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Description**FIELD OF THE INVENTION**

5 **[0001]** The present invention generally relates to bacteriologically pure bacterial cultures of novel strains of plant growth-promoting bacteria, and inoculums comprising the same. The invention is also directed to plant seeds coated with the inoculums, kits comprising the inoculums and methods for stimulating plant growth by using the claimed bacterial cultures and/or inoculums.

10 **BACKGROUND OF THE INVENTION**

[0002] Plant growth-promoting bacteria (PGPB) are associated with many, if not all, plant species and are commonly present in many environments. The most widely studied group of PGPB is plant growth-promoting rhizobacteria (PGPR), which colonize the root surfaces and the closely adhering soil interface, the rhizosphere. Inside the rhizosphere is a zone where bacteria, fungi, and other organisms compete for nutrients and for binding to the root structures of the plant. Both detrimental and beneficial bacteria can occupy the plant's roots. The presence of plant growth-promoting bacteria (PGPB) within or near the roots or seeds can lead to a healthier rhizosphere environment and healthier plants. These free living bacteria promote plant growth in agricultural crops and lead to increased growth and yield at harvest. The bacteria that colonize the roots and maintain their benefits throughout the growth cycle of the plant are especially desired for application during early growth or as a seed coating agent to agricultural crops.

[0003] The mechanisms that PGPBs use in promoting plant growth are diverse and often plant-or cultivar-specific. Several PGPB growth-promoting mechanisms are known, which can influence the plant in a direct or indirect manner. The direct mechanism involves increasing plant growth by supplying the plant with nutrients and hormones, such as by fixing nitrogen that is available to plants, synthesizing phytohormones, and providing nutrients such as phosphate to the plant. The indirect mechanism of action for PGPB occurs through the ability to control detrimental fungal and bacterial pathogens from establishing or surviving within the rhizosphere. This is usually achieved through the beneficial secretion of antifungals and other antibiotics by the PGPB. As an additional advantage, PGPBs can also lead to extensive remodeling of the plant root systems.

[0004] In recent years, a significant effort has been expanded to identify novel strains of plant growth-promoting bacteria, and use them to promote plant growth, thereby increasing the yield of plant product, reducing the use and amounts of fertilizers and herbicides, and providing other benefits for agricultural and horticultural communities.

[0005] López-Bucio et al 2007 Mol Plant Microbe Interact discloses that *Bacillus megaterium* rhizobacteria promotes growth and alters root-system architecture through an auxin- and ethylene-independent signaling mechanism in *Arabidopsis thaliana*.

35 **[0006]** Raddadi et al 2008 Annals of Microbiology discusses the screening of plant growth promoting traits of *Bacillus thuringiensis*.

[0007] Freitas et al 1997 Biology and Fertility of Soils discloses that phosphate-solubilizing rhizobacteria enhance the growth and yield but not phosphorus uptake of canola (*Brassica napus* L.).

40 **[0008]** Selvakumar et al 2008 Curr Microbiol discusses the isolation and characterization of nonrhizobial plant growth promoting bacteria from nodules of Kudzu (*Pueraria thunbergiana*) and their effect on wheat seedling growth.

SUMMARY OF THE INVENTION

45 **[0009]** The invention is as defined by the claims and provides:

an inoculum for application to plants, plant seeds, or a plant growth medium, wherein the inoculum comprises an agriculturally acceptable carrier and an effective amount of a biologically pure bacterial culture, wherein the bacteria in the bacterial culture is *Bacillus thuringiensis* strain BT013A (NRRL No. B-50924);

50 a method for stimulating plant growth comprising applying the inoculum of the invention to a plant, plant seed, or plant growth medium;

a plant seed coated with the inoculum of the invention; and

55 a kit for stimulating plant growth comprising an inoculum of the invention and instructions for applying the inoculum to plants, plant seeds, or a plant growth medium.

[0010] Also disclosed is a biologically pure bacterial culture wherein the bacteria in the bacterial culture is: (a) *Bacillus*

aryabhatai strain CAP53 (NRRL No. B-50819); (b) *Bacillus aryabhatai* strain CAP56 (NRRL No. B-50817); (c) *Bacillus flexus* strain BT054 (NRRL No. B-50816); (d) *Paracoccus kondralievae* strain NC35 (NRRL No. B-50820); (e) *Bacillus mycoides* strain BT155 (NRRL No. B-50921); (f) *Enterobacter cloacae* strain CAP12 (NRRL No. B-50822); (g) *Bacillus nealsonii* strain BOBA57 (NRRL No. B-50821); (h) *Bacillus mycoides* strain EE118 (NRRL No. B-50918); (i) *Bacillus subtilis* strain EE148 (NRRL No. B-50927); (j) *Alcaligenes faecalis* strain EE107 (NRRL No. B-50920); (k) *Bacillus mycoides* strain EE141 (NRRL No. B-50916); (l) *Bacillus mycoides* strain BT46-3 (NRRL No. B-50922); (m) *Bacillus cereus* family member strain EE128 (NRRL No. B-50917); (n) *Bacillus thuringiensis* strain BT013A (NRRL No. B-50924); (o) *Paenibacillus massiliensis* strain BT23 (NRRL No. B-50923); (p) *Bacillus cereus* family member strain EE349 (NRRL No. B-50928); (q) *Bacillus subtilis* strain EE218 (NRRL No. B-50926); (r) *Bacillus megaterium* strain EE281 (NRRL No. B-50925); (s) salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; (t) salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; (u) thiram-resistant *Bacillus aryabhatai* CAP53; (v) thiram-resistant *Bacillus thuringiensis* BT013A; or (w) glyphosate-tolerant *Bacillus aryabhatai* CAP53.

[0011] Also disclosed are biologically pure bacterial cultures wherein bacteria in the bacterial cultures are mutants of any of the foregoing strains comprising one or more mutations which retain the ability to promote plant growth.

[0012] The present disclosure is also directed to an inoculum for application to plants, plant seeds, or a plant growth medium, wherein the inoculum comprises an effective amount of a biologically pure bacterial culture disclosed herein and an agriculturally acceptable carrier. In the invention, the biologically pure bacterial culture is a culture of *Bacillus thuringiensis* strain BT013A (NRRL No. B-50924).

[0013] Also disclosed is a method for stimulating plant growth by applying the biologically pure bacterial culture or the inoculum as disclosed herein to a plant, plant seed, or plant growth medium.

[0014] Also disclosed is a method for stimulating plant growth by applying glycerol, pyruvate, yeast extract, a polyol (e.g., mannitol, sorbitol, galactitol, fucitol, iditol, inositol, arabitol, xylitol, ribitol), polyethylene glycol, or a combination thereof to a plant growth medium, and applying at least one bacterial culture or at least one inoculum to a plant or plant seed in the plant growth medium, or to the plant growth medium, wherein the at least one bacterial culture or at least one inoculum is capable of stimulating plant growth.

[0015] Also disclosed is a plant seed coated with the inoculum or with the bacterial culture disclosed herein.

[0016] Yet another aspect of the present invention is a kit for stimulating plant growth comprising an inoculum disclosed herein and instructions for applying the inoculum to plants, plant seeds, or a plant growth medium.

[0017] Other objects and features will be in part apparent and in part pointed out hereinafter.

DEFINITIONS

[0018] A "biologically pure bacterial culture" refers to a culture of bacteria containing no other bacterial species in quantities sufficient to interfere with the replication of the culture or be detected by normal bacteriological techniques. Stated another way, it is a culture wherein virtually all of the bacterial cells present are of the selected strain.

[0019] The term "rhizosphere" is used interchangeably with "root zone" to denote that segment of the soil that surrounds the roots of a plant and is influenced by them.

[0020] The term "inoculant" as described in this invention is defined in several Federal, or State regulations as (1) "soil or plant inoculants shall include any carrier or culture of a specific micro-organism or mixture of micro-organisms represented to improve the soil or the growth, quality, or yield of plants, and shall also include any seed or fertilizer represented to be inoculated with such a culture" (New York State 10-A Consolidated Law); (2) "substances other than fertilizers, manufactured, sold or represented for use in the improvement of the physical condition of the soil or to aid plant growth or crop yields" (Canada Fertilizers Act); (3) "a formulation containing pure or predetermined mixtures of living bacteria, fungi or virus particles for the treatment of seed, seedlings or other plant propagation material for the purpose of enhancing the growth capabilities or disease resistance or otherwise altering the properties of the eventual plants or crop" (Ad hoc European Working Group, 1997) or (4) "meaning any chemical or biological substance of mixture of substances or device distributed in this state to be applied to soil, plants or seeds for soil corrective purposes; or which is intended to improve germination, growth, quality, yield, product quality, reproduction, flavor, or other desirable characteristics of plants or which is intended to produce any chemical, biochemical, biological or physical change in soil" (Section 14513 of the California Food and Agriculture Code).

[0021] The term "effective amount" refers to a quantity which is sufficient to result in a statistically significant increase of growth and/or of protein yield and/or of grain yield of a plant as compared to the growth, protein yield and grain yield of the control-treated plant.

[0022] The terms "agriculturally acceptable carrier" and "carrier" are used interchangeably herein.

[0023] The terms "promoting plant growth" and "stimulating plant growth" are used interchangeably herein, and refer to the ability to enhance or increase at least one of the plant's height, weight, leaf size, root size, or stem size, to increase protein yield from the plant or to increase grain yield of the plant.

DETAILED DESCRIPTION

[0024] Disclosed herein are biologically pure bacterial cultures of plant growth-promoting bacteria (PGPB) wherein the bacteria, i.e. the bacterial strain in each of the bacterial cultures, are selected from the group consisting of (a) *Bacillus aryabhattai* strain CAP53 (NRRL No. B-50819), (b) *Bacillus aryabhattai* strain CAP56 (NRRL No. B-50817), (c) *Bacillus flexus* strain BT054 (NRRL No. B-50816), (d) *Paracoccus kondratievae* strain NC35 (NRRL No. B-50820), (e) *Bacillus mycoides* strain BT155 (NRRL No. B-50921), (f) *Enterobacter cloacae* strain CAP12 (NRRL No. B-50822), (g) *Bacillus nealsonii* strain BOBA57 (NRRL No. B-50821), (h) *Bacillus mycoides* strain EE118 (NRRL No. B-50918), (i) *Bacillus subtilis* strain EE148 (NRRL No. B-50927), (j) *Alcaligenes faecalis* strain EE107 (NRRL No. B-50920), (k) *Bacillus mycoides* strain EE141 (NRRL No. B-50916), (l) *Bacillus mycoides* strain BT46-3 (NRRL No. B-50922), (m) *Bacillus cereus* family member strain EE128 (NRRL No. B-50917), (n) *Bacillus thuringiensis* strain BT013A (NRRL No. B-50924), (o) *Paenibacillus massiliensis* strain BT23 (NRRL No. B-50923), (p) *Bacillus cereus* family member strain EE349 (NRRL No. B-50928), (q) *Bacillus subtilis* strain EE218 (NRRL No. B-50926), (r) *Bacillus megaterium* strain EE281 (NRRL No. B-50925), (s) salt-tolerant and thiram-resistant *Paracoccus sp.* NC35 (NRRL No. B-50948), (t) salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155 (NRRL No. B-50949), (u) thiram-resistant *Bacillus aryabhattai* CAP53 (NRRL No. B-50946), (v) thiram-resistant *Bacillus thuringiensis* BT013A (NRRL No. B-50947), or (w) glyphosate-tolerant *Bacillus aryabhattai* CAP53 (NRRL No. B-50945).

[0025] The foregoing strains (a)-(d) and (f)-(g) were deposited with the United States Department of Agriculture (USDA) Agricultural Research Service (ARS), having the address 1815 North University Street, Peoria, Illinois 61604 U.S.A., on March 11, 2013, and are identified by the NRRL numbers provided in parentheses. The strains (e) and (h)-(r) were deposited with the United States Department of Agriculture (USDA) Agricultural Research Service (ARS) on March 10, 2014, and are also identified by the NRRL numbers provided in parentheses. The strains (s)-(w) were deposited with the United States Department of Agriculture (USDA) Agricultural Research Service (ARS) on March 17, 2014, and are also identified by the NRRL numbers provided in parentheses.

[0026] As shown in the Examples and Comparative Examples, the present strains were isolated from rhizospheres of various vigorous plants, and were shown to be most promising among a large number of isolates by *in vitro* culturing and application to plants. The novel strains disclosed herein were identified by 16S RNA sequencing and biochemical assays. Thus, *Bacillus aryabhattai* strain CAP53 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 1; *Bacillus aryabhattai* strain CAP56 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 2; *Bacillus flexus* strain BT054 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 3; *Paracoccus kondratievae* strain NC35 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 4; *Bacillus mycoides* strain BT155 (NRRL No. B-50921) has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 21; *Enterobacter cloacae* strain CAP12 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 5; *Bacillus nealsonii* strain BOBA57 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 6; *Bacillus mycoides* strain EE118 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 10; *Bacillus subtilis* strain EE148 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 11; *Alcaligenes faecalis* strain EE107 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 12; *Bacillus mycoides* strain EE141 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 13; *Bacillus mycoides* strain BT46-3 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 14; *Bacillus cereus* family member strain EE128 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 15; *Bacillus thuringiensis* strain BT013A has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 16; *Paenibacillus massiliensis* strain BT23 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 17; *Bacillus cereus* family member strain EE349 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 18; *Bacillus subtilis* strain EE218 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 19; and *Bacillus megaterium* strain EE281 has a 16S ribosomal RNA sequence having at least about 98%, at least about 99%, or 100% sequence identity with the sequence of SEQ ID NO: 20. These sequences are shown in Table 1 below.

TABLE 1

Strain (SEQ ID. NO)	Partial 16S ribosomal RNA sequence
5 <i>Bacillus aryabhatai</i> CAP53 (SEQ ID NO: 1) 10 15 20	GGNNCAACGCCGCGTGAGTGATGAAGGCTTTCGGGGTCGTAAAACT CTGTTGTTAGGGAAGAACAAGTACGAGAGTAACTGCTCGTACCTT GACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGC CGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCG TAAAGCGCGCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCAC GGCTCAACCGTGGAGGGTCATTGGAAACTGGGGAACTTGAGTGCA GAAGAGAAAAGCGGAATTCCACGTGTAGCGGTGAAATGCGTAGAG ATGTGGAGGAACACCAGTGGCGAAGGCGGCTTTTTGGTCTGTAAC TGACGCTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATAC CCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGAGGG TTTCCGCCCTTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCTG GGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGGGC CCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAA GAACCTTACCAGGTCTTGACATCCTCTGACAACTCTAGAGATAGAG CGTTCCCCTTCGGGGGACAGAGTGACAGGTGGTGCATGGTTGTCGT CAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACG
25 <i>Bacillus aryabhatai</i> CAP56 (SEQ ID NO: 2) 30 35 40	TCTGANGGNNCACGCCGCGTGAGTGATGAAGGCTTTCGGGGTCGT AAACTCTGTTGTTAGGGAAGAACAAGTACGAGAGTAACTGCTCGT ACCTTGACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGCCA GCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATT GGGCGTAAAGCGCGCGCAGGCGGTTTCTTAAGTCTGATGTGAAAG CCCACGGCTCAACCGTGGAGGGTCATTGGAAACTGGGGAACTTGA GTGCAGAAGAGAAAAGCGGAATTCCACGTGTAGCGGTGAAATGCG TAGAGATGTGGAGGAACACCAGTGGCGAAGGCGGCTTTTTGGTCT GTAAGTACGCTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTA GATACCCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTA GAGGGTTTCCGCCCTTTAGTGCTGCAGCTAACGCATTAAGCACTCC GCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACG GGGGCCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAAC GCGAAGAACCTTACCAGGTCTTGACATCCTCTGACAACTCTAGAGA TAGAGCGTTCCCCTTCGGGGGACAGAGTGACAGGTGGTGCATGGT TGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGC
45 <i>Bacillus flexus</i> BT054 (SEQ ID NO: 3) 50 55	GGANCAACGCCGCGTGAGTGANGAAGGCTTTCGGGGTCGTAAAACT CTGTTGTTAGGGAAGAACAAGTACAAGAGTAACTGCTTGTACCTTG ACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGCC GCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCGT AAAGCGCGCGCAGGCGGTTTCTTAAGTCTGATGTGAAAGCCACG GCTCAACCGTGGAGGGTCATTGGAAACTGGGGAACTTGAGTGCA AAGAGAAAAGCGGAATTCCACGTGTAGCGGTGAAATGCGTAGAGA TGTGGAGGAACACCAGTGGCGAAGGCGGCTTTTTGGTCTGTAAC GACGCTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATAC CCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGAGGG TTTCCGCCCTTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCTG GGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGGGC CCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAA GAACCTTACCAGGTCTTGACATCCTCTGACAACTCTAGAGATAGAG CGTTCCCCTTCGGGGGACAGAGTGACAGGTGGTGCATGGTTGTCGT CAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAAC

(continued)

Strain (SEQ ID. NO)	Partial 16S ribosomal RNA sequence
5 <i>Paracoccus kondratievae</i> NC35 (SEQ ID NO: 4) 10 15 20	GCCGCGTGAGTGNNNAAGNCCCTAGGGTTGTAAAGCTCTTTCANC TGGGAAGATAATGACTGTACCAGCAGAAGAAGCCCCGGCTAACTC CGTGCCAGCAGCCGCGGTAATACGGAGGGGGGCTAGCGTTGTTTCGG AATTACTGGGCGTAAAGCGCACGTAGGCGGACCGGAAAGTTGGGG GTGAAATCCCCGGGGCTCAACCCCGGAAGTGCCTTCAAAACTATCG GTCTGGAGTTCGAGAGAGGTGAGTGGAATTCCGAGTGTAGAGGTG AAATTCGTAGATATTCGGAGGAACACCAGTGGCGAAGGCGGCTCA CTGGCTCGATACTGACGCTGAGGTGCGAAAGCGTGGGGGAGCAAAC AGGATTAGATAACCCTGGTAGTCCACGCCGTAAACGATGAATGCCA GTCGTCGGGCAGCATGCTGTTCGGTGACACACCTAACGGATTAAG CATTCCGCCTGGGGAGTACGGTCGCAAGATTAAAACTCAAAGGAA TTGACGGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAAATTCGA AGCAACGCGCAGAACCTTACCAACCCTTGACATCCCAGGACAGCC CGAGAGATCGGGTCTCCACTTCGGTGGCCTGGAGACAGGTGCTGC ATGGCTGTCGTCAGCTCGTGTCGTGAGATGTTTCGGTTAAGTCCGGC

(continued)

Strain (SEQ ID. NO)	Partial 16S ribosomal RNA sequence
5 <i>Bacillus mycoides</i> EE118 (SEQ ID NO: 10) 10 15 20	GGAGCACGCCGCGTGAGTGNNGAAGGCTTTCGGGTCGTAAAACCTC TGTTGTTAGGGAAGAACAAGTGCTAGTTGAATAAGCTGGCACCTT GACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGC CGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCG TAAAGCGCGCGCAGGTGGTCTTCTTAAGTCTGATGTGAAAGCCAC GGCTCAACCGTGGAGGGTCATTGGAAACTGGGAGACTTGAGTGCA GAAGAGGAAAGTGGAATTCCATGTGTAGCGGTGAAATGCGTAGAG ATATGGAGGAACACCAGTGGCGAAGGCGACTTTCTGGTCTGTAAAC TGACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATAC CCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGAGGG TTTCCGCCCTTTAGTGCTGAAGTTAACGCATTAAGCACTCCGCCTG GGGAGTACGGCCGCAAGGCTGAAACTCAAAGGAATTGACGGGGG CCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGA AGAACCTTACCAGGTCTTGACATCCTCTGAAAACCTCTAGAGATAGA GCTTCTCCTTCGGGAGCAGAGTGACAGGTGGTGCATGGTTGTCGTC AGCTCGTGTCGTGAGATGTTGGGTAAAGTCCCGC
25 <i>Bacillus subtilis</i> EE148 (SEQ ID NO: 11) 30 35	CGCCGCGTGAGTGATGAAGGTTTTTCGGATCGTAAAGCTCTGTTGTT AGGGAAGAACAAGTGCCGTTCAAATAGGGCGGCACCTTGACGGTA CCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGCCGCGGTA ATACGTAGGTGGCAAGCGTTGTCCGGAATTATTGGGCGTAAAGGG CTCGCAGGCGGTTTTCTTAAGTCTGATGTGAAAGCCCCCGGCTCAAC CGGGGAGGGTCATTGGAAACTGGGGAACCTTGAGTGCAGAAGAGG AGAGTGGAATTCCACGTGTAGCGGTGAAATGCGTAGAGATGTGGA GGAACACCAGTGGCGAAGGCGACTCTCTGGTCTGTAACTGACGCT GAGGAGCGAAAGCGTGGGGAGCGAACAGGATTAGATACCCTGGT AGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGGGGGGTTTCCG CCCCTTAGTGCTGCAGCTAACGCATTAAGCACTCCGCCTGGGGAGT ACGGTCGCAAGACTGAAACTCAAAGGAATTGACGGGGGGCCCGCAC AAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGAAGAACCT TACCAGGTCTTGACATCCTCTGACAATCCTAGAGATAGGACGTCCC CTTCGGGGGGCAGAGTGACAGGTGGTGCATGGTTGTCGTCAGCTCGT GTCGTGAGATGTTGGGTAAAGTCC
40 <i>Alcaligenes faecalis</i> EE107 (SEQ ID NO: 12) 45 50 55	CTTCGGGTTGTAAAGTACTTTTGGCAGAGAAGAAAAGGTATCTCCT AATACGAGATACTGCTGACGGTATCTGCAGAATAAGCACCGGCTA ACTACGTGCCANCAGCCGCGGTAATACGTAGGGTGCAAGCGTTAA TCGGAATTACTGGGCGTAAAGCGTGTGTAGGCGGTTCGGAAAGAA AGATGTGAAATCCCAGGGCTCAACCTTGGAACCTGCATTTTTTAAC CCGAGCTAGAGTATGTCAGAGGGGGGGTAGAATTCNNNTGTAGCAN NGAAATGCGTAGATATGTGGAGGAATACCGATGGCGAAGGCAGCC CCCTGGGATAATACTGACGCTCAGACACGAAAGCGTGGGGAGCAA ACAGGATTAGATACCCTGGTAGTCCACGCCCTAAACGATGTCAACT AGCTGTTGGGGCCGTTAGGCCTTAGTAGCGCAGCTAACGCGTGAA GTTGACCGCCTGGGGAGTACGGTCGCAAGATTAAACTCAAAGGA ATTGACGGGGACCCGCACAAGCGGTGGATGATGTGGATTAAATTCG ATGCAACGCGAAAAACCTTACCTACCCTTGACATGTCTGGAAAGC CGAAGAGATTTGGCCGTGCTCGCAAGAGAACCGGAACACAGGTGC TGCATGGCTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTAAAGTCC C

(continued)

Strain (SEQ ID. NO)	Partial 16S ribosomal RNA sequence
5 <i>Bacillus mycoides</i> EE141 (SEQ ID NO: 13) 10 15 20	AAAGTCTGACGGAGCACGCCGCGTGAGTGATGAAGGCTTTCGGGT CGTAAAACTCTGTTGTTAGGGAAGAACAAGTGCTAGTTGAATAAG CTGGCACCTTGACGGTACCTAACCAGAAAGCCACGGCTAACTACG TGCCAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAA TTATTGGGCGTAAAGCGCGCGCAGGTGGTTTTCTTAAGTCTGATGTG AAAGCCCACGGCTCAACCGTGGAGGGTCATTGGAAACTGGGAGAC TTGAGTGCAGAAGAGGAAAGTGGAATTCCATGTGTAGCGGTGAAA TGCGTAGAGATATGGAGGAACACCAGTGGCGAAGGCGACTTTCTG GTCTGTAACCTGACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGG ATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGT GTTAGAGGGTTTTCCGCCCTTTAGTGCTGAAGTTAACGCATTAAGCA CTCCGCCTGGGGAGTACGGCCGCAAGGCTGAAACTCAAAGGAAT TGACGGGGGGCCCGCACAAAGCGGTGGAGCATGTGGTTTAATTCGAA GCAACGCGAAGAACCTTACCAGGTCTTGACATCCTCTGAAAACNC TAGAGATANNNCTTCTCCTTCGGGAGCAGAGTGACAGGTGGTGCA TGGTTGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCC
25 <i>Bacillus mycoides</i> BT46-3 (SEQ ID NO: 14) 30 35	GGAGCACGCCGCGTGAGTGATGAAGGCTTTCGGGTCGTAAAACTC TGTTGTTAGGGAAGAACAAGTGCTAGTTGAATAAGCTGGCACCTT GACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGC CGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCG TAAAGCGCGCGCAGGTGGTTTTCTTAAGTCTGATGTGAAAGCCAC GGCTCAACCGTGGAGGGTCATTGGAAACTGGGAGACTTGAGTGCA GAAGAGGAAAGTGGAATTCCATGTGTAGCGGTGAAATGCGTAGAG ATATGGAGGAACACCAGTGGCGAAGGCGACTTTCTGGTCTGTAACT TGACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATAC CCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGAGGG TTTCCGCCCTTTAGTGCTGAAGTTAACGCATTAAGCACTCCGCCTG GGGAGTACGGCCGCAAGGCTGAAACTCAAAGGAATTGACGGGGG CCCGCACAAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGA AGAACCTTACCAGGTCTTGACATCCTCTGACAACCCTAGAGATAGG GCTTCCCCTTCGGGGGGCAGAGTGACAGGTGGTGTCATGGTTGTCGTC AGCTCGTGTCGTGAGATGTTGGGTTAAGTCCC
40 <i>Bacillus cereus</i> family member EE128 (SEQ ID NO: 15) 45 50 55	GGANCAACGCCGCGTGAGTGANGAAGGCTTTCGGGTCGTAAAACT CTGTTGTTAGGGAAGAACAAGTGCTAGTTGAATAAGCTGGCACCTT GACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGCCAGCAGC CGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTATTGGGCG TAAAGCGCGCGCAGGTGGTTTTCTTAAGTCTGATGTGAAAGCCAC GGCTCAACCGTGGAGGGTCATTGGAAACTGGGAGACTTGAGTGCA GAAGAGGAAAGTGGAATTCCATGTGTAGCGGTGAAATGCGTAGAG ATATGGAGGAACACCAGTGGCGAAGGCGACTTTCTGGTCTGTAACT TGACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATTAGATAC CCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTTAGAGGG TTTCCGCCCTTTAGTGCTGAAGTTAACGCATTAAGCACTCCGCCTG GGGAGTACGGCCGCAAGGCTGAAACTCAAAGGAATTGACGGGGG CCCGCACAAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAACGCGA AGAACCTTACCAGGTCTTGACATCCTCTGAAAACCTCTAGAGATAGA GCTTCTCCTTCGGGAGCAGAGTGACAGGTGGTGTCATGGTTGTCGTC AGCTCGTGTCGTGAGATGNTGGGTTAAGTCCCGCA

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Strain (SEQ ID. NO)	Partial 16S ribosomal RNA sequence
<i>Bacillus thuringiensis</i> BT013A (SEQ ID NO: 16)	TCTGACGGAGCAACGCCGCGTGAGTGATGAAGGCTTTCGGGTCGT AAAACCTCTGTTGTTAGGGAAGAACAAGTGCTAGTTGAATAAGCTG GCACCTTGACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGC CAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTA TTGGGCGTAAAGCGCGCGCAGGTGGTTCCTTAAGTCTGATGTGAAA GCCCACGGCTCAACCGTGGAGGGTCATTGGAAACTGGGAGACTTG AGTGCAGAAGAGGAAAGTGGAATTCCATGTGTAGCGGTGAAATGC GTAGAGATATGGAGGAACACCAGTGGCGAAGGCGACTTTCTGGTC TGTAACCTGACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATT AGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTT AGAGGGTTTCCGCCCTTTAGTGCTGAAGTTAACGCATTAAGCACTC CGCCTGGGGAGTACGGCCGCAAGGCTGAAACTCAAAGGAATTGAC GGGGGCCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAA CGCGAAGAACCTTACCAGGTCTTGACATCCTCTGAAAACCCTAGA GATAGGGCTTCTCCTTCGGGAGCAGAGTGACAGGTGGTGCATGGT TGTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCGCAACGA GCGCAACCCTTGATCTTAGTTGCCATCATTAAAGTTGGGCACTCTAA GGTGACTGCCGGTGACAAACCGGAGGAAGGTGGGGATGACGTCAA ATCATCATGCCCCTTATGACCTGGGCTACACACGTGCTACAATGGA CGGTACAAAGAGCTGC
<i>Paenibacillus massiliensis</i> BT23 (SEQ ID NO: 17)	CTTANNGNNTGANNNNNCTTGNNNAANAAAGCCCCGGCTAACTACN TGCCANCANCCGCGGTAATACNTANGGNGCAAGCGTTGTCCGGAA TTATTGGGCGTAAAGCGCGCGCAGGCGGTCNTTTAAGTCTGGTGTT TAAGCCCCGGGGCTCAACCCCCGGATCNCNCGGGAAACTGGATGACT TGANTGCNNAANAAGAGAGTGGAATTCCNNGTGTANCGGTGAAAT GCNTANANATGTGNANGAACACCANTGGCNAANGCNACTCTCTGG GCTGTAACCTGACNCTGANGCNCGAAAGCGTGGGGAGCAAACANG ATTANATACCCTGGTANTCCACGCCNTANACNATNANTGCTAGGT GTTNNGGGTTTCNATACCCTTGNTGCCNAANTTAACACATTAANCA CTCCGCCTGGNNANTACNGTCNCAANANTGAAACTCNNANGAANT GACNGGGACCCGCACAAGCNNTGNANTATGTGGTTTAANTNNNNN CAACNCNAANAANCTTACCNNGNCTTGACATCTNAATGACCNGNG CANANATGTNCCTTTCCTTCNGNACATTCNNGACAGGTGGTGCATG GNTGTCNTCNNCTCNTGTCNNGNGATGTTGGGTTAANTCCCCGCAN CNANNNN

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Strain (SEQ ID. NO)	Partial 16S ribosomal RNA sequence
5 <i>Bacillus cereus</i> family member EE349 (SEQ ID NO: 18) 10 15 20	AAGGCTTTCGGGTCGTAAAACTCTGTTGTTAGGGAAGAACAAGTG CTAGTTGAATAAGCTGGCACCTTGACGGTACCTAACCAGAAAGCC ACGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAA GCGTTATCCGGAATTATTGGGCGTAAAGCGCGCGCAGGTGGTTTCT TAAGTCTGATGTGAAAGCCCACGGCTCAACCGTGGAGGGGTCATTG GAAACTGGGAGACTTGAGTGCAGAAGAGGAAAGTGGAATTCCATG TGTAGCGGTGAAATGCGTAGAGATATGGAGGAACACCAGTGGCGA AGGCGACTTTTCTGGTCTGTA ACTGACACTGAGGCGCGAAAGCGTG GGGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACG ATGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGAAGTT AACGCATTAAGCACTCCGCCTGGGGAGTACGGCCGCAAGGCTGAA ACTCAAAGGAATTGACGGGGGGCCCGCACAAAGCGGTGGAGCATGTG GTTTAATTCGAAGCAACGCGAAGAACCTTACCAGGTCTTGACATCC TCTGAAAACCCTAGAGATAGGGCTTCTCCTTCGGGAGCAGAGTGA CAGGTGGTGCATGGTTGTCGTCAGCTCGTGTCGTGAGATGTTGGGT TAAGTCC
25 <i>Bacillus subtilis</i> EE218 (SEQ ID NO: 19) 30 35	AAGCTCTGTTGTTAGGGAAGAACAAGTACCGTTCGAATAGGGCGG TACCTTGACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGCC AGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTGTCCGGAATTAT TGGGCGTAAAGGGCTCGCAGGCGGTTTCTTAAGTCTGATGTGAAA GCCCCCGGCTCAACCGGGGAGGGTCATTGGAAACTGGGGAACTTG AGTGCAGAAGAGGAGAGTGGAATTCCACGTGTAGCGGTGAAATGC GTAGAGATGTGGAGGAACACCAGTGGCGAAGGCGACTCTCTGGTC TGTA ACTGACGCTGAGGAGCGAAAGCGTG GGGGAGCGAACAGGATT AGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTT AGGGGGTTTCCGCCCTTANTGCTGCAGCTAACGCATTAAGCACTC CGCCTGGGGAGTACGGTCGCAAGACTGAAACTCAAAGGAATTGAC GGGGGCCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAA CGCGAANAACCTTACCAGGTCTTGACATCCTCTGACAATCCTAGAG ATAGGACGTCCCCTTCGGGGGGCAGAGTGACAGGTGGTGCATGGTT GTCGTCANCTCGTGTCGTGAGATGTTGGNTTAAGTCCCGCAACGAG
40 <i>Bacillus megaterium</i> EE281 (SEQ ID NO: 20) 45 50 55	AAGNCTTTCGGNNCGTAAAACTCTGTTGTTAGGGAAGAACAAGTA CGAGAGTAACTGCTCGTACCTTGACGGTACCTAACCAGAAAGCCA CGGCTAACTACGTGCCAGCAGCCGCGGTAATACGTAGGTGGCAAG CGTTATCCGGAATTATTGGGCGTAAAGCGCGCGCAGGCGGTTTCTT AAGTCTGATGTGAAAGCCCACGGCTCAACCGTGGAGGGGTCATTGG AAACTGGGGAACTTGAGTGCAGAAGAGAAAAGCGGAATTCCACGT GTAGCGGTGAAATGCGTAGAGATGTGGAGGAACACCAGTGGCGAA GGCGGCTTTTTTGGTCTGTA ACTGACGCTGAGGCGCGAAAGCGTGG GGAGCAAACAGGATTAGATACCCTGGTAGTCCACGCCGTAAACGA TGAGTGCTAAGTGTTAGAGGGTTTCCGCCCTTTAGTGCTGCAGCTA ACGCATTAAGCACTCCGCCTGGGGAGTACGGTCGCAAGACTGAAA CTCAAAGGAATTGACGGGGGGCCCGCACAAAGCGGTGGAGCATGTGG TTTAATTCGAAGCAACGCGAAGAACCTTACCAGGTCTTGACATCCT CTGACAACTCTAGAGATAGAGCGTTCCCCTTCGGGGGGACAGAGTG ACAGGTGGTGCATGGTTGTCGTCAGCTCGTGTCGTGAGATGTTGGG TTAAGTCCCNNNNNNNNNNNNNNNNNNNNTCTNAGANNCGNGCT GACNANNCCANGCACCNNGG

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Strain (SEQ ID. NO)	Partial 16S ribosomal RNA sequence
5 <i>Bacillus mycoides</i> BT155 (SEQ ID NO: 21)	GTCTGANGGANACGCCGCGTGAGTGATGAAGGCTTTCGGGTCGT AAAACCTCTGTTGTTAGGGAAGAACAAGTGCTAGTTGAATAAGCTG GCACCTTGACGGTACCTAACCAGAAAGCCACGGCTAACTACGTGC CAGCAGCCGCGGTAATACGTAGGTGGCAAGCGTTATCCGGAATTA 10 TTGGGCGTAAAGCGCGCGCAGGTGGTTTCTTAAGTCTGATGTGAAA GCCCACGGCTCAACCGTGGAGGGTCATTGGAAACTGGGAGACTTG AGTGCAGAAGAGGAAAGTGGAATTCCATGTGTAGCGGTGAAATGC GTAGAGATATGGAGGAACACCAGTGGCGAAGGCGACTTTCTGGTC TGTAACCTGACACTGAGGCGCGAAAGCGTGGGGAGCAAACAGGATT 15 AGATACCCTGGTAGTCCACGCCGTAAACGATGAGTGCTAAGTGTT AGAGGGTTTCCGCCCTTTAGTGCTGAAGTTAACGCATTAAGCACTC CGCCTGGGGAGTACGGCCGCAAGGCTGAAACTCAAAGGAATTGAC GGGGGCCCCGCACAAGCGGTGGAGCATGTGGTTTAATTCGAAGCAA CGCGAAGAACCTTACCAGGTCTTGACATCCTCTGACAACCCTAGAG 20 ATAGGGCTTCCCCTTCGGGGGCAGAGTGACAGGTGGTGCATGGTT GTCGTCAGCTCGTGTCGTGAGATGTTGGGTTAAGTCCCG

[0027] Methods for determining sequence identity are well known by one of ordinary skill in the art. By way of example and not of limitation, the BLASTn algorithm available through National Center for Biotechnology Information (NCBI) can be used to align sequences and determine their identity.

[0028] The foregoing bacterial strains were identified at least to their genus designation by means of conventional biochemistry and morphological indicators. Furthermore, the biochemical assays for confirmed Gram-negative strains such as *Paracoccus kondratievae*, *Alcaligenes faecalis*, and *Enterobacter cloacae* included growth on MacConkey medium and nutrient agar, microscopic examination, growth on 5% and 7.5% NaCl medium, growth at pH 5 and pH 9, growth at 42°C and 50°C, the ability to produce acid upon fermentation with cellobiose, lactose, glycerol, glucose, sucrose, d-mannitol, and starch; fluorescent pigment production; gelatin hydrolysis; nitrate reduction; starch hydrolysis; oxidase reaction, catalase production, urease production and motility. Similarly, the biochemical assays for confirmed Gram-positive strains such as *Bacillus* and *Paenibacillus* included growth in phenylethyl alcohol (PEA) medium and nutrient agar, microscopic examination, growth on 5% and 7.5% NaCl medium, growth at pH 5 and pH 9, growth at 42°C and 50°C, the ability to produce acid upon fermentation with cellobiose, lactose, glycerol, glucose, sucrose, d-mannitol, and starch; fluorescent pigment production; gelatin hydrolysis; nitrate reduction; catalase production, starch hydrolysis; oxidase reaction, urease production and motility.

[0029] Also disclosed is a biologically pure bacterial culture wherein bacteria, i.e. the bacterial strain in the bacterial culture, are mutants of any of the foregoing bacterial strains, which comprises one or more mutations that retain the ability to promote plant growth. Thus, the mutant of any of the foregoing strains will be capable of promoting plant growth when compared to plants to which the mutant was not applied. For example, the mutant can comprise a salt-tolerant mutant (e.g., salt-tolerant mutant of *Paracoccus kondratievae* strain NC35 or a salt-tolerant mutant of *Bacillus mycoides* strain BT155), a thiram-resistant mutant (e.g., a thiram-resistant mutant of *Paracoccus kondratievae* strain NC35, a thiram-resistant mutant of *Bacillus aryabhathi* strain CAP53, a thiram-resistant mutant of *Bacillus mycoides* strain BT155, or a thiram-resistant mutant of *Bacillus thuringiensis* BT013A), or a glyphosate-tolerant mutant (e.g., a glyphosate-tolerant mutant of *Bacillus aryabhathi* strain CAP53).

[0030] The following assays can be used, either individually or in concert, to identify mutant strains, which are capable under gnotobiotic conditions to increase leaf area in 3- to 4-week old whole plants (beaker assay), to increase shoot length or shoot dry weight (soil-plate assay), or to increase root length, root dry weight, shoot dry weight and shoot length (growth pouch assay), which are positively correlated with the ability to promote growth directly in whole plants grown in raw (nonsterilized) soil. In a "Beaker Assay," a mixture of field soil and perlite is sterilized, e.g., by gamma-irradiation (about 1 mRad has proved suitable). Samples of the mix are transferred aseptically to sterile, covered beakers, to which enough water or nutrient solution is added to achieve a moisture content of roughly 25%. Surface-sterilized seed of a test plant, such as rape (*Brassica napus* and *Brassica campestris*), radish, wheat, soybean, corn or cotton, are then sown (1 seed per beaker) after briefly incubating in an aqueous bacterial cell suspension of the strain under study. (A bacterial concentration in the range of 10⁹ colony forming units (CFU) per ml of suspension has proved suitable for this purpose.) After seedlings have developed, under controlled conditions suitable to the test plant, to a point where mature leaves have grown, those plants subjected to bacterial treatments are compared against uninoculated controls to as-

certain differences in leaf area between test and control groups.

[0031] In a "Soil-plate Assay," Petri plates are filled with ground, air-dried soil that has been sterilized by autoclaving, gamma-irradiation, etc. The soil in each plate is then moistened and left covered overnight to assure a uniform moisture distribution through the soil. Inoculated (test) and control seeds, as described above, are thereafter sown in each plate, some six to eight seeds per plate at about 1 cm depth, and grown in the dark under appropriate conditions of temperature and humidity until shoots develop. At the end of incubation, the shoot lengths are determined. In a "Growth Pouch Assay," cellophane containers of the type heretofore used as seed-pack growth pouches are filled with a small volume of deionized water or mineral solution and autoclaved to assure sterility. Test seeds incubated in a bacterial suspension, as previously described, and control seeds not exposed to the bacteria are aseptically sown, about six seeds to a pouch, respectively, and are germinated in the dark under suitable, controlled conditions. After shoots have developed, the pouches are opened and the seedling root length, root dry weight, shoot length and shoot dry weight determined for both tests and controls. Alternatively, the mutant strains can be tested as shown in the Examples for any of the foregoing bacterial strains with respect to the ability to promote plant growth.

[0032] Also disclosed is an inoculum for application to plants, plant seeds, or a plant growth medium, wherein the inoculum comprises an effective amount of a biologically pure bacterial culture of *Bacillus aryabhatai* strain CAP53 (NRRL No. B-50819), *Bacillus aryabhatai* strain CAP56 (NRRL No. B-50817), *Bacillus flexus* strain BT054 (NRRL No. B-50816), *Paracoccus kondratievae* strain NC35 (NRRL No. B-50820), *Bacillus mycoides* strain BT155 (NRRL No. B-50921), *Enterobacter cloacae* strain CAP12 (NRRL No. B-50822), *Bacillus nealsonii* strain BOBA57 (NRRL No. B-50821), *Bacillus mycoides* strain EE118 (NRRL No. B-50918), *Bacillus subtilis* strain EE148 (NRRL No. B-50927), *Alcaligenes faecalis* strain EE107 (NRRL No. B-50920), *Bacillus mycoides* strain EE141 (NRRL No. B-50916), *Bacillus mycoides* strain BT46-3 (NRRL No. B-50922), *Bacillus cereus* family member strain EE128 (NRRL No. B-50917), *Bacillus thuringiensis* strain BT013A (NRRL No. B-50924), *Paenibacillus massiliensis* strain BT23 (NRRL No. B-50923), *Bacillus cereus* strain family member EE349 (NRRL No. B-50928), *Bacillus subtilis* strain EE218 (NRRL No. B-50926), *Bacillus megaterium* strain EE281 (NRRL No. B-50925) or a mutant of any of the foregoing strains, and an agriculturally acceptable carrier. Alternatively, the inoculum can include an effective amount of a mixture comprising at least two biologically pure bacterial cultures described herein. Thus, the mixture of two biologically pure bacterial cultures can include *Bacillus aryabhatai* strain CAP53 and *Bacillus aryabhatai* strain CAP56; *Bacillus aryabhatai* strain CAP53 and *Bacillus flexus* strain BT054; *Bacillus aryabhatai* strain CAP53 and *Paracoccus kondratievae* strain NC35; *Bacillus aryabhatai* strain CAP53 and *Bacillus mycoides* strain BT155; *Bacillus aryabhatai* strain CAP53 and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhatai* strain CAP53 and *Bacillus nealsonii* strain BOBA57; *Bacillus aryabhatai* strain CAP53 and *Bacillus mycoides* strain EE118; *Bacillus aryabhatai* strain CAP53 and *Bacillus subtilis* strain EE148; *Bacillus aryabhatai* strain CAP53 and *Alcaligenes faecalis* strain EE107; *Bacillus aryabhatai* strain CAP53 and *Bacillus mycoides* strain EE141; *Bacillus aryabhatai* strain CAP53 and *Bacillus mycoides* strain BT46-3; *Bacillus aryabhatai* strain CAP53 and *Bacillus cereus* family member strain EE128; *Bacillus aryabhatai* strain CAP53 and *Bacillus thuringiensis* strain BT013A; *Bacillus aryabhatai* strain CAP53 and *Paenibacillus massiliensis* strain BT23; *Bacillus aryabhatai* strain CAP53 and *Bacillus cereus* family member strain EE349; *Bacillus aryabhatai* strain CAP53 and *Bacillus subtilis* strain EE218; *Bacillus aryabhatai* strain CAP53 and *Bacillus megaterium* strain EE281; *Bacillus aryabhatai* strain CAP53 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Bacillus aryabhatai* strain CAP53 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Bacillus aryabhatai* strain CAP53 and thiram-resistant *Bacillus aryabhatai* CAP53; *Bacillus aryabhatai* strain CAP53 and thiram-resistant *Bacillus thuringiensis* BT013A; *Bacillus aryabhatai* strain CAP53 and glyphosate-tolerant *Bacillus aryabhatai* CAP53; *Bacillus aryabhatai* strain CAP56 and *Bacillus flexus* strain BT054; *Bacillus aryabhatai* strain CAP56 and *Paracoccus kondratievae* strain NC35; *Bacillus aryabhatai* strain CAP56 and *Bacillus mycoides* strain BT155; *Bacillus aryabhatai* strain CAP56 and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhatai* strain CAP56 and *Bacillus nealsonii* strain BOBA57; *Bacillus aryabhatai* strain CAP56 and *Bacillus mycoides* strain EE118; *Bacillus aryabhatai* strain CAP56 and *Bacillus subtilis* strain EE148; *Bacillus aryabhatai* strain CAP56 and *Alcaligenes faecalis* strain EE107; *Bacillus aryabhatai* strain CAP56 and *Bacillus mycoides* strain EE141; *Bacillus aryabhatai* strain CAP56 and *Bacillus mycoides* strain BT46-3; *Bacillus aryabhatai* strain CAP56 and *Bacillus cereus* family member strain EE128; *Bacillus aryabhatai* strain CAP56 and *Bacillus thuringiensis* strain BT013A; *Bacillus aryabhatai* strain CAP56 and *Paenibacillus massiliensis* strain BT23; *Bacillus aryabhatai* strain CAP56 and *Bacillus cereus* family member strain EE349; *Bacillus aryabhatai* strain CAP56 and *Bacillus subtilis* strain EE218; *Bacillus aryabhatai* strain CAP56 and *Bacillus megaterium* strain EE281; *Bacillus aryabhatai* strain CAP56 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Bacillus aryabhatai* strain CAP56 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Bacillus aryabhatai* strain CAP56 and thiram-resistant *Bacillus aryabhatai* CAP53; *Bacillus aryabhatai* strain CAP56 and thiram-resistant *Bacillus thuringiensis* BT013A; *Bacillus aryabhatai* strain CAP56 and glyphosate-tolerant *Bacillus aryabhatai* CAP53; *Bacillus flexus* strain BT054 and *Paracoccus kondratievae* strain NC35; *Bacillus flexus* strain BT054 and *Bacillus mycoides* strain BT155; *Bacillus flexus* strain BT054 and *Enterobacter cloacae* strain CAP12; *Bacillus flexus* strain BT054 and *Bacillus nealsonii* strain BOBA57; *Bacillus flexus* strain BT054 and *Bacillus mycoides* strain EE118; *Bacillus flexus* strain BT054 and *Bacillus subtilis* strain EE148; *Bacillus flexus* strain BT054 and

Alcaligenes faecalis strain EE107; *Bacillus flexus* strain BT054 and *Bacillus mycoides* strain EE141; *Bacillus flexus* strain BT054 and *Bacillus mycoides* strain BT46-3; *Bacillus flexus* strain BT054 and *Bacillus cereus* family member strain EE128; *Bacillus flexus* strain BT054 and *Bacillus thuringiensis* strain BT013A; *Bacillus flexus* strain BT054 and *Paenibacillus massiliensis* strain BT23; *Bacillus flexus* strain BT054 and *Bacillus cereus* family member strain EE349; *Bacillus flexus* strain BT054 and *Bacillus subtilis* strain EE218; *Bacillus flexus* strain BT054 and *Bacillus megaterium* strain EE281; *Bacillus flexus* strain BT054 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Bacillus flexus* strain BT054 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Bacillus flexus* strain BT054 and thiram-resistant *Bacillus aryabhattai* CAP53; *Bacillus flexus* strain BT054 and thiram-resistant *Bacillus thuringiensis* BT013A; *Bacillus flexus* strain BT054 and glyphosate-tolerant *Bacillus aryabhattai* CAP53; *Paracoccus kondratievae* strain NC35 and *Bacillus mycoides* strain BT155; *Paracoccus kondratievae* strain NC35 and *Enterobacter cloacae* strain CAP12; *Paracoccus kondratievae* strain NC35 and *Bacillus nealsonii* strain BOBA57; *Paracoccus kondratievae* strain NC35 and *Bacillus mycoides* strain EE118; *Paracoccus kondratievae* strain NC35 and *Bacillus subtilis* strain EE148; *Paracoccus kondratievae* strain NC35 and *Alcaligenes faecalis* strain EE107; *Paracoccus kondratievae* strain NC35 and *Bacillus mycoides* strain EE141; *Paracoccus kondratievae* strain NC35 and *Bacillus mycoides* strain BT46-3; *Paracoccus kondratievae* strain NC35 and *Bacillus cereus* family member strain EE128; *Paracoccus kondratievae* strain NC35 and *Bacillus thuringiensis* strain BT013A; *Paracoccus kondratievae* strain NC35 and *Paenibacillus massiliensis* strain BT23; *Paracoccus kondratievae* strain NC35 and *Bacillus cereus* family member strain EE349; *Paracoccus kondratievae* strain NC35 and *Bacillus subtilis* strain EE218; *Paracoccus kondratievae* strain NC35 and *Bacillus megaterium* strain EE281; *Paracoccus kondratievae* strain NC35 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Paracoccus kondratievae* strain NC35 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Paracoccus kondratievae* strain NC35 and thiram-resistant *Bacillus aryabhattai* CAP53; *Paracoccus kondratievae* strain NC35 and thiram-resistant *Bacillus thuringiensis* BT013A; *Paracoccus kondratievae* strain NC35 and glyphosate-tolerant *Bacillus aryabhattai* CAP53; *Bacillus mycoides* strain BT155 and *Enterobacter cloacae* strain CAP12; *Bacillus mycoides* strain BT155 and *Bacillus nealsonii* strain BOBA57; *Bacillus mycoides* strain BT155 and *Bacillus mycoides* strain EE118; *Bacillus mycoides* strain BT155 and *Bacillus subtilis* strain EE148; *Bacillus mycoides* strain BT155 and *Alcaligenes faecalis* strain EE107; *Bacillus mycoides* strain BT155 and *Bacillus mycoides* strain EE141; *Bacillus mycoides* strain BT155 and *Bacillus mycoides* strain BT46-3; *Bacillus mycoides* strain BT155 and *Bacillus cereus* family member strain EE128; *Bacillus mycoides* strain BT155 and *Bacillus thuringiensis* strain BT013A; *Bacillus mycoides* strain BT155 and *Paenibacillus massiliensis* strain BT23; *Bacillus mycoides* strain BT155 and *Bacillus cereus* family member strain EE349; *Bacillus mycoides* strain BT155 and *Bacillus subtilis* strain EE218; *Bacillus mycoides* strain BT155 and *Bacillus megaterium* strain EE281; *Bacillus mycoides* strain BT155 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Bacillus mycoides* strain BT155 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Bacillus mycoides* strain BT155 and thiram-resistant *Bacillus aryabhattai* CAP53; *Bacillus mycoides* strain BT155 and thiram-resistant *Bacillus thuringiensis* BT013A; *Bacillus mycoides* strain BT155 and glyphosate-tolerant *Bacillus aryabhattai* CAP53; *Enterobacter cloacae* strain CAP12 and *Bacillus nealsonii* strain BOBA57; *Enterobacter cloacae* strain CAP12 and *Bacillus mycoides* strain EE118; *Enterobacter cloacae* strain CAP12 and *Bacillus subtilis* strain EE148; *Enterobacter cloacae* strain CAP12 and *Alcaligenes faecalis* strain EE107; *Enterobacter cloacae* strain CAP12 and *Bacillus mycoides* strain EE141; *Enterobacter cloacae* strain CAP12 and *Bacillus mycoides* strain BT46-3; *Enterobacter cloacae* strain CAP12 and *Bacillus cereus* family member strain EE128; *Enterobacter cloacae* strain CAP12 and *Bacillus thuringiensis* strain BT013A; *Enterobacter cloacae* strain CAP12 and *Paenibacillus massiliensis* strain BT23; *Enterobacter cloacae* strain CAP12 and *Bacillus cereus* family member strain EE349; *Enterobacter cloacae* strain CAP12 and *Bacillus subtilis* strain EE218; *Enterobacter cloacae* strain CAP12 and *Bacillus megaterium* strain EE281; *Enterobacter cloacae* strain CAP12 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Enterobacter cloacae* strain CAP12 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Enterobacter cloacae* strain CAP12 and thiram-resistant *Bacillus aryabhattai* CAP53; *Enterobacter cloacae* strain CAP12 and thiram-resistant *Bacillus thuringiensis* BT013A; *Enterobacter cloacae* strain CAP12 and glyphosate-tolerant *Bacillus aryabhattai* CAP53; *Bacillus nealsonii* strain BOBA57 and *Bacillus mycoides* strain EE118; *Bacillus nealsonii* strain BOBA57 and *Bacillus subtilis* strain EE148; *Bacillus nealsonii* strain BOBA57 and *Alcaligenes faecalis* strain EE107; *Bacillus nealsonii* strain BOBA57 and *Bacillus mycoides* strain EE141; *Bacillus nealsonii* strain BOBA57 and *Bacillus mycoides* strain BT46-3; *Bacillus nealsonii* strain BOBA57 and *Bacillus cereus* family member strain EE128; *Bacillus nealsonii* strain BOBA57 and *Bacillus thuringiensis* strain BT013A; *Bacillus nealsonii* strain BOBA57 and *Paenibacillus massiliensis* strain BT23; *Bacillus nealsonii* strain BOBA57 and *Bacillus cereus* family member strain EE349; *Bacillus nealsonii* strain BOBA57 and *Bacillus subtilis* strain EE218; *Bacillus nealsonii* strain BOBA57 and *Bacillus megaterium* strain EE281; *Bacillus nealsonii* strain BOBA57 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Bacillus nealsonii* strain BOBA57 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Bacillus nealsonii* strain BOBA57 and thiram-resistant *Bacillus aryabhattai* CAP53; *Bacillus nealsonii* strain BOBA57 and thiram-resistant *Bacillus thuringiensis* BT013A; *Bacillus nealsonii* strain BOBA57 and glyphosate-tolerant *Bacillus aryabhattai* CAP53; *Bacillus mycoides* strain EE118 and *Bacillus subtilis* strain EE148; *Bacillus mycoides* strain EE118 and *Alcaligenes faecalis* strain EE107; *Bacillus mycoides* strain EE118 and *Bacillus*

[illegible]

Bacillus subtilis strain EE218; *Paenibacillus massiliensis* strain BT23 and *Bacillus megaterium* strain EE281; *Paenibacillus massiliensis* strain BT23 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Paenibacillus massiliensis* strain BT23 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Paenibacillus massiliensis* strain BT23 and thiram-resistant *Bacillus* *aryabhattai* CAP53; *Paenibacillus massiliensis* strain BT23 and thiram-resistant *Bacillus* *thuringiensis* BT013A; *Paenibacillus massiliensis* strain BT23 and glyphosate-tolerant *Bacillus aryabhattai* CAP53; *Bacillus cereus* family member strain EE349 and *Bacillus subtilis* strain EE218; *Bacillus cereus* family member strain EE349 and *Bacillus megaterium* strain EE281; *Bacillus cereus* family member strain EE349 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Bacillus cereus* family member strain EE349 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Bacillus cereus* family member strain EE349 and thiram-resistant *Bacillus aryabhattai* CAP53; *Bacillus cereus* family member strain EE349 and thiram-resistant *Bacillus thuringiensis* BT013A; *Bacillus cereus* family member strain EE349 and glyphosate-tolerant *Bacillus aryabhattai* CAP53; *Bacillus subtilis* strain EE218 and *Bacillus megaterium* strain EE281; *Bacillus subtilis* strain EE218 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Bacillus subtilis* strain EE218 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Bacillus subtilis* strain EE218 and thiram-resistant *Bacillus aryabhattai* CAP53; *Bacillus subtilis* strain EE218 and thiram-resistant *Bacillus thuringiensis* BT013A; *Bacillus subtilis* strain EE218 and glyphosate-tolerant *Bacillus aryabhattai* CAP53; *Bacillus megaterium* strain EE281 and salt-tolerant and thiram-resistant *Paracoccus* sp. NC35; *Bacillus megaterium* strain EE281 and salt-tolerant and thiram-resistant *Bacillus mycoides* strain BT155; *Bacillus megaterium* strain EE281 and thiram-resistant *Bacillus aryabhattai* CAP53; *Bacillus megaterium* strain EE281 and thiram-resistant *Bacillus thuringiensis* BT013A; or *Bacillus megaterium* strain EE281 and glyphosate-tolerant *Bacillus aryabhattai* CAP53..

[0033] An inoculum which includes an effective amount of a mixture of three bacteriologically pure bacterial cultures can include *Bacillus aryabhattai* strain CAP53, *Bacillus aryabhattai* strain CAP56, and *Bacillus flexus* strain BT054; *Bacillus aryabhattai* strain CAP53, *Bacillus aryabhattai* strain CAP56, and *Paracoccus kondratievae* strain NC35; *Bacillus aryabhattai* strain CAP53, *Bacillus aryabhattai* strain CAP56, and *Bacillus mycoides* strain BT155; *Bacillus aryabhattai* strain CAP53, *Bacillus aryabhattai* strain CAP56, and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhattai* strain CAP53, *Bacillus aryabhattai* strain CAP56, and *Bacillus nealsonii* strain BOBA57; *Bacillus aryabhattai* strain CAP53, *Bacillus flexus* strain BT054, and *Paracoccus kondratievae* strain NC35; *Bacillus aryabhattai* strain CAP53, *Bacillus flexus* strain BT054, and *Bacillus mycoides* strain BT155; *Bacillus aryabhattai* strain CAP53, *Bacillus flexus* strain BT054, and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhattai* strain CAP53, *Bacillus flexus* strain BT054, and *Bacillus nealsonii* strain BOBA57; *Bacillus aryabhattai* strain CAP53, *Paracoccus kondratievae* strain NC35, and *Bacillus mycoides* strain BT155; *Bacillus aryabhattai* strain CAP53, *Paracoccus kondratievae* strain NC35, and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhattai* strain CAP53, *Bacillus mycoides* strain BT155, and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhattai* strain CAP53, *Bacillus mycoides* strain BT155, and *Bacillus nealsonii* strain BOBA57; *Bacillus aryabhattai* strain CAP56, *Bacillus flexus* strain BT054, and *Paracoccus kondratievae* strain NC35; *Bacillus aryabhattai* strain CAP56, *Bacillus flexus* strain BT054, and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhattai* strain CAP56, *Bacillus flexus* strain BT054, and *Bacillus mycoides* strain BT155; *Bacillus aryabhattai* strain CAP56, *Paracoccus kondratievae* strain NC35, and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhattai* strain CAP56, *Paracoccus kondratievae* strain NC35, and *Bacillus nealsonii* strain BOBA57; *Bacillus aryabhattai* strain CAP56, *Bacillus mycoides* strain BT155, and *Enterobacter cloacae* strain CAP12; *Bacillus aryabhattai* strain CAP56, *Enterobacter cloacae* strain CAP12, and *Bacillus nealsonii* strain BOBA57; *Bacillus flexus* strain BT054, *Paracoccus kondratievae* strain NC35, and *Bacillus mycoides* strain BT155; *Bacillus flexus* strain BT054, *Paracoccus kondratievae* strain NC35, and *Enterobacter cloacae* strain CAP12; *Bacillus flexus* strain BT054, *Paracoccus kondratievae* strain NC35, and *Bacillus nealsonii* strain BOBA57; *Bacillus flexus* strain BT054, *Bacillus mycoides* strain BT155, and *Bacillus nealsonii* strain BOBA57; *Bacillus flexus* strain BT054, *Enterobacter cloacae* strain CAP12, and *Bacillus nealsonii* strain BOBA57; *Paracoccus kondratievae* strain NC35, *Bacillus mycoides* strain BT155, and *Enterobacter cloacae* strain CAP12; *Paracoccus kondratievae* strain NC35, *Bacillus mycoides* strain BT155, and *Bacillus nealsonii* strain BOBA57; *Paracoccus kondratievae* strain NC35, *Enterobacter cloacae* strain CAP12, and *Bacillus nealsonii* strain BOBA57; or *Bacillus mycoides* strain BT155, *Enterobacter cloacae* strain CAP12, and *Bacillus nealsonii* strain BOBA57.

[0034] The following mixtures of bacteriologically pure bacterial cultures are favorable for use in stimulating plant growth. They include, without limitation, mixtures of: (1) *Enterobacter cloacae* CAP12 and *Bacillus aryabhattai* CAP53; (2) *Enterobacter cloacae* CAP12 and *Bacillus aryabhattai* CAP56; (3) *Enterobacter cloacae* CAP12 and *Bacillus flexus* BT054; (4) *Enterobacter cloacae* CAP12 and *Bacillus nealsonii* BOBA57; (5) *Bacillus aryabhattai* CAP53 and *Bacillus aryabhattai* CAP56; (6) *Bacillus flexus* BT054 and *Bacillus aryabhattai* CAP56, (7) *Paracoccus kondratievae* NC35 and *Bacillus mycoides* BT155, (8) *Bacillus subtilis* EE218 and *Paracoccus kondratievae* NC35, and (9) *Bacillus subtilis* EE218

and *Bacillus mycoides* BT155.

[0035] Rhizobacteria are root-colonizing bacteria that form symbiotic relationships with many plants, and as such are useful in promoting plant growth. Accordingly, any of the inoculums disclosed herein, regardless of whether they contains a single bacterial strain disclosed herein or a mixture of two or more such bacterial strains, can also include an effective amount of rhizobacteria.

[0036] Such rhizobacteria can be present as a biologically pure bacterial culture. Alternatively, rhizobacteria that are used in the inoculums disclosed herein can include two or more strains of rhizobacteria. By way of example and not of limitation, the rhizobacteria can include *Bradyrhizobium* genus bacteria, *Rhizobium* genus bacteria, or a combination thereof. Also, the *Bradyrhizobium* genus bacteria can comprise *Bradyrhizobium japonicum*, and the *Rhizobium* genus bacteria can comprise *Rhizobium phaseoli*, *Rhizobium leguminosarum*, or a combination thereof. The inclusion of rhizobacteria in the present compositions and methods is especially advantageous in so-called "virgin soils" which do not contain an indigenous population of PGPB such as nitrogen fixing rhizobia. This may occur e.g. where nitrogen-fixing legume crops have not been previously or recently grown.

[0037] In addition to one or more biologically pure bacterial cultures as described in the foregoing sections, an inoculum disclosed herein also comprises an agriculturally acceptable carrier. The carrier can include a dispersant, a surfactant, an additive, water, a thickener, an anti-caking agent, residue breakdown, a composting formulation, a granular application, diatomaceous earth, an oil, a coloring agent, a stabilizer, a preservative, a polymer, a coating, or a combination thereof. One of ordinary skill in the art can readily determine the appropriate carrier to be used taking into consideration factors such as a particular bacterial strain, plant to which the inoculum is to be applied, type of soil, climate conditions, and whether the inoculum is in liquid, solid or powder form.

[0038] The additive can comprise an oil, a gum, a resin, a clay, a polyoxyethylene glycol, a terpene, a viscid organic, a fatty acid ester, a sulfated alcohol, an alkyl sulfonate, a petroleum sulfonate, an alcohol sulfate, a sodium alkyl butane diamate, a polyester of sodium thiobutane dioate, a benzene acetonitrile, a proteinaceous material, or a combination thereof.

[0039] The proteinaceous material can include a milk product, wheat flour, soybean meal, alfalfa meal, yeast extract, blood, albumin, gelatin, or a combination thereof.

[0040] The thickener can comprise a long chain alkylsulfonate of polyethylene glycol, polyoxyethylene oleate, or a combination thereof.

[0041] The surfactant can contain a heavy petroleum oil, a heavy petroleum distillate, a polyol fatty acid ester, a polyethoxylated fatty acid ester, an aryl alkyl polyoxyethylene glycol, an alkyl amine acetate, an alkyl aryl sulfonate, a polyhydric alcohol, an alkyl phosphate, or a combination thereof.

[0042] The anti-caking agent can include a sodium salt such as a sodium sulfite, a sodium sulfate, a sodium salt of monomethyl naphthalene sulfonate, a sodium salt of dimethyl naphthalene sulfonate, or a combination thereof; or a calcium salt such as calcium carbonate, diatomaceous earth, or a combination thereof.

[0043] Any agriculturally acceptable carrier can be used. Such carriers include, but are not limited to, vermiculite, charcoal, sugar factory carbonation press mud, rice husk, carboxymethyl cellulose, peat, perlite, fine sand, calcium carbonate, flour, alum, a starch, talc, polyvinyl pyrrolidone, or a combination thereof.

[0044] Inoculants can be prepared as solid, liquid or powdered formulations as is known in the art. The inoculum of the present invention can be formulated as a seed coating formulation, a liquid formulation for application to plants or to a plant growth medium, or a solid formulation for application to plants or to a plant growth medium.

[0045] When the inoculum is prepared as a liquid formulation for application to plants or to a plant growth medium, it can be prepared in a concentrated formulation or a ready-to-use formulation. In some instances, the seed coating formulation of the present invention is an aqueous or oil-based solution for application to seeds.

[0046] When the inoculum of the present invention is prepared as a solid formulation for application to plants or to a plant growth medium, it can be prepared as a granular formulation or a powder agent. The seed coating formulation can be a powder or granular formulation for application to seeds.

[0047] The inoculum can further include an agrochemical such as a fertilizer, a micronutrient fertilizer material, an insecticide, a herbicide, a plant growth amendment, a fungicide, a molluscicide, an algicide, a bacterial inoculant, a fungal inoculant, or a combination thereof. In some instances, the fertilizer is a liquid fertilizer. The agrochemical can either be applied to a plant growth medium or to plants and/or seeds. Liquid fertilizer can include, without limitation, ammonium sulfate, ammonium nitrate, ammonium sulfate nitrate, ammonium chloride, ammonium bisulfate, ammonium polysulfide, ammonium thiosulfate, aqueous ammonia, anhydrous ammonia, ammonium polyphosphate, aluminum sulfate, calcium nitrate, calcium ammonium nitrate, calcium sulfate, calcined magnesite, calcitic limestone, calcium oxide, calcium nitrate, dolomitic limestone, hydrated lime, calcium carbonate, diammonium phosphate, monoammonium phosphate, magnesium nitrate, magnesium sulfate, potassium nitrate, potassium chloride, potassium magnesium sulfate, potassium sulfate, sodium nitrates, magnesian limestone, magnesia, urea, urea-formaldehydes, urea ammonium nitrate, sulfur-coated urea, polymer-coated urea, isobutylidene diurea, $K_2SO_4 \cdot 2MgSO_4$, kainite, sylvinite, kieserite, Epsom salts, elemental sulfur, marl, ground oyster shells, fish meal, oil cakes, fish manure, blood meal, rock phosphate, super phos-

phates, slag, bone meal, wood ash, manure, bat guano, peat moss, compost, green sand, cottonseed meal, feather meal, crab meal, fish emulsion, or a combination thereof.

[0048] The micronutrient fertilizer material can comprise boric acid, a borate, a boron frit, copper sulfate, a copper frit, a copper chelate, a sodium tetraborate decahydrate, an iron sulfate, an iron oxide, iron ammonium sulfate, an iron frit, an iron chelate, a manganese sulfate, a manganese oxide, a manganese chelate, a manganese chloride, a manganese frit, a sodium molybdate, molybdic acid, a zinc sulfate, a zinc oxide, a zinc carbonate, a zinc frit, zinc phosphate, a zinc chelate, or a combination thereof.

[0049] The insecticide can include an organophosphate, a carbamate, a pyrethroid, an acaricide, an alkyl phthalate, boric acid, a borate, a fluoride, sulfur, a haloaromatic substituted urea, a hydrocarbon ester, a biologically-based insecticide, or a combination thereof.

[0050] The herbicide can comprise a chlorophenoxy compound, a nitrophenolic compound, a nitrocresolic compound, a dipyridyl compound, an acetamide, an aliphatic acid, an anilide, a benzamide, a benzoic acid, a benzoic acid derivative, anisic acid, an anisic acid derivative, a benzonitrile, benzothiadiazinone dioxide, a thiocarbamate, a carbamate, a carbamate, chloropyridinyl, a cyclohexenone derivative, a dinitroaminobenzene derivative, a fluorodinitrotoluidine compound, isoxazolidinone, nicotinic acid, isopropylamine, an isopropylamine derivative, oxadiazolinone, a phosphate, a phthalate, a picolinic acid compound, a triazine, a triazole, a uracil, a urea derivative, endothall, sodium chlorate, or a combination thereof.

[0051] The fungicide can comprise a substituted benzene, a thiocarbamate, an ethylene bis dithiocarbamate, a thiophthalidamide, a copper compound, an organomercury compound, an organotin compound, a cadmium compound, anilazine, benomyl, cyclohexamide, dodine, etridiazole, iprodione, metlaxyl, thiamimefon, triforine, or a combination thereof.

[0052] The fungal inoculant can comprise a fungal inoculant of the family Glomeraceae, a fungal inoculant of the family Clavariaceae, a fungal inoculant of the family Gigasporaceae, a fungal inoculant of the family Acaulosporaceae, a fungal inoculant of the family Sacculosporaceae, a fungal inoculant of the family Entrophosporaceae, a fungal inoculant of the family Pacidsporaceae, a fungal inoculant of the family Diversisporaceae, a fungal inoculant of the family Paraglomeraceae, a fungal inoculant of the family Archaeosporaceae, a fungal inoculant of the family Geosiphonaceae, a fungal inoculant of the family Ambisporaceae, a fungal inoculant of the family Scutellosporaceae, a fungal inoculant of the family Dentiscultataceae, a fungal inoculant of the family Racocetraceae, a fungal inoculant of the phylum Basidiomycota, a fungal inoculant of the phylum Ascomycota, a fungal inoculant of the phylum Zygomycota, or a combination thereof.

[0053] The bacterial inoculant can include a bacterial inoculant of the genus *Rhizobium*, a bacterial inoculant of the genus *Bradyrhizobium*, a bacterial inoculant of the genus *Mesorhizobium*, a bacterial inoculant of the genus *Azorhizobium*, a bacterial inoculant of the genus *Allorhizobium*, a bacterial inoculant of the genus *Sinorhizobium*, a bacterial inoculant of the genus *Kluyvera*, a bacterial inoculant of the genus *Azotobacter*, a bacterial inoculant of the genus *Pseudomonas*, a bacterial inoculant of the genus *Azospirillum*, a bacterial inoculant of the genus *Bacillus*, a bacterial inoculant of the genus *Streptomyces*, a bacterial inoculant of the genus *Paenibacillus*, a bacterial inoculant of the genus *Paracoccus*, a bacterial inoculant of the genus *Enterobacter*, a bacterial inoculant of the genus *Alcaligenes*, a bacterial inoculant of the genus *Mycobacterium*, a bacterial inoculant of the genus *Trichoderma*, a bacterial inoculant of the genus *Gliocladium*, a bacterial inoculant of the genus *Glomus*, a bacterial inoculant of the genus *Klebsiella*, or a combination thereof.

[0054] All of the biologically pure bacterial cultures and inoculums disclosed herein can be used in methods for stimulating plant growth. Such methods include applying the foregoing cultures and inoculums to a plant, plant seed, or plant growth medium in order to stimulate growth of the plant. Techniques for applying inoculants to plants are known in the art, including appropriate modes of administration, frequency of administration, and dosages. The inoculant can be applied to the soil prior to, contemporaneously with, or after sowing seeds, after planting, or after plants have emerged from the ground. The inoculant can also be applied to seeds themselves prior to or at the time of planting (e.g., packaged seed may be sold with the inoculant already applied). The inoculant can also be applied to the plant after it has emerged from the ground, or to the leaves, stems, roots, or other parts of the plant.

[0055] The method for stimulating plant growth can include applying a substance such as glycerol, pyruvate, yeast extract, a polyol (e.g., mannitol, sorbitol, galactitol, fucitol, iditol, inositol, arabitol, xylitol, ribitol), polyethylene glycol or combination thereof to the plant growth medium. Any of the polyols can be used, with the preferred ones being mannitol and sorbitol. For the preparation of yeast extract, *Saccharomyces cerevisiae* is a preferred yeast starting material, although several other yeast strains may be useful to produce yeast ferment materials used in the compositions and methods described herein. Additional yeast strains that can be used instead of or in addition to *Saccharomyces cerevisiae* include *Kluyveromyces marxianus*, *Kluyveromyces lactis*, *Candida utilis* (Torula yeast), *Zygosaccharomyces*, *Pichia pastoris*, and *Hansanula polymorpha*, and others known to those skilled in the art.

[0056] In instances in which the substance is applied to a plant growth medium, at least one bacterial culture or at least one inoculum of the present invention can be applied to a plant or plant seed in the plant growth medium, or to the plant growth medium. Preferably, the inoculum is applied to the plant growth medium as a solid or liquid formulation.

The bacterial culture or inoculum and the chemical can be applied contemporaneously or at separate times. The exact order is not of great relevance, and the optimal combination can be determined empirically by one of ordinary skill in the art without due experimentation. For example, a skilled artisan can set up experimental conditions wherein: (1) the inoculum or bacterial culture and the substance are administered concurrently, (2) the inoculum or bacterial culture is administered on a separate occasion after the substance is added to a plant growth medium, (3) the inoculum or bacterial culture is administered on a separate occasion prior to the substance being added to a plant growth medium. The results of such and similar experimental designs can easily demonstrate the most suitable methods for application of the bacterial strain or inoculum and the substance. Thus, the bacterial culture or inoculum disclosed herein can be applied to a plant growth medium prior to, concurrently with, or after planting of seeds, seedlings, cuttings, bulbs, or plants in the plant growth medium.

[0057] The plant growth medium includes soil, water, an aqueous solution, sand, gravel, a polysaccharide, mulch, compost, peat moss, straw, logs, clay, or a combination thereof. Preferably, the plant growth medium is soil or compost. As is known in the art, the plant growth medium can be stored for future planting.

[0058] For purposes of the compositions and methods of the present invention, the plant can be a dicotyledon, a monocotyledon or a gymnosperm.

[0059] The dicotyledon can be selected from the group consisting of bean, pea, tomato, pepper, squash, alfalfa, almond, aniseed, apple, apricot, arracha, artichoke, avocado, bambara groundnut, beet, bergamot, black pepper, black wattle, blackberry, blueberry, bitter orange, bok-choi, Brazil nut, breadfruit, broccoli, broad bean, Brussels sprouts, buckwheat, cabbage, camelina, Chinese cabbage, cacao, cantaloupe, caraway seeds, cardoon, carob, carrot, cashew nuts, cassava, castor bean, cauliflower, celeriac, celery, cherry, chestnut, chickpea, chicory, chili pepper, chrysanthemum, cinnamon, citron, clementine, clove, clover, coffee, cola nut, colza, corn, cotton, cottonseed, cowpea, crambe, cranberry, cress, cucumber, currant, custard apple, drumstick tree, earth pea, eggplant, endive, fennel, fenugreek, fig, filbert, flax, geranium, gooseberry, gourd, grape, grapefruit, guava, hemp, hempseed, henna, hop, horse bean, horseradish, indigo, jasmine, Jerusalem artichoke, jute, kale, kapok, kenaf, kohlrabi, kumquat, lavender, lemon, lentil, lespe-deza, lettuce, lime, liquorice, litchi, loquat, lupine, macadamia nut, mace, mandarin, mangel, mango, medlar, melon, mint, mulberry, mustard, nectarine, niger seed, nutmeg, okra, olive, opium, orange, papaya, parsnip, pea, peach, peanut, pear, pecan nut, persimmon, pigeon pea, pistachio nut, plantain, plum, pomegranate, pomelo, poppy seed, potato, sweet potato, prune, pumpkin, quebracho, quince, trees of the genus *Cinchona*, quinoa, radish, ramie, rapeseed, raspberry, rhea, rhubarb, rose, rubber, rutabaga, safflower, sainfoin, salsify, sapodilla, Satsuma, scorzonera, sesame, shea tree, soybean, spinach, squash, strawberry, sugar beet, sugarcane, sunflower, swede, sweet pepper, tangerine, tea, teff, tobacco, tomato, trefoil, tung tree, turnip, urena, vetch, walnut, watermelon, yerba mate, wintercress, shepherd's purse, garden cress, peppergrass, watercress, pennycress, star anise, laurel, bay laurel, cassia, jamun, dill, tamarind, peppermint, oregano, rosemary, sage, soursop, pennywort, calophyllum, balsam pear, kukui nut, Tahitian chestnut, basil, huckleberry, hibiscus, passionfruit, star apple, sassafras, cactus, St. John's wort, loosestrife, hawthorn, cilantro, curry plant, kiwi, thyme, zucchini, ulluco, jicama, waterleaf, spiny monkey orange, yellow mombin, starfruit, amaranth, wasabi, Japanese pepper, yellow plum, mashua, Chinese toon, New Zealand spinach, bower spinach, ugu, tansy, chickweed, jocote, Malay apple, paracress, sowthistle, Chinese potato, horse parsley, hedge mustard, campion, agate, cassod tree, thistle, burnet, star gooseberry, saltwort, glasswort, sorrel, silver lace fern, collard greens, primrose, cowslip, purslane, knotgrass, terebinth, tree lettuce, wild betel, West African pepper, yerba santa, tarragon, parsley, chervil, land cress, burnet saxifrage, honeyherb, butterbur, shiso, water pepper, perilla, bitter bean, oca, kampong, Chinese celery, lemon basil, Thai basil, water mimosa, cicely, cabbage-tree, moringa, mauka, ostrich fern, rice paddy herb, yellow sawah lettuce, lovage, pepper grass, maca, bottle gourd, hyacinth bean, water spinach, catsear, fishwort, Okinawan spinach, lotus sweetjuice, gallant soldier, culantro, arugula, cardoon, caigua, mitsuba, chipilin, samphire, mampat, ebolo, ivy gourd, cabbage thistle, sea kale, chaya, huauzontle, Ethiopian mustard, magenta spreen, good king henry, epazole, lamb's quarters, centella plumed cockscomb, caper, rapini, napa cabbage, mizuna, Chinese savoy, kai-lan, mustard greens, Malabar spinach, chard, marshmallow, climbing wattle, China jute, paprika, annatto seed, spearmint, savory, marjoram, cumin, chamomile, lemon balm, allspice, bilberry, cherimoya, cloudberry, damson, pitaya, durian, elderberry, feijoa, jackfruit, jambul, jujube, physalis, purple mangosteen, rambutan, redcurrant, blackcurrant, salal berry, satsuma, ugli fruit, azuki bean, black bean, black-eyed pea, borlotti bean, common bean, green bean, kidney bean, lima bean, mung bean, navy bean, pinto bean, runner bean, mangetout, snap pea, broccoflower, calabrese, nettle, bell pepper, raddichio, daikon, white radish, skirret, tat soi, broccolini, black radish, burdock root, fava bean, broccoli raab, lablab, lupin, sterculia, velvet beans, winged beans, yam beans, mulga, ironweed, umbrella bush, tjuntjula, wakalpulka, witchetty bush, wiry wattle, chia, beech nut, candlenut, colocynth, mamoncillo, Maya nut, mongongo, ogbono nut, paradise nut, and cempedak.

[0060] The dicotyledon can be from a family selected from the group consisting of Acanthaceae (acanthus), Aceraceae (maple), Achariaceae, Achatocarpaceae (achatocarpus), Actinidiaceae (Chinese gooseberry), Adoxaceae (moschatel), Aextoxicaceae, Aizoaceae (fig marigold), Akaniaceae, Alangiaceae, Alseuosmiaceae, Alzateaceae, Amaranthaceae (amaranth), Amborellaceae, Anacardiaceae (sumac), Ancistrocladaceae, Anisophylleaceae, Annonaceae (custard ap-

ple), Apiaceae (carrot), Apocynaceae (dogbane), Aquifoliaceae (holly), Araliaceae (ginseng), Aristolochiaceae (birthwort), Asclepiadaceae (milkweed), Asteraceae (aster), Austrobaileyaceae, Balanopaceae, Balanophoraceae (balanophora), Balsaminaceae (touch-me-not), Barbeyaceae, Barclayaceae, Basellaceae (basella), Bataceae (saltwort), Begoniaceae (begonia), Berberidaceae (barberry), Betulaceae (birch), Bignoniaceae (trumpet creeper), Bixaceae (lipstick tree), Bombacaceae (kapok tree), Boraginaceae (borage), Brassicaceae (mustard, also Cruciferae), Bretschneideraceae, Brunelliaceae (brunellia), Bruniaceae, Brunoniaceae, Buddlejaceae (butterfly bush), Burseraceae (frankincense), Buxaceae (boxwood), Byblidaceae, Cabombaceae (water shield), Cactaceae (cactus), Caesalpiniaceae, Callitrichaceae (water starwort), Calycanthaceae (strawberry shrub), Calyceraceae (calycera), Campanulaceae (bellflower), Canellaceae (canella), Cannabaceae (hemp), Capparaceae (caper), Caprifoliaceae (honeysuckle), Cardiopteridaceae, Caricaceae (papaya), Caryocaraceae (souari), Caryophyllaceae (pink), Casuarinaceae (she-oak), Cecropiaceae (cecropia), Celastraceae (bittersweet), Cephalotaceae, Ceratophyllaceae (hornwort), Cercidiphyllaceae (katsura tree), Chenopodiaceae (goosefoot), Chloranthaceae (chloranthus), Chrysobalanaceae (cocoa plum), Circaeasteraceae, Cistaceae (rockrose), Clethraceae (clethra), Clusiaceae (mangosteen, also Guttiferae), Cneoraceae, Columelliaceae, Combretaceae (Indian almond), Compositae (aster), Connaraceae (cannarus), Convolvulaceae (morning glory), Coriariaceae, Cornaceae (dogwood), Corynocarpaceae (karaka), Crassulaceae (stonecrop), Crossosomataceae (crossosoma), Crypteroniaceae, Cucurbitaceae (cucumber), Cunoniaceae (cunonia), Cuscutaceae (dodder), Cyrillaceae (cyrilla), Daphniphyllaceae, Datis-caceae (datisca), Davidsoniaceae, Degeneriaceae, Dialypetalanthaceae, Diapensiaceae (diapensia), Dichapetalaceae, Didiereaceae, Didymelaceae, Dilleniaceae (dillenia), Dioncophyllaceae, Dipentodontaceae, Dipsacaceae (teasel), Diterocarpaceae (meranti), Donatiaceae, Droseraceae (sundew), Duceodendraceae, Ebenaceae (ebony), Elaeagnaceae (oleaster), Elaeocarpaceae (elaecarpus), Elatinaceae (waterwort), Empetraceae (crowberry), Epacridaceae (epacris), Eremolepidaceae (catkin-mistletoe), Ericaceae (heath), Erythroxylaceae (coca), Eucommiaceae, Eucryphiaceae, Euphorbiaceae (spurge), Eupomatiaceae, Eupteleaceae, Fabaceae (pea or legume), Fagaceae (beech), Flacourtiaceae (flacourtia), Fouquieriaceae (ocotillo), Frankeniaceae (frankenian), Fumariaceae (fumitory), Garryaceae (silk tassel), Geis-solomataceae, Gentianaceae (gentian), Geraniaceae (geranium), Gesneriaceae (gesneriad), Globulariaceae, Gomortegaceae, Goodeniaceae (goodenia), Greyiaceae, Grossulariaceae (currant), Grubbiaceae, Gunneraceae (gunnera), Gyrostemonaceae, Haloragaceae (water milfoil), Hamamelidaceae (witch hazel), Hernandiaceae (hernandia), Himantandraceae, Hippocastanaceae (horse chestnut), Hippocrateaceae (hippocratea), Hippuridaceae (mare's tail), Hoplostigmataceae, Huaceae, Hugoniaceae, Humiriaceae, Hydnoaceae, Hydrangeaceae (hydrangea), Hydrophyllaceae (waterleaf), Hydrostachyaceae, Icacinaceae (icacina), Idiospermaceae, Illiciaceae (star anise), Ixonanthaceae, Juglandaceae (walnut), Julianiaceae, Krameriaceae (krameria), Lacistemataceae, Lamiaceae (mint, also Labiatae), Lardizabalaceae (lardizabala), Lauraceae (laurel), Lecythidaceae (brazil nut), Leeaceae, Leitneriaceae (corkwood), Lennoaceae (lennoa), Lentibulariaceae (bladderwort), Limnanthaceae (meadow foam), Linaceae (flax), Lissocarpaceae, Loasaceae (loasa), Loganiaceae (logania), Loranaceae (showy mistletoe), Lythraceae (loosestrife), Magnoliaceae (magnolia), Malesherbiaceae, Malpighiaceae (barbados cherry), Malvaceae (mallow), Marcgraviaceae (shingle plant), Medusagynaceae, Medusandraceae, Melastomataceae (melastome), Meliaceae (mahogany), Melianthaceae, Mendonciaceae, Menispermaceae (moonseed), Menyanthaceae (buckbean), Mimosaceae, Misodendraceae, Mitrastemonaceae, Molluginaceae (carpetweed), Monimiaceae (monimia), Monotropaceae (Indian pipe), Moraceae (mulberry), Moringaceae (horseradish tree), Myoporaceae (myoporum), Myricaceae (bayberry), Myristicaceae (nutmeg), Myrothamnaceae, Myrsinaceae (myrsine), Myrtaceae (myrtle), Nelumbonaceae (lotus lily), Nepenthaceae (East Indian pitcherplant), Neuradaceae, Nolanaceae, Nothofagaceae, Nyctaginaceae (four-o'clock), Nymphaeaceae (water lily), Nyssaceae (sour gum), Ochnaceae (ochna), Olacaceae (olax), Oleaceae (olive), Oliniaceae, Onagraceae (evening primrose), Oncothecaceae, Opiliaceae, Orobanchaceae (broom rape), Oxalidaceae (wood sorrel), Paeoniaceae (peony), Pandaceae, Papaveraceae (poppy), Papilionaceae, Paracryphiaceae, Passifloraceae (passionflower), Pedaliaceae (sesame), Pellicieraceae, Penaeaceae, Pentaphragmataceae, Pentaphylacaceae, Peridiscaceae, Physenaceae, Phytolaccaceae (pokeweed), Piperaceae (pepper), Pittosporaceae (pittosporum), Plantaginaceae (plantain), Platanaceae (plane tree), Plumbaginaceae (leadwort), Podostemaceae (river weed), Polemoniaceae (phlox), Polygalaceae (milkwort), Polygonaceae (buckwheat), Portulacaceae (purslane), Primulaceae (primrose), Proteaceae (protea), Punicaceae (pomegranate), Pyrolaceae (shinleaf), Quiinaceae, Rafflesiaceae (rafflesia), Ranunculaceae (buttercup or ranunculus), Resedaceae (mignonette), Retziaceae, Rhabdodendraceae, Rhamnaceae (buckthorn), Rhizophoraceae (red mangrove), Rhoipteleaceae, Rhynchocalycaceae, Rosaceae (rose), Rubiaceae (madder), Rutaceae (rue), Sabiaceae (sabia), Saccifoliaceae, Salicaceae (willow), Salvadoraceae, Santalaceae (sandalwood), Sapindaceae (soapberry), Sapotaceae (sapodilla), Sarcolaenaceae, Sargentodoxaceae, Sarracenaceae (pitcher plant), Saururaceae (lizard's tail), Saxifragaceae (saxifrage), Schisandraceae (schisandra), Scrophulariaceae (figwort), Scyphostegiaceae, Scyttopetalaceae, Simaroubaceae (quassia), Simmondsiaceae (jojoba), Solanaceae (potato), Sonneratiaceae (sonneratia), Sphaerosepalaceae, Sphenocleaceae (spenoclea), Stackhousiaceae (stackhousia), Stachyuraceae, Staphyleaceae (bladdernut), Sterculiaceae (cacao), Stylidiaceae, Styracaceae (storax), Surianaceae (suriana), Symplocaceae (sweetleaf), Tamaricaceae (tamarix), Tepuianthaceae, Tetracentraceae, Tetrameristaceae, Theaceae (tea), Theligonaceae, Theophrastaceae (theophrasta), Thymelaeaceae (mezereum), Ticodendraceae, Tiliaceae (linden), Tovariaceae, Trapaceae (water chestnut), Treman-

draceae, Trigoniaceae, Trimeniaceae, Trochodendraceae, Tropaeolaceae (nasturtium), Turneraceae (turnera), Ulmaceae (elm), Urticaceae (nettle), Valerianaceae (valerian), Verbenaceae (verbena), Violaceae (violet), Viscaceae (Christmas mistletoe), Vitaceae (grape), Vochysiaceae, Winteraceae (wintera), Xanthophyllaceae, and Zygophyllaceae (creosote bush).

5 **[0061]** The monocotyledon can be selected from the group consisting of corn, wheat, oat, rice, barley, millet, banana, onion, garlic, asparagus, ryegrass, millet, fonio, raishan, nipa grass, turmeric, saffron, galangal, chive, cardamom, date palm, pineapple, shallot, leek, scallion, water chestnut, ramp, Job's tears, bamboo, ragi, spotless watermeal, arrowleaf elephant ear, Tahitian spinach, abaca, areca, bajra, betel nut, broom millet, broom sorghum, citronella, coconut, cocoyam, maize, dasheen, durra, durum wheat, edo, fique, formio, ginger, orchard grass, esparto grass, Sudan grass, guinea
10 corn, Manila hemp, henequen, hybrid maize, jowar, lemon grass, maguey, bulrush millet, finger millet, foxtail millet, Japanese millet, proso millet, New Zealand flax, oats, oil palm, palm palmyra, sago palm, redtop, sisal, sorghum, spelt wheat, sweet corn, sweet sorghum, taro, teff, timothy grass, triticale, vanilla, wheat, and yam.

[0062] Alternatively, the monocotyledon can be selected from a family selected from the group consisting of Acoraceae (calamus), Agavaceae (century plant), Alismataceae (water plantain), Aloeaceae (aloe), Aponogetonaceae (cape pond-
15 weed), Araceae (arum), Arecaceae (palm), Bromeliaceae (bromeliad), Burmanniaceae (burmannia), Butomaceae (flowering rush), Cannaceae (canna), Centrolepidaceae, Commelinaceae (spiderwort), Corsiaceae, Costaceae (costus), Cyanastraceae, Cyclanthaceae (Panama hat), Cymodoceaceae (manatee grass), Cyperaceae (sedge), Dioscoreaceae (yam), Eriocaulaceae (pipewort), Flagellariaceae, Geosiridaceae, Haemodoraceae (bloodwort), Hanguanaceae (hanguana), Heliconiaceae (heliconia), Hydatellaceae, Hydrocharitaceae (tape grass), Iridaceae (iris), Joinvilleaceae
20 (joinvillea), Juncaceae (rush), Juncaginaceae (arrow grass), Lemnaceae (duckweed), Liliaceae (lily), Limnocharitaceae (water poppy), Lowiaceae, Marantaceae (prayer plant), Mayacaceae (mayaca), Musaceae (banana), Najadaceae (water nymph), Orchidaceae (orchid), Pandanaceae (screw pine), Petrosaviaceae, Philydraceae (philydraceae), Poaceae (grass), Pontederiaceae (water hyacinth), Posidoniaceae (posidonia), Potamogetonaceae (pondweed), Rapateaceae, Restionaceae, Ruppiaceae (ditch grass), Scheuchzeriaceae (scheuchzeria), Smilacaceae (catbrier), Sparganiaceae
25 (bur reed), Stemonaceae (stemona), Strelitziaceae, Taccaceae (tacca), Thurniaceae, Triuridaceae, Typhaceae (cattail), Velloziaceae, Xanthorrhoeaceae, Xyridaceae (yellow-eyed grass), Zannichelliaceae (horned pondweed), Zingiberaceae (ginger), and Zosteraceae (eelgrass).

[0063] The gymnosperm can be selected from a family selected from the group consisting of Araucariaceae, Bowe-
niaceae, Cephalotaxaceae, Cupressaceae, Cycadaceae, Ephedraceae, Ginkgoaceae, Gnetaceae, Pinaceae, Podocar-
30 paceae, Taxaceae, Taxodiaceae, Welwitschiaceae, and Zamiaceae.

[0064] The stimulation of plant growth achieved by the present methods can be measured and demonstrated in a number of ways. Stimulation of plant growth can be shown in instances wherein the average height of the plant is increased by at least about 5%, by at least about 10%, by at least about 15% or by at least about 20% as compared to the average height of plants grown under the same conditions but that have not been treated with the bacterial culture
35 or inoculant. Also, stimulation of plant growth can be shown in instances wherein the average leaf diameter of the leaves of plant is increased by at least about 5%, by at least about 10%, by at least about 15% or by at least about 20% as compared to the average leaf diameter of plants grown under the same conditions but that have not been treated with the bacterial culture or inoculant. Similarly, stimulation of plant growth can be shown in instances wherein the average root length of the plant is increased by at least about 5%, by at least about 10%, by at least about 15% or by at least
40 about 20% as compared to the average root length of the plants grown under the same conditions but that have not been treated with the bacterial culture or inoculant.

[0065] The present disclosure is also directed to plant seeds, which are coated with any of the inoculums or bacteri-
ologically pure bacterial cultures disclosed herein. The seed can be from any of the plants discussed in the foregoing sections belonging to monocotyledons, dicotyledons or gymnosperms. The bacterial inoculant or culture can be applied
45 to the seeds through the use of a suitable coating mechanism prior to the seeds being sold into commerce for planting. The process of coating seeds with such an inoculum is generally well known to those skilled in the art. For example, the bacteria can be mixed with a porous, chemically inert granular carrier as described by U.S. Pat. No. 4,875,921. Alternatively, the bacterial inoculant can be prepared with or without a carrier and sold as a separate inoculant to be inserted directly into the furrows into which the seed is planted. The process for inserting such inoculants directly into the furrows
50 during seed planting is also generally well known in the art. The density of inoculation of these bacterial cultures onto seeds or into the furrows should be sufficient to populate the sub-soil region adjacent to the roots of the plant with viable bacterial growth.

[0066] The present invention also relates to kits for stimulating plant growth, which include an inoculum of the invention, and instructions for applying the inoculum to plants, plant seeds, or a plant growth medium. Kits containing inoculants
55 of the invention will typically include one or more containers of the inoculant, and printed instructions for using the inoculant for promoting plant growth. The kit can also include tools or instruments for reconstituting, measuring, mixing, or applying the inoculant, and will vary in accordance with the particular formulation and intended use of the inoculant.

[0067] As shown in Example 10 and other tables, bacteria provide a good system in which to select mutations for

desired characteristics. It is possible to force such mutations through proper selection of desirable traits, while retaining the desired plant growth-promoting capabilities in bacteria. Accordingly, traits that may be desirable to induce in bacterial strains disclosed herein by forcing mutations without affecting plant growth promotion include, but are not limited to, antibiotic resistance, heavy metal resistance, tolerance to heat and cold, high and low salt tolerance, metabolic deficiencies (such as requirements for certain amino acids), metabolic gain-of-function (such as the ability to metabolize polysaccharides or plastic compounds), ability to withstand dessication, resistance to UV radiation, tolerance of man-made chemicals, ability to bind more tightly to plant roots, higher affinity for plants, increased ability to colonize plants, motility, ability to accept recombinant DNA, and ability to express exogenous proteins. These attributes can be garnered by use of selective pressure or through man-made manipulation of plant growth promoting bacteria's genetics.

[0068] Further details concerning the preparation of bacterial inoculants and methods for inoculating plants with bacterial inoculants are found in e.g. U.S. Pat. Nos. 5,586,411; 5,697,186; 5,484,464; 5,906,929; 5,288,296; 4,875,921; 4,828,600; 5,951,978; 5,183,759; 5,041,383; 6,077,505; 5,916,029; 5,360,606; 5,292,507; 5,229,114; 4,421,544; and 4,367,609.

EXAMPLES

[0069] The following non-limiting examples are provided to further illustrate the present invention.

Comparative Example 1: Isolation and identification of bacterial strains.

[0070] Soil samples from rhizospheres of the healthiest and most resistant potato (*Solanum tuberosum*), yellow summer squash (*Cucurbita pepo*), tomato (*Solanum lycopersicum*), and pole bean (*Phaseolus coccineus*) plants were collected, diluted in sterile water, and spread onto nutrient agar plates. Bacterial isolates that demonstrated high growth rates and were able to be passaged and propagated were selected for further study. The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in an equal amount of distilled water. Ten lettuce seeds per treatment were planted at a depth of 1 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting in 4 cm pots with 0.5 µl of resuspended bacteria in water mixed into 10 ml of H₂O. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. After one week, plant heights and leaf diameters, as well as overall health of the plants were collected. Initial screening of rhizosphere isolates resulted in obtaining greater than 200 distinct species of bacteria and fungi from the rhizosphere of the four plants. Some of the bacterial species are described in Table 2. Identified strains are indicated by their proper bacterial identifications. Other strains are indicated by their unknown identification number. Inoculants giving results near control (+/- 2%) were left out of the table.

TABLE 2

	Butterhead Lettuce		
Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
Uninoculated	1.8	Control	.07
<i>Paracoccus kondratiaevae</i> NC35	2	111.1%	.05
<i>B. aryabhatai</i> CAP53	3.65	202.8%	.45
<i>B. flexus</i> BT054	2.45	136.1%	.11
<i>Bacillus mycoides</i> strain BT155	2.17	120.4%	.21
<i>B. aryabhatai</i> CAP56	2.1	116.7%	.20
<i>B. nealsonii</i> BOBA57	2.8	155.6%	.03
<i>E. cloacae</i> CAP12	2.4	133.3%	.41
Unknown 8	1.77	77.8%	.65
Unknown 122	1.9	105.6%	.11
Unknown 15	1.4	77.8%	.41

(continued)

	Butterhead Lettuce		
Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
Unknown 39	1.8	100.0%	.20
Unknown 401	2	111.1%	.21
Unknown 402	1.53	85.2%	.27
Unknown 41	1.45	80.6%	.31
Unknown 42	1.4	77.8%	.15
Unknown 44	2.2	133.3%	.08
Unknown 51	1.83	102.9%	.21

[0071] Bacterial strains that produced the greatest effect on the overall plant health and plant height in the initial lettuce trial were subjected to further identification. Bacterial strains were grown overnight in Luria Bertani broth at 37°C, and overnight cultures were spun down in a centrifuge. Media was decanted and the remaining bacterial pellet was subjected to chromosomal DNA isolation using the Qiagen Bacterial Chromosomal DNA Isolation kit. Chromosomal DNA was subjected to PCR amplification of the 16S rRNA coding regions using the primers E338F 5'-ACT CCT ACG GGA GGC AGC AGT-3' (SEQ ID NO: 7), E1099R A 5'-GGG TTG CGC TCG TTG C-3' (SEQ ID NO: 8), and E1099R B 5'-GGG TTG CGC TCG TTA C-3' (SEQ ID NO: 9). PCR amplicons were purified using a Promega PCR purification kit, and the resultant amplicons were diluted and sent to the University of Missouri DNA Core for DNA sequencing. DNA sequences were compared to the NCBI BLAST database of bacterial isolates, and genus and species were identified by direct comparison to known strains. Top identified species are indicated in Table 2. In many cases, 16S rRNA DNA sequences were only able to delineate the genus of the selected bacterial strain. In cases where a direct identification was not forthcoming, additional biochemistry analyses, using methods standard in the field, were performed to differentiate strains at the species and strain levels, and are listed in Table 3.

TABLE 3

Test	<i>E. cloacae</i> CAP12	<i>P. kondratiaevae</i> NC35	<i>B. aryabhatai</i> CAP53	<i>B. flexus</i> BT054	<i>B. mycoides</i> BT155	<i>B. aryabhatai</i> CAP56	<i>B. sp.</i> BOBA57
Urease	-	-	-	-	-	-	+
Catalase	+	+	+	+	+	+	+
Oxidase	-	+	+	+	-	-	-
Nitrate	+	+	-	+	+	-	+
Growth, 5% NaCl	+	-	+	+	-	+	+
Growth, 7.5% NaCl	-	-	+	+	-	+	-
Growth, 42°C	+	+	+	+	+	+	+
Growth, 50°C	-	-	+	+	-	+	-
Growth, pH 5	+	-	+	+	-	+	-
Growth, pH 9	+	+	+	+	+	+	+
Acid, Cellobiose	+	-	+	+	+	+	-
Acid, Lactose	+	-	+	+	+	-	+
Acid, Starch	-	-	-	+	-	+	-

Example 1: Isolation and identification of additional bacterial strains.

[0072] Soil samples from agricultural fields near Gas, Kansas were collected, diluted in sterile water, and spread onto nutrient agar plates. Bacterial isolates that demonstrated high growth rates and were able to be passaged and propagated were selected for further study. The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15g, CaCl₂ 2H₂O 0.013g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in an equal amount of distilled water. Corn seeds were coated with commercial seed polymer mixed with water alone (1.6 µl per seed total) or commercial seed polymer containing selected bacterial strains (1.6 µl per seed total). Coated seeds were planted in (3 inch) 7.62 cm diameter pots at a depth of 1 inch (2.54 cm) in loam top soil (Columbia, MO) that was sieved to remove large debris. Plants were grown at temperatures between 18-24°C (65-75°F) with 11 hours of light/day, and 50 ml of watering at planting and every 3 days. After two weeks, plant heights and leaf diameters, as well as overall health of the plants were collected. For germination assays and determining 3 day root length, seeds were coated as indicated above and evenly dispersed at 10 seeds per paper towel. The paper towels were wetted with 10 mls of water, rolled up, placed in a small plastic bag and incubated at 30°C or placed on a germination heat mat at 27-30°C (80-85°F). Root measurements were recorded after 3 days. Initial screening of rhizosphere isolates resulted in obtaining greater than 100 distinct species of bacteria and fungi from the rhizosphere. Some of the bacterial species are described in Table 4. Identified strains are indicated by their proper bacterial identifications.

TABLE 4

	Corn Seed Treatments	
Bacterial Inoculant	Avg. Height (2 weeks) normalized to polymer control (%)	Avg. Root Length (3 days) normalized to polymer control (%)
Polymer control	100	100
<i>B. mycoides</i> EE118	111.1	189.1
<i>B. subtilis</i> EE148	99.4	172.8
<i>Alcaligenes faecalis</i> EE107	111.5	129.2
<i>B. mycoides</i> EE141	109.2	143.5
<i>B. mycoides</i> BT46-3	105.6	141.3
<i>B. cereus</i> family member EE128	105.6	-
<i>B. thuringiensis</i> BT013A	101.8	103.8
<i>Paenibacillus massiliensis</i> BT23	104.2	139.4
<i>B. cereus</i> family member EE349	105.2	-
<i>B. subtilis</i> EE218	106.6	-
<i>B. megaterium</i> EE281	107.8	-

[0073] Bacterial strains that produced the greatest effect on plant health are described in Table 4. Bacterial strains were grown overnight in Luria Bertani broth at 37°C, and overnight cultures were spun down in a centrifuge. Media was decanted and the remaining bacterial pellet was subjected to chromosomal DNA isolation using the Qiagen Bacterial Chromosomal DNA Isolation kit. Chromosomal DNA was subjected to PCR amplification of the 16S rRNA coding regions using the primers E338F 5'-ACT CCT ACG GGA GGC AGC AGT-3', E1099R A 5'-GGG TTG CGC TCG TTG C-3' and E1099R B 5'-GGG TTG CGC TCG TTA C-3'. PCR amplicons were purified using a Promega PCR purification kit, and the resultant amplicons were diluted and sent to the University of Missouri DNA Core for DNA sequencing. DNA sequences were compared to the NCBI BLAST database of bacterial isolates, and genus and species were identified by direct comparison to known strains. Top identified species are indicated in Table 4. In many cases, 16S rRNA DNA sequences were only able to delineate the genus of the selected bacterial strain. In cases where a direct identification was not forthcoming, additional biochemistry analyses, using methods standard in the field, were performed to differentiate strains

at the species and strain levels, and the differentiated strains are listed in Table 5.

TABLE 5

Test	<i>B. thuringiensis</i> BT013A	<i>B. cereus</i> family member EE349	<i>B. subtilis</i> EE148	<i>B. subtilis</i> EE218	<i>B. megaterium</i> EE281	<i>Paenibacillus</i> <i>massiliensis</i> BT23	<i>B. mycoides</i> BT46-3	<i>Alcaligenes faecalis</i> EE107	<i>B. mycoides</i> EE118	<i>B. cereus</i> family member EE128	<i>B. mycoides</i> EE141
Motility	+	+	+	+	+	+	-	+	-	-	-
Rhizoid Colony	-	-	-	-	-	+	+	-	+	-	+
Catalase	+	+	+	+	+	+	+	+	+	+	+
Oxidase	+	-	-	-	-	-	-	+	-	-	-
Nitrate	+	+	wk	-	-	-	+	+	+	+	+
Growth, 5% NaCl	+	wk	-	+	+	-	+	+	-	+	-
Growth, 7.5% NaCl	wk	-	-	+	+	-	-	-	-	-	-
Growth, 42°C	-	+	+	+	+	+	+	+	-	+	-
Growth, 50°C	-	-	-	-	-	-	-	-	-	-	-
Growth, pH 5	wk	-	+	+	+	-	wk	+	-	+	-
Growth, pH 9	+	+	-	+	+	-	wk	+	+	+	-
Acid, Cellobiose	-	-	wk	+	-	+	+	wk	+	-	wk
Acid, Lactose	-	+	+	+	+	-	+	+	-	+	wk
Acid, Starch	-	+	-	+	+	-	+	wk	+	+	-

wk = weak growth or low growth

Comparative Example 2: Testing of bacteria disclosed herein on alfalfa.

[0074] The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and bacteria resuspended in an equal amount of distilled water. Ten Zeba-coated alfalfa seeds were planted for each treatment at a depth of 0.6 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 μl of resuspended bacteria in water mixed into 10 ml of H₂O. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Alfalfa was allowed to grow for 1 week to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications and final height data are listed in Table 6.

TABLE 6

	Alfalfa		
Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
Uninoculated	4.82	-	.008
<i>B. aryabhattai</i> CAP56	4.85	101.20%	.016

(continued)

	Alfalfa		
Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
<i>B. nealsonii</i> BOBA57	4.86	101.70%	.021
<i>E. cloacae</i> CAP12	5.6	116.23%	.020

Comparative Example 3: Testing of bacteria disclosed herein on cucumbers.

[0075] The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in equal amount of distilled water. Ten cucumber seeds were planted for each treatment at a depth of 1 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 µl of resuspended bacteria in water mixed into 10 ml of H₂O. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Cucumbers were allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications and final height data are listed in Table 7.

TABLE 7

	Cucumbers		
Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
Uninoculated	11.23	-	.067
<i>B. aryabhattai</i> CAP53	11.5	102.00%	.023
<i>B. aryabhattai</i> CAP56	11.35	101.20%	.035
<i>B. nealsonii</i> BOBA57	11.33	101.10%	.014

Comparative Example 4: Testing of bacteria disclosed herein on yellow squash.

[0076] The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in an equal amount of distilled water. Ten yellow squash seeds were planted for each treatment at a depth of 1 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 µl of resuspended bacteria in water mixed into 10 ml of H₂O. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Squash was allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications, final height data, and final leaf diameter (by span of the two leaves) data are listed in Table 8.

TABLE 8

		Yellow Squash			
Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM	Leaf Diameter (cm)	Comparison
Uninoculated	10.16	-	.028	5.08	-
<i>B. aryabhattai</i> CAP53	11.75	115.60%	.055	7.25	142.60%
<i>B. flexus</i> BT054	11.88	116.90%	.017	6.36	125.20%
<i>Bacillus mycoides</i> BT155	11.92	117.20%	.051	6.33	124.60%
<i>B. aryabhattai</i> CAP56	11.95	117.60%	.027	6.33	124.60%

(continued)

		Yellow Squash			
	Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM	Leaf Diameter (cm)
	<i>B. nealsonii</i> BOBA57	11.89	117.00%	.118	6.42
	<i>E. cloacae</i> CAP12	11.42	112.30%	.039	6.83

Comparative Example 5: Testing of bacteria disclosed herein on ryegrass.

[0077] The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in an equal amount of distilled water. Thirty ryegrass seeds were planted for each treatment at a depth of 0.3 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 μl of resuspended bacteria in water mixed into 10 ml of H₂O. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Ryegrass was allowed to grow for 1.5 weeks to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications and height data are listed in Table 9.

TABLE 9

		Ryegrass		
	Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
	Uninoculated	1.61	-	.023
	<i>B. aryabhattai</i> CAP53	2.01	124.70%	.012
	<i>B. flexus</i> BT054	2.21	137.30%	.034
	<i>Bacillus mycoides</i> BT155	2.29	142.20%	
	<i>B. aryabhattai</i> CAP56	2.19	136.00%	.009
	<i>B. nealsonii</i> BOBA57	2.29	142.40%	.045
	<i>E. cloacae</i> CAP12	1.98	122.50%	.015

Comparative Example 6: Testing of bacteria disclosed herein on corn.

[0078] The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in an equal amount of distilled water. Ten corn seeds were planted for each treatment at a depth of 2.5 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 μl of resuspended bacteria in water mixed into 10 ml of H₂O. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Corn was allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications and final height data are listed in Table 10.

TABLE 10

		Corn		
	Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
	Uninoculated	8.9	-	.039
	<i>B. aryabhattai</i> CAP53	11.01	123.60%	.081
	<i>B. flexus</i> BT054	9.96	112.00%	.095

(continued)

	Corn		
Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
<i>Bacillus mycoides</i> strain BT155	9.6	107.90%	.041
<i>B. aryabhattai</i> CAP56	9.54	107.10%	.088
<i>B. nealsonii</i> BOBA57	9.23	103.70%	.077

Comparative Example 7: Testing of bacteria on soybeans

[0079] The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight, or for *Bradyrhizobium* or *Rhizobium* on yeast mannitol media). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in equal amount of distilled water. Ten soybean seeds were planted for each treatment at a depth of 2.5 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 µl of resuspended bacteria in water mixed into 10 ml of H₂O. When testing two bacterial strains, 0.5 µl of each resuspended bacteria was mixed into 10 ml of H₂O. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Soybeans were allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications and final height data are listed in Table 11. Co-inoculation of bacteria strains in the present invention with members of the *Bradyrhizobium* sp. or *Rhizobium* sp. lead to an increase in plant growth compared to either inoculant alone.

TABLE 11

	Soybeans		
Bacterial Inoculant	Avg. Height (cm)	Comparison	SEM
Uninoculated	13.94	-	.089
<i>B. aryabhattai</i> CAP53	16.32	117.1%	.146
<i>B. flexus</i> BT054	17.85	128.0%	.177
<i>Bacillus mycoides</i> strain BT155	18.93	135.8%	.117
<i>B. aryabhattai</i> CAP56	17.23	123.6%	.133
<i>B. aryabhattai</i> CAP53	16.32	117.1%	.077
<i>B. aryabhattai</i> CAP53 and <i>Bradyrhizobium</i> sp.	16.72	119.9%	.182
<i>B. aryabhattai</i> CAP53 and <i>Rhizobium</i> sp.	17.32	124.2%	.086
<i>Bradyrhizobium</i> sp.	14.25	102.2%	
<i>Rhizobium</i> sp.	14.75	105.8%	

Comparative Example 8: Testing of bacteria disclosed herein on soybeans with the additive effect of plant-growth-promoting chemicals.

[0080] The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in equal amount of distilled water. Ten soybean seeds were planted for each treatment at a depth of 2.5 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 µl of resuspended bacteria in water mixed into 10 ml of H₂O alone, into 10 ml of H₂O with 0.5 µl glycerol, into 10 ml of H₂O with 0.5 µl 2,3-butanediol, or into 10 ml of H₂O with 0.5 mg yeast extract. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Soybeans were allowed to grow for 2 weeks to analyze emergence and initial outgrowth

of plants under described conditions. Identified strains indicated by their proper bacterial identifications and final height data are listed in Table 12.

TABLE 12

		Soybeans			
	Bacterial Inoculant	Avg. Height (cm)	Comparison to Uninoculated	Comparison to Inoculant	SEM
	Uninoculated	11.24	-		.153
	Uninoculated and Glycerol	12.34	109.8%		.107
	Uninoculated and Yeast Extract	14.03	124.8%		.212
	<i>B. aryabhatai</i> CAP53	12.56	111.7%	-	.146
	<i>B. aryabhatai</i> CAP53 and Glycerol	13.22	117.6%	105.3%	.118
	<i>B. aryabhatai</i> CAP53 and Yeast Extract	14.73	131.0%	117.3%	.119
	<i>Paracoccus</i> sp. NC35	13.32	118.5%	-	.027
	<i>Paracoccus</i> sp. NC35 and 2,3-butanediol	15.09	134.3%	113.3%	.210
	<i>Paracoccus</i> sp. NC35 and Yeast Extract	15.83	140.8%	118.8%	.145

Comparative Example 9: Testing of bacteria disclosed herein on corn with the additive effect of plant-growth-promoting chemicals.

[0081] The selected strains were grown in minimal media (KH₂PO₄ 3 g, Na₂HPO₄ 6 g, NH₄Cl 1 g, NaCl 0.50 g, MgSO₄ 7H₂O 0.15 g, CaCl₂ 2H₂O 0.013 g, and glucose 1 g, per L dry weight). Overnight cultures (30°C) of selected strains were spun down, media decanted off, and resuspended in an equal amount of distilled water. Ten corn seeds were planted for each treatment at a depth of 2.5 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 μl of resuspended bacteria in water mixed into 10 ml of H₂O alone, into 10 ml of H₂O with 0.5 μl 2,3-butanediol, or into 10 ml of H₂O with 0.5 mg yeast extract. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Corn was allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications and final height data are listed in Table 13.

TABLE 13

		Corn			
	Bacterial Inoculant	Avg. Height (cm)	Comparison to Uninoculated	Comparison to Inoculant	SEM
	Uninoculated	15.15	-		.156
	Uninoculated and 2,3-butanediol	16.03	105.8%		.078
	Uninoculated and Yeast Extract	17.04	112.5%		.101
	<i>Paracoccus</i> sp. NC35	16.04	105.9%	-	.023
	<i>Paracoccus</i> sp. NC35 and 2,3-butanediol	16.24	107.2%	101.2%	.111
	<i>Paracoccus</i> sp. NC35 and Yeast Extract	17.96	118.5%	112.0%	.127

Comparative Example 10: Generation of mutants able to grow in high salt conditions that retain plant growth-promoting ability using selective pressure.

[0082] *Paracoccus kondratiaevae* NC35 and *Bacillus mycoides* BT155 were found to be salt sensitive and not very active in high salt soil types (Table 3). To induce salt tolerance in these plant growth promoting bacteria, a selective pressure using successively higher concentrations of NaCl was used to find mutants that could tolerate these high salt soil types. The selected strains were grown in Luria Bertani liquid media at 37°C overnight and plated on 1% NaCl salt agar media and allowed to grow for 48 hours at 30°C. Individual colonies of strains that survived on the 1% salt LB agar plates were grown in Luria Bertani with 1% NaCl liquid media at 37°C overnight and plated on 3% NaCl salt LB agar media for 48 hours at 30°C. Colonies of strains that survived on the 3% NaCl salt LB agar were grown in Luria Bertani with 3% NaCl liquid media at 37°C overnight and plated on 5% NaCl salt LB agar media and allowed to grow for 48 hours at 30°C. Bacterial colonies selected from the 5% salt LB agar plates were grown in Luria Bertani overnight plus 5% NaCl media at 37°C. Overnight cultures of original strains in minimal media and salt-tolerant mutants were grown overnight and were spun down, media decanted off, and resuspended in equal amount of distilled water. Nine soybean seeds were planted for each treatment at a depth of 2.5 cm in loam top soil (Columbia, MO) or loam supplemented with 5% salt solution (w/w), wherein both of the soils were sieved to remove large debris. Seeds were inoculated at planting with 0.5 µl of resuspended bacteria in water mixed into 10 ml of H₂O. Ten ml of H₂O was sufficient to deliver the bacteria into the 3 in³ (7.62 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-75°F (18-24°C) with 11 hours of light/day, and 5 ml of watering every 3 days. Soybeans were allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications and final height data are listed in Table 14. Mutations were forced in a salt-sensitive strain that retained plant growth-promoting ability.

TABLE 14

Bacterial Inoculant	Avg. Height, Loam Soil (cm)	Percentage	Avg. height, 5% Salt Soil (cm)	Percentage	SEM, Loam Soil	SEM, 5% Salt Soil
H ₂ O Control	10.54	100%	7.34	100%	.322	.117
<i>Paracoccus</i> sp NC35, salt sensitive	11.91	113%	7.15	97.4%	.115	.215
<i>Paracoccus</i> sp. NC35 salt-tolerant	12.02	114%	9.23	125.7%	.451	.105
<i>Bacillus mycoides</i> strain BT155, salt sensitive	12.75	120.9%	7.45	101.5%	.212	.279
<i>Bacillus mycoides</i> strain BT155, salt-tolerant	12.92	122.6%	8.93	121.7%	.185	.056

Comparative Example 11: Generation of mutants able to grow in the presence of thiram that retained plant growth-promoting ability using selective pressure.

[0083] Thiram is a common well-established insecticide and fungicide used on a wide range of agriculture crops. Thiram is also quite antibacterial, and can have deleterious effects on most plant growth promoting bacteria, including those described herein. All strains tested were found to have a high degree of thiram sensitivity. Several of the strains, including the salt resistant *Paracoccus kondratiaevae* NC35 from Example 11, *Bacillus aryabhattai* CAP53, the salt resistant *Bacillus mycoides* BT155 from Example 11, and *Bacillus thuringiensis* BT0013A were grown in LB media with minute quantities of thiram (0.05 mg/L). Growth under these condition took four days, rather than the typical 12 hours in the absence of thiram. The mutated bacteria that began to grow in the presence of thiram were sequentially subjected to higher and higher concentrations of thiram (0.25 mg/L, 0.50 mg/L, 1 mg/L, 5 mg/L). Upon achieving a mutated culture that could withstand the highest amount of thiram on the seed (0.05 mg/seed), *Paracoccus kondratiaevae* NC35, the NC35 thiram (ThR)/salt resistant mutant, *Bacillus aryabhattai* CAP53, the CAP53 ThR mutant, *Bacillus mycoides* BT155, the BT155 salt/ThR mutant, *Bacillus thuringiensis* BT013A and BT013A ThR mutant were applied on seeds in the presence of thiram at 0.05 mg/seed. Overnight cultures of original strains in minimal media and thiram-resistant mutants were

grown overnight and were spun down, media decanted off, and resuspended in equal amount of distilled water. Ten soybean seeds were planted for each treatment at a depth of 2.5 cm in loam top soil (Columbia, MO) in standard seed treatment with 0.5 mg/seed thiram. Plants were grown at temperatures between 18-24°C (65-75°F) with 11 hours of light/day, and 5 ml of watering every 3 days. Soybeans were allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Mutations were forced in all strains that retained plant growth-promoting ability. Results are shown in Table 15 below.

TABLE 15

Bacterial Inoculant	Percentage	SEM,
H ₂ O Control	100%	3.4%
<i>Paracoccus</i> sp NC35, thiram sensitive	114.3%	7.1%
<i>Paracoccus</i> sp. NC35 ThR, thiram-resistant	135.3%	9.2%
<i>Bacillus aryabhattai</i> CAP53, thiram sensitive	107.6%	10.1%
<i>Bacillus aryabhattai</i> CAP53 ThR thiram-resistant	111.9%	7.2%
<i>Bacillus mycoides</i> BT155, thiram sensitive	103.9%	6.9%
<i>Bacillus mycoides</i> BT 155, ThR thiram-resistant	124.1%	12.3%
<i>Bacillus thuringiensis</i> BT013A, thiram sensitive	100.1%	5.7%
<i>Bacillus thuringiensis</i> BT013A, ThR, thiram-resistant	104.9%	8.3%

Comparative Example 12: Generation of mutants able to grow in the presence of glyphosate that retained plant growth-promoting ability using selective pressure.

[0084] Glyphosate is a common well-established herbicide used on a wide range of agriculture crops. Glyphosate is also quite inhibitory to various bacteria, and can have deleterious effects on some plant growth promoting bacteria, including those described herein. Two of the strains described herein had a degree of inhibition in the presence of glyphosate. *Bacillus arybhattai* CAP53 were grown in LB media with minute quantities of glyphosate (0.05 mg/L). Growth under these conditions took two days, rather than the typical 12 hours in the absence of glyphosate. The mutated bacteria that began to grow in the presence of glyphosate were sequentially subjected to higher and higher concentrations of glyphosate (0.25 mg/L, 0.50 mg/L, 1 mg/L, 5 mg/L). Upon achieving a mutated culture that could withstand a higher amount of glyphosate in the soil (100 ppm), *Bacillus aryhbhattai* CAP53 glyphosate-tolerant and wild type strains were applied to seeds at a rate of 1x 10⁵ CFU/seed with a standard seed treatment. Ten soybean seeds were planted for each treatment at a depth of 2.5 cm in loam top soil (Columbia, MO) in standard seed treatment with 100 ppm glyphosate. Plants were grown at temperatures between 18-24°C (65-75°F) with 11 hours of light/day, and 5 ml of watering every 3 days. Soybeans were allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Mutations were forced in all strains that retained plant growth-promoting ability.

TABLE 16

Bacterial Inoculant	Percentage	SEM,
H ₂ O Control	100%	7.4%
<i>Bacillus aryabhattai</i> CAP53, glyphosate sensitive	107.4%	5.8%
<i>Bacillus aryabhattai</i> CAP53, glyphosate-tolerant, GlyR	110.5%	4.6%

Example 2: *Bacillus cereus* family members with plant growth promoting attributes

[0085] *Bacillus mycoides* strain BT155, *Bacillus mycoides* strain EE118, *Bacillus mycoides* strain EE141, *Bacillus mycoides* strain BT46-3, *Bacillus cereus* family member strain EE349, *Bacillus thuringiensis* strain BT013A, and *Bacillus megaterium* strain EE281 were grown in Luria Bertani broth at 37°C and overnight cultures were spun down, media decanted off, and resuspended in equal amount of distilled water. 20 corn seeds were planted for each treatment at a depth of 2.5 cm in loam top soil (Columbia, MO) that was sieved to remove large debris. Seeds were inoculated at planting with 0.5 µl of resuspended bacteria in water mixed into 10 ml of H₂O. Fifty ml of H₂O was sufficient to deliver

the bacteria into the 29 in³ (442.5 cm³) of soil as well as saturate the soil for proper germination of seeds. Plants were grown at temperatures between 65-72°F with 13 hours of light/day, and 5 ml of watering every 3 days. Seedlings were allowed to grow for 2 weeks to analyze emergence and initial outgrowth of plants under described conditions. Identified strains indicated by their proper bacterial identifications and final height data are listed in Table 17.

TABLE 17

Bacterial Inoculant	Avg. Height, cm, Corn	Percentage	SEM,
H2O Control	11.41	100%	.123
<i>B. mycoides</i> EE118	12.43	108.9%	.207
<i>B. mycoides</i> EE141	12.84	112.5%	.231
<i>B. mycoides</i> BT46-3	11.81	103.5%	.089
<i>Bacillus thuringiensis</i> BT013A	12.05	105.6%	.148
<i>Bacillus cereus</i> family member EE128	13.12	114.9%	.159
<i>Bacillus mycoides</i> BT155	12.85	112.6%	.163
<i>Bacillus megaterium</i> EE281	11.99	105.1%	.098

[0086] All plant growth promoting bacteria tested had a beneficial effect on corn height at two weeks under the described conditions. The *Bacillus cereus* family member EE128 strain had the greatest effect in this trial, giving a greater than at 14% boost in corn height. When introducing elements of the present invention, the articles "a", "an", "the" and "said" are intended to mean that there are one or more of the elements. The terms "comprising", "including" and "having" are intended to be inclusive and mean that there may be additional elements other than the listed elements.

[0087] In view of the above, it will be seen that the several objects of the invention are achieved and other advantageous results attained.

REFERENCES CITED IN THE DESCRIPTION

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Patentkrav

1. Inokulum til påføring på planter, plantefrø eller et plantevækstmedium, hvor inokulummet omfatter et landbrugsmæssigt acceptabelt bærestof og en effektiv mængde af en biologisk ren bakteriekultur, hvor bakterierne i bakteriekulturen er *Bacillus thuringiensis*-stamme BT013A
5 (NRRL nr. B-50924).

2. Inokulum ifølge krav 1, hvor inokulummet omfatter en effektiv mængde af en blanding af mindst to bakteriekulturer, idet blandingen omfatter:

Bacillus aryabhattai-stamme CAP53 (NRRL nr. B-50819) og *Bacillus thuringiensis*-
10 stamme BT013A (NRRL nr. B-50924);

Bacillus aryabhattai-stamme CAP56 (NRRL nr. B-50817) og *Bacillus thuringiensis*-
stamme BT013A (NRRL nr. B-50924);

Bacillus flexus-stamme BT054 (NRRL nr. B-50816) og *Bacillus thuringiensis*-
stamme BT013A (NRRL nr. B-50924);

15 *Paracoccus kondratievae*-stamme NC35 (NRRL nr. B-50820) og *Bacillus*
thuringiensis-stamme BT013A (NRRL nr. B-50924);

Bacillus mycoides-stamme BT155 (NRRL nr. B-50921) og *Bacillus thuringiensis*-
stamme BT013A (NRRL nr. B-50924);

Bacillus nealsonii-stamme BOBA57 (NRRL nr. B-50821) og *Bacillus thuringiensis*-
20 stamme BT013A (NRRL nr. B-50924);

Bacillus mycoides-stamme EE118 (NRRL nr. B-50918) og *Bacillus thuringiensis*-
stamme BT013A (NRRL nr. B-50924);

Bacillus subtilis-stamme EE148 (NRRL nr. B-50927) og *Bacillus thuringiensis*-
stamme BT013A (NRRL nr. B-50924);

25 *Bacillus mycoides*-stamme EE141 (NRRL nr. B-50916) og *Bacillus thuringiensis*-
stamme BT013A (NRRL nr. B-50924);

Bacillus mycoides-stamme BT46-3 (NRRL nr. B-50922) og *Bacillus thuringiensis*-
stamme BT013A (NRRL nr. B-50924);

Bacillus cereus-familiemedlemsstamme EE128 (NRRL nr. B-50917) og *Bacillus*
30 *thuringiensis*-stamme BT013A (NRRL nr. B-50924);

Bacillus thuringiensis-stamme BT013A (NRRL nr. B-50924) og *Paenibacillus*
massiliensis-stamme BT23 (NRRL nr. B-50923);

Bacillus thuringiensis-stamme BT013A (NRRL nr. B-50924) og *Bacillus cereus*-familiemedlemsstamme EE349 (NRRL nr. B-50928);

Bacillus thuringiensis-stamme BT013A (NRRL nr. B-50924) og *Bacillus subtilis*-stamme EE218 (NRRL nr. B-50926); eller

5 *Bacillus thuringiensis*-stamme BT013A (NRRL nr. B-50924) og *Bacillus megaterium*-stamme EE281 (NRRL nr. B-50925).

10 3. Inokulum ifølge krav 1 eller 2, hvor inokulummet yderligere omfatter en effektiv mængde af en rhizobakterie.

4. Inokulum ifølge et hvilket som helst af kravene 1-3, hvor det landbrugsmæssigt acceptable bærestof omfatter et dispergeringsmiddel, et overfladeaktivt middel, et tilsætningsstof, vand, et fortykkelsesmiddel, et antiklumpningsmiddel, restnedbrydning, en
15 komposteringsformulering, en granulatpåføring, diatoméjord, en olie, et farvegivende middel, en stabilisator, et konserveringsmiddel, en polymer, et overtræk eller en kombination deraf.

5. Inokulum ifølge krav 4, hvor:

20 tilsætningsstoffet omfatter en olie, en gummi, en harpiks, et ler, en polyoxyethylenglycol, en terpen, et viskøst organisk stof, en fedtsyreester, en sulfateret alkohol, et alkylsulfonat, et jordoliesulfonat, et alkoholsulfat, et natriumalkylbutandiamat, en polyester af natriumthiobutandioat, et proteinholdigt materiale eller en kombination deraf;
25 fortykkelsesmidlet omfatter et langkædet alkylsulfonat af polyethylenglycol, polyoxyethylenoleat eller en kombination deraf;
det overfladeaktive middel omfatter en tung jordolie, et tungt petroleumsdestillat, en polyolfedtsyreester, en polyethoxileret fedtsyreester, en arylalkylpolyoxyethylenglycol, et alkylaminacetat, et alkylarylsulfonat, en polyvalent alkohol, et alkylphosphat eller en kombination deraf; eller
30 antiklumpningsmidlet omfatter et natriumsalt, et calciumcarbonat, diatoméjord eller en kombination deraf.

6. Inokulum ifølge et hvilket som helst af kravene 1-5, hvor det landbrugsmæssigt acceptable bærestof omfatter vermiculit, trækul, karboneringspressemudder fra en sukkerfabrik, risskal, carboxymethylcellulose, tørv, perlit, finsand, calciumcarbonat, mel, alun, en stivelse, talkum, polyvinylpyrrolidon eller en kombination deraf.

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7. Inokulum ifølge et hvilket som helst af kravene 1-6, hvor inokulummet er formuleret som en frøovertræksformulering, en flydende formulering til påføring på planter eller på et plantevækstmedium eller en fast formulering til påføring på planter eller på et plantevækstmedium.

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8. Inokulum ifølge krav 7, hvor:

frøovertræksformuleringen er en vandig eller oliebaseret opløsning til påføring på frø eller en pulver- eller en granulatformulering til påføring på frø;
den flydende formulering til påføring på planter eller på et plantevækstmedium er i en koncentreret formulering eller en brugsklar formulering; eller
den faste formulering til påføring på planter eller på et plantevækstmedium er en granulatformulering eller et pulvermiddel.

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9. Inokulum ifølge et hvilket som helst af kravene 1-8, hvor inokulummet yderligere omfatter et gødningsstof, et gødningsmateriale med mikronæringsstoffer, et insekticid, et herbicid, en plantevækstændring, et fungicid, et molluskicid, et algicid, et bakteriepodestof, et svampepodestof eller en kombination deraf.

20

10. Inokulum ifølge krav 9, hvor:

gødningsstoffet omfatter ammoniumsulfat, ammoniumnitrat, ammoniumsulfatnitrat, ammoniumchlorid, ammoniumbisulfat, ammoniumpolysulfid, ammoniumthiosulfat, vandig ammoniak, vandfri ammoniak, ammoniumpolyphosphat, aluminiumsulfat, calciumnitrat, calciumammoniumnitrat, calciumsulfat, kalcineret magnesit, calcitkalksten, calciumoxid, calciumnitrat, dolomitkalksten, hydratkalk, calciumcarbonat, diammoniumphosphat, monoammoniumphosphat, magnesiumnitrat, magnesiumsulfat, kaliumnitrat, kaliumchlorid, kaliummagnesiumsulfat, kaliumsulfat, natriumnitrater, magnesiakalksten, magnesia, urinstof, urinstofformaldehyder, urinstofammoniumnitrat, svovlovertrukket urinstof, polymerovertrukket urinstof,

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isobutylidendiurea, $K_2SO_4 \cdot 2MgSO_4$, kainit, sylvinit, kieserit, bittersalte, elementært svovl, mergel, formalede østersskaller, fiskemel, oliekgager, fiskegødning, blodmel, råphosphat, superphosphater, slagge, benmel, træaske, gødning, flagermuseguano, tørvemos, kompost, grønsand, bomuldsfrømel, fjermel, krabbemel, fiskeemulsion

5 eller en kombination deraf;

gødningsmaterialet med mikronæringsstoffer omfatter borsyre, et borat, en borfritte, kobbersulfat, en kobberfritte, et kobberchelate, et natriumtetraaboratdecahydrat, et jernsulfat, et jernoxid, jernammoniumsulfat, en jernfritte, et jernchelate, et mangansulfat, et manganoxid, et manganchelate, et manganchlorid, en manganfritte, et

10 natriummolybdat, molybdensyre, et zinksulfat, et zinkoxid, et zinkcarbonat, en zinkfritte, zinkphosphat, et zinkchelate eller en kombination deraf;

insekticidet omfatter et organophosphat, et carbamat, et pyrethroid, et acaricid, et alkylphthalat, borsyre, et borat, et fluorid, svovl, et halogenaromatisk substitueret urinstof, en carbonhydridester, et biologisk baseret insekticid eller en kombination

15 deraf;

herbicidet omfatter en chlorphenoxyforbindelse, en nitrophenolforbindelse, en nitrocresolforbindelse, en dipyridylforbindelse, et acetamid, en alifatisk syre, et anilid, et benzamid, en benzoesyre, anissyre, et benzonitril, benzothiadiazinondioxid, et thiocarbamat, et carbamat, et carbanilat, chlorpyridinyl, en

20 fluordinitrotoluidinforbindelse, isoxazolidinon, nicotinsyre, isopropylamin, oxadiazolinon, et phosphat, et phthalat, en picolinsyreforbindelse, et triazin, et triazol, et uracil, endothall, natriumchlorat eller en kombination deraf;

fungicidet omfatter en substitueret benzen, et thiocarbamat, et ethylen-bis-dithiocarbamat, et thiophthalidamid, en kobberforbindelse, en organisk

25 kviksølvforbindelse, en organisk tinforbindelse, en cadmiumforbindelse, anilazin, benomyl, cyclohexamid, dodin, etridiazol, iprodion, metlaxyl, thiamimefon, triforin eller en kombination deraf;

svampepodestoffet omfatter et svampepodestof af familien Glomeraceae, et svampepodestof af familien Clavariaceae, et svampepodestof af familien Gigasporaceae, et svampepodestof af familien Acaulosporaceae, et svampepodestof af familien Sacculosporaceae, et svampepodestof af familien Entrophosporaceae, et svampepodestof af familien Pacidsporaceae, et svampepodestof af familien Diversisporaceae, et svampepodestof af familien Paraglomeraceae, et svampepodestof af familien Archaeosporaceae, et svampepodestof af familien Geosiphonaceae, et

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- svampepodestof af familien Ambisporaceae, et svampepodestof af familien Scutellosporaceae, et svampepodestof af familien Dentiscultataceae, et svampepodestof af familien Racocetraceae, et svampepodestof af phylum Basidiomycota, et svampepodestof af phylum Ascomycota, et svampepodestof af phylum Zygomycota eller en kombination deraf; eller
- 5 bakteriepodestoffet omfatter et bakteriepodestof af genus *Rhizobium*, et bakteriepodestof af genus *Bradyrhizobium*, et bakteriepodestof af genus *Mesorhizobium*, et bakteriepodestof af genus *Azorhizobium*, et bakteriepodestof af genus *Allorhizobium*, et bakteriepodestof af genus *Sinorhizobium*, et
- 10 bakteriepodestof af genus *Kluyvera*, et bakteriepodestof af genus *Azotobacter*, et bakteriepodestof af genus *Pseudomonas*, et bakteriepodestof af genus *Azospirillum*, et bakteriepodestof af genus *Bacillus*, et bakteriepodestof af genus *Streptomyces*, et bakteriepodestof af genus *Paenibacillus*, et bakteriepodestof af genus *Paracoccus*, et bakteriepodestof af genus *Enterobacter*, et bakteriepodestof af
- 15 genus *Alcaligenes*, et bakteriepodestof af genus *Mycobacterium*, et bakteriepodestof af genus *Trichoderma*, et bakteriepodestof af genus *Gliocladium*, et bakteriepodestof af genus *Glomus*, et bakteriepodestof af genus *Klebsiella* eller en kombination deraf.
- 20 **11.** Fremgangsmåde til stimulering af plantevækst, hvilken fremgangsmåde omfatter påføring af inokulummet ifølge et hvilket som helst af kravene 1-10 på en plante, et plantefrø eller et plantevækstmedium.
- 12.** Fremgangsmåde ifølge krav 11, der omfatter:
- 25 påføring af bakteriekulturen eller inokulummet på plantevækstmediet;
- påføring af bakteriekulturen eller inokulummet på plantevækstmediet inden, samtidigt med eller efter såning, plantning eller lægning af frø, frøplanter, stiklinger, løg eller planter i plantevækstmediet;
- påføring af bakteriekulturen eller inokulummet på plantblade, -rødder eller -stilke;
- 30 eller
- påføring af bakteriekulturen eller inokulummet på plantefrø.

13. Plantefrø overtrukket med inokulummet ifølge et hvilket som helst af kravene 1-10.

14. Kit til stimulering af plantevækst, hvilket kit omfatter inokulummet ifølge et hvilket som helst af kravene 1-10 og instruktioner til påføring af inokulummet på planter, plantefrø eller

5 et plantevækstmedium.