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- (71) **Applicant:** BIONOVION HOLDING B.V. [NL/NL];  
Molenweg 79, RE 1129, NL-5349 AC Oss (NL).
- (72) **Inventors:** VAN EENENNAAM, Hans; Bandoengstraat  
12, NL-6524 KZ Nijmegen (NL). VAN ELSAS, Andrea;  
Ruusbroecgaarde 139, NL-5343 JL Oss (NL).
- (74) **Agent:** TAUCHNER, Paul; Vossius & Partner,  
Siebertstrasse 4, D-81675 München (DE).
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(54) **Title:** COMBINING CD27 AGONISTS AND IMMUNE CHECKPOINT INHIBITION FOR IMMUNE STIMULATION

(57) **Abstract:** The present invention relates to treatments of conditions ameliorated by stimulation of an immune response, in particular by the stimulation of antigen-specific T-lymphocytes. Treatment of such conditions according to the invention is effected by the combination of an anti-human CD27 agonistic antibody together with a number of immune checkpoint inhibitors.



## FIELD OF THE INVENTION

20 BACKGROUND

CD27 is exclusively expressed by hematopoietic cells, in particular those of the lymphocyte lineage, i.e. T-, B- and NK cells. CD27 was originally defined as a human T-cell co-stimulatory molecule that increments the proliferative response to TCR stimulation (van Lier et al., 1987, *J Immunol* 139:1589-96). Presence of CD70, the ligand of CD27, dictates the timing and persistence of CD27-mediated co-stimulation.

Transgenic expression of CD70 in immature dendritic cells sufficed to convert immunological tolerance to virus or tumors into CD8<sup>+</sup> T cell responsiveness. Likewise, agonistic soluble CD70 promoted the CD8<sup>+</sup> T cell response upon  
5 such peptide immunization (Rowley et al., 2004, *J Immunol* 172:6039-6046) and in CD70 transgenic mice, CD4<sup>+</sup> and CD8<sup>+</sup> effector cell formation in response to TCR stimulation was greatly facilitated (Arens et al. 2001, *Immunity* 15:801-12; Tesselaar et al., 2003, *Nat Immunol* 4:49-54; Keller et al.  
10 2008, *Immunity* 29: 334-346). In mouse lymphoma models, tumor rejection was improved upon CD70 transgenesis or injection of an anti-mouse CD27 antibody (Arens et al., 2003, *J Exp Med* 199:1595-1605; French et al., 2007, *Blood* 109: 4810-15; Sakanishi and Yagita, 2010, *Biochem. Biophys. Res. Comm.*  
15 393: 829-835; WO 2008/051424; WO 2012/004367).

In WO2012/004367 the first anti-human agonistic antibody (designated hCD27.15) was described that does not require crosslinking to activate CD27-mediated co-stimulation of the immune response. In addition, an anti-  
20 human CD27 antibody, designated 1F5 was disclosed that activates CD27 upon crosslinking (WO2011/130434 and Vitale et al., *Clin. Cancer Res*, 2012, 18(14): 3812-3821).

Recently, the first clinical successes of agents that modulate cancer immunity have validated cancer  
25 immunotherapy as a novel path to obtain durable and long-lasting clinical responses in cancer patients (Mellman et al., *Nature*, 2011, 480:480-489). The first such agent, ipilimumab (Yervoy, BMS), that obtained marketing approval for treatment of metastatic melanoma is an antibody blocking  
30 the CTLA4 receptor, an immune checkpoint protein. Further immune checkpoint inhibitors under development are antibodies that block the interaction between the PD-1 receptor and its ligands PD-L1 and PD-L2 (Mullard, *Nat. Rev.*

Drug Disc., 2013, 12:489-492). Several antibodies targeting the PD-1 pathway are currently in clinical development for treatment of melanoma, renal cell cancer, non-small cell lung cancer, diffuse large B cell lymphoma and other tumors. Although these agents have not yet been filed for marketing approval, impressive results have been obtained in early clinical studies, for example with Lambrolizumab (anti-PD1) in melanoma (Hamid et al., 2013, *New. Eng. J. Med.*, 369:134-44).

The current state of the art does not suggest that agonists of the CD27 receptor would be rationally combined with immune checkpoint inhibitors, such as anti-PD1, anti-PDL1 or anti-CTLA4 antibodies to improve cancer immunity. In fact, available data suggest that CD27 acts, at least in part via overruling PD-1 and CTLA-4 signals or via downregulation of these immune checkpoints. First, in a T-cell tolerance model based on LCMV-derived epitopes that was demonstrated to be highly dependent on PD-1 and CTLA4 receptors the forced expression of CD70 ligand was sufficient to turn T cell tolerance into activation of T-cell immunity. Apparently these data strongly suggest that CD27 activation overrules tolerance mediated via PD-1 and CTLA4 (Keller et al., *Immunity*, 2008, 29:934-946. In a second model, activation of CD27 using a rat anti-mouse CD27 agonistic antibody was demonstrated to support the maintenance of CD8<sup>+</sup> T-cells, to reduce the frequency of FoxP3-expressing CD4<sup>+</sup> T-cells within tumors and to potentiate the ability of NK1.1<sup>+</sup> and CD8<sup>+</sup> tumor infiltrating cells to secrete IFN- $\gamma$  in coculture with tumor cells. This enhanced function correlated with decreased levels of PD-1 expression on CD8<sup>+</sup> T-cells (Roberts et al., *J Immunother.*, 2010, 33: 769-79).

SUMMARY

The current state of the art thus is, that stimulation of CD27 overrules PD-1 and CTLA-4 mediated immune tolerance and that activation using anti-CD27 antibodies results in down-regulation of these immune checkpoint receptors. Therefore, according to the state of the art the combination of an agent that activates the CD27 receptor together with one or more immune checkpoint inhibitors would result in either efficacy that is equal or similar to CD27 activation or immune checkpoint inhibition alone.

The inventors of the present invention however have surprisingly found that the combination of a CD27 agonistic antibody together with an immune checkpoint inhibitor does have additional effects on T-cell stimulation in comparison to a CD27 agonistic antibody or an immune checkpoint inhibitor alone. In particular, the additional effects of the combination of a CD27 agonistic antibody together with an immune checkpoint inhibitor has been tested in established assays that have been clinically validated to predict anti-cancer immune responses. In these assays immune checkpoint inhibitory antibodies were demonstrated to induce elevated levels of the T-cell cytokine IL-2 upon stimulation with Staphylococcus Enterotoxin B in either human peripheral blood mononuclear cells or whole human blood (Dulos *et al.*, *J. Immunother*, 2012, 35:169-78). The clinical validation and the predictive value of this assay was subsequently established in Phase I/II clinical studies (Patnaik *et al.*, ASCO, Chicago, 2012; Ribas *et al.*, PEGS Summit, Boston, 2013; Hamid *et al.*, *N. Engl. J. Med.*, 2013, 369: 134-144).

The present invention is thus based on the surprising discovery that the combination of an anti-human CD27 agonistic antibody together with an immune checkpoint

inhibitor results in immune stimulation to an unexpected level. In view of the so far known relation between CD27 and immune checkpoint inhibitors the higher level of immune stimulation resulting from the combination is surprising. In  
5 view of its surprising discovery the present invention relates to treatments of conditions ameliorated by stimulation or enhancement of the immune response, in particular the treatments of conditions that result in the stimulation or enhancement of antigen-specific T-  
10 lymphocytes, such as cancer and infectious disease. More specifically, the present invention is aimed at treatment of any condition known or expected to be ameliorated by stimulation of CD27+ immune cells or by inhibition of one or more immune checkpoint protein(s). Treatment of these  
15 conditions may be further improved by using a combination of a CD27 agonist and one or more immune checkpoint blocking agents.

According to a first aspect the invention relates to an anti-human CD27 agonistic antibody, such as hCD27.15  
20 or 1F5, or an antibody derived therefrom, for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of a condition ameliorated by stimulation of antigen-specific T-lymphocytes wherein in said treatment a number of immune checkpoint  
25 protein inhibitors is administered. By this co-administration of the anti-human CD27 agonistic antibody with a number of immune checkpoint protein inhibitors, surprising effects are obtained.

According to a further aspect the invention  
30 relates to an immune checkpoint protein inhibitor for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of a condition ameliorated by stimulation of antigen-specific T-

lymphocytes, wherein in said treatment an anti-human CD27 agonistic antibody, such as hCD27.15 or 1F5, or an antibody derived therefrom, is administered. By this co-administration of the immune checkpoint protein inhibitor with an anti-human CD27 agonistic antibody, surprising effects are obtained.

Yet a further aspect of the invention relates to a combination of an anti-human CD27 agonistic antibody, such as hCD27.15 or 1F5, or an antibody derived therefrom, together with a number of immune checkpoint protein inhibitors for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of a condition ameliorated by stimulation of antigen-specific T-lymphocytes.

Still a further aspect of the invention relates to a method for treating a condition ameliorated by stimulation of an immune response, in particular the treatment of a condition ameliorated by stimulation of antigen-specific T-lymphocytes, said method comprising administering an anti-human CD27 agonistic antibody, such as hCD27.15 or 1F5, or an antibody derived therefrom, in combination with a number of immune checkpoint inhibitors.

An anti-human CD27 antibody of the invention may be selected from hCD27.15 or analogues thereof, in particular analogues comprising the CDRs of hCD27.15, analogues (cross-)blocking the binding of hCD27.15 to human CD27, analogues binding to the same epitope of hCD27.15 or humanized analogues of hCD27.15.

In another embodiment, the anti-human CD27 antibody is administered in combination with an anti-PD1 antibody. In one embodiment, the anti-human CD27 antibody is administered in combination with nivolumab. In another embodiment, the anti-human CD27 antibody is administered in

combination with pembrolizumab. In another embodiment, the anti-human CD27 antibody is administered in combination with an anti-CTLA4 antibody. In another embodiment, the anti-human CD27 antibody is administered in combination with an anti-LAG3 antibody. In another embodiment, the anti-human CD27 antibody is administered in combination with an anti-LAG3 antibody comprising the heavy chain amino acid sequence of SEQ ID NO: 23 and the light chain amino acid sequence of SEQ ID NO: 24.

As the skilled person will know the published sequences for the heavy and light chain of nivolumab and pembrolizumab are as presented in SEQ ID NO: 21, 22, 19 and 20 respectively.

**Table 1:** Sequence Listing

SEQ ID NO:	Description
1	hCD27.15 heavy chain CDR1 (AA)
2	hCD27.15 heavy chain CDR2 (AA)
3	hCD27.15 heavy chain CDR3 (AA)
4	hCD27.15 light chain CDR1 (AA)
5	hCD27.15 light chain CDR2 (AA)
6	hCD27.15 light chain CDR3 (AA)
7	hCD27.15 heavy chain variable region (DNA)
8	hCD27.15 heavy chain variable region (AA)
9	hCD27.15 light chain variable region (DNA)
10	hCD27.15 light chain variable region (AA)
11	1F5 heavy chain CDR1 (AA)
12	1F5 heavy chain CDR2 (AA)
13	1F5 heavy chain CDR3 (AA)
14	1F5 light chain CDR1 (AA)
15	1F5 light chain CDR2 (AA)
16	1F5 light chain CDR3 (AA)
17	1F5 heavy chain variable region (AA)



18	1F5 light chain variable region (AA)
19	Pembrolizumab heavy chain (AA)
20	Pembrolizumab light chain (AA)
21	Nivolumab heavy chain (AA)
22	Nivolumab light chain (AA)
23	Anti-human LAG3 mature heavy chain (AA)
24	Anti-human LAG3 mature light chain (AA)

#### BRIEF DESCRIPTION OF THE FIGURES

**Figure 1.** CD27 agonistic antibodies in combination with immune checkpoint blockers unexpectedly enhance SEB induced T-cell activation in human PBMCs in comparison to CD27 agonistic antibodies and immune checkpoint inhibitors alone. Figure 1A. CD27 agonistic antibodies in combination with anti-PD1 antibodies enhance SEB induced T-cell activation in human PBMCs. Equal amounts of the indicated antibodies were added, with a final concentration as indicated on the X-axis. Human IgG4 was used as an isotype-matched control. Figure 1B. CD27 agonistic antibodies in combination with anti-PDL1 antibodies enhance SEB induced T-cell activation in human PBMCs. Equal amounts of the indicated antibodies were added, with a final concentration as indicated on the X-axis. Human IgG4 and/or mouse IgG1 were used as isotype-matched controls.

**Figure 2.** CD27 agonistic antibodies in combination with immune checkpoint blockers unexpectedly enhance SEB induced T-cell activation in human whole blood in comparison to CD27 agonistic antibodies and immune checkpoint inhibitors alone. Figure 2A. CD27 agonistic antibodies in combination with anti-PD1 antibodies enhance SEB induced T-cell activation in human whole blood. Equal amounts of the indicated antibodies were added, with a final concentration as indicated on the X-axis. Human IgG4 was used as isotype-matched control. Figure 2B. CD27 agonistic antibodies in

combination with anti-PDL1 antibodies enhance SEB induced T-cell activation in human whole blood. Equal amounts of the indicated antibodies were added, with a final concentration as indicated on the X-axis. Human IgG4 and/or mouse IgG1 were used as isotype-matched controls. Figure 2C. CD27 agonistic antibodies in combination with anti-CTLA-4 antibodies enhance SEB induced T-cell activation in human whole blood. Equal amounts of the indicated antibodies were added, with a final concentration as indicated on the X-axis. Human IgG4 and/or mouse IgG2A were used as isotype-matched controls.

**Figure 3.** CD27 agonistic antibodies in combination with immune checkpoint blockers enhance SEB induced T-cell activation in human whole blood in comparison to CD27 agonistic antibodies and immune checkpoint inhibitors alone. Figures 3A-3D present data for the effect of a hCD27.15 analogue in combination with anti-LAG3 (left panel) and anti-PD1 (right panel). Combination of the antibodies enhances SEB induced T-cell activation in human whole blood of different donors.

#### DETAILED DESCRIPTION

In a first aspect the invention relates to an anti-human CD27 agonistic antibody for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of a condition ameliorated by stimulation of antigen-specific T-lymphocytes, wherein in said treatment a number of immune checkpoint protein inhibitors is administered.

Anti-human CD27 agonistic antibodies should be construed as meaning an antibody demonstrating activation of the CD27 receptor on CD27<sup>+</sup> immune cells. Agonistic properties

of anti-human CD27 antibodies can be assayed by CD27 receptor activation using for example the NF- $\kappa$ B luciferase reporter assay as described in WO2012/004367. Activation of CD27 receptor using activating anti-human CD27 antibodies has been shown to induce activation, proliferation and/or survival of human CD27<sup>+</sup> immune cells. By displaying its CD27 receptor-stimulating effect the CD27 agonist is capable of inducing and/or enhancing an immune response (e.g. an antigen-specific T-cell mediated immune response). An anti-human CD27 antibody used in the present invention may exert its agonistic activity when in soluble form. Alternatively an anti-human CD27 antibody used in the present invention may exert its agonistic activity when cross-linked. For cross-linking the anti-human CD27 antibody may be adapted in respect of its Fc function. The use of anti-human CD27 antibodies exerting agonistic activity when in soluble form is preferred.

Anti-human CD27 agonistic antibodies are known in the art. For example hCD27.15 is disclosed in WO2012/004367 and 1F5 is disclosed in WO2011/130434 and Vitale et al., *Clin. Cancer Res*, 2012, 18(14): 3812-3821. The use of 1F5 or hCD27.15 or an antibody derived from one of these known antibodies is preferred. The use of hCD27.15 or an antibody analogue therefrom is most preferred in view of its beneficial binding properties and its ability to display good agonistic activity when in soluble form (without any further cross-linking). Within the present invention an antibody derived from a certain antibody may be considered an analogue. The skilled person will understand that for a proper functioning of an antibody analogue within the context of this invention, a derived antibody (or antibody analogue), according to certain embodiments, will comprise antigen binding regions of its originating antibody or will

at least bind to the same epitope. Antibody analogues may (cross-)block binding of the anti-CD27 antibody, for example hCD27.15 or 1F5, to human CD27. Antibody analogues in particular comprise antibody fragments, antibodies having  
5 modified effector function, chimeric antibodies and humanized antibodies as defined below. The antibody analogue according to the invention maintains CD27 agonistic functionality.

The heavy chain CDR1, CDR2 and CDR3 amino acid  
10 sequences and light chain CDR1, CDR2 and CDR3 amino acid sequences of hCD27.15, identifying the antigen binding region of this antibody, are already disclosed in WO2012/004367. These sequences have also been presented in SEQ ID NO: 1-6 of the sequence listing of this description,  
15 together with the amino acid sequences of the heavy chain variable region and the light chain variable region (SEQ ID NO: 8 and 10 respectively). Antibody analogues of hCD27.15 comprising these CDR sequences, or sequence variants thereof, are particularly considered for use in the  
20 invention.

The heavy chain CDR1, CDR2 and CDR3 amino acid sequences and light chain CDR1, CDR2 and CDR3 amino acid sequences of 1F5, identifying the antigen binding region of this antibody, are already disclosed in WO2011/130434. These  
25 sequences have also been presented in SEQ ID NO: 11-16 of the sequence listing of this description, together with the amino acid sequences of the heavy chain variable region and the light chain variable region (SEQ ID NO: 17 and 18). Antibody analogues of 1F5 comprising these CDR sequences, or  
30 sequence variants thereof, are particularly considered for use in the invention.

Alternatively, analogues of hCD27.15 or 1F5 binding to the same epitope of these antibodies, but having

differing CDRs may also be selected. To screen for antibodies that bind to the hCD27.15 or 1F5 epitope on human CD27, a routine cross-blocking assay such as that described in "Antibodies, A Laboratory Manual, Cold Spring Harbor Laboratory, Ed Harlow and David Lane (1988)", can be performed. Antibodies that bind to the same epitope are likely to cross-block in such assays, but not all cross-blocking antibodies will necessarily bind at precisely the same epitope since cross-blocking may result from steric hindrance of antibody binding by antibodies binding at overlapping epitopes, or even nearby non-overlapping epitopes. Such cross-blocking antibodies maintaining CD27 agonistic functionality are also within the scope of the present invention.

Alternatively, the known technique of epitope mapping, e.g., as described in Champe *et al.*, 1995, *J. Biol. Chem.* 270:1388-1394, can be performed to determine whether the antibody binds an epitope of interest. "Alanine scanning mutagenesis," as described by Cunningham and Wells, 1989, *Science* 244: 1081-1085, or some other form of point mutagenesis of amino acid residues in human CD27 may also be used to determine the functional epitope for anti-CD27 antibodies of the present invention.

Another known method to map the epitope of an antibody is to study binding of the antibody to synthetic linear and CLIPS peptides that can be screened using credit-card format mini PEPSCAN cards as described by Slootstra *et al.* (Slootstra *et al.*, 1996, *Mol. Diversity* 1: 87-96) and Timmerman *et al.* (Timmerman *et al.*, 2007, *J. Mol. Recognit.* 20: 283-299). The binding of antibodies to each peptide is determined in a PEPSCAN-based enzyme-linked immuno assay (ELISA).

Additional antibodies binding to the same epitope as hCD27.15 may be obtained with known techniques, for example, by screening of antibodies raised against CD27 for binding to the epitope, or by immunization of an animal with a peptide comprising a fragment of human CD27 comprising the epitope sequences. Antibodies that bind to the same functional epitope might be expected to exhibit similar biological activities, such as CD27 agonistic activity, and such activities can be confirmed by functional assays of the antibodies. For analogues of 1F5 binding to the same epitope of this antibody, these techniques can also be used in analogy.

According to an embodiment an analogue of hCD27.15 binding to the same epitope of this antibody, but having differing CDRs may block binding of hCD27.15 to human CD27 with an  $IC_{50}$  of about 50 nM or lower. Alternatively an analogue of hCD27.15 binding to the same epitope of this antibody, but having differing CDRs may be blocked in its binding to human CD27 by hCD27.15 with an  $IC_{50}$  of about 50 nM or lower. Similarly analogues of 1F5 binding to the same epitope of this antibody, but having differing CDRs may block binding of 1F5 to human CD27 with an  $IC_{50}$  of about 50 nM or lower or alternatively may be blocked in its binding to human CD27 by 1F5 with an  $IC_{50}$  of about 50 nM or lower. About 50 nM or lower is to be understood to include  $50 \times 10^{-9}$  to  $0.1 \times 10^{-12}$  M, such as  $20 \times 10^{-9}$  to  $1.0 \times 10^{-11}$  M,  $10 \times 10^{-9}$  to  $1.0 \times 10^{-10}$  M or  $10 \times 10^{-9}$  to  $1.0 \times 10^{-19}$ .

The differing CDRs may be sequence variants of the known CDRs of the human CD27 binding antibody used in the invention, for example hCD27.15 or 1F5. As used herein, a sequence "variant" refers to a sequence that differs from the disclosed sequence at one or more amino acid residues but which retains the biological activity of the resulting

molecule. The invention includes the variants of antibodies explicitly disclosed by the various sequences, for example hCD27.15 or 1F5. For the V<sub>H</sub> domain CDR1, CDR2 and CDR3 sequences, according to some embodiments, variant sequences may comprise up to 6 amino acid substitutions, such as 1, 2, 3, 4, 5 or 6 amino acid substitutions, for the CDR1, CDR2 and CDR3 sequences taken together. Similarly for the V<sub>L</sub> domain CDR1, CDR2 and CDR3 sequences, according to some embodiments, variant sequences may comprise up to 6 amino acid substitutions, such as 1, 2, 3, 4, 5 or 6 amino acid substitutions, for the CDR1, CDR2 and CDR3 sequences taken together. The skilled person will understand that in particular conservative amino acid substitutions may result in maintaining biological activity. For all amino acid and DNA sequences disclosed, the sequence variants are also envisaged within this invention.

"Conservatively modified variants" or "conservative amino acid substitution" refers to substitutions of amino acids are known to those of skill in this art and may be made generally without altering the biological activity of the resulting molecule. Those of skill in this art recognize that, in general, single amino acid substitutions in non-essential regions of a polypeptide do not substantially alter biological activity (see, e.g., Watson, et al., *Molecular Biology of the Gene*, The Benjamin/Cummings Pub. Co., p. 224 (4th Edition 1987)). Such exemplary substitutions are preferably made in accordance with those set forth above in **Table 2**.

**Table 2:** Exemplary Conservative Amino Acid Substitutions

Original residue	Conservative substitution
Ala (A)	Gly; Ser
Arg (R)	Lys, His

Asn (N)	Gln; His
Asp (D)	Glu; Asn
Cys (C)	Ser; Ala
Gln (Q)	Asn
Glu (E)	Asp; Gln
Gly (G)	Ala
His (H)	Asn; Gln
Ile (I)	Leu; Val
Leu (L)	Ile; Val
Lys (K)	Arg; His
Met (M)	Leu; Ile; Tyr
Phe (F)	Tyr; Met; Leu
Pro (P)	Ala
Ser (S)	Thr
Thr (T)	Ser
Trp (W)	Tyr; Phe
Tyr (Y)	Trp; Phe
Val (V)	Ile; Leu

The anti-human CD27 agonistic antibody of the invention is intended for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of conditions which are ameliorated by stimulation of antigen-specific T-lymphocytes. Characteristic of this treatment is that a number of immune checkpoint protein inhibitors is administered. Thus the CD27 agonistic antibody is co-administered with a number of immune checkpoint protein inhibitors. Within the present invention the term "number of" should be understood as meaning at least one or alternatively one or more, such as, 1, 2, 3, 4, 5 or 6.

The term "immune checkpoint protein" is known in the art. Within the known meaning of this term it will be



clear to the skilled person that on the level of "immune checkpoint proteins" the immune system provides inhibitory signals to its components in order to balance immune reactions. Known immune checkpoint proteins comprise CTLA-4, PD1 and its ligands PD-L1 and PD-L2 and in addition LAG-3, BTLA, B7H3, B7H4, TIM3, KIR. The pathways involving LAG3, BTLA, B7H3, B7H4, TIM3, and KIR are recognized in the art to constitute immune checkpoint pathways similar to the CTLA-4 and PD-1 dependent pathways (see e.g. Pardoll, 2012. *Nature Rev Cancer* 12:252-264; Mellman et al., 2011. *Nature* 480:480-489).

Within the present invention an immune checkpoint protein inhibitor is any compound inhibiting the function of an immune checkpoint protein. Inhibition includes reduction of function and full blockade. In particular the immune checkpoint protein is a human immune checkpoint protein. Thus the immune checkpoint protein inhibitor preferably is an inhibitor of a human immune checkpoint protein. Immune checkpoint proteins are described in the art (see for instance Pardoll, 2012. *Nature Rev. cancer* 12: 252-264). The designation immune checkpoint includes the experimental demonstration of stimulation of an antigen-receptor triggered T lymphocyte response by inhibition of the immune checkpoint protein *in vitro* or *in vivo*, e.g. mice deficient in expression of the immune checkpoint protein demonstrate enhanced antigen-specific T lymphocyte responses or signs of autoimmunity (such as disclosed in Waterhouse et al., 1995. *Science* 270:985-988; Nishimura et al., 1999. *Immunity* 11:141-151). It may also include demonstration of inhibition of antigen-receptor triggered CD4+ or CD8+ T cell responses due to deliberate stimulation of the immune checkpoint protein *in vitro* or *in vivo* (e.g. Zhu et al., 2005. *Nature Immunol.* 6:1245-1252).

Preferred immune checkpoint protein inhibitors are antibodies that specifically recognize immune checkpoint proteins. A number of CTLA-4, PD1, PDL-1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 and KIR inhibitors are known and in analogy of these known immune checkpoint protein inhibitors, alternative immune checkpoint inhibitors may be developed in the (near) future. For example ipilimumab is a fully human CTLA-4 blocking antibody presently marketed under the name Yervoy (Bristol-Myers Squibb). A second CTLA-4 inhibitor is tremelimumab (referenced in Ribas et al., 2013, *J. Clin. Oncol.* 31:616-22). Examples of PD-1 inhibitors include without limitation humanized antibodies blocking human PD-1 such as lambrolizumab (e.g. disclosed as hPD109A and its humanized derivatives h409A11, h409A16 and h409A17 in WO2008/156712; Hamid et al., *N. Engl. J. Med.* 369: 134-144 2013,)), or pidilizumab (disclosed in Rosenblatt et al., 2011. *J Immunother.* 34:409-18), as well as fully human antibodies such as nivolumab (previously known as MDX-1106 or BMS-936558, Topalian et al., 2012. *N. Eng. J. Med.* 366:2443-2454, disclosed in US8008449 B2). Other PD-1 inhibitors may include presentations of soluble PD-1 ligand including without limitation PD-L2 Fc fusion protein also known as B7-DC-Ig or AMP-244 (disclosed in Mkrtichyan M, et al. *J Immunol.* 189:2338-47 2012) and other PD-1 inhibitors presently under investigation and/or development for use in therapy. In addition, immune checkpoint inhibitors may include without limitation humanized or fully human antibodies blocking PD-L1 such as MEDI-4736 (disclosed in WO2011066389 A1), MPDL3280A (disclosed in US8217149 B2) and MIH1 (Affymetrix obtainable via eBioscience (16.5983.82)) and other PD-L1 inhibitors presently under investigation. According to this invention an immune checkpoint inhibitor is preferably selected from a CTLA-4, PD-1 or PD-L1

inhibitor, such as selected from the known CTLA-4, PD-1 or PD-L1 inhibitors mentioned above (ipilimumab, tremelimumab, labrolizumab, nivolumab, pidilizumab, AMP-244, MEDI-4736, MPDL3280A, MIH1). Known inhibitors of these immune

5 checkpoint proteins may be used as such or analogues may be used, in particular chimerized, humanized or human forms of antibodies.

As the skilled person will know, alternative and/or equivalent names may be in use for certain antibodies  
10 mentioned above. Such alternative and/or equivalent names are interchangeable in the context of the present invention. For example it is known that lambrolizumab is also known under the alternative and equivalent names MK-3475 and pembrolizumab.

15 The selection of an immune checkpoint inhibitor from PD1 and PD-L1 inhibitors, such as a known PD-1 or PD-L1 inhibitor mentioned above, is more preferred and most preferably a selection is made from a PD-1 inhibitor, such as a known PD1 inhibitor mentioned above. In preferred  
20 embodiments, the PD1 inhibitor is nivolumab or pembrolizumab or another antagonist antibody against human PD1.

The invention also includes the selection of other immune checkpoint inhibitors that are known in the art to stimulate immune responses. This includes inhibitors that  
25 directly or indirectly stimulate or enhance antigen-specific T-lymphocytes. These other immune checkpoint inhibitors include, without limitation, agents targeting immune checkpoint proteins and pathways involving PD-L2, LAG3, BTLA, B7H4 and TIM3. For example, human PD-L2 inhibitors  
30 known in the art include MIH18 (disclosed in (Pfistershammer et al., 2006. Eur J Immunol. 36:1104-13). Another example, LAG3 inhibitors known in the art include soluble LAG3 (IMP321, or LAG3-Ig disclosed in WO2009044273 A2, and in

Brignon et al. 2009. Clin. Cancer Res. 15:6225-6231) as well as mouse or humanized antibodies blocking human LAG3 (for instance IMP701 disclosed in and derived from WO2008132601 A1), or fully human antibodies blocking human LAG3 (such as  
5 disclosed in EP 2320940 A2). Another example is provided by the use of blocking agents towards BTLA, including without limitation antibodies blocking human BTLA interaction with its ligand (such as 4C7 disclosed in WO2011014438). Yet another example is provided by the use of agents  
10 neutralizing B7H4 including without limitation antibodies to human B7H4 (disclosed in WO 2013025779 A1, and in WO 2013067492 A1) or soluble recombinant forms of B7H4 (such as disclosed in US20120177645 A1 or Anti-human B7H4 clone H74: eBiocience # 14-5948). Yet another example is provided by  
15 agents neutralizing B7-H3, including without limitation antibodies neutralizing human B7-H3 (e.g. MGA271 disclosed as BRCA84D and derivatives in US 20120294796 A1). Yet another example is provided by agents targeting TIM3, including without limitation antibodies targeting human TIM3  
20 (e.g. as disclosed in WO 2013006490 A2 or the anti-human TIM3, blocking antibody F38-2E2 disclosed by Jones et al., J Exp Med. 2008 Nov 24;205(12):2763-79). Known inhibitors of immune checkpoint proteins may be used in their known form or analogues may be used, in particular chimerized forms of  
25 antibodies, most preferably humanized forms.

The invention also includes the selection of more than one immune checkpoint inhibitor selected from CTLA-4, PD-1 or PDL1 inhibitors for combination with an anti-human CD27 agonistic antibody within the various aspects of the  
30 invention. For example concurrent therapy of ipilimumab (anti-CTLA4) with Nivolumab (anti-PD1) has demonstrated clinical activity that appears to be distinct from that obtained in monotherapy (Wolchok et al., 2013, *N. Eng. J.*

*Med.*, 369:122-33). Also included are combinations of agents that have been shown to improve the efficacy of checkpoint inhibitors, such as Lirilumab (also known as anti-KIR, BMS-986015 or IPH2102, as disclosed in US8119775 B2 and Benson et al., *Blood* 120:4324-4333 (2012)) in combination with ipilimumab (Rizvi et al., ASCO 2013, and [clinicaltrials.gov](http://clinicaltrials.gov) NCT01750580) or in combination with nivolumab (Sanborn et al., ASCO 2013, and [clinicaltrials.gov](http://clinicaltrials.gov) NCT01714739), agents targeting LAG3 combined with anti-PD-1 (Woo et al., 2012 Cancer Res. 72:917-27) or anti-PD-L1 (Butler NS et al., *Nat Immunol.* 2011 13:188-95), agents targeting ICOS in combination with anti-CTLA-4 (Fu et al., *Cancer Res.* 2011 71:5445-54, or agents targeting 4-1BB in combination with anti-CTLA-4 (Curran et al., *PLoS One.* 2011 6(4):e19499).

Combinations of anti-human CD27 agonistic antibodies and immune checkpoint inhibitors envisaged within the various embodiments of the present invention include those presented in Table 3. In this table the names of anti-human CD27 agonistic antibodies and immune checkpoint inhibitors refers to both the known compound and analogues thereof. For antibodies, analogues include analogues having modified Fc-fuction, chimerized antibodies and humanized antibodies. For antibodies preferably human or humanized forms are selected. CD27 agonist refers to an anti-human CD27 agonistic antibody.



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In the table above presenting combinations of anti-human CD27 antibodies with Immune checkpoint inhibitors of the invention, numbers indicated (N) refer to the number of immune checkpoint inhibitors (N=1 or N=2) combined with an anti-human CD27 agonistic antibody.

- 5 When a number is presented the combination comprises the immune checkpoint inhibitor(s) listed in the column at the position of the number.

Most of the immune checkpoint protein inhibitors  
 10 presently known are antibodies. In view of the advances in antibody technology the use of an antibody as an immune checkpoint protein inhibitor is preferred in the present invention. However, the use of alternative immune checkpoint inhibitors based on other technologies is also envisaged in  
 15 alternative embodiments.

As the skilled person will know other technologies are available for developing binding compounds interfering with the function of proteins such as immune checkpoint

proteins and thus acting as an inhibitor. For example as the skilled person will understand, a library of binding peptides engineered on non-immunoglobulin protein scaffolds can be used to select binding peptides that inhibit immune checkpoint proteins. Examples of such protein scaffolds include, but are not restricted to Adnectins, Affibodies, Anticalins and DARPins (Gebauer and Skerra, *Current opinion Chem. Biol.*, 2009, 13:245-255 and Caravella and Lugovskoy, *Current opinion Chem. Biol.*, 2010, 14:520-528). Selection methods for example include phage display to identify protein scaffolds that express peptides binding to immune check point proteins. In addition, combinatorial peptide libraries comprising peptides potentially presenting immune checkpoint protein inhibitory functions may be screened for suitable immune checkpoint protein inhibitors. From such a library For example, one-bead-one-compound combinatorial libraries expressing a broad set of peptides on beads, where one bead is binding one peptide, may be used. After selection procedures, beads are recovered and the peptide is identified (Lam *et al.*, *Methods*, 1996, 9:482-93; Xiao *et al.*, *Comb. Chem. High Throughput Screen*, 2013 16:441-8). For example using mass-spectrometry methods.

In the present invention the term "antibody" is used in the broadest sense and specifically covers, but is not limited to, monoclonal antibodies (including full length monoclonal antibodies), polyclonal antibodies, and multispecific antibodies (e.g., bispecific antibodies) and binding fragments thereof.

"Antibody fragment" and "antibody binding fragment" mean antigen-binding fragments of an antibody, typically including at least a portion of the antigen binding or variable regions (e.g. one or more CDRs) of the parental antibody. An antibody fragment retains at least

some of the binding specificity of the parental antibody. Typically, an antibody fragment retains at least 10% of the parental binding activity when that activity is expressed on a molar basis. Preferably, an antibody fragment retains at least 20%, 50%, 70%, 80%, 90%, 95% or 100% or more of the parental antibody's binding affinity for the target.

Therefore, as is clear for the skilled person, "antibody fragments" in many applications may substitute antibodies and the term "antibody" should be understood as including "antibody fragments" when such a substitution is suitable. Examples of antibody fragments include, but are not limited to, Fab, Fab', F(ab')<sub>2</sub>, and Fv fragments; diabodies; linear antibodies; single-chain antibody molecules, e.g., sc-Fv, unibodies or duobodies (technology from Genmab); nanobodies (technology from Ablynx); domain antibodies (technology from Domantis); and multispecific antibodies formed from antibody fragments. Engineered antibody variants are reviewed in Holliger and Hudson, 2005, Nat. Biotechnol. 23:1126-1136.

An "Fab fragment" is comprised of one light chain and the CH1 and variable regions of one heavy chain. The heavy chain of a Fab molecule cannot form a disulfide bond with another heavy chain molecule.

An "Fc" region contains two heavy chain fragments comprising the C<sub>H</sub>1 and C<sub>H</sub>2 domains of an antibody. The two heavy chain fragments are held together by two or more disulfide bonds and by hydrophobic interactions of the C<sub>H</sub>3 domains.

An "Fab' fragment" contains one light chain and a portion of one heavy chain that contains the V<sub>H</sub> domain and the C<sub>H</sub>1 domain and also the region between the C<sub>H</sub>1 and C<sub>H</sub>2 domains, such that an interchain disulfide bond can be formed between the two heavy chains of two Fab' fragments to form a F(ab')<sub>2</sub> molecule.

An "F(ab')<sub>2</sub> fragment" contains two light chains and two heavy chains containing a portion of the constant region between the C<sub>H1</sub> and C<sub>H2</sub> domains, such that an interchain disulfide bond is formed between the two heavy chains. A  
5 F(ab')<sub>2</sub> fragment thus is composed of two Fab' fragments that are held together by a disulfide bond between the two heavy chains.

The "Fv region" comprises the variable regions from both the heavy and light chains, but lacks the constant  
10 regions.

A "single-chain Fv antibody" (or "scFv antibody") refers to antibody fragments comprising the V<sub>H</sub> and V<sub>L</sub> domains of an antibody, wherein these domains are present in a single polypeptide chain. Generally, the Fv polypeptide  
15 further comprises a polypeptide linker between the V<sub>H</sub> and V<sub>L</sub> domains which enables the scFv to form the desired structure for antigen binding. For a review of scFv, see Pluckthun, 1994, The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenberg and Moore eds. Springer-Verlag, New York, pp. 269-  
20 315. See also, International Patent Application Publication No. WO 88/01649 and U.S. Pat. Nos. 4,946, 778 and 5,260,203.

A "diabody" is a small antibody fragment with two antigen-binding sites. The fragments comprises a heavy chain variable domain (V<sub>H</sub>) connected to a light chain variable  
25 domain (V<sub>L</sub>) in the same polypeptide chain (V<sub>H</sub>-V<sub>L</sub> or V<sub>L</sub>-V<sub>H</sub>). By using a linker that is too short to allow pairing between the two domains on the same chain, the domains are forced to pair with the complementary domains of another chain and create two antigen-binding sites. Diabodies are described  
30 more fully in, e.g., EP 404,097; WO 93/11161; and Holliger et al., 1993, Proc. Natl. Acad. Sci. USA 90: 6444-6448.

A "domain antibody fragment" is an immunologically functional immunoglobulin fragment containing only the

variable region of a heavy chain or the variable region of a light chain. In some instances, two or more  $V_H$  regions are covalently joined with a peptide linker to create a bivalent domain antibody fragment. The two  $V_H$  regions of a bivalent domain antibody fragment may target the same or different antigens.

An antibody fragment of the invention may comprise a sufficient portion of the constant region to permit dimerization (or multimerization) of heavy chains that have reduced disulfide linkage capability, for example where at least one of the hinge cysteines normally involved in inter-heavy chain disulfide linkage is altered with known methods available to the skilled person. In another embodiment, an antibody fragment, for example one that comprises the Fc region, retains at least one of the biological functions normally associated with the Fc region when present in an intact antibody, such as FcRn binding, antibody half life modulation, ADCC (antibody dependent cellular cytotoxicity) function, and/or complement binding (for example, where the antibody has a glycosylation profile necessary for ADCC function or complement binding).

The antibody is directed against human CD27 and thus comprises binding domains that bind to and/or interact with human CD27. The antibody may be raised in an animal from a non-human species suitable for eliciting antibodies against human antigens. Alternatively, the antibody may be isolated from antibody phage libraries generated using the techniques described in McCafferty *et al.*, 1990, *Nature*, 348:552-554. Clackson *et al.*, 1991, *Nature*, 352:624-628, and Marks *et al.*, 1991, *J. Mol. Biol.* 222:581-597. The skilled person will be able to select a suitable non-human species for eliciting antibodies against human antigens. For example a selection may be made from a non-human mammal, such as a

rodent, including murine (rat or mouse) or hamster species, or alternatively a camelid species.

The antibody, when raised in a non-human species, preferably is chimerized with methods and techniques known in the art to form a "chimeric antibody". The term "chimeric" antibody refers to antibodies in which a portion of the heavy and/or light chain is identical with or homologous to corresponding sequences in antibodies derived from a particular species or belonging to a particular antibody class or subclass, while the remainder of the chain(s) is identical with or homologous to corresponding sequences in antibodies derived from another species or belonging to another antibody class or subclass, as well as fragments of such antibodies, so long as they exhibit the desired biological activity (See, for example, U.S. Pat. No. 4,816,567 and Morrison et al., 1984, Proc. Natl. Acad. Sci. USA 81:6851-6855). Within the present invention a "chimeric antibody" preferably is a "humanized antibody".

As used herein, the term "humanized antibody" refers to forms of antibodies that contain sequences from non-human (e.g., murine) antibodies as well as human antibodies. Such antibodies contain minimal sequence derived from non-human immunoglobulin. In general, the humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the hypervariable loops correspond to those of a non-human immunoglobulin and all or substantially all of the FR regions are those of a human immunoglobulin sequence. The humanized antibody optionally also will comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin. The humanized forms of rodent antibodies will essentially comprise the same CDR sequences of the parental rodent

antibodies, although certain amino acid substitutions may be included to increase affinity, increase stability of the humanized antibody, or for other reasons. However, as CDR loop exchanges do not uniformly result in an antibody with the same binding properties as the antibody of origin, changes in framework residues (FR), residues involved in CDR loop support, might also be introduced in humanized antibodies to preserve antigen binding affinity (Kabat et al., 1991, J. Immunol. 147:1709).

The term "antibody" also includes "fully human" antibodies, i.e., antibodies that comprise human immunoglobulin protein sequences only. A fully human antibody may contain non-human, such as murine (rat or mouse) carbohydrate chains if produced in a non-human cell (e.g. mouse or hamster), or in a hybridoma derived from a murine cell. Similarly, "mouse antibody" or "rat antibody" refer to an antibody that comprises only mouse or rat immunoglobulin sequences, respectively.

A fully human antibody may be generated in a human being, in a transgenic non-human animal having human immunoglobulin germline sequences, by phage display or other molecular biological methods. Also, recombinant immunoglobulins may also be made in transgenic mice. See Mendez et al., 1997, Nature Genetics 15:146-156. See also Abgenix, Medarex, MeMo and Kymab technologies.

The antibodies of the present invention also include antibodies with modified (or blocked) Fc regions to provide altered effector functions. See, e.g. U.S. Pat. No. 5,624,821; WO2003/086310; WO2005/120571; WO2006/0057702; Presta, 2006, *Adv. Drug Delivery Rev.* 58:640-656; Vincent and Zurini, *Biotechnol. J.*, 2012, 7:1444-50; Kaneko and Niwa, *Biodrugs*, 2011, 25: 1-11. Such modification can be used to enhance or suppress various reactions of the immune



system, with possible beneficial effects in diagnosis and therapy. Alterations of the Fc region include amino acid changes (substitutions, deletions and insertions), glycosylation or deglycosylation, and adding multiple Fc.

5 Preferably, Fc regions displaying reduced Fc effector functions are used. The antibodies of the present invention also include antibodies that have a human IgG4 containing Fc regions. And/or Fc regions carrying a N297Q glycosylation deficient mutant are used.

10 Changes to the Fc can also alter the half-life of antibodies in therapeutic antibodies, and a longer half-life would result in less frequent dosing, with the concomitant increased convenience and decreased use of material. See Presta, 2005, J. Allergy Clin. Immunol. 116:731 at 734-35.

15 The antibodies of the present invention, although less preferred, also include antibodies with intact Fc regions that provide full effector functions, e.g. antibodies of isotype IgG1, which induce complement-dependent cytotoxicity (CDC) or antibody dependent cellular  
20 cytotoxicity (ADCC) in the a cell associated with the target for the antibody.

The antibodies may also be conjugated (e.g., covalently linked) to molecules that improve stability of the antibody during storage or increase the half-life of the  
25 antibody *in vivo*. Examples of molecules that increase the half-life are albumin (e.g., human serum albumin) and polyethylene glycol (PEG). Albumin-linked and PEGylated derivatives of antibodies can be prepared using techniques well known in the art. See, e.g. Chapman, 2002, Adv. Drug  
30 Deliv. Rev. 54:531-545; Anderson and Tomasi, 1988, J. Immunol. Methods 109:37-42; Suzuki et al., 1984, Biochim. Biophys. Acta 788:248-255; and Brekke and Sandlie, 2003, Nature Rev. 2:52-62.

As used herein, the term "about" refers to a value that is within an acceptable error range for the particular value as determined by one of ordinary skill in the art, which will depend in part on how the value is measured or  
5 determined, i.e. the limitations of the measurement system. For example, "about" can mean within 1 or more than 1 standard deviation per the practice in the art.

Alternatively, "about" or "comprising essentially of" can mean a range of up to 20%. Furthermore, particularly with  
10 respect to biological systems or processes, the terms can mean up to an order of magnitude or up to 5-fold of a value. When particular values are provided in the application and claims, unless otherwise stated, the meaning of "about" or "comprising essentially of" should be assumed to be within  
15 an acceptable error range for that particular value.

The antibody can be selected from any class of immunoglobulins, including IgM, IgG, IgD, IgA, and IgE. Preferably, the antibody is an IgG antibody. Any isotype of IgG can be used, including IgG1, IgG2, IgG3, and IgG4.

20 Variants of the IgG isotypes are also contemplated. The humanized antibody may comprise sequences from more than one class or isotype. Optimization of the necessary constant domain sequences to generate the desired biologic activity is readily achieved by screening the antibodies in the  
25 biological assays described in the Examples.

Likewise, either class of light chain can be used in the compositions and methods herein. Specifically, kappa, lambda, or variants thereof are useful in the present compositions and methods.

30 The term "monoclonal antibody" as used herein refers to an antibody obtained from a population of substantially homogeneous antibodies, i.e., the individual antibodies comprising the population are identical except

for possible naturally occurring mutations that may be present in minor amounts. Monoclonal antibodies are highly specific, being directed against a single antigenic site. Furthermore, in contrast to conventional (polyclonal) antibody preparations that typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. The modifier "monoclonal" indicates the character of the antibody as being obtained from a substantially homogeneous population of antibodies, and is not to be construed as requiring production of the antibody by any particular method. For example, the monoclonal antibodies to be used in accordance with the present invention may be made by the hybridoma method first described by Kohler et al., 1975, *Nature* 256:495, or may be made by recombinant DNA methods (see, for example, U.S. Pat. No. 4,816,567). The "monoclonal antibodies" may also be isolated from phage antibody libraries using the techniques described in Clackson et al., 1991, *Nature* 352:624-628 and Marks et al., 1991, *J. Mol. Biol.* 222:581-597, for example. The monoclonal antibodies herein specifically include "chimeric" antibodies.

Monoclonal antibodies can be made according to knowledge and skill in the art of injecting test subjects with human antigen and then generating hybridomas expressing antibodies having the desired sequence or functional characteristics. DNA encoding the monoclonal antibodies is readily isolated and sequenced using conventional procedures (e.g., by using oligonucleotide probes that are capable of binding specifically to genes encoding the heavy and light chains of the monoclonal antibodies). The hybridoma cells serve as a preferred source of such DNA.

For therapeutic applications the antibodies may be used as such or as a treatment conjugate. As used herein, a treatment "conjugate" refers to an antibody, or a fragment thereof, conjugated to a therapeutic moiety, such as a bacterial toxin, a cytotoxic drug or a radiotoxin. Toxic moieties can be conjugated to antibodies, of the invention using methods available in the art.

In view of the fact that the present invention resides in the surprising immune stimulatory effects of the combination of an anti-human CD27 agonistic antibody together with an immune checkpoint inhibitor, the present invention, in its various embodiments is suitable for treatment of a condition known or expected to be ameliorated by immune stimulation, in particular stimulation of antigen-specific T-lymphocytes. In this invention the term antigen-specific T-lymphocyte in particular includes CD4+ and/or CD8+ T cells.

The immune stimulation, in particular the stimulation of antigen-specific T-lymphocytes, may be achieved by stimulation of CD27<sup>+</sup> immune cells or by inhibition of an immune checkpoint protein, such as CTLA-4, PD1, PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 and KIR. Thus according to certain embodiments the present invention, is suitable for treatment of a condition known or expected to be ameliorated by stimulation of CD27<sup>+</sup> immune cells. According to certain alternative embodiments the present invention, is suitable for treatment of a condition known or expected to be ameliorated by inhibition of an immune checkpoint protein, such as CTLA-4, PD1, PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 or KIR.

The meaning of the term stimulation in the context of an immune response will be known to the skilled person. It will be clear that this term includes enhancement and

thus relates to both elevation of existing immune responses and induction or *de novo* generation of an immune response.

The skilled person will know which cells are associated with CD27, CTLA-4, PD1, PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 and KIR. In particular it is known that CD27 positive cells may or may not express on their cell surface CTLA-4, PD1, PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 and KIR (and *vice versa*). Thus conditions treatable within the present invention are most certainly not restricted to conditions involving cells expressing both CD27 and an immune checkpoint protein, such as CTLA-4, PD-1 or PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 or KIR.

The various aspects of the invention are aimed at "treatment of a condition ameliorated by stimulation of an immune response, in particular stimulation of antigen-specific T-lymphocytes". "The treatment of a condition ameliorated by stimulation of an immune response, in particular stimulation of antigen-specific T-lymphocytes" may alternatively be defined as "a treatment wherein stimulation of an immune response, in particular stimulation of antigen-specific T-lymphocytes, is beneficial". The terms "amelioration" and "beneficial" in the context of treatment both refer to clinically meaning full improvement as can be predicted and/or established by a physician.

Within the present invention the treatment of the "condition" includes any therapeutic use including prophylactic and curative uses of the anti-human CD27 agonistic antibody and the number of immune checkpoint inhibitors. Therefore the term "condition" may refer to disease states but also to physiological states in the prophylactic setting where physiology is not altered to a detrimental state. Conditions in the setting of prophylactic use of the anti-human CD27 agonistic antibody and the number

of immune checkpoint inhibitors include for example immunization (the physiological process of inducing and/or generating immune memory against an antigen) after vaccination. The treatment within the context of the present invention thus may be aimed at supporting prophylactically induced physiological processes.

Conditions ameliorated by immune stimulation, in particular stimulation of antigen-specific T-lymphocytes include infectious diseases, such as bacterial, fungal, viral and parasitic infectious diseases. In addition immunization after vaccination may also be ameliorated by immune stimulation, in particular stimulation of antigen-specific T-lymphocytes. The vaccination may be against a pathogen, such as pathogen selected from bacteria, fungi, viruses or parasites, or against toxins, or self-antigens, including antigens expressed on benign or malignant tumors, including cancers. Also conditions associated with uncontrolled proliferation of cells such as cancers may be ameliorated by immune stimulation, in particular stimulation of antigen-specific T-lymphocytes. These conditions may be ameliorated by stimulation of CD27<sup>+</sup> immune cells and/or by inhibition of an immune checkpoint protein, such as CTLA-4, PD1, PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 or KIR.

Cancers within the present invention include, but are not limited to, leukemia, acute lymphocytic leukemia, acute myelocytic leukemia, myeloblasts promyelocyte, myelomonocytic monocytic erythroleukemia, chronic leukemia, chronic myelocytic (granulocytic) leukemia, chronic lymphocytic leukemia, mantle cell lymphoma, primary central nervous system lymphoma, Burkitt's lymphoma and marginal zone B cell lymphoma, Polycythemia vera Lymphoma, Hodgkin's disease, non-Hodgkin's disease, multiple myeloma, Waldenstrom's macroglobulinemia, heavy chain disease, solid

tumors, sarcomas, and carcinomas, fibrosarcoma, myxosarcoma, liposarcoma, chondrosarcoma, osteogenic sarcoma, osteosarcoma, chordoma, angiosarcoma, endotheliosarcoma, lymphangiosarcoma, lymphangioendotheliosarcoma, synovioma, mesothelioma, Ewing's tumor, leiomyosarcoma, rhabdomyosarcoma, colon sarcoma, colorectal carcinoma, pancreatic cancer, breast cancer, ovarian cancer, prostate cancer, squamous cell carcinoma, basal cell carcinoma, adenocarcinoma, sweat gland carcinoma, sebaceous gland carcinoma, papillary carcinoma, papillary adenocarcinomas, cystadenocarcinoma, medullary carcinoma, bronchogenic carcinoma, renal cell carcinoma, hepatoma, bile duct carcinoma, choriocarcinoma, seminoma, embryonal carcinoma, Wilm's tumor, cervical cancer, uterine cancer, testicular tumor, lung carcinoma, small cell lung carcinoma, non-small cell lung carcinoma, bladder carcinoma, epithelial carcinoma, glioma, astrocytoma, medulloblastoma, craniopharyngioma, ependymoma, pinealoma, hemangioblastoma, acoustic neuroma, oligodendroglioma, menangioma, melanoma, neuroblastoma, retinoblastoma, nasopharyngeal carcinoma, esophageal carcinoma, basal cell carcinoma, biliary tract cancer, bladder cancer, bone cancer, brain and central nervous system (CNS) cancer, cervical cancer, choriocarcinoma, colorectal cancers, connective tissue cancer, cancer of the digestive system, endometrial cancer, esophageal cancer, eye cancer, head and neck cancer, gastric cancer, intraepithelial neoplasm, kidney cancer, larynx cancer, liver cancer, lung cancer (small cell, large cell), melanoma, neuroblastoma; oral cavity cancer(for example lip, tongue, mouth and pharynx), ovarian cancer, pancreatic cancer, retinoblastoma, rhabdomyosarcoma, rectal cancer; cancer of the respiratory system, sarcoma, skin cancer,

stomach cancer, testicular cancer, thyroid cancer, uterine cancer, and cancer of the urinary system.

Less preferred cancers include CD27-expressing tumors, such as those selected from the group consisting of chronic lymphocytic leukemia, mantle cell lymphoma, primary central nervous system lymphoma, Burkitt's lymphoma and marginal zone B cell lymphoma.

Some examples of pathogenic viruses causing infections ameliorated by immune stimulation, in particular stimulation of antigen-specific T-lymphocytes include HIV, hepatitis (A, B, C, D or E), herpes virus (e.g., VZV, HSV-1, HAV-6, HSV-II, and CMV, Epstein Barr virus), adenovirus, influenza virus, flaviviruses, echovirus, rhinovirus, coxsackie virus, cornovirus, respiratory syncytial virus, mumps virus, rotavirus, measles virus, rubella virus, parvovirus, vaccinia virus, HTLV virus, dengue virus, papillomavirus, molluscum virus, poliovirus, rabies virus, JC virus and arboviral encephalitis virus.

Some examples of pathogenic bacteria causing infections ameliorated by immune stimulation, in particular stimulation of antigen-specific T-lymphocytes, include chlamydia, rickettsial bacteria, mycobacteria, staphylococci, streptococci, pneumococci, meningococci and conococci, klebsiella, proteus, serratia, pseudomonas, legionella, diphtheria, salmonella, bacilli, cholera, tetanus, botulism, anthrax, plague, leptospirosis, and Lyme disease bacteria.

Some examples of pathogenic fungi causing infections ameliorated by immune stimulation, in particular stimulation of antigen-specific T-lymphocytes, include Candida (albicans, krusei, glabrata, tropicalis, etc.), Cryptococcus neoformans, Aspergillus (fumigatus, niger, etc.), Genus Mucorales (mucor, absidia, rhizophus),



*Sporothrix schenckii*, *Blastomyces dermatitidis*,  
*Paracoccidioides brasiliensis*, *Coccidioides immitis* and  
*Histoplasma capsulatum*.

Some examples of pathogenic parasites causing  
5 infections ameliorated by immune stimulation, in particular  
stimulation of antigen-specific T-lymphocytes, include  
*Entamoeba histolytica*, *Balantidium coli*, *Naegleria fowleri*,  
*Acanthamoeba* sp., *Giardia lamblia*, *Cryptosporidium* sp.,  
*Pneumocystis carinii*, *Plasmodium vivax*, *Babesia microti*,  
10 *Trypanosoma brucei*, *Trypanosoma cruzi*, *Leishmania donovani*,  
*Toxoplasma gondi*, and *Nippostrongylus brasiliensis*.

If the condition ameliorated by immune stimulation,  
in particular stimulation of antigen-specific T-lymphocytes  
is immunization against an antigen, the anti-human CD27  
15 agonistic antibody and the number of immune checkpoint  
inhibitors are in general administered in combination with a  
vaccine. The vaccine may comprise a number of antigen or  
antigenic determinants specific for, a pathogen, such as a  
pathogen selected from bacteria, fungi, viruses or  
20 parasites, or of a toxin, or a self-antigen, including  
antigens expressed on benign or malignant tumors, such as  
cancers. The pathogens and cancers against which the  
vaccination is directed may be selected as indicated above.

According to certain embodiments the vaccination is  
25 directed against HIV, Hepatitis (A, B, C, D, E), Influenza,  
Herpes, *Giardia*, Malaria, *Leishmania*, *Staphylococcus aureus*,  
or *Pseudomonas Aeruginosa*, preferably HIV. For these  
pathogens there is currently no effective vaccine, or  
existing vaccines are less than completely effective.

30 The inventors of the present invention have found  
unexpected immune stimulatory effects when combining an  
anti-human CD27 agonistic antibody and a number of immune  
checkpoint protein inhibitors, in particular an inhibitor of

CTLA4 or PD1, PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 or KIR. Tests indicate synergistic and/or potentiating effects of the combinations. Thus according to certain embodiments the anti-human CD27 agonistic antibody is for synergistic and/or potentiating stimulation of the immune response, particularly synergistic and/or potentiating stimulation of antigen-specific T-lymphocytes, together with the number of immune checkpoint inhibitors.

The anti-human CD27 agonistic antibody of the present invention preferably is presented in a composition comprising the antibody. The composition comprises the antibody or antibodies together with a carrier. The composition according to certain embodiments preferably is a pharmaceutical composition.

To prepare pharmaceutical or sterile compositions, the antibody, antibodies or fragment thereof, is admixed with a pharmaceutically acceptable carrier and/or excipient, see, e.g., Remington's Pharmaceutical Sciences and U.S. Pharmacopeia: National Formulary, Mack Publishing Company, Easton, PA (1984). Formulations of therapeutic and diagnostic agents may be prepared by mixing with physiologically acceptable carriers, excipients, or stabilizers in the form of, e.g., lyophilized powders, slurries, aqueous solutions or suspensions (see, e.g., Hardman, et al., 2001, *Goodman and Gilman's The Pharmacological Basis of Therapeutics*, McGraw-Hill, New York, NY; Gennaro, 2000, Remington: *The Science and Practice of Pharmacy*, Lippincott, Williams, and Wilkins, New York, NY; Avis, et al. (eds.), 1993, *Pharmaceutical Dosage Forms: Parenteral Medications*, Marcel Dekker, NY; Lieberman, et al. (eds.), 1990, *Pharmaceutical Dosage Forms: Tablets*, Marcel Dekker, NY; Lieberman, et al. (eds.), 1990, *Pharmaceutical Dosage Forms: Disperse Systems*, Marcel Dekker, NY; Weiner

and Kotkoskie, 2000, *Excipient Toxicity and Safety*, Marcel Dekker, Inc., New York, NY).

Toxicity and therapeutic efficacy of the binding compound, in particular antibody, compositions, administered alone or in combination with another agent, such as the usual anti-cancer drugs, can be determined by standard pharmaceutical procedures in cell cultures or experimental animals, e.g., for determining the LD<sub>50</sub> (the dose lethal to 50% of the population) and the ED<sub>50</sub> (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio between LD<sub>50</sub> and ED<sub>50</sub>. The data obtained from these cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED<sub>50</sub> with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized.

Suitable routes of administration include parenteral administration, such as intramuscular, intravenous, or subcutaneous administration and oral administration. Administration of antibodies, used in the pharmaceutical composition or to practice the method of the present invention can be carried out in a variety of conventional ways, such as oral ingestion, inhalation, topical application or cutaneous, subcutaneous, intraperitoneal, parenteral, intraarterial or intravenous injection. In one embodiment, the binding compound of the invention is administered intravenously. In another embodiment, the binding compound of the invention is administered subcutaneously.

Alternatively, one may administer the antibodies in a local rather than systemic manner, for example, via injection of the antibody or antibodies directly into the site of action, often in a depot or sustained release formulation. Furthermore, one may administer the antibody in a targeted drug delivery system.

Guidance in selecting appropriate doses of antibodies, cytokines, and small molecules are available (see, e.g., Wawrzynczak, 1996, *Antibody Therapy*, Bios Scientific Pub. Ltd, Oxfordshire, UK; Kresina (ed.), 1991, *Monoclonal Antibodies, Cytokines and Arthritis*, Marcel Dekker, New York, NY; Bach (ed.), 1993, *Monoclonal Antibodies and Peptide Therapy in Autoimmune Diseases*, Marcel Dekker, New York, NY; Baert, et al., 2003, *New Engl. J. Med.* 348:601-608; Milgrom, et al., 1999, *New Engl. J. Med.* 341:1966-1973; Slamon, et al., 2001, *New Engl. J. Med.* 344:783-792; Beniaminovitz, et al., 2000, *New Engl. J. Med.* 342:613-619; Ghosh, et al., 2003, *New Engl. J. Med.* 348:24-32; Lipsky, et al., 2000, *New Engl. J. Med.* 343:1594-1602).

Determination of the appropriate dose is made by the clinician, e.g., using parameters or factors known or suspected in the art to affect treatment or predicted to affect treatment. Generally, the dose begins with an amount somewhat less than the optimum dose and it is increased by small increments thereafter until the desired or optimum effect is achieved relative to any negative side effects. Important diagnostic measures include those of symptoms of, e.g., the inflammation or level of inflammatory cytokines produced.

A preferred dose protocol is one involving the maximal dose or dose frequency that avoids significant undesirable side effects. A total weekly dose is generally at least 0.05 µg/kg body weight, more generally at least 0.2

µg/kg, most generally at least 0.5 µg/kg, typically at least 1 µg/kg, more typically at least 10 µg/kg, most typically at least 100 µg/kg, preferably at least 0.2 mg/kg, more preferably at least 1.0 mg/kg, most preferably at least 2.0 mg/kg, optimally at least 10 mg/kg, more optimally at least 25 mg/kg, and most optimally at least 50 mg/kg (see, e.g., Yang, et al., 2003, *New Engl. J. Med.* 349:427-434; Herold, et al., 2002, *New Engl. J. Med.* 346:1692-1698; Liu, et al., 1999, *J. Neurol. Neurosurg. Psych.* 67:451-456; Portielji, et al., 2003, *Cancer Immunol. Immunother.* 52:133-144). The desired dose of a small molecule therapeutic, e.g., a peptide mimetic, natural product, or organic chemical, is about the same as for an antibody or polypeptide, on a moles/kg basis.

"Administration", "therapy" and "treatment," as it applies to an animal, human, experimental subject, cell, tissue, organ, or biological fluid, refers to contact of an exogenous pharmaceutical, therapeutic, diagnostic agent, or composition to the animal, human, subject, cell, tissue, organ, or biological fluid. "Administration", "therapy" and "treatment" can refer, e.g., to therapeutic, pharmacokinetic, diagnostic, research, and experimental methods. Treatment of a cell encompasses contact of a reagent to the cell, as well as contact of a reagent to a fluid, where the fluid is in contact with the cell. "Administration", "therapy" and "treatment" also mean *in vitro* and *ex vivo* treatments, e.g., of a cell, by a reagent, diagnostic, binding composition, or by another cell.

As used herein, "inhibit" or "treat" or "treatment" includes a postponement of development of the symptoms associated with disease and/or a reduction in the severity of such symptoms that will or are expected to develop with said disease. The terms further include

ameliorating existing symptoms, preventing additional symptoms, and ameliorating or preventing the underlying causes of such symptoms. Thus, the terms denote that a beneficial result has been conferred on a vertebrate subject with a disease.

As used herein, the term "therapeutically effective amount" or "effective amount" refers to an amount of antibody or antibodies, that when administered alone or in combination with an additional therapeutic agent to a cell, tissue, or subject is effective to prevent or ameliorate the disease or condition to be treated. A therapeutically effective dose further refers to that amount of the compound sufficient to result in amelioration of symptoms, e.g., treatment, healing, prevention or amelioration of the relevant medical condition, or an increase in rate of treatment, healing, prevention or amelioration of such conditions. When applied to an individual active ingredient administered alone, a therapeutically effective dose refers to that ingredient alone. When applied to a combination, a therapeutically effective dose refers to combined amounts of the active ingredients that result in the therapeutic effect, whether administered in combination, serially or simultaneously. An effective amount of therapeutic will decrease the symptoms typically by at least 10%; usually by at least 20%; preferably at least about 30%; more preferably at least 40%, and most preferably by at least 50%.

Methods for co-administration or treatment with a second therapeutic agent are well known in the art, see, e.g., Hardman, et al. (eds.), 2001, *Goodman and Gilman's The Pharmacological Basis of Therapeutics*, 10th ed., McGraw-Hill, New York, NY; Poole and Peterson (eds.), 2001, *Pharmacotherapeutics for Advanced Practice: A Practical*

*Approach*, Lippincott, Williams & Wilkins, Phila., PA;  
Chabner and Longo (eds.), 2001, *Cancer Chemotherapy and  
Biotherapy*, Lippincott, Williams & Wilkins, Phila., PA.

Within the present invention the term 'co-  
5 administered' should be understood as meaning that the  
individual active components (here the anti-human CD27  
agonistic antibody and the number of immune checkpoint  
inhibitors) are administered in the same subject. Such  
administration may be simultaneously or alternatively the  
10 active components may be administered within a time frame of  
up to 3 months, in view of the fact that most antibody  
therapeutics, including those targeting immune checkpoint  
inhibitors and CD27 display a terminal half-life in human  
subjects of 2-4 weeks. Therefore, pharmacodynamic responses  
15 to such antibody may be detectable for months after  
administration.

The pharmaceutical composition of the invention  
may also contain other agents, including but not limited to  
a cytotoxic, chemotherapeutic, cytostatic, anti-angiogenic  
20 or antimetabolite agents, a tumor targeted agent, an immune  
stimulating or immune modulating agent or an antibody  
conjugated to a cytotoxic, cytostatic, or otherwise toxic  
agent. The pharmaceutical composition can also be employed  
with other therapeutic modalities such as surgery,  
25 chemotherapy and radiation.

According to a preferred embodiment the anti-human  
CD27 agonistic antibody is provided, in a pharmaceutical  
composition comprising the anti-human CD27 agonistic  
antibody together with a pharmaceutically acceptable  
30 carrier, said pharmaceutical composition being packed in a  
container and said container being associated with an  
information carrier, wherein said information carrier  
comprises information indicating the combined use of the

anti-human CD27 agonistic antibody, preferably in the pharmaceutical composition, together with an immune checkpoint inhibitor for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular stimulation of antigen-specific T-lymphocytes. The container may be any container suitable for holding a pharmaceutical composition, preferably a sterile pharmaceutical composition, more preferably an injectable pharmaceutical composition. According to a preferred embodiment the container comprises a unit dosage of the anti-human CD27 agonistic antibody.

The information carrier may be any suitable information carrier such as an object having a surface suitable for carrying information, for example paper or cardboard. Alternatively a data carrier comprising machine readable information may be used. The information may be presented in visual form such as in the form of written information or in the form of a number of pictures. Alternatively the information may be presented in a form "readable" by touch, such as in braille. In case the information is on a data carrier comprising machine readable information, the information may be stored as machine code or analogous signals which can be converted to visual and/or audio information.

The information carrier and the container comprising the pharmaceutical composition are associated such that they can be presented together in a single product. For example they may be associated by being enclosed together in an enclosure, such as a container, including a box.

A further aspect of the invention relates to an immune checkpoint inhibitor for use in the treatment of a condition ameliorated by stimulation of an immune response,



in particular the treatment of a condition that benefits from the stimulation of antigen-specific T-lymphocytes, wherein in said treatment an anti-human CD27 agonistic antibody is administered. This immune checkpoint inhibitor together with the anti-human CD27 agonistic antibody forms a therapeutic combination resulting in unexpected immune stimulation. According to certain embodiments the immune checkpoint inhibitor is for synergistic and/or potentiating stimulation of the immune response, particularly synergistic and/or potentiating stimulation of antigen-specific T-lymphocytes, together with the anti-human CD27 agonistic antibody. The technical details of the various features and preferred embodiments of the immune checkpoint inhibitor of the invention is similar to what has already been discussed in connection to the anti-human CD27 agonistic antibody of the invention.

Similar to the anti-human CD27 agonistic antibody, the immune checkpoint inhibitor of the invention may be provided, in a pharmaceutical composition comprising the immune check point inhibitor together with a pharmaceutically acceptable carrier, said pharmaceutical composition being packed in a container and said container being associated with an information carrier, wherein said information carrier comprises information indicating the combined use of the immune checkpoint inhibitor, preferably in the pharmaceutical composition, together with an anti-human CD27 agonistic antibody for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of a condition that benefits from stimulation of antigen-specific T-lymphocytes. The container may be any container suitable for holding a pharmaceutical composition, preferably a sterile pharmaceutical composition more preferably an injectable

pharmaceutical. According to a preferred embodiment the container comprises a unit dosage of the anti-human CD27 agonistic antibody. Also for this embodiment of the invention the details of the various technical features and preferred embodiments will be clear from the parts of the description relating to the anti-human CD27 agonistic antibody.

Yet a further aspect of the invention relates to a combination of an anti-human CD27 agonistic antibody together with a number of immune checkpoint inhibitors for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of a condition that benefits from the stimulation of antigen-specific T-lymphocytes. This combination according to the invention is primarily a functional combination and any physical combination considered suitable by the skilled person for making this functional combination is to be considered within the scope of this invention. For example in the combination the anti-human CD27 agonistic antibody may be co-administered with the number of immune checkpoint inhibitors to a subject. Alternatively, the combination may be a kit of parts comprising:

(i) a first container holding a first pharmaceutical composition comprising the anti-human CD27 agonistic antibody together with a pharmaceutically acceptable carrier;

(ii) a second container holding a second pharmaceutical composition comprising the immune checkpoint inhibitor together with a pharmaceutically acceptable carrier

(iii) optionally an information carrier comprising information indicating the combined use of the anti-human CD27 agonistic antibody, preferably in the first

pharmaceutical composition, together with an immune checkpoint inhibitor, preferably in the second pharmaceutical composition, for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of conditions that benefit from the stimulation or enhancement of antigen-specific T-lymphocytes.

According to an alternative embodiment the first and second container and the first and second pharmaceutical composition coincide thus presenting a kit of parts comprising:

(A) a container holding a pharmaceutical composition comprising the anti-human CD27 agonistic antibody and the immune checkpoint inhibitor together with a pharmaceutically acceptable carrier;

(B) optionally an information carrier comprising information indicating the combined use of the anti-human CD27 agonistic antibody, preferably in the pharmaceutical composition, together with an immune checkpoint inhibitor, preferably in the pharmaceutical composition, for use in the treatment of a condition ameliorated by stimulation of an immune response, in particular the treatment of a condition that benefits from the stimulation of antigen-specific T-lymphocytes.

Also for the combination of the invention the details of the various technical features and preferred embodiments is similar to what has been discussed in the parts of the description relating to the anti-human CD27 agonistic antibody.

A further aspect of the invention relates to a method for treating a condition ameliorated by stimulation of an immune response, in particular the treatments of a condition that result in the stimulation of antigen-specific

T-lymphocytes, said method comprising administering a CD27 agonist, preferably an anti-human CD27 agonistic antibody, in combination with a number of immune checkpoint inhibitors.

5           Also for the method of treatment of the invention the details of the various technical features and preferred embodiments are similar to what has been discussed above in relation to the anti-human CD27 agonistic antibody.

          The invention and its effects will now be further  
10 illustrated with reference to the following non-limiting examples.

### EXAMPLES

#### **EXAMPLE 1**

15   *CD27 agonism combined with checkpoint protein blockade results in unexpected immune stimulation*

          To study the effect of combining CD27 agonists with PD-1 or PD-L1 blocking antibodies human Peripheral Blood Mononuclear cells (PBMCs) were isolated from buffy  
20 coat. First, the Buffy coat was diluted to a total volume of 300 ml with DMEM/F12 medium (Gibco, 11320) supplemented with heparin solution (Leo Pharma, DB6132, 5000 U/ml) at RT. After mixing the cell suspension, aliquotes were loaded on a Ficoll-Paque Plus gradient in conical tubes and centrifuged  
25 at 450 g for 30 min, at 20°C without a brake. Next, plasma was removed by aspiration and PBMCs were recovered from the plasma/Ficoll interface. PBMCs were washed twice with DMEM/F12 medium and resuspended in RPMI 1640 medium (Gibco, 52400) supplemented with 10% Foetal Calf Serum. PBMCs were  
30 plated at  $2 \times 10^5$  cells/well in 96-well flat bottom plates (Nuclon). Antibodies (anti-PD1 (hIgG4 chimera, see below, of hPD109A, WO/2008/156712), anti-hCD27 (hIgG4 chimera of hCD27.15). See below for chimerization), anti-PDL1 (MIH1,

eBioscience 16.5983.82), human IgG4 isotype control (Sigma, 14639) and mouse IgG1 isotype control (eBioscience, 16.4714.85)) and dilutions thereof were diluted in PBS and added to the PBMCs. Human IgG4 chimeric versions of hPD1.09A and hCD27.15 were constructed by cloning of the VH and VL genes upstream of the human IgG4 and human Kappa constant domain encoding cDNA, respectively. Full cDNAs were subcloned in pcDNA3.1 (Invitrogen) and transiently transfected in 293T/17 cells (ATCC) using Lipofectamine 2000 (Invitrogen) following the manufacturer's instructions. After 5-7 days, supernatants were harvested and antibodies were purified using standard protein A purification. Finally, Staphylococcus Enterotoxin B (Sigma S4881) diluted in RPMI 1640 medium supplemented with 10% Foetal Calf Serum was added to the wells in a final concentration of 25 ng/ml. Cells were incubated for two days at 37°C, 5% CO<sub>2</sub> and 95% humidity. To assess the level of immune activation, IL-2 secretion levels were determined in the supernatant in accordance with the method described in Dulos *et al.*, *J. Immunother*, 2012, 35:169-78. To that aim, supernatants were aspirated and cleared from any cell material by centrifugation. Next, supernatants were added to Nunc maxisorp ELISA plates that had been coated with 2 µg/ml anti-hIL-2 antibody (BD Pharmingen, 555051) in PBS by overnight incubation at 4°C. Prior to addition of the supernatants, wells were emptied and blocked with 200 µl/well PBS/1%BSA for one hour at Room Temperature (RT). Supernatants were incubated for one hour at RT, washed three times with PBST (PBS with 0.01% Tween 20). Subsequently, 100 µl of 0.5 µg/ml of anti-hIL2-biotin (BD Pharmingen 555040) was added in PBS/PBS-1%BSA (1:1) and incubated for one hour at RT. After three washes with PBST, 1:5000 diluted streptavidin-HRP (BD Pharmingen, 554066) was added in 100 µl

PBS/PBS-1% BSA (1:1). After three washes with PBST and one wash with water, IL-2 was detected by addition of 100  $\mu$ l/well TMB stabilized chromogen (Invitrogen, SB02).

Reactions were stopped with 100  $\mu$ l 0.5 M H<sub>2</sub>SO<sub>4</sub> and

5 absorbances were read at 450 and 620 nm. In this assay, recombinant human IL-2 (Sigma, H7041) was used to quantify IL-2 protein levels in the supernatants. In **Figure 1A** the cooperative action between CD27 agonistic antibody and PD-1 blocking antibody and in **Figure 1B** the cooperative action  
10 between CD27 agonistic antibody and PD-L1 blocking antibody is shown.

To study the effect of combining CD27 agonist with PD-1 and PD-L1 blocking antibody in whole human blood, blood was diluted 10 times in RPMI 1640 medium (Gibco, 52400)  
15 supplemented with 10% Foetal Bovine Serum (Hyclone). Diluted blood was plated in 96-well Nunclon delta surface flat bottom plates (100  $\mu$ l/well). Antibodies anti-hCD27 (hIgG4 chimera of hCD27.15, see above), anti-PD-1 (hIgG4 chimera of hPD1.09A, see above), anti-PDL1 (MIH1, eBioscience  
20 16.5983.82), human IgG4 isotype control (Sigma, 14639) and mouse IgG1 isotype control (eBioscience, 16.4714.85) and dilutions thereof were diluted in PBS and added to the diluted blood. Finally, Staphylococcus Enterotoxin B (Sigma S4881) diluted in RPMI 1640 medium supplemented with 10%  
25 Foetal Calf Serum was added to the wells in a final concentration of 25 ng/ml. Cells were incubated for two days at 37°C, 5% CO<sub>2</sub> and 95% humidity. To assess the level of immune activation, IL-2 secretion levels were determined in the supernatant. To that aim, supernatants were aspirated  
30 and cleared from any cell material by centrifugation. Next, supernatants were added to Nunc maxisorp ELISA plates that had been coated with 2  $\mu$ g/ml anti-hIL-2 antibody (BD Pharmingen, 555051) in PBS by overnight incubation at 4°C.

Prior to addition of the supernatant, wells were emptied and blocked with 200 µl/well PBS/1%BSA for one hour at Room Temperature (RT). Supernatants were incubated for one hour at RT, washed three times with PBST (PBS with 0.01% Tween 20). Subsequently, 100 µl of 0.5 µg/ml of anti-hIL2-biotin (BD Pharmingen 555040) was added in PBS/PBS-1%BSA (1:1) and incubated for one hour at RT. After three washes with PBST, 1:5000 diluted streptavidin-HRP (BD Pharmingen, 554066) was added in 100 µl PBS/PBS-1% BSA (1:1). After three washes with PBST and one wash with water, IL-2 was detected by addition of 100 µl/well TMB stabilized chromogen (Invitrogen, SB02). Reactions were stopped with 100 µl 0.5 M H<sub>2</sub>SO<sub>4</sub> and absorbances were read at 450 and 620 nm. In this assay, recombinant human IL-2 (Sigma, H7041) was used to quantify IL-2 protein levels in the supernatants. Also in this test the IL-2 level was increased to an unexpected level by the combination of the CD27 agonistic antibody and the PD-1 or PDL-1 inhibiting antibody in comparison to these antibodies alone.

The expectation presented by these results, that combinations of a CD27 agonistic antibody with other immune check point inhibitors will give similar effects, was confirmed in additional experiments performed with the same whole blood test protocol, which included the combination of a CD27 agonist (hIgG4 chimera of hCD27.15, see above) with anti-CTLA4 (14D3, eBioscience 16.1529.82). The results of these experiments (see **Figures 2A-2C**) showed a similar cooperative action by the combination of the CD27 agonistic antibody and the CTLA4 inhibiting antibody in comparison to the individual antibodies alone (IgG4 isotype control as above; IgG2A isotype control: BD Pharmingen 554126).

On the basis of these results it may be expected that the combination of an anti-human CD27 agonistic

antibody together with at least one immune checkpoint inhibitor will have beneficial effects in conditions ameliorated by stimulation of an immune response, such as conditions ameliorated by stimulation of antigen-specific T-lymphocytes.

## Example 2

*Evaluating the activity of anti-CD27, anti-LAG-3, and anti-PD1 antibodies in a human PBMC SEB superantigen-induced IL2 secretion*

An additional confirmatory experiment was conducted to further show the effects of the combination of anti-human CD27 agonistic antibodies with immune checkpoint protein inhibitors. In this experiment a hCD27.15 analogue was used in combination with an anti-LAG3 antibody and an anti-PD1 antibody. The analogue used contains functional heavy chain CDR1, CDR2, CDR3 (SEQ ID NO: 1, 2, 3, respectively) and light chain CDR1, CDR2, CDR3 (SEQ ID NO: 4, 5, 6, respectively) of hCD27.15. In addition it contains functionally linked functional human IgG4 constant domains (CH1 - CH3, GenBank accession #K01316). Functionality of the hCD27.15 analogue was established in a CD27 binding assay corresponding to the one disclosed in example 2 (page 49-50) of WO2012/004367 and a CD27 signaling test corresponding to the NF- $\kappa$ B tests disclosed in example 3 (page 54-55) of WO2012/004367.

The human anti-LAG3 antibody used comprised a heavy chain amino acid sequence according to SEQ ID NO: 23 and a light chain amino acid sequence according to SEQ ID NO: 24. The human anti-PD1 antibody used comprised a heavy chain amino acid sequence according to SEQ ID NO: 21 and a light chain amino acid sequence according to SEQ ID NO: 22. To study the effect of the combination of anti-human CD27 agonistic antibodies with



immune checkpoint protein inhibitors human PBMCs were isolated from heparinized human blood from healthy donors using SepMate® tubes (Stem Cell). 15 ml of Ficoll-1077 was added to SepMate® tubes, which was carefully overlaid with 25 ml of human blood and centrifuged at 1,250 g for 12 min. at RT with light brake (5 out of 10 - 10 being max brake). White blood cells were isolated at the interphase of the Ficoll and dilute into 40 ml of Hanks Balances Salt Solution (HBSS) at RT. Next, cells were centrifuged at 300 g for 10 min. at 4 °C and the pellet was resuspended in 50 ml of HBSS. Finally, the cell suspension was centrifuged at 250 g for 10 min to remove platelets and the pellet was resuspended in 12 ml complete media (RPMI 1640 containing HEPES and Penn/Step and 10% human A+B+ serum)). Cells were quantified by Vi-cell. Next, 50 µl of PBMC cell suspension ( $1 \times 10^7$  cells/ml (final  $5 \times 10^5$  cells/well)) were plated in a 96-well round bottom plate. 50 µl of antibody (per antibody treatment, so 100 µl total antibody added for combination conditions) at 10 µg/ml end concentrations for 30 min. at 37°C. Next, a 4x concentration of 50 µl *Staphylococcus* enterotoxin B (SEB) superantigen (Toxin Technology, Sarasota, FL) at 0.5, 1, 8, 16, 32, 64 and 80 ng/ml end concentration was added and incubate 72 hr at 37°C. Supernatants were collected and cleared from any cell material by centrifugation. IL-2 secretion levels as a measure for immune activation was detected using Human IL-2 V-PLEX Kit (Catalog No. K151QQD-4, from Meso Scale Discovery, Rockville, MD), according to the manufacturer's instructions. As a negative control, (sub)-isotype matched antibodies were used.

**Figures 3A-3D** show the results obtained from four separate donors using the experimental methods described above. For each donor the results of the anti-CD27/Anti-LAG3 combination is shown in the left panel and the results of

the anti-CD27/Anti-PD1 combination is shown in the right panel. For each SEB concentration the bars represent from left to right: isotype control; immune checkpoint inhibitor (anti-Lag3 or anti-PD1); anti-CD27; immune checkpoint inhibitor (anti-Lag3 or anti-PD1) + anti-CD27. The anti-CD27 antibody demonstrates approximately 1.5-2 fold IL-2 increase above isotype controls as a single agent. The anti-CD27 antibody showed combination activity with anti-LAG-3 antibody and with anti-PD1 antibody. Similar effects may be expected for combinations of anti-human CD27 agonistic antibody (including analogues) with other immune checkpoint protein inhibitors.

CLAIMS

1. An anti-human CD27 agonistic antibody, such as hCD27.15  
5 or 1F5, or an antibody analogue thereof, for use in the  
treatment of a condition ameliorated by stimulation of  
an immune response, in particular stimulation of  
antigen-specific T-lymphocytes, wherein in said  
treatment a number of immune checkpoint protein  
10 inhibitors is administered.
2. An anti-human CD27 agonistic antibody according to  
claim 1, wherein an immune checkpoint protein inhibitor  
is selected from an inhibitor of CTLA-4, PD1, PD-L1,  
PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 or KIR.
- 15 3. An anti-human CD27 agonistic antibody according to any  
of the claims 1-2, wherein the condition ameliorated by  
immune stimulation, in particular stimulation of  
antigen-specific T-lymphocytes is selected from  
infectious diseases, such as bacterial, fungal, viral  
20 and parasitic infectious diseases, immunization against  
a pathogen, such as a pathogen selected from bacteria,  
fungi, viruses or parasites, or vaccination against  
toxins, or self-antigens, including antigens expressed  
on benign or malignant tumors, such as cancers, or  
25 conditions associated with uncontrolled proliferation  
of cells such as cancers.
4. An anti-human CD27 agonistic antibody according to any  
of the claims 1-3, wherein the treatment is vaccination  
and a vaccine is administered in the treatment.
- 30 5. Immune checkpoint inhibitor for use in the treatment of  
a condition ameliorated by stimulation of an immune  
response, in particular stimulation of antigen-specific  
T-lymphocytes, wherein in said treatment an anti-human

CD27 agonistic antibody, such as hCD27.15 or 1F5, or an antibody analogue thereof, is administered.

6. Immune checkpoint inhibitor according to claim 5,

wherein the immune checkpoint inhibitor is selected

5 from an inhibitor of CTLA-4, PD1, PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 or KIR.

7. Immune checkpoint inhibitor according to any of the claims 5-6, wherein the condition ameliorated by immune stimulation is selected from infectious diseases, such

10 as bacterial, fungal, viral and parasitic infectious diseases, immunization against a pathogen, such as a pathogen selected from bacteria, fungi, viruses or parasites, or vaccination against toxins, or self-

15 antigens, including antigens expressed on benign or malignant tumors, such as cancers, or conditions associated with uncontrolled proliferation of cells such as cancers.

8. An immune checkpoint inhibitor according to any of the claims 5-7, wherein the treatment is vaccination and

20 wherein a vaccine is administered in the treatment.

9. Combination of an anti-human CD27 agonistic antibody, such as hCD27.15 or 1F5, or an antibody analogue

thereof, together with a number of immune checkpoint inhibitors for use in the treatment of a condition

25 ameliorated by stimulation of an immune response, particularly stimulation of antigen-specific T-lymphocytes.

10. Combination according to claim 9, wherein an immune checkpoint inhibitor is selected from an inhibitor of

30 CTLA-4, PD1, PD-L1, PD-L2, LAG-3, BTLA, B7H3, B7H4, TIM3 or KIR.

11. Combination according to any of the claims 9-10,

wherein the condition ameliorated by immune stimulation

is selected from infectious diseases, such as bacterial, fungal, viral and parasitic infectious diseases, immunization against a pathogen, such as a pathogen selected from bacteria, fungi, viruses or parasites, or vaccination against toxins, or self-antigens, including antigens expressed on benign or malignant tumors, such as cancers, or conditions associated with uncontrolled proliferation of cells such as cancers.

12. Combination according to any of the claims 9-11, wherein the treatment is vaccination and wherein a vaccine is administered in the treatment.
13. A method of treating a condition ameliorated by stimulation of an immune response, comprising administering to a subject in need thereof a therapeutically effective amount of an anti-human CD27 agonistic antibody and further an immune checkpoint protein inhibitor.
14. The method of claim 13, wherein said anti-human CD27 agonistic antibody is selected from the group consisting of: an anti-human CD27 agonistic antibody comprising the CDR amino acid sequences of SEQ ID NO: 1, 2, 3, 4, 5, 6, or a variant sequence; a humanized analogue of antibody hCD27.15; an analogue of antibody hCD27.15 that binds to the same epitope as hCD27.15; antibody 1F5; an anti-human CD27 agonistic antibody that does not require cross-linking.
15. The method of any of the claims 13-14, wherein said further immune checkpoint inhibitor protein is selected from the group consisting of: an CTLA-4 antibody, an anti-PD1 antibody, an anti-PD-L1 antibody, an anti-PD-L2 antibody, an anti LAG-3 antibody, an anti-BTLA

antibody, an anti-B7H3 antibody, an anti-B7H4 antibody, an anti-TIM3 antibody and an anti-KIR antibody.

16. The method of any of the claims 13-15, wherein said further immune checkpoint inhibitor protein is an anti-PD1 antibody.

17. The method of any of the claims 13-16, wherein said anti-PD-1 antibody is pembrolizumab.

18. The method of any of the claims 13-17, wherein said anti-PD-1 antibody is nivolumab.

19. The method of any of the claims 13-18, wherein said further immune checkpoint inhibitor protein is an anti-LAG3 antibody.

20. The method of claim 19, wherein said anti-LAG3 antibody comprises the heavy chain and light chain amino acid sequences of SEQ ID NO: 23 and SEQ ID NO: 24, respectively.

21. The method of any of the claims 13-20, wherein the subject in need of treatment suffers from cancer.

22. The method of any of the claims 13-20, wherein the subject in need of treatment suffers from an infection (such as a bacterial, fungal, viral and parasitic infectious diseases).

23. A vaccine comprising an anti-human CD27 agonistic antibody and further comprising an immune checkpoint protein inhibitor.

24. The vaccine of claim 23, wherein said further immune checkpoint protein inhibitor is selected from the group consisting of: an CTLA-4 antibody, an anti-PD1 antibody, an anti-PD-L1 antibody, an anti-PD-L2 antibody, an anti LAG-3 antibody, an anti-BTLA antibody, an anti-B7H3 antibody, an anti-B7H4 antibody, an anti-TIM3 antibody and an anti-KIR antibody.

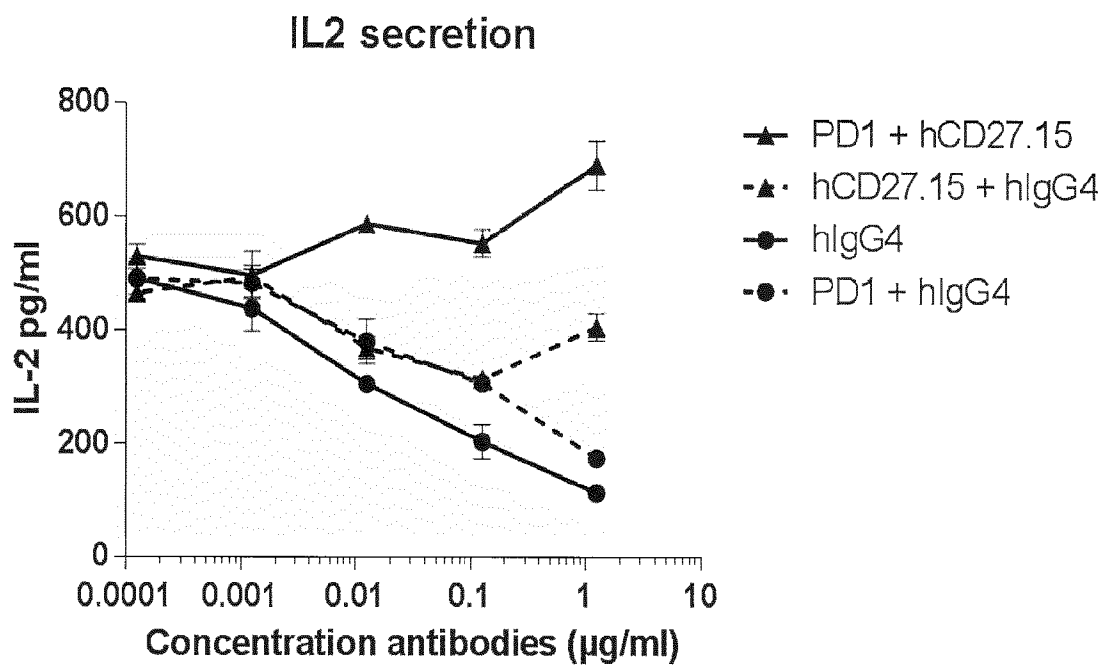
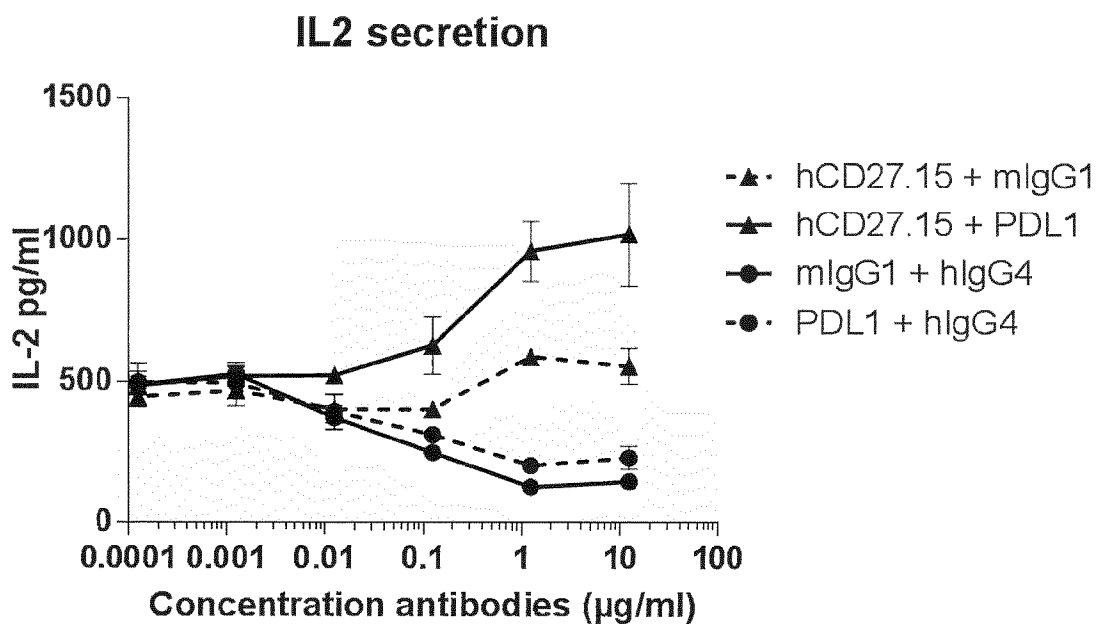
25.The vaccine of any of the claims 23-24, wherein said further immune checkpoint inhibitor protein is an anti-PD1 antibody.

26.The vaccine of any of the claims 23-25, wherein said anti-PD-1 antibody is pembrolizumab.

27.The vaccine of claims 23-26, wherein said anti-PD-1 antibody is nivolumab.

28.The vaccine of any of the claims 23-27, wherein said further immune checkpoint inhibitor protein is an anti-LAG3 antibody.

29.The vaccine of any of the claims 23-28, wherein said anti-LAG3 antibody comprises the heavy chain and light chain amino acid sequences of SEQ ID NO:23 and SEQ ID NO:24, respectively.

**FIG. 1A****FIG. 1B**



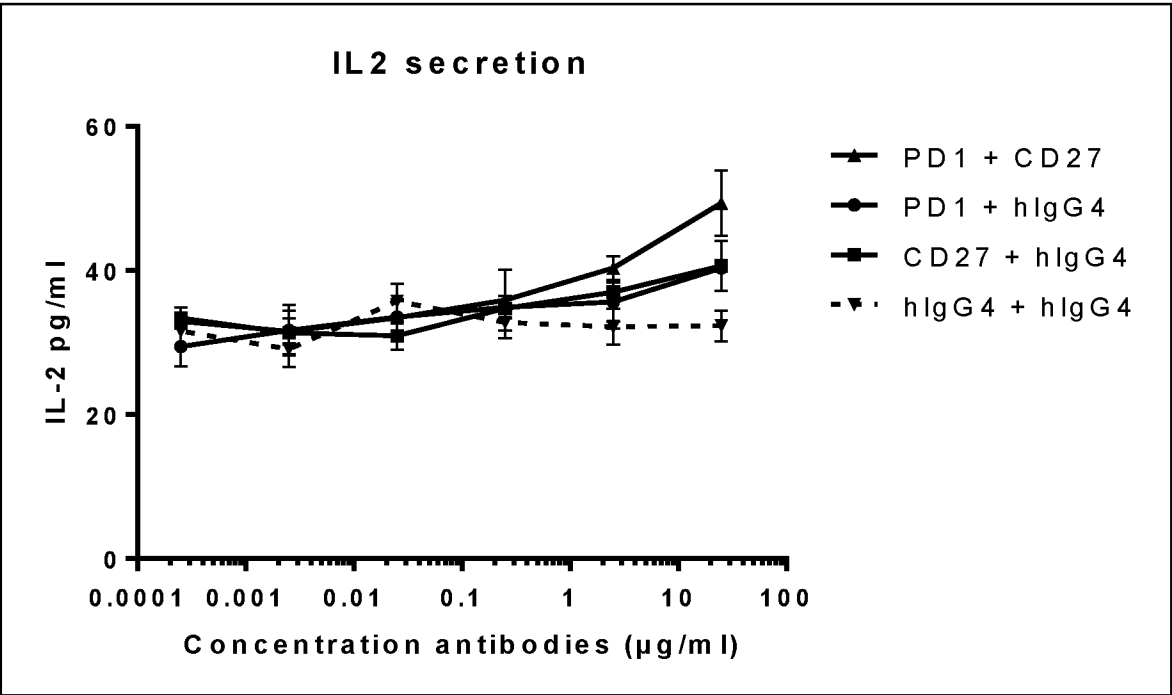


FIG. 2A

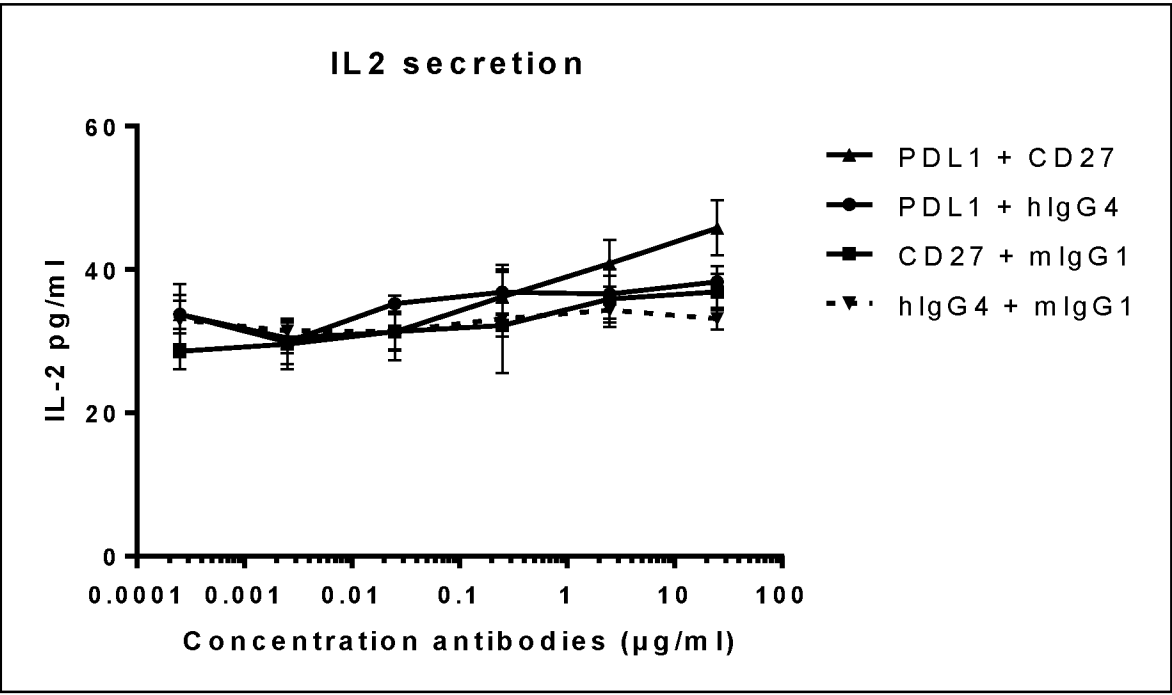


FIG. 2B

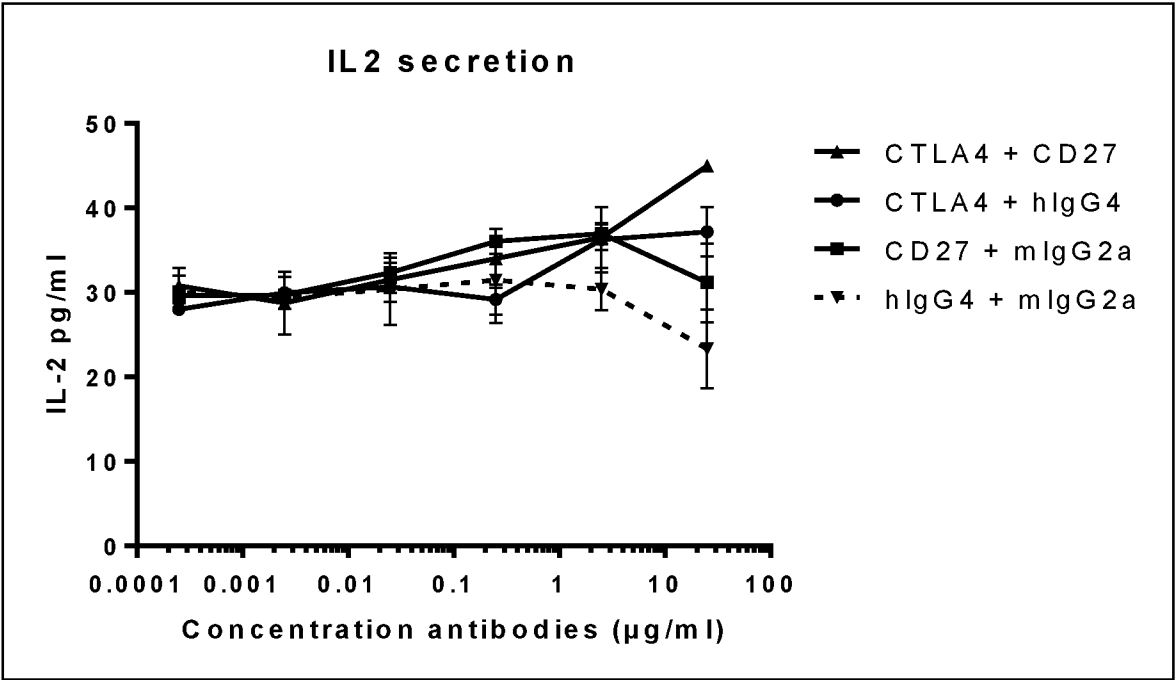


FIG. 2C

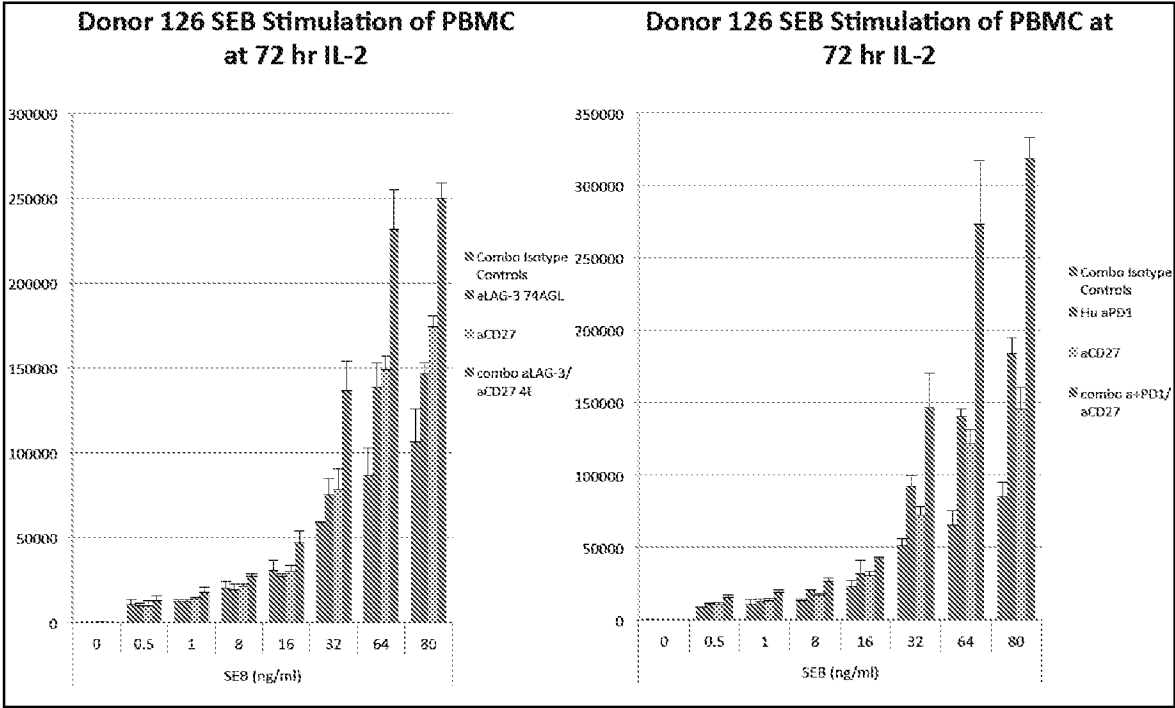


FIG. 3A

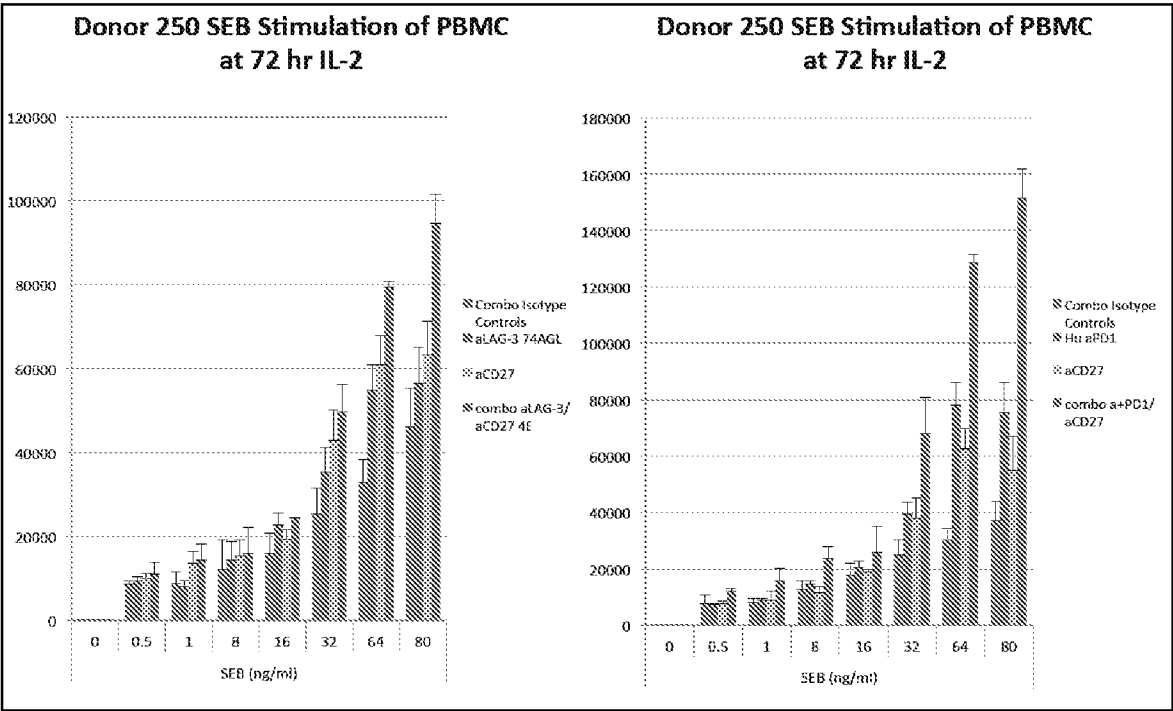


FIG. 3B

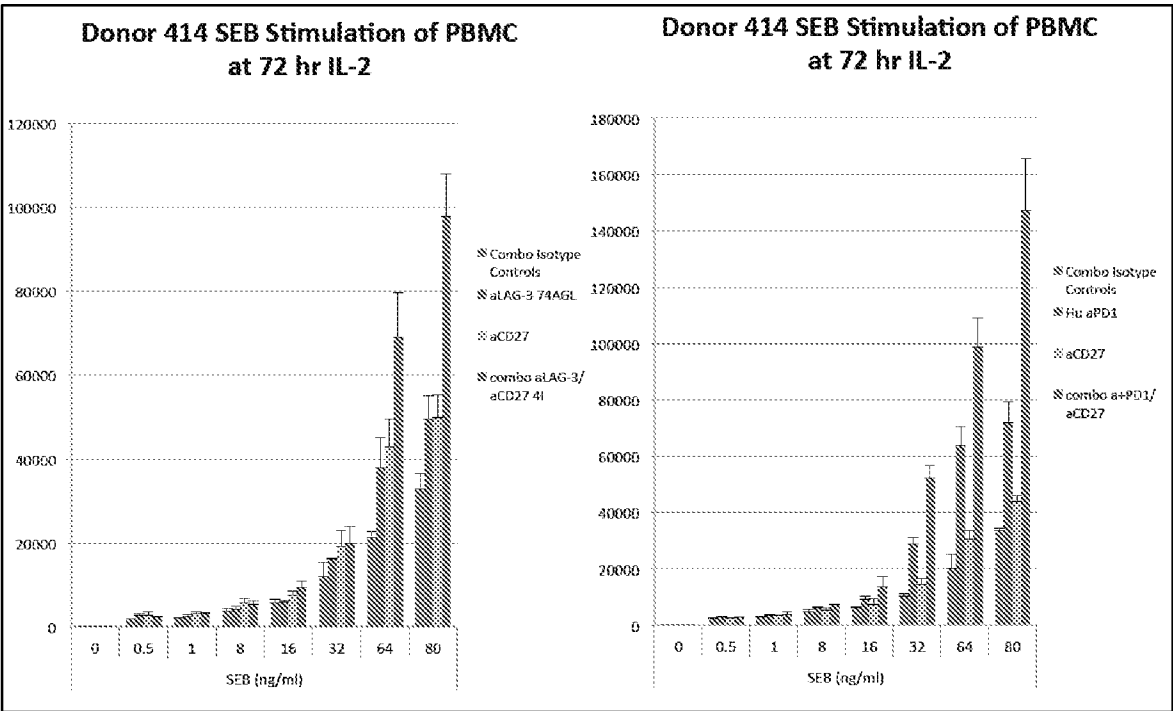


FIG. 3C

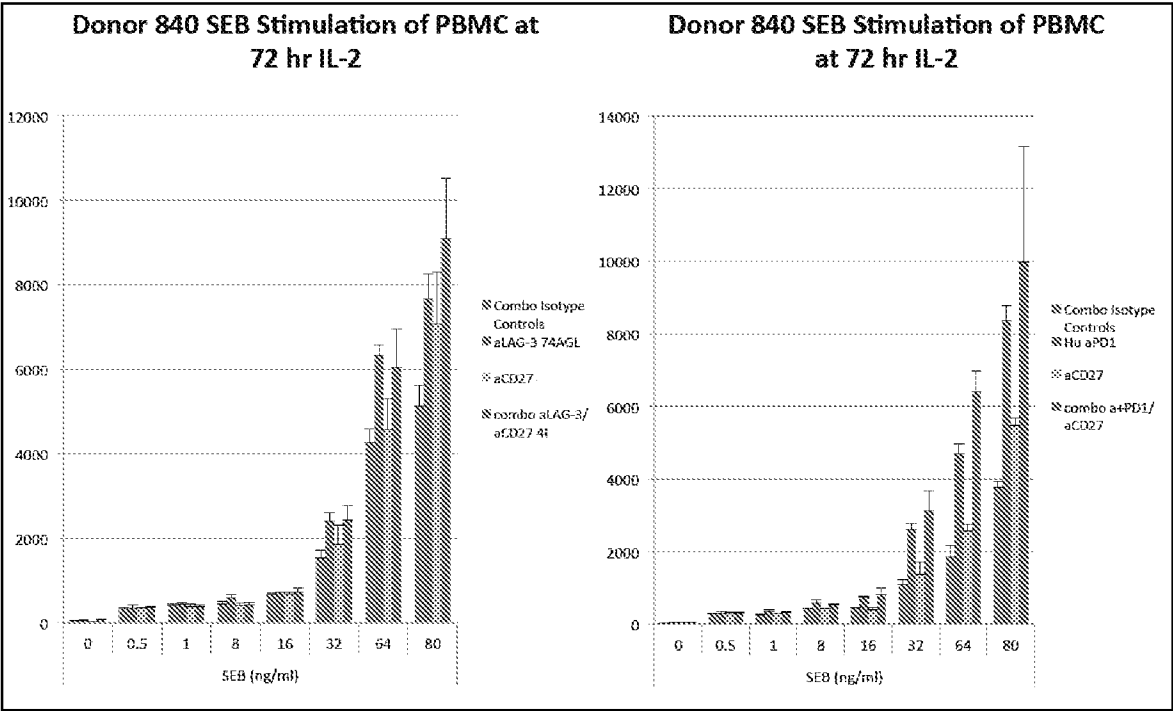


FIG. 3D

# INTERNATIONAL SEARCH REPORT

International application No  
PCT/NL2014/050543

**A. CLASSIFICATION OF SUBJECT MATTER**  
INV. A61K39/395 C07K16/28  
ADD. A61K39/00

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)  
A61K C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EP0-Internal, EMBASE, WPI Data

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	L. A. VITALE ET AL: "Development of a Human Monoclonal Antibody for Potential Therapy of CD27-Expressing Lymphoma and Leukemia", CLINICAL CANCER RESEARCH, vol. 18, no. 14, 15 May 2012 (2012-05-15), pages 3812-3821, XP055132599, ISSN: 1078-0432, DOI: 10.1158/1078-0432.CCR-11-3308 the whole document ----- -/--	1-29

☒ Further documents are listed in the continuation of Box C.

☐ See patent family annex.

\* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

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"&" document member of the same patent family

Date of the actual completion of the international search

3 December 2014

Date of mailing of the international search report

09/12/2014

Name and mailing address of the ISA/

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+31-70) 340-2040,  
Fax: (+31-70) 340-3016

Authorized officer

Lechner, Oskar

# INTERNATIONAL SEARCH REPORT

International application No

PCT/NL2014/050543

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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X	MICHAEL A. CURRAN ET AL: "Combination CTLA-4 Blockade and 4-1BB Activation Enhances Tumor Rejection by Increasing T-Cell Infiltration, Proliferation, and Cytokine Production", PLOS ONE, vol. 6, no. 4, 29 April 2011 (2011-04-29), page e19499, XP055136232, DOI: 10.1371/journal.pone.0019499 abstract -----	1-29
X	SIMEONE-E ET A.: "Immunomodulating antibodies in the treatment of metastatic melanoma: The experience with anti-CTLA-4, anti-CD137 and anti-PD1.", JOURNAL OF IMMUNOTOXICOLOGY, vol. 9, no. 3, 2012, XP009181173, abstract page 246, left-hand column, last paragraph -----	1-29
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International application No

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A	----- DREW M. PARDOLL: "The blockade of immune checkpoints in cancer immunotherapy", NATURE REVIEWS CANCER, vol. 12, no. 4, 22 March 2012 (2012-03-22) , pages 252-264, XP055150744, ISSN: 1474-175X, DOI: 10.1038/nrc3239 the whole document page 206, left-hand column, paragraph 1 page 206, right-hand column, paragraph 2 page 207, left-hand column, paragraph 1	1-29
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International application No

PCT/NL2014/050543

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	S.-R. WOO ET AL: "Immune Inhibitory Molecules LAG-3 and PD-1 Synergistically Regulate T-cell Function to Promote Tumoral Immune Escape", CANCER RESEARCH, vol. 72, no. 4, 20 December 2011 (2011-12-20), pages 917-927, XP055151722, ISSN: 0008-5472, DOI: 10.1158/0008-5472.CAN-11-1620 abstract	1-29
A	----- K. SAKUISHI ET AL: "Targeting Tim-3 and PD-1 pathways to reverse T cell exhaustion and restore anti-tumor immunity", NATURE REVIEWS IMMUNOLOGY, vol. 6, no. 10, 27 September 2010 (2010-09-27), pages 715-2194, XP055151727, ISSN: 1474-1733, DOI: 10.1038/nri1936 abstract	1-29
A	----- LAWRENCE J THOMAS ET AL: "Anti-tumor Activity of a Fully Human anti-CD27 Monoclonal Antibody in a Transgenic Mouse Model CD27 AS A TARGET FOR IMMUNOTHERAPY HUMAN ANTI-HUMAN CD27 ANTIBODIES", AMERICAN ASSOCIATION FOR CANCER RESEARCH 102ND ANNUAL MEETING 2011, 1 April 2011 (2011-04-01), XP055151917, abstract	1-29
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A	----- JULIET C. GRAY ET AL: "Optimising anti-tumour CD8 T-cell responses using combinations of immunomodulatory antibodies", EUROPEAN JOURNAL OF IMMUNOLOGY, vol. 38, no. 9, 1 September 2008 (2008-09-01), pages 2499-2511, XP055151985, ISSN: 0014-2980, DOI: 10.1002/eji.200838208 abstract	1-29
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International application No

PCT/NL2014/050543

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A,P	<p>Li He ET AL: "Combination therapies augment the anti-tumor activity of agonist anti-CD27 mAb in human CD27 transgenic mouse models IFNg + CD4 T cells",  10 November 2013 (2013-11-10),  XP055132852,  Retrieved from the Internet:  URL:http://www.celldex.com/docs/SITCPoster2013finalversion_Preclinical-Combination-Therapies.pdf  [retrieved on 2014-08-01]  the whole document</p>	1-29
A,P	<p>LI-ZHEN HE ET AL: "Combination therapies augment the anti-tumor activity of agonist CD27 mAb in human CD27 transgenic mouse models",  JOURNAL FOR IMMUNOTHERAPY OF CANCER,  BIOMED CENTRAL LTD, LONDON, UK,  vol. 1, no. Suppl 1,  7 November 2013 (2013-11-07), page P76,  XP021167289,  ISSN: 2051-1426, DOI:  10.1186/2051-1426-1-S1-P76  the whole document</p>	1-29
A,P	<p>LAWRENCE J THOMAS ET AL: "Targeting human CD27 with an agonist antibody stimulates T-cell activation and antitumor immunity",  ONCOIMMUNOLOGY,  vol. 3, no. 1, 1 January 2014 (2014-01-01),  page e27255, XP055132845,  ISSN: 2162-4011, DOI: 10.4161/onci.27255  the whole document</p>	1-29
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# INTERNATIONAL SEARCH REPORT

International application No

PCT/NL2014/050543

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A,P	ZHIQIANG GUO ET AL: "Combined TIM-3 blockade and CD137 activation affords the long-term protection in a murine model of ovarian cancer", JOURNAL OF TRANSLATIONAL MEDICINE, BIOMED CENTRAL, LONDON, GB, vol. 11, no. 1, 17 September 2013 (2013-09-17), page 215, XP021164386, ISSN: 1479-5876, DOI: 10.1186/1479-5876-11-215 the whole document	1-18
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A,P	----- LEI LU ET AL: "Combined PD-1 blockade and GITR triggering induce a potent antitumor immunity in murine cancer models and synergizes with chemotherapeutic drugs", JOURNAL OF TRANSLATIONAL MEDICINE, BIOMED CENTRAL, LONDON, GB, vol. 12, no. 1, 7 February 2014 (2014-02-07), page 36, XP021176018, ISSN: 1479-5876, DOI: 10.1186/1479-5876-12-36 the whole document -----	1-18