

(19) **DANMARK**

(10) **DK/EP 3473296 T3**



(12) **Oversættelse af
europæisk patentskrift**

Patent- og
Varemærkestyrelsen

-
- (51) Int.Cl.: **A 61 N 1/30 (2006.01)** **A 61 K 39/00 (2006.01)** **A 61 M 37/00 (2006.01)**
A 61 N 1/04 (2006.01) **A 61 N 1/32 (2006.01)** **C 12 M 1/42 (2006.01)**
C 12 N 15/87 (2006.01)
- (45) Oversættelsen bekendtgjort den: **2020-11-09**
- (80) Dato for Den Europæiske Patentmyndigheds bekendtgørelse om meddelelse af patentet: **2020-08-05**
- (86) Europæisk ansøgning nr.: **18190065.5**
- (86) Europæisk indleveringsdag: **2012-06-28**
- (87) Den europæiske ansøgnings publiceringsdag: **2019-04-24**
- (30) Prioritet: **2011-06-28 US 201161502198 P**
- (62) Stamansøgningsnr: **12846201.7**
- (84) Designerede stater: **AL AT BE BG CH CY CZ DE DK EE ES FI FR GB GR HR HU IE IS IT LI LT LU LV MC MK MT NL NO PL PT RO RS SE SI SK SM TR**
- (73) Patenthaver: **Inovio Pharmaceuticals, Inc., 660 W. Germantown Pike , Suite 110, Plymouth Meeting, PA 19462, USA**
- (72) Opfinder: **BRODERICK, Kate, c/o Inovio Pharmaceuticals, Inc., 660 W. Germantown Pike, Suite 110, Plymouth Meeting, PA 19462, USA**
KEMMERRER, Stephen, c/o Inovio Pharmaceuticals, Inc., 660 W. Germantown Pike, Suite 110, Plymouth Meeting, PA 19462, USA
MCCOY, Jay, c/o Inovio Pharmaceuticals, Inc., 660 W. Germantown Pike, Suite 110, Plymouth Meeting, PA 19462, USA
- (74) Fuldmægtig i Danmark: **Patrade A/S, Ceresbyen 75, 8000 Århus C, Danmark**
- (54) Benævnelse: **MINIMALT INVASIV DERMAL ELEKTROPORERINGSSENHED**
- (56) Fremdragne publikationer:
EP-A1- 1 163 024
WO-A1-2011/109406
CN-A- 101 563 132
US-A1- 2001 023 330
US-A1- 2005 048 651
US-A1- 2007 185 432

DESCRIPTION

FIELD OF THE INVENTION

[0001] The present invention relates to an electroporation device that is capable of delivering one or more plasmid vaccines simultaneously at spatially separate sites in a tolerable manner.

BACKGROUND

[0002] A major obstacle to effective vaccination via antigenic plasmids is the need of the DNA vaccine to be delivered intracellularly. The delivery of naked DNA through a standard intramuscular injection is notoriously inefficient outside of rodent models. Historically, this has led to an inability to achieve robust immune responses in large mammals and humans. Several strategies have been developed to enhance the expression of DNA-based vaccines, such as codon-optimization, RNA optimization, leader sequence addition and the development of optimized consensus sequences. These optimization strategies can lead to improved, cross-reactive immune responses. The addition of co-delivered gene-based molecular adjuvants is another area where an augmentation of resulting immune responses frequently occurs. Despite the improvements in vector design and use of molecular adjuvants, there is still a clear need for an efficient method of administration of DNA vaccines that results in high level expression of the plasmid in the desired cell type of the desired tissue, most commonly, muscle, tumor or skin.

[0003] Drug delivery to dermal tissue (intradermal) is an attractive method in a clinical setting for a number of reasons. The skin is the largest organ of the human body, the most accessible, and easily monitored, as well as being highly immuno-competent. However, the impervious, barrier function of the skin has been a major obstacle to efficient trans-dermal drug delivery.

[0004] Human skin comprises approximately 2 m² in area and is around 2.5 mm thick on average, making it the largest organ of the human body. Conventionally, the skin has two broad tissue types, the epidermis and the dermis. The epidermis is a continually keratinizing stratified epithelium. The outermost layer of skin is the stratum corneum (SC) and functions as the primary barrier. The SC is a 15-30 cell thick layer of non-viable but biochemically active corneocytes. The other three strata of the epidermis (S. granulosum, S. spinosum, S. basale) all contain keratinocytes at different stages of differentiation as well as the immune Langerhans cells and dermal dendritic cells.

[0005] Both physical and chemical methods for trans-dermal drug delivery and gene delivery have been detailed by groups worldwide. Iontophoresis, lipid delivery and gene gun are such examples. A physical method to temporarily increase skin permeability is electroporation ("EP"). Electroporation involves the application of brief electrical pulses that result in the creation of aqueous pathways within the lipid bi-layer membranes of mammalian cells. This allows the passage of large molecules, including DNA, through the cell membrane which would otherwise be less permeable. As such, electroporation increases the uptake or the extent to which drugs and DNA are delivered to their target tissue.

[0006] Although the precise mechanism by which electroporation enables cell transformation has not been elucidated, a proposed theoretical model involves a poration event due to the destabilization of the membrane, followed by the electrophoretic movement of charged molecules into the cell. For electroporation to occur, the formation of pores requires that a threshold energy be achieved and the movement produced by the electrophoretic effect depends upon both the electric field and the pulse length.

[0007] In the case of DNA vaccines, electroporation has been shown to quantitatively enhance immune responses, increase the breadth of those immune responses as well as improve the efficiency of dose. More recently, the DNA-EP platform has been successfully translated into the human clinical setting and has demonstrated significantly improved immune responses in several vaccine studies. Therefore, there has developed a need for a dermal electroporation device that would be considered tolerable, user-friendly and easily amenable to mass production, while continuing to achieve high transfection rates resulting in robust immune responses.

[0008] Although a number of intramuscular devices have now successfully entered clinical trials, the procedure is generally considered invasive and painful. To be considered amenable to mass vaccination, especially in a pediatric setting, a solution for a more tolerable electroporation method is needed. Accordingly, an effective dermal electroporation device that is capable of delivering a multi-agent DNA vaccine in a tolerable manner is desirable. US 2007/185432 and US 2001/023330 address electroporation devices.

SUMMARY OF THE INVENTION

[0009] The present disclosure is directed to a device for electroporating and delivering one or more antigens. The device comprises a housing, a plurality of electrode arrays projecting from the housing, each electrode array including at least one electrode, a pulse generator electrically coupled to the electrodes, a programmable microcontroller electrically coupled to the pulse generator, and an electrical power source coupled to the pulse generator and the microcontroller. The electrode arrays define spatially separate sites. The electrodes are configured to deliver an electroporating pulse to cells of epidermal tissues. The microcontroller is configured to adjust parameters of the electroporating pulse of each electrode array independently.

[0010] The present description is also directed to a method of electroporating and delivering one or more antigens to cells of epidermal tissues using the device described herein. In embodiments, the antigens generally include DNA vaccine plasmids, peptides, small molecules, and combinations thereof. The method comprises administering the one or more antigens to the cells of the epidermal tissues, contacting the epidermal tissues with the electrodes, and delivering the electroporating pulses.

BRIEF DESCRIPTION OF THE DRAWINGS

[0011]

Figures 1(A), 1(B), and 1(C) are perspective views of a minimally invasive device (MID) for EP

according to an embodiment.

Figure 2 is a schematic illustration of an electrical system of the embodiment of Figs. 1(A), 1(B), and 1(C).

Figures 3(A), 3(B), and 3(C) are perspective views of a MID for EP according to another embodiment.

Figure 4 is a schematic illustration of an electrical system of the embodiment of Figs. 3(A), 3(B), and 3(C).

Figures 5(A), 5(B), and 5(C) are fluorescent micrographs of green fluorescent protein (GFP) expression following injection and electroporation of plasmids using an MID with (A) stainless steel electrodes or (B) gold electrodes or with (C) injection only and no electroporation as a control. GFP pixel intensity was calculated (D).

Figures 6(A) and 6(B) are views of a MID for EP as (A) a side view of a 1 mm-spaced array (top) and 1.5 mm-spaced array (bottom) electrode hand piece, and as (B) a close up of the face of a 1 mm-spaced array showing all 25 needle electrodes.

Figures 7(A), 7(B), and 7(C) are fluorescent micrographs of GFP expression following injection and electroporation of plasmids using an MID with 1.5 mm or 1 mm electrode spacing and (A) stainless steel electrodes or (B) gold electrodes. A set with injection only and no electroporation was used as a control. GFP pixel intensity was calculated (C).

Figure 8 shows graphs of current and resistance for MIDs with 1.5 mm or 1 mm electrode spacing. Electrodes were either gold or stainless steel (SS) in composition.

Figure 9 shows graphs of current and impedance for MIDs with 1.5 mm or 1 mm electrode spacing at 5 or 15 volts.

Figure 10(A) shows fluorescent micrographs of GFP expression following injection and electroporation of plasmids using an MID with 1.5 mm or 1 mm electrode spacing at 5 or 15 volts. GFP pixel intensity was calculated (B).

Figure 11(A) shows fluorescent micrographs of GFP expression following injection and electroporation of different concentrations (0.5, 0.25, and 0.1 mg/mL) of plasmid using an MID with 1.5 mm or 1 mm electrode spacing at 15 volts. GFP pixel intensity was calculated (B).

Figures 12(A), 12(B), and 12(C) are fluorescent micrographs of lymphocyte staining following injection and electroporation of plasmids using an MID. Fig. 12(A) is a skin biopsy in an untreated animal at 20x magnification. Fig. 12(B) is a skin biopsy from an animal treated with plasmid expression GFP at 20x magnification. Fig. 12(C) is the sample in Fig. 12(B) but at 40x magnification.

Figure 13(A) is a perspective view of a MID for EP according to an embodiment, Figure 13(B) is a photograph of an MID with a 4x4 array and 1.5 mm spacing, Figure 13(C) is a photograph of an MID with a 5x5 array and 1.0 mm spacing, and Figure 13(D) is a photograph showing a side by side comparison of the MIDs in Figs. 13(B) and 13(C).

Figure 14 is a fluorescent micrograph showing the GFP expression following an intradermal administration of a reporter gene plasmid and EP with the dual-head MIDs of Figure 13(B), compared to the GFP expression following the intradermal injection alone (no EP).

DETAILED DESCRIPTION

[0012] The present invention is directed to an electroporation device that can provide heterogeneous intradermal delivery of antigens to a mammal. One or more antigens can be delivered simultaneously at spatially separated sites in a tolerable manner via a minimally invasive device (MID) having a plurality of electrode arrays. The electrode arrays are configured to vary the electroporating pulse from array to array. For example, each electrode array can be independently and selectively activated or controlled. Thus, the MID enables a heterogeneous delivery of antigens. Dermal electroporation via this MID reflects a clinically acceptable method to effectively deliver vaccines to the skin of a subject. This device is amenable to delivering multiple vaccines in multiple forms (nucleic acid, protein, small molecules, or a combination thereof) simultaneously while removing potential concerns with immune-interference resulting from the co-delivery of multiple antigens. It also allows the ability to deliver higher doses of a single antigen during a single treatment.

1. Definitions

[0013] The terminology used herein is for the purpose of describing particular embodiments only and is not intended to be limiting. As used in the specification and the appended claims, the singular forms "a," "and" and "the" include plural references unless the context clearly dictates otherwise.

[0014] For the recitation of numeric ranges herein, each intervening number therebetween with the same degree of precision is explicitly contemplated. For example, for the range of 6-9, the numbers 7 and 8 are contemplated in addition to 6 and 9, and for the range 6.0-7.0, the number 6.0, 6.1, 6.2, 6.3, 6.4, 6.5, 6.6, 6.7, 6.8, 6.9, and 7.0 are explicitly contemplated.

[0015] The term "electroporation" as used herein refers to the use of a transmembrane electric field pulse to induce microscopic pathways (pores) in a bio-membrane; the pores' presence allows biomolecules such as plasmids, oligonucleotides, siRNA, drugs, ions, and water to temporarily pass from one side of the cellular membrane to the other.

[0016] The term "minimally invasive" as used herein refers to a limited penetration by the needle electrodes of the provided EP device, and can include noninvasive electrodes (or nonpenetrating needles). Preferably, the penetration is to a degree that penetrates through stratum corneum, and preferably enters into the outermost living tissue layer, the stratum granulosum, but does not penetrate the basal layer. The penetration depth preferably does not exceed 0.1 mm, and in some embodiments the penetration depth ranges from about 0.01 mm to about 0.04 mm to break through stratum corneum. This can be accomplished using an electrode that has a trocar end ground to provide a sharp point that allows penetration through the stratum corneum but avoids a deeper penetration.

[0017] The terms "tolerable" and "nearly painless" are used interchangeably herein, and when referring to EP, mean a substantially lower level of pain associated with EP than with typically available EP devices. More specifically, a tolerable or near painless EP is the result of combination of using the

device described herein, avoiding EP of muscle, along with delivering low electrical fields to the epidermal layers between the stratum corneum and the basal layers. Preferably the electrical fields will comprise low voltage levels, for example from 0.01 V to 70 V, or from 1 V to 15 V. When measured using a visual analog scale, subjects experiencing EP with the device described herein according to the methods provided herein experience pain levels that are within 20% (of the full scale) from their painless or pain free score, or for example, within 2 points, with 0 - 10 full scale, and preferably within 10% from their painless score.

[0018] The term "substantially prevents damage" is used herein to refer to an amount of energy that is delivered by the described devices to the target cells to electroporate said cells and cause minimal discernible damage to same cells. Preferably, there is no discernable macroscopic histological damage or alteration to such cells.

2. Minimally Invasive Device

[0019] The present invention is directed to a minimally invasive device (MID) having a plurality of electrode arrays that are configured to vary the electroporating pulse from array to array. **Figs. 1(A), 1(B), and 1(C)** disclose an MID **100** having a plurality of electrode arrays for electroporating and delivering one or more antigens. The MID **100** comprises a housing **102** having a tip portion **104**, a plurality of electrode arrays **106, 108**, coupled to the tip portion **104**, each electrode array **106, 108** including electrodes **110** arranged in a square 4×4 pattern. In some embodiments, one or both of the tip portion **104** and the electrodes **110** can be detached from the rest of the MID **100**, e.g., for sterilizing after use so that the detached parts can be used again. Alternatively, one or both of the tip portion **104** and the electrodes **110** can be for a single use.

[0020] The electrode arrays **106, 108** define spatially separate sites. Although **Figs. 1(A), 1(B), and 1(C)** illustrate the MID **100** as including two electrode arrays **106, 108**, in other embodiments the MID **100** can include more than two electrode arrays, e.g., three or more, four or more, five or more, six or more, seven or more, eight or more, nine or more, or ten or more electrode arrays.

[0021] In some embodiments, each array **106, 108** can include a 4×4 array of electrodes **110** having a respective length of 0.1 mm, 0.2 mm, 0.3 mm, 0.4 mm, 0.5 mm, 1.0 mm, 1.5 mm, 2.0 mm, 2.5 mm, 3.0 mm, 3.5 mm, 4.0 mm, 4.5 mm, 5 mm, 6.0 mm, 7.0 mm, 8.0 mm, 9.0 mm, or 10.0 mm. Although in the illustrated embodiment each array **106, 108** includes a 4×4 array of electrodes **110**, i.e., 16 electrodes **110**, in other embodiments, each of the arrays **106, 108** can respectively include other numbers and/or patterns of electrodes **110**. For example, each of the arrays **106, 108** can respectively include electrodes **110** arranged in a pattern of 1×1, 1×2, 1×3, 1×4, 1×5, 1×6, 1×7, 1×8, 1×9, 1×10, 2×1, 2×2, 2×3, 2×4, 2×5, 2×6, 2×7, ×8, 2×9, 2×10, 3×1, 3×2, 3×3, 3×4, 3×5, 3×6, 3×7, 3×8, 3×9, 3×10, 4×1, 4×2, 4×3, 4×4, 4×5, 4×6, 4×7, 4×8, 4×9, 4×10, 5×1, 5×2, 5×3, 5×4, 5×5, 5×6, 5×7, 5×8, 5×9, 5×10, 6×1, 6×2, 6×3, 6×4, 6×5, 6×6, 6×7, 6×8, 6×9, 6×10, 7×1, 7×2, 7×3, 7×4, 7×5, 7×6, 7×7, 7×8, 7×9, 7×10, 8×1, 8×2, 8×3, 8×4, 8×5, 8×6, 8×7, 8×8, 8×9, 8×10, 9×1, 9×2, 9×3, 9×4, 9×5, 9×6, 9×7, 9×8, 9×9, 9×10, 10×1, 10×2, 10×3, 10×4, 10×5, 10×6, 10×7, 10×8, 10×9, 10×10, or multiples of 11-100 and any combination thereof. The patterns can be arranged in various shapes such as squares, triangles, rectangles, parallelograms, circles or any other geometric shape. The needle-shaped electrodes **110** can comprise gold, platinum, titanium, stainless steel, aluminum, or any other conductive metal. The electrodes can be coated or plated with a metal such as gold,

copper, platinum, silver, or any other conductive metal.

[0022] In some embodiments, each electrode **110** is needle-shaped. That is, the electrodes **110** each include a shaft **112** and a tapered tissue-penetrating or trocar end **114**. Although **Fig. 2(B)** illustrates the electrodes **110** as being generally cylindrical, in other embodiments, at least one of the electrodes **110** can assume any geometric form, including, but not limited to, a semi-cylindrical, a regular polyhedral, and an irregular polyhedral shape, derivatives thereof, and combinations thereof. The tissue-penetrating end **114** can facilitate the electrodes **110** penetrating through the stratum corneum and reaching the stratum granulosum. In some embodiments, the tissue-penetrating end **114** allows the electrode **110** to penetrate through the stratum corneum but avoids deep penetration. To this end, the tissue-penetrating end **114** can have a length of about 0.1 mm or less, or about 0.01 mm to about 0.04 mm.

[0023] The illustrated electrodes **110** are configured to deliver an electroporating pulse to cells of epidermal tissues. In some embodiments, the electroporating pulses are associated with an electrical field that substantially prevents damage in the cells of the epidermal tissues. In further embodiments, the electroporating pulses are associated with an electrical potential that is nearly painless as measured by a visual analog scale. The visual analog scale is essentially a 100-mm-long horizontal line on which 0 mm indicates no pain and 100 mm indicates the worst pain. Near painless is a score using the visual analog scale that produces a mean score of about 20 mm or less (within a 95% confidence interval), and preferably 10 mm or less (within a 95% confidence interval).

[0024] In some embodiments, adjoining electrodes **110** are spaced apart from one another at a distance of no more than about 1.5 mm. In further embodiments, adjoining electrodes **110** are spaced apart from one another at a distance of no more than about 1.0 mm. A shorter distance between the electrodes **110** means that the electrodes **110** are packed in a more compact manner, which can increase the efficacy of the MID **100** and therefore can be desirable. In some embodiments, each electrode **110** can be spaced apart from each adjacent electrode **110** at a distance of 150 mm or less, from 100 mm to 1.0 mm, from 50 mm to 1.0 mm, from 40 mm to 1.0 mm, from 30 mm to 1.0 mm, from 20 mm to 1.0 mm, from 10 mm to 1.0 mm, from 5.0 mm to 2.0 mm, from 5.0 mm to 1.0 mm, approximately 2.0 mm, approximately 1.5 mm, or approximately 1.0 mm.

[0025] Referring also to **Fig. 2**, pulse generators **116**, **118** are electrically coupled to respective electrode arrays **106**, **108**. In some embodiments, at least one of the pulse generators **116**, **118** can be the Elgen 1000 (Inovio Pharmaceuticals, Inc., Blue Bell, PA) pulse generator (not shown). In other embodiments, however, the electroporating pulse can be generated using other suitable mechanisms. In the illustrated embodiment, a programmable microcontroller **120** is electrically coupled to the pulse generators **116**, **118**. In response to an input condition/signal, the microcontroller **120** is capable of adjusting EP parameters of each electrode array **106**, **108** independently, depending on the usage requirements or preferences for each electrode array **106**, **108**. Thus, the microcontroller **120** is configured to vary the electroporating pulse from array to array. For example, the pulse voltage, current, duration, and quantity of the applied electrical pulses can be varied from array to array so as to vary the Joules per cm³ applied at each injection site. In some embodiments, the microcontroller **120** is configured to deliver the electroporating pulses substantially simultaneously. In the illustrated embodiment, each electrode array **106**, **108** is driven with a respective pulse generator **116**, **118**. The MID **100** also includes an electrical power source **122** coupled to the pulse generators **116**, **118** and the microcontroller **120** for providing electrical power. In the illustrated embodiment, the electrical

power source **122** is a high and low voltage supply, although other power sources performing the same function as the electrical power source **122** disclosed herein can be used instead.

[0026] In some embodiments, the electroporating pulse of each electrode array **106, 108** is associated with an electrical potential of 0.01 V to 70 V, 0.01 V to 50 V, 0.01 V to 40 V, 0.01 V to 30 V, 0.01 V to 20 V, 0.01 V to 15 V, 0.1 V to 70 V, 0.1 V to 50 V, 0.1 V to 40 V, 0.1 V to 30 V, 0.1 V to 20 V, 0.1 V to 15 V, 1V to 30V, 1V to 20V, 1 V to 15 V, 15V to 30V, or 15V to 30V. In further embodiments, the electrical potential is preferably low so that the EP is tolerable or nearly painless as measured by a visual analog scale, yet sufficiently high so as to effect transfection of the cells in the epidermal tissues. For example, the electrical potential can be 5V, 10V, 15 V, or in some embodiments 20V, when adjacent electrodes **110** of the MID **100** are spaced apart by 1.0 mm to 2.0 mm.

[0027] In some embodiments, each electroporating pulse of each electrode array **106, 108** is associated with an electrical current of 0.1 mA to 100 mA, 0.2 mA to 100 mA, 0.5 mA to 100 mA, 1 mA to 100 mA, 1 mA to 80 mA, 1 mA to 60 mA, 1 mA to 50 mA, 1 mA to 40 mA, 1 mA to 30 mA, 10 mA to 50 mA, 10 mA to 40 mA, 10 mA to 30 mA, 10 mA to 20 mA, or 10 mA to 15 mA, or approximately 10 mA, or in some embodiments approximately 20 mA. Like the electrical potential, the electrical current is preferably low so that the EP is tolerable or nearly painless as measured by a visual analog scale, yet sufficiently high so as to effect transfection of the cells in the epidermal tissues.

[0028] In some embodiments, the electroporating pulse of each electrode array **106, 108** is associated with a duration of from 5 ms to 250 ms, 10 ms to 250 ms, 20 ms to 250 ms, 40 ms to 250 ms, 60 ms, to 250 ms, 80 ms to 250 ms, 100 ms to 250 ms, 20 ms to 150 ms, 40 ms to 150 ms, 60 ms to 150 ms, 80 ms to 150 ms, 100 ms to 150 ms, 100 ms to 140 ms, 100 ms to 130 ms, 100 ms to 120 ms, 100 ms to 110 ms, or approximately 100 ms. In some embodiments, the duration is preferably short so that the EP is tolerably or nearly painless as measured by a visual analog scale, yet sufficiently long so as to effect transfection of the cells in the epidermal tissues.

[0029] In some embodiments, the microcontroller **120** is configured to adjust a respective quantity of electroporating pulses for each electrode array **106, 108** independently, and the number of electrical pulses can be 1 or more, 2 or more, 3 or more, 4 or more, 5 or more, 6 or more, 7 or more, 8 or more, 9 or more, 10 or more, 20 or more, 30 or more, 40 or more, 50 or more, 60 or more, 70 or more, 80 or more, 90 or more, or 100 or more. By increasing the quantity of electroporating pulses and reducing the energy per pulse, the amount of pain perceived or experienced by a subject can also be reduced as compared with fewer pulses at higher energy. Preferably, a lower number of pulses, which does not reduce immune response that is generate, is used as it results in less pain experienced by the subject. Furthermore, less pain and better tolerability results by using lower energy per pulse. In some embodiments, the quantity of electroporating pulses is about 1 to about 10, preferably about 1 to about 10, and more preferably about 1 to about 3. In some embodiments 3 pulses are used.

[0030] In some embodiments, the electrode arrays **106, 108** are spaced apart from one another at least by a distance so as to substantially prevent interference of the two antigens delivered by the two arrays **106, 108** when the electroporating pulses are delivered. Plasmid interference has been observed for a number of antigens when they are delivered sequentially at the same site in the skin. Though not wishing to be bound by a particular theory, this could be due to interference at either the transcriptional level (possible competition at the promoter, etc.) or the translational level (mis-folding

or dimerization at the protein level). The MID **100** having a plurality of spaced electrode arrays **106**, **108** could eliminate this interference effect, which can negatively affect the resulting immune response. Moreover, the MID **100** having a plurality of electrode arrays **106**, **108** could negate the need for two separate treatments, allowing a treated subject to experience one incident of treatment, thus reducing the pain that is experienced.

[0031] In some embodiments, the MID **100** includes switches (not shown) electrically coupled to each electrode array **106**, **108** for selectively activating each electrode array **106**, **108**. For example, the MID **100** can be triggered by a foot pedal or a trigger button, or any other trigger connected to an electrical circuit.

[0032] **Figs. 3** and **4** illustrate an MID **200** including a pulse generator **202** according to another embodiment. This embodiment employs much of the same structure and has many of the same properties as the embodiment of the MID **100** described above in connection with **Figs. 1(A)-1(C)**. Accordingly, the following description focuses primarily upon the structure and features that are different than the embodiment described above in connection with **Figs. 1(A)-1(C)**. Structure and features of the embodiment shown in **Figs. 3** and **4** that correspond to structure and features of the embodiment of **Figs. 1(A)-1(C)** are designated hereinafter with like reference numbers.

[0033] In this embodiment, the pulse generator **202** is powered by a battery **204**. In the illustrated embodiment, the battery **204** is within the housing **102**. As such, the MID **200** can be portable. The battery **204** can be a lithium ion, nickel metal hydride, lead acid, or nickel cadmium battery.

[0034] Referring to Fig. 4, the pulse generator **202** is a high and low voltage driver. The pulse generator **202** in this embodiment drives both electrode arrays **106**, **108**. A microcontroller **206** is electrically coupled to the pulse generator **202**. The microcontroller **206** is capable of adjusting the electroporation parameters of each electrode array **106**, **108** independently for example in response to an input condition/signal. The pulse generator **202** can generate pulses with EP parameters as adjusted by the microcontroller **206**, and, in cooperation with the battery **204**, amplify the generated pulses as needed.

3. Method of Electroporating and Delivering One or More Antigens

[0035] In an aspect, the MID **100**, **200** having a plurality of electrode arrays **106**, **108** described herein can be used in a method of electroporating and delivering one or more antigens, as discussed below, through the skin, organs, or other body parts of a subject. That is, the MID **100**, **200** can be used to apply a transmembrane electric field pulse that induces microscopic pathways (pores) in a bio-membrane, allowing the delivery of one or more antigens from one side of the cellular membrane to the other. The method can comprise the steps of administering the antigen to the cells of the epidermal tissues, contacting the epidermal tissues with the electrodes, and delivering an electroporating pulse to generate an immune response. The method can further comprise simultaneously delivering antigen to the cells and delivering an electroporating pulse to generate an immune response.

• Administering the Antigen to the Cells of the Epidermal Tissues

[0036] A plurality of antigens is first injected intradermally at spatially separated sites. In some embodiments, the antigen is intradermally delivered to the target tissue using the Mantoux technique, e.g., using a 29 gauge injection needle.

- **Contacting the Epidermal Tissues with the Electrodes**

[0037] Next, the epidermal tissues are penetrated with at least one electrode **110** at a depth of about 0.1 mm or less, or about 0.01 mm to about 0.04 mm. The injection sites and the tissue-penetration sites are preferably co-localized, in some examples, to facilitate co-localizing or centering the injection sites and the tissue-penetration sites, the epidermal tissues can be marked or indented before the intradermal injection.

- **Delivering an Electroporating Pulse**

[0038] Once the epidermal tissues are penetrated, the epidermal tissues are contacted with the electrodes **110** and an electroporating pulse is delivered. In some embodiments, the electroporating pulses are associated with an electrical field that substantially prevents damage to the cells of the epidermal tissues. In further embodiments, the electroporating pulses are associated with an electrical potential that is nearly painless as measured by a visual analog scale. For example, the electroporating pulses are associated with an electrical potential of about 1 volts to about 30 volts, or preferably about 15 volts to about 20 volts, an electrical current of about 1 mA to about 50 mA, or preferably about 10 mA to about 15 mA, and a duration ranging from about 80 ms to about 150 ms, or preferably 100 ms, or a combination thereof. These pulses can be delivered in a series, preferably 1-10 pulses, and more preferably 1-3 pulses.

[0039] A drawback to conventional intradermal delivery is a limitation to the volume that can be delivered to the skin. For a single needle injection, generally volumes no larger than 100-150 μ L can be delivered directly to the skin due to issues with dermal delamination. Because the antigen can only be produced at concentrations not in excess of 10mg/mL, the volume limitation can constrain the resulting dose.

[0040] In some embodiments, the MID **100, 200** can preferably deliver higher doses of a single vaccine. The use of the MID **100, 200** avoids the single injection volume limitation of 100-150 μ L. In some embodiments, significantly higher doses can be delivered simultaneously with a single treatment without any added discomfort to the patient. The ability to deliver higher doses could have significant positive effects on the resulting immune responses for specific vaccines. The use of a multi-head device also has the added benefit of directly targeting more cells than a single array device. Increased numbers of cells transfected with an antigen could result in improved immune responses through increased presentation to antigen presenting cells.

[0041] In some embodiments, the disclosed method can be administered to a subject such as a mammal. The mammal can be a human, monkey, dog, cat, livestock, guinea pig, mouse, or a rat. The livestock can be bovine, a pig, a sheep, or a cow, for example.

4. Antigen

[0042] The present invention is also directed to methods of delivering at least one antigen using the MID **100, 200** having a plurality of electrode arrays **106, 108**, as discussed above. The method can be directed to delivery of two or more antigens or a combination thereof using heterogeneous delivery by the MID **100, 200**. In certain embodiments, the MID **100, 200** described herein can be used to enhance delivery of an antigen. As used herein, "antigen" refers to any substance or organism that provokes an immune response (produces immunity) when introduced into the body.

[0043] In some embodiments, the antigen can be derived from an infectious agent or a self-antigen, e.g., a prostate cancer antigen such as prostate-specific antigen (PSA) or prostate-specific membrane antigen (PSMA). The particular antigen used is not critical. Antigens are known in the art and can be incorporated for use in the methods and compositions provided herein using any common method. Non-limiting lists of suitable antigens for use in the various aspects and embodiments described herein can be found in the literature, for example, BioCarb Chemicals Catalogue; and The Jordan Report: Accelerated Development of Vaccine 1995 NIH, Bethesda, Md., 1995, both of which are incorporated herein by reference. Antigens can include, but are not limited to, nucleic acids, peptides, small molecules, chemotherapeutics, immunotherapeutics, or combinations thereof. An antigen can include an immunogen.

[0044] In some embodiments, the antigen comprises a nucleic acid. Nucleic acid refers to a polynucleotide compound, which includes oligonucleotides, comprising nucleosides or nucleoside analogs that have nitrogenous heterocyclic bases or base analogs, covalently linked by standard phosphodiester bonds or other linkages. Nucleic acids can include RNA, DNA, chimeric DNA-RNA polymers, or analogs thereof. The DNA can be a plasmid expressing a particular antigen of interest. For example, the plasmid can be a SynCon influenza construct (Inovio Pharmaceuticals, Inc., Blue Bell PA).

[0045] In some embodiments, the antigen comprises a peptide. Peptides include any amino acid sequence. Peptides can be synthetic or isolated from a natural source. The peptide can be a protein. The peptide can be an antibody or antibody fragment.

[0046] In some embodiments, the antigen comprises a small molecule. Small molecules include organic and inorganic compounds.

[0047] In some embodiments the antigen comprises a chemotherapeutic. Chemotherapeutics can include cytotoxic or cytostatic drugs such as, for example, methotrexate (amethopterin), doxorubicin (adrimycin), daunorubicin, cytosinarabioside, etoposide, 5-fluorouracil, melphalan, chlorambucil, and other nitrogen mustards (e.g. cyclophosphamide), cis-platinum, vindesine (and other vinca alkaloids), mitomycin, bleomycin, purothionin (barley flour oligopeptide), macromomycin, 1,4-benzoquinone derivatives, and trenimon.

[0048] In some embodiments, the antigen includes a cytokine. Cytokine refers to a substance secreted by cells of the immune system that carry signals locally between cells. Cytokines include proteins, peptides, and glycoproteins. Cytokines include, but are not limited to, interferons, chemokines, TGF- β , TNF- α , and interleukins. Interleukins include IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7,

IL-8, IL-9, IL-10, IL-11, IL-12, IL-13, IL-14, IL-15, IL-15, IL-16, IL-17, IL-18, IL-19, IL-20, IL-21, IL-22, IL-23, IL-24, IL-25, IL-26, IL-27, IL-28, IL-29, IL-30, IL-31, IL-32, IL-33, IL-35, and IL-36. Cytokines can be derived from a human source or a transgenic non-human source expressing a human gene.

[0049] Antigens can include, but are not limited to, microbial antigens such as parasitic antigens, viral antigens, bacterial antigens, fungal antigens, cancer antigens, vaccine antigen additive drugs such as cocaine and nicotine derivatives, attenuated or killed bacteria, attenuated or killed virus, autoimmune antigens, or nonstructural protein antigens, or any combination thereof. In some embodiments, the antigen comprises at least one flu, autoimmune, cocaine, or cancer antigen.

[0050] In some embodiments an antigen comprises any antigen derived from bacterial surface polysaccharides which can be used in carbohydrate-based vaccines. Bacteria typically express carbohydrates on the cell surface as part of glycoproteins, glycolipids, O-specific side chains of lipopolysaccharides, capsular polysaccharides and the like. Non-limiting examples of suitable bacterial strains include *Streptococcus pneumoniae*, *Neisseria meningitidis*, *Haemophilus influenzae*, *Klebsiella* spp., *Pseudomonas* spp., *Salmonella* spp., *Shigella* spp., and *Group B* streptococci. In some embodiments any known bacterial carbohydrate epitope (e.g., those described in Sanders, et al. *Pediatr. Res.* 1995, 37, 812-819; Bartoloni, et al. *Vaccine* 1995, 13, 463-470; Pirofski, et al., *Infect. Immun.* 1995, 63, 2906-2911; U.S. Pat. No. 6,413,935; and International Publication No. WO 93/21948) can be used as an antigen in the compositions and methods herein described.

[0051] Some embodiments provide for an antigen that comprises a viral antigen. Non-limiting examples of viral antigens include those derived from HIV (e.g., gp120, nef, tat, pol), influenza, and West Nile Virus (WNV). In some embodiments, the antigen can comprise whole killed virus or attenuated virus.

[0052] Some aspects provide for a fungal antigen. Non-limiting examples of fungal antigens include those derived from *Candida albicans*, *Cryptococcus neoformans*, *Coccidioides* spp., *Histoplasma* spp., and *Aspergillus* spp.

[0053] Some embodiments provide for an antigen derived from a parasite. Non-limiting examples of parasitic antigens include those derived from *Plasmodium* spp., *Trypanosoma* spp., *Schistosoma* spp., *Leishmania* spp. and the like.

[0054] In some embodiments the antigen comprises a carbohydrate epitope. Non-limiting examples of carbohydrate epitopes that can be used in the aspects and embodiments described herein include: Gal α 1,4Gal β (for bacterial vaccines); GalNAc α (for cancer vaccines); Man β 1,2(Man β)_nMan β - (for fungal vaccines useful against, for example, *C. albicans*), wherein n is any integer, including zero; GalNAc β 1,4(NeuAc α 2,3)Gal β 1,4Glc β -O-ceramide (for cancer vaccines); Gal α 1,2(Ty α 1,3)Man α 1,4Rha α 1,3Gal α 1,2-(Ty α 1,3)Man α 4Rha- and Gal α 1,2(Abe α 1,3)Man α 1,4Rha α 1,3Gal α 1,2(Abe α 1,3)Man α 1,4Rha α 1,3Gal α 1,2(Abe α 1,3)Man α 1,4Rha (both of which are useful against, for example, *Salmonella* spp.). Description of other exemplary carbohydrate epitopes as antigens or immunogens and the synthesis thereof are described further in U.S. Pat. No. 6,413,935, incorporated herein by reference.

[0055] Other examples of antigens include, but are not limited to, those that produce an immune response or antigenic response to the following diseases and disease-causing agents: anthrax;

adenoviruses; *Bordetella pertussis*; Botulism; bovine rhinotracheitis; *Branhamella catarrhalis*; canine hepatitis; canine distemper; Chlamydiae; Cholera; coccidiomycosis; cowpox; cytomegalovirus; cytomegalovirus; Dengue fever; dengue toxoplasmosis; Diphtheria; encephalitis; Enterotoxigenic *Escherichia coli*; Epstein Barr virus; equine encephalitis; equine infectious anemia; equine influenza; equine pneumonia; equine rhinovirus; feline leukemia; flavivirus; Globulin; *Haemophilus influenza* type b; *Haemophilus influenzae*; *Haemophilus pertussis*; *Helicobacter pylori*; *Hemophilus* spp.; hepatitis; hepatitis A; hepatitis B; hepatitis C; herpes viruses; HIV; HTV-1 viruses; HIV-2 viruses; HTLV; Influenza; Japanese encephalitis; *Klebsiellae* spp. *Legionella pneumophila*; leishmania; leprosy; lyme disease; malaria immunogen; measles; meningitis; meningococcal; Meningococcal Polysaccharide Group A, Meningococcal Polysaccharide Group C; mumps; Mumps Virus; mycobacteria; *Mycobacterium tuberculosis*; *Neisseria* spp; *Neisseria gonorrhoeae*; *Neisseria meningitidis*; ovine blue tongue; ovine encephalitis; papilloma; parainfluenza; paramyxovirus; paramyxoviruses; Pertussis; Plague; *Pneumococcus* spp.; *Pneumocystis carinii*; Pneumonia; Poliovirus; Proteus species; *Pseudomonas aeruginosa*; rabies; respiratory syncytial virus; rotavirus; Rubella; Salmonellae; schistosomiasis; Shigellae; simian immunodeficiency virus; Smallpox; *Staphylococcus aureus*; *Staphylococcus* spp.; *Streptococcus pneumoniae*; *Streptococcus pyogenes*; *Streptococcus* spp.; swine influenza; tetanus; *Treponema pallidum*; Typhoid; Vaccinia; varicella-zoster virus; and *Vibrio cholerae*. The antigens or immunogens can include various toxoids, viral antigens and/or bacterial antigens such as antigens commonly employed in the following vaccines: chickenpox vaccine; diphtheria, tetanus, and pertussis vaccines; haemophilus influenzae type b vaccine (Hib); hepatitis A vaccine; hepatitis B vaccine; influenza vaccine; measles, mumps, and rubella vaccines (MMR); pneumococcal vaccine; polio vaccines; rotavirus vaccine; anthrax vaccines; and tetanus and diphtheria vaccine (Td) (see, e.g., U.S.

[0056] Patent No. 6,309,633).

[0057] In some embodiments, antigens can include any type of antigen associated with cancer such as, for example, tumor associated antigens (TSAs) (including antigens associated with leukemias and lymphomas) such as carcinoembryonic antigen, prostatic acid phosphatase, PSA, PSMA, and the like, and antigens that are associated with agents that can cause cancer (e.g., tumorigenic viruses such as, for example, adenovirus, HBV, HCV, HTLV, Kaposi's sarcoma-associated herpes virus, HPV (Gardasil), and the like).

[0058] Antigens can include combinations of antigens such as combinations of peptides, polysaccharides, lipids, nucleic acids, and the like. Antigens can include glycoproteins, glycolipids, lipoproteins, lipopolysaccharides, and the like.

[0059] Antigens that are used to carry out the disclosed EP methods include those that are derivatized or modified in some way, such as by conjugating or coupling one or more additional groups thereto to enhance function or achieve additional functions such as targeting or enhanced delivery thereof, including techniques known in the art such as, for example, those described in U.S. Patent No. 6,493,402 to Pizzo et al. (α -2 macroglobulin complexes); U.S. Patent No. 6,309,633; U.S. Patent No. 6,207,157; and U.S. Patent No. 5,908,629.

[0060] Illustrative examples of the MID 100, 200 and the method of using the MID 100, 200 are described in greater detail below.

EXAMPLES

Example 1

Effect of Electrode Composition on Transfection Efficiency

[0061] To address the effect of electrode material on reporter gene localization, transfection efficiency was compared for two minimally invasive devices (MID) with different electrode compositions (gold and stainless steel). This comparison was to assess whether a cheaper alternative (stainless steel) to gold electrodes could be used while still maintaining transfection efficacy. The electrode composition was easily tested, because the gold-plated electrodes were easily removed from their sockets and replaced with stainless steel electrodes of the same gauge and length. This produced an identical electrode head differing only in the electrode composition.

[0062] The experimental outline is detailed in Table 1.

Table 1

Electrode Composition	DNA Delivered	DNA Concentration (mg/ml)	Number of Treatment Sites	Biopsy Removal Time (hours)	Number of Animals Required	Final Analysis
Gold-plated	pgWIZ-GFP	0.5	10	12, 24, 48	4	Gross Visualization/Histology
		1	10	12, 24, 48		
		2	10	12, 24, 48		
Stainless steel	pgWIZ-GFP	0.5	10	12, 24, 48	4	Gross Visualization/Histology
		1	10	12, 24, 48		
		2	10	12, 24, 48		

[0063] A series of *in vivo* expression localization studies were completed. All *in vivo* experiments were conducted in Hartley guinea pigs (Charles River Laboratories, Wilmington, MA), which are considered an excellent model for dermatologic applications. All experiments were conducted under institutional IACUC protocols. All animal experiments were conducted in accordance with U.S. Department of Defense (DoD) 3216.1 "Use of Laboratory Animals in DoD Programs," 9 CFR parts 1-4 "Animal Welfare Regulations," National Academy of Sciences Publication "Guide for the Care & Use of Laboratory Animals," as amended, and the Department of Agriculture rules implementing the Animal Welfare Act (7 U.S.C. 2131-2159), as well as other applicable federal and state laws and regulations and DoD instructions. All animal treatments were carried out under anesthesia.

[0064] A plasmid expressing the reporter gene GFP was injected intradermally (0.5, 1, and 2 mg/mL) to guinea pig skin. Immediately following injection, the skin was electroporated at the injection site using a MID device with either gold or stainless steel electrodes. Animals were sacrificed three days post treatment. Skin was excised and visualized under a fluorescent microscope. High resolution photographs were taken and subsequently analyzed for pixel intensity using standard software (Adobe Photoshop CS5). The level of expression was calculated through pixel counts of pre-defined treatment areas. A "gated region" of electrode contact for pixel analysis was established on the presumption that transfection occurs only where the electric field is applied and that the electric field is formed only where the electrodes are in direct contact with the skin. The distance between the first and fourth electrode in the MID device was 4.5 mm. As such, the 'ruler tool' in Adobe Photoshop CS5 was used to isolate a 4.5 mm² region, which was defined as approximately 95 pixels in length. Adobe Photoshop CS5 recognized pixel intensities ranging from 0-255 (darkest-brightest) in three different channels (Red, Green, Blue). Since positive GFP signal would predominate in the green channel, pixel analysis was restricted to this channel. The CS5 version of Adobe Photoshop was able to automatically calculate mean and median pixel intensity of the selected region. Since the distribution of pixel intensity was not symmetrical in most cases, the median was deemed to give a better representation of central tendency for the histogram. To ensure accurate results, pooled data from multiple treatment sites on multiple animals was analyzed.

[0065] Results are shown in **Figure 5**. Results were also compared to GFP expression following intradermal injection without subsequent electroporation. There was no statistically significant difference between the gold and stainless steel groups (P value < 0.05 between both treatment groups and the ID injection-only control). The results suggested that the cheaper stainless steel electrode was as effective as the gold electrode at eliciting reporter gene expression.

Example 2

Effect of Electrode Spacing on Transfection Efficiency

[0066] To assess the effect of electrode spacing on transfection efficiency and reporter gene localization, a 1 mm spaced circuit board was created and fitted in a head-piece housing and compared with a similar MID with a 1.5 mm spaced circuit board. To ensure that the surface treatment area remained the same between the two hand pieces at the different spacings, an additional row of electrodes was added to the 1 mm spaced circuit board. Therefore, a 1.5 mm spacing hand piece with 4x4 rows of electrodes (16 electrodes) was compared to a 1 mm spacing hand piece with 5x5 rows of electrodes (25 electrodes). As such, each hand piece had an approximate treatment surface area of approximately 4-4.5 mm². **Figure 6A** shows a photograph of both hand pieces from a side perspective. The top hand piece is the 1 mm spacing, and the bottom hand piece is the 1.5 mm spacing. **Figure 6B** shows a close-up view of the face of the 1 mm hand piece.

[0067] A series of *in vivo* expression localization studies were completed, as described in Example 1. Specifically, following an intradermal injection of a known dose (0.5, 1, and 2 mg/mL) of plasmid DNA expressing the reporter gene GFP into guinea pig skin, the 1 mm or 1.5 mm prototype device was used to almost immediately, within 10 seconds after injection, electroporate the resulting injection

bubble. Electrodes were either gold or stainless steel in composition. Animals were sacrificed three days post treatment. Skin was excised and visualized under a fluorescent microscope. High resolution photographs were taken and subsequently analyzed for pixel intensity using standard software (Adobe Photoshop CS5). The level of expression was calculated through pixel counts of pre-defined treatment areas. To ensure accurate results, pooled data from multiple treatment sites on multiple animals was analyzed. Expression of the GFP was monitored over different time periods (12, 24, and 48 hours) to allow assessment of expression kinetics.

[0068] The experimental outline is detailed in Table 2.

Table 2

Electrode Spacing	DNA Delivered	DNA Concentration (mg/ml)	Number of Treatment Sites	Biopsy Removal Time (hours)	Number of Animals Required	Final Analysis
1 mm	pgWIZ-GFP	0.5	10	12, 24, 48	4	Gross Visualization/ Histology
		1	10	12, 24, 48		
		2	10	12, 24, 48		
1.5 mm	pgWIZ-GFP	0.5	10	12, 24, 48	4	Gross Visualization/ Histology
		1	10	12, 24, 48		
		2	10	12, 24, 48		

[0069] Results are shown in **Figure 7**. Results are shown for 6 treatments for each condition. Results were also compared to GFP expression following intradermal injection without subsequent electroporation. There was no statistically significant difference between results for the device with the 1.5 mm spaced circuit board and the 1 mm spaced circuit board (P value < 0.05 between the treatment groups and the ID injection-only control). The results are representative of multiple experiments and demonstrated that successful, robust transfection was achieved with a minimally invasive device electroporation (MID EP) using either 1.5 mm or 1 mm electrode spacing.

[0070] These results suggested that electrode spacing does not impact GFP expression in skin because no visible difference (as determined by eye and quantifiably through pixel counting) was observed between the two electrode spacings. Thus, electroporation with MID EP using either 1.5 mm or 1 mm electrode spacing resulted in robust reporter gene expression.

Example 3

Effect of Electrode Spacing on Current

[0071] A device as described herein can have the capacity to capture and store all electrical parameters real time as they occur during each electroporation pulse. A series of *in vivo* expression localization studies were completed, as described in Example 1, to examine current and voltage for electroporation with devices of different electrode spacing and composition. While the applied voltage

remained constant (15 volts), the impedance (resistance) and current delivered for each treatment was examined for each electrode spacing and each electrode composition.

[0072] **Figure 8** shows both the resulting impedance (resistance in Ohms) and Current (in milli Amps).

[0073] The current was approximately three times greater in the 1 mm hand piece (average 85 mA) compared to the 1.5 mm hand piece (average 23 mA), yet the applied voltage was the same across all conditions. The increased current in the 1.5 mm hand piece resulted in a large reduction (approximately 75%) in the impedance of the tissue. From the perspective of producing a tolerable dermal device, increased current can be problematic by causing more pain or sensation to the patient. Increased current can cause more pain or sensation in a patient, and thus, increased current can be problematic in producing a tolerable dermal device. These results suggested that while the electrode spacing did not appear to impact the resulting GFP expression, either the spacing or the presence of the additional electrodes can impact the current flow and, as such, impact the impedance of the tissue.

[0074] To further address the issue of increased current, it was investigated whether the applied voltage could be reduced by a third (to 5 volts) and still result in 10-20 mA current. Electrodes were stainless steel. Results are shown in **Figure 9**. The results suggested that additional electrodes, and not the actual electrode spacing, affected the resulting current.

[0075] To assess whether a reduced voltage affected the transfection efficacy, a DNA plasmid expressing the reporter gene GFP was delivered intradermally to guinea pig skin and immediately followed with electroporation using a MID device with either 1.5 mm (4x4) electrode spacing or 1 mm (4x4 or 5x5) electrode spacing at 15 or 5 volts. The animals were sacrificed three days post treatment. The skin was excised and visualized under a fluorescence microscope. High resolution photographs were taken and subsequently analyzed for pixel intensity using standard software (Adobe Photoshop CS5).

[0076] Results are shown in **Figure 10**. These results suggested that the input voltage can be reduced on larger electrode hand pieces while still maintaining transfection efficacy. As such, the results suggested that robust transfection can be achieved with a larger array while maintaining pain-free and low voltage parameters. It is likely that in a human clinical device, at least 25 electrodes would be required to maintain optimal coverage depending on the usage requirements or preferences, e.g., the injection volume.

Example 4

Effect of Plasmid Concentration on Transfection Efficiency

[0077] The effect of lower concentrations of plasmid expressing the reporter gene GFP on transfection efficiency was examined with devices of different composition and electrode spacing.

[0078] A series of *in vivo* expression localization studies were completed, as described in Example 1.

A plasmid expressing the reporter gene GFP was injected intradermally (0.5, 0.25, and 0.1 mg/mL) to guinea pig skin and immediately followed with electroporation using a MID device with either gold or stainless steel electrodes and either 1 mm- (5x5) or 1.5 mm-(4x4) spaced electrodes at 15 volts. The animals were sacrificed three days post treatment. The skin was excised and visualized under a fluorescence microscope. High resolution photographs were taken and subsequently analyzed for pixel intensity using standard software (Adobe Photoshop CS5). The level of expression was calculated through pixel counts of pre-defined treatment areas. To ensure accurate results, pooled data from multiple treatment sites on multiple animals was analyzed.

[0079] Results are shown in **Figure 11**. The GFP expression following ID injection alone (no EP) was also observed but minimal expression was detected (data not shown). No statistically significant differences between either spacing or electrode composition was observed at any of the concentrations of the plasmid expressing the reporter gene GFP.

Example 5

Electroporation Efficiency Analyzed at the Cellular Level

[0080] Skin samples removed from treated animals from experiments detailed in the above Examples were analyzed immunohistochemically. Skin post-treatment was excised post-mortem, sectioned, and paraffin mounted. GFP-expressing cells were observed and counted using a high powered fluorescent microscope (Olympus - BX51 TF). The number and region (i.e., layer of strata in epidermis) of GFP-expressing cells were noted. Histological sections were also counter-stained with a collection of commercially available antibodies prior to mounting to allow the direct identification of transfected cell types, such as lymphocyte IHC, keratinocytes (the majority of cells in the epidermis), and Langerhans cells (most common APC's in the epidermis). The antibodies were also used to observe the effect of electroporation on lymphocyte infiltration.

[0081] Robust keratinocyte staining was achieved in the epidermis. The epidermal region in the skin sections was clearly defined. Overall, the results suggested that the additional electrodes massively impacted the current flow and, as such, impacted the impedance of the tissue. This increased current flow did not appear to affect the resulting expression of the reporter gene. However, it was apparent that the voltage could be reduced on the 5x5 1 mm hand piece while still achieving strong currents and robust transfection.

[0082] To see positive staining for the Langerhans specific antibody in the skin, spleen and lymph nodes were removed from a sacrificed animal to use as positive controls for the antibody. Strong antibody staining was detected in both the spleen and the lymph nodes but not in the skin. Although not wishing to be bound by a particular theory, this suggested that the antibody was working but that either the signal in the skin was too weak to detect, or there were no Langerhans cells present in the tissue tested.

[0083] Additional results are shown in **Figure 12**. It was observed that both electroporation and expression of reporter genes resulted in infiltration of lymphocytes to the treatment area. The combination of EP and reporter gene expression resulted in the largest infiltration. While co-

localization of keratinocyte staining and reporter gene expression was observed, the staining was not consistent. However, co-localization of lymphocyte IHC and reporter gene expression was consistently confirmed.

Example 6

Dual-head Device

[0084] A dual-head device having two arrays side by side with a small buffer zone was manufactured. The arrays were designed to deliver pulses simultaneously. Alternatively, each head can be pulsed independently with additional equipment modifications. Plastic housings and custom electrical components were prototyped. The devices are shown in **Figure 13**.

[0085] A series of *in vivo* expression localization studies were completed, as described in Example 1. A plasmid expressing the reporter gene GFP or RFP was injected intradermally at a concentration of 1mg/mL to guinea pig skin. Injection was immediately followed with electroporation using the dual head device (with 16 stainless steel electrodes in a 4×4 array and a spacing of 1.5 mm, at 25 V). The results are shown in **Figure 14**, which is a fluorescent micrograph showing the GFP expression following an intradermal administration of a reporter gene plasmid and EP with the MID **100, 200**, compared to the GFP expression following the intradermal injection alone (no EP). The electrodes **110** of the MID **100, 200** were spaced from one another by a spacing of 1.5 mm. The animal was sacrificed post treatment and the skin excised and visualized under a fluorescence microscope. The fluorescent micrograph confirms that multiple plasmids were delivered simultaneously at spatially separated sites.

Example 7

Kinetics of Transfection

[0086] A series of *in vivo* expression localization studies will be completed, using the methods as described in Example 1. Specifically, following an intradermal injection of a known dose (0.5, 1, and 2 mg/mL) of plasmid DNA expressing the reporter gene GFP into guinea pig skin, a 1 mm or 1.5 mm prototype device will be used to immediately electroporate the resulting injection bubble. Electrodes of the MID will be either gold or stainless steel in composition. Animals will be sacrificed at different time points (12 hours, 24 hours, and 48 hours, 3 days) post treatment. Skin will be excised and visualized under a fluorescent microscope for each time point. High resolution photographs will be taken and subsequently analyzed for pixel intensity using standard software (Adobe Photoshop CS5). The level of expression will be calculated through pixel counts of pre-defined treatment areas. To ensure accurate results, pooled data from multiple treatment sites on multiple animals will be analyzed. Expression of the GFP will be compared over the different time periods to facilitate assessment of expression kinetics.

[0087] The invention is set out in the appended claims.

REFERENCES CITED IN THE DESCRIPTION

This list of references cited by the applicant is for the reader's convenience only. It does not form part of the European patent document. Even though great care has been taken in compiling the references, errors or omissions cannot be excluded and the EPO disclaims all liability in this regard.

Patent documents cited in the description

- US2007185432A [0008]
- US2001023330A [0008]
- US6413935B [0050] [0054]
- WO9321948A [0050]
- US6309633B [0056] [0059]
- US6493402B [0059]
- US6207157B [0059]
- US5908629A [0059]

Non-patent literature cited in the description

- The Jordan Report: Accelerated Development of Vaccine 1995NIH19950000 [0043]
- SANDERS et al. *Pediatr. Res.*, 1995, vol. 37, 812-819 [0050]
- BARTOLONI et al. *Vaccine*, 1995, vol. 13, 463-470 [0050]
- PIROFSKI et al. *Infect. Immun.*, 1995, vol. 63, 2906-2911 [0050]

Patentkrav

1. Indretning (100, 200) til elektroporering og administration af et eller flere antigener, hvilken indretning omfatter:

et hus (102), der har en spidsdel (104),

5 en flerhed af elektrodearrays (106, 108), der hver er koblet til spidsdelen (104), hvor elektrodearrayene (106, 108) rager ud fra spidsdelen (104) og definerer rumligt adskilte steder, idet hver elektrodearray (106, 108) omfatter en flerhed af elektroder (110), og spidsdelen (104) er aftagelig fra indretningen (100, 200) med henblik på
10 sterilisering mellem anvendelser af indretningen (100, 200),

en impulsgenerator (116, 118, 202), der er elektrisk koblet til mindst én elektrode (110) af hvert elektrodearray (106, 108), og

en programmerbar mikrostyreenhed (120, 206), der er elektrisk koblet til impulsgeneratoren (116, 118, 202), hvor hver elektrode (110) er
15 konfigureret til at afgive en elektroporerende impuls til epidermisvævsceller, og hvor mikrostyreenheden (120, 206) er konfigureret til at justere parametre for hvert elektrodearrays (106, 108) elektroporerende impuls uafhængigt.

2. Indretning (100, 200) ifølge krav 1, hvor den elektroporerende impuls er
20 tilknyttet et elektrisk potential, og hvor mikrostyreenheden (120, 206) er konfigureret til at variere det elektriske potential fra array (106, 108) til array (106, 108).

3. Indretning (100, 200) ifølge krav 1, hvor den elektroporerende impuls er
25 tilknyttet en elektrisk strøm, og hvor mikrostyreenheden (120, 206) er konfigureret til at variere den elektriske strøm fra array (106, 108) til array (106, 108).

4. Indretning (100, 200) ifølge krav 1, hvor den elektroporerende impuls er
30 tilknyttet en varighed, og hvor mikrostyreenheden (120, 206) er konfigureret til

at variere varigheden fra array (106, 108) til array (106, 108).

5 **5.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor mikrostyreenheden (120, 206) er konfigureret til at justere en respektiv mængde af elektroporerende impulser for hvert elektrodearray (106, 108) uafhængigt, og hvor mængden er 1 til 10.

10 **6.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor de rumligt adskilte steder i det væsentlige forhindrer interferens med anti-generne, når de elektroporerende impulser afgives.

15 **7.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor flerheden af elektroder (110) i hvert elektrodearray (106, 108) er arrangeret i et respektivt mønster.

20 **8.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor mindst én elektrode (110) af flerheden af elektroder (110) i hvert elektrodearray (106, 108) indbefatter en vævспенetrerende ende (114) med en længde på 0,1 mm eller mindre.

25 **9.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor mindst én elektrode (110) af flerheden af elektroder (110) i hvert array (106, 108) indbefatter en vævспенetrerende ende (114) med en længde på fra 0,01 mm til 0,04 mm.

10. Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor de elektroporerende impulser i det væsentlige forebygger skade i epidermis-vævscellerne.

30 **11.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor de elektroporerende impulser er tilknyttet et elektrisk potential, der er på fra 0,01 V til 70 V, hvor det elektriske potential er næsten smertefrit som målt ved hjælp af en visuel analog skala.

- 12.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor tilstødende elektroder (110) er anbragt med en indbyrdes afstand på højst 1,5 mm.
- 5 **13.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor tilstødende elektroder (110) er anbragt med en indbyrdes afstand på højst 1,0 mm.
- 14.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, hvor
10 flerheden af elektroder (110) for hvert elektrodearray (106, 108) er konfigureret til at afgive de elektroporerende impulser samtidigt.
- 15.** Indretning (100, 200) ifølge et hvilket som helst af de foregående krav, der endvidere omfatter:
- 15 en elektrisk strømkilde (122), der er koblet til impuls-
 generatoren (116, 118, 202) og mikrostyreenheden (120, 206), hvor den
 elektriske strømkilde (122) er et batteri (204), der bæres inde i
 huset (102), idet batteriet (204) er konfigureret til at drive impuls-
 generatoren (116, 118, 202), og batteriet (204) gør det muligt for
20 indretningen (100, 200) at være bærbar, og
 en udløserknap til at udløse afgivelse af den elektroporerende impuls.

DRAWINGS

FIGURE 1

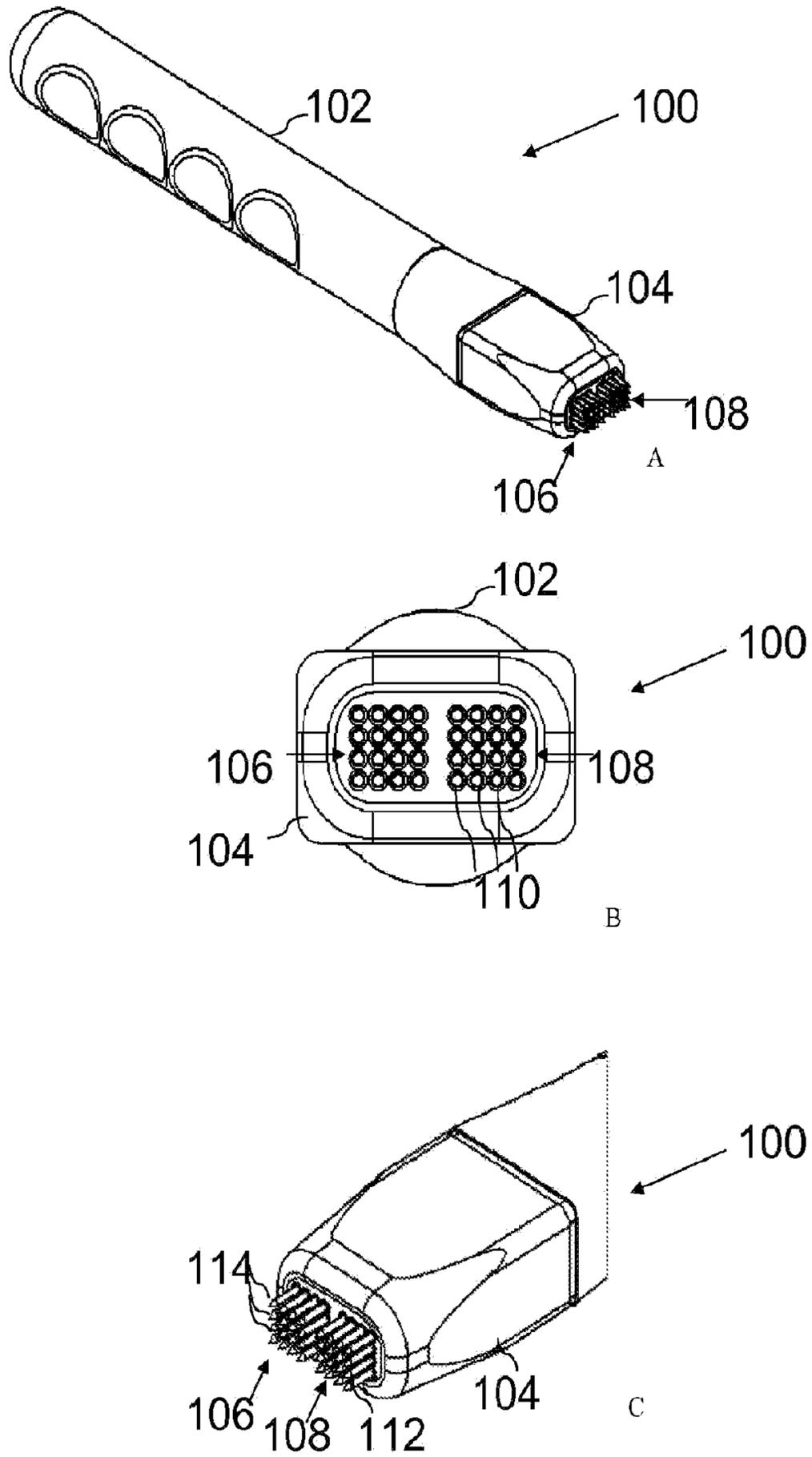


FIGURE 2

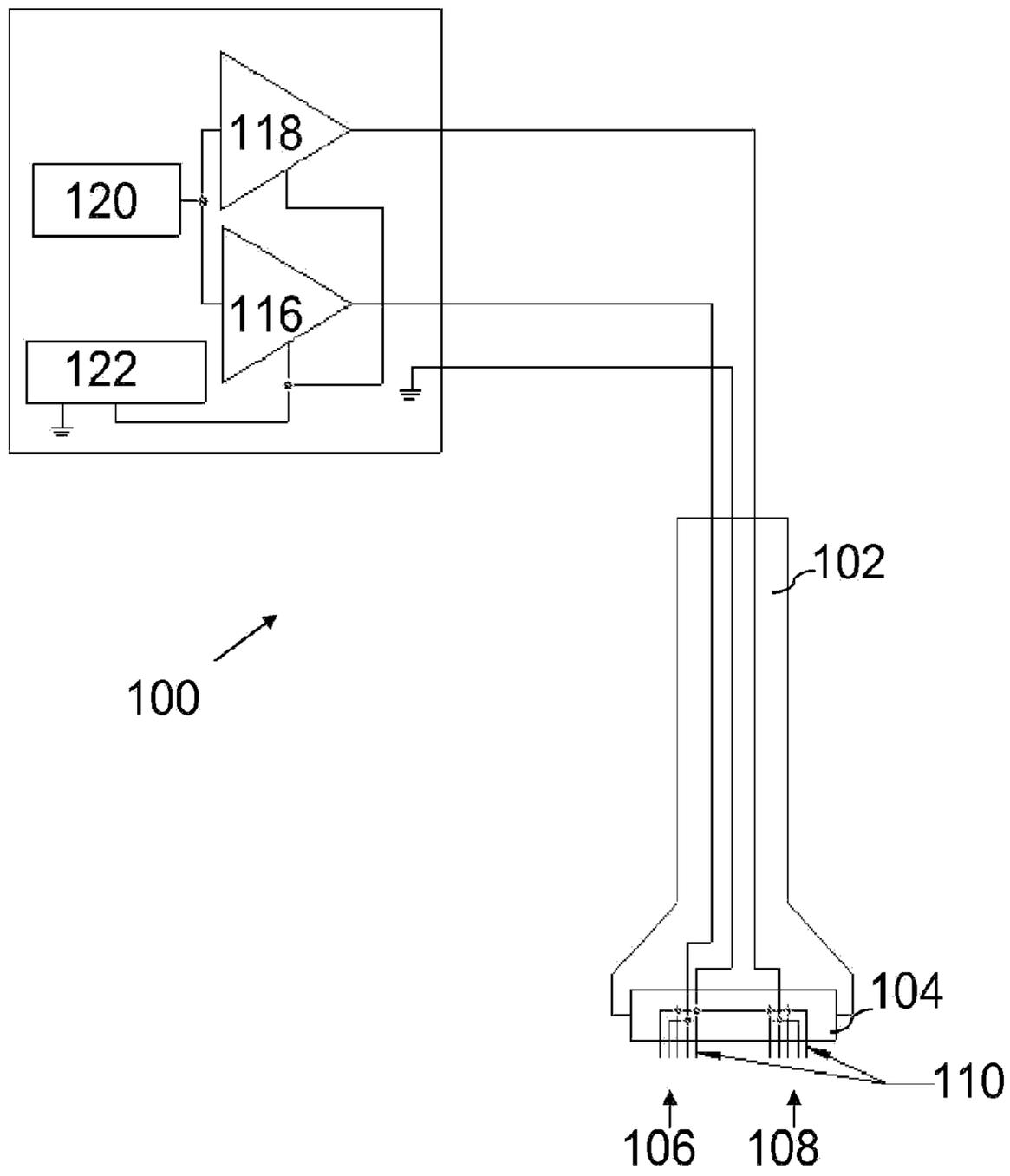


FIGURE 3

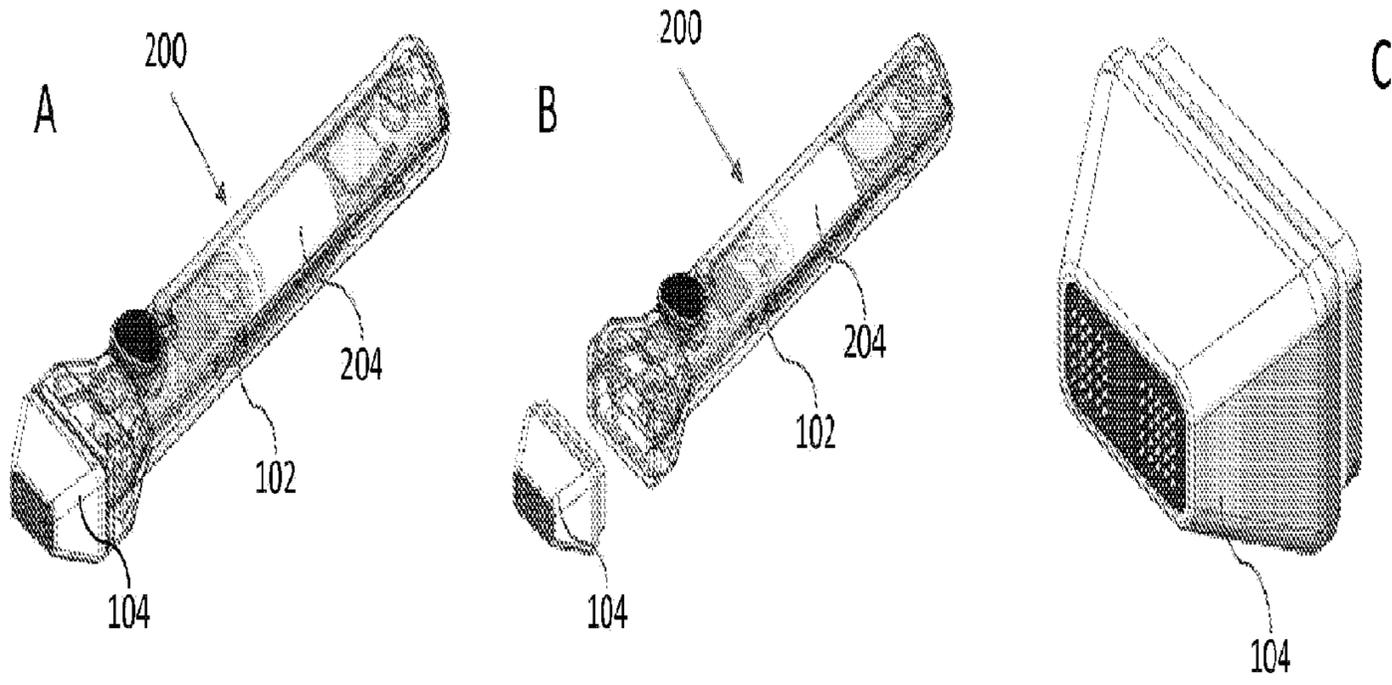


FIGURE 4

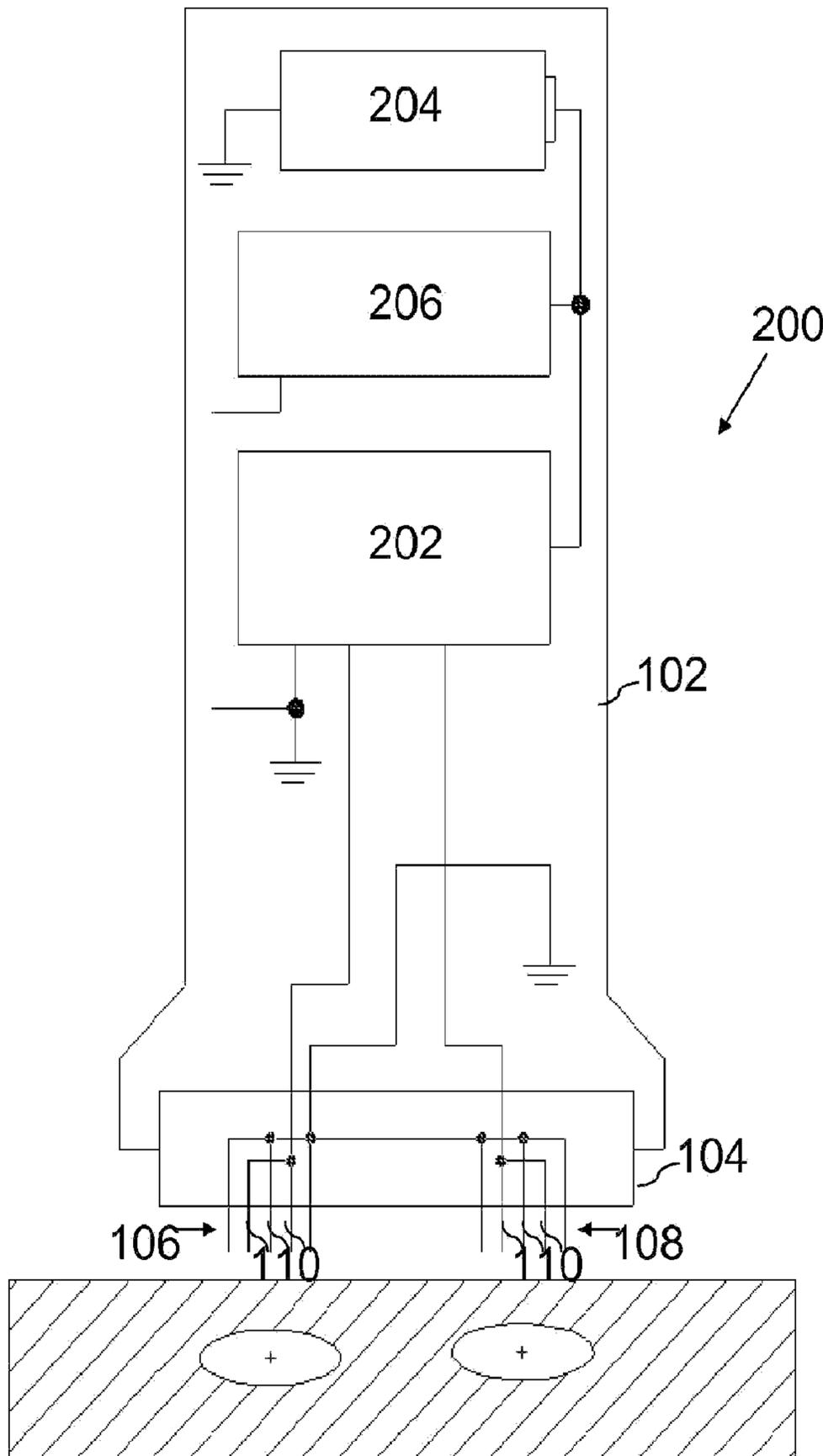


FIGURE 5

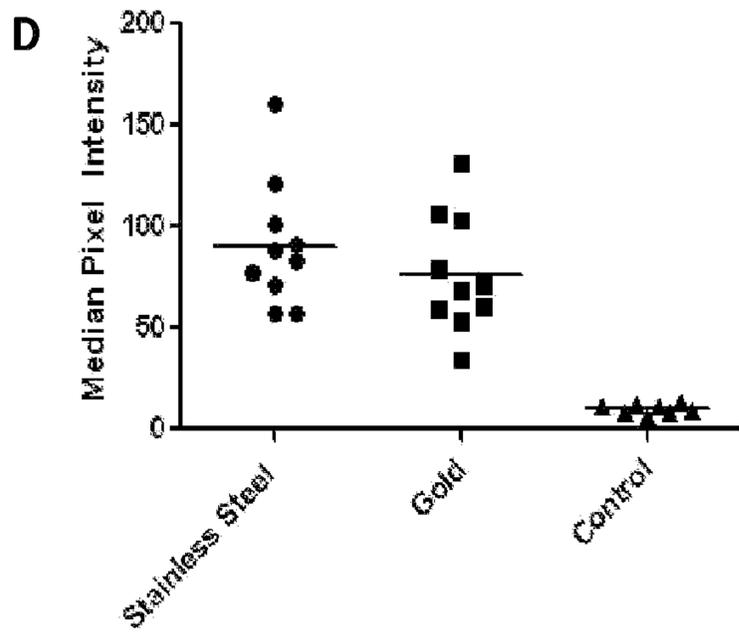
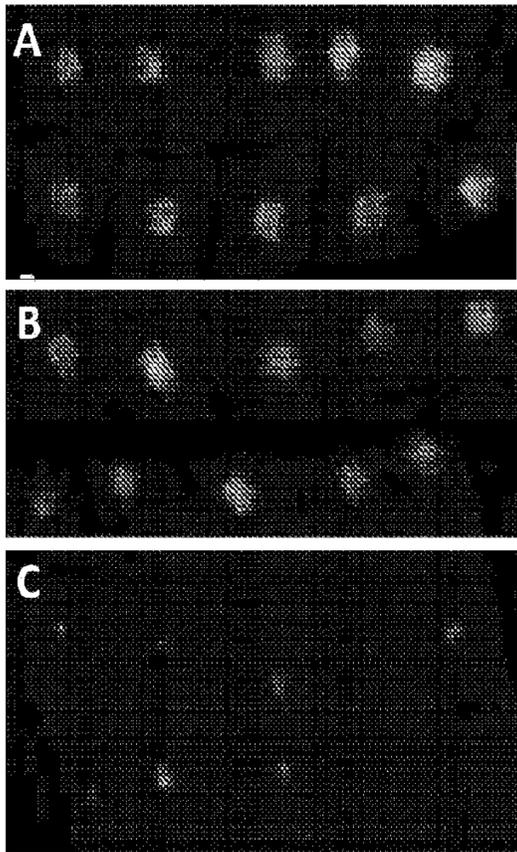
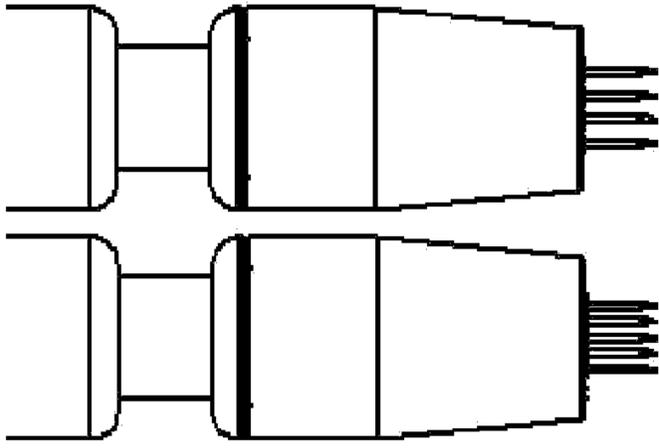
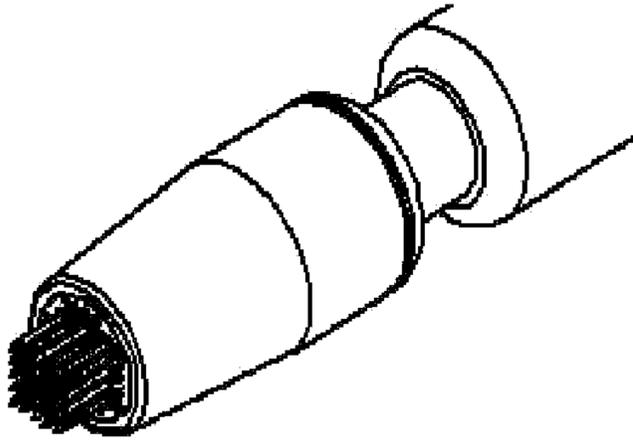


FIGURE 6



A



B

FIGURE 7

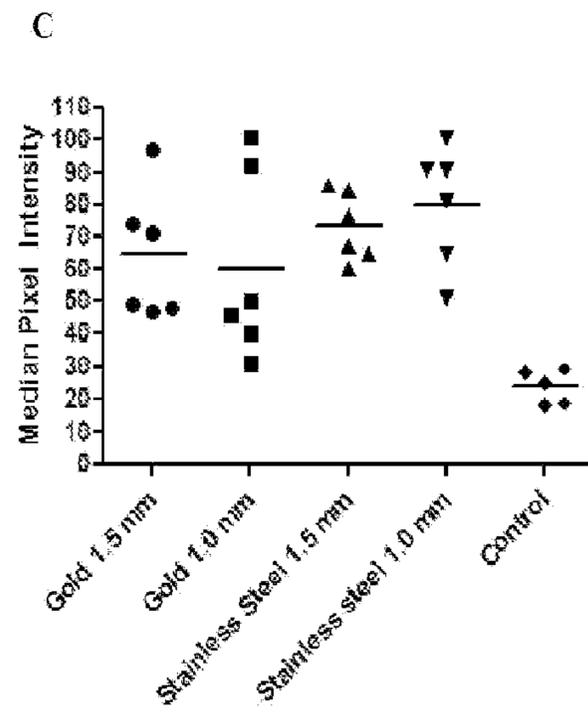
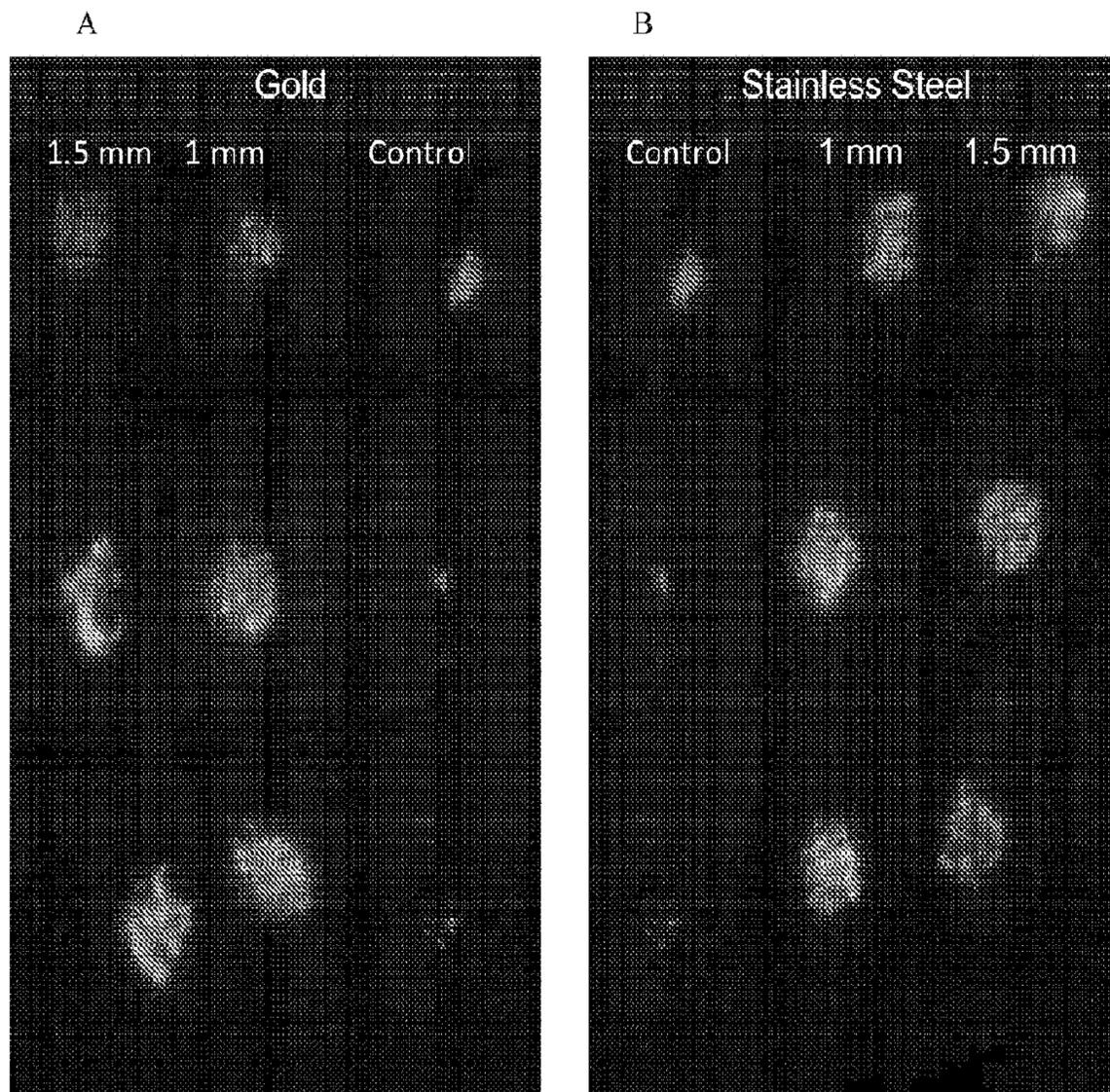


FIGURE 8

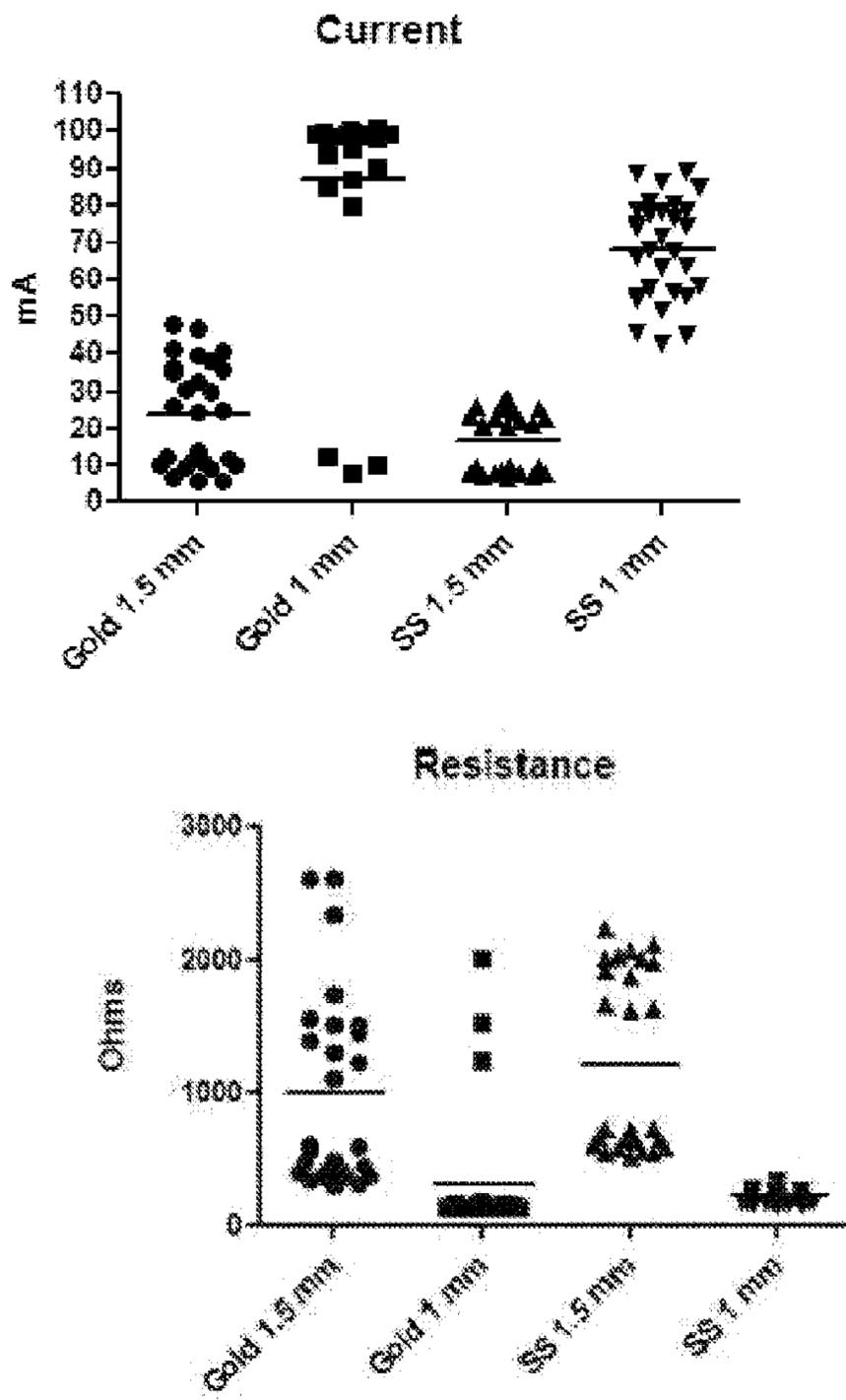


FIGURE 9

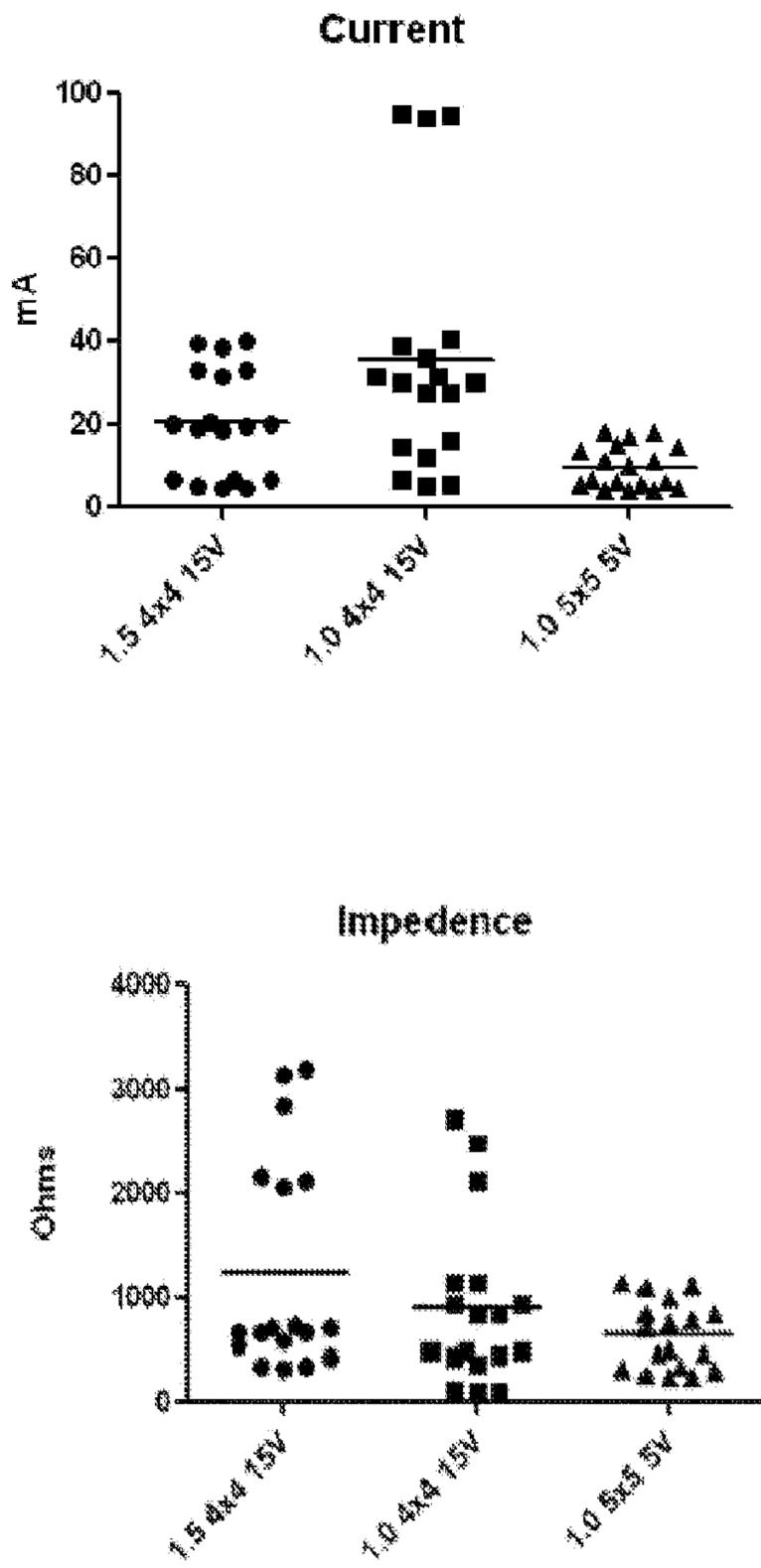


FIGURE 10

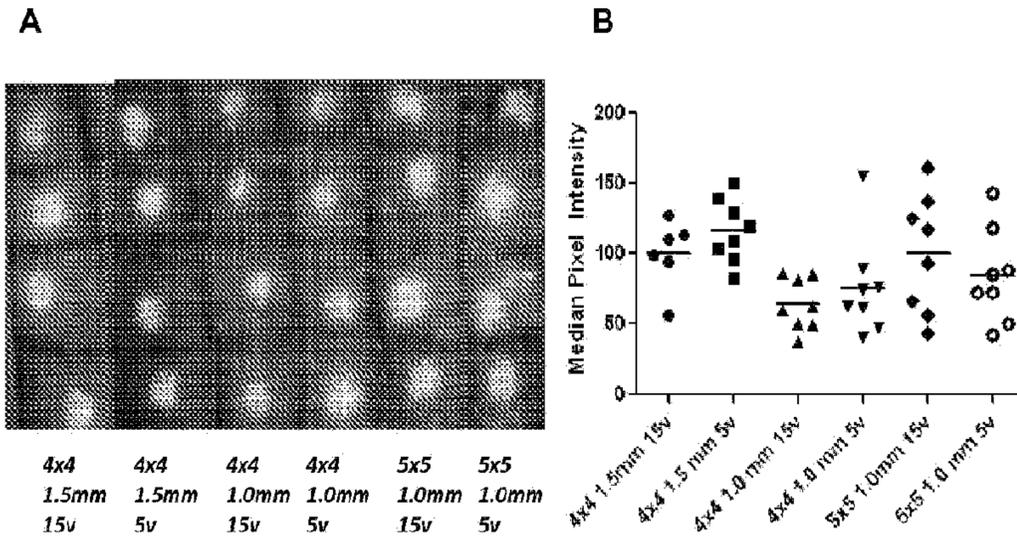


FIGURE 11

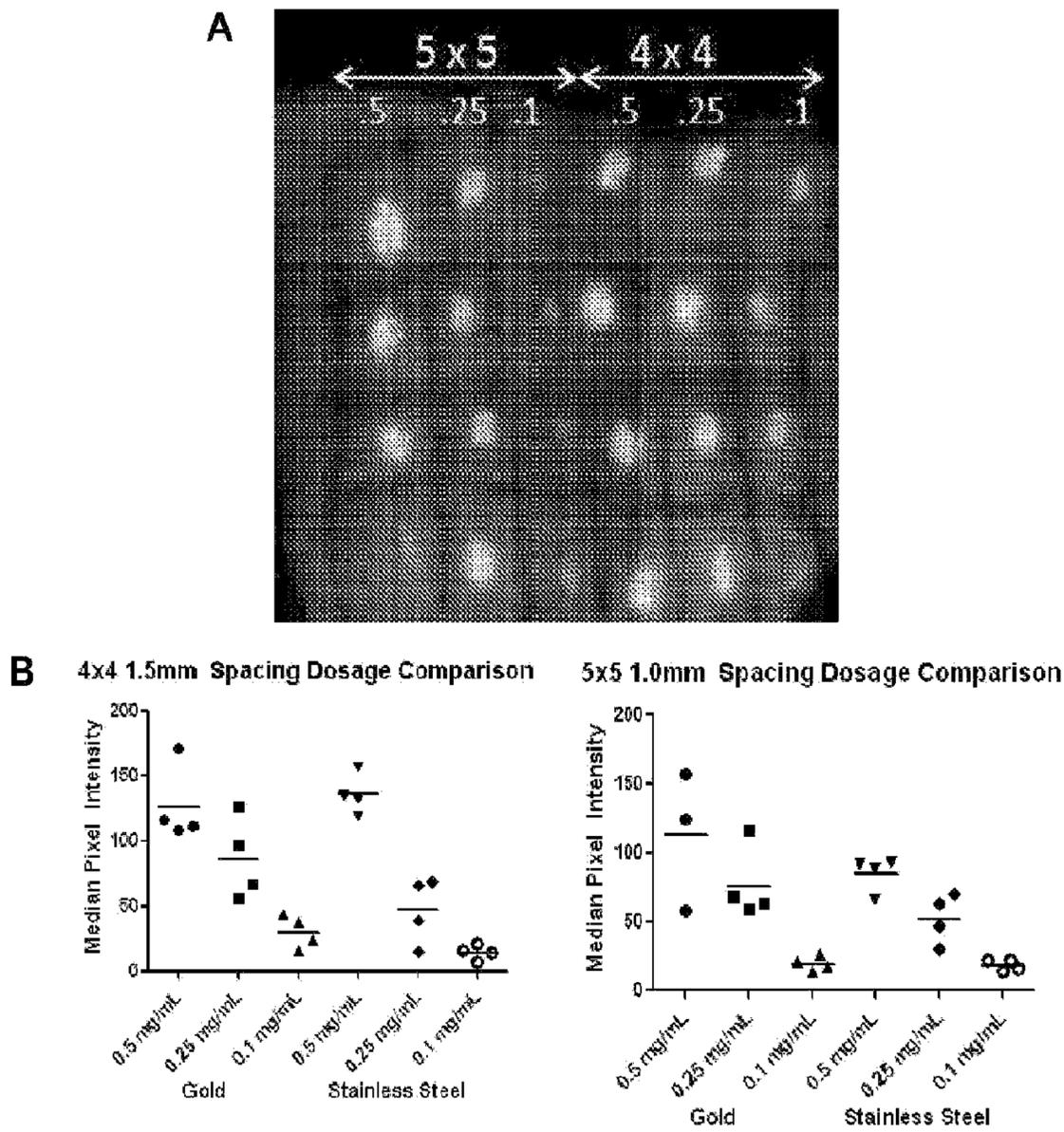


FIGURE 12

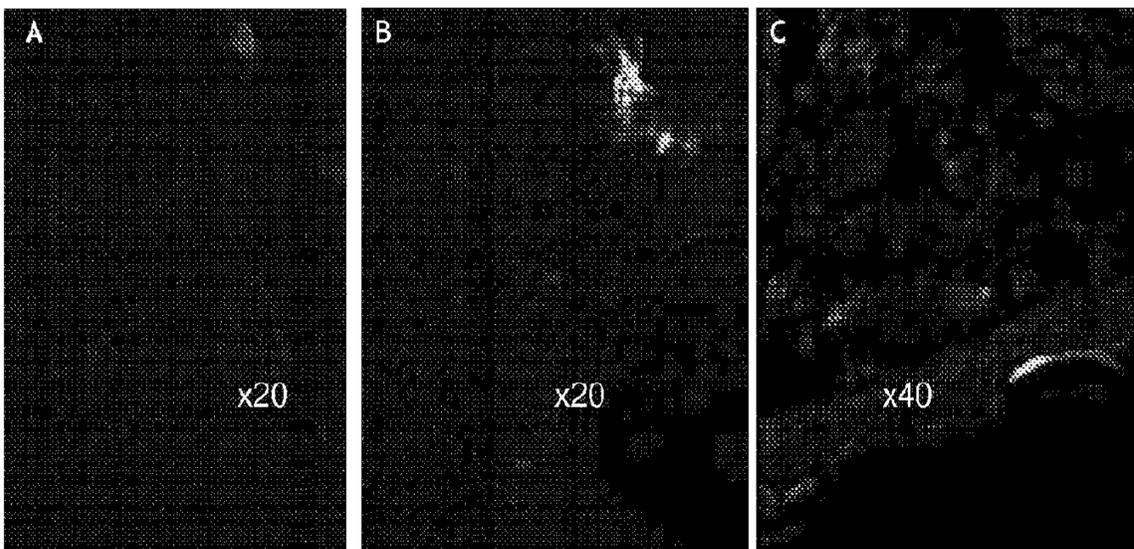


FIGURE 13

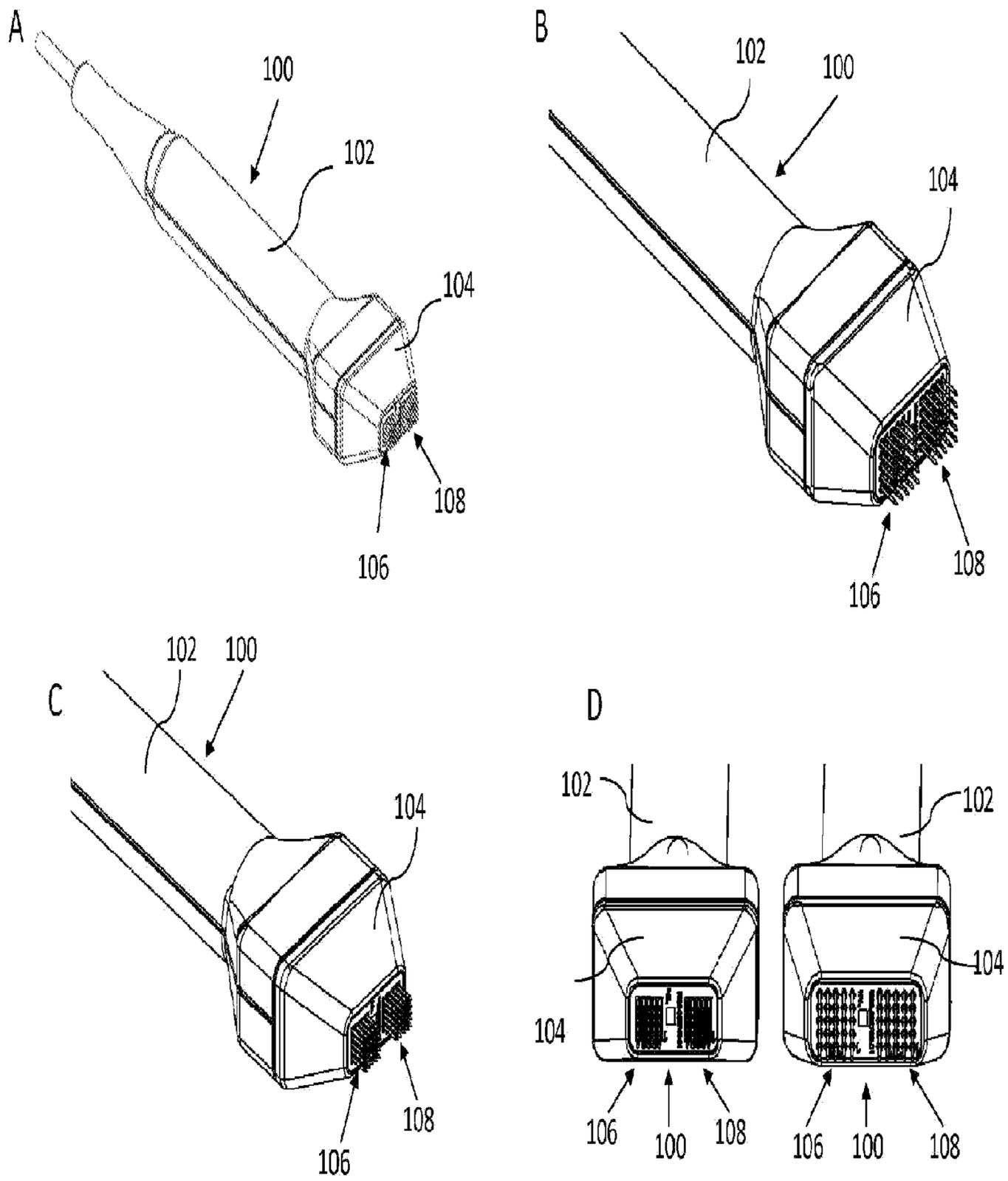
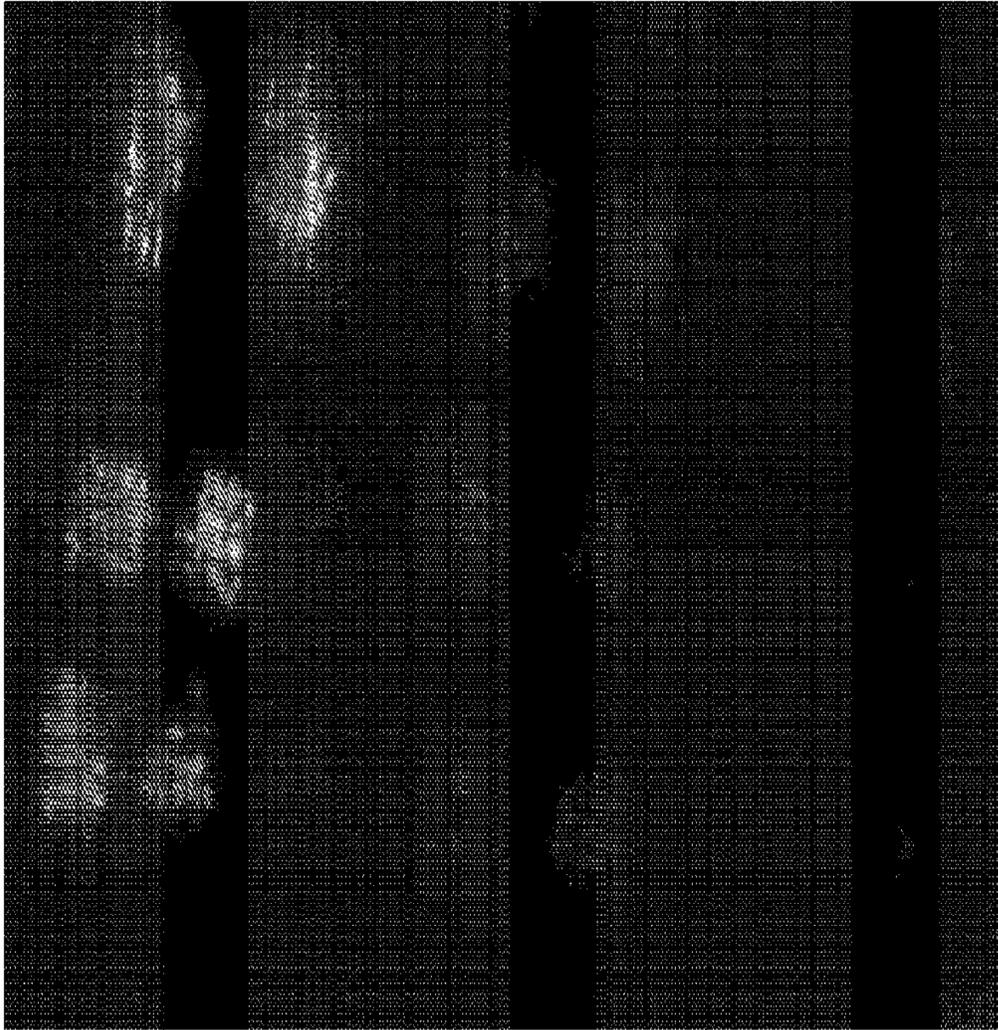


FIGURE 14



GFP

RFP

GFP

RFP

Controls
(Injection Only)