



(86) Date de dépôt PCT/PCT Filing Date: 1999/10/21
(87) Date publication PCT/PCT Publication Date: 2000/05/04
(45) Date de délivrance/Issue Date: 2010/08/03
(85) Entrée phase nationale/National Entry: 2001/04/09
(86) N° demande PCT/PCT Application No.: US 1999/024647
(87) N° publication PCT/PCT Publication No.: 2000/024904
(30) Priorités/Priorities: 1998/10/23 (US60/105,359);
1998/10/23 (US60/105,408)

(51) Cl.Int./Int.Cl. *C12N 15/32* (2006.01),
A01N 63/02 (2006.01), *C07K 14/325* (2006.01),
C12N 15/82 (2006.01), *A01H 5/00* (2006.01)
(72) Inventeurs/Inventors:
CARDINEAU, GUY A., US;
STELMAN, STEVEN J., US;
NARVA, KENNETH E., US
(73) Propriétaire/Owner:
MYCOGEN CORPORATION, US
(74) Agent: MACRAE & CO.

(54) Titre : POLYNUCLEOTIDES OPTIMISES POUR LES VEGETAUX CODANT POUR DES PROTEINES PESTICIDES
D'ENVIRON 15 kDA ET D'ENVIRON 45 kDA
(54) Title: PLANT-OPTIMIZED POLYNUCLEOTIDES ENCODING APPROXIMATELY 15 kDA AND APPROXIMATELY 45
kDA PESTICIDAL PROTEINS

(57) **Abrégé/Abstract:**

The subject invention provides new, plant-optimized polynucleotide sequences that encode pesticidal toxins. The polynucleotide sequences of the subject invention have certain modifications, compared to wild-type sequences, for example, that make them particularly well-suited for optimized expression in plants. Using the polynucleotide sequences described herein, the transformation of plants can be accomplished, using techniques known to those skilled in the art, in order to confer pest resistance upon said plants. In preferred embodiments, the subject invention provides plant-optimized polynucleotide sequences which encode approximately 15 kDa and approximately 45 kDa pesticidal proteins.



PCTWORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁷ : C12N 15/32, 15/82, A01N 63/02	A1	(11) International Publication Number: WO 00/24904 (43) International Publication Date: 4 May 2000 (04.05.00)
(21) International Application Number: PCT/US99/24647 (22) International Filing Date: 21 October 1999 (21.10.99) (30) Priority Data: 60/105,408 23 October 1998 (23.10.98) US 60/105,359 23 October 1998 (23.10.98) US (71) Applicant: MYCOGEN CORPORATION [US/US]; 5501 Oberlin Drive, San Diego, CA 92121 (US). (72) Inventors: CARDINEAU, Guy, A.; 17041 Cloudcroft Drive, Poway, CA 92064 (US). STELMAN, Steven, J.; 7954 Westbury Avenue, San Diego, CA 92126 (US). NARVA, Kenneth, E.; 12123 Caminito Mira Del Mar, San Diego, CA 92130 (US). (74) Agents: SANDERS, Jay, M. et al.; Saliwanchik, Lloyd & Saliwanchik, Suite A-1, 2421 N.W. 41st Street, Gainesville, FL 32606-6669 (US).		(81) Designated States: AE, AL, AU, BA, BB, BG, BR, CA, CN, CR, CU, CZ, DM, EE, GD, GE, HR, HU, ID, IL, IN, IS, JP, KP, KR, LC, LK, LR, LT, LV, MA, MG, MK, MN, MX, NO, NZ, PL, RO, SG, SI, SK, SL, TR, TT, TZ, UA, UZ, VN, YU, ZA, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>With international search report.</i> <i>Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
(54) Title: PLANT-OPTIMIZED POLYNUCLEOTIDES ENCODING APPROXIMATELY 15 kDa AND APPROXIMATELY 45 kDa PESTICIDAL PROTEINS (57) Abstract <p>The subject invention provides new, plant-optimized polynucleotide sequences that encode pesticidal toxins. The polynucleotide sequences of the subject invention have certain modifications, compared to wild-type sequences, for example, that make them particularly well-suited for optimized expression in plants. Using the polynucleotide sequences described herein, the transformation of plants can be accomplished, using techniques known to those skilled in the art, in order to confer pest resistance upon said plants. In preferred embodiments, the subject invention provides plant-optimized polynucleotide sequences which encode approximately 15 kDa and approximately 45 kDa pesticidal proteins.</p>		

DESCRIPTION

PLANT-OPTIMIZED POLYNUCLEOTIDES ENCODING
APPROXIMATELY 15 kDa AND APPROXIMATELY 45 kDa
PESTICIDAL PROTEINS

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Background of the Invention

Insects and other pests cost farmers billions of dollars annually in crop losses and in the expense of keeping these pests under control. The losses caused by insect pests in agricultural production environments include decrease in crop yield, reduced crop quality, and increased harvesting costs.

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Chemical pesticides have provided an effective method of pest control; however, the public has become concerned about the amount of residual chemicals which might be found in food, ground water, and the environment. Therefore, synthetic chemical pesticides are being increasingly scrutinized, and correctly so, for their potential toxic environmental consequences. Synthetic chemical pesticides can poison the soil and underlying aquifers, pollute surface waters as a result of runoff, and destroy non-target life forms. Synthetic chemical control agents have the further disadvantage of presenting public safety hazards when they are applied in areas where pets, farm animals, or children may come into contact with them. They may also provide health hazards to applicants, especially if the proper application techniques are not followed. Regulatory agencies around the world are restricting and/or banning the uses of many pesticides and particularly the synthetic chemical pesticides which are persistent in the environment and enter the food chain. Examples of widely used synthetic chemical pesticides include the organochlorines, *e.g.*, DDT, mirex, kepone, lindane, aldrin, chlordane, aldicarb, and dieldrin; the organophosphates, *e.g.*, chlorpyrifos, parathion, malathion, and diazinon; and carbamates. Stringent new restrictions on the use of pesticides and the elimination of some effective pesticides from the market place could limit economical and effective options for controlling costly pests.

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Because of the problems associated with the use of synthetic chemical pesticides, there exists a clear need to limit the use of these agents and a need to identify alternative control agents. The replacement of synthetic chemical pesticides, or combination of

these agents with biological pesticides, could reduce the levels of toxic chemicals in the environment.

5 A biological pesticidal agent that is being used with increasing popularity is the soil microbe *Bacillus thuringiensis* (*B.t.*). The soil microbe *Bacillus thuringiensis* (*B.t.*) is a Gram-positive, spore-forming bacterium. Most strains of *B.t.* do not exhibit pesticidal activity. Some *B.t.* strains produce, and can be characterized by, parasporal crystalline protein inclusions. These “ δ -endotoxins,” which typically have specific pesticidal activity, are different from exotoxins, which have a non-specific host range. These inclusions often appear microscopically as distinctively shaped crystals. The proteins can be highly toxic to pests and are specific in their toxic activity.

10 Preparations of the spores and crystals of *B. thuringiensis* subsp. *kurstaki* have been used for many years as commercial insecticides for lepidopteran pests. For example, *B. thuringiensis* var. *kurstaki* HD-1 produces a crystalline δ -endotoxin which is toxic to the larvae of a number of lepidopteran insects.

15 The cloning and expression of a *B.t.* crystal protein gene in *Escherichia coli* was described in the published literature more than 15 years ago (Schnepf, H.E., H.R. Whiteley [1981] *Proc. Natl. Acad. Sci. USA* 78:2893-2897.). U.S. Patent No. 4,448,885 and U.S. Patent No. 4,467,036 both disclose the expression of *B.t.* crystal protein in *E. coli*. Recombinant DNA-based *B.t.* products have been produced and approved for use.

20 Commercial use of *B.t.* pesticides was originally restricted to a narrow range of lepidopteran (caterpillar) pests. More recently, however, investigators have discovered *B.t.* pesticides with specificities for a much broader range of pests. For example, other species of *B.t.*, namely *israelensis* and *morrisoni* (a.k.a. *tenebrionis*, a.k.a. *B.t.* M-7), have been used commercially to control insects of the orders Diptera and Coleoptera, respectively (Gaertner, F.H. [1989] “Cellular Delivery Systems for Insecticidal Proteins: Living and Non-Living Microorganisms,” in *Controlled Delivery of Crop Protection Agents*, R.M. Wilkins, ed., Taylor and Francis, New York and London, 1990, pp. 245-255).

25 New subspecies of *B.t.* have now been identified, and genes responsible for active δ -endotoxin proteins have been isolated and sequenced (Höfte, H., H.R. Whiteley [1989] *Microbiological Reviews* 52(2):242-255). Höfte and Whiteley classified *B.t.* crystal protein genes into four major classes. The classes were *cryI* (Lepidoptera-specific), *cryII*

(Lepidoptera- and Diptera-specific), *cryIII* (Coleoptera-specific), and *cryIV* (Diptera-specific). The discovery of strains specifically toxic to other pests has been reported (Feitelson, J.S., J. Payne, L. Kim [1992] *Bio/Technology* 10:271-275). For example, the designations CryV and CryVI have been proposed for two new groups of nematode-active toxins.

The 1989 nomenclature and classification scheme of Höfte and Whiteley was based on both the deduced amino acid sequence and the host range of the toxin. That system was adapted to cover 14 different types of toxin genes which were divided into five major classes. The number of sequenced *Bacillus thuringiensis* crystal protein genes currently stands at more than 50. A revised nomenclature scheme has been proposed which is based solely on amino acid identity (Crickmore *et al.* [1996] Society for Invertebrate Pathology, 29th Annual Meeting, IIIrd International Colloquium on *Bacillus thuringiensis*, University of Cordoba, Cordoba, Spain, September 1-6, 1996, abstract). The mnemonic "cry" has been retained for all of the toxin genes except cytA and cytB, which remain a separate class. Roman numerals have been exchanged for Arabic numerals in the primary rank, and the parentheses in the tertiary rank have been removed. Many of the original names have been retained, although a number have been reclassified.

With the use of genetic engineering techniques, new approaches for delivering *B.t.* toxins to agricultural environments are under development, including the use of plants genetically engineered with *B.t.* toxin genes for insect resistance and the use of stabilized, microbial cells as delivery vehicles of *B.t.* toxins (Gaertner, F.H., L. Kim [1988] *TIBTECH* 6:S4-S7). Thus, isolated *B.t.* endotoxin genes are becoming commercially valuable.

Various improvements have been achieved by modifying *B.t.* toxins and/or their genes. For example, U.S. Patent Nos. 5,380,831 and 5,567,862 relate to the production of synthetic insecticidal crystal protein genes having improved expression in plants.

Obstacles to the successful agricultural use of *B.t.* toxins include the development of resistance to *B.t.* toxins by insects. In addition, certain insects can be refractory to the effects of *B.t.* The latter includes insects such as boll weevil and black cutworm as well as adult insects of most species which heretofore have demonstrated no apparent significant sensitivity to *B.t.* δ -endotoxins.

Thus, resistance management strategies in *B.t.* plant technology have become of great interest, and there remains a great need for new toxin genes. For example, WO 97/40162 (published PCT application) discloses 15 kDa and 45 kDa coleopteran-active proteins obtainable from *B.t.* isolates PS80JJ1 and PS149B1.

5 As a result of extensive research and resource investment, patents continue to issue for new *B.t.* isolates, toxins, and genes, and for new uses of *B.t.* isolates. See Feitelson *et al.*, *supra*, for a review. U.S. Patent No. 5,589,382 discloses *B.t.* isolate PS80JJ1 as having activity against nematodes. U.S. Patent No. 5,632,987 discloses *B.t.* isolate PS80JJ1 as having activity against corn rootworm. However, the discovery of
10 new *B.t.* isolates and new uses of known *B.t.* isolates remains an empirical, unpredictable art.

There remains a great need for new toxin genes that can be successfully expressed at adequate levels in plants in a manner that will result in the effective control of insects and other pests.

15 Brief Summary of the Invention

The subject invention concerns materials and methods useful in the control of pests and, particularly, plant pests. More specifically, the subject invention provides new, plant-optimized polynucleotide sequences that encode pesticidal proteins. The polynucleotide sequences of the subject invention have certain modifications, compared
20 to wild-type sequences, that make them particularly well-suited for optimized expression in plants. Using the polynucleotide sequences described herein, the transformation of plants can be accomplished, using techniques known to those skilled in the art, in order to confer pest resistance upon said plants. In a preferred embodiment, the subject invention provides plant-optimized polynucleotide sequences which encode
25 approximately 15 kDa and approximately 45 kDa pesticidal proteins.

Description of the Sequences

SEQ ID NO. 1 is a polynucleotide sequence for a gene designated 80JJ1-15-PO5, which is optimized for expression in maize. This gene encodes an approximately 15 kDa protein. This gene and protein were disclosed in WO 97/40162.

30 **SEQ ID NO. 2** is a novel polynucleotide sequence for a gene designated 80JJ1-15-PO7, which is optimized for expression in maize. This is an alternative gene that encodes an approximately 15 kDa protein.

SEQ ID NO. 3 is an amino acid sequence for a novel pesticidally active protein encoded by the gene designated 80JJ1-15-PO7.

SEQ ID NO. 4 is a polynucleotide sequence for a gene designated 80JJ1-45-PO, which is optimized for expression in maize. This gene encodes an approximately 45 kDa protein. This gene was disclosed in WO 97/40162.

SEQ ID NO. 5 is a novel polynucleotide sequence for a gene designated 149B1-15-PO, which is optimized for expression in *Zea mays*. This gene encodes an approximately 15 kDa protein obtainable from PS149B1 that is disclosed in WO 97/40162.

SEQ ID NO. 6 is a novel polynucleotide sequence for a gene designated 149B1-45-PO, which is optimized for expression in *Zea mays*. This gene encodes an approximately 45 kDa protein obtainable from PS149B1 that is disclosed in WO 97/40162.

Detailed Disclosure of the Invention

The subject invention concerns materials and methods useful in the control of pests and, particularly, plant pests. More specifically, the subject invention provides new, plant-optimized polynucleotide sequences that encode pesticidal proteins. The polynucleotide sequences of the subject invention have certain modifications, compared to wild-type sequences, that make them particularly well-suited for optimized expression in plants. Using the polynucleotide sequences described herein, the transformation of plants can be accomplished, using techniques known to those skilled in the art, in order to confer pest resistance upon said plants. In a preferred embodiment, the subject invention provides plant-optimized polynucleotide sequences which encode approximately 15 kDa and approximately 45 kDa pesticidal proteins.

Using techniques such as computer- or software-assisted sequence alignments, differences can be noted in the nucleotide sequence of the subject plant-optimized genes as compared to the wild-type genes or to previously known genes. Similarly, differences in the unique amino acid sequences of the subject invention can be noted as compared to wild-type toxins or to previously known toxins.

It should be apparent to a person skilled in this art that, given the sequences of the genes as set forth herein, the genes of the subject invention can be obtained through several means. In preferred embodiments, the subject genes may be constructed

synthetically by using a gene synthesizer, for example. The specific genes exemplified herein can also be obtained by modifying, according to the teachings of the subject invention, certain wild-type genes (for example, by point-mutation techniques) from certain isolates deposited at a culture depository as discussed below.

Certain cultures discussed in this application have been deposited in the Agricultural Research Service Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois 61604, USA. The deposited strains listed below are disclosed in the patent references as discussed above in the section entitled "Background of the Invention."

<u>Subculture</u>	<u>Accession Number</u>	<u>Deposit Date</u>
<i>B.t.</i> PS80JJ1	NRRL B-18679	July 17, 1990
<i>E. coli</i> (NM522) (pMYC2421) (PS80JJ1 14 kDa & 45 kDa)	NRRL B-21555	March 28, 1996
<i>E. coli</i> (NM522) (pMYC2426) (PS80JJ1 14 kDa & 45 kDa)	NRRL B-21671	March 26, 1997
<i>B.t.</i> PS149B1	NRRL B-21553	March 28, 1996
<i>E. coli</i> (NM522) (pMYC2429) (PS149B1 15 kDa & 45 kDa)	NRRL B-21673	March 26, 1997

It should be understood that the availability of a deposit does not constitute a license to practice the subject invention in derogation of patent rights granted by governmental action.

Genes and toxins. The subject invention includes, in preferred embodiments, polynucleotide sequences optimized for expression in plants, wherein said sequences are selected from the group consisting of SEQ ID NO. 2, SEQ ID NO. 5, and SEQ ID NO. 6. SEQ ID NO. 2 encodes a preferred protein that is shown in SEQ ID NO. 3.

The polynucleotides of the subject invention can be used to form complete "genes" to encode proteins or peptides in a desired host cell. For example, as the skilled artisan would readily recognize, SEQ ID NO. 2, SEQ ID NO. 5, and SEQ ID NO. 6 are shown without stop codons. SEQ ID NO. 2, SEQ ID NO. 5, and/or SEQ ID NO. 6 can

be appropriately placed under the control of a promoter in a host of interest, as is readily known in the art.

As the skilled artisan would readily recognize, DNA can exist in a double-stranded form. In this arrangement, one strand is complementary to the other strand and vice versa. The "coding strand" is often used in the art to refer to the strand having a series of codons (a codon is three nucleotides that can be read three-at-a-time to yield a particular amino acid) that can be read as an open reading frame (ORF) to form a protein or peptide of interest. In order to express a protein *in vivo*, a strand of DNA is typically translated into a complementary strand of RNA which is used as the template for the protein. As DNA is replicated in a plant (for example) additional, complementary strands of DNA are produced. Thus, the subject invention includes the use of either the exemplified polynucleotides shown in the attached sequence listing or the complementary strands. RNA and PNA (peptide nucleic acids) that are functionally equivalent to the specifically exemplified, novel DNA molecules are included in the subject invention.

Certain DNA sequences of the subject invention have been specifically exemplified herein. These sequences are exemplary of the subject invention. It should be readily apparent that the subject invention includes not only the genes and sequences specifically exemplified herein but also equivalents and variants thereof (such as mutants, fusions, chimerics, truncations, fragments, and smaller genes) that exhibit the same or similar characteristics relating to expressing toxins in plants, as compared to those specifically disclosed herein. As used herein, "variants" and "equivalents" refer to sequences which have nucleotide (or amino acid) substitutions, deletions (internal and/or terminal), additions, or insertions which do not materially affect the expression of the subject genes, and the resultant pesticidal activity, in plants. Fragments of polynucleotide proteins retaining pesticidal activity, and "pesticidal portions" of full-length proteins, are also included in this definition.

Genes can be modified, and variations of genes may be readily constructed, using standard techniques. For example, techniques for making point mutations are well known in the art. In addition, commercially available exonucleases or endonucleases can be used according to standard procedures, and enzymes such as *Bal*31 or site-directed

mutagenesis can be used to systematically cut off nucleotides from the ends of these genes. Useful genes can also be obtained using a variety of restriction enzymes.

It should be noted that equivalent genes will encode toxins that have high amino acid identity or homology with the toxins encoded by the subject genes. The amino acid homology will be highest in critical regions of the toxin which account for biological activity or are involved in the determination of three-dimensional configuration which ultimately is responsible for the biological activity. In this regard, certain substitutions are acceptable and can be expected if these substitutions are in regions which are not critical to activity or are conservative amino acid substitutions which do not affect the three-dimensional configuration of the molecule. For example, amino acids may be placed in the following classes: non-polar, uncharged polar, basic, and acidic. Conservative substitutions whereby an amino acid of one class is replaced with another amino acid of the same type fall within the scope of the subject invention so long as the substitution does not materially alter the biological activity of the compound. Table 1 provides a listing of examples of amino acids belonging to each class.

Table 1.

Class of Amino Acid	Examples of Amino Acids
Nonpolar	Ala, Val, Leu, Ile, Pro, Met, Phe, Trp
Uncharged Polar	Gly, Ser, Thr, Cys, Tyr, Asn, Gln
Acidic	Asp, Glu
Basic	Lys, Arg, His

In some instances, non-conservative substitutions can also be made. The critical factor is that these substitutions must not significantly detract from the ability of plants to express the subject DNA sequences or from the biological activity of the toxin.

As used herein, reference to "isolated" polynucleotides and/or "purified" toxins refers to these molecules when they are not associated with the other molecules with which they would be found in nature and would include their use in plants. Thus, reference to "isolated" and/or "purified" signifies the involvement of the "hand of man" as described herein.

Recombinant hosts. The toxin-encoding genes of the subject invention can be introduced into a wide variety of microbial or plant hosts. In some embodiments of the subject invention, transformed microbial hosts can be used in preliminary steps for preparing precursors, for example, that will eventually be used to transform, in preferred
5 embodiments, plant cells and plants so that they express the toxins encoded by the genes of the subject invention. Microbes transformed and used in this manner are within the scope of the subject invention. Recombinant microbes may be, for example, a *B.t.*, *E. coli*, or *Pseudomonas*. Transformations can be made by those skilled in the art using standard techniques. Materials necessary for these transformations are disclosed herein
10 or are otherwise readily available to the skilled artisan.

Thus, in preferred embodiments, expression of a gene of this invention results, directly or indirectly, in the intracellular production and maintenance of the protein of interest. When transformed plants are ingested by the pest, the pests will ingest the toxin. The result is a control of the pest.

15 The *B.t.* toxin gene can be introduced via a suitable vector into a host, preferably a plant host. There are many crops of interest, such as corn, wheat, rice, cotton, soybeans, and sunflowers. The genes of the subject invention are particularly well suited for providing stable maintenance and expression, in the transformed plant, of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of
20 the pesticide from environmental degradation and inactivation.

Thus, the subject invention includes recombinant hosts comprising a polynucleotide sequence optimized for expression in a plant, wherein said sequence is selected from the group consisting of SEQ ID NO. 2, SEQ ID NO. 5, and SEQ ID NO. 6. The recombinant host can be, for example, a plant cell. Entire plants comprising the
25 subject polynucleotides are also within the scope of the subject invention. In particularly preferred embodiments, a plant can be made resistant to corn rootworm damage by being transformed to express a polynucleotide, such as SEQ ID NO. 2, that encodes an approximately 15 kDa protein, together with a second polynucleotide, such as SEQ ID NO. 4, which encodes a 45 kDa protein. Likewise, SEQ ID NO. 5 and SEQ ID NO. 6
30 can be used together, under one promoter or separate promoters, such as the ubiquitin promoter. For that matter, the polynucleotide of SEQ ID NO. 2 and SEQ ID NO. 6 can be used together, or SEQ ID NO. 4 and SEQ ID NO. 5 can be used, for example.

While the subject invention provides specific embodiments of synthetic genes, other genes that are functionally equivalent to the genes exemplified herein can also be used to transform hosts, preferably plant hosts. Additional guidance for the production of synthetic genes can be found in, for example, U.S. Patent No. 5,380,831.

5 Following is an example which illustrates procedures for practicing the invention. This example should not be construed as limiting.

Example 1- Insertion of Toxin Genes Into Plants

One aspect of the subject invention is the transformation of plants with the subject polynucleotide sequences encoding insecticidal toxins. The transformed plants are resistant
10 to attack by the target pest. The genes of the subject invention are optimized for use in plants.

Obviously, a promoter region capable of expressing the gene in a plant is needed. Thus, for *in planta* expression, the DNA of the subject invention is under the control of an appropriate promoter region. Techniques for obtaining *in planta* expression by using such
15 constructs is known in the art. A preferred promoter region used for expression of both 15 kDa and 45 kDa transgenes is the *Zea mays* ubiquitin promoter plus *Z. mays* exon 1 and *Z. mays* intron 1 (Christensen, A. H., *et al.* 1992 Plant Mol. Biol. 18:675-689). A preferred transcriptional terminator for both transgenes is the potato proteinase inhibitor II (PinII) terminator (An, G. *et al.* 1989 Plant Cell 1:115-22).

20 Genes encoding pesticidal toxins, as disclosed herein, can be inserted into plant cells using a variety of techniques which are well known in the art. For example, in preferred embodiments, maize plants containing 14 kDa and 44 kDa transgenes were obtained by microprojectile bombardment using the Biolistics® PDS-100He particle gun manufactured by Bio-Rad, essentially as described by Klein *et al.* *Nature*, 327:70-73
25 (1987).

A large number of cloning vectors comprising a replication system in *E. coli* and a marker that permits selection of the transformed cells are available for preparation for the insertion of foreign genes into higher plants. The vectors comprise, for example, pBR322, pUC series, M13mp series, pACYC184, etc. Accordingly, the sequence encoding the *B.t.*
30 toxin can be inserted into the vector at a suitable restriction site. The resulting plasmid is

used for transformation into *E. coli*. The *E. coli* cells are cultivated in a suitable nutrient medium, then harvested and lysed. The plasmid is recovered. Sequence analysis, restriction analysis, electrophoresis, and other biochemical-molecular biological methods are generally carried out as methods of analysis. After each manipulation, the DNA sequence
5 used can be cleaved and joined to the next DNA sequence. Each plasmid sequence can be cloned in the same or other plasmids.

Depending on the method of inserting desired genes into the plant, other DNA sequences may be necessary. If, for example, the Ti or Ri plasmid is used for the transformation of the plant cell, then at least the right border, but often the right and the left
10 border of the Ti or Ri plasmid T-DNA, has to be joined as the flanking region of the genes to be inserted. The use of T-DNA for the transformation of plant cells has been intensively researched and sufficiently described in EP 120 516; Hoekema (1985) In: *The Binary Plant Vector System*, Offset-durkkerij Kanters B. V., Alblaserdam, Chapter 5; Fraley *et al.* (1986), *Crit. Rev. Plant Sci.* 4:1-46; and An *et al.* (1985) *EMBO J.* 4:277-287.

15 Once the inserted DNA has been integrated in the genome, it is relatively stable there and, as a rule, does not come out again. It normally contains a selection marker that confers on the transformed plant cells resistance to a biocide or an antibiotic, such as kanamycin, G 418, bleomycin, hygromycin, or chloramphenicol, *inter alia*. The individually employed marker should accordingly permit the selection of transformed cells
20 rather than cells that do not contain the inserted DNA.

A large number of techniques are available for inserting DNA into a plant host cell. Those techniques include transformation with T-DNA using *Agrobacterium tumefaciens* or *Agrobacterium rhizogenes* as transformation agent, fusion, injection, biolistics (microparticle bombardment), or electroporation as well as other possible methods. If
25 *Agrobacteria* are used for the transformation, the DNA to be inserted has to be cloned into special plasmids, namely either into an intermediate vector or into a binary vector. The intermediate vectors can be integrated into the Ti or Ri plasmid by homologous recombination owing to sequences that are homologous to sequences in the T-DNA. The Ti or Ri plasmid also comprises the *vir* region necessary for the transfer of the T-DNA.
30 Intermediate vectors cannot replicate themselves in *Agrobacteria*. The

intermediate vector can be transferred into *Agrobacterium tumefaciens* by means of a helper plasmid (conjugation). Binary vectors can replicate themselves both in *E. coli* and in *Agrobacteria*. They comprise a selection marker gene and a linker or polylinker which are framed by the right and left T-DNA border regions. They can be transformed directly
5 into *Agrobacteria* (Holsters *et al.* [1978] *Mol. Gen. Genet.* 163:181-187). The *Agrobacterium* used as host cell is to comprise a plasmid carrying a *vir* region. The *vir* region is necessary for the transfer of the T-DNA into the plant cell. Additional T-DNA may be contained. The bacterium so transformed is used for the transformation of plant cells. Plant explants can advantageously be cultivated with *Agrobacterium tumefaciens*
10 or *Agrobacterium rhizogenes* for the transfer of the DNA into the plant cell. Whole plants can then be regenerated from the infected plant material (for example, pieces of leaf, segments of stalk, roots, but also protoplasts or suspension-cultivated cells) in a suitable medium, which may contain antibiotics or biocides for selection. The plants so obtained can then be tested for the presence of the inserted DNA. No special demands
15 are made of the plasmids in the case of injection and electroporation. It is possible to use ordinary plasmids, such as, for example, pUC derivatives.

The transformed cells grow inside the plants in the usual manner. They can form germ cells and transmit the transformed trait(s) to progeny plants. Such plants can be grown in the normal manner and crossed with plants that have the same transformed
20 hereditary factors or other hereditary factors. The resulting hybrid individuals have the corresponding phenotypic properties.

It should be understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof
25 will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and the following claims.

WO 00/24904

PCT/US99/24647

1

SEQUENCE LISTING

<110> Mycogen Corporation

<120> Plant-Optimized Polynucleotides Encoding Approximately
15 kDa and Approximately 45 kDa Pesticidal Proteins

<130> MA723/4X

<140>

<141>

<150> 60/105,408

<151> 1998-10-23

<150> 60/105,359

<151> 1998-10-23

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<170> PatentIn Ver. 2.1

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<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence: Synthetic
Bacillus thuringiensis toxin gene

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gacaccatca agacgttcgt ggcggagtc caccggcttca tgaccggcgt cgagggcatc 180

atctacttct ccgtgaacgg cgacgccgag atctccctcc acttcgacaa cccgtacatc 240

ggctccaaca agtgcgacgg ctccctccgac aagcccagat acgaggtgat caccagtc 300

ggctccggcg acaagtccca cgtgacctac accatccaga ccgtgtccct ccgcctc 357

<210> 2

<211> 357

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Synthetic
Bacillus thuringiensis toxin gene

<400> 2

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2

atgtccgccc gcgaggtgca catcgagatc aacaacaaga cccgccacac cctccagctc 60
 gaggacaaga ccaagctctc cggcggcagg tggcgcacct ccccgaccaa cgtggcccgc 120
 gacaccatca agacgttcgt ggcggagtcc cacggcttca tgaccggcgt cgagggcatc 180
 atctacttct ccgtgaacgg cgacgccgag atctccctcc acttcgacaa cccgtacatc 240
 ggctccaaca agtccgacgg ctctccgac aagcccagat acgaggtgat caccaggtcc 300
 ggctccggcg acaagtccca cgtgacctac accatccaga ccgtgtccct ccgcctc 357

<210> 3

<211> 119

<212> PRT

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Toxin encoded
 by synthetic *Bacillus thuringiensis* gene

<400> 3

Met	Ser	Ala	Arg	Glu	Val	His	Ile	Glu	Ile	Asn	Asn	Lys	Thr	Arg	His
1				5				10					15		

Thr	Leu	Gln	Leu	Glu	Asp	Lys	Thr	Lys	Leu	Ser	Gly	Gly	Arg	Trp	Arg
		20						25					30		

Thr	Ser	Pro	Thr	Asn	Val	Ala	Arg	Asp	Thr	Ile	Lys	Thr	Phe	Val	Ala
		35					40					45			

Glu	Ser	His	Gly	Phe	Met	Thr	Gly	Val	Glu	Gly	Ile	Ile	Tyr	Phe	Ser
		50				55					60				

Val	Asn	Gly	Asp	Ala	Glu	Ile	Ser	Leu	His	Phe	Asp	Asn	Pro	Tyr	Ile
	65				70					75					80

Gly	Ser	Asn	Lys	Ser	Asp	Gly	Ser	Ser	Asp	Lys	Pro	Glu	Tyr	Glu	Val
			85						90					95	

Ile	Thr	Gln	Ser	Gly	Ser	Gly	Asp	Lys	Ser	His	Val	Thr	Tyr	Thr	Ile
			100					105					110		

Gln	Thr	Val	Ser	Leu	Arg	Leu
						115

<210> 4

<211> 1155

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Synthetic

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3

Bacillus thuringiensis toxin gene

<400> 4

atgctcgaca ccaacaaggt gtacgagatc tccaacctcg ccaacggcct ctacacctcc 60
acctacctct ccctcgacga ctccggcgtg tccctcatgt ccaagaagga cgaggacatc 120
gacgactaca acctcaagtg gttcctcttc ccgatcgaca acaaccagta catcatcacc 180
tcctacggcg ccaacaactg caaggtgtgg aacgtgaaga acgacaagat caacgtgtcc 240
acctactcct ccaccaactc cgtgcagaag tggcagatca aggccaagga ctctcctac 300
atcatccagt ccgacaacgg caaggtgctc accgcgggcg tgggccagtc cctcggcatc 360
gtgcgcctca ccgacgagtt cccggagaac tccaaccagc aatggaacct caccgccgtg 420
cagaccatcc agtccccgca gaagccgaag atcgacgaga agtcaagga ccaccggag 480
tactccgaga ccggcaacat caaccgaag accaccgcc agtcatggg ctggaccctc 540
gtgccgtgca tcatggtgaa cgactccaag atcgacaaga acaccagat caagaccacc 600
ccgtactaca tcttcaagaa atacaagtac tggaacctcg ccaagggctc caacgtgtcc 660
ctctccgcc accagaagcg cagctacgac tacgagtggg gcaccgagaa gaaccagaag 720
accaccatca tcaacaccgt gggcctgcag atcaacatcg actcggggat gaagtctgag 780
gtgccggagg tgggcggcgg caccgaggac atcaagacc agtccaccga ggagctgaag 840
gtggagtact ccaccgagac caagatcatg accaagtacc aggagcactc cgagatcgac 900
aaccgacca accagccgat gaactccatc ggcctcctca tctacacctc cctcgagctg 960
taccgctaca acggcaccga gatcaagatc atggacatcg agacctccga ccacgacacc 1020
tacacctca cctcctaccc gaaccacaag gaggcgctgc tgctgctgac caaccactcc 1080
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cactacttca agaag 1155

<210> 5

<211> 369

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Synthetic
Bacillus thuringiensis toxin gene

<400> 5

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4

gaggacaaga ccaagctcga cggcggcagg tggcgcacct ccccgaccaa cgtggccaac 120
 gaccagatca agaccttcgt ggccgaatcc aacggcttca tgaccggcac cgagggcacc 180
 atctactact ccatcaacgg cgaggccgag atcagcctct acttcgacaa cccgttcgcc 240
 ggctccaaca aatacgacgg ccactccaac aagtcccagt acgagatcat caccagggc 300
 ggctccggca accagtccca cgtgacctac accatccaga ccacctctc ccgctacggc 360
 cacaagtcc 369

<210> 6

<211> 1149

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence: Synthetic
 Bacillus thuringiensis toxin gene

<400> 6

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 acctacctct ccctcgacga ctccggcgtg tcctcatga acaagaacga cgacgacatc 120
 gacgactaca acctcaagtg gttcctcttc ccgatcgacg acgaccagta catcatcacc 180
 tcctacgccg ccaacaactg caaggtgtgg aacgtgaaca acgacaagat caacgtgtcc 240
 acctactcct ccaccaactc catccagaag tggcagatca aggccaacgg ctctcctac 300
 gtgatccagt ccgacaacgg caaggtgctc accgccggca ccggccaggc cctcggcctc 360
 atccgcctca ccgacgagtc ctccaacaac ccgaaccagc aatggaacct gacgtccgtg 420
 cagaccatcc agtccccgca gaagccgatc atcgacacca agtcaagga ctaccggaag 480
 tactccccga ccggcaacat cgacaacggc acctccccgc agtcatggg ctggaccctc 540
 gtgccgtgca tcatggtgaa cgacccgaac atcgacaaga acaccagat caagaccacc 600
 ccgtactaca tcctcaagaa gtaccagtac tggcagaggg ccgtgggctc caacgtcgcg 660
 ctccgcccgc acgagaagaa gtcctacacc tacgagtggg gcaccgagat cgaccagaag 720
 accaccatca tcaacaccct cggcttccag atcaacatcg acagcggcat gaagttcgac 780
 atcccggagg tgggcggcgg taccgacgag atcaagacct agtcaacga ggagctcaag 840
 atcgagtact cccacgagac gaagatcatg gagaagtacc aggagcagtc cgagatcgac 900
 aacccgaccg accagtccat gaactccatc ggcttcctca ccatcacctc cctggagctc 960

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5

taccgctaca acggctccga gatccgcatc atgcagatcc agacctccga caacgacacc 1020

tacaacgtga cctcctaccc gaaccaccag caggccctgc tgctgctgac caaccactcc 1080

tacgaggagg tggaggagat caccaacatc ccgaagtcca cctcaagaa gctcaagaag 1140

tactacttc 1149

CLAIMS:

1. A polynucleotide which comprises nucleotide sequence SEQ ID NO:5.
2. A recombinant host cell that comprises the polynucleotide according to claim 1.
3. The host cell according to claim 2 wherein said cell is a plant cell.
- 5 4. The host cell according to claim 2 wherein said cell is a maize plant cell.
5. A method of producing the recombinant host cell of claim 2 wherein said method comprises introducing the polynucleotide of claim 1 into the genome of said cell.
6. The method according to claim 5 wherein said cell is a plant cell.
7. The method according to claim 5 wherein said cell is a maize plant cell.
- 10 8. A method for controlling a plant pest wherein said method comprises contacting said pest with a protein encoded by the polynucleotide according to claim 1 wherein said protein is produced by said polynucleotide in a recombinant host cell.
9. The method according to claim 8 wherein said cell is a plant cell.
10. The method according to claim 8 wherein said cell is a *Zea mays* plant cell.
- 15 11. The method according to claim 8, 9 or 10 wherein said method further comprises contacting said pest with a 45kDa pesticidal protein.