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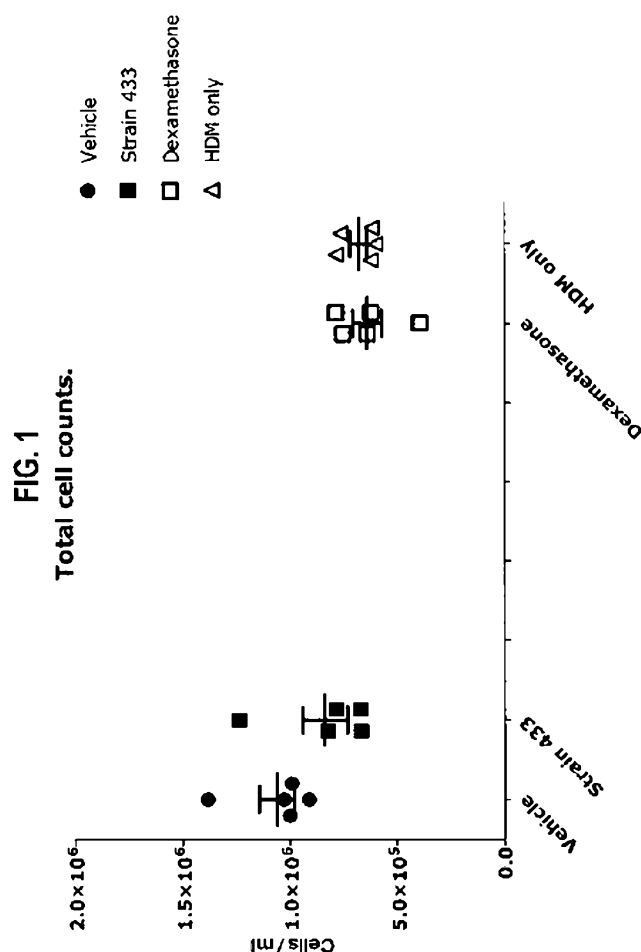
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54 Titre : Compositions comprising bacterial strains.

57 Abrégé :

The invention provides compositions comprising bacterial strains for treating and preventing inflammatory and autoimmune diseases.

Fig. 1



COMPOSITIONS COMPRISING BACTERIAL STRAINS

TECHNICAL FIELD

This invention is in the field of compositions comprising bacterial strains isolated from the mammalian digestive tract and the use of such compositions in the treatment of disease.

BACKGROUND TO THE INVENTION

The human intestine is thought to be sterile *in utero*, but it is exposed to a large variety of maternal and environmental microbes immediately after birth. Thereafter, a dynamic period of microbial colonization and succession occurs, which is influenced by factors such as delivery mode, environment, diet and host genotype, all of which impact upon the composition of the gut microbiota, particularly during early life. Subsequently, the microbiota stabilizes and becomes adult-like [1]. The human gut microbiota contains more than 500-1000 different phylotypes belonging essentially to two major bacterial divisions, the Bacteroidetes and the Firmicutes [2]. The successful symbiotic relationships arising from bacterial colonization of the human gut have yielded a wide variety of metabolic, structural, protective and other beneficial functions. The enhanced metabolic activities of the colonized gut ensure that otherwise indigestible dietary components are degraded with release of by-products providing an important nutrient source for the host. Similarly, the immunological importance of the gut microbiota is well-recognized and is exemplified in germfree animals which have an impaired immune system that is functionally reconstituted following the introduction of commensal bacteria [3-5].

Dramatic changes in microbiota composition have been documented in gastrointestinal disorders such as inflammatory bowel disease (IBD). For example, the levels of *Clostridium* cluster XIVa bacteria are reduced in IBD patients whilst numbers of *E. coli* are increased, suggesting a shift in the balance of symbionts and pathobionts within the gut [6-9]. Interestingly, this microbial dysbiosis is also associated with imbalances in T effector cell populations.

In recognition of the potential positive effect that certain bacterial strains may have on the animal gut, various strains have been proposed for use in the treatment of various diseases (see, for example, [10-13]). Also, certain strains, including mostly *Lactobacillus* and *Bifidobacterium* strains, have been proposed for use in treating various inflammatory and autoimmune diseases that are not directly linked to the intestines (see [14] and [15] for reviews). However, the relationship between different diseases and different bacterial strains, and the precise effects of particular bacterial strains on the gut and at a systemic level and on any particular types of diseases, are poorly characterised.

There is a requirement in the art for new methods of treating inflammatory and autoimmune diseases. There is also a requirement for the potential effects of gut bacteria to be characterised so that new therapies using gut bacteria can be developed.

SUMMARY OF THE INVENTION

The inventors have developed new therapies for treating and preventing inflammatory and autoimmune diseases. In particular, the inventors have developed new therapies for treating and preventing diseases and conditions mediated by IL-17 or the Th17 pathway. In particular, the inventors have identified that bacterial strains from the genus *Roseburia* can be effective for reducing the Th17 inflammatory response. As described in the examples, oral administration of compositions comprising *Roseburia hominis* may reduce the severity of the inflammatory response, including the Th17 inflammatory response, in mouse models of asthma, rheumatoid arthritis and multiple sclerosis.

Therefore, in a first embodiment, the invention provides a composition comprising a bacterial strain of the genus *Roseburia*, for use in a method of treating or preventing a disease or condition mediated by IL-17 or the Th17 pathway. The inventors have identified that treatment with bacterial strains from this genus can reduce levels of cytokines that are part of the Th17 pathway, including IL-17, can alleviate the Th17 inflammatory response and can provide clinical benefits in mouse models of inflammatory and autoimmune diseases mediated by IL-17 and the Th17 pathway.

In particular embodiments, the invention provides a composition comprising a bacterial strain of the genus *Roseburia*, for use in a method of treating or preventing a disease or condition selected from the group consisting of: multiple sclerosis; arthritis, such as rheumatoid arthritis, osteoarthritis, psoriatic arthritis, or juvenile idiopathic arthritis; neuromyelitis optica (Devic's disease); ankylosing spondylitis; spondyloarthritis; psoriasis; systemic lupus erythematosus; inflammatory bowel disease, such as Crohn's disease or ulcerative colitis; celiac disease; asthma, such as allergic asthma or neutrophilic asthma; chronic obstructive pulmonary disease (COPD); cancer, such as breast cancer, colon cancer, lung cancer or ovarian cancer; uveitis; scleritis; vasculitis; Behcet's disease; atherosclerosis; atopic dermatitis; emphysema; periodontitis; allergic rhinitis; and allograft rejection. The effect shown for the bacterial strains from the genus *Roseburia* on the Th17 inflammatory response may provide therapeutic benefits for diseases and conditions mediated by IL-17 and the Th17 pathway, such as those listed above.

In preferred embodiments, the invention provides a composition comprising a bacterial strain of the genus *Roseburia*, for use in a method of treating or preventing asthma, such as neutrophilic asthma or allergic asthma. The inventors have identified that treatment with *Roseburia* strains can reduce recruitment of neutrophils and eosinophils into the lungs, which can help treat or prevent asthma. Furthermore, the inventors have tested and demonstrated the efficacy of *Roseburia* strains in mouse models of asthma. In certain embodiments, the composition is for use in a method of treating or preventing neutrophilic asthma or eosinophilic asthma. The effect shown for the compositions of the invention on neutrophils and eosinophils mean that they may be particularly effective for treating or preventing neutrophilic asthma and eosinophilic asthma. Indeed, in certain embodiments, the

composition is for use in a method of reducing a neutrophilic inflammatory response in the treatment or prevention of asthma, or the composition is for use in a method of reducing an eosinophilic inflammatory response in the treatment or prevention of asthma. In preferred embodiments, the invention provides a composition comprising a bacterial strain of the species *Roseburia hominis* for use in the treatment of asthma, and in particular eosinophilic or allergic asthma. *Roseburia hominis* is shown to have a particularly pronounced effect on eosinophils in asthma models and treatment with *Roseburia hominis* may be particularly effective for treating eosinophilic or allergic asthma.

In further preferred embodiments, the invention provides a composition comprising a bacterial strain of the genus *Roseburia*, for use in a method of treating or preventing rheumatoid arthritis. The inventors have identified that treatment with *Roseburia* strains can provide clinical benefits in a mouse model of rheumatoid arthritis and can reduce joint swelling. In preferred embodiments, the invention provides a composition comprising a bacterial strain of the species *Roseburia hominis*, for use in the treatment of rheumatoid arthritis. Compositions using *Roseburia hominis* may be particularly effective for treating rheumatoid arthritis.

In further preferred embodiments, the invention provides a composition comprising a bacterial strain of the genus *Roseburia*, for use in a method of treating or preventing multiple sclerosis. The inventors have identified that treatment with *Roseburia* strains can reduce disease incidence and disease severity in a mouse model of multiple sclerosis. In preferred embodiments, the invention provides a composition comprising a bacterial strain of the species *Roseburia hominis*, for use in the treatment of multiple sclerosis. Compositions using *Roseburia hominis* may be particularly effective for treating multiple sclerosis.

In certain embodiments, the compositions of the invention are for use in a method of reducing IL-17 production or reducing Th17 cell differentiation in the treatment or prevention of a disease or condition mediated by IL-17 or the Th17 pathway. In particular, the compositions of the invention may be used in reducing IL-17 production or reducing Th17 cell differentiation in the treatment or prevention of asthma, rheumatoid arthritis or multiple sclerosis. Preferably, the invention provides compositions comprising a bacterial strain of the species *Roseburia hominis*, for use in reducing IL-17 production or reducing Th17 cell differentiation in the treatment or prevention of asthma, rheumatoid arthritis or multiple sclerosis.

In certain embodiments, the composition is for use in a patient with elevated IL-17 levels or Th17 cells. The effect on the Th17 inflammatory response shown for *Roseburia* strains may be particularly beneficial for such patients.

In preferred embodiments of the invention, the bacterial strain in the composition is of *Roseburia hominis*. Closely related strains may also be used, such as bacterial strains that have a 16s rRNA sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to the 16s rRNA

sequence of a bacterial strain of *Roseburia hominis*. Preferably, the bacterial strain has a 16s rRNA sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to SEQ ID NO:1, 2 or 3. Preferably, the sequence identity is to SEQ ID NO:3. Preferably, the bacterial strain for use in the invention has the 16s rRNA sequence represented by SEQ ID NO:3.

5 In certain embodiments, the composition of the invention is for oral administration. Oral administration of the strains of the invention can be effective for treating IL-17- or Th17 pathway-mediated diseases and conditions. Also, oral administration is convenient for patients and practitioners and allows delivery to and / or partial or total colonisation of the intestine.

10 In certain embodiments, the composition of the invention comprises one or more pharmaceutically acceptable excipients or carriers.

In certain embodiments, the composition of the invention comprises a bacterial strain that has been lyophilised. Lyophilisation is an effective and convenient technique for preparing stable compositions that allow delivery of bacteria.

15 In certain embodiments, the invention provides a food product comprising the composition as described above.

In certain embodiments, the invention provides a vaccine composition comprising the composition as described above.

20 Additionally, the invention provides a method of treating or preventing a disease or condition mediated by IL-17 or the Th17 pathway, comprising administering a composition comprising a bacterial strain of the genus *Roseburia*.

25 In developing the above invention, the inventors have identified and characterised a bacterial strain that is particularly useful for therapy. The *Roseburia hominis* strain of the invention is shown to be effective for treating the diseases described herein, such as arthritis, asthma and multiple sclerosis. Therefore, in another aspect, the invention provides a cell of the *Roseburia hominis* strain deposited under accession number NCIMB 42383, or a derivative thereof. The invention also provides compositions comprising such cells, or biologically pure cultures of such cells. The invention also provides a cell of the *Roseburia hominis* strain deposited under accession number NCIMB 42383, or a derivative thereof, for use in therapy, in particular for the diseases described herein.

BRIEF DESCRIPTION OF DRAWINGS

30 **Figure 1:** Mouse model of house dust mite-induced asthma - Total BAL fluid cell counts.

Figure 2: Mouse model of house dust mite-induced asthma - Total eosinophil count in BALF.

Figure 3: Mouse model of house dust mite-induced asthma - Proportion of eosinophils in BALF.

Figure 4: Mouse model of house dust mite-induced asthma - Total macrophage count in BALF.

Figure 5: Mouse model of house dust mite-induced asthma - Proportion of macrophages in BALF.

Figure 6: Mouse model of house dust mite-induced asthma - Total neutrophil count in BALF.

Figure 7: Mouse model of house dust mite-induced asthma - Proportion of neutrophils in BALF.

5 **Figure 8:** Mouse model of house dust mite-induced asthma - Total lymphocyte count in BALF.

Figure 9: Mouse model of house dust mite-induced asthma - Proportion of lymphocytes in BALF.

Figure 10: Mouse model of severe neutrophilic asthma - Total BAL fluid cell counts.

Figure 11: Mouse model of severe neutrophilic asthma - Total eosinophil count in BALF.

Figure 12: Mouse model of severe neutrophilic asthma - Proportion of eosinophils in BALF.

10 **Figure 13:** Mouse model of severe neutrophilic asthma - Total macrophage count in BALF.

Figure 14: Mouse model of severe neutrophilic asthma - Proportion of macrophages in BALF.

Figure 15: Mouse model of severe neutrophilic asthma - Total neutrophil count in BALF.

Figure 16: Mouse model of severe neutrophilic asthma - Proportion of neutrophils in BALF.

Figure 17: Mouse model of severe neutrophilic asthma - Total lymphocyte count in BALF.

15 **Figure 18:** Mouse model of severe neutrophilic asthma - Proportion of lymphocytes in BALF.

Figure 19: Mouse model of rheumatoid arthritis - Bodyweights, days -14 to 0. Data are presented as Mean $\pm$ SEM percentages of the initial (Day -14) bodyweights.

Figure 20: Mouse model of rheumatoid arthritis - Bodyweights, days 0 to 42. Data are presented as Mean $\pm$ SEM percentages of the initial (Day 0) bodyweights.

20 **Figure 21:** Mouse model of rheumatoid arthritis - Clinical Scores. Data are presented as Mean $\pm$ SEM. **** $p < 0.0001$ when compared to Day 21 in the vehicle-treated group.

Figure 22: Mouse model of rheumatoid arthritis - Splenocyte proliferative response to Collagen II. Media background subtracted [CII-stimulated - media background] counts per minute based on ³H-TdR incorporation. All data are presented as Mean $\pm$ SEM. *** $p < 0.001$ compared to Vehicle group.

25

Figure 23: Mouse model of rheumatoid arthritis - Levels of IFN γ in tissue culture supernatants. Lines represent group median values.

Figure 24: Mouse model of rheumatoid arthritis - Levels of IL-17A in tissue culture supernatants. Lines represent group median values.

Figure 25: Mouse model of rheumatoid arthritis - Levels of IL-10 in tissue culture supernatants. Lines represent group median values.

5 **Figure 26:** Mouse model of rheumatoid arthritis - Levels of IL-6 in tissue culture supernatants. Lines represent group median values.

Figure 27: Mouse model of house dust mite-induced asthma – Total IgE in Serum

Figure 28: Mouse model of house dust mite-induced asthma – HDM specific IgG1 in Serum

Figure 29: Mouse model of house dust mite-induced asthma – Total IgE in BALF

10 **Figure 30:** Mouse model of house dust mite-induced asthma – HDM specific IgG1 in BALF

Figure 31: Mouse model of house dust mite-induced asthma – Histological Analysis – Mean Peribronchiolar Infiltration Score

Figure 32: Mouse model of house dust mite-induced asthma – Histological Analysis – Mean Perivascular Infiltration Score

15 **Figure 33:** Mouse model of house dust mite-induced asthma – Histological Analysis – Mean Inflammatory Score (Average of both Peribronchiolar and Perivascular Infiltration Score)

Figure 34: Mouse model of house dust mite-induced asthma – Histological Analysis – Mucus Score

Figure 35: Mouse model of house dust mite-induced asthma – IL-9 level in lung tissue

Figure 36: Mouse model of house dust mite-induced asthma – IL-1a level in lung tissue

20 **Figure 37:** Mouse model of house dust mite-induced asthma – IFN γ level in lung tissue

Figure 38: Mouse model of house dust mite-induced asthma – IL-17A level in lung tissue

Figure 39: Mouse model of house dust mite-induced asthma – IL-4 level in lung tissue

Figure 40: Mouse model of house dust mite-induced asthma – IL-5 level in lung tissue

Figure 41: Mouse model of house dust mite-induced asthma - IL-1b level in lung tissue

25 **Figure 42:** Mouse model of house dust mite-induced asthma - RANTES level in lung tissue

Figure 43: Mouse model of house dust mite-induced asthma - MIP-1a level in lung tissue

Figure 44: Mouse model of house dust mite-induced asthma - KC level in lung tissue

Figure 45: Mouse model of house dust mite-induced asthma - MIP-2 level in lung tissue

Figure 46: Mouse model of severe neutrophilic asthma – HDM specific IgG1 in Serum

Figure 47: Mouse model of severe neutrophilic asthma – HDM specific IgG2a in Serum

Figure 48: Mouse model of severe neutrophilic asthma – HDM specific IgG1 in BALF

5 **Figure 49:** Mouse model of severe neutrophilic asthma – HDM specific IgG2a in BALF

Figure 50: Mouse model of severe neutrophilic asthma – Histological Analysis – Mean Peribronchiolar Infiltration Score

Figure 51: Mouse model of severe neutrophilic asthma – Histological Analysis – Mean Perivascular Infiltration Score

10 **Figure 52:** Mouse model of severe neutrophilic asthma – Histological Analysis – Mean Inflammatory Score (Average of both Peribronchiolar and Perivascular Infiltration Score)

Figure 53: Mouse model of severe neutrophilic asthma – TNF α level in lung tissue

Figure 54: Mouse model of severe neutrophilic asthma – IL-1 α level in lung tissue

Figure 55: Mouse model of severe neutrophilic asthma – IFN γ level in lung tissue

15 **Figure 56:** Mouse model of severe neutrophilic asthma – IL-17F level in lung tissue

Figure 57: Mouse model of severe neutrophilic asthma – IL-1 β level in lung tissue

Figure 58: Mouse model of severe neutrophilic asthma - RANTES level in lung tissue

Figure 59: Mouse model of severe neutrophilic asthma - MIP-2 level in lung tissue

Figure 60: Mouse model of severe neutrophilic asthma - KC level in lung tissue

20 **Figure 61:** Mouse model of severe neutrophilic asthma – IL-17A level in lung tissue

Figure 62: Mouse model of severe neutrophilic asthma –MIP-1 α level in lung tissue

Figure 63: Mouse model of severe neutrophilic asthma – IL-33 level in lung tissue

25 **Figure 64:** Mouse model of rheumatoid arthritis - Visual Template for Histopathology Scoring. Representative images showing composite scores from mouse tarsal joints in a collagen-induced arthritis study.

Figure 65: Mouse model of rheumatoid arthritis - Histopathology: Inflammation Scores. Data are presented as Mean $\pm$ SEM.

Figure 66: Mouse model of rheumatoid arthritis - Histopathology: Cartilage Scores. Data are presented as Mean $\pm$ SEM.

Figure 67: Mouse model of rheumatoid arthritis - Histopathology: Bone Scores. Data are presented as Mean $\pm$ SEM.

5 **Figure 68:** Mouse model of rheumatoid arthritis - Histopathology: Total Scores. Data are presented as Mean $\pm$ SEM.

10 **Figure 69:** Mouse model of rheumatoid arthritis - Histopathology: Representative Pictures. Animal ID (#n.n) and limb (R for right, L for left) are indicated between brackets. Left image (vehicle): extensive joint and bone destruction with inflammation and fibrosis extending to the peri-articular soft tissues.

Figure 70: Mouse model of multiple sclerosis - clinical score.

Figure 71: Mouse model of multiple sclerosis - disease incidence.

DISCLOSURE OF THE INVENTION

Bacterial strains

15 The compositions of the invention comprise a bacterial strain of the genus *Roseburia*. The examples demonstrate that bacteria of this genus are useful for treating or preventing diseases and conditions mediated by IL-17 or the Th17 pathway. The preferred bacterial strains are of the species *Roseburia hominis*.

20 Examples of *Roseburia* species for use in the invention include *Roseburia hominis*, *Roseburia cecicola*, *Roseburia faecis*, *Roseburia intestinalis*, and *Roseburia inulinivorans*. *Roseburia* bacteria are slightly curved rod-shaped cells that are strictly anaerobic and indigenous to the mammalian intestine. They are of the phylogenetic cluster XIVa within the Firmicutes phylum. The bacteria are butyrate-producing and are actively motile through multiple flagella present along the concave side and in a cluster at one end [16]. *Roseburia hominis* and *Roseburia intestinalis* are recently described
25 examples.

An example of *Roseburia hominis* is the strain deposited under the terms of the Budapest Treaty at National Collections of Industrial, Food and Marine Bacteria (NCIMB) at NCIMB Ltd, Ferguson Building, Craibstone Estate, Bucksburn, Aberdeen, UK, AB21 9YA, on 21 October 2004 by the Rowett Research Institute under the accession number NCIMB 14029^T *Roseburia hominis*
30 A2-183^T (DSM = 16839^T). Other exemplary *Roseburia hominis* strains are described in [17]. GenBank/EMBL/DBJ accession numbers for the 16S rRNA gene sequence of strains of *Roseburia hominis* are AY804148 and AJ270482 (disclosed herein as SEQ ID NO:1 and SEQ ID NO:2).

An example of *Roseburia intestinalis* is the strain deposited under the accession number NCIMB 13810 *Roseburia intestinalis* L1-82^T (DSM = 14610^T). Another example is the *Roseburia intestinalis* strain as described in [17]. Reference [17] also describes exemplary *Roseburia faecis* and *Roseburia inulinivorans* strains.

5 The *Roseburia hominis* bacterium deposited under accession number NCIMB 42383 was tested in the Examples and is also referred to herein as strain 433. A 16S rRNA sequence for the 433 strain that was tested is provided in SEQ ID NO:3. Strain 433 was deposited with the international depositary authority NCIMB, Ltd. (Ferguson Building, Aberdeen, AB21 9YA, Scotland) by GT
10 Biologics Ltd. (Life Sciences Innovation Building, Aberdeen, AB25 2ZS, Scotland) on 12th March 2015 as “*Roseburia hominis* 433” and was assigned accession number NCIMB 42383. GT Biologics Ltd. subsequently changed its name to 4D Pharma Research Limited.

A genome sequence for strain 433 is provided in SEQ ID NO:4 of WO2016203221. This sequence was generated using the PacBio RS II platform.

15 Bacterial strains closely related to the strain tested in the examples are also expected to be effective for treating or preventing diseases and conditions mediated by IL-17 or the Th17 pathway. In certain embodiments, the bacterial strain for use in the invention has a 16s rRNA sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to the 16s rRNA sequence of a bacterial strain of *Roseburia intestinalis*. Preferably, the bacterial strain for use in the invention has a 16s rRNA
20 sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to SEQ ID NO:1, 2 or 3. Preferably, the sequence identity is to SEQ ID NO:3. Preferably, the bacterial strain for use in the invention has the 16s rRNA sequence represented by SEQ ID NO:3.

Bacterial strains that are biotypes of the bacterium deposited under accession number 42383 are also expected to be effective for treating or preventing diseases and conditions mediated by IL-17 or the Th17 pathway. A biotype is a closely related strain that has the same or very similar physiological
25 and biochemical characteristics.

Strains that are biotypes of the bacterium deposited under accession number NCIMB 42383 and that are suitable for use in the invention may be identified by sequencing other nucleotide sequences for the bacterium deposited under accession number NCIMB 42383. For example, substantially the whole genome may be sequenced and a biotype strain for use in the invention may have at least 95%,
30 96%, 97%, 98%, 99%, 99.5% or 99.9% sequence identity across at least 80% of its whole genome (e.g. across at least 85%, 90%, 95% or 99%, or across its whole genome). Other suitable sequences for use in identifying biotype strains may include hsp60 or repetitive sequences such as BOX, ERIC, (GTG)₅, or REP or [18]. Biotype strains may have sequences with at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% sequence identity to the corresponding sequence of the bacterium deposited
35 under accession number NCIMB 42383.

In certain embodiments, the bacterial strain for use in the invention has a genome with sequence identity to SEQ ID NO:4 of WO2016203221. In preferred embodiments, the bacterial strain for use in the invention has a genome with at least 90% sequence identity (e.g. at least 92%, 94%, 95%, 96%, 97%, 98%, 99% or 100% sequence identity) to SEQ ID NO:4 across at least 60% (e.g. at least 65%, 70%, 75%, 80%, 85%, 95%, 96%, 97%, 98%, 99% or 100%) of SEQ ID NO:4. For example, the bacterial strain for use in the invention may have a genome with at least 90% sequence identity to SEQ ID NO:4 across 70% of SEQ ID NO:4, or at least 90% sequence identity to SEQ ID NO:4 across 80% of SEQ ID NO:4, or at least 90% sequence identity to SEQ ID NO:4 across 90% of SEQ ID NO:4, or at least 90% sequence identity to SEQ ID NO:4 across 100% of SEQ ID NO:4, or at least 95% sequence identity to SEQ ID NO:4 across 70% of SEQ ID NO:4, or at least 95% sequence identity to SEQ ID NO:4 across 80% of SEQ ID NO:4, or at least 95% sequence identity to SEQ ID NO:4 across 90% of SEQ ID NO:4, or at least 95% sequence identity to SEQ ID NO:4 across 100% of SEQ ID NO:4, or at least 98% sequence identity to SEQ ID NO:4 across 70% of SEQ ID NO:4, or at least 98% sequence identity to SEQ ID NO:4 across 80% of SEQ ID NO:4, or at least 98% sequence identity to SEQ ID NO:4 across 90% of SEQ ID NO:4, or at least 98% sequence identity to SEQ ID NO:4 across 100% of SEQ ID NO:4.

Alternatively, strains that are biotypes of the bacterium deposited under accession number NCIMB 42383 and that are suitable for use in the invention may be identified by using the accession number NCIMB 42383 deposit and restriction fragment analysis and/or PCR analysis, for example by using fluorescent amplified fragment length polymorphism (FAFLP) and repetitive DNA element (rep)-PCR fingerprinting, or protein profiling, or partial 16S or 23s rDNA sequencing. In preferred embodiments, such techniques may be used to identify other *Roseburia hominis* strains.

In certain embodiments, strains that are biotypes of the bacterium deposited under accession number NCIMB 42383 and that are suitable for use in the invention are strains that provide the same pattern as the bacterium deposited under accession number NCIMB 42383 when analysed by amplified ribosomal DNA restriction analysis (ARDRA), for example when using Sau3AI restriction enzyme (for exemplary methods and guidance see, for example,[19]). Alternatively, biotype strains are identified as strains that have the same carbohydrate fermentation patterns as the bacterium deposited under accession number NCIMB 42383.

Other *Roseburia* strains that are useful in the compositions and methods of the invention, such as biotypes of the bacteria deposited under accession number NCIMB 42383, may be identified using any appropriate method or strategy, including the assays described in the examples. For instance, strains for use in the invention may be identified by culturing in anaerobic YCFA and/or administering the bacteria to the type II collagen-induced arthritis mouse model and then assessing cytokine levels. In particular, bacterial strains that have similar growth patterns, metabolic type

and/or surface antigens to the bacterium deposited under accession number NCIMB 42383 may be useful in the invention. A useful strain will have comparable immune modulatory activity to the NCIMB 42383 strain. In particular, a biotype strain will elicit comparable effects on the asthma, arthritis and multiple sclerosis disease models and comparable effects on cytokine levels to the effects shown in the Examples, which may be identified by using the culturing and administration protocols described in the Examples.

A particularly preferred strain of the invention is the *Roseburia hominis* strain deposited under accession number NCIMB 42383. This is the exemplary 433 strain tested in the examples and shown to be effective for treating disease. Therefore, the invention provides a cell, such as an isolated cell, of the *Roseburia hominis* strain deposited under accession number NCIMB 42383, or a derivative thereof. The invention also provides a composition comprising a cell of the *Roseburia hominis* strain deposited under accession number NCIMB 42383, or a derivative thereof. The invention also provides a biologically pure culture of the *Roseburia hominis* strain deposited under accession number NCIMB 42383. The invention also provides a cell of the *Roseburia hominis* strain deposited under accession number NCIMB 42383, or a derivative thereof, for use in therapy, in particular for the diseases described herein.

A derivative of the strain deposited under accession number NCIMB 42383 may be a daughter strain (progeny) or a strain cultured (subcloned) from the original. A derivative of a strain of the invention may be modified, for example at the genetic level, without ablating the biological activity. In particular, a derivative strain of the invention is therapeutically active. A derivative strain will have comparable immune modulatory activity to the original NCIMB 42383 strain. In particular, a derivative strain will elicit comparable effects on the asthma, arthritis and multiple sclerosis disease models and comparable effects on cytokine levels to the effects shown in the Examples, which may be identified by using the culturing and administration protocols described in the Examples. A derivative of the NCIMB 42383 strain will generally be a biotype of the NCIMB 42383 strain.

References to cells of the *Roseburia hominis* strain deposited under accession number NCIMB 42383 encompass any cells that have the same safety and therapeutic efficacy characteristics as the strains deposited under accession number NCIMB 42383, and such cells are encompassed by the invention.

In preferred embodiments, the bacterial strains in the compositions of the invention are viable and capable of partially or totally colonising the intestine.

Therapeutic uses

As demonstrated in the examples, the bacterial compositions of the invention are effective for reducing the Th17 inflammatory response. In particular, treatment with compositions of the invention achieves a reduction in IL-17A levels and other Th17 pathway cytokines, and clinical improvements

in animal models of conditions mediated by IL-17 and the Th17 pathway. Therefore, the compositions of the invention may be useful for treating or preventing inflammatory and autoimmune diseases, and in particular diseases or conditions mediated by IL-17. In particular, the compositions of the invention may be useful for reducing or preventing elevation of the IL-17 inflammatory response.

Th17 cells are a subset of T helper cells that produce, for example, IL-17A, IL-17F, IL-21 and IL-22. Th17 cell differentiation and IL-17 expression may be driven by IL-23. These cytokines and others form important parts of the Th17 pathway, which is a well-established inflammatory signalling pathway that contributes to and underlies a number of inflammatory and autoimmune diseases (as described in, for example, [20-25]). Diseases wherein the Th17 pathway is activated are Th17 pathway-mediated diseases. Th17 pathway-mediated diseases can be ameliorated or alleviated by repressing the Th17 pathway, which may be through a reduction in the differentiation of Th17 cells or a reduction in their activity or a reduction in the level of Th17 pathway cytokines. Diseases mediated by the Th17 pathway may be characterised by increased levels of cytokines produced by Th17 cells, such as IL-17A, IL-17F, IL-21, IL-22, IL-26, IL-9 (reviewed in [26]). Diseases mediated by the Th17 pathway may be characterised by increased expression of Th-17-related genes, such as Stat3 or IL-23R. Diseases mediated by the Th17 pathway may be associated with increased levels of Th17 cells.

IL-17 is a pro-inflammatory cytokine that contributes to the pathogenesis of several inflammatory and autoimmune diseases and conditions. IL-17 as used herein may refer to any member of the IL-17 family, including IL-17A, IL-17B, IL-17C, IL-17D, IL-17E, and IL-17F. IL-17-mediated diseases and conditions are characterised by high expression of IL-17 and/or the accumulation or presence of IL-17-positive cells in a tissue affected by the disease or condition. Similarly, IL-17-mediated diseases and conditions are diseases and conditions that are exacerbated by high IL-17 levels or an increase in IL-17 levels, and that are alleviated by low IL-17 levels or a reduction in IL-17 levels. The IL-17 inflammatory response may be local or systemic.

Examples of diseases and conditions that may be mediated by IL-17 or the Th17 pathway include multiple sclerosis; arthritis, such as rheumatoid arthritis, osteoarthritis, psoriatic arthritis, or juvenile idiopathic arthritis; neuromyelitis optica (Devic's disease); ankylosing spondylitis; spondyloarthritis; psoriasis; systemic lupus erythematosus; inflammatory bowel disease, such as Crohn's disease or ulcerative colitis; celiac disease; asthma, such as allergic asthma or neutrophilic asthma; chronic obstructive pulmonary disease (COPD); cancer, such as breast cancer, colon cancer, lung cancer or ovarian cancer; uveitis; scleritis; vasculitis; Behcet's disease; atherosclerosis; atopic dermatitis; emphysema; periodontitis; allergic rhinitis; and allograft rejection. In preferred embodiments, the compositions of the invention are used for treating or preventing one or more of these conditions or

diseases. In further preferred embodiments, these conditions or diseases are mediated by IL-17 or the Th17 pathway.

In certain embodiments, the compositions of the invention are for use in a method of reducing IL-17 production or reducing Th17 cell differentiation in the treatment or prevention of a disease or condition mediated by IL-17 or the Th17 pathway. In certain embodiments, the compositions of the invention are for use in treating or preventing an inflammatory or autoimmune disease, wherein said treatment or prevention is achieved by reducing or preventing elevation of the Th17 inflammatory response. In certain embodiments, the compositions of the invention are for use in treating a patient with an inflammatory or autoimmune disease, wherein the patient has elevated IL-17 levels or elevated Th17 cells or is exhibiting a Th17 inflammatory response. In certain embodiments, the patient may have been diagnosed with a chronic inflammatory or autoimmune disease or condition, or the composition of the invention may be for use in preventing an inflammatory or autoimmune disease or condition developing into a chronic inflammatory or autoimmune disease or condition. In certain embodiments, the disease or condition may not be responsive to treatment with TNF- α inhibitors. These uses of the invention may be applied to any of the specific disease or conditions listed in the preceding paragraph.

IL-17 and the Th17 pathway are often associated with chronic inflammatory and autoimmune diseases, so the compositions of the invention may be particularly useful for treating or preventing chronic diseases or conditions as listed above. In certain embodiments, the compositions are for use in patients with chronic disease. In certain embodiments, the compositions are for use in preventing the development of chronic disease.

The compositions of the invention may be useful for treating diseases and conditions mediated by IL-17 or the Th17 pathway and for addressing the Th17 inflammatory response, so the compositions of the invention may be particularly useful for treating or preventing chronic disease, treating or preventing disease in patients that have not responded to other therapies (such as treatment with TNF- α inhibitors), and/or treating or preventing the tissue damage and symptoms associated with IL-17 and Th17 cells. For example, IL-17 is known to activate matrix destruction in cartilage and bone tissue and IL-17 has an inhibitory effect on matrix production in chondrocytes and osteoblasts, so the compositions of the invention may be useful for treating or preventing bone erosion or cartilage damage.

In certain embodiments, treatment with compositions of the invention provides a reduction or prevents an elevation in IL-17 levels, in particular IL-17A levels. In certain embodiments, treatment with compositions of the invention provides a reduction or prevents an elevation in IFN- γ or IL-6 levels. Such reduction or prevention of elevated levels of these cytokines may be useful for treating

or preventing inflammatory and autoimmune diseases and conditions, in particular those mediated by IL-17 or the Th17 pathway.

Asthma

In preferred embodiments, the compositions of the invention are for use in treating or preventing asthma. The examples demonstrate that the compositions of the invention achieve a reduction in the recruitment of neutrophils and/or eosinophils into the airways following sensitisation and challenge with house dust mite extract and so they may be useful in the treatment or prevention of asthma. Asthma is a chronic disease characterised by inflammation and restriction of the airways. The inflammation in asthma may be mediated by IL-17 and/or Th17 cells, and so the compositions of the invention may be particularly effective for preventing or treating asthma. The inflammation in asthma may be mediated by eosinophils and/or neutrophils.

In certain embodiments, the asthma is eosinophilic or allergic asthma. Eosinophilic and allergic asthma are characterised by increased numbers of eosinophils in peripheral blood and in airway secretions and is associated pathologically with thickening of the basement membrane zone and pharmacologically by corticosteroid responsiveness [27]. Compositions that reduce or inhibit eosinophil recruitment or activation may be useful for treating or preventing eosinophilic and allergic asthma.

In additional embodiments, the compositions of the invention are for use in treating or preventing neutrophilic asthma (or non-eosinophilic asthma). High neutrophil numbers are associated with severe asthma that may be insensitive to corticosteroid treatment. Compositions that reduce or inhibit neutrophil recruitment or activation may be useful for treating or preventing neutrophilic asthma.

Eosinophilic and neutrophilic asthma are not mutually exclusive conditions and treatments that help address either the eosinophil and neutrophil responses may be useful for treating asthma in general.

Increased IL-17 levels and activation of the Th17 pathway are associated with severe asthma, so the compositions of the invention may be useful for preventing the development of severe asthma or for treating severe asthma.

In certain embodiments, the compositions of the invention are for use in methods reducing an eosinophilic inflammatory response in the treatment or prevention of asthma, or for use in methods of reducing a neutrophilic inflammatory response in the treatment or prevention of asthma. As noted above, high levels of eosinophils in asthma is associated pathologically with thickening of the basement membrane zone, so reducing eosinophilic inflammatory response in the treatment or prevention of asthma may be able to specifically address this feature of the disease. Also, elevated neutrophils, either in combination with elevated eosinophils or in their absence, is associated with

severe asthma and chronic airway narrowing. Therefore, reducing the neutrophilic inflammatory response may be particularly useful for addressing severe asthma.

In certain embodiments, the compositions reduce peribronchiolar infiltration in allergic asthma, or are for use in reducing peribronchiolar infiltration in the treatment of allergic asthma. In certain
5 embodiments, the compositions reduce peribronchiolar and/or perivascular infiltration in neutrophilic asthma, or are for use in reducing peribronchiolar and/or perivascular infiltration in the treatment of allergic neutrophilic asthma.

In certain embodiments, treatment with compositions of the invention provides a reduction or prevents an elevation in IFN γ levels.

10 In certain embodiments, the compositions of the invention are for use in a method of treating asthma that results in a reduction of the eosinophilic and/or neutrophilic inflammatory response. In certain embodiments, the patient to be treated has, or has previously been identified as having, elevated neutrophil or eosinophil levels, for example as identified through blood sampling or sputum analysis.

The compositions of the invention may be useful for preventing the development of asthma in a new-
15 born when administered to the new-born, or to a pregnant woman. The compositions may be useful for preventing the development of asthma in children. The compositions of the invention may be useful for treating or preventing adult-onset asthma. The compositions of the invention may be useful for managing or alleviating asthma. The compositions of the invention may be particularly useful for reducing symptoms associated with asthma that is aggravated by allergens, such as house dust mites.

20 Treatment or prevention of asthma may refer to, for example, an alleviation of the severity of symptoms or a reduction in the frequency of exacerbations or the range of triggers that are a problem for the patient.

Arthritis

In preferred embodiments, the compositions of the invention are for use in treating or preventing
25 rheumatoid arthritis (RA). The examples demonstrate that the compositions of the invention achieve a reduction in the clinical signs of RA in a mouse model, reduce cartilage and bone damage, and reduce the IL-17 inflammatory response, and so they may be useful in the treatment or prevention of RA. RA is a systemic inflammatory disorder that primarily affects joints. RA is associated with an inflammatory response that results in swelling of joints, synovial hyperplasia, and destruction of
30 cartilage and bone. IL-17 and Th17 cells may have a key role in RA, for example because IL-17 inhibits matrix production in chondrocytes and osteoblasts and activates the production and function of matrix metalloproteinases and because RA disease activity is correlated to IL-17 levels and Th-17 cell numbers [28,29], so the compositions of the invention may be particularly effective for preventing or treating RA.

In certain embodiments, the compositions of the invention are for use in lowering IL-17 levels or preventing elevation of IL-17 levels in the treatment or prevention of RA. In certain embodiments, treatment with compositions of the invention provides a reduction or prevents an elevation in IL-17 levels, in particular IL-17A levels. In certain embodiments, treatment with compositions of the invention provides a reduction or prevents an elevation in IFN- γ or IL-6 levels.

In certain embodiments, treatment with the compositions of the invention results in a reduction in the swelling of joints. In certain embodiments, the compositions of the invention are for use in patients with swollen joints or patients identified as at risk of having swollen joints. In certain embodiments, the compositions of the invention are for use in a method of reducing joint swelling in RA.

In certain embodiments, treatment with the compositions of the invention results in a reduction in cartilage damage or bone damage. In certain embodiments, the compositions of the invention are for use in reducing or preventing cartilage or bone damage in the treatment of RA. In certain embodiments, the compositions are for use in treating patient with severe RA that are at risk of cartilage or bone damage.

Increased IL-17 levels and Th17 cell numbers are associated with cartilage and bone destruction in RA [28,29]. IL-17 is known to activate matrix destruction in cartilage and bone tissue and IL-17 has an inhibitory effect on matrix production in chondrocytes and osteoblasts. Therefore, in certain embodiments, the compositions of the invention are for use in preventing bone erosion or cartilage damage in the treatment of RA. In certain embodiments, the compositions are for use in treating patients that exhibit bone erosion or cartilage damage or patients identified as at risk of bone erosion or cartilage damage.

TNF- α is also associated with RA, but TNF- α is not involved in the pathogenesis of the later stages of the disease. In contrast, IL-17 has a role throughout all stages of chronic disease [30]. Therefore, in certain embodiments the compositions of the invention are for use in treating chronic RA or late-stage RA, such as disease that includes joint destruction and loss of cartilage. In certain embodiments, the compositions of the invention are for treating patients that have previously received anti-TNF- α therapy. In certain embodiments, the patients to be treated do not respond or no longer respond to anti-TNF- α therapy.

The compositions of the invention may be useful for modulating a patient's immune system, so in certain embodiments the compositions of the invention are for use in preventing RA in a patient that has been identified as at risk of RA, or that has been diagnosed with early-stage RA. The compositions of the invention may be useful for preventing the development of RA.

The compositions of the invention may be useful for managing or alleviating RA. The compositions of the invention may be particularly useful for reducing symptoms associated with joint swelling or

bone destruction. Treatment or prevention of RA may refer to, for example, an alleviation of the severity of symptoms or a reduction in the frequency of exacerbations or the range of triggers that are a problem for the patient.

Multiple sclerosis

5 In preferred embodiments, the compositions of the invention are for use in treating or preventing multiple sclerosis. The examples demonstrate that the compositions of the invention achieve a reduction in the disease incidence and disease severity in a mouse model of multiple sclerosis (the EAE model), and so they may be useful in the treatment or prevention of multiple sclerosis. Multiple sclerosis is an inflammatory disorder associated with damage to the myelin sheaths of neurons,
10 particularly in the brain and spinal column. Multiple sclerosis is a chronic disease, which is progressively incapacitating and which evolves in episodes. IL-17 and Th17 cells may have a key role in multiple sclerosis, for example because IL-17 levels may correlate with multiple sclerosis lesions, IL-17 can disrupt blood brain barrier endothelial cell tight junctions, and Th17 cells can migrate into the central nervous system and cause neuronal loss [31,32]. Therefore, the compositions
15 of the invention may be particularly effective for preventing or treating multiple sclerosis.

In certain embodiments, treatment with the compositions of the invention results in a reduction in disease incidence or disease severity. In certain embodiments, the compositions of the invention are for use in reducing disease incidence or disease severity. In certain embodiments, treatment with the compositions of the invention prevents a decline in motor function or results in improved motor
20 function. In certain embodiments, the compositions of the invention are for use in preventing a decline in motor function or for use in improving motor function. In certain embodiments, treatment with the compositions of the invention prevents the development of paralysis. In certain embodiments, the compositions of the invention are for use in preventing paralysis in the treatment of multiple sclerosis.

25 The compositions of the invention may be useful for modulating a patient's immune system, so in certain embodiments the compositions of the invention are for use in preventing multiple sclerosis in a patient that has been identified as at risk of multiple sclerosis, or that has been diagnosed with early-stage multiple sclerosis or "relapsing-remitting" multiple sclerosis. The compositions of the invention may be useful for preventing the development of sclerosis. Indeed, the examples show that
30 administration of compositions of the invention prevented the development of disease in many mice.

The compositions of the invention may be useful for managing or alleviating multiple sclerosis. The compositions of the invention may be particularly useful for reducing symptoms associated with multiple sclerosis. Treatment or prevention of multiple sclerosis may refer to, for example, an alleviation of the severity of symptoms or a reduction in the frequency of exacerbations or the range
35 of triggers that are a problem for the patient.

Modes of administration

Preferably, the compositions of the invention are to be administered to the gastrointestinal tract in order to enable delivery to and / or partial or total colonisation of the intestine with the bacterial strain of the invention. Generally, the compositions of the invention are administered orally, but they
5 may be administered rectally, intranasally, or via buccal or sublingual routes.

In certain embodiments, the compositions of the invention may be administered as a foam, as a spray or a gel.

In certain embodiments, the compositions of the invention may be administered as a suppository, such as a rectal suppository, for example in the form of a theobroma oil (cocoa butter), synthetic hard
10 fat (e.g. suppcire, witepsol), glycerol-gelatin, polyethylene glycol, or soap glycerin composition.

In certain embodiments, the composition of the invention is administered to the gastrointestinal tract via a tube, such as a nasogastric tube, orogastric tube, gastric tube, jejunostomy tube (J tube), percutaneous endoscopic gastrostomy (PEG), or a port, such as a chest wall port that provides access to the stomach, jejunum and other suitable access ports.

15 The compositions of the invention may be administered once, or they may be administered sequentially as part of a treatment regimen. In certain embodiments, the compositions of the invention are to be administered daily.

In certain embodiments of the invention, treatment according to the invention is accompanied by assessment of the patient's gut microbiota. Treatment may be repeated if delivery of and / or partial
20 or total colonisation with the strain of the invention is not achieved such that efficacy is not observed, or treatment may be ceased if delivery and / or partial or total colonisation is successful and efficacy is observed.

In certain embodiments, the composition of the invention may be administered to a pregnant animal, for example a mammal such as a human in order to prevent an inflammatory or autoimmune disease
25 developing in her child *in utero* and / or after it is born.

The compositions of the invention may be administered to a patient that has been diagnosed with a disease or condition mediated by IL-17 or the Th17 pathway, or that has been identified as being at risk of a disease or condition mediated by IL-17 or the Th17 pathway. The compositions may also be administered as a prophylactic measure to prevent the development of diseases or conditions
30 mediated by IL-17 or the Th17 pathway in a healthy patient.

The compositions of the invention may be administered to a patient that has been identified as having an abnormal gut microbiota. For example, the patient may have reduced or absent colonisation by *Roseburia*, and in particular *Roseburia hominis*.

The compositions of the invention may be administered as a food product, such as a nutritional supplement.

Generally, the compositions of the invention are for the treatment of humans, although they may be used to treat animals including monogastric mammals such as poultry, pigs, cats, dogs, horses or rabbits. The compositions of the invention may be useful for enhancing the growth and performance of animals. If administered to animals, oral gavage may be used.

Compositions

Generally, the composition of the invention comprises bacteria. In preferred embodiments of the invention, the composition is formulated in freeze-dried form. For example, the composition of the invention may comprise granules or gelatin capsules, for example hard gelatin capsules, comprising a bacterial strain of the invention.

Preferably, the composition of the invention comprises lyophilised bacteria. Lyophilisation of bacteria is a well-established procedure and relevant guidance is available in, for example, references [33-35].

Alternatively, the composition of the invention may comprise a live, active bacterial culture.

In preferred embodiments, the composition of the invention is encapsulated to enable delivery of the bacterial strain to the intestine. Encapsulation protects the composition from degradation until delivery at the target location through, for example, rupturing with chemical or physical stimuli such as pressure, enzymatic activity, or physical disintegration, which may be triggered by changes in pH.

Any appropriate encapsulation method may be used. Exemplary encapsulation techniques include entrapment within a porous matrix, attachment or adsorption on solid carrier surfaces, self-aggregation by flocculation or with cross-linking agents, and mechanical containment behind a microporous membrane or a microcapsule. Guidance on encapsulation that may be useful for preparing compositions of the invention is available in, for example, references [36] and [37].

The composition may be administered orally and may be in the form of a tablet, capsule or powder. Encapsulated products are preferred because *Roseburia* are anaerobes. Other ingredients (such as vitamin C, for example), may be included as oxygen scavengers and prebiotic substrates to improve the delivery and / or partial or total colonisation and survival *in vivo*. Alternatively, the probiotic composition of the invention may be administered orally as a food or nutritional product, such as milk or whey based fermented dairy product, or as a pharmaceutical product.

The composition may be formulated as a probiotic.

A composition of the invention includes a therapeutically effective amount of a bacterial strain of the invention. A therapeutically effective amount of a bacterial strain is sufficient to exert a beneficial

effect upon a patient. A therapeutically effective amount of a bacterial strain may be sufficient to result in delivery to and / or partial or total colonisation of the patient's intestine.

A suitable daily dose of the bacteria, for example for an adult human, may be from about 1×10^3 to about 1×10^{11} colony forming units (CFU); for example, from about 1×10^7 to about 1×10^{10} CFU; in another example from about 1×10^6 to about 1×10^{10} CFU.

In certain embodiments, the composition contains the bacterial strain in an amount of from about 1×10^6 to about 1×10^{11} CFU/g, respect to the weight of the composition; for example, from about 1×10^8 to about 1×10^{10} CFU/g. The dose may be, for example, 1 g, 3g, 5g, and 10g.

Typically, a probiotic, such as the composition of the invention, is optionally combined with at least one suitable prebiotic compound. A prebiotic compound is usually a non-digestible carbohydrate such as an oligo- or polysaccharide, or a sugar alcohol, which is not degraded or absorbed in the upper digestive tract. Known prebiotics include commercial products such as inulin and transgalacto-oligosaccharides.

In certain embodiments, the probiotic composition of the present invention includes a prebiotic compound in an amount of from about 1 to about 30% by weight, respect to the total weight composition, (e.g. from 5 to 20% by weight). Carbohydrates may be selected from the group consisting of: fructo- oligosaccharides (or FOS), short-chain fructo-oligosaccharides, inulin, isomalt-oligosaccharides, pectins, xylo-oligosaccharides (or XOS), chitosan-oligosaccharides (or COS), beta-glucans, arable gum modified and resistant starches, polydextrose, D-tagatose, acacia fibers, carob, oats, and citrus fibers. In one aspect, the prebiotics are the short-chain fructo-oligosaccharides (for simplicity shown herein below as FOSs-c.c); said FOSs-c.c. are not digestible carbohydrates, generally obtained by the conversion of the beet sugar and including a saccharose molecule to which three glucose molecules are bonded.

The compositions of the invention may comprise pharmaceutically acceptable excipients or carriers. Examples of such suitable excipients may be found in the reference [38]. Acceptable carriers or diluents for therapeutic use are well known in the pharmaceutical art and are described, for example, in reference [39]. Examples of suitable carriers include lactose, starch, glucose, methyl cellulose, magnesium stearate, mannitol, sorbitol and the like. Examples of suitable diluents include ethanol, glycerol and water. The choice of pharmaceutical carrier, excipient or diluent can be selected with regard to the intended route of administration and standard pharmaceutical practice. The pharmaceutical compositions may comprise as, or in addition to, the carrier, excipient or diluent any suitable binder(s), lubricant(s), suspending agent(s), coating agent(s), solubilising agent(s). Examples of suitable binders include starch, gelatin, natural sugars such as glucose, anhydrous lactose, free-flow lactose, beta-lactose, corn sweeteners, natural and synthetic gums, such as acacia, tragacanth or sodium alginate, carboxymethyl cellulose and polyethylene glycol. Examples of suitable lubricants

include sodium oleate, sodium stearate, magnesium stearate, sodium benzoate, sodium acetate, sodium chloride and the like. Preservatives, stabilizers, dyes and even flavouring agents may be provided in the pharmaceutical composition. Examples of preservatives include sodium benzoate, sorbic acid and esters of p-hydroxybenzoic acid. Antioxidants and suspending agents may be also used.

The compositions of the invention may be formulated as a food product. For example, a food product may provide nutritional benefit in addition to the therapeutic effect of the invention, such as in a nutritional supplement. Similarly, a food product may be formulated to enhance the taste of the composition of the invention or to make the composition more attractive to consume by being more similar to a common food item, rather than to a pharmaceutical composition. In certain embodiments, the composition of the invention is formulated as a milk-based product. The term "milk-based product" means any liquid or semi-solid milk- or whey- based product having a varying fat content. The milk-based product can be, e.g., cow's milk, goat's milk, sheep's milk, skimmed milk, whole milk, milk recombined from powdered milk and whey without any processing, or a processed product, such as yoghurt, curdled milk, curd, sour milk, sour whole milk, butter milk and other sour milk products. Another important group includes milk beverages, such as whey beverages, fermented milks, condensed milks, infant or baby milks; flavoured milks, ice cream; milk-containing food such as sweets.

In certain embodiments, the compositions of the invention contain a single bacterial strain or species and do not contain any other bacterial strains or species. Such compositions may comprise only *de minimis* or biologically irrelevant amounts of other bacterial strains or species. Such compositions may be a culture that is substantially free from other species of organism.

The compositions for use in accordance with the invention may or may not require marketing approval.

In some cases, the lyophilised bacterial strain is reconstituted prior to administration. In some cases, the reconstitution is by use of a diluent described herein.

The compositions of the invention can comprise pharmaceutically acceptable excipients, diluents or carriers.

In certain embodiments, the invention provides a pharmaceutical composition comprising: a bacterial strain of the invention; and a pharmaceutically acceptable excipient, carrier or diluent; wherein the bacterial strain is in an amount sufficient to treat a disorder when administered to a subject in need thereof; and wherein the disorder is selected from the group consisting of asthma, allergic asthma, neutrophilic asthma, osteoarthritis, psoriatic arthritis, juvenile idiopathic arthritis, neuromyelitis optica (Devic's disease), ankylosing spondylitis, spondyloarthritis, systemic lupus erythematosus,

celiac disease, chronic obstructive pulmonary disease (COPD), cancer, breast cancer, colon cancer, lung cancer, ovarian cancer, uveitis, scleritis, vasculitis, Behcet's disease, atherosclerosis, atopic dermatitis, emphysema, periodontitis, allergic rhinitis, and allograft rejection.

5 In certain embodiments, the invention provides pharmaceutical composition comprising: a bacterial strain of the invention; and a pharmaceutically acceptable excipient, carrier or diluent; wherein the bacterial strain is in an amount sufficient to treat or prevent a disease or condition mediated by IL-17 or the Th17 pathway. In preferred embodiments, said disease or condition is selected from the group consisting of rheumatoid arthritis, multiple sclerosis, psoriasis, inflammatory bowel disease, Crohn's disease, ulcerative colitis, celiac disease, asthma, allergic asthma, neutrophilic asthma, osteoarthritis, 10 psoriatic arthritis, juvenile idiopathic arthritis, neuromyelitis optica (Devic's disease), ankylosing spondylitis, spondyloarthritis, systemic lupus erythematosus, chronic obstructive pulmonary disease (COPD), cancer, breast cancer, colon cancer, lung cancer, ovarian cancer, uveitis, scleritis, vasculitis, Behcet's disease, atherosclerosis, atopic dermatitis, emphysema, periodontitis, allergic rhinitis, and allograft rejection.

15 In certain embodiments, the invention provides the above pharmaceutical composition, wherein the amount of the bacterial strain is from about 1×10^3 to about 1×10^{11} colony forming units per gram with respect to a weight of the composition.

In certain embodiments, the invention provides the above pharmaceutical composition, wherein the composition is administered at a dose of 1 g, 3 g, 5 g or 10 g.

20 In certain embodiments, the invention provides the above pharmaceutical composition, wherein the composition is administered by a method selected from the group consisting of oral, rectal, subcutaneous, nasal, buccal, and sublingual.

In certain embodiments, the invention provides the above pharmaceutical composition, comprising a carrier selected from the group consisting of lactose, starch, glucose, methyl cellulose, magnesium 25 stearate, mannitol and sorbitol.

In certain embodiments, the invention provides the above pharmaceutical composition, comprising a diluent selected from the group consisting of ethanol, glycerol and water.

In certain embodiments, the invention provides the above pharmaceutical composition, comprising an excipient selected from the group consisting of starch, gelatin, glucose, anhydrous lactose, free-flow lactose, beta-lactose, corn sweetener, acacia, tragacanth, sodium alginate, carboxymethyl 30 cellulose, polyethylene glycol, sodium oleate, sodium stearate, magnesium stearate, sodium benzoate, sodium acetate and sodium chloride.

In certain embodiments, the invention provides the above pharmaceutical composition, further comprising at least one of a preservative, an antioxidant and a stabilizer.

In certain embodiments, the invention provides the above pharmaceutical composition, comprising a preservative selected from the group consisting of sodium benzoate, sorbic acid and esters of p-hydroxybenzoic acid.

In certain embodiments, the invention provides the above pharmaceutical composition, wherein said bacterial strain is lyophilised.

In certain embodiments, the invention provides the above pharmaceutical composition, wherein when the composition is stored in a sealed container at about 4°C or about 25°C and the container is placed in an atmosphere having 50% relative humidity, at least 80% of the bacterial strain as measured in colony forming units, remains after a period of at least about: 1 month, 3 months, 6 months, 1 year, 1.5 years, 2 years, 2.5 years or 3 years.

Culturing methods

The bacterial strains for use in the present invention can be cultured using standard microbiology techniques as detailed in, for example, references [40-42].

The solid or liquid medium used for culture may be YCFA agar or YCFA medium. YCFA medium may include (per 100ml, approximate values): Casitone (1.0 g), yeast extract (0.25 g), NaHCO₃ (0.4 g), cysteine (0.1 g), K₂HPO₄ (0.045 g), KH₂PO₄ (0.045 g), NaCl (0.09 g), (NH₄)₂SO₄ (0.09 g), MgSO₄ · 7H₂O (0.009 g), CaCl₂ (0.009 g), resazurin (0.1 mg), hemin (1 mg), biotin (1 µg), cobalamin (1 µg), *p*-aminobenzoic acid (3 µg), folic acid (5 µg), and pyridoxamine (15 µg).

Bacterial strains for use in vaccine compositions

The inventors have identified that the bacterial strains of the invention are useful for treating or preventing diseases or conditions mediated by IL-17 or the Th17 pathway. This is likely to be a result of the effect that the bacterial strains of the invention have on the host immune system. Therefore, the compositions of the invention may also be useful for preventing diseases or conditions mediated by IL-17 or the Th17 pathway, when administered as vaccine compositions. In certain such embodiments, the bacterial strains of the invention may be killed, inactivated or attenuated. In certain such embodiments, the compositions may comprise a vaccine adjuvant. In certain embodiments, the compositions are for administration via injection, such as via subcutaneous injection.

General

The practice of the present invention will employ, unless otherwise indicated, conventional methods of chemistry, biochemistry, molecular biology, immunology and pharmacology, within the skill of

the art. Such techniques are explained fully in the literature. See, *e.g.*, references [43] and [44-50], *etc.*

The term “comprising” encompasses “including” as well as “consisting” *e.g.* a composition “comprising” X may consist exclusively of X or may include something additional *e.g.* X + Y.

5 The term “about” in relation to a numerical value x is optional and means, for example, $x \pm 10\%$.

The word “substantially” does not exclude “completely” *e.g.* a composition which is “substantially free” from Y may be completely free from Y. Where necessary, the word “substantially” may be omitted from the definition of the invention.

10 References to a percentage sequence identity between two nucleotide sequences means that, when aligned, that percentage of nucleotides are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in section 7.7.18 of ref. [51]. A preferred alignment is determined by the Smith-Waterman homology search algorithm using an affine gap search with a gap open penalty of 12 and a gap extension penalty of 2, BLOSUM matrix of 62. The Smith-
15 Waterman homology search algorithm is disclosed in ref. [52].

Unless specifically stated, a process or method comprising numerous steps may comprise additional steps at the beginning or end of the method, or may comprise additional intervening steps. Also, steps may be combined, omitted or performed in an alternative order, if appropriate.

20 Various embodiments of the invention are described herein. It will be appreciated that the features specified in each embodiment may be combined with other specified features, to provide further embodiments. In particular, embodiments highlighted herein as being suitable, typical or preferred may be combined with each other (except when they are mutually exclusive).

MODES FOR CARRYING OUT THE INVENTION

Example 1 – Efficacy of bacterial inocula in a mouse model of house dust mite-induced asthma

25 Summary

Mice were administered with compositions comprising bacterial strains according to the invention and were subsequently challenged with house dust mite (HDM) extract to elicit an allergic inflammatory response. The inflammatory response to HDM includes eosinophilic and neutrophilic components, is mediated by IL-17 and the Th17 pathway, and is a model for asthma. The magnitude
30 and characteristics of the inflammatory response exhibited by mice treated with compositions of the invention were compared to control groups. The compositions of the invention were found to alleviate the inflammatory response, and to reduce recruitment of eosinophils and neutrophils,

indicating that they may be useful for treating IL-17- and Th17-mediated conditions such as eosinophilia, neutrophilia and asthma.

Strain

433: *Roseburia hominis*

Study design

Groups:

1. Negative control group. Treatment with vehicle control (per oral).
2. Treatment with therapeutic bacteria inoculum strain 433 (per oral).
7. Positive control group. Treatment with Dexamethasone (i.p.).
8. Untreated Control Group.

Number of mice per group = 5

Day -14 to day 13: Daily administration of vehicle control per oral (Group 1).

Day -14 to day 13: Daily administration of therapeutic bacteria inoculum per oral (Group 2-6).

Day 0, 2, 4, 7, 9, 11 Administration of 15ug HDM (house dust mite extract – Catalogue number: XPB70D3A25, Lot number: 231897, Greer Laboratories, Lenoir, NC, USA) in a volume of 30ul PBS per nasal (Group 1-8).

Day 0, 2, 4, 7, 9, 11 Administration of Dexamethasone (i.p., 3mg/kg, Sigma-Aldrich, Catalogue number D1159) (Group 7).

Day 14 Sacrifice of all animals for analysis.

Total number of mice = 40.

Endpoints and analysis

On day 14 animals were sacrificed by lethal intraperitoneal injection with pentobarbitol (Streuli Pharma AG, Uznach, Cat: 1170139A) immediately followed by a bronchoalveolar lavage (BAL).

Cells were isolated from the BAL (bronchoalveolar lavage) fluid and differential cell counts performed (200 cell counts/ samples).

Material and Methods

Mice. Female 7 week old BALB/c mice were purchased from Charles River Laboratories and randomly allocated to cages totally 5 mice per cage (Ventilated cages sourced from Indulab AG, Gams, Switzerland Cage type: “The Sealsafe™ – IVC cage. Product number 1248L). Cages were labeled with study number, group number and experimental starting date. Mice were monitored weekly and acclimatized to facility for 7 days prior to initiation of study (Study Day -14). Animals were 8 weeks old on Study Day -14. Potable water and food were available ad libitum. Cage enrichment was present. Daily care of the animals was performed according to local authorization license number 2283.1 (issued and approved by: Service de la consommation et des affaires vétérinaires du Canton de Vaud). Potable water and food were available ad libitum and refreshed once daily. Cage enrichment was present. Animal welfare regulations were observed as given by

official authorities of Switzerland under ordinance 455.163 of the FVO (Federal Veterinary Office) on laboratory animal husbandry, production of genetically modified animals, and methods of animal experimentation.

Culturing of bacteria inoculum. Within a sterile workstation, a cryo-vial of bacteria was thawed by warming in gloved hand and ~0.7 ml of contents injected into a Hungate tube (Cat Number, 1020471, Glasgerätebau Ochs, Bovenden-Lenglern, Germany), containing 8 ml of anaerobic YCFA. Two tubes per strain were usually prepared. The Hungate tubes were then incubated (static) at 37°C for up to 24-26 hours (for strain 433).

Culturing of vehicle control. A Hungate tube containing 8 ml of anaerobic YCFA was incubated (static) at 37°C for 16h.

Administration of bacteria inoculum or vehicle control. 400ul of cultured bacteria inoculum or vehicle control were administered per day per oral gavage.

Intranasal sensitization. Mice were anesthetized by i.p. injection with 9.75 mg xylasol and 48.75 mg ketasol per kg (Dr. E. Graeub AG, Bern, Switzerland) and administered with 15ug of HDM (Catalogue number: XPB70D3A25, Lot number: 231897, Greer Laboratories, Lenoir, NC, USA) in a volume of 30ul PBS per nasal.

Preparation and administration of positive control compound Dexamethasone. Dexamethasone 21-phosphate disodium salt (Sigma-Aldrich, Catalogue number D1159, Lot N° SLBD.1030V) was solved in H₂O and administered to the animals in a dose of 3mg/kg in a volume of 200ul per oral at days indicated in study protocol above.

Terminal procedure. On day 14 animals were sacrificed by lethal i.p. injection with pentobarbitol (Streuli Pharma AG, Uznach, Cat: 1170139A) immediately followed by bronchoalveolar lavage (BAL) in 500 ul of saline.

Measurement of cellular infiltrates into BAL. Cells were isolated from the BAL fluid and differential cell counts were performed based upon standard morphological and cytochemical criteria.

Graphs and statistical analysis. All graphs were generated with Graphpad Prism Version 6 and a one-way ANOVA was applied. Results from the statistical analysis were provided with the individual data tables. Error bars represent Standard Error of the Mean (SEM).

Results and analysis

The results of the experiments are shown in Figures 1-9.

No morbidity or mortality was noted in the mice treated with the bacteria or the vehicle. The two controls, vehicle treatment (negative control) and the dexamethasone treatment (positive control) behaved as expected, with impaired eosinophilia and neutrophilia noted following dexamethasone treatment.

Strain 433 was efficacious in alleviating the magnitude of the allergic inflammatory response. As shown in Figures 2 and 3, administration of strain 433 reduced total eosinophils and the proportion of eosinophils in BAL, which indicates reduced eosinophilia. In addition, administration of strain 433 resulted in a statistically significant reduction in total neutrophils and the proportion of neutrophils in BAL relative to the vehicle-only control, as shown in Figures 6 and 7.

Example 2 – Efficacy of bacterial inocula in a mouse model of severe neutrophilic asthma

Summary

Mice were administered with compositions comprising bacterial strains according to the invention and were subsequently sensitised with subcutaneous administrations of house dust mite (HDM) extract and challenged with an intranasal administration of HDM in order to model the inflammatory response of severe neutrophilic asthma. The magnitude and characteristics of the inflammatory response exhibited by mice treated with compositions of the invention were compared to control groups. The compositions of the invention were found to alleviate the inflammatory response, and in particular to reduce recruitment of neutrophils, in a manner comparable to the positive control comprising administrations of anti-IL-17 antibodies. The data therefore indicate that the compositions of the invention may be useful for treating IL-17- and Th17-mediated conditions such as neutrophilia and asthma.

Strain

433: *Roseburia hominis*

Study design

Groups:

1. Negative control group. Treatment with vehicle control (per oral).
2. Treatment with therapeutic bacteria inoculum strain 433 (per oral).
7. Positive control group. Treatment anti-IL-17 (i.p.).
8. Untreated Control Group.

9: Healthy mice (baseline).

Number of mice per group (Group 1-8) = 5

Day -14 to day 17: Daily administration of vehicle control per oral (Group 1).

Day -14 to day 17: Daily administration of therapeutic bacteria inoculum per oral (Group 2-6).

Day 0: Sensitization with HDM in CFA (s.c.) (Group 1-8).

Day 7: Sensitization with HDM in CFA (s.c.) (Group 1-8).

Day 13, 15, 17: Administration of anti IL-17 neutralizing antibody per i.p. (Group 7).

Day 14, 15, 16, 17: Challenge with HDM in 30ul PBS per nasal (Group 1-8).

Day 18: Sacrifice of all animals for analysis.

Endpoints and analysis:

- 5 On day 14 animals were sacrificed by lethal intraperitoneal injection with pentobarbitol (Streuli Pharma AG, Uznach, Cat: 1170139A) immediately followed by a bronchoalveolar lavage (BAL). Cells were isolated from the BAL fluid and differential cell counts performed (200 cell counts/samples).

Material and Methods.

- 10 **Mice.** Female 7 week old C57BL/6 mice were purchased from Charles River Laboratories and randomly allocated to cages totally 5 mice per cage (Ventilated cages sourced from Indulab AG, Gams, Switzerland Cage type: “The Sealsafe™ – IVC cage. Product number 1248L). Cages were labelled with study number, group number and experimental starting date. Mice were monitored weekly and acclimatized to facility for 7 days prior to initiation of study (Study Day -14). Animals
15 were 8 weeks old on Study Day -14. Potable water and food were available ad libitum. Cage enrichment was present. Daily care of the animals was performed according to local authorization license number 2283.1 (issued and approved by: Service de la consommation et des affaires vétérinaires du Canton de Vaud). Potable water and food were available ad libitum and refreshed once daily. Cage enrichment was present. Animal welfare regulations were observed as given by
20 official authorities of Switzerland under ordinance 455.163 of the FVO (Federal Veterinary Office) on laboratory animal husbandry, production of genetically modified animals, and methods of animal experimentation.

- Culturing of bacteria inoculum.** Within a sterile workstation, a cryo-vial of bacteria was thawed by warming in gloved hand and ~0.7 ml of contents injected into a Hungate tube (Cat Number,
25 1020471, Glasgerätebau Ochs, Bovenden-Lengler, Germany), containing 8 ml of anaerobic YCFA. Two tubes per strain were usually prepared. The Hungate tubes were then incubated (static) at 37°C for up to 24-26 hours (for strain 433).

Culturing of vehicle control. A Hungate tube containing 8 ml of anaerobic YCFA was incubated (static) at 37°C for 16h.

- 30 **Administration of bacteria inoculum or vehicle control.** 400ul of cultured bacteria inoculum or vehicle control were administered per day per oral gavage.

HDM sensitization. 50 µg of HDM (Catalogue number: XPB70D3A25, Lot number: 231897, Greer Laboratories, Lenoir, NC, USA) in PBS was emulsified in equal volume of complete Freund’s adjuvant (CFA Chondrex Inc. Washington, USA) and administered subcutaneously in a volume of

200 µl, twice over two weeks on opposite flanks. A week after the second immunization, mice were anesthetized by i.p. injection with 9.75 mg xylasol and 48.75 mg ketasol per kg (Dr. E. Graeb AG, Bern, Switzerland) and then given intranasal challenges of 15 µg of HDM in a volume of 30 µl PBS on 4 consecutive days. Analysis was performed one day after the final challenge.

- 5 **Preparation and administration of positive control compound anti mouse IL-17 antibody.** Anti-IL-17 neutralizing antibody was sourced from Bio X Cell and was stored at 4°C (Clone 17F3, Cat. Number BE0173, Bio X Cell) and administered per i.p. at a dose of 12.5 mg/kg at days indicated in study protocol above.

- 10 **Terminal procedure.** On day 18 animals were sacrificed by lethal i.p. injection with pentobarbital (Streuli Pharma AG, Uznach, Cat: 1170139A) immediately followed by bronchoalveolar lavage (BAL) in 500 µl of saline.

Measurement of cellular infiltrates into BAL. Cells were isolated from the BAL fluid and differential cell counts were performed based upon standard morphological and cytochemical criteria.

- 15 **Graphs and statistical analysis.** All graphs were generated with Graphpad Prism Version 6 and a one-way ANOVA was applied. Results from the statistical analysis are provided with the individual data tables. Error bars represent Standard Error of the Mean (SEM).

Results and analysis

The results of the experiment are shown in Figures 10-18.

- 20 No morbidity or mortality was noted in the mice treated with the bacteria or the vehicle. As shown in Figures 11, 12, 15 and 16, certain mice treated with strain 433 exhibited reduced eosinophilia and neutrophilia.

Example 3 – Efficacy of bacterial inocula to treat arthritis in a type II collagen-induced arthritis mouse model

- 25 Materials and methods

Strain

433: *Roseburia hominis*

Bacterial cultures

- 30 Bacterial cultures were grown up for administration in an anaerobic workstation (Don Whitley Scientific).

Bacterial strain #433 was grown using glycerol stocks. The glycerol stocks were stored at -80°C. Three times per week, glycerol stocks were thawed at room temperature and streaked on YCFA plates. A new glycerol aliquot was used on each occasion. Bacteria were allowed to grow on a given plate for up to 72 hours.

Solutions to be administered to the animals were prepared twice daily with an eight hour interval for morning (AM) and afternoon (PM) treatments. A bacterial colony was picked from the streaked plate and transferred into a tube containing YCFA media. Bacterial strain #433 was allowed to grow for 24 hours before AM administrations. Bacteria were sub-cultured at 1% into YCFA media for PM administrations. OD values were recorded for each strain after morning and afternoon treatment preparations.

Type II collagen-induced arthritis mouse model

Adult male DBA/1 mice were randomly allocated to experimental groups and allowed to acclimatise for two weeks. On Day 0, animals were administered by subcutaneous injection with 100 microliters of an emulsion containing 100 micrograms of type II collagen (CII) in incomplete's Freund's adjuvant supplemented with 4 mg/ml *Mycobacterium tuberculosis* H37Ra. On Day 21, animals were administered by subcutaneous injection with a booster emulsion containing 100 µg of type II collagen in incomplete Freund's adjuvant.

Treatments were given according to the administration schedule below. From Day -14 until the end of the experiment on Day 45, animals were weighed three times per week. From Day 21 until the end of the experiment, animals were scored three times per week for clinical signs of arthritis to include swelling of the hind- and front paws, radio-carpal (wrist) joints and tibio-tarsal (ankle) joints.

On Day 45 mice were culled and terminal blood samples were taken for cytokine analysis.

On Day -14, Day 0 and Day 45, faecal samples were collected for microbiological analysis, immediately snap-frozen and stored at -80°C.

The collagen-induced arthritis (CIA) mouse model is a well-established mouse model for rheumatoid arthritis [53]. Immunisation with CII causes a pathogenesis that includes several important pathological features of rheumatoid arthritis, including synovial hyperplasia, mononuclear cell infiltration and cartilage degradation. Significantly, the development of CIA is mediated by Th17 cells through secretion of IL-17A [54]. The immune response underlying the arthritis model is enhanced by the use of Freund's adjuvant supplemented with *Mycobacterium tuberculosis*.

On Day 21, spleens were collected from three satellite animals in each group. Cells were cultured for 72 hours in the presence or absence of type II collagen. Cytokines, including TNF-α, IL-6, IFN-γ, IL-

4, IL-10 and IL-17, were quantified in the culture supernatants and in terminal serum by Luminex. Cell proliferation was quantified using a tritiated thymidine incorporation method.

Treatment Groups and Dosages

All Groups were n=15 (n=12 for the main study group and n=3 for satellite groups)

5 The vehicle used for the biotherapeutics was Yeast extract-Casitone-Fatty Acids (YCFA) medium.

	<i>Group</i>	<i>Dose</i>	<i>Administration</i>		<i>Disease Induction</i>
			<i>Route</i>	<i>Regimen</i>	
1	<i>Vehicle</i>	5 ml/kg	<i>PO</i>	<i>BID: Day -14-End</i>	<i>Day 0: Collagen/CFA, once, SC</i>
6	<i>Biotherapeutic #433</i>	5 ml/kg			<i>Day 21: Collagen/IFA, once, SC</i>

PO: oral gavage, SC: subcutaneous injection, BID: twice a day, CFA: complete Freund's adjuvant.

Bodyweights

10 From Day -14 until the end of the experiment, animals were weighed three times per week. Data were graphed (Mean $\pm$ SEM).

Non-specific clinical observations

From Day -14 until the end of the experiment, animals were checked daily for non-specific clinical signs to include abnormal posture (hunched), abnormal coat condition (piloerection) and abnormal activity levels (reduced or increased activity).

15 Clinical Observations

20 From Day 21 until the end of the experiment on Day 45, animals were scored three times per week for clinical signs of arthritis to include swelling of the hind- and front paws, radio-carpal (wrist) joints and tibio-tarsal (ankle) joints. Each limb was scored using the following scale: (0) normal, (1) slight swelling, (2) mild swelling, (3) moderate swelling and (4) severe swelling. A clinical score was calculated by adding each limb score. The maximum possible clinical score for an animal was (16). Animals with a score equal to (12) on two consecutive occasions and animals with a score greater than (12) on any one occasion were culled. Data were graphed (Mean $\pm$ SEM).

Cell proliferation analysis

25 On Day 21, three satellite animals per group were culled and spleens were dissected out. Spleen cells were cultured for 72 hours in presence or absence of type II Collagen. After 72 hours, cells were

pulsed overnight in the presence of tritiated thymidine. Cell proliferation was quantified by measuring thymidine incorporation. Data were graphed (Mean $\pm$ SEM). Supernatants were taken and tested for the presence of key cytokines.

Cytokine analysis

- 5 Terminal supernatants from the spleen cell cultures were tested in order to quantitate TNF- α , IL-6, IFN- γ , IL-4, IL-10 and IL-17 by Luminex. Data were graphed (Mean $\pm$ SEM).

Microbiological analysis

- 10 On Day -14, Day 0 and Day 45, faecal samples were collected from each animal, immediately snap-frozen, and stored at -80°C. Caeca (including content) were immediately snap-frozen and stored at -80°C. A bacterial identification test was performed daily by plating the bacteria.

Histopathology

- 15 At the end of the experiment, hind paws were stored in tissue fixative. Samples were transferred into decalcification solution. Tissue samples were processed, sectioned and stained with Haematoxylin & Eosin. Sections were scored by a qualified histopathologist, blind to the experimental design, for signs of arthritis to include inflammation, articular cartilage damage and damage to the underlying metaphyseal bone. A detailed scoring system was used (see below). Data were graphed (Mean $\pm$ SEM). Raw and analysed data were provided as well as representative pictures.

Table 1: Histopathology Scoring System

Grade	Description
Inflammation	
0	Normal joint
1	Mild synovial hyperplasia with inflammation dominated by neutrophils. Low numbers of neutrophils and macrophages in joint space.
2	Synovial hyperplasia with moderate to marked inflammation involving both neutrophils and macrophages. Neutrophils and macrophages in joint space; may be some necrotic tissue debris.
3	Synovial hyperplasia with marked inflammation involving both neutrophils and macrophages. Loss of synoviocyte lining. Inflammation may extend from synovium to surrounding tissue including muscle. Numerous neutrophils and macrophages in joint space, together with significant necrotic tissue debris.
Articular cartilage damage	
0	Normal joint

- 1 Articular cartilage shows only mild degenerative change. Early pannus formation may be present peripherally.
- 2 Articular cartilage shows moderate degenerative change and focal loss. Pannus formation is present focally.
- 3 Significant disruption and loss of articular cartilage with extensive pannus formation.

Damage to the underlying metaphyseal bone

- 0 Normal joint
 - 1 No change to underlying metaphyseal bone.
 - 2 May be focal necrosis or fibrosis of metaphyseal bone.
 - 3 Disruption or collapse of metaphyseal bone. Extensive inflammation, necrosis or fibrosis extending to medullary space of the metaphysis.
-

Results and analysis
Survival and Non-specific Clinical Observations

Some animals were culled prior to the scheduled end of the study due to the severity of the clinical signs of arthritis or due to the severity of the non-specific clinical observations.

- 5 Two animals were culled or found dead during the pre-treatment period (Day -14 to Day 0): one animal in Group 1 (vehicle-treated, animal arrived from supplier with broken leg and was culled) and one animal in Group 6 (biotherapeutic #433-treated).

Ten animals were culled due to the severity of the clinical signs of arthritis: five animals in Group 1 (vehicle-treated) and five animals in Group 6 (biotherapeutic #433-treated).

- 10 Four animals were culled due to the severity of the non-specific clinical signs including abnormal posture (hunched), abnormal coat condition (piloerection), abnormal activity levels (reduced activity): three animals in Group 1 (vehicle-treated) and one animal in Group 6 (biotherapeutic #433-treated).

Bodyweights

- 15 Bodyweight data recorded from Day -14 until Day 0 and expressed as a percentage of the initial (Day -14) bodyweights were analysed by two-way ANOVA followed by Dunnett's post-test for multiple comparisons with Day -14 then for multiple comparison with the vehicle-treated group. The data are presented in Figure 19. Data from animals culled prior to the scheduled end of the experiment were excluded from the analyses.
- 20 When compared to Day -14, twice daily administrations by oral gavage induced a significant bodyweight loss in the vehicle-treated group on Day -9 and Day -7.

Bodyweight data recorded from Day 0 until Day 28 and expressed as a percentage of the initial (Day 0) bodyweights were analysed by two-way ANOVA followed by Dunnett's post-test for multiple comparisons with Day 0 in the Vehicle group then for multiple comparison with the vehicle-treated group. The data are presented in Figure 20. Data from animals culled prior to the scheduled end of the experiment and from Satellite animals were excluded from the analyses. Day 28, Day 35 and Day 42 data were further analysed by one-way ANOVA followed by Dunnett's post-test for multiple comparisons to the vehicle-treated group.

The onset of clinical signs of arthritis was associated with a significant bodyweight loss on Day 26 and Day 28 ($p < 0.0001$) when compared to Day 0 in the vehicle-treated group.

Clinical Observations

Clinical score data were analysed by two-way ANOVA followed by Dunnett's post-test for multiple comparisons between days in the vehicle-treated group then for multiple comparisons between experimental groups and the vehicle-treated group each day. The data are presented in Figure 21. Data recorded from animals culled prior to the end of the experiment were excluded from the analysis. When animals were culled due to the severity of the clinical signs of arthritis, the last recorded score was reported for the following days and used in the statistical analyses.

A significant increase of the clinical scores was observed in the vehicle-treated group from Day 28 until Day 45 ($p < 0.0001$) when compared to Day 21.

Biotherapeutic #433 induced a reduction of the clinical scores when compared to the vehicle-treated group from Day 28 until Day 45, although the difference was non-significant.

Cell proliferation analysis

To validate the assay, splenocytes were cultured in the presence of soluble anti-CD3 and anti-CD28 (anti-CD3/CD28) as positive control stimuli to confirm the proliferative potential of the cells.

Strong proliferative responses to anti-CD3/CD28 were seen in all experimental groups, showing cells were healthy, viable and able to respond to activation signals.

To test the proliferative response in presence of Collagen II (CII), splenocytes were cultured in the presence of CII at 50 $\mu\text{g/ml}$. Splenocyte proliferative response to CII were analysed by two-way ANOVA followed by Sydak's post-test for multiple comparisons between unstimulated and CII-stimulated splenocytes and one-way ANOVA followed by Dunnett's post-test for comparison of CII-stimulated response in different experimental groups with the vehicle-treated group. The data are presented in Figure 22.

CII induced a highly significant increase of ^3H -thymidine incorporation (cpm) when compared to the unstimulated splenocytes in the vehicle-treated group ($p < 0.0001$).

The groups treated with biotherapeutic #433 demonstrated significantly lower levels of CII-induced splenocyte proliferation than the vehicle-treated group.

5 Cytokine levels in tissue culture supernatants

Levels of each cytokine were measured in tissue culture supernatants derived from anti-CD3/CD28 stimulated cultures by luminex analysis. These showed robust responses for all cytokines measured (mean levels in vehicle group were as follows: IL-4 = 6,406 pg/ml; IL-6 = 306 pg/ml; IL-10 = 10,987 pg/ml; IL-17A = 11,447 pg/ml; IFN- γ = 15,581 pg/ml; TNF- α = 76 pg/ml).

10 The following sections summarise the data obtained from the Collagen II-stimulated cultures. Where applicable, statistical analyses of the differences between cytokine levels in supernatants of unstimulated and CII-stimulated splenocytes were conducted using two-way ANOVA followed by Sidak's post-test for multiple comparisons, while one-way ANOVA followed by Dunnett's post-test was used for comparison of CII-stimulated response in biotherapeutic-treated groups with the
15 vehicle-treated group. There was no significant difference in cytokine levels between the groups in both cases. This is likely due to the small sample size used ($n=3$).

In order to more accurately present the distribution of the data for the cytokines with substantial spread of the data, these are presented as scatter plots.

The group means of IL-4 in tissue culture supernatants after stimulation with CII were $<5\text{pg/ml}$.
20 These are not considered biologically significant and not included here. The group means of TNF- α in tissue culture supernatants after stimulation with collagen were below limit of quantitation.

Supernatant levels of IFN- γ (Figure 23)

Along with IL-17, IFN- γ is the major cytokine driving disease in the CIA model. The scatter plot in Figure 23 demonstrates IFN- γ levels after CII stimulation, with group median being higher for the
25 Vehicle-treated group compared to the biotherapeutic.

Supernatant levels of IL-17A (Figure 24)

Levels of IL-17A were 50pg/ml in CII-stimulated cultures for the Vehicle-treated group. The levels of this cytokine appeared to be lower in the biotherapeutic group compared to the Vehicle-treated.

Supernatant levels of IL-10 (Figure 25)

30 Levels of IL-10 in Vehicle-treated group were 13 pg/ml and 2.1 pg/ml for CII-stimulated, and media control cultures, respectively. Higher levels of IL-10 (which is an anti-inflammatory cytokine) for the

CII induced a highly significant increase of ^3H -thymidine incorporation (cpm) when compared to the unstimulated splenocytes in the vehicle-treated group ($p < 0.0001$).

The groups treated with biotherapeutic #433 demonstrated significantly lower levels of CII-induced splenocyte proliferation than the vehicle-treated group.

5 Cytokine levels in tissue culture supernatants

Levels of each cytokine were measured in tissue culture supernatants derived from anti-CD3/CD28 stimulated cultures by luminex analysis. These showed robust responses for all cytokines measured (mean levels in vehicle group were as follows: IL-4 = 6,406 pg/ml; IL-6 = 306 pg/ml; IL-10 = 10,987 pg/ml; IL-17A = 11,447 pg/ml; IFN- γ = 15,581 pg/ml; TNF- α = 76 pg/ml).

10 The following sections summarise the data obtained from the Collagen II-stimulated cultures. Where applicable, statistical analyses of the differences between cytokine levels in supernatants of unstimulated and CII-stimulated splenocytes were conducted using two-way ANOVA followed by Sidak's post-test for multiple comparisons, while one-way ANOVA followed by Dunnett's post-test was used for comparison of CII-stimulated response in biotherapeutic-treated groups with the
15 vehicle-treated group. There was no significant difference in cytokine levels between the groups in both cases. This is likely due to the small sample size used ($n=3$).

In order to more accurately present the distribution of the data for the cytokines with substantial spread of the data, these are presented as scatter plots.

The group means of IL-4 in tissue culture supernatants after stimulation with CII were $<5\text{pg/ml}$.
20 These are not considered biologically significant and not included here. The group means of TNF- α in tissue culture supernatants after stimulation with collagen were below limit of quantitation.

Supernatant levels of IFN- γ (Figure 23)

Along with IL-17, IFN- γ is the major cytokine driving disease in the CIA model. The scatter plot in Figure 23 demonstrates IFN- γ levels after CII stimulation, with group median being higher for the
25 Vehicle-treated group compared to the biotherapeutic.

Supernatant levels of IL-17A (Figure 24)

Levels of IL-17A were 50pg/ml in CII-stimulated cultures for the Vehicle-treated group. The levels of this cytokine appeared to be lower in the biotherapeutic group compared to the Vehicle-treated.

Supernatant levels of IL-10 (Figure 25)

30 Levels of IL-10 in Vehicle-treated group were 13 pg/ml and 2.1 pg/ml for CII-stimulated, and media control cultures, respectively. Higher levels of IL-10 (which is an anti-inflammatory cytokine) for the

vehicle-treated group may be expected because inflammation and pro-inflammatory cytokine induction could be accompanied by an anti-inflammatory feedback mechanism.

Supernatant levels of IL-6 (Figure 26)

Inflammatory cytokines such as IL-6 and TNF- α are not typically produced at high levels in anti-CII cultures. However, their levels may be altered as a result of immune modulation. Levels of IL-6 in CII-stimulated cultures were modest, reaching 10pg/ml. Although higher than in media control cultures, these differences were too small to provide rationale for performing statistical analyses.

Microbiological analysis

Bacterial growth was confirmed by measuring the optical density at 600 nm using a spectrophotometer. Bacterial identity was confirmed by comparing streaked plate pictures to reference pictures.

Following the improved bacterial preparation method, consistently high doses of bacterial strain were administered from Day -2 and Day -3 as indicated by the high OD values measured.

Faecal samples were collected and snap-frozen on Day -14, Day 0 and at termination.

Histopathology

The histopathology results are shown in Figures 65-69. As expected for this model, intra-individual and inter-individual variability was observed in terms of the presence/absence of arthritis or the severity of change present.

The nature of the pathology was as expected for this model, with extensive mixed chronic-active inflammation of the synovium and bursa extending to involve the peri-articular soft tissues (muscle, adipose tissue, dermal collagen). In the most severely affected joints there was articular cartilage degeneration and loss with intra-articular debris and inflammation and disruption of the joint and bone structure by fibrosis and inflammation.

The incidence of histopathological changes was: vehicle – 80% (16/20); Biotherapeutic #433 – 55% (12/22). Treatment with Biotherapeutic #433 reduced the incidence of histopathological scores in mouse hind limbs when compared to the vehicle-treated group (see Figures 65-68). Histopathology scores were analysed by one-way ANOVA for non-parametric data (Kruskal-Wallis test) followed by Dunn's post-test for multiple comparisons to the vehicle-treated group, although the reduction achieved by Biotherapeutic #433 was not statistically significant in this analysis. Biotherapeutic #433 induced a reduction of the joint inflammation scores observed in histopathology when compared to the vehicle-treated group. Biotherapeutic #433 induced a reduction of the cartilage damage scores observed in histopathology when compared to the vehicle-treated group. Biotherapeutic #433 induced a reduction of the bone damage scores observed in histopathology when compared to the

vehicle-treated group. Biotherapeutic #433 induced a reduction of the total histopathology scores when compared to the vehicle-treated group.

Summary

Increased clinical scores were observed from Day 28 after the first administration of type II collagen, as expected in this model of arthritis in DBA/1 mice. Biotherapeutic #433 was shown to be effective at treating arthritis in this model. Biotherapeutic #433 was effective for reducing the severity of the clinical scores and for reducing pathological disease in the joints, as demonstrated in the histopathological analysis.

Proliferative recall responses to Collagen II were seen in splenocyte cultures from all experimental groups. The collagen-specific response was significantly reduced following treatment with biotherapeutic #433 (Group 5).

Most of the T cell cytokines tested showed detectable increases between Collagen II-stimulated and media controls in the Vehicle-treated group. These increases were not as obvious in the biotherapeutic-treated group. This broadly supports the proliferative recall responses to Collagen II described above.

There was evidence of suppression of the Th1/Th17 axis, which is the pathogenic response in this model and in human RA. Correlation of reduced levels of cytokines with reduced proliferation is suggestive of immune modulation. There was no evidence that this modulation resulted either from enhanced levels of Th2 associated IL-4 or with increases in the immune modulating cytokine, IL-10.

Example 4 – Further analysis of the effect of bacterial inocula in the mouse model of house dust mite-induced asthma

The mice tested in Example 1 were subjected to further analyses to further characterise the effect of the compositions of the invention on the allergic asthma inflammatory response.

Materials and methods

Blood withdrawal and serum preparation on day 14. Blood samples of animals were collected via cardiac puncture. Serum was isolated from the blood sample by centrifugation for 5 min at 14000g and stored at -20 °C.

Organ removal on day 14. Collection of the left lung lobe in formalin for follow-on histological analysis. Collection of the right lung lobes (all remaining lobes) and removal of serum for snap freezing and follow-on analysis. Remaining BAL fluid was snap frozen for follow-on analysis.

Measurement of antibody levels in serum and BAL fluid

Total IgE and house-dust-mite (HDM) specific IgG1 antibody production were measured in the BAL

and serum by ELISA assay.

Isolation of lung and histological analysis

Left lung lobes were fixed in formalin followed by embedment in paraffin, sectioning, and staining with hematoxylin and eosin and PAS. Subsequent histological scoring was performed blinded as followed: Five random fields of view per sample were scored for inflammation (peribronchial infiltration and perivascular infiltration) and mucus production. Inflammatory infiltration was scored with the following grading system:

0 - normal

1 - mild inflammatory infiltrates

2 - moderate inflammatory infiltrates

3 - marked inflammatory infiltrates

4 - severe inflammatory infiltrates

5 - very severe inflammatory infiltrates

In each field of view, airways were measured in size and mucus cell numbers were quantified/ um.

Measurement of inflammatory mediators in lung tissue

Right lung lobes (all remaining lobes) isolated for quantification of inflammatory mediators were snap frozen for subsequent measurement of CCL11, IFN-gamma, IL-1 alpha, IL-1 beta, IL-4, IL-5, IL-9, IL-17A, CXCL1, CCL3, CXCL2 and CCL5 by commercially available multiplex assay (Merck-Millipore). Analysis was performed according to the manufacturer's instructions.

Results and analysis

The results of the experiments are shown in Figures 27-45.

In support of the findings described in Example 1, analysis of the cellular infiltrates in the lung tissue of mice treated with strain 433 showed a notable and statistically significant reduction in mean inflammation score (see Figures 31 and 33).

Antibody levels in the BAL fluid and serum were analysed (see Figures 27-30). No clear effect of the bacterial treatment on serum antibody levels was observed. This may reflect a failure in the experiment, because the spread of data and the error bars for each treatment are large, and the positive and negative controls do not appear to have behaved as would be expected. Also, the baseline serum antibody levels could have masked any changes.

Similarly, no clear effect of the bacterial treatment on cytokine levels in lung tissue was observed (see Figures 35-45). Again, this may reflect a failure in the experiment, because the spread of data and the error bars for each treatment are large, and the positive and negative controls do not appear to have behaved as would be expected. It is also possible that the mechanism of action involved influences earlier cytokine responses that were no longer detectable on day 4 post the final HDM airway challenge. Some care should be taken when interpreting the cytokine data in the current study, due to the variability in the levels detected. This variability could in part be explained by the fact that the lung tissue was separated for the different analyses, and thus one lung lobe might not have been fully representative or comparable to the same lobe in other mice due to patchy distribution of the inflammation.

Example 5 – Further analysis of the effect of bacterial inocula in the mouse model of severe neutrophilic asthma

The mice tested in Example 2 were subjected to further analyses to further characterise the effect of the compositions of the invention on the neutrophilic response associated with severe asthma.

Materials and methods

Organ removal on day 18. Collection of the left lung lobe in formalin for follow-on histological analysis. Collection of the right lung lobes (all remaining lobes) and removal of serum for snap freezing and follow-on analysis. Remaining BAL fluid was snap frozen for follow-on analysis.

Measurement of inflammatory mediators in lung tissue (follow-on analysis). Right lung lobes (all remaining lobes) isolated for quantification of inflammatory mediators were snap frozen for subsequent measurement of IFN-gamma, IL-1 alpha, IL-1 beta, CXCL1, CCL3, CXCL2, CCL5, IL-17A, TNF-alpha, IL-17F, IL-23 and IL-33 by commercially available multiplex assay (Merck-Millipore). Analysis was performed according to the manufacturer's instructions.

Measurement of antibody levels in serum and BAL fluid (follow-on analysis). House-dust-mite (HDM) specific IgG1 and IgG2a antibody production were measured in the BAL and serum by ELISA assay.

Isolation of lung and histological analysis (follow-on analysis). Left lung lobes were fixed in formalin followed by embedment in paraffin, sectioning, and staining with hematoxylin and eosin and PAS. Subsequent histological scoring was performed blinded as followed: Five random fields of view per sample were scored for inflammation (peribronchial infiltration and perivascular infiltration) and mucus production. Inflammatory infiltration was scored with the following grading system:

0 - normal

1 - mild inflammatory infiltrates

2 - moderate inflammatory infiltrates

3 - marked inflammatory infiltrates

4 - severe inflammatory infiltrates

5 5 - very severe inflammatory infiltrates

Results and analysis

The results of the experiments are shown in Figures 46-63.

Further analysis of antibody levels revealed that the efficacy of bacterial strain 433 was also reflected in reduced HDM-specific IgG1 levels in the BAL fluid and serum (see Figures 46 and 48). Firm
10 conclusions regarding an effect on IgG2a levels cannot be drawn. Overall, the data from the antibody analysis is suggestive of a reduction related to an overall reduced inflammatory response, as opposed to a selective effect on antibody isotype switching.

In relation to cytokine levels, as for Example 4, the spread of data and the error bars for each treatment are large, and the positive and negative controls do not appear to have behaved as
15 necessarily would be expected. It is also possible that the mechanism of action involves influencing earlier cytokine responses that were no longer detectable on day 4 post the final HDM airway challenge. Some care should be taken when interpreting the cytokine data in the current study, due to the variability in the levels detected. This variability could in part be explained by the fact that the lung tissue was separated for the different analyses, and thus one lung lobe might not have been fully
20 representative or comparable to the same lobe in other mice due to patchy distribution of the inflammation. Despite this variability, a clear anti-inflammatory effect on cytokine levels for strain 433 was shown, and the positive control anti-IL-17 Ab generally behaved as expected.

With the above caveats, the data in Figure 55 suggests that treatment with strain 433 may achieve a reduction in the levels of IFN γ , which may be indicative of a mechanism of action related to
25 influences on chemokine release (and thus recruitment of cells) by stromal or innate immune cells. IFN γ is involved in the Th17 pathway. Taking this dataset together, a clear conclusion can be drawn that strains 433 was effective at protecting mice against inflammation in this mouse model of severe neutrophilic asthma.

Example 6 – Efficacy of bacterial inocula in a mouse model of multiple sclerosis

Summary

Mice were administered with compositions comprising bacterial strains according to the invention and the mice were subsequently immunised with myelin oligodendrocyte glycoprotein to induce

experimental autoimmune encephalomyelitis (EAE). EAE is the most commonly used experimental model for human multiple sclerosis. The compositions of the invention were found to have a striking effect on disease incidence and disease severity.

Strain

5 433: bacteria deposited under accession number NCIMB 43283

Study design

Groups:

- 1. Negative control group. Treatment with vehicle control (per oral).
 - 5. Treatment with therapeutic bacteria inoculum strain 433 (per oral).
 - 10 9. Positive control group. Treatment with Dexamethasone (i.p.).
 - 10. Untreated Control Group.
- Number of mice per group = 10

Days -14 to day 27: Daily administration of vehicle control per oral (Group 1).

15 Days -14 to day 27: Daily administration of therapeutic bacteria inoculum per oral (Group 5).

Days 0-28: administration of Dexamethasone (i.p.) three times a week (Group 9)

Day 0: MOG35-55 (myelin oligodendrocyte glycoprotein - 2mg/ml) and CFA (2mg/ml MTB) were mixed 1:1 resulting in 1mg/ml solutions. 100µl of the peptide-CFA mixture was injected subcutaneously into each hind leg. Administration of pertussis toxin intraperitoneally (300ng).

20 Day 1: Administration of pertussis toxin intraperitoneally (300ng).

Days 7-onwards: Measurement of disease incidence and weight three times a week.

Endpoints and analysis

25 Mice were analysed for disease incidence and disease severity three times a week. Scoring was performed blind. Disease severity was assessed using a clinical score ranging from 0 to 5, with 5 indicating a dead mouse (see clinical scoring system below).

Monitoring

On the indicated days mice were weighed and observed for disease activity score and disease incidence.

30 Disease activity score observations:

0 - No obvious changes in motor function compared to non-immunized mice.

0.5 - Tip of tail is limp.

1.0 - Limp tail.

1.5 - Limp tail and hind leg inhibition.

35 2.0 - Limp tail and weakness of hind legs.

OR - There are obvious signs of head tilting when the walk is observed. The balance is poor.

2.5 - Limp tail and dragging of hind legs.

- OR - There is a strong head tilt that causes the mouse to occasionally fall over.

3.0 - Limp tail and complete paralysis of hind legs.

3.5 - Limp tail and complete paralysis of hind legs.

5 In addition to: Mouse is moving around the cage, but when placed on its side, is unable to right itself.

Hind legs are together on one side of body.

4.0 - Limp tail, complete hind leg and partial front leg paralysis.

- Mouse is minimally moving around the cage but appears alert and feeding

4.5 - Complete hind and partial front leg paralysis, no movement around the cage.

10 Mouse is immediately euthanized and removed from cage.

5.0 Mouse is euthanized due to severe paralysis.

When an animal has equal or greater disease activity score of 1, it is considered to have a positive disease incidence score.

Results

15 The results of the study are shown in Figures 70 and 71.

Disease induction in the negative control groups was successful with high scores shown by the vehicle control and the untreated control. The effect of treatment with strain 433 was striking and the mice treated with strain 433 exhibited notably reduced disease incidence and disease severity.. These data indicate the strain 433 may be useful for treating or preventing multiple sclerosis.

20 ***Example 7 – Stability testing***

A composition described herein containing at least one bacterial strain described herein is stored in a sealed container at 25°C or 4°C and the container is placed in an atmosphere having 30%, 40%, 50%, 60%, 70%, 75%, 80%, 90% or 95% relative humidity. After 1 month, 2 months, 3 months, 6 months, 1 year, 1.5 years, 2 years, 2.5 years or 3 years, at least 50%, 60%, 70%, 80% or 90% of the bacterial strain shall remain as measured in colony forming units determined by standard protocols.

Sequences

SEQ ID NO:1 (*Roseburia hominis* strain A2-181 16S ribosomal RNA gene, partial sequence - AY804148)

30 1 taaagggttga tcttggtcga ggatgaacgc tggaggcgtg cttaacacat gcaagtcgaa
61 cgaagcactt taattgattt cttcggaatg aagtttttgt gactgagtgg cggacgggtg
121 agtaacgcgt gggtaacctc gctcatcacg ggggataaca gttggaaacg actgctaata
181 ccgcataagc gcacaggatt gcatgatcca gtgtgaaaaa ctccggtggt atgagatgga
241 cccgcgtctg attagccagt tggcggggta acggcccacc aaagcgacga tcagtagccg

301 acctgagagg gtgaccggcc acattggggac tgagacacgg cccaaactcc tacgggagggc
 361 agcagtgggt aatattgcac aatgggggaa accctgatgc agcgacgccg agtgagcgaa
 421 gaagtatttc ggtatgtaaa gctctatcag caggaagaag aatgacggta cctgactaaa
 481 aagcaccggc taaatacgtg ccagcagccg cggtaatacg tatgggtgcaa gcgttatccg
 5 541 gatttactgg gtgtaaaggg agcgcaggcg gtacggcaag tctgatgtga aatcccgggg
 601 ctcaaccccg gtactgcatt ggaaactgtc ggactagggg gtctgagggg taagtggaaat
 661 tcctagtgtg gcggtgaaat gcgtagatat taggaggaac accagtggcg aaggcggcctt
 721 actggacgat tactgacgct gaggctcgaa agcgtgggga gcaaacagga ttagataccc
 781 tggtagtcca cgccgtaaac gatgaatact aggtgtcggg gagcattgct cttcgggtgcc
 10 841 gcagcaaacg caataagtat tccacctggg gagtacgttc gcaagaatga aactcaaagg
 901 aattgacggg gacccgcaca agcgggtggg catgtggttt aattcgaagc aacgcgaaga
 961 accttaccaa gtcttgacat cccactgaca aagtatgtaa tgtactttct cttcggagca
 1021 gtgggtgacag gtgggtgcatg gttgtcgtca gctcgtgtcg tgagatgttg ggttaagtcc
 1081 cgcaacgagc gcaaccccta ttcttagtag ccagcggttt ggccgggcac tctagggaga
 15 1141 ctgccaggga taacctggag gaagggtggg atgacgtcaa atcatcatgc cccttatgac
 1201 ttgggctaca cacgtgctac aatggcgtaa acaaagggaa gcaatccgcg gagggggagc
 1261 aaatctcaaa aataacgtct cagttcggac tgtagtctgc aactcgacta cacgaagctg
 1321 gaatcgctag taatcgcgaa tcagaatgtc gcggtgaata cgttcccggg tcttgtagac
 1381 accgcccgtc acaccatggg agttggtaat gcccgaaagtc agtgacccaa ccgcaaggag
 20 1441 ggagctgccg aagcaggact gataactggg gtgaagtcgt aacaagt

SEQ ID NO:2 (*Roseburia hominis* A2-183 16S rRNA gene, type strain A2-183T - AJ270482)

1 gatcctggct caggatgaac gctggcgggc tgcttaacac atgcaagtcg aacgaagcac
 61 ttttaattgat ttcttcggaa tgaagttttt gtgactgagt ggccggacggg tgagtaaacgc
 25 121 gtgggtaacc tgccctatac agggggataa cagttggaaa cgactgctaa taccgcataa
 181 gcgcacagga ttgcatgata cagtgtgaaa aactccgggtg gtatgagatg gacccgcgtc
 241 tgattagcca gttggcgggg taacggccca ccaaagcgac gatcagtagc cgacctgaga
 301 ggggtgaccgg ccacattggg actgagacac ggcccaaact cctacgggag gcagcagtgg
 361 ggaatattgc acaatggggg aaacctgat gcagcgacgc cgcgtagcg aagaagtatt
 30 421 tcggtatgta aagctctatc agcagggaag aagaatgcgg tacctgacta agaagcaccg
 481 gctaaatacg tgccagcagc cgcggtaata cgtatggtgc aagcgttatc cggatttact
 541 ggggtgtaaag ggagcgcagg cggtagcgca agtctgatgt gaaatcccgg ggctcaaccc
 601 cggtagtgca ttggaaactg tcggactaga gtgtcggagg ggtaagtgga attcctagtg
 661 tagcggtgaa atgcgtagat attaggagga acaccagtgg cgaaggcggc ttactggacg
 35 721 attactgacg ctgaggctcg aaagcgtggg gagcaaacag gattagatac cctggtagtc
 781 cacgccgtaa acgatgaata ctaggtgtcg gggagcattg ctcttcgggtg ccgcagcaaa
 841 cgcaataagt attccacctg gggagtagct tcgcaagaat gaaactcaaa ggaattgacg
 901 gggacccgca caagcgggtg agcatgtggt ttaattcgaa gcaacgcgaa gaaccttacc
 961 aagtcttgac atcccactga cagagtatgt aatgtacttt ctcttcggag cagtggtagc
 40 1021 aggtggtgca tggttgtcgt cagctcgtgt cgtgagatgt tgggttaaagt cccgcaacga
 1081 gcgcaacccc tattcttagt agccagcggg tcggccgggg actctaggga gactgccagg

1141 gataacctgg aggaaggtgg g gatgacgtc aaatcatcat gccccttatg acttgggcta
 1201 cacacgtgct acaatggcgt aaacaaaggg aagcaatccc gcgaggggga gcaaacttca
 1261 aaaataacgt ctacgttcgg actgtagtct gcaactcgac tacacgaagc tggaatcgct
 1321 agtaatcgcg aatcagaatg tcgcggtgaa tacgttcccc ggtcttgtac acaccgcccc
 5 1381 tcacaccatg ggagttggta atgcccgaag tcagtgaacc aaccgcaagg agggagctgc
 1441 cgaaggcagg actgataact ggggtgaagt cgtaacaagg gtacg

SEQ ID NO:3 (consensus 16S rRNA sequence for *Roseburia hominis* strain 433)

10 AAGAGTTTGGGHCAGGCTCAGGATGAACGCTGGCGGCGTGCTTAACACATGCAAGTCGAACGAAGCACTTTAATTGA
 TTTCTTCGGAATGAAGTTTTTGTGACTGAGTGGCGGACGGGTGAGTAACGCGTGGGTAACTGCCTCATACAGGGG
 ATAACAGTTGGAAACGACTGCTAATACCGCATAAGCGCACAGGATTGCATGATCCAGTGTGAAAACTCCGGTGGTA
 TGAGATGGACCCGCGTCTGATTAGCCAGTTGGCGGGTAACGGCCCCACAAAGCGACGATCAGTAGCCGACCTGAGA
 GGGTGACCGGCCACATTGGGACTGAGACACGGCCAACTCCTACGGGAGGCAGCAGTGGGGAATATTGCACAATGG
 GGGAAACCCTGATGCAGCGACGCCGCGTGAGCGAAGAAGTATTTCCGGTATGTAAAGCTCTATCAGCAGGGAAGAAGA
 15 ATGACGGTACCTGACTAAGAAGCACCGGCTAAATACGTGCCAGCAGCCGCGGTAATACGTATGGTGCAAGCGTTATC
 CGGATTTACTGGGTGTAAAGGGAGCGCAGGCGGTACGGCAAGTCTGATGTGAAATCCCCGGGGCTCAACCCCGGTACT
 GCATTGGAACTGTCTGGACTAGAGTGTCTGGAGGGGTAAGTGGAAATTCCTAGTGTAGCGGTGAAATGCGTAGATATTA
 GGAGGAACACCAGTGGCGAAGGCGGCTTACTGGACGATTACTGACGCTGAGGCTCGAAAGCGTGGGGAGCAAACAGG
 ATTAGATACCCTGGTAGTCCACGCCGTAAACGATGAATACTAGGTGTCTGGGAGCATTGCTCTTCGGTGCCGCAGCA
 20 AACGCAATAAGTATNCCACCTGGGGAGTACGTTTCGAAGAATGAAACTCAAAGGAATTGACGGGGACCCGCACAAGC
 GGTGGAGCNTGTGGTTTAATTTCGAAGCAACGCGAAGAACCTTACCAAGTCTTGACATCCCACTGACAGAGTATGTAA
 TGTACTTTCTCTTCGGAGCAGTGGTGACAGGTGGTGATGGTTGTCTGCTCAGCTCGTGTCTGAGATGTTGGGTTAAG
 TCCCGCAACGAGCGCAACCCCTATTCTTAGTAGCCAGCGGTTTGGCCGGGCACTCTAGGGAGACTGCCAGGGATAAC
 CTGGAGGAAGGTGGGGATGACGTCAAATCATCATGCCCCCTTATGACTTGGGCTACACACGTGCTACAATGGCGTAAA
 25 CAAAGGGAAGCAATCCCGCGAGGGGGAGCAAATCTCAAAAATAACGTCTCAGTTCGGACTGTAGTCTGCAACTCGAC
 TACACGAAGCTGGAATCGCTAGTAATCGCGAATCAGAATGTCTCGCGGTGAATACGTTCCCGGGTCTTGTACACACCGC
 CCGTCACACCATGGGAGTTGGTAATGCCCCAAGTCAGTGACCCAACCGCAAGGAGGGAGCTGCCGAAGGCAGGACTG
 ATA ACTGGGGTGAAGTCTACRSAGGGTAGCCGTRMMC

30 SEQ ID NO:4 (strain 433 genome sequence) – see WO2016203221.

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CLAIMS

1. A composition comprising a bacterial strain of the genus *Roseburia*, for use in a method of treating or preventing asthma.

2. The composition of claim 1, wherein the composition is for use in a method of treating or preventing neutrophilic asthma or allergic asthma.

3. The composition of claim 1 or 2, wherein the composition is for use in a method of reducing neutrophilia or eosinophilia in the treatment of asthma.

4. The composition of any preceding claim, wherein the composition is for use in a method of reducing IL-17 production or reducing Th17 cell differentiation in the treatment or prevention of a disease or condition mediated by IL-17 or the Th17 pathway.

5. The composition of any preceding claim, wherein the composition is for use in a patient with elevated IL-17 levels or Th17 cells.

6. The composition of any preceding claim, wherein the bacterial strain is of *Roseburia hominis*.

7. The composition of any of claims 1-6, wherein the bacterial strain has a 16s rRNA sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to SEQ ID NO:1, 2 or 3.

8. The composition of claim 7, wherein the bacterial strain has a 16s rRNA sequence that is at least 95%, 96%, 97%, 98%, 99%, 99.5% or 99.9% identical to SEQ ID NO:3, or wherein the bacterial strain has the 16s rRNA sequence represented by SEQ ID NO:3.

9. The composition of claim 1, wherein the composition comprises a bacterial strain of the species *Roseburia hominis*, for use in a method of treating or preventing neutrophilic asthma or allergic asthma.

10. The composition of any preceding claim, wherein the bacterial strain is lyophilised.

11. A food product or vaccine composition comprising the composition of any preceding claim, for the use of any preceding claim.

12. A cell of the *Roseburia hominis* strain deposited under accession number NCIMB 42383, or a derivative thereof.

13. A biologically pure culture of the *Roseburia hominis* strain deposited under accession
5 number NCIMB 42383, or a derivative thereof.

14. A cell of the *Roseburia hominis* strain deposited under accession number NCIMB 42383, or a derivative thereof, for use in therapy.

10 15. The cell of claim 14, wherein the cell is for use in a method defined in any of claims 1-5.

ABSTRACT

The invention provides compositions comprising bacterial strains for treating and preventing inflammatory and autoimmune diseases.

FIG. 1
Total cell counts.

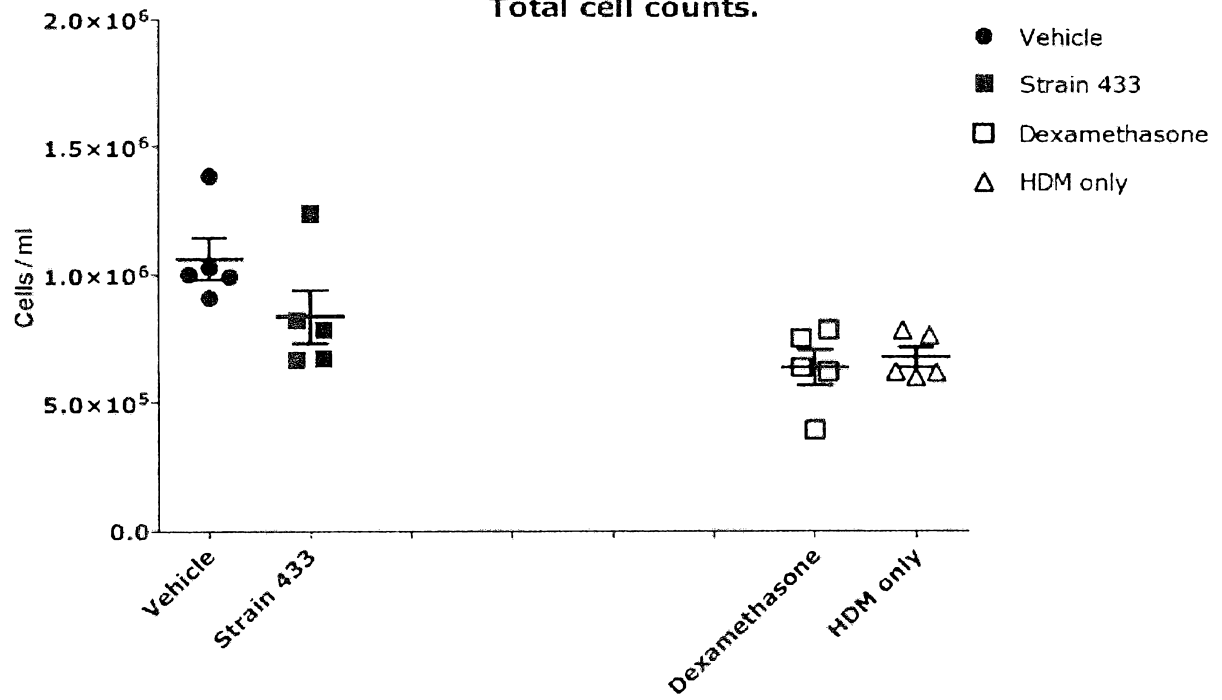


FIG. 1
Total cell counts.

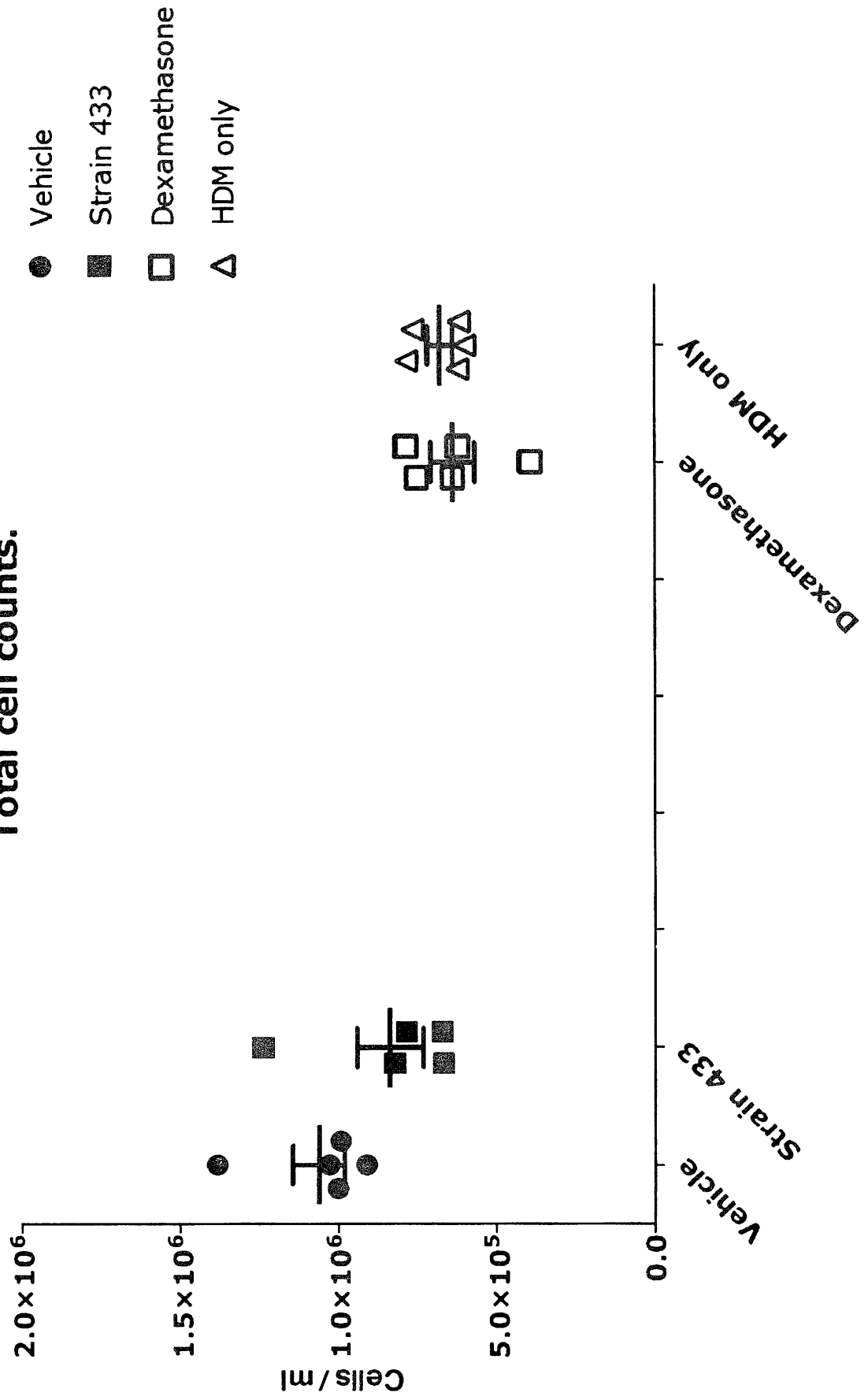
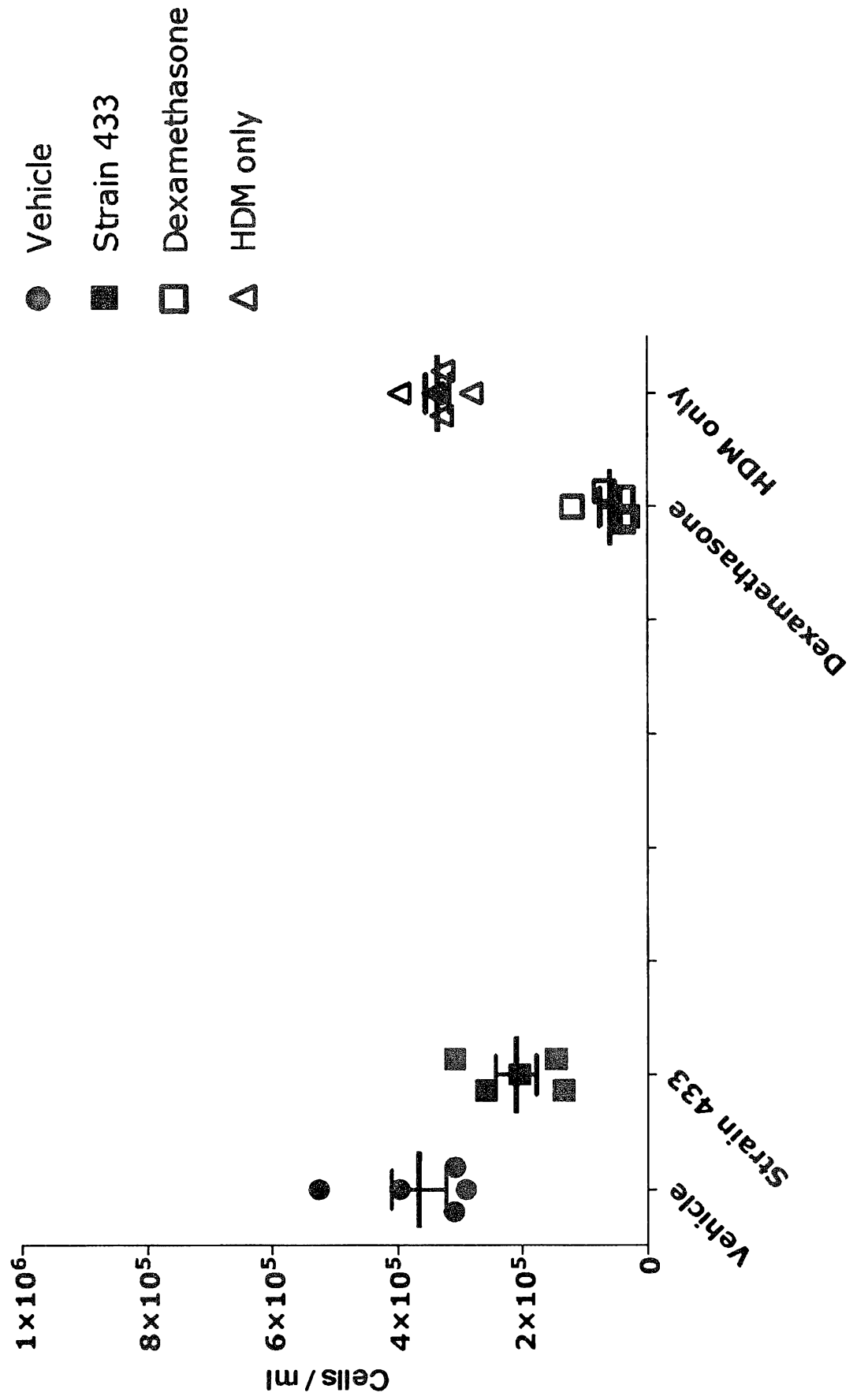


FIG. 2
Total eosinophils.



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FIG. 3
Eosinophils %.

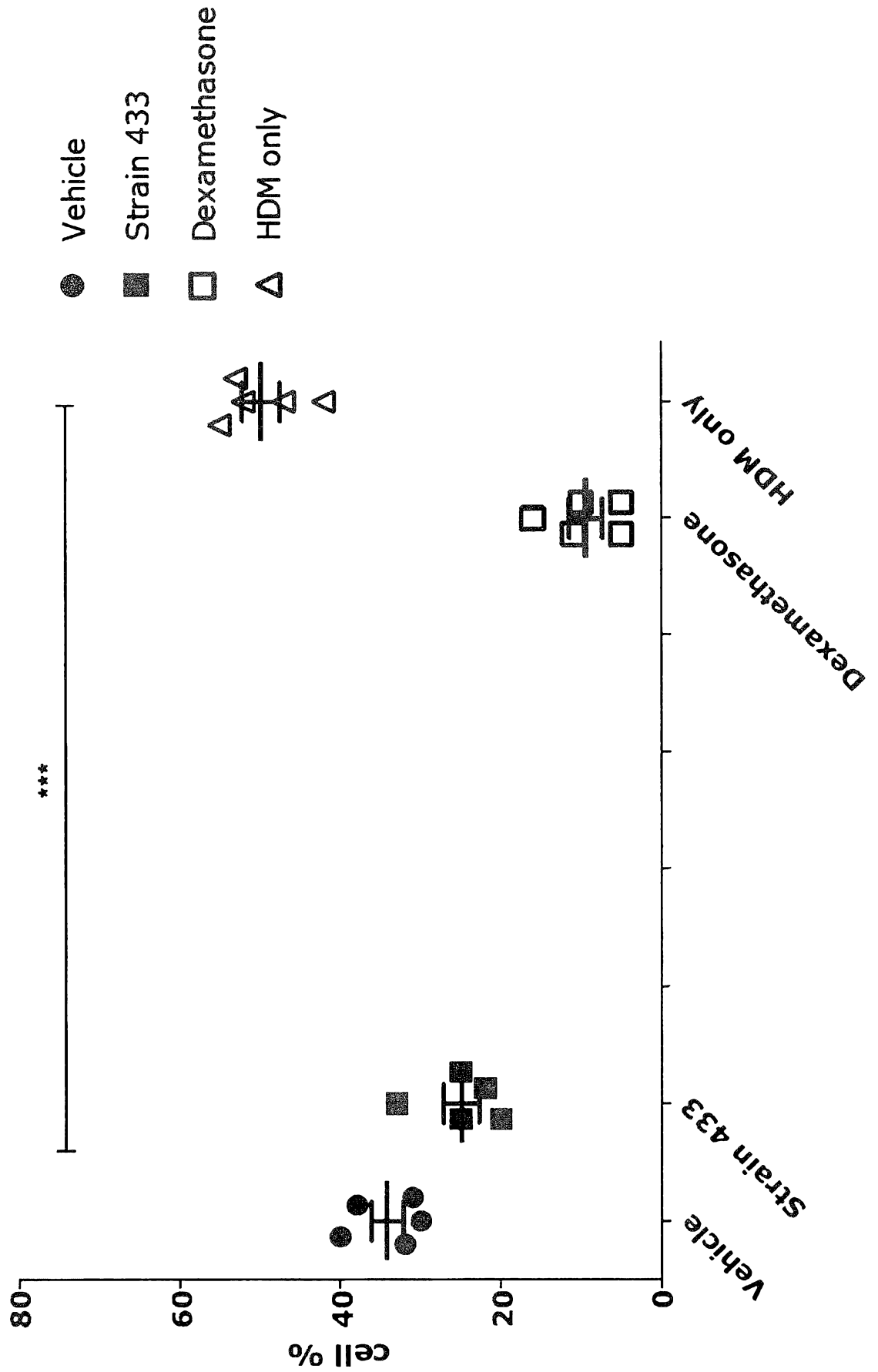
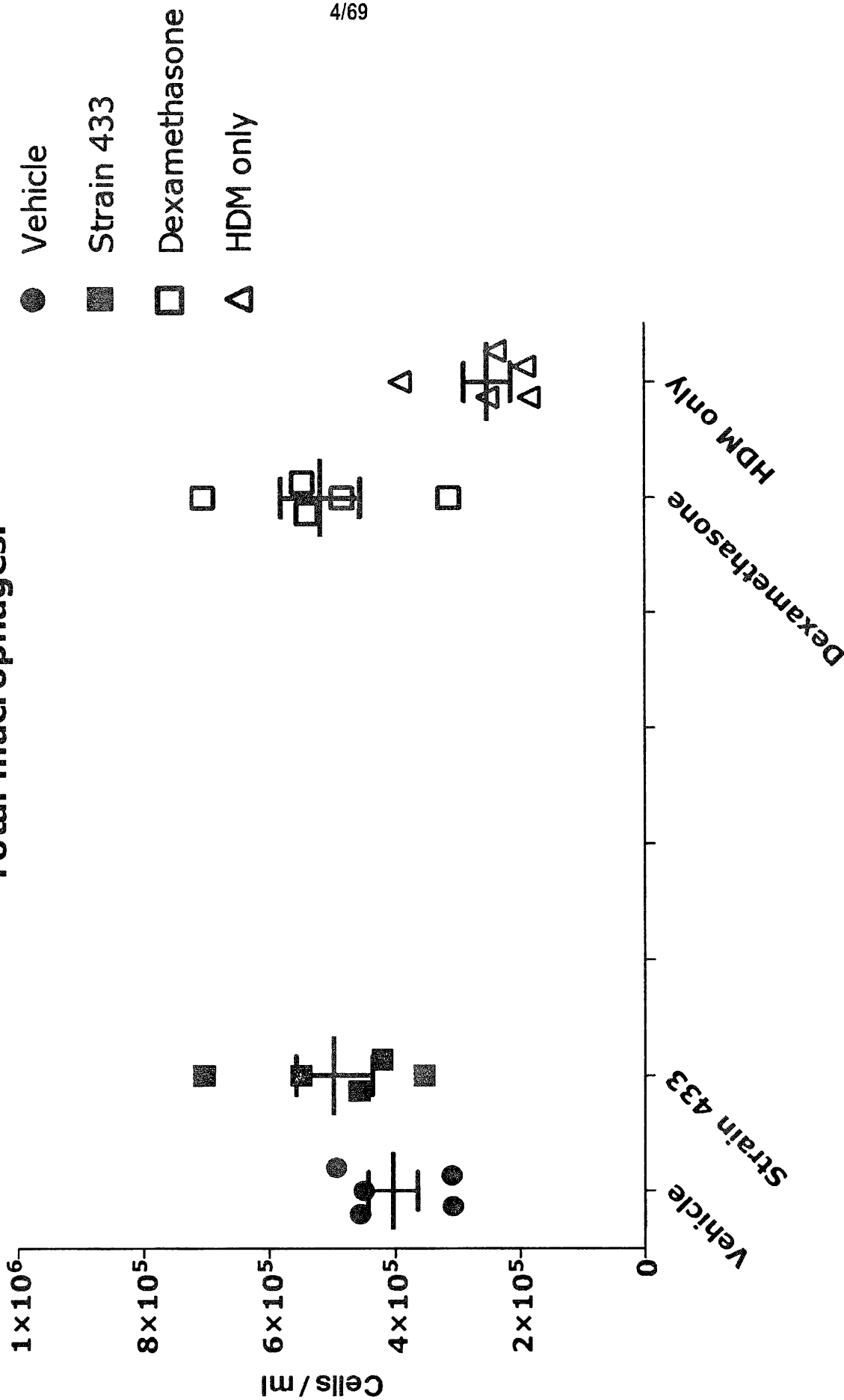
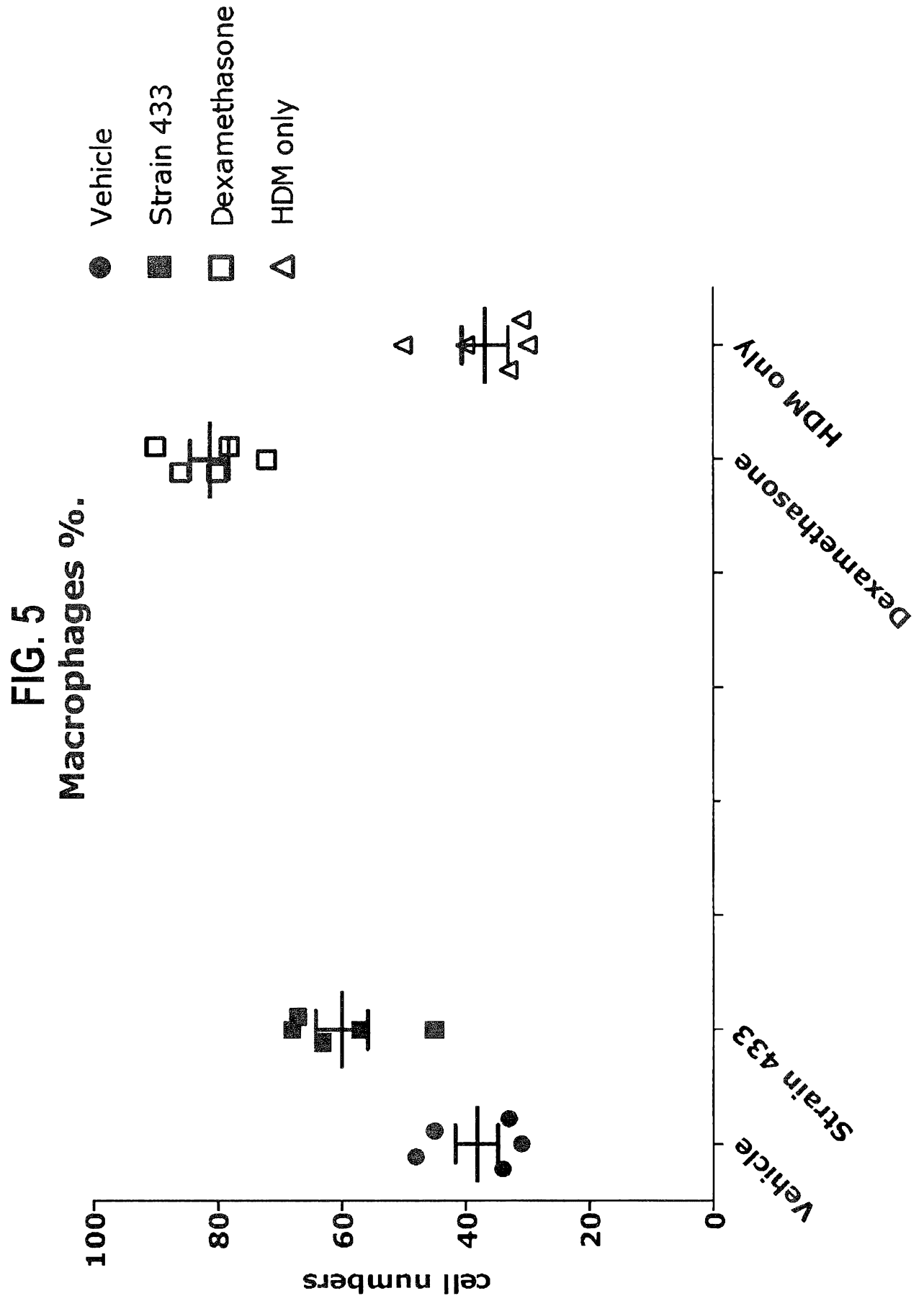


FIG. 4
Total macrophages.



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FIG. 6
Total neutrophils.

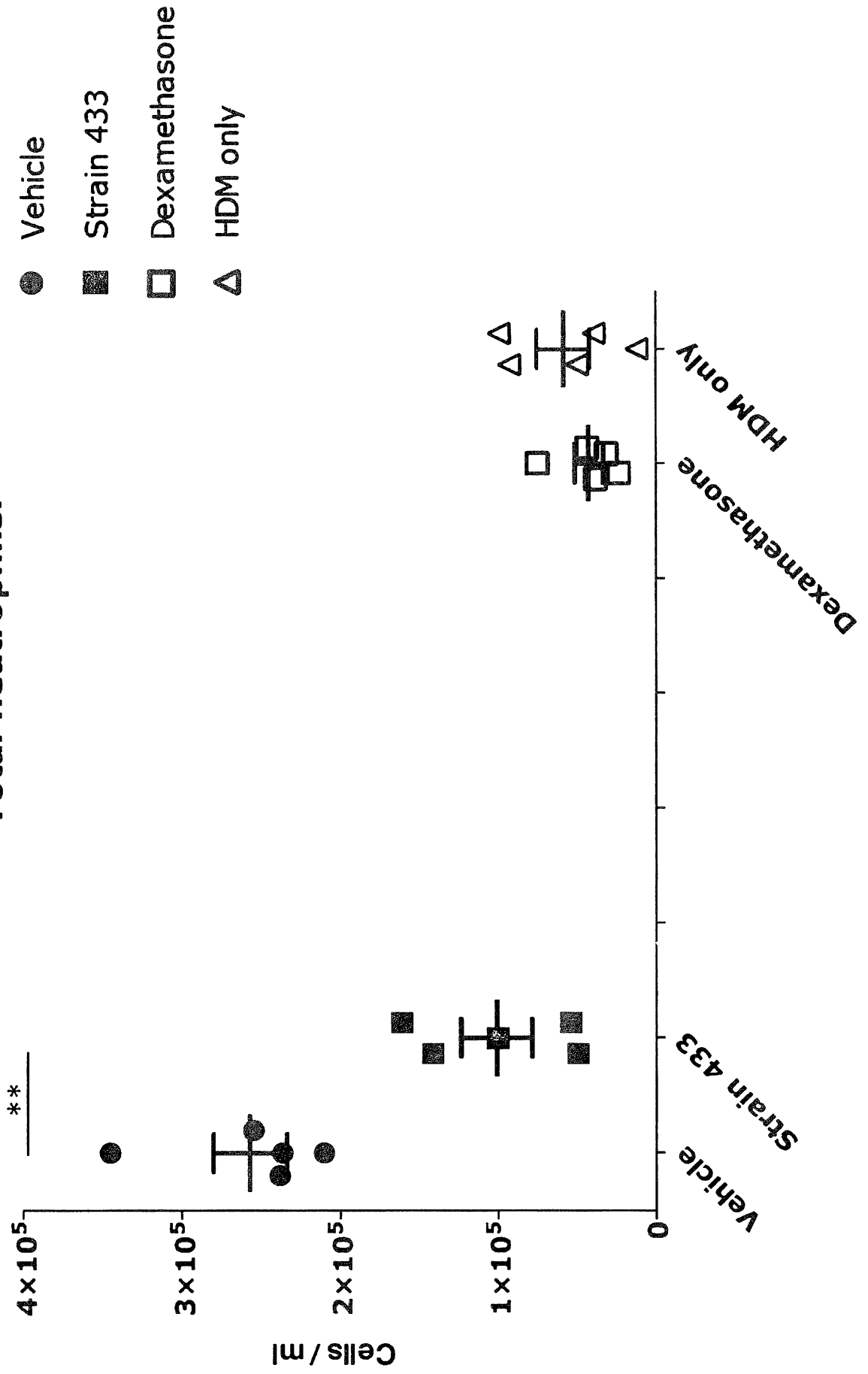


FIG. 7
Neutrophils %.

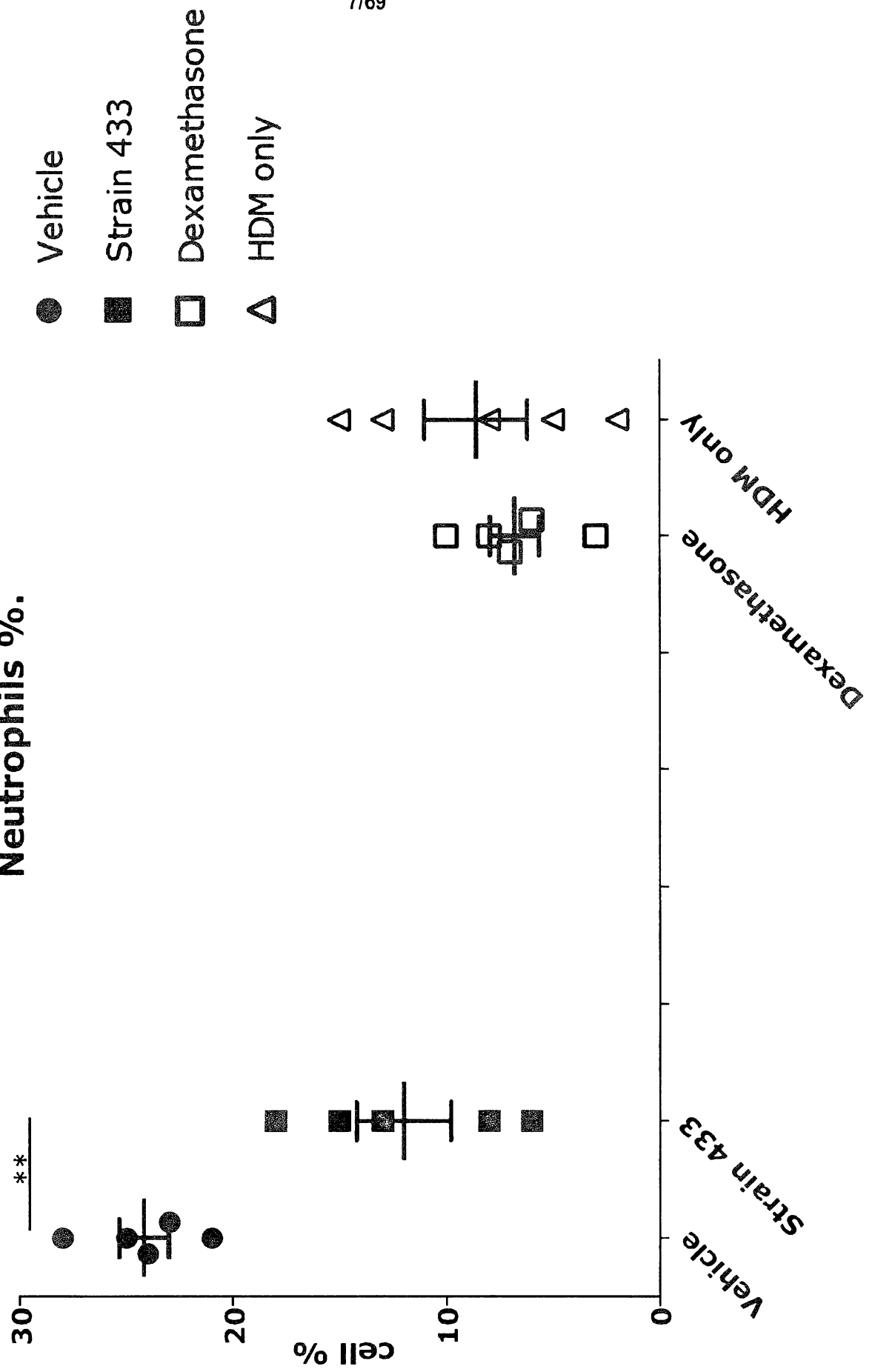
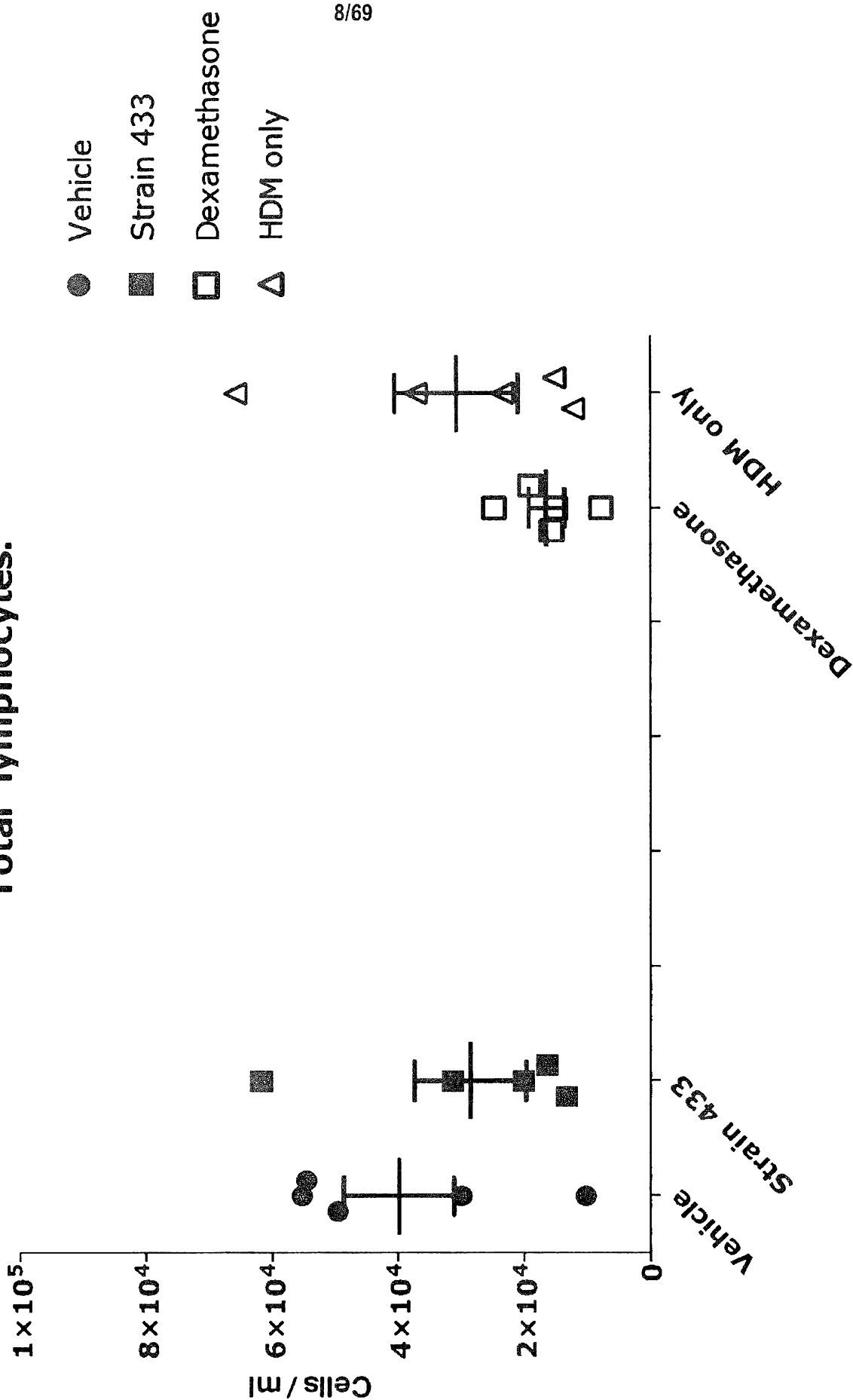


FIG. 8
Total lymphocytes.



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FIG. 9
Lymphocytes %.

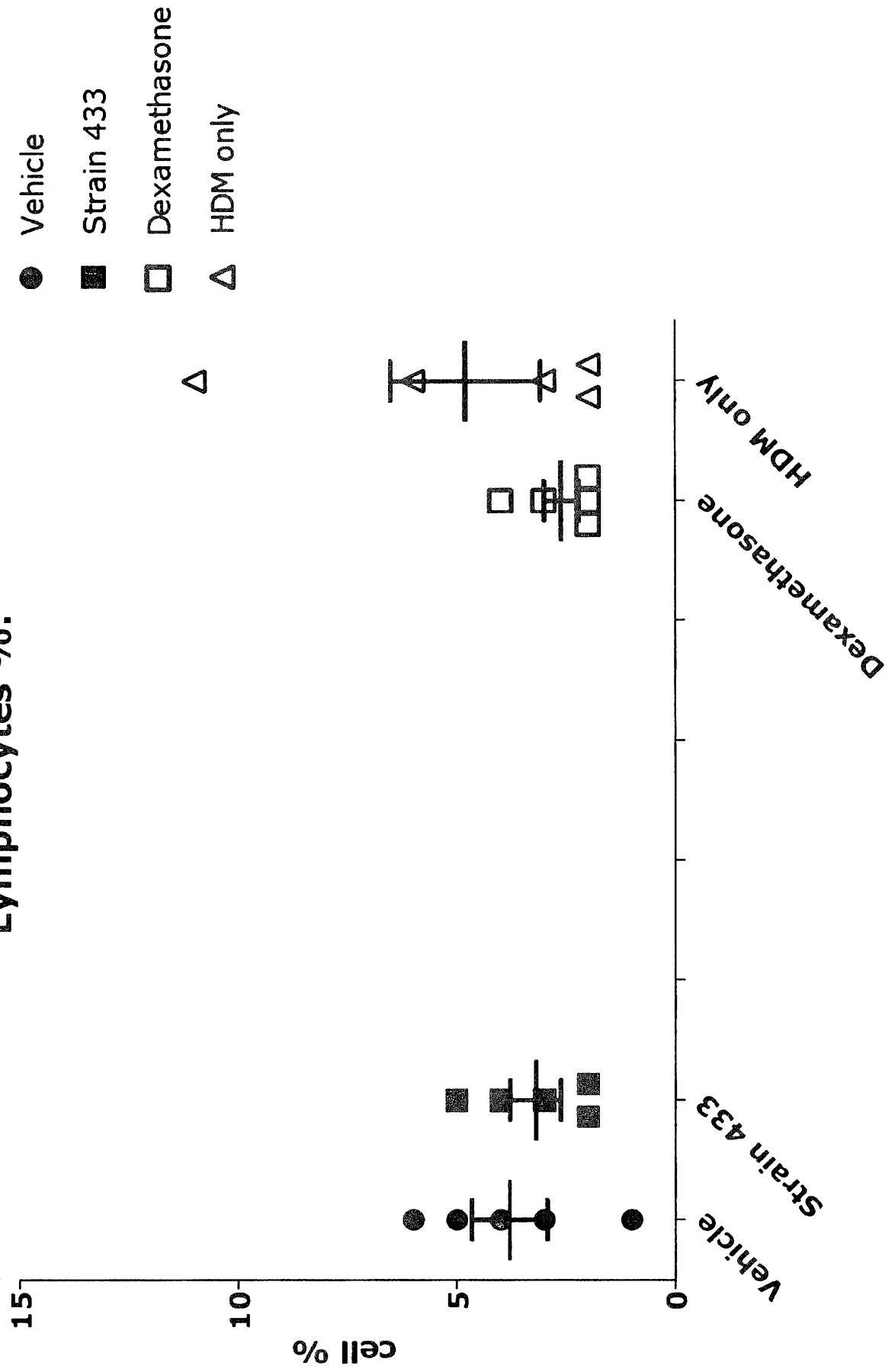


FIG. 10
Total cell counts.

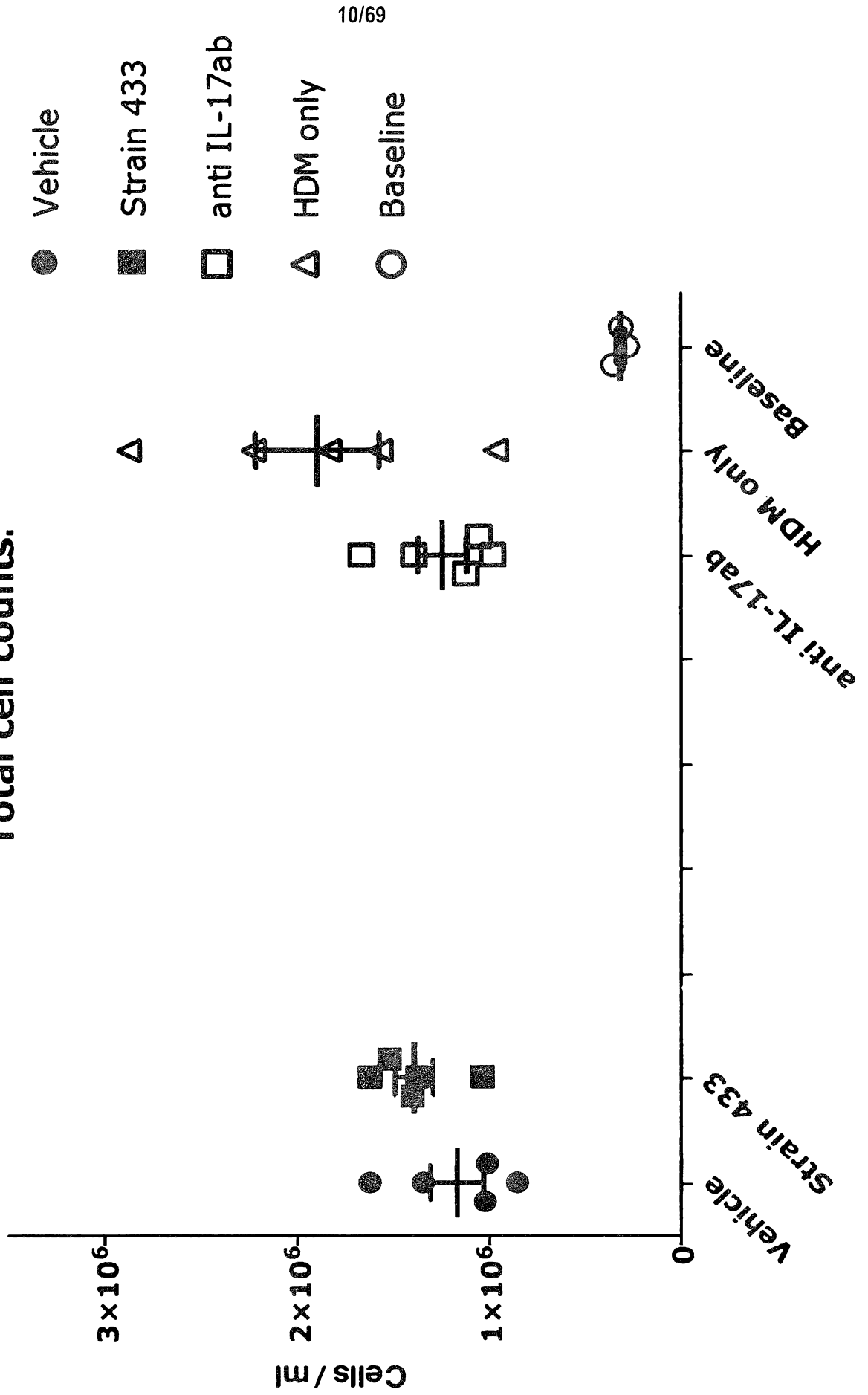


FIG. 11
Total eosinophils.

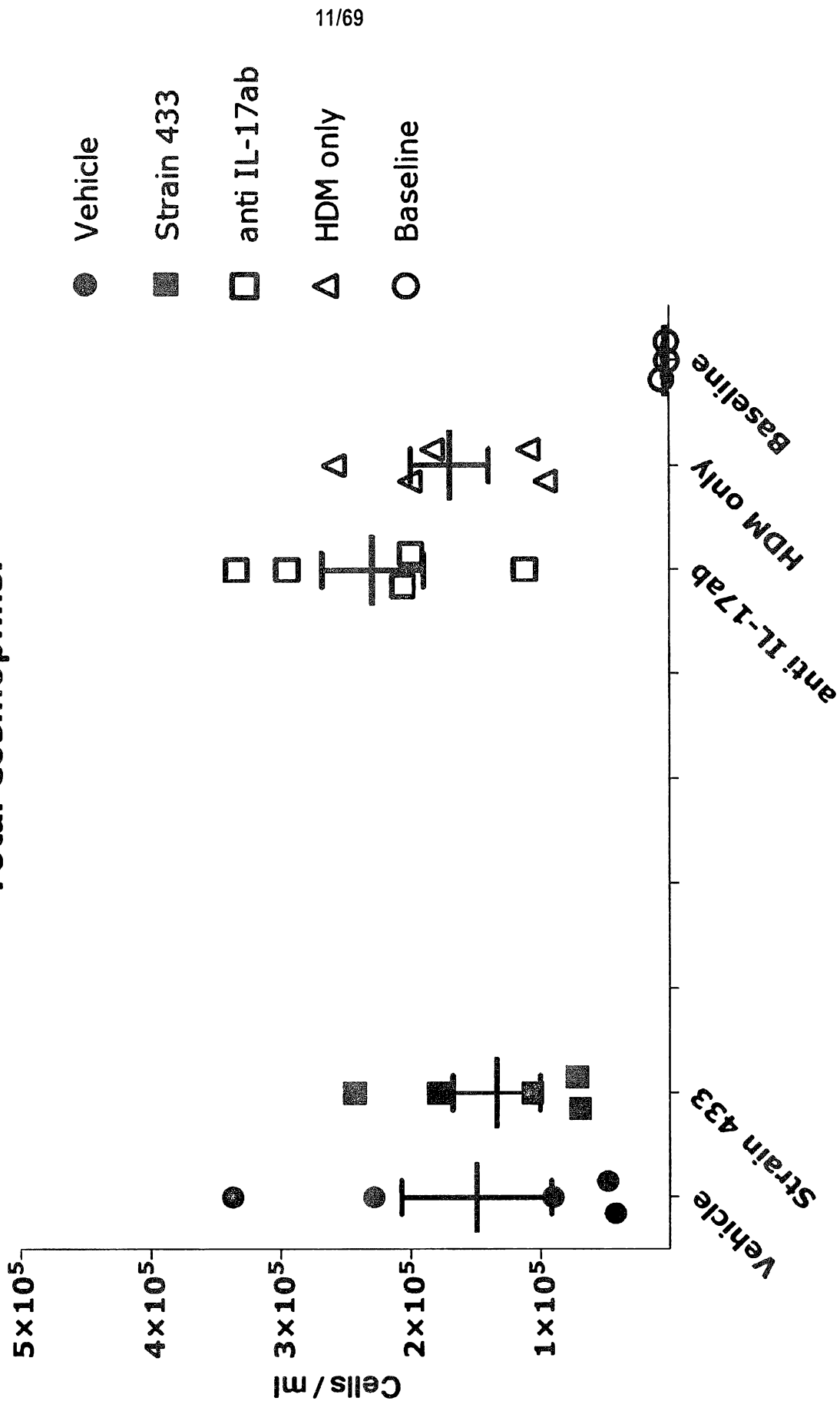


FIG. 12
Eosinophils %.

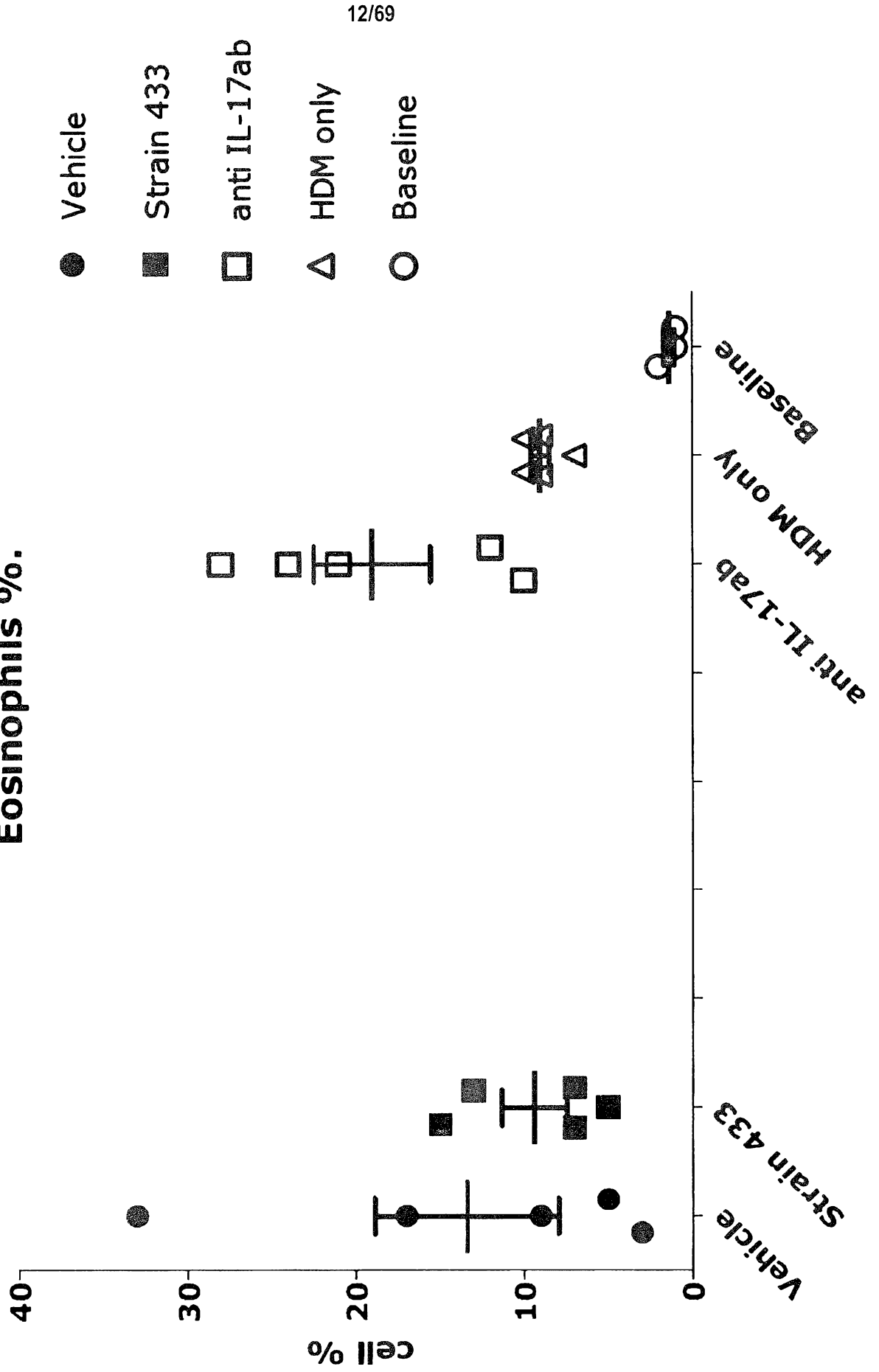


FIG. 13
Total macrophages.

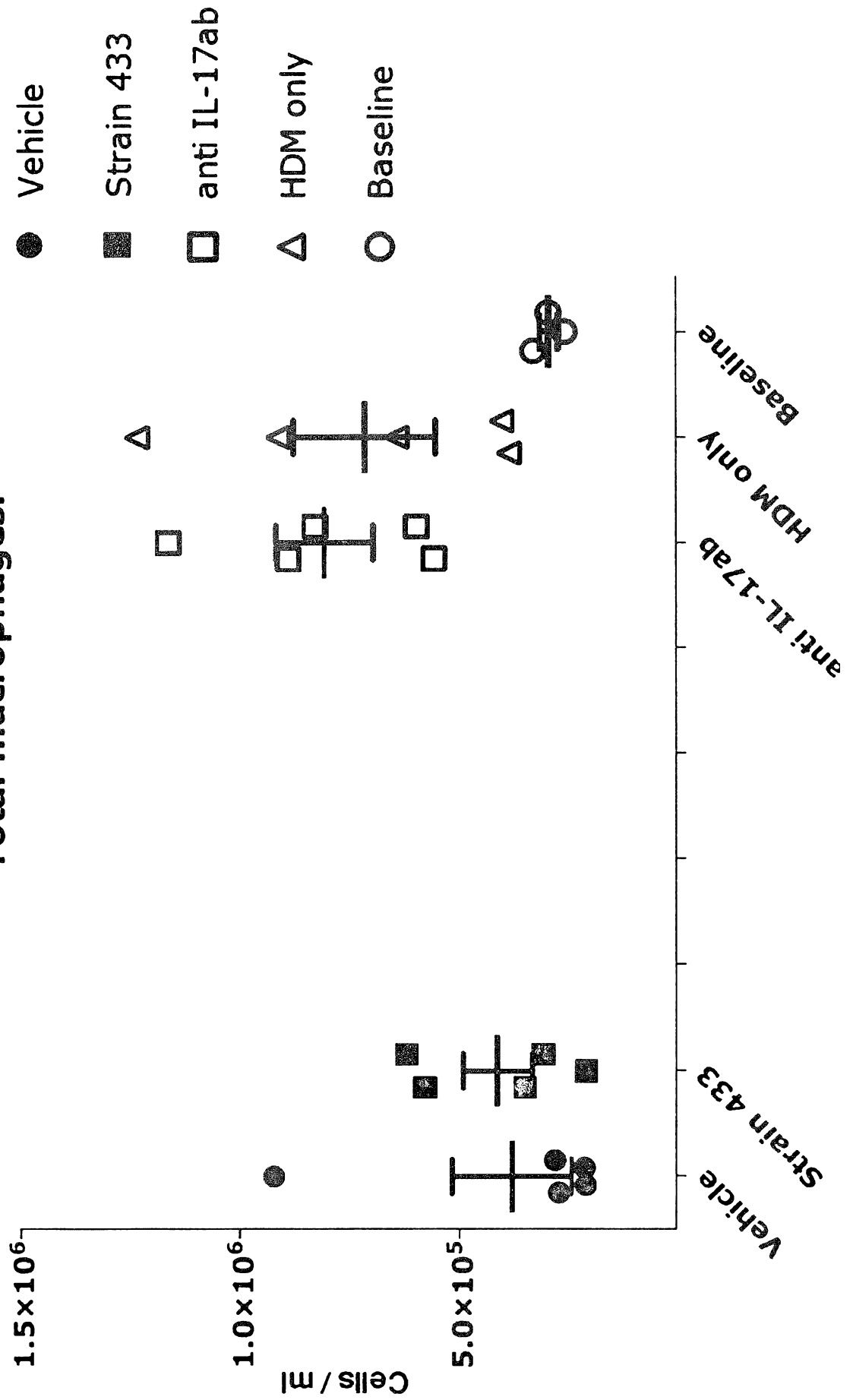


FIG. 14
Macrophages %.

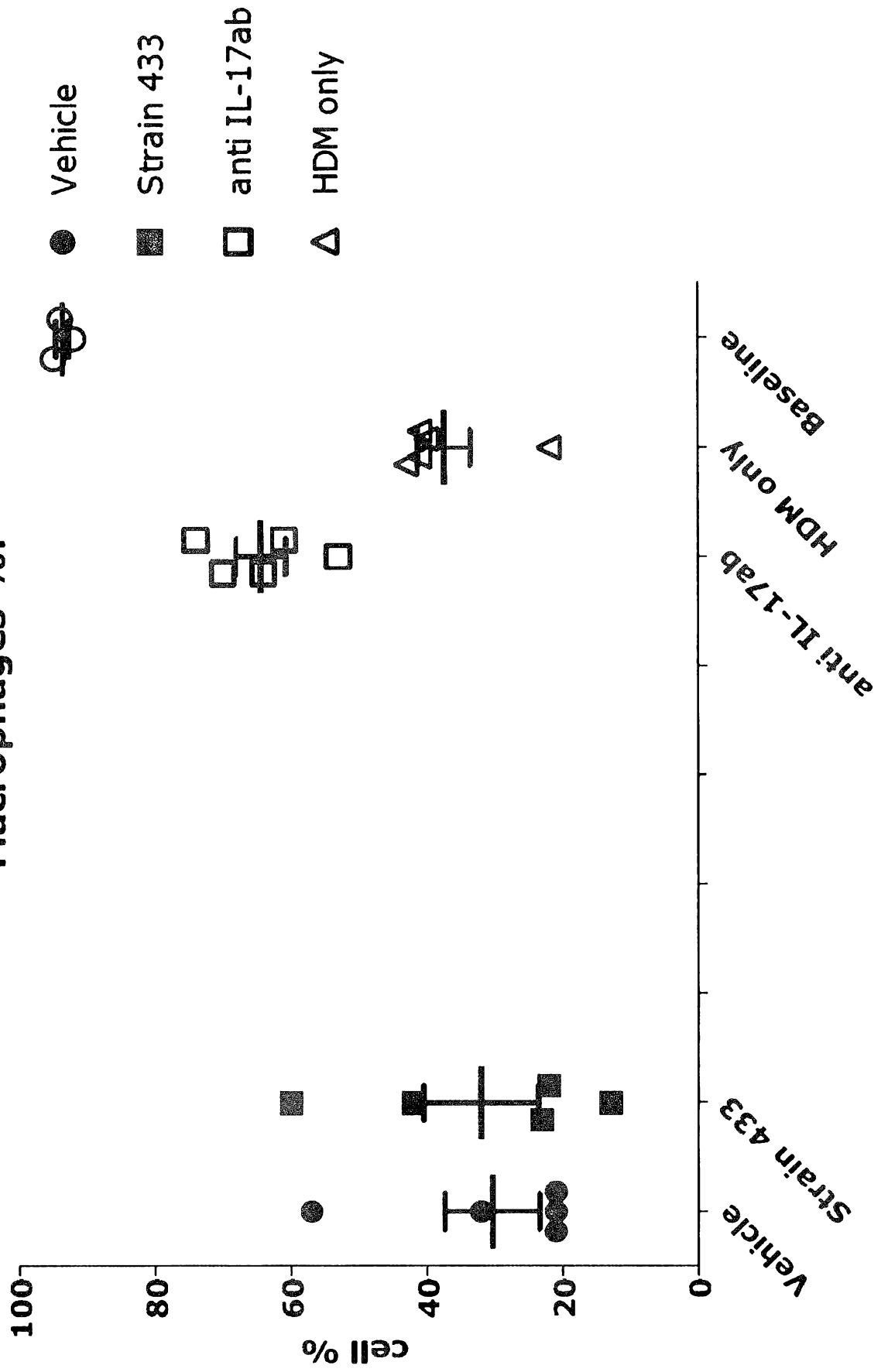
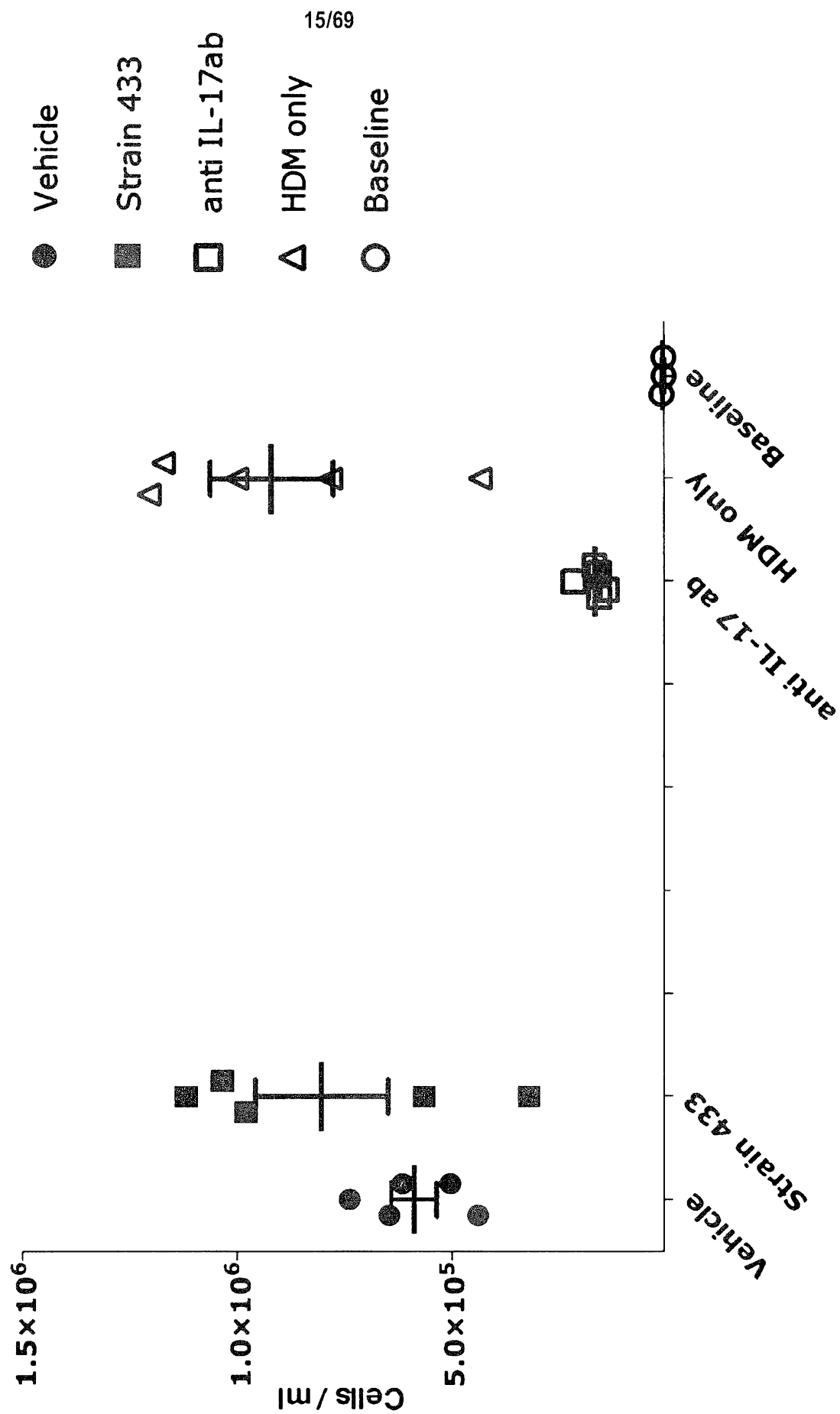


FIG. 15
Total neutrophils.



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FIG. 16
Neutrophils %.

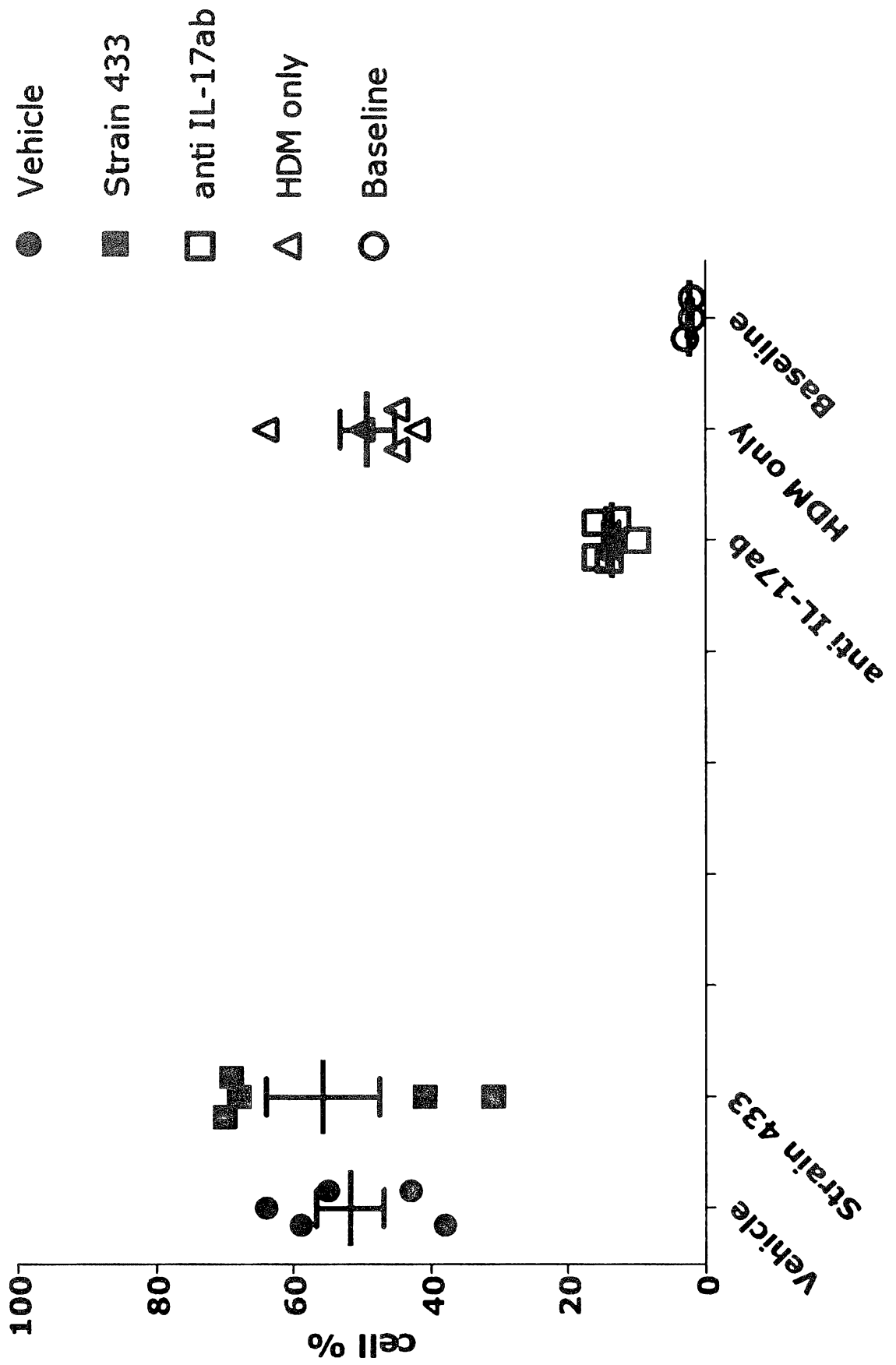
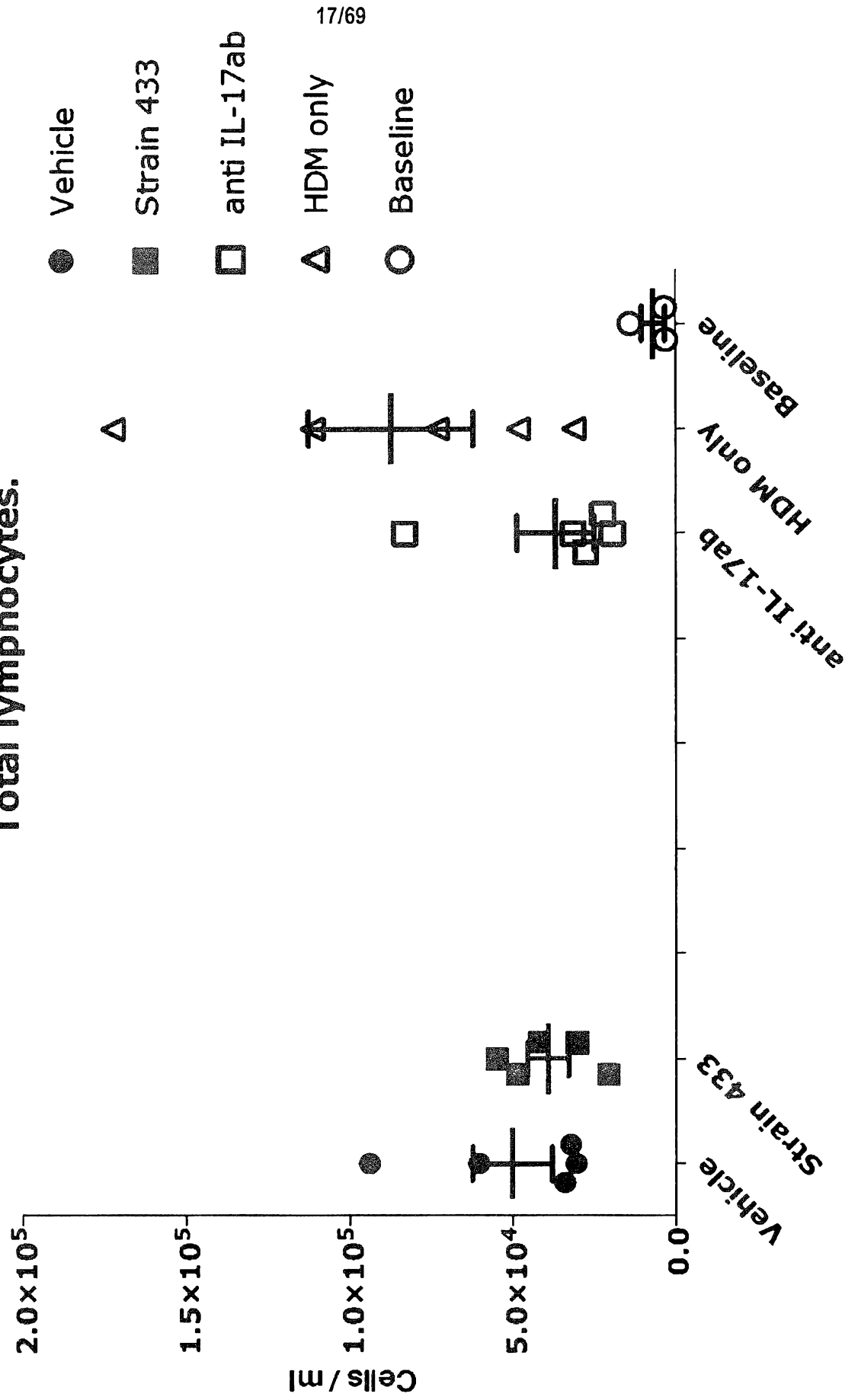
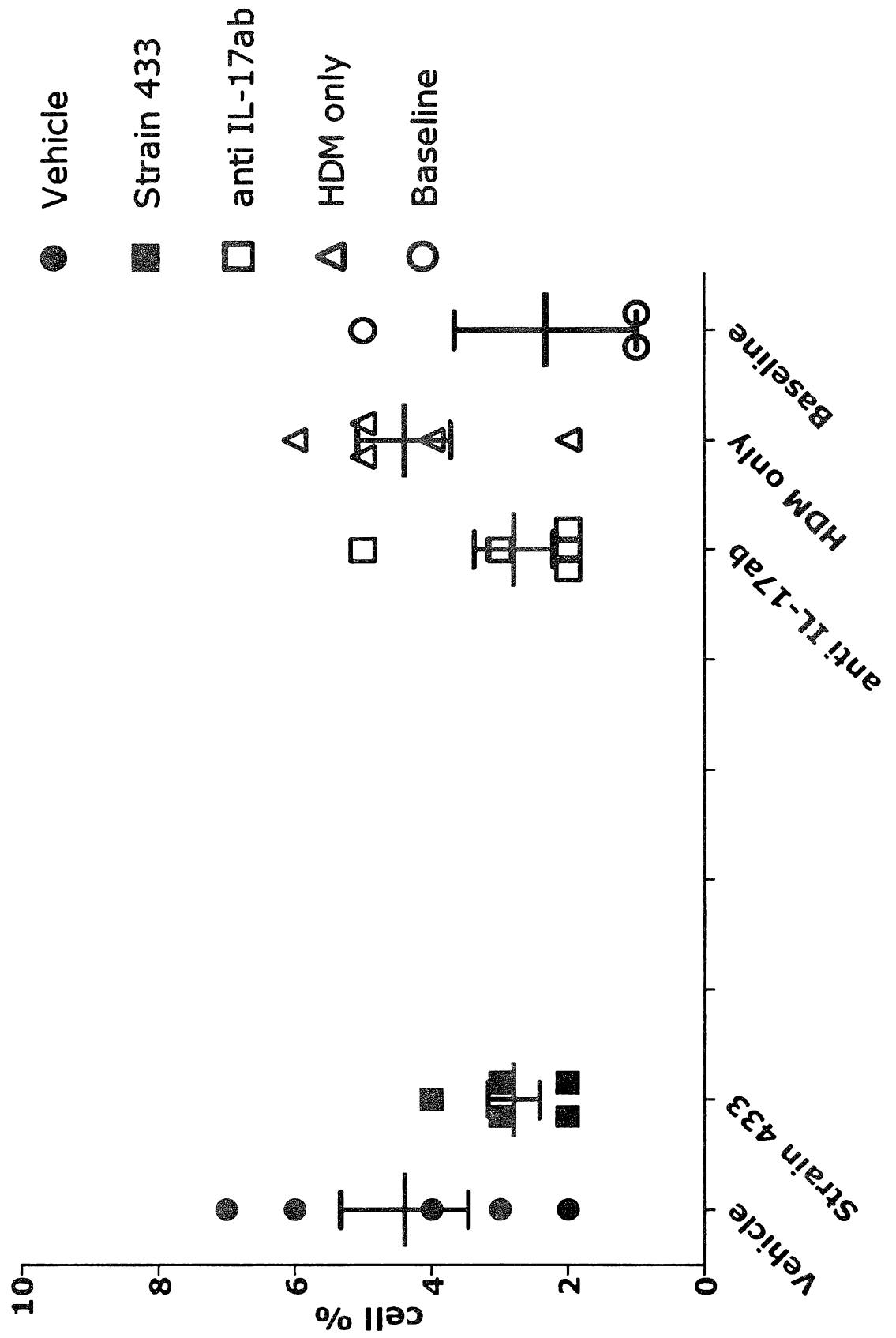


FIG. 17
Total lymphocytes.



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FIG. 18
Lymphocytes %.



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FIG. 19
Bodyweights

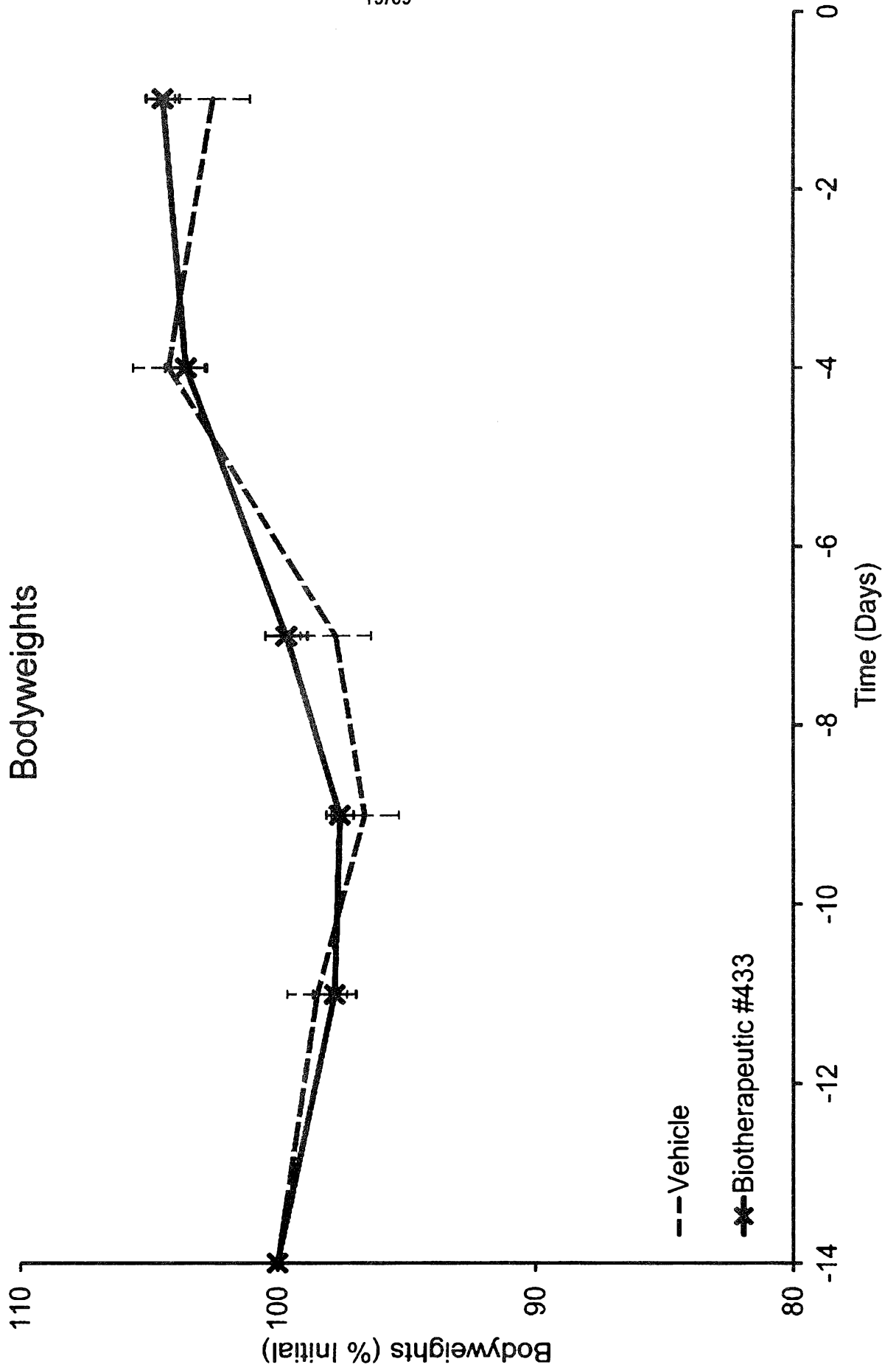


FIG. 20
Bodyweights

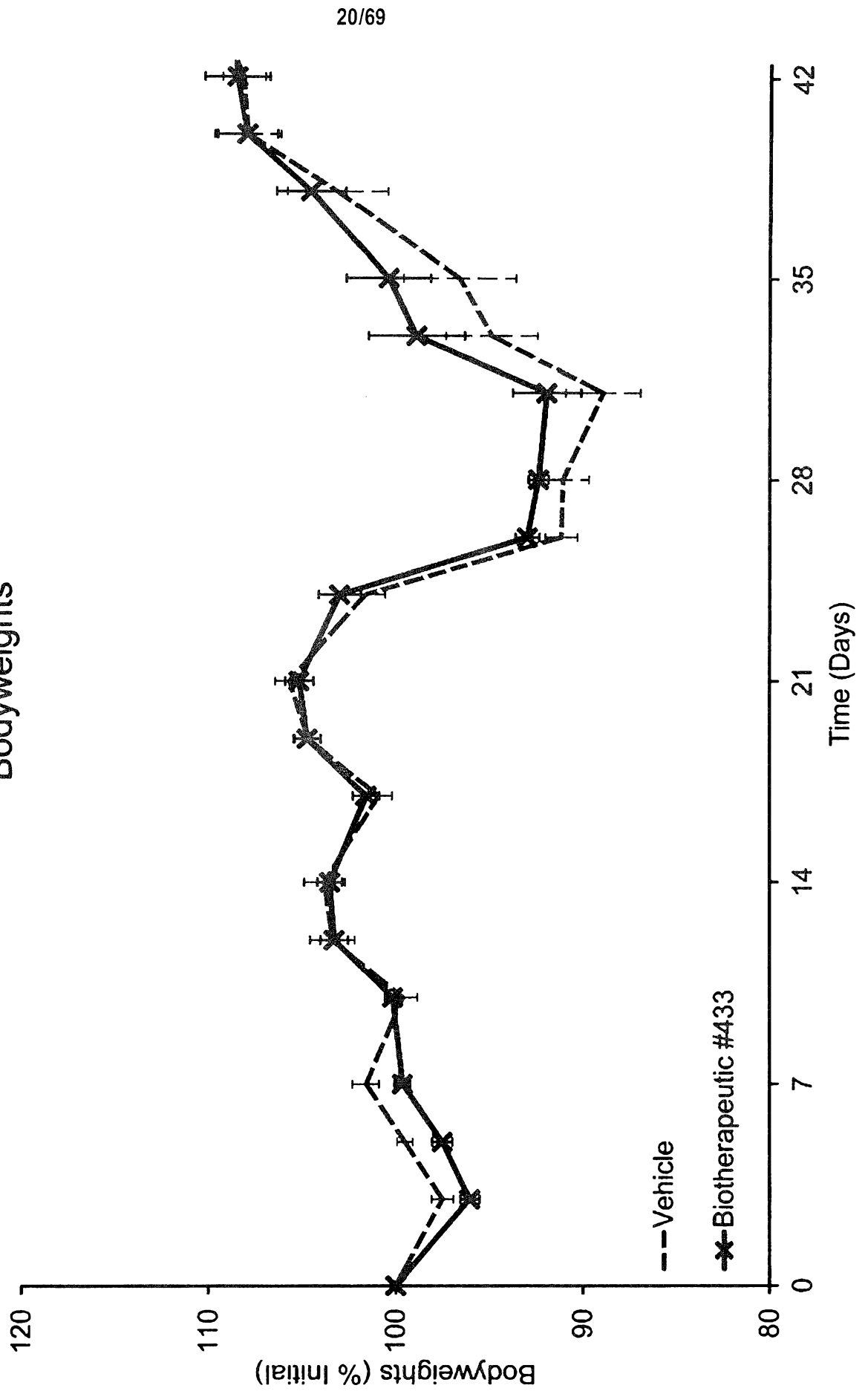


FIG. 21 Clinical Scores

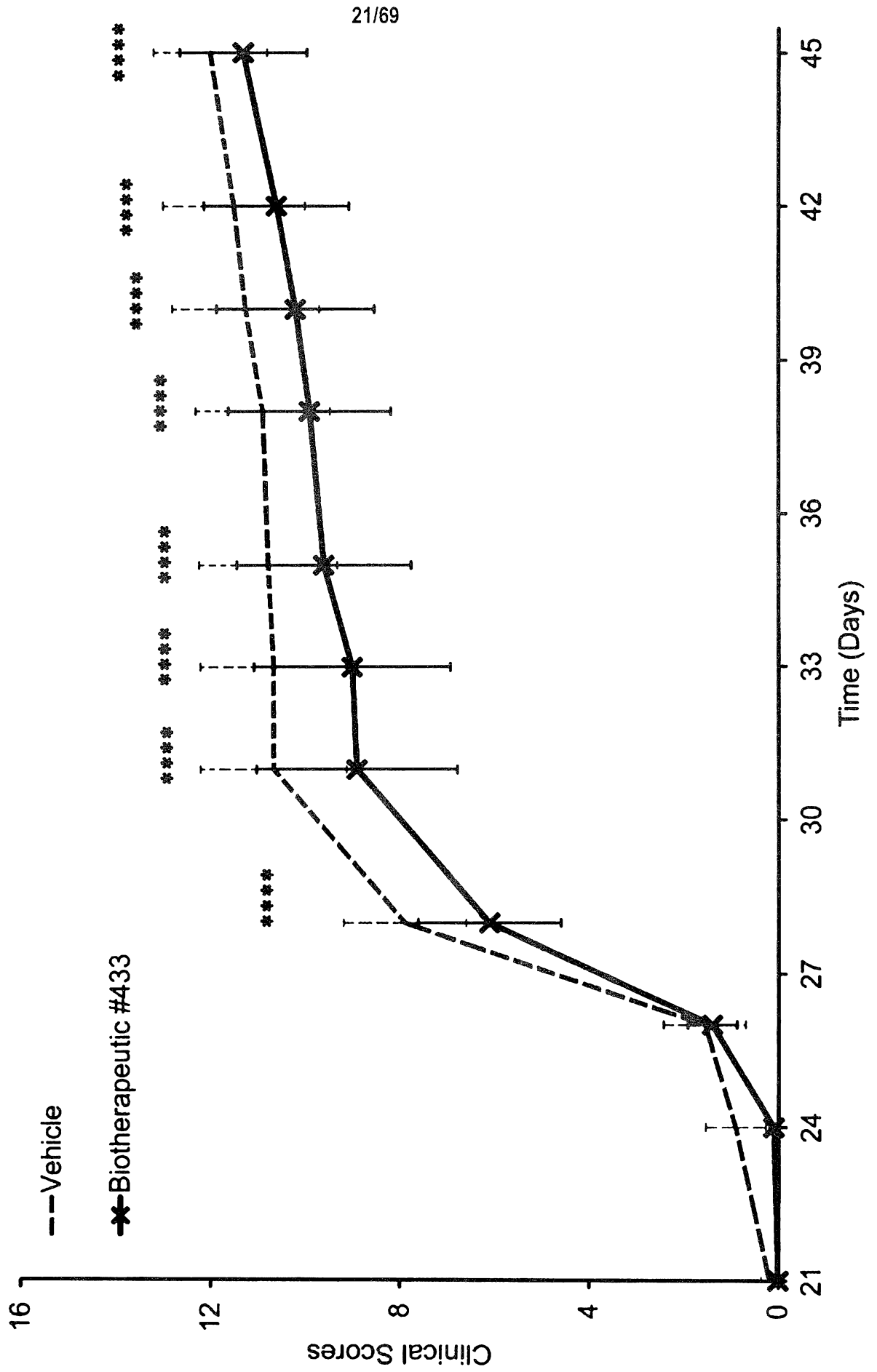


FIG. 23

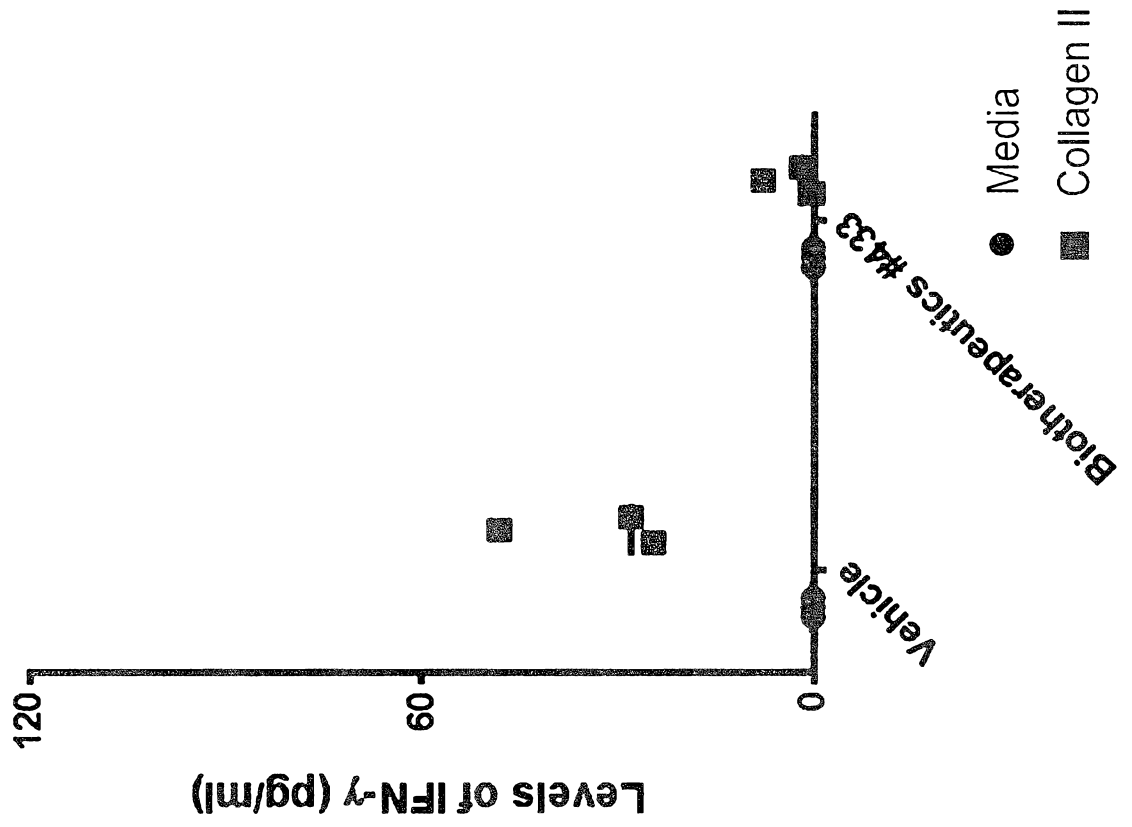
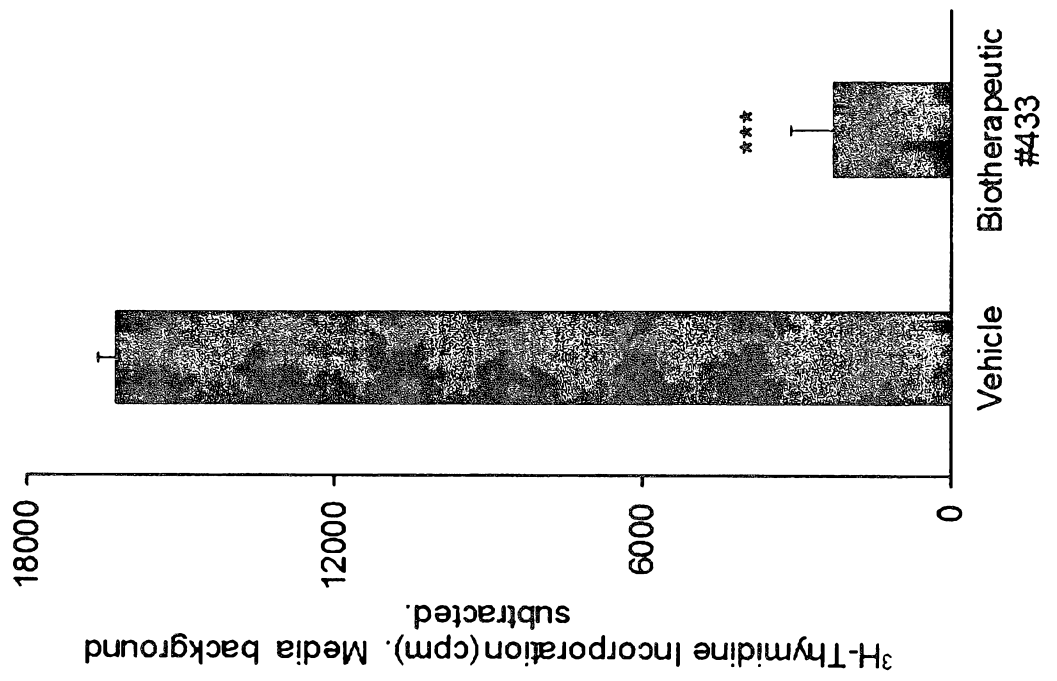
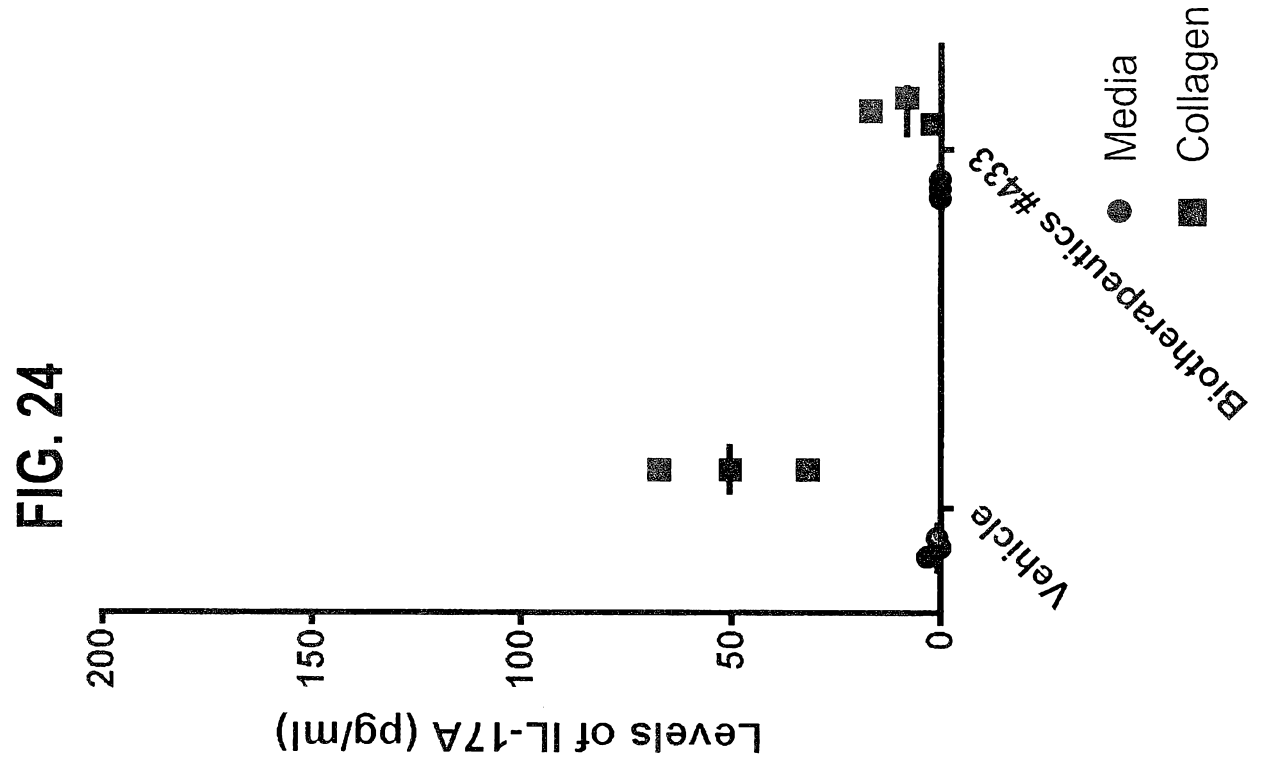
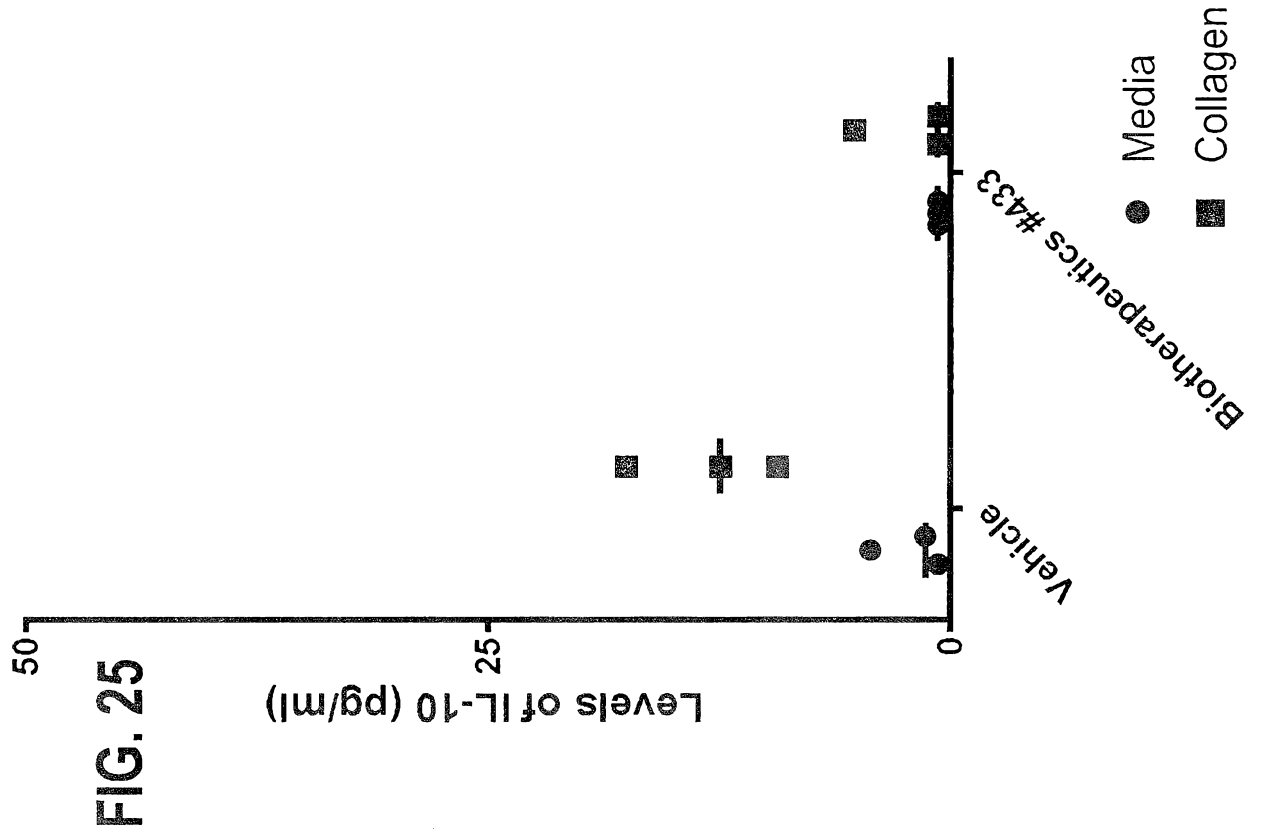


FIG. 22

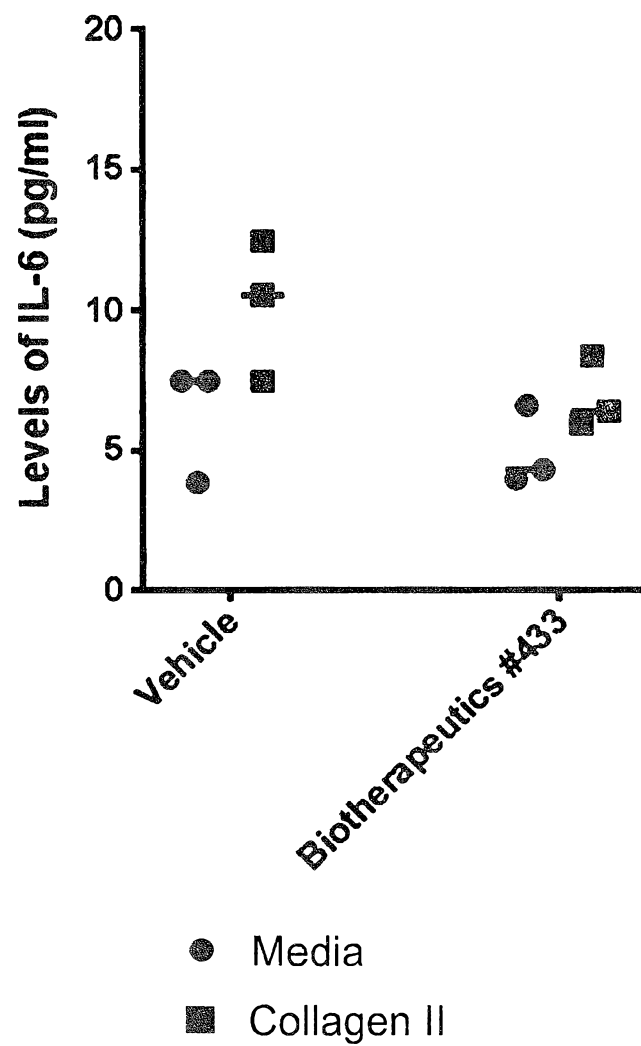
Splenocyte proliferation to Collagen II



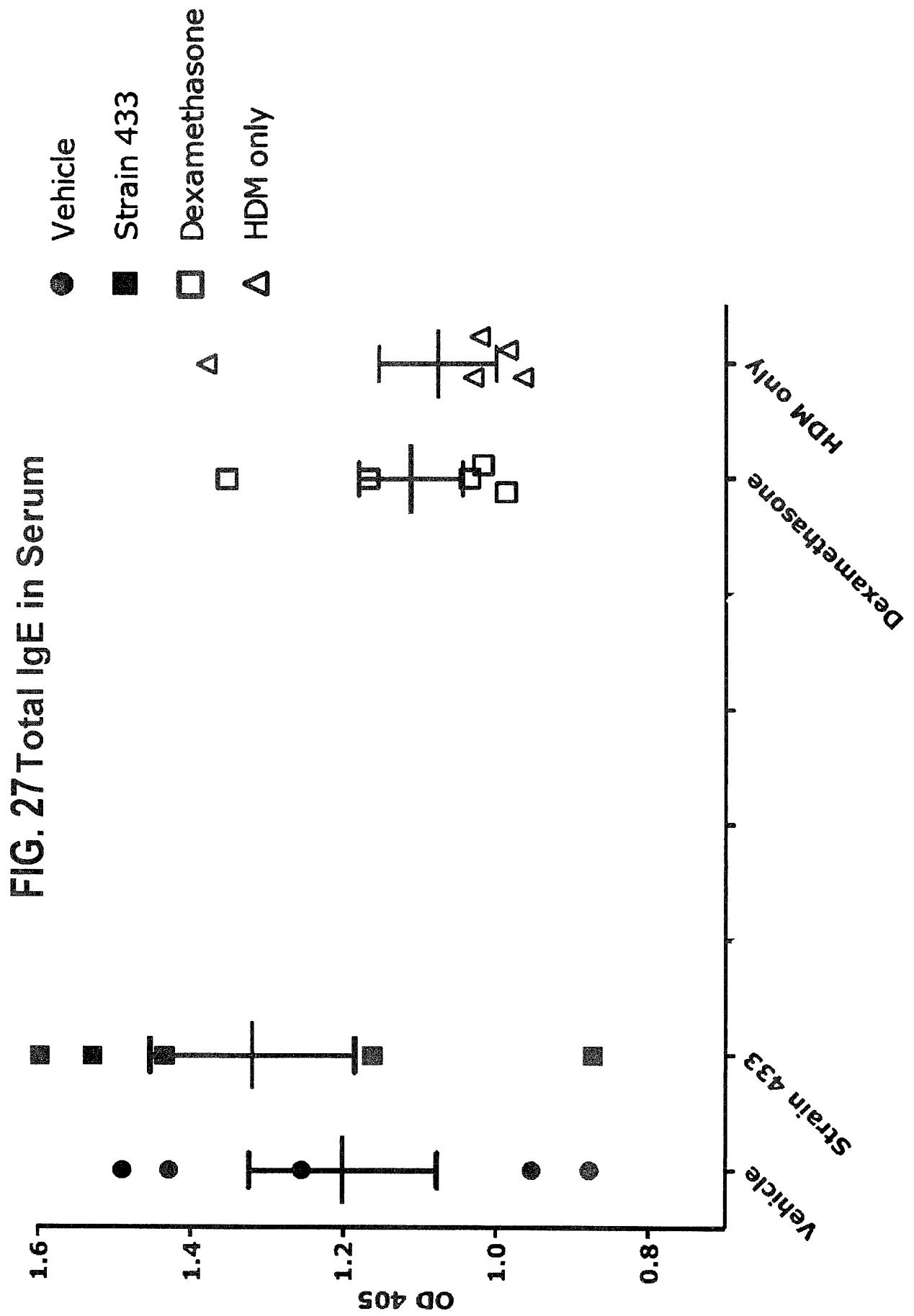


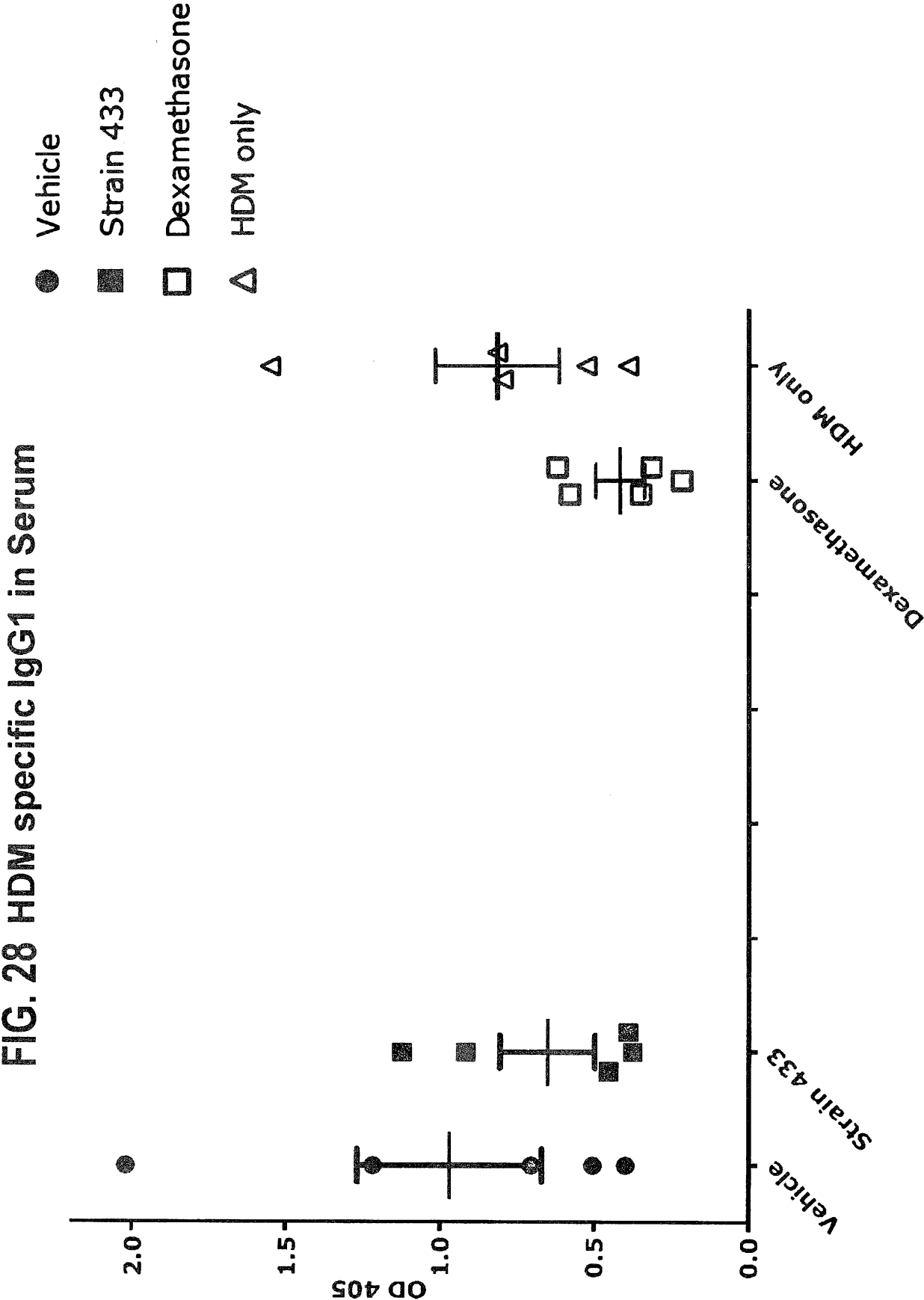
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FIG. 26



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FIG. 29 Total IgE in BALF

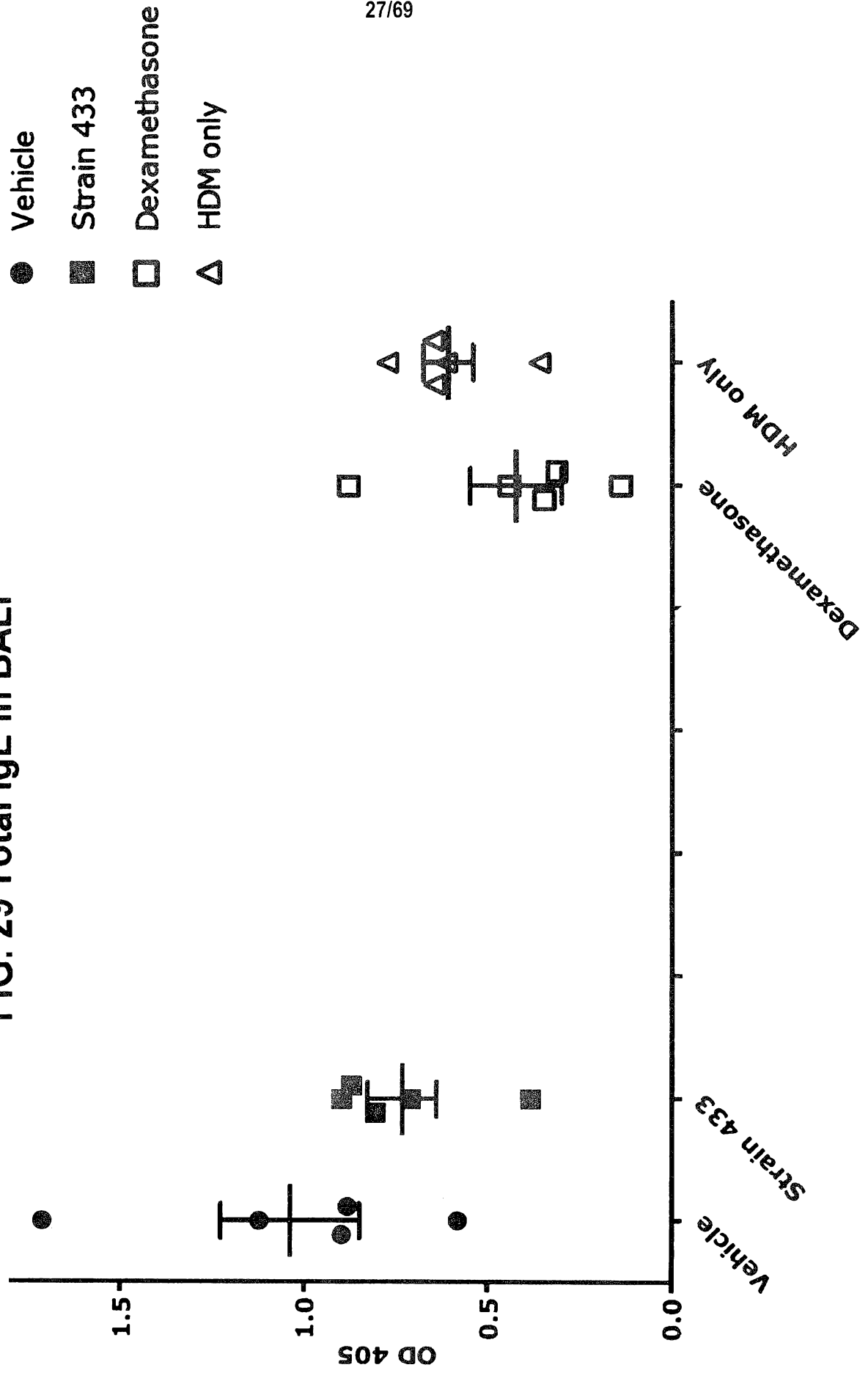
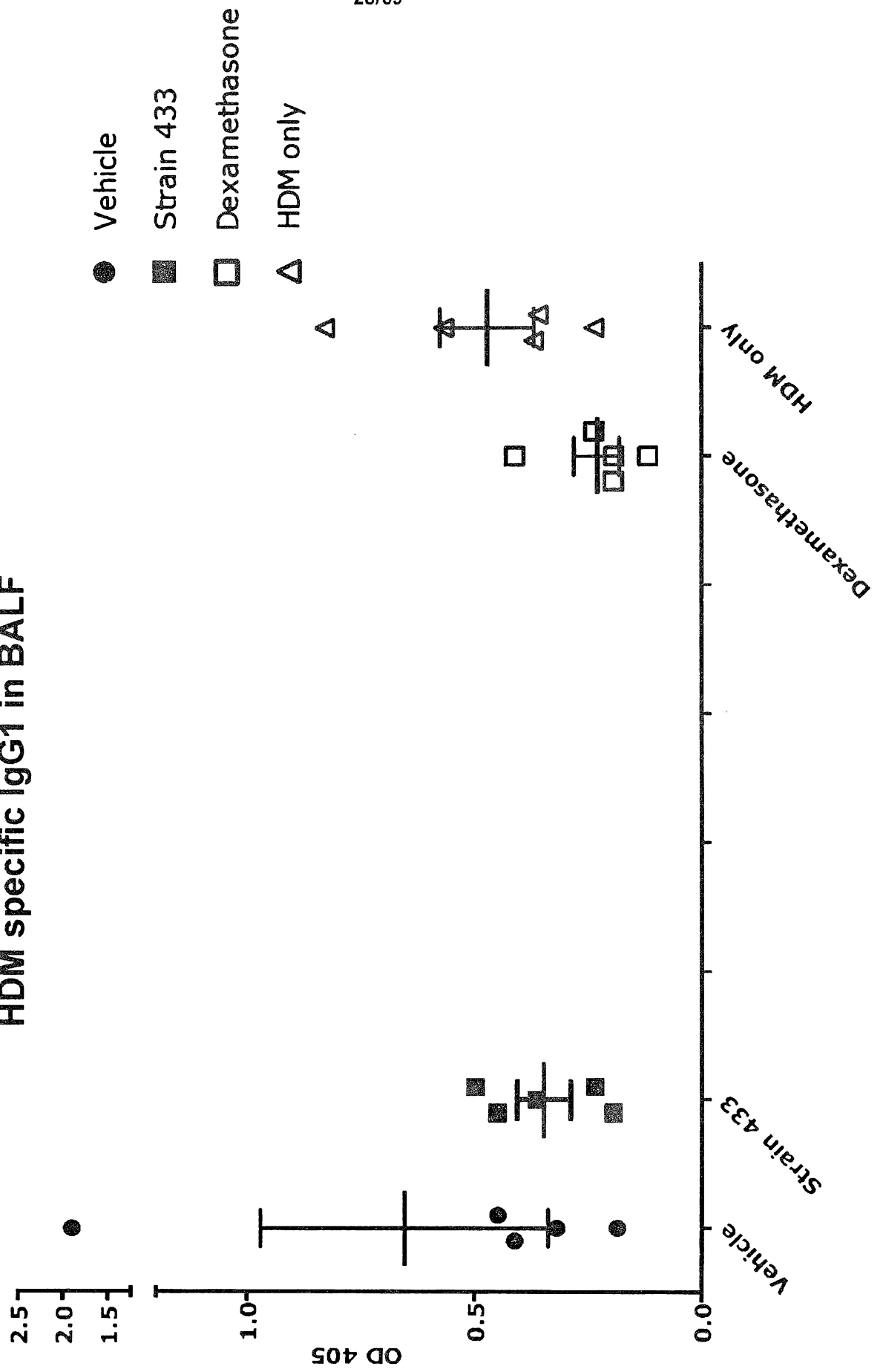
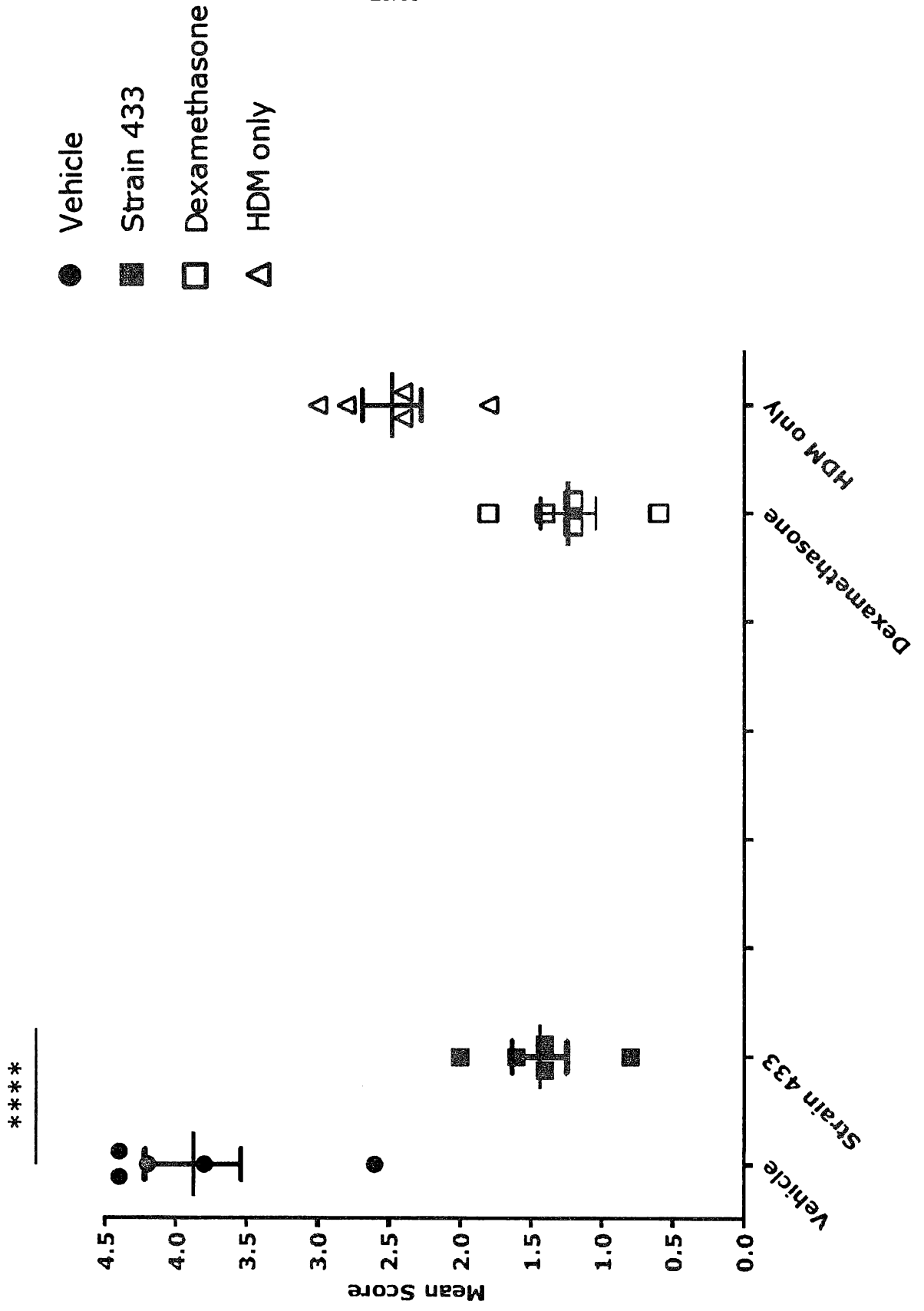


FIG. 30
HDM specific IgG1 in BALF



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FIG. 31 Histological Analysis – Mean Peribronchiolar Infiltration Score



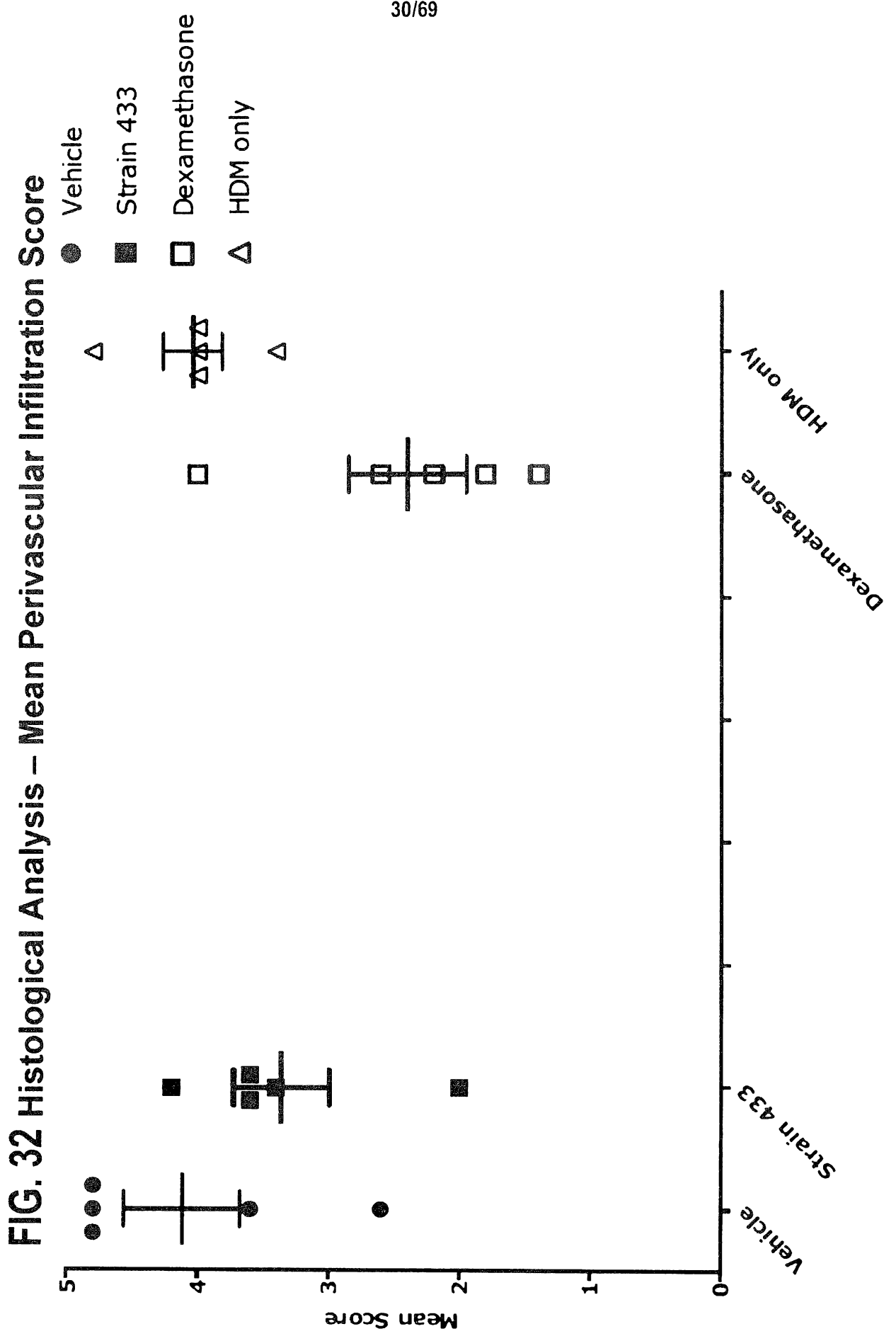
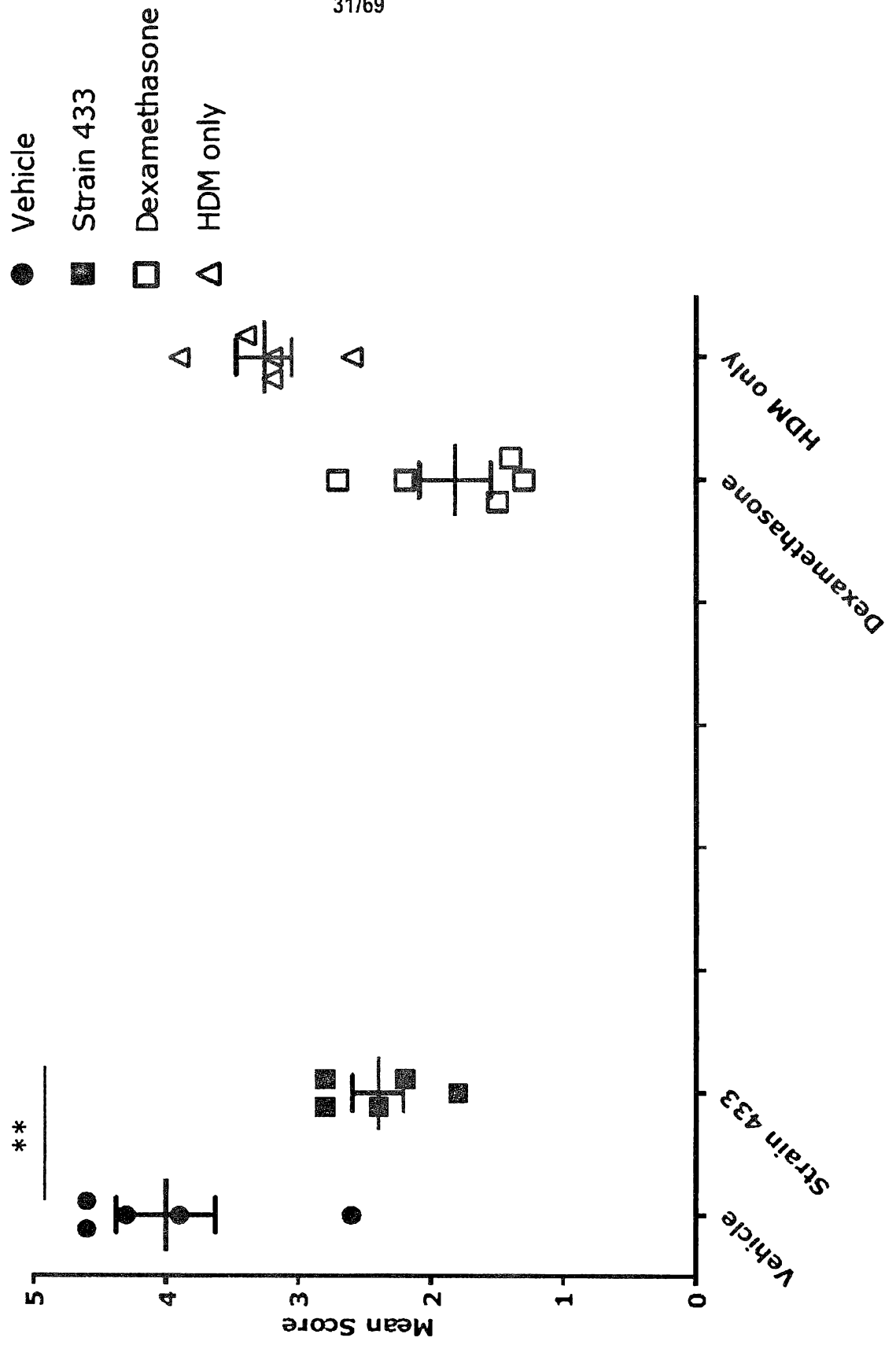


FIG. 33 Histological Analysis – Mean Inflammatory Score (Average of both Peribronchiolar and Perivascular Infiltration Score)



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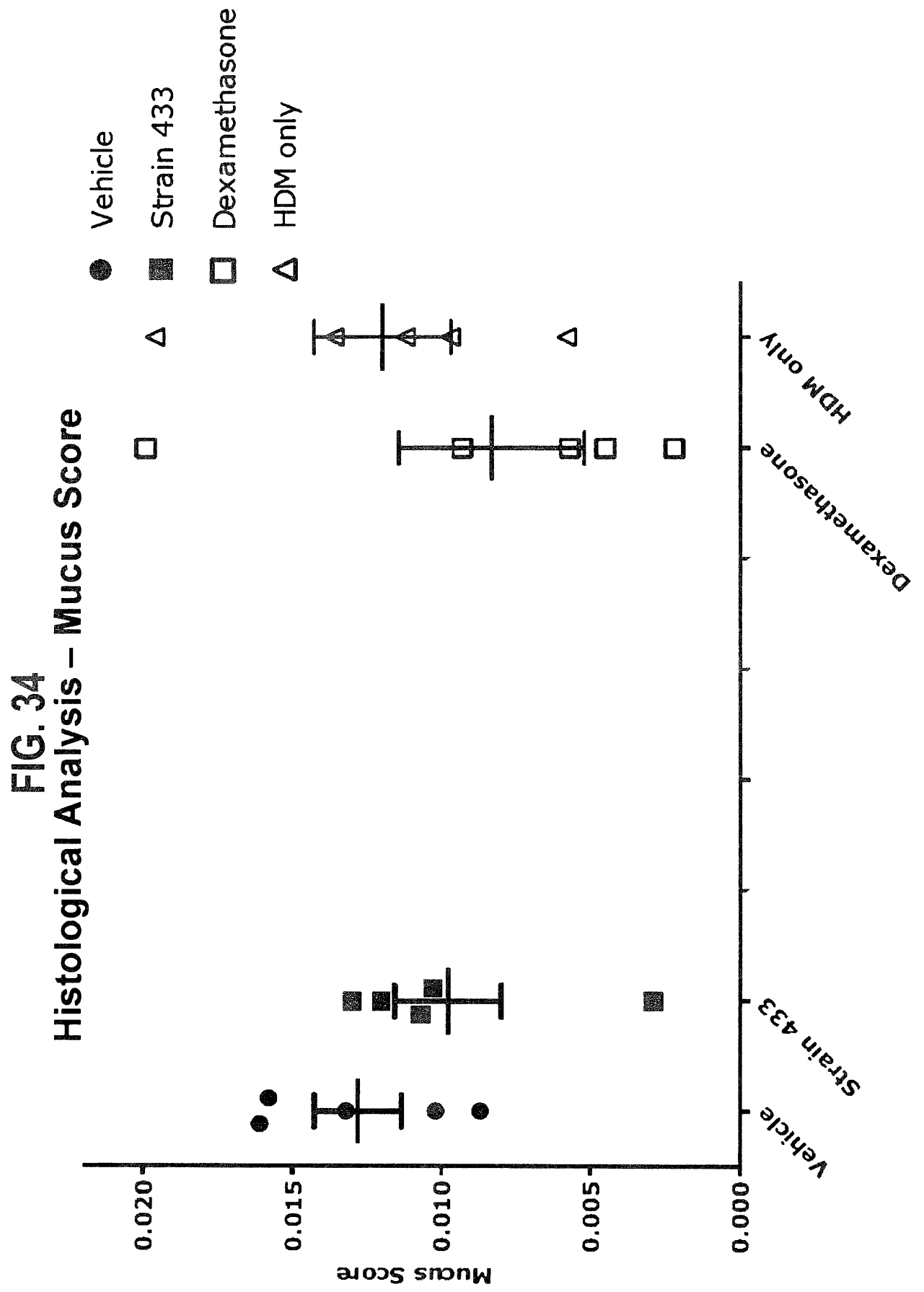
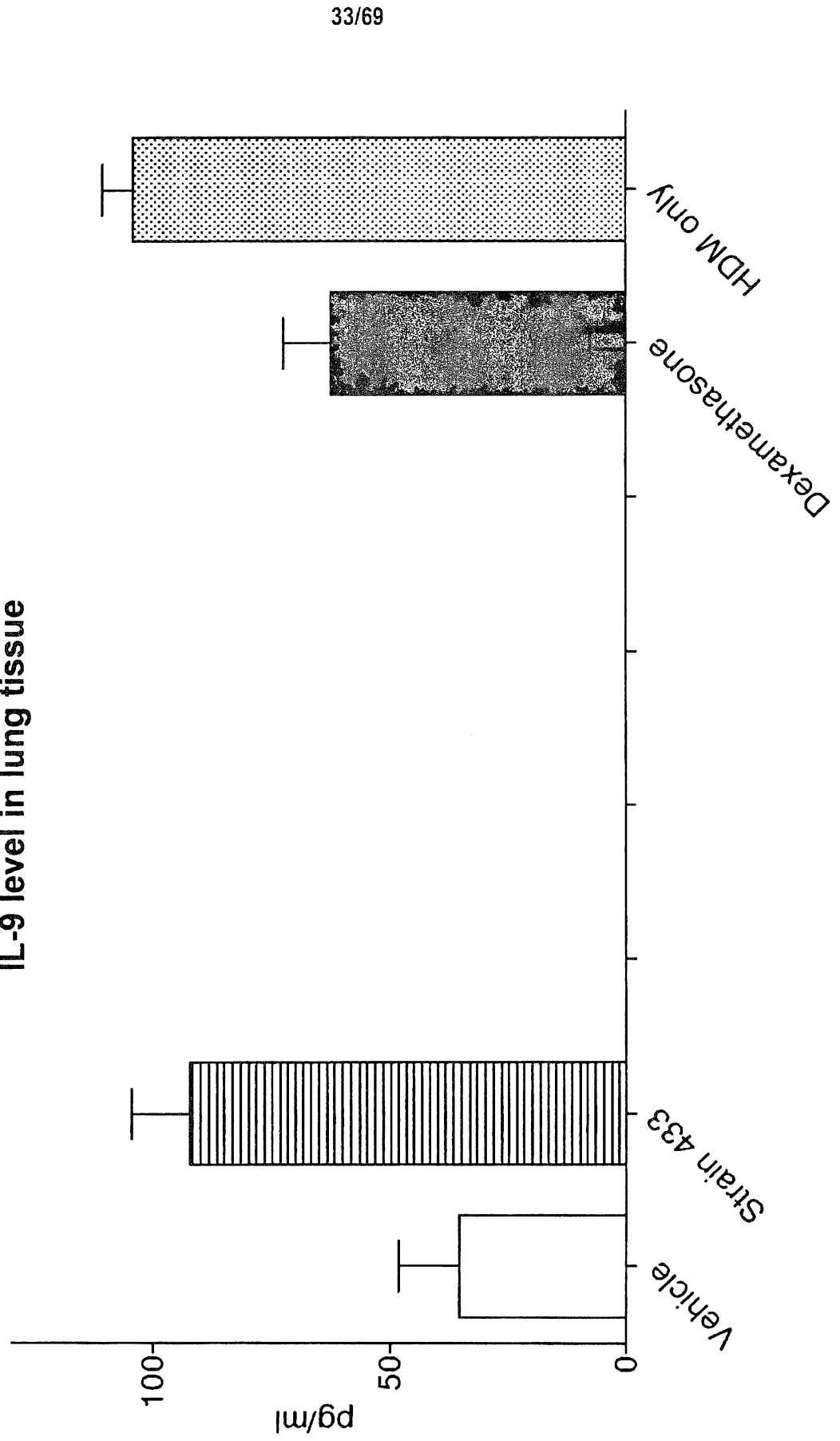


FIG. 35
IL-9 level in lung tissue



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FIG. 36
IL-1a level in lung tissue

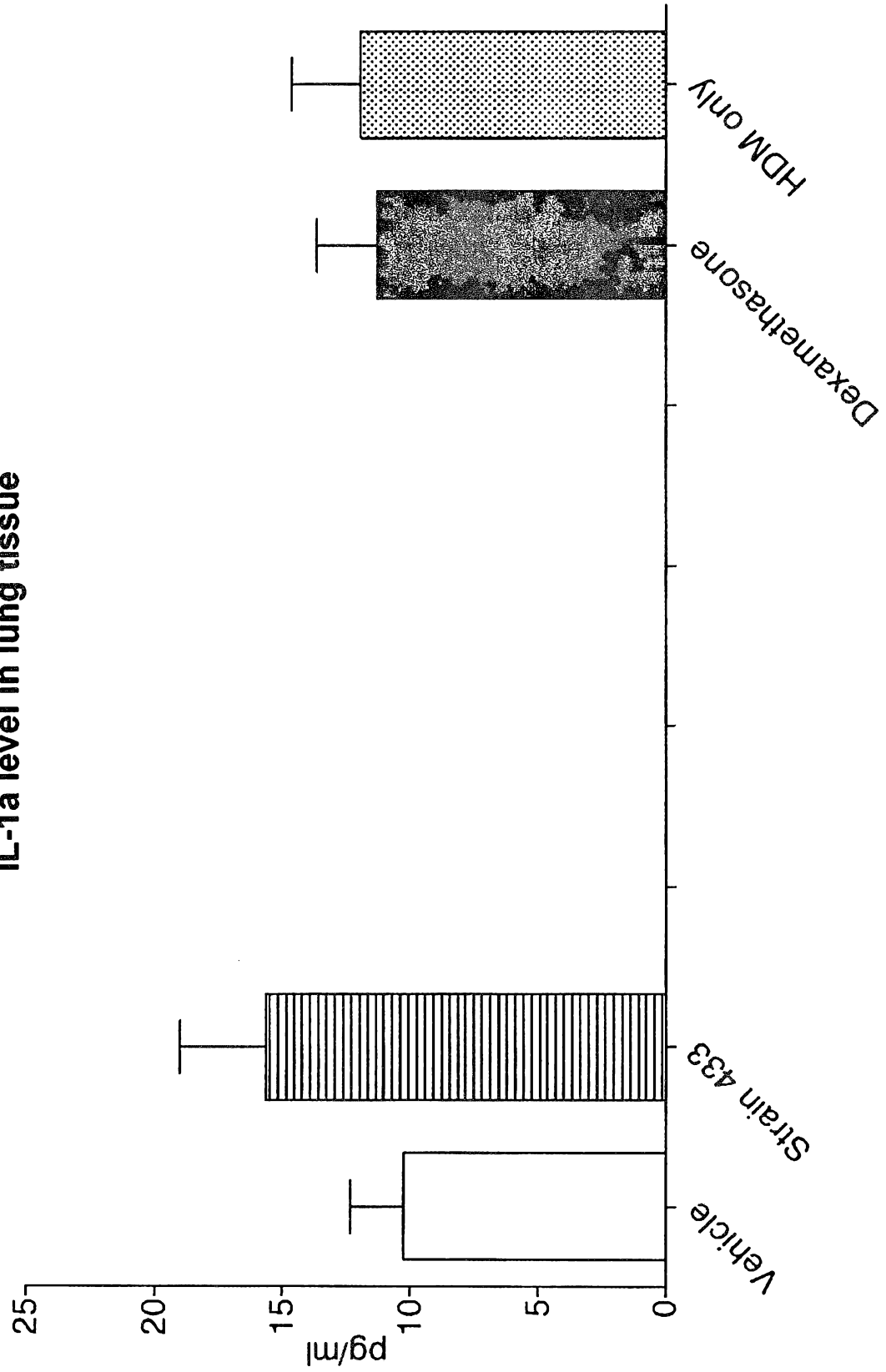
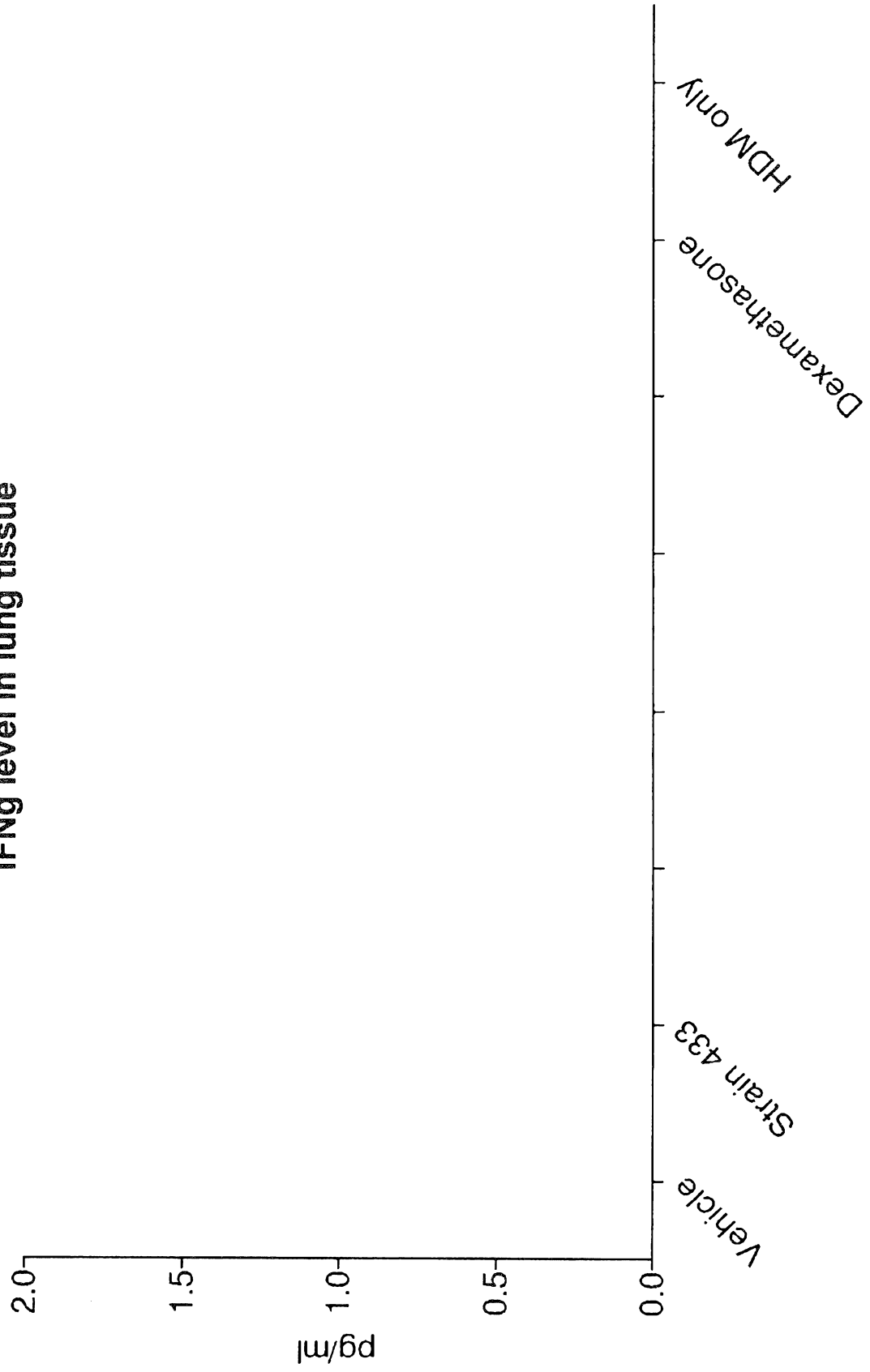
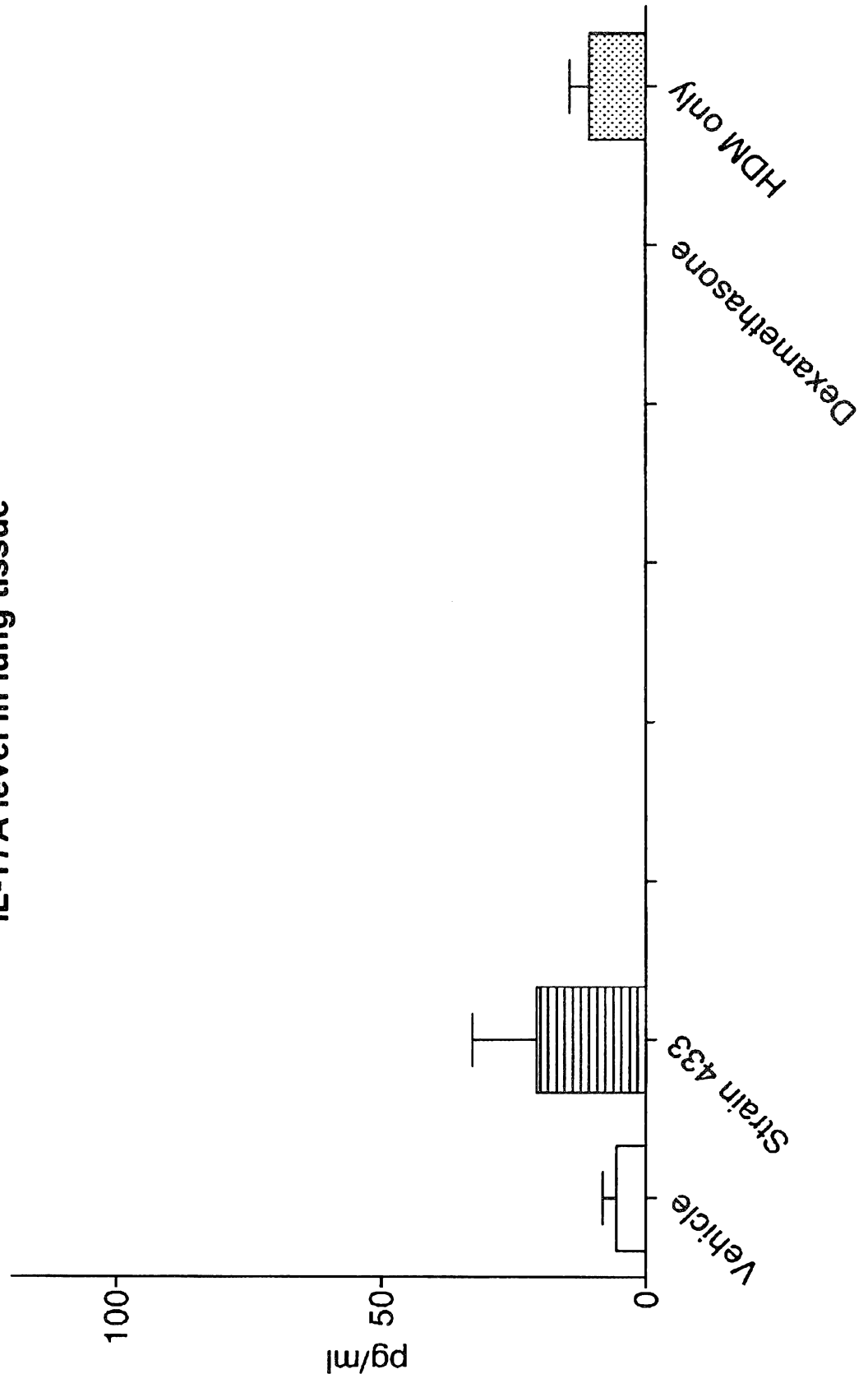


FIG. 37
IFNg level in lung tissue



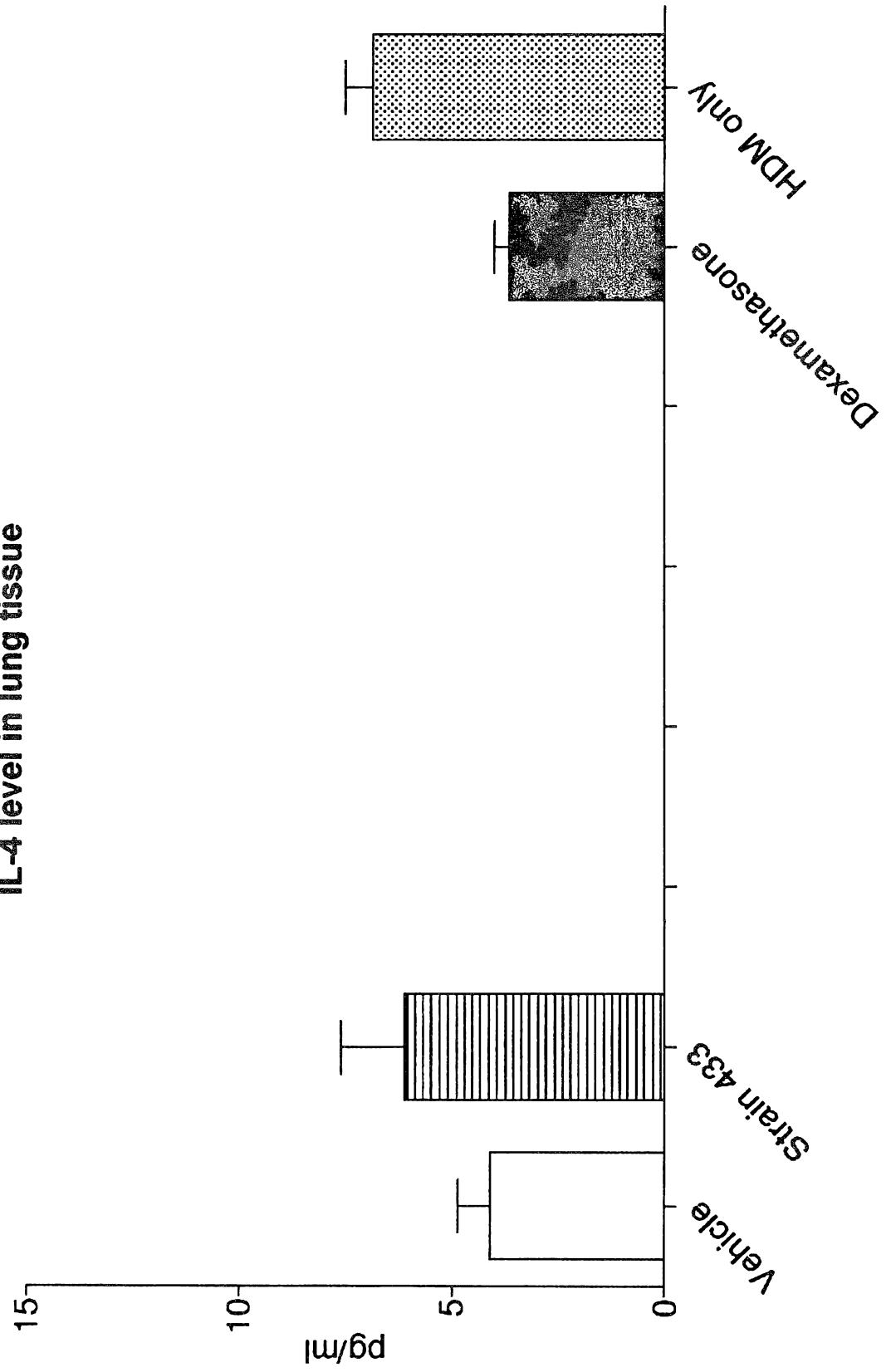
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FIG. 38
IL-17A level in lung tissue



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FIG. 39
IL-4 level in lung tissue



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FIG. 40
IL-5 level in lung tissue

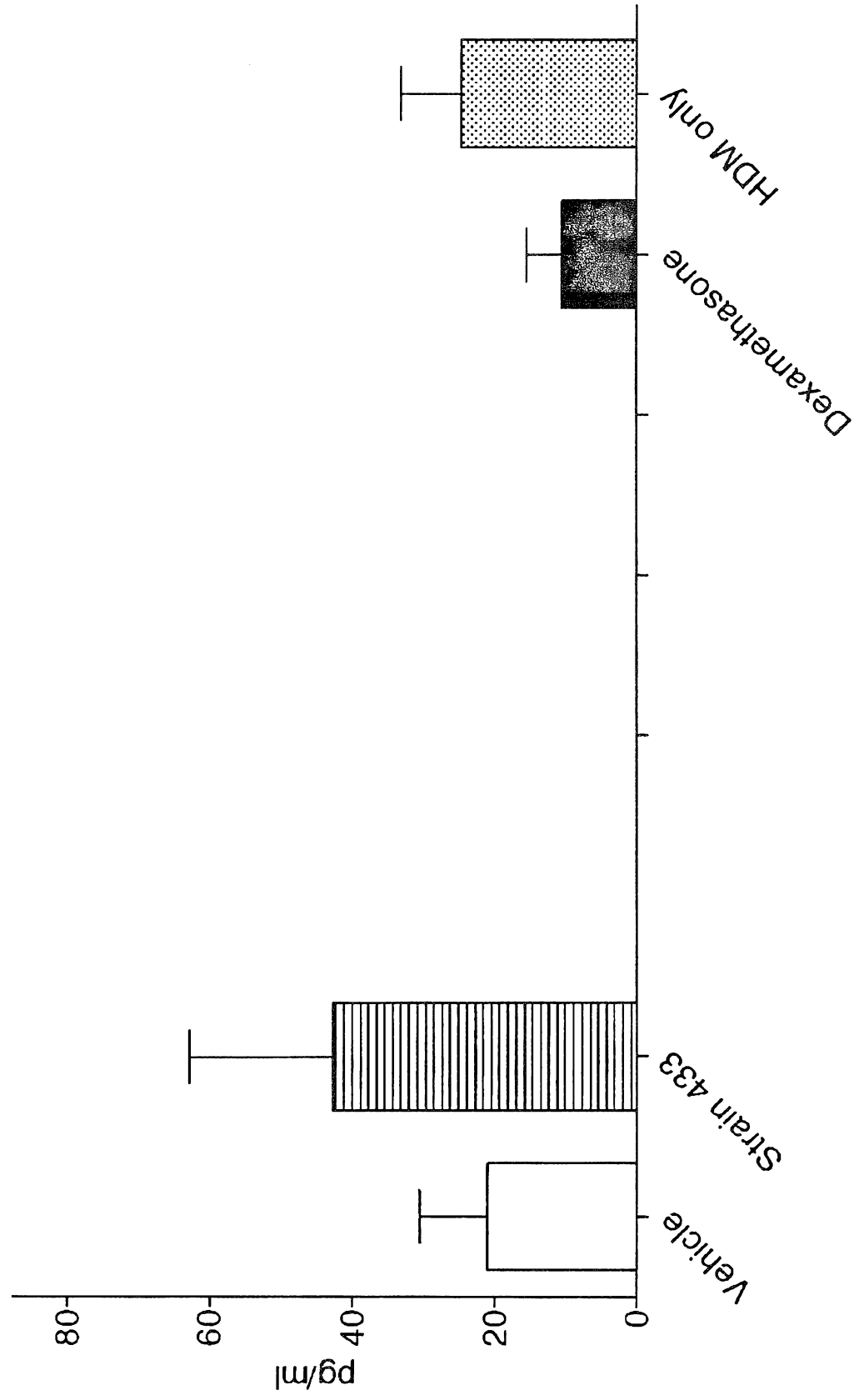
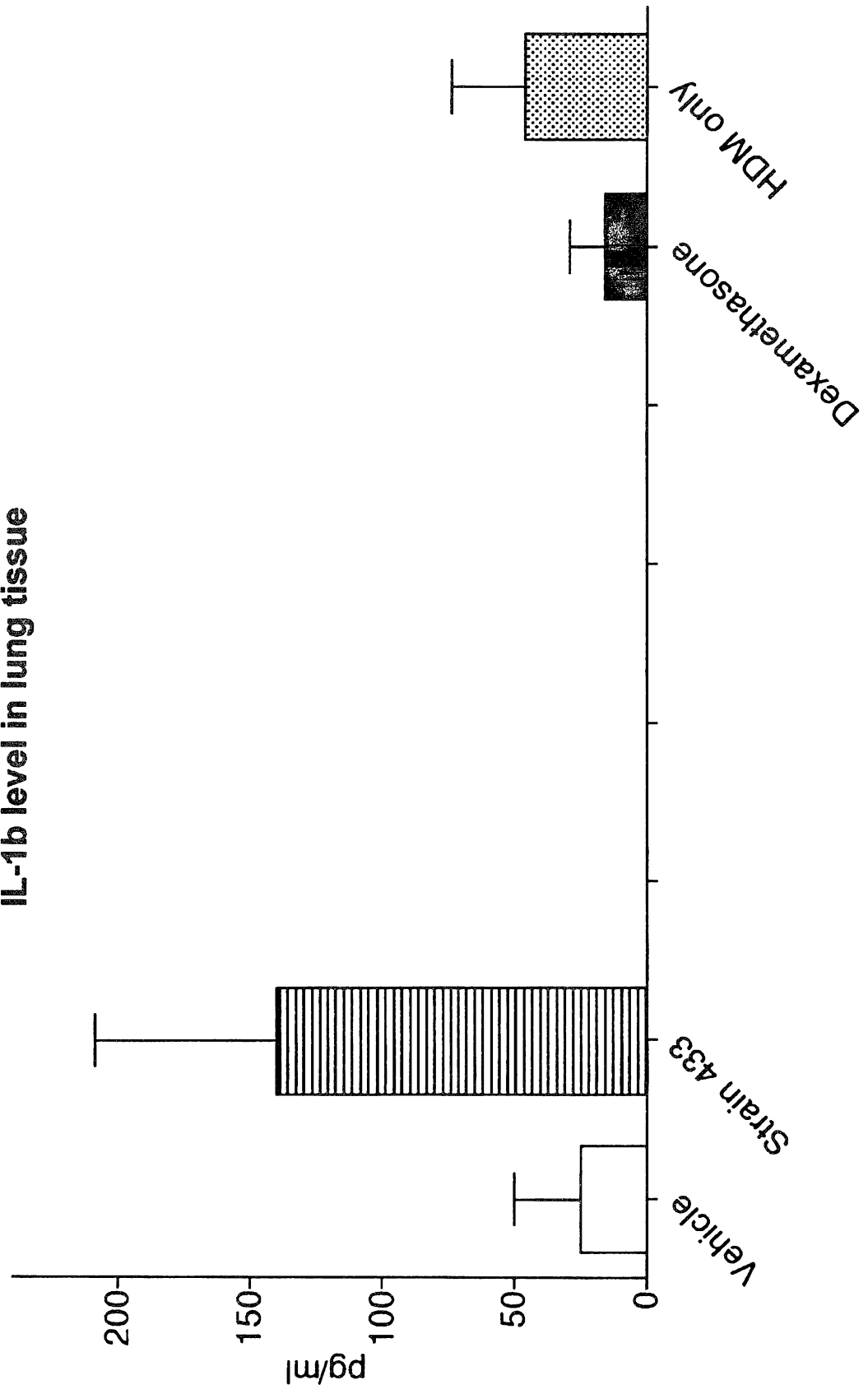
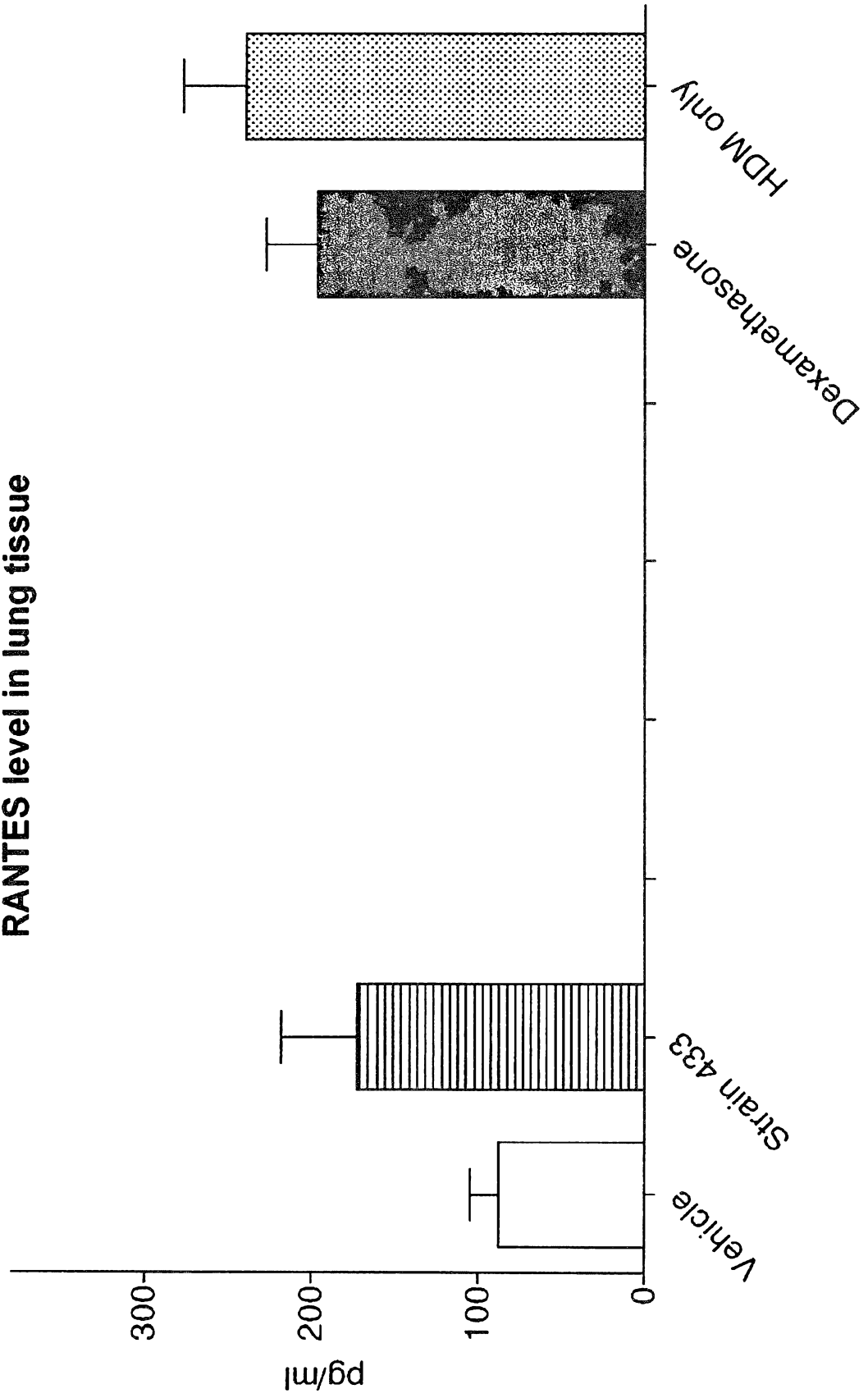


FIG. 41
IL-1b level in lung tissue



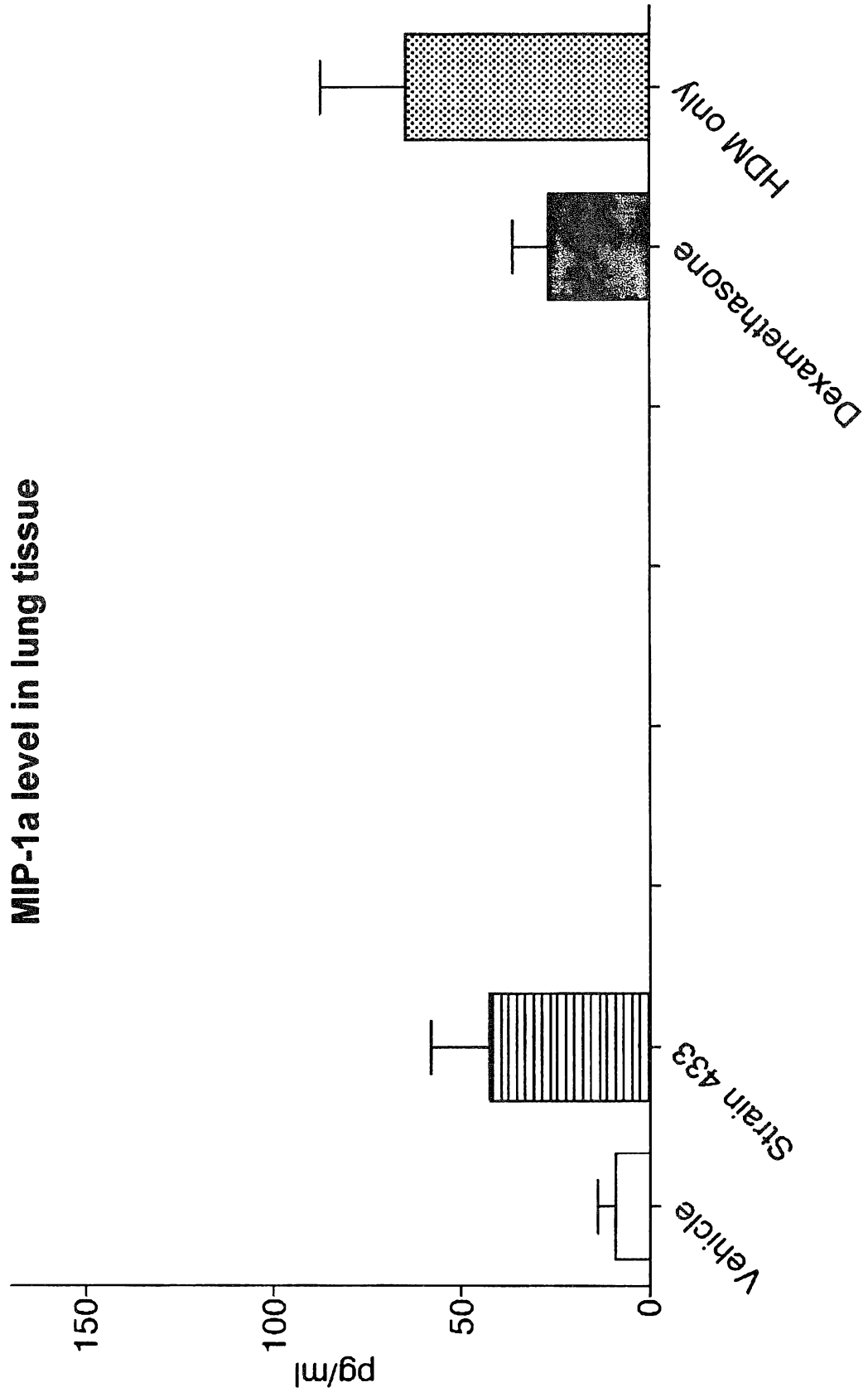
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FIG. 42
RANTES level in lung tissue



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FIG. 43
MIP-1a level in lung tissue



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FIG. 44
KC level in lung tissue

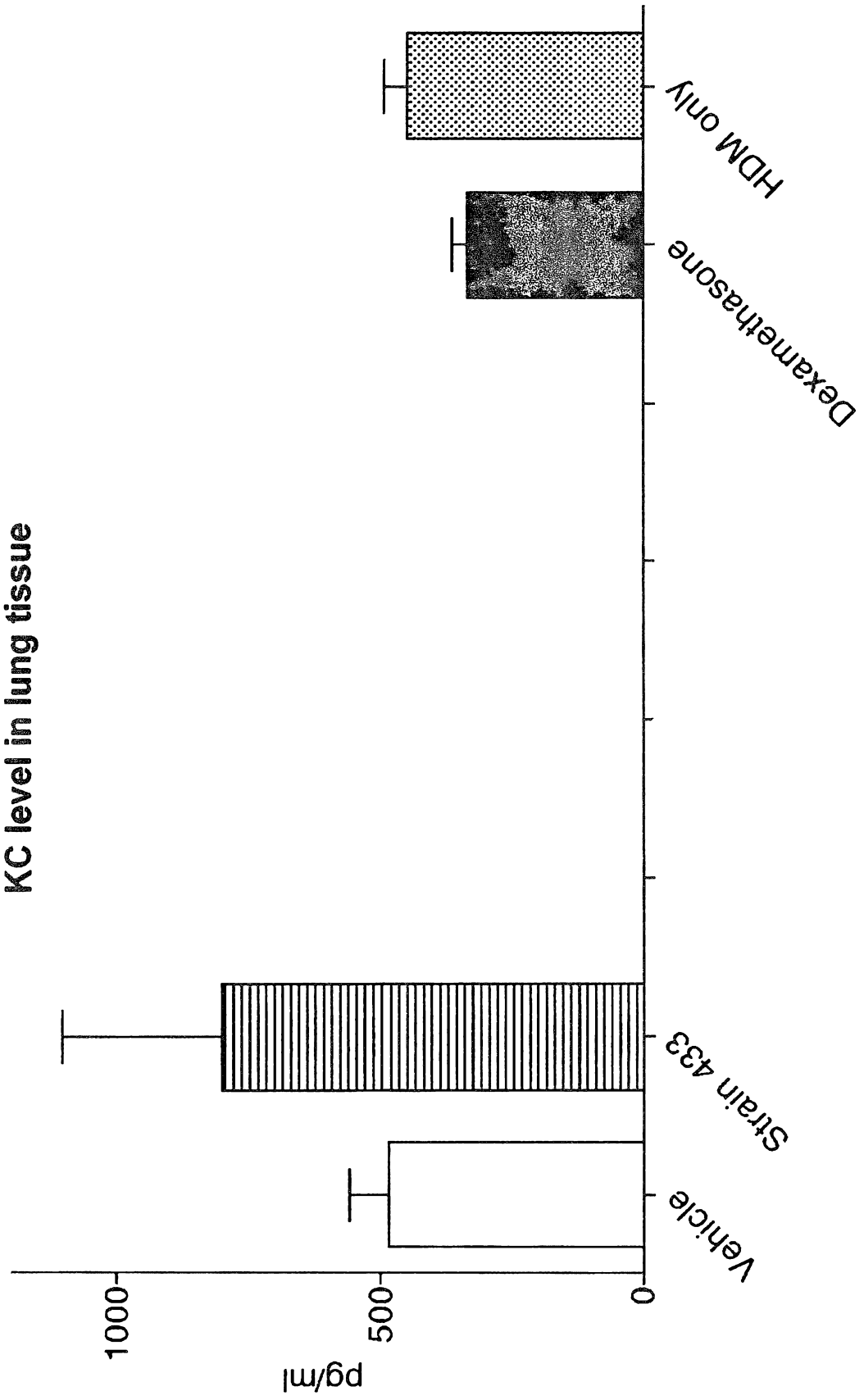
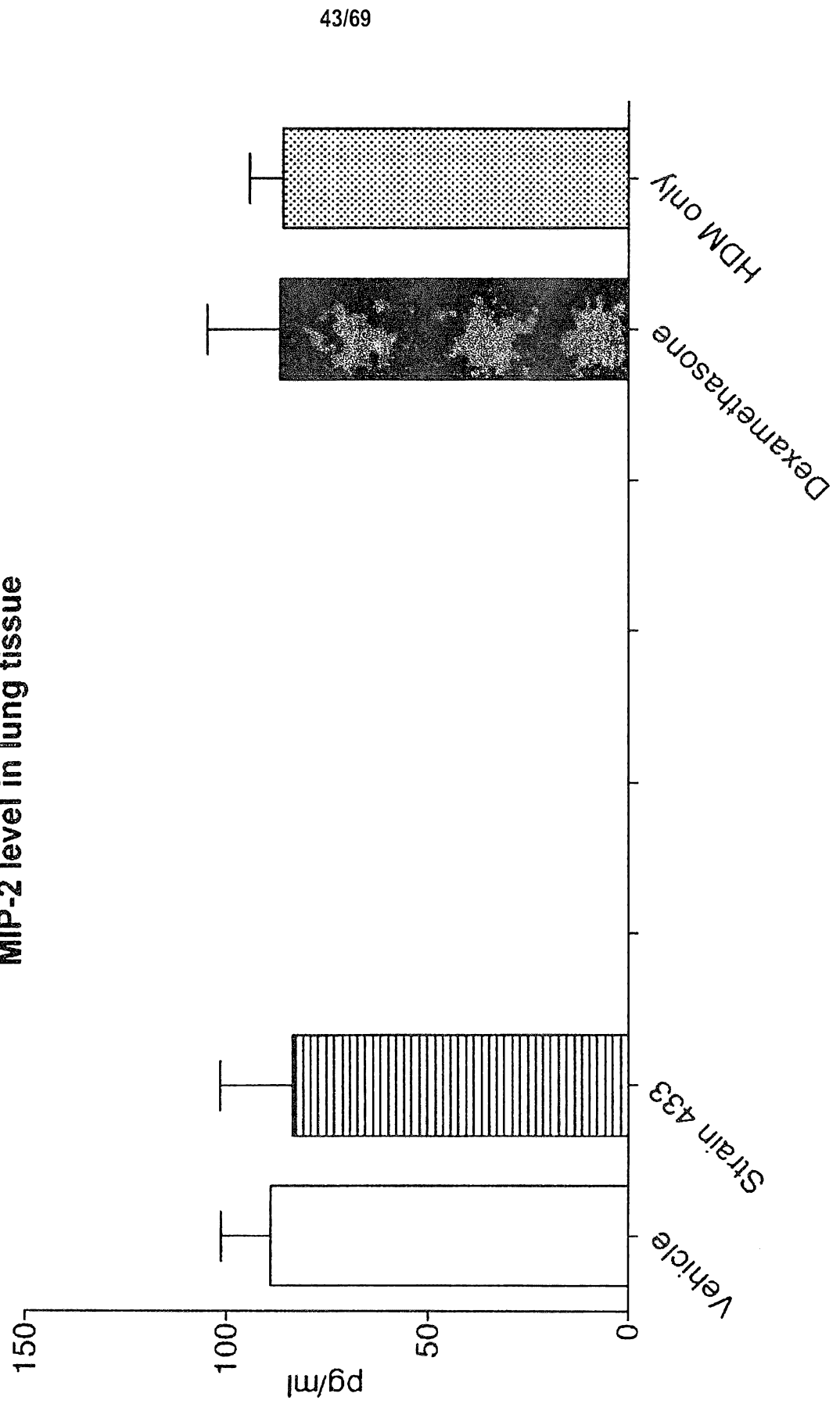


FIG. 45
MIP-2 level in lung tissue



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FIG. 46 HDM specific IgG1 in Serum

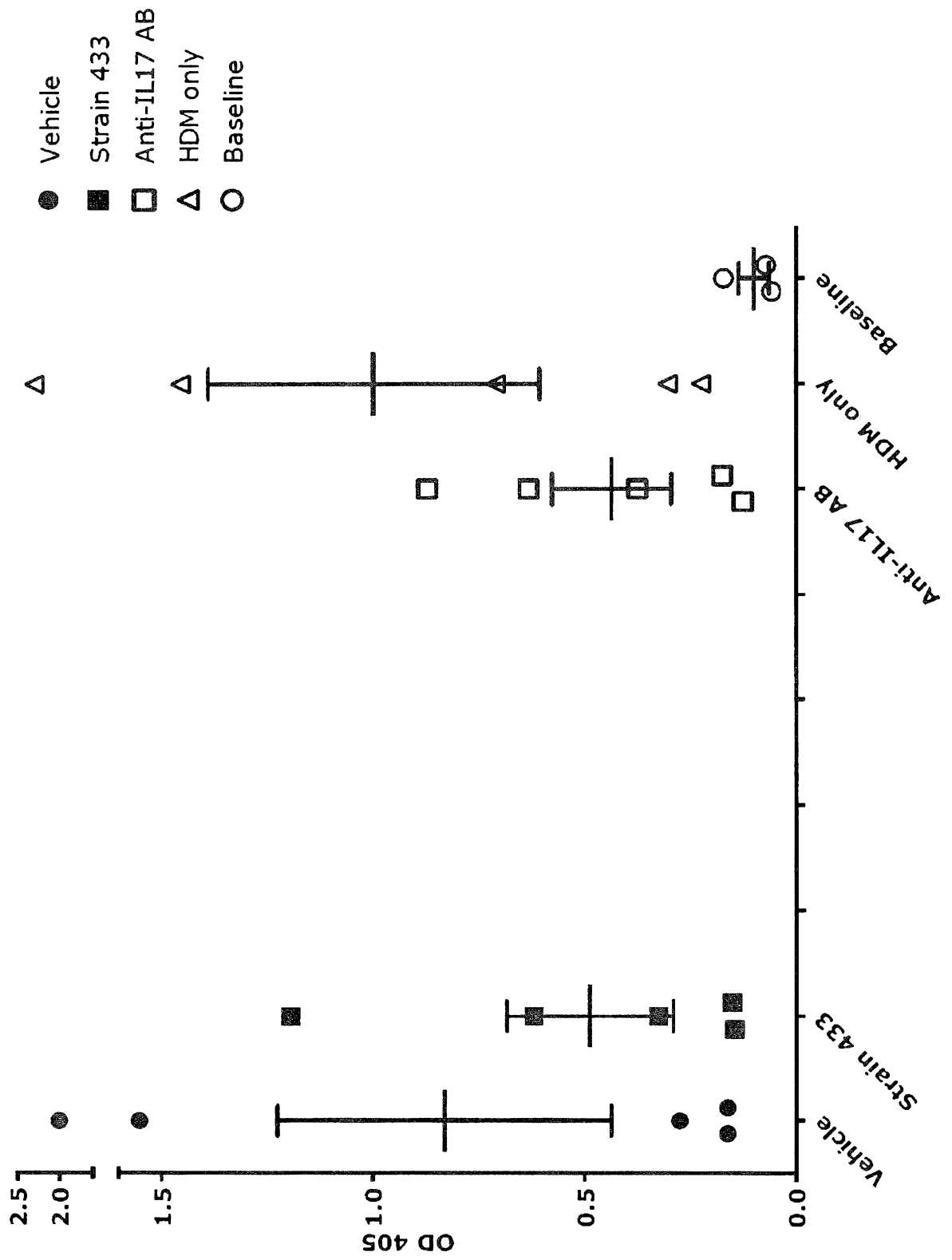
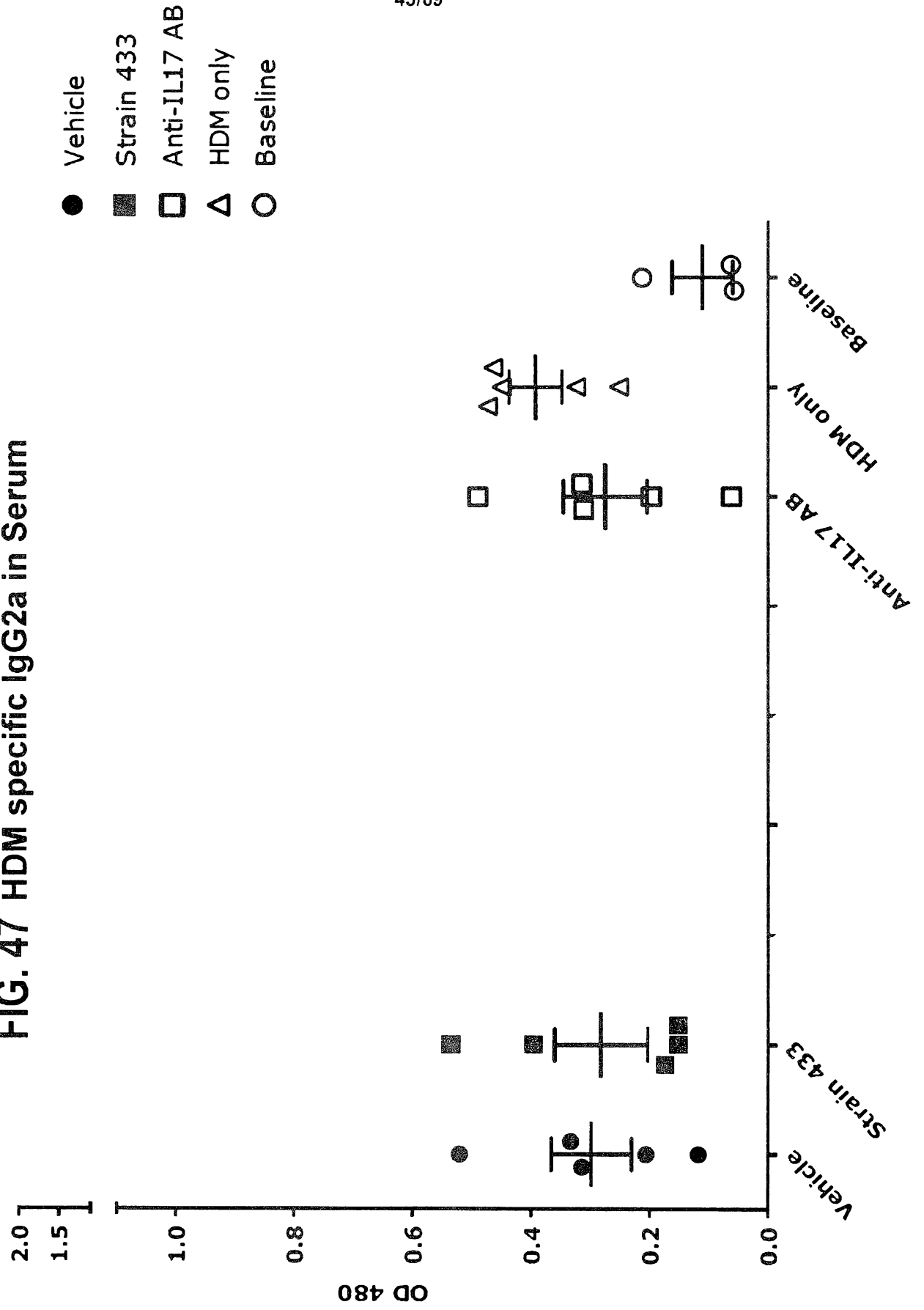
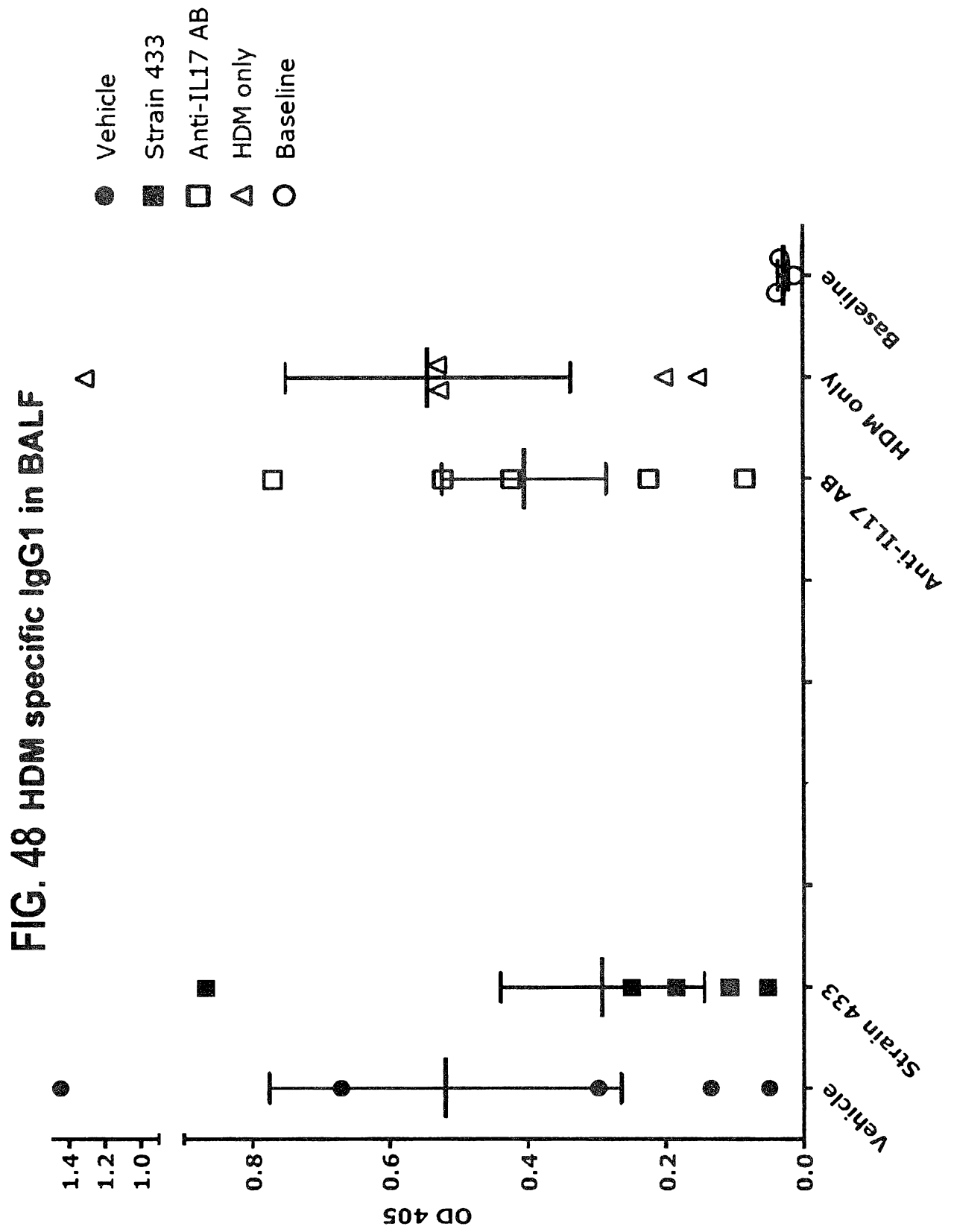


FIG. 47 HDM specific IgG2a in Serum

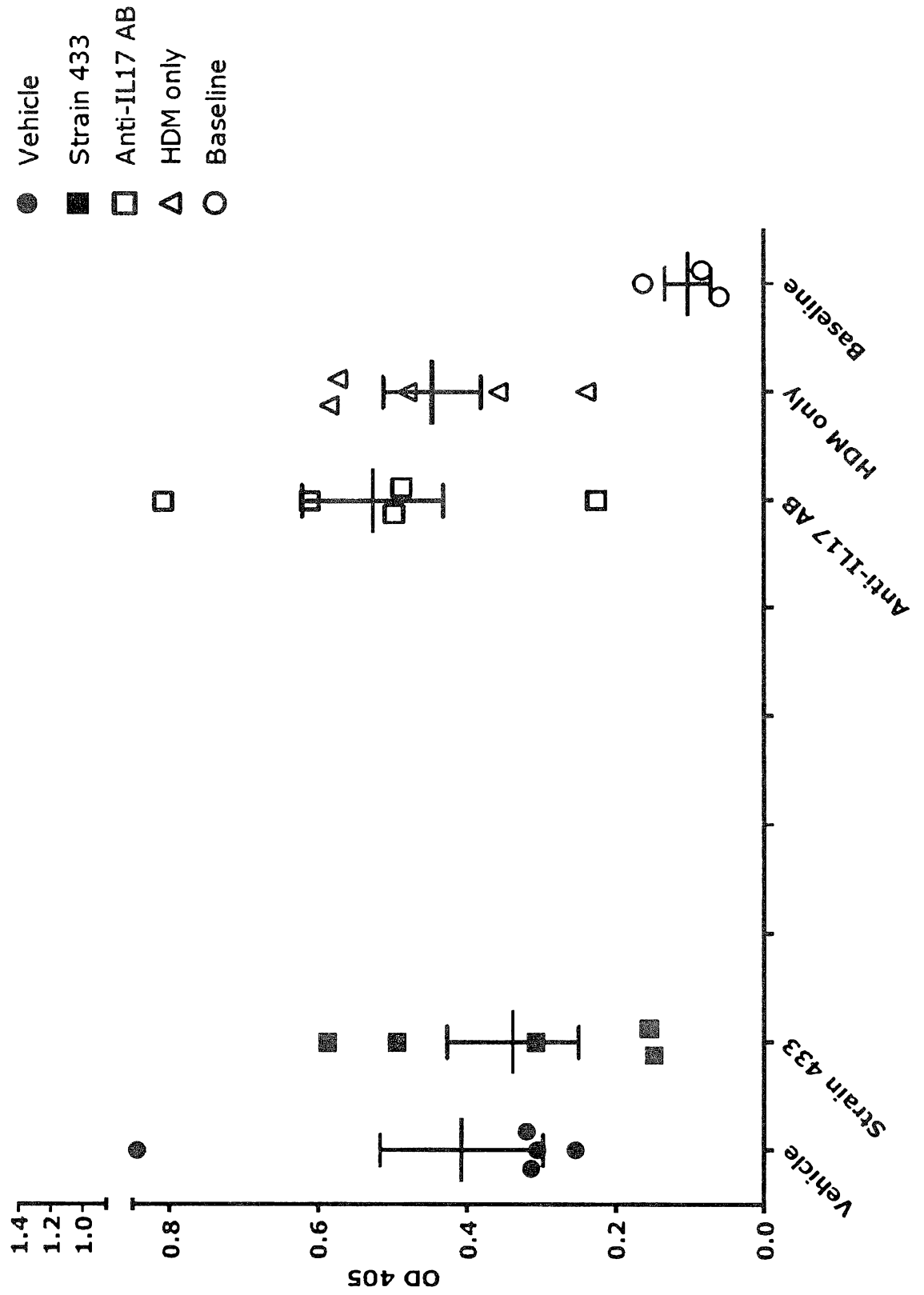


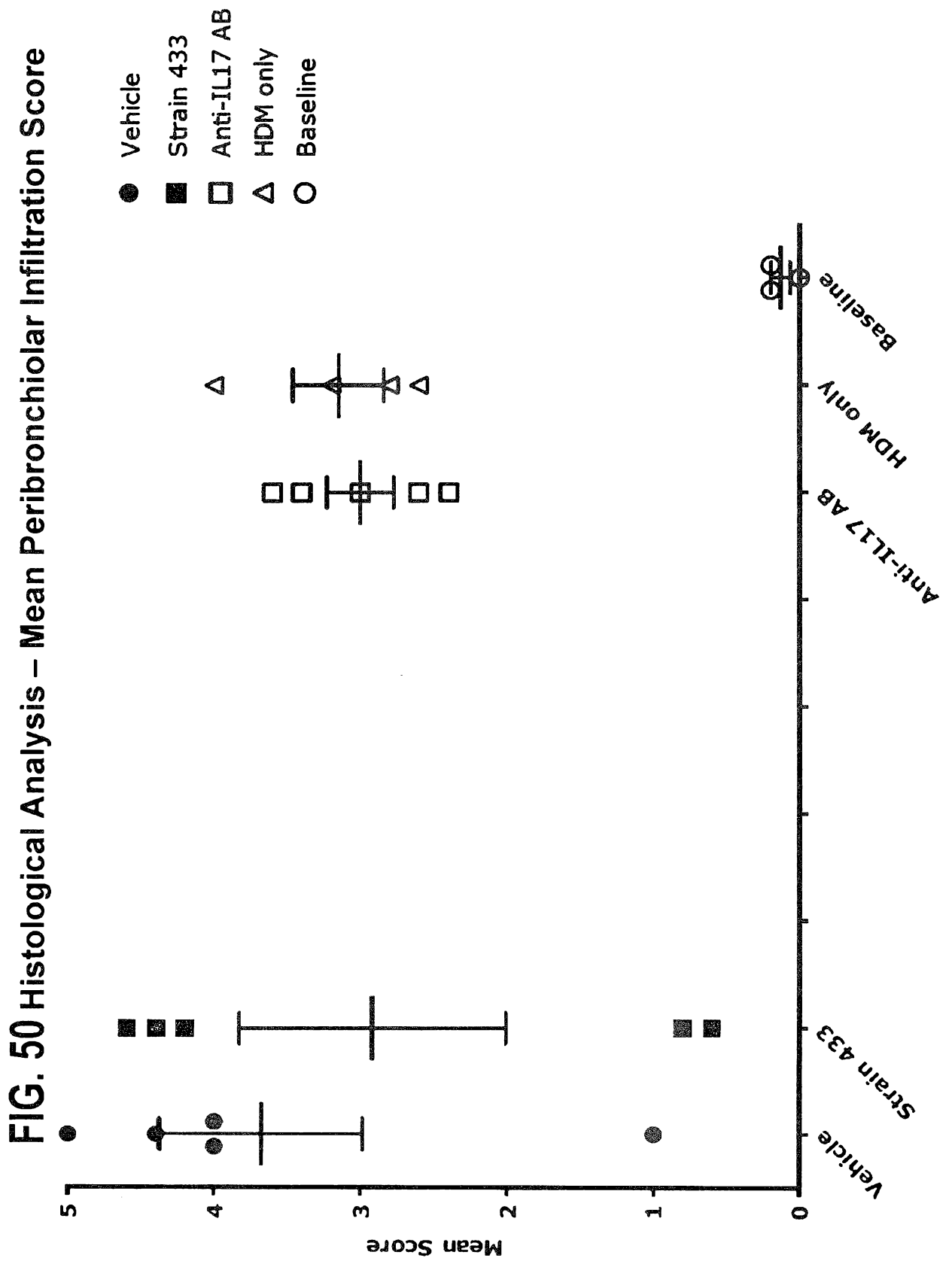
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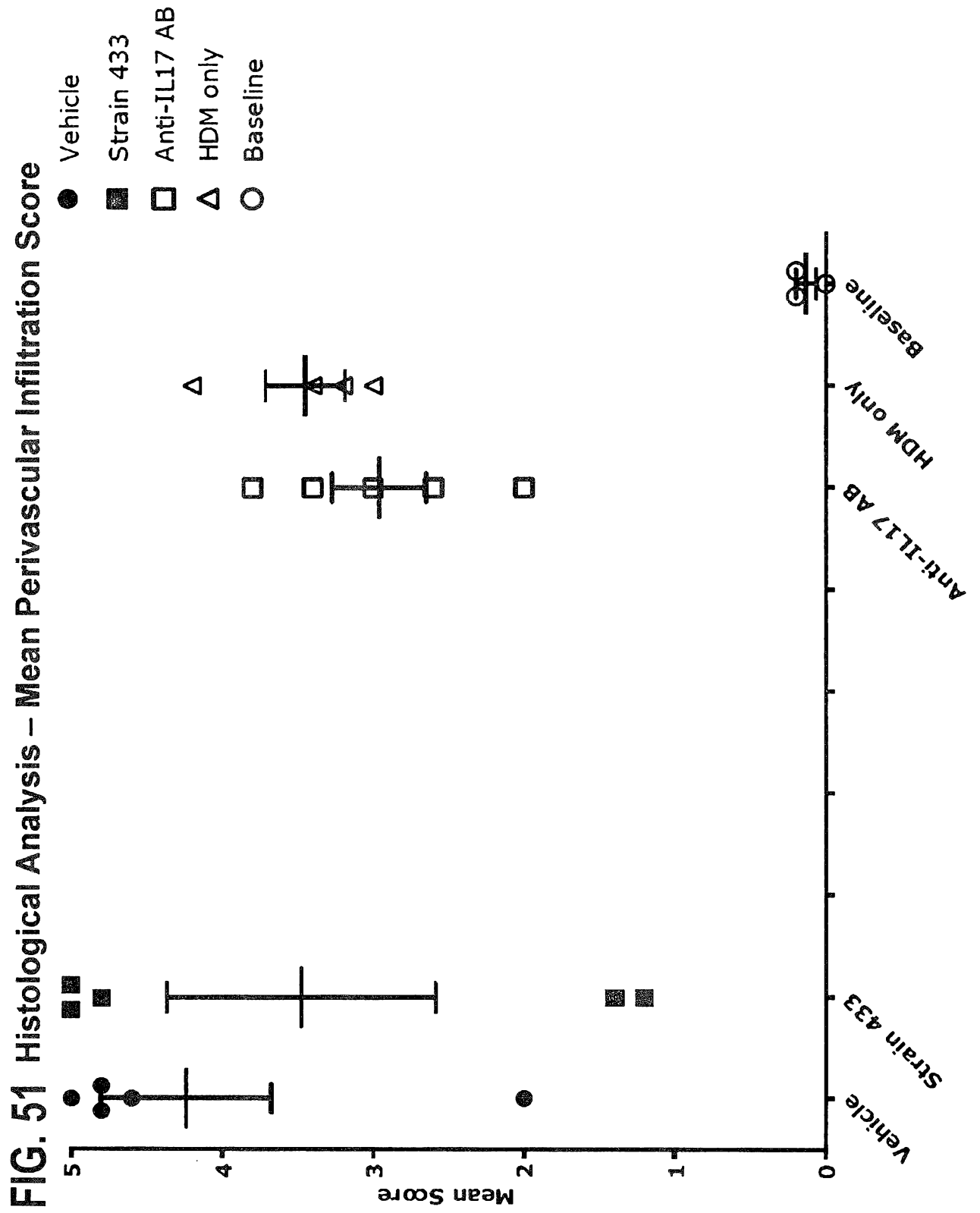


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FIG. 49 HDM specific IgG2a in BALF







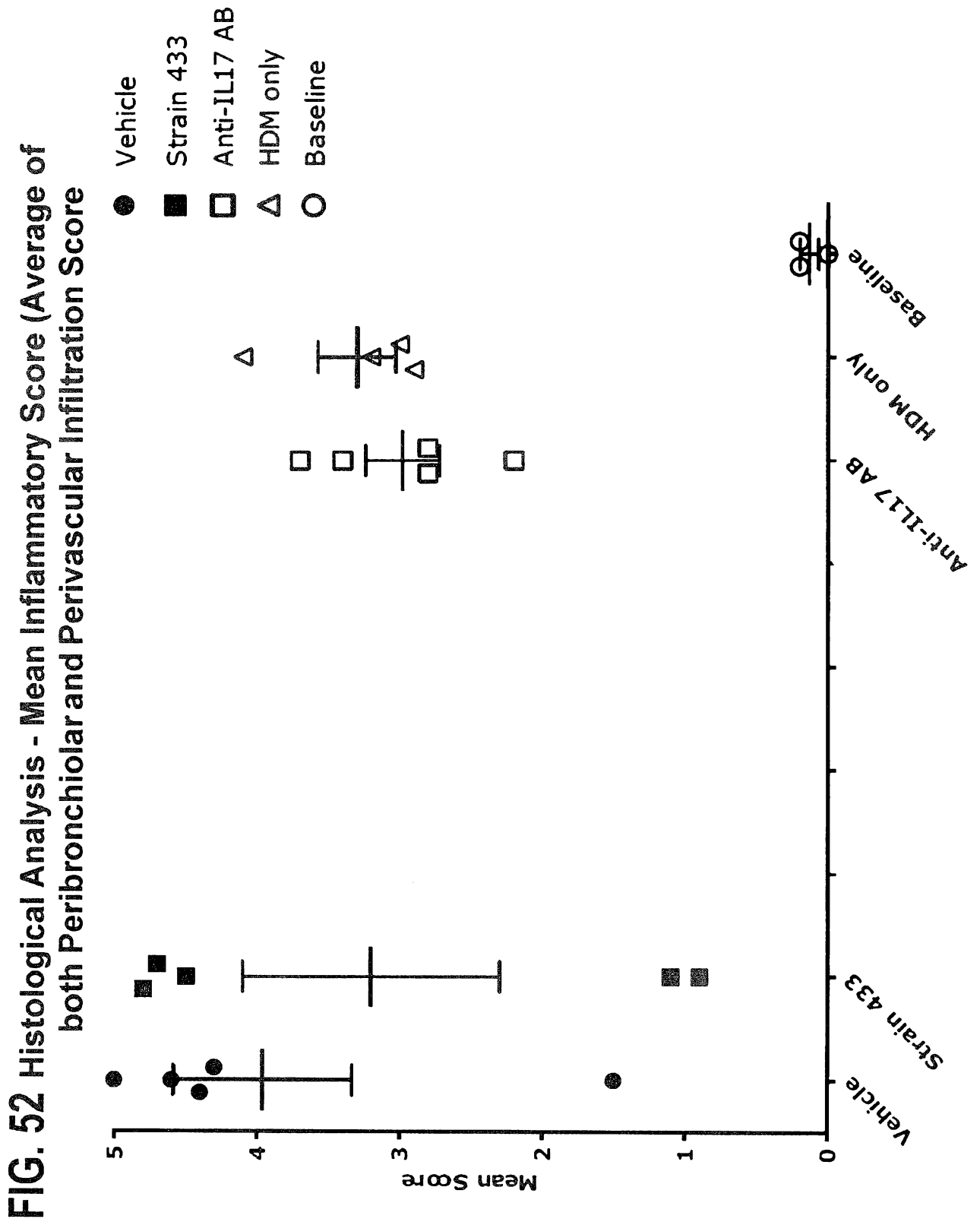
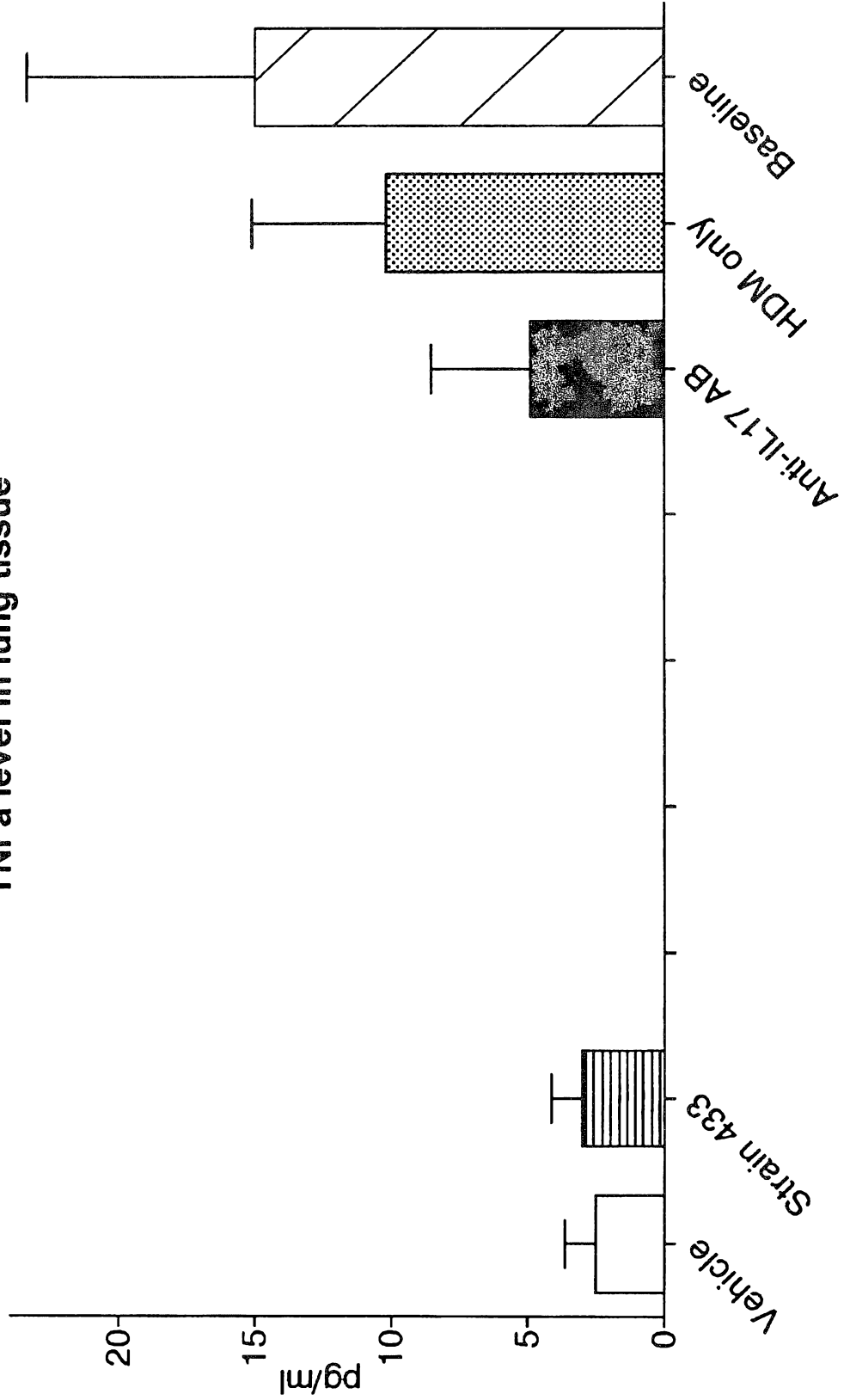
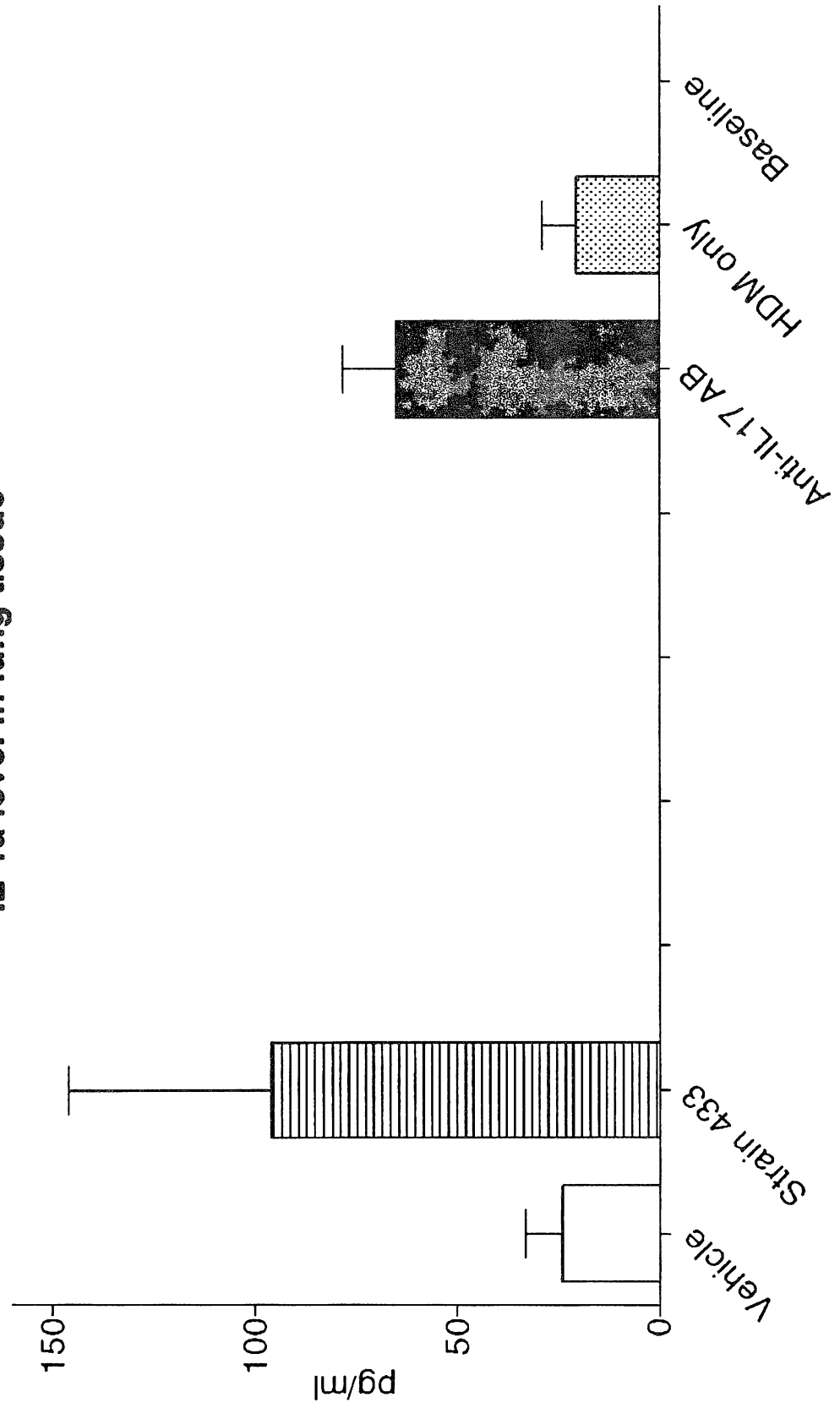


FIG. 53
TNFα level in lung tissue



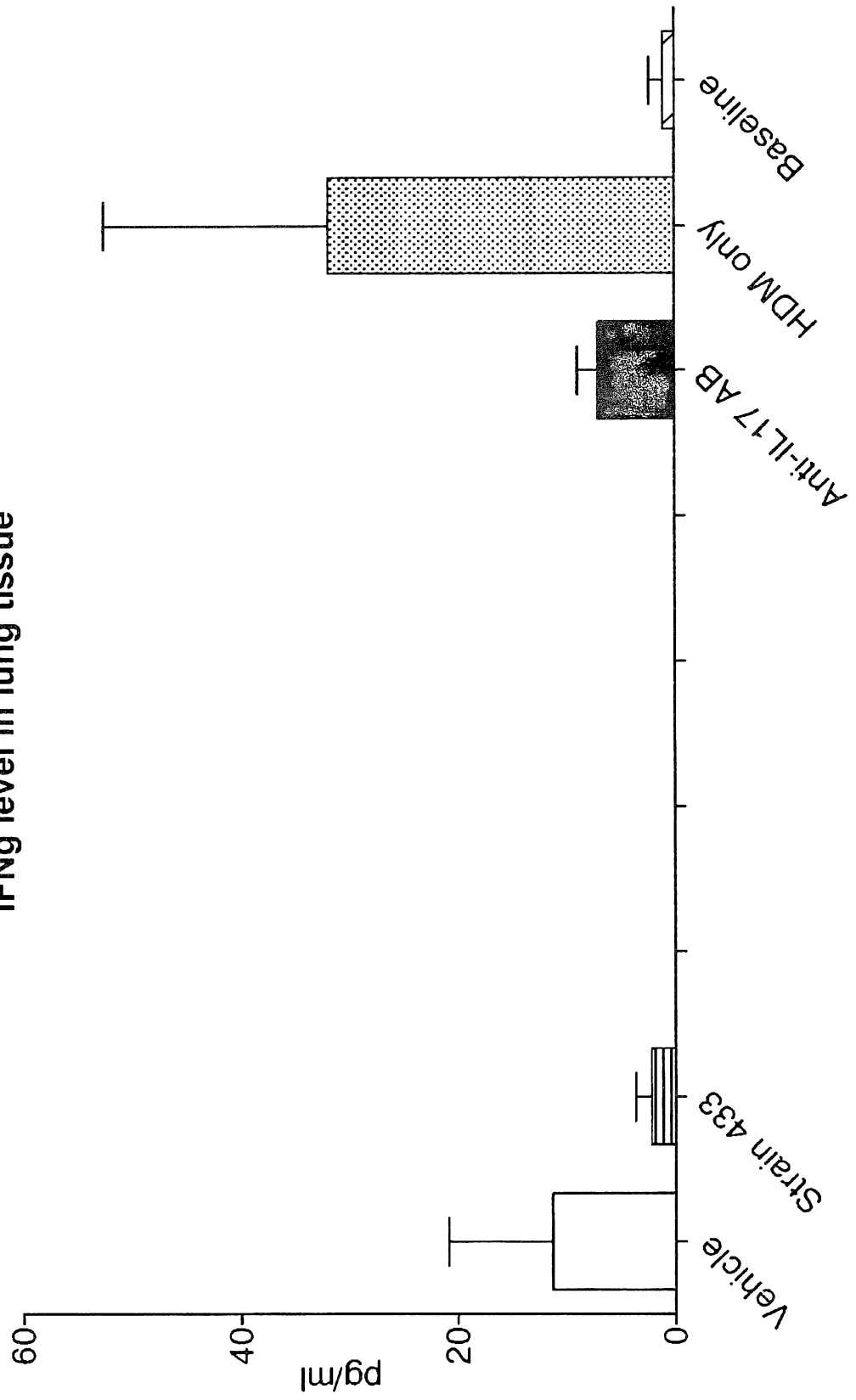
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FIG. 54
IL-1a level in lung tissue



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FIG. 55
IFN γ level in lung tissue



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FIG. 56
IL-17F level in lung tissue

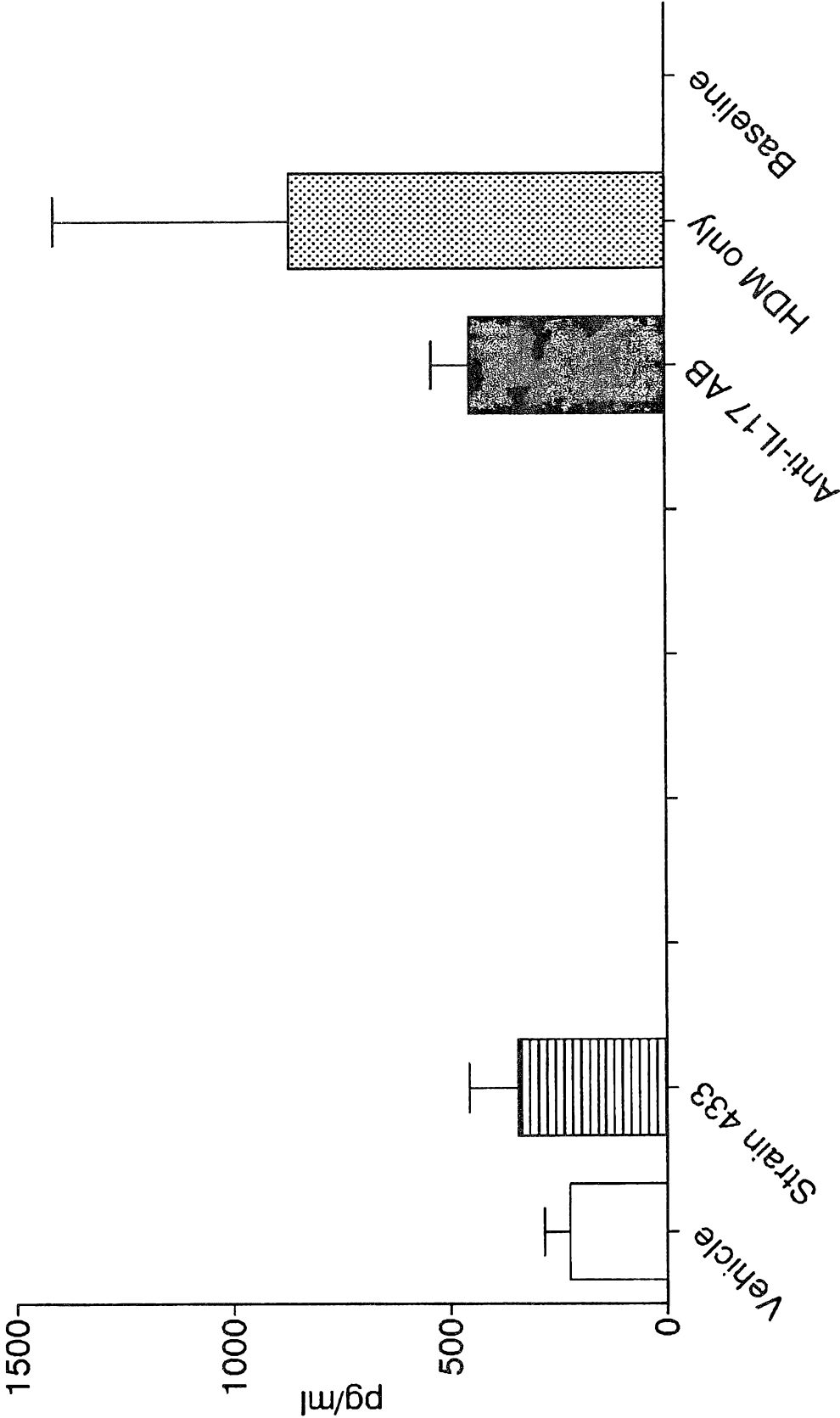
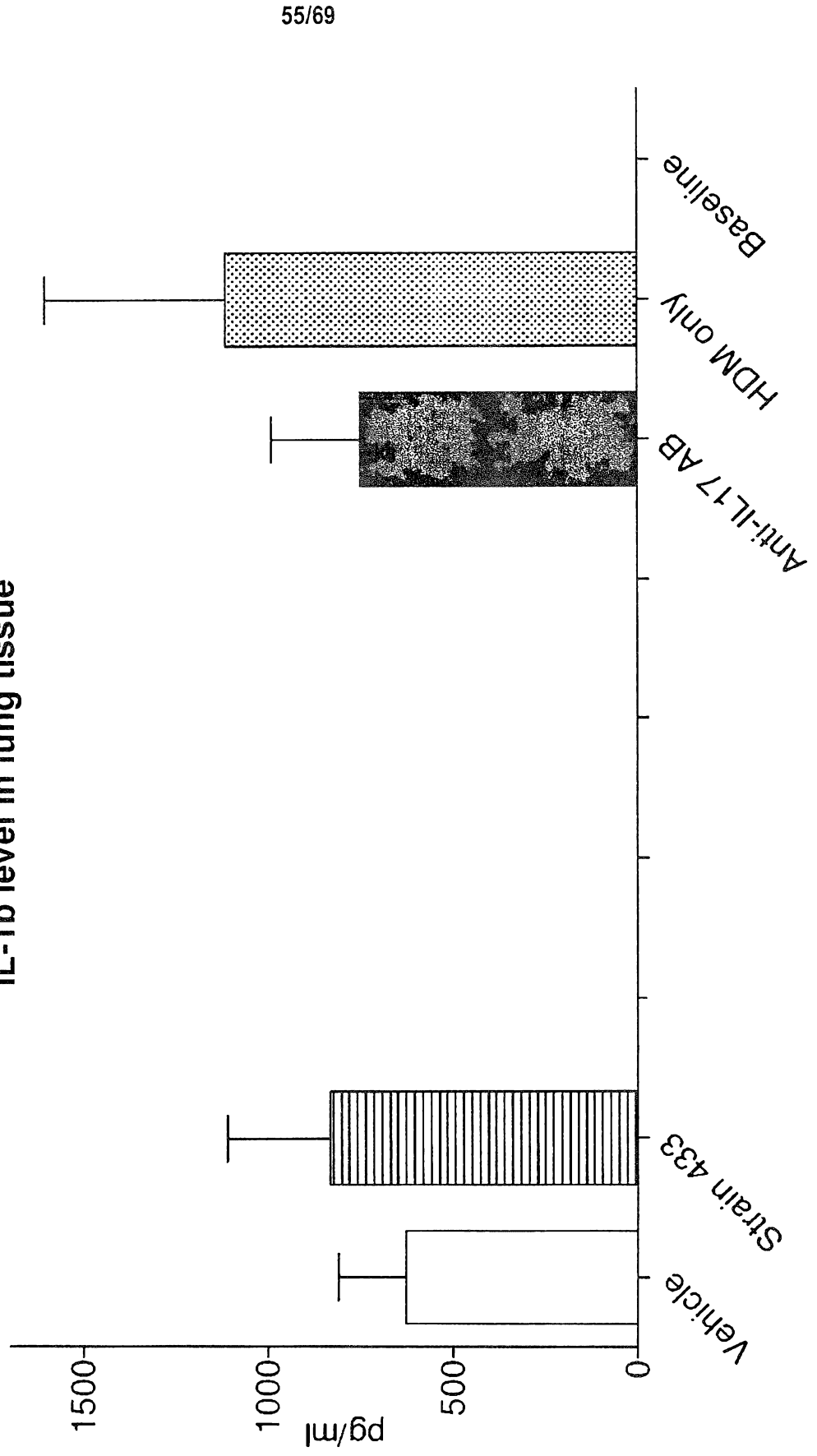
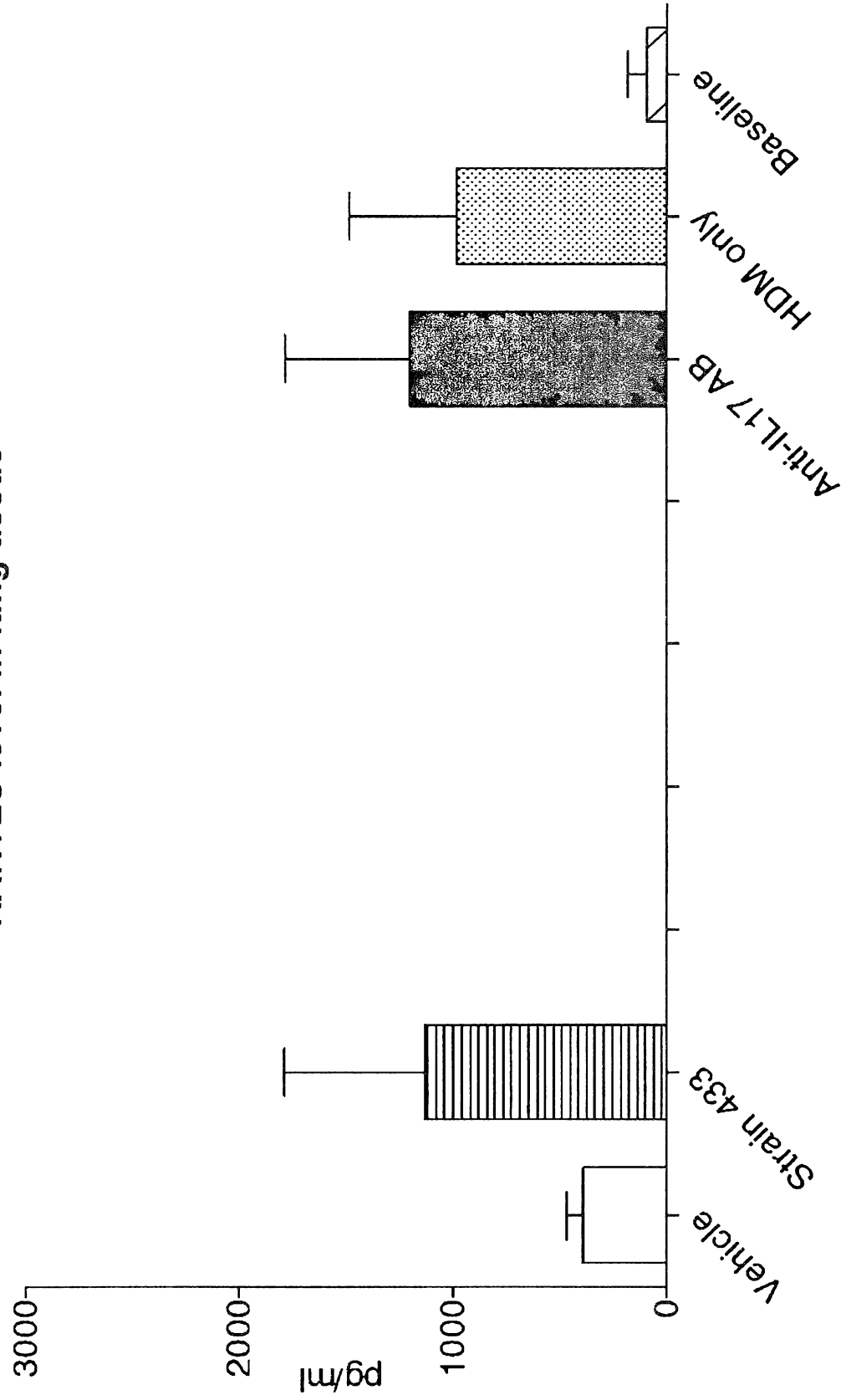


FIG. 57
IL-1b level in lung tissue



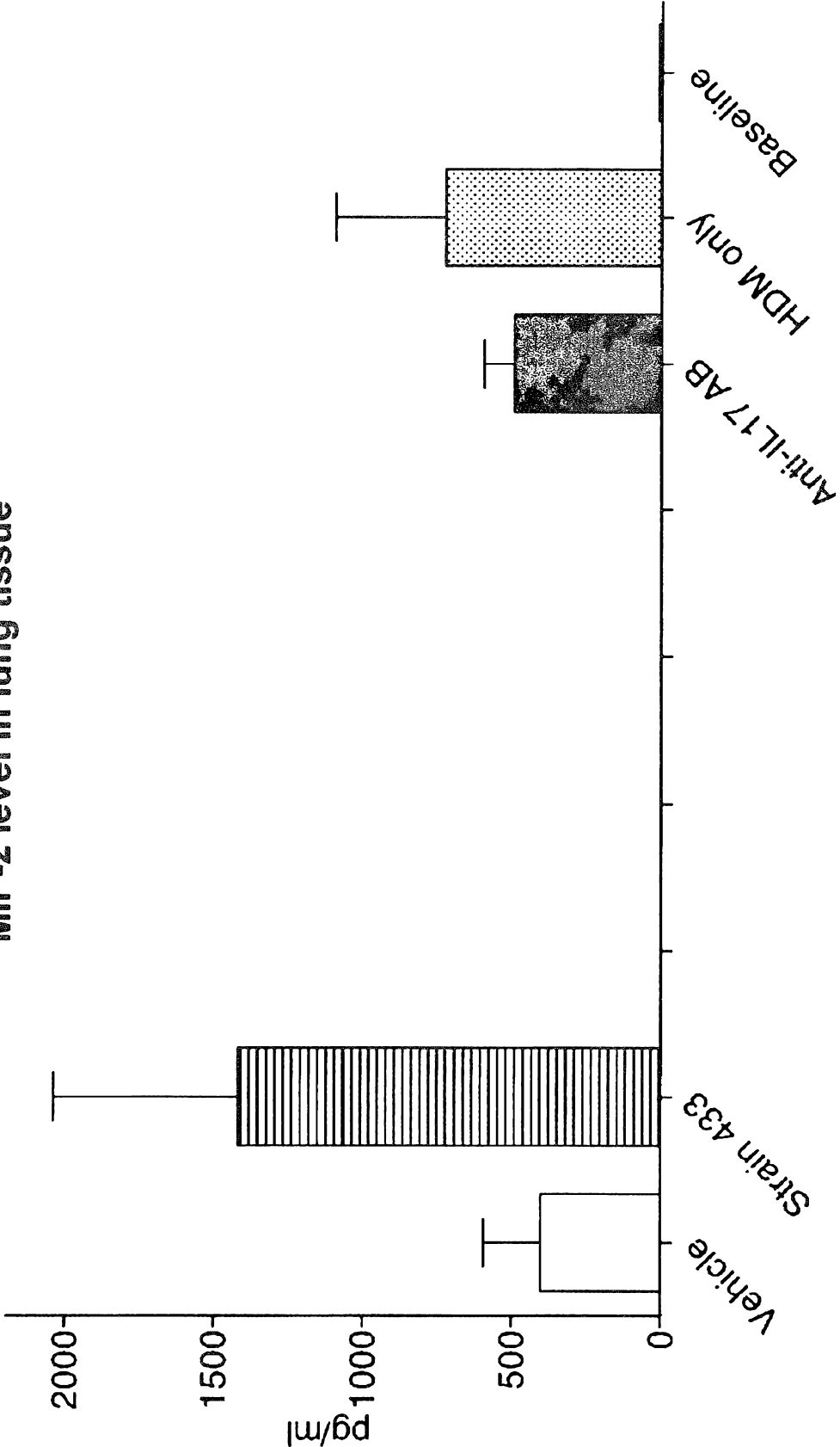
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FIG. 58
RANTES level in lung tissue



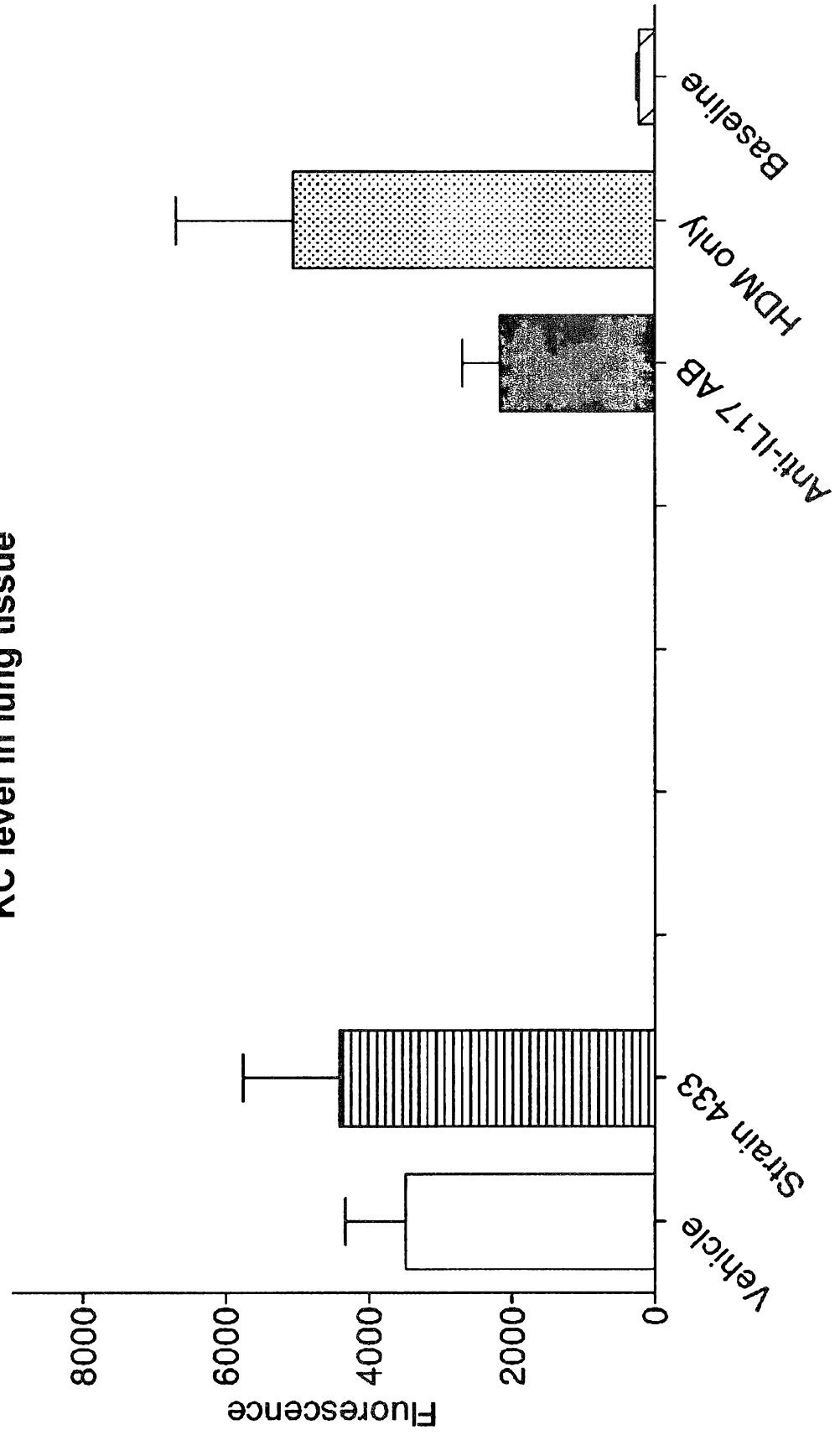
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FIG. 59
MIP-2 level in lung tissue



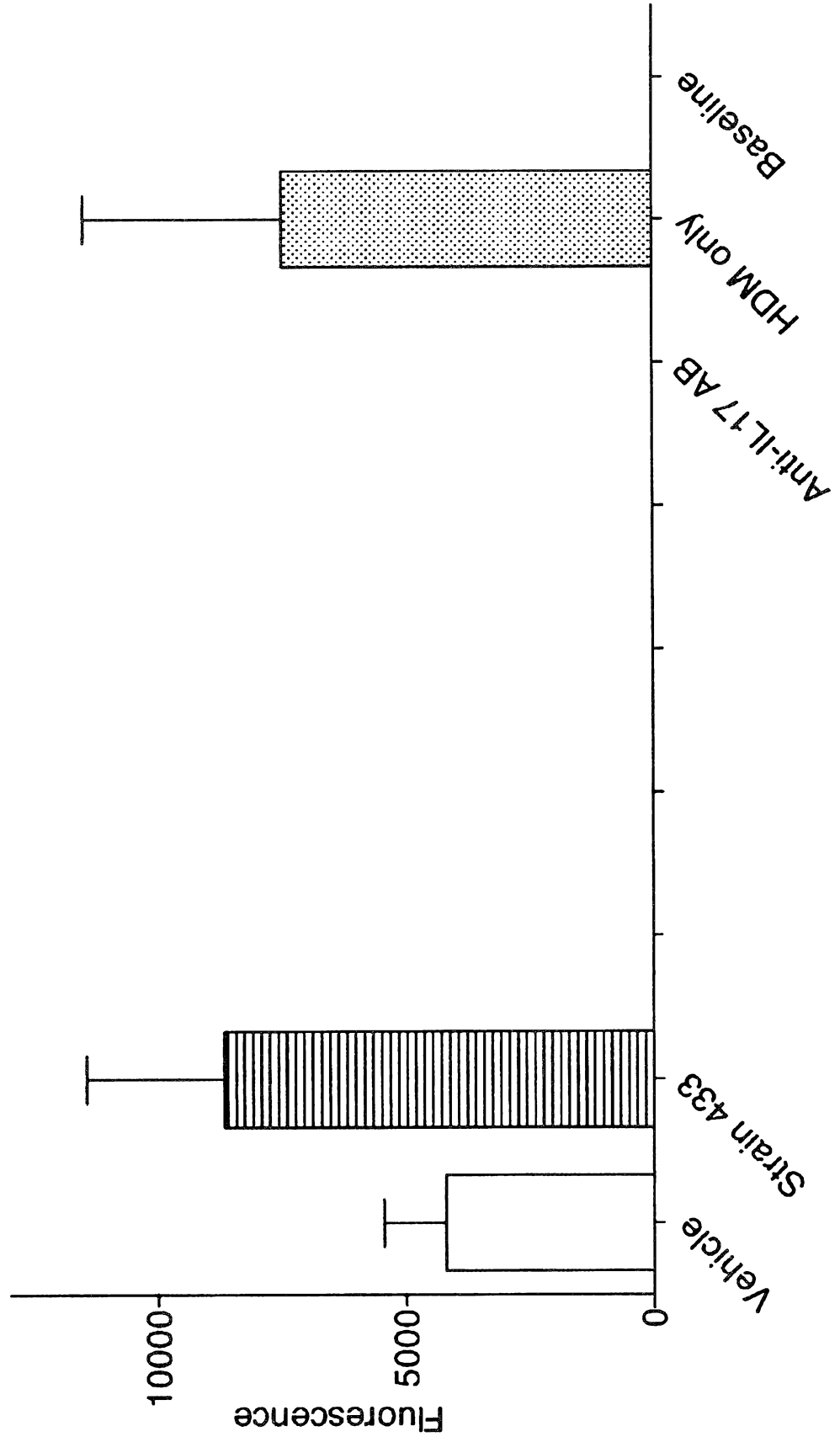
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FIG. 60
KC level in lung tissue



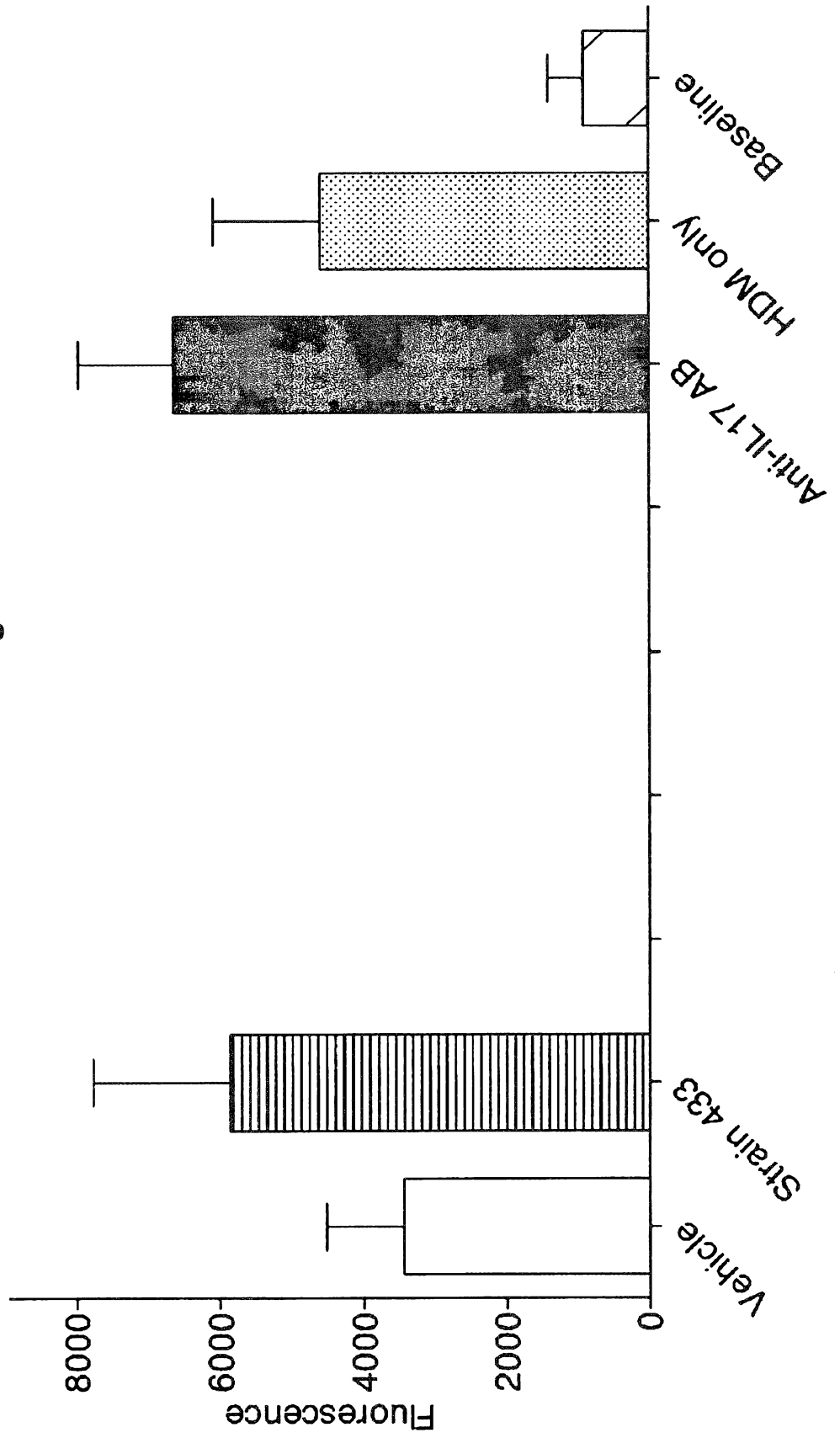
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FIG. 61
IL-17A level in lung tissue



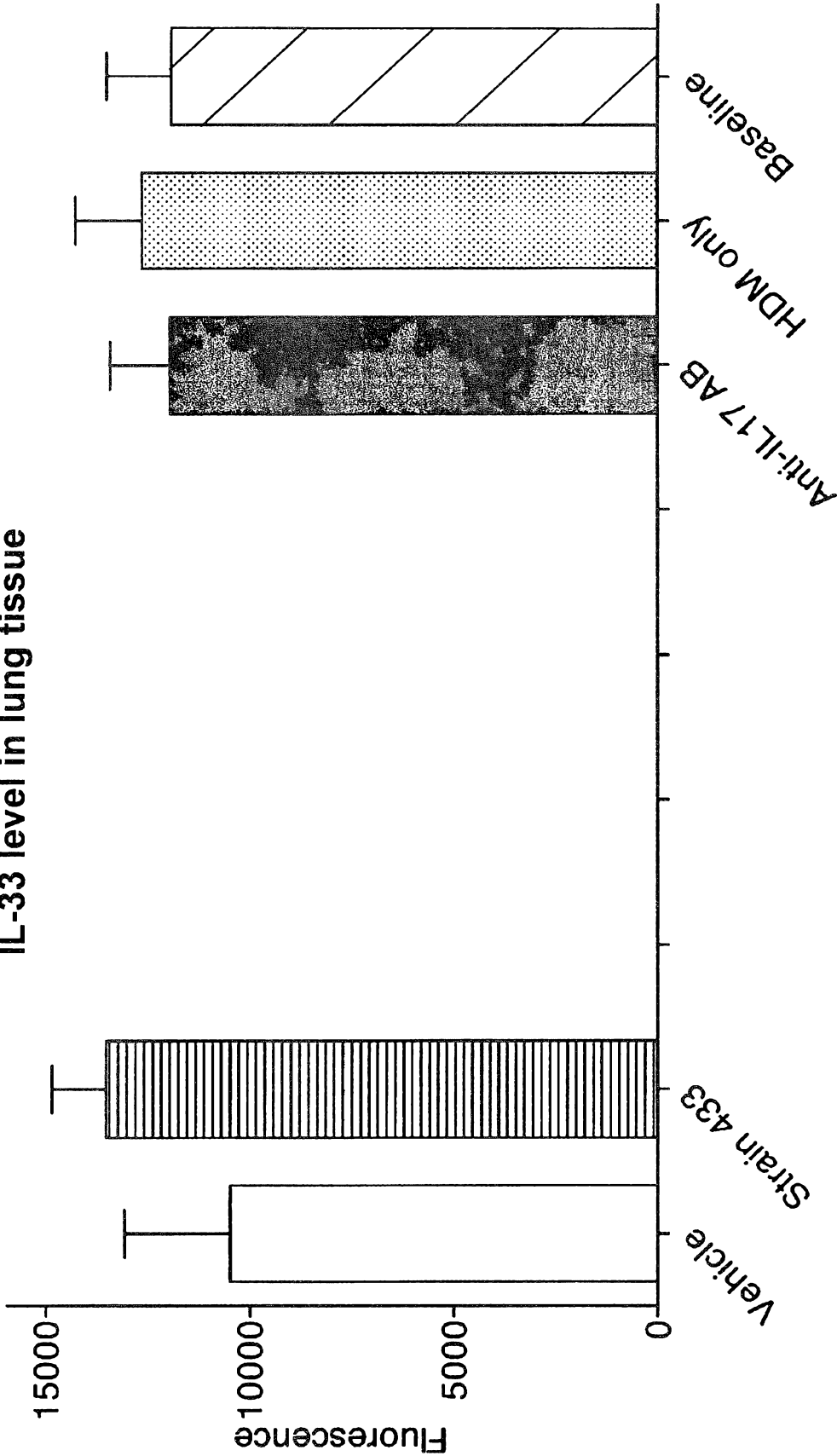
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FIG. 62
MIP-1a level in lung tissue



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FIG. 63
IL-33 level in lung tissue

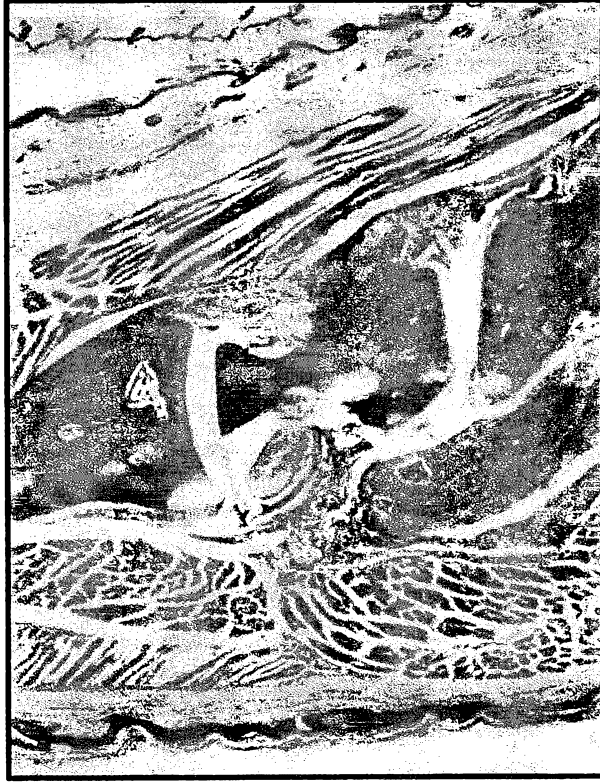


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FIG. 64
Visual Template for Histopathology Scoring



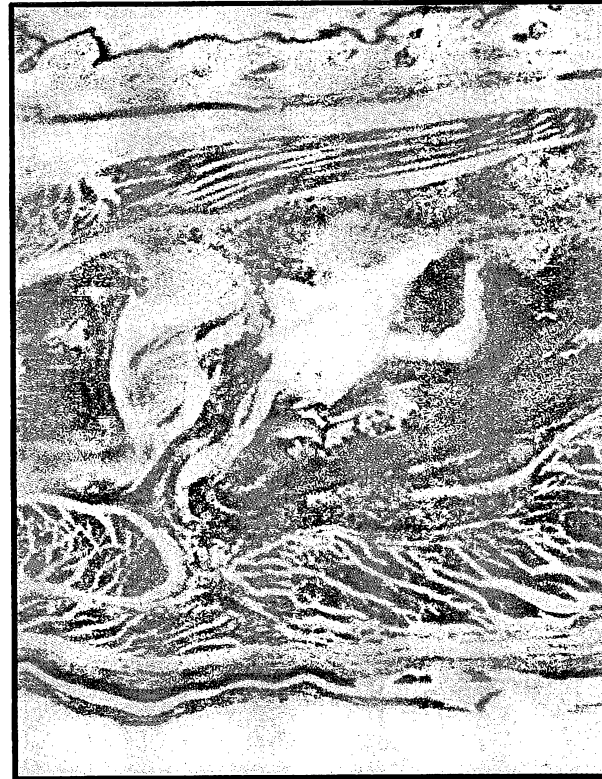
Grade 0



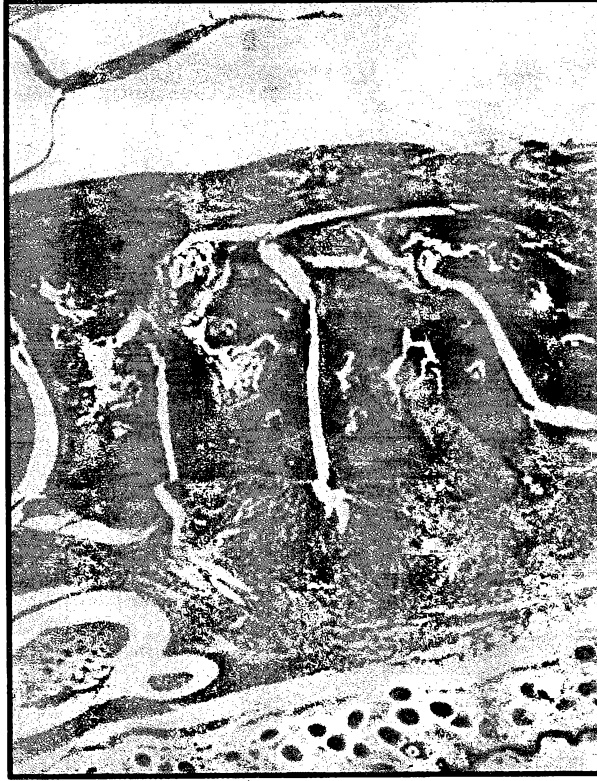
Grade 1

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FIG. 64(contd.)



Grade 4



Grade 7

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Grade 9

FIG. 64(contd.)

FIG. 65
Histopathology: Inflammation Scores

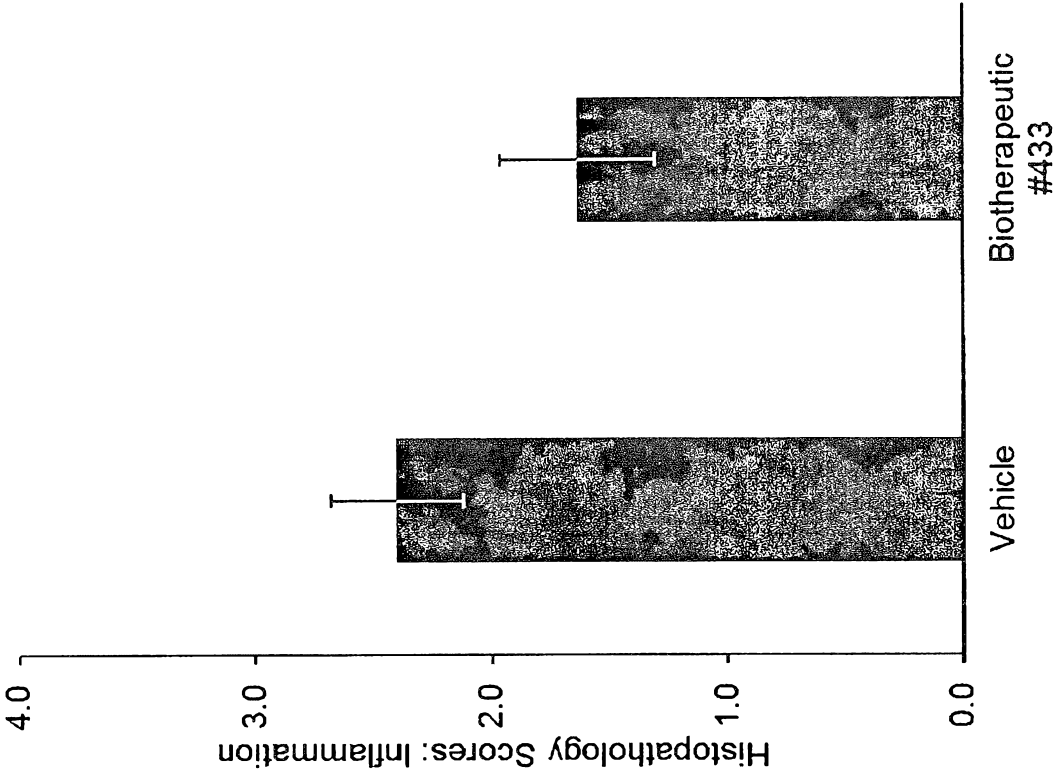
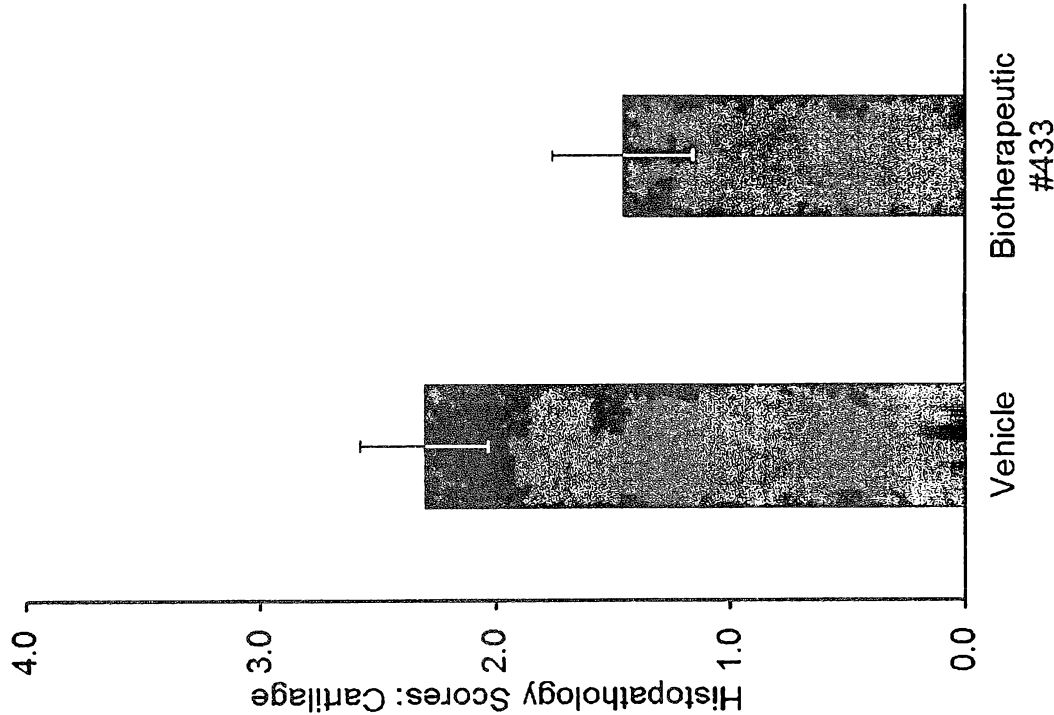
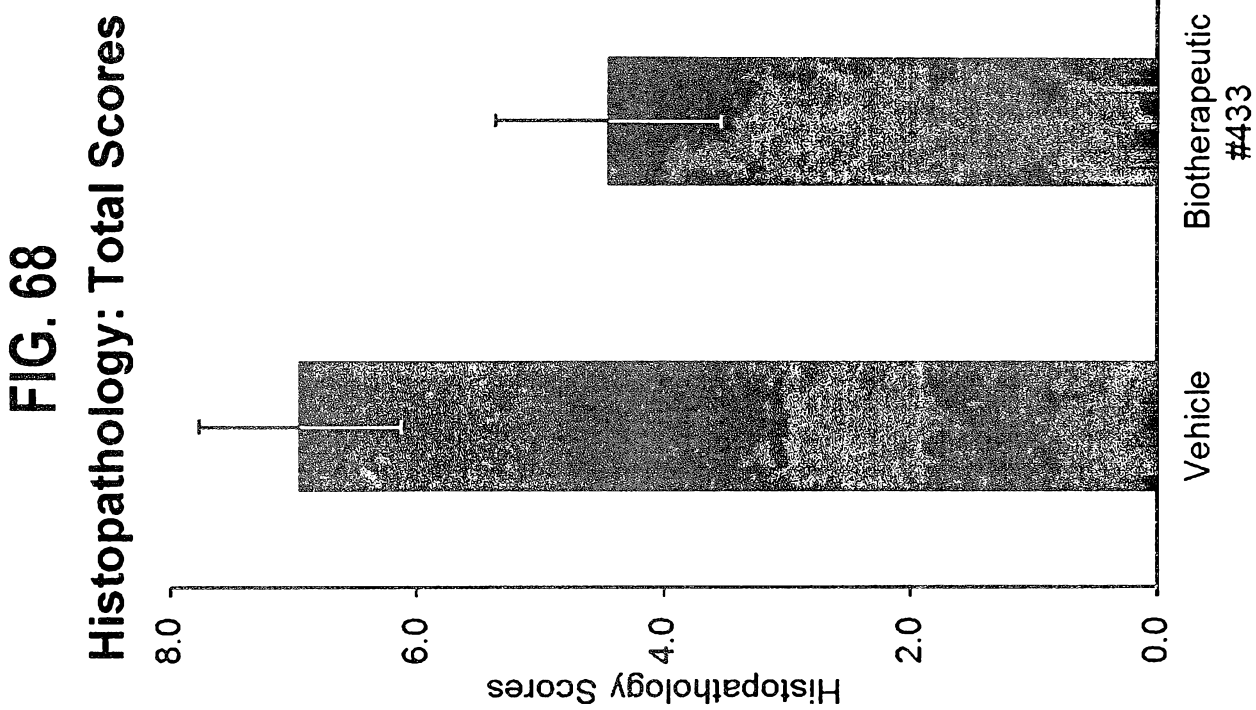
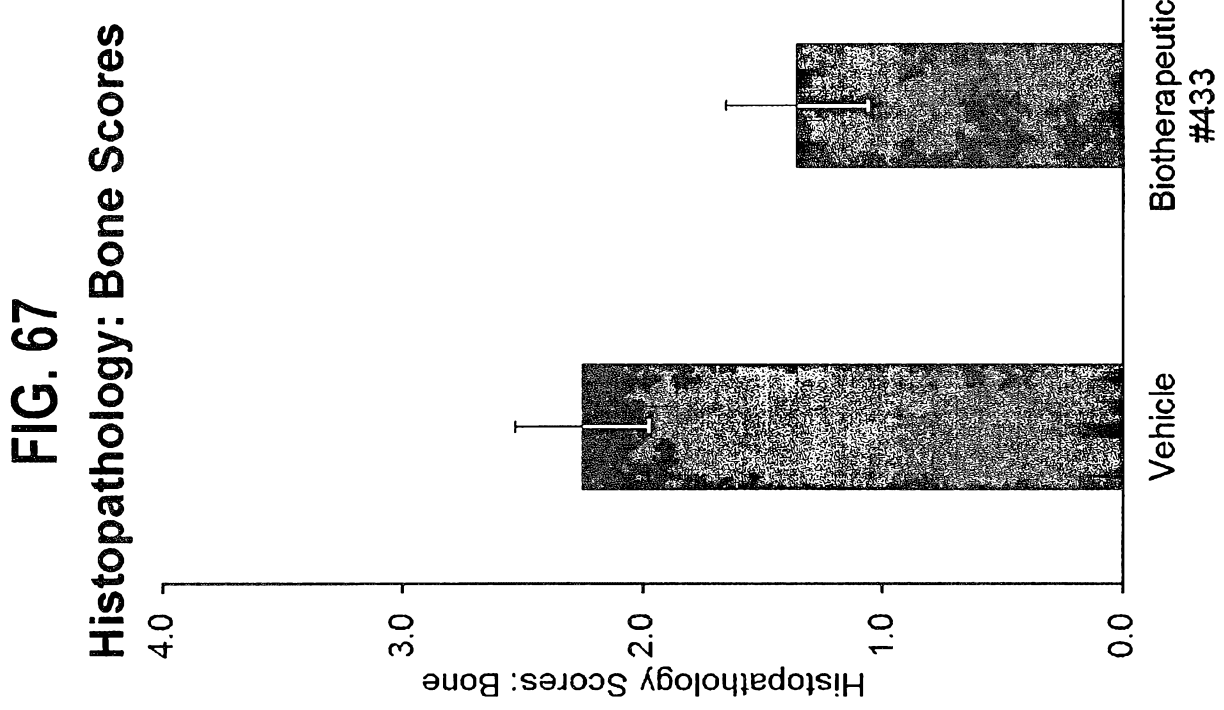


FIG. 66
Histopathology: Cartilage Scores





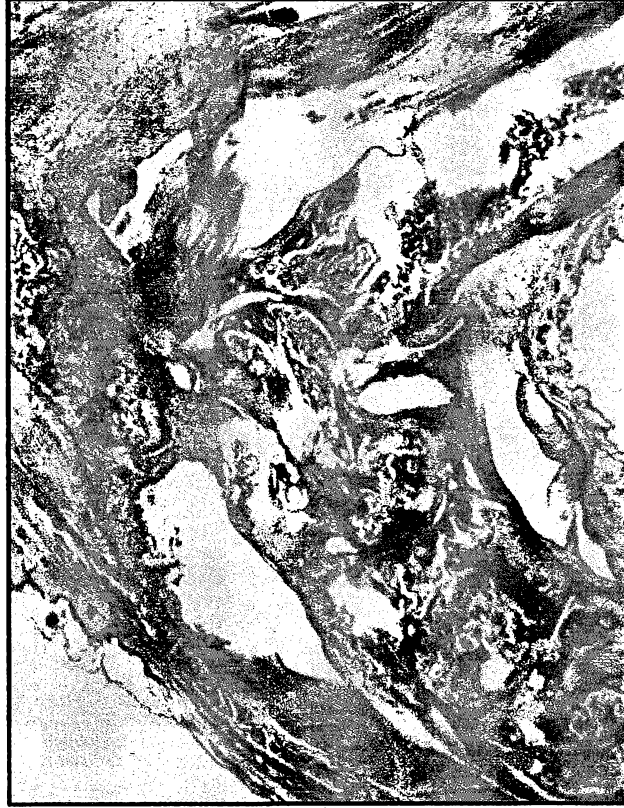
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FIG. 69

Vehicle (#1.1 1R, Grade 9)



Strain #433 (#6.1R, Grade 9)

FIG. 70

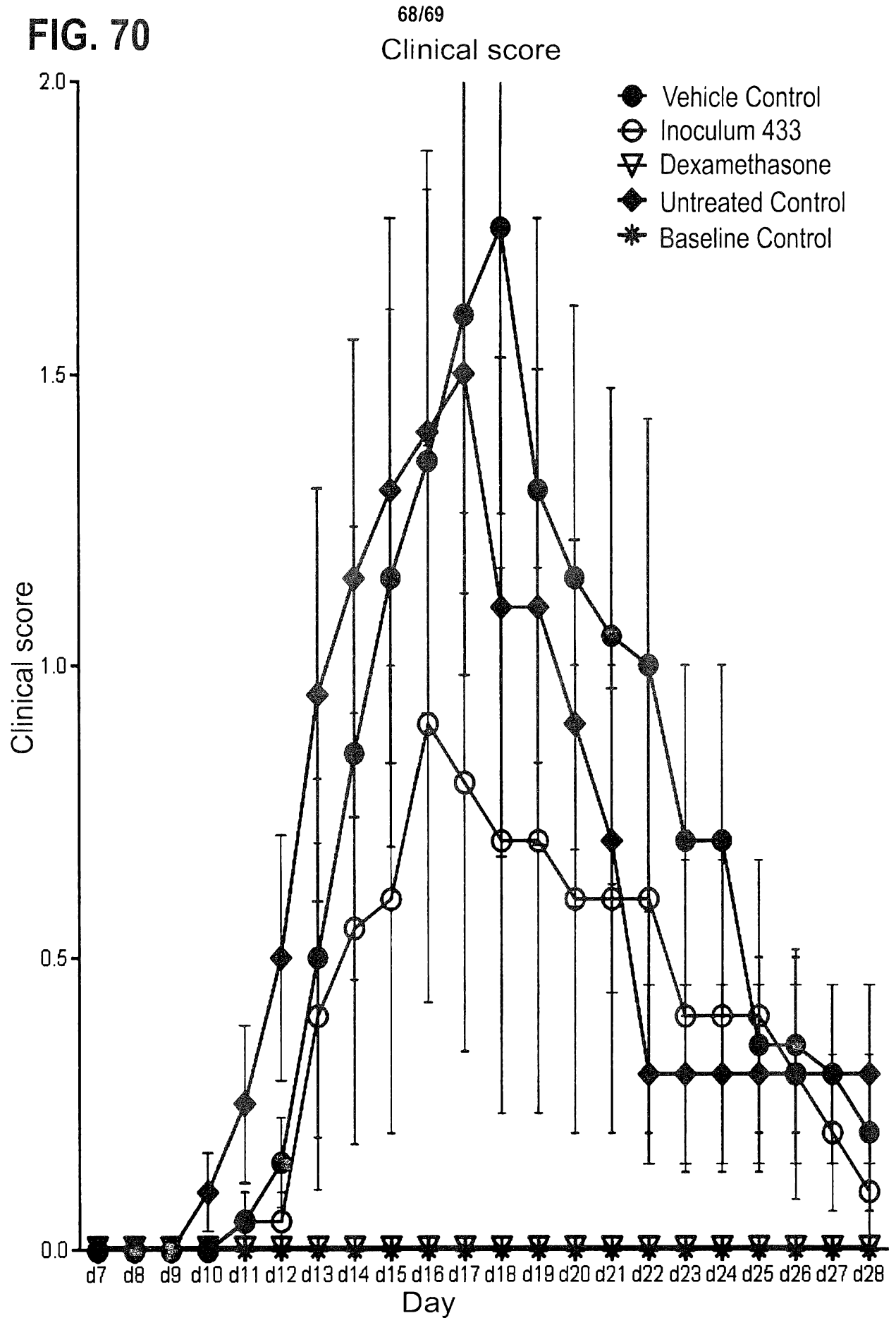
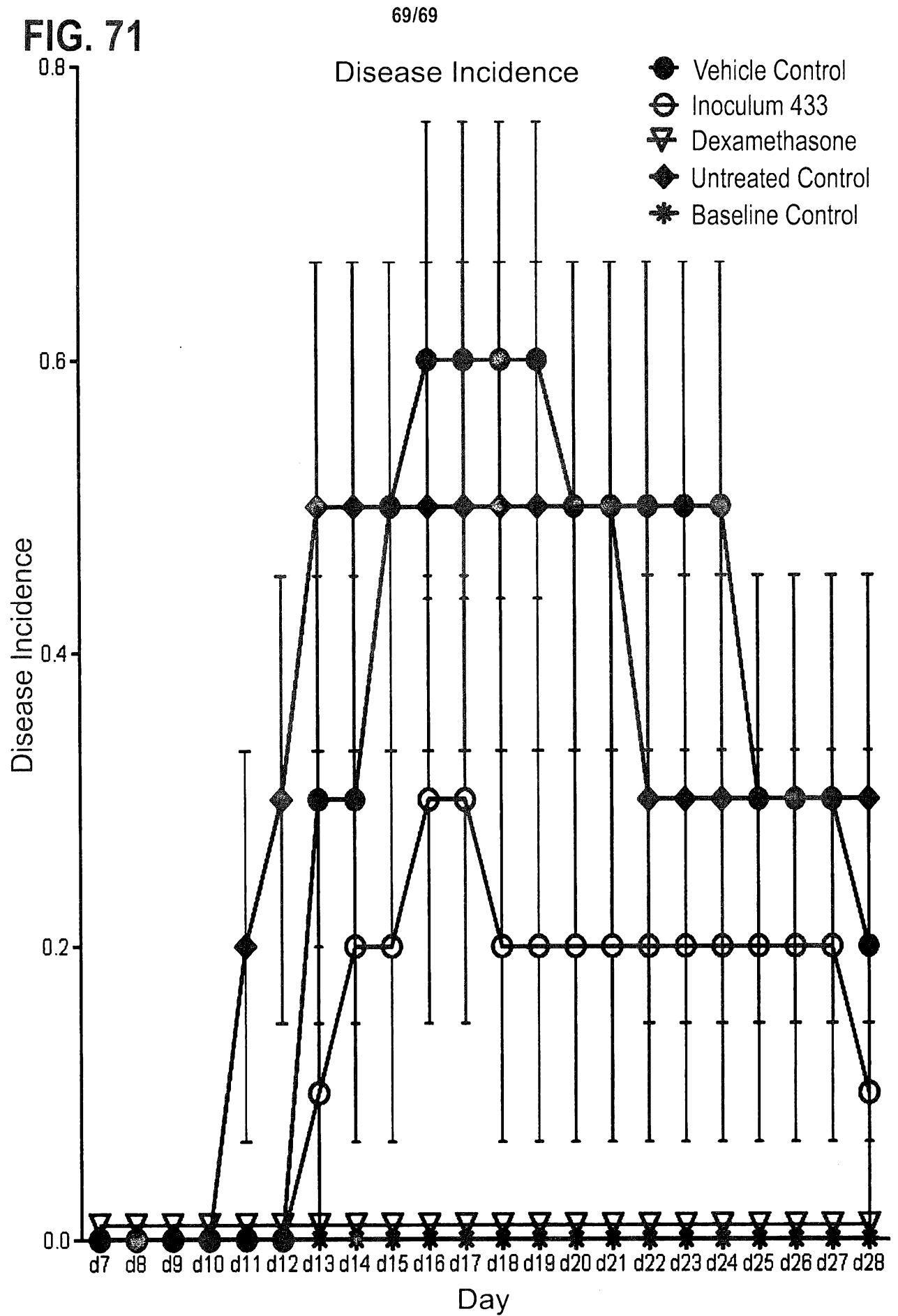


FIG. 71



p065780wo sequence listing.txt

SEQUENCE LISTING

<110> 4D PHARMA RESEARCH LIMITED

<120> COMPOSITIONS COMPRISING BACTERIAL STRAINS

<130> P065780wo

<150> GB 1510468.0

<151> 2015-06-15

<150> GB 1606807.4

<151> 2016-04-19

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c	c	c	g	c	g	t	c	t	g					300
a	c	c	t	g	a	g	a	g						360
a	g	c	a	g	t	g	g	g	t					420
g	a	a	g	t	a	t	t	t	c					480
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c	t	g	c	c	a	g	g	a						1200
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t	g	a	t	t	a	c	c	a						300
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g	g	a	a	t	a	t	t	g						420
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g	g	g	t	g	t	a	a	a						600
c	g	g	t	a	c	t	g	c						660
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a	t	t	a	c	t	g	a	c						780

p065780wo sequence listing.txt

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 <211> 1500
 <212> DNA
 <213> Roseburia hominis

<400> 3

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