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(54) **HUMAN AND MAMMALIAN STEM CELL-DERIVED NEURON SURVIVAL FACTORS**

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Related U.S. Application Data

(63) Continuation-in-part of application No. 10/493,393, filed as application No. PCT/JP02/10936 on Oct. 22, 2002, now Pat. No. 7,273,725.

(30) **Foreign Application Priority Data**

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C12N 5/10 (2006.01)
C12N 15/63 (2006.01)
C07H 21/00 (2006.01)

(52) **U.S. Cl.** **435/320.1**; 435/325; 536/23.1; 536/23.5; 536/24.3

(58) **Field of Classification Search** None
See application file for complete search history.

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(57) **ABSTRACT**

The present invention relates to human, rat and mouse stem cell-derived neuron survival factor polypeptides (SDNSF), a process for producing them, cDNA encoding SDNSF, a vector comprising the cDNA, host cells transformed by the vector, an antibody against SDNSF, pharmaceutical compositions containing SDNSF or the antibody, a method of assaying SDNSF, a reagent for assaying SDNSF, and a screening method using SDNSF. The polypeptides are effective in the survival of nerve cells and neuronal stem cells, therefore, efficacious in treating injury to the central nerve system and cancer.

4 Claims, 8 Drawing Sheets

FIG. 1(A)

rSDNSF	1	M A S L Q L L R G P F L C V L L W A F C V - - - - - P G A R A Q E - - - - - H G A	31
mSDNSF	1	M A T L Q L L R A P L L C V L L W V F C A = = = = P G A R A H D = = = = = H G A	31
hSDNSF	1	M T M R S L L R T P F L C G L L W A F C A - - - - - P G A R A E E - - - - - P A A	31
NP_505967	1	M A A N I L V V S - - - C L I L G S F A H Q P Q Q F P G S N Q Q Q P Q Q G G Q A E Q A	40
CG12817	1	M C N L S N L L N F I T C I A S F S Q N F - - - - - D A T L A V K - - - - - R G P	31
rSDNSF	32	G V H H - G S V G L D K S T V H D Q E H I M E H L E G V I N Q - P E A E M S P Q E L Q	72
mSDNSF	32	D V H H - G S V G L D K S T V H D Q E H I M E H L E G V I D Q - P E A E M S P Q E L Q	72
hSDNSF	32	S F S Q P G S M G L D K N T V H D Q E H I M E H L E G V I N K - P E A E M S P Q E L Q	73
NP_505967	41	Q H A Q P G Q Q Q F G G E Q A R D E H H I K E H L D G K V D = P T A N M T P E Q L Q	81
CG12817	32	H H P R G E T R R V D Q H L T H E E H R I D D D L K D M G V Q A N L D D L S E E E K I	74
EF hand 1			
rSDNSF	73	L H Y F K M H D Y D G N S L L D G L E L S T A I T H V H K E E - G - - - - -	104
mSDNSF	73	L H Y F K M H D Y D G N S L L D G L E L S I A I T H V H K E E - G - - - - -	104
hSDNSF	74	L H Y F K M H D Y D G N N L L D G L E L S T A I T H V H K E E - G - - - - -	105
NP_505967	83	F H Y F N M H D L D K N G K L D G V E L I K A I T H F H A E N P G P Q H T Q N N A N A	134
CG12817	75	F Y M F K A H D N D N N A L D G L E M I Q S A M H H N Y D Y F K - - - - - N	108
EF hand 2			
rSDNSF	105	S E Q - - - - - V P P M S E D E - L I S I I D G V L R D D D K M N D G Y I D Y A E F A	141
mSDNSF	105	S K Q - - - - - A P V M S E D E - L V S I I D G V L R D D D K M N D G Y I D Y A E F A	141
hSDNSF	106	S E Q - - - - - A P L M S E D E - L I N I I D G V L R D D D K M N D G Y I D Y A E F A	142
NP_505967	125	N H Q - - - - - P P L P S E V E - L E T M I D S I L K D D D F M A D G F I D Y G E E L	162
CG12817	109	N E R D A Y L - Q N A T D E L E H F I E A I D K F L L T A D D M N D G L L H Y P E F V	150
rSDNSF	142	K S L Q	145
mSDNSF	142	K S L Q	145
hSDNSF	143	K S L Q	146
NP_505967	160	K A Q K L R E D Q A R S H Q E Q M Q K A G G T Q	106
CG12817	151	K A I T G G K E Q P N V D R N I L R	168

FIG. 1(B)

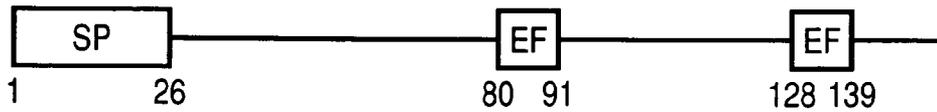


FIG. 2

	HELIX	LOOP	HELIX
rSDNSF 1	LQLHYFKMHDYDGNSSLLDGLELSTAITHV		
rSDNSF 2	IIDGVLRDDDKNNDGYIDYAEFAKSLQ*		
calmodulin 1	EFKEAFALFDKDDGTITTKELGTVMRSL		
calmodulin 2	ELQDMINEVDADGNGTIDFPFLSLMARK		
calmodulin 3	ELIEAFKVFDRDGNLSAAELRHVMTNL		
calmodulin 4	EVDEMIREADIDGDGHINYEYFVRMMVAK		
consensus EF	DXDGDGXIDXXE		

FIG. 3

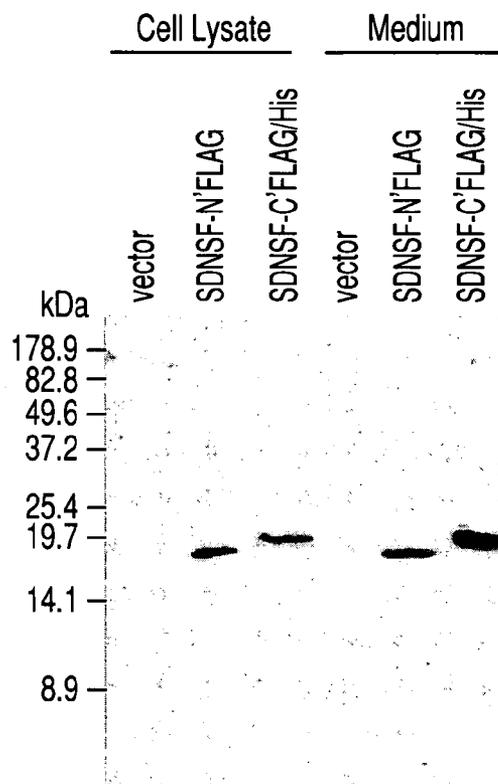


FIG. 4

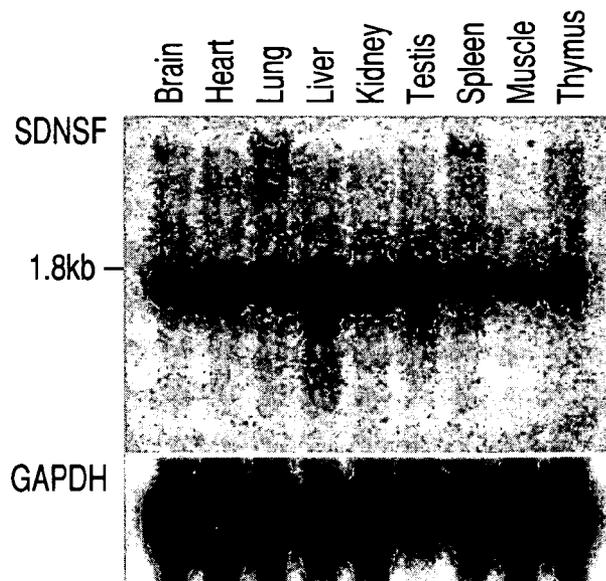


FIG. 5

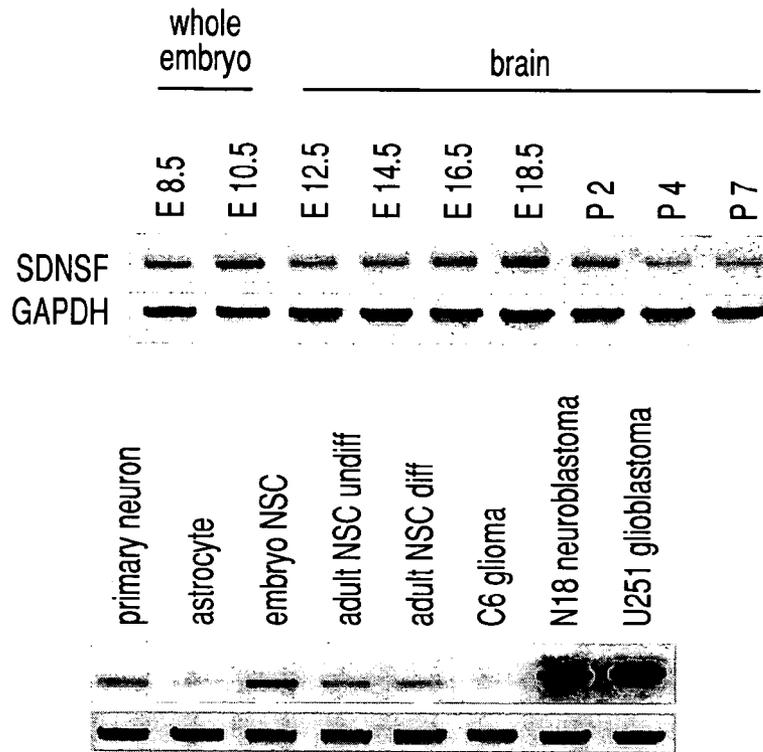


FIG. 6

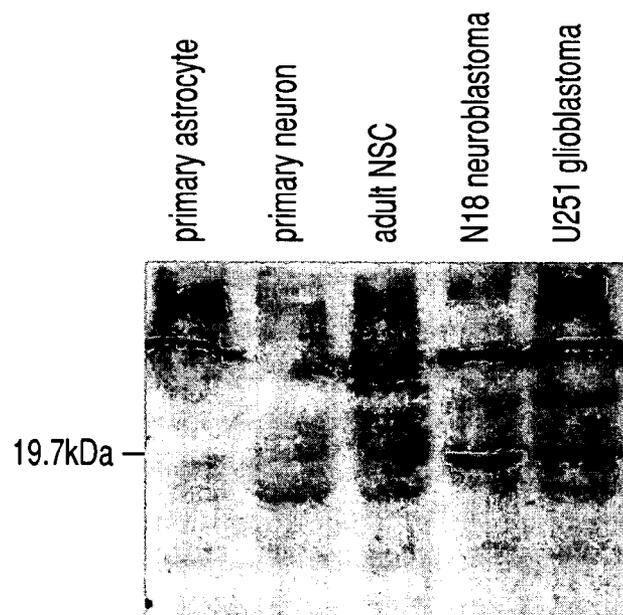


FIG. 7(A)

PRIMARY NEURONS

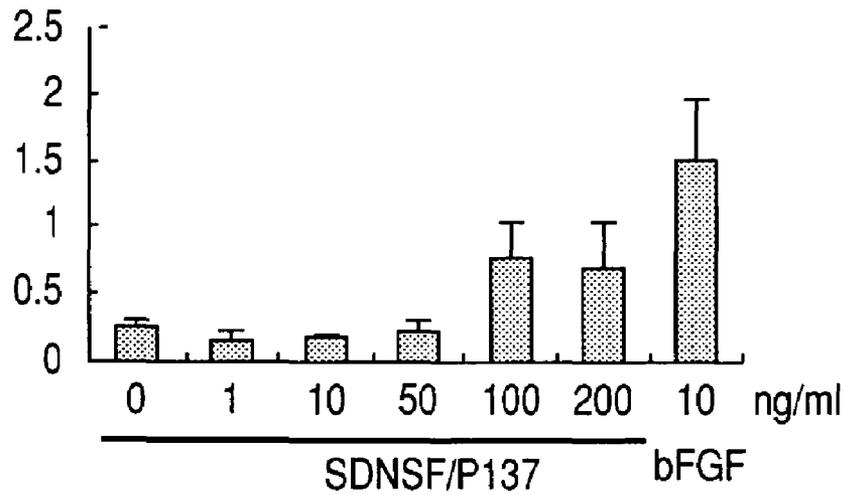


FIG. 7(B)

NEURAL STEM CELLS

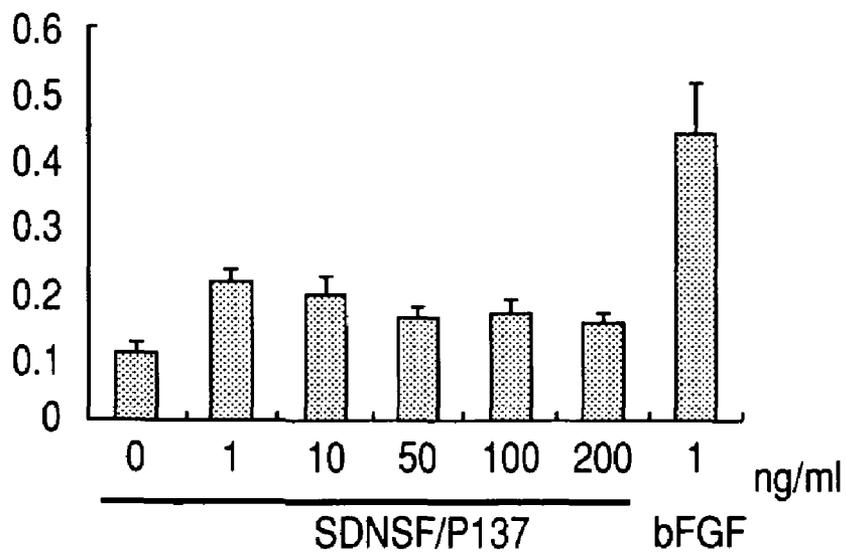


FIG. 8

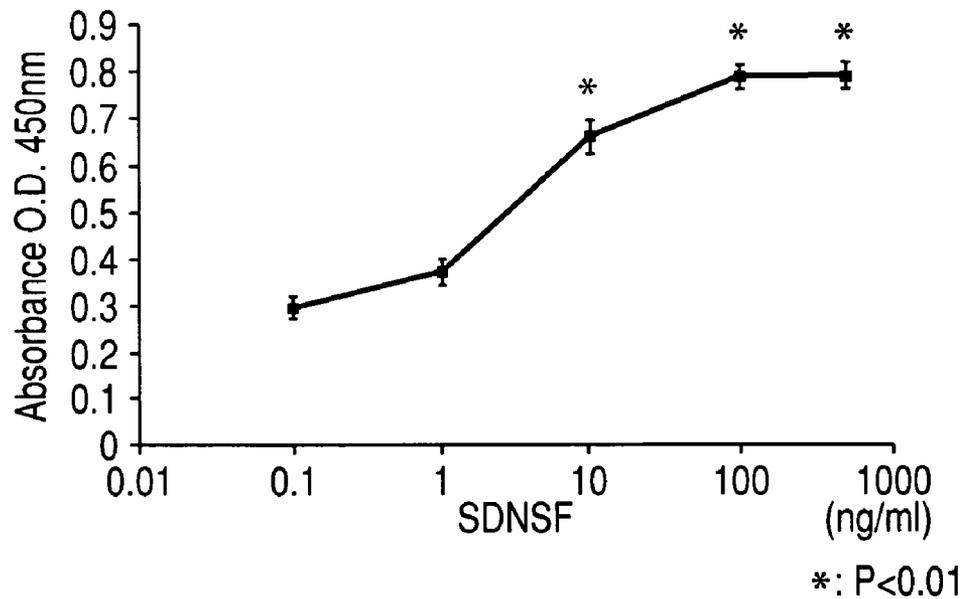


FIG. 9

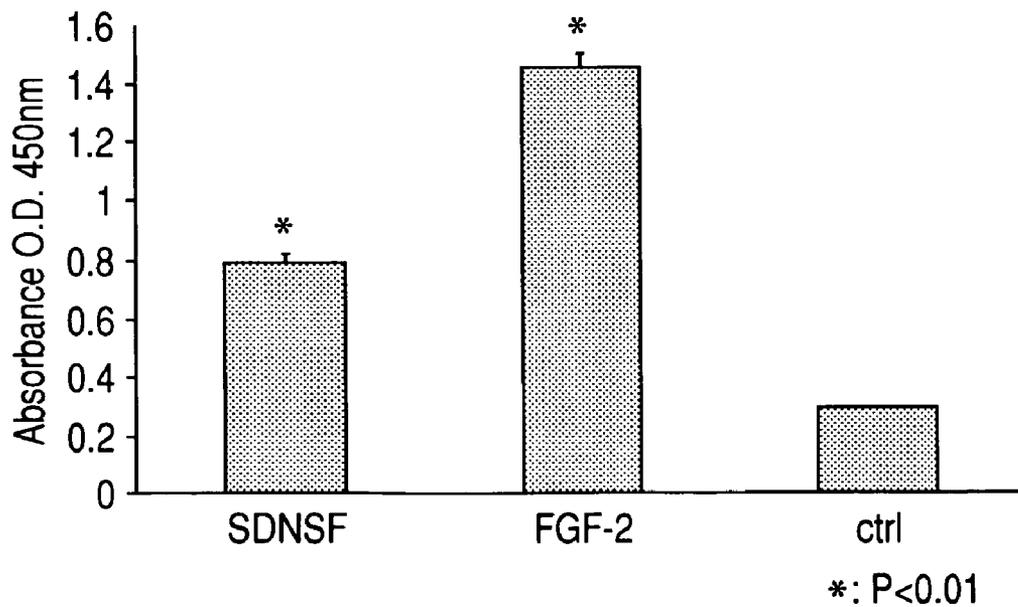


FIG. 10

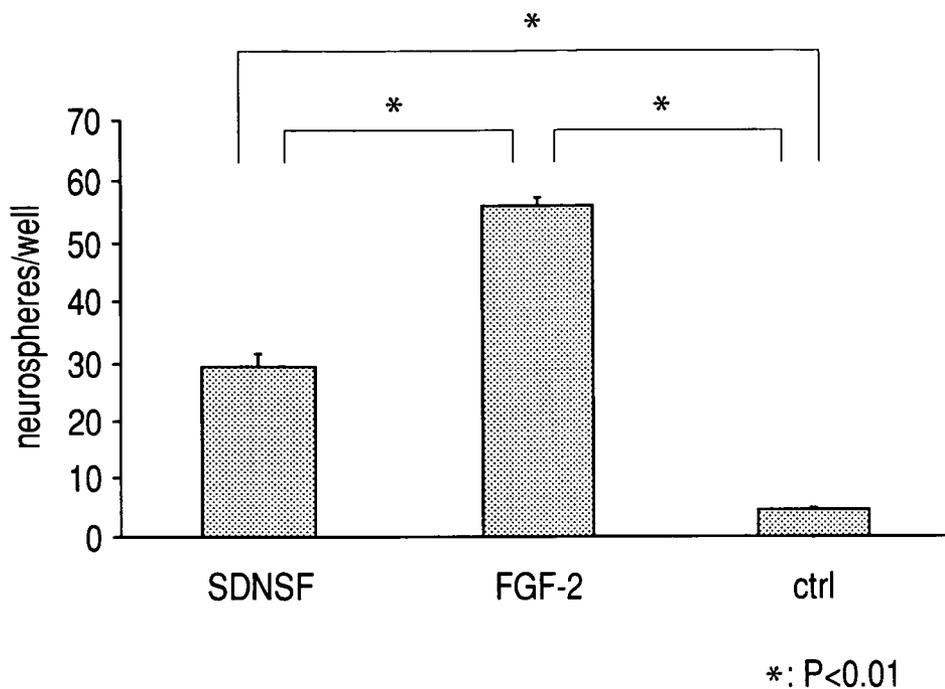


FIG. 11

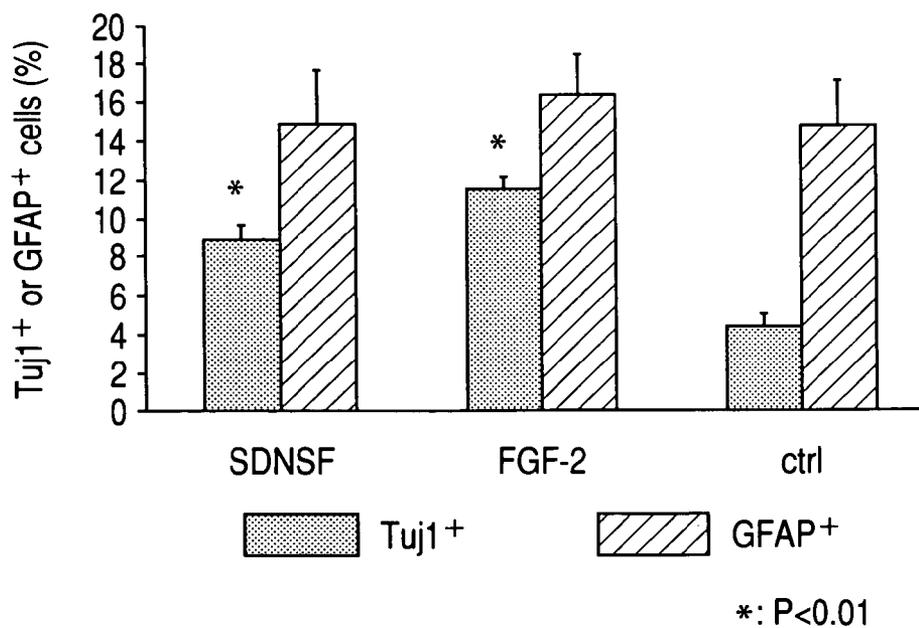


FIG. 12

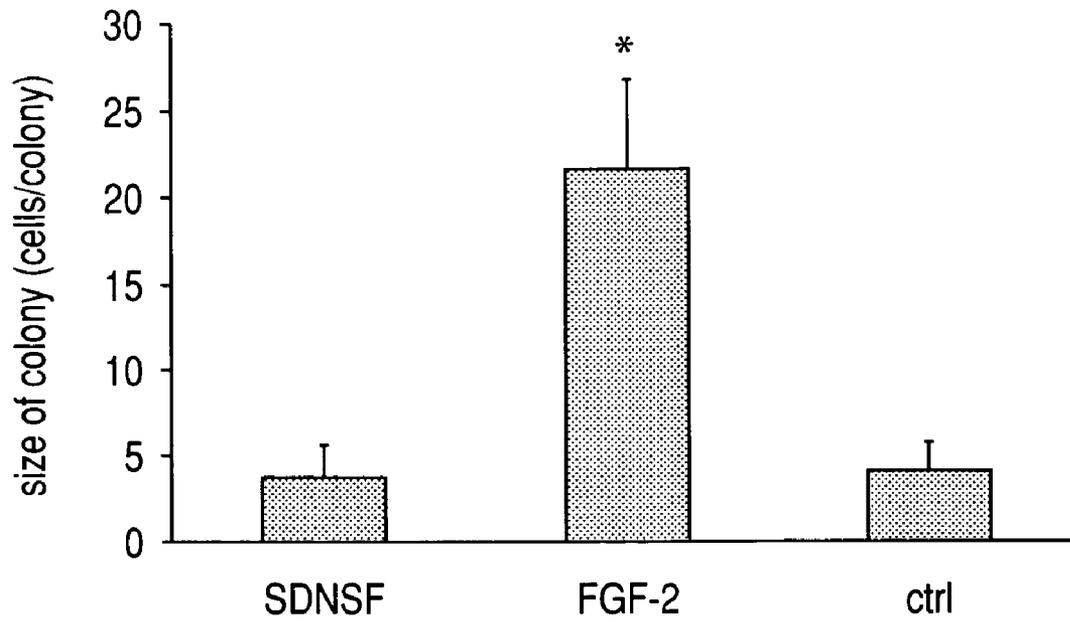


FIG. 13

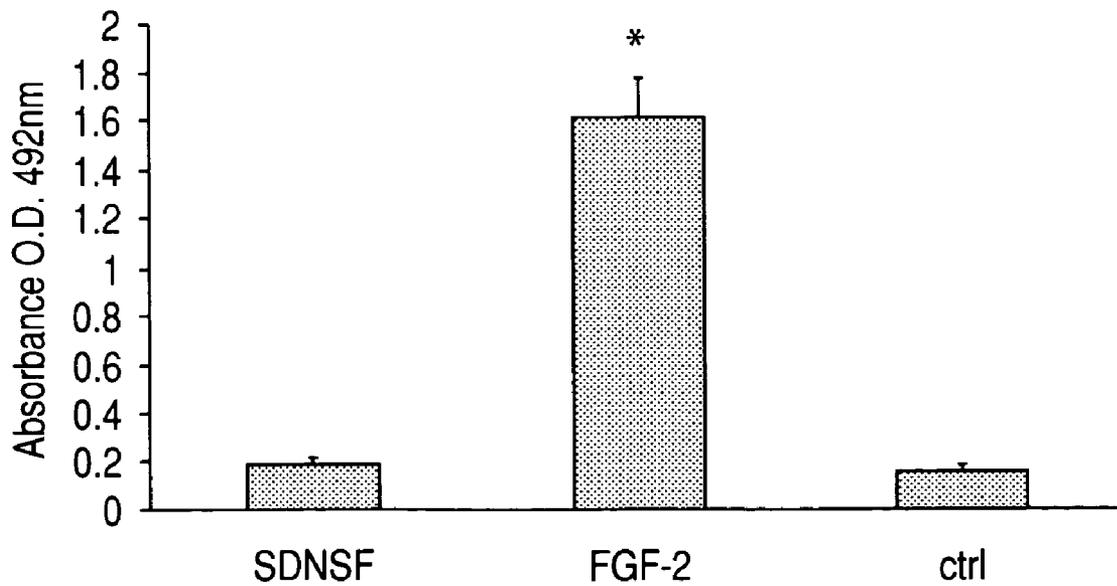


FIG. 14

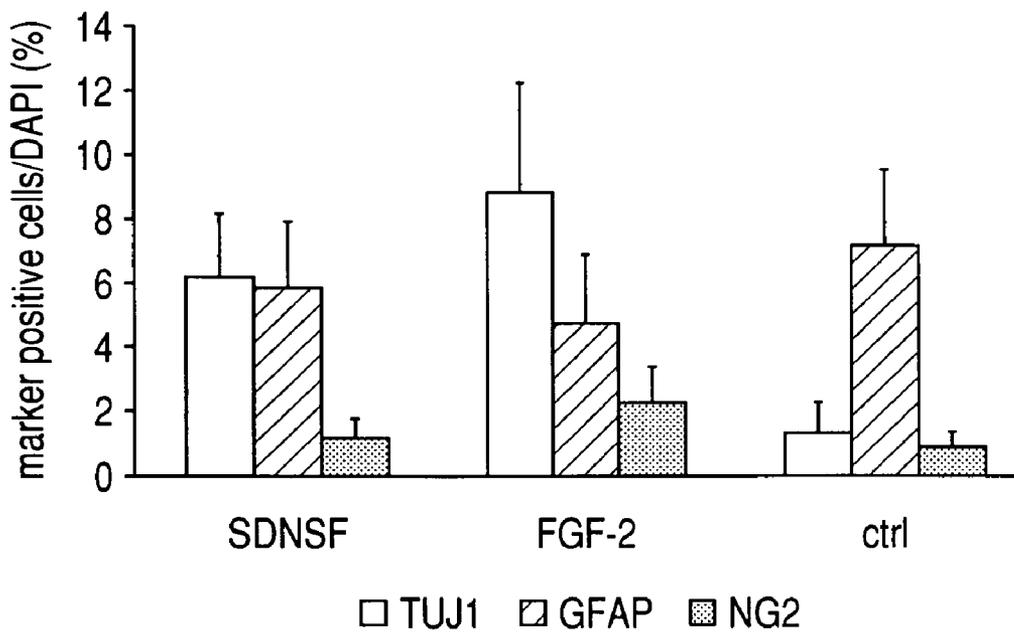
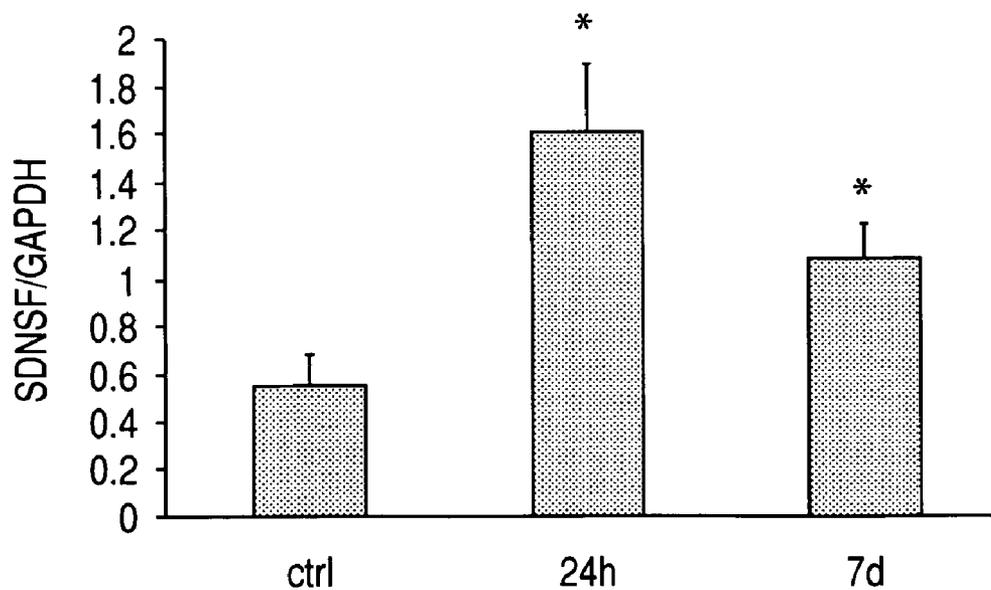


FIG. 15



HUMAN AND MAMMALIAN STEM CELL-DERIVED NEURON SURVIVAL FACTORS

This is a Continuation-In-Part of U.S. application Ser. No. 10/493,393 filed Apr. 22, 2004, now U.S. Pat. No. 7,273,725 of Tasuku HONDO, Kei TASHIRO, Jun TAKAHASHI and Hiroki TODA, entitled HUMAN AND MAMMALIAN STEM CELL-DERIVED NEURON SURVIVAL FACTORS, which is a 371 of PCT/JP02/10936 filed Oct. 22, 2002, the disclosures of which are incorporated herein by reference.

TECHNICAL FIELD

The present invention relates to human, rat and mouse Stem cell-Derived Neuron Survival Factors (hereinafter, simply referred to as "SDNSF"). In more detail, it relates to human, rat and mouse SDNSF, a process for producing them, cDNA encoding SDNSF, a vector comprising the cDNA, host cells transformed by the vector, an antibody against SDNSF, a pharmaceutical compositions containing SDNSF or the antibody or the cDNA or the vector, a method of assaying SDNSF, a reagent for assaying SDNSF, and a screening method using SDNSF.

BACKGROUND ART

In adult brain, it is a long established theory that a new neuron is not generated, and that in an injury to the central nerve system caused by brain infarction, brain hemorrhage, spinal cord injury, etc. and in a neurodegenerative disease such as Parkinson's disease and amyotrophic lateral sclerosis (ALS), recovery of the function which the movement was lost according to the cell death of a neuron was difficult. In recent years, however, it was shown that a neuron was newly generated in adult brain (hippocampus, cerebral cortex association area, lateral cerebral ventricle) of higher mammals such as human and monkey, and that a new neuron in these regions was generated from a neuronal stem cell. It was also demonstrated that a neuronal stem cell existed in aged people's brain and it could differentiate into a neuron. These facts suggest that cerebral regenerative medical treatment is not limited to cell therapy which transplants cells, but therapy which activates inherent neuronal stem cells directly by administering a drug medicine containing protein or compound or by gene therapy technology is possible.

In order to obtain a specific polypeptide or a cDNA encoding it, there have been generally employed methods comprising confirming the target biological activity in a tissue or a cell culture medium and then cloning of a gene through the isolation and purification of a polypeptide and further methods comprising expression-cloning of a gene with the guidance of the biological activity. However, it is frequently observed that a gene, which has been cloned with the guidance of a certain activity, codes for a known polypeptide since many physiologically active polypeptides occurring in vivo have various biological activities. Further, most intravital physiologically active factors are generated only in a trace amount or under a specific physiological condition, which makes the isolation and purification thereof and the confirmation of biological activity difficult.

DISCLOSURE OF THE INVENTION

The inventors focused on novel factors (polypeptides), especially secretory proteins and membrane proteins which have secretion signals, useful for the medical treatment or

diagnosis of an injury to the central nerve system or a neurodegenerative disease, diagnosis or research of a brain tumor, and examined repeatedly and wholeheartedly. Consequently, the inventors have isolated the novel polypeptide molecule concerning a neuronal stem cell, and found out that it is available for regenerative medical treatment above-mentioned and for specific marker of a neuron.

The present inventors have studied the cloning of genes for proliferation and differentiation factors in hemetopoietic and immune systems. They have paid attention to the fact that most secretory proteins such as proliferation and/or differentiation factors (for example various cytokines) and membrane proteins such as receptors thereof (hereinafter these proteins will be referred to generally as secretory proteins and the like) have sequences called signal peptides in the N-termini. Extensive studies have been conducted to provide a process for efficiently and selectively cloning genes encoding signal peptides. As a result, a process (signal sequence trap (SST) method) has been devised using animal cells whereby the existence of a signal peptide can be easily examined (Japanese Patent No. 2,879,303). Furthermore, a process (the yeast SST method) for massively and easily isolating genes encoding signal peptides by using yeast was also developed based on the same concept (U.S. Pat. No. 5,536,637).

By using this method, the inventors have identified successfully a novel secretory protein which is generated by neural stem cells derived from adult rat hippocampus and a cDNA encoding the protein, and found out a full-length cDNA from neural stem cells derived from adult rat hippocampus based on the information.

The inventors have confirmed that the polypeptide had survival supporting activity to some cerebral neurons (primary cultured cells from hippocampus nerve and stem cells derived from hippocampus), as might be explained in detail behind, and completed the invention. The polypeptide is the useful factor as which the function was specified.

The cDNA sequence which this invention offers was identified as a rat SDNSF clone shown in SEQ ID No.1 or 2, and isolated from cDNA library prepared from stem cells derived from hippocampus based on the information obtained by yeast SST method. The rat SDNSF clone shown in SEQ ID No.1 is a full-length cDNA encoding the secretory protein (it is indicated as rat SDNSF protein in the invention).

The cDNA sequence which this invention offers was identified as a human SDNSF clone shown in SEQ ID No.5 or 6, and isolated from cDNA library prepared from stem cells derived from hippocampus based on the information obtained by yeast SST method. The human SDNSF clone shown in SEQ ID No.5 is a full-length cDNA encoding the secretory protein (it is indicated as human SDNSF protein in the invention).

The cDNA sequence which this invention offers was identified as a mouse SDNSF clone shown in SEQ ID No.9 or 10, and isolated from cDNA library prepared from stem cells derived from hippocampus based on the information obtained by yeast SST method. The mouse SDNSF clone shown in SEQ ID No.9 is a full-length cDNA encoding the secretory protein (it is indicated as mouse SDNSF protein in the invention).

The nucleotide sequence encoding rat SDNSF was compared with sequences in GenBank and NCBI utilizing the programs BLASTN, FASTA and UNIGENE, and the amino acid sequence of rat SDNSF was compared with sequences utilizing the programs BLASTP, Fly Database and SwissProt, to reveal no identical sequence. From the results, it became clear that the polypeptide of the invention was a novel secretory protein.

From the facts that the polypeptide of the invention has sustentation activity on neuron in a portion of brain, has EF hand motifs in spite of secretory protein, is a possible cytokine which is regulated by extracellular calcium or calcium from organelle involved in secretory pathway, and has no sequence homology to known neurotrophins, it is thought that the polypeptide is a cytokine which promotes formation or support of survival of neurons other than sympathetic nerve, sensory nerve, neuron in spinal cord motor nerve nucleus and cholinergic nerve in basal ganglia which are known to be neurotrophin dependence by the analysis using deficit mouse of known neurotrophin gene, and that the polypeptide leads to elucidate etiologies of neurodegenerative diseases and to treat them. From the unique characteristic of the polypeptide having survival effect on adult neural stem cells without proliferation activity, it is thought that the polypeptide can also be applied for the protection of neural stem cells in cancer therapy, for example, cancer radiotherapy, cancer chemotherapy and the like. Furthermore, from the facts that the transcripts of the polypeptide are highly up-regulated in ischemically treated hippocampi, it is thought that the polypeptide can be applied for the protection of neural stem cells in ischemic injury, for example, ischemic stroke, ischemic cerebrovascular disease, cerebral ischemic syndrome, and the like.

The present invention relates to:

(1) A substantially purified form of a polypeptide comprising the amino acid sequence shown in SEQ ID NO. 4, 8 or 12, a homologue thereof, a fragment thereof or a homologue of the fragment,

(2) The polypeptide according to (1) which comprises the amino acid sequence shown in SEQ ID NO. 4, 8 or 12,

(3) A cDNA encoding the polypeptide according to (1) or (2),

(4) The cDNA according to (3), comprising the nucleotide sequence shown in SEQ ID NO. 1, 2, 5, 6, 9 or 10, or a fragment selectively hybridized to the sequence,

(5) A replication or expression vector comprising the cDNA according to (3) or (4),

(6) A host cell transformed with the replication or expression vector according to (5),

(7) A process for producing the polypeptide according to (1) or (2), which comprises culturing the host cell according to (6) under a condition effective to express the polypeptide according to (1) or (2),

(8) A monoclonal or polyclonal antibody against the polypeptide according to (1) or (2),

(9) A pharmaceutical composition comprising the polypeptide according to (1) or (2), or the antibody according to (8), or the cDNA according to (3) or (4), or the replication or expression vector according to (5), in association with a pharmaceutically acceptable excipient and/or carrier,

(10) A pharmaceutical composition effective for the medical treatment of a neurodegenerative disease, or a cancer or an ischemic injury, comprising the polypeptide according to (1) or (2) or the cDNA according to (3) or (4) or the replication or expression vector according to (5), in association with a pharmaceutically acceptable excipient and/or carrier,

(11) The pharmaceutical composition according to (10), in which the neurodegenerative disease is an injury to the central nerve system by brain infarction, brain hemorrhage, spinal cord injury, etc.,

(12) A method for measuring the polypeptide according to (1) or (2),

(13) A method for measuring immunochemically the polypeptide according to (1) or (2), comprising using the antibody according to (8),

(14) A reagent for detecting the polypeptide according to (1) or (2), which is used in the method according to (12) or (13),

15. A reagent for testing tumor, comprising using the method according to (12) or (13),

(16) The reagent according to (14), in which the tumor is a brain tumor,

(17) A method for screening a compound having agonistic or antagonistic activity against the polypeptide, comprising using the polypeptide according to (1) or (2),

(18) An agent for the treatment of an injury to the central nerve system which comprises, as an active ingredient, the polypeptide according to (1) or (2), or the cDNA according to (3) or (4), or the replication or expression vector according to (5),

(19) The agent according to (18), in which the injury to the central nerve system is the one caused by a brain infarction,

(20) The agent according to (18), in which the injury to the central nerve system is the one caused by a brain hemorrhage,

(21) The agent according to (18), in which the injury to the central nerve system is the one caused by a spinal cord injury,

(22) An agent for the treatment of cancer which comprises, as an active ingredient, the polypeptide according to (1) or (2) or the cDNA according to (3) or (4) or the replication or expression vector according to (5),

(23) An agent for the treatment of an ischemic injury which comprises, as an active ingredient, the polypeptide according to (1) or (2), or the cDNA according to (3) or (4) or the replication or expression vector according to (5),

(24) The agent according to (23), in which the ischemic injury is the one caused by an ischemic stroke,

(25) The agent according to (23), in which the ischemic injury is the one caused by an ischemic cerebrovascular disease, and

(26) The agent according to (23), in which the ischemic injury is the one caused by a cerebral ischemic syndrome.

DETAILED DESCRIPTION

A polypeptide of SEQ ID NO. 4, 8 or 12 in substantially purified form will generally comprise the polypeptide in a preparation in which more than 90%, eg. 95%, 98% or 99% of the polypeptide in the preparation is that of the SEQ ID NO. 4, 8 or 12.

A polypeptide homologue of the SEQ ID NO. 4, 8 or 12 will be at least 70%, preferably at least 80% or 90%, and more preferably 95% homologous to the polypeptide over a region of at least 20, preferably at least 30 for instance 40, 60, 80 or 100 contiguous amino acids. Such polypeptide homologues will be referred to below as a polypeptide according to the invention.

Furthermore, fragments of SEQ ID NO. 4, 8 or 12 or its homologues will be at least 10, preferably at least 15 for example 20, 25, 30, 40, 50 or 60 amino acids in length of the polypeptide.

A cDNA capable of selectively hybridizing to the cDNA of SEQ ID NO. 1, 2, 5, 6, 9 or 10 will be generally at least 70%,

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preferably at least 80% or 90%, and more preferably at least 95% homologous to the cDNA of SEQ ID NO. 1, 2, 5, 6, 9 or 10 over a region of at least 20, preferably at least 30, for instance 40, 60, 80 or 100 contiguous nucleotides. Such cDNA will be encompassed by the term "cDNA according to the invention".

Fragments of SEQ ID NO. 1, 2, 5, 6, 9 or 10 will be at least 10, preferably at least 15 for example 20, 25, 30 or 40 nucleotides in length of the cDNA. Such fragments will be encompassed by the term "cDNA according to the invention".

Because SDNSF protein of the invention is secreted in great quantities from undifferentiated neuroblastoma and glioblastoma among brain tumors, but not secreted from differentiated glioma, and there is no available neural tumor marker secreted in circulating blood or spinal fluid, SDNSF protein could be the first marker for undifferentiated neural tumor of which detection is possible in blood.

A further embodiment of the invention provides replication and expression vectors comprising cDNA according to the invention. The vectors may be, for example, plasmid, virus or phage vectors provided with an origin of replication, optionally a promoter for the expression of the said DNA and optionally a regulator of the promoter. The vector may contain one or more selectable marker genes, for example an ampicillin resistance gene. The vector may be used in vitro, for example for the production of RNA corresponding to the cDNA, or used to transform a host cell.

A further embodiment of the invention provides host cells transformed or transfected with the vectors for the replication and expression of cDNA according to the invention, including the cDNA of SEQ ID NO. 1, 2, 5, 6, 9 or 10 or the open reading frame thereof. The cells will be chosen to be compatible with the vector and may for example be bacterial, yeast, insect or mammalian.

A further embodiment of the invention provides a method of producing a polypeptide which comprises culturing host cells of the present invention under conditions effective to express a polypeptide of the invention. Preferably, cultivation is carried out under conditions in which the polypeptide of the invention is expressed and then generated from the host cells.

A cDNA according to the invention may also be inserted into the vectors described above in an antisense orientation in order to provide for the production of antisense RNA. Such antisense RNA maybe used in a method of controlling the levels of a polypeptide of the invention in a cell.

An embodiment of the invention also provides monoclonal or polyclonal antibodies against a polypeptide of the present invention. A further embodiment of the invention provides a process for production of monoclonal or polyclonal antibodies to the polypeptides of the present invention. Monoclonal antibodies may be prepared by common hybridoma technology using polypeptides of the present invention or fragments thereof as an immunogen. Polyclonal antibodies may also be prepared by common means which comprises inoculating host animals, for example a rat or a rabbit, with polypeptides of the invention and recovering immune serum.

An embodiment of the invention also provides pharmaceutical compositions containing a polypeptide of the present invention or an antibody against the polypeptide or a cDNA encoding the polypeptide or a vector comprising the cDNA, in association with a pharmaceutically acceptable excipient and/or carrier.

(A) As the polypeptide of the present invention referred to above 1, those which have deficiency in a part of their amino acid sequence (e.g., a mature polypeptide consisted of the only essential sequence for revealing a biological activity in an amino acid sequence shown in SEQ ID NO. 4), those

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which have a part of their amino acid sequence replaced by other amino acids (e.g., those replaced by an amino acid having a similar property) and those which have other amino acids added or inserted into a part of their amino acid sequence, as well as those comprising the amino acid sequence shown in SEQ ID NO. 4, 8 or 12.

As known well, there are one to six kinds of codon encoding one amino acid (for example, one kind of codon for Methionine, and six kinds of codon for leucine are known). Accordingly, the nucleotide sequence of cDNA can be changed without changing the amino acid sequence of the polypeptide.

(B) The cDNA of the present invention referred to above 3 includes every group of nucleotide sequences encoding polypeptides of SEQ ID NO. 4, 8 or 12 shown in (A). There is a probability that yield of a polypeptide is improved by changing a nucleotide sequence.

(C) The cDNA specified in SEQ ID NO. 2, 6 or 10 is an embodiment of the cDNA shown in (B), and indicates the sequence of natural form.

(D) The cDNA shown in SEQ ID NO. 1, 5 or 9 indicates the sequence in which natural non-coding region is added to the cDNA specified in (C).

A cDNA carrying nucleotide sequence shown in SEQ ID NO. 1, 5 or 9 is prepared by the following method.

First, Yeast SST method (see U.S. Pat. No. 5,536,637) is briefly described below.

Yeast such as *Saccharomyces cerevisiae* should secrete invertase into the medium in order to take sucrose or raffinose as a source of energy or carbon (Invertase is an enzyme to cleave raffinose into sucrose and melibiose, sucrose into fructose and glucose.). It is known that many of known mammalian signal sequences make yeast secrete its invertase.

From this knowledge, SST method was developed as a screening method to find a novel signal peptide which enables invertase secretion of yeast from mammalian cDNA library with growth of the yeast on raffinose medium as an index.

Non-secretory type invertase gene SUC2 (GENBANK Accession No. V01311) lacking initiation codon ATG was inserted to yeast expression vector to prepare yeast SST vector pSUC2. In this expression vector, ADH promoter, ADH terminator (both were derived from AAH5 plasmid (Gamerer, Methods in Enzymol. 101, 192-201, 1983)), 2 μ ori (as a yeast replication origin), TRP1 (as a yeast selective marker), ColE1 ori (as a *E. Coli* replication origin) and ampicillin resistance gene (as a drug resistance marker) were inserted. Mammalian cDNA was inserted into the upstream of SUC2 gene to prepare yeast SST cDNA library. Yeast lacking secretory type invertase, was transformed with this library.

If inserted mammalian cDNA encodes a signal peptide, the yeast could survive on raffinose medium as a result of restoring secretion of invertase. By culturing yeast in colonies to prepare plasmids and determine the nucleotide sequence of the insert cDNAs, it is possible to identify novel signal peptide rapidly and easily.

Preparation of yeast SST cDNA library is as follows:

(1) mRNA is isolated from targeted cells, a double-strand cDNA is synthesized by using random primer with certain restriction enzyme (enzyme I) recognition site,

(2) the double-strand cDNA is ligated to adapter containing certain restriction endonuclease (enzyme II) recognition site different from enzyme I, digested with enzyme I and fractionated in a appropriate size,

(3) the obtained cDNA fragment is inserted into yeast expression vector on the upstream region of invertase gene of which signal peptide is deleted and the library is transformed.

Detailed description of each step is as follows:

In step (1), mRNA is isolated from mammalian organs or cell lines after stimulating them with appropriate stimulator if necessary by known methods (as described in Molecular Cloning (Sambrook, J., Fritsch, E. F. and Maniatis, T., Cold Spring Harbor Laboratory Press, 1989) or Current Protocol in Molecular Biology (F. M. Ausubel et al, John Wiley & Sons, Inc.) unless otherwise specified).

A suitable tissue may be heart of fetal mouse. Double-strand cDNA synthesis using random primer is performed by known methods.

Any sites may be used as restriction endonuclease recognition site I which is linked to adapter and restriction endonuclease recognition site II which is used in step (2), insofar as both sites are different each other. Preferably, XhoI is used as enzyme I and EcoRI as enzyme II.

In step (2), ends of cDNA are blunted with T4 DNA polymerase, and ligated to enzyme II adapter and digested with enzyme I. Fragment cDNA is analyzed with agarose-gel electrophoresis (AGE) and cDNA fraction ranging in size from 300 to 800 bp is selected. As mentioned above, any enzyme may be used as enzyme II insofar as it is not same with the enzyme I.

In step (3), cDNA fragment obtained in step (2) is inserted into yeast expression vector on the upstream region of invertase gene of which signal peptide is deleted. *E. coli* is transformed with the expression vector. Many vectors are known as yeast expression plasmid vector. For example, YEp24 is also functioned in *E. Coli*. Preferably pSUC2 as described above is used.

Many host *E. Coli* strains are known as usable for transformation, preferably DH10B competent cell is used. Any known transformation method is available, preferably it is performed by electroporation method. Transformant is cultured by conventional methods to obtain cDNA library for yeast SST method.

However, not all of the cloned cDNA fragments are introduced into this cDNA library. Further, not all of the gene fragments encode unknown (novel) signal peptides. It is therefore necessary to screen a gene fragment encoding for an unknown signal peptide from the library.

Therefore, screening of fragments containing a sequence encoding an appropriate signal peptide is performed by transformation of the cDNA library into *Saccharomyces cerevisiae* (e.g. YT455 strain) lacking the invertase gene or strain which artificially lack the gene (it may be prepared by known methods.). Transformation of yeast is performed by known methods, e.g. lithium acetate method. Transformant is cultured in a selective medium, then transferred to a medium containing raffinose as a carbon source. Survival colonies are selected and then plasmid is collected. Survival colonies on a raffinose-medium indicates that some signal peptide of secretory protein was inserted to this clone.

With respect to isolated positive clones, the nucleotide is determined. As to a cDNA encoding unknown protein, full-length clone may be isolated by using cDNA fragment as a probe, and then the full-length nucleotide sequence is determined. The manipulation is performed by known methods.

Once the nucleotide sequences shown in SEQ ID NO. 1, 5 or 9 are determined partially or preferably fully, it is possible to obtain cDNA encoding mammalian protein itself, homologue or subset. By screening cDNA library or mRNA derived from mammals by PCR method with any synthesized

oligonucleotide primers or by hybridization with any fragment as a probe, it is possible to obtain cDNA encoding other mammalian homologue protein from other mammalian cDNA or genome library.

5 If the cDNA obtained above contains a nucleotide sequence of cDNA fragment obtained by SST (or homologous sequence thereof), it implies that the cDNA encodes signal peptide. Accordingly, it is clear that the length of the cDNA is full or almost full. (All signal sequences exist at N-termini of a protein and are encoded at 5'-termini of open reading frame of cDNA.)

10 By known methods, the confirmation of full-length may be carried out by Northern analysis with the said cDNA as a probe. The cDNA is assumed to have almost complete length if the length of the cDNA is almost the same with the length of the mRNA obtained in the hybridizing band.

15 The present invention provides both types of protein, i.e., full-length and mature. The full-length proteins are specified with the amino acid sequences translated from the nucleotides shown in SEQ ID NO. 4, 8 or 12. The mature proteins are obtained by expression in suitable mammal cells or other host cells transformed by the full-length DNA shown in SEQ ID NO.1, 5 or 9. Sequences of mature proteins could be predicted from full-length amino acid sequences.

20 Once the nucleotide sequences shown in SEQ ID No. 1, 2, 5, 6, 9 or 10 are determined, cDNAs of the present invention are obtained by chemical synthesis, or by hybridization making use of nucleotide fragments which are chemically synthesized as a probe. Furthermore, cDNAs of the invention are obtained in desired amount by transforming a vector that contains the DNA into a proper host, and culturing the transformant.

The polypeptides of the present invention may be prepared by:

- 35 (1) isolating and purifying from an organism or a cultured cell,
 - (2) chemically synthesizing, or
 - 40 (3) using recombinant DNA technology,
- preferably, by the method described in (3) in an industrial production.

Examples of expression system (host-vector system) for producing a polypeptide by using recombinant DNA technology are the expression systems of bacteria, yeast, insect cells and mammalian cells.

In the expression of the polypeptide, for example, in *E. Coli*, the expression vector is prepared by adding the initiation codon (ATG) to 5' end of a cDNA encoding mature peptide, connecting the cDNA thus obtained to the downstream of a proper promoter (e.g., trp promoter, lac promoter, λ PL promoter, and T7 promoter), and then inserting it into a vector (e.g., pBR322, pUC18 and pUC19) which functions in an *E. coli* strain.

55 Then, an *E. coli* strain (e.g., *E. coli* DH1 strain, *E. coli* JM109 strain and *E. coli* HB101 strain) which is transformed with the expression vector described above may be cultured in an appropriate medium to obtain the desired polypeptide. When a signal peptide of bacteria (e.g., signal peptide of pel B) is utilized, the desired polypeptide may be also released in periplasm. Furthermore, a fusion protein with other polypeptide may also be produced.

In the expression of the polypeptide, for example, in mammalian cells, for example, the expression vector is prepared by inserting the DNA encoding nucleotide shown in SEQ ID NO. 1, 2, 5, 6, 9 or 10 into the downstream of a proper promoter (e.g., SV40 promoter, LTR promoter and metal-

lothionein promoter) in a proper vector (e.g., retrovirus vector, papilloma virus vector, vaccinia virus vector and SV40 vector). A proper mammalian cell (e.g., monkey COS-1 cell, COS-7 cell, Chinese hamster CHO cell, mouse L cell etc.) is transformed with the expression vector thus obtained, and then the transformant is cultured in a proper medium, the secretory protein of the present invention can be secreted into the culture medium as the aimed polypeptide. Then, by linking to cDNA fragment coding other polypeptides, for example, common region (Fc portion) of antibody, fusion proteins can be produced. Polypeptides obtained by the method above can be isolated and purified by conventional biochemical methods.

INDUSTRIAL APPLICABILITY

A polypeptide and cDNA encoding it of the invention are thought to have one or more effects or biological activities (The effects or biological activities relevant to assay enumerated below are included).

Administration or use of the protein or of cDNA coding the protein (for example, gene therapy (including regenerative therapy) or vectors suitable for cDNA transfection) may provide the effect or biological activities described about the protein of the invention.

The polypeptide of the invention has survival effect for neuron in a portion of brain (primary cultured cells from hippocampus nerve and stem cells derived from hippocampus) and, therefore, can be applied in treating neurodegenerative disease (injury in the central nerve system by brain infarction, brain hemorrhage, spinal cord injury, etc.) . As the polypeptide has survival effect for adult neural stem cells without proliferation activity, it can also be applied for the protection of neural stem cells in cancer therapy, for example, cancer radiotherapy, cancer chemotherapy and the like. Furthermore, the transcripts of the polypeptide are highly up-regulated in ischemically treated hippocampi and, therefore, the polypeptide can be applied for the protection of neural stem cells in ischemic injury, for example, ischemic stroke, ischemic cerebrovascular disease, cerebral ischemic syndrome, and the like.

Quantitative analysis of the polypeptide of the present invention in vivo can be performed using polyclonal or monoclonal antibodies against the polypeptide of the present invention. It can be used in studies on relationship between this polypeptide and disease, or diagnosis of disease, etc. The polyclonal and the monoclonal antibodies can be prepared using this polypeptide or its fragment as an antigen by conventional methods.

Identification, purification or gene cloning of known or unknown proteins (ligands) which are connected with the polypeptide of the present invention can be performed using the polypeptide of the present invention by, for example, preparation of the affinity-column.

Identification of molecules which interact with the polypeptide, molecular cloning of the gene may be conducted, for example, by west-western blot, using the polypeptide, or by yeast two-hybrid method, using the cDNA (desirably cDNA coding the polypeptide).

Screening, which can identify agonists or antagonists against the polypeptide receptor and inhibitors against interaction between receptors and signal transduction molecules can be performed by using the polypeptide.

For example, the screening could be performed by the following steps:

(a) The polypeptide of the invention, compound to be screened and reaction mixture including cells are mixed (the

reaction mixture includes markers which are transferred into cells as the cell grows and peptides except for the polypeptide for efficient observation of the function of the polypeptide.) under condition which the cells are normally stimulated by the polypeptide, then,

(b) It is determined whether the compound is a useful agonist or antagonist by measuring the cell growth.

The cDNA of the invention may be useful not only as an important and essential template in production of the polypeptide of the present invention which is expected to have a considerable utility, but also for diagnoses and treatments of hereditary diseases (treatments of gene deficiency or treatments which anti-sense DNA(RNA)s intercept expression of polypeptides, etc) . In addition, genomic DNAs may be isolated by using cDNA of the invention as a probe.

For the usage for above mentioned diseases, administration of the polypeptide of the invention or the antibodies against the polypeptide of the invention can be carried out in systemic or local, generally peroral or parenteral ways. Oral, intravenous and intracerebroventricular administration are preferred.

The dosage to be administered depends upon age, body weight, symptom, desired therapeutic effect, route of administration, and duration of the treatment etc. In human adults, one dose per person is generally between 100 µg and 100 mg by oral administration up to several times per day, or between 10 µg and 100 mg by parenteral administration up to several times per day.

As mentioned above, the doses to be used depend upon various conditions. Therefore, there are cases in which doses lower than or greater than the ranges specified above may be used.

The compounds of the present invention may be administered as solid compositions, liquid compositions or other compositions for oral administration, or as injections, liniments or suppositories etc. for parenteral administration.

Examples of solid compositions for oral administration include compressed tablets, pills, capsules, dispersible powders and granules etc. Examples of capsules include hard capsules and soft capsules.

In such solid compositions, one or more of the active compound(s) is or are admixed with at least one inert diluent (such as lactose, mannitol, glucose, hydroxypropyl cellulose, microcrystalline cellulose, starch, polyvinylpyrrolidone, magnesium metasilicate aluminate, etc.) . The compositions may also comprise, as is normal practice, additional substances other than inert diluents: e.g. lubricating agents (such as magnesium stearate etc.), disintegrating agents (such as cellulose calcium glycolate, etc.), stabilizing agents (such as human serum albumin, lactose etc.), and assisting agents for dissolving (such as arginine, asparaginic acid etc.).

The tablets or pills may, if desired, be coated with a film of gastric or enteric materials (such as sugar, gelatin, hydroxypropyl cellulose or hydroxypropylmethyl cellulose phthalate, etc.), or be coated with more than two films. And then, coating may include containment within capsules of absorbable materials such as gelatin.

Liquid compositions for oral administration may contain pharmaceutically-acceptable emulsions, solutions, suspensions, syrups and elixirs, and also may contain inert diluent(s) commonly used (purified water, ethanol etc.) Besides inert diluents, such compositions may also comprise adjuvants (such as wetting agents, suspending agents, etc.), sweetening agents, flavoring agents, perfuming agents, and preserving agents.

Other compositions for oral administration include spray compositions which may be prepared by known methods and which comprise one or more of the active compound(s). Spray compositions may comprise additional substances other than inert diluents: e.g. stabilizing agents such as sodium sulfite etc., stabilizing agents providing for isotonic behavior, isotonic buffer (sodium chloride, sodium citrate, citric acid, etc.). For preparation of such spray compositions, for example, the methods described in the U.S. Pat. Nos. 2,868,691 and 3,095,355 may be used.

Injections for parenteral administration include sterile aqueous or non-aqueous solutions, suspensions and emulsions. In aqueous or non-aqueous solutions or suspensions, one or more active compound(s) is or are admixed with at least one inert diluent(s). Aqueous diluents may be distilled water for injection, physiological salt solution, etc. Inert non-aqueous diluents(s) maybe propylene glycol, polyethylene glycol, oil of the plant such as olive oil, alcohol such as ethanol, POLYSOLBATE 80™, etc.

Such compositions may comprise additional preserving agents, wetting agents, emulsifying agents, dispersing agents, stabilizing agent (such as human serum albumin, lactose, etc.), and assisting agents such as assisting agents for dissolving (arginine, asparaginic acid, etc.).

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 (A) shows an alignment of deduced amino acid sequences of human, mouse, and rat SDNSF, (B) existence of two EF hand motifs (calcium binding motif) in the downstream of the signal peptide,

FIG. 2 shows the EF hand motifs of SDNSF and those of calmodulin, calcium secretory protein, have common sequences,

FIG. 3 is the result of Western Blot analysis with anti-SDNSF antibody and anti-FLAG antibody, showing SDNSF protein is secreted into the culture medium,

FIG. 4 is the result of blotting with the cDNA fragment ³²P-labeled and purified by gel as a probe,

FIG. 5 shows the expression of SDNSF mRNA in primary neurons, astrocytes, neural stem cells (embryo NSCs, adult NSCs undifferentiated and adult NSCs differentiated), rat glioma C6, mouse neuroblast N18 and human glioblastoma UG251,

FIG. 6 shows the expression of SDNSF protein in primary astrocytes, primary neurons, adult NSCs, mouse neuroblast N18 and human glioblastoma UG251,

FIG. 7 (A) (B) shows the effect of SDNSF addition on the survival of primary hippocampal neurons and neural stem cells,

FIG. 8 shows that SDNSF improves primary adult neural stem cells viability in a dose dependent manner (Significant differences versus control are indicated by an asterisk (*) (P<0.01)),

FIG. 9 shows the effect of SDNSF on the survival of neural stem cells cultured in the absence of FGF-2 (Significant differences versus control are indicated by an asterisk (*) (P<0.01)),

FIG. 10 shows the effect of SDNSF on self-renewal of neural stem cells by counting neurospheres formed in the absence of FGF-2 (Significant differences versus control are indicated by an asterisk (*) (P<0.01)),

FIG. 11 shows the effect of SDNSF on the differentiation of neural stem cells into neurons in the neurospheres formed in the presence of SDNSF without FGF-2 (Significant differences between SDNSF-treated, FGF-2-treated and control groups are indicated by an asterisk (*) (P<0.01)),

FIG. 12 shows the result of colony size assay at DIV6 to assess cell proliferation in the presence of 100 ng/ml of SDNSF,

FIG. 13 shows the result of BrdU cell proliferation ELISA at DIV4 to assess cell proliferation in the presence of 100 ng/ml of SDNSF,

FIG. 14 shows the differentiation profiles of the adult neural stem cells that were treated with SDNSF in the absence of FGF-2, and

FIG. 15 shows the result of quantitation of SDNSF transcripts in ischemically treated hippocampi, in which quantitated amounts of SDNSF transcript normalized with corresponding GAPDH are plotted in each group.

BEST MODE FOR CARRYING OUT THE INVENTION

The present invention is more specifically explained by means of the following examples regarding SDNSF, but is not limited only to these examples.

Example 1

Preparation of Poly(A)⁺ RNA

Total RNA was extracted from PZ5 cells, which was cloned from neural stem cells derived from adult rat hippocampus, with TRIzol Reagent™ (purchased from Life Technologies, Inc), and poly(A)⁺ RNA was purified with Oligotex-dT30<Super>™ (purchased from Roche).

Example 2

Construction of Yeast SST cDNA Library

Double-stranded cDNA was synthesized from above-described poly(A)⁺ RNA with a primer, in which XhoI site 9 mer was connected with,

5'-TCC CGA TTG AAT TCT AGA CCT GCC TCG AGN NNN NNN NN-3' (SEQ ID NO.13)

by using Super Script Choice System™ (purchased from Life Technologies, Inc). It was connected with EcoRI adaptor (purchased from GIBCOERL) by using DNA ligation kit Ver.2™ (purchased from TAKARA SYUZO hereinafter this kit was used for ligation of cDNA), digested with XhoI, electrophoresed in agarose gel and 400-800 bp cDNAs were cut off from the gel. The cDNAs were inserted into EcoRI/XhoI site of pSuc2t71ori (see U.S. Pat. No. 5,536,637), introduced into *E. coli* DH10 by electroporation to obtain cDNA library for yeast SST method.

Example 3

Screening by SST and Sequencing of Positive Clones

Plasmids were prepared from the cDNA library, yeast YTK12 was transformed with the plasmids by lithium acetate method (see Current Protocols in Molecular Biology 13.7.1) and plated onto selection medium for yeast transformants (CMO-Trp medium) lacking tryptophan. After 48 hours incubation at 30° C., colonies (transformants) were replicated onto YPR plates, of which carbon source is raffinose, by using Accutran Replica Plater™ (purchased from Schleicher & Schuell) and incubated for 14 days at 30° C.

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On day 3 or later, each yeast colony was purified by streaking onto YPR plates and incubated for 48 hours at 30° C. Single colony was inoculated into YPD medium and incubated for further 48 hours at 30° C., then plasmid was prepared. PCR reaction was performed with two kinds of primers having the sequences at the ends of pSUC2 cloning site (primer for sense-strand is biotin-labelled) by known method to amplify insert cDNA, biotin-labelled single-stranded cDNA was purified using Dynabeads™ (purchased from DYNAL) and then sequenced by cycle-sequencing method using fluorescence-dye terminator with DNA Sequencing kit (Dye Terminator Cycle Sequencing Ready Reaction™) (purchased from Applied Biosystems Inc.). DNA sequencer 373 (Applied Biosystems Inc.) was used for reading the nucleotide sequence (hereinafter sequencing was carried out by this method).

DNA sequences thus obtained and deduced amino acid sequences were compared with sequences in data bases, it became clear that a clone named SDNSF was a novel cDNA. Therefore, full-length cDNA cloning was tried with this fragment cDNA of SDNSF clone (hereinafter referred to as "SDNSF SST fragment cDNA"). A comparison of the deduced amino acid sequence with those of known signal peptides indicated that SDNSF SST fragment cDNA had signal peptide both functionally and structurally.

Example 4

Full-Length cDNA Cloning and Sequencing

One million plaques obtained from PZ5 cDNA library were transferred to nylon membrane. Hybridization was carried out with 32^P-labelled rat SDNSF SST fragment cDNA as a probe by known method to obtain many positive clones. One clone among them was isolated, introduced into *E. coli* DH5α, and its plasmid was prepared. After sequencing of 5' region of the insert and confirming the DNA contained the sequence of rat SDNSF SST fragment cDNA, full length sequencing was performed to obtain the sequence shown in SEQ ID No.1. The open reading frame was also determined to obtain the translated amino acid sequence shown in SEQ ID No.2 and the deduced amino acid sequence shown in SEQ ID No.4.

The amino acid sequence and nucleotide sequence encoding the polypeptide of the invention (referred to as rat SDNSF polypeptide) were compared with sequences in NCBI data base to reveal no identical sequence. Furthermore, it became clear that rat SDNSF polypeptide had no transmembrane region and that rat SDNSF polypeptide was a novel secretory protein.

The result of motif search revealed that SDNSF had a signal peptide and two EF hand motifs (calcium binding motif) downstream of the signal peptide (FIG. 1). EF hand motif of SDNSF and that of calmodulin, a calcium secretory protein, have common sequences (FIG. 2). It is rare that a secretory protein has the EF hand motif, however, it is reported that BM-40 and its related proteins have a similar structure. The EF hands in BM-40 is suspected to be involved in the conformation change of the protein depending on the concentration of calcium in vesicle and secretion efficiency of BM-40 (Literature 1: Busch E et al., Calcium affinity cooperativity and domain interaction of extracellular EF-hands present in BM-40., *J. Biol. Chem.*, 275(33), 25508-15 (2000)).

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Example 5

Sequencing of Human and Mouse SDNSF Genes

Homology searches on mammalian ESTs and UNIGENE DNA data bases revealed human and mouse ESTs homologous to rat SDNSF.

Consequently, the inventors isolated full length human and mouse SDNSF genes using the sequence information by known method and sequenced completely to obtain the nucleotide sequences shown in SEQ ID No.5 and 9, respectively. The open reading frame was also determined to obtain the deduced amino acid sequence shown in SEQ ID No.8 and 12, respectively. It was revealed from above information that said human and mouse clones were full length and their amino acid sequences were 87% and 90% identical to that of rat SDNSF, respectively.

The nucleotide sequences and amino acid sequences of the human and mouse SDNSF were compared with sequences on nucleotide and amino acid data bases to reveal no identical sequence as was the case with rat SDNSF. From this, it became clear that the polypeptides of the invention were novel secretory proteins as well.

Example 6

Homology Searches on Data Bases for Non-Mammals

Homology searches on nematode and *drosophila* data bases revealed that F55A11.1, which had been reported to be a virtual protein of nematode, and CG12817, which had been reported to be a gene product of *drosophila*, had 20 to 30% identities in amino acid sequence with the polypeptides. These data base searches suggest that SDNSF genes are highly conserved.

Example 7

Preparation of Anti-SDNSF Polyclonal Antibody

Three kinds of rat SDNSF partial polypeptides were synthesized by solid-phase method and conjugated to Keyhole limpet hemocyanin (KLH),

Asp Lys Ser Thr Val His Asp Gln Glu His Ile Met Glu His Leu Glu Cys-KLH

(amino acid sequence 15-30 in SEQ ID No.4) His Lys Glu Glu Gly Ser Glu Gln Val Pro Pro Met Ser Glu Asp Glu Cys-KLH

(amino acid sequence 74-89 in SEQ ID No.4) KLH-Cys Asp Gly Tyr Ile Asp Tyr Ala Glu Phe Ala Lys Ser Leu Gln

(amino acid sequence 106-119 in SEQ ID No.4) and immunized rabbits as an immunogen to obtain serum after measuring levels of antibody to the protein. Anti-SDNSF polyclonal antibodies were purified using affinity column, in which each peptide fragment used as an immunogen was bound.

Example 8

Investigation of Secretory Pathway of SDNSF

Modified SDNSF proteins, which were tagged with inserted FLAG at N terminus (FLAG-SDNSF) or with FLAG-6His at C terminus (SDNSF-C' FLAG-6His) of rat SDNSF, were expressed in 293T cells. The secretion of these tagged SDNSF proteins into culture supernatant was exam-

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ined with Western blot analysis using both anti-SDNSF and anti-FLAG antibodies (FIG. 3).

Example 9

Localization of Rat SDNSF

Total RNA was extracted from adult rat brain, heart, lung, liver, spleen, kidney, testis, skeletal muscle and thymus with TRIzol Reagent™ (purchased from Life Technologies, Inc), and poly(A)⁺ RNA was purified with Oligotex-dT30 <Super>™ (purchased from Roche).

The poly(A)⁺ RNA from various tissues were subjected to formaldehyde-gel electrophoresis and blotting according to the method of Sambrook et al. (Molecular Cloning (1989)). The detection using gel-purified and ³²P-labelled cDNA fragment as a probe revealed that SDNSF were expressed in all tissues tested as shown in FIG. 4.

Total RNA was extracted from rat whole embryo and embryonic brain, brain up to postnatal day 7, primary neurons and cell lines with TRIzol Reagent™ (purchased from Life Technologies, Inc), and mRNA expression were examined by using RT-PCR. As shown in FIG. 5, the SDNSF transcript was expressed in cultured stem cells, primary neurons and neural stem cells. The SDNSF transcript was also expressed in human glioblastoma UG251 cells and mouse neuroblastoma N18 cells, but not in primary glia cells and rat glioma C6 cells. Furthermore, the expression of SDNSF protein was detected in human glioblastoma UG251 cells and mouse neuroblastoma N18 cells by Western blot analysis (FIG. 6).

Example 10

Viability Assays of SDNSF on Neurons and Stem Cells

Based on the localization of SDNSF, biological effects of SDNSF on neurons and stem cells were examined.

Modified SDNSF protein (SDNSF-FLAG-6His), which was purified by using Ni-NTA method utilizing His structure in the molecule, was added to the cultures of rat primary hippocampal neurons and neural stem cells derived from rat hippocampus, and WST reduction assay was performed to measure the number of viable cell at day 4. As shown in FIG. 7, in primary neurons, SDNSF at the concentration of 100 ng/ml was effective on cell survival as compared with control group. In neural stem cells, it was shown that SDNSF tended to improve viable cell numbers as compared with control group. It was also shown that SDNSF improved the viability of neural stem cells derived from rat hippocampus in a dose-dependent manner when cells were cultured in FGF-2 (fibroblast growth factor-2) minus growth medium for 5 days (FIGS. 8 and 9). To see whether this difference of viability came from proliferation activity of SDNSF, mitogenic activity was examined by tracking single cell proliferation via BrdU incorporation. In addition, the progeny from adult neural stem cells (ANSCs) were tracked by marking single ANSCs with replication-deficient GFP-expressing recombinant retrovirus. ANSCs treated with 100 ng/ml of SDNSF did not proliferate as much as FGF-2-treated ANSC, and the sizes of the colonies in SDNSF group were similar to that of the control group (FIG. 12). BrdU ELISA assay did not reveal an increased uptake of BrdU in the ANSCs treated with SDNSF (FIG. 13).

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Example 11

Test for SDNSF's Effect on Self-Renewal and Differentiation of Neural Stem Cells

After neural stem cells derived from rat hippocampus were cultured in the SDNSF⁺/FGF-2⁻ medium for 5 days, cells were replated on noncoated plates and grown for 6 days in growth medium containing FGF-2 (20 ng/ml), and the number of neurospheres were counted. A neuronal marker, Tuj-1-positive cells were also counted. AS shown in FIGS. 10 and 11, the number of neurospheres/well from SDNSF-treated group was significantly larger than that from the control group. Proportion of neurons in the neurospheres from SDNSF-treated group was significantly larger than that from the control group. These results indicate that SDNSF have activities for retaining self-renewal potentials of neural stem cells and for promoting the differentiation of neural stem cells into neuronal and glial phenotypes as well as FGF-2.

Example 12

Assessment of Multipotency of SDNSF-Treated ANSCs

To test the differentiation potential of SDNSF-treated ANSCs, after ANSCs were cultured in the SDNSF⁺/FGF-2⁻ medium (100 ng/ml of SDNSF without FGF-2) for 6 days, those ANSCs were replated on noncoated Nunc 48-well plates at the density of 2,000 cells/cm² and grown for 6 days in growth medium containing 20 ng/ml of FGF-2. The newly formed neurospheres were replated onto poly-L-ornithine/laminin-coated Labtek 4-well chamber Slides™ (purchased from Nunc) at the density of 10-30 spheres/well and cultured in differentiation medium (containing 0.5 micromolar retinoic acid) for further 6 days. Under confocal laser microscope or fluorescent microscope, positively stained cells (Tuj-1-positive cells, a neuronal marker, GFAP-positive cells, a glial marker, and NG-2-positive cells, an oligodendrocyte marker) were quantified at least 20 fields systematically across the coverslips from three to four independent experiments of parallel cultures. The neurospheres from SDNSF-treated ANSCs could differentiate into three different cellular types: neurons, astrocytes, and oligodendrocytes (FIG. 14). From these results, the surviving cells in SDNSF-treated ANSCs still retained self renewal potentials and maintained multipotency to differentiate into neuronal and glial phenotypes as has been seen in FGF-2-treated ANSCs.

Example 13

Quantitation of SDNSF Transcripts in Ischemically Treated Hippocampi

Transient global ischemia was induced on male Sprague-Dawley rats (300-350 g; Charles River Laboratories, Wilmington, Mass.) by bilateral common carotid artery occlusion and induced hypotension using the modified two-vessel occlusion method (Smith M. L. et al., Acta Neurol. Scand. 69:385-401 (1984)). In brief, the animals were anesthetized with 1.5% isoflurane, 68.5% nitrous oxide, and 30% oxygen and monitored from the femoral artery with PE-50 catheter (427410; Becton Dickinson). After exposure of the right jugular vein and both common carotid arteries, 150 IU/kg

heparin was intravenously injected, and blood was quickly withdrawn via the jugular vein. When the mean arterial blood pressure became 30 mm Hg, both common carotid arteries were clamped with surgical clips. Mean arterial blood pressure was maintained at 30-35 mm Hg for 5 min. After ischemic treatment, the clips were removed, and the blood was reinfused. Body temperature was monitored with a rectal probe and controlled at 37° C. Sham-operated animals underwent exposure of vessels without blood withdrawal or clamping of carotid arteries.

For RNA extraction, the animals were deeply anesthetized, sacrificed with sodium pentobarbital, and then dissected immediately. Tissues were frozen in liquid nitrogen immediately. All animals were treated in accordance with Kyoto University Animal Research Committee guidelines.

For semiquantitative RT-PCR, total RNA was extracted from ischemic hippocampi at two time points, postoperative day 1 and 7 with sham operated control (n=6/time point) with TRIzol Reagent™ (purchased from Life Technologies, Inc), and synthesis of cDNA was performed as above using 1 microgram of total RNA from each sample. Then cDNA was amplified in 50 microliter of PCRs that contained 1.5 mM MgCl₂, 0.2 mM dNTP mixture, 2.5 units of Taq DNA in PCR buffer (Invitrogen), and 0.5 micromolar gene-specific primers using i-Cycler™ (Bio-Rad) between 22 and 40 cycles using SDNSF and GAPDH primers as below.

5'-GCG TCA GGG GGA CGC AGC TGG-3'

(SEQ ID NO.14)

5'-GTC AGC TCC GAT TGC ACA AAT ACT TGA-3'

(SEQ ID NO.15)

5'-TGC ATC CTG CAC CAC CAA CT-3'

(SEQ ID NO.16)

5'-CGC CTG CTT CAC CAC CTT C-3'

(SEQ ID NO.17)

The resultant PCR products were electrophoresed in 1.5% agarose gel and stained with ethidium bromide, the fluorescent bands were scanned, and the volume density of SDNSF was then quantified using NIH Image 1.62. These conditions produced amplicons within the linear exponential phase of the PCR curve. The quantifications of SDNSF were normalized with GAPDH by dividing time point band density by that of its matched PCR.

As shown in FIG. 15, semiquantitative RT-PCR results showed on postoperative days 1 and 7 that the SDNSF transcripts were highly up-regulated. The results indicated that the tissues that have more neurogenesis have more expression of SDNSF transcripts, suggesting that SDNSF may play a role in maintaining the pool of ANSCs within the neural stem cell microenvironment.

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 24

<210> SEQ ID NO 1

<211> LENGTH: 1771

<212> TYPE: DNA

<213> ORGANISM: Rattus rattus

<400> SEQUENCE: 1

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tctgctctgg gccttttttg ttctctgtgc cagggcccag gagcatgggg ctggtgtcca      180
ccatggcagc gtgggccttg acaagagcac agtgcaagc caagagcaca ttatggaaca      240
tctggaaggt gtcacatcaacc agccagaggc ggagatgtcc ccacaggaac tacagctcca      300
ttatttcaaa atgcatgatt acgatggcaa cagtttgctt gatggcttag agctctcgac      360
ggccatcact cacgtgcaca aggaggaggg gactgagcag gtcccaccca tgagcgagga      420
cgagctcacc agcatcatag acggtgtcct gagagacgac gacaagaaca atgacggcta      480
catcgactat gcagagtttg ccaagtcgct gcagtaggcg gcaggccctt tcctgtatgc      540
acacgtgacc cttgctaagt tgatggacat tctggtaatg agaagcagct tatttctgtc      600
tactgctgca gcgctggtaa agcctgtggc agtctgttag actggggtag gaggaagcca      660
caaggaatac ggagagaagt ggggcagtggt caatgtgtgt ttaaacctgt tggacaagag      720
ctcgaacctt ccgaagggtg gtggggtatc tcaagctccc gggaacctga ctctagatgc      780
cactctaact tcttgatggt atttcatgct acctgaaaag taaagacagt ctgctttgcc      840
aagtggagac ttcagtgacg gtggaggag agccaaaagc cgcgatatctt ccagttggg      900
tcctgctctg ggcagatgtg gtcagtatgc tgttcccag gcatacagca tcacgtccta      960

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aagccacagc aggagaagaa tgtcaccac ggagtccacc agacacagag tgaagactcc 1020
ttaccactg gcattttgga agcgaagcac cactggcctg aatacttagc cttttcagat 1080
cttcagtttc cttcacaact actgccacac cctgtgctct gtcatttcag cccgagagaa 1140
acctgaatt ggggtgtgctc tccgctcacc acccaccgtt tgagctccct gaccttgtgt 1200
tttatccttg cccccagggc tcccttcttg gcttatgaac tattaacttg gtatcgcagg 1260
tttaaaactgt cagctgctct agcctaagtc agaccagaaa agatcagtc ttaagggtgg 1320
tggetaacct tatccaagtt ttgaaggaat gtttttaaaa ttacctctt gagcctgaat 1380
atgataattc ttttaatttc agggaagaac agaaaaggaa gagcagtagt agctgaaaga 1440
gaaacagcca taggtcgtagc tttgcgttgt gaaacgtcat agacttactg taaacgaatc 1500
cagaatgatg gtgggatcag aaaaagaaac tgaatcaaat ttgctttacg atgtatagag 1560
acttattttc tttattaag tattcttgta agaaaactta cgtatttgta aaacagtttt 1620
ctgtgtcaag tatttgtgca atcggagctg acttgtaaac tattcttgta agatctcatt 1680
attttgaaag atttatataa tgaactctga ctatctgaca ataaaatgga tgaaaaagta 1740
aaaaaaaaa aaaaaaaaaa aaaaaaaaaa a 1771

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<210> SEQ ID NO 2
<211> LENGTH: 435
<212> TYPE: DNA
<213> ORGANISM: Rattus rattus

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<400> SEQUENCE: 2

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atggcatccc tgcagctgct cagaggtccc ttcctgtgtg ttctgctctg ggccttttgt 60
gttctgtgtg ccagggccca ggagcatggg gctggtgtcc accatggcag cgtgggcctg 120
gacaagagca cagtgcacga ccaagagcac attatggaac atctggaagg tgatcaaac 180
cagccagagg cggagatgct cccacaggaa ctacagctcc attatttcaa aatgcatgat 240
tacgatggca acagtttgct tgatggctta gagctctcga cggccatcac tcacgtgcac 300
aaggaggagg ggagtgagca ggtcccacc atgagcgagg acgagctcat cagcatcata 360
gacggtgtcc tgagagacga cgacaagaac aatgacggct acatcgacta tgcagagttt 420
gccaagtcgc tgcag 435

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<210> SEQ ID NO 3
<211> LENGTH: 1771
<212> TYPE: DNA
<213> ORGANISM: Rattus rattus
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (80)..(514)
<220> FEATURE:
<221> NAME/KEY: sig_peptide
<222> LOCATION: (80)..(157)
<220> FEATURE:
<221> NAME/KEY: mat_peptide
<222> LOCATION: (158)..(514)

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<400> SEQUENCE: 3

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gcgtcagggg gacgcagctg gcaaggttca tccacaagtg ctctcgcgact gcgtcagggg 60
ttatcagggg actggaagc atg gca tcc ctg cag ctg ctc aga ggt ccc ttc 112
Met Ala Ser Leu Gln Leu Leu Arg Gly Pro Phe
-25 -20
ctg tgt gtt ctg ctc tgg gcc ttt tgt gtt cct ggt gcc agg gcc cag 160
Leu Cys Val Leu Leu Trp Ala Phe Cys Val Pro Gly Ala Arg Ala Gln
-15 -10 -5 -1 1

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gag cat ggg gct ggt gtc cac cat ggc agc gtg ggc ctg gac aag agc	208
Glu His Gly Ala Gly Val His His Gly Ser Val Gly Leu Asp Lys Ser	
5 10 15	
aca gtg cac gac caa gag cac att atg gaa cat ctg gaa ggt gtc atc	256
Thr Val His Asp Gln Glu His Ile Met Glu His Leu Glu Gly Val Ile	
20 25 30	
aac cag cca gag gcg gag atg tcc cca cag gaa cta cag ctc cat tat	304
Asn Gln Pro Glu Ala Glu Met Ser Pro Gln Glu Leu Gln Leu His Tyr	
35 40 45	
ttc aaa atg cat gat tac gat ggc aac agt ttg ctt gat ggc tta gag	352
Phe Lys Met His Asp Tyr Asp Gly Asn Ser Leu Leu Asp Gly Leu Glu	
50 55 60 65	
ctc tcg acg gcc atc act cac gtg cac aag gag gag ggg agt gag cag	400
Leu Ser Thr Ala Ile Thr His Val His Lys Glu Glu Gly Ser Glu Gln	
70 75 80	
gtc cca ccc atg agc gag gac gag ctc atc agc atc ata gac ggt gtc	448
Val Pro Pro Met Ser Glu Asp Glu Leu Ile Ser Ile Ile Asp Gly Val	
85 90 95	
ctg aga gac gac gac aag aac aat gac ggc tac atc gac tat gca gag	496
Leu Arg Asp Asp Asp Lys Asn Asn Asp Gly Tyr Ile Asp Tyr Ala Glu	
100 105 110	
ttt gcc aag tcg ctg cag tagggggcag gccctttcct gtatgcacac	544
Phe Ala Lys Ser Leu Gln	
115	
gtgacccttg ctaatgtgat ggacattctg gtaatgagaa gcagcttatt tctgtctact	604
gctgcagcgc tggtaaagcc tgtggcagtc tgttagactg gggtaggagg aagccacaag	664
gaatacggag agaagtgggg cagtgtcaat gtgtgtttaa acctgttgga caagagctcg	724
aaccttccga aggggtggtgg ggtatctcaa gctccccgga acctgactct agatgccact	784
ctaacttctt gatgttattt catgctacct gaaaagtaaa gacagtctgc tttgccaagt	844
ggagacttca gtgacggtgg agggagagcc aaaagccgcy tatcttccca gttgggtcct	904
gctctgggca gatgtggtca gtatgctggt cccagggcat acagcatcac gtcctaaagc	964
cacagcagga gaagaatgtc acccaccggag tccaccagac acagagtgaa gactccttac	1024
ccactggcat tttggaagcg aagcaccact ggcctgaata cttagccttt tcagatcttc	1084
agtttccctc acaactactg ccacacctg tgctctgtca tttcagcccg agagaaacct	1144
tgaattgggt gtgctctccg ctcaccaccc accgtttgag ctocctgacc ttgtgtttaa	1204
tccttctcc cagggctccc ttcttggett atgaactatt aacttggtat cgcaggttta	1264
aactgtcagc tgctctagcc taagtacagc cagaaaagat cagtcattaa ggggtggtggc	1324
taaccttatc caagttttga aggaatggtt ttaaaattac ctctttgagc ctgaatatga	1384
taattctttt aatttcaggg aagaacagaa aaggaagagc agtagtagct gaaagagaaa	1444
cagccatagg tcgtactttg cgttgtgaaa cgtcatagac ttactgtaaa cgaatccaga	1504
atgatggtgg gatcagaaaa agaaaactgaa tcaaatgtgc tttacgatgt atagagactt	1564
atcttcttta ttaaagtatt cttgtaagaa aacttaogta tttgtaaaac agttttctgt	1624
gtcaagtatt tgtgcaatcg gagctgactt gtaaaactatt cttgtaagat ctcattattt	1684
tgaaagattt atataatgaa ctctgactat ctgacaataa aatggatgaa aaagtaaaaa	1744
aaaaaaaaa aaaaaaaaaa aaaaaaa	1771

<210> SEQ ID NO 4

<211> LENGTH: 145

<212> TYPE: PRT

<213> ORGANISM: Rattus rattus

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<400> SEQUENCE: 4

Met Ala Ser Leu Gln Leu Leu Arg Gly Pro Phe Leu Cys Val Leu Leu
 -25 -20 -15

Trp Ala Phe Cys Val Pro Gly Ala Arg Ala Gln Glu His Gly Ala Gly
 -10 -5 -1 1 5

Val His His Gly Ser Val Gly Leu Asp Lys Ser Thr Val His Asp Gln
 10 15 20

Glu His Ile Met Glu His Leu Glu Gly Val Ile Asn Gln Pro Glu Ala
 25 30 35

Glu Met Ser Pro Gln Glu Leu Gln Leu His Tyr Phe Lys Met His Asp
 40 45 50

Tyr Asp Gly Asn Ser Leu Leu Asp Gly Leu Glu Leu Ser Thr Ala Ile
 55 60 65 70

Thr His Val His Lys Glu Glu Gly Ser Glu Gln Val Pro Pro Met Ser
 75 80 85

Glu Asp Glu Leu Ile Ser Ile Ile Asp Gly Val Leu Arg Asp Asp Asp
 90 95 100

Lys Asn Asn Asp Gly Tyr Ile Asp Tyr Ala Glu Phe Ala Lys Ser Leu
 105 110 115

Gln

<210> SEQ ID NO 5

<211> LENGTH: 823

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 5

tggtgaggcc cgaggcgttg gagggcttcg cgtctgcttc ggagaccgta aggatattga 60

tgaccatgag atccctgctc agaaccacct tcctgtgtgg cctgctctgg gccttttgtg 120

ccccaggcgc cagggctgag gagcctgcag ccagcttctc ccaaccggc agcatgggccc 180

tggataagaa cacagtgcac gaccaagagc atatcatgga gcatctagaa ggtgtcatca 240

acaaaccaga ggcggagatg tcgccacaag aattgcagct ccattacttc aaaatgcatg 300

attatgatgg caataatttg cttgatggct tagaactctc cacagccatc actcatgtcc 360

ataaggagga agggagtga caggcaccac taatgagtga agatgaactg attaacataa 420

tagatggtgt tttgagagat gatgacaaga acaatgatgg atacattgac tatgctgaat 480

ttgcaaaatc actgcagtag atgttatttg gccatctcct ggttatatac aaatgtgacc 540

cgtgataatg tgattgaaca ctttagtaat gcaaaataac tcatttccaa ctactgctgc 600

agcatttttg taaaaacctg tagcgattcg ttactctggg gtgagaagag ataagagaaa 660

tgaaagagaa gagaaatggg acatctaata gtcocctaagt gctattaaat accttattgg 720

acaaggaaaa acaacaaaaa aaaatattag tctgtattaa tgctgctgat aaagacgtac 780

ccaagactgg gaagaaaaaa aaaaaaaaaa aaaaaaaaaa aaa 823

<210> SEQ ID NO 6

<211> LENGTH: 438

<212> TYPE: DNA

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 6

atgaccatga gatccctgct cagaaccccc ttctgtgtg gctgctctg ggccttttgt 60

gccccaggcg ccagggtgga ggagcctgca gccagcttct ccaaccggc cagcatgggc 120

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ctggataaga acacagtgca cgaccaagag catatcatgg agcatctaga aggtgtcatc 180
aacaaccag agggcgagat gtcgccacaa gaattgcagc tccattactt caaaatgcat 240
gattatgatg gcaataattt gcttgatggc ttagaactct ccacagccat cactcatgtc 300
cataaggagg aaggagtgga acaggcacca ctaatgagtg aagatgaact gattaacata 360
atagatggtg ttttgagaga tgatgacaag aacaatgatg gatacattga ctatgctgaa 420
ttgcaaaat cactgcag 438

<210> SEQ ID NO 7
<211> LENGTH: 823
<212> TYPE: DNA
<213> ORGANISM: Homo sapiens
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (60)..(497)
<220> FEATURE:
<221> NAME/KEY: sig_peptide
<222> LOCATION: (60)..(137)
<220> FEATURE:
<221> NAME/KEY: mat_peptide
<222> LOCATION: (138)..(497)

<400> SEQUENCE: 7

tggtgaggcc cgaggcgttg gagggtctcg cgtctgcttc ggagaccgta aggatattg 59

atg acc atg aga tcc ctg ctc aga acc ccc ttc ctg tgt ggc ctg ctc 107
Met Thr Met Arg Ser Leu Leu Arg Thr Pro Phe Leu Cys Gly Leu Leu
-25 -20 -15

tgg gcc ttt tgt gcc cca ggc gcc agg gct gag gag cct gca gcc agc 155
Trp Ala Phe Cys Ala Pro Gly Ala Arg Ala Glu Glu Pro Ala Ala Ser
-10 -5 -1 1 5

ttc tcc caa ccc ggc agc atg ggc ctg gat aag aac aca gtg cac gac 203
Phe Ser Gln Pro Gly Ser Met Gly Leu Asp Lys Asn Thr Val His Asp
10 15 20

caa gag cat atc atg gag cat cta gaa ggt gtc atc aac aaa cca gag 251
Gln Glu His Ile Met Glu His Leu Glu Gly Val Ile Asn Lys Pro Glu
25 30 35

gcg gag atg tcg cca caa gaa ttg cag ctc cat tac ttc aaa atg cat 299
Ala Glu Met Ser Pro Gln Glu Leu Gln Leu His Tyr Phe Lys Met His
40 45 50

gat tat gat ggc aat aat ttg ctt gat ggc tta gaa ctc tcc aca gcc 347
Asp Tyr Asp Gly Asn Asn Leu Leu Asp Gly Leu Glu Leu Ser Thr Ala
55 60 65 70

atc act cat gtc cat aag gag gaa ggg agt gaa cag gca cca cta atg 395
Ile Thr His Val His Lys Glu Glu Gly Ser Glu Gln Ala Pro Leu Met
75 80 85

agt gaa gat gaa ctg att aac ata ata gat ggt gtt ttg aga gat gat 443
Ser Glu Asp Glu Leu Ile Asn Ile Ile Asp Gly Val Leu Arg Asp Asp
90 95 100

gac aag aac aat gat gga tac att gac tat gct gaa ttt gca aaa tca 491
Asp Lys Asn Asn Asp Gly Tyr Ile Asp Tyr Ala Glu Phe Ala Lys Ser
105 110 115

ctg cag tagatgttat ttggccatct cctggttata tacaatgtg acccgtgata 547
Leu Gln
120

atgtgattga acactttagt aatgcaaaat aactcatttc caactactgc tgcagcattt 607

tggtaaaaac ctgtagcgat tcgttacact ggggtgagaa gagataagag aatgaaaga 667

gaagagaaat gggacatcta atagtccecta agtgctatta aataccttat tggacaagga 727

aaaacaacaa aaaaaaatat tagtctgtat taatgctgct gataaagacg tacccaagac 787

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tggaagaaa aaaaaaaaaa aaaaaaaaaa aaaaaa 823

<210> SEQ ID NO 8
 <211> LENGTH: 146
 <212> TYPE: PRT
 <213> ORGANISM: Homo sapiens

<400> SEQUENCE: 8

Met Thr Met Arg Ser Leu Leu Arg Thr Pro Phe Leu Cys Gly Leu Leu
 -25 -20 -15

Trp Ala Phe Cys Ala Pro Gly Ala Arg Ala Glu Glu Pro Ala Ala Ser
 -10 -5 -1 1 5

Phe Ser Gln Pro Gly Ser Met Gly Leu Asp Lys Asn Thr Val His Asp
 10 15 20

Gln Glu His Ile Met Glu His Leu Glu Gly Val Ile Asn Lys Pro Glu
 25 30 35

Ala Glu Met Ser Pro Gln Glu Leu Gln Leu His Tyr Phe Lys Met His
 40 45 50

Asp Tyr Asp Gly Asn Asn Leu Leu Asp Gly Leu Glu Leu Ser Thr Ala
 55 60 65 70

Ile Thr His Val His Lys Glu Glu Gly Ser Glu Gln Ala Pro Leu Met
 75 80 85

Ser Glu Asp Glu Leu Ile Asn Ile Ile Asp Gly Val Leu Arg Asp Asp
 90 95 100

Asp Lys Asn Asn Asp Gly Tyr Ile Asp Tyr Ala Glu Phe Ala Lys Ser
 105 110 115

Leu Gln
 120

<210> SEQ ID NO 9
 <211> LENGTH: 1815
 <212> TYPE: DNA
 <213> ORGANISM: Mus musculus

<400> SEQUENCE: 9

gtgcgagaaa aagcgtccca gggacggcag ctggcaaggt tcacgttga gtgcttcgcg 60
 actgcgtcgg ggattatcgg ggtaccacc cggaagcatg gcaaccctac agctgctcag 120
 agctcccttg ctgtgtgtcc tgctttgggt cttttgtgct ccaggtgcca gagcccatga 180
 ccatggggct gatgtccatc atggcagcgt gggcctggat aagagcacag tgcacgacca 240
 agagcacatc atggaacatc tggaggtgt catcgaccag ccagaggcgg agatgtcccc 300
 acaggaactg cagctccatt actcaaaat gcatgattac gacggcaaca gtttgcttga 360
 cggcctagag ctctocatag ccatcactca cgtgcacaag gaggagggga gtgagcaggc 420
 gccagtcatg agcagagatg agctcgtcag catcatagat ggtgtcctga gggacgatga 480
 caagaacaat gacggctaca tcgactacgc tgagtttgca aagtcactgc agtagaccgt 540
 tggtcttttc ctttgtgcac atgtgaccct tgctaatgtg atggacgtgt ctggtaatgc 600
 gaaacaactt atttcgtct actgctcagc actttggtaa gacgctgtgg cagtctgtaa 660
 gagtggggtg aggaagaagc cacatgactg tggagagaag tgggacaggc ctcagtccct 720
 agaggtgtgt ttaagcttgt tgggcaagag ccggatgagg atcttcggaa gggcgggtggg 780
 tatcccgagt tctcaggaat ccgactgtag aatgccactc tgacttcttg atgttaatcc 840
 atgctaccta aagtaaagac aggctgcttg gccaaagtga cacacttgag aaacagtgga 900

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gggagagtgt gaaagccaca cgcttgccct ggttggtcct gtctttaggc agatgtggtc 960
agtattctgt tccccaggca tacagcatca tatattaaag ccacagcaga agaggaatgt 1020
cgcccactga ggccaccag atgcagagtc taggattcct tgcccactgg ccttttgaa 1080
atgaagcacc actggcctga ataattagca tttccagat cttcagatc tccacaact 1140
actgccatac cctgtgtgt atcatttgac caggaggaa acctgaatt ggggtgtgtt 1200
ctctaateac tttccactgt ctgagcttcc ctgaccctg tattgtatcc ttgctcccag 1260
ggctcccttc atggctgtg aactgttaac ttggtatctc aggttaaact gtcagctggt 1320
ctagcctgag cgaggcctga gaccatcagt cactaagagc agtggctaac ctcatcgaag 1380
ttggaaggaa tgtttttaa attacctctt cgagcctgaa tacaagaat aaaagaataa 1440
aagaattctt ttaatttcag ggaagatcag aaaagaaagc ctaaagccct ttagcgttgt 1500
gaacctcagt agtagctgaa agagaagctg ccacaggtt gacttgctct gtgagatgtt 1560
gtagacattc cgtaagagaa tccagaatga tagcaggatc aggaaagaaa tggagccaaa 1620
tctgctctaa ggtgaataga gacttatttt tctttattaa agtattcttg taagacagtt 1680
ttctgtgcca agtatttggt aatcagagc tgacatgtaa gctattcttg taatatctca 1740
ttattttgaa agatttatat aatgaactct ggctatctga caataaaatg gatgaaaag 1800
caaaaaaaaa aaaaa 1815
    
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<210> SEQ ID NO 10
<211> LENGTH: 435
<212> TYPE: DNA
<213> ORGANISM: Mus musculus
    
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<400> SEQUENCE: 10

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atggcaacc tacagctgct cagagctccc ttgctgtgtg tctgctttg ggtctttgt 60
gctccagggt ccagagccca tgaccatggg gctgatgtcc atcatggcag cgtgggcctg 120
gataagagca cagtgcacga ccaagagcac atcatggaac atctggaagg tgtcatcgac 180
cagccagagg cggagatgtc cccacaggaa ctgcagctcc attacttcaa aatgcatgat 240
tacgacggca acagtttctg tgacggccta gagctctcca tagccatcac tcacgtgcac 300
aaggaggagg ggagttagca ggcgccagtc atgagcgagg atgagctcgt cagcatcata 360
gatggtgtcc tgagggacga tgacaagaac aatgacggct acatcgacta cgctgagttt 420
gcaaagtcac tgcag 435
    
```

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<210> SEQ ID NO 11
<211> LENGTH: 1815
<212> TYPE: DNA
<213> ORGANISM: Mus musculus
<220> FEATURE:
<221> NAME/KEY: CDS
<222> LOCATION: (98)..(532)
<220> FEATURE:
<221> NAME/KEY: sig_peptide
<222> LOCATION: (98)..(175)
<220> FEATURE:
<221> NAME/KEY: mat_peptide
<222> LOCATION: (176)..(532)
    
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<400> SEQUENCE: 11

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gtgcgagaaa aagegtccca gggacggcag ctggcaaggt tcacgttga gtgcttcgag 60
actgcgtcgg ggattatcgg ggtaccacc ccgaagc atg gca acc cta cag ctg 115
Met Ala Thr Leu Gln Leu
    
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ctc aga gct ccc ttg ctg tgt gtc ctg ctt tgg gtc ttt tgt gct cca	163
Leu Arg Ala Pro Leu Leu Cys Val Leu Leu Trp Val Phe Cys Ala Pro	
-20 -15 -10 -5	
ggg gcc aga gcc cat gac cat ggg gct gat gtc cat cat ggc agc gtg	211
Gly Ala Arg Ala His Asp His Gly Ala Asp Val His His Gly Ser Val	
-1 1 5 10	
ggc ctg gat aag agc aca gtg cac gac caa gag cac atc atg gaa cat	259
Gly Leu Asp Lys Ser Thr Val His Asp Gln Glu His Ile Met Glu His	
15 20 25	
ctg gaa ggt gtc atc gac cag cca gag gcg gag atg tcc cca cag gaa	307
Leu Glu Gly Val Ile Asp Gln Pro Glu Ala Glu Met Ser Pro Gln Glu	
30 35 40	
ctg cag ctc cat tac ttc aaa atg cat gat tac gac ggc aac agt ttg	355
Leu Gln Leu His Tyr Phe Lys Met His Asp Tyr Asp Gly Asn Ser Leu	
45 50 55 60	
ctt gac ggc cta gag ctc tcc ata gcc atc act cac gtg cac aag gag	403
Leu Asp Gly Leu Glu Leu Ser Ile Ala Ile Thr His Val His Lys Glu	
65 70 75	
gag ggg agt gag cag gcg cca gtc atg agc gag gat gag ctc gtc agc	451
Glu Gly Ser Glu Gln Ala Pro Val Met Ser Glu Asp Glu Leu Val Ser	
80 85 90	
atc ata gat ggt gtc ctg agg gac gat gac aag aac aat gac ggc tac	499
Ile Ile Asp Gly Val Leu Arg Asp Asp Lys Asn Asn Asp Gly Tyr	
95 100 105	
atc gac tac gct gag ttt gca aag tca ctg cag tagaccgttg gctctttcct	552
Ile Asp Tyr Ala Glu Phe Ala Lys Ser Leu Gln	
110 115	
ttgtgcacat gtgacccttg ctaatgtgat ggacgtgtct ggtaatgcca aacaacttat	612
ttccgtctac tgctcagcac tttggtaaga gcctgtggca gtctgtaaga gtggggtgag	672
gaagaagcca catgactgtg gagagaagtg ggacaggcct cagtcacctag aggtgtgttt	732
aagcttgttg ggcaagagcc ggatgcggat cttcggaagg gcggtgggta tcccagattc	792
tcaggaatcc gactgtagaa tgccactctg acttcttgat gttaatccat getacctaaa	852
gtaaagacag gctgcttggc caagtggaca cacttgagaa acagtggagg gagagtgtga	912
aagccacaag cttgccttgg ttggtcctgt ctttaggcag atgtggtcag tattctgttc	972
cccaggcata cagcatcata tattaaagcc acagcagaag aggaatgtcg cccactgagg	1032
ccaccagat gcagagtcta ggattccttg cccactggcc ttttggaat gaagcaccac	1092
tggcctgaat aattagcatt ttccagatct tcagtatctt ccacaactac tgccataccc	1152
tgtgttgtat catttgacca ggagggaaac cttgaattgg ggtgtgttct ctaatcactt	1212
tccactgtct gagctttcct gacccctgta ttgtatcctt gctcccaggg ctcccttcac	1272
ggcttgtgaa ctgttaactt ggtatctcag gttaaactgt cagctggtct agcctgagcg	1332
aggcctgaga ccatcagtca ctaagagcag tggctaacct catcgaagtt ggaaggaatg	1392
tttttaaaat tacctcttcg agcctgaata caaagaataa aagaataaaa gaattctttt	1452
aatttcaggg aagatcagaa aagaaagcct aaagcccttt agcgttgtga acctcagtag	1512
tagctgaaag agaagctgcc acaggttgta cttgctctgt gagatgttgt agacattccg	1572
taagagaatc cagaatgata gcaggatcag gaaagaaatg gagccaaatc tgctetaagg	1632
tgaatagaga cttatttttc tttattaaag tattcttgta agacagtttt ctgtgtcaag	1692
tatttgtgaa atcagagctg acatgtaagc tattcttgta atatctcatt attttgaaag	1752
atztatataa tgaactctgg ctatctgaca ataaaatgga tgaaaaagca aaaaaaaaaa	1812
aaa	1815

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<210> SEQ ID NO 12
<211> LENGTH: 145
<212> TYPE: PRT
<213> ORGANISM: Mus musculus

<400> SEQUENCE: 12

Met Ala Thr Leu Gln Leu Leu Arg Ala Pro Leu Leu Cys Val Leu Leu
  -25                -20                -15

Trp Val Phe Cys Ala Pro Gly Ala Arg Ala His Asp His Gly Ala Asp
 -10                -5                -1 1                5

Val His His Gly Ser Val Gly Leu Asp Lys Ser Thr Val His Asp Gln
      10                15                20

Glu His Ile Met Glu His Leu Glu Gly Val Ile Asp Gln Pro Glu Ala
      25                30                35

Glu Met Ser Pro Gln Glu Leu Gln Leu His Tyr Phe Lys Met His Asp
      40                45                50

Tyr Asp Gly Asn Ser Leu Leu Asp Gly Leu Glu Leu Ser Ile Ala Ile
  55                60                65                70

Thr His Val His Lys Glu Glu Gly Ser Glu Gln Ala Pro Val Met Ser
      75                80                85

Glu Asp Glu Leu Val Ser Ile Ile Asp Gly Val Leu Arg Asp Asp Asp
      90                95                100

Lys Asn Asn Asp Gly Tyr Ile Asp Tyr Ala Glu Phe Ala Lys Ser Leu
      105                110                115

Gln

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<210> SEQ ID NO 13
<211> LENGTH: 38
<212> TYPE: DNA
<213> ORGANISM: Rattus rattus
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (30)..(30)
<223> OTHER INFORMATION: n = a, c, g or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (31)..(31)
<223> OTHER INFORMATION: n = a, c, g or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (32)..(32)
<223> OTHER INFORMATION: n = a, c, g or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (33)..(33)
<223> OTHER INFORMATION: n = a, c, g or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (34)..(34)
<223> OTHER INFORMATION: n = a, c, g or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (35)..(35)
<223> OTHER INFORMATION: n = a, c, g or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (36)..(36)
<223> OTHER INFORMATION: n = a, c, g or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (37)..(37)
<223> OTHER INFORMATION: n = a, c, g or t
<220> FEATURE:
<221> NAME/KEY: misc_feature
<222> LOCATION: (38)..(38)
<223> OTHER INFORMATION: n = a, c, g or t

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<400> SEQUENCE: 13
 tccccgattga attctagacc tgcctcgagn nnnnnnnn 38

<210> SEQ ID NO 14
 <211> LENGTH: 21
 <212> TYPE: DNA
 <213> ORGANISM: Rattus rattus

<400> SEQUENCE: 14
 gcgtcagggg gacgcagctg g 21

<210> SEQ ID NO 15
 <211> LENGTH: 27
 <212> TYPE: DNA
 <213> ORGANISM: Rattus rattus

<400> SEQUENCE: 15
 gtcagctccg attgcacaaa tacttga 27

<210> SEQ ID NO 16
 <211> LENGTH: 20
 <212> TYPE: DNA
 <213> ORGANISM: Rattus rattus

<400> SEQUENCE: 16
 tgcacacctgc accaccaact 20

<210> SEQ ID NO 17
 <211> LENGTH: 19
 <212> TYPE: DNA
 <213> ORGANISM: Rattus rattus

<400> SEQUENCE: 17
 cgctgcttc accaccttc 19

<210> SEQ ID NO 18
 <211> LENGTH: 186
 <212> TYPE: PRT
 <213> ORGANISM: Caenorhabditis elegans

<400> SEQUENCE: 18

Met	Ala	Ala	Asn	Ile	Leu	Val	Val	Ser	Cys	Leu	Ile	Leu	Gly	Ser	Phe
1			5					10						15	
Ala	His	Gln	Pro	Gln	Gln	Phe	Pro	Gly	Ser	Asn	Gln	Gln	Gln	Pro	Gln
			20					25						30	
Gln	Gly	Gly	Gln	Ala	Glu	Gln	Ala	Gln	His	Ala	Gln	Pro	Gly	Gln	Gln
			35				40						45		
Gln	Phe	Gly	Gly	Glu	Gln	Ala	Arg	Asp	Glu	His	His	Ile	Lys	Glu	His
			50			55						60			
Leu	Asp	Gly	Lys	Val	Asp	Pro	Thr	Ala	Asn	Met	Thr	Pro	Glu	Gln	Leu
					70					75					80
Pro	Phe	His	Tyr	Phe	Asn	Met	His	Asp	Leu	Asp	Lys	Asn	Gly	Lys	Leu
				85					90					95	
Asp	Gly	Val	Glu	Leu	Ile	Lys	Ala	Ile	Thr	His	Phe	His	Ala	Glu	Asn
				100				105						110	
Pro	Gly	Pro	Gln	His	Thr	Gln	Asn	Asn	Ala	Asn	Ala	Asn	His	Gln	Pro
				115			120						125		
Pro	Pro	Leu	Pro	Ser	Glu	Val	Glu	Leu	Glu	Thr	Met	Ile	Asp	Ser	Ile
				130			135								140

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Leu Lys Asp Asp Asp Phe Asn Ala Asp Gly Phe Ile Asp Tyr Gly Glu
145 150 155 160

Phe Leu Lys Ala Gln Lys Leu Arg Glu Asp Gln Ala Arg Ser His Gln
165 170 175

Glu Gln Met Gln Lys Ala Gly Gly Thr Gln
180 185

<210> SEQ ID NO 19

<211> LENGTH: 168

<212> TYPE: PRT

<213> ORGANISM: *Drosophila melanogaster*

<400> SEQUENCE: 19

Met Cys Asn Leu Ser Asn Leu Leu Asn Phe Ile Ile Cys Ile Ala Ser
1 5 10 15

Phe Ser Gln Asn Phe Asp Ala Thr Leu Ala Val Lys Arg Gly Pro His
20 25 30

His Pro Arg Gly Glu Thr Arg Arg Val Asp Gln His Leu Thr His Glu
35 40 45

Glu His Arg Ile Asp Asp Asp Leu Lys Asp Met Gly Val Gln Ala Asn
50 55 60

Leu Asp Asp Leu Ser Glu Glu Glu Lys Ile Phe Tyr Met Phe Lys Ala
65 70 75 80

His Asp Asn Asp Asn Asn Asn Ala Leu Asp Gly Leu Glu Met Ile Gln
85 90 95

Ser Ala Met His His Asn Tyr Asp Tyr Phe Lys Asn Asn Glu Arg Asp
100 105 110

Ala Tyr Leu Gln Asn Ala Thr Asp Glu Leu Glu His Phe Ile Glu Ala
115 120 125

Ile Asp Lys Phe Leu Leu Ile Ala Asp Asp Asn Asn Asp Gly Leu Leu
130 135 140

His Tyr Pro Glu Phe Val Lys Ala Ile Thr Gly Gly Lys Glu Gln Pro
145 150 155 160

Asn Val Asp Arg Asn Ile Leu Arg
165

<210> SEQ ID NO 20

<211> LENGTH: 29

<212> TYPE: PRT

<213> ORGANISM: *Rattus rattus*

<400> SEQUENCE: 20

Glu Phe Lys Glu Ala Phe Ala Leu Phe Asp Lys Asp Gly Asp Gly Thr
1 5 10 15

Ile Thr Thr Lys Glu Leu Gly Thr Val Met Arg Ser Leu
20 25

<210> SEQ ID NO 21

<211> LENGTH: 29

<212> TYPE: PRT

<213> ORGANISM: *Rattus rattus*

<400> SEQUENCE: 21

Glu Leu Gln Asp Met Ile Asn Glu Val Asp Ala Asp Gly Asn Gly Thr
1 5 10 15

Ile Asp Phe Pro Glu Phe Leu Ser Leu Met Ala Arg Lys
20 25

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<210> SEQ ID NO 22
<211> LENGTH: 29
<212> TYPE: PRT
<213> ORGANISM: Rattus rattus

<400> SEQUENCE: 22

Glu Leu Ile Glu Ala Phe Lys Val Phe Asp Arg Asp Gly Asn Gly Leu
1             5             10             15

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Ile Ser Ala Ala Glu Leu Arg His Val Met Thr Asn Leu
                20             25

```

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<210> SEQ ID NO 23
<211> LENGTH: 29
<212> TYPE: PRT
<213> ORGANISM: Rattus rattus

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<400> SEQUENCE: 23

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Glu Val Asp Glu Met Ile Arg Glu Ala Asp Ile Asp Gly Asp Gly His
1             5             10             15

```

```

Ile Asn Tyr Glu Phe Val Arg Met Met Val Ala Lys
                20             25

```

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<210> SEQ ID NO 24
<211> LENGTH: 12
<212> TYPE: PRT
<213> ORGANISM: Artificial Sequence
<220> FEATURE:
<223> OTHER INFORMATION: Loop consensus sequence
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (2)..(2)
<223> OTHER INFORMATION: X = any amino acid
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (7)..(7)
<223> OTHER INFORMATION: X = any amino acid
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (10)..(10)
<223> OTHER INFORMATION: X = any amino acid
<220> FEATURE:
<221> NAME/KEY: MISC_FEATURE
<222> LOCATION: (11)..(11)
<223> OTHER INFORMATION: X = any amino acid

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<400> SEQUENCE: 24

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Asp Xaa Asp Gly Asp Gly Xaa Ile Asp Xaa Xaa Glu
1             5             10

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The invention claimed is:

1. An isolated polynucleotide comprising a nucleotide sequence encoding the polypeptide of SEQ ID NO: 4 or the nucleotide sequence complementary to SEQ ID NO: 4.

2. An isolated polynucleotide comprising a nucleotide sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 9, the complement of SEQ ID

50

NO: 1, the complement of SEQ ID NO: 2, and the complement of SEQ ID NO: 9.

3. A replication or expression vector comprising the isolated polynucleotide according to claim 1.

55

4. A host cell transformed with the replication or expression vector according to claim 3.

* * * * *