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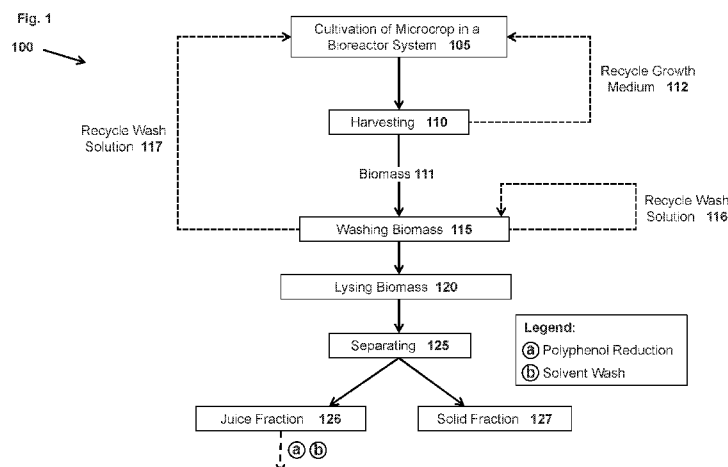
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(54) Title: METHODS AND SYSTEMS FOR EXTRACTING A POLYSACCHARIDE PRODUCT FROM A MICROCROP AND COMPOSITIONS THEREOF



(57) Abstract: The present disclosure relates, according to some embodiments, to methods and systems for purifying proteins, carbohydrate rich products, and polysaccharide products from a microcrop (e.g., photosynthetic aquatic species) and compositions thereof. For example, the present disclosure relates, in some embodiments to methods and systems for extracting proteins, dry biocrude, carbohydrate-rich meal, and polysaccharide products from Lemna.

METHODS AND SYSTEMS FOR EXTRACTING A POLYSACCHARIDE PRODUCT FROM A MICROCROP AND COMPOSITIONS THEREOF

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Patent Application No. 62/189,040 filed on July 6, 2015 and U.S. Provisional Patent Application No. 62/240,279
5 filed on October 12, 2015 which are incorporated herein by reference in their entirety as set forth in full.

FIELD OF THE DISCLOSURE

The present disclosure relates, in some embodiments, to methods and systems for extracting a polysaccharide product (e.g., apiogalacturonan, oligogalacturonide,
10 polygalacturonide) from a microcrop (e.g., photosynthetic aquatic species) and compositions thereof.

BACKGROUND OF THE DISCLOSURE

An ever-increasing global population continues to fuel a plethora of sustainability
15 concerns including sufficient and affordable access to carbohydrate sources for human and animal consumption among other uses. Additionally, concerns relating to climate change and fossil fuel consumption continue to drive innovators to develop and improve both carbohydrate sources and extraction efficiency. Moreover, water conservation concerns—particularly in equatorial and arid regions—are a driving factor in identifying suitable
20 processes and sources for the production of carbohydrates.

SUMMARY

Accordingly, a need has arisen for improved methods and systems for the production of carbohydrate-rich products and polysaccharide products. A need has arisen for improved
25 methods and systems for the production of carbohydrate-rich products in a manner requiring decreased water and/or energy expenditures. Additionally, a need has arisen for efficiently producing polysaccharide products comprising polygalacturonides, apiogalacturonan and/or oligogalacturonide.

The present disclosure relates, in some embodiments, to methods and systems for
30 extracting a polysaccharide product (e.g., apiogalacturonan, oligogalacturonide, polygalacturonide) from *Lemna*. In some embodiment, the present disclosure relates to

compositions of a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide) isolated from a microcrop (*e.g.*, photosynthetic aquatic species).

The present disclosure relates, in some embodiments to a method of treating a biomass comprising a microcrop, the method comprising: (a) lysing the biomass to form a
5 lysed biomass; (b) separating the lysed biomass to generate a juice fraction and a solid fraction; (c) separating the solid fraction to generate a first solid and a second juice; and (d) extracting the first solid with an extraction solution to form a polysaccharide extract. According to some embodiments, an extraction solution may comprise an acid solution, an oxalic acid solution, an oxalate solution, or combinations thereof. A polysaccharide extract
10 may comprise an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof. A polysaccharide extract, according to some embodiments, may comprise at least 30% (w/v) of an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or the combination thereof.

In some embodiments, a method may further comprise concentrating a polysaccharide
15 extract to form a polysaccharide product. A polysaccharide product may comprise an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof. According to some embodiments, concentrating a polysaccharide extract may consist of a process selected from evaporation, precipitation, dialysis, filtration, ultrafiltration, nanofiltration, reverse osmosis filtration, centrifugation, dewatering, and combinations
20 thereof. A polysaccharide product, according to some embodiments, may comprise at least 80% (w/v) of an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or the combination thereof.

In some embodiments, a method of treating a biomass comprising a microcrop may comprise decoloring a biomass. A method, in some embodiments, may comprise
25 concentrating a polysaccharide extract to form a polysaccharide product, the polysaccharide comprising an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof. Concentrating a polysaccharide extract, in some embodiments, may consist of a process selected from evaporation, precipitation, dialysis, filtration, ultrafiltration, nanofiltration, reverse osmosis filtration, centrifugation, dewatering, and
30 combinations thereof. A polysaccharide product, according to some embodiments, may comprise at least 80% (w/v) of an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or the combination thereof.

According to some embodiments, a method may comprise washing a biomass with at least one of a first wash solution, a second wash solution, a third wash solution, or a combination thereof. A method, in some embodiments, may comprise separating a juice fraction to generate a first juice and a first cake, and filtering the first juice to generate a soluble microcrop protein and a reject stream. In some embodiments, a method may comprise (1) separating a first cake; (2) separating a second juice, or (3) separating the first cake and separating the second juice, in each case, to generate a third juice and a second cake; and combining the second cake with a first solid prior to extracting the first solid with an extraction solution to form a polysaccharide extract. According to some embodiments, a method may comprise subjecting a first juice to a polyphenol reduction process to generate a soluble microcrop protein having a reduced concentration of at least one polyphenol. According to some embodiments, a microcrop may be *Lemna*.

According to some embodiments, a polysaccharide product may be produced from a biomass comprising a microcrop by a process comprising: lysing the biomass to form a lysed biomass; separating the lysed biomass to generate a juice fraction and a solid fraction, separating the solid fraction to generate a first solid and a second juice, extracting the first solid with an extraction solution to form a polysaccharide extract, and concentrating the polysaccharide extract to form a polysaccharide product. An extraction solution, in some embodiments, may comprise an acid solution, an oxalic acid solution, an oxalate solution, or combinations thereof. A polysaccharide product may comprise a apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof, according to some embodiments. In some embodiments, concentrating a polysaccharide extract may consist of a process selected from evaporation, precipitation, dialysis, filtration, ultrafiltration, nanofiltration, reverse osmosis filtration, centrifugation, dewatering, and combinations thereof. A polysaccharide product, in some embodiments, may comprise at least 80% (w/v) of an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof. In some embodiments, a process may comprise decoloring a biomass. In some embodiments, a microcrop may comprise *Lemna*.

According to some embodiments, a polysaccharide product from a biomass comprising a microcrop, may comprise at least about 85 wt. % of an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof. In some embodiments, a polysaccharide product may comprise a protein content of about 5 wt. % or less. A polysaccharide product, according to some embodiments, may comprise a fat content of

lower than about 1 wt. %. According to some embodiments, a polysaccharide product may comprise an ash content of lower than about 10 wt. %.

BRIEF DESCRIPTION OF THE DRAWINGS

5 The file of this patent contains at least one drawing executed in color. Copies of this patent with color drawing(s) will be provided by the Patent and Trademark Office upon request and payment of the necessary fee.

Some embodiments of the disclosure may be understood by referring, in part, to the present disclosure and the accompanying drawings, wherein:

10 FIGURE 1 is a flow diagram illustrating a system for cultivating, harvesting, and processing a microcrop for the production of protein concentrate and/or carbohydrate rich products according to a specific example embodiment of the disclosure;

FIGURE 2A is a flow diagram illustrating a process for producing a protein concentrate and/or carbohydrate rich products from a biomass according to a specific
15 example embodiment of the disclosure;

FIGURE 2B is a flow diagram illustrating a process for producing a protein concentrate and/or carbohydrate rich products from a biomass according to a specific example embodiment of the disclosure;

FIGURE 3A is a flow diagram illustrating a process for producing a protein
20 concentrate and/or carbohydrate rich products from a biomass according to a specific example embodiment of the disclosure;

FIGURE 3B is a flow diagram illustrating a process for producing a protein concentrate and/or carbohydrate rich products from a biomass according to a specific example embodiment of the disclosure;

25 FIGURE 4 is a flow diagram illustrating a process for continuously producing a protein concentrate and/or carbohydrate rich products from a biomass according to a specific example embodiment of the disclosure;

FIGURE 5A is a flow diagram illustrating a process for producing a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide) and/or a protein
30 concentrate from a biomass according to a specific example embodiment of the disclosure;
and

FIGURE 5B is a flow diagram illustrating a process for producing a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide) and/or a protein concentrate from a biomass according to a specific example embodiment of the disclosure.

DETAILED DESCRIPTION

The present disclosure relates to compositions, systems, and methods for producing a protein concentrate (*e.g.*, soluble protein, dry protein concentrate) and/or carbohydrate rich products and/or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide) from a microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species). For example, a method may comprise growing, harvesting, and/or separating a microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) for the production of a protein concentrate (*e.g.*, soluble protein, a dry protein concentrate) and/or a carbohydrate rich product and/or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide) according to specific example embodiments of the disclosure. A method may be performed, in some embodiments, in a series of steps, one or more of which may be repeated. For example, a method may comprise a single cycle (*e.g.*, no step is repeated) resulting in the production of protein concentrate (*e.g.*, soluble protein, dry protein concentrate) and/or carbohydrate rich products and/or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide). In some embodiments, a method may comprise multiple cycles (*e.g.*, first portion, second portion) or a continuous process for the production of protein concentrate (*e.g.*, soluble protein, dry protein concentrate) and/or carbohydrate rich products and/or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide) such that products, intermediates and/or byproducts of an earlier cycle of the process may be recycled into one or more subsequent cycles of the process.

Microcrop

In some embodiments, a microcrop may comprise a single aquatic species including, for example, a single photosynthetic aquatic species (*e.g.*, *Lemna* species, *Salvinia* species). A microcrop may include species of *Lemna* (*e.g.*, duckweed), *Spirodela*, *Landoltia*, *Wolffiella*, *Salvinia* (*e.g.*, floating fern), *Wolffia* (*e.g.*, watermeal), *Azolla* (*e.g.*, mosquito fern), *Pistia* (*e.g.*, water lettuce), or any combination thereof. According to some embodiments, a microcrop may be a species of *Lemna*, for example, *Lemna minor*, *Lemna obscura*, *Lemna minuta*, *Lemna gibba*, *Lemna valdiviana*, or *Lemna aequinoctialis*. A microcrop may

comprise, according to some embodiments, a combination of two or more types of aquatic specie including, for example, a combination of two or more types of photosynthetic aquatic species. In some embodiments, a microcrop may be selected from a local photosynthetic aquatic species based on identified compositional and growth characteristics that have developed within the local environmental conditions. Local species may out-compete other species in open ponds or bioreactors based on their adaptation to the local environmental conditions. A microcrop, in some embodiments, may be adjusted in response to seasonal variations in temperature and light availability.

A microcrop may have characteristics that are advantageous in comparison to other aquatic species (*e.g.*, rapid growth rate; reduced nutritional requirements; ease of harvesting and/or processing; enhanced amino acid profile; enhanced palatability; reduced evapotranspiration rate; increased protein composition).

For example, *Lemna* is a genus of free-floating aquatic plants from the Lemnaceae family (*e.g.*, duckweed) that grow rapidly. *Lemna* protein has an essential amino acid profile that more closely resembles animal protein than most other plant proteins. Table 1 shows a typical essential amino acid compositional profile of *Lemna* protein. Additionally, *Lemna* provides high protein yields, with freshly harvested *Lemna* containing up to about 43% protein by dry weight. Furthermore, compared with most other plants, *Lemna* leaves have a low fiber content (*e.g.*, about 5% to about 15% in dry matter) and are highly digestible, even for monogastric animals. This contrasts with the compositions of many crop species (*e.g.*, soy beans, rice, maize) which have fiber contents of approximately 50% and low digestibility.

TABLE 1: Essential Amino Acid Profile of Lemna Protein Concentration	
Essential Amino Acid	Protein (g/ 100g)
Lysine	5.9
Leucine	9.7
Isoleucine	5.1
Methionine	2.4
Phenylalanine	6.3
Threonine	4.4
Tryptophan	2.0
Valine	6.3
Histidine	2.7
Arginine*	6.8

*Conditionally essential amino acid

Cultivation of a Microcrop

In some embodiments a microcrop may be asexually propagated (*e.g.*, cultivated) by contacting the microcrop with an aqueous nutrient composition under conditions that permit expansion. A microcrop may be cultivated in a bioreactor system, according to some embodiments. A bioreactor system may contain a growth medium. In some embodiments a growth medium may comprise water and/or a nutrient composition. A growth medium (*e.g.*, water) may be provided in and/or added to a bioreactor (*e.g.*, a pond) and may be maintained at a desired set-point level (*e.g.*, specific volume), according to some embodiments. A bioreactor system, in some embodiments, may be configured to collect rainfall and/or to intake water from a source of ground, surface, or recycled water (*e.g.*, storm water, recycled water) or any other suitable water source. According to some embodiments, a bioreactor system may further comprise an additional storage container (*e.g.*, container or pond) for excess growth medium. A bioreactor system may be configured to insert additional nutrients (*e.g.*, nitrogen, phosphorus, potassium) or gases (*e.g.*, oxygen; carbon dioxide) at specified time indicators or in response to sensor readings. In some embodiments, one or more smaller bioreactors (*e.g.*, pond) may be designed and sized to adequately serve as "feeder" bioreactors to a larger bioreactor. Smaller bioreactors, in some embodiments, may be first inoculated and grown to high density at which point they may optimally seed a larger bioreactor in a manner that supports faster growth.

In some embodiments, a bioreactor system may comprise a monitoring system. A monitoring system may be configured to display and/or provide one or more user alerts regarding bioreactor condition(s) (*e.g.*, nutrient concentrations, pH, dissolved oxygen levels, growth medium levels, microcrop distribution, flow rate, temperature) and/or adjust operating
5 conditions (*e.g.*, growth medium flow rate and /or timing and/or quantity of nutrient addition; “feeder” microcrop addition; oxygen or carbon dioxide addition), in some embodiments. Adjustments may be made continuously, semi-continuously, periodically, intermittently, as needed, at set or variable times, or any other interval. In some embodiments, adjustments may be selected to optimize growth rates and/or yield of the aquatic species. For example, a
10 microcrop species may be grown in large-scale, open bioreactors with monitoring systems configured to adjust the introduction of materials (*e.g.*, fresh or recycled water, fresh or recycled growth media) based on, for example, exposure to light, which may thereby regulate nutrient consumption rates.

A bioreactor system may comprise, in some embodiments, a single container in which
15 a microcrop may be cultivated. In some embodiments, the bioreactor system may comprise multiple cultivation containers that may be connected, partially connected, or disconnected. A bioreactor (*e.g.*, a pond), in some embodiments, may be an earthen basin with at least one embankment (*e.g.*, made of compacted dirt removed from the interior bottom of the bioreactor). According to some embodiments the bioreactor may be an artificial container
20 (*e.g.*, metal, plastic, resin). A bioreactor system may comprise an open bioreactor, a closed bioreactor, a semi-open bioreactor, or any combination thereof. In some embodiments, a bioreactor system may be configured to divide the container(s) into channels or cells. A bioreactor system may be configured to permit a flow of growth medium, in some embodiments. A bioreactor system, in some embodiments, may include a propulsion system
25 (*e.g.*, paddle wheels, bubbling, submerged or surface water jets, submerged mixers) and/or a recirculation system. In some embodiments, a bioreactor system may be configured to adjust a flow rate of a growth medium (*e.g.*, to redistribute nutrient concentrations or microcrop growth patterns).

In some embodiments a bioreactor system may be open (*e.g.*, in a horizontal plane
30 relative to the ground) such that a growth medium contained within a bioreactor container (*e.g.*, a serpentine raceway) and/or a microcrop growing on a top surface of the growth medium may be exposed to a wind initiating from an exterior of the bioreactor container. A bioreactor system, according to some embodiments, may be partially open (*e.g.*, in a

horizontal plane relative to the ground) with at least 90% or at least 80%, or at least 70%, or at least 60%, or at least 50%, or at least 40%, or at least 30%, or at least 20%, or at least 10% of the top surface of the contained culture media being open. A top surface may be open, according to some embodiments, where the surface is substantially free (*e.g.*, free) of any covering or other barrier, where the surface is directly exposed to ambient weather conditions, where there is substantially no membrane, glass, cover or other barrier (whether or not such barrier has pores or apertures) between the surface and the atmosphere, and/or where ambient atmosphere is the only occupant of the space immediately and directly above the surface for a distance of at least about 1 meter above the surface.

A bioreactor system, in some embodiments, may monitor and adjust a thickness and distribution of a microcrop mat. For example, when a microcrop reaches a specified thickness or distribution a bioreactor system may initiate harvest procedures. In some embodiments, a minimum thickness of a microcrop may be maintained such that a desired evapotranspiration rate of a growth medium within a bioreactor system may be maintained. A minimum thickness of a microcrop may be maintained, in some embodiments, such that less sunlight is capable of penetrating a surface of a growth medium (*i.e.*, reducing a growth potential of submerged aquatic species such as algae).

Harvesting of a Microcrop

A microcrop may be harvested in whole or in part at any desired time(s). For example, a microcrop may be harvested at one or more specific times, at regular or irregular intervals and/or continuously. Selection of harvest time(s) and/or intervals may be based on environmental conditions (*e.g.*, precipitation, relative humidity, temperature range, average, low or high threshold and/or light intensity, wavelength range, duration of exposure) and/or the microcrop exhibiting one or more desired characteristics (*e.g.*, mat thickness, mat distribution, maturation). Harvesting a microcrop may be manual or automated. In some embodiments, an automated skimmer system may collect a microcrop from a bioreactor system and transfer a harvested microcrop (*e.g.*, via a pumping system) onto an inclined vibrating screen to separate a biomass from growth medium and debris. A microcrop, in some embodiments, may be harvested by vacuum skimming the microcrop from a bioreactor system through a stationary or mobile screen filter. According to some embodiments, a biomass slurry, including a harvested microcrop (*e.g.*, *Lemna*) and a growth medium (*e.g.*,

water), may be conveyed to an inclined vibrating screen where a biomass (*e.g.*, microcrop) may be separated from the growth medium.

During harvesting, a separated growth medium may be recycled back into a bioreactor system or to an additional storage container (*e.g.*, container or pond), according to some
5 embodiments. In some embodiments, at least about 40%, or at least about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, or at least about 95% of a growth medium (*e.g.*, water) separated from a biomass may be recycled for further use in cultivating, harvesting, and/or processing a microcrop.

10 Decoloring a Biomass

According to some embodiments, processing a microcrop or biomass (*e.g.*, first portion, second portion) may include a decoloring procedure to remove at least some of a chlorophyll component of the microcrop or biomass. Decoloring a biomass may increase
15 purity of a protein product and/ or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide). In some embodiments, decoloring a biomass may reduce a fat content of a protein concentrate (*e.g.*, soluble protein, dry protein concentrate) and/or carbohydrate rich products and/or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide).

Decoloring a biomass, in some embodiments, may comprise blanching a biomass at a
20 temperature of about 70° C, or about 75° C, or about 80° C, or about 85° C, or about 90° C, or about 95° C, or about 100° C for about 1 min., or about 3 min., or about 5 min., or about 7 min., or about 10 min., or about 15 min., or about 20 min., or about 25 min., or about 30 min., or about 35 min., or about 40 min., or about 45 min., or about 50 min., or about 55 min., or about 1 hour. In some embodiment, decoloring a biomass may comprise exposing (*e.g.*,
25 submerging, spraying, dripping) at least one surface of a biomass to a solvent solution (*e.g.*, ethanol, methanol, acetone, hexane, dichloromethane, propanol, isopropanol, glycerol). A solvent solution, in some embodiments, may be combined with a biomass (*e.g.*, first portion, second portion) to form a slurry. In some embodiments a solvent solution may include one or more alcohols (*e.g.*, ethanol, methanol, propanol, isopropanol, glycerol), acetone, hexane, or
30 ketones. A solvent solution may comprise at least about 10% (v/v), or at least about 20% (v/v), or at least about 30% (v/v), or at least about 40% (v/v), or at least about 50% (v/v), or at least about 60% (v/v), or at least about 70% (v/v), or at least about 80% (v/v), or at least about 90% (v/v) of one or more alcohols (*e.g.*, ethanol, methanol), acetone, hexane, or

ketones, or any combination thereof. According to some embodiments, a decoloring procedure may include exposing (*e.g.*, submerging, spraying, dripping, slurry) at least one surface of a biomass to a solvent solution for at least: about 5 sec., or about 15 sec., or about 30 sec., or about 45 sec., or about 1 min., or about 2 min., or about 3 min., or about 5 min., or about 10 min., or about 20 min., or about 30 min., or about 40 min., or about 50 min., or about 1 hour, or about 2 hours, or about 3 hours, or about 4 hours, or about 5 hours, or about 6 hours, or about 12 hours, or about 24 hours. A decoloring procedure, in some embodiments, may include moving (*e.g.*, agitating, stirring, propelling) at least a portion of a solvent solution at a specified time, or intermittently, or continually.

Washing a Biomass

In some embodiments, processing a microcrop or biomass (*e.g.*, first portion, second portion) may include a wash procedure to remove excess growth medium, a solvent solution, debris, contaminants, microorganisms, and/or toxins. Washing a biomass may increase a purity and/or yield of a protein product and/ or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide). A wash procedure may disinfect and/or disinfest a biomass, reducing or removing bacteria, fungi, viruses, insects, and any combination thereof which are on or around the surfaces of the biomass. In some embodiments a wash procedure may be performed by exposing (*e.g.*, submerging, spraying) at least one surface of a biomass to a wash solution (*e.g.*, water, growth medium, antimicrobial solution). A wash solution, in some embodiments, may be combined with a biomass (*e.g.*, first portion, second portion) to form a slurry.

In some embodiments, a wash solution may comprise any desired portion of recycled fluid. For example, a wash solution may comprise at least about 10% (v/v), or at least about 20% (v/v), or at least about 30% (v/v), or at least about 40% (v/v), or at least about 50% (v/v), or at least about 60% (v/v), or at least about 70% (v/v), or at least about 80% (v/v), or at least about 90% (v/v) fluid recycled from another stage of the process (*e.g.*, recycled wash solution FIGURE 1, **116**; a reject stream from filtration (*e.g.*, FIGURE 2A, **252**; FIGURE 4, **452**, **457**)). In some embodiments a wash solution may be an aqueous solution or solvent. A wash solution, according to some embodiments, may contain one or more antimicrobials, deinfestation compounds, fatty acids, alcohols, chlorine, oxidizing compounds, and any combination thereof (*e.g.*, ozonated water).

According to some embodiments a wash solution may be applied at an elevated temperature and/or high pressure. A wash solution, in some embodiments, may remain in contact with a biomass for at least about 1 second, or for at least about 5 seconds, or for at least about 10 seconds, or for at least about 20 seconds, or for at least about 30 seconds, or for at least about 1 minute, or for at least about 5 minutes. In some embodiments, a second wash solution (*e.g.*, water, ozonated water, a recycled wash solution (FIGURE 1, **116**)) may be applied to a biomass. A third wash solution (*e.g.*, water, ozonated water, recycled wash solution) may be applied to a biomass, in some embodiments. A composition of a first wash solution, a second wash solution, and a third wash solution may be the same or different from one another. In some embodiments a first wash solution may be or may comprise a reject stream from a filtration process (*e.g.*, FIGURE 2A, **252**; FIGURE 4, **452**, **457**), a second wash solution may be water, and a third wash solution may be ozonated water. Some or all of a wash solution (*e.g.*, a first, second, and/or third wash solution), in some embodiments, may be separated from a biomass (*e.g.*, using an inclined screen or vibratory screen).

In some embodiments, some or all of a wash solution, a first wash solution, a second wash solution, and/or third wash solution may be collected and reused/recycled (*e.g.*, FIGURE 1, **116/117**). At least about 40%, or at least about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, or at least about 95% of a wash solution, a first wash solution, a second wash solution, and/or third wash solution (*e.g.*, water) separated from the biomass may be recycled for future use (*e.g.*, recycled wash solution (FIGURE 1, **116**), used as growth medium in the bioreactor system (FIGURE 1, **117**)), according to some embodiments.

A wash solution, a first wash solution, a second wash solution, and/or third wash solution, in some embodiments, may have or may be adjusted to have any desired pH. For example, the pH of a wash solution, a second wash solution, and/or a third wash solution may be neutral or basic (*e.g.*, about 7.0, or about 7.5, or about 8.0, or about 8.5, or about 9.0, or about 9.5, or about 10.0). According to some embodiments, the pH of a wash solution, a first wash solution, a second wash solution, and/or a third wash solution may be from about 7.0 to about 7.5, or from about 7.5 to about 8.0, or from about 8.0 to about 8.5, or from about 8.5 to about 9.0, or from about 9.0 to about 9.5, or from about 9.5 to about 10.0. The pH of a wash solution, a first wash solution, a second wash solution, and/or a third wash solution may be from about 7.0 to about 10.0, or from about 7.0 to about 9.5, or from about 7.0 to about 9.0,

or from about 7.0 to about 8.5, or from about 7.0 to about 8.0, or from about 7.0 to about 7.5, in some embodiments.

A wash solution (*e.g.*, a first, second, and/or third wash solution) may have a temperature below room temperature (*e.g.*, about 12° C) at the time of use. Cooling a wash solution, and thereby a microcrop, may improve product recovery efficiency and/or decrease proteolytic activity of product-degrading enzymes. In some embodiments, a wash solution (*e.g.*, a first, a second, and/or a third wash solution) may have a temperature below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C at the time of use. A wash solution (*e.g.*, a first, second, and/or third wash solution) may have a temperature at the time of use between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or 15° C and about 25° C, or between about 20° C and about 30° C, in some embodiments.

In some embodiments, a wash solution (*e.g.*, a first, a second, and/or a third wash solution) may have a temperature above room temperature (*e.g.*, about 50° C) at the time of use. Heating a wash solution, and thereby a microcrop, may improve product recovery efficiency, decrease activity of product-degrading enzymes (*e.g.*, denature proteolytic enzymes), and/or decrease microbial contamination (*e.g.*, pasteurization). In some embodiments, a wash solution (*e.g.*, a first, a second, and/or a third wash solution) at the time of use may have a temperature above about 20° C, or above about 25° C, or above about 30° C, or above about 35° C, or above about 40° C, or above about 45° C, or above about 50° C, or above about 55° C, or above about 60° C, or above about 65° C, or above about 70° C, or above about 75° C, or above about 80° C, or above about 85° C, or above about 90° C, or above about 95° C, or above about 100° C. A wash solution (*e.g.*, a first, a second, and/or a third wash solution) at the time of use may have a temperature between about 40° C and about 50° C, or between about 45° C and about 55° C, or between about 50° C and about 60° C, in some embodiments. According to some embodiments, a wash solution (*e.g.*, a first, second, and/or third wash solution) at the time of use may have a temperature between about 75° C and about 80° C, or between about 80° C and about 85° C, or between about 85° C and about 90° C, or between about 90° C and about 95° C, or between about 95° C and about 100° C. In some embodiments, a wash solution (*e.g.*, a first, second, and/or third wash solution) may have a temperature between about 50° C and about 80° C, or between about 55° C and

about 85° C, or between about 60° C and about 90° C, or between about 65° C and about 95° C, or between about 70° C and about 100° C at the time of use.

Lysing a Biomass

5 According to some embodiments a biomass (*e.g.*, washed or un-washed) may be lysed to form a lysed biomass (*e.g.*, first portion, second portion). As used herein, lysing may include mechanical, chemical, and/or ultrasonic (*e.g.*, sonication) procedures that disturb the organization of the organism on the level of individual cells or multicellular structures. Lysing may include, in some embodiments, rendering carbohydrates, proteins, and
10 micronutrients present in a microcrop more available for downstream processing to purified protein, carbohydrate-containing materials, and/or micronutrient-containing fluids. According to some embodiments, lysing may be achieved using a combination of mechanical, chemical, and/or ultrasonic (*e.g.*, sonication) methods.

In some embodiments, lysing may be performed at temperatures below room
15 temperature. Lysing a microcrop at a lower temperature may improve yields, for example, by limiting or decreasing undesired enzymatic activity (*e.g.*, proteolytic activity). Lysing may be performed, in some embodiments, at a temperature below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C.

20 A lysing fluid (*e.g.*, water, recycled water, reverse osmosis water) may be added to a biomass, washed or unwashed, before or during lysing according to some embodiments. For example, at least about 10%, or at least about 20%, or at least about 30%, or at least about 40%, or at least about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90% of lysing fluid may be water generated as the result of reverse
25 osmosis/nanofiltration of a filtration product (*e.g.*, FIGURE 3, **362**; FIGURE 4, **462**). In some embodiments a lysing fluid may be at a temperature below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. A lysing fluid, in some embodiments, may include buffers, protease inhibitors, anti-microbial agents, chelators (*e.g.*, EDTA), reducing agents, or
30 any combination thereof.

According to some embodiments, lysing may be performed at temperatures above room temperature (*e.g.*, about 40° C), for example, to enhance cellulosic breakdown and/or denature undesired enzymes (*e.g.*, proteolytic enzymes). Lysing may be performed at a

temperature above about 30° C, or above about 35° C, or above about 37° C, or above about 40° C, in some embodiments.

In some embodiments, a lysing fluid may have or may be adjusted to have any desired pH. For example, a lysing fluid may have a neutral or basic pH (*e.g.*, about 7.0, or about 7.5, or about 8.0, or about 8.5, or about 9.0, or about 9.5, or about 10.0). According to some
5 embodiments, a lysing fluid may have a pH of from about 7.0 to about 7.5, or from about 7.5 to about 8.0, or from about 8.0 to about 8.5, or from about 8.5 to about 9.0, or from about 9.0 to about 9.5, or from about 9.5 to about 10.0. The pH of a lysing fluid may be from about 7.0 to about 10.0, or from about 7.0 to about 9.5, or from about 7.0 to about 9.0, or from about
10 7.0 to about 8.5, or from about 7.0 to about 8.0, or from about 7.0 to about 7.5, in some embodiments.

Lysing may be performed with a biomass having any desired pH. For example, a biomass may have or may be adjusted to have a neutral or basic pH (*e.g.*, about 7.0, or about 7.5, or about 8.0, or about 8.5, or about 9.0, or about 9.5, or about 10.0). According to some
15 embodiments, lysing may be performed with a biomass having a pH from about 7.0 to about 7.5, or from about 7.5 to about 8.0, or from about 8.0 to about 8.5, or from about 8.5 to about 9.0, or from about 9.0 to about 9.5, or from about 9.5 to about 10.0. In some embodiments, lysing may be performed with a biomass having a pH from about 7.0 to about 10.0, or from about 7.0 to about 9.5, or from about 7.0 to about 9.0, or from about 7.0 to about 8.5, or from
20 about 7.0 to about 8.0, or from about 7.0 to about 7.5, in some embodiments. Adjusting or maintaining a pH value of a biomass prior to or during a lysing procedure may increase protein yield (*e.g.*, increased protein solubility).

Lysing may include, for example, chopping, shredding, smashing, pressing, tearing, ultrasonic treatment (*e.g.*, sonication), lysis by osmotic pressure, chemical treatments that
25 degrade biological structures, or any combination thereof. In some embodiments, lysing is achieved in a mechanical way (also referred to as milling), for example, by milling, grinding, or shredding a biomass to generate a lysed biomass. A lysing process may be achieved using, for example, a shear mill, a ball mill, a colloid mill, a knife mill, a hammer mill, a grinding mill, a puree machine, a filter press, a mechanical press, or any combination thereof.

In some embodiments, entry into or exit from a lysing (*e.g.*, milling) process may be metered at any desired volume, mass, or other rate or interval (*e.g.*, a constant rate, a variable rate, continuously, semi-continuously, periodically, intermittently). A feed rate and/or mode
30 may be determined based on considerations including, for example: a target production rate;

apparatus(es) employed in a process; properties of a feedstock, or any combination thereof. A feed rate, in some embodiments, may be at least about 10 kg/hour, or at least about 50 kg/hour, or at least about 100 kg/hour, or at least about 200 kg/hour, or at least about 300 kg/hour, or at least about 400 kg/hour, or at least about 500 kg/hour, or at least about 600 kg/hour, or at least about 700 kg/hour, or at least about 800 kg/hour, or at least about 900 kg/hour, or at least about 1000 kg/hour, or at least about 1200 kg/hour, or at least about 1400 kg/hour, or at least about 1600 kg/hour, or at least about 1800 kg/hour, or at least about 2000 kg/hour, or at least about 2200 kg/hour. In some embodiments, the feeding rate is from about 10 kg/hour to about 200 kg/hour, or from about 200 kg/hour to about 400 kg/hour, or from about 400 kg/hour to about 600 kg/hour, or from about 600 kg/hour to about 800 kg/hour, or from about 800 kg/hour to about 1000 kg/hour, or from about 1000 kg/hour to about 1200 kg/hour, or from about 1200 kg/hour to about 1400 kg/hour, or from about 1400 kg/hour to about 1600 kg/hour, or from about 1600 kg/hour to about 1800 kg/hour, or from about 1800 kg/hour to about 2000 kg/hour, or from about 2000 kg/hour to about 2200 kg/hour.

Chemical methods may be employed, in some embodiments, (*e.g.*, alone or in combination with mechanical methods) to lyse a biomass or washed biomass. According to some embodiments, an amphiphilic compound (*e.g.*, detergent) may be employed to lyse a biomass or a washed biomass. In some embodiments, an amphiphilic chemical compound may comprise lecithin. Enzymes (*e.g.*, cellulase) may be used, in some embodiments to breakdown or assist in breakdown of cellular structures. According to some embodiments, sonication may be performed to breakdown or assist in breakdown of cellular structures, for example, by a sonicator. In some embodiments, manual grinding may be employed to lyse a biomass or a washed biomass. According to some embodiments, a freeze-thaw method may be employed to lyse a biomass or a washed biomass, wherein iterative cycles of freezing and thawing may disrupt cells. In some embodiments lysing may be performed, for example, by changing the pH value of a biomass (*e.g.*, harvested microcrop). The pH value, in some embodiments, may be raised to higher than about 7.0, or higher than about 7.5, or higher than about 8.0, or higher than about 8.5, or higher than about 9.0, or higher than about 9.5, or higher than about 10.0. According to some embodiments, the pH value of a biomass may be maintained from about 7.0 to about 7.5, or from about 7.5 to about 8.0, or from about 8.0 to about 8.5, or from about 8.5 to about 9.0, or from about 9.0 to about 9.5, or from about 9.5 to about 10.0. The pH value of a biomass may be maintained from about 7.0 to about 14.0, or from about 7.0 to about 13.0, or from about 7.0 to about 12.0, or from about 7.0 to about

11.0, or from about 7.0 to about 10.0, or from about 7.0 to about 10.5, or from about 7.0 to about 10.0, or from about 7.0 to about 9.5, or from about 7.0 to about 9.0, or from about 7.0 to about 8.5, or from about 7.0 to about 8.0, or from about 7.0 to about 7.5, in some embodiments. In some embodiments the pH value may be lowered to below about 7.0, or below about 6.5, or below about 6.0, or below about 5.5, or below about 5.0, or below about 4.5, or below about 4.0, or below about 3.5, or below about 3.0. The pH value of a biomass, in some embodiments, may be maintained from about 3.0 to about 3.5, or from about 3.5 to about 4.0, or from about 4.0 to about 4.5, or from about 4.5 to about 5.0, or from about 5.0 to about 5.5, or from about 5.5 to about 6.0, or from about 6.0 to about 6.5, or from about 6.5 to about 7.0. The pH value of a biomass may be maintained from about 3.0 to about 7.0, or from about 3.5 to about 7.0, or from about 4.0 to about 7.0, or from about 4.5 to about 7.0, or from about 5.0 to about 7.0, or about 50 from about 5.5 to about 7.0, or from about 6.0 to about 7.0, or from about 6.5 to about 7.0, according to some embodiments.

In some embodiments, a lysed biomass (*e.g.*, a mechanically lysed biomass, a chemically lysed biomass) may pass to the next step or procedure for isolating protein and/or other product(s) with or without neutralization. For example, a lysed biomass may be fed directly to the next procedure or it may be first pH-adjusted (*e.g.*, neutralized). Precipitating agents (*e.g.*, salts) may be added, in some embodiments, to a lysed microcrop to precipitate dissolved compounds.

A lysed biomass (*e.g.*, first portion, second portion) may be at a temperature below room temperature (*e.g.*, about 12° C), in some embodiments. Cooling a lysed biomass may improve protein recovery efficiency and/or decrease proteolytic activity. A lysed biomass, in some embodiments, may have a temperature below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C at the time of use. According to some embodiments, a lysed biomass may have a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C at the time of use.

In some embodiments, a lysed biomass (*e.g.*, first portion, second portion) may have a temperature above room temperature (*e.g.*, about 50° C) at the time of use. Heating a lysed biomass may improve protein recovery efficiency, decrease proteolytic activity (*e.g.*, denature proteolytic enzymes), and/or decrease microbial contamination (*e.g.*, pasteurization). In some embodiments, a lysed biomass may have a temperature above about

20° C, or above about 25° C, or above about 30° C, or above about 35° C, or above about 40° C, or above about 45° C, or above about 50° C, or above about 55° C, or above about 60° C, or above about 65° C, or above about 70° C, or above about 75° C, or above about 80° C, or above about 85° C, or above about 90° C at the time of use. A lysed biomass may have a temperature between about 40° C and about 50° C, or between about 45° C and about 55° C, or between about 50° C and about 60° C, or between about 75° C and about 80° C, or between about 80° C and about 85° C at the time of use, in some embodiments.

Separating a Biomass

A biomass (*e.g.*, *Lemna*), washed biomass, lysed biomass, or any combination thereof may be separated to generate a juice fraction and a solid fraction. A juice fraction (*e.g.*, first portion, second portion) may include a protein-rich liquid and/or at least about some solid particles (*e.g.*, carbohydrates, fiber). A solid fraction (*e.g.*, first portion, second portion) may include a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide). In some embodiments a biomass (*e.g.*, washed, lysed) may be diluted with a dilution fluid (*e.g.*, water, recycled water, reverse osmosis water) prior to separation.

A dilution fluid may be at a temperature below room temperature (*e.g.*, about 12° C), in some embodiments. Cooling a dilution fluid may improve protein recovery efficiency and/or decrease proteolytic activity. A dilution fluid, in some embodiments, may have a temperature below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C at the time of use. According to some embodiments, a dilution fluid may have a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C at the time of use.

In some embodiments, a dilution fluid may have a temperature above room temperature (*e.g.*, about 50° C) at the time of use. Heating a dilution fluid may improve protein recovery efficiency, decrease proteolytic activity (*e.g.*, denature proteolytic enzymes), and/or decrease microbial contamination (*e.g.*, pasteurization). In some embodiments, a dilution fluid may have a temperature above about 20° C, or above about 25° C, or above about 30° C, or above about 35° C, or above about 40° C, or above about 45° C, or above about 50° C, or above about 55° C, or above about 60° C, or above about 65° C, or above about 70° C, or above about 75° C, or above about 80° C, or above about 85° C, or above

about 90° C at the time of use. A dilution fluid may have a temperature between about 40° C and about 50° C, or between about 45° C and about 55° C, or between about 50° C and about 60° C at the time of use, in some embodiments. According to some embodiments, a dilution fluid may have a temperature between about 75° C and about 80° C, or between about 80° C and about 85° C at the time of use.

A dilution fluid, in some embodiments, may include buffers, protease inhibitors, anti-microbial agents, chelators (*e.g.*, EDTA), reducing agents, or any combination thereof. In some embodiments a lysed biomass or diluted lysed biomass may be sonicated prior to separation. Sonication may increase protein yield.

Separating a biomass (*e.g.*, washed, lysed) to form a juice fraction and a solid fraction may involve pressing (*e.g.*, belt press, filter press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating a biomass (*e.g.*, harvested microcrop), washed biomass, and/or lysed biomass include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

In some embodiments, a biomass (*e.g.*, harvested microcrop), a washed biomass, a lysed biomass, or any combination thereof may be metered to a separating mechanism at any desired volume, mass, or other rate or interval (*e.g.*, a constant rate, a variable rate, continuously, semi-continuously, periodically, intermittently). A feed rate and/or mode may be determined based on considerations including, for example: the target production rate; apparatus(es) employed in the process; properties of the feedstock; or any combination thereof. A feeding rate, in some embodiments, may be at least about 10 kg/hour, or at least about 50 kg/hour, or at least about 100 kg/hour, or at least about 200 kg/hour, or at least about 300 kg/hour, or at least about 400 kg/hour, or at least about 500 kg/hour, or at least about 600 kg/hour, or at least about 700 kg/hour, or at least about 800 kg/hour, or at least about 900 kg/hour, or at least about 1000 kg/hour, or higher than about 1000 kg/hour. According to some embodiments, the feeding rate may be from about 10 kg/hour to about 200 kg/hour, or from about 200 kg/hour to about 400 kg/hour, or from about 400 kg/hour to about 600 kg/hour, or from about 600 kg/hour to about 800 kg/hour, or from about 800 kg/hour to about 1000 kg/hour, or from about 1000 kg/hour to about 1200 kg/hour, or from about 1200 kg/hour to about 1400 kg/hour, or from about 1400 kg/hour to about 1600 kg/hour, or from about 1600 kg/hour to about 1800 kg/hour, or from about 1800 kg/hour to about 2000 kg/hour, or from about 2000 kg/hour to about 2200 kg/hour.

Separating a biomass may be performed at any desired temperature. Separating may be performed at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments, separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed, for example, at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C.

Separating a Solid Fraction

In some embodiments, a solid fraction may be further separated to extract additional juice (*e.g.*, a second juice (FIGURE 3A, **332**)). Separation of a solid fraction (*e.g.*, first portion, second portion) may form a second juice (*e.g.*, FIGURE 3A, **332**) and a first solid (*e.g.*, FIGURE 3A, **331**). A second juice (*e.g.*, first portion, second portion) may include a protein-rich liquid and/or at least some solid particles (*e.g.*, carbohydrates, fiber).

Separating a solid fraction to form a second juice and a first solid may involve pressing (*e.g.*, screw press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating a solid fraction include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

In some embodiments, a solid fraction may be metered to a separating mechanism at any desired volume, mass, or other rate or interval (*e.g.*, a constant rate, a variable rate, continuously, semi-continuously, periodically, intermittently). A feed rate and/or mode may be determined based on considerations including, for example: the target production rate; apparatus(es) employed in the process; properties of the feedstock; or any combination thereof. A feeding rate, in some embodiments, may be at least about 10 kg/hour, or at least about 50 kg/hour, or at least about 100 kg/hour, or at least about 200 kg/hour, or at least about 300 kg/hour, or at least about 400 kg/hour, or at least about 500 kg/hour, or at least about 600 kg/hour, or at least about 700 kg/hour, or at least about 800 kg/hour, or at least about 900 kg/hour, or at least about 1000 kg/hour, or higher than about 1000 kg/hour. According to some embodiments, the feeding rate may be from about 10 kg/hour to about 200 kg/hour, or from about 200 kg/hour to about 400 kg/hour, or from about 400 kg/hour to

about 600 kg/hour, or from about 600 kg/hour to about 800 kg/hour, or from about 800 kg/hour to about 1000 kg/hour, or higher than about 1000 kg/hour, or from about 1000 kg/hour to about 1200 kg/hour, or from about 1200 kg/hour to about 1400 kg/hour, or from about 1400 kg/hour to about 1600 kg/hour, or from about 1600 kg/hour to about 1800 kg/hour, or from about 1800 kg/hour to about 2000 kg/hour, or from about 2000 kg/hour to about 2200 kg/hour.

Separating a solid fraction may be performed at any desired temperature. According to some embodiments, separating may be performed at temperatures below room temperature, for example, to decrease proteolytic activity and/or microbial growth. In some embodiments, separating may be performed at a temperature below about 40° C, below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed, for example, at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C.

In some embodiments, a separation apparatus (*e.g.*, screw press) selected to separate a solid fraction may be the same apparatus used to separate a biomass (*e.g.*, lysed) to form a juice fraction and a solid fraction. A separation apparatus (*e.g.*, screw press) selected to separate a solid fraction may be a different apparatus than that used to separate (*e.g.*, decanter centrifuge) a biomass (*e.g.*, lysed) to form a juice fraction and a solid fraction, in some embodiments. In some embodiments, a separation apparatus (*e.g.*, screw press) may be used multiple times to extract additional second juice from a solid fraction.

According to some embodiments, a process for growing, harvesting, and separating a microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) may be single cycle: (a) at least one of a first cake (*e.g.*, FIGURE 3, **342**) and a second cake (*e.g.*, FIGURE 3, **347**) which are collected at other stages in the cycle (*e.g.*, separation of a juice fraction yields a first cake) may be combined with a first solid to form a solid mixture, and (b) the solid mixture may be further processed (*e.g.*, FIGURE 2A, FIGURE 2B, FIGURE 3A, FIGURE 3B).

In some embodiments a process for growing, harvesting, and separating a microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) may be multiple cycles or a continuous process such that one or more of a first cake (*e.g.*, FIGURE 4,

442; FIGURE 5A, 542) and a second cake (*e.g.*, FIGURE 4, 447; FIGURE 5A, 547) that are collected in an earlier cycle may be combined with a solid fraction from a subsequent cycle prior to separation (*e.g.*, FIGURE 4, 430; FIGURE 5A, 530) of the solid fraction.

Increasing the extraction of a second juice from a solid fraction may decrease the overall moisture content of a first solid and may thereby lower the energy expenditure required to further process the first solid (*e.g.*, energy required to dry). Additionally, increasing the extraction of juice from a solid fraction and/or solid mixture may improve the yield of a protein-rich product and/or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide).

In some embodiments, a moisture content of a solid fraction and/or solid mixture may be less than about 90%, or less than about 80%, or less than about 70%, or less than about 60%, or less than about 50%, or less than about 40%, or less than about 30%, or less than about 20%, or less than about 10% by weight.

Separating a Juice Fraction

A juice fraction (*e.g.*, first portion, second portion) may be separated to generate a first juice and a first cake, according to some embodiments. A first juice (*e.g.*, first portion, second portion) may include a dissolved protein. In some embodiments, buffers, protease inhibitors, anti-microbial agents, chelators (*e.g.*, EDTA), reducing agents, or any combination thereof may be added to a juice fraction and/or a first juice. Separating a juice fraction, in some embodiments, may include centrifugation, filtration, pressurized filtration, or any combination thereof. Two or more unit operations (*e.g.*, interchangeable unit operations) may be used to separate a juice fraction including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

Microfiltration may be used, in some embodiments, to separate a juice fraction into a first juice and a first cake. In some embodiments, suitable filter sizes may include $\leq$ about 10 μm , or $\leq$ about 5 μm , or $\leq$ about 3 μm , or $\leq$ about 2 μm , or $\leq$ about 1 μm , or $\leq$ about 0.5 μm , or $\leq$ about 0.4 μm , or $\leq$ about 0.3 μm , or $\leq$ about 0.2 μm , or $\leq$ about 0.1 μm . A filter may have a filter size of not less than about 0.1 μm , in some embodiments. Microfiltration may reduce the concentration of suspended solids (*e.g.*, fats, fiber), microbial contamination (*e.g.*,

Escherichia coli), and/or fungal contamination (*e.g.*, yeast) in the first juice, according to some embodiments.

In some embodiments, a vacuum may be implemented during at least some of a separating process.

5 Separating may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed at
10 a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

15 A first juice may be pumped into a storage tank, for example, a chilled storage tank, until further processing. In some embodiments a chilled storage tank may be maintained at a temperature below room temperature (*e.g.*, 12° C). Storage of a first juice at a low temperature may reduce proteolytic activity and thereby improve protein recovery efficiency. A chilled storage tank, in some embodiments, may be maintained at a temperature below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or
20 below about 2° C, or below about 1° C, or below about 0° C. According to some embodiments, a chilled storage tank may be maintained at a temperature of about 5° C, or about 6° C, or about 7° C, or about 8° C, or about 9° C, or about 10° C, or about 11° C, or about 12° C, or about 13° C, or about 14° C, or about 15° C. In some embodiments, a first juice may be fed directly to further processing without being stored in a storage tank.

25 Any one or more of the liquid phases (*e.g.*, a juice fraction, a first juice, a second juice, a third juice) or any one or more of the solid phases (*e.g.*, a solid fraction, a first cake, a second cake) generated in one procedure may be stored in a storage tank before being fed to one or more downstream procedures or apparatuses. In some embodiments a homogeneous liquid phase or solid phase may be generated for a downstream procedure(s) or apparatus(es).
30 This may accommodate different operation schedules or modes including, for example, continuous mode, batch mode, or multiple feeding streams to one or more downstream procedure(s) and/or apparatus(es). A liquid phase or a solid phase may be maintained in a

storage tank at a desirable temperature (*e.g.*, below room temperature, such as 12° C) to reduce degradation and maintain high quality until further processing.

Separating a First Cake and/or a Second Juice

5 In some embodiments, further processing of a first cake (*e.g.*, first portion, second portion) and/or a second juice (*e.g.*, first portion, second portion) may be performed. Such additional processing may increase product yield and/or quality. In some embodiments, a first cake and a second juice may be combined and further separated to form a third juice and a second cake. A first cake and a second juice may be independently subjected to further
10 separation, according to some embodiments.

Separating a first cake, a second juice, or any combination thereof may involve vibratory separation, centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit operations may be used to separate including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a
15 linear/inclined motion shaker, a decanter centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

In some embodiments, filtration (*e.g.*, a vibratory separator) may be used to separate a first cake, a second juice, or any combination thereof to form a third juice and a second cake. Suitable filter sizes may include, in some embodiments, $\leq$ about 800 μm , or $\leq$ about 600 μm ,
20 or $\leq$ about 500 μm , or $\leq$ about 400 μm , or $\leq$ about 300 μm , or $\leq$ about 200 μm , or $\leq$ about 180 μm , or $\leq$ about 150 μm , or $\leq$ about 120 μm , or $\leq$ about 100 μm , or $\leq$ about 90 μm , or $\leq$ about 80 μm , or $\leq$ about 70 μm , or $\leq$ about 60 μm , or $\leq$ about 50 μm , or $\leq$ about 40 μm , or $\leq$ about 30 μm , or $\leq$ 25 μm , or $\leq$ about 20 μm , or $\leq$ about 15 μm , or $\leq$ about 10 μm , or $\leq$ about 5 μm , or $\leq$ about 1 μm . A filter may have a filter size of not more than about 800 μm , in
25 some embodiments. Pore size of a filter may be selected larger or smaller as desired. For example, a larger pore size may be desirable where removal of contaminant material is of interest. A smaller pore size may be desirable where limiting the number of cycles of the process and/or protein yield is of interest. In some embodiments, a pore size of a filter may be selected based on lysing conditions, for example, an average particle size of a lysed
30 biomass. A pore size of a filter may be selected based on one or more characteristics of a microcrop (*e.g.* cell wall composition, protein composition), according to some embodiments.

Microfiltration may be used, in some embodiments, to separate a first cake, a second juice, or any combination thereof to form a third juice and a second cake. In some

embodiments, suitable filter sizes may include $\leq$ about 10 μm , or $\leq$ about 5 μm , or $\leq$ about 3 μm , or $\leq$ about 2 μm , or $\leq$ about 1 μm , or $\leq$ about 0.5 μm , or $\leq$ about 0.4 μm , or $\leq$ about 0.3 μm , or $\leq$ about 0.2 μm , or $\leq$ about 0.1 μm . A microfilter may have a filter size of not less than about 0.1 μm , in some embodiments.

5 In some embodiments, a vacuum may be implemented during at least some of a separating process.

Separating may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or
10 below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30°
15 C and about 40° C, in some embodiments.

According to some embodiments, a process for growing, harvesting, and separating a microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) may comprise a single cycle. In a single cycle process, in some embodiments, at least one of a first cake (*e.g.*, FIGURE 3, **342**) and a second cake (*e.g.*, FIGURE 3, **347**) may be
20 combined with a first solid to form a solid mixture, and the solid mixture may be further processed (*e.g.*, FIGURE 2A, FIGURE 2B, FIGURE 3A, FIGURE 3B). In some embodiments of a single cycle process, a third juice may be combined with a first juice prior to further processing.

In some embodiments a process for growing, harvesting, and separating a microcrop
25 (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) may comprise multiple cycles (*e.g.*, a continuous process). In a multiple cycle or continuous process, according to some embodiments, one or more of a first cake (*e.g.*, FIGURE 4, **442**; FIGURE 5A, **542**) and a second cake (*e.g.*, FIGURE 4, **447**; FIGURE 5A, **547**) that are collected in an earlier cycle may be combined with a solid fraction from a subsequent cycle prior to separation (*e.g.*, FIGURE 4, **430**; FIGURE 5A, **530**) of the solid fraction. In some
30 embodiments of a multiple cycle or continuous process a third juice collected in an earlier cycle may be combined with a juice fraction from a subsequent cycle prior to further processing.

Filtering a First Juice, a Third Juice, or any Combination Thereof

A first juice (*e.g.*, first portion, second portion), a third juice (*e.g.*, first portion, second portion), or any combination thereof may be filtered one or more times to generate a soluble protein product (*e.g.*, a soluble protein (*e.g.*, FIGURE 2, **251**), a first soluble protein (*e.g.*, FIGURE 3, **351**), a second soluble protein (*e.g.*, FIGURE 3, **356**)). Filtration may involve microfiltration, ultrafiltration, nanofiltration, or reverse osmosis filtration either individually or in combination.

According to some embodiments, diafiltration may be used in conjunction with ultrafiltration and/or nanofiltration, by an addition of water to a first soluble protein (*e.g.*, FIGURE 2A, **251**; FIGURE 2B, **251**; FIGURE 3A, **351**; FIGURE 3B, **351**; FIGURE 4, **451**) or a second soluble protein (FIGURE 3A, **356**; FIGURE 3B, **356**; FIGURE 4, **456**). In some embodiments, using diafiltration in conjunction with ultrafiltration and/or nanofiltration by an addition of water to a first soluble protein, a second soluble protein, or a combination thereof may further reduce a concentration of permeable solutes from a first soluble protein, a second soluble protein, or a combination thereof. In some embodiments, reducing a concentration of permeable solutes from a first soluble protein, a second soluble protein, or a combination thereof may increase a protein purity of a first soluble protein, a second soluble protein, or a combination thereof. In some embodiments, a permeate of ultrafiltration (*e.g.*, FIGURE 3A, FIGURE 3B, FIGURE 4) may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

Microfiltration may reduce the concentration of suspended solids (*e.g.*, fats, fiber), microbial contamination (*e.g.*, *Escherichia coli*), and/or fungal contamination (*e.g.*, yeast) in a first juice, a third juice, or any combination thereof, according to some embodiments. Suitable filter sizes for microfiltration may include, in some embodiments, $\leq$ about 10 μm , or $\leq$ about 5 μm , or $\leq$ about 3 μm , or $\leq$ about 2 μm , or $\leq$ about 1 μm , or $\leq$ about 0.5 μm , or $\leq$ about 0.4 μm , or $\leq$ about 0.3 μm , or $\leq$ about 0.2 μm , or $\leq$ about 0.1 μm . In some embodiments, a first juice, a third juice, or any combination thereof may be filtered using microfiltration to generate a soluble protein in the permeate.

Ultrafiltration may involve membrane filtration using pressure, concentration gradients, or a combination thereof. Suitable nominal molecular weight cut-offs (NMWCO) for ultrafiltration may be, in some embodiments, at most about 100 kDa, or at most about 90 kDa, or at most about 80 kDa, or at most about 70 kDa, or at most about 60 kDa, or at most

about 55 kDa, or at most about 50 kDa, or at most about 45 kDa, or at most about 40 kDa, or at most about 30 kDa, or at most about 20 kDa, or at most about 15 kDa, or at most about 14 kDa, or at most about 13 kDa, or at most about 12 kDa, or at most about 11 kDa, or at most about 10 kDa, or at most about 9 kDa, or at most about 8 kDa, or at most about 7 kDa, or at most about 6 kDa, or at most about 5 kDa, or at most about 4 kDa, or at most about 3 kDa, or at most about 2 kDa, or at most about 1 kDa. In some embodiments, suitable NMWCO cut-offs for ultrafiltration may be in a range of at most about 1 kDa to at most about 10 kDa, at most about 2 kDa to at most about 10 kDa, at most about 3 kDa to at most about 10 kDa, at most about 3 kDa to at most about 15 kDa, or at most about 3 kDa to at most about 20 kDa, or at most about 3 kDa to at most about 60 kDa, or at most about 3 kDa to at most about 55 kDa, or at most about 10 kDa to at most about 55 kDa. In some embodiments a NMWCO for ultrafiltration may be at least 1 kDa, or at least 3 kDa, or at least 5 kDa, or at least 10 kDa, or at least 15 kDa, or at least 20 kDa, or at least 25 kDa, or at least 30 kDa, or at least 35 kDa, or at least 40 kDa, or at least 45 kDa, or at least 50 kDa, or at least 55 kDa. A suitable NMWCO for ultrafiltration may vary depending on a manufacturing specification of an ultrafilter. In some embodiments a suitable NMWCO for ultrafiltration may vary depending on a rate of hydrolysis.

In some embodiments, suitable filter sizes for nanofiltration may include $\leq$ about 0.01 μm , or $\leq$ about 0.009 μm , or $\leq$ about 0.008 μm , or $\leq$ about 0.007 μm , or $\leq$ about 0.006 μm , or $\leq$ about 0.005 μm , or $\leq$ about 0.004 μm , or $\leq$ about 0.003 μm , or $\leq$ about 0.002 μm , or $\leq$ about 0.001 μm . A nanofiltration filter may have a filter size of not more than about 0.01 μm , in some embodiments.

According to some embodiments, suitable filter sizes for reverse osmosis filtration may include $\leq$ about 0.001 μm , or $\leq$ about 0.0009 μm , or $\leq$ about 0.0008 μm , or $\leq$ about 0.0007 μm , or $\leq$ about 0.0006 μm , or $\leq$ about 0.0005 μm , or $\leq$ about 0.0004 μm , or $\leq$ about 0.0003 μm , or $\leq$ about 0.0002 μm , or $\leq$ about 0.0001 μm . A reverse osmosis filter may have a filter size of not more than about 0.001 μm , in some embodiments.

Buffers, protease inhibitors, anti-microbial agents, chelators (*e.g.*, EDTA), reducing agents, or any combination thereof may be added, in some embodiments, to a soluble protein product. A soluble protein product may be chilled and/or stored at a temperature below about 30° C, or below about 25° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C, or below about -2° C, or below about -5° C, or below about -10° C, in some embodiments.

Chilling and/or storing a soluble protein product at reduced temperatures may reduce degradation and/or improve protein recovery efficiency.

Polyphenol Reduction

5 In some embodiments, a polyphenol-rich product may be subjected to a polyphenol reduction process to generate a product having a reduced concentration of at least one polyphenol (*e.g.*, a tannin). A polyphenol-rich product may include, according to some embodiments, a juice fraction (*e.g.*, FIGURE 1, **126**), a soluble protein (*e.g.*, FIGURE 2A, **251**; FIGURE 2B, **251**), a first soluble protein (*e.g.*, FIGURE 3A, **351**; FIGURE 3B, **351**,
10 FIGURE 4, **451**), a second soluble protein (*e.g.*, FIGURE 3A, **356**; FIGURE 3B, **356**; FIGURE 4, **456**), a concentrated protein (*e.g.*, FIGURE 3A, **361**; FIGURE 3B, **361**; FIGURE 4, **461**), a first juice (*e.g.*, FIGURE 2A, **241**; FIGURE 2B, **241**; FIGURE 3A, **341**; FIGURE 3B, **341**, FIGURE 4, **441**, FIGURE 5A, **541**, FIGURE 5B, **541**), a second juice (*e.g.*, FIGURE 2A, **232**; FIGURE 2B, **232**; FIGURE 3A, **332**; FIGURE 3B, **332**, FIGURE 4, **432**,
15 FIGURE 5A, **532**, FIGURE 5B, **532**), a third juice (*e.g.*, FIGURE 2A, **246**; FIGURE 2B, **246**; FIGURE 3A, **346**; FIGURE 3B, **346**, FIGURE 4, **446**, FIGURE 5A, **546**, FIGURE 5B, **546**), a juice fraction (*e.g.*, FIGURE 1, **126**; FIGURE 2A, **226**; FIGURE 2B, **226**; FIGURE 3A, **326**; FIGURE 3B, **326**, FIGURE 4, **426**, FIGURE 5A, **526**, FIGURE 5B, **526**), or any combination thereof. A polyphenol reduction process may be configured to reduce a
20 concentration of at least one polyphenol (*e.g.*, at least one tannin), according to some embodiments. A polyphenol reduction process may be configured, in some embodiments, to minimize a reduction in yield or quality of a downstream soluble protein product.

According to some embodiments a polyphenol reduction process may comprise passing a polyphenol-rich product through an ion exchange resin. In some embodiments, a
25 polyphenol reduction process may comprise passing a polyphenol-rich product through a series (*e.g.*, at least two, at least three) of ion exchange resins. Each ion exchange resin in a series may be the same or different than the other ion exchange resins in the series. In some embodiments an ion exchange resin may be a strongly acidic resin, a strongly basic resin (*e.g.*, DIAION PA308), a weakly acidic resin (*e.g.*, Relite JA800), a weakly basic resin, a
30 weak anion exchange resin (*e.g.*, Relite RAM2), a strong anion exchange resin, a weak cation exchange resin, a strong cation exchange resin, or any combination thereof. According to some embodiments a polyphenol reduction process may comprise passing a polyphenol-rich product through an ion exchange column selected from a weakly acidic resin (*e.g.*, Relite

JA800), an anion exchange resin (*e.g.*, Relite RAM2), a strongly basic resin (*e.g.*, DIAION PA308), or a combination thereof. A polyphenol reduction process, in some embodiments, may comprise: first passing a polyphenol-rich product through an ion exchange column selected from a weak anion exchange and a strong anion exchange resin, and second passing the polyphenol-rich product through an ion exchange column selected from a weak anion exchange resin and a strong anion exchange resin. Ion exchange resins may be used in a batch mode or arranged in a continuous process, whereby resins may be cycled through polyphenol extraction and regeneration processes. In some embodiments a polyphenol reduction process may further comprise adjusting a pH of a polyphenol-rich product or a product yielded from an ion exchange column. A polyphenol reduction process may be performed alone or in combination with other purification processes and/or steps.

In some embodiments a polyphenol reduction process may reduce a polyphenol (*e.g.*, a tannin) content of a polyphenol-rich product by at least 5%, or at least 10%, or at least 15%, or at least 20%, or at least 25%, or at least 30%, or at least 35%, or at least 40%, or at least 45%, or at least 50%, or at least 55%, or at least 60%, or at least 65%, or at least 70%. A polyphenol reduction process, according to some embodiments, may reduce a polyphenol content of a polyphenol-rich product from about 5% to about 10%, from about 15% to about 20%, from about 20% to about 30%, from about 30% to about 40%, from about 35% to about 45%, from about 40% to about 50%, from about 45% to about 55%, from about 50% to about 60%, from about 55% to about 65%, or from about 60% to about 70%.

In some embodiments, a soluble protein product (*e.g.*, a soluble protein, a first soluble protein, a second soluble protein), may comprise polyphenol (*e.g.*, total polyphenol) at a concentration of about 0.05 g/100 g of soluble protein product, or about 0.1 g/100 g of soluble protein product, or about 0.5 g/100 g of soluble protein product, or about 1 g/100 g of soluble protein product, or about 5 g/100 g of soluble protein product, or about 10 g/100 g of soluble protein product, or about 20 g/100 g of protein concentrate. According to some embodiments, a 100 g sample of a soluble protein product resulting from a polyphenol reduction process may include at least about 65 g of protein and not more than 1.092 g of polyphenols (expressed as gallic acid equivalents).

Reducing a Moisture Content of Soluble Protein Products

In some embodiments a process may be used to reduce a moisture content of a soluble protein (*e.g.*, FIGURE 2, **251**), a first soluble protein (*e.g.*, FIGURE 3, **351**), a second soluble

protein (*e.g.*, FIGURE 3, **356**), or any combination thereof (collectively “a soluble protein product”). Reducing a moisture content of a soluble protein product may reduce capital and operational expenditures, for example, by reducing the energy needed to dry an end protein product (*e.g.*, concentrated soluble protein).

5 In some embodiments an evaporation process may be used to reduce a moisture content of a soluble protein product. Evaporation may be performed by, for example, a thermal (evaporative) means such as: a rising film evaporator, a falling film evaporator, a natural circulation evaporator (vertical or horizontal), an agitated-film evaporator, a multiple-effect evaporator, by vacuum evaporation, or any combination thereof. Heat may be supplied
10 directly into the evaporator, or indirectly through a heat jacket. Heat may either come from a raw source (*e.g.*, combustion of natural gas, steam from a boiler) or from a waste heat stream (*e.g.*, dryer exhaust) or from heat transferred by cooling the input stream.

A moisture content of a soluble protein product (*e.g.*, a soluble protein, a second soluble protein) may be reduced, in some embodiments, by nanofiltration or reverse osmosis
15 filtration. In some embodiments, suitable filter sizes for nanofiltration may include $\leq$ about 0.01 μm , or $\leq$ about 0.009 μm , or $\leq$ about 0.008 μm , or $\leq$ about 0.007 μm , or $\leq$ about 0.006 μm , or $\leq$ about 0.005 μm , or $\leq$ about 0.004 μm , or $\leq$ about 0.003 μm , or $\leq$ about 0.002 μm , or $\leq$ about 0.001 μm . A moisture content of a soluble protein product (*e.g.*, a soluble protein, a second soluble protein) may be reduced, in some embodiments, using nanofiltration with the
20 soluble protein product (*e.g.*, a soluble protein, a second soluble protein) in a retentate. According to some embodiments, suitable filter sizes for reverse osmosis filtration may include $\leq$ about 0.001 μm , or $\leq$ about 0.0009 μm , or $\leq$ about 0.0008 μm , or $\leq$ about 0.0007 μm , or $\leq$ about 0.0006 μm , or $\leq$ about 0.0005 μm , or $\leq$ about 0.0004 μm , or $\leq$ about 0.0003 μm , or $\leq$ about 0.0002 μm , or $\leq$ about 0.0001 μm . A moisture content of a soluble protein
25 product (*e.g.*, a soluble protein, a second soluble protein), in some embodiments, may be reduced using reverse osmosis filtration with the soluble protein product in a retentate. A permeate of nanofiltration or reverse osmosis filtration may be recycled (*e.g.*, dilution fluid for lysis; wash solution), according to some embodiments.

In some embodiments an antioxidant (*e.g.*, rosemary extract) may be mixed with a
30 soluble protein product (*e.g.*, a soluble protein, a second soluble protein) prior to drying to improve shelf life of a product when packaged.

Drying a Soluble Protein Product

A soluble protein product (*e.g.*, a soluble protein, a first soluble protein, a second soluble protein) may be dried to generate a dry protein concentrate (*e.g.*, first portion, second portion), according to some embodiments. A drying procedure, in some embodiments, may reduce the moisture content of a soluble protein product to a desired level (*e.g.*, higher or lower moisture content, a desired moisture content). A moisture content of a dry protein concentrate may be, for example, below 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the dry protein concentrate, in some embodiments. According to some embodiments, a protein concentration of a dry protein concentrate may be from about 30% to about 95%, or from about 40% to about 90%, or from about 50% to about 85%, or from about 60% to about 80%, or from about 70% to about 75% by weight of the dry protein concentrate. A drying procedure may be performed using a mechanism including, for example, a spray dryer, double drum dryer, flash dryer, an evaporator, or any combination thereof.

In some embodiments, an inlet temperature of a dryer mechanism (the temperature at the entrance to a dryer) may be above 25° C, or above 50° C, or above 75° C, or above 100° C, or above 125° C, or above 150° C, or above 175° C, or above 200° C, or above 225° C, or above 250° C, or above 275° C, or above 300° C, or above 325° C, or above 350° C, or above 375° C, or above 400° C, or above 425° C, or above 450° C, or above 475° C, or above 500° C. An inlet temperature, in some embodiments, may be from about 25° C to about 50° C, or from about 50° C to about 75° C, or from about 75° C to about 100° C, or from about 100° C to about 125° C, or from about 125° C to about 150° C, or from about 150° C to about 175° C, or from about 175° C to about 200° C, or from about 200° C to about 225° C, or from about 225° C to about 250° C, or from about 250° C to about 275° C, or from about 275° C to about 300° C, or from about 300° C to about 325° C, or from about 325° C to about 350° C, or from about 350° C to about 375° C, or from about 375° C to about 400° C, or from about 400° C to about 425° C, or from about 425° C to about 450° C, or from about 450° C to about 475° C, or from about 475° C to about 500° C, or above 500° C. An inlet temperature may be from about 50° C to about 100° C, or from about 100° C to about 150° C, or from about 150° C to about 200° C, or from about 200° C to about 250° C, or from about 250° C to about 300° C, or from about 300° C to about 350° C, or from about 350° C to about 400° C, or from about 400° C to about 450° C, or from about 450° C to about 500° C, or above 500° C, in some

embodiments. According to some embodiments, an inlet temperature of a dryer mechanism may be about 225° C.

According to some embodiments, an outlet temperature of a dryer mechanism (the temperature at the exit from a dryer) may be below about 300° C, or below about 275° C, or
5 below about 250° C, or below about 225° C, or below about 200° C, or below about 175° C, or below about 150° C, or below about 125° C, or below about 100° C, or below about 75° C, or below about 50° C, or below about 25° C. An outlet temperature may be from about 300° C to about 275° C, or from about 275° C to about 250° C, or from about 250° C to about 225° C, or from about 225° C to about 200° C, or from about 200° C to about 175° C, or from about
10 175° C to about 150° C, or from about 150° C to about 125° C, or from about 125° C to about 100° C, or from about 100° C to about 75° C, or from about 75° C to about 50° C, or from about 50° C to about 25° C, in some embodiments. An outlet temperature, in some embodiments, may be from about 300° C to about 250° C, or from about 250° C to about 200° C, or from about 200° C to about 150° C, or from about 150° C to about
15 100° C, from about 100° C to about 50° C, or from about 50° C to about 25° C, or below about 25° C. According to some embodiments, an outlet temperature of a dryer mechanism may be about 75° C.

In some embodiments, a volume of a soluble protein product (*e.g.*, a soluble protein, a first soluble protein, a second soluble protein) may be mixed with a volume of a dry protein
20 concentrate prior to drying. This process, known as back-mixing, may be employed when, for example, the moisture content of a soluble protein exceeds the level that a dryer mechanism is capable of accepting. By back-mixing a dry protein concentrate with a soluble protein product, a total moisture content may be kept within the specifications of a dryer mechanism, thereby reducing operational costs (*e.g.*, wear and tear on equipment).

25 An antioxidant (*e.g.*, rosemary extract) may be mixed with a dry protein concentrate before packaging, according to some embodiments.

Solvent Washing a Soluble Protein Product or a Dry Protein Concentrate

A soluble protein product (*e.g.*, a soluble protein, a first soluble protein, a second
30 soluble protein) and/or a dry protein concentrate (*e.g.*, first portion, second portion) may be washed with at least one solvent (*e.g.*, ethanol, methanol) to generate a washed protein product, according to some embodiments.

A washed protein product, in some embodiments, may have a reduced fat content (*e.g.*, about 2% of a dry protein concentrate or less by weight) and/or a reduced chlorophyll content (*e.g.*, visually perceivable reduction in green coloration) compared to unwashed counterparts. In some embodiments, a washed protein product may appear colorless, white, substantially white, or have reduced green coloration. A washed protein product, in some embodiments, may exhibit improved taste, color, shelf life (*e.g.*, reduced oxidation of fats), protein density, malleability, or combinations thereof. In some embodiments, a washed protein product may be extruded to form a texturized protein product.

According to some embodiments, a solvent may comprise methanol, ethanol, acetone, hexane, dichloromethane, ethyl acetate, propanol, isopropanol, glycerol, or combinations thereof.

In some embodiments, a washed protein product may have a fat content comprising lower than about 50%, or lower than about 40%, or lower than about 30%, or lower than about 25%, or lower than about 20%, or lower than about 15%, or lower than about 10%, or lower than about 5%, or lower than about 4%, or lower than about 3%, or lower than about 2%, or lower than about 1% by weight of the washed protein product. According to some embodiments, a washed protein product may have a fat content comprising from about 1% to about 10%, or from about 10% to about 20%, or from about 20% to about 30%, or from about 30% to about 40%, or from about 40% to about 50% by weight of the protein concentrate in some embodiments.

In some embodiments, a washed protein product may have a fat content comprising $\leq$ about 15% of a dry protein concentrate or less by weight (DMB), or $\leq$ about 10% DMB, or $\leq$ about 8% DMB, or $\leq$ about 6% DMB, or $\leq$ about 4% DMB, or $\leq$ about 2% DMB, or $\leq$ about 1% DMB, or $\leq$ about 0.5% DMB, or $\leq$ about 0.2% DMB, or $\leq$ about 0.1% DMB. In some embodiments, a washed protein product may have a fat content comprising from about 0.1% DMB to about 0.2% DMB.

Protein Concentrate

Some embodiments relate to a process for production of a soluble protein product (*e.g.*, a soluble protein, a first soluble protein, a second soluble protein) and/or a dry protein concentrate (collectively “a protein concentrate”) from a biomass of a harvested microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species). A process may be configured or performed to achieve any desired protein yield (*e.g.*, maximal yield, a

selected yield). In some embodiments, a protein concentration of a protein concentrate is higher than about 30%, or higher than about 40%, or higher than about 50%, or higher than 55%, or higher than about 60%, or higher than 65%, or higher than about 70%, or higher than about 75%, or higher than about 80% by weight of the protein concentrate. A remainder of a protein concentrate may include carbohydrates, fiber, fats, minerals, or any combination thereof. A protein concentrate may be suitable for animal feed and/or human consumption. For example, a protein concentrate may serve as an effective replacement for protein isolates (*e.g.*, soy, pea, whey) which are presently used in a large number of human food products either individually or as ingredients and additives. According to some embodiments, a protein composition of a protein concentrate may be in native or near native form. For example, a protein composition of a protein concentrate may include < 2% denatured protein, or < 4% denatured protein, < 6% denatured protein, or < 8% denatured protein, or < 10% denatured protein, or < 12% denatured protein, or < 14% denatured protein, or < 16% denatured protein, or < 18% denatured protein, or < 20% denatured protein, or < 22% denatured protein, or < 24% denatured protein, or < 26% denatured protein, or < 28% denatured protein, or < 30% denatured protein.

In some embodiments, a protein concentrate may comprise one or more essential amino acids. For example, a protein concentrate may include one or more amino acids selected from leucine, isoleucine, lysine, methionine, phenylalanine, threonine, tryptophan, valine, histidine, arginine, aspartic acid, serine, glutamic acid, proline, glycine, alanine, tyrosine and cysteine. The concentration of an essential amino acid may be at least about 1 g/100 g of protein concentrate, or at least about 1.5 g/100 g of protein concentrate, or at least about 2 g/100 g of protein concentrate, or at least about 2.5 g/100 g of protein concentrate, or at least about 3 g/100 g of protein concentrate, or at least about 4 g/100 g of dry at least about 2.5 g/100 g of protein concentrate, or at least about 3 g/100 g of protein concentrate, or at least about 4 g/100 g of protein concentrate, or at least about 5 g/ 100 g of protein concentrate, or at least about 6 g/100 g of protein concentrate, or at least about 7 g/100 g of protein concentrate, or at least about 8 g/100 g of protein concentrate, or at least about 9 g/100 g of protein concentrate, or at least about 10 g/100 g of protein concentrate in some embodiments.

The concentration of an amino acid (*e.g.*, an essential amino acid), in some embodiments, may be expressed as a weight fraction of the protein recovered from a protein concentrate, and is at least about 1 g/100 g of protein, or at least about 1.5 g/100 g of protein,

or at least about 2 g/100 g of protein, or at least about 2.5 g/100 g of protein, or at least about 3 g/100 g of protein, or at least about 4 g/100 g of protein, or at least about 5 g/100 g of protein, or at least about 6 g/100 g of protein, or at least about 7 g/100 g of protein, or at least about 8 g/100 g of protein, or at least about 9 g/100 g of protein, or at least about 10 g/100 g of protein.

In some embodiments, a protein concentrate may comprise one or more branched-chain amino acids (BCAAs). For example, a protein concentrate may include one or more amino acids selected from leucine, isoleucine, valine, and combinations thereof. According to some embodiments, a BCAA content may be evaluated using ion exchange chromatography of an amino acid profile based on AOAC Official Method 994.12. According to some embodiments, a concentration of a BCAA per 100 g of a protein concentrate may be at least about 1 g, or at least about 1.5 g, or at least about 2 g, or at least about 2.5 g, or at least about 3 g, or at least about 4 g, or at least about 5 g, or at least about 6 g, or at least about 7 g, or at least about 8 g, or at least about 9 g, or at least about 10 g, or at least about 11 g, or at least about 12 g, or at least about 13 g, or at least about 14 g, or at least about 15 g. In some embodiments, a BCAA protein content of a protein concentrate may be higher than a percentage of total amino acid content of the protein concentrate (TAA). For example, in some embodiments a BCAA protein content of a protein concentrate may be > about 10% TAA, or > about 11% TAA, or > about 12% TAA, or > about 13% TAA, or > about 14% TAA, or > about 15% TAA, or > about 20% TAA, or > about 25% TAA, or > about 30% TAA, or > about 35% TAA, or > about 40% TAA, or > about 45% TAA, or > about 50% TAA, or > about 55% TAA, or > about 60% TAA. In some embodiments, a BCAA protein content of a protein concentrate(e.g., derived from *Lemna*) may be at least 20% TAA, about 11% higher than a BCAA content of alternative protein products derived from pea or soy beans (about 18% TAA).

In some embodiments, a protein concentrate may have a fat content lower than about 50%, or lower than about 40%, or lower than about 30%, or lower than about 25%, or lower than about 20%, or lower than about 15%, or lower than about 10%, or lower than about 5%, or lower than about 4%, or lower than about 3%, or lower than about 2%, or lower than about 1% by weight of the protein concentrate. A protein concentrate may have a fat content from about 1% to about 10%, or from about 10% to about 20%, or from about 20% to about 30%, or from about 30% to about 40%, or from about 40% to about 50% by weight of the protein concentrate in some embodiments. A protein concentrate, in some embodiments, may have a

fat content from about 1% to about 50%, or from about 2% to about 40%, or from about 5% to about 30%, or from about 8% to about 20%, or from about 10% to about 15% by weight of the protein concentrate. A protein concentrate may be further processed to meet a desired fat content (*e.g.*, higher or lower concentration, a desired fat composition).

5 According to some embodiments, a protein concentrate may include an ash content consisting of a residue containing inorganic mineral elements. An ash content in some embodiments may be determined by combusting a protein concentrate at a high temperature (*e.g.*, $\geq 500^{\circ}\text{C}$) to remove organic matter. A protein concentrate may have an ash content lower than about 50%, or lower than about 40%, or lower than about 30%, or lower than about 25%, or lower than about 20%, or lower than about 15%, or lower than about 10%, or
10 lower than about 5%, or lower than about 4%, or lower than about 3%, or lower than about 2%, or lower than about 1% by weight of the protein concentrate in some embodiments. In some embodiments, a protein concentrate may have an ash content from about 1% to about 10%, or from about 10% to about 20%, or from about 20% to about 30%, or from about 30%
15 to about 40%, or from about 40% to about 50% by weight of the protein concentrate. A protein concentrate, in some embodiments, may have an ash content from about 1% to about 50%, or from about 2% to about 40%, or from about 3% to about 30%, or from about 3% to about 20%, or from about 3% to about 15%, or from about 3% to about 10%, or from about 5% to about 10%, or from about 5% to about 15% by weight of the protein concentrate. A
20 protein concentrate may be further processed to meet a desired ash content (*e.g.*, higher or lower concentration, a desired ash composition).

 According to some embodiments, a protein concentrate may have a carbohydrate content lower than about 50%, or lower than about 40%, or lower than about 30%, or lower than about 25%, or lower than about 20%, or lower than about 15%, or lower than about
25 10%, or lower than about 5%, or lower than about 4%, or lower than about 3%, or lower than about 2%, or lower than about 1% by weight of the protein concentrate. A protein concentrate, in some embodiments, may have a carbohydrate content from about 1% to about 10%, or from about 10% to about 20%, or from about 20% to about 30%, or from about 30% to about 40%, or from about 40% to about 50% by weight of the protein concentrate. In some
30 embodiments, a protein concentrate may have a carbohydrate content from about 1% to about 50%, or from about 2% to about 40%, or from about 5% to about 30%, or from about 8% to about 20%, or from about 10% to about 15% by weight of the protein concentrate. A protein

concentrate may be further processed to meet a desired carbohydrate content (*e.g.*, higher or lower concentration, a desired carbohydrate composition).

In some embodiments, a protein concentrate may have a fiber content lower than about 20%, or lower than about 15%, or lower than about 10%, or lower than about 8%, or lower than about 5%, or lower than about 4%, or lower than about 3%, or lower than about 2%, or lower than about 1% by weight of the protein concentrate. A protein concentrate may be further processed to meet a desired fiber content (*e.g.*, higher or lower concentration, a desired fiber composition).

For example, a dry protein concentrate produced by the processes described herein may include the contents summarized in Table 2.

TABLE 2: Example Contents of Dry Protein Concentrate Product			
Wt. Percent	Product 1	Product 2	Product 3
Solids	$\geq \sim 90$	$\geq \sim 88 - \sim 90$	$\geq \sim 95$
Moisture	$\leq \sim 10$	$\leq \sim 12 - \sim 10$	$\leq \sim 5$
Protein	$\geq \sim 50$	from ~ 60 to ~ 80	$\geq \sim 65 - \sim 75$
Fat	$\leq \sim 20$	from ~ 5 to ~ 20	$\leq \sim 5 - \sim 15$
Ash	$\leq \sim 15$	from ~ 1 to ~ 10	$\leq \sim 2 - \sim 10$
Carbohydrate	$\leq \sim 20$	from ~ 5 to ~ 20	$\leq \sim 10 - \sim 15$
Fiber	$\leq \sim 10$	$\leq \sim 5$	$\leq \sim 5$
Other	~ 10	$\sim 5 - \sim 20$	$\sim 10 - \sim 15$

A product and/or process, in some embodiments, may be configured or performed so other characteristics of a protein concentrate, (*e.g.*, particle size, bacterial specification) meet desired criteria and/or may be suitable for an intended purpose.

According to some embodiments a protein concentrate may have a mesh size (*e.g.*, most or all gross particles of the protein concentrate will pass through a mesh having an average pore size) of about 30 μm , or about 40 μm , or about 50 μm , or about 60 μm , or about 70 μm , or about 80 μm , or about 90 μm , or about 100 μm , or about 110 μm , or about 120 μm , or about 130 μm , or about 140 μm , or about 150 μm , or about 160 μm , or about 170 μm , or about 180 μm , or about 190 μm , or about 200 μm , or about 225 μm , or about 250 μm , or about 275 μm , or about 300 μm , or about 325 μm , or about 350 μm , or about 375 μm , or about 400 μm , or about 425 μm , or about 450 μm , or about 475 μm , or about 500 μm . A protein concentrate may have mesh size range of, in some embodiments, about 30 μm to about 500 μm , or about 30 μm to about 300 μm , or about 50 μm to about 300 μm , or about

70 μm to about 300 μm , or about 100 μm to about 300 μm , or about 30 μm to about 200 μm , or about 50 μm to about 200 μm , or about 70 μm to about 200 μm , or about 100 μm to about 200 μm , or about 30 μm to about 190 μm , or about 50 μm to about 190 μm , or about 70 μm or about 190 μm , or about 100 μm to about 190 μm , or about 30 μm to about 180 μm , or about 50 μm to about 180 μm , or about 70 μm to about 180 μm , or about 100 μm to about 180 μm , or about 30 μm to about 170 μm , or about 50 μm to about 170 μm , or about 70 μm to about 170 μm , or about 100 μm to about 170 μm .

A protein concentrate, according to some embodiments may have a density of about 400 kg/m^3 , or about 405 kg/m^3 , or about 410 kg/m^3 , or about 415 kg/m^3 , or about 420 kg/m^3 , or about 425 kg/m^3 , or about 430 kg/m^3 , or about 435 kg/m^3 , or about 440 kg/m^3 , or about 445 kg/m^3 , or about 450 kg/m^3 .

In some embodiments a protein concentrate may have a solubility value (% water soluble nitrogen) of at least 35%, or at least 40%, or at least 45%, or at least 50%, or at least 55%, or at least 60%, or at least 65%, or at least 70%, or at least 75%. A solubility value may be determined using the Nitrogen Solubility Index (NSI) method as described in F. Vojdani, Methods of Testing Protein Functionality 11-60 (G.M. Hall, ed., 1996).

According to some embodiments, a protein concentrate may have a dispersibility value (%water dispersible protein/ % total protein) of at least 35%, or at least 40%, or at least 45%, or at least 50%, or at least 55%, or at least 60%, or at least 65%, or at least 70%, or at least 75%. A dispersibility value may be determined using the Protein Dispersibility Index (PDI) as described in F. Vojdani, Methods of Testing Protein Functionality 11-60 (G.M. Hall, ed., 1996).

In some embodiments, a standard plate count of bacteria may be lower than about 100,000 cfu/g, or lower than about 80,000 cfu/g, or lower than about 60,000 cfu/g, or lower than about 50,000 cfu/g, or lower than about 40,000 cfu/g, or lower than about 30,000 cfu/g, or lower than about 25,000 cfu/g, or lower than about 20,000 cfu/g, or lower than about 15,000 cfu/g, or lower than about 10,000 cfu/g, or lower than about 5,000 cfu/g, or lower than about 1000 cfu/g, or lower than about 500 cfu/g. If a protein concentrate comprises any *Escherichia coli*, the bacteria may be present at such low levels as to be undetectable and/or noninfectious. If a protein concentrate comprises any *Salmonella spp.*, the bacteria may be present at such low levels as to be undetectable and/or noninfectious. If a protein concentrate comprises any yeast/mold, the microorganism count may be lower than about 500/g, or lower

than about 400/g, or lower than about 300/g, or lower than about 250/g, or lower than about 200/g, or lower than about 150/g, or lower than about 100/g, or lower than about 50/g.

In some embodiments, a protein concentrate may be packed and/or sealed in either an industry standard bag or drum of varying sizes. A sealing method of industry-standard grade may be used to ensure proper shelf-life and shipping conditions. A bag or drum may include printed instructions or specifications regarding, for example, its intended use, shelf-life, suggested storage conditions, shipping conditions, compositions, or the like, or a combination thereof. An antioxidant (e.g., rosemary extract) may be mixed with a protein concentrate before packaging, according to some embodiments.

Processing a First Solid and/or Solid Mixture

A first solid (e.g., first portion, second portion) and/or solid mixture (e.g., first portion, second portion) may be processed to generate one or more carbohydrate-rich products and/or a polysaccharide product (e.g., apiogalacturonan, oligogalacturonide, polygalacturonide). As described previously, a solid mixture may include one or more of a first solid (e.g., FIGURE 2, **231**), a first cake (e.g., **242**), a second cake (e.g., FIGURE 2, **247**), or any combination thereof that remain after one or more separation processes (e.g., FIGURE 2, **230/240/245**, FIGURE 5A **530/540/545**). Carbohydrate rich products may include a dry biocrude product suitable as a fuel feedstock or a carbohydrate-rich meal suitable as a human or animal feed supplement (e.g., Lemna meal). A polysaccharide product may include at least one apiogalacturonan, and/or at least one oligogalacturonide, and/or at least one polygalacturonide, or any combination thereof.

Generating a Dry Biocrude and/or a Carbohydrate-Rich Meal

A first solid and/or solid mixture may be processed (e.g., drying, pelletization), in some embodiments, to generate at least one of a dry biocrude and a carbohydrate-rich meal. According to some embodiments, processing a first solid and/or solid mixture involves drying and/or pelletization.

A process for generating a carbohydrate-rich product (e.g., dry biocrude and/or carbohydrate-rich) meal may be varied depending upon the specific characteristics desired, including, for example, moisture content, particle size, protein content, fat content, fiber content, ash content, shelf-life, pellet size, texture, or any combination thereof.

In some embodiments, a first solid and/or a solid mixture may be dried to reduce a moisture content of a resulting carbohydrate-rich product (e.g., dry biocrude and/or

carbohydrate-rich meal). A drying procedure, in some embodiments, may be performed in conjunction with (*e.g.*, prior to, following) other processing of a first solid and/or a solid mixture, such that the end product is a carbohydrate-rich product with a reduced moisture content. A drying procedure may be performed, in some embodiments, using a dryer mechanism including, for example, a spray dryer, double drum dryer, flash dryer, evaporator, or any combination thereof.

In some embodiments, an inlet temperature of a dryer mechanism (*e.g.*, the temperature at the entrance to a dryer) may be above or above 25° C, or above 50° C, or above 75° C, or above 100° C, or above 125° C, or above 150° C, or above 175° C, or above 200° C, or above 225° C, or above 250° C, or above 275° C, or above 300° C, or above 325° C, or above 350° C, or above 375° C, or above 400° C, or above 425° C, or above 450° C, or above 475° C, or above 500° C. An inlet temperature may be from about 25° C to about 50° C, or from about 50° C to about 75° C, or from about 75° C to about 100° C, or from about 100° C to about 125° C, or from about 125° C to about 150° C, or from about 150° C to about 175° C, or from about 175° C to about 200° C, or from about 200° C to about 225° C, or from about 225° C to about 250° C, or from about 250° C to about 275° C, or from about 275° C to about 300° C, or from about 300° C to about 325° C, or from about 325° C to about 350° C, or from about 350° C to about 375° C, or from about 375° C to about 400° C, or from about 400° C to about 425° C, or from about 425° C to about 450° C, or from about 450° C to about 475° C, or from about 475° C to about 500° C, or above 500° C, in some embodiments. An inlet temperature, according to some embodiments, may be from about 50° C to about 100° C, or from about 100° C to about 150° C, or from about 150° C to about 200° C, or from about 200° C to about 250° C, or from about 250° C to about 300° C, or from about 300° C to about 350° C, or from about 350° C to about 400° C, or from about 400° C to about 450° C, or from about 450° C to about 500° C, or above 500° C.

According to some embodiments, an outlet temperature of a dryer mechanism (*e.g.*, the temperature at the exit from a dryer) may be below about 300° C, or below about 275° C, or below about 250° C, or below about 225° C, or below about 200° C, or below about 175° C, or below about 150° C, or below about 125° C, or below about 100° C, or below about 75° C, or below about 50° C, or below about 25° C. An outlet temperature, in some embodiments, may be from about 300° C to about 275° C, or from about 275° C to about 250° C, or from about 250° C to about 225° C, or from about 225° C to about 200° C, or from about 200° C to about 175° C, or from about 175° C to about 150° C, or from about 150° C to about

125° C, or from about 125° C to about 100° C, or from about 100° C to about 75° C, or from about 75° C to about 50° C, or from about 50° C to about 25° C, or below about 25° C. In some embodiments, an outlet temperature may be from about 300° C to about 250° C, or from about 250° C to about 200° C, or from about 200° C to about 150° C, or from about 150° C to about 100° C, from about 100° C to about 50° C, or from about 50° C to about 25° C, or below about 25° C.

A volume of a first solid and/or a solid mixture may be mixed with a volume of a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal) prior to drying, in some embodiments. This process, known as back-mixing, may be employed when, for example, the moisture content of a first solid and/or a solid mixture exceeds the level that a dryer mechanism is capable of accepting. By back-mixing a carbohydrate-rich product with a first solid and/or a solid mixture the total moisture content may be kept within the specifications of a dryer mechanism, thereby reducing operational costs (*e.g.*, wear and tear on equipment).

A moisture content of a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal) may be below about 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the product, in some embodiments.

According to some embodiments a first solid and/or a solid mixture may be pelletized (*e.g.*, steam pelletization). Pelletization may be performed before or after a drying procedure, in some embodiments.

Lutein Rich Products

According to some embodiments, the present disclosure relates to processes for a production of lutein rich products from a harvested microcrop (*e.g.*, a photosynthetic aquatic species, an aquatic plant species, *Lemna*, algal species) according to specific example embodiments of the disclosure. Lutein rich products may include a dry biocrude product (*e.g.*, suitable as a fuel feedstock), or a soluble protein concentrate (*e.g.*, suitable as a human or an animal feed supplement), or a dry protein concentrate (*e.g.*, suitable as a human or an animal feed supplement). A dry biocrude may be used as fuel feedstock (*e.g.*, power plant, refinery, coker) for combustion, co-combustion with other hydrocarbon-based fuels (*e.g.*, coal), and feedstock for biofuel conversion and/or fermentation. Whereas a soluble protein

product, a dry protein concentrate, or any combination thereof may be suitable may be used as a feed or feed supplement for animal (*e.g.*, fish, swine, cattle) or human consumption. A lutein rich product may also be used in an animal bedding and/or litter.

5 In some embodiments, the present disclosure relates to processes, methods, and systems for a production of lutein rich products, a soluble protein, or a dry protein concentrate.

A soluble protein product or a dry protein concentrate, according to some embodiments, may comprise a lutein concentration of at least 100 mg/kg or at least 150 mg/kg, or at least 200 mg/kg, or at least 250 mg/kg, or at least 300 mg/kg, or at least 350
10 mg/kg, or at least 400 mg/kg, or at least 450 mg/kg, or at least 500 mg/kg, or at least 550 mg/kg, or at least 600 mg/kg, or at least 650 mg/kg, or at least 700 mg/kg, or at least 750 mg/kg. In some embodiments, a soluble protein product or a dry protein concentrate may comprise a lutein concentration of at least about 500 mg/kg, or at least about at least about 550 mg/kg, or at least about 560 mg/kg, or at least about 570 mg/kg, or at least about 580
15 mg/kg, at least about 585 mg/kg, at least about 590 mg/kg, at least about 595 mg/kg, at least about 600 mg/kg, at least about 605 mg/kg, at least about 610 mg/kg, at least about 615 mg/kg, at least about 620 mg/kg, at least about 625 mg/kg, and at least about 630 mg/kg, or at least about 640 mg/kg, or at least about 650 mg/kg, or at least about 660 mg, or at least about 670 mg/kg, or at least about 680 mg/kg, or at least about 690 mg/kg, or at least about 700
20 mg/kg.

Generating a Polysaccharide Product

A first solid and/or solid mixture may be processed (*e.g.*, extracted, separated), in some embodiments, to generate at least one polysaccharide product (*e.g.*, a product
25 comprising a polysaccharide). According to some embodiments, a first solid and/or solid mixture may be dried (*e.g.*, fully dried, partially dried) prior to processing (*e.g.*, extracting, separating). A polysaccharide product may include an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof.

According to some embodiments, processing a first solid and/or solid mixture may
30 involve at least one of: (1) extracting with an acid solution or (2) extracting with an oxalic acid solution or an oxalate solution to form a polysaccharide extract. A polysaccharide extract may include an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof. In some embodiments, at least a portion of a polysaccharide extract

may be concentrated (*e.g.*, evaporation). A polysaccharide extract may be separated, according to some embodiments, to form a polysaccharide product including at least one apiogalacturonan, at least one oligogalacturonide, and/or at least one polygalacturonide. For example, in some embodiments, a method for generating a polysaccharide product comprises

5 (a) extracting a first solid and/or a solid mixture with an acid solution to form a polysaccharide extract, (b) precipitating the polysaccharide extract with at least one solvent (*e.g.*, ethanol) to form a precipitant and a supernatant, (c) separating the precipitant from the supernatant by, for example, centrifugation or filtration to form a polysaccharide product.

(1) Extracting with an Acid Solution

10 In some embodiments, processing a first solid and/or solid mixture to form a polysaccharide extract and/or a polysaccharide product (*e.g.*, at least one apiogalacturonan, oligogalacturonide, and/or polygalacturonide) may involve extracting with an acid solution. Extracting with an acid solution may be performed by exposing (*e.g.*, submerging) a first solid and/or solid mixture to an acid solution (*e.g.*, hydrochloric acid). An acid solution, in

15 some embodiments, may be combined with a first solid and/or solid mixture to form a slurry. An acid solution may include at least one of hydrochloric acid, sulfuric acid, perchloric acid, hydroiodic acid, hydrobromic acid, *p*-toluenesulfonic acid, methanesulfonic acid, nitric acid, chloric acid, citric acid, and trifluoroacetic acid (TFA).

According to some embodiments, extracting with an acid may include exposing (*e.g.*, submerging) a first solid and/or solid mixture to an acid solution (*e.g.*, hydrochloric acid) until carbohydrates are no longer detectable (*e.g.*, phenol/sulfuric acid detection method; anion exchange liquid phase chromatography (HPAEC), gas phase chromatography (GPC)). Extracting with an acid, in some embodiments, may include exposing (*e.g.*, submerging) a first solid and/or solid mixture to an acid solution (*e.g.*, hydrochloric acid) for a specified

25 period of time. For example, a first solid and/or solid mixture may be exposed to an acid solution (*e.g.*, hydrochloric acid) for about 0.25 hours, or about 0.5 hours, or about 0.75 hours, or about 1 hour, or about 2 hours, or about 3 hours, or about 4 hours, or about 5 hours, or about 6 hours, or about 7 hours, or about 8 hours, or about 9 hours, or about 10 hours. In some embodiments, extracting with an acid may include moving (*e.g.*, agitating, stirring, propelling) at least a portion of a an acid solution at a specified time, intermittently, or

30 continually.

Acid extraction may be performed at any desirable temperature. According to some embodiments, acid extraction may be performed at room temperature, above room

temperatures, or below room temperature. In some embodiments acid extraction may be performed at (*e.g.*, an acid solution is maintained at a temperature of) a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Acid extraction may be performed, in some embodiments, at (*e.g.*, an acid solution is maintained at a temperature of) a temperature above about 30° C, or above about 35° C, or above about 40° C, or above about 45° C, or above about 50° C, or above about 55° C, or above about 60° C, or above about 65° C, or above about 70° C, or above about 75° C, or above about 80° C, or above about 85° C, or above about 90° C, or above about 95° C. In some embodiments, acid extraction may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, or between about 35° C and about 45° C, or between about 40° C and about 50° C, or between about 45° C and about 55° C, or between about 50° C and about 60° C, or between about 55° C and about 65° C, or between about 60° C and about 70° C, or between about 65° C and about 75° C, or between about 70° C and about 80° C, or between about 75° C and about 85° C, or between about 80° C and about 90° C, or between about 85° C and about 95° C, or between about 90° C and about 100° C.

According to some embodiments, a first solid and/or solid mixture may be exposed to an acid solution (*e.g.*, hydrochloric acid), wherein the acid solution may have a composition of acid (*e.g.*, hydrochloric acid) by weight/volume (w/v) of about 0.1%, or about 0.2%, or about 0.3%, or about 0.4%, or about 0.5%, or about 0.6%, or about 0.7%, or about 0.8%, or about 0.9%, or about 1%, or about 1.1%, or about 1.2%, or about 1.3%, or about 1.4%, or about 1.5%, or about 1.6%, or about 1.7%, or about 1.8%, or about 1.9%, or about 2%.

In some embodiments, acid extractions may be performed on a first solid and/or solid mixture that has undergone drying (*e.g.*, FIGURE 3, **335**). For example, a first solid and/or solid mixture that has undergone drying, may be extracted with an acid to dried solid ratio comprising about 15:1, about 14:1, about 13:1, about 12:1, about 11:1, about 10:1, about 9:1, about 8:1, about 7:1, about 6:1, about 5:1, about 4:1, about 3:1, about 2:1, about 1:1, about 1:2, about 1:3, about 1:4, about 1:5, about 1:6, about 1:7, about 1:8, about 1:9, about 1:10, about 1:11, about 1:12, about 1:13, about 1:14, or about 1:15.

Acid extraction may be performed on a first solid and/or solid mixture, according to some embodiments, at a temperature of about 85 ° C, wherein the first solid and/or solid mixture may be exposed to an acid solution (e.g., hydrochloric acid) with a composition of acid by weight volume (w/v) of about 0.4%; wherein the first solid and/or solid mixture may be exposed to the acid solution for about 0.5 hours.

According to some embodiments, an extraction acid may be removed from a solid and/or a solid mixture by any desired method. For example, an acid may be removed by actual removal from the material (e.g., evaporation) and/or it may be titrated to a desired pH (e.g., less acidic, neutral, or basic) by addition of a suitable base or buffer.

(2) Extracting with an Oxalic Acid Solution or an Oxalate Solution

In some embodiments, processing a first solid and/or solid mixture to form a polysaccharide product (e.g., at least one apiogalacturonan, oligogalacturonide, and/or polygalacturonides) may involve extracting with an oxalic acid solution or an oxalate solution (e.g., ammonium oxalate). Extracting with an oxalic acid solution or an oxalate solution may be performed by exposing (e.g., submerging) a first solid and/or solid mixture to an oxalic acid solution (e.g., 0.5% oxalic acid solution) or an oxalate solution (e.g., 1% ammonium oxalate solution). An oxalic acid solution or an oxalate solution, in some embodiments, may be combined with a first solid and/or solid mixture to form a slurry. In some embodiments, an oxalic acid solution may have a composition of oxalic acid by weight/volume (w/v) (e.g., with water as the solvent) of about 0.1%, or about 0.2%, or about 0.3%, or about 0.5%, or about 1%, or about 5%, or about 10%, or about 15%, or about 20%, or about 25%, or about 30%, or about 35%, or about 40%, or about 45%, or about 50%, or about 55%, or about 60%. An oxalate solution, in some embodiments, may have a composition of oxalate (exclusive of any counter-ion(s) that may be present) by weight/volume (w/v) (e.g., with water as the solvent) of about 0.3%, or about 0.4%, or about 0.5%, or about 0.6%, or about 0.7%, or about 0.8%, or about 0.9%, or about 1%, or about 1.1%, or about 1.2%, or about 1.3%, or about 1.4%, or about 1.5%, or about 1.6%, or about 1.7%, or about 1.8%, or about 1.9%, or about 2%.

According to some embodiments, extracting with an oxalic acid solution or an oxalate solution may include exposing (e.g., submerging) a first solid and/or solid mixture to an oxalic acid (e.g., 0.5% oxalic acid solution) or an oxalate solution (e.g., 1% ammonium oxalate solution) until carbohydrates are no longer detectable (e.g., phenol/sulfuric acid detection method; HPAEC, GPC). Extracting with an oxalic acid solution or an oxalate

solution, in some embodiments, may include exposing (*e.g.*, submerging) a first solid and/or solid mixture to an oxalic acid solution (*e.g.*, 0.5% oxalic acid) or an oxalate solution (*e.g.*, 1% ammonium oxalate) for a specified period of time. For example, a first solid and/or solid mixture may be exposed to an oxalate solution (*e.g.*, 1% ammonium oxalate) for about 1 hour or about 1.5 hours, or about 2 hours, or about 2.5 hours, or about 3 hours, or about 3.5 hours, or about 4 hours, or about 4.5 hours, or about 5 hours. In some embodiments, extracting with an oxalic acid solution or an oxalate solution may include moving (*e.g.*, agitating, stirring, propelling) at least a portion of the oxalic acid or the oxalate solution at a specified time, intermittently, or continually.

Extraction with an oxalic acid solution or an oxalate solution may be performed, according to some embodiments, at temperatures at or below room temperature. In some embodiments extraction with an oxalic acid solution or an oxalate solution may be performed at (*e.g.*, an oxalate solution is maintained at a temperature of) a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. In some embodiments, extraction with an oxalic acid solution or an oxalate solution may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C.

(3) Processing a Polysaccharide Extract

In some embodiments, at least a portion of a polysaccharide extract may be concentrated (*e.g.*, by evaporation, precipitation, dialysis, filtration, centrifugation, dewatering or any other desired water removal technique).

According to some embodiments, at least a portion of a polysaccharide extract may be concentrated by evaporation (*e.g.*, FIGURE 5A and 5B **595**). Evaporation may be performed by, for example, a thermal (evaporative) means such as: a rising film evaporator, a falling film evaporator, a natural circulation evaporator (vertical or horizontal), an agitated-film evaporator, a multiple-effect evaporator, by vacuum evaporation, by rotoevaporation, or any combination thereof.

Concentration of at least a portion of a polysaccharide extract may involve separating at least one apiogalacturonan and/or oligogalacturonide from the polysaccharide extract (*e.g.*, filtration, precipitation) to form a polysaccharide product, according to some embodiments.

According to some embodiments, concentration of at least a portion of a polysaccharide extract may involve precipitation (*e.g.*, ethanol). In some embodiments, at least a portion of a polysaccharide may be precipitated by adding a volume (*e.g.*, two-fold volume) of ethanol (*e.g.*, 50%, 60%, 70%, 80%, 90%, 95%, 98%, 99%) and isolating a pellet of polysaccharide by centrifugation. At least a portion of a polysaccharide may be precipitated using a ratio of at least one solvent (*e.g.*, ethanol) to a polysaccharide extract, the ratio comprising about 15:1, about 14:1, about 13:1, about 12:1, about 11:1, about 10:1, about 9:1, about 8:1, about 7:1, about 6:1, about 5:1, about 4:1, about 3:1, about 2:1, about 1:1, about 1:2, about 1:3, about 1:4, about 1:5, about 1:6, about 1:7, about 1:8, about 1:9, about 1:10, about 1:11, about 1:12, about 1:13, about 1:14, or about 1:15. In some embodiments, a portion of a polysaccharide extract may be precipitated (*i.e.*, recrystallized) at a temperature above about -20° C, above about -10° C, above about 0° C, above about 20° C, or above about 25° C, or above about 30° C, or above about 35° C, or above about 40° C, or above about 45° C, or above about 50° C, or above about 55° C, or above about 60° C, or above about 65° C, or above about 70° C, or above about 75° C, or above about 80° C, or above about 85° C, or above about 90° C, or above about 95° C, or above about 100° C, or above about 110° C, above about 120° C.

In some embodiments, concentration of at least a portion of a polysaccharide extract may involve ultrafiltration of the polysaccharide extract to form a polysaccharide product. Ultrafiltration may involve membrane filtration using pressure, concentration gradients, diafiltration with deionized (DI) water, or a combination thereof. Suitable nominal molecular weight cut-offs (NMWCO) for ultrafiltration of a polysaccharide extract may be, in some embodiments, at most about 30 kDa, or at most about 20 kDa, or at most about 15 kDa, or at most about 14 kDa, or at most about 13 kDa, or at most about 12 kDa, or at most about 11 kDa, or at most about 10 kDa, or at most about 9 kDa, or at most about 8 kDa, or at most about 7 kDa, or at most about 6 kDa, or at most about 5 kDa, or at most about 4 kDa, or at most about 3 kDa, or at most about 2 kDa, or at most about 1 kDa. In some embodiments, suitable NMWCO cut-offs for ultrafiltration may be in a range of at most about 1 kDa to at most about 10 kDa, at most about 2 kDa to at most about 10 kDa, at most about 3 kDa to at most about 10 kDa, at most about 3 kDa to at most about 15 kDa, or at most about 3 kDa to at most about 20 kDa, or at most about 3 kDa to at most about 30 kDa. In some embodiments, concentration of at least a portion of a polysaccharide extract may involve nanofiltration or reverse osmosis filtration of the polysaccharide extract to form a polysaccharide product. In

some embodiments, suitable filter sizes for nanofiltration may include $\leq$ about 0.01 μm , or $\leq$ about 0.009 μm , or $\leq$ about 0.008 μm , or $\leq$ about 0.007 μm , or $\leq$ about 0.006 μm , or $\leq$ about 0.005 μm , or $\leq$ about 0.004 μm , or $\leq$ about 0.003 μm , or $\leq$ about 0.002 μm , or $\leq$ about 0.001 μm . A nanofiltration filter may have a filter size of not more than about 0.01 μm , in some
 5 embodiments. According to some embodiments, suitable filter sizes for reverse osmosis filtration may include $\leq$ about 0.001 μm , $\leq$ about 0.0009 μm , $\leq$ about 0.0008 μm , $\leq$ about 0.0007 μm , $\leq$ about 0.0006 μm , $\leq$ about 0.0005 μm , $\leq$ about 0.0004 μm , $\leq$ about 0.0003 μm , $\leq$ about 0.0002 μm , or $\leq$ about 0.0001 μm . A reverse osmosis filter may have a filter size of not more than about 0.001 μm , in some embodiments.

Carbohydrate Rich Products

The present disclosure, in some embodiments, relates to processes for production of carbohydrate rich products (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) from a harvested microcrop (*e.g.*, photosynthetic aquatic
 15 species, aquatic plant species, *Lemna*, algal species) according to specific example embodiments of the disclosure. Carbohydrate rich products may include a dry biocrude product suitable as a fuel feedstock or a carbohydrate-rich meal suitable as an animal feed supplement. A dry biocrude has many potential uses including: fuel feedstock (*e.g.*, power plant, refinery, coker) for combustion; co-combustion with other hydrocarbon-based fuels
 20 (*e.g.*, coal); and feedstock for biofuel conversion and/or fermentation. A carbohydrate meal (*e.g.*, *Lemna* meal) may be used as a feed or feed supplement for animal (*e.g.*, fish, swine, cattle) or human consumption. A carbohydrate-rich meal may also be used in an animal bedding and/or litter. A carbohydrate-rich product, according to some embodiments, may include a solid fraction (*e.g.*, FIGURE 5B, 527), or a first solid (*e.g.*, FIGURE 4, 431), or a
 25 solid mixture, or a polysaccharide extract, or a polysaccharide product.

A carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have a protein content lower than about 50 wt. %, or lower than about 40 wt. %, or lower than about 30 wt. %, or lower than about 25 wt. %, or lower than about 20 wt. %, or lower than about 15 wt. %, or lower than about 14
 30 wt. %, or lower than about 13 wt. %, or lower than about 12 wt. %, or lower than about 11 wt. %, or lower than about 10 wt. %, or lower than about 5 wt. % by weight of the product. In some embodiments, a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have a protein content from about

1 wt. % to about 10 wt. %, or from about 10 wt. % to about 20 wt. %, or from about 20 wt. % to about 30 wt. %, or from about 30 wt. % to about 40 wt. %, or from about 40 wt. % to about 50 wt. % by weight of the product. In some embodiments, a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have a protein content from about 1 wt. % to about 50 wt. %, or from about 5 wt. % to about 40 wt. %, or from about 5 wt. % to about 30 wt. %, or from about 5 wt. % to about 20 wt. %, or from about 5 wt. % to about 15 wt. %, or from about 5 wt. % to about 10 wt. %, or from about 10 wt. % to about 50 wt. %, or from about 10 wt. % to about 40 wt. %, or from about 10 wt. % to about 30 wt. %, or from about 10 wt. % to about 20 wt. %, or from about 10 wt. % to about 15 wt. % by weight of the product. A carbohydrate-rich product may be further processed to meet a desired protein content (*e.g.*, higher or lower concentration, a desired amino acid composition).

In some embodiments a carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal polysaccharide extract, polysaccharide product) may have a fiber content lower than about 70%, or lower than about 60%, or lower than about 50 wt. %, or lower than about 40 wt. %, or lower than about 30 wt. %, or lower than about 20 wt. %, or lower than about 15 wt. %, or lower than about 10 wt. % by weight of the product. A carbohydrate-rich product, in some embodiments, may have a fiber content from about 1 wt. % to about 10 wt. %, or from about 10 wt. % to about 20 wt. %, or from about 15 wt. % to about 25 wt. %, or from about 20 wt. % to about 30 wt. %, or from about 25 wt. % to about 35 wt. %, or from about 30 wt. % to about 40 wt. %, or from about 35 wt. % to about 45 wt. %, or from about 40 wt. % to about 50 wt. %, or from about 45 wt. % to about 55 wt. %, or from about 50 wt. % to about 60 wt. %, or from about 55 wt. % to about 65 wt. % by weight of the product. A carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have a fiber content from about 5 wt. % to about 65 wt. %, or from about 5 wt. % to about 60 wt. %, or from about 5 wt. % to about 55 wt. %, or from about 5 wt. % to about 50 wt. %, or from about 5 wt. % to about 40 wt. %, or from about 10 wt. % to about 65 wt. %, or from about 10 wt. % to about 55 wt. %, or from about 10 wt. % to about 50 wt. %, or from about 10 wt. % to about 45 wt. %, or from about 10 wt. % to about 40 wt. %, or from about 20 wt. % to about 65 wt. %, or from about 20 wt. % to about 60 wt. %, or from about 20 wt. % to about 55 wt. %, or from about 20 wt. % to about 50 wt. %, or from about 20 wt. % to about 45 wt. %, or from about 20 wt. % to about 40 wt. %, or from about 30 wt. % to about 65 wt. %, or from about 30 wt. % to about 60 wt. %, or from about

30 wt. % to about 55 wt. %, or from about 30 wt. % to about 50 wt. %, or from about 30 wt. % to about 45 wt. %, or from about 30 wt. % to about 40 wt. %, or from about 40 wt. % to about 65 wt. %, or from about 40 wt. % to about 60 wt. %, or from about 40 wt. % to about 55 wt. %, or from about 40 wt. % to about 50 wt. %, or from about 40 wt. % to about 45 wt. % by weight of the product, according to some embodiments. According to some embodiments, a carbohydrate-rich product may be further processed to meet a desired fiber content (*e.g.*, higher or lower concentration, a desired fiber composition).

A carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have, in some embodiments, an ash content lower than 50 wt. %, or lower than about 40 wt. %, or lower than about 30 wt. %, or lower than about 25 wt. %, or lower than about 20 wt. %, or lower than about 15 wt. %, or lower than about 10 wt. %, or lower than about 5 wt. % by weight of the product. In some embodiments, a carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have an ash content from about 1 wt. % to about 10 wt. %, or from about 10 wt. % to about 20 wt. %, or from about 20 wt. % to about 30 wt. %, or from about 30 wt. % to about 40 wt. %, or from about 40 wt. % to about 50 wt. % by weight of the product. A carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have an ash content from about 1 wt. % to about 50 wt. %, or from about 2 wt. % to about 40 wt. %, or from about 3 wt. % to about 30 wt. %, or from about 3 wt. % to about 20 wt. %, or from about 3 wt. % to about 15 wt. %, or from about 3 wt. % to about 10 wt. %, or from about 5 wt. % to about 10 wt. %, or from about 5 wt. % to about 15 wt. %, or from about 5 wt. % to about 20 wt. % by weight of the product, according to some embodiments. A carbohydrate-rich product may be further processed to meet a desired ash content (*e.g.*, higher or lower concentration, a desired ash composition).

In some embodiments, a carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have a fat content lower than about 50 wt. %, or lower than about 40 wt. %, or lower than about 30 wt. %, or lower than about 25 wt. %, or lower than about 20 wt. %, or lower than about 15 wt. %, or lower than about 10 wt. %, or lower than about 5 wt. % by weight of the product. A carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have a fat content from about 1 wt. % to about 10 wt. %, or from about 5 wt. % to about 10 wt. %, or from about 10 wt. % to about 20 wt. %, or from about 20 wt. % to about

30 wt. %, or from about 30 wt. % to about 40 wt. %, or from about 40 wt. % to about 50 wt. % by weight of the product. According to some embodiments, a carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have a fat content from about 1 wt. % to about 50 wt. %, or from about 1 wt. % to about 40 wt. %, or from about 1 wt. % to about 30 wt. %, or from about 1 wt. % to about 20 wt. %, or from about 1 wt. % to about 15 wt. %, or from about 1 wt. % to about 10 wt. %, or from about 1 wt. % to about 5 wt. %, or from about 2 wt. % to about 40 wt. %, or from about 2 wt. % to about 30 wt. %, or from about 2 wt. % to about 20 wt. %, or from about 2 wt. % to about 15 wt. %, or from about 2 wt. % to about 10 wt. %, or from about 2 wt. % to about 5 wt. %, or from about 3 wt. % to about 30 wt. %, or from about 3 wt. % to about 20 wt. %, or from about 3 wt. % to about 15 wt. %, or from about 3 wt. % to about 10 wt. %, or from about 3 wt. % to about 5 wt. %, or from about 5 wt. % to about 10 wt. %, or from about 5 wt. % to about 15 wt. %, or from about 5 wt. % to about 20 wt. % by weight of the product. A carbohydrate-rich product may be further processed to meet a desired fat content (*e.g.*, higher or lower concentration, a desired fat composition).

A carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have, according to some embodiments, a carbohydrate content higher than about 30 wt. %, or higher than about 40 wt. %, or higher than about 50 wt. %, or higher than about 60 wt. %, or higher than about 65 wt. %, or higher than about 70 wt. %, or higher than about 75 wt. %, or higher than about 80 wt. %, or higher than about 85 wt. % by weight of a dry bio-crude. In some embodiments, a carbohydrate-rich product may have a carbohydrate content from about 30 wt. % to about 90 wt. %, or from about 40 wt. % to about 90 wt. %, or from about 50 wt. % to about 90 wt. %, or from about 60 wt. % to about 90 wt. %, or from about 70 wt. % to about 90 wt. %, or from about 80 wt. % to about 90 wt. %, or from about 30 wt. % to about 85 wt. %, or from about 40 wt. % to about 85 wt. %, or from about 50 wt. % to about 85 wt. %, or from about 60 wt. % to about 85 wt. %, or from about 70 wt. % to about 85 wt. %, or from about 30 wt. % to about 80 wt. %, or from about 40 wt. % to about 80 wt. %, or from about 50 wt. % to about 80 wt. %, or from about 60 wt. % to about 80 wt. %, or from about 70 wt. % to about 80 wt. % by weight of the product. A carbohydrate-rich product may be further processed to meet a desired carbohydrate content (*e.g.*, higher or lower concentration, a desired carbohydrate composition).

According to some embodiments, a carbohydrate-rich product may comprise at least one apiogalacturonan, at least one oligogalacturonan, at least one polygalacturonide, or a combination thereof. In some embodiments, a carbohydrate-rich product may have a concentration of at least one apiogalacturonan, at least one oligogalacturonan, at least one polygalacturonide, or a combination thereof of at least 5% (w/v), or at least 10% (w/v), or at least 15% (w/v), or at least 20% (w/v), or at least 25% (w/v), or at least 30% (w/v), or at least 35% (w/v), or at least 40% (w/v), or at least 45% (w/v), or at least 50% (w/v), or at least 55% (w/v), or at least 60% (w/v), or at least 65% (w/v), or at least 70% (w/v), or at least 75% (w/v), or at least 80% (w/v), or at least 85% (w/v), or at least 90% (w/v), or at least 95% (w/v). A first solid or a solid mixture, in some embodiments, may have a concentration of at least one apiogalacturonan, at least one oligogalacturonan, at least one polygalacturonide, or a combination thereof of up to or more than 30% (w/v). A first solid or a solid mixture, in some embodiments, may have a concentration of at least one apiogalacturonan, at least one oligogalacturonan, at least one polygalacturonide, or a combination thereof of up to or more than 35% (w/v).

According to some embodiments, a carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have a negligible amount of volatile matter. A carbohydrate-rich product may have a volatile matter content lower than about 1 wt. %, or lower than about 2 wt. %, or lower than about 5 wt. %, or lower than about 10 wt. %, or lower than about 15 wt. %, or lower than about 20 wt. % by weight of the product, in some embodiments. A carbohydrate-rich product may have, in some embodiments, a volatile matter content from about 1 wt. % to about 5 wt. %, or from about 1 wt. % to about 10 wt. %, or from about 1 wt. % to about 15 wt. %, or from about 1 wt. % to about 20 wt. %, from about 2 wt. % to about 10 wt. %, or from about 2 wt. % to about 15 wt. %, or from about 2 wt. % to about 20 wt. %, from about 5 wt. % to about 10 wt. %, or from about 5 wt. % to about 15 wt. %, or from about 5 wt. % to about 20 wt. % by weight of the product.

A carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) may have, in some embodiments, an energy content higher than about 3 MJ/kg, or higher than about 5 MJ/kg, or higher than about 8 MJ/kg, or higher than about 10 MJ/kg, or higher than about 12 MJ/kg, or higher than about 15 MJ/kg, or higher than about 50 MJ/kg, or higher than about 20 MJ/kg. A dry bio-crude

may be further processed to meet a desired energy content (*e.g.*, a higher or lower energy content, a desired energy content).

For example, a carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal, polysaccharide extract, polysaccharide product) produced by a process described herein may include the contents summarized in Table 3.

TABLE 3: Example Contents of Dry Biocrude and/or Carbohydrate-Rich Meal			
	Product A	Product B	Product C
Wt. % Solids	$\geq \sim 90$	$\geq \sim 88$ to ~ 92	$\geq \sim 90$
% Moisture	$\leq \sim 10$	$\leq \sim 8$ - ~ 12	$\leq \sim 10$
% Protein	$\leq \sim 20$	from ~ 10 to ~ 20	$\leq \sim 14$
% Fat	from ~ 5 to ~ 10	from ~ 5 to ~ 20	$\leq \sim 5$
% Ash	$\leq \sim 15$	from ~ 1 to ~ 20	$\leq \sim 12$
% Carbohydrate	$\geq \sim 50$	from ~ 60 to ~ 90	$\geq \sim 65$ to ~ 70
% Fiber	$\geq \sim 50$	$\geq \sim 40$	$\geq \sim 40$ to ~ 60
Energy (MJ/kg)	$\geq \sim 10$	$\geq \sim 10$	$\geq \sim 10$

Component Ratios

According to some embodiments, the present disclosure relates to compositions comprising a soluble microcrop protein. For example, a composition comprising a soluble microcrop protein, may comprise two or more of the following components: protein, lutein, fats, carbohydrates, polyphenols, and fiber, wherein each component may be present at a concentration disclosed herein. Table 4 and Table 5 illustrate example embodiments of compositions comprising a soluble microcrop protein. A soluble microcrop protein composition, for example, may include two or more such components at a desirable ratio to each other. A desirable ratio may be selected from any disclosed concentration. Using Table 4 to illustrate, a ratio of protein to fat may be about 30:0.2 (w/w), or about 30:2 (w/w), or about 30:20 (w/w), or about 60:0.2 (w/w), or about 60:2 (w/w), or about 60:20 (w/w), or about 90:0.2 (w/w), or about 90:2 (w/w), or about 90:20 (w/w). A ratio of any other component to protein may be derived from Table 4 in the same manner. Using Table 5 to illustrate, a ratio of carbohydrate to fat may be about 1:0.2 (w/w), or about 1:2 (w/w), or about 1:20 (w/w), about 10:0.2 (w/w), or about 10:2 (w/w), or about 10:20 (w/w), about 50:0.2 (w/w), or about 50:2 (w/w), or about 50:20 (w/w). A ratio of any other component to protein may be derived from Table 5 in the same manner.

According to some embodiments, the protein to polyphenol ratio of a composition comprising a soluble microcrop protein may be greater than the protein to polyphenol ratio in a living, intact microcrop (*e.g.*, *Lemna*), from which the soluble microcrop protein is derived. For example, the protein to polyphenol ratio of a composition comprising a soluble microcrop protein may be about 70:1 (w/w), about 65:1 (w/w), about 60:1 (w/w), about 55:1 (w/w), about 50:1 (w/w), about 45:1 (w/w), about 35:1 (w/w), about 25:1 (w/w), about 10:1 (w/w), about 5:1 (w/w), about 4:1 (w/w), about 3:1 (w/w), about 2:1 (w/w), and about 1.5:1 (w/w).

TABLE 4: Examples of Concentration Ratios (Protein: Fats, Lutein, Carbohydrates, Fiber, or Polyphenols) (w/w)		
Protein 30, 60, 90	0.2, 2, 20	Fats
	0.01, 0.05, 0.1	Lutein
	1, 10, 50	Carbohydrates
	1, 20, 70	Fiber
	0.05, 0.5, 2	Polyphenols

TABLE 5: Examples of Concentration Ratios (Carbohydrates: Fats, Lutein, Proteins, Fiber, or Polyphenols) (w/w)		
Carbohydrates 1, 10, 50	0.2, 2, 20	Fats
	0.01, 0.05, 0.1	Lutein
	30, 60, 90	Proteins
	1, 20, 70	Fiber
	0.05, 0.5, 2	Polyphenols

A Polysaccharide Product

The present disclosure, in some embodiments, relates to processes for production of a polysaccharide products (*e.g.*, apiogalacturonan, oligogalacturonide, and/or polygalacturonide) from a harvested microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) according to specific example embodiments of the disclosure. A polysaccharide products may include at least one apiogalacturonan and/or at least one oligogalacturonide. A polysaccharide product has many potential uses including: a cosmetic or dermatological additive (*e.g.*, topical formulations, serums, make-up, lotions, sunscreen, cleansers); cryoconservation applications; agricultural applications (*e.g.*, spray, treatment) for stimulation of plant disease resistance and defense mechanisms.

According to some embodiments, a polysaccharide product may have a concentration of at least one apiogalacturonan of at least 5% (w/v), or at least 10% (w/v), or at least 15% (w/v), or at least 20% (w/v), or at least 25% (w/v), or at least 30% (w/v), or at least 35% (w/v), or at least 40% (w/v), or at least 45% (w/v), or at least 50% (w/v), or at least 55% (w/v), or at least 60% (w/v), or at least 65% (w/v), or at least 70% (w/v), or at least 75% (w/v), or at least 80% (w/v), or at least 85% (w/v), or at least 90% (w/v), or at least 95% (w/v). A polysaccharide product, in some embodiments, may have a concentration of at least one apiogalacturonan of up to or more than 30% (w/v). In some embodiments, a polysaccharide product may have a concentration of at least one apiogalacturonan of up to or more than 35% (w/v). Concentrations recited in this paragraph may refer to a single apiogalacturonan or to the combined (total) concentration of two or more (up to all) apiogalacturonans present, according to some embodiments.

A polysaccharide product, in some embodiments, may have a concentration of at least one oligogalacturonide of at least 5% (w/v), or at least 10% (w/v), or at least 15% (w/v), or at least 20% (w/v), or at least 25% (w/v), or at least 30% (w/v), or at least 35% (w/v), or at least 40% (w/v), or at least 45% (w/v), or at least 50% (w/v), or at least 55% (w/v), or at least 60% (w/v), or at least 65% (w/v), or at least 70% (w/v), or at least 75% (w/v), or at least 80% (w/v), or at least 85% (w/v), or at least 90% (w/v), or at least 95% (w/v). A polysaccharide product, in some embodiments, may have a concentration of at least one oligogalacturonide up to or more than 30% (w/v). In some embodiments, a polysaccharide product may have a concentration of at least one oligogalacturonide of up to or more than 35% (w/v). Concentrations recited in this paragraph may refer to a single oligogalacturonide or to the combined (total) concentration of two or more (up to all) oligogalacturonides present, according to some embodiments.

A polysaccharide product, in some embodiments, may have a concentration of at least one polygalacturonide of at least 5% (w/v), or at least 10% (w/v), or at least 15% (w/v), or at least 20% (w/v), or at least 25% (w/v), or at least 30% (w/v), or at least 35% (w/v), or at least 40% (w/v), or at least 45% (w/v), or at least 50% (w/v), or at least 55% (w/v), or at least 60% (w/v), or at least 65% (w/v), or at least 70% (w/v), or at least 75% (w/v), or at least 80% (w/v), or at least 85% (w/v), or at least 90% (w/v), or at least 95% (w/v). A polysaccharide product, in some embodiments, may have a concentration of at least one polygalacturonide up to or more than 80% (w/v). In some embodiments, a polysaccharide product may have a concentration of at least one polygalacturonide of up to or more than 85% (w/v).

Concentrations recited in this paragraph may refer to a single polygalacturonide or to the combined (total) concentration of two or more (up to all) polygalacturonides present, according to some embodiments.

According to some embodiments, a concentration of a polysaccharide product may be determined by a phenol-sulfuric acid method, such as that described in Dubois, M., Gilles, K.A., Hamilton, J.K., *et al.*, *Anal. Chem.*, 1956, vol. 28, no. 2, 350-356. A concentration of a polysaccharide product may be determined using UV spectrophotometry, such as that described in Albalasmeh, A., Berhe, A., and Ghezzeher, T., *Carbohydrate Polymers*, 2013, vol. 97, no. 2, 253-261, in some embodiments. Any desired method may be used to determine a concentration of a polysaccharide product.

A monosaccharide composition of a polysaccharide product may be determined by HPAEC, according to some embodiments. For example, HPAEC may be performed using a Dionex CarboPac PA1 column with amperometric detection of polysaccharide hydrolysis where hydrolysis was performed under the following conditions: (1) hydrolysis with 2N Trifluoroacetic acid (TFA) at 100° C for 0.5 hours; (2) hydrolysis with 2N TFA at 100° C for 4 hours; (3) hydrolysis with 2N H₂SO₄ at 100° C for 6 hours; (4) hydrolysis with 2N H₂SO₄ at 100° C for 6 hours after overnight exposure to 26N H₂SO₄ at room temperature.

In some embodiments, a monosaccharide composition of a polysaccharide product may be determined by gas phase chromatography. For example, the relative composition of a monosaccharide of a polysaccharide product may be identified and quantified by (1) hydrolyzing the polysaccharide product to form monosaccharides by methanolysis; (2) trimethylsilylation of the monosaccharides to form a volatilized monosaccharides derivative; and (3) quantifying and identifying the volatilized monosaccharides as O-methylglycosides by gas phase chromatography.

Any desired method may be used to determine a composition of a polysaccharide product. According to some embodiments, a polysaccharide product may comprise homopolysaccharides, heteropolysaccharides, or combinations thereof. In some embodiments, a polysaccharide product may comprise at least one monosaccharide, the at least one monosaccharide comprising L-glucose, D-glucose, L-fructose, D-fructose, L-galactose, D-galactose, L-mannose, D-mannose, L-glucuronic acid, D-glucuronic acid, L-fructuronic acid, D-fructuronic acid, L-galacturonic acid, D-galacturonic acid, L-mannuronic acid, D-mannuronic acid, and combinations thereof. In some embodiments, a polysaccharide product may comprise alpha monosaccharides (*e.g.*, alpha-D-glucose), beta monosaccharides

(*e.g.*, beta-D-glucose), and combinations thereof. According to some embodiments, a polysaccharide product may comprise substituted carbohydrate monomers, the substituted carbohydrate monomers comprising glucuronide, galacturonide, fructuronide, mannuronide, and combinations thereof.

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Heat Exchange

According to some embodiments, thermal energy exchange mechanisms (*e.g.*, heat exchanger) may decrease an overall energy input required for the production of concentrated proteins and/or carbohydrate-rich products and/or a polysaccharide product from a microcrop (*e.g.*, *Lemna*). In some embodiments, a chilled stream (*e.g.*, recipient stream) may be directed to flow in proximity to a donor stream having thermal energy such that the chilled stream absorbs at least some of the donor stream thermal energy. A recipient stream, according to some embodiments, may be directed to flow in proximity to a donor stream having thermal energy such that the recipient stream absorbs at least some of the donor stream thermal energy.

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In some embodiments, a recipient stream may be at least one of a lysed biomass (*e.g.*, first portion, second portion), a juice fraction (*e.g.*, first portion, second portion), a first juice (*e.g.*, first portion, second portion), a first soluble protein fraction (*e.g.*, first portion, second portion), a first reject stream, a second soluble protein fraction (*e.g.*, first portion, second portion), a second reject stream, and a permeate. A recipient stream may be a chilled stream, in some embodiments. According to some embodiments at least one of a lysed biomass (*e.g.*, first portion, second portion), a juice fraction (*e.g.*, first portion, second portion), a first juice (*e.g.*, first portion, second portion), a first soluble protein fraction (*e.g.*, first portion, second portion), a first reject stream, a second soluble protein fraction (*e.g.*, first portion, second portion), a second reject stream, and a permeate may be chilled to form a chilled stream. A recipient stream (*e.g.*, a chilled stream) may have a temperature below room temperature (*e.g.*, about 12° C) at the time of use. In some embodiments, a recipient stream (*e.g.*, a chilled stream) may have a temperature below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C at the time of use. A recipient stream (*e.g.*, a chilled stream) may have a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or 15° C and about 25° C, or between about 20° C and about 30° C at the time of use, in some embodiments. In some embodiments, a recipient

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stream (*e.g.*, a chilled stream) may have a temperature of about 12° C. According to some embodiments, a recipient stream (*e.g.*, a chilled stream) may have a temperature that is lower than a donor stream.

A donor stream, in some embodiments, may comprise at least one of a lysed biomass (*e.g.*, first portion, second portion), a juice fraction (*e.g.*, first portion, second portion), or a first juice (*e.g.*, first portion, second portion). According to some embodiments, a donor stream may have a temperature that is higher than a recipient stream. In some embodiments, a donor stream may have a temperature above room temperature (*e.g.*, about 50° C). In some embodiments, a donor stream may have a temperature above about 20° C, or above about 25° C, or above about 30° C, or above about 35° C, or above about 40° C, or above about 45° C, or above about 50° C, or above about 55° C, or above about 60° C, or above about 65° C, or above about 70° C, or above about 75° C, or above about 80° C, or above about 85° C, or above about 90° C, or above about 95° C, or above about 100° C at the time of use. A donor stream may have a temperature between about 40° C and about 50° C, or between about 45° C and about 55° C, or between about 50° C and about 60° C at the time of use, in some embodiments. According to some embodiments, a donor stream may have a temperature between about 75° C and about 80° C, or between about 80° C and about 85° C, or between about 85° C and about 90° C, or between about 90° C and about 95° C, or between about 95° C and about 100° C. In some embodiments, a donor stream may have a temperature between about 50° C and about 80° C, or between about 55° C and about 85° C, or between about 60° C and about 90° C, or between about 65° C and about 95° C, or between about 70° C and about 100° C.

In some embodiments, a thermal energy may be generated by one or more processes during production of concentrated proteins and/or carbohydrate-rich products and/or a polysaccharide product from a microcrop (*e.g.*, *Lemna*). For example, a thermal energy may be generated by (1) drying a concentrated protein, (2) drying a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal), and/or (3) chilling at least one of a lysed biomass (*e.g.*, first portion, second portion), a juice fraction (*e.g.*, first portion, second portion), a first juice (*e.g.*, first portion, second portion), a first soluble protein fraction (*e.g.*, first portion, second portion), a first reject stream, a second soluble protein fraction (*e.g.*, first portion, second portion), a second reject stream, and a permeate to generate a chilled stream. According to some embodiments, a thermal energy may be generated in thermal communication with a heat exchanger. For example, chilling at least one of a juice fraction

(*e.g.*, first portion, second portion), a first juice (*e.g.*, first portion, second portion), a first soluble protein fraction (*e.g.*, first portion, second portion), a second soluble protein fraction (*e.g.*, first portion, second portion) may be performed in thermal communication with a heat exchanger. Heating at least one of a wash solution, a first reject stream, a second reject stream, and a permeate may be performed in thermal communication with a heat exchanger, in some embodiments. In some embodiments, drying a concentrated protein and/or drying a carbohydrate-rich product may be performed in thermal communication with a heat exchanger.

FIGURE 1

FIGURE 1 is a schematic diagram illustrating a process **100** for growing, harvesting, and separating a microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) for the production of protein concentrate and/or carbohydrate rich products according to a specific example embodiment of the disclosure. A microcrop (*e.g.*, *Lemna*) may be cultivated in a bioreactor system **105**, harvested **110**, and separated **125** to form a juice fraction **126** and a solid fraction **127**. In some embodiments, a juice fraction may be processed to produce one or more protein concentrate products and/or a solid fraction may be processed to produce one or more carbohydrate rich products. Protein concentrate products may include products suitable for animal feed and/or human consumption. Carbohydrate rich products may include a dry biocrude suitable as a fuel feedstock or a carbohydrate-rich meal suitable as a feed or supplement for animal and/or human consumption. A process **100** may be performed indoors, outdoors, and any combination thereof based, for example, on the specific environmental characteristics of the location(s).

As shown in **FIGURE 1**, a microcrop may be cultivated in a bioreactor system **105** (*e.g.*, open bioreactor, closed bioreactor). A bioreactor system may contain a growth medium (*e.g.*, water, a nutrient composition). In some embodiments, a bioreactor system, in some embodiments, may be configured to collect rainfall and/or to intake water from a source of recycled or ground water (*e.g.*, storm water, recycled water) or any other suitable source. A bioreactor system may be configured, in some embodiments, to insert additional nutrients (*e.g.*, nitrogen, phosphorus, potassium) or gases (*e.g.*, oxygen; carbon dioxide) at desired time indicators or in response to sensor readings. In some embodiments, a bioreactor system may comprise a monitoring system. A bioreactor system, in some embodiments, may monitor and adjust a thickness and distribution of a microcrop mat. For example, when a microcrop

reaches a desired thickness or distribution a bioreactor system may initiate harvest procedures.

As shown in FIGURE 1, at specified times (*e.g.*, based on environmental conditions) or after a microcrop develops desired characteristics (*e.g.*, mat thickness; mat distribution; maturation), a microcrop may be harvested **110** (*e.g.*, manual, automated) from a bioreactor system, forming a biomass **111**. An automated skimmer system, in some embodiments, may collect a microcrop from a bioreactor system and transfer a harvested microcrop (*e.g.*, via a pumping system) onto an inclined vibrating screen to separate a biomass from growth medium and debris. In some embodiments a microcrop may be harvested by vacuum skimming the microcrop from a bioreactor system through a stationary screen filter. A microcrop may be harvested manually, according to some embodiments. A biomass slurry, including a harvested microcrop (*e.g.*, *Lemna*) and a growth medium (*e.g.*, water), may be conveyed to an inclined screen, which may optionally vibrate, where a biomass (*e.g.*, microcrop) may be separated from the growth medium.

During harvesting **110**, a separated growth medium may be recycled **112** back into the bioreactor system or to an additional storage container (*e.g.*, container or pond), according to some embodiments. In some embodiments, at least about 40% (v/v), or at least about 50% (v/v), or at least about 60% (v/v), or at least about 70% (v/v), or at least about 80% (v/v), or at least about 90% (v/v), or at least about 95% (v/v) of a growth medium (*e.g.*, water) separated from a biomass may be recycled for future use.

As shown in FIGURE 1, a biomass **111** may go through a wash procedure **115** (*e.g.*, submerging, spraying, slurry) to remove debris, contaminants, microorganisms, and/or toxins. In some embodiments a wash procedure may be performed by exposing (*e.g.*, submerging, spraying) at least about one surface of a biomass to a wash solution (*e.g.*, water, growth medium, antimicrobial solution). A wash solution (*e.g.*, water, ozonated water), in some embodiments, may be combined with a biomass to form a slurry. According to some embodiments, a wash solution may comprise by volume at least about 10%, or at least about 20%, or at least about 30%, or at least about 40%, or at least about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90% recycled from another stage of process **100** (*e.g.*, recycled wash solution **116**, a reject stream from filtration stages (FIGURE 2A, **252**)). In some embodiments, a second wash solution (*e.g.*, water, ozonated water, recycled wash solution **116**) may be applied to a biomass. A third wash solution (*e.g.*, water, ozonated water, recycled wash solution) may be applied to a biomass, in

some embodiments. In some embodiments a first wash solution may be or may comprise a reject stream from a filtration process (*e.g.*, FIGURE 2A, **252**), a second wash solution may be or may comprise water, and a third wash solution may be or may comprise ozonated water. Some or all of a wash solution (*e.g.*, a first, second, and/or third wash solution), in
5 some embodiments, may be separated from a biomass (*e.g.*, using an inclined screen or vibratory screen).

In some embodiments, some or all of a wash solution, second wash solution, and/or third wash solution may be collected and reused/recycled **116/117**. By volume, at least about 40%, or at least about 50%, or at least about 60%, or at least about 70%, or at least about
10 80%, or at least about 90%, or at least about 95% of a wash solution, second wash solution, and/or third wash solution (*e.g.*, water) separated from the biomass may be recycled for future use (*e.g.*, recycled wash solution **116**, used as growth medium in the bioreactor system **117**), according to some embodiments.

As shown in FIGURE 1, a biomass, either washed or unwashed, may be lysed **120**
15 (*e.g.*, pressing, tearing, ultrasonic treatment). A lysing process may be achieved using, for example, a shear mill, a ball mill, colloid mill, knife mill, hammer mill, grinding mill, puree machine, filter press, or any combination thereof.

A lysed biomass **121** may be separated **125**, as shown in FIGURE 1, to form a juice fraction **126** and a solid fraction **127**. Separating **125** a lysed biomass **121** or biomass may
20 involve pressing (*e.g.*, belt press), centrifugation (*e.g.*, decanter centrifuge), filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating the lysed biomass and/or biomass include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

In some embodiments, a juice fraction **126**, may undergo a processing step for the reduction of at least one polyphenol (**a**). A polyphenol reduction process may comprise passing a juice fraction **126**, through a series (*e.g.*, at least two, at least three) of ion exchange resins. In some embodiments, a polyphenol reduction process may reduce a polyphenol (*e.g.*, a tannin) content of a juice fraction **126** by at least 5%, or at least 10%, or at least 15%, or at
25 least 20%, or at least 25%, or at least 30%, or at least 35%, or at least 40%, or at least 45%, or at least 50%, or at least 55%, or at least 60%, or at least 65%, or at least 70%.

According to some embodiments, a juice fraction **126** may undergo a solvent wash (**b**). A solvent wash of a juice fraction **126**, may comprise at least one solvent (*e.g.*, ethanol,

methanol), in some embodiments. According to some embodiments, a solvent wash of a juice fraction **126**, may result in a reduced fat content (e.g., about 2% of a dry protein concentrate or less by weight) and/or a reduced chlorophyll content (e.g., visually perceivable reduction in green coloration) compared to unwashed counterparts.

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FIGURES 2A and 2B

FIGURE 2A and **FIGURE 2B** illustrate separate embodiments for processing a microcrop (e.g. *Lemna*), for the production of protein concentrate and/or carbohydrate rich products. In some embodiments a microcrop may be harvested (e.g., **FIGURE 1, 110**) forming a biomass (e.g., **FIGURE 1, 111**).

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Washing a Biomass

A biomass may be washed **215** prior to processing, according to some embodiments. A wash procedure may remove debris, contaminants, nutrients, microorganisms, and/or toxins. A wash solution, in some embodiments, may be combined with a biomass to form a slurry. According to some embodiments, a wash solution (e.g., a first, second, and/or third wash solution) may have a temperature below room temperature (e.g., about 12° C). A wash solution, in some embodiments, may have a temperature above room temperature (e.g., 50° C). In some embodiments, a wash solution may comprise any desired portion of recycled fluid. For example, a wash solution may comprise at least about 10% (v/v), at least about 20% (v/v), at least about 30% (v/v), at least about 40% (v/v), at least about 50% (v/v), at least about 60% (v/v), at least about 70% (v/v), at least about 80% (v/v), or at least about 90% (v/v) recycled from another stage of the process (e.g., recycled wash solution **216**, a reject stream from filtration (e.g., **FIGURE 2A, 252**)). In some embodiments, a second wash solution (e.g., water, ozonated water, recycled wash solution **FIGURE 1, 116**) may be applied to a biomass. A third wash solution (e.g., water, ozonated water, recycled wash solution) may be applied to a biomass, in some embodiments. In some embodiments a first wash solution may be a reject stream from a filtration process (e.g., **FIGURE 2A, 252**), a second wash solution may be water, and a third wash solution may be ozonated water. Some or all of a wash solution (e.g., a first wash solution, second wash solution, and/or third wash solution) may be separated, in some embodiments, from a biomass (e.g., using an inclined screen or vibratory screen).

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Some or all of a wash solution, second wash solution, and/or third wash solution may be collected and/or reused, according to some embodiments. At least about 40%, or at least

about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, or at least about 95% of the wash solution and/or second wash solution separated from a biomass may be recycled for future use.

Lysing a Washed or Unwashed Biomass

5 As shown in FIGURE 2A and FIGURE 2B, a washed biomass may be lysed **220** to form a lysed biomass **221**. In some embodiments, a biomass (*e.g.*, unwashed) may be lysed to form a lysed biomass **221**. According to some embodiments, lysing may be achieved using a combination of mechanical (*e.g.*, milling), chemical (*e.g.*, pH adjustment), and/or ultrasonic (*e.g.*, sonication) methods. A lysing process may be achieved using, for example, a
10 shear mill, a ball mill, a colloid mill, a knife mill, a hammer mill, shear mill, a grinding mill, a puree machine, a filter press, or any combination thereof.

In some embodiments, lysing may be performed at temperatures below room temperature. A lysing fluid (*e.g.*, water, recycled water, reverse osmosis water) may be added to a biomass or microcrop before or during lysing, according to some embodiments. For
15 example, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90% of lysing fluid may be water generated as the result of reverse osmosis/nanofiltration of a filtration product (*e.g.*, FIGURE 3A, **362**). In some embodiments a lysing fluid may be at a temperature below room temperature (*e.g.*, about 12° C).

Separating a Biomass

20 As shown in FIGURE 2A and FIGURE 2B, a lysed biomass **221** may be separated **225** to generate a juice fraction **226** and a solid fraction **227**. In some embodiments, a biomass (*e.g.*, *Lemna*), a washed biomass, a lysed biomass **221**, or any combination thereof may be separated to generate a juice fraction and a solid fraction. A juice fraction **226** may
25 include a protein-rich liquid and/or at least some solid particles (*e.g.*, carbohydrates, fiber).

Separating **225** a lysed biomass **221** may involve pressing (*e.g.*, belt press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating **225** a biomass (*e.g.*, harvested microcrop), washed biomass, and/or lysed biomass include, for example, a decanter centrifuge, a belt press, a fan press, a
30 rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

Separating **225** may be performed at any desired temperature. Separating **225** may be performed at temperatures below room temperature (*e.g.*, 12° C), for example, to decrease proteolytic activity. In some embodiments, separating may be performed at a temperature

below about 40° C, below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating **225** may be performed, for example, at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C.

Separating a Solid Fraction

As shown in FIGURE 2A and FIGURE 2B, a solid fraction **227** may be further separated **230** to extract additional juice, forming a second juice **232** and a first solid **231**. A second juice may include a protein-rich liquid and/or at least some solid particles (*e.g.*, carbohydrates, fiber).

Separating a solid fraction to form a second juice and a first solid may involve pressing (*e.g.*, belt press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating a solid fraction include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

In some embodiments, as shown in FIGURE 2A and FIGURE 2B for example, other solid portions (*e.g.*, a first cake **242**, a second cake **247**) which are collected at other stages in the process (*e.g.*, separation of a juice fraction yields a first cake) may be combined with a first solid to form a solid mixture and the solid mixture may be further processed.

In some embodiments, a moisture content of a solid fraction and/or solid mixture is less than about 90%, or less than about 80%, or less than about 70%, or less than about 60%, or less than about 50%, or less than about 40%, or less than about 30%, or less than about 20%, or less than about 10% by weight.

Separating a Juice Fraction

As shown in FIGURE 2A and FIGURE 2B, a juice fraction **226** may be separated **240** to generate a first juice **241** and a first cake **242**. A first juice may include a soluble protein. Separating **240** a juice fraction, in some embodiments, may involve centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit operations may be used to separate a juice fraction including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter

centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

Separating **240** may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

Separating a First Cake and/or a Second Juice

As shown in FIGURE 2A and FIGURE 2B, a first cake **242** and a second juice **232** may be combined and further separated to form a third juice **246** and a second cake **247**. In some embodiments, a first cake and a second juice may be independently subjected to further separation. Separating a first cake, a second juice, or any combination thereof may involve centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit operations may be used to separate including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

Separating may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

As shown in FIGURE 2A and FIGURE 2B, in some embodiments a third juice **246** may be combined with a first juice **241** prior to further processing. A second cake **247** may

be combined with a first solid **231** to form a solid mixture, in some embodiments, prior to further processing.

Filtering a First Juice, a Third Juice, or any Combination Thereof

As shown in FIGURE 2A and FIGURE 2B, a first juice **241**, a third juice **246**, or any
5 combination thereof may be filtered to generate a soluble protein **251** and a reject stream **252**.
Filtration **250** may involve microfiltration, ultrafiltration, nanofiltration, or reverse osmosis
filtration.

Suitable filter sizes for microfiltration may include, in some embodiments, $\leq$ about 10
 μm , or $\leq$ about 5 μm , or $\leq$ about 3 μm , or $\leq$ about 2 μm , or $\leq$ about 1 μm , or $\leq$ about 0.5 μm ,
10 or $\leq$ about 0.4 μm , or $\leq$ about 0.3 μm , or $\leq$ about 0.2 μm , or $\leq$ about 0.1 μm . As shown in
FIGURE 2B, in some embodiments, a first juice, a third juice, or any combination thereof
may be filtered **250** using microfiltration to generate a soluble protein **251** in the permeate
and a reject stream product **253** in the retentate.

Ultrafiltration may involve membrane filtration using pressure, concentration
15 gradients, or a combination thereof. Suitable nominal molecular weight cut-offs (NMWCO)
for ultrafiltration may be, in some embodiments, at most in some embodiments, at most about
100 kDa, or at most about 90 kDa, or at most about 80 kDa, or at most about 70 kDa, or at
most about 60 kDa, or at most about 55 kDa, or at most about 50 kDa, or at most about 45
kDa, or at most about 40 kDa, or at most about 30 kDa, or at most about 20 kDa, or at most
20 about 15 kDa, or at most about 14 kDa, or at most about 13 kDa, or at most about 12 kDa, or
at most about 11 kDa, or at most about 10 kDa, or at most about 9 kDa, or at most about 8
kDa, or at most about 7 kDa, or at most about 6 kDa, or at most about 5 kDa, or at most about
4 kDa, or at most about 3 kDa, or at most about 2 kDa, or at most about 1 kDa. In some
embodiments, suitable NMWCO cut-offs for ultrafiltration may be in a range of at most
25 about 1 kDa to at most about 10 kDa, at most about 2 kDa to at most about 10 kDa, at most
about 3 kDa to at most about 10 kDa, at most about 3 kDa to at most about 15 kDa, or at most
about 3 kDa to at most about 20 kDa, or at most about 3 kDa to at most about 60 kDa, or at
most about 3 kDa to at most about 55 kDa, or at most about 10 kDa to at most about 55 kDa.
A first juice **241**, a third juice **246**, or any combination thereof may be filtered **250** using
30 ultrafiltration to generate a soluble protein **251** and a reject stream **252**, according to some
embodiments. A soluble protein may be in a retentate, as shown in FIGURE 2A, or a
permeate, as shown in FIGURE 2B, depending upon the molecular weight cut-offs for

ultrafiltration. When reject stream **252** is a permeate of ultrafiltration (*e.g.*, FIG 2A) it may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

In some embodiments, suitable filter sizes for nanofiltration may include $\leq$ about 0.01 μm , or $\leq$ about 0.009 μm , or $\leq$ about 0.008 μm , or $\leq$ about 0.007 μm , or $\leq$ about 0.006 μm , or $\leq$ about 0.005 μm , or $\leq$ about 0.004 μm , or $\leq$ about 0.003 μm , or $\leq$ about 0.002 μm , or $\leq$ about 0.001 μm . According to some embodiments, suitable filter sizes for reverse osmosis filtration may include $\leq$ about 0.001 μm , $\leq$ about 0.0009 μm , $\leq$ about 0.0008 μm , $\leq$ about 0.0007 μm , $\leq$ about 0.0006 μm , $\leq$ about 0.0005 μm , $\leq$ about 0.0004 μm , $\leq$ about 0.0003 μm , $\leq$ about 0.0002 μm , or $\leq$ about 0.0001 μm . As shown in FIGURE 2B, in some embodiments, a first juice, a third juice, or any combination thereof may be filtered **250** using nanofiltration or reverse osmosis filtration to generate a soluble protein **251** in the retentate and a permeate **252** of nanofiltered water or reverse osmosis water, respectively. In some embodiments reject stream **252** may be a permeate of nanofiltration or reverse osmosis filtration and may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

Buffers, protease inhibitors, anti-microbial agents, chelators (*e.g.*, EDTA), reducing agents, or any combination thereof may be added, in some embodiments, to a soluble protein **251**. A soluble protein may be chilled and/or stored at a temperature below about 30° C, or below about 25° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C, or below -2° C, or below -5° C, or below -10° C, in some embodiments. Chilling and/or storing a soluble protein at reduced temperatures may reduce degradation and/or improve protein recovery efficiency.

Reducing a Moisture Content of a Soluble Protein

In some embodiments a process may be used to reduce a moisture content of a soluble protein **251**. In some embodiments, an evaporation process may be used to reduce a moisture content of soluble protein. In some embodiments, a moisture content of a soluble protein **251** may be reduced by nanofiltration or reverse osmosis filtration. In some embodiments, a moisture content of soluble protein **251** may be reduced using nanofiltration with the soluble protein **251** in a retentate. A moisture content of soluble protein **251**, in some embodiments, may be reduced using reverse osmosis filtration with the soluble protein **251** in a retentate. A permeate of nanofiltration or reverse osmosis filtration may be recycled (*e.g.*, dilution fluid for lysis; wash solution), according to some embodiments.

Drying a Soluble Protein

A soluble protein **251** may be dried **255** to generate a dry protein concentrate, according to some embodiments. A drying procedure, in some embodiments, may reduce the moisture content of the soluble protein to a desired level (*e.g.*, higher or lower moisture content, a desired moisture content). A moisture content of a dry protein concentrate, in some embodiments, may be, for example, below about 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the dry protein concentrate. In some embodiments, the protein concentration of a dry protein concentrate may be from about 30% to about 95%, or from about 40% to about 90%, or from about 50% to about 85%, or from about 60% to about 80%, or from about 70% to about 75% by weight of the dry protein concentrate. A drying procedure may be performed using a mechanism including, for example, a spray dryer, double drum dryer, flash dryer, an evaporator, or any combination thereof.

Processing a First Solid and/or Solid Mixture

A first solid and/or solid mixture may be further processed to generate one or more carbohydrate-rich products. As described previously, a solid mixture may include one or more of a first solid **231**, a first cake **242**, a second cake **247**, or any combination thereof that remain after one or more separation processes **230/240/245**. Carbohydrate rich products may include a dry biocrude product suitable as a fuel feedstock or a carbohydrate-rich meal suitable as a human or animal feed supplement (*e.g.*, Lemna meal).

A first solid and/or solid mixture may be processed (*e.g.*, drying, pelletization), in some embodiments, to generate at least one of a dry biocrude and a carbohydrate-rich meal. According to some embodiments, processing a first solid and/or solid mixture involves drying and/or pelletization.

A process for generating a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich) meal may be varied depending upon the specific characteristics desired, including, for example, moisture content, particle size, protein content, fat content, fiber content, ash content, shelf-life, pellet size, texture, or any combination thereof.

In some embodiments, a first solid and/or a solid mixture may be dried to reduce a moisture content of a resulting carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal). A drying procedure, in some embodiments, may be performed in conjunction with (*e.g.*, prior to, following) other processing of a first solid and/or a solid mixture, such that the end product is a carbohydrate-rich product with a reduced moisture

content. A drying procedure may be performed, in some embodiments, using a dryer mechanism including, for example, a spray dryer, double drum dryer, flash dryer, evaporator, or any combination thereof.

A moisture content of a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal) may be below about 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the product, in some embodiments.

Polyphenol Reduction

In some embodiments, a juice fraction **226**, a first juice **241**, a second juice **232**, a third juice **246**, and/or a soluble protein **251** may undergo a processing step for the reduction of at least one polyphenol (a). A polyphenol reduction process may comprise passing a juice fraction **226**, a first juice **241**, a second juice **232**, a third juice **246**, and/or a soluble protein **251** through a series (*e.g.*, at least two, at least three) of ion exchange resins. In some embodiments, a polyphenol reduction process may reduce a polyphenol (*e.g.*, a tannin) content of a juice fraction **226**, a first juice **241**, a second juice **232**, a third juice **246**, and/or a soluble protein **251** by at least 5%, or at least 10%, or at least 15%, or at least 20%, or at least 25%, or at least 30%, or at least 35%, or at least 40%, or at least 45%, or at least 50%, or at least 55%, or at least 60%, or at least 65%, or at least 70%.

Solvent Wash

In some embodiments, a soluble protein **251** may undergo a solvent wash (b). Solvent washing (b) may also follow drying **255**. A solvent wash of a soluble protein product **251** and/or a solvent wash following drying **255**, may comprise at least one solvent (*e.g.*, ethanol, methanol), in some embodiments. According to some embodiments, a solvent wash of a soluble protein product **251** and/or a solvent wash following drying **255**, may result in a reduced fat content (*e.g.*, about 2% of a dry protein concentrate or less by weight) and/or a reduced chlorophyll content (*e.g.*, visually perceivable reduction in green coloration) compared to unwashed counterparts.

FIGURES 3A and 3B

FIGURE 3A and **FIGURE 3B** illustrate an embodiment for processing a microcrop (*e.g.*, *Lemna*) for production of protein concentrate and/or carbohydrate rich products. A microcrop may be harvested (*e.g.*, **FIGURE 1, 110**) forming a biomass (*e.g.*, **FIGURE 1, 111**).

Washing a Biomass

As shown in FIGURE 3A and FIGURE 3B, a biomass may be washed **315** prior to processing, according to some embodiments. A wash procedure may remove debris, contaminants, nutrients, microorganisms, and/or toxins. A wash solution, in some
5 embodiments, may be combined with a biomass to form a slurry. According to some embodiments, a wash solution (*e.g.*, a first, second, and/or third wash solution) may have a temperature below room temperature (*e.g.*, about 12° C). A wash solution, in some embodiments, may have a temperature above room temperature (*e.g.*, 50° C). In some
10 embodiments, a wash solution may comprise any desired portion of recycled fluid. For example, a wash solution may comprise at least about 10% (v/v), at least about 20% (v/v), at least about 30% (v/v), at least about 40% (v/v), at least about 50% (v/v), at least about 60% (v/v), at least about 70% (v/v), at least about 80% (v/v), or at least about 90% (v/v) recycled
15 from another stage of the process (*e.g.*, recycled wash solution **316**, a first reject stream from filtration (*e.g.*, FIGURE 3A, **352**), a second reject stream from filtrations (*e.g.*, FIGURE 3A, **357**)). In some embodiments, a second wash solution (*e.g.*, water, ozonated water, recycled wash solution FIGURE 1, **316**) may be applied to a biomass. A third wash solution (*e.g.*, water, ozonated water, recycled wash solution) may be applied to a biomass, in some
20 embodiments. In some embodiments a first wash solution may be a reject stream from a filtration process (*e.g.*, FIGURE 3A, **352**, **357**), a second wash solution may be water, and a third wash solution may be ozonated water. Some or all of a wash solution (*e.g.*, a first wash solution, second wash solution, and/or third wash solution) may be separated, in some
embodiments, from a biomass (*e.g.*, using an inclined screen or vibratory screen).

Some or all of a wash solution, second wash solution, and/or third wash solution may be collected and/or reused, according to some embodiments. At least about 40%, or at least
25 about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, or at least about 95% of the wash solution and/or second wash solution separated from a biomass may be recycled for future use.

Lysing a Washed or Unwashed Biomass

As shown in FIGURE 3A and FIGURE 3B, a washed biomass may be lysed **320** to
30 form a lysed biomass **321**. In some embodiments, a biomass (*e.g.*, unwashed) may be lysed to form a lysed biomass **321**. According to some embodiments, lysing may be achieved using a combination of mechanical (*e.g.*, milling), chemical (*e.g.*, pH adjustment), and/or ultrasonic (*e.g.*, sonication) methods. A lysing process may be achieved using, for example, a

shear mill, a ball mill, a colloid mill, a knife mill, a hammer mill, a grinding mill, a puree machine, a filter press, or any combination thereof.

In some embodiments, lysing may be performed at temperatures below room temperature. A lysing fluid (*e.g.*, water, recycled water, reverse osmosis water) may be added to a biomass or microcrop before or during lysing, according to some embodiments. As shown in FIGURE 3A and FIGURE 3B, a permeate **362** from reverse osmosis/nan-filtration of a second soluble protein may be recycled as a lysing fluid. For example, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90% of lysing fluid may be water generated as the result of reverse osmosis/nanofiltration of a filtration product (*e.g.*, FIGURE 3A, **362**). In some embodiments a lysing fluid may be at a temperature below room temperature (*e.g.*, about 12° C). A lysing fluid may be at a temperature above room temperature (*e.g.*, 50 °C).

Separating a Biomass

As shown in FIGURE 3A and FIGURE 3B, a lysed biomass **321** may be separated **325** to generate a juice fraction **326** and a solid fraction **327**. In some embodiments, a biomass (*e.g.*, *Lemna*), a washed biomass, a lysed biomass **321**, or any combination thereof may be separated to generate a juice fraction and a solid fraction. A juice fraction **326** may include a protein-rich liquid and/or at least some solid particles (*e.g.*, carbohydrates, fiber).

Separating **325** a lysed biomass **321** may involve pressing (*e.g.*, belt press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating **325** a biomass (*e.g.*, harvested microcrop), washed biomass, and/or lysed biomass include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

Separating **325** may be performed at any desired temperature. Separating **325** may be performed at temperatures below room temperature (*e.g.*, 12° C), for example, to decrease proteolytic activity. In some embodiments, separating may be performed at a temperature below about 40° C, below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 0° C. Separating **325** may be performed, for example, at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25°

C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C.

Separating a Solid Fraction

As shown in FIGURE 3, a solid fraction **327** may be further separated **330** to extract
5 additional juice, forming a second juice **332** and a first solid **331**. A second juice may include a protein-rich liquid and/or at least some solid particles (*e.g.*, carbohydrates, fiber).

Separating a solid fraction to form a second juice and a first solid may involve pressing (*e.g.*, belt press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating a solid fraction include, for example,
10 a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

In some embodiments, as shown in FIGURE 3 for example, other solid fractions (*e.g.*, a first cake **342**, a second cake **347**) which are collected at other stages in the process (*e.g.*, separation of a juice fraction yields a first cake) may be combined with a first solid to form a
15 solid mixture and the solid mixture may be further processed.

In some embodiments, a moisture content of a solid fraction and/or solid mixture is less than about 90%, or less than about 80%, or less than about 70%, or less than about 60%, or less than about 50%, or less than about 40%, or less than about 30%, or less than about 20%, or less than about 10% by weight.

Separating a Juice Fraction

As shown in FIGURE 3A and FIGURE 3B, a juice fraction **326** may be separated to generate a first juice **341** and a first cake **342**. A first juice may include a soluble protein. Separating a juice fraction, in some embodiments, may involve centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit
25 operations may be used to separate a juice fraction including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

Separating may be performed, according to some embodiments, at temperatures
30 below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below

about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

5 Separating a First Cake and/or a Second Juice

As shown in FIGURE 3A and FIGURE 3B, a first cake **342** and a second juice **332** may be combined and further separated to form a third juice **346** and a second cake **347**. In some embodiments, a first cake and a second juice may be independently subjected to further separation. Separating a first cake, a second juice, or any combination thereof may involve
10 centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit operations may be used to separate including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

15 Separating may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below
20 about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

As shown in FIGURE 3A and FIGURE 3B, in some embodiments a third juice **346**
25 may be combined with a first juice **341** prior to further processing. A second cake **347** may be combined with a first solid **331** to form a solid mixture, in some embodiments, prior to further processing.

Filtering a First Juice, a Third Juice, or any Combination Thereof

As shown in FIGURE 3A and FIGURE 3B, a first juice **341**, a third juice **346**, or any
30 combination thereof may be filtered to generate a first soluble protein **351** and a first reject stream **352**. Filtration **350** may involve microfiltration, ultrafiltration, nanofiltration, or reverse osmosis filtration.

Suitable filter sizes for microfiltration may include, in some embodiments, $\leq$ about 10 μm , or $\leq$ about 5 μm , or $\leq$ about 3 μm , or $\leq$ about 2 μm , or $\leq$ about 1 μm , or $\leq$ about 0.5 μm , or $\leq$ about 0.4 μm , or $\leq$ about 0.3 μm , or $\leq$ about 0.2 μm , or $\leq$ about 0.1 μm . As shown in FIGURE 3B, in some embodiments, a first juice, a third juice, or any combination thereof may be filtered **350** using microfiltration to generate a first soluble protein **351** in a permeate and a reject stream product **353** in the retentate.

Ultrafiltration may involve membrane filtration using pressure, concentration gradients, or a combination thereof. Suitable NMWCOs for ultrafiltration may be, in some embodiments, at most in some embodiments, at most about 100 kDa, or at most about 90 kDa, or at most about 80 kDa, or at most about 70 kDa, or at most about 60 kDa, or at most about 55 kDa, or at most about 50 kDa, or at most about 45 kDa, or at most about 40 kDa, or at most about 30 kDa, or at most about 20 kDa, or at most about 15 kDa, or at most about 14 kDa, or at most about 13 kDa, or at most about 12 kDa, or at most about 11 kDa, or at most about 10 kDa, or at most about 9 kDa, or at most about 8 kDa, or at most about 7 kDa, or at most about 6 kDa, or at most about 5 kDa, or at most about 4 kDa, or at most about 3 kDa, or at most about 2 kDa, or at most about 1 kDa. In some embodiments, suitable NMWCO cut-offs for ultrafiltration may be in a range of at most about 1 kDa to at most about 10 kDa, at most about 2 kDa to at most about 10 kDa, at most about 3 kDa to at most about 10 kDa, at most about 3 kDa to at most about 15 kDa, or at most about 3 kDa to at most about 20 kDa, or at most about 3 kDa to at most about 60 kDa, or at most about 3 kDa to at most about 55 kDa, or at most about 10 kDa to at most about 55 kDa.

A first juice **341**, a third juice **346**, or any combination thereof may be filtered **350** using ultrafiltration to generate a first soluble protein **351** and a reject stream **352**, according to some embodiments. A first soluble protein may be in a retentate, as shown in FIGURE 3A, or a permeate, as shown in FIGURE 3B, depending upon the molecular weight cut-offs for ultrafiltration. When first reject stream **352** is a permeate of ultrafiltration (*e.g.*, FIG 3A) it may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

In some embodiments, suitable filter sizes for nanofiltration may include $\leq$ about 0.01 μm , or $\leq$ about 0.009 μm , or $\leq$ about 0.008 μm , or $\leq$ about 0.007 μm , or $\leq$ about 0.006 μm , or $\leq$ about 0.005 μm , or $\leq$ about 0.004 μm , or $\leq$ about 0.003 μm , or $\leq$ about 0.002 μm , or $\leq$ about 0.001 μm . According to some embodiments, suitable filter sizes for reverse osmosis filtration may include $\leq$ about 0.001 μm , $\leq$ about 0.0009 μm , $\leq$ about 0.0008 μm , $\leq$ about

0.0007 μm , $\leq$ about 0.0006 μm , $\leq$ about 0.0005 μm , $\leq$ about 0.0004 μm , $\leq$ about 0.0003 μm , $\leq$ about 0.0002 μm , or $\leq$ about 0.0001 μm . As shown in FIGURE 3B, in some embodiments, a first juice, a third juice, or any combination thereof may be filtered **350** using nanofiltration or reverse osmosis filtration to generate a first soluble protein **351** in the retentate and a permeate **352** of nanofiltered water or reverse osmosis water, respectively. In some embodiments first reject stream **352** may be a permeate of nanofiltration or reverse osmosis filtration and may be recycled to a bioreactor system for cultivation of a microcrop (e.g., FIGURE 1, **105**).

Buffers, protease inhibitors, anti-microbial agents, chelators (e.g., EDTA), reducing agents, or any combination thereof may be added, in some embodiments, to a soluble protein **351**. A soluble protein may be chilled and/or stored at a temperature below about 30° C, or below about 25° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C, or below -2° C, or below -5° C, or below -10° C, in some embodiments. Chilling and/or storing a soluble protein at reduced temperatures may reduce degradation and/or improve protein recovery efficiency.

Filtering a First Soluble Protein

As shown in FIGURE 3A and FIGURE 3B, first soluble protein **351** may be filtered **355** to generate a second soluble protein **356** and a second reject stream **357**. Filtration **355** may involve microfiltration, ultrafiltration, nanofiltration, or reverse osmosis filtration.

Suitable filter sizes for microfiltration may include, in some embodiments, $\leq$ about 10 μm , or $\leq$ about 5 μm , or $\leq$ about 3 μm , or $\leq$ about 2 μm , or $\leq$ about 1 μm , or $\leq$ about 0.5 μm , or $\leq$ about 0.4 μm , or $\leq$ about 0.3 μm , or $\leq$ about 0.2 μm , or $\leq$ about 0.1 μm . In some embodiments, a first soluble protein may be filtered **355** using microfiltration to generate a second soluble protein **356** in the permeate and a reject stream product in the retentate.

Ultrafiltration may involve membrane filtration using pressure, concentration gradients, or a combination thereof. Suitable NMWCOs for ultrafiltration may be, in some embodiments, at most in some embodiments, at most about 100 kDa, or at most about 90 kDa, or at most about 80 kDa, or at most about 70 kDa, or at most about 60 kDa, or at most about 55 kDa, or at most about 50 kDa, or at most about 45 kDa, or at most about 40 kDa, or at most about 30 kDa, or at most about 20 kDa, or at most about 15 kDa, or at most about 14 kDa, or at most about 13 kDa, or at most about 12 kDa, or at most about 11 kDa, or at most about 10 kDa, or at most about 9 kDa, or at most about 8 kDa, or at most about 7 kDa, or at

most about 6 kDa, or at most about 5 kDa, or at most about 4 kDa, or at most about 3 kDa, or at most about 2 kDa, or at most about 1 kDa. In some embodiments, suitable NMWCO for ultrafiltration may be in a range of at most about 1 kDa to at most about 10 kDa, at most about 2 kDa to at most about 10 kDa, at most about 3 kDa to at most about 10 kDa, at most about 3 kDa to at most about 15 kDa, or at most about 3 kDa to at most about 20 kDa, or at most about 3 kDa to at most about 60 kDa, or at most about 3 kDa to at most about 55 kDa, or at most about 10 kDa to at most about 55 kDa.

A first soluble protein may be filtered **355** using ultrafiltration to generate a second soluble protein **355** and a reject stream **357**, according to some embodiments. A second soluble protein may be in a retentate, as shown in FIGURE 3A and FIGURE 3B, or a permeate (not shown) depending upon the molecular weight cut-offs for ultrafiltration. When second reject stream **357** is a permeate of ultrafiltration (*e.g.*, FIG 3A) it may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

In some embodiments, suitable filter sizes for nanofiltration may include $\leq$ about 0.01 μm , or $\leq$ about 0.009 μm , or $\leq$ about 0.008 μm , or $\leq$ about 0.007 μm , or $\leq$ about 0.006 μm , or $\leq$ about 0.005 μm , or $\leq$ about 0.004 μm , or $\leq$ about 0.003 μm , or $\leq$ about 0.002 μm , or $\leq$ about 0.001 μm . According to some embodiments, suitable filter sizes for reverse osmosis filtration may include $\leq$ about 0.001 μm , $\leq$ about 0.0009 μm , $\leq$ about 0.0008 μm , $\leq$ about 0.0007 μm , $\leq$ about 0.0006 μm , $\leq$ about 0.0005 μm , $\leq$ about 0.0004 μm , $\leq$ about 0.0003 μm , $\leq$ about 0.0002 μm , or $\leq$ about 0.0001 μm . As shown in FIGURE 3A and FIGURE 3B, in some embodiments, a first soluble protein may be filtered **355** using nanofiltration or reverse osmosis filtration to generate a second soluble protein **356** in the retentate and a permeate **357** of nanofiltered water or reverse osmosis water, respectively. In some embodiments second reject stream **357** may be a permeate of nanofiltration or reverse osmosis filtration and may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

According to some embodiments, diafiltration may be used to further eliminate permeable solutes from a first soluble protein **351**, a second soluble protein **356**, or a combination thereof. In some embodiments, diafiltration may increase a protein purity of a first soluble protein **351**, a second soluble protein **356**, or a combination thereof. When second reject stream **357** is a permeate of ultrafiltration (*e.g.*, FIG 3A) it may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

In some embodiments buffers, protease inhibitors, anti-microbial agents, chelators (*e.g.*, EDTA), reducing agents, or any combination thereof may be added to the second

soluble protein **356**. A second soluble protein may be chilled and/or stored at a temperature below about 30° C, or below about 25° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C, or below -2° C, or below -5° C, or below -10° C. Chilling and/or storing the second soluble protein at reduced temperatures may reduce degradation and/or improve protein recovery efficiency.

Reducing a Moisture Content of a First Soluble Protein and/or Second Soluble Protein

In some embodiments a process may be used to reduce a moisture content of a first soluble protein **351** and/or a second soluble protein **356**.

In some embodiments an evaporation process may be used to reduce a moisture content of a first soluble protein **351** or a second soluble protein **356** and generate a concentrated protein **361**. Reducing a moisture content of a first soluble protein **351** or a second soluble protein **356** may reduce capital and operational expenditures, for example, by reducing an energy needed to dry a concentrated protein **361**. Evaporation may be performed by, for example, a thermal (evaporative) means such as: a rising film evaporator, a falling film evaporator, a natural circulation evaporator (vertical or horizontal), an agitated-film evaporator, a multiple-effect evaporator, by vacuum evaporation, or any combination thereof.

As shown in FIGURE 3, a moisture content of a first soluble protein **351** or a second soluble protein **356** may be reduced, in some embodiments, by nanofiltration or reverse osmosis filtration. In some embodiments, a moisture content of a first soluble protein **351** or a second soluble protein **356** may be reduced using nanofiltration with the first soluble protein **351** or a second soluble protein **356**, respectively, in a retentate. A moisture content of a first soluble protein **351** or a second soluble protein **356**, in some embodiments, may be reduced using reverse osmosis filtration with the first soluble protein **351** or a second soluble protein **356**, respectively in a retentate. A permeate **362** of nanofiltration or reverse osmosis filtration may be recycled (*e.g.*, dilution fluid for lysis; wash solution), according to some embodiments. In some embodiments, reverse osmosis filtration and/or nanofiltration may reduce the concentration of undesirable dissolved compounds (*e.g.*, salts, calcium ions) in the resulting concentrated protein **361**.

Drying a Soluble Protein

A concentrated protein **361** may be dried **365** to generate a dry protein concentrate, according to some embodiments. A drying procedure, in some embodiments, may reduce the moisture content of a concentrated protein **361** to a desired level (*e.g.*, higher or lower

moisture content, a desired moisture content). A moisture content of a dry protein concentrate, in some embodiments, may be, for example, below about 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the dry protein concentrate. In some embodiments, the protein concentration of a dry protein concentrate may be from about 30% to about 95%, or from about 40% to about 90%, or from about 50% to about 85%, or from about 60% to about 80%, or from about 70% to about 75% by weight of the dry protein concentrate. A drying procedure may be performed using a mechanism including, for example, a spray dryer, double drum dryer, flash dryer, an evaporator, or any combination thereof.

Processing a First Solid and/or Solid Mixture

A first solid and/or solid mixture may be further processed to generate one or more carbohydrate-rich products. As described previously, a solid mixture may include one or more of a first solid **331**, a first cake **342**, a second cake **347**, or any combination thereof that remain after one or more separation processes **330/340/345**. Carbohydrate rich products may include a dry biocrude product suitable as a fuel feedstock or a carbohydrate-rich meal suitable as a human or animal feed supplement (*e.g.*, Lemna meal).

A first solid and/or solid mixture may be processed (*e.g.*, drying, pelletization), in some embodiments, to generate at least one of a dry biocrude and a carbohydrate-rich meal. According to some embodiments, processing a first solid and/or solid mixture involves drying and/or pelletization.

A process for generating a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich) meal may be varied depending upon the specific characteristics desired, including, for example, moisture content, particle size, protein content, fat content, fiber content, ash content, shelf-life, pellet size, texture, or any combination thereof.

In some embodiments, a first solid and/or a solid mixture may be dried to reduce a moisture content of a resulting carbohydrate-rich product (*e.g.*, a dry biocrude, a carbohydrate-rich meal, a polysaccharide extract, a polysaccharide product). A drying procedure, in some embodiments, may be performed in conjunction with (*e.g.*, prior to, following) other processing of a first solid and/or a solid mixture, such that the end product is a carbohydrate-rich product with a reduced moisture content. A drying procedure may be performed, in some embodiments, using a dryer mechanism including, for example, a spray dryer, double drum dryer, flash dryer, evaporator, or any combination thereof.

A moisture content of a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal) may be below about 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the product, in some embodiments.

Polyphenol Reduction

In some embodiments, a juice fraction **326**, a first juice **341**, a second juice **332**, a third juice **346**, a first soluble protein **351**, a second soluble protein **356**, and/or a concentrated protein **361** may undergo a processing step for the reduction of at least one polyphenol (**a**). A polyphenol reduction process may comprise passing a juice fraction **326**, a first juice **341**, a second juice **332**, a third juice **346**, and/or a soluble protein **351** through a series (*e.g.*, at least two, at least three) of ion exchange resins. In some embodiments, a polyphenol reduction process may reduce a polyphenol (*e.g.*, a tannin) content of a juice fraction **326**, a first juice **341**, a second juice **332**, a third juice **346**, and/or a soluble protein **351** by at least 5%, or at least 10%, or at least 15%, or at least 20%, or at least 25%, or at least 30%, or at least 35%, or at least 40%, or at least 45%, or at least 50%, or at least 55%, or at least 60%, or at least 65%, or at least 70%.

Solvent Wash

In some embodiments, a first soluble protein **351**, a second soluble protein **356**, and/or a concentrated protein **361** may undergo a solvent wash (**b**). Solvent washing (**b**) may also follow drying **365**. A solvent wash of a soluble protein product **351** and/or a solvent wash following drying **355**, may comprise at least one solvent (*e.g.*, ethanol, methanol), in some embodiments. According to some embodiments, a solvent wash of a soluble protein product **351** and/or a solvent wash following drying **365**, may result in a reduced fat content (*e.g.*, about 2% of a dry protein concentrate or less by weight) and/or a reduced chlorophyll content (*e.g.*, visually perceivable reduction in green coloration) compared to unwashed counterparts.

FIGURE 4

FIGURE 4 is a flow diagram illustrating a process for continuously producing a protein concentrate and/or carbohydrate rich products from a biomass according to a specific example embodiment of the disclosure. In some embodiments a process for growing, harvesting, and separating a microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) may be multiple cycles or a continuous process for the

production of protein concentrate (*e.g.*, soluble protein, dry protein concentrate) and/or carbohydrate rich products such that byproducts of an earlier cycle of the process may be recycled into one or more subsequent cycles of the process. Recycling of one or more by-products may reduce the overall water requirement for a process. In some embodiments, a process may be configured to minimize the net energy input required (*i.e.*, energy conservation).

A microcrop may be harvested (*e.g.*, FIGURE 1, **110**) forming a biomass (*e.g.*, FIGURE 1, **111**).

Washing a Biomass

As shown in FIGURE 4, a biomass may be washed **415** prior to processing, according to some embodiments. A wash procedure may remove debris, contaminants, nutrients, microorganisms, and/or toxins. A wash solution, in some embodiments, may be combined with a biomass to form a slurry. According to some embodiments, a wash solution (*e.g.*, a first, second, and/or third wash solution) may have a temperature below room temperature (*e.g.*, about 12° C). A wash solution, in some embodiments, may have a temperature above room temperature (*e.g.*, 50° C). In some embodiments, a wash solution may comprise any desired portion of recycled fluid. For example, a wash solution may comprise at least about 10% (v/v), at least about 20% (v/v), at least about 30% (v/v), at least about 40% (v/v), at least about 50% (v/v), at least about 60% (v/v), at least about 70% (v/v), at least about 80% (v/v), or at least about 90% (v/v) recycled from another stage of the process (*e.g.*, recycled wash solution **416**, a first reject stream from filtration (*e.g.*, FIGURE 3A, **352**), a second reject stream from filtrations (*e.g.*, FIGURE 4, **457**)).

According to some embodiments, a wash solution recycled, at least in part, from another stage of the process (*e.g.*, recycled wash solution **416**, a first reject stream from filtration (*e.g.*, FIGURE 3A, **352**), a second reject stream from filtrations (*e.g.*, FIGURE 4, **457**)) may have a temperature below room temperature (*e.g.*, about 12° C) as the result of chilling of an earlier product in the process (*e.g.*, chilling a first juice, chilling a first soluble protein, chilling a second soluble protein). A wash solution recycled, at least in part, from another stage of the process (*e.g.*, recycled wash solution **416**, a first reject stream from filtration (*e.g.*, FIGURE 3A, **352**), a second reject stream from filtrations (*e.g.*, FIGURE 4, **457**)) may have a temperature above room temperature (*e.g.*, about 50° C) as the result of a heat exchange mechanism. For example, a heat exchange mechanism may be configured to use released energy from a drying process **335** to heat a first reject stream from filtration

(*e.g.*, FIGURE 3A, **352**), a second reject stream from filtrations (*e.g.*, FIGURE 4, **457**)). Such recycling may be configured to reduce an energy input requirement of a process (*e.g.*, chilled recycled fluids cool the process they are added to; heat exchanger reduces energy requirement to heat the first reject stream).

5 In some embodiments, a second wash solution (*e.g.*, water, ozonated water, recycled wash solution FIGURE 4, **416**) may be applied to a biomass. A third wash solution (*e.g.*, water, ozonated water, recycled wash solution) may be applied to a biomass, in some embodiments. In some embodiments a first wash solution may be a reject stream from a filtration process (*e.g.*, FIGURE 3A, **352**; FIGURE 4, **457**), a second wash solution may be
10 water, and a third wash solution may be ozonated water. Some or all of a wash solution (*e.g.*, a first wash solution, second wash solution, and/or third wash solution) may be separated, in some embodiments, from a biomass (*e.g.*, using an inclined screen or vibratory screen).

Some or all of a wash solution, second wash solution, and/or third wash solution may be collected and/or reused, according to some embodiments. At least about 40%, or at least
15 about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, or at least about 95% of the wash solution and/or second wash solution separated from a biomass may be recycled for future use.

Lysing a Washed or Unwashed Biomass

As shown in FIGURE 4, a washed biomass may be lysed **420** to form a lysed biomass
20 **421**. In some embodiments, a biomass (*e.g.*, unwashed) may be lysed to form a lysed biomass. According to some embodiments, lysing may be achieved using a combination of mechanical (*e.g.*, milling), chemical (*e.g.*, pH adjustment), and/or ultrasonic (*e.g.*, sonication) methods. A lysing process may be achieved using, for example, a shear mill, a ball mill, a colloid mill, a knife mill, a hammer mill, a grinding mill, a puree machine, a filter press, or
25 any combination thereof.

In some embodiments, lysing may be performed at temperatures below room temperature. A lysing fluid (*e.g.*, water, recycled water, reverse osmosis water) may be added to a biomass or microcrop before or during lysing, according to some embodiments. As shown in FIGURE 4, a permeate **462** from reverse osmosis/nanofiltration of a second soluble
30 protein may be recycled as a lysing fluid. For example, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90% of lysing fluid may be water generated as the result of reverse osmosis/nanofiltration of a filtration product (*e.g.*, FIGURE 4, **462**).

In some embodiments a lysing fluid may be at a temperature below room temperature (*e.g.*, about 12° C). A lysing fluid may be at a temperature above room temperature (*e.g.*, 50° C). According to some embodiments, a lysing fluid recycled, at least in part, from another stage of the process (*e.g.*, FIGURE 4, **462**) may have a temperature below room temperature (*e.g.*, about 12° C) as the result of chilling of an earlier product in the process (*e.g.*, chilling a first juice, chilling a first soluble protein, chilling a second soluble protein). A lysing fluid recycled, at least in part, from another stage of the process (*e.g.*, FIGURE 4, **462**) may have a temperature above room temperature (*e.g.*, about 50° C) as the result of a heat exchange mechanism. For example, a heat exchange mechanism may be configured to use released energy from a drying process **335** to heat a permeate (*e.g.*, FIGURE 4, **462**) resulting from a reverse osmosis or nanofiltration process (*e.g.*, FIGURE 4, **460**).

Separating a Biomass

As shown in FIGURE 4, a lysed biomass may be separated **425** to generate a juice fraction **426** and a solid fraction **427**. In some embodiments, a biomass (*e.g.*, *Lemna*), a washed biomass, a lysed biomass, or any combination thereof may be separated to generate a juice fraction and a solid fraction. A juice fraction **426** may include a protein-rich liquid and/or at least some solid particles (*e.g.*, carbohydrates, fiber).

Separating **425** a lysed biomass may involve pressing (*e.g.*, belt press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating **425** a biomass (*e.g.*, harvested microcrop), washed biomass, and/or lysed biomass include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

Separating **425** may be performed at any desired temperature. Separating **425** may be performed at temperatures below room temperature (*e.g.*, 12° C), for example, to decrease proteolytic activity. In some embodiments, separating may be performed at a temperature below about 40° C, below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating **425** may be performed, for example, at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C.

Separating a Solid Fraction

As shown in FIGURE 4, a solid fraction **427** may be further separated **430** to extract additional juice, forming a second juice **432** and a first solid **431**. A second juice may include a protein-rich liquid and/or at least some solid particles (*e.g.*, carbohydrates, fiber).

5 Separating a solid fraction to form a second juice and a first solid may involve pressing (*e.g.*, belt press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating a solid fraction include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

10 As shown in FIGURE 4, according to some embodiments, one or more of a first cake (*e.g.*, FIGURE 4, **442**) and a second cake (*e.g.*, FIGURE 4, **447**) that are collected in an earlier cycle may be combined with a solid fraction from a subsequent cycle prior to separation (*e.g.*, FIGURE 4, **430**) of the solid fraction.

In some embodiments, a moisture content of a solid fraction and/or solid mixture is less than about 90%, or less than about 80%, or less than about 70%, or less than about 60%,
15 or less than about 50%, or less than about 40%, or less than about 30%, or less than about 20%, or less than about 10% by weight.

Separating a Juice Fraction

As shown in FIGURE 4, a juice fraction **426** may be separated **440** to generate a first juice **441** and a first cake **442**. A first juice may include a soluble protein. Separating a juice
20 fraction, in some embodiments, may involve centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit operations may be used to separate a juice fraction including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination
25 thereof.

Separating may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 15° C, or
30 below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or

between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

Separating a First Cake and/or a Second Juice

As shown in FIGURE 4, a first cake **442** and a second juice **432** may be combined
5 and further separated to form a third juice **446** and a second cake **447**. In some embodiments, a first cake and a second juice may be independently subjected to further separation. Separating a first cake, a second juice, or any combination thereof may involve centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit operations may be used to separate including, for example, a high-speed
10 disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

Separating may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments
15 separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or
20 between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

As shown in FIGURE 4, according to some embodiments, a third juice collected in an earlier cycle may be combined with a juice fraction from a subsequent cycle prior to further processing. In some embodiments a third juice may be combined with a first juice prior to
25 further processing (*e.g.*, FIGURE 3A). A second cake may be combined with a first solid to form a solid mixture, in some embodiments, prior to further processing (*e.g.*, FIGURE 3A).

Filtering a First Juice, a Third Juice, or any Combination Thereof

As shown in FIGURE 4, a first juice **441** may be filtered **450** to generate a first soluble protein **451** and a first reject stream **452**. In some embodiments, a first juice, a third
30 juice, or any combination thereof may be filtered to generate a first soluble protein and a first reject stream (*e.g.*, FIGURE 3A). Filtration **450** may involve microfiltration, ultrafiltration, nanofiltration, or reverse osmosis filtration.

Suitable filter sizes for microfiltration may include, in some embodiments, $\leq$ about 10 μm , or $\leq$ about 5 μm , or $\leq$ about 3 μm , or $\leq$ about 2 μm , or $\leq$ about 1 μm , or $\leq$ about 0.5 μm , or $\leq$ about 0.4 μm , or $\leq$ about 0.3 μm , or $\leq$ about 0.2 μm , or $\leq$ about 0.1 μm . As shown in FIGURE 4, a first juice, a third juice, or any combination thereof may be filtered **450** using microfiltration to generate a first soluble protein **451** in the permeate and a reject stream product **453** in the retentate first reject stream **452**.

Ultrafiltration may involve membrane filtration using pressure, concentration gradients, or a combination thereof. Suitable NMWCOs for ultrafiltration may be, in some embodiments, at most about 100 kDa, or at most about 90 kDa, or at most about 80 kDa, or at most about 70 kDa, or at most about 60 kDa, or at most about 55 kDa, or at most about 50 kDa, or at most about 45 kDa, or at most about 40 kDa, or at most about 30 kDa, or at most about 20 kDa, or at most about 15 kDa, or at most about 14 kDa, or at most about 13 kDa, or at most about 12 kDa, or at most about 11 kDa, or at most about 10 kDa, or at most about 9 kDa, or at most about 8 kDa, or at most about 7 kDa, or at most about 6 kDa, or at most about 5 kDa, or at most about 4 kDa, or at most about 3 kDa, or at most about 2 kDa, or at most about 1 kDa. In some embodiments, suitable NMWCO cut-offs for ultrafiltration may be in a range of at most about 1 kDa to at most about 10 kDa, at most about 2 kDa to at most about 10 kDa, at most about 3 kDa to at most about 10 kDa, at most about 3 kDa to at most about 15 kDa, or at most about 3 kDa to at most about 20 kDa, or at most about 3 kDa to at most about 60 kDa, or at most about 3 kDa to at most about 55 kDa, or at most about 10 kDa to at most about 55 kDa.

A first juice **441**, a third juice **446**, or any combination thereof may be filtered **450** using ultrafiltration to generate a first soluble protein **451** and a reject stream **452**, according to some embodiments. A first soluble protein may be in a retentate (*e.g.*, FIGURE 3A) or a permeate, as shown in FIGURE 4, depending upon the molecular weight cut-offs for ultrafiltration. When first reject stream **452** is a permeate of ultrafiltration (*e.g.*, FIG 3A) it may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

In some embodiments, suitable filter sizes for nanofiltration may include $\leq$ about 0.01 μm , or $\leq$ about 0.009 μm , or $\leq$ about 0.008 μm , or $\leq$ about 0.007 μm , or $\leq$ about 0.006 μm , or $\leq$ about 0.005 μm , or $\leq$ about 0.004 μm , or $\leq$ about 0.003 μm , or $\leq$ about 0.002 μm , or $\leq$ about 0.001 μm . According to some embodiments, suitable filter sizes for reverse osmosis filtration may include $\leq$ about 0.001 μm , $\leq$ about 0.0009 μm , $\leq$ about 0.0008 μm , $\leq$ about 0.0007 μm , $\leq$ about 0.0006 μm , $\leq$ about 0.0005 μm , $\leq$ about 0.0004 μm , $\leq$ about 0.0003 μm ,

≤ about 0.0002 μm, or ≤ about 0.0001 μm. In some embodiments, a first juice, a third juice, or any combination thereof may be filtered **450** using nanofiltration or reverse osmosis filtration to generate a first soluble protein (*e.g.*, FIGURE 3A, **351**) in the retentate and a permeate (*e.g.*, FIGURE 3A, **352**) of nanofiltered water or reverse osmosis water, respectively. In some embodiments a first reject stream (*e.g.*, FIGURE 3A, **352**) may be a permeate of nanofiltration or reverse osmosis filtration and may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

Buffers, protease inhibitors, anti-microbial agents, chelators (*e.g.*, EDTA), reducing agents, or any combination thereof may be added, in some embodiments, to a soluble protein **451**. A soluble protein may be chilled and/or stored at a temperature below about 30° C, or below about 25° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C, or below -2° C, or below -5° C, or below -10° C, in some embodiments. Chilling and/or storing a soluble protein at reduced temperatures may reduce degradation and/or improve protein recovery efficiency.

Filtering a First Soluble Protein

As shown in FIGURE 4, first soluble protein **451** may be filtered **455** to generate and second soluble protein **456** and a second reject stream **457**. Filtration **455** may involve microfiltration, ultrafiltration, nanofiltration, or reverse osmosis filtration.

Suitable filter sizes for microfiltration may include, in some embodiments, ≤ about 10 μm, or ≤ about 5 μm, or ≤ about 3 μm, or ≤ about 2 μm, or ≤ about 1 μm, or ≤ about 0.5 μm, or ≤ about 0.4 μm, or ≤ about 0.3 μm, or ≤ about 0.2 μm, or ≤ about 0.1 μm. In some embodiments, a first soluble protein may be filtered **455** using microfiltration to generate a second soluble protein in the permeate (not shown) and a reject stream product in the retentate.

Ultrafiltration may involve membrane filtration using pressure, concentration gradients, or a combination thereof. Suitable NMWCOs for ultrafiltration may be, in some embodiments, at most about 100 kDa, or at most about 90 kDa, or at most about 80 kDa, or at most about 70 kDa, or at most about 60 kDa, or at most about 55 kDa, or at most about 50 kDa, or at most about 45 kDa, or at most about 40 kDa, or at most about 30 kDa, or at most about 20 kDa, or at most about 15 kDa, or at most about 14 kDa, or at most about 13 kDa, or at most about 12 kDa, or at most about 11 kDa, or at most about 10 kDa, or at most about 9 kDa, or at most about 8 kDa, or at most about 7 kDa, or at most about 6 kDa, or at most about

5 kDa, or at most about 4 kDa, or at most about 3 kDa, or at most about 2 kDa, or at most about 1 kDa. In some embodiments, suitable NMWCO cut-offs for ultrafiltration may be in a range of at most about 1 kDa to at most about 10 kDa, at most about 2 kDa to at most about 10 kDa, at most about 3 kDa to at most about 10 kDa, at most about 3 kDa to at most about 15 kDa, or at most about 3 kDa to at most about 20 kDa, or at most about 3 kDa to at most about 60 kDa, or at most about 3 kDa to at most about 55 kDa, or at most about 10 kDa to at most about 55 kDa. A first soluble protein may be filtered **455** using ultrafiltration to generate a second soluble protein **455** and a reject stream **357**, according to some embodiments. A second soluble protein may be in a retentate, as shown in FIGURE 4, or a permeate (not shown) depending upon the molecular weight cut-offs for ultrafiltration. When second reject stream **457** is a permeate of ultrafiltration, as shown in FIGURE 4, it may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

In some embodiments, suitable filter sizes for nanofiltration may include $\leq$ about 0.01 μm , or $\leq$ about 0.009 μm , or $\leq$ about 0.008 μm , or $\leq$ about 0.007 μm , or $\leq$ about 0.006 μm , or $\leq$ about 0.005 μm , or $\leq$ about 0.004 μm , or $\leq$ about 0.003 μm , or $\leq$ about 0.002 μm , or $\leq$ about 0.001 μm . According to some embodiments, suitable filter sizes for reverse osmosis filtration may include $\leq$ about 0.001 μm , $\leq$ about 0.0009 μm , $\leq$ about 0.0008 μm , $\leq$ about 0.0007 μm , $\leq$ about 0.0006 μm , $\leq$ about 0.0005 μm , $\leq$ about 0.0004 μm , $\leq$ about 0.0003 μm , $\leq$ about 0.0002 μm , or $\leq$ about 0.0001 μm . As shown in FIGURE 4, in some embodiments, a first soluble protein may be filtered **455** using nanofiltration or reverse osmosis filtration to generate a second soluble protein **456** in the retentate and a permeate **457** of nanofiltered water or reverse osmosis water, respectively. In some embodiments second reject stream **457** may be a permeate of nanofiltration or reverse osmosis filtration and may be recycled to a bioreactor system for cultivation of a microcrop (*e.g.*, FIGURE 1, **105**).

In some embodiments buffers, protease inhibitors, anti-microbial agents, chelators (*e.g.*, EDTA), reducing agents, or any combination thereof may be added to the second soluble protein **456**. A second soluble protein may be chilled and/or stored at a temperature below about 30° C, or below about 25° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C, or below -2° C, or below -5° C, or below -10° C. Chilling and/or storing the second soluble protein at reduced temperatures may reduce degradation and/or improve protein recovery efficiency.

Reducing a Moisture Content of a First Soluble Protein and/or Second Soluble Protein

In some embodiments a process may be used to reduce a moisture content of a first soluble protein **451** and/or a second soluble protein **456** (*e.g.*, dewatering).

In some embodiments an evaporation process may be used to reduce a moisture content of a first soluble protein **451** or a second soluble protein **456** and generate a concentrated protein **461**. Reducing a moisture content of a first soluble protein **451** or a second soluble protein **456** may reduce capital and operational expenditures, for example, by reducing an energy needed to dry a concentrated protein **461**. Evaporation may be performed by, for example, a thermal (evaporative) means such as: a rising film evaporator, a falling film evaporator, a natural circulation evaporator (vertical or horizontal), an agitated-film evaporator, a multiple-effect evaporator, by vacuum evaporation, or any combination thereof.

As shown in FIGURE 4, a moisture content of a first soluble protein **451** or a second soluble protein **456** may be reduced, in some embodiments, by nanofiltration or reverse osmosis filtration. In some embodiments, a moisture content of a first soluble protein **451** or a second soluble protein **456** may be reduced using nanofiltration with the first soluble protein **451** or a second soluble protein **456**, respectively, in a retentate. A moisture content of a first soluble protein **451** or a second soluble protein **456**, in some embodiments, may be reduced using reverse osmosis filtration with the first soluble protein **451** or a second soluble protein **456**, respectively in a retentate. A permeate **462** of nanofiltration or reverse osmosis filtration may be recycled (*e.g.*, dilution fluid for lysis; wash solution), according to some embodiments. In some embodiments, reverse osmosis filtration and/or nanofiltration may reduce the concentration of undesirable dissolved compounds (*e.g.*, salts, calcium ions) in the resulting concentrated protein **461**.

Drying a Soluble Protein

A concentrated protein **461** may be dried **465** to generate a dry protein concentrate, according to some embodiments. A drying procedure, in some embodiments, may reduce the moisture content of a concentrated protein **461** to a desired level (*e.g.*, higher or lower moisture content, a desired moisture content). A moisture content of a dry protein concentrate, in some embodiments, may be, for example, below about 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the dry protein concentrate. In some embodiments, the protein concentration of a dry protein concentrate may be from about 30% to about 95%, or from about 40% to about 90%, or from about 50% to about 85%, or from about 60% to about 80%,

or from about 70% to about 75% by weight of the dry protein concentrate. A drying procedure may be performed using a mechanism including, for example, a spray dryer, double drum dryer, flash dryer, an evaporator, or any combination thereof.

Processing a First Solid and/or Solid Mixture

5 A first solid and/or solid mixture may be further processed to generate one or more carbohydrate-rich products. As described previously, a solid mixture may include one or more of a first solid **431**, a first cake **442**, a second cake **447**, or any combination thereof that remain after one or more separation processes **430/440/445**. Carbohydrate rich products may include a dry biocrude product suitable as a fuel feedstock or a carbohydrate-rich meal
10 suitable as a human or animal feed supplement (*e.g.*, Lemna meal).

A first solid and/or solid mixture may be processed (*e.g.*, drying, pelletization), in some embodiments, to generate at least one of a dry biocrude and a carbohydrate-rich meal. According to some embodiments, processing a first solid and/or solid mixture involves drying and/or pelletization.

15 A process for generating a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich) meal may be varied depending upon the specific characteristics desired, including, for example, moisture content, particle size, protein content, fat content, fiber content, ash content, shelf-life, pellet size, texture, or any combination thereof.

In some embodiments, a first solid and/or a solid mixture may be dried to reduce a
20 moisture content of a resulting carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal). A drying procedure, in some embodiments, may be performed in conjunction with (*e.g.*, prior to, following) other processing of a first solid and/or a solid mixture, such that the end product is a carbohydrate-rich product with a reduced moisture content. A drying procedure may be performed, in some embodiments, using a dryer
25 mechanism including, for example, a spray dryer, double drum dryer, flash dryer, evaporator, or any combination thereof.

A moisture content of a carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal) may be below about 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or
30 below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the product, in some embodiments.

Polyphenol Reduction

In some embodiments, a juice fraction **426**, a first juice **441**, a second juice **432**, a third juice **446**, a first soluble protein **451**, a second soluble protein **456**, and/or a concentrated protein **461** may undergo a processing step for the reduction of at least one polyphenol **(a)**. A polyphenol reduction process may comprise passing a juice fraction **226**, a first juice **241**, a second juice **232**, a third juice **246**, and/or a soluble protein **251** through a series (*e.g.*, at least two, at least three) of ion exchange resins. In some embodiments, a polyphenol reduction process may reduce a polyphenol (*e.g.*, a tannin) content of a juice fraction **226**, a first juice **241**, a second juice **232**, a third juice **246**, and/or a soluble protein **251** by at least 5%, or at least 10%, or at least 15%, or at least 20%, or at least 25%, or at least 30%, or at least 35%, or at least 40%, or at least 45%, or at least 50%, or at least 55%, or at least 60%, or at least 65%, or at least 70%.

Solvent Wash

In some embodiments, a first soluble protein **451**, a second soluble protein **456**, and/or a concentrated protein **461** may undergo a solvent wash **(b)**. Solvent washing **(b)** may also follow drying **465**. A solvent wash of a soluble protein product **451** and/or a solvent wash following drying **465**, may comprise at least one solvent (*e.g.*, ethanol, methanol), in some embodiments. According to some embodiments, a solvent wash of a soluble protein product **451** and/or a solvent wash following drying **455**, may result in a reduced fat content (*e.g.*, about 2% of a dry protein concentrate or less by weight) and/or a reduced chlorophyll content (*e.g.*, visually perceivable reduction in green coloration) compared to unwashed counterparts.

FIGURES 5A and 5B

FIGURES 5A and 5B are flow diagrams illustrating processes for producing a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide) and/or a protein concentrate from a biomass according to a specific example embodiments of the disclosure.

In some embodiments a process for growing, harvesting, and separating a microcrop (*e.g.*, photosynthetic aquatic species, aquatic plant species, *Lemna*, algal species) may be multiple cycles or a continuous process for the production of protein concentrate (*e.g.*, soluble protein, dry protein concentrate) and/or carbohydrate rich products such that byproducts of an earlier cycle of the process may be recycled into one or more subsequent cycles of the process. Recycling of one or more by-products may reduce the overall water

requirement for a process. In some embodiments, a process may be configured to minimize the net energy input required (*i.e.*, energy conservation).

A microcrop may be harvested (*e.g.*, FIGURE 1, **110**) forming a biomass (*e.g.*, FIGURE 1, **111**).

5 Washing a Biomass

As shown in FIGURES 5A and 5B, a biomass may be washed **515** prior to processing, according to some embodiments. A wash procedure may remove debris, contaminants, nutrients, microorganisms, and/or toxins. A wash solution, in some embodiments, may be combined with a biomass to form a slurry. According to some
10 embodiments, a wash solution (*e.g.*, a first, second, and/or third wash solution) may have a temperature below room temperature (*e.g.*, about 12° C). A wash solution, in some embodiments, may have a temperature above room temperature (*e.g.*, 50° C). In some
15 embodiments, a wash solution may comprise any desired portion of recycled fluid. For example, a wash solution may comprise at least about 10% (v/v), at least about 20% (v/v), at least about 30% (v/v), at least about 40% (v/v), at least about 50% (v/v), at least about 60%
(v/v), at least about 70% (v/v), at least about 80% (v/v), or at least about 90% (v/v) recycled from another stage of the process (*e.g.*, recycled wash solution **516**, a first reject stream from filtration (*e.g.*, FIGURE 3A, **352**), a second reject stream from filtrations (*e.g.*, FIGURE 4, **457**)).

20 According to some embodiments, a wash solution recycled, at least in part, from another stage of the process (*e.g.*, recycled wash solution **516**, a first reject stream from filtration (*e.g.*, FIGURE 3A, **352**), a second reject stream from filtrations (*e.g.*, FIGURE 4, **457**)) may have a temperature below room temperature (*e.g.*, about 12° C) as the result of chilling of an earlier product in the process (*e.g.*, chilling a first juice, chilling a first soluble
25 protein, chilling a second soluble protein). A wash solution recycled, at least in part, from another stage of the process (*e.g.*, recycled wash solution **516**, a first reject stream from filtration (*e.g.*, FIGURE 3A, **352**), a second reject stream from filtrations (*e.g.*, FIGURE 4, **457**)) may have a temperature above room temperature (*e.g.*, about 50° C).

In some embodiments, a second wash solution (*e.g.*, water, ozonated water, recycled
30 wash solution FIGURE 5A, **516**) may be applied to a biomass. A third wash solution (*e.g.*, water, ozonated water, recycled wash solution) may be applied to a biomass, in some embodiments. In some embodiments a first wash solution may be a reject stream from a filtration process (*e.g.*, FIGURE 3A, **352**; FIGURE 4, **457**), a second wash solution may be

water, and a third wash solution may be ozonated water. Some or all of a wash solution (*e.g.*, a first wash solution, second wash solution, and/or third wash solution) may be separated, in some embodiments, from a biomass (*e.g.*, using an inclined screen or vibratory screen).

Some or all of a wash solution, second wash solution, and/or third wash solution may
 5 be collected and/or reused, according to some embodiments. At least about 40%, or at least about 50%, or at least about 60%, or at least about 70%, or at least about 80%, or at least about 90%, or at least about 95% of the wash solution and/or second wash solution separated from a biomass may be recycled for future use.

Decoloring a Washed or Unwashed Biomass

10 As shown in FIGURE 5B, in some embodiments, processing a microcrop or biomass (*e.g.*, first portion, second portion) may include a decoloring procedure **570** to remove at least some of a chlorophyll component of the microcrop or biomass. A decoloring procedure **570** may be performed by exposing (*e.g.*, submerging, spraying, dripping) at least one surface of a biomass to a solvent solution (*e.g.*, ethanol, methanol, acetone). A solvent solution, in some
 15 embodiments, may be combined with a biomass (*e.g.*, first portion, second portion) to form a slurry. In some embodiments a solvent solution may include one or more alcohols (*e.g.*, ethanol, methanol), acetone, hexane, or ketones. According to some embodiments, a decoloring procedure **570** may include exposing (*e.g.*, submerging, spraying, dripping, slurry) at least one surface of a biomass to a solvent solution for at least about 5 sec., about 15 sec.,
 20 about 30 sec., about 45 sec., about 1 min., about 2 min., about 3 min., about 5 min., about 10 min., about 20 min., about 30 min., about 40 min., about 50 min., about 1 hour, about 2 hours, about 3 hours, about 4 hours, about 5 hours, about 6 hours, about 12 hours, or about 24 hours. A decoloring procedure **570**, in some embodiments, may include moving (*e.g.*, agitating, stirring, propelling) at least a portion of a solvent solution at a specified time,
 25 intermittently, or continually.

Lysing a Washed or Unwashed Biomass

As shown in FIGURES 5A and 5B, a washed biomass may be lysed **520** to form a lysed biomass **521**. In some embodiments, a biomass (*e.g.*, unwashed) may be lysed to form a lysed biomass. According to some embodiments, lysing may be achieved using a
 30 combination of mechanical (*e.g.*, milling), chemical (*e.g.*, pH adjustment), and/or ultrasonic (*e.g.*, sonication) methods. A lysing process may be achieved using, for example, a shear mill, a ball mill, a colloid mill, a knife mill, a hammer mill, a grinding mill, a puree machine, a filter press, or any combination thereof.

In some embodiments, lysing may be performed at temperatures below room temperature. A lysing fluid (*e.g.*, water, recycled water, reverse osmosis water) may be added to a biomass or microcrop before or during lysing, according to some embodiments. In some embodiments, a permeate (*e.g.*, FIGURE 4, **462**) from reverse osmosis/nanofiltration of a second soluble protein may be recycled as a lysing fluid. For example, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, at least about 70%, at least about 80%, or at least about 90% of lysing fluid may be water generated as the result of reverse osmosis/nanofiltration of a filtration product (*e.g.*, FIGURE 4, **462**). In some embodiments a lysing fluid may be at a temperature below room temperature (*e.g.*, about 12° C). A lysing fluid may be at a temperature above room temperature (*e.g.*, 50° C). According to some embodiments, a lysing fluid recycled, at least in part, from another stage of the process (*e.g.*, FIGURE 4, **462**) may have a temperature below room temperature (*e.g.*, about 12° C) as the result of chilling of an earlier product in the process (*e.g.*, chilling a first juice, chilling a first soluble protein, chilling a second soluble protein). A lysing fluid recycled, at least in part, from another stage of the process (*e.g.*, FIGURE 4, **462**) may have a temperature above room temperature (*e.g.*, about 50° C) as the result of a heat exchange mechanism. For example, a heat exchange mechanism may be configured to use released energy from a drying process to heat a permeate (*e.g.*, FIGURE 4, **462**) resulting from a reverse osmosis or nanofiltration process (*e.g.*, FIGURE 4, **460**).

Separating a Biomass

As shown in FIGURES 5A and 5B, a lysed biomass may be separated **525** to generate a juice fraction **526** and a solid fraction **527**. In some embodiments, a biomass (*e.g.*, *Lemna*), a washed biomass, a lysed biomass, or any combination thereof may be separated to generate a juice fraction and a solid fraction. A juice fraction **526** may include a protein-rich liquid and/or at least some solid particles (*e.g.*, carbohydrates, fiber). A solid fraction **527** may include at least one of an apiogalacturonan and/or an oligogalacturonide.

Separating **525** a lysed biomass may involve pressing (*e.g.*, belt press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating **525** a biomass (*e.g.*, harvested microcrop), washed biomass, and/or lysed biomass include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

Separating **525** may be performed at any desired temperature. Separating **525** may be performed at temperatures below room temperature (*e.g.*, 12° C), for example, to decrease

proteolytic activity. In some embodiments, separating may be performed at a temperature below about 40° C, below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating **525** may be performed, for example, at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C.

Separating a Solid Fraction

As shown in FIGURES 5A and 5B, a solid fraction **527** may be further separated **530** to extract additional juice, forming a second juice **532** and a first solid **531**. A second juice may include a protein-rich liquid and/or at least some solid particles (e.g., carbohydrates, fiber).

Separating a solid fraction to form a second juice and a first solid may involve pressing (e.g., belt press), centrifugation, filtration, pressurized filtration, or any combination thereof. Interchangeable unit operations for separating a solid fraction include, for example, a decanter centrifuge, a belt press, a fan press, a rotary press, a screw press, a filter press, a finisher press, or any combination thereof.

As shown in FIGURES 5A and 5B, according to some embodiments, one or more of a first cake **542** and a second cake **547** that are collected in an earlier cycle may be combined with a solid fraction from a subsequent cycle prior to separation **530** of the solid fraction.

In some embodiments, a moisture content of a solid fraction and/or solid mixture is less than about 90%, or less than about 80%, or less than about 70%, or less than about 60%, or less than about 50%, or less than about 40%, or less than about 30%, or less than about 20%, or less than about 10% by weight.

Separating a Juice Fraction

As shown in FIGURES 5A and 5B, a juice fraction **526** may be separated **540** to generate a first juice **541** and a first cake **542**. A first juice may include a soluble protein. Separating a juice fraction, in some embodiments, may involve centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit operations may be used to separate a juice fraction including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter

centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

Separating may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

Separating a First Cake and/or a Second Juice

As shown in FIGURES 5A and 5B, a first cake **542** and a second juice **532** may be combined and further separated to form a third juice **546** and a second cake **547**. In some embodiments, a first cake and a second juice may be independently subjected to further separation. Separating a first cake, a second juice, or any combination thereof may involve centrifugation, filtration, pressurized filtration, or any combination thereof. Several different interchangeable unit operations may be used to separate including, for example, a high-speed disc stack centrifuge, a circular vibratory separator, a linear/inclined motion shaker, a decanter centrifuge, a filter press, pressurized filtration mechanisms, microfiltration, vacuum filtration, or any combination thereof.

Separating may be performed, according to some embodiments, at temperatures below room temperature, for example, to decrease proteolytic activity. In some embodiments separating may be performed at a temperature below about 40° C, or below about 30° C, or below about 20° C, or below about 15° C, or below about 10° C, or below about 5° C, or below about 2° C, or below about 1° C, or below about 0° C. Separating may be performed at a temperature between about 0° C and about 10° C, or between about 5° C and about 15° C, or between about 10° C and about 20° C, or between about 15° C and about 25° C, or between about 20° C and about 30° C, or between about 25° C and about 35° C, or between about 30° C and about 40° C, in some embodiments.

As shown in FIGURES 5A and 5B, according to some embodiments, a third juice collected in an earlier cycle may be combined with a juice fraction from a subsequent cycle prior to further processing. In some embodiments a third juice may be combined with a first

juice prior to further processing (*e.g.*, FIGURE 3A). A second cake may be combined with a first solid to form a solid mixture, in some embodiments, prior to further processing (*e.g.*, FIGURE 3A).

Protein Extraction

5 In some embodiments, a first juice, a third juice, or any combination thereof may be further processed (*e.g.*, filtered) to generate a first soluble protein, a second soluble protein, a concentrated protein, and/or a dry protein concentrate according to any of the mechanisms described in Examples 1-4 above.

Processing a First Solid and/or Solid Mixture

10 As shown in FIGURES 5A and 5B, a first solid (*e.g.*, first portion, second portion) **531** may be processed to generate one or more carbohydrate-rich products and/or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide). In some embodiments, a solid mixture may be processed to generate one or more carbohydrate-rich products and/or a polysaccharide product (*e.g.*, apiogalacturonan, oligogalacturonide, polygalacturonide) (*e.g.*, FIGURE 2, **231**). Carbohydrate rich products may include a dry biocrude product suitable as a fuel feedstock or a carbohydrate-rich meal suitable as a human
15 or animal feed supplement (*e.g.*, Lemna meal). A polysaccharide product may include at least one apiogalacturonan and/or at least one oligogalacturonide.

Carbohydrate Rich Products

20 A first solid and/or solid mixture may be processed (*e.g.*, drying, pelletization), in some embodiments, to generate at least one of a dry biocrude and a carbohydrate-rich meal. According to some embodiments, processing a first solid and/or solid mixture involves drying and/or pelletization.

A process for generating a carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich) meal may be varied depending upon the specific characteristics desired, including, for example, moisture content, particle size, protein content, fat content, fiber
25 content, ash content, shelf-life, pellet size, texture, or any combination thereof.

In some embodiments, a first solid and/or a solid mixture may be dried to reduce a moisture content of a resulting carbohydrate-rich product (*e.g.*, dry biocrude and/or carbohydrate-rich meal). A drying procedure, in some embodiments, may be performed in
30 conjunction with (*e.g.*, prior to, following) other processing of a first solid and/or a solid mixture, such that the end product is a carbohydrate-rich product with a reduced moisture content. A drying procedure may be performed, in some embodiments, using a dryer

mechanism including, for example, a spray dryer, double drum dryer, flash dryer, evaporator, or any combination thereof.

A moisture content of a carbohydrate-rich product (*e.g.*, dry biocrude, carbohydrate-rich meal) may be below about 90%, or below about 80%, or below about 70%, or below about 60%, or below about 50%, or below about 40%, or below about 30%, or below about 20%, or below about 10%, or below about 5%, or below about 1% by weight of the product, in some embodiments.

Generating a Polysaccharide Product

As shown in FIGURES 5A and 5B, a first solid and/or solid mixture may be processed (*e.g.*, extracting, separating), in some embodiments, to generate at least one polysaccharide product (*e.g.*, at least one apiogalacturonan, at least one oligogalacturonide, at least one polygalacturonide).

Processing a first solid and/or solid mixture to form a polysaccharide extract and/or a polysaccharide product (*e.g.*, at least one apiogalacturonan) may involve extracting with an acid solution **580**. Extracting with an acid solution **580** may be performed by exposing (*e.g.*, submerging) a first solid and/or solid mixture to an acid solution (*e.g.*, hydrochloric acid). According to some embodiments, extracting with an acid **580** may include exposing (*e.g.*, submerging) a first solid and/or solid mixture to an acid solution (*e.g.*, hydrochloric acid) until carbohydrates are no longer detectable (*e.g.*, phenol/sulfuric acid detection method; HPAEC, GPC). Extracting with an acid **580**, in some embodiments, may include exposing (*e.g.*, submerging) a first solid and/or solid mixture to an acid solution (*e.g.*, hydrochloric acid) for a specified period of time. Acid extraction may be performed, according to some embodiments, at temperatures at or below room temperature.

As shown in FIGURES 5A and 5B, processing a first solid and/or solid mixture to form a polysaccharide product (*e.g.*, at least one apiogalacturonan) may involve extracting with an oxalic acid solution (*e.g.*, 0.5% oxalic acid) or an oxalate solution **590** (*e.g.*, ammonium oxalate). Extracting with an oxalic acid solution or an oxalate solution **590** may be performed by exposing (*e.g.*, submerging) a first solid and/or solid mixture to an oxalic acid solution or an oxalate solution (*e.g.*, 1% ammonium oxalate solution). An oxalic acid solution or an oxalate solution, in some embodiments, may be combined with a first solid and/or solid mixture to form a slurry. According to some embodiments, extracting with an oxalic acid solution or an oxalate solution **590** may include exposing (*e.g.*, submerging) a first solid and/or solid mixture to an oxalic acid solution (*e.g.*, 0.5% oxalic acid solution) or

an oxalate solution (*e.g.*, 1% ammonium oxalate solution) until carbohydrates are no longer detectable. Extracting with an oxalic acid solution or an oxalate solution, in some embodiments, may include exposing (*e.g.*, submerging) a first solid and/or solid mixture to an oxalic acid solution (*e.g.*, 0.5% oxalic acid solution) or an oxalate solution (*e.g.*, 1% ammonium oxalate) for a specified period of time. Extraction with an oxalic acid solution or an oxalate solution **590** may be performed, according to some embodiments, at temperatures at or below room temperature.

In some embodiments, at least a portion of a polysaccharide extract may be concentrated **595** (*e.g.*, evaporation). Concentration **595** of at least a portion of a polysaccharide extract may involve separating at least one apiogalacturonan or at least one oligogalacturonide or at least one polygalacturonide from the polysaccharide extract (*e.g.*, filtration) to form a polysaccharide product **598**, according to some embodiments. In some embodiments, concentration **595** of at least a portion of a polysaccharide extract may involve ultrafiltration, nanofiltration, or reverse osmosis filtration of the polysaccharide extract to form a polysaccharide product **598**. In some embodiments, concentration **595** of at least a portion of a polysaccharide extract may involve precipitating (*i.e.*, recrystallizing) with at least one solvent to form a polysaccharide product **595**, wherein the at least one solvent comprises ethanol, hexane, dichloromethane, methanol, isopropanol, ethyl acetate, water, acetone, and combinations thereof.

Suitable NMWCO for ultrafiltration, nanofiltration, or reverse osmosis filtration of a polysaccharide extract may be, in some embodiments, the same as or differ from the NMWCOs for filtration of a protein extract.

Systems of Extracting Proteins and Carbohydrate-rich Products from Aquatic Species

Embodiments of the disclosure also provide systems of extracting proteins and carbohydrate rich products from aquatic species. Such systems may include, for example: a lysing unit (*e.g.*, **220/320/420**) for lysing a biomass (*e.g.*, washed, unwashed) to generate a lysed biomass; a first separating unit (*e.g.*, **225/325/425**) for separating the lysed biomass to generate a juice fraction and a solid fraction; a second separating unit (*e.g.*, **240/340/440**) for forming a first juice and a first cake; a third separating unit (*e.g.*, **230/330/430**) for forming a first solid and a second juice; a fourth separating unit (*e.g.*, **245/345/445**) for forming a second cake and a third juice; a protein concentration unit (*e.g.*, **250/350/355/450/455**) for forming: a soluble protein and a reject stream, or for forming a first soluble protein and a first

- reject stream, or for forming a second soluble protein and second reject stream; a dewatering unit (*e.g.*, **360/460**) for forming a concentrated protein and a permeate; a protein drying unit (*e.g.*, **255/365/465**) for drying a soluble protein product to generate a dry protein concentrate; and a carbohydrate drying unit (*e.g.*, **235/335/435**) for drying a first solid or solid mixture to generate at least one carbohydrate-rich product (*e.g.*, dry bio-crude, carbohydrate-rich meal). Summarized in Table 6 are apparatuses that can be included in the units described above.

TABLE 6: Example Apparatuses	
Lysing Unit (<i>e.g.</i> , 220/320)	Shear Mill, Ball Mill, Colloid Mill, Knife Mill, Hammer Mill, Grinding Mill, Puree Machine, Filter Press
First Separating Unit (<i>e.g.</i> , 225/325)	Decanter Centrifuge, Belt Press, Fan Press, Rotary Press, Screw Press, Filter Press, Finisher Press
Second Separating Unit (<i>e.g.</i> , 240/340)	High-speed Disc Stack Centrifuge, Circular Vibratory Separator, Linear/inclined Motion Shaker, Decanter Centrifuge, Filter Press, Pressurized Filtration Mechanisms, Microfiltration Module, Vacuum Filtration Apparatus
Third Separating Unit (<i>e.g.</i> , 230/330)	Belt Press, Fan Press, Rotary Press, Screw Press, Filter Press, Finisher Press, Decanter Centrifuge
Fourth Separating Unit (<i>e.g.</i> , 245/345)	High-speed Disc Stack Centrifuge, Circular Vibratory Separator, Linear/inclined Motion Shaker, Decanter Centrifuge, Filter Press, Pressurized Filtration Mechanisms, Microfiltration, Vacuum Filtration Apparatus
Protein Concentration Unit (<i>e.g.</i> , 250/350/355)	Microfiltration Module, Ultrafiltration Module, Nanofiltration Module, Reverse Osmosis Filtration Module *Any of the above modules may be configured as single or multistage crossflow membrane filtration systems.
Dewatering Unit	Rising Film Evaporator, Falling Film Evaporator, Natural Circulation Evaporator (vertical or horizontal), Agitated-Film Evaporator, Multiple-effect Evaporator, Vacuum Evaporation Apparatus, Nano-filtration Module, Reverse Osmosis Filtration Module
Protein Drying Unit	Spray dryer, Drum dryer, Flash dryer
Carbohydrate Drying Unit	Spray dryer, Drum dryer, Flash dryer

It is understood that the listed apparatuses for each unit are for illustration purposes only, and this is not intended to limit the scope of the application. A specific combination of these or other apparatuses or units can be configured in such a system for the intended use based on the teachings in the application.

Persons skilled in the art may make various changes in the shape, size, number, separation characteristic, and/or arrangement of parts without departing from the scope of the instant disclosure. Each disclosed method and method step may be performed in association with any other disclosed method or method step and in any order according to some embodiments. Where the verb “may” appears, it is intended to convey an optional and/or permissive condition, but its use is not intended to suggest any lack of operability unless otherwise indicated. Persons skilled in the art may make various changes in methods of preparing and using a composition, device, and/or system of the disclosure. Where desired, some embodiments of the disclosure may be practiced to the exclusion of other embodiments.

Also, where ranges have been provided, the disclosed endpoints may be treated as exact and/or approximations as desired or demanded by the particular embodiment. Where the endpoints are approximate, the degree of flexibility may vary in proportion to the order of magnitude of the range. For example, on one hand, a range endpoint of about 50 in the context of a range of about 5 to about 50 may include 50.5, but not 52.5 or 55 and, on the other hand, a range endpoint of about 50 in the context of a range of about 0.5 to about 50 may include 55, but not 60 or 75. In addition, it may be desirable, in some embodiments, to mix and match range endpoints. Also, in some embodiments, each figure disclosed (*e.g.*, in one or more of the examples, tables, and/or drawings) may form the basis of a range (*e.g.*, depicted value +/- about 10%, depicted value +/- about 50%, depicted value +/- about 100%) and/or a range endpoint. With respect to the former, a value of 50 depicted in an example, table, and/or drawing may form the basis of a range of, for example, about 45 to about 55, about 25 to about 100, and/or about 0 to about 100.

These equivalents and alternatives along with obvious changes and modifications are intended to be included within the scope of the present disclosure. Accordingly, the foregoing disclosure is intended to be illustrative, but not limiting, of the scope of the disclosure as illustrated by the appended claims.

The title, abstract, background, and headings are provided in compliance with regulations and/or for the convenience of the reader. They include no admissions as to the scope and content of prior art and no limitations applicable to all disclosed embodiments.

EXAMPLES

Some specific example embodiments of the disclosure may be illustrated by one or more of the examples provided herein.

5

EXAMPLE 1: Apiogalacturonan

Apiogalacturonan is an apiogalacturonic pectic polysaccharide which consists in one backbone of linear chain of pectin (α 1-4 linked D-Galacturonic acids: D-GalpA- α 1-4 D-GalpA) with ramifications:

- 10
- Chains of β 1-5 linked apiose: Api β 1-5Api β
 - Chains of α 1-5 linked Arabinose: Ara α 1-5Ara α
 - Chains of Galactose alternatively linked in β 1-3 and β 1-4: Galp β 1-3Galp β 1-4Galp β
 - Chains of β 1-4 linked xylose with O-Methyl-xylose: Xylp β 1-4Xyl β -4Xylp β 2-OMeXylp

15 Apiogalacturonan consists of the following monosaccharides: apiose, galacturonic acid, arabinose, galactose, xylose and O-Methyl-xylose.

EXAMPLE 2: Apiogalacturonan Extracted from *Lemna*

Monosaccharide analysis was performed on *Lemna* samples prepared in accordance with certain embodiments of the disclosure. A biomass was cultivated by growing *Lemna* in a bioreactor system with a growth medium and a propulsion system. A portion of the biomass was harvested and subjected to a first wash of water followed by removal of the first wash water using a shaking mechanism (*e.g.*, slotted or mesh shaker). The biomass was then subjected to a second wash using ozonated water and the second wash solution was removed using a shaking mechanism (*e.g.*, slotted or mesh shaker) to form a washed biomass. A first portion of the washed biomass was used as a sample for polysaccharide analysis described below ("Raw Biomass"). A second portion of the washed biomass was diluted with reverse osmosis and lysed in a shear mill at room temperature. The lysed biomass was separated to form a juice fraction and a solid fraction using a decanter centrifuge at room temperature.

30 The solid fraction was further separated using a screw press to form a first solid and a second juice. The juice fraction was separated by disc stack centrifuge to form a first cake and a first juice. The first cake and the second juice were combined and further separated using a vibratory separator to form a third juice and a second cake. The third juice was

combined with the first juice. The second cake was combined with the first solid to form a solid mixture.

The solid mixture was dried to 10% moisture content forming a carbohydrate rich product. A sample of the carbohydrate rich product ("Lemna Meal" or "LM") was submitted for polysaccharide testing described below.

The first juice/third juice mixture was filtered using ultrafiltration to form a first soluble protein. The first soluble protein was then subjected to reverse osmosis filtration to form a concentrated protein. A sample of the concentrated protein ("Lemna Protein Concentrate" or "LPC") was submitted for polysaccharide testing described below.

Quantification of the monosaccharide composition of the Raw Biomass, LPC, and LM was performed by high pressure anion exchange chromatography (HPAEC). HPAEC was performed using a Dionex CarboPac PA1 column with amperometric detection of polysaccharide hydrolysis where hydrolysis was performed under the following conditions: (1) hydrolysis with 2N Trifluoroacetic acid (TFA) at 100° C for 0.5 hours; (2) hydrolysis with 2N TFA at 100° C for 4 hours; (3) hydrolysis with 2N H₂SO₄ at 100° C for 6 hours; (4) hydrolysis with 2N H₂SO₄ at 100° C for 6 hours after overnight exposure to 26N H₂SO₄ at room temperature. The results of one experiment are shown in Table 7, with each value representing the higher monosaccharide content of the four hydrolysis methods.

Table 7: Monosaccharide Composition of Lemna Products using HPAEC

Sample	Monosaccharide Quantification (% w/w)										
	Fuc	Ara	Rha	Gal	Glc	Man	Xyl	Gal Ac	Glu Ac	Glc Ac	Man Ac
Raw Biomass	0.2	0.9	0.4	1.6	7.5	0.4	2.1	4.6	N.D.	N.D.	N.D.
LPC	0.1	1.1	N.D.	1.8	1.2	N.D.	0.6	N.D.	N.D.	N.D.	N.D.
LM	0.2	1.1	0.7	1.4	13.4	N.D.	4.0	10.8	N.D.	N.D.	N.D.

*N.D. None detected.

The HPAEC analysis revealed the presence of two main monosaccharides, glucose and galacturonic acid, thus suggesting the presence of two different polysaccharides (one with glucose and one with galacturonic). Apiose could not be quantified using the HPAEC method described above; therefore, it could not be confirmed that the polysaccharide with galacturonic acid was apiogalacturonan.

The failure to detect galacturonic acid in the LPC indicates that the vast majority of the galacturonic acid polysaccharide is retained in the solid fraction.

Because apiose could not be quantified using the HPAEC method described, to confirm that the galacturonic acid polysaccharide was apiogalacturonan, gas phase chromatography (GPC) was used. Samples of LM were prepared by (1) hydrolyzing the polysaccharide product to form monosaccharides by methanolysis; (2) trimethylsilylation of the monosaccharides to form a volatilized monosaccharides derivative; and (3) quantifying and identifying the volatilized monosaccharides as O-methylglycosides by gas phase chromatography.

Table 8: Monosaccharide Quantification of Lemna LM using GPC

Sample	Monosaccharide Quantification (% w/w)					
	Rha	Gal	Glc	Xyl	Gal Ac	Apiose
LM	2.0	2.3	3.8	7.9	25.2	8.7

Table 9: Monosaccharide Molar Ratio of Lemna LM using GPC

Sample	Monosachharides Molar Ratio					
	Rha	Gal	Glc	Xyl	Gal Ac	Apiose
LM	0.15	0.13	0.24	0.64	1.00	0.43

Tables 8 and 9 show the monosaccharide as either a quantification or a molar ratio of the LM as analyzed using GPC. The GPC analysis of LM confirmed the presence of both apiose and galacturonic acid and thereby the composition of the polysaccharide as apiogalacturonan. The presence of xylose and arabinose also supports this conclusion as these are side chains of the apiogalacturonan molecule. However, it is also possible that the sample contained a residual presence of pectin as suggested by the detection of low quantities of rhamnose, the monosaccharide marker of pectin.

The content of apiogalacturonan in the LM sample tested was calculated as 33.9% (w/w) (This quantification does not take into account the contribution of galacturonic acid coming from a possible presence of pectin in the product).

EXAMPLE 3: Composition of a Polysaccharide Product Extracted from Lemna

Compositional analysis was performed on a polysaccharide product sample extracted from *Lemna*, according to certain embodiments of the disclosure. A biomass was cultivated

by growing *Lemna* in a bioreactor system with a growth medium and a propulsion system. A portion of the biomass was harvested and subjected to a first wash of water followed by removal of the first wash water using a shaking mechanism (*e.g.*, slotted or mesh shaker). The biomass was then subjected to a second wash using ozonated water and the second wash solution was removed using a shaking mechanism (*e.g.*, slotted or mesh shaker) to form a washed biomass. A second portion of the washed biomass was diluted with reverse osmosis water and lysed in a shear mill at room temperature forming a lysed biomass. The lysed biomass was separated to form a juice fraction and a solid fraction using a decanter centrifuge at room temperature.

The solid fraction was further separated using a screw press to form a first solid and a second juice. The juice fraction was separated by disc stack centrifuge to form a first cake and a first juice. The first cake and the second juice were combined and further separated using a vibratory separator to form a third juice and a second cake. The third juice was combined with the first juice. The second cake was combined with the first solid to form a solid mixture.

The solid mixture was dried to about 10% moisture content forming a carbohydrate rich product, which was then exposed to 0.5% oxalic acid and titrated to pH 4.0. At least a portion of the oxalic acid extract was precipitated out of solution by the addition of ethanol and separated (*e.g.*, centrifugation), to form a polysaccharide product, which was analyzed via GPC.

Table 10 illustrates the composition of the polysaccharide product sample based on a gas chromatographic analysis.

Table 10: Composition of Polysaccharide Product Following Oxalic Acid Extraction

Sample	Compositional Analysis (% w/w)				
	Protein	Ash	Fat	Pectin	Total
Polysaccharide Product	2.77	7.38	0.55	89.36	100.00

CLAIMS

What is claimed is:

1. A method of treating a biomass comprising a microcrop, the method comprising:
 - lysing the biomass to form a lysed biomass;
 - separating the lysed biomass to generate a juice fraction and a solid fraction;
 - separating the solid fraction to generate a first solid and a second juice; and
 - extracting the first solid with an extraction solution to form a polysaccharide extract,wherein the extraction solution comprises an acid solution, an oxalic acid solution, an oxalate solution, or combinations thereof, and
wherein the polysaccharide extract comprises an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof.
2. The method of claim 1 further comprising concentrating the polysaccharide extract to form a polysaccharide product,
wherein the polysaccharide product comprises the apiogalacturonan, the oligogalacturonide, the polygalacturonide, or the combination thereof.
3. The method of claim 2, wherein concentrating the polysaccharide extract consists of a process selected from evaporation, precipitation, dialysis, filtration, ultrafiltration, nanofiltration, reverse osmosis filtration, centrifugation, dewatering, and combinations thereof.
4. The method of claim 1, wherein the polysaccharide extract comprises at least 30% (w/w) of the apiogalacturonan, the oligogalacturonide, the polygalacturonide, or the combination thereof.
5. The method of claim 2, wherein the polysaccharide product comprises at least 80% (w/w) of the apiogalacturonan, the oligogalacturonide, the polygalacturonide, or the combination thereof.
6. The method of claim 1 further comprising decoloring the biomass.

7. The method of claim 6 further comprising concentrating the polysaccharide extract to form a polysaccharide product,

wherein the polysaccharide product comprises the apiogalacturonan, the oligogalacturonide, the polygalacturonide, or the combination thereof.

8. The method of claim 7, wherein concentrating the polysaccharide extract consists of a process selected from evaporation, precipitation, dialysis, filtration, ultrafiltration, nanofiltration, reverse osmosis filtration, centrifugation, dewatering, and combinations thereof.

9. The method of claim 7, wherein the polysaccharide product comprises at least 80% (w/w) of the apiogalacturonan, the oligogalacturonide, the polygalacturonide, or the combination thereof.

10. The method of claim 1, further comprising washing the biomass with at least one of a first wash solution, a second wash solution, a third wash solution, or a combination thereof.

11. The method of claim 1 further comprising separating the juice fraction to generate a first juice and a first cake, and filtering the first juice to generate a soluble microcrop protein and a reject stream.

12. The method of claim 11 further comprising:

- (1) separating the first cake,
- (2) separating the second juice, or
- (3) separating the first cake and separating the second juice, in each case, to generate a third juice and a second cake;

combining the second cake with the first solid prior to extracting the first solid with the extraction solution to form the polysaccharide extract.

14. The method of claim 11 further comprising subjecting the first juice to a polyphenol reduction process to generate a soluble microcrop protein having a reduced concentration of at least one polyphenol.

15. The method of claim 1, wherein the microcrop comprises *Lemna*.

16. A polysaccharide product from a biomass comprising a microcrop, the polysaccharide product produced by a process comprising:

lysing the biomass to form a lysed biomass;

separating the lysed biomass to generate a juice fraction and a solid fraction;

separating the solid fraction to generate a first solid and a second juice;

extracting the first solid with an extraction solution to form a polysaccharide extract,

wherein the extraction solution comprises an acid solution, an oxalic acid solution, an oxalate solution, or combinations thereof, and

concentrating the polysaccharide extract to form a polysaccharide product,

wherein the polysaccharide product comprises an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or a combination thereof.

17. The polysaccharide product of claim 16, wherein concentrating the polysaccharide extract consists of a process selected from evaporation, precipitation, dialysis, filtration, ultrafiltration, nanofiltration, reverse osmosis filtration, centrifugation, dewatering, and combinations thereof.

18. The polysaccharide product of claim 16, wherein the polysaccharide product comprises at least 80% (w/w) of the apiogalacturonan, the oligogalacturonide, the polygalacturonide, or the combination thereof.

19. The polysaccharide product of claim 16, wherein the process further comprises decoloring the biomass.

20. The polysaccharide product of claim 16, wherein concentrating the polysaccharide extract consists of a process selected from evaporation, precipitation, dialysis,

filtration, ultrafiltration, nanofiltration, reverse osmosis filtration, centrifugation, dewatering, and combinations thereof.

21. The polysaccharide product of claim 19, wherein the polysaccharide product comprises at least 80% (w/w) of the apiogalacturonan, the oligogalacturonide, the polygalacturonide, or the combination thereof.

22. The polysaccharide product of claim 16, wherein the process further comprises washing the biomass with at least one of a first wash solution, a second wash solution, a third wash solution, or a combination thereof.

23. The method of claim 16, wherein the microcrop comprises *Lemna*.

24. The method of claim 19, wherein the microcrop comprises *Lemna*.

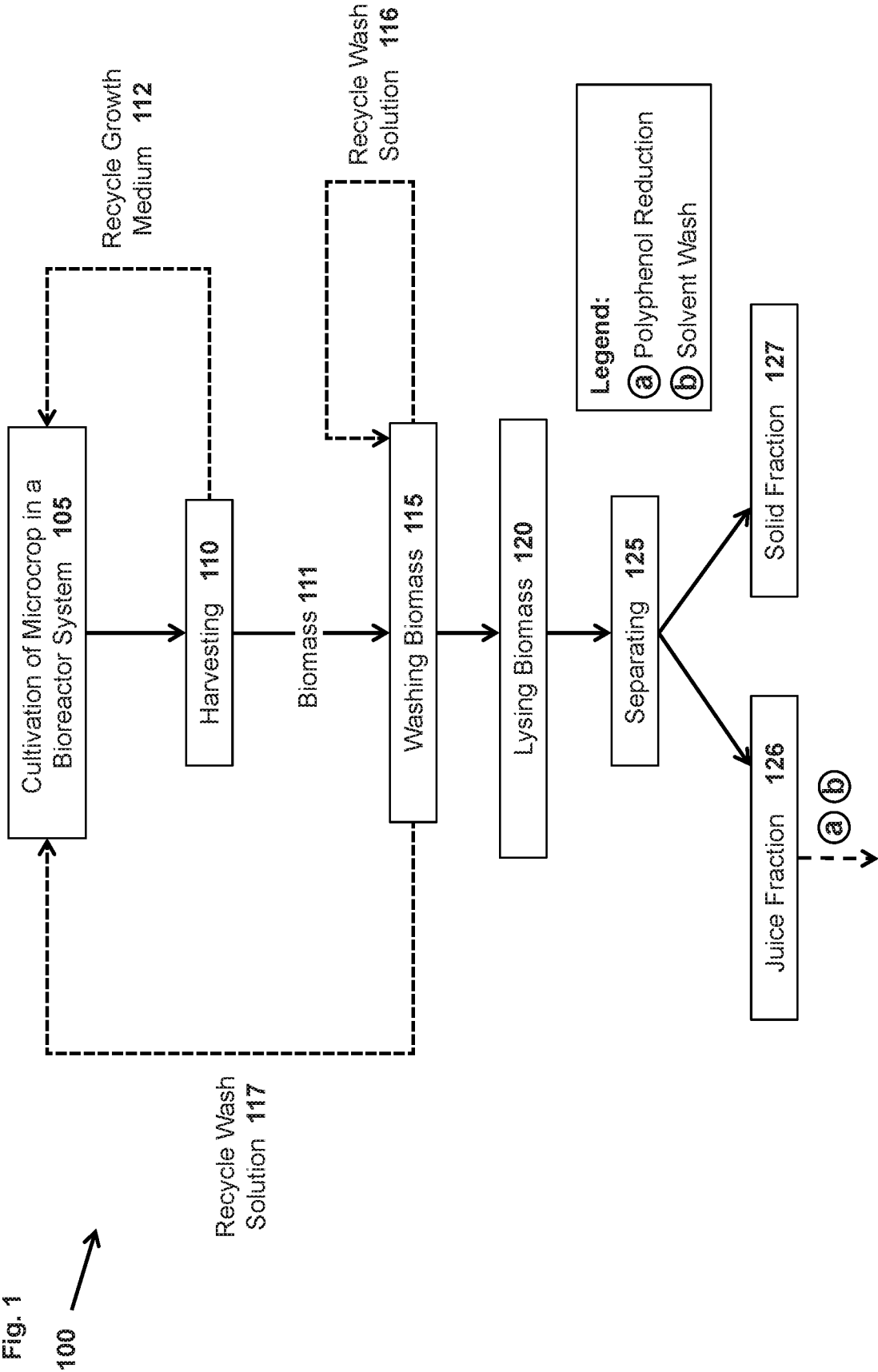
25. The method of claim 16 further comprising decoloring the biomass.

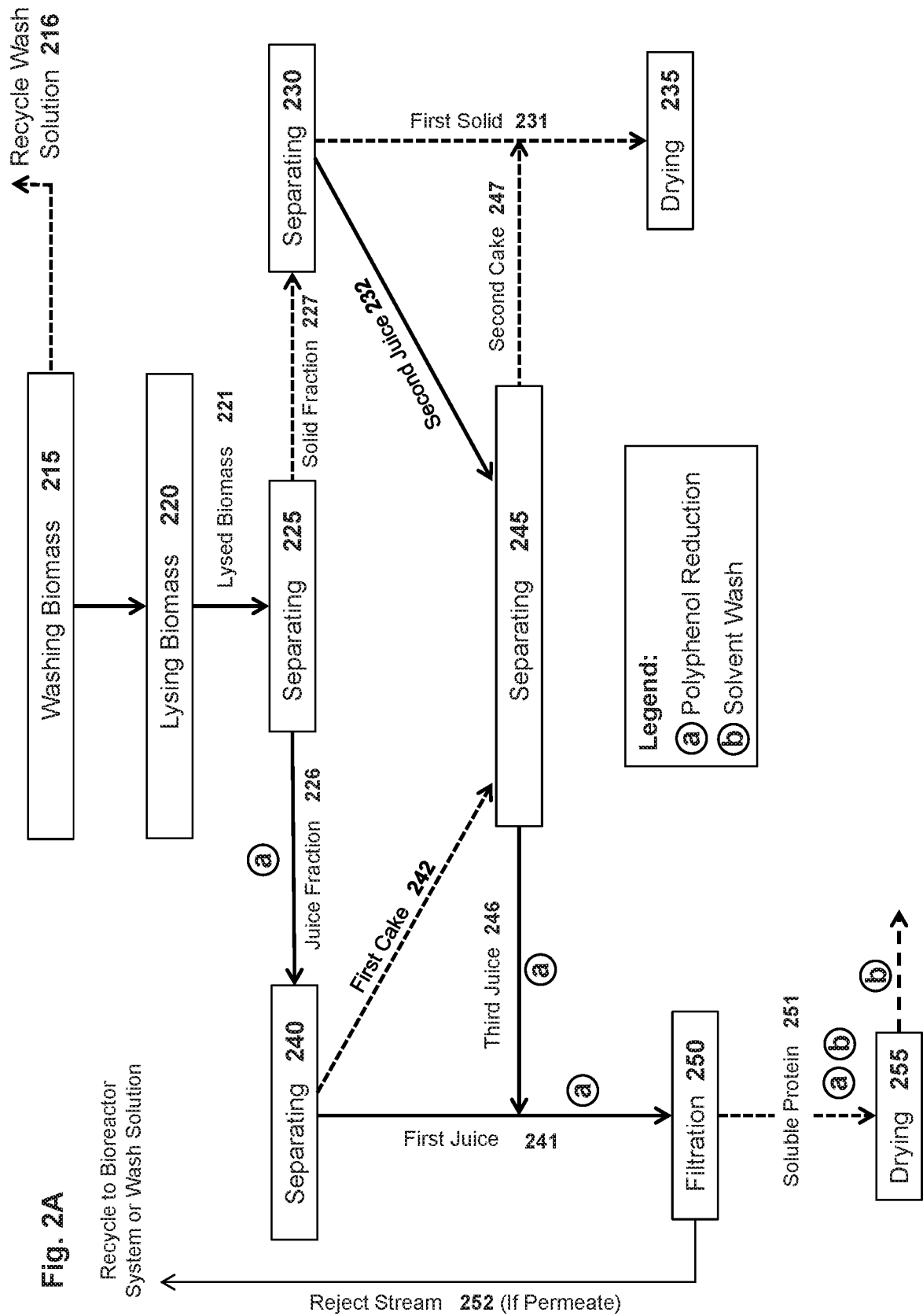
26. A polysaccharide product from a biomass comprising a microcrop, wherein the polysaccharide product comprises at least 80% (w/w) of an apiogalacturonan, an oligogalacturonide, a polygalacturonide, or the combination thereof.

27. The polysaccharide product of claim 26, wherein the polysaccharide product further comprises a protein content of about 5 wt. % or less.

28. The polysaccharide product of claim 26, wherein the polysaccharide product further comprises a fat content of lower than about 1 wt. %.

29. The polysaccharide product of claim 26, wherein the polysaccharide product further comprises an ash content of lower than about 10 wt. %.





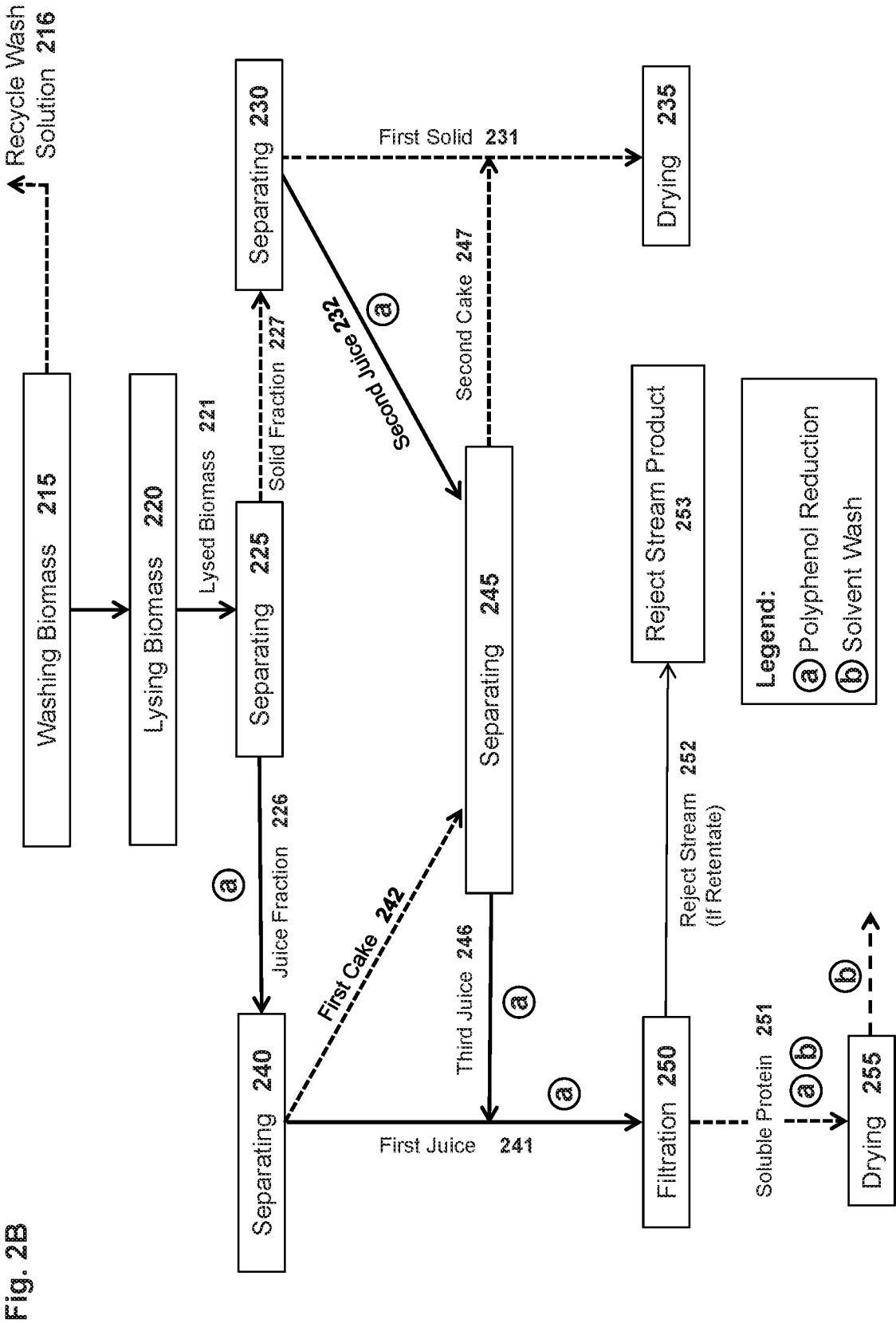
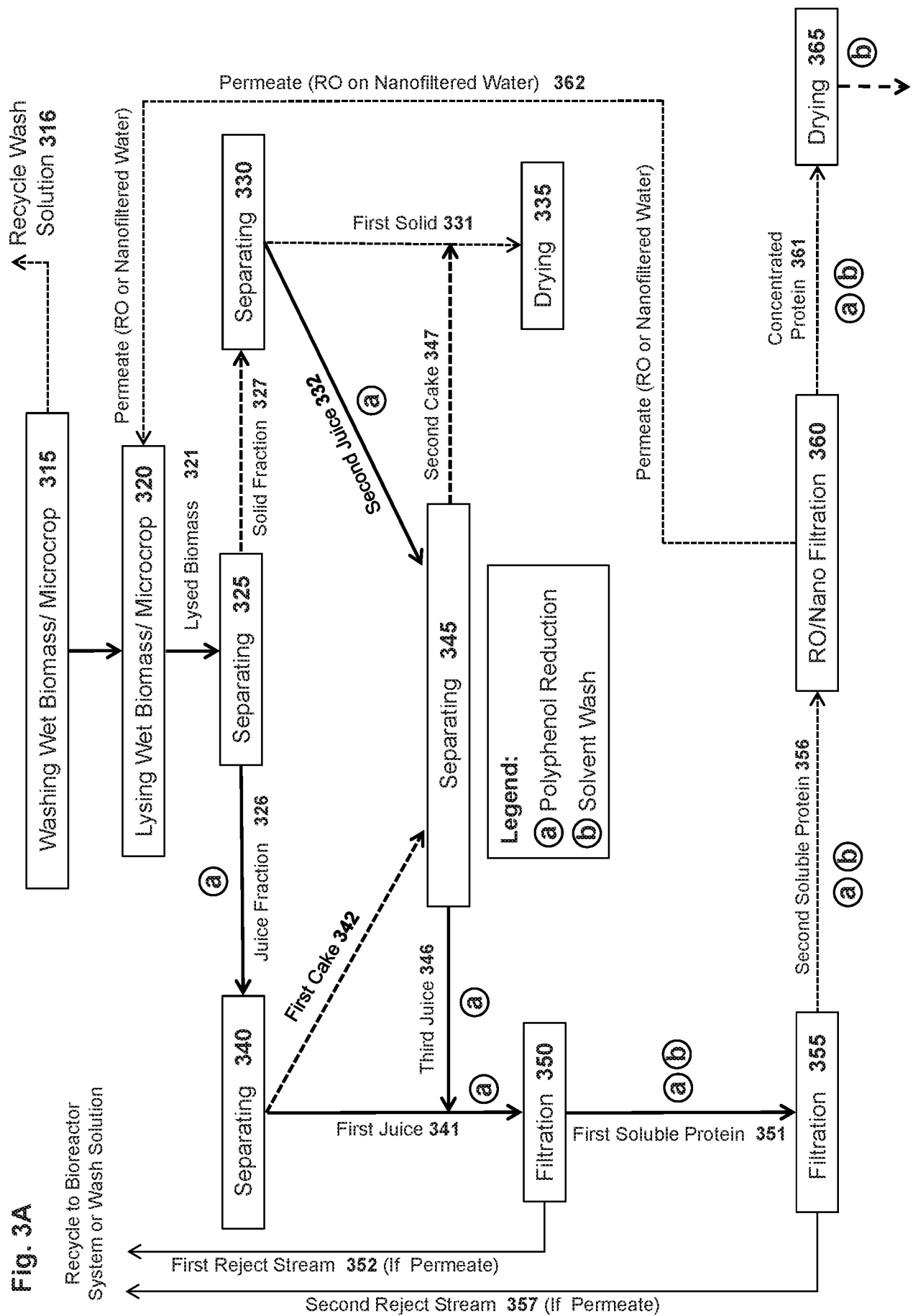
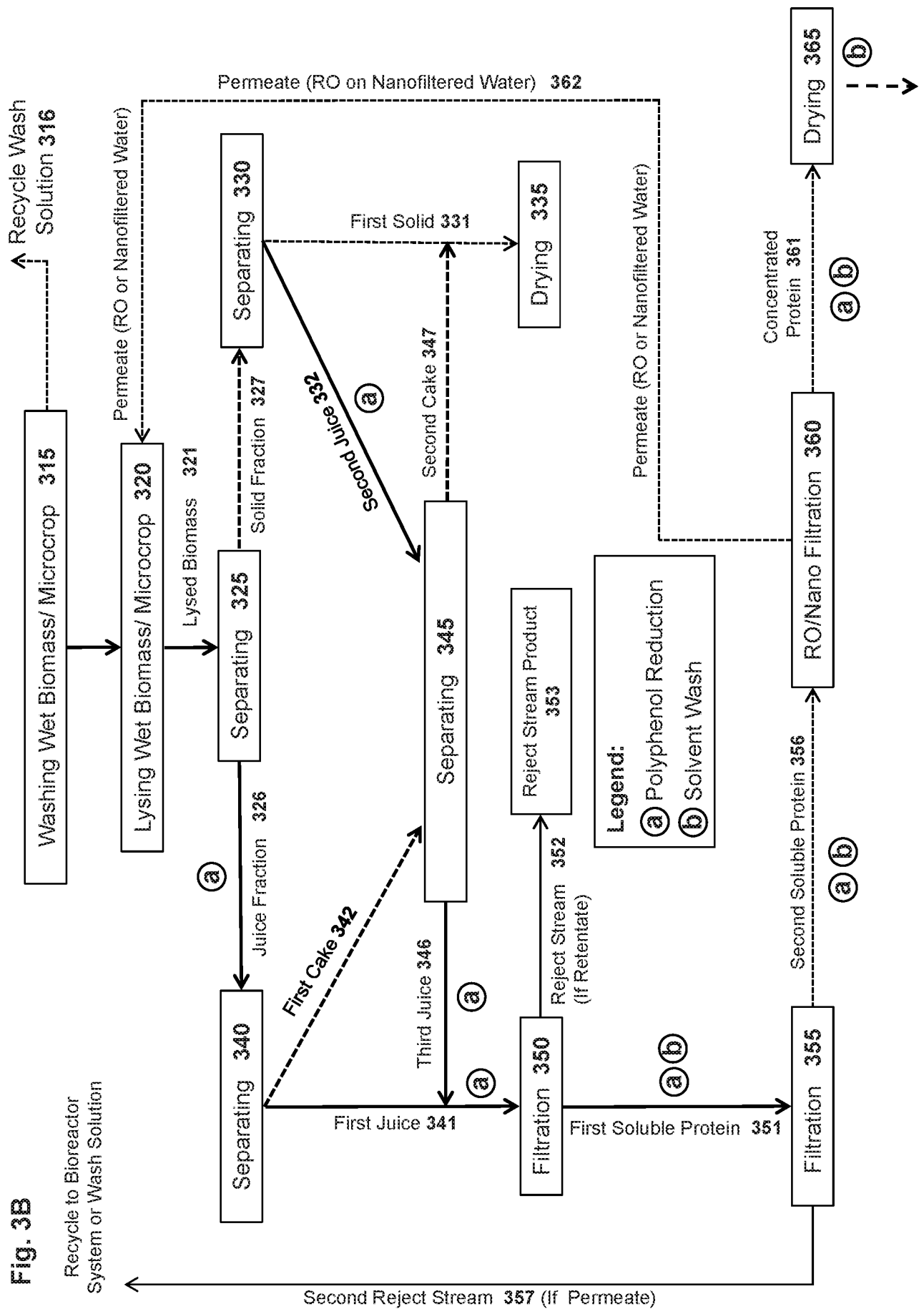
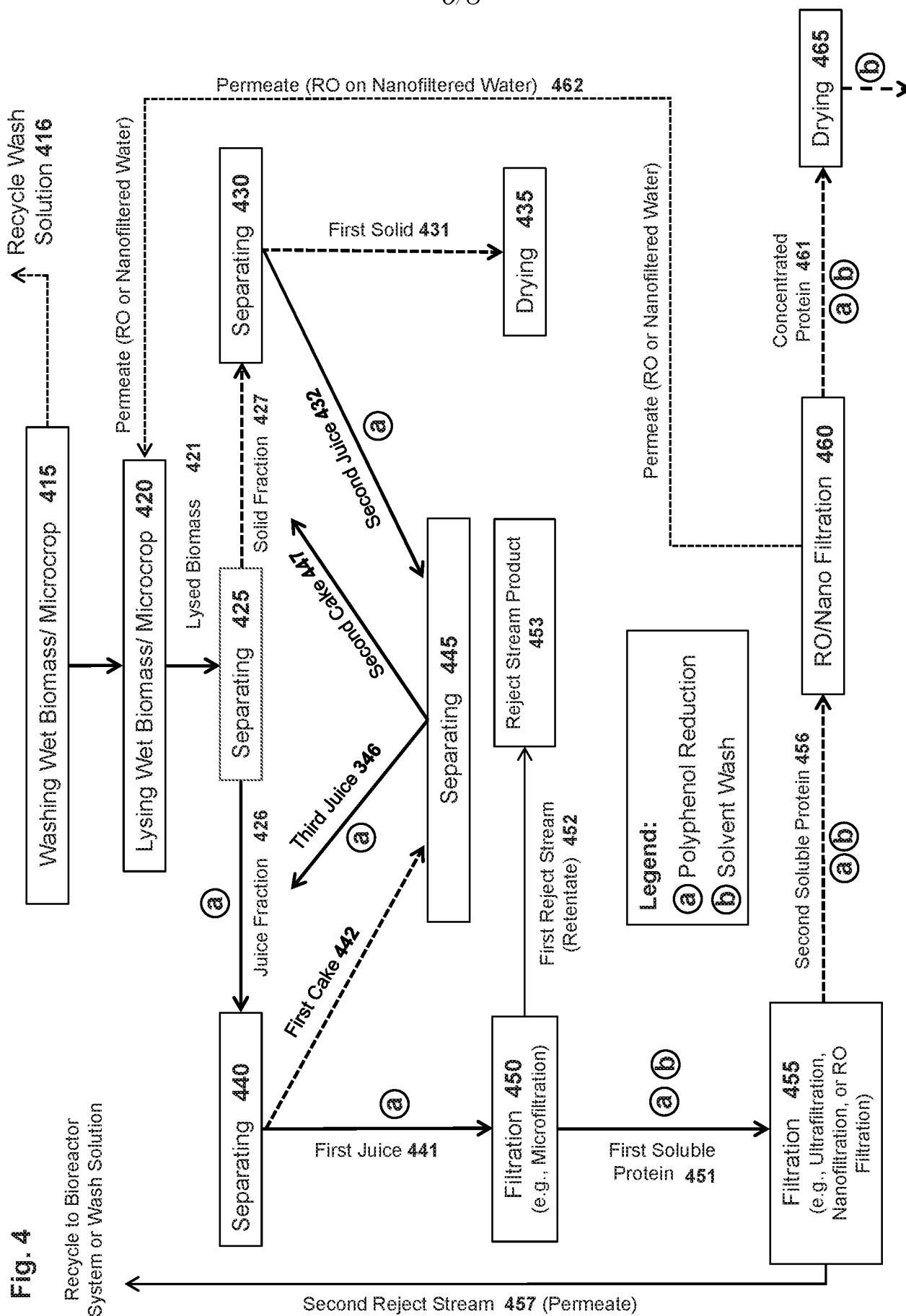


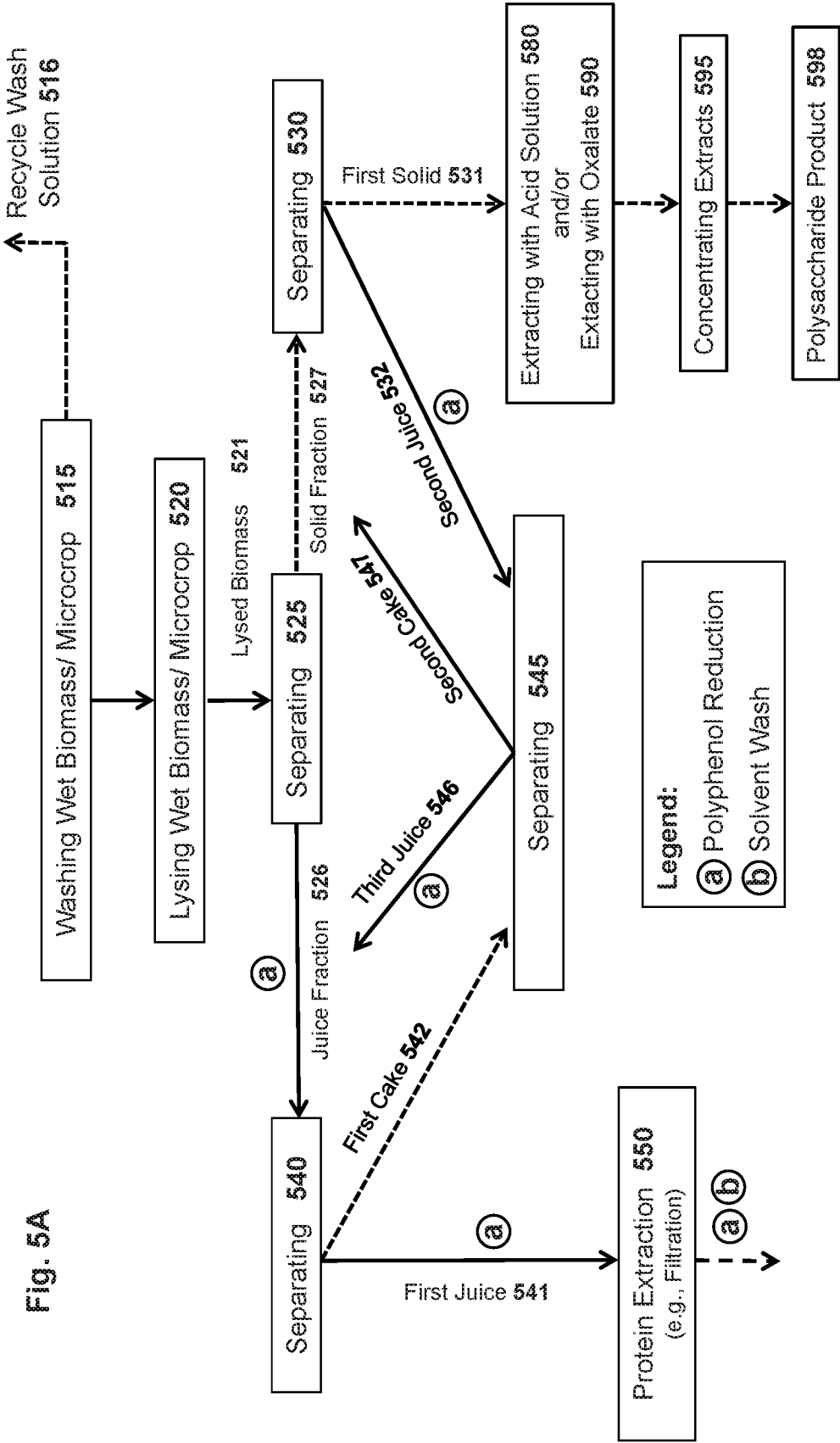
Fig. 3A

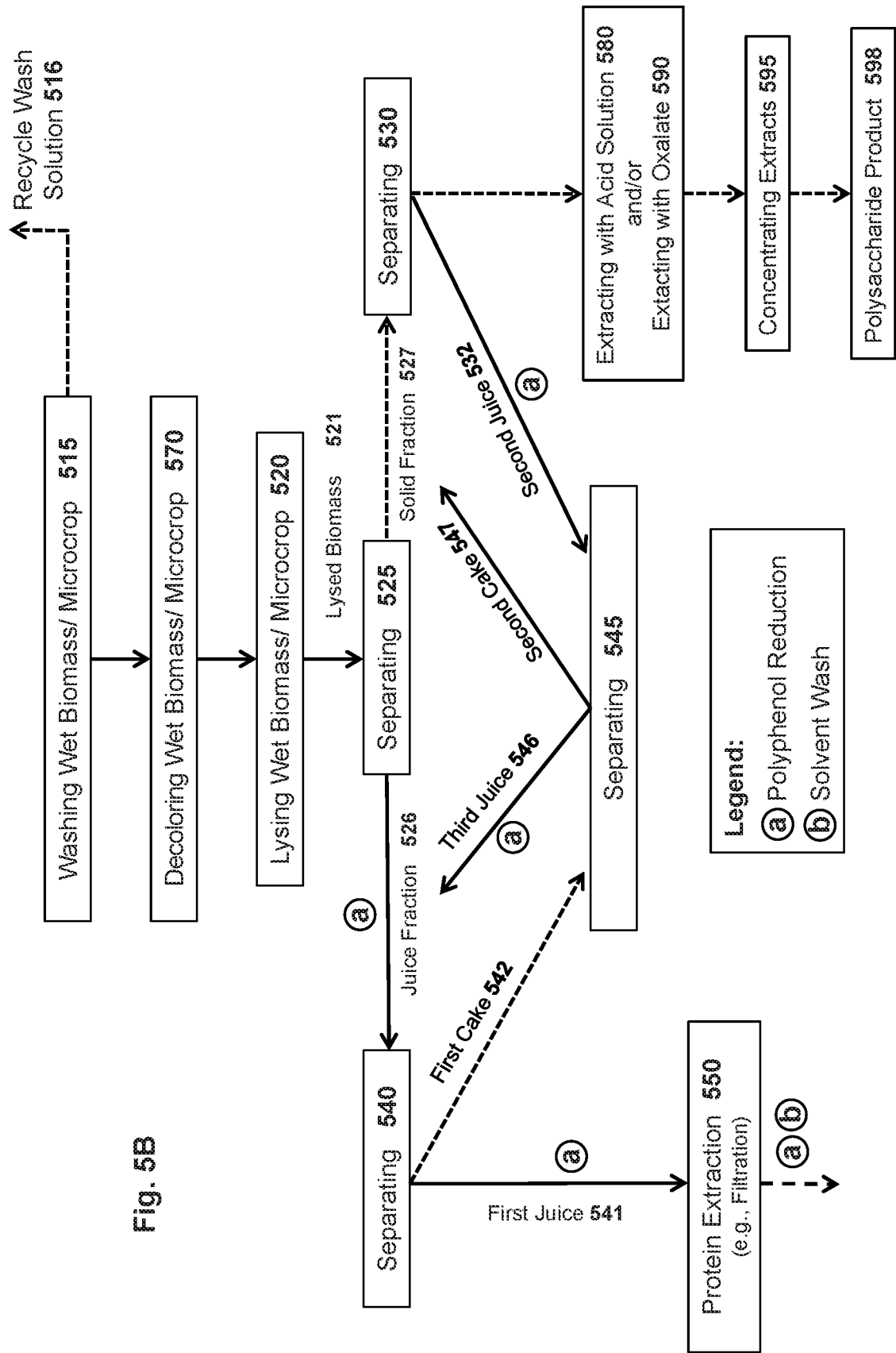
Recycle to Bioreactor System or Wash Solution











INTERNATIONAL SEARCH REPORT

International application No.
PCT/US2016/041156**A. CLASSIFICATION OF SUBJECT MATTER****C08B 37/00(2006.01)i, B01D 61/16(2006.01)i, B01D 61/02(2006.01)i, B01D 61/04(2006.01)i**

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C08B 37/00; C12P 19/04; C07H 3/00; C07H 1/00; C08B 37/06; A61K 31/737; A01N 43/16; B01D 61/16; B01D 61/02; B01D 61/04

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Korean utility models and applications for utility models

Japanese utility models and applications for utility models

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

eKOMPASS(KIPO internal) & Keywords: biomass, microcrop, polysaccharide, apiogalacturonan, oligogalacturonide, polygalacturonide, Lemna**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2011-0172102 A1 (JACOB, BRUNO et al.) 14 July 2011 See claims 1-2; paragraphs [0052]-[0057].	1-12, 14-29
A	US 2014-0023675 A1 (LINA, BRUNO et al.) 23 January 2014 See claim 42.	1-12, 14-29
A	CN 101595943 B (SHUDE HONGLONG BIOLOGY AND TECHNOLOGY CO., LTD.) 13 March 2013 See the whole document.	1-12, 14-29
A	US 8722878 B2 (RAINES, RONALD T. et al.) 13 May 2014 See claim 1.	1-12, 14-29
A	US 2008-0032349 A1 (VISKOV, CHRISTIAN et al.) 07 February 2008 See claim 1.	1-12, 14-29



Further documents are listed in the continuation of Box C.



See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

17 October 2016 (17.10.2016)

Date of mailing of the international search report

18 October 2016 (18.10.2016)

Name and mailing address of the ISA/KR

International Application Division

Korean Intellectual Property Office

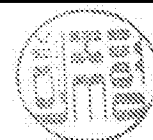
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INTERNATIONAL SEARCH REPORTInternational application No.
PCT/US2016/041156**Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)**

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☒ Claims Nos.: 13
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
Claim 13 is unclear, because claim 13 is missing in this application (PCT Article 6).
3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fees, this Authority did not invite payment of any additional fees.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/US2016/041156

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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