



(51) International Patent Classification:

C12N 11/14 (2006.01) G01N 27/327 (2006.01)
C12Q 1/32 (2006.01)

(21) International Application Number:

PCT/US2016/049367

(22) International Filing Date:

30 August 2016 (30.08.2016)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

14/840,694 31 August 2015 (31.08.2015) US

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(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Published:

— with international search report (Art. 21(3))

(54) Title: FORMALDEHYDE GRAPHENE SENSOR

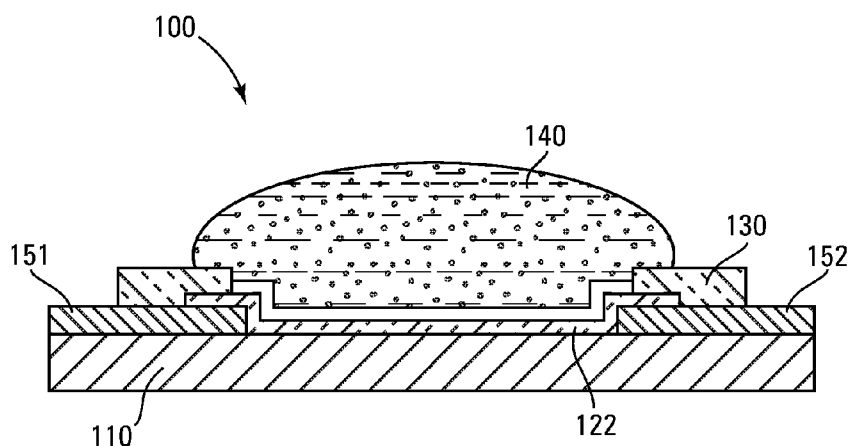


Fig. 2

(57) Abstract: A formaldehyde electrochemical sensor (100) employing a formaldehyde sensitive assembly (120) of formaldehyde dehydrogenase (126) attached to graphene (122) in fluid communication with a source of NAD⁺ (140), and a method of measuring formaldehyde (F) utilizing the sensor (100).

FORMALDEHYDE GRAPHENE SENSOR

BACKGROUND

[0001] Formaldehyde is a naturally occurring volatile organic compound with the formula CH_2O . Formaldehyde is an important precursor in the production of many other materials and chemical compounds, such as particle board and various coatings.

[0002] Formaldehyde is toxic and recognized as a carcinogen. The United States National Institute for Occupational Safety and Health (NIOSH) considers 20 ppm formaldehyde in the atmosphere to be immediately dangerous to life and health (IDLH). The United States Occupational Safety and Health Administration has established a permissible exposure limit (PEL) for formaldehyde in the workplace at 0.75 parts formaldehyde per million parts of air (0.75 ppm) measured as an 8-hour time-weighted average (TWA). Other agencies and organizations have recommended that the acceptable level be reduced to less than about 0.10 ppm (100 ppb).

[0003] An excellent review of the various formaldehyde gas sensors developed over the years is provided in Chung, P.; Tzeng, C.; Ke, M.; Lee, C., Formaldehyde Gas Sensors: A Review, *Sensors*, 2013, 13, 4468-4484. Such sensors, while generally effective for measuring formaldehyde concentrations in air, tend to lack the necessary sensitivity and/or are far too expensive for widespread use. Accordingly, a substantial need exists for an economical yet robust and highly sensitive formaldehyde sensor capable of providing a quick response.

SUMMARY OF THE INVENTION

[0004] A first aspect of the invention is a formaldehyde sensitive assembly suitable for use in the manufacture of a formaldehyde electrochemical sensor. The assembly includes formaldehyde dehydrogenase attached to graphene.

[0005] A second aspect of the invention is a formaldehyde electrochemical sensor. The sensor has a formaldehyde interactive material located between and in electrical communication with a working electrode and a counter electrode. The formaldehyde interactive material includes a layer of graphene at least partially coated with immobilized formaldehyde dehydrogenase which is in fluid communication with a source of nicotinamide adenine dinucleotide.

[0006] The formaldehyde electrochemical sensor is preferably supported upon a structural substrate, and preferably also includes measurement circuitry in electrical communication with the working and counter electrodes operable for detecting the presence of any formaldehyde in fluid communication with the formaldehyde interactive material and generating an electrical signal representative of the amount of detected formaldehyde.

[0007] A third aspect of the invention is a method of measuring formaldehyde concentration within a sample. A first embodiment of the third aspect includes at least the steps of (i) obtaining a formaldehyde electrochemical sensor in accordance with the second aspect of the invention, (ii) placing the sensor into sensible fluid communication with the sample, and (iii) ascertaining a formaldehyde concentration within the sample by detecting the presence of any formaldehyde in the sample with the formaldehyde electrochemical sensor, generating an electrical signal representative of the amount of formaldehyde detected in the sample, and converting the electrical signal to a formaldehyde concentration based upon a known conversion algorithm.

[0008] A second embodiment of the third aspect includes at least the steps of (i) applying a voltage to a working electrode of a formaldehyde electrochemical sensor, the formaldehyde electrochemical sensor comprising at least (-) a working electrode and a counter electrode, (-) a formaldehyde interactive material comprising at least a layer of graphene at least partially coated with immobilized formaldehyde dehydrogenase, located between and in electrical communication with the working and counter electrodes, (-) a source of nicotinamide adenine dinucleotide in fluid communication with the formaldehyde interactive material, and (-) measurement circuitry in electrical communication with the working and counter electrodes operable for detecting the presence of any formaldehyde in fluid communication with the formaldehyde interactive material and generating an electrical signal representative of the amount of detected formaldehyde, (ii) measuring an electrical signal generated by the electrodes with the formaldehyde interactive material in sensing communication with the sample, and (iii) determining a formaldehyde concentration within the sample by converting the electrical signal to a formaldehyde concentration based upon a known conversion algorithm.

BRIEF DESCRIPTION OF THE DRAWINGS

[0009] Figure 1 is a schematic top-view of one embodiment of a formaldehyde electrochemical sensor in accordance with the second aspect of the invention.

[0010] Figure 2 is an enlarged schematic cross-section side-view of the formaldehyde interactive component of the formaldehyde electrochemical sensor depicted in Figure 1 taken along line 2-2, and further depicting a supply of NAD^+ in fluid communication with the formaldehyde interactive component.

[0011] Figures 3 is a schematic representations of a formaldehyde electrochemical sensor in accordance with the second aspect of the invention connected to measurement circuitry.

[0012] Figures 4A-D are schematic representations of an exemplary method of fabricating a formaldehyde electrochemical sensor in accordance with the second aspect of the invention.

[0013] Figure 5 is a grossly enlarged schematic representation of an exemplary formaldehyde sensitive assembly in accordance with the first aspect of the invention, secured to a structural substrate by alternating layers of positively charged polydiallyldimethylammonium chloride (PDDA) and negatively charged poly(sodium styrene sulfonate) (PSS).

[0014] Figure 6, is a graph depicting changes in resistance over time measured in Experiment Two.

[0015] Figures 6A, 6B and 6C are enlarged portions of the graph depicted in Figure 6 encompassing each measureable shift in resistance.

[0016] Figures 7A and 7B are graphs depicting changes in resistance over time measured in Experiment Three for gaseous test samples containing 1 and 2 ppm formaldehyde respectively.

[0017] Figure 7C is a graph of shift in resistance v. formaldehyde concentration for the two test samples measured in Experiment Three, from which an algorithm for converting the electrical signal generated by the sensor to a formaldehyde concentration can be derived.

DETAILED DESCRIPTION OF A PREFERRED EMBODIMENT

Definitions

Nomenclature

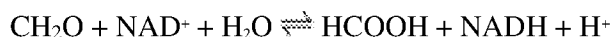
REFERENCE NO.	NAME
100	Formaldehyde Electrochemical Sensor

110	Structural Substrate / Wafer
120	Formaldehyde Sensitive Assembly / Formaldehyde Interactive Material
122	Graphene
124	Polymeric Electrolyte Linking Agent
126	Formaldehyde Dehydrogenase (FALDH)
130	Dielectric Material
140	Source of Nicotinamide Adenine Dinucleotide (NDA ⁺)
150	Electrodes
150'	Electrical Leads
151	Working Electrode
151'	Electrical Lead for Working Electrode
152	Counter Electrode
152'	Electrical Lead for Counter Electrode
200	Measurement Circuitry
210	Computer
F	Formaldehyde

Construction

[0018] Referring to Figures 1 and 2, a first aspect of the invention is a formaldehyde sensitive assembly or formaldehyde interactive material **120**, suitable for use in the manufacture of a formaldehyde electrochemical sensor **100**. The assembly **120** includes a formaldehyde dehydrogenase (FALDH) **126** attached to graphene **122**.

[0019] Formaldehyde dehydrogenase (FALDH) is an enzyme that catalyzes the chemical reaction



[0020] FALDH is commercially available from a number of sources, including specifically but not exclusively, Cayman Chemical of Ann Arbor, Michigan, Sigma-Aldrich of St. Louis, Missouri, and Fisher Scientific of Hampton, New Hampshire.

[0021] The use of FALDH in combination with a source of NAD⁺ in the construction of electrochemical formaldehyde biosensors is known. *See*, Marzuki, N.; Bakar, F.; Salleh, A.; Heng, L.; Yusof, N.; Siddiquee, S., Development of Electrochemical Biosensor for Formaldehyde Determination Based on Immobilized Enzyme, *Int. J. Electrochem. Sci.*, 2012, 7, 6070-6083, and Wang, M.; Jiang, S.; Chen, Y.; Chen, X.; Zhao, L.; Zhang, J.; Xu, J,

Formaldehyde Biosensor with Formaldehyde Dehydrogenase Adsorbed on Carbon Electrode Modified with Polypyrrole and Carbon Nanotube, *Engineering*, 2012, 5, 135-138. Both publications are incorporated herein by reference.

[0022] Graphene is fundamentally a single layer of sp² bonded carbon atoms arranged in a honeycomb (hexagonal) lattice. Graphene is the strongest material ever recorded, more than three hundred times stronger than structural steel and more than forty times stronger than diamond. Graphene provides superior levels of electronic conduction due to its single layer structure and the occurrence of a free pi (π) electron for each carbon atom in the lattice.

[0023] There are a number of ways to produce graphene, including specifically but not exclusively, mechanical exfoliation (the adhesive tape technique), chemical vapor deposition (CVD), thermolytic growth from a solid carbon source, sonication, carbon dioxide reduction, and graphite oxide reduction. Chemical vapor deposition is the generally preferred technique.

[0024] The FALDH **126** is immobilized upon the graphene **122**, preferably via a linking agent **124**. The linking agent **124** is preferably a polymeric electrolyte linking agent, such as polydiallyldimethylammonium chloride (PDDA), for facilitating transport of the electron signal resulting from the chemical reaction catalyzed by FALDH **126** to the graphene **122**. Other linking agents may also be employed, such as poly(allylamine hydrochloride), but we have found positively charged PDDA, self-assembled layer-by-layer (LbL) in alternating layers with negatively charged poly(sodium styrene sulfonate) (PSS) to provide an acceptable combination of retention strength and electron transport. We found poly-L-Lysine to be effective for immobilizing FALDH onto graphene, but ineffective for purposes of the invention as the resultant assembly was generally nonresponsive to formaldehyde.

[0025] Referring to Figures 1 and 3, a second aspect of the invention is a formaldehyde electrochemical sensor **100** employing the formaldehyde sensitive assembly **120** of the first aspect of the invention. The formaldehyde sensitive assembly **120** is positioned between and in electrical communication with a working electrode **151** and a counter electrode **152** (collectively referenced as electrodes **150**). The electrodes **150** are preferably Au/Cr electrodes. The formaldehyde sensitive assembly **120** is in fluid communication with a source of the coenzyme nicotinamide adenine dinucleotide (NAD⁺) **140**, as a necessary reactant in the FALDH catalyzed chemical reaction



[0026] The source of NAD⁺ is preferably an aqueous phosphate buffered saline solution of β -NAD⁺.

[0027] The sensor **100** preferably includes a structural substrate **110** to support the formaldehyde sensitive assembly **120** and electrodes **150**, such as a wafer used in fabrication of the sensor **100**.

[0028] Referring to Figures 1 and 2, a dielectric material **130**, such as KMPR, is used as necessary to electrically insulate and isolate various components of the sensor **100** from one another.

[0029] The sensor **100** can be sized as desired for each particular application, with a general preference for a sensor **100** between about 10 to 20 mm wide, 10 to 20 mm long and a height as minimally necessary to house the required supply of NAD⁺ atop the formaldehyde sensitive assembly **120**, typically 5 to 20 mm.

[0030] Referring to Figure 3, the electrodes **150** can be placed into electrical communication with standard measurement circuitry **200** via electrical leads **151'** and **152'** for detecting and measuring any electrical signal generated by the oxidation of any formaldehyde **F** in fluid communication with the formaldehyde interactive material **120**, and generating an electrical signal representative of the amount of detected formaldehyde **F**. An exemplary measurement circuitry **200** suitable for use in the present invention is the 34970A data acquisition resistance analyzer available from Agilent Technologies, Inc. of Santa Clara, California. The generated electrical signal is then converted, typically via a software program running on a general purpose computer **210** in communication with the analyzer, to a formaldehyde concentration based upon an empirically derived conversion algorithm.

[0031] Referring to Figures 4A-D, a preferred method of fabricating the formaldehyde sensitive assembly **120** involves the steps of photolithographic formation of working **151** and counter **152** electrodes (collectively electrodes **150**) (*e.g.*, Cr/50nm + Au/100nm) on a structural substrate **110** (*e.g.*, 300nm SiO₂), (-) photolithographic formation of an LbL self-assembled layer of graphene **122** between and in electrical communication with both of the electrodes **150** (*e.g.*, [PDDA (10 min) + PSS (10 min)]₂ + [PDDA (10 min) + graphene (20 min)]₅, (-) photolithographic formation of a dielectric coating (*e.g.*, KMPR) as necessary and appropriate to prevent short circuiting between the electrodes **150** while leaving an area of the graphene layer **122** exposed, and (-) formation of an LbL layer of immobilized enzyme on the exposed area of the graphene layer **122** (*e.g.*, [PDDA (10 min) + FALDH (20 min)]₂).

Referring to Figure 2, construction of the electrochemical sensor **100** is completed by placing a supply of NAD^+ in fluid communication with the enzyme coated exposed area of the graphene layer **122** (e.g., a reservoir containing an aqueous phosphate buffered saline solution of $\beta\text{-NAD}^+$).

Experimental

EXPERIMENT ONE

Sensor Fabrication

[0032] Thirty sensors, each approximately 10 mm by 13 mm and having the layout set forth in Figure 1, were formed on a single 4 in diameter 300 nm thick SiO_2 wafer.

[0033] A 50 nm layer of Chromium and a 100 nm layer of Gold (Cr/Au) were sputter coated onto a major surface of the wafer and patterned by photolithography using Microposit S1813 as the photoresist and wet chemical etching, to form the electrodes **150** and electrical leads **150'** as depicted in Figure 1, for each of the thirty sensors.

[0034] A layer of LbL self-assembled graphene was then formed over the wafer [PDDA (10 min) + PSS (10 min)]₂ + [PDDA (10 min) + graphene (20 min)]₅ and patterned by photolithography using Microposit S1813 as the photoresist and oxygen plasma etching, to form the graphene layer **122** of the formaldehyde sensitive assembly **120** as depicted in Figure 1, for each of the thirty sensors.

[0035] A coating of KMPR was then placed over the wafer and patterned by photolithography using wet chemical etching, to form the dielectric protective coating **130** as depicted in Figure 1 (noting removal of dielectric coating over electrodes to facilitate viewing of internal components) and Figure 2, for each of the thirty sensors.

[0036] An incomplete layer of formaldehyde dehydrogenase (FALDH) was then immobilized onto the exposed area of the graphene layer **122** using polydiallyldimethylammonium chloride (PDDA) as the linking agent ([PDDA (10 min) + FALDH (20 min)]₂).

[0037] Each sensor was then stamped out from the wafer.

EXPERIMENT TWO

Sensing of Formaldehyde in Aqueous Solution

[0038] A silicone o-ring was sealing secured with silicone rubber around the formaldehyde sensitive assembly of a sensor constructed in Experiment One so as to form a reservoir directly over the formaldehyde sensitive assembly of the sensor. A 20 μ l volume of phosphate buffered saline solution of β -NAD⁺ at pH 8.0, N7004 purchased from Sigma-Aldrich, was placed into the reservoir into direct fluid communication with the formaldehyde sensitive assembly.

[0039] A 34970A data acquisition resistance analyzer purchased from Agilent Technologies, Inc., connected to a general purpose computer, was electrically connected to the electrical leads on the sensor for detecting and measuring shifts in resistance [$k\Omega$] at the sensor.

[0040] The phosphate buffered saline solution of β -NAD⁺ in the reservoir was sequentially injected with 20 μ l of 0.006 mM, 0.06 mM, 0.6 mM and 6 mM solutions of formaldehyde over time as set forth in Figure 6 to form samples containing the equivalent of a gas phase concentration of 0.9 ppb, 5.9 ppb, 38.5 ppb and 264.5 ppb formaldehyde based upon the transformation algorithms established by S. Dong and P. K. Dasgupta, *Environmental Science and Technology* 20 (1986), 637-640) set forth below, and shifts in resistance measured over time as displayed in Figure 6.

$$[\text{CH}_2\text{O} (aq)] = 16650 [\text{CH}_2\text{O} (g)]^{1.0789}$$

$$\log [\text{CH}_2\text{O} (g)] = ((0.9261)(\log(x_{\text{CH}_2\text{O} (aq)}) - 2.2942) \pm 0.0332$$

[0041] The sensor was capable of detecting formaldehyde as low as a converted concentration of 5.9 ppb as shown in Figure 6, with a T(90) response time (*i.e.*, the time to reach 90% of the final value) averaging 31.5 sec as depicted in Figures 6A, 6B and 6C.

EXPERIMENT THREE

Sensing of Formaldehyde in Gas

[0042] A silicone o-ring was sealing secured with silicone rubber around the formaldehyde sensitive assembly of a sensor constructed in Experiment One so as to form a reservoir directly over the formaldehyde sensitive assembly of the sensor. An aqueous phosphate buffered saline solution of β -NAD⁺, N7004 purchased from Sigma-Aldrich, was

placed into the reservoir into direct fluid communication with the formaldehyde sensitive assembly.

[0043] A 34970A data acquisition resistance analyzer purchased from Agilent Technologies, Inc., connected to a general purpose computer, was electrically connected to the electrical leads on the sensor for detecting and measuring shifts in resistance [$k\Omega$] at the sensor.

[0044] The reservoir on the sensor was covered with a polytetrafluorethylene (PTFE) membrane to reduce evaporation of the aqueous phosphate buffered saline solution of β -NAD⁺ retained within the reservoir. Carrier gas samples of N₂ heated to approximately 60°C and containing 1 ppm and 2 ppm formaldehyde, formed using a model 8990 Permeation Calibrator from Mocon, Inc. of Minneapolis, Minnesota and a formaldehyde containing Trace Source™ Permeation Tube from Kin-Tek Laboratories, Inc. of La Marque, Texas, were separately passed over the PTFE membrane at a flow rate of approximately 760 cm³/min for 60 seconds. Shifts in resistance measured over time for each sample as the sample was passed over the PTFE membrane are displayed in Figures 7A and 7B.

[0045] The sensor detected a 12 ohm shift in resistance with a T(95) of 46 seconds at a formaldehyde concentration of 1 ppm in the gas sample, and a 25 ohm shift in resistance with a T(95) of 48 seconds at a formaldehyde concentration of 2 ppm in the gas sample. Figure 7C is a graph of shift in resistance v. formaldehyde concentration for these two test samples, from which an algorithm for converting the electrical signal generated by the sensor to a formaldehyde concentration can be derived.

We claim:

1. A formaldehyde sensitive assembly suitable for use in the manufacture of a formaldehyde electrochemical sensor, comprising formaldehyde dehydrogenase attached to graphene.
2. The formaldehyde sensitive assembly of claim 1 wherein the formaldehyde dehydrogenase is attached to the graphene by a polymeric electrolyte linking agent.
3. The formaldehyde sensitive assembly of claim 2 wherein the polymeric electrolyte linking agent is polydiallyldimethylammonium chloride.
4. A formaldehyde electrochemical sensor having a formaldehyde interactive material located between and in electrical communication with a working electrode and a counter electrode, the formaldehyde interactive material comprising a layer of graphene at least partially coated with immobilized formaldehyde dehydrogenase which is in fluid communication with a source of nicotinamide adenine dinucleotide.
5. The formaldehyde electrochemical sensor of claim 4 further comprising measurement circuitry in electrical communication with the working and counter electrodes operable for detecting the presence of any formaldehyde in fluid communication with the formaldehyde interactive material and generating an electrical signal representative of the amount of detected formaldehyde.
6. The formaldehyde electrochemical sensor of claim 5 wherein the measured electrical property is a shift in resistance.
7. The formaldehyde electrochemical sensor of claim 4 wherein the sensor is supported upon a major surface of a structural substrate.
8. The formaldehyde electrochemical sensor of claim 7 wherein the structural substrate is a wafer.

9. The formaldehyde electrochemical sensor of claim 4 wherein the formaldehyde dehydrogenase is immobilized upon the layer of graphene by a polymeric electrolyte linking agent.
10. The formaldehyde electrochemical sensor of claim 9 wherein the polymeric electrolyte linking agent is polydiallyldimethylammonium chloride.
11. The formaldehyde electrochemical sensor of claim 4 wherein the source of nicotinamide adenine dinucleotide is an aqueous buffered solution of β -NAD⁺.
12. The formaldehyde electrochemical sensor of claim 11 wherein the aqueous buffered solution of nicotinamide adenine dinucleotide is a phosphate buffered saline solution of nicotinamide adenine dinucleotide.
13. The formaldehyde electrochemical sensor of claim 4 wherein the graphene layer is layer-by-layer self-assembled on a wafer.
14. The formaldehyde electrochemical sensor of claim 9 wherein the polymeric electrolyte linking agent and the formaldehyde dehydrogenase are assembled layer-by-layer on the layer of graphene.
15. A method of measuring formaldehyde concentration within a sample, comprising the steps of:
 - (a) obtaining a formaldehyde electrochemical sensor in accordance with claim 5,
 - (b) placing the formaldehyde electrochemical sensor into sensible fluid communication with the sample, and
 - (c) ascertaining a formaldehyde concentration within the sample by detecting the presence of any formaldehyde in the sample with the formaldehyde electrochemical sensor, generating an electrical signal representative of the amount of formaldehyde detected in the sample, and converting the electrical

signal to a formaldehyde concentration based upon a known conversion algorithm.

16. The method of claim 15 wherein the sample is a gaseous sample.
17. A method of measuring formaldehyde concentration within a sample, comprising the steps of:
 - (a) applying a voltage to a working electrode of a formaldehyde electrochemical sensor, the formaldehyde electrochemical sensor comprising at least:
 - (i) a working electrode and a counter electrode,
 - (ii) a formaldehyde interactive material comprising at least a layer of graphene at least partially coated with immobilized formaldehyde dehydrogenase, located between and in electrical communication with the working and counter electrodes,
 - (iii) a source of nicotinamide adenine dinucleotide in fluid communication with the formaldehyde interactive material, and
 - (iv) measurement circuitry in electrical communication with the working and counter electrodes operable for detecting the presence of any formaldehyde in fluid communication with the formaldehyde interactive material and generating an electrical signal representative of the amount of detected formaldehyde,
 - (b) measuring an electrical signal generated by the electrodes with the formaldehyde interactive material in sensing communication with the sample, and
 - (c) determining a formaldehyde concentration within the sample by converting the electrical signal to a formaldehyde concentration based upon a known conversion algorithm.
18. The method of claim 17 wherein the sample is a gaseous sample.

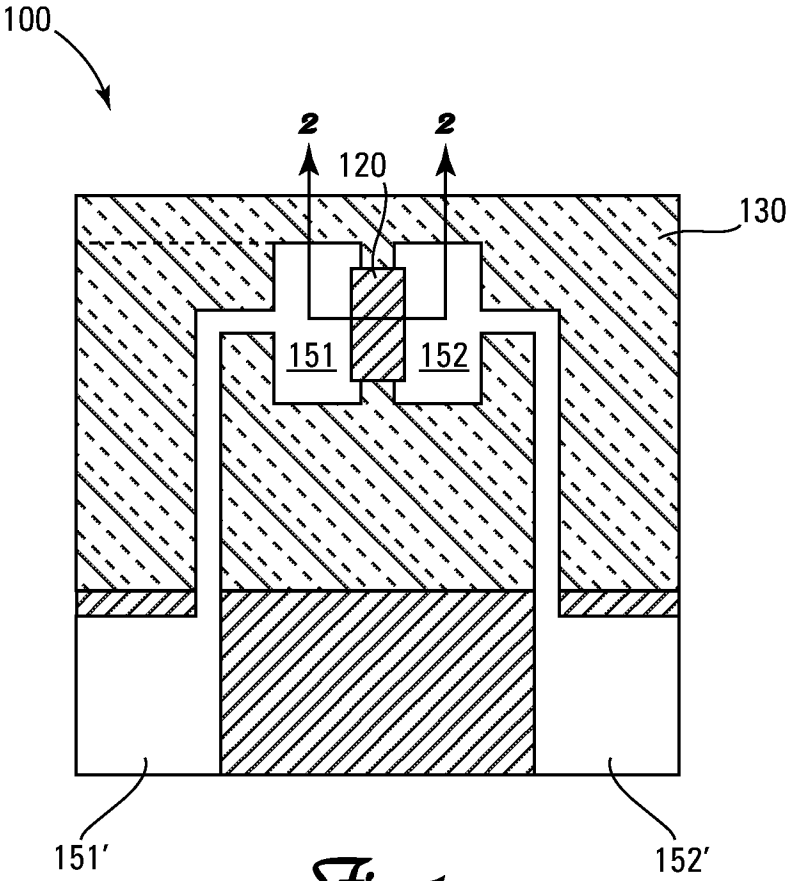


Fig. 1

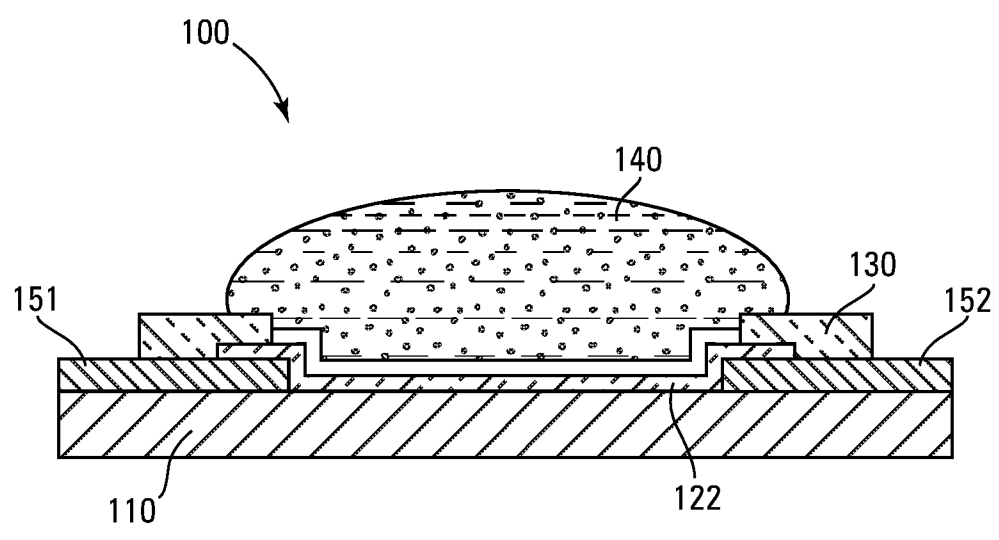


Fig. 2

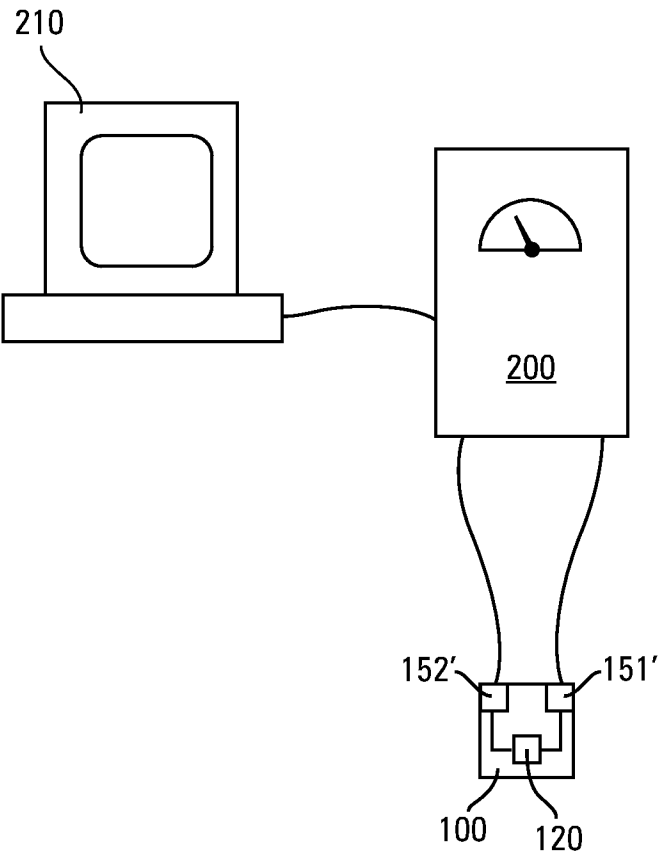


Fig. 3

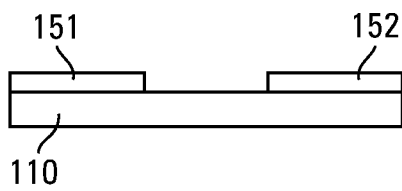


Fig. 4A

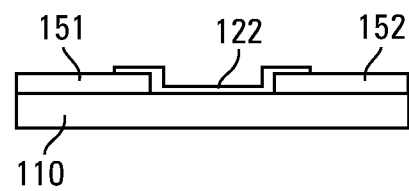


Fig. 4B

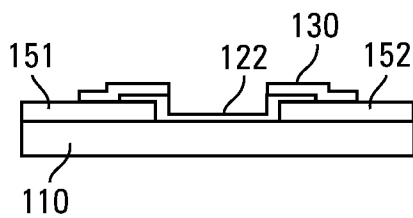


Fig. 4C

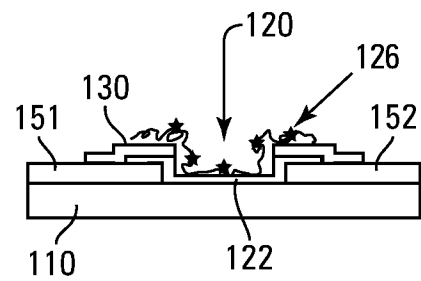


Fig. 4D

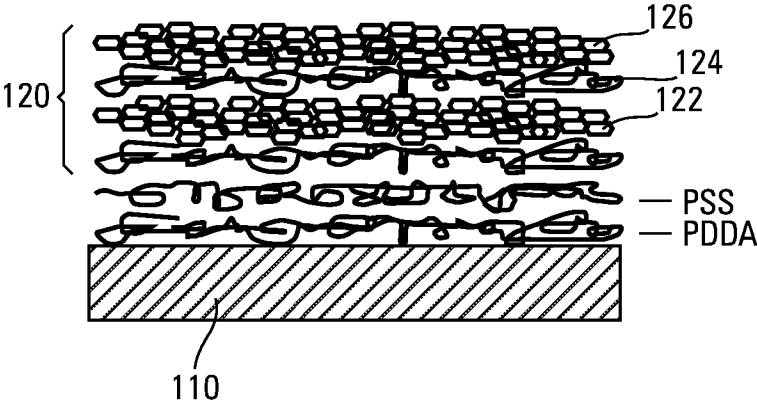


Fig. 5

Fig. 6

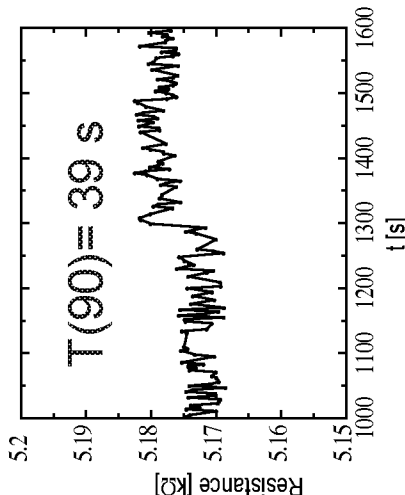
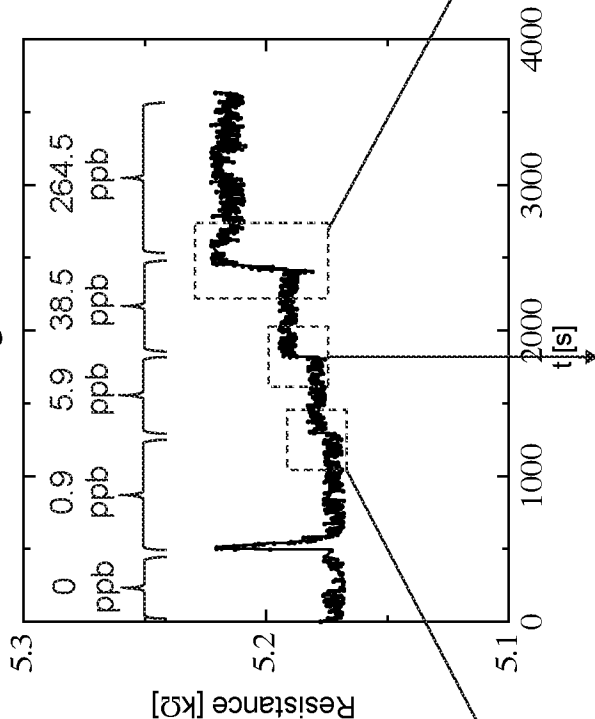


Fig. 6A

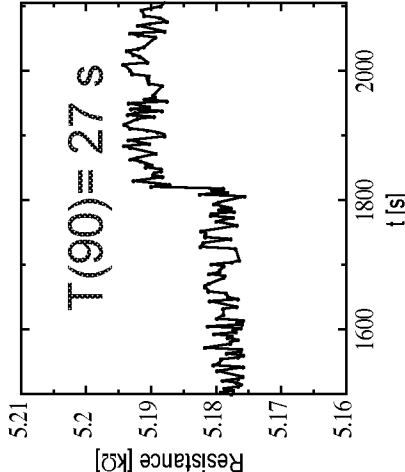


Fig. 6B

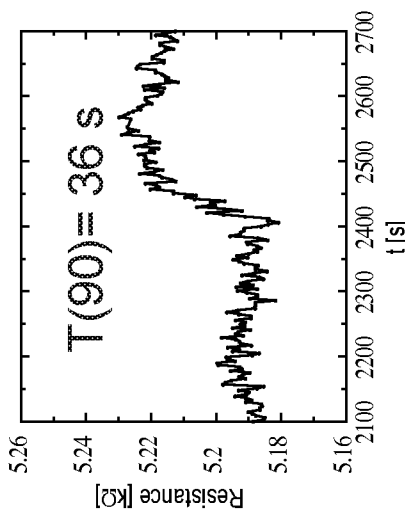
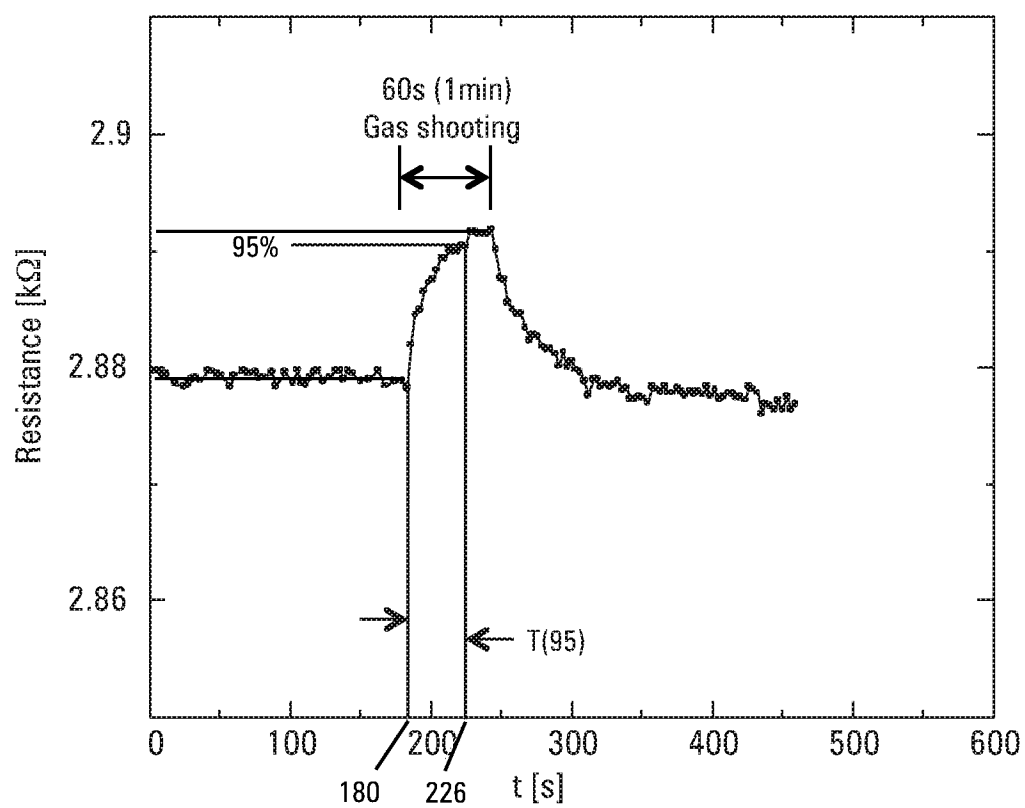
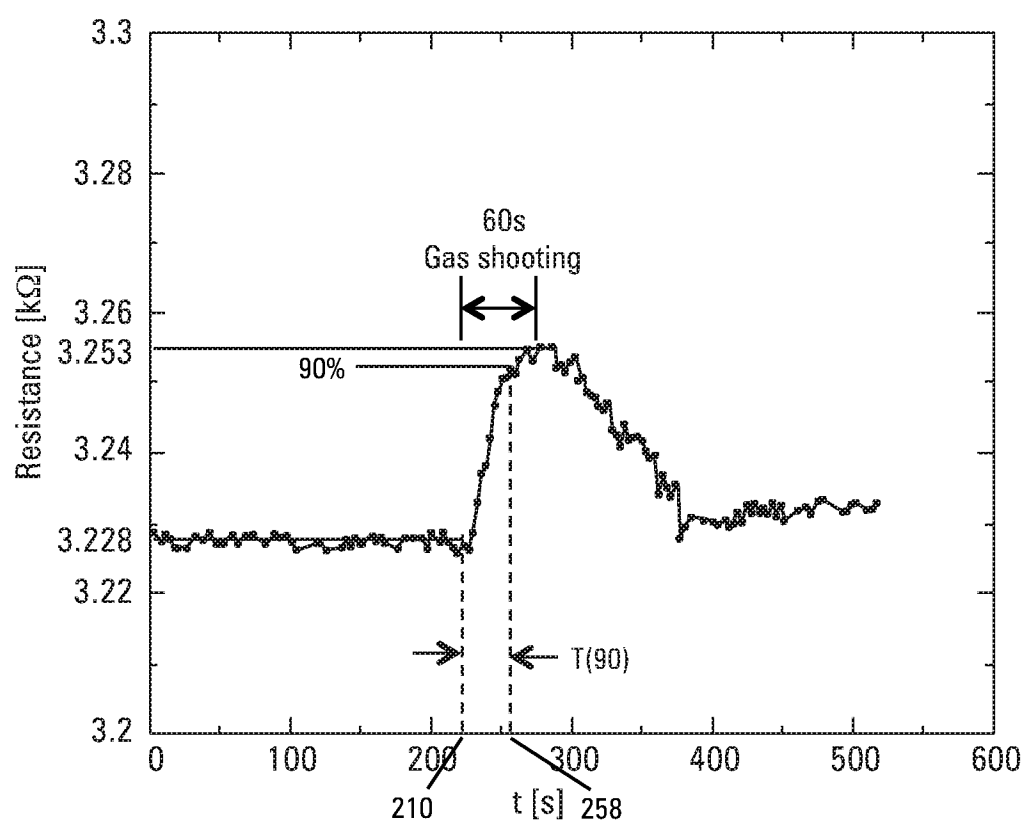


Fig. 6C

*Fig. 7A*

*Fig. 7B*

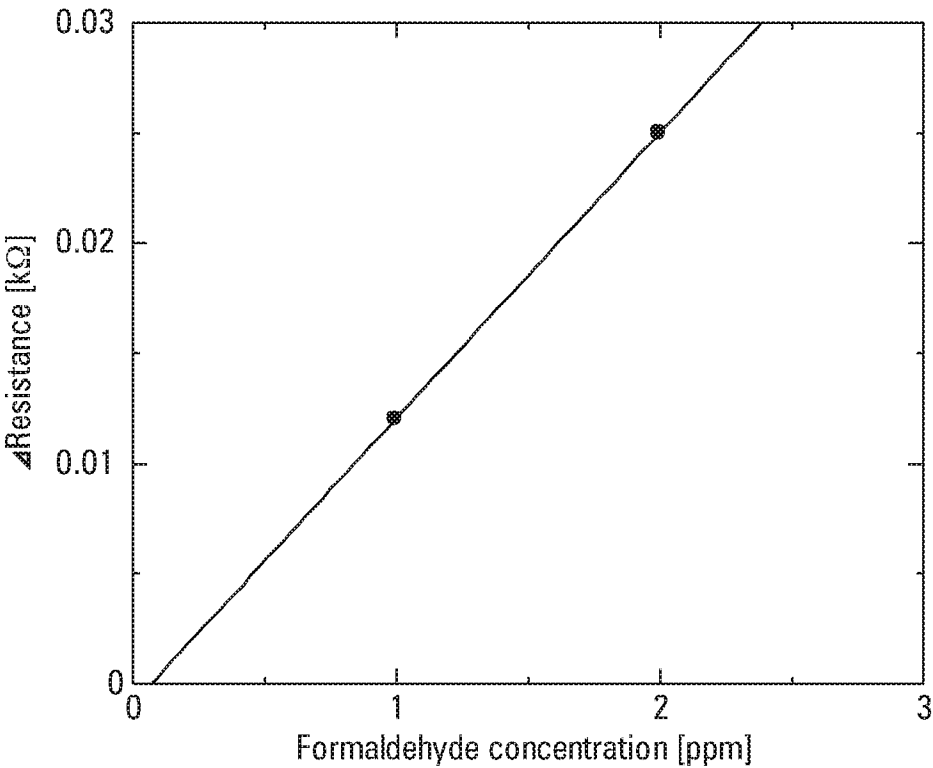


Fig. 7C

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2016/049367

A. CLASSIFICATION OF SUBJECT MATTER

IPC(8) - C12N 11/14; C12Q 1/32; G01N 27/327 (2016.01)

CPC - C12N 11/14; C12Q 1/003; C12Q 1/005; C12Q 1/32; G01N 27/308; G01N 27/327 (2016.08)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC(8) - C12N 11/14; C12Q 1/32; G01N 27/327 (2016.01)

CPC - C12N 11/14; C12Q 1/003; C12Q 1/005; C12Q 1/32; G01N 27/308; G01N 27/327 (2016.08)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

USPC - 204/403.14; 204/403.15; 205/786; 205/787; IPC(8) - C12N 11/14; C12Q 1/32; G01N 27/327; CPC - C12N 11/14; C12Q 1/003; C12Q 1/005; C12Q 1/32; G01N 27/308; G01N 27/327 (keyword delimited)

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Orbit, Google Patents, Google

Search terms used: formaldehyde, sensor, dehydrogenase, graphene, carbon nanotube, CNT, graphite, polydiallyldimethylammonium chloride, electrochemical, β -NAD, beta, resistance, wafer, layer by layer, lbl,

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2009/0008248 A1 (SHIMOMURA et al) 08 January 2009 (08.01.2009) entire document	1-13, 15-18
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Y		14
Y	WANG et al., Formaldehyde Biosensor with Formaldehyde Dehydrogenase Adsorped on Carbon Electrode Modified with Polypyrrole and Carbon Nanotube, Engineering, Vol. 5, October 2012, [retrieved on 19 October 2016]. Retrieved from the Internet: <URL: http://file.scirp.org/pdf/ENG_2013102114222979.pdf >, Pgs.135-138	14

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Date of the actual completion of the international search

24 October 2016

Date of mailing of the international search report

14 NOV 2016

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