

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
7 June 2001 (07.06.2001)

PCT

(10) International Publication Number
WO 01/40802 A2

(51) International Patent Classification⁷: **G01N 33/68**

Gunnels Wood Road, Stevenage, Hertfordshire SG1 2NY (GB).

(21) International Application Number: PCT/GB00/04606

(22) International Filing Date: 1 December 2000 (01.12.2000)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
9928539.7 2 December 1999 (02.12.1999) GB
0012699.5 24 May 2000 (24.05.2000) GB

(71) Applicant (for all designated States except US): **GLAXO GROUP LIMITED** [GB/GB]; Glaxo Wellcome House, Berkeley Avenue, Greenford, Middlesex UB6 0NN (GB).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **COUSENS, Diane, Joan** [GB/GB]; Glaxo Wellcome plc, Gunnels Wood Road, Stevenage, Hertfordshire SG1 2NY (GB). **IGNAR, Diane, Michele** [US/US]; Glaxo Wellcome Inc., Five Moore Drive, Research Triangle Park, NC 27709 (US). **SANSEAU, Philippe** [FR/GB]; Glaxo Wellcome plc, Gunnels Wood Road, Stevenage, Hertfordshire SG1 2NY (GB). **VOLPE, Filippo** [IT/GB]; Glaxo Wellcome plc,

(74) Agent: **REES, Marion, L.**; Glaxo Wellcome plc, Glaxo Wellcome House, Berkeley Avenue, Greenford, Middlesex UB6 0NN (GB).

(81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.

(84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

Published:

— Without international search report and to be republished upon receipt of that report.

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: NOVEL PROTEIN

(57) Abstract: A method for the identification of a compound which modulates leukotriene B₄ like (LTRGW1) receptor activity comprises contacting an LTRGW1 polypeptide comprising i) The amino acid sequence of SEQ ID #2, or ii) A variant of (i) which is capable of binding leukotrienes; or iii) A fragment of (i) or (ii) which is capable of binding leukotrienes. Additionally, LTRGW1 has been determined to enhance the response of the leukotriene B₄ Receptor (BLTR) to LTB₄. Hence LTRGW1 may be used in methods to identify compounds which modulate BLTR receptor activity.



WO 01/40802 A2

NOVEL PROTEIN

Field of the Invention

The present invention relates to methods of screening for modulators of leukotriene-B₄ receptor-like polypeptides, and use of leukotriene-B₄ receptor like polypeptides in methods of screening modulators of the leukotriene B₄ receptor, BLTR.

Background of the Invention

Phospholipid undergoes metabolic degradation to form arachidonic acid which may be further metabolised to produce leukotrienes (LT) such as LTB₄, LTD₄, LTE₄, LTC₄ and LTF₄. There are two main classes of leukotriene receptor, the cysteinyl receptors and leukotriene-B₄ (BLT) receptors. One leukotriene-B₄ receptor, BLTR, has been cloned and pharmacologically characterised (Yokomitso *et al.* (1997) Nature 387, 620-624.) Analysis of the amino acid sequence of BLTR suggests that it belongs the G-protein coupled receptor superfamily, characterised by seven predicted transmembrane domains.

LTB₄ is a chemoattractant for neutrophils and primed eosinophils. LTB₄ enhances neutrophil-endothelial interactions and activates neutrophils leading to degranulation and release of mediators, enzymes and superoxides. LTB₄ is also able to bind and activate a nuclear transcription factor (PPAR_α). This activation results in the transcription of genes that are responsible for the termination of the immune response. In addition, the cloned LTB₄ receptor, BLTR, has been reported to mediate HIV-1 entry in CD4 positive T cells.

Diseases such as asthma are associated with deregulation of the host immune response. Hyper-responsiveness in asthmatic patients is characterised by eosinophilia, oedema and mucus production in the lung. Leukotriene receptor antagonists such as zafirlukast, pranlukast and montelukast are currently used to control the inflammatory response in asthmatic patients. Neutrophilic inflammation is a symptom of chronic obstructive pulmonary disease (COPD) and it is likely that LTB₄ receptor antagonists may also have clinical benefit in COPD patients.

Summary of the Invention

2

A leukotriene-B₄ receptor-like polypeptide (LTRGW1) is now provided which the inventors have shown to enhance the activity of BLTR in response to LTB₄ stimulation, as well as acting as a leukotriene receptor in its own right. Novel assays are provided which utilise the interaction between LTRGW1 and BLTR to
5 identify and develop novel pharmaceutical agents, including agonists and antagonists of the BLTR leukotriene-B₄ receptor. LTRGW1 may also be utilised as a screening target for the identification and development of novel pharmaceutical agents, including agonists and antagonists of the receptor. LTRGW1 may further be utilised in a screen to discover modulators of the interaction between BLTR and LBT4. Also
10 provided is a method of enhancing BLTR response to LBT4 comprising contacting a polypeptide of SEQ ID #2 or a variant thereof with BLTR. The present invention also provides a heterodimer comprising a polypeptide according to SEQ ID #2 and a polypeptide according to SEQ ID #8.

Accordingly, the present invention provides a method for the identification of
15 a compound which modulates leukotriene B₄ like receptor (LTRGW1) activity, which method comprises contacting an LTRGW1 polypeptide comprising

- (i) the amino acid sequence of SEQ ID NO: 2; or
- (ii) a variant of (i) which is capable of binding leukotrienes; or
- 20 (iii) a fragment of (i) or (ii) which is capable of binding leukotrienes.

The invention also provides:

- the use of an LTRGW1 polypeptide as herein defined to
25 enhance BLTR responses to leukotrienes.
- An isolated heterodimer comprising a LTRGW1 polypeptide as herein defined, and a BLTR polypeptide as herein defined.
- A method for increasing the responsiveness of a screen for identification of a substance that modulates the activity of the
30 leukotriene B₄ receptor (BLTR) comprising the addition to said screen of an LTRGW1 polypeptide as herein defined.
- a method for identification of a substance that modulates BLTR activity, which method comprises contacting a polypeptide of the invention and a

BLTR polypeptide comprising

- (i) the amino acid sequence of SEQ ID NO: 8, or
- (ii) A variant of (i) which is capable of binding LTB₄; or
- (iii) A fragment of (i) or (ii), which is capable of binding LTB₄

5 with a test substance and monitoring for LTB₄ binding to the said polypeptides;

- a method for identification of a substance that modulates leukotriene-B₄ receptor activity, which method comprises contacting an LTRGW1 polypeptide as herein defined and a BLTR polypeptide as herein defined with
10 LTB₄ in the presence of a test substance and monitoring for leukotriene-B₄ receptor activity;
- a method for identification of a substance that modulates leukotriene-B₄ receptor activity, which method comprises:
 - (i) providing
 - 15 (a) an LTRGW1 polypeptide as herein defined;
 - (b) a BLTR polypeptide as herein defined; and
 - (c) a test substanceunder conditions that would permit the interaction of (a) and (b) in the absence of (c);
20 (ii) monitoring the interaction between (a) and (b); and
(iii) determining whether (c) modulates the interaction between (a) and (b) and thereby determining whether the test substance is a modulator of leukotriene-B₄ receptor activity;
- a test kit suitable for identification of a substance that modulates leukotriene-B₄ receptor activity, which kit comprises:
25 (a) an LTRGW1 polypeptide as herein defined which is capable of potentiating the activity of BLTR in response to LTB₄; and
(b) a BLTR polypeptide as herein defined.
- a substance identified by one of the methods referred to above which
30 stimulates or modulates leukotriene-B₄ receptor activity;
- a method of treating a subject having a disorder that is responsive to LTRGW1 or BLTR receptor modulation, or modulation of the interaction between

LTRGW1 and BLTR, which method⁴ comprises administering to said patient an effective amount of a substance of the invention; and

- use of a substance of the invention in the manufacture of a medicament for the treatment or prophylaxis of a disorder that is responsive to modulation of LTRGW1 or BLTR receptor activity or modulation of the interaction between LTRGW1 and BLTR;

Preferably the disorder is selected from acute and chronic inflammatory diseases, asthma, chronic obstructive pulmonary disease (COPD), allergic rhinitis, hayfever, immune deficiency disorder, AIDS, rheumatoid arthritis, multiple sclerosis, leukemia, myasthenia gravis, graves disease, systemic lupus erythematosus, inflammatory bowel disease, encephalomyelitis, psoriasis, atopic dermatitis, septic shock, stroke, ischaemia reperfusion injury or cardiovascular disease. More preferred is when the disorder is an acute or chronic inflammatory diseases, asthma, chronic obstructive pulmonary disease (COPD) or psoriasis. Particularly preferred is when the disorder is asthma.

The invention further provides

- A method of treating a patient with a respiratory disorder in particular asthma or chronic obstructive pulmonary disease (COPD), said method comprising the administration of a therapeutically effective amount of an antagonist of LTRGW1
- A method of treating a respiratory disorder in particular asthma or chronic obstructive pulmonary disease (COPD), said method comprising the administration to a patient of a therapeutically effective amount of a substance that downregulates the interaction between LTRGW1 and BLTR, hence decreasing the response of BLTR to its ligand.
- Use of a therapeutically effective amount of an antagonist of LTRGW1 in the manufacture of a medicament for the treatment or prophylaxis of respiratory diseases, in particular asthma or chronic obstructive pulmonary disease (COPD)
- Use of an effective amount of a substance that downregulates the interaction between LTRGW1 and BLTR in the manufacture of a medicament for the

5
treatment or prophylaxis of respiratory disorders, in particular asthma
or chronic obstructive pulmonary disease (COPD)

Brief Description of the Figures

5 Figure 1 shows the dose dependent enhancement of BLTR activity by
LTRGW1.

Figure 2 shows the enhancement of LTB₄ binding to cells co-expressing
BLTR and LTRGW1.

Figure 3 shows the tissue distribution of LTRGW1 mRNA.

10 Figure 4 shows the tissue distribution of BLTR mRNA.

Figure 5 shows the presence of BLTR and LTRGW1 transcripts in three
different tissues.

Figure 6 shows that BLTR and LTRGW1 co-immunoprecipitate.

Brief Description of the Sequences

SEQ ID No 1 shows the DNA and amino acid sequences of human
LTRGW1.

SEQ ID No 2 is the amino acid sequence alone of LTRGW1.

SEQ ID NO: 3 shows the DNA and amino acid sequences of human
20 LTRGW1-32.

SEQ ID NO: 4 is the amino acid sequence alone of LTRGW1-32.

SEQ ID NO: 5 shows the DNA and amino acid sequences of the short splice
variant of human BLTR.

SEQ ID NO: 6 is the amino acid sequence alone of the short variant of
25 BLTR.

SEQ ID NO: 7 shows the DNA and amino acid sequences of the long splice
variant of human BLTR.

SEQ ID NO: 8 is the amino acid sequence alone of long variant of BLTR.

Detailed Description of the Invention

30 Throughout the present specification and the accompanying claims the words
"comprise" and "include" and variations such as "comprises", "comprising",
"includes" and "including" are to be interpreted inclusively. That is, these words are

intended to convey the possible inclusion⁶ of other elements or integers not specifically recited, where the context allows.

The present invention relates to methods using a human leukotriene-B₄ receptor-like polypeptide, referred to herein as LTRGW1. LTRGW1 is also
5 sometimes referred to as BLTR2. Sequence information for LTRGW1 is provided in SEQ ID NO: 1 (nucleotide and amino acid) and in SEQ ID NO: 2. Sequence information for a preferred fragment of LTRGW1 is provided in SEQ ID NO: 3 (nucleotide and amino acid) and in SEQ ID NO: 4.

The terms "LTRGW1 polypeptide", "LTRGW1 receptor" and "LTRGW1" as
10 used throughout the specification refer to a polypeptide comprising:

- (i) the amino acid sequence of SEQ ID NO: 2; or
- (ii) a variant of (i) which is capable of binding leukotrienes; or
- (iii) a fragment of (i) or (ii) which is capable of binding
15 leukotrienes.

Preferably, said variant or fragment is capable of binding LTB₄, 12-epi-LTB₄, LTB₃, LTB₅, LTD₄, LTE₄, LTC₄, or LTF₄, particularly preferred is when said variant or fragment is capable of binding LTB₄.

The polypeptides are provided in isolated form. The term "isolated" is
20 intended to convey that the polypeptide is not in its native state, insofar as it has been purified at least to some extent or has been synthetically produced, for example by recombinant methods. The term "isolated" therefore includes the possibility of the polypeptide being in combination with other biological or non-biological material, such as cells, suspensions of cells or cell fragments, proteins, peptides, expression
25 vectors, organic or inorganic solvents, or other materials where appropriate, but excludes the situation where the polypeptide is in a state as found in nature.

AN LTRGW1 polypeptide may also be in a substantially purified form, in which case it will generally comprise the polypeptide in a preparation in which more than 50%, e.g. more than 80%, 90%, 95% or 97%, 98% , 99%, by weight of the
30 polypeptide in the preparation is a polypeptide of the invention. Routine methods can be employed to purify and/or synthesise the proteins according to the invention. Such methods are well understood by persons skilled in the art, and include techniques such as those disclosed in Sambrook *et al*, Molecular Cloning: a

Laboratory Manual, 2nd Edition, CSH ⁷ Laboratory Press (1989), the disclosure of which is included herein in its entirety by way of reference.

The term "variant" in relation to LTRGW1 refers to a polypeptide which has the same essential character or basic biological functionality as LTRGW1. It is preferred that fragments of LTRGW1 and/or fragments of a variant of LTRGW1 also possess the same essential character or basic biological functionality as LTRGW1. Any such variants or fragments are herein included within the definition of LTRGW1 polypeptides.

In one aspect, the essential character of LTRGW1 can be defined as follows:
LTRGW1 is a leukotriene-B₄ receptor-like polypeptide which interacts with BLTR. In this aspect, a polypeptide having the same essential character as LTRGW1 typically potentiates the activity of LTB₄ at BLTR. A polypeptide having the same essential character as LTRGW1 may be identified by co-expressing the polypeptide with BLTR and monitoring the effect on BLTR activity in response to LTB₄. Any of the assays described herein as suitable for identifying a modulator of BLTR receptor activity may be performed in the absence of a test compound to determine whether a polypeptide has the same essential character as LTRGW1.

Preferably a polypeptide with the same essential character as LTRGW1 enhances LTB₄ mediated activity of BLTR in a dose dependent manner. Typically, when cells are transfected with equal amounts by weight of BLTR and a polypeptide with the same essential character as LTRGW1, the maximal response of BLTR to LTB₄ is enhanced from 30 to 70 fold, preferably from 40 to 60 fold, more preferably from 45 to 55 fold by the LTRGW1 polypeptide.

Typically a polypeptide with the same essential character as LTRGW1 is capable of binding BLTR. Preferably binding of LTRGW1 and BLTR occurs when LTRGW1 is recruited to a BLTR-LTB₄ receptor complex. In this aspect, the LTRGW1 polypeptide may enhance BLTR activity, or indeed may not be active in this respect, and may be used to bind to BLTR to prevent BLTR from binding to active LTRGW1. An LTRGW1 polypeptide may form heteromultimers with a BLTR polypeptide, such that one or more LTRGW1 polypeptides complex with one or more BLTR polypeptides. Such multimers are termed LTRGW1/BLTR throughout the specification. Preferably, the heteromultimer is a heterodimer, comprising one LTRGW1 polypeptide and one BLTR polypeptide.

Binding of the LTRGW1 polypeptide to BLTR may be measured by any suitable means. For example, binding of an LTRGW1 polypeptide to BLTR may be measured by immunoprecipitating said LTRGW1 polypeptide and detecting co-immunoprecipitation of BLTR or by immunoprecipitating BLTR and detecting co-immunoprecipitation of said LTRGW1 polypeptide.

Immunoprecipitation techniques are well known in the art. The binding assay may be carried out in the presence of a BLTR ligand, such as LTB₄ or leukotriene-B₄-3-aminopropylamide (LTB₄-APA). LTB₄-APA may be crosslinked to BLTR (Goldman (1991) *J. Immunol.* **146**, 2671-2677).

Alternatively, the interaction between LTRGW1 and BLTR may be monitored using fluorescence resonance energy transfer (FRET) (Guo *et al.*, (1995) *J. Biol. Chem.* **270**, 27562-27568) or bioluminescence resonance energy transfer (BRET) (Angers, S *et al* 2000 Proceedings of the National Academy of Sciences of the United States of America, **97**, 3684-3689).

In another aspect, a polypeptide with the same essential character as LTRGW1 may also be defined as one which binds to the same ligand as LTRGW1. This may be, for example, LTB₄, 12-epi-LTB₄, LTB₃, LTB₅, LTD₄, LTE₄, LTC₄, or LTF₄. Preferably an LTRGW1 polypeptide will bind LTB₄. In this aspect, a polypeptide having the same essential character as LTRGW1 may be identified by monitoring for binding of leukotrienes, for example, using radiolabelled LTB₄ or other leukotrienes. Any of the assays described herein as suitable for identifying a modulator of LTRGW1 activity may be performed in the absence of a test compound to determine whether a polypeptide has the same essential character as LTRGW1.

Preferably a polypeptide with the same essential character as LTRGW1 enhances binding of LTB₄ to the membranes of cells co-expressing BLTR and the LTRGW1 polypeptide compared to cells expressing only BLTR or the LTRGW1 polypeptide. A typical assay for determining whether an LTRGW1 polypeptide enhances the binding of LTB₄ in this manner comprises preparing membranes from mammalian cells or *Xenopus* oocytes co-expressing the LTRGW1 polypeptide and BLTR, performing a scintillation proximity assay using wheat germ agglutinin beads and [³H]LTB₄ and comparing the binding data obtained to that obtained with membranes from cells expressing only BLTR or only LTRGW1 (Yokomizo *et al.* (1997) *Nature* **387**, 620-624).

Preferably a polypeptide with the same essential character as LTRGW1 enhances LTB₄ binding from two to six fold, more preferably from four to five fold, or most preferably three fold in cells co-expressing the said polypeptide and BLTR compared to cells expressing BLTR alone. LTRGW1 may be used to discover modulators of the interaction between BLTR and LBT4

A full length protein is preferably one which includes a seven transmembrane region. Preferably, the full length receptor may couple to a G-protein to mediate intracellular responses.

Throughout the present specification the terms "BLTR polypeptide", "BLTR receptor" and "BLTR" refer to the leukotriene-B₄ receptor polypeptide, comprising

- (i) the amino acid sequence of SEQ ID NO: 8, or
- (ii) A variant of (i) which is capable of binding LTB₄; or
- (iii) A fragment of (i) or (ii), which is capable of binding LTB₄

A preferred fragment of BLTR has the amino acid sequence shown in SEQ ID NO:6. The term "variant" and "fragment" in relation to BLTR refer to a polypeptide which has the same essential character or basic biological functionality as BLTR. The essential character of BLTR can be defined as follows: BLTR is a G-protein coupled leukotriene-B₄ receptor which is activated by LTB₄. LTB₄ activation of a BLTR polypeptide can be enhanced by LTRGW1. Any of the assays described herein as suitable for identifying a modulator of BLTR receptor activity may be performed in the absence of a test compound to determine whether a polypeptide has the same essential character as BLTR.

A typical assay for determining whether the response of a BLTR polypeptide to LTB₄ is potentiated by LTRGW1 comprises co-expressing the BLTR polypeptide with LTRGW1 in mammalian cells, incubating cells with a calcium indicator dye, measuring the activity of the BLTR polypeptide in response to LTB₄ using a Fluorescence Imaging Plate Reader (FLIPR) and comparing the response to that obtained in cells expressing the BLTR polypeptide alone or the BLTR polypeptide and a different level of LTRGW1.

Preferably LTRGW1 enhances LTB₄ mediated activity of a polypeptide with the same essential character as BLTR in a dose dependent manner. Typically, when cells are transfected with equal amounts by weight of LTRGW1 and a polypeptide

with the same essential character as ¹⁰ BLTR, the maximal response of BLTR to LTB₄ is enhanced from 30 to 70 fold, preferably from 40 to 60 fold, more preferably from 45 to 55 fold by LTRGW1.

Typically a polypeptide with the same essential character as BLTR is capable
5 of binding LTRGW1. Binding of the BLTR polypeptide to LTRGW1 may be measured by any suitable means. For example, binding of the BLTR polypeptide to LTRGW1 may be measured by immunoprecipitating said BLTR polypeptide and detecting co-immunoprecipitation of LTRGW1 or by immunoprecipitating LTRGW1 and detecting co-immunoprecipitation of said BLTR polypeptide.
10 Immunoprecipitation techniques are well known in the art. The binding assay may be carried out in the presence of a BLTR ligand, such as LTB₄ or leukotriene-B₄-3-aminopropylamide (LTB₄-APA).

Alternatively, the interaction between LTRGW1 and BLTR may be monitored using fluorescence resonance energy transfer (FRET) (Guo *et al.*, (1995)
15 *J. Biol. Chem.* **270**, 27562-27568) or bioluminescence resonance energy transfer (BRET) (Angers, S *et al* 2000 Proceedings of the National Academy of Sciences of the United States of America, **97**, 3684-3689

In another aspect, a polypeptide with the same essential character as BLTR is one which binds to the same ligand as BLTR. Preferably a polypeptide with the
20 same essential character as BLTR will bind LTB₄. In this aspect, a polypeptide having the same essential character as BLTR may be identified by monitoring for binding of a BLTR ligand, for example, using radiolabelled LTB₄. Typically binding of LTB₄ to BLTR is enhanced in the presence of LTRGW1. Preferably the affinity of BLTR for LTB₄ is enhanced by the recruitment of LTRGW1 to the BLTR-LTB₄
25 receptor complex.

Preferably a polypeptide with the same essential character as BLTR is capable of coupling to a G-protein.

The following description of variants and fragments of the LTRGW1 polypeptide applies also to variants and fragments of BLTR except that rather than
30 being in relation to the amino acid sequences shown in SEQ ID NO: 2 and SEQ ID NO: 4, the sequence identities are in relation to the amino acid sequences shown in SEQ ID NO: 8 and SEQ ID NO: 6 and the basic biological functionality is that of the BLTR receptor.

Typically, polypeptides with more than about 65% identity preferably at least 80% or at least 90% and particularly preferably at least 95% at least 96% at least 97% at least 98% or at least 99% identity, with the amino acid sequences of SEQ ID NO: 2 or SEQ ID NO: 4 are considered as variants of

- 5 LTRGW1, provided that they retain the basic biological functionality or essential character of the LTRGW1 polypeptide as herein defined. Such variants may include allelic variants and the deletion, modification or addition of single amino acids or groups of amino acids within the protein sequence, as long as the peptide maintains the basic biological functionality of the LTRGW1 receptor.

- 10 Amino acid substitutions may be made, for example from 1, 2 or 3 to 10, 20 or 30 substitutions. The modified polypeptide generally retains activity as an LTRGW1 receptor. Conservative substitutions may be made, for example according to the following Table. Amino acids in the same block in the second column and preferably in the same line in the third column may be substituted for each other.

15

ALIPHATIC	Non-polar	G A P
		I L V
	Polar-uncharged	C S T M
		N Q
	Polar-charged	D E
		K R
AROMATIC		H F W Y

- Shorter polypeptide sequences are within the scope of the invention. For example, a peptide of at least 20 amino acids or up to 50, 60, 70, 80, 100, 150 or 200 amino acids in length is considered to fall within the scope of the invention as long as it demonstrates the basic biological functionality of LTRGW1. In particular, but not exclusively, this aspect of the invention encompasses the situation when the protein is a fragment of the complete protein sequence and may represent a ligand-
- 20

binding region (N-terminal extracellular domain) or an effector binding region (C-terminal intracellular domain). Such fragments can be used to construct chimeric receptors preferably with another 7-transmembrane receptor, more preferably with another leukotriene-B₄ receptor. Such fragments can also be used to raise anti-
5 LTRGW1 antibodies. In this embodiment the fragment may comprise an epitope of the LTRGW1 polypeptide and may otherwise not demonstrate the ligand binding or other properties of LTRGW1.

Polypeptides of the invention may be chemically modified, e.g. post-translationally modified. For example, they may be glycosylated or comprise
10 modified amino acid residues. They may also be modified by the addition of histidine residues to assist their purification or by the addition of a signal sequence to promote insertion into the cell membrane. Polypeptides of the invention may be tagged to aid detection, for example using a VSV, HA, T7, myc or flag tag. Such modified polypeptides fall within the scope of the term "polypeptide" of the
15 invention.

The invention also includes cells that have been modified to express an LTRGW1 polypeptide and which are also modified to express a BLTR polypeptide. Such cells include transient, or preferably stable higher eukaryotic cell lines, such as mammalian cells or insect cells, lower eukaryotic cells, such as yeast or prokaryotic
20 cells such as bacterial cells. Particular examples of cells which may be modified by insertion of vectors encoding for LTRGW1 and BLTR polypeptides include mammalian HEK293T, CHO, HeLa and COS cells. Stable cell lines expressing LTRGW1 and BLTR may be isolated using a chemotaxis assay. Preferably the cell line selected will be one which is not only stable, but also allows for mature
25 glycosylation and cell surface expression of a polypeptide. Expression may be achieved in transformed oocytes. A polypeptide of the invention may be expressed in cells of a transgenic non-human animal, preferably a mouse. A transgenic non-human animal expressing a polypeptide of the invention is included within the scope of the invention. A polypeptide of the invention may also be expressed in *Xenopus*
30 *laevis* oocytes or melanophores, in particular for use in an assay of the invention.

It is also possible for the polypeptides of the invention to be transiently expressed in a host cell or on a membrane, such as for example in a baculovirus expression system. Such systems, which are adapted to express the polypeptides

13
according to the invention, are also included within the scope of the present invention. Preferably such systems are adapted to co-express an LTRGW1 polypeptide and BLTR.

An important aspect of the present invention is the use of polypeptides
5 according to the invention in screening methods to identify substances that may act as agonists or antagonists which may modulate leukotriene-B₄ receptor activity. This may take three forms –

- i) screening for substances that modulate LTRGW1's leukotriene activity,
- 10 ii) using LTRGW1 polypeptides to enhance the responsiveness of BLTR to LTB₄ and hence screening for substances that modulate BLTR's leukotriene activity, or modulate the ability of LTRGW1 to enhance the activity of BLTR,
- iii) screening for substances that modulate the interaction between
15 LTRGW1 and BLTR.

The term modulators as used herein should be interpreted to mean substances that are agonists or antagonists of the interaction of either receptor and its respective ligands, or that upregulate or downregulate the interaction between LTRGW1 and BLTR. Preferably, modulators are antagonists of the LTRGW1 receptor, or are able to
20 downregulate the interaction between LTRGW1 and BLTR.

To identify modulators of LTRGW1's leukotriene binding ability, any suitable form may be used for the assay. In general terms, such screening methods may involve contacting an LTRGW1 polypeptide with a test compound and then measuring receptor activity or may involve incubating an LTRGW1 polypeptide with
25 a test substance and then detecting modulation of leukotriene activity at the LTRGW1 receptor. Agents which bind to the LTRGW1 polypeptides can also be identified by binding assays.

Modulator activity can be determined by contacting cells expressing an LTRGW1 polypeptide with a substance under investigation and by monitoring the
30 effect mediated by the LTRGW1 polypeptides. The cells expressing the LTRGW1 polypeptide may be *in vitro* or *in vivo*. The LTRGW1 polypeptide may be naturally or recombinantly expressed. Preferably, the assay is carried out *in vitro* using cells expressing recombinant LTRGW1 polypeptide. Typically, LTRGW1 receptor

activity can be monitored indirectly by ¹⁴ measuring a Gi-coupled readout. Gi coupled readout can typically be monitored using an electrophysiological method to determine the activity of G-protein regulated Ca²⁺ or K⁺ channels or by using a fluorescent dye to measure changed in intracellular Ca²⁺ levels. Other methods that
5 can typically be used to monitor LTRGW1 receptor activity involved measuring levels of or activity of GTPγS, cAMP or chemotaxis. An assay of the invention may be carried out using a known leukotriene agonist or leukotriene antagonist to provide a comparison with a modulator under test.

For example, Kamohara *et al* (Journal of Biological Chemistry Vol. 275 Issue
10 35 pp 27000-27004, 10/7/00) have shown that leukotrienes LTB₄, LTB₃, LTB₅ and 12-epi-LTB₄ when binding to LTRGW1 cause inhibition of forskolin-stimulated intracellular cAMP accumulation, and further that LTB₄ induces chemotaxis. This paper both demonstrates the functionality of LTRGW1 and indicates that such functional assays may be used to measure modulation of the leukotriene mediated
15 activity of LTRGW1. Addition of a suspected modulator to such an assay allows determination of whether the functional response is increased or decreased, or whether there is no change. Yokomizo *et al* (Journal of Experimental Medicine Vol. 192, number 3 pp 421-432 7/8/00) have also shown forskolin stimulated intracellular cAMP accumulation and chemotaxis when LTRGW1 is exposed to leukotrienes, and
20 further demonstrate that LTB₄ increases cellular calcium, demonstrating that this is another suitable assay to measure modulator activity at the LTRGW1 receptor.

To identify a modulator of BLTR's leukotriene activity by using an LTRGW1 polypeptide, or to identify a modulator of LTRGW1's ability to bind to, and enhance the activity of, BLTR any suitable form may be used for the assay. In
25 general terms, such screening methods may involve contacting an LTRGW1 polypeptide and a BLTR polypeptide, with a test substance and then measuring BLTR receptor activity or may involve incubating an LTRGW1 polypeptide and a BLTR polypeptide with a test substance and then detecting modulation of leukotriene activity at the BLTR receptor. Substances which bind to LTRGW1 or BLTR
30 polypeptides can also be identified by binding assays. Preferably the assay may be carried out in a single well of a microtitre plate. Assay formats which allow high throughput screening are preferred.

Modulator activity can be determined by contacting cells co-expressing an LTRGW1 polypeptide and a BLTR polypeptide, with a substance under investigation and by monitoring the effect mediated by the BLTR receptor. To determine whether a test substance acts as an antagonist of LTB₄ at the BLTR/LTRGW1 receptor the effect of a test substance on the activation of BLTR by LTB₄ or another BLTR agonist may be monitored. A typical method for determining whether a test substrate acts as a BLTR agonist comprises monitoring stimulation of BLTR activity by contacting an LTRGW1 polypeptide and a BLTR polypeptide with a test substance and monitoring for BLTR activity. The cells expressing the polypeptide may be *in vitro* or *in vivo*. The LTRGW1 polypeptide and/or the BLTR polypeptide may be naturally or recombinantly expressed. Preferably, the assay is carried out *in vitro* using cells expressing recombinant BLTR polypeptide. More preferably, the cells express both recombinant LTRGW1 polypeptide and recombinant BLTR polypeptide.

Typically, receptor activity can be monitored indirectly by measuring a G_q/G_i-coupled readout. G_q/G_i coupled readout can typically be monitored using an electrophysiological method to determine the activity of G-protein regulated Ca²⁺ or K⁺ channels or by using a fluorescent dye to measure changed in intracellular Ca²⁺ levels. Other methods that can typically be used to monitor receptor activity involve measuring levels of or activity of GTPγS, cAMP or chemotaxis. An assay of the invention may be carried out using a known leukotriene-B₄ agonist or leukotriene-B₄ antagonist to provide a comparison with a modulator under test.

A standard assay for measuring activation of the G_i family of G proteins is the GTPγS binding assay. Agonist binding to G protein-coupled receptors promotes the exchange of GTP for GDP bound to the α subunit of coupled heterotrimeric G proteins. Binding of the poorly hydrolysable GTP analogue, [³⁵S]GTPγS, to membranes has been used extensively as a functional assay to measure agonism at a wide variety of receptors. Furthermore, the assay is largely restricted to measuring function of receptors coupled to the G_i family of G proteins due to their ability to bind and hydrolyse guanine nucleotide at significantly higher rates than members of the G_q, G_s and G₁₂ families. See Wieland and Jakobs, *Methods Enzymol.* 237, 3-13, 1994.

G protein coupled receptors (GPCRs) have been shown to activate MAPK

signalling pathways. Host cells overexpressing the LTRGW1 and BLTR polypeptides with MAPK reporter genes may be utilised as assays for receptor activation or inhibition. For example, yeast assays may be used to screen for agents that modulate the activity of an LTRGW1/BLTR receptor. A typical yeast assay
5 involves heterologously expressing an LTRGW1/BLTR receptor in a modified yeast strain containing multiple reporter genes, typically FUS1-HIS3 and FUS1-lacZ, each linked to an endogenous MAPK cascade-based signal transduction pathway. This pathway is normally linked to pheromone receptors, but can be coupled to foreign receptors by replacement of the yeast G protein with yeast/mammalian G protein
10 chimeras. Strains may also contain further gene deletions, such as deletions of SST2 and FAR1, to potentiate the assay. Ligand activation of the heterologous receptor can be monitored for example either as cell growth in the absence of histidine or with a suitable substrate such as beta-galactosidase (lacZ).

Alternatively melanophore assays may be used to screen for activators of an
15 LTRGW1/BLTR receptor. An LTRGW1/BLTR receptor can be heterologously expressed in *Xenopus laevis* melanophores and their activation can be measured by either melanosome dispersion or aggregation. Basically, melanosome dispersion is promoted by activation of adenylate cyclase or phospholipase C, i.e. G_s and G_q mediated signalling respectively, whereas aggregation results from activation of G_i-
20 protein resulting in inhibition of adenylate cyclase. Hence, ligand activation of the heterologous receptor can be measured simply by measuring the change in light transmittance through the cells or by imaging the cell response.

Assays may also be carried out by incubating a cell expressing a BLTR polypeptide and an LTRGW1 polypeptide with a test substance in the presence of
25 neutrophils or other cells of the immune system. Chemotaxis of the neutrophils associated with stimulation of the receptor of the invention can be monitored. Similarly, neutrophil degranulation and release of mediators, enzymes and superoxides from neutrophils can be measured to monitor or assess activation of the BLTR/LTRGW1 receptor in the presence of a test substance.

30 Preferably, control experiments are carried out on cells which do not express LTRGW1 to establish whether the observed responses are the result of activation or inhibition of the polypeptide. More preferably, control experiments are carried out on cells which express BLTR but which do not express LTRGW1.

All assays may be carried out utilising cells expressing only BLTR and/or only LTRGW1 and the results of those experiments may be compared to the results of parallel experiments on cells expressing both BLTR and LTRGW1 to ensure any effects of a test substance are dependent on the presence of LTRGW1 in the cells.

The binding of a modulator to an LTRGW1 polypeptide or BLTR polypeptide can also be determined directly. For example, a radiolabelled test substance can be incubated with LTRGW1 polypeptide or BLTR polypeptide and binding of the test substance to the polypeptide can be monitored. Typically, the radiolabelled test substance can be incubated with cell membranes or cells containing the polypeptide until equilibrium is reached. The membranes can then be separated from a non-bound test substance and dissolved in scintillation fluid to allow the radioactive content to be determined by scintillation counting. Non-specific binding of the test substance may also be determined by repeating the experiment in the presence of a saturating concentration of a non-radioactive ligand. Preferably such binding assays are carried out on cells co-expressing an LTRGW1 polypeptide and a BLTR polypeptide. Preferably the binding of a test substance to cells co-expressing an LTRGW1 polypeptide and a BLTR polypeptide is compared to the binding of a test substance to cells expressing only BLTR and/or to cells expressing only an LTRGW1 polypeptide.

Alternatively, ligand binding may be monitored by binding a fluorescent ligand such as LTB₄-APA-fluorescein to cells expressing the polypeptides of interest and detecting bound ligand by fluorescence activated cell sorting (FACS).

A test substance may modulate leukotriene mediated activity by disrupting the interaction between LTRGW1 and BLTR. An assay which monitors the interaction between LTRGW1 and BLTR may be used to screen for substances that modulate leukotriene activity. For example, an immunoprecipitation assay, a pull-down assay, an affinity-purification assay or a fluorescence resonance energy transfer (FRET) assay may be used to determine the effect of a test substance on the interaction between BLTR and LTRGW1. An LTRGW1 polypeptide for use in such an assay is capable of binding to BLTR but may, or may not, possess other essential characteristics of LTRGW1. A BLTR polypeptide for use in such an assay is capable

of binding to LTRGW1 but may, or may not, possess other essential characteristics of BLTR.

Suitable test substances which can be tested in the above assays include combinatorial libraries, defined chemical entities, peptide and peptide mimetics, oligonucleotides and natural product libraries, such as display (e.g. phase display libraries) and antibody products.

Test substances may be used in an initial screen of, for example, 10 substances per reaction, and the substances of these batches which show inhibition or activation tested individually. Test substances may be used at a concentration of from 1nM to 1000µM, preferably from 1µM to 100µM, more preferably from 1µM to 10µM.

Another aspect of the present invention is the use of the substances that have been identified by screening techniques referred to above in the treatment or prophylaxis of disorders which are responsive to regulation of leukotriene-B₄ receptor activity. Typically modulators useful in the therapeutic or prophylactic treatment of such disorders are inhibitors of leukotriene-B₄ receptor activity. In particular, such substances may be used in the treatment of acute and chronic inflammatory diseases, such as asthma, chronic obstructive pulmonary disease (COPD), allergic rhinitis, hayfever, immune deficiency disorder, AIDS, rheumatoid arthritis, multiple sclerosis, leukaemia, myasthenia gravis, graves disease, systemic lupus erythematosus, inflammatory bowel disease, encephalomyelitis, psoriasis, atopic dermatitis, septic shock, stroke, ischaemia reperfusion injury and cardiovascular diseases. Preferably, such substances are used in the treatment of asthma, COPD, Rheumatoid arthritis and psoriasis. Particularly preferred is when such substances are used in the treatment of asthma. It is to be understood that mention of these specific disorders is by way of example only and is not intended to be limiting on the scope of the invention as described.

The substances identified according to the screening methods outlined above may be formulated with standard pharmaceutically acceptable carriers and/or excipients as is routine in the pharmaceutical art, and as fully described in Remington's Pharmaceutical Sciences, Mack Publishing Company, Eastern Pennsylvania 17th Ed. 1985, the disclosure of which is included herein of its entirety by way of reference. The carrier or excipient may be an isotonic saline solution but

will depend more generally upon the ¹⁹ particular agent concerned and the route by which the agent is to be administered.

The substances may be administered by enteral or parenteral routes such as via oral, buccal, anal, pulmonary, intravenous, intra-arterial, intramuscular, 5 intraperitoneal, topical or other appropriate administration routes. A therapeutically effective amount of a modulator is administered to a patient. The dose of a modulator may be determined according to various parameters and especially according to the substance used; the age, weight and condition of the patient to be treated; the route of administration; and the required regimen. A physician will be 10 able to determine the required route of administration and dosage for any particular patient. A typical daily dose is from about 0.1 to 50 mg per kg of body weight, according to the activity of the specific modulator, the age, weight and conditions of the subject to be treated, the type and severity of the degeneration and the frequency and route of administration. Preferably, daily dosage levels are from 5 mg to 2 g.

15 Alternatively substances which down-regulate LTRGW1 expression or nucleic acid encoding a polypeptide, preferably an LTRGW1 variant polypeptide, which inhibits the function of LTRGW1 may be administered to the mammal. Nucleic acid, such as RNA or DNA, preferably DNA, is provided in the form of a vector, which may be expressed in the cells of a human or other mammal under 20 treatment. Preferably such down-regulation or expression following nucleic acid administration will inhibit LTRGW1 mediated potentiation of BLTR activity.

Nucleic acid encoding the LTRGW1 or variant polypeptide may be administered to a human or other mammal by any available technique. For example, the nucleic acid may be introduced by injection, preferably intradermally, 25 subcutaneously or intramuscularly. Alternatively, the nucleic acid may be delivered directly across the skin using a nucleic acid delivery device such as particle-mediated gene delivery. The nucleic acid may be administered topically to the skin, or to the mucosal surfaces for example by intranasal, oral, intravaginal, intrarectal administration.

30 Uptake of nucleic acid constructs may be enhanced by several known transfection techniques, for example those including the use of transfection agents. Examples of these agents includes cationic agents, for example, calcium phosphate and DEAE-Dextran and lipofectants, for example, lipofectam and transfectam. The

dosage of the nucleic acid to be administered can be altered. Typically the nucleic acid is administered in the range of 1pg to 1mg, preferably to 1pg to 10µg nucleic acid for particle mediated gene delivery and 10µg to 1mg for other routes.

Polynucleotides encoding LTRGW1 or a variant polypeptide can also be used to identify mutation(s) in LTRGW1 genes which may be implicated in human disorders. Identification of such mutation(s) may be used to assist in diagnosis of acute and chronic inflammatory diseases, such as asthma, chronic obstructive pulmonary disease (COPD), allergic rhinitis, hayfever, immune deficiency disorder, AIDS, rheumatoid arthritis, multiple sclerosis, leukaemia, myasthenia gravis, graves disease, systemic lupus erythematosus, inflammatory bowel disease, encephalomyelitis, psoriasis, atopic dermatitis, septic shock, stroke, ischaemia reperfusion injury, cardiovascular diseases or susceptibility to such disorders and in assessing the physiology of such disorders. Preferably, the disorder is asthma, COPD, rheumatoid arthritis or psoriasis.

Antibodies (either polyclonal or preferably monoclonal antibodies, chimeric, single chain, Fab fragments) which are specific for the LTRGW1 polypeptide or a variant thereof can be generated. Such antibodies may for example be useful in purification, isolation or screening methods involving immunoprecipitation techniques and may be used as tools to elucidate further the function of LTRGW1 or a variant thereof, or indeed as therapeutic agents in their own right. Such antibodies may be used to block ligand binding to the receptor. A variety of protocols for competitive binding or immunoradiometric assays to determine the specific binding capability of an antibody are well known in the art (see for example Maddox *et al*, J. Exp. Med. 158, 1211 *et seq*, 1993).

25

The following Examples illustrate the invention.

Example 1: LTRGW1 potentiation of BLTR activity in response to LTB₄

The 388 amino acids (aa) encoding for LTRGW1 peptide were aligned to other known seven transmembrane proteins and putative transmembrane domains were identified as follows: TM1 aa 55-77, TM2 aa 91-113, TM3 aa 124-148, TM4 aa 168-187, TM6 aa 256-277, TM7 aa 305-324. Hydrophobicity plot analysis confirmed these areas as putative transmembrane domains. LTRGW1 does not

contain a signal peptide at its amino terminal end. The closest protein is the BLT receptor (BLTR) with 50% similarity and 45% identity throughout their length. LTRGW1 gene is localised in the near proximity of the already described BLTR suggesting that it could have been evolved as a result of gene duplication. Indeed they share a 61% identity at the DNA level over a stretch of 885 bases. Expression constructs were generated from both putative methionine (position 1 or 32 in SEQ ID NO: 1) using the following 5' end primers respectively:

GGAATTCGCACCATGGCACCTTCTCATCGGGCATCACAG (LL1) and
GGAATTCGCACCATGTTCGGTCTGCTACCGTCCCCCA (LL2).

Genomic DNA was used as template in PCR reaction using either LL1 or LL2 primer together with a 3' end primer with the following sequence:
GCTCTAGATCAAAGGTCCCATTCGGACCGTCCTTC (LL6).

PCR products were digested with EcoRI and XbaI and subcloned into pcDNA3 (pLTRGW1-1 and pLTRGW1-32 for corresponding methionines).

pLTRGW1-1 and pLTRGW1-32 were fully sequenced and their pharmacological properties when co-expressed with BLTR were assessed in *Xenopus laevis* oocyte and CHO over-expression systems.

10mg total DNA was transfected into CHO cells using the following transfection mix: 5µg BLTR, pcDNA3 as needed to keep constant the amount of total DNA used, 1µg pCMV-luciferase and BLTR-2 at various concentration. Responses to LTB₄ (between 10⁻¹⁰ and 10⁻⁷ M) were measured using a FLIPR assay (Fluorescence Imaging Plate Reader - Molecular Devices) and values were normalised relative to the luciferase reading. The results are shown in Table 1. Under these particular experimental conditions LTRGW1 expressed alone does not induce a detectable mobilisation of intracellular calcium in response to LTB₄. However, as shown by Kamohara *et al* (Journal of Biological Chemistry Vol. 275 Issue 35 pp 27000-27004, 10th July 2000) and Yokomizo *et al* (Journal of Experimental Medicine Vol. 192, number 3 pp 421-432 7th August 2000), it is possible to detect responses such as inhibition of forskolin-stimulated intracellular cAMP accumulation, and chemotaxis, indicating that LTRGW1 is responsive to leukotrienes. Yokomizo *et al* also manage to show that LTB₄ increases intracellular calcium.

The response of BLTR to LTB₄ is increased when LTRGW1 is co-expressed with BLTR. The size of the increased response is proportional to the amount of LTRGW1 in the transfection mix. This enhancement of LTB₄ activation of BLTR in the presence of LTRGW1 is illustrated in Figure 1.

5

Table 1

CHO cells transfected with	Response to LTB ₄
Mock transfected	Inactive
BLTR	Active
LTRGW1 ₍₂₇₅₎	Inactive
LTRGW1 ₍₂₇₄₎	Inactive
LTRGW1 ₍₂₇₆₎ [Signal Peptide-FLAG-LTRGW1 ₍₂₇₆₎]	Inactive [FACS showed surface localisation]
BLTR/LTRGW1 ₍₂₇₅₎ 1 µg	Active +
BLTR/LTRGW1 ₍₂₇₅₎ 3 µg	Active ++
BLTR/LTRGW1 ₍₂₇₅₎ 5 µg	Active +++
BLTR/LTRGW1 ₍₂₇₆₎	Active

10 **Example 2: Enhanced binding of LTB₄ to membranes of cells co-expressing LTRGW1 and BLTR**

CHO cells were transfected with DNA as described in Example 1. [³H] LTB₄ binding to membrane preparations from transfected cells was measured using wheat germ agglutinin beads in a scintillation proximity assay. The assay was carried out in 50 :g Hepes, 20 :g MgCl₂ in a total assay volume of 100 :g. [³H] LTB₄ was used to measure total binding. [³H] LTB₄ was then displaced with unlabelled LTB₄ to measure non-specific binding (NSB). The binding of LTB₄ to cells expressing BLTR cells co-expressing BLTR and LTRGW1 and cells expressing LTRGW1 is illustrated in Figure 2.

20

Example 3: Tissue distribution of LTRGW1 and BLTR

LTRGW1 mRNA tissue distribution was studied using the following oligonucleotides as forward, reverse and probe primers respectively:

5'GCGCGAGCGGGA ACTA-3'

5'AGCGGTGAAGACG²³TAGAGCAC-3'
 5'CCTTGGCCTTCTTCAGTTCTAGCGTCAA-3'

using TaqmanTM PCR analysis (P. E. Biosystems). The results of this analysis on normal human tissues is shown in Figure 3.

- 5 BLTR mRNA tissue distribution was also studied using TaqmanTM PCR analysis. The tissue distribution of BLTR mRNA is shown in Figure 4.

LTRGW1 and BLTR have an identical tissue distribution. Both appear to be ubiquitously expressed, with highest levels in skin, tonsil, spleen and adenoid.

10 **Example 4: Comparison of presence of full length transcripts of BLTR and LTRGW1.**

The presence of full length transcripts of the short and long forms of both BLTR and LTRGW1 was analysed in the spleen, testis and skin. The control was no DNA. The following primer sets were used:

- 15 LTRGW1 long: LL1 ATGGCACCTTCTCATCGGGCATCACAG
 LL6b TCAAAGGTCCCATTCCGGACCGTCCTTC
- LTRGW1 short: LL2 ATGTCGGTCTGCTACCGTGGGGGA
 LL6b As above
- 20 BLTR long: BLTR17 ATGGCGTCAGGAAACCCTTGGTCCTC
 BLTR15 CTAGTTCAGTTCGTTTAACTTGAGAGGGC
- BLTR short: BLTR4 AACACTACATCTTCTGCAGCACCCCCCT
 25 BLTR15 As above

Spleen and Testis libraries were from Life Technologies (cat.No 10425-015 and 10426-103 respectively) Skin library was from Invitrogen (Cat. No. A900-14)

- 30 75ng of DNA from each library was used in the PCR reaction, and 35 cycles of 94°C for 60 seconds, 62°C for 60 seconds, 70°C for 120 seconds were carried out.

The results can be seen in figure 5. They show that cDNAs for both BLTR and

LTRGW1 are present in all the tissues²⁴ tested, further confirming the overlapping distribution suggested by the results of example 3. Interestingly, LTRGW1 is represented by the short form (Seq ID #4) only.

5 **Example 5: Immunoprecipitation of BLTR and LTRGW1**

- To test whether BLTR and LTRGW1 form heterodimers, immunoprecipitation experiments were carried out. Transiently transfected CHO cells were harvested from 60 mm culture dishes. Cells from each dish were resuspended in 1 ml of 50 mM Tris-HCl, 150 mM NaCl, 1 % (v/v) Nonidet[®] P40, 0.5 % (w/v) sodium deoxycholate, pH 7.5 (lysis buffer) supplemented with Complete[™] protease inhibitor cocktail tablets (1 tablet/25 ml) (Roche). Cell lysis and membrane protein solubilisation was achieved by passage through a 25-gauge needle followed by gentle mixing for 30 min at 4°C. Insoluble debris was removed by microcentrifugation at 16,000 g for 15 min at 4°C and the supernatant was precleared by incubating with 50 µl of Protein A-agarose (Roche) for 3 h at 4°C on a helical wheel to reduce background caused by non-specific adsorption of cellular proteins. The solubilised supernatant was then divided into 2 x 500 µl aliquots and 20 µl of either BLTR (221), BLTR2 (287) or Myc antisera was added to each. Immunoprecipitation was allowed to proceed for 1 h at 4°C on a helical wheel prior to the addition of 50 µl of Protein A-agarose suspension. Capture of immune complexes was progressed overnight at 4°C on a helical wheel. Complexes were then collected by microcentrifugation 12,000 g for 1 min at 4°C and supernatant was discarded. Beads were then washed by gentle resuspension and agitation sequentially in 1 ml of 50 mM Tris-HCl, pH 7.5, 500 mM NaCl, 0.1 % (v/v) Nonidet[®] P40 and 0.05 % (w/v) sodium deoxycholate followed by 1 ml of 50 mM Tris-HCl, pH 7.5, 0.1 % (v/v) Nonidet[®] P40 and 0.05 % (w/v) sodium deoxycholate. Immunoprecipitated proteins were released from Protein A-agarose by incubation in 30 µl of SDS-PAGE sample buffer at 70°C for 10 min and analysed by SDS-PAGE followed by immunoblotting.
- The results can be seen in figure 6. This experiment clearly demonstrates that BLTR and LTRGW1 co-precipitate, indicating that they have formed a dimer. This heterodimerisation was specific for the two receptors, since immunoprecipitation of GABAb, an unrelated G-protein coupled receptor did not pull down either

leukotriene receptor when co-expressed. Furthermore, this experiment also demonstrates that BLTR and LTRGW1 receptors heterodimerise following overexpression even in the absence of ligand stimulation.

5

Example 6: Screening for substances which exhibit protein modulating activity

(i) Transfection of cells

Mammalian cells, such as HEK293, CHO or COS7 cells or *Xenopus laevis* oocytes over-expressing either a BLTR polypeptide, an LTRGW1 polypeptide, or
10 both a BLTR polypeptide and an LTRGW1 polypeptide, are generated for use in the assays.

For example, *Xenopus* oocyte expression may be determined as follows. Adult female *Xenopus laevis* (Blades Biologicals) are anaesthetised using 0.2% tricaine (3-aminobenzoic acid ethyl ester), killed and the ovaries rapidly removed.
15 Oocytes are then de-folliculated by collagenase digestion (Sigma type I, 1.5 mg ml⁻¹) in divalent cation-free OR2 solution (82.5mM NaCl, 2.5mM KCl, 1.2mM NaH₂PO₄, 5mM HEPES; pH 7.5 at 25°C). Single stage V and VI oocytes are transferred to ND96 solution (96mM NaCl, 2mM KCl, 1mM MgCl₂, 5mM HEPES, 2.5mM sodium pyruvate; pH 7.5 at 25°C) which contains 50µg ml⁻¹ gentamycin and stored at
20 18°C.

LTRGW1 DNA and/or BLRT DNA (in pcDNA₃, Invitrogen) is linearised and transcribed to RNA using T7 (Promega Wizard kit). m⁷G(5')pp(5')GTP capped cRNA is injected into oocytes (20-50ng per oocyte) and whole-cell currents are recorded using two-microelectrode voltage-clamp (Geneclamp amplifier, Axon
25 instruments Inc.) 3 to 7 days post-RNA injection. Microelectrodes have a resistance of 0.5 to 2MΩ when filled with 3M KCl.

(ii) Ligand binding

Cells are transiently transfected with both tagged and untagged polypeptides. The binding of various concentrations of [³H] LTB₄ to membranes derived from cells
30 overexpressing the relevant polypeptides is measured and normalised to the level of expression of tagged receptors. Non-specific binding of [³H]LTB₄ is determined by monitoring [³H]LTB₄ binding in the presence of non-radioactive LTB₄.

Test substances are screened for their ability to displace [³H]LTB₄ from

BLTR, LTRGW1 and/or BLTR/LTRGW1 receptors by repeating the experiment in the presence of non-radioactive test substance. (See Yokomizo T *et al.* 1997 Nature 387, 620-624).

Alternatively, ligand binding may be monitored by binding a fluorescent ligand such as LTB₄-APA-fluorescein to cells expressing the polypeptides of interest and detecting bound ligand by fluorescence activated cell sorting (FACS).

(iii) Ca²⁺ immobilisation (FLIPR)

96 and 384 well plate, high throughput screens (HTS) are employed using fluorescence based calcium indicator molecules, including but not limited to dyes such as Fura-2, Fura-Red, Fluo 3 and Fluo 4 (Molecular Probes). Secondary screening involves the same technology.

A screening assay may be conducted as follows. Mammalian cells stably over-expressing the protein(s) are cultured in black wall, clear bottom, tissue culture coated 96 or 384 well plates with a volume of 100µl cell culture medium in each well 3 days before use in a FLIPR assay. Cells are incubated with 4µM FLUO-3AM at 30°C in 5%CO₂ for 90 mins and then washed once in Tyrodes buffer containing 3mM probenecid. Basal fluorescence is determined prior to substance additions. The protein is activated upon the addition of a known agonist such as LTB₄. Activation results in an increase in intracellular calcium which can be measured directly in the FLIPR. For antagonist studies, substances are preincubated with the cells for 4 minutes following dye loading and washing and fluorescence is measured for 4 minutes. Agonists are then added and cell fluorescence is measured for a further 1 minute.

Transiently transfected CHO-Gα and wild type CHO cells are ideal for such experiments. Results can be compared to results of parallel experiments run in the presence of petussis toxin (PTX).

(iv) Accumulation of cAMP

Following leukotriene stimulation, cyclic AMP accumulation can be measured in forskolin stimulated cells such as CHO-Gα and wild-type CHO cells transfected with the LTRGW1 receptor either directly, by SPA assay, or indirectly by monitoring the expression of co-transfected reporter gene, the expression of which will be controlled by cyclic AMP response elements. Results are compared to parallel experiments run in the presence of PTX.

(v) Chemotaxis

A typical chemotaxis assay will measure the movement of LTRGW1 and BLTR transfected cells such as CHO cells through a polycarbonate filter with 8- μ m pores towards the side in contact with the leukotriene ligand (Yokomizo T, et al 1997

5 Nature, 387, 620-624).

(vi) Fluorescence resonance energy transfer (FRET)

The association of BLTR polypeptides and LTRGW1 polypeptides may be monitored using FRET. The polypeptides are co-expressed in cells and may be labelled with a donor probe, fluorescein or with an acceptor carbocyanine probe (Cy3). A typical FRET assay utilises cells co-expressing VSV-tagged BLTR and FLAG-tagged LTRGW1. The cells are stained with Cy3-labelled anti-VSV antibody and biotin-labelled anti-FLAG antibody and then with fluorescein-streptavidin. FACS analysis of FRET between BLTR and LTRGW1 is then carried out in the absence of stimulation and/or following stimulation with LTB₄ or LTB₄-APA.

15 Alternatively, a typical FRET assay may measure FRET between LTB₄-APA-fluorescein and flag-tagged LTRGW1 stained with cy3 conjugated anti-FLAG antibody in cells expressing BLTR and flag-tagged LTRGW1.

(vii) Tertiary screening

Tertiary screens involve the study of modulators in rat, mouse and guinea-pig
20 models of disease relevant to the target.

CLAIMS

1. A method for the identification of a compound which modulates leukotriene
5 B₄ like (LTRGW1) receptor activity, which method comprises contacting an
LTRGW1 polypeptide comprising:
- i) The amino acid sequence of SEQ ID # 2, or
 - ii) A variant of (i) which is capable of binding leukotrienes; or
 - iii) A fragment of (i) or (ii) which is capable of binding
10 leukotrienes.
- with a test compound in the presence of a leukotriene such as LTB₄, LTD₄,
LTE₄, LTC₄, and LTT₄.
- 15 2. A method according to claim 1 wherein the variant (ii) has at least 80%
identity to the seq of SEQ ID#2.
3. A method according to claim 1 or 2 which comprises monitoring the
interaction between the LTRGW1 polypeptide and the leukotriene.
20
4. A method according to any preceeding claim wherein the LTRGW1
polypeptide is expressed in a cell.
5. Use of an LTRGW1 polypeptide as defined in claim 1 or 2 to enhance the
25 leukotriene B₄ Receptor (BLTR) response to leukotrienes.
6. An isolated heterodimer comprising an LTRGW1 polypeptide as defined in
claim 1 or 2 and a BLTR polypeptide comprising:
- i) The amino acid sequence of SEQ ID # 8, or
 - ii) A variant of (i) which is capable of binding leukotrienes or
30
 - iii) A fragment of (i) or (ii) which is capable of binding
leukotrienes.

7. A method for increasing the responsiveness of a screen for identification of a substance that modulates the activity of BLTR, comprising the addition to said screen of an LTRGW1 polypeptide as defined in claim 1 or 2.
8. A method for identification of a substance that modulates BLTR activity, which method comprises contacting an LTRGW1 polypeptide as defined in claim 1 or 2, and a BLTR polypeptide as defined in claim 6, with a test substance and monitoring for LTB₄ binding to the said polypeptides.
9. A method for identification of a substance that modulates BLTR receptor activity, which method comprises contacting an LTRGW1 polypeptide as defined in claim 1 or 2, and a BLTR polypeptide as defined in claim 6, with LTB₄ in the presence of a test substance and monitoring for BLTR activity.
10. A method according to claim 8 or 9 which comprises monitoring the activation of a G-protein.
11. A method for identification of a substance that modulates BLTR activity, which method comprises:
- (i) providing
 - (a) an LTRGW1 polypeptide as defined in claim 1 or 2; and
 - (b) a BLTR polypeptide as defined in claim 6; and
 - (c) a test substance
 - (ii) monitoring the interaction between (a) and (b); and
 - (iii) determining whether (c) modulates the interaction between (a) and (b) and thereby determining whether the test substance is a modulator of BLTR receptor activity.
12. A substance identified by a method according to any one of claims 1 to 4, 8, 9, 10 or 11.

- 5 13. A method of treating a subject having a disorder that is responsive to modulation of LTRGW1 or BLTR activity, which method comprises administering to said subject an effective amount of a substance according to claim 12.
14. A method according to claim 13 wherein said substance is an antagonist of LTRGW1 or BLTR.
- 10 15. A method of treating a subject having a disorder that is responsive to modulation of the interaction between LTRGW1 and BLTR, which method comprises administering to said subject an effective amount of a substance according to claim 14.
- 15 16. A method according to claim 15 wherein said substance downregulates the interaction between LTRGW1 and BLTR.
- 20 17. Use of a substance as defined in claim 12 in the manufacture of a medicament for treatment or prophylaxis of a disorder that is responsive to stimulation or modulation of LTRGW1 or BLTR activity.
18. Use according to claim 17 wherein said substance is an antagonist of LTRGW1 or BLTR
- 25 19. Use of a substance as defined in claim 12 in the manufacture of a medicament for treatment or prophylaxis of a disorder that is responsive to stimulation or modulation of the interaction between BLTR and LTRGW1
- 30 20. Use according to claim 19 wherein said substance downregulates the interaction between BLTR and LTRGW1.

21. A method according to any of ³¹ claims 13 to 16, or a use according to any of claims 17 to 20 wherein the disorder is an acute or chronic inflammatory disease, asthma, chronic obstructive pulmonary disease or psoriasis.
- 5 22. A method of treating a patient with a respiratory disorder, said method comprising the administration of a therapeutically effective amount of an antagonist of LTRGW1
- 10 23. A method of treating a respiratory disorder, said method comprising the administration to a patient of a therapeutically effective amount of a substance that downregulates the interaction between LTRGW1 and BLTR, hence decreasing the response of BLTR to its ligand.
- 15 24. Use of a therapeutically effective amount of an antagonist of LTRGW1 in the manufacture of a medicament for the treatment or prophylaxis of respiratory diseases.
- 20 25. Use of an effective amount of a substance that downregulates the interaction between LTRGW1 and BLTR in the manufacture of a medicament for the treatment or prophylaxis of respiratory disorders.
- 25 26. A method according to claim 22 or 23, or a use according to claim 24 or 25 wherein said respiratory disorder is asthma or chronic obstructive pulmonary disorder.

1 / 5

FIG. 1

LTRGW1 enhances BLTR (5ug) response in a dose dependent manner.

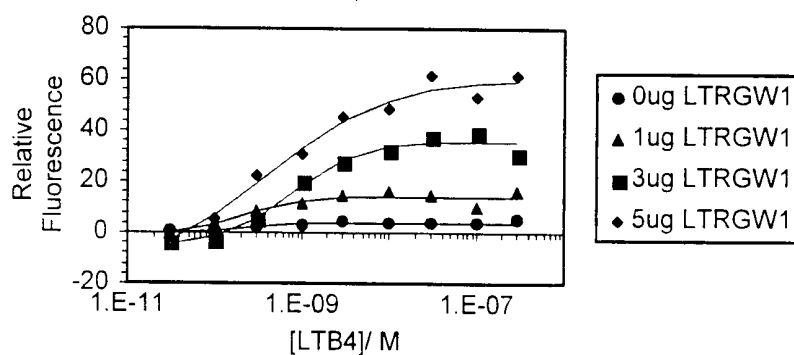
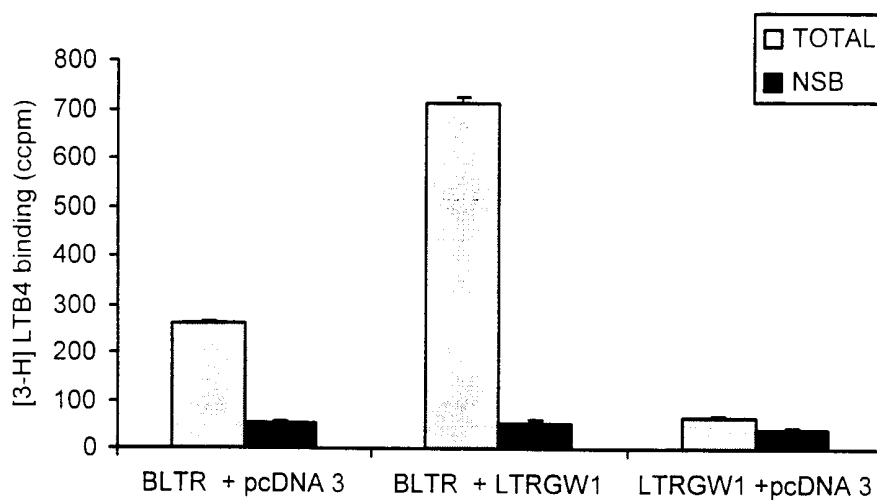


FIG. 2

LTB4 membrane binding
(10ug mems/0.5mg/well bead)



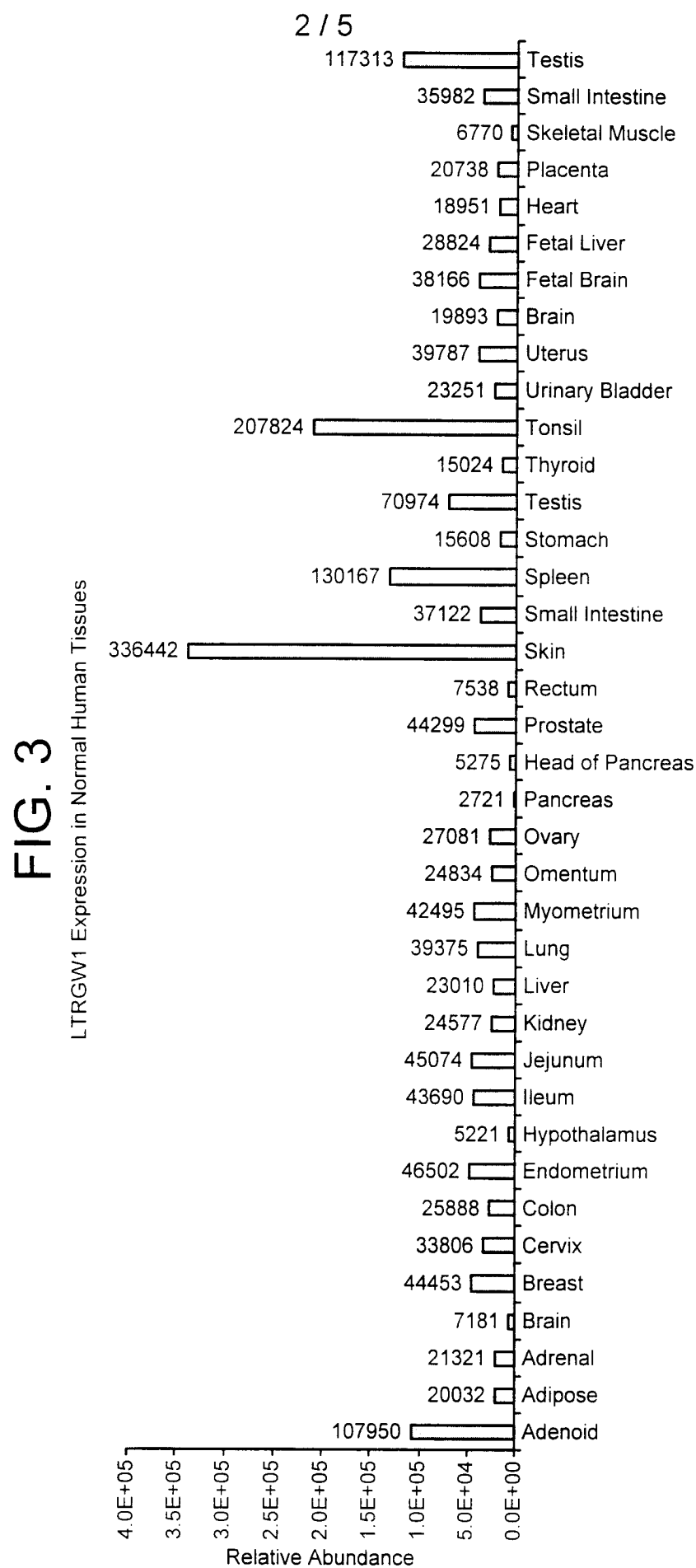


FIG. 4
BLTR Expression in Normal Human Tissues

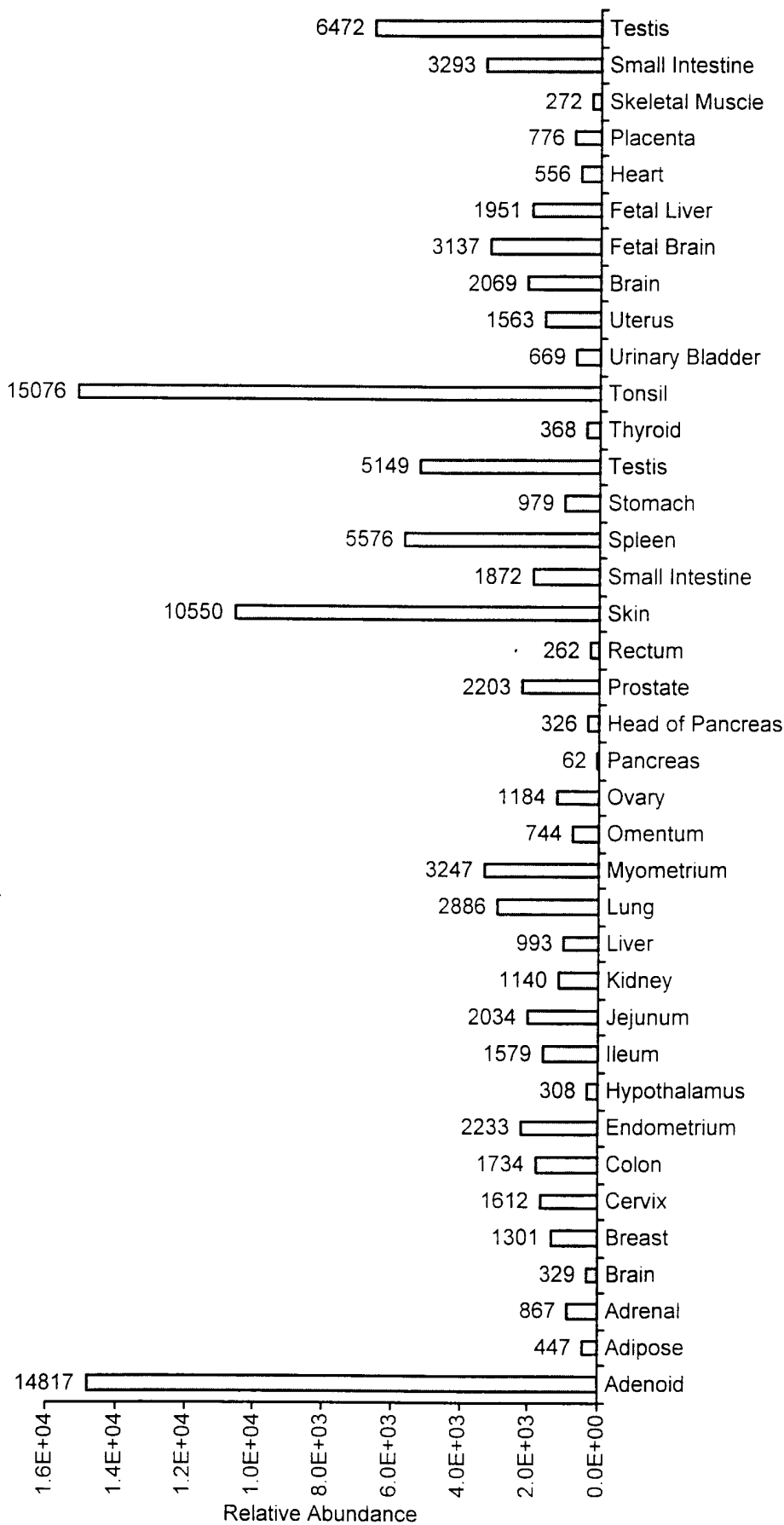
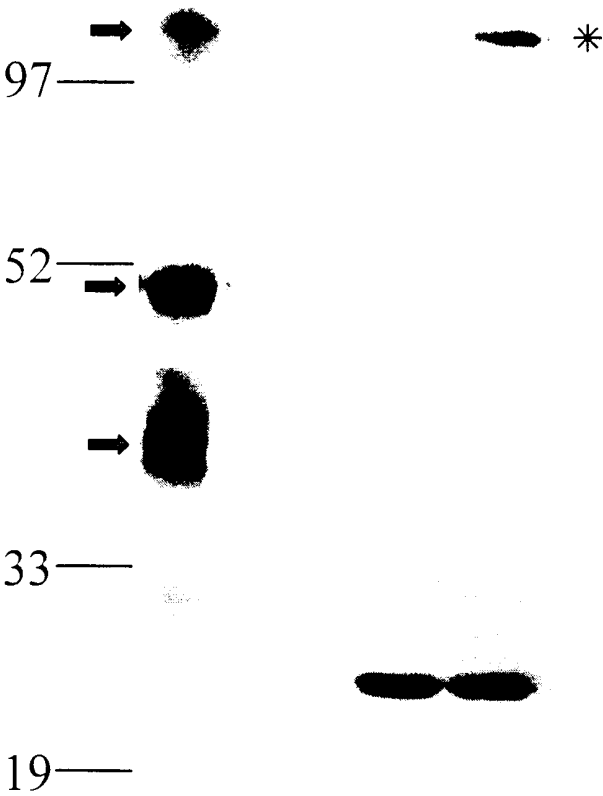


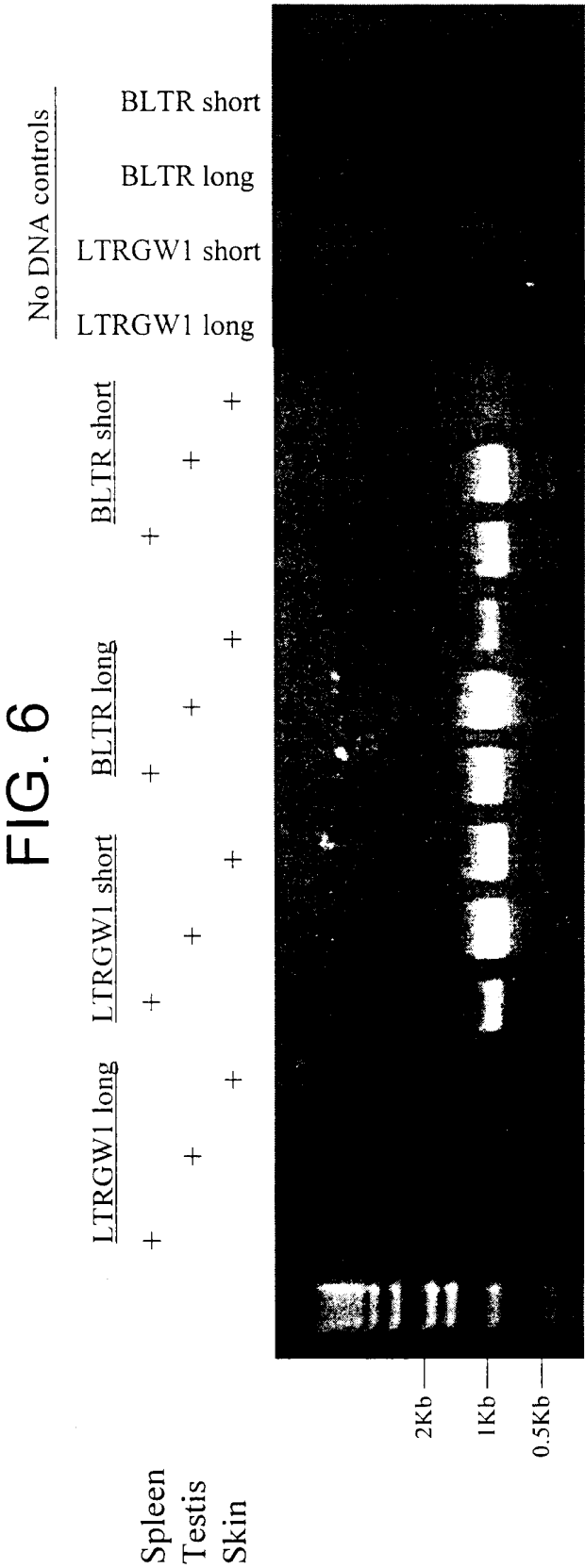
FIG. 5

	IP: α -		IP: α -	
Myc-GABA _B R1b		+	+	+
HA- GABA _B R2 *				+
VSV- BLTR	+			
HA- LTRGW1 ➡	+	+	+	+



WB: α -HA Blot

FIG. 6



SEQUENCE LISTING

5 <160> 6
 <170> PatentIn Ver. 2.1

 <210> 1
 <211> 1170
 10 <212> DNA
 <213> Homo sapiens

 <220>
 <221> CDS
 15 <222> (1)..(1170)

 <400> 1
 atg gca cct tct cat cgg gca tca cag gtg ggg ttt tgc ccc acc cct 48
 Met Ala Pro Ser His Arg Ala Ser Gln Val Gly Phe Cys Pro Thr Pro
 20 1 5 10 15

 gaa cgc cct ctg tgg cgc ctt cca ccc acc tgt agg ccc aga agg atg 96
 Glu Arg Pro Leu Trp Arg Leu Pro Pro Thr Cys Arg Pro Arg Arg Met
 25 20 25 30

 tcg gtc tgc tac cgt ccc cca ggg aac gag aca ctg ctg agc tgg aag 144
 Ser Val Cys Tyr Arg Pro Pro Gly Asn Glu Thr Leu Leu Ser Trp Lys
 35 40 45

 30 act tcg cgg gcc aca ggc aca gcc ttc ctg ctg ctg gcg gcg ctg ctg 192
 Thr Ser Arg Ala Thr Gly Thr Ala Phe Leu Leu Leu Ala Ala Leu Leu
 50 55 60

 ggg ctg cct ggc aac ggc ttc gtg gtg tgg agc ttg gcg ggc tgg cgg 240
 35 Gly Leu Pro Gly Asn Gly Phe Val Val Trp Ser Leu Ala Gly Trp Arg
 65 70 75 80

 cct gca cgg ggg cga ccg ctg gcg gcc acg ctt gtg ctg cac ctg gcg 288
 40 Pro Ala Arg Gly Arg Pro Leu Ala Ala Thr Leu Val Leu His Leu Ala
 85 90 95

 ctg gcc gac ggc gcg gtg ctg ctg ctc acg ccg ctc ttt gtg gcc ttc 336
 Leu Ala Asp Gly Ala Val Leu Leu Leu Thr Pro Leu Phe Val Ala Phe
 100 105 110
 45
 ctg acc cgg cag gcc tgg ccg ctg ggc cag gcg ggc tgc aag gcg gtg 384
 Leu Thr Arg Gln Ala Trp Pro Leu Gly Gln Ala Gly Cys Lys Ala Val
 115 120 125

 50 tac tac gtg tgc gcg ctc agc atg tac gcc agc gtg ctg ctc acc ggc 432
 Tyr Tyr Val Cys Ala Leu Ser Met Tyr Ala Ser Val Leu Leu Thr Gly
 130 135 140

 ctg ctc agc ctg cag cgc tgc ctc gca gtc acc cgc ccc ttc ctg gcg 480
 55 Leu Leu Ser Leu Gln Arg Cys Leu Ala Val Thr Arg Pro Phe Leu Ala
 145 150 155 160

	cct	cgg	ctg	cgc	agc	ccg	gcc	ctg	gcc	cgc	cgc	ctg	ctg	ctg	gcg	gtc	528
	Pro	Arg	Leu	Arg	Ser	Pro	Ala	Leu	Ala	Arg	Arg	Leu	Leu	Leu	Ala	Val	
				165						170					175		
5	tgg	ctg	gcc	gcc	ctg	ttg	ctc	gcc	gtc	ccg	gcc	gcc	gtc	tac	cgc	cac	576
	Trp	Leu	Ala	Ala	Leu	Leu	Leu	Ala	Val	Pro	Ala	Ala	Val	Tyr	Arg	His	
			180					185						190			
10	ctg	tgg	agg	gac	cgc	gta	tgc	cag	ctg	tgc	cac	ccg	tcg	ccg	gtc	cac	624
	Leu	Trp	Arg	Asp	Arg	Val	Cys	Gln	Leu	Cys	His	Pro	Ser	Pro	Val	His	
			195					200					205				
15	gcc	gcc	gcc	cac	ctg	agc	ctg	gag	act	ctg	acc	gct	ttc	gtg	ctt	cct	672
	Ala	Ala	Ala	His	Leu	Ser	Leu	Glu	Thr	Leu	Thr	Ala	Phe	Val	Leu	Pro	
		210						215				220					
20	ttc	ggg	ctg	atg	ctc	ggc	tgc	tac	agc	gtg	acg	ctg	gca	cgg	ctg	cgg	20
	Phe	Gly	Leu	Met	Leu	Gly	Cys	Tyr	Ser	Val	Thr	Leu	Ala	Arg	Leu	Arg	
	225					230					235					240	
25	ggc	gcc	cgc	tgg	ggc	tcc	ggg	cgg	cac	ggg	gcg	cgg	gtg	ggc	cgg	ctg	768
	Gly	Ala	Arg	Trp	Gly	Ser	Gly	Arg	His	Gly	Ala	Arg	Val	Gly	Arg	Leu	
				245						250				255			
30	gtg	agc	gcc	atc	gtg	ctt	gcc	ttc	ggc	ttg	ctc	tgg	gcc	ccc	tac	cac	816
	Val	Ser	Ala	Ile	Val	Leu	Ala	Phe	Gly	Leu	Leu	Trp	Ala	Pro	Tyr	His	
			260					265					270				
35	gca	gtc	aac	ctt	ctg	cag	gcg	gtc	gca	gcg	ctg	gct	cca	ccg	gaa	ggg	864
	Ala	Val	Asn	Leu	Leu	Gln	Ala	Val	Ala	Ala	Leu	Ala	Pro	Pro	Glu	Gly	
			275					280					285				
40	gcc	ttg	gcg	aag	ctg	ggc	gga	gcc	ggc	cag	gcg	gcg	cga	gcg	gga	act	912
	Ala	Leu	Ala	Lys	Leu	Gly	Gly	Ala	Gly	Gln	Ala	Ala	Arg	Ala	Gly	Thr	
		290						295					300				
45	acg	gcc	ttg	gcc	ttc	ttc	agt	tct	agc	gtc	aac	ccg	gtg	ctc	tac	gtc	960
	Thr	Ala	Leu	Ala	Phe	Phe	Ser	Ser	Ser	Val	Asn	Pro	Val	Leu	Tyr	Val	
	305					310					315				320		
50	ttc	acc	gct	gga	gat	ctg	ctg	ccc	cgg	gca	ggt	ccc	cgt	ttc	ctc	acg	1008
	Phe	Thr	Ala	Gly	Asp	Leu	Leu	Pro	Arg	Ala	Gly	Pro	Arg	Phe	Leu	Thr	
				325						330				335			
55	cgg	ctc	ttc	gaa	ggc	tct	ggg	gag	gcc	cga	ggg	ggc	ggc	cgc	tct	agg	1056
	Arg	Leu	Phe	Glu	Gly	Ser	Gly	Glu	Ala	Arg	Gly	Gly	Gly	Arg	Ser	Arg	
			340						345					350			
60	gaa	ggg	acc	atg	gag	ctc	cga	act	acc	cct	cag	ctg	aaa	gtg	gtg	ggg	1104
	Glu	Gly	Thr	Met	Glu	Leu	Arg	Thr	Thr	Pro	Gln	Leu	Lys	Val	Val	Gly	
			355					360					365				
65	cag	ggc	cgc	ggc	aat	gga	gac	ccg	ggg	ggt	ggg	atg	gag	aag	gac	ggt	1152
	Gln	Gly	Arg	Gly	Asn	Gly	Asp	Pro	Gly	Gly	Gly	Met	Glu	Lys	Asp	Gly	
		370						375				380					

3
1170

ccg gaa tgg gac ctt tga
Pro Glu Trp Asp Leu
385 390

5

<210> 2
<211> 389
10 <212> PRT
<213> Homo sapiens

<400> 2
Met Ala Pro Ser His Arg Ala Ser Gln Val Gly Phe Cys Pro Thr Pro
15 1 5 10 15

Glu Arg Pro Leu Trp Arg Leu Pro Pro Thr Cys Arg Pro Arg Arg Met
20 20 25 30

Ser Val Cys Tyr Arg Pro Pro Gly Asn Glu Thr Leu Leu Ser Trp Lys
25 35 40 45

Thr Ser Arg Ala Thr Gly Thr Ala Phe Leu Leu Leu Ala Ala Leu Leu
50 55 60

Gly Leu Pro Gly Asn Gly Phe Val Val Trp Ser Leu Ala Gly Trp Arg
65 70 75 80

Pro Ala Arg Gly Arg Pro Leu Ala Ala Thr Leu Val Leu His Leu Ala
30 85 90 95

Leu Ala Asp Gly Ala Val Leu Leu Leu Thr Pro Leu Phe Val Ala Phe
100 105 110

Leu Thr Arg Gln Ala Trp Pro Leu Gly Gln Ala Gly Cys Lys Ala Val
35 115 120 125

Tyr Tyr Val Cys Ala Leu Ser Met Tyr Ala Ser Val Leu Leu Thr Gly
40 130 135 140

Leu Leu Ser Leu Gln Arg Cys Leu Ala Val Thr Arg Pro Phe Leu Ala
145 150 155 160

Pro Arg Leu Arg Ser Pro Ala Leu Ala Arg Arg Leu Leu Leu Ala Val
45 165 170 175

Trp Leu Ala Ala Leu Leu Leu Ala Val Pro Ala Ala Val Tyr Arg His
180 185 190

Leu Trp Arg Asp Arg Val Cys Gln Leu Cys His Pro Ser Pro Val His
50 195 200 205

Ala Ala Ala His Leu Ser Leu Glu Thr Leu Thr Ala Phe Val Leu Pro
210 215 220

Phe Gly Leu Met Leu Gly Cys Tyr Ser Val Thr Leu Ala Arg Leu Arg
55 225 230 235 240

Gly Ala Arg Trp Gly Ser Gly Arg His Gly Ala Arg Val Gly Arg Leu
 245 250 255
 5 Val Ser Ala Ile Val Leu Ala Phe Gly Leu Leu Trp Ala Pro Tyr His
 260 265 270
 Ala Val Asn Leu Leu Gln Ala Val Ala Ala Leu Ala Pro Pro Glu Gly
 275 280 285
 10 Ala Leu Ala Lys Leu Gly Gly Ala Gly Gln Ala Ala Arg Ala Gly Thr
 290 295 300
 Thr Ala Leu Ala Phe Phe Ser Ser Ser Val Asn Pro Val Leu Tyr Val
 15 305 310 315 320
 Phe Thr Ala Gly Asp Leu Leu Pro Arg Ala Gly Pro Arg Phe Leu Thr
 325 330 335
 20 Arg Leu Phe Glu Gly Ser Gly Glu Ala Arg Gly Gly Gly Arg Ser Arg
 340 345 350
 Glu Gly Thr Met Glu Leu Arg Thr Thr Pro Gln Leu Lys Val Val Gly
 355 360 365
 25 Gln Gly Arg Gly Asn Gly Asp Pro Gly Gly Gly Met Glu Lys Asp Gly
 370 375 380
 Pro Glu Trp Asp Leu
 30 385
 <210> 3
 <211> 1170
 <212> DNA
 35 <213> Homo sapiens
 <220>
 <221> CDS
 <222> (94)..(1170)
 40
 <400> 3
 atg gca cct tct cat cgg gca tca cag gtg ggg ttt tgc ccc acc cct 48
 45 gaa cgc cct ctg tgg cgc ctt cca ccc acc tgt agg ccc aga agg atg 96
 Met
 1
 tcg gtc tgc tac cgt ccc cca ggg aac gag aca ctg ctg agc tgg aag 144
 50 Ser Val Cys Tyr Arg Pro Pro Gly Asn Glu Thr Leu Leu Ser Trp Lys
 5 10 15
 act tcg cgg gcc aca ggc aca gcc ttc ctg ctg ctg gcg gcg ctg ctg 192
 Thr Ser Arg Ala Thr Gly Thr Ala Phe Leu Leu Leu Ala Ala Leu Leu
 55 20 25 30
 ggg ctg cct ggc aac ggc ttc gtg gtg tgg agc ttg gcg ggc tgg cgg 240

SUBSTITUTE SHEET (RULE 26)

6
270

260 265

5 275 280 285

acg gcc ttg gcc ttc ttc agt tct agc gtc aac ccg gtg ctc tac gtc 960
 Thr Ala Leu Ala Phe Phe Ser Ser Ser Val Asn Pro Val Leu Tyr Val

10 290 295 300 305

ttc acc gct gga gat ctg ctg ccc cgg gca ggt ccc cgt ttc ctc acg 1008
 Phe Thr Ala Gly Asp Leu Leu Pro Arg Ala Gly Pro Arg Phe Leu Thr

1056
 cgg ctc ttc gaa ggc tct ggg gag gcc cga ggg ggc ggc cgc tct agg
 Arg Leu Phe Glu Gly Ser Gly Glu Ala Arg Gly Gly Gly Arg Ser Arg

15 310 315 320

gaa ggg acc atg gag ctc cga act acc cct cag ctg aaa gtg gtg ggg 1104
 Glu Gly Thr Met Glu Leu Arg Thr Thr Pro Gln Leu Lys Val Val Gly

20 325 330 335

cag ggc cgc ggc aat gga gac ccg ggg ggt ggg atg gag aag gac ggt 1152
 Gln Gly Arg Gly Asn Gly Asp Pro Gly Gly Gly Met Glu Lys Asp Gly

25 340 345 350

ccg gaa tgg gac ctt tga 1170
 Pro Glu Trp Asp Leu

30 355

30 <210> 4
 <211> 358
 <212> PRT
 <213> Homo sapiens

35 <400> 4
 Met Ser Val Cys Tyr Arg Pro Pro Gly Asn Glu Thr Leu Leu Ser Trp
 1 5 10 15

40 20 25 30

Lys Thr Ser Arg Ala Thr Gly Thr Ala Phe Leu Leu Leu Ala Ala Leu

45 35 40 45

Leu Gly Leu Pro Gly Asn Gly Phe Val Val Trp Ser Leu Ala Gly Trp

50 50 55 60

Arg Pro Ala Arg Gly Arg Pro Leu Ala Ala Thr Leu Val Leu His Leu

55 65 70 75 80

Ala Leu Ala Asp Gly Ala Val Leu Leu Leu Thr Pro Leu Phe Val Ala

85 90 95

Phe Leu Thr Arg Gln Ala Trp Pro Leu Gly Gln Ala Gly Cys Lys Ala

100 105 110

Val Tyr Tyr Val Cys Ala Leu Ser Met Tyr Ala Ser Val Leu Leu Thr

Gly Leu Leu Ser Leu Gln Arg Cys Leu Ala Val Thr Arg Pro Phe Leu

<400> 5																	
	atg aac act aca tct tct gca gca ccc ccc tca cta ggt gta gag ttc	48															
	Met Asn Thr Thr Ser Ser Ala Ala Pro Pro Ser Leu Gly Val Glu Phe																
	1 5 10 15																
5																	
	atc tct ctg ctg gct atc atc ctg ctg tca gtg gcg ctg gct gtg ggg	96															
	Ile Ser Leu Leu Ala Ile Ile Leu Leu Ser Val Ala Leu Ala Val Gly																
	20 25 30																
10																	
	ctt ccc ggc aac agc ttt gtg gtg tgg agt atc ctg aaa agg atg cag	144															
	Leu Pro Gly Asn Ser Phe Val Val Trp Ser Ile Leu Lys Arg Met Gln																
	35 40 45																
15																	
	aag cgc tct gtc act gcc ctg atg gtg ctg aac ctg gcc ctg gcc gac	192															
	Lys Arg Ser Val Thr Ala Leu Met Val Leu Asn Leu Ala Leu Ala Asp																
	50 55 60																
20																	
	ctg gcc gta ttg ctc act gct ccc ttt ttc ctt cac ttc ctg gcc caa	240															
	Leu Ala Val Leu Leu Thr Ala Pro Phe Phe Leu His Phe Leu Ala Gln																
	65 70 75 80																
25																	
	ggc acc tgg agt ttt gga ctg gct ggt tgc cgc ctg tgt cac tat gtc	288															
	Gly Thr Trp Ser Phe Gly Leu Ala Gly Cys Arg Leu Cys His Tyr Val																
	85 90 95																
30																	
	tgc gga gtc agc atg tac gcc agc gtc ctg ctt atc acg gcc atg agt	336															
	Cys Gly Val Ser Met Tyr Ala Ser Val Leu Leu Ile Thr Ala Met Ser																
	100 105 110																
35																	
	cta gac cgc tca ctg gcg gtg gcc cgc ccc ttt gtg tcc cag aag cta	384															
	Leu Asp Arg Ser Leu Ala Val Ala Arg Pro Phe Val Ser Gln Lys Leu																
	115 120 125																
40																	
	cgc acc aag gcg atg gcc cgg cgg gtg ctg gca ggc atc tgg gtg ttg	432															
	Arg Thr Lys Ala Met Ala Arg Arg Val Leu Ala Gly Ile Trp Val Leu																
	130 135 140																
45																	
	tcc ttt ctg ctg gcc aca ccc gtc ctc gcg tac cgc aca gta gtg ccc	480															
	Ser Phe Leu Leu Ala Thr Pro Val Leu Ala Tyr Arg Thr Val Val Pro																
	145 150 155 160																
50																	
	tgg aaa acg aac atg agc ctg tgc ttc ccg cgg tac ccc agc gaa ggg	528															
	Trp Lys Thr Asn Met Ser Leu Cys Phe Pro Arg Tyr Pro Ser Glu Gly																
	165 170 175																
55																	
	cac cgg gcc ttc cat cta atc ttc gag gct gtc acg ggc ttc ctg ctg	576															
	His Arg Ala Phe His Leu Ile Phe Glu Ala Val Thr Gly Phe Leu Leu																
	180 185 190																
60																	
	ccc ttc ctg gct gtg gtg gcc agc tac tcg gac ata ggg cgt cgg cta	624															
	Pro Phe Leu Ala Val Val Ala Ser Tyr Ser Asp Ile Gly Arg Arg Leu																
	195 200 205																
65																	
	cag gcc cgg cgc ttc cgc cgc agc cgc cgc acc ggc cgc ctg gtg gtg	672															
	Gln Ala Arg Arg Phe Arg Arg Ser Arg Arg Thr Gly Arg Leu Val Val																
	210 215 220																

SUBSTITUTE SHEET (RULE 26)

Gly Thr Trp Ser Phe Gly Leu Ala Gly Cys Arg Leu Cys His Tyr Val
 85 90 95
 5 Cys Gly Val Ser Met Tyr Ala Ser Val Leu Leu Ile Thr Ala Met Ser
 100 105 110
 Leu Asp Arg Ser Leu Ala Val Ala Arg Pro Phe Val Ser Gln Lys Leu
 115 120 125
 10 Arg Thr Lys Ala Met Ala Arg Arg Val Leu Ala Gly Ile Trp Val Leu
 130 135 140
 Ser Phe Leu Leu Ala Thr Pro Val Leu Ala Tyr Arg Thr Val Val Pro
 145 150 155 160
 15 Trp Lys Thr Asn Met Ser Leu Cys Phe Pro Arg Tyr Pro Ser Glu Gly
 165 170 175
 20 His Arg Ala Phe His Leu Ile Phe Glu Ala Val Thr Gly Phe Leu Leu
 180 185 190
 Pro Phe Leu Ala Val Val Ala Ser Tyr Ser Asp Ile Gly Arg Arg Leu
 195 200 205
 25 Gln Ala Arg Arg Phe Arg Arg Ser Arg Arg Thr Gly Arg Leu Val Val
 210 215 220
 Leu Ile Ile Leu Thr Phe Ala Ala Phe Trp Leu Pro Tyr His Val Val
 225 230 235 240
 30 Asn Leu Ala Glu Ala Gly Arg Ala Leu Ala Gly Gln Ala Ala Gly Leu
 245 250 255
 35 Gly Leu Val Gly Lys Arg Leu Ser Leu Ala Arg Asn Val Leu Ile Ala
 260 265 270
 Leu Ala Phe Leu Ser Ser Ser Val Asn Pro Val Leu Tyr Ala Cys Ala
 275 280 285
 40 Gly Gly Gly Leu Leu Arg Ser Ala Gly Val Gly Phe Val Ala Lys Leu
 290 295 300
 Leu Glu Gly Thr Gly Ser Glu Ala Ser Ser Thr Arg Arg Gly Gly Ser
 305 310 315 320
 45 Leu Gly Gln Thr Ala Arg Ser Gly Pro Ala Ala Leu Glu Pro Gly Pro
 325 330 335
 Ser Glu Ser Leu Thr Ala Ser Ser Pro Leu Lys Leu Asn Glu Leu Asn
 340 345 350
 50
 <210> 7
 <211> 1134
 55 <212> DNA
 <213> homo sapiens

<220>
 <221> CDS
 <222> (1)..(1134)

5
 <400> 7
 atg gcg tca gga aac cct tgg tcc tct act ctc atg cgt gtg tcc gcc 48
 Met Ala Ser Gly Asn Pro Trp Ser Ser Thr Leu Met Arg Val Ser Ala
 1 5 10 15

10
 ctc act ctc cag gtc ctc ccg acg gcc atg aac act aca tct tct gca 96
 Leu Thr Leu Gln Val Leu Pro Thr Ala Met Asn Thr Thr Ser Ser Ala
 20 25 30

15
 gca ccc ccc tca cta ggt gta gag ttc atc tct ctg ctg gct atc atc 144
 Ala Pro Pro Ser Leu Gly Val Glu Phe Ile Ser Leu Leu Ala Ile Ile
 35 40 45

20
 ctg ctg tca gtg gcg ctg gct gtg ggg ctt ccc gcc aac agc ttt gtg 192
 Leu Leu Ser Val Ala Leu Ala Val Gly Leu Pro Gly Asn Ser Phe Val
 50 55 60

25
 gtg tgg agt atc ctg aaa agg atg cag aag cgc tct gtc act gcc ctg 240
 Val Trp Ser Ile Leu Lys Arg Met Gln Lys Arg Ser Val Thr Ala Leu
 65 70 75 80

30
 atg gtg ctg aac ctg gcc ctg gcc gac ctg gcc gta ttg ctc act gct 288
 Met Val Leu Asn Leu Ala Leu Ala Asp Leu Ala Val Leu Leu Thr Ala
 85 90 95

35
 ccc ttt ttc ctt cac ttc ctg gcc caa ggc acc tgg agt ttt gga ctg 336
 Pro Phe Phe Leu His Phe Leu Ala Gln Gly Thr Trp Ser Phe Gly Leu
 100 105 110

40
 gct ggt tgc cgc ctg tgt cac tat gtc tgc gga gtc agc atg tac gcc 384
 Ala Gly Cys Arg Leu Cys His Tyr Val Cys Gly Val Ser Met Tyr Ala
 115 120 125

45
 agc gtc ctg ctt atc acg gcc atg agt cta gac cgc tca ctg gcg gtg 432
 Ser Val Leu Leu Ile Thr Ala Met Ser Leu Asp Arg Ser Leu Ala Val
 130 135 140

50
 gcc cgc ccc ttt gtg tcc cag aag cta cgc acc aag gcg atg gcc cgg 480
 Ala Arg Pro Phe Val Ser Gln Lys Leu Arg Thr Lys Ala Met Ala Arg
 145 150 155 160

55
 cgg gtg ctg gca ggc atc tgg gtg ttg tcc ttt ctg ctg gcc aca ccc 528
 Arg Val Leu Ala Gly Ile Trp Val Leu Ser Phe Leu Leu Ala Thr Pro
 165 170 175

60
 gtc ctc gcg tac cgc aca gta gtg ccc tgg aaa acg aac atg agc ctg 576
 Val Leu Ala Tyr Arg Thr Val Val Pro Trp Lys Thr Asn Met Ser Leu
 180 185 190

65
 tgc ttc ccg cgg tac ccc agc gaa ggg cac cgg gcc ttc cat cta atc 624
 Cys Phe Pro Arg Tyr Pro Ser Glu Gly His Arg Ala Phe His Leu Ile
 195 200 205

	ttc gag gct gtc acg ggc ttc ctg ctg ccc ttc ctg gct gtg gtg gcc	672
	Phe Glu Ala Val Thr Gly Phe Leu Leu Pro Phe Leu Ala Val Val Ala	
	210 215 220	
5	agc tac tcg gac ata ggg cgt cgg cta cag gcc cgg cgc ttc cgc cgc	720
	Ser Tyr Ser Asp Ile Gly Arg Arg Leu Gln Ala Arg Arg Phe Arg Arg	
	225 230 235 240	
10	agc cgc cgc acc ggc cgc ctg gtg gtg ctc atc atc ctg acc ttc gcc	768
	Ser Arg Arg Thr Gly Arg Leu Val Val Leu Ile Ile Leu Thr Phe Ala	
	245 250 255	
15	gcc ttc tgg ctg ccc tac cac gtg gtg aac ctg gct gag gcg ggc cgc	816
	Ala Phe Trp Leu Pro Tyr His Val Val Asn Leu Ala Glu Ala Gly Arg	
	260 265 270	
20	gcg ctg gcc ggc cag gcc gcc ggg tta ggg ctc gtg ggg aag cgg ctg	864
	Ala Leu Ala Gly Gln Ala Ala Gly Leu Gly Leu Val Gly Lys Arg Leu	
	275 280 285	
	agc ctg gcc cgc aac gtg ctc atc gca ctc gcc ttc ctg agc agc agc	912
	Ser Leu Ala Arg Asn Val Leu Ile Ala Leu Ala Phe Leu Ser Ser Ser	
	290 295 300	
25	gtg aac ccc gtg ctg tac gcg tgc gcc ggc ggc ggc ctg ctg cgc tcg	960
	Val Asn Pro Val Leu Tyr Ala Cys Ala Gly Gly Gly Leu Leu Arg Ser	
	305 310 315 320	
30	gcg ggc gtg ggc ttc gtc gcc aag ctg ctg gag ggc acg ggc tcc gag	1008
	Ala Gly Val Gly Phe Val Ala Lys Leu Leu Glu Gly Thr Gly Ser Glu	
	325 330 335	
35	gcg tcc agc acg cgc cgc ggg ggc agc ctg ggc cag acc gct agg agc	1056
	Ala Ser Ser Thr Arg Arg Gly Gly Ser Leu Gly Gln Thr Ala Arg Ser	
	340 345 350	
40	ggc ccc gcc gct ctg gag ccc ggc cct tcc gag agc ctc act gcc tcc	1104
	Gly Pro Ala Ala Leu Glu Pro Gly Pro Ser Glu Ser Leu Thr Ala Ser	
	355 360 365	
	agc cct ctc aag tta aac gaa ctg aac tag	1134
	Ser Pro Leu Lys Leu Asn Glu Leu Asn	
	370 375	
45		
	<210> 8	
	<211> 377	
	<212> PRT	
50	<213> homo sapiens	
	<400> 8	
	Met Ala Ser Gly Asn Pro Trp Ser Ser Thr Leu Met Arg Val Ser Ala	
	1 5 10 15	
55	Leu Thr Leu Gln Val Leu Pro Thr Ala Met Asn Thr Thr Ser Ser Ala	
	20 25 30	

Ala Pro Pro Ser Leu Gly Val Glu Phe Ile Ser Leu Leu Ala Ile Ile
 35 40 45
 5 Leu Leu Ser Val Ala Leu Ala Val Gly Leu Pro Gly Asn Ser Phe Val
 50 55 60
 Val Trp Ser Ile Leu Lys Arg Met Gln Lys Arg Ser Val Thr Ala Leu
 65 70 75 80
 10 Met Val Leu Asn Leu Ala Leu Ala Asp Leu Ala Val Leu Leu Thr Ala
 85 90 95
 Pro Phe Phe Leu His Phe Leu Ala Gln Gly Thr Trp Ser Phe Gly Leu
 15 100 105 110
 Ala Gly Cys Arg Leu Cys His Tyr Val Cys Gly Val Ser Met Tyr Ala
 115 120 125
 20 Ser Val Leu Leu Ile Thr Ala Met Ser Leu Asp Arg Ser Leu Ala Val
 130 135 140
 Ala Arg Pro Phe Val Ser Gln Lys Leu Arg Thr Lys Ala Met Ala Arg
 145 150 155 160
 25 Arg Val Leu Ala Gly Ile Trp Val Leu Ser Phe Leu Leu Ala Thr Pro
 165 170 175
 Val Leu Ala Tyr Arg Thr Val Val Pro Trp Lys Thr Asn Met Ser Leu
 30 180 185 190
 Cys Phe Pro Arg Tyr Pro Ser Glu Gly His Arg Ala Phe His Leu Ile
 195 200 205
 35 Phe Glu Ala Val Thr Gly Phe Leu Leu Pro Phe Leu Ala Val Val Ala
 210 215 220
 Ser Tyr Ser Asp Ile Gly Arg Arg Leu Gln Ala Arg Arg Phe Arg Arg
 225 230 235 240
 40 Ser Arg Arg Thr Gly Arg Leu Val Val Leu Ile Ile Leu Thr Phe Ala
 245 250 255
 Ala Phe Trp Leu Pro Tyr His Val Val Asn Leu Ala Glu Ala Gly Arg
 45 260 265 270
 Ala Leu Ala Gly Gln Ala Ala Gly Leu Gly Leu Val Gly Lys Arg Leu
 275 280 285
 50 Ser Leu Ala Arg Asn Val Leu Ile Ala Leu Ala Phe Leu Ser Ser Ser
 290 295 300
 Val Asn Pro Val Leu Tyr Ala Cys Ala Gly Gly Gly Leu Leu Arg Ser
 305 310 315 320
 55 Ala Gly Val Gly Phe Val Ala Lys Leu Leu Glu Gly Thr Gly Ser Glu
 325 330 335

14

Ala Ser Ser Thr Arg Arg Gly Gly Ser Leu Gly Gln Thr Ala Arg Ser
340 345 350

5 Gly Pro Ala Ala Leu Glu Pro Gly Pro Ser Glu Ser Leu Thr Ala Ser
355 360 365

Ser Pro Leu Lys Leu Asn Glu Leu Asn
370 375

10