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(54) Title: DISPERSIBLE PHOSPHOLIPID STABILIZED MICROPARTICLES

(57) Abstract

Rapidly dispersing solid dry therapeutic dosage form comprised of a water insoluble compound existing as a nanometer or micrometer particulate solid which is surface stabilized by the presence of at least one phospholipid, the particulate solid being dispersed throughout a bulking matrix. When the dosage form is introduced into an aqueous environment the bulking matrix is substantially completely dissolved within less than 2 minutes thereby releasing the water insoluble particulate solid in an unaggregated and/or unagglomerated state. The matrix is composed of a water insoluble substance or therapeutically useful water insoluble or poorly water soluble compound, a phospholipid and optionally also at least one non–ionic, anionic, cationic or amphipathic surfactant, together with a matrix or bulking agent and if needed a release agent. The volume weighted mean particle size of the water insoluble particle is 5 micrometers or less.

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DISPERSIBLE PHOSPHOLIPID STABILIZED MICROPARTICLES

FIELD OF THE INVENTION

This invention relates to compositions comprised of water-insoluble or poorly soluble drug particles of a size of about 0.05 to 10 micrometers having a surface modifying agent or combination of agents, of which at least one is a phospholipid, adsorbed on to the surface thereof. The composition includes a matrix-forming agent(s) which is present in an amount sufficient to allow freeze-drying and subsequent release of the surface coated drug particles upon contact with an aqueous environment. Small surface coated particles are sometimes referred to as MicroCrystals (in US Pat. No. 5,091,187 and 5,091,188), MicroParticles (WO 98/07414), NanoParticles (US 5,145,684 and 5,302,401 and US 5,145,684).

This invention further provides methods of making dried compositions of water-insoluble or poorly soluble drug particles having surface modifying agents or combinations of agents, of which at least one is a phospholipid, adsorbed on the surface thereof and matrix-forming agent(s). The matrix-forming agent(s) is present in an amount sufficient to allow freeze-drying, such as by lyophilization, with subsequent release of the surface coated drug particles upon contact with an aqueous environment. The method comprises contacting said phospholipid coated particle with the matrix-forming agent(s) for a time and under conditions sufficient to allow the phospholipid coated drug particles to be freeze-dried.

BACKGROUND OF THE INVENTION

Poor bioavailability of water insoluble compounds has long been a problem in the pharmaceutical and diagnostics industry. While compounds with an aqueous solubility of greater than 1% w/v are not expected to present dissolution-related bioavailability and absorption problems, many new chemical entities exhibit aqueous solubility much below this value (see Pharmaceutical Dosage Forms – Tablets, Vol 1, page 13, Edited by H. Lieberman, Marcel Dekker, Inc, 1980). Many highly useful compounds are dropped from development or are formulated in a manner otherwise undesirable due to poor water solubility. A great number of these compounds are unstable in aqueous media and some require dissolution in oil, rendering the dosage form often unpleasant to take or even painful to use via the parenteral

route of administration. This can lead to poor patient compliance and potentially an overall greater expense in treatment due to unnecessary hospitalizations. It is therefore desirable to develop a formulation of these water insoluble compounds that can be dosed in the simplest possible form: a rapidly dispersing solid dosage form.

Many methods exist for preparing rapidly dispersing solid dosage medicaments. Traditional approaches to this problem have involved the dispersion of a biological active ingredient with pharmaceutically acceptable excipients using mix techniques and/or granulation techniques. Specific functional excipients known in the art can be employed to aid in liberating the medicament, as for example effervescent disintegration agents(s) as taught by U.S. 5,178,878.

As a method of improving the disintegration of the solid dosage form, thereby liberating the medicament, freeze drying techniques have been previously employed as described by U.S. Pat Nos. 4,371,516; 4,758,598; 5,272,137. Additionally, spray drying techniques have been employed for similar purposes as for example, US Pat 5,776,491 which describes the use of a polymeric component, a solubilizing component and a bulking agent as a matrix forming composition upon spray drying. This particulate matrix rapidly disintegrates upon introduction to an aqueous environment to release the medicament. Although these approaches produce rapidly drug liberating solid dosage forms, they suffer from a number of disadvantages particularly with medicaments that are water insoluble or poorly water-soluble. In these cases, suspensions of water insoluble compounds are likely to sediment prior to completion of the freeze-drying or spray drying process leading to particle aggregation and potentially inhomogeneous dry dosage forms. Additionally, large macromolecules of polysaccharides, typified by dextrans, when utilized as matrix formers have been implicated in agglomeration tendencies in reconstituted freeze-dried suspensions of liposomes (Miyajima, 1997). Therefore, the proper selection and employment of saccharide matrix formers remains elusive, we believe it is linked to the surface physicochemical nature of the water insoluble particle under consideration.

Additionally, suspensions of water insoluble compounds will be subjected to unwanted particle size growth as a result of the process of Ostwald ripening. In order to curtail this process, stabilization of these micronized materials suspended in an aqueous environment can

be achieved using compositions of a variety of pharmaceutically acceptable excipients known to those skilled in the art. Such approaches can be found, as example, in the commonly assigned U.S. Pat. Nos. 5,631,023 and 5,302,401 and EP0193208.

For instance, US Patent 5,631,023 discloses a method to prepare rapidly dissolving tablets (10 seconds) using Xantham gum at a maximum weight percent of 0.05 as the suspending and flocculating agent with gelatin in which are dispersed water insoluble drug particles. Mannitol is used as the preferred cryoprotectant. The suspension is freeze-dried in molds to generate the solid dosage form.

In US Patent 5,302,401 describes a method to reduce particle size growth during lyophilization. It discloses a composition containing particles having a surface modifier adsorbed onto the surface together with a cryoprotectant, the cryoprotectant present in an amount sufficient to form a nanoparticle-cryoprotectant composition. A preferred surface modifier is polyvinylpyrrolidone, and a preferred cryoprotectant is a carbohydrate such as sucrose. Also described are methods of making particles having a surface modifier adsorbed on to the surface and a cryoprotectant associated with it. The patent refers specifically to 5% Danazol with 1.5% PVP and sucrose (2%) or mannitol (2%) as the cryoprotectant. Thus while various cyroprotectants are available and function adequately to protect the active agent during lyophilization, the solid product that results is often difficult to redisperse in aqueous media.

EP 0193208 describes a method of lyophilizing reagent-coated latex particles to allow for reconstitution without aggregation and discusses the need to incorporate a zwitterionic buffer such as an amino acid, a stabilizer such as PVP or bovine albumin and a cryoprotectant such as Dextran T10 or other polysaccharide.

SUMMARY OF THE INVENTION

This invention is directed to an improvement in the dispersibility of micronized particles through the specific selection of excipients and methodology necessary to recover the primary particles. Inherent in this approach is the ability to produce stable aqueous suspensions of micron or submicron sized particles of water insoluble or poorly water-soluble compounds. These particles, which are required in the practice of the present invention, can be prepared according to the methods disclosed in U.S. Pat. No. 5,091,187 and 5,091,188 as well as WO

98/07414, whose disclosure is incorporated herein by reference. Briefly, water insoluble or poorly soluble compounds are dispersed in an aqueous medium in the presence of surface modifying agents or combinations of agents of which at least one is a phospholipid adsorbed on the surface thereof. Particle fragmentation occurs when the aforementioned suspension is subjected to stress as a result of processing with the use of various methods known in the art including, but not limited to, sonication, milling, homogenization, microfluidization, and antisolvent and solvent precipitation. The particle so produced is referred to as a microparticle which is defined herein as a solid particle of irregular, non-spherical or spherical shape having a nominal diameter of from nanometers to micrometers on to which is adsorbed a least one surface modifying agent of which one is a phospholipid.

According to this invention the microparticle suspension so produced is further admixed with surface modifying agent(s) and/or matrix-forming agent(s) which are present in an amount sufficient to allow freeze-drying and subsequent release of the surface coated drug particles upon contact with an aqueous environment. The selection of these components serves to minimize the tendency of microparticles to aggregate upon drying. Such aggregates are extremely difficult to redisperse due to the very high particle surface area which facilitates the degree of contact available to interacting particles resulting in irreversible lattices.

Small particle sizes of drugs are often needed in drug formulation development in order to maximize surface area, bioavailability, and dissolution requirements. The introduction of a suitable matrix-forming agent(s) in the above noted process serves to stabilize the phospholipid coated drug particle during the freeze-drying process and in the resulting freeze-dried product by suppressing any tendency of particle agglomeration or particle growth.

DESCRIPTION OF THE INVENTION

The present invention provides a rapidly disintegrating solid dosage form for water insoluble compounds, which releases primary particles stabilized with one or more surface modifiers, including but not limited to phospholipids. Examples of some preferred water-insoluble drugs include antifungal agents, immunosuppressive and immunoactive agents, antiviral agents, antineoplastic agents, analgesic and anti-inflammatory agents, antibiotics, antiepileptics, anesthetics, hypnotics, sedatives, antipsychotic agents, neuroleptic agents,

antidepressants, anxiolytics, anticonvulsant agents, antagonists, neuron blocking agents, anticholinergic and cholinomimetic agents, antimuscarinic and muscarinic agents, antiadrenergic and, antarrhythmics, antihypertensive agents, hormones, and nutrients. A detailed description of these drugs may be found in Remington's Pharmaceutical Sciences, 18th Edition, 1990, Mack Publishing Co., PA. The concentration of the water insoluble ingredient in the aqueous suspension can vary between 0.1% w/w and 60% w/w, preferably between 5% w/w and 30% w/w.

The water insoluble compound is first prepared as an aqueous suspension in the presence of one or more surface stabilizing agents, of which at least one is a phospholipid. The phospholipid may be any natural or synthetic phospholipid, including but not limited to, phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinoistol, phosphatidylglycerol, phosphatidic acid, lysophospholipids, egg or soybean phospholipid or a combination thereof. The phospholipid may be salted or desalted, hydrogenated or partially hydrogenated or natural, semisynthetic or synthetic. The concentration of the phospholipid ingredient in the aqueous suspension can vary between 0.1% w/w and 90% w/w, preferably between 0.5% w/w and 50% w/w and more preferably between 1% w/w and 20% w/w.

Examples of some suitable second and additional surface modifiers include: (a) natural surfactants such as casein, gelatin, natural phospholipids, tragacanth, waxes, enteric resins, paraffin, acacia, gelatin, and cholesterol, (b) nonionic surfactants such as polyoxyethylene fatty alcohol ethers, sorbitan fatty acid esters, polyoxyethylene fatty acid esters, sorbitan esters, glycerol monostearate, polyethylene glycols, cetyl alcohol, cetostearyl alcohol, stearyl alcohol, poloxamers, polaxamines, methylcellulose, hydroxycellulose, hydroxy propylcellulose, hydroxy propylmethylcellulose, noncrystalline cellulose, and synthetic phospholipids, (c) anionic surfactants such as potassium laurate, triethanolamine stearate, sodium lauryl sulfate, alkyl polyoxyethylene sulfates, sodium alginate, dioctyl sodium sulfosuccinate, negatively charged phospholipids (phosphatidyl glycerol, phosphatidyl inositol, phosphatidylserine, phosphatidic acid and their salts), and negatively charged glyceryl esters, sodium carboxymethylcellulose, and calcium carboxymethylcellulose, (d) cationic surfactants such as quaternary ammonium compounds, benzalkonium chloride, cetyltrimethylammonium bromide, and lauryldimethylbenzyl-ammonium chloride, (e) colloidal clays such as bentonite and veegum. A detailed description of these surfactants may be found in Remington's

Pharmaceutical Sciences, 18th Edition, 1990, Mack Publishing Co., PA; and Theory and Practice of Industrial Pharmacy, Lachman et al., 1986. The concentration of additional surfactants in the aqueous suspension can vary between 0.1% w/w and 90% w/w, preferably between 0.5% w/w and 50% w/w and more preferably between 1% w/w and 20% w/w. These surfactants may be either added initially during compounding or added post processing prior to freeze-drying or a combination of both depending on the nature, concentration and number of the surfactant(s).

The resulting coarse dispersion is primarily intended to distribute the surfactant(s) throughout the aqueous medium using traditional mixing methods involving shear, extrusion, agitation and/or cavitation. The coarse dispersion is referred to as a pre-mix for purposes of this disclosure.

The premix is then subjected to a process which facilitates particle fragmentation including but not limited to sonication, milling, homogenization, microfluidization, and antisolvent and solvent precipitation. The attrition time may vary and is dependent on the physicochemical characteristics of the medicament, the physicochemical characteristics of the surfactant(s) and the selected attrition process. As an example, high pressure homogenization processes can be employed as typified by the use of equipment such as APV Gaulin E15, Avestin C50 or MFIC Microfluidizer M110EH. In this process, the particles in the premix are reduced in size at a pressure and temperature which does not significantly compromise the stability of the medicament and/or the surfactant(s). Processing pressures of about 2000 psi to 30,000 psi, preferably of about 5,000 psi to 20,000 psi, more preferably of about 10,000 psi to 18,000 psi and operating temperatures of about 2°C to 65° C, more preferably 10°C to 45° C are suitable. The processing fluid is cycled through the homogenization chamber in such a manner as to ensure the entire fluid admixture is subjected to discrete homogenization resulting in a homogeneous suspension of micron or submicron particles. The mean volume weighted particle size of the resulting suspended therapeutic agent is measured to be between 0.05 micrometers to 10 micrometers, preferably between 0.2 micrometers to 5 micrometers using a laser light diffraction based instrument, Malvern Mastersizer Microplus.

The resulting homogeneous suspension of microparticles stabilized by one or more surface modifiers is then mixed with matrix-forming bulking and/or releasing agents (dry or as

an aqueous solution) and is then dried. The bulking or matrix-forming agent provides a mass in which the particles of drug are embedded or retain. The release agent assists in disintegration of the matrix when it contacts aqueous media. The bulking/releasing agents are chosen in order to produce a support matrix that, upon drying, will yield rapidly dispersible tablets that release the primary particles upon reconstitution in an aqueous medium. Examples of matrix-forming/release agents include (a) saccharides and polysaccharides such as mannitol, trehalose, lactose, sucrose, sorbitol, maltose; (b) humectants such as glycerol, propylene glycol, polyethylene glycol; (c) natural or synthetic polymers such as gelatin, dextran, starches, polyvinylpyrrolidone, poloxamers, acrylates; (d) inorganic additives such as colloidal silica, tribasic calcium phosphate and; (e) cellulose based polymers such as microcrystalline cellulose, hydroxymethyl cellulose, hydroxypropyl cellulose, methylcelluloses. Matrix forming agents may be added prior to producing the micronized particles of the therapeutic agent (formulation) or to the homogeneous suspension of microparticles prior to freeze-drying. The concentration of the matrix forming agents in the aqueous suspension can vary between 0.1% w/w and 90% w/w, preferably between 0.5% w/w and 50% w/w and more preferably between 1% w/w and 20% w/w.

The prepared aqueous suspension can be dried using several methods well known in the art. Spray drying, spray coating and freeze-drying are among the most common methods. The examples cited in Table 1 all use freeze drying as the drying method but this is not intended to be in any way limiting. The preferred method of freeze-drying is by lyophilization involving the sublimation of the frozen water from the aqueous suspension medium under reduced pressure. Lyophilization of this suspension may be performed in suitable containers such as glass vials, open trays, unit dosage form molds or *in-situ* spraying onto a supporting matrix. By way of example of the lyophilization process, the prepared suspension of microparticles containing matrix forming agents is distributed into stainless steel trays which are placed onto pre-equilibrated shelves held at a temperature of 5°C within a pressure rated, sealed chamber. The prepared suspension is then subjected to decreasing temperature at a rate of 5°C/min to –50°C until all of the suspension medium is completely solidified. This procedure uses only moderate temperature gradients because of the energy losses between different boundaries (shelf-tray-liquid). As a general rule, the typical time for freezing a 1 cm layer of a dilute aqueous suspension is 40-90 min at a temperature of -50°C. Freezing outside of the

lyophilization chamber may also be accomplished by: (a) freezing on cooled plates, e.g., in trays or in the form of small particles on a drum cooler, (b) dropping in liquid nitrogen or some other cooling liquid, (c) co-spraying with liquid CO_2 or liquid nitrogen, or (d) freezing with circulating cold air.

Separate cooling is necessary for the performance of continuous freeze-drying. Equipment producing small pellets by dropping the solution into liquid nitrogen is commercially available as the Cryopel® process (Buchmuller and Weyermanns, 1990). Direct freezing inside the lyophilization chamber is advantageous if the product requires handling under aseptic conditions as may be the situation in the preparation of injectable dried formulations.

The so-obtained solidified prepared suspension is held at this temperature for a period of 2 hours to ensure all crystallization has been completed. The pressure inside the chamber is reduced to a pressure of approximately 5 mm of Hg and preferably to about 0.1 mm Hg. The sublimation of the frozen water is accomplished by raising the shelf temperature of the lyophilizer to about –30°C to –10 °C and holding the material at this temperature for about 20 hours until the primary drying stage is completed. The drying time depends on a number of factors, some of them fairly constant and can be approximated as the heat of sublimation of ice, thermal conductivity of the frozen suspension and, the mass transfer coefficient. Other factors such as temperature or pressure in the chamber may vary considerably. The temperature of the shelves may be further increased to effect secondary drying as deemed necessary according to the composition of the sample.

Material is harvested from the lyophilizing cycle upon returning the chamber to ambient conditions. The harvested dried material may be passed through a coarse milling operation to facilitate handling or further blending operations with other excipients necessary to complete the required solid dosage form. These may include tableting aids for compression, glidants for hard gelatin encapsulation or dispersants for dry powder inhalers.

The matrix-forming agent used in the present invention must dissolve or disperse upon contact with an aqueous environment and release the phospholipid coated therapeutic agent particle. Upon reconstitution, the product reverts to a suspension having the same degree of

dispersity as the pre-dried suspension, with preferably no more than 20% by weight and more preferably no more than 10% by weight and ideally less than 1% by weight of aggregated primary particles as revealed by the particle sizing and microscopic methods known in the art. Surprisingly, the freeze-dried suspension prepared according to the present invention can be stored for extended periods of time, even at high temperature and humidity. without loss of this redispersibility characteristic upon reconstitution and thus is essentially devoid of particle aggregation. Freeze-dried suspensions prepared in accordance with the composition of Examples 6-10 herein can be stored for at least 60 days at room temperature indicating the possibility of long term storage consistent with pharmaceutical dosage form shelf life.

Solid dosage material prepared according to the present invention is defined as possessing the characteristic of being rapidly dispersible. This characteristic is identified as the time required for the complete disintegration of a freeze-dried cake arising from this invention when subjected to an aqueous medium as occurs upon administration of the dosage form to *in-vivo* systems. Disintegration time can be measured by carrying out an *in-vitro* test such as observing the disintegration time in water at 37 °C. The dosage material is immersed in the water without forcible agitation whereupon the time required for the material to substantially disperse by observation is noted. In the context of the definition of "rapid", the disintegration time is expected to be less than 2 minutes and preferably less than 30 seconds and most preferably less than 10 seconds.

The rate of dissolution or release of the active ingredient may also be affected by the nature of the medicament and the microparticle composition such that it may be rapid (5-60 sec) or intermediate (on the order of 75% disintegration in 15 minutes) or sustained-released.

In some cases, visual microscopic observation or scanning electron micrographs may reveal the presence of aggregates of particles however these particles are small in size and consist of aggregates of the original pre-freeze dried suspension particles. These aggregates are easily dispersed by low levels of energy such as short periods of sonication or physical agitation and as such display the key feature of this invention i.e. the prevention of particle size growth and irreversible aggregation and/or agglomeration.

EXAMPLES

The present invention of a rapidly dispersing, solid medicament is illustrated by way of the examples summarized in Table 1. Compositions noted in this table are expressed on % weight basis of the dried product. It is understood that the bulking agent may be added to the suspension prior to the homogenization step or prior to the drying step.

Table 1. Composition (% w/w) and Attributes of Solid Dosage Form Examples

Formulation Number	Phospholipids		Additional Surfactants				Active Ingredient			Bulking Agent				Attributes		
	E80	P100H	Myrj 52	PVP 17	NaDeox	Tween 80	CyA	ITR	FEN	Type	Qty	Type	Qty	Disintegration Time (sec)	Particle Size pre-Lyo (micron)	Particle Size post-Lyo (micron)
1	-	-	-	-	-	-	33	-	-	LAC	67	-	-	5	10.6	13.3
2	-	-	-	-	-	-	62.5	-	-	PVP17	37.5	-	-	5	10.2	17.4
3	-	-	4.6	-	1.1	-	23	-	-	MAN	5.5	LAC	46	60	0.66	48.9
4	23.1	-	-	-	-	-	-	-	76.9	-	-	-	-	>2 min.	0.91	85.50
5	-	5.6	-	-	-	5.6	-	-	27.8	MAN	61.0	-	-	10	0.97	6.73
6	9.1	-	-	-	-	-	-	-	33.3	SUC	45.5	SOR	15.1	5	0.97	0.98
7	11.1	-	-	-	-	-	-	27.8	-	TRE	33.3	LAC	27.8	5	1.15	1.15
8	15.4	-	-	-	-	-	-	38.4	-	TRE	46.2	-	-	5	1.15	1.12
9	-	8.4	4.2	-	1.0	-	21.1	-	-	MAN	23.2	LAC	42.1	15	0.92	1.33
10	-	11.9	6.0	17.9	1.5	-	29.9	-	-	MAN	32.8	-	-	5	0.91	1.08

Symbols and Note:

CyA=Cyclosporine; E80=Lipoid E80; FEN=Fenofibrate; ITR=Itraconazole; MAN=Mannitol; NaDeox=Sodium deoxycholate; P100H=Phospholipon 100H; PVP17=Polyvinyl pyrrolidone; SOR=Sorbitol; SUC=Sucrose; TRE=Trehalose

Formulations 1 and 2 as show in the above table illustrate that reconstitutable particulates are obtained from these compositions, indicating that the relatively large size of the particulates (approximately 10 micrometers) poses little problem from an aggregation perspective. These relatively large particulates are easily achieved by traditional particle fracturing techniques. However, in order to appreciably affect bioavailability, particles which are an order of magnitude less in size are required. These particulates are obtained using procedures described in US Pat. No. 5,091,187 and 5,091,188 as Microcrystals, WO 98/07414 as microparticles, and US 5,145,684, US 5,302,401 and US 5,145,684 as nanocrystals. It is the particulates arising from these compositions that require specific excipient selection and processing conditions in order to recover the original suspension particle. Examples 3 to 5 illustrate that certain microparticle compositions do not reconstitute favorably when traditional freeze drying cryoprotectants such as lactose or PVP17 are used as described in US pat 5,302,401. For these examples, large aggregates are formed comprised of adhering primary particles.

Examples 6 to 10 illustrate that the original suspension particle is easily and rapidly recovered upon reconstitution of the dried powder requiring no excessive agitation. These examples require careful selection of the bulking agent which may also act as a cryoprotectant as well as a humectant, such as, trehalose in formulation 8 and mannitol in formulation 10.

Alternatively, when a single matrix forming bulking agent is not suitable, as in the case of sucrose, the composition may include a mixture of bulking agents selected from pharmaceutically acceptable agents such as sucrose, trehalose, mannitol, sorbitol, or lactose. Example formulations 6, 7, and 9 demonstrate this type of composition. Volume weighted particle size distribution profiles of fenofibrate formulation 6 are shown in Figures 6 and 7, respectively, before and after the lyophilization/reconstitution step. This example demonstrates the ideal scenario of no change in the particle size distribution profile following lyophilization and reconstitution.

With no intention to propose any particular theoretical explanation, it may be speculated that the components of the bulking agent mixture may simultaneously serve to

inhibit the particle size increase on lyophilization/reconstitution by one or more mechanisms including cryoprotection, humectant action, dispersibility, and others.

These criteria are surprisingly important considerations when attempting to recover the unaggregated particulate suspension following reconstitution of a dried dosage form that comprises a phospholipid as one of the surface stabilizers.

In addition to the example compositions mentioned above, the formulations of this invention may additionally contain suitable amounts of pH buffering salts and pH adjusting agents such as sodium hydroxide and/or pharmaceutically acceptable acids. It is known to those skilled in the chemistry of phospholipids that at pH lower than 4 and higher than 10 the phospholipid molecules undergo extensive hydrolysis. Therefore, the pH of the suspension is usually adjusted to within this range prior to homogenization. If necessary the pH can be readjusted prior to the drying step.

While the invention and the examples have been described in connection with what is presently considered to be the most practical and preferred embodiment, it is to be understood that the invention is not to be limited to the disclosed embodiments, but on the contrary, is intended to cover various modifications and equivalent arrangements included within the spirit and scope of the following claims.

CLAIMS

We claim:

1. A rapidly dispersing solid therapeutic dosage form comprised of a water insoluble compound existing as a nanometer or micrometer particulate solid which is surface stabilized with one or more surface modifiers of which at least one may be a phospholipid, the particulate solid dispersed throughout a bulking matrix optionally also including a releasing agent forming a therapeutic dosage form when dried which when the dosage form is introduced into an aqueous environment the bulking/releasing matrix is substantially completely disintegrated within less than 2 minutes thereby releasing the water insoluble particulate solid in an unaggregated and/or unagglomerated state.

2. The rapidly dispersing solid dosage form of claim 1 wherein the water insoluble particulate solid component consisting essentially of a composition of a water insoluble substance comprising particles of a therapeutically useful water insoluble or poorly water soluble compound, a phospholipid and optionally also at least one non-ionic, anionic, cationic or amphipathic surfactant, wherein a volume weighted mean particle size of the water insoluble particle is 5 micrometers or less.

3. The rapidly dispersing solid dosage form of claim 1 wherein the bulking/releasing matrix component is selected from saccharides, polysaccharides, humectants, natural or synthetic polymers inorganic additives, or cellulose based polymers.

4. The rapidly dispersing solid dosage form of claim 3 wherein the polyof, saccharide or polysaccharide is mannitol, trehalose, lactose, sucrose, sorbitol, dentrose, mulodextrose or maltose.

5. The rapidly dispersing solid dosage form of claim 3 wherein the humectant is glycerol, propylene glycol or polyethylene glycol.

6. The rapidly dispersing solid dosage form of claim 3 wherein the natural or synthetic polymer is gelatin, dextran, starches, polyvinylpyrrolidone, a poloxamer or an acrylate.

7. The rapidly dispersing solid dosage form of claim 3 wherein the inorganic additive

is colloidal silica or tribasic calcium phosphate.

8. The rapidly dispersing solid dosage form of claim 3 wherein the cellulose based polymer is microcrystalline cellulose, hydroxymethyl cellulose, hydroxypropyl cellulose or methylcellulose.

9. The rapidly dispersing solid dosage form of claim 1 wherein the disintegration time in an aqueous medium is less than 2 minutes and preferably less than 60 seconds, more preferably less than 30 seconds, and the most preferably less than 10 seconds.

10. The rapidly dispersing solid dosage form of claim 1 further containing an effervescent agent, a binding agent, a flavor, a polymeric coating on the external surface of the dosage form, a color or combinations thereof.

INTERNATIONAL SEARCH REPORT

Inter onal Application No
PCT/US 99/27436

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 A61K9/14 A61K9/19

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98 07414 A (RES TRIANGLE PHARM LTD) 26 February 1998 (1998-02-26) cited in the application abstract page 1 page 4, line 1 -page 7, line 2 page 8 -page 11; example 1 page 14; examples 4.4,4.5 page 18; examples 5.5,5.6 claims 2,5,6,8-13,16,17	1-6,8-10
E	WO 99 61001 A (RTP PHARMA INC) 2 December 1999 (1999-12-02) page 2, line 14 -page 5, line 3 page 5; example 1 page 12; example 7 claims 1,2,6-8,11,13,15,18	1-4

☐ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

° Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

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"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
14 March 2000	21/03/2000

Name and mailing address of the ISA	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL – 2280 HV Rijswijk Tel. (+31–70) 340–2040, Tx. 31 651 epo nl, Fax: (+31–70) 340–3016	Muller, S

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INTERNATIONAL SEARCH REPORT

Information on patent family members

Inter onal Application No
PCT/US 99/27436

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9807414 A	26-02-1998	AU 2587197 A	06-03-1998
		CN 1228021 A	08-09-1999
		CZ 9900596 A	16-06-1999
		EP 0925061 A	30-06-1999
		NO 990790 A	19-04-1999
		PL 331715 A	02-08-1999
		US 5922355 A	13-07-1999
WO 9961001 A	02-12-1999	NONE	