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<table style="width: 100%; border: none;"> <tr> <td style="width: 50%; vertical-align: top; padding: 5px;"> (21) International Application Number: PCT/CA00/00566 (22) International Filing Date: 8 May 2000 (08.05.00) (30) Priority Data: 2,271,190 6 May 1999 (06.05.99) CA (71) Applicant (for all designated States except US): VASOGEN INC. [CA/CA]; 2155 Dunwin Drive, Suite 10, Mississauga, Ontario L5L 4M1 (CA). (72) Inventor; and (75) Inventor/Applicant (for US only): BOLTON, Anthony, E. [GB/GB]; Ivy Cottage, Sherwood Road, Tideswell, Derbyshire SK17 8HS (GB). (74) Agent: JOACHIM, Roland, H.; Ridout & Maybee, One Queen Street East, Suite 2400, Toronto, Ontario M5C 3B1 (CA). </td> <td style="width: 50%; vertical-align: top; padding: 5px;"> (81) Designated States: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>Without international search report and to be republished upon receipt of that report.</i> </td> </tr> </table>			(21) International Application Number: PCT/CA00/00566 (22) International Filing Date: 8 May 2000 (08.05.00) (30) Priority Data: 2,271,190 6 May 1999 (06.05.99) CA (71) Applicant (for all designated States except US): VASOGEN INC. [CA/CA]; 2155 Dunwin Drive, Suite 10, Mississauga, Ontario L5L 4M1 (CA). (72) Inventor; and (75) Inventor/Applicant (for US only): BOLTON, Anthony, E. [GB/GB]; Ivy Cottage, Sherwood Road, Tideswell, Derbyshire SK17 8HS (GB). (74) Agent: JOACHIM, Roland, H.; Ridout & Maybee, One Queen Street East, Suite 2400, Toronto, Ontario M5C 3B1 (CA).	(81) Designated States: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG). Published <i>Without international search report and to be republished upon receipt of that report.</i>																		
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(54) Title: IMPROVED METHOD FOR TREATING MAMMALS WITH MODIFIED MAMMALIAN BLOOD																						
(57) Abstract An improved method for treating or preventing a pathological condition in a mammalian subject in which modified mammalian blood is administered to the subject. The method comprises administering to the subject from two to four aliquots of modified mammalian blood, with the administration of any pair of consecutive treatments either being on consecutive days or being separated by a rest period of 1 to 21 days. The treatment is useful for treating or preventing a wide range of pathological conditions in which modified mammalian blood is effective, including preconditioning a mammalian subject to better withstand the adverse effects of ischemic stress, atherosclerosis and rheumatoid arthritis.																						
Group F Survival Rate <table border="1" style="margin-top: 10px; border-collapse: collapse; width: 100%;"> <caption>Group F Survival Rate Data</caption> <thead> <tr> <th>Days after surgery</th> <th>Survival Rate (%)</th> </tr> </thead> <tbody> <tr><td>0</td><td>100</td></tr> <tr><td>1</td><td>100</td></tr> <tr><td>2</td><td>100</td></tr> <tr><td>3</td><td>100</td></tr> <tr><td>4</td><td>100</td></tr> <tr><td>5</td><td>100</td></tr> <tr><td>6</td><td>100</td></tr> <tr><td>7</td><td>100</td></tr> <tr><td>8</td><td>100</td></tr> </tbody> </table> Days after surgery			Days after surgery	Survival Rate (%)	0	100	1	100	2	100	3	100	4	100	5	100	6	100	7	100	8	100
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IMPROVED METHOD FOR TREATING MAMMALS WITH MODIFIED MAMMALIAN BLOOD

FIELD OF THE INVENTION

5 This invention relates to the field of medicine and medical treatments. In particular, the invention relates to improved methods for treating the mammalian body, including the human body, by administration thereto of modified mammalian blood.

BACKGROUND OF THE INVENTION

10 Mammalian blood modified by exposure to one or more certain stressors has been found useful for the treatment and prevention of a wide variety of pathological conditions. The stressors to which the blood is exposed are selected from one or more of an oxidative environment, a temperature stressor and ultraviolet (UV) light. The following is a brief discussion of the prior art relating to uses of mammalian blood
15 which has been modified by one or more of the above-mentioned stressors.

 U.S. Patent No. 4,968,483 to Mueller et al. describes an apparatus for oxygenating blood by treating an aliquot of a patient's blood extracorporeally, with an
20 oxygen/ozone mixture and UV light, at a controlled temperature. The apparatus taught by Mueller is proposed for use in hematological oxidation therapy.

 U.S. Patent No. 5,591,457 to Bolton discloses a method of inhibiting the aggregation of blood platelets in a human, a method of stimulating the immune system
25 and a method of treating peripheral vascular diseases such as Raynaud's disease, by extracting an aliquot of blood from a patient, subjecting it to an ozone/oxygen gas mixture and UV light at a temperature in the range of about 37 to 43 °C, and then re-injecting the treated blood in the human patient.

30 U.S. Patent No. 5,834,030 to Bolton describes a similar process for increasing

-2-

5 the content of nitric oxide in the blood of mammalian subject, potentially useful in treating conditions such as high blood pressure in mammalian subjects. Example 5 of this patent discloses a course of treatment comprising ten injections of modified mammalian blood administered over a period of 2 to 4 weeks.

10 International Publication No. WO 98/07436 describes an autoimmune vaccine for administration to human patients to alleviate the symptoms of autoimmune diseases such as rheumatoid arthritis. The vaccine comprises an aliquot of the subject's blood which has been subjected extracorporeally to an oxidizing environment, UV light and elevated temperature. This application discloses a course of treatment comprising from
15 30 to 60 injections of modified mammalian blood.

International Publication No. WO 96/34613 relates to treatment of vascular disorders associated with deficient endothelial function, in a mammalian subject, by administration to the patient of an aliquot of blood which has been modified by having
20 been subjected to at least one stressor selected from elevated temperature in the range of 37° to 55°C, UV light and an oxidative environment. This application discloses a number of different treatment methods. For example, Example 1 discloses ten injections of modified mammalian blood administered over a period of 2 weeks; Example 2 discloses ten injections over a period of 2 to 4 weeks; Example 3 discloses a
25 treatment schedule comprising two courses of treatment, the first course comprising ten injections over a period of 2 to 4 weeks and the second course comprising five injections; and Example 5 discloses administration of five injections at 2 to 3 day intervals.

30 U.S. Patent Application Serial No. 09/190,236, filed November 13, 1998, describes a method for lowering levels of lipids in mammals by injecting a mammalian subject with an aliquot of mammalian blood which has been treated extracorporeally by one or more stressors selected from heat, UV light and oxidative environments. This application describes a study in which animals were subjected to a course of treatment
35 comprising a total of 10 injections over 12 days, with two sets of 5 daily injections

-3-

5 being separated by a rest period of two days.

U.S. Patent Application Serial No. 09/151,653, filed November 9, 1998,
discloses a method for treatment of stress and preconditioning against stress by
injecting a mammalian subject with an aliquot of mammalian blood having been
10 subjected extracorporeally to at least one stressor selected from an oxidative
environment, UV light and elevated temperature up to about 45°C. This application
discloses various treatment methods. Examples 1 and 3 disclose a single course of
treatment comprising ten injections administered over a period of 10 days; and
Examples 4 and 6 to 8 disclose a treatment method comprising two courses of
15 treatment, each comprising ten injections administered over a period of 10 days,
separated by a rest period of about three weeks.

Although the treatments described above have been shown to be useful in the
treatment and prevention of a wide range of pathological conditions, there is a desire to
20 develop a treatment schedule which is less costly to administer and more convenient to
patients, and which either improves or at least does not reduce the effectiveness of the
treatment.

25 SUMMARY OF THE INVENTION

The present invention provides a novel method of treating a mammalian subject
with modified mammalian blood which provides advantages over previously known
methods of treatment, which have typically required subjects to be treated with a
relatively large number of injections of modified blood.

30 The present invention is based upon the discovery that an effective treatment
may be provided in which the number of aliquots of modified blood administered to the
mammalian subject is substantially reduced from that disclosed in the prior art, while
the efficacy of the treatment is significantly enhanced.

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-4-

5 Accordingly, in one aspect, the present invention provides a method of
treatment or prophylaxis of a condition in a mammalian subject in which modified
mammalian blood is administered to said subject, said blood being modified
extracorporeally by exposure to at least one stressor selected from the group consisting
of an oxidative environment, an electromagnetic emission and a temperature above or
10 below body temperature, said method comprising: administering to said subject from
two to four aliquots of said modified mammalian blood, with the administration of any
pair of consecutive aliquots either being on consecutive days or being separated by a
rest period of 1 to 21 days on which no aliquots are administered to the subject; the rest
period preferably being from about 3 to about 15 days.

15 The modified aliquot of blood is preferably prepared by exposing it to one or
more stressors selected from an oxidative stressor, a temperature stressor and an
electromagnetic emission, alone or in combinations of two or three of such stressors,
applied simultaneously or sequentially.

20 BRIEF DESCRIPTION OF THE DRAWINGS

The invention will now be more fully described, by way of example only, with
reference to the accompanying drawings, in which:

25 Figures 1 and 2 are graphical representations of the results obtained from
Example 1 described below;

30 Figures 3 and 4 are graphical representations of the results obtained from
Example 2 described below;

Figures 5 and 6 are graphical representations of the results obtained from
Example 3 described below;

35 Figures 7 and 8 are graphical representations of the results obtained from

-5-

5 Example 4 described below;

Figure 9 is a graphical representation of the results obtained from Example 5 described below; and

10 Figure 10 is a graphical representation of the results of Example 8 described below.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENTS

15 According to a preferred process of the present invention, an aliquot of blood is extracted from a mammalian subject, preferably a human, and the aliquot of blood is treated ex vivo with certain stressors, described in more detail below. The terms “aliquot”, “aliquot of blood” or similar terms used herein include whole blood, separated cellular fractions of the blood including platelets, separated non-cellular
20 fractions of the blood including plasma, plasma components, and combinations thereof. The effect of the stressors is to modify the blood, and/or the cellular or non-cellular fractions thereof, contained in the aliquot. The modified aliquot is then re-introduced into the subject’s body by any suitable method, most preferably intramuscular injection, but also including subcutaneous injection, intraperitoneal injection, intra-arterial
25 injection, intravenous injection and oral, nasal or rectal administration.

The stressors to which the aliquot of blood is subjected ex vivo according to the method of the present invention are selected from temperature stress (blood temperature above or below body temperature), an oxidative environment and an electromagnetic
30 emission, individually or in any combination, simultaneously or sequentially. Suitably, in human patients, the aliquot has a sufficient volume that, when re-introduced into the patient’s body, has the desired effect. Preferably, the volume of the aliquot is up to about 400 ml, preferably from about 0.1 to about 100 ml, more preferably from about 5 to about 15 ml, even more preferably from about 8 to about 12 ml, and most preferably
35 about 10 ml.

-6-

5 It is preferred, according to the invention, to apply all three of the
aforementioned stressors simultaneously to the aliquot under treatment, in order to
ensure the appropriate modification to the blood. It may also be preferred in some
embodiments of the invention to apply any two of the above stressors, for example to
10 apply temperature stress and oxidative stress, temperature stress and an electromagnetic
emission, or an electromagnetic emission and oxidative stress. Care must be taken to
utilize an appropriate level of the stressors to thereby effectively modify the blood to
achieve the desired effect.

15 The temperature stressor warms the aliquot being treated to a temperature above
normal body temperature or cools the aliquot below normal body temperature. The
temperature is selected so that the temperature stressor does not cause excessive
hemolysis in the blood contained in the aliquot and so that, when the treated aliquot is
injected into a subject, the desired effect will be achieved. Preferably, the temperature
stressor is applied so that the temperature of all or a part of the aliquot is up to about
20 55°C, and more preferably in the range of from about -5°C to about 55°C.

In some preferred embodiments of the invention, the temperature of the aliquot
is raised above normal body temperature, such that the mean temperature of the aliquot
does not exceed a temperature of about 55°C, more preferably from about 40°C to
25 about 50°C, even more preferably from about 40°C to about 44°C, and most preferably
about $42.5 \pm 1^\circ\text{C}$.

In other preferred embodiments, the aliquot is cooled below normal body
temperature such that the mean temperature of the aliquot is within the range of from
30 about -5°C to about 36.5°C, even more preferably from about 10°C to about 30°C, and
even more preferably from about 15°C to about 25°C.

The oxidative environment stressor can be the application to the aliquot of solid,
liquid or gaseous oxidizing agents. Preferably, it involves exposing the aliquot to a
35 mixture of medical grade oxygen and ozone gas, most preferably by bubbling through

-7-

the aliquot, at the aforementioned temperature range, a stream of medical grade oxygen gas having ozone as a minor component therein. The ozone content of the gas stream and the flow rate of the gas stream are preferably selected such that the amount of ozone introduced to the blood aliquot, either on its own or in combination with other stressors, does not give rise to excessive levels of cell damage such that the treatment is rendered ineffective. Suitably, the gas stream has an ozone content of up to about 300 $\mu\text{g/ml}$, preferably up to about 100 $\mu\text{g/ml}$, more preferably about 30 $\mu\text{g/ml}$, even more preferably up to about 20 $\mu\text{g/ml}$, particularly preferably from about 10 $\mu\text{g/ml}$ to about 20 $\mu\text{g/ml}$, and most preferably about $14.5 \pm 1.0 \mu\text{g/ml}$. The gas stream is suitably supplied to the aliquot at a rate of up to about 2.0 litres/min, preferably up to about 0.5 litres/min, more preferably up to about 0.4 litres/min, even more preferably up to about 0.33 litres/min, and most preferably about 0.24 ± 0.024 litres/min. The lower limit of the flow rate of the gas stream is preferably not lower than 0.01 litres/min, more preferably not lower than 0.1 litres/min, and even more preferably not lower than 0.2 litres/min.

The electromagnetic emission stressor is suitably applied by irradiating the aliquot under treatment from a source of an electromagnetic emission while the aliquot is maintained at the aforementioned temperature and/or while the oxygen/ozone gaseous mixture is being bubbled through the aliquot. Preferred electromagnetic emissions are selected from photonic radiation, more preferably ultraviolet (UV), visible and infrared light, and even more preferably UV light. The most preferred sources of UV light are UV lamps emitting primarily UV-C band wavelengths, i.e. wavelengths shorter than about 280 nm. Such lamps may also emit amounts of visible and infrared light. Sources of UV light corresponding to standard UV-A (wavelengths from about 315 to about 400 nm) and UV-B (wavelengths from about 280 to about 315) can also be used. For example, an appropriate dosage of such UV light, applied simultaneously with one or both of the aforementioned temperature and oxidative environment stressors, can be obtained from up to eight lamps arranged to surround the sample container holding the aliquot, operated at an intensity to deliver a total UV light energy at the surface of the blood of from about 0.025 to about 10 joules/cm²,

-8-

5 preferably from about 0.1 to about 3.0 joules/cm². Preferably, four such lamps are used.

10 The time for which the aliquot is subjected to the stressors is normally within the time range of up to about 60 minutes. The time depends to some extent upon the chosen intensity of the electromagnetic emission, the temperature, the concentration of the oxidizing agent and the rate at which it is supplied to the aliquot. Some experimentation to establish optimum times may be necessary on the part of the operator, once the other stressor levels have been set. Under most stressor conditions, preferred times will be in the approximate range of from about 2 to about 5 minutes, more preferably about 3 minutes. The starting blood temperature, and the rate at which
15 it can be warmed or cooled to a predetermined temperature, tends to vary from subject to subject. Such a treatment provides a modified blood aliquot which is ready for injection into the subject.

20 In one preferred embodiment of the present invention, the aliquot of blood is stressed by being simultaneously subjected to all three of the above stressors using an apparatus of the type described in aforementioned U.S. Patent No. 4,968,483, issued on November 6, 1990 to Mueller. The aliquot is placed in a suitable, sterile, UV light-transmissive container, which is fitted into the machine. The UV lamps are switched on for a fixed period before the gas flow is applied to the aliquot providing the
25 oxidative stress, to allow the output of the UV lamps to stabilize. The UV lamps are typically on while the temperature of the aliquot is adjusted to the predetermined value, e.g. 42.5 ± 1 °C. Then the oxygen/ozone gas mixture, of known composition and controlled flow rate, is applied to the aliquot, for the predetermined duration of up to about 60 minutes, preferably 2 to 5 minutes and most preferably about 3 minutes as
30 discussed above, so that the aliquot experiences all three stressors simultaneously. In this way, blood is appropriately modified according to the present invention to achieve the desired effects.

35 In the preferred method of the invention, each course of treatment comprises the administration to a mammalian subject of from two to four aliquots of mammalian

-9-

5 blood which has been modified as discussed above.

For optimum effectiveness of the treatment, it is preferred that no more than one aliquot of modified blood be administered to the subject per day. In some preferred embodiments of the invention, at least one rest period is provided during the course of treatment. As used herein, the term "rest period" is defined as the number of days
10 between consecutive aliquots on which no aliquots of modified blood are administered to the subject. The maximum rest period between any two consecutive aliquots during the course of treatment is preferably no greater than about 21 days, and is more preferably from about 3 to about 15 days. .

15 In some preferred embodiments, aliquots of modified blood are administered to the subject on consecutive days, i.e. without an intervening rest period. Some preferred embodiments of the invention (comprising administration of 3 or 4 aliquots) include administration of a pair of aliquots on consecutive days and also provide for a rest
20 period between a pair of consecutive aliquots.

More preferably, a course of treatment comprises administration of two or three aliquots to the subject, the course of treatment optionally including at least one rest period, with the longer (in the case of three aliquots) or the only (in the case of two
25 aliquots) rest period between consecutively administered aliquots being from about 5 to 15 days.

Where the course of treatment comprises the administration of two aliquots to the subject, it is most preferred that the aliquots be administered on consecutive days
30 without an intervening rest period.

Where the course of treatment comprises the administration of three aliquots to the subject, it may be preferred to provide two rest periods, including a longer rest period having a length of 9 to 13 days, more preferably about 10 to 12 days, and most
35 preferably about 11 days; and a shorter rest period of 1 to 3 days.

-10-

5 However, where the course of treatment comprises three injections, it is most preferred
to administer two of the three aliquots on consecutive days, and also to provide a rest
period between two aliquots having a length of 9 to 13 days, more preferably about 10
to 12 days, and most preferably about 11 days. In a particularly preferred example, the
10 first and second aliquots are administered on consecutive days without an intervening
rest period, and the second and third aliquots are separated by a rest period of 11 days.

 Although it may be sufficient to administer only one course of treatment as
described above to the subject, it may be preferred in some circumstances to administer
15 two or more courses of treatment, or to follow the above-described course of treatment
by periodic "booster" treatments, if necessary, to maintain the desired effects of the
present invention. In particular, it may be preferred to administer booster treatments or
to administer a second course of treatments to the subject following a time period of
several weeks or months. For example, it may be preferred to provide a time period of
20 from about 1 to about 4 months between consecutive courses of treatment, with 6
weeks being particularly preferred in some embodiments of the invention.

 In one preferred embodiment, it is preferred to administer a first course of
therapy comprising three injections, with the first and second aliquots being
25 administered on consecutive days without an intervening rest period, and the second
and third aliquots are separated by a rest period of 11 days. Following a period of six
weeks from the end of the first course of therapy, an identical second course of therapy
is administered to the subject.

30 The invention is further illustrated and described with reference to the following
specific examples, comprising animal studies conducted in an approved manner.

 In following Examples 1 to 5, the beneficial effects of the present invention are
demonstrated *in vivo* by clinical experiments on juvenile and adult rats, specifically rats
35 of an inbred strain of genetically hypertensive rats (SHR's). These genetically

-11-

hypertensive rats are the most widely used animal model for hypertension research, and are well known and readily available to researchers in this field. SHR's have several genetic defects, one of the most important being failure to produce appropriate amounts of heat shock proteins when subjected to stress. SHR's develop hypertension rapidly and exhibit exaggerated increases in heart rate, blood pressure and body temperature in response to stress. They represent a model of hypersensitivity to stress. The results obtained using SHR's provide reliable indications of potential results obtainable with human patients.

EXAMPLE 1

Blood from sacrificed SHR's of the same strain as the test animals was collected, treated with sodium citrate anticoagulant and cooled. A portion of the blood was then placed in a sterile container, and subjected simultaneously to UV light, an ozone/oxygen gas oxidative environment and elevated temperature stressors, in an apparatus as generally described in the above-mentioned Mueller patent. More specifically, the blood sample in the sterile, UV-transparent container was heated using infrared lamps to 42.5°C, and while maintained at that temperature, was subjected to UV light emitting UV-C band wavelengths under the preferred conditions previously described. Simultaneously, a gaseous mixture of medical grade oxygen and ozone, the mixture having an ozone content of 13.5 to 15.5 µg/ml, was bubbled through the blood sample at a flow rate of about 240 ml/min. The time of simultaneous UV exposure and gas mixture feed was 3 minutes.

To provide a control, a sterile aqueous physiological saline solution was also prepared.

A total of 60 seven week old SHR's were selected and divided into five groups, Groups A, E, F, G and H, each containing 12 animals. For a period of 10 days (at 7 to 9 weeks of age), each animal of Group A received a daily intragluteal injection of 150 µl of the UV, heat and ozone treated blood, followed by a rest period of 21 days. After the

-12-

5 rest period, the animals were subjected to a second series of 10 daily injections of the modified blood, each injection having a volume of 150 μ l.

10 One day following the second course of injections, the rats of Group A were anesthetized under light gas anesthesia, and the right kidney of each animal was removed through a back incision. An occlusive clip was placed on the remaining renal artery and vein to expose the left kidney to transient ischemia, for 60 minutes. Following the exposure to ischemia, the skin was definitively closed. The animals were then followed with respect to the degree of injury resulting from the ischemia and/or the subsequent reperfusion by determining survival rates and by observing the urine output
15 of the animals, which is a measure of renal function.

The survival rate was monitored by daily count of survivors, and is graphically represented in Figure 1. After eight days, 58% (7 animals) of the sub-group A animals which had received the treated blood injections were surviving. Figure 2 illustrates that
20 5 animals were oligo-anuric, meaning that they produced less than 2 ml of urine in the 24 hour period following surgery.

EXAMPLE 2

25 The twelve animals of Group E were treated as discussed above in Example 1, with the exception that the course of therapy comprised a single course of treatment, each comprising two injections separated by a 5 day rest period (i.e. injection on day 1, followed by a five day rest period, followed by an injection on day 7).

30 Ischemia was induced one day following the final injection as described in Example A in the Group E animals, and the results are shown in Figures 3 and 4. Specifically, Figure 3 shows that after 8 days following surgery, 67% (8 animals) of the Group E animals were surviving, and Figure 4 illustrates that only 4 of the animals of Group E were oligo-anuric.

35

5 EXAMPLE 3

10 The Group F animals were treated as set out in Example 1, with the exception that the course of treatment comprised a total of three injections, with the first two injections being administered on consecutive days, with 11 days rest separating the second and third injections (i.e. injections on days 1 and 2, 11 day rest period, injection on day 14).

15 Ischemia was induced one day following the final injection in the Group F animals as described above in Example 1, with the results being shown in Figures 5 and 6. Figure 5 illustrates a significant increase in survival rate, with 100% of the animals surviving eight days after the surgery. In addition, only 3 of the animals of Group F were shown to be oligo-anuric in the 24 hour period following surgery.

20 EXAMPLE 4

25 The animals of Group G received injections of physiological saline in place of modified blood, following an injection schedule identical to that in Example 1. Ischemia was induced one day following the final injection as described above in Example 1, and the results are shown in Figures 7 and 8. Only 42% (5 animals) of the Group G animals were surviving 8 days after the surgery, and 7 of the animals were shown to be oligo-anuric in the 24 hour period following surgery.

30 EXAMPLE 5

35 The animals of Group H were treated with a single course of treatment, comprising two injections separated by a 12 day rest period (i.e. injection on day 1, 12 day rest period, injection on day 14). The results of the ischemic stress on the animals of Group H is illustrated in Figure 9, which illustrates that 67% (8 animals) of the animals in Group H were surviving eight days after the surgery. Also, 3 of the animals in Group H were oligo-anuric in the 24 hour period following surgery.

The results of the above animal studies are tabulated below.

TABLE I

Group	# of Courses	# of Injections	Rest Period (Days)	Survival Rate After 8 Days	# of Anuric Animals
A	2	10 + 10	21	58%	5
E	1	1 + 1	5	67%	4
F	1	2 + 1	11	100%	3
G*	2	10 + 10	21	42%	7
H	1	1 + 1	12	67%	3

* Saline Control

The above results demonstrate that the method of the present invention protects the kidney from injury resulting from ischemia and/or reperfusion, as measured by survival rate and urine flow in the 24 hours following surgery in the SHR. This indicates use of the procedure in protecting other body organs from ischemia/reperfusion injury, including the heart, the liver, the brain, the spinal cord and other vital organs, and indicates practical application of the procedure on patients scheduled to undergo surgical procedures involving ischemia/reperfusion of a body organ, such as surgical repair, removal or transplantation of a body organ.

In particular, ischemic acute renal failure is an important clinical problem with high morbidity and high mortality. The process of the present invention presents a novel approach to combatting this disorder. It can be adopted prior to kidney transplantation, on either or both the donor or recipient. It can be adopted prior to kidney revascularization. It can be adopted prior to invasive evaluation in high risk subjects, e.g. angiography in diabetics. It can be adopted prior to abdominal aortic surgery such as aortic aneurism repair and renal bench surgery (i.e. where the kidney is

-15-

5 temporarily removed and operated on ex vivo, and then re-implanted).

As regards its use in connection with procedures involving the heart, the procedure of the invention can be conducted prior to coronary angioplasty, and bypass, or prior to transplantation, as in the case of the kidney. It is indicated for use with
10 patients about to undergo open heart surgery with cardio-pulmonary bypass for coronary artery bypass grafting, valve replacement or surgical repair of congenital or acquired cardiac structural abnormalities. In the case of the brain or other vital organs and tissues including the intestines, the kidneys and limbs, the procedure of the invention can be used prior to angioplasty or endarterectomy, in high risk subjects.

15 In connection with body organs destined for transplantation, treatment of the donor body by the process of the present invention serves to protect body organs against damage resulting from the inevitable ischemia which the organ will suffer on removal from the donor body, transportation and subsequent surgical introduction into the
20 recipient body. The treatment according to the invention extends the useful life of the transplant organ between its removal from the donor body and its surgical introduction into the recipient body, thereby reducing losses of viable transplant organs due to transportation delays.

25 The use of the process of the present invention prior to general anaesthesia in connection with major surgery can be viewed as general pre-conditioning of the body, to better withstand ischemia-reperfusion injuries to which the major organs will later be subjected. It is indicated for use prior to conducting major surgical procedures involving general anaesthesia in patients known to have or likely to have a significant
30 degree of underlying atherosclerosis in the arteries supplying the brain, heart, liver, intestine, spinal cord, kidneys or limbs, the atherosclerosis rendering them more susceptible to a thrombo-ischemic event in the operative or post-operative period. In addition, it is known that repetitive mild ischemic (anginal) episodes can render tissues and organs less susceptible to stress-induced damage, by ischemic preconditioning,
35 although application of ischemic preconditioning by current methods is largely

-16-

5 impractical. The process of the present invention can take the place of ischemic preconditioning, ischemia being a species of physical stress. Accordingly, the process of the present invention offers potential for treatment of unstable angina and decrease of infarct size, a treatment not effectively addressed by available therapies.

10 A further, specific clinical application of the process of the invention is in treatment of patients suffering from transient ischemic attacks (TIA's, pre-strokes), which are due to temporary obstruction of blood flow to certain areas of the brain. They commonly indicate the likelihood of suffering a major stroke in the near future. Subjection of such patients to treatment according to the process of the invention, at the
15 onset of TIA's, will precondition the brain to avoid or at least to lessen the severity of the effects of the forthcoming major stroke.

Similar general pre-conditioning of the body by the process of the invention is also indicated for use in alleviating the effects of subsequently encountered shock,
20 leading to under-perfusion of vital organs and tissues through failure of cardiac action due to loss of blood or other body fluids, excessive dilation of blood vessels and excessively low blood pressure. Examples include major blood loss, trauma, sepsis and cardiogenic shock. Individuals likely to be exposed to such hazards, including patients awaiting surgery, rescue and relief crews for natural disasters, would be beneficiaries of
25 the process of the invention.

It now appears that common physiological events underlie all stress responses, including responses to ischemic stresses. These physiological events include the induction and upregulation or synthesis, in all body cells, of a group of specialized
30 intracellular proteins known as heat stress proteins or heat shock proteins (HSP's). These HSPs function to protect the cells from potential damage caused by whatever form of stress is being applied. Therefore, it is expected that the method of the present invention is of general application and will be effective to provide a subject with resistance to a wide variety of different types of stresses, whether evident at the time the
35 treatment is administered, or whether subsequently encountered. For example,

-17-

5 psychological stresses induced by restraint, confinement, sudden exposure to danger, shock and the like translate into physical stresses affecting one or more organs of the body. Similarly, physical stress such as exposure to heat or cold, over-exertion and the like, result in abnormal functioning of body organs.

10 A subject who has undergone a treatment according to the method of the present invention will exhibit notably reduced adverse reactions to subsequently encountered stress, such as a notably reduced rise in body temperature, a reduced increase in heart rate and/or a reduced increase in diastolic blood pressure in response to stress, as compared to a similar but untreated subject.

15 In addition, it is expected that the improved results obtained by using the process of the invention would also be applicable to other conditions which may be treated by administration to a subject of modified mammalian blood, such as those discussed above with reference to the prior art. Specific examples of such conditions
20 are diseases of the circulatory system caused by atherosclerosis, for example coronary artery disease, cerebrovascular disease, peripheral vascular disease and diseases of other vascular systems; vasospastic disorders including primary and secondary Raynaud's disease, cardiac syndrome X (microvascular angina), migraine headache, cluster headache, hypertension, pre-eclampsia, and thrombotic disorders related to increased
25 platelet aggregation and/or coagulation abnormalities and/or related endothelial dysfunction; immune system disorders such as rheumatoid arthritis, asthma, graft-versus-host disease, diabetes mellitus, organ rejection, miscarriage, systemic lupus erythematosus, atopic allergy, multiple sclerosis, allergic dermatitis, inflammatory bowel disease and psoriasis.

30 EXAMPLE 6

Model:

The purpose of the experiment is to determine the effects of treatment according
the present invention on the development of atherosclerosis in the LDL receptor (LDL-
35 R) deficient mouse model, a widely used transgenic atherosclerosis model created by

-18-

5 targeted disruption of the LDL receptor. This animal model is analogous to familial
hypercholesterolemia, an inherited condition in which a mutation results in complete
lack of functional LDL-R. In the human disease, homozygous individuals demonstrate
a marked increase in serum cholesterol and develop severe premature atherosclerosis,
often succumbing to this disease at an early age. In patients with this disease, currently
10 used lipid lowering agents do not have a significant effect in terms of lowering
cholesterol levels.

The LDL-R deficient mouse model shows intolerance to cholesterol feeding and
develops widespread atherosclerotic changes which progress to mature fibrous lesions
15 morphologically indistinguishable from established human atherosclerosis. Apart from
the defined genetic abnormality causing predisposition to atherosclerosis, this model
has the advantage of rapid development of widespread atherosclerosis within 6 to 8
weeks following institution of cholesterol feeding.

20 Protocol:

LDL-R deficient mice were purchased from Jackson Laboratories. The mice
were entered into the study at 22 weeks of age. The mice were maintained on a 12 hour
dark/12 hour light cycle with free access to food and water, and were fed a specified
diet as follows.

25 Group A (control) - fed a normal diet;

Group B1 - fed a high cholesterol diet as described below for 8 weeks;

30 Group B2 - fed a high cholesterol diet as described below for 12 weeks;

Group C1 - fed a high cholesterol diet as described below for 8 weeks, and treated by
the preferred method of the present invention as described below at 4 weeks of dietary
intervention; and

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-19-

5 Group C2 - fed a high cholesterol diet as described below for 12 weeks. and treated by the preferred method of the present invention as described below at 8 weeks of dietary intervention.

10 The high cholesterol group were fed a diet containing 1.25% cholesterol, 7.5% cocoa butter, 7.5% casein, and 0.5% sodium cholate. To ensure proper food intake, food consumption and animal weight were monitored on a weekly basis. In previous experiments, it was demonstrated that 8 weeks of feeding with the high cholesterol diet results in substantial atherosclerosis development, particularly in the aortic arch and the descending thoracic aorta.

15 Treatment:

Animals fed the high cholesterol diet were selected at random to undergo a course of treatment by the preferred method of the invention. The treatments began four or eight weeks after initiation of the study, with each of the animals on the high cholesterol diet receiving a total of 3 treatments (injections on days 1, 2 and 14 of the treatment). Each individual injection administered to the animals treated by the method of the present invention consisted of the collection of 10 ml of blood from genetically compatible donor animals fed on a normal diet, the blood being collected into sodium citrate anticoagulant. In order to collect each 10 ml aliquot of blood, about 1 ml of blood was extracted from each of 10 animals. The blood was extracted by cardiac puncture, with the animals being under full xylazine/ketamine anesthesia during the blood extraction procedure, and being given T-61 immediately following extraction. The blood aliquot was transferred to a sterile, disposable, low-density polyethylene vessel for ex vivo treatment, and was then treated simultaneously with a gaseous oxygen/ozone mixture and ultraviolet light at elevated temperature using an apparatus as generally described in aforementioned U.S. Patent No. 4,968,483 to Mueller et al.

35 The constitution of the gas mixture was $14.5 \pm 1.0 \mu\text{g}$ ozone/ml, with the remainder of the mixture comprising medical grade oxygen. The gas mixture was bubbled through the aliquot at a rate of 240 ± 24 ml/min for a period of 3 minutes. The

-20-

5 temperature of the aliquot was held steady at $42.5 \pm 1.0^{\circ}\text{C}$. The UV light was within the UV-C band, and included a wavelength of 253.7 nm.

10 After treatment by the preferred method of the present invention, 30 μl of the treated blood was re-injected intramuscularly into each animal undergoing treatment according to the present invention. As discussed above, 3 treatments were administered to each animal over a 14 day period, with injections being administered on days 1, 2 and 14.

15 In the sham treatments, 30 μl of untreated blood was injected intramuscularly into each of the remaining five animals on the high cholesterol diet.

Assessment of Atherosclerosis:

20 After 8 or 12 weeks, the animals were anesthetized with zylaxine/ketamine and the heart was exposed. After nicking the vena cava to obtain blood samples, the animals were perfused via ventricular puncture, first with PBS to flush out the blood and then with 10% neutral buffered formalin for 3 minutes to fix the aorta. The thoracic aorta was dissected away from the thorax en bloc and stored in 10% formalin at 4°C . Pressure-fixed (10% formalin) aortae were removed en bloc and opened to allow a longitudinal full length inversion. The aortae were then mounted internally
25 exposed on glass slides and stained with oil red O. The bright red staining (indicating lipid deposition) was then quantified using a computer assisted morphometric system, and expressed as a percentage of total aortic intimal surface.

Statistical Analysis:

30 Continuous variables are reported as mean $\pm$ SD. Differences in atherosclerotic lesion area among groups were tested using the one-way ANOVA test in conjunction with the Bonferroni correction.

Results:

35 As demonstrated by measurement of atherosclerotic area, the animals of group

-21-

5 B (high cholesterol diet alone) exhibited substantial aortic lipid deposition, with group
B1 animals having atherosclerosis ratios (AA/TA) of 0.16 ± 0.1 at eight weeks and
group B2 animals having ratios of 0.17 ± 0.1 at 12 weeks of dietary intervention. In
contrast, the animals of group C (high cholesterol diet with treatment according to the
invention) exhibited profoundly reduced lipid deposition, with group C1 animals
10 having ratios of 0.04 ± 0.02 ($p < 0.05$) at eight weeks of dietary intervention, and group
C2 animals having ratios of 0.04 ± 0.02 ($p < 0.01$) at twelve weeks of dietary
intervention.

15 The animals of group C also exhibited a marked reduction in xanthelasma and
limb swelling as compared to animals of group B.

As shown in Example 6, the treatment according to the present invention
substantially inhibited the development of atherosclerosis in a mouse model of familial
hypercholesterolemia. These improvements in cardiovascular health were accompanied
20 by improvements in the animals' general overall appearance and appetite.

EXAMPLE 7

25 In this example, LDL-R deficient mice were fed a high cholesterol diet as in
Example 6 for a period of 12 weeks and divided into two treatment groups, Group I and
Group II. All animals of both groups received a first course of treatment comprising
injections on days 1, 2 and 14 as described above in Example 6. In addition, the
animals of Group II received a second, identical course of treatment 6 weeks after the
end of the first course of treatment.

30 All animals were sacrificed 24 weeks after initiation of the high cholesterol diet.
Calculation of percent plaque area by en face examination of oil red O stained aortas
revealed a 38% reduction in plaque area in Group II animals ($0.32 \pm 0.03\%$ for sham
versus $0.20 \pm 0.03\%$ for treated, $p = 0.016$) but no reduction in plaque area in the Group
35 I animals.

5 The above example indicates that in some embodiments of the invention it may be preferred to administer two or more courses of therapy at spaced intervals.

EXAMPLE 8

10 This example demonstrates the beneficial effects of the present invention in in vivo clinical experiments on rats, specifically male Lewis rats in which rheumatoid-like arthritis has been induced. An animal model used for studying rheumatoid arthritis is adjuvant-induced arthritis in a rat model (see, for example, Pearson, C., 1956, "Development of Arthritis, peri-arthritis and periostitis in rats given adjuvant", Proc. Soc. Exp. Biol. Med., 91:95). According to this model, arthritis is induced in rats by
15 injecting them with adjuvant containing Mycobacterium butyricum.

 Male Lewis rats, 4 to 5 weeks of age, 100 to 120 g, were obtained from Charles River Laboratories, quarantined one week and entered into the study. An adjuvant
20 mixture was prepared for induction of arthritis by suspending 50 mg M. butyricum (Difco Laboratories, Inc., Detroit MI) in 5 ml light white paraffin oil - m3516 (Sigma Chemical Co., St. Louis, MO) and thoroughly mixed using a homogenizer.

 Aliquots of the mixture sufficient to supply 0.15 mg M. butyricum were injected
25 into each animal subcutaneously, at the base of the tail. Symptoms of arthritis appeared about 12 days after induction, in each animal, as evidenced by limb swelling.

 Blood was collected from donor animals, by cardiac puncture. 10 ml of citrated donor blood was transferred to a sterile, low density polyethylene vessel for ex vivo
30 treatment with stressors according to the invention. Using an apparatus as described in the above-mentioned Mueller patent (more specifically, a VasoCare™ V7000 apparatus), the blood was heated to $42.5 \pm 1^\circ\text{C}$ and at that temperature irradiated with ultraviolet light, principally at a wavelength of 254 nm, while a gas mixture of medical grade oxygen containing $14.5 \pm 1.0 \mu\text{g/ml}$ of ozone was bubbled through the blood at a
35 flow rate of $240 \pm 24 \text{ ml/min}$.

-23-

5 Six animals were given a course of 2 injections of 0.2 ml aliquots of the treated
blood, on days 1 and 2 after disease induction. Eighteen animals were injected with
untreated blood or saline as controls. Hind paw volumes and clinical scores of the
animals were measured every 2 to 3 days for 5 weeks after disease induction. Hind paw
volumes were measured by water displacement in a 250 ml beaker using a top-loaded
10 Mettler balance.

 The results of this study showed that the incidence of arthritis in the treated
group was decreased as compared with the control group, as measured both by clinical
scores and foot volumes. The results of the foot volume measurements for each group
15 of animals were averaged and are presented graphically on the accompanying Figure
10, a plot of mean foot volume against days after induction of arthritis. The upper
curve is derived from the control group of animals, the lower curve from the animals
which received the course of injections of treated blood. A significant decrease in the
severity of the arthritis, as indicated by lower foot volumes, is apparent for the treated
20 animals as compared to the animals of the control group.

 Although the invention has been described in connection with certain preferred
embodiments, it is to be appreciated that it is not limited thereto. Rather, the present
invention includes within its scope all embodiments which may fall within the scope of
25 the following claims.

5

What is claimed is:

10

1. A method of treatment or prophylaxis of a condition in a mammalian subject in which modified mammalian blood is administered to said subject, said blood being modified extracorporeally by exposure to at least one stressor selected from the group consisting of an oxidative environment, an electromagnetic emission and a temperature above or below body temperature, said method comprising:

15

administering to said subject from two to four aliquots of said modified mammalian blood, with the administration of any pair of consecutive aliquots either being on consecutive days or being separated by a rest period of 1 to 21 days on which no aliquots are administered to the subject.

20

2. The method according to claim 1, comprising the administration to said subject of from two to four aliquots of said modified mammalian blood, said method including a rest period of from about 3 to about 15 days separating a selected pair of consecutive aliquots.

25

3. The method according to claim 2, wherein two or three aliquots of the modified mammalian blood are administered to the subject, with a selected two consecutive aliquots being separated by a rest period of from about 5 to about 15 days.

30

4. The method according to claim 3, wherein a total of two aliquots of the modified mammalian blood are administered to the subject, the administration of the two aliquots being separated by a rest period of from about 5 to about 12 days.

5. The method according to claim 3, wherein a total of three aliquots of the modified mammalian blood are administered to the subject.

35

6. The method according to claim 3, wherein a total of three aliquots of the

-25-

5 modified mammalian blood are administered to the subject, with a selected two consecutive aliquots being separated by a rest period of from about 10 to about 12 days.

7. The method according to claim 6, wherein two of said aliquots are administered on consecutive days.

10 8. The method according to claim 7, wherein the three aliquots comprise first, second and third aliquots, with the first and second aliquots being administered on consecutive days and a rest period of 11 days being provided between the second and third aliquots.

15 9. The method according to claim 1, wherein said method comprises the administration of two aliquots of said mammalian blood to the subject, the aliquots being administered on consecutive days.

20 10. The method according to claim 1, wherein said method is used to precondition the mammalian subject to better withstand the adverse effects of ischemic stress to be encountered as a result of subsequent ischemia-reperfusion of a body organ of the subject.

25 11. The method according to claim 10, wherein the ischemic stress is ischemia-reperfusion of the subject's kidney, heart, liver, intestine or brain.

12. The method according to claim 11, wherein the ischemic stress is ischemia-reperfusion of the kidney.

30 13. The method according to claim 10, wherein the ischemic stress is encountered within 5 days following the administration of the last one of said aliquots.

35 14. The method of according to claim 13, wherein the ischemic stress is encountered on the day following the administration of the last one of said aliquots.

-26-

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15. The method of claim 1, wherein the condition is atherosclerosis.

16. The method of claim 1, wherein the condition is rheumatoid arthritis.

10

17. A method of treatment or prophylaxis of a condition in a mammalian subject in which modified mammalian blood is administered to said subject, said blood being modified extracorporeally by exposure to at least one stressor selected from the group consisting of an oxidative environment, an electromagnetic emission and a temperature above or below body temperature, said method comprising two or more consecutive

15

courses of treatment, each course of treatment comprising:

administering to said subject from two to four aliquots of said modified mammalian blood, with the administration of any pair of consecutive aliquots either being on consecutive days or being separated by a rest period of 1 to 21 days on which

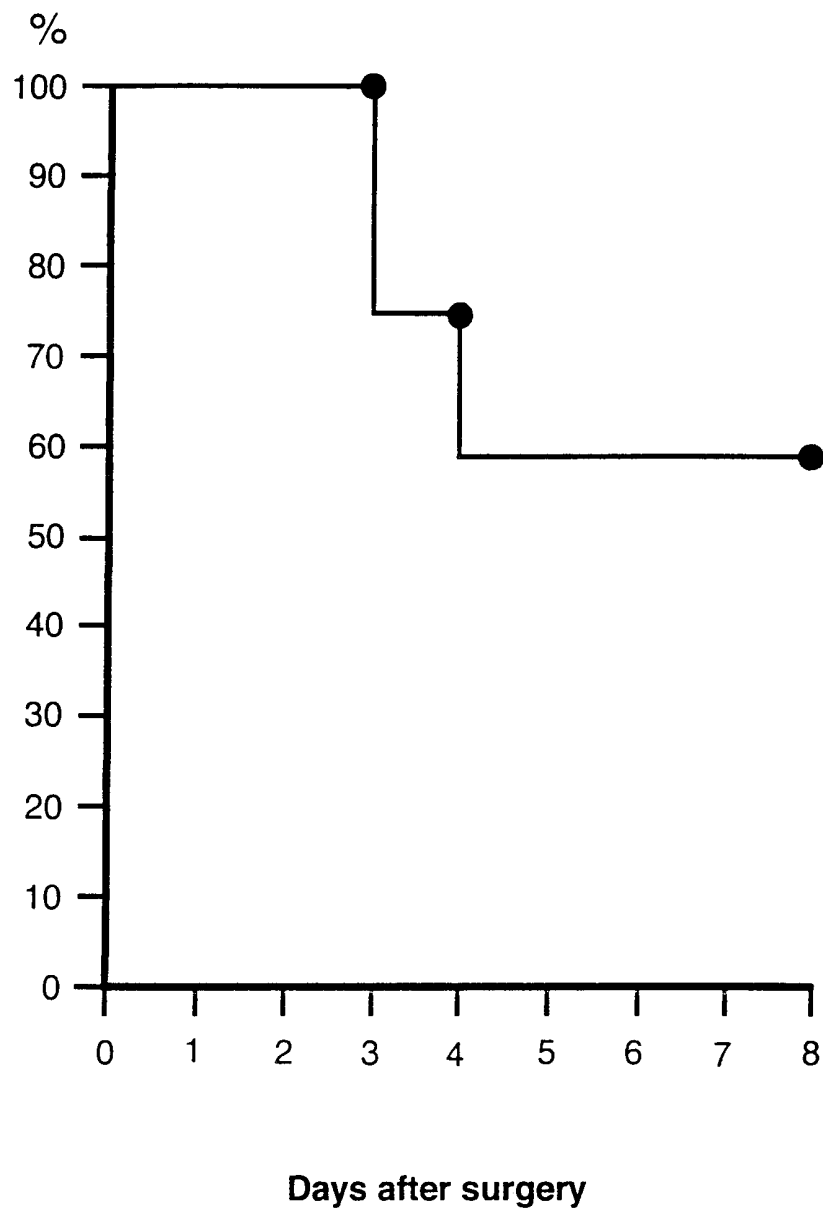
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no aliquots are administered to the subject.

18. The method according to claim 17, wherein said consecutive courses of treatment are separated from one another by a time period of from about 1 to about 4 months.

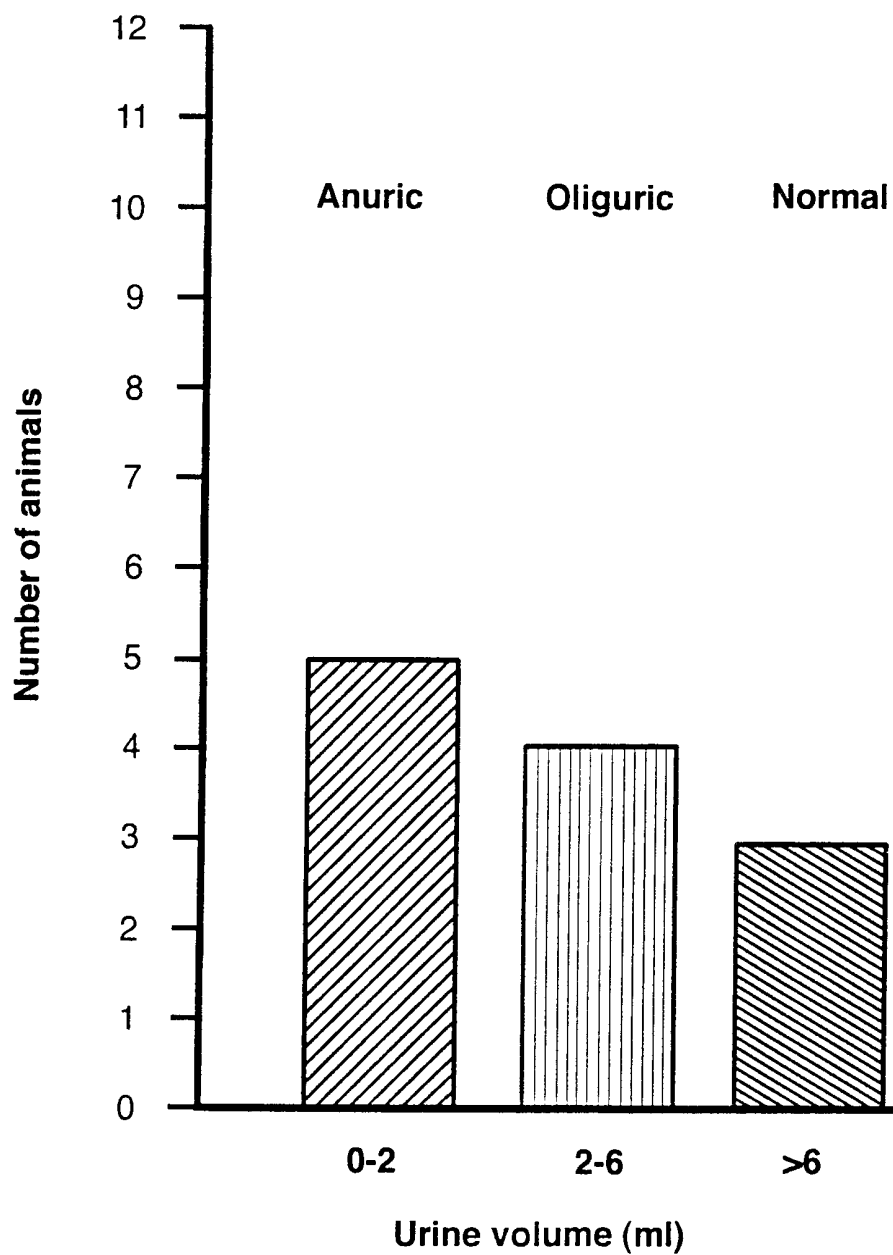
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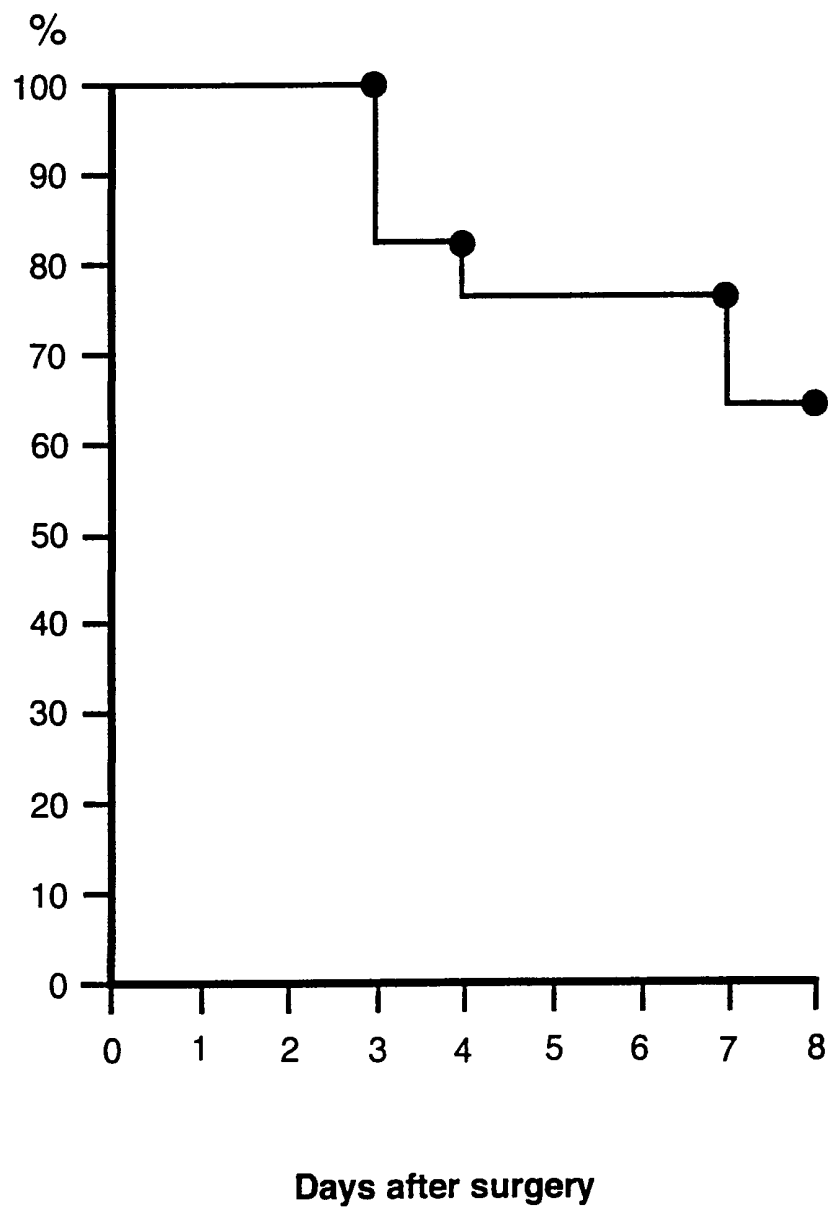
19. The method according to claim 18, wherein the time period is about 6 weeks.

1/10
Fig.1**Group A Survival Rate**

2/10 Fig.2

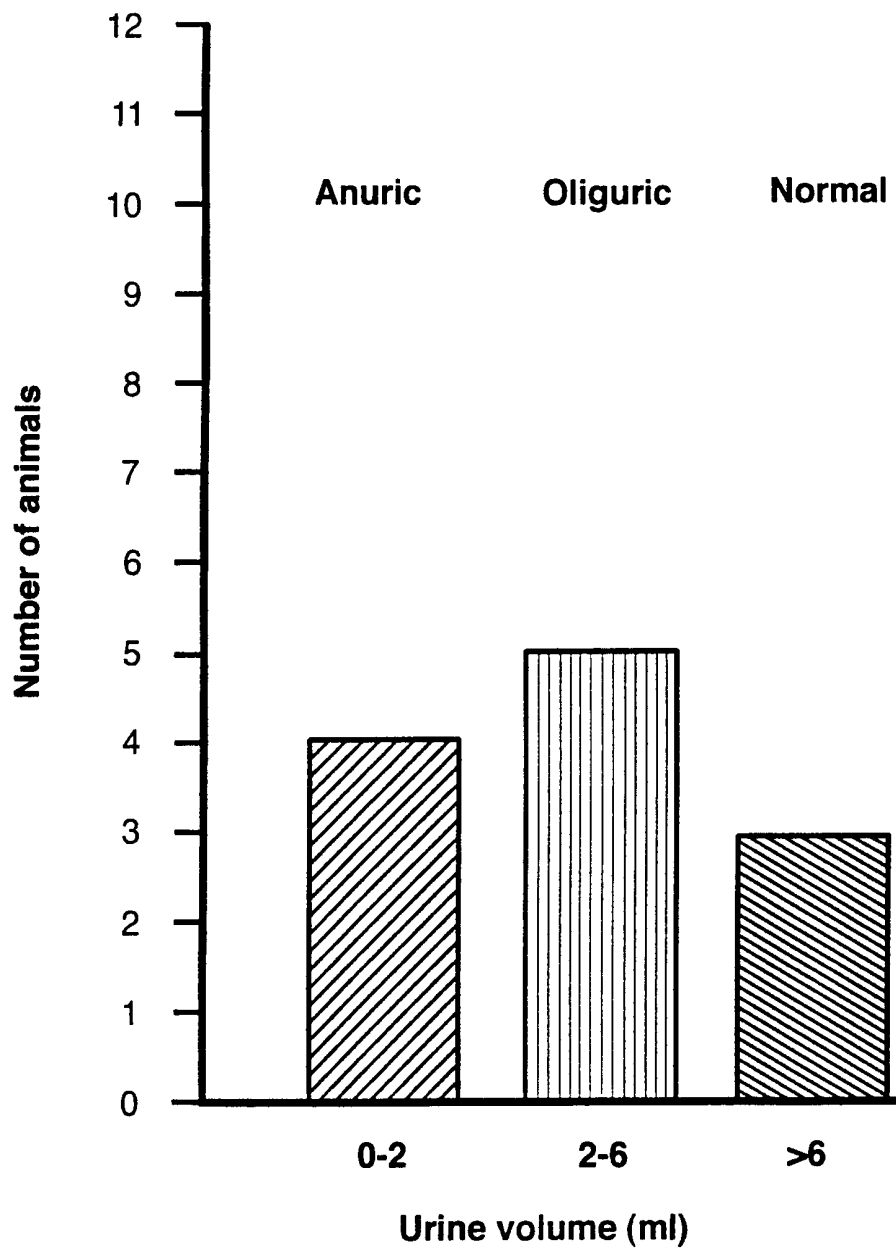
**Group A 24h Diuresis after
renal ischemia/reperfusion injury**

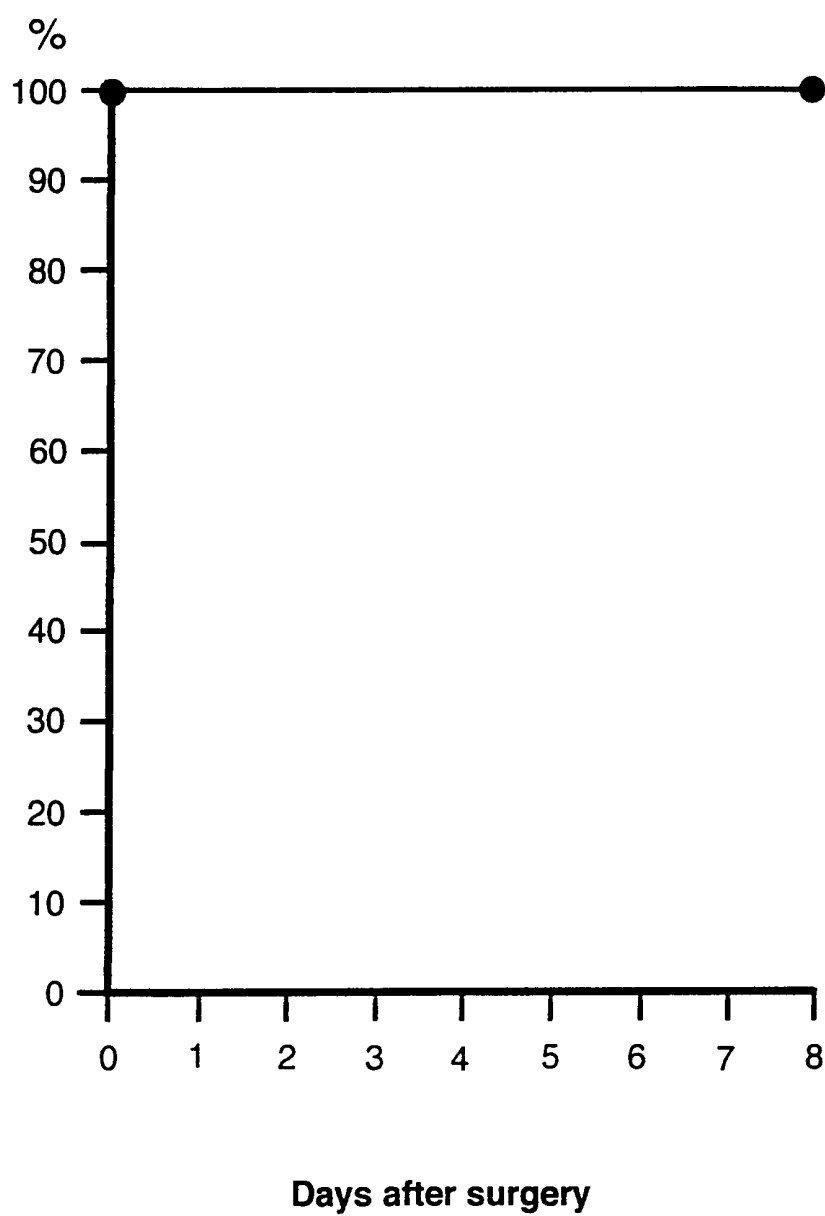


3/10
Fig.3**Group E Survival Rate**

4/10
Fig.4

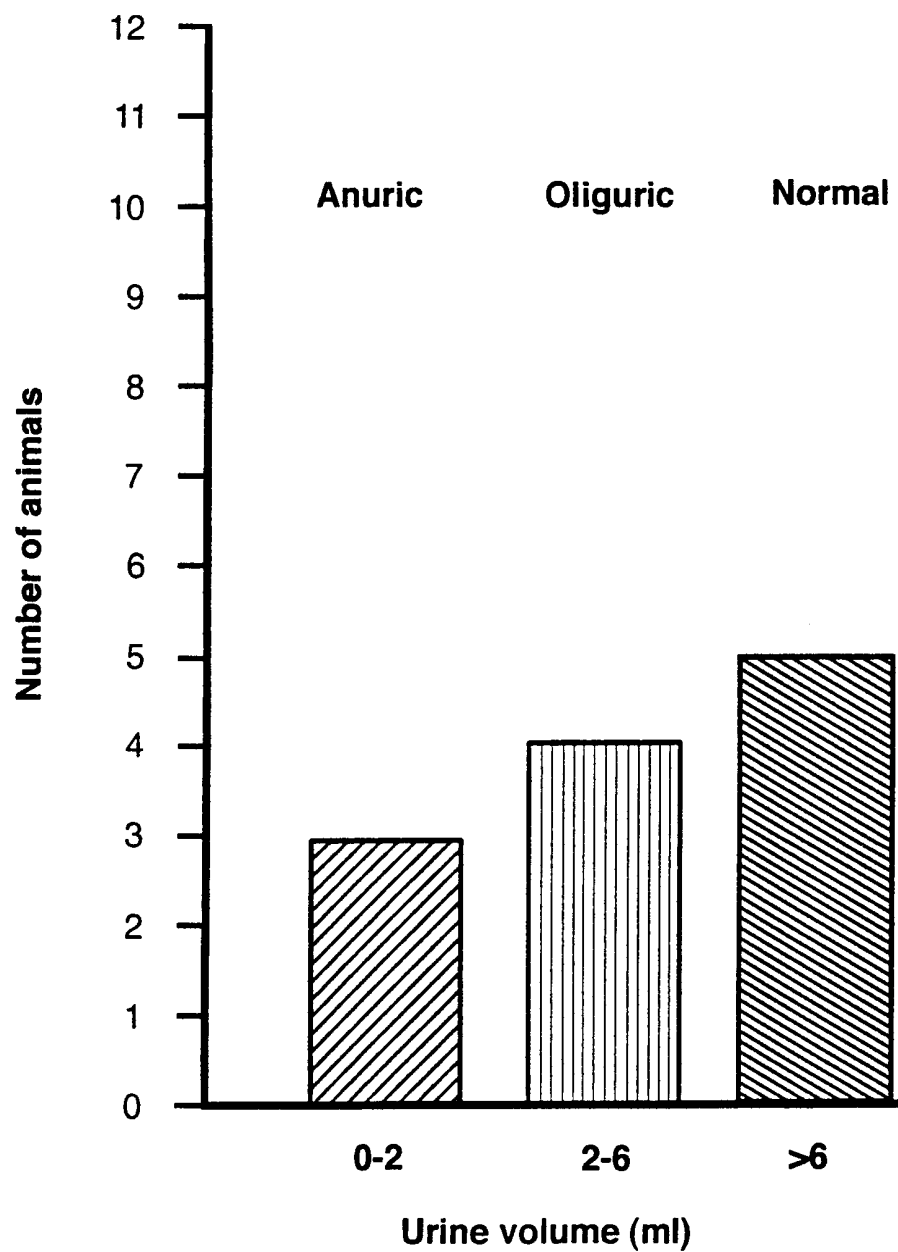
**Group E 24h Diuresis after
renal ischemia/reperfusion injury**



5/10
Fig.5**Group F Survival Rate**

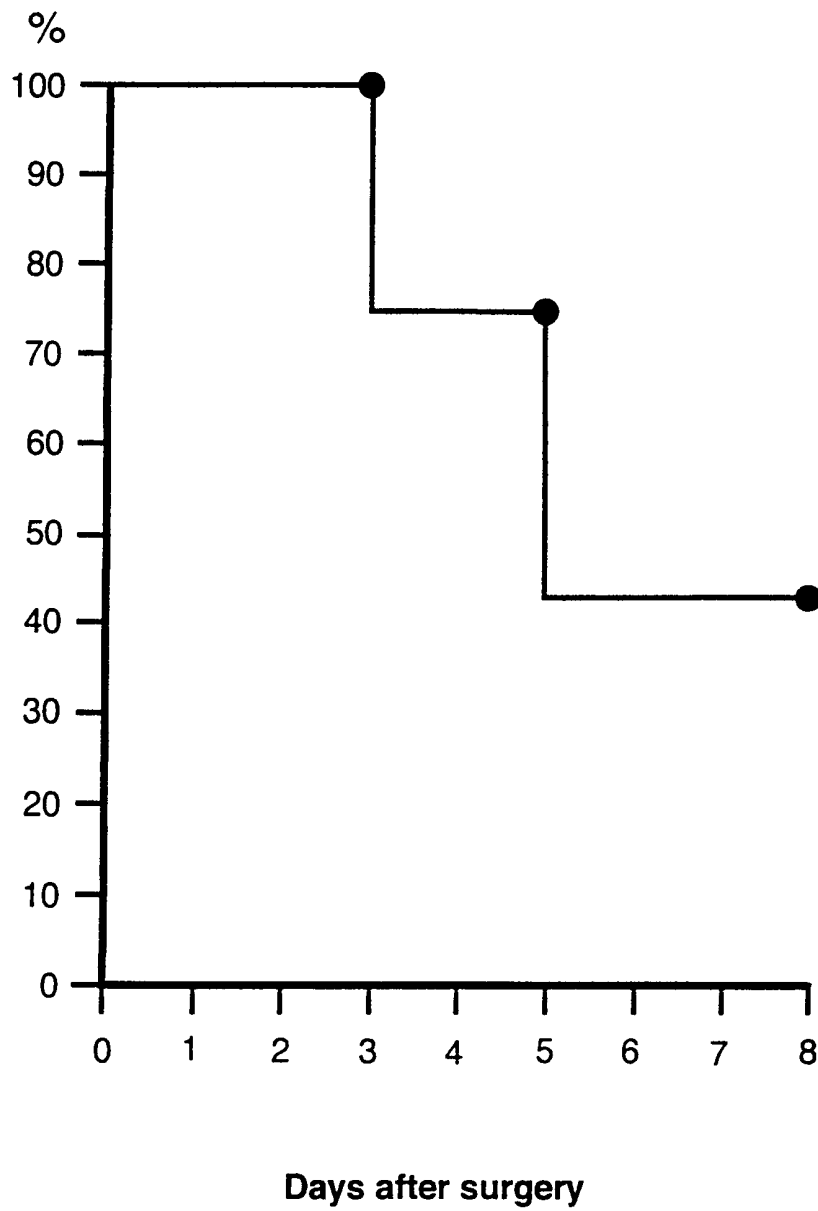
6/10
Fig.6

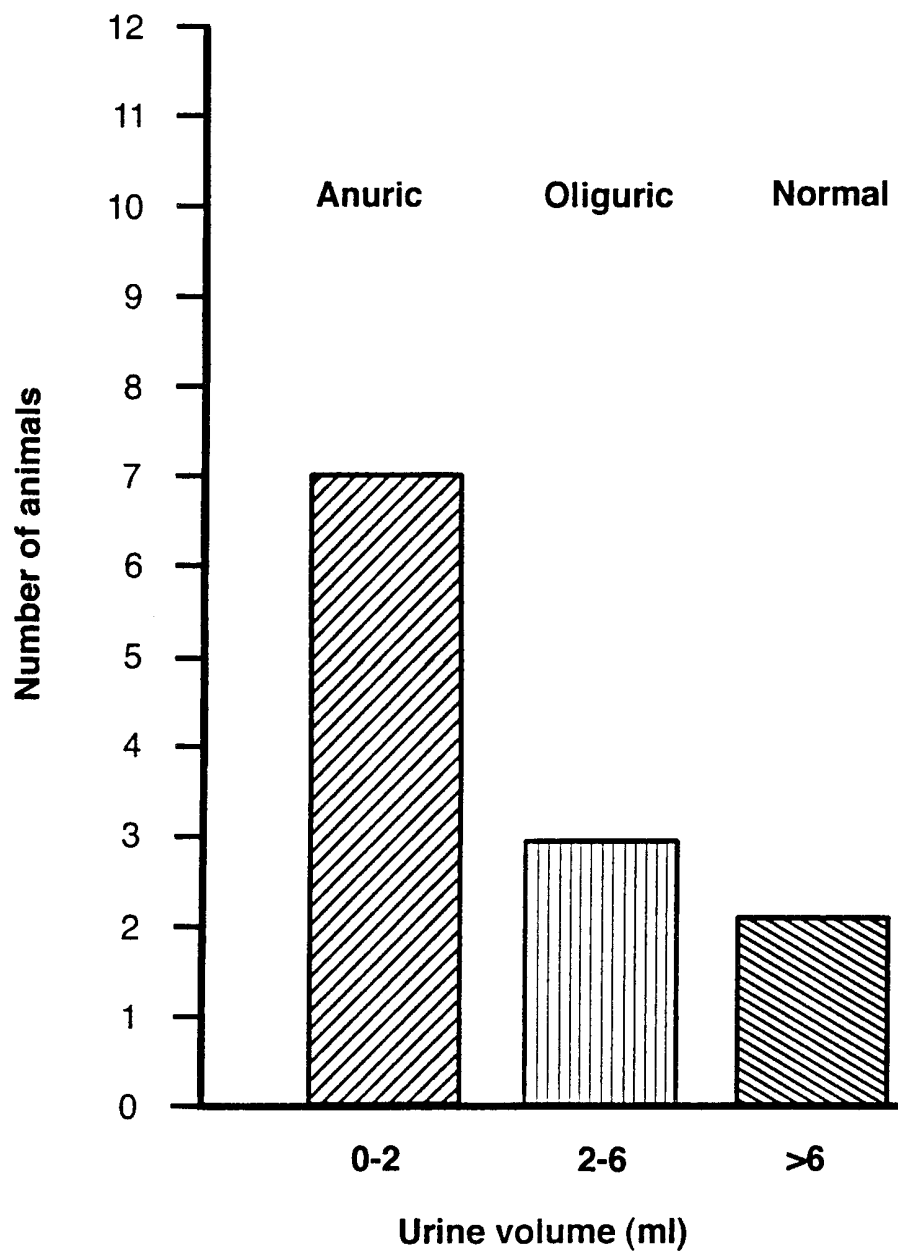
**Group F 24h Diuresis after
renal ischemia/reperfusion injury**



7/10
Fig.7

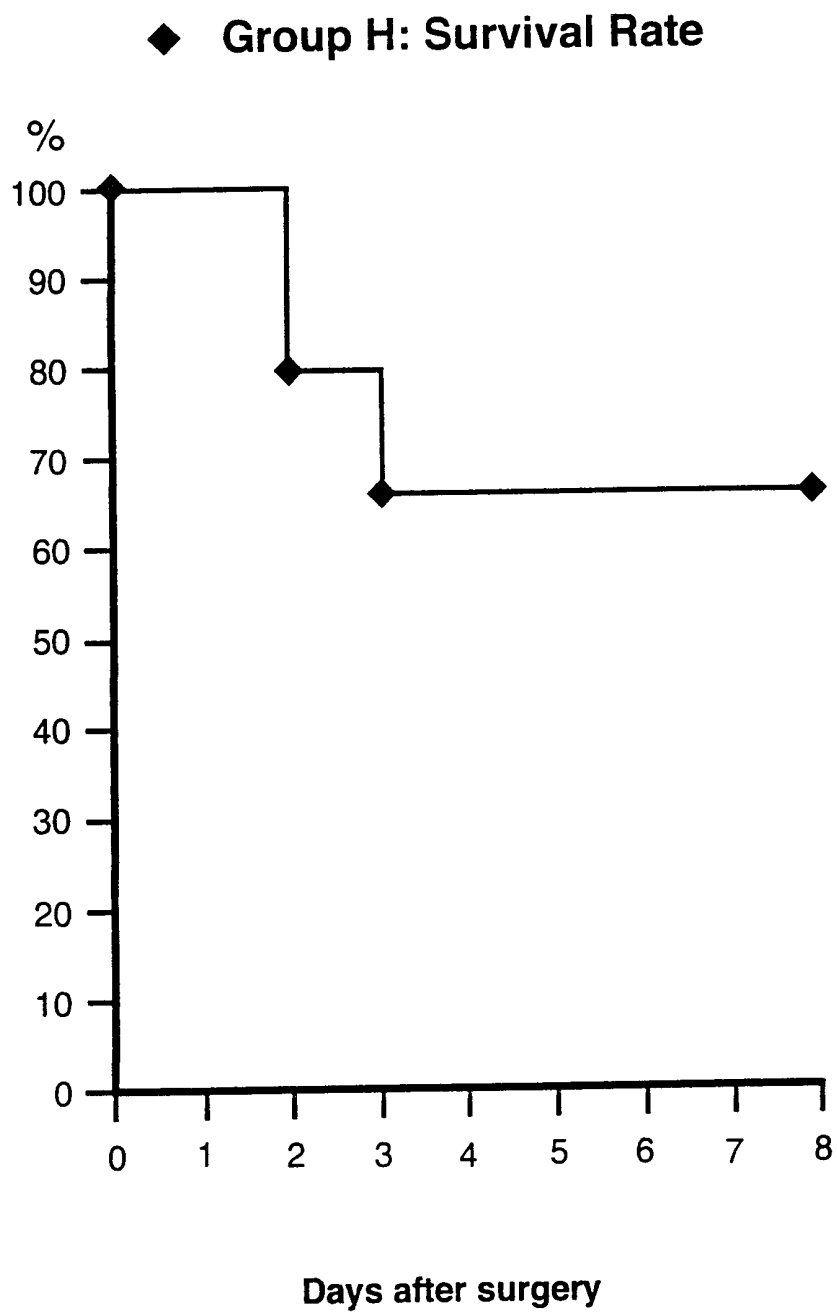
Group G Survival Rate



8/10
Fig.8**Group G 24h Diuresis after
renal ischemia/reperfusion injury**

9/10

Fig.9



10 / 10

Fig.10

