

(12) STANDARD PATENT
(19) AUSTRALIAN PATENT OFFICE

(11) Application No. **AU 2009290452 B2**

(54) Title
A method for predicting athletic performance potential

(51) International Patent Classification(s)
C12Q 1/68 (2006.01)

(21) Application No: **2009290452** (22) Date of Filing: **2009.09.11**

(87) WIPO No: **WO10/029527**

(30) Priority Data

(31) Number	(32) Date	(33) Country
61/213,125	2009.05.08	US
61/136,533	2008.09.11	US

(43) Publication Date: **2010.03.18**

(44) Accepted Journal Date: **2015.01.29**

(71) Applicant(s)
University College Dublin - National University of Ireland, Dublin

(72) Inventor(s)
Hill, Emmeline;McHugh, David;Orr, Nick;Gu, Jingjing;Katz, Lisa

(74) Agent / Attorney
Phillips Ormonde Fitzpatrick, 367 Collins Street, Melbourne, VIC, 3000

(56) Related Art
WO 2008/034177 A1

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
18 March 2010 (18.03.2010)

PCT

(10) International Publication Number
WO 2010/029527 A1

(51) International Patent Classification:
C12Q 1/68 (2006.01)

(21) International Application Number:
PCT/IE2009/000062

(22) International Filing Date:
11 September 2009 (11.09.2009)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
61/136,533 11 September 2008 (11.09.2008) US
61/213,125 8 May 2009 (08.05.2009) US

(71) Applicant (for all designated States except US): **UNIVERSITY COLLEGE DUBLIN - NATIONAL UNIVERSITY OF IRELAND, DUBLIN** [IE/IE]; Belfield, Dublin 4 (IE).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **HILL, Emmeline** [IE/IE]; University College Dublin - National University of Ireland, Dublin, Belfield, Dublin 4 (IE). **McHUGH, David** [IE/IE]; University College Dublin - National University of Ireland, Dublin, Belfield, Dublin 4 (IE). **ORR, Nick** [GB/IE]; University College Dublin - National University of Ireland, Dublin, Belfield, Dublin 4 (IE). **GU, JingJing** [CN/IE]; University College Dublin - National University of Ireland, Dublin, Belfield, Dublin 4 (IE). **KATZ, Lisa** [US/IE]; University College Dublin - Na-

tional University of Ireland, Dublin, Belfield, Dublin 4 (IE).

(74) Agents: **O'BRIEN, John A.** et al.; John A. O'Brien & Associates, Third Floor, Duncairn House, 14 Carysfort Avenue, Blackrock, County Dublin (IE).

(81) Designated States (unless otherwise indicated, for every kind of national protection available): AF, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PE, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

— with international search report (Art. 21(3))

[Continued on next page]

(54) Title: A METHOD FOR PREDICTING ATHLETIC PERFORMANCE POTENTIAL

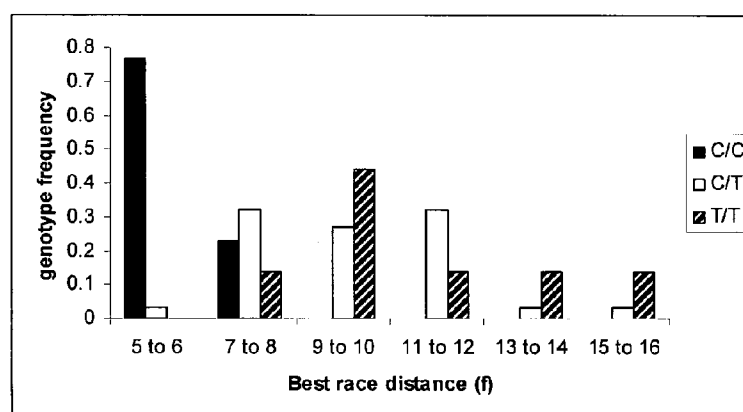


Fig. 7

(57) Abstract: A method and assay for predicting athletic performance potential of a subject, such as a thoroughbred race horse, comprising the steps of assaying a biological sample from a subject for the presence of a single nucleotide polymorphism in one or more genes associated with athletic performance. The athletic performance genes may be selected from one or more of MSTN, COX4I2, PDK4, CKM and COX4I1.

WO 2010/029527 A1



— before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments (Rule 48.2(h)) — with sequence listing part of description (Rule 5.2(a))

“A method for predicting athletic performance potential”

The invention relates to a method for predicting the athletic performance potential of a subject.

5 Introduction

The Thoroughbred horse industry is a multi-billion euro international industry involved in the breeding, training and racing of Thoroughbred horses. Often multi-million euro decisions are made on the purchase of individual animals with perceived racing potential. The integration of genomics information into the Thoroughbred racing and breeding industries has huge potential
10 for early ‘talent identification’. Thoroughbreds are traditionally selected for racing and breeding based on pedigree information as well as numerous phenotypic characteristics. Early identification of genetic potential, by traditional or new means, is paramount to success. Within the industry the quest to find an ‘edge’ pushes those involved to constantly consider new methods and techniques. Therefore, genomics information has the potential to directly assist
15 breeders and trainers to fine-tune often multi-million dollar decisions by providing previously inaccessible information.

Oxygen is an essential regulator of muscle function, influencing energy production, muscle contraction and removal of by-products. During exercise the requirement for energy is greatly
20 limited by the availability of oxygen. Mammalian cells have evolved elaborate adaptive mechanisms to respond to low cellular oxygen environments (Taylor & Colgan 1999). In studies of human exercise, adaptation to such a hypoxic environment in trained skeletal muscle causes a shift in substrate selection to increased oxidation of carbohydrates and stimulates cells to improve conditions for oxygen transport and utilisation (Hoppeler & Vogt 2001). In
25 Thoroughbred horses, despite a number of structural and functional adaptations in the cardiovascular and respiratory systems that improve oxygen carrying capacity and delivery during high-intensity short-duration exercise, the oxygen transport system lags far behind peripheral demand reflected in the routine development of an exercise-induced arterial hypoxemia and hypercapnia (Dempsey & Wagner, 1999; Seaman, 1995). The Thoroughbred
30 response is extreme in comparison to other animal species, including trained human athletes, reflecting the enormous requirement of the musculature for energy. Remarkably, even faced with a limited oxygen supply, Thoroughbreds remain elite athletes exquisitely adapted to extreme exercise.

Thoroughbred horses excel in both sprint (<1 mile) and longer distance (>1 mile) races. The physiological requirements for these disciplines differ and are regulated by the partitioning of metabolic pathways. During the first 75 seconds of exercise at supramaximal intensities (105-125% VO₂max) horses experience an oxygen deficit because oxygen supply cannot meet the demand of exercising muscles (Dempsey & Wagner, 1999; Seaman, 1995). Despite this, it has been estimated that during sprint races (< 1000m) approximately 70% of the total energy will be supplied aerobically. Horses competing over longer distances and for longer duration (>75 seconds) reach steady-state VO₂ and therefore are not oxygen deficient.

10 A range of approaches has been taken to investigate measurable associations with athletic performance phenotype in Thoroughbred racehorses including assessment of heart size (Young et al 2005), muscle fibre type (Rivero et al. 2007) musculoskeletal conformation (Love et al 2006), speed at maximum heart rate (Gramkow & Evans 2006), haematological (Revington 1983) and other physiological variables (Harkins et al 1993).

15 WO2006003436 describes the association between performance and gene variants encoded by the mitochondrial genome. However, mitochondrial DNA (mtDNA) haplotypes are inherited strictly from the maternal parent and therefore relate solely to female contributions to the phenotype. As there is a limited number of mtDNA haplotypes ($n = 17$) in the Thoroughbred population and just 10 females contribute to 74% of present maternal lineages (Cunningham et al 2002) it is unlikely that these haplotype variants have a significant effect as the favourable haplotypes would become 'fixed' quickly in a population where there is targeted selection for performance; in addition, the effective population size (of mtDNA variants) is one third of nuclear-encoded variants (Ballard and Dean 2001, Blier et al 2001, Das 2006, Meiklejohn et al 2007). Also, mtDNA haplotypes can be directly inferred from pedigree information.

It is an object of the invention to provide a method for predicting the athletic performance potential of a subject that overcomes some of these problems.

30 Statements of Invention

This invention provides DNA-based tests for detecting variation in nuclear-encoded genes. This approach is superior to mitochondrial DNA (mtDNA) testing because variation in nuclear encoded genes reflects inheritance of favourable gene variants from all possible ancestors whereas mtDNA testing is restricted to female ancestry.

The methods and assays described herein are performed *ex vivo* and can be considered to be *ex vivo* or *in vitro* methods and assays.

- 5 Any suitable biological sample which contains genetic material for example, blood, saliva, hair, skin, bone marrow, soft tissue, internal organs, biopsy sample, semen, skeletal muscle tissue and the like, may be used as a biological sample for the methods described herein. Blood and hair samples are particularly suitable as a biological sample.
- 10 "Athletic performance" as used herein includes racing such as competitive racing and equestrian sports such as racing, showjumping, eventing, dressage, endurance events, riding, hunting and the like. The equestrian sports may be competitive sports.

Competitive racing species include equines (horses), camels, dogs, elephants, hares, kangaroos,
15 ostriches, pigeons, *Homo sapiens* and birds of prey such as hawks or falcons. The competitive racing species may be a Thoroughbred race horse or a showjumping horse.

By "primer" we mean a nucleic acid sequence containing between about 15 to about 40 for example between about 18 to about 25 contiguous nucleotides from a nucleic acid sequence of
20 interest. The primer may be a forward (5' or 3') or reverse (3' to 5') primer or a primer designed on a complementary nucleic acid sequence to the sequence of interest. In the present invention, the sequence of interest is the genomic sequence of a gene associated with athletic performance, for example a gene listed in the appendices or one or more of the COX4I1, COX4I2, PDK4, CKM or MSTN genes. In one embodiment, the primer may comprise between about 15 to about
25 40 for example between about 18 to about 25 contiguous nucleotides from SEQ ID No. 1, SEQ ID No. 2, SEQ ID No. 3, SEQ ID No. 31 or SEQ ID No. 32 or between about 15 to about 40 for example between about 18 to about 25 contiguous nucleotides from a complementary sequence to SEQ ID No. 1, SEQ ID No. 2, SEQ ID No. 3, SEQ ID No. 31 or SEQ ID No. 32. By
30 "complementary sequence" we mean a sequence that binds to the sequence of interest using conventional Watson-Crick base pairing i.e. adenine binds to thymine and cytosine binds to guanine.

The invention provides single nucleotide polymorphisms (SNPs) that are associated with elite athletic performance. The invention provides a method of predicting the athletic performance of

a subject comprising the step of assaying a biological sample from the subject for the presence of a single nucleotide polymorphism (SNP) in one or more of the genes listed in the appendices wherein the SNP has a significant association with athletic performance.

- 5 According to the invention there is provided a method for predicting the athletic performance potential of a subject comprising the step of assaying a biological sample from a subject for the presence of a single nucleotide polymorphism (SNP) in one or more of the *MSTN* gene, *COX4I2* gene, *PDK4* gene, *CKM* gene or *COX4I1* gene.
- 10 The SNP may be *MSTN*_66493737 (T/C). The presence of a C allele is indicative of elite athletic performance. The presence of a homozygous CC genotype may be indicative of elite athletic performance. The elite athletic performance may be elite sprinting performance

- The SNP may be *COX4I2*_22684390 (C/T). The presence of a T allele may be indicative of elite athletic performance. The presence of a homozygous TT genotype may be indicative of elite athletic performance.
- 15

- The SNP may be *PDK4*_38973231 (A/G). The presence of an A allele may be indicative of elite athletic performance. The presence of a homozygous AA genotype may be indicative of elite athletic performance.
- 20

- The SNP may be *CKM*_15884567 (G/A). The presence of an A allele may be indicative of elite athletic performance. The presence of a homozygous AA genotype may be indicative of elite athletic performance.
- 25

- The SNP may be *COX4I1*_32772871 (T/C). The presence of a T allele may be indicative of elite athletic performance. The presence of a homozygous TT genotype may be indicative of elite athletic performance.

- 30 The biological sample of the subject may be selected from the group comprising: blood, saliva, skeletal muscle, skin, semen, biopsy, bone marrow, soft tissue, internal organs and hair.

The subject may be from a competitive racing species. The subject may be an equine such as a Thoroughbred race horse.

The invention further provides an assay for determining the athletic performance potential of a subject comprising the steps of:

- 5 - obtaining a sample;
 - extracting or releasing DNA from the sample; and
 - identifying a single nucleotide polymorphism (SNP) in a target sequence from a gene associated with athletic performance in the extracted or released DNA
- 10 wherein the athletic performance potential of a subject is associated with the SNP.

The gene associated with athletic performance may be selected from one or more of *MSTN*, *COX4I2*, *PDK4*, *CKM* or *COX4I1*.

- 15 The assay may comprise the step of:
- amplifying a target sequence from a gene associated with athletic performance in the extracted or released DNA
- 20 prior to the step of identifying a single nucleotide polymorphism.

The DNA may be genomic DNA

- The invention further provides an assay for use in determining the athletic performance potential
- 25 of a subject comprising means for detecting the presence of a single nucleotide polymorphism (SNP) in one or more of the *MSTN* gene, *COX4I2* gene, *PDK4* gene, *CKM* gene or *COX4I1* gene.

- The SNP may be *MSTN*_66493737 (T/C). The presence of a C allele is indicative of elite
- 30 athletic performance. The presence of a homozygous CC genotype may indicative of elite athletic performance. The elite athletic performance may be elite sprinting performance.

The SNP may be COX4I2_22684390 (C/T). The presence of a T allele may be indicative of elite athletic performance. The presence of a homozygous TT genotype may be indicative of elite athletic performance.

- 5 The SNP may be PDK4_38973231 (A/G). The presence of an A allele may be indicative of elite athletic performance. The presence of a homozygous AA genotype may be indicative of elite athletic performance.

- 10 The SNP may be CKM_15884567 (G/A). The presence of an A allele may be indicative of elite athletic performance. The presence of a homozygous AA genotype may be indicative of elite athletic performance.

- 15 The SNP may be COX4I1_32772871 (T/C). The presence of a T allele may be indicative of elite athletic performance. The presence of a homozygous TT genotype may be indicative of elite athletic performance.

The invention also provides an assay for determining the athletic potential of a subject comprising the step of:

- 20 - obtaining a sample;
 - extracting or releasing DNA from the sample;
 - identifying the genotype of the MSTN_66493737 (T/C) SNP in the extracted or released DNA

- 25 wherein the presence of a C allele in the MSTN_66493737 (T/C) SNP is indicative of elite athletic performance.

The assay may comprise the step of:

- 30 - amplifying a target sequence encoding the MSTN_66493737 (T/C) SNP in the extracted or released DNA

prior to the step of identifying the genotype of the MSTN_66493737 (T/C) SNP.

The presence of a homozygous CC genotype indicative of elite athletic performance.

The elite athletic performance may be elite sprinting performance.

- 5 The DNA may be genomic DNA.

The sample from the subject may be selected from the group comprising: blood, saliva, skeletal muscle skin, bone marrow, biopsy, soft tissue, semen, internal organ and hair.

- 10 The subject may be from a competitive racing species. The subject may be an equine such as a Thoroughbred race horse.

- We have also shown that homozygous carriers of the T allele of the *COX4I2* gene (EquCab2.0 22676361-C/T) single nucleotide polymorphism (SNP), i.e. those that have the polymorphism in
15 both alleles of the *COX4I2* gene, are statistically more likely to be elite sprinting racehorses compared to subjects that are heterozygous for the SNP. i.e. subjects that have the polymorphism in one of the alleles of the *COX4I2* gene, or subjects that do not have the SNP in either allele of the *COX4I2* gene.

- 20 We describe a method of predicting athletic performance of a subject comprising the step of assaying a biological sample from the subject for the presence or absence of a single nucleotide polymorphism (SNP) in the *COX4I2* gene. The SNP may be EquCab 2.0 COX4I2-22676361 – C/T. The presence of a homozygous TT genotype may be indicative of elite athletic performance. The presence of a homozygous TT genotype may be indicative of elite aerobic
25 performance. The presence of a homozygous TT genotype may be indicative of elite sprinting performance. The biological sample of the subject may be selected from the group comprising: blood, saliva, skeletal muscle, semen, biopsy, internal organ, skin, bone marrow (or any other biological tissue) and hair. The subject may be from a competitive racing species. The subject may be an equine. The subject may be a Thoroughbred race horse.

30

We also describe an assay for use in determining athletic performance of a subject comprising means for detecting the presence or absence of a single nucleotide polymorphism (SNP) in the *COX4I2* gene. The SNP may be EquCab 2.0 COX4I2-22676361 – C/T. The presence of a homozygous TT genotype may be indicative of elite athletic performance. The biological

sample of the subject may be selected from the group comprising: blood, saliva, skeletal muscle, semen, biopsy, internal organ, skin, bone marrow (or any other biological tissue) and hair. The subject may be from a competitive racing species. The subject may be an equine. The subject may be a Thoroughbred race horse.

5

Brief Description of the Drawings

The invention will be more clearly understood from the following description of an embodiment thereof, given by way of example only, with reference to the accompanying drawings, in which:-

10 Fig. 1 is a schematic of the partitioning of energy during exercise in horses;

Fig. 2 is a bar chart showing the distribution of *COX4I2* 22676361 (C/T) SNP genotypes in Thoroughbred subpopulations (TBE_EN: elite performing Thoroughbreds over distances >8f; TBE_SP: elite performing Thoroughbreds over distances <8f; TBO: other
15 Thoroughbreds that have raced but have never won a race and have a handicap rating < 70) and in non-Thoroughbred horses (AH: Akhal-Teke; CON: Connemara Pony; TU: Tuva). Elite Thoroughbreds that have successfully competed over distances <8f have a significantly higher frequency of the TT genotype than other Thoroughbred subpopulations and non-Thoroughbreds;

20

Fig. 3 is a schematic showing the relationship between three of the main metabolic pathways contributing to energy production during exercise, the function of three genes CKM, *COX4I2* and *PDK4* associated with elite racing performance are shown;

25 Fig. 4 (A) to (D) are graphs showing the allele frequency distribution among elite (hatched bar) and non-elite Thoroughbreds for CKM 22684390 (C/T) SNP (A), *COX4I2* 22684390 (C/T) SNP (B) and *PDK4* 38973231 (A/G) SNP (C) and among elite sprinters (hatched bar) and elite endurance Thoroughbreds for *MSTN* 66493737 (T/C) SNP (D);

30 Fig. 5 (A) to (D) are graphs showing the genotype frequency distributions among elite (hatched bar) and non-elite Thoroughbreds for CKM 22684390 (C/T) SNP (A), *COX4I2* 22684390 (C/T) SNP (B) and *PDK4* 38973231 (A/G) SNP (C) and among elite sprinters (hatched bar) and elite endurance Thoroughbreds for *MSTN* 66493737 (T/C) SNP (D);

Fig. 6 (A) to (C) are graphs showing the genotype frequency for best race distance for the MSTN 66493737 (T/C) SNP in which (A) shows the C/C genotype frequency; (B) shows the C/T genotype frequency; and (C) shows the T/T genotype frequency;

5 Fig. 7 is a graph showing the genotype frequency for best race distance for the MSTN 66493737 (T/C) SNP in which the best race distance for horses that had won their group race as a two-year-old was replaced with the average distance of their three-year-old races;

10 Fig. 8 is a graph showing the genotype frequency for the MSTN 66493737 (T/C) SNP in a non-thoroughbred population known for endurance exercise capabilities (Egyptian Arabian horse) and a thoroughbred population; and

15 Fig 9 is a graph showing the genotype frequency for the MSTN 66493737 (T/C) SNP for stallions with a Stamina Index 6 - 8f, 8-10 f, 10-12f; and

Fig 10 is a graph showing the relative expression of MSTN gene for the MSTN 66493737 (T/C) SNP C/C, C/T and T/T genotypes.

20 Detailed Description

Intense selection for elite racing performance in the Thoroughbred horse (*Equus caballus*) has resulted in a number of adaptive physiological phenotypes relevant to exercise, however the underlying molecular mechanisms responsible for these characteristics are not well understood.

25 Eivers et al (2009) investigated adaptive changes in mRNA expression in equine skeletal muscle for a panel of candidate exercise-response genes following a standardised incremental-step treadmill exercise test in eight unconditioned Thoroughbred horses. In the study, biopsy samples were obtained from the *gluteus medius* pre-exercise (T_0), immediately post-exercise (T_1) and four hours post-exercise (T_2). They detected significant ($P < 0.05$) fold differences relative to T_0 in
30 eight genes (*CKM*, *COX4I1*, *COX4I2*, *PDK4*, *PPARGC1A*, *PRKAA1*, *SLC2A1*, and *SLC2A4*) at T_2 . By studying the relationships between mRNA and velocity at maximum heart rate (VHR_{max}) and peak post-exercise plasma lactate concentration ($[La]T_1$), they demonstrated significant ($P < 0.05$) associations with *COX4I1* and *PPARGC1A* at T_2 and between $[La]T_1$ and *COX4I1* at T_0 . In a follow-on study they investigated gene expression changes in a second cohort of horses after a

ten month period of conditioning. They showed that in resting samples, the *COX4I1* gene had a significant increase in abundance following conditioning and, after exercise in the conditioned cohort, significant fold differences were identified in *COX4I2*, *PDK4* and *PPARGC1A* at T₂. They also detected significant relationships with VHR_{max} and [La]T₁ for *PPARGC1A* and

5 *COX4I1*.

The present invention relates to a previously unknown relationship between sequence variants (such as SNPs) in a number of candidate exercise response genes (listed in the appendices) and retrospective athletic performance (given as racecourse success i.e. Group winner or non-winner,

10 handicap rating (RPR) and best race distance for Group winners) in Thoroughbred race horses. In some aspects, the invention relates to SNPs in the *COX4I1*, *COX4I2*, *PDK4*, *CKM* and *MSTN* genes.

COX4I1 and *COX4I2*

15 Cytochrome C oxidase (COX) is a multi-subunit enzyme (Complex IV) that catalyzes the electron transfer from reduced cytochrome C to oxygen in mitochondrial respiration. COX is a dimer in which each monomer is made up of 13 subunits, three of which are encoded by the mitochondrial genome (COX1, 2 and 3). Nuclear encoded COX4 is responsible for the regulation and assembly of mitochondrially encoded subunits on the inner mitochondrial

20 membrane (Fukuda et al. 2007). In human skeletal muscle, *COX4* mRNA levels have been shown to be associated with mitochondrial volume and, by extension, VO_{2max}. COX4 comprises two isoforms (COX4-1 and COX4-2) encoded by the *COX4I1* and *COX4I2* genes that are differentially regulated in normoxic and hypoxic environments (Fukuda et al. 2007). In normal oxygen environments *COX4I1* is preferentially transcribed. In limited oxygen environments HIF-

25 1 activates transcription of *COX4I2* and the mitochondrial *LON* gene. As *LON* inhibits the expression of *COX4I1*, these control mechanisms result in increased *COX4I2* transcription and protein synthesis and decreased COX4-1 availability. This mechanism has been postulated to be a strategy to maximise the efficiency of cellular respiration in limited oxygen environments (Fukuda et al. 2007).

30

The physiological requirements during a race differ depending on the energy demand and are regulated by the partitioning of metabolic pathways to provide energy in the most efficient manner. During the first 75 seconds of exercise at supramaximal intensities (105 – 125% VO_{2max}) horses experience an oxygen deficit because oxygen supply cannot meet the demand of

exercising muscles (Dempsey and Wagner 1999; Seaman et al. 1995). Over longer distances and for longer duration (> 75 seconds) horses reach steady-state VO_2 and rely principally on aerobic metabolism. At the end of a race anaerobic demand increases as horses pass the 'lactate threshold'. During short distance races (< 1,000 m) approximately 70% of the total energy in the form of ATP, necessary for muscle contraction, is generated by aerobic metabolic pathways (Eaton et al. 1995). In Thoroughbred horses exercising at supramaximal intensities over short distances this hypoxic environment may trigger the well-conserved metabolic switch from COX4-1 to COX4-2 utilisation (Fukuda et al. 2007). This environmental regulation of COX4-2 may increase the efficiency of cellular respiration. COX4-2 may therefore be an important regulator of energy supply in the early stages of a race and towards the end of a race when oxygen is limited. As can be seen from Fig. 1, large amounts of energy are required while peripheral physiological systems (i.e. skeletal muscle) are operating in limited oxygen environments in the early stages of exercise and towards the end of a race. Generation of energy via COX4-2 may be important during both these stages.

It has been suggested that regulation of mitochondrial biogenesis may be mediated by glucocorticoid hormone (Weber et al 2002). The *COX4I2* gene contains a glucocorticoid receptor element (TGTT) which may be targeted to increase COX4-2 expression and therefore increase mitochondrial volume. Also, the *COX4I2* gene contains a p53 tumor suppressor binding site (CATG). Recent studies have suggested that p53 may play a role in regulation of mitochondrial biogenesis and aerobic metabolism via COX (Matoba et al. 2006; Saleem et al. 2009).

CKM

Creatine kinase (CK), also known as creatine phosphokinase (CPK) or phosphocreatine kinase, is an enzyme (EC 2.7.3.2) expressed by various tissue types. It catalyses the conversion of creatine and consumes adenosine triphosphate (ATP) to create phosphocreatine and adenosine diphosphate (ADP). In tissues that consume ATP rapidly, especially skeletal muscle, but also brain and smooth muscle, phosphocreatine serves as an energy reservoir for the rapid regeneration of ATP. Thus creatine kinase is an important enzyme in such tissues.

In most cells the CK enzyme consists of two subunits, which can be either *B* (brain type) or *M* (muscle type). There are, therefore, three different isoenzymes: CK-MM, CK-BB and CK-MB. The genes for these subunits are located on different chromosomes. In addition, there are two

mitochondrial creatine kinases, the ubiquitous and sarcomeric form. The different types of CK isoenzymes are listed in Table 1.

Table 1 – Isoenzymes of creatine kinase

gene	protein
CKB	creatine kinase, brain
CKBE	creatine kinase, ectopic expression
CKM	creatine kinase, muscle
CKMT1A	creatine kinase, mitochondrial 1A
CKMT1B	creatine kinase, mitochondrial 1B
CKMT2	creatine kinase, mitochondrial 2 (sarcomeric)

5

Isoenzyme patterns differ depending on tissue type. For example, CK-BB occurs mainly in brain tissues, and its levels rarely have any significance in skeletal muscle. Skeletal muscle expresses CK-MM (98%) and low levels of CK-MB (1%) whereas in contrast the myocardium (heart muscle) expresses CK-MM at about 70% and CK-MB at 25-30%.

10

The mitochondrial creatine kinase (CK_m), which produces ATP from ADP by converting creatine phosphate to creatine, is present in the mitochondrial intermembrane space. Apart from the mitochondrial form, there are three forms present in the cytosol—CK_a (in times of acute need, produces ATP in the cytosol at the cost of creatine phosphate), CK_c (maintains critical concentration of creatine and creatine phosphate in the cytosol by coupling their phosphorylation and dephosphorylation respectively with ATP and ADP) and CK_g (which couples direct phosphorylation of creatine to the glycolytic pathway).

15

The creatine kinase, muscle gene (*CKM*) encodes a muscle type isozyme of creatine kinase found exclusively in striated muscle. The encoded protein is involved in cellular energetics. During exercise *CKM* gene knockout mice show a lack of burst activity but maintain normal absolute muscle force (van Deursen et al. 1993). We have found that *CKM* gene transcripts are the most abundant transcripts in the Thoroughbred horse skeletal muscle transcriptome, supporting the pivotal role played by the *CKM* gene in exercise adaptation in the horse.

25

PDK4

The regulation of glucose utilisation is tightly controlled by the uptake of glucose by glucose transporters, the rate of glycolytic flux and the conversion of pyruvate to acetyl-CoA in mitochondria via the catalytic function of the pyruvate dehydrogenase complex (PDC). The critical rate limiting step in the oxidation of glucose is the regulation of assembly of the PDC which is controlled by pyruvate dehydrogenase kinase (PDK). PDK blocks the formation of the PDC resulting in the beta-oxidation of fatty acids to acetyl-CoA as the substrate for oxidative phosphorylation. Three genes (*PDK2*, *PDK3* and *PDK4*) of the four genes that encode PDK isoforms are located in positively selected genomic regions in Thoroughbred (Gu et al 2009). The *PDK4* gene promoter contains a binding site for the *FOXO1A* transcription factor, a key regulator of insulin signalling in liver and adipose tissue. Single nucleotide polymorphisms in *FOXO1A* have been found to have a protective effect on T2DM development and related phenotypes in humans. *FOXO1A* has also been found among positively selected genomic regions in Thoroughbred and its *PDK4* promoter binding site sequence is conserved in horse. The transcription factors FOXO1 and SMAD have also been shown to be responsible for myostatin (*MSTN*) gene regulation and therefore play key roles in the regulation of muscle growth.

In a genome scan for positive selection, Gu et al (2009) detected a region that deviated very significantly from neutral expectations in two independent statistical tests (F_{ST} and Ewens-Watterson test). This region contained the *PDK4* gene. *PDK4* gene expression is co-ordinated by the transcriptional co-activator PGC-1 α via ERR α (estrogen-related receptor alpha) binding. PGC-1 α , encoded by the *PPARGC1A* (peroxisome proliferator-activated receptor gamma, coactivator 1 alpha) gene, is a key regulator of energy metabolism that regulates insulin sensitivity by controlling glucose transport via SLC2A4 (solute carrier family 2 (facilitated glucose transporter), member 4; previously GLUT4) and drives the formation of oxidative muscle fibres and co-ordinates mitochondrial biogenesis via its interaction with nuclear encoded mitochondrial protein genes.

MSTN

Myostatin is also known as growth/differentiation factor 8 precursor (GDF-8). In several mammalian species (including cattle, sheep and dogs), the double muscling trait is caused by mutations in the myostatin (*MSTN*) gene. In dogs, *MSTN* gene mutations in racing whippets have been associated with the 'bully' phenotype and heterozygous individuals are significantly faster than individuals carrying the wild-type genotype (Mosher et al 2007). Mutations in the *MSTN* gene may be associated with athletic power.

We have analysed a number of single nucleotide polymorphisms (SNPs) in genes associated with athletic performance and have developed a simple DNA based method of predicting the athletic performance potential of a subject based on the SNPs.

- 5 The invention will be more clearly understood from the following examples.

Examples

Subjects

- 10 A Thoroughbred is a registered racehorse that can trace its ancestry to one of three foundation stallions and the approximately 30 foundation mares entered in The General Studbook, 1791 (Weatherby and Sons 1791). There are two types of Thoroughbred race: National Hunt races are run over hurdles or steeplechase fences over distances of up to 4.5 miles (7,200 m), while Flat races have no obstacles and are run over distances ranging from five furlongs (5/8 mile or 1,006
- 15 m) to 20 furlongs (4,024 m). The highest standard and most valuable elite Flat races are known as Group (Europe and Australasia) or Stakes races (North America). The most prestigious of these races include The Breeders' Cup races (United States), The Kentucky Derby (United States), The Epsom Derby (United Kingdom) *et cetera*.
- 20 Three hundred and fifty Group races are run in Europe (Britain, Ireland (incl. Northern Ireland), France, Germany, Italy) annually including 84 Group 1, 93 Group 2 and 173 Group 3 races. In the United Kingdom and Ireland 196 Group races are competed annually (43 Group 1, 50 Group 2 and 103 Group 3). Britain has the highest number of Group races (139) in Europe per annum, with 57% run over distances ≤ 1 mile (1609 meters) and 43% run over distances > 1 mile.
- 25 Australia has approximately 540-550 Group races per season from a total of almost 21,000 races and New Zealand hosts 78 Group races per season. After Group races, Listed races are the next highest grade of race.

- Horses that compete over distances ≤ 1 mile are known as 'sprinters' whereas horses that
- 30 compete over distances > 1 mile are known as 'stayers'. Horses competing in 1 mile races ('milers' and 'middle distance') may be considered either sprinters or stayers and the way in which a race is executed by the rider often reflects the trainers perceived ability ('sprinter' or 'stayer') of the horse. The International Federation of Horseracing Authorities recognizes five race distance categories: Sprint (5–6.5 f, $\leq 1,300$ m), Mile (6.51–9.49 f, 1,301–1,900 m),

Intermediate (9.5–10.5 f, 1,901–2,112 m), Long (10.51–13.5 f, 2,114–2,716 m) and Extended (> 13.51 f, > 2,717 m); S-M-I-L-E [Note: 1 furlong = 1/8 mile = 201.2 meters].

- To minimise confounding effects of racing over obstacles only horses with performance records
- 5 in Flat races were considered for inclusion in the study cohorts. In all cases pedigree information was used to control for genetic background by exclusion of samples sharing relatives within two generations. Also, overrepresentation of popular sires within the pedigrees was avoided where possible.
- 10 Samples from Thoroughbred horses were collected with informed owner's consent from racing, breeding and sales establishments in Ireland, Britain and New Zealand during 1997-2006. All horses were categorized based on retrospective racecourse performance records as "elite Thoroughbreds" (TBE) or "other Thoroughbreds" (TBO). Elite Thoroughbreds were flat race horses that had won at least one Group (Group 1, Group 2 or Group 3) race. Other
- 15 Thoroughbreds were those that had competed on the racetrack but had never won a flat race or had a handicap rating (Racing Post Racing (RPR)) of less than 89.

Example 1 – COX4I2

- During sprint exercise, energy in the form of ATP, necessary for muscle contraction, is generated
- 20 principally by aerobic metabolic pathways (70% aerobic, 30% anaerobic) albeit in a limited oxygen environment. We suggest that this relative hypoxic environment triggers the well-conserved metabolic switch from COX4-1 to COX4-2 thereby increasing the efficiency of cellular respiration. COX4-2 is therefore an important regulator of energy supply during sprinting, but not necessarily in longer distance competitions where oxygen demands are met.
- 25 This switch is mediated by the transcription factor HIF-1 α in the cell that has been well-characterised as the master regulator of hypoxia-dependent gene expression (Semenza 1998). HIF-1 α activates the transcription of genes encoding PDK1, LDHA, COX4-2 and LON and controls the switch from COX4-1 to COX4-2. In Thoroughbred muscle that is deprived of oxygen during intense exercise an enhanced response to reduced oxygen and the ability to
- 30 generate ATP in the most efficient manner will provide a significant advantage to that individual.

Also, increased mitochondrial volume has been shown to be associated with higher aerobic capacity (Fluck 2006). Increased amount of glucocorticoid hormone has been shown to stimulate mitochondrial biogenesis, either by specifically targeting the mitochondrial genome or by an

unknown mechanism (Weber et al 2002). Glucocorticoid receptor elements (Glucocorticoid responsive and related elements) contain the recognition sequence (TGTT). The COX4I2 gene contains one of these elements in Intron 2. Therefore glucocorticoid binding may stimulate increased gene expression leading to increased mitochondrial volume and therefore aerobic
5 energy capacity.

In some aspects, the present invention relates to a single nucleotide polymorphism (SNP) in COX4I2 that is significantly ($P < 0.01$) associated with elite sprinting performance. The significant association of the COX4I2 homozygous TT genotype (EquCab2.0 22676361-C/T) in
10 elite sprint race winners may be utilized in DNA-based tests of genetic potential for elite athletic performance in Thoroughbred horses.

Precise location of the COX4I2 gene (EquCab2.0 22676361-C/T) polymorphism

The exact location of the COX4I2 polymorphism is on *Equus caballus* chromosome 22 at
15 position 22676361 of the Horse Genome Sequence (Equus caballus Version .2.0) which can be viewed at www.broad.mit.edu/mammals/horse/. The COX4I2 polymorphism may be identified as EquCab2.0 COX4I2_22676361 (C/T) SNP.

The horse genome EquCab2 assembly is a Whole Genome Shotgun (WGS) assembly at 6.79x
20 and was released in September 2007. A female Thoroughbred named "Twilight" was selected as the representative horse for genome sequencing. The project coordination and genome sequencing and assembly is provided by the Broad Institute. The N50 size is the length such that 50% of the assembled genome lies in blocks of the N50 size or longer. The N50 size of the contigs is 112.38 kb, and the total length of all contigs is 2.43 Gb. When the gaps between
25 contigs in scaffolds are included, the total span of the assembly is 2.68 Gb. The horse EquCab2 was annotated using a standard Ensembl mammalian pipeline. Predictions from vertebrate mammals as well as horse proteins have been given priority over predictions from non-vertebrate mammals. The set of predictions has been compared to 1:1 homologues genes in human and mouse, and missing homologs in the horse annotation have been recovered using exonerate.
30 Horse and human cDNAs have been used to add UTRs to protein based predictions. The final gene-set comprises 20,737 protein-coding genes, 2,863 identified as pseudogenes and 1,580 classified as retro-transposed genes.

Genotyping of the EquCab2.0 COX4I2-22676361-C/T polymorphism

Genotyping of SNPs was conducted in a sample of Thoroughbreds ($n = 149$) comprising both elite ($n = 79$) and non-elite performers ($n = 70$). The elite performer group contained a subset of animals ($n = 70$) that competed preferentially in short distance (≤ 1 mile; $n = 34$) and long distance (> 1 mile; $n = 36$) races.

5

Genomic DNA was extracted from either fresh whole blood or hair samples. Blood samples were collected in 7 ml Vacutainer K₃EDTA blood collection tubes (Becton Dickinson, Franklin Lakes, NJ). Hair samples with visible hair roots were collected in labelled, airtight zip-lock bags. Samples were stored at 4°C prior to DNA extraction using a modified version of a standard
10 phenol / chloroform method (Sambrook and Russell 2001). DNA concentrations for all samples were estimated using a NanoDrop ND-1000 UV-Vis Spectrophotometer (NanoDrop Technologies, Wilmington, DE).

The flanking sequence and SNP (bold and square brackets) is as follows;

15 (bases indicated in lower case indicate that the sequence read was not optimal for this region of the flanking sequence)

caagagtggagtgtgctccaagaactggaggctagcatgtagcagaggaggcagtagcagaggaggagaggttgatgggggagctgc
atttggagagtctggcaggcaggacctgaatgccaggctaaggagtttATTGGGAGGCAAGTGGGTGCTGATAA
20 AGGCTCAAGGATTCCATCAGGCTGTTCCACAAAGACC[C/T]GGGCCACCTCAGGGC
ACCATATCCCCATATCCAGGAGCCAGTTGTGTCCCAGAGAAAACAAGGGACTGGAC
CTTGAGACTTGGCCAGTGTCTTCACATCCTACCCTGTGCACGCCCTGTTTGGCCTG
TGGTGCAGAAGGCCCTGGGAGACCTGAAGCAGAAGCTGCAGACCATTCAGGTTA
GTGTGGAGCCCCAGA

25 (SEQ. ID No. 1)

Genotyping of the *COX4I2* gene was performed by KBiosciences (www.kbioscience.co.uk) using either competitive allele specific PCR (KASPar) or Taqman (Applied Biosystems). KASPar is a proprietary in house homogeneous fluorescent genotyping system.

30

25 μ L of total DNA was supplied to KBiosciences at a concentration of 15 ng/ μ L in “v-bottomed” 96 well micro-titre plates. Also included were a number of samples for set-up and assay validation ($n = 24$) and blank ($n = 1$ per 96 well plate) samples to check for reproducibility and to control for errors in sample handling

Genetic analysis study

Following genotyping, a genetic analysis was carried out on the subjects (n = 149) described above.

5

Individual dichotomous logistic regression models were fitted for each SNP. Genotype trend effects were modeled by estimating the risk associated with a linear trend in magnitude of effect relative to the common homozygote, heterozygote, and rare homozygote genotypes. *P*-values were determined from a likelihood ratio test statistic and approximated according to an asymptotic χ^2 distribution with one degree of freedom. The best genetic model for significantly associated SNPs was determined by repeating the analysis with coding variables for additive, recessive and overdominant models.

10

Table 2 shows the EquCab2.0 *COX4I2*_22676361 (C/T) SNP genotype frequencies amongst the subjects.

<u>Elite sprinters (less than 8 furlongs) (n = 39) Vs Other elite race winners (n=36)</u>				
	Sp (<8f) vs En			
<u>Genotype</u>	OR	lower	upper	p-value
dominant (CC v CT-TT)	1.77	0.55	5.76	3.37E-01
recessive (CC-CT v TT)	4.89	1.21	19.75	<u>1.56E-02*</u>
over-dominant (CC-TT v CT)	0.56	0.2	1.52	2.50E-01
<u>Elite sprinters (less than 7 furlongs) (n = 28) Vs Other elite race winners (n=36)</u>				
	Sp (<7f) vs En			
<u>Genotype</u>	OR	lower	upper	p-value
dominant (CC v CT-TT)	2.05	0.54	7.69	2.78E-01
recessive (CC-CT v TT)	5.6	1.31	23.86	<u>1.25E-02*</u>
over-dominant (CC-TT v CT)	0.53	0.18	1.57	2.53E-01

15

Wherein: **Sp = sprinter**
 En = endurance (or 'stayer')
 OR = Odds ration

Lower = lower confidence interval

upper = upper confidence interval

Referring to Table 2 the EquCab2.0 *COX4I2*_22676361 (C/T) SNP homozygote TT genotype is significantly associated with elite racing (sprinting) performance over distances < 8 furlongs (less than 1 mile) ($P < 0.02$) (Fig. 2) and this association is more pronounced over distances < 7f ($P < 0.01$).

Thoroughbred horses carrying the homozygote T allele (TT) of the *COX4I2*_22676361 (C/T) SNP have a greater sprinting ability compared to Thoroughbred horses carrying the heterozygous T allele (TC) or to Thoroughbred horses that do not carry the T allele (CC). Therefore, the sprinting performance of a Thoroughbred horse can be predicted by testing a biological sample for the presence or absence of the homozygote T allele of the *COX4I2*_22676361 (C/T) SNP.

15 **Example 2 – Genes associated with Thoroughbred racing performance**

We investigated associations between 80 SNPs in the following genes: *ACN9*, *ACSS1*, *ACTA1*, *ACTN2*, *ADHFE1*, *GGPS1*, *GSN*, *MC3R*, *MTFR1*, *NDUFA8*, *PDK4*, *PON1*, *PTGS1*, *PTPN1*, *TNC*, *TOMM20*, *UGCG CKM*, *COX4I2*, *COX4I1*, *HIF1A*, *MYEF2*, and *PRKAA1* (details of the SNPs are given in the appendices with racing performance in Thoroughbreds).

20

The present invention identifies significant associations between SNPs and athletic performance phenotypes in a set of these genes including *ACN9*, *ACSS1*, *ACTN2*, *ADHFE1*, *CKM*, *COX4I2*, *GSN*, *MSTN*, *PON1*, *PTGS1* and *PTPN1* (see the appendices). Because of the known gene expression response to exercise in equine skeletal muscle (Eivers et al 2009) and evidence for association with performance in dogs (Mosher et al 2008) and response to training, four of the genes (*CKM*, *COX4I2*, *PDK4* and *MSTN*) that had a significant association with Thoroughbred racing performance were investigated in detail. SNPs in three of those genes (*CKM*, *COX4I2* and *PDK4*) are associated with elite (Group race winning) performance and a SNP in the *MSTN* gene is associated with elite sprint race performance.

30

In this example, the following sample set was used:

Table 3 – Details of samples included in each subpopulation.

Sample Set B	<i>n</i>	No. sires	No. Gr race winners	Mean RPR	Range RPR	Total no. races	Mean no. races	No. races won	No. Gr races won	No. Gr 1 races won	Mean no. Gr races won
TB	148	136									
TBE	86	86	84	115	87-134	1170	13.8	425	215	91	2.5
TBO	62	62	0	59	21 - 89	537	8.7	15	0	0	0

In which TB is Thoroughbred, TBE is Elite Group race winning Thoroughbred; and TBO is non-elite (i.e other non-winning) Thoroughbred. The TBE cohort was further subdivided into TBE_sprinter (*n* = 39) and TBE_endurance or 'stayer' (*n* = 32).

5

Genetic analysis study

Tests for association of SNPs with athletic performance were performed using the program PLINK (<http://pngu.mgh.harvard.edu/purcell/plink/> Purcell et al., 2007)

10 Standard case/control association analysis

To perform a standard case/control association analysis, the option: `plink --file mydata --assoc` was used, which generates a file `plink.assoc` containing the fields:

	CHR	Chromosome
15	SNP	SNP ID
	BP	Physical position (base-pair)
	A1	Minor allele name (based on whole sample)
	F_A	Frequency of this allele in cases
	F_U	Frequency of this allele in controls
20	A2	Major allele name
	CHISQ	Basic allelic test chi-square (1df)
	P	Asymptotic p-value for this test
	OR	Estimated odds ratio (for A1)

25 Association analysis

It is possible to perform tests of association between a disease and a variant other than the basic allelic test (which compares frequencies of alleles in cases versus controls), by using the `--model` option. The tests offered here are (in addition to the basic allelic test):

- Cochran-Armitage trend test
- 30 • Genotypic (2 df) test

- Dominant gene action (1df) test
- Recessive gene action (1df) test

One advantage of the Cochran-Armitage test is that it does not assume Hardy-Weinberg equilibrium, as the individual, not the allele, is the unit of analysis (although the permutation-based empirical p-values from the basic allelic test also have this property). SNPs showing severe deviations from Hardy-Weinberg are often likely to be bad SNPs, or reflect stratification in the sample, however, and so are probably best excluded in many cases.

10 The genotypic test provides a general test of association in the 2-by-3 table of disease-by-genotype. The dominant and recessive models are tests for the minor allele (which is the minor allele can be found in the output of either the --assoc or the --freq commands. That is, if D is the minor allele (and d is the major allele):

15 Allelic: D versus d
 Dominant: (DD, Dd) versus dd
 Recessive: DD versus (Dd, dd)
 Genotypic: DD versus Dd versus dd

20 As mentioned above, these tests are generated with option plink --file mydata --model which generates a file plink.model containing the following fields:

25 CHR Chromosome number
 SNP SNP identifier
 TEST Type of test
 AFF Genotypes/alleles in cases
 UNAFF Genotypes/alleles in controls
 CHISQ Chi-squared statistic
 DF Degrees of freedom for test
 30 P Asymptotic p-value

Each SNP will feature on five rows of the output, corresponding to the five tests applied. The column TEST refers to either ALLELIC, TREND, GENO, DOM or REC, referring to the

different types of test mentioned above. The genotypic or allelic counts are given for cases and controls separately. For recessive and dominant tests, the counts represent the genotypes, with two of the classes pooled.

- 5 These tests only consider diploid genotypes: that is, for the X chromosome males will be excluded even from the ALLELIC test. This way the same data are used for the five tests presented here. Note that, in contrast, the basic association commands (--assoc and --linear, etc) include single male X chromosomes, and so the results may differ.
- 10 The genotypic and dominant/recessive tests will only be conducted if there is a minimum number of observations per cell in the 2-by-3 table: by default, if at least one of the cells has a frequency less than 5, then the alternate tests are skipped (NA is written in the results file). The Cochran-Armitage and allelic tests are performed in all cases. This threshold can be altered with the --cell option: plink --file mydata --model --cell 20

15

Results of the association tests are provided in full in the appendices. A number of SNPs in the four genes *PDK4*, *COX4I2*, *CKM* and *MSTN* were investigated. For each gene we selected the SNP with the most significant association (P value) (Table 4) for the trait for further investigation. The 4 SNPs with greatest association with athletic performance were

20 *PDK4*_38973231 – A/G, *COX4I2*_22684390 – C/T, *CKM*_15884567 – G/A, and *MSTN*_66493737 – T/C. SNPs in *PDK4*, *CKM* and *COX4I2* were associated with elite (Group race winning) performance and a SNP in *MSTN* was associated with elite sprinting performance in Thoroughbred racehorses. The best fit genotypic models were assigned based on the results in Table 5 below.

25

Table 4 - Results of SNP association tests for elite (Group winning) performance (*PDK4*, *CKM* and *COX4I2*) and elite sprinting performance (*MSTN*) in Thoroughbred racehorses. SNPs with the most significant association in each gene are shown here.

CHR	SNP	BP	A1	A2	F _{A(A1)}	F _{A(A2)}	F _{U(A1)}	F _{U(A2)}	CHISQ	P	OR
4	<i>PDK4</i> _38973231	3924	A	G	0.464	0.536	0.282	0.718	9.874	0.001676	2.2
10	<i>CKM</i> _15884567	2716	G	A	0.074	0.926	0.164	0.836	5.355	0.02066	0.4089
22	<i>COX4I2</i> _22684390	1164	C	T	0.325	0.675	0.455	0.546	4.654	0.03098	0.5778
18	<i>MSTN</i> _	212	T	C	0.282	0.718	0.641	0.359	18.31	1.88E-	4.5

	66493737								05	
--	----------	--	--	--	--	--	--	--	----	--

In which A1: allele 1; A2: Allele 2; F_A(A1): frequency of allele 1 in elite TB (PDK4, CKM and COX4I2) and elite sprinters (MSTN); F_A(A2): frequency of allele 2 in elite TB (PDK4, CKM and COX4I2) and elite sprinters (MSTN); F_U(A1): frequency of allele 1 in non-elite TB
 5 (PDK4, CKM and COX4I2) and elite endurance (MSTN); F_U(A2): frequency of allele 2 in non-elite TB (PDK4, CKM and COX4I2) and elite endurance (MSTN).

The SNPs that were chosen for further investigation were as follows:

- 10 PDK4_38973231 (A/G)
 COX4I2_22684390 (C/T)
 CKM_15884567 (G/A)
 MSTN_66493737 (T/C)

15 Table 5 - Association test results for best-fit model

CHR	SNP	A1	A2	TEST	AFF	UNAFF	CHISQ	DF	P
4	PDK4_38973231	A	G	GENO	18/41/24	6/23/33	9.644	2	0.008049
4	PDK4_38973231	A	G	TREND	77/89	35/89	9.237	1	0.002372
4	PDK4_38973231	A	G	ALLELIC	77/89	35/89	9.874	1	0.001676
4	PDK4_38973231	A	G	DOM	59/24	29/33	8.791	1	0.003027
4	PDK4_38973231	A	G	REC	18/65	6/56	3.706	1	0.05422
22	COX4I2_22684390	C	T	GENO	4/44/32	10/30/15	6.979	2	0.03052
22	COX4I2_22684390	C	T	TREND	52/108	50/60	5.58	1	0.01817
22	COX4I2_22684390	C	T	ALLELIC	52/108	50/60	4.654	1	0.03098
22	COX4I2_22684390	C	T	DOM	48/32	40/15	2.326	1	0.1272
22	COX4I2_22684390	C	T	REC	4/76	10/45	6.093	1	0.01357
10	CKM_15884567	G	A	GENO	1/10/70	2/14/39	5.03	2	0.08087
10	CKM_15884567	G	A	TREND	12/150	18/92	4.865	1	0.02741
10	CKM_15884567	G	A	ALLELIC	12/150	18/92	5.355	1	0.02066

	15884567								
10	CKM_15884567	G	A	DOM	11/70	16/39	4.953	1	0.02605
10	CKM_15884567	G	A	REC	1/80	2/53	0.876	1	0.3493
18	MSTN_66493737	T	C	GENO	3/16/20	9/23/0	23.8	2	0.000006799
18	MSTN_66493737	T	C	TREND	22/56	41/23	20.64	1	0.000005545
18	MSTN_66493737	T	C	ALLELIC	22/56	41/23	18.31	1	0.00001875
18	MSTN_66493737	T	C	DOM	19/20	32/0	22.85	1	0.000001755
18	MSTN_66493737	T	C	REC	3/36	9/23	5.225	1	0.02226

The best fit genotypic models were assigned based on the results in Table 5. The best model for association of the SNPs with athletic performance was concluded as follows:

5 *PDK4* - Allelic → A allele is preferred i.e. AA or AG

CKM - Allelic → A allele is preferred i.e. AA or AG

COX4I2 - Recessive → T allele is preferred i.e. TT

10

MSTN - Genotypic → Genotype predicts distance category. (in the cohort used in this example, none of the 'stayers' were CC but 50% sprinters were CC)

The allele frequency distributions among Elite and Non-elite Thoroughbreds for *CKM*, *COX4I2*

15 and *PDK4* and among Elite Sprinters and Elite Endurance for *MSTN* are shown in Fig. 4. Table 6 below shows the allele frequencies for the four SNPs in Thoroughbreds (TBE and TBO).

Table 6 - Allele frequencies for the four SNPs in Thoroughbreds (TBE and TBO)

CHR	SNP	A1	A2	MAF	NCHROBS
4	PDK4_38973231	A	G	0.3862	290
10	CKM_15884567	G	A	0.1103	272
22	COX4I2_22684390	C	T	0.3778	270
18	MSTN_66493737	T	C	0.4353	278

In which MAF is Minor Allele Frequency and NCHROBS Number of Chromosomes analysed.

The genotype frequency distributions among Elite and Non-elite Thoroughbreds for the SNPs in
5 *CKM*, *COX4I2* and *PDK4* and among Elite Sprinters and Elite Endurance for the SNP in *MSTN*.
The results of this study are shown in Fig. 5 and Table 7 below.

Table 7 - Genotype frequencies in elite and non-elite Thoroughbred sub-populations for SNPs:
PDK4 (PDK4_38973231); COX4I2 (COX4I2_22684390); CKM (CKM_15884567) and in elite
10 sprinters and elite endurance Thoroughbreds for SNP: MSTN (MSTN_66493737).

		AA	AG	GG		AA	AG	GG
	ALL	24	64	57	145	0.17	0.44	0.39
PDK4	TBE	18	41	24	83	0.22	0.49	0.29
	TBO	6	23	33	62	0.10	0.37	0.53
		CC	CT	TT		CC	CT	TT
	ALL	14	74	47	135	0.10	0.55	0.35
COX4I2	TBE	4	44	32	80	0.05	0.55	0.40
	TBO	10	30	15	55	0.18	0.55	0.27
		GG	GA	AA		GG	GA	AA
	ALL	3	24	109	136	0.02	0.18	0.80
CKM	TBE	1	10	70	80	0.01	0.13	0.88
	TBO	2	14	39	55	0.04	0.25	0.71
		TT	TC	CC		TT	TC	CC
	ALL	23	75	41	139	0.17	0.54	0.29
MSTN	TBE_SP	3	16	20	39	0.08	0.41	0.51
	TBE_EN	9	23	0	32	0.28	0.72	0.00

Deviations from Hardy-Weinberg equilibrium (HWE) for the four SNPs in the sample cohort were investigated to determine departure from expected neutral genetic drift. Deviation from
15 HWE may be an indicator of selection and may alter the expected distribution of genotypes in a population given the allele frequencies. This information is required to correctly assign genotype frequencies to enable the test for performance.

Table 8 - Tests for deviations from Hardy-Weinberg equilibrium in ALL (All TB); AFF (Elite
20 and elite sprinters); UNAFF (non-elite and elite endurance).

CHR	SNP	TEST	A1	A2	GENO	O(HET)	E(HET)	P
4	PDK4	ALL	A	G	24/64/57	0.4414	0.4741	0.3872

	38973231							
4	PDK4_38973231	AFF	A	G	18/41/24	0.494	0.4974	1
4	PDK4_38973231	UNAFF	A	G	6/23/33	0.371	0.4052	0.5332
10	CKM_15884567	ALL	G	A	3/24/109	0.1765	0.1963	0.2033
10	CKM_15884567	AFF	G	A	1/10/70	0.1235	0.1372	0.3553
10	CKM_15884567	UNAFF	G	A	2/14/39	0.2545	0.2737	0.6188
22	COX4I2_22684390	ALL	C	T	14/74/47	0.5481	0.4701	0.06812
22	COX4I2_22684390	AFF	C	T	4/44/32	0.55	0.4388	0.03941
22	COX4I2_22684390	UNAFF	C	T	10/30/15	0.5455	0.4959	0.5891
18	MSTN_66493737	ALL	T	C	23/75/41	0.5396	0.4916	0.3022
18	MSTN_66493737	AFF	T	C	3/16/20	0.4103	0.405	1
18	MSTN_66493737	UNAFF	T	C	9/23/0	0.7188	0.4604	0.001815

Deviation from HWE was identified in COX4I2_22684390-C/T in the Elite Thoroughbred sub-population. Thus genotype frequencies may be adjusted in the test to account for the over-representation of the TT genotype among elite racehorses. Also, deviation from HWE was

5 identified in MSTN_66493737-T/C in the elite endurance Thoroughbred cohort. Thus genotype frequencies may be adjusted in the test to account for the under-representation of the CC genotype among elite endurance racehorses.

Example 3 - A multi-gene prediction test for elite (Group race winning) performance in

10 **Thoroughbred horses**

The present invention provides a simple DNA based method (genotypic test) for predicting the athletic performance of a thoroughbred race horse based on the presence or absence of a SNP in one or more exercise response gene. The exercise response gene may be one or more of the genes listed in the appendices. For example the genotypic test may be based on a SNP in one or

15 more of the *PDK4*, *CKM*, *COX4I2*, *COX4I1*, *MSTN*, *ACSSI*, *ACTN2* or *PTGSI* genes. Details of some of the SNPs that may be used to predict the athletic performance of a thoroughbred horse

are given in the appendices. It will be appreciated that the genotypic test may be based on a combination of any one or more of these SNPs.

Referring to Fig. 3, the three main metabolic pathways contributing to energy production during exercise and the location in the pathways and the function of three genes (*CKM*, *COX4I2* and *PDK4*) associated with elite racing performance are shown. Using knowledge of known function, the knowledge that the genes are expressed in skeletal muscle in response to exercise and the results in Example 2 above, in this non-limiting example, we developed a simple DNA based genotypic test for predicting elite performance in Thoroughbred horses based on SNPs in the *PDK4*, *COX4I2* and *CKM* genes.

COX4I2 SNP (*COX4I2*_22684390 (C/T) SNP)

This SNP is located on Chromosome 22 of *Equus caballus* at position 22,684,390 bp forward strand of the Horse Genome Sequence (*Equus caballus* Version 2.0) which can be viewed at www.broad.mit.edu/mammals/horse/.

The horse genome EquCab2 assembly is a Whole Genome Shotgun (WGS) assembly at 6.79x and was released in September 2007. A female Thoroughbred named "Twilight" was selected as the representative horse for genome sequencing. The project coordination and genome sequencing and assembly is provided by the Broad Institute. The N50 size is the length such that 50% of the assembled genome lies in blocks of the N50 size or longer. The N50 size of the contigs is 112.38 kb, and the total length of all contigs is 2.43 Gb. When the gaps between contigs in scaffolds are included, the total span of the assembly is 2.68 Gb. The horse EquCab2 was annotated using a standard Ensembl mammalian pipeline. Predictions from vertebrate mammals as well as horse proteins have been given priority over predictions from non-vertebrate mammals. The set of predictions was been compared to 1:1 homologues genes in human and mouse, and missing homologs in the horse annotation have been recovered using exonerate. Horse and human cDNAs have been used to add UTRs to protein based predictions. The final gene-set comprises 20,737 protein-coding genes, 2,863 identified as pseudogenes and 1,580 classified as retro-transposed genes.

Further details of the SNP are as follows:

- SNP: *COX4I2*_22684390 (C/T) $P = 0.03098$ OR = 0.5778
- EquCab2.0 SNP_ID: BIEC2-617568

- Genomic location of SNP: Intron 2
- Substitution type: Transition
- The SNP disrupts a glucocorticoid response element (GRE) binding site (**C/T**GTT). The favourable allele (T) retains the site (TGTT), therefore enabling GRE binding and increasing expression of the gene. The less favourable allele (C) disrupts the site (CGTT), therefore disabling GRE binding and not increasing expression of the gene. Alternatively, the SNP disrupts a p53 tumor suppressor binding site (**CAC/T**G). The favourable allele (T) retains the site (CATG), therefore enabling p53 binding and contributing to mitochondrial biogenesis. The less favourable allele (C) disrupts the site (CACG) therefore disabling the p53 binding and not contributing to mitochondrial biogenesis.

The flanking sequence and SNP (bold and square brackets) is as follows:

GCTGGGCGATCCTGGGGACATAAAAGTGAATCACCTGGATGGTTCTTGCCCTCAGG
 GTGCTCCAGTCCAGTGGGGGAACCAACACAAGCCCAGATAACTGTAATATAGGAT
 15 ATGTGGCGAGGGTGAAGTGTGTTCAAGGGGCTGTGAGGACCCAAAGGAGAGAGAG
 ATGAAATCCTGGTGGGCCTTCCAGAGGAGGGCA[T/C]GTTCTAGTTGACCTTGAATG
 GTGAGGCTGAGGGTGCTGCCAGGTGGTGGGAACAGCATGGGTAAGGGTATGGGAGC
 GGAAGAGCATGGAGGGTCCTAGGCATCAGTAAGTGCTGTAGGGGAAGGAACAGAG
 AGAGGCGGTGAGGTGGCCAGGAAAGAAGGGGGCCTGACCCTGGGGAGCAGGAGGG
 20 ATGTGTGACTCCAA

(SEQ ID No. 2)

CKM SNP (CKM_15884567 (G/A) SNP)

- 25 This SNP is located on Chromosome 10 of *Equus caballus* at position 15,884,567 bp of the Horse Genome Sequence (*Equus caballus* Version 2.0) which can be viewed at www.broad.mit.edu/mammals/horse/.

- 30 The horse genome EquCab2 assembly is a Whole Genome Shotgun (WGS) assembly at 6.79x and was released in September 2007. A female Thoroughbred named "Twilight" was selected as the representative horse for genome sequencing. The project coordination and genome sequencing and assembly is provided by the Broad Institute. The N50 size is the length such that 50% of the assembled genome lies in blocks of the N50 size or longer. The N50 size of the contigs is 112.38 kb, and the total length of all contigs is 2.43 Gb. When the gaps between

contigs in scaffolds are included, the total span of the assembly is 2.68 Gb. The horse EquCab2 was annotated using a standard Ensembl mammalian pipeline. Predictions from vertebrate mammals as well as horse proteins have been given priority over predictions from non-vertebrate mammals. The set of predictions was been compared to 1:1 homologues genes in human and
 5 mouse, and missing homologs in the horse annotation have been recovered using exonerate. Horse and human cDNAs have been used to add UTRs to protein based predictions. The final gene-set comprises 20,737 protein-coding genes, 2,863 identified as pseudogenes and 1,580 classified as retro-transposed genes.

10 Further details of the SNP are as follows:

- SNP: CKM_15884567 (G/A) P = 0.02066 OR = 0.4089
- EquCab2.0 SNP_ID: BIEC2-109575
- Genomic location of SNP: Intron 4
- Substitution type: Transition
- 15 ○ The SNP disrupts an Interferon regulatory factor binding site (**GCAAGA**). The A allele retains the site (GCAA). The G allele disrupts the site (GCGA).

The flanking sequence and SNP (bold and square brackets) is as follows:

CTGTCCCTAACAGACCTGGACCTTGGCCCCGTGGAGGTCCTAAAGGCRACCTATACGC
 20 GATGTAAACCCAAATTCATGACATCCCCTGAAGCATGCTCTTCCCCTGTCTGCCCCGG
 GTCCCCGGAACAGCCACCCCAAGTGCTCTCTCCCAAGTGGACTCTCCCTTCACACCC
 TGCCCCCTCGCATCCAGTGCACCGGCAAGC**[A/G]**AACTATCCCGGTGCCCACTCCAG
 AAAGTCAATGTCTCAGGAATCTGGGGAGCCATCAGTCAAAATTACTATCATAACAGT
 ATATATAGGATTCGCATATATTCCTATGCATAATAATTATACGTTTTGTGGATAATA
 25 AATATATGTATATATGCATAATTTACATAATATATACATATTTATATACATTTTAT
 ACATT

(SEQ ID No. 3)

30 PK4 (PK4_38973231 (A/G) SNP)

This SNP is located on Chromosome 4 of *Equus caballus* at position 38,973,231 bp of the Horse Genome Sequence (*Equus caballus* Version 2.0) which can be viewed at www.broad.mit.edu/mammals/horse/.

The horse genome EquCab2 assembly is a Whole Genome Shotgun (WGS) assembly at 6.79x and was released in September 2007. A female Thoroughbred named "Twilight" was selected as the representative horse for genome sequencing. The project coordination and genome sequencing and assembly is provided by the Broad Institute. The N50 size is the length such that 5 50% of the assembled genome lies in blocks of the N50 size or longer. The N50 size of the contigs is 112.38 kb, and the total length of all contigs is 2.43 Gb. When the gaps between contigs in scaffolds are included, the total span of the assembly is 2.68 Gb. The horse EquCab2 was annotated using a standard Ensembl mammalian pipeline. Predictions from vertebrate mammals as well as horse proteins have been given priority over predictions from non-vertebrate mammals. The set of predictions was been compared to 1:1 homologues genes in human and mouse, and missing homologs in the horse annotation have been recovered using exonerate. Horse and human cDNAs have been used to add UTRs to protein based predictions. The final gene-set comprises 20,737 protein-coding genes, 2,863 identified as pseudogenes and 1,580 classified as retro-transposed genes.

15

Further details of the SNP are as follows:

- SNP: PDK4_38973231 (A/G) P = 0.001676 OR = 2.2
- EquCab2.0 SNP_ID: BIEC2-903605
- Genomic location of SNP: Intron 2
- 20 ○ Substitution type: Transition
- The SNP is 4 bp upstream from a transcription factor (Two-handed zinc finger homeodomain transcription factors) binding site (ta**g**gtgTGTTCaga).

The flanking sequence and SNP (bold and square brackets) is as follows:

25 ACTTTAACCCTCAACTTTCTAACTTAAAATTTATGTTTAACTATTCCAGAGCAATATT
CAGTTTTATTTGGCAAATGTTTTTCATTTTTTATAGCAAAAGTATTTAGAAATTTTAA
GAAAGATTTTCATATTTCTTTCTACTTCATTCATTCATGTGTGGGTAGAAGTCTCGAAA
GCAGCAGTAAAGACTATGGATTGAAT[**A/G**]TGTGTTTCAGATTGTCATTGTTTAATGG
GTATGGAATGCATATATTTCTTGAATCAATGAACAAAACGCTGTATAGTCAGCAGAT
30 TAGGGTGAGGCTCTGGTGCATATCTGCTGCAGTGCATATCCTGGCTCTATTCTCTGA
AAATCTGCTCTTGTGGGTCATCTACCCTCTCTAAGCTTMAGCACCCCTATTTGTAAAA

(SEQ ID No. 4)

Risk prediction

The prediction of 'risk' for complex traits is greatly enhanced by testing multiple genes contributing to a trait, rather than relying on single gene SNPs (Yang et al 2003) if the additive genetic variance is small. However, single SNPs may be used where the effect is large (i.e. high odds ratio). Based on subpopulation prediction using population allele frequencies (for SNPs in HWE) or observed genotype frequencies (for SNPs deviating from HWE) and Bayes Theorem we investigated the probability of being a member of one or other subpopulation (elite or non-elite) given a certain combination of genotypes for the sequence variants in the *PDK4*, *CKM* and *COX4I2* genes. Results are provided in Table 9 below as a percentage chance of being a member of each of the two subpopulations.

Table 9 - Predictive test for Elite racing ability using SNPs in the genes *PDK4*, *CKM* and *COX4I2*. (The genotype combinations are ranked by most to least favourable for racing ability)

				TBE					TBO				
				Population 1: allele, genotype freqs					Population 2: allele, genotype freqs				
	AA	AB	BB	A	AA	AB	BB	P(G C)	A	AA	AB	BB	P(G C)
Locus													
PDK4	1			0.464	0.22	0.50	0.29	0.22	0.282	0.08	0.41	0.52	0.08
COX4I2			1	0.325	0.05	0.55	0.40	0.40	0.455	0.21	0.50	0.30	0.30
CKM			1	0.074	0.01	0.14	0.86	0.86	0.164	0.03	0.27	0.70	0.70
						P(G C)		0.074					0.02
PDK4	AA	AG	GG			P(C)		0.5					0.5
COX4I2	CC	CT	TT			P(G C) P(C)		0.037					0.01
CKM	GG	GA	AA										
						P(C G)		0.817					0.18
Sub-population prediction from genotype based on obs population genotype frequencies and Bayes Theorem													
	PDK4	COX4I2	CKM		TBE	TBO							
	AA	TT	AA		0.82	0.18							
	AA	CT	AA		0.79	0.21							
	AG	TT	AA		0.67	0.33							
	AA	TT	GA		0.65	0.35							
	AG	CT	AA		0.63	0.37							
	AA	CT	GA		0.60	0.40							
	GG	TT	AA		0.48	0.52							
	AG	TT	GA		0.45	0.55							
	AA	CC	AA		0.44	0.56							
	AA	TT	GG		0.44	0.56							
	GG	CT	AA		0.43	0.57							
	AG	CT	GA		0.40	0.60							
	AA	CT	GG		0.38	0.62							
	AG	CC	AA		0.27	0.73							
	GG	TT	GA		0.27	0.73							
	AA	CC	GA		0.25	0.75							
	AG	TT	GG		0.25	0.75							
	GG	CT	GA		0.23	0.77							
	AG	CT	GG		0.22	0.78							
	GG	CC	AA		0.14	0.86							

	AG	CC	GA			0.13	0.87										
	GG	TT	GG			0.13	0.87										
	AA	CC	GG			0.11	0.89										
	GG	CT	GG			0.11	0.89										
	AG	CC	GG			0.06	0.94										
	GG	CC	GA			0.06	0.94										
	GG	CC	GG			0.03	0.97										

In which for *PDK4*: AA represents genotype AA, AB represents genotype AG and BB represents genotype GG; for *COX4I2*: AA represents genotype CC, AB represents genotype CT, and BB represents genotype TT; and for *CKM*: AA represents genotype GG, AB represents genotype GA, and BB represents genotype AA.

From Table 9 it can be seen that the most favourable combination of genotypes at these three genes is AA, TT, AA for *PDK4*, *COX4I2* and *CKM* respectively (82% chance of being an elite racehorse, 18% chance of being a non-elite) and the least favourable combination of genotypes at these three genes is GG, CC, GG for *PDK4*, *COX4I2* and *CKM* respectively (3% chance of being an elite racehorse, 97% chance of being a non-elite).

The risk prediction test may be performed using one or more of the SNPs listed in the appendices.

Example 4 - Quantitative association of SNPs with handicap rating (RPR)

Racing Post Ratings (RPR) are a handicap rating determined by a horse's overall performance in a given race with respect to the race level, field quality, weight carried and time of the race. RPR are not directly comparable to speed ratings, rather the rating is intended to represent the weight a horse would be required to carry in a handicap. For example, in races restricted to horses of the same age and sex, a horse with a Racing Post Rating of 120 would, in a handicap, carry three pounds more than a horse rated 117. In open races, sex and weight-for-age allowances are factored in. Thus, in a handicap, if a horse carrying 120 pounds defeats a horse carrying 128 pounds by a length, the horse carrying 128 pounds will generally receive a Racing Post Rating six or seven pounds higher than the horse who carried 120 pounds. Guideline values to help determine a good rating for winners of races in different divisions are given in Table 10 below.

Table 10 – Guideline RPR for winning horses

	2-Year-Olds	3-Year-Olds	4-Year-Olds & Up
Group 1	120	125	130
Group 2	115	117	120
Group 3	105	110	115
Listed Race	95	105	110
Maidens	80	85	---

We examined whether there was a significant relationship between some of the SNPs that have shown a significant association with athletic performance and RPR in a quantitative association test analysis. Table 11 below shows three SNPs that are significantly associated with RPR.

5

Table 11 – SNPs having a significant association with RPR

CHR	SNP	STAT	EMP1	NP
4	PDK4_38973231	8.095	0.005052	4750
4	PDK4_38969307	6.825	0.009441	2541
3	COX4I1_32772871	6.748	0.009681	2478

Table 12 below shows the mean RPR for each genotype for the three significantly associated SNPs listed in Table 11 above.

10

Table 12 – Mean RPR for each genotype of the SNPs from Table 11

CHR	SNP	VALUE	G11	G12	G22
4	PDK4_38973231	GENO	A/A	A/G	G/G
4	PDK4_38973231	COUNTS	19	46	44
4	PDK4_38973231	FREQ	0.1743	0.422	0.4037
4	PDK4_38973231	MEAN	99.95	97.7	80.3
4	PDK4_38973231	SD	33.78	28.9	28.85
4	PDK4_38969307	GENO	A/A	A/C	C/C
4	PDK4_38969307	COUNTS	16	42	42
4	PDK4_38969307	FREQ	0.16	0.42	0.42
4	PDK4_38969307	MEAN	97.19	99.21	79.9
4	PDK4_38969307	SD	36.23	28.06	28.45

3	COX4I1_32772871	GENO	T/T	T/C	C/C
3	COX4I1_32772871	COUNTS	13	42	50
3	COX4I1_32772871	FREQ	0.1238	0.4	0.4762
3	COX4I1_32772871	MEAN	100.6	99.71	83.3
3	COX4I1_32772871	SD	29.46	28.92	30.49

Referring to Table 12 above, at PDK4_38973231-(A/G) the AA genotype has a mean RPR of 99.95, the AG genotype has a mean RPR of 97.7 and the GG genotype has a mean RPR of 80.3. Therefore we conclude that the AA and AG genotypes are the favourable genotypes correlated with higher RPR. At COX4I1_32772871-(C/T) the TT genotype has a mean RPR of 100.6, the TC genotype has a mean RPR of 99.71 and the CC genotype has a mean RPR of 83.3. Therefore we conclude that the TT and TC genotypes are the favourable genotypes correlated with higher RPR.

10 COX4I1 SNP (COX4I1_32772871 (C/T) SNP)

This SNP is located on Chromosome 3 of *Equus caballus* at position 32,772,871 bp of the Horse Genome Sequence (*Equus caballus* Version 2.0) which can be viewed at www.broad.mit.edu/mammals/horse/.

- 15 The horse genome EquCab2 assembly is a Whole Genome Shotgun (WGS) assembly at 6.79x and was released in September 2007. A female Thoroughbred named "Twilight" was selected as the representative horse for genome sequencing. The project coordination and genome sequencing and assembly is provided by the Broad Institute. The N50 size is the length such that 50% of the assembled genome lies in blocks of the N50 size or longer. The N50 size of the contigs is 112.38 kb, and the total length of all contigs is 2.43 Gb. When the gaps between contigs in scaffolds are included, the total span of the assembly is 2.68 Gb. The horse EquCab2 was annotated using a standard Ensembl mammalian pipeline. Predictions from vertebrate mammals as well as horse proteins have been given priority over predictions from non-vertebrate mammals. The set of predictions was been compared to 1:1 homologues genes in human and mouse, and missing homologs in the horse annotation have been recovered using exonerate. Horse and human cDNAs have been used to add UTRs to protein based predictions. The final gene-set comprises 20,737 protein-coding genes, 2,863 identified as pseudogenes and 1,580 classified as retro-transposed genes.

Further details of the SNP are as follows:

- SNP: COX4I1_32772871 (C/T) EMP1 = 0.009681
- EquCab2.0 SNP_ID: BIEC2-816028
- 5 ○ Genomic location of SNP: Intron
- Substitution type: Transition

The flanking sequence and SNP (bold and square brackets) is as follows in which M is A or C:

10 TCAGGTCTCAGTCGCACCAGAGCTGGATGGAGCCAGCGCAGCTCCATCTCTCAGTGG
 CTGGGAGTGGGCTGCAGGGTGGTCCTCACACAAGATGGGCACCTCCCTCCTGGGCTC
 CATCCCAGGACTGTTTCCCAGGTTTGGGAAACTGGCTCGCATTAGCCGAGTGGCGTG
 AGCCGGAATMTGATTTACTCACAGTGC GC[T/C]GTGCTTGGTGGGGAACGACTTCCC
 TGCTTTGTACAGCACCTGCGTTTCCAGTGGTGGTTTGTCTGGTCACTAGTCTTTTAT
 15 CAAGAGATAGTATAGTGAAGGTTAGGTCAAGGAAAAGGGAACTCTGACTTGTCAGA
 GGGCTGTTTGAAGTGTATGGGGACTGCATCTCGATAACCAGGATTCTGGGTCTCCAG
 ACCCA

(SEQ ID No. 32)

20

In a quantitative association test analysis this SNP is significantly associated with RPR.

Example 5 - A gene prediction test for elite sprint race performance in Thoroughbred horses.

Thoroughbred horses excel in both sprint (< 1,500 m) and longer distance (> 1,800 m) races.

- 25 Horses competing in middle distance races ('milers' and 'middle distance') may be considered either 'sprinters' or 'stayers' and the way in which a race is executed by the rider often reflects the trainer's perceived sprinting and endurance ability of the horse. Within the industry horses may be described as sprinters based on their conformation and usually have a stockier and more muscular stature and are faster maturing. They usually race as 2 year olds and over shorter
- 30 distances as 3 year olds. Individuals perceived to be longer distance animals may be referred to as 'backward' requiring more time to mature and running over longer distances as 3 year olds. In some regions (e.g. Australia) breeders attempt to breed only faster 'sprint' type horses.

- 35 In some aspects, the invention provides a simple DNA based method (genotype test) for predicting the elite sprint race performance of a thoroughbred race horse based on the presence

or absence of a SNP in one or more exercise response gene. The exercise response gene may be one or more of the genes listed in the appendices. For example the genotype test may be based on a SNP in one or more of the *MSTN*, *ACN9*, *PTPN1*, *PON1*, *ADHFE1*, or *GSN* genes. Details of some of the SNPs that may be used to predict the elite sprint race performance of a thoroughbred race horse are given in the appendices. It will be appreciated that the genotypic test may be based on a combination of any one or more of these SNPs.

In this non-limiting example, we studied the *MSTN* gene.

10 Methods

Association sample

The International Federation of Horseracing Authorities recognizes five distance categories: Sprint (5–6.5 f, $\leq 1,300$ m), Mile (6.51–9.49 f, 1,301–1,900 m), Intermediate (9.5–10.5 f, 1,901–2,112 m), Long (10.51–13.5 f, 2,114–2,716 m) and Extended (> 13.51 f, $> 2,717$ m) races (International Federation of Horseracing Authorities Classifications, www.horseracingintfed.com) [Note: 1 furlong = 1/8 mile = 201.2 meters]. For the case-control investigations we compared two cohorts: samples were subdivided into short (≤ 8 f and ≤ 7 f) and long (> 8 f) distance racing cohorts. To avoid animals with excessive consanguinity (within two generations) and over-representation of popular sires within the pedigrees, a set of Thoroughbred DNA samples ($n = 148$) was selected from a large DNA sample repository ($n > 1,000$) collected with informed owners' consent from Thoroughbred training, breeding and sales establishments in Ireland and New Zealand during 1998–2008.

Replication sample

To minimize non-genetic influences on performance the findings were validated by genotyping elite (Group and Listed race winning) racehorse samples ($n = 39$) selected from a repository of DNA samples ($n = 419$) from horses trained by the same trainer in Ireland during 2004–2008. A subset ($n = 142$) of this repository was evaluated for genotypic trends with parameters of racecourse success in two-year-old racehorses. Race records were derived from three sources: European race records, The Racing Post on-line database (www.racingpost.co.uk); Australasian and South East Asian race records, Arion Pedigrees (www.arion.co.nz); and North American race records: Pedigree Online Thoroughbred database (www.pedigreequery.com). The replication samples had some sharing of relatives, accounted for in the analyses.

DNA extraction, re-sequencing and genotyping

Genomic DNA was extracted from either fresh whole blood or hair samples using a modified version of a standard phenol/chloroform method (Sambrook & Russell 2001). Thirteen pairs of overlapping PCR primers were designed to cover the entire *MSTN* genomic sequence using the
5 PCR Suite extension to the Primer3 web-based primer design tool (Rozen & Skaletsky 2000; van Baren & Heutink 2004) (Table 13). Twenty-four unrelated Thoroughbred DNA samples were included in a re-sequencing panel to identify Thoroughbred-specific sequence variants. As such this study was powered to detect 95% of SNPs with MAF > 0.05 in the Thoroughbred population (Kruglyak & Nickerson 2001). Bidirectional DNA sequencing of PCR products was outsourced
10 to Macrogen Inc. (Seoul, Korea) and carried out using AB 3730xl sequencers (Applied Biosystems, Foster City, CA). Sequence variants were detected by visual examination of sequences following alignment using Consed version 19.0 (090206)[Gordon et al 1998]. Genotyping was carried out using Sequenom (San Diego, USA) iPLEX technology at Sequenom facilities in San Diego, USA (Association samples) and Hamburg, Germany (Replication
15 samples).

Statistical analyses

All statistical analyses, including tests of association were performed using PLINK Version 1.05 (Purcell et al 2007). Quality control analyses included computation of sample allele frequency,
20 percent missing genotypes and deviation from Hardy-Weinberg equilibrium. The series of case-control association tests were performed for two loci (g.66493737C>T and g.66494218A>C). Statistical significance was assessed using the Cochran-Armitage test for trend and an unconditioned genotypic model. Odds ratios and 95% CIs were calculated for the two most significant associations. The linear regression model was used to evaluate quantitative trait
25 association at locus g.66493737C>T using the phenotypes: best race distance and kg / cm ratio.

Results

Horses, in particular Thoroughbreds, have a very high muscle mass to body weight ratio (55%) compared to other mammalian species (30 - 40%) (Gunn 1987). Myostatin gene (*MSTN*) variants
30 have previously been shown to contribute to muscle hypertrophy; therefore, sequence variation in the equine *MSTN* gene, which contains three exons and spans 6,172 bp on chromosome 18 (reverse strand nt 66489608—66495780, EquCab2.0) was investigated. To-date, no sequence variants have been reported in genomic *MSTN* sequence in Thoroughbred horses and no *MSTN* SNPs are documented in the EquCab2.0 SNP database. Novel sequence variants were identified

by re-sequencing the equine *MSTN* gene in 24 unrelated Thoroughbred horses using 14 overlapping primer pairs (Table 13) spanning all three exons and 288 bp of the 5' upstream region. Although no exonic sequence variants were detected, six SNPs were detected in intron 1 of *MSTN* [nt 66492979—66494807] (Table 14).

5

Population genetic diversity analyses suggest that selection for the region containing the *MSTN* gene has been strong in the Thoroughbred population. Thirteen microsatellite loci spanning equine chromosome 18 were genotyped in three populations of unrelated Thoroughbred ($n = 106$), Akhal-Teke ($n = 18$), Connemara ($n = 17$) and Tuva ($n = 17$) horses. In Thoroughbred, evaluation of linkage disequilibrium indicated conserved haplotypes encompassing the two loci in closest proximity to the *MSTN* gene: *TKY101* (nt.63528459) and *TKY016* (nt.66838920). Among population differentiation (F_{ST}) was high at *TKY016* ($F_{ST} = 0.23$), which was among the top 10% of ($n = 394$) genome-wide loci when ranked by F_{ST} (Gu et al 2009). Interestingly, the highest F_{ST} score on chromosome 18 was for *TKY303* (nt.31.1 Mb; $F_{ST} = 0.31$), which is in close proximity to the *ACVR2A* gene encoding activin A receptor, type IIA, a key signaling molecule for myostatin. The high F_{ST} at *TKY016*, located 350 kb from *MSTN*, results from divergent allele frequency distributions among Thoroughbred and non-Thoroughbred populations and redistribution of Thoroughbred samples into distance cohorts ($TBE \leq 8$ f; $n = 25$ and $TBE > 8$ f $n = 22$) identified a significant difference (Pearson's chi-square test; $\chi^2 = 5.809$; $df = 1$; $P = 0.0159$) in allele 144 frequency ($TBE \leq 8$ f = 70%; $TBE > 8$ f = 45%).

To investigate associations between *MSTN* sequence variants and racing phenotypes we genotyped $n = 148$ Thoroughbred horses. Four of the six *MSTN* sequence polymorphisms displayed $MAF < 0.05$ in Thoroughbreds (Table 15) and were excluded from the association analyses. A series of population-based case-control investigations by separating the Thoroughbreds on the basis of retrospective racecourse performance into discrete cohorts containing unrelated animals (Table 16) was performed. Individual genotypes at the two SNPs used for the analyses (g.66493737C>T and g.66494218A>C) were not more common among elite Group race winning Thoroughbreds (Thoroughbred-elite, TBE) than horses that had never won a race (Thoroughbred-other, TBO) (Table 17). Also, no association was detected when handicap ratings, reflecting retrospective racing ability, were evaluated as a quantitative phenotype. However, considering the relative contribution of muscle power to sprint and longer distance racing, the elite Group race winning animals were subdivided into those that had won

30

their best (most valuable or highest grade) race over distances ≤ 8 f ($n = 51$) and those that had won their best race over distances > 8 f ($n = 35$) and found highly significant associations [Note: 1 furlong = 1/8 mile = 201.2 meters]. In Britain, of the 139 Group races per annum 57% are run over distances ≤ 1 mile and 43% are run over distances > 1 mile. The elite performer cohort contained a subset of animals ($n = 71$) that competed preferentially in short distance (≤ 8 f, $n = 39$) and long distance (> 8 f $n = 32$) races.

For all analyses the significance of association was consistently higher for g.66493737C>T than g.66494218A>C and the linkage disequilibrium between these SNPs was relatively high ($r^2 = 0.50$). Conditioning on each SNP using a logistic regression model identified an independent effect for g.66493737C>T on g.66494218A>C ($P = 0.0108$) but not for g.66494218A>C on g.66493737C>T ($P = 0.7388$) and therefore only the results for g.66493737C>T were considered further. Among the two distance cohorts we found a highly significant ($P = 3.70 \times 10^{-5}$) association with g.66493737C>T and this association became marginally stronger ($P = 1.88 \times 10^{-5}$) when the short distance cohort was further subdivided into animals ($n = 43$) that had won their best race over distances ≤ 7 f (Table 17a).

The C allele was twice as frequent in the short distance (≤ 7 f) than in the long distance (> 8 f) cohort (0.72 and 0.36 respectively) corresponding to an odds ratio of 4.54 (95% C.I. 2.23—9.23). When all Thoroughbreds were considered together the locus conformed to expected Hardy-Weinberg proportions (Table 18). However, there was a significant ($P = 0.0018$) deviation from HWE in the longer distance cohort, possibly due to selection at this locus. Conversely, the C/C genotype was the most common genotype among sprinters (≤ 7 f, $> 51\%$). Genotype trend effects were modeled by estimating the risk associated with a linear trend in magnitude of effect relative to the common homozygote, heterozygote, and rare homozygote genotype using the Cochran-Armitage test for the trend model. The most parsimonious model was the genotypic model ($P = 1.18 \times 10^{-6}$) indicating that genotypes are predictive of racing distance (Table 17b).

Table 13: PCR primer details for SNP discovery in the equine myostatin (MSTN) gene

Amplicon	Primer sequences (5')	Primer sequences (3')	PCR product size (bp)	Chr location (EquCab2.0)	No. of SNPs identified
MSTN_1	ATAAATGCAATTGT	CCATATGCAAGTT	399	chr18:66489320	-

	CTCAAAGTC (SEQ ID NO. 5)	TCCATTCC (SEQ ID NO. 6)		+66489718	
MSTN_2	TCAGCCATTCAGCC TATTTG (SEQ ID NO. 7)	ACGGTTGGCATT AACCATC (SEQ ID NO. 8)	422	chr18:66489629 +66490050	-
MSTN_3	GGAGACTTGCTTTC ATTTACCTG (SEQ ID NO. 9)	GAAGCTTTTGGAT GGGATTG (SEQ ID NO. 10)	552	chr18:66489914 +66490465	-
MSTN_4	CTCTGGGGTTTGCT TGGTG (SEQ ID NO. 11)	ACCTAGGGAATG GAGGATGG (SEQ ID NO. 12)	695	chr18:66490336 +66491030	-
MSTN_5	GAAGAGGAGGGAG GGAAGAG (SEQ ID NO. 13)	TTCAGTCTTCATG TGGTCTTGG (SEQ ID NO. 14)	762	chr18:66490908 +66491669	-
MSTN_7	AAGGTATTGTCATC TGCTTGG (SEQ ID NO. 15)	CCAAGACCAGGA GAAGATGG (SEQ ID NO. 16)	783	chr18:66491846 +66492628	-
MSTN_8	GCTTGTTAGCATAG GAAACTGG (SEQ ID NO. 17)	CTGAGACCCGTCA AGACTCC (SEQ ID NO. 18)	376	chr18:66492499 +66492874	-
MSTN_9	CGTCTTTCATGGGT TTGATG (SEQ ID NO. 19)	ATGTTCTCCACG GTGTCTC (SEQ ID NO. 20)	530	chr18:66492805 +66493334	1 (Indel)
MSTN_10	TGAAGGAATGAAC TGTGGATG (SEQ ID NO. 21)	GTCTGCGATCCTG CTTTACC (SEQ ID NO. 22)	580	chr18:66493261 +66493840	5
MSTN_11	TTTTGAAACTGTTG TGTCTG (SEQ ID NO. 23)	TCATAATTGCGTT TGGTTGC (SEQ ID NO. 24)	674	chr18:66493779 +66494452	1
MSTN_12	GCAAATGCTCAAA TGACCTAAAC (SEQ ID NO. 25)	TGTGCTGATTCTT GCTGGTC (SEQ ID NO. 26)	799	chr18:66494344 +66495142	-
MSTN_13	TGAAGATTAGTGT TTTGTCTCC (SEQ ID NO. 27)	CGAGATTCAATTGT GGAGCAG (SEQ ID NO. 28)	382	chr18:66495028 +66495409	-
MSTN_14	GAGACAACTTGCC ACACCAG (SEQ ID NO. 29)	TGCCCTGGTAATA ACAATGAAG (SEQ ID NO. 30)	786	chr18:66495287 +66496072	-

Table 14: Details of SNPs discovered in equine myostatin gene following resequencing in a panel of 24 unrelated Thoroughbred horses. None of these SNPs are among the SNPs in the EquCab2.0 SNP Map and have not been previously reported in any publically available literature.

SNP ID	Chr	EquCab2.0 SNP location	Allele 1	Allele 2	Gene structure	Amplicon	Substitution type
MSTN_9_383-386	chr18	66493222-66493225	delACTT		Intron 1	MSTN_9	

- 41 -

MSTN_10_227	chr18	66493525	T	G	Intron 1	MSTN_10	Transversion
MSTN_10_284	chr18	66493582	T	G	Intron 1	MSTN_10	Transversion
MSTN_10_439	chr18	66493737	T	C	Intron 1	MSTN_10	Transition
MSTN_10_447	chr18	66493745	A	G	Intron 1	MSTN_10	Transition
MSTN_10_477	chr18	66493775	A	G	Intron 1	MSTN_10	Transition
MSTN_11_404	chr18	66494218	A	C	Intron 1	MSTN_11	Transversion

Table 15: Genotyping results for *MSTN* SNPs

Assay (SNP_ID)	Coverage	NA.	Total	allele	COMMON	HET	RARE	p	q	F _p	F _q (MAF)	n
MSTN_66493525 (SEQ ID No. 33)	96.67%	5	145	2	138	5	2	281	9	0.969	0.031	290
MSTN_66493582 (SEQ ID No. 34)	92%	12	138	2	135	3	0	273	3	0.989	0.011	276
MSTN_66493737 (SEQ ID No. 31)	93.33%	10	140	2	42	75	23	159	121	0.568	0.432	280
MSTN_66493745 (SEQ ID No. 35)	97.33%	4	146	2	139	6	1	284	8	0.973	0.027	292
MSTN_66493775 (SEQ ID No. 36)	96.67%	5	145	2	139	5	1	283	7	0.976	0.024	290
MSTN_66494218 (SEQ ID No. 37)	91.33%	13	137	2	59	67	11	185	89	0.675	0.325	274

5

Table 16: Population summary including details of retrospective racecourse success for each cohort.

	n	No. sires	No. Males	No. Females	Mean RPR	Range RPR	Total no. races	Mean no. races	No. races won	No. Gr races won	No. Gr 1 races won	Mean no. Gr races won
TBE	86	86	37	49	115	87-134	1170	13.8	425	215	91	2.5
TBE > 8 f	35	35	12	23	119	107-134	-	-	-	89	42	-
TBE ≤ 8 f	51	51	25	26	114	87-129	-	-	-	129	49	-

TBE ≤ 7 f	43	43	20	23	113	87-129	-	-	-	76	23	-
TBO	62	62	22	40	59	21 - 89	537	8.7	15	0	0	0

Table 17: a. Case-control association test results for a series of cohort comparisons for g.66493737C>T. TBE: elite Group race winning Thoroughbreds; TBO: other non-winning Thoroughbreds; TBE > 8 f, TBE ≤ 8 f and TBE ≤ 7 f: elite Group race winning Thoroughbreds that won their best (most valuable or highest grade) races over distances > 8 f, ≤ 8 f and ≤ 7 f. In each case the frequency of the g.66493737-T allele is given. Odds ratios were calculated for the two most significant results. **b.** Best-fit model results for g.66493737C>T association with elite Group race winning performance over distances ≤ 7 f.

a.	Pop 1 vs Pop 2	Freq T Pop 1	Freq T Pop 2	CHISQ	P	OR
	TBE vs TBO	0.443	0.425	0.09	0.764	-
	TBE > 8 f vs TBE ≤ 8 f	0.641	0.309	17.02	3.70E-05	3.996
	TBE > 8 f vs TBE ≤ 7 f	0.641	0.282	18.31	1.88E-05	4.538
	TBE > 8 f vs TBO	0.641	0.425	7.76	0.005	-
	TBE ≤ 8 f vs TBO	0.309	0.425	3.06	0.080	-
	TBE ≤ 7 f vs TBO	0.282	0.425	4.15	0.042	-
b.		TBE > 8 f	TBE ≤ 7 f	P		
	Genotypic (C/C, C/T, T/T)	0/23/9	21/23/3	1.18E-06		
	Trend (C, T)	23/41	65/29	5.23E-06		

10 **Table 18** Hardy-Weinberg equilibrium test results for locus g.66493737C>T.

TEST	A1	A2	GENO	O(HET)	E(HET)	P
ALL	T	C	23/75/41	0.5396	0.4916	0.3022
TBE <7 f	T	C	3/16/20	0.4103	0.4050	1
TBE >8 f	T	C	9/23/0	0.7188	0.4604	0.0018

MSLN SNP (MSLN_66493737 (T/C) SNP)

This SNP is located on Chromosome 18 of *Equus caballus* at position 66,490,208-66,495,180 reverse strand of the Horse Genome Sequence (*Equus caballus* Version 2.0) which can be viewed at www.broad.mit.edu/mammals/horse/.

The horse genome EquCab2 assembly is a Whole Genome Shotgun (WGS) assembly at 6.79x and was released in September 2007. A female Thoroughbred named "Twilight" was selected as the representative horse for genome sequencing. The project coordination and genome sequencing and assembly is provided by the Broad Institute. The N50 size is the length such that 50% of the assembled genome lies in blocks of the N50 size or longer. The N50 size of the

contigs is 112.38 kb, and the total length of all contigs is 2.43 Gb. When the gaps between contigs in scaffolds are included, the total span of the assembly is 2.68 Gb. The horse EquCab2 was annotated using a standard Ensembl mammalian pipeline. Predictions from vertebrate mammals as well as horse proteins have been given priority over predictions from non-vertebrate mammals. The set of predictions was been compared to 1:1 homologues genes in human and mouse, and missing homologs in the horse annotation have been recovered using exonerate. Horse and human cDNAs have been used to add UTRs to protein based predictions. The final gene-set comprises 20,737 protein-coding genes, 2,863 identified as pseudogenes and 1,580 classified as retro-transposed genes.

10

Further details of the SNP are as follows:

- SNP: MSTN_66493737 (T/C) $P = 1.88E-05$ OR = 4.54
- EquCab2.0 SNP_ID: not detected in EquCab2.0 database. No report of SNP in on-line bioinformatics resources.
- 15 ○ In house SNP_ID: MSTN-10_439
- Genomic location of SNP: Intron 1
- Substitution type: Transition

The flanking sequence and SNP (bold and square brackets) is as follows:

20 AGCTAAGCAAGTAATTAGCACAAAAATTTGAATGTTATATTCAGGCTATCTCAAAAG
 TTAGAAAATACTGTCTTTAGAGCCAGGCTGTCATTGTGAGCAAAATCACTAGCAATT
 TCTTTTATTTTGGTTCCCCAAGATTGTTTATAAATAAGGTAAATCTACTCCAGGACTA
 TTTGATAGCAGAGTCATAAAGGAAAATTA[T/C]TTGGTGCATTATAACCTGATTACTT
 AATAAGGAGAACAATATTTTGAACTGTTGTGTCCTGTTTAAAGTAGATAAAGCACT
 25 GGGTAAAGCAGGATCGCAGACACATGGCACAGAATCTTCCGTGTCATGCCTTCTCTG
 TGAAGGTGTCTGTCTCCCTTTCCCTTGAGTGTAGTTATGAACTGACTGCAAAAAGAAT
 ATATG

(SEQ ID No. 31)

30

Considering best race distance (BRD) as a quantitative trait, we analyzed the data for the elite cohort using the distance (furlongs) of the highest grade or most valuable Group race won as the phenotype ($n = 79$). BRD was highly significantly associated ($P = 4.85 \times 10^{-8}$) with the g.66493737C>T SNP (Table 19). This result was independently validated ($P = 0.0047$) in a

cohort of 37 elite racehorses ($n = 27$ Group race winners and $n = 10$ Listed race winners) produced by the same trainer. For each genotype we determined the mean BRD (Table 20): C/C mean = 6.2 ± 0.8 f; C/T mean = 9.1 ± 2.4 f; and T/T mean = 10.5 ± 2.7 f. A distribution of the genotypes in two furlong increments is shown in Figs 6A to 6. It is important to note that a bias may be introduced to these distances as two-year-old Group races are limited to ≤ 8 f in Ireland and the United Kingdom (there are only three Group races for two-year-olds in Europe > 8 f). Therefore we replaced BRD for horses that had won their only Group race as two-year-olds with the average distance of their three-year-old races ($n = 73$), which resulted in a marginal increase in the means for the three genotypes (C/C mean = 6.4 ± 1.0 f; C/T mean = 9.7 ± 2.0 f; and T/T mean = 10.9 ± 2.4 f) and an increase in the significance of association ($P = 5.45 \times 10^{-9}$) (Fig. 7).

Eight National Hunt (races over obstacles and distances 16 - 36 f) racehorses were also genotyped for the g.66493737C>T SNP and the results support an association of the T allele with stamina (T/T, $n = 7$; and C/T, $n = 1$). Also, the genotype frequencies among a non-Thoroughbred population known for endurance exercise capabilities ($n = 31$, Egyptian Arabian horse) were considerably different to the Thoroughbred population (Fig. 8). Together these findings indicate that the C/C genotype is particularly suited to sprint racing.

In Thoroughbred breeding considerable weight is given to the contribution of the sire in the predicted best race distance of offspring. For breeding stallions a 'Stamina Index' (S.I.) is estimated as the average winning distance of all racing progeny. Therefore we investigated the distribution of genotypes for the $n = 19$ unrelated breeding stallions with S.I. in our sample (Fig. 9). All (100%) stallions with S.I. = 6 - 8 f had the C/C genotype; 83.3% and 75% stallions with S.I. = 9 - 10 f and 10 - 12 f respectively were C/T; and 25% stallions with S.I. = 10 - 12 f were T/T. While the sample size is small there is a clear indication that g.66493737C>T genotypes are predictive of S.I. in breeding stallions (Fig. 9).

These data indicate that genotypic information at this locus may have practical applications in the Thoroughbred horse racing and breeding industry. To evaluate this further, two-year-old racing form for $n = 142$ horses-in-training with the same trainer during 2007 and 2008 ($n = 63$, 2007; $n = 79$, 2008) (Table 20a) was investigated. Consistently, for each parameter of racing success, C/C and C/T genotypes were more successful two-year-old racehorses than T/T animals (Table 20b). In terms of earnings, the greatest returns on training investment were for animals

that were C/C or C/T; on average these horses earned 5.5-fold more than T/T horses. Even when individuals that had won > Sterling£100,000 (US\$165,000) were excluded, on average C/C individuals earned 1.6-fold more than T/T individuals. The bulk of keeping and training expenses are not returned in prize money (72% Ireland, 78% United Kingdom for horses that have run in at least one race) [International Federation of Horseracing Authorities, www.horseracingintfed.com]; therefore, employing a strategy to train and race only C/C and C/T individuals as two-year-olds may be beneficial.

To eliminate potential confounding effects of shared sires, the racing successes of 41 half-sibs (progeny of a single sire) [C/T, $n = 22$; T/T, $n = 19$] (Table 20c) that were trained by the same trainer as two-year-olds was investigated. A significant genotype association with racing performance (Pearson's chi-square test: $\chi^2 = 7.235$; $df = 1$; $P = 0.0071$) was identified; five of the progeny were two-year-old Group race winners and all displayed the C/T genotype.

In many instances the goal of breeders is to breed a Derby winner. The Derby distance (12 f) predicts that individuals must have at least one copy of the T allele at g.66493737C>T. There were $n = 7$ Derby winners in our sample: C/T $n = 6$; and T/T $n = 1$. Furthermore, $n = 51$ progeny from a highly successful commercial breeding stallion that had won both the Epsom Derby and the Irish Derby were genotyped and had a S.I. = 11.3 f. Among the progeny just two genotypes ($n = 29$, C/T; and $n = 22$, T/T) were identified suggesting that this individual (while not genotyped here) is T/T (both the sire and dam were genotyped: sire: T/T; dam: C/T). We estimated the mean BRD for the genotypes in $n = 9$ of the stallion's progeny that had won Group races, further reinforcing the g.66493737C>T genotype trend (C/T: $n = 6$, mean BRD = 8 f; T/T: $n = 3$, mean BRD = 10.7 f).

Similar to their human counterparts, sprint racing Thoroughbreds are generally more compact and muscular than horses suited to longer distance races. Therefore, to investigate whether *MSTN* genotypes influence body mass, mass (kg) and height at withers (cm) measurements that were taken during two two-year-old racing seasons for $n = 97$ ($n = 37$ males, $n = 60$ females) horses-in-training with the same trainer were used. Mass to height ratio displayed a significant ($P = 0.0147$) relationship with g.66493737C>T genotype (2.94 kg/cm, C/C; 2.88 kg/cm, C/T; and 2.83 kg/cm, T/T). This association became stronger when males were considered independently

($P = 0.0025$) of females ($P = 0.2272$) [Table 21]. On average C/C males had 6.7% (*i.e.* 3.033 kg/cm versus 2.843 kg/cm) greater mass per cm than T/T males.

Table 19: Quantitative trait association tests and best race distance (BRD) means for **a.** association test sample; **b.** association test sample using mean three year old distances as phenotype (for two-year-olds that won their best race ≤ 8 f) ; and **c.** replication sample.

	<i>n</i>	Quantitative association test results					Best race distance means			
		<i>BETA</i>	<i>SE</i>	<i>R2</i>	<i>T</i>	<i>P</i>	GENO	C/C	C/T	T/T
a.	79	2.308	0.381	0.322	6.052	4.85E-08	COUNTS	21	46	12
							FREQ	0.266	0.582	0.152
							MEAN	6.167	9.087	10.540
							SD	0.827	2.365	2.742
b.	73	2.390	0.360	0.383	6.635	5.46E-09	COUNTS	19	42	12
							FREQ	0.260	0.575	0.164
							MEAN	6.421	9.682	10.930
							SD	1.022	2.081	2.441
c.	37	-1.500	0.497	0.207	-3.021	0.005	COUNTS	7	23	7
							FREQ	0.189	0.622	0.189
							MEAN	6.714	8.217	9.714
							SD	1.704	1.930	1.890

Table 20: Parameters of two-year-old racing (Ireland and United Kingdom) success for $n = 142$ horses-in-training with the same trainer during 2007 and 2008. *a.* Two year old horses-in-training 2007 & 2008; *b.* two year old horses- in-training 2007 & 2008 comparing C/C and C/T versus T/T genotypes; *c.* Half-sib two year old horses-in-training sharing a single sire.

<i>a.</i>	<i>n</i>	no. runners	no. winners	total no. races	total no. races won	% runners	% winners to runners	% wins to runners	% winners to total	% wins to runs	mean no. races per runner	total earnings (£)	mean earnings (£)	mean earnings excl. earnings >£100k	no. earnings >£100k
CC	40	21	11	87	17	52.5	52.4	81.0	27.5	19.5	4.1	51114	20440	8203	1
CT	67	32	18	115	26	47.8	56.3	81.3	26.9	22.6	3.6	1801103	36968	4925	5
TT	35	13	6	40	6	37.1	46.2	46.2	17.1	15.0	3.1	87461	5175	5175	0
b.															
CC/ CT	10 7	53	29	202	43	49.5	54.7	81.1	27.1	21.3	3.8	2312217	28704	6564	6
TT	35	13	6	40	6	37.1	46.2	46.2	17.1	15.0	3.1	87461	5175	5175	0
c.															
CT	22	12	9	46	18	54.5	75.0	150.0	40.9	39.1	3.8	1620087	73640	-	6
TT	19	9	5	23	5	47.4	55.6	55.6	26.3	21.7	2.6	67864	3572	-	0

Table 21 Quantitative association test results for g.66493737C>T with kg/cm ratio as phenotype

	<i>n</i>	Quantitative association test results				<i>P</i>	Kg/cm means			
		<i>BETA</i>	<i>SE</i>	<i>R2</i>	<i>T</i>		GENO	C/C	T/C	T/T
Two year olds-in-training	97	-								
		0.05671	0.02282	0.06104	-2.485	0.015	COUNTS	29	47	21
							FREQ	0.299	0.485	0.217
							MEAN	2.939	2.875	2.826
							SD	0.155	0.162	0.168
Males only	37	-								
		0.09575	0.02941	0.2325	-3.256	0.003	COUNTS	10	18	9
							FREQ	0.270	0.487	0.243
							MEAN	3.033	2.918	2.843
							SD	0.169	0.101	0.134
Females only	60	-								
		0.03773	0.03091	0.02505	-1.221	0.227	COUNTS	19	29	12
							FREQ	0.317	0.483	0.200
							MEAN	2.889	2.848	2.814
							SD	0.124	0.187	0.194

5

Risk prediction

Based on subpopulation prediction using population observed genotype frequencies and Bayes Theorem the probability of being a member of one or other subpopulation (elite sprinter or elite endurance or 'stayer') given a certain genotype at the *MSTN* gene was investigated. Results are

10 give in Table 22 and are provided as a percentage chance of being a member of each of the two subpopulations.

Table 22 - Predictive test for Elite Sprint racing ability

15

				TBE SP						TBE EN					
				Population 1: allele, genotype frequencies						Population 2: allele, genotype frequencies					
				A		AA	AB	BB	P(G C)	A	AA	AB	BB	P(G C)	
<i>Locus</i>															
MSTN		1		0.28		0.08	0.41	0.51	0.41	0.64	0.28	0.72	0	0.72	
MSTN	TT	TC	CC			P(G C)			0.41					0.72	
						P(C)			0.5					0.5	
						P(G C) P(C)			0.21					0.36	

[illegible]

In which AA represents genotype TT, AB represents genotype CT, and BB represents genotype CC

From Table 22 it can be seen that subjects with the genotype CC in MSTN_66493737 (T/C) SNP
5 have the greatest chance of being sprinters given that they are elite Thoroughbreds.

MSTN gene expression

MSTN mRNA expression in two independent real-time qRT-PCR assays (Table 23) has been investigated in resting skeletal muscle (*gluteus medius*) from biopsy samples that had been collected for $n = 60$ untrained yearlings (C/C, $n = 15$; C/T, $n = 28$; T/T, $n = 17$).

Table 23: Primer sequences for qRT-PCR assays for MSTN gene expression and TTN reference gene expression

Primer Name	Target Gene	Location	Sequence
TTN_FOR	Titin (TTN)	Exon 357	gcatgacacaactggaaagc (SEQ ID No. 38)
TTN_REV	Titin (TTN)	Exon 357	aactttgccctcatcaatgc (SEQ ID No. 39)
MSTN1-2_FOR	Myostatin (MSTN)	Exon 1	tgacagcagtgatggctctt (SEQ ID No. 40)
MSTN1-2_REV	Myostatin (MSTN)	Exon 2	ttgggttttcctccacttg (SEQ ID No. 41)
MSTN2-3_FOR	Myostatin (MSTN)	Exon 2	ttcccaagaccaggagaaga (SEQ ID No. 42)
MSTN2-3_REV	Myostatin (MSTN)	Exon 3	cagcatcgagattctgtgga (SEQ ID No. 43)

15

We found a significant association with genotype for the MSTN 66493737 (T/C) SNP ($P = 0.003559$). The C/C genotype cohort had higher *MSTN* mRNA levels (654.3 ± 354.3 ; 613.7 ± 327.0) than either of the C/T (405.7 ± 234.1 ; 368.6 ± 213.6) and T/T (350.1 ± 185.5 ; 348.1 ± 167.2) cohorts (Fig. 10).

20

It was also found that MSTN gene expression is significantly down-regulated (-4.2-fold, $P = 0.0043$) following a period of training. In the Thoroughbred horse skeletal muscle transcriptome

the greatest reduction in gene expression following a period of training is MSTN gene expression.

Quantitative association with best race distance

- 5 It was examined whether there was a significant relationship between SNPs in the examined genes and best race distance (i.e. distance of the highest quality / most valuable Group race won) in a quantitative association test analysis using the subset of individuals that had won a Group race (i.e. TBE n = 86). Table 24 below shows the SNPs significantly associated with best race distance.

10

Table 24 – SNPs associated with best race distance

CHR	SNP	STAT	EMP1	NP
18	MSTN_66493737	36.63	1.00E-06	1000000
18	MSTN_66494218	15.97	0.0001744	137586
22	COX4I2_22684844	6.495	0.01146	2094
22	COX4I2_22684390	5.783	0.02526	949
22	PTPN1_38585796	4.963	0.0377	609
4	PON1_38697145	4.596	0.03938	583
22	PTPN1_38597033	4.64	0.04406	521
22	COX4I2_22684676	4.51	0.04742	484

- Table 25 below shows the mean best race distance for each genotype for four of the SNPs from
15 Table 24 above.

Table 25 – Mean best race distance for each genotype

CHR	SNP	VALUE	G11	G12	G22
18	MSTN_66493737	GENO	T/T	T/C	C/C
18	MSTN_66493737	COUNTS	12	46	21
18	MSTN_66493737	FREQ	0.1519	0.5823	0.2658
18	MSTN_66493737	MEAN	10.54	9.087	6.167
18	MSTN_66493737	SD	2.742	2.365	0.8266
18	MSTN_66494218	GENO	C/C	C/A	A/A
18	MSTN_66494218	COUNTS	8	39	31
18	MSTN_66494218	FREQ	0.1026	0.5	0.3974
18	MSTN_66494218	MEAN	10.56	9.179	7.403
18	MSTN_66494218	SD	2.872	2.48	2.043
22	COX4I2_22684844	GENO	C/C	C/T	T/T

22	COX4I2_22684844	COUNTS	4	39	40
22	COX4I2_22684844	FREQ	0.04819	0.4699	0.4819
22	COX4I2_22684844	MEAN	11.12	8.91	7.975
22	COX4I2_22684844	SD	2.097	2.762	2.247
22	COX4I2_22684390	GENO	C/C	C/T	T/T
22	COX4I2_22684390	COUNTS	4	44	32
22	COX4I2_22684390	FREQ	0.05	0.55	0.4
22	COX4I2_22684390	MEAN	10.62	8.943	7.875
22	COX4I2_22684390	SD	2.056	2.783	2.254

Wherein, at MSTN_66493737-(T/C) the TT genotype has a mean best race distance of 10.54 furlongs (f), the TC genotype has a mean best race distance of 9.087 f and the CC genotype has a mean best race distance of 6.167 f. Overall the mean best race distance was 8.55 f.

5

There are many practical applications for the genotypic test based on the MSTN genotype. These include:

1. Young stock (foals and yearlings)

10 *Informed selection and sales decisions can be made to:*

- identify sprinters
- identify middle-distance / potential Derby winners with speed
- identify individuals with enhanced stamina

15 2. Horses-in-training

Operating costs can be reduced and racing strategy can be fine tuned by:

- identifying the most precocious two-year olds
- horses can be trained and raced for optimal racing distance

20 3. Broodmares

Breeding outcomes can be optimised by:

- focusing on optimal breeding mares
- selecting compatible stallions

25 4. Stallions

A stallions potential can be promoted by:

- predicting stamina index for young stallions (5 year advantage)

- attracting compatible mares to enhance stallion profile

For example, for the MSTN-66493737 TIC SNP for foals, young stock and horses-in-training selection of individuals may be made for individuals most likely to perform well as two year olds (C/C and C/T) and against 'backward' individuals (industry terminology for less physically developed young Thoroughbreds) that may benefit from waiting to race until they are three years old (T/T). Breeding objectives may be more confidently met by selecting C/C individuals for short distance racing, C/T individuals for middle-distance racing and T/T individuals for racing requiring greater stamina. For stallion owners, prediction of a stallion's genetic stamina index at the outset of a stud career (five years are required to estimate S.I. from retrospective three year old progeny racing performance) will immediately enhance a young stallion's profile and promote their genetic potential to mare owners. This in turn will enable mare owners, with targeted breeding strategies, to better select stallions to achieve specific breeding objectives. To eliminate uncertainty from a mating outcome (unless both sire and dam are homozygous) it will be necessary to genotype the foal, enabling selection of individuals for a targeted breeding outcome.

Example 6 – Detecting SNPs

SNPs may be determined by any SNP genotyping method including for example the following non limiting methods:

Sequenom Inc

The iPLEX® Gold assay based on multiplex PCR followed by a single base primer extension reaction. After the PCR, remaining nucleotides are deactivated by SAP treatment. The single base primer extension step is performed, and the primer extension products analyzed using MALDI TOF MS.

KBiosciences

KBiosciences uses both its own novel form of competitive allele specific PCR system (KASPar) and Taqman™ chemistries for genotyping. KASPar assays are a proprietary in-house system developed to replace the previously used Amplifluor system.

Taqman assays

TaqMan® SNP Genotyping Assays make it easy to perform SNP genotyping with the precision of TaqMan® reagent-based chemistry. TaqMan® Assays provide SNP detection capabilities. Also, the TaqMan® Sample-to-SNP™ Kit provides a streamlined protocol for performing TaqMan chemistry-based genotyping analysis from any sample with a single kit. The kit is
5 comprised of two parts: the DNA Extract All Lysis Reagents and the TaqMan® GTXpress™ Master Mix. The DNA Extract All Lysis Reagents reduce prolonged procedures for the release of real-time PCR-ready DNA to a 5 minute protocol. They can process a wide variety of samples ranging from blood to buccal swabs. The TaqMan GTXpress Master Mix enables robust PCR amplification in less than 50 minutes.

10

Industrial application

The predictive tests described herein may be applied to select individuals with high or low genetic potential for racing success. These tests can be performed on an individual at any stage in the life cycle e.g. Day 1 (birth), prior to sales (i.e. yearlings, 2 year olds etc), during racing career
15 (i.e. from 2 years old), during breeding (i.e. up to approx 25 years). Also, the tests may be applied to select appropriate stallion - mare matches for mating based on the genetic make-up of mare and stallion.

The invention is not limited to the embodiment hereinbefore described, with reference to the
20 accompanying drawings, which may be varied in construction and detail.

References

- Ballard J.W. & Dean M.D. (2001) The mitochondrial genome: mutation, selection and recombination. *Curr Opin Genet Dev* 11, 667-72.
- 5 Blier P.U., Dufresne F. & Burton R.S. (2001) Natural selection and the evolution of mtDNA-encoded peptides: evidence for intergenomic co-adaptation. *Trends Genet* 17, 400-6.
- Bray MS, Hagberg JM, Pérusse L, Rankinen T, Roth SM, Wolfarth B, Bouchard C. The human
10 gene map for performance and health-related fitness phenotypes: the 2006-2007 update. *Med Sci Sports Exerc.* 2009 Jan;41(1):35-73.
- Cunningham EP, Dooley JJ, Splan RK, Bradley DG. Microsatellite diversity, pedigree relatedness and the contributions of founder lineages to Thoroughbred horses. *Anim Genet.* 2001
15 Dec;32(6):360-4.
- Das J. (2006) The role of mitochondrial respiration in physiological and evolutionary adaptation. *Bioessays* 28, 890-901.
- 20 Dempsey and Wagner 1999 Exercise-induced arterial hypoxemia. *J Appl Physiol.* 1999 Dec;87(6):1997-2006. Review. PMID: 10601141
- Suzanne S. Eivers, Beatrice A. McGivney, Rita G. Fonseca, David E. MacHugh, Katie Menson, Stephen D. Park, Jose-Luis L. Rivero, Cormac T. Taylor, Lisa M. Katz and Emmeline W. Hill*
25 Exercise-induced skeletal muscle gene expression in unconditioned and conditioned Thoroughbred horses and associations with physiological variables. *Physiological Genomics*, In Preparation (2009)
- Martin Flück 2006 Functional, structural and molecular plasticity of mammalian skeletal muscle
30 in response to exercise stimuli. *The Journal of Experimental Biology* 209, 2239-2248

Fukuda R, Zhang H, Kim JW, Shimoda L, Dang CV, Semenza GL. HIF-1 regulates cytochrome oxidase subunits to optimize efficiency of respiration in hypoxic cells. *Cell*. 2007 Apr 6;129(1):111-22.

- 5 | Gordon D, Abajian C, Green P. Consed: a graphical tool for sequence finishing. *Genome Res*. 1998 Mar;8(3):195-202.

Gramkow and Evans 2006 Gramkow HL, Evans DL. Correlation of race earnings with velocity at maximal heart rate during a field exercise test in Thoroughbred racehorses. *Equine Vet J* 10 Suppl. 2006 Aug;(36):118-22. PMID: 17402405

Gu J, Orr N, Park SD, Katz LM, Sulimova G, MacHugh DE, Hill EW. A genome scan for positive selection in thoroughbred horses. *PLoS One*. 2009 Jun 2;4(6):e5767.

- 15 Gunn HM. Muscle, bone and fat proportions and muscle distribution of thoroughbreds and quarter horses. In: Gillespie JR, Robinson NE eds. *Equine exercise physiology 2*. Davis, CA: ICEEP; 1987:253-264.

Harkins et al., 1993 Harkins JD, Hackett RP, Ducharme NG. Effect of furosemide on physiologic 20 variables in exercising horses. *Am J Vet Res*. 1993 Dec;54(12):2104-9. PMID: 8116946

Hoppeler and Vogt, 2001 Muscle tissue adaptations to hypoxia. *J Exp Biol*. 2001 Sep;204(Pt 18):3133-9. Review. PMID: 11581327

- 25 Love S, Wyse CA, Stirk AJ, Stear MJ, Calver P, Voute LC, Mellor DJ. Prevalence, heritability and significance of musculoskeletal conformational traits in Thoroughbred yearlings. *Equine Vet J*. 2006 Nov;38(7):597-603. PMID: 17228572

Matoba S, Kang JG, Patino WD, Wragg A, Boehm M, Gavrilova O, Hurley PJ, Bunz F, Hwang 30 PM. p53 regulates mitochondrial respiration. *Science*. 2006 Jun 16;312(5780):1650-3. Epub 2006 May 25.

Meiklejohn C.D., Montooth K.L. & Rand D.M. (2007) Positive and negative selection on the mitochondrial genome. *Trends Genet* 23, 259-63.

- Mosher DS, Quignon P, Bustamante CD, Sutter NB, Mellersh CS, Parker HG, Ostrander EA. A mutation in the myostatin gene increases muscle mass and enhances racing performance in heterozygote dogs. *PLoS Genet.* 2007 May 25;3(5):e79. Epub 2007 Apr 30
- 5 Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MAR, Bender D, Maller J, Sklar P, de Bakker PIW, Daly MJ & Sham PC (2007) PLINK: a toolset for whole-genome association and population-based linkage analysis. *American Journal of Human Genetics*, 81.
- 10 Revington M. Haematology of the racing Thoroughbred in Australia 2: haematological values compared to performance. *Equine Vet J.* 1983 Apr;15(2):145-8. PMID: 6873047
- Rivero JL, Ruz A, Martí-Korff S, Estepa JC, Aguilera-Tejero E, Werkman J, Sobotta M, Lindner A. Effects of intensity and duration of exercise on muscular responses to training of Thoroughbred racehorses. *J Appl Physiol.* 2007 May;102(5):1871-82. Epub 2007 Jan 25. PMID: 17255370
- 15 Saleem A, Adhihetty PJ, Hood DA. Role of p53 in mitochondrial biogenesis and apoptosis in skeletal muscle. *Physiol Genomics.* 2009 Mar 3;37(1):58-66. Epub 2008 Dec 23. Links
- 20 Sambrook, J. and D. Russell (2001). *Molecular Cloning; A Laboratory Manual*, Cold Spring Harbor Laboratory.
- Seaman J, Erickson BK, Kubo K, Hiraga A, Kai M, Yamaya Y, Wagner PD. Exercise induced ventilation/perfusion inequality in the horse. *Equine Vet J.* 1995 Mar;27(2):104-9. PMID: 7607141
- 25 Taylor CT, Colgan SP. Therapeutic targets for hypoxia-elicited pathways. *Pharm Res.* 1999 Oct;16(10):1498-505. Review. PMID: 10554089
- 30 van Deursen et al. 1993 Skeletal muscles of mice deficient in muscle creatine kinase lack burst activity *Cell* 74: 621-631

Weber K, Brück P, Mikes Z, Küpper JH, Klingenspor M, Wiesner RJ. Glucocorticoid hormone stimulates mitochondrial biogenesis specifically in skeletal muscle. *Endocrinology*. 2002 Jan;143(1):177-84.

- 5 Yang Q, Khoury MJ, Botto L, Friedman JM, Flanders WD. Improving the prediction of complex diseases by testing for multiple disease-susceptibility genes. *Am J Hum Genet*. 2003 Mar;72(3):636-49. Epub 2003 Feb 14.

- Young LE, Rogers K, Wood JL. Left ventricular size and systolic function in Thoroughbred
10 racehorses and their relationships to race performance. *J Appl Physiol*. 2005 Oct;99(4):1278-85. Epub 2005 May 26. PMID: 15920096

CHR	SNP	BP	A1	F_A	F_U	A2	CHISQ	P	OR
4	PDK4_38973231	3924	A	0.4639	0.2823	G	9.874	0.001676	2.2
4	PDK4_38968139	0	T	0.4146	0.582	C	7.842	0.005106	0.5088
4	PDK4_38969307	1168	A	0.4304	0.2712	C	7.409	0.006488	2.031
10	CKM_15884567	2716	G	0.07407	0.1636	A	5.355	0.02066	0.4089
18	MSTN_66493525	0	G	0.01205	0.05833	T	4.896	0.02692	0.1969
22	COX4I2_22684390	1164	C	0.325	0.4545	T	4.654	0.03098	0.5778
22	COX4I2_22684676	286	C	0.3415	0.4655	T	4.384	0.03628	0.5953
22	COX4I2_22683226	6865	T	0.3636	0.4828	G	3.868	0.04922	0.6122
22	ACSS1_759076	0	G	0.2317	0.1379	C	3.838	0.05009	1.885
3	COX4I1_32772871	0	T	0.3642	0.2586	C	3.462	0.0628	1.642
9	ADHFE1_18802749	66	A	0.03659	0.08197	T	2.728	0.0986	0.4253
18	MSTN_66493775	30	G	0.01205	0.04167	A	2.559	0.1097	0.2805
25	PTGS1_26007699	2168	C	0.1325	0.2016	T	2.494	0.1143	0.605
25	PTGS1_25991437	1489	C	0.1098	0.175	T	2.49	0.1146	0.5812
22	ACSS1_780613	12338	C	0.2625	0.3475	T	2.341	0.126	0.6685
1	ACTN2_74842283	0	G	0.08537	0.1417	A	2.259	0.1329	0.5655
25	GSN_25024464	0	T	0.09259	0.15	A	2.199	0.1381	0.5782
25	GSN_25028755	4291	A	0.0875	0.1441	G	2.193	0.1386	0.5697
10	CKM_15887661	3094	C	0.3025	0.386	T	2.088	0.1485	0.6899
1	GGPS1_76001872	0	A	0.439	0.3559	C	1.967	0.1607	1.416

22	PTPN1_38591965	1580	G	0.3025	0.3814	A	1.905	0.1675	0.7034
4	ACN9_40285196	5470	G	0.3861	0.3083	T	1.806	0.179	1.411
1	TOMM20_76186624	1936	C	0.2062	0.2727	T	1.61	0.2044	0.6929
4	ACN9_40267593	0	A	0.02907	0.008065	T	1.601	0.2058	3.683
18	MSTN_66493745	8	G	0.01765	0.04237	A	1.577	0.2092	0.406
21	PRKAA1_25374247	9959	A	0.04321	0.01667	G	1.572	0.2099	2.665
22	COX4I2-5900_22676361	0	C	0.4259	0.5	T	1.495	0.2215	0.7419
9	ADHFE1_18793538	5477	C	0.142	0.1949	T	1.394	0.2378	0.6835
9	ADHFE1_18802683	9145	G	0.05696	0.09322	A	1.321	0.2504	0.5876
1	GGPS1_76002021	149	C	0.4304	0.3644	A	1.223	0.2688	1.318
10	CKM_15881851	0	A	0.06329	0.0339	G	1.212	0.2709	1.926
18	MSTN_66494218	443	C	0.3526	0.2895	A	1.193	0.2747	1.337
1	ACTN2_74900867	19039	T	0.0625	0.09821	A	1.179	0.2775	0.6121
25	TNC_19737599	6101	C	0.4145	0.4815	G	1.149	0.2837	0.7623
22	PTPN1_38585796	0	G	0.1975	0.25	C	1.106	0.2929	0.7385
1	MYEF2_141647593	20636	T	0.1456	0.1034	C	1.065	0.302	1.477
9	MTFR1_19456942	17072	A	0.04819	0.07627	T	0.9663	0.3256	0.6132
1	MYEF2_141651362	394	G	0.141	0.1017	A	0.9562	0.3282	1.45

9	MTFR1_19472498	15556	A	0.05422	0.08333	G	0.9521	0.3292	0.6306
24	HIF1A_8984849	4922	G	0.1582	0.2034	A	0.9436	0.3314	0.7362
25	GSN_25033440	4685	G	0.3916	0.3361	A	0.9313	0.3345	1.271
1	ACTN2_74853540	11257	T	0.2785	0.2288	G	0.8721	0.3504	1.301
1	ACTN2_74872377	18837	G	0.09494	0.1293	T	0.8106	0.3679	0.7063
25	TNC_19737816	217	A	0.425	0.475	G	0.6937	0.4049	0.8169
9	ADHFE1_18787798	2714	G	0.03205	0.05172	A	0.6635	0.4153	0.6071
22	MC3R-530_43059660	0	C	0.2692	0.3148	T	0.6469	0.4212	0.8019
1	MYEF2_141626957	0	C	0.1386	0.1083	T	0.5781	0.447	1.324
1	TOMM20_76184688	0	T	0.3253	0.2833	A	0.5759	0.4479	1.22
25	PTGS1_25989948	0	C	0.1392	0.1724	T	0.5672	0.4514	0.7765
25	TNC_19741797	3981	C	0.3036	0.2564	G	0.5027	0.4783	1.264
25	TNC_19716930	0	G	0.09375	0.1167	A	0.3879	0.5334	0.7833
1	MYEF2_141650968	3375	C	0.1341	0.1102	T	0.3629	0.5469	1.251
4	PON1_38681590	784	T	0.08537	0.06667	C	0.3391	0.5604	1.307
25	NDUFA8_25799774	680	T	0.5064	0.4712	C	0.3103	0.5775	1.152
21	PRKAA1_25364288	0	A	0.03659	0.025	G	0.3031	0.5819	1.481
4	PON1_38680806	0	A	0.1098	0.09016	G	0.2947	0.5872	1.244
25	NDUFA8_25799094	3431	G	0.4661	0.5	T	0.2544	0.614	0.873
9	MTFR1_19439870	129	G	0.3072	0.2807	A	0.228	0.633	1.136

1	ACTN2_74881828	9451	C	0.275	0.25	T	0.2204	0.6387	1.138
4	PON1_38693816	12226	A	0.1084	0.09167	G	0.2149	0.643	1.205
25	UGCG_16689693	0	T	0.3494	0.375	C	0.1981	0.6562	0.8951
24	HIF1A_8973233	0	C	0.3293	0.3534	A	0.1772	0.6738	0.898
1	ACTA1+50243_68459659	50311	C	0.4634	0.4407	T	0.1431	0.7052	1.096
25	NDUFA8_25801538	1764	C	0.4872	0.5086	T	0.1223	0.7265	0.9178
25	NDUFA8_25795663	0	A	0.4873	0.5085	G	0.1207	0.7283	0.9189
25	PTGS1_26005531	14094	T	0.4444	0.4237	C	0.1192	0.7299	1.088
4	PON1_38697145	3329	T	0.07738	0.06667	C	0.1189	0.7303	1.174
22	PTPN1_38590385	4589	A	0.3063	0.325	C	0.1119	0.738	0.9168
9	ADHFE1_18788061	263	C	0.04938	0.05833	T	0.1097	0.7405	0.8386
24	HIF1A_8979927	6694	G	0.175	0.1897	A	0.09736	0.755	0.9063
1	TOMM20_76191732	5108	T	0.4337	0.4153	C	0.09633	0.7563	1.079
18	MSTN_66493737	212	T	0.443	0.425	C	0.09028	0.7638	1.076
22	ACSS1_762559	120	A	0.4767	0.4597	T	0.08426	0.7716	1.071
1	ACTA1_68409348	0	C	0.1524	0.1404	T	0.07816	0.7798	1.102
9	MTFR1_19439741	0	A	0.3086	0.2931	G	0.0774	0.7808	1.077
25	UGCG_16710063	178	G	0.08537	0.07627	C	0.07577	0.7831	1.13
4	ACN9_40305424	20228	T	0.2831	0.2966	G	0.06101	0.8049	0.9366
4	ACN9_40279726	12133	C	0.3291	0.3421	T	0.0502	0.8227	0.9434

22	PTPN1_ 38597033	5068	T	0.2615	0.2745	C	0.04911	0.8246	0.936
1	MYEF2_ 141653617	2255	T	0.4177	0.4068	G	0.03336	0.8551	1.046
25	UGCG_ 16709885	20192	G	0.3812	0.3917	A	0.0314	0.8593	0.957
22	ACSS1_762439	0	G	0.4753	0.4831	A	0.0164	0.8981	0.9695
22	COX4I2_ 22684844	168	C	0.2831	0.2881	T	0.008468	0.9267	0.9758
9	ADHFE1_ 18785084	0	T	0.05921	0.06122	C	0.004285	0.9478	0.965
22	ACSS1_768275	5716	T	0.07407	0.075	G	0.000858	0.9766	0.9867
25	TNC_19731498	14568	G	0.3214	0	T	7.193	0.007319	NA

Appendix II: TBE_SP (elite sprinter) V TBE_EN (elite stayer) association test. SNPs ranked by P value

CHR	SNP	BP	A1	F_A	F_U	A2	CHISQ	P	OR
18	MSTN_66493737	212	T	0.2821	0.6406	C	18.31	1.88E-05	0.2204
18	MSTN_66494218	443	C	0.2368	0.4844	A	9.357	0.002221	0.3304
4	ACN9_40279726	12133	C	0.4079	0.25	T	4.026	0.0448	2.067
22	PTPN1_38585796	0	G	0.225	0.1029	C	3.901	0.04826	2.53
22	COX4I2_22684844	168	C	0.2195	0.3529	T	3.283	0.07001	0.5156
22	PTPN1_38597033	5068	T	0.2931	0.1552	C	3.173	0.07488	2.257
4	PON1_38697145	3329	T	0.04878	0.1286	C	3.074	0.07955	0.3476
22	COX4I2-5900_22676361	0	C	0.359	0.5	T	2.957	0.0855	0.56
22	COX4I2_22684390	1164	C	0.2692	0.3971	T	2.69	0.101	0.5595
4	ACN9_40305424	20228	T	0.3415	0.2206	G	2.656	0.1032	1.832
1	MYEF2_141650968	3375	C	0.0875	0.1765	T	2.6	0.1068	0.4475
18	MSTN_66493525	0	G	0	0.02941	T	2.444	0.1179	0
18	MSTN_66493775	30	G	0	0.02941	A	2.444	0.1179	0
25	UGCG_16710063	178	G	0.05	0.1176	C	2.258	0.133	0.3947
9	ADHFE1_18787798	2714	G	0.01351	0.06061	A	2.246	0.1339	0.2123
10	CKM_15884567	2716	G	0.05128	0.1176	A	2.121	0.1453	0.4054
4	PON1_38681590	784	T	0.0625	0.1324	C	2.094	0.1479	0.437

1	MYEF2_141626957	0	C	0.09756	0.1765	T	2.003	0.157	0.5045
25	GSN_25028755	4291	A	0.05263	0.1176	G	1.986	0.1588	0.4167
22	COX4I2_22683226	6865	T	0.3108	0.4242	G	1.938	0.1639	0.612
1	MYEF2_141651362	394	G	0.09722	0.1765	A	1.872	0.1712	0.5026
25	GSN_25024464	0	T	0.05263	0.1143	A	1.836	0.1754	0.4306
24	HIF1A_8984849	4922	G	0.1053	0.1818	A	1.711	0.1909	0.5294
4	ACN9_40267593	0	A	0.01163	0.04286	T	1.506	0.2197	0.2627
22	PTPN1_38590385	4589	A	0.2692	0.3636	C	1.483	0.2233	0.6447
1	MYEF2_141653617	2255	T	0.3553	0.4545	G	1.449	0.2287	0.6612
4	PON1_38693816	12226	A	0.08537	0.1471	G	1.408	0.2354	0.5413
22	COX4I2_22684676	286	C	0.3049	0.3971	T	1.395	0.2376	0.666
1	MYEF2_141647593	20636	T	0.1081	0.1765	C	1.369	0.2421	0.5657
4	PON1_38680806	0	A	0.0875	0.1471	G	1.282	0.2574	0.5562
25	TNC_19737599	6101	C	0.4583	0.3636	G	1.274	0.2591	1.481
10	CKM_15887661	3094	C	0.2692	0.3529	T	1.194	0.2745	0.6754
1	ACTA1+50243_68459659	50311	C	0.425	0.5147	T	1.189	0.2756	0.6969
9	ADHFE1_18802749	66	A	0.025	0.05882	T	1.081	0.2985	0.4103
9	MTRF1_19456942	17072	A	0.03659	0.07353	T	1.005	0.3161	0.4785
24	HIF1A_8979927	6694	G	0.1316	0.1912	A	0.9498	0.3298	0.641

1	ACTN2_74900867	19039	T	0.03846	0.07576	A	0.9478	0.3303	0.488
25	TNC_19737816	217	A	0.4605	0.3824	G	0.8982	0.3433	1.379
22	PTPN1_38591965	1580	G	0.3125	0.2424	A	0.8793	0.3484	1.42
9	ADHFE1_18802683	9145	G	0.08108	0.04412	A	0.8156	0.3665	1.912
25	PTGS1_25991437	1489	C	0.0875	0.1324	T	0.7669	0.3812	0.6286
21	PRKAA1_25364288	0	A	0.0375	0.01471	G	0.7262	0.3941	2.61
21	PRKAA1_25374247	9959	A	0.0375	0.01515	G	0.6779	0.4103	2.532
9	ADHFE1_18793538	5477	C	0.1282	0.1765	T	0.6613	0.4161	0.6863
1	GGPS1_76001872	0	A	0.4756	0.4091	C	0.6549	0.4184	1.31
18	MSTN_66493745	8	G	0.01163	0.02941	A	0.6288	0.4278	0.3882
1	TOMM20_76186624	1936	C	0.175	0.2273	T	0.6208	0.4307	0.7212
25	GSN_25033440	4685	G	0.378	0.4412	A	0.614	0.4333	0.7699
4	PDK4_38969307	1168	A	0.3919	0.4559	C	0.5947	0.4406	0.7692
9	MTFR1_19439741	0	A	0.3375	0.2794	G	0.579	0.4467	1.314
9	ADHFE1_18785084	0	T	0.07895	0.04839	C	0.5231	0.4695	1.686
22	ACSS1_780613	12338	C	0.3026	0.25	T	0.4955	0.4815	1.302
22	ACSS1_762559	120	A	0.5116	0.4571	T	0.4585	0.4983	1.244
9	MTFR1_19472498	15556	A	0.04878	0.07353	G	0.4037	0.5252	0.6462
25	PTGS1	2168	C	0.122	0.1571	T	0.3928	0.5308	0.7449

	26007699								
1	GGPS1_76002021	149	C	0.4605	0.4091	A	0.3799	0.5376	1.233
1	ACTN2_74853540	11257	T	0.3026	0.2576	G	0.3544	0.5516	1.251
25	UGCG_16709885	20192	G	0.4189	0.3714	A	0.3392	0.5603	1.22
3	COX4I1_32772871	0	T	0.3846	0.3382	C	0.338	0.561	1.223
9	MTFR1_19439870	129	G	0.2805	0.3235	A	0.3279	0.5669	0.8151
25	UGCG_16689693	0	T	0.3659	0.3235	C	0.294	0.5877	1.206
25	TNC_19741797	3981	C	0.2778	0.3261	G	0.2761	0.5993	0.7949
1	ACTA1_68409348	0	C	0.1625	0.1324	T	0.264	0.6074	1.272
25	TNC_19731498	14568	G	0.35	0.25	T	0.262	0.6088	1.615
1	ACTN2_74881828	9451	C	0.2875	0.2576	T	0.1628	0.6866	1.163
4	PDK4_38973231	3924	A	0.439	0.4706	G	0.1494	0.6991	0.8804
22	MC3R-530_43059660	0	C	0.2692	0.2424	T	0.1346	0.7138	1.151
22	ACSS1_762439	0	G	0.5	0.4697	A	0.1329	0.7154	1.129
24	HIF1A_8973233	0	C	0.3125	0.3382	A	0.1111	0.7389	0.8893
4	PDK4_38968139	0	T	0.4375	0.4118	C	0.09958	0.7523	1.111
4	ACN9_40285196	5470	G	0.3684	0.3939	T	0.09761	0.7547	0.8974
1	TOMM20_76191732	5108	T	0.4512	0.4265	C	0.09241	0.7611	1.106
25	TNC_19716930	0	G	0.07692	0.09091	A	0.09154	0.7622	0.8333

1	ACTN2_ 74842283	0	G	0.075	0.08824	A	0.08642	0.7688	0.8378
22	ACSS1_ 759076	0	G	0.2375	0.2206	C	0.05941	0.8074	1.101
25	PTGS1_ 25989948	0	C	0.1351	0.1471	T	0.04164	0.8383	0.9062
9	ADHFE1_ 18788061	263	C	0.05128	0.05882	T	0.03989	0.8417	0.8649
25	NDUFA8_ 25799094	3431	G	0.4833	0.5	T	0.02746	0.8684	0.9355
25	NDUFA8_ 25801538	1764	C	0.5135	0.5	T	0.02549	0.8732	1.056
25	NDUFA8_ 25795663	0	A	0.5132	0.5	G	0.02486	0.8747	1.054
22	ACSS1_ 768275	5716	T	0.07895	0.08571	G	0.02212	0.8818	0.9143
10	CKM_ 15881851	0	A	0.06757	0.07353	G	0.01924	0.8897	0.913
1	ACTN2_ 74872377	18837	G	0.09459	0.08824	T	0.01723	0.8956	1.08
1	TOMM20_ 76184688	0	T	0.3333	0.3235	A	0.01636	0.8982	1.045
25	PTGS1_ 26005531	14094	T	0.4625	0.4559	C	0.006481	0.9358	1.027
25	NDUFA8_ 25799774	680	T	0.4865	0.4853	C	0.000202	0.9887	1.005

Appendix III: Significant associations between SNP and phenotype

TBE - elite (Group race winning) Thoroughbred

TBO - other (non-winning) Thoroughbred

TBE_SP8 - elite (Group race winning) Thoroughbred that won best race over a distance <8f

5 TBE_SP7 - elite (Group race winning) Thoroughbred that won best race over a distance <7f

TBE_EN- elite (Group race winning) Thoroughbred that won best race over a distance >8f

SNPs are given by GeneSymbol_chromosomeposition(bp)

TBE V TBO	P	OR
PDK4_38973231	0.001676	2.2
CKM_15884567	0.02066	0.4089
COX4I2_22684390	0.03098	0.5778
TBE SP8 V TBE EN	P	OR
PON1_38697145	0.03584	0.2884
PTPN1_38585796	0.01011	3.157
MSTN_66493737	3.70E-05	0.2503
TBE SP7 V TBE EN	P	OR
ACN9_40279726	0.0448	2.067
PTPN1_38585796	0.04826	2.53
MSTN_66493737	1.88E-05	0.2204
TBE EN V TBO	P	OR
PDK4_38973231	0.008833	2.26
PTPN1_38585796	0.01482	0.3443
MSTN_66493737	0.005334	2.412
TBE SP8 V TBO	P	OR
ADHFE1_18802749	0.04945	0.2383
PDK4_38973231	0.006401	2.159
CKM_15884567	0.00545	0.2272
COX4I2_22684390	0.007404	0.4478
P - TBE SP7 V TBO	P	OR
GSN_25024464	0.03537	0.3148
PDK4_38973231	0.02048	1.99
MSTN_66493737	0.04163	0.5315
CKM_15884567	0.01821	0.2763
COX4I2_22684390	0.009814	0.4421
P TBE V TBO males	P	OR
ACTN2_74842283	0.04372	0.308
PDK4_38973231	0.003429	3.4
PTGS1_26007699	0.005124	0.2174
PTPN1_38590385	0.03461	0.3966
COX4I1_32772871	0.04415	2.229

Appendix IV: Case control association test and best fit model for significantly associated SNPs**TBE V TBO (Case - control association test) P < 0.05**

CHR	SNP	BP	A1	F A	F U	A2	CHISQ	P	OR
4	PDK4_38973231	3924	A	0.4639	0.2823	G	9.874	0.001676	2.2
4	PDK4_38968139	0	T	0.4146	0.582	C	7.842	0.005106	0.5088
4	PDK4_38969307	1168	A	0.4304	0.2712	C	7.409	0.006488	2.031
10	CKM_15884567	2716	G	0.07407	0.1636	A	5.355	0.02066	0.4089
18	MSTN_66493525	0	G	0.01205	0.05833	T	4.896	0.02692	0.1969
22	COX4I2_22684390	1164	C	0.325	0.4545	T	4.654	0.03098	0.5778
22	COX4I2_22684676	286	C	0.3415	0.4655	T	4.384	0.03628	0.5953
22	COX4I2_22683226	6865	T	0.3636	0.4828	G	3.868	0.04922	0.6122
22	ACSS1_759076	0	G	0.2317	0.1379	C	3.838	0.05009	1.885

5 TBE V TBO Best-fit Model for significantly associated SNPs

CHR	SNP	A1	A2	TEST	AFF	UNAFF	CHISQ	DF	P
4	PDK4_38973231	A	G	ALLELIC	77/89	35/89	9.874	1	0.001676
4	PDK4_38973231	A	G	TREND	77/89	35/89	9.237	1	0.002372
4	PDK4_38973231	A	G	DOM	59/24	29/33	8.791	1	0.003027
4	PDK4_38973231	A	G	GENO	18/41/24	6/23/33	9.644	2	0.008049
4	PDK4_38973231	A	G	REC	18/65	6/56	3.706	1	0.05422
4	PDK4_38968139	T	C	ALLELIC	68/96	71/51	7.842	1	0.005106
4	PDK4_38968139	T	C	TREND	68/96	71/51	6.841	1	0.008907
4	PDK4_38968139	T	C	REC	16/66	23/38	5.837	1	0.01569
4	PDK4_38968139	T	C	GENO	16/36/30	23/25/13	7.029	2	0.02977
4	PDK4_38968139	T	C	DOM	52/30	48/13	3.881	1	0.04884

4	PDK4_38969307	A	C	DOM	54/25	26/33	8.177	1	0.004243
4	PDK4_38969307	A	C	ALLELIC	68/90	32/86	7.409	1	0.006488
4	PDK4_38969307	A	C	TREND	68/90	32/86	6.996	1	0.008169
4	PDK4_38969307	A	C	GENO	14/40/25	6/20/33	8.245	2	0.01621
4	PDK4_38969307	A	C	REC	14/65	19511	1.554	1	0.2125
10	CKM_15884567	G	A	ALLELIC	12/150	18/92	5.355	1	0.02066
10	CKM_15884567	G	A	DOM	11/70	16/39	4.953	1	0.02605
10	CKM_15884567	G	A	TREND	12/150	18/92	4.865	1	0.02741
10	CKM_15884567	G	A	GENO	1/10/70	2/14/39	5.03	2	0.08087
10	CKM_15884567	G	A	REC	1/80	2/53	0.876	1	0.3493
18	MSTN_66493525	G	T	ALLELIC	2/164	7/113	4.896	1	0.02692
18	MSTN_66493525	G	T	TREND	2/164	7/113	3.432	1	0.06394
18	MSTN_66493525	G	T	REC	0/83	21217	2.806	1	0.09392
18	MSTN_66493525	G	T	DOM	29618	20210	2.625	1	0.1052
18	MSTN_66493525	G	T	GENO	0/2/81	20150	3.563	2	0.1683
22	COX4I2_22684390	C	T	REC	4/76	10/45	6.093	1	0.01357
22	COX4I2_22684390	C	T	TREND	52/108	50/60	5.58	1	0.01817
22	COX4I2_22684390	C	T	GENO	4/44/32	10/30/15	6.979	2	0.03052
22	COX4I2_22684390	C	T	ALLELIC	52/108	50/60	4.654	1	0.03098
22	COX4I2_22684390	C	T	DOM	48/32	40/15	2.326	1	0.1272
22	COX4I2_22684676	C	T	REC	28246	17472	5.557	1	0.01841
22	COX4I2_22684676	C	T	TREND	56/108	54/62	5.268	1	0.02172
22	COX4I2_22684676	C	T	ALLELIC	56/108	54/62	4.384	1	0.03628
22	COX4I2_22684676	C	T	GENO	5/46/31	11/32/15	6.402	2	0.04072

22	COX4I2_22684676	C	T	DOM	51/31	43/15	2.196	1	0.1383
22	COX4I2_22683226	T	G	REC	26420	17137	6.057	1	0.01385
22	COX4I2_22683226	T	G	TREND	56/98	56/60	4.776	1	0.02887
22	COX4I2_22683226	T	G	GENO	5/46/26	12/32/14	6.449	2	0.03978
22	COX4I2_22683226	T	G	ALLELIC	56/98	56/60	3.868	1	0.04922
22	COX4I2_22683226	T	G	DOM	51/26	44/14	1.471	1	0.2252
22	ACSS1_759076	G	C	DOM	35/47	15/43	4.187	1	0.04075
22	ACSS1_759076	G	C	TREND	38/126	16/100	4.063	1	0.04382
22	ACSS1_759076	G	C	ALLELIC	38/126	16/100	3.838	1	0.05009
22	ACSS1_759076	G	C	GENO	3/32/47	1/14/43	4.231	2	0.1206
22	ACSS1_759076	G	C	REC	28915	20821	0.458	1	0.4986

TBE SP7 V TBE EN (Case - control association test) P < 0.05

CHR	SNP	BP	A1	F_A	F_U	A2	CHISQ	P	OR
18	MSTN_66493737	212	T	0.2821	0.6406	C	18.31	1.88E-05	0.22044.54
18	MSTN_66494218	443	C	0.2368	0.4844	A	9.357	0.002221	0.3304
4	ACN9_40279726	12133	C	0.4079	0.25	T	4.026	0.0448	2.067
22	PTPN1_38585796	0	G	0.225	0.1029	C	3.901	0.04826	2.53

TBE SP7 V TB EN Best-fit Model for significantly associated SNPs

CHR	SNP	A1	A2	TEST	AFF	UNAFF	CHISQ	DF	P
18	MSTN_66493737	T	C	GENO	3/16/20	9/23/0	23.8	2	6.80E-06
18	MSTN_66493737	T	C	TREND	22/56	41/23	20.64	1	5.55E-06
18	MSTN_66493737	T	C	ALLELIC	22/56	41/23	18.31	1	1.88E-05
18	MSTN_66493737	T	C	DOM	19/20	32/0	22.85	1	1.76E-06
18	MSTN_66493737	T	C	REC	3/36	9/23	5.225	1	0.02226
18	MSTN_66494218	C	A	GENO	2/14/22	6/19/7	10.08	2	0.006487
18	MSTN_66494218	C	A	TREND	18/58	31/33	9.708	1	0.001835
18	MSTN_66494218	C	A	ALLELIC	18/58	31/33	9.357	1	0.002221
18	MSTN_66494218	C	A	DOM	16/22	40019	9.288	1	0.002306

18	MSTN_66494218	C	A	REC	13181	46174	3.122	1	0.07726
4	ACN9_40279726	C	T	GENO	6/19/13	2/13/19	4.04	2	0.1326
4	ACN9_40279726	C	T	TREND	31/45	17/51	4.026	1	0.0448
4	ACN9_40279726	C	T	ALLELIC	31/45	17/51	4.026	1	0.0448
4	ACN9_40279726	C	T	DOM	25/13	15/19	3.413	1	0.06467
4	ACN9_40279726	C	T	REC	11841	11720	1.783	1	0.1817
22	PTPN1_38585796	G	C	GENO	0/18/22	0/7/27	NA	NA	NA
22	PTPN1_38585796	G	C	TREND	18/62	7/61	4.896	1	0.02692
22	PTPN1_38585796	G	C	ALLELIC	18/62	7/61	3.901	1	0.04826
22	PTPN1_38585796	G	C	DOM	18/22	7/27	4.896	1	0.02692
22	PTPN1_38585796	G	C	REC	0/40	0/34	NA	NA	NA

TBE Best Race Distance Quantitative Trait Association

CHR	SNP	STAT	EMPI	NP
18	MSTN_66493737	36.63	1.00E-06	1000000
18	MSTN_66494218	15.97	0.000174	137586
22	COX4I2_22684844	6.495	0.01146	2094
22	COX4I2_22684390	5.783	0.02526	949
22	PTPN1_38585796	4.963	0.0377	609
4	PON1_38697145	4.596	0.03938	583
22	PTPN1_38597033	4.64	0.04406	521
22	COX4I2_22684676	4.51	0.04742	484
22	COX4I2_22683226	4.248	0.05263	436
4	ACN9_40279726	3.628	0.0545	421

TBE Best Race Distance Means (where distances are furlongs) for significantly associated**5 SNPs**

CHR	SNP	VALUE	G11	G12	G22
18	MSTN_66493737	GENO	T/T	T/C	C/C
18	MSTN_66493737	COUNTS	12	46	21
18	MSTN_66493737	FREQ	0.1519	0.5823	0.2658

18	MSTN_66493737	MEAN	10.54	9.087	6.167
18	MSTN_66493737	SD	2.742	2.365	0.8266
18	MSTN_66494218	GENO	C/C	C/A	A/A
18	MSTN_66494218	COUNTS	8	39	31
18	MSTN_66494218	FREQ	0.1026	0.5	0.3974
18	MSTN_66494218	MEAN	10.56	9.179	7.403
18	MSTN_66494218	SD	2.872	2.48	2.043
22	COX4I2_22684844	GENO	C/C	C/T	T/T
22	COX4I2_22684844	COUNTS	4	39	40
22	COX4I2_22684844	FREQ	0.04819	0.4699	0.4819
22	COX4I2_22684844	MEAN	11.12	8.91	7.975
22	COX4I2_22684844	SD	2.097	2.762	2.247
22	COX4I2_22684390	GENO	C/C	C/T	T/T
22	COX4I2_22684390	COUNTS	4	44	32
22	COX4I2_22684390	FREQ	0.05	0.55	0.4
22	COX4I2_22684390	MEAN	10.62	8.943	7.875
22	COX4I2_22684390	SD	2.056	2.783	2.254
22	PTPN1_38585796	GENO	G/G	G/C	C/C
22	PTPN1_38585796	COUNTS	1	30	50
22	PTPN1_38585796	FREQ	0.01235	0.3704	0.6173
22	PTPN1_38585796	MEAN	8	7.75	9.09
22	PTPN1_38585796	SD	0	2.176	2.736
4	PON1_38697145	GENO	T/T	T/C	C/C
4	PON1_38697145	COUNTS	1	11	72
4	PON1_38697145	FREQ	0.0119	0.131	0.8571
4	PON1_38697145	MEAN	6	10.82	8.278
4	PON1_38697145	SD	0	2.892	2.359
22	PTPN1_38597033	GENO	T/T	T/C	C/C
22	PTPN1_38597033	COUNTS	2	30	33
22	PTPN1_38597033	FREQ	0.03077	0.4615	0.5077
22	PTPN1_38597033	MEAN	9.5	7.917	9.5
22	PTPN1_38597033	SD	2.121	2.275	2.339
22	COX4I2_22684676	GENO	C/C	C/T	T/T
22	COX4I2_22684676	COUNTS	5	46	31
22	COX4I2_22684676	FREQ	0.06098	0.561	0.378
22	COX4I2_22684676	MEAN	10.1	8.859	7.903
22	COX4I2_22684676	SD	2.133	2.75	2.286

22	COX4I2_22683226	GENO	T/T	T/G	G/G
22	COX4I2_22683226	COUNTS	5	46	26
22	COX4I2_22683226	FREQ	0.06494	0.5974	0.3377
22	COX4I2_22683226	MEAN	10.9	8.641	8.077
22	COX4I2_22683226	SD	1.884	2.491	2.331
4	ACN9_40279726	GENO	C/C	C/T	T/T
4	ACN9_40279726	COUNTS	8	36	35
4	ACN9_40279726	FREQ	0.1013	0.4557	0.443
4	ACN9_40279726	MEAN	7.875	8.264	9.3
4	ACN9_40279726	SD	2.1	2.628	2.544

Racing Post Handicap Rating (Best RPR) Quantitative Trait Association

CHR	SNP	STAT	EMP1	NP
4	PDK4_38973231	8.095	0.005052	4750
4	PDK4_38969307	6.825	0.009441	2541
3	COX4I1_32772871	6.748	0.009681	2478

5 Racing Post Handicap Rating (Best RPR) Means for significantly associated SNPs

CHR	SNP	STAT	EMP1	NP	
4	PDK4_38973231	GENO	A/A	A/G	G/G
4	PDK4_38973231	COUNTS	19	46	44
4	PDK4_38973231	FREQ	0.1743	0.422	0.4037
4	PDK4_38973231	MEAN	99.95	97.7	80.3
4	PDK4_38973231	SD	33.78	28.9	28.85
4	PDK4_38969307	GENO	A/A	A/C	C/C
4	PDK4_38969307	COUNTS	16	42	42
4	PDK4_38969307	FREQ	0.16	0.42	0.42
4	PDK4_38969307	MEAN	97.19	99.21	79.9
4	PDK4_38969307	SD	36.23	28.06	28.45
3	COX4I1_32772871	GENO	T/T	T/C	C/C
3	COX4I1_32772871	COUNTS	13	42	50
3	COX4I1_32772871	FREQ	0.1238	0.4	0.4762
3	COX4I1_32772871	MEAN	100.6	99.71	83.3
3	COX4I1_32772871	SD	29.46	28.92	30.49

The claims defining the invention are as follows:

1. A method for predicting the best race distance performance potential of a Thoroughbred race horse comprising the step of assaying a biological sample from a Thoroughbred race horse for the presence of a MSTN_66493737 (T/C) single nucleotide polymorphism (SNP).
2. A method as claimed in claim 1 wherein the presence of a C allele is indicative of elite sprinting performance potential.
3. A method as claimed in claim 1 or 2 wherein the presence of a homozygous CC genotype is indicative of elite sprinting performance potential.
4. A method as claimed in claim 1 or 2 wherein the presence of a T allele is indicative of stamina performance potential.
5. A method as claimed in claim 1 wherein the presence of a homozygous TT allele is indicative of stamina performance potential.
6. A method as claimed in any one of claims 1 to 5 wherein the biological sample of the subject is selected from the group comprising: blood, saliva, skeletal muscle, hair, semen, bone marrow, soft tissue, internal organ biopsy sample and skin.
7. A method as claimed in any of claims 1 to 6 comprising the steps of:
 - extracting or releasing DNA from an *ex-vivo* biological sample; and
 - identifying the genotype of the MSTN-66493737 (T/C) SNP in the extracted or released DNA

wherein the presence of a C allele in the MSTN-66493737 (T/C) SNP is indicative of elite sprinting performance potential and wherein the presence of a T allele in the MSTN-66493737 (T/C) SNP is indicative of stamina performance potential.

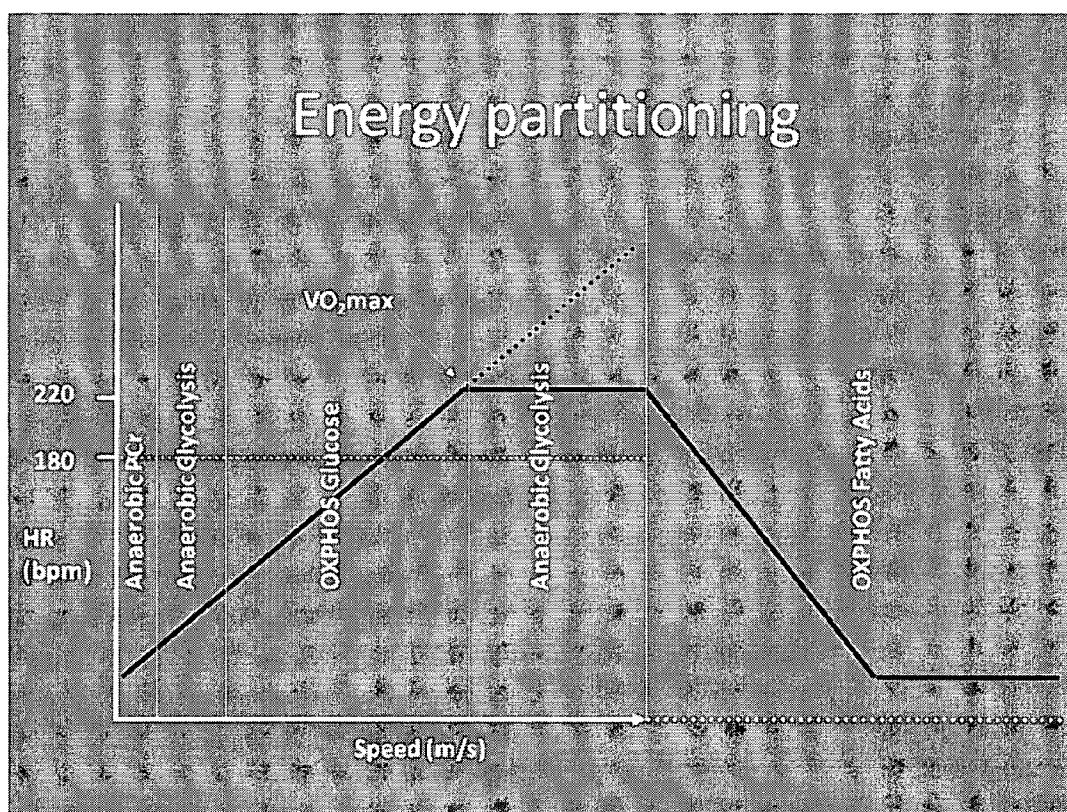
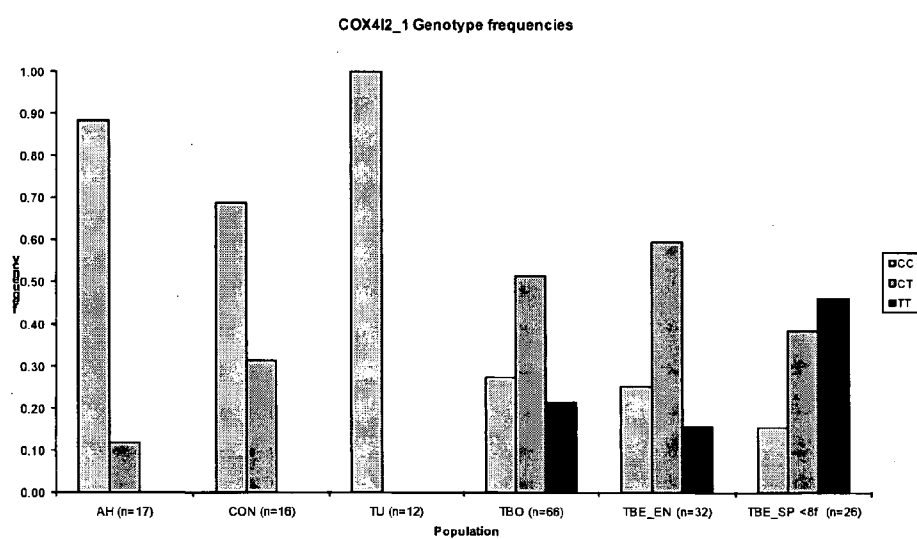
8. A method as claimed in claim 7 comprising the step of:

- amplifying a target sequence encoding the MSTN-66493737 (T/C) SNP in the extracted or released DNA

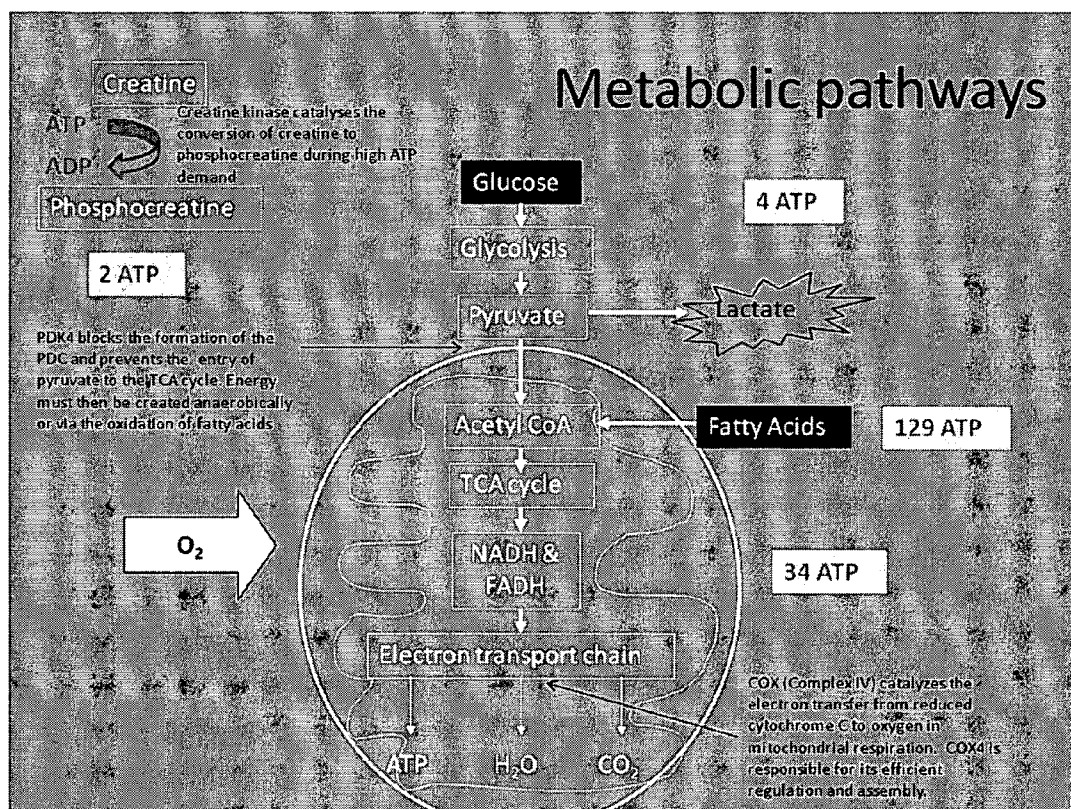
prior to the step of identifying the genotype of the MSTN-66493737 (T/C) SNP

- 5 9. A method as claimed in claim 7 or 8 wherein the DNA is genomic DNA.
- 10 10. An isolated polynucleotide consisting of the nucleic acid sequence of SEQ ID No. 31.
- 10 11. An isolated polynucleotide as claimed in claim 10 wherein the nucleotide at position 202 is c.
- 15 12. Use of an isolated polynucleotide consisting of the nucleic acid sequence of SEQ ID No. 31 as a genetic marker for predicting the best race distance performance potential of a Thoroughbred race horse.
13. Use as claimed in claim 12 wherein the presence of a C allele is indicative of elite sprinting performance potential.
- 20 14. Use as claimed in claim 12 or 13 wherein the presence of a T allele is indicative of stamina performance potential.
- 25 15. Use as claimed in claim 12 wherein the presence of a homozygous CC genotype is indicative of elite sprinting performance potential and the presence of a homozygous TT genotype is indicative of stamina performance potential.

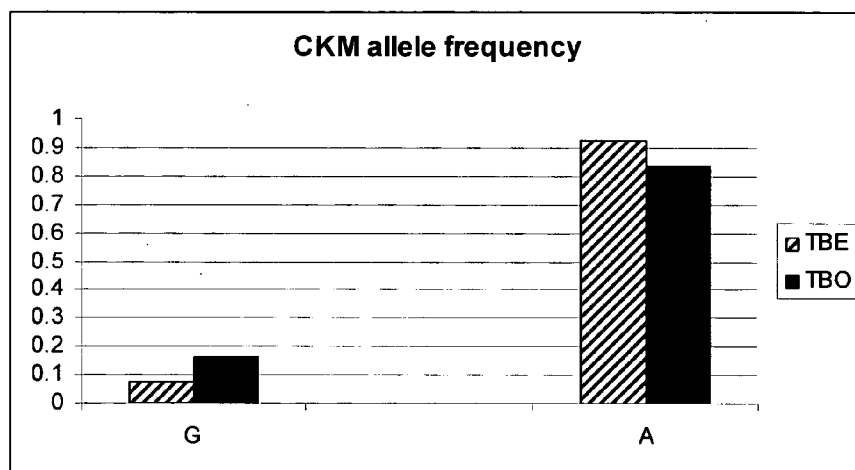
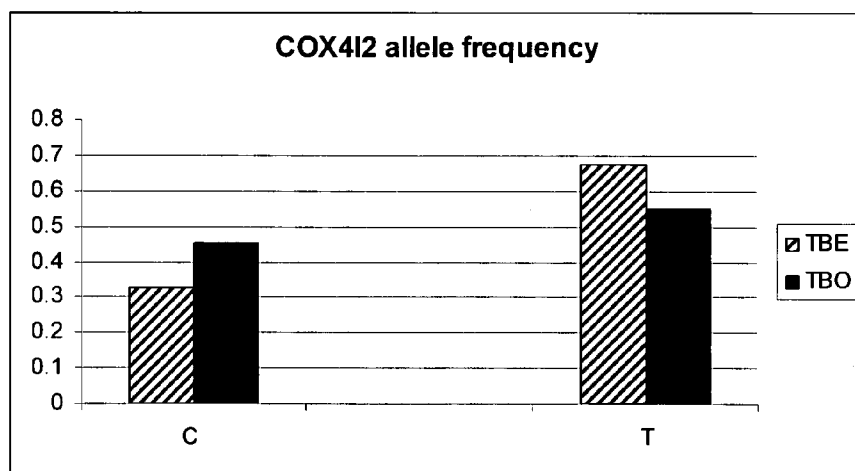
1 / 10

**Fig. 1****FIG.2**

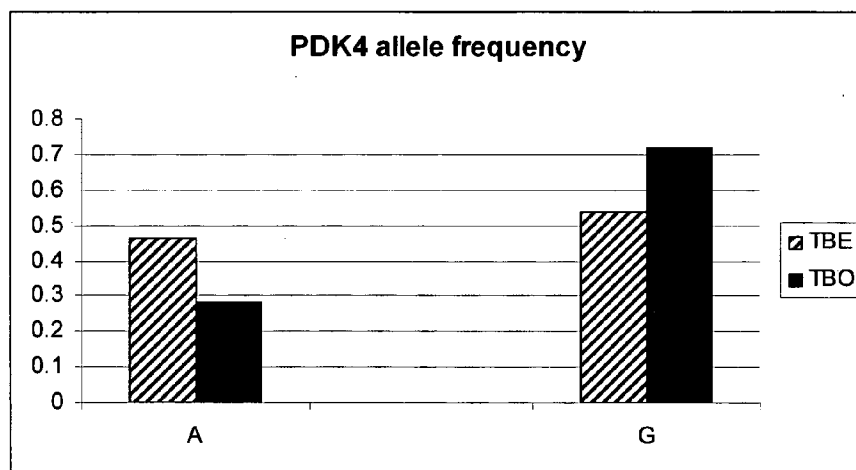
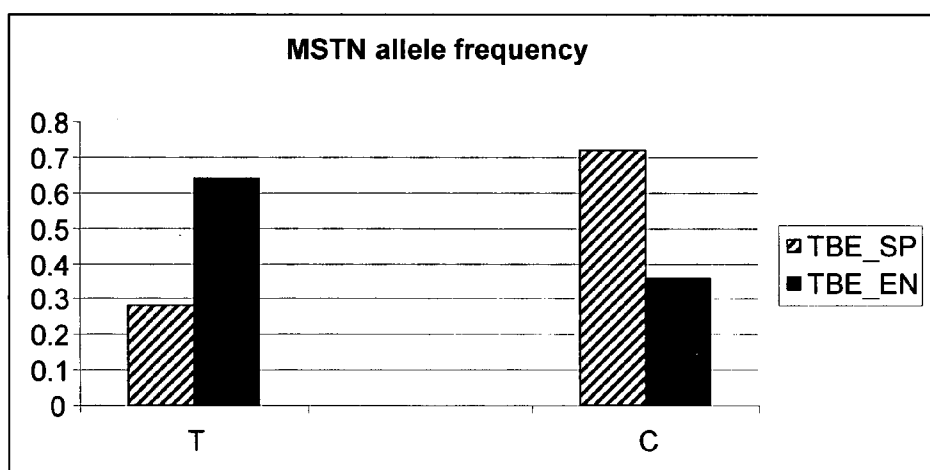
2 / 10

**Fig. 3**

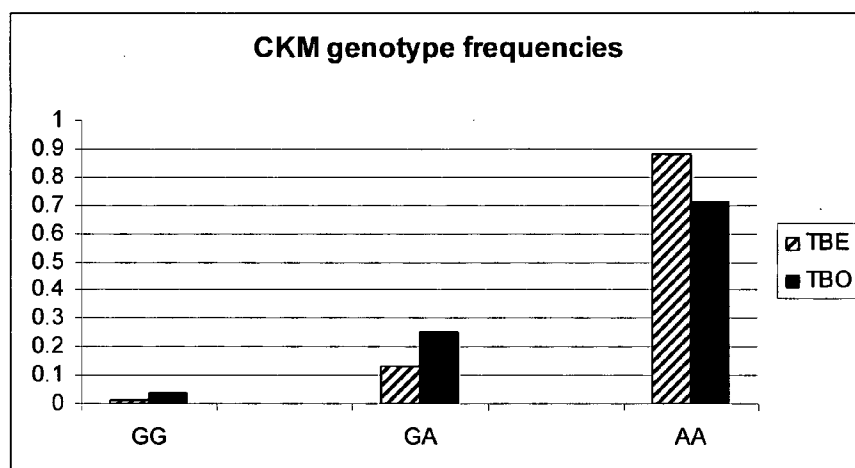
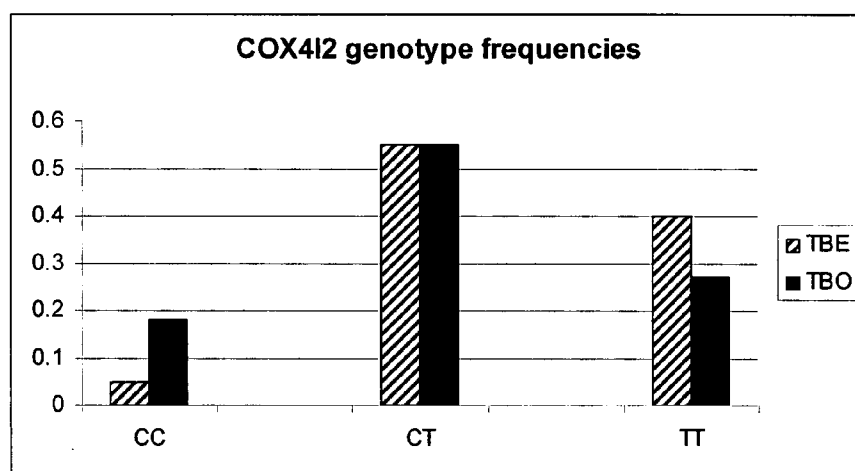
3 / 10

A**B**

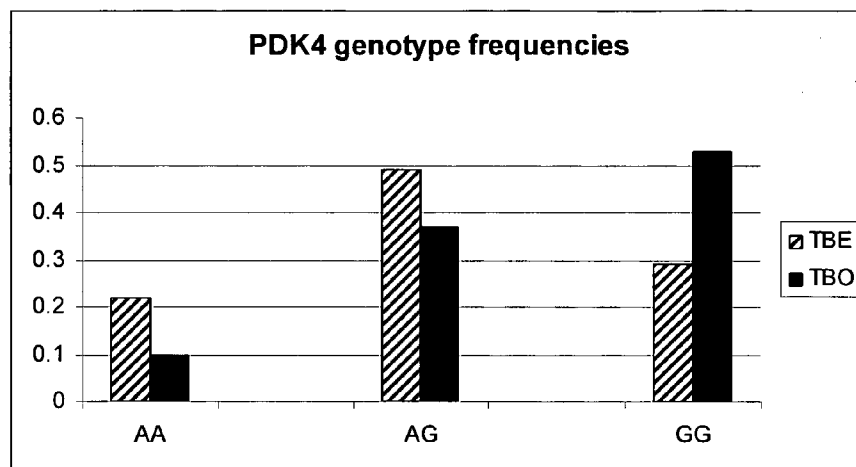
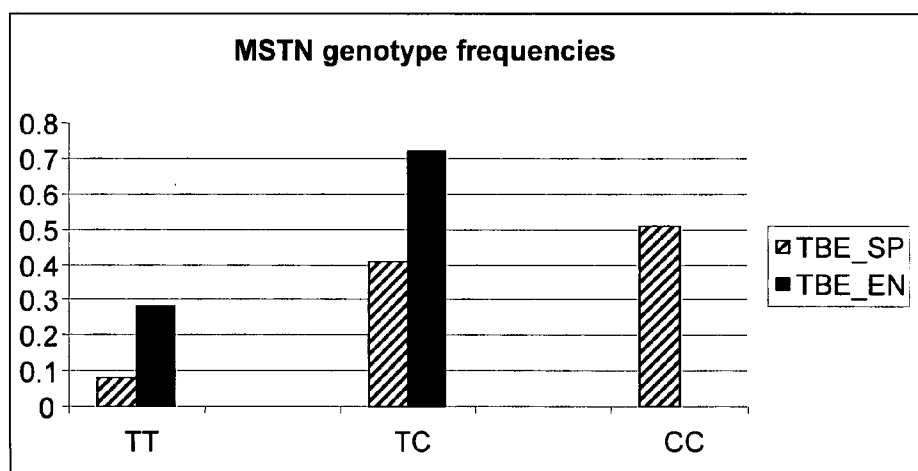
4 / 10

CDFig. 4

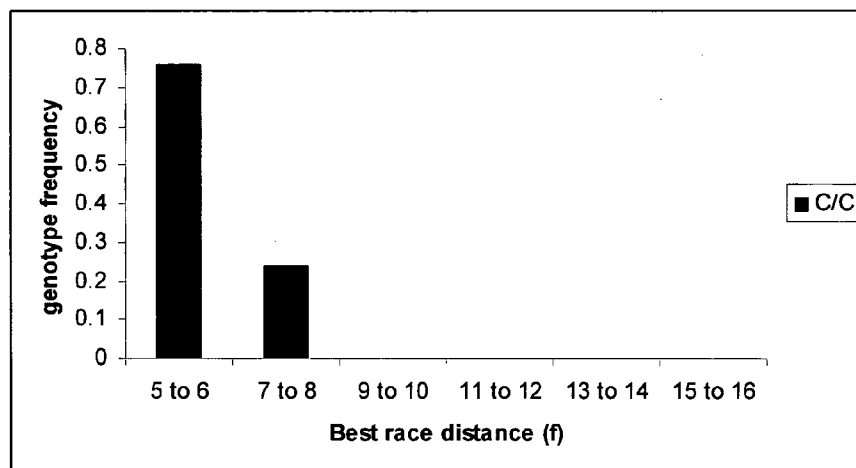
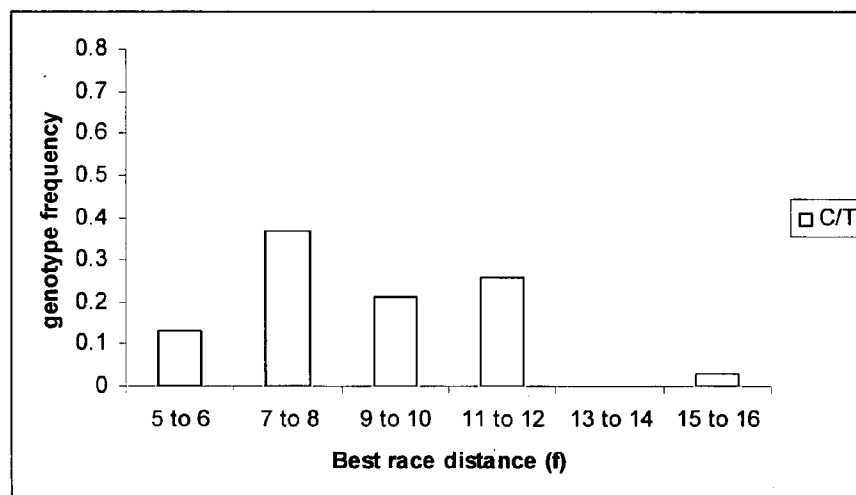
5 / 10

AB

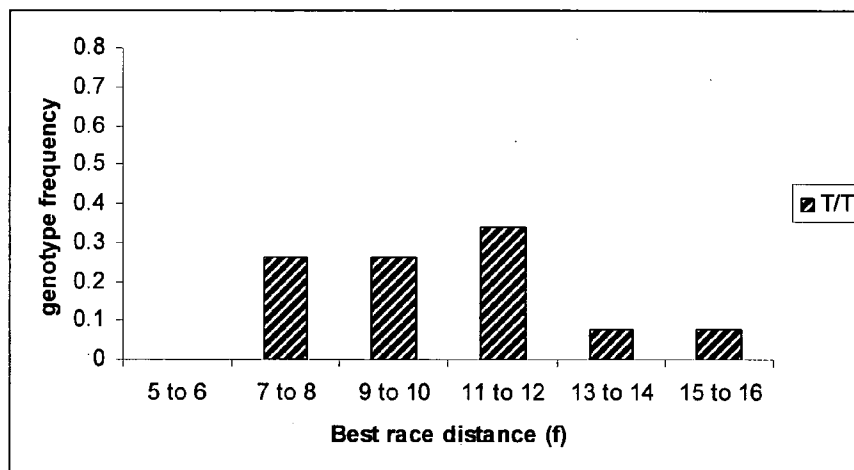
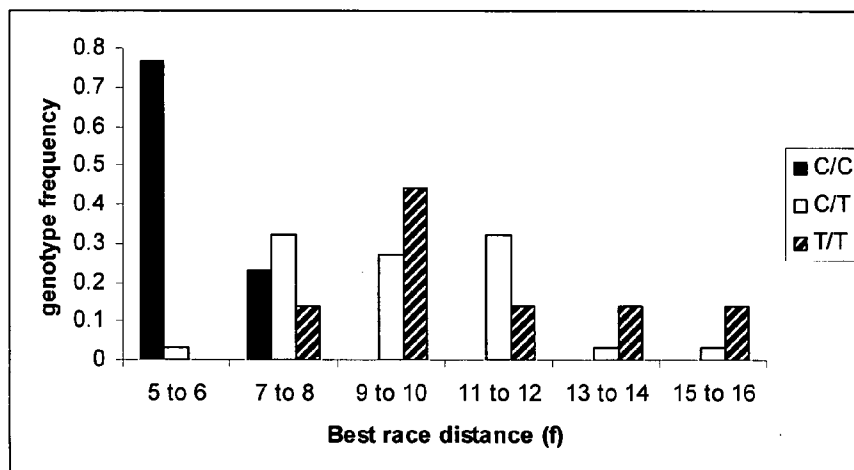
6 / 10

CDFig. 5

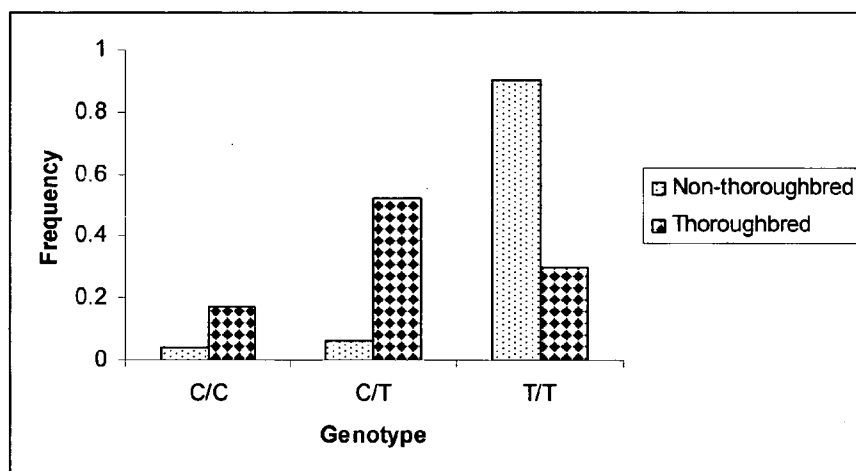
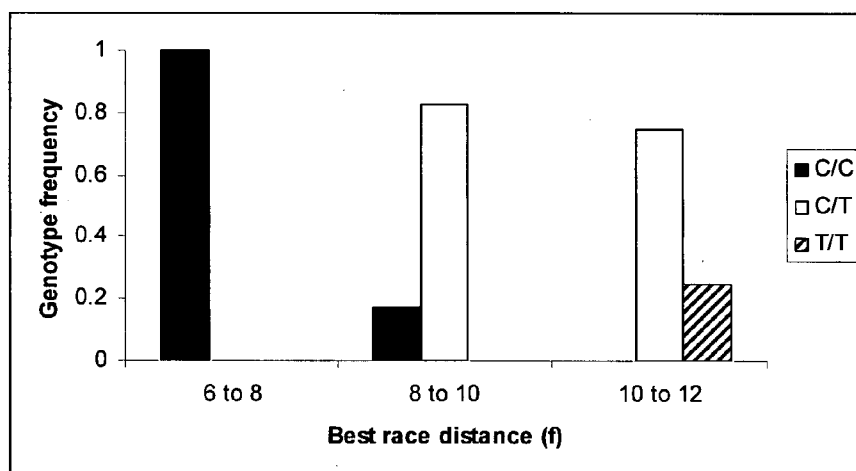
7 / 10

A**B**

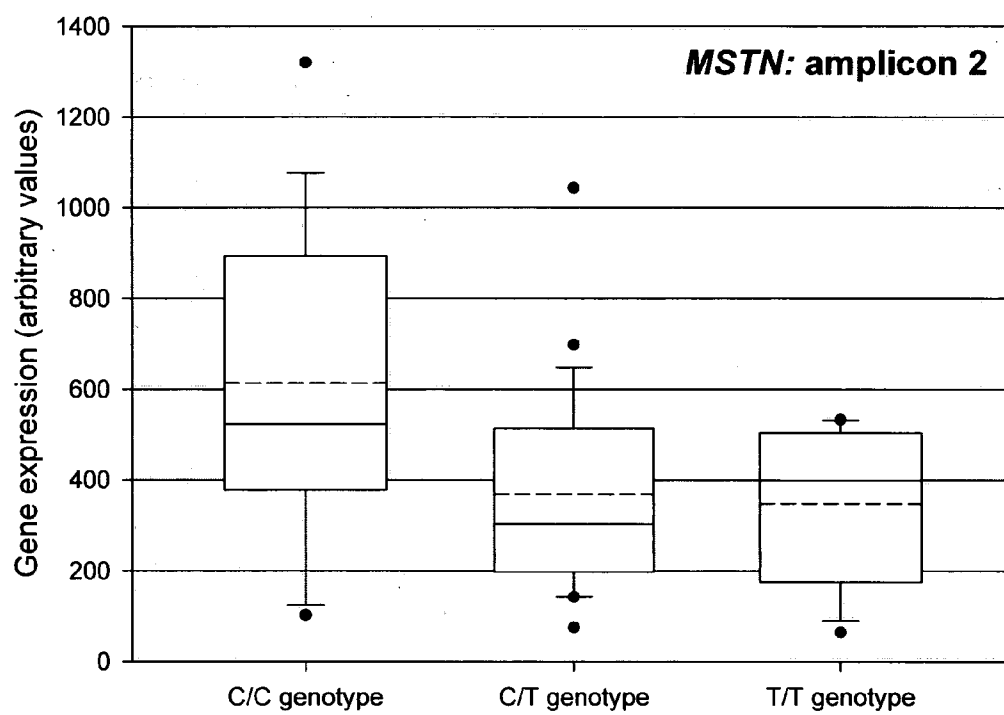
8 / 10

C**Fig. 6****Fig. 7**

9 / 10

**Fig. 8****Fig. 9**

10 / 10

**Fig. 10**

UCD011.C.WO.ST25
SEQUENCE LISTING

<110> University College Dublin - National University of Ireland,
Dublin

<120> A method for predicting athletic performance potential

<130> UCD011/c/wo

<150> US 61/136,553

<151> 2008-09-11

<150> US 61/213,125

<151> 2009-05-07

<160> 43

<170> Patentin version 3.3

<210> 1

<211> 401

<212> DNA

<213> Equus caballus

<220>

<221> misc_feature

<223> COX4I2 EquCab2.0 22676361_C/T SNP

<220>

<221> variation

<222> (201)..(201)

<223> y is c or t

<400> 1

caagagtgga ggtgtctcca agaactggag gctagcatgt agcagaggag gcgttagcag 60

aggagagag gttgatgggg gagctgcatt tggagagtct ggcaggcagg accttgaatg 120

ccaggcraag gattttattg ggaggcaagt ggtgtctgat aaaggctcaa ggattccatc 180

aggtgtttcc cacaagagac ygggcccact cagggcacca tatccctata tccaggagcc 240

agttgtgtcc cagagaaaac aagggactgg accttgagac ttggccagtg tccctcacct 300

cttaccttgt gcargccctt gtttggcttg tgggtcagaa ggcccttggg agacctgaag 360

cagaagctgc agaccattcc aggttagtgt ggagcccag a 401

<210> 2

<211> 401

<212> DNA

<213> Equus caballus

<220>

<221> misc_feature

<223> COX4I2 EquCab 2.0 22684390_C/T SNP

<220>

<221> variation

<222> (201)..(201)

<223> y is t or c

<400> 2

gctgggagat cctggggaca taaaagtga tcacctggat ggttcttgcc ctccaggggc 60

UCD011.C.WO.ST25

```

tcccagtcac gtgggggaac caacacaagc ccagataact gtaatatagg atatgtggcg 120
aggggtgaagt gtgttcaagg ggctgtgagg acccaaagga gagagagatg aaatccrpgt 180
gggcctttcca gaggagggca ygttctagtt gaccttgaat ggtgaggctg aggggtgctgc 240
caggtggttg gaacagcatg ggttaagggt tgggagcgga agagcargga gggtcttagg 300
catcagtaag tgctgtaggg gaaggaacag agagagggcg tgaggtggcc aggaagaag 360
ggggcctgac cctggggagc aggagggatg tgtgaccca a 401

```

<210> 3
<211> 401
<212> DNA
<213> Equus caballus

<220>
<221> misc_feature
<223> CKM EquCab 2.0 15884567_G/A SNP

<220>
<221> variation
<222> (48)..(48)
<223> r is g or a

<220>
<221> variation
<222> (201)..(201)
<223> r is g or a

```

<400> 3
ctgtccctaa cagacctgga ccttggcccc gtggaggtcc taaaggcrac tatacgggat 60
gtaaaccctaa attcatgaca tcccttgaag catgtcttc cctgtcttgc ccgggtcccc 120
ggaacagcca cccaagtgc tcttcccaa gtggactctc ccttcacacc ctgccctcgc 180
catccagtgc accggcaagc racactatcc cgttgccacc tccagaaagt caatgtctca 240
ggaatctggg gagccatcag tcaaaattac tatcatcac tatatatagg attcgcatat 300
attcctatgc ataataatta tacgttttgt ggataataaa tatatgtata tatgcataat 360
atttacataa tatatacata tttatataca ttttatatat t 401

```

<210> 4
<211> 401
<212> DNA
<213> Equus caballus

<220>
<221> misc_feature
<223> PDK4 EquCab 2.0 38973211_A/G SNP

<220>
<221> variation
<222> (201)..(201)
<223> r is g or a

```

<400> 4
actttaaccc tcaactttct aacttaaaat ttatgtttta ctattccaga gcaatattca 60
gttttatttg gcaaatgttt tcatttttta tagcaaaagt atttagaaat ttttaagaaa 120

```

UCD011.C.WO.5T25

gatttcatat tictttctac ttcattcatt caigtgtggg tagaagtctc gaaagcagca 180
 gtaaagacta tggattgaat ctgtgtttca gattgtcatt gtttaattggg tatgggaatgc 240
 atatatctct tgaatcaatg aacaaaaagc tgtatagtca gcagattagg gtgaggctct 300
 ggtgcataat tgcctgcagt catatccrgg ctctattctc tgaaaatctg ctcttgtggg 360
 tcatctaccc tctctaagct tmacacccc tatttgtaa a 401

<210> 5
 <211> 23
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_1 forward primer

<400> 5
 ataaatgcaa ttgtctcaaa gtc 23

<210> 6
 <211> 21
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_1 reverse primer

<400> 6
 ccatatgcaa gtttccattc c 21

<210> 7
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_2 forward primer

<400> 7
 tcagccattc agcctatttg 20

<210> 8
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_2 reverse primer

<400> 8
 acggttggca ttttaaccatc 20

UCD011.C.WO.ST25

<210> 9
 <211> 23
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_3 forward primer

<400> 9
 ggagacttgc ttctatttac ctg

23

<210> 10
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_3 reverse primer

<400> 10
 gaagcttttg gatgggattg

20

<210> 11
 <211> 19
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_4 forward primer

<400> 11
 ctctgggggtt tgcttggtg

19

<210> 12
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_4 reverse primer

<400> 12
 acctagggaa tggaggatgg

20

<210> 13
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_5 forward primer

<400> 13
 gaagaggagg gaggggaagag

20

UCD011.C.WO.ST25

<210> 14
 <211> 22
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_5 reverse primer

<400> 14
 ttcagtcttc atgtgggtctt gg 22

<210> 15
 <211> 21
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_7 forward primer

<400> 15
 aaggtattgt catctgcttg g 21

<210> 16
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_7 reverse primer

<400> 16
 ccaagaccag gagaagatgg 20

<210> 17
 <211> 22
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_8 forward primer

<400> 17
 gcttgtagc ataggaaact gg 22

<210> 18
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_8 reverse primer

UCD011.C.WO.ST25

<400> 18
ctgagacctg tcaagactcc

20

<210> 19
<211> 20
<212> DNA
<213> Equus caballus

<220>
<221> misc_feature
<223> MSTN_9 forward primer

<400> 19
cgtctttcat gggtttgatg

20

<210> 20
<211> 20
<212> DNA
<213> Equus caballus

<220>
<221> misc_feature
<223> MSTN_9 reverse primer

<400> 20
atgttctctc acggtgtctc

20

<210> 21
<211> 21
<212> DNA
<213> Equus caballus

<220>
<221> misc_feature
<223> MSTN_10 forward primer

<400> 21
tgaaggaatg aactgtggat g

21

<210> 22
<211> 20
<212> DNA
<213> Equus caballus

<220>
<221> misc_feature
<223> MSTN_10 reverse primer

<400> 22
gtctgcgac ctcgtttacc

20

<210> 23
<211> 21
<212> DNA
<213> Equus caballus

<220>
<221> misc_feature

UCD011.C.WO.ST25

<223> MSTN_11 forward primer

<400> 23

ttttgaaact gttgtgtcct g

21

<210> 24

<211> 20

<212> DNA

<213> Equus caballus

<220>

<221> misc_feature

<223> MSTN_11 reverse primer

<400> 24

tcataattgc gtttggttgc

20

<210> 25

<211> 23

<212> DNA

<213> Equus caballus

<220>

<221> misc_feature

<223> MSTN_12 forward primer

<400> 25

gcaaatgttc aaatgaccta aac

23

<210> 26

<211> 20

<212> DNA

<213> Equus caballus

<220>

<221> misc_feature

<223> MSTN_12 reverse primer

<400> 26

tgtgtgatt ctgtgtggtc

20

<210> 27

<211> 23

<212> DNA

<213> Equus caballus

<220>

<221> misc_feature

<223> MSTN_13 forward primer

<400> 27

tgaagattta gtgttttgtc tcc

23

<210> 28

<211> 20

<212> DNA

<213> Equus caballus

UCD011.C.WO.ST25

<220>
 <221> misc_feature
 <223> MSTN_13 reverse primer

<400> 28
 cgagattcat tgtggagcag

20

<210> 29
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_14 forward primer

<400> 29
 gagacaactt gccacaccag

20

<210> 30
 <211> 22
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN_14 reverse primer

<400> 30
 tgcccttgga ataacaatga ag

22

<210> 31
 <211> 403
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN EquCab2.0 66493737_T/C SNP

<220>
 <221> variation
 <222> (202)..(202)
 <223> y is t or c

<400> 31
 agctaagcaa gtaattagca caaaaatttg aatgttatat tcaggctatc tcaaaagtta 60
 gaaaatactg tcttttagagc caggctgtca ttgtgagcaa aatcactagc aatttcitrt 120
 attttgggtc cccaagattg ttataaata aggtaaatct actccaggac tatttgatag 180
 cagagtcata aaggaaaatt aytfggtgca ttataacctg attacttaat aaggagaaca 240
 atattttgaa actgttgtgt cctgtttaaa gtagataaag cactgggtaa agcaggatcg 300
 cagacacatg gcacagaatc ttccgtgtca tgccttctct gtgaagggtg ctgtctccct 360
 ttcccttgagt gtagttaatga actgactgca aaaagaatat atg 403

<210> 32

UCD011.C.WO.ST25

<211> 401
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> COX4I1 EquCab2.0 32772871_C/T SNP

<220>
 <221> variation
 <222> (181)..(181)
 <223> m is a or c

<220>
 <221> variation
 <222> (201)..(201)
 <223> y is c or t

<400> 32
 tcaggctctca gtcgcaccag agctggatgg agccagcgca gcttcatctc tcagtggctg 60
 ggagtggggt gcagggtggt cctcacacaa gatgggcacc tccctctctg gctccatccc 120
 aggactgttt cccaggtttg ggaactggc tcgcattagc cgagtggcgt gagccggaat 180
 mtgatttact cacagtgcgc ygtgcttggg ggggaacgac tccctgctt tgtacagcac 240
 cctgcgtttc cagtggttgg tigtctggtc actagcttt tatcaagaga tagtaragtg 300
 aaggtraggc caagaaaaag ggaactctga ctgtcagag ggctgtttga actgtatggg 360
 gaetgcatct cgataaccag gattctgggt ctcagaccc a 401

<210> 33
 <211> 403
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN EquCab2.0 66493525_T/G SNP

<220>
 <221> variation
 <222> (202)..(202)
 <223> k is t or g

<400> 33
 tggaggaaca tccacttaga attctttggg aatctgagta gttacactta ctgagcagct 60
 gtaccaatca gtctggaaga aggaaccctt ccccaggcc tgaattacct ggggacaaga 120
 cacactgagg aactaactga gccctgggaa ttaagagaaa atatagtraca tctgtrtgt 180
 ttggcctttg gaatagcctt tkaaagggaac aaagctaagc aagtaattag cacaaaaatt 240
 tgaatgttat attcaggcta tctcaaaagt tagaaaatac tgtctttaga gccaggctgt 300
 cattgtgagc aaaatcacta gcaatttctt ttattttggg tcccaagat tgtttataaa 360
 taaggtaaat ctactccagg actatrtgat agcagagtca taa 403

<210> 34
 <211> 403

UCD011.C.WO.ST25

<212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN EquCab2.0 66493582_T/G SNP

<220>
 <221> variation
 <222> (202)..(202)
 <223> k is t or g

<400> 34
 gctgtaccaa tcagtctgga agaaggaacc ctccccccag gcctgaatta cctggggaca 60
 agacacactg aggaactaac tgagcctcgg gaattaagag aaaatatagt acatctgtta 120
 tgttttggct ttgggaatagc cttttaaagg aacaaagcta agcaagtaat tagcacaaaa 180
 atttgaatgt tatattcagg ckatctcaaa agttagaaaa tactgtcttt agagccaggc 240
 tgtcattgtg agcaaaatca ctagcaattt cttttatttt ggttccccaa gattgtttat 300
 aaataaggta aatctactcc aggactatit gatagcagag tcataaagga aaattatttg 360
 gtgcattata acctgattac ttaataagga gaacaatatt ttg 403

<210> 35
 <211> 403
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN EquCab2.0 66493745_A/G SNP

<220>
 <221> variation
 <222> (202)..(202)
 <223> r is a or g

<400> 35
 aagtaattag cacaaaaatt tgaatgttat attcaggcta tctcaaaagt tagaaaatac 60
 tgtctttaga gccaggctgt cattgtgagc aaaatcacta gcaatttcit ttatttttgt 120
 rccccaagat tgtttataaa taaggtaaat ctactccagg actattttgat agcagagtca 180
 taaaggaaaa ttattttgtg ctttataacc tgattactta ataaggagaa caatattttg 240
 aaactgttgt gtcctgttta aagtagataa agcactgggt aaagcaggat cgcagacaca 300
 tggcacagaa tcttccgtgt catgccttct ctgtgaaggt gtctgtctcc ctttccttga 360
 gtgtayttat gaactgactg caaaaagaat atatgaata tat 403

<210> 36
 <211> 403
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature

UCD011.C.WO.ST2S

<223> MSTN EquCab2.0 66493775_A/G SNP

<220>

<221> variation

<222> (202)..(202)

<223> r i s a o r g

<400> 36

attcaggcta tctcaaaagt tagaaaaatac tgcctttaga gccaggctgt cattgtgagc 60

aaaatcacta gcaatttctt ttatttttgt tccccaagat tgrttataaa taaggtaaat 120

cractccagg actattttgat agcagagaca taaaggaaaa ttatttttgt cattataacc 180

tgattaccta ataaggagaa cratatatttg aaactgttgt gtcctgttta aagtagataa 240

agcactgggt aaagcaggat cgcagacaca tggcacagaa tcttcgtgt catgccttct 300

ctgtgaagggt gtctgtctcc ctctcttga gtgtagttat gaactgactg caaaaagaat 360

atatgaaata tattatcttt cagaagcaat atagatacta cag 403

<210> 37

<211> 403

<212> DNA

<213> Equus caballus

<220>

<221> misc_feature

<223> MSTN EquCab2.0 66494218_A/C SNP

<220>

<221> variation

<222> (202)..(202)

<223> m i s a o r c

<400> 37

aggagattat taagcaatgt gccctgcttg aaatgtgcac cccgggtgct ctcaacaata 60

gtactatggt caagggtgaa graggactct gagctataac ctctttgatt aaaatgttta 120

tttattaggt attttatgat aattagctca tgattatcat targctatgt ttacttcatc 180

atttttctta ctaatacatt amattttaaa aaatatTTTT cctaataccc aggggaataa 240

ctttcaaaat ctaatatgtt aatttgrgaa gaacataaaa acactatgag aaatagtitt 300

gagtaacaga agtcattttg gtgttcagca aatgctcaaa tgacctaac gtctacaaat 360

ttcttcttc tctattatta gtgaaaaaaa ctgtttatta taa 403

<210> 38

<211> 20

<212> DNA

<213> Equus caballus

<220>

<221> misc_feature

<223> TTN forward primer

<400> 38

gcattgacaca actggaaagc 20

GCCD011.C.WO.ST25

<210> 39
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> TTN reverse primer

<400> 39
 aactttgccc tcatcaatgc

20

<210> 40
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN1-2 forward primer

<400> 40
 tgacagcagt gatggctctt

20

<210> 41
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN1-2 reverse primer

<400> 41
 ttgggttttc ctccacttg

20

<210> 42
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN2-3 forward primer

<400> 42
 ttcccaagac caggagaaga

20

<210> 43
 <211> 20
 <212> DNA
 <213> Equus caballus

<220>
 <221> misc_feature
 <223> MSTN2-3 reverse primer

<400> 43
 cagcatcgag attctgtgga

20