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(54) **AP2 DOMAIN TRANSCRIPTION FACTOR
ODP2 (OVULE DEVELOPMENT PROTEIN 2)
AND METHODS OF USE**

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800/320.3; 435/468

ABSTRACT

Methods and compositions for modulating plant development are provided. Nucleotide sequences and amino acid sequences encoding Ovule Development Protein 2 (ODP2) proteins are provided. The sequences can be used in a variety of methods including modulating development, developmental pathways, altering oil content in a plant, increasing transformation efficiencies, modulating stress tolerance, and modulating the regenerative capacity of a plant. Transformed plants, plant cells, tissues, and seed are also provided.

ZM-ODP2	(1)	1	50
OsAnt (BAB89946)	(1)	MATVNNWLAFSLSPQELPPSQTTDSTLISAAT-----ADHVS	GDVCFN
OSBNM (AAL47205)	(1)	MATMNNWLAFSLSPQDQLPPSQTNSTLISAAT--TTAGDSSTGDVCFN	
OSODP (CAE05555)	(1)	MATMNNWLAFSLSPQDQLPPSQTNSTFISAAT--TTAGDSSTGDVCFN	
AtODP (NP_197245)	(1)	MASADNWLGFSLSGQGNPQHHQNGPSAAGDA-----AIDISGSGDFY	G
AtBBM (AAM33803)	(1)	MNSMNNWLGFSLSPHDQNHHRTDVSSTTRTA-----VDVAGGYCFDL	
BnBBM1 (AAM33800)	(1)	MNSMNNWLGFSLSPHDQNHHRTDVSSTTRTA-----VDVAGGYCFDL	
BnBBM2 (AAM33801)	(1)	--MNNNWLGFSLSPPYEQNHHRKDVYSSTTTV-----VDVAGEYCYD	P
AtODP (NP_175530)	(1)	--MNNNWLGFSLSPPYEQNHHRKDVCSSTTTA-----VDVAGEYCYD	
AtODP (BAB02492)	(1)	MNSNNWLAFPLSPTHSSLPPHIHSSQNSHFPNLGLVNDNIDNPQFQNGW	N
AtODP (AAD30633)	(1)	(1) -----MDNPFPQTQEWN	
Consensus	(1)	M S NNWLGFSLSP DQ	S S A
			VD A F
ZM-ODP2	(44)	51	100
OsAnt (BAB89946)	(49)	IPQDWSMRGSELSALVAEPKLEDFLGGISFS-EQHHKANCNMI PSTSSTV	
OSBNM (AAL47205)	(49)	IPQDWSMRGSELSALVAEPKLEDFLGGISFS-EQHHKANCNMI PSTSSTV	
OSODP (CAE05555)	(45)	IP-----QAHPS	
AtODP (NP_197245)	(44)	LPTPDAHIGMAGEDAPYGVMDAFNRGTHETQDWAMRGLDYGGGSDLSM	
AtBBM (AAM33803)	(44)	AAPSDESSAVQTSFLSPFGVTLEAFTRDN--NSHSRDWDINGACNNIN	
BnBBM1 (AAM33800)	(42)	AAPSDESSAVQTSFLSPFGVTLEAFTRDN--NSHSRDWDINGACNNIN	
BnBBM2 (AAM33801)	(42)	TAASDESSAIQTSFPSFGVVDAFTRDN--NSHSRDWDINGSACNNIH	
AtODP (NP_175530)	(50)	TAASDESSAIQTSFPSFGVVDAFTRDN--NSHSRDWDINGSACNNIH	
AtODP (BAB02492)	(12)	MINPHGGGGE---GGEVPKVADFLGVS---KSGDHHTDHNLVPYNDIH	
AtODP (AAD30633)	(1)	MINPHGGGDE---GGEVPKVADFLGVS---KPDENQS-----NLHV	
Consensus	(51)	I G S V DF	NSH R D N A N I
ZM-ODP2	(93)	101	150
OsAnt (BAB89946)	(99)	CYASSGASTGYHHQLYHQPTSSALHFADSVMVASSAGVHDGGAMLSAAA	
OSBNM (AAL47205)	(56)	CYASSGSSV---GYLYPPPSSSSLQFADSVMVATSSPVVAHDGVSGGGMV	
OSODP (CAE05555)	(95)	(56) -----	
AtODP (NP_197245)	(91)	LVGSSGGRRTVAGDGVGEAPKLENFLDGNFSFDVHGQAAGGYLYSGSAV	
AtBBM (AAM33803)	(91)	NNEQNG-----P--KLENFLGRTTTIYNTNETVVDGNG-----	
BnBBM1 (AAM33800)	(89)	NDEQDG-----P--KLENFLGRTTTIYNTNETVVDGNG-----	
BnBBM2 (AAM33801)	(89)	NDEQDG-----P--KLENFLGRTTTIYNTNETVVDGNG-----	
AtODP (NP_175530)	(92)	QTNAS-----DYYFQTNSLLP-----	
AtODP (BAB02492)	(48)	AYNDS-----DYYFHTNSLMP-----V-----	
AtODP (AAD30633)	(43)	QTNAS-----DYYFQTNSLLP-----	
Consensus	(101)	N SG	P ENF S I
ZM-ODP2	(143)	151	200
OsAnt (BAB89946)	(146)	NGVA--GAASANGGGIGLMSIKNWLRSPQAPMQPRVAAAEGAQGLSLSMNN	
OSBNM (AAL47205)	(56)	SAAA---AAAASNGGGIGLMSIKNWLRSPQAPQP-----AQALSLSMN	
OSODP (CAE05555)	(145)	(56) -----TPAIGNGGIGLMSIKNWLRSPQAPQP-----AQALSLSMN	
AtODP (NP_197245)	(122)	GGAGGYSNGGGGGTIELMSIKTWLRSNQSQQQP-----SPPQHADQG	
AtBBM (AAM33803)	(122)	---DCGGGDGGGGGSGLGLSMIKTWLRSNHSVANANH-----	
BnBBM1 (AAM33800)	(121)	---DCGGGDGGGGGSGLGLSMIKTWLRSNHSVANANH-----	
BnBBM2 (AAM33801)	(121)	---GCYGGGDGGGGSGLGLSMIKTWLRSNQPVVDNVN-----	
AtODP (NP_175530)	(108)	---GCYGGGDGGGGSGLGLSMIKTWLRSNQPVVDNVN-----	
AtODP (BAB02492)	(66)	-----TVVTCASNAPNN-----	
AtODP (AAD30633)	(59)	-----QSNDVVVAACDSNTPNNSSY-----	
Consensus	(151)	GG GGG IGLSMIKTWLRSNQ P	N

FIG. 1-1

		201			250
ZM-ODP2	(191)	MAGTTQGAAG--MPLLAGERAR----APESVSTSAQGGAVVVTPAKEDS			
OsAnt (BAB89946)	(186)	MAGTTAQGGGAMALLAGAGERGRTTPASESLSTSAGATTATMAGGRKE			
OSBNM (AAL47205)	(91)	MAGTTAQGGGAMALLAGAGERGRTTPASESLSTSAGATTATMAGGRKE			
OSODP (CAE05555)	(188)	MSTDASASSYACSDVLVGSCGGGG---AGGTASSHGQGLALSMSTGSVA			
AtODP (NP_197245)	(154)	-----QDNGNGARGLSSLMSMNSSTS-D			
AtBBM (AAM33803)	(154)	-----QDNGNGARGLSSLMSMNSSTS-D			
BnBBM1 (AAM33800)	(153)	-----QENGNAAKGLSSLMSMNSSTSCD			
BnBBM2 (AAM33801)	(153)	-----QENGNGAKGLSSLMSMNSSTSCD			
AtODP (NP_175530)	(120)	-----YELQESAHNLQSLTLSMGSTGA-A			
AtODP (BAB02492)	(86)	-----HELQESAHNLQSLTLSMGSTTT---			
AtODP (AAD30633)	(71)	-----YELQESAHNLQSLTLSMGSTGA-A			
Consensus	(201)	QE S A GLTSLM SS S D			
		251			300
ZM-ODP2	(234)	G---GSGVAGALVAVSTDGGG---GGASADNTARKTVDTFGQRTSIYR			
OsAnt (BAB89946)	(236)	INEEGSGSAGAVVAVGSESGGGAVVEAGAAAAAARKSVDTFGQRTSIYR			
OSBNM (AAL47205)	(141)	INEEGSGSAGAVVAVGSESGGGAVVEAGAAAAAARKSVDTFGQRTSIYR			
OSODP (CAE05555)	(235)	AG---GGGAVVAAESSSENKRDSP-GGAVDGAVPRKSIDTGFQRTSIYR			
AtODP (NP_197245)	(174)	SNNYNNNNDDVVQEKTIVDVVET-----TPK--KTIESFGQRTSIYR			
AtBBM (AAM33803)	(174)	SNNYNNNNDDVVQEKTIVDVVET-----TPK--KTIESFGQRTSIYR			
BnBBM1 (AAM33800)	(174)	NNNDNNNNVAQGKTIDDSVEA-----TPK--KTIESFGQRTSIYR			
BnBBM2 (AAM33801)	(174)	NNYSSNNLVAQGKTIDDSVEA-----TPK--KTIESFGQRTSIYR			
AtODP (NP_175530)	(143)	AAEVATVKASPAETSADNSSTNTSGGAI	VEATP	PRRTLET	FGQRTSIYR
AtODP (BAB02492)	(106)	AGNNVVDKASPSETTGDNAS--G-GALAVVETATP	PRALDT	FGQRTSIYR	
AtODP (AAD30633)	(94)	AAEVATVKASPAETSADNSSTNTSGGAI	VEATP	PRRTLET	FGQRTSIYR
Consensus	(251)	AN GS A A E T DS T GA A RKTIE	FGQRTSIYR		
		301			350
ZM-ODP2	(277)	GVTRHWTGRYEALWDNSCREGQTRKGRQVYLGGYDKEEKAARAYDLA			
OsAnt (BAB89946)	(286)	GVTRHWTGRYEALWDNSCREGQTRKGR---QGGYDKEEKAARAYDLA			
OSBNM (AAL47205)	(191)	GVTRHWTGRYEALWDNSCREGQTRKGR---QGGYDKEEKAARAYDLA			
OSODP (CAE05555)	(282)	GVTRHWTGRYEALWDNSCREGQSRKGR---QGGYDKEEKAARAYDLA			
AtODP (NP_197245)	(213)	GVTRHWTGRYEALWDNSCKREGQTRKGR---QGGYDKEEKAARAYDLA			
AtBBM (AAM33803)	(213)	GVTRHWTGRYEALWDNSCKREGQTRKGRQVYLGGYDKEEKAARAYDLA			
BnBBM1 (AAM33800)	(213)	GVTRHWTGRYEALWDNSCKREGQTRKGRQVYLGGYDKEEKAARAYDLA			
BnBBM2 (AAM33801)	(213)	GVTRHWTGRYEALWDNSCKREGQTRKGRQVYLGGYDKEEKAARAYDLA			
AtODP (NP_175530)	(193)	GVTRHWTGRYEALWDNSCREGQSRKGR---QGGYDKEEKAARAYDLA			
AtODP (BAB02492)	(153)	GVTRHWTGRYEALWDNSCREGQSRKGR---QGGYDKEEKAARAYDLA			
AtODP (AAD30633)	(144)	GVTRHWTGRYEALWDNSCREGQSRKGR---QGGYDKEEKAARAYDLA			
Consensus	(301)	GVTRHWTGRYEALWDNSCREGQTRKGR QGGYDKEEKAARAYDLA			
		351			400
ZM-ODP2	(327)	ALKYWGATTTTNFPVSNYEKELEDMKHMTRQEFVASLRRKSSGFSRGASI			
OsAnt (BAB89946)	(333)	ALKYWGPTTTTNFPVNYYEKELEEMKHMTRQEFVASLRRKSSGFSRGASI			
OSBNM (AAL47205)	(238)	ALKYWGPTTTTNFPVNYYEKELEEMKHMTRQEFVASLRRKSSGFSRGASI			
OSODP (CAE05555)	(329)	ALKYWGTTTTNFPMSENYEKELEEMKHMTRQEFVIAHRRNSSGFSRGASK			
AtODP (NP_197245)	(260)	ALKYWGTTTTNFPLSEYEKEVEEMKHMTRQEFVIAHRRNSSGFSRGASI			
AtBBM (AAM33803)	(263)	ALKYWGPTTTNFPLSEYEKEVEEMKHMTRQEFVIAHRRNSSGFSRGASI			
BnBBM1 (AAM33800)	(263)	ALKYWGTTTTNFPMSEYEKEVEEMKHMTRQEFVIAHRRNSSGFSRGASI			
BnBBM2 (AAM33801)	(263)	ALKYWGTTTTNFPMSEYEKEIEEMKHMTRQEFVIAHRRNSSGFSRGASI			
AtODP (NP_175530)	(240)	ALKYWGPTTTNFPI	TNYEKEVEEMKHMTRQEFVIAIRRKSSGFSRGASM		
AtODP (BAB02492)	(200)	ALKYWGPTTTNFPI	TNYEKEVEEMKHMTRQEFVAAIRRKSSGFSRGASM		
AtODP (AAD30633)	(191)	ALKYWGPTTTNFPI	TNYEKEVEEMKHMTRQEFVASIRRKSSGFSRGASM		
Consensus	(351)	ALKYWGPTTTNFPI	SNYEKEVEEMKHMTRQEFVASLRRKSSGFSRGASI		

FIG. 1-2

		551		450
ZM-ODP2	(377)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTGSTQEEAAEAYDIAAIKFRG		
OsAnt (BAB89946)	(383)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTGSTQEEAAEAYDIAAIKFRG		
OSBNM (AAL47205)	(288)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTGSTQEEAAEAYDIAAIKFRG		
OSODP (CAE05555)	(379)	YRGVTRHHQHGRWQARIGRVAGNKDIYLGTGSTEEAAEAYDIAAIKFRG		
AtODP (NP_197245)	(310)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTFGTGSTQEEAAEAYDIAAIKFRG		
AtBBM (AAM33803)	(313)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTFGTGSTQEEAAEAYDIAAIKFRG		
BnBBM1 (AAM33800)	(313)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTFGTGSTQEEAAEAYDIAAIKFRG		
BnBBM2 (AAM33801)	(313)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTFGTGSTQEEAAEAYDIAAIKFRG		
AtODP (NP_175530)	(290)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTGSTEEAAEAYDIAAIKFRG		
AtODP (BAB02492)	(250)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTGSTEEAAEAYDIAAIKFRG		
AtODP (AAD30633)	(241)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTGSTEEAAEAYDIAAIKFRG		
Consensus	(401)	YRGVTRHHQHGRWQARIGRVAGNKDLYLGTGSTQEEAAEAYDIAAIKFRG		
		551		500
ZM-ODP2	(427)	LNAVTNFDMSRYDVKSILDSSALPIG-SAAKRLKEAEAAAASAQHHHAGVV		
OsAnt (BAB89946)	(433)	LNAVTNFDMSRYDVKSILDSSALPIG-TAAKRLKDAEAAA-----		
OSBNM (AAL47205)	(338)	LNAVTNFDMSRYDVKSILDSSALPIG-TAAKRLKDAEAAA-----		
OSODP (CAE05555)	(429)	LNAVTNFDMSRYDVKSILDSSALPIG-GAARRLKEAEVAA-----A-		
AtODP (NP_197245)	(360)	LSAVTNFDMNRYNVKAILESPSLPIG-SSAKRLKDVNNPVP-----		
AtBBM (AAM33803)	(363)	LSAVTNFDMNRYNVKAILESPSLPIG-SSAKRLKDVNNPVP-----		
BnBBM1 (AAM33800)	(363)	LTAVTNFDMNRYNVKAILESPSLPIG-SAAKRLKEANRPVPS-----		
BnBBM2 (AAM33801)	(363)	LTAVTNFDMNRYNVKAILESPSLPIG-SAAKRLKEANRPVPS-----		
AtODP (NP_175530)	(340)	LNAVTNFEINRYDVKAILESNTLPIGGAAKRLKEAQALESSRKR-----		
AtODP (BAB02492)	(300)	LNAVTNFEINRYDVKAILESNTLPIGGAAKRLKEAQALESSRKR-----		
AtODP (AAD30633)	(291)	LNAVTNFEINRYDVKAILESNTLPIGGAAKRLKEAQALESSRKR-----		
Consensus	(451)	LNAVTNFDMNRYDVKAILES SLPIG SAAKRLKEANA S		
		551		550
ZM-ODP2	(476)	SYDVGRIASQLGDGGALA-AAYCAHYHG-AAWPTIAFQPGAAS----T		
OsAnt (BAB89946)	(472)	AYDVGRIASHLGGDGAYA-AHYGHHHSAAAAPWPTIAFQAAAAPPHAA		
OSBNM (AAL47205)	(377)	AYDVGRIASHLGGDGAYA-AHYGHHHSAAAAPWPTIAFQAAAAPPHAA		
OSODP (CAE05555)	(469)	AAGGGVIVSHLADGG-----VGGYYYG---CGPTIAFGGGGQQPAPLA		
AtODP (NP_197245)	(400)	---AMMISNNVSESEN-----NVSGWQNTAFQHHQGMDLSLLQQQERYV		
AtBBM (AAM33803)	(403)	---AMMISNNVSESEN-----NVSGWQNTAFQHHQGMDLSLLQQQERYV		
BnBBM1 (AAM33800)	(404)	---MMMISSNNVSESEN-----SASGWQNAAVQHHQGVDLSLLHQHQERYN		
BnBBM2 (AAM33801)	(404)	---MMMISSNNVSESEN-----NASGWQNAAVQHHQGVDLSLLHQHQERYN		
AtODP (NP_175530)	(385)	-EEMIALGSNFHQYGAASSSSVASSRQLQPYPLSIQQPFEHLHHHQP		
AtODP (BAB02492)	(346)	-EEMIALGSNFHQYGAASSSSVASSRQLQPYPLSIQQPFEHLHHHQP		
AtODP (AAD30633)	(336)	-EEMIALGSNFHQYCAASCSSSVASSRQLQPYPLSIQQPFEHLHHHQP		
Consensus	(501)	DMM ISSNL E GA A SG A Q HP I Q		
		551		600
ZM-ODP2	(518)	GLYHPYAQPMRGGGWCKQEVDHAVIAAAHSLQDLHHNLG-AAGAHDF		
OsAnt (BAB89946)	(520)	GLYHPYAQPLR---GWCKQEVDHAVIAAAHSLQDLHHNLG-AAAAAHD		
OSBNM (AAL47205)	(425)	GLYHPYAQPLR---GWCKQEVDHAVIAAAHSLQDLHHNLG-AAAAAHD		
OSODP (CAE05555)	(509)	VHYPYQGQASG---WCKPE-QDAVIAAGHCATDLQHLHLSGGAAATHN		
AtODP (NP_197245)	(442)	GYYN-GGNLST-----ESTRVCFKQEEEQQHFLRN-SPSHMTN		
AtBBM (AAM33803)	(445)	GYYN-GGNLST-----ESTRVCFKQEEEQQHFLRN-SPSHMTN		
BnBBM1 (AAM33800)	(446)	GYYYNGGNLSS-----ESARACFKQEDDQHHFLSN-TQSLMTN		
BnBBM2 (AAM33801)	(446)	GYYYNGGNLSS-----ESARACFKQEDDQHHFLSN-TQSLMTN		
AtODP (NP_175530)	(434)	LLTLQNNN-----DISQYHDSFSYIQTQLHLHQO-QTNNYLQ		
AtODP (BAB02492)	(394)	NDISHYNNNNNA-----HDSSSFNHSYIQTQLHLHQO-TNNYLQ		
AtODP (AAD30633)	(385)	LLTLQNNN-----DISQYHDSFSYIQTQLHLHQO-QTNNYLQ		
Consensus	(551)	G Y GN S AAF IQDQ HL N S N		

FIG. 1-3

			601		650
ZM-ODP2	(566)	FSAGQQAAAAAMHGLSIDSASLEHSTGSNSVYNGVGDSNGASAVGGS			
OsAnt (BAB89946)	(565)	FFS---QAMQQQHGLGSIDNASLEHSTGSNSVYNGDNG-----GGG			
OSBNM (AAL47205)	(470)	FFS---QAMQQQHGLGSIDNASLEHSTGSNSVYNGDNG-----GGG			
OSODP (CAE05555)	(554)	FFQ----QPASS-----SAVYGNNGG-----GG			
AtODP (NP_197245)	(478)	VDHHS-----STSDDSVTVCNVVVS-----YGG			
AtBBM (AAM33803)	(481)	VDHHS-----STSDDSVTVCNVVVS-----YGG			
BnBBM1 (AAM33800)	(483)	IDHQS-----SVSDDSVTVCNVVVG-----YGG			
BnBBM2 (AAM33801)	(483)	IDHQS-----SVSDDSVTVCNVVVG-----YGG			
AtODP (NP_175530)	(470)	SSS-----HTSQLYNAYLQS-N-----PGL			
AtODP (BAB02492)	(433)	QSSQ-----NSQQLYNAYLHS-N-----PAL			
AtODP (AAD30633)	(421)	SSS-----HTSQLYNAYLQS-N-----PGL			
Consensus	(601)	S	S	SVVYNG V	GG
			651		700
ZM-ODP2	(616)	GGGYMMPMSAAGATTSAMVSHEQVHARAYDEAKQAAQMYESYLVNAEN			
OsAnt (BAB89946)	(604)	GGYIMAPMSAVSATATAVASSHDHG----GDGGKQVQMGYDSYLVGADA			
OSBNM (AAL47205)	(509)	GGYIMAPMSAVSATATAVASSHDHG----GDGGKQVQMGYDSYLVGADA			
OSODP (CAE05555)	(573)	GNAFMMMPGAVVAAADHGGQSSAYGG----GDESGRLVGYDGVVDPYAA			
AtODP (NP_197245)	(501)	YQGFAIPVGTSVNYDPFTAEEIAYN-----AR-NHYYAQHQQ			
AtBBM (AAM33803)	(504)	YQGFAIPVGTSVNYDPFTAEEIAYN-----AR-NHYYAQHQQ			
BnBBM1 (AAM33800)	(506)	YQGFAAPV---NCDAYAASEFDYN-----AR-NHYYFAQQQQ			
BnBBM2 (AAM33801)	(506)	YQGFAAPV---NCDAYAASEFDYN-----AR-NHYYFAQQQQ			
AtODP (NP_175530)	(489)	LHGFFS----DNNNTSG-----FLGNNGIGIGSSSTVGSSAE			
AtODP (BAB02492)	(453)	LHGLVSTS-IVDNNNNNGGSSGSYNTAA--FLGNHGIGIGSSSTVGS--T			
AtODP (AAD30633)	(440)	LHGFFS----DNNNTSG-----FLGNNGIGIGSSSTVGSSAE			
Consensus	(651)	GFMAPM N GAS Y G IAIG SYVA			
			701		745
ZM-ODP2	(666)	NGGGRMSAWGTVVSSAAAAAASSNDNMAADVGHGGAQLFSWNNDT			
OsAnt (BAB89946)	(649)	YGGGGAGRMPSWAMTPASAPAATSSSDMTGVCHG-AQLFSWNNDT			
OSBNM (AAL47205)	(554)	YGGGGAGRMPSWAMTPASAPAATSSSDMTGVCHG-AQLFSWNNDT			
OSODP (CAE05555)	(619)	MRSAYELSQGSSSSSVAKAANGYPDNWSSPFNGMG-----			
AtODP (NP_197245)	(538)	QQQIQQSPGGDFPVAISNNHSSNMYFHGEGGGEG-APTFSWNDT			
AtBBM (AAM33803)	(541)	QQQIQQSPGGDFPVAISNNHSSNMYFHGEKGEG-APTFSWNDT			
BnBBM1 (AAM33800)	(539)	TQ---QSPGGDFPAAAMTNNVGSNMYHGEKGEGEV-APTFVWNND			
BnBBM2 (AAM33801)	(539)	TQ---HSPGGDFPAAAMTNNVGSNMYHGEKGEGEV-APTFVWNND			
AtODP (NP_175530)	(522)	EEFPNAVVDYDMPPSGGATGYGGWNSGESAQGSNPGGVFTMWNE-			
AtODP (BAB02492)	(498)	EEFPTVKTDYDMPPSSDGTGGYSGWTS-ESVQGSNPGGVFTMWNE-			
AtODP (AAD030633)	(473)	EEFPNAVVDYDMPPSGGATGYGGWNSGESAQGSNPGGVFTMWNE-			
Consensus	(701)	GDFP A A AS G G A LFSWNND			

FIG. 1-4

		1	50
ZM-ODP2_unmodifiedPEP		(1) MATIVNNWLAFLSFCIPPSOTTDSTILSAATAADIVSCDVCNLPQDWSM	
ZM-ODP2_modifiedPEP_id_97.3		(1) MATIVNNWLAFLSFCIPPSOTTDSTILSAATAADIVSCDVCNLPQDWSM	
ZM-ODP2_modifiedPEP_id_92.4		(1) MATIVNNWLAFLSFCIPPSOTTDSTILSAATAADIVSCDVCNLPQDWSM	
ZM-ODP2_modifiedPEP_id_87.3		(1) MATIVNNWLAFLSFCIPPSOTTDSTILSAATAADIVSCDVCNLPQDWSM	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(1) MATIVNNWLAFLSFCIPPSOTTDSTILSAATAADIVSCDVCNLPQDWSM	
ZM-ODP2_unmodifiedPEP		51	100
ZM-ODP2_modifiedPEP_id_97.3		(51) RCSEISALVAPKIEDFIGGLSFEQEHKANCNMLPSTSSTICYASSGAS	
ZM-ODP2_modifiedPEP_id_92.4		(51) RCSEISAIWAKIEPKIEDFIGGLSFEQEHKANCNMLPSTSSTICYASSGAS	
ZM-ODP2_modifiedPEP_id_87.3		(51) RCSEISAIIAEPKIEDFIGGLSFEQEHKANCNMLPSTSSTICYASSGAS	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(51) RCSEISIGIIGPKIEDFIGGLSFEQEHKANCNMLPSTSSTICYASSGAS	
ZM-ODP2_unmodifiedPEP		(51) RCSEISIGIIGPKIEDFIGGLSFEQEHKANCNMLPSTSSTICYASSGAS	150
ZM-ODP2_modifiedPEP_id_97.3		(101) TGYVHHOYHOPHTSSAIIHADDSVIVASSACVHOGAMISAIAAANGVAGAAS	
ZM-ODP2_modifiedPEP_id_92.4		(101) TGYVHHOYHOPHTSSAIIHADDSVIVASSACVHOGAMISAIAAANGVAGAAS	
ZM-ODP2_modifiedPEP_id_87.3		(101) TGYVHHOYHOPHTSSAIIHADDSVIVASSACVHOGAMISAIAAANGVAGAAS	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(101) TGYVHHOYHOPHTSSAIIHADDSVIVASSACVHOGAMISAIAAANGVAGAAS	
ZM-ODP2_unmodifiedPEP		151	200
ZM-ODP2_modifiedPEP_id_97.3		(151) ANGGGIGLSMIKNNWLRSQPAPMOPRVAAGAAGISIISNNMACTTQGAAC	
ZM-ODP2_modifiedPEP_id_92.4		(151) ANGGGIGLSMIKNNWLRSQPAPMOPRVAAGAAGISIISNNMACTTQGAAC	
ZM-ODP2_modifiedPEP_id_87.3		(151) ANGGGIGLSMIKNNWLRSQPAPMOPRVAAGAAGISIISNNMACTTQGAAC	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(151) ANGGGIGLSMIKNNWLRSQPAPMOPRVAAGAAGISIISNNMACTTQGAAC	
ZM-ODP2_unmodifiedPEP		201	250
ZM-ODP2_modifiedPEP_id_97.3		(201) MELIAGERARAPESVTSADGGAAVVTIAKEDSGGSGVAGAVLAVSTDTG	
ZM-ODP2_modifiedPEP_id_92.4		(201) MELIAGERARAPESVTSADGGAAVVTIAKEDSGGSGVAGAVLAVSTDTG	
ZM-ODP2_modifiedPEP_id_87.3		(201) MELIAGERARAPESVTSADGGAAVVTIAKEDSGGSGVAGAVLAVSTDTG	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(201) MELIAGERARAPESVTSADGGAAVVTIAKEDSGGSGVAGAVLAVSTDTG	
ZM-ODP2_unmodifiedPEP		251	300
ZM-ODP2_modifiedPEP_id_97.3		(251) GSGGAADNTARKTVDFGQRTSIYRGVTRHWTGRYEAHLWDNSCRREG	
ZM-ODP2_modifiedPEP_id_92.4		(251) GSGGAADNTARKTVDFGQRTSIYRGVTRHWTGRYEAHLWDNSCRREG	
ZM-ODP2_modifiedPEP_id_87.3		(251) GSGGAADNTARKTVDFGQRTSIYRGVTRHWTGRYEAHLWDNSCRREG	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(251) GSGGAADNTARKTVDFGQRTSIYRGVTRHWTGRYEAHLWDNSCRREG	
ZM-ODP2_unmodifiedPEP		301	350
ZM-ODP2_modifiedPEP_id_97.3		(301) QTRKGROVYLGGYDKEEKAARAYDLAAIKYWGATTITNEPVSNYEKELED	
ZM-ODP2_modifiedPEP_id_92.4		(301) QTRKGROVYLGGYDKEEKAARAYDLAAIKYWGATTITNEPVSNYEKELED	
ZM-ODP2_modifiedPEP_id_87.3		(301) QTRKGROVYLGGYDKEEKAARAYDLAAIKYWGATTITNEPVSNYEKELED	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(301) QTRKGROVYLGGYDKEEKAARAYDLAAIKYWGATTITNEPVSNYEKELED	
ZM-ODP2_unmodifiedPEP		351	400
ZM-ODP2_modifiedPEP_id_97.3		(351) MHHMTROEFVASLRRKSSGFSRGASIYRGVTRHJQHGRWQARIGRVAGNK	
ZM-ODP2_modifiedPEP_id_92.4		(351) MHHMTROEFVASLRRKSSGFSRGASIYRGVTRHJQHGRWQARIGRVAGNK	
ZM-ODP2_modifiedPEP_id_87.3		(351) MHHMTROEFVASLRRKSSGFSRGASIYRGVTRHJQHGRWQARIGRVAGNK	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(351) MHHMTROEFVASLRRKSSGFSRGASIYRGVTRHJQHGRWQARIGRVAGNK	
ZM-ODP2_unmodifiedPEP		401	450
ZM-ODP2_modifiedPEP_id_97.3		(401) DLYLGTFSTOEAAEAYDIAAIKFRGLNAVTFDMSRYDVKSILDSALP	
ZM-ODP2_modifiedPEP_id_92.4		(401) DLYLGTFSTOEAAEAYDIAAIKFRGLNAVTFDMSRYDVKSILDSALP	
ZM-ODP2_modifiedPEP_id_87.3		(401) DLYLGTFSTOEAAEAYDIAAIKFRGLNAVTFDMSRYDVKSILDSALP	
ZM-ODP2_modifiedPEP_id_82.4	Consensus	(401) DLYLGTFSTOEAAEAYDIAAIKFRGLNAVTFDMSRYDVKSILDSALP	

FIG. 2-1

451		500
ZM-ODP2_unmodifiedPEP	(451) IGSAAKRLKEAEAAAASAOHIIAGVVSYDVGRVIA ZM-ODP2_modifiedPEP_id_97.3	IGSAAKRLKEAEAAAASAOHIIAGVVSYDVGRVIA ZM-ODP2_modifiedPEP_id_92.4
ZM-ODP2_modifiedPEP_id_92.4	(451) IGSAAKRLKEAEAAAASAOHIIAGLLSYDLGRV ZM-ODP2_modifiedPEP_id_87.3	IGSAAKRLKEAEAAAASAOHIIAGLLSYDLGRV ZM-ODP2_modifiedPEP_id_82.4
ZM-ODP2_modifiedPEP_id_87.3	(451) IGSAAKRLKEAEAAAASAOHIIAGLLSYDLGRV Consensus	IGSAAKRLKEAEAAAASAOHIIAGLLSYDLGRV ZM-ODP2_unmodifiedPEP
ZM-ODP2_modifiedPEP_id_82.4	(451) IGSAAKRLKEAEAAAASAOHIIAGLLSYDLGRV Consensus	(501) HGAAPWTIAFQPGAA ZM-ODP2_modifiedPEP_id_97.3
Consensus		(501) HGAAPWTIAFQPGAA ZM-ODP2_modifiedPEP_id_92.4
		(501) HGAAPWTIAFQPGAA ZM-ODP2_modifiedPEP_id_87.3
		(501) HGAAPWTIAFQPGAA ZM-ODP2_modifiedPEP_id_82.4
		Consensus
551		600
ZM-ODP2_unmodifiedPEP	(551) DHHHNILCAAGAHDFESAGC ZM-ODP2_modifiedPEP_id_97.3	DHHHNILCAAGAHDFESAGC ZM-ODP2_modifiedPEP_id_92.4
ZM-ODP2_modifiedPEP_id_92.4	(551) DHHHNILCAAGAHDFESAGC ZM-ODP2_modifiedPEP_id_87.3	DHHHNILCAAGAHDFESAGC ZM-ODP2_modifiedPEP_id_82.4
ZM-ODP2_modifiedPEP_id_87.3	(551) DHHHNILCAAGAHDFESAGC Consensus	DHHHNILCAAGAHDFESAGC ZM-ODP2_unmodifiedPEP
ZM-ODP2_modifiedPEP_id_82.4	(551) DHHHNILCAAGAHDFESAGC Consensus	(601) GGVDSNGASAV ZM-ODP2_modifiedPEP_id_97.3
Consensus		(601) GGVDSNGASAV ZM-ODP2_modifiedPEP_id_92.4
		(601) GGVDSNGASAV ZM-ODP2_modifiedPEP_id_87.3
		(601) GGVDSNGASAV ZM-ODP2_modifiedPEP_id_82.4
		Consensus
651		700
ZM-ODP2_unmodifiedPEP	(651) AAQMGYESYL ZM-ODP2_modifiedPEP_id_97.3	AAQMGYESYL ZM-ODP2_modifiedPEP_id_92.4
ZM-ODP2_modifiedPEP_id_92.4	(651) AAQMGYESYL ZM-ODP2_modifiedPEP_id_87.3	AAQMGYESYL ZM-ODP2_modifiedPEP_id_82.4
ZM-ODP2_modifiedPEP_id_87.3	(651) AAQMGYESYL Consensus	AAQMGYESYL ZM-ODP2_unmodifiedPEP
ZM-ODP2_modifiedPEP_id_82.4	(651) AAQMGYESYL Consensus	(701) AQLFSVWNDT ZM-ODP2_modifiedPEP_id_97.3
Consensus		(701) AQLFSVWNDT ZM-ODP2_modifiedPEP_id_92.4
		(701) AQLFSVWNDT ZM-ODP2_modifiedPEP_id_87.3
		(701) AQLFSVWNDT ZM-ODP2_modifiedPEP_id_82.4
		Consensus
701		

FIG. 2-2

**AP2 DOMAIN TRANSCRIPTION FACTOR
ODP2 (OVULE DEVELOPMENT PROTEIN 2)
AND METHODS OF USE**

**CROSS-REFERENCE TO RELATED
APPLICATIONS**

[0001] This application is a continuation of U.S. application Ser. No. 11/045,802, filed on Jan. 28, 2005, which claims priority to U.S. Provisional Application No. 60/541,122, filed on Feb. 2, 2004, the contents of which are hereby incorporated by reference in their entirety.

**REFERENCE TO A SEQUENCE LISTING
SUBMITTED AS A TEXT FILE VIA EFS-WEB**

[0002] The official copy of the sequence listing is submitted electronically via EFS-Web as an ASCII formatted sequence listing with a file named 376742SEQULIST.TXT, created on Jul. 15, 2009, and having a size of 243 kilobytes and is filed concurrently with the specification. The sequence listing contained in this ASCII formatted document is part of the specification and is herein incorporated by reference in its entirety.

FIELD OF THE INVENTION

[0003] The invention relates to the field of the genetic manipulation of plants, particularly the modulation of gene activity and development in plants.

BACKGROUND OF THE INVENTION

[0004] Cell division plays a crucial role during all phases of plant development. The continuation of organogenesis and growth responses to a changing environment requires precise spatial, temporal and developmental regulation of cell division activity in meristems. Such control of cell division is also important in organs themselves for example, leaf expansion, and secondary growth. A complex network controls cell proliferation in eukaryotes. Various regulatory pathways communicate environmental constraints, such as nutrient availability, mitogenic signals such as growth factors or hormones, or developmental cues such as the transition from vegetative to reproductive. Ultimately, these regulatory pathways control the timing, frequency (rate), plane and position of cell divisions. The regulation of cell division impacts a variety of developmental pathways including transformation and plant regeneration.

[0005] Current transformation technology provides an opportunity to engineer plants with desired traits. Major advances in plant transformation have occurred over the last few years. However, in many major crop plants, serious genotype limitations still exist. Transformation of some agronomically important crop plants continues to be both difficult and time consuming.

[0006] For example, it is difficult to obtain a culture response from some maize genotypes. Typically, a suitable culture response has been obtained by optimizing medium components and/or explant material and source. This has led to success in some genotypes. While, transformation of model genotypes is efficient, the process of introgressing transgenes into production inbreds is laborious, expensive and time consuming. It would save considerable time and money if genes could be more efficiently introduced into and evaluated directly into inbreds. Accordingly, methods are needed in the art to increase transformation efficiencies of plants.

[0007] Influencing cell cycle and cell division can also affect various developmental pathways in a plant. Pathways of interest include those that influence embryo development. The AP2/ERF family of proteins is a plant-specific class of putative transcription factors that have been shown to regulate a wide-variety of developmental processes and are characterized by the presence of a AP2/ERF DNA binding domain. The AP2/ERF proteins have been subdivided into two distinct subfamilies based on whether they contain one (ERF subfamily) or two (AP2 subfamily) DNA binding domains.

[0008] One member of the AP2 family that has been implicated in a variety of critical plant cellular functions is the Baby Boom protein (BBM). The BBM protein from *Arabidopsis* is preferentially expressed in seed and has been shown to play a central role in regulating embryo-specific pathways. Overexpression of BBM has been shown to induce spontaneous formation of somatic embryos and cotyledon-like structures on seedlings. See, Boutilier et al. (2002) *The Plant Cell* 14:1737-1749. Thus, members of the AP2 protein family promote cell proliferation and morphogenesis during embryogenesis. Such activity finds potential use in promoting apomixis in plants.

[0009] Apomixis refers to the production of a seed from the maternal ovule tissue in the absence of egg cell fertilization (Koltunow (1995) *Plant Physiol* 108:1345-1352). Apomixis is a valuable trait for crop improvement since apomictic seeds give rise to clonal offspring and can therefore be used to genetically fix hybrid lines. The production of hybrid lines is intensive and costly. Production of seed through apomixis avoids these problems in that once a hybrid has been produced, it can be maintained clonally, thereby eliminating the need to maintain and cross separate parent lines. The use of apomictic seeds also eliminates the use of cuttings or tissue culture techniques to propagate lines, reduces the spread of disease which are easily spread through vegetative-propagated tissues and in many species reduces the size of the propagule leading to lower shipping and planting costs. Methods are therefore needed for the efficient production of apomictic seed.

[0010] Members of the APETALA2 (AP2) family of proteins play critical roles in a variety of important biological events including development, plant regeneration, cell division, etc. Accordingly, it is valuable to the field of agronomic development to identify and characterize novel AP2 family members and develop novel methods to modulate embryogenesis, transformation efficiencies, oil content, starch content and yield in a plant.

BRIEF SUMMARY OF THE INVENTION

[0011] Methods and compositions are provided to modulate plant development using DNA, RNA or protein derived from the maize AP2 family member ZmODP2. The present invention provides an isolated polypeptide comprising an amino acid sequence selected from the group consisting of: (a) the polypeptide comprising the amino acid sequence of SEQ ID NO:2, 26, or 28; (b) the polypeptide having at least 50% sequence identity to SEQ ID NO:2, 26, or 28, wherein the polypeptide has Ovule Development Protein 2 (ODP2) activity; (c) the polypeptide encoded by a polynucleotide that hybridizes under stringent conditions to a polynucleotide comprising the complement of SEQ ID NOS: 1, 3, 25, or 27, wherein the stringent conditions comprise hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37° C., and a wash in 0.1×SSC at 60° C. to 65° C.; and, (d) the polypeptide having

at least 70 consecutive amino acids of SEQ ID NO:2, 26, or 28, wherein the polypeptide retains ODP2 activity.

[0012] Further compositions of the invention include an isolated polynucleotide selected from the group consisting of: (a) the polynucleotide comprising SEQ ID NO:1, 3, 25 or 27; (b) the polynucleotide encoding the amino acid sequence of SEQ ID NO:2, 26 or 28; (c) the polynucleotide having at least 50% sequence identity to SEQ ID NO:1, 3, 25 or 27, wherein the polynucleotide encodes a polypeptide having ODP2 activity; (d) the polynucleotide having at least 200 consecutive nucleotides of SEQ ID NO:1, 3, 25 or 27 or a complement thereof, and, (e) the polynucleotide that hybridizes under stringent conditions to the complement of the polynucleotide of (a), wherein the stringent conditions comprise hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37° C., and a wash in 0.1×SSC at 60° C. to 65° C. Nucleotide constructs comprising the polynucleotide of the invention are also provided.

[0013] Additional compositions of the invention include plants having a heterologous polynucleotide of the invention operably linked to a promoter that drives expression in the plant. The plant can be a plant cell, a plant part, a seed, or a grain. Methods are provided to modulate development in a plant. In one embodiment, the plant of the invention has an altered oil phenotype. In specific embodiments the oil content of the plant is decreased. In other embodiments, starch production of the plant is modified. In specific embodiments, the starch content of the plant is increased. In another embodiment, the regenerative capacity of the plant is modified. In yet another embodiment, the plant produces an asexually derived embryo. In still another embodiment, the transformation efficiency of the plant is increased. In another embodiment, the seed set is increased or maintained during periods of abiotic stress. In still another embodiment, haploid embryos are produced from male or female gametes.

[0014] Methods of the invention comprise methods for modulating the activity and/or level of a polypeptide in a plant. This method comprises providing to the plant an ODP2 sequence of the invention.

[0015] The present invention further provides a method for altering the oil phenotype in a plant. The method comprises providing to the plant an ODP2 sequence of the invention; and, thereby altering the oil phenotype of the plant.

[0016] The present invention further provides a method for modifying starch production in a plant. The method comprises providing to the plant an ODP2 sequence of the invention; and, thereby modifying starch production of the plant.

[0017] The present invention further provides a method for producing asexually derived embryos. The method comprises introducing into a plant ODP2 sequence of the present invention; and, thereby producing asexually derived embryos. The asexually derived embryos can be somatic embryos, adventitious embryos, or gametophytic embryos.

[0018] The present invention also provides a method for modifying the regenerative capacity of a plant. The method comprises introducing into the plant an ODP2 nucleotide sequence of the invention, and thereby modifying the regenerative capacity of the plant.

[0019] The present invention also provides a method of transforming a plant. The method comprises providing to target plant an ODP2 sequence of the invention, and, transforming into the target plant a nucleotide sequence of interest. The regenerative capacity can be modified to include tissues

normally not amenable to culture including but not limited to leaves, stems, and mature seed.

[0020] The invention further provides a method for increasing transformation efficiency in a plant. The method comprises providing to the plant an ODP2 nucleotide sequence of the invention, and thereby increasing the transformation efficiency of the plant.

[0021] The invention further provides a method for increasing or maintaining yield in a plant under abiotic stress. The method comprises providing to the plant an ODP2 nucleotide sequence of the invention, and thereby increasing the stress tolerance of the plant.

BRIEF DESCRIPTION OF THE DRAWINGS

[0022] FIG. 1 shows an alignment of the amino acid sequence of maize Ovule Development Protein 2 (Zm-ODP2) (SEQ ID NO:2) with OsAnt (Accession No. BAB89946; SEQ ID NO:26), OSBNM (Accession No. AAL47205; SEQ ID NO:28); OSODP (Accession No. CAE05555; SEQ ID NO:29); AtODP (NP_197245; SEQ ID NO:30); ATBBM (Accession No. AAM33803; SEQ ID NO:31); BnBBM1 (AAM33800; SEQ ID NO:32); BnBBM2 (Accession No. AAM33801; SEQ ID NO:33); ATODP (Accession No. NP_175530; SEQ ID NO:34); AtODP (Accession No. BAB02492; SEQ ID NO:35); AtODP (Accession No. AAD30633; SEQ ID NO:36). All 11 proteins present in the alignment have two AP2 (APETALA2; pfam00847.8) domains. Using the amino acid numbering of the Zm-ODP2 polypeptide, the first AP2 domain is from about amino acid 273 to about 343 and the second AP2 domain is from about amino acid 375 to about 437. A consensus sequence for all 11 aligned polypeptides is also provided (SEQ ID NO:37).

[0023] FIG. 2 provides an amino acid alignment of the Zm-ODP2 amino acid sequence (ZM-ODP2_unmodified-PEP; SEQ ID NO:2) with four polypeptide variants of the Zm-ODP2 sequence. The variant amino acid sequences include ZM-ODP2_modifiedPEP_id_97.3 (SEQ ID NO:20) which shares 97.3% amino acid sequence identity with SEQ ID NO:2; ZM-ODP2_modifiedPEP_id_92.4 (SEQ ID NO:21) which shares 92.4% amino acid sequence identity with SEQ ID NO:2; ZM-ODP2_modifiedPEP_id_87.3 (SEQ ID NO:22) which shares 87.3% amino acid sequence identity with SEQ ID NO:2; and, ZM-ODP2_modifiedPEP_id_82.4 (SEQ ID NO:23) which shares 82.4% amino acid sequence identity with SEQ ID NO:2. The consensus sequence is set forth in SEQ ID NO:24.

DETAILED DESCRIPTION OF THE INVENTION

[0024] The present inventions now will be described more fully hereinafter with reference to the accompanying drawings, in which some, but not all embodiments of the invention are shown. Indeed, these inventions may be embodied in many different forms and should not be construed as limited to the embodiments set forth herein; rather, these embodiments are provided so that this disclosure will satisfy applicable legal requirements. Like numbers refer to like elements throughout.

[0025] Many modifications and other embodiments of the inventions set forth herein will come to mind to one skilled in the art to which these inventions pertain having the benefit of the teachings presented in the foregoing descriptions and the associated drawings. Therefore, it is to be understood that the inventions are not to be limited to the specific embodiments

disclosed and that modifications and other embodiments are intended to be included within the scope of the appended claims. Although specific terms are employed herein, they are used in a generic and descriptive sense only and not for purposes of limitation.

[0026] The article "a" and "an" are used herein to refer to one or more than one (i.e., to at least one) of the grammatical object of the article. By way of example, "an element" means one or more element.

Compositions

[0027] Compositions of the invention include polynucleotide sequence and amino acid sequence of Ovule Development Protein 2 (ODP2) proteins that are involved in regulating plant growth and development. In particular, the present invention provides for isolated nucleic acid molecules comprising nucleotide sequences encoding the amino acid sequences shown in SEQ ID NO:2, 26, or 28. Further provided are polypeptides having an amino acid sequence encoded by a nucleic acid molecule (SEQ ID NO: 1, 3, 25, or 27) described herein, and fragments and variants thereof.

[0028] The ODP2 polypeptides of the invention contain two predicted APETALA2 (AP2) domains and are members of the AP2 protein family (PFAM Accession PF00847). The AP2 domains of the maize ODP2 polypeptide are located from about amino acids S273 to N343 and from about S375 to R437 of SEQ ID NO:2). The AP2 family of putative transcription factors have been shown to regulate a wide range of developmental processes, and the family members are characterized by the presence of an AP2 DNA binding domain. This conserved core is predicted to form an amphipathic alpha helix that binds DNA. The AP2 domain was first identified in APETALA2, an *Arabidopsis* protein that regulates meristem identity, floral organ specification, seed coat development, and floral homeotic gene expression. The AP2 domain has now been found in a variety of proteins.

[0029] The ODP2 polypeptides of the invention share homology with several polypeptides within the AP2 family. FIG. 1 provides an alignment of the maize and rice ODP2 polypeptides of the present invention with 8 other proteins having two AP2 domains. A consensus sequence of all proteins appearing in the alignment is also provided in FIG. 1. The alignment of FIG. 1 was generated using Align X® which employs a modified Clustal W algorithm to generate multiple sequence alignments. FIG. 1 demonstrates that the maize ODP2 polypeptide of the present invention (SEQ ID NO:2) shares about 51.7% sequence identity and 62.3% sequence similarity across the full sequence with the rice sequences of OsBNM3 (ovule development aintegumenta-like protein) (Genbank Accession No. AAL47205; SEQ ID NO:28). In addition, the ODP2 polypeptide of SEQ ID NO:2 shares 65.4% sequence identity and 72.7% sequence similarity across the full sequence to a putative ovule development protein from rice (OS) (Genbank Accession No. BAB89946; SEQ ID NO:26).

[0030] The OsBNM3 polypeptide sequence (SEQ ID NO:28), the OS polypeptide (SEQ ID NO:26), as well as the ODP2 sequence (SEQ ID NO:2) share homology with *Arabidopsis* Baby Boom (AtBBM, AAM33803; SEQ ID NO:31). Blast alignments demonstrate that Zm-ODP2 shares about 38.1% sequence identity and about 46.3% sequence similarity across the full length of the *Arabidopsis* Baby Boom polypeptide (AtBBM). See FIG. 1. The AtBBM polypeptide encodes an AP2 domain transcription factor and

is optimally expressed in the developing embryo and seeds. AtBBM has been shown to trigger formation of somatic embryos and cotyledon-like structures on seedlings and thus activates signal transduction pathways leading to the induction of embryo development from differentiated somatic cells. See, for example, Boutilier et al. (2002) *Plant Cell* 14:1737-49), herein incorporated by reference. Accordingly, the ODP2 sequences of the present invention also find use in modifying the regenerative capabilities of plants and rendering the plant embryogenic.

[0031] In addition, other polypeptides that influence ovule and embryo development and stimulate cell growth, such as, Lec1, Kn1 family, WUSCHEL, Zwille, and Aintegumenta (ANT) allow for increased transformation efficiencies when expressed in plants. See, for example, U.S. Application No. 2003/0135889, herein incorporated by reference. In fact, a maize Lec1 homologue of the *Arabidopsis* embryogenesis controlling gene AtLEC1, has been shown to increase oil content and transformation efficiencies in plants. See, for example, WO 03001902 and U.S. Pat. No. 6,512,165. Accordingly, the Zm-ODP2 sequences of the invention find further use in increasing transformation efficiencies in plants.

[0032] The invention encompasses isolated or substantially purified nucleic acid or protein compositions. An "isolated" or "purified" nucleic acid molecule or protein, or biologically active portion thereof, is substantially or essentially free from components that normally accompany or interact with the nucleic acid molecule or protein as found in its naturally occurring environment. Thus, an isolated or purified nucleic acid molecule or protein is substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. Optimally, an "isolated" nucleic acid is free of sequences (optimally protein encoding sequences) that naturally flank the nucleic acid (i.e., sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb, or 0.1 kb of nucleotide sequences that naturally flank the nucleic acid molecule in genomic DNA of the cell from which the nucleic acid is derived. A protein that is substantially free of cellular material includes preparations of protein having less than about 30%, 20%, 10%, 5%, or 1% (by dry weight) of contaminating protein. When the protein of the invention or biologically active portion thereof is recombinantly produced, optimally culture medium represents less than about 30%, 20%, 10%, 5%, or 1% (by dry weight) of chemical precursors or non-protein-of-interest chemicals.

[0033] Fragments and variants of the disclosed nucleotide sequences and proteins encoded thereby are also encompassed by the present invention. By "fragment" is intended a portion of the nucleotide sequence or a portion of the amino acid sequence and hence protein encoded thereby. Fragments of a nucleotide sequence may encode protein fragments that retain the biological activity of the native protein and hence have ODP2 activity. Alternatively, fragments of a nucleotide sequence that are useful as hybridization probes generally do not encode fragment proteins retaining biological activity. Thus, fragments of a nucleotide sequence may range from at least about 20 nucleotides, about 50 nucleotides, about 100 nucleotides, and up to the full-length nucleotide sequence encoding the proteins of the invention.

[0034] By “ODP2 activity” or “Ovule Development Protein 2 activity” is intended the ODP2 polypeptide has at least one of the following exemplary activities: increases the regenerative capability of a plant cell, renders the plant cell embryogenic, increases the transformation efficiencies of a plant cell, alters the oil content of a plant cell, binds DNA, increases abiotic stress tolerance, increases or maintains yield under abiotic stress, increases asexual embryo formation, alters starch content, alters embryo size or activates transcription. Methods to assay for such activity are known in the art and are described more fully below.

[0035] A fragment of an ODP2 nucleotide sequence that encodes a biologically active portion of an ODP2 protein of the invention will encode at least 15, 25, 30, 50, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 709 contiguous amino acids, or up to the total number of amino acids present in a full-length ODP2 protein of the invention (for example, 710 amino acids for SEQ ID NO: 2, 692 amino acids for SEQ ID NO: 25 and 597 for SEQ ID NO:27). Fragments of an ODP2 nucleotide sequence that are useful as hybridization probes or PCR primers generally need not encode a biologically active portion of an ODP2 protein.

[0036] Thus, a fragment of an ODP2 nucleotide sequence may encode a biologically active portion of an ODP2 protein, or it may be a fragment that can be used as a hybridization probe or PCR primer using methods disclosed below. A biologically active portion of an ODP2 protein can be prepared by isolating a portion of one of the ODP2 nucleotide sequences of the invention, expressing the encoded portion of the ODP2 protein (e.g., by recombinant expression in vitro), and assessing the activity of the encoded portion of the ODP2 protein. Nucleic acid molecules that are fragments of an ODP2 nucleotide sequence comprise at least 16, 20, 50, 75, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 800, 900, 1,000, 1,100, 1,200, 1,300, or 1,400, 1,500, 1,600, 1,700, 1,800, 1,900, 2,000, 2,100, 2,200 contiguous nucleotides, or up to the number of nucleotides present in a full-length ODP2 nucleotide sequence disclosed herein (for example, 2,260, 2133, 2079, and 1794 nucleotides for SEQ ID NOS:1, 3, 25 and 27, respectively).

[0037] “Variants” is intended to mean substantially similar sequences. For polynucleotides, a variant comprises a deletion and/or addition of one or more nucleotides at one or more internal sites within the native polynucleotide and/or a substitution of one or more nucleotides at one or more sites in the native polynucleotide. As used herein, a “native” polynucleotide or polypeptide comprises a naturally occurring nucleotide sequence or amino acid sequence, respectively. For polynucleotides, conservative variants include those sequences that, because of the degeneracy of the genetic code, encode the amino acid sequence of one of the ODP2 polypeptides of the invention. Naturally occurring variants such as these can be identified with the use of well-known molecular biology techniques, as, for example, with polymerase chain reaction (PCR) and hybridization techniques as outlined below. Variant polynucleotides also include synthetically derived polynucleotide, such as those generated, for example, by using site-directed mutagenesis but which still encode an ODP2 protein of the invention. Generally, variants of a particular polynucleotide of the invention will have at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or

more sequence identity to that particular polynucleotide as determined by sequence alignment programs and parameters described elsewhere herein.

[0038] Variants of a particular polynucleotide of the invention (i.e., the reference polynucleotide) can also be evaluated by comparison of the percent sequence identity between the polypeptide encoded by a variant polynucleotide and the polypeptide encoded by the reference polynucleotide. Thus, for example, an isolated polynucleotide that encodes a polypeptide with a given percent sequence identity to the polypeptide of SEQ ID NO:2, 26, or 28 are disclosed. Percent sequence identity between any two polypeptides can be calculated using sequence alignment programs and parameters described elsewhere herein. Where any given pair of polynucleotides of the invention is evaluated by comparison of the percent sequence identity shared by the two polypeptides they encode, the percent sequence identity between the two encoded polypeptides is at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity.

[0039] “Variant” protein is intended to mean a protein derived from the native protein by deletion or addition of one or more amino acids at one or more internal sites in the native protein and/or substitution of one or more amino acids at one or more sites in the native protein. Variant proteins encompassed by the present invention are biologically active, that is they continue to possess the desired biological activity of the native protein, that is, the polypeptide has ODP2 activity (i.e., modulating the regenerative capability of a plant, rendering the plant embryogenic, increasing the transformation efficiency of a plant, altering oil content of a plant, increasing cell proliferation, increasing abiotic stress tolerance, increasing or maintaining yield under abiotic stress, modifying starch content, increasing asexual embryo formation, binding DNA or regulating transcription) as described herein. Such variants may result from, for example, genetic polymorphism or from human manipulation. Biologically active variants of a native ODP2 protein of the invention will have at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity to the amino acid sequence for the native protein as determined by sequence alignment programs and parameters described elsewhere herein. A biologically active variant of a protein of the invention may differ from that protein by as few as 1-15 amino acid residues, as few as 1-10, such as 6-10, as few as 5, as few as 4, 3, 2, or even 1 amino acid residue.

[0040] The proteins of the invention may be altered in various ways including amino acid substitutions, deletions, truncations, and insertions. Methods for such manipulations are generally known in the art. For example, amino acid sequence variants of the ODP2 proteins can be prepared by mutations in the DNA. Methods for mutagenesis and nucleotide sequence alterations are well known in the art. See, for example, Kunkel (1985) *Proc. Natl. Acad. Sci. USA* 82:488-492; Kunkel et al. (1987) *Methods in Enzymol.* 154:367-382; U.S. Pat. No. 4,873,192; Walker and Gaastra, eds. (1983) *Techniques in Molecular Biology* (MacMillan Publishing Company, New York) and the references cited therein. Guidance as to appropriate amino acid substitutions that do not affect biological activity of the protein of interest may be found in the model of Dayhoff et al. (1978) *Atlas of Protein Sequence and Structure* (Nat'l. Biomed. Res. Found., Washington, D.C.), herein incorporated by reference. Conservative substi-

tutions, such as exchanging one amino acid with another having similar properties, may be optimal.

[0041] Thus, the genes and nucleotide sequences of the invention include both the naturally occurring sequences as well as mutant forms. Likewise, the proteins of the invention encompass both naturally occurring proteins as well as variations and modified forms thereof. Such variants will continue to possess the desired ODP2 activity. Obviously, the mutations that will be made in the DNA encoding the variant must not place the sequence out of reading frame and optimally will not create complementary regions that could produce secondary mRNA structure. See, EP Patent Application Publication No. 75,444.

[0042] The deletions, insertions, and substitutions of the protein sequences encompassed herein are not expected to produce radical changes in the characteristics of the protein. However, when it is difficult to predict the exact effect of the substitution, deletion, or insertion in advance of doing so, one skilled in the art will appreciate that the effect will be evaluated by routine screening assays. Various methods for screening for ODP2 activity are discussed in detail elsewhere herein.

[0043] Variant nucleotide sequences and proteins also encompass sequences and proteins derived from a mutagenic and recombinogenic procedure such as DNA shuffling. With such a procedure, one or more different ODP2 coding sequences can be manipulated to create a new ODP2 possessing the desired properties. In this manner, libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides comprising sequence regions that have substantial sequence identity and can be homologously recombined in vitro or in vivo. For example, using this approach, sequence motifs encoding a domain of interest may be shuffled between the ODP2 gene of the invention and other known ODP2 genes to obtain a new gene coding for a protein with an improved property of interest, such as an increased K_m in the case of an enzyme. Strategies for such DNA shuffling are known in the art. See, for example, Stemmer (1994) *Proc. Natl. Acad. Sci. USA* 91:10747-10751; Stemmer (1994) *Nature* 370:389-391; Crameri et al. (1997) *Nature Biotech.* 15:436-438; Moore et al. (1997) *J. Mol. Biol.* 272:336-347; Zhang et al. (1997) *Proc. Natl. Acad. Sci. USA* 94:4504-4509; Crameri et al. (1998) *Nature* 391:288-291; and U.S. Pat. Nos. 5,605,793 and 5,837,458.

[0044] The nucleotide sequences of the invention can be used to isolate corresponding sequences from other organisms, particularly other plants including other monocots. In this manner, methods such as PCR, hybridization, and the like can be used to identify such sequences based on their sequence homology to the sequence set forth herein. Sequences isolated based on their sequence identity to the entire ODP2 sequence set forth herein or to fragments thereof are encompassed by the present invention. Such sequences include sequences that are orthologs of the disclosed sequences. By "orthologs" is intended genes derived from a common ancestral gene and which are found in different species as a result of speciation. Genes found in different species are considered orthologs when their nucleotide sequences and/or their encoded protein sequences share substantial identity as defined elsewhere herein. Functions of orthologs are often highly conserved among species. Thus, isolated sequences that encode for an ODP2 protein and which hybridize under stringent conditions to the ODP2

sequence disclosed herein, or to fragments thereof, are encompassed by the present invention.

[0045] In a PCR approach, oligonucleotide primers can be designed for use in PCR reactions to amplify corresponding DNA sequences from cDNA or genomic DNA extracted from any plant of interest. Methods for designing PCR primers and PCR cloning are generally known in the art and are disclosed in Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, N.Y.). See also Innis et al., eds. (1990) *PCR Protocols: A Guide to Methods and Applications* (Academic Press, New York); Innis and Gelfand, eds. (1995) *PCR Strategies* (Academic Press, New York); and Innis and Gelfand, eds. (1999) *PCR Methods Manual* (Academic Press, New York). Known methods of PCR include, but are not limited to, methods using paired primers, nested primers, single specific primers, degenerate primers, gene-specific primers, vector-specific primers, partially-mismatched primers, and the like.

[0046] In hybridization techniques, all or part of a known nucleotide sequence is used as a probe that selectively hybridizes to other corresponding nucleotide sequences present in a population of cloned genomic DNA fragments or cDNA fragments (i.e., genomic or cDNA libraries) from a chosen organism. The hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides, and may be labeled with a detectable group such as ^{32}P , or any other detectable marker. Thus, for example, probes for hybridization can be made by labeling synthetic oligonucleotides based on the ODP2 sequences of the invention. Methods for preparation of probes for hybridization and for construction of cDNA and genomic libraries are generally known in the art and are disclosed in Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, N.Y.).

[0047] For example, the entire ODP2 sequence disclosed herein, or one or more portions thereof, may be used as a probe capable of specifically hybridizing to corresponding ODP2 sequences and messenger RNAs. To achieve specific hybridization under a variety of conditions, such probes include sequences that are unique among ODP2 sequences and are optimally at least about 10 nucleotides in length, and at least about 20 nucleotides in length. Such probes may be used to amplify corresponding ODP2 sequences from a chosen plant by PCR. This technique may be used to isolate additional coding sequences from a desired plant or as a diagnostic assay to determine the presence of coding sequences in a plant. Hybridization techniques include hybridization screening of plated DNA libraries (either plaques or colonies; see, for example, Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, N.Y.).

[0048] Hybridization of such sequences may be carried out under stringent conditions. By "stringent conditions" or "stringent hybridization conditions" is intended conditions under which a probe will hybridize to its target sequence to a detectably greater degree than to other sequences (e.g., at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences that are 100% complementary to the probe can be identified (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing).

Generally, a probe is less than about 1000 nucleotides in length, optimally less than 500 nucleotides in length.

[0049] Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30° C. for short probes (e.g., 10 to 50 nucleotides) and at least about 60° C. for long probes (e.g., greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37° C., and a wash in 1 \times to 2 \times SSC (20 \times SSC=3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55° C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37° C., and a wash in 0.5 \times to 1 \times SSC at 55 to 60° C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37° C., and a wash in 0.1 \times SSC at 60 to 65° C. Optionally, wash buffers may comprise about 0.1% to about 1% SDS. Duration of hybridization is generally less than about 24 hours, usually about 4 to about 12 hours. The duration of the wash time will be at least a length of time sufficient to reach equilibrium.

[0050] Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_m can be approximated from the equation of Meinkoth and Wahl (1984) *Anal. Biochem.* 138:267-284: $T_m = 81.5^\circ\text{C} + 16.6 (\log M) + 0.41 (\% \text{GC}) - 0.61 (\% \text{form}) - 500/L$; where M is the molarity of monovalent cations, % GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The T_m is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe. T_m is reduced by about 1° C. for each 1% of mismatching; thus, T_m , hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with $\geq 90\%$ identity are sought, the T_m can be decreased 10° C. Generally, stringent conditions are selected to be about 5° C. lower than the thermal melting point (T_m) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4° C. lower than the thermal melting point (T_m); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 11° C. lower than the thermal melting point (T_m); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20° C. lower than the thermal melting point (T_m). Using the equation, hybridization and wash compositions, and desired T_m , those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T_m of less than 45° C. (aqueous solution) or 32° C. (formamide solution), it is optimal to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen (1993) *Laboratory Techniques in Biochemistry and Molecular Biology-Hybridization with Nucleic Acid Probes*, Part I, Chapter 2 (Elsevier, New York); and Ausubel et al., eds. (1995) *Current Protocols in Molecular Biology*, Chapter 2 (Greene Publishing and

Wiley-Interscience, New York). See Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, N.Y.).

[0051] The following terms are used to describe the sequence relationships between two or more nucleic acids or polynucleotides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", (d) "percentage of sequence identity", and (e) "substantial identity".

[0052] (a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.

[0053] (b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and is subtracted from the number of matches.

[0054] Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent sequence identity between any two sequences can be accomplished using a mathematical algorithm. Non-limiting examples of such mathematical algorithms are the algorithm of Myers and Miller (1988) *CABIOS* 4:11-17; the local alignment algorithm of Smith et al. (1981) *Adv. Appl. Math.* 2:482; the global alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453; the search-for-local alignment method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci.* 85:2444-2448; the algorithm of Karlin and Altschul (1990) *Proc. Natl. Acad. Sci. USA* 87:2264, modified as in Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA* 90:5873-5877.

[0055] Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Such implementations include, but are not limited to: CLUSTAL in the PC/Gene program (available from Intelligenetics, Mountain View, Calif.); the ALIGN program (Version 2.0) and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the GCG Wisconsin Genetics Software Package, Version 10 (available from Accelrys Inc., 9685 Scranton Road, San Diego, Calif., USA). Alignments using these programs can be performed using the default parameters. The CLUSTAL program is well described by Higgins et al. (1988) *Gene* 73:237-244 (1988); Higgins et al. (1989) *CABIOS* 5:151-153; Corpet et al. (1988) *Nucleic Acids Res.* 16:10881-90; Huang et al. (1992) *CABIOS* 8:155-65; and Pearson et al. (1994) *Meth. Mol. Biol.* 24:307-331. The ALIGN program is based on the algorithm of Myers and Miller (1988) supra. A PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used with the ALIGN program when comparing amino acid sequences. The BLAST programs of Altschul et al (1990) *J. Mol. Biol.* 215:403 are based on the algorithm of Karlin and Altschul (1990) supra. BLAST nucleotide searches can be performed with the BLASTN program, score=100, wordlength=12, to obtain nucleotide sequences homologous to a nucleotide

sequence encoding a protein of the invention. BLAST protein searches can be performed with the BLASTX program, score=50, wordlength=3, to obtain amino acid sequences homologous to a protein or polypeptide of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul et al. (1997) *Nucleic Acids Res.* 25:3389. Alternatively, PSI-BLAST (in BLAST 2.0) can be used to perform an iterated search that detects distant relationships between molecules. See Altschul et al. (1997) *supra*. When utilizing BLAST, Gapped BLAST, PSI-BLAST, the default parameters of the respective programs (e.g., BLASTN for nucleotide sequences, BLASTX for proteins) can be used. See <http://www.ncbi.nlm.nih.gov>. Alignment may also be performed manually by inspection.

[0056] Unless otherwise stated, sequence identity/similarity values provided herein refer to the value obtained using GAP Version 10 using the following parameters: % identity and % similarity for a nucleotide sequence using GAP Weight of 50 and Length Weight of 3, and the nwsgapdna.cmp scoring matrix; % identity and % similarity for an amino acid sequence using GAP Weight of 8 and Length Weight of 2, and the BLOSUM62 scoring matrix; or any equivalent program thereof. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide or amino acid residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by GAP Version 10.

[0057] GAP uses the algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453, to find the alignment of two complete sequences that maximizes the number of matches and minimizes the number of gaps. GAP considers all possible alignments and gap positions and creates the alignment with the largest number of matched bases and the fewest gaps. It allows for the provision of a gap creation penalty and a gap extension penalty in units of matched bases. GAP must make a profit of gap creation penalty number of matches for each gap it inserts. If a gap extension penalty greater than zero is chosen, GAP must, in addition, make a profit for each gap inserted of the length of the gap times the gap extension penalty. Default gap creation penalty values and gap extension penalty values in Version 10 of the GCG Wisconsin Genetics Software Package for protein sequences are 8 and 2, respectively. For nucleotide sequences the default gap creation penalty is 50 while the default gap extension penalty is 3. The gap creation and gap extension penalties can be expressed as an integer selected from the group of integers consisting of from 0 to 200. Thus, for example, the gap creation and gap extension penalties can be 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65 or greater.

[0058] GAP presents one member of the family of best alignments. There may be many members of this family, but no other member has a better quality. GAP displays four figures of merit for alignments: Quality, Ratio, Identity, and Similarity. The Quality is the metric maximized in order to align the sequences. Ratio is the quality divided by the number of bases in the shorter segment. Percent Identity is the percent of the symbols that actually match. Percent Similarity is the percent of the symbols that are similar. Symbols that are across from gaps are ignored. A similarity is scored when the scoring matrix value for a pair of symbols is greater than or equal to 0.50, the similarity threshold. The scoring matrix used in Version 10 of the GCG Wisconsin Genetics Software

Package is BLOSUM62 (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

[0059] (c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences makes reference to the residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g., charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences that differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, Calif.).

[0060] (d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

[0061] (e)(i) The term "substantial identity" of polynucleotide sequences means that a polynucleotide comprises a sequence that has at least 70% sequence identity, optimally at least 80%, more optimally at least 90%, and most optimally at least 95%, compared to a reference sequence using one of the alignment programs described using standard parameters. One of skill in the art will recognize that these values can be appropriately adjusted to determine corresponding identity of proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning, and the like. Substantial identity of amino acid sequences for these purposes normally means sequence identity of at least 60%, more optimally at least 70%, 80%, 90%, and most optimally at least 95%.

[0062] Another indication that nucleotide sequences are substantially identical is if two molecules hybridize to each other under stringent conditions. Generally, stringent conditions are selected to be about 5° C. lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. However, stringent conditions encompass temperatures in the range of about 1° C. to about 20° C. lower than the T_m , depending upon the desired degree of stringency

as otherwise qualified herein. Nucleic acids that do not hybridize to each other under stringent conditions are still substantially identical if the polypeptides they encode are substantially identical. This may occur, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code. One indication that two nucleic acid sequences are substantially identical is when the polypeptide encoded by the first nucleic acid is immunologically cross reactive with the polypeptide encoded by the second nucleic acid.

[0063] (e)(ii) The term “substantial identity” in the context of a peptide indicates that a peptide comprises a sequence with at least 70% sequence identity to a reference sequence, optimally 80%, more optimally 85%, most optimally at least 90% or 95% sequence identity to the reference sequence over a specified comparison window. Optimally, optimal alignment is conducted using the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453. An indication that two peptide sequences are substantially identical is that one peptide is immunologically reactive with antibodies raised against the second peptide. Thus, a peptide is substantially identical to a second peptide, for example, where the two peptides differ only by a conservative substitution. Peptides that are “substantially similar” share sequences as noted above except that residue positions that are not identical may differ by conservative amino acid changes.

[0064] The invention further provides plants, plant cells, and plant parts having altered levels and/or activities of the ODP2 polypeptides of the invention. In some embodiments, the plants of the invention have stably incorporated the ODP2 sequences of the invention. As discussed elsewhere herein, altering the level/activity of the ODP2 sequences of the invention can produce a variety of phenotypes. As used herein, the term plant includes plant cells, plant protoplasts, plant cell tissue cultures from which plants can be regenerated, plant calli, plant clumps, and plant cells that are intact in plants or parts of plants such as embryos, pollen, ovules, seeds, leaves, flowers, branches, fruit, kernels, ears, cobs, husks, stalks, roots, root tips, anthers, grain and the like. As used herein “grain” is intended the mature seed produced by commercial growers for purposes other than growing or reproducing the species. Progeny, variants, and mutants of the regenerated plants are also included within the scope of the invention, provided that these parts comprise the introduced nucleic acid sequences.

[0065] A “subject plant or plant cell” is one in which an alteration, such as transformation or introduction of a polypeptide, has occurred, or is a plant or plant cell which is descended from a plant or cell so altered and which comprises the alteration. A “control” or “control plant” or “control plant cell” provides a reference point for measuring changes in phenotype of the subject plant or plant cell.

[0066] A control plant or plant cell may comprise, for example: (a) a wild-type plant or cell, i.e., of the same genotype as the starting material for the alteration which resulted in the subject plant or cell; (b) a plant or plant cell of the same genotype as the starting material but which has been transformed with a null construct (i.e. with a construct which has no known effect on the trait of interest, such as a construct comprising a marker gene); (c) a plant or plant cell which is a non-transformed segregant among progeny of a subject plant or plant cell; (d) a plant or plant cell genetically identical to the subject plant or plant cell but which is not exposed to

conditions or stimuli that would induce expression of the gene of interest; or (e) the subject plant or plant cell itself, under conditions in which the gene of interest is not expressed.

Methods

[0067] I. Providing Sequences

[0068] The use of the term “nucleotide constructs” or “polynucleotide” herein is not intended to limit the present invention to nucleotide constructs comprising DNA. Those of ordinary skill in the art will recognize that nucleotide constructs, particularly polynucleotides and oligonucleotides, comprised of ribonucleotides and combinations of ribonucleotides and deoxyribonucleotides may also be employed in the methods disclosed herein. Thus, the nucleotide constructs of the present invention encompass all nucleotide constructs that can be employed in the methods of the present invention for transforming plants including, but not limited to, those comprised of deoxyribonucleotides, ribonucleotides, and combinations thereof. Such deoxyribonucleotides and ribonucleotides include both naturally occurring molecules and synthetic analogues. The nucleotide constructs of the invention also encompass all forms of nucleotide constructs including, but not limited to, single-stranded forms, double-stranded forms, hairpins, stem-and-loop structures, and the like.

[0069] The nucleic acid sequences of the present invention can be introduced/expressed in a host cell such as bacteria, yeast, insect, mammalian, or optimally plant cells. It is expected that those of skill in the art are knowledgeable in the numerous systems available for the introduction of a polypeptide or a nucleotide sequence of the present invention. No attempt to describe in detail the various methods known for providing proteins in prokaryotes or eukaryotes will be made.

[0070] As used herein, “heterologous” in reference to a nucleic acid is a nucleic acid that originates from a foreign species, or, if from the same species, is substantially modified from its native form in composition and/or genomic locus by deliberate human intervention. For example, a promoter operably linked to a heterologous structural gene is from a species different from that from which the structural gene was derived, or, if from the same species, one or both are substantially modified from their original form and/or genomic location.

[0071] By “host cell” is meant a cell, which comprises a heterologous nucleic acid sequence of the invention. Host cells may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, insect, amphibian, or mammalian cells. Optimally, host cells are monocotyledonous or dicotyledonous plant cells. A particularly optimal monocotyledonous host cell is a maize host cell.

[0072] The ODP2 sequences of the invention can be provided in expression cassettes for expression in the plant of interest. The cassette can include 5' and 3' regulatory sequences operably linked to an ODP2 sequence of the invention. “Operably linked” is intended to mean a functional linkage between two or more elements. For example, an operable linkage between a polynucleotide of interest and a regulatory sequence (i.e., a promoter) is functional link that allows for expression of the polynucleotide of interest. Operably linked elements may be contiguous or non-contiguous. When used to refer to the joining of two protein coding regions, by operably linked is intended that the coding regions are in the same reading frame. The cassette may additionally contain at least one additional gene to be cotransformed into the organ-

ism. Alternatively, the additional gene(s) can be provided on multiple expression cassettes. Such an expression cassette is provided with a plurality of restriction sites for insertion of the ODP2 sequence to be under the transcriptional regulation of the regulatory regions. The expression cassette may additionally contain selectable marker genes.

[0073] The expression cassette can include in the 5'-3' direction of transcription, a transcriptional initiation region (i.e., a promoter) and translational initiation region, an ODP2 sequence of the invention, and a transcriptional and translational termination region (i.e., termination region) functional in plants. The promoter may be native/analogous or foreign to the plant host and/or to the ODP2 sequence of the invention. In one embodiment, the promoter employed in the methods of the invention is the native ODP2 promoter. See, for example, U.S. Provisional Application No. 60/541,171, entitled "ODP2 Promoter and Methods of Use", filed on Feb. 2, 2004. Additionally, the promoter may be a natural sequence or alternatively a synthetic sequence. Where the promoter is "foreign" to the plant host, it is intended that the promoter is not found in the native plant into which the promoter is introduced. Where the promoter is "foreign" to the ODP2 sequence of the invention, it is intended that the promoter is not the native or naturally occurring promoter for the operably linked ODP2 sequence of the invention. As used herein, a chimeric gene comprises a coding sequence operably linked to a transcription initiation region that is heterologous to the coding sequence.

[0074] While it may be optimal to express the sequences using foreign promoters, the native promoter sequences may be used. Such constructs would change expression levels of ODP2 in the plant or plant cell. Thus, the phenotype of the plant or plant cell can be altered.

[0075] The termination region may be native with the transcriptional initiation region, may be native with the operably linked ODP2 sequence of interest, may be native with the plant host, or may be derived from another source (i.e., foreign to the promoter, the ODP2 sequence of interest, the plant host, or any combination thereof). Convenient termination regions are available from the Ti-plasmid of *A. tumefaciens*, such as the octopine synthase and nopaline synthase termination regions. See also Guerineau et al. (1991) *Mol. Gen. Genet.* 262:141-144; Proudfoot (1991) *Cell* 64:671-674; Sanfacon et al. (1991) *Genes Dev.* 5:141-149; Mogen et al. (1990) *Plant Cell* 2:1261-1272; Munroe et al. (1990) *Gene* 91:151-158; Ballas et al. (1989) *Nucleic Acids Res.* 17:7891-7903; and Joshi et al. (1987) *Nucleic Acid Res.* 15:9627-9639.

[0076] Where appropriate, the gene(s) may be optimized for increased expression in the transformed plant. That is, the genes can be synthesized using plant-preferred codons for improved expression. See, for example, Campbell and Gowri (1990) *Plant Physiol.* 92:1-11 for a discussion of host-preferred codon usage. Methods are available in the art for synthesizing plant-preferred genes. See, for example, U.S. Pat. Nos. 5,380,831, and 5,436,391, and Murray et al. (1989) *Nucleic Acids Res.* 17:477-498, herein incorporated by reference.

[0077] Additional sequence modifications are known to enhance gene expression in a cellular host. These include elimination of sequences encoding spurious polyadenylation signals, exon-intron splice site signals, transposon-like repeats, and other such well-characterized sequences that may be deleterious to gene expression. The G-C content of the sequence may be adjusted to levels average for a given cellu-

lar host, as calculated by reference to known genes expressed in the host cell. When possible, the sequence is modified to avoid predicted hairpin secondary mRNA structures.

[0078] The expression cassettes may additionally contain 5' leader sequences in the expression cassette construct. Such leader sequences can act to enhance translation. Translation leaders are known in the art and include: picornavirus leaders, for example, EMCV leader (Encephalomyocarditis 5' non-coding region) (Elroy-Stein et al. (1989) *Proc. Natl. Acad. Sci. USA* 86:6126-6130); potyvirus leaders, for example, TEV leader (Tobacco Etch Virus) (Gallie et al. (1995) *Gene* 165(2):233-238), MDMV leader (Maize Dwarf Mosaic Virus), and human immunoglobulin heavy-chain binding protein (BiP) (Macejak et al. (1991) *Nature* 353:90-94); untranslated leader from the coat protein mRNA of alfalfa mosaic virus (AMV RNA 4) (Jobling et al. (1987) *Nature* 325:622-625); tobacco mosaic virus leader (TMV) (Gallie et al. (1989) in *Molecular Biology of RNA*, ed. Cech (Liss, New York), pp. 237-256); and maize chlorotic mottle virus leader (MCMV) (Lommel et al. (1991) *Virology* 81:382-385). See also, Della-Cioppa et al. (1987) *Plant Physiol.* 84:965-968.

[0079] In preparing the expression cassette, the various DNA fragments may be manipulated, so as to provide for the DNA sequences in the proper orientation and, as appropriate, in the proper reading frame. Toward this end, adapters or linkers may be employed to join the DNA fragments or other manipulations may be involved to provide for convenient restriction sites, removal of superfluous DNA, removal of restriction sites, or the like. For this purpose, in vitro mutagenesis, primer repair, restriction, annealing, resubstitutions, e.g., transitions and transversions, may be involved.

[0080] Generally, the expression cassette will comprise a selectable marker gene for the selection of transformed cells. Selectable marker genes are utilized for the selection of transformed cells or tissues. Marker genes include genes encoding antibiotic resistance, such as those encoding neomycin phosphotransferase II (NEO) and hygromycin phosphotransferase (HPT), as well as genes conferring resistance to herbicidal compounds, such as glufosinate ammonium, bromoxynil, imidazolinones, and 2,4-dichlorophenoxyacetate (2,4-D). See generally, Yarranton (1992) *Curr. Opin. Biotech.* 3:506-511; Christoperson et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:6314-6318; Yao et al. (1992) *Cell* 71:63-72; Reznikoff (1992) *Mol. Microbiol.* 6:2419-2422; Barkley et al. (1980) in *The Operon*, pp. 177-220; Hu et al. (1987) *Cell* 48:555-566; Brown et al. (1987) *Cell* 49:603-612; Figge et al. (1988) *Cell* 52:713-722; Deuschle et al. (1989) *Proc. Natl. Acad. Aci. USA* 86:5400-5404; Fuerst et al. (1989) *Proc. Natl. Acad. Sci. USA* 86:2549-2553; Deuschle et al. (1990) *Science* 248:480-483; Gossen (1993) Ph.D. Thesis, University of Heidelberg; Reines et al. (1993) *Proc. Natl. Acad. Sci. USA* 90:1917-1921; Labow et al. (1990) *Mol. Cell. Biol.* 10:3343-3356; Zambretti et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:3952-3956; Baim et al. (1991) *Proc. Natl. Acad. Sci. USA* 88:5072-5076; Wyborski et al. (1991) *Nucleic Acids Res.* 19:4647-4653; Hilleman-Wissman (1989) *Topics Mol. Struc. Biol.* 10:143-162; Degenkolb et al. (1991) *Antimicrob. Agents Chemother.* 35:1591-1595; Kleinschmidt et al. (1988) *Biochemistry* 27:1094-1104; Bonin (1993) Ph.D. Thesis, University of Heidelberg; Gossen et al. (1992) *Proc. Natl. Acad. Sci. USA* 89:5547-5551; Oliva et al. (1992) *Antimicrob. Agents Chemother.* 36:913-919; Hlavka et al. (1985) *Handbook of Experimental Pharmacology*, Vol. 78 (Springer-Verlag, Berlin); Gill et al. (1988) *Nature* 334:721-724. Such disclosures

are herein incorporated by reference. The above list of selectable marker genes is not meant to be limiting. Any selectable marker gene can be used in the present invention.

[0081] A number of promoters can be used in the practice of the invention. The promoters can be selected based on the desired outcome. That is, the nucleic acid can be combined with constitutive, tissue-preferred, developmentally regulated, or other promoters for expression in plants. Constitutive promoters include, for example, the core promoter of the Rsyn7 promoter and other constitutive promoters disclosed in WO 99/43838 and U.S. Pat. No. 6,072,050; the core CaMV 35S promoter (Odell et al. (1985) *Nature* 313:810-812); rice actin (McElroy et al. (1990) *Plant Cell* 2:163-171); ubiquitin (Christensen et al. (1989) *Plant Mol. Biol.* 12:619-632 and Christensen et al. (1992) *Plant Mol. Biol.* 18:675-689); pEMU (Last et al. (1991) *Theor. Appl. Genet.* 81:581-588); MAS (Velten et al. (1984) *EMBO J.* 3:2723-2730); ALS promoter (U.S. Pat. No. 5,659,026), and the like. Other constitutive promoters include, for example, U.S. Pat. Nos. 5,608,149; 5,608,144; 5,604,121; 5,569,597; 5,466,785; 5,399,680; 5,268,463; 5,608,142; and 6,177,611.

[0082] Chemical-regulated promoters can be used to modulate the expression of a gene in a plant through the application of an exogenous chemical regulator. Depending upon the objective, the promoter may be a chemical-inducible promoter, where application of the chemical induces gene expression, or a chemical-repressible promoter, where application of the chemical represses gene expression. Chemical-inducible promoters are known in the art and include, but are not limited to, the maize ln2-2 promoter, which is activated by benzenesulfonamide herbicide safeners, the maize GST promoter, which is activated by hydrophobic electrophilic compounds that are used as pre-emergent herbicides, and the tobacco PR-1a promoter, which is activated by salicylic acid. Other chemical-regulated promoters of interest include steroid-responsive promoters (see, for example, the glucocorticoid-inducible promoter in Schena et al. (1991) *Proc. Natl. Acad. Sci. USA* 88:10421-10425 and McNellis et al. (1998) *Plant J.* 14(2):247-257) and tetracycline-inducible and tetracycline-repressible promoters (see, for example, Gatz et al. (1991) *Mol. Gen. Genet.* 227:229-237, and U.S. Pat. Nos. 5,814,618 and 5,789,156), herein incorporated by reference.

[0083] Tissue-preferred promoters can be utilized to target enhanced ODP2 expression within a particular plant tissue. Tissue-preferred promoters include Yamamoto et al. (1997) *Plant J.* 12(2):255-265; Kawamata et al. (1997) *Plant Cell Physiol.* 38(7):792-803; Hansen et al. (1997) *Mol. Gen. Genet.* 254(3):337-343; Russell et al. (1997) *Transgenic Res.* 6(2):157-168; Rinehart et al. (1996) *Plant Physiol.* 112(3): 1331-1341; Van Camp et al. (1996) *Plant Physiol.* 112(2): 525-535; Canevascini et al. (1996) *Plant Physiol.* 112(2): 513-524; Yamamoto et al. (1994) *Plant Cell Physiol.* 35(5): 773-778; Lam (1994) *Results Probl. Cell Differ.* 20:181-196; Orozco et al. (1993) *Plant Mol. Biol.* 23(6):1129-1138; Matsuoka et al. (1993) *Proc. Natl. Acad. Sci. USA* 90(20):9586-9590; and Guevara-Garcia et al. (1993) *Plant J.* 4(3):495-505. Such promoters can be modified, if necessary, for weak expression.

[0084] "Seed-preferred" promoters include both "seed-specific" promoters (those promoters active during seed development such as promoters of seed storage proteins) as well as "seed-germinating" promoters (those promoters active during seed germination). See Thompson et al. (1989) *BioEssays* 10:108, herein incorporated by reference. Such

seed-preferred promoters include, but are not limited to, Cim1 (cytokinin-induced message); cZ19B1 (maize 19 kDa zein); and, milps (myo-inositol-1-phosphate synthase); (see WO 00/11177 and U.S. Pat. No. 6,225,529; herein incorporated by reference). Gamma-zein is another endosperm-specific promoter (Boronat et al. (1986) *Plant Science* 47:95-102). Globulin-1 (Glob-1) is a preferred embryo-specific promoter. For dicots, seed-specific promoters include, but are not limited to, bean β -phaseolin, napin, β -conglycinin, soybean lectin, cruciferin, and the like. For monocots, seed-specific promoters include, but are not limited to, maize 15 kDa, 22 kDa zein, 27 kDa zein, gamma-zein, waxy, shrunken 1, shrunken 2, globulin 1, etc. See also WO 00/12733, where seed-preferred promoters from end1 and end2 genes are disclosed; herein incorporated by reference. Additional seed-preferred promoters include the oleosin promoter (WO 00/028058), the lipid transfer protein (LTP) promoter (U.S. Pat. No. 5,525,716). Additional seed-preferred promoters include the Lec1 promoter, the Jip1 promoter, and the milp3 promoter (see, WO 02/42424).

[0085] The methods of the invention involve introducing a nucleotide construct or a polypeptide into a plant. By "introducing" is intended presenting to the plant the nucleotide construct (i.e., DNA or RNA) or a polypeptide in such a manner that the nucleic acid or the polypeptide gains access to the interior of a cell of the plant. The methods of the invention do not depend on a particular method for introducing the nucleotide construct or the polypeptide to a plant, only that the nucleotide construct gains access to the interior of at least one cell of the plant. Methods for introducing nucleotide constructs and/or polypeptide into plants are known in the art including, but not limited to, stable transformation methods, transient transformation methods, and virus-mediated methods.

[0086] By "stable transformation" is intended that the nucleotide construct introduced into a plant integrates into the genome of the plant and is capable of being inherited by progeny thereof. By "transient transformation" is intended that a nucleotide construct or the polypeptide introduced into a plant does not integrate into the genome of the plant.

[0087] Thus the ODP2 sequences of the invention can be provided to a plant using a variety of transient transformation methods including, but not limited to, the introduction of ODP2 protein or variants thereof directly into the plant and the introduction of the an ODP2 transcript into the plant. Such methods include, for example, microinjection or particle bombardment. See, for example, Crossway et al. (1986) *Mol. Gen. Genet.* 202:179-185; Nomura et al. (1986) *Plant Sci.* 44:53-58; Hepler et al. (1994) *Proc. Natl. Acad. Sci.* 91: 2176-2180 and Hush et al. (1994) *The Journal of Cell Science* 107:775-784, all of which are herein incorporated by reference. Alternatively, the various viral vector systems can be used for transient expression or the ODP2 nucleotide construct can be precipitated in a manner that precludes subsequent release of the DNA (thus, transcription from the particle-bound DNA can occur, but the frequency with which its released to become integrated into the genome is greatly reduced). Such methods include the use of PEI, as outlined in more detail in Example 13.

[0088] The nucleotide constructs of the invention may be introduced into plants by contacting plants with a virus or viral nucleic acids. Generally, such methods involve incorporating a nucleotide construct of the invention within a viral DNA or RNA molecule. It is recognized that the an ODP2

polypeptide of the invention may be initially synthesized as part of a viral polyprotein, which later may be processed by proteolysis in vivo or in vitro to produce the desired recombinant protein. Further, it is recognized that promoters of the invention also encompass promoters utilized for transcription by viral RNA polymerases. Methods for introducing nucleotide constructs into plants and expressing a protein encoded therein, involving viral DNA or RNA molecules, are known in the art. See, for example, U.S. Pat. Nos. 5,889,191, 5,889,190, 5,866,785, 5,589,367 and 5,316,931; herein incorporated by reference.

[0089] Transformation protocols as well as protocols for introducing nucleotide sequences into plants may vary depending on the type of plant or plant cell, i.e., monocot or dicot, targeted for transformation. Suitable methods of introducing nucleotide sequences into plant cells and subsequent insertion into the plant genome include microinjection (Crossway et al. (1986) *Biotechniques* 4:320-334), electroporation (Riggs et al. (1986) *Proc. Natl. Acad. Sci. USA* 83:5602-5606, *Agrobacterium*-mediated transformation (Townsend et al., U.S. Pat. No. 5,563,055; Zhao et al., U.S. Pat. No. 5,981,840), direct gene transfer (Paszkowski et al. (1984) *EMBO J.* 3:2717-2722), and ballistic particle acceleration (see, for example, Sanford et al., U.S. Pat. No. 4,945,050; Tomes et al., U.S. Pat. No. 5,879,918; Tomes et al., U.S. Pat. No. 5,886,244; Bidney et al., U.S. Pat. No. 5,932,782; Tomes et al. (1995) "Direct DNA Transfer into Intact Plant Cells via Microprojectile Bombardment," in *Plant Cell, Tissue, and Organ Culture: Fundamental Methods*, ed. Gamborg and Phillips (Springer-Verlag, Berlin); McCabe et al. (1988) *Biotechnology* 6:923-926); and Lec1 transformation (WO 00/28058). Also see Weissinger et al. (1988) *Ann. Rev. Genet.* 22:421-477; Sanford et al. (1987) *Particulate Science and Technology* 5:27-37 (onion); Christou et al. (1988) *Plant Physiol.* 87:671-674 (soybean); McCabe et al. (1988) *Biotechnology* 6:923-926 (soybean); Finer and McMullen (1991) *In Vitro Cell Dev. Biol.* 27P:175-182 (soybean); Singh et al. (1998) *Theor. Appl. Genet.* 96:319-324 (soybean); Datta et al. (1990) *Biotechnology* 8:736-740 (rice); Klein et al. (1988) *Proc. Natl. Acad. Sci. USA* 85:4305-4309 (maize); Klein et al. (1988) *Biotechnology* 6:559-563 (maize); Tomes, U.S. Pat. No. 5,240,855; Buisling et al., U.S. Pat. Nos. 5,322,783 and 5,324,646; Tomes et al. (1995) "Direct DNA Transfer into Intact Plant Cells via Microprojectile Bombardment," in *Plant Cell, Tissue, and Organ Culture: Fundamental Methods*, ed. Gamborg (Springer-Verlag, Berlin) (maize); Klein et al. (1988) *Plant Physiol.* 91:440-444 (maize); Fromm et al. (1990) *Biotechnology* 8:833-839 (maize); Hooykaas-Van Slogteren et al. (1984) *Nature* (London) 311:763-764; Bowen et al., U.S. Pat. No. 5,736,369 (cereals); Bytebier et al. (1987) *Proc. Natl. Acad. Sci. USA* 84:5345-5349 (Liliaceae); De Wet et al. (1985) in *The Experimental Manipulation of Ovule Tissues*, ed. Chapman et al. (Longman, New York), pp. 197-209 (pollen); Kaeppler et al. (1990) *Plant Cell Reports* 9:415-418 and Kaeppler et al. (1992) *Theor. Appl. Genet.* 84:560-566 (whisker-mediated transformation); D'Halluin et al. (1992) *Plant Cell* 4:1495-1505 (electroporation); Li et al. (1993) *Plant Cell Reports* 12:250-255 and Christou and Ford (1995) *Annals of Botany* 75:407-413 (rice); Osjoda et al. (1996) *Nature Biotechnology* 14:745-750 (maize via *Agrobacterium tumefaciens*); all of which are herein incorporated by reference.

[0090] Methods are known in the art for the targeted insertion of a polynucleotide at a specific location in the plant

genome. In one embodiment, the insertion of the polynucleotide at a desired genomic location is achieved using a site-specific recombination system. See, for example, WO99/25821, WO99/25854, WO99/25840, WO99/25855, and WO99/25853, all of which are herein incorporated by reference. Briefly, the polynucleotide of the invention can be contained in transfer cassette flanked by two non-recombinogenic recombination sites. The transfer cassette is introduced into a plant having stably incorporated into its genome a target site which is flanked by two non-recombinogenic recombination sites that correspond to the sites of the transfer cassette. An appropriate recombinase is provided and the transfer cassette is integrated at the target site. The polynucleotide of interest is thereby integrated at a specific chromosomal position in the plant genome.

[0091] The cells that have been transformed may be grown into plants in accordance with conventional ways. See, for example, McCormick et al. (1986) *Plant Cell Reports* 5:81-84. These plants may then be grown, and either pollinated with the same transformed strain or different strains, and the resulting hybrid having constitutive expression of the desired phenotypic characteristic identified. Two or more generations may be grown to ensure that expression of the desired phenotypic characteristic is stably maintained and inherited and then seeds harvested to ensure expression of the desired phenotypic characteristic has been achieved. In this manner, the present invention provides transformed seed (also referred to as "transgenic seed") having a nucleotide construct of the invention, for example, an expression cassette of the invention, stably incorporated into their genome.

[0092] The present invention may be used for transformation of any plant species, including, but not limited to, monocots and dicots. Examples of plant species of interest include, but are not limited to, corn (*Zea mays*), *Brassica* sp. (e.g., *B. napus*, *B. rapa*, *B. juncea*), particularly those *Brassica* species useful as sources of seed oil, alfalfa (*Medicago sativa*), rice (*Oryza sativa*), rye (*Secale cereale*), sorghum (*Sorghum bicolor*, *Sorghum vulgare*), millet (e.g., pearl millet (*Pennisetum glaucum*), proso millet (*Panicum miliaceum*), foxtail millet (*Setaria italica*), finger millet (*Eleusine coracana*)), sunflower (*Helianthus annuus*), safflower (*Carthamus tinctorius*), wheat (*Triticum aestivum*), soybean (*Glycine max*), tobacco (*Nicotiana tabacum*), potato (*Solanum tuberosum*), peanuts (*Arachis hypogaea*), cotton (*Gossypium barbadense*, *Gossypium hirsutum*), sweet potato (*Ipomoea batatas*), cassava (*Manihot esculenta*), coffee (*Coffea* spp.), coconut (*Cocos nucifera*), pineapple (*Ananas comosus*), citrus trees (*Citrus* spp.), cocoa (*Theobroma cacao*), tea (*Camellia sinensis*), banana (*Musa* spp.), avocado (*Persea americana*), fig (*Ficus carica*), guava (*Psidium guajava*), mango (*Mangifera indica*), olive (*Olea europaea*), papaya (*Carica papaya*), cashew (*Anacardium occidentale*), macadamia (*Macadamia integrifolia*), almond (*Prunus amygdalus*), sugar beets (*Beta vulgaris*), sugarcane (*Saccharum* spp.), oats, barley, vegetables, ornamentals, and conifers.

[0093] Vegetables include tomatoes (*Lycopersicon esculentum*), lettuce (e.g., *Lactuca sativa*), green beans (*Phaseolus vulgaris*), lima beans (*Phaseolus limensis*), peas (*Lathyrus* spp.), and members of the genus *Cucumis* such as cucumber (*C. sativus*), cantaloupe (*C. cantalupensis*), and musk melon (*C. melo*). Ornamentals include azalea (*Rhododendron* spp.), hydrangea (*Macrophylla hydrangea*), hibiscus (*Hibiscus rosasanensis*), roses (*Rosa* spp.), tulips (*Tulipa* spp.), daffodils (*Narcissus* spp.), petunias (*Petunia hybrida*),

carnation (*Dianthus caryophyllus*), poinsettia (*Euphorbia pulcherrima*), and chrysanthemum.

[0094] Conifers that may be employed in practicing the present invention include, for example, pines such as loblolly pine (*Pinus taeda*), slash pine (*Pinus elliottii*), ponderosa pine (*Pinus ponderosa*), lodgepole pine (*Pinus contorta*), and Monterey pine (*Pinus radiata*); Douglas-fir (*Pseudotsuga menziesii*); Western hemlock (*Tsuga canadensis*); Sitka spruce (*Picea glauca*); redwood (*Sequoia sempervirens*); true firs such as silver fir (*Abies amabilis*) and balsam fir (*Abies balsamea*); and cedars such as Western red cedar (*Thuja plicata*) and Alaska yellow-cedar (*Chamaecyparis nootkatensis*). Optimally, plants of the present invention are crop plants (for example, corn, alfalfa, sunflower, *Brassica*, soybean, cotton, safflower, peanut, sorghum, wheat, millet, tobacco, etc.), more optimally corn and soybean plants, yet more optimally corn plants.

[0095] Plants of particular interest include grain plants that provide seeds of interest, oil-seed plants, and leguminous plants. Seeds of interest include grain seeds, such as corn, wheat, barley, rice, sorghum, rye, etc. Oil-seed plants include cotton, soybean, safflower, sunflower, *Brassica*, maize, alfalfa, palm, coconut, etc. Leguminous plants include beans and peas. Beans include guar, locust bean, fenugreek, soybean, garden beans, cowpea, mungbean, lima bean, fava bean, lentils, chickpea, etc.

[0096] Typically, an intermediate host cell will be used in the practice of this invention to increase the copy number of the cloning vector. With an increased copy number, the vector containing the nucleic acid of interest can be isolated in significant quantities for introduction into the desired plant cells. In one embodiment, plant promoters that do not cause expression of the polypeptide in bacteria are employed.

[0097] Prokaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be used. Commonly used prokaryotic control sequences which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding sequences, include such commonly used promoters as the beta lactamase (penicillinase) and lactose (lac) promoter systems (Chang et al. (1977) *Nature* 198: 1056), the tryptophan (trp) promoter system (Goeddel et al. (1980) *Nucleic Acids Res.* 8:4057) and the lambda derived P L promoter and N-gene ribosome binding site (Shimatake et al. (1981) *Nature* 292:128). The inclusion of selection markers in DNA vectors transfected in *E. coli*. is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol.

[0098] The vector is selected to allow introduction into the appropriate host cell. Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA. Expression systems for expressing a protein of the present invention are available using *Bacillus* sp. and *Salmonella* (Palva et al. (1983) *Gene* 22:229-235); Mosbach et al. (1983) *Nature* 302: 543-545).

[0099] A variety of eukaryotic expression systems such as yeast, insect cell lines, plant and mammalian cells, are known to those of skill in the art. As explained briefly below, a polynucleotide of the present invention can be expressed in these eukaryotic systems. In some embodiments, transformed/transfected plant cells, as discussed infra, are

employed as expression systems for production of the proteins of the instant invention. Synthesis of heterologous polynucleotides in yeast is well known (Sherman et al. (1982) *Methods in Yeast Genetics*, Cold Spring Harbor Laboratory). Two widely utilized yeasts for production of eukaryotic proteins are *Saccharomyces cerevisiae* and *Pichia pastoris*. Vectors, strains, and protocols for expression in *Saccharomyces* and *Pichia* are known in the art and available from commercial suppliers (e.g., Invitrogen). Suitable vectors usually have expression control sequences, such as promoters, including 3-phosphoglycerate kinase or alcohol oxidase, and an origin of replication, termination sequences and the like as desired.

[0100] A protein of the present invention, once expressed, can be isolated from yeast by lysing the cells and applying standard protein isolation techniques to the lists. The monitoring of the purification process can be accomplished by using Western blot techniques or radioimmunoassay of other standard immunoassay techniques.

[0101] The sequences of the present invention can also be ligated to various expression vectors for use in transfecting cell cultures of, for instance, mammalian, insect, or plant origin. Illustrative cell cultures useful for the production of the peptides are mammalian cells. A number of suitable host cell lines capable of expressing intact proteins have been developed in the art, and include the HEK293, BHK21, and CHO cell lines. Expression vectors for these cells can include expression control sequences, such as an origin of replication, a promoter (e.g. the CMV promoter, a HSV tk promoter or pgk (phosphoglycerate kinase) promoter), an enhancer (Queen et al. (1986) *Immunol. Rev.* 89:49), and necessary processing information sites, such as ribosome binding sites, RNA splice sites, polyadenylation sites (e.g., an SV40 large T Ag poly A addition site), and transcriptional terminator sequences. Other animal cells useful for production of proteins of the present invention are available, for instance, from the American Type Culture Collection.

[0102] Appropriate vectors for expressing proteins of the present invention in insect cells are usually derived from the SF9 baculovirus. Suitable insect cell lines include mosquito larvae, silkworm, armyworm, moth and *Drosophila* cell lines such as a Schneider cell line (See, Schneider (1987) *J. Embryol. Exp. Morphol.* 27:353-365).

[0103] As with yeast, when higher animal or plant host cells are employed, polyadenylation or transcription terminator sequences are typically incorporated into the vector. An example of a terminator sequence is the polyadenylation sequence from the bovine growth hormone gene. Sequences for accurate splicing of the transcript may also be included. An example of a splicing sequence is the VP 1 intron from SV40 (Sprague et al. (1983) *J. Virol.* 45:773-781). Additionally, gene sequences to control replication in the host cell may be incorporated into the vector such as those found in bovine papilloma virus type-vectors (Saveria-Campo (1985) *DNA Cloning Vol. II a Practical Approach*, D. M. Glover, Ed., IRL Press, Arlington, Va., pp. 213-238).

[0104] Animal and lower eukaryotic (e.g., yeast) host cells are competent or rendered competent for transfection by various means. There are several well-known methods of introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextrin, electroporation, biolistics, and micro-injection of the DNA directly into the cells. The transfected cells are cultured by

means well known in the art (Kuchler (1997) *Biochemical Methods in Cell Culture and Virology*, Dowden, Hutchinson and Ross, Inc.).

[0105] In some embodiments, the content and/or composition of polypeptides of the present invention in a plant may be modulated by altering, *in vivo* or *in vitro*, the promoter of a gene to up- or down-regulate gene expression. In some embodiments, the coding regions of native genes of the present invention can be altered via substitution, addition, insertion, or deletion to decrease activity of the encoded enzyme. See, e.g., Kmiec, U.S. Pat. No. 5,565,350; Zarling et al., PCT/US93/03868. In other embodiments, the polypeptide of the invention is introduced. And in some embodiments, an isolated nucleic acid (e.g., a vector) comprising a promoter sequence is transfected into a plant cell. Subsequently, a plant cell comprising the promoter operably linked to a polynucleotide of the present invention is selected for by means known to those of skill in the art such as, but not limited to, Southern blot, DNA sequencing, or PCR analysis using primers specific to the promoter and to the gene and detecting amplicons produced therefrom. A plant or plant part altered or modified by the foregoing embodiments is grown under plant forming conditions for a time sufficient to modulate the concentration and/or composition of polypeptides of the present invention in the plant. Plant forming conditions are well known in the art and discussed briefly, *supra*.

[0106] A method for modulating the concentration and/or activity of the polypeptide of the present invention is provided. By “modulation” is intended any alteration in the level and/or activity (i.e., increase or decrease) that is statistically significant compared to a control plant or plant part. In general, concentration, composition or activity is increased or decreased by at least 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, or 90% relative to a control plant, plant part, or cell. The modulation may occur during and/or subsequent to growth of the plant to the desired stage of development. Modulating nucleic acid expression temporally and/or in particular tissues can be controlled by employing the appropriate promoter operably linked to a polynucleotide of the present invention in, for example, sense or antisense orientation as discussed in greater detail, *supra*. Induction of expression of a polynucleotide of the present invention can also be controlled by exogenous administration of an effective amount of inducing compound. Inducible promoters and inducing compounds, which activate expression from these promoters, are well known in the art. In specific embodiments, the polypeptides of the present invention are modulated in monocots, particularly maize.

[0107] The level of the ODP2 polypeptide may be measured directly, for example, by assaying for the level of the ODP2 polypeptide in the plant, or indirectly, for example, by measuring the ODP2 activity of the ODP2 polypeptide in the plant. Methods for determining the presence of ODP2 activity are described elsewhere herein.

[0108] In specific embodiments, the polypeptide or the polynucleotide of the invention is introduced into the plant cell. Subsequently, a plant cell having the introduced sequence of the invention is selected using methods known to those of skill in the art such as, but not limited to, Southern blot analysis, DNA sequencing, PCR analysis, or phenotypic analysis. A plant or plant part altered or modified by the foregoing embodiments is grown under plant forming conditions for a time sufficient to modulate the concentration and/or activity of polypeptides of the present invention in the

plant. Plant forming conditions are well known in the art and discussed briefly elsewhere herein.

[0109] It is also recognized that the level and/or activity of the polypeptide may be modulated by employing a polynucleotide that is not capable of directing, in a transformed plant, the expression of a protein or an RNA. For example, the polynucleotides of the invention may be used to design polynucleotide constructs that can be employed in methods for altering or mutating a genomic nucleotide sequence in an organism. Such polynucleotide constructs include, but are not limited to, RNA:DNA vectors, RNA:DNA mutational vectors, RNA:DNA repair vectors, mixed-duplex oligonucleotides, self-complementary RNA:DNA oligonucleotides, and recombinogenic oligonucleobases. Such nucleotide constructs and methods of use are known in the art. See, U.S. Pat. Nos. 5,565,350; 5,731,181; 5,756,325; 5,760,012; 5,795,972; and 5,871,984; all of which are herein incorporated by reference. See also, WO 98/49350, WO 99/07865, WO 99/25821, and Beetham et al. (1999) *Proc. Natl. Acad. Sci. USA* 96:8774-8778; herein incorporated by reference.

[0110] It is therefore recognized that methods of the present invention do not depend on the incorporation of the entire polynucleotide into the genome, only that the plant or cell thereof is altered as a result of the introduction of the polynucleotide into a cell. In one embodiment of the invention, the genome may be altered following the introduction of the polynucleotide into a cell. For example, the polynucleotide, or any part thereof, may incorporate into the genome of the plant. Alterations to the genome of the present invention include, but are not limited to, additions, deletions, and substitutions of nucleotides into the genome. While the methods of the present invention do not depend on additions, deletions, and substitutions of any particular number of nucleotides, it is recognized that such additions, deletions, or substitutions comprises at least one nucleotide.

[0111] In some embodiments, the activity and/or level of the ODP2 polypeptide of the invention is increased. An increase in the level or activity of the ODP2 polypeptide of the invention can be achieved by providing to the plant an ODP2 polypeptide. As discussed elsewhere herein, many methods are known the art for providing a polypeptide to a plant including, but not limited to, direct introduction of the polypeptide into the plant and/or introducing into the plant (transiently or stably) a nucleotide construct encoding a polypeptide having ODP2 activity. In other embodiments, the level or activity of an ODP2 polypeptide may be increased by altering the gene encoding the ODP2 polypeptide or its promoter. See, e.g. U.S. Pat. No. 5,565,350 and PCT/US93/03868. The invention therefore encompasses mutagenized plants that carry mutations in ODP2 genes, where the mutations increase expression of the ODP2 gene or increase the ODP2 activity of the encoded ODP2 polypeptide.

[0112] In some embodiments, the activity and/or level of the ODP2 polypeptide of the invention is reduced or eliminated by introducing into a plant a polynucleotide that inhibits the level or activity of the ODP2 polypeptide of the invention. The polynucleotide may inhibit the expression of ODP2 directly, by preventing translation of the ODP2 messenger RNA, or indirectly, by encoding a polypeptide that inhibits the transcription or translation of an ODP2 gene encoding an ODP2 protein. Methods for inhibiting or eliminating the expression of a gene in a plant are well known in the art, and any such method may be used in the present invention to inhibit the expression of ODP2 in a plant. In other embodi-

ments of the invention, the activity of ODP2 polypeptide is reduced or eliminated by transforming a plant cell with an expression cassette comprising a polynucleotide encoding a polypeptide that inhibits the activity of the ODP2 polypeptide. In other embodiments, the activity of an ODP2 polypeptide may be reduced or eliminated by disrupting the gene encoding the ODP2 polypeptide. The invention encompasses mutagenized plants that carry mutations in ODP2 genes, where the mutations reduce expression of the ODP2 gene or inhibit the ODP2 activity of the encoded ODP2 polypeptide.

[0113] Reduction of the activity of specific genes (also known as gene silencing or gene suppression) is desirable for several aspects of genetic engineering in plants. Methods for inhibiting gene expression are well known in the art and include, but are not limited to, homology-dependent gene silencing, antisense technology, RNA interference (RNAi), and the like. The general term homology-dependent gene silencing encompasses the phenomenon of *cis*-inactivation, *trans*-inactivation, and cosuppression. See Finnegan et al. (1994) *Biotech.* 12:883-888; and Matzke et al. (1995) *Plant Physiol.* 107:679-685; both incorporated herein in their entirety by reference. These mechanisms represent cases of gene silencing that involve transgene/transgene or transgene/endogenous gene interactions that lead to reduced expression of protein in plants. A "transgene" is a recombinant DNA construct that has been introduced into the genome by a transformation procedure. As one alternative, incorporation of antisense RNA into plants can be used to inhibit the expression of endogenous genes and produce a functional mutation within the genome. The effect is achieved by introducing into the cell(s) DNA that encodes RNA that is complementary to the sequence of mRNA of the target gene. See e.g. Bird et al. (1991) *Biotech and Gen. Eng. Rev.* 9:207-226; incorporated herein in its entirety by reference. See also the more detailed discussion herein below addressing these and other methodologies for achieving inhibition of expression or function of a gene.

[0114] Many techniques for gene silencing are well known to one of skill in the art, including, but not limited to, antisense technology (see, e.g., Sheehy et al. (1988) *Proc. Natl. Acad. Sci. USA* 85:8805-8809; and U.S. Pat. Nos. 5,107,065; 5,453,566; and 5,759,829); cosuppression (e.g., Taylor (1997) *Plant Cell* 9:1245; Jorgensen (1990) *Trends Biotech.* 8(12):340-344; Flavell (1994) *Proc. Natl. Acad. Sci. USA* 91:3490-3496; Finnegan et al. (1994) *Bio/Technology* 12:883-888; and Neuhuber et al. (1994) *Mol. Gen. Genet.* 244:230-241); RNA interference (Napoli et al. (1990) *Plant Cell* 2:279-289; U.S. Pat. No. 5,034,323; Sharp (1999) *Genes Dev.* 13:139-141; Zamore et al. (2000) *Cell* 101:25-33; and Montgomery et al. (1998) *Proc. Natl. Acad. Sci. USA* 95:15502-15507), virus-induced gene silencing (Burton et al. (2000) *Plant Cell* 12:691-705; and Baulcombe (1999) *Curr. Op. Plant Biol.* 2:109-113); target-RNA-specific ribozymes (Haseloff et al. (1988) *Nature* 334: 585-591); hairpin structures (Smith et al. (2000) *Nature* 407:319-320; WO 99/53050; WO 02/00904; WO 98/53083; Chuang and Meyerowitz (2000) *Proc. Natl. Acad. Sci. USA* 97:4985-4990; Stoutjesdijk et al. (2002) *Plant Physiol.* 129:1723-1731; Waterhouse and Helliwell (2003) *Nat. Rev. Genet.* 4:29-38; Pandolfini et al. *BMC Biotechnology* 3:7, U.S. Patent Publication No. 20030175965; Panstruga et al. (2003) *Mol. Biol. Rep.* 30:135-140; Wesley et al. (2001) *Plant J.* 27:581-590; Wang and Waterhouse (2001) *Curr. Opin. Plant Biol.* 5:146-150; U.S. Patent Publication No. 20030180945; and, WO 02/00904, all of which are herein

incorporated by reference); ribozymes (Steinecke et al. (1992) *EMBO J.* 11: 1525; and Perriman et al. (1993) *Antisense Res. Dev.* 3:253); oligonucleotide-mediated targeted modification (e.g., WO 03/076574 and WO 99/25853); Zn-finger targeted molecules (e.g., WO 01/52620; WO 03/048345; and WO 00/42219); transposon tagging (Maes et al. (1999) *Trends Plant Sci.* 4:90-96; Dharmapuri and Sonti (1999) *FEMS Microbiol. Lett.* 179:53-59; Meissner et al. (2000) *Plant J.* 22:265-274; Phogat et al. (2000) *J. Biosci.* 25:57-63; Walbot (2000) *Curr. Opin. Plant Biol.* 2:103-107; Gai et al. (2000) *Nucleic Acids Res.* 28:94-96; Fitzmaurice et al. (1999) *Genetics* 153:1919-1928; Bensen et al. (1995) *Plant Cell* 7:75-84; Mena et al. (1996) *Science* 274:1537-1540; and U.S. Pat. No. 5,962,764); each of which is herein incorporated by reference; and other methods or combinations of the above methods known to those of skill in the art.

[0115] It is recognized that with the polynucleotides of the invention, antisense constructions, complementary to at least a portion of the messenger RNA (mRNA) for the ODP2 sequences can be constructed. Antisense nucleotides are constructed to hybridize with the corresponding mRNA. Modifications of the antisense sequences may be made as long as the sequences hybridize to and interfere with expression of the corresponding mRNA. In this manner, antisense constructions having 70%, optimally 80%, more optimally 85% sequence identity to the corresponding antisensed sequences may be used. Furthermore, portions of the antisense nucleotides may be used to disrupt the expression of the target gene. Generally, sequences of at least 50 nucleotides, 100 nucleotides, 200 nucleotides, 300, 400, 450, 500, 550, or greater may be used.

[0116] The polynucleotides of the present invention may also be used in the sense orientation to suppress the expression of endogenous genes in plants. Methods for suppressing gene expression in plants using polynucleotides in the sense orientation are known in the art. The methods generally involve transforming plants with a DNA construct comprising a promoter that drives expression in a plant operably linked to at least a portion of a polynucleotide that corresponds to the transcript of the endogenous gene. Typically, such a nucleotide sequence has substantial sequence identity to the sequence of the transcript of the endogenous gene, optimally greater than about 65% sequence identity, more optimally greater than about 85% sequence identity, most optimally greater than about 95% sequence identity. See, U.S. Pat. Nos. 5,283,184 and 5,034,323; herein incorporated by reference. Thus, many methods may be used to reduce or eliminate the activity of an ODP2 polypeptide. More than one method may be used to reduce the activity of a single ODP2 polypeptide. In addition, combinations of methods may be employed to reduce or eliminate the activity of the ODP2 polypeptides.

[0117] Furthermore, it is recognized that the methods of the invention may employ a nucleotide construct that is capable of directing, in a transformed plant, the expression of at least one protein, or at least one RNA, such as, for example, an antisense RNA that is complementary to at least a portion of an mRNA. Typically such a nucleotide construct is comprised of a coding sequence for a protein or an RNA operably linked to 5' and 3' transcriptional regulatory regions. Alternatively, it is also recognized that the methods of the invention may employ a nucleotide construct that is not capable of directing, in a transformed plant, the expression of a protein or an RNA.

[0118] The ODP2 polynucleotides of the present invention can also be combined with genes implicated in transcriptional regulation, homeotic gene regulation, stem cell maintenance and proliferation, cell division, and/or cell differentiation such as other ODP2 homologues; Wuschel (see, e.g., Mayer et al. (1998) *Cell* 95:805-815); clavata (e.g., CLV1, CLV2, CLV3) (see, e.g., WO 03/093450; Clark et al. (1997) *Cell* 89:575-585; Jeong et al. (1999) *Plant Cell* 11:1925-1934; Fletcher et al. (1999) *Science* 283:1911-1914); Clavata and Embryo Surround region genes (e.g., CLE) (see, e.g., Sharma et al. (2003) *Plant Mol. Biol.* 51:415-425; Hobe et al. (2003) *Dev Genes Evol* 213:371-381; Cock & McCormick (2001) *Plant Physiol* 126:939-942; and Casamitjana-Martinez et al. (2003) *Curr Biol* 13:1435-1441); baby boom (e.g., BNM3, BBM) (see, e.g., WO 00/75530; Boutiller et al. (2002) *Plant Cell* 14:1737-1749); Zwillie (Lynn et al. (1999) *Dev* 126:469-481); leafy cotyledon (e.g., Lec1, Lec2) (see, e.g., Lotan et al. (1998) *Cell* 93:1195-1205; WO 00/28058; Stone et al. (2001) *PNAS* 98:11806-11811; and U.S. Pat. No. 6,492,577); Shoot Meristem-less (STM) (Long et al. (1996) *Nature* 379:66-69); ultrapetala (ULT) (see, e.g., Fletcher (2001) *Dev* 128:1323-1333); mitogen activated protein kinase (MAPK) (see, e.g., Jonak et al. (2002) *Curr Opin Plant Biol* 5:415); kinase associated protein phosphatase (KAPP) (see, e.g., Williams et al. (1997) *PNAS* 94:10467-10472; and Trotochaud et al. (1999) *Plant Cell* 11:393-406); ROP GTPase (see, e.g., Wu et al. (2001) *Plant Cell* 13:2841-2856; and Trotochaud et al. (1999) *Plant Cell* 11:393-406); fasciata (e.g., FAS1, FAS2) (see, e.g., Kaya et al. (2001) *Cell* 104:131-142); cell cycle genes (see, e.g., U.S. Pat. No. 6,518,487; WO 99/61619; and WO 02/074909); Shepherd (SHD) (see, e.g., Ishiguro et al. (2002) *EMBO J.* 21:898-908); Poltergeist (see, e.g., Yu et al. (2000) *Dev* 127:1661-1670; Yu et al. (2003) *Curr Biol* 13:179-188); Pickle (PKL) (see, e.g., Ogas et al. (1999) *PNAS* 96:13839-13844); knox genes (e.g., KN1, KNAT1) (see, e.g., Jackson et al. (1994) *Dev* 120:405-413; Lincoln et al. (1994) *Plant Cell* 6:1859-1876; Venglat et al. (2002) *PNAS* 99:4730-4735); fertilization independent endosperm (FIE) (e.g., Ohad et al. (1999) *Plant Cell* 11:407-415), and the like, the disclosures of which are herein incorporated by reference. The combinations generated can also include multiple copies of any one of the polynucleotides of interest. The combinations may have any combination of up-regulating and down-regulating expression of the combined polynucleotides. The combinations may or may not be combined on one construct for transformation of the host cell, and therefore may be provided sequentially or simultaneously. The host cell may be a wild-type or mutant cell, in a normal or aneuploid state.

[0119] II. Altering the Oil Content in Plants

[0120] The present invention provides a method for altering the oil content of a plant. By “altering the oil phenotype” of a plant is intended any modulation (increase or decrease) in the overall level of oil in the plant or plant part (i.e., seed) when compared to a control plant. The altered oil phenotype can comprise any statistically significant increase or decrease in oil when compared to a control plant. For example, altering the oil phenotype can comprise either an increase or a decrease in overall oil content of about 0.1%, 0.5%, 1%, 3% 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, or greater when compared to a control plant or plant part that has not been transformed with the ODP2 sequence of the invention. Alternatively, the alteration in oil phenotype can include about a 0.5 fold, 1 fold, 2 fold, 4 fold, 8 fold, 16 fold, or 32 fold

increase in overall oil phenotype in the plant or plant part when compared to a control plant that has not been transformed with the ODP2 sequence.

[0121] It is further recognized that the alteration in the oil phenotype need not be an overall increase/decrease in oil content, but also includes a change in the ratio of various components of the plant oil (i.e., a change in the ratio of any of the various fatty acids that compose the plant oil). For example, the ratio of various fatty acids such as linoleic acid, oleic acid, palmitic acid, stearic acid, myristic acid, linolenic acid, lauric acid, and the like, could be altered and thereby change the oil phenotype of the plant or plant part when compared to a control plant lacking the ODP2 sequence of the invention.

[0122] The method for altering the oil phenotype of a plant comprises providing an ODP2 sequence of the invention. An ODP2 polypeptide can be provided by introducing the polypeptide into the plant, and thereby modifying the oil content of the plant or plant part. Alternatively, an ODP2 nucleotide sequence can be provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 nucleotide sequence of the invention, expressing the ODP2 sequence, and thereby modifying the oil content of the plant. In yet other embodiments, the ODP2 nucleotide construct introduced into the plant is stably incorporated into the genome of the plant.

[0123] Methods for determining if the oil phenotype of the plant has been altered are known in the art. For example, the oil phenotype can be determined using NMR. Briefly, data for plant or plant part oil percentage, total plant or plant part oil, and plant or plant part weight are collected and analyzed by NMR. If changes from the control (a plant not transformed with ODP2) are observed above base-line, a PCR co-segregation analysis can be performed to determine if the changes are correlated with the presence of the ODP2 sequence. In specific embodiments, the plant part is an embryo. Alternatively, fatty acid content and composition can be determined by gas chromatography (GC). See, for example, WO 03/001902, herein incorporated by reference.

[0124] As discussed above, one of skill will recognize the appropriate promoter to use to alter the oil content of the plant in the desired manner. Exemplary promoters for this embodiment include the ubiquitin promoter (Christensen et al. (1992) *Plant Molecular Biology* 18:675-680), a lipid transfer protein (LTP) promoter (U.S. Pat. No. 5,525,716), a gamma-zein promoter (GZP) (Boronat et al. (1986) *Plant Sciences* 47:95-102), and the oleosin promoter (WO 00/28058), the lec1 promoter (WO 02/42424), and the Zm-ODP2 promoter (U.S. Provisional Application No. 60/541,171, entitled “ODP2 Promoter and Methods of Use” filed on Feb. 2, 2004, herein incorporated by reference in its entirety).

[0125] In specific embodiments, the oil content of the plant is decreased upon increasing level/activity of the ODP2 polypeptide in a plant. A decreased oil content finds use in the wet milling industry and in the ethanol dry grind industry. In the dry grind process, raw corn is ground, mixed with water, cooked, saccharified, fermented, and then distilled to make ethanol. The process also recovers distillers dried gains with solubles that can be used in feed products. Various methods of ethanol dry grind are known in the art. See, for example, U.S. Pat. No. 6,592,921, U.S. Pat. No. 6,433,146, Taylor et al. (2003) *Applied Biochemistry and Biotechnology* 104:141-148; Taylor et al. (2000) *Biotechnol Prog.* 16:541-7, and Taylor et al. (2001) *Appl Biochem Biotechnol* 94:41-9.

[0126] In the wet milling process, the purpose is to fractionate the kernel and isolate chemical constituents of economic value into their component parts. The process allows for the fractionation of starch into a highly purified form, as well as, for the isolation in crude forms of other material including, for example, unrefined oil, or as a wide mix of materials which commonly receive little to no additional processing beyond drying. Hence, in the wet milling process grain is softened by steeping and cracked by grinding to release the germ from the kernels. The germ is separated from the heavier density mixture of starch, hulls and fiber by “floating” the germ segments free of the other substances in a centrifugation process. This allows a clean separation of the oil-bearing fraction of the grain from tissue fragments that contain the bulk of the starch. Since it is not economical to extract oil on a small scale, many wet milling plants ship their germ to large, centralized oil production facilities. Oil is expelled or extracted with solvents from dried germs and the remaining germ meal is commonly mixed into corn gluten feed (CGF), a coproduct of wet milling. Hence, starch contained within the germ is not recovered as such in the wet milling process and is channeled to CGF. See, for example, Anderson et al. (1982) “*The Corn Milling Industry*”; *CRC Handbook of Processing and Utilization in Agriculture*, A. Wolff, Boca Raton, Fla., CRC Press, Inc., Vol. 11, Part 1, *Plant Products*: 31-61 and Eckhoff (Jun. 24-26, 1992) *Proceedings of the 4th Corn Utilization Conference*, St. Louis, Mo., printed by the National Corn Growers Association, CIBA-GEIGY Seed Division, and the USDA, both of which are herein incorporated by reference.

[0127] In other embodiments, the oil content of the plant or plant part is increased. Plants containing an increase in oil content can be used in a variety of applications. For example, high oil plants have an improved food efficiency, which results in greater amounts of energy in the germ. In addition, high oil plants can have an increase in lysine levels, reduced dust during grinding, and improved feed product when compared with normal plants. High oil content in seeds also yields greater amounts of oil when grain is processed into oil and provides economic advantages to starch wet milling.

[0128] Accordingly, the present invention further provides plants having an altered oil phenotype when compared to the oil phenotype of a control plant. In specific embodiments, the altered oil phenotype is in a grain. In some embodiments, the plant of the invention has an increased level/activity of the ODP2 polypeptide of the invention and has a decreased oil content. In other embodiments, such plants have stably incorporated into their genome a heterologous nucleic acid molecule comprising an ODP2 nucleotide sequence of the invention operably linked to a promoter that drives expression in the plant cell.

[0129] III. Altering Starch Production in Plants

[0130] The present invention provides a method for modifying the starch production of a plant. By “starch” is intended a polymer of glucose and normally comprises amylose, amylopectin or a mixture of these two polymer types. Functionally analogous chemical compounds, also included within the definition of starch, include phytoglycogen (which occurs in select types of corn) and water soluble polysaccharides (glucose polymers lacking the crystalline structure of starch granules).

[0131] By “modify starch production” of a plant is intended any modulation (increase or decrease) in the overall level of starch in the plant or plant part (i.e., seed, grain, etc.) when

compared to a control plant. The modification in starch production can comprise any statistically significant increase or decrease in starch levels when compared to a control plant. For example, modifying starch production can comprise either an increase or a decrease in overall starch content of about 0.1%, 0.5%, 1%, 3% 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 110%, 125% or greater when compared to a control plant or plant part that has not been transformed with the ODP2 sequence of the invention. Alternatively, the modification of starch production can include about a 0.2 fold, 0.5 fold, 1 fold, 2 fold, 4 fold, 8 fold, 16 fold, or 32 fold increase in overall starch content in the plant or plant part when compared to a control plant that has not been transformed with the ODP2 sequence.

[0132] The method for modifying the starch production in a plant comprises providing an ODP2 sequence of the invention. An ODP2 polypeptide can be provided by introducing the polypeptide into the plant, and thereby modifying the starch production of the plant or plant part. Alternatively, an ODP2 nucleotide sequence can be provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 nucleotide sequence of the invention, expressing the ODP2 sequence, and thereby modifying the starch production of the plant. In yet other embodiments, the ODP2 nucleotide construct introduced into the plant is stably incorporated into the genome of the plant.

[0133] Methods for determining if the starch production in the plant or plant part has been altered are known in the art. For example, total starch measurement can be performed as outlined in McCleary et al. (1994) *Journal of Cereal Science* 20:51-58, McCleary et al. (1997) *J. Assoc. Off. Anal. Chem.* 80:571-579, and McCleary et al. (2002) *J. AOAC International* 85:1103-1111, each of which is herein incorporated by reference.

[0134] As discussed above, one of skill will recognize the appropriate promoter to use to modify starch production in a plant in the desired manner. Exemplary promoters for this embodiment include the ubiquitin promoter (Christensen et al. (1992) *Plant Molecular Biology* 18:675-680), a lipid transfer protein (LTP) promoter (U.S. Pat. No. 5,525,716), a gamma-zein promoter (GZP) (Boronat et al. (1986) *Plant Sciences* 47:95-102), and the oleosin promoter (WO 00/28058), the lec1 promoter (WO 02/42424), and the Zm-ODP2 promoter (U.S. Provisional Application No. 60/541,171, entitled “ODP2 Promoter and Methods of Use” filed on Feb. 2, 2004).

[0135] In specific embodiments, the modification of starch production results in an increase in starch content in the plant or plant part upon increasing level/activity of the ODP2 polypeptide in a plant. An increased starch content finds use in the wet milling industry and in the ethanol dry grind industry. In other embodiments, the starch production results in a decrease in starch content in the plant or plant part upon decreasing the level/activity of the ODP2 polypeptide in the plant.

[0136] Accordingly, the present invention further provides plants or plant parts having modified starch production when compared to the starch production of a control plant or plant part. In specific embodiments, the plant having the altered starch production is a grain. In some embodiments, the plant of the invention has an increased level/activity of the ODP2 polypeptide of the invention and has an increase in starch accumulation. In other embodiments, such plants have stably

incorporated into their genome a heterologous nucleic acid molecule comprising an ODP2 nucleotide sequence of the invention operably linked to a promoter that drives expression in the plant cell.

[0137] IV. Modifying the Regenerative Capacity of Plants
[0138] The present invention further provides methods to modify the regenerative capacity of a plant. As used herein “regeneration” refers to a morphogenic response that results in the production of new tissues, organs, embryos, whole plants or parts of whole plants that are derived from a single cell or a group of cells. Regeneration may proceed indirectly via a callus phase or directly, without an intervening callus phase. “Regenerative capacity” refers to the ability of a plant cell to undergo regeneration.

[0139] In this embodiment, the method of modifying the regenerative capacity of a plant comprises providing an ODP2 sequence of the invention. In one embodiment, the regenerative capacity of the plant is modified by increasing the level and/or activity of an ODP2 polypeptide. The ODP2 sequence can be provided by introducing an ODP2 polypeptide into the plant, and thereby modifying the regenerative capacity of said plant. Alternatively, an ODP2 nucleotide sequence can be provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 polynucleotide of the invention, expressing the ODP2 sequence, and thereby modifying the regenerative capacity of the plant. In yet other embodiments, the ODP2 nucleotide construct introduced into the plant is stably incorporated into the genome of the plant.

[0140] It is further recognized that providing the ODP2 sequences may be used to enhance the regenerative capacity of plant tissues both in vitro and in vivo and thereby stimulating cell proliferation and/or differentiation. In one embodiment, a method of initiating meristem formation is provided.

[0141] As discussed in further detail below, the promoter used to express the ODP2 sequence of the invention will depend, in part, on the target tissue used for regeneration. Various promoters of interest include constitutive promoters, tissue-preferred promoters, developmentally regulated promoters, and chemically-inducible systems. Various promoters that regulate ovule and embryo expression, nucellus expression, and inner integument expression are discussed in further detail below.

[0142] The ODP2 sequences of the invention also will be useful for inducing apomixis in plants. In specific embodiments, increasing the level and/or activity of the ODP2 polypeptide induces apomixis. Apomixis and methods of conferring apomixis into plants are discussed in U.S. Pat. Nos. 5,710,367; 5,811,636; 6,028,185; 6,229,064; and 6,239,327 as well as WO 00/24914, all of which are incorporated herein by reference. Reproduction in plants is ordinarily classified as sexual or asexual. The term apomixis is generally accepted as the replacement of sexual reproduction by various forms of asexual reproduction (Rieger et al. (1976) *Glossary of Genetics and Cytogenetics*, Springer-Verlag, New York, N.Y.). In general, the initiation of cell proliferation in the embryo and endosperm are uncoupled from fertilization. Apomixis is a genetically controlled method of reproduction in plants where the embryo is formed without the union of an egg and a sperm. There are three basic types of apomictic reproduction: 1) apospory-embryo develops from a chromosomally unreduced egg in an embryo sac derived from a somatic cell in the nucellus; 2) diplospory-embryo develops from an unreduced egg in an embryo sac derived from the

megasporic mother cell; and, 3) adventitious embryony-embryo develops directly from a somatic cell. In most forms of apomixis, pseudogamy or fertilization of the polar nuclei to produce endosperm is necessary for seed viability.

[0143] These types of apomixis have economic potential because they can cause any genotype, regardless of how heterozygous, to breed true. It is a reproductive process that bypasses female meiosis and syngamy to produce embryos genetically identical to the maternal parent. With apomictic reproduction, progeny of specially adaptive or hybrid genotypes would maintain their genetic fidelity throughout repeated life cycles. In addition to fixing hybrid vigor, apomixis can make possible commercial hybrid production in crops where efficient male sterility or fertility restoration systems for producing hybrids are not known or developed. Apomixis can make hybrid development more efficient. It also simplifies hybrid production and increases genetic diversity in plant species with good male sterility. It also provides a system for the production of hybrid seed in species, or between genotypes of the same species in which crossing between separate parent plants is impractical on a large scale.

[0144] In another embodiment, methods for producing embryogenic cells are provided. By “embryogenic cell” is intended a cell that has completed the transition from either a somatic or a gametophytic cell to a state where no further applied stimuli are necessary to produce an embryo. In this embodiment, the method comprises providing an ODP2 sequence of the invention. In one embodiment, the level and/or activity of the ODP2 polypeptide is increased and thereby allows for an increased production of embryogenic cells. In one embodiment, the ODP2 sequence is an ODP2 polypeptide which is provided by introducing the polypeptide into the plant, and thereby producing an embryogenic cell. Alternatively, an ODP2 nucleotide sequence can be provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 nucleotide sequence of the invention, expressing the ODP2 sequence, and thereby producing an embryogenic cell. In yet other embodiments, the ODP2 nucleotide construct introduced into the plant is stably incorporated into the genome of the plant.

[0145] Further provided is a method for producing asexually derived embryos. As used herein, the term “asexually derived embryo” refers to an embryo that is generated in the absence of fertilization. The term is inclusive of apomictic and somatic embryos. The term “somatic embryogenesis” refers to non-zygotic embryogenesis. The method comprises introducing into a plant an ODP2 sequence of the invention and thereby producing asexually derived embryos. As discussed above, the embryo can be a somatic embryo, an adventitious embryos, or a gametophytic embryo.

[0146] Methods are also provided for an increase in the production of somatic embryos in a plant. In one embodiment, the level and/or activity of the ODP2 polypeptide is increased and thereby allowing for the production of somatic embryos. In one embodiment, an ODP2 sequence of the invention is provided. The polypeptide can be provided by introducing the polypeptide into the plant, and thereby increasing the production of somatic embryos. Alternatively, an ODP2 nucleotide sequence can be provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 nucleotide sequence of the invention, expressing the ODP2 sequence, and thereby increasing the production of somatic embryos. In yet other embodiments, the ODP2 nucle-

otide construct introduced into the plant is stably incorporated into the genome of the plant.

[0147] The somatic embryo structures may form as individual embryos or as a cluster of structures. In specific embodiments, the plants (i.e., the root, leaf, seedling) expressing the ODP2 sequences are cultured in vitro. The embryos, non-embryogenic callus or both are transferred to appropriate media for the production of embryos or plantlets. While the somatic embryo can be formed independent of additional growth regulators, it is recognized that in some embodiments, growth regulators can be added to the media and include, but are not limited to, 2,4-D (Mordhorst et al. (1998) *Genetics* 149:549-563).

[0148] An increase in asexually derived embryos can be assayed by determining if embryogenesis or embryonic callus is initiated at a higher frequency from transgenic lines expressing ODP2 sequences of the invention compared to a control plant or plant part. See, for example, Boutiller et al. (2002) *The Plant Cell* 14:1737-1749, herein incorporated by reference.

[0149] It is recognized that the plant having the somatic embryo structures may form only a limited number of somatic embryo structures and then resume additional post germination growth. In other embodiments, expression of the ODP2 sequence leads to the reiteration of the embryo forming process, with the result that new embryos or cotyledons are formed continuously.

[0150] In particular embodiments, the level and/or activity of the ODP2 polypeptide will be reduced prior to the regeneration of a plant from these various embryogenic cell types. Methods for reducing the activity of the ODP2 polypeptide are discussed in detail elsewhere herein.

[0151] Embryogenesis can be induced in haploid cells, such as pollen cells, egg cells, or cells from haploid lines, to produce haploid plants. Methods of inducing embryogenesis in haploid cells comprise providing an ODP2 sequence of the invention to a plant. In one embodiment, the level and/or activity of the ODP2 polypeptide is increased and thereby allows for the induction of embryogenesis in haploid cells. An ODP2 polypeptide can be provided by introducing the polypeptide into the plant, and thereby inducing embryogenesis. Alternatively, an ODP2 nucleotide sequence can be provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 nucleotide sequence of the invention, expressing the ODP2 sequence, and thereby inducing embryogenesis. In yet other embodiments, the ODP2 nucleotide construct introduced into the plant is stably incorporated into the genome of the plant.

[0152] In one embodiment, the ODP2 nucleotide sequence introduced into the plant is under the control of a tissue specific promoter that is active in a haploid cell or tissue or a promoter that is active during microspore development (such as, the maize PG47 promoter (Allen et al. (1993) *Plant J.* 3:261-71), the zm-G13 promoter (Hamilton et al. (1992) *Plant Mol. Biol.* 18:211-218). In other embodiments, the ODP2 nucleotide sequence is under the control of an inducible promoter and the application of the inducer allows expression of the ODP2 sequence therein. Alternatively, the promoter used can be both inducible and tissue-preferred, giving greater control over the process. For example, the promoter can be both haploid-tissue specific and inducible. In one embodiment, the promoter is an inducible pollen-specific promoter used to induce somatic embryogenesis in pollen cells. In still other embodiments, site-specific recombination

systems can be used in combination with promoters (i.e., constitutive promoters or inducible promoters) to regulate the appropriate time and level of ODP2 expression. Thus, the methods of the invention find use in promoting embryogenesis in microspore and anther cultures.

[0153] Providing the ODP2 sequence to a haploid tissue or cell results in the formation of haploid somatic embryos, which can be grown into haploid plants using standard techniques. When an inducible promoter is used (whether tissue specific or not), an optimal method comprises exposing excised transgenic tissue containing the haploid cells (e.g., pollen or ovules) to the inducer specific for the inducible promoter for a time sufficient to induce the formation of a somatic embryo, withdrawing the inducer, and growing the somatic embryo into a transgenic haploid plant in the absence of the inducer.

[0154] Diploidization of the haploid plants to form dihaploids, either spontaneously or by treatment with the appropriate chemical (e.g. colchicine) will significantly expedite the process of obtaining homozygous plants as compared to a method of conventional genetic segregation. This technology will not only be beneficial for breeding purposes but also for basic research such as studies of mutagenesis and other genetic studies, because dihaploids are truly homozygous down to the DNA level, containing two identical copies of each gene.

[0155] In yet another embodiment, adventitious embryony can be achieved by providing an ODP2 sequence of the invention to sporophytic ovule tissues such as the nucellus, the inner integuments, or other tissues lying adjacent to or in proximity to the developing embryo sac.

[0156] The ODP2 sequences of the invention may also be used as a selectable marker to recover transgenic plants. In one embodiment, the level and/or activity of the ODP2 sequence is increased. In this embodiment, a plant is transformed with the ODP2 sequences along with a nucleotide sequence of interest. Upon expression of the ODP2 sequences, the plants can be selected based on their ability to regenerate under conditions in which wild type explants are unable to. For example, the transgenic plants may be able to regenerate in the absence of growth regulators. If the ODP2 sequence and the polynucleotide of interest are carried on separate plasmids, the ODP2 sequence can be subsequently removed from transgenic plants by routine breeding methods.

[0157] One of skill in the art will recognize that a variety of promoters can be used in the various methods of the invention. Somatic or gametophytic embryos can be obtained expressing the ODP2 polypeptide under the control of constitutive promoters, tissue-preferred, developmentally regulated, or various inducible promoters including chemical induction systems (i.e., tetracycline-inducible systems, steroid inducible promoters, and ethanol-inducible promoters). Temporal and/or spatial restriction of ODP2 is optimal when recurrent embryogenesis is not a desirable trait. Promoters of interest when microspore-derived embryo production is desired include, but are not limited to, microspore/pollen expressed genes such as NTM19 (EP 790,311), BCP1 (Xu et al. (1995) *Plant Mol. Biol.* 22:573-588, PG47 (Allen et al. (1993) *Plant J.* 3:261-71), ZmG13 (Hamilton et al. (1992) *Plant Mol. Biol.* 18:211-218), and BNM1 (Treacy et al. (1997) *Plant Mol. Biol.* 34:603-611), each reference is herein incorporated by reference. Promoters of interest when the production of somatic embryos are desired include, but are not limited to, cytokinin inducible IB6 and CK11 promoters

(Brandstatter et al. (1998) *Plant Cell* 10: 1009-1019). Exemplary promoters of interest when adventitious embryony, diplospory or haploid parthenogenesis of embryo sac components, include, the AtDMC1 gene (WO 98/28431), promoters that direct expression in the ovule, such as the AGL11 promoter (Rounsley et al. (1995) *Plant Cell* 10: 1009-1019) and the SERK promoter (Schmidt et al. (1997) *Development* 124:2049-2062), promoters that direct expression in the nucellus such as the NUC1 promoter (WO 98/08961), promoters that regulate expression of inner integument genes such as the FBP7 promoter (Angenent et al. (1995) *Plant Cell* 7:1569-1582), microspore/pollen-preferred promoters (discussed above) and chemical induction systems. Each of these references is herein incorporated by reference.

[0158] Accordingly, the present invention further provides plants having a modified regenerative capacity, including plants that are capable of producing asexually derived embryos. In some embodiments, the plants having a modified regenerative capacity have an increased level/activity of the ODP2 polypeptide of the invention. In other embodiments, the plant comprises a heterologous ODP2 nucleotide sequence of the invention operably linked to a promoter that drives expression in the plant cell. In other embodiments, such plants have stably incorporated into their genome a heterologous nucleic acid molecule comprising an ODP2 nucleotide sequence of the invention operably linked to a promoter that drives expression in the plant cell.

[0159] In other embodiments, the ODP2 sequences of the invention can be used to modify the tolerance of a plant to abiotic stress. In one embodiment, a method is provided to increase or maintain seed set during abiotic stress episodes. During periods of stress (i.e., drought, salt, heavy metals, temperature, etc.) embryo development is often aborted. In maize, halted embryo development results in aborted kernels on the ear. Preventing this kernel loss will maintain yield. Accordingly, methods are provided to increase the stress resistance in a plant (i.e., an early developing embryo).

[0160] The method comprises providing an ODP2 sequence of the invention. The polypeptide can be provided by introducing the polypeptide into the plant, and thereby modifying the plants tolerance to abiotic stress. Alternatively, an ODP2 nucleotide sequence can be provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 nucleotide sequence of the invention, expressing the ODP2 sequence, and thereby modifying the plants tolerance to abiotic stress. In yet other embodiments, the ODP2 nucleotide construct introduced into the plant is stably incorporated into the genome of the plant.

[0161] A variety of promoters can be employed in this method. In one embodiment, the ODP2 sequence is under the control of an early promoter. An early embryo is defined as the stages of embryo development including the zygote and the developing embryo up to the point where embryo maturation begins. An "early embryo promoter" is a promoter that drives expression predominately during the early stages of embryo development (i.e., before 15-18 DAP). Alternatively, the early embryo promoter can drive expression during both early and late stages. Early embryo promoters include, but are not limited to, to Lec 1 (WO 02/42442); cim1, a pollen and whole kernel specific promoter (WO 00/11177); the seed-preferred promoter end1 (WO 00/12733); and, the seed-preferred promoter end2 (WO 00/12733) and lpt2 (U.S. Pat. No. 5,525,716). Additional promoter include, sm1ps, an embryo specific promoter and cz19B1a whole kernel specific promoter.

See, for example, WO 00/11177, which is herein incorporated by reference. All of these references is herein incorporated by reference.

[0162] Methods to assay for an increase in seed set during abiotic stress are known in the art. For example, plants having the ODP2 sequences of the invention can be monitored under various stress conditions and compared to controls plants (not having had the ODP2 introduced). For instance, the plant having the ODP2 sequence can be subjected to various degrees of stress during flowering and seed set. Under identical conditions, the genetically modified plant having the ODP2 sequences will have a higher number of developing kernels than a wild type (non-transformed) plant.

[0163] Accordingly, the present invention further provides plants having increased yield or maintaining their yield during periods of abiotic stress (i.e. drought, salt, heavy metals, temperature, etc.). In some embodiments, the plants having an increased or maintained yield during abiotic stress have an increased level/activity of the ODP2 polypeptide of the invention. In other embodiments, the plant comprises a heterologous ODP2 nucleotide sequence of the invention operably linked to a promoter that drives expression in the plant cell. In other embodiments, such plants have stably incorporated into their genome a heterologous nucleic acid molecule comprising an ODP2 nucleotide sequence of the invention operably linked to a promoter that drives expression in the plant cell.

[0164] V. Modifying the Transformation Efficiency in Plants

[0165] The present invention provides novel methods for transformation and for increasing transformation frequencies. As used herein "responsive target plant cell" is a plant cell that exhibits increased transformation efficiency after the introduction of the ODP2 sequences of the invention when compared to a control plant or plant part. The increase in transformation efficiency can comprise any statistically significant increase when compared to a control plant. For example, an increase in transformation efficiency can comprises about 0.2%, 0.5%, 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 120%, 125% or greater increase when compared to a control plant or plant part. Alternatively, the increase in transformation efficiency can include about a 0.2 fold, 0.5 fold, 1 fold, 2 fold, 4 fold, 8 fold, 16 fold, or 32 fold or greater increase in transformation efficiency in the plant when compared to a control plant or plant part.

[0166] Many maize genotypes, and in particular elite germplasm developed in commercial breeding programs, are recalcitrant to in vitro culture and transformation. Such genotypes do not produce an appropriate embryogenic or organogenic culture response on culture media developed to elicit such responses from typically suitable explants such as immature embryos. Furthermore, when exogenous DNA is introduced into these immature embryos (for example, using particle bombardment or *Agrobacterium*), no transgenic events are recovered after selection (or so few events are recovered as to make transformation of such a genotype impractical). When the ODP2 gene is expressed (either transiently or stably) in immature embryos of such genotypes, vigorously growing transgenic events can be readily recovered.

[0167] Thus, the present invention finds use in increasing the transformation of a recalcitrant plant or explants. As used herein "recalcitrant plant or explant" means a plant or explant that is more difficult to transform than model systems. In

maize such a model system is High type-II maize. Elite maize inbreds are typically recalcitrant. In soybeans such model systems are Peking or Jack.

[0168] In one embodiment of the invention, a method for increasing the transformation efficiency in a plant is provided. The method comprises providing an ODP2 sequence of the invention. An ODP2 polypeptide can be provided by introducing the polypeptide into the plant, and thereby increasing the transformation efficiency of the plant. Alternatively, an ODP2 nucleotide sequence can be provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 nucleotide sequence of the invention, expressing the ODP2 sequence, and thereby increasing the transformation efficiency of the plant. In yet other embodiments, the ODP2 nucleotide construct introduced into the plant is stably incorporated into the genome of the plant. Through the introduction of an ODP2 into a recalcitrant plant and producing a positive influence on transformation, the methods of the invention provide the potential to increase the overall genetic transformation throughput of various recalcitrant germplasm.

[0169] Accordingly, the present invention further provides plants having increased transformation efficiencies when compared to the transformation efficiency of a control plant. In some embodiments, the plants having increased transformation efficiencies have an increased level/activity of the ODP2 polypeptide of the invention. In other embodiments, the plant comprises a heterologous ODP2 nucleotide sequence of the invention operably linked to a promoter that drives expression in the plant cell. In other embodiments, such plants have stably incorporated into their genome a heterologous nucleic acid molecule comprising an ODP2 nucleotide sequence of the invention operably linked to a promoter that drives expression in the plant cell.

[0170] In another embodiment, a method of transforming in a plant is provided. The method comprises providing a target plant, where the target plant had been provided an ODP2 sequence of the invention. In some embodiments, the ODP2 nucleotide sequence is provided by introducing into the plant a heterologous polynucleotide comprising an ODP2 nucleotide sequence of the invention, expressing the ODP2 sequence. In yet other embodiments, the ODP2 nucleotide construct introduced into the target plant is stably incorporated into the genome of the plant. The target plant is transformed with a polynucleotide of interest. It is recognized that the target plant having had the ODP2 sequence introduced (referred to herein as a "modified target plant"), can be grown under conditions to produce at least one cell division to produce a progeny cell expressing the ODP2 sequence prior to transformation with one or more polynucleotides of interest. As used herein "re-transformation" refers to the transformation of a modified cell.

[0171] The modified target cells having been provided the ODP2 sequence can be obtained from T0 transgenic cultures, regenerated plants or progeny whether grown in vivo or in vitro so long as they exhibit stimulated growth compared to a corresponding cell that does not contain the modification. This includes but is not limited to transformed callus, tissue culture, regenerated T0 plants or plant parts such as immature embryos or any subsequent progeny of T0 regenerated plants or plant parts.

[0172] Once the target cell is provided with the ODP2 nucleotide sequence it is re-transformed with at least one gene of interest. The transformed cell can be from transformed callus, transformed embryo, T0 regenerated plants or

its parts, progeny of T0 plants or parts thereof as long as the ODP2 sequence of the invention is stably incorporated into the genome.

[0173] Methods to determine transformation efficiencies or the successful transformation of the polynucleotide of interest are known in the art. For example, transgenic plants expressing a selectable marker can be screened for transmission of the gene(s) of interest using, for example, chemical selection, phenotype screening standard immunoblot and DNA detection techniques. Transgenic lines are also typically evaluated on levels of expression of the heterologous nucleic acid. Expression at the RNA level can be determined initially to identify and quantitate expression-positive plants. Standard techniques for RNA analysis can be employed and include PCR amplification assays using oligonucleotide primers designed to amplify only the heterologous RNA templates and solution hybridization assays using heterologous nucleic acid-specific probes.

[0174] The RNA-positive plants can then be analyzed for protein expression by Western immunoblot analysis using the specifically reactive antibodies of the present invention. In addition, in situ hybridization and immunocytochemistry according to standard protocols can be done using heterologous nucleic acid specific polynucleotide probes and antibodies, respectively, to localize sites of expression within transgenic tissue. Generally, a number of transgenic lines are usually screened for the incorporated nucleic acid to identify and select plants with the most appropriate expression profiles.

[0175] Seeds derived from plants regenerated from re-transformed plant cells, plant parts or plant tissues, or progeny derived from the regenerated plants, may be used directly as feed or food, or further processing may occur.

[0176] Any polynucleotide of interest can be used in the methods of the invention. Various changes in phenotype are of interest including modifying the fatty acid composition in a plant, altering the amino acid content, starch content, or carbohydrate content of a plant, altering a plant's pathogen defense mechanism, affecting kernel size, sucrose loading, and the like. The gene of interest may also be involved in regulating the influx of nutrients, and in regulating expression of phytate genes particularly to lower phytate levels in the seed. These results can be achieved by providing expression of heterologous products or increased expression of endogenous products in plants. Alternatively, the results can be achieved by providing for a reduction of expression of one or more endogenous products, particularly enzymes or cofactors in the plant. These changes result in a change in phenotype of the transformed plant.

[0177] Genes of interest are reflective of the commercial markets and interests of those involved in the development of the crop. Crops and markets of interest change, and as developing nations open up world markets, new crops and technologies will emerge also. In addition, as our understanding of agronomic traits and characteristics such as yield and heterosis increase, the choice of genes for transformation will change accordingly. General categories of genes of interest include, for example, those genes involved in information, such as zinc fingers, those involved in communication, such as kinases, and those involved in housekeeping, such as heat shock proteins. More specific categories of transgenes, for example, include genes encoding important traits for agronomics, insect resistance, disease resistance, herbicide resistance, sterility, grain characteristics, and commercial prod-

ucts. Genes of interest include, generally, those involved in oil, starch, carbohydrate, or nutrient metabolism as well as those affecting kernel size, sucrose loading, and the like.

[0178] Agronomically important traits such as oil, starch, and protein content can be genetically altered in addition to using traditional breeding methods. Modifications include increasing content of oleic acid, saturated and unsaturated oils, increasing levels of lysine and sulfur, providing essential amino acids, and also modification of starch. Hordothionin protein modifications are described in U.S. Pat. Nos. 5,703,049, 5,885,801, 5,885,802, and 5,990,389, herein incorporated by reference. Another example is lysine and/or sulfur rich seed protein encoded by the soybean 2S albumin described in U.S. Pat. No. 5,850,016, and the chymotrypsin inhibitor from barley, described in Williamson et al. (1987) *Eur. J. Biochem.* 165:99-106, the disclosures of which are herein incorporated by reference.

[0179] Derivatives of the coding sequences can be made by site-directed mutagenesis to increase the level of preselected amino acids in the encoded polypeptide. For example, methionine-rich plant proteins such as from sunflower seed (Lilley et al. (1989) *Proceedings of the World Congress on Vegetable Protein Utilization in Human Foods and Animal Feedstuffs*, ed. Applewhite (American Oil Chemists Society, Champaign, Ill.), pp. 497-502; herein incorporated by reference); corn (Pedersen et al. (1986) *J. Biol. Chem.* 261:6279; Kirihara et al. (1988) *Gene* 71:359; both of which are herein incorporated by reference); and rice (Musumura et al. (1989) *Plant Mol. Biol.* 12:123, herein incorporated by reference) could be used. Other agronomically important genes encode latex, Floury 2, growth factors, seed storage factors, and transcription factors.

[0180] Insect resistance genes may encode resistance to pests that have great yield drag such as rootworm, cutworm, European Corn Borer, and the like. Such genes include, for example, *Bacillus thuringiensis* toxic protein genes (U.S. Pat. Nos. 5,366,892; 5,747,450; 5,736,514; 5,723,756; 5,593,881; and Geiser et al. (1986) *Gene* 48:109); and, the like.

[0181] Genes encoding disease resistance traits include detoxification genes, such as against fumonosin (U.S. Pat. No. 5,792,931); avirulence (avr) and disease resistance (R) genes (Jones et al. (1994) *Science* 266:789; Martin et al. (1993) *Science* 262:1432; and Mindrinos et al. (1994) *Cell* 78:1089); and the like.

[0182] Herbicide resistance traits may include genes coding for resistance to herbicides that act to inhibit the action of acetolactate synthase (ALS), in particular the sulfonylurea-type herbicides (e.g., the acetolactate synthase (ALS) gene containing mutations leading to such resistance, in particular the S4 and/or Hra mutations), genes coding for resistance to herbicides that act to inhibit action of glutamine synthase, such as phosphinothricin or basta (e.g., the bar gene), glyphosate (e.g., the EPSPS gene and the GAT gene; see, for example, U.S. Publication No. 20040082770 and WO 03/092360) or other such genes known in the art. The bar gene encodes resistance to the herbicide basta, the nptII gene encodes resistance to the antibiotics kanamycin and gentamicin, and the ALS-gene mutants encode resistance to the herbicide chlorsulfuron.

[0183] Sterility genes can also be encoded in an expression cassette and provide an alternative to physical detasseling. Examples of genes used in such ways include male tissue-preferred genes and genes with male sterility phenotypes such as QM, described in U.S. Pat. No. 5,583,210. Other

genes include kinases and those encoding compounds toxic to either male or female gametophytic development.

[0184] The quality of grain is reflected in traits such as levels and types of oils, saturated and unsaturated, quality and quantity of essential amino acids, and levels of cellulose. In corn, modified hordothionin proteins are described in U.S. Pat. Nos. 5,703,049, 5,885,801, 5,885,802, and 5,990,389.

[0185] Commercial traits can also be encoded on a gene or genes that could increase for example, starch for ethanol production, or provide expression of proteins. Another important commercial use of transformed plants is the production of polymers and bioplastics such as described in U.S. Pat. No. 5,602,321. Genes such as β -Ketothiolase, PHBBase (polyhydroxybutyrate synthase), and acetoacetyl-CoA reductase (see Schubert et al. (1988) *J. Bacteriol.* 170:5837-5847) facilitate expression of polyhydroxyalkanoates (PHAs).

[0186] Exogenous products include plant enzymes and products as well as those from other sources including prokaryotes and other eukaryotes. Such products include enzymes, cofactors, hormones, and the like. The level of proteins, particularly modified proteins having improved amino acid distribution to improve the nutrient value of the plant, can be increased. This is achieved by the expression of such proteins having enhanced amino acid content.

[0187] The following examples are offered by way of illustration and not by way of limitation.

EXPERIMENTAL

Example 1

Cloning of ZM-ODP2

[0188] The protein encoded by maize EST clone cpf1c_pk009.f4 was initially identified as the homologue of a rice putative ovule development protein (BAB89946). The EST clone was subjected to full-insert sequencing. Comparison of rice BAB89946 and the protein sequence encoded by the longest open reading frame (ORF) from cpf1c_pk009.f4 suggests that this clone may have an internal deletion which causes premature termination of the protein by at least 120 amino acids. A genomic fragment encompassing the potential deletion was amplified by PCR using DNA isolated from Hi II callus. Sequencing results confirm the presence of an extra 146 base pairs in the genomic fragment. When added to cDNA clone cpf1c_pk009.f4, this 146-bp can be read through in the same reading frame and the ORF is extended to encode a protein very similar to BAB89946 in length.

[0189] The full-length Zm-ODP2 (SEQ ID NO:1) used in the transformation was created by combining the 5' end of cDNA clone cpf1c_pk009.f4 and part of the genomic clone from Hi II callus that contains the missing 146-bp. More specifically, a 1790-bp EcoRI-SbfI fragment from cpf1c_pk009.f4 and a 582-bp SbfI-SalI genomic fragment were ligated into pBluescript II KS+digested with EcoRI and SalI to form PHP20430.

[0190] The full-length Zm-ODP2 sequence is 2260 nucleotides in length. The open reading frame is 2133 nucleotides in length and starts at nucleotide 128 and ends at nucleotide 2260 of SEQ ID NO:1. The nucleotide sequence of the Zm-ODP2 open reading frame is set forth in SEQ ID NO:3. The 710 amino acid sequence encoded by the Zm-ODP2 sequence is set forth in SEQ ID NO:2.

Example 2

Sequence Analysis of Zm-ODP2

[0191] The ZM-ODP2 sequence of the invention was analyzed for conserved domains.

[0192] FIG. 1 shows an alignment of the amino acid sequence of Zm-ODP2 (SEQ ID NO:2) with various polypeptides sharing sequence similarity to the Zm-ODP2 sequence. Specifically, Zm-ODP2 shares over its full-length about 65.4% sequence identity and 72.7% sequence similarity with OsAnt (Accession No. BAB89946; SEQ ID NO:25). Zm-ODP2 shares over its full-length about 57.1% sequence identity and about 62.3% sequence similarity to OSBNM (Accession No. AAL47205; SEQ ID NO:27). Zm-ODP2 shares over its full-length about 42% sequence identity and about 53.2% sequence similarity to OSODP (Accession No. CAE05555; SEQ ID NO:29). Zm-ODP2 shares over its full-length about 37% sequence identity and about 45% sequence similarity to BnBBM2 (Accession No. AAM33801; SEQ ID NO:33). Zm-ODP2 shares over its full-length about 38% sequence identity and about 47% sequence similarity to BnBBM1 (AAM33800; SEQ ID NO:32). Zm-ODP2 shares over its full-length about 38.1% sequence identity and about 46.3% sequence similarity to ATBBM (Accession No. AAM33803; SEQ ID NO:31). Zm-ODP2 shares over its full-length about 40% sequence identity and about 43% sequence similarity to AtODP (Accession No. AAD30633; SEQ ID NO:36). Zm-ODP2 shares over its full-length about 35.6% sequence identity and about 50% sequence similarity to ATODP (Accession No. NP_175530; SEQ ID NO:34). Zm-ODP2 shares over its full-length about 34.9% sequence identity and about 44.6% sequence similarity to AtODP (Accession No. BAB02492; SEQ ID NO:35). Zm-ODP2 shares over its full-length about 38.4% sequence identity and about 46% sequence similarity to AtODP (NP_197245; SEQ ID NO:30). A consensus sequence for all 11 aligned polypeptides is also provided (SEQ ID NO:37).

[0193] All 11 proteins present in the alignment have two AP2 (APETALA2; pfam00847.8) domains. Using the amino acid numbering of the Zm-ODP2, the first AP2 domain is from about amino acid 273 to about 343 and the second AP2 domain is from about amino acid 375 to about 437. The consensus sequence for the APETALA2 PFAM family is

(SEQ ID NO: 38)
SKYRGVRQRPGKVAEIRDPRKGTRVWLGTFDTEAAARAYDVAALKR
GPSAVLNFPNEL.

Example 3

Variants of Zm-ODP2

[0194] A. Variant Nucleotide Sequences of Zm-ODP2 (SEQ ID NO:1) That Do Not Alter the Encoded Amino Acid Sequence

[0195] The Zm-ODP2 nucleotide sequence set forth in SEQ ID NO:1 was used to generate 6 variant nucleotide sequences having the nucleotide sequence of the open reading frame with about 70.6%, 76.1%, 81.2%, 86.3%, 92.1%, and 97.1% nucleotide sequence identity when compared to the starting unaltered ORF nucleotide sequence of SEQ ID NO: 1. These functional variants were generated using a standard codon table. While the nucleotide sequence of the

variant was altered, the amino acid sequence encoded by the open reading frame did not change.

[0196] The variants of Zm-ODP2 using this method are set forth in SEQ ID NOS:6-11. Specifically, SEQ ID NO: 6 shares about 97.1% nucleic acid sequence identity to the Zm-ODP2 sequence of SEQ ID NO:1; SEQ ID NO:7 shares about 92.1% nucleic acid sequence identity to SEQ ID NO:1, SEQ ID NO:8 shares about 86.3% nucleic acid sequence identity to SEQ ID NO:1; SEQ ID NO:9 shares about 81.2% nucleic acid sequence identity to SEQ ID NO:1; SEQ ID NO:10 shares about 76.1% nucleic acid sequence identity to SEQ ID NO:1; and SEQ ID NO:11 shares about 70.6% nucleic acid sequence identity to SEQ ID NO:1.

[0197] B. Variant Amino Acid Sequences of Zm-ODP2

[0198] Variant amino acid sequences of Zm-ODP2 were generated. In this example, one amino acid was altered. Specifically, the open reading frame set forth in SEQ ID NO:3 was reviewed to determine the appropriate amino acid alteration. The selection of the amino acid to change was made by consulting the protein alignment (with the other orthologs and other gene family members from various species). See FIG. 1. An amino acid was selected that was deemed not to be under high selection pressure (not highly conserved) and which could be rather easily substituted by an amino acid with similar chemical characteristics (i.e., similar functional side-chain). Using the protein alignment set forth in FIG. 1 and focusing at the N-terminus (amino acids 1-50), the serine at amino acid position 37 (shaded) was changed to a threonine, which is chemically similar. Thus, the “TCC” serine codon in the nucleic acid sequence is changed to an “ACC” codon for threonine. The Zm-ODP2 sequence having the single change from “TCC” to “ACC” is set forth in SEQ ID NO:12.

[0199] Once the targeted amino acid was identified, the procedure outlined in Example 3A was followed. Variants having about 70.4% (SEQ ID NO:18), 75.9% (SEQ ID NO:17), 81.5% (SEQ ID NO:16), 86.6% (SEQ ID NO:15), 91.9% (SEQ ID NO:14), and 97.3% (SEQ ID NO:13) nucleic acid sequence identity to SEQ ID NO:3 were generated using this method. SEQ ID NOS: 13-18 all encode the same polypeptide, which is set forth in SEQ ID NO: 19.

[0200] C. Additional Variant Amino Acid Sequences of Zm-ODP2

[0201] In this example, artificial protein sequences were created at a narrower interval range (82.5%, 87.5%, 92.5%, and 97.5% identity relative to the reference protein sequence). This latter effort requires identifying conserved and variable regions from the alignment set forth in FIG. 1 and then the judicious application of an amino acid substitutions table. These parts will be discussed in more detail below.

[0202] Largely, the determination of which amino acid sequences were altered was made based on the conserved regions among AP2 protein or among the other ODP-like genes. See FIG. 1. Based on the sequence alignment, the various regions of the Zm-ODP2 that can likely be altered are represented in lower case letters, while the conserved regions are represented by capital letters. It is recognized that conservative substitutions can be made in the conserved regions below without altering function. In addition, one of skill will understand that functional variants of the ODP2 sequence of the invention can have minor non-conserved amino acid alterations in the conserved domain. This sequence is set forth in SEQ ID NO:2.

The conserved regions are found between about amino acid 1-2; 5-14; 33-34; 38-43; 153-169; 262-463; 591-602; 614-619; 655-661; and 695-800 of SEQ ID NO: 2. The non-conserved regions are from about amino acids 3-4; 15-32; 35-37; 44-152; 170-261; 464-590; 603-613; 620-654; and 662-694 of SEQ ID NO: 2.

[0203] The goal was to create four artificial protein sequences that are different from the original in the intervals of 80-85%, 85-90%, 90-95%, and 95-100% identity. Mid-points of these intervals were targeted, with liberal latitude of plus or minus 1%, for example. The amino acids substitutions will be effected by a custom Perl script. The substitution table is provided below in Table 1.

TABLE 1

Substitution Table			
Amino Acid	Strongly Similar and Optimal Substitution	Rank of Order to Change	Comment
I	L, V	1	50:50 substitution
L	I, V	2	50:50 substitution
V	I, L	3	50:50 substitution
A	G	4	
G	A	5	
D	E	6	
E	D	7	
W	Y	8	
Y	W	9	
S	T	10	
T	S	11	
K	R	12	
R	K	13	
N	Q	14	
Q	N	15	
F	Y	16	

TABLE 1-continued

Substitution Table				
Amino Acid	Strongly Similar and Optimal Substitution	Rank of Order to Change	Comment	
M	L	17	First methionine cannot change	
H		Na	No good substitutes	
C		Na	No good substitutes	
P		Na	No good substitutes	

[0204] First, any conserved amino acids in the protein that should not be changed was identified and “marked off” for insulation from the substitution. The start methionine will of course be added to this list automatically. Next, the changes were made.

[0205] H, C, and P will not be changed in any circumstance. The changes will occur with isoleucine first, sweeping N-terminal to C-terminal. Then leucine, and so on down the list until the desired target is reached. Interim number substitutions can be made so as not to cause reversal of changes. The list is ordered 1-17, so start with as many isoleucine changes as needed before leucine, and so on down to methionine. Clearly many amino acids will in this manner not need to be changed. L, I and V will involve a 50:50 substitution of the two alternate optimal substitutions.

[0206] Four amino acid sequences were written as output. Perl script was used to calculate the percent identities. Using this procedure, variants of Zm-ODP2 were generating having about 82.4% (SEQ ID NO:23), 87.3% (SEQ ID NO:22), 92.4% (SEQ ID NO:21), and 97.3% (SEQ ID NO:20) amino acid identity to the starting unaltered ORF nucleotide sequence of SEQ ID NO:2. FIG. 2 provides an amino acid alignment of SEQ ID NO:2 and the modified proteins set forth in SEQ ID NOS: 20-23.

TABLE 2

Summary of the ODP2 sequences and exemplary variants thereof (SEQ ID NOS 1-25)		
SEQ ID NO	Nucleotide or Amino Acid	Description of Sequence
1	nucleic acid	ZM-ODP2 full length
2	amino acid	ZM-ODP2 full length
3	nucleic acid	ZM-ODP2 - open reading frame
4	nucleic acid	ZM-ODP2 cDNA insert from EST clone cfp1c.pk009.f4
5	nucleic acid	cDNA insert from EST clone cpc1c.pk005.c19
6	nucleic acid	Nucleic acid variant of Zm-ODP2 having 97.2% nucleic acid sequence identity to SEQ ID NO: 2
7	nucleic acid	Nucleic acid variant of Zm-ODP2 having 92.1% nucleic acid sequence identity to SEQ ID NO: 2

TABLE 2-continued

Summary of the ODP2 sequences and exemplary variants thereof (SEQ ID NOS 1-25)		
SEQ ID NO	Nucleotide or Amino Acid	Description of Sequence
8	nucleic acid	Nucleic acid variant of Zm-ODP2 having 86.3% nucleic acid sequence identity to SEQ ID NO: 2
9	nucleic acid	Nucleic acid variant of Zm-ODP2 having 81.2% nucleic acid sequence identity to SEQ ID NO: 2
10	nucleic acid	Nucleic acid variant of Zm-ODP2 having 76.1% nucleic acid sequence identity to SEQ ID NO: 2
11	nucleic acid	Nucleic acid variant of Zm-ODP2 having 70.6% nucleic acid sequence identity to SEQ ID NO: 2
12	nucleic acid	Variant of Zm-ODP2 having the serine 37 codon altered from "tcc" to the threonine codon of "acc". The ORF encodes the amino acid sequence set forth in SEQ ID NO: 19.
13	nucleic acid	Variant of Zm-ODP2 having 97.3% nucleic acid sequence identity to SEQ ID NO: 3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO: 2 with a single amino acid alteration (i.e., S37 to T37). The ORF encodes the amino acid sequence set forth in SEQ ID NO: 19.
14	nucleic acid	Variant of Zm-ODP2 having 91.9% nucleic acid sequence identity to SEQ ID NO: 3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO: 2 with a single amino acid alteration (i.e., S37 to T37).
15	nucleic acid	Variant of Zm-ODP2 having 86.6% nucleic acid sequence identity to SEQ ID NO: 3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO: 2 with a single amino acid alteration (i.e., S37 to T37).
16	nucleic acid	Variant of Zm-ODP2 having 81.5% nucleic acid sequence identity to SEQ ID NO: 3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO: 2 with a single amino acid alteration (i.e., S37 to T37).
17	nucleic acid	Variant of Zm-ODP2 having 75.9% nucleic acid sequence identity to SEQ ID NO: 3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO: 2 with a single amino acid alteration (i.e., S37 to T37).
18	nucleic acid	Variant of Zm-ODP2 having 70.4% nucleic acid sequence identity to SEQ ID NO: 3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO: 2 with a single amino acid alteration (i.e., S37 to T37).
19	Amino acid	Variant of Zm-ODP2 having a single amino acid alteration at S37 to T37.
20	Amino acid	Variant of Zm-ODP2 having 97.3% amino acid sequence identity to SEQ ID NO: 2 (Zm-ODP2).
21	Amino acid	Variant of Zm-ODP2 having 92.4% amino acid sequence identity to SEQ ID NO: 2 (Zm-ODP2).
22	Amino acid	Variant of Zm-ODP2 having 87.3% amino acid sequence identity to SEQ ID NO: 2 (Zm-ODP2).
23	Amino acid	Variant of Zm-ODP2 having 82.4% amino acid sequence identity to SEQ ID NO: 2 (Zm-ODP2).
24	Amino acid	Consensus sequence of FIG. 2.

Example 4

Agrobacterium-mediated Transformation

[0207] For *Agrobacterium*-mediated transformation of maize with a plasmid containing the Zm-ODP2 operably linked to an oleosin promoter and the selectable marker gene PAT, optimally the method of Zhao is employed (U.S. Pat. No. 5,981,840, and PCT patent publication WO98/32326; the contents of which are hereby incorporated by reference). Briefly, immature embryos are isolated from maize and the embryos contacted with a suspension of *Agrobacterium*, where the bacteria are capable of transferring the ODP2 sequence to at least one cell of at least one of the immature embryos (step 1: the infection step). In this step the immature embryos are optimally immersed in an *Agrobacterium* suspension for the initiation of inoculation. The embryos are co-cultured for a time with the *Agrobacterium* (step 2: the co-cultivation step). Optimally the immature embryos are cultured on solid medium following the infection step. Following this co-cultivation period an optional "resting" step is contemplated. In this resting step, the embryos are incubated in the presence of at least one antibiotic known to inhibit the growth of *Agrobacterium* without the addition of a selective

agent for plant transformants (step 3: resting step). Optimally the immature embryos are cultured on solid medium with antibiotic, but without a selecting agent, for elimination of *Agrobacterium* and for a resting phase for the infected cells. Next, inoculated embryos are cultured on medium containing a selective agent and growing transformed callus is recovered (step 4: the selection step). Optimally, the immature embryos are cultured on solid medium with a selective agent resulting in the selective growth of transformed cells. The callus is then regenerated into plants (step 5: the regeneration step), and optimally calli grown on selective medium are cultured on solid medium to regenerate the plants.

Example 5

Altering Oil Content and Starch Content of Maize

[0208] The full length ODP2 sequence described in Example 1, was used for construction of the oleosin driven expression cassette: OLE PRO::ZM-ODP2::NOS TERM. This cassette was inserted into a final transformation plasmid using standard protocols. The final transformation vector contains OLE PRO::ZM-ODP2:: NOS TERM and MO-PAT selection marker is transformed into High type-II maize/

PHR03 via *Agrobacterium* transformation. Methods of *Agrobacterium* transformation are outlined in Example 4.

[0209] Transgenic events are recovered and advanced to the greenhouse. The plants are self-pollinated. At maturity, ears are collected and a portion of seeds (typically 20 kernels from each ear) dissected to separate the embryo from the endosperm. Dissected seeds are dried down in a lyophilizer overnight. The amount of oil in each embryo is determined using NMR. Data for embryo oil %, total embryo oil and embryo weight are collected and analyzed. If changes from High type-II maize/PHR03 baseline are observed, a PCR co-segregation analysis is performed to determine if the changes are correlated with the presence of transgene (ZM-ODP2).

[0210] In addition, germs are also isolated from mature kernels for determination of starch and oil concentrations of the seed part. Individual dry seed are soaked overnight at 4° C. in 1 mL of solution containing 20 mM acetate (pH 6.5) and 10 mM mercuric chloride. (Adkins et al. (1966) *Starch* 7: 213-218). Intact germ is dissected from the seed, dried by lyophilization and recorded for dry weight. Individual germ is ground for 10 sec in a Silamet amalgam mixer and transferred with hexane washing into a microcentrifuge tube. The tissue is extracted by stirring with 1 mL of hexane 3×60 min and centrifuged after each extraction period. The supernatant of extractions is collected and placed into a preweighed aluminum pan. After evaporation of hexane from the weigh pans in a fumehood, final traces of solvent are removed in a forced draft oven at 105° C. for 15 minutes. Cooled weigh pans are reweighed to determine the total weight of oil extracted from the germ. The meal remaining after oil extraction is twice washed with water and centrifugation (10 min; 1,000×g) and analyzed for starch by a modified procedure for total starch measurement (McCleany et al. (1994) *Journal of Cereal Science* 20: 51-58). Free sugars are removed by extraction with 80% ethanol and the starch dissolved in 90% dimethylsulfoxide. Heat stable α -amylase and high purity amyloglucosidase (very low in β -glucanase activities) are used to degrade the starch to monomeric carbohydrate. The resulting glucose will be quantitated according to (Jones et al. (1977) *Plant Physiol.* 60: 379-383) with modification to a microplate format.

Example 6

Placing ODP2 Sequence Under the Control of a Tissue-Preferred Promoter

[0211] The ODP2 gene can be placed under control of an inducible expression system, as described in Zuo et al. (2000) *Plant J* 24:265-273 and in U.S. Patent Application Publication No. US 2003/0082813 A1, the entire contents of which are herein incorporated by reference. The G10-90 promoter in the XVE vector can be replaced with a tissue-preferred promoter (e.g. a pollen-, root- stem- or leaf-specific promoter). A variety of tissue-preferred promoters are well known to those of skill in the art. Because expression of a transgene is activated by the chimeric XVE gene which is controlled by a tissue-preferred promoter in this Example, the O^{lex4} -46 promoter controlling the ODP2 transgene is therefore tissue-preferred in an inducer-dependent manner. This means that ODP2 will be induced only in the presence of an inducer and only in the specific tissue corresponding to the tissue specific promoter. Appropriate tissues or cell types, can then be collected from the transgenic plants and used for induction of somatic embryos and regeneration of plants.

[0212] Particularly when pollen derived from transgenic plants carrying a pollen-specific promoter-XVE/ O^{lex4} -46-ODP2 vector is used, progeny plants generated from pollen-derived somatic embryos should be haploid instead of diploid (see, e.g., Twell et al. (1989) *Mol. Gen. Genetics* 217:240-245 and Twell et al. (1990) *Development* 109:705-714 for pollen-specific promoters). In this embodiment of the invention, a transgenic plant having in its genome a ODP2 gene under the control of an inducible, pollen-specific promoter would not normally express the gene. Pollen from such a plant can be cultured in the presence of the inducer until somatic embryogenesis occurs, after which the inducer is removed and the haploid embryos are permitted to develop into haploid clones according to standard techniques.

Example 7

Generating an Apomictic Plant

[0213] Apomixis can be induced by introducing ODP2 into a plant cell in such a manner that the ODP2 gene is expressed in the appropriate tissues (e.g., nucellus tissue). This can be by means of, but is not limited to, placing the ODP2 gene under the control of a tissue-preferred promoter (e.g., a nucellus-specific promoter), an inducible promoter, or a promoter that is both inducible and tissue-preferred. Inducing expression of the ODP2 gene, e.g. in the nucellus, produces fertilization-independent embryo formation leading to an apomictic plant. This plant may then be used to establish a true-breeding plant line. Additionally, the vector utilized to transfer ODP2 into the plant cell can include any other desired heterologous gene in addition to ODP2, including but not limited to, a marker gene or a gene to confer a desirable trait upon the plant, e.g., a gene resulting in larger plants, faster growth, resistance to stress, etc. This would lead to the development of an apomictic line with the desired trait.

[0214] In a variation of the scheme, plant expression cassettes, including but not limited to monocot or dicot expression cassettes, directing ODP2 expression to the inner integument or nucellus can easily be constructed. An expression cassette directing expression of the ODP2 DNA sequences to the nucellus can be made using the barley Nuc1 promoter (Doan et al. (1996) *Plant Mol. Biol.* 2:276-284). Such an expression can be used for plant transformation. Other genes which confer desirable traits can also be included in the cassette.

[0215] It is anticipated that transgenic plants carrying the expression cassette will then be capable of producing de novo embryos from ODP2 expressing nucellar cells. In the case of maize, this is complemented by pollinating the ears to promote normal central cell fertilization and endosperm development. In another variation of this scheme, Nuc1:ODP2 transformations could be done using a fie (fertility-independent endosperm)-null genetic background which would promote both de novo embryo development and endosperm development without fertilization (Ohad et al. (1999) *The Plant Cell* 11:407-416). Upon microscopic examination of the developing embryos it will be apparent that apomixis has occurred by the presence of embryos budding off the nucellus. In yet another variation of this scheme the ODP2 DNA sequences could be delivered as described above into a

homozygous zygotic-embryo-lethal genotype. Only the adventive embryos produced from somatic nucellus tissue would develop in the seed.

Example 8

Transformation and Regeneration of Maize Embryos

[0216] Immature maize embryos from greenhouse donor plants are bombarded with a plasmid containing the ODP2 sequence of the invention operably linked to a promoter. This could be a weak promoter such as nos, a tissue-specific promoter, such as globulin-1, an inducible promoter such as In2, or a strong promoter such as ubiquitin plus a plasmid containing the selectable marker gene PAT (Wohllaben et al. (1988) *Gene* 70:25-37) that confers resistance to the herbicide Bialaphos. Transformation is performed as follows.

[0217] Maize ears are harvested 8-14 days after pollination and surface sterilized in 30% Chlorox bleach plus 0.5% Micro detergent for 20 minutes, and rinsed two times with sterile water. The immature embryos are excised and placed embryo axis side down (scutellum side up), 25 embryos per plate. These are cultured on 560L medium 4 days prior to bombardment in the dark. Medium 560L is an N6-based medium containing Eriksson's vitamins, thiamine, sucrose, 2,4-D, and silver nitrate. The day of bombardment, the embryos are transferred to 560Y medium for 4 hours and are arranged within the 2.5-cm target zone. Medium 560Y is a high osmoticum medium (560L with high sucrose concentration).

[0218] A plasmid vector comprising the ODP2 sequence operably linked to the selected promoter is constructed. This plasmid DNA plus plasmid DNA containing a PAT selectable marker is precipitated onto 1.1 μ m (average diameter) tungsten pellets using a CaCl_2 precipitation procedure as follows: 100 μ l prepared tungsten particles in water, 10 μ l (1 μ g) DNA in TrisEDTA buffer (1 μ g total), 100 μ l 2.5M CaCl_2 , 10 μ l 0.1M spermidine.

[0219] Each reagent is added sequentially to the tungsten particle suspension, while maintained on the multitube vortexer. The final mixture is sonicated briefly and allowed to incubate under constant vortexing for 10 minutes. After the precipitation period, the tubes are centrifuged briefly, liquid removed, washed with 500 μ l 100% ethanol, and centrifuged for 30 seconds. Again the liquid is removed, and 105 μ l 100% ethanol is added to the final tungsten particle pellet. For particle gun bombardment, the tungsten/DNA particles are briefly sonicated and 10 μ l spotted onto the center of each macrocarrier and allowed to dry about 2 minutes before bombardment.

[0220] The sample plates are positioned 2 levels below the stooping plate for bombardment in a DuPont Helium Particle Gun. All samples receive a single shot at 650 PSI, with a total of ten aliquots taken from each tube of prepared particles/DNA. As a control, embryos are bombarded with DNA containing the PAT selectable marker as described above without the gene of invention.

[0221] Following bombardment, the embryos are kept on 560Y medium, an N6 based medium, for 2 days, then transferred to 560R selection medium, an N6 based medium containing 3 mg/liter Bialaphos, and subcultured every 2 weeks. After approximately 10 weeks of selection, bialaphos-resistant callus clones are sampled for PCR and activity of the gene of interest. In treatments containing the ODP2 gene, it is expected that growth will be stimulated and transformation

frequencies increased, relative to the control. Positive lines are transferred to 288J medium, an MS based medium with lower sucrose and hormone levels, to initiate plant regeneration. Following somatic embryo maturation (2-4 weeks), well-developed somatic embryos are transferred to medium for germination and transferred to the lighted culture room. Approximately 7-10 days later, developing plantlets are transferred to medium in tubes for 7-10 days until plantlets are well established. Plants are then transferred to inserts in flats (equivalent to 2.5" pot) containing potting soil and grown for 1 week in a growth chamber, subsequently grown an additional 1-2 weeks in the greenhouse, then transferred to ClassicTM 600 pots (1.6 gallon) and grown to maturity. Plants are monitored for expression of the gene of interest.

Example 9

Ectopic Expression of Maize ODP2 to Induce Embryogenesis

[0222] Using the genotype High type II as an example, immature embryos are isolated 15 days after pollination and cultured on 560P medium for 3-5 days. At this developmental stage the embryos are too large for callus initiation under standard culture conditions (see above). Twelve hours before bombardment these embryos are transferred to high osmotic 560Y medium. Expression cassettes containing the ODP2 cDNA are then co-introduced into the scutella of these embryos along with an expression cassette containing genes encoding a screenable markers, such as green fluorescent protein (GFP) or cyan fluorescent protein (CFP) using methods well described in the art for particle gun transformations. Twelve to 24 hours following bombardment, embryos are then transferred back to 560P culture medium and incubated in the dark at 26° C. Cultures are then transferred every two weeks until transformed colonies appear. It is expected that expression of ODP2 will stimulate adventive embryo formation. This will be apparent when the cultures are compared to controls (transformed without the ODP2 cDNA). Using either inducible expression cassettes, tissue specific promoters, or promoters of varying strengths it will be possible to control the levels of expression to maximize the formation of adventive embryos. Using either non-responsive genotypes or sub-optimal culture conditions with responsive genotypes, only the transformed cells expressing the ODP2 cDNA will form embryos and regenerate plants. In this manner, ODP2-induced embryo proliferation can be used as a positive selective marker (only the cells expressing the gene will form embryos) and transformants can be recovered without a negative selective agent (i.e. bialaphos, basta, kanamycin, etc.).

Example 10

Ectopic Expression of Maize ODP2 is Sufficient to Stimulate Organogenesis/Embryogenesis and Increases Transformation Frequencies in Recalcitrant Tissues

[0223] There exists only a small developmental window in which maize embryos are amenable to tissue culture growth, a prerequisite for transformation. Normally this occurs between 9-12 days after pollination when the immature embryos are between 1.0-1.5 mm in length. Older, larger embryos fail to produce embryogenic callus and thus cannot be transformed. To demonstrate that ODP2 can be used to induce embryogenesis, embryos from the maize inbred

PH581, ATCC deposit PTA-4432, were isolated 17 days after pollination and used for transformation experiments. Isolated embryos were cultured on 605J medium (a medium containing both full strength MS salts (macro and micronutrient) and 0.6×N6 macronutrient salts plus additional B5 micronutrients, with a mixture of SH and Eriksson's vitamin, L-proline and casamino acids, silver nitrate, 0.3 mg/l 2,4-D and 1.2 mg/l Dicamba, 2% sucrose and 0.06% glucose, solidified with agar). The embryos were incubated in the dark at 28° C. overnight. Embryos were shot in a method similar to that in Example 8 substituting 0.6 μ m gold particles for tungsten. DNA was delivered using co-transformation, as noted above. As a control, embryos were shot with a 1:1 mixture of plasmid DNA's containing a Ubiquitin driven yellow fluorescence protein (YFP) and a plasmid containing a Ubiquitin driven uidA gene (GUS). In the ODP2 treatment the embryos were bombarded with a 1:1 mixture of plasmid DNA's containing the Ubiquitin promoter driving expression of YFP (Ubi:YFP) and a plasmid containing ODP2 (SEQ ID NO: 3) driven by the maize Ubiquitin promoter (Ubi:ODP2). Each treatment contained 20 embryos. After one month of culture the embryos were observed under the dissecting microscope using epifluorescence.

[0224] As mentioned above, it is well known in the art that there is a narrow window in embryo ontogeny where embryos are culture/transformation responsive and this window occurs when embryos are in 1-2 mm in length which is typically 9-12 days after pollination. Since these embryos were taken at 17 days after pollination no multicellular colonies were expected in the control treatment. As expected, hundreds of cells transiently expressing the YFP protein were visible under a fluorescent microscope in the control treatment, and in this population of fluorescing cells, cell division was very rare. Cells transiently expressing YFP were also apparent in the ODP2 treatment. However, in the ODP2 treatment, cell division was apparent in all of the bombarded embryos with up to 50 multicellular colonies observed per embryo (data not shown). No events were observed in the control treatment while 100% of the ODP2 embryos were transformed with 5-50 events/embryo. Embryo morphology was clearly visible in many of these growing transgenic colonies.

[0225] As mentioned above ODP2 expression was sufficient to induce embryogenesis in larger and normally non-responsive embryos. In a similar manner, controlled ODP2 expression should allow transformation of other vegetative tissues such as leaves, stems, and even seed. ODP2 driven by the ubiquitin promoter was used to transform stem tissues. Transformed embryos were recovered from stem tissues (data not shown).

Example 11

Transient Expression of the ODP2 Gene Product to Induce Embryogenesis

[0226] It may be desirable to "kick start" meristem formation by transiently expressing the ODP2 gene product. This can be done by delivering ODP2 5' capped polyadenylated RNA, expression cassettes containing ODP2 DNA, or ODP2 protein. All of these molecules can be delivered using a biolistics particle gun. For example, 5' capped polyadenylated ODP2 RNA can easily be made in vitro using Ambion's mMessage mMachine kit. Following a delivery procedure outlined above, RNA is co-delivered along with DNA containing an agronomically useful expression cassette. It is

expected that cells receiving ODP2 will form embryos and a large portion of these will have integrated the agronomic gene. Plants regenerated from these embryos can then be screened for the presence of the agronomic gene.

Example 12

Modifying the Regenerative Capacity of a Plant

[0227] To demonstrate that ODP2 improves the regenerative capacity of maize tissues transformants were produced in the genotype High Type II with constructs containing the ODP2 gene driven by the maize Oleosin promoter. The Oleosin promoter is highly specific and is expressed only in scutella of developing embryos. Transformants were produced using both particle gun (as described in example 4 above) and *Agrobacterium* (U.S. Pat. No. 5,981,840). Putative transformants were grown in the greenhouse and were completely normal in phenotype. Ears were pollinated and segregating embryos were isolated from a particle gun event at 17 DAP (days after pollination) and from *Agrobacterium* derived events at 24 DAP. Embryos cultured at such late stages would be expected to germinate on regeneration medium. This was observed in the wild-type segregates but germination was delayed in the transformed embryos. In addition to delayed germination, somatic embryos proliferated from the scutella of the transformed embryos (data not shown) when cultured on regeneration medium. The maize Oleosin promoter is highly expressed at these late stages of development and this result demonstrates that the maize ODP2 gene is sufficient to induce embryogenesis in a normally non-responsive tissue.

Example 13

Transient Expression of ODP2 Enhances Transformation

[0228] Parameters of the transformation protocol can be modified to insure that the increased ODP2 activity is transient. One such method involves precipitating the ODP2-containing plasmid in a manner that precludes subsequent release of the DNA (thus, transcription from the particle-bound DNA can occur, but the frequency with which its released to become integrated into the genome is greatly reduced. Such a precipitation relies on the chemical PEI, and it could be used as discussed below.

[0229] The ODP2 plasmid is precipitated onto gold particles with PEI, while the transgenic expression cassette (UBI:3:moPAT~GFPm::pinII) to be integrated is precipitated onto gold particles using the standard Ca^{++} method. Briefly, coating gold particles with PEI is done as follows. First, the gold particles are washed. Thirty-five mg of gold particles, for example 1.0 μ M in average diameter (A.S.I. #162-0010), are weighed out in a microcentrifuge tube, and 1.2 ml absolute EtOH is added and vortexed for one minute. The tube is set aside for 15 minutes at room temperature and then centrifuged at high speed using a microfuge for 15 minutes at 4° C. The supernatant is discarded and a fresh 1.2 ml aliquot of EtOH is added, vortexed for one minute, centrifuged for one minute and the supernatant again discarded (this is repeated twice). A fresh 1.2 ml aliquot of EtOH is added, and this suspension (gold particles in EtOH) can be stored at -20° C. for weeks. To coat particles with polyethylenimine (PEI; Sigma #P3143), start with 250 μ l of washed gold particle/EtOH, centrifuge and discard EtOH. Wash once in 100 μ l ddH₂O to

remove residual ethanol. Add 250 μ l of 0.25 mM PEI, pulse-sonicate to suspend particles and then plunge tube into dry ice/EtOH bath to flash-freeze suspension into place. Lyophilize overnight. At this point, dry, coated particles can be stored at -80° C. for at least 3 weeks. Before use, rinse particles 3 times with 250 μ l aliquots of 2.5 mM HEPES buffer, pH 7.1, with 1x pulse-sonication and then quick vortex before each centrifugation. Suspend in final volume of 250 μ l HEPES buffer. Aliquot 25 μ l to fresh tubes before attaching DNA. To attach uncoated DNA, pulse-sonicate the particles, then add DNA's and mix by pipetting up and down a few times with a PipettemanTM. Let sit for at least 2 minutes, spin briefly (i.e. 10 seconds), remove supernatant and add 60 μ l EtOH. Spot onto macrocarriers and bombard following standard protocol. The Ca⁺⁺ precipitation and bombardment follows standard protocol for the PDS-1000.

[0230] The two particle preparations are mixed together, and the mixture is bombarded into scutellar cells on the surface of immature embryos (some cells receiving only an ODP2 particle, some cells receiving only a PAT~GFP particle and some cells receiving both). PEI-mediated precipitation results in a high frequency of transiently expressing cells on the surface of the immature embryo and extremely low frequencies of recovery of stable transformants (relative to the Ca⁺⁺ method). Thus, the PEI-precipitated ODP2 cassette expresses transiently and stimulates a burst of embryogenic growth on the bombarded surface of the tissue (i.e. the scutellar surface), but this plasmid does not integrate. The PAT~GFP plasmid released from the Ca⁺⁺/gold particles integrates and expresses the selectable marker at a frequency that result in substantially improved recovery of transgenic events.

[0231] As a control treatment, PEI-precipitated particles containing a UBI::GUS::pinII (instead of ODP2) are mixed with the PAT~GFP/Ca⁺⁺ particles. Immature embryos from both treatments are moved onto culture medium containing 3 mg/l bialaphos. After 6-8 weeks, GFP+, bialaphos-resistant calli are observed in the PEI/ODP2 treatment at a much higher frequency relative to the control treatment (PEI/GUS).

[0232] The ODP2 plasmid is precipitated onto gold particles with PEI, and then introduced into scutellar cells on the surface of immature embryos, and subsequent transient expression of the ODP2 gene elicits a rapid proliferation of embryogenic growth. During this period of induced growth, the explants are treated with *Agrobacterium* using standard methods for maize (Zhao et al., U.S. Pat. No. 5,981,840), with T-DNA delivery into the cell introducing a transgenic expression cassette such as UBI::moPAT~GFPm::pinII. After co-cultivation, explants are allowed to recover on normal culture medium, and then are moved onto culture medium containing 3 mg/l bialaphos. After 6-8 weeks, GFP+, bialaphos-resistant calli are observed in the PEI/ODP2 treatment at a much higher frequency relative to the control treatment (PEI/GUS).

Example 14

Transient Expression of the ODP2 Polynucleotide Product to Induce Somatic Embryogenesis

[0233] It may be desirable to "kick start" somatic embryogenesis by transiently expressing the ODP2 polynucleotide product. This can be done by delivering ODP2 5' capped polyadenylated RNA, expression cassettes containing ODP2 DNA, or ODP2 protein. All of these molecules can be delivered using a biolistics particle gun. For example 5'capped

polyadenylated ODP2 RNA can easily be made in vitro using Ambion's mMessage mMachine kit. Following the procedure outline above RNA is co-delivered along with DNA containing an agronomically useful expression cassette, and a marker used for selection/screening such as Ubi::moPAT~GFPm::pinII. The cells receiving the RNA will immediately form somatic embryos and a large portion of these will have integrated the agronomic gene, and these can further be validated as being transgenic clonal colonies because they will also express the PAT~GFP fusion protein (and thus will display green fluorescence under appropriate illumination). Plants regenerated from these embryos can then be screened for the presence of the agronomic gene.

Example 15

Ectopic Expression of ODP2 in Early Zygotic Embryos Increases Seed Set During Abiotic Stress Episodes

[0234] During periods of abiotic stress such as during a drought episode, embryo development often is halted resulting in aborted kernels on the ear. Preventing this kernel loss will increase or maintain yield. To increase seed set during periods of abiotic stress, the ODP2 gene is cloned into an expression cassette behind an early-embryo promoter such as LEC1, and this expression cassette is cloned along with a selectable/screenable marker into an *Agrobacterium* T-DNA region. For example, the following T-DNA is constructed: RB-LEC1::ODP2::pinII/Ubi::moPAT~GFPm::pinII-LB. This T-DNA is introduced into a maize inbred using standard *Agrobacterium* transformation methods. Transgenic plants are screened for single-copy integrations, and then planted in individual pots in the greenhouse. Transgenic plants are selfed and out-crossed to wild-type plants. Plants transgenic for the ODP2 expression cassette are easily tracked (using the cosegregating marker) through either BASTA resistance or green fluorescence conferred by the PAT~GFP fusion protein. Transgenic plants are planted in the field, and subjected to various degrees of drought stress during flowering and seed-set. Under identical stress regimes, the transgenic plants have much higher numbers of developed kernels relative to wild-type (non-transgenic) plants.

Example 16

Expression of ODP2 in Double-Haploid Production

[0235] There are two necessary steps in the production of double-haploid germplasm from maize inbreds. The first is induction of embryogenesis from a haploid cell, and the second is chromosome doubling to convert the haploid to a doubled-haploid.

[0236] The ODP2 gene can be used to generate haploid plants at high frequencies (i.e. improving the efficiency of step one of the process). Various strategies for accomplishing this are described below.

[0237] A. The following expression cassettes are placed in between a single set of T-DNA borders. T-DNA cassette #1 comprises RB-loxP/gal::FLP::pinII/PG47::C1-GAL-EcR::pinII/Ubi::PAT::pinII/Ubi::frt:YFP::pinII::frt:ODP2::pinII/LEC1::Cre::pinII/loxP-LB. To use this construct, it is first transformed into a maize genotype using *Agrobacterium* methods for 2-T-DNA transformation into immature embryos (Miller et al. (2002) *Transgene Research* 11:381-96).

[0238] In addition to the T-DNA diagramed above, this method also introduces T-DNA cassette #2 containing RB-Ole::WUS2::pinII/Ubi::CFP::pinII-LB, but which integrates at an unlinked location in the genome. T-DNA cassette #2 provides a means of recovering transformed events without chemical selection and then later segregating the T-DNA cassette #2 away from #1. Standard tissue culture and regeneration methods are used.

[0239] Transgenic plants are grown until the microspores in the developing tassel are at the uninucleate stage. At this point, the tassel is excised and pretreated by wrapping in moist paper towel and incubated for 14-17 days at 8-10° C. Following pre-treatment, tassels are surface sterilized by soaking for 10 minutes in sodium hypochlorite solution (i.e. 50% Chlorox), and then rinsed twice in sterile water. The anthers are then excised from the tassel and placed on solid anther culture medium using standard media formulations developed for maize anther culture (see Petolino and Genvovesi (1994) in *The Maize Handbook*, (Walbot and Freeling, eds), pages 701-704). Once the anthers are on solid medium, the inducing agent methoxifenozone is pipetted directly onto the solid medium (for example, a 10 mM stock of methoxifenozone is diluted to 10 uM by pipetting 30 ul of the stock onto the surface of 30 ml of solid medium and allowed to equilibrate before adding plant tissue). This will induce expression of FLP recombinase in the uninucleate microspores in the anther. FLP activity would excise the YFP gene, functionally linking the strong Ubiquitin promoter with the ODP2 gene. This burst of ODP2 expression will induce embryogenesis at high frequencies in the haploid uninucleate microspores. After the embryos begin developing, the embryogenic-specific promoter LEC1 will turn on Cre expression and this recombinase will excise the entire transgene cassette. Excision of the expression cassette and the concomitant loss of ODP2 expression will permit embryo maturation and subsequent plant regeneration to occur. During embryo development stimulated by the process described above, colchicine can be added (i.e. a 0.01 to 1.0% solution) to induce chromosome doubling. Doubled haploid plants are recovered that no longer contain T-DNA cassette #2 (because it was segregated away) and only contain the RB-loxP-LB sequence left behind after excision of almost all of cassette #1.

[0240] An alternative way to accomplish the above scenario would be to place the ODP2 gene behind a promoter that is active during microspore development. For example the maize promoters PG47, Zm-POL67 and Zm-POL95 are all promoters active during microspore development. In transgenic plants containing the PG47::ODP2 expression cassette, embryo formation is initiated in the microspores of the developing tassel. An embryo-specific promoter such as LEC1 or Glb1 is then used to drive expression of the Cre gene, which excises the loxP-flanked ODP2 and Cre expression cassettes. These embryos are then capable of maturing and germinating into haploid plants, or if exposed to a doubling agent such as colchicines, double-haploid plants are generated.

Example 17

ODP2 Expression for Positive Selection

[0241] It is expected that transformants can be recovered using ODP2 expression to provide a positive selection means

under reduced auxin levels or in the absence of auxins in the medium, and in the absence of herbicide or antibiotic selection.

[0242] To determine if ODP2 can be used in a positive selection scheme, transformation experiments, using any standard method including particle gun or *Agrobacterium*, can be performed. Transformants are selected on medium with normal auxin levels, or on medium with reduced or no auxin, or visually (using GFP) on medium without bialaphos. Transformation frequencies are based on numbers of embryos with one or more multicellular GFP positive cell clusters. For example, one can test this concept using two treatment variables. The first is that immature embryos are bombarded with a control plasmid (UBI:PAT~GFP) or with UBI:PAT~GFP+In2:ODP2. The second variable is that the bombarded embryos are divided onto either normal bialaphos-containing selection medium (with normal auxin levels of 2 mg/L 2,4-D), or medium with no bialaphos and reduced 2,4-D levels (0.5 mg/L). It is expected from previous studies of positive selection that on bialaphos selection the ODP2 treatment will result in higher transformation frequency than the control. It is also anticipated that the low auxin medium (0.5 mg/L 2,4-D) will result in reduced growth rates. Consistent with this, it is expected that for the control plasmid treatment (UBI:PAT~GFP), recovery of GFP-expressing (fluorescent) colonies will be reduced relative to highly effective bialaphos selection treatment. In contrast, it is expected that ODP2 expression, through its stimulation of embryogenesis, may compensate for the low auxin environment, providing a growth advantage to the transgenic colonies, and maintaining the efficiency of transformant recovery at approximately the same range as the ODP2/bialaphos-selected treatment.

[0243] On medium completely devoid of auxin, it is expected that colonies will only be observed in the ODP2 treatment. In this experiment, immature embryos are transformed with either the control plasmid (UBI:PAT~GFP) or with UBI:PAT~GFP+In2:ODP2, and then plated either onto 3.0 mg/L bialaphos, 2.0 mg/L 2,4-D medium or onto no-bialaphos, no 2,4-D medium (in this latter treatment, wild-type maize callus will not exhibit embryonic growth). Again, it is expected that expression of the ODP2 polynucleotide will increase transformation significantly over the control plasmid value on normal auxin-containing, bialaphos selection medium. Also, it is expected that no transformants will be recovered with the control plasmid on medium devoid of exogenous auxin.

[0244] Even on auxin-containing medium, the ODP2 polynucleotide in combination with GFP+ expression can be used to recover transformants without chemical selection. For example, under these conditions it is expected that the recovery of transformants will be relatively efficient, but may require more diligence than the low- or no-auxin treatments above to separate the GFP-expressing colonies from the growing callus population.

Example 18

Soybean Embryo Transformation

[0245] Soybean embryos are bombarded with a plasmid containing the ODP2 sequence operably linked to a promoter. This could be a weak promoter such as nos, a tissue-specific promoter, such as globulin-1, an inducible promoter such as In2, or a strong promoter such as ubiquitin plus a plasmid

containing the selectable marker gene PAT (Wohlleben et al. (1988) *Gene* 70:25-37) that confers resistance to the herbicide Bialaphos. Transformation is performed as follows.

[0246] To induce somatic embryos, cotyledons, 3-5 mm in length dissected from surface-sterilized, immature seeds of the soybean cultivar A2872, are cultured in the light or dark at 26° C. on an appropriate agar medium for six to ten weeks. Somatic embryos producing secondary embryos are then excised and placed into a suitable liquid medium. After repeated selection for clusters of somatic embryos that multiplied as early, globular-staged embryos, the suspensions are maintained as described below.

[0247] Soybean embryogenic suspension cultures can be maintained in 35 ml liquid media on a rotary shaker, 150 rpm, at 26° C. with fluorescent lights on a 16:8 hour day/night schedule. Cultures are subcultured every two weeks by inoculating approximately 35 mg of tissue into 35 ml of liquid medium.

[0248] Soybean embryogenic suspension cultures may then be transformed by the method of particle gun bombardment (Klein et al. (1987) *Nature* (London) 327:70-73, U.S. Pat. No. 4,945,050). A Du Pont Biolistic PDS1000/HE instrument (helium retrofit) can be used for these transformations.

[0249] A selectable marker gene that can be used to facilitate soybean transformation is a transgene composed of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812), the hygromycin phosphotransferase gene from plasmid pJR225 (from *E. coli*; Gritz et al. (1983) *Gene* 25:179-188), and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*. The expression cassette comprising the ODP2 operably linked to the promoter can be isolated as a restriction fragment. This fragment can then be inserted into a unique restriction site of the vector carrying the marker gene.

[0250] To 50 µl of a 60 mg/ml 1 µm gold particle suspension is added (in order): 5 µl DNA (1 µg/µl), 20 µl spermidine (0.1 M), and 50 µl CaCl₂ (2.5 M). The particle preparation is then agitated for three minutes, spun in a microfuge for 10 seconds and the supernatant removed. The DNA-coated particles are then washed once in 400 µl 70% ethanol and resuspended in 40 µl of anhydrous ethanol. The DNA/particle suspension can be sonicated three times for one second each. Five microliters of the DNA-coated gold particles are then loaded on each macro carrier disk.

[0251] Approximately 300-400 mg of a two-week-old suspension culture is placed in an empty 60×15 mm petri dish and the residual liquid removed from the tissue with a pipette. For each transformation experiment, approximately 5-10 plates of tissue are normally bombarded. Membrane rupture pressure is set at 1100 psi, and the chamber is evacuated to a vacuum of 28 inches mercury. The tissue is placed approximately 3.5 inches away from the retaining screen and bombarded three times. Following bombardment, the tissue can be divided in half and placed back into liquid and cultured as described above.

[0252] Five to seven days post bombardment, the liquid media may be exchanged with fresh media, and eleven to twelve days post-bombardment with fresh media containing 50 mg/ml hygromycin. This selective media can be refreshed weekly. Seven to eight weeks post-bombardment, green, transformed tissue may be observed growing from untransformed, necrotic embryogenic clusters. Isolated green tissue is removed and inoculated into individual flasks to generate

new, clonally propagated, transformed embryogenic suspension cultures. Each new line may be treated as an independent transformation event. These suspensions can then be subcultured and maintained as clusters of immature embryos or regenerated into whole plants by maturation and germination of individual somatic embryos.

Example 19

Sunflower Meristem Tissue Transformation Prophetic Example

[0253] Sunflower meristem tissues are transformed with an expression cassette containing the ODP2 sequence operably linked to a promoter. This could be a weak promoter such as nos, a tissue-specific promoter, such as globulin-1, an inducible promoter such as In2, or a strong promoter such as ubiquitin plus a plasmid containing the selectable marker gene PAT (Wohlleben et al. (1988) *Gene* 70:25-37) that confers resistance to the herbicide Bialaphos. Transformation is performed as follows. See also European Patent Number EP 0 486233, herein incorporated by reference, and Malone-Schoneberg et al. (1994) *Plant Science* 103:199-207).

[0254] Mature sunflower seed (*Helianthus annuus* L.) are dehulled using a single wheat-head thresher. Seeds are surface sterilized for 30 minutes in a 20% Clorox bleach solution with the addition of two drops of Tween 20 per 50 ml of solution. The seeds are rinsed twice with sterile distilled water.

[0255] Split embryonic axis explants are prepared by a modification of procedures described by Schrammeijer et al. (Schrammeijer et al. (1990) *Plant Cell Rep.* 9:55-60). Seeds are imbibed in distilled water for 60 minutes following the surface sterilization procedure. The cotyledons of each seed are then broken off, producing a clean fracture at the plane of the embryonic axis. Following excision of the root tip, the explants are bisected longitudinally between the primordial leaves. The two halves are placed, cut surface up, on GBA medium consisting of Murashige and Skoog mineral elements (Murashige et al. (1962) *Physiol. Plant.*, 15: 473-497), Shepard's vitamin additions (Shepard (1980) in *Emergent Techniques for the Genetic Improvement of Crops* (University of Minnesota Press, St. Paul, Minn.), 40 mg/l adenine sulfate, 30 g/l sucrose, 0.5 mg/l 6-benzyl-aminopurine (BAP), 0.25 mg/l indole-3-acetic acid (IAA), 0.1 mg/l gibberellic acid (GA3), pH 5.6, and 8 g/l Phytagar.

[0256] The explants are subjected to microprojectile bombardment prior to *Agrobacterium* treatment (Bidney et al. (1992) *Plant Mol. Biol.* 18:301-313). Thirty to forty explants are placed in a circle at the center of a 60×20 mm plate for this treatment. Approximately 4.7 mg of 1.8 mm tungsten micro-projectiles are resuspended in 25 ml of sterile TE buffer (10 mM Tris HCl, 1 mM EDTA, pH 8.0) and 1.5 ml aliquots are used per bombardment. Each plate is bombarded twice through a 150 mm nytex screen placed 2 cm above the samples in a PDS 1000® particle acceleration device.

[0257] Disarmed *Agrobacterium tumefaciens* strain EHA105 is used in all transformation experiments. A binary plasmid vector comprising the expression cassette that contains the ODP2 gene operably linked to the promoter is introduced into *Agrobacterium* strain EHA105 via freeze-thawing as described by Holsters et al. (1978) *Mol. Gen. Genet.* 163: 181-187. This plasmid further comprises a kanamycin selectable marker gene (i.e., nptII). Bacteria for plant transformation experiments are grown overnight (28° C. and 100 RPM

continuous agitation) in liquid YEP medium (10 gm/l yeast extract, 10 gm/l Bactopeptone, and 5 gm/l NaCl, pH 7.0) with the appropriate antibiotics required for bacterial strain and binary plasmid maintenance. The suspension is used when it reaches an OD₆₀₀ of about 0.4 to 0.8. The *Agrobacterium* cells are pelleted and resuspended at a final OD₆₀₀ of 0.5 in an inoculation medium comprised of 12.5 mM MES pH 5.7, 1 gm/l NH₄Cl, and 0.3 gm/l MgSO₄.

[0258] Freshly bombarded explants are placed in an *Agrobacterium* suspension, mixed, and left undisturbed for 30 minutes. The explants are then transferred to GBA medium and co-cultivated, cut surface down, at 26° C. and 18-hour days. After three days of co-cultivation, the explants are transferred to 374B (GBA medium lacking growth regulators and a reduced sucrose level of 1%) supplemented with 250 mg/l cefotaxime and 50 mg/l kanamycin sulfate. The explants are cultured for two to five weeks on selection and then transferred to fresh 374B medium lacking kanamycin for one to two weeks of continued development. Explants with differentiating, antibiotic-resistant areas of growth that have not produced shoots suitable for excision are transferred to GBA medium containing 250 mg/l cefotaxime for a second 3-day phytohormone treatment. Leaf samples from green, kanamycin-resistant shoots are assayed for the presence of NPTII by ELISA and for the presence of transgene expression by assaying for ODP2 activity.

[0259] NPTII-positive shoots are grafted to Pioneer® hybrid 6440 in vitro-grown sunflower seedling rootstock. Surface sterilized seeds are germinated in 48-0 medium (half-strength Murashige and Skoog salts, 0.5% sucrose, 0.3% gelrite, pH 5.6) and grown under conditions described for explant culture. The upper portion of the seedling is removed, a 1 cm vertical slice is made in the hypocotyl, and the transformed shoot inserted into the cut. The entire area is wrapped with parafilm to secure the shoot. Grafted plants can be transferred to soil following one week of in vitro culture. Grafts in soil are maintained under high humidity conditions followed by a slow acclimatization to the greenhouse environment. Transformed sectors of T₀ plants (parental generation) maturing in the greenhouse are identified by NPTII ELISA and/or by ODP2 activity analysis of leaf extracts while transgenic seeds harvested from NPTII-positive T₀ plants are identified by ODP2 activity analysis of small portions of dry seed cotyledon.

[0260] An alternative sunflower transformation protocol allows the recovery of transgenic progeny without the use of chemical selection pressure. Seeds are dehulled and surface-sterilized for 20 minutes in a 20% Clorox bleach solution with the addition of two to three drops of Tween 20 per 100 ml of solution, then rinsed three times with distilled water. Sterilized seeds are imbibed in the dark at 26° C. for 20 hours on filter paper moistened with water. The cotyledons and root radical are removed, and the meristem explants are cultured on 374E (GBA medium consisting of MS salts, Shepard vitamins, 40 mg/l adenine sulfate, 3% sucrose, 0.5 mg/l 6-BAP, 0.25 mg/l IAA, 0.1 mg/l GA, and 0.8% Phytagar at pH 5.6) for 24 hours under the dark. The primary leaves are removed to expose the apical meristem, around 40 explants are placed with the apical dome facing upward in a 2 cm circle in the center of 374M (GBA medium with 1.2% Phytagar), and then cultured on the medium for 24 hours in the dark.

[0261] Approximately 18.8 mg of 1.8 µm tungsten particles are resuspended in 150 µl absolute ethanol. After sonication, 8 µl of it is dropped on the center of the surface of macrocar-

rier. Each plate is bombarded twice with 650 psi rupture discs in the first shelf at 26 mm of Hg helium gun vacuum.

[0262] The plasmid of interest is introduced into *Agrobacterium tumefaciens* strain EHA105 via freeze thawing as described previously. The pellet of overnight-grown bacteria at 28° C. in a liquid YEP medium (10 g/l yeast extract, 10 g/l Bactopeptone, and 5 g/l NaCl, pH 7.0) in the presence of 50 µg/l kanamycin is resuspended in an inoculation medium (12.5 mM 2-mM 2-(N-morpholino) ethanesulfonic acid, MES, 1 g/l NH₄Cl and 0.3 g/l MgSO₄ at pH 5.7) to reach a final concentration of 4.0 at OD 600. Particle-bombarded explants are transferred to GBA medium (374E), and a drop of bacteria suspension is placed directly onto the top of the meristem. The explants are co-cultivated on the medium for 4 days, after which the explants are transferred to 374C medium (GBA with 1% sucrose and no BAP, IAA, GA3 and supplemented with 250 µg/ml cefotaxime). The plantlets are cultured on the medium for about two weeks under 16-hour day and 26° C. incubation conditions.

[0263] Explants (around 2 cm long) from two weeks of culture in 374C medium are screened for ODP2 activity using assays known in the art. After positive (i.e., for ODP2 expression) explants are identified, those shoots that fail to exhibit ODP2 activity are discarded, and every positive explant is subdivided into nodal explants. One nodal explant contains at least one potential node. The nodal segments are cultured on GBA medium for three to four days to promote the formation of auxiliary buds from each node. Then they are transferred to 374 C medium and allowed to develop for an additional four weeks. Developing buds are separated and cultured for an additional four weeks on 374 C medium. Pooled leaf samples from each newly recovered shoot are screened again by the appropriate protein activity assay. At this time, the positive shoots recovered from a single node will generally have been enriched in the transgenic sector detected in the initial assay prior to nodal culture.

[0264] Recovered shoots positive for ODP2 expression are grafted to Pioneer hybrid 6440 in vitro-grown sunflower seedling rootstock. The rootstocks are prepared in the following manner. Seeds are dehulled and surface-sterilized for 20 minutes in a 20% Clorox bleach solution with the addition of two to three drops of Tween 20 per 100 ml of solution, and are rinsed three times with distilled water. The sterilized seeds are germinated on the filter moistened with water for three days, then they are transferred into 48 medium (half-strength MS salt, 0.5% sucrose, 0.3% gelrite pH 5.0) and grown at 26° C. under the dark for three days, then incubated at 16-hour-day culture conditions. The upper portion of selected seedling is removed, a vertical slice is made in each hypocotyl, and a transformed shoot is inserted into a V-cut. The cut area is wrapped with parafilm. After one week of culture on the medium, grafted plants are transferred to soil. In the first two weeks, they are maintained under high humidity conditions to acclimatize to a greenhouse environment.

[0265] All publications and patent applications mentioned in the specification are indicative of the level of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

[0266] Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

SEQUENCE LISTING

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 including the 5' untranslated region

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					Ser	Ser
					Leu	Pro
					10	
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Gln	Glu	Leu	Pro	Pro	Ser	
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ccc	acc	agc	tca	ggc	ctc	505
Pro	Thr	Ser	Ala	Leu	His	
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tcg	gcc	ggg	gtc	cac	gac	553
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Ala Val Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val			
225	230	235	
gcc ggc gct cta gta gcc gtg agc acg gac acg ggt ggc agc ggc ggc		889	
Ala Gly Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly			
240	245	250	
gct tcg gct gac aac acg gca agg aag acg gtg gac acg ttc ggg cag		937	
Ala Ser Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln			
255	260	265	270
cgc acg tcg att tac cgt ggc gtg aca agg cat aga tgg act ggg aga		985	
Arg Thr Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg			
275	280	285	
tat gag gca cat ctt tgg gat aac agt tgc aga agg gaa ggg caa act		1033	
Tyr Glu Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr			
290	295	300	
cgt aag ggt cgt caa gtc tat tta ggt ggc tat gat aaa gag gag aaa		1081	
Arg Lys Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys			
305	310	315	
gct gct agg get tat gat ctt gct gct ctg aag tac tgg ggt gcc aca		1129	
Ala Ala Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr			
320	325	330	
aca aca aca aat ttt cca gtg agt aac tac gaa aag gag ctc gag gac		1177	
Thr Thr Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp			
335	340	345	350
atg aag cac atg aca agg cag gag ttt gta gcg tct ctg aga agg aag		1225	
Met Lys His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys			
355	360	365	
agc agt ggt ttc tcc aga ggt gca tcc att tac agg gga gtg act agg		1273	
Ser Ser Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg			
370	375	380	
cat cac caa cat gga aga tgg caa gca cgg att gga cga gtt gca ggg		1321	
His His Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly			
385	390	395	
aac aag gat ctt tac ttg ggc acc ttc agc acc cag gag gag gca gcg		1369	
Asn Lys Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala			
400	405	410	
gag gcg tac gac atc gcg gcg atc aag ttc cgc ggc ctc aac gcc gtc		1417	
Glu Ala Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val			
415	420	425	430
acc aac ttc gac atg agc cgc tac gac gtg aag agc atc ctg gac agc		1465	
Thr Asn Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser			
435	440	445	
agc gcc ctc ccc atc ggc agc gcc gcc aag cgc ctc aag gag gcc gag		1513	
Ser Ala Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu			
450	455	460	
gcc gca gcg tcc gcg cag cac cac gcc ggc gtg gtg agc tac gac		1561	
Ala Ala Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp			
465	470	475	
gtc ggc cgc atc gcc tcg cag ctc ggc gac ggc gga gca ctg gcg gcg		1609	
Val Gly Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala			
480	485	490	
gcg tac ggc gcg cac tac cac ggc gcc tgg ccc acc atc gcg ttc		1657	
Ala Tyr Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe			
495	500	505	510
cag ccg ggc gcc gcc agc aca ggc ctg tac cac ccg tac gcg cag cag		1705	

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Gln Pro Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln			
515	520	525	
cca atg cgc ggc ggc ggg tgg tgc aag cag gag cag gac cac gcg gtc	1753		
Pro Met Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val			
530	535	540	
atc gcg gcc gcg cac agc ctg cag gac ctc cac cac ctg aac ctg ggc	1801		
Ile Ala Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly			
545	550	555	
gcg gcc ggc gcg cac gac ttt ttc tcg gca ggg cag cag gcc gcc gcc	1849		
Ala Ala Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala			
560	565	570	
gct gcg atg cac ggc ctg ggt agc atc gac agt gcg tcg ctc gag cac	1897		
Ala Ala Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His			
575	580	585	590
agc acc ggc tcc aac tcc gtc gtc tac aac ggc ggg gtc ggc gac agc	1945		
Ser Thr Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser			
595	600	605	
aac ggc gcc agc gcc gtc ggc ggc agt ggc ggt ggc tac atg atg ccg	1993		
Asn Gly Ala Ser Ala Val Gly Ser Gly Gly Tyr Met Met Pro			
610	615	620	
atg agc gct gcc gga gca acc act aca tcg gca atg gtg agc cac gag	2041		
Met Ser Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu			
625	630	635	
cag gtg cat gca cgg gcc tac gac gaa gcc aag cag gct gct cag atg	2089		
Gln Val His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met			
640	645	650	
ggg tac gag agc tac ctg gtg aac gcg gag aac aat ggt ggc gga agg	2137		
Gly Tyr Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg			
655	660	665	670
atg tct gca tgg ggg act gtc gtg tct gca gcc gcg gca gca gca	2185		
Met Ser Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala			
675	680	685	
agc agc aac gac aac atg gcc gac gtc ggc cat ggc ggc gcg cag	2233		
Ser Ser Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln			
690	695	700	
ctc ttc agt gtc tgg aac gac act taa	2260		
Leu Phe Ser Val Trp Asn Asp Thr *			
705	710		
<210> SEQ_ID NO 2			
<211> LENGTH: 710			
<212> TYPE: PRT			
<213> ORGANISM: Zea mays			
<400> SEQUENCE: 2			
Met Ala Thr Val Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu			
1	5	10	15
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr			
20	25	30	
Ala Asp His Val Ser Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp			
35	40	45	
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu			
50	55	60	
Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala			
65	70	75	80
Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser			
85	90	95	

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Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr
 100 105 110
 Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala
 115 120 125
 Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Ala Asn Gly
 130 135 140
 Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Gly Ile Gly Leu Ser Met
 145 150 155 160
 Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val
 165 170 175
 Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala
 180 185 190
 Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg
 195 200 205
 Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val
 210 215 220
 Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly
 225 230 235 240
 Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser
 245 250 255
 Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr
 260 265 270
 Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
 275 280 285
 Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys
 290 295 300
 Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
 305 310 315 320
 Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr
 325 330 335
 Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys
 340 345 350
 His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser
 355 360 365
 Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His
 370 375 380
 Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys
 385 390 395 400
 Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala
 405 410 415
 Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn
 420 425 430
 Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala
 435 440 445
 Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala
 450 455 460
 Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly
 465 470 475 480
 Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Ala Tyr
 485 490 495

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Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro
 500 505 510

Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met
 515 520 525

Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala
 530 535 540

Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala
 545 550 555 560

Gly Ala His Asp Phe Phe Ser Ala Gly Gln Ala Ala Ala Ala
 565 570 575

Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr
 580 585 590

Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly
 595 600 605

Ala Ser Ala Val Gly Ser Gly Gly Tyr Met Met Pro Met Ser
 610 615 620

Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val
 625 630 635 640

His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr
 645 650 655

Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser
 660 665 670

Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser
 675 680 685

Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe
 690 695 700

Ser Val Trp Asn Asp Thr
 705 710

<210> SEQ ID NO 3
<211> LENGTH: 2133
<212> TYPE: DNA
<213> ORGANISM: Zea mays
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (1)....(2133)
<221> NAME/KEY: misc_feature
<222> LOCATION: (0)....(0)
<223> OTHER INFORMATION: Open reading frame of Zm-ODP2

<400> SEQUENCE: 3

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Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu	
1 5 10 15	
ctg ccg ccc tcc cag acg acg gac tcc aca ctc atc tcg gcc gcc acc	96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr	
20 25 30	
gcc gac cat gtc tcc ggc gat gtc tgc ttc aac atc ccc caa gat tgg	144
Ala Asp His Val Ser Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp	
35 40 45	
agc atg agg gga tca gag ctt tcg gcg ctc gtc gcg gag ccg aag ctg	192
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu	
50 55 60	
gag gac ttc ctc ggc ggc atc tcc ttc tcc gag cag cat cac aag gcc	240
Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala	
65 70 75 80	

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aac tgc aac atg ata ccc agc act agc agc aca gtt tgc tac gcg agc Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser 85 90 95	288
tca ggt gct agc acc ggc tac cat cac cag ctg tac cac cag ccc acc Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr 100 105 110	336
agc tca gcg ctc cac ttc gcg gac tcc gta atg gtc gcc tcc tcg gcc Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala 115 120 125	384
ggt gtc cac gac ggc ggt gcc atg ctc agc gcg gcc gct aac ggt Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly 130 135 140	432
gtc gct ggc gct gcc agt gcc aac ggc ggc atc ggg ctg tcc atg Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met 145 150 155 160	480
att aag aac tgg ctg cgg agc caa ccg ggc ccc atg cag ccg agg gtc Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val 165 170 175	528
gcg gcg gct gag ggc gcg cag ggg ctc tct ttg tcc atg aac atg gcg Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala 180 185 190	576
ggg acg acc caa ggc gct gtc atg cca ctt ctc gct gga gag cgc Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg 195 200 205	624
gca cgg gcg ccc gag agt gta tcg acg tca gca cag ggt gga gcc gtc Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val 210 215 220	672
gtc gtc acg gcg ccc aag gag gat agc ggt ggc agc ggt gtt gcc ggc Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly 225 230 235 240	720
gct cta gta gcc gtg agc acg gac acg ggt ggc agc ggc ggc gtc Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser 245 250 255	768
gct gac aac acg gca agg aag acg gtg gac acg ttc ggg cag cgc acg Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr 260 265 270	816
tcg att tac cgt ggc gtg aca agg cat aga tgg act ggg aga tat gag Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu 275 280 285	864
gca cat ctt tgg gat aac agt tgc aga agg gaa ggg caa act cgt aag Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys 290 295 300	912
ggt cgt caa gtc tat tta ggt ggc tat gat aaa gag gag aaa gct gct Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala 305 310 315 320	960
agg gct tat gat ctt gct gct ctg aag tac tgg ggt gcc aca aca aca Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr 325 330 335	1008
aca aat ttt cca gtg agt aac tac gaa aag gag ctc gag gac atg aag Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys 340 345 350	1056
cac atg aca agg cag gag ttt gta gcg tct ctg aga agg aag agc agt His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser 355 360 365	1104
ggt ttc tcc aga ggt gca tcc att tac agg gga gtg act agg cat cac Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His 370 375 380	1152

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caa cat gga aga tgg caa gca cgg att gga cga gtt gca ggg aac aag Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys 385 390 395 400	1200
gat ctt tac ttg ggc acc ttc agc acc cag gag gag gca gcg gag gcg Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala 405 410 415	1248
tac gac atc gcg gcg atc aag ttc cgc ggc ctc aac gcc gtc acc aac Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn 420 425 430	1296
ttc gac atg agc cgc tac gac gtg aag agc atc ctg gac agc agc gcc Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala 435 440 445	1344
ctc ccc atc ggc agc gcc gcc aag cgc ctc aag gag gcc gag gcc gca Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala 450 455 460	1392
gcg tcc gcg cag cac cac cac ggc ggc gtg agc tac gac gtc ggc Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly 465 470 475 480	1440
cgc atc gcc tcg cag ctc ggc gac ggc gga gcc ctg gcg gcg tac Arg Ile Ala Ser Gln Leu Gly Asp Gly Ala Leu Ala Ala Tyr 485 490 495	1488
ggc gcg cac tac cac ggc gcc tgg ccg acc atc gcg ttc cag ccc Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro 500 505 510	1536
ggc gcc gcc agc aca ggc ctg tac cac ccg tac gcg cag cag cca atg Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met 515 520 525	1584
cgc ggc ggc ggg tgg tgc aag cag gag cag gac cac gcg gtc atc gcg Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala 530 535 540	1632
gcc ggc cac agc ctg cag gac ctc cac ctg aac ctg ggc gcg gcc Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala 545 550 555 560	1680
ggc gcg cac gac ttt ttc tcg gca ggg cag cag gac ggc gcc gct gcg Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala 565 570 575	1728
atg cac ggc ctg ggt agc atc gac agt gcg tcg ctc gag cac agc acc Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr 580 585 590	1776
ggc tcc aac tcc gtc gtc tac aac ggc ggg gtc ggc gac agc aac ggc Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly 595 600 605	1824
gcc agc gcc gtc ggc ggc agt ggc ggt ggc tac atg atg ccg atg agc Ala Ser Ala Val Gly Ser Gly Gly Tyr Met Met Pro Met Ser 610 615 620	1872
gct gcc gga gca acc act aca tcg gca atg gtg agc cac gag cag gtc Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val 625 630 635 640	1920
cat gca cgg gcc tac gac gaa aag cag gag gct gct cag atg ggg tac His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr 645 650 655	1968
gag agc tac ctg gtg aac gcg gag aac aat ggt ggc gga agg atg tct Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser 660 665 670	2016
gca tgg ggg act gtc gtg tct gca gcc gcg gca gca gca agc agc Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ser Ser 675 680 685	2064

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aac gac aac atg gcc gac gtc ggc cat ggc ggc ggc cag ctc ttc	2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Gly Ala Gln Leu Phe	
690 695 700	
agt gtc tgg aac gac act taa	2133
Ser Val Trp Asn Asp Thr *	
705 710	
<210> SEQ ID NO 4	
<211> LENGTH: 2392	
<212> TYPE: DNA	
<213> ORGANISM: Zea mays	
<220> FEATURE:	
<221> NAME/KEY: misc_feature	
<222> LOCATION: (0)...(0)	
<223> OTHER INFORMATION: ZM-ODP2 cDNA insert from EST clone	
cpflc.pk009.f4	
<400> SEQUENCE: 4	
cttccctaac ctttgactg tccaaaatgg cttcctgatc ccctcacttc ctcgaatcaa	60
tctaagaaga aactcaagcc gcaaccatta ggggcagatt aattgctgca ctttcagata	120
atcaaccatg gcaactgtga acaactggct cgctttctcc ctctcccgcc aggagctgcc	180
gccctcccaag acgacggact ccacactcat ctccggccgcc accggccgacc atgtctccgg	240
cgatgtctgc ttcaacatcc cccaagattg gagcatgagg ggatcagagc tttcggcgct	300
cgtcgccggag ccgaagctgg aggacttcct cggcgccatc tccttctccg agcagoatca	360
caaggccaaac tgcaacatga taccacgac tagcagcaca gtttgctacg cgagtcagg	420
tgcttagcacc ggctaccatc accagctgtt ccaccagccc accagctca ggcgtcoactt	480
cgcggactcc gtaatggtgg cctcctccggc cggtgtccac gacggcggtg ccatgtctcg	540
cgcggccggcc gctaacgggtg tgcgtggccg tgccagtggcc aacggggggccg gcatcgccgt	600
gtccatgatt aagaactggc tgoggagcca accggccccc atgcagccga ggggtggccggc	660
ggctgagggc ggcggccggcc tctctttgtc catgaacatg gccccggccgccaaggccgc	720
tgctggcatg ccacttctcg ctggagagcg cgacacggccg cccgagatgt tatcgacgtc	780
agcacagggt ggagccgtcg tgcgtcaaggcc gccgaaggag gatagcggtg gcagcggtgt	840
tgcggccgtct ctagtageccg tgagcaegga cacgggtggc agcgccggccg cgtcggtctga	900
caacacggca aggaagacgg tggcacacgtt cggccggccg acgtcgattt accgtggcggt	960
gacaaggcat agatggactg ggagatatga ggcacatctt tggataaca gttgcagaag	1020
ggaaggccaa actcgtaagg gtcgtcaagt ctattttaggt ggctatgata aagaggagaa	1080
agctgctagg gcttatgatc ttgctgtct gaagttactgg ggtgccacaa caacaacaaa	1140
ttttccagtg agtaactacg aaaaggagct cgaggacatg aagcacatga caaggccagga	1200
gtttttagcg tctctgagaa ggaagagccg tggtttctcc agaggtgcat ccatttacag	1260
gggagtgtact aggcacatcacc aacatggaa atggcaagca cggattggac gagttcgagg	1320
gaacaaggat ctttacttgg gcaccttcag cacccaggag gaggcagccg aggccgtacga	1380
catcgccggccg atcaagttcc gccccctcaaa cggccgtccacc aacttcgaca tgagccgcta	1440
cgacgtgaag agcatcctgg acagcagccgc cctcccccattc ggcagccggcc ccaagccct	1500
caaggaggcc gaggccgcag cgtccggccgca gcaccaccac gccggcggtgg tgagctacga	1560
cgtcgccggcc atcgccctcgcc agctcgccgca cggccggagcc ctggccggccg cgtacggccgc	1620
gcactaccac ggccgcgcgtt ggccgaccat cgcgttccag cccggccgcgc ccagcacagg	1680

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cctgtaccac ccgtacgcgc agcagccaat gcgcggcgcc gggtgtgca agcaggagca	1740
ggaccacgcg gtgatcgcgg ccgcgcacag cctgcaggac ctccaccacc tgaacctggg	1800
cacggccggc gcgcacgact tttctcggc ggtggctaca tgatgccat gagcgtgcc	1860
ggagcgcacca ctacatcggc aatggtgagc cacgagcaga tgcgcacg ggcctacgc	1920
gaagccaagc aggctgctca gatggggta gagagctacc tggtaaacgc ggagaacaat	1980
ggtgtggcgaa ggatgtctgc atggggact gtcgtgtctg cagccgcggc ggcagcagca	2040
agcagcaacg acaacatggc cgccgcacgta ggccatggcg ggcgcacgct ctgcgtgtc	2100
tggaaacgaca cttaaagctac gcgtacgtgc cggcctggct ctccgaattc gaaccgatcg	2160
atgcgttgta aaaccgtaca ctgacataag taacaacact tagggttctt catggagagg	2220
tggccagtaa gttgttactt gtcataatgtt ttaagttctc aattttgtac tggaaaggaaa	2280
gttagggttt cttctgaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aa	2340
aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aa	2392

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<210> SEQ ID NO 5
<211> LENGTH: 224
<212> TYPE: DNA
<213> ORGANISM: Zea mays
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (0)...(0)
<223> OTHER INFORMATION: EST clone cpclc.pk005.c19
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<400> SEQUENCE: 5  
  
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gcaaggcaggag gcaggaccac gcggtgtatcg cggccgcgcga cagcctgtcag gacctccacc 120  
acctgaacct gggcacggcc ggccgcacg acttttctc ggcagggcag caggccgccc 180  
ccacccacccat gatgcacggac ctggatadca ttgacaaatgc atca 224
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<210> SEQ ID NO 6
<211> LENGTH: 2133
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Variant of Zm-ODP2 having 97.1% nucleic acid
      sequence identity to SEQ ID NO:1 (Zm-ODP2). The
      ORF encodes the amino acid sequence set forth in
      SEQ ID NO:2.
<221> NAME/KEY: CDS
<222> LOCATION: (1) .. (2133)
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<400> SEQUENCE: 6
atg gcc act gtg aac aac tgg ctc gct ttc tcc ctc tcc ccg cag gag 48
Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu

```

ctg ccg ccc tcc cag acg acg gac tcc aca ctc atc tcg gcc gcg acg 96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr
 20          25          30

```

```

gcc gac cat gtc agc ggc gat gtc tgc ttc aac atc ccc caa gat tgg      144
Ala Asp His Val Ser Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp
          35           40           45

```

```

agc atg agg gga tca gag ctt tcg gcg ctg gtc gcg gag ccg aag ctg 192
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu
      50          55          60

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gag gac ttc ctc ggc ggc atc agc ttt tcc gag cag cat cat aag gcc	240
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Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser	
85 90 95	
tca ggt gct agc acc ggc tac cat cac cag ctg tac cat cag ccc acc	336
Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr	
100 105 110	
agc tca gcg ctc cac ttc gcg gac tcc gtt atg gtc gcg tcc tcc gcc	384
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Ala Ser Ser Ala	
115 120 125	
ggt gtc cac gac ggc ggt gcc atg ctc agc gcg gcc gct aac ggt	432
Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly	
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gtc gct ggc gca gcg agt gcc aac ggc ggc ggc atc ggg ctc tcc atg	480
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met	
145 150 155 160	
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Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val	
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Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala	
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ggg acg acc caa ggc gct gct ggc atg cca ctt ctc gct gga gag cgc	624
Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg	
195 200 205	
gca cgg gcg ccc gag agt gta tcg acg tca gca cag ggt gga gcc gtc	672
Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val	
210 215 220	
gtc gtc acg gcc ccg aag gag gat agc ggt ggc agc ggc ggt gtt gcc ggc	720
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly	
225 230 235 240	
gct cta gta gcc gtg agc acg gac acg ggt ggc agc ggc ggc gtc	768
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser	
245 250 255	
gct gac aac acg gca agg aag acg gtg gac acg ttc ggg cag cgc acg	816
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr	
260 265 270	
tcg att tac cgt ggg gtc aca agg cat aga tgg aca ggg aga tac gag	864
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu	
275 280 285	
gca cat ctt tgg gat aac agt tgc aga agg gaa ggg caa act cgt aag	912
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys	
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ggt cgt caa gtc tat tta ggt ggc tat gat aaa gag gag aaa gct gct	960
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala	
305 310 315 320	
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Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr	
325 330 335	
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Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys	
340 345 350	
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His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser	
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gct gcc gga gca acc act act tcg gca atg gtg agc cac gag cag gtg Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val 625 630 635 640	1920
cat gca cgg gcc tac gat gaa gca aag cag gct gct cag atg ggc tac His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr 645 650 655	1968
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gca tgg ggg aca gtc gtg tct gca gcc gcg gca gca agc agc Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser 675 680 685	2064
aac gac aac atg gcg gcc gat gtc ggc cat ggc ggc gcg cag ctc ttc Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe 690 695 700	2112
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ctg ccc ccc tcc cag acc acg gac tcc aca ctc atc tcc gcg gcc acc Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr 20 25 30	96
gcc gac cac gtg tcc ggc gac gtg tcc ttt aat att ccc cag gat tgg Ala Asp His Val Ser Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp 35 40 45	144
agc atg agg gga tca gag ctt tcc gcg ctc gtc gcg gag ccg aag ctg Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu 50 55 60	192
gag gac ttc ctc ggc ggg atc tcc ttc tcc gaa caa cat cac aag gcc Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala 65 70 75 80	240
aac tgt aac atg ata ccc tcc act agc agc aca gtt tcc tac gcg agc Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser 85 90 95	288
tca ggt gca agc acc ggg tat cat cac cag ctg tac cac cag ccc acc Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr 100 105 110	336
agc tca gcg ctc cac ttc gcg gac tcc gtt atg gtg gcc tcc tcc gcc Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala 115 120 125	384
gga gtc cac gac ggc ggt gcc atg ctg tcc gcc gcc gcc gct aac gga Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly 130 135 140	432
gtg gct ggc gct gcc tcc gcc aac ggc ggc ggg atc ggg ctg agc atg Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met 145 150 155 160	480
atc aag aac tgg ctc cgg agc caa ccg gcg ccc atg cag ccg agg gtg Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val 165 170 175	528
gcg gcg gct gag ggg gcc cag ggc ctc tct ttg tcc atg aac atg gcg Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala 180 185 190	576
ggg acg acc caa ggc gca gct ggc atg cct ctt ctg gct gga gag cgc	624

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gtc gtc acc gcg ccg aag gaa gat tcc ggt ggc agc ggt gtt gcc ggc	720
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly	
225 230 235 240	
gct ctc gta gcg gtc agc acg gac acc ggt ggc agc ggg ggc gcg tcg	768
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser	
245 250 255	
gca gac aat acg gct agg aag acg gtg gac acg ttc ggg cag cgg acg	816
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr	
260 265 270	
tcg atc tac cgt ggg gtg aca agg cac aga tgg aca ggg aga tat gag	864
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu	
275 280 285	
gca cac ctt tgg gat aac agt tgc agg gaa ggg caa act cgt aaa	912
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys	
290 295 300	
ggt aga caa gtc tat tta ggt ggc tat gac aaa gag gag aag gct gct	960
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala	
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Arg Ala Tyr Asp Leu Ala Ala Lys Tyr Trp Gly Ala Thr Thr Thr	
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340 345 350	
cac atg act agg caa gag ttc gtt gcg tcc ctg aga aga aag agc agt	1104
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser	
355 360 365	
ggt ttc tcc aga ggt gca tcc att tac agg gga gtg act agg cat cac	1152
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His	
370 375 380	
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Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys	
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gat ctg tac ttg ggc acc ttt agc acc cag gag gaa gca gcg gag gcg	1248
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala	
405 410 415	
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Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn	
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Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala	
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Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala	
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Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly	
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Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Tyr	
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Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met			
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cgc ggg ggc ggg tgg tgc aag cag gag caa gac cac gcc gtg atc gcg			1632
Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala			
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gcc gcg cac agc ctg cag gac ctg cat cac ctg aac ctc ggg gcg gcc			1680
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala			
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gac gcc cac gac ttt ttc tcg gca ggg caa cag gcc gcg gcc gct gcc			1728
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala			
565	570	575	
atg cac ggc ctg ggt agc atc gat agt gcg tcg ctc gaa cac agc acc			1776
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr			
580	585	590	
ggc agc aac tcc gtc gtc tat aac ggc ggc gtc gac agc aat ggg			1824
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly			
595	600	605	
gcc agc gcg gtc ggc ggc tcc ggg ggt ggc tat atg atg ccc atg agc			1872
Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser			
610	615	620	
gct gcc ggt gca acg aca aca agc gca atg gtg agc cac gag caa gtc			1920
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val			
625	630	635	640
cat gca cgg gcc tac gac gaa gcc aag cag gct gct cag atg ggg tac			1968
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr			
645	650	655	
gag agc tac ctg gtc aac gcg gag aac aat ggt ggc gga aga atg tct			2016
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser			
660	665	670	
gca tgg ggg act gtc gtg tct gca gcc gcg gca gct gct tcc agc			2064
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser			
675	680	685	
aat gac aac atg gcc gcc gac gtg ggc cat ggg ggg ggc cag ctc ttt			2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe			
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1 5 10 15			
ctg ccc ccg tcc cag acg acg gat agc aca ctg att agc gcg gcc acc			96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr			

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agc atg aga gga agc gag ctt tcg gcc ctg gtc gcg gag ccc aag ctg Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu 50 55 60			192
gaa gat ttt ctg ggc ggc att agc ttc tcc gag cag cat cat aag gcg Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala 65 70 75 80			240
aat tgc aac atg ata ccg agc act tcc tcc act gtt tgt tac gcg agc Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser 85 90 95			288
agc ggt gct agc acg ggc tat cac cat caa ctg tac cac cag ccg acc Ser Gly Ala Ser Thr Gly His His Gln Leu Tyr His Gln Pro Thr 100 105 110			336
agc tca gcg ctc cac ttc gcg gat agc gta atg gtg gcc tcc tcg gcc Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala 115 120 125			384
ggt gtc cac gat ggg ggt gcc atg ctc tcc gcg gcc gct aat ggt Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly 130 135 140			432
gtc gct ggc gct gcc agt gcg aac ggc ggg ggg atc ggg ctc tcc atg Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met 145 150 155 160			480
att aag aac tgg ctg cgg agc cag ccg gcc ccc atg caa ccg agg gtg Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val 165 170 175			528
gcg gcc gca gaa ggc gcg cag ggg ctc tcc ttg tcc atg aac atg gcc Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala 180 185 190			576
ggc acc acg caa ggc gct gca ggg atg cct ctt ctc gct ggt gag cgg Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg 195 200 205			624
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agc atc tat cgt ggc gtc aca aga cat aga tgg act ggc agg tat gaa Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu 275 280 285			864
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gga cgt caa gtc tac ctg ggt ggg tac gat aaa gag gaa aaa gct gct Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala 305 310 315 320			960
aga gct tac gac ctt gct gtc aca aaa tac tgg gga gcg aca aca aca Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr			1008

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ggt ttc tcc aga ggt gct tcc atc tat aga gga gtc aca agg cat cat Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His 370 375 380			1152
caa cat gga aga tgg cag gct cgg atc gga cgc gtc gca ggg aac aag Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys 385 390 395 400			1200
gat ctt tat ctc ggc acg ttc acg acg caa gaa gaa gca gcc gaa gcg Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala 405 410 415			1248
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ctc ccc atc ggg agc gcc gcc aag cgg ctg aaa gag gcc gag gcc gct Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala 450 455 460			1392
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ggc gcg cac tac cat ggc gcc gcg tgg ccc acc atc gcg ttc caa ccg Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro 500 505 510			1536
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ggc tcc aat tcc gtg gtc tac aac ggc ggc gtg ggg gat agc aac ggc Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly 595 600 605			1824
gcc tcc gcc gtc ggc ggg tcc ggc ggt ggc tac atg atg ccg atg agc Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser 610 615 620			1872
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645	650	655		
gaa tcc tac ctg gtg aac gcg gag aat aat gga ggc gga aga atg tct				2016
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser				
660	665	670		
gct tgg ggg aca gtc gtg tcc gct gcc gcg gct gca gct agc agc				2064
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser				
675	680	685		
aat gac aat atg gcg gcg gac gtc ggc cac ggc ggc gcg cag ctg ttc				2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Gly Ala Gln Leu Phe				
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Ser Val Trp Asn Asp Thr *				
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1	5	10	15	
ctg ccg ccg tcc caa acg acc gac agc aca ctg att agc gcg gcg acg			96	
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr				
20	25	30		
gcg gat cac gtg agc ggc gat gtg tgc att ccc cag gac tgg			144	
Ala Asp His Val Ser Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp				
35	40	45		
tcc atg agg ggt agc gaa ctt agc gcg ctc gtg gcg gaa ccg aaa ctg			192	
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu				
50	55	60		
gag gac ttc ctg ggg ggc atc tcc ttt tcc gag cag cat cac aaa gcg			240	
Glu Asp Phe Leu Gly Ile Ser Phe Ser Glu Gln His His Lys Ala				
65	70	75	80	
aac tgt aac atg atc ccc agc act agc agc aca gtt tgt tat gcc tcc			288	
Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser				
85	90	95		
tca ggt gct tcc acg ggc tac cac cat cag ctg tat cac caa ccg acc			336	
Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr				
100	105	110		
agc tca gcg ctc cac ttt gcg gat tcc gta atg gtg gcc tcc agc gcc			384	
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala				
115	120	125		
gga gtc cac gac ggg ggt gcg atg ctc agc gcg gcc gct aat gga			432	
Gly Val His Asp Gly Ala Met Leu Ser Ala Ala Ala Asn Gly				
130	135	140		
gtg gca ggg gct gcg agt gcg aac ggc ggc att ggg ctc tcc atg			480	
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met				
145	150	155	160	

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atc aaa aat tgg ctg cgg tcc cag ccg gcg ccg atg cag ccc aga gtg Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val 165 170 175	528
gcc gcc gct gaa ggc gcc caa ggc ctg tcc ctc agc atg aac atg gcg Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala 180 185 190	576
ggg acg acc cag ggc gca gca ggg atg cca ctt ctc gca ggt gaa cgc Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg 195 200 205	624
gct cgc gcg ccc gag tcc gta agc acc agc gca cag gga ggt gcg gtg Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val 210 215 220	672
gtg gtc acg gcc ccg aag gaa gat tcc gga ggg agc gga gtg gcc ggg Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly 225 230 235 240	720
gca ctc gta gcc gtg agc acg gac acc gga ggg tcc ggg ggg gcc tcg Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser 245 250 255	768
gca gat aat acc gct aga aaa acg gtc gac acg ttt ggc cag cgc acg Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr 260 265 270	816
tcg atc tat aga ggc gtg aca agg cac aga tgg aca ggc aga tac gaa Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu 275 280 285	864
gct cac ctt tgg gat aac agt tgc aga agg gaa ggg caa act aga aag Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys 290 295 300	912
gga cgt cag gtc tat ctg gga ggc tac gac aaa gag gag aag gca gca Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala 305 310 315 320	960
aga gca tac gat ctg gct gca ctg aaa tac tgg gga gcc aca act act Arg Ala Tyr Asp Leu Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr 325 330 335	1008
act aat ttt cca gtg agt aac tac gag aaa gaa ctg gag gac atg aag Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys 340 345 350	1056
cac atg act aga caa gaa ttc gta gcg tct ctg agg aga aag agc tcc His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser 355 360 365	1104
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gat ctg tat ctc ggc acg ttt tcc acc cag gaa gaa gca gcc gag gcg Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala 405 410 415	1248
tac gac atc gcg gcc atc aaa ttt cgc ggc ctc aat gcc gtc acg aat Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn 420 425 430	1296
ttc gat atg agc cgc tat gac gtg aag tcc att ctc gat agc tcc gcg Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala 435 440 445	1344
ctc ccc atc ggg tcc gcg gcg aag cgc ctg aag gag gac gag gcg gct Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala 450 455 460	1392

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gcc tcc gcc caa cat cat cat gcc ggc gtc gtg tcc tac gat gtc ggg Ala Ser Ala Gln His His His Ala Gly Val Val Ser Tyr Asp Val Gly 465 470 475 480	1440
cg ^g atc g ^c g ^c a ^g c ^a g ^c ctg g ^g g ^g a ^g g ^c c ^t g ^c g ^c g ^c t ^t a ^t Arg Ile Ala Ser Gln Leu Gly Asp Gly Ala Leu Ala Ala Tyr 485 490 495	1488
g ^g g ^g c ^c c ^a t ^a c ^a c ^a g ^g g ^c g ^c t ^g c ^c a ^c g ^t a ^c g ^c t ^t t ^t c ^a g ^c c ^c Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro 500 505 510	1536
g ^g c ^c g ^c a ^g c ^a t ^t g ^c t ^c c ^a c ^c t ^t a ^t g ^c c ^a a ^a c ^c t ^t a ^t g ^c Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met 515 520 525	1584
cg ^g g ^g g ^g g ^g t ^g t ^g c ^a a ^a c ^a g ^g a ^g c ^a t ^t g ^c g ^t a ^t t ^t g ^c Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala 530 535 540	1632
g ^c g ^c c ^a t ^c c ^t c ^a g ^c a ^t g ^c c ^t a ^t c ^t g ^g g ^c g ^c g ^c Ala Ala His Ser Leu Gln Asp Leu His Leu Asn Leu Gly Ala Ala 545 550 555 560	1680
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at ^g c ^a t ^g g ^g c ^t g ^g a ^g c ^a t ^c g ^t t ^c g ^g a ^g c ^t a ^t c ^t a ^c g ^c Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr 580 585 590	1776
g ^g t ^c c ^a a ^t a ^g c ^t g ^t t ^t a ^t g ^g g ^g g ^g g ^c g ^c a ^g c ^a a ^a g ^g Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly 595 600 605	1824
g ^c a ^g c ^c g ^t g ^g g ^g a ^g t ^t g ^c g ^g g ^g t ^c a ^t g ^t a ^t g ^c a ^t g ^c Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser 610 615 620	1872
g ^c t ^t g ^g g ^g g ^c a ^c a ^c a ^t t ^c g ^c a ^t g ^t a ^g c ^a c ^a g ^g c ^a g ^c g ^t Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val 625 630 635 640	1920
c ^a t ^t g ^c g ^c g ^c t ^t a ^t g ^g a ^a g ^g g ^c a ^a c ^a g ^c a ^a a ^t g ^g g ^c t ^c His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr 645 650 655	1968
g ^a g ^c a ^t c ^t g ^t a ^a g ^c g ^a a ^a t ^a a ^c g ^g t ^t g ^g g ^g g ^t a ^g g ^t t ^c Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser 660 665 670	2016
g ^c t ^t g ^g g ^g a ^c a ^c g ^t g ^c t ^c g ^c Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser 675 680 685	2064
a ^a c ^a g ^a a ^t g ^c g ^c g ^c a ^c g ^t g ^g g ^g g ^g g ^c c ^a g ^t c ^t t ^c Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe 690 695 700	2112
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<223> OTHER INFORMATION: Variant of Zm-ODP2 having 76.1% nucleic acid
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ctg ccc ccg tcc caa acc acc gat tcc act ctg att agc gcg gcc acg	96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr	
20 25 30	
gcc gat cat gtg tcc ggc gat gtc tgc ttt aat att ccg cag gac tgg	144
Ala Asp His Val Ser Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp	
35 40 45	
agc atg aga gga tca gaa ctg agc gcc ctc gtg gcg gaa ccc aaa ctc	192
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu	
50 55 60	
gaa gat ttc ctg ggg ggg att agc ttc agc gaa cag cac cac aaa gcg	240
Glu Asp Phe Leu Gly Ile Ser Phe Ser Glu Gln His His Lys Ala	
65 70 75 80	
aat tgt aat atg ata ccg tcc act agc tcc aca gtt tgt tat gcc agc	288
Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser	
85 90 95	
agc gga gca agc acg ggg tac cac cat cag ctc tat cat caa ccc acg	336
Ser Gly Ala Ser Thr Gly His His Gln Leu Tyr His Gln Pro Thr	
100 105 110	
tcc agc gcc ctg cac ttt gcc gat agc gtt atg gtg gcg tcc agc gcg	384
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala	
115 120 125	
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Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly	
130 135 140	
gtg gca ggg gct gcc tcc gcg aat ggc ggg ggg atc ggg ctc agc atg	480
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met	
145 150 155 160	
att aaa aac tgg ctc cgc tcc caa ccc gcc ccg atg caa ccc aga gtc	528
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val	
165 170 175	
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Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg	
195 200 205	
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210 215 220	
gtg gtg acg gcg ccg aag gaa gac agc ggt ggg agc ggt gtg gcg ggg	720
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly	
225 230 235 240	
gct cta gta gcg gtc tcc acc gac acc gga ggc agc ggc ggc gcg tcg	768
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser	
245 250 255	
gct gat aat acc gct aga aag acg gtg gat acc ttc ggc cag cgg acc	816
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr	
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Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu	
275 280 285	

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ggt aga cag gtg tac tta ggt ggc tac gac aag gaa gaa aag gca gca Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala 305 310 315 320	960
agg gca tac gac ctg gca gct ctc aag tat tgg gga gcg act act aca Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr 325 330 335	1008
aca aat ttc cca gtc agt aac tat gag aaa gaa ctg gag gat atg aaa Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys 340 345 350	1056
cat atg aca aga caa gaa ttc gtt gcc tcc ctc agg aga aaa agc agt His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser 355 360 365	1104
gga ttc tcc agg ggt gct tcc atc tac agg ggt gtc aca agg cac cat Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His 370 375 380	1152
cag cat gga agg tgg cag gca cgc atc gga cgc gtt gca ggc aat aaa Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys 385 390 395 400	1200
gat ctg tac ctc ggg acg ttc tcc acg cag gaa gca gca gag gcg Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala 405 410 415	1248
tat gac att gcg gcc att aag ttt cgg ggc ctc aat gcg gtc acg aat Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn 420 425 430	1296
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ctc ccc att ggc tcc gcg aag cgc ctc aaa gaa gcc gaa gca gct Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala 450 455 460	1392
gcc agc gcg caa cac cat cat gcc ggg gtg gtc tcc tat gac gtc ggg Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly 465 470 475 480	1440
cgg att gcc tcg cag ctg ggg gat ggg ggt gcc ctg gcc gca tat Arg Ile Ala Ser Gln Leu Gly Asp Gly Ala Leu Ala Ala Ala Tyr 485 490 495	1488
ggg gcc cat tac cat ggc gcg gcc tgg ccg acg atc gcc ttt cag ccc Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro 500 505 510	1536
ggg gcg gcg agc act ggg ctg tac cat ccc tac gcg caa caa cct atg Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met 515 520 525	1584
cgc ggg ggg ggc tgg tgt aaa caa gaa cag gac cat gcc gtc att gcc Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala 530 535 540	1632
gcg gcc cac tcc ctc cag gac ctg cat cac ctg aat ctc ggg gcg gca Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala 545 550 555 560	1680
ggg gcc cat gat ttc ttc tcg gct ggc caa cag gcg gcg gca gca Gly Ala His Asp Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala 565 570 575	1728
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gca gcg ggt gca acc aca act tcg gct atg gtc tcc cat gaa caa gtg Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val 625 630 635 640	1920
cac gct cgg gcg tat gat gag gcg aaa caa gca gca caa atg ggg tat His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr 645 650 655	1968
gag tcc tat ctg gtg aat gcc gaa aat aac gga ggg ggt aga atg tcc Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser 660 665 670	2016
gct tgg ggg aca gtg gtc tcc gca gcg gcg gct gct gca agc tcc Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ser Ser 675 680 685	2064
aac gat aat atg gcc gcg gat gtg ggg cac ggg ggg gcc caa ctg ttc Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe 690 695 700	2112
agt gtg tgg aat gac aca taa Ser Val Trp Asn Asp Thr * 705 710	2133
<210> SEQ ID NO 11 <211> LENGTH: 2133 <212> TYPE: DNA <213> ORGANISM: Artificial Sequence <220> FEATURE: <223> OTHER INFORMATION: Variant of Zm-ODP2 having 70.6% nucleic acid sequence identity to SEQ ID NO:1 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO:2. <221> NAME/KEY: CDS <222> LOCATION: (1)...(2133) <400> SEQUENCE: 11	
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ctc ccc ccc tcc caa acg acc gat agc act ctg att agc gcg gcc acg Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr 20 25 30	96
gcc gat cac gtg agc ggg gac gtg tgt ttt aat att ccg cag gac tgg Ala Asp His Val Ser Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp 35 40 45	144
agc atg aga gga tca gaa ctt tcg gcc ctg gtg gcc gag ccc aaa ctc Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu 50 55 60	192
gaa gac ttt ctg ggg ggg att agc ttt agc gaa caa cac cac aaa gcc Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala 65 70 75 80	240
aat tgt aac atg atc ccg tcc act agc tcc aca gtg tgc tat gcc tcc Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser 85 90 95	288
agc gga gct tcc acg ggg tac cac cat caa ctc tat cat caa ccg acg Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr 100 105 110	336
agc agc gcc ctg cat ttt gcc gat agc gtt atg gtc gcc agc agc gcg	384

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Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala			
115	120	125	
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Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly			
130	135	140	
gtg gca ggc gca gcc tcc gcg aat ggg ggg ggg att ggc ctc agc atg			480
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met			
145	150	155	160
atc aaa aat tgg ctc cgc tcc cag ccc gcc ccg atg caa ccc aga gtc			528
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val			
165	170	175	
gcc gcc gca gag ggg gcc caa ggg ctg tct ctc agc atg aat atg gcg			576
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ggc acc acg cag ggg gca gca ggg atg cct ctc gca ggt gaa cgg			624
Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg			
195	200	205	
gca cgc gcc ccg gaa aat gtt agc acc tca gct caa gga ggt gcc gtc			672
Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val			
210	215	220	
gtg gtg acc gcg ccc aaa gaa gac tcc gga ggg tcc gga gtg gcc ggg			720
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly			
225	230	235	240
gct ctc gtt gcc gtg tcc acc gat acc ggt ggg tcc ggg ggc gcc agc			768
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser			
245	250	255	
gca gat aat acc gct aga aaa acg gtc gat acc ttt ggc caa cgg acg			816
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr			
260	265	270	
tcg atc tat aga ggg gtc act agg cac agg tgg aca ggc agg tac gag			864
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu			
275	280	285	
gca cac ctg tgg gac aat agt tgt agg aga gaa ggc cag aca aga aaa			912
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys			
290	295	300	
gga cgt cag gtc tat ctg gga ggg tac gac aag gaa aag gca gca			960
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala			
305	310	315	320
aga gca tat gac ctg gca gct ctg aaa tat tgg ggt gcg act act act			1008
Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr			
325	330	335	
act aat ttc cct gtc tcc aat tat gag aaa gaa ctg gaa gac atg aaa			1056
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys			
340	345	350	
cat atg act aga caa gaa ttt gtt gcc tcc ctc agg aga aaa tcc tcc			1104
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser			
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gga ttt agc agg ggt gct agc atc tat aga ggt gtc aca agg cac cat			1152
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His			
370	375	380	
caa cac gga aqa tgg cag gct cgc atc ggt cgc gtg gct ggc aat aaa			1200
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys			
385	390	395	400
gac ctg tat ctc ggg acg ttt tcc acg caa gaa gag gca gcc gaa gcc			1248
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala			
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Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala			
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Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala			
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Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly			
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cgg att gcg agc caa ctg ggg gat ggg ggt gcc ctc gcc gcc tat			1488
Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Ala Tyr			
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Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met			
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Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala			
530	535	540	
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545	550	555	560
ggg gcc cat gac ttc ttc tcg gtc ggc caa caa gcg gcg gca gca			1728
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala			
565	570	575	
atg cac ggg ctc gga tcc atc gac tcc gcc agc ctc gag cat tcc acc			1776
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr			
580	585	590	
ggg agc aat agc gtg gtg tat aat ggg ggc gtg ggc gat tcc aat ggg			1824
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly			
595	600	605	
gct tcc gcg gtc ggg ggg agt ggg gga ggg tat atg atg ccc atg tcc			1872
Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser			
610	615	620	
gca gcg ggt gct acg aca act tcg gca atg gtg tcc cat gaa caa gtc			1920
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val			
625	630	635	640
cac gct cgg gcg tat gat gag ggc aaa caa gca gca caa atg ggc tat			1968
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr			
645	650	655	
gaa agc tat ctc gtc aac gcc gaa aac aac gga ggg ggt aga atg tcc			2016
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser			
660	665	670	
gct tgg ggc act gtc gtc tcc gct ggc gcc gct gct gca tcc agc			2064
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser			
675	680	685	
aat gat aac atg gcg gcg gat gtg ggg cac ggg ggg gca ctc ttt			2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe			
690	695	700	
tcc gtg tgg aat gac aca taa			2133
Ser Val Trp Asn Asp Thr *			
705	710		

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<210> SEQ ID NO 12
<211> LENGTH: 2133
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Variant of Zm-ODP2 having the serine 37 codon
      altered from "tcc" to the threonine codon of
      "acc". The ORF encodes the amino acid sequence
      set forth in SEQ ID NO: 19.
<221> NAME/KEY: CDS
<222> LOCATION: (1)...(2133)

<400> SEQUENCE: 12

atg gcc act gtg aac aac tgg ctc gct ttc tcc ctc tcc ccg cag gag      48
Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu
1           5           10          15

ctg ccg ccc tcc cag acg acg gac tcc aca ctc atc tcg gcc gcc acc      96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr
20          25          30

gcc gac cat gtc acc ggc gat gtc tgc ttc aac atc ccc caa gat tgg      144
Ala Asp His Val Thr Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp
35          40          45

agc atg agg gga tca gag ctt tcg gcg ctc gtc gcg gag ccg aag ctg      192
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu
50          55          60

gag gac ttc ctc ggc ggc atc tcc ttc gag cag cat cac aag gcc      240
Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala
65          70          75          80

aac tgc aac atg ata ccc agc act agc agc aca gtt tgc tac gcg agc      288
Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser
85          90          95

tca ggt gct agc acc ggc tac cat cac cag ctg tac cac cag ccc acc      336
Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr
100         105         110

agc tca gcg ctc cac ttc gcg gac tcc gta atg gtg gcc tcc tcg gcc      384
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala
115         120         125

ggc gtc cac gac ggc ggt gcc atg ctc agc gcg gcc gct aac ggt      432
Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly
130         135         140

gtc gct ggc gct gcc agt gcc aac ggc ggc ggc atc ggg ctg tcc atg      480
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Gly Ile Gly Leu Ser Met
145         150         155         160

att aag aac tgg ctg cgg agc caa ccg gcg ccc atg cag ccg agg gtg      528
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val
165         170         175

gct ggc gct gag ggc gcg cag ggg ctc tct ttg tcc atg aac atg gcg      576
Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala
180         185         190

ggc acg acc caa ggc gct gct ggc atg cca ctt ctc gct gga gag cgc      624
Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg
195         200         205

gca cgg gcg ccc gag agt gta tcg acg tca gca cag ggt gga gcc gtc      672
Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val
210         215         220

gtc gtc acg gcg ccc aag gag gat agc ggt ggc agc ggt gtt gcc ggc      720
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly
225         230         235         240

gct cta gta gcc gtg agc acg gac acg ggt ggc agc ggc ggc gcg tcg      768
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser

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245	250	255	
gct gac aac acg gca agg aag acg gtg gac acg ttc ggg cag cgc acg Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr	260	265	816
270			
tcg att tac cgt ggc gtg aca agg cat aga tgg act ggg aga tat gag Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu	275	280	864
285			
gca cat ctt tgg gat aac agt tgc aga agg gaa ggg caa act cgt aag Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys	290	295	912
300			
ggt cgt caa gtc tat tta ggt ggc tat gat aaa gag gag aaa gct gct Gly Arg Gln Val Tyr Leu Gly Tyr Asp Lys Glu Glu Lys Ala Ala	305	310	960
315	320		
agg gct tat gat ctt gct gct ctg aag tac tgg ggt gcc aca aca aca Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr	325	330	1008
335			
aca aat ttt cca gtg agt aac tac gaa aag gag ctc gag gac atg aag Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys	340	345	1056
350			
cac atg aca agg cag gag ttt gta gcg tct ctg aga agg aag agc agt His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser	355	360	1104
365			
ggt ttc tcc aga ggt gca tcc att tac agg gga gtg act agg cat cac Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His	370	375	1152
380			
caa cat gga aga tgg caa gca cgg att gga cga gtt gca ggg aac aag Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys	385	390	1200
395	400		
gat ctt tac ttg ggc acc ttc agc acc cag gag gag gca gcg gag gcg Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala	405	410	1248
415			
tac gac atc gcg gcg atc aag ttc cgc ggc ctc aac gcc gtc acc aac Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn	420	425	1296
430			
ttc gac atg agc cgc tac gac gtg aag agc atc ctg gac agc agc gcc Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala	435	440	1344
445			
ctc ccc atc ggc agc gcc gcc aag cgc ctc aag gag gcc gag gcc gca Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala	450	455	1392
460			
gcg tcc gcg cag cac cac gac ggc ggc gtg gtg agc tac gac gtc ggc Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly	465	470	1440
475	480		
cgc atc gcc tcg cag ctc ggc gac ggc gga gcc ctg gcg gcg gcg tac Arg Ile Ala Ser Gln Leu Gly Asp Gly Ala Leu Ala Ala Tyr	485	490	1488
495			
ggc ggc cac tac cac ggc gcc tgg ccg acc atc gcg ttc cag ccg Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro	500	505	1536
510			
ggc gcc gcc agc aca ggc ctg tac cac ccg tac gcg cag cag cca atg Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met	515	520	1584
525			
cgc ggc ggc ggg tgg tgc aag cag gag cag gag gac cac gcg gtg atc gcg Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala	530	535	1632
540			
gcc gcg cac agc ctg cag gac ctc cac cac ctg aac ctg ggc gcg gcc Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala			1680

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545	550	555	560	
ggc gcg cac gac ttt ttc tcg gca ggg cag cag gcc gcc gct gcg				1728
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala				
565	570	575		
atg cac ggc ctg ggt agc atc gac agt gcg tcg ctc gag cac agc acc				1776
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr				
580	585	590		
ggc tcc aac tcc gtc gtc tac aac ggc ggg gtc ggc gac agc aac ggc				1824
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly				
595	600	605		
gcc agc gcc gtc ggc ggc agt ggc ggt ggc tac atg atg ccg atg agc				1872
Ala Ser Ala Val Gly Ser Gly Gly Tyr Met Met Pro Met Ser				
610	615	620		
gct gcc gga gca acc act aca tcg gca atg gtg agc cac gag cag cag gtg				1920
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val				
625	630	635	640	
cat gca cgg gcc tac gac gaa gcc aag cag gct gct cag atg ggg tac				1968
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr				
645	650	655		
gag agc tac ctg gtg aac gcg gag aac aat ggt ggc gga agg atg tct				2016
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser				
660	665	670		
gca tgg ggg act gtc gtg tct gca gcc gcg gca gca gca agc agc				2064
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser				
675	680	685		
aac gac aac atg gcc gcc gac gtc ggc cat ggc ggc gcg cag ctc ttc				2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe				
690	695	700		
agt gtc tgg aac gac act taa				2133
Ser Val Trp Asn Asp Thr *				
705	710			

<210> SEQ ID NO 13
 <211> LENGTH: 2133
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Variant of Zm-ODP2 having 97.3% nucleic acid sequence identity to SEQ ID NO:3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO:2 with a single amino acid alteration (i.e., S37 to T37). The ORF encodes the amino acid sequence set forth in SEQ ID NO: 19.
 <221> NAME/KEY: CDS
 <222> LOCATION: (1)...(2133)

<400> SEQUENCE: 13				
atg gcc act gtg aac aac tgg ctc gct ttc tcc ctc tcc ccg cag gag				48
Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu				
1	5	10	15	
ctg ccg ccc tcc caa acc acg gac tcc aca ctc atc tcg gcc gcc acc				96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr				
20	25	30		
gcc gac cat gtc acc ggc gat gtc tgc ttc aac atc ccc caa gat tgg				144
Ala Asp His Val Thr Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp				
35	40	45		
agc atg agg gga tca gag ctt tcg gcg ctc gtc gcg gag cgg aaa ctg				192
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu				
50	55	60		
gag gac ttc ctc ggg ggc att tcc ttc tcc gag cag cat cac aag gcc				240

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Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala	65	70	75	80	
aac tgc aac atg ata ccc tcc act agc tcc aca gtt tgc tac gcg agc					288
Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser					
85	90	95			
tca ggt gct agc acc ggc tac cat cac cag ctg tac cac cag ccc acc					336
Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr					
100	105	110			
tcc tca gcg ctc cac ttc gcg gac tcc gta atg gtc gac tcc tcg gac					384
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala					
115	120	125			
ggt gtc cac gac ggc ggt gcc atg ctc agc ggc gcc gct aac ggt					432
Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly					
130	135	140			
gtc gct ggc gca gcc agt gcc aac ggg ggc ggc atc ggg ctg tcc atg					480
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met					
145	150	155	160		
att aag aac tgg ctg cgg agc caa ccg gcg ccc atg cag ccg agg gtc					528
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val					
165	170	175			
gcg gcg gct gag ggc gcg cag ggg ctc tct ttg tcc atg aat atg gtc					576
Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala					
180	185	190			
ggg acg acc caa ggc gct gca ggc atg cca ctt ctc gct gga gag ggc					624
Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg					
195	200	205			
gca cgg gcg ccc gag agt gta tcc acg tca gca cag ggt gga gcc gtc					672
Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val					
210	215	220			
gtc gtc acc gcg ccc aag gag gat agc ggt ggc agc ggt gtt gcc ggc					720
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly					
225	230	235	240		
gct cta gta gcc gtc agc acg gac acg ggt ggc agc ggg ggc gcg tcg					768
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser					
245	250	255			
gct gac aac acg gca agg aag acg gtc gac acg ttt ggg cag ccg acg					816
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr					
260	265	270			
tcg atc tac cgt ggc gtc aca aga cat aga tgg act ggg aga tat gag					864
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu					
275	280	285			
gca cat ctt tgg gat aac agt tcc aga agg gaa ggg caa act cgt aag					912
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys					
290	295	300			
ggt cgt caa gtc tat tta ggt ggc tat gat aaa gag gag aaa gct gtc					960
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala					
305	310	315	320		
agg gct tat gac ctt gct ctc aag tac tgg ggt ggc aca aca aca					1008
Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr					
325	330	335			
aca aat ttc cca gtg agt aac tac gaa aag gag ctc gag qac atg aag					1056
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys					
340	345	350			
cac atg aca agg cag gag ttt gta gcg tct ctg aga agg aag agc tcc					1104
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser					
355	360	365			
ggt ttc tcc aga ggt gca tcc att tac agg gga gtc act agg cat cac					1152

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Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His			
370	375	380	
caa cat gga aga tgg caa gct cgg att gga cga gtt gca ggg aac aag		1200	
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys			
385	390	395	400
gat ctt tac ctc ggc acc ttc agc acc cag gag gaa gct gcg gag gcg		1248	
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala			
405	410	415	
tac gac atc gcg gcg atc aaa ttc cgc ggc ctc aac gcc gtc acc aac		1296	
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn			
420	425	430	
ttc gac atg agc cgc tac gac gtg aag agc atc ctg gac agc agc gcc		1344	
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala			
435	440	445	
ctg ccc atc ggc agc gcc gcc aag cgc ctg aag gag gac ggc gag gca		1392	
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala			
450	455	460	
gcg tcc gcg cag cac cac gcc ggc gtg gtg agc tac gac gtc ggg		1440	
Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly			
465	470	475	480
cgc atc gcc tcg cag ctc ggc gac ggc gga gcc ctg gcg gcg gcg tat		1488	
Arg Ile Ala Ser Gln Leu Gly Asp Gly Ala Leu Ala Ala Ala Tyr			
485	490	495	
ggc ggc cac tac cac ggg gcc gcc tgg ccg acc atc gcg ttc cag ccg		1536	
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro			
500	505	510	
ggc gcc gcc agc aca ggc ctg tac cac ccc tac gcg cag cag cca atg		1584	
Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met			
515	520	525	
cgc ggc ggc ggg tgg tgc aag cag gag cag gag gac cac gcg gtg atc gcg		1632	
Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala			
530	535	540	
gcg gcg cac agc ctg cag gac ctc cac cac ctg aac ctg ggc gcc gcc		1680	
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala			
545	550	555	560
ggc gcg cac gac ttt ttt tcg gca ggg cag cag gcc gcc gca gcc		1728	
Gly Ala His Asp Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala			
565	570	575	
atg cac ggc ctg ggt agc atc gac agt gcg tcg ctc gaa cac tcc acc		1776	
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr			
580	585	590	
ggc agc aac tcc gtc gtc tac aat ggc ggg gtc ggc gac agc aac ggc		1824	
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly			
595	600	605	
gcc agc gcc gtc ggc ggc tcc ggc gga ggc tat atg atg ccg atg agc		1872	
Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser			
610	615	620	
gct gcc gga gca acc aca aca tcg gca atg gtg agc cac gag cag gtc		1920	
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val			
625	630	635	640
cat gca cgg gcc tac gac gaa gcc aag cag gct gca cag atg ggg tac		1968	
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr			
645	650	655	
gag agc tac ctg gtg aac gcg gag aac aat ggt ggc gga agg atg tct		2016	
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser			
660	665	670	
gca tgg ggg act gtc gtg tct gca gcg gcg gca gca agc agc		2064	

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Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser
 675 680 685

aac gac aac atg gcc gac gtc ggc cat ggc ggc gcg cag ctc ttc 2112
 Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Gly Ala Gln Leu Phe
 690 695 700

agt gtc tgg aac gac act taa 2133
 Ser Val Trp Asn Asp Thr *
 705 710

<210> SEQ ID NO 14
 <211> LENGTH: 2133
 <212> TYPE: DNA
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Variant of Zm-ODP2 having 91.9% nucleic acid
 sequence identity to SEQ ID NO:3 (Zm-ODP2). The
 ORF encodes the amino acid sequence set forth in
 SEQ ID NO:2 with a single amino acid alteration
 (i.e., S37 to T37).
 <221> NAME/KEY: CDS
 <222> LOCATION: (1)...(2133)

<400> SEQUENCE: 14

atg gcc aca gtg aac aac tgg ctc gct ttt agc ctg agc ccg cag gaa	48
Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu	
1 5 10 15	
ctg ccg ccc tcc cag acc acg gac tcc act ctc atc tcg gcc gcc acc	96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr	
20 25 30	
gcc gat cat gtc acc ggc gac gtc tgt ttc aat att ccc caa gat tgg	144
Ala Asp His Val Thr Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp	
35 40 45	
tcc atg agg gga tca gag ctt tcg gcg ctg gtc gcg gaa ccg aaa ctg	192
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu	
50 55 60	
gaa gac ttc ctc ggc ggc atc tcc ttt tcc gaa cag cat cat aag gcc	240
Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala	
65 70 75 80	
aac tgc aac atg ata ccc agc act agc agc aca gtg tgc tac gcg agc	288
Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser	
85 90 95	
tca ggt gct tcc acc ggc tac cac cat caa ctc tac cac caa ccg acg	336
Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr	
100 105 110	
agc tca gcg ctc cat ttc gcc gat tcc gta atg gtc gcc tcc tcg gcc	384
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala	
115 120 125	
ggt gtc cac gac ggg ggt gcc atg ctc tcc gcg gcc gcc gct aac ggt	432
Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly	
130 135 140	
gtc gct ggc gct gcc agt gcg aac ggc ggc ggc atc ggg ctg agc atg	480
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Gly Ile Gly Leu Ser Met	
145 150 155 160	
atc aag aat tgg ctg cgg agc caa ccg gcg ccc atg caa ccg agg gtg	528
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val	
165 170 175	
gcc gcg gct gag ggg gcg cag ggg ctg tct ttg agc atg aat atg gcc	576
Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala	
180 185 190	
ggg acc acg caa ggg gct gct ggc atg cca ctt ctc gct ggt gag ccg	624

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Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg		
195 200 205		
gca cgc gcc ccc gag agt gtt tcg acg tca gca cag gga ggt gcg gtg	672	
Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val		
210 215 220		
gtc gtc acg gcg ccg aag gag gat agc ggt ggc acg ggt gtt gcg ggc	720	
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly		
225 230 235 240		
gct ctc gta gcc gtg agc acg gac acg ggt ggc acg ggc ggc gcg tcg	768	
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser		
245 250 255		
gct gac aac acg gca agg aag acg gtg gac acg ttc ggg cag cgg acg	816	
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr		
260 265 270		
tcg att tac cgt ggc gtg aca agg cat aga tgg act ggg agg tat gag	864	
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu		
275 280 285		
gca cat ctt tgg gat aac agt tgc aga agg gag ggg caa act cgt aag	912	
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys		
290 295 300		
ggt aga cag gtc tac ctg ggt ggc tat gat aaa gag gag aag gct gct	960	
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala		
305 310 315 320		
agg gct tat gat ctt gct gca ctg aag tac tgg ggt gcc act act aca	1008	
Arg Ala Tyr Asp Leu Ala Ala Lys Tyr Trp Gly Ala Thr Thr Thr		
325 330 335		
aca aac ttt cct gtc agt aac tat gaa aag gag ctc gag gac atg aag	1056	
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys		
340 345 350		
cac atg aca agg caa gaa ttt gtt gcg tct ctg aga agg aag agc agt	1104	
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser		
355 360 365		
ggt ttc tcc aga ggt gca tcc att tac agg gga gtg act agg cat cac	1152	
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His		
370 375 380		
caa cat ggt aga tgg cag gca cgc att ggt cga gtt gca ggg aac aaa	1200	
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys		
385 390 395 400		
gat ctg tat ttg ggc acc ttt agc acc caa gag gag gca gcc gag gcg	1248	
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala		
405 410 415		
tac gac atc gcg gcg atc aaa ttc cgc ggc ctc aac gcc gtc acg aac	1296	
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn		
420 425 430		
ttc gat atg agc cgc tac gac gtc aag agc atc ctg gac agc agc gcc	1344	
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala		
435 440 445		
ctc ccg atc ggc agc gcc gcg aaa cgc ctc aag gag gcc gag gcc gca	1392	
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala		
450 455 460		
gcg tcc gcg cag cac cac cac gcc ggc gtg gtc agc tac gac gtc ggc	1440	
Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly		
465 470 475 480		
cgc att gcc tcg cag ctc ggc gac ggc gga gcc ctg gcc gcg ggc tac	1488	
Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Tyr		
485 490 495		
ggg gcg cac tac cac ggg gcc tgg ccg acc atc gcc ttt cag ccg	1536	

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Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro			
500	505	510	
ggc gcc gcc agc aca ggc ctg tac cat ccg tac gcg caa caa cca atg	1584		
Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met			
515	520	525	
cgc ggc ggc ggg tgg tgc aaa cag gag cag gac cac gcg gtc att gcg	1632		
Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala			
530	535	540	
gcg gcc cat agc ctg cag gac ctc cac cac ctc aac ctg ggc gcg gcg	1680		
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala			
545	550	555	560
ggc gcg cac gac ttt ttc tcg gca ggg caa cag gcc gcg gcc gct gcg	1728		
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala			
565	570	575	
atg cac ggc ctg ggt agc att gac tcc gcg tcg ctg gag cac agc acg	1776		
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr			
580	585	590	
ggc agc aac tcc gtc gtg tac aac ggc ggc gtg ggc gac agc aac ggc	1824		
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly			
595	600	605	
gcc agc gcc gtc ggg ggg agt ggg ggt ggg tac atg atg ccc atg agc	1872		
Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser			
610	615	620	
gca gcc gga gca acc act act agc gca atg gtg agc cat gag cag gtg	1920		
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val			
625	630	635	640
cat gca cgg gcc tac gac gag gcc aaa cag gca gca caa atg ggg tat	1968		
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr			
645	650	655	
gag agc tac ctg gtc aat gcc gag aac aat ggt ggg ggt aga atg tct	2016		
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser			
660	665	670	
gca tgg ggc act gtc gtg tct gca gcc gcg gca gca gct agc tcc	2064		
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser			
675	680	685	
aac gat aac atg gcc gcc gac gtc ggc cat ggc ggg ggc caa ctc ttt	2112		
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe			
690	695	700	
tcc gtg tgg aac gat act taa	2133		
Ser Val Trp Asn Asp Thr *			
705	710		
<210> SEQ ID NO 15			
<211> LENGTH: 2133			
<212> TYPE: DNA			
<213> ORGANISM: Artificial Sequence			
<220> FEATURE:			
<223> OTHER INFORMATION: Variant of Zm-ODP2 having 86.6% nucleic acid sequence identity to SEQ ID NO:3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO:2 with a single amino acid alteration (i.e., S37 to T37).			
<221> NAME/KEY: CDS			
<222> LOCATION: (1)...(2133)			
<400> SEQUENCE: 15			
atg gcg act gtg aat aat tgg ctc gct ttc tcc ctc tcc ccc cag gag	48		
Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu			
1	5	10	15
ctc ccc ccc tcc cag acc acg gat agc aca ctc atc tcg gcc gcc acc	96		

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Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr		
20	25	30
gcg gat cat gtc acc ggc gac gtc tgt ttc aac att ccg cag gac tgg	144	
Ala Asp His Val Thr Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp		
35	40	45
agc atg aga ggt tca gag ctt agc gcg ctc gtc gcg gaa ccg aaa ctc	192	
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu		
50	55	60
gag gac ttt ctc ggc ggc atc tcc ttc tcc gag cag cat cac aaa gcg	240	
Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala		
65	70	75
80		
aac tgt aac atg atc ccc agc act agc tcc act gtt tgc tac gcc agc	288	
Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser		
85	90	95
tca ggt gct agc acg ggc tat cat cac cag ctg tat cat cag ccc acc	336	
Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr		
100	105	110
agc tca gcg ctg cat ttt gcc gat agc gta atg gtg gcg tcc tcg gcg	384	
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala		
115	120	125
ggt gtg cac gac ggg gga gcc atg ctc agc gcg gcc gcg gct aat ggt	432	
Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Ala Asn Gly		
130	135	140
gtc gca ggc gca gcg tcc gcg aac ggc ggg ggc att ggg ctg tcc atg	480	
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met		
145	150	155
160		
att aaa aac tgg ctg cgc agc cag ccg gcg ccc atg caa ccg aga gtg	528	
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val		
165	170	175
gcc gcg gca gaa ggc gcg caa ggc ctc tcc ctc agc atg aac atg gcc	576	
Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala		
180	185	190
ggg acc acg cag ggc gct gca ggg atg cca ctg ctg gca ggt gaa cgg	624	
Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg		
195	200	205
gca cgg gcg ccc gaa agt gta agc acg tca gca cag ggt gga gcc gtc	672	
Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val		
210	215	220
gtc gtc acg gcg ccg aag gag gac tcc ggt ggc agc ggt gtg gcg ggc	720	
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly		
225	230	235
240		
gca ctc gtt gcc gtg agc acc gat acg ggt ggc agc ggg ggc gcc agc	768	
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser		
245	250	255
gca gac aac acc gca agg aag acg gtc gac acc ttc ggg caa ccg acg	816	
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr		
260	265	270
agc att tac cgt ggg gtg aca aga cac agg tgg aca ggg aga tat gag	864	
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu		
275	280	285
gct cac ctg tgg gat aat tcc tgc aga agg gag ggc caa act cgt aag	912	
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys		
290	295	300
ggt cgt caa gtg tat tta gga ggg tat gat aaa gag gag aaa gct gct	960	
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala		
305	310	315
320		
aga gct tat gat ctt gct gct ctg aag tac tgg ggt gcc aca act aca	1008	

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Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr		
325	330	335
aca aac ttt cca gtg tcc aac tat gag aag gag ctc gaa gac atg aag	1056	
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys		
340	345	350
cat atg aca agg caa gaa ttt gtt gcg tcc ctg agg aga aag tcc agt	1104	
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser		
355	360	365
gga ttc tcc agg gga gct agc atc tat agg gga gtc aca agg cat cac	1152	
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His		
370	375	380
caa cac gga aga tgg caa gct cgc att ggt cga gtt gct ggc aac aag	1200	
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys		
385	390	395
400		
gat ctt tac ttg ggc acg ttt agc acc caa gag gag gca gcg gaa gcg	1248	
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala		
405	410	415
tat gat atc gcc gcg atc aaa ttc cgc ggg ctg aat gcc gtc acg aac	1296	
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn		
420	425	430
ttc gac atg tcc cgc tac gat gtg aag agc att ctc gac agc agc gcg	1344	
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala		
435	440	445
ctg ccg atc ggg agc gcc gcg aag cgc ctg aag gaa gcg gag gcc gct	1392	
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala		
450	455	460
gcc tcc gcc cag cat cat cac gcc ggg gtg gtg tcc tac gat gtc ggc	1440	
Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly		
465	470	475
480		
cgc att gcc tcg cag ctc ggg gac ggg gga gcc ctc gcg gcc tac	1488	
Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Tyr		
485	490	495
ggc gcc cac tat cac ggc gcc tgg ccg acc atc gcg ttc caa ccg	1536	
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro		
500	505	510
ggc gcc ggc act ggg ctc tat cat ccg tat gcc cag caa cct atg	1584	
Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met		
515	520	525
cgc ggc ggc ggg tgg tgc aaa cag gag caa gat cac gcc gtc att gcg	1632	
Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala		
530	535	540
gcg gcg cac agc ctc cag gac ctg cat cac ctg aac ctg ggc gcg gcg	1680	
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala		
545	550	555
560		
ggc gcg cat gac ttt ttc tcg gct ggg cag caa gcc gcg gcc gca gcg	1728	
Gly Ala His Asp Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala		
565	570	575
atg cat ggc ctg ggt tcc atc gat tcc gcg tcg ctg gag cac agc acc	1776	
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr		
580	585	590
ggc tcc aac tcc gtc gtg tat aac ggc ggc gtg ggg gac agc aat ggc	1824	
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly		
595	600	605
gct gcc gga gct acc act act tcg gca atg gtg tcc cac gag cag gtg	1920	
610	615	620

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Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val	
625 630 635 640	
cat gct cgc gcc tac gac gaa gcg aag cag gca gct caa atg ggc tat	1968
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr	
645 650 655	
gaa agc tac ctc gtg aat gcg gaa aac aac gga ggc gga agg atg tct	2016
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser	
660 665 670	
gca tgg ggc act gtc gtg tcc gca gcc gcc gcg gct gct gct agc agc	2064
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ser Ser	
675 680 685	
aac gac aat atg gcc gcc gac gtc ggc cat ggc ggc gcg cag ctc ttc	2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe	
690 695 700	
agt gtg tgg aat gat act taa	2133
Ser Val Trp Asn Asp Thr *	
705 710	

<210> SEQ ID NO 16

<211> LENGTH: 2133

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Variant of Zm-ODP2 having 81.5% nucleic acid sequence identity to SEQ ID NO:3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO:2 with a single amino acid alteration (i.e., S37 to T37).

<221> NAME/KEY: CDS

<222> LOCATION: (1)...(2133)

<400> SEQUENCE: 16

atg gcg act gtc aac aat tgg ctg gca ttc agc ctg tcc ccc caa gag	48
Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu	
1 5 10 15	
ctg ccc ccg agc caa acg acc gac agc aca ctc atc tcg gcc gcg acc	96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr	
20 25 30	
gcg gac cac gtc acg ggc gac gtc tgc ttc aat att ccg cag gac tgg	144
Ala Asp His Val Thr Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp	
35 40 45	
agc atg agg ggt tca gag ctg tcg gcg ctg gtg gcg gaa ccc aag ctc	192
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu	
50 55 60	
gaa gat ttc ctc ggg ggg atc agc ttt agc gag cag cat cat aaa gcg	240
Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala	
65 70 75 80	
aac tgc aat atg atc ccc tcc act agc tcc act gtt tgt tat gcg tcc	288
Asn Cys Asn Met Ile Pro Ser Thr Ser Thr Val Cys Tyr Ala Ser	
85 90 95	
tca gga gca agc acg ggg tac cac cat caa ctg tat cac caa ccg acg	336
Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr	
100 105 110	
tcc tca gcc ctc cat ttc gcc gac tcc gtt atg gtc gcc agc tcg gcg	384
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala	
115 120 125	
ggt gtg cat gac ggg ggt gcg atg ctc agc gcc gcc gcc gct aat ggt	432
Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly	
130 135 140	
gtc gca ggc gct gcg tcc gcc aac ggg ggg ggg atc ggc ctg tcc atg	480

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Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Gly Ile Gly Leu Ser Met			
145	150	155	160
att aag aat tgg ctg cgc tcc caa ccg gcc ccc atg cag ccc aga gtc			528
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val			
165	170	175	
gcc gcc gca gaa ggg gcc cag ggc ctc tct ctc tcc atg aat atg gcg			576
Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala			
180	185	190	
ggg acc acg cag ggg gca gca ggc atg cct ctg ctg gct gga gaa cgc			624
Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg			
195	200	205	
gca cgg gcc ccc gag agt gtt agc acg agc gct cag ggt ggt gcc gtc			672
Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val			
210	215	220	
gtg gtg acc gcc ccg aaa gag gac tcc gga ggc tcc gga gtt gcc ggc			720
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly			
225	230	235	240
gct cta gtt gcc gtg agc acg gat acg ggt ggc tcc ggc ggg gcg agc			768
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser			
245	250	255	
gct gat aat acc gca aga aag acc gtc gac acc ttt ggg cag cgc acg			816
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr			
260	265	270	
tcg atc tac aga ggc gtc act aga cat agg tgg aca ggc aga tac gaa			864
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu			
275	280	285	
gca cac ctt tgg gat aac agt tgt agg agg gaa ggc caa aca cgt aaa			912
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys			
290	295	300	
ggt aga caa gtc tat tta gga ggc tac gat aag gaa gag aag gtc gca			960
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala			
305	310	315	320
agg gca tac gac ctt gct gca ctc aag tat tgg ggt gcc aca act act			1008
Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr			
325	330	335	
aca aac ttt cca gtg agt aac tac gaa aaa gaa ctc gag gat atg aaa			1056
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys			
340	345	350	
cac atg act agg cag gag ttt gta gcc tcc ctc aga aga aaa tcc tcc			1104
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser			
355	360	365	
gga ttt agc agg ggt gct tcc att tac aga gga gtg aca aga cac cac			1152
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His			
370	375	380	
cag cat ggt agg tgg cag gca cgg att gga cga gtg gca ggc aac aaa			1200
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys			
385	390	395	400
gac ctt tat ctc ggc acc ttt agc acg cag gaa gag gca gcg gag gcg			1248
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala			
405	410	415	
tac gac att gcc gcg att aaa ttc cgg ggc ctc aat gcg gtc acg aac			1296
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn			
420	425	430	
ttt gat atg tcc cgc tat gat gtg aaa agc atc ctc gac agc agc gcc			1344
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala			
435	440	445	
ctc ccc att ggc agc gcg gcg aaa cgg ctc aaa gaa gcg gaa gcg gct			1392

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Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala			
450	455	460	
gcc agc gcc cag cac cat cat gcc ggg gtg gtc agc tac gat gtg ggg			1440
Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly			
465	470	475	480
cgc atc gcg agc caa ctg ggc gat ggg ggt gcc ctc gcc gcg gcc tat			1488
Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Ala Tyr			
485	490	495	
ggc gcc cat tac cat ggc gcc gcg tgg ccg acc atc gcc ttc caa ccc			1536
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro			
500	505	510	
gac gcc gcc agc act ggc ctc tac cac ccc tat gcc caa caa cca atg			1584
Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met			
515	520	525	
cgc ggc ggc ggc tgg tgt aag cag gag caa gat cat gcc gtg att gcg			1632
Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala			
530	535	540	
gcg gcg cac tcc ctc cag gac ctg cat cac ctg aat ctg ggc gcc gcg			1680
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala			
545	550	555	560
ggg gcc cat gat ttc ttt agc gct ggg cag caa gcg gcc gca gca gcc			1728
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala			
565	570	575	
atg cac ggg ctc ggt agc att gac agt gcc tcg ctg gaa cat agc acg			1776
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr			
580	585	590	
ggg agc aac tcc gtg gtc tac aac ggg ggc gtg ggc gat agc aac ggc			1824
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly			
595	600	605	
gcc agc gcg gtg ggc ggc agt ggc ggt ggc tat atg atg ccg atg agc			1872
Ala Ser Ala Val Gly Ser Gly Gly Tyr Met Met Pro Met Ser			
610	615	620	
gct gcg gga gct acg aca act agc gca atg gtc tcc cac gag caa gtc			1920
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val			
625	630	635	640
cac gct cgc gcg tat gat gaa gcc aaa cag gca gct cag atg ggc tac			1968
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr			
645	650	655	
gaa tcc tac ctg gtg aat gcc gaa aat aac ggt ggc gga aga atg tcc			2016
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser			
660	665	670	
gct tgg ggc aca gtg gtg tct gca gcc gcc gcg gct gct gca tcc agc			2064
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser			
675	680	685	
aat gac aac atg gcg gcc gac gtg ggc cat ggc ggg gcg caa ctc ttt			2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Gly Ala Gln Leu Phe			
690	695	700	
agt gtc tgg aat gac act taa			2133
Ser Val Trp Asn Asp Thr *			
705	710		

<210> SEQ ID NO 17

<211> LENGTH: 2133

<212> TYPE: DNA

<213> ORGANISM: Artificial Sequence

<220> FEATURE:

<223> OTHER INFORMATION: Variant of Zm-ODP2 having 75.9% nucleic acid sequence identity to SEQ ID NO:3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in

-continued

SEQ ID NO:2 with a single amino acid alteration
(i.e., S37 to T37).
<221> NAME/KEY: CDS
<222> LOCATION: (1)...(2133)

<400> SEQUENCE: 17

atg	gcg	act	gtc	aat	aat	tgg	ctg	gca	ttc	agc	ctg	ago	ccg	caa	gag	48	
Met	Ala	Thr	Val	Asn	Asn	Trp	Leu	Ala	Phe	Ser	Leu	Ser	Pro	Gln	Glu		
1	5			10			15										
ctg	ccc	ccg	tcc	caa	acg	acg	gat	tcc	act	ctg	att	agc	gcg	gcc	acg	96	
Leu	Pro	Pro	Ser	Gln	Thr	Thr	Asp	Ser	Thr	Leu	Ile	Ser	Ala	Ala	Thr		
20	25				25			30									
gcc	gat	cac	gtg	acg	ggg	gac	gtg	tgt	ttt	aac	att	ccc	cag	gat	tgg	144	
Ala	Asp	His	Val	Thr	Gly	Asp	Val	Cys	Phe	Asn	Ile	Pro	Gln	Asp	Trp		
35	40			40			45										
tcc	atg	aga	gga	tca	gag	ctg	tcg	gcc	ctg	gtg	gcc	gag	ccg	aaa	ctc	192	
Ser	Met	Arg	Gly	Ser	Glu	Leu	Ser	Ala	Leu	Val	Ala	Glu	Pro	Lys	Leu		
50	55			55			60										
gaa	gat	ttt	ctc	ggc	ggg	att	agc	ttt	agc	gaa	caa	cac	cat	aaa	gcg	240	
Glu	Asp	Phe	Leu	Gly	Gly	Ile	Ser	Phe	Ser	Glu	Gln	His	His	Lys	Ala		
65		70				75		75		80							
aac	tgc	aac	atg	atc	ccc	agc	aca	tcc	tcc	act	gtt	tgc	tat	gcc	agc	288	
Asn	Cys	Asn	Met	Ile	Pro	Ser	Thr	Ser	Ser	Thr	Val	Cys	Tyr	Ala	Ser		
85			85		90			90		95							
agc	gga	gca	tcc	acc	ggg	tat	cac	cat	caa	ctc	tat	cat	cag	ccc	acg	336	
Ser	Gly	Ala	Ser	Thr	Gly	Tyr	His	His	Gln	Leu	Tyr	His	Gln	Pro	Thr		
100			100		105			105		110							
agc	agc	gcc	ctg	cac	ttt	gcc	gat	agc	gta	atg	gtc	gcg	agc	agc	gcg	384	
Ser	Ser	Ala	Leu	His	Phe	Ala	Asp	Ser	Val	Met	Val	Ala	Ser	Ser	Ala		
115			115		120			120		125							
ggt	gtg	cat	gac	ggg	gga	gcg	atg	ctg	tcc	gcc	gcg	gct	aat	ggt		432	
Gly	Val	His	Asp	Gly	Gly	Ala	Met	Leu	Ser	Ala	Ala	Ala	Asn	Gly			
130			130		135			135		140							
gtg	gca	ggg	gca	gct	aat	ggg	ggg	ggc	att	ggc	ctc	agc	atg			480	
Val	Ala	Gly	Ala	Ala	Ser	Ala	Asn	Gly	Gly	Gly	Ile	Gly	Leu	Ser	Met		
145			145		150			150		155		160					
atc	aaa	aac	tgg	ctc	ccg	tcc	caa	ccg	gcg	ccc	atg	cag	ccc	aga	gtc	528	
Ile	Lys	Asn	Trp	Leu	Arg	Ser	Gln	Pro	Ala	Pro	Met	Gln	Pro	Arg	Val		
165			165		170			170		175							
gcg	gcc	gca	gaa	ggc	gcc	caa	ggc	ctg	tcc	ctc	agc	atg	aat	atg	gcg	576	
Ala	Ala	Ala	Glu	Gly	Ala	Gln	Gly	Leu	Ser	Leu	Ser	Met	Asn	Met	Ala		
180			180		185			185		190							
ggc	acc	acg	cag	ggg	gct	gca	ggg	atg	cct	ctg	ctg	gct	ggt	gag	cg	624	
Gly	Thr	Thr	Gln	Gly	Ala	Ala	Gly	Met	Pro	Leu	Leu	Ala	Gly	Glu	Arg		
195			195		200			200		205							
gct	ccg	ccg	ccg	aaa	gag	gac	agc	ggt	ggg	tcc	gga	gtg	gct	ggg		672	
Ala	Arg	Ala	Pro	Glu	Ser	Val	Ser	Thr	Ser	Ala	Gln	Gly	Gly	Ala	Val		
210			210		215			215		220							
gtg	gtg	acc	gcc	ccc	aaa	gag	gac	agc	ggt	ggg	tcc	gga	gtg	gct	ggg	720	
Val	Val	Thr	Ala	Pro	Lys	Glu	Asp	Ser	Gly	Gly	Ser	Gly	Ser	Gly	Ala	Gly	
225			225		230			230		235		240					
gca	ctc	gtt	gct	tcc	acc	gat	acc	gga	ggg	agc	ggg	ggg	gcc	tcc		768	
Ala	Leu	Val	Ala	Val	Ser	Thr	Asp	Thr	Gly	Gly	Ser	Gly	Gly	Ala	Ser		
245			245		250			250		255							
gca	gat	aac	acc	gct	aga	aag	acc	gtc	gac	acc	ttt	ggc	cag	ccg	acg	816	
Ala	Asp	Asn	Thr	Ala	Arg	Lys	Thr	Val	Asp	Thr	Phe	Gly	Gln	Arg	Thr		
260			260		265			265		270							
agc	atc	tac	aga	ggc	gtc	aca	aga	cac	aga	tgg	act	ggc	agg	tac	gaa	864	

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Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu		
275	280	285
gca cac ctt tgg gac aac agt tgt agg aga gag ggc caa aca aga aaa		912
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys		
290	295	300
gga aga cag gtg tat tta gga ggc tac gac aag gaa aag gca gca		960
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala		
305	310	315
agg gca tat gac ctg gca gca ctc aaa tat tgg gga gcc act aca act		1008
Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr		
325	330	335
aca aac ttt cct gtc agt aac tac gaa aaa gaa ctc gag gat atg aaa		1056
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys		
340	345	350
cac atg act aga cag gaa ttc gta gcc tcc ctc agg aga aag tcc agt		1104
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser		
355	360	365
gga ttc agc aga gga gca tcc atc tat aga ggt gtg act aga cac cac		1152
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His		
370	375	380
caa cac ggt agg tgg caa gct cgc atc ggt cgc gtg gct ggc aat aaa		1200
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys		
385	390	395
gat ctg tat ctc ggg acc ttt agc acg caa gaa gag gct gcc gaa gcc		1248
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala		
405	410	415
tac gac att gcg gcc att aaa ttt cgc ggg ctc aat gcg gtc acc aac		1296
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn		
420	425	430
ttt gat atg tcc cgc tac gat gtg aag tcc atc ctc gac agc tcc gcc		1344
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala		
435	440	445
ctc ccg atc ggg tcc gcc gcg aaa cgc ctg aaa gag gcg gaa gcg gct		1392
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala		
450	455	460
gcc tcc gcc caa cat cat cat gcg ggg gtc gtc tcc tac gac gtg ggc		1440
Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly		
465	470	475
480		
cgg atc gcg agc cag ctg ggg gat ggc ggt gcg ctg gcc gcc tat		1488
Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Tyr		
485	490	495
ggc gcc cac tat cac ggg gcg gcg tgg ccc acg att gcg ttt caa ccg		1536
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro		
500	505	510
ggg gcg gcg agc act ggg ctg tac cat ccc tat gcg caa caa cca atg		1584
Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met		
515	520	525
cgc ggg ggg ggc tgg tgc aaa caa gaa cag gat cat gac gtc att gcc		1632
Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala		
530	535	540
gac gcg cac aac ctg caa gac ctc cat cat ctc aac ctc ggc gcc gcg		1680
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala		
545	550	555
560		
ggc gcg cac gat ttc ttc tcg gct ggg cag caa gcg gcc gcg gct gcc		1728
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala		
565	570	575
atg cat ggc ctc gga tcc atc gac tcc gcc agc ctg gaa cac agc acc		1776

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Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr	
580 585 590	
ggg tcc aac agc gtc gtg tat aac ggg ggg gtc ggg gac tcc aat ggc	1824
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly	
595 600 605	
gcg agc gcg gtg ggg ggg agt ggc gga ggg tat atg atg ccc atg agc	1872
Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser	
610 615 620	
gct gcg gga gct acg aca aca tcg gct atg gtc agc cat gaa caa gtc	1920
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val	
625 630 635 640	
cat gct cgg gcc tat gat gag gcg aaa caa gca gca caa atg ggg tac	1968
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr	
645 650 655	
gag tcc tat ctc gtc aat gcc gaa aat aat ggt ggc gga agg atg tcc	2016
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser	
660 665 670	
gca tgg ggg aca gtg gtc tcc gct gcc gcg gct gca gct tcc tcc	2064
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser	
675 680 685	
aat gat aat atg gcg gcg gat gtc ggg cac ggg ggg gcc cag ctg ttt	2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe	
690 695 700	
tcc gtg tgg aac gat act taa	2133
Ser Val Trp Asn Asp Thr *	
705 710	

<210> SEQ ID NO 18
<211> LENGTH: 2133
<212> TYPE: DNA
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Variant of Zm-ODP2 having 70.4% nucleic acid sequence identity to SEQ ID NO:3 (Zm-ODP2). The ORF encodes the amino acid sequence set forth in SEQ ID NO:2 with a single amino acid alteration (i.e., S37 to T37).
<221> NAME/KEY: CDS
<222> LOCATION: (1) . . . (2133)

<400> SEQUENCE: 18	
atg gcg aca gtg aat aat tgg ctg gca ttt agc ctg agc ccg caa gaa	48
Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu	
1 5 10 15	
ctc ccg ccg agc caa acc acc gat tcc act ctg att tcg gcg gcc acc	96
Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr	
20 25 30	
gcg gac cac gtg acg ggg gac gtg tgt ttc aat att ccg caa gac tgg	144
Ala Asp His Val Thr Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp	
35 40 45	
tcc atg aga gga agc gaa ctg agc gcc ctg gtg gcc gaa ccc aaa ctc	192
Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu	
50 55 60	
gaa gac ttt ctg ggg ggg att agc ttc agc gaa caa cac cat aag gcc	240
Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala	
65 70 75 80	
aat tgt aat atg atc ccg tcc act tcc tcc act gtg tgt tat gcc tcc	288
Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser	
85 90 95	
agc gga gca agc acg ggc tat cac cat cag ctc tac cat caa ccg acg	336

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Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr			
100	105	110	
tcc tca gcc ctc cac ttt gcc gat agc gtt atg gtc gcg agc agc gcg		384	
Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala			
115	120	125	
ggt gtg cat gat ggg gga gcg atg ctg tcc gcc gcg gca aat gga		432	
Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly			
130	135	140	
gtg gca ggg gca gcc agt gcg aat ggg ggg ggg att ggc ctc agc atg		480	
Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met			
145	150	155	160
atc aag aat tgg ctc cgc tcc caa ccg gcc ccc atg caa ccg aga gtc		528	
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val			
165	170	175	
gcc gcc gca gaa ggg gcc caa ggc ctg tcc ctc agc atg aac atg gcc		576	
Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala			
180	185	190	
ggg acg acg cag ggg gca gca ggg atg cct ctg ctg gca gga gaa cgg		624	
Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg			
195	200	205	
gca cgc gcc ccg gaa agt gtt agc acc agc gct caa gga ggt gcg gtg		672	
Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val			
210	215	220	
gtg gtg acc gcc ccc aag gaa gat tcc gga ggg tcc gga gtg gcg ggg		720	
Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly			
225	230	235	240
gca ctc gtt gcg gtc tcc acc gat acc ggt ggg tcc ggg ggg gcc agc		768	
Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser			
245	250	255	
gca gat aat acc gct aga aaa acc gtc gat acc ttt ggc caa cgc acc		816	
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr			
260	265	270	
agc atc tac aga ggg gtc act aga cac agg tgg aca ggc aga tac gaa		864	
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu			
275	280	285	
gct cac ctg tgg gac aat agt tgt agg aga gag ggc cag aca aga aaa		912	
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys			
290	295	300	
ggt aga cag gtg tac ctg gga ggc tac gac aaa gaa gaa aag gct gca		960	
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala			
305	310	315	320
aga gca tac gac ctg gca gct ctc aaa tac tgg gga gcg aca act aca		1008	
Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr			
325	330	335	
act aac ttc cct gtc tcc aat tat gag aaa gag ctc gaa gat atg aag		1056	
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys			
340	345	350	
cat atg act aga caa gaa ttt gtt gcg tcc ctc agg aga aaa tcc agt		1104	
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser			
355	360	365	
gga ttt agc agg gga gct agc atc tat aga ggt gtg aca aga cac cac		1152	
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His			
370	375	380	
cag cac ggt agg tgg caa gct cgc atc gga cgc gtg gct ggc aat aaa		1200	
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys			
385	390	395	400
gac ctt tat ctc ggg acg ttt tcc acg caa gaa gaa gct gcc gaa gcc		1248	

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Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala			
405	410	415	
tac gat att gcc gcc att aaa ttt cgg ggg ctg aat gcc gtg acg aac			1296
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn			
420	425	430	
ttt gat atg tcc cgg tat gat gtc aaa tcc att ctc gat tcc tcc gcg			1344
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala			
435	440	445	
ctg ccg atc ggg agc gcc gcg aaa cgg ctc aag gag gcg gaa gcg gca			1392
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala			
450	455	460	
gcc agc gcc cag cat cat cac gcg ggc gtc gtg tcc tat gac gtg ggg			1440
Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly			
465	470	475	480
cgc atc gcc agc caa ctg ggg gat ggg ggt gcg ctc gcc gcc tat			1488
Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Ala Tyr			
485	490	495	
ggc gcc cat tat cat ggg gcg gcg tgg ccc acc att gcg ttt cag ccc			1536
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro			
500	505	510	
ggg gcg gcg tcc act ggc ctc tat cat ccc tat gcg caa caa cct atg			1584
Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met			
515	520	525	
cgg ggg ggg tgg tgt aaa caa gaa caa gac cat gcg gtc att gcc			1632
Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala			
530	535	540	
gcg gcc cat tcc ctc caa gat ctg cat cat ctg aac ctc ggg gcc gcc			1680
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala			
545	550	555	560
ggg gcc cat gat ttt ttt agc gct ggc caa cag gcg gcg gct gcc			1728
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala			
565	570	575	
atg cat ggg ctc gga tcc att gat agt gcg agc ctg gaa cat tcc acg			1776
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr			
580	585	590	
ggg tcc aac agc gtg gtg tat aat ggg ggc gtg ggc gat agc aat ggc			1824
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly			
595	600	605	
gcg tcc gcg gtc ggg ggc tcc ggg ggt ggg tat atg atg ccc atg tcc			1872
Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser			
610	615	620	
gct gcc ggt gct acg aca act tcg gct atg gtc tcc cat gaa caa gtc			1920
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val			
625	630	635	640
cac gct cgc gcc tat gat gag ggc aaa caa gca gca cag atg ggc tat			1968
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr			
645	650	655	
gaa tcc tat ctc gtc aat gcc gaa aat aat gga ggg ggt aga atg tcc			2016
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser			
660	665	670	
gct tgg ggc act gtg gtc tcc gct gcg gcc gcc gct gca gct tcc tcc			2064
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser			
675	680	685	
aat gat aac atg gcg gcg gac gtg ggg cac ggc ggg gcc caa ctc ttt			2112
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Gly Ala Gln Leu Phe			
690	695	700	
agt gtg tgg aat gat aca taa			2133

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Ser Val Trp Asn Asp Thr *
705 710

<210> SEQ ID NO 19
<211> LENGTH: 710
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Variant of Zm-ODP2 polypeptide having the amino acid sequence set forth in SEQ ID NO:2 with a single amino acid alteration (i.e., S37 to T37).

<400> SEQUENCE: 19

Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu
1 5 10 15

Leu Pro Pro Ser Gln Thr Thr Asp Ser Thr Leu Ile Ser Ala Ala Thr
20 25 30

Ala Asp His Val Thr Gly Asp Val Cys Phe Asn Ile Pro Gln Asp Trp
35 40 45

Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val Ala Glu Pro Lys Leu
50 55 60

Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala
65 70 75 80

Asn Cys Asn Met Ile Pro Ser Thr Ser Ser Thr Val Cys Tyr Ala Ser
85 90 95

Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr
100 105 110

Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala
115 120 125

Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Asn Gly
130 135 140

Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met
145 150 155 160

Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val
165 170 175

Ala Ala Ala Glu Gly Ala Gln Gly Leu Ser Leu Ser Met Asn Met Ala
180 185 190

Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg
195 200 205

Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val
210 215 220

Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly
225 230 235 240

Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser
245 250 255

Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr
260 265 270

Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
275 280 285

Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys
290 295 300

Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
305 310 315 320

Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr

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325	330	335	
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys 340	345	350	
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser 355	360	365	
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His 370	375	380	
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys 385	390	395	400
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala 405	410	415	
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn 420	425	430	
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala 435	440	445	
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala 450	455	460	
Ala Ser Ala Gln His His Ala Gly Val Val Ser Tyr Asp Val Gly 465	470	475	480
Arg Ile Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Ala Tyr 485	490	495	
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Ile Ala Phe Gln Pro 500	505	510	
Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met 515	520	525	
Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala 530	535	540	
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala 545	550	555	560
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala 565	570	575	
Met His Gly Leu Gly Ser Ile Asp Ser Ala Ser Leu Glu His Ser Thr 580	585	590	
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly 595	600	605	
Ala Ser Ala Val Gly Ser Gly Gly Tyr Met Met Pro Met Ser 610	615	620	
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val 625	630	635	640
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr 645	650	655	
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser 660	665	670	
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ser Ser 675	680	685	
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Gly Ala Gln Leu Phe 690	695	700	
Ser Val Trp Asn Asp Thr 705	710		

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<211> LENGTH: 710
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Variant of Zm-ODP2 having 97.3% amino acid
sequence identity to SEQ ID NO:2 (Zm-ODP2).

<400> SEQUENCE: 20

Met Ala Thr Val Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu
1 5 10 15

Ile Pro Pro Ser Gln Thr Thr Asp Ser Thr Ile Leu Ser Ala Ala Thr
20 25 30

Ala Asp His Val Ser Gly Asp Val Cys Phe Asn Leu Pro Gln Asp Trp
35 40 45

Ser Met Arg Gly Ser Glu Ile Ser Ala Ile Val Ala Glu Pro Lys Ile
50 55 60

Glu Asp Phe Ile Gly Gly Leu Ser Phe Ser Glu Gln His His Lys Ala
65 70 75 80

Asn Cys Asn Met Leu Pro Ser Thr Ser Thr Val Cys Tyr Ala Ser
85 90 95

Ser Gly Ala Ser Thr Gly Tyr His His Gln Ile Tyr His Gln Pro Thr
100 105 110

Ser Ser Ala Ile His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala
115 120 125

Gly Val His Asp Gly Gly Ala Met Ile Ser Ala Ala Ala Asn Gly
130 135 140

Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met
145 150 155 160

Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val
165 170 175

Ala Ala Ala Glu Gly Ala Gln Gly Ile Ser Ile Ser Met Asn Met Ala
180 185 190

Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Leu Leu Ala Gly Glu Arg
195 200 205

Ala Arg Ala Pro Glu Ser Val Ser Thr Ser Ala Gln Gly Gly Ala Val
210 215 220

Val Val Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Val Ala Gly
225 230 235 240

Ala Leu Val Ala Val Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser
245 250 255

Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr
260 265 270

Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
275 280 285

Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys
290 295 300

Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
305 310 315 320

Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr
325 330 335

Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys
340 345 350

His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser

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355	360	365	
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His			
370	375	380	
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys			
385	390	395	400
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala			
405	410	415	
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn			
420	425	430	
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala			
435	440	445	
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala			
450	455	460	
Ala Ser Ala Gln His His His Ala Gly Val Val Ser Tyr Asp Val Gly			
465	470	475	480
Arg Val Ala Ser Gln Leu Gly Asp Gly Gly Ala Leu Ala Ala Ala Tyr			
485	490	495	
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Val Ala Phe Gln Pro			
500	505	510	
Gly Ala Ala Ser Thr Gly Leu Tyr His Pro Tyr Ala Gln Gln Pro Met			
515	520	525	
Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Val Ala			
530	535	540	
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala			
545	550	555	560
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala			
565	570	575	
Met His Gly Leu Gly Ser Val Asp Ser Ala Ser Leu Glu His Ser Thr			
580	585	590	
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Val Gly Asp Ser Asn Gly			
595	600	605	
Ala Ser Ala Val Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser			
610	615	620	
Ala Ala Gly Ala Thr Thr Ser Ala Met Val Ser His Glu Gln Val			
625	630	635	640
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr			
645	650	655	
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser			
660	665	670	
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser			
675	680	685	
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe			
690	695	700	
Ser Val Trp Asn Asp Thr			
705	710		

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<210> SEQ_ID NO 21
<211> LENGTH: 710
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Variant of Zm-ODP2 having 92.4% amino acid
sequence identity to SEQ ID NO:2 (Zm-ODP2).

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<400> SEQUENCE: 21

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Met Ala Thr Ile Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu
1           5           10           15

Ile Pro Pro Ser Gln Thr Thr Asp Ser Thr Ile Leu Ser Ala Ala Thr
20          25          30

Ala Asp His Ile Ser Gly Asp Val Cys Phe Asn Leu Pro Gln Asp Trp
35          40          45

Ser Met Arg Gly Ser Glu Ile Ser Ala Ile Ile Ala Glu Pro Lys Ile
50          55          60

Glu Asp Phe Ile Gly Gly Leu Ser Phe Ser Glu Gln His His Lys Ala
65          70          75          80

Asn Cys Asn Met Leu Pro Ser Thr Ser Ser Thr Ile Cys Tyr Ala Ser
85          90          95

Ser Gly Ala Ser Thr Gly Tyr His His Gln Ile Tyr His Gln Pro Thr
100         105         110

Ser Ser Ala Ile His Phe Ala Asp Ser Ile Met Ile Ala Ser Ser Ala
115         120         125

Gly Ile His Asp Gly Gly Ala Met Ile Ser Ala Ala Ala Asn Gly
130         135         140

Ile Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met
145         150         155         160

Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Ile
165         170         175

Ala Ala Ala Glu Gly Ala Gln Gly Ile Ser Ile Ser Met Asn Met Ala
180         185         190

Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Ile Val Ala Gly Glu Arg
195         200         205

Ala Arg Ala Pro Glu Ser Ile Ser Thr Ser Ala Gln Gly Gly Ala Ile
210         215         220

Ile Ile Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Leu Ala Gly
225         230         235         240

Ala Val Leu Ala Leu Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser
245         250         255

Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr
260         265         270

Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
275         280         285

Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys
290         295         300

Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
305         310         315         320

Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr
325         330         335

Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys
340         345         350

His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser
355         360         365

Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His
370         375         380

Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys

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385	390	395	400
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala			
405	410	415	
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn			
420	425	430	
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala			
435	440	445	
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala			
450	455	460	
Ala Ser Ala Gln His His Ala Gly Leu Leu Ser Tyr Asp Leu Gly			
465	470	475	480
Arg Val Ala Ser Gln Val Gly Asp Gly Gly Ala Val Ala Ala Tyr			
485	490	495	
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Val Ala Phe Gln Pro			
500	505	510	
Gly Ala Ala Ser Thr Gly Val Tyr His Pro Tyr Ala Gln Gln Pro Met			
515	520	525	
Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Leu Val Ala			
530	535	540	
Ala Ala His Ser Val Gln Asp Val His His Val Asn Val Gly Ala Ala			
545	550	555	560
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala			
565	570	575	
Met His Gly Val Gly Ser Val Asp Ser Ala Ser Val Glu His Ser Thr			
580	585	590	
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Leu Gly Asp Ser Asn Gly			
595	600	605	
Ala Ser Ala Leu Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser			
610	615	620	
Ala Ala Gly Ala Thr Thr Ser Ala Met Leu Ser His Glu Gln Val			
625	630	635	640
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr			
645	650	655	
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser			
660	665	670	
Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser			
675	680	685	
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe			
690	695	700	
Ser Val Trp Asn Asp Thr			
705	710		

<210> SEQ ID NO 22
 <211> LENGTH: 710
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Variant of Zm-ODP2 having 87.3% amino acid sequence identity to SEQ ID NO:2 (Zm-ODP2).

<400> SEQUENCE: 22

Met Ala Thr Ile Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu
 1 5 10 15

-continued

Ile Pro Pro Ser Gln Thr Thr Asp Ser Thr Ile Leu Ser Gly Gly Thr
 20 25 30

Ala Asp His Ile Ser Gly Asp Val Cys Phe Asn Leu Pro Gln Asp Trp
 35 40 45

Ser Met Arg Gly Ser Glu Ile Ser Gly Ile Ile Gly Glu Pro Lys Ile
 50 55 60

Glu Asp Phe Ile Gly Gly Leu Ser Phe Ser Glu Gln His His Lys Gly
 65 70 75 80

Asn Cys Asn Met Leu Pro Ser Thr Ser Thr Ile Cys Tyr Gly Ser
 85 90 95

Ser Gly Gly Ser Thr Gly Tyr His His Gln Ile Tyr His Gln Pro Thr
 100 105 110

Ser Ser Gly Ile His Phe Gly Asp Ser Ile Met Ile Gly Ser Ser Gly
 115 120 125

Gly Ile His Asp Gly Gly Met Ile Ser Gly Gly Gly Asn Gly
 130 135 140

Ile Gly Gly Gly Ser Gly Asn Gly Gly Ile Gly Leu Ser Met
 145 150 155 160

Ile Lys Asn Trp Leu Arg Ser Gln Pro Gly Pro Met Gln Pro Arg Ile
 165 170 175

Gly Gly Gly Glu Gly Gln Gly Ile Ser Ile Ser Met Asn Met Gly
 180 185 190

Gly Thr Thr Gln Gly Gly Gly Met Pro Ile Val Gly Gly Glu Arg
 195 200 205

Gly Arg Gly Pro Glu Ser Ile Ser Thr Ser Gly Gln Gly Gly Ile
 210 215 220

Ile Ile Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Leu Ala Gly
 225 230 235 240

Ala Val Leu Ala Leu Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser
 245 250 255

Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr
 260 265 270

Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
 275 280 285

Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys
 290 295 300

Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
 305 310 315 320

Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr
 325 330 335

Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys
 340 345 350

His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser
 355 360 365

Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His
 370 375 380

Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys
 385 390 395 400

Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala
 405 410 415

Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn

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420	425	430
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala		
435	440	445
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala		
450	455	460
Ala Ser Ala Gln His His Ala Gly Leu Leu Ser Tyr Asp Leu Gly		
465	470	475
Arg Val Ala Ser Gln Val Gly Asp Gly Gly Ala Val Ala Ala Ala Tyr		
485	490	495
Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Val Ala Phe Gln Pro		
500	505	510
Gly Ala Ala Ser Thr Gly Val Tyr His Pro Tyr Ala Gln Gln Pro Met		
515	520	525
Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Leu Val Ala		
530	535	540
Ala Ala His Ser Val Gln Asp Val His His Val Asn Val Gly Ala Ala		
545	550	555
Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala Ala		
565	570	575
Met His Gly Val Gly Ser Val Asp Ser Ala Ser Val Glu His Ser Thr		
580	585	590
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Leu Gly Asp Ser Asn Gly		
595	600	605
Ala Ser Ala Leu Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser		
610	615	620
Ala Ala Gly Ala Thr Thr Ser Ala Met Leu Ser His Glu Gln Leu		
625	630	635
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr		
645	650	655
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser		
660	665	670
Ala Trp Gly Thr Leu Leu Ser Ala Ala Ala Ala Ala Ala Ser Ser		
675	680	685
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Gly Ala Gln Leu Phe		
690	695	700
Ser Val Trp Asn Asp Thr		
705	710	

<210> SEQ ID NO 23
 <211> LENGTH: 710
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Variant of Zm-ODP2 having 82.4% amino acid
 sequence identity to SEQ ID NO:2 (Zm-ODP2).
 <400> SEQUENCE: 23

Met Ala Thr Ile Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu		
1	5	10
Ile Pro Pro Ser Gln Thr Thr Asp Ser Thr Ile Leu Ser Gly Gly Thr		
20	25	30
Ala Asp His Ile Ser Gly Asp Val Cys Phe Asn Leu Pro Gln Asp Trp		
35	40	45

-continued

Ser Met Arg Gly Ser Glu Ile Ser Gly Ile Ile Gly Glu Pro Lys Ile
 50 55 60
 Glu Asp Phe Ile Gly Gly Leu Ser Phe Ser Glu Gln His His Lys Gly
 65 70 75 80
 Asn Cys Asn Met Leu Pro Ser Thr Ser Ser Thr Ile Cys Tyr Gly Ser
 85 90 95
 Ser Gly Gly Ser Thr Gly Tyr His His Gln Ile Tyr His Gln Pro Thr
 100 105 110
 Ser Ser Gly Ile His Phe Gly Asp Ser Ile Met Ile Gly Ser Ser Gly
 115 120 125
 Gly Ile His Asp Gly Gly Met Ile Ser Gly Gly Gly Asn Gly
 130 135 140
 Ile Gly Gly Gly Ser Gly Asn Gly Gly Ile Gly Leu Ser Met
 145 150 155 160
 Ile Lys Asn Trp Leu Arg Ser Gln Pro Gly Pro Met Gln Pro Arg Ile
 165 170 175
 Gly Gly Gly Glu Gly Gln Gly Ile Ser Ile Ser Met Asn Met Gly
 180 185 190
 Gly Thr Thr Gln Gly Gly Gly Met Pro Ile Val Gly Gly Glu Arg
 195 200 205
 Gly Arg Gly Pro Glu Ser Ile Ser Thr Ser Gly Gln Gly Gly Ile
 210 215 220
 Ile Ile Thr Gly Pro Lys Glu Asp Ser Gly Gly Ser Gly Leu Gly Gly
 225 230 235 240
 Gly Val Leu Gly Leu Ser Thr Asp Thr Gly Gly Ser Gly Gly Ser
 245 250 255
 Gly Asp Asn Thr Gly Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr
 260 265 270
 Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
 275 280 285
 Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys
 290 295 300
 Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
 305 310 315 320
 Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr
 325 330 335
 Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys
 340 345 350
 His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser
 355 360 365
 Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His
 370 375 380
 Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys
 385 390 395 400
 Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala
 405 410 415
 Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn
 420 425 430
 Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala
 435 440 445
 Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Gly

-continued

450	455	460
Gly Ser Gly Gln His His His Gly Gly Leu Leu Ser Tyr Asp Leu Gly		
465	470	475
480		
Arg Val Gly Ser Gln Val Gly Asp Gly Gly Val Gly Gly Tyr		
485	490	495
Gly Gly His Tyr His Gly Gly Trp Pro Thr Val Gly Phe Gln Pro		
500	505	510
Gly Gly Gly Ser Thr Gly Val Tyr His Pro Tyr Gly Gln Gln Pro Met		
515	520	525
Arg Gly Gly Gly Trp Cys Lys Gln Glu Gln Asp His Gly Leu Val Gly		
530	535	540
Gly Gly His Ser Val Gln Asp Val His His Val Asn Val Gly Gly Gly		
545	550	555
560		
Gly Gly His Asp Phe Phe Ser Gly Gly Gln Gln Gly Gly Gly Ala		
565	570	575
Met His Gly Val Gly Ser Val Asp Ser Ala Ser Val Glu His Ser Thr		
580	585	590
Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Leu Gly Asp Ser Asn Gly		
595	600	605
Ala Ser Ala Leu Gly Gly Ser Gly Gly Tyr Met Met Pro Met Ser		
610	615	620
Ala Ala Gly Ala Thr Thr Ser Ala Met Leu Ser His Glu Gln Leu		
625	630	635
640		
His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr		
645	650	655
Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser		
660	665	670
Ala Trp Gly Thr Leu Leu Ser Ala Ala Ala Ala Ala Ala Ser Ser		
675	680	685
Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe		
690	695	700
Ser Val Trp Asn Asp Thr		
705	710	

<210> SEQ ID NO 24
 <211> LENGTH: 710
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: Consensus sequence of Figure 2.

<400> SEQUENCE: 24

Met Ala Thr Ile Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Glu		
1	5	10
15		
Ile Pro Pro Ser Gln Thr Thr Asp Ser Thr Ile Leu Ser Ala Ala Thr		
20	25	30
Ala Asp His Ile Ser Gly Asp Val Cys Phe Asn Leu Pro Gln Asp Trp		
35	40	45
Ser Met Arg Gly Ser Glu Ile Ser Ala Ile Ile Ala Glu Pro Lys Ile		
50	55	60
Glu Asp Phe Ile Gly Gly Leu Ser Phe Ser Glu Gln His His Lys Ala		
65	70	75
80		
Asn Cys Asn Met Leu Pro Ser Thr Ser Ser Thr Ile Cys Tyr Ala Ser		

-continued

85	90	95	
Ser Gly Ala Ser Thr Gly Tyr His His Gln Ile Tyr His Gln Pro Thr			
100	105	110	
Ser Ser Ala Ile His Phe Ala Asp Ser Ile Met Ile Ala Ser Ser Ala			
115	120	125	
Gly Ile His Asp Gly Gly Ala Met Ile Ser Ala Ala Ala Asn Gly			
130	135	140	
Ile Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met			
145	150	155	160
Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Ile			
165	170	175	
Ala Ala Ala Glu Gly Ala Gln Gly Ile Ser Ile Ser Met Asn Met Ala			
180	185	190	
Gly Thr Thr Gln Gly Ala Ala Gly Met Pro Ile Val Ala Gly Glu Arg			
195	200	205	
Ala Arg Ala Pro Glu Ser Ile Ser Thr Ser Ala Gln Gly Gly Ala Ile			
210	215	220	
Ile Ile Thr Ala Pro Lys Glu Asp Ser Gly Gly Ser Gly Leu Ala Gly			
225	230	235	240
Ala Val Leu Ala Leu Ser Thr Asp Thr Gly Gly Ser Gly Gly Ala Ser			
245	250	255	
Ala Asp Asn Thr Ala Arg Lys Thr Val Asp Thr Phe Gly Gln Arg Thr			
260	265	270	
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu			
275	280	285	
Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys			
290	295	300	
Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala			
305	310	315	320
Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Ala Thr Thr Thr			
325	330	335	
Thr Asn Phe Pro Val Ser Asn Tyr Glu Lys Glu Leu Glu Asp Met Lys			
340	345	350	
His Met Thr Arg Gln Glu Phe Val Ala Ser Leu Arg Arg Lys Ser Ser			
355	360	365	
Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His			
370	375	380	
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys			
385	390	395	400
Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln Glu Glu Ala Ala Glu Ala			
405	410	415	
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn			
420	425	430	
Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser Ser Ala			
435	440	445	
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Glu Ala Ala			
450	455	460	
Ala Ser Ala Gln His His Ala Gly Leu Leu Ser Tyr Asp Leu Gly			
465	470	475	480
Arg Val Ala Ser Gln Val Gly Asp Gly Gly Ala Val Ala Ala Tyr			
485	490	495	

-continued

Gly Ala His Tyr His Gly Ala Ala Trp Pro Thr Val Ala Phe Gln Pro
 500 505 510

Gly Ala Ala Ser Thr Gly Val Tyr His Pro Tyr Ala Gln Gln Pro Met
 515 520 525

Arg Gly Gly Trp Cys Lys Gln Glu Gln Asp His Ala Leu Val Ala
 530 535 540

Ala Ala His Ser Val Gln Asp Val His His Val Asn Val Gly Ala Ala
 545 550 555 560

Gly Ala His Asp Phe Phe Ser Ala Gly Gln Gln Ala Ala Ala Ala
 565 570 575

Met His Gly Val Gly Ser Val Asp Ser Ala Ser Val Glu His Ser Thr
 580 585 590

Gly Ser Asn Ser Val Val Tyr Asn Gly Gly Leu Gly Asp Ser Asn Gly
 595 600 605

Ala Ser Ala Leu Gly Ser Gly Gly Tyr Met Met Pro Met Ser
 610 615 620

Ala Ala Gly Ala Thr Thr Ser Ala Met Leu Ser His Glu Gln Val
 625 630 635 640

His Ala Arg Ala Tyr Asp Glu Ala Lys Gln Ala Ala Gln Met Gly Tyr
 645 650 655

Glu Ser Tyr Leu Val Asn Ala Glu Asn Asn Gly Gly Arg Met Ser
 660 665 670

Ala Trp Gly Thr Val Val Ser Ala Ala Ala Ala Ala Ala Ser Ser
 675 680 685

Asn Asp Asn Met Ala Ala Asp Val Gly His Gly Ala Gln Leu Phe
 690 695 700

Ser Val Trp Asn Asp Thr
 705 710

<210> SEQ ID NO 25
 <211> LENGTH: 2079
 <212> TYPE: DNA
 <213> ORGANISM: Oryza sativa
 <220> FEATURE:
 <221> NAME/KEY: misc_feature
 <222> LOCATION: (0)...(0)
 <223> OTHER INFORMATION: predicted cDNA of OsANT, Genbank Accession No.
 AP003313 (rice ODP2)
 <221> NAME/KEY: CDS
 <222> LOCATION: (1)...(2079)

<400> SEQUENCE: 25

atg gcc acc atg aac aac tgg ctg gcc ttc tcc ctc tcc ccg cag gat	48
Met Ala Thr Met Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Asp	
1 5 10 15	
cag ctc ccg ccg tct cag acc aac tcc act ctc atc tcc gcc gcc gcc	96
Gln Leu Pro Pro Ser Gln Thr Asn Ser Thr Leu Ile Ser Ala Ala Ala	
20 25 30	
acc acc acc acc gcc gac tcc tcc acc ggc gac gtc tgc ttc aac	144
Thr Thr Thr Thr Ala Gly Asp Ser Ser Thr Gly Asp Val Cys Phe Asn	
35 40 45	
atc ccc caa gat tgg agc atg agg gga tcg gag ctc tcg gcg ctc gtc	192
Ile Pro Gln Asp Trp Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val	
50 55 60	
gcc gag ccg aag ctg gag gac ttc ctc ggc ggc atc tcc ttc tcg gag	240
Ala Glu Pro Lys Leu Glu Asp Phe Leu Gly Ile Ser Phe Ser Glu	

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65	70	75	80	
cag cag cat cat cac ggc ggc aag ggc ggc gtg atc ccg agc agc ggc Gln Gln His His His Gly Gly Lys Gly Gly Val Ile Pro Ser Ser Ala				288
85 90 95				
gcc gct tgc tac gcg agc tcc ggc agc agc gtc ggc tac ctg tac tac cct Ala Ala Cys Tyr Ala Ser Ser Gly Ser Ser Val Gly Tyr Leu Tyr Pro				336
100 105 110				
cct cca agc tca tcc tcg ctc cag ttc gcc gac tcc gtc atg gtg gcc Pro Pro Ser Ser Ser Leu Gln Phe Ala Asp Ser Val Met Val Ala				384
115 120 125				
acc tcc tcg ccc gtc gtc gcc cac gac ggc gtc agc ggc ggc ggc atg Thr Ser Ser Pro Val Val Ala His Asp Gly Val Ser Gly Gly Met				432
130 135 140				
gtg agc gcc gcc ggc gcg gcg agt ggc aac ggc ggc att ggc Val Ser Ala Ala Ala Ala Ala Ser Gly Asn Gly Gly Ile Gly				480
145 150 155 160				
ctg tcc atg atc aag aac tgg ctc cgg agc cag ccg cgg cag ccg Leu Ser Met Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Gln Pro				528
165 170 175				
gcg cag gcg ctg tct ctg tcc atg aac atg gcg ggg acg acg acg gcg Ala Gln Ala Leu Ser Leu Ser Met Asn Met Ala Gly Thr Thr Ala				576
180 185 190				
cag ggc ggc ggc atg gcg ctc ctc gcc gca ggg gag cga ggc Gln Gly Gly Ala Met Ala Leu Leu Ala Gly Ala Gly Glu Arg Gly				624
195 200 205				
ccg acg acg ccc gcg tca gag agc ctg tcc acg tcc gcg cac gga gcg Arg Thr Thr Pro Ala Ser Glu Ser Leu Ser Thr Ser Ala His Gly Ala				672
210 215 220				
acg acg acg acg atg ggt ggt cgc aag gag att aac gag gaa ggc Thr Thr Ala Thr Met Ala Gly Gly Arg Lys Glu Ile Asn Glu Glu Gly				720
225 230 235 240				
agc ggc agc gcc ggc gtg gtt gcc gtc ggc tcg gag tca ggc ggc Ser Gly Ser Ala Gly Ala Val Val Ala Val Gly Ser Glu Ser Gly Gly				768
245 250 255				
agc ggc gcc gtg gtg gag gcc ggc gcg gcg gcg gcg agg aag Ser Gly Ala Val Val Glu Ala Gly Ala Ala Ala Ala Ala Arg Lys				816
260 265 270				
tcc gtc gac acg ttc ggc cag aga aca tcg atc tac cgc ggc gtg aca Ser Val Asp Thr Phe Gly Gln Arg Thr Ser Ile Tyr Arg Gly Val Thr				864
275 280 285				
agg cat aga tgg aca ggg agg tat gag gct cat ctt tgg gac aac agc Arg His Arg Trp Thr Gly Arg Tyr Glu Ala His Leu Trp Asp Asn Ser				912
290 295 300				
tgc aga aga gag ggc caa act cgc aag ggt cgt caa ggt ggt tat gac Cys Arg Arg Glu Gly Gln Thr Arg Lys Gly Arg Gln Gly Gly Tyr Asp				960
305 310 315 320				
aaa gag gaa aaa gct gct aga gct tat gat ttg gct gct ctc aaa tac Lys Glu Glu Lys Ala Ala Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr				1008
325 330 335				
tgg ggc ccg acg acg acg aca aat ttt ccg gta aat aac tat gaa aag Trp Gly Pro Thr Thr Thr Asn Phe Pro Val Asn Asn Tyr Glu Lys				1056
340 345 350				
gag ctg gag gag atg aag cac atg aca agg cag gag ttc gta gcc tct Glu Leu Glu Glu Met Lys His Met Thr Arg Gln Glu Phe Val Ala Ser				1104
355 360 365				
ttg aga agg aag agc agt ggt ttc tcc aga ggt gca tcc att tac cgt Leu Arg Arg Lys Ser Ser Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg				1152

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370	375	380	
gga gta act agg cat cac cag cat ggg aga tgg caa gca agg ata gga Gly Val Thr Arg His His Gln His Gly Arg Trp Gln Ala Arg Ile Gly 385 390 395 400			1200
aga gtt gca ggg aac aag gac ctc tac ttg ggc acc ttc agc acg cag Arg Val Ala Gly Asn Lys Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln 405 410 415			1248
gag gag gcg gcg gag gcg tac gac atc gcg gcg atc aag ttc cgg ggg Glu Glu Ala Ala Glu Ala Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly 420 425 430			1296
ctc aac gcc gtc acc aac ttc gac atg agc cgc tac gac gtc aag agc Leu Asn Ala Val Thr Asn Phe Asp Met Ser Arg Tyr Asp Val Lys Ser 435 440 445			1344
atc ctc gac agc gct gcc ctc ccc gtc ggc acc gcc gcc aag cgc ctc Ile Leu Asp Ser Ala Ala Leu Pro Val Gly Thr Ala Ala Lys Arg Leu 450 455 460			1392
aag gac gcc gag gcc gcc gcc tac gac gtc ggc cgc atc gcc tcg Lys Asp Ala Glu Ala Ala Ala Tyr Asp Val Gly Arg Ile Ala Ser 465 470 475 480			1440
cac ctc ggc gac ggc gcc tac gcc gcg cat tac ggc cac cac cac His Leu Gly Asp Gly Ala Tyr Ala Ala His Tyr Gly His His His 485 490 495			1488
cac tcg gcc gcc gcc tgg ccg acc atc gcg ttc cag gcg gcg gcg His Ser Ala Ala Ala Ala Trp Pro Thr Ile Ala Phe Gln Ala Ala Ala 500 505 510			1536
gcg ccg ccg cac gcc gcc ggg ctt tac cac ccg tac gcg cag ccg Ala Pro Pro Pro His Ala Ala Gly Leu Tyr His Pro Tyr Ala Gln Pro 515 520 525			1584
ctg cgt ggg tgg tgc aag cag gag cag gac cac gcc gtg atc gcg gcg Leu Arg Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala Ala 530 535 540			1632
gcg cac agc ctg cag gat ctc cac cac aac ctc ggc gcc gcc gcc Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala Ala 545 550 555 560			1680
gcc gcg cat gac ttc ttc tcg cag gcg atg cag cag cag cac ggc ctc Ala Ala His Asp Phe Ser Gln Ala Met Gln Gln His Gly Leu 565 570 575			1728
ggc agc atc gac aac gcg tcg ctc gag cac agc acc ggc tcc aac tcc Gly Ser Ile Asp Asn Ala Ser Leu Glu His Ser Thr Gly Ser Asn Ser 580 585 590			1776
gtc gtc tac aac ggc gac aat ggc ggc gga ggc ggc ggc tac atc atg Val Val Tyr Asn Gly Asp Asn Gly Gly Gly Gly Gly Tyr Ile Met 595 600 605			1824
gcg ccg atg agc gcc gtg tcg gcc acg gcc acc gcg gtg gcg agc agc Ala Pro Met Ser Ala Val Ser Ala Thr Ala Thr Ala Val Ala Ser Ser 610 615 620			1872
cac gat cac ggc ggc gac ggc ggg aag cag gtg cag atg ggg tac gac His Asp His Gly Gly Asp Gly Gly Lys Gln Val Gln Met Gly Tyr Asp 625 630 635 640			1920
agc tac ctc gtc ggc gca gac gcc tac ggc ggc ggc ggc ggg agg Ser Tyr Leu Val Gly Ala Asp Ala Tyr Gly Gly Gly Ala Gly Arg 645 650 655			1968
atg cca tcc tgg gcg atg acg ccg gcg tcg cgc ccg gcc acg agc Met Pro Ser Trp Ala Met Thr Pro Ala Ser Ala Pro Ala Ala Thr Ser 660 665 670			2016
agc agc gac atg acc gga gtc tgc cat ggc gca cag ctc ttc agc gtc Ser Ser Asp Met Thr Gly Val Cys His Gly Ala Gln Leu Phe Ser Val			2064

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675	680	685	
			2079
tgg aac gac aca taa			
Trp Asn Asp Thr *			
690			
 <210> SEQ ID NO 26			
<211> LENGTH: 692			
<212> TYPE: PRT			
<213> ORGANISM: Oryza sativa			
 <400> SEQUENCE: 26			
Met Ala Thr Met Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Asp			
1	5	10	15
Gln Leu Pro Pro Ser Gln Thr Asn Ser Thr Leu Ile Ser Ala Ala Ala			
20	25	30	
Thr Thr Thr Ala Gly Asp Ser Ser Thr Gly Asp Val Cys Phe Asn			
35	40	45	
Ile Pro Gln Asp Trp Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val			
50	55	60	
Ala Glu Pro Lys Leu Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu			
65	70	75	80
Gln Gln His His His Gly Gly Lys Gly Gly Val Ile Pro Ser Ser Ala			
85	90	95	
Ala Ala Cys Tyr Ala Ser Ser Gly Ser Ser Val Gly Tyr Leu Tyr Pro			
100	105	110	
Pro Pro Ser Ser Ser Leu Gln Phe Ala Asp Ser Val Met Val Ala			
115	120	125	
Thr Ser Ser Pro Val Val Ala His Asp Gly Val Ser Gly Gly Met			
130	135	140	
Val Ser Ala Ala Ala Ala Ala Ala Ser Gly Asn Gly Gly Ile Gly			
145	150	155	160
Leu Ser Met Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Gln Pro			
165	170	175	
Ala Gln Ala Leu Ser Leu Ser Met Asn Met Ala Gly Thr Thr Thr Ala			
180	185	190	
Gln Gly Gly Gly Ala Met Ala Leu Leu Ala Gly Ala Gly Glu Arg Gly			
195	200	205	
Arg Thr Thr Pro Ala Ser Glu Ser Leu Ser Thr Ser Ala His Gly Ala			
210	215	220	
Thr Thr Ala Thr Met Ala Gly Gly Arg Lys Glu Ile Asn Glu Glu Gly			
225	230	235	240
Ser Gly Ser Ala Gly Ala Val Val Ala Val Gly Ser Glu Ser Gly Gly			
245	250	255	
Ser Gly Ala Val Val Glu Ala Gly Ala Ala Ala Ala Ala Arg Lys			
260	265	270	
Ser Val Asp Thr Phe Gly Gln Arg Thr Ser Ile Tyr Arg Gly Val Thr			
275	280	285	
Arg His Arg Trp Thr Gly Arg Tyr Glu Ala His Leu Trp Asp Asn Ser			
290	295	300	
Cys Arg Arg Glu Gly Gln Thr Arg Lys Gly Arg Gln Gly Gly Tyr Asp			
305	310	315	320
Lys Glu Glu Lys Ala Ala Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr			
325	330	335	

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Trp Gly Pro Thr Thr Thr Asn Phe Pro Val Asn Asn Tyr Glu Lys
 340 345 350
 Glu Leu Glu Glu Met Lys His Met Thr Arg Gln Glu Phe Val Ala Ser
 355 360 365
 Leu Arg Arg Lys Ser Ser Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg
 370 375 380
 Gly Val Thr Arg His His Gln His Gly Arg Trp Gln Ala Arg Ile Gly
 385 390 395 400
 Arg Val Ala Gly Asn Lys Asp Leu Tyr Leu Gly Thr Phe Ser Thr Gln
 405 410 415
 Glu Glu Ala Ala Glu Ala Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly
 420 425 430
 Leu Asn Ala Val Thr Asn Phe Asp Met Ser Arg Tyr Asp Val Lys Ser
 435 440 445
 Ile Leu Asp Ser Ala Ala Leu Pro Val Gly Thr Ala Ala Lys Arg Leu
 450 455 460
 Lys Asp Ala Glu Ala Ala Ala Ala Tyr Asp Val Gly Arg Ile Ala Ser
 465 470 475 480
 His Leu Gly Gly Asp Gly Ala Tyr Ala Ala His Tyr Gly His His His
 485 490 495
 His Ser Ala Ala Ala Ala Trp Pro Thr Ile Ala Phe Gln Ala Ala Ala
 500 505 510
 Ala Pro Pro Pro His Ala Ala Gly Leu Tyr His Pro Tyr Ala Gln Pro
 515 520 525
 Leu Arg Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala Ala
 530 535 540
 Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala Ala
 545 550 555 560
 Ala Ala His Asp Phe Phe Ser Gln Ala Met Gln Gln Gln His Gly Leu
 565 570 575
 Gly Ser Ile Asp Asn Ala Ser Leu Glu His Ser Thr Gly Ser Asn Ser
 580 585 590
 Val Val Tyr Asn Gly Asp Asn Gly Gly Gly Gly Tyr Ile Met
 595 600 605
 Ala Pro Met Ser Ala Val Ser Ala Thr Ala Thr Ala Val Ala Ser Ser
 610 615 620
 His Asp His Gly Gly Asp Gly Gly Lys Gln Val Gln Met Gly Tyr Asp
 625 630 635 640
 Ser Tyr Leu Val Gly Ala Asp Ala Tyr Gly Gly Gly Ala Gly Arg
 645 650 655
 Met Pro Ser Trp Ala Met Thr Pro Ala Ser Ala Pro Ala Ala Thr Ser
 660 665 670
 Ser Ser Asp Met Thr Gly Val Cys His Gly Ala Gln Leu Phe Ser Val
 675 680 685
 Trp Asn Asp Thr
 690

<210> SEQ_ID NO 27
 <211> LENGTH: 1794
 <212> TYPE: DNA
 <213> ORGANISM: oryza sativa
 <220> FEATURE:

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<221> NAME/KEY: misc_feature
<222> LOCATION: (0)...(0)
<223> OTHER INFORMATION: predicted OsBNM, Genbank Accession No.
    AY062180, (rice ODP2)
<221> NAME/KEY: CDS
<222> LOCATION: (1)...(1794)

<400> SEQUENCE: 27

atg gcc acc atg aac aac tgg ctg gcc ttc tcc ctc tcc ccg cag gat      48
Met Ala Thr Met Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Asp
1          5          10          15

cag ctc ccg ccg tct cag acc aac tcc act ttc atc tcc gcc gcc gcc      96
Gln Leu Pro Pro Ser Gln Thr Asn Ser Thr Phe Ile Ser Ala Ala Ala
20          25          30

acc acc acc acc gcc ggc gac tcc tcc acc ggc gac gtc tgc ttc aac      144
Thr Thr Thr Ala Gly Asp Ser Ser Thr Gly Asp Val Cys Phe Asn
35          40          45

atc ccc caa gct cac ccc tcc acg ccg gcc att ggc aac ggc ggc att      192
Ile Pro Gln Ala His Pro Ser Thr Pro Ala Ile Gly Asn Gly Gly Ile
50          55          60

ggc ctg tcc atg atc aag aac tgg ctc ccg acg ccg ggg ccg cag cag      240
Gly Leu Ser Met Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Gln
65          70          75          80

ccg gcg cag ggc ctg tct ctg atg aac atg gcg ggg acg acg acg      288
Pro Ala Gln Ala Leu Ser Leu Ser Met Asn Met Ala Gly Thr Thr Thr
85          90          95

gct cag ggc ggc ggc atg gcg ctc ctc gcc ggc gca ggg gag cga      336
Ala Gln Gly Gly Ala Met Ala Leu Leu Ala Gly Ala Gly Glu Arg
100         105         110

ggc cgg acg acg ccc ggc tca gag acg ctg tcc acg tgc gcg cac gga      384
Gly Arg Thr Thr Pro Ala Ser Glu Ser Leu Ser Thr Ser Ala His Gly
115         120         125

gct acg acg ggc acg atg gct ggt ggt cgc aag gag att aac gag gaa      432
Ala Thr Thr Ala Thr Met Ala Gly Gly Arg Lys Glu Ile Asn Glu Glu
130         135         140

ggc acg ggc acg gcc ggc gtc gtg gtt gcc gtc tgc gag tca ggc      480
Gly Ser Gly Ser Ala Gly Ala Val Val Ala Val Gly Ser Glu Ser Gly
145         150         155         160

ggc acg ggc ggc gtg gag gcc ggc gcg ggc gcg ggc gcg agg      528
Gly Ser Gly Ala Val Val Glu Ala Gly Ala Ala Ala Ala Ala Arg
165         170         175

aag tcc gtc gac acg ttc ggc cag aga aca tcg atc tac cgc ggc gtg      576
Lys Ser Val Asp Thr Phe Gly Gln Arg Thr Ser Ile Tyr Arg Gly Val
180         185         190

aca agg cat aca tgg aca ggg agg tat gag gct cat ctt tgg gac aac      624
Thr Arg His Arg Trp Thr Gly Arg Tyr Glu Ala His Leu Trp Asp Asn
195         200         205

agc tgc aga aga gag ggc caa act cgc aag ggt cgt caa ggt ggt tat      672
Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys Gly Arg Gln Gly Gly Tyr
210         215         220

gac aaa gag gaa aaa gct gct aga gct tat gat ttg gct gct ctc aaa      720
Asp Lys Glu Glu Lys Ala Ala Arg Ala Tyr Asp Leu Ala Ala Leu Lys
225         230         235         240

tac tgg ggc ccg acg acg aca aat ttt ccg gta aat aac tat gaa      768
Tyr Trp Gly Pro Thr Thr Thr Asn Phe Pro Val Asn Asn Tyr Glu
245         250         255

aag gag ctg gag gag atg aag cac atg aca agg gag ttc gta gcc      816
Lys Glu Leu Glu Glu Met Lys His Met Thr Arg Gln Glu Val Ala
260         265         270

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tct ttg aga agg aag agc agt ggt ttc tcc aga ggt gca tcc att tac	864
Ser Leu Arg Arg Lys Ser Ser Gly Phe Ser Arg Gly Ala Ser Ile Tyr	
275 280 285	
cgt gga gta act agg cat cac cag cat ggg aga tgg caa gca agg ata	912
Arg Gly Val Thr Arg His His Gln His Gly Arg Trp Gln Ala Arg Ile	
290 295 300	
gga aga gtt gca ggg aac aag gac ctc tac ttg ggc acc ttc agc acg	960
Gly Arg Val Ala Gly Asn Lys Asp Leu Tyr Leu Gly Thr Phe Ser Thr	
305 310 315 320	
cag gag gag gcg gcg gag gac atc gac gcg gcg atc aag ttc cgg	1008
Gln Glu Ala Ala Glu Ala Tyr Asp Ile Ala Ala Ile Lys Phe Arg	
325 330 335	
ggg ctc aac gcc gtc acc aac ttc gac atg agc cgc tac gac gtc aag	1056
Gly Leu Asn Ala Val Thr Asn Phe Asp Met Ser Arg Tyr Asp Val Lys	
340 345 350	
agc atc ctc gac agc gct gcc ctc ccc gtc ggc acc gcc gcc aag cgc	1104
Ser Ile Leu Asp Ser Ala Ala Leu Pro Val Gly Thr Ala Ala Lys Arg	
355 360 365	
ctc aag gac gcc gag gcc gac gtc tac gac gtc ggc cgc atc gcc	1152
Leu Lys Asp Ala Glu Ala Ala Ala Tyr Asp Val Gly Arg Ile Ala	
370 375 380	
tcg cac ctc ggc gac ggc tac gcc gcg cat tac ggc cac cac	1200
Ser His Leu Gly Gly Asp Gly Ala Tyr Ala Ala His Tyr Gly His His	
385 390 395 400	
cac cac tcg gcc gcc gcc tgg ccg acc atc gcg ttc cag gcg gcg	1248
His His Ser Ala Ala Ala Ala Trp Pro Thr Ile Ala Phe Gln Ala Ala	
405 410 415	
gcg gcg ccg ccg cac gcc ggg ctt tac cac ccg tac gcg cag	1296
Ala Ala Pro Pro His Ala Ala Gly Leu Tyr His Pro Tyr Ala Gln	
420 425 430	
ccg ctg cgt ggg tgg tgc aag cag gag cag gac cac gcc gtg atc gcg	1344
Pro Leu Arg Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala	
435 440 445	
gcg gcg cac agc ctg cag gat ctc cac ctc aac ctc ggc gcc gcc	1392
Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala	
450 455 460	
gcc gcc gcg cat gac ttc ttc tcg cag gcg atg cag cag cag cac ggc	1440
Ala Ala Ala His Asp Phe Phe Ser Gln Ala Met Gln Gln His Gly	
465 470 475 480	
ctc ggc agc atc gac aac gcg tcg ctc gag cac agc acc ggc tcc aac	1488
Leu Gly Ser Ile Asp Asn Ala Ser Leu Glu His Ser Thr Gly Ser Asn	
485 490 495	
tcc gtc gtc tac aac ggc gac aat ggc gga ggc ggc ggc tac atc	1536
Ser Val Val Tyr Asn Gly Asp Asn Gly Gly Gly Gly Gly Tyr Ile	
500 505 510	
atg gcg ccg atg agc gcc gtg tcg gcc acg gcc acc gcg gtg gcg agc	1584
Met Ala Pro Met Ser Ala Val Ser Ala Thr Ala Thr Ala Val Ala Ser	
515 520 525	
agc cac gat cac ggc gac ggc ggg aag cag gtg cag atg ggg tac	1632
Ser His Asp His Gly Gly Asp Gly Gly Lys Gln Val Gln Met Gly Tyr	
530 535 540	
gac agc tac ctc gtc ggc gca gac ggc tac ggc ggc ggc gcc ggg	1680
Asp Ser Tyr Leu Val Gly Ala Asp Ala Tyr Gly Gly Gly Gly Ala Gly	
545 550 555 560	
agg atg cca tcc tgg gcg atg acg ccg gcg tcg gcg ccg gcc acg	1728
Arg Met Pro Ser Trp Ala Met Thr Pro Ala Ser Ala Pro Ala Ala Thr	
565 570 575	

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agc agc agc gac acc gga gtc tgc cat ggc gca cag ctc ttc agc 1776
 Ser Ser Ser Asp Met Thr Gly Val Cys His Gly Ala Gln Leu Phe Ser
 580 585 590

 gtc tgg aac gac aca taa 1794
 Val Trp Asn Asp Thr *
 595

 <210> SEQ ID NO 28
 <211> LENGTH: 597
 <212> TYPE: PRT
 <213> ORGANISM: oryza sativa

 <400> SEQUENCE: 28

 Met Ala Thr Met Asn Asn Trp Leu Ala Phe Ser Leu Ser Pro Gln Asp
 1 5 10 15

 Gln Leu Pro Pro Ser Gln Thr Asn Ser Thr Phe Ile Ser Ala Ala Ala
 20 25 30

 Thr Thr Thr Ala Gly Asp Ser Ser Thr Gly Asp Val Cys Phe Asn
 35 40 45

 Ile Pro Gln Ala His Pro Ser Thr Pro Ala Ile Gly Asn Gly Gly Ile
 50 55 60

 Gly Leu Ser Met Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Gln
 65 70 75 80

 Pro Ala Gln Ala Leu Ser Leu Ser Met Asn Met Ala Gly Thr Thr Thr
 85 90 95

 Ala Gln Gly Gly Ala Met Ala Leu Leu Ala Gly Ala Gly Glu Arg
 100 105 110

 Gly Arg Thr Thr Pro Ala Ser Glu Ser Leu Ser Thr Ser Ala His Gly
 115 120 125

 Ala Thr Thr Ala Thr Met Ala Gly Gly Arg Lys Glu Ile Asn Glu Glu
 130 135 140

 Gly Ser Gly Ser Ala Gly Ala Val Val Ala Val Gly Ser Glu Ser Gly
 145 150 155 160

 Gly Ser Gly Ala Val Val Glu Ala Gly Ala Ala Ala Ala Ala Arg
 165 170 175

 Lys Ser Val Asp Thr Phe Gly Gln Arg Thr Ser Ile Tyr Arg Gly Val
 180 185 190

 Thr Arg His Arg Trp Thr Gly Arg Tyr Glu Ala His Leu Trp Asp Asn
 195 200 205

 Ser Cys Arg Arg Glu Gly Gln Thr Arg Lys Gly Arg Gln Gly Tyr
 210 215 220

 Asp Lys Glu Glu Lys Ala Ala Arg Ala Tyr Asp Leu Ala Ala Leu Lys
 225 230 235 240

 Tyr Trp Gly Pro Thr Thr Thr Asn Phe Pro Val Asn Asn Tyr Glu
 245 250 255

 Lys Glu Leu Glu Glu Met Lys His Met Thr Arg Gln Glu Phe Val Ala
 260 265 270

 Ser Leu Arg Arg Lys Ser Ser Gly Phe Ser Arg Gly Ala Ser Ile Tyr
 275 280 285

 Arg Gly Val Thr Arg His His Gln His Gly Arg Trp Gln Ala Arg Ile
 290 295 300

 Gly Arg Val Ala Gly Asn Lys Asp Leu Tyr Leu Gly Thr Phe Ser Thr
 305 310 315 320

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Gln Glu Glu Ala Ala Glu Ala Tyr Asp Ile Ala Ala Ile Lys Phe Arg
 325 330 335
 Gly Leu Asn Ala Val Thr Asn Phe Asp Met Ser Arg Tyr Asp Val Lys
 340 345 350
 Ser Ile Leu Asp Ser Ala Ala Leu Pro Val Gly Thr Ala Ala Lys Arg
 355 360 365
 Leu Lys Asp Ala Glu Ala Ala Ala Tyr Asp Val Gly Arg Ile Ala
 370 375 380
 Ser His Leu Gly Gly Asp Gly Ala Tyr Ala Ala His Tyr Gly His His
 385 390 395 400
 His His Ser Ala Ala Ala Ala Trp Pro Thr Ile Ala Phe Gln Ala Ala
 405 410 415
 Ala Ala Pro Pro Pro His Ala Ala Gly Leu Tyr His Pro Tyr Ala Gln
 420 425 430
 Pro Leu Arg Gly Trp Cys Lys Gln Glu Gln Asp His Ala Val Ile Ala
 435 440 445
 Ala Ala His Ser Leu Gln Asp Leu His His Leu Asn Leu Gly Ala Ala
 450 455 460
 Ala Ala Ala His Asp Phe Phe Ser Gln Ala Met Gln Gln Gln His Gly
 465 470 475 480
 Leu Gly Ser Ile Asp Asn Ala Ser Leu Glu His Ser Thr Gly Ser Asn
 485 490 495
 Ser Val Val Tyr Asn Gly Asp Asn Gly Gly Gly Gly Gly Tyr Ile
 500 505 510
 Met Ala Pro Met Ser Ala Val Ser Ala Thr Ala Thr Ala Val Ala Ser
 515 520 525
 Ser His Asp His Gly Gly Asp Gly Gly Lys Gln Val Gln Met Gly Tyr
 530 535 540
 Asp Ser Tyr Leu Val Gly Ala Asp Ala Tyr Gly Gly Gly Gly Ala Gly
 545 550 555 560
 Arg Met Pro Ser Trp Ala Met Thr Pro Ala Ser Ala Pro Ala Ala Thr
 565 570 575
 Ser Ser Ser Asp Met Thr Gly Val Cys His Gly Ala Gln Leu Phe Ser
 580 585 590
 Val Trp Asn Asp Thr
 595

<210> SEQ ID NO 29
 <211> LENGTH: 655
 <212> TYPE: PRT
 <213> ORGANISM: Oryza sativa

<400> SEQUENCE: 29

Met Ala Ser Ala Asp Asn Trp Leu Gly Phe Ser Leu Ser Gly Gln Gly
 1 5 10 15
 Asn Pro Gln His His Gln Asn Gly Ser Pro Ser Ala Ala Gly Asp Ala
 20 25 30
 Ala Ile Asp Ile Ser Gly Ser Gly Asp Phe Tyr Gly Leu Pro Thr Pro
 35 40 45
 Asp Ala His His Ile Gly Met Ala Gly Glu Asp Ala Pro Tyr Gly Val
 50 55 60
 Met Asp Ala Phe Asn Arg Gly Thr His Glu Thr Gln Asp Trp Ala Met

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65	70	75	80
Arg Gly Leu Asp Tyr Gly Gly Ser Ser Asp Leu Ser Met Leu Val			
85	90	95	
Gly Ser Ser Gly Gly Arg Arg Thr Val Ala Gly Asp Gly Val Gly			
100	105	110	
Glu Ala Pro Lys Leu Glu Asn Phe Leu Asp Gly Asn Ser Phe Ser Asp			
115	120	125	
Val His Gly Gln Ala Ala Gly Gly Tyr Leu Tyr Ser Gly Ser Ala Val			
130	135	140	
Gly Gly Ala Gly Gly Tyr Ser Asn Gly Gly Cys Gly Gly Thr Ile			
145	150	155	160
Glu Leu Ser Met Ile Lys Thr Trp Leu Arg Ser Asn Gln Ser Gln Gln			
165	170	175	
Gln Pro Ser Pro Pro Gln His Ala Asp Gln Gly Met Ser Thr Asp Ala			
180	185	190	
Ser Ala Ser Ser Tyr Ala Cys Ser Asp Val Leu Val Gly Ser Cys Gly			
195	200	205	
Gly Gly Ala Gly Gly Thr Ala Ser Ser His Gly Gln Gly Leu Ala			
210	215	220	
Leu Ser Met Ser Thr Gly Ser Val Ala Ala Gly Gly Gly Ala			
225	230	235	240
Val Val Ala Ala Glu Ser Ser Ser Glu Asn Lys Arg Val Asp Ser			
245	250	255	
Pro Gly Gly Ala Val Asp Gly Ala Val Pro Arg Lys Ser Ile Asp Thr			
260	265	270	
Phe Gly Gln Arg Thr Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp			
275	280	285	
Thr Gly Arg Tyr Glu Ala His Leu Trp Asp Asn Ser Cys Arg Arg Glu			
290	295	300	
Gly Gln Ser Arg Lys Gly Arg Gln Gly Gly Tyr Asp Lys Glu Asp Lys			
305	310	315	320
Ala Ala Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Thr Thr			
325	330	335	
Thr Thr Thr Asn Phe Pro Met Ser Asn Tyr Glu Lys Glu Leu Glu Glu			
340	345	350	
Met Lys His Met Thr Arg Gln Glu Tyr Ile Ala His Leu Arg Arg Asn			
355	360	365	
Ser Ser Gly Phe Ser Arg Gly Ala Ser Lys Tyr Arg Gly Val Thr Arg			
370	375	380	
His His Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly			
385	390	395	400
Asn Lys Asp Ile Tyr Leu Gly Thr Phe Ser Thr Glu Glu Ala Ala			
405	410	415	
Glu Ala Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val			
420	425	430	
Thr Asn Phe Asp Met Ser Arg Tyr Asp Val Lys Ser Ile Leu Asp Ser			
435	440	445	
Ser Thr Leu Pro Val Gly Gly Ala Ala Arg Arg Leu Lys Glu Ala Glu			
450	455	460	
Val Ala Ala Ala Ala Gly Gly Val Ile Val Ser His Leu Ala			
465	470	475	480

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Asp Gly Gly Val Gly Gly Tyr Tyr Gly Cys Gly Pro Thr Ile Ala
 485 490 495

Phe Gly Gly Gly Gly Gln Gln Pro Ala Pro Leu Ala Val His Tyr Pro
 500 505 510

Ser Tyr Gly Gln Ala Ser Gly Trp Cys Lys Pro Glu Gln Asp Ala Val
 515 520 525

Ile Ala Ala Gly His Cys Ala Thr Asp Leu Gln His Leu His Leu Gly
 530 535 540

Ser Gly Gly Ala Ala Ala Thr His Asn Phe Phe Gln Gln Pro Ala Ser
 545 550 555 560

Ser Ser Ala Val Tyr Gly Asn Gly Gly Gly Gly Asn Ala Phe
 565 570 575

Met Met Pro Met Gly Ala Val Val Ala Ala Asp His Gly Gly Gln
 580 585 590

Ser Ser Ala Tyr Gly Gly Asp Glu Ser Gly Arg Leu Val Val Gly
 595 600 605

Tyr Asp Gly Val Val Asp Pro Tyr Ala Ala Met Arg Ser Ala Tyr Glu
 610 615 620

Leu Ser Gln Gly Ser Ser Ser Ser Val Ser Val Ala Lys Ala Ala
 625 630 635 640

Asn Gly Tyr Pro Asp Asn Trp Ser Ser Pro Phe Asn Gly Met Gly
 645 650 655

<210> SEQ ID NO 30

<211> LENGTH: 581

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 30

Met Asn Ser Met Asn Asn Trp Leu Gly Phe Ser Leu Ser Pro His Asp
 1 5 10 15

Gln Asn His His Arg Thr Asp Val Asp Ser Ser Thr Thr Arg Thr Ala
 20 25 30

Val Asp Val Ala Gly Gly Tyr Cys Phe Asp Leu Ala Ala Pro Ser Asp
 35 40 45

Glu Ser Ser Ala Val Gln Thr Ser Phe Leu Ser Pro Phe Gly Val Thr
 50 55 60

Leu Glu Ala Phe Thr Arg Asp Asn Asn Ser His Ser Arg Asp Trp Asp
 65 70 75 80

Ile Asn Gly Ala Cys Asn Asn Ile Asn Asn Asn Glu Gln Asn Gly
 85 90 95

Pro Lys Leu Glu Asn Phe Leu Gly Arg Thr Thr Ile Tyr Asn Thr
 100 105 110

Asn Glu Thr Val Val Asp Gly Asn Gly Asp Cys Gly Gly Asp Gly
 115 120 125

Gly Gly Gly Ser Leu Gly Leu Ser Met Ile Lys Thr Trp Leu Ser
 130 135 140

Asn His Ser Val Ala Asn Ala Asn His Gln Asp Asn Gly Asn Gly Ala
 145 150 155 160

Arg Gly Leu Ser Leu Ser Met Asn Ser Ser Thr Ser Asp Ser Asn Asn
 165 170 175

Tyr Asn Asn Asn Asp Asp Val Val Gln Glu Lys Thr Ile Val Asp Val

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180	185	190	
Val Glu Thr Thr Pro Lys Lys Thr Ile Glu Ser Phe Gly Gln Arg Thr			
195	200	205	
Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu			
210	215	220	
Ala His Leu Trp Asp Asn Ser Cys Lys Arg Glu Gly Gln Thr Arg Lys			
225	230	235	240
Gly Arg Gln Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala Arg Ala Tyr			
245	250	255	
Asp Leu Ala Ala Leu Lys Tyr Trp Gly Thr Thr Thr Thr Asn Phe			
260	265	270	
Pro Leu Ser Glu Tyr Glu Lys Glu Val Glu Glu Met Lys His Met Thr			
275	280	285	
Arg Gln Glu Tyr Val Ala Ser Leu Arg Arg Lys Ser Ser Gly Phe Ser			
290	295	300	
Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His Gln His Gly			
305	310	315	320
Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys Asp Leu Tyr			
325	330	335	
Leu Gly Thr Phe Gly Thr Gln Glu Glu Ala Ala Glu Ala Tyr Asp Ile			
340	345	350	
Ala Ala Ile Lys Phe Arg Gly Leu Ser Ala Val Thr Asn Phe Asp Met			
355	360	365	
Asn Arg Tyr Asn Val Lys Ala Ile Leu Glu Ser Pro Ser Leu Pro Ile			
370	375	380	
Gly Ser Ser Ala Lys Arg Leu Lys Asp Val Asn Asn Pro Val Pro Ala			
385	390	395	400
Met Met Ile Ser Asn Asn Val Ser Glu Ser Ala Asn Asn Val Ser Gly			
405	410	415	
Trp Gln Asn Thr Ala Phe Gln His His Gln Gly Met Asp Leu Ser Leu			
420	425	430	
Leu Gln Gln Gln Glu Arg Tyr Val Gly Tyr Tyr Asn Gly Gly Asn			
435	440	445	
Leu Ser Thr Glu Ser Thr Arg Val Cys Phe Lys Gln Glu Glu Gln			
450	455	460	
Gln His Phe Leu Arg Asn Ser Pro Ser His Met Thr Asn Val Asp His			
465	470	475	480
His Ser Ser Thr Ser Asp Asp Ser Val Thr Val Cys Gly Asn Val Val			
485	490	495	
Ser Tyr Gly Gly Tyr Gln Gly Phe Ala Ile Pro Val Gly Thr Ser Val			
500	505	510	
Asn Tyr Asp Pro Phe Thr Ala Ala Glu Ile Ala Tyr Asn Ala Arg Asn			
515	520	525	
His Tyr Tyr Tyr Ala Gln His Gln Gln Gln Gln Ile Gln Gln Ser			
530	535	540	
Pro Gly Gly Asp Phe Pro Val Ala Ile Ser Asn Asn His Ser Ser Asn			
545	550	555	560
Met Tyr Phe His Gly Glu Gly Gly Glu Gly Ala Pro Thr Phe Ser			
565	570	575	
Val Trp Asn Asp Thr			
580			

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<210> SEQ ID NO 31
<211> LENGTH: 584
<212> TYPE: PRT
<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 31

Met Asn Ser Met Asn Asn Trp Leu Gly Phe Ser Leu Ser Pro His Asp
1 5 10 15

Gln Asn His His Arg Thr Asp Val Asp Ser Ser Thr Thr Arg Thr Ala
20 25 30

Val Asp Val Ala Gly Gly Tyr Cys Phe Asp Leu Ala Ala Pro Ser Asp
35 40 45

Glu Ser Ser Ala Val Gln Thr Ser Phe Leu Ser Pro Phe Gly Val Thr
50 55 60

Leu Glu Ala Phe Thr Arg Asp Asn Asn His Ser Arg Asp Trp Asp
65 70 75 80

Ile Asn Gly Gly Ala Cys Asn Thr Leu Thr Asn Asn Glu Gln Asn Gly
85 90 95

Pro Lys Leu Glu Asn Phe Leu Gly Arg Thr Thr Ile Tyr Asn Thr
100 105 110

Asn Glu Thr Val Val Asp Gly Asn Gly Asp Cys Gly Gly Asp Gly
115 120 125

Gly Gly Gly Ser Leu Gly Leu Ser Met Ile Lys Thr Trp Leu Ser
130 135 140

Asn His Ser Val Ala Asn Ala Asn His Gln Asp Asn Gly Asn Gly Ala
145 150 155 160

Arg Gly Leu Ser Leu Ser Met Asn Ser Ser Thr Ser Asp Ser Asn Asn
165 170 175

Tyr Asn Asn Asn Asp Val Val Gln Glu Lys Thr Ile Val Asp Val
180 185 190

Val Glu Thr Thr Pro Lys Lys Thr Ile Glu Ser Phe Gly Gln Arg Thr
195 200 205

Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
210 215 220

Ala His Leu Trp Asp Asn Ser Cys Lys Arg Glu Gly Gln Thr Arg Lys
225 230 235 240

Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
245 250 255

Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Pro Thr Thr Thr
260 265 270

Thr Asn Phe Pro Leu Ser Glu Tyr Glu Lys Glu Val Glu Glu Met Lys
275 280 285

His Met Thr Arg Gln Glu Tyr Val Ala Ser Leu Arg Arg Lys Ser Ser
290 295 300

Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His
305 310 315 320

Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys
325 330 335

Asp Leu Tyr Leu Gly Thr Phe Gly Thr Gln Glu Glu Ala Ala Glu Ala
340 345 350

Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Ser Ala Val Thr Asn

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-continued

355	360	365	
Phe Asp Met Asn Arg Tyr Asn Val Lys Ala Ile Leu Glu Ser Pro Ser			
370	375	380	
Leu Pro Ile Gly Ser Ser Ala Lys Arg Leu Lys Asp Val Asn Asn Pro			
385	390	395	400
Val Pro Ala Met Met Ile Ser Asn Asn Val Ser Glu Ser Ala Asn Asn			
405	410	415	
Val Ser Gly Trp Gln Asn Thr Ala Phe Gln His His Gln Gly Met Asp			
420	425	430	
Leu Ser Leu Leu Gln Gln Gln Glu Arg Tyr Val Gly Tyr Tyr Asn			
435	440	445	
Gly Gly Asn Leu Ser Thr Glu Ser Thr Arg Val Cys Phe Lys Gln Glu			
450	455	460	
Glu Glu Gln His Phe Leu Arg Asn Ser Pro Ser His Met Thr Asn			
465	470	475	480
Val Asp His His Ser Ser Thr Ser Asp Asp Ser Val Thr Val Cys Gly			
485	490	495	
Asn Val Val Ser Tyr Gly Tyr Gln Gly Phe Ala Ile Pro Val Gly			
500	505	510	
Thr Ser Val Asn Tyr Asp Pro Phe Thr Ala Ala Glu Ile Ala Tyr Asn			
515	520	525	
Ala Arg Asn His Tyr Tyr Ala Gln His Gln Gln Gln Gln Ile			
530	535	540	
Gln Gln Ser Pro Gly Gly Asp Phe Pro Val Ala Ile Ser Asn Asn His			
545	550	555	560
Ser Ser Asn Met Tyr Phe His Gly Glu Gly Gly Glu Gly Ala Pro			
565	570	575	
Thr Phe Ser Val Trp Asn Asp Thr			
580			

<210> SEQ ID NO 32

<211> LENGTH: 579

<212> TYPE: PRT

<213> ORGANISM: Brassica napus

<400> SEQUENCE: 32

Met Asn Asn Asn Trp Leu Gly Phe Ser Leu Ser Pro Tyr Glu Gln Asn			
1	5	10	15
His His Arg Lys Asp Val Tyr Ser Ser Thr Thr Thr Thr Val Val Asp			
20	25	30	
Val Ala Gly Glu Tyr Cys Tyr Asp Pro Thr Ala Ala Ser Asp Glu Ser			
35	40	45	
Ser Ala Ile Gln Thr Ser Phe Pro Ser Pro Phe Gly Val Val Val Asp			
50	55	60	
Ala Phe Thr Arg Asp Asn Asn Ser His Ser Arg Asp Trp Asp Ile Asn			
65	70	75	80
Gly Cys Ala Cys Asn Asn Ile His Asn Asp Glu Gln Asp Gly Pro Lys			
85	90	95	
Leu Glu Asn Phe Leu Gly Arg Thr Thr Ile Tyr Asn Thr Asn Glu			
100	105	110	
Asn Val Gly Asp Gly Ser Gly Ser Gly Cys Tyr Gly Gly Asp Gly			
115	120	125	

-continued

Gly Gly Gly Ser Leu Gly Leu Ser Met Ile Lys Thr Trp Leu Arg Asn
 130 135 140
 Gln Pro Val Asp Asn Val Asp Asn Gln Glu Asn Gly Asn Ala Ala Lys
 145 150 155 160
 Gly Leu Ser Leu Ser Met Asn Ser Ser Thr Ser Cys Asp Asn Asn Asn
 165 170 175
 Asp Ser Asn Asn Asn Val Val Ala Gln Gly Lys Thr Ile Asp Asp Ser
 180 185 190
 Val Glu Ala Thr Pro Lys Lys Thr Ile Glu Ser Phe Gly Gln Arg Thr
 195 200 205
 Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
 210 215 220
 Ala His Leu Trp Asp Asn Ser Cys Lys Arg Glu Gly Gln Thr Arg Lys
 225 230 235 240
 Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
 245 250 255
 Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Thr Thr Thr Thr
 260 265 270
 Thr Asn Phe Pro Met Ser Glu Tyr Glu Lys Glu Val Glu Glu Met Lys
 275 280 285
 His Met Thr Arg Gln Glu Tyr Val Ala Ser Leu Arg Arg Lys Ser Ser
 290 295 300
 Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His
 305 310 315 320
 Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys
 325 330 335
 Asp Leu Tyr Leu Gly Thr Phe Gly Thr Gln Glu Glu Ala Ala Glu Ala
 340 345 350
 Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Thr Ala Val Thr Asn
 355 360 365
 Phe Asp Met Asn Arg Tyr Asn Val Lys Ala Ile Leu Glu Ser Pro Ser
 370 375 380
 Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Asn Arg Pro
 385 390 395 400
 Val Pro Ser Met Met Ile Ser Asn Asn Val Ser Glu Ser Glu Asn
 405 410 415
 Ser Ala Ser Gly Trp Gln Asn Ala Ala Val Gln His His Gln Gly Val
 420 425 430
 Asp Leu Ser Leu Leu His Gln His Gln Glu Arg Tyr Asn Gly Tyr Tyr
 435 440 445
 Tyr Asn Gly Gly Asn Leu Ser Ser Glu Ser Ala Arg Ala Cys Phe Lys
 450 455 460
 Gln Glu Asp Asp Gln His His Phe Leu Ser Asn Thr Gln Ser Leu Met
 465 470 475 480
 Thr Asn Ile Asp His Gln Ser Ser Val Ser Asp Asp Ser Val Thr Val
 485 490 495
 Cys Gly Asn Val Val Gly Tyr Gly Tyr Gln Gly Phe Ala Ala Pro
 500 505 510
 Val Asn Cys Asp Ala Tyr Ala Ala Ser Glu Phe Asp Tyr Asn Ala Arg
 515 520 525
 Asn His Tyr Tyr Phe Ala Gln Gln Gln Thr Gln Gln Ser Pro Gly

-continued

530 535 540

Gly Asp Phe Pro Ala Ala Met Thr Asn Asn Val Gly Ser Asn Met Tyr
 545 550 555 560
 Tyr His Gly Glu Gly Gly Glu Val Ala Pro Thr Phe Thr Val Trp
 565 570 575

Asn Asp Asn

<210> SEQ ID NO 33

<211> LENGTH: 579

<212> TYPE: PRT

<213> ORGANISM: Brassica napus

<400> SEQUENCE: 33

Met Asn Asn Asn Trp Leu Gly Phe Ser Leu Ser Pro Tyr Glu Gln Asn
 1 5 10 15

His His Arg Lys Asp Val Cys Ser Ser Thr Thr Thr Ala Val Asp
 20 25 30

Val Ala Gly Glu Tyr Cys Tyr Asp Pro Thr Ala Ala Ser Asp Glu Ser
 35 40 45

Ser Ala Ile Gln Thr Ser Phe Pro Ser Pro Phe Gly Val Val Leu Asp
 50 55 60

Ala Phe Thr Arg Asp Asn Asn Ser His Ser Arg Asp Trp Asp Ile Asn
 65 70 75 80

Gly Ser Ala Cys Asn Asn Ile His Asn Asp Glu Gln Asp Gly Pro Lys
 85 90 95

Leu Glu Asn Phe Leu Gly Arg Thr Thr Thr Ile Tyr Asn Thr Asn Glu
 100 105 110

Asn Val Gly Asp Ile Asp Gly Ser Gly Cys Tyr Gly Gly Asp Gly
 115 120 125

Gly Gly Gly Ser Leu Gly Leu Ser Met Ile Lys Thr Trp Leu Arg Asn
 130 135 140

Gln Pro Val Asp Asn Val Asp Asn Gln Glu Asn Gly Asn Gly Ala Lys
 145 150 155 160

Gly Leu Ser Leu Ser Met Asn Ser Ser Thr Ser Cys Asp Asn Asn Asn
 165 170 175

Tyr Ser Ser Asn Asn Leu Val Ala Gln Gly Lys Thr Ile Asp Asp Ser
 180 185 190

Val Glu Ala Thr Pro Lys Lys Thr Ile Glu Ser Phe Gly Gln Arg Thr
 195 200 205

Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu
 210 215 220

Ala His Leu Trp Asp Asn Ser Cys Lys Arg Glu Gly Gln Thr Arg Lys
 225 230 235 240

Gly Arg Gln Val Tyr Leu Gly Gly Tyr Asp Lys Glu Glu Lys Ala Ala
 245 250 255

Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Thr Thr Thr Thr
 260 265 270

Thr Asn Phe Pro Met Ser Glu Tyr Glu Lys Glu Ile Glu Glu Met Lys
 275 280 285

His Met Thr Arg Gln Glu Tyr Val Ala Ser Leu Arg Arg Lys Ser Ser
 290 295 300

Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His

-continued

305	310	315	320
Gln His Gly Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys			
325	330	335	
Asp Leu Tyr Leu Gly Thr Phe Gly Thr Gln Glu Glu Ala Ala Glu Ala			
340	345	350	
Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Thr Ala Val Thr Asn			
355	360	365	
Phe Asp Met Asn Arg Tyr Asn Val Lys Ala Ile Leu Glu Ser Pro Ser			
370	375	380	
Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Glu Ala Asn Arg Pro			
385	390	395	400
Val Pro Ser Met Met Ile Ser Asn Asn Val Ser Glu Ser Glu Asn			
405	410	415	
Asn Ala Ser Gly Trp Gln Asn Ala Ala Val Gln His His Gln Gly Val			
420	425	430	
Asp Leu Ser Leu Leu Gln Gln His Gln Glu Arg Tyr Asn Gly Tyr Tyr			
435	440	445	
Tyr Asn Gly Gly Asn Leu Ser Ser Glu Ser Ala Arg Ala Cys Phe Lys			
450	455	460	
Gln Glu Asp Asp Gln His His Phe Leu Ser Asn Thr Gln Ser Leu Met			
465	470	475	480
Thr Asn Ile Asp His Gln Ser Ser Val Ser Asp Asp Ser Val Thr Val			
485	490	495	
Cys Gly Asn Val Val Gly Tyr Gly Tyr Gln Gly Phe Ala Ala Pro			
500	505	510	
Val Asn Cys Asp Ala Tyr Ala Ala Ser Glu Phe Asp Tyr Asn Ala Arg			
515	520	525	
Asn His Tyr Tyr Phe Ala Gln Gln Gln Thr Gln His Ser Pro Gly			
530	535	540	
Gly Asp Phe Pro Ala Ala Met Thr Asn Asn Val Gly Ser Asn Met Tyr			
545	550	555	560
Tyr His Gly Glu Gly Gly Glu Val Ala Pro Thr Phe Thr Val Trp			
565	570	575	
Asn Asp Asn			

<210> SEQ ID NO 34

<211> LENGTH: 565

<212> TYPE: PRT

<213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 34

Met Asn Ser Asn Asn Trp Leu Ala Phe Pro Leu Ser Pro Thr His Ser			
1	5	10	15
Ser Leu Pro Pro His Ile His Ser Ser Gln Asn Ser His Phe Asn Leu			
20	25	30	
Gly Leu Val Asn Asp Asn Ile Asp Asn Pro Phe Gln Asn Gln Gly Trp			
35	40	45	
Asn Met Ile Asn Pro His Gly Gly Gly Glu Gly Glu Val Pro			
50	55	60	
Lys Val Ala Asp Phe Leu Gly Val Ser Lys Ser Gly Asp His His Thr			
65	70	75	80
Asp His Asn Leu Val Pro Tyr Asn Asp Ile His Gln Thr Asn Ala Ser			

-continued

85	90	95	
Asp Tyr Tyr Phe Gln Thr Asn Ser Leu Leu Pro Thr Val Val Thr Cys			
100	105	110	
Ala Ser Asn Ala Pro Asn Asn Tyr Glu Leu Gln Glu Ser Ala His Asn			
115	120	125	
Leu Gln Ser Leu Thr Leu Ser Met Gly Ser Thr Gly Ala Ala Ala Ala			
130	135	140	
Glu Val Ala Thr Val Lys Ala Ser Pro Ala Glu Thr Ser Ala Asp Asn			
145	150	155	160
Ser Ser Ser Thr Thr Asn Thr Ser Gly Gly Ala Ile Val Glu Ala Thr			
165	170	175	
Pro Arg Arg Thr Leu Glu Thr Phe Gly Gln Arg Thr Ser Ile Tyr Arg			
180	185	190	
Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu Ala His Leu Trp			
195	200	205	
Asp Asn Ser Cys Arg Arg Glu Gly Gln Ser Arg Lys Gly Arg Gln Gly			
210	215	220	
Gly Tyr Asp Lys Glu Glu Lys Ala Ala Arg Ala Tyr Asp Leu Ala Ala			
225	230	235	240
Leu Lys Tyr Trp Gly Pro Ser Thr Thr Asn Phe Pro Ile Thr Asn			
245	250	255	
Tyr Glu Lys Glu Val Glu Glu Met Lys Asn Met Thr Arg Gln Glu Phe			
260	265	270	
Val Ala Ser Ile Arg Arg Lys Ser Ser Gly Phe Ser Arg Gly Ala Ser			
275	280	285	
Met Tyr Arg Gly Val Thr Arg His His Gln His Gly Arg Trp Gln Ala			
290	295	300	
Arg Ile Gly Arg Val Ala Gly Asn Lys Asp Leu Tyr Leu Gly Thr Phe			
305	310	315	320
Ser Thr Glu Glu Ala Ala Glu Ala Tyr Asp Ile Ala Ala Ile Lys			
325	330	335	
Phe Arg Gly Leu Asn Ala Val Thr Asn Phe Glu Ile Asn Arg Tyr Asp			
340	345	350	
Val Lys Ala Ile Leu Glu Ser Asn Thr Leu Pro Ile Gly Gly Ala			
355	360	365	
Ala Lys Arg Leu Lys Glu Ala Gln Ala Leu Glu Ser Ser Arg Lys Arg			
370	375	380	
Glu Glu Met Ile Ala Leu Gly Ser Asn Phe His Gln Tyr Gly Ala Ala			
385	390	395	400
Ser Gly Ser Ser Ser Val Ala Ser Ser Arg Leu Gln Leu Gln Pro			
405	410	415	
Tyr Pro Leu Ser Ile Gln Gln Pro Phe Glu His Leu His His His Gln			
420	425	430	
Pro Leu Leu Thr Leu Gln Asn Asn Asp Ile Ser Gln Tyr His Asp			
435	440	445	
Ser Phe Ser Tyr Ile Gln Thr Gln Leu His Leu His Gln Gln Gln Thr			
450	455	460	
Asn Asn Tyr Leu Gln Ser Ser Ser His Thr Ser Gln Leu Tyr Asn Ala			
465	470	475	480
Tyr Leu Gln Ser Asn Pro Gly Leu Leu His Gly Phe Val Ser Asp Asn			
485	490	495	

-continued

Asn	Asn	Thr	Ser	Gly	Phe	Leu	Gly	Asn	Asn	Gly	Ile	Gly	Ile	Gly	Ser
500								505					510		

Ser	Ser	Thr	Val	Gly	Ser	Ser	Ala	Glu	Glu	Glu	Phe	Pro	Ala	Val	Lys
515							520					525			

Val	Asp	Tyr	Asp	Met	Pro	Pro	Ser	Gly	Gly	Ala	Thr	Gly	Tyr	Gly	Gly
530							535					540			

Trp	Asn	Ser	Gly	Glu	Ser	Ala	Gln	Gly	Ser	Asn	Pro	Gly	Gly	Val	Phe
545				550				555					560		

Thr	Met	Trp	Asn	Glu											
				565											

<210> SEQ_ID NO 35
 <211> LENGTH: 540
 <212> TYPE: PRT
 <213> ORGANISM: Arabidopsis thaliana

<400> SEQUENCE: 35

Met	Asp	Asn	Pro	Phe	Gln	Thr	Gln	Glu	Trp	Asn	Met	Ile	Asn	Pro	His
1				5				10				15			

Gly	Gly	Gly	Gly	Asp	Glu	Gly	Gly	Glu	Val	Pro	Lys	Val	Ala	Asp	Phe
20					25						30				

Leu	Gly	Val	Ser	Lys	Pro	Asp	Glu	Asn	Gln	Ser	Asn	His	Leu	Val	Ala
35					40						45				

Tyr	Asn	Asp	Ser	Asp	Tyr	Tyr	Phe	His	Thr	Asn	Ser	Leu	Met	Pro	Ser
50					55				60						

Val	Gln	Ser	Asn	Asp	Val	Val	Val	Ala	Ala	Cys	Asp	Ser	Asn	Thr	Pro
65					70				75				80		

Asn	Asn	Ser	Ser	Tyr	His	Glu	Leu	Gln	Glu	Ser	Ala	His	Asn	Leu	Gln
85					90				95						

Ser	Leu	Thr	Leu	Ser	Met	Gly	Thr	Thr	Ala	Gly	Asn	Asn	Val	Val	Asp
100					105						110				

Lys	Ala	Ser	Pro	Ser	Glu	Thr	Thr	Gly	Asp	Asn	Ala	Ser	Gly	Gly	Ala
115					120						125				

Leu	Ala	Val	Val	Glu	Thr	Ala	Thr	Pro	Arg	Arg	Ala	Leu	Asp	Thr	Phe
130				135					140						

Gly	Gln	Arg	Thr	Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	Arg	Trp	Thr
145					150				155			160			

Gly	Arg	Tyr	Glu	Ala	His	Leu	Trp	Asp	Asn	Ser	Cys	Arg	Arg	Glu	Gly
165					170				175						

Gln	Ser	Arg	Lys	Gly	Arg	Gln	Gly	Gly	Tyr	Asp	Lys	Glu	Asp	Lys	Ala
180					185						190				

Ala	Arg	Ser	Tyr	Asp	Leu	Ala	Ala	Leu	Lys	Tyr	Trp	Gly	Pro	Ser	Thr
195					200					205					

Thr	Thr	Asn	Phe	Pro	Ile	Thr	Asn	Tyr	Glu	Lys	Glu	Val	Glu	Glu	Met
210					215					220					

Lys	His	Met	Thr	Arg	Gln	Glu	Phe	Val	Ala	Ala	Ile	Arg	Arg	Lys	Ser
225					230				235			240			

Ser	Gly	Phe	Ser	Arg	Gly	Ala	Ser	Met	Tyr	Arg	Gly	Val	Thr	Arg	His
245					250					255					

His	Gln	His	Gly	Arg	Trp	Gln	Ala	Arg	Ile	Gly	Arg	Val	Ala	Gly	Asn
260					265					270					

Lys	Asp	Leu	Tyr	Leu	Gly	Thr	Phe	Ser	Thr	Glu	Glu	Ala	Ala	Glu	
-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	-----	--

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275	280	285
Ala Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr		
290	295	300
Asn Phe Glu Ile Asn Arg Tyr Asp Val Lys Ala Ile Leu Glu Ser Ser		
305	310	315
320		
Thr Leu Pro Ile Gly Gly Ala Ala Lys Arg Leu Lys Glu Ala Gln		
325	330	335
Ala Leu Glu Ser Ser Arg Lys Arg Glu Ala Glu Met Ile Ala Leu Gly		
340	345	350
Ser Ser Phe Gln Tyr Gly Gly Ser Ser Thr Gly Ser Gly Ser Thr		
355	360	365
Ser Ser Arg Leu Gln Leu Gln Pro Tyr Pro Leu Ser Ile Gln Gln Pro		
370	375	380
Leu Glu Pro Phe Leu Ser Leu Gln Asn Asn Asp Ile Ser His Tyr Asn		
385	390	395
400		
Asn Asn Asn Ala His Asp Ser Ser Ser Phe Asn His His Ser Tyr Ile		
405	410	415
Gln Thr Gln Leu His Leu His Gln Gln Thr Asn Asn Tyr Leu Gln Gln		
420	425	430
Gln Ser Ser Gln Asn Ser Gln Leu Tyr Asn Ala Tyr Leu His Ser		
435	440	445
Asn Pro Ala Leu Leu His Gly Leu Val Ser Thr Ser Ile Val Asp Asn		
450	455	460
Asn Asn Asn Gly Gly Ser Ser Gly Ser Tyr Asn Thr Ala Ala Phe		
465	470	475
480		
Leu Gly Asn His Gly Ile Gly Ile Gly Ser Ser Ser Thr Val Gly Ser		
485	490	495
Thr Glu Glu Phe Pro Thr Val Lys Thr Asp Tyr Asp Met Pro Ser Ser		
500	505	510
Asp Gly Thr Gly Gly Tyr Ser Gly Trp Thr Ser Glu Ser Val Gln Gly		
515	520	525
Ser Asn Pro Gly Gly Val Phe Thr Met Trp Asn Glu		
530	535	540

<210> SEQ ID NO 36

<211> LENGTH: 516

<212> TYPE: PRT

<213> ORGANISM: *Arabidopsis thaliana*

<400> SEQUENCE: 36

Met Ile Asn Pro His Gly Gly Gly Glu Gly Gly Glu Val Pro Lys
1 5 10 15

Val Ala Asp Phe Leu Gly Val Ser Lys Ser Gly Asp His His Thr Asp
20 25 30

His Asn Leu Val Pro Tyr Asn Asp Ile His Gln Thr Asn Ala Ser Asp
35 40 45

Tyr Tyr Phe Gln Thr Asn Ser Leu Leu Pro Thr Val Val Thr Cys Ala
50 55 60

Ser Asn Ala Pro Asn Asn Tyr Glu Leu Gln Glu Ser Ala His Asn Leu
65 70 75 80

Gln Ser Leu Thr Leu Ser Met Gly Ser Thr Gly Ala Ala Ala Glu
85 90 95

-continued

Val Ala Thr Val Lys Ala Ser Pro Ala Glu Thr Ser Ala Asp Asn Ser
 100 105 110

Ser Ser Thr Thr Asn Thr Ser Gly Gly Ala Ile Val Glu Ala Thr Pro
 115 120 125

Arg Arg Thr Leu Glu Thr Phe Gly Gln Arg Thr Ser Ile Tyr Arg Gly
 130 135 140

Val Thr Arg His Arg Trp Thr Gly Arg Tyr Glu Ala His Leu Trp Asp
 145 150 155 160

Asn Ser Cys Arg Arg Glu Gly Gln Ser Arg Lys Gly Arg Gln Gly Gly
 165 170 175

Tyr Asp Lys Glu Glu Lys Ala Ala Arg Ala Tyr Asp Leu Ala Ala Leu
 180 185 190

Lys Tyr Trp Gly Pro Ser Thr Thr Asn Phe Pro Ile Thr Asn Tyr
 195 200 205

Glu Lys Glu Val Glu Glu Met Lys Asn Met Thr Arg Gln Glu Phe Val
 210 215 220

Ala Ser Ile Arg Arg Lys Ser Ser Gly Phe Ser Arg Gly Ala Ser Met
 225 230 235 240

Tyr Arg Gly Val Thr Arg His His Gln His Gly Arg Trp Gln Ala Arg
 245 250 255

Ile Gly Arg Val Ala Gly Asn Lys Asp Leu Tyr Leu Gly Thr Phe Ser
 260 265 270

Thr Glu Glu Glu Ala Ala Glu Ala Tyr Asp Ile Ala Ala Ile Lys Phe
 275 280 285

Arg Gly Leu Asn Ala Val Thr Asn Phe Glu Ile Asn Arg Tyr Asp Val
 290 295 300

Lys Ala Ile Leu Glu Ser Asn Thr Leu Pro Ile Gly Gly Ala Ala
 305 310 315 320

Lys Arg Leu Lys Glu Ala Gln Ala Leu Glu Ser Ser Arg Lys Arg Glu
 325 330 335

Glu Met Ile Ala Leu Gly Ser Asn Phe His Gln Tyr Gly Ala Ala Ser
 340 345 350

Gly Ser Ser Ser Val Ala Ser Ser Ser Arg Leu Gln Leu Gln Pro Tyr
 355 360 365

Pro Leu Ser Ile Gln Gln Pro Phe Glu His Leu His His Gln Pro
 370 375 380

Leu Leu Thr Leu Gln Asn Asn Asn Asp Ile Ser Gln Tyr His Asp Ser
 385 390 395 400

Phe Ser Tyr Ile Gln Thr Gln Leu His Leu His Gln Gln Gln Thr Asn
 405 410 415

Asn Tyr Leu Gln Ser Ser Ser His Thr Ser Gln Leu Tyr Asn Ala Tyr
 420 425 430

Leu Gln Ser Asn Pro Gly Leu Leu His Gly Phe Val Ser Asp Asn Asn
 435 440 445

Asn Thr Ser Gly Phe Leu Gly Asn Asn Gly Ile Gly Ile Gly Ser Ser
 450 455 460

Ser Thr Val Gly Ser Ser Ala Glu Glu Phe Pro Ala Val Lys Val
 465 470 475 480

Asp Tyr Asp Met Pro Pro Ser Gly Gly Ala Thr Gly Tyr Gly Gly Trp
 485 490 495

Asn Ser Gly Glu Ser Ala Gln Gly Ser Asn Pro Gly Gly Val Phe Thr

-continued

500

505

510

Met Trp Asn Glu
515

<210> SEQ ID NO 37
<211> LENGTH: 65
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: consensus sequence of Figure 1

<400> SEQUENCE: 37

Met Ser Asn Asn Trp Leu Gly Phe Ser Leu Ser Pro Asp Gln Ser Ser
1 5 10 15
Ala Val Asp Ala Phe Ile Gly Ser Val Asp Phe Asn Ser His Arg Asp
20 25 30
Asn Ala Asn Ile Asn Ser Gly Pro Glu Asn Phe Ser Ile Gly Gly Gly
35 40 45
Gly Gly Ile Gly Leu Ser Met Ile Lys Thr Trp Leu Arg Asn Gln Pro
50 55 60

Asn
65

<210> SEQ ID NO 38
<211> LENGTH: 62
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: APETALA2 PFAM consensus sequence

<400> SEQUENCE: 38

Ser Lys Tyr Arg Gly Val Arg Gln Arg Pro Trp Gly Lys Trp Val Ala
1 5 10 15
Glu Ile Arg Asp Pro Arg Lys Gly Thr Arg Val Trp Leu Gly Thr Phe
20 25 30
Asp Thr Ala Glu Glu Ala Ala Arg Ala Tyr Asp Val Ala Ala Leu Lys
35 40 45
Leu Arg Gly Pro Ser Ala Val Leu Asn Phe Pro Asn Glu Leu
50 55 60

1. An isolated polynucleotide selected from the group consisting of:

- a polynucleotide comprising SEQ ID NO:1 or 3;
- a polynucleotide encoding the amino acid sequence of SEQ ID NO:2;
- a polynucleotide having at least 75% sequence identity to SEQ ID NO:1 or 3, wherein said polynucleotide encodes a polypeptide comprising two APETALA2 (AP2) domains and having Ovule Development Protein 2 (ODP2) activity; and
- a polynucleotide encoding an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said polynucleotide encodes a polypeptide comprising two APETALA2 (AP2) domains and having ODP2 activity.

2. The isolated polynucleotide of claim 1, wherein said polynucleotide encodes an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said

polynucleotide encodes a polypeptide comprising two APETALA2 (AP2) domains and having ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO: 2 by one amino acid residue;
- the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO: 2 by one amino acid residue; and
- the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

3. The isolated polynucleotide of claim **1**, wherein said polynucleotide encodes an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said polynucleotide encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity, and wherein said polypeptide comprises the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 by one amino acid residue.

4. The isolated polynucleotide of claim **3**, wherein said polynucleotide further comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 by one amino acid residue; and
- b) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 by one amino acid residue.

5. An expression cassette comprising the polynucleotide of claim **1**, wherein said polynucleotide is operably linked to a promoter that drives expression in a plant.

6. A plant comprising a heterologous polynucleotide comprising a first nucleotide sequence operably linked to a promoter that drives expression in the plant, wherein said first nucleotide sequence is selected from the group consisting of:

- a) a nucleotide sequence comprising SEQ ID NO:1, 3, 25 or 27;
- b) a nucleotide sequence encoding the amino acid sequence of SEQ ID NO:2, 26 or 28;
- c) a nucleotide sequence having at least 75% sequence identity to SEQ ID NO:1, 3, 25 or 27, wherein said nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity; and
- d) a nucleotide sequence encoding an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, 26 or 28, wherein said nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity.

7. The plant of claim **6**, wherein said first nucleotide sequence encodes an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said first nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 by one amino acid residue;
- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 by one amino acid residue; and
- c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 by one amino acid residue.

responding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

8. The plant of claim **6**, wherein said plant comprises a plant part selected from the group consisting of a cell, a seed, and a grain.

9. The plant of claim **6**, wherein said promoter is a constitutive promoter, a tissue-preferred promoter, an inducible promoter, or a developmentally regulated promoter.

10. The plant of claim **6**, wherein said plant is a monocot.

11. The plant of claim **10**, wherein said monocot is maize, wheat, rice, barley, sorghum, or rye.

12. The plant of claim **6**, wherein said plant is a dicot.

13. The plant of claim **6**, wherein said heterologous polynucleotide is stably incorporated into the genome of the plant.

14. A method of increasing the activity of a polypeptide in a plant comprising providing to said plant a polypeptide selected from the group consisting of:

- (a) a polypeptide comprising the amino acid sequence of SEQ ID NO:2, or 28; and,
- (b) a polypeptide having at least 75% sequence identity to SEQ ID NO:2, or 28, wherein said polypeptide comprises two APETELA2 (AP2) domains and has ODP2 activity;

and wherein increasing the activity of said polypeptide increases the transformation efficiency of the plant.

15. The method of claim **14**, wherein said polypeptide has at least 75% sequence identity to SEQ ID NO:2, wherein said polypeptide comprises two APETELA2 (AP2) domains and has ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 by one amino acid residue;
- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 by one amino acid residue; and
- c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 by one amino acid residue.

16. The method of claim **14**, wherein providing the polypeptide comprises introducing into said plant a heterologous polynucleotide selected from the group consisting of:

- (a) a polynucleotide comprising SEQ ID NO:1, 3, or 27;
- (b) a polynucleotide encoding the amino acid sequence of SEQ ID NO:2, or 28;
- (c) a polynucleotide having at least 75% sequence identity to SEQ ID NO:1, or 27, wherein said polynucleotide encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity; and,
- (d) a polynucleotide encoding an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2 or 28, wherein said polynucleotide encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity.

17. The method of claim **16**, wherein said polynucleotide encodes an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said polynucle-

otide encodes a polypeptide comprising two APETALA2 (AP2) domains and having ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO: 2 by one amino acid residue;
- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO: 2 by one amino acid residue; and
- c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

18. The method of claim 16, wherein said heterologous polynucleotide is transiently expressed.

19. The method of claim 16, wherein said heterologous polynucleotide is stably integrated into the genome of the plant.

20. The method of claim 16, wherein said heterologous polynucleotide is operably linked to a constitutive promoter, a tissue-preferred promoter, an inducible promoter, or a developmentally regulated promoter.

21. The method of claim 14, wherein said plant is a dicot.

22. The method of claim 14, wherein said plant is a monocot.

23. The method of claim 22, wherein said monocot is maize, wheat, rice, barley, sorghum, or rye.

24. A method of transforming a target plant comprising

- a) providing to a plant a polypeptide comprising an amino acid sequence selected from the group consisting of:
 - i) the amino acid sequence of SEQ ID NO:2, 26, or 28; and,
 - ii) an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, 26, or 28, wherein said polypeptide comprises two APETALA2 (AP2) domains and has ODP2 activity;

thereby producing a target plant; and

- b) transforming into said target plant a polynucleotide of interest.

25. The method of claim 24, wherein said polypeptide has at least 75% sequence identity to SEQ ID NO:2, wherein said polypeptide comprises two APETALA2 (AP2) domains and has ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO: 2 by one amino acid residue;
- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO: 2 by one amino acid residue; and
- c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

responding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

26. The method of claim 24, wherein providing the polypeptide comprises introducing into said plant a heterologous polynucleotide selected from the group consisting of:

- a) a polynucleotide comprising SEQ ID NO:1, 3, 25, or 27;
- b) a polynucleotide encoding the amino acid sequence of SEQ ID NO:2, 26, or 28;
- c) a polynucleotide having at least 75% sequence identity to SEQ ID NO:1, 3, 25, or 27, wherein said polynucleotide encodes a polypeptide comprising two APETALA2 (AP2) domains and having ODP2 activity; and
- d) a polynucleotide encoding an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, 26 or 28, wherein said polynucleotide encodes a polypeptide comprising two APETALA2 (AP2) domains and having ODP2 activity.

27. The method of claim 26, wherein said polynucleotide encodes an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said polynucleotide encodes a polypeptide comprising two APETALA2 (AP2) domains and having ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO: 2 by one amino acid residue;
- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO: 2 by one amino acid residue; and
- c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

28-31. (canceled)

32. The expression cassette of claim 5, wherein said expression cassette further comprises a polynucleotide encoding a Wuschel polypeptide, wherein said polynucleotide encoding a Wuschel polypeptide is operably linked to a promoter that drives expression in a plant.

33. The expression cassette of claim 5, wherein said promoter is an oleosin promoter, an In2 promoter, or an ubiquitin promoter.

34. An expression cassette comprising a polynucleotide comprising a first nucleotide sequence operably linked to a promoter, wherein said first nucleotide sequence is selected from the group consisting of:

- a) a nucleotide sequence comprising SEQ ID NO:1 or 3;
- b) a nucleotide sequence encoding the amino acid sequence of SEQ ID NO:2;
- c) a nucleotide sequence having at least 75% sequence identity to SEQ ID NO:1 or 3, wherein said nucleotide sequence encodes a polypeptide comprising two APETALA2 (AP2) domains and having ODP2 activity; and
- d) a nucleotide sequence encoding an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said nucleotide sequence encodes a polypeptide

comprising two APETELA2 (AP2) domains and having ODP2 activity, and wherein said first nucleotide sequence is flanked by recombination sites.

35. The expression cassette of claim **34**, wherein said first nucleotide sequence encodes an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said first nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO: 2 by one amino acid residue;
- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO: 2 by one amino acid residue; and
- c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

36. The expression cassette of claim **34**, wherein said recombination sites comprise frt or loxP sites.

37. The expression cassette of claim **34**, wherein said recombination sites flank said polynucleotide.

38. The expression cassette of claim **37**, wherein said polynucleotide further comprises a second nucleotide sequence encoding a Wuschel polypeptide, wherein said polynucleotide encoding a Wuschel polypeptide is operably linked to a promoter that drives expression in a plant.

39. A plant comprising the expression cassette of claim **34**.

40. The plant of claim **6**, wherein said plant further comprises a polynucleotide encoding a Wuschel polypeptide, wherein said polynucleotide encoding a Wuschel polypeptide is operably linked to a promoter that drives expression in the plant.

41. The plant of claim **6**, wherein said first nucleotide sequence is flanked by recombination sites.

42. The plant of claim **41**, wherein said recombination sites comprise frt or loxP sites.

43. The plant of claim **41**, wherein said recombination sites flank said heterologous polynucleotide.

44. The plant of claim **43**, wherein said heterologous polynucleotide further comprises a second nucleotide sequence, wherein said second nucleotide sequence encodes a Wuschel polypeptide, and wherein said second nucleotide sequence is operably linked to a promoter that drives expression in the plant.

45. The plant of claim **41**, wherein said plant further comprises a polynucleotide encoding a recombinase, wherein said polynucleotide encoding a recombinase is operably linked to a promoter that drives expression in the plant.

46. The plant of claim **45**, wherein said recombinase comprises Cre or Flp.

47. The plant of claim **11**, wherein said plant is an elite maize inbred.

48. A method of transiently increasing the activity of a polypeptide in a plant cell, said method comprising the steps of:

a) providing to a plant cell a polypeptide selected from the group consisting of:

- (i) a polypeptide comprising the amino acid sequence of SEQ ID NO:2, 26, or 28; and,
- (ii) a polypeptide having at least 75% sequence identity to SEQ ID NO:2, 26, or 28, wherein said polypeptide comprises two APETELA2 (AP2) domains and has ODP2 activity; and

b) reducing the activity, level, or activity and level of said polypeptide in said plant cell prior to regeneration of a plant from said plant cell.

49. The method of claim **48**, wherein said polypeptide has at least 75% sequence identity to SEQ ID NO:2, wherein said polypeptide comprises two APETELA2 (AP2) domains and has ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO: 2 by one amino acid residue;
- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO: 2 by one amino acid residue; and
- c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

50. The method of claim **48**, wherein providing the polypeptide comprises introducing into said plant cell a heterologous polynucleotide comprising a first nucleotide sequence selected from the group consisting of:

- (a) a nucleotide sequence comprising SEQ ID NO:1, 3, 25, or 27;
- (b) a nucleotide sequence encoding the amino acid sequence of SEQ ID NO:2, 26, or 28;
- (c) a nucleotide sequence having at least 75% sequence identity to SEQ ID NO:1, 3, 25, or 27, wherein said nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity; and,
- (d) a nucleotide sequence encoding an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, 26, or 28, wherein said nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity.

51. The method of claim **50**, wherein said first nucleotide sequence encodes an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said first nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO: 2 by one amino acid residue;
- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid

sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO: 2 by one amino acid residue; and

c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

52. The method of claim **50**, wherein said first nucleotide sequence is operably linked to a constitutive promoter, a tissue-preferred promoter, an inducible promoter, or a developmentally regulated promoter.

53. The method of claim **48**, wherein said plant is a dicot.

54. The method of claim **48**, wherein said plant is a monocot.

55. The method of claim **54**, wherein said monocot is maize, wheat, rice, barley, sorghum, or rye.

56. The method of claim **50**, wherein said reducing the level of said polypeptide in said plant cell prior to regeneration of a plant from said plant cell comprises excising said first nucleotide sequence.

57. The method of claim **56**, wherein said first nucleotide sequence is flanked by recombination sites.

58. The method of claim **57**, wherein said recombination sites comprise frt or loxP sites.

59. The method of claim **56**, wherein said plant cell further comprises a second nucleotide sequence encoding a recombinase and wherein said method further comprises expressing said recombinase prior to regeneration of a plant from said plant cell.

60. The method of claim **59**, wherein said recombinase comprises Cre or Flp.

61. A method of transforming a plant cell comprising:

- (a) transforming a plant cell with a polynucleotide of interest and a heterologous polynucleotide comprising a first nucleotide sequence selected from the group consisting of:
 - a) a nucleotide sequence comprising SEQ ID NO:1, 3, 25, or 27;
 - b) a nucleotide sequence encoding the amino acid sequence of SEQ ID NO:2, 26, or 28;
 - c) a nucleotide sequence having at least 75% sequence identity to SEQ ID NO:1, 3, 25, or 27, wherein said nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity; and
 - d) a nucleotide sequence encoding an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, 26 or 28, wherein said nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity.

62. The method of claim **61**, wherein said first nucleotide sequence encodes an amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, wherein said first nucleotide sequence encodes a polypeptide comprising two APETELA2 (AP2) domains and having ODP2 activity, and wherein said polypeptide comprises at least one of the amino acid sequences selected from the group consisting of:

- a) the amino acid sequence corresponding to amino acid residues 5-14 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence cor-

responding to amino acid residues 5-14 of SEQ ID NO: 2 by one amino acid residue;

- b) the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 156-169 of SEQ ID NO: 2 by one amino acid residue; and
- c) the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO:2 or an amino acid sequence that differs from the amino acid sequence corresponding to amino acid residues 704-709 of SEQ ID NO: 2 by one amino acid residue.

63. The method of claim **61**, wherein said method further comprises transforming said plant cell with a polynucleotide encoding a Wuschel polypeptide.

64. The method of claim **61**, wherein said method further comprises excising said first nucleotide sequence prior to regenerating said plant cell into a plant.

65. The method of claim **64**, wherein said first nucleotide sequence is flanked by recombination sites.

66. The method of claim **65**, wherein said recombination sites comprise frt or loxP sites.

67. The method of claim **65**, wherein said recombination sites flank said heterologous polynucleotide.

68. The method of claim **67**, wherein said heterologous polynucleotide further comprises a second nucleotide sequence, wherein said second nucleotide sequence encodes a Wuschel polypeptide.

69. The method of claim **64**, wherein said method further comprises introducing into said plant cell a polynucleotide encoding a recombinase and expressing said recombinase prior to regenerating a plant from said plant cell.

70. The method of claim **69**, wherein said recombinase comprises Cre or Flp.

71. The method of claim **61**, wherein said plant cell is a cell of a mature seed, a leaf, or a stem

72. The method of claim **61**, wherein said plant cell is a cell of a recalcitrant plant or recalcitrant explant.

73. The method of claim **72**, wherein said recalcitrant plant is an elite maize inbred.

74. An isolated polypeptide comprising an amino acid sequence selected from the group consisting of:

- (a) the polypeptide comprising the amino acid sequence of SEQ ID NO:2; and,
- (b) the polypeptide having at least 75% sequence identity to SEQ ID NO:2, wherein said polypeptide comprises two AP2 domains and has ODP2 activity.

75. A method for producing haploid plant embryos comprising providing to a haploid plant cell, a polypeptide comprising an amino acid sequence selected from the group consisting of:

- (a) the amino acid sequence of SEQ ID NO:2, 26, or 28;
- (b) the amino acid sequence having at least 75% sequence identity to SEQ ID NO:2, 26, or 28, wherein said polypeptide comprises two AP2 domains and has ODP2 activity; and,

thereby inducing embryogenesis.