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(54) Title: CYANOBACTERIA SAXITOXIN GENE CLUSTER AND DETECTION OF CYANOTOXIC ORGANISMS

(57) Abstract: The present invention relates to methods for the detection of cyanobacteria, dinoflagellates, and in particular, methods for the detection of cyanotoxic organisms. Kits for the detection of cyanobacteria, dinoflagellates, and cyanotoxic organisms are provided. The invention further relates to methods of screening for compounds that modulate the activity of polynucleotides and/or polypeptides of the saxitoxin and cylindrospermopsin biosynthetic pathways.



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Cyanobacteria saxitoxin gene cluster and detection of cyanotoxic organisms.**Technical Field**

The present invention relates to methods for the detection of cyanobacteria, dinoflagellates, and in particular, methods for the detection of cyanotoxic organisms. Kits for the detection of cyanobacteria, dinoflagellates, and cyanotoxic organisms are provided. The invention further relates to methods of screening for compounds that modulate the activity of polynucleotides and/or polypeptides of the saxitoxin and cylindrospermopsin biosynthetic pathways.

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Background

Cyanobacteria, also known as blue-green algae, are photosynthetic bacteria widespread in marine and freshwater environments. Of particular significance for water quality and human and animal health are those cyanobacteria which produce toxic compounds. Under eutrophic conditions cyanobacteria tend to form large blooms which drastically promote elevated toxin concentrations. Cyanobacterial blooms may flourish and expand in coastal waters, streams, lakes, and in drinking water and recreational reservoirs. The toxins they produce can pose a serious health risk for humans and animals and this problem is internationally relevant since most toxic cyanobacteria have a global distribution.

A diverse range of cyanobacterial genera are well known for the formation of toxic blue-green algal blooms on water surfaces. Saxitoxin (*SXT*) and its analogues cause the paralytic shellfish poisoning (PSP) syndrome, which afflicts human health and impacts on coastal shellfish economies worldwide. PSP toxins are unique alkaloids, being produced by both prokaryotes and eukaryotes. PSP toxins are among the most potent and pervasive algal toxins and are considered a serious toxicological health-risk that may affect humans, animals and ecosystems worldwide. These toxins block voltage-gated sodium and calcium channels, and prolong the gating of potassium channels preventing the transduction of neuronal signals. It has been estimated that more than 2000 human cases of PSP occur globally every year. Moreover, coastal blooms of producing microorganisms result in millions of dollars of economic damage due to PSP toxin contamination of seafood and the continuous requirement for costly biotoxin monitoring programs. Early warning systems to anticipate paralytic shellfish toxin (PST)-producing

algal blooms, such as PCR and ELISA-based screening, are as yet unavailable due to the lack of data on the genetic basis of PST production.

SXT is a tricyclic perhydropurine alkaloid which can be substituted at various positions leading to more than 30 naturally occurring *SXT* analogues. Although *SXT* biosynthesis seems complex and unique, organisms from two kingdoms, including certain species of marine dinoflagellates and freshwater cyanobacteria, are capable of producing these toxins, apparently by the same biosynthetic route. In spite of considerable efforts none of the enzymes or genes involved in the biosynthesis and modification of *SXT* have been previously identified.

The occurrence of the cyanobacterial genus *Cylindrospermopsis* has been documented on all continents and therefore poses a significant public health threat on a global scale. The major toxin produced by *Cylindrospermopsis* is cylindrospermopsin (CYR). Besides posing a threat to human health, cylindrospermopsin also causes significant economic losses for farmers due to the poisoning of livestock with cylindrospermopsin-contaminated drinking water. Cylindrospermopsin has hepatotoxic, general cytotoxic and neurotoxic effects and is a potential carcinogen. Its toxicity is due to the inhibition of glutathione and protein synthesis as well as inhibiting cytochrome P450. Six cyanobacterial species have so far been identified to produce cylindrospermopsin; *Cylindrospermopsis raciborskii*, *Aphanizomenon ovalisporum*, *Aphanizomenon flos-aquae*, *Umezakia natans*, *Rhaphidiopsis curvata* and *Anabaena bergii*. Incidents of human poisoning with cylindrospermopsin have only been reported in sub-tropical Australia to date, however *C. raciborskii* and *A. flos-aquae* have recently been detected in areas with more temperate climates. The tendency of *C. raciborskii* to form dense blooms and the invasiveness of the producer organisms gives rise to global concerns for drinking water quality and necessitates the monitoring of drinking water reserves for the presence of cylindrospermopsin producers.

There is a need for rapid and accurate methods detecting cyanobacteria, and in particular those strains which are capable of producing cyanotoxins such as saxitoxin and cylindrospermopsin. Rapid and accurate methods for detecting cyanotoxic organisms are needed for assessing the potential health hazard of cyanobacterial blooms and for the implementation of effective water management strategies to minimize the effects of toxic bloom outbreaks.

Summary

In a first aspect, there is provided an isolated polynucleotide comprising a sequence according to SEQ ID NO: 1 or a variant or fragment thereof, wherein said fragment encodes a protein of a saxitoxin biosynthetic pathway.

5 In one embodiment of the first aspect, the fragment comprises a sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, 10 SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof.

In a second aspect, there is provided an isolated ribonucleic acid or an isolated 15 complementary DNA encoded by a sequence according to the first aspect.

In a third aspect, there is provided an isolated polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, 20 SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, and variants and fragments thereof.

25 In one embodiment, there is provided a probe or primer that hybridises specifically with one or more of: a polynucleotide according to the first aspect, a ribonucleic acid or complementary DNA according to the second aspect, or a polypeptide according the third aspect.

In another embodiment, there is provided a vector comprising a polynucleotide 30 according to the first aspect, or a ribonucleic acid or complementary DNA according the second aspect. The vector may be an expression vector.

In another embodiment, a host cell is provided comprising the vector.

In another embodiment, there is provided an isolated antibody capable of binding specifically to a polypeptide according to the third aspect.

In a fourth aspect, there is provided a method for the detection of cyanobacteria and/or dinoflagellates, the method comprising the steps of obtaining a sample for use in the method and analyzing the sample for the presence of one or more of:

- 5 (i) a polynucleotide comprising a sequence according to the first aspect
- (ii) a ribonucleic acid or complementary DNA according to the second aspect
- (iii) a polypeptide comprising a sequence according to third aspect

wherein said presence is indicative of cyanobacteria and/or dinoflagellates in the sample.

10 In a fifth aspect, there is provided a method for detecting a cyanotoxic organism, the method comprising the steps of obtaining a sample for use in the method and analyzing the sample for the presence of one or more of:

- (i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 36, and variants thereof sharing at least 80% sequence homology with SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, or SEQ ID NO: 36,

15 (ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i)

- (iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 37, and variants thereof sharing at least 80% sequence homology with SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, or SEQ ID NO: 37,

20 wherein said presence is indicative of cyanotoxic organisms in the sample.

In one embodiment of the fifth aspect, the cyanotoxic organism is a cyanobacteria or a dinoflagellate.

25 In one embodiment of the fourth and fifth aspects, analyzing the sample comprises amplification of DNA from the sample by polymerase chain reaction and detecting the amplified sequences. The polymerase chain reaction may utilise one or more primers comprising a sequence selected from the group consisting of SEQ ID NO: 70, SEQ ID NO: 71, SEQ ID NO: 72, SEQ ID NO: 73, SEQ ID NO: 74, SEQ ID NO: 75, SEQ ID NO: 76, SEQ ID NO: 77, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 113, SEQ ID NO: 114, SEQ ID NO: 115, SEQ ID NO: 116, SEQ ID NO: 117, SEQ ID NO: 118, SEQ ID NO: 119, SEQ ID NO: 120, SEQ ID NO: 121, SEQ ID NO: 122, SEQ ID NO: 123, SEQ ID NO: 124, SEQ ID NO: 125, SEQ ID NO: 126, SEQ ID NO: 127, SEQ ID NO:

128, SEQ ID NO: 129, SEQ ID NO: 130, SEQ ID NO: 131, SEQ ID NO: 132, SEQ ID NO: 133, SEQ ID NO: 134, and variants and fragments thereof.

In another embodiment of the fourth and fifth aspects, the method comprises further analyzing the sample for the presence of one or more of:

5 (i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants and fragments thereof,

10 (ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, 15 SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, and SEQ ID NO: 110, and variants and fragments thereof.

The further analysis of the sample may comprise amplification of DNA from the sample by polymerase chain reaction. The polymerase chain reaction may utilise one or more primers comprising a sequence selected from the group consisting of SEQ ID NO: 20 111, SEQ ID NO: 112, or variants or fragments thereof.

Described herein is a method for the detection of dinoflagellates, the method comprising the steps of obtaining a sample for use in the method and analyzing the sample for the presence of one or more of:

(i) a polynucleotide comprising a sequence according to the first aspect,
25 (ii) a ribonucleic acid or complementary DNA according to the second aspect,
(iii) a polypeptide comprising a sequence according to the third aspect,

wherein said presence is indicative of dinoflagellates in the sample.

Analysing the sample may comprise amplification of DNA from the sample by polymerase chain reaction and detecting the amplified sequences.

30 In one embodiment of the fourth and fifth aspects, the detection comprises one or both of gel electrophoresis and nucleic acid sequencing. The sample may comprise one or more isolated or cultured organisms. The sample may be an environmental sample. The environmental sample may be derived from salt water, fresh water or a blue-green algal bloom.

In a sixth aspect, there is provided a kit for the detection of cyanobacteria and/or dinoflagellates, the kit comprising at least one agent for detecting the presence of one or more of:

- (i) a polynucleotide comprising a sequence according to the first aspect,
- 5 (ii) a ribonucleic acid or complementary DNA according to the second aspect,
- (iii) a polypeptide comprising a sequence according to the third aspect,

wherein said presence is indicative of cyanobacteria and/or dinoflagellates in the sample.

In a seventh aspect, there is provided a kit for the detection of cyanotoxic organisms, the kit comprising at least one agent for detecting the presence of one or more
10 of:

(i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 36, and variants thereof sharing at least 90% sequence homology with SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, or SEQ ID NO: 36,

15 (ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 37, and variants thereof sharing at least 90% sequence homology with SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, or SEQ ID NO: 37,
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wherein said presence is indicative of cyanotoxic organisms in the sample.

In one embodiment of the sixth and seventh aspects, the at least one agent is a primer, antibody or probe. The primer or probe may comprise a sequence selected from the group consisting of SEQ ID NO: 70, SEQ ID NO: 71, SEQ ID NO: 72, SEQ ID NO: 73, SEQ ID NO: 74, SEQ ID NO: 75, SEQ ID NO: 76, SEQ ID NO: 77, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 113, SEQ ID NO: 114, SEQ ID NO: 115, SEQ ID NO: 116, SEQ ID NO: 117, SEQ ID NO: 118, SEQ ID NO: 119, SEQ ID NO: 120, SEQ ID NO: 121, SEQ ID NO: 122, SEQ ID NO: 123, SEQ ID NO: 124, SEQ ID NO: 125, SEQ ID NO: 126, SEQ ID NO: 127, SEQ ID NO: 128, SEQ ID NO: 129, SEQ ID NO: 130, SEQ ID NO: 131, SEQ ID NO: 132, SEQ ID NO: 133, SEQ ID NO: 134, and variants and fragments thereof.
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In another embodiment of the sixth and seventh aspects, the kit further comprises at least one additional agent for detecting the presence of one or more of:

(i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants and fragments thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, and SEQ ID NO: 110, and variants and fragments thereof.

The at least one additional agent may be a primer, antibody or probe. The primer or probe may comprise a sequence selected from the group consisting of SEQ ID NO: 109, SEQ ID NO: 110, and variants and fragments thereof.

Described herein is a kit for the detection of dinoflagellates, the kit comprising at least one agent for detecting the presence of one or more of:

(i) a polynucleotide comprising a sequence according to the first aspect,

(ii) a ribonucleic acid or complementary DNA according to the second aspect,

(iii) a polypeptide comprising a sequence according to the third aspect,

wherein said presence is indicative of dinoflagellates in the sample.

In an eighth aspect, there is provided a method of screening for a compound that modulates the expression or activity of one or more polypeptides according to the third aspect, the method comprising contacting the polypeptide with a candidate compound under conditions suitable to enable interaction of the candidate compound and the polypeptide, and assaying for activity of the polypeptide.

In one embodiment of the eighth aspect, modulating the expression or activity of one or more polypeptides comprises inhibiting the expression or activity of said polypeptide.

In another embodiment of the eighth aspect, modulating the expression or activity of one or more polypeptides comprises enhancing the expression or activity of said polypeptide.

Described herein is an isolated polynucleotide comprising a sequence according to SEQ ID NO: 80 or a variant or fragment thereof.

The fragment may comprise a sequence selected from the group consisting of SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and
5 variants and fragments thereof.

Described herein is a ribonucleic acid or complementary DNA encoded by a sequence according to the present invention.

Described herein is an isolated polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, SEQ ID NO: 110, and variants and fragments thereof.
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In one embodiment, there is provided a probe or primer that hybridises specifically with one or more of: a polynucleotide according to the present invention, a ribonucleic acid or complementary DNA according to the present invention, or a polypeptide according to the present invention.
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In another embodiment, there is provided a vector comprising a polynucleotide according to the present invention, or a ribonucleic acid or complementary DNA according to the present invention. The vector may be an expression vector. In one embodiment, a host cell is provided comprising the vector.
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In another embodiment, there is provided an isolated antibody capable of binding specifically to a polypeptide according to the present invention.

Described herein is a method for the detection of cyanobacteria, the method comprising the steps of obtaining a sample for use in the method and analyzing the sample for the presence of one or more of:
25

- (i) a polynucleotide comprising a sequence according to the present invention,
- (ii) a ribonucleic acid or complementary DNA according to the present invention,
- (iii) a polypeptide comprising a sequence according to present invention,

wherein said presence is indicative of cyanobacteria in the sample.

Described herein is a method for detecting a cyanotoxic organism, the method comprising the steps of obtaining a sample for use in the method and analyzing the sample for the presence of one or both of:
30

- (i) a polynucleotide comprising a sequence according to SEQ ID NO: 95 or a variant or fragment thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence according to SEQ ID NO: 96, or a variant or fragment thereof,

5 wherein said presence is indicative of a cyanotoxic organism in the sample.

The cyanotoxic organism may be a cyanobacterium.

Analyzing the sample may comprise amplification of DNA from the sample by polymerase chain reaction and detecting the amplified sequences. The polymerase chain reaction may utilise one or more primers comprising a sequence selected from the group consisting of SEQ ID NO: 111, SEQ ID NO: 112 and variants and fragments thereof.

The method may comprise analyzing the sample for the presence of one or more of:

(i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, and variants and fragments thereof.

The further analysis of the sample may comprise amplification of DNA from the sample by polymerase chain reaction. The polymerase chain reaction may utilise one or more primers comprising a sequence selected from the group consisting of SEQ ID NO: 70, SEQ ID NO: 71, SEQ ID NO: 72, SEQ ID NO: 73, SEQ ID NO: 74, SEQ ID NO: 75,

SEQ ID NO: 76, SEQ ID NO: 77, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 113, SEQ ID NO: 114, SEQ ID NO: 115, SEQ ID NO: 116, SEQ ID NO: 117, SEQ ID NO: 118, SEQ ID NO: 119, SEQ ID NO: 120, SEQ ID NO: 121, SEQ ID NO: 122, SEQ ID NO: 123, SEQ ID NO: 124, SEQ ID NO: 125, SEQ ID NO: 126, SEQ ID NO: 127, SEQ ID NO: 128, SEQ ID NO: 129, SEQ ID NO: 130, SEQ ID NO: 131, SEQ ID NO: 132, SEQ ID NO: 133, SEQ ID NO: 134, and variants and fragments thereof.

Described herein is a method for detecting a cylindrospermopsin-producing organism, the method comprising the steps of obtaining a sample for use in the method and analyzing the sample for the presence of one or both of:

(i) a polynucleotide comprising a sequence according to SEQ ID NO: 95 or a variant or fragments thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence according to SEQ ID NO: 96, or a variant or fragments thereof,

wherein said presence is indicative of a cylindrospermopsin-producing organism in the sample.

The cyanotoxic organism may be a cyanobacterium. Analyzing the sample may comprise amplification of DNA from the sample by polymerase chain reaction and detecting the amplified sequences. The polymerase chain reaction may utilise one or more primers comprising a sequence selected from the group consisting of SEQ ID NO: 111, SEQ ID NO: 112 and variants and fragments thereof.

The detection may comprise one or both of gel electrophoresis and nucleic acid sequencing. The sample may comprise one or more isolated or cultured organisms. The sample may be an environmental sample. The environmental sample may be derived from salt water, fresh water or a blue-green algal bloom.

Described herein is a kit for the detection of cyanobacteria, the kit comprising at least one agent for detecting the presence of one or more of:

(i) a polynucleotide comprising a sequence according to the present invention,

(ii) a ribonucleic acid or complementary DNA according to the present invention,

(iii) a polypeptide comprising a sequence according to the present invention,

wherein said presence is indicative of cyanobacteria in the sample.

Described herein is a kit for the detection of cyanotoxic organisms, the kit comprising at least one agent for detecting the presence of one or more of:

(i) a polynucleotide comprising a sequence according to SEQ ID NO: 95 or a variant or fragment thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

5 (iii) a polypeptide comprising a sequence according to SEQ ID NO: 96, or a variant or fragment thereof,

wherein said presence is indicative of cyanotoxic organisms in the sample.

The at least one agent may be a primer, antibody or probe. The primer or probe may comprise a sequence selected from the group consisting of SEQ ID NO: 111, SEQ ID
10 NO: 112 and variants and fragments thereof.

The kit may further comprise at least one additional agent for detecting the presence of one or more nucleotide sequences selected from the group consisting of:

(i) a polynucleotide comprising a sequence selected from the group consisting of:
15 SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID
20 NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of:
25 SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, and variants and fragments thereof.
30

The at least one additional agent may be a primer, antibody or probe. The primer or probe may comprise a sequence selected from the group consisting of SEQ ID NO: 70, SEQ ID NO: 71, SEQ ID NO: 72, SEQ ID NO: 73, SEQ ID NO: 74, SEQ ID NO: 75,

SEQ ID NO: 76, SEQ ID NO: 77, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 113, SEQ ID NO: 114, SEQ ID NO: 115, SEQ ID NO: 116, SEQ ID NO: 117, SEQ ID NO: 118, SEQ ID NO: 119, SEQ ID NO: 120, SEQ ID NO: 121, SEQ ID NO: 122, SEQ ID NO: 123, SEQ ID NO: 124, SEQ ID NO: 125, SEQ ID NO: 126, SEQ ID NO: 127, SEQ ID NO: 128, SEQ ID NO: 129, SEQ ID NO: 130, SEQ ID NO: 131, SEQ ID NO: 132, SEQ ID NO: 133, SEQ ID NO: 134, and variants and fragments thereof.

Described herein is a kit for the detection of cylindrospermopsin-producing organisms, the kit comprising at least one agent for detecting the presence of one or more of:

(i) a polynucleotide comprising a sequence according to SEQ ID NO: 95 or a variant or fragment thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence according to SEQ ID NO: 96, or a variant or fragment thereof,

wherein said presence is indicative of a cylindrospermopsin-producing organism in the sample.

Described herein is a method of screening for a compound that modulates the expression or activity of one or more polypeptides according to the present invention, the method comprising contacting the polypeptide with a candidate compound under conditions suitable to enable interaction of the candidate compound and the polypeptide, and assaying for activity of the polypeptide.

Modulating the expression or activity of one or more polypeptides may comprise inhibiting the expression or activity of said polypeptide.

Modulating the expression or activity of one or more polypeptides may comprise enhancing the expression or activity of said polypeptide.

Definitions

As used in this application, the singular form “a”, “an” and “the” include plural references unless the context clearly dictates otherwise. For example, the term “a stem cell” also includes a plurality of stem cells.

5 As used herein, the term “comprising” means “including.” Variations of the word “comprising”, such as “comprise” and “comprises,” have correspondingly varied meanings. Thus, for example, a polynucleotide “comprising” a sequence encoding a protein may consist exclusively of that sequence or may include one or more additional sequences.

10 As used herein, the terms “antibody” and “antibodies” include IgG (including IgG1, IgG2, IgG3, and IgG4), IgA (including IgA1 and IgA2), IgD, IgE, or IgM, and IgY, whole antibodies, including single-chain whole antibodies, and antigen-binding fragments thereof. Antigen-binding antibody fragments include, but are not limited to, Fab, Fab' and F(ab')₂, Fd, single-chain Fvs (scFv), single-chain antibodies, disulfide-linked Fvs (sdFv)
15 and fragments comprising either a VL or VH domain. The antibodies may be from any animal origin. Antigen-binding antibody fragments, including single-chain antibodies, may comprise the variable region(s) alone or in combination with the entire or partial of the following: hinge region, CH1, CH2, and CH3 domains. Also included are any combinations of variable region(s) and hinge region, CH1, CH2, and CH3 domains.
20 Antibodies may be monoclonal, polyclonal, chimeric, multispecific, humanized, and human monoclonal and polyclonal antibodies which specifically bind the biological molecule.

As used herein, the terms “polypeptide” and “protein” are used interchangeably and are taken to have the same meaning.

25 As used herein, the terms “nucleotide sequence” and “polynucleotide sequence” are used interchangeably and are taken to have the same meaning.

As used herein, the term “kit” refers to any delivery system for delivering materials. In the context of the detection assays described herein, such delivery systems include systems that allow for the storage, transport, or delivery of reaction reagents (for example
30 labels, reference samples, supporting material, etc. in the appropriate containers) and/or supporting materials (for example, buffers, written instructions for performing the assay etc.) from one location to another. For example, kits include one or more enclosures, such as boxes, containing the relevant reaction reagents and/or supporting materials.

Any discussion of documents, acts, materials, devices, articles or the like which has been included in the present specification is solely for the purpose of providing a context for the present invention. It is not to be taken as an admission that any or all of these matters form part of the prior art base or were common general knowledge in the field relevant to the present invention before the priority date of this application.

For the purposes of description all documents referred to herein are incorporated by reference unless otherwise stated.

Brief Description of the Drawings

A preferred embodiment of the present invention will now be described, by way of an example only, with reference to the accompanying drawings wherein:

Figure 1A is a table showing the distribution of the *sxt* genes in toxic and non-toxic cyanobacteria. PSP, saxitoxin; CYLN, cylindrospermopsin; +, gene fragment amplified; - no gene detected.

Figure 1B is a table showing primer sequences used to amplify various *SXT* genes.

Figure 2 is a table showing *sxt* genes from the saxitoxin gene cluster of *C. raciborskii* T3, their putative length, their BLAST similarity match with similar protein sequences from other organisms, and their predicted function.

Figure 3 is a diagram showing the structural organisation of the *sxt* gene cluster from *C. raciborskii* T3. Abbreviations used are: IS4, insertion sequence 4; at, aminotransferase; dmt, drug metabolite transporter; ompR, transcriptional regulator of *ompR* family; penP, penicillin binding; smf, gene predicted to be involved in DNA uptake. The scale indicates the gene cluster length in base pairs.

Figure 4 is a flow diagram showing the pathway for *SXT* biosynthesis and the putative functions of *sxt* genes.

Figure 5 shows MS/MS spectra of selected ions from cellular extracts of *Cylindrospermopsis raciborskii* T3. The predicted fragmentation of ions and the corresponding *m/z* values are indicated. Figure 5A, arginine (*m/z* 175); Figure 5B, saxitoxin (*m/z* 300); Figure 5C, intermediate A' (*m/z* 187); Figure 5D, intermediate C' (*m/z* 211); Figure 5E, intermediate E' (*m/z* 225).

Figure 6 is a table showing the *cyr* genes from the cylindrospermopsin gene cluster of *C. raciborskii* AWT205, their putative length, their BLAST similarity match with similar protein sequences from other organisms, and their predicted function.

Figure 7 is a table showing the distribution of the sulfotransferase gene (*cyrJ*) in toxic and non-toxic cyanobacteria. 16S rRNA gene amplification is shown as a positive control. CYLN, cylindrospermopsin; SXT, saxitoxin; N.D., not detected; +, gene fragment amplified; -, no gene detected; NA, not available; AWQC, Australian Water
5 Quality Center.

Figure 8 is a flow diagram showing the biosynthetic pathway of cylindrospermopsin biosynthesis.

Figure 9 is a diagram showing the structural organization of the cylindrospermopsin gene cluster from *C. raciborskii* AWT205. Scale indicates gene cluster length in base
10 pairs.

Description

The inventors have identified a gene cluster responsible for saxitoxin biosynthesis (the *SXT* gene cluster) and a gene cluster responsible for cylindrospermopsin biosynthesis (the *CYR* gene cluster). The full sequence of each gene cluster has been determined and
15 functional activities assigned to each of the genes identified therein. Based on this information, the inventors have elucidated the full saxitoxin and cylindrospermopsin biosynthetic pathways.

Accordingly, the invention provides polynucleotide and polypeptide sequences
20 derived from each of the *SXT* and *CYR* gene clusters and in particular, sequences relating to the specific genes within each pathway. Methods and kits for the detection of cyanobacterial strains in a sample are provided based on the presence (or absence) in the sample of one or more of the sequences of the invention. The inventors have determined that certain open-reading frames present in the *SXT* gene cluster of saxitoxin-producing
25 microorganisms are absent in the *SXT* gene cluster of microorganisms that do not produce saxitoxin. Similarly, it has been discovered that one open-reading frame present in the *CYR* gene cluster of cylindrospermopsin-producing microorganisms is absent in non-cylindrospermopsin-producing microorganisms. Accordingly, the invention provides methods and kits for the detection of toxin-producing microorganisms.

Also provided by the invention are screening methods for the identification of
30 compounds capable of modulating the expression or activity of proteins in the saxitoxin and/or cylindrospermopsin biosynthetic pathways.

Polynucleotides and polypeptides

The inventors have determined the full polynucleotide sequence of the saxitoxin (*SXT*) gene cluster and the cylindrospermopsin (*CYR*) gene cluster.

In accordance with aspects and embodiments of the invention, the *SXT* gene cluster may have, but is not limited to, the polynucleotide sequence as set forth SEQ ID NO: 1 (GenBank accession number DQ787200), or display sufficient sequence identity thereto to hybridise to the sequence of SEQ ID NO: 1.

The *SXT* gene cluster comprises 31 genes and 30 intergenic regions.

Gene 1 of the *SXT* gene cluster is a 759 base pair (bp) nucleotide sequence set forth in SEQ ID NO: 4. The nucleotide sequence of *SXT* Gene 1 ranges from the nucleotide in position 1625 up to the nucleotide in position 2383 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 1 (*SXTD*) is set forth in SEQ ID NO: 5.

Gene 2 of the *SXT* gene cluster is a 396 bp nucleotide sequence set forth in SEQ ID NO: 6. The nucleotide sequence of *SXT* Gene 2 ranges from the nucleotide in position 2621 up to the nucleotide in position 3016 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 2 (*ORF3*) is set forth in SEQ ID NO: 7.

Gene 3 of the *SXT* gene cluster is a 360 bp nucleotide sequence set forth in SEQ ID NO: 8. The nucleotide sequence of *SXT* Gene 3 ranges from the nucleotide in position 2955 up to the nucleotide in position 3314 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 3 (*ORF4*) is set forth in SEQ ID NO: 9.

Gene 4 of the *SXT* gene cluster is a 354 bp nucleotide sequence set forth in SEQ ID NO: 10. The nucleotide sequence of *SXT* Gene 4 ranges from the nucleotide in position 3647 up to the nucleotide in position 4000 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 4 (*SXTC*) is set forth in SEQ ID NO: 11.

Gene 5 of the *SXT* gene cluster is a 957 bp nucleotide sequence set forth in SEQ ID NO: 12. The nucleotide sequence of *SXT* Gene 5 ranges from the nucleotide in position 4030 up to the nucleotide in position 4986 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 5 (*SXTB*) is set forth in SEQ ID NO: 13.

Gene 6 of the *SXT* gene cluster is a 3738 bp nucleotide sequence set forth in SEQ ID NO: 14. The nucleotide sequence of *SXT* Gene 6 ranges from the nucleotide in position 5047 up to the nucleotide in position 8784 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 6 (*SXTA*) is set forth in SEQ ID NO: 15.

Gene 7 of the *SXT* gene cluster is a 387 bp nucleotide sequence set forth in SEQ ID NO: 16. The nucleotide sequence of *SXT* Gene 7 ranges from the nucleotide in position

9140 up to the nucleotide in position 9526 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 7 (*SXTE*) is set forth in SEQ ID NO: 17.

Gene 8 of the *SXT* gene cluster is a 1416 bp nucleotide sequence set forth in SEQ ID NO: 18. The nucleotide sequence of *SXT* Gene 8 ranges from the nucleotide in position 9686 up to the nucleotide in position 11101 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 8 (*SXTF*) is set forth in SEQ ID NO: 19.

Gene 9 of the *SXT* gene cluster is an 1134 bp nucleotide sequence set forth in SEQ ID NO: 20. The nucleotide sequence of *SXT* Gene 9 ranges from the nucleotide in position 11112 up to the nucleotide in position 12245 of SEQ ID NO: 1. The polypeptide sequence encoded by *SXT* Gene 9 (*SXTG*) is set forth in SEQ ID NO: 21.

Gene 10 of the *SXT* gene cluster is a 1005 bp nucleotide sequence set forth in SEQ ID NO: 22. The nucleotide sequence of *SXT* Gene 10 ranges from the nucleotide in position 12314 up to the nucleotide in position 13318 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 10 (*SXTH*) is set forth in SEQ ID NO: 23.

Gene 11 of the *SXT* gene cluster is an 1839 bp nucleotide sequence set forth in SEQ ID NO: 24. The nucleotide sequence of *SXT* Gene 11 ranges from the nucleotide in position 13476 up to the nucleotide in position 15314 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 11 (*SXTI*) is set forth in SEQ ID NO: 25.

Gene 12 of the *SXT* gene cluster is a 444 bp nucleotide sequence set forth in SEQ ID NO: 26. The nucleotide sequence of *SXT* Gene 12 ranges from the nucleotide in position 15318 up to the nucleotide in position 15761 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 12 (*SXTJ*) is set forth in SEQ ID NO: 27.

Gene 13 of the *SXT* gene cluster is a 165 bp nucleotide sequence set forth in SEQ ID NO: 28. The nucleotide sequence of *SXT* Gene 13 ranges from the nucleotide in position 15761 up to the nucleotide in position 15925 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 13 (*SXTK*) is set forth in SEQ ID NO: 29.

Gene 14 of the *SXT* gene cluster is a 1299 bp nucleotide sequence set forth in SEQ ID NO: 30. The nucleotide sequence of *SXT* Gene 14 ranges from the nucleotide in position 15937 up to the nucleotide in position 17235 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 14 (*SXTL*) is set forth in SEQ ID NO: 31.

Gene 15 of the *SXT* gene cluster is a 1449 bp nucleotide sequence set forth in SEQ ID NO: 32. The nucleotide sequence of *SXT* Gene 15 ranges from the nucleotide in position 17323 up to the nucleotide in position 18771 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 16 (*SXTM*) is set forth in SEQ ID NO: 33.

Gene 16 of the *SXT* gene cluster is an 831 bp nucleotide sequence set forth in SEQ ID NO: 34. The nucleotide sequence of *SXT* Gene 16 ranges from the nucleotide in position 19119 up to the nucleotide in position 19949 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 16 (*SXTN*) is set forth in SEQ ID NO: 35.

5 Gene 17 of the *SXT* gene cluster is a 774 bp nucleotide sequence set forth in SEQ ID NO: 36. The nucleotide sequence of *SXT* Gene 17 ranges from the nucleotide in position 20238 up to the nucleotide in position 21011 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 17 (*SXTX*) is set forth in SEQ ID NO: 37.

10 Gene 18 of the *SXT* gene cluster is a 327 bp nucleotide sequence set forth in SEQ ID NO: 38. The nucleotide sequence of *SXT* Gene 18 ranges from the nucleotide in position 21175 up to the nucleotide in position 21501 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 18 (*SXTW*) is set forth in SEQ ID NO: 39.

15 Gene 19 of the *SXT* gene cluster is a 1653 bp nucleotide sequence set forth in SEQ ID NO: 40. The nucleotide sequence of *SXT* Gene 219 ranges from the nucleotide in position 21542 up to the nucleotide in position 23194 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 19 (*SXTV*) is set forth in SEQ ID NO: 41.

20 Gene 20 of the *SXT* gene cluster is a 750 bp nucleotide sequence set forth in SEQ ID NO: 42. The nucleotide sequence of *SXT* Gene 20 ranges from the nucleotide in position 23199 up to the nucleotide in position 23948 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 20 (*SXTU*) is set forth in SEQ ID NO: 43.

 Gene 21 of the *SXT* gene cluster is a 1005 bp nucleotide sequence set forth in SEQ ID NO: 44. The nucleotide sequence of *SXT* Gene 21 ranges from the nucleotide in position 24091 up to the nucleotide in position 25095 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 21 (*SXTT*) is set forth in SEQ ID NO: 45.

25 Gene 22 of the *SXT* gene cluster is a 726 bp nucleotide sequence set forth in SEQ ID NO: 46. The nucleotide sequence of *SXT* Gene 22 ranges from the nucleotide in position 25173 up to the nucleotide in position 25898 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 22 (*SXTS*) is set forth in SEQ ID NO: 47.

30 Gene 23 of the *SXT* gene cluster is a 576 bp nucleotide sequence set forth in SEQ ID NO: 48. The nucleotide sequence of *SXT* Gene 23 ranges from the nucleotide in position 25974 up to the nucleotide in position 26549 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 23 (*ORF24*) is set forth in SEQ ID NO: 49.

 Gene 24 of the *SXT* gene cluster is a 777 bp nucleotide sequence set forth in SEQ ID NO: 50. The nucleotide sequence of *SXT* Gene 24 ranges from the nucleotide in

position 26605 up to the nucleotide in position 27381 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 24 (*SXTR*) is set forth in SEQ ID NO: 51.

Gene 25 of the *SXT* gene cluster is a 777 bp nucleotide sequence set forth in SEQ ID NO: 52. The nucleotide sequence of *SXT* Gene 25 ranges from the nucleotide in
5 position 27392 up to the nucleotide in position 28168 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 25 (*SXTQ*) is set forth in SEQ ID NO: 53.

Gene 26 of the *SXT* gene cluster is a 1227 bp nucleotide sequence set forth in SEQ ID NO: 54. The nucleotide sequence of *SXT* Gene 26 ranges from the nucleotide in position 28281 up to the nucleotide in position 29507 of SEQ ID NO: 1. The polypeptide
10 sequence encoded by Gene 26 (*SXTP*) is set forth in SEQ ID NO: 55.

Gene 27 of the *SXT* gene cluster is a 603 bp nucleotide sequence set forth in SEQ ID NO: 56. The nucleotide sequence of *SXT* Gene 27 ranges from the nucleotide in position 29667 up to the nucleotide in position 30269 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 27 (*SXTO*) is set forth in SEQ ID NO: 57.

15 Gene 28 of the *SXT* gene cluster is a 1350 bp nucleotide sequence set forth in SEQ ID NO: 58. The nucleotide sequence of *SXT* Gene 28 ranges from the nucleotide in position 30612 up to the nucleotide in position 31961 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 28 (*ORF29*) is set forth in SEQ ID NO: 59.

Gene 29 of the *SXT* gene cluster is a 666 bp nucleotide sequence set forth in SEQ
20 ID NO: 60. The nucleotide sequence of *SXT* Gene 29 ranges from the nucleotide in position 32612 up to the nucleotide in position 33277 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 29 (*SXTY*) is set forth in SEQ ID NO: 61.

Gene 30 of the *SXT* gene cluster is a 1353 bp nucleotide sequence set forth in SEQ ID NO: 62. The nucleotide sequence of *SXT* Gene 30 ranges from the nucleotide in
25 position 33325 up to the nucleotide in position 34677 of SEQ ID NO: 1. The polypeptide sequence encoded by Gene 30 (*SXTZ*) is set forth in SEQ ID NO: 63.

Gene 31 of the *SXT* gene cluster is an 819 bp nucleotide sequence set forth in SEQ ID NO: 64. The nucleotide sequence of *SXT* Gene 31 ranges from the nucleotide in position 35029 up to the nucleotide in position 35847 of SEQ ID NO: 1. The polypeptide
30 sequence encoded by Gene 31 (*OMPR*) is set forth in SEQ ID NO: 65.

The 5' border region of *SXT* gene cluster comprises a 1320 bp gene (*orf1*), the sequence of which is set forth in SEQ ID NO: 2. The nucleotide sequence of *orf1* ranges from the nucleotide in position 1 up to the nucleotide in position 1320 of SEQ ID NO: 1. The polypeptide sequence encoded by *orf1* is set forth in SEQ ID NO: 3.

The 3' border region of *SXT* gene cluster comprises a 774 bp gene (*hisA*), the sequence of which is set forth in SEQ ID NO: 66. The nucleotide sequence of *hisA* ranges from the nucleotide in position 35972 up to the nucleotide in position 36745 of SEQ ID NO: 1. The polypeptide sequence encoded by *hisA* is set forth in SEQ ID NO: 67.

5 The 3' border region of *SXT* gene cluster also comprises a 396 bp gene (*orfA*), the sequence of which is set forth in in SEQ ID NO: 68. The nucleotide sequence of *orfA* ranges from the nucleotide in position 37060 up to the nucleotide in position 37455 of SEQ ID NO: 1. The polypeptide sequence encoded by *orfA* is set forth in SEQ ID NO: 69.

10 In accordance with other aspects and embodiments of the invention, the *CYR* gene cluster may have, but is not limited to, the nucleotide sequence as set forth SEQ ID NO: 80 (GenBank accession number EU140798), or display sufficient sequence identity thereto to hybridise to the sequence of SEQ ID NO: 80.

The *CYR* gene cluster comprises 15 genes and 14 intergenic regions.

15 Gene 1 of the *CYR* gene cluster is a 5631 bp nucleotide sequence set forth in SEQ ID NO: 81. The nucleotide sequence of *CYR* Gene 1 ranges from the nucleotide in position 444 up to the nucleotide in position 6074 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 1 (*CYRD*) is set forth in SEQ ID NO: 82.

20 Gene 2 of the *CYR* gene cluster is a 4074 bp nucleotide sequence set forth in SEQ ID NO: 83. The nucleotide sequence of *CYR* Gene 2 ranges from the nucleotide in position 6130 up to the nucleotide in position 10203 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 2 (*CYRF*) is set forth in SEQ ID NO: 84.

25 Gene 3 of the *CYR* gene cluster is a 1437 bp nucleotide sequence set forth in SEQ ID NO: 85. The nucleotide sequence of *CYR* Gene 3 ranges from the nucleotide in position 10251 up to the nucleotide in position 11687 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 3 (*CYRG*) is set forth in SEQ ID NO: 86.

Gene 4 of the *CYR* gene cluster is an 831 bp nucleotide sequence set forth in SEQ ID NO: 87. The nucleotide sequence of *CYR* Gene 4 ranges from the nucleotide in position 11741 up to the nucleotide in position 12571 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 4 (*CYRI*) is set forth in SEQ ID NO: 88.

30 Gene 5 of the *CYR* gene cluster is a 1398 bp nucleotide sequence set forth in SEQ ID NO: 89. The nucleotide sequence of *CYR* Gene 5 ranges from the nucleotide in position 12568 up to the nucleotide in position 13965 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 5 (*CYRK*) is set forth in SEQ ID NO: 90.

Gene 6 of the *CYR* gene cluster is a 750 bp nucleotide sequence set forth in SEQ ID NO: 91. The nucleotide sequence of *CYR* Gene 6 ranges from the nucleotide in position 14037 up to the nucleotide in position 14786 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 6 (*CYRL*) is set forth in SEQ ID NO: 92.

5 Gene 7 of the *CYR* gene cluster is a 1431 bp nucleotide sequence set forth in SEQ ID NO: 93. The nucleotide sequence of *CYR* Gene 7 ranges from the nucleotide in position 14886 up to the nucleotide in position 16316 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 7 (*CYRH*) is set forth in SEQ ID NO: 94.

10 Gene 8 of the *CYR* gene cluster is a 780 bp nucleotide sequence set forth in SEQ ID NO: 95. The nucleotide sequence of *CYR* Gene 8 ranges from the nucleotide in position 16893 up to the nucleotide in position 17672 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 8 (*CYRJ*) is set forth in SEQ ID NO: 96.

15 Gene 9 of the *CYR* gene cluster is an 1176 bp nucleotide sequence set forth in SEQ ID NO: 97. The nucleotide sequence of *CYR* Gene 9 ranges from the nucleotide in position 18113 up to the nucleotide in position 19288 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 9 (*CYR4*) is set forth in SEQ ID NO: 98.

20 Gene 10 of the *CYR* gene cluster is an 8754 bp nucleotide sequence set forth in SEQ ID NO: 99. The nucleotide sequence of *CYR* Gene 10 ranges from the nucleotide in position 19303 up to the nucleotide in position 28056 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 10 (*CYRB*) is set forth in SEQ ID NO: 100.

 Gene 11 of the *CYR* gene cluster is a 5667 bp nucleotide sequence set forth in SEQ ID NO: 101. The nucleotide sequence of *CYR* Gene 11 ranges from the nucleotide in position 28061 up to the nucleotide in position 33727 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 11 (*CYRE*) is set forth in SEQ ID NO: 102.

25 Gene 12 of the *CYR* gene cluster is a 5004 bp nucleotide sequence set forth in SEQ ID NO: 103. The nucleotide sequence of *CYR* Gene 12 ranges from the nucleotide in position 34299 up to the nucleotide in position 39302 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 12 (*CYRC*) is set forth in SEQ ID NO: 104.

30 Gene 13 of the *CYR* gene cluster is a 318 bp nucleotide sequence set forth in SEQ ID NO: 105. The nucleotide sequence of *CYR* Gene 13 ranges from the nucleotide in position 39366 up to the nucleotide in position 39683 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 13 (*CYRM*) is set forth in SEQ ID NO: 106.

 Gene 14 of the *CYR* gene cluster is a 600 bp nucleotide sequence set forth in SEQ ID NO: 107. The nucleotide sequence of *CYR* Gene 14 ranges from the nucleotide in

position 39793 up to the nucleotide in position 40392 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 14 (*CYRM*) is set forth in SEQ ID NO: 108.

Gene 15 of the *CYR* gene cluster is a 1548 bp nucleotide sequence set forth in SEQ ID NO: 109. The nucleotide sequence of *CYR* Gene 15 ranges from the nucleotide in position 40501 up to the nucleotide in position 42048 of SEQ ID NO: 80. The polypeptide sequence encoded by Gene 15 (*CYRO*) is set forth in SEQ ID NO: 110.

In general, the nucleic acids and polypeptides of the invention are of an isolated or purified form.

In addition to the *SXT* and *CYR* polynucleotides and polypeptide sequences set forth herein, also included within the scope of the present invention are variants and fragments thereof.

SXT and *CYR* polynucleotides disclosed herein may be deoxyribonucleic acids (DNA), ribonucleic acids (RNA) or complementary deoxyribonucleic acids (cDNA).

RNA may be derived from RNA polymerase-catalyzed transcription of a DNA sequence. The RNA may be a primary transcript derived transcription of a corresponding DNA sequence. RNA may also undergo post-transcriptional processing. For example, a primary RNA transcript may undergo post-transcriptional processing to form a mature RNA. Messenger RNA (mRNA) refers to RNA derived from a corresponding open reading frame that may be translated into protein by the cell. cDNA refers to a double-stranded DNA that is complementary to and derived from mRNA. Sense RNA refers to RNA transcript that includes the mRNA and so can be translated into protein by the cell. Antisense RNA refers to an RNA transcript that is complementary to all or part of a target primary transcript or mRNA and may be used to block the expression of a target gene.

The skilled addressee will recognise that RNA and cDNA sequences encoded by the *SXT* and *CYR* DNA sequences disclosed herein may be derived using the genetic code. An RNA sequence may be derived from a given DNA sequence by generating a sequence that is complementary the particular DNA sequence. The complementary sequence may be generated by converting each cytosine ('C') base in the DNA sequence to a guanine ('G') base, each guanine ('G') base in the DNA sequence to a cytosine ('C') base, each thymidine ('T') base in the DNA sequence to an adenine ('A') base, and each adenine ('A') base in the DNA sequence to a uracil ('U') base.

A complementary DNA (cDNA) sequence may be derived from a DNA sequence by deriving an RNA sequence from the DNA sequence as above, then converting the RNA sequence into a cDNA sequence. An RNA sequence can be converted into a cDNA

sequence by converting each cytosine ('C') base in the RNA sequence to a guanine ('G') base, each guanine ('G') base in the RNA sequence to a cytosine ('C') base, each uracil ('U') base in the RNA sequence to an adenine ('A') base, and each adeneine ('A') base in the RNA sequence to a thymidine ('T') base.

5 The term "variant" as used herein refers to a substantially similar sequence. In general, two sequences are "substantially similar" if the two sequences have a specified percentage of amino acid residues or nucleotides that are the same (percentage of "sequence identity"), over a specified region, or, when not specified, over the entire sequence. Accordingly, a "variant" of a polynucleotide and polypeptide sequence
10 disclosed herein may share at least 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 83% 85%, 88%, 90%, 93%, 95%, 96%, 97%, 98% or 99% sequence identity with the reference sequence.

 In general, polypeptide sequence variants possess qualitative biological activity in common. Polynucleotide sequence variants generally encode polypeptides which
15 generally possess qualitative biological activity in common. Also included within the meaning of the term "variant" are homologues of polynucleotides and polypeptides of the invention. A polynucleotide homologue is typically from a different bacterial species but sharing substantially the same biological function or activity as the corresponding polynucleotide disclosed herein. A polypeptide homologue is typically from a different
20 bacterial species but sharing substantially the same biological function or activity as the corresponding polypeptide disclosed herein. For example, homologues of the polynucleotides and polypeptides disclosed herein include, but are not limited to those from different species of cyanobacteria.

 Further, the term "variant" also includes analogues of the polypeptides of the
25 invention. A polypeptide "analogue" is a polypeptide which is a derivative of a polypeptide of the invention, which derivative comprises addition, deletion, substitution of one or more amino acids, such that the polypeptide retains substantially the same function. The term "conservative amino acid substitution" refers to a substitution or replacement of one amino acid for another amino acid with similar properties within a
30 polypeptide chain (primary sequence of a protein). For example, the substitution of the charged amino acid glutamic acid (Glu) for the similarly charged amino acid aspartic acid (Asp) would be a conservative amino acid substitution.

 In general, the percentage of sequence identity between two sequences may be determined by comparing two optimally aligned sequences over a comparison window.

The portion of the sequence in the comparison window may, for example, comprise deletions or additions (*i.e.* gaps) in comparison to the reference sequence (for example, a polynucleotide or polypeptide sequence disclosed herein), which does not comprise deletions or additions, in order to align the two sequences optimally. A percentage of sequence identity may then be calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

10 In the context of two or more nucleic acid or polypeptide sequences, the percentage of sequence identity refers to the specified percentage of amino acid residues or nucleotides that are the same over a specified region, (or, when not specified, over the entire sequence), when compared and aligned for maximum correspondence over a comparison window, or designated region as measured using one of the following sequence comparison algorithms or by manual alignment and visual inspection.

For sequence comparison, typically one sequence acts as a reference sequence, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are entered into a computer, subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. Default program parameters can be used, or alternative parameters can be designated. The sequence comparison algorithm then calculates the percent sequence identities for the test sequences relative to the reference sequence, based on the program parameters. Methods of alignment of sequences for comparison are well known in the art. Optimal alignment of sequences for comparison can be determined conventionally using known computer programs, including, but not limited to: CLUSTAL in the PC/Gene program (available from Intelligenetics, Mountain View, California); the ALIGN program (Version 2.0) and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the GCG Wisconsin Genetics Software Package, Version 10 (available from Accelrys Inc., 9685 Scranton Road, San Diego, California, USA).

30 The BESTFIT program (Wisconsin Sequence Analysis Package, for Unix, Genetics Computer Group, University Research Park, 575 Science Drive, Madison, Wis. 53711) uses the local homology algorithm of Smith and Waterman to find the best segment of homology between two sequences (Smith and Waterman, *Advances in Applied Mathematics* 2:482-489 (1981)). When using BESTFIT or any other sequence alignment

program to determine the degree of homology between sequences, the parameters may be set such that the percentage of identity is calculated over the full length of the reference nucleotide sequence and that gaps in homology of up to 5% of the total number of nucleotides in the reference sequence are allowed.

5 GAP uses the algorithm described in Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453, to find the alignment of two complete sequences that maximizes the number of matches and minimizes the number of gaps. GAP considers all possible alignments and gap positions and creates the alignment with the largest number of matched bases and the fewest gaps. It allows for the provision of a gap creation penalty and a gap extension
10 penalty in units of matched bases. GAP presents one member of the family of best alignments.

Another method for determining the best overall match between a query sequence and a subject sequence, also referred to as a global sequence alignment, can be determined using the FASTDB computer program based on the algorithm of Brutlag and
15 colleagues (*Comp. App. Biosci.* 6:237-245 (1990)). In a sequence alignment the query and subject sequences are both DNA sequences. An RNA sequence can be compared by converting U's to T's. The result of said global sequence alignment is in percent identity.

The BLAST and BLAST 2.0 algorithms, may be used for determining percent sequence identity and sequence similarity. These are described in Altschul et al. (1977)
20 *Nuc. Acids Res.* 25:3389-3402, and Altschul et al (1990) *J. Mol. Biol.* 215:403-410, respectively. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when
25 aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul et al, *supra*). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide
30 sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the

accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, M=5, N=-4 and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength of 3, and expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1989) Proc. Natl. Acad. Sci USA 89:10915) alignments (B) of 50, expectation (E) of 10, M=5, N=-4, and a comparison of both strands. [0028] The BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, e.g., Karlin and Altschul (1993) Proc. Natl. Acad. Sci. USA 90:5873-5787). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.2, more preferably less than about 0.01, and most preferably less than about 0.001.

The invention also contemplates fragments of the polypeptides disclosed herein. A polypeptide "fragment" is a polypeptide molecule that encodes a constituent or is a constituent of a polypeptide of the invention or variant thereof. Typically the fragment possesses qualitative biological activity in common with the polypeptide of which it is a constituent. The peptide fragment may be between about 5 to about 3000 amino acids in length, between about 5 to about 2750 amino acids in length, between about 5 to about 2500 amino acids in length, between about 5 to about 2250 amino acids in length, between about 5 to about 2000 amino acids in length, between about 5 to about 1750 amino acids in length, between about 5 to about 1500 amino acids in length, between about 5 to about 1250 amino acids in length, between about 5 to about 1000 amino acids in length, between about 5 to about 900 amino acids in length, between about 5 to about 800 amino acids in length, between about 5 to about 700 amino acids in length, between about 5 to about 600 amino acids in length, between about 5 to about 500 amino acids in length, between about 5 to about 450 amino acids in length, between about 5 to about 400 amino acids in length, between about 5 to about 350 amino acids in length, between about 5 to about 300 amino acids in length, between about 5 to about 250 amino acids in length, between about 5 to about 200 amino acids in length, between about 5 to about 175 amino acids in length, between about 5 to about 150 amino acids in length, between about 5 to

about 125 amino acids in length, between about 5 to about 100 amino acids in length, between about 5 to about 75 amino acids in length, between about 5 to about 50 amino acids in length, between about 5 to about 40 amino acids in length, between about 5 to about 30 amino acids in length, between about 5 to about 20 amino acids in length, and
5 between about 5 to about 15 amino acids in length. Alternatively, the peptide fragment may be between about 5 to about 10 amino acids in length.

Also contemplated are fragments of the polynucleotides disclosed herein. A polynucleotide "fragment" is a polynucleotide molecule that encodes a constituent or is a constituent of a polynucleotide of the invention or variant thereof. Fragments of a
10 polynucleotide do not necessarily need to encode polypeptides which retain biological activity. The fragment may, for example, be useful as a hybridization probe or PCR primer. The fragment may be derived from a polynucleotide of the invention or alternatively may be synthesized by some other means, for example by chemical synthesis.

15 Certain embodiments of the invention relate to fragments of SEQ ID NO: 1. A fragment of SEQ ID NO: 1 may comprise, for example, a constituent of SEQ ID NO: 1 in which the 5' gene border region gene *orf1* is absent. Alternatively, a fragment of SEQ ID NO: 1 may comprise, for example, a constituent of SEQ ID NO: 1 in which the 3' gene border region gene *hisA* is absent. Alternatively, a fragment of SEQ ID NO: 1 may
20 comprise, for example, a constituent of SEQ ID NO: 1 in which the 3' gene border region gene *orfA* is absent. Alternatively, a fragment of SEQ ID NO: 1 may comprise, for example, a constituent of SEQ ID NO: 1 in which the 5' gene border region gene *orf1* is absent and the 3' border region gene *orfA* is absent. Alternatively, a fragment of SEQ ID NO: 1 may comprise, for example, a constituent of SEQ ID NO: 1 in which the 5' gene
25 border region gene *orf1* is absent and the 3' border region genes *hisA* and *orfA* are absent.

In other embodiments, a fragment of SEQ ID NO: 1 may comprise one or more *SXT* open reading frames. The *SXT* open reading frame may be selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID

NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants thereof.

Additional embodiments of the invention relate to fragments of SEQ ID NO: 80. The fragment of SEQ ID NO: 80 may comprise one or more *CYR* open reading frames. The *CYR* open reading frame may be selected from the group consisting of of SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants thereof.

In particular embodiments, the polynucleotides of the invention may be cloned into a vector. The vector may comprise, for example, a DNA, RNA or complementary DNA (cDNA) sequence. The vector may be a plasmid vector, a viral vector, or any other suitable vehicle adapted for the insertion of foreign sequences, their introduction into cells and the expression of the introduced sequences. Typically the vector is an expression vector and may include expression control and processing sequences such as a promoter, an enhancer, ribosome binding sites, polyadenylation signals and transcription termination sequences. The invention also contemplates host cells transformed by such vectors. For example, the polynucleotides of the invention may be cloned into a vector which is transformed into a bacterial host cell, for example *E. coli*. Methods for the construction of vectors and their transformation into host cells are generally known in the art, and described in, for example, *Molecular Cloning: A Laboratory Manual* (2nd ed., Cold Spring Harbor Laboratory Press, Plainview, New York, and, Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc.

Nucleotide Probes, Primers and Antibodies

The invention contemplates nucleotides and fragments based on the sequences of the polynucleotides disclosed herein for use as primers and probes for the identification of homologous sequences.

The nucleotides and fragments may be in the form of oligonucleotides. Oligonucleotides are short stretches of nucleotide residues suitable for use in nucleic acid amplification reactions such as PCR, typically being at least about 5 nucleotides to about 80 nucleotides in length, more typically about 10 nucleotides in length to about 50 nucleotides in length, and even more typically about 15 nucleotides in length to about 30 nucleotides in length.

Probes are nucleotide sequences of variable length, for example between about 10 nucleotides and several thousand nucleotides, for use in detection of homologous sequences, typically by hybridization. Hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides.

5 Methods for the design and/or production of nucleotide probes and/or primers are generally known in the art, and are described in Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2nd ed., Cold Spring Harbor Laboratory Press, Plainview, New York; Itakura K. et al. (1984) *Annu. Rev. Biochem.* 53:323; Innis et al., (Eds) (1990) *PCR Protocols: A Guide to Methods and Applications* (Academic Press, 10 New York); Innis and Gelfand, (Eds) (1995) *PCR Strategies* (Academic Press, New York); and Innis and Gelfand, (Eds) (1999) *PCR Methods Manual* (Academic Press, New York). Nucleotide primers and probes may be prepared, for example, by chemical synthesis techniques for example, the phosphodiester and phosphotriester methods (see for example Narang S. A. et al. (1979) *Meth. Enzymol.* 68:90; Brown, E. L. (1979) et al. 15 *Meth. Enzymol.* 68:109; and U.S. Patent No. 4356270), the diethylphosphoramidite method (see Beaucage S.L et al. (1981) *Tetrahedron Letters*, 22:1859-1862). A method for synthesizing oligonucleotides on a modified solid support is described in U.S. Patent No. 4458066.

20 The nucleic acids of the invention, including the above-mentioned probes and primers, may be labelled by incorporation of a marker to facilitate their detection. Techniques for labelling and detecting nucleic acids are described, for example, in Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc. Examples of suitable markers include fluorescent molecules (e.g. acetylaminofluorene, 5-bromodeoxyuridine, digoxigenin, fluorescein) and radioactive 25 isotopes (e.g. ³²P, ³⁵S, ³H, ³³P). Detection of the marker may be achieved, for example, by chemical, photochemical, immunochemical, biochemical, or spectroscopic techniques.

30 The probes and primers of the invention may be used, for example, to detect or isolate cyanobacteria and/or dinoflagellates in a sample of interest. Additionally or alternatively, the probes and primers of the invention may be used to detect or isolate a cyanotoxic organism and/or a cylindrospermopsis-producing organism in a sample of interest. Additionally or alternatively, the probes or primers of the invention may be used to isolate corresponding sequences in other organisms including, for example, other bacterial species. Methods such as the polymerase chain reaction (PCR), hybridization, and the like can be used to identify such sequences based on their sequence homology to

the sequences set forth herein. Sequences that are selected based on their sequence identity to the entire sequences set forth herein or to fragments thereof are encompassed by the embodiments. Such sequences include sequences that are orthologs of the disclosed sequences. The term "orthologs" refers to genes derived from a common
5 ancestral gene and which are found in different species as a result of speciation. Genes found in different species are considered orthologs when their nucleotide sequences and/or their encoded protein sequences share substantial identity as defined elsewhere herein. Functions of orthologs are often highly conserved among species.

In hybridization techniques, all or part of a known nucleotide sequence is used to
10 generate a probe that selectively hybridizes to other corresponding nucleic acid sequences present in a given sample. The hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides, and may be labelled with a detectable marker. Thus, for example, probes for hybridization can be made by labelling synthetic oligonucleotides based on the sequences of the invention.

15 The level of homology (sequence identity) between probe and the target sequence will largely be determined by the stringency of hybridization conditions. In particular the nucleotide sequence used as a probe may hybridize to a homologue or other variant of a polynucleotide disclosed herein under conditions of low stringency, medium stringency or high stringency. There are numerous conditions and factors, well known to those skilled
20 in the art, which may be employed to alter the stringency of hybridization. For instance, the length and nature (DNA, RNA, base composition) of the nucleic acid to be hybridized to a specified nucleic acid; concentration of salts and other components, such as the presence or absence of formamide, dextran sulfate, polyethylene glycol etc; and altering the temperature of the hybridization and/or washing steps.

25 Typically, stringent hybridization conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (*e.g.*, 10 to 50 nucleotides) and at least about 60°C for long probes (*e.g.*, greater than 50 nucleotides). Stringent conditions may also be achieved with the addition
30 of destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30% to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulfate) at 37 °C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50°C to 55 °C. Exemplary moderate stringency conditions include hybridization in 40% to 45% formamide, 1.0 M NaCl, 1% SDS at

37 °C, and a wash in 0.5X to 1X SSC at 55°C to 60 °C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37 °C, and a final wash in 0.1X SSC at 60°C to 65 °C for at least about 20 minutes. Optionally, wash buffers may comprise about 0.1% to about 1% SDS. The duration of hybridization is
5 generally less than about 24 hours, usually about 4 to about 12 hours.

Under a PCR approach, oligonucleotide primers can be designed for use in PCR reactions to amplify corresponding DNA sequences from cDNA or genomic DNA extracted from any organism of interest. Methods for designing PCR primers and PCR cloning are generally known in the art and are disclosed in Sambrook *et al.* (1989)
10 *Molecular Cloning: A Laboratory Manual* (2nd ed., Cold Spring Harbor Laboratory Press, Plainview, New York); Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc; Maniatis et al. *Molecular Cloning* (1982), 280-281; Innis et al. (Eds) (1990) *PCR Protocols: A Guide to Methods and Applications* (Academic Press, New York); Innis and Gelfand, (Eds) (1995) *PCR Strategies* (Academic
15 Press, New York); and Innis and Gelfand, (Eds) (1999) *PCR Methods Manual* (Academic Press, New York). Known methods of PCR include, but are not limited to, methods using paired primers, nested primers, single specific primers, degenerate primers, gene-specific primers, vector-specific primers, partially-mismatched primers, and the like.

The skilled addressee will recognise that the primers described herein for use in
20 PCR or RT-PCR may also be used as probes for the detection of *SXT* or *CYR* sequences.

Also contemplated by the invention are antibodies which are capable of binding specifically to the polypeptides of the invention. The antibodies may be used to qualitatively or quantitatively detect and analyse one or more *SXT* or *CYR* polypeptides in a given sample. By "binding specifically" it will be understood that the antibody is
25 capable of binding to the target polypeptide or fragment thereof with a higher affinity than it binds to an unrelated protein. For example, the antibody may bind to the polypeptide or fragment thereof with a binding constant in the range of at least about 10^{-4} M to about 10^{-10} M. Preferably the binding constant is at least about 10^{-5} M, or at least about 10^{-6} M, more preferably the binding constant of the antibody to the *SXT* or *CYR*
30 polypeptide or fragment thereof is at least about 10^{-7} M, at least about 10^{-8} M, or at least about 10^{-9} M or more.

Antibodies of the invention may exist in a variety of forms, including for example as a whole antibody, or as an antibody fragment, or other immunologically active fragment thereof, such as complementarity determining regions. Similarly, the antibody

may exist as an antibody fragment having functional antigen-binding domains, that is, heavy and light chain variable domains. Also, the antibody fragment may exist in a form selected from the group consisting of, but not limited to: Fv, F_{ab}, F(ab)₂, scFv (single chain Fv), dAb (single domain antibody), chimeric antibodies, bi-specific antibodies, 5 diabodies and triabodies.

An antibody 'fragment' may be produced by modification of a whole antibody or by synthesis of the desired antibody fragment. Methods of generating antibodies, including antibody fragments, are known in the art and include, for example, synthesis by recombinant DNA technology. The skilled addressee will be aware of methods of 10 synthesising antibodies, such as those described in, for example, US Patent No. 5296348 and Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc.

Preferably antibodies are prepared from discrete regions or fragments of the *SXT* or *CYR* polypeptide of interest. An antigenic portion of a polypeptide of interest may be of 15 any appropriate length, such as from about 5 to about 15 amino acids. Preferably, an antigenic portion contains at least about 5, 6, 7, 8, 9, 10, 11, 12, 13 or 14 amino acid residues.

In the context of this specification reference to an antibody specific to a *SXT* or *CYR* polypeptide of the invention includes an antibody that is specific to a fragment of the 20 polypeptide of interest.

Antibodies that specifically bind to a polypeptide of the invention can be prepared, for example, using the purified *SXT* or *CYR* polypeptides or their nucleic acid sequences using any suitable methods known in the art. For example, a monoclonal antibody, typically containing Fab portions, may be prepared using hybridoma technology 25 described in Harlow and Lane (Eds) *Antibodies - A Laboratory Manual*, (1988), Cold Spring Harbor Laboratory, N.Y; Coligan, *Current Protocols in Immunology* (1991); Goding, *Monoclonal Antibodies: Principles and Practice* (1986) 2nd ed; and Kohler & Milstein, (1975) *Nature* 256: 495-497. Such techniques include, but are not limited to, antibody preparation by selection of antibodies from libraries of recombinant antibodies 30 in phage or similar vectors, as well as preparation of polyclonal and monoclonal antibodies by immunizing rabbits or mice (see, for example, Huse et al. (1989) *Science* 246: 1275-1281; Ward et al. (1989) *Nature* 341: 544-546).

It will also be understood that antibodies of the invention include humanised antibodies, chimeric antibodies and fully human antibodies. An antibody of the invention

may be a bi-specific antibody, having binding specificity to more than one antigen or epitope. For example, the antibody may have specificity for one or more *SXT* or *CYR* polypeptide or fragments thereof, and additionally have binding specificity for another antigen. Methods for the preparation of humanised antibodies, chimeric antibodies, fully human antibodies, and bispecific antibodies are known in the art and include, for example as described in United States Patent No. 6995243 issued February 7, 2006 to Garabedian, et al. and entitled "Antibodies that recognize and bind phosphorylated human glucocorticoid receptor and methods of using same".

Generally, a sample potentially comprising *SXT* or *CYR* polypeptides can be contacted with an antibody that specifically binds the *SXT* or *CYR* polypeptide or fragment thereof. Optionally, the antibody can be fixed to a solid support to facilitate washing and subsequent isolation of the complex, prior to contacting the antibody with a sample. Examples of solid supports include, for example, microtitre plates, beads, ticks, or microbeads. Antibodies can also be attached to a ProteinChip array or a probe substrate as described above.

Detectable labels for the identification of antibodies bound to the *SXT* or *CYR* polypeptides of the invention include, but are not limited to fluorochromes, fluorescent dyes, radiolabels, enzymes such as horse radish peroxidase, alkaline phosphatase and others commonly used in the art, and colorimetric labels including colloidal gold or coloured glass or plastic beads. Alternatively, the marker in the sample can be detected using an indirect assay, wherein, for example, a second, labelled antibody is used to detect bound marker-specific antibody.

Methods for detecting the presence of or measuring the amount of, an antibody-marker complex include, for example, detection of fluorescence, chemiluminescence, luminescence, absorbance, birefringence, transmittance, reflectance, or refractive index such as surface plasmon resonance, ellipsometry, a resonant mirror method, a grating coupler wave guide method or interferometry. Radio frequency methods include multipolar resonance spectroscopy. Electrochemical methods include amperometry and voltametry methods. Optical methods include imaging methods and non-imaging methods and microscopy.

Useful assays for detecting the presence of or measuring the amount of, an antibody-marker complex include, include, for example, enzyme-linked immunosorbent assay (ELISA), a radioimmune assay (RIA), or a Western blot assay. Such methods are described in, for example, Clinical Immunology (Stites & Terr, eds., 7th ed. 1991);

Methods in Cell Biology: Antibodies in Cell Biology, volume 37 (Asai, ed. 1993); and Harlow & Lane, supra.

5

Methods and kits for detection

The invention provides methods and kits for the detection and/or isolation of *SXT* nucleic acids and polypeptides. Also provided are methods and kits for the detection and/or isolation *CYR* nucleic acids and polypeptides.

10 In one aspect, the invention provides a method for the detection of cyanobacteria. The skilled addressee will understand that the detection of "cyanobacteria" encompasses the detection of one or more cyanobacteria. The method comprises obtaining a sample for use in the method, and detecting the presence of one or more *SXT* polynucleotides or polypeptides as disclosed herein, or a variant or fragment thereof. The presence of *SXT*
15 polynucleotides, polypeptides, or variants or fragments thereof, is indicative of cyanobacteria in the sample.

The *SXT* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO:
20 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and
25 variants and fragments thereof.

Alternatively, the *SXT* polynucleotide may be an RNA or cDNA encoded by a sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26,
30 SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66,

SEQ ID NO: 68, and variants and fragments thereof and/or polypeptides as disclosed herein, or a variant or fragment thereof.

The *SXT* polypeptide may comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, and variants and fragments thereof.

The inventors have determined that several genes of the *SXT* gene cluster exist in saxitoxin-producing organisms, and are absent in organisms with the *SXT* gene cluster that do not produce saxitoxin. Specifically, the inventors have identified that gene 6 (*sxtA*) (SEQ ID NO: 14), gene 9 (*sxtG*) (SEQ ID NO: 20), gene 10 (*sxtH*) (SEQ ID NO: 22), gene 11 (*sxtI*) (SEQ ID NO: 24) and gene 17 (*sxtX*) (SEQ ID NO: 36) of the *SXT* gene cluster are present only in organisms that produce saxitoxin.

Accordingly, in another aspect the invention provides a method of detecting a cyanotoxic organism. The method comprises obtaining a sample for use in the method, and detecting a cyanotoxic organism based on the detection of one or more *SXT* polynucleotides comprising a sequence set forth in SEQ ID NO: 14 (*sxtA*, gene 6), SEQ ID NO: 20 (*sxtG*, gene 9), SEQ ID NO: 22 (*sxtH*, gene 10), SEQ ID NO: 24 (*sxtI*, gene 11), SEQ ID NO: 36 (*sxtX*, gene 17), or variants or fragments thereof. Additionally or alternatively, a cyanotoxic organism may be detected based on the detection of an RNA or cDNA comprising a sequence encoded by SEQ ID NO: 14 (*sxtA*, gene 6), SEQ ID NO: 20 (*sxtG*, gene 9), SEQ ID NO: 22 (*sxtH*, gene 10), SEQ ID NO: 24 (*sxtI*, gene 11), SEQ ID NO: 36 (*sxtX*, gene 17), or variants or fragments thereof. Additionally or alternatively, a cyanotoxic organism may be detected based on the detection of one or more polypeptides comprising a sequence set forth in SEQ ID NO: 15 (*SXTA*), SEQ ID NO: 21 (*SXTG*), SEQ ID NO: 23 (*SXTH*), SEQ ID NO: 25 (*SXTI*), SEQ ID NO: 37 (*SXTX*), or variants or fragments thereof, in a sample suspected of comprising one or more cyanotoxic organisms. The cyanotoxic organism may be any organism capable of producing saxitoxin. In a preferred embodiment of the invention, the cyanotoxic organism is a cyanobacteria or a dinoflagellate.

In certain embodiments of the invention, the methods for detecting cyanobacteria or the methods for detecting cyanotoxic organisms may further comprise the detection of one or more *CYR* polynucleotides or *CYR* polypeptides as disclosed herein, or a variant or fragment thereof. The *CYR* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants or fragments thereof.

Alternatively, the *CYR* polynucleotide may be an RNA or cDNA encoded by a polynucleotide sequence selected from the group consisting of SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants or fragments thereof.

The *CYR* polypeptide may comprise a sequence selected from the group consisting of SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, and SEQ ID NO: 110, and variants or fragments thereof.

The inventors have determined gene 8 (*cyrJ*) (SEQ ID NO: 95) of the *CYR* gene cluster exists in cylindrospermopsin-producing organisms, and is absent in organisms with the *CYR* gene cluster that do not produce cylindrospermopsin. Accordingly, the methods for detecting cyanobacteria or the methods for detecting cyanotoxic organisms may further comprise the detection of a cylindrospermopsin-producing organism based on the detection of a *CYR* polynucleotide comprising a sequence set forth in SEQ ID NO: 95, or a variant or fragment thereof. Additionally or alternatively, the methods for detecting cyanobacteria or the methods for detecting cyanotoxic organisms may further comprise the detection of a cylindrospermopsin-producing organism based on the detection of an RNA or cDNA comprising a sequence encoded by SEQ ID NO: 95, or a variant or fragment thereof. Additionally or alternatively, the methods for detecting cyanobacteria or the methods for detecting cyanotoxic organisms may further comprise the detection of a cylindrospermopsin-producing organism based on the detection of a *CYR* polypeptide comprising a sequence set forth in SEQ ID NO: 96, or a variant or fragment thereof.

In another aspect, the invention provides a method for the detection of cyanobacteria. The skilled addressee will understand that the detection of "cyanobacteria" encompasses the detection of one or more cyanobacteria. The method comprises obtaining a sample for use in the method, and detecting the presence of one or more *CYR* polynucleotides or polypeptides as disclosed herein, or a variant or fragment thereof. The presence of *CYR* polynucleotides, polypeptides, or variants or fragments thereof, is indicative of cyanobacteria in the sample.

The *CYR* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109 and variants and fragments thereof.

Alternatively, the *CYR* polynucleotide may be an RNA or cDNA encoded by a sequence selected from the group consisting of SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109 and variants and fragments thereof.

The *CYR* polypeptide may comprise a sequence selected from the group consisting of SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, and SEQ ID NO: 110, and variants or fragments thereof.

In another aspect of the invention there is provided a method of detecting a cylindrospermopsin-producing organism based on the detection of *CYR* gene 8 (*cyrJ*). The method comprises obtaining a sample for use in the method, and detecting the presence of a *CYR* polynucleotide comprising a sequence set forth in SEQ ID NO: 95, or a variant or fragment thereof. Additionally or alternatively, the method for detecting a cylindrospermopsin-producing organism based on the detection of *CYR* gene 8 (*cyrJ*) may comprise the detection of an RNA or cDNA comprising a sequence encoded by SEQ ID NO: 95, or a variant or fragment thereof. Additionally or alternatively, the method for detecting a cylindrospermopsin-producing organism based on the detection of *CYR* gene 8 (*cyrJ*) may comprise the detection of a *CYR* polypeptide comprising a sequence set forth in SEQ ID NO: 96, or a variant or fragment thereof.

In certain embodiments of the invention, the methods for detecting cyanobacteria comprising the detection of *CYR* sequences or variants or fragments thereof further comprise the detection of one or more *SXT* polynucleotides or *SXT* polypeptides as disclosed herein, or a variant or fragment thereof.

5 The *SXT* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38,
10 SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof.

 Alternatively, the *SXT* polynucleotide may be an RNA or cDNA encoded by a
15 sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46,
20 SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof and/or polypeptides as disclosed herein, or a variant or fragment thereof.

 The *SXT* polypeptide may comprising an amino acid sequence selected from the
25 group consisting of SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID
30 NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, and variants and fragments thereof.

 In another aspect, the invention provides a method for the detection of dinoflagellates. The skilled addressee will understand that the detection of

“dinoflagellates” encompasses the detection of one or more dinoflagellates. The method comprises obtaining a sample for use in the method, and detecting the presence of one or more *SXT* polynucleotides or polypeptides as disclosed herein, or a variant or fragment thereof. The presence of *SXT* polynucleotides, polypeptides, or variants or fragments thereof, is indicative of dinoflagellates in the sample.

The *SXT* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof.

Alternatively, the *SXT* polynucleotide may be an RNA or cDNA encoded by a sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof and/or polypeptides as disclosed herein, or a variant or fragment thereof.

The *SXT* polypeptide may comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, and variants and fragments thereof.

A sample for use in accordance with the methods described herein may be suspected of comprising one or more cyanotoxic organisms. The cyanotoxic organisms may be one or more cyanobacteria and/or one or more dinoflagellates. Additionally or alternatively, a sample for use in accordance with the methods described herein may be suspected of comprising one more cyanobacteria and/or one or more dinoflagellates. A sample for use in accordance with the methods described herein may be a comparative or control sample, for example, a sample comprising a known concentration or density of a cyanobacteria and/or dinoflagellates, or a sample comprising one or more known species or strains of cyanobacteria and/or dinoflagellates.

A sample for use in accordance with the methods described herein may be derived from any source. For example, a sample may be an environmental sample. The environmental sample may be derived, for example, from salt water, fresh water or a blue-green algal bloom. Alternatively, the sample may be derived from a laboratory source, such as a culture, or a commercial source.

It will be appreciated by those in the art that the methods and kits disclosed herein are generally suitable for detecting any organisms in which the *SXT* and/or *CYR* gene clusters are present. Suitable cyanobacteria to which the methods of the invention are applicable may be selected from the orders Oscillatoriales, Chroococcales, Nostocales and Stigonematales. For example, the cyanobacteria may be selected from the genera *Anabaena*, *Nostoc*, *Microcystis*, *Planktothrix*, *Oscillatoria*, *Phormidium*, and *Nodularia*. For example, the cyanobacteria may be selected from the species *Cylindrospermopsis raciborskii* T3, *Cylindrospermopsis raciborskii* AWT205, *Aphanizomenon ovalisporum*, *Aphanizomenon flos-aquae*, *Aphanizomenon sp.*, *Umezakia natans*, *Raphidiopsis curvata*, *Anabaena bergii*, *Lyngbya wollei*, and *Anabaena circinalis*. Examples of suitable dinoflagellates to which the methods and kits of the invention are applicable may be selected from the genera *Alexandrium*, *Pyrodinium* and *Gymnodinium*. The methods and kits of the invention may also be employed for the discovery of novel hepatotoxic species or genera in culture collections or from environmental samples. The methods and kits of the invention may also be employed to detect cyanotoxins that accumulate in other animals, for example, fish and shellfish.

Detection of *SXT* and *CYR* polynucleotides and polypeptides disclosed herein may be performed using any suitable method. For example, methods for the detection of *SXT* and *CYR* polynucleotides and/or polypeptides disclosed herein may involve the use of a primer, probe or antibody specific for one or more *SXT* and *CYR* polynucleotides and

polypeptides. Suitable techniques and assays in which the skilled addressee may utilise a primer, probe or antibody specific for one or more *SXT* and *CYR* polynucleotides and polypeptides include, for example, the polymerase chain reaction (and related variations of this technique), antibody based assays such as ELISA and flow cytometry, and fluorescent microscopy. Methods by which the *SXT* and *CYR* polypeptides disclosed herein may be identified are generally known in the art, and are described for example in Coligan J. E. et al. (Eds) *Current Protocols in Protein Science* (2007), John Wiley and Sons, Inc; Walker, J. M., (Ed) (1988) *New Protein Techniques: Methods in Molecular Biology*, Humana Press, Clifton, N.J; and Scopes, R. K. (1987) *Protein Purification: Principles and Practice*, 3rd. Ed., Springer-Verlag, New York, N.Y. For example, *SXT* and *CYR* polypeptides disclosed herein may be detected by western blot or spectrophotometric analysis. Other examples of suitable methods for the detection of *SXT* and *CYR* polypeptides are described, for example, in US Patent No. 4683195, US Patent No. 6228578, US Patent No. 7282355, US Patent No. 7348147 and PCT publication No. W0/2007/056723.

In a preferred embodiment of the invention, the detection of *SXT* and *CYR* polynucleotides and polypeptides is achieved by amplification of DNA from the sample of interest by polymerase chain reaction, using primers that hybridise specifically to the *SXT* and/or *CYR* sequence, or a variant or fragment thereof, and detecting the amplified sequence.

Nucleic acids and polypeptides for analysis using methods and kits disclosed herein may be extracted from organisms either in mixed culture or as individual species or genus isolates. Accordingly, the organisms may be cultured prior to nucleic acid and/or polypeptide isolation or alternatively nucleic acid and/or polypeptides may be extracted directly from environmental samples, such as water samples or blue-green algal blooms.

Suitable methods for the extraction and purification of nucleic acids for analysis using the methods and kits invention are generally known in the art and are described, for example, in Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc; Neilan (1995) *Appl. Environ. Microbiol.* 61:2286-2291; and Neilan et al. (2002) *Astrobiol.* 2:271-280. The skilled addressee will readily appreciate that the invention is not limited to the specific methods for nucleic acid isolation described therein and other suitable methods are encompassed by the invention. The invention may be performed without nucleic acid isolation prior to analysis of the nucleic acid.

Suitable methods for the extraction and purification of polypeptides for the purposes of the invention are generally known in the art and are described, for example, in Coligan J. E. et al. (Eds) *Current Protocols in Protein Science* (2007), John Wiley and Sons, Inc; Walker, J. M., (Ed) (1988) *New Protein Techniques: Methods in Molecular Biology*, Humana Press, Clifton, N.J; and Scopes, R. K. (1987) *Protein Purification: Principles and Practice*, 3rd. Ed., Springer-Verlag, New York, N.Y. Examples of suitable techniques for protein extraction include, but are not limited to dialysis, ultrafiltration, and precipitation. Protein purification techniques suitable for use include, but are not limited to, reverse-phase chromatography, hydrophobic interaction chromatography, centrifugation, gel filtration, ammonium sulfate precipitation, and ion exchange.

In accordance with the methods and kits of the invention, *SXT* and *CYR* polynucleotides or variants or fragments thereof may be detected by any suitable means known in the art. In a preferred embodiment of the invention, *SXT* and *CYR* polynucleotides are detected by PCR amplification. Under the PCR approach, oligonucleotide primers can be designed for use in PCR reactions to amplify *SXT* and *CYR* polynucleotides of the invention. Also encompassed by the invention is the PCR amplification of complementary DNA (cDNA) amplified from messenger RNA (mRNA) derived from reverse-transcription of *SXT* and *CYR* sequences (RT-PCR). Known methods of PCR include, but are not limited to, methods using paired primers, nested primers, single specific primers, degenerate primers, gene-specific primers, vector-specific primers, partially-mismatched primers, and the like. Methods for designing PCR and RT-PCR primers are generally known in the art and are disclosed, for example, in Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc; Maniatis et al. *Molecular Cloning* (1982), 280-281; Innis et al. (Eds) (1990) *PCR Protocols: A Guide to Methods and Applications* (Academic Press, New York); Innis and Gelfand, (Eds) (1995) *PCR Strategies* (Academic Press, New York); Innis and Gelfand, (Eds) (1999) *PCR Methods Manual* (Academic Press, New York); and Sambrook et al. (1989) *Molecular Cloning: A Laboratory Manual* (2nd ed., Cold Spring Harbor Laboratory Press, Plainview, New York).

The skilled addressee will readily appreciate that various parameters of PCR and RT-PCR procedures may be altered without affecting the ability to achieve the desired product. For example, the salt concentration may be varied or the time and/or temperature of one or more of the denaturation, annealing and extension steps may be varied. Similarly, the amount of DNA, cDNA, or RNA template may also be varied depending on

the amount of nucleic acid available or the optimal amount of template required for efficient amplification. The primers for use in the methods and kits of the present invention are typically oligonucleotides typically being at least about 5 nucleotides to about 80 nucleotides in length, more typically about 10 nucleotides in length to about 50 nucleotides in length, and even more typically about 15 nucleotides in length to about 30 nucleotides in length. The skilled addressee will recognise that the primers described herein may be useful for a number of different applications, including but not limited to PCR, RT-PCR, and use of probes for the detection of *SXT* or *CYR* sequences.

Such primers can be prepared by any suitable method, including, for example, direct chemical synthesis or cloning and restriction of appropriate sequences. Not all bases in the primer need reflect the sequence of the template molecule to which the primer will hybridize, the primer need only contain sufficient complementary bases to enable the primer to hybridize to the template. A primer may also include mismatch bases at one or more positions, being bases that are not complementary to bases in the template, but rather are designed to incorporate changes into the DNA upon base extension or amplification. A primer may include additional bases, for example in the form of a restriction enzyme recognition sequence at the 5' end, to facilitate cloning of the amplified DNA.

The invention provides a method of detecting a cyanotoxic organism based on the detection of one or more of *SXT* gene 6 (*sxtA*), *SXT* gene 9 (*sxtG*), *SXT* gene 10 (*sxtH*), *SXT* gene 11 (*sxtI*) and *SXT* gene 17 (*sxtX*) (SEQ ID NOS: 14, 20, 22, 24, and 36 respectively), or fragments or variants thereof. Additionally or alternatively, a cyanotoxic organism may be detected based on the detection of one or more of the following *SXT* polypeptides: *SXTA* (SEQ ID NO: 15), *SXTG* (SEQ ID NO: 21), *SXTH* (SEQ ID NO: 23), *SXTI* (SEQ ID NO: 25), *SXTX* (SEQ ID NO: 37), or fragments or variants thereof.

The skilled addressee will recognise that any primers capable of the amplifying the stated *SXT* and/or *CYR* sequences, or variants or fragments thereof, are suitable for use in the methods of the invention. For example, suitable oligonucleotide primer pairs for the PCR amplification of *SXT* gene 6 (*sxtA*) may comprise a first primer comprising the sequence of SEQ ID NO: 70 and a second primer comprising the sequence of SEQ ID NO: 71, a first primer comprising the sequence of SEQ ID NO: 72 and a second primer comprising the sequence of SEQ ID NO: 73, a first primer comprising the sequence of SEQ ID NO: 74 and a second primer comprising the sequence of SEQ ID NO: 75, a first primer comprising the sequence of SEQ ID NO: 76 and a second primer comprising the

sequence of SEQ ID NO: 77, a first primer comprising the sequence of SEQ ID NO: 78 and a second primer comprising the sequence of SEQ ID NO: 79, a first primer comprising the sequence of SEQ ID NO: 113 and a second primer comprising the sequence of SEQ ID NO: 114, or a first primer comprising the sequence of SEQ ID NO: 115 or SEQ ID NO: 116 and a second primer comprising the sequence of SEQ ID NO: 117.

Suitable oligonucleotide primer pairs for the amplification of *SXT* gene 9 (*sxtG*) may comprise a first primer comprising the sequence of SEQ ID NO: 118 and a second primer comprising the sequence of SEQ ID NO: 119, or a first primer comprising the sequence of SEQ ID NO: 120 and a second primer comprising the sequence of SEQ ID NO: 121.

Suitable oligonucleotide primer pairs for the amplification of *SXT* gene 10 (*sxtH*) may comprise a first primer comprising the sequence of SEQ ID NO: 122 and a second primer comprising the sequence of SEQ ID NO: 123.

Suitable oligonucleotide primer pairs for the amplification of *SXT* gene 11 (*sxtI*) may comprise a first primer comprising the sequence of SEQ ID NO: 124 or SEQ ID NO: 125 and a second primer comprising the sequence of SEQ ID NO: 126, or a first primer comprising the sequence of SEQ ID NO: 127 and a second primer comprising the sequence of SEQ ID NO: 128.

Suitable oligonucleotide primer pairs for the amplification of *SXT* gene 17 (*sxtX*) may comprise a first primer comprising the sequence of SEQ ID NO: 129 and a second primer comprising the sequence of SEQ ID NO: 130, or a first primer comprising the sequence of SEQ ID NO: 131 and a second primer comprising the sequence of SEQ ID NO: 132.

The skilled addressee will recognise that fragments and variants of the above-mentioned primer pairs may also efficiently amplify *SXT* gene 6 (*sxtA*), *SXT* gene 9 (*sxtG*), *SXT* gene 10 (*sxtH*), *SXT* gene 11 (*sxtI*) or *SXT* gene 17 (*sxtX*) sequences.

In certain embodiments of the invention, polynucleotide sequences derived from the *CYR* gene are detected based on the detection of *CYR* gene 8 (*cyrJ*) (SEQ ID NO: 95). Suitable oligonucleotide primer pairs for the PCR amplification of *CYR* gene 8 (*cyrJ*) may comprise a first primer having the sequence of SEQ ID NO: 111 or a fragment or variant thereof and a second primer having the sequence of SEQ ID NO: 112 or a fragment thereof.

Also included within the scope of the present invention are variants and fragments of the exemplified oligonucleotide primers. The skilled addressee will also recognise that the invention is not limited to the use of the specific primers exemplified, and alternative primer sequences may also be used, provided the primers are designed appropriately so as to enable the amplification of *SXT* and/or *CYR* sequences. Suitable primer sequences can be determined by those skilled in the art using routine procedures without undue experimentation. The location of suitable primers for the amplification of *SXT* and/or *CYR* sequences may be determined by such factors as G+C content and the ability for a sequence to form unwanted secondary structures.

Suitable methods of analysis of the amplified nucleic acids are well known to those skilled in the art and are described for example, in, Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual* (2nd ed., Cold Spring Harbor Laboratory Press, Plainview, New York); Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc; and Maniatis et al. *Molecular Cloning* (1982), 280-281. Suitable methods of analysis of the amplified nucleic acids include, for example, gel electrophoresis which may or may not be preceded by restriction enzyme digestion, and/or nucleic acid sequencing. Gel electrophoresis may comprise agarose gel electrophoresis or polyacrylamide gel electrophoresis, techniques commonly used by those skilled in the art for separation of DNA fragments on the basis of size. The concentration of agarose or polyacrylamide in the gel in large part determines the resolution ability of the gel and the appropriate concentration of agarose or polyacrylamide will therefore depend on the size of the DNA fragments to be distinguished.

In other embodiments of the invention, *SXT* and *CYR* polynucleotides and variants or fragments thereof may be detected by the use of suitable probes. The probes of the invention are based on the sequences of *SXT* and/or *CYR* polynucleotides disclosed herein. Probes are nucleotide sequences of variable length, for example between about 10 nucleotides and several thousand nucleotides, for use in detection of homologous sequences, typically by hybridization. Hybridization probes of the invention may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides.

Methods for the design and/or production of nucleotide probes are generally known in the art, and are described, for example, in Robinson P. J.. et al. (Eds) *Current Protocols in Cytometry* (2007), John Wiley and Sons, Inc; Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc; Sambrook et

al. (1989) *Molecular Cloning: A Laboratory Manual* (2nd ed., Cold Spring Harbor Laboratory Press, Plainview, New York; and Maniatis et al. *Molecular Cloning* (1982), 280-281. Nucleotide probes may be prepared, for example, by chemical synthesis techniques, for example, the phosphodiester and phosphotriester methods (see for
5 example Narang S. A. et al. (1979) *Meth. Enzymol.* 68:90; Brown, E. L. (1979) et al. *Meth. Enzymol.* 68:109; and U.S. Patent No. 4356270), the diethylphosphoramidite method (see Beaucage S.L et al. (1981) *Tetrahedron Letters*, 22:1859-1862). A method for synthesizing oligonucleotides on a modified solid support is described in U.S. Patent No. 4458066.

10 The probes of the invention may be labelled by incorporation of a marker to facilitate their detection. Techniques for labelling and detecting nucleic acids are described, for example, in Ausubel F. M. et al. (Eds) *Current Protocols in Molecular Biology* (2007), John Wiley and Sons, Inc. Examples of suitable markers include
15 fluorescent molecules (e.g. acetylaminofluorene, 5-bromodeoxyuridine, digoxigenin, fluorescein) and radioactive isotopes (e.g. ^{32}P , ^{35}S , ^3H , ^{33}P). Detection of the marker may be achieved, for example, by chemical, photochemical, immunochemical, biochemical, or spectroscopic techniques.

The methods and kits of the invention also encompass the use of antibodies which are capable of binding specifically to the polypeptides of the invention. The antibodies
20 may be used to qualitatively or quantitatively detect and analyse one or more *SXT* or *CYR* polypeptides in a given sample. Methods for the generation and use of antibodies are generally known in the art and described in, for example, Harlow and Lane (Eds) *Antibodies - A Laboratory Manual*, (1988), Cold Spring Harbor Laboratory, N.Y: Coligan, *Current Protocols in Immunology* (1991); Goding, *Monoclonal Antibodies: Principles and Practice* (1986) 2nd ed; and Kohler & Milstein, (1975) *Nature* 256: 495-
25 497. The antibodies may be conjugated to a fluorochrome allowing detection, for example, by flow cytometry, immunohistochemistry or other means known in the art. Alternatively, the antibody may be bound to a substrate allowing colorimetric or chemiluminescent detection. The invention also contemplates the use of secondary
30 antibodies capable of binding to one or more antibodies capable of binding specifically to the polypeptides of the invention.

The invention also provides kits for the detection of cyanotoxic organisms and/or cyanobacteria, and/or dinoflagellates. In general, the kits of the invention comprise at least one agent for detecting the presence of one or more *SXT* and/or *CYR* polynucleotide

or polypeptides disclosed herein, or a variant or fragment thereof. Any suitable agent capable of detecting *SXT* and/or *CYR* sequences of the invention may be included in the kit. Non-limiting examples include primers, probes and antibodies.

In one aspect, the invention provides a kit for the detection of cyanobacteria, the kit comprising at least one agent for detecting the presence the presence of one or more *SXT* polynucleotides or polypeptides as disclosed herein, or a variant or fragment thereof.

The *SXT* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof.

Alternatively, the *SXT* polynucleotide may be an RNA or cDNA encoded by a sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof and/or polypeptides as disclosed herein, or a variant or fragment thereof.

The *SXT* polypeptide may comprise an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, and variants and fragments thereof.

Also provided is a kit for the detection of cyanotoxic organisms. The kit comprises at least one agent for detecting the presence of one or more *SXT* polynucleotides or polypeptides as disclosed herein, or a variant or fragment thereof.

The *SXT* polynucleotide may comprise a sequence selected from the group
5 consisting of SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 36, and variants and fragments thereof.

Alternatively, the *SXT* polynucleotide may be an RNA or cDNA encoded by a sequence selected from the group consisting of SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 36, and variants and fragments thereof.

10 The *SXT* polypeptide may comprising an amino acid sequence selected from the group consisting of consisting of SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 37, and variants and fragments thereof.

The at least one agent may be any suitable reagent for the detection of *SXT* polynucleotides and/or polypeptides disclosed herein. For example, the agent may be a
15 primer, an antibody or a probe. By way of exemplification only, the primers or probes may comprise a sequence selected from the group consisting of SEQ ID NO: 70, SEQ ID NO: 71, SEQ ID NO: 72, SEQ ID NO: 73, SEQ ID NO: 74, SEQ ID NO: 75, SEQ ID NO: 76, SEQ ID NO: 77, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 113, SEQ ID NO: 114, SEQ ID NO: 115, SEQ ID NO: 116, SEQ ID NO: 117, SEQ ID NO: 118, SEQ
20 ID NO: 119, SEQ ID NO: 120, SEQ ID NO: 121, SEQ ID NO: 122, SEQ ID NO: 123, SEQ ID NO: 124, SEQ ID NO: 125, SEQ ID NO: 126, SEQ ID NO: 127, SEQ ID NO: 128, SEQ ID NO: 129, SEQ ID NO: 130, SEQ ID NO: 131, SEQ ID NO: 132, SEQ ID NO: 133, SEQ ID NO: 134, and variants and fragments thereof.

In certain embodiments of the invention, the kits for the detection of cyanobacteria
25 or cyanotoxic organisms may further comprise at least one additional agent capable of detecting one or more *CYR* polynucleotide and/or *CYR* polypeptide sequences as disclosed herein, or a variant or fragment thereof.

The *CYR* polynucleotide may comprise a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 83,
30 SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants and fragments thereof.

Alternatively, the *CYR* polynucleotide may comprise a ribonucleic acid or complementary DNA encoded by a sequence selected from the group consisting of: SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants and fragments thereof.

The *CYR* polypeptide may comprise a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, and SEQ ID NO: 110, and variants and fragments thereof.

The at least one additional agent may be selected, for example, from the group consisting of primers, antibodies and probes. A suitable primer or probe may comprise a sequence selected from the group consisting of SEQ ID NO: 111, SEQ ID NO: 112, and variants and fragments thereof.

In another aspect, the invention provides a kit for the detection of cyanobacteria, the kit comprising at least one agent for detecting the presence the presence of one or more *CYR* polynucleotides or polypeptides as disclosed herein, or a variant or fragment thereof.

The *CYR* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants and fragments thereof.

Alternatively, the *CYR* polynucleotide may be an RNA or cDNA encoded by a sequence selected from the group consisting of SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants and fragments thereof.

The *CYR* polypeptide may comprise a sequence selected from the group consisting of SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, and SEQ ID NO: 110, and variants or fragments thereof.

In certain embodiments of the invention, the kits for detecting cyanobacteria comprising one or more agents for the detection of *CYR* sequences or variants or fragments thereof, may further comprise at least one additional agent capable of detecting one or more of the *SXT* polynucleotides and/or *SXT* polypeptides disclosed herein, or
5 variants or fragments thereof.

The *SXT* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28,
10 SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof.

Alternatively, the *SXT* polynucleotide may be an RNA or cDNA encoded by a sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36,
20 SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof and/or polypeptides as disclosed herein, or a variant or fragment thereof.

The at least one agent may be any suitable reagent for the detection of *CYR* polynucleotides and/or polypeptides disclosed herein. For example, the agent may be a primer, an antibody or a probe. By way of exemplification only, the primers or probes may comprise a sequence selected from the group consisting of SEQ ID NO: 111, SEQ ID NO: 112, and variants and fragments thereof.

Also provided is a kit for the detection of dinoflagellates, the kit comprising at least one agent for detecting the presence one or more *SXT* polynucleotides or polypeptides as disclosed herein, or a variant or fragment thereof.

The *SXT* polynucleotide may comprise a sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID

NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, 5 SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof.

Alternatively, the *SXT* polynucleotide may be an RNA or cDNA encoded by a sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, 15 SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, SEQ ID NO: 68, and variants and fragments thereof and/or polypeptides as disclosed herein, or a variant or fragment thereof.

The *SXT* polypeptide may comprise an amino acid sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, SEQ ID NO: 69, and variants 25 and fragments thereof.

In general, the kits of the invention may comprise any number of additional components. By way of non-limiting examples the additional components may include, reagents for cell culture, reference samples, buffers, labels, and written instructions for 30 performing the detection assay.

Methods of screening

The polypeptides and polynucleotides of the present invention, and fragments and analogues thereof are useful for the screening and identification of compounds and agents that interact with these molecules. In particular, desirable compounds are those that modulate the activity of these polypeptides and polynucleotides. Such compounds may exert a modulatory effect by activating, stimulating, increasing, inhibiting or preventing expression or activity of the polypeptides and/or polynucleotides. Suitable compounds may exert their effect by virtue of either a direct (for example binding) or indirect interaction.

Compounds which bind, or otherwise interact with the polypeptides and polynucleotides of the invention, and specifically compounds which modulate their activity, may be identified by a variety of suitable methods. Non limiting methods include the two-hybrid method, co-immunoprecipitation, affinity purification, mass spectroscopy, tandem affinity purification, phage display, label transfer, DNA microarrays/gene coexpression and protein microarrays.

For example, a two-hybrid assay may be used to determine whether a candidate agent or plurality of candidate agents interacts or binds with a polypeptide of the invention or a variant or fragment thereof. The yeast two-hybrid assay system is a yeast-based genetic assay typically used for detecting protein-protein interactions (Fields and Song., *Nature* 340: 245-246 (1989)). The assay makes use of the multi-domain nature of transcriptional activators. For example, the DNA-binding domain of a known transcriptional activator may be fused to a polypeptide of the invention or a variant or fragment thereof, and the activation domain of the transcriptional activator fused to the candidate agent. Interaction between the candidate agent and the polypeptide of the invention or a variant or fragment thereof, will bring the DNA-binding and activation domains of the transcriptional activator into close proximity. Subsequent transcription of a specific reporter gene activated by the transcriptional activator allows the detection of an interaction.

In a modification of the technique above, a fusion protein may be constructed by fusing the polypeptide of the invention or a variant or fragment thereof to a detectable tag, for example alkaline phosphatase, and using a modified form of immunoprecipitation as described by Flanagan and Leder (Flanagan and Leder, *Cell* 63:185-194 (1990))

Alternatively, co-immunoprecipitation may be used to determine whether a candidate agent or plurality of candidate agents interacts or binds with polypeptide of the invention or a variant or fragment thereof. Using this technique, cyanotoxic organisms,

cyanobacteria and/or dinoflagellates may be lysed under nondenaturing conditions suitable for the preservation of protein-protein interactions. The resulting solution can then be incubated with an antibody specific for a polypeptide of the invention or a variant or fragment thereof and immunoprecipitated from the bulk solution, for example by capture with an antibody-binding protein attached to a solid support. Immunoprecipitation of the polypeptide of the invention or a variant or fragment thereof by this method facilitates the co-immunoprecipitation of an agent associated with that protein. The identification an associated agent can be established using a number of methods known in the art, including but not limited to SDS-PAGE, western blotting, and mass spectrometry.

Alternatively, the phage display method may be used to to determine whether a candidate agent or plurality of candidate agents interacts or binds with a polypeptide of the invention or a variant or fragment thereof. Phage display is a test to screen for protein interactions by integrating multiple genes from a gene bank into phage. Under this method, recombinant DNA techniques are used to express numerous genes as fusions with the coat protein of a bacteriophage such the peptide or protein product of each gene is displayed on the surface of the viral particle. A whole library of phage-displayed peptides or protein products of interest can be produced in this way. The resulting libraries of phage-displayed peptides or protein products may then be screened for the ability to bind a polypeptide of the invention or a variant or fragment thereof. DNA extracted from interacting phage contains the sequences of interacting proteins.

Alternatively, affinity chromatography may be used to to determine whether a candidate agent or plurality of candidate agents interacts or binds with a polypeptide of the invention or a variant or fragment thereof. For example, a polypeptide of the invention or a variant or fragment thereof, may be immobilised on a support (such as sepharose) and cell lysates passed over the column. Proteins binding to the immobilised polypeptide of the invention or a variant or fragment thereof may then be eluted from the column and identified, for example by N-terminal amino acid sequencing.

Potential modulators of the activity of the polypeptides of the invention may be generated for screening by the above methods by a number of techniques known to those skilled in the art. For example, methods such as X-ray crystallography and nuclear magnetic resonance spectroscopy may be used to model the structure of polypeptide of the invention or a variant or fragment thereof, thus facilitating the design of potential modulating agents using computer-based modeling. Various forms of combinatorial chemistry may also be used to generate putative modulators.

Polypeptides of the invention and appropriate variants or fragments thereof can be used in high-throughput screens to assay candidate compounds for the ability to bind to, or otherwise interact therewith. These candidate compounds can be further screened against functional polypeptides to determine the effect of the compound on polypeptide activity.

The present invention also contemplates compounds which may exert their modulatory effect on polypeptides of the invention by altering expression of the polypeptide. In this case, such compounds may be identified by comparing the level of expression of the polypeptide in the presence of a candidate compound with the level of expression in the absence of the candidate compound.

It will be appreciated that the methods described above are merely examples of the types of methods that may be utilised to identify agents that are capable of interacting with, or modulating the activity of polypeptides of the invention or variants or fragments thereof. Other suitable methods will be known by persons skilled in the art and are within the scope of this invention.

Using the methods described above, an agent may be identified that is an agonist of a polypeptide of the invention or a variant or fragment thereof. Agents which are agonists enhance one or more of the biological activities of the polypeptide. Alternatively, the methods described above may identify an agent that is an antagonist of a polypeptide of the invention or a variant or fragment thereof. Agents which are antagonists retard one or more of the biological activities of the polypeptide.

Antibodies may act as agonists or antagonists of a polypeptide of the invention or a variant or fragment thereof. Preferably suitable antibodies are prepared from discrete regions or fragments of the polypeptides of the invention or variants or fragments thereof. An antigenic portion of a polynucleotide of interest may be of any appropriate length, such as from about 5 to about 15 amino acids. Preferably, an antigenic portion contains at least about 5, 6, 7, 8, 9, 10, 11, 12, 13 or 14 amino acid residues.

Methods for the generation of suitable antibodies will be readily appreciated by those skilled in the art. For example, monoclonal antibody specific for a polypeptide of the invention or a variant or fragment thereof typically containing Fab portions, may be prepared using hybridoma technology described in *Antibodies-A Laboratory Manual*, Harlow and Lane, eds., Cold Spring Harbor Laboratory, N.Y. (1988).

In essence, in the preparation of monoclonal antibodies directed toward polypeptide of the invention or a variant or fragment thereof, any technique that provides for the

production of antibody molecules by continuous cell lines in culture may be used. These include the hybridoma technique originally developed by Kohler *et al.*, *Nature*, 256:495-497 (1975), as well as the trioma technique, the human B-cell hybridoma technique (Kozbor *et al.*, *Immunology Today*, 4:72 (1983)), and the EBV-hybridoma technique to
5 produce human monoclonal antibodies (Cole *et al.*, in *Monoclonal Antibodies and Cancer Therapy*, pp. 77- 96, Alan R. Liss, Inc., (1985)). Immortal, antibody-producing cell lines can be created by techniques other than fusion, such as direct transformation of B lymphocytes with oncogenic DNA, or transfection with Epstein-Barr virus. See, for example, M. Schreier *et al.*, "*Hybridoma Techniques*" Cold Spring Harbor Laboratory,
10 (1980); Hammerling *et al.*, "*Monoclonal Antibodies and T-cell Hybridomas*" Elsevier/North-Holland Biochemical Press, Amsterdam (1981); and Kennett *et al.*, "*Monoclonal Antibodies*", Plenum Press (1980).

In brief, a means of producing a hybridoma from which the monoclonal antibody is produced, a myeloma or other self-perpetuating cell line is fused with lymphocytes
15 obtained from the spleen of a mammal hyperimmunised with a recognition factor-binding portion thereof, or recognition factor, or an origin-specific DNA-binding portion thereof. Hybridomas producing a monoclonal antibody useful in practicing this invention are identified by their ability to immunoreact with the present recognition factors and their ability to inhibit specified transcriptional activity in target cells.

20 A monoclonal antibody useful in practicing the invention can be produced by initiating a monoclonal hybridoma culture comprising a nutrient medium containing a hybridoma that secretes antibody molecules of the appropriate antigen specificity. The culture is maintained under conditions and for a time period sufficient for the hybridoma to secrete the antibody molecules into the medium. The antibody-containing medium is
25 then collected. The antibody molecules can then be further isolated by well-known techniques.

Similarly, there are various procedures known in the art which may be used for the production of polyclonal antibodies. For the production of polyclonal antibodies against a polypeptide of the invention or a variant or fragment thereof, various host animals can be
30 immunized by injection with a polypeptide of the invention, or a variant or fragment thereof, including but not limited to rabbits, chickens, mice, rats, sheep, goats, etc. Further, the polypeptide variant or fragment thereof can be conjugated to an immunogenic carrier (e.g., bovine serum albumin (BSA) or keyhole limpet hemocyanin (KLH)). Also, various adjuvants may be used to increase the immunological response, including but not

limited to Freund's (complete and incomplete), mineral gels such as aluminium hydroxide, surface active substances such as rylenelecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and *Corynebacterium* parvum.

Screening for the desired antibody can also be accomplished by a variety of techniques known in the art. Assays for immunospecific binding of antibodies may include, but are not limited to, radioimmunoassays, ELISAs (enzyme-linked immunosorbent assay), sandwich immunoassays, immunoradiometric assays, gel diffusion precipitation reactions, immunodiffusion assays, in situ immunoassays, Western blots, precipitation reactions, agglutination assays, complement fixation assays, immunofluorescence assays, protein A assays, and immunoelectrophoresis assays, and the like (see, for example, Ausubel *et al.*, Current Protocols in Molecular Biology, Vol. 1, John Wiley & Sons, Inc., New York (1994)). Antibody binding may be detected by virtue of a detectable label on the primary antibody. Alternatively, the antibody may be detected by virtue of its binding with a secondary antibody or reagent which is appropriately labelled. A variety of methods are known in the art for detecting binding in an immunoassay and are included in the scope of the present invention.

The antibody (or fragment thereof) raised against a polypeptide of the invention or a variant or fragment thereof, has binding affinity for that protein. Preferably, the antibody (or fragment thereof) has binding affinity or avidity greater than about 10^5M^{-1} , more preferably greater than about 10^6M^{-1} , more preferably still greater than about 10^7M^{-1} and most preferably greater than about 10^8M^{-1} .

In terms of obtaining a suitable amount of an antibody according to the present invention, one may manufacture the antibody(s) using batch fermentation with serum free medium. After fermentation the antibody may be purified via a multistep procedure incorporating chromatography and viral inactivation/removal steps. For instance, the antibody may be first separated by Protein A affinity chromatography and then treated with solvent/detergent to inactivate any lipid enveloped viruses. Further purification, typically by anion and cation exchange chromatography may be used to remove residual proteins, solvents/detergents and nucleic acids. The purified antibody may be further purified and formulated into 0.9% saline using gel filtration columns. The formulated bulk preparation may then be sterilised and viral filtered and dispensed.

Embodiments of the invention may utilise antisense technology to inhibit the expression of a nucleic acid of the invention or a fragment or variant thereof by blocking translation of the encoded polypeptide. Antisense technology takes advantage of the fact that nucleic acids pair with complementary sequences. Suitable antisense molecules can
5 be manufactured by chemical synthesis or, in the case of antisense RNA, by transcription *in vitro* or *in vivo* when linked to a promoter, by methods known to those skilled in the art.

For example, antisense oligonucleotides, typically of 18-30 nucleotides in length, may be generated which are at least substantially complementary across their length to a
10 region of the nucleotide sequence of the polynucleotide of interest. Binding of the antisense oligonucleotide to their complementary cellular nucleotide sequences may interfere with transcription, RNA processing, transport, translation and/or mRNA stability. Suitable antisense oligonucleotides may be prepared by methods well known to those of skill in the art and may be designed to target and bind to regulatory regions of the
15 nucleotide sequence or to coding (gene) or non-coding (intergenic region) sequences. Typically antisense oligonucleotides will be synthesized on automated synthesizers. Suitable antisense oligonucleotides may include modifications designed to improve their delivery into cells, their stability once inside a cell, and/or their binding to the appropriate target. For example, the antisense oligonucleotide may be modified by the addition of
20 one or more phosphorothioate linkages, or the inclusion of one or morpholine rings into the backbone (so-called 'morpholino' oligonucleotides).

An alternative antisense technology, known as RNA interference (RNAi), may be used, according to known methods in the art (see for example WO 99/49029 and WO 01/70949), to inhibit the expression of a polynucleotide. RNAi refers to a means of
25 selective post-transcriptional gene silencing by destruction of specific mRNA by small interfering RNA molecules (siRNA). The siRNA is generated by cleavage of double stranded RNA, where one strand is identical to the message to be inactivated. Double-stranded RNA molecules may be synthesised in which one strand is identical to a specific region of the p53 mRNA transcript and introduced directly. Alternatively corresponding
30 dsDNA can be employed, which, once presented intracellularly is converted into dsRNA. Methods for the synthesis of suitable molecule for use in RNAi and for achieving post-transcriptional gene silencing are known to those of skill in the art.

A further means of inhibiting expression may be achieved by introducing catalytic antisense nucleic acid constructs, such as ribozymes, which are capable of cleaving

mRNA transcripts and thereby preventing the production of wild type protein. Ribozymes are targeted to and anneal with a particular sequence by virtue of two regions of sequence complementarity to the target flanking the ribozyme catalytic site. After binding the ribozyme cleaves the target in a site-specific manner. The design and testing
5 of ribozymes which specifically recognise and cleave sequences of interest can be achieved by techniques well known to those in the art (see for example Lieber and Strauss, 1995, *Molecular and Cellular Biology*, 15:540-551).

The invention will now be described with reference to specific examples, which should not be construed as in any way limiting the scope of the invention.

10

Examples

The invention will now be described with reference to specific examples, which
15 should not be construed as in any way limiting the scope of the invention.

Example 1: Cyanobacterial cultures and characterisation of the *SXT* gene cluster.

Cyanobacterial strains used in the present study (Figure 1) were grown in Jaworski
20 medium in static batch culture at 26°C under continuous illumination ($10 \mu\text{mol m}^{-2} \text{s}^{-1}$). Total genomic DNA was extracted from cyanobacterial cells by lysozyme/SDS/proteinase K lysis following phenol-chloroform extraction as described in Neilan, B. A. 1995.. *Appl Environ Microbiol* 61:2286-2291. DNA in the supernatant was precipitated with 2 volumes - 20°C ethanol, washed with 70% ethanol, dissolved in TE-buffer (10:1), and
25 stored at - 20°C. PCR primer sequences used for the amplification of *sxt* ORFs are shown in Figure 1B).

PCR amplicons were separated by agarose gel electrophoresis in TAE buffer (40 mM Tris-acetate, 1 mM EDTA, pH 7.8), and visualised by UV transillumination after staining in ethidium bromide (0.5 $\mu\text{g/ml}$). Sequencing of unknown regions of DNA was
30 performed by adaptor-mediated PCR as described in Moffitt et al. (2004) *Appl. Environ. Microbiol.* 70:6353-6362. Automated DNA sequencing was performed using the PRISM Big Dye cycle sequencing system and a model 373 sequencer (Applied Biosystems). Sequence data were analysed using ABI Prism-Autoassembler software, and percentage similarity and identity to other translated sequences determined using BLAST in

conjunction with the National Center for Biotechnology Information (NIH), Fugue blast (<http://www-cryst.bioc.cam.ac.uk/fugue/>) was used to identify distant homologs via sequence-structure comparisons. The *sxt* gene clusters were assembled using the software Phred, Phrap, and Consed (<http://www.phrap.org/phredphrapconsed.html>), and open reading frames manually identified. GenBank accession numbers for the *sxt* gene cluster from *C. raciborskii* T3 is DQ787200.

Example 2: Mass spectrometric analysis of SXT intermediates.

Bacterial extracts and SXT standards were analysed by HPLC (Thermo Finnigan Surveyor HPLC and autosampler) coupled to an ion trap mass spectrometer (Thermo Finnigan LCQ Deca XP Plus) fitted with an electrospray source. Separation of analytes was obtained on a 2.1 mm x 150 mm Phenomenex Luna 3 micron C18 column at 100 mL/min. Analysis was performed using a gradient starting at 5% acetonitrile in 10 mM heptafluorobutyric acid (HFBA). This was maintained for 10 min, then ramped to 100% acetonitrile, over 30 min. Conditions were held at 100% acetonitrile for 10 min to wash the column and then returned to 5% acetonitrile in 10 mM HFBA and again held for 10 min to equilibrate the column for the next sample. This resulted in a runtime of 60 min per sample. Sample volumes of 10-100 mL were injected for each analysis. The HPLC eluate directly entered the electrospray source, which was programmed as follows: electrospray voltage 5 kV, sheath gas flow rate 30 arbitrary units, auxiliary gas flow rate 5 arbitrary units. The capillary temperature was 200°C and had a voltage of 47 V. Ion optics were optimised for maximum sensitivity before sample analysis using the instruments autotune function with a standard toxin solution. Mass spectra were acquired in the centroid mode over the m/z range 145-650. Mass range setting was 'normal', with 200 ms maximum ion injection time and automatic gain control (AGC) on. Tandem mass spectra were obtained over a m/z range relevant to the precursor ion. Collision energy was typically 20-30 ThermoFinnigan arbitrary units, and was optimised for maximal information using standards where available.

Example 3: Identification and sequencing of the SXT gene cluster in *Cylindrospermopsis raciborskii* T3

O-carbamoyltransferase was initially detected in *C. raciborskii* T3 via degenerate PCR, and later named *sxtI*. Further investigation showed that homologues of *sxtI* were exclusively present in SXT toxin-producing strains of four cyanobacterial genera (Table

1), thus representing a good candidate gene in *SXT* toxin biosynthesis. The sequence of the complete putative *SXT* biosynthetic gene cluster (*sxt*) was then obtained by genome walking up- and downstream of *sxtI* in *C. raciborskii* T3 (Figure 3). In *C. raciborskii* T3, this *sxt* gene cluster spans approximately 35000 bp, encoding 31 open reading frames (Figure 2). The cluster also included other genes encoding *SXT*-biosynthesis enzymes, including a methyltransferase (*sxtA1*), a class II aminotransferase (*sxtA4*), an amidinotransferase (*sxtG*), dioxygenases (*sxtH*), in addition to the Ocarbamoyltransferase (*sxtI*). PCR screening of selected *sxt* open reading frames in toxic and non-toxic cyanobacteria strains showed that they were exclusively present in *SXT* toxin-producing isolates (Figure 1A), indicating the association of these genes with the toxic phenotype. In the following passages we describe the open reading frames in the putative *sxt* gene cluster and their predicted functions, based on bioinformatic analysis, LCMS/ MS data on biosynthetic intermediates and *in vitro* biosynthesis, when applicable.

15 **Example 4: Functional prediction of the parent molecule *SXT* biosynthetic genes**

Bioinformatic analysis of the *sxt* gene cluster revealed that it contains a previously undescribed example of a polyketide synthase (PKS) like structure, named *sxtA*. *SxtA* possesses four catalytic domains, *SxtA1* to *SxtA4*. An iterated PSI-blast search revealed low sequence homology of *SxtA1* to S-adenosylmethionine (SAM)-dependent methyltransferases. Further analysis revealed the presence of three conserved sequence motifs in *SxtA1* (278-ITDMGCGDG- 286, 359-DPENILHI-366, and 424-VVKNHGLMIL-433) that are specific for SAMdependent methyltransferases. *SxtA2* is related to GCN5-related N-acetyl transferases (GNAT). GNAT catalyse the transfer of acetate from acetyl-CoA to various heteroatoms, and have been reported in association with other unconventional PKSs, such as *PedI*, where they load the acyl carrier protein (ACP) with acetate. *SxtA3* is related to an ACP, and provides a phosphopantetheinyl-attachment site. *SxtA4* is homologous to class II aminotransferases and was most similar to 8-amino-7-oxononanoate synthase (AONS). Class II aminotransferases are a monophyletic group of pyridoxal phosphate (PLP)-dependent enzymes, and the only enzymes that are known to perform Claisen-condensations of amino acids. We therefore reasoned that *sxtA* performs the first step in *SXT* biosynthesis, involving a Claisen-condensation.

The predicted reaction sequence of *SxtA*, based on its primary structure, is the loading of the ACP (*SxtA3*) with acetate from acetyl-CoA, followed by the *SxtA1*-

catalysed methylation of acetyl-ACP, converting it to propionyl-ACP. The class II aminotransferase domain, SxtA4, would then perform a Claisen-condensation between propionyl-ACP and arginine (Figure 4). The putative product of SxtA is thus 4-amino-3-oxoguanidinoheptane which is here designated as Compound A', (Figure 4). To verify this pathway for SXT biosynthesis based on comparative gene sequence analysis, cell extracts of *C. raciborskii* T3 were screened by LC-MS/MS for the presence of compound A' (Figure 5) as well as arginine and SXT as controls. Arginine and SXT were readily detected (Figure 5) and produced the expected fragment ions. On the other hand, LC-MS/MS data obtained from m/z 187 was consistent with the presence of structure A from *C. raciborskii* T3 (Figure 5). MS/MS spectra showed the expected fragment ion (m/z 170, m/z 128) after the loss of ammonia and guanidine from A'. LC-MS/MS data strongly supported the predicted function of SxtA and thus a revised initiating reaction in the SXT biosynthesis pathway.

sxtG encodes a putative amidinotransferase, which had the highest amino acid sequence similarity to L-arginine:lysine amidinotransferases. It is proposed that the product of SxtA is the substrate for the amidinotransferase SxtG, which transfers an amidino group from arginine to the α -amino group A' (Figure 4), thus producing 4,7-diguanidino-3-oxoheptane designated compound B' (Figure 3). This hypothetical sequence of reactions was also supported by the detection of C' by LC-MS/MS (Figure 4). Cell extracts from *C. raciborskii* T3, however, did not contain any measurable levels of B' (4,7-diguanidino-3-oxoheptane). A likely explanation for the failure to detect the intermediate B' is its rapid cyclisation to form C' via the action of SxtB.

The *sxt* gene cluster encodes an enzyme, sxtB, similar to the cytidine deaminase-like enzymes from γ -proteobacteria. The catalytic mechanism of cytidine deaminase is a retro-aldol cleavage of ammonia from cytidine, which is the same reaction mechanism in the reverse direction as the formation of the first heterocycle in the conversion from B' to C' (Figure 4). It is therefore suggested that SxtB catalyses this retroaldol-like condensation (step 4, Figure 4).

The incorporation of methionine methyl into SXT, and its hydroxylation was studied. Only one methionine methyl-derived hydrogen is retained in SXT, and a 1,2-H shift has been observed between acetate-derived C-5 and C-6 of SXT. Hydroxylation of the methyl side-chain of the SXT precursor proceeds via epoxidation of a double-bond between the SAM-derived methyl group and the acetate derived C-6. This incorporation pattern may result from an electrophilic attack of methionine methyl on the double bond

between C-5 and C-6, which would have formed during the preceding cyclisation. Subsequently, the new methylene side-chain would be epoxidated, followed by opening to an aldehyde, and subsequent reduction to a hydroxyl. Retention of only one methionine methyl-derived hydrogen, the 1,2-H shift between C-5 and C-6, and the lacking 1,2-H shift between C-1 and C-5 is entirely consistent with the results of this study, whereby the introduction of methionine methyl precedes the formation of the three heterocycles.

sxtD encodes an enzyme with sequence similarity to sterol desaturase and is the only candidate desaturase present in the *sxt* gene cluster, SxtD is predicted to introduce a double bond between C-1 and C-5 of C', and cause a 1,2-H shift between C-5 and C-6 (compound D', Figure 3). The gene product of *sxtS* has sequence homology to non-heme iron 2-oxoglutarate-dependent (2OG) dioxygenases. These are multifunctional enzymes that can perform hydroxylation, epoxidation, desaturation, cyclisation, and expansion reactions. 2OG dioxygenases have been reported to catalyse the oxidative formation of heterocycles. SxtS could therefore perform the consecutive epoxidation of the new double bond, and opening of the epoxide to an aldehyde with concomitant bicyclisation. This explains the retention of only one methionine methyl-derived hydrogen, and the lack of a 1,2-H shift between C-1 and C-5 of SXT (steps 5 to 7, Figure 4). SxtU has sequence similarity to short-chain alcohol dehydrogenases. The most similar enzyme with a known function is clavuldehyde dehydrogenase (AAF86624), which reduces the terminal aldehyde of clavulanate-9-aldehyde to an alcohol. SxtU is therefore predicted to reduce the terminal aldehyde group of the SXT precursor in step 8 (Figure 4), forming compound E'.

The concerted action of SxtD, SxtS and SxtU is therefore the hydroxylation and bicyclisation of compound C' to E' (Figure 4). In support for this proposed pathway of SXT biosynthesis, LC-MS/MS obtained from *m/z* 211 and *m/z* 225 allowed the detection of compounds C' and E' from *C. raciborskii* T3 (Figure 5). On the other hand, no evidence could be found by LC-MS/MS for intermediates B (*m/z* 216), and C (*m/z* 198). MS/MS spectra showed the expected fragment ions after the loss of ammonia and guanidine from C', as well as the loss of water in the case of E'.

The detection of E' indicated that the final reactions leading to the complete SXT molecule are the O-carbamoylation of its free hydroxyl group and a oxidation of C-12. The actual sequence of these final reactions, however, remains uncertain. The gene product of *sxtI* is most similar to a predicted Ocarbamoyltransferase from *Trichodesmium erythraeum* (accession ABG50968) and other predicted O-carbamoyltransferases from

cyanobacteria. O-carbamoyltransferases invariably transfer a carbamoyl group from carbamoylphosphate to a free hydroxyl group. Our data indicate that SxtI may catalyse the transfer of a carbamoyl group from carbamoylphosphate to the free hydroxy group of E'. Homologues of *sxtJ* and *sxtK* with a known function were not found in the databases, however it was noted that *sxtJ* and *sxtK* homologues were often encoded adjacent to O-carbamoyltransferase genes.

The *sxt* gene cluster contains two genes, *sxtH* and *sxtT*, each encoding a terminal oxygenase subunit of bacterial phenyl-propionate and related ring-hydroxylating dioxygenases. The closest homologue with a predicted function was capreomycin hydroxylase from *Streptomyces vinaceus*, which hydroxylates a ring carbon (C-6) of capreomycin. SxtH and SxtT may therefore perform a similar function in SXT biosynthesis, that is, the oxidation or hydroxylation and oxidation of C-12, converting F' into SXT.

Members belonging to bacterial phenylpropionate and related ring-hydroxylating dioxygenases are multi-component enzymes, as they require an oxygenase reductase for their regeneration after each catalytic cycle. The *sxt* gene cluster provides a putative electron transport system, which would fulfill this function. *sxtV* encodes a 4Fe-4S ferredoxin with high sequence homology to a ferredoxin from *Nostoc punctiforme*. *sxtW* was most similar to fumarate reductase/succinate dehydrogenase-like enzymes from *A. variabilis* and *Nostoc punctiforme*, followed by AsfA from *Pseudomonas putida*. AsfA and AsfB are enzymes involved in the transport of electrons resulting from the catabolism of aryl sulfonates. SxtV could putatively extract an electron pair from succinate, converting it to fumarate, and then transfer the electrons via ferredoxin (SxtW) to SxtH and SxtT.

Example 5: Comparative sequence analysis and functional assignment of SXT tailoring genes

Following synthesis of the parent molecule SXT, modifying enzymes introduce various functional groups. In addition to SXT, *C. raciborskii* T3 produces N-1 hydroxylated (neoSXT), decarbamoylated (dcSXT), and N-sulfurylated (GTX-5) toxins, whereas *A. circinalis* AWQC131C produces decarbamoylated (dcSXT), O-sulfurylated (GTX-3/2, dcGTX-3/2), as well as both O-and N-sulfurylated toxins (C- 1/2), but no N-1 hydroxylated toxins.

sxtX encodes an enzyme with homology to cephalosporin hydroxylase. *sxtX* was only detected in *C. raciborskii* T3, *A. flos-aquae* NH- 5, and *Lyngbya wollei*, which produce N-1 hydroxylated analogues of *SXT*, such as neo*SXT*. This component of the gene cluster was not present in any strain of *A. circinalis*, and therefore probably the reason why this species does not produce N-1 hydroxylated *PSP* toxins (Figure 1A). The predicted function of SxtX is therefore the N-1 hydroxylation of *SXT*.

A. circinalis AWQC131C and *C. raciborskii* T3 also produces N- and O-sulfated analogues of *SXT* (GTX-5, C-2/3, (dc)GTX- 3/4). The activity of two 3'-phosphate 5'-phosphosulfate (PAPS)-dependent sulfotransferases, which were specific for the N- 21 of *SXT* and GTX-3/2, and O-22 of 11- hydroxy *SXT*, respectively, has been described from the *SXT* toxin-producing dinoflagellate *Gymnodinium catenatum*. The *sxt* gene cluster from *C. raciborskii* T3 encodes a putative sulfotransferase, SxtN. A PSI-BLAST search with SxtN identified only 25 hypothetical proteins of unknown function with an E value above the threshold (0.005). A profile library search, however, revealed significant structural relatedness of SxtN to estrogen sulfotransferase (1AQU) (Z-score=24.02) and other sulfotransferases. SxtN has a conserved N-terminal region, which corresponds to the adenosine 3'-phosphate 5'-phosphosulfate (PAPS) binding region in 1AQU. It is not known, however, whether SxtN transfers a sulfate group to N-21 or O-22. Interestingly, the *sxt* gene cluster encodes an adenylylsulfate kinase (APSK), SxtO, homologues of which are involved in the formation of PAPS (Figure 2). APSK phosphorylates the product of ATPsulfurylase, adenylylsulfate, converting it to PAPS. Other biosynthetic gene clusters that result in sulfated secondary metabolites also contain genes required for the production of PAPS.

Decarbamoylated analogues of *SXT* could be produced via either of two hypothetical scenarios. Enzymes that act downstream of the carbamoyltransferase, SxtI, in the biosynthesis of *PSP* toxins are proposed to have broad substrate specificity, processing both carbamoylated and decarbamoylated precursors of *SXT*. Alternatively, hydrolytic cleavage of the carbamoyl moiety from *SXT* or its precursors may occur. SxtL is related to GDSL-lipases, which are multifunctional enzymes with thioesterase, arylesterase, protease and lysophospholipase activities. The function of SxtL could therefore include the hydrolytic cleavage of the carbamoyl group from *SXT* analogues.

Example 6: Cluster-associated *SXT* genes involved in metabolite transport

sxtF and *sxtM* encoded two proteins with high sequence similarity to sodium-driven multidrug and toxic compound extrusion (MATE) proteins of the NorM family. Members of the NorM family of MATE proteins are bacterial sodium-driven antiporters, that export cationic substances. All of the PSP toxins are cationic substances, except for the C-toxins which are zwitterionic. It is therefore probable that SxtF and SxtM are also involved in the export of PSP toxins. A mutational study of NorM from *V. parahaemolyticus* identified three conserved negatively charged residues (D32, E251, and D367) that confer substrate specificity, however the mechanism of substrate recognition remains unknown. In SxtF, the residue corresponding to E251 of NorM is conserved, whereas those corresponding to D32 and D367 are replaced by the neutral amino acids asparagine and tyrosine, respectively. Residues corresponding to D32 and E251 are conserved in SxtM, but D367 is replaced by histidine. The changes in substrate-binding residues may reflect the differences in PSP toxin substrates transported by these proteins.

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Example 7: Putative transcriptional regulators of saxitoxin synthase

Environmental factors, such as nitrogen and phosphate availability have been reported to regulate the production of PSP toxins in dinoflagellates and cyanobacteria. Two transcriptional factors, *sxtY* and *sxtZ*, related to PhoU and OmpR, respectively, as well as a two component regulator histidine kinase were identified proximal to the 3'-end of the *sxt* gene cluster in *C. raciborskii* T3. PhoU-related proteins are negative regulators of phosphate uptake whereas OmpR-like proteins are involved in the regulation of a variety of metabolisms, including nitrogen and osmotic balance. It is therefore likely that PSP toxin production in *C. raciborskii* T3 is regulated at the transcriptional level in response to the availability of phosphate, as well as, other environmental factors.

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Example 8: Phylogenetic origins of the SXT genes

The *sxt* gene cluster from *C. raciborskii* T3 has a true mosaic structure. Approximately half of the *sxt* genes of *C. raciborskii* T3 were most similar to counterparts from other cyanobacteria, however the remaining genes had their closest matches with homologues from proteobacteria, actinomycetes, sphingobacteria, and firmicutes. There is an increasing body of evidence that horizontal gene transfer (HGT) is a major driving force behind the evolution of prokaryotic genomes, and cyanobacterial genomes are known to be greatly affected by HGT, often involving transposases and

30

phages. The fact that the majority of *sxt* genes are most closely related to homologues from other cyanobacteria, suggests that *SXT* biosynthesis may have evolved in an ancestral cyanobacterium that successively acquired the remaining genes from other bacteria via HGT. The structural organisation of the investigated *sxt* gene cluster, as well as the presence of several transposases related to the IS4-family, suggests that small cassettes of *sxt* genes are mobile.

Example 9: Cyanobacterial cultures and characterisation of the *CYR* gene cluster.

Cyanobacterial strains were grown in Jaworski medium as described in Example 1 above. Total genomic DNA was extracted from cyanobacterial cells by lysozyme/SDS/proteinase K lysis following phenol-chloroform extraction as described previously Neilan, B. A. 1995.. Appl Environ Microbiol 61:2286-2291. DNA in the supernatant was precipitated with 2 volumes -20°C ethanol, washed with 70% ethanol, dissolved in TE-buffer (10:1), and stored at -20°C.

Characterization of unknown regions of DNA flanking the putative cylindrospermopsin biosynthesis genes was performed using an adaptor-mediated PCR as described in Moffitt et al. (2004) Appl. Environ. Microbiol. 70:6353-6362. PCRs were performed in 20 µl reaction volumes containing 1 x *Taq* polymerase buffer 2.5 mM MgCl₂, 0.2 mM deoxynucleotide triphosphates, 10 pmol each of the forward and reverse primers, between 10 and 100 ng genomic DNA and 0.2 U of *Taq* polymerase (Fischer Biotech, Australia). Thermal cycling was performed in a GeneAmp PCR System 2400 Thermal cycler (Perkin Elmer Corporation, Norwalk, CT). Cycling began with a denaturing step at 94°C for 3 min followed by 30 cycles of denaturation at 94°C for 10 s, primer annealing between 55° and 65°C for 20 s and a DNA strand extension at 72°C for 1-3 min. Amplification was completed by a final extension step at 72°C for 7 min. Amplified DNA was separated by agarose gel electrophoresis in TAE buffer (40 mM Tris-acetate, 1 mM EDTA, pH 7.8), and visualized by UV transillumination after staining with ethidium bromide (0.5 µg/ml).

Automated DNA sequencing was performed using the PRISM Big Dye cycle sequencing system and a model 373 sequencer (Applied Biosystems, Foster City, CA). Sequence data were analyzed using ABI Prism-Autoassembler software, while identity/similarity values to other translated sequences were determined using BLAST in conjunction with the National Center for Biotechnology Information (NIH, Bethesda,

MD). Fugue blast (<http://www-cryst.bioc.cam.ac.uk/fugue/>) was used to identify distant homologs via sequence-structure comparisons. The gene clusters were assembled using the software Phred, Phrap, and Consed (<http://www.phrap.org/phredphrapconsed.html>), open reading frames were manually identified. Polyketide synthase and non-ribosomal peptide synthetase domains were determined using the specialized databases based on crystal structures (<http://www-ab.informatik.uni-tuebingen.de/software/NRPSpredictor>; <http://www.tigr.org/jravel/nrps/>, <http://www.nii.res.in/nrps-pks.html>).

10 **Example 10: Genetic screening of Cyindrospermopsin-producing and non-producing cyanobacterial strains**

Cyindrospermopsin-producing and non-producing cyanobacterial strains were screened for the presence of the sulfotransferase gene *cyrJ* using the primer set *cynsulfF* (5' ACTTCTCTCCTTTCCCTATC 3') (SEQ ID NO: 111) and *cynamR* (5' GAGTGAAAATGCGTAGAACTTG 3') (SEQ ID NO: 112). Genomic DNA was tested for positive amplification using the 16S rRNA gene primers 27F and 809 as described in Neilan et al. (1997) Int. J. Syst. Bacteriol. 47:693-697. Amplicons were sequenced, as described in Example 9 above, to verify the identity of the gene fragment.

The biosynthesis of cyindrospermopsin involves an amidinotransferase, a NRPS, and a PKS (*AoaA*, *AoaB* and *AoaC*, respectively). In order to obtain the entire sequence of the cyindrospermopsin biosynthesis gene cluster, we used adaptor-mediated 'gene-walking' technology, initiating the process from a partial sequence of the amidinotransferase gene from *C. raciborskii* AWT205. Successive outward facing primers were designed and the entire gene cluster spanning 43 kb was sequenced, together with a further 3.5 kb on either side of the toxin gene cluster.

25 These flanking regions encode putative accessory genes (*hyp* genes), which include molecular chaperons involved in the maturation of hydrogenases. Due to the fact that these genes are flanking the cyindrospermopsin gene cluster at both ends, we postulate that the toxin gene cluster was inserted into this area of the genome thus interrupting the HYP gene cluster. This genetic rearrangement is mechanistically supported by the presence of transposase-like sequences within the cyindrospermopsin cluster.

Bioinformatic analysis of the toxin gene cluster was performed and based on gene function inference using sequence alignments (NCBI BLAST), predicted structural homologies (Fugue Blast), and analysis of PKS and NRPS domains using specialized blast servers based on crystal structures. The cyindrospermopsin biosynthesis cluster

contains 15 ORFs, which encode all the functions required for the biosynthesis, regulation and export of the toxin cylindrospermopsin (Figure 6).

Example 11: Formation of the **CYR** carbon skeleton

5 The first step in formation of the carbon skeleton of cylindrospermopsin involves the synthesis of guanidinoacetate via transamidation of glycine. CyrA, the AoaA homolog, which encodes an amidinotransferase similar to the human arginine:glycine amidinotransferase GATM, transfers a guanidino group from a donor molecule, most likely arginine, onto an acceptor molecule of glycine thus forming guanidinoacetate
10 (Figure 8, step 1).

 The next step (Figure 8, step 2) in the biosynthesis is carried out by CyrB (AoaB homolog), a mixed NRPS-PKS. CyrB spans 8.7 kb and encodes the following domains; adenylation domain (A domain) and a peptidyl carrier protein (PCP) of an NRPS followed by a β ketosynthase domain (KS), acyltransferase domain (AT), dehydratase
15 domain (DH), methyltransferase domain (MT), ketoreductase domain (KR), and an acyl carrier protein (ACP) of PKS origin. CyrB therefore must catalyse the second reaction since it is the only gene containing an A domain that could recruit a starter unit for subsequent PKS extensions. The specific amino acid activated by the CyrB A domain cannot be predicted as its substrate specificity conferring residues do not match any in the
20 available databases (<http://www-ab.informatik.uni-tuebingen.de/software/NRPSpredictor/>; <http://www.tigr.org/jravel/nrps/>, <http://www.nii.res.in/nrps-pks.html>). So far, no other NRPS has been described that utilizes guanidinoacetate as a substrate. The A domain is thought to activate guanidinoacetate, which is then transferred via the swinging arm of the peptidyl carrier protein (PCP) to the KS domain. The AT domain activates malonyl-CoA and attaches it to the ACP. This is followed by a condensation reaction between the
25 activated guanidinoacetate and malonyl-CoA in the KS domain. CyrB contains two reducing modules, KR and DH. Their concerted reaction reduces the keto group to a hydroxyl followed by elimination of H₂O, resulting in a double bond between C13 and C14. The methyl transferase (MT) domain identified in CyrB via the NRPS/PKS
30 databases (Example 9 above), is homologous to S-adenosylmethionine (SAM) dependent MT. It is therefore suggested that the MT methylates C13. It is proposed that a nucleophilic attack of the amidino group at N19 onto the newly formed double bond between C13 and C14 occurs via a 'Michael addition'. The cyclization follows Baldwin's rules for ring closure (Baldwin et al. (1997) J. Org. Chem 42;3846-3852), resulting in the

formation of the first ring in cylindrospermopsin. This reaction could be spontaneous and may not require enzymatic catalysis, as it is energetically favourable. This is the first of three ring formations.

The third step (Figure 8, step 3) in the biosynthesis involves CyrC (AoaC homolog), which encodes a PKS with KS, AT, KR, and ACP domains. The action of these domains results in the elongation of the growing chain by an acetate via activation of malonyl-CoA by the AT domain, its transfer to ACP and condensation at the KS domain with the product of CyrB. The elongated chain is bound to the ACP of CyrC and the KR domain reduces the keto group to a hydroxyl group on C12. The PKS module carrying out this step contains a KR domain and does not contain a DH domain, this corresponds only to CyrC.

Following the catalysis of enzyme CyrC is CyrD (Figure 8, step 4), a PKS with five modules; KS, AT, DH, KR, and an ACP. The action of this PKS module on the product of CyrC results in the addition of one acetate and the reduction of the keto group on C10 to a hydroxyl and dehydration to a double bond between C9 and C10. This double bond is the site of a nucleophilic attack by the amidino group N19 via another Michael addition that again follows Baldwin's rules of ring closure, resulting in the formation of the second ring, the first 6-membered ring made in cylindrospermopsin.

The product of CyrD is the substrate for CyrE (step 5 in Figure 8), a PKS containing a KS, AT, DH, KR domains and an ACP. Since this sequence of domains is identical to that of CyrD, it is not possible at this stage to ascertain which PKS acts first, but as their action is proposed to be identical it is immaterial at this point. CyrE catalyzes the addition of one acetate and the formation of a double bond between C7 and C8. This double bond is attacked by N18 via a Michael addition and the third cyclisation occurs, resulting in the second 6-member ring.

CyrF is the final PKS module (step 6 of Figure 8) and is a minimal PKS containing only a KS, AT, and ACP. CyrF acts on the product of CyrE and elongates the chain by an acetate, leaving C4 and C6 unreduced.

Step 7 in the pathway (Figure 8) involves the formation of the uracil ring, a reaction that is required for the toxicity of the final cylindrospermopsin compound. The cylindrospermopsin gene cluster encodes two enzymes with high sequence similarity (87%) that have been denoted CyrG and CyrH. A Psi-blast search (NCBI) followed by a Fugue profile library search (see materials and methods) revealed that CyrG and CyrH are most similar to the enzyme family of amidohydrolases/ureases/dihydroorotases, whose

members catalyze the formation and cleavage of N-C bonds. It is proposed that these enzymes transfer a second guanidino group from a donor molecule, such as arginine or urea, onto C6 and C4 of cylindrospermopsin resulting in the formation of the uracil ring. These enzymes carry out two or three reactions depending on the guanidino donor. The first reaction consists of the formation of a covalent bond between the N of the guanidino donor and C6 of cylindrospermopsin followed by an elimination of H₂O forming a double bond between C5 and C6. The second reaction catalyses the formation of a bond between the second N on the guanidino donor and C4 of cylindrospermopsin, co-committently with the breaking of the thioester bond between the acyl carrier protein of CyrE and cylindrospermopsin, causing the release of the molecule from the enzyme complex. Feeding experiments with labeled acetate have shown that the oxygen at C4 is of acetate origin and is not lost during biosynthesis, therefore requiring the *de novo* formation of the uracil ring. The third reaction - if required - would catalyze the cleavage of the guanidino group from a donor molecule other than urea. The action of CyrG and CyrH in the formation of the uracil ring in cylindrospermopsin describes a novel biosynthesis pathway of a pyrimidine.

One theory suggest a linear polyketide which readily assumes a favorable conformation for the formation of the rings. Cyclization may thus be spontaneous and not under enzymatic control. These analyses show that this may happen step-wise, with successive ring formation of the appropriate intermediate as it is synthesized. This mechanism also explains the lack of a thioesterase or cyclization domain, which are usually associated with NRPS/PKS modules and catalyze the release and cyclization of the final product from the enzyme complex.

Example 12: CYR tailoring reactions

Cylindrospermopsin biosynthesis requires the action of tailoring enzymes in order to complete the biosynthesis, catalyzing the sulfation at C12 and hydroxylation at C7. Analysis of the cylindrospermopsin gene cluster revealed three candidate enzymes for the tailoring reactions involved in the biosynthesis of cylindrospermopsin, namely CyrI, CyrJ, and CyrN. The sulfation of cylindrospermopsin at C12 is likely to be carried out by the action of a sulfotransferase. CyrJ encodes a protein that is most similar to human 3'-phosphoadenylyl sulfate (PAPS) dependent sulfotransferases. The cylindrospermopsin gene cluster also encodes an adenylylsulfate kinase (ASK), namely CyrN. ASKs are enzymes that catalyze the formation of PAPS, which is the sulfate donor for

sulfotransferases. It is proposed that CyrJ sulfates cylindrospermopsin at C12 while CyrN creates the pool of PAPS required for this reaction. Screening of cylindrospermopsin producing and non-producing strains revealed that the sulfotransferase genes were only present in cylindrospermopsin producing strains, further affirming the involvement of this entire cluster in the biosynthesis of cylindrospermopsin (Figure 7). The *cyrJ* gene might therefore be a good candidate for a toxin probe, as it is more unique than NRPS and PKS genes and would presumably have less cross-reactivity with other gene clusters containing these genes, which are common in cyanobacteria. The final tailoring reaction is carried out by CyrI. A Fugue search and an iterated Psi-Blast revealed that CyrI is similar to a hydroxylase belonging to the 2-oxoglutarate and Fe(II)-dependent oxygenase superfamily, which includes the mammalian Prolyl 4-hydroxylase alpha subunit that catalyze the hydroxylation of collagen. It is proposed that CyrI catalyzes the hydroxylation of C7, a residue that, along with the uracil ring, seems to confer much of the toxicity of cylindrospermopsin. The hydroxylation at C7 by CyrI is probably the final step in the biosynthesis of cylindrospermopsin.

Example 13: CYR toxin transport

Cylindrospermopsin and other cyanobacterial toxins appear to be exported out of the producing cells. The cylindrospermopsin gene cluster contains an ORF denoted CyrK, the product of which is most similar to sodium ion driven multi-drug and toxic compound extrusion proteins (MATE) of the NorM family. It is postulated that CyrK is a transporter for cylindrospermopsin, based on this homology and its central location in the cluster. Heterologous expression and characterization of the protein are currently being undertaken to verify its putative role in cylindrospermopsin export.

Example 14: Transcriptional regulation of the toxin gene cluster

Cylindrospermopsin production has been shown to be highest when fixed nitrogen is eliminated from the growth media (Saker et al. (1999) J. Phycol 35:599-606). Flanking the cylindrospermopsin gene cluster are “*hyp*” gene homologs involved in the maturation of hydrogenases. In the cyanobacterium *Nostoc* PCC73102 they are under the regulation of the global nitrogen regulator NtcA, that activates transcription of nitrogen assimilation genes. It is plausible that the cylindrospermopsin gene cluster is under the same regulation, as it is located wholly within the “*hyp*” gene cluster in *C. raciborskii*

AWT205, and no obvious promoter region in the cylindrospermopsin gene cluster could be identified.

Finally, the cylindrospermopsin cluster also includes an ORF at its 3' -end designated CyrO. By homology, it encodes a hypothetical protein that appears to possess an ATP binding cassette, and is similar to WD repeat proteins, which have diverse regulatory and signal transduction roles. CyrO may also have a role in transcriptional regulation and DNA binding. It also shows homology to AAA family proteins that often perform chaperone-like functions and assist in the assembly, operation, or disassembly of protein complexes. Further insights into the role of CyrO are hindered due to low sequence homology with other proteins in databases.

The foregoing describes preferred forms of the present invention. It is to be understood that the present invention should not be restricted to the particular embodiment(s) shown above. Modifications and variations, obvious to those skilled in the art can be made thereto without departing from the scope of the present invention.

Related Application

This application claims benefit from Australian Provisional Application Number 2008902056 entitled "Detection of Cyanotoxic Organisms" which was filed on 24 April 2008, the entire contents of which are incorporated herein by reference.

CLAIMS:

1. An isolated polynucleotide comprising a nucleotide sequence sharing at least 90% sequence homology with SEQ ID NO: 1 or a fragment thereof, wherein said fragment encodes a protein of a saxitoxin biosynthetic pathway.

2. The polynucleotide according to claim 1, wherein said fragment comprises a nucleotide sequence sharing at least 90% sequence homology with a sequence selected from the group consisting of SEQ ID NO: 2, SEQ ID NO: 4, SEQ ID NO: 6, SEQ ID NO: 8, SEQ ID NO: 10, SEQ ID NO: 12, SEQ ID NO: 14, SEQ ID NO: 16, SEQ ID NO: 18, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 26, SEQ ID NO: 28, SEQ ID NO: 30, SEQ ID NO: 32, SEQ ID NO: 34, SEQ ID NO: 36, SEQ ID NO: 38, SEQ ID NO: 40, SEQ ID NO: 42, SEQ ID NO: 44, SEQ ID NO: 46, SEQ ID NO: 48, SEQ ID NO: 50, SEQ ID NO: 52, SEQ ID NO: 54, SEQ ID NO: 56, SEQ ID NO: 58, SEQ ID NO: 60, SEQ ID NO: 62, SEQ ID NO: 64, SEQ ID NO: 66, and SEQ ID NO: 68.

3. An isolated ribonucleic acid or an isolated complementary DNA encoded by a sequence according to claim 1 or claim 2.

4. An isolated saxitoxin biosynthetic pathway polypeptide comprising an amino acid sequence sharing at least 90% sequence homology with a sequence selected from the group consisting of SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 11, SEQ ID NO: 13, SEQ ID NO: 15, SEQ ID NO: 17, SEQ ID NO: 19, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 27, SEQ ID NO: 29, SEQ ID NO: 31, SEQ ID NO: 33, SEQ ID NO: 35, SEQ ID NO: 37, SEQ ID NO: 39, SEQ ID NO: 41, SEQ ID NO: 43, SEQ ID NO: 45, SEQ ID NO: 47, SEQ ID NO: 49, SEQ ID NO: 51, SEQ ID NO: 53, SEQ ID NO: 55, SEQ ID NO: 57, SEQ ID NO: 59, SEQ ID NO: 61, SEQ ID NO: 63, SEQ ID NO: 65, SEQ ID NO: 67, and SEQ ID NO: 69.

5. A probe or primer that hybridises specifically with one or more of:

- (i) a polynucleotide according to claim 1 or 2,
- (ii) a ribonucleic acid or complementary DNA according to claim 3,
- (iii) a polypeptide according to claim 4.

6. A vector comprising a polynucleotide according to claim 1 or claim 2, or a ribonucleic acid or complementary DNA according to claim 3.

7. A host cell comprising the vector according to claim 6.

8. A method for the detection of cyanobacteria and/or dinoflagellates, the method comprising the steps of obtaining a sample for use in the method and analyzing the sample for the presence of one or more of:

- (i) a polynucleotide comprising a sequence according to claim 1 or 2,

(ii) a ribonucleic acid or complementary DNA according to claim 3,

(iii) a polypeptide comprising a sequence according to claim 4,

wherein said presence is indicative of cyanobacteria and/or dinoflagellates in the sample.

9. A method for detecting a cyanotoxic organism, the method comprising the steps of obtaining a sample for use in the method and analyzing the sample for the presence of one or more of:

(i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 36, and variants thereof sharing at least 80% sequence homology with SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, or SEQ ID NO: 36,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 37, and variants thereof sharing at least 80% sequence homology with SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, or SEQ ID NO: 37,

wherein said presence is indicative of cyanotoxic organisms in the sample.

10. The method according to claim 9, wherein said cyanotoxic organism is a cyanobacterium or a dinoflagellate.

11. The method according to any one of claims 8 to 10, further comprising analyzing the sample for the presence of one or more of:

(i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants and fragments thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, and SEQ ID NO: 110, and variants and fragments thereof.

12. The method according to any one of claims 8 to 11, wherein said analyzing comprises amplification of DNA from the sample by polymerase chain reaction.

13. The method according to claim 12, wherein said polymerase chain reaction utilises:

(i) one or more primers comprising a sequence selected from the group consisting of SEQ ID NO: 111, SEQ ID NO: 112, and variants and fragments thereof; and/or

(ii) one or more primers comprising a sequence selected from the group consisting of SEQ ID NO: 70, SEQ ID NO: 71, SEQ ID NO: 72, SEQ ID NO: 73, SEQ ID NO: 74, SEQ ID NO: 75, SEQ ID NO: 76, SEQ ID NO: 77, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 113, SEQ ID NO: 114, SEQ ID NO: 115, SEQ ID NO: 116, SEQ ID NO: 117, SEQ ID NO: 118, SEQ ID NO: 119, SEQ ID NO: 120, SEQ ID NO: 121, SEQ ID NO: 122, SEQ ID NO: 123, SEQ ID NO: 124, SEQ ID NO: 125, SEQ ID NO: 126, SEQ ID NO: 127, SEQ ID NO: 128, SEQ ID NO: 129, SEQ ID NO: 130, SEQ ID NO: 131, SEQ ID NO: 132, SEQ ID NO: 133, SEQ ID NO: 134, and variants and fragments thereof.

14. An isolated antibody capable of binding specifically to a polypeptide according to claim 4.

15. A kit for the detection of cyanobacteria and/or dinoflagellates, the kit comprising at least one agent for detecting the presence of one or more of:

- (i) a polynucleotide comprising a sequence according to claim 1 or 2,
- (ii) a ribonucleic acid or complementary DNA according to claim 3,
- (iii) a polypeptide comprising a sequence according to claim 4,

wherein said presence is indicative of cyanobacteria and/or dinoflagellates in the sample.

16. A kit for the detection of cyanotoxic organisms, the kit comprising at least one agent for detecting the presence of one or more of:

(i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, SEQ ID NO: 36, and variants thereof sharing at least 90% sequence homology with SEQ ID NO: 14, SEQ ID NO: 20, SEQ ID NO: 22, SEQ ID NO: 24, or SEQ ID NO: 36,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, SEQ ID NO: 37, and variants thereof sharing at least 90% sequence homology with SEQ ID NO: 15, SEQ ID NO: 21, SEQ ID NO: 23, SEQ ID NO: 25, or SEQ ID NO: 37,

wherein said presence is indicative of cyanotoxic organisms in the sample.

17. The kit according to claim 15 or claim 16, further comprising at least one additional agent for detecting the presence of one or more of:

(i) a polynucleotide comprising a sequence selected from the group consisting of: SEQ ID NO: 80, SEQ ID NO: 81, SEQ ID NO: 83, SEQ ID NO: 85, SEQ ID NO: 87, SEQ ID NO: 89, SEQ ID NO: 91, SEQ ID NO: 93, SEQ ID NO: 95, SEQ ID NO: 97, SEQ ID NO: 99, SEQ ID NO: 101, SEQ ID NO: 103, SEQ ID NO: 105, SEQ ID NO: 107, SEQ ID NO: 109, and variants and fragments thereof,

(ii) a ribonucleic acid or complementary DNA encoded by a sequence according to (i),

(iii) a polypeptide comprising a sequence selected from the group consisting of: SEQ ID NO: 82, SEQ ID NO: 84, SEQ ID NO: 86, SEQ ID NO: 88, SEQ ID NO: 90, SEQ ID NO: 92, SEQ ID NO: 94, SEQ ID NO: 96, SEQ ID NO: 98, SEQ ID NO: 100, SEQ ID NO: 102, SEQ ID NO: 104, SEQ ID NO: 106, SEQ ID NO: 108, and SEQ ID NO: 110, and variants and fragments thereof.

18. The kit according to any one of claims 15 to 17, wherein said at least one additional agent is a primer, antibody or probe.

19. The kit according to claim 18, wherein said primer or probe comprises:

(i) a sequence selected from the group consisting of SEQ ID NO: 109, SEQ ID NO: 110, and variants and fragments thereof; or

(ii) a sequence selected from the group consisting of SEQ ID NO: 70, SEQ ID NO: 71, SEQ ID NO: 72, SEQ ID NO: 73, SEQ ID NO: 74, SEQ ID NO: 75, SEQ ID NO: 76, SEQ ID NO: 77, SEQ ID NO: 78, SEQ ID NO: 79, SEQ ID NO: 113, SEQ ID NO: 114, SEQ ID NO: 115, SEQ ID NO: 116, SEQ ID NO: 117, SEQ ID NO: 118, SEQ ID NO: 119, SEQ ID NO: 120, SEQ ID NO: 121, SEQ ID NO: 122, SEQ ID NO: 123, SEQ ID NO: 124, SEQ ID NO: 125, SEQ ID NO: 126, SEQ ID NO: 127, SEQ ID NO: 128, SEQ ID NO: 129, SEQ ID NO: 130, SEQ ID NO: 131, SEQ ID NO: 132, SEQ ID NO: 133, SEQ ID NO: 134, and variants and fragments thereof.

20. A method of screening for a compound that modulates the expression or activity of one or more polypeptides according to claim 4, the method comprising:

contacting the polypeptide with a candidate compound under conditions suitable to enable interaction of the candidate compound and the polypeptide; and

assaying for activity of the polypeptide.

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Cyanobacteria Strains	Toxicity (Ref)	svtA	svtG	svtH	svtI	svtX
<i>A. circinalis</i> AWQC118C	PSP (54)	+	+	+	+	-
<i>A. circinalis</i> AWQC131C	PSP (25)	+	+	+	+	-
<i>A. circinalis</i> AWQC134C	PSP (54)	+	+	+	+	-
<i>A. circinalis</i> AWQC150E	PSP (54)	+	+	+	+	-
<i>A. circinalis</i> AWQC173A	PSP (54)	+	+	+	+	-
<i>A. circinalis</i> AWQC271C	- (54)	-	-	-	-	-
<i>A. circinalis</i> AWQC306A	- (54)	-	-	-	-	-
<i>A. circinalis</i> AWQC310F	- (54)	-	-	-	-	-
<i>A. circinalis</i> AWQC342D	- (54)	-	-	-	-	-
<i>Aph. flos-aquae</i> NH-5	PSP (26)	+	+	+	+	+
<i>Aph. ovalisporum</i> APH028A	CYLN (46)	-	-	-	-	-
<i>C. raciborskii</i> T3	PSP (23)	+	+	+	+	+
<i>C. raciborskii</i> 23B	CYLN (58)	-	-	-	-	-
<i>C. raciborskii</i> GOON	CYLN (43)	-	-	-	-	-
<i>C. raciborskii</i> GERM1	- (30)	-	-	-	-	-
<i>C. raciborskii</i> MARAU1	- (30)	-	-	-	-	-
<i>L. wollei</i>	PSP (7)	+	+	+	+	+

FIGURE 1A

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Primer	From	To	Direction	Sequence	Gene	
SEQ ID NO:133	1917	1937	→	GCAAAATTTTCAGGAGTAAAG	sterole desaturase	sxtD
SEQ ID NO:134	2744	2763	→	AGAGAATGCTATGCTTTTCAA	InsB	orf3
SEQ ID NO:135	2889	2911	→	TTTTGGGTAAACTTTATAGCCAT	InsB	orf3
SEQ ID NO:136	3020	3041	←	IGGGTCTGGACAGTTGTAGATA	InsA	orf4
SEQ ID NO:137	3306	3328	←	AAGGGGAAAACAAAATTATCAAT	InsA	orf4
SEQ ID NO:138	3396	3415	→	GGCGATCGCCIGCTAAAAAT	InsA	orf4
SEQ ID NO:139	3717	3739	→	CCCTATTTCATTTCTAGACGTT	SPUR	sxtC
SEQ ID NO:140	4201	4220	→	CCACTTCAACTAAAACAGCA	cytidine deaminase	sxtB
SEQ ID NO:141	4362	4381	←	AAAAATTTTGGAGGGTATGC	cytidine deaminase	sxtB
SEQ ID NO:142	4932	4951	→	ATCCAAGATGGACACACACT	cytidine deaminase	sxtB
SEQ ID NO: 70	5193	5212	→	TTAATTGCTTGGTCTATCTC	PKS	sxtA
SEQ ID NO: 71	5206	5225	←	CAATACCGAAGAGGAGATAG	PKS	sxtA
SEQ ID NO: 72	5345	5364	←	TAGCGTGTTAGTGGGAGAT	PKS	sxtA
SEQ ID NO: 73	5415	5434	→	TGTGTAAACCAATTGTGTAGT	PKS	sxtA
SEQ ID NO: 74	5478	5497	←	TTAGCCGGATTACAGGTGA	PKS	sxtA
SEQ ID NO: 75	6136	6155	←	CTGGACTCGGCTTGTGTCTT	PKS	sxtA
SEQ ID NO: 76	6933	6952	→	CAGCGAGTTACACCCACCAC	PKS	sxtA
SEQ ID NO: 76	7035	7054	←	CTCGCACTAAATATTTCTACC	PKS	sxtA
SEQ ID NO: 78	7434	7452	→	AAAACCTCAGCTTCCACAA	PKS	sxtA
SEQ ID NO: 79	7537	7558	←	ATGATTTTGGAGGTCCATTGTT	PKS	sxtA
SEQ ID NO: 113	7820	7841	→	CCCAATATCTCCCTGTAAACT	PKS	sxtA
SEQ ID NO: 114	8170	8189	←	TGGCAATTGTCTCTCCGTAT	PKS	sxtA
SEQ ID NO: 115	8742	8761	←	CTCGCCGATGAAGAATCTCTCT	PKS	sxtA
SEQ ID NO: 116	8772	8791	←	GCGTGTGAGAGAAAGGTGT	PKS	sxtA
SEQ ID NO: 117	8782	8801	→	CTCGACACGCAGAAATACG	PKS	sxtA
SEQ ID NO: 143	9390	9410	→	GGTCCCTTGGCAGATAGAGTG	chaperon-like	sxtE
SEQ ID NO: 144	9390	9410	←	CACICTATCTGCGCAAGGACC	chaperon-like	sxtE
SEQ ID NO: 145	9856	9876	←	TGACICGATTCGCTGTATAAA	MATE I	sxtF
SEQ ID NO: 118	10080	10100	→	ATGCTTCTGCTTGGCATGGC	amidinotransferase	sxtG
SEQ ID NO: 119	11468	11488	←	TAACTCGACGAACTTTGACCC	amidinotransferase	sxtG

FIGURE 1B

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Primer	From	To	Direction	Sequence	Gene	
SEQ ID NO: 120	11551	11569	-->	GCCGCCAATCTCGCGATG	amidontransferase	sxtG
SEQ ID NO: 121	12256	12277	<--	GAACGTCTAATGTTGCACAGTG	amidontransferase	sxtG
SEQ ID NO: 122	12410	12432	-->	CTGGTACGTAGTCGCAAGGTGG	dioxygenase I	sxtH
SEQ ID NO: 123	13292	13317	-->	CTGACGGTACATGATTTCTCTGTGAC	dioxygenase I	sxtH
SEQ ID NO: 124	13540	13561	-->	egtcctATGCGAGATCTTAGGAATTCAG	carbamoyltransferase	sxtI
SEQ ID NO: 125	13561	13585	-->	GCTTACTACCGACGATAGTGCTGCCG	carbamoyltransferase	sxtI
SEQ ID NO: 126	14451	14472	-->	TCTATGTTAGCAGGTGGTGTCTC	carbamoyltransferase	sxtI
SEQ ID NO: 127	14735	14754	<--	TTCTGCAAGACGAGCCATAA	carbamoyltransferase	sxtI
SEQ ID NO: 128	15211	15230	<--	GGTTCGCCGCGGACATTAAA	carbamoyltransferase	sxtI
SEQ ID NO: 146	15709	15730	-->	TTCATAAGACGGCTGTTGAATC	hypothetical protein	sxtJ
SEQ ID NO: 147	15966	15989	<--	ctcgagTTAAAAAGAGGTGTAATGAAAGG	hypothetical protein	sxtK
SEQ ID NO: 148	16326	16348	<--	TTCTATAACTGCTGCCAAATTTT	GDSL-lipase	sxtL
SEQ ID NO: 149	16400	16422	-->	AATTTTGGAGTGACTGGTTATGG	GDSL-lipase	sxtL
SEQ ID NO: 150	16400	16422	<--	CCATAACCAGTCACTCCCAAATTT	GDSL-lipase	sxtL
SEQ ID NO: 151	16929	16949	-->	TTTTAGTTGTTACTTTTGGCG	GDSL-lipase	sxtL
SEQ ID NO: 152	17215	17234	-->	ACAGCAGATGAGAGAAAAGTA	GDSL-lipase	sxtL
SEQ ID NO: 153	18054	18073	-->	GGGTTGCTTGTGCTGATTTTC	MATE II	sxtM
SEQ ID NO: 154	18721	18742	<--	CATTAAATAAGTCCGGACAGG	MATE II	sxtM
SEQ ID NO: 155	19133	19152	<--	TTAAACAGAATGAGGAGCAA	MATE II	sxtM
SEQ ID NO: 156	19260	19279	<--	AAACAACACACCCATCTAAG	sulfotransferase	sxtN
SEQ ID NO: 157	19531	19550	-->	TTAATAAGGCATCCCCAAGA	sulfotransferase	sxtN
SEQ ID NO: 158	19728	19747	<--	GAAATGGCTGTGTA AAAACT	sulfotransferase	sxtN
SEQ ID NO: 129	20584	20603	<--	ATGCTAATGCGGTGGGAGTA	cephalosporin hydroxylase	sxtX
SEQ ID NO: 130	20643	20662	-->	AAAGCAGTCCGACGACATTT	cephalosporin hydroxylase	sxtX
SEQ ID NO: 131	20831	20853	-->	CCTATTTGATTATTGTTTTCGG	cephalosporin hydroxylase	sxtX
SEQ ID NO: 132	21252	21271	<--	GATACCGATCATAACTACG	cephalosporin hydroxylase	sxtX
SEQ ID NO: 159	21290	21309	-->	TCTGCCATATCCCCAACCTA	cephalosporin hydroxylase	sxtX
SEQ ID NO: 160	21445	21464	<--	GATCGCCCGACAGGAAGACT	ferredoxin	sxtW
SEQ ID NO: 161	22020	22039	-->	TCCGGCTTGACCTGCTGGAC	ferredoxin	sxtW
			-->		succinate dehydrogenase	sxtV

FIGURE 1B (cont)

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Primer	From	To	Direction	Sequence	Gene	
SEQ ID NO: 162	22715	22734	-->	TGCGATGATTTTGCTCTGT	succinate dehydrogenase	sxtV
SEQ ID NO: 163	22801	22820	-->	AAATTTGCACACCCACAG	succinate dehydrogenase	sxtV
SEQ ID NO: 164	22942	22968	-->	TTGGATTGAACGTGTAATTGAAAAGC	succinate dehydrogenase	sxtV
SEQ ID NO: 186	22942	22968	<--	GCTTTTTCATTAACACGTTCAATCCAA	succinate dehydrogenase	sxtV
SEQ ID NO: 165	23434	23453	<--	GTTTAGTCGATACGCCATT	succinate dehydrogenase	sxtV
SEQ ID NO: 166	23434	23453	-->	AAATGGCGTATCGACTAAC	succinate dehydrogenase	sxtV
SEQ ID NO: 167	24095	24115	-->	ATATAGGAGCGCATAAAGTGC	succinate dehydrogenase	sxtV
SEQ ID NO: 168	24728	24747	-->	CTTGGTATAAGTCTTTGTGAT	dioxygenase II	sxtT
SEQ ID NO: 169	25426	25445	<--	AACACTCATTAGATTCACT	phytanoyl-CoA dioxygenase	sxtS
SEQ ID NO: 170	25979	25999	<--	TCCACTAAATCCTTTGAATTG	phytanoyl-CoA dioxygenase	sxtS
SEQ ID NO: 171	26279	26299	-->	TGTTTGTCTGGATGCGATCCT	unknown protein	orf24
SEQ ID NO: 172	26451	26470	-->	GCAGTTCAGGTCCATGAAAC	unknown protein	orf25
SEQ ID NO: 173	27155	27174	-->	AGCCCAAGTCACAACCTTCGT	GNAT transferase	sxtR
SEQ ID NO: 174	27508	27528	-->	TCTGGAAGTACTTGCACTGTC	unknown protein	sxtQ
SEQ ID NO: 175	28197	28218	-->	TGTAACCTCCGTCAGGACATAAA	unknown protein	sxtQ
SEQ ID NO: 176	28395	28417	<--	TGCAAAATTTTAGTAGCAATAACG	RTX-toxin like	sxtP
SEQ ID NO: 177	29532	29558	<--	CTTTACTAATTATAGCGGGATATTAT	RTX-toxin like	sxtP
SEQ ID NO: 178	29868	29887	<--	CAGTGGGAAATAGATGGAT	adenylylsulphate kinase	sxtO
SEQ ID NO: 179	30249	30268	<--	TGGTCATAAAAGCGGGATTC	adenylylsulphate kinase	sxtO
SEQ ID NO: 180	31745	31762	-->	GGATCTTGGCGCAATTTA	IS4	orf29
SEQ ID NO: 181	33031	33053	<--	GTTAGAGACTTGGAAACGTATTGG	PhoU	sxtY
SEQ ID NO: 182	34711	34729	-->	CCAAACCCAGAAAGAAATCC	histidine kinase	sxtZ
SEQ ID NO: 183	35100	35121	-->	AATCTATAGCCAAACCCCTAA	ribotide isomerase	
SEQ ID NO: 184	36447	36465	-->	ACTGTGTGAACAAATTCCCC	ribotide isomerase	
SEQ ID NO: 185	36652	36680	-->	GCAACAAGACTACATTTAGTAGATTTAGA	ribotide isomerase	

FIGURE 1B (cont)

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Name	Enzyme Family	Size (bp)	Blast Similarity Match	(%)	Putative Function
<i>orf1</i>	unknown protein	1320	BAB76734.1 <i>Noroc PCC7120</i>	82	unknown
<i>orfD</i>	sterol desaturase-like	759	ABG52264.1 <i>Trichodesmium erythraeum</i>	63	desaturation
<i>orf3</i>	transposase InsB	392	CAE11915.2 <i>Microcystis aeruginosa</i>	86	transposition
<i>orf4</i>	transposase InsA	360	CAE11914.1 <i>Microcystis aeruginosa</i>	71	transposition
<i>orfC</i>	unknown protein	354	no similarity found		regulatory
<i>orfB</i>	cytidine deaminase	957	EAS64681.1 <i>Vibrio angustum</i>	62	cyclisation
<i>orfA</i>	methyltransferase	1506	ABF89568.1 <i>Mycrococcus xanthus</i>	64	methylation
	GNAT	633	AAT70096.1 <i>CitA Lyngbya majuscula</i>	64	loading of ACP
	acyl carrier protein	324	AAV97870 <i>OmnB Theonella swinhoei</i>	59	ACP
	AONS	1275	ABD13093.1 <i>Frankia</i> sp. Ccl3	61	Claisen condensation
<i>orfE</i>	unknown protein	387	ABE53436.1 <i>Shewanella denitrificans</i>	52	unknown
<i>orfF</i>	MATE	1416	NCIM ABC44739.1 <i>Salinibacter ruber</i>	52	export of PST's
<i>orfG</i>	amidinotransferase	1134	ABA05575.1 <i>Nitrobacter winogradskyi</i>	71	amidinotransfer
<i>orfH</i>	phenylpropionate dioxygenase	1005	ZP_00243439.1 <i>Rubrivivax gelatinosus</i>	50	C-12 hydroxylation
<i>orfI</i>	carbamoyltransferase	1839	ABG50968.1 <i>Trichodesmium erythraeum</i>	82	carbamoylation

FIGURE 2

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<i>xxJ</i>	unknown protein	444	EAM51043.1 <i>Crocospira waizonii</i>	72	regulatory
<i>xxK</i>	unknown protein	165	ABG50954.1 <i>Trichodesmium erythraeum</i>	81	regulatory
<i>xxL</i>	GDSL lipase	1299	ABG50952.1 <i>Trichodesmium erythraeum</i>	60	cyclisation
<i>xxM</i>	MATE	1449	NorM ABC44739.1 <i>Salinibacter ruber</i>	53	export of PSTs
<i>xxN</i>	sulfotransferase	831	ABG53102.1 <i>Trichodesmium erythraeum</i>	57	sulfotransfer
<i>xxX</i>	cephalosporin hydroxylase	774	ABG50679.1 <i>Trichodesmium erythraeum</i>	77	N-1 hydroxylation
<i>xxW</i>	ferredoxin	327	ZP_00106179.2 <i>Nostoc punctiforme</i>	99	electron carrier
<i>xxV</i>	succinate dehydrogenase	1653	ABA24604.1 <i>Anabaena variabilis</i>	92	dioxygenase reductase
<i>xxU</i>	alcohol dehydrogenase	750	ZP_00111652.1 <i>Nostoc punctiforme</i>	83	reduction of C-1
<i>xxT</i>	phenylpropionate dioxygenase	1005	ZP_00243439.1 <i>Rubrivivax gelatinosus</i>	48	C-12 hydroxylation
<i>xxS</i>	phytanoyl-CoA dioxygenase	726	ABG30370.1 <i>Roseobacter denitrificans</i>	41	ring formation
<i>orf24</i>	unknown protein	576	no similarity found		unknown
<i>xxR</i>	acyl transferase	777	AAU26161.1 <i>Legionella pneumophila</i>	54	unknown
<i>xxQ</i>	unknown protein	777	EAR64935.1 <i>Bacillus</i> sp. NRRL B-14911	46	unknown
<i>xxP</i>	RTX-toxin	1227	ABA20206.1 <i>Anabaena variabilis</i>	68	binding of PSTs
<i>xxO</i>	adenylylsulfate kinase	603	ZP_00053494.2 <i>Magnetospirillum magnetotacticum</i>	76	PAPS biosynthesis
<i>orf29</i>	transposase, IS4	1350	EA022367.1 <i>Synitrophobacter fumaroxidans</i>	61	transposition
<i>xxY</i>	PhoU	666	BAB76300.1 <i>Nostoc PCC7120</i>	87	signal transduction
<i>xxZ</i>	histidine kinase	1353	ABA22975.1 <i>Anabaena variabilis</i>	78	signal transduction
<i>ompR</i>	OmpR	819	ZP_00108178.2 <i>Nostoc punctiforme</i>	91	signal transduction
<i>hisA</i>	PROFAR isomerase	774	ABA22979.1 <i>Anabaena variabilis</i>	90	histidine biosynthesis
<i>orf34</i>	unknown protein	396	ZP_00343366.1 <i>Nostoc punctiforme</i>	84	unknown

FIGURE 2 (cont)

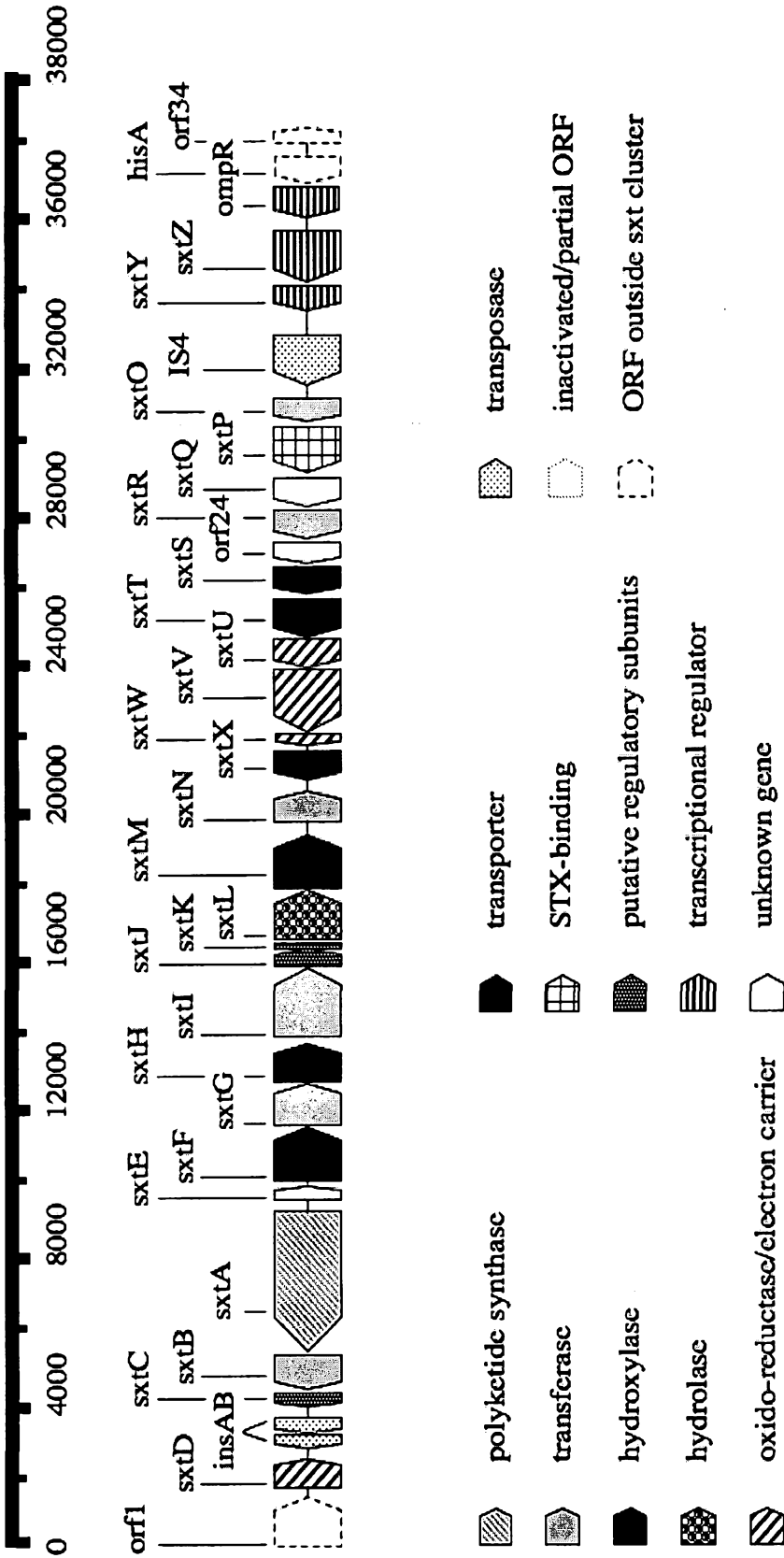
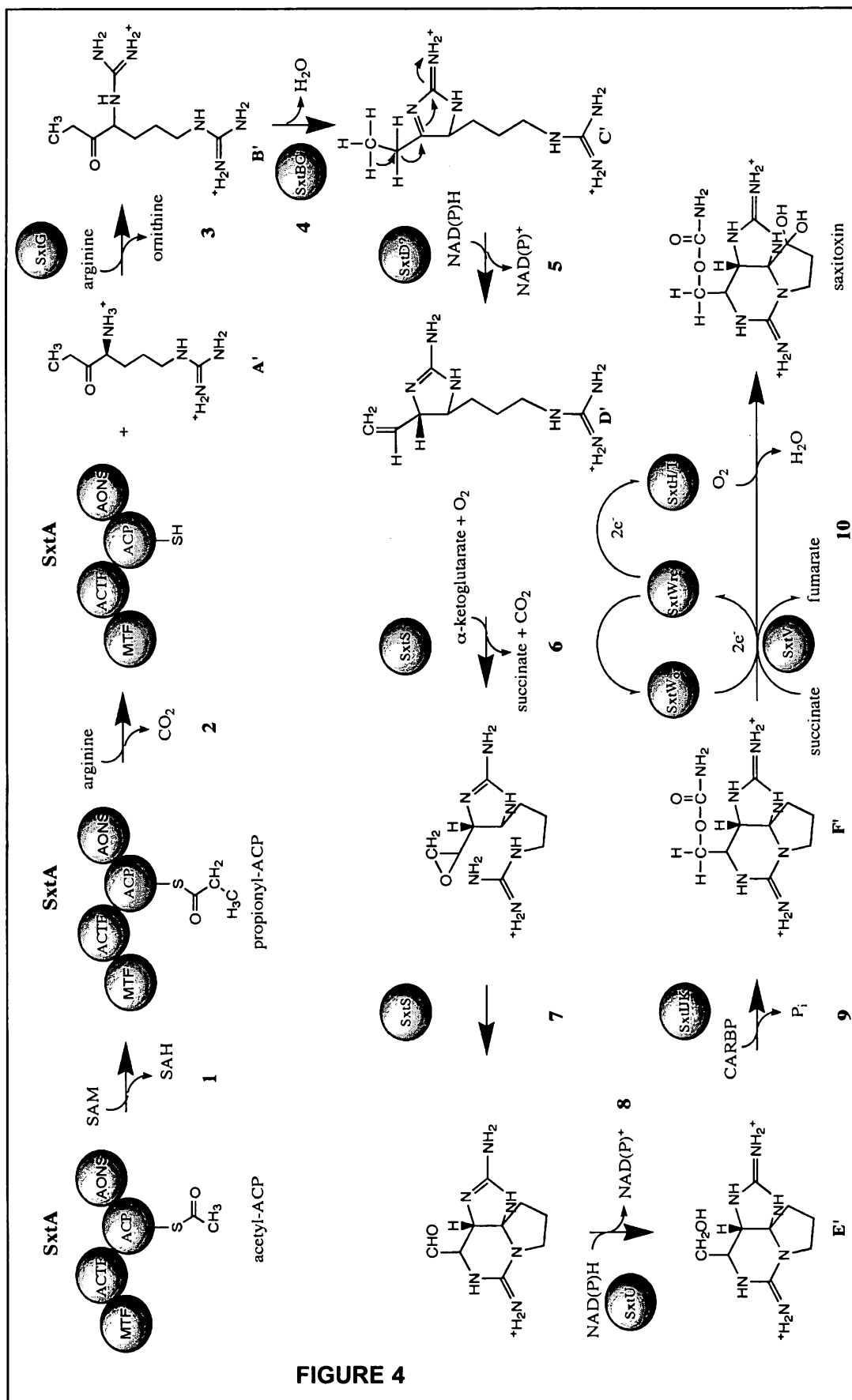


FIGURE 3

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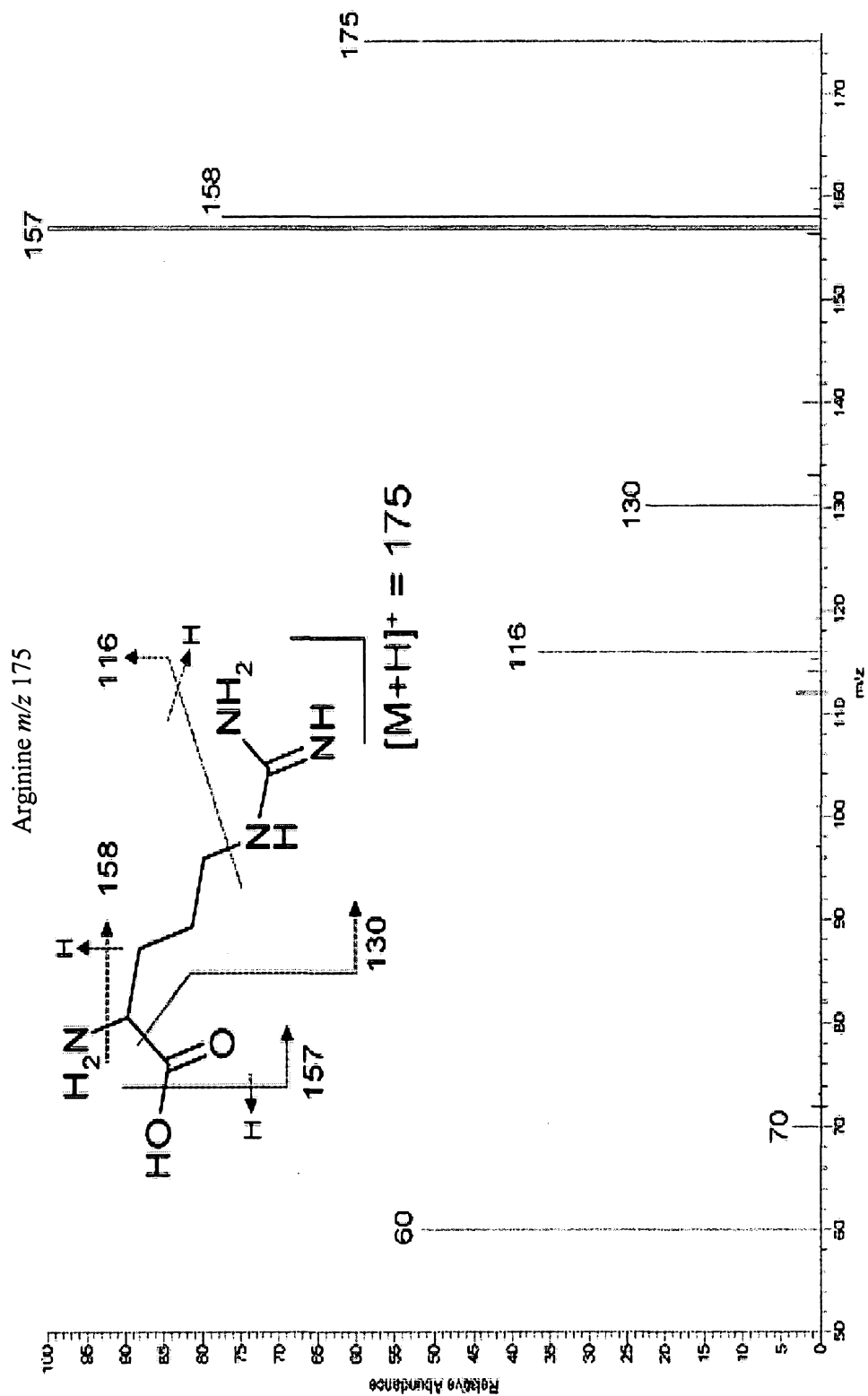


FIGURE 5A

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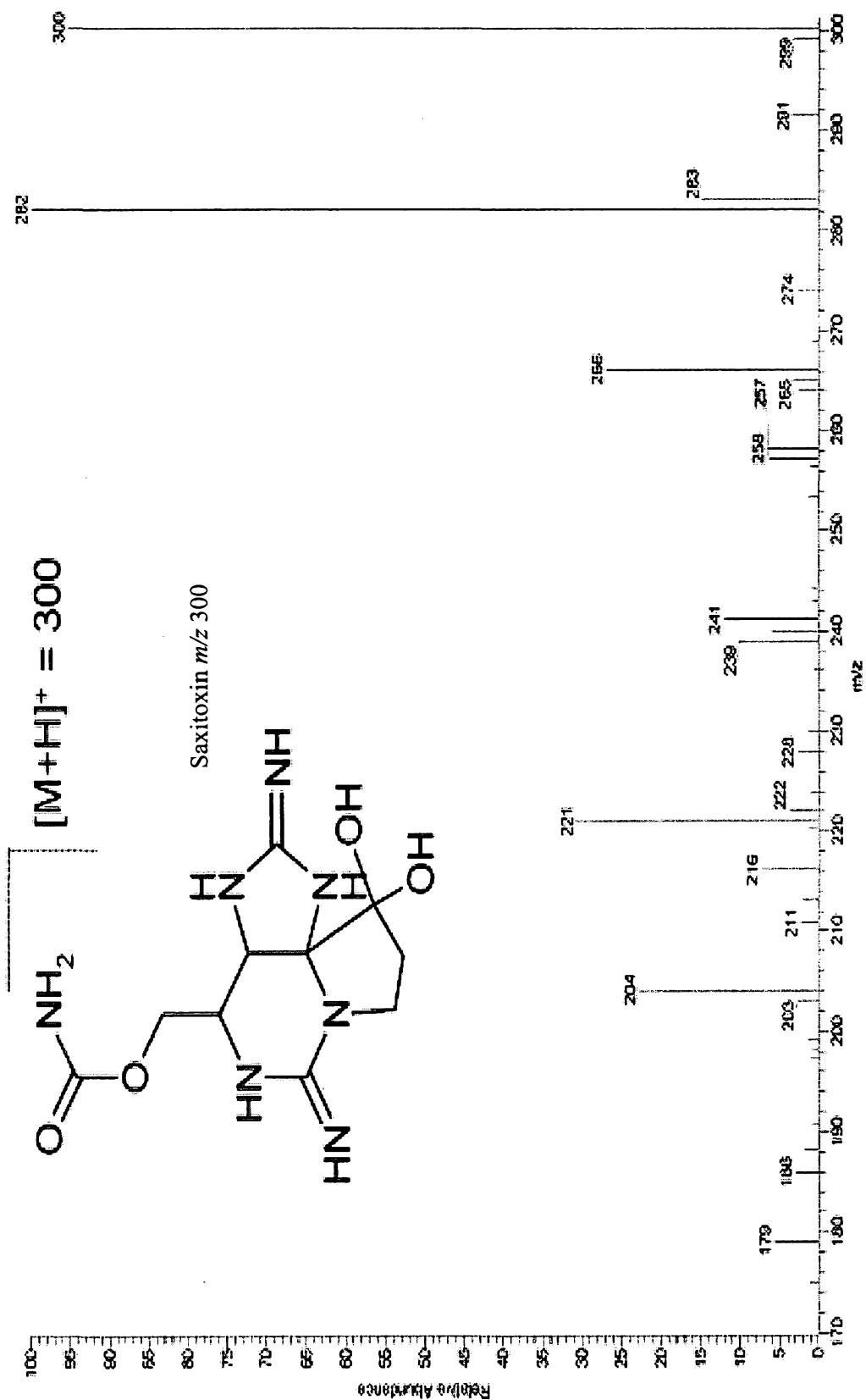


FIGURE 5B

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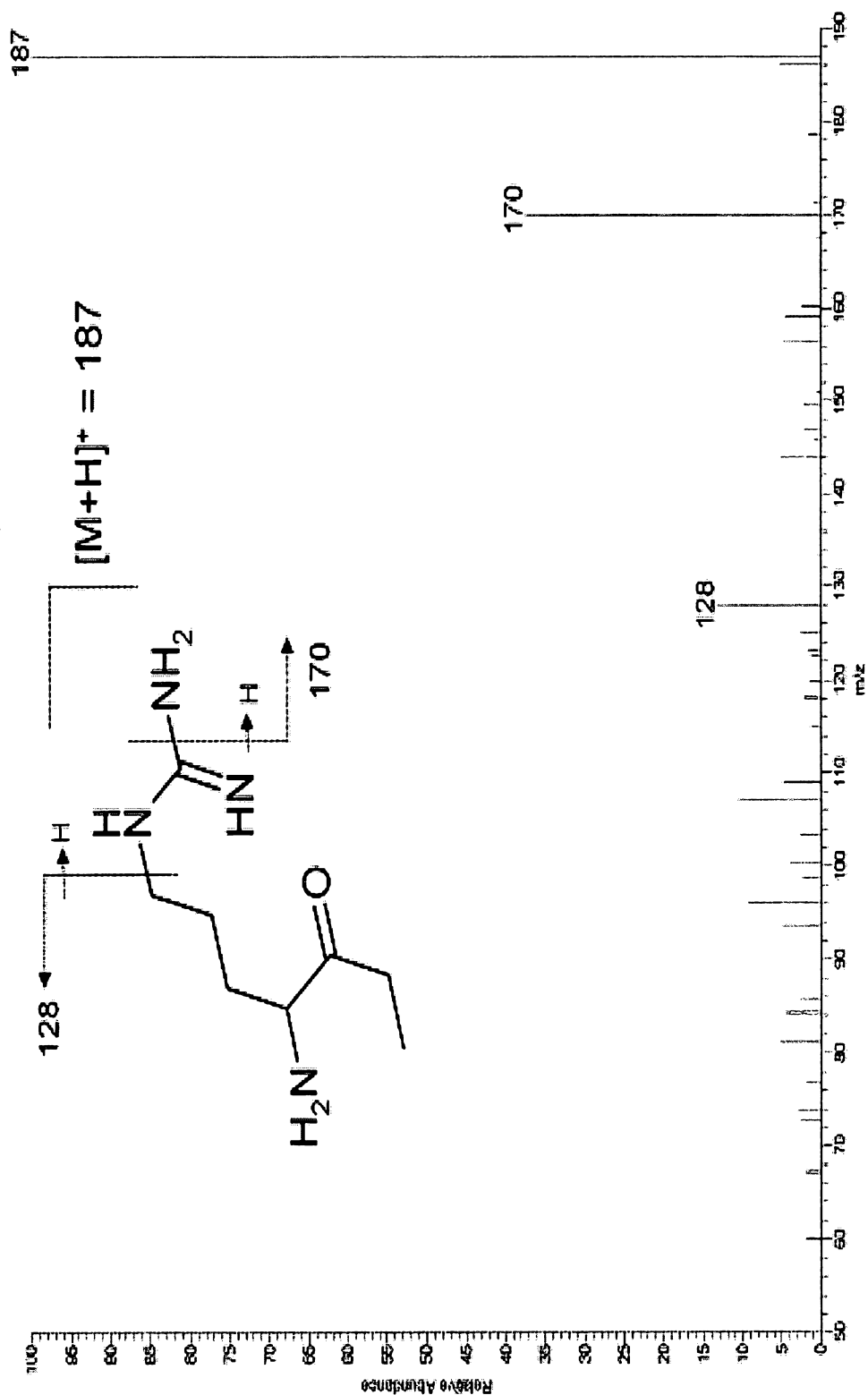


FIGURE 5C

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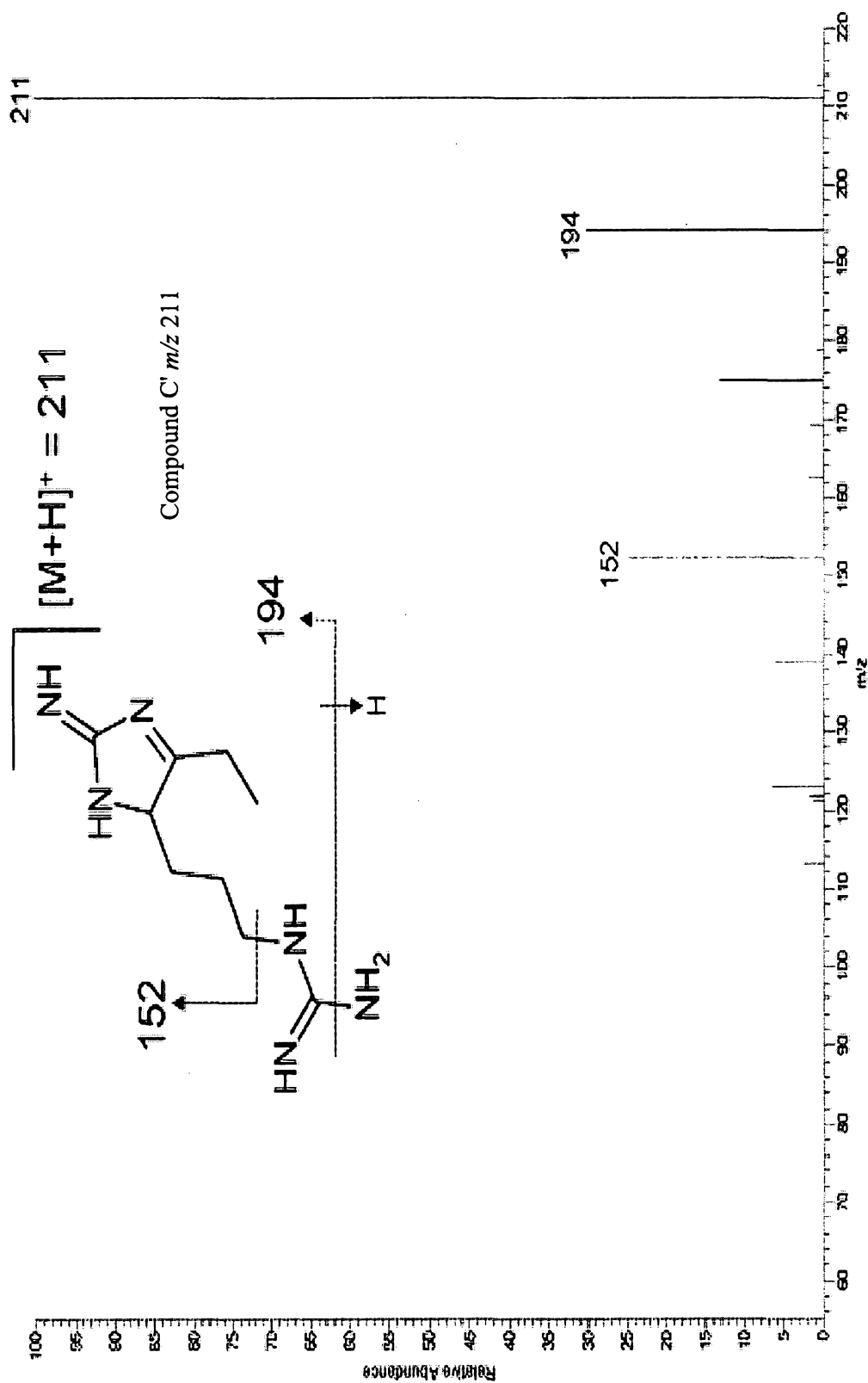


FIGURE 5D

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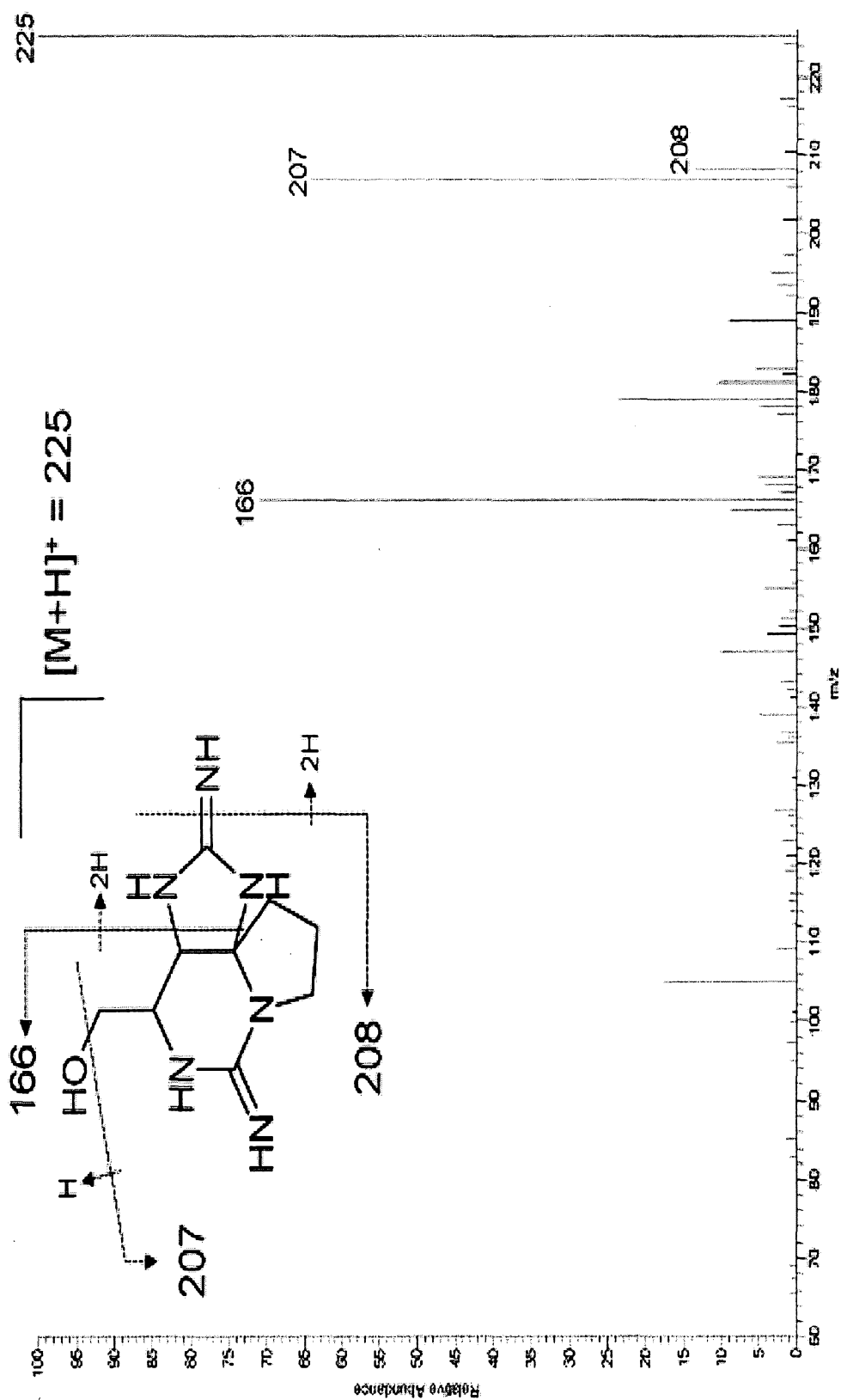


FIGURE 5E

Name	Enzyme Family	Size (bp)	Psi-Blast similarity match	% ID	Putative function
<i>cyrD</i>	PKS CrpB	5631	ABM21570.1 <i>Nostoc</i> sp. ATCC 53789	58	PKS KS-AT-DH-KR-ACP
<i>cyrF</i>	PKS CrpB	4074	ABM21570.1 <i>Nostoc</i> sp. ATCC 53789	68	PKS KS-AT-ACP
<i>cyrG</i>	cytosine deaminase /Aminohydrolase/ Dihydroorotase	1437	BAF59909.1 <i>Pelotomaculum thermopropionicum</i> SI	50	Uracil ring formation
<i>cyrI</i>	Prolyl 4-Hydroxylase	831	ABB06365.1 <i>Burkholderia</i> sp. 383	43	Hydroxylation of C7
<i>cyrK</i>	MatE Na ⁺ -driven multidrug efflux pump	1398	EAW39051.1 <i>Lyngbya</i> sp. PCC 8106	65	Exporter
<i>cyrL</i>	Transposase	750	ABG50981.1 <i>Trichodesmium erythraeum</i> IMS101	70	Transposase
<i>cyrH</i>	cytosine deaminase /Aminohydrolase/ Dihydroorotase	1431	BAF59909.1 <i>Pelotomaculum thermopropionicum</i> SI	50	Uracil ring formation
<i>cyrJ</i>	branched-chain amino acid aminotransferase	780	<i>Trichodesmium erythraeum</i> IMS101	53	sulfotransferase
<i>cyrA</i>	Amidinotransferase AoaA	1176	AAX81898.1 <i>Cylindrospermopsis raciborskii</i>	100	amidinotransferase
<i>cyrB</i>	NRPS/PKS AoaB	8754	AAM33468.1 <i>Aphanizomenon ovalisporum</i>	97	NRPS/PKS A-domain, pp, KS, AT, DH, Met, KR, ACP
<i>cyrE</i>	PKS	5667	ABA23591.1 <i>Anabaena variabilis</i> ATCC 29413	62	PKS KS-AT-DH-KR-ACP
<i>cyrC</i>	PKS AoaC	5005	AAM33470.1 <i>Aphanizomenon ovalisporum</i>	97	PKS KS-AT-KR-ACP
<i>cyrM</i>	Partial Transposase	318	ABG50981.1 <i>Trichodesmium erythraeum</i> IMS101	70	Transposase
<i>cyrN</i>	Adenylylsulfate kinase (PAPS)	600	CAM76460.1 <i>Magnetospirillum gryphiswaldense</i> MSR-1	75	Adenylylsulfate kinase (PAPS)
<i>cyrO</i>	hypothetical protein	1548	EAW46978.1 <i>Nodularia spumigena</i> CCY9414	74	Regulator

FIGURE 6

Cyanobacterial Strain	16s rRNA	<i>cyrJ</i>	Toxicity	Reference
<i>Cylindrospermopsis raciborskii</i> T3	+	-	SXT	Lagos et al. (1999)
<i>Anabaena circinalis</i> 344B	+	-	N.D.	AWQC
<i>Cylindrospermopsis raciborskii</i> Germ1	+	-	N.D.	Neilan et al. (2003)
<i>Anabaena circinalis</i> 310F	+	-	N.D.	AWQC
<i>Cylindrospermopsis raciborskii</i> 44D	+	-	N.D.	NA
<i>Anabaena circinalis</i> 118C	+	-	SXT	Fergusson et al. (2000)
<i>Anabaena circinalis</i> 323A	+	-	N.D.	AWQC
<i>Anabaena circinalis</i> 323H	+	-	N.D.	AWQC
<i>Cylindrospermopsis raciborskii</i> VOLL2	+	-	N.D.	Neilan et al. (2003)
<i>Cylindrospermopsis raciborskii</i> VOLL1	+	-	N.D.	Neilan et al. (2003)
<i>Cylindrospermopsis raciborskii</i> HUNG1	+	-	N.D.	NA
<i>Cylindrospermopsis raciborskii</i> 023B	+	+	CYLN	Wilson et al. (2000)
<i>Cylindrospermopsis raciborskii</i> 05E	+	+	CYLN	Schembri et al. (2001)
<i>Cylindrospermopsis raciborskii</i> 4799	+	+	CYLN	Neilan et al. (2003)
<i>Cylindrospermopsis raciborskii</i> 24C	+	+	CYLN	Schembri et al. (2001)
<i>Cylindrospermopsis raciborskii</i> AWT 205	+	+	CYLN	Hawkins et al. (1997)
<i>Aphanizomenon ovalisporum</i> AO/QH	+	+	CYLN	NA

FIGURE 7

FIGURE 8

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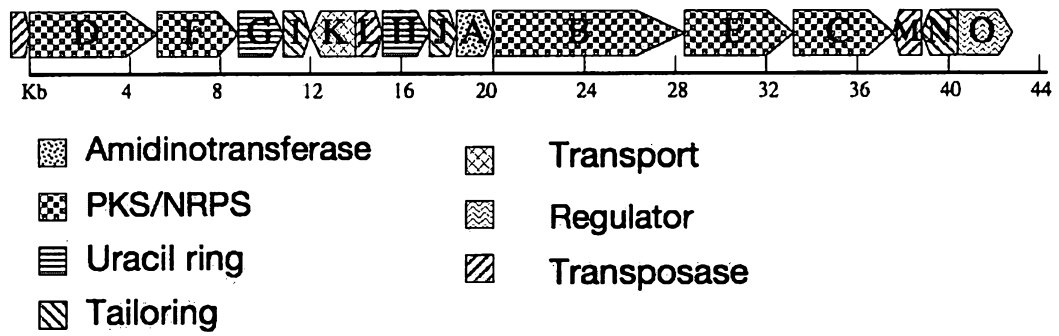


FIGURE 9

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<211> 1320

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 2

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<210> 3

<211> 439

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 3

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Ile Ser Pro Phe Leu Ser Met Ile Val Gly Ile Tyr Glu Asn Ile Lys
 20 25 30

Phe Arg Val Leu Phe Asp Leu Val Val Arg Ala Leu Met Val Val Asp
 35 40 45

Cys Phe Asn Ile Lys Lys His Arg Val Lys Ile Ser Arg Gln Leu Pro
 50 55 60

Leu Arg Leu Ser Ile Gly Arg Glu Asn Leu Val Ile Leu Lys Val Glu
 65 70 75 80

Ser Gly Asn Val Asn Ser Ala Ile Gln Ile Arg Asp Tyr Tyr Pro Thr
85 90 95

Glu Phe Pro Val Ser Thr Ser Asn Leu Ile Val Asn Leu Pro Pro Asn
100 105 110

His Thr Gln Glu Val Lys Tyr Thr Ile Arg Pro Asn Gln Arg Gly Glu
115 120 125

Phe Trp Trp Gly Asn Ile Gln Val Arg Gln Leu Gly Asn Trp Ser Leu
130 135 140

Gly Trp Asp Asn Trp Gln Ile Pro Gln Lys Thr Val Ala Lys Val Tyr
145 150 155 160

Pro Asp Leu Leu Gly Leu Arg Ser Leu Ala Ile Arg Leu Thr Leu Gln
165 170 175

Ser Ser Gly Ser Ile Thr Lys Leu Arg Gln Arg Gly Met Gly Thr Glu
180 185 190

Phe Ala Glu Leu Arg Asn Tyr Cys Met Gly Asp Asp Leu Arg Leu Ile
195 200 205

Asp Trp Lys Ala Thr Ala Arg Arg Ala Tyr Gly Asn Leu Ser Pro Leu
210 215 220

Val Arg Val Leu Glu Pro Gln Gln Glu Gln Thr Leu Leu Ile Leu Leu
225 230 235 240

Asp Arg Gly Arg Leu Met Thr Ala Asn Val Gln Gly Leu Lys Arg Tyr
245 250 255

Asp Trp Gly Leu Asn Thr Thr Leu Ser Leu Ala Leu Ala Gly Leu His
260 265 270

Arg Gly Asp Arg Val Gly Val Gly Val Phe Asp Ser Gln Leu His Thr

275 280 285
 Trp Ile Pro Pro Glu Arg Gly Gln Asn His Leu Asn Arg Leu Ile Asp
 290 295 300
 Arg Leu Thr Pro Ile Glu Pro Val Leu Val Glu Ser Asp Tyr Leu Asn
 305 310 315 320
 Ala Ile Thr Tyr Val Val Lys Gln Gln Thr Arg Arg Ser Leu Val Val
 325 330 335
 Leu Ile Thr Asp Leu Val Asp Val Thr Ala Ser His Glu Leu Leu Val
 340 345 350
 Ala Leu Cys Lys Leu Val Pro Arg Tyr Leu Pro Phe Cys Val Thr Leu
 355 360 365
 Arg Asp Pro Gly Ile Asp Lys Ile Ala His Asn Phe Ser Gln Asp Leu
 370 375 380
 Thr Gln Ala Tyr Asn Arg Ala Val Ser Leu Asp Leu Ile Ser Gln Arg
 385 390 395 400
 Glu Ile Ala Phe Ala Gln Leu Lys Gln Gln Gly Val Leu Val Leu Asp
 405 410 415
 Ala Pro Ala Asn Gln Ile Ser Glu Gln Leu Val Glu Arg Tyr Leu Gln
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 Ile Lys Ala Lys Asn Gln Ile
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<210> 5

<211> 252

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 5

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Leu Thr Gly Leu Ala Phe Trp Leu Trp Glu Ile Arg Ser Pro Phe His
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Gln Ile Glu Tyr Lys Ala Lys Phe Phe Lys Glu Leu Gly Trp Ala Gly
 35 40 45

Ile Ser Phe Val Phe Arg Asn Val Tyr Ala Tyr Val Ser Val Ala Ile
 50 55 60

Ile Lys Leu Leu Ser Ser Leu Phe Met Gly Glu Ser Ala Asn Phe Ala

65

70

75

80

Gly Val Met Tyr Val Pro Leu Trp Leu Arg Ile Ile Thr Ala Tyr Ile
 85 90 95

Leu Gln Asp Leu Thr Asp Tyr Leu Leu His Arg Thr Met His Ser Asn
 100 105 110

Gln Phe Leu Trp Leu Thr His Lys Trp His His Ser Thr Lys Gln Ser
 115 120 125

Trp Trp Leu Ser Gly Asn Lys Asp Ser Phe Thr Gly Gly Leu Leu Tyr
 130 135 140

Thr Val Thr Ala Leu Trp Phe Pro Leu Leu Asp Ile Pro Ser Glu Val
 145 150 155 160

Met Ser Val Val Ala Val His Gln Val Ile His Asn Asn Trp Ile His
 165 170 175

Leu Asn Val Lys Trp Asn Ser Trp Leu Gly Ile Ile Glu Trp Ile Tyr
 180 185 190

Val Thr Pro Arg Ile His Thr Leu His His Leu Asp Thr Gly Gly Arg
 195 200 205

Asn Leu Ser Ser Met Phe Thr Phe Ile Asp Arg Leu Phe Gly Thr Tyr
 210 215 220

Val Phe Pro Glu Asn Phe Asp Ile Glu Lys Ser Lys Asn Arg Leu Asp
 225 230 235 240

Asp Gln Ser Val Thr Val Lys Thr Ile Leu Gly Phe
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<210> 6

<211> 396

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 6

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aatttggtct ccacggggaa taaattttgg gtaaacttta tagccatccg taatccaaaa  240
ataggatttc caatgctcta tctttttcca taatttggca aatgttttgg cacttctatc  300
tcccactaca tattgaataa ttcccgaacg ttgttatct acaactgtcc agaccatat  360
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<210> 7

<211> 131

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 7

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1         5         10        15
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Thr Val Val Asp Asn Lys Arg Ser Gly Ile Ile Gln Tyr Val Val Gly
        20        25        30
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Asp Arg Ser Ala Lys Thr Phe Ala Lys Leu Trp Lys Lys Ile Glu His
        35        40        45
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Trp Lys Ser Tyr Phe Trp Ile Thr Asp Gly Tyr Lys Val Tyr Pro Lys
        50        55        60
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Phe Ile Pro Asp Gly Asp Gln Ile Ile Ser Lys Thr Ser Met Thr Arg
        65        70        75        80
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Val Glu Gly Glu Asn Ser Arg Leu Arg His Tyr Leu Ala Arg Leu His
        85        90        95
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Arg Lys Thr Phe Cys Tyr Ser Lys Ser Lys Glu Met Leu Cys Phe Ser
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100 105 110

Ile Lys Leu Leu Ile Tyr Tyr Leu Arg Glu Lys Ser Phe Ser Ala Ile
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Ser Trp Gly
130

<210> 8
<211> 360
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<213> *Cylindrospermopsis raciborskii* T3

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tagctttttg acccaacgaa tgactgtatt gtgatttact ttagtcattc ttcaattgc 180
cctaaatcca ttccattta catacatggg taaacatgct tcctttactt cttgggaata 240
acctctagga gaataagatt caataaattg acgaccacaa ttcttgcat gataatttg 300
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<210> 9
<211> 119
<212> PRT
<213> *Cylindrospermopsis raciborskii* T3

<400> 9

Met Asn Cys Ser Asn Cys Asp Ser Asn Asn Ile Arg Lys Asn Gly Gln
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Arg Arg Gly Lys Gln Asn Tyr Gln Cys Lys Asn Cys Gly Arg Gln Phe
20 25 30

Ile Glu Ser Tyr Ser Pro Arg Gly Tyr Ser Gln Glu Val Lys Glu Ala
35 40 45

Cys Leu Thr Met Tyr Val Asn Gly Asn Gly Phe Arg Ala Ile Glu Arg

50 55 60
 Met Thr Lys Val Asn His Asn Thr Val Ile Arg Trp Val Lys Lys Leu
 65 70 75 80
 Gly Arg Gln Leu Ser Asp Ser Asn Asn Asn Ser Gln Thr Pro Glu Val
 85 90 95
 Cys Gln Leu Asp Glu Leu Glu Thr Phe Ile Gly Lys Lys Lys Gln Asp
 100 105 110
 Met Gly Leu Asp Ser Cys Arg
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<210> 10
 <211> 354
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 <213> *Cylindrospermopsis raciborskii* T3
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 acctttatca tcaacaaaaa ccatctgctc gcaccaatct acaaatccgg aattagtcac 180
 ctcatagact aaaatgatgg gaggaagtgc tgccaatccc atttttcaa tgacttccat 240
 acaaaccagc ttaaataactt gttcgtttgt caattcatta gacataaaga atttccctt 300
 aatcaattct gtttctaact ctaccacaga gtaataactc ttggtctgga acat 354

<210> 11
 <211> 117
 <212> PRT
 <213> *Cylindrospermopsis raciborskii* T3
 <400> 11

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 1 5 10 15

Leu Ile Lys Gly Lys Phe Phe Met Ser Asn Glu Leu Thr Asn Glu Gln

20 25 30

Val Phe Lys Leu Val Cys Met Glu Val Ile Glu Lys Met Gly Phe Ala
35 40 45

His Phe Pro Pro Ile Ile Leu Val Tyr Glu Met Thr Asn Ser Gly Phe
50 55 60

Val Asp Trp Cys Glu Gln Met Val Phe Val Asp Asp Lys Gly Lys Leu
65 70 75 80

Asp Glu Gly Glu Lys Phe Leu Leu Asp Trp Met Arg Arg Asn Val Gly
85 90 95

Asn Phe Asp Leu Ile Arg Glu Leu Met Pro Val Ala Glu Arg Leu Glu
100 105 110

Met Lys Met Arg Ser
115

<210> 12

<211> 957

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

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aacagcatca caaatgtcga atagtgattg agaatatcta ttcatttca tgaaagtcag 180

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<210> 13

<211> 318

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 13

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 20 25 30

Ser Ser Val Val Ala Ser Trp Ile Asp Arg Leu Gly Thr Asn Ile Ser
 35 40 45

Thr Leu Met Ile Gln Leu Leu Pro Val Ala Ala Thr Tyr Ala Arg Val
 50 55 60

Pro Ile Ser Gln Phe Tyr Val Gly Ala Ile Ala Leu Gly Lys Pro Gln
 65 70 75 80

Ser Lys Asn Gln Leu Gly Ser Gly Thr Leu Tyr Phe Gly Ala Asp Met
 85 90 95

Glu Phe Val Gly Gln Ala Leu Ser Phe Ser Val His Ala Glu Gln Ser
 100 105 110

Ala Thr Ile Asn Ala Trp Leu His Gly Glu Thr Gly Leu Gln Ala Leu
 115 120 125

Ala Ile His Glu Ala Pro Cys Gly Tyr Cys Arg Gln Phe Leu Tyr Glu
 130 135 140

Met Ala Thr Val Asn Gln Asn Phe Val Leu Leu Val Lys Ser Asn Glu
 145 150 155 160

Ser Gln Pro Glu Gln Thr Tyr Thr Ser Asn Lys Leu Pro His Phe Leu
 165 170 175

Pro Glu Pro Phe Gly Pro Ala Asp Leu Gly Leu Thr Gly Gly Leu Met
 180 185 190

Gln Thr Val Phe His Asp Leu Glu Thr Tyr Ser Thr Asp Asp Val Val
 195 200 205

Leu Ala Ala Leu Ser Ala Ala Asn Gln Ser Tyr Ala Pro Tyr Thr Lys
 210 215 220

Asn Phe Ala Gly Val Ala Leu Lys Asp Ser His Gly Asn Ile Phe Thr
 225 230 235 240

Gly Arg Tyr Ala Glu Asn Ala Ala Phe Asn Ser Ser Met Ser Pro Met
 245 250 255

Glu Ser Ala Leu Thr Phe Met Asn Met Asn Arg Tyr Ser Gln Ser Leu
 260 265 270

Phe Asp Ile Cys Asp Ala Val Leu Val Glu Val Glu Thr Gly Ile Ser
 275 280 285

Gln Arg Pro Val Thr Glu Ala Phe Leu Ser Ser Ile Ala Pro Lys Val
 290 295 300

Lys Leu Arg Tyr Ala Pro Ala Thr Pro Ser Ser Asn Lys Leu

305 310 315

<210> 14

<211> 3738

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

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gagtgc tttt cccctagcag acttaaatg aatggttcc cagactcgtt tcaggagagt 2880
tccatcgcca caccatgt cagtaatgta ttgggtgtt tcttcta atg gcaactgatt 2940
gaatactgag aggatacttt cttctaaatc ggcaaaatat ttctggtgtt gaaatccact 3000
cccgatcacg ttaaggggtgc gatcaatgtg cctttcgtga ccggaagcat ctctttggaa 3060
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cgccactatg gctgtattca aggctcgctc tccataaat cgaccaagtt cggttatggt 3180
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attaatcttt tgtaacat 3738

<210> 15
<211> 1245
<212> PRT
<213> *Cylindrospermopsis raciborskii* T3

<400> 15

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Val Ile Leu Ala Cys Arg Glu Lys Gly Val Phe Glu Leu Leu Ala Asp
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Glu Ser Pro Leu Ser Leu Asn Gln Met Val Glu His Leu Gly Ala Asn
 35 40 45

Ser Gly His Phe Gln Val Ala Leu Arg Met Leu Glu Ser Leu His Trp
 50 55 60

Leu Ser Arg Asn Lys Glu Leu Lys Tyr Ser Leu Thr Ala Glu Ala Ala
 65 70 75 80

Ile His Asn Lys Ile Ser Glu Asp Ile Leu Gln Leu Tyr Asn Leu Pro
 85 90 95

Ile Gln Ser Tyr Leu Glu Gly Lys Gln Gly Asn Leu Leu Gly Arg Trp
 100 105 110

Ile Glu Arg Ser Cys Gln Leu Trp Asn Leu Asp Asn Pro Leu Met Ala
 115 120 125

Asp Phe Leu Asp Gly Leu Leu Val Ile Pro Leu Leu Leu Ala Leu His
 130 135 140

Lys His Asn Leu Leu Ala Asp Ser Glu Asp Lys Pro Leu Leu Ser Ser
 145 150 155 160

Leu Ser Ser Thr Val Gln Glu Glu Leu Gly Lys Leu Phe Leu His Leu
 165 170 175

Gly Trp Ala Asp Leu Thr Ala Gly Arg Leu Thr Ile Thr Glu Leu Gly
 180 185 190

Arg Phe Met Gly Glu Arg Ala Leu Asn Thr Ala Ile Val Ala Ser Tyr
 195 200 205

Thr Pro Met Leu Ser Arg Ile His Asp Val Leu Phe Gly Asn Cys Leu
 210 215 220

Ser Val Phe Gln Arg Asp Ala Ser Gly His Glu Arg His Ile Asp Arg

225 230 235 240

Thr Leu Asn Val Ile Gly Ser Gly Phe Gln His Gln Lys Tyr Phe Ala
 245 250 255

Asp Leu Glu Glu Ser Ile Leu Ser Val Phe Asn Gln Leu Pro Leu Glu
 260 265 270

Glu Gln Pro Lys Tyr Ile Thr Asp Met Gly Cys Gly Asp Gly Thr Leu
 275 280 285

Leu Lys Arg Val Trp Glu Thr Ile Gln Phe Lys Ser Ala Arg Gly Lys
 290 295 300

Ala Leu Glu Gln Tyr Pro Leu Arg Leu Ile Gly Val Asp Tyr Asn Glu
 305 310 315 320

Ala Ser Leu Lys Ala Thr Thr Arg Thr Leu Ala Ser Leu Pro His Leu
 325 330 335

Val Leu Gln Gly Asp Ile Gly Asn Pro Glu Gln Met Val Arg Ser Leu
 340 345 350

Glu Ala His Gly Ile His Asp Pro Glu Asn Ile Leu His Ile Arg Ser
 355 360 365

Phe Leu Asp His Asp Arg Leu Phe Ile Pro Pro Gln Lys Arg Asn Glu
 370 375 380

Leu Lys Glu Arg Ala His Leu Pro Tyr Gln Ser Val Cys Val Asp Asp
 385 390 395 400

Gln Gly Glu Leu Ile Pro Pro His Val Met Val Gln Ser Leu Val Glu
 405 410 415

His Leu Glu Arg Trp Ser Gln Val Val Asn Lys His Gly Leu Met Ile
 420 425 430

Leu Glu Val His Cys Leu Glu Pro Arg Val Val Tyr Gln Phe Leu Asp
 435 440 445

Lys Ser Glu Asn Leu His Phe Asp Ala Phe Gln Gly Phe Ser Gln Gln
 450 455 460

Tyr Leu Val Glu Ala Glu Val Phe Leu Met Ser Ala Ala Gln Val Gly
 465 470 475 480

Leu Phe Pro Lys Leu Glu Leu Ser Lys Arg Tyr Pro Lys Thr Phe Pro
 485 490 495

Phe Thr Arg Ile Thr Leu Asn Tyr Phe Glu Lys Arg Pro Tyr Lys Ile
 500 505 510

Ser His Ala Tyr Leu Ser Asp Leu Pro Ala Leu Val Asp Leu Glu Val
 515 520 525

Lys Cys Trp Pro Glu Asn Leu Arg Ala Ser Thr His Glu Ile Arg Arg
 530 535 540

Arg Leu Glu Leu Asn Pro Gln Gly Asn Leu Val Leu Ile Ile Glu Asp
 545 550 555 560

Gln Ile Ile Gly Ala Ile Tyr Ser Gln Thr Ile Thr Ser Thr Glu Ala
 565 570 575

Leu Glu Asn Val Lys Tyr Ala Gln Val Pro Thr Leu His Thr Pro Gln
 580 585 590

Gly Ser Val Ile Gln Leu Leu Ala Leu Asn Ile Leu Pro Glu Phe Gln
 595 600 605

Ala Arg Gly Leu Gly Asn Glu Leu Arg Asp Phe Met Leu Tyr Tyr Cys
 610 615 620

Thr Leu Lys Gly Gly Ile Glu Ser Val Val Gly Val Thr Arg Cys Arg

625 630 635 640

Asn Tyr Val Asn Tyr Ser Gln Met Pro Met Met Glu Tyr Leu Lys Leu
 645 650 655

His Asn Glu Gln Arg Gln Leu Leu Asp Pro Ile Val Gly Phe His Val
 660 665 670

Ser Gly Gly Ala Glu Ile Arg Gly Ile Ile Ala Asn Tyr Arg Pro Glu
 675 680 685

Asp Thr Asp Asn Leu Gly Met Gly Ile Leu Ile Glu Tyr Asn Leu Arg
 690 695 700

Asp Ser Ala Leu His Ser Pro Gly Asp Arg Lys Gly Pro Tyr Ile Asn
 705 710 715 720

Ser Ala Ile Gly Ser Leu Val Pro Lys Ala Thr Ser Ala Thr Lys Glu
 725 730 735

Asn Lys Thr Val Ala Asp Leu Val Lys Glu Cys Ile Leu Lys Val Met
 740 745 750

Gly Ser Gln Arg Gln Ala Ala Tyr Ala Pro Gln Gln Lys Leu Leu Asp
 755 760 765

Met Gly Leu Asp Ser Leu Asp Leu Leu Glu Leu Gln Thr Leu Leu Glu
 770 775 780

Glu Arg Leu Gly Ile Asn Leu Ser Gly Thr Phe Phe Leu Gln Lys Asn
 785 790 795 800

Thr Pro Thr Ala Ile Ile Thr Tyr Phe Gln Asn Gln Val Val Gln Glu
 805 810 815

Lys Gln Ser Asp Leu Ala Pro Pro Val Asp Ser Ala Asn Glu Ile Asn
 820 825 830

Thr Leu Glu Asn Val Val Asn Gln Gln Lys Ile Pro Gln Val Thr Arg
835 840 845

Val Val Thr Glu Gln Gln Gly Arg Lys Val Leu Ile Asp Gly His Trp
850 855 860

Val Ile Asp Phe Ala Ser Cys Asn Tyr Leu Gly Leu Asp Leu His Pro
865 870 875 880

Lys Val Lys Glu Ala Ile Pro Pro Ala Leu Asp Lys Trp Gly Thr His
885 890 895

Pro Ser Trp Thr Arg Leu Val Ala Ser Pro Ala Ile Tyr Glu Glu Leu
900 905 910

Glu Glu Glu Leu Ser Lys Leu Leu Gly Val Pro Asp Val Leu Val Phe
915 920 925

Pro Ala Val Thr Leu Leu Gln Ile Gly Ile Leu Pro Leu Leu Thr Gly
930 935 940

Asn Asn Gly Val Ile Phe Gly Asp Ile Ala Ala His Arg Cys Ile Tyr
945 950 955 960

Glu Ala Cys Cys Leu Ala Gln His Lys Gly Ala Gln Phe Ile Gln Tyr
965 970 975

Arg His Asn Asp Leu Asn Asp Leu Ala Glu Lys Leu Ala Lys Tyr Pro
980 985 990

Pro Glu Gln Val Lys Ile Ile Val Ile Asp Gly Val Tyr Ser Met Ser
995 1000 1005

Ala Asp Phe Pro Asp Leu Pro Ala Tyr Val His Leu Ala Lys Glu
1010 1015 1020

Tyr Asn Ala Leu Ile Tyr Met Asp Asp Ala His Gly Phe Gly Ile

1025	1030	1035
Leu Gly	Glu Asn Pro Ser Ser	Asp Met Pro Tyr Gly Tyr Lys Gly
1040	1045	1050
Asn Gly	Met Val Asn Tyr Phe	Asp Leu Arg Phe Ala Glu Asp Asn
1055	1060	1065
Ile Ile	Tyr Val Ala Gly Leu	Ser Lys Ala Tyr Ser Ser Tyr Ala
1070	1075	1080
Ala Phe	Leu Thr Cys Gly Asp	Arg Arg Ile Lys Thr Asn Phe Arg
1085	1090	1095
Asn Ala	Trp Thr Ala Ile Phe	Ser Gly Pro Ser Pro Val Ala Ser
1100	1105	1110
Leu Ala	Ser Ala Leu Ala Gly	Leu Gln Val Asn Arg Gln Glu Gly
1115	1120	1125
Glu Gln	Leu Arg Lys Gln Ile	Tyr His Leu Thr His Lys Leu Val
1130	1135	1140
Thr Gln	Ala Arg Ala Ile Gly	Phe Glu Val Asp Asn Tyr Gly Tyr
1145	1150	1155
Val Pro	Ile Val Gly Val Leu	Val Gly Asp Ala Gln His Met Ile
1160	1165	1170
Asp Val	Cys Gln Leu Leu Trp	Glu Tyr Gly Ile Leu Ile Thr Pro
1175	1180	1185
Ala Ile	Phe Pro Ile Val Pro	Leu Asn Lys Ser Ala Leu Arg Phe
1190	1195	1200
Ser Ile	Thr Ala Ala Asn Thr	Glu Glu Glu Ile Asp Gln Ala Ile
1205	1210	1215

Lys Ser Leu Lys Ala Val Trp Asp Leu Leu Gln Lys Arg Lys Ala
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Leu Pro Cys Lys Gln Glu Glu Asn Ile Leu Lys His
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<210> 16

<211> 387

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 16

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attaccaatg ttcaaatac ttcttctaac ggtgataatt atgccgtag tatcagcgg 180

gggtttggtc ctgcgcaga tagagtgatt atcctaccaa cttcaggagt gataaatcga 240

gacattcata tgcgtggcta tgaagccgca ttaactgcac tatccaatgg ctttttagta 300

gatatttacg actatactgg ctcttcttgc agcaatgggtg gccaaactaac tattaccaac 360

caattaggta agctaatacag caattag 387

<210> 17

<211> 128

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 17

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Phe Ala Phe Gly Ile Phe Leu Thr Thr Gly Val Gly Ile Ala Lys Ala
 20 25 30

Asp Tyr Leu Val Lys Gly Gly Lys Ile Thr Asn Val Gln Asn Thr Ser
 35 40 45

Ser Asn Gly Asp Asn Tyr Ala Val Ser Ile Ser Gly Gly Phe Gly Pro

50 55 60
 Cys Ala Asp Arg Val Ile Ile Leu Pro Thr Ser Gly Val Ile Asn Arg
 65 70 75 80
 Asp Ile His Met Arg Gly Tyr Glu Ala Ala Leu Thr Ala Leu Ser Asn
 85 90 95
 Gly Phe Leu Val Asp Ile Tyr Asp Tyr Thr Gly Ser Ser Cys Ser Asn
 100 105 110
 Gly Gly Gln Leu Thr Ile Thr Asn Gln Leu Gly Lys Leu Ile Ser Asn
 115 120 125
 <210> 18
 <211> 1416
 <212> DNA
 <213> *Cylindrospermopsis raciborskii* T3
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 gcagtcatga tgggtttact tggtagcгаа gttttggccg ccggtgcttt gggcgcgctc 180
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 gccgaagctt ttggggcaaa taaaatagat gaagttagtc gtattgcttc cgggcaaata 300
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 attttatggg gctttccgc tgcgcttagt atttgacat taagaggcat tgcctctgct 480
 ctcaacgttc cccgattgat aactattact atgctcactc agctgatatt gaataccgcc 540
 gccgattatg tgtaaatatt cggtaaattt ggtcttcctc aacttggttt ggctggaata 600
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 ttctccctga aagttagaga ttataaactt ttccgctact tgcacagtt tgataaacag 720
 atctttgtca aaattttca aactggatgg cccatggggg ttcaatgggg ggcggaacg 780

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gttattttaa ctgttcgttt ttatcgagtg attgacaatc ttctgcca cagtgatgat 1380
atgttacaga atgcgtctgt tactactcta ggctga 1416

<210> 19

<211> 471

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 19

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Arg Ala Ser Leu Lys Leu Gly Ile Pro Leu Val Ile Ser Gln Met Cys
20 25 30

Glu Thr Gly Ile Tyr Thr Ala Asn Ala Val Met Met Gly Leu Leu Gly
35 40 45

Thr Gln Val Leu Ala Ala Gly Ala Leu Gly Ala Leu Ala Phe Leu Thr
50 55 60

Leu Leu Phe Ala Cys His Gly Ile Leu Ser Val Gly Gly Ser Leu Ala
65 70 75 80

Ala Glu Ala Phe Gly Ala Asn Lys Ile Asp Glu Val Ser Arg Ile Ala
 85 90 95

Ser Gly Gln Ile Trp Leu Ala Val Thr Leu Ser Leu Pro Ala Met Leu
 100 105 110

Leu Leu Trp His Gly Asp Thr Ile Leu Leu Leu Phe Gly Gln Glu Glu
 115 120 125

Ser Asn Val Leu Leu Thr Lys Thr Tyr Leu His Ser Ile Leu Trp Gly
 130 135 140

Phe Pro Ala Ala Leu Ser Ile Leu Thr Leu Arg Gly Ile Ala Ser Ala
 145 150 155 160

Leu Asn Val Pro Arg Leu Ile Thr Ile Thr Met Leu Thr Gln Leu Ile
 165 170 175

Leu Asn Thr Ala Ala Asp Tyr Val Leu Ile Phe Gly Lys Phe Gly Leu
 180 185 190

Pro Gln Leu Gly Leu Ala Gly Ile Gly Trp Ala Thr Ala Leu Gly Phe
 195 200 205

Trp Val Ser Phe Thr Leu Gly Leu Ile Leu Leu Ile Phe Ser Leu Lys
 210 215 220

Val Arg Asp Tyr Lys Leu Phe Arg Tyr Leu His Gln Phe Asp Lys Gln
 225 230 235 240

Ile Phe Val Lys Ile Phe Gln Thr Gly Trp Pro Met Gly Phe Gln Trp
 245 250 255

Gly Ala Glu Thr Ala Leu Phe Asn Val Thr Ala Trp Val Ala Gly Tyr
 260 265 270

Leu Gly Thr Val Thr Leu Ala Ala His Asp Ile Gly Phe Gln Thr Ala

275 280 285
 Glu Leu Ala Met Val Ile Pro Leu Gly Val Gly Asn Val Ala Met Thr
 290 295 300
 Arg Val Gly Gln Ser Ile Gly Glu Lys Asn Pro Leu Gly Ala Arg Arg
 305 310 315 320
 Val Ala Ser Ile Gly Ile Thr Ile Val Gly Ile Tyr Ala Ser Ile Val
 325 330 335
 Ala Leu Val Phe Trp Leu Phe Pro Tyr Gln Ile Ala Gly Ile Tyr Leu
 340 345 350
 Asn Ile Asn Asn Pro Glu Asn Ile Glu Ala Ile Lys Lys Ala Thr Thr
 355 360 365
 Phe Ile Pro Leu Ala Gly Leu Phe Gln Met Phe Tyr Ser Ile Gln Ile
 370 375 380
 Ile Ile Val Gly Ala Leu Val Gly Leu Arg Asp Thr Phe Val Pro Val
 385 390 395 400
 Ser Met Asn Leu Ile Val Trp Gly Leu Gly Leu Ala Gly Ser Tyr Phe
 405 410 415
 Met Ala Ile Ile Leu Gly Trp Gly Gly Ile Gly Ile Trp Leu Ala Met
 420 425 430
 Val Leu Ser Pro Leu Leu Ser Ala Val Ile Leu Thr Val Arg Phe Tyr
 435 440 445
 Arg Val Ile Asp Asn Leu Leu Ala Asn Ser Asp Asp Met Leu Gln Asn
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 Ala Ser Val Thr Thr Leu Gly
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<210> 20

<211> 1134

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 20

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ggtgcaatgt taccggccct agaaccaatc acaaatgga cattcccttt tgaagaattg  180
gaatctgccc aaaagatact ctctgagagg ggaggagttc cttatccacc agagatgatt  240
acattagcac acaaagaact aaatgaattt attcacattc ttgaagcaga aggggtcaaa  300
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tggggattta agcctattgt ttgccattt gaaagctact atccatttt aggatcattt 1080
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<210> 21

<211> 377

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 21

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Lys Gln Pro Cys Pro Val Asn Ser Tyr Asn Glu Trp Asp Thr Leu Glu
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Glu Val Ile Val Gly Ser Val Glu Gly Ala Met Leu Pro Ala Leu Glu
 35 40 45

Pro Ile Asn Lys Trp Thr Phe Pro Phe Glu Glu Leu Glu Ser Ala Gln
 50 55 60

Lys Ile Leu Ser Glu Arg Gly Gly Val Pro Tyr Pro Pro Glu Met Ile
 65 70 75 80

Thr Leu Ala His Lys Glu Leu Asn Glu Phe Ile His Ile Leu Glu Ala
 85 90 95

Glu Gly Val Lys Val Arg Arg Val Lys Pro Val Asp Phe Ser Val Pro
 100 105 110

Phe Ser Thr Pro Ala Trp Gln Val Gly Ser Gly Phe Cys Ala Ala Asn
 115 120 125

Pro Arg Asp Val Phe Leu Val Ile Gly Asn Glu Ile Ile Glu Ala Pro
 130 135 140

Met Ala Asp Arg Asn Arg Tyr Phe Glu Thr Trp Ala Tyr Arg Glu Met
 145 150 155 160

Leu Lys Glu Tyr Phe Gln Ala Gly Ala Lys Trp Thr Ala Ala Pro Lys
 165 170 175

Pro Gln Leu Phe Asp Ala Gln Tyr Asp Phe Asn Phe Gln Phe Pro Gln
 180 185 190

Leu Gly Glu Pro Pro Arg Phe Val Val Thr Glu Phe Glu Pro Thr Phe
 195 200 205

Asp Ala Ala Asp Phe Val Arg Cys Gly Arg Asp Ile Phe Gly Gln Lys
 210 215 220

Ser His Val Thr Asn Gly Leu Gly Ile Glu Trp Leu Gln Arg His Leu
 225 230 235 240

Glu Asp Glu Tyr Arg Ile His Ile Ile Glu Ser His Cys Pro Glu Ala
 245 250 255

Leu His Ile Asp Thr Thr Leu Met Pro Leu Ala Pro Gly Lys Ile Leu
 260 265 270

Val Asn Pro Glu Phe Val Asp Val Asn Lys Leu Pro Lys Ile Leu Lys
 275 280 285

Ser Trp Asp Ile Leu Val Ala Pro Tyr Pro Asn His Ile Pro Gln Asn
 290 295 300

Gln Leu Arg Leu Val Ser Glu Trp Ala Gly Leu Asn Val Leu Met Leu
 305 310 315 320

Asp Glu Glu Arg Val Ile Val Glu Lys Asn Gln Glu Gln Met Ile Lys
 325 330 335

Ala Leu Lys Asp Trp Gly Phe Lys Pro Ile Val Cys His Phe Glu Ser
 340 345 350

Tyr Tyr Pro Phe Leu Gly Ser Phe His Cys Ala Thr Leu Asp Val Arg
 355 360 365

Arg Arg Gly Thr Leu Gln Ser Tyr Phe
 370 375

<210> 22

<211> 1005

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 22

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 agtcgtgaac agaattcccc catacagata tggcaagact actgccctca ccgaggtgtg 180
 gctctgtcta tgggagaaat tgtaataat actttgggtt gtccgtatca cggatggaga 240
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 agtgcccaag ccaagatcta tcattgccag gagcgatagc gattagtatg ggtgtgctta 360
 ggtgatcctg tcaatgatat accttcatta cccgaatggg acgatccgaa ttatcataat 420
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 gatgtatctc attttccttt tgtccacgac ggtggggttag gtgatcgcaa ccacgcacaa 540
 attgaagaat ttgaggtaaa agtagacaaa gatggcatta gcataggtaa ccttaaactc 600
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 aacatgcaat ggttgtcaca ggaaatacat gtaccgtcag atcgatgcac agttgcctat 960
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<210> 23

<211> 334

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 23

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Lys Val Glu Asp Cys Lys Pro Gly Ser Ile Thr Thr Ala Leu Leu Leu
 20 25 30

Gly Val Lys Leu Val Leu Trp Arg Ser Arg Glu Gln Asn Ser Pro Ile
 35 40 45

Gln Ile Trp Gln Asp Tyr Cys Pro His Arg Gly Val Ala Leu Ser Met
 50 55 60

Gly Glu Ile Val Asn Asn Thr Leu Val Cys Pro Tyr His Gly Trp Arg
 65 70 75 80

Tyr Asn Gln Ala Gly Lys Cys Val His Ile Pro Ala His Pro Asp Met
 85 90 95

Thr Pro Pro Ala Ser Ala Gln Ala Lys Ile Tyr His Cys Gln Glu Arg
 100 105 110

Tyr Gly Leu Val Trp Val Cys Leu Gly Asp Pro Val Asn Asp Ile Pro
 115 120 125

Ser Leu Pro Glu Trp Asp Asp Pro Asn Tyr His Asn Thr Cys Thr Lys
 130 135 140

Ser Tyr Phe Ile Gln Ala Ser Ala Phe Arg Val Met Asp Asn Phe Ile
 145 150 155 160

Asp Val Ser His Phe Pro Phe Val His Asp Gly Gly Leu Gly Asp Arg
 165 170 175

Asn His Ala Gln Ile Glu Glu Phe Glu Val Lys Val Asp Lys Asp Gly
 180 185 190

Ile Ser Ile Gly Asn Leu Lys Leu Gln Met Pro Arg Phe Asn Ser Ser
 195 200 205

Asn Glu Asp Asp Ser Trp Thr Leu Tyr Gln Arg Ile Ser His Pro Leu

210 215 220
 Cys Gln Tyr Tyr Ile Thr Glu Ser Ser Glu Ile Arg Thr Ala Asp Leu
 225 230 235 240
 Met Leu Val Thr Pro Ile Asp Glu Asp Asn Ser Leu Val Arg Met Leu
 245 250 255
 Val Thr Trp Asn Arg Ser Glu Ile Leu Glu Ser Thr Val Leu Glu Glu
 260 265 270
 Phe Asp Glu Thr Ile Glu Gln Asp Ile Pro Ile Ile His Ser Gln Gln
 275 280 285
 Pro Ala Arg Leu Pro Leu Leu Pro Ser Lys Gln Ile Asn Met Gln Trp
 290 295 300
 Leu Ser Gln Glu Ile His Val Pro Ser Asp Arg Cys Thr Val Ala Tyr
 305 310 315 320
 Arg Arg Trp Leu Lys Glu Leu Gly Val Thr Tyr Gly Val Cys
 325 330

<210> 24

<211> 1839

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 24

atgcagatct taggaatttc agcttactac cacgatatgt ctgccgcgat gggtatcgat 60

ggcgaaattg ttgctgcagc tcaggaagaa cgtttctcaa gacgaaagca cgatgctggg 120

ttccgactg gagcgattac ttactgtcta aaacaagtag gaaccaagtt acaatatatc 180

gatcaaattg tttttacga caagccatta gtcaaattg agcggttgct agaaacatat 240

ttagcatatg ccccaaaggg atttggtctg ttattactg ctatgcccggt ttggctcaaa 300

gaaaagcttt acctaaaaac acttttaaaa aaagaattgg cgcttttggg ggagtgcaaa 360

gcttctcaat tgcctcctct actgtttacc tcacatcacc aagcccatgc ggccgctgct 420

tttttccca gtccttttca gcggtgctgcc gttctgtgct tagatgggtg aggagagtgg 480
 gcaactactt ctgtctgggt gggagaagga aataaactca caccacaatg ggaaattgat 540
 ttccccatt cctcgggtt gctttactca gcgtttacct actacactgg gttcaaagtt 600
 aactcaggtg agtacaaact catgggttta gcacctacg gggaacccaa atatgtggac 660
 caaatttca agcatttgtt ggaatcctaaa gaagatggta ctttaggtt gaatatggac 720
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 ggaccaccac gccaggcgga aggaaaaatc tccaaagag acatggatct ggcaagttcg 840
 atccaaaagg tgactgaaga agtcatactg cgtctggcta gaactatcaa aaaagaactg 900
 ggtgtagagt atctatgttt agcaggtggt gtcggtctca attgcgtggc taacggacga 960
 attctccgag aaagtgtttt caaagatatt tggattcaac ccgcagcagg agatgccggt 1020
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<210> 25
 <211> 612
 <212> PRT
 <213> *Cylindrospermopsis raciborskii* T3

<400> 25

Met Gln Ile Leu Gly Ile Ser Ala Tyr Tyr His Asp Ser Ala Ala Ala
 1 5 10 15

Met Val Ile Asp Gly Glu Ile Val Ala Ala Ala Gln Glu Glu Arg Phe
 20 25 30

Ser Arg Arg Lys His Asp Ala Gly Phe Pro Thr Gly Ala Ile Thr Tyr
 35 40 45

Cys Leu Lys Gln Val Gly Thr Lys Leu Gln Tyr Ile Asp Gln Ile Val
 50 55 60

Phe Tyr Asp Lys Pro Leu Val Lys Phe Glu Arg Leu Leu Glu Thr Tyr
 65 70 75 80

Leu Ala Tyr Ala Pro Lys Gly Phe Gly Ser Phe Ile Thr Ala Met Pro
 85 90 95

Val Trp Leu Lys Glu Lys Leu Tyr Leu Lys Thr Leu Leu Lys Lys Glu
 100 105 110

Leu Ala Leu Leu Gly Glu Cys Lys Ala Ser Gln Leu Pro Pro Leu Leu
 115 120 125

Phe Thr Ser His His Gln Ala His Ala Ala Ala Ala Phe Phe Pro Ser
 130 135 140

Pro Phe Gln Arg Ala Ala Val Leu Cys Leu Asp Gly Val Gly Glu Trp
 145 150 155 160

Ala Thr Thr Ser Val Trp Leu Gly Glu Gly Asn Lys Leu Thr Pro Gln
 165 170 175

Trp Glu Ile Asp Phe Pro His Ser Leu Gly Leu Leu Tyr Ser Ala Phe
 180 185 190

Thr Tyr Tyr Thr Gly Phe Lys Val Asn Ser Gly Glu Tyr Lys Leu Met
 195 200 205

Gly Leu Ala Pro Tyr Gly Glu Pro Lys Tyr Val Asp Gln Ile Leu Lys
 210 215 220

His Leu Leu Asp Leu Lys Glu Asp Gly Thr Phe Arg Leu Asn Met Asp
 225 230 235 240

Tyr Phe Asn Tyr Thr Val Gly Leu Thr Met Thr Asn His Lys Phe His
 245 250 255

Ser Met Phe Gly Gly Pro Pro Arg Gln Ala Glu Gly Lys Ile Ser Gln
 260 265 270

Arg Asp Met Asp Leu Ala Ser Ser Ile Gln Lys Val Thr Glu Glu Val
 275 280 285

Ile Leu Arg Leu Ala Arg Thr Ile Lys Lys Glu Leu Gly Val Glu Tyr
 290 295 300

Leu Cys Leu Ala Gly Gly Val Gly Leu Asn Cys Val Ala Asn Gly Arg
 305 310 315 320

Ile Leu Arg Glu Ser Asp Phe Lys Asp Ile Trp Ile Gln Pro Ala Ala
 325 330 335

Gly Asp Ala Gly Ser Ala Val Gly Ala Ala Leu Ala Ile Trp His Glu
 340 345 350

Tyr His Lys Lys Pro Arg Thr Ser Thr Ala Gly Asp Arg Met Lys Gly
 355 360 365

Ser Tyr Leu Gly Pro Ser Phe Ser Glu Ala Glu Ile Leu Gln Phe Leu

370 375 380
 Asn Ser Val Asn Ile Pro Tyr His Arg Cys Val Asp Asn Glu Leu Met
 385 390 395 400
 Ala Arg Leu Ala Glu Ile Leu Asp Gln Gly Asn Val Val Gly Trp Phe
 405 410 415
 Ser Gly Arg Met Glu Phe Gly Pro Arg Ala Leu Gly Gly Arg Ser Ile
 420 425 430
 Ile Gly Asp Ser Arg Ser Pro Lys Met Gln Ser Val Met Asn Leu Lys
 435 440 445
 Ile Lys Tyr Arg Glu Ser Phe Arg Pro Phe Ala Pro Ser Val Leu Ala
 450 455 460
 Glu Arg Val Ser Asp Tyr Phe Asp Leu Asp Arg Pro Ser Pro Tyr Met
 465 470 475 480
 Leu Leu Val Ala Gln Val Lys Glu Asn Leu His Ile Pro Met Thr Gln
 485 490 495
 Glu Gln His Glu Leu Phe Gly Ile Glu Lys Leu Asn Val Pro Arg Ser
 500 505 510
 Gln Ile Pro Ala Val Thr His Val Asp Tyr Ser Ala Arg Ile Gln Thr
 515 520 525
 Val His Lys Glu Thr Asn Pro Arg Tyr Tyr Glu Leu Ile Arg His Phe
 530 535 540
 Glu Ala Arg Thr Gly Cys Ala Val Leu Val Asn Thr Ser Phe Asn Val
 545 550 555 560
 Arg Gly Glu Pro Ile Val Cys Thr Pro Glu Asp Ala Tyr Arg Cys Phe
 565 570 575

Met Arg Thr Glu Met Asp Tyr Leu Val Met Glu Asn Phe Leu Leu Val
 580 585 590

Lys Ser Glu Gln Pro Arg Gly Asn Ser Asp Glu Ser Trp Gln Lys Glu
 595 600 605

Phe Glu Leu Asp
 610

<210> 26

<211> 444

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 26

atgagtgaat tttccaca aaaaagtggt aaattaaaga tggaacagat aaaagaactt 60

gacaaaaaag gattgcgtga gtttgactg attggcgggt ctatagtggc gggtttattc 120

gggtttttac tgccagttat acgccatcat tccttatcag ttatcccttg ggttggtgct 180

ggatttctct ggatttgggc aataatcgca cctacgactt taagttttat ttaccaaata 240

tggatgagga ttggacttgt ttaggatgg atacaaacac gaattathtt gggagtthta 300

ttttatataa tgatcacacc aataggattc ataagacggc tgttgaatca agatccaatg 360

acgcgaatct tcgagccaga gttgccaact tatcgccaat tgagtaagtc aagaactaca 420

caaagtatgg agaaaccatt ctaa 444

<210> 27

<211> 147

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 27

Met Ser Glu Phe Phe Pro Gln Lys Ser Gly Lys Leu Lys Met Glu Gln
 1 5 10 15

Ile Lys Glu Leu Asp Lys Lys Gly Leu Arg Glu Phe Gly Leu Ile Gly
 20 25 30

Gly Ser Ile Val Ala Val Leu Phe Gly Phe Leu Leu Pro Val Ile Arg
 35 40 45

His His Ser Leu Ser Val Ile Pro Trp Val Val Ala Gly Phe Leu Trp
 50 55 60

Ile Trp Ala Ile Ile Ala Pro Thr Thr Leu Ser Phe Ile Tyr Gln Ile
 65 70 75 80

Trp Met Arg Ile Gly Leu Val Leu Gly Trp Ile Gln Thr Arg Ile Ile
 85 90 95

Leu Gly Val Leu Phe Tyr Ile Met Ile Thr Pro Ile Gly Phe Ile Arg
 100 105 110

Arg Leu Leu Asn Gln Asp Pro Met Thr Arg Ile Phe Glu Pro Glu Leu
 115 120 125

Pro Thr Tyr Arg Gln Leu Ser Lys Ser Arg Thr Thr Gln Ser Met Glu
 130 135 140

Lys Pro Phe
 145

<210> 28

<211> 165

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 28

atgctaaaag acacttgga tttattaaa gacattgccg gattattaa agaacaaaaa 60

aactatttgt tgattccct aattatcacc ctggtatcct tgggggcgct gattgtcttt 120

gctcaatctt ctgcgatcgc accttcatt tacactcttt tttaa 165

<210> 29

<211> 54

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 29

Met Leu Lys Asp Thr Trp Asp Phe Ile Lys Asp Ile Ala Gly Phe Ile
1 5 10 15

Lys Glu Gln Lys Asn Tyr Leu Leu Ile Pro Leu Ile Ile Thr Leu Val
20 25 30

Ser Leu Gly Ala Leu Ile Val Phe Ala Gln Ser Ser Ala Ile Ala Pro
35 40 45

Phe Ile Tyr Thr Leu Phe
50

<210> 30

<211> 1299

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 30

atgagtaact tcaaggggtc ggtaaagata gcattgatgg gaatattgat ttttgtggg 60
ctaattcttg gcgtagcatt tgttgaaatt ggggtacgta ttgccgggat cgaacacata 120
gcattccata gcattgatga acacaggggg tgggtagggc gacctcatgt ttccgggtgg 180
tatagaaccg aaggtgaagc tcacatccaa atgaatagtg atggctttcg agatcgagaa 240
cacatcaagg tcaaaccaga aaataccttc aggatagcgc tgttgggaga ttcctttgta 300
gagtccatgc aagtaccgtt ggagcaaaat ttggcagcag ttatagaagg agaaatcagt 360
agttgtatag cttagctgg acgaaaggcg gaagtgatta atttggagt gactggttat 420
ggaacagacc aagaactaat tactctacgg gagaaagttt gggactattc acctgatata 480
gtagtgctag atttttatag tggcaacgac attgttgata actcccgtgc gctgagtcag 540
aaattctatc ctaatgaact aggttcacta aagccgtttt ttatacttag agatggtaat 600
ctggtggttg atgcttcgtt tatcaatacg gataattatc gctcaaagct gacatggtgg 660
ggcaaaactt atatgaaaat aaaagaccac tcacggattt tacaggtttt aaacatggta 720
cgggatgctc ttaacaactc tagtagaggg ttttcttctc aagctataga ggaaccgtta 780

tttagtgatg gaaaacagga tacaaaattg agcggggttt ttgatatcta caaaccacct 840
 actgacctg aatggcaaca ggcattggcaa gtcacagaga aactgattag ctcaatgcaa 900
 cacgaggtga ctgcgaagaa agcagatttt ttagttgta ctttggcgg tcccttcaa 960
 cgagaacctt tagtgctga aaaagaaatg caagaattgg gtctgactga ttggttttac 1020
 ccagagaagc gaattacacg ttgggtgag gatgaggggt tcagtgtact caatctcagc 1080
 ccaaatttgc aggtttatc tgagcagaac aatgcttgcc tatatgggtt tgatgatact 1140
 caaggctgtg tagggcattg gaatgcttta ggacatcagg tagcaggaaa aatgattgca 1200
 tcgaagattt gtcaacagca gatgagagaa agtatattgc ctcataagca cgaccttca 1260
 agccaaagct cacctattac ccaatcagtg atccaataa 1299

<210> 31

<211> 432

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 31

Met Ser Asn Phe Lys Gly Ser Val Lys Ile Ala Leu Met Gly Ile Leu
 1 5 10 15

Ile Phe Cys Gly Leu Ile Phe Gly Val Ala Phe Val Glu Ile Gly Leu
 20 25 30

Arg Ile Ala Gly Ile Glu His Ile Ala Phe His Ser Ile Asp Glu His
 35 40 45

Arg Gly Trp Val Gly Arg Pro His Val Ser Gly Trp Tyr Arg Thr Glu
 50 55 60

Gly Glu Ala His Ile Gln Met Asn Ser Asp Gly Phe Arg Asp Arg Glu
 65 70 75 80

His Ile Lys Val Lys Pro Glu Asn Thr Phe Arg Ile Ala Leu Leu Gly
 85 90 95

Asp Ser Phe Val Glu Ser Met Gln Val Pro Leu Glu Gln Asn Leu Ala
 100 105 110

Ala Val Ile Glu Gly Glu Ile Ser Ser Cys Ile Ala Leu Ala Gly Arg
 115 120 125

Lys Ala Glu Val Ile Asn Phe Gly Val Thr Gly Tyr Gly Thr Asp Gln
 130 135 140

Glu Leu Ile Thr Leu Arg Glu Lys Val Trp Asp Tyr Ser Pro Asp Ile
 145 150 155 160

Val Val Leu Asp Phe Tyr Thr Gly Asn Asp Ile Val Asp Asn Ser Arg
 165 170 175

Ala Leu Ser Gln Lys Phe Tyr Pro Asn Glu Leu Gly Ser Leu Lys Pro
 180 185 190

Phe Phe Ile Leu Arg Asp Gly Asn Leu Val Val Asp Ala Ser Phe Ile
 195 200 205

Asn Thr Asp Asn Tyr Arg Ser Lys Leu Thr Trp Trp Gly Lys Thr Tyr
 210 215 220

Met Lys Ile Lys Asp His Ser Arg Ile Leu Gln Val Leu Asn Met Val
 225 230 235 240

Arg Asp Ala Leu Asn Asn Ser Ser Arg Gly Phe Ser Ser Gln Ala Ile
 245 250 255

Glu Glu Pro Leu Phe Ser Asp Gly Lys Gln Asp Thr Lys Leu Ser Gly
 260 265 270

Phe Phe Asp Ile Tyr Lys Pro Pro Thr Asp Pro Glu Trp Gln Gln Ala
 275 280 285

Trp Gln Val Thr Glu Lys Leu Ile Ser Ser Met Gln His Glu Val Thr

290 295 300
 Ala Lys Lys Ala Asp Phe Leu Val Val Thr Phe Gly Gly Pro Phe Gln
 305 310 315 320
 Arg Glu Pro Leu Val Arg Gln Lys Glu Met Gln Glu Leu Gly Leu Thr
 325 330 335
 Asp Trp Phe Tyr Pro Glu Lys Arg Ile Thr Arg Leu Gly Glu Asp Glu
 340 345 350
 Gly Phe Ser Val Leu Asn Leu Ser Pro Asn Leu Gln Val Tyr Ser Glu
 355 360 365
 Gln Asn Asn Ala Cys Leu Tyr Gly Phe Asp Asp Thr Gln Gly Cys Val
 370 375 380
 Gly His Trp Asn Ala Leu Gly His Gln Val Ala Gly Lys Met Ile Ala
 385 390 395 400
 Ser Lys Ile Cys Gln Gln Gln Met Arg Glu Ser Ile Leu Pro His Lys
 405 410 415
 His Asp Pro Ser Ser Gln Ser Ser Pro Ile Thr Gln Ser Val Ile Gln
 420 425 430
 <210> 32
 <211> 1449
 <212> DNA
 <213> *Cylindrospermopsis raciborskii* T3
 <400> 32
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 agatcggagg cacgagttag cctccaactg gcaattccct tagtccttgt cgaaatatgc 120
 ggaacgagta ttaatgtggt ggatgtagtc atgatgggct tacttggtac tcaagttttg 180
 gctgctggtg ccttgggtgc gatcgctttt ttatctgtat cgaatacttg ttataatatg 240
 cttttgtcgg gggtagcaaa ggcactgtag gcttttgggg caaacaaaat agatcaggtt 300

agtcgtattg cttctgggca aatatggctg gcactcacct tgtctttgcc tgcaatgctt 360
 ttgctttggt atatggatac tatattgggtg ctatttggtc aagttgaaag caacacatta 420
 attgcaaaaa cgtatttaca ctcaattgtg tggggatttc cggcggcagt tggatatttg 480
 atattaagag gcattgcctc tgctgtgaac gtccccaat tggtaactgt gacgatgcta 540
 gtagggctgg tcttgaatgc cccggccaat tatgtattaa tgttcggtaa atttggcttt 600
 cctgaacttg gtttagctgg aataggctgg gcaagtactt tggtttttg gattagtttt 660
 ctagtggggg ttgtcttgc t gattttctcc caaaagtta gagattataa actttccgc 720
 tacttgcatc agttgatcg acagacgggt gtggaaattt ttcaaactgg atggcctatg 780
 ggtttctac tgggagtggg atcagtagta ttgagcctca ccgcttggtt aacaggctat 840
 ttgggaacag taacattagc agctcatgag atcgcatcc aaacagcaga actggcgata 900
 gtgataccac tcggaatcgg gaatgttgcc gtcacgagag taggtcagac tataggagaa 960
 aaaaaccctt tgggtgctag aagggcagca ttgattggga ttatgattgg tggcatttat 1020
 gccagtcttg tggcagtcat ttctgggtg ttccatatac agattgcggg actttattta 1080
 aaaataaacg atccagagag tatggaagca gtaagacag caactaattt tctctcttg 1140
 gcgggattat tccaatttt tcatagcgtt caaataattg ttgtgggggt ttaatatagg 1200
 ttgcaggata cgtttatccc attgttaatg aatttggtag gctgggggtct tggcttggca 1260
 gtaagctatt acatgggaat cattttatgt tggggaggta tgggtatctg gttaggtctg 1320
 gttttgagtc cactcctgtc cggacttatt ttaatggttc gttttatca agagattgcc 1380
 aataggattg ccaatagtga tgatgggcaa gagagtatat ctattgacaa cgttgaagaa 1440
 ctctcctga 1449

<210> 33

<211> 482

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 33

Met Thr Asn Thr Glu Arg Gly Leu Ala Glu Ile Thr Ser Thr Gly Tyr

1 5 10 15

Lys Ser Glu Leu Arg Ser Glu Ala Arg Val Ser Leu Gln Leu Ala Ile
20 25 30

Pro Leu Val Leu Val Glu Ile Cys Gly Thr Ser Ile Asn Val Val Asp
35 40 45

Val Val Met Met Gly Leu Leu Gly Thr Gln Val Leu Ala Ala Gly Ala
50 55 60

Leu Gly Ala Ile Ala Phe Leu Ser Val Ser Asn Thr Cys Tyr Asn Met
65 70 75 80

Leu Leu Ser Gly Val Ala Lys Ala Ser Glu Ala Phe Gly Ala Asn Lys
85 90 95

Ile Asp Gln Val Ser Arg Ile Ala Ser Gly Gln Ile Trp Leu Ala Leu
100 105 110

Thr Leu Ser Leu Pro Ala Met Leu Leu Leu Trp Tyr Met Asp Thr Ile
115 120 125

Leu Val Leu Phe Gly Gln Val Glu Ser Asn Thr Leu Ile Ala Lys Thr
130 135 140

Tyr Leu His Ser Ile Val Trp Gly Phe Pro Ala Ala Val Gly Ile Leu
145 150 155 160

Ile Leu Arg Gly Ile Ala Ser Ala Val Asn Val Pro Gln Leu Val Thr
165 170 175

Val Thr Met Leu Val Gly Leu Val Leu Asn Ala Pro Ala Asn Tyr Val
180 185 190

Leu Met Phe Gly Lys Phe Gly Leu Pro Glu Leu Gly Leu Ala Gly Ile
195 200 205

Gly Trp Ala Ser Thr Leu Val Phe Trp Ile Ser Phe Leu Val Gly Val
 210 215 220

Val Leu Leu Ile Phe Ser Pro Lys Val Arg Asp Tyr Lys Leu Phe Arg
 225 230 235 240

Tyr Leu His Gln Phe Asp Arg Gln Thr Val Val Glu Ile Phe Gln Thr
 245 250 255

Gly Trp Pro Met Gly Phe Leu Leu Gly Val Glu Ser Val Val Leu Ser
 260 265 270

Leu Thr Ala Trp Leu Thr Gly Tyr Leu Gly Thr Val Thr Leu Ala Ala
 275 280 285

His Glu Ile Ala Ile Gln Thr Ala Glu Leu Ala Ile Val Ile Pro Leu
 290 295 300

Gly Ile Gly Asn Val Ala Val Thr Arg Val Gly Gln Thr Ile Gly Glu
 305 310 315 320

Lys Asn Pro Leu Gly Ala Arg Arg Ala Ala Leu Ile Gly Ile Met Ile
 325 330 335

Gly Gly Ile Tyr Ala Ser Leu Val Ala Val Ile Phe Trp Leu Phe Pro
 340 345 350

Tyr Gln Ile Ala Gly Leu Tyr Leu Lys Ile Asn Asp Pro Glu Ser Met
 355 360 365

Glu Ala Val Lys Thr Ala Thr Asn Phe Leu Phe Leu Ala Gly Leu Phe
 370 375 380

Gln Phe Phe His Ser Val Gln Ile Ile Val Val Gly Val Leu Ile Gly
 385 390 395 400

Leu Gln Asp Thr Phe Ile Pro Leu Leu Met Asn Leu Val Gly Trp Gly

405 410 415

Leu Gly Leu Ala Val Ser Tyr Tyr Met Gly Ile Ile Leu Cys Trp Gly
 420 425 430

Gly Met Gly Ile Trp Leu Gly Leu Val Leu Ser Pro Leu Leu Ser Gly
 435 440 445

Leu Ile Leu Met Val Arg Phe Tyr Gln Glu Ile Ala Asn Arg Ile Ala
 450 455 460

Asn Ser Asp Asp Gly Gln Glu Ser Ile Ser Ile Asp Asn Val Glu Glu
 465 470 475 480

Leu Ser

<210> 34

<211> 831

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 34

atgaaaacaa acaaacatat agctatgtgg gcttgccta gaagtcgtc tactgtaatt 60

accctgtcct ttgagaactt agatgggtgt gttgtttatg atgagcctct agaggctccg 120

aatgtcttga tgacaactta cacgatgagt aacagtcgta cgtagcaga agaagactta 180

aagcaattaa tactgcaaaa taatgtagaa acagacctca agaaagttat agaacaattg 240

actggagatt taccggacgg aaaattatc tcatttcaaa aatgataac aggtgactat 300

agatctgaat ttggaataga ttgggcaaaa aagctaacta acttctttt aataaggcat 360

ccccaagata ttatttttcc ttctgatata gcggagagaa agacaggtat cacagaacca 420

ttcacacaac aaaatcttgg catgaaaaca ctttatgaag tttccaaca aattgaagtt 480

attacagggc aaacaccttt agttattcac tcagatgata taattaaaaa ccctccttct 540

gctttgaaat ggctgtgtaa aaacttaggg ctgcatttg atgaaaagat gctgacatgg 600

aaagcaaatc tagaagactc caatttaaag tatacaaaat tatatgctaa ttctgcgtct 660

ggcagttcag aaccttgggt tgaaacttta agatcgacca aaacatttct cgcctatgaa 720

aagaaggaga aaaaattacc agctcgggta atacctctac tagatgaatc tattccttac 780

tatgaaaaac tcttacagca ttgtcatatt ttgtaatggc cagaacactg a 831

<210> 35

<211> 276

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 35

Met Lys Thr Asn Lys His Ile Ala Met Trp Ala Cys Pro Arg Ser Arg
1 5 10 15

Ser Thr Val Ile Thr Arg Ala Phe Glu Asn Leu Asp Gly Cys Val Val
20 25 30

Tyr Asp Glu Pro Leu Glu Ala Pro Asn Val Leu Met Thr Thr Tyr Thr
35 40 45

Met Ser Asn Ser Arg Thr Leu Ala Glu Glu Asp Leu Lys Gln Leu Ile
50 55 60

Leu Gln Asn Asn Val Glu Thr Asp Leu Lys Lys Val Ile Glu Gln Leu
65 70 75 80

Thr Gly Asp Leu Pro Asp Gly Lys Leu Phe Ser Phe Gln Lys Met Ile
85 90 95

Thr Gly Asp Tyr Arg Ser Glu Phe Gly Ile Asp Trp Ala Lys Lys Leu
100 105 110

Thr Asn Phe Phe Leu Ile Arg His Pro Gln Asp Ile Ile Phe Ser Phe
115 120 125

Asp Ile Ala Glu Arg Lys Thr Gly Ile Thr Glu Pro Phe Thr Gln Gln
130 135 140

Asn Leu Gly Met Lys Thr Leu Tyr Glu Val Phe Gln Gln Ile Glu Val
 145 150 155 160

Ile Thr Gly Gln Thr Pro Leu Val Ile His Ser Asp Asp Ile Ile Lys
 165 170 175

Asn Pro Pro Ser Ala Leu Lys Trp Leu Cys Lys Asn Leu Gly Leu Ala
 180 185 190

Phe Asp Glu Lys Met Leu Thr Trp Lys Ala Asn Leu Glu Asp Ser Asn
 195 200 205

Leu Lys Tyr Thr Lys Leu Tyr Ala Asn Ser Ala Ser Gly Ser Ser Glu
 210 215 220

Pro Trp Phe Glu Thr Leu Arg Ser Thr Lys Thr Phe Leu Ala Tyr Glu
 225 230 235 240

Lys Lys Glu Lys Lys Leu Pro Ala Arg Leu Ile Pro Leu Leu Asp Glu
 245 250 255

Ser Ile Pro Tyr Tyr Glu Lys Leu Leu Gln His Cys His Ile Phe Glu
 260 265 270

Trp Ser Glu His
 275

<210> 36

<211> 774

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 36

ctaaaaattt ttttctactc ttttcaggat agaattccag tttctagagc cgttgtaacc 60

gtacatatct tgatagtacg tatcgatgag gtactcattt tcgtggagca ttaaccagct 120

ttttaactcc gctaatttct gctctccttt ttctattaat tcttgetcat ccaaatac 180

cctgtccaac tcttcctgt ccaactccca catagttttg ttggtatctt cgacaatcaa 240

gtagtctcca ctttttagac cgttttcgtg aaaatattca actactccca ccgcattagc 300
 atgggcatct tctacgatca accagggatg agcaagccca gaaagcagtt ccgacgacat 360
 tattgcaccc atattgttac aatccccctc taaaaaatga acgcgagagt cagtttttgc 420
 tttctcgtcg agtagggaaa gatcgatata gatacagtag acacaacctt ctatttgga 480
 cagttctaag tgatcggcta gccaaatcgc gctgccaccg cttaatgctc ctatttcgat 540
 tattgttttc gggcgaagct catacaggag cattgaataa agagctattt cgggtgcaccc 600
 tttcaggaag ggtatccctt tccaagtga caaatcgcg tttgccaaga gcgctctcca 660
 agctggcact ggaatagcac atttatcttc tcttcagaa attttgcaa accgattagg 720
 ttgaaaggt gcaactttat aggcggcttc ttgaacaaat tttggaagc tcat 774

<210> 37

<211> 257

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 37

Met Ser Phe Gln Lys Phe Val Gln Glu Ala Ala Tyr Lys Val Ala Pro
 1 5 10 15

Phe Lys Pro Asn Arg Phe Ala Lys Ile Ser Glu Arg Glu Asp Lys Cys
 20 25 30

Ala Ile Pro Val Pro Ala Trp Arg Ala Leu Leu Ala Asn Arg Asp Leu
 35 40 45

Phe Thr Trp Lys Gly Ile Pro Phe Leu Lys Gly Cys Thr Glu Ile Ala
 50 55 60

Leu Tyr Ser Met Leu Leu Tyr Glu Leu Arg Pro Lys Thr Ile Ile Glu
 65 70 75 80

Ile Gly Ala Leu Ser Gly Gly Ser Ala Ile Trp Leu Ala Asp His Leu
 85 90 95

Glu Leu Phe Gln Ile Glu Gly Cys Val Tyr Cys Ile Asp Ile Asp Leu
 100 105 110

Ser Leu Leu Asp Glu Lys Ala Lys Thr Asp Ser Arg Val His Phe Leu
 115 120 125

Glu Gly Asp Cys Asn Asn Met Gly Ala Ile Met Ser Ser Glu Leu Leu
 130 135 140

Ser Gly Leu Ala His Pro Trp Leu Ile Val Glu Asp Ala His Ala Asn
 145 150 155 160

Ala Val Gly Val Val Glu Tyr Phe His Glu Asn Gly Leu Lys Ser Gly
 165 170 175

Asp Tyr Leu Ile Val Glu Asp Thr Asn Lys Thr Met Trp Glu Leu Asp
 180 185 190

Arg Glu Glu Leu Asp Arg Asp Asp Leu Asp Glu Gln Glu Leu Ile Glu
 195 200 205

Lys Gly Glu Gln Lys Leu Ala Glu Leu Lys Ser Trp Leu Met Leu His
 210 215 220

Glu Asn Glu Tyr Leu Ile Asp Thr Tyr Tyr Gln Asp Met Tyr Gly Tyr
 225 230 235 240

Asn Gly Ser Arg Asn Trp Asn Ser Ile Leu Lys Arg Val Glu Lys Asn
 245 250 255

Phe

<210> 38

<211> 327

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 38

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gttgactgca acattggtat gagattgcgg cgcaacatag agcgcacccg caggacaata 180

tgcttcacag atgaaacaag ttgacagtc ttctgtcgg gcgatcgcag gcggttggtt 240

gggaactgca tcaaagacat tggtagggca tacttgagcg caaacattac aattaataca 300

gagtttatgg ctgacaagct cgatcat 327

<210> 39

<211> 108

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 39

Met Ile Glu Leu Val Ser His Lys Leu Cys Ile Asn Cys Asn Val Cys

1 5 10 15

Val Gln Val Cys Pro Thr Asn Val Phe Asp Ala Val Pro Asn Gln Pro

20 25 30

Pro Ala Ile Ala Arg Gln Glu Asp Cys Gln Thr Cys Phe Ile Cys Glu

35 40 45

Ala Tyr Cys Pro Ala Asp Ala Leu Tyr Val Ala Pro Gln Ser His Thr

50 55 60

Asn Val Ala Val Asn Glu Asp Asp Leu Ile Asp Ser Gly Ile Met Gly

65 70 75 80

Glu Tyr Arg Arg Ile Leu Gly Trp Gly Tyr Gly Arg Lys Asn Asn Ser

85 90 95

Glu Leu Asp Thr Asp His Lys Leu Arg Leu Phe Glu

100 105

<210> 40

<211> 1653

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 40

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gtctatgcga atatgttcgc tacgcgttc cttgcgatgt aaagcgctaa aatatgccca   180
tcgtgctaca gacacaagag cagccgctcg acgagaaaat tccagatcgc gcactgtatc   240
ttgttcggg ttcccttgta cttgctgcca cagcatttct aatttggcga gggaatccaa   300
aagtcctgc tcacagcgca agtaattctt ctctaattgg aacatctcgg cttgtacacc   360
gcggacaact gcctcgctat cgaatgttc ggaaccaggg tactgggaac gtaatccggc   420
ttgacctgct ggacgcacaa cccgttcatt gacatgagcg cccaaactct tggcaaaggc   480
ggctgcacct tcccctgccc attgtcctgt agagattgcc caagcagcat taggaccatc   540
acccccagaa gctatcccag ctaaaaactc ccgcgatgct gcatctccgg cggcatacag   600
tccaggaact ttgtaccac aactatcatt cacaatccga attccacctg taccacggac   660
tgtaccttct aaaaccagtg ttacaggtac tcgttctgta taagggtcaa tgccagcttt   720
tttatagggt agaaaggcga tgaagtgaga ctttcaacc aatgcttggg ttccaggtgt   780
ggctcgatcc aaacgagcat aaacgggacc ttccaggagg gcattgggca ggaacgatgg   840
atcgcgacga ccattgatat agccaccaag atcgttacct gcctcatcgg tgtaactagc   900
ccagtaaaag ggagcagccc ttgtcactgt ggcatgaaa gcggtcgaga tggatatgtg   960
actggaagct tccatactgg agagtgcgc gccagcttcc accgccatca gcagtccatc  1020
gcctgtattg gtattgcaac ctaaagcttt acttaggaat gcacaaccgc cattcgctag  1080
aactactgca ccagcgcgaa cggtataggt gcgatgatt tgctctgta cacctctagc  1140
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ttgcacaccc acacgaaca ggttcttgcg aagtaccgcg atatattccg gaccataata  1260
actctggcgc acggattccc cattttctt ggggaaacga tagccccaat ctccactaa  1320
gggcaaactc agccaagctt ttcaattac acgttcaatc caacgtaagt tagcgaggtt  1380

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gacgctattg ccaactggcag cagctgcacc gctcgtacct agaaaacctt tatcaacaat 1500
gatgactttg acaccttggg ctccagccgc ccatgctgcc catgcggcgg caggaccacc 1560
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<210> 41
<211> 550
<212> PRT
<213> *Cylindrospermopsis raciborskii* T3

<400> 41

Met Asn Leu Thr Leu Asn Lys Glu Glu Lys Gln Leu Leu Thr Ala Tyr
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Ser Gly Thr Glu Leu Gln Leu Thr Ala Asp Val Leu Val Ile Gly Gly
20 25 30

Gly Pro Ala Ala Ala Trp Ala Ala Trp Ala Ala Gly Ala Gln Gly Val
35 40 45

Lys Val Ile Ile Val Asp Lys Gly Phe Leu Gly Thr Ser Gly Ala Ala
50 55 60

Ala Ala Ser Gly Asn Ser Val Met Ala Pro Ser Pro Glu Asn Trp Glu
65 70 75 80

Lys Asp Val Ser Glu Cys Tyr Ser Lys Gly Asn Asn Leu Ala Asn Leu
85 90 95

Arg Trp Ile Glu Arg Val Ile Glu Lys Ala Trp Leu Ser Leu Pro Leu
100 105 110

Val Glu Asp Trp Gly Tyr Arg Phe Pro Lys Glu Asn Gly Glu Ser Val
115 120 125

Arg Gln Ser Tyr Tyr Gly Pro Glu Tyr Met Arg Val Leu Arg Lys Asn
 130 135 140

Leu Leu Arg Val Gly Val Gln Ile Phe Asp Gln Ser Pro Ala Leu Glu
 145 150 155 160

Leu Leu Leu Ala Gln Asp Gly Ser Val Ala Gly Ala Arg Gly Val Gln
 165 170 175

Arg Gln Asn His Arg Thr Tyr Thr Val Arg Ala Gly Ala Val Val Leu
 180 185 190

Ala Asn Gly Gly Cys Ala Phe Leu Ser Lys Ala Leu Gly Cys Asn Thr
 195 200 205

Asn Thr Gly Asp Gly Leu Leu Met Ala Val Glu Ala Gly Gly Glu Leu
 210 215 220

Ser Ser Met Glu Ala Ser Ser His Tyr Thr Ile Ser Thr Ala Phe Asn
 225 230 235 240

Ala Thr Val Thr Arg Ala Ala Pro Phe Tyr Trp Ala Ser Tyr Thr Asp
 245 250 255

Glu Ala Gly Asn Asp Leu Gly Gly Tyr Ile Asn Gly Arg Arg Asp Pro
 260 265 270

Ser Phe Leu Pro Asn Ala Leu Leu Lys Gly Pro Val Tyr Ala Arg Leu
 275 280 285

Asp Arg Ala Thr Pro Glu Ile Gln Ala Leu Val Glu Lys Ser His Phe
 290 295 300

Ile Ala Phe Leu Pro Tyr Lys Lys Ala Gly Ile Asp Pro Tyr Thr Glu
 305 310 315 320

Arg Val Pro Val Thr Leu Val Leu Glu Gly Thr Val Arg Gly Thr Gly

325 330 335

Gly Ile Arg Ile Val Asn Asp Ser Cys Gly Thr Lys Val Pro Gly Leu
340 345 350

Tyr Ala Ala Gly Asp Ala Ala Ser Arg Glu Phe Leu Ala Gly Ile Ala
355 360 365

Ser Gly Gly Asp Gly Pro Asn Ala Ala Trp Ala Ile Ser Thr Gly Gln
370 375 380

Trp Ala Gly Glu Gly Ala Ala Ala Phe Ala Lys Ser Leu Gly Ala His
385 390 395 400

Val His Glu Arg Val Val Arg Pro Ala Gly Gln Ala Gly Leu Arg Ser
405 410 415

Gln Tyr Pro Gly Ser Glu Thr Phe Asp Ser Glu Ala Val Val Arg Gly
420 425 430

Val Gln Ala Glu Met Phe Pro Leu Glu Lys Asn Tyr Leu Arg Cys Glu
435 440 445

Gln Gly Leu Leu Asp Ser Leu Ala Lys Leu Glu Met Leu Trp Gln Gln
450 455 460

Val Gln Gly Asn Pro Lys Gln Asp Thr Val Arg Asp Leu Glu Phe Ser
465 470 475 480

Arg Arg Ala Ala Ala Leu Val Ser Val Ala Arg Trp Ala Tyr Phe Ser
485 490 495

Ala Leu His Arg Lys Glu Thr Arg Ser Glu His Ile Arg Ile Asp Tyr
500 505 510

Pro Glu Thr Asp Pro Asn Gln Leu Tyr Tyr Gln Ala Thr Gly Gly Leu
515 520 525

Glu Arg Leu Trp Val Arg Arg Asp Trp Val Lys Asp Ala Ser Ala Thr
 530 535 540

Pro Pro Val Leu Thr Thr
 545 550

<210> 42

<211> 750

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 42

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 aagttgttct ttactaagct gtttcgtgat cgggtcagaa attaagtcac taaatggcgt 180
 atcgactaaa cctggctcaa tgatggtaac gcgaatgttg tctaaagata cctcctggcg 240
 taatgcttct gaaagagcat tgacgcctga ttggcagca ctataaacga ccgcaccgga 300
 ctgcgctatc ctgccatcga cagaagatat attgactata tgaccggatt tttgggcctt 360
 cagaagaggc aaaactgcgt ggatagcata taaaactccc agaacattca catcgaatgc 420
 tcgcctccag tctgcgggat ttccagtatc aattgcacca aacacaccaa ttctgcatt 480
 attcaccaaa atatctacat gtcttagctc aaccttggtc ttttgacta gatgatttac 540
 ttgagattcg tctgtaatat ctgtaacaat aggcaatgct tgaccaccac tggcttcaat 600
 ccgttttgcg agtgcacgca aaagctcagc acgtcttgcg gcgatcgcaa cttttgcccc 660
 ctccgcagct aaagcaaatg ctgtagcctc tccaatccca gaggaagctc cagtaataat 720
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<210> 43

<211> 249

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 43

Met Ala Gly Lys Leu Asp Gly Lys Val Ala Ile Ile Thr Gly Ala Ser

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Ser Gly Ile Gly Glu Ala Thr Ala Phe Ala Leu Ala Ala Glu Gly Ala
20 25 30

Lys Val Ala Ile Ala Ala Arg Arg Ala Glu Leu Leu His Ala Leu Ala
35 40 45

Lys Arg Ile Glu Ala Ser Gly Gly Gln Ala Leu Pro Ile Val Thr Asp
50 55 60

Ile Thr Asp Glu Ser Gln Val Asn His Leu Val Gln Lys Thr Lys Val
65 70 75 80

Glu Leu Gly His Val Asp Ile Leu Val Asn Asn Ala Gly Ile Gly Val
85 90 95

Phe Gly Ala Ile Asp Thr Gly Asn Pro Ala Asp Trp Arg Arg Ala Phe
100 105 110

Asp Val Asn Val Leu Gly Val Leu Tyr Ala Ile His Ala Val Leu Pro
115 120 125

Leu Leu Lys Ala Gln Lys Ser Gly His Ile Val Asn Ile Ser Ser Val
130 135 140

Asp Gly Arg Ile Ala Gln Ser Gly Ala Val Val Tyr Ser Ala Ala Lys
145 150 155 160

Ser Gly Val Asn Ala Leu Ser Glu Ala Leu Arg Gln Glu Val Ser Leu
165 170 175

Asp Asn Ile Arg Val Thr Ile Ile Glu Pro Gly Leu Val Asp Thr Pro
180 185 190

Phe Asn Asp Leu Ile Ser Asp Pro Ile Thr Lys Gln Leu Ser Lys Glu
195 200 205

Gln Leu Ser Thr Ile Thr Pro Leu Gln Ser Glu Asp Ile Ala Arg Ala
 210 215 220

Ile Ile Tyr Ala Val Thr Gln Pro Asp His Val Asn Val Asn Glu Ile
 225 230 235 240

Leu Ile Arg Pro Thr Ala Glu Asp Asn
 245

<210> 44

<211> 1005

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 44

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taacagtggg aaacgtaccg gctgtgtgac atgtaagatc cgaatatctt gttctattgt 180

ttcgatcatat tcagttagca tctttgactc taacgtttca taccggtcc acattatcaa 240

catacgcaat aactatcttt cctcatcaat cgggtgtgac gtcattaaat ccacaatcct 300

catttcaggg gattctgaaa cgcagtattg acataaagga tgactaagcc tgaaccaatt 360

aacccaagag tcattcttga tatggctgac aatccttgat gtctggaatt gatacttacc 420

catagtaagg ccattcttat ctaatttcac ctcaaattct tccacttttg tataattgcg 480

atcacctaac caaccgtcat ggataaaagg aaaatgagac acgtctaagg aattatccat 540

cacacgaaac gcactagctt taatcaagta agacttggtg taagtcttgt gataattcgg 600

atcatcccat tcaggaaatg aaggatatatc attaacagga tcgccaagc acaccacac 660

taagccatag cgctcctggg agtgatatgt cctggcttca gcacttgccg gtggtaccat 720

gccagggtga gctgggatct gtatgcattt accagcctca ttgtatctcc atccgtgata 780

cggacaaact aaagtattat tcgtaatttc tccatagac agaggaaacac ctcggtgggg 840

gcagtagtca agccatacct gtatgggtga atttgttca taactgcgcc ataataccaa 900

cttactccc aacaaacgag atctggtgat acttcagggt ttacagtctt ctacattggc 960

gactacgtgc cagttattga ttaagattgg gtcggtagtt gtcac 1005

<210> 45

<211> 334

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 45

Met Thr Thr Thr Asp Pro Ile Leu Ile Asn Asn Trp His Val Val Ala
1 5 10 15

Asn Val Glu Asp Cys Lys Pro Gly Ser Ile Thr Arg Ser Arg Leu Leu
20 25 30

Gly Val Lys Leu Val Leu Trp Arg Ser Tyr Glu Gln Asn Ser Pro Ile
35 40 45

Gln Val Trp Leu Asp Tyr Cys Pro His Arg Gly Val Pro Leu Ser Met
50 55 60

Gly Glu Ile Thr Asn Asn Thr Leu Val Cys Pro Tyr His Gly Trp Arg
65 70 75 80

Tyr Asn Glu Ala Gly Lys Cys Ile Gln Ile Pro Ala His Pro Gly Met
85 90 95

Val Pro Pro Ala Ser Ala Glu Ala Arg Thr Tyr His Ser Gln Glu Arg
100 105 110

Tyr Gly Leu Val Trp Val Cys Leu Gly Asp Pro Val Asn Asp Ile Pro
115 120 125

Ser Phe Pro Glu Trp Asp Asp Pro Asn Tyr His Lys Thr Tyr Thr Lys
130 135 140

Ser Tyr Leu Ile Lys Ala Ser Ala Phe Arg Val Met Asp Asn Ser Leu
145 150 155 160

Asp Val Ser His Phe Pro Phe Ile His Asp Gly Trp Leu Gly Asp Arg
 165 170 175

Asn Tyr Thr Lys Val Glu Glu Phe Glu Val Lys Leu Asp Lys Asp Gly
 180 185 190

Leu Thr Met Gly Lys Tyr Gln Phe Gln Thr Ser Arg Ile Val Ser His
 195 200 205

Ile Glu Asp Asp Ser Trp Val Asn Trp Phe Arg Leu Ser His Pro Leu
 210 215 220

Cys Gln Tyr Cys Val Ser Glu Ser Pro Glu Met Arg Ile Val Asp Leu
 225 230 235 240

Met Thr Ile Thr Pro Ile Asp Glu Glu Asn Ser Val Leu Arg Met Leu
 245 250 255

Ile Met Trp Asn Gly Tyr Glu Thr Leu Glu Ser Lys Met Leu Thr Glu
 260 265 270

Tyr Asp Glu Thr Ile Glu Gln Asp Ile Arg Ile Leu His Ala Gln Gln
 275 280 285

Pro Val Arg Leu Pro Leu Leu Thr Pro Lys Gln Ile Asn Thr Gln Leu
 290 295 300

Phe Ser His Glu Ile His Val Pro Ser Asp Arg Cys Thr Leu Ala Tyr
 305 310 315 320

Arg Arg Trp Leu Lys Gln Leu Gly Val Thr Tyr Gly Val Cys
 325 330

<210> 46

<211> 726

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 46

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 tgggtgataa aacattgctg cccctcctgg aagtgaaga catatttctg catttagtga 180
 attggcagaa gatgaatcta atgagtgttc ccattgggtgg ctacttgga taactcgcat 240
 tgtaccata gtattatctg taccctgtaa gtatatagtt atgaatacca tggcttgatt 300
 ggctactgga accaacaacc gaagcgcgtc gtcatttaac tcgtttttg acatggatgc 360
 aagtgcgttc aatactcaa ctacatatcc atggtcttga tgccaagcaa tgtatcctgt 420
 acctgcacga attatggcta gatcgggtgat caataggaag atacagacc caattagagc 480
 ctgtactggt cccatcacag ttggaagctc taaaagcctc tgaattatct ttgataacct 540
 aactggatct gggatagtat gctcagacca ccactcatag tcacccgcca atactcccc 600
 acgtttttgt tcggtaataa gttctacttc atgccgtatt tcttcaatta acgcttttgg 660
 tacagcttct tcaactgtga aataaccatc atttgtgtaa gcttgttttt gttccgctgt 720
 gagcat 726

<210> 47

<211> 241

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 47

Met Leu Thr Ala Glu Gln Lys Gln Ala Tyr Thr Asn Asp Gly Tyr Phe

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Thr Val Glu Glu Ala Val Pro Lys Ala Leu Ile Glu Glu Ile Arg His

20 25 30

Glu Val Glu Leu Ile Thr Glu Gln Lys Arg Gly Gly Val Leu Ala Gly

35 40 45

Asp Tyr Glu Trp Trp Ser Glu His Thr Ile Pro Asp Pro Val Arg Tyr

50 55 60

Gln Lys Ile Ile Gln Arg Leu Leu Glu Leu Pro Thr Val Met Gly Pro
 65 70 75 80

Val Gln Ala Leu Ile Gly Ser Asp Ile Phe Leu Leu Ile Thr Asp Leu
 85 90 95

Ala Ile Ile Arg Ala Gly Thr Gly Tyr Ile Ala Trp His Gln Asp His
 100 105 110

Gly Tyr Val Val Glu Val Leu Asn Ala Leu Ala Ser Met Ser Lys Asn
 115 120 125

Glu Leu Asn Asp Asp Ala Leu Arg Leu Leu Val Pro Val Ala Asn Gln
 130 135 140

Ala Met Val Phe Ile Thr Ile Tyr Leu Gln Asp Thr Asp Asn Thr Met
 145 150 155 160

Gly Thr Met Arg Val Ile Pro Ser Ser His Gln Trp Glu His Ser Leu
 165 170 175

Asp Ser Ser Ser Ala Asn Ser Leu Asn Ala Glu Ile Cys Leu Ser Leu
 180 185 190

Pro Gly Gly Ala Ala Met Phe Tyr Thr Pro Thr Val Trp His Thr Ala
 195 200 205

Ala Ala Asn Thr Ser Ile Thr Asp Tyr Arg Met Leu Thr Leu Ile Phe
 210 215 220

Thr Lys Asn Asn Ile Lys Pro Leu Leu Val Asp Ala Leu Lys Arg Ile
 225 230 235 240

Ile

<210> 48

<211> 576

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 48

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agaagctcgc tccgacttca tttagccaa ttttctgca gaccaatact gaaaatatcc 120

caatcttaat aattcatcac tagcctcttg taactggctg aatgactgta ctgatgctaa 180

aacatactta gggtagagta tgattacgtt attcacattc tccgcgtcat caccaacata 240

ttgtttgtct ggatgcgac ctaaagctac caaatcgat tctggtaata cataattcgc 300

cttggaatg tacctttcca acctctgtgc atctaggtt tgagggtcgc agccaaaaat 360

caccatttca aagtcattat tccatgttct tatctgttcc attagaagct ctggcagttc 420

aggtccatga aaccaacgaa cactaacacg gttatttaac caagctgcct tcgcgtaagg 480

acagggtgga aaatttctg ttagaggatt gggaatgctg acaacattga taatccaatc 540

ctctatttct tggcgaaatt gttcgatatt tatcat 576

<210> 49

<211> 191

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 49

Met Ile Asn Ile Glu Gln Phe Arg Gln Glu Ile Glu Asp Trp Ile Ile

1 5 10 15

Asn Val Val Ser Ile Pro Asn Pro Leu Thr Gly Asn Phe Pro Pro Cys

20 25 30

Pro Tyr Ala Lys Ala Ala Trp Leu Asn Asn Arg Val Ser Val Arg Trp

35 40 45

Phe His Gly Pro Glu Leu Pro Glu Leu Leu Met Glu Gln Ile Arg Thr

50 55 60

Trp Asn Asn Asp Phe Glu Met Val Ile Phe Gly Cys Asp Pro Gln Asn

65

70

75

80

Leu Asp Ala Gln Arg Leu Glu Arg Tyr Ile Thr Lys Ala Asn Tyr Val
 85 90 95

Leu Pro Glu Tyr Asp Leu Val Ala Leu Gly Ser His Pro Asp Lys Gln
 100 105 110

Tyr Val Gly Asp Asp Ala Glu Asn Val Asn Asn Val Ile Ile Thr His
 115 120 125

Pro Lys Tyr Val Leu Ala Ser Val Gln Ser Phe Ser Gln Leu Gln Glu
 130 135 140

Ala Ser Asp Glu Leu Leu Arg Leu Gly Tyr Phe Gln Tyr Trp Ser Ala
 145 150 155 160

Glu Lys Leu Ala Glu Met Lys Ser Glu Arg Ala Ser His Asn Leu Ser
 165 170 175

Ser Ile Gln Arg Lys Asn Ser Tyr Arg Ile Ile Pro Thr Asn His
 180 185 190

<210> 50

<211> 777

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 50

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atagtcaatt ttgacaggta ctaagccttt gggaataata tagtcaccag ttcttgaaa 120

acgcatccca actctatctt cccaaccgtc aatagtatca ttaattgttg tggatttaa 180

acagatccct gcaattttag ccccatgttt gacattaact cgtaaccaag ggtcaaatat 240

aagaccattt ttatctcgcc aggtaataata ccgctctatg ggtataagtg ggtaaagata 300

ttttaggctt ggacgtgcag ccatgatcaa agaattaaga ccgtggtatt gagcaagttc 360

tttcatgtat ccaatcagat actgactcaa gttttgcct tgatactctg gtaggattga 420

aatcgatact acacataacg cattaggcag gcggttctgt tctcggtctt caagccactt 480
 ggctaaagcc cagtcacaac ctctgtccgg taactcatca aaacggcttt cataagttaa 540
 agggatacag ttccttgcg ctatcataag ctgtgtggta gcttctacta acccaaactg 600
 gaattctgga taaatttcaa atagagctaa ggaagctgga tctgcccaga catcatgtat 660
 caaaaatttt gggatgctt gatcaaagac actcatcgtc cttccacaa aatcagaagt 720
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<210> 51

<211> 258

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 51

Met Thr Ile Gln Ile Val Gln His Asn Leu Glu Tyr Ser Phe Val Thr
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Pro Lys Glu Thr Ser Asp Phe Val Glu Arg Thr Met Ser Val Phe Asp
 20 25 30

Gln Ala Tyr Pro Lys Phe Leu Ile His Asp Val Trp Ala Asp Pro Ala
 35 40 45

Ser Leu Ala Leu Phe Glu Ile Tyr Pro Glu Phe Gln Phe Gly Leu Val
 50 55 60

Glu Ala Thr Thr Gln Leu Met Ile Ala Gln Gly Asn Cys Ile Pro Leu
 65 70 75 80

Thr Tyr Glu Ser Arg Phe Asp Glu Leu Pro Asp Glu Gly Cys Asp Trp
 85 90 95

Ala Leu Ala Lys Trp Leu Glu Asp Arg Glu Gln Asn Arg Leu Pro Asn
 100 105 110

Ala Leu Cys Val Val Ser Ile Ser Ile Leu Pro Glu Tyr Gln Gly Lys

115 120 125
 Asn Leu Ser Gln Tyr Leu Ile Gly Tyr Met Lys Glu Leu Ala Gln Tyr
 130 135 140
 His Gly Leu Asn Ser Leu Ile Met Ala Ala Arg Pro Ser Leu Lys Tyr
 145 150 155 160
 Leu Tyr Pro Leu Ile Pro Ile Glu Arg Tyr Ile Thr Trp Arg Asp Lys
 165 170 175
 Asn Gly Leu Ile Phe Asp Pro Trp Leu Arg Val Asn Val Lys His Gly
 180 185 190
 Ala Lys Ile Ala Gly Ile Cys Phe Lys Ser Thr Thr Ile Asn Asp Thr
 195 200 205
 Ile Asp Gly Trp Glu Asp Arg Val Gly Met Arg Phe Pro Glu Thr Gly
 210 215 220
 Asp Tyr Ile Ile Pro Lys Gly Leu Val Pro Val Lys Ile Asp Tyr Pro
 225 230 235 240
 Asn Asn Met Gly Ile Tyr Ile Glu Pro Asn Ile Trp Leu Tyr Tyr Asp
 245 250 255

Leu Asp

<210> 52

<211> 777

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 52

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tacttgcaact gtcaattcat taccgacttt cccattccca ggcataatta ataagtagg 120

gtgaggtgga atgccgtcgt actgtcggac gcggcgaaaa atgctcgaat tctcgccacc 180

atgtttattc aagaggactt caactgggtg gatgacaaaa gtcattcctg acccaaggtg 240
 gcgcgatcgc cgcttttcat ttgctggagt ggaaacacta acaaataagg cacaccctcc 300
 tagagaataa gaccagttag cagactgcgg atcggcagac caatggcagg gacaagacac 360
 cgcataagg ctatgtaacg cattcaaaaa atcaaatgct tgacctgcat attcctctac 420
 tgtaagaact gttggttcag gtgggaaaaa gatgacaagt gtcagaagat ccgcattttc 480
 gtgctgaagc aattcgtttt cattaacttc atcaatgtat ttgtagatac cctcaagcgt 540
 atgctcaacc aagatcgggt cagttaaaga tgagactatc aggtatctaa tcattccctt 600
 ctgtccccc atagttcccc agaagcaagg gaaggcagaa tcgctgattg ttcaacaaa 660
 tgttgagtag ctagtgcgta cccaagcagg aaggcactcc tctagaagag aggattccat 720
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<210> 53

<211> 258

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 53

Met Val Ile Lys Asn Leu Cys Pro Asp Gly Val Thr Pro Ile Trp Asn
 1 5 10 15

Lys Ser Gln Met Glu Ser Ser Leu Leu Glu Glu Cys Leu Pro Ala Trp
 20 25 30

Val Arg Thr Ser Tyr Ser Thr Phe Val Glu Thr Ile Ser Asp Ser Ala
 35 40 45

Phe Pro Cys Phe Trp Gly Thr Ile Gly Glu Gln Lys Gly Met Ile Arg
 50 55 60

Tyr Leu Ile Val Ser Ser Leu Thr Asp Pro Ile Leu Val Glu His Thr
 65 70 75 80

Leu Glu Gly Ile Tyr Lys Tyr Ile Asp Glu Val Asn Glu Asn Glu Leu

85 90 95

Leu Gln His Glu Asn Ala Asp Leu Leu Thr Leu Val Ile Phe Phe Pro
100 105 110

Pro Glu Pro Thr Val Leu Thr Val Glu Glu Tyr Ala Gly Gln Ala Phe
115 120 125

Asp Phe Leu Asn Ala Leu His Ser Leu Asp Ala Val Ser Cys Pro Cys
130 135 140

His Trp Ser Ala Asp Pro Gln Ser Ala Asn Trp Ser Tyr Ser Leu Gly
145 150 155 160

Gly Cys Ala Leu Phe Val Ser Val Ser Thr Pro Ala Asn Gln Lys Arg
165 170 175

Arg Ser Arg His Leu Gly Ser Gly Met Thr Phe Val Ile Thr Pro Val
180 185 190

Glu Val Leu Leu Asn Lys His Gly Gly Glu Asn Ser Ser Ile Phe Arg
195 200 205

Arg Val Arg Gln Tyr Asp Gly Ile Pro Pro His Pro Asn Leu Leu Ile
210 215 220

Met Pro Gly Asn Gly Lys Val Gly Asn Glu Leu Thr Val Gln Val Leu
225 230 235 240

Pro Asp Asn Asn Asp Ser Glu Ile Ser Phe Asp Phe Gln Tyr Lys Phe
245 250 255

Lys Asp

<210> 54

<211> 1227

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 54

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agattcagga ttagtaccga ctggaaagt agtagctatg ccaaagacc cattaccatt  180

tcctggtaag accgagacgt tattgtact ataattgta acagccaggt caagtttact  240

gtcgccattc acatctctaa tcgtacaga gtagggatta gtaccggctg gaaagttagt  300

ggctgcgcca aaagacccat taccatttc cagtaagacc gagacgttat tgctgctagt  360

atttgaaca gccagggtcaa gcttgctgc gccattaca tccccagttg ctacaaatat  420

gggattagta ccgactggaa agttagtggc tgcgcaaaa gaccattac catttccag  480

taagaccgag acgttattgc tgaccaatt tgtaatagca aggtcgagct tactgtcgct  540

attaaaatcc gcaatcgcta cggaaatcga ataagtatcg acaggaagc tgctggctgc  600

gccaaaagac ccattaccat ttccagtaa aaccaagacc ttattgtcga accaatttgt  660

aaaagcaagg tcaagctcac tatcgttatt cacatctcca atggctacag aataagggtt  720

agtaccaact gaaaagttag tggtgcgcc aaaagacca ttaccatttc ctagtaagac  780

cgagacgtta ttgtactaa aattgcaac agccagggtca agcttgctgt cgccatttac  840

atccccagtc actacaaaga cgggattagt accgactgga aagttagtgg ctgcgcaaaa  900

agaccatta ccatttcca gtaagaccga gacgttattg tcgaaccaat ttgtaacagc  960

caggtcgagc ttactatcg tattgaaatc ccaactgct acagagtcag catcaagacc 1020

agttgggaag ttaatagcag tagcataact actcgttg gcaaattca ctctacgga 1080

caaattaacc ggaacactaa attgccaga aagcttttca ttctcagat aatagtcagt 1140

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tataattagt aaagtgagcc tttcac                                     1227

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<210> 55

<211> 408

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 55

Met Lys Arg Leu Thr Leu Leu Ile Ile Ala Gly Ile Leu Ser Val Ser
 1 5 10 15

Thr Phe Leu Cys Ile Thr Pro Val Ala Leu Ala Asn Ile Thr Asp Tyr
 20 25 30

Tyr Leu Lys Asn Glu Lys Leu Ser Gly Gln Phe Ser Val Pro Val Asn
 35 40 45

Leu Ser Val Gly Val Arg Phe Ala His Arg Ser Ser Tyr Ala Thr Ala
 50 55 60

Ile Asn Phe Pro Thr Gly Leu Asp Ala Asp Ser Val Ala Val Gly Asp
 65 70 75 80

Phe Asn Ser Asp Ser Lys Leu Asp Leu Ala Val Thr Asn Trp Phe Asp
 85 90 95

Asn Asn Val Ser Val Leu Leu Gly Asn Gly Asn Gly Ser Phe Gly Ala
 100 105 110

Ala Thr Asn Phe Pro Val Gly Thr Asn Pro Val Phe Val Val Thr Gly
 115 120 125

Asp Val Asn Gly Asp Ser Lys Leu Asp Leu Ala Val Ala Asn Phe Ser
 130 135 140

Ser Asn Asn Val Ser Val Leu Leu Gly Asn Gly Asn Gly Ser Phe Gly
 145 150 155 160

Ala Ala Thr Asn Phe Ser Val Gly Thr Asn Pro Tyr Ser Val Ala Ile
 165 170 175

Gly Asp Val Asn Asn Asp Ser Glu Leu Asp Leu Ala Phe Thr Asn Trp
 180 185 190

Phe Asp Asn Lys Val Leu Val Leu Leu Gly Asn Gly Asn Gly Ser Phe
 195 200 205

Gly Ala Ala Ser Ser Phe Pro Val Asp Thr Tyr Ser Ile Ser Val Ala
 210 215 220

Ile Ala Asp Phe Asn Ser Asp Ser Lys Leu Asp Leu Ala Ile Thr Asn
 225 230 235 240

Trp Val Ser Asn Asn Val Ser Val Leu Leu Gly Asn Gly Asn Gly Ser
 245 250 255

Phe Gly Ala Ala Thr Asn Phe Pro Val Gly Thr Asn Pro Ile Phe Val
 260 265 270

Ala Thr Gly Asp Val Asn Gly Asp Ser Lys Leu Asp Leu Ala Val Ala
 275 280 285

Asn Thr Ser Ser Asn Asn Val Ser Val Leu Leu Gly Asn Gly Asn Gly
 290 295 300

Ser Phe Gly Ala Ala Thr Asn Phe Pro Ala Gly Thr Asn Pro Tyr Ser
 305 310 315 320

Val Ala Ile Arg Asp Val Asn Gly Asp Ser Lys Leu Asp Leu Ala Val
 325 330 335

Thr Asn Tyr Ser Ser Asn Asn Val Ser Val Leu Pro Gly Asn Gly Asn
 340 345 350

Gly Ser Phe Gly Ile Ala Thr Asn Phe Pro Val Gly Thr Asn Pro Glu
 355 360 365

Ser Ile Ala Ile Ala Asp Phe Asn Gly Asp Ser Lys Leu Asp Leu Ala
 370 375 380

Val Thr Asn Ser Gly Asn Asn Asn Val Ser Ile Leu Leu Asn Asn Phe

385 390 395 400

Gln Gly Leu Pro Lys Asn Lys Ile
405

<210> 56

<211> 603

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 56

ctattgttg aaaattgtga attgttttc cacgtattg agtagttgtt ctaggctttc 60

ctcgacgggtg agttcggatg ttccacca taaatctggg ctattgggtg gttcataagg 120

ggcgctgatt cccgtaaatc catctatttc ccactgcgt gcttttagat aaagacctt 180

cggatcacgc tgctcacaaa gttccagtgg agttgcaatg tatacttcat gaaatagatc 240

tccagctagt ctacgcacct gttctcggtc attcctgtag ggtgagatga aggcagtgat 300

cactaggcat cctgactccg caaagagttt ggcaacctca ccaaacgac ggatattttc 360

tgagcgatca ctagcagaaa atcctaaatc ggaacacagt ccatgacgaa cactatcacc 420

atctaaaaca aaggtagacc atcctttctc gaacaaagtc tgctctaatt ttaaagccaa 480

tgtgtttta ccagccccgg acagtccagt aaacctaga atcccgcttt tatgaccatt 540

cttagataa cgatcatatg gagatataag atgttttgta tagtgaatat tagttgattt 600

cat 603

<210> 57

<211> 200

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 57

Met Lys Ser Thr Asn Ile His Tyr Thr Lys His Leu Ile Ser Pro Tyr
1 5 10 15

Asp Arg Tyr Leu Lys Asn Gly His Lys Ser Gly Ile Leu Trp Phe Thr
20 25 30

Gly Leu Ser Gly Ala Gly Lys Thr Thr Leu Ala Leu Lys Leu Glu Gln
 35 40 45

Thr Leu Phe Glu Lys Gly Trp Ser Thr Phe Val Leu Asp Gly Asp Ser
 50 55 60

Val Arg His Gly Leu Cys Ser Asp Leu Gly Phe Ser Ala Ser Asp Arg
 65 70 75 80

Ser Glu Asn Ile Arg Arg Leu Gly Glu Val Ala Lys Leu Phe Ala Glu
 85 90 95

Ser Gly Cys Leu Val Ile Thr Ala Phe Ile Ser Pro Tyr Arg Asn Asp
 100 105 110

Arg Glu Gln Val Arg Arg Leu Ala Gly Asp Leu Phe His Glu Val Tyr
 115 120 125

Ile Ala Thr Pro Leu Glu Leu Cys Glu Gln Arg Asp Pro Lys Gly Leu
 130 135 140

Tyr Leu Lys Ala Arg Ser Gly Glu Ile Asp Gly Phe Thr Gly Ile Ser
 145 150 155 160

Ala Pro Tyr Glu Pro Pro Asn Ser Pro Asp Leu Trp Val Glu Thr Ser
 165 170 175

Glu Leu Thr Val Glu Glu Ser Leu Glu Gln Leu Leu Lys Tyr Val Glu
 180 185 190

Asn Lys Phe Thr Ile Phe Lys Gln
 195 200

<210> 58

<211> 1350

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 58

ttaagaaaaa attatttcaa actcgctcgc caaacgctcc ataatcaaat taatttcaga 60
 cgaaaaagga cagtaatatg gtagctctac caacaccctt ctgcggaag ctgtcacctt 120
 cgctgctatt ttgataatcg ttcccttaa cctaggaacc tgggctttag ccagttttgt 180
 tccctgtgct gcttgccgaa ttccaacat taaaatgtaa gctgcttgag ataaaaataa 240
 ccgaaactga ttgacaataa atttctcaca gctgagtcta tctgatttta tcccagttt 300
 taattcctta attctatgct ctgaagtagc tcctctttga acataaaatt tatcgataa 360
 atcctgagct tctgtttcca agctagtaat tataaatcta ggattgggtc cttttctag 420
 ccattctgct ttcataatta ctgcccaggg ttctgaccaa ctccgagctg cgtaatacac 480
 atcatcaaat aaacgaactt ttctcctgt gcgacaatat tccagtctgg ctcggtcaag 540
 aaggttaatta attttcgtt ttaagacatc attattgctg aatccaaaaa catatccaac 600
 cccgcttttt tcacaaacct caatgatttc tggtaacgag aaacccccgt ctcccctcag 660
 aacaattcta atttcaggta aggctctttt gattcgcaa aataaccatt ttagaatgcc 720
 agctactcct ttaccagagt gagaatttcc cgcccttagt tgtagaacta atggataacc 780
 actggaagct tcattaatca gaactggaaa gtatataatca tgcctatggt aaccattaaa 840
 taagctcagt tgttgatgac catgagttag agcatccac gcatctatgt ccaggacaat 900
 ctcttttgat tcccaggat aggattctag gaatttatca acaataacc gacgaatttg 960
 ttgatatct tttgagtca cctgatttcc taaacgactc atagtgggt gactagctaa 1020
 taagtttct cctactgtgg gaacttgatt acaactagc ttaaaaattg gatcttggcg 1080
 caatttatta ctatcgtgc tatctcata gccagcaatt atttgataaa ttcgttggt 1140
 aattaattga gaaagagaat gttgacttt agtttggtcc cgattatccg tcaaacaatc 1200
 tgccatatct tgacaaattt ttaccttct tctacttgt cgtgccagaa taattccgcc 1260
 atcactactt aaactcatat cagaaaaagt cagatctaaa gttttttat cgaagaaatt 1320
 taaagataat cttaggaag atttagcat 1350

<210> 59

<211> 449

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 59

Met Thr Lys Ser Ser Ser Arg Leu Ser Leu Asn Phe Phe Asp Lys Lys

1 5 10 15

Thr Leu Asp Leu Thr Phe Ser Asp Met Ser Leu Ser Ser Asp Gly Gly

20 25 30

Ile Ile Leu Ala Arg Gln Val Glu Glu Lys Val Lys Ile Cys Gln Asp

35 40 45

Met Ala Asp Cys Leu Thr Asp Asn Arg Asp Gln Thr Lys Val Lys His

50 55 60

Ser Leu Ser Gln Leu Ile Ser Gln Arg Ile Tyr Gln Ile Ile Ala Gly

65 70 75 80

Tyr Glu Asp Ser Asn Asp Ser Asn Lys Leu Arg Gln Asp Pro Ile Phe

85 90 95

Lys Leu Val Cys Asn Gln Val Pro Thr Val Gly Glu Asn Leu Leu Ala

100 105 110

Ser Gln Pro Thr Met Ser Arg Leu Glu Asn Gln Val Thr Gln Lys Asp

115 120 125

Ile Lys Gln Ile Arg Arg Leu Phe Val Asp Lys Phe Leu Glu Ser Tyr

130 135 140

Pro Arg Glu Ser Lys Glu Ile Val Leu Asp Ile Asp Ala Trp Asp Ala

145 150 155 160

Leu Thr His Gly His Gln Gln Leu Ser Leu Phe Asn Gly Tyr His Arg

165 170 175

His Asp Ile Tyr Phe Pro Val Leu Ile Asn Glu Ala Ser Ser Gly Tyr

180 185 190

Pro Leu Val Leu Gln Leu Arg Ala Gly Asn Ser His Ser Gly Lys Gly
195 200 205

Val Ala Gly Ile Leu Lys Trp Leu Phe Leu Arg Ile Lys Arg Ala Leu
210 215 220

Pro Glu Ile Arg Ile Val Leu Arg Gly Asp Gly Gly Phe Ser Leu Pro
225 230 235 240

Glu Ile Ile Glu Val Cys Glu Lys Ser Gly Val Gly Tyr Val Phe Gly
245 250 255

Phe Ser Asn Asn Asp Val Leu Lys Arg Lys Ile Asn Tyr Leu Leu Asp
260 265 270

Arg Ala Arg Leu Glu Tyr Cys Arg Thr Gly Glu Lys Val Arg Leu Phe
275 280 285

Asp Asp Val Tyr Tyr Ala Ala Arg Ser Trp Ser Glu Pro Arg Arg Val
290 295 300

Ile Met Lys Ala Glu Trp Leu Glu Lys Gly Pro Asn Pro Arg Phe Ile
305 310 315 320

Ile Thr Ser Leu Glu Thr Glu Ala Gln Asp Leu Tyr Asp Lys Phe Tyr
325 330 335

Val Gln Arg Gly Ala Thr Ser Glu His Arg Ile Lys Glu Leu Lys Leu
340 345 350

Gly Ile Lys Ser Asp Arg Leu Ser Cys Glu Lys Phe Ile Val Asn Gln
355 360 365

Phe Arg Leu Phe Leu Ser Gln Ala Ala Tyr Ile Leu Met Leu Gly Ile
370 375 380

Arg Gln Ala Ala Gln Gly Thr Lys Leu Ala Lys Ala Gln Val Pro Arg
 385 390 395 400

Leu Arg Glu Thr Ile Ile Lys Ile Ala Ala Lys Val Thr Val Ser Ala
 405 410 415

Arg Arg Val Leu Val Glu Leu Pro Tyr Tyr Cys Pro Phe Ser Ser Glu
 420 425 430

Ile Asn Leu Ile Met Glu Arg Leu Ala Ser Glu Phe Glu Ile Ile Phe
 435 440 445

Ser

<210> 60

<211> 666

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 60

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cattctctct aaacactgaa ttgctaattg taatagtaaa atgggctcca ctaccccgagg 120

aacatctttc tgctgcgcca aattacgata taactttttg taagcatcat ctactgtatc 180

atctaataat ttaatccttc taccactaat ctgctctaaa tccgctaaag ctactaggct 240

ggtagccaac atagattggg catgatcgga cataatggca acctcccca aagtaggatg 300

gggggggatag ggaaatattt tcattgctat ttctgcaaaa tctttggcat agtccccaat 360

acgttccaag tctctaacta attgcatgaa tgagcttaaa caccgagatt cttggtctgt 420

gggagcttga ctgctcataa ttgtggcaca atcgacttct attgtctgtt agaagcgatc 480

aatttttttg tctaactccc gtatttgctc agctgctgtt aaatcccgat tgaatagagc 540

ttggtgactc agacggaatg actgctctac taaagcacc atacgcaaaa catctcgttc 600

cagtctttta atggcacgta taggttgagg ttttcaaaa attgtatatt tcacaacagc 660

tttcat

666

<210> 61

<211> 221

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 61

Met Lys Ala Val Val Lys Tyr Thr Ile Phe Glu Lys Pro Gln Pro Ile
 1 5 10 15

Arg Ala Ile Lys Arg Leu Glu Arg Asp Val Leu Arg Met Gly Ala Leu
 20 25 30

Val Glu Gln Ser Phe Arg Leu Ser His Gln Ala Leu Phe Asn Arg Asp
 35 40 45

Leu Thr Ala Ala Glu Gln Ile Arg Arg Leu Asp Lys Lys Ile Asp Arg
 50 55 60

Phe Tyr Arg Gln Ile Glu Val Asp Cys Ala Thr Ile Met Ser Ser Gln
 65 70 75 80

Ala Pro Thr Asp Gln Glu Ser Arg Cys Leu Ser Ser Phe Met Gln Leu
 85 90 95

Val Arg Asp Leu Glu Arg Ile Gly Asp Tyr Ala Lys Asp Leu Ala Glu
 100 105 110

Ile Ala Met Lys Ile Phe Pro Tyr Pro Pro His Pro Thr Leu Gly Glu
 115 120 125

Val Ala Ile Met Ser Asp His Ala Gln Ser Met Leu Ala Thr Ser Leu
 130 135 140

Val Ala Leu Ala Asp Leu Asp Glu Ile Ser Gly Arg Arg Ile Lys Leu
 145 150 155 160

Leu Asp Asp Thr Val Asp Asp Ala Tyr Lys Lys Leu Tyr Arg Asn Leu

165

170

175

Ala Gln Gln Lys Asp Val Pro Gly Val Val Glu Pro Ile Leu Leu Leu
 180 185 190

Thr Leu Ala Ile Gln Cys Leu Glu Arg Met Ala Asp His Ala Thr Asn
 195 200 205

Ile Gly Gln Arg Val Ala Tyr Ile Val Thr Gly Gln Arg
 210 215 220

<210> 62

<211> 1353

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 62

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accggtatca ggatggttca tggccctgat ttgccacca tgagctataa ttatttggcg 120

gacaatggat aaccctaaac cactaccagt aatttctact gtttcattct cagagcggga 180

ctcgcggtgt ctagctttgt cccccgata aaatctttga aagacatggg gtagatccat 240

gggagcaaat ccaaccccg aatcaataat gtaatttct aaaatctgat ttgatacttg 300

gtttaatatt gtatctgctt ctggatcaac cccattaata gacttctccc caciaactgg 360

attcatttca atgaaaatag taccgttcag gttgctgtat ttaatacagt tatctaacag 420

attaagaaac acttgataaa ttctggactt atcagcacat atatagacct ttccggggcc 480

ggagtaagaa atactaagat gctgattagc ggctaggggc tctaaattct cccagactga 540

aaaaattagg gagcggactt ctagcatttc caaattcagt tgtatggagg aggttatttc 600

catctgggtc aggtctaacc aattttggac taaattaatt agtctgtcaa cctctgcat 660

caagcggatg acccaacggt ttagaggggg atctaagcga gtttcaggg ttctgcgac 720

cagacgaatg gaagtcagag gtgttctcag ttcatgggcc aggtctgaaa aagagcggtc 780

acgttgctga tgaatgtcta caaattgttg gtgactttct agaaacacac ccactgtcc 840

ccccggtagg ggaaaactgt tagctgctaa agacaatggc tttaatccta aaataccctg 900

accatgatct cgggaagggt gaaaaatcca ctcttgcat tgcgggtttt gccaatcccg 960
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<210> 63
 <211> 450
 <212> PRT
 <213> *Cylindrospermopsis raciborskii* T3

<400> 63

Met Phe Leu Leu Gly Phe Leu Leu Gly Leu Ala Val Gly Phe Gly Phe
 1 5 10 15

Trp Leu Trp Gln Gln Phe Gln Leu Asn Ser His Leu Glu Gln Leu Thr
 20 25 30

Gln Pro Leu Asn Pro His Ala Glu Lys Ile Leu Leu Pro Leu Leu Ala
 35 40 45

Gly Leu His Arg Lys Ile Ser Thr Val Arg Asp Glu Gln Gln Asn Leu
 50 55 60

Arg Leu Ser Leu Lys Ala Tyr Glu Gln Leu Leu Asp Ala Ala Pro Leu
 65 70 75 80

Gly Tyr Leu Gln Val Asp Glu Glu Asn Gln Leu Leu Trp Cys Asn Gln
 85 90 95

Cys Ala Arg Glu Met Leu Tyr Leu Gln Arg Trp Gln Pro Gly Gln Val

100	105	110
Arg Leu Leu Leu Glu Leu Val Arg Ser Tyr Glu Leu Asp Gln Leu Ile		
115	120	125
Glu Gln Thr Arg Asp Trp Gln Lys Pro Gln Met Gln Glu Trp Ile Phe		
130	135	140
His Pro Ser Arg Asp His Gly Gln Gly Ile Leu Gly Leu Lys Pro Leu		
145	150	155
160		
Ser Leu Ala Ala Asn Ser Phe Pro Leu Pro Gly Gly Gln Val Gly Val		
165	170	175
Phe Leu Glu Ser His Gln Gln Phe Val Asp Ile His Gln Gln Arg Asp		
180	185	190
Arg Ser Phe Ser Asp Leu Ala His Glu Leu Arg Thr Pro Leu Thr Ser		
195	200	205
Ile Arg Leu Val Ala Glu Thr Leu Gln Thr Arg Leu Asp Pro Pro Leu		
210	215	220
Asn Arg Trp Val Ile Arg Leu Met Gln Glu Val Asp Arg Leu Ile Asn		
225	230	235
240		
Leu Val Gln Asn Trp Leu Asp Leu Thr Gln Met Glu Ile Thr Ser Ser		
245	250	255
Ile Gln Leu Asn Leu Glu Met Leu Glu Val Arg Ser Leu Ile Phe Ser		
260	265	270
Val Trp Glu Asn Leu Glu Pro Leu Ala Ala Asn Gln His Leu Ser Ile		
275	280	285
Ser Tyr Ser Gly Pro Glu Lys Val Tyr Ile Cys Ala Asp Lys Ser Arg		
290	295	300

Ile Tyr Gln Val Phe Leu Asn Leu Leu Asp Asn Cys Ile Lys Tyr Ser
 305 310 315 320

Asn Leu Asn Gly Thr Ile Phe Ile Glu Met Asn Pro Val Cys Gly Glu
 325 330 335

Lys Ser Ile Asn Gly Val Asp Pro Glu Ala Asp Thr Ile Leu Asn Gln
 340 345 350

Val Ser Asn Gln Ile Leu Glu Ile Asn Ile Ile Asp Ser Gly Val Gly
 355 360 365

Phe Ala Pro Met Asp Leu Pro His Val Phe Gln Arg Phe Tyr Arg Gly
 370 375 380

Asp Lys Ala Arg His Arg Glu Ser Arg Ser Glu Asn Glu Thr Val Glu
 385 390 395 400

Ile Thr Gly Ser Gly Leu Gly Leu Ser Ile Val Arg Gln Ile Ile Ile
 405 410 415

Ala His Gly Gly Lys Ile Arg Ala Met Asn His Pro Asp Thr Gly Gly
 420 425 430

Ala Trp Ile Gln Ile His Leu Pro Gln Val Val Gln His Asp Gly Gly
 435 440 445

Tyr Phe
 450

<210> 64

<211> 819

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

<400> 64

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taactctaattttccctca gccatcgaat gtgaacatcc accgttttac tgcaccaac 120

aaaatcagga ccccaaacct ggtctaataa ctgttcccg gaccacacc tcgagacata 180
 actcataaat agttctagta accggaattc ttctggtagc aagtcacct cctccctct 240
 cactaacacc cgacattcct gaggatttaa actgatatcc ttatattta aagtgggtat 300
 caagggcaaa ttagaaaacc gctgacgacg taacagggcg cgacacctag ccaccatttc 360
 ccgtacgcta aaaggcttag ttaggtaatc atccgccct acctctaaac ccagcacccg 420
 gtcagtttca ctaccttcg cactcagaat taaaatcggg atggaattac cctggtagc 480
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 gtcgaaggat aactcaccag gttgggtctc taaattcctg attaattcca cagcacaacg 600
 accatcctta gcagtcacaa ctctataacc ttaccctct aaggctacta caagcatctc 660
 tcggatcagt tcttcgtctt ccactattaa aacgcgacta actggttcaa tatccgatt 720
 agtgaagtat ctagggtaat tcagtagtat acattgataa caaaaatttg taagaatgta 780
 ctggtctggg ttcccaacta gtatatgat ctcactcat 819

<210> 65

<211> 272

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 65

Met Ser Glu Asp His Ile Leu Val Gly Asn Pro Asp Gln Tyr Ile Leu
 1 5 10 15

Thr Asn Phe Cys Tyr Gln Cys Ile Leu Leu Asn Tyr Pro Arg Tyr Phe
 20 25 30

Thr Lys Ser Asp Ile Glu Pro Val Ser Arg Val Leu Ile Val Glu Asp
 35 40 45

Glu Glu Leu Ile Arg Glu Met Leu Val Val Ala Leu Glu Gly Glu Gly
 50 55 60

Tyr Glu Val Val Thr Ala Lys Asp Gly Arg Cys Ala Val Glu Leu Ile

65 70 75 80
 Arg Asn Leu Glu Thr Gln Pro Gly Glu Leu Ser Phe Asp Leu Leu Val
 85 90 95
 Leu Asp Leu Met Leu Pro Gln Ile Asn Gly Leu Asp Ile Cys Arg Leu
 100 105 110
 Leu Arg His Gln Gly Asn Ser Ile Pro Ile Leu Ile Leu Ser Ala Lys
 115 120 125
 Gly Ser Glu Thr Asp Arg Val Leu Gly Leu Glu Val Gly Ala Asp Asp
 130 135 140
 Tyr Leu Thr Lys Pro Phe Ser Val Arg Glu Met Val Ala Arg Cys Arg
 145 150 155 160
 Ala Leu Leu Arg Arg Gln Arg Phe Ser Asn Leu Pro Leu Ile Pro Thr
 165 170 175
 Leu Lys Tyr Lys Asp Ile Ser Leu Asn Pro Gln Glu Cys Arg Val Leu
 180 185 190
 Val Arg Gly Arg Glu Val Ser Leu Ser Pro Lys Glu Phe Arg Leu Leu
 195 200 205
 Glu Leu Phe Met Ser Tyr Ala Arg Arg Val Trp Ser Arg Glu Gln Leu
 210 215 220
 Leu Asp Gln Val Trp Gly Pro Asp Phe Val Gly Asp Ser Lys Thr Val
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 Asp Val His Ile Arg Trp Leu Arg Glu Lys Leu Glu Leu Asp Pro Ser
 245 250 255
 His Pro Glu Tyr Ile Val Thr Val Arg Gly Phe Gly Tyr Arg Phe Gly
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<211> 774

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

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<210> 67

<211> 257

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 67

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20 25 30

Val Glu Thr Ala Lys Met Trp Ala Asp Gln Gly Ala Thr Arg Leu His

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Thr Ile Glu Ala Ile Thr Asn Ala Ile Ser Ile Pro Ile Glu Val Gly					
65	70	75	80		
Gly Gly Ile Arg Asp Ser Ser Ser Val Ile Gln Leu Phe Asn Leu Gly					
	85	90	95		
Val Gln Trp Ala Ile Leu Gly Thr Val Ala Val Glu Gln Pro Asp Leu					
100	105	110			
Val Gln Gly Leu Cys Glu Gln Phe Pro Gln Gln Ile Ile Ile Gly Ile					
115	120	125			
Asp Ala Arg Asn Gly Leu Val Ala Thr Arg Gly Trp Leu Glu Thr Ser					
130	135	140			
Gln Ile Leu Ala Pro Gln Leu Ala Thr Gln Met Gln Glu Leu Gly Ala					
145	150	155	160		
Ala Ala Ile Ile Tyr Thr Asp Ile Asn Arg Asp Gly Thr Leu Gln Gly					
165	170	175			
Pro Asn Leu Arg Ala Leu Arg Asp Leu Val Ser Ala Ile Ser Ile Pro					
180	185	190			
Val Ile Ala Ser Gly Gly Val Gly Ser Val Thr Asp Leu Leu Thr Leu					
195	200	205			
Leu Ser Leu Glu His Gln Gly Val Thr Gly Val Ile Val Gly Lys Ala					
210	215	220			
Leu Tyr Thr Gly Asp Ile Ser Leu Pro Glu Ala Leu Arg Ala Ile Gly					
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Pro Gly Arg Ile Gln Asp Ile Pro Pro Thr Leu Asp Phe Ser Arg Phe
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<210> 68

<211> 396

<212> DNA

<213> *Cylindrospermopsis raciborskii* T3

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<210> 69

<211> 131

<212> PRT

<213> *Cylindrospermopsis raciborskii* T3

<400> 69

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Ile Ile His Ile Ser Ser Val Ile Glu Trp Val Val Ala Ile Ser Leu
 35 40 45

Ile Trp Lys Tyr Gly Glu Leu Thr Gln Asn His Ser Trp Arg Gly Phe

50 55 60

Ala Leu Gly Met Ile Pro Ala Leu Ile Ser Ala Leu Ser Ala Cys Thr
65 70 75 80

Trp His Tyr Phe Asp Asn Pro Gln Ser Leu Glu Trp Leu Val Thr Leu
85 90 95

Gln Ala Thr Thr Thr Leu Ile Gly Asn Phe Thr Leu Trp Ala Ala Ala
100 105 110

Val Trp Val Trp Arg Ser Thr Arg Pro Asn Glu Val Leu Ser Ile Ser
115 120 125

Asn Lys Glu
130

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<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 71

caataccgaa gaggagatag 20

<210> 72

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 72

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<212> DNA

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<211> 20

<212> DNA

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<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

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<210> 75

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 75

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<210> 76

<211> 20

<212> DNA

<213> Artificial

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<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 76

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<211> 20

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<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

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ctcgactaa atattctacc 20

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<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 78

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<210> 80

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<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 80

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 945 950 955 960

Asn Lys Val Val Phe Glu Thr Glu Phe Gly Val Arg Gln Met Pro His
 965 970 975

Ile Ser Asp His Gln Ile Tyr Gly Glu Val Ile Val Pro Gly Ala Val
 980 985 990

Leu Ala Ser Leu Ile Phe Asn Ala Ala Gln Val Leu Tyr Pro Asp Tyr
 995 1000 1005

Gln His Glu Leu Thr Asp Ile Ala Phe Tyr Gln Pro Ile Ile Phe
 1010 1015 1020

His Asp Asp Asp Thr Val Ile Val Gln Ala Ile Phe Ser Pro Asp
 1025 1030 1035

Lys Ser Gln Glu Asn Gln Ser His Gln Thr Phe Pro Pro Met Ser
 1040 1045 1050

Phe Gln Ile Ile Ser Phe Met Pro Asp Gly Pro Leu Glu Asn Lys
 1055 1060 1065

Pro Lys Val His Val Thr Gly Cys Leu Arg Met Leu Arg Asp Ala
 1070 1075 1080

Gln Pro Pro Thr Leu Ser Pro Thr Glu Ile Arg Gln Arg Cys Pro
 1085 1090 1095

His Thr Val Asn Gly His Asp Trp Tyr Asn Ser Leu Val Lys Gln
 1100 1105 1110

Lys Phe Glu Met Gly Pro Ser Phe Arg Trp Val Gln Gln Leu Trp
 1115 1120 1125

His Gly Glu Asn Glu Ala Leu Thr Arg Leu His Ile Pro Asp Val
 1130 1135 1140

Val Gly Ser Val Ser Gly His Gln Leu His Gly Ile Leu Leu Asp
 1145 1150 1155

Gly Ser Leu Ser Thr Thr Ala Val Met Glu Tyr Glu Tyr Gly Asp
 1160 1165 1170

Ser Ala Thr Arg Val Pro Leu Ser Phe Ala Ser Leu Gln Leu Tyr
 1175 1180 1185

Lys Pro Val Thr Gly Thr Glu Trp Trp Cys Tyr Ala Arg Lys Ile
 1190 1195 1200

Gly Glu Phe Lys Tyr Asp Phe Gln Ile Met Asn Glu Ile Gly Glu
 1205 1210 1215

Thr Leu Val Lys Ala Ile Gly Phe Val Leu Arg Glu Ala Ser Pro
 1220 1225 1230

Glu Lys Phe Leu Arg Thr Thr Tyr Val His Asn Trp Leu Val Asp
 1235 1240 1245

Ile Glu Trp Gln Ala Gln Ser Thr Ser Leu Val Pro Ser Asp Gly
 1250 1255 1260

Thr Ile Ser Gly Ser Cys Leu Val Leu Ser Asp Gln His Gly Thr
 1265 1270 1275

Gly Ala Ala Leu Ala Gln Arg Leu Asp Asn Ala Gly Val Pro Val
 1280 1285 1290

Thr Met Ile Tyr Ala Asp Leu Ile Leu Asp Asn Tyr Glu Leu Ile
 1295 1300 1305

Phe Arg Thr Leu Pro Asp Leu Gln Gln Val Val Tyr Leu Trp Gly
 1310 1315 1320

Leu Asp Gln Lys Glu Asp Cys His Pro Met Lys Gln Ala Glu Asp
 1325 1330 1335

Asn Cys Thr Ser Val Leu Tyr Leu Val Gln Ala Leu Leu Asn Thr
 1340 1345 1350

Tyr Ser Thr Pro Pro Ser Leu Leu Ile Val Thr Cys Asp Ala Gln
 1355 1360 1365

Ala Val Val Glu Gln Asp Arg Val Asn Gly Phe Ala Gln Ser Ser
 1370 1375 1380

Leu Leu Gly Leu Ala Lys Val Ile Met Leu Glu His Pro Glu Leu
 1385 1390 1395

Ser Cys Val Tyr Met Asp Val Glu Ala Gly Tyr Leu Gln Gln Asp
 1400 1405 1410

Val Ala Asn Thr Ile Phe Thr Gln Leu Lys Arg Gly His Leu Ser
 1415 1420 1425

Lys Asp Gly Glu Glu Ser Gln Leu Ala Trp Arg Asn Gly Gln Ala
 1430 1435 1440

Tyr Val Ala Arg Leu Ser Gln Tyr Lys Pro Lys Ser Glu Gln Leu
 1445 1450 1455

Val Glu Ile Arg Ser Asp Arg Ser Tyr Leu Ile Thr Gly Gly Arg
 1460 1465 1470

Gly Gly Val Gly Leu Gln Ile Ala Arg Trp Leu Val Glu Lys Gly
 1475 1480 1485

Ala Lys His Leu Val Leu Leu Gly Arg Ser Gln Thr Ser Ser Glu
 1490 1495 1500

Val Ser Leu Val Leu Asp Glu Leu Glu Ser Ala Gly Ala Gln Ile
 1505 1510 1515

Ile Val Ala Gln Ala Asp Ile Ser Asp Glu Lys Val Leu Ala Gln
 1520 1525 1530

Ile Leu Thr Asn Leu Thr Val Pro Leu Cys Gly Val Ile His Ala
 1535 1540 1545

Ala Gly Val Leu Asp Asp Ala Ser Leu Leu Gln Gln Thr Pro Ala
 1550 1555 1560

Lys Leu Lys Lys Val Leu Leu Pro Lys Ala Glu Gly Ala Trp Ile
 1565 1570 1575

Leu His Asn Leu Thr Leu Glu Gln Arg Leu Asp Phe Phe Val Leu
 1580 1585 1590

Phe Ser Ser Ala Ser Ser Leu Leu Gly Ala Pro Gly Gln Ala Asn
 1595 1600 1605

Tyr Ser Ala Ala Asn Ala Phe Leu Asp Gly Leu Ala Ala Tyr Arg
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Arg Gly Arg Gly Leu Pro Cys Leu Ser Ile Cys Trp Gly Ala Trp
 1625 1630 1635

Asp Gln Val Gly Met Ala Ala Arg Gln Gly Leu Leu Asp Lys Leu
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Pro Gln Arg Gly Glu Glu Ala Ile Pro Leu Gln Lys Gly Leu Asp
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Leu Phe Gly Glu Leu Leu Asn Glu Pro Ala Ala Gln Ile Gly Val
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Ile Pro Ile Gln Trp Thr Arg Phe Leu Asp His Gln Lys Gly Asn
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Leu Pro Phe Tyr Glu Lys Phe Ser Lys Ser Ser Arg Lys Ala Gln
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Ser Tyr Asp Ser Met Ala Val Ser His Thr Glu Asp Ile Gln Arg
 1715 1720 1725

Lys Leu Lys Gln Ala Ala Val Gln Asp Arg Pro Lys Leu Leu Glu
 1730 1735 1740

Val His Leu Arg Ser Gln Val Ala Gln Leu Leu Gly Ile Asn Val
 1745 1750 1755

Ala Glu Leu Pro Asn Glu Glu Gly Ile Gly Phe Val Thr Leu Gly
 1760 1765 1770

Leu Asp Ser Leu Thr Ser Ile Glu Leu Arg Asn Ser Leu Gln Arg
 1775 1780 1785

Thr Leu Asp Cys Ser Leu Pro Val Thr Phe Ala Phe Asp Tyr Pro
 1790 1795 1800

Thr Ile Glu Ile Ala Val Lys Tyr Leu Thr Gln Val Val Ile Ala
 1805 1810 1815

Pro Met Glu Ser Thr Ala Ser Gln Gln Thr Asp Ser Leu Ser Ala
 1820 1825 1830

Met Phe Thr Asp Thr Ser Ser Ile Gly Arg Ile Leu Asp Asn Glu
 1835 1840 1845

Thr Asp Val Leu Asp Ser Glu Met Gln Ser Asp Glu Asp Glu Ser
 1850 1855 1860

Leu Ser Thr Leu Ile Gln Lys Leu Ser Thr His Leu Asp
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<210> 83

<211> 4074

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 83

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<211> 1357

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 84

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Leu Asn Lys Ile Glu Glu Leu Gln Ala Glu Leu Asp Arg Leu Lys Tyr

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Ala Gln Arg Glu Pro Ile Ala Ile Ile Gly Met Gly Cys Arg Phe Pro
 35 40 45

Gly Ala Asp Thr Pro Glu Ala Phe Trp Lys Leu Leu His Asn Gly Val
 50 55 60

Asp Ala Ile Gln Glu Ile Pro Lys Ser Arg Trp Asp Ile Asp Asp Tyr
 65 70 75 80

Tyr Asp Pro Thr Pro Ala Thr Pro Gly Lys Met Tyr Thr Arg Phe Gly
 85 90 95

Gly Phe Leu Asp Gln Ile Ala Ala Phe Asp Pro Glu Phe Phe Arg Ile
 100 105 110

Ser Thr Arg Glu Ala Ile Ser Leu Asp Pro Gln Gln Arg Leu Leu Leu
 115 120 125

Glu Val Ser Trp Glu Ala Leu Glu Arg Ala Gly Leu Thr Gly Asn Lys
 130 135 140

Leu Thr Thr Gln Thr Gly Val Phe Val Gly Ile Ser Glu Ser Asp Tyr
 145 150 155 160

Arg Asp Leu Ile Met Arg Asn Gly Ser Asp Leu Asp Val Tyr Ser Gly
 165 170 175

Ser Gly Asn Cys His Ser Thr Ala Ser Gly Arg Leu Ser Tyr Tyr Leu
 180 185 190

Gly Leu Thr Gly Pro Asn Leu Ser Leu Asp Thr Ala Cys Ser Ser Ser
 195 200 205

Leu Val Cys Val Ala Leu Ala Val Lys Ser Leu Arg Gln Gln Glu Cys
 210 215 220

Asp Leu Ala Leu Ala Gly Gly Val Gln Ile Gln Val Ile Pro Asp Gly
 225 230 235 240

Phe Ile Lys Ala Cys Gln Ser Arg Met Leu Ser Pro Asp Gly Arg Cys
 245 250 255

Lys Thr Phe Asp Phe Gln Ala Asp Gly Tyr Ala Arg Ala Glu Gly Cys
 260 265 270

Gly Met Val Val Leu Lys Arg Leu Ser Asp Ala Ile Ala Asp Asn Asp
 275 280 285

Asn Ile Leu Ala Leu Ile Arg Gly Ala Ala Val Asn His Asp Gly Tyr
 290 295 300

Thr Ser Gly Leu Thr Val Pro Ser Gly Pro Ser Gln Arg Ala Val Ile
 305 310 315 320

Gln Gln Ala Leu Ala Asp Ala Gly Ile His Pro Asp Gln Ile Ser Tyr
 325 330 335

Ile Glu Ala His Gly Thr Gly Thr Ser Leu Gly Asp Pro Ile Glu Met
 340 345 350

Gly Ala Ile Gly Gln Val Phe Gly Gln Arg Ser Gln Met Leu Phe Val
 355 360 365

Gly Ser Val Lys Thr Asn Ile Gly His Thr Glu Ala Ala Ala Gly Ile
 370 375 380

Ala Gly Leu Ile Lys Val Val Leu Ser Met Gln His Gly Glu Ile Pro
 385 390 395 400

Ala Asn Leu His Phe Asp Gln Pro Ser Pro Tyr Ile Asn Trp Asp Gln
 405 410 415

Leu Pro Val Ser Ile Pro Thr Glu Thr Ile Pro Trp Ser Thr Ser Asp
 420 425 430

Arg Phe Ala Gly Val Ser Ser Phe Gly Phe Ser Gly Thr Asn Ser His
 435 440 445

Ile Val Leu Glu Ala Ala Pro Asn Ile Glu Gln Pro Thr Asp Asp Ile
 450 455 460

Asn Gln Thr Pro His Ile Leu Thr Leu Ala Ala Lys Thr Pro Ala Ala
 465 470 475 480

Leu Gln Glu Leu Ala Arg Arg Tyr Ala Thr Gln Ile Glu Thr Ser Pro
 485 490 495

Asp Val Pro Leu Ala Asp Ile Cys Phe Thr Ala His Ile Gly Arg Lys
 500 505 510

His Phe Lys His Arg Phe Ala Val Val Thr Glu Ser Lys Glu Gln Leu
 515 520 525

Arg Leu Gln Leu Asp Ala Phe Ala Gln Ser Gly Gly Val Gly Arg Glu
 530 535 540

Val Lys Ser Leu Pro Lys Ile Ala Phe Leu Phe Thr Gly Gln Gly Ser
 545 550 555 560

Gln Tyr Val Gly Met Gly Arg Gln Leu Tyr Glu Asn Gln Pro Thr Phe
 565 570 575

Arg Lys Ala Leu Ala His Cys Asp Asp Ile Leu Arg Ala Gly Ala Tyr
 580 585 590

Phe Asp Arg Ser Leu Leu Ser Ile Leu Tyr Pro Glu Gly Lys Ser Glu
 595 600 605

Ala Ile His Gln Thr Ala Tyr Thr Gln Pro Ala Leu Phe Ala Leu Glu
 610 615 620

Tyr Ala Ile Ala Gln Leu Trp His Ser Trp Gly Ile Lys Pro Asp Ile
 625 630 635 640

Val Met Gly His Ser Val Gly Glu Tyr Val Ala Ala Cys Val Ala Gly
 645 650 655

Ile Phe Ser Leu Glu Asp Gly Leu Lys Leu Ile Ala Thr Arg Gly Arg
 660 665 670

Leu Met Gln Ser Leu Pro Gln Asp Gly Thr Met Val Ser Ser Leu Ala
 675 680 685

Ser Glu Ala Arg Ile Gln Glu Ala Ile Thr Pro Tyr Arg Asp Asp Val
 690 695 700

Ser Ile Ala Ala Ile Asn Gly Thr Glu Ser Val Val Ile Ser Gly Lys
 705 710 715 720

Arg Thr Ser Val Met Ala Ile Ala Glu Gln Leu Ala Thr Val Gly Ile
 725 730 735

Lys Thr Arg Gln Leu Thr Val Ser His Ala Phe His Ser Pro Leu Met
 740 745 750

Thr Pro Ile Leu Asp Glu Phe Arg Gln Val Ala Ala Ser Ile Thr Tyr
 755 760 765

His Gln Pro Lys Leu Leu Leu Val Ser Asn Val Ser Gly Lys Val Ala
 770 775 780

Gly Pro Glu Ile Thr Arg Pro Asp Tyr Trp Val Arg His Val Arg Glu
 785 790 795 800

Ala Val Arg Phe Ala Asp Gly Val Arg Thr Leu Asn Glu Gln Gly Val
 805 810 815

Asn Ile Phe Leu Glu Ile Gly Ser Thr Ala Thr Leu Leu Gly Met Ala
 820 825 830

Leu Arg Val Asn Glu Glu Asp Ser Asn Ala Ser Lys Gly Thr Ser Ser
 835 840 845

Cys Tyr Leu Pro Ser Leu Arg Glu Ser Gln Lys Asp Cys Gln Gln Met
 850 855 860

Phe Thr Ser Leu Gly Glu Leu Tyr Val His Gly Tyr Asp Ile Asp Trp
 865 870 875 880

Gly Ala Phe Asn Arg Gly Tyr Gln Gly Arg Lys Val Ile Leu Pro Thr
 885 890 895

Tyr Pro Phe Gln Arg Gln Arg Tyr Trp Leu Pro Asp Pro Lys Leu Ala
 900 905 910

Gln Ser Ser Asp Leu Asp Thr Phe Gln Ala Gln Ser Ser Ala Ser Ser
 915 920 925

Gln Asn Pro Ser Ala Val Ser Thr Leu Leu Met Glu Tyr Leu Gln Ala
 930 935 940

Gly Asp Val Gln Ser Leu Val Gly Leu Leu Asp Asp Glu Arg Lys Leu
 945 950 955 960

Ser Ala Ala Glu Arg Ile Ala Leu Pro Ser Ile Leu Glu Phe Leu Val
 965 970 975

Glu Glu Gln Gln Arg Gln Ile Ser Ser Thr Thr Thr Pro Gln Thr Val
 980 985 990

Leu Gln Lys Ile Ser Gln Thr Ser His Glu Asp Arg Tyr Glu Ile Leu
 995 1000 1005

Lys Asn Leu Ile Lys Ser Glu Ile Glu Thr Ile Ile Lys Ser Val
 1010 1015 1020

Pro Ser Asp Glu Gln Met Phe Ser Asp Leu Gly Ile Asp Ser Leu
 1025 1030 1035

Met Ala Ile Glu Leu Arg Asn Lys Leu Arg Ser Ala Ile Gly Leu
 1040 1045 1050

Glu Leu Pro Val Ala Ile Val Phe Asp His Pro Thr Ile Lys Gln
 1055 1060 1065

Leu Thr Asn Phe Val Leu Asp Arg Ile Val Pro Gln Ala Asp Gln
 1070 1075 1080

Lys Asp Val Pro Thr Glu Ser Leu Phe Ala Ser Lys Gln Glu Ile
 1085 1090 1095

Ser Val Glu Glu Gln Ser Phe Ala Ile Thr Lys Leu Gly Leu Ser
 1100 1105 1110

Pro Ala Ser His Ser Leu His Leu Pro Pro Trp Thr Val Arg Pro
 1115 1120 1125

Ala Val Met Ala Asp Val Thr Lys Leu Ser Gln Leu Glu Arg Glu
 1130 1135 1140

Ala Tyr Gly Trp Ile Gly Glu Gly Ala Ile Ala Pro Pro His Leu
 1145 1150 1155

Ile Ala Asp Arg Ile Asn Leu Leu Asn Ser Gly Asp Met Pro Trp
 1160 1165 1170

Phe Trp Val Met Glu Arg Ser Gly Glu Leu Gly Ala Trp Gln Val
 1175 1180 1185

Leu Gln Pro Thr Ser Val Asp Pro Tyr Thr Tyr Gly Ser Trp Asp
 1190 1195 1200

Glu Val Thr Asp Gln Gly Lys Leu Gln Ala Thr Phe Asp Pro Ser
 1205 1210 1215

Gly Arg Asn Val Tyr Ile Val Ala Gly Gly Ser Ser Asn Leu Pro
 1220 1225 1230

Thr Val Ala Ser His Leu Met Thr Leu Gln Thr Leu Leu Met Leu
 1235 1240 1245

Arg Glu Thr Gly Arg Asp Thr Ile Phe Val Cys Leu Ala Met Pro
 1250 1255 1260

Gly Tyr Ala Lys Tyr His Ser Gln Thr Gly Lys Ser Pro Glu Glu
 1265 1270 1275

Tyr Ile Ala Leu Thr Asp Glu Asp Gly Ile Pro Met Asp Glu Phe
 1280 1285 1290

Ile Ala Leu Ser Val Tyr Asp Trp Pro Val Thr Pro Ser Phe Arg
 1295 1300 1305

Val Leu Arg Asp Gly Tyr Pro Pro Asp Arg Asp Ser Gly Gly His
 1310 1315 1320

Ala Val Ser Thr Val Phe Gln Leu Asn Asp Phe Asp Gly Ala Ile
 1325 1330 1335

Glu Glu Thr Tyr Arg Arg Ile Ile Arg His Ala Asp Val Leu Gly
 1340 1345 1350

Leu Glu Arg Gly
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<210> 85

<211> 1437

<212> DNA

<213> cylindrospermopsis raciborskii awt205

<400> 85

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<210> 86

<211> 478

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 86

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Thr Met Gln Gly Asn Gly Leu Gly Tyr Ile Ala Asp Gly Ala Ile Ala
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Val Gln Gly Ser Gln Ile Val Ala Val Asp Ser Thr Glu Ala Leu Leu
 35 40 45

Ser His Phe Glu Gly Asn Lys Thr Ile Asn Ala Val Asn Cys Ala Val
 50 55 60

Leu Pro Gly Leu Ile Asp Ala His Ile His Thr Thr Cys Ala Ile Leu
 65 70 75 80

Arg Gly Val Ala Gln Asp Val Thr Asn Trp Leu Met Asp Ala Thr Ile
 85 90 95

Pro Tyr Ala Leu Gln Met Thr Pro Ala Val Asn Ile Ala Gly Thr Arg
 100 105 110

Leu Ser Val Leu Glu Gly Leu Lys Ala Gly Thr Thr Thr Phe Gly Asp
 115 120 125

Ser Glu Thr Pro Tyr Pro Leu Trp Gly Glu Phe Phe Asp Glu Ile Gly
 130 135 140

Val Arg Ala Ile Leu Ser Pro Ala Phe Asn Ala Phe Pro Leu Glu Trp
 145 150 155 160

Ser Ala Trp Lys Glu Gly Asp Leu Tyr Pro Phe Asp Met Lys Ala Gly
 165 170 175

Arg Arg Gly Met Glu Glu Ala Val Asp Phe Ala Cys Ala Trp Asn Gly
 180 185 190

Ala Ala Glu Gly Arg Ile Thr Thr Met Leu Gly Leu Gln Ala Ala Asp
 195 200 205

Met Leu Pro Leu Glu Ile Leu His Ala Ala Lys Glu Ile Ala Gln Arg
 210 215 220

Glu Gly Leu Met Leu His Ile His Val Ala Gln Gly Asp Arg Glu Thr
 225 230 235 240

Lys Gln Ile Val Lys Arg Tyr Gly Lys Arg Pro Ile Ala Phe Leu Ala
 245 250 255

Glu Ile Gly Tyr Leu Asp Glu Gln Leu Leu Ala Val His Leu Thr Asp
 260 265 270

Ala Thr Asp Glu Glu Val Ile Gln Val Ala Lys Ser Gly Ala Gly Met
 275 280 285

Ala Leu Cys Ser Gly Ala Ile Gly Ile Ile Asp Gly Leu Val Pro Pro
 290 295 300

Ala His Val Phe Arg Gln Ala Gly Gly Ser Val Ala Leu Gly Ser Asp
 305 310 315 320

Gln Ala Cys Gly Asn Asn Cys Cys Asn Ile Phe Asn Glu Met Lys Leu
 325 330 335

Thr Ala Leu Phe Asn Lys Ile Lys Tyr His Asp Pro Thr Ile Met Pro
 340 345 350

Ala Trp Glu Val Leu Arg Met Ala Thr Ile Glu Gly Ala Gln Ala Ile
 355 360 365

Gly Leu Asp His Lys Ile Gly Ser Leu Gln Val Gly Lys Glu Ala Asp
 370 375 380

Leu Ile Leu Ile Asp Leu Ser Ser Pro Asn Leu Ser Pro Thr Leu Leu
 385 390 395 400

Asn Pro Ile Arg Asn Leu Val Pro Asn Leu Val Tyr Ala Ala Ser Gly
 405 410 415

His Glu Val Lys Ser Val Met Val Ala Gly Lys Leu Leu Val Glu Asp
 420 425 430

Tyr Gln Val Leu Thr Val Asp Glu Ser Ala Ile Leu Ala Glu Ala Gln
 435 440 445

Val Gln Ala Gln Gln Leu Cys Gln Arg Val Thr Ala Asp Pro Ile His
 450 455 460

Lys Lys Met Val Leu Met Glu Ala Met Ala Lys Gly Lys Leu
 465 470 475

<210> 87

<211> 831

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 87

atgaccatat atgaaaataa gttgagtagt tatcaaaaa atcaagatgc cataatatct 60

gcaaaagaac tcgaagaatg gcatttaatt ggacttctag accattcaat agatgcggtg 120

atagtaccga attattttct tgagcaagag tgtatgacaa tttagagag aataaaaaag 180

agtaaatatt ttagcgctta tcccggtcat ccatcagtaa gtagcttggg acaagagttg 240

tatgaatgcg aaagtgcgct tgaattagca aagtatcaag aagacgcacc cacattgatt 300

aaagaaatgc ggaggctggt acatccgtac ataagtcaa ttgatagact tagggttgaa 360

gttgatgata ttggaggtta tggctgtaat ttagcaaaac ttggtgataa aaaactgttt 420

gcgggtatcg ttagagagtt taaagaagat aaccctggcg caccacattg tgacgtaatg 480

gcatgggggt ttctgaata ttataaagat aaaccaaata tcataaatca aatcgagca 540

aatgtatatt taaaacgtc tgcacagga ggagaaatag tgctttggga tgaatggcca 600

actcaaagcg aatatatagc atacaaaaca gatgatccag ctagtttcgg tcttgatagc 660

aaaaagatcg cacaacaaa acttgagatc caaccgaacc agggagattt aattctattc 720

aattccatga gaattcatgc ggtgaaaaag atagaaactg gtgtacgtat gacatgggga 780

tgtttgattg gatactctgg aactgataaa ccgcttgta tttggactta a 831

<210> 88

<211> 276

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 88

Met Thr Ile Tyr Glu Asn Lys Leu Ser Ser Tyr Gln Lys Asn Gln Asp
1 5 10 15

Ala Ile Ile Ser Ala Lys Glu Leu Glu Glu Trp His Leu Ile Gly Leu
20 25 30

Leu Asp His Ser Ile Asp Ala Val Ile Val Pro Asn Tyr Phe Leu Glu
35 40 45

Gln Glu Cys Met Thr Ile Ser Glu Arg Ile Lys Lys Ser Lys Tyr Phe
50 55 60

Ser Ala Tyr Pro Gly His Pro Ser Val Ser Ser Leu Gly Gln Glu Leu
65 70 75 80

Tyr Glu Cys Glu Ser Glu Leu Glu Leu Ala Lys Tyr Gln Glu Asp Ala
85 90 95

Pro Thr Leu Ile Lys Glu Met Arg Arg Leu Val His Pro Tyr Ile Ser
100 105 110

Pro Ile Asp Arg Leu Arg Val Glu Val Asp Asp Ile Trp Ser Tyr Gly
115 120 125

Cys Asn Leu Ala Lys Leu Gly Asp Lys Lys Leu Phe Ala Gly Ile Val
130 135 140

Arg Glu Phe Lys Glu Asp Asn Pro Gly Ala Pro His Cys Asp Val Met
 145 150 155 160

Ala Trp Gly Phe Leu Glu Tyr Tyr Lys Asp Lys Pro Asn Ile Ile Asn
 165 170 175

Gln Ile Ala Ala Asn Val Tyr Leu Lys Thr Ser Ala Ser Gly Gly Glu
 180 185 190

Ile Val Leu Trp Asp Glu Trp Pro Thr Gln Ser Glu Tyr Ile Ala Tyr
 195 200 205

Lys Thr Asp Asp Pro Ala Ser Phe Gly Leu Asp Ser Lys Lys Ile Ala
 210 215 220

Gln Pro Lys Leu Glu Ile Gln Pro Asn Gln Gly Asp Leu Ile Leu Phe
 225 230 235 240

Asn Ser Met Arg Ile His Ala Val Lys Lys Ile Glu Thr Gly Val Arg
 245 250 255

Met Thr Trp Gly Cys Leu Ile Gly Tyr Ser Gly Thr Asp Lys Pro Leu
 260 265 270

Val Ile Trp Thr
 275

<210> 89

<211> 1398

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 89

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actatcattc tcaactacac tetcaaatgt cctaggtaac tgtgccccaa acatcagcat 120

tccaatggcg ttgaacaaaa agaaagccaa ccacaagata tggttactct caaatTTAAC 180

agcagctaca tccgcaggta aaaatcctac accaaacgcg attaagttaa cattgcggag 240

agtatgccct tgagccaaac ccaagaagta cccacatagt atgcaacata ctgaattgca 300
 tactaggaca agtaccaacc agggaataaa aatatcaata ttctcaataa ttctgcgtg 360
 gttgggtaac aacccaaaaa catcatcggg aaatagccaa cacgctccgc cgaaaaccag 420
 actcactagc agagccattc ccacagaaac tttgccaga ggtgctaact gttctgtggc 480
 tcctttccct ttaaatttc ctgccagagt ttctgtacag aatcccaatc cttcaacaat 540
 gtagatgctc aaagcccata tctgtaagag caaggcattt tgagcgtaga taattgtccc 600
 catttgtgcc ccttcgtagt taaacgttaa gttggtaaac atacaaacta aattgtgac 660
 aaagatgttt ccattgagag ttaaggtgga gcgtatagct ttatgtccc aaattttcc 720
 agctaattct ttacctctt gccacgggat ttcttgcag acaaaaaaca atcccacaa 780
 tagggtgaga tattgacttg cagcagaagc tactctgcc cccatgctcg accagttaa 840
 gtggataata aacaagtagt cgagtgcgat attggcagca ttgccacaa ccgacaacaa 900
 cacaactaag ccatttttt cccgtcccag aaaccagcca agcaggacaa agttgagcaa 960
 aatggcaggc gtcceccaac tctgggtgtt aaaatacgtt tgagctgaag acttcacctc 1020
 tgggccgaca tctagtatag aaaaccccaa caccctaac ggtactgta acagtatgat 1080
 cgccaccccc agcaccagag caattaaacc attaagcagt cccgccaaca gtacgccctc 1140
 tcggtcatct cgtccgactg cttgtgctgt taacgcagtg gtaccattc gtaaaaacga 1200
 taaaacaaag tagagaaagt taagcaggtt tccagcaagg gctactccag ctaggtagt 1260
 gatttccgag agatgaccta agaactgat actgactaaa ttactcagtg gtactataat 1320
 attcgatagg acgttggtta aagctagtcg gaagtagcgg ggtataaagt catactggct 1380
 tggaaatgtc aggctcat 1398

<210> 90

<211> 465

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 90

Met Ser Leu Thr Phe Pro Ser Gln Tyr Asp Phe Ile Pro Arg Tyr Phe

1

5

10

15

Arg Leu Ala Phe Thr Asn Val Leu Ser Asn Ile Ile Val Pro Leu Ser
 20 25 30

Asn Leu Val Ser Ile Met Phe Leu Gly His Leu Ser Glu Ile His Tyr
 35 40 45

Leu Ala Gly Val Ala Leu Ala Gly Asn Leu Leu Asn Phe Leu Tyr Phe
 50 55 60

Val Leu Ser Phe Leu Arg Met Gly Thr Thr Ala Leu Thr Ala Gln Ala
 65 70 75 80

Val Gly Arg Asp Asp Arg Glu Gly Val Leu Leu Ala Gly Leu Leu Asn
 85 90 95

Gly Leu Ile Ala Leu Val Leu Gly Val Ala Ile Ile Leu Leu Gln Tyr
 100 105 110

Pro Leu Gly Val Leu Gly Phe Ser Ile Leu Asp Val Gly Pro Glu Val
 115 120 125

Lys Ser Ser Ala Gln Ala Tyr Phe Asn Thr Gln Ser Trp Gly Ala Pro
 130 135 140

Ala Ile Leu Leu Asn Phe Val Leu Leu Gly Trp Phe Leu Gly Arg Glu
 145 150 155 160

Lys Asn Gly Leu Val Val Leu Leu Ser Val Val Gly Asn Ala Ala Asn
 165 170 175

Ile Ala Leu Asp Tyr Leu Phe Ile Ile His Leu Asp Trp Ser Ser Met
 180 185 190

Gly Ala Gly Val Ala Ser Ala Ala Ser Gln Tyr Leu Thr Leu Leu Val
 195 200 205

Gly Leu Phe Phe Val Cys Lys Glu Ile Pro Trp Gln Glu Val Lys Glu
 210 215 220

Leu Ala Gly Lys Ile Trp Asp Ile Lys Ala Ile Arg Ser Thr Leu Thr
 225 230 235 240

Leu Asn Gly Asn Ile Phe Val Ser Asn Leu Val Cys Met Phe Thr Asn
 245 250 255

Leu Thr Phe Asn Tyr Glu Gly Ala Gln Met Gly Thr Ile Ile Tyr Ala
 260 265 270

Gln Asn Ala Leu Leu Leu Gln Ile Trp Ala Leu Ser Ile Tyr Ile Val
 275 280 285

Glu Gly Leu Gly Phe Cys Thr Glu Thr Leu Ala Gly Asn Phe Lys Gly
 290 295 300

Lys Gly Ala Thr Glu Gln Leu Ala Pro Leu Ala Lys Val Ser Val Gly
 305 310 315 320

Met Ala Leu Leu Val Ser Leu Val Phe Gly Gly Ala Cys Trp Leu Phe
 325 330 335

Pro Asp Asp Val Phe Gly Leu Leu Thr Asn His Ala Glu Ile Ile Glu
 340 345 350

Asn Ile Asp Ile Phe Ile Pro Trp Leu Val Leu Val Leu Val Cys Asn
 355 360 365

Ser Val Cys Cys Ile Leu Cys Gly Tyr Phe Leu Gly Leu Ala Gln Gly
 370 375 380

His Thr Leu Arg Asn Val Asn Leu Ile Ala Phe Gly Val Gly Phe Leu
 385 390 395 400

Pro Ala Asp Val Ala Ala Val Lys Phe Glu Ser Asn His Ile Leu Trp
 405 410 415

Leu Ala Phe Phe Leu Phe Asn Ala Ile Gly Met Leu Met Phe Gly Ala
 420 425 430

Gln Leu Pro Arg Thr Phe Glu Ser Val Val Glu Asn Asp Ser Val Ser
 435 440 445

Ile Pro Ala Leu Glu Ala Ser Arg Ala Leu Thr Gln Met Glu Thr Leu
 450 455 460

His
 465

<210> 91

<211> 750

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 91

atgttgaact tagaccgcat cctgaatcaa gagcgactgc tacgagaaat gactggactt 60

aaccgccaag cattcaacga gctgttatct cagtttgctg atacctatga acgcaccgtg 120

ttcaactcct tagcaaaccg caaacgtgcg cccggggggcg gacgcaagcc tacactcaga 180

agtatagagg aaaaactatt ttatatcctg ctgtactgca aatgttatcc gacgtttgac 240

ttgctgagtg tgttgttcaa ctttgaccgc tcctgtgctc atgattgggt acatcgacta 300

ctgtctgtgc tagaaaccac ttaggagaa aagcaagttt tgccagcacg caaactcagg 360

agcatggagg aattcaccaa aaggtttcca gatgtgaagg aggtgattgt ggatggtagc 420

gagcgtccag tccagcgtcc tcaaaaccga gaacgcaaaa aagagtatta ctctggcaag 480

aaaaagcggc atacatgcaa gcagattaca gtcagcaciaa gggagaaacg agtgattatt 540

cggacggaaa ccagagcagg taaagtgcac gacaaacggc tactccatga atcagagata 600

gtgcaataca ttctgatga agtagcaata gagggagatt tgggttttca tgggttggag 660

aaagaatttg tcaatgtcca ttaccacac aagaaccga aaggtatcga agcaaggagg 720

catggcggcg ggatgggtca gttttataa

750

<210> 92
 <211> 249
 <212> PRT
 <213> *Cylindrospermopsis raciborskii* AWT205

<400> 92

Met Leu Asn Leu Asp Arg Ile Leu Asn Gln Glu Arg Leu Leu Arg Glu
 1 5 10 15

Met Thr Gly Leu Asn Arg Gln Ala Phe Asn Glu Leu Leu Ser Gln Phe
 20 25 30

Ala Asp Thr Tyr Glu Arg Thr Val Phe Asn Ser Leu Ala Asn Arg Lys
 35 40 45

Arg Ala Pro Gly Gly Gly Arg Lys Pro Thr Leu Arg Ser Ile Glu Glu
 50 55 60

Lys Leu Phe Tyr Ile Leu Leu Tyr Cys Lys Cys Tyr Pro Thr Phe Asp
 65 70 75 80

Leu Leu Ser Val Leu Phe Asn Phe Asp Arg Ser Cys Ala His Asp Trp
 85 90 95

Val His Arg Leu Leu Ser Val Leu Glu Thr Thr Leu Gly Glu Lys Gln
 100 105 110

Val Leu Pro Ala Arg Lys Leu Arg Ser Met Glu Glu Phe Thr Lys Arg
 115 120 125

Phe Pro Asp Val Lys Glu Val Ile Val Asp Gly Thr Glu Arg Pro Val
 130 135 140

Gln Arg Pro Gln Asn Arg Glu Arg Gln Lys Glu Tyr Tyr Ser Gly Lys
 145 150 155 160

Lys Lys Arg His Thr Cys Lys Gln Ile Thr Val Ser Thr Arg Glu Lys
 165 170 175

Arg Val Ile Ile Arg Thr Glu Thr Arg Ala Gly Lys Val His Asp Lys
 180 185 190

Arg Leu Leu His Glu Ser Glu Ile Val Gln Tyr Ile Pro Asp Glu Val
 195 200 205

Ala Ile Glu Gly Asp Leu Gly Phe His Gly Leu Glu Lys Glu Phe Val
 210 215 220

Asn Val His Leu Pro His Lys Lys Pro Lys Gly Ile Glu Ala Arg Arg
 225 230 235 240

His Gly Gly Gly Met Gly Gln Phe Leu
 245

<210> 93

<211> 1431

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 93

atgaatctta taacaacaaa aaaacaggta gatacattag tgatacacgc tcattctttt 60

accatgcagg gaaatggtgt gggatatatt gcagatgggg cacttgcggt tgagggtagc 120

cgtatttag cagttgattc gacggaggcg ttgctgagtc atttgaggg cagaaaggtt 180

attgagtccg cgaattgtgc cgtcttgctt gggctgatta atgctcacgt agacacaagt 240

ttggtgctga tgcgtggggc ggcgcaagat gtaactaatt ggctaattga cgcgaccatg 300

ccttattttg ctcacatgac acccgtggcg agtatggctg caacacgctt aagggtggta 360

gaagagtga aagcaggcac aacaacattc tgtgacaata aaattattag cccctgtgg 420

ggcgaatttt tcgatgaaat tgggtgtacgg gctagttag ctctatgtt cgatgcactc 480

ccactggaga tgccaccgt tcaagacggg gagctttatc cttcgatat caaggcggga 540

cggcgggcga tggcagaggc tgtggatttt gcctgtgggt ggaatggggc agcagagggg 600

cgtatcacta ccatgttagg aatgtattcg ccagatatga tgccgcttga gatgctacgc 660

gcagccaaag agattgctca acgggaaggc ttaatgctgc atttcatgt agcgaggga 720
gatcgggaaa cagagcaa atcgtaaacga tatggtaagc gtccgatcg attttagct 780
gagattggct acttggaaga acagttgctg gcagttcacc tcaccgatgc caccgatgaa 840
gaggtgatac aagtagccaa aagtggcgct ggcatggtag tctgttcggg aatgattggc 900
actattgacg gtatcgtgcc gcccgctcat gtgttcggc aagcaggcgg acccgttgcg 960
ctaggcagca gctacaataa tatttccat gagatgaagc tgaccgcctt attcaacaaa 1020
ataaaatata acgatccaac cattatgccg gcttgggaag tcctgcgtat ggctaccatc 1080
gaaggagcgc gggcgattgg ttagatcac aagattggct ctcttgaagt tggcaaagaa 1140
gccgacctga tcttaataga cctcagcacc cctaacctct caccactct gcttaacccc 1200
attcgtaacc ttgtacctaa tttcgtgtac gctgctcag gacatgaagt taaaagtgct 1260
atggtggcgg gaaaactgtt attggaagac taccaagtc tcacagtaga tgagtctgct 1320
atcattgctg aagcacaatt gcaagcccaa cagatttctc aatgcgtagc atctgacct 1380
atccacaaaa aaatggtgct gatggcggcg atggcaagg gccaattgta g 1431

<210> 94

<211> 476

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 94

Met Asn Leu Ile Thr Thr Lys Lys Gln Val Asp Thr Leu Val Ile His
1 5 10 15

Ala His Leu Phe Thr Met Gln Gly Asn Gly Val Gly Tyr Ile Ala Asp
20 25 30

Gly Ala Leu Ala Val Glu Gly Ser Arg Ile Val Ala Val Asp Ser Thr
35 40 45

Glu Ala Leu Leu Ser His Phe Glu Gly Arg Lys Val Ile Glu Ser Ala
50 55 60

Asn Cys Ala Val Leu Pro Gly Leu Ile Asn Ala His Val Asp Thr Ser
 65 70 75 80

Leu Val Leu Met Arg Gly Ala Ala Gln Asp Val Thr Asn Trp Leu Met
 85 90 95

Asp Ala Thr Met Pro Tyr Phe Ala His Met Thr Pro Val Ala Ser Met
 100 105 110

Ala Ala Thr Arg Leu Arg Val Val Glu Glu Leu Lys Ala Gly Thr Thr
 115 120 125

Thr Phe Cys Asp Asn Lys Ile Ile Ser Pro Leu Trp Gly Glu Phe Phe
 130 135 140

Asp Glu Ile Gly Val Arg Ala Ser Leu Ala Pro Met Phe Asp Ala Leu
 145 150 155 160

Pro Leu Glu Met Pro Pro Leu Gln Asp Gly Glu Leu Tyr Pro Phe Asp
 165 170 175

Ile Lys Ala Gly Arg Arg Ala Met Ala Glu Ala Val Asp Phe Ala Cys
 180 185 190

Gly Trp Asn Gly Ala Ala Glu Gly Arg Ile Thr Thr Met Leu Gly Met
 195 200 205

Tyr Ser Pro Asp Met Met Pro Leu Glu Met Leu Arg Ala Ala Lys Glu
 210 215 220

Ile Ala Gln Arg Glu Gly Leu Met Leu His Phe His Val Ala Gln Gly
 225 230 235 240

Asp Arg Glu Thr Glu Gln Ile Val Lys Arg Tyr Gly Lys Arg Pro Ile
 245 250 255

Ala Phe Leu Ala Glu Ile Gly Tyr Leu Asp Glu Gln Leu Leu Ala Val
 260 265 270

His Leu Thr Asp Ala Thr Asp Glu Glu Val Ile Gln Val Ala Lys Ser
 275 280 285

Gly Ala Gly Met Val Leu Cys Ser Gly Met Ile Gly Thr Ile Asp Gly
 290 295 300

Ile Val Pro Pro Ala His Val Phe Arg Gln Ala Gly Gly Pro Val Ala
 305 310 315 320

Leu Gly Ser Ser Tyr Asn Asn Ile Phe His Glu Met Lys Leu Thr Ala
 325 330 335

Leu Phe Asn Lys Ile Lys Tyr His Asp Pro Thr Ile Met Pro Ala Trp
 340 345 350

Glu Val Leu Arg Met Ala Thr Ile Glu Gly Ala Arg Ala Ile Gly Leu
 355 360 365

Asp His Lys Ile Gly Ser Leu Glu Val Gly Lys Glu Ala Asp Leu Ile
 370 375 380

Leu Ile Asp Leu Ser Thr Pro Asn Leu Ser Pro Thr Leu Leu Asn Pro
 385 390 395 400

Ile Arg Asn Leu Val Pro Asn Phe Val Tyr Ala Ala Ser Gly His Glu
 405 410 415

Val Lys Ser Val Met Val Ala Gly Lys Leu Leu Leu Glu Asp Tyr Gln
 420 425 430

Val Leu Thr Val Asp Glu Ser Ala Ile Ile Ala Glu Ala Gln Leu Gln
 435 440 445

Ala Gln Gln Ile Ser Gln Cys Val Ala Ser Asp Pro Ile His Lys Lys
 450 455 460

Met Val Leu Met Ala Ala Met Ala Arg Gly Gln Leu
 465 470 475

<210> 95
 <211> 780
 <212> DNA
 <213> *Cylindrospermopsis raciborskii* AWT205

<400> 95
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 caagcctggt cgagtcggcc agataccgta gtctttgatg aacttctctc ctttcctat 120
 ctctttatca aagggaaaga tatgggcttt acttggacag accttgattc tagccaaatg 180
 cccacgcag attggcgatc cgtcatcgat ctgttaaagg ctcccctgcc tgaagggaaa 240
 tcaatcatcg atctgttaaa ggctcccctg cctgaaggga aatcaatttg ctatcagaag 300
 catcaagcgt atcatttaat cgaagagacc atggggattg agtggatatt gcccttcagc 360
 aactgcttgc tgattcgcca acccaaagaa atgctcttat ctttcgtaa gattgtgcca 420
 cattttacct ttgaagaaac aggctggatc gaattaaaac ggctgtttga ctatgtacat 480
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 agaatgctct ccaagctttg tcagggtgta ggggttgagt ttaccgagac aatgctcagt 600
 tggcccccca tggaggtcga gtgaacgaa aaactagccc cttggtacag caccgtagca 660
 agttctacgc atttcactc gtatcagaat aaaaatgagt cgttgccgct atatctgtc 720
 gatatttgta aacgctgcga tgaaatatat caggaattat atcaatttcg actttattag 780

<210> 96
 <211> 259
 <212> PRT
 <213> *Cylindrospermopsis raciborskii* AWT205

<400> 96

Met Gln Glu Lys Arg Ile Ala Met Trp Ser Val Pro Arg Ser Leu Gly
 1 5 10 15

Thr Val Leu Leu Gln Ala Trp Ser Ser Arg Pro Asp Thr Val Val Phe
 20 25 30

Asp Glu Leu Leu Ser Phe Pro Tyr Leu Phe Ile Lys Gly Lys Asp Met
 35 40 45

Gly Phe Thr Trp Thr Asp Leu Asp Ser Ser Gln Met Pro His Ala Asp
 50 55 60

Trp Arg Ser Val Ile Asp Leu Leu Lys Ala Pro Leu Pro Glu Gly Lys
 65 70 75 80

Ser Ile Ile Asp Leu Leu Lys Ala Pro Leu Pro Glu Gly Lys Ser Ile
 85 90 95

Cys Tyr Gln Lys His Gln Ala Tyr His Leu Ile Glu Glu Thr Met Gly
 100 105 110

Ile Glu Trp Ile Leu Pro Phe Ser Asn Cys Phe Leu Ile Arg Gln Pro
 115 120 125

Lys Glu Met Leu Leu Ser Phe Arg Lys Ile Val Pro His Phe Thr Phe
 130 135 140

Glu Glu Thr Gly Trp Ile Glu Leu Lys Arg Leu Phe Asp Tyr Val His
 145 150 155 160

Gln Thr Ser Gly Val Ile Pro Pro Val Ile Asp Ala His Asp Leu Leu
 165 170 175

Asn Asp Pro Arg Arg Met Leu Ser Lys Leu Cys Gln Val Val Gly Val
 180 185 190

Glu Phe Thr Glu Thr Met Leu Ser Trp Pro Pro Met Glu Val Glu Leu
 195 200 205

Asn Glu Lys Leu Ala Pro Trp Tyr Ser Thr Val Ala Ser Ser Thr His
 210 215 220

Phe His Ser Tyr Gln Asn Lys Asn Glu Ser Leu Pro Leu Tyr Leu Val
 225 230 235 240

Asp Ile Cys Lys Arg Cys Asp Glu Ile Tyr Gln Glu Leu Tyr Gln Phe
 245 250 255

Arg Leu Tyr

<210> 97

<211> 1176

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 97

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 gataagaaca ttgcaaaat gttctctttt ccagggggtc cgaaaaagca agaggtaaca 180
 gagaaagcta atgaggagtt gaatgggctg gtagcgcttc tagaatcaca gggcgtaact 240
 gtacgccgcc cagagaaaca taactttggc ctgtctgtga agacaccatt ctttgagga 300
 gagaatcaat attgtgcggt ctgccacgt gatgttatga tcaccttgg gaacgaaatt 360
 ctggaagcaa ctatgtcacg gcggtcacgc ttctttgagt attacccta tcgcaaacta 420
 gtctatgaat attggcataa agatccagat atgatctgga atgctgcgcc taaaccgact 480
 atgcaaaatg ccatgtaccg cgaagatttc tgggagtgtc cgatggaaga tcgatttgag 540
 agtatgcatg attttgagtt ctgcgtcacc caggatgagg tgattttga cgcagcagac 600
 ttagccgct ttggccgtga tttttgtg caggagtcaa tgacgactaa tcgtgcaggg 660
 attcgctggc tcaaacggca ttagagccg cgtcgcttcc gcgtgcatga tattcattc 720
 ccactagata tttcccatc ccacattgat tgtacttttg tccccttagc acctgggggt 780
 gtgttagtga atccagatcg ccccatcaaa gaggggtgaag agaaactctt catggataac 840
 gggtggcaat tcacgaagc accctcccc attccaccg acgatgagat gcctatgttc 900
 tgccagtcca gtaagtgggt ggcatgaat gtgtaagca tttccccaa gaaggtcatc 960

tgtgaagagc aagagcatcc gcttcatgag ttgctagata aacacggctt tgaggtctat 1020

ccaattccct ttcgcaatgt ctttgagttt ggcggttcgc tccattgtgc cacctgggat 1080

atccatcgca cgggaacctg tgaggattac ttcctaaac taaactatac gccggtaact 1140

gcaccaacca atggcgtttc tcgcttcac atttag 1176

<210> 98

<211> 391

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 98

Met Gln Thr Arg Ile Val Asn Ser Trp Asn Glu Trp Asp Glu Leu Lys
1 5 10 15

Glu Met Val Val Gly Ile Ala Asp Gly Ala Tyr Phe Glu Pro Thr Glu
20 25 30

Pro Gly Asn Arg Pro Ala Leu Arg Asp Lys Asn Ile Ala Lys Met Phe
35 40 45

Ser Phe Pro Arg Gly Pro Lys Lys Gln Glu Val Thr Glu Lys Ala Asn
50 55 60

Glu Glu Leu Asn Gly Leu Val Ala Leu Leu Glu Ser Gln Gly Val Thr
65 70 75 80

Val Arg Arg Pro Glu Lys His Asn Phe Gly Leu Ser Val Lys Thr Pro
85 90 95

Phe Phe Glu Val Glu Asn Gln Tyr Cys Ala Val Cys Pro Arg Asp Val
100 105 110

Met Ile Thr Phe Gly Asn Glu Ile Leu Glu Ala Thr Met Ser Arg Arg
115 120 125

Ser Arg Phe Phe Glu Tyr Leu Pro Tyr Arg Lys Leu Val Tyr Glu Tyr
130 135 140

Trp His Lys Asp Pro Asp Met Ile Trp Asn Ala Ala Pro Lys Pro Thr
 145 150 155 160

Met Gln Asn Ala Met Tyr Arg Glu Asp Phe Trp Glu Cys Pro Met Glu
 165 170 175

Asp Arg Phe Glu Ser Met His Asp Phe Glu Phe Cys Val Thr Gln Asp
 180 185 190

Glu Val Ile Phe Asp Ala Ala Asp Cys Ser Arg Phe Gly Arg Asp Ile
 195 200 205

Phe Val Gln Glu Ser Met Thr Thr Asn Arg Ala Gly Ile Arg Trp Leu
 210 215 220

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Leu Pro Thr Ser Thr Asp Asp Glu Met Pro Met Phe Cys Gln Ser Ser
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Lys Trp Leu Ala Met Asn Val Leu Ser Ile Ser Pro Lys Lys Val Ile
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Cys Glu Glu Gln Glu His Pro Leu His Glu Leu Leu Asp Lys His Gly
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Ser Leu His Cys Ala Thr Trp Asp Ile His Arg Thr Gly Thr Cys Glu
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Glu Tyr Met Leu Arg Asp Ser Asp Ala Arg Ile Leu Leu Thr Ser Thr
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His Asn Leu Pro Asp Leu Thr Asn Glu Lys Phe Ile Pro Asn Pro Phe
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Gly Ala Asn Glu Asn Ala Lys Lys Leu Tyr Arg Thr Gly Asp Leu Ala
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Glu Ile Ala Gly His Thr Gln Leu Val Gly Tyr Ile Ile Ala Lys Asp
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Thr Leu Asn Leu Ser Phe Asp Lys Leu Glu Pro Ile Leu Arg Gln Tyr
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Gln Thr Ala Cys Ser Thr Ser Leu Val Ala Val His Met Ala Cys Gln
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Leu Asp Met Gly Pro Met Met Arg Gly Val Lys Gln Leu Trp Arg
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Lys Val Ser Leu Pro Glu Ile Leu Leu His Glu Ser Asn Ala Tyr
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Gln Phe His Pro Val Ile Leu Asp Ala Gly Leu Gln Met Ile Thr
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 1850 1855 1860

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 1865 1870 1875

Gln Ser Val Leu Val Gly Arg Glu Ala Met Leu Arg Ser Gln Asp
 1880 1885 1890

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Cys Phe Gly Leu Leu Pro Asn Tyr Leu Pro Thr Pro Asp Lys Ile
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Leu Arg Gly Val Ile His Ala Ala Gly Thr Leu Asn Asp Gly Ile
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<213> *Cylindrospermopsis raciborskii* AWT205

<400> 101

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<213> *Cylindrospermopsis raciborskii* AWT205

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Gln	Cys	Glu	Pro	Ile	Ala	Ile	Val	Gly	Met	Gly	Cys	Arg	Phe	Pro	Gly
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Pro Arg Glu Ala Val Ser Leu Asp Pro Gln Gln Arg Leu Leu Leu Glu
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Ser Thr Ala Ser Gly Arg Leu Ser Tyr Phe Leu Gly Leu Thr Gly Pro
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Cys Leu Ser Ile Asp Thr Ala Cys Ser Ser Ser Leu Val Ala Val His
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Gly Gly Val His Arg Leu Ile Ala Pro Glu Glu Ser Val Ser Leu Ala
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Lys Arg Leu Ser Asp Ala Gln Ala Asp Gly Asp Lys Ile Leu Ala Leu
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Ile Arg Gly Ser Ala Ile Asn Gln Asp Gly Arg Thr Ser Gly Leu Thr
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Asn Ser Gly Ile Arg Pro Glu Gln Val Asn Tyr Val Glu Ala His Gly
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Thr Gly Thr Ser Leu Gly Asp Pro Ile Glu Val Gly Ala Leu Gly Thr
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Ile Phe Asn Gln Arg Ser Gln Pro Leu Ile Ile Gly Ser Val Lys Thr
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Val Val Leu Ala Met Gln His Gly Glu Ile Pro Pro Asn Leu His Phe
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His Gln Pro Asn Pro Arg Ile Asn Trp Asp Lys Leu Pro Ile Arg Ile
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Pro Thr Glu Arg Thr Ala Trp Pro Thr Gly Asp Arg Ile Ala Gly Ile
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Ser Ser Phe Gly Phe Ser Gly Thr Asn Ser His Val Val Leu Glu Glu
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Gln Arg Tyr Val Thr Tyr Leu Thr Glu His Leu Gln Glu Ser Leu Ala
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755 760 765

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785 790 795 800

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Gln Ser Ser Glu Asn Pro Phe Ser Met Pro Met Met Met Pro Ser Leu
835 840 845

Arg Gln Asn Arg Ser Asp Trp Gln Gln Met Leu Glu Ser Leu Ser Gln
 850 855 860

Leu Tyr Val His Gly Val Glu Ile Asp Trp Ile Gly Phe Asn Lys Asp
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Tyr Val Arg His Lys Val Val Leu Pro Thr Tyr Pro Trp Gln Lys Glu
 885 890 895

Arg Tyr Trp Val Glu Leu Asp Gln Gln Lys His Ala Ala Lys Asn Leu
 900 905 910

His Pro Leu Leu Asp Arg Cys Met Lys Leu Pro Arg His Asn Glu Thr
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Ile Phe Glu Lys Glu Phe Ser Leu Glu Thr Leu Pro Phe Leu Ala Asp
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Tyr Arg Ile Tyr Gly Ser Val Val Ser Pro Gly Ala Ser Tyr Leu Ser
 945 950 955 960

Met Ile Leu Ser Ile Ala Glu Ser Tyr Ala Asn Gly His Leu Asn Gly
 965 970 975

Gly Asn Ser Ala Lys Gln Thr Thr Tyr Leu Leu Lys Asp Val Thr Phe
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Pro Val Pro Leu Val Ile Ser Asp Glu Ala Asn Tyr Met Val Gln Val
 995 1000 1005

Ala Cys Ser Leu Ser Cys Ala Ala Pro His Asn Arg Gly Asp Glu
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Thr Gln Phe Glu Leu Phe Ser Phe Ala Glu Asn Val Pro Glu Ser
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Ser Ser Ile Asn Ala Asp Phe Gln Thr Pro Ile Ile His Ala Lys
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Gly Gln Phe Lys Leu Glu Asp Thr Ala Pro Pro Lys Val Glu Leu
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Glu Glu Leu Gln Ala Gly Cys Pro Gln Glu Ile Asp Leu Asn Leu
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Phe Tyr Gln Thr Phe Thr Asp Lys Gly Phe Val Phe Gly Ser Arg
 1085 1090 1095

Phe Arg Trp Leu Glu Gln Ile Trp Val Gly Asp Gly Glu Ala Leu
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Ala Arg Leu Arg Gln Pro Glu Ser Ile Glu Ser Phe Lys Gly Tyr
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Val Ile His Pro Gly Leu Leu Asp Ala Cys Thr Gln Val Pro Phe
 1130 1135 1140

Ala Ile Ser Ser Asp Asp Glu Asn Arg Gln Ser Glu Thr Thr Met
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Pro Phe Ala Leu Asn Glu Leu Arg Cys Tyr Gln Pro Ala Asn Gly
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Gln Met Trp Trp Val His Ala Thr Glu Lys Asp Arg Tyr Thr Trp
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Asp Val Ser Leu Phe Asp Glu Ser Gly Gln Val Ile Ala Glu Phe
 1190 1195 1200

Ile Gly Leu Glu Val Arg Ala Ala Met Pro Glu Gly Leu Leu Arg
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Ala Asp Phe Trp His Asn Trp Leu Tyr Thr Val Asn Trp Arg Ser
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Gln Pro Leu Gln Ile Pro Glu Val Leu Asp Ile Asn Lys Thr Gly
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Ala Glu Thr Trp Leu Leu Phe Ala Gln Pro Glu Gly Ile Gly Ala
 1250 1255 1260

Asp Leu Ala Glu Tyr Leu Gln Ser Gln Gly Lys His Cys Val Phe
 1265 1270 1275

Val Val Pro Gly Ser Glu Tyr Thr Val Thr Glu Gln His Ile Gly
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Arg Thr Gly His Leu Asp Val Thr Lys Leu Thr Lys Ile Val Thr
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Ile Asn Pro Ala Ser Pro His Asp Tyr Lys Tyr Phe Leu Glu Thr
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Leu Thr Asp Ile Arg Leu Pro Cys Glu His Ile Leu Tyr Leu Trp
 1325 1330 1335

Asn Arg Tyr Asp Leu Thr Asn Thr Ser Asn His Arg Thr Glu Leu
 1340 1345 1350

Thr Val Pro Asp Ile Val Leu Asn Leu Cys Thr Ser Leu Thr Tyr
 1355 1360 1365

Leu Val Gln Ala Leu Ser His Met Gly Phe Ser Pro Lys Leu Trp
 1370 1375 1380

Leu Ile Thr Gln Asn Ser Gln Ala Val Gly Ser Asp Leu Ala Asn
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Leu Glu Ile Glu Gln Ser Pro Leu Trp Ala Leu Gly Arg Ser Ile
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Arg Ala Glu His Pro Glu Phe Asp Cys Arg Cys Leu Asp Phe Asp
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Thr Leu Ser Asn Ile Ala Pro Leu Leu Leu Lys Glu Met Gln Ala
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Ile Asp Tyr Glu Ser Gln Ile Ala Tyr Arg Gln Gly Thr Arg Tyr
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Val Ala Arg Leu Ile Arg Asn Gln Ser Glu Cys His Ala Pro Ile
 1460 1465 1470

Gln Thr Gly Ile Arg Pro Asp Gly Ser Tyr Leu Ile Thr Gly Gly
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Leu Gly Gly Leu Gly Leu Gln Val Ala Leu Ala Leu Ala Asp Ala
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Gly Ala Arg His Leu Ile Leu Asn Ser Arg Arg Gly Thr Val Ser
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Lys Glu Ala Gln Leu Ile Ile Asp Arg Leu Arg Gln Glu Asp Val
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Arg Val Asp Leu Ile Ala Ala Asp Val Ser Asp Ala Ala Asp Ser
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Glu Arg Leu Leu Val Glu Ser Gln Arg Lys Thr Ser Leu Arg Gly
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Gln Asn Gln Glu Arg Phe Glu Lys Val Met Ala Ala Lys Val Arg
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Gly Ala Trp His Leu Asp Gln Gln Ser Gln Thr Leu Asp Leu Asp
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Phe Phe Val Ala Phe Ser Ser Val Ala Ser Leu Ile Glu Glu Pro
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Gly Gln Ala Asn Tyr Ala Ala Ala Asn Ala Phe Leu Asp Ser Leu
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Met Tyr Tyr Arg His Ile Lys Gly Ser Asn Ser Leu Ser Ile Asn
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Trp Gly Ala Trp Ala Glu Val Gly Met Ala Ala Asn Leu Ser Trp
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Glu Gln Arg Gly Ile Ala Ala Ile Ser Pro Lys Gln Gly Arg His
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Ile Leu Val Gln Leu Ile Gln Lys Leu Asn Gln His Thr Ile Pro
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Gln Val Ala Val Gln Pro Thr Asn Trp Ala Glu Tyr Leu Ser His
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Asp Gly Val Asn Met Pro Phe Tyr Glu Tyr Phe Thr His His Leu
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Arg Asn Glu Lys Glu Ala Lys Leu Arg Gln Thr Ala Gly Ser Thr
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Ser Glu Glu Val Ser Leu Arg Gln Gln Leu Gln Thr Leu Ser Glu
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Lys Asp Arg Asp Ala Leu Leu Met Glu His Leu Gln Lys Thr Ala
 1760 1765 1770

Ile Arg Val Leu Gly Leu Ala Ser Asn Gln Lys Ile Asp Pro Tyr
 1775 1780 1785

Gln Gly Leu Met Asn Met Gly Leu Asp Ser Leu Met Ala Val Glu
 1790 1795 1800

Phe Arg Asn His Leu Ile Arg Ser Leu Glu Arg Pro Leu Pro Ala
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Thr Leu Leu Phe Asn Cys Pro Thr Leu Asp Ser Leu His Asp Tyr
 1820 1825 1830

Leu Val Ala Lys Met Phe Asp Asp Ala Pro Gln Lys Ala Glu Gln
 1835 1840 1845

Met Ala Gln Pro Thr Thr Leu Thr Ala His Ser Ile Ser Ile Glu
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Ser Lys Ile Asp Asp Asn Glu Ser Val Asp Asp Ile Ala Gln Met
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Leu Ala Gln Ala Leu Asn Ile Ala Phe Glu
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<210> 103

<211> 5004

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 103

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<211> 1667

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 104

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Glu Ser Ile Ala Ile Val Gly Met Ser Cys Arg Phe Pro Gly Gly Ala
 35 40 45

Thr Thr Pro Glu Arg Phe Trp Val Leu Leu Arg Glu Gly Ile Ser Ala
 50 55 60

Ile Thr Glu Ile Pro Ala Asp Arg Trp Asp Val Asp Lys Tyr Tyr Asp
 65 70 75 80

Ala Asp Pro Thr Ser Ser Gly Lys Met His Thr Arg Tyr Gly Gly Phe
 85 90 95

Leu Asn Glu Val Asp Thr Phe Glu Pro Ser Phe Phe Asn Ile Ala Ala
 100 105 110

Arg Glu Ala Val Ser Met Asp Pro Gln Gln Arg Leu Leu Leu Glu Val
 115 120 125

Ser Trp Glu Ala Leu Glu Ser Gly Asn Ile Val Pro Ala Thr Leu Phe
 130 135 140

Asp Ser Ser Thr Gly Val Phe Ile Gly Ile Gly Gly Ser Asn Tyr Lys
 145 150 155 160

Ser Leu Met Ile Glu Asn Arg Ser Arg Ile Gly Lys Thr Asp Leu Tyr
 165 170 175

Glu Leu Ser Gly Thr Asp Val Ser Val Ala Ala Gly Arg Ile Ser Tyr
 180 185 190

Val Leu Gly Leu Met Gly Pro Ser Phe Val Ile Asp Thr Ala Cys Ser
 195 200 205

Ser Ser Leu Val Ser Val His Gln Ala Cys Gln Ser Leu Arg Gln Arg
 210 215 220

Glu Cys Asp Leu Ala Leu Ala Gly Gly Val Gly Leu Leu Ile Asp Pro
 225 230 235 240

Asp Glu Met Ile Gly Leu Ser Gln Gly Gly Met Leu Ala Pro Asp Gly
 245 250 255

Ser Cys Lys Thr Phe Asp Ala Asn Ala Asn Gly Tyr Val Arg Gly Glu
 260 265 270

Gly Cys Gly Met Ile Val Leu Lys Arg Leu Ser Asp Ala Thr Ala Asp
 275 280 285

Gly Asp Asn Ile Leu Ala Ile Ile Arg Gly Ser Met Val Asn His Asp
 290 295 300

Gly His Ser Ser Gly Leu Thr Ala Pro Arg Gly Pro Ala Gln Val Ser
 305 310 315 320

Val Ile Lys Gln Ala Leu Asp Arg Ala Gly Ile Ala Pro Asp Ala Val
 325 330 335

Ser Tyr Leu Glu Ala His Gly Thr Gly Thr Pro Leu Gly Asp Pro Ile
 340 345 350

Glu Met Asp Ser Leu Asn Glu Val Phe Gly Arg Arg Thr Glu Pro Leu
 355 360 365

Trp Val Gly Ser Val Lys Thr Asn Ile Gly His Leu Glu Ala Ala Ser
 370 375 380

Gly Ile Ala Gly Leu Ile Lys Val Val Leu Met Leu Lys Asn Lys Gln
 385 390 395 400

Ile Pro Pro His Leu His Phe Lys Thr Pro Asn Pro Tyr Ile Asp Trp
 405 410 415

Lys Asn Leu Pro Val Glu Ile Pro Thr Thr Leu His Ala Trp Asp Asp
 420 425 430

Lys Thr Leu Lys Asp Arg Lys Arg Ile Ala Gly Val Ser Ser Phe Ser
 435 440 445

Phe Ser Gly Thr Asn Ala His Ile Val Leu Ser Glu Ala Pro Ser Ser
 450 455 460

Glu Leu Ile Ser Asn His Ala Ala Val Glu Arg Pro Trp His Leu Leu
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Thr Leu Ser Ala Lys Asn Glu Glu Ala Leu Ala Asn Leu Val Gly Leu
 485 490 495

Tyr Gln Ser Phe Ile Ser Thr Thr Asp Ala Ser Leu Ala Asp Ile Cys
 500 505 510

Tyr Thr Ala Asn Thr Ala Arg Thr His Phe Ser His Arg Leu Ala Leu
 515 520 525

Ser Ala Thr Ser His Ile Gln Ile Glu Ala Leu Leu Ala Ala Tyr Lys
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Glu Gly Ser Val Ser Leu Ser Ile Asn Gln Gly Cys Val Leu Ser Asn
 545 550 555 560

Ser Arg Ala Pro Lys Val Ala Phe Leu Phe Thr Gly Gln Gly Ser Gln
 565 570 575

Tyr Val Gln Met Ala Gly Glu Leu Tyr Glu Thr Gln Pro Thr Phe Arg
 580 585 590

Asn Cys Leu Asp Arg Cys Ala Glu Ile Leu Gln Ser Ile Phe Ser Ser
 595 600 605

Arg Asn Ser Pro Trp Gly Asn Pro Leu Leu Ser Val Leu Tyr Pro Asn
 610 615 620

His Glu Ser Lys Glu Ile Asp Gln Thr Ala Tyr Thr Gln Pro Ala Leu
 625 630 635 640

Phe Ala Val Glu Tyr Ala Leu Ala Gln Met Trp Arg Ser Trp Gly Ile
 645 650 655

Glu Pro Asp Ile Val Met Gly His Ser Ile Gly Glu Tyr Val Ala Ala
 660 665 670

Cys Val Ala Gly Ile Phe Ser Leu Glu Asp Gly Leu Lys Leu Ala Ala
 675 680 685

Glu Arg Gly Arg Leu Met Gln Ala Leu Pro Gln Asn Gly Glu Met Val
 690 695 700

Ala Ile Ser Ala Ser Leu Glu Glu Val Lys Pro Ala Ile Gln Ser Asp
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Gln Arg Val Val Ile Ala Ala Val Asn Gly Pro Arg Ser Val Val Ile
 725 730 735

Ser Gly Asp Arg Gln Ala Val Gln Val Phe Thr Asn Thr Leu Glu Asp
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Gln Gly Ile Arg Cys Lys Arg Leu Ser Val Ser His Ala Phe His Ser
 755 760 765

Pro Leu Met Lys Pro Met Glu Gln Glu Phe Ala Gln Val Ala Arg Glu
 770 775 780

Ile Asn Tyr Ser Pro Pro Lys Ile Ala Leu Val Ser Asn Leu Thr Gly
 785 790 795 800

Asp Leu Ile Ser Pro Glu Ser Ser Leu Glu Glu Gly Val Ile Ala Ser
 805 810 815

Pro Gly Tyr Trp Val Asn His Leu Cys Asn Pro Val Leu Phe Ala Asp
 820 825 830

Gly Ile Ala Thr Met Gln Ala Gln Asp Val Gln Val Phe Leu Glu Val
 835 840 845

Gly Pro Lys Pro Thr Leu Ser Gly Leu Val Gln Gln Tyr Phe Asp Glu
 850 855 860

Val Ala His Ser Asp Arg Pro Val Thr Ile Pro Thr Leu Arg Pro Lys
 865 870 875 880

Gln Pro Asn Trp Gln Thr Leu Leu Glu Ser Leu Gly Gln Leu Tyr Ala
 885 890 895

Leu Gly Val Gln Val Asn Trp Ala Gly Phe Asp Arg Asp Tyr Thr Arg
 900 905 910

Arg Lys Val Ser Leu Pro Thr Tyr Ala Trp Lys Arg Gln Arg Tyr Trp
 915 920 925

Leu Glu Lys Gln Ser Ala Pro Arg Leu Glu Thr Thr Gln Val Arg Pro
 930 935 940

Ala Thr Ala Ile Val Glu His Leu Glu Gln Gly Asn Val Pro Lys Ile
 945 950 955 960

Val Asp Leu Leu Ala Ala Thr Asp Val Leu Ser Gly Glu Ala Arg Lys
 965 970 975

Leu Leu Pro Ser Ile Ile Glu Leu Leu Val Ala Lys His Arg Glu Glu
 980 985 990

Ala Thr Gln Lys Pro Ile Cys Asp Trp Leu Tyr Glu Val Val Trp Gln
 995 1000 1005

Pro Gln Leu Leu Thr Leu Ser Thr Leu Pro Ala Val Glu Thr Glu
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Gly Arg Gln Trp Leu Ile Phe Ala Asp Ala Ser Gly His Gly Glu
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Ala Leu Ala Ala Gln Leu Arg Gln Gln Gly Asp Ile Ile Thr Leu
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Val Tyr Ala Gly Leu Lys Tyr His Ser Ala Asn Asn Lys Gln Asn
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Thr Gly Gly Asp Ile Pro Tyr Phe Gln Ile Asp Pro Ile Gln Arg
 1070 1075 1080

Glu Asp Tyr Glu Arg Leu Phe Ala Ala Leu Pro Pro Leu Tyr Gly
 1085 1090 1095

Ile Val His Leu Trp Ser Leu Asp Ile Leu Ser Leu Asp Lys Val
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Ser Asn Leu Ile Glu Asn Val Gln Leu Gly Ser Gly Thr Leu Leu
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Asn Leu Ile Gln Thr Val Leu Gln Leu Glu Thr Pro Thr Pro Ser
 1130 1135 1140

Leu Trp Leu Val Thr Lys Asn Ala Gln Ala Val Arg Lys Asn Asp
 1145 1150 1155

Ser Leu Val Gly Val Leu Gln Ser Pro Leu Trp Gly Met Gly Lys
 1160 1165 1170

Val Ile Ala Leu Glu His Pro Glu Leu Asn Cys Val Ser Ile Asp
 1175 1180 1185

Leu Asp Gly Glu Gly Leu Pro Asp Glu Gln Ala Lys Phe Leu Ala
 1190 1195 1200

Ala Glu Leu Arg Ala Ala Ser Glu Phe Arg His Thr Thr Ile Pro
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His Glu Ser Gln Val Ala Trp Arg Asn Arg Thr Arg Tyr Val Ser
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Arg Phe Lys Gly Tyr Gln Lys His Pro Ala Thr Ser Ser Lys Met
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Pro Ile Arg Pro Asp Ala Thr Tyr Leu Ile Thr Gly Gly Phe Gly
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Gly Leu Gly Leu Leu Val Ala Arg Trp Met Val Glu Gln Gly Ala
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Gln Lys Gln Leu Gln Glu Ile Ala Ala Leu Gly Ala Thr Val Thr
 1295 1300 1305

Val Val Gln Ala Asp Val Gly Ile Arg Ser Gln Val Ala Asn Val
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Leu Ala Gln Ile Asp Lys Ala Tyr Pro Leu Ala Gly Ile Ile His
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Thr Ala Gly Val Leu Asp Asp Gly Ile Leu Leu Gln Gln Asn Trp
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Ala Arg Phe Ser Lys Val Phe Ala Pro Lys Leu Glu Gly Ala Trp
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His Leu His Thr Leu Thr Glu Glu Met Pro Leu Asp Phe Phe Ile
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Cys Phe Ser Ser Thr Ala Gly Leu Leu Gly Ser Gly Gly Gln Ala
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Asn Tyr Ala Ala Ala Asn Ala Phe Leu Asp Ala Phe Ala His His
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Arg Arg Ile Gln Gly Leu Pro Ala Leu Ser Ile Asn Trp Asp Ala
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Trp Ser Gln Val Gly Met Thr Val Arg Leu Gln Gln Ala Ser Ser
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Gln Ser Thr Thr Val Gly Gln Asp Ile Ser Thr Leu Glu Ile Ser
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Pro Glu Gln Gly Leu Gln Ile Phe Ala Tyr Leu Leu Gln Gln Pro
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Ser Ala Gln Ile Ala Ala Ile Ser Thr Asp Gly Leu Arg Lys Met
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Tyr Asp Thr Ser Ser Ala Phe Phe Ala Leu Leu Asp Leu Asp Arg
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Gly Leu Thr Leu Leu Glu Gln Leu Gln Gln Ala Arg Pro Lys Glu
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Arg Glu Lys Met Leu Leu Arg His Leu Gln Thr Gln Val Ala Ala
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Val Leu Arg Ser Pro Glu Leu Pro Ala Val His Gln Pro Phe Thr
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Asp Leu Gly Met Asp Ser Leu Met Ser Leu Glu Leu Met Arg Arg
 1565 1570 1575

Leu Glu Glu Ser Leu Gly Ile Gln Met Pro Ala Thr Leu Ala Phe
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Asp Tyr Pro Met Val Asp Arg Leu Ala Lys Phe Ile Leu Thr Gln
 1595 1600 1605

Ile Cys Ile Asn Ser Glu Pro Asp Thr Ser Ala Val Leu Thr Pro
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Asp Gly Asn Gly Glu Glu Lys Asp Ser Asn Lys Asp Arg Ser Thr
 1625 1630 1635

Ser Thr Ser Val Asp Ser Asn Ile Thr Ser Met Ala Glu Asp Leu
 1640 1645 1650

Phe Ala Leu Glu Ser Leu Leu Asn Lys Ile Lys Arg Asp Gln
 1655 1660 1665

<210> 105

<211> 318

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 105

ttatgctgca tctaaataga agttccatag ccctgcactg accaacaatca attgatcatc 60

aaaatcggtc acacgattcc tatatgtggg ataaaatttg cagtacagca ggatataaaa 120

tagtttttcc tctatacttc tgagtgtagg cttgcgtccg cccccgggcg cacgtttgcg 180

gtttgctaag gagttgaaca cggtcggttc ataggtatca gcaaactgag ataacagctc 240

gttgaatgct tggcgggtaa gtccagtcac tgctcgtagc agtcgctctt gattcaggat 300

gcggtctaag ttcaacat 318

<210> 106

<211> 105

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 106

Met Leu Asn Leu Asp Arg Ile Leu Asn Gln Glu Arg Leu Leu Arg Ala
1 5 10 15

Met Thr Gly Leu Asn Arg Gln Ala Phe Asn Glu Leu Leu Ser Gln Phe
20 25 30

Ala Asp Thr Tyr Glu Arg Thr Val Phe Asn Ser Leu Ala Asn Arg Lys
35 40 45

Arg Ala Pro Gly Gly Gly Arg Lys Pro Thr Leu Arg Ser Ile Glu Glu
50 55 60

Lys Leu Phe Tyr Ile Leu Leu Tyr Cys Lys Phe Tyr Pro Thr Tyr Arg
65 70 75 80

Asn Arg Val Thr Asp Phe Asp Asp Gln Leu Met Leu Val Ser Ala Gly
85 90 95

Leu Trp Asn Phe Tyr Leu Asp Ala Ala
100 105

<210> 107

<211> 600

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 107

ctactgagtg aaagtgaact tctttccac gtattcgagt agctgttgta agctggcctc 60

gatggaaagt tccgaagttt ccaccagtaa atctggtggt ctcggtggtt cgtagggagc 120
 gctaattccc gtaaaagact caatttctcc acggcgtgct ttgcataga gacccttggg 180
 gtcacgttgt tcacaaattt ccatcggagt tgcaatatat acttcatgaa acagatctcc 240
 ggacagaata cggatttgct cccggtcttt cctgtaaggt gaaatgaaag cagtaatcac 300
 taaacaaccc gaatccgcaa aaagtttggc cacctcgcca atacgacgaa tattttccgc 360
 acgatcagca gcagaaaatc ccaagtcagc acataatcca tgacggatat tgtcaccatc 420
 aaggacaaaa gtataccaac ctttctggaa caaatccgc tctaattcta gagccaatgt 480
 tgttttacct gatcctgata atccagtga ccatagaatt ccatttcggt gaccattctt 540
 taaacaacga tcaaatgggg acacaagatg tttgtatgt tgaatattgc ttgattcat 600

<210> 108

<211> 199

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 108

Met Lys Ser Ser Asn Ile Gln His Thr Lys His Leu Val Ser Pro Phe
 1 5 10 15

Asp Arg Cys Leu Lys Asn Gly His Arg Asn Gly Ile Leu Trp Phe Thr
 20 25 30

Gly Leu Ser Gly Ser Gly Lys Thr Thr Leu Ala Leu Glu Leu Glu Arg
 35 40 45

Ile Leu Phe Gln Lys Gly Trp Tyr Thr Phe Val Leu Asp Gly Asp Asn
 50 55 60

Ile Arg His Gly Leu Cys Ala Asp Leu Gly Phe Ser Ala Ala Asp Arg
 65 70 75 80

Ala Glu Asn Ile Arg Arg Ile Gly Glu Val Ala Lys Leu Phe Ala Asp
 85 90 95

Ser Gly Cys Leu Val Ile Thr Ala Phe Ile Ser Pro Tyr Arg Lys Asp
 100 105 110

Arg Glu Gln Ile Arg Ile Leu Ser Gly Asp Leu Phe His Glu Val Tyr
 115 120 125

Ile Ala Thr Pro Met Glu Ile Cys Glu Gln Arg Asp Pro Lys Gly Leu
 130 135 140

Tyr Ala Lys Ala Arg Arg Gly Glu Ile Glu Ser Phe Thr Gly Ile Ser
 145 150 155 160

Ala Pro Tyr Glu Pro Pro Arg Thr Pro Asp Leu Leu Val Glu Thr Ser
 165 170 175

Glu Leu Ser Ile Glu Ala Ser Leu Gln Gln Leu Leu Glu Tyr Val Gly
 180 185 190

Lys Lys Phe Thr Phe Thr Gln
 195

<210> 109

<211> 1548

<212> DNA

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 109

atgcctaaat actttaatac tgctggaccc tgtaaaccg aaatccacta tatgctctct 60

cccacagctc gactaccgga ttgaaagca ctaattgacg gagaaaacta ctttataatt 120

cacgcgccgc gacaagtcgg caaaactaca gctatgatag ccttagcacg agaattgact 180

gatagtggaa aatataccgc agttattctt tccgttgaag tgggatcagt attctcccat 240

aatccccagc aagcggagca gggtatttta gaagaatgga aacaggcaat caaatTTTat 300

ttacccaaag aactacaacc atcctattgg ccagagcgtg aaacagactc aggaataggc 360

aaaactTTaa gtgagtggtc cgcacaatct ccaagacctc ttgtaatttt ttacatgaa 420

atcgattccc taacagatga agctttaatc ctaattTTaa gacaattacg ctcaggTTTT 480

ccccgctgc ctcggggatt tccccattcg gtgggggttaa ttggtatgcg ggatgtgcgg 540
 gactataagg ttaaatctgg tggaagtga cgaactgaata cgtcaagtcc tttaatatc 600
 aaagcggaat ccttgacttt aagtaatttc actctgtcag aggtggaaga actttactta 660
 caacatacgc aagctacagg acaaattttt accccggaag caattaaaca agcattttat 720
 ttaaccgatg ggcaaccatg gttagtaaac gccctagctc gtcaagccac tcaggtgtta 780
 gtgaaagata ttactcaacc cattaccgct gaagtaatta accaagccaa agaagttctg 840
 attcagcgcc aggataccca ttgggatagt ttggcagagc gcttacggga agatcgggtc 900
 aaagccatta ttcaacctat gttagctgga tcggacttac cagatacccc agaggatgat 960
 cgccgtttct tgctagattt aggcttggt aagcgcagtc ccttgggagg actaaccatt 1020
 gccaatccca ttaccagga ggtgattcct cgtgtttgt cccagggtag tcaggatagt 1080
 ctaccccaga ttcaacctac ttgggtaaat actgataata ctttaaatec tgacaaactc 1140
 ttaaatgctt tcctagagt ttggcgacaa catggggaac cattactcaa aagtgcgcct 1200
 tatcatgaaa ttgctcccca tttagttttg atggcgtttt tacatcgggt agtgaatggt 1260
 ggtggcactt tagaacggga atatgccgtt ggttctggaa gaatggatat ttgtttacgc 1320
 tatggcaagg tagtgatggg catagagtta aaggtttggg ggggaaaatc ggatccgtta 1380
 acgaagggtt tgaccaat ggataaatat ctgggtgggt taggattaga tagaggttgg 1440
 ttagtaattt ttgatcccg tccgggatta ccacccatgg gtgagaggat tagtatggaa 1500
 caggccatta gtccagaggg aagaaccatt acagtgattc gtagctag 1548

<210> 110

<211> 515

<212> PRT

<213> *Cylindrospermopsis raciborskii* AWT205

<400> 110

Met Pro Lys Tyr Phe Asn Thr Ala Gly Pro Cys Lys Ser Glu Ile His
 1 5 10 15

Tyr Met Leu Ser Pro Thr Ala Arg Leu Pro Asp Leu Lys Ala Leu Ile
 20 25 30

Asp Gly Glu Asn Tyr Phe Ile Ile His Ala Pro Arg Gln Val Gly Lys
 35 40 45

Thr Thr Ala Met Ile Ala Leu Ala Arg Glu Leu Thr Asp Ser Gly Lys
 50 55 60

Tyr Thr Ala Val Ile Leu Ser Val Glu Val Gly Ser Val Phe Ser His
 65 70 75 80

Asn Pro Gln Gln Ala Glu Gln Val Ile Leu Glu Glu Trp Lys Gln Ala
 85 90 95

Ile Lys Phe Tyr Leu Pro Lys Glu Leu Gln Pro Ser Tyr Trp Pro Glu
 100 105 110

Arg Glu Thr Asp Ser Gly Ile Gly Lys Thr Leu Ser Glu Trp Ser Ala
 115 120 125

Gln Ser Pro Arg Pro Leu Val Ile Phe Leu His Glu Ile Asp Ser Leu
 130 135 140

Thr Asp Glu Ala Leu Ile Leu Ile Leu Arg Gln Leu Arg Ser Gly Phe
 145 150 155 160

Pro Arg Arg Pro Arg Gly Phe Pro His Ser Val Gly Leu Ile Gly Met
 165 170 175

Arg Asp Val Arg Asp Tyr Lys Val Lys Ser Gly Gly Ser Glu Arg Leu
 180 185 190

Asn Thr Ser Ser Pro Phe Asn Ile Lys Ala Glu Ser Leu Thr Leu Ser
 195 200 205

Asn Phe Thr Leu Ser Glu Val Glu Glu Leu Tyr Leu Gln His Thr Gln
 210 215 220

Ala Thr Gly Gln Ile Phe Thr Pro Glu Ala Ile Lys Gln Ala Phe Tyr
 225 230 235 240

Leu Thr Asp Gly Gln Pro Trp Leu Val Asn Ala Leu Ala Arg Gln Ala
 245 250 255

Thr Gln Val Leu Val Lys Asp Ile Thr Gln Pro Ile Thr Ala Glu Val
 260 265 270

Ile Asn Gln Ala Lys Glu Val Leu Ile Gln Arg Gln Asp Thr His Leu
 275 280 285

Asp Ser Leu Ala Glu Arg Leu Arg Glu Asp Arg Val Lys Ala Ile Ile
 290 295 300

Gln Pro Met Leu Ala Gly Ser Asp Leu Pro Asp Thr Pro Glu Asp Asp
 305 310 315 320

Arg Arg Phe Leu Leu Asp Leu Gly Leu Val Lys Arg Ser Pro Leu Gly
 325 330 335

Gly Leu Thr Ile Ala Asn Pro Ile Tyr Gln Glu Val Ile Pro Arg Val
 340 345 350

Leu Ser Gln Gly Ser Gln Asp Ser Leu Pro Gln Ile Gln Pro Thr Trp
 355 360 365

Leu Asn Thr Asp Asn Thr Leu Asn Pro Asp Lys Leu Leu Asn Ala Phe
 370 375 380

Leu Glu Phe Trp Arg Gln His Gly Glu Pro Leu Leu Lys Ser Ala Pro
 385 390 395 400

Tyr His Glu Ile Ala Pro His Leu Val Leu Met Ala Phe Leu His Arg
 405 410 415

Val Val Asn Gly Gly Gly Thr Leu Glu Arg Glu Tyr Ala Val Gly Ser
 420 425 430

Gly Arg Met Asp Ile Cys Leu Arg Tyr Gly Lys Val Val Met Gly Ile
 435 440 445

Glu Leu Lys Val Trp Gly Gly Lys Ser Asp Pro Leu Thr Lys Gly Leu
 450 455 460

Thr Gln Leu Asp Lys Tyr Leu Gly Gly Leu Gly Leu Asp Arg Gly Trp
 465 470 475 480

Leu Val Ile Phe Asp His Arg Pro Gly Leu Pro Pro Met Gly Glu Arg
 485 490 495

Ile Ser Met Glu Gln Ala Ile Ser Pro Glu Gly Arg Thr Ile Thr Val
 500 505 510

Ile Arg Ser
 515

<210> 111

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* AWT205 sequence

<400> 111

acttctctcc ttccctatc

20

<210> 112

<211> 22

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* AWT205 sequence

<400> 112

gagtgaaaat gcgtagaact tg

22

<210> 113
 <211> 22
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 113
 cccaatatct ccctgtaaaa ct 22

<210> 114
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 114
 tggcaattgt ctctccgtat 20

<210> 115
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 115
 ctgccgatg aaagtcctct 20

<210> 116
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 116
 gcgtgtcgag aaaaaggtgt 20

<210> 117
 <211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 117

ctcgacacgc aagaataacg

20

<210> 118

<211> 21

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 118

atgcttctgc ttggcatgg c

21

<210> 119

<211> 21

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 119

taactcgacg aactttgacc c

21

<210> 120

<211> 19

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3

<400> 120

gccgccaatc ctcgcatg

19

<210> 121

<211> 22

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 121

gaacgtctaa tgtgcacag tg

22

<210> 122

<211> 23

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 122

ctggtacgta gtcgcaaagg tgg

23

<210> 123

<211> 26

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 123

ctgacggtac atgtattcc tgtgac

26

<210> 124

<211> 30

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 124

cgtctcatat gcagatctta ggaatttcag

30

<210> 125

<211> 25

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 125

gcttactacc acgatagtc tgccg

25

<210> 126

<211> 22

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 126

tctatgttta gcaggtggtg tc

22

<210> 127

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 127

ttctgcaaga cgagccataa

20

<210> 128

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 128

gggtcgccgc ggacattaa

20

<210> 129

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 129
 atgctaatagc ggtgggagta 20

<210> 130
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 130
 aaagcagttc cgacgacatt 20

<210> 131
 <211> 23
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 131
 cctatttcga ttattgtttt cgg 23

<210> 132
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 132
 gataccgatc ataaactacg 20

<210> 133
 <211> 21
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 133
 gcaaatttg caggagtaat g 21

<210> 134
 <211> 21
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 134

gcaaattttg caggagtaat g

21

<210> 135
 <211> 23
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 135

tttgggtaa actttatagc cat

23

<210> 136
 <211> 22
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 136

tgggtctgga cagttgtaga ta

22

<210> 137
 <211> 23
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 137

aaggggaaaa caaaattatc aat

23

<210> 138
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 138
 ggcgatcgcc tgctaaaaat 20

<210> 139
 <211> 23
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 139
 cctcatttc atttctagac gtt 23

<210> 140
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 140
 ccacttcaac taaaacagca 20

<210> 141
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 141
 aaaaattttg gaggggtagc 20

<210> 142
 <211> 20

<212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 142
 atccaagatg cgacaacact 20

<210> 143
 <211> 21
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 143
 ggtccttgcg cagatagagt g 21

<210> 144
 <211> 21
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 144
 cactctatct gcgcaaggac c 21

<210> 145
 <211> 21
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 145
 tgactgcatt cgctgtataa a 21

<210> 146
 <211> 22
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 146

ttcataagac ggctgttgaa tc

22

<210> 147

<211> 30

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 147

ctcgagttaa aaaagagtgt aaatgaaagg

30

<210> 148

<211> 23

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 148

ttctataact gctgccaaat ttt

23

<210> 149

<211> 23

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 149

aattttggag tgactggta tgg

23

<210> 150

<211> 23

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 150

ccataaccag tcactccaaa att

23

<210> 151

<211> 21

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 151

ttttagttgt tacttttggc g

21

<210> 152

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 152

acagcagatg agagaaagta

20

<210> 153

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 153

gggttgctt gctgatttc

20

<210> 154

<211> 22

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 154
cattaaaata agtccggaca gg 22

<210> 155
<211> 20
<212> DNA
<213> Artificial

<220>
<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 155
ttaaacagaa tgaggagcaa 20

<210> 156
<211> 20
<212> DNA
<213> Artificial

<220>
<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 156
aaacaacaca cccatctaag 20

<210> 157
<211> 20
<212> DNA
<213> Artificial

<220>
<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 157
ttaataaggc atccccaaga 20

<210> 158
<211> 20
<212> DNA
<213> Artificial

<220>
<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 158
gaaatggctg tgtaaaaact 20

<210> 159
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 159
 tctgccatat cccaaccta 20

<210> 160
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 160
 gatcgcccga caggaagact 20

<210> 161
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 161
 tccggcttga cctgctggac 20

<210> 162
 <211> 20
 <212> DNA
 <213> Artificial

<220>
 <223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 162
 tgcgatgatt ttgcctctgt 20

<210> 163
 <211> 20
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 163

aaaatttgca cacccacacg

20

<210> 164
 <211> 27
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 164

ttggattgaa cgtgtaattg aaaaagc

27

<210> 165
 <211> 27
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 165

gctttttcaa ttacacgttc aatccaa

27

<210> 166
 <211> 19
 <212> DNA
 <213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 166

aaatggcgta tcgactaac

19

<210> 167
 <211> 21

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 167

atataggagc gcataaagtg c

21

<210> 168

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 168

cttggataa gtcttgat

20

<210> 169

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 169

aacactcatt agattcatct

20

<210> 170

<211> 21

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 170

tccactaat ccttgaatt g

21

<210> 171

<211> 21

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 171

tgttgtctg gatcgatcc t 21

<210> 172

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 172

gcagttcagg tccatgaaac 20

<210> 173

<211> 20

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 173

agcccagtc caaccttcgt 20

<210> 174

<211> 21

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 174

tctggaagta ctgcactgt c 21

<210> 175

<211> 22

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 175

tgtaactccg tcaggacata aa

22

<210> 176

<211> 23

<212> DNA

<213> Artificial

<220>

<223> Based on *Cylindrospermopsis raciborskii* T3 sequence

<400> 176

tgcaaatttt agtagcaata acg

23

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