Ile Ile Ser Asp Ile
  225                              230                              235                              240

Phe Ala Tyr Thr Ser Ala Lys Met Pro Lys Phe Asn Ser Ile Ser Ile

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245					250					255					
Ser	Gly	Tyr	His	Ile	Gln	Glu	Ala	Gly	Ala	Thr	Ala	Asp	Leu	Glu	Leu
			260					265					270		
Ala	Tyr	Thr	Leu	Ala	Asp	Gly	Val	Asp	Tyr	Ile	Arg	Ala	Gly	Leu	Asn
		275					280					285			
Ala	Gly	Leu	Asp	Ile	Asp	Ser	Phe	Ala	Pro	Arg	Leu	Ser	Phe	Phe	Trp
	290					295					300				
Gly	Ile	Gly	Met	Asn	Phe	Phe	Met	Glu	Val	Ala	Lys	Leu	Arg	Ala	Gly
305				310						315					320
Arg	Leu	Leu	Trp	Ser	Glu	Leu	Val	Ala	Gln	Phe	Ala	Pro	Lys	Ser	Ala
				325					330					335	
Lys	Ser	Leu	Ser	Leu	Arg	Thr	His	Ser	Gln	Thr	Ser	Gly	Trp	Ser	Leu
			340					345					350		
Thr	Ala	Gln	Asp	Val	Phe	Asn	Asn	Val	Ala	Arg	Thr	Cys	Ile	Glu	Ala
		355					360					365			
Met	Ala	Ala	Thr	Gln	Gly	His	Thr	Gln	Ser	Leu	His	Thr	Asn	Ala	Leu
	370					375					380				
Asp	Glu	Ala	Leu	Ala	Leu	Pro	Thr	Asp	Phe	Ser	Ala	Arg	Ile	Ala	Arg
385				390						395					400
Asn	Thr	Gln	Leu	Val	Leu	Gln	Gln	Glu	Ser	Gly	Thr	Thr	Arg	Pro	Ile
			405						410					415	
Asp	Pro	Trp	Gly	Gly	Ser	Tyr	Tyr	Val	Glu	Trp	Leu	Thr	His	Arg	Leu
			420					425					430		
Ala	Arg	Arg	Ala	Arg	Ala	His	Ile	Ala	Glu	Val	Ala	Glu	His	Gly	Gly
		435					440					445			
Met	Ala	Gln	Ala	Ile	Ser	Asp	Gly	Ile	Pro	Lys	Leu	Arg	Ile	Glu	Glu
	450					455					460				
Ala	Ala	Ala	Arg	Thr	Gln	Ala	Arg	Ile	Asp	Ser	Gly	Gln	Gln	Pro	Val
465				470						475					480
Val	Gly	Val	Asn	Lys	Tyr	Gln	Val	Pro	Glu	Asp	His	Glu	Ile	Glu	Val
			485						490					495	
Leu	Lys	Val	Glu	Asn	Ser	Arg	Val	Arg	Ala	Glu	Gln	Leu	Ala	Lys	Leu
			500					505					510		
Gln	Arg	Leu	Arg	Ala	Gly	Arg	Asp	Glu	Pro	Ala	Val	Arg	Ala	Ala	Leu
		515					520					525			
Ala	Glu	Leu	Thr	Arg	Ala	Ala	Ala	Glu	Gln	Gly	Arg	Ala	Gly	Ala	Asp
	530					535					540				
Gly	Leu	Gly	Asn	Asn	Leu	Leu	Ala	Leu	Ala	Ile	Asp	Ala	Ala	Arg	Ala
545				550						555					560
Gln	Ala	Thr	Val	Gly	Glu	Ile	Ser	Glu	Ala	Leu	Glu	Lys	Val	Tyr	Gly
				565					570					575	
Arg	His	Arg	Ala	Glu	Ile	Arg	Thr	Ile	Ser	Gly	Val	Tyr	Arg	Asp	Glu
			580					585					590		
Val	Gly	Lys	Ala	Pro	Asn	Ile	Ala	Ala	Ala	Thr	Glu	Leu	Val	Glu	Lys
		595					600					605			
Phe	Ala	Glu	Ala	Asp	Gly	Arg	Arg	Pro	Arg	Ile	Leu	Ile	Ala	Lys	Met
	610					615					620				
Gly	Gln	Asp	Gly	His	Asp	Arg	Gly	Gln	Lys	Val	Ile	Ala	Thr	Ala	Phe
625				630						635					640
Ala	Asp	Ile	Gly	Phe	Asp	Val	Asp	Val	Gly	Ser	Leu	Phe	Ser	Thr	Pro
				645					650					655	

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Glu Glu Val Ala Arg Gln Ala Ala Asp Asn Asp Val His Val Ile Gly
660 665 670

Val Ser Ser Leu Ala Ala Gly His Leu Thr Leu Val Pro Ala Leu Arg
675 680 685

Asp Ala Leu Ala Gln Val Gly Arg Pro Asp Ile Met Ile Val Val Gly
690 695 700

Gly Val Ile Pro Pro Gly Asp Phe Asp Glu Leu Tyr Ala Ala Gly Ala
705 710 715 720

Thr Ala Ile Phe Pro Pro Gly Thr Val Ile Ala Asp Ala Ala Ile Asp
725 730 735

Leu Leu His Arg Leu Ala Glu Arg Leu Gly Tyr Thr Leu Asp
740 745 750

<210> SEQ ID NO 65
 <211> LENGTH: 616
 <212> TYPE: PRT
 <213> ORGANISM: Corynebacterium glutamicum
 <300> PUBLICATION INFORMATION:
 <308> DATABASE ACCESSION NUMBER: NCBI / YP_225814
 <309> DATABASE ENTRY DATE: 2010-12-14
 <313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(616)

<400> SEQUENCE: 65

Met Thr Asp Leu Thr Lys Thr Ala Val Pro Glu Glu Leu Ser Glu Asn
1 5 10 15

Leu Glu Thr Trp Tyr Lys Ala Val Ala Gly Val Phe Ala Arg Thr Gln
20 25 30

Lys Lys Asp Ile Gly Asp Ile Ala Val Asp Val Trp Lys Lys Leu Ile
35 40 45

Val Thr Thr Pro Asp Gly Val Asp Ile Asn Pro Leu Tyr Thr Arg Ala
50 55 60

Asp Glu Ser Gln Arg Lys Phe Thr Glu Val Pro Gly Glu Phe Pro Phe
65 70 75 80

Thr Arg Gly Thr Thr Val Asp Gly Glu Arg Val Gly Trp Gly Val Thr
85 90 95

Glu Thr Phe Gly His Asp Ser Pro Lys Asn Ile Asn Ala Ala Val Leu
100 105 110

Asn Ala Leu Asn Ser Gly Thr Thr Thr Leu Gly Phe Glu Phe Ser Glu
115 120 125

Glu Phe Thr Ala Ala Asp Leu Lys Val Ala Leu Glu Gly Val Tyr Leu
130 135 140

Asn Met Ala Pro Leu Leu Ile His Ala Gly Gly Ser Thr Ser Glu Val
145 150 155 160

Ala Ala Ala Leu Tyr Thr Leu Ala Glu Glu Ala Gly Thr Phe Phe Ala
165 170 175

Ala Leu Thr Leu Gly Ser Arg Pro Leu Thr Ala Gln Val Asp Gly Ser
180 185 190

His Ser Asp Thr Ile Glu Glu Ala Val Gln Leu Ala Val Asn Ala Ser
195 200 205

Lys Arg Ala Asn Val Arg Ala Ile Leu Val Asp Gly Ser Ser Phe Ser
210 215 220

Asn Gln Gly Ala Ser Asp Ala Gln Glu Ile Gly Leu Ser Ile Ala Ala
225 230 235 240

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Gly Val Asp Tyr Val Arg Arg Leu Val Asp Ala Gly Leu Ser Thr Glu
 245 250 255
 Ala Ala Leu Lys Gln Val Ala Phe Arg Phe Ala Val Thr Asp Glu Gln
 260 265 270
 Phe Ala Gln Ile Ser Lys Leu Arg Val Ala Arg Arg Leu Trp Ala Arg
 275 280 285
 Val Cys Glu Val Leu Gly Phe Pro Glu Leu Ala Val Ala Pro Gln His
 290 295 300
 Ala Val Thr Ala Arg Ala Met Phe Ser Gln Arg Asp Pro Trp Val Asn
 305 310 315 320
 Met Leu Arg Ser Thr Val Ala Ala Phe Ala Ala Gly Val Gly Gly Ala
 325 330 335
 Thr Asp Val Glu Val Arg Thr Phe Asp Asp Ala Ile Pro Asp Gly Val
 340 345 350
 Pro Gly Val Ser Arg Asn Phe Ala His Arg Ile Ala Arg Asn Thr Asn
 355 360 365
 Leu Leu Leu Leu Glu Glu Ser His Leu Gly His Val Val Asp Pro Ala
 370 375 380
 Gly Gly Ser Tyr Phe Val Glu Ser Phe Thr Asp Asp Leu Ala Glu Lys
 385 390 395 400
 Ala Trp Ala Val Phe Ser Gly Ile Glu Ala Glu Gly Gly Tyr Ser Ala
 405 410 415
 Ala Cys Ala Ser Gly Thr Val Thr Ala Met Leu Asp Gln Thr Trp Glu
 420 425 430
 Gln Thr Arg Ala Asp Val Ala Ser Arg Lys Lys Lys Leu Thr Gly Ile
 435 440 445
 Asn Glu Phe Pro Asn Leu Ala Glu Ser Pro Leu Pro Ala Asp Arg Arg
 450 455 460
 Val Glu Pro Ala Gly Val Arg Arg Trp Ala Ala Asp Phe Glu Ala Leu
 465 470 475 480
 Arg Asn Arg Ser Asp Ala Phe Leu Glu Lys Asn Gly Ala Arg Pro Gln
 485 490 495
 Ile Thr Met Ile Pro Leu Gly Pro Leu Ser Lys His Asn Ile Arg Thr
 500 505 510
 Gly Phe Thr Ser Asn Leu Leu Ala Ser Gly Gly Ile Glu Ala Ile Asn
 515 520 525
 Pro Gly Gln Leu Val Pro Gly Thr Asp Ala Phe Ala Glu Ala Ala Gln
 530 535 540
 Ala Ala Gly Ile Val Val Val Cys Gly Thr Asp Gln Glu Tyr Ala Glu
 545 550 555 560
 Thr Gly Glu Gly Ala Val Glu Lys Leu Arg Glu Ala Gly Val Glu Arg
 565 570 575
 Ile Leu Leu Ala Gly Ala Pro Lys Ser Phe Glu Gly Ser Ala His Ala
 580 585 590
 Pro Asp Gly Tyr Leu Asn Met Thr Ile Asp Ala Ala Ala Thr Leu Ala
 595 600 605
 Asp Leu Leu Asp Ala Leu Gly Ala
 610 615

<210> SEQ ID NO 66

<211> LENGTH: 737

<212> TYPE: PRT

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<213> ORGANISM: Corynebacterium glutamicum
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / YP_225813
<309> DATABASE ENTRY DATE: 2010-12-14
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(737)

<400> SEQUENCE: 66

Met Thr Ser Ile Pro Asn Phe Ser Asp Ile Pro Leu Thr Ala Glu Thr
 1                               5                               10          15

Arg Ala Ser Glu Ser His Asn Val Asp Ala Gly Lys Val Trp Asn Thr
 20                               25                               30

Pro Glu Gly Ile Asp Val Lys Arg Val Phe Thr Gln Ala Asp Arg Asp
 35                               40                               45

Glu Ala Gln Ala Ala Gly His Pro Val Asp Ser Leu Pro Gly Gln Lys
 50                               55                               60

Pro Phe Met Arg Gly Pro Tyr Pro Thr Met Tyr Thr Asn Gln Pro Trp
 65                               70                               75          80

Thr Ile Arg Gln Tyr Ala Gly Phe Ser Thr Ala Ala Glu Ser Asn Ala
 85                               90

Phe Tyr Arg Arg Asn Leu Ala Ala Gly Gln Lys Gly Leu Ser Val Ala
 100                              105          110

Phe Asp Leu Ala Thr His Arg Gly Tyr Asp Ser Asp Asn Glu Arg Val
 115                              120          125

Val Gly Asp Val Gly Met Ala Gly Val Ala Ile Asp Ser Ile Leu Asp
 130                              135          140

Met Arg Gln Leu Phe Asp Gly Ile Asp Leu Ser Ser Val Ser Val Ser
 145                              150          155          160

Met Thr Met Asn Gly Ala Val Leu Pro Ile Leu Ala Phe Tyr Ile Val
 165                              170          175

Ala Ala Glu Glu Gln Gly Val Gly Pro Glu Gln Leu Ala Gly Thr Ile
 180                              185          190

Gln Asn Asp Ile Leu Lys Glu Phe Met Val Arg Asn Thr Tyr Ile Tyr
 195                              200          205

Pro Pro Lys Pro Ser Met Arg Ile Ile Ser Asn Ile Phe Glu Tyr Thr
 210                              215          220

Ser Leu Lys Met Pro Arg Phe Asn Ser Ile Ser Ile Ser Gly Tyr His
 225                              230          235          240

Ile Gln Glu Ala Gly Ala Thr Ala Asp Leu Glu Leu Ala Tyr Thr Leu
 245                              250          255

Ala Asp Gly Ile Glu Tyr Ile Arg Ala Gly Lys Glu Val Gly Leu Asp
 260                              265          270

Val Asp Lys Phe Ala Pro Arg Leu Ser Phe Phe Trp Gly Ile Ser Met
 275                              280          285

Tyr Thr Phe Met Glu Ile Ala Lys Leu Arg Ala Gly Arg Leu Leu Trp
 290                              295          300

Ser Glu Leu Val Ala Lys Phe Asp Pro Lys Asn Ala Lys Ser Gln Ser
 305                              310          315          320

Leu Arg Thr His Ser Gln Thr Ser Gly Trp Ser Leu Thr Ala Gln Asp
 325                              330          335

Val Tyr Asn Asn Val Ala Arg Thr Ala Ile Glu Ala Met Ala Ala Thr
 340                              345          350

Gln Gly His Thr Gln Ser Leu His Thr Asn Ala Leu Asp Glu Ala Leu
 355                              360          365

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Ala Leu Pro Thr Asp Phe Ser Ala Arg Ile Ala Arg Asn Thr Gln Leu
370 375 380

Leu Leu Gln Gln Glu Ser Gly Thr Val Arg Pro Val Asp Pro Trp Ala
385 390 395 400

Gly Ser Tyr Tyr Val Glu Trp Leu Thr Asn Glu Leu Ala Asn Arg Ala
405 410 415

Arg Lys His Ile Asp Glu Val Glu Glu Ala Gly Gly Met Ala Gln Ala
420 425 430

Thr Ala Gln Gly Ile Pro Lys Leu Arg Ile Glu Glu Ser Ala Ala Arg
435 440 445

Thr Gln Ala Arg Ile Asp Ser Gly Arg Gln Ala Leu Ile Gly Val Asn
450 455 460

Arg Tyr Val Ala Glu Glu Asp Glu Glu Ile Glu Val Leu Lys Val Asp
465 470 475 480

Asn Thr Lys Val Arg Ala Glu Gln Leu Ala Lys Leu Ala Gln Leu Lys
485 490 495

Ala Glu Arg Asn Asp Ala Glu Val Lys Ala Ala Leu Asp Ala Leu Thr
500 505 510

Ala Ala Ala Arg Asn Glu His Lys Glu Pro Gly Asp Leu Asp Gln Asn
515 520 525

Leu Leu Lys Leu Ala Val Asp Ala Ala Arg Ala Lys Ala Thr Ile Gly
530 535 540

Glu Ile Ser Asp Ala Leu Glu Val Val Phe Gly Arg His Glu Ala Glu
545 550 555 560

Ile Arg Thr Leu Ser Gly Val Tyr Lys Asp Glu Val Gly Lys Glu Gly
565 570 575

Thr Val Ser Asn Val Glu Arg Ala Ile Ala Leu Ala Asp Ala Phe Glu
580 585 590

Ala Glu Glu Gly Arg Arg Pro Arg Ile Phe Ile Ala Lys Met Gly Gln
595 600 605

Asp Gly His Asp Arg Gly Gln Lys Val Val Ala Ser Ala Tyr Ala Asp
610 615 620

Leu Gly Met Asp Val Asp Val Gly Pro Leu Phe Gln Thr Pro Ala Glu
625 630 635 640

Ala Ala Arg Ala Ala Val Asp Ala Asp Val His Val Val Gly Met Ser
645 650 655

Ser Leu Ala Ala Gly His Leu Thr Leu Leu Pro Glu Leu Lys Lys Glu
660 665 670

Leu Ala Ala Leu Gly Arg Asp Asp Ile Leu Val Thr Val Gly Gly Val
675 680 685

Ile Pro Pro Gly Asp Phe Gln Asp Leu Tyr Asp Met Gly Ala Ala Ala
690 695 700

Ile Tyr Pro Pro Gly Thr Val Ile Ala Glu Ser Ala Ile Asp Leu Ile
705 710 715 720

Thr Arg Leu Ala Ala His Leu Gly Phe Asp Leu Asp Val Asp Val Asn
725 730 735

Glu

<210> SEQ ID NO 67
<211> LENGTH: 631
<212> TYPE: PRT

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<213> ORGANISM: Rhodococcus erythropolis
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / YP_002766535
<309> DATABASE ENTRY DATE: 2010-05-12
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(631)

<400> SEQUENCE: 67

Met Ser Leu Ala Ser Glu Ala Glu Ala Val Glu Gln Ala Tyr Ala Glu
1          5          10          15

Trp Gln Arg Ser Val Ala Gly Val Leu Ala Lys Ser Arg Arg Val Asp
20          25          30

Ala Ala Glu Leu Gly Pro Glu Pro Gln Lys Leu Leu Glu Thr Val Thr
35          40          45

Tyr Asp Gly Val Thr Val Ala Pro Leu Tyr Ser Pro Arg Asp Glu Arg
50          55          60

Pro Glu Gln Ser Leu Pro Gly Thr Phe Pro Tyr Val Arg Gly Val Asp
65          70          75          80

Ala His Arg Asp Val Asn Ala Gly Trp Leu Val Ser Ala Ala Phe Gly
85          90          95

Thr Ala Ser Ala Ala Glu Thr Asn Arg Ala Ile Leu Asp Ala Leu Glu
100         105         110

Asn Gly Val Ser Ala Leu Trp Leu Lys Val Gly Ala Asp Gly Val Pro
115         120         125

Val Thr Asp Leu Ala Ala Ala Leu Glu Gly Val Leu Leu Asp Leu Ala
130         135         140

Pro Leu Thr Leu Asp Ala Gly Ala Glu Val Asn Asp Ala Ala Arg Ala
145         150         155         160

Leu Phe Ser Leu Leu Asp Ala Arg Gly Glu Ala Gly Asp Gly Val Ser
165         170         175

Asp Arg Ser Ser Ile Arg Val His Leu Gly Ala Ala Pro Leu Thr Ser
180         185         190

Ser Phe Ser Gly Ala Ala Asp Val Glu Phe Ala Gly Ala Val Glu Leu
195         200         205

Ala Ala Leu Ala Ala Ala Arg Ala Glu Thr Val His Ala Ile Thr Val
210         215         220

Asp Gly Thr Ala Phe His Asn Ala Gly Ala Gly Asp Ala Glu Glu Leu
225         230         235         240

Gly Ala Ala Ile Ala Ala Gly Leu Glu Tyr Leu Arg Ala Leu Thr Ala
245         250         255

Glu Ser Gly Leu Thr Ile Gly Ala Ala Leu Ser Gln Leu Ala Phe Arg
260         265         270

Tyr Ser Ala Thr Asp Asp Gln Phe Gln Thr Ile Ala Lys Phe Arg Ala
275         280         285

Ala Arg Leu Val Trp Ala Arg Ile Ala Gln Val Cys Gly Ala Ser Asp
290         295         300

Phe Gly Gly Ala Pro Gln His Ala Val Thr Ser Ala Ala Met Met Ala
305         310         315         320

Gln Arg Asp Pro Trp Val Asn Met Leu Arg Thr Thr Leu Ala Ala Phe
325         330         335

Gly Ala Gly Val Gly Gly Ala Asp Ala Val Thr Val Leu Pro Phe Asp
340         345         350

Val Ala Leu Ala Asp Gly Thr Leu Gly Val Ser Lys Ser Phe Ser Ser
355         360         365

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Arg Ile Ala Arg Asn Thr Gln Leu Leu Leu Leu Glu Glu Ser His Leu
 370                375                380

Gly Arg Val Leu Asp Pro Ser Ala Gly Ser Trp Tyr Val Glu Asp Leu
 385                390                395                400

Thr Gln Gln Ile Ala Ala Thr Ala Trp Glu Phe Phe Gln Glu Ile Glu
                405                410                415

Ala Ala Gly Gly Tyr Leu Ala Ala Leu Glu Ala Gly Ile Val Ser Gly
                420                425                430

Arg Ile Ala Ala Thr Lys Ala Lys Arg Asp Ser Asp Ile Ala His Arg
 435                440                445

Lys Thr Thr Val Thr Gly Val Asn Glu Phe Pro Asn Leu Gly Glu Thr
 450                455                460

Pro Leu Ser Ala Glu Ala Val Glu Pro Gly Gln Ser Val Ala Arg Tyr
 465                470                475                480

Ala Ala Ala Phe Glu Ala Leu Arg Asp Arg Ser Asp Ala Phe Leu Ala
                485                490                495

Ala Gly Gly Ala Arg Pro Thr Ala Leu Leu Ala Pro Leu Gly Ser Val
                500                505                510

Ala Glu His Asn Val Arg Thr Thr Phe Ala Ser Asn Leu Leu Ala Ser
 515                520                525

Gly Gly Ile Asp Ala Val Asn Pro Gly Pro Leu Glu Val Gly Ala Glu
 530                535                540

Ala Ile Ser Ala Ala Val Lys Ala Ser Gly Val Thr Val Ala Val Leu
 545                550                555                560

Cys Gly Thr Asp Lys Arg Tyr Gly Glu Ser Ala Ala Ala Val Ala
 565                570                575

Glu Leu Arg Ala Ala Gly Ile Thr Lys Val Leu Leu Ala Gly Pro Glu
 580                585                590

Lys Ala Val Ala Asp Ala Thr Gly Glu Ser Arg Pro Asp Gly Phe Leu
 595                600                605

Thr Ala Arg Ile Asp Ala Val Ser Ala Leu Thr Glu Leu Leu Asp Phe
 610                615                620

Ile Glu Thr Gly Ser Ser Lys
 625                630

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<210> SEQ ID NO 68

<211> LENGTH: 750

<212> TYPE: PRT

<213> ORGANISM: Rhodococcus erythropolis

<300> PUBLICATION INFORMATION:

<308> DATABASE ACCESSION NUMBER: NCBI / YP_002766536

<309> DATABASE ENTRY DATE: 2010-05-12

<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(750)

<400> SEQUENCE: 68

```

Met Thr Thr Arg Glu Val Lys His Val Ile Gly Ser Phe Ala Glu Val
 1                5                10                15

Pro Leu Glu Asp Pro Gln Ser Pro Ala Pro Thr Pro Pro Ser Val Glu
 20                25                30

Gln Ala Gln Ala Leu Ile Glu Glu Gly Ala Asn Ala Asn Asn Tyr Ala
 35                40                45

Ala Glu Gln Val Val Trp Ser Thr Pro Glu Gly Ile Asp Val Lys Pro
 50                55                60

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Val	Tyr	Thr	Gly	Ala	Asp	Arg	Thr	Ala	Ala	Ala	Glu	Ser	Gly	Tyr	Pro
65					70					75					80
Leu	Asp	Ser	Phe	Pro	Gly	Ala	Ala	Pro	Phe	Leu	Arg	Gly	Pro	Tyr	Pro
				85					90					95	
Thr	Met	Tyr	Val	Asn	Gln	Pro	Trp	Thr	Ile	Arg	Gln	Tyr	Ala	Gly	Phe
			100					105					110		
Ser	Thr	Ala	Ala	Glu	Ser	Asn	Ala	Phe	Tyr	Arg	Arg	Asn	Leu	Ala	Ala
		115					120					125			
Gly	Gln	Lys	Gly	Leu	Ser	Val	Ala	Phe	Asp	Leu	Ala	Thr	His	Arg	Gly
		130				135					140				
Tyr	Asp	Ser	Asp	His	Pro	Arg	Val	Ala	Gly	Asp	Val	Gly	Met	Ala	Gly
145					150					155					160
Val	Ala	Ile	Asp	Ser	Ile	Leu	Asp	Met	Arg	Gln	Leu	Phe	Asp	Gly	Ile
				165					170					175	
Asp	Leu	Ser	Gln	Val	Ser	Val	Ser	Met	Thr	Met	Asn	Gly	Ala	Val	Leu
			180					185					190		
Pro	Ile	Leu	Ala	Leu	Tyr	Val	Ala	Ala	Ala	Gly	Glu	Gln	Gly	Val	Thr
		195				200						205			
Pro	Asp	Lys	Leu	Ala	Gly	Thr	Ile	Gln	Asn	Asp	Ile	Leu	Lys	Glu	Phe
		210				215					220				
Met	Val	Arg	Asn	Thr	Tyr	Ile	Tyr	Pro	Pro	Lys	Pro	Ser	Met	Arg	Ile
225					230					235					240
Ile	Ser	Asp	Ile	Phe	Ala	Tyr	Ser	Ser	Ala	Glu	Met	Pro	Lys	Tyr	Asn
				245					250					255	
Ser	Ile	Ser	Ile	Ser	Gly	Tyr	His	Ile	Gln	Glu	Ala	Gly	Ala	Thr	Ala
			260					265					270		
Asp	Leu	Glu	Leu	Ala	Tyr	Thr	Leu	Ala	Asp	Gly	Val	Glu	Tyr	Ile	Arg
		275					280					285			
Ala	Gly	Leu	Asp	Ala	Gly	Met	Asp	Ile	Asp	Lys	Phe	Ala	Pro	Arg	Leu
		290				295					300				
Ser	Phe	Phe	Trp	Ala	Ile	Gly	Met	Asn	Phe	Phe	Met	Glu	Val	Ala	Lys
305					310					315					320
Leu	Arg	Ala	Gly	Arg	Leu	Leu	Trp	Ala	Glu	Leu	Val	Ala	Lys	Phe	Asp
				325					330					335	
Pro	Lys	Ser	Ala	Lys	Ser	Leu	Ser	Leu	Arg	Thr	His	Ser	Gln	Thr	Ser
			340					345					350		
Gly	Trp	Ser	Leu	Thr	Ala	Gln	Asp	Val	Phe	Asn	Asn	Val	Pro	Arg	Thr
		355					360					365			
Cys	Val	Glu	Ala	Met	Ala	Ala	Thr	Gln	Gly	His	Thr	Gln	Ser	Leu	His
	370					375						380			
Thr	Asn	Ala	Leu	Asp	Glu	Ala	Ile	Ala	Leu	Pro	Thr	Asp	Phe	Ser	Ala
385					390					395					400
Arg	Ile	Ala	Arg	Asn	Thr	Gln	Leu	Leu	Leu	Gln	Gln	Glu	Ser	Gly	Thr
				405					410					415	
Val	Arg	Pro	Ile	Asp	Pro	Trp	Gly	Gly	Ser	Tyr	Tyr	Val	Glu	Trp	Leu
			420					425					430		
Thr	Asn	Glu	Leu	Ala	Asn	Arg	Ala	Arg	Lys	His	Ile	Glu	Glu	Val	Glu
		435					440					445			
Glu	Ala	Gly	Gly	Met	Ala	Gln	Ala	Ile	Asn	Glu	Gly	Ile	Pro	Lys	Leu
	450					455					460				
Arg	Ile	Glu	Glu	Ala	Ala	Ala	Arg	Thr	Gln	Ala	Arg	Ile	Asp	Ser	Gly

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465                470                475                480
Arg Gln Pro Leu Val Gly Val Asn Lys Tyr Val Pro Asp Glu Val Asp
      485                490                495
Thr Ile Glu Val Leu Lys Val Glu Asn Ser Lys Val Arg Lys Glu Gln
      500                505                510
Leu Glu Lys Leu Val Arg Leu Arg Ala Glu Arg Asp Pro Glu Ala Val
      515                520                525
Glu Ala Ala Leu Ala Asn Leu Thr Arg Ala Ala Ala Ser Thr Glu Gly
      530                535                540
Gly Met Glu Asn Asn Leu Leu Ala Leu Ala Val Val Ala Ala Arg Ala
      545                550                555                560
Met Ala Thr Val Gly Glu Ile Ser Asp Ala Leu Glu Lys Val Tyr Gly
      565                570                575
Arg His Gln Ala Glu Ile Arg Thr Ile Ser Gly Val Tyr Arg Asp Glu
      580                585                590
Ala Gly Thr Val Ser Asn Ile Ser Lys Ala Met Glu Leu Val Glu Lys
      595                600                605
Phe Ala Glu Asp Glu Gly Arg Arg Pro Arg Ile Leu Val Ala Lys Met
      610                615                620
Gly Gln Asp Gly His Asp Arg Gly Gln Lys Val Ile Ser Thr Ala Phe
      625                630                635                640
Ala Asp Ile Gly Phe Asp Val Asp Val Gly Pro Leu Phe Gln Thr Pro
      645                650                655
Glu Glu Val Ala Asn Gln Ala Ala Asp Asn Asp Val His Val Val Gly
      660                665                670
Val Ser Ser Leu Ala Ala Gly His Leu Thr Leu Val Pro Ala Leu Arg
      675                680                685
Glu Ala Leu Ala Ala Ala Gly Arg Pro Asp Ile Met Ile Val Val Gly
      690                695                700
Gly Val Ile Pro Pro Gly Asp Phe Asp Glu Leu Tyr Glu Ala Gly Ala
      705                710                715                720
Ala Ala Ile Phe Pro Pro Gly Thr Val Ile Ala Asp Ala Ala Ser Gly
      725                730                735
Leu Leu Glu Lys Leu Ser Ala Gln Leu Gly His Asp His Ser
      740                745                750

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<210> SEQ ID NO 69
<211> LENGTH: 618
<212> TYPE: PRT
<213> ORGANISM: Porphyromonas gingivalis
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / NP_905776
<309> DATABASE ENTRY DATE: 2010-06-29
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(618)

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<400> SEQUENCE: 69

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Met Ala Lys Glu Lys Glu Lys Leu Phe Ser Glu Phe Pro Pro Val Ser
 1                5                10                15
Arg Glu Ala Trp Ile Asp Lys Ile Thr Ala Asp Leu Lys Gly Val Pro
 20                25                30
Phe Glu Lys Lys Leu Val Trp Arg Thr Asn Glu Gly Phe Asn Val Asn
 35                40                45
Pro Phe Tyr Arg Arg Glu Asp Ile Glu Asp Leu Lys Thr Thr Thr Ser
 50                55                60

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Leu Pro Asp Glu Tyr Pro Tyr Val Arg Ser Thr Arg Met His Asn Glu
 65 70 75 80

Trp Leu Val Arg Gln Asp Ile Val Val Gly Asp Asn Val Ala Glu Ala
 85 90 95

Asn Glu Lys Ala Leu Asp Leu Leu Asn Lys Gly Val Asp Ser Leu Gly
 100 105 110

Phe Tyr Leu Lys Lys Val His Ile Asn Val Asp Thr Leu Ala Ala Leu
 115 120 125

Leu Lys Asp Ile Glu Leu Thr Ala Val Glu Leu Asn Phe Asn Cys Cys
 130 135 140

Ile Thr Arg Ala Ala Asp Leu Leu Ser Ala Phe Ser Ala Tyr Val Lys
 145 150 155 160

Lys Val Gly Ala Asp Pro Asn Lys Cys His Gly Ser Val Ser Tyr Asp
 165 170 175

Pro Phe Lys Lys Gln Leu Val Arg Gly Val Ser Asn Pro Asp Trp Val
 180 185 190

Lys Met Thr Leu Pro Val Met Asp Ala Ala Arg Glu Leu Pro Ala Phe
 195 200 205

Arg Val Leu Asn Val Asn Ala Val Asn Leu Ser Asp Ala Gly Ala Phe
 210 215 220

Ile Thr Gln Glu Leu Gly Tyr Ala Leu Ala Trp Gly Ala Glu Leu Leu
 225 230 235 240

Asp Lys Leu Thr Asp Ala Gly Tyr Lys Pro Glu Glu Ile Ala Ser Arg
 245 250 255

Ile Lys Phe Asn Phe Gly Ile Gly Ser Asn Tyr Phe Met Glu Ile Ala
 260 265 270

Lys Phe Arg Ala Ala Arg Trp Leu Trp Ala Gln Ile Val Gly Ser Tyr
 275 280 285

Gly Asp Gln Tyr Lys Asn Glu Thr Ala Lys Ile His Gln His Ala Thr
 290 295 300

Thr Ser Met Trp Asn Lys Thr Val Phe Asp Ala His Val Asn Leu Leu
 305 310 315 320

Arg Thr Gln Thr Glu Thr Met Ser Ala Ala Ile Ala Gly Val Asp Ser
 325 330 335

Ile Thr Val Leu Pro Phe Asp Val Thr Tyr Gln Gln Ser Asp Asp Phe
 340 345 350

Ser Glu Arg Ile Ala Arg Asn Gln Gln Leu Leu Lys Glu Glu Cys
 355 360 365

His Phe Asp Lys Val Ile Asp Pro Ser Ala Gly Ser Tyr Tyr Ile Glu
 370 375 380

Thr Leu Thr Asn Ser Ile Gly Glu Glu Ala Trp Lys Leu Phe Leu Ser
 385 390 395 400

Val Glu Asp Ala Gly Gly Phe Thr Gln Ala Ala Glu Thr Ala Ser Ile
 405 410 415

Gln Lys Ala Val Asn Ala Ser Asn Ile Lys Arg His Gln Ser Val Ala
 420 425 430

Thr Arg Arg Glu Ile Phe Leu Gly Thr Asn Gln Phe Pro Asn Phe Thr
 435 440 445

Glu Val Ala Gly Asp Lys Ile Thr Leu Ala Gln Gly Glu His Asp Cys
 450 455 460

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Asn Cys Val Lys Ser Ile Glu Pro Leu Asn Phe Ser Arg Gly Ala Ser
465                               470                               475                               480

Glu Phe Glu Ala Leu Arg Leu Ala Thr Glu Lys Ser Gly Lys Thr Pro
                               485                               490                               495

Val Val Phe Met Leu Thr Ile Gly Asn Leu Ala Met Arg Leu Ala Arg
                               500                               505                               510

Ser Gln Phe Ser Ser Asn Phe Phe Gly Cys Ala Gly Tyr Lys Leu Ile
                               515                               520                               525

Asp Asn Leu Gly Phe Lys Ser Val Glu Glu Gly Val Asp Ala Ala Leu
                               530                               535                               540

Ala Ala Lys Ala Asp Ile Val Val Leu Cys Ser Ser Asp Asp Glu Tyr
545                               550                               555                               560

Ala Glu Tyr Ala Pro Ala Ala Phe Asp Tyr Leu Ala Gly Arg Ala Glu
                               565                               570                               575

Phe Val Val Ala Gly Ala Pro Ala Cys Met Ala Asp Leu Glu Ala Lys
                               580                               585                               590

Gly Ile Arg Asn Tyr Val His Val Lys Ser Asn Val Leu Glu Thr Leu
                               595                               600                               605

Arg Ala Phe Asn Asp Lys Phe Gly Ile Arg
                               610                               615

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<210> SEQ ID NO 70
<211> LENGTH: 715
<212> TYPE: PRT
<213> ORGANISM: Porphyromonas gingivalis
<300> PUBLICATION INFORMATION:
<308> DATABASE ACCESSION NUMBER: NCBI / NP_905777
<309> DATABASE ENTRY DATE: 2010-06-29
<313> RELEVANT RESIDUES IN SEQ ID NO: (1)..(715)

<400> SEQUENCE: 70

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```

Met Lys Pro Asn Tyr Lys Asp Ile Asp Ile Lys Ser Ala Gly Phe Val
1                               5                               10                               15

Ala Lys Asp Ala Thr Arg Trp Ala Glu Glu Lys Gly Ile Val Ala Asp
                               20                               25                               30

Trp Arg Thr Pro Glu Gln Ile Met Val Lys Pro Leu Tyr Thr Lys Asp
                               35                               40                               45

Asp Leu Glu Gly Met Glu His Leu Asp Tyr Val Ser Gly Leu Pro Pro
                               50                               55                               60

Phe Leu Arg Gly Pro Tyr Ser Gly Met Tyr Pro Met Arg Pro Trp Thr
65                               70                               75                               80

Ile Arg Gln Tyr Ala Gly Phe Ser Thr Ala Glu Glu Ser Asn Ala Phe
                               85                               90                               95

Tyr Arg Arg Asn Leu Ala Ser Gly Gln Lys Gly Leu Ser Val Ala Phe
                               100                              105                              110

Asp Leu Ala Thr His Arg Gly Tyr Asp Ala Asp His Ser Arg Val Val
                               115                              120                              125

Gly Asp Val Gly Lys Ala Gly Val Ser Ile Cys Ser Leu Glu Asp Met
                               130                              135                              140

Lys Val Leu Phe Asp Gly Ile Pro Leu Ser Lys Met Ser Val Ser Met
145                              150                              155                              160

Thr Met Asn Gly Ala Val Leu Pro Ile Leu Ala Phe Tyr Ile Asn Ala
                               165                              170                              175

Gly Leu Glu Gln Gly Ala Lys Leu Glu Glu Met Ala Gly Thr Ile Gln

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180					185					190					
Asn	Asp	Ile	Leu	Lys	Glu	Phe	Met	Val	Arg	Asn	Thr	Tyr	Ile	Tyr	Pro
	195						200					205			
Pro	Glu	Phe	Ser	Met	Arg	Ile	Ile	Ala	Asp	Ile	Phe	Glu	Tyr	Thr	Ser
	210					215					220				
Gln	Asn	Met	Pro	Lys	Phe	Asn	Ser	Ile	Ser	Ile	Ser	Gly	Tyr	His	Met
	225					230					235				240
Gln	Glu	Ala	Gly	Ala	Thr	Ala	Asp	Ile	Glu	Met	Ala	Tyr	Thr	Leu	Ala
				245					250					255	
Asp	Gly	Met	Gln	Tyr	Leu	Lys	Ala	Gly	Ile	Asp	Ala	Gly	Ile	Asp	Val
			260					265					270		
Asp	Ala	Phe	Ala	Pro	Arg	Leu	Ser	Phe	Phe	Trp	Ala	Ile	Gly	Val	Asn
		275					280					285			
His	Phe	Met	Glu	Ile	Ala	Lys	Met	Arg	Ala	Ala	Arg	Leu	Leu	Trp	Ala
	290					295					300				
Lys	Ile	Val	Lys	Ser	Phe	Gly	Ala	Lys	Asn	Pro	Lys	Ser	Leu	Ala	Leu
	305					310					315				320
Arg	Thr	His	Ser	Gln	Thr	Ser	Gly	Trp	Ser	Leu	Thr	Glu	Gln	Asp	Pro
				325					330					335	
Phe	Asn	Asn	Val	Gly	Arg	Thr	Cys	Ile	Glu	Ala	Met	Ala	Ala	Ala	Leu
			340					345					350		
Gly	His	Thr	Gln	Ser	Leu	His	Thr	Asn	Ala	Leu	Asp	Glu	Ala	Ile	Ala
		355					360					365			
Leu	Pro	Thr	Asp	Phe	Ser	Ala	Arg	Ile	Ala	Arg	Asn	Thr	Gln	Ile	Tyr
	370					375					380				
Ile	Gln	Glu	Glu	Thr	Leu	Val	Cys	Lys	Glu	Ile	Asp	Pro	Trp	Gly	Gly
	385					390					395				400
Ser	Tyr	Tyr	Val	Glu	Ser	Leu	Thr	Asn	Glu	Leu	Val	His	Lys	Ala	Trp
				405					410					415	
Thr	Leu	Ile	Lys	Glu	Val	Gln	Glu	Met	Gly	Gly	Met	Ala	Lys	Ala	Ile
			420					425					430		
Glu	Thr	Gly	Leu	Pro	Lys	Leu	Arg	Ile	Glu	Glu	Ala	Ala	Ala	Arg	Thr
		435					440					445			
Gln	Ala	Arg	Ile	Asp	Ser	His	Gln	Gln	Val	Ile	Val	Gly	Val	Asn	Lys
	450					455					460				
Tyr	Arg	Leu	Pro	Lys	Glu	Asp	Pro	Ile	Asp	Ile	Leu	Glu	Ile	Asp	Asn
	465					470					475				480
Thr	Ala	Val	Arg	Lys	Gln	Gln	Ile	Glu	Arg	Leu	Asn	Asp	Leu	Arg	Ser
				485					490					495	
His	Arg	Asp	Glu	Lys	Ala	Val	Gln	Glu	Ala	Leu	Glu	Ala	Ile	Thr	Lys
			500					505					510		
Cys	Val	Glu	Thr	Lys	Glu	Gly	Asn	Leu	Leu	Asp	Leu	Ala	Val	Lys	Ala
		515					520					525			
Ala	Gly	Leu	Arg	Ala	Ser	Leu	Gly	Glu	Ile	Ser	Asp	Ala	Cys	Glu	Lys
	530					535					540				
Val	Val	Gly	Arg	Tyr	Lys	Ala	Val	Ile	Arg	Thr	Ile	Ser	Gly	Val	Tyr
	545					550					555				560
Ser	Ser	Glu	Ser	Gly	Glu	Asp	Lys	Asp	Phe	Ala	His	Ala	Lys	Glu	Leu
				565					570					575	
Ala	Glu	Lys	Phe	Ala	Lys	Lys	Glu	Gly	Arg	Gln	Pro	Arg	Ile	Met	Ile
			580					585					590		

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Ala	Lys	Met	Gly	Gln	Asp	Gly	His	Asp	Arg	Gly	Ala	Lys	Val	Val	Ala
		595					600					605			
Thr	Gly	Tyr	Ala	Asp	Cys	Gly	Phe	Asp	Val	Asp	Met	Gly	Pro	Leu	Phe
	610					615					620				
Gln	Thr	Pro	Glu	Glu	Ala	Ala	Arg	Gln	Ala	Val	Glu	Asn	Asp	Val	His
625					630					635				640	
Val	Met	Gly	Val	Ser	Ser	Leu	Ala	Ala	Gly	His	Lys	Thr	Leu	Ile	Pro
				645					650					655	
Gln	Val	Ile	Ala	Glu	Leu	Glu	Lys	Leu	Gly	Arg	Pro	Asp	Ile	Leu	Val
			660					665					670		
Thr	Ala	Gly	Gly	Val	Ile	Pro	Ala	Gln	Asp	Tyr	Asp	Phe	Leu	Tyr	Gln
		675					680					685			
Ala	Gly	Val	Ala	Ala	Ile	Phe	Gly	Pro	Gly	Thr	Pro	Val	Ala	Tyr	Ser
		690				695					700				
Ala	Ala	Lys	Val	Leu	Glu	Ile	Leu	Leu	Glu	Glu					
705				710						715					

What is claimed is:

1. A method for producing branched-chain fatty acid comprising a methyl on one or more even number carbons, the method comprising culturing a cell comprising

(aa) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of propionyl-CoA to methylmalonyl-CoA and/or (bb) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a polypeptide that catalyzes the conversion of succinyl-CoA to methylmalonyl-CoA,

under conditions allowing expression of the polynucleotide(s) and production of branched-chain fatty acid, wherein the cell produces more branched-chain fatty acid comprising a methyl on one or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s).

2. The method of claim 1 further comprising extracting from culture the branched-chain fatty acid or a product of the branched-chain fatty acid.

3. The method of claim 2, wherein the polypeptide that catalyzes the conversion of propionyl-CoA to methylmalonyl-CoA is a propionyl-CoA carboxylase and/or the polypeptide that catalyzes the conversion of succinyl-CoA to methylmalonyl-CoA is a methylmalonyl-CoA mutase.

4. The method of claim 3, wherein (i) the propionyl-CoA carboxylase is *Streptomyces coelicolor* PccB and AccA1 or PccB and AccA2 and/or (ii) the methylmalonyl-CoA mutase is *Janibacter* sp. HTCC2649 methylmalonyl-CoA mutase, *S. cinnamomensis* MutA and MutB, or *E. coli* Sbm.

5. The method of claim 3, wherein (i) the methylmalonyl-CoA mutase comprises an amino acid sequence having at least about 80% sequence identity to the amino acid sequence set forth in SEQ ID NOs: 3, 4, or 28 and/or (ii) the propionyl-CoA carboxylase comprises an amino acid sequence having at least about 80% sequence identity to the amino acid sequence set forth in SEQ ID NOs: 9 and 10.

6. The method of claim 3, wherein the cell comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA

mutase and further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA epimerase.

7. The method of claim 2, wherein the cell further comprises an exogenous or overexpressed polynucleotide encoding an acyl transferase lacking polyketide synthesis activity and/or an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a thioesterase.

8. The method of claim 7, wherein the acyl transferase is FabD, an acyl transferase domain of a polyketide synthase, or an acyl transferase domain of *Mycobacterium* mycocerosic acid synthase.

9. The method of claim 2, wherein the cell has been modified to attenuate endogenous methylmalonyl-CoA mutase activity, endogenous methylmalonyl-CoA decarboxylase activity, and/or endogenous acyl transferase activity.

10. The method of claim 2, wherein the cell produces a Type II fatty acid synthase.

11. The method of claim 10, wherein the cell is *Escherichia coli*.

12. A branched-chain fatty acid produced by the method of claim 1.

13. A cell comprising:

(i) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding an acyl transferase lacking polyketide synthesis activity, and

(ii) an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a propionyl-CoA carboxylase and/or an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA mutase,

wherein the polynucleotide(s) are expressed and the cell produces more branched-chain fatty acid comprising a methyl on one or more even number carbons than an otherwise similar cell that does not comprise the polynucleotide(s).

14. The cell of claim 13, wherein (i) the propionyl-CoA carboxylase is *Streptomyces coelicolor* PccB and AccA1 or PccB and AccA2 and/or (ii) the methylmalonyl-CoA mutase

is *Janibacter* sp. HTCC2649 methylmalonyl-CoA mutase, *S. cinnamomensis* MutA and MutB, or *E. coli* Sbm.

15. The cell of claim **13**, wherein (i) the methylmalonyl-CoA mutase comprises an amino acid sequence having at least about 80% sequence identity to the amino acid sequence set forth in SEQ ID NOs: 3, 4, or 28 and/or (ii) the propionyl-CoA carboxylase comprises an amino acid sequence having at least about 80% sequence identity to the amino acid sequence set forth in SEQ ID NOs: 9 and 10.

16. The cell of claim **13**, wherein the cell comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA mutase and further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a methylmalonyl-CoA epimerase.

17. The cell of claim **13**, wherein the acyl transferase is FabD, an acyl transferase domain of a polyketide synthase, or an acyl transferase domain of *Mycobacterium* mycocerosic acid synthase.

18. The cell of claim **13**, wherein the cell further comprises an exogenous or overexpressed polynucleotide comprising a nucleic acid sequence encoding a thioesterase.

19. The cell of claim **13**, wherein the cell has been modified to attenuate endogenous methylmalonyl-CoA mutase activity, endogenous methylmalonyl-CoA decarboxylase activity, and/or endogenous acyl transferase activity.

20. The cell of claim **13**, wherein the cell is *Escherichia coli*.

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