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PATENTS ACT 1990
NOTICE OF ENTITLEMENT

We, **Beecham Group p.l.c.**, the applicant/Nominated Person in respect of
Application No. 73483/91 state the following:-

The Nominated Person is entitled to the grant of the patent
because the Nominated Person would, on the grant of a patent for
the invention to the inventor, be entitled to have the patent
assigned to the Nominated Person.

The Nominated Person is entitled to claim priority from the
application listed in the declaration under Article 8 of the PCT
because the Nominated Person made the application listed in the
declaration under Article 8 of the PCT, and because that
application was the first application made in a Convention country
in respect of the invention.

DATED this FOURTEENTH day of AUGUST 1992



.....
a member of the firm of
DAVIES COLLISON
CAVE for and on behalf
of the applicant(s)

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(57) Claim

1. A composition for controlled and sustained release of a pharmaceutically acceptable drug comprising a drug-resin complex formed from an ion-exchange resin and a pharmacologically active drug, characterised in that at least 20% by weight of the resinate particles have a particle size below 35 μ m.

13. A process for preparing a composition as defined in any one of claims 1 to 12 comprising the admixture of ion-exchange resin, the pharmacologically active drug and a suitable solvent.

15. A method of expediting and sustaining therapeutic levels of a pharmaceutically acceptable drug in humans or animals following a single dose, which method comprises administering to a patient an effective single dose of a composition as defined in any one of claims 1 to 12.

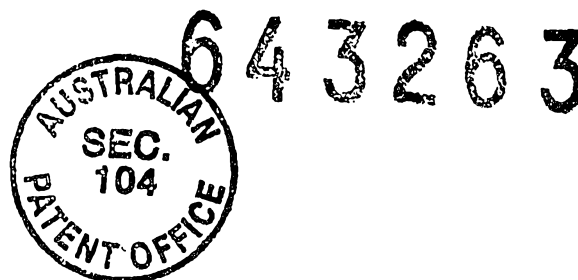
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(21) International Application Number: PCT/GB91/00324 (22) International Filing Date: 4 March 1991 (04.03.91) (30) Priority data: 9005498.2 12 March 1990 (12.03.90) GB (71) Applicant (for all designated States except US): BEECHAM GROUP P.L.C. [GB/GB]; SB House, Great West Road, Brentford, Middlesex TW8 9BD (GB). (72) Inventor; and (75) Inventor/Applicant (for US only) : GRATTAN, Timothy, James [GB/GB]; SmithKline Beecham Consumer Brands, St. George's Avenue, Weybridge, Surrey KT13 0DE (GB). (71) ADDRESS - FOUR NEW HORIZONS COURT HARLEQUIN AVENUE BRENTFORD, MIDDLESEX TW8 9EP UNITED KINGDOM		(74) Agents: LOCKWOOD, Barbara, Ann; SmithKline Beecham, Corporate Patents, Great Burgh, Yew Tree Bottom Road, Epsom, Surrey KT18 5XQ (GB) et al. (81) Designated States: AT (European patent), AU, BE (European patent), CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, LU (European patent), NL (European patent), SE (European patent), US. Published With international search report.
(54) Title: COMPOSITION (57) Abstract A composition for controlled and sustained release of a pharmaceutically acceptable drug comprising a drug-resin complex formed from an ion-exchange resin and a pharmacologically active drug, characterised in that at least 20 % by weight of the resinate particles have a particle size below 35µm.		



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COMPOSITION

The present invention relates to controlled, sustained release compositions for pharmaceutical use and in particular to compositions containing an ion-exchange resin having a pharmacologically active drug adsorbed thereon.

Ion-exchange is known to have utility in providing improved drug delivery systems for orally administered drug products.

10

US Patent 2,900,332 (Keating) discloses drug-resin complexes prepared by interaction of cationic ion-exchange resins with basic drugs in their cationic form, such as amphetamine and codeine. When a drug-resin complex of this type is orally administered, drug release is initiated when the acidic environment of the gastro-intestinal tract is encountered.

Drug-resin complexes thus offered a method for retarding the rate of drug release not available using conventional drug delivery systems. The degree of control of drug release from drug-resin complexes was however found to be generally unsatisfactory since for many drugs only a relatively short delay in drug release was obtained.

Coating of drug-resin particles was therefore undertaken so as to provide a further diffusion barrier to the drug and hence offer greater control over the drug release profile. For example, US Patent 4,221,778 (Raghunathan) discloses a range of drug-resin complexes coated with a water-permeable diffusion barrier coating.

Uncoated complexes formed with the cationic, cross-linked sulphonated polystyrene ion-exchange resins Amberlite XE-69 and Amberlite EX-120, prepared for the purposes of

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comparison, are described as having faster dissolution profiles than are desirable for sustained release of drug.

US Patent 4,788,055 (Fischer et al.) describes uncoated
5 complexes prepared from a cross-linked sulphonated polystyrene ion-exchange resin (Amberlite IRP-69) and an acid addition salt of dextromethorphan, conferring a drug-release profile suitable for once in 12 hour dosing, sustained release being achieved by the selection of a drug
10 to resin ratio within the range 1:5 to 1:10.

The above-mentioned publications present results which demonstrate that the rate of release of drug from ion-exchange complexes is influenced by the resinate
15 particle size, with small resinate particles releasing drug more rapidly than large resinate particles.

US 4,788,055 (Fischer et al.) presents results which demonstrate that small resinate particles are considered
20 unsuitable for use in drug-resin complexes where drug release over a time period in excess of 8 hours is desired. A preference is expressed for average particle size in the range 40-80µm and more preferably in the range 50-70µm.

25 Surprisingly, it has now been found that, contrary to the teaching of the prior art, controlled and sustained drug release from drug-resin complexes can be achieved using small resinate particles. Moreover, the use of small resinate particles has been found to confer advantages of
30 practical benefit.

Accordingly, the present invention provides a composition suitable for controlled, sustained release of a pharmaceutically acceptable drug comprising a drug-resin
35 complex formed from an ion-exchange resin and a pharmacologically active drug, characterised in that at least 20% by weight of the resinate particles have a

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particle size below 35 μ m.

Suitably, a composition of the invention has at least 30% by weight and preferably at least 40% by weight of resinate particles having a particle size below 35 μ m.

A preferred composition according to the invention is further characterised in that at least 50% by weight and preferably at least 60% by weight of resinate particles have a particle size below 50 μ m.

In vivo tests have shown that targetted blood plasma-time profiles may not be achievable from compositions in which resinate particles greater than 50 μ m predominate in view of insufficient initial drug release, attributable to the retardation of the rate of release per unit time with increasing particle size.

The present invention overcomes this potential problem. Small resinate particles have been found to confer on a drug-resin complex an increased initial release of drug per unit time which enables therapeutic drug levels to be achieved more rapidly.

Preparations containing suspensions of drug-resin complexes wherein the resinate particle size is greater than about 120 μ m are known to impart a gritty texture which renders the product unpalatable for oral consumption, (Irwin et al., Drug Dev. and Ind. Pharm, 13(9-11), 2064, (1987). Panel tests have now shown that particle sizes greater than 50 μ m also impart an unpalatable gritty texture to the product. Compositions according to the present invention, in liquid form, also therefore have the advantage over known drug-resin complexes in that they provide a product for oral administration with improved mouth feel.

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Furthermore, compositions according to the present invention may confer the additional advantage of enhancing the in vivo bioavailability and/or the duration of action of the drug.

5 Firstly, release of drug from resinate particles is a diffusion-controlled equilibrium process, and small resin particles offer a more complete release of drug from the resin.

10 Secondly, the villous structure of the mucosal wall of the gastro-intestinal tract presents a large available surface area. Resinate particles greater than for example 50µm may be too large to enter the interstices between individual villi, such that a large part of the surface area of the
15 mucosal surface may not be available for drug adsorption. It is however possible that small resinate particles may enter between individual villi and hence take a more tortuous path down the gastro-intestinal tract, offering greater availability of the dosage form with the associated
20 potential for a longer duration of action.

In general, all ionisable, orally administrable pharmacologically active drugs that are absorbed under conditions encountered in the gastro-intestinal tract are
25 suitable for use in compositions according to the present invention. Particularly suitable drugs are, for example, those having basic character, including for example phenylpropanolamine, chlorpheniramine, dextromethorphan, noscapine and pseudoephedrine.

30

Similarly, a wide range of ion-exchange resins is potentially available to form drug-resin complexes. Cationic ion-exchange resins are suitable for use with basic drugs and anionic ion-exchange resins are suitable for use
35 with drug compounds having acidic character. The complex resulting from a cationic ion-exchange resin or an anionic ion-exchange resin will release drug on encountering the

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ionic environment of the gastro-intestinal tract.

Suitable ion-exchange resins, which it will be appreciated will be pharmaceutically acceptable, are commercially
5 available and include a range of synthetic ion-exchange resins with different polymeric matrices which may be cross-linked.

Ion-exchange resins particularly suitable for use with basic
10 drugs include strongly cationic resins having sulphonic acid groups on a polystyrene polymer matrix, which resins are suitably cross-linked, preferably by a divinyl or polyvinyl compound such as divinylbenzene. The proportion of cross-linking may vary from 1 to 20%, suitably from 1 to
15 12%, for example from 5 to 10%, and preferably from 7 to 9% by weight cross-linking.

Suitable resins are those commercially available under the trade names Amberlite and Duolite from Rohm and Haas Co. and
20 Dowex from the Dow Chemical Company. A preferred resin is Amberlite IRP-69.

A basic drug may be presented for complex formation in the form of a pharmaceutically acceptable acid addition salt, in
25 which case the chosen resin will be in its 'sodium' form. Alternatively, the drug may be presented as the free base, in which case the resin will be in its 'hydrogen' form.

Acidic drugs may be presented for complex formation with a
30 suitable anionic ion-exchange resin either as the free acid or in salt form. Suitable salts include inorganic salts for example sodium, potassium, calcium or magnesium salts, or organic salts, for example salts derived from amino acid residues or choline.

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It will be appreciated that the invention extends to compositions comprising two or more drugs and two or more ion-exchange resins.

5 Commercially available ion-exchange resins generally have a particle size distribution in which the average particle size is greater than that required by the present invention, for example Amberlite IRP-69. The size of the ion-exchange particles may therefore be reduced as necessary to achieve
10 the desired particle size distribution. Size reduction may be carried out using standard milling methods. Suitable milling methods include the use of micronisers, commutating mills, pin mills, air-jet mills for example fluid energy mills, and wet milling and ultrasonic techniques.

15

It will be appreciated that the spread of particle sizes generally present in commercially available ion-exchange resins may allow ion-exchange resin suitable for use in compositions of the present invention to be obtained by
20 sieving. Preferably, ion-exchange resin for use in compositions of the invention is obtained using a combination of size reduction and sieving techniques.

Size reduction and/or sieving may be carried out either
25 before or after loading the ion-exchange resin with drug.

The release profile of drug from a drug-resin complex is influenced by the physico-chemical properties of the drug. Thus the particle size distribution of resinate particles in
30 compositions of the invention may be selected to match the properties of a particular drug, the ion-exchange resin on which it is loaded, and the desired drug-release profile.

Compositions of the invention suitably have a particle size
35 distribution comprising from about 20 to 75% by weight of

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particles below 35 μ m and from about 50 to 100% by weight of particles below 50 μ m, preferably from about 30 to 50% by weight below 35 μ m and from about 60 to 100% by weight below 50 μ m.

5

It will be appreciated that where a drug inherently gives rise to a more rapid release than is desirable for effective sustained release, a proportion (% by weight) of resinate particles of larger size, for example up to 130 μ m, may be included to sustain release of drug over the desired time period.

The desired particle size distribution may be achieved by a combination of standard milling and sieving techniques.

15 Where size reduction is carried out before loading the ion-exchange resin with drug, further screening by sieving to achieve a desired composition may also be carried out before drug loading. Alternatively, further screening may be carried out after preparation of the drug-resin complex.
20 This will of course be the procedure adopted where particle size reduction is carried out after loading the drug onto the ion-exchange resin.

Pre-conditioning of ion-exchange resins prior to complex
25 formation may be desirable in order to remove any potentially toxic extractibles and/or to facilitate easier processing and analysis of the resulting resinates.

Adsorption of drug onto ion-exchange resins to form a
30 drug-resin complex may be carried out by standard procedures well known in the art, for example as described in US Patent 2,990,332 (Keating).

Thus, for example, a commercially available ion-exchange
35 resin may be added directly to a solution of drug in an appropriate solvent. The resulting drug-resin complex may

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then be dried and the resinate particles reduced by milling, and/or screened by sieving into particle size ranges, to achieve the desired particle size distribution.

5 Alternatively, the drug-resin complex formed initially as described above may be wet-milled and then dried and, as necessary, screened by sieving into particle size ranges.

In a further process for the preparation of drug-resin
10 complexes of the invention, the ion-exchange resin particles are first reduced by milling and/or screened by sieving. The ion-exchange resin having the required particle size distribution is then added to a solution of drug in an appropriate solvent, followed by drying.

15

Compositions of the invention are suitably administered as liquid suspensions.

The invention further provides a novel process specifically
20 adapted for the preparation of a composition of the invention as a liquid suspension, which process comprises the addition of ion-exchange resin (having the desired particle size distribution wherein at least 20% by weight of the resin particles have a particle size below 35 μ m) to a
25 mixture of a pharmacologically active drug and a pharmaceutically acceptable liquid carrier.

The mixture of drug and liquid carrier may be in the form of a solution or, where the drug is insoluble in the carrier,
30 in the form of a suspension. It will be appreciated that in situ loading of the drug onto the ion-exchange resin in a liquid carrier is advantageous in that it obviates the need for a separate drying stage.

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Compositions according to the present invention will contain drug and ion-exchange resin in a ratio suitable for achieving release of drug over the desired time period. For example, to achieve drug release for up to 12 hours from dosing, it will generally be suitable to use a drug to resin ratio in the range 1:2 to 1:12 by weight, (calculated on a moisture-free basis), the exact ratio being selected to suit the chosen drug.

10 The release of drug from drug-resin complexes approximates to first-order kinetics. Thus, release rates decrease as the concentration of the drug in the complex decreases. A low drug loading, for example a drug to resin ratio up to 1:30, may therefore be selected to achieve a more prolonged duration of action. However, in order to reduce the cost of materials and the dosage volume of drug-resin complex administered, a drug to resin ratio of 1:2 to 1:10 is generally preferred.

20 Compositions according to the present invention may be formulated for oral administration in liquid form, for example as a syrup, or alternatively in solid form, for example as a capsule or tablet. Compositions may therefore be formulated in admixture with an appropriate pharmaceutically acceptable vehicle, additionally containing, as desired, pharmaceutically acceptable adjuvants including inter alia thickeners, preservatives, and colouring and flavouring agents.

30 Accordingly, the present invention provides a pharmaceutical composition comprising a drug-resin complex formed from a pharmaceutically acceptable drug and a pharmaceutically acceptable ion-exchange resin wherein at least 20% by weight of the resinate particles have a particle size below 35µm, in admixture with a pharmaceutically acceptable carrier.

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The quantity of resinate in a formulation may be adjusted to deliver an effective dose of a selected drug to a patient in need of treatment. Compositions of the invention are
5 suitably formulated for administration in unit dose form.

Thus, the amount of resinate in a composition for oral administration, for example in the form of a capsule or tablet, may be calculated accordingly. As described above, compositions of the invention are particularly suitable for
10 oral administration in liquid form. Liquid compositions may similarly be prepared in unit dose form by presentation for consumption in single-dose liquid-dosage units.

The invention also provides a method of expediting and
15 sustaining therapeutic levels of a pharmaceutically acceptable drug in humans or animals following a single dose, which method comprises administering an effective single dose of a composition comprising a drug-resin complex formed from an ion-exchange resin and a pharmacologically
20 active drug, whereby at least 20% by weight of the resinate particles have a particle size below 35 μ m.

~~In another aspect, the invention provides the use of a composition comprising a drug-resin complex formed from an
25 ion-exchange resin and a pharmacologically active drug for the manufacture of a medicament for use in expediting and sustaining therapeutic drug levels after a single dose, whereby at least 20% by weight of the resinate particles have a particle size below 35 μ m.~~

30

The Examples which follow illustrate but do not limit the scope of the present invention.

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Example 1Preparation of small particle ion-exchange resin

5 Samples of ion-exchange resin having particle sizes below
35 μ m and 50 μ m were prepared from Amberlite IRP-69
ion-exchange resin (from Rohm and Haas), a strongly acidic
(sodium form), 100-500 wet mesh resin. All samples of
Amberlite IRP-69 were conditioned before use according to
10 the following procedure.

Conditioning Procedure: Amberlite IRP-69 was added to an
aqueous solution of 2M sodium hydroxide and stirred for one
hour. The conditioned resin was isolated by Buchner
15 filtration, washed with successive aliquots of distilled
water, and then dried to constant weight at 60°C.
[Residual water content = 4.76%].

Resin Preparation

20

1(a) Amberlite IRP-69 (100g) was sieved through a 50 μ
screen. 42g of resin particles below 50 μ m were collected.

1(b) Resin obtained from Example 1(a) (20g) was sieved
25 through a 35 μ m screen. 9.2g of resin particles below 35 μ m
were collected.

1(c) Amberlite IRP-69 (2kg) was processed using a fluid
energy mill to give 1.6kg of milled material.

30

1(d) Resin obtained from Example 1(c) (500g) was sieved
through a 50 μ m screen. 455g of resin particles below 50 μ m
were collected.

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1(e) Resin obtained from Example 1(d) (20g) was sieved through a 35 μ m screen. 9.2g of resin particles below 35 μ m were collected.

5 1(f) Amberlite IRP-69 (20kg) was processed using a fluid energy mill to give 18kg of milled material.

1(g) Resin obtained from Example 1(f), 10kg, was sieved through a 63 μ m screen. 9.2kg of resin particles below 63 μ m
10 were collected.

1(h) Resin obtained from Example 1(g), 20g was sieved through a 35 μ m screen. 7.5g of resin particles below 35 μ m were collected.

15

Example 2

Preparation of dextromethorphan-Amberlite IRP-69 resinates

20 2(a) Dextromethorphan hydrobromide (0.5kg) was dissolved in water (15 litres) at 35°C. Amberlite IRP-69, prepared according to the method of Example 1(d), (2kg; equivalent to 1.9kg of moisture-free resin) was added to the solution. After stirring the mixture for 2 hours, the resinate was
25 isolated by Buchner filtration and washed with successive aliquots of distilled water. The resinate was dried at 50°C under vacuum to give 2.35kg of dextromethorphan-Amberlite IRP-69 resinate particles below 50 μ m.

30 2(b) The above procedure was repeated using Amberlite IRP-69 (2kg; equivalent to 1.9kg of moisture-free resin) as received from the supplier. The resulting dried resinate was sieved through a 50 μ m screen to give 0.9kg of dextromethorphan- Amberlite IRP-69 resinate particles below
35 50 μ m. Resinate particles above 50 μ m were retained for dissolution analysis.

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2(c) Dextromethorphan-Amberlite IRP-69 resinate particles below 50 μ m, prepared according to Example 2(b), were sieved through a 35 μ m screen.

5 2(d) Dextromethorphan-Amberlite IRP-69 resinate particles prepared from dextromethorphan HBr (0.5kg) and ion-exchange resin as received from the supplier (2kg; equivalent to 1.9kg of moisture-free resin) were processed using a fluid energy mill to give 0.8kg of milled material. The isolated
10 material was subsequently sieved through a 50 μ m screen to give 0.74kg of resinate particles below 50 μ m.

2(e) Dextromethorphan hydrobromide (50g) was dissolved in water (1.5 litres) at 35°C. Amberlite IRP-69, prepared
15 according to the method of Example 1(g), (210g; equivalent to 200g of moisture-free resin) was added to the solution. After stirring the mixture for 1 hour, the resinate was isolated by Buchner filtration and washed with successive aliquots of distilled water. The resinate was dried at 50°C
20 under vacuum to give 230g of dextromethorphan - Amberlite IRP-69 resinate particles below 63 μ m.

Example 3

25 Preparation of pharmaceutical syrups containing dextromethorphan-Amberlite IRP-69 resinate

3(a) A suitable syrup base was prepared from sorbitol, keltrol, sodium benzoate, propylene glycol, sodium citrate,
30 citric acid, flavour, colour and water.

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3(b) 900ml of syrup base obtained from Example 3(a) was mixed with 19g of the resinate from example 2(d) and sufficient water to give 1 litre of syrup containing the equivalent to 40mg dextromethorphan HBr per 10ml.

5

3(c) Dextromethorphan hydrobromide (4g) was dissolved in 900ml of the syrup obtained from Example 3(a). The resulting solution was mixed with 16g of resin obtained from Example 1(d) (equivalent to 15.24g of moisture-free resin) and sufficient water to give 1 litre of syrup containing the equivalent to 40mg dextromethorphan HBr per 10ml.

3(d) Dextromethorphan hydrobromide (4g) was dissolved in 900ml of the syrup obtained from Example 3(a). The resulting syrup was mixed with 16.8g of resin obtained from Example 1(g) (equivalent to 16.0g of moisture-free resin) and sufficient water to give 1 litre of syrup containing the equivalent to 40mg dextromethorphan HBr per 10ml.

3(e) Dextromethorphan hydrobromide (3g) was dissolved in 900ml of the syrup obtained from 3(a). The resulting solution was mixed with 12.6g of resin obtained from Example 1(g) (equivalent to 12.0g of moisture-free resin) and sufficient water to give 1 litre of syrup containing the equivalent to 30mg dextromethorphan HBr per 10ml.

3(f) Dextromethorphan hydrobromide (4g) and chlorpheniramine maleate (0.80g) were dissolved in 900ml of the syrup obtained from 3(a). The resulting solution was mixed with 16.0g of resin obtained from Example 1(d) (equivalent to 15.2g of moisture-free resin) and sufficient water to give 1 litre of syrup containing the equivalent to 40mg dextromethorphan HBr per 10ml and 8mg chlorpheniramine maleate per 10ml.

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Example 4In-vitro dissolution profile of dextromethorphan-Amberlite
IRP-69 resinate particles

5

Dextromethorphan-Amberlite IRP-69 resinate particles were prepared as described in Example 2. The in vitro dissolution profile of four samples, each having a drug to resin ratio of 1:3.8 (calculated on a moisture-free basis) and a different particle size distribution, was evaluated by flow-through analysis using a Langenbacher Flow-Through Cell.

Samples of resinate (190mg; equivalent to 40mg of dextromethorphan HBr) were eluted with 0.15M sodium chloride at 4ml/min. The percentage drug release at intervals over an 8 hour time period was determined by UV analysis.

Sample A: particle size distribution as received from supplier.

Sample B: particle size >50 μ m.

Sample C: particle size <50 μ m; (at least 20% of particles below 35 μ m).

Sample D: particle size <35 μ m.

25

Results % Drug Release

Sample	1hr	2hr	4hr	6hr	8hr
A	39	58	75	84	91
B	29	47	66	74	78
C	49	69	84	91	96
D	54	76	88	92	97

30

35

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The results show that for Samples A and B, less than 40% of the dose is released within the first hour. In contrast samples C and D, comprising small resinate particles, have released approximately 50% and 55% respectively of their drug load within 1 hour, thus demonstrating the enhanced initial availability of the drug.

Example 5

10 Preparation of noscapine-Amberlite IRP-69 resinate

(Drug to resin ratio 1:3.8, calculated on a moisture-free basis using Amberlite IRP-69 with a 4.76% residual moisture content)

15 5(a) Noscapine HCl (5g) was dissolved in water (200ml) at 25°C. Amberlite IRP-69 as received from the supplier (20; equivalent to 19g of moisture-free resin) was added to the solution. After stirring for 2 hours the resinate was isolated by Buchner filtration and washed with successive
20 aliquots of distilled water. The resinate was dried at 50°C under vacuum to give 23g of noscapine-Amberlite IRP-69 resinate particles.

5(b) Noscapine-Amberlite IRP-69 resinate particles obtained
25 from Example 5(a) (10g) were sieved through a 50µm screen. 4.0g of resinate particles below 50µm were collected.

Example 6

30 In-Vitro dissolution profile of noscapine-Amberlite IRP-69 resinate

Noscapine-Amberlite IRP-69 resinate particles were prepared as described in Examples 5(a) and 5(b). The in vitro
35 dissolution profile of the two samples, each having a different particle size distribution, was evaluated by flow through analysis using a Langenbacher Flow Through Cell.

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Samples of resinate (190mg; equivalent to 40mg noscapine hydrochloride) were eluted using the method described in Example 4.

5 Sample A: Particle size distribution as
received from supplier

Sample B: Particle size <50 μ m; (at least 20% of
particles below 35 μ m).

10 Results% Drug Release

	Sample	1hr	2hr	4hr	6hr
	A	44	63	80	87
15	B	55	75	87	90

Example 7

20 Preparation of phenylpropanolamine-Amberlite IRP-69 resinate
(Drug to resin ratio (1:28.5), calculated on a moisture-free
basis using Amberlite IRP-69 with 4.76% residual moisture
content)

25 7(a) Phenylpropanolamine HCl (3g) was dissolved in water
(200ml) at 25°C. Amberlite IRP-69, as received from the
supplier, (90g; equivalent to 85.5g of moisture-free resin)
was added to the solution. After stirring for 2 hours the
resinate was isolated by Buchner filtration and washed with
30 successive aliquots of purified water. The resinate was
dried at 50°C under vacuum to give 91g of
phenylpropanolamine-Amberlite IRP-69 resinate particles.

7(b) Phenylpropanolamine-Amberlite IRP-69 resinate
35 particles obtained from Example 7(a) (10g) were sieved
through a 50 μ m screen. 4.0g of resinate particles below
50 μ m were collected.

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7(c) The procedure described in example 7(a) was repeated using milled and sieved Amberlite IRP-69 resin (Example 1d) instead of the standard grade Amberlite IRP-69. 91g of phenylpropanolamine-Amberlite IRP-69 resinate particles 5 under 50 μ m were obtained.

Example 8

In-Vitro dissolution profile of 10 phenylpropanolamine-Amberlite IRP-69 resinsates

Phenylpropanolamine-Amberlite IRP-69 resinate particles were prepared as described in Example 7.

15 Samples of resinate (1540mg; equivalent to 50mg phenylpropanolamine hydrochloride) were eluted using the method described in Example 4.

Sample A : particle size distribution as received from
20 supplier.

Sample B : particle size > 50 μ m.

Sample C : particle size < 50 μ m; (at least 20%
of particles below 35 μ m).

25 Results

% Drug Release

30	Sample	1hr	2hr	4hr	6hr
	A	44	59	79	88
	B	29	44	67	76
	C	47	66	92	97

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Example 9Preparation of pseudoephedrine-Amberlite IRP-69 resinate particles

5 (Drug to resin ratio (1:14.2 calculated on a moisture-free basis using Amberlite IRP-69 with 4.76% residual moisture content)

9(a) Pseudoephedrine hydrochloride (3g) was dissolved in
10 water (200ml) at 25°C. Amberlite IRP-69 as received from supplier (45g; equivalent to 42.8g of moisture-free resin) was added to the solution. After stirring for 2 hours the resinate was isolated by Buchner filtration and washed with successive aliquots of purified water. The resinate was
15 dried at 50°C under vacuum to give 47g of pseudoephedrine-Amberlite IRP-69 resinate particles.

9(b) Pseudoephedrine-Amberlite IRP-69 resinate particles obtained from Example 9(a) (10g) were sieved through a 50µm
20 screen. 4.1g of resinate particles below 50µm were collected.

9(c) The procedure given in Example 9(a) was repeated using milled Amberlite IRP-69 (Example 1d) in place of the
25 standard material. 47g of pseudoephedrine-Amberlite IRP-69 resinate particles below 50µm were collected.

Example 10

30 In-Vitro dissolution profile of pseudoephedrine-Amberlite IRP-69 resinates.

Pseudoephedrine-Amberlite IRP-69 resinate particles were prepared as described in Example 9.

-20-

Samples of resinate (1910mg; equivalent to 120mg psuedoephedrine hydrochloride) were eluted using the method described in Example 4.

5 Sample A : particle size distribution as received from supplier.

Sample B : particle size > 50 μ .

Sample C : particle size < 50 μ ; (at least 20% particles below 35 μ).

10

Results% Drug Release

Sample	% Drug Release			
	1hr	2hr	4hr	6hr
A	42	63	83	96
B	37	59	82	92
C	54	76	91	95

15

20 Example 11In-Vitro dissolution profile of dextromethorphan-Amberlite
IRP-69 resinate particles

25 Dextromethorphan-Amberlite IRP-69 resinate particles were prepared as described in Example 2(e). The in-vito dissolution profile of two samples, each having a different drug-load, was evaluated by flow-through analysis using a Langenbacher Flow-Through Cell.

30

The samples of resinate were eluted using the method of Example 4.

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Sample A : 148mg; equivalent to 30mg of
dextromethorphan HBr.

Particle size < 63 μ m (at least 20% of
particles below 35 μ m

5 Sample B : 197mg; equivalent to 40mg of
dextromethorphan HBr.

Particle size < 63 μ m (at least 20% of
particles below 35 μ m.

10 Results

% Drug Release

Sample	1hr	2hr	4hr	6hr
A	48	74	85	93
B	45	64	80	88

Example 12

20 In-Vitro dissolution profile of dextromethorphan-
chlorpheniramine-Amberlite IRP-69 resinate particles

A sample of dextromethorphan-Amberlite IRP-69 particles as
prepared in Example 2(b) (190mg; equivalent to 40mg of
25 dextromethorphan HBr) with a particle size below 50 μ m (at
least 20% by weight below 35 μ m) was mixed with
chlorpheniramine maleate powder (8mg) and water (5ml). The
mixture was allowed to stand for 12 hours, prior to
dissolution by the method described in Example 4, in order
30 to effect the loading of the chlorpheniramine onto the
resinate.

-22-

Results% Drug Release

5	Drug	1 hr	2 hr	4 hr
	Dextromethorphan HBr	52	67	78
	Chlorpheniramine maleate	54	68	81

10 Example 13Preparation of dextromethorphan-Amberlite XE-625 resinate

Amberlite XE-625 (250g) was milled using an Apex commutating
15 mill. The resulting powder was sieved through a 50µm
screen. 70g of resin particles below 50µm were collected.
The 50µm sieve-cut was shown by further sieve analysis to
have 32% by weight of particles below 35µm.

20 A 10g sample of the 50µm sieve-cut was conditioned with
sodium hydroxide solution as described in Example 1. The
resulting resin (4g; equivalent to 3.8g moisture-free resin)
was treated with dextromethorphan HBr (1g) in water (100ml)
according to the method described in Example 2(a).

25

Example 14In-Vitro dissolution profile of dextromethorphan-
Amberlite XE-625 resinate particles

30

A sample of the resinate prepared in Example 13 (190mg;
equivalent to 40mg of dextromethorphan HBr) was eluted
using the method of Example 4.

-23-

Results% Drug Release

	1 hr	2 hr	4 hr	6 hr
5	42	59	73	82

Example 15

10

Particle size distribution (by weight) of Ion-Exchange Resinates by Sieve Analysis

	Particle Size Range (μm)	Example No.	
		2(a)	2(e)
	Passed through 63 μm and retained by 40 μm .	42	50
20	Passed through 40 μm and retained by 35 μm .	11	7
	Passed through 35 μm .	47	43

25

Claims

1. A composition for controlled and sustained release of a pharmaceutically acceptable drug comprising a drug-resin
5 complex formed from an ion-exchange resin and a pharmacologically active drug, characterised in that at least 20% by weight of the resinate particles have a particle size below 35 μ m.
- 10 2. A composition as claimed in claim 1 wherein at least 40% by weight of resinate particles have a particle size below 35 μ m.
3. A composition as claimed in claim 1 or 2 wherein at
15 least 50% by weight of resinate particles have a particle size below 50 μ m.
4. A composition as claimed in any one of claims 1 to 3 wherein the pharmacologically active drug has basic
20 character.
5. A composition as claimed in claim 4 wherein the pharmacologically active drug is dextromethorphan, phenylpropanolamine, noscapine, pseudoephedrine or
25 chlorpheniramine, or a pharmaceutically acceptable salt thereof.
6. A composition as claimed in claim 3 or 4 wherein the ion-exchange resin is a strongly cationic resin having
30 sulphonic acid groups on a polystyrene polymer matrix.
7. A composition as claimed in claim 6 wherein the cationic resin is cross-linked with from 1 to 20% by weight of a divinyl or polyvinyl compound.
35
8. A composition as claimed in claim 7 wherein the cationic resin is cross-linked with from 7-9% by weight of

divinylbenzene.

9. A composition as claimed in claim 8 wherein the ratio of drug to resin is in the range 1:2 to 1:12 by weight, 5 calculated on a moisture-free basis.

10. A composition as claimed in claim 9 comprising a pharmacologically active drug which is dextromethorphan or a pharmaceutically acceptable salt thereof, wherein the ratio 10 of drug to resin is in the range 1:3 to 1:5 by weight, calculated on a moisture-free basis, and the particle size distribution comprises from 30 to 50% by weight of particles below 35 μ m and from 60 to 100% by weight of particles below 50 μ m.

15

11. A composition as claimed in any one of claims 1 to 10 further comprising a pharmaceutically acceptable carrier.

12. A composition as claimed in claim 11 for oral 20 administration in liquid form.

13. A process for preparing a composition as defined in any one of claims 1 to 12 comprising the admixture of ion-exchange resin, the pharmacologically active drug and a 25 suitable solvent.

14. A process for preparing a composition as defined in claim 12 comprising adding ion-exchange resin as defined in any one of claims 1 to 10 to a mixture of a 30 pharmacologically active drug as defined in any one of claims 1 to 10 and a pharmaceutically acceptable liquid carrier.

15. A method of expediting and sustaining therapeutic 35 levels of a pharmaceutically acceptable drug in humans or animals following a single dose, which method comprises

administering to a patient an effective single dose of a composition as defined in any one of claims 1 to 12.

16

~~17~~. Compositions substantially as hereinbefore described with reference to the accompanying examples.

17

~~18~~. Processes substantially as hereinbefore described with reference to the accompanying examples.

10

15 DATED this 7th day of September, 1993
Beecham Group p.l.c.
By Its Patent Attorneys
DAVIES COLLISON CAVE

INTERNATIONAL SEARCH REPORT

International Application No

PCT/GB 91/00324

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all) ⁶		
According to International Patent Classification (IPC) or to both National Classification and IPC		
IPC ⁵ : A 61 K 9/18		
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁷		
Classification System	Classification Symbols	
IPC ⁵	A 61 K	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁸		
III. DOCUMENTS CONSIDERED TO BE RELEVANT ⁹		
Category ⁹	Citation of Document, ¹¹ with Indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
Y	EP, A, 0225615 (CIBA-GEIGY) 16 June 1987 see claims cited in the application ---	1-14,16-19
Y	EP, A, 0294103 (TAKEDA) 7 December 1988 see claims 1-4,8,10-13; page 2, lines 44-55; page 3, lines 12-16; page 4, lines 33-36; page 12, example 9 ---	1-14,16-19
Y	US, A, 2990332 (J.W. KEATING) 27 June 1961 see claims; column 4, lines 4-25; column 13, table VIII; column 36, lines 52-61 cited in the application ---	1-14,16-19
	./.	
⁹ Special categories of cited documents: ¹⁰ "A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention "X" document of particular relevance: the claimed invention cannot be considered novel or cannot be considered to involve an inventive step "Y" document of particular relevance: the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art. "&" document member of the same patent family		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report	
27th May 1991	11.07.91	
International Searching Authority	Signature of Authorized Officer	
EUROPEAN PATENT OFFICE	Natalie Weinberg	

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

Y	US, A, 4221778 (Y. RAGHUNATHAN) 9 September 1980 see claims 1,4-8; column 2, lines 3-15; column 3, lines 30-38; column 15, lines 4-20 cited in the application ---	1-14,16-19
P,Y	EP, A, 0367746 (RICHARDSON VICKS) 9 May 1990 see claims; page 4, lines 32-34; page 5, lines 32-35,42-46 -----	1-14,16-19

V. ☒ OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE ¹

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. ☒ Claim numbers ...15... because they relate to subject matter not required to be searched by this Authority, namely:

See PCT Rule 39.1(iv)
Methods for treatment of the human or animal body by means
of surgery or therapy, as well as diagnostic methods.

2. ☐ Claim numbers because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:

3. ☐ Claim numbers because they are dependent claims and are not drafted in accordance with the second and third sentences of PCT Rule 6.4(a).

VI. ☐ OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING ²

This International Searching Authority found multiple inventions in this international application as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.
2. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:
3. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:
4. ☐ As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remark on Protest

- ☐ The additional search fees were accompanied by applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

**ANNEX TO THE INTERNATIONAL SEARCH REPORT
ON INTERNATIONAL PATENT APPLICATION NO.**

GB 9100324

SA 45122

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on 28/06/91
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information.

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP-A- 0225615	16-06-87	US-A- 4788055	29-11-88
		AU-B- 597316	31-05-90
		AU-A- 6618186	11-06-87
		JP-A- 62138428	22-06-87

EP-A- 0294103	07-12-88	JP-A- 1079111	24-03-89
		US-A- 4894239	16-01-90

US-A- 2990332		None	

US-A- 4221778	09-09-80	None	

EP-A- 0367746	09-05-90	US-A- 4996047	26-12-91
		AU-A- 4430689	10-05-90
		JP-A- 2172912	04-07-90
