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**CROUCH, N.P. ET AL.: "A Mechanistic Rationalisation for the Substrate Specificity of Recombinant Mammalian 4-Hydroxyphenylpyruvate Dioxygenase (4-HPPD)", TETRAHEDRON, vol. 53, no. 20, 19 May 1997 (1997-05-19), pages 6993-7010, XP004105682,**  
**LEE, I.Y. ET AL.: "Conversion of beta-Methylbutyric Acid to beta-Hydroxy-beta-Methylbutyric Acid by Galactomyces reessii", APPLIED AND ENVIRONMENTAL MICROBIOLOGY, vol. 63, no. 11, November 1997 (1997-11), pages 4191-4195, XP002569951, cited in the application**  
**MIDDLETON, B.: "The Kinetic Mechanism of 3-Hydroxy-3-methylglutaryl-Coenzyme A Synthase from Baker's Yeast", BIOCHEMICAL JOURNAL, vol. 126, no. 1, 1972, pages 35-47, XP002569952,**



## DESCRIPTION

**[0001]** The present invention relates to a method for the production of 3-hydroxy-3-methylbutyric acid (also referred to as beta-hydroxyisovalerate or HIV) from acetone and acetyl-coenzyme A comprising the enzymatic conversion of acetone and acetyl-coenzyme A into 3-hydroxy-3-methylbutyric acid. The conversion makes use of an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the methyl group of acetyl-coenzyme A. The enzyme employed in the process is an enzyme with the activity of a HMG CoA synthase (EC 2.3.3.10). The present invention also relates to organisms able to produce 3-hydroxy-3-methylbutyric acid from acetone and acetyl-coenzyme A and to the use of the above-mentioned enzyme and organisms for the production of 3-hydroxy-3-methylbutyric acid. Finally, the present invention relates to the use of acetone for the production of 3-hydroxy-3-methylbutyric acid.

**[0002]** 3-hydroxy-3-methylbutyric acid (also referred to as beta-hydroxyisovalerate or HIV; see Figure 1) is a metabolite of the essential amino acid leucine and is synthesized in the human body. It can be found in small quantities in grapefruit, alfalfa and catfish. It is also known to occur in some metabolic disorders of leucine catabolism, i.e. hypovaleric acidemia. It has been shown that 3-hydroxy-3-methylbutyric acid may have an effect on increasing muscle weight and strength (Nissen et al., J. Appl. Physiol. 81 (1996), 2095-2104). Wilson et al. (Nutrition & Metabolism 5 (2008)) proposes as the mechanisms of action the following:

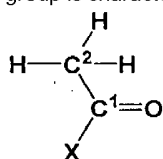
- increased sarcolemmal integrity via conversion by HMG CoA reductase
- enhanced protein synthesis via the mTOR pathway
- depression of protein degradation through inhibition of the ubiquitin pathway. 3-hydroxy-3-methylbutyric acid is supposed to help muscles combat protein breakdown, assist in muscle repair and support increased endurance. It has been described to help patients with chronic obstructive pulmonary disease in hospital intensive care units, muscle wasting associated with HIV and cancer and trauma victims with severe injuries. Thus, it is of commercial interest because of its use as a muscle enhancer for bodybuilding and as a medicament for avoiding muscle wasting. United States Patent 7026507 describes a process for preparing solid formulations of sodium 3-hydroxy-3-methylbutyrate in which, in a first process step, 4,4-dimethyloxetan-2-one is reacted with aqueous sodium hydroxide to form a solution of sodium 3-hydroxy-3-methylbutyrate, and then, if appropriate after concentration, the solution is applied, in a further process step, to synthetic silica, and in which the resultant product is, if appropriate, dried.

**[0003]** It would be desirable to provide a process for the production of 3-hydroxy-3-methylbutyrate which would be independent of inorganic production steps and which could be effected in living organisms thereby being environmentally sound and inexpensive. In this context, Lee et al. (Appl. Environ. Microbiol. 63 (1997), 4191-4195) describes a method for the production of 3-hydroxy-3-methylbutyrate by converting 3-methylbutyric acid to 3-hydroxy-3-methylbutyric acid using the microorganism *Gatactomyces reessii*. Further, Crouch et al. (Tetrahedron 53 (1997), 6993-7010) describes a method for the production of 3-hydroxy-3-methylbutyrate from 2-oxo-4-methylpentanoate using the enzyme alpha-ketoisocaproate dioxygenase. However, although these processes allowed the production of 3-hydroxy-3-methylbutyrate there is still a need to provide alternative efficient and cost effective ways of producing 3-hydroxy-3-methylbutyrate in particular by biological processes.

**[0004]** The present invention meets this demand for an alternative process for the production of 3-hydroxy-3-methylbutyrate and provides a method which is based on biological resources and allows to produce 3-hydroxy-3-methylbutyrate in vitro or in vivo in a microorganism and other species.

**[0005]** In particular, the present invention relates to a method for the production of 3-hydroxy-3-methylbutyric acid (also referred to as beta-hydroxyisovalerate or HIV) from acetone and acetyl-coenzyme A comprising the enzymatic conversion of acetone and acetyl coenzyme A into 3-hydroxy-3-methylbutyric acid.

**[0006]** Acetone is represented by the following formula:  $\text{CH}_3\text{-(C=O)-CH}_3$ . The compound which provides an activated acetyl group is characterized by the following formula (I):



,wherein X is S-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH(OH)-C(CH<sub>3</sub>)<sub>2</sub>-CH<sub>2</sub>-O-P<sub>0</sub>2H-O-P<sub>0</sub>2H-C<sub>10</sub>H<sub>13</sub>N<sub>5</sub>O<sub>7</sub>P (coenzyme A).

**[0007]** The conversion makes use of an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group according to formula (I). According to this reaction scheme the oxo group of acetone reacts as an electrophile and the methyl group of the compound which provides an activated acetyl group according to formula (I) reacts as a nucleophile. The reaction of the conversion of acetone and a compound which provides an activated acetyl group according to formula (I) is shown in Figure 5.

**[0008]** The reaction can occur in one step, i.e. 3-hydroxy-3-methylbutyrate can be the direct product of a reaction catalyzed by the above described enzyme. Alternatively, the reaction may comprise two steps, when acetyl CoA is used as the compound which provides an activated acetyl group, in the sense that first an adduct of 3-hydroxy-3-methylbutyrate and the compound which provides an activated acetyl group is produced, e.g. 3-hydroxy-3-methylbutyryl-CoA, which is subsequently hydrolyzed, e.g. to 3-hydroxy-3-methylbutyrate and CoA. Thus, in the first alternative the enzyme catalyzes the complete reaction as shown in Figure 5. In the second alternative, the enzyme catalyzes the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group according to formula (I) but X stays in the molecule. X is then removed subsequently from the molecule by hydrolysis.

**[0009]** The present invention shows for the first time that it is possible to produce 3-hydroxy-3-methylbutyrate by making use of an enzyme which can transfer an activated acetyl group to acetone. In the prior art production of 3-hydroxy-3-methylbutyrate from isovaleric acid through bioconversion using the fungus *Galactomyces reessii* has been reported. However, considering that isovaleric acid is obtained from leucine through decarboxylation and that leucine itself derives in metabolism from the overall condensation of two molecules of pyruvate and one molecule of acetyl CoA, this production process is energetically unfavorable. The process of the present invention avoids this disadvantage. The compound which provides an activated acetyl group is acetyl CoA. Acetyl CoA (also known as acetyl Coenzyme A) in chemical structure is the thioester between coenzyme A (a thiol) and acetic acid.

**[0010]** The enzyme employed in the process is an enzyme with the activity of a HMG CoA synthase (EC 2.3.3.10).

**[0011]** The method according to the present invention comprises the enzymatic conversion of acetone and acetyl CoA into 3-hydroxy-3-methylbutyrate with an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom C<sup>2</sup> of acetyl CoA according to formula (I). The enzyme employed in the process according to the invention is an enzyme which has the activity of a HMG CoA synthase (EC 2.3.3.10).

**[0012]** In particular, it has been shown in the context of the present invention that HMG CoA synthase can accept acetone instead of its normal substrate acetoacetyl-CoA thereby allowing the conversion of acetyl-CoA (or a compound according to formula (I)) and acetone into 3-hydroxy-3-methylbutyrate.

**[0013]** In the context of the present application the term "HMG CoA synthase" or "a protein/enzyme having the activity of a HMG CoA synthase" refers to any enzyme which is classified in the EC number EC 2.3.3.10 (formerly, HMG-CoA synthase has been classified as EC 4.1.3.5 but has been transferred to EC 2.3.3.10), in particular it refers to any enzyme which is able to catalyze the reaction where acetyl-CoA condenses with acetoacetyl-CoA to form 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) (see Figure 2) and the term also refers to any enzyme which is derived from such a HMG CoA synthase and which is capable of catalyzing the conversion of acetone and a compound which provides an activated acetyl group as defined above, preferably acetyl CoA, into 3-hydroxy-3-methylbutyrate.

**[0014]** The enzymatic activity of condensing acetyl-CoA with acetoacetyl-CoA to form 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) can be measured by methods well known in the art. One possible and preferably used assay is described, e.g., in Clinkenbeard et al. (J. Biol. Chem. 250 (1975), 3108-3116). In this assay HMG-CoA synthase activity is measured by monitoring the decrease in absorbance at 303 nm that accompanies the acetyl-CoA-dependent disappearance of the enolate form of acetoacetyl-CoA. Preferably HMG CoA synthase activity is assayed as described in Example 3.

**[0015]** HMG CoA synthase is part of the mevalonate pathway. Two pathways have been identified for the synthesis of isopentenyl pyrophosphate (IPP), i.e. the mevalonate pathway and the glyceraldehyde 3-phosphate-pyruvate pathway. HMG CoA synthase catalyzes the biological Claisen condensation of acetyl-CoA with acetoacetyl-CoA and is a member of a superfamily of acyl-condensing enzymes that includes beta-ketothiolases, fatty acid synthases (beta-ketoacyl carrier protein synthase) and polyketide synthases.

[0016] HMG CoA synthase has been described for various organisms. Also amino acid and nucleic-acid sequences encoding HMG CoA synthases from numerous sources are available. Generally, the sequences only share a low degree of overall sequence identity. For example, the enzymes from *Staphylococcus* or *Streptococcus* show only about 20% identity to those of human and avian HMG CoA synthase. In some sources it is reported that the bacterial HMG CoA synthases and their animal counterparts exhibit only about 10% overall sequence identity (Sutherlin et al, *J. Bacteriol.* 184 (2002), 4065-4070). However, the amino acid residues involved in the acetylation and condensation reactions are conserved among bacterial and eukaryotic HMG CoA synthases (Campobasso et al., *J. Biol. Chem.* 279 (2004), 44883-44888). The three-dimensional structure of three HMG CoA synthase enzymes has been determined and the amino acids crucial for the enzymatic reaction are in principle well characterized (Campobasso et al., loc. cit.; Chun et al., *J. Biol.Chem.* 275 (2000), 17946-17953; Nagegowda et al., *Biochem. J.* 383 (2004), 517-527; Hegardt, *Biochem. J.* 338 (1999), 569-582). In eukaryotes there exist two forms of the HMG CoA synthase, i.e. a cytosolic and a mitochondrial form. The cytosolic form plays a key role in the production of cholesterol and other isoprenoids and the mitochondrial form is involved in the production of ketone bodies.

[0017] In principle any HMG CoA synthase enzyme can be used in the context of the present invention, in particular from prokaryotic or eukaryotic organisms.

[0018] Prokaryotic HMG CoA synthases are described, e.g., from *Staphylococcus aureus* (Campobasso et al., loc. cit.; Uniprot accession number Q9FD87), *Staphylococcus epidermidis* (Uniprot accession number Q9FD76), *Staphylococcus haemolyticus* (Uniprot accession number Q9FD82), *Enterococcus faecalis* (Sutherlin et al., loc. cit.; Uniprot accession number Q9FD7), *Enterococcus faecium* (Uniprot accession number Q9FD66), *Streptococcus pneumoniae* (Uniprot accession number Q9FD56), *Streptococcus pyogenes* (Uniprot accession number Q9FD61) and *Methanobacterium thermoautotrophicum* (accession number AE000857), *Borrelia burgdorferi* (NCBI accession number BB0683).

[0019] Moreover, the following Table A lists some known HMG CoA synthases from prokaryotes:

Table A:

Swissprot/TrEmbl Accession number	Organisme
Q9YAS0	<i>Aeropyrum pernix</i>
A7Z4Y2	<i>Bacillus amyloliquefaciens</i>
P40830 2874037340	<i>Bacillus subtilis</i>
B8G795	<i>Chloroflexus aggregans</i>
A5EUV4	<i>Dichelobacter nodosus</i>
A5FM54	<i>Flavobacterium johnsoniae</i>
Q18GC4	<i>Haloquadratum walsbyi</i>
B9LS15	<i>Halorubrum lacusprofundi</i>
A9B8F0	<i>Herpetosiphon aurantiacus</i>
A2BMY8	<i>Hyperthermus butylicus</i>
Q5FLB7	<i>Lactobacillus acidophilus</i>
Q03QR0	<i>Lactobacillus brevis</i>
Q1GAH5	<i>Lactobacillus delbrueckii</i>
B2GBL1	<i>Lactobacillus fermentum</i>
B1MZ51	<i>Leuconostoc citreum</i>
Q03WZ0	<i>Leuconostoc mesenteroides</i>
A4YH99	<i>Metallosphaera sedula</i>
A5UNI8	<i>Methanobrevibacter smithii</i>
Q58941	<i>Methanocaldococcus jannaschii</i>
Q12UR3	<i>Methanococcoides burtonii</i>
A6USZ1	<i>Methanococcus aeolicus</i>
A4FWW6	<i>Methanococcus maripaludis</i>
A6UPL1	<i>Methanosarcina mazei</i>
A2STY2	<i>Methanocorpusculum labreanum</i>

Swissprot/TrEmbl Accession number	Organisme
Q8TVL0	Methanopyrus_andleri
Q8PYJ0	Methanosarcina mazei
Q2NHU7	Methanosphaera stadtmanae
Q2FPH4	Methanospirillum hungatei
B2HGT6	Mycobacterium marinum
Q3IMZ7	Natronomonas pharaonis
Q8EP69	Oceanobacillus iheyensis
Q04F95	Oenococcus oeni
Q03FU5	Pediococcus pentosaceus
Q6L233	Picrophilus torridus
A6G7N7	Plesiocystis pacifica
A4WJ12	Pyrobaculum arsenaticum
A7NHZ7	Roseiflexus castenholzii
Q8CN06	Staphylococcus epidermidis
Q4L958	Staphylococcus haemolyticus
Q4A0D6	Staphylococcus saprophyticus
B4U364	Streptococcus equi
Q8DUJ5	Streptococcus mutans
Q4J933	Sulfolobus acidocaldarius
Q971K8	Sulfolobus tokodaii
Q9HI87	Thermoplasma acidophilum
Q31EW2	Thiomicrospira crunogena
Q51798	Pyrococcus furiosus
A5VJB7	Lactobacillus reuteri
Q7CF79	Streptococcus pyogenes
Q9UWU0	Sulfolobus solfataricus

**[0020]** Eukaryotic HMG CoA synthases are described, e.g., from fungi, such as *Schizosaccharomyces pombe* (accession numbers U32187 and P54874), *Saccharomyces cerevisiae* (accession number P54839), plants, such as *Arabidopsis thaliana* (accession numbers X83882 and P54873), *Pinus sylvestris* (accession number X96386) and animals, such as *Caenorhabditis elegans* (accession number P54871), *Mus musculus* (mitochondrial; accession number P54869 and Hegardt, *Biochem. J.* 338 (1999), 569-582), *Rattus norvegicus* (mitochondrial; accession number P22791 and Hegardt, *Biochem. J.* 338 (1999); cytosolic; accession number P17425), 569-582), Chinese hamster (*Cricetulus griseus*; accession number P13704), *Sus scrofa* (mitochondrial; accession number U90884 and Hegardt, *Biochem. J.* 338 (1999), 569-582), *Homo sapiens* (mitochondrial; accession number P54868 and Hegardt, *Biochem. J.* 338 (1999), 569-582; cytosolic; accession number Q01581), *Blattella germanica* (cytosolic form 1; accession number P54961), *Blattella germanica* (cytosolic form 2; accession number P54870) and *Gallus gallus* (cytosolic; accession number P23228).

**[0021]** Examples of HMG CoA synthases from different organisms are given in SEQ ID NO: 1 to 14. SEQ ID NO: 1 shows the sequence of the cytoplasmic HMG CoA synthase of *Caenorhabditis elegans* (P54871, gene bank F25B4.6), SEQ ID NO: 2 shows the sequence of the cytoplasmic HMG CoA synthase of *Schizosaccharomyces pombe* (fission yeast; P54874), SEQ ID NO: 3 shows the sequence of the cytoplasmic HMG CoA synthase of *Saccharomyces cerevisiae* (baker's yeast; P54839, gene bank CAA65437.1), SEQ ID NO: 4 shows the sequence of the cytoplasmic HMG CoA synthase of *Arabidopsis thaliana* (Mouse-ear cress; P54873), SEQ ID NO: 5 shows the sequence of the cytoplasmic HMG CoA synthase of *Dictyostelium discoideum* (Slime mold; P54872, gene bank L2114), SEQ ID NO: 6 shows the sequence of the cytoplasmic HMG CoA synthase of *Blattella germanica* (German cockroach; P54961, gene bank X73679), SEQ ID NO: 7 shows the sequence of the cytoplasmic HMG CoA synthase of *Gallus gallus* (Chicken; P23228, gene bank CHKHMGOAS), SEQ ID NO: 8 shows the sequence of the cytoplasmic HMG CoA synthase of *Homo sapiens* (Human; Q01581, gene bank X66435), SEQ ID NO: 9 shows the sequence of the

mitochondrial HMG CoA synthase of Homo sapiens (Human; P54868, gene bank X83618), SEQ ID NO: 10 shows the sequence of the mitochondrial HMG CoA synthase of Dictyostelium discoideum (Slime mold; Q86HL5, gene bank XM\_638984), SEQ ID NO: 11 shows the sequence of the HMG CoA synthase of Staphylococcus epidermidis (Q9FD76), SEQ ID NO: 12 shows the sequence of the HMG CoA synthase of Lactobacillus fermentum (B2GBL1), SEQ ID, NO: 13 shows the sequence of the HMG CoA synthase of Hyperthermus butylicus (A2BMY8), SEQ ID NO: 14 shows the sequence of the HMG CoA synthase of Chloroflexus aggregans (B8G795), SEQ ID NO: 24 shows the sequence of the HMG CoA synthase of Lactobacillus delbrueckii (Q1GAH5) and SEQ ID NO: 25 shows the sequence of the HMG CoA synthase of Staphylococcus haemolyticus Q4L958 (198>V difference compared to wild type protein).

**[0022]** In a preferred embodiment of the present invention the HMG CoA synthase is an enzyme comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 1 to 14 or a sequence which is at least n % identical to any of SEQ ID NOs: 1 to 14 and having the activity of a HMG CoA synthase with n being an integer between 10 and 100, preferably 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 91, 92, 93, 94, 95, 96, 97, 98 or 99.

**[0023]** Preferably, the degree of identity is determined by comparing the respective sequence with the amino acid sequence of any one of the above-mentioned SEQ ID NOs. When the sequences which are compared do not have the same length, the degree of identity preferably either refers to the percentage of amino acid residues in the shorter sequence which are identical to amino acid residues in the longer sequence or to the percentage of amino acid residues in the longer sequence which are identical to amino acid residues in the shorter sequence. The degree of sequence identity can be determined according to methods well known in the art using preferably suitable computer algorithms such as CLUSTAL.

**[0024]** When using the Clustal analysis method to determine whether a particular sequence is, for instance, 80% identical to a reference sequence default settings may be used or the settings are preferably as follows: Matrix: blosum 30; Open gap penalty: 10.0; Extend gap penalty: 0.05; Delay divergent: 40; Gap separation distance: 8 for comparisons of amino acid sequences. For nucleotide sequence comparisons, the Extend gap penalty is preferably set to 5.0.

**[0025]** Preferably, the degree of identity is calculated over the complete length of the sequence.

**[0026]** The HMG CoA synthase employed in the process according to the invention can be a naturally occurring HMG CoA synthase or it can be a HMG CoA synthase which is derived from a naturally occurring HMG CoA synthase, e.g. by the introduction of mutations or other alterations which, e.g., alter or improve the enzymatic activity, the stability, etc.

**[0027]** The term "HMG CoA synthase" or "a protein/enzyme having the activity of a HMG CoA synthase" in the context of the present application also covers enzymes which are derived from a HMG CoA synthase, which are capable of producing 3-hydroxy-3-methylbutyrate by an enzymatic conversion of acetone and acetyl-coenzyme A, but which only have a low affinity to acetoacetyl-CoA as a substrate or do no longer accept acetoacetyl-CoA as a substrate. Such a modification of the preferred substrate of a HMG CoA synthase allows to improve the conversion of acetone into 3-hydroxy-3-methylbutyrate and to reduce the production of the by-product, e.g. HMG-CoA. Methods for modifying and/or improving the desired enzymatic activities of proteins are well-known to the person skilled in the art and include, e.g., random mutagenesis or site-directed mutagenesis and subsequent selection of enzymes having the desired properties or approaches of the so-called "directed evolution".

**[0028]** For example, for genetic engineering in prokaryotic cells, a nucleic acid molecule encoding HMG CoA synthase can be introduced into plasmids which permit mutagenesis or sequence modification by recombination of DNA sequences. Standard methods (see Sambrook and Russell (2001), Molecular Cloning: A Laboratory Manual, CSH Press, Cold Spring Harbor, NY, USA ) allow base exchanges to be performed or natural or synthetic sequences to be added. DNA fragments can be connected to each other by applying adapters and linkers to the fragments. Moreover, engineering measures which provide suitable restriction sites or remove surplus DNA or restriction sites can be used. In those cases, in which insertions, deletions or substitutions are possible, *in vitro* mutagenesis, "primer repair", restriction or ligation can be used. In general, a sequence analysis, restriction analysis and other methods of biochemistry and molecular biology are carried out as analysis methods. The resulting HMG CoA synthase variants are then tested for their enzymatic activity and in particular for their capacity to prefer acetone as a substrate rather than acetoacetylCoA. An assay for measuring the capacity of a HMG CoA synthase to use acetone as a substrate is described in Example 5. The formation of 3-hydroxy-3-methylbutyrate can be detected by comparison with standard compound, e.g. after separation by thin-layer chromatography, LC/MS and colorimetric assay after its derivatization or by mass spectrometry.

**[0029]** In particular, a reaction is carried out in a reaction mixture containing 40 mM Tris-HCl pH 8, 5 to 50 mM acetyl-CoA, 100 to 500 mM acetone, 1 MgCl<sub>2</sub> (except for mitochondria HMG-CoA synthase), 0.5 mM DTT and enzyme varying in the range from 0.2 to 8 mg/ml. Control reactions are carried in the absence of enzyme and one of the substrates.

**[0030]** The progress of synthesis is followed by analyzing aliquots taken after increasing period of incubation at 30 or 37°C. Typically, an aliquot of 50 µl is removed after 48 h of incubation, heated for 1 min at 100°C to eliminate the proteins, centrifuged and the supernatant is transferred to a clean vial for HIV detection by mass spectrometry. A solution of 3-hydroxy-3-methylbutyrate is prepared in 40 mM Tris-HCl pH 8, 1 mM MgCl<sub>2</sub>, 0.5 mM DTT, heated as described above and used as reference.

**[0031]** The samples are analyzed on a PE SCIEX API 2000 triple quadrupole mass spectrometer in negative ion mode with H<sub>2</sub>O/acetonitrile=60/40 containing 0.1% triethylamine as mobile phase, flow rate was 40 µl/min. 10 µl of each supernatant are mixed with an equal quantity of mobile phase and directly injected into the mass spectrometer. The presence of [3-hydroxy-3-methylbutyrate-H]<sup>-</sup> ion is monitored. 3-hydroxy-3-methylbutyrate synthesis can also be carried out in the presence of radiolabeled [2-<sup>14</sup>C] acetone. The formation of product is analyzed after separation of the reaction mixture by TLC or HPLC.

**[0032]** In a preferred embodiment the HMG CoA synthase employed in the present invention is an enzyme which has a K<sub>M</sub> value for acetone of 300 mM or lower, preferably of 250 mM or lower even more preferably of 200 mM or lower and particularly preferred of 150 mM or lower. It is preferred that the K<sub>M</sub> value is determined under the conditions described in Example 7. In another preferred embodiment the HMG CoA synthase employed in the present invention has a k<sub>cat</sub> value for the described reaction of at least 0.1 x 10<sup>-4</sup> sec<sup>-1</sup>, preferably at least 0.2 x 10<sup>-4</sup> sec<sup>-1</sup> even more preferably at least 0.5 x 10<sup>-4</sup> sec<sup>-1</sup> and particularly preferred at least 1 x 10<sup>-4</sup> sec<sup>-1</sup>, at least 2 x 10<sup>-4</sup> sec<sup>-1</sup>, at least 3 x 10<sup>-4</sup> sec<sup>-1</sup> or at least 5 x 10<sup>-4</sup> sec<sup>-1</sup>. It is preferred that the k<sub>cat</sub> value is determined under the conditions described in Example 7

**[0033]** It is known in the art that His264 of avian HMG CoA synthase plays a role in the interaction of the enzyme with acetoacetyl-CoA and that the Ala264 variant lacks interaction with the oxygen of the thioester moiety of acetoacetyl-CoA ( Misraa et al., Biochem. 35 (1996), 9610-9616). Thus, in order to develop variants of HMG CoA synthase which show a lower acceptance of acetoacetyl-CoA as a substrate but which accept acetone as a substrate, it is conceivable to systematically mutate in a HMG CoA synthase the histidine residue which corresponds to His264 of the avian HMG CoA synthase described in Misraa et al. (loc. cit.) so as to reduce or disable the acceptance of acetoacetyl-CoA as substrate.

**[0034]** In addition, HMG CoA synthase variants can be provided which show an increased activity. Steussy et al. (Biochemistry 45 (2006), 14407-14414), for example, describe a mutant of the Enterococcus faecalis HMG CoA synthase in which Ala110 was changed to Gly110 and which shows an 140-fold increase of the overall reaction rate.

**[0035]** Methods for identifying variants with improved enzymatic properties as regards the production of 3-hydroxy-3-methylbutyrate may also be carried out in the presence of a cofactor which allows for a steric and/or electronic complementation in the catalytic site of the enzyme/enzymes due to the fact that the substrate acetone is shorter than the natural substrate acetoacetyl-CoA of, HMG CoA synthase. One example of such a cofactor would be coenzyme A or a structurally closely related molecule such as S-nitroso-CoA.

**[0036]** The modified version of the HMG CoA synthase accepting acetone as a substrate but having a low affinity to acetoacetyl-CoA as a substrate or no longer accepting acetoacetyl-CoA as a substrate may be derived from a naturally occurring HMG CoA synthase or from an already modified, optimized or synthetically synthesized HMG CoA synthase.

**[0037]** The HMG CoA synthase employed in the process according to the present invention can be a natural version of the protein or a synthetic protein as well as a protein which has been chemically synthesized or produced in a biological system or by recombinant processes. The HMG CoA synthase may also be chemically modified, for example in order to improve its/their stability, resistance, e.g. to temperature, for facilitating its purification or its immobilization on a support. The enzyme may be used in isolated form, purified form, in immobilized form, as a crude or partially purified extract obtained from cells synthesizing the enzyme as chemically synthesized enzyme, as recombinantly produced enzyme, in the form of microorganisms producing them etc.

**[0038]** The process according to the present invention may be carried out in vitro or in vivo. An in vitro reaction is understood to be a reaction in which no cells are employed, i.e. an acellular reaction.

**[0039]** For carrying out the process in vitro the substrates for the reaction and the enzyme are incubated under conditions (buffer, temperature, cofactors etc.) allowing the enzyme to be active and the enzymatic conversion to occur. The reaction is allowed to proceed for a time sufficient to produce 3-hydroxy-3-methylbutyrate. The production of 3-hydroxy-3-methylbutyrate

and/or 3-hydroxy-3-methylbutyryl-CoA can be detected by comparison with standard compound after separation by thin-layer chromatography, LC/MS and colorimetric assay after its derivatization.

**[0040]** The enzyme may be in any suitable form allowing the enzymatic reaction to take place. It may be purified or partially purified or in the form of crude cellular extracts or partially purified extracts. It is also possible that the enzyme is immobilized on a suitable carrier.

**[0041]** Since the substrate acetone is in general shorter than the natural substrate used by the enzyme, acetoacetyl-CoA used by HMG CoA synthase, it may be advantageous to add to the reaction mixture a cofactor which allows for a steric and/or electronic complementation in the catalytic site of the enzyme. One example of such a cofactor would be coenzyme A or a structurally closely related molecule such as S-nitroso-CoA.

**[0042]** For carrying out the process in vivo use is made of a suitable organism/microorganism(s) which is/are capable of providing the substrates, i.e. acetone and acetyl-coenzyme A, and an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group according to formula (I), which is a HMG CoA synthase.

**[0043]** Thus, in the case of this embodiment the method according to the invention is characterised in that the conversion of acetone and a compound which provides an activated acetyl group is realized in the presence of an organism, preferably a microorganism capable of producing acetone and expressing an enzyme which is capable of the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group according to formula (I) expressing an enzyme with the activity of a HMG CoA synthase (EC 2.3.3.10).

**[0044]** The term "which is capable of producing acetone" in the context of the present invention means that the organism/microorganism has the capacity to produce acetone within the cell due to the presence of enzymes providing enzymatic activities allowing the production of acetone from metabolic precursors.

**[0045]** Acetone is produced by certain microorganisms, such as *Clostridium acetobutylicum*, *Clostridium beijerinckii*, *Clostridium cellulolyticum*, *Bacillus polymyxa* and *Pseudomonas putida*. The synthesis of acetone is best characterized in *Clostridium acetobutylicum*. It starts out with a reaction (reaction step 1) in which two molecules of acetyl-CoA are condensed into acetoacetyl-CoA. This reaction is catalyzed by acetyl-CoA acetyltransferase (EC 2.3.1.9). Acetoacetyl-CoA is then converted into acetoacetate by a reaction with acetic acid or butyric acid resulting also in the production of acetyl-CoA or butyryl-CoA (reaction step 2). This reaction is catalyzed e.g. by acetoacetylCoA transferase (EC 2.8.3.8). AcetoacetylCoA transferase is known from various organisms, e.g. from *E. coli* in which it is encoded by the *atoAD* gene or from *Clostridium acetobutylicum* in which it is encoded by the *ctfAB* gene. However, also other enzymes can catalyze this reaction, e.g. 3-oxoacid CoA transferase (EC 2.8.3.5) or succinate CoA ligase (EC 6.2.1.5).

**[0046]** Finally, acetoacetate is converted into acetone by a decarboxylation, step (reaction step 3) catalyzed by acetoacetate decarboxylase (EC 4.1.1.4).

**[0047]** The above described reaction steps 1 and 2 and the enzymes catalyzing them are not characteristic for the acetone synthesis and can be found in various organism. In contrast, reaction step 3 which is catalyzed by acetoacetate decarboxylase (EC 4.1.1.4) is only found in those organisms which are capable of producing acetone.

**[0048]** In one preferred embodiment the organism employed in the method according to the invention is an organism, preferably a microorganism, which naturally has the capacity to produce acetone. Thus, preferably the microorganism belongs to the genus *Clostridium*, *Bacillus* or *Pseudomonas*, more preferably to the species *Clostridium acetobutylicum*, *Clostridium beijerinckii*, *Clostridium cellulolyticum*, *Bacillus polymyxa* or *Pseudomonas putida*.

**[0049]** In a further preferred embodiment, the organism employed in the method according to the invention is an organism, preferably a microorganism, which naturally has the capacity to produce acetone and which is recombinant in the sense that it has further been genetically modified so as to express an enzyme as defined above. The term "recombinant" in one embodiment means that the organism is genetically modified so as to contain a foreign nucleic acid molecule encoding an enzyme as defined above. In a preferred embodiment the organism has been genetically modified so as to contain a foreign nucleic acid molecule encoding an enzyme as defined above, i.e. a HMG CoA synthase.

**[0050]** The term "foreign" in this context means that the nucleic acid molecule does not naturally occur in said organism/microorganism. This means that it does not occur in the same structure or at the same location in the organism/microorganism. In one preferred embodiment, the foreign nucleic acid molecule is a recombinant molecule comprising a promoter and a coding sequence encoding the respective enzyme, i.e. a HMG CoA synthase, in which the promoter driving expression of the coding sequence is heterologous with respect to the coding sequence. Heterologous in this context means that the promoter is not the promoter naturally driving the expression of said coding sequence but is a promoter naturally driving expression of a different coding sequence, i.e., it is derived from another gene, or is a synthetic promoter or a chimeric promoter. Preferably, the promoter is a promoter heterologous to the organism/microorganism, i.e. a promoter which does not naturally occur in the respective organism/microorganism. Even more preferably, the promoter is an inducible promoter. Promoters for driving expression in different types of organisms, in particular in microorganisms, are well known to the person skilled in the art.

**[0051]** In another preferred embodiment the nucleic acid molecule is foreign to the organism/microorganism in that the encoded enzyme, i.e. the HMG CoA-synthase is not endogenous to the organism/microorganism, i.e. are naturally not expressed by the organism/microorganism when it is not genetically modified. In other words, the encoded HMG CoA synthase is heterologous with respect to the organism/microorganism.

**[0052]** The term "recombinant" in another embodiment means that the organism is genetically modified in the regulatory region controlling the expression of an enzyme as defined above which naturally occurs in the organism so as to lead to an increase in expression of the respective enzyme in comparison to a corresponding non-genetically modified organism. The meaning of the term high "higher expression" is described further below.

**[0053]** Such a modification of a regulatory region can be achieved by methods known to the person skilled in the art. One example is to exchange the naturally occurring promoter by a promoter which allows for a higher expression or to modify the naturally occurring promoter so as to show a higher expression. Thus, in this embodiment the organism contains in the regulatory region of the gene encoding an enzyme as defined above a foreign nucleic acid molecule which naturally does not occur in the organism and which leads to a higher expression of the enzyme in comparison to a corresponding non-genetically modified organism.

**[0054]** The foreign nucleic acid molecule may be present in the organism/microorganism in extrachromosomal form, e.g. as plasmid, or stably integrated in the chromosome. A stable integration is preferred.

**[0055]** In a further preferred embodiment the organism/microorganism is characterized in that the expression/activity of an enzyme as defined above, i.e. of a HMG CoA synthase, is higher in the organism/microorganism genetically modified with the foreign nucleic acid molecule in comparison to the corresponding non-genetically modified organism/microorganism. A "higher" expression/activity means that the expression/activity of the enzyme, i.e. of the HMG CoA synthase, in the genetically modified microorganism is at least 10%, preferably at least 20%, more preferably at least 30% or 50%, even more preferably at least 70% or 80% and particularly preferred at least 90% or 100% higher than in the corresponding non-genetically modified organism/microorganism. In even more preferred embodiments the increase in expression/activity may be at least 150%, at least 200% or at least 500%. In particularly preferred embodiments the expression is at least 10-fold, more preferably at least 100-fold and even more preferred at least 1000-fold higher than in the corresponding non-genetically, modified organism/microorganism.

**[0056]** The term "higher" expression/activity also covers the situation in which the corresponding non-genetically modified organism/microorganism does not express a corresponding enzyme, i.e. a HMG CoA synthase, so that the corresponding expression/activity in the non-genetically modified organism/microorganism is zero.

**[0057]** Methods for measuring the level of expression of a given protein in a cell are well known to the person skilled in the art. In one embodiment, the measurement of the level of expression is done by measuring the amount of the corresponding protein. Corresponding methods are well known to the person skilled in the art and include Western Blot, ELISA etc. In another embodiment the measurement of the level of expression is done by measuring the amount of the corresponding RNA. Corresponding methods are well known to the person skilled in the art and include, e.g., Northern Blot.

**[0058]** Methods for measuring the enzymatic activity of the above-mentioned enzyme, HMG CoA synthase, are known in the art and have already been described above.

**[0059]** In another preferred embodiment, the organism employed in the method according to the invention is a genetically modified organism, preferably a microorganism, derived from an organism/microorganism which naturally does not produce acetone but which has been genetically modified so as to produce acetone, i.e. by introducing the gene(s) necessary for allowing

the production of acetone in the organism/microorganism. In principle any microorganism can be genetically modified in this way. The enzymes responsible for the synthesis of acetone have been described above. Genes encoding corresponding enzymes are known in the art and can be used to genetically modify a given microorganism so as to produce acetone. As described above, the reaction steps 1 and 2 of the acetone synthesis occur naturally in most organisms. However, reaction step 3 is characteristic and crucial for acetone synthesis. Thus, in a preferred embodiment, a genetically modified organism/microorganism derived from an organism/microorganism which naturally does not produce acetone is modified so as to contain a nucleotide sequence encoding an enzyme catalyzing the conversion of acetoacetate into acetone by decarboxylation, e.g. an acetoacetate decarboxylase (EC 4.1.1.4). Nucleotide sequences from several organisms encoding this enzyme are known in the art, e.g. the *adc* gene from *Clostridium acetobutylicum* (Uniprot accession numbers P23670 and P23673), *Clostridium beijerinckii* (*Clostridium* MP; Q9RPK1), *Clostridium pasteurianum* (Uniprot accession number P81336), *Bradyrhizobium* sp. (strain BTAi1 / ATCC BAA-1182; Uniprot accession number A5EBU7), *Burkholderia mallei* (ATCC 10399 A9LBS0), *Burkholderia mallei* (Uniprot accession number A3MAE3), *Burkholderia mallei* FMH A5XJB2, *Burkholderia cenocepacia* (Uniprot accession number A0B471), *Burkholderia ambifaria* (Uniprot accession number Q0b5P1), *Burkholderia phytofirmans* (Uniprot accession number B2T319), *Burkholderia* spec. (Uniprot accession number Q38ZU0), *Clostridium botulinum* (Uniprot accession number B2TLN8), *Ralstonia pickettii* (Uniprot accession number B2UIG7), *Streptomyces nogalater* (Uniprot accession number Q9EY17), *Streptomyces avermitilis* (Uniprot accession number Q82NF4), *Legionella pneumophila* (Uniprot accession number Q5ZXQ9), *Lactobacillus salivarius* (Uniprot accession number Q1WVG5), *Rhodococcus* spec. (Uniprot accession number Q0S7W4), *Lactobacillus plantarum* (Uniprot accession number Q890G0), *Rhizobium leguminosarum* (Uniprot accession number Q1M911), *Lactobacillus casei* (Uniprot accession number Q03B66), *Francisella tularensis* (Uniprot accession number Q0BLC9), *Saccharopolyspora erythraea* (Uniprot accession number A4FKR9), *Korarchaeum cryptofilum* (Uniprot accession number B1L3N6), *Bacillus amyloliquefaciens* (Uniprot accession number A7Z8K8), *Cochliobolus heterostrophus* (Uniprot accession number Q8NJ03), *Sulfolobus islandicus* (Uniprot accession number C3ML22) and *Francisella tularensis* subsp. *holarctica* (strain OSU18).

**[0060]** More preferably, the organism, preferably microorganism, is genetically modified so as to be transformed with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 2 of the acetone synthesis, i.e. the conversion of acetoacetyl CoA into acetoacetate.

**[0061]** Even more preferably, the organism, preferably microorganism, is genetically modified so as to be transformed with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 1 of the acetone synthesis, i.e. the condensation of two molecules of acetyl CoA into acetoacetyl CoA.

**[0062]** In a particularly preferred embodiment the organism/microorganism is genetically modified so as to be transformed with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 1 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 2 of the acetone synthesis or with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 1 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 3 of the acetone synthesis or with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 2 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 3 of the acetone synthesis or with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 1 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 2 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 3 of the acetone synthesis.

**[0063]** Methods for preparing the above mentioned genetically modified organism, preferably microorganisms, are well known in the art. Thus, generally, the organism/microorganism is transformed with a DNA construct allowing expression of the respective enzyme in the microorganism. Such a construct normally comprises the coding sequence in question linked to regulatory sequences allowing transcription and translation in the respective host cell, e.g. a promoter and/enhancer and/or transcription terminator and/or ribosome binding sites etc. The prior art already describes microorganisms which have been genetically modified so as to be able to produce acetone. In particular genes from, e.g., *Clostridium acetobutylicum* have been introduced into *E. coli* thereby allowing the synthesis of acetone in *E. coli*, a bacterium which naturally does not produce acetone (Bermejo et al., *Appl. Environ. Microbiol.* 64 (1998), 1079-1085; Hanai et al., *Appl. Environ. Microbiol.* 73 (2007), 7814-7818). In particular Hanai et al. (loc. cit.) shows that it is sufficient to introduce a nucleic acid sequence encoding an acetoacetate decarboxylase (such as that from *Clostridium acetobutylicum*) in order to achieve acetone production in *E. coli* indicating that the endogenous enzymes in *E. coli* catalyzing the above-mentioned reaction steps 1 and 2 (i.e. the expression products of the *E. coli* *atoB* and *atoAD* genes) are sufficient to provide substrate for the acetone production.

**[0064]** In a particularly preferred embodiment the organism, preferably a microorganism, employed in the method according to

the invention is a recombinant organism/microorganism derived from an organism/microorganism which naturally does not produce acetone but which has been genetically modified, as described above, so as to produce acetone and which expresses an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group as defined above. The term "recombinant" in this context preferably means that the organism is recombinant in the sense that it has further been genetically modified so as to express an enzyme as defined above. The term "recombinant" in one embodiment means that the organism is genetically modified so as to contain a foreign nucleic acid molecule encoding an enzyme as defined above, i.e. a HMG CoA synthase.

**[0065]** As regards the definition of the term "foreign nucleic acid molecule" the same applies what has already been set forth above.

**[0066]** The term "recombinant" in another embodiment means that the organism is genetically modified in the regulatory region controlling the expression of an enzyme as defined above which naturally occurs in the organism so as to lead to an increase in expression of the respective enzyme in comparison to a corresponding non-genetically modified organism. The meaning of the term high "higher expression" is described further below.

**[0067]** Such a modification of a regulatory region can be achieved by methods known to the person skilled in the art. One example is to exchange the naturally occurring promoter by a promoter which allows for a higher expression or to modify the naturally occurring promoter so as to show a higher expression. Thus, in this embodiment the organism contains in the regulatory region of the gene encoding an enzyme as defined above a foreign nucleic acid molecule which naturally does not occur in the organism and which leads to a higher expression of the enzyme in comparison to a corresponding non-genetically modified organism.

**[0068]** Preferably such an organism/microorganism is characterized in that the expression/activity of said enzyme, i.e. the HMG CoA synthase, is higher in the recombinant organism/microorganism in comparison to the corresponding non-genetically modified organism/microorganism. A "higher" expression/activity means that the expression/activity of the enzyme, i.e. the HMG CoA synthase, in the genetically modified organism/microorganism is at least 10%, preferably at least 20%, more preferably at least 30% or 50%, even more preferably at least 70% or 80% and particularly preferred at least 90% or 100% higher than in the corresponding non-genetically modified organism/microorganism. In even more preferred embodiments the increase in expression/activity may be at least 150%, at least 200% or at least 500%. In particularly preferred embodiments the expression is at least 10-fold, more preferably at least 100-fold and even more preferred at least 1000-fold higher than in the corresponding non-genetically modified organism/microorganism.

**[0069]** The term "higher" expression/activity also covers the situation in which the corresponding non-genetically modified organism/microorganism does not express said enzyme, i.e. a HMG CoA synthase, so that the corresponding expression/activity in the non-genetically modified organism/microorganism is zero. As regards the methods for measuring the level of expression or activity, the same applies what has already been said above.

**[0070]** The term "organism" as used in the context of the present invention refers in general to any possible type of organism, in particular eukaryotic organisms, prokaryotic organisms and archaeobacteria. The term includes non-human animal, plants, fungi, bacteria and J archaeobacteria. The term also includes isolated cells or cell aggregates of such organisms, like tissue or calli.

**[0071]** In one preferred embodiment, the organism is a microorganism. The term "microorganism" in the context of the present invention refers to prokaryotic cells, in particular bacteria, as well as to fungi, such as yeasts, and also to algae and archaeobacteria. In one preferred embodiment, the microorganism is a bacterium. In principle any bacterium can be used. Preferred bacteria to be employed in the process according to the invention are bacteria of the genus *Bacillus*, *Clostridium*, *Pseudomonas* or *Escherichia*. In a particularly preferred embodiment the bacterium belongs to the genus *Escherichia* and even more preferred to the species *Escherichia coli*.

**[0072]** In another preferred embodiment the microorganism is a fungus, more preferably a fungus of the genus *Saccharomyces*, *Schizosaccharomyces*, *Aspergillus* or *Trichoderma* and even more preferably of the species *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Aspergillus niger* or of the species *Trichoderma reesei*.

**[0073]** In still another preferred embodiment the microorganism is a photosynthetically active microorganism such as bacteria which are capable of carrying out photosynthesis or micro-algae.

**[0074]** In a particularly preferred embodiment the microorganism is an algae, more preferably an algae belonging to the diatomeae.

**[0075]** If microorganism are used in the context of the method of the present invention, it is also conceivable to carry out the method according to the invention in a manner in which two types of microorganisms are employed, i.e. one type which produces acetone and one type which uses the acetone produced by the first type of microorganisms to convert it with the help of an enzyme as defined herein above.

**[0076]** When the process according to the invention is carried out in vivo by using microorganisms providing the respective enzyme activity/activities, the microorganisms are cultivated under suitable culture conditions allowing the occurrence of the enzymatic reaction(s). The specific culture conditions depend on the specific microorganism employed but are well known to the person skilled in the art. The culture conditions are generally chosen in such a manner that they allow the expression of the genes encoding the enzymes for the respective reactions. Various methods are known to the person skilled in the art in order to improve and fine-tune the expression of certain genes at certain stages of the culture such as induction of gene expression by chemical inducers or by a temperature shift.

**[0077]** In another preferred embodiment the organism employed in the method according to the invention is an organism which is capable of photosynthesis, such as a plant or microalgae. In principle any possible plant can be used, i.e. a monocotyledonous plant or a dicotyledonous plant. It is preferable to use a plant which can be cultivated on an agriculturally meaningful scale and which allows to produce large amounts of biomass. Examples are grasses like *Lolium*, cereals like rye, barley, oat, millet, maize, other starch storing plants like potato or sugar storing plants like sugar cane or sugar beet. Conceivable is also the use of tobacco or of vegetable plants such as tomato, pepper, cucumber, egg plant etc. Another possibility is the use of oil storing plants such as rape seed, olives etc. Also conceivable is the use of trees, in particular fast growing trees such as eucalyptus, poplar or rubber tree (*Hevea brasiliensis*).

**[0078]** The present invention also relates to an organism, preferably a microorganism, which is characterized by the following features:

1. (a) it is capable of producing acetone; and
2. (b) it expresses an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group as defined above, i.e. an enzyme with the activity of HMG CoA synthase (EC 2.3.3.10).

**[0079]** As regards the source, nature, properties, sequence etc. of the enzyme, i.e. the HMG CoA synthase, expressed in the organism according to the invention, the same applies as has been set forth above in connection With the method according to the invention.

**[0080]** In one preferred embodiment, the organism according to the invention is an organism, preferably a microorganism which naturally has the capacity to produce acetone, i.e., feature (a) mentioned above is a feature which the organism, preferably microorganism, shows naturally. Thus, preferably the organism is a microorganism which belongs to the genus *Clostridium*, *Bacillus* or *Pseudomonas*, more preferably to the species *Clostridium acetobutylicum*, *Clostridium beijerinckii*, *Clostridium cellulolyticum*, *Bacillus polymyxa* or *Pseudomonas putida*.

**[0081]** In another preferred embodiment, the organism, preferably microorganism, according to the invention is a genetically modified organism/microorganism derived from an organism/microorganism which naturally does not produce acetone but which has been genetically modified so as to produce acetone, i.e. by introducing the gene(s) necessary for allowing the production of acetone in the organism/microorganism. In principle any organism/microorganism can be genetically modified in this way. The enzymes responsible for the synthesis of acetone have been described above. Genes encoding corresponding enzymes are known in the art and can be used to genetically modify a given organism, preferably microorganism so as to produce acetone.

**[0082]** In a preferred embodiment, a genetically modified organism/microorganism derived from an organism/microorganism which naturally does not produce acetone is modified so as to contain a nucleotide sequence encoding an enzyme catalyzing the conversion of acetoacetate into acetone by decarboxylation, e.g. an acetoacetate decarboxylase (EC 4.1.1.4). Nucleotide sequences from several organisms encoding this enzyme are known in the art, e.g. the *adc* gene from *Clostridium acetobutylicum*. More preferably, the organism/microorganism is genetically modified so as to be transformed with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 2 of the acetone synthesis, i.e. the conversion of

acetoacetyl CoA into acetoacetate.

**[0083]** Even more preferably, the organism/microorganism is genetically modified so as to be transformed with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 1 of the acetone synthesis, i.e. the condensation of two molecules of acetyl CoA into acetoacetyl CoA.

**[0084]** In a particularly preferred embodiment the organism/microorganism is genetically modified so as to be transformed with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 1 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 2 of the acetone synthesis or with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 1 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 3 of the acetone synthesis or with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 2 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 3 of the acetone synthesis or with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 1 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 2 of the acetone synthesis and with a nucleic acid molecule encoding an enzyme capable of catalyzing the above mentioned reaction step 3 of the acetone synthesis.

**[0085]** Methods for preparing the above mentioned genetically modified organisms/microorganisms are well known in the art. Thus, generally, the organism/microorganism is transformed with a DNA construct allowing expression of the respective enzyme in the organism/microorganism. Such a construct normally comprises the coding sequence in question linked to regulatory sequences allowing transcription and translation in the respective host cell, e.g. a promoter and/enhancer and/or transcription terminator and/or ribosome binding sites etc. The prior art already describes organism, in particular microorganisms which have been genetically modified so as to be able to produce acetone. In particular genes from, e.g., *Clostridium acetobutylicum* have been introduced into *E. coli* thereby allowing the synthesis of acetone in *E. coli*, a bacterium which naturally does not produce acetone (Bermejo et al., *Appl. Environ. Microbiol.* 64 (1998); 1079-1085; Hanal et al., *Appl. Environ. Microbiol.* 73 (2007), 7814-7818). In particular Hanai et al. (loc. cit.) shows that it is sufficient to introduce a nucleic acid sequence encoding an acetoacetate decarboxylase (such as that from *Clostridium acetobutylicum*) in order to achieve acetone production in *E. coli* indicating that the endogenous enzymes in *E. coli* catalyzing the above-mentioned reaction steps 1 and 2 (i.e. the expression products of the *E. coli* *atoB* and *atoAD* genes) are sufficient to provide substrate for the acetone production.

**[0086]** In a further preferred embodiment the organism, preferably a microorganism, according to the invention is genetically modified so as to express an enzyme as defined above which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound as defined above which provides an activated acetyl group. In this context, the term "recombinant" means in a first aspect that the organism contains a foreign nucleic acid molecule encoding an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group, i.e. a foreign nucleic acid molecule encoding a HMG CoA synthase. The term "foreign" in this context means that the nucleic acid molecule does not naturally occur in said organism/microorganism. This means that it does not occur in the same structure or at the same location in the organism/microorganism. In one preferred embodiment, the foreign nucleic acid molecule is a recombinant molecule comprising a promoter and a coding sequence encoding said enzyme, i.e. the HMG CoA synthase, in which the promoter driving expression of the coding sequence is heterologous with respect to the coding sequence. Heterologous in this context means that the promoter is not the promoter naturally driving the expression of said coding sequence but is a promoter naturally driving expression of a different coding sequence, i.e., it is derived from another gene, or is a synthetic promoter or a chimeric promoter. Preferably, the promoter is a promoter heterologous to the organism/microorganism, i.e. a promoter which does not naturally occur in the respective organism/microorganism. Even more preferably, the promoter is an inducible promoter. Promoters for driving expression in different types of organisms, in particular microorganisms, are well known to the person skilled in the art. In another preferred embodiment the nucleic acid molecule is foreign to the organism/microorganism in that the encoded enzyme, i.e. the HMG CoA synthase, is not endogenous to the organism/microorganism, i.e. is not naturally expressed by the organism/microorganism when it is not genetically modified. In other words, the encoded enzyme, i.e. the HMG CoA synthase, is heterologous with respect to the organism/microorganism.

**[0087]** The term "recombinant" in another aspect means that the organism is genetically modified in the regulatory region controlling the expression of an enzyme as defined above which naturally occurs in the organism so as to lead to an increase in expression of the respective enzyme in comparison to a corresponding non-genetically modified organism. The meaning of the term high "higher expression" is described further below.

**[0088]** Such a modification of a regulatory region can be achieved by methods known to the person skilled in the art. One example is to exchange the naturally occurring promoter by a promoter which allows for a higher expression or to modify the naturally occurring promoter so as to show a higher expression. Thus, in this embodiment the organism contains in the regulatory region of the gene encoding an enzyme as defined above a foreign nucleic acid molecule which naturally does not occur in the organism and which leads to a higher expression of the enzyme in comparison to a corresponding non-genetically modified organism.

**[0089]** In a further preferred embodiment the organism/microorganism is characterized in that the expression/activity of said enzyme, i.e. the HMG CoA synthase, is higher in the organism/microorganism genetically modified with the foreign nucleic acid molecule in comparison to the corresponding non-genetically modified organism/microorganism. A "higher" expression/activity means that the expression/activity of the enzyme, i.e. the HMG CoA synthase, in the genetically modified organism/microorganism is at least 10%, preferably at least 20%, more preferably at least 30% or 50%, even more preferably at least 70% or 80% and particularly preferred at least 90% or 100% higher than in the corresponding non-genetically modified organism/microorganism.. In even more preferred embodiments the increase in expression/activity may be at least 150%, at least 200% or at least 500%. In particularly preferred embodiments the expression is at least 10-fold, more preferably at least 100-fold and even more preferred at least 1000-fold higher than in the corresponding non-genetically modified organism/microorganism.

**[0090]** The term "higher" expression/activity also covers the situation in which the corresponding non-genetically modified organism/microorganism does not express a corresponding enzyme, i.e. a HMG CoA synthase, so that the corresponding expression/activity in the non-genetically modified organism/microorganism is zero.

**[0091]** Methods for measuring the level of expression of a given protein in a cell are well known to the person skilled in the art. In one embodiment, the measurement of the level of expression is done by measuring the amount of the corresponding protein. Corresponding methods are well known to the person skilled in the art and include Western Blot, ELISA etc. In another embodiment the measurement of the level of expression is done by measuring the amount of the corresponding RNA. Corresponding methods are well known to the person skilled in the art and include, e.g., Northern Blot.

**[0092]** Methods for measuring the enzymatic activity of the above-mentioned enzyme, i.e. of a HMG CoA synthase, are known in the art and have already been described above.

**[0093]** The term "organism" as used in the context of the present invention refers in general to any possible type of organism, in particular eukaryotic organisms, prokaryotic organisms and archaeobacteria. The term includes non-human animal, plants, fungi, bacteria and archaeobacteria. The term also includes isolated cells or cell aggregates of such organisms, like tissue or calli.

**[0094]** In one preferred embodiment, the organism is a microorganism. The term "microorganism" in the context of the present invention refers to prokaryotic cells, in particular bacteria, as well as to fungi, such as yeasts, and also to algae and archaeobacteria. In one preferred embodiment, the microorganism is a bacterium. In principle any bacterium can be used. Preferred bacteria to be employed in the process according to the invention are bacteria of the genus *Bacillus*, *Clostridium*, *Pseudomonas* or *Escherichia*. In a particularly preferred embodiment the bacterium belongs to the genus *Escherichia* and even more preferred to the species *Escherichia coli*.

**[0095]** In another preferred embodiment the microorganism is a fungus, more preferably a fungus of the genus *Saccharomyces*, *Schizosaccharomyces*, *Aspergillus* or *Trichoderma* and even more preferably of the species *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Aspergillus niger* or of the species *Trichoderma reesei*.

**[0096]** In still another preferred embodiment the microorganism is a photosynthetically active microorganism such as bacteria which are capable of carrying out photosynthesis or micro-algae.

**[0097]** In a particularly preferred embodiment the microorganism is an algae, more preferably an algae from the genus belonging to the diatomeae.

**[0098]** In another preferred embodiment the organism according to the invention is an organism which is capable of photosynthesis, such as a plant or micro-algae. In principle, it can be any possible plant, i.e. a monocotyledonous plant or a dicotyledonous plant. It is preferably a plant which can be cultivated on an agriculturally meaningful scale and which allows to produce large amounts of biomass. Examples are grasses like *Lolium*, cereals like rye, barley, oat, millet, maize, other starch storing plants like potato or sugar storing plants like sugar cane or sugar beet. Conceivable is also the use of tobacco or of vegetable plants such as tomato, pepper, cucumber, egg plant etc. In another preferred embodiment the plant is an oil storing

plants such as rape seed, olives etc. Also conceivable is the use of trees, in particular fast growing trees such as eucalyptus, poplar or rubber tree (*Hevea brasiliensis*).

**[0099]** The present invention also relates to the use of an organism, preferably a microorganism, which is characterized by the following features:

1. (a) it is capable of producing acetone; and
2. (b) it expresses an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group as defined herein above, i.e. an enzyme with the activity of a HMG CoA synthase (EC 2.3.3.10) for the production of 3-hydroxy-3-methylbutyric acid.  
i.e., the present invention also relates to the use of an organism/microorganism according to the invention for the production of 3-hydroxy-3-methylbutyric acid.

**[0100]** The present invention also relates to a composition comprising an organism according to the present invention.

**[0101]** Moreover, the present invention also relates to a composition comprising (i) acetone; and (ii) a compound which provides an activated acetyl group as defined herein above; and (iii) an enzyme which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group as defined herein above.

**[0102]** Moreover, the present invention also relates to the use of an enzyme as defined above which is capable of catalyzing the formation of a covalent bond between the carbon atom of the oxo (i.e. the C=O) group of acetone and the carbon atom (C<sup>2</sup>) corresponding to the methyl group of the compound which provides an activated acetyl group as defined herein above for the production of 3-hydroxy-3-methylbutyric acid.

**[0103]** Finally, the present invention also relates to the use of acetone for the production of 3-hydroxy-3-methylbutyric acid, comprising the enzymatic conversion of acetone and a compound which provides an activated acetyl group as defined herein above wherein the enzymatic conversion is achieved by an enzyme as described above in connection with the method according to the invention, preferably with an enzyme having the enzymatic activity of a HMG CoA synthase, and more preferably the conversion is achieved by the use of an organism according to the invention.

**Figure 1:**

Chemical structure of 3-hydroxy-3-methylbutyric acid (also referred to as beta-hydroxyisovalerate)

**Figure 2:**

Reaction scheme of the reaction catalysed by HMG-CoA synthase

**Figure 3:**

Reaction scheme of the reaction catalysed by HMG-CoA lyase (falling outside the scope of the claims)

**Figure 4:**

Reaction schemes of the reactions of the pksX pathway including the reaction catalysed by the PksG protein (falling outside the scope of the claims).

**Figure 5:**

Reaction scheme of the reaction of the conversion of acetone and a compound containing an activated acetyl group into 3-hydroxy-3-methylbutyric acid X stands for S-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH(OH)-C(CH<sub>3</sub>)<sub>2</sub>-CH<sub>2</sub>-O-P<sub>0</sub>2H-O-P<sub>0</sub>2H-C<sub>10</sub>H<sub>13</sub>N<sub>5</sub>O<sub>7</sub>P (coenzyme A).

**Figure 6:**

Mass spectra of commercial available 3-hydroxy-3-methylbutyrate

**Figure 7:**

Mass spectra of formation of 3-hydroxy-3-methylbutyrate from acetyl-CoA and acetone in the presence of HMG-CoA synthase from *Gallus gallus* (P23228).

**Figure 8:**

Mass spectra of the control assay without enzyme.

**Figure 9:**

Michaelis-Menten plot for the reaction with the HMG CoA synthase of *S. epidermidis* described in Example 7

[0104] The following examples serve to illustrate the invention.

**Example 1****Bioinformatic method used to create HMG-CoA synthases and HMG-CoA lyases database (falling outside the scope of the claims)**

[0105] A panel of 12 HMG-CoA synthases and 8 HMG-CoA lyases were selected to create a non-redundant set of proteins aiming to represent the diversity of these enzyme classes as found across eukaryotic organisms. These proteins were identified by performing multiple sequence-based and text-based searches on the Universal Protein Resource Database Uniprot (<http://www.uniprot.org/>). They all contain unique features such as conserved protein domains and motifs characteristic to the enzyme class of interest. In order to effectively cover the sequence diversity without having to screen a large set of proteins, the initial pool of enzymes was narrowed down by grouping them into clusters of sequences with more than 85% homology and then selecting one single candidate sequence representative of each cluster. Protein sequence identity ranged from 30% to 80% and from 50% to 80% between any two proteins from the HMG-CoA synthases panel and the lyases panel respectively.

[0106] The same approach was applied to select the HMG-CoA synthases and HMG-CoA lyases from prokaryotic organisms. The created set contained 50 proteins homologues to HMG-CoA synthases, including pksG proteins, and 59 proteins homologues to HMG-CoA lyases.

**Example 2****Cloning, expression and purification of a collection of HMG-CoA Leases and HMG-CoA synthases (falling outside the scope of the claims)****Gene cloning:**

[0107] The nucleic acid sequences coding for HMG-CoA synthase and lyase from eukaryotic organism were optimized for E.coli codon preference and the genes were obtained by chemical synthesis (GeneArt).

[0108] The genes encoding for HMG-CoA synthases and lyases from prokaryotic organisms were cloned from genomic DNA of different origins by routine recombinant techniques. These genes were then inserted in a His-tag containing pET 25b and pET 22b vectors (Novagen), respectively, for eukaryotic and prokaryotic organisms.

**Overexpression in E. coli:**

[0109] Plasmids are electroporated into E. coli BL21 bacteria (Novagen) that are then spread on an ampicillin containing LB-Agar Petri dish. The cultures are grown at 30°C on TB medium, containing 0.5 M sorbitol, 5 mM betaine, 100 µg/ml ampicillin under moderate shaking. When OD (600 nm) reached 0.8, IPTG is added to a final concentration of 1 mM, and expression is run for 16 hours at 20°C under moderated shaking. The bacteria cells are then harvested by centrifugation at 4°C, 10.000 rpm, 20 minutes and frozen at -80°C.

**Cell extract preparation:**

[0110] Cell extracts are prepared by resuspending 1.6 g of cell pellet in 5 ml 50 mM Na<sub>2</sub>HPO<sub>4</sub> buffer, containing 300 mM NaCl, 5 mM MgCl<sub>2</sub>, 1 mM DTT pH 8. 20 µl lysonase (Novagen) is then added to the preparations, which are incubated for 10 min at room

temperature and 20 min on ice. The cell lysis is achieved by triple sonication treatment of 5 minutes in ultrasonic water-bath on ice and homogenization of extract between each pulse. The crude extracts are then clarified by centrifugation at 4°C, 10.000 rpm, 20 minutes.

#### **Protein purification:**

[0111] The clear supernatants are loaded onto the PROTINO-1000 Ni-IDA column (Macherey-Nagel) which enables the specific immobilization of proteins carrying 6-histidine tails. The columns are washed and the enzymes are eluted with 4 ml 50 mM - Na<sub>2</sub>HPO<sub>4</sub> buffer, containing 300 mM NaCl, 5 mM MgCl<sub>2</sub>, 1 mM DTT, 250 mM imidazole pH 8. The enzyme containing fractions are then concentrated and desalted on Amicon Ultra-4 10 kDa filter unit (Millipore) and resuspended in 250 µl 40 mM Tris-HCl pH8, containing 0.5 mM DTT. The protein concentration is determined by the Bradford method.

[0112] The homogeneity of purified enzymes varied from 20 % to 75 %.

#### **Example 3**

#### **Measure of the HMG-CoA synthase activity using natural substrates acetoacetyl-CoA and acetyl-coma**

[0113] The HMG-CoA synthase activity is measured according to Clinkenbeard et al. (J. Biol.Chem. 250 (1975), 3108-3116). The standard assay medium mixture for HMG-CoA synthases contains 40 mM Tris-HCl pH 8, 1 mM MgCl<sub>2</sub>, 100 µM acetoacetyl-CoA, 200 µM acetyl-CoA, 0.5mM DTT in a total volume of 1 ml. Mitochondria HMG-CoA synthases are assayed in the absence of MgCl<sub>2</sub> to avoid the inhibition observed for this enzyme (Reed et al., J. Biol.Chem. 250. (1975), 3117-3123). Reaction is initiated by addition of 0.02 mg/mL enzyme.

[0114] A Control assay was carried out in the absence of enzyme. HMG-CoA synthase activity was measured by monitoring the decrease in absorbance at 303 nm that accompanies the acetyl-CoA-dependent disappearance of the enolate form of acetoacetyl-CoA. To account for non-specific disappearance of acetoacetyl-CoA, results obtained in a control assay lacking enzyme were subtracted from results obtained in test samples. The apparent absorption coefficient for acetoacetyl-CoA under the assay conditions was 5600 M<sup>-1</sup>

**Table 1:** Physiological activity of some purified HMG-CoA synthases or enzymes homologous to HMG CoA synthases

Uniprot Accession number	Organism	Physiological activity µmol/min - mg protein
P54961	Blattella germanica (German cockroach)	0,02
P23228	Gallus gallus (Chicken)	0,02
Q01581	Homo sapiens (Human)	0,03
P54873	Arabidopsis thaliana	1,19
P54871	Caenorhabditis elegans	0,23
P54874	Schizosaccharomyces pombe (Fission yeast)	0,61
P54839	Saccharomyces cerevisiae (Baker's yeast)	0,28
P54872	Dictyostelium discoideum (Slime mold)	0,09
Q86HL5	Dictyostelium discoideum (Slime mold)	0,02
Q9M6U3	Brassica juncea	0,02
A5FM54	Flavobacterium johnsoniae	0,02
Q03WZ0	Leuconostoc mesenteroides	0,28
Q2NHU7	Methanosphaera stadtmanae	0,02
Q8CN06	Staphylococcus epidermidis	0,07
Q03QR0	Lactobacillus brevis	0,18

Uniprot Accession number	Organism	Physiological activity $\mu\text{mol}/\text{min} \cdot \text{mg}$ protein
A6UPL1	Methanosarcina mazei	0,01
B2HGT6	Mycobacterium marinum	0,01
Q4L958	Staphylococcus haemolyticus	0,18
Q4A0D6	Staphylococcus saprophyticus	0,08
Q1GAH5	Lactobacillus delbrueckii	0,32

#### Example 4

#### Measuring of the HMG-CoA lyase activity using natural substrate HMG-CoA (falling outside the scope of the claims)

[0115] HMG-CoA lyase activity is measured according to Mellanby J et al. (Methods of Enzymatic Analysis; Bergmeyer Ed. (1963), 454-458). The complete reaction mixture (1ml) containing 40 mM Tris-HCl pH 8, 1 mM  $\text{MgCl}_2$ , 0.5 mM DTT, 0.4 mM HMG-CoA, 0.2 mM NADH, 5 units of 3-hydroxybutyrate dehydrogenase is incubated for 5 min before adding 0.005 mg/ml of HMG-CoA lyase and then the progress of the reaction is monitored by the decrease in absorbance at 340 nm. A control assay was carried out in the absence of enzyme.

[0116] To account for non-specific disappearance of NADH, results obtained in a control assay lacking enzyme were subtracted from results obtained in test samples. Specific activities were calculated as  $\Delta\mu\text{mol NADH}/\text{min mg}$  protein.

**Table 2:** Physiological activity of some purified HMG-CoA lyases

Uniprot Accession number	Organism	Physiological activity $\mu\text{mol}/\text{min} \cdot \text{mg}$ protein
A8WG57	Danio rerio (Zebrafish) (Brachydanio rerio)	4,05
Q29448	Bos taurus (Bovine)	5,79
B6U7B9	Zea mays	13,31
A5FHS2	Flavobacterium johnsoniae	2,89
A1VJH1	Polaromonas naphthalenivorans	34,92
A9IFQ7	Bordetella petrii	9,84
A9IR28	Bordetella petrii	1,74
A1VT25	Polaromonas naphthalenivorans	0,39

#### Example 5

#### 3-hydroxy-3-methylbutyrate production

[0117] The complete reaction for 3-hydroxy-3-methylbutyrate synthesis contained 40 mM Tris-HCl pH 8, 5 to 50 mM acetyl-CoA, 100 to 500 mM acetone, 1  $\text{MgCl}_2$  (except for mitochondria HMG-CoA synthase), 0.5 mM DTT and enzyme varying in the range from 0.2 to 8 mg/ml. Control reactions were carried in the absence of enzyme and one of the substrates.

[0118] The progress of synthesis was followed by analyzing aliquots taken after increasing period of incubation at 30 or 37°C. Typically, an aliquot of 50  $\mu\text{l}$  was removed after 48 h of incubation, heated for 1 min at 100°C to eliminate the proteins, centrifuged and the supernatant was transferred to a clean vial for HIV detection by mass spectrometry. A solution of 3-hydroxy-3-methylbutyrate was prepared in 40 mM Tris-HCl pH 8, 1 mM  $\text{MgCl}_2$ , 0.5 mM DTT, heated as described early and used as reference.

**[0119]** The samples were analyzed on a PE SCIEX API 2000 triple quadrupole mass spectrometer in negative ion mode with H<sub>2</sub>O/acetonitrile=60/40 containing 0.1% triethylamine as mobile phase, flow rate was 40 µl/min. 10 µl of each supernatant were mixed with an equal quantity of mobile phase and directly injected into the mass spectrometer. The presence of [3-hydroxy-3-methylbutyrate-H]<sup>-</sup> ion was monitored. A peak corresponding to 3-hydroxy-3-methylbutyrate was observed for the following enzymes:

*Blattella germanica* (German cockroach) P54961 (SEQ ID NO: 6)

*Gallus gallus* (Chicken) P23228 (SEQ ID NO: 7)

*Homo sapiens* (Human) Q01581 (SEQ ID NO: 8)

*Arabidopsis thaliana* P54873 (CAA58763) (SEQ ID NO: 4)

*Caenorhabditis elegans* P54871 (SEQ ID NO: 1)

*Schizosaccharomyces pombe* (Fission yeast) P54874 (SEQ ID NO: 2)

*Saccharomyces cerevisiae* (Baker's yeast) P54839 (SEQ ID NO: 3)

*Dictyostelium discoideum* (Slime mold) Q86HL5 (SEQ ID NO: 10)

*Leuconostoc mesenteroides* Q03WZ0

*Staphylococcus epidermidis* Q8CN06 (SEQ ID NO: 11)

*Lactobacillus delbrueckii* Q1GAH5 (SEQ ID NO: 24)

*Staphylococcus haemolyticus* Q4L958 (198>V difference compared to wild type protein) (SEQ ID NO: 25)

**[0120]** Figures 6 to 8 show representative results for commercially available 3-hydroxy-3-methylbutyrate, for the reaction using the HMG CoA synthase from *Gallus gallus* (P23228) and for the control assay without enzyme.

#### **Example 6**

#### **3-hydroxy-3-methylbutyryl-CoA production using lyases (falling outside the scope of the claims)**

**[0121]** 3-hydroxy-3-methylbutyryl-CoA synthesis is carried out in the presence of radiolabeled [2-<sup>14</sup>C] acetone. The complete reaction for 3-hydroxy-3-methylbutyryl-CoA synthesis contains 40 mM Tris-HCl pH 8, 5 to 50 mM acetyl-CoA, 100 to 500 mM acetone, 1 to 10 mM MgCl<sub>2</sub>, 0.5 mM DTT and enzyme varying in the range from 0.5 to 7 mg/ml. The formation of product is analyzed after separation of reaction mixture by TLC or HPLC.

**[0122]** 3-hydroxy-3-methylbutyryl-CoA is also analyzed by TLC method (Stadtman E.R., J. Biol. Chem. 196 (1952), 535-546). An aliquot of reaction is deposited on a cellulose plate and chromatographed in the following solvent system: ethanol/0.1 M sodium acetate pH 4.5 (1/1). Co-A and acetyl-CoA are used as internal standards. R<sub>f</sub> reported for 3-hydroxy-3-methylbutyryl-CoA is 0.88.

#### **Example 7**

#### **Kinetic parameters for the enzymatic reaction between acetyl-CoA and acetone in the case of HMG synthases**

**[0123]** The kinetic parameters were measured using a variable concentration of acetone and a constant concentration of acetyl-CoA (10 mM) in following conditions:

40 mM Tris-HCl pH 8

2 mM MgCl<sub>2</sub>

0 - 1 M acetone

The final pH was adjusted to 8.

[0124] The reaction was initiated by the addition of 3 mg of purified enzyme to the 1 ml reaction mixture. The mixture was then incubated without shaking at 37°C for 40 h.

**Analysis of 3-hydroxy-3-methylbutyrate production**

[0125] Thermochemical conditions leading to the decomposition of 3-hydroxy-3-methylbutyrate into isobutene were applied (Pressman et al., JACS, 1940, 2069-2080): the pH of the reaction mixtures was first adjusted to pH 4 using 6N HCl and the samples were then transferred into gas chromatography vials (Interchim). The vials were sealed and incubated at 110°C for 4 hours, thus leading to the decomposition of 3-hydroxy-3-methylbutyrate into isobutene.

[0126] The calibration curve was prepared in the same conditions using commercial 3-hydroxy-3-methylbutyrate.

[0127] One milliliter of headspace gas was collected and injected into a HP5890 gas chromatograph (HP) equipped with a FID detector and a CP SilicaPlot column (Varian). Commercial isobutene was used as reference. From the isobutene signal the amount of 3-hydroxy-3-methylbutyrate initially present in the sample was calculated.

[0128] The kinetics parameters for several of the studied HMG-CoA synthases are presented in the following Table.

Organism	$K_M$ for acetone, mM	$k_{cat} \times 10^{-4}$ , Sec <sup>-1</sup>	$k_{cat} / K_M \times 10^{-6}$ , mM <sup>-1</sup> x sec <sup>-1</sup>
<i>Gallus gallus</i>	250	5	2
<i>Staphylococcus epidermidis</i>	200	0.6	0.3
<i>Schizosaccharomyces pombe</i>	200	0.2	0.1

[0129] For the enzyme from *S. epidermidis* Figure 9 shows a corresponding Michaelis-Menten plot.

**SEQUENCE LISTING**

[0130]

<110> MARLIÈRE, Philippe

<120> Method for the production of 3-hydroxy-3-methylbutyric acid from acetone and an activated acetyl compound

<130> R1889 PCT S3

<150> EP 09 17 0312.4

<151> 2009-09-15

<160> 25

<170> PatentIn version 3.5

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 35 40 45

Gly Gln Gln Gln Met Gly Phe Cys Ser Asp Asn Glu Asp Ile Val Ser  
 50 55 60

Ile Ser Leu Thr Val Thr Arg Lys Leu Ile Glu Thr Tyr Lys Ile Ser  
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Thr Asp Ser Ile Gly Cys Leu Val Val Gly Thr Glu Thr Met Ile Asp  
 85 90 95

Lys Ser Lys Ser Val Lys Thr Ala Leu Met Asp Leu Phe Pro Gly Asn  
 100 105 110

Ser Asp Ile Glu Gly Val Asp Ile Lys Asn Ala Cys Phe Gly Gly Ala  
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Gln Ala Leu Leu His Ala Ile Asp Trp Val Thr Val Asn His Pro Leu  
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Asp Lys Lys Asn Ala Ile Val Val Val Ala Asp Ile Ala Ile Tyr Glu  
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Glu Gly Pro Ala Arg Cys Thr Gly Gly Ala Gly Ala Ile Ala Phe Leu  
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Ile Cys Pro Asp Ala Ser Ile Pro Ile Asp Arg Gln Phe Ser Ala Cys  
 180 185 190

His Met Lys Asn Thr Trp Asp Phe Phe Lys Pro Ile Thr Pro Ile Pro  
 195 200 205

Ser Glu Tyr Pro Val Val Asp Gly Ser Leu Ser Leu Ser Ser Tyr Leu  
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Glu Ala Val Arg Met Thr Tyr Thr Tyr Phe Ile Ser Lys Val Asn Arg  
 225 230 235 240

His Thr Thr Gly Ile Asp Gly Leu Asn Ser Phe Asp Gly Val Phe Leu  
 245 250 255

His Ser Pro Phe Thr Lys Met Val Gln Lys Gly Leu Ala Val Met Asn  
 260 265 270

Tyr Thr Asp Ser Gln Leu Arg His Lys Gln Leu Asn Gly Asn Gly Val  
 275 280 285

Asp His Lys Leu Asp Glu Asn Asp Arg Ala Gly Leu Ala Lys Met Ile  
 290 295 300

Glu Leu Ser Ala Gln Val Trp Lys Glu Lys Thr Asp Pro Tyr Leu Val  
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Phe Asn Arg Arg Ile Gly Asn Met Tyr Thr Pro Ser Leu Phe Ala Gln  
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Leu Leu Ala Tyr Leu Ala Ala Asp Asp Cys Val Thr Gly Glu Lys Ser  
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Leu Glu Val Gly Thr Glu Thr Ile Ile Asp Lys Ser Lys Ser Val Lys  
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Asp Cys Val Asn Ala Cys Tyr Gly Gly Val Asn Ala Leu Phe Asn Thr  
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Gly Gly Ala Gly Cys Val Ala Leu Leu Val Gly Pro Asn Ala Pro Ile  
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Val Phe Glu Pro Gly Leu Arg Gly Thr Tyr Met Gln His Ala Tyr Asp  
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Phe Tyr Lys Pro Asp Leu Thr Ser Glu Tyr Pro Tyr Val Asp Gly His  
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Tyr Asn Val Arg Asp Val Ala Lys Asn Gly Lys Ser Gln Gly Leu Gly



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 195 200 205  
 Met Trp Ile Gly Pro Asp Ala Pro Ile Val Phe Asp Ser Val Arg Ala  
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 Ser Tyr Met Glu His Ala Tyr Asp Phe Tyr Lys Pro Asp Phe Thr Ser  
 225 230 235 240  
 Glu Tyr Pro Tyr Val Asp Gly His Phe Ser Leu Thr Cys Tyr Val Lys  
 245 250 255  
 Ala Leu Asp Gln Val Tyr Lys Ser Tyr Ser Lys Lys Ala Ile Ser Lys  
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Leu Phe Pro Glu Val Asp Ala Glu Leu Ala Thr Arg Asp Tyr Asp Glu  
 325 330 335

Ser Leu Thr Asp Lys Asn Ile Glu Lys Thr Phe Val Asn Val Ala Lys  
 340 345 350

Pro Phe His Lys Glu Arg Val Ala Gln Ser Leu Ile Val Pro Thr Asn  
 355 360 365

Thr Gly Asn Met Tyr Thr Ala Ser Val Tyr Ala Ala Phe Ala Ser Leu  
 370 375 380

Leu Asn Tyr Val Gly Ser Asp Asp Leu Gln Gly Lys Arg Val Gly Leu  
 385 390 395 400

Phe Ser Tyr Gly Ser Gly Leu Ala Ala Ser Leu Tyr Ser Cys Lys Ile  
 405 410 415

Val Gly Asp Val Gln His Ile Ile Lys Glu Leu Asp Ile Thr Asn Lys  
 420 425 430

Leu Ala Lys Arg Ile Thr Glu Thr Pro Lys Asp Tyr Glu Ala Ala Ile  
 435 440 445

Glu Leu Arg Glu Asn Ala His Leu Lys Lys Asn Phe Lys Pro Gln Gly  
 450 455 460

Ser Ile Glu His Leu Gln Ser Gly Val Tyr Tyr Leu Thr Asn Ile Asp  
 465 470 475 480

Asp Lys Phe Arg Arg Ser Tyr Asp Val Lys Lys  
 485 490

<210> 4

<211> 461

<212> PRT

<213> Arabidopsis thaliana

<400> 4

Met Ala Lys Asn Val Gly Ile Leu Ala Met Asp Ile Tyr Phe Pro Pro  
 1 5 10 15

Thr Cys Val Gln Gln Glu Ala Leu Glu Ala His Asp Gly Ala Ser Lys  
 20 25 30

Gly Lys Tyr Thr Ile Gly Leu Gly Gln Asp Cys Leu Ala Phe Cys Thr  
 35 40 45

Glu Leu Glu Asp Val Ile Ser Met Ser Phe Asn Ala Val Thr Ser Leu  
 50 55 60  
 Phe Glu Lys Tyr Lys Ile Asp Pro Asn Gln Ile Gly Arg Leu Glu Val  
 65 70 75 80  
 Gly Ser Glu Thr Val Ile Asp Lys Ser Lys Ser Ile Lys Thr Phe Leu  
 85 90 95  
 Met Gln Leu Phe Glu Lys Cys Gly Asn Thr Asp Val Glu Gly Val Asp  
 100 105 110  
 Ser Thr Asn Ala Cys Tyr Gly Gly Thr Ala Ala Leu Leu Asn Cys Val  
 115 120 125  
 Asn Trp Val Glu Ser Asn Ser Trp Asp Gly Arg Tyr Gly Leu Val Ile  
 130 135 140  
 Cys Thr Asp Ser Ala Val Tyr Ala Glu Gly Pro Ala Arg Pro Thr Gly  
 145 150 155 160  
 Gly Ala Ala Ala Ile Ala Met Leu Ile Gly Pro Asp Ala Pro Ile Val  
 165 170 175  
 Phe Glu Ser Lys Leu Arg Ala Ser His Met Ala His Val Tyr Asp Phe  
 180 185 190  
 Tyr Lys Pro Asn Leu Ala Ser Glu Tyr Pro Val Val Asp Gly Lys Leu  
 195 200 205  
 Ser Gln Thr Cys Tyr Leu Met Ala Leu Asp Ser Cys Tyr Lys His Leu  
 210 215 220  
 Cys Asn Lys Phe Glu Lys Ile Glu Gly Lys Glu Phe Ser Ile Asn Asp  
 225 230 235 240  
 Ala Asp Tyr Ile Val Phe His Ser Pro Tyr Asn Lys Leu Val Gln Lys  
 245 250 255  
 Ser Phe Ala Arg Leu Leu Tyr Asn Asp Phe Leu Arg Asn Ala Ser Ser  
 260 265 270  
 Ile Asp Glu Ala Ala Lys Glu Lys Phe Thr Pro Tyr Ser Ser Leu Thr  
 275 280 285  
 Leu Asp Glu Ser Tyr Gln Ser Arg Asp Leu Glu Lys Val Ser Gln Gln  
 290 295 300  
 Ile Ser Lys Pro Phe Tyr Asp Ala Lys Val Gln Pro Thr Thr Leu Ile  
 305 310 315 320  
 Pro Lys Glu Val Gly Asn Met Tyr Thr Ala Ser Leu Tyr Ala Ala Phe  
 325 330 335  
 Ala Ser Leu Ile His Asn Lys His Asn Asp Leu Ala Gly Lys Arg Val  
 340 345 350  
 Val Met Phe Ser Tyr Gly Ser Gly Ser Thr Ala Thr Met Phe Ser Leu  
 355 360 365  
 Arg Leu Asn Asp Asn Lys Pro Pro Phe Ser Ile Ser Asn Ile Ala Ser  
 370 375 380  
 Val Met Asp Val Gly Gly Lys Leu Lys Ala Arg His Glu Tyr Ala Pro  
 385 390 395 400  
 Glu Lys Phe Val Glu Thr Met Lys Leu Met Glu His Arg Tyr Gly Ala  
 405 410 415  
 Lys Asp Phe Val Thr Thr Lys Glu Gly Ile Ile Asp Leu Leu Ala Pro  
 420 425 430  
 Gly Thr Tyr Tyr Leu Lys Glu Val Asp Ser Leu Tyr Arg Arg Phe Tyr  
 435 440 445  
 Gly Lys Lys Gly Glu Asp Gly Ser Val Ala Asn Gly His  
 450 455 460

<210> 5

<211> 482

<212> PRT

<213> Dictyostelium discoideum

<400> 5

Met Thr Lys Pro Glu Asn Ile Gly Ile His Gly Ile Glu Val Tyr Phe  
 1 5 10 15  
 Pro Ser Thr Tyr Val Ala Gln Glu Asp Leu Glu Lys Phe Asp Gly Val  
 20 25 30  
 Ser Gln Gly Lys Tyr Thr Leu Gly Leu Gly Gln Thr Asn Met Ala Phe  
 35 40 45  
 Cys Gly Asp Arg Glu Asp Ile Tyr Ser Leu Ser Leu Asn Ala Val Asn  
 50 55 60  
 Asn Leu Met Asp Lys Phe Asn Val Asp Pro Asn Ser Ile Gly Arg Leu  
 65 70 75 80  
 Glu Val Gly Thr Glu Thr Val Ile Asp Lys Ser Lys Ser Val Lys Thr  
 85 90 95  
 Val Leu Met Asp Leu Phe Ala Lys His Gly Asn Thr Ser Ile Asp Gly  
 100 105 110  
 Ile Asp Thr Ile Asn Ala Cys Tyr Gly Gly Thr Ser Ala Leu His Asn  
 115 120 125  
 Ala Leu Gln Trp Met Glu Ser Ser Tyr Trp Asp Gly Arg Asn Ala Ile  
 130 135 140  
 Val Val Ala Gly Asp Ile Ala Val Tyr Glu Lys Gly Pro Ala Arg Pro  
 145 150 155 160  
 Thr Gly Gly Ala Gly Val Val Ala Met Leu Ile Gly Pro Asn Ala Pro  
 165 170 175  
 Ile Thr Phe Glu Ser Gly Leu Arg Gly Val His Met Glu Asn Val Tyr  
 180 185 190  
 Asp Phe Tyr Lys Pro Asp Met Asp Ser Glu Tyr Pro Arg Val Asp Gly  
 195 200 205  
 Lys Leu Ser Ile Ser Cys Tyr Phe Arg Ala Ile Asp Asn Cys Tyr Asn  
 210 215 220  
 Arg Tyr Ala Lys Ala Phe Glu Lys Lys Tyr Gly Lys Ser Phe Ser Leu  
 225 230 235 240  
 Asp Gln Val Asp Phe Ala Leu Phe His Ser Pro Tyr Asn Lys Leu Val  
 245 250 255  
 Gln Lys Ser Phe Gly Arg Met Leu Tyr Asn Asp Phe Leu Asn Asn Pro  
 260 265 270  
 Asn Asp Ser Arg Tyr Ala Ser Leu Glu Ala Tyr Lys Asn Val Lys Pro  
 275 280 285  
 Glu Asp Thr Tyr Phe Asp Ser Val Leu Glu Lys Ala Leu Ser Ala Ile  
 290 295 300  
 Thr Lys Asn Asp Tyr Ala Thr Lys Val Ala Pro Thr Thr Leu Leu Ala  
 305 310 315 320  
 Lys Gln Leu Gly Asn Thr Tyr Cys Gly Ser Thr Tyr Ser Gly Leu Leu  
 325 330 335  
 Ser Leu Leu Asp Glu Lys Ser Asn Asp Leu Val Gly Lys Arg Val Leu  
 340 345 350  
 Thr Phe Ser Tyr Gly Ser Gly Leu Ala Ala Ser Ala Phe Ser Phe Lys  
 355 360 365

Val Glu Lys Pro Ile Asn His Ile Val Glu Lys Val Asp Leu Lys Asn  
370 375 380

Arg Leu Ala Lys Arg Val Arg Val Glu Pro Glu Ile Phe Thr Glu Lys  
385 390 395 400

Leu Ser Leu Arg Glu Thr Arg His Asn Leu Lys Asn Tyr Val Pro Ser  
405 410 415

Asp Glu Thr Thr Asn Met Phe Pro Gly Ser Phe Tyr Leu Ser Ser Val  
420 425 430

Asp Asn Ala Gly Ile Arg Lys Tyr Asp Arg Thr Tyr Ser Thr Ser Ala  
435 440 445

Val Leu Gly Ala Phe Gln Arg Arg Gln Gln Ile Ser Gln Ser Thr Ile  
450 455 460

Lys Ser Leu Asn Leu Phe Arg Ala Thr Lys Ser Val Leu Ser Ile Leu  
465 470 475 480

Lys Lys

<210> 6

<211> 453

<212> PRT

<213> Blattella germanica

<400> 6

Met Trp Pro Ser Asp Val Gly Ile Val Ala Leu Glu Leu Ile Phe Pro  
1 5 10 15

Ser Gln Tyr Val Asp Gln Val Asp Leu Glu Val Tyr Asp Asn Val Ser  
20 25 30

Ala Gly Lys Tyr Thr Val Gly Leu Gly Gln Ala Arg Met Gly Phe Cys  
35 40 45

Thr Asp Arg Glu Asp Ile Asn Ser Leu Cys Leu Thr Val Val Ser Arg  
50 55 60

Leu Met Glu Arg Trp Ser Ile Pro Tyr Ser Gln Ile Gly Arg Leu Glu  
65 70 75 80

Val Gly Thr Glu Thr Leu Leu Asp Lys Ser Lys Ser Val Lys Thr Val  
85 90 95

Leu Met Gln Leu Phe Lys Asp Asn Thr Asp Ile Glu Gly Val Asp Thr  
100 105 110

Val Asn Ala Cys Tyr Gly Gly Thr Ser Ala Leu Phe Asn Ala Ile Ser



Met Pro Gly Ser Leu Pro Val Asn Thr Glu Ser Cys Trp Pro Lys Asp  
 1 5 10 15

Val Gly Ile Val Ala Leu Glu Ile Tyr Phe Pro Ser Gln Tyr Val Asp  
 20 25 30

Gln Thr Glu Leu Glu Lys Tyr Asp Gly Val Asp Ala Gly Lys Tyr Thr  
 35 40 45

Ile Gly Leu Gly Gln Ser Lys Met Gly Phe Cys Ser Asp Arg Glu Asp  
 50 55 60

Ile Asn Ser Leu Cys Leu Thr Val Val Gln Lys Leu Met Glu Arg Asn  
 65 70 75 80

Ser Leu Ser Tyr Asp Cys Ile Gly Arg Leu Glu Val Gly Thr Glu Thr  
 85 90 95

Ile Ile Asp Lys Ser Lys Ser Val Lys Thr Val Leu Met Gln Leu Phe  
 100 105 110

Glu Glu Ser Gly Asn Thr Asp Val Glu Gly Ile Asp Thr Thr Asn Ala  
 115 120 125

Cys Tyr Gly Gly Thr Ala Ala Leu Phe Asn Ala Ile Asn Trp Ile Glu  
 130 135 140

Ser Ser Ser Trp Asp Gly Arg Tyr Ala Leu Val Val Ala Gly Asp Ile  
 145 150 155 160

Ala Val Tyr Ala Thr Gly Asn Ala Arg Pro Thr Gly Gly Ala Gly Ala  
 165 170 175

Val Ala Met Leu Val Gly Ser Asn Ala Pro Leu Ile Phe Glu Arg Gly  
 180 185 190

Leu Arg Gly Thr His Met Gln His Ala Tyr Asp Phe Tyr Lys Pro Asp  
 195 200 205

Met Val Ser Glu Tyr Pro Val Val Asp Gly Lys Leu Ser Ile Gln Cys  
 210 215 220

Tyr Leu Ser Ala Leu Asp Arg Cys Tyr Ser Val Tyr Arg Asn Lys Ile  
 225 230 235 240

His Ala Gln Trp Gln Lys Glu Gly Thr Asp Arg Gly Phe Thr Leu Asn  
 245 250 255

Asp Phe Gly Phe Met Ile Phe His Ser Pro Tyr Cys Lys Leu Val Gln  
 260 265 270

Lys Ser Val Ala Arg Leu Leu Leu Asn Asp Phe Leu Ser Asp Gln Asn  
 275 280 285

Ala Glu Thr Ala Asn Gly Val Phe Ser Gly Leu Glu Ala Phe Arg Asp  
 290 295 300

Val Lys Leu Glu Asp Thr Tyr Phe Asp Arg Asp Val Glu Lys Ala Phe  
 305 310 315 320

Met Lys Ala Ser Ala Glu Leu Phe Asn Gln Lys Thr Lys Ala Ser Leu  
 325 330 335

Leu Val Ser Asn Gln Asn Gly Asn Met Tyr Thr Pro Ser Val Tyr Gly  
 340 345 350

Cys Leu Ala Ser Leu Leu Ala Gln Tyr Ser Pro Glu His Leu Ala Gly  
 355 360 365

Gln Arg Ile Ser Glu Phe Ser Tyr Gly Ser Gly Phe Ala Ala Thr Leu  
 370 375 380

Tyr Ser Ile Arg Val Thr Gln Asp Ala Thr Pro Gly Ser Ala Leu Asp  
 385 390 395 400

Lys Ile Thr Ala Ser Leu Ser Asp Leu Lys Ala Arg Leu Asp Ser Arg  
 405 410 415

Lys Cys Ile Ala Pro Asp Val Phe Ala Glu Asn Met Lys Ile Arg Gln  
 420 425 430

Glu Thr His His Leu Ala Asn Tyr Ile Pro Gln Cys Ser Val Glu Asp  
 435 440 445

Leu Phe Glu Gly Thr Trp Tyr Leu Val Arg Val Asp Glu Lys His Arg  
 450 455 460

Arg Thr Tyr Ala Arg Arg Pro Val Met Gly Asp Gly Pro Leu Glu Ala  
 465 470 475 480

Gly Val Glu Val Val His Pro Gly Ile Val His Glu His Ile Pro Ser  
 485 490 495

Pro Ala Lys Lys Val Pro Arg Ile Pro Ala Thr Thr Glu Ser Glu Gly  
 500 505 510

Val Thr Val Ala Ile Ser Asn Gly Val His  
 515 520

<210> 8

<211> 520

<212> PRT

<213> Homo sapiens

<400> 8

Met Pro Gly Ser Leu Pro Leu Asn Ala Glu Ala Cys Trp Pro Lys Asp  
 1 5 10 15

Val Gly Ile Val Ala Leu Glu Ile Tyr Phe Pro Ser Gln Tyr Val Asp  
 20 25 30

Gln Ala Glu Leu Glu Lys Tyr Asp Gly Val Asp Ala Gly Lys Tyr Thr  
 35 40 45

Ile Gly Leu Gly Gln Ala Lys Met Gly Phe Cys Thr Asp Arg Glu Asp  
 50 55 60

Ile Asn Ser Leu Cys Met Thr Val Val Gln Asn Leu Met Glu Arg Asn  
 65 70 75 80

Asn Leu Ser Tyr Asp Cys Ile Gly Arg Leu Glu Val Gly Thr Glu Thr  
 85 90 95

Ile Ile Asp Lys Ser Lys Ser Val Lys Thr Asn Leu Met Gln Leu Phe  
 100 105 110

Glu Glu Ser Gly Asn Thr Asp Ile Glu Gly Ile Asp Thr Thr Asn Ala  
 115 120 125

Cys Tyr Gly Gly Thr Ala Ala Val Phe Asn Ala Val Asn Trp Ile Glu  
 130 135 140

Ser Ser Ser Trp Asp Gly Arg Tyr Ala Leu Val Val Ala Gly Asp Ile  
 145 150 155 160

Ala Val Tyr Ala Thr Gly Asn Ala Arg Pro Thr Gly Gly Val Gly Ala  
 165 170 175

Val Ala Leu Leu Ile Gly Pro Asn Ala Pro Leu Ile Phe Glu Arg Gly  
 180 185 190

Leu Arg Gly Thr His Met Gln His Ala Tyr Asp Phe Tyr Lys Pro Asp  
 195 200 205

Met Leu Ser Glu Tyr Pro Ile Val Asp Gly Lys Leu Ser Ile Gln Cys  
 210 215 220

Tyr Leu Ser Ala Leu Asp Arg Cys Tyr Ser Val Tyr Cys Lys Lys Ile  
 225 230 235 240

His Ala Gln Trp Gln Lys Glu Gly Asn Asp Lys Asp Phe Thr Leu Asn  
 245 250 255

Asp Phe Gly Phe Met Ile Phe His Ser Pro Tyr Cys Lys Leu Val Gln  
 260 265 270

Lys Ser Leu Ala Arg Met Leu Leu Asn Asp Phe Leu Asn Asp Gln Asn  
 275 280 285

Arg Asp Lys Asn Ser Ile Tyr Ser Gly Leu Glu Ala Phe Gly Asp Val  
 290 295 300

Lys Leu Glu Asp Thr Tyr Phe Asp Arg Asp Val Glu Lys Ala Phe Met  
 305 310 315 320

Lys Ala Ser Ser Glu Leu Phe Ser Gln Lys Thr Lys Ala Ser Leu Leu  
 325 330 335

Val Ser Asn Gln Asn Gly Asn Met Tyr Thr Ser Ser Val Tyr Gly Ser  
 340 345 350

Leu Ala Ser Val Leu Ala Gln Tyr Ser Pro Gln Gln Leu Ala Gly Lys  
 355 360 365

Arg Ile Gly Val Phe Ser Tyr Gly Ser Gly Leu Ala Ala Thr Leu Tyr  
 370 375 380

Ser Leu Lys Val Thr Gln Asp Ala Thr Pro Gly Ser Ala Leu Asp Lys  
 385 390 395 400

Ile Thr Ala Ser Leu Cys Asp Leu Lys Ser Arg Leu Asp Ser Arg Thr  
 405 410 415

Gly Val Ala Pro Asp Val Phe Ala Glu Asn Met Lys Leu Arg Glu Asp  
 420 425 430

Thr His His Leu Val Asn Tyr Ile Pro Gln Gly Ser Ile Asp Ser Leu  
 435 440 445

Phe Glu Gly Thr Trp Tyr Leu Val Arg Val Asp Glu Lys His Arg Arg  
 450 455 460

Thr Tyr Ala Arg Arg Pro Thr Pro Asn Asp Asp Thr Leu Asp Glu Gly  
 465 470 475 480

Val Gly Leu Val His Ser Asn Ile Ala Thr Glu His Ile Pro Ser Pro  
 485 490 495

Ala Lys Lys Val Pro Arg Leu Pro Ala Thr Ala Ala Glu Pro Glu Ala  
 500 505 510

Ala Val Ile Ser Asn Gly Val Trp  
 515 520

<210> 9

<211> 508

<212> PRT

<213> Homo sapiens

<400> 9

Met Gln Arg Leu Leu Thr Pro Val Lys Arg Ile Leu Gln Leu Thr Arg  
 1 5 10 15

Ala Val Gln Glu Thr Ser Leu Thr Pro Ala Arg Leu Leu Pro Val Ala  
 20 25 30

His Gln Arg Phe Ser Thr Ala Ser Ala Val Pro Leu Ala Lys Thr Asp  
 35 40 45

Thr Trp Pro Lys Asp Val Gly Ile Leu Ala Leu Glu Val Tyr Phe Pro  
 50 55 60

Ala Gln Tyr Val Asp Gln Thr Asp Leu Glu Lys Tyr Asn Asn Val Glu  
 65 70 75 80

Ala Gly Lys Tyr Thr Val Gly Leu Gly Gln Thr Arg Met Gly Phe Cys  
 85 90 95

Ser Val Gln Glu Asp Ile Asn Ser Leu Cys Leu Thr Val Val Gln Arg  
 100 105 110

Leu Met Glu Arg Ile Gln Leu Pro Trp Asp Ser Val Gly Arg Leu Glu  
 115 120 125

Val Gly Thr Glu Thr Ile Ile Asp Lys Ser Lys Ala Val Lys Thr Val  
 130 135 140

Leu Met Glu Leu Phe Gln Asp Ser Gly Asn Thr Asp Ile Glu Gly Ile  
 145 150 155 160

Asp Thr Thr Asn Ala Cys Tyr Gly Gly Thr Ala Ser Leu Phe Asn Ala  
 165 170 175

Ala Asn Trp Met Glu Ser Ser Ser Trp Asp Gly Arg Tyr Ala Met Val  
 180 185 190

Val Cys Gly Asp Ile Ala Val Tyr Pro Ser Gly Asn Ala Arg Pro Thr  
 195 200 205

Gly Gly Ala Gly Ala Val Ala Met Leu Ile Gly Pro Lys Ala Pro Leu  
 210 215

Ala Leu Glu Arg Gly Leu Arg Gly Thr His Met Glu Asn Val Tyr Asp  
 225 230 235 240

Phe Tyr Lys Pro Asn Leu Ala Ser Glu Tyr Pro Ile Val Asp Gly Lys  
 245 250 255

Leu Ser Ile Gln Cys Tyr Leu Arg Ala Leu Asp Arg Cys Tyr Thr Ser  
 260 265 270

Tyr Arg Lys Lys Ile Gln Asn Gln Trp Lys Gln Ala Gly Ser Asp Arg  
 275 280 285

Pro Phe Thr Leu Asp Asp Leu Gln Tyr Met Ile Phe His Thr Pro Phe  
 290 295 300

Cys Lys Met Val Gln Lys Ser Leu Ala Arg Leu Met Phe Asn Asp Phe  
 305 310 315 320

Leu Ser Ala Ser Ser Asp Thr Gln Thr Ser Leu Tyr Lys Gly Leu Glu  
 325 330 335

Ala Phe Gly Gly Leu Lys Leu Glu Asp Thr Tyr Thr Asn Lys Asp Leu  
 340 345 350

Asp Lys Ala Leu Leu Lys Ala Ser Gln Asp Met Phe Asp Lys Lys Thr  
 355 360 365

Lys Ala Ser Leu Tyr Leu Ser Thr His Asn Gly Asn Met Tyr Thr Ser  
 370 375 380

Ser Leu Tyr Gly Cys Leu Ala Ser Leu Leu Ser His His Ser Ala Gln  
 385 390 395 400

Glu Leu Ala Gly Ser Arg Ile Gly Ala Phe Ser Tyr Gly Ser Gly Leu  
 405 410 415

Ala Ala Ser Phe Phe Ser Phe Arg Val Ser Gln Asp Ala Ala Pro Gly  
 420 425 430

Ser Pro Leu Asp Lys Leu Val Ser Ser Thr Ser Asp Leu Pro Lys Arg  
 435 440 445

Leu Ala Ser Arg Lys Cys Val Ser Pro Glu Glu Phe Thr Glu Ile Met  
 450 455 460

Asn Gln Arg Glu Gln Phe Tyr His Lys Val Asn Phe Ser Pro Pro Gly  
 465 470 475 480

Asp Thr Asn Ser Leu Phe Pro Gly Thr Trp Tyr Leu Glu Arg Val Asp  
 485 490 495

Glu Gln His Arg Arg Lys Tyr Ala Arg Arg Pro Val  
 500 505

<210> 10

<211> 468

<212> PRT

<213> Dictyostelium discoideum

<400> 10

Met Lys Lys Thr Lys Asp Ile Gly Ile Cys Ala Ile Asp Ile Tyr Phe  
 1 5 10 15

Pro Gln Thr Tyr Val Asn Gln Ser Glu Leu Lys Lys Tyr Asp Lys Val  
 20 25 30

Ser Asn Gly Lys Tyr Thr Ile Gly Leu Gly Gln Thr Asn Met Ser Phe  
 35 40 45

Val Gly Asp Arg Glu Asp Ile Val Ser Met Ala Met Thr Ser Val Lys  
 50 55 60

Met Met Met Ser Lys Tyr Ser Ile Asp Tyr Gln Ser Ile Gly Arg Leu  
 65 70 75 80

Glu Val Gly Thr Glu Thr Ile Ile Asp Lys Ser Lys Ser Val Lys Ser  
 85 90 95

Ser Ile Met Ser Leu Phe Gln Glu Tyr Gly Asn Thr Ser Leu Glu Gly  
 100 105 110

Val Asp Thr Leu Asn Ala Cys Tyr Gly Gly Thr Asn Ala Leu Phe Asn  
 115 120 125

Ser Leu Gln Trp Ile Glu Ser Ser Tyr Trp Asp Gly Arg Tyr Ala Leu  
 130 135 140

Val Val Thr Gly Asp Ile Ala Val Tyr Ser Lys Gly Ala Ala Arg Pro  
 145 150 155 160

Thr Gly Gly Ala Gly Val Val Thr Met Leu Ile Gly Pro Asn Ala Thr  
 165 170 175

Leu Ile Phe Asp Gln Ser Leu Arg Gly Thr His Met Glu Asn Val Asn  
 180 185 190

Asp Phe Tyr Lys Pro Asp Leu Ser Ser Glu Tyr Pro Tyr Val Asp Gly  
 195 200 205

Lys Leu Ser Ile Glu Cys Tyr Leu Arg Ala Leu Asp Lys Cys Tyr Leu  
 210 215 220

Glu Tyr Lys Lys Lys Phe Glu Ser Ile Asn Asp Asp Asn Lys Phe Ser  
 225 230 235 240

Met Asp Ser Phe Asp Tyr Val Cys Phe His Ser Pro Tyr Asn Arg Leu  
 245 250 255

Val Gln Lys Ser Tyr Ala Arg Leu Ile Tyr Asn Asp Phe Leu Gln Asn  
 260 265 270

Pro Asn Asn Pro Lys Tyr Gln Asp Leu Leu Pro Phe Lys Asp Leu Ser  
 275 280 285

Thr Gly Lys Asp Ser Tyr Ile Asn Ser Lys Leu Asp Gln Ile Thr Leu  
 290 295 300

Lys Leu Ser Leu Asp Asp Phe Lys Thr Lys Val Asn Pro Ser Thr Leu  
 305 310 315 320

Leu Ser Lys Glu Cys Gly Asn Ser Tyr Cys Gly Ser Val Tyr Ser Gly  
 325 330 335

Ile Leu Ser Leu Leu Ser Asn Val Asn Asp Leu Asn Asn Lys Lys Val  
 340 345 350

Leu Val Phe Ser Tyr Gly Ser Gly Leu Ala Ala Ser Leu Phe Ser Phe  
 355 360 365

Arg Ile Asn Asn Asn Lys Asn Arg Asn Asn Asn Asn Asn Asn Asn  
 370 375 380

Cys Phe Phe Lys Thr Thr Asn Asp Ile Gly Lys Ile Ser Asn Ile Lys  
 385 390 395 400

Glu Arg Leu Ser Asn Arg Val Lys Val Ser Pro Glu Glu Phe Thr Arg  
 405 410 415

Ile Leu Asp Ile Arg Glu Lys Ser His Gln Met Val Gly Ala Arg Thr



Tyr Pro Leu Val Ala Gly Ala Leu Ser Lys Asp Ala Tyr Ile Lys Ser  
 195 200 205

Phe Gln Glu Ser Trp Asn Glu Tyr Ala Arg Arg His Asn Lys Thr Leu  
 210 215 220

Ala Asp Phe Ala Ser Leu Cys Phe His Val Pro Phe Thr Lys Met Gly  
 225 230 235 240

Gln Lys Ala Leu Asp Ser Ile Ile Asn His Ala Asp Glu Thr Thr Gln  
 245 250 255

Asp Arg Leu Asn Ser Ser Tyr Gln Asp Ala Val Asp Tyr Asn Arg Tyr  
 260 265 270

Val Gly Asn Ile Tyr Thr Gly Ser Leu Tyr Leu Ser Leu Ile Ser Leu  
 275 280 285

Leu Glu Thr Arg Asp Leu Lys Gly Gly Gln Thr Ile Gly Leu Phe Ser  
 290 295 300

Tyr Gly Ser Gly Ser Val Gly Glu Phe Phe Ser Gly Thr Leu Val Asp  
 305 310 315 320

Gly Phe Lys Glu Gln Leu Asp Val Glu Arg His Lys Ser Leu Leu Asn  
 325 330 335

Asn Arg Ile Glu Val Ser Val Asp Glu Tyr Glu His Phe Phe Lys Arg  
 340 345 350

Phe Asp Gln Leu Glu Leu Asn His Glu Leu Glu Lys Ser Asn Ala Asp  
 355 360 365

Arg Asp Ile Phe Tyr Leu Lys Ser Ile Asp Asn Asn Ile Arg Glu Tyr  
 370 375 380

His Ile Ala Glu  
 385

<210> 12

<211> 389

<212> PRT

<213> Lactobacillus fermentum

<400> 12

Met Lys Ile Gly Ile Asp Lys Leu Ala Phe Ala Thr Thr Pro Tyr Tyr  
 1 5 10 15

Leu Ala Met Glu Asp Leu Ala Gln Gly Arg Asn Val Asp Pro Asn Lys  
 20 25 30

Tyr Leu Ile Gly Ile Gly Gln Ser Lys Gln Ala Val Val Pro Pro Thr  
 35 40 45  
 Gln Asp Val Val Thr Leu Ala Ala Ala Ala Ala Asp Lys Leu Leu Asp  
 50 55 60  
 Pro Val Glu Arg Asp Gln Val Ser Thr Val Ile Val Ala Thr Glu Ser  
 65 70 75 80  
 Gly Ile Asp Asn Ser Lys Ala Ala Ala Val Tyr Val Lys His Leu Leu  
 85 90 95  
 Lys Leu Ser Asp Phe Thr Arg Ala Val Glu Val Lys Glu Ala Cys Tyr  
 100 105 110  
 Ser Ala Thr Ala Ala Leu Gln Phe Ala Arg Gly Leu Val Ala Leu Asn  
 115 120 125  
 Pro Gln Glu Lys Ile Leu Val Ile Ala Ser Asp Ile Ala Arg Tyr Gly  
 130 135 140  
 Leu Glu Thr Gly Gly Glu Val Thr Gln Gly Ala Gly Ala Val Ala Met  
 145 150 155 160  
 Leu Ile Thr Ala Asn Pro Arg Val Leu Ala Ile Glu Pro Thr Ser Val  
 165 170 175  
 Ala Tyr Thr Lys Asp Val Met Asp Phe Trp Arg Pro Leu Tyr Ala Glu  
 180 185 190  
 Glu Ala Leu Val Asn Gly Lys Tyr Ser Thr Asn Val Tyr Ile Asp Phe  
 195 200 205  
 Phe Lys Gln Cys Trp Thr Arg Tyr Gln Gln Leu Ala Gly Tyr Gly Leu  
 210 215 220  
 Glu Asp Phe Ala Ala Leu Ala Phe His Leu Pro Phe Thr Lys Met Gly  
 225 230 235 240  
 Lys Lys Ala Leu Glu Ala Glu Leu Gly Asp Arg Asp Asp Gln Val Ala  
 245 250 255  
 Thr Arg Leu Arg Ala Asn Leu Thr Ala Gly Gln Glu Ala Cys Arg Gln  
 260 265 270  
 Val Gly Asn Leu Tyr Thr Gly Ser Leu Tyr Leu Gly Leu Met Ser Leu  
 275 280 285  
 Leu Thr Glu Gly Asp Val Lys Pro Gly Glu Arg Ile Gly Leu Phe Ser  
 290 295 300  
 Tyr Gly Ser Gly Ala Glu Gly Glu Phe Phe Ala Gly Ile Leu Gln Pro  
 305 310 315 320  
 Gly Tyr Gln Glu Gly Leu Gly Asp Leu Asn Glu Gln Leu Ala Ala Arg  
 325 330 335  
 Thr Gln Val Ser Leu Ala Glu Tyr Glu Asp Leu Phe Asn Gln Gln Leu  
 340 345 350  
 Gly Leu Lys Glu Glu Asp Val Thr Phe Lys Thr Pro Ala Ala Gly Gln  
 355 360 365  
 Arg Phe Val Leu Val Gly Gln Lys Asp His Gln Arg Gln Tyr Arg Asp  
 370 375 380

Leu Ala Glu Arg Asp  
385

<210> 13

<211> 351

<212> PRT

<213> Hyperthermus butylicus

<400> 13

Met Pro Arg Gly Ser Gly Ile Val Gly Trp Gly Gly Tyr Val Pro Arg  
 1 5 10 15  
 Tyr Arg Ile Lys Ala Ala Glu Ile Val Arg Val Trp Gly Trp Glu Pro  
 20 25 30  
 Ser Val Pro Ala Gly Leu Gly Val Lys Glu Lys Ala Val Glu Asn Val  
 35 40 45  
 Asp Glu Asp Ser Val Thr Met Gly Tyr Glu Ala Ala Arg Asn Ala Ile  
 50 55 60  
 Ala Arg Ala Asn Val Asp Pro Arg Glu Ile Lys Ala Val Phe Phe Gly  
 65 70 75 80  
 Thr Glu Ser Lys Pro Tyr Ala Val Lys Pro Ser Ala Thr Ile Ile Ala  
 85 90 95  
 Glu Ala Leu Gly Ile Thr Pro Glu Thr Met Ala Ser Asp Leu Glu Phe  
 100 105 110  
 Ala Cys Arg Ala Ala Ser Glu Gly Leu Arg Ala Ser Leu Ala Leu Val  
 115 120 125  
 Glu Ala Gly Tyr Met Lys Tyr Ala Leu Val Val Ala Ser Asp Thr Ala  
 130 135 140  
 Gln Ala Asn Pro Gly Asp Val Leu Glu Phe Thr Ala Ala Ser Gly Ala  
 145 150 155 160  
 Ala Ala Phe Val Val Gly Pro Ala Ser Glu Ser Val Ala Val Leu Glu  
 165 170 175  
 Gly Val Tyr Thr Tyr Val Thr Asp Thr Pro Asp Phe Trp Arg Gly Gln  
 180 185 190  
 His Ser Arg Tyr Pro Met His Gly Glu Ala Phe Thr Gly Glu Pro Ala  
 195 200 205  
 Tyr Phe His His Ile Glu Ser Ala Val Lys Gly Leu Met Glu Lys Leu  
 210 215 220  
 Gly Leu Lys Pro Glu Asp Phe Asp Tyr Ala Val Phe His Gln Pro Asn  
 225 230 235 240  
 Gly Lys Phe Pro Leu Arg Val Gly Ala Arg Leu Gly Phe Pro Lys Glu  
 245 250 255  
 Lys Ile Leu Pro Gly Leu Leu Thr Pro Ile Ile Gly Asn Thr Tyr Asn  
 260 265 270  
 Ala Ser Ala Leu Leu Gly Phe Ala Arg Ile Leu Asp Gln Ala Lys Pro  
 275 280 285  
 Gly Gln Arg Ile Leu Val Ala Pro Phe Gly Ser Gly Ala Gly Ser Asp  
 290 295 300  
 Ala Tyr Ser Phe Ile Val Thr Asp Arg Ile Glu Glu Ala Arg Asn Arg  
 305 310 315 320  
 Ala Pro Lys Val Asp Asp Tyr Val Asn Trp Lys Arg Tyr Ile Asp Tyr  
 325 330 335  
 Ala Met His Ala Arg Met Arg Lys Leu Tyr Asp Arg Arg Pro Val  
 340 345 350

<210> 14

<211> 349

<212> PRT

<213> Chloroflexus aggregans

<400> 14

Met Met Lys Pro Asn Gln Pro Val Gly Ile Ile Gly Tyr Gly Val Tyr  
 1 5 10 15  
 Ile Pro Arg Tyr Arg Ile Ala Ala Arg Glu Ile Ala Arg Ile Trp Thr  
 20 25 30  
 Asp Gly Gln Asn Gly Val Pro Val Glu Ala Lys Ser Val Pro Gly Pro  
 35 40 45  
 Asp Glu Asp Thr Ile Thr Met Ala Ile Glu Ala Ala Arg Asn Ala Leu  
 50 55 60  
 Val Arg Ala Asp Ile Pro Ala Ser Ala Leu Gly Ala Val Trp Ile Gly  
 65 70 75 80  
 Ser Glu Ser His Pro Tyr Ser Val Lys Pro Ser Gly Thr Val Val Ala  
 85 90 95  
 Asp Ala Leu Gly Ala Gly Pro Trp Val Ser Ala Ala Asp Trp Glu Phe  
 100 105 110  
 Ala Cys Lys Ala Gly Ser Glu Ala Leu Thr Ala Ala Met Ala Leu Val  
 115 120 125  
 Gly Ser Gly Met Gln Arg Tyr Ala Leu Ala Ile Gly Ala Asp Thr Ala  
 130 135 140  
 Gln Gly Arg Pro Gly Asp Ala Leu Glu Tyr Thr Ala Ser Ala Gly Ala  
 145 150 155 160  
 Ala Ala Leu Ile Val Gly Pro Ala Thr Glu Ala Leu Ala Thr Ile Asp  
 165 170 175  
 Ala Thr Val Ser Tyr Val Thr Asp Thr Pro Asp Phe Tyr Arg Arg Ala  
 180 185 190  
 Asp Arg Pro Tyr Pro Val His Gly Asn Arg Phe Thr Gly Glu Pro Ala  
 195 200 205  
 Tyr Phe His Gln Ile Gln Ser Ala Ala Ser Glu Leu Leu Arg Gln Leu  
 210 215 220  
 Asn Arg Thr Ala Ala Asp Phe Thr Tyr Ala Val Phe His Gln Pro Asn  
 225 230 235  
 Ala Lys Phe Pro Gln Thr Val Ala Lys Arg Leu Gly Phe Thr Asp Ala  
 245 250 255  
 Gln Ile Ala Pro Gly Leu Leu Ser Pro Gln Ile Gly Asn Thr Tyr Ser  
 260 265 270  
 Gly Ala Ala Leu Leu Gly Leu Cys Ala Ile Leu Asp Val Ala Lys Pro  
 275 280 285  
 Gly Asp Thr Ile Phe Val Thr Ser Tyr Gly Ser Gly Ala Gly Ser Asp  
 290 295 300  
 Ala Tyr Ala Leu Thr Val Thr Glu Ala Ile Val Glu Arg Arg Glu Arg  
 305 310 315 320  
 Ala Pro Leu Thr Ala Ala Tyr Leu Ala Arg Lys Val Met Ile Asp Tyr  
 325 330 335  
 Ala Met Tyr Ala Lys Trp Arg Gly Lys Leu Val Met Gly  
 340 345

<210> 15

<211> 420

<212> PRT

<213> Bacillus subtilis

<400> 15

Met Val Ser Ala Gly Ile Glu Ala Met Asn Val Phe Gly Gly Thr Ala  
1 5 10

Tyr Leu Asp Val Met Glu Leu Ala Lys Tyr Arg His Leu Asp Thr Ala  
20 25 30

Arg Phe Glu Asn Leu Leu Met Lys Glu Lys Ala Val Ala Leu Pro Tyr  
35 40 45

Glu Asp Pro Val Thr Phe Gly Val Asn Ala Ala Lys Pro Ile Ile Asp  
50 55 60

Ala Leu Ser Glu Ala Glu Lys Asp Arg Ile Glu Leu Leu Ile Thr Cys  
65 70 75 80

Ser Glu Ser Gly Ile Asp Phe Gly Lys Ser Leu Ser Thr Tyr Ile His  
85 90 95

Glu Tyr Leu Gly Leu Asn Arg Asn Cys Arg Leu Phe Glu Val Lys Gln  
100 105 110

Ala Cys Tyr Ser Gly Thr Ala Gly Phe Gln Met Ala Val Asn Phe Ile  
115 120 125

Leu Ser Gln Thr Ser Pro Gly Ala Lys Ala Leu Val Ile Ala Ser Asp  
130 135 140

Ile Ser Arg Phe Leu Ile Ala Glu Gly Gly Asp Ala Leu Ser Glu Asp  
145 150 155 160

Trp Ser Tyr Ala Glu Pro Ser Ala Gly Ala Gly Ala Val Ala Val Leu  
165 170 175

Val Gly Glu Asn Pro Glu Val Phe Gln Ile Asp Pro Gly Ala Asn Gly  
180 185 190

Tyr Tyr Gly Tyr Glu Val Met Asp Thr Cys Arg Pro Ile Pro Asp Ser  
195 200 205

Glu Ala Gly Asp Ser Asp Leu Ser Leu Met Ser Tyr Leu Asp Cys Cys  
210 215 220

Glu Gln Thr Phe Leu Glu Tyr Gln Lys Arg Val Pro Gly Ala Asn Tyr  
225 230 235 240

Gln Asp Thr Phe Gln Tyr Leu Ala Tyr His Thr Pro Phe Gly Gly Met  
245 250 255

Val Lys Gly Ala His Arg Thr Met Met Arg Lys Val Ala Lys Val Lys  
260 265 270

Thr Ser Gly Ile Glu Thr Asp Phe Leu Thr Arg Val Lys Pro Gly Leu  
275 280 285

Asn Tyr Cys Gln Arg Val Gly Asn Ile Met Gly Ala Ala Leu Phe Leu  
290 295 300

Ala Leu Ala Ser Thr Ile Asp Gln Gly Arg Phe Asp Thr Pro Lys Arg  
305 310 315 320

Ile Gly Cys Phe Ser Tyr Gly Ser Gly Cys Cys Ser Glu Phe Tyr Ser  
325 330 335

Gly Ile Thr Thr Pro Gln Gly Gln Glu Arg Gln Arg Thr Phe Gly Ile  
340 345 350

Glu Lys His Leu Asp Arg Arg Tyr Gln Leu Ser Met Glu Glu Tyr Glu  
355 360 365

Leu Leu Phe Lys Gly Ser Gly Met Val Arg Phe Gly Thr Arg Asn Val  
370 375 380

Lys Leu Asp Phe Glu Met Ile Pro Gly Ile Met Gln Ser Thr Gln Glu  
385 390 395 400

Lys Pro Arg Leu Phe Leu Glu Glu Ile Ser Glu Phe His Arg Lys Tyr  
405 410 415

Arg Trp Ile Ser  
420

&lt;210&gt; 16

&lt;211&gt; 388

&lt;212&gt; PRT

&lt;213&gt; Mycobacterium marinum

&lt;400&gt; 16

Met Val Ser Ile Gly Ile His Asp Leu Ser Ile Ala Thr Ala His Tyr  
 1 5 10 15  
 Val Leu Asp His Ala Thr Leu Ala Glu His His Gly Val Asp Val Asn  
 20 25 30  
 Lys Tyr Leu Ile Gly Leu Gly Gln Gln Gln Met Ser Ile Val Ala Pro  
 35 40 45  
 Asp Glu Asp Ile Val Thr Leu Ala Ala Ala Ala Asp Pro Ile Ile  
 50 55 60  
 Lys Arg His Gly Ser Gln Lys Ile Arg Thr Ile Val Ile Gly Thr Glu  
 65 70 75 80  
 Thr Gly Val Asp Gln Ser Lys Ser Ala Gly Ile Trp Val Ser Ser Leu  
 85 90 95  
 Leu Gly Leu Pro Ser Ser Ala Arg Val Leu Glu Val Lys Gln Ala Cys  
 100 105 110  
 Tyr Gly Ala Thr Gly Ala Leu Gln Leu Ala Leu Ala Leu Val His Arg  
 115 120 125  
 Asp Pro Thr Gln Gln Val Leu Val Ile Ala Ala Asp Val Ala Arg Tyr  
 130 135 140  
 Asp Leu Asp Ser Pro Gly Glu Pro Thr Gln Gly Ala Ala Ala Ala Ala  
 145 150 155 160  
 Met Leu Val Ser Ala Asp Pro Ala Leu Leu Arg Leu Glu Glu Pro Thr  
 165 170 175  
 Gly Ile Tyr Thr Ala Asp Ile Met Asp Phe Trp Arg Pro Asn Tyr Arg  
 180 185 190  
 Ser Thr Ala Leu Val Asp Gly Lys Ala Ser Val Thr Ala Tyr Met Glu  
 195 200 205  
 Ala Ala Ser Gly Ala Trp Lys Asp Tyr Thr Glu Arg Gly Gly Arg Ala  
 210 215 220  
 Phe Gly Glu Phe Ala Ala Phe Cys Tyr His Gln Pro Phe Thr Lys Met  
 225 230 235 240  
 Ala Tyr Lys Ala His Lys Gln Leu Ala Ala Glu Ala Gly Glu Asp Ala  
 245 250 255  
 Ser Gly Ala Ala Val Gln Ala Ala Val Gly Asn Thr Val Glu Tyr Asn  
 260 265 270  
 Arg Arg Ile Gly Asn Ser Tyr Thr Ala Ser Leu Tyr Leu Ala Leu Ala  
 275 280 285

Ala Leu Leu Asp Gln Ala Asp Asp Leu Ser Asp Gln Pro Ile Ala Met  
 290 295 300  
 Leu Ser Tyr Gly Ser Gly Cys Val Ala Glu Leu Phe Ala Gly Thr Val  
 305 310 315 320  
 Thr Pro Gly Tyr Gln Gln His Leu Arg Thr Asp Gln His Arg Ala Ala  
 325 330 335  
 Leu Glu Thr Arg Ile Pro Leu Ser Tyr Glu His Tyr Arg Arg Leu His  
 340 345 350  
 Asn Leu Thr Leu Pro Thr Asn Gly Asn His His Ser Leu Pro Val Glu  
 355 360 365  
 Thr Ser Arg Pro Phe Arg Leu Thr Ala Ile Ser Glu His Lys Arg Met  
 370 375 380  
 Tyr Gly Ala Val  
 385

<210> 17

<211> 435

<212> PRT

<213> Zea mays

<400> 17

Met Leu Ala Ala Ser Thr Lys Val Gly Ser Arg Leu Ala Ser Pro His  
 1 5 10 15  
 Ala Ser Leu Ser Ala Gly Ala Ala Ala Ala Ala Leu Ala Ser Ser Pro  
 20 25 30  
 Val Leu Gly Ser Gly Met Leu Pro Gly Ala Gly Phe Gly Glu Thr Gly  
 35 40 45  
 Asn His His Ala Ala Asp Ala Pro Pro Pro Leu Pro Cys Ser Ser Ser  
 50 55 60  
 Gly Asp Ser Arg Glu Tyr Tyr Gln Trp Lys Arg Leu Val Asn Gln Arg  
 65 70 75 80  
 Gln Ser Thr Leu His Val Gly Glu Val Pro Ala Ala Leu Gly His His  
 85 90 95  
 Val Phe Gly Ala Gly Cys Ser Ser Arg Lys Gln His Ile Tyr Arg Tyr  
 100 105 110  
 Phe Ser Ser Ser His Gln Gly Ser Ile Trp Ala Arg Ser Lys Ile  
 115 120 125

Leu His Asp Leu Pro Gly Tyr Val Lys Ile Val Glu Val Gly Pro Arg  
 130 135 140  
 Asp Gly Leu Gln Asn Glu Lys Asp Ile Val Pro Thr Pro Val Lys Val  
 145 150 155 160  
 Glu Leu Ile Arg Arg Leu Ala Thr Ser Gly Leu Pro Val Val Glu Ala  
 165 170 175  
 Thr Ser Phe Val Ser Pro Lys Trp Val Pro Gln Leu Ala Asp Ala Lys  
 180 185 190  
 Asp Val Met Glu Ala Val Arg Thr Ile Gly Gly Val Arg Phe Pro Val  
 195 200 205  
 Leu Thr Pro Asn Leu Lys Gly Phe Glu Ala Ala Ile Ala Ala Gly Ala  
 210 215 220  
 Lys Glu Ile Ala Ile Phe Ala Ser Ala Ser Glu Gly Phe Ser Lys Ser  
 225 230 235 240  
 Asn Ile Asn Cys Thr Ile Lys Glu Ser Ile Ala Arg Tyr Asn Asp Val  
 245 250 255  
 Ala Leu Ala Ala Lys Glu Lys Glu Ile Pro Val Arg Gly Tyr Val Ser  
 260 265 270  
 Cys Val Val Gly Cys Pro Val Asp Gly Pro Val Pro Pro Ser Asn Val  
 275 280 285  
 Ala Tyr Val Ala Lys Glu Leu Tyr Asp Met Gly Cys Tyr Glu Val Ser  
 290 295 300  
 Leu Gly Asp Thr Ile Gly Val Gly Thr Pro Gly Thr Val Val Pro Met  
 305 310 315 320  
 Leu Glu Ala Ala Ile Ser Val Val Pro Val Glu Lys Leu Ala Val His  
 325 330 335  
 Phe His Asp Thr Tyr Gly Gln Ser Leu Ser Asn Ile Leu Ile Ser Leu  
 340 345 350  
 Gln Met Gly Val Ser Val Val Asp Ser Ser Val Ala Gly Leu Gly Gly  
 355 360 365  
 Cys Pro Tyr Ala Lys Gly Ala Ser Gly Asn Val Ala Thr Glu Asp Val  
 370 375 380  
 Val Tyr Met Leu Asn Gly Leu Gly Val Lys Thr Gly Val Asp Leu Gly  
 385 390 395 400  
 Lys Val Met Ala Ala Gly Glu Phe Ile Cys Arg His Leu Gly Arg Gln  
 405 410 415  
 Ser Gly Ser Lys Ala Ala Thr Ala Leu Ser Lys Val Thr Ala Asn Ala  
 420 425 430  
 Ser Lys Leu  
 435

<210> 18

<211> 335

<212> PRT

<213> Danio rerio (Brachydanio rerio)

<400> 18

Met Gly Asn Val Ser Ser Ala Val Lys His Cys Leu Ser Tyr Glu Thr  
 1 5 10  
 Phe Leu Arg Asp Tyr Pro Trp Leu Pro Arg Leu Leu Trp Glu Glu Lys  
 20 25 30  
 Cys Ser Glu Leu Pro Lys Leu Pro Val Tyr Val Lys Ile Val Glu Val  
 35 40 45  
 Gly Pro Arg Asp Gly Leu Gln Asn Glu Lys Glu Ile Val Pro Thr Glu  
 50 55 60  
 Val Lys Ile Gln Leu Ile Asp Leu Leu Ser Gln Thr Gly Leu Pro Val  
 65 70 75 80  
 Ile Glu Ala Thr Ser Phe Val Ser Ser Lys Trp Val Ala Gln Met Ala  
 85 90 95  
 Asp His Thr Ala Val Leu Lys Gly Ile Lys Arg Ser Pro Asp Val Arg  
 100 105 110  
 Tyr Pro Val Leu Thr Pro Asn Ile Gln Gly Phe Gln Ala Ala Val Ala  
 115 120 125  
 Ala Gly Ala Asn Glu Val Ala Val Phe Gly Ser Ala Ser Glu Thr Phe  
 130 135 140  
 Ser Arg Lys Asn Ile Asn Cys Ser Ile Glu Glu Ser Leu Gln Arg Phe  
 145 150 155 160  
 Glu Gln Val Val Ser Ala Ala Lys Gln Glu Gly Ile Pro Val Arg Gly  
 165 170 175  
 Tyr Val Ser Cys Ala Leu Gly Cys Pro Tyr Glu Gly Gln Val Lys Pro  
 180 185 190  
 Ser Gln Val Thr Lys Val Ala Lys Arg Leu Phe Glu Leu Gly Cys Tyr  
 195 200 205  
 Glu Val Ser Leu Gly Asp Thr Ile Gly Val Gly Thr Ala Gly Ser Met  
 210 215 220  
 Ala Glu Met Leu Ser Asp Val Leu Thr Glu Val Pro Ala Gly Ala Leu  
 225 230 235 240  
 Ala Val His Cys His Asp Thr Tyr Gly Gln Ala Leu Pro Asn Ile Leu  
 245 250 255  
 Ile Ala Leu Gln Met Gly Val Ser Val Val Asp Ala Ser Val Ala Gly  
 260 265 270  
 Leu Gly Gly Cys Pro Phe Ala Lys Gly Ala Ser Gly Asn Val Ser Thr  
 275 280 285  
 Glu Asp Leu Leu Tyr Met Leu His Gly Leu Gly Ile Glu Thr Gly Val  
 290 295 300  
 Asp Leu Leu Lys Val Met Glu Ala Gly Asp Phe Ile Cys Lys Ala Leu  
 305 310 315 320  
 Asn Arg Lys Thr Asn Ser Lys Val Ser Gln Ala Thr Arg Asn Asn  
 325 330 335

<210> 19

<211> 325

<212> PRT

<213> Bos taurus

<400> 19

Met Ala Thr Val Lys Lys Val Leu Pro Arg Arg Leu Val Gly Leu Ala  
 1 5 10 15  
 Thr Leu Arg Ala Val Ser Thr Ser Ser Val Gly Thr Phe Pro Lys Gln  
 20 25 30  
 Val Lys Ile Val Glu Val Gly Pro Arg Asp Gly Leu Gln Asn Glu Lys  
 35 40 45  
 Asn Ile Val Pro Thr Pro Val Lys Ile Lys Leu Ile Asp Met Leu Ser  
 50 55 60  
 Glu Ala Gly Leu Pro Val Val Glu Ala Thr Ser Phe Val Ser Pro Lys  
 65 70 75 80  
 Trp Val Pro Gln Met Ala Asp His Ala Glu Val Leu Lys Gly Ile Gln  
 85 90 95  
 Lys Phe Pro Gly Val Asn Tyr Pro Val Leu Thr Pro Asn Phe Lys Gly  
 100 105 110  
 Phe Gln Ala Ala Val Ala Ala Gly Ala Lys Glu Val Ala Ile Phe Gly  
 115 120 125  
 Ala Ala Ser Glu Leu Phe Thr Lys Lys Asn Ile Asn Cys Ser Ile Asp  
 130 135 140  
 Glu Ser Leu Gln Arg Phe Asp Glu Ile Leu Lys Ala Ala Arg Ala Ala  
 145 150 155 160  
 Gly Ile Ser Val Arg Gly Tyr Val Ser Cys Val Leu Gly Cys Pro Tyr  
 165 170 175  
 Glu Gly Lys Ile Ser Pro Ala Lys Val Ala Glu Val Thr Lys Lys Leu  
 180 185 190  
 Tyr Ser Met Gly Cys Tyr Glu Ile Ser Leu Gly Asp Thr Ile Gly Val  
 195 200 205  
 Gly Thr Pro Gly Ala Met Lys Asp Met Leu Ser Ala Val Leu Gln Glu  
 210 215 220  
 Val Pro Val Thr Ala Leu Ala Val His Cys His Asp Thr Tyr Gly Gln  
 225 230 235 240  
 Ala Leu Ala Asn Thr Leu Thr Ala Leu Gln Met Gly Val Ser Val Met  
 245 250 255  
 Asp Ser Ser Val Ala Gly Leu Gly Gly Cys Pro Tyr Ala Gln Gly Ala  
 260 265 270  
 Ser Gly Asn Leu Ala Thr Glu Asp Leu Val Tyr Met Leu Ala Gly Leu  
 275 280 285  
 Gly Ile His Thr Gly Val Asn Leu Gln Lys Leu Leu Glu Ala Gly Ala  
 290 295 300  
 Phe Ile Cys Gln Ala Leu Asn Arg Arg Thr Asn Ser Lys Val Ala Gln  
 305 310 315 320  
 Ala Thr Cys Lys Leu  
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<210> 20

<211> 325

<212> PRT

<213> Homo sapiens

<400> 20

Met Ala Ala Met Arg Lys Ala Leu Pro Arg Arg Leu Val Gly Leu Ala  
 1 5 10 15

Ser Leu Arg Ala Val Ser Thr Ser Ser Met Gly Thr Leu Pro Lys Arg  
 20 25 30  
 Val Lys Ile Val Glu Val Gly Pro Arg Asp Gly Leu Gln Asn Glu Lys  
 35 40 45  
 Asn Ile Val Ser Thr Pro Val Lys Ile Lys Leu Ile Asp Met Leu Ser  
 50 55 60  
 Glu Ala Gly Leu Ser Val Ile Glu Thr Thr Ser Phe Val Ser Pro Lys  
 65 70 75 80  
 Trp Val Pro Gln Met Gly Asp His Thr Glu Val Leu Lys Gly Ile Gln  
 85 90 95  
 Lys Phe Pro Gly Ile Asn Tyr Pro Val Leu Thr Pro Asn Leu Lys Gly  
 100 105 110  
 Phe Glu Ala Ala Val Ala Ala Gly Ala Lys Glu Val Val Ile Phe Gly  
 115 120 125  
 Ala Ala Ser Glu Leu Phe Thr Lys Lys Asn Ile Asn Cys Ser Ile Glu  
 130 135 140  
 Glu Ser Phe Gln Arg Phe Asp Ala Ile Leu Lys Ala Ala Gln Ser Ala  
 145 150 155 160  
 Asn Ile Ser Val Arg Gly Tyr Val Ser Cys Ala Leu Gly Cys Pro Tyr  
 165 170 175  
 Glu Gly Lys Ile Ser Pro Ala Lys Val Ala Glu Val Thr Lys Lys Phe  
 180 185 190  
 Tyr Ser Met Gly Cys Tyr Glu Ile Ser Leu Gly Asp Thr Ile Gly Val  
 195 200 205  
 Gly Thr Pro Gly Ile Met Lys Asp Met Leu Ser Ala Val Met Gln Glu  
 210 215 220  
 Val Pro Leu Ala Ala Leu Ala Val His Cys His Asp Thr Tyr Gly Gln  
 225 230 235 240  
 Ala Leu Thr Asn Thr Leu Met Ala Leu Gln Met Gly Val Ser Val Val  
 245 250 255  
 Asp Ser Ser Val Ala Gly Leu Gly Gly Cys Pro Tyr Ala Gln Gly Ala  
 260 265 270  
 Ser Gly Asn Leu Ala Thr Glu Asp Leu Val Tyr Met Leu Glu Gly Leu  
 275 280 285  
 Gly Ile His Thr Gly Val Asn Leu Gln Lys Leu Leu Glu Ala Gly Asn  
 290 295 300  
 Phe Ile Cys Gln Ala Leu Asn Arg Lys Thr Ser Ser Lys Val Ala Gln  
 305 310 315 320  
 Ala Thr Cys Lys Leu  
 325

&lt;210&gt; 21

&lt;211&gt; 299

&lt;212&gt; PRT

&lt;213&gt; Pseudomonas putida Q88H25

&lt;400&gt; 21

Met Ser Leu Pro Lys His Val Arg Leu Val Glu Val Gly Pro Arg Asp  
 1 5 10 15  
 Gly Leu Gln Asn Glu Ala Gln Pro Ile Ser Val Ala Asp Lys Val Arg  
 20 25 30  
 Leu Val Asn Asp Leu Thr Glu Ala Gly Leu Ala Tyr Ile Glu Val Gly  
 35 40 45  
 Ser Phe Val Ser Pro Lys Trp Val Pro Gln Met Ala Gly Ser Ala Glu  
 50 55 60  
 Val Phe Ala Gly Ile Gln Gln Arg Pro Gly Val Thr Tyr Ala Ala Leu  
 65 70 75 80  
 Ala Pro Asn Leu Arg Gly Phe Glu Asp Ala Leu Ala Ala Gly Val Lys  
 85 90 95  
 Glu Val Ala Val Phe Ala Ala Ala Ser Glu Ala Phe Ser Gln Arg Asn  
 100 105 110  
 Ile Asn Cys Ser Ile Ser Glu Ser Leu Lys Arg Phe Glu Pro Ile Met  
 115 120 125  
 Asp Ala Ala Arg Ser His Gly Met Arg Val Arg Gly Tyr Val Ser Cys  
 130 135 140  
 Val Leu Gly Cys Pro Tyr Glu Gly Lys Val Ser Ala Glu Gln Val Ala  
 145 150 155 160  
 Pro Val Ala Arg Ala Leu His Asp Met Gly Cys Tyr Glu Val Ser Leu  
 165 170 175  
 Gly Asp Thr Ile Gly Thr Gly Thr Ala Gly Asp Thr Arg Arg Leu Phe  
 180 185 190  
 Glu Val Val Ser Ala Gln Val Pro Arg Glu Gln Leu Ala Gly His Phe  
 195 200 205  
 His Asp Thr Tyr Gly Gln Ala Leu Ala Asn Val Tyr Ala Ser Leu Leu  
 210 215 220  
 Glu Gly Ile Ser Val Phe Asp Ser Ser Val Ala Gly Leu Gly Gly Cys  
 225 230 235 240  
 Pro Tyr Ala Lys Gly Ala Thr Gly Asn Ile Ala Ser Glu Asp Val Val  
 245 250 255  
 Tyr Leu Leu Gln Gly Leu Gly Ile Glu Thr Gly Ile Asp Leu Gly Leu  
 260 265 270  
 Leu Ile Ala Ala Gly Gln Arg Ile Ser Gly Val Leu Gly Arg Asp Asn  
 275 280 285  
 Gly Ser Arg Val Ala Arg Ala Cys Ser Ala Gln  
 290 295

<210> 22

<211> 312

<212> PRT

<213> Acinetobacter baumannii B7H4C6

<400> 22

Met Thr Ala Phe Ser Asp Leu Leu Val Val Gln Glu Val Ser Pro Arg  
 1 5 10 15  
 Asp Gly Leu Gln Ile Glu Pro Thr Trp Val Pro Thr Asp Lys Lys Ile  
 20 25 30  
 Asp Leu Ile Asn Gln Leu Ser Thr Met Gly Phe Ser Arg Ile Glu Ala  
 35 40 45  
 Gly Ser Phe Val Ser Pro Lys Ala Ile Pro Asn Leu Arg Asp Gly Glu  
 50 55 60  
 Glu Val Phe Thr Gly Ile Thr Arg His Lys Asp Ile Ile Tyr Val Gly  
 65 70 75 80  
 Leu Ile Pro Asn Leu Lys Gly Ala Leu Arg Ala Val Glu Ala Asn Ala  
 85 90 95  
 Asn Glu Leu Asn Leu Val Leu Ser Ala Ser Gln Thr His Asn Leu Ala  
 100 105 110  
 Asn Met Arg Met Thr Lys Ala Gln Ser Phe Ala Gly Phe Thr Glu Ile  
 115 120 125  
 Val Glu Gln Leu Gln Gly Lys Thr Gln Phe Asn Gly Thr Val Ala Thr  
 130 135 140  
 Thr Phe Gly Cys Pro Phe Glu Gly Lys Ile Ser Glu Arg Glu Val Phe  
 145 150 155 160  
 Ser Leu Val Glu His Tyr Leu Lys Leu Gly Ile His Asn Ile Thr Leu  
 165 170 175  
 Ala Asp Thr Thr Gly Met Ala Asn Pro Val Gln Val Lys Arg Ile Val  
 180 185 190  
 Ser His Val Leu Ser Leu Ile Ser Pro Glu Gln Leu Thr Leu His Phe  
 195 200 205  
 His Asn Thr Arg Gly Leu Gly Leu Thr Asn Val Leu Ala Ala Tyr Glu  
 210 215 220  
 Val Gly Ala Arg Arg Phe Asp Ala Ala Leu Gly Gly Leu Gly Gly Cys  
 225 230 235 240  
 Pro Phe Ala Pro Gly Ala Ser Gly Asn Ile Cys Thr Glu Asp Leu Val  
 245 250 255  
 Asn Met Cys Glu Glu Ile Gly Ile Pro Thr Thr Ile Asp Leu Asp Ala  
 260 265 270  
 Leu Ile Gln Leu Ser Arg Thr Leu Pro Ala Leu Leu Gly His Asp Thr  
 275 280 285  
 Pro Ser Gln Leu Ala Lys Ala Gly Arg Asn Thr Asp Leu His Pro Ile  
 290 295 300  
 Pro Asp Tyr Ile Lys Ser Leu Asn  
 305 310

<210> 23

<211> 286

<212> PRT

<213> Thermus thermophilus Q72IH0

<400> 23

Met Lys Ala Ser Val Arg Trp Val Glu Cys Pro Arg Asp Ala Trp Gln  
 1 5 10 15  
 Gly Phe Ser Arg Phe Ile Pro Thr Glu Glu Lys Val Ala Phe Leu Asn  
 20 25 30  
 Glu Leu Leu Glu Ala Gly Phe Ala His Leu Asp Leu Thr Ser Phe Val  
 35 40 45  
 Ser Pro Lys Trp Val Pro Gln Met Gln Asp Ala Glu Glu Val Leu Lys  
 50 55 60

Ala Leu Pro Pro Pro Asn Gly Arg Thr Tyr Leu Ala Ile Val Ala Asn  
65 70 75 80

Glu Lys Gly Leu Glu Arg Ala Leu Ala Ala Pro Asn Leu Thr His Val  
85 90 95

Gly Tyr Pro Phe Ser Leu Ser Glu Thr Phe Gln Gln Arg Asn Thr Asn  
100 105 110

Arg Ser Ile Glu Ala Ser Trp Pro Leu Val Gly Ala Met Val Glu Arg  
115 120 125

Thr Glu Gly Arg Leu Gly Leu Val Val Tyr Leu Ser Met Ala Phe Gly  
130 135 140

Asn Pro Tyr Gly Asp Pro Trp Ser Val Glu Ala Val Leu Glu Ala Leu  
145 150 155 160

Ala Arg Leu Lys Glu Met Gly Val Arg Glu Ile Ala Leu Ala Asp Thr  
165 170 175

Tyr Gly Val Ala Glu Pro Glu Arg Ile His Glu Val Leu Lys Ala Ala  
180 185 190

Val Ala Arg Phe Gly Pro Glu Gly Leu Gly Ala His Leu His Ala Arg  
195 200 205

Pro Glu Gly Ala Leu Ala Lys Val Glu Ala Val Leu Ala Ala Gly Val  
210 215 220

Thr Trp Leu Glu Gly Ala Leu Ala Gly Val Gly Gly Cys Pro Phe Ala  
225 230 235 240

Gly Asp Glu Leu Val Gly Asn Leu Pro Thr Glu Val Val Leu Pro His  
245 250 255

Leu Glu Lys Arg Gly Leu Ala Thr Gly Val Asp Leu Ser Arg Leu Pro  
260 265 270

Leu Leu Ala Glu Glu Ala Ala Arg Leu Lys Ala Leu Tyr Ala  
275 280 285

<210> 24

<211> 387

<212> PRT

<213> Lactobacillus delbrueckii

<400> 24

Met Asp Ile Gly Ile Asp Gln Ile Gly Phe Tyr Thr Pro Asn Lys Phe  
1 5 10 15

Val Asp Met Val Asp Leu Ala Asn Ala Arg Asn Gln Asp Pro Asn Lys  
 20 25 30  
 Phe Leu Ile Gly Ile Gly Gln Asp Arg Met Ala Val Ala Asp Lys Thr  
 35 40 45  
 Gln Asp Ala Val Ser Met Gly Ile Asn Ala Thr Ala Glu Tyr Leu Asp  
 50 55 60  
 Gln Val Asp Leu Glu Gln Leu Gly Leu Leu Ile Phe Ala Thr Glu Ser  
 65 70 75 80  
 Gly Ile Asp Gln Ser Lys Ser Ala Ser Leu Phe Val Lys Glu Ala Leu  
 85 90 95  
 Asn Leu Pro Ala Arg Ile Arg Thr Phe Glu Ile Lys Glu Ala Cys Phe  
 100 105 110  
 Ala Leu Thr Ala Ser Leu Gln Val Ala Arg Asp Tyr Val Arg Ala His  
 115 120 125  
 Pro His His Ser Ala Met Ile Ile Gly Ser Asp Ile Ala Arg Tyr Gly  
 130 135 140  
 Leu Ala Thr Ala Gly Glu Val Thr Gln Gly Ala Gly Ala Ile Ser Met  
 145 150 155 160  
 Leu Ile Lys Glu Asn Pro Ala Ile Ile Ala Leu Glu Asp Gly His Thr  
 165 170 175  
 Ser His Ser Glu Asn Ile Asn Asp Phe Trp Arg Pro Asn Asn Leu Ala  
 180 185 190  
 Thr Ala Val Val Asp Gly His Tyr Ser Arg Asp Val Tyr Leu Asp Phe  
 195 200 205  
 Phe Lys Ser Thr Phe Lys Pro Phe Leu Ala Glu Lys Gln Leu Gln Val  
 210 215 220  
 Ser Asp Phe Ala Gly Ile Cys Tyr His Leu Pro Tyr Thr Lys Met Gly  
 225 230 235 240  
 Tyr Lys Ala His Lys Ile Ala Ile Glu Gly Gln Asp Asp Glu Thr Val  
 245 250 255  
 Lys Arg Leu Ser Asp Asn Phe Gln Leu Ser Ala Lys Tyr Ser Arg Gln  
 260 265 270  
 Val Gly Asn Ile Tyr Thr Ala Ser Leu Tyr Met Ser Val Leu Ser Leu  
 275 280 285  
 Leu Glu Asn Gly Asp Leu Glu Ala Gly Asp Arg Ile Gly Phe Phe Ser  
 290 295 300  
 Tyr Gly Ser Gly Ala Met Ala Glu Phe Phe Ser Gly Lys Val Val Ala  
 305 310 315 320  
 Gly Tyr Gln Lys Arg Leu Arg Pro Ala Leu His Ala Arg Met Leu Lys  
 325 330 335  
 Glu Arg Ile Arg Leu Gly Val Gly Gln Tyr Glu Asp Ile Phe Thr Glu  
 340 345 350  
 Gly Leu Glu Ala Leu Pro Glu Asn Val Glu Phe Thr Ser Asp Ala Asn  
 355 360 365  
 His Gly Thr Trp Tyr Leu Ala Gly Gln Glu Gly Tyr Val Arg Gln Tyr  
 370 375 380  
 Lys Gln Lys  
 385

<210> 25

<211> 388

<212> PRT

<213> Staphylococcus haemolyticus

<400> 25

Met Ser Ile Gly Ile Asp Lys Ile Asn Phe Tyr Val Pro Lys Tyr Tyr  
 1 5 10 15  
 Val Asp Met Ala Lys Leu Ala Glu Ala Arg Gln Val Asp Pro Asn Lys  
 20 25 30  
 Phe Leu Ile Gly Ile Gly Gln Thr Gln Met Ala Val Ser Pro Val Ser  
 35 40 45  
 Gln Asp Ile Val Ser Met Gly Ala Asn Ala Ala Lys Asp Ile Ile Thr  
 50 55 60  
 Asp Asp Asp Lys Lys His Ile Gly Met Val Ile Val Ala Thr Glu Ser  
 65 70 75 80  
 Ala Ile Asp Asn Ala Lys Ala Ala Ala Val Gln Ile His Asn Leu Leu  
 85 90 95  
 Gly Val Gln Pro Phe Ala Arg Cys Phe Glu Met Lys Glu Ala Cys Tyr  
 100 105 110  
 Ala Ala Thr Pro Ala Ile Gln Leu Ala Lys Asp Tyr Ile Glu Lys Arg  
 115 120 125  
 Pro Asn Glu Lys Val Leu Val Ile Ala Ser Asp Thr Ala Arg Tyr Gly  
 130 135 140  
 Ile Gln Ser Gly Gly Glu Pro Thr Gln Gly Ala Gly Ala Val Ala Met  
 145 150 155 160  
 Leu Ile Ser Asn Asn Pro Ser Ile Leu Glu Leu Asn Asp Asp Ala Val  
 165 170 175  
 Ala Tyr Thr Glu Asp Val Tyr Asp Phe Trp Arg Pro Thr Gly His Lys  
 180 185 190  
 Tyr Pro Leu Val Ala Gly Ala Leu Ser Lys Asp Ala Tyr Ile Lys Ser  
 195 200 205  
 Phe Gln Glu Ser Trp Asn Glu Tyr Ala Arg Arg Glu Asp Lys Thr Leu  
 210 215 220  
 Ser Asp Phe Glu Ser Leu Cys Phe His Val Pro Phe Thr Lys Met Gly  
 225 230 235 240  
 Lys Lys Ala Leu Asp Ser Ile Ile Asn Asp Ala Asp Glu Thr Thr Gln  
 245 250 255  
 Glu Arg Leu Thr Ser Gly Tyr Glu Asp Ala Val Tyr Tyr Asn Arg Tyr  
 260 265 270  
 Val Gly Asn Ile Tyr Thr Gly Ser Leu Tyr Leu Ser Leu Ile Ser Leu  
 275 280 285  
 Leu Glu Asn Arg Ser Leu Lys Gly Gly Gln Thr Ile Gly Leu Phe Ser  
 290 295 300  
 Tyr Gly Ser Gly Ser Val Gly Glu Phe Phe Ser Ala Thr Leu Val Glu  
 305 310 315 320  
 Gly Tyr Glu Lys Gln Leu Asp Ile Glu Gly His Lys Ala Leu Leu Asn  
 325 330 335  
 Glu Arg Gln Glu Val Ser Val Glu Asp Tyr Glu Ser Phe Phe Lys Arg  
 340 345 350  
 Phe Asp Asp Leu Glu Phe Asp His Ala Thr Glu Gln Thr Asp Asp Asp  
 355 360 365  
 Lys Ser Ile Tyr Tyr Leu Glu Asn Ile Gln Asp Asp Ile Arg Gln Tyr  
 370 375 380  
 His Ile Pro Lys  
 385

## REFERENCES CITED IN THE DESCRIPTION

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#### Patent documents cited in the description

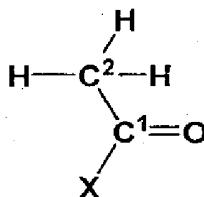
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## PATENTKRAV

1. Fremgangsmåde til fremstilling af 3-hydroxy-3-methylsmørsyre omfattende enzymatisk omdannelse af acetone og en forbindelse, som tilvejebringer en aktiveret acetylgruppe og er **kendetegnet ved** følgende formel I:



- hvor X er S-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH(OH)-C(CH<sub>3</sub>)<sub>2</sub>-CH<sub>2</sub>-O-PO<sub>2</sub>H-O-PO<sub>2</sub>H-C<sub>10</sub>H<sub>13</sub>N<sub>5</sub>O<sub>7</sub>P (coenzym A),

- under anvendelse af et enzym, som er i stand til at katalysere dannelsen af en covalent binding mellem carbonatomet i oxogruppen (dvs. C=O-gruppen) i acetone og det carbonatom (C<sup>2</sup>), der svarer til methylgruppen i den forbindelse, som tilvejebringer en aktiveret acetylgruppe med formlen I, idet enzymet er et enzym, der har en HMG-CoA-syntases (EC 2.3.3.10) aktivitet.

2. Fremgangsmåde ifølge krav 1, **kendetegnet ved, at** omdannelsen foregår i nærvær af en organisme, der er i stand til at producere acetone og udtrykke enzymet som defineret i krav 1.

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3. Rekombinant organisme, **kendetegnet ved** følgende træk:

- (a) den er i stand til at producere acetone; og  
 (b) den udtrykker et enzym, som er i stand til at katalysere dannelsen af en covalent binding mellem carbonatomet i oxogruppen (dvs. C=O-gruppen) i acetone og det carbonatom (C<sup>2</sup>), der svarer til methylgruppen i den forbindelse, som tilvejebringer en aktiveret acetylgruppe med formlen I, idet X er coenzym A, og idet enzymet er et enzym med en HMG-CoA-syntases (EC 2.3.3.10) enzymatiske aktivitet.

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4. Organisme ifølge krav 3, hvor organismen er en organisme, som naturligt har evne til at producere acetone.

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5. Organisme ifølge krav 4, hvor organismen er en mikroorganisme, der hører til slægten Clostridium, Bacillus eller Pseudomonas.

6. Mikroorganisme ifølge krav 5, hvor mikroorganismen er en mikroorganisme, der hører til arten Clostridium acetobutylicum, Clostridium beijerinckii, Clostridium cellulolyticum, Bacillus polymyxa eller Pseudomonas putida.

7. Organisme ifølge krav 3, hvor organismen er afledt af en organisme, som naturligt ikke producerer acetone og genetisk er modificeret til at være i stand til at producere acetone ved indføring af ét eller flere gener, der er nødvendige for at muliggøre produktion af acetone i organismen.

8. Organisme ifølge krav 7, hvor organismen er en organisme, der er i stand til fotosyntese.

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9. Anvendelse af en organisme ifølge et hvilket som helst af kravene 3 til 8 fremstilling af 3-hydroxy-3-methylsmørsyre.

10. Sammensætning omfattende en organisme ifølge et hvilket som helst af kravene 3 til 8.

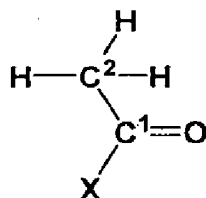
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11. Sammensætning omfattende

- (i) acetone; og
- (ii) en forbindelse, der tilvejebringer en aktiveret acetylgruppe og er **kendetegnet**

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**ved** følgende formel I:



hvor X er S-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH(OH)-C(CH<sub>3</sub>)<sub>2</sub>-CH<sub>2</sub>-O-PO<sub>2</sub>H-O-PO<sub>2</sub>H-C<sub>10</sub>H<sub>13</sub>N<sub>5</sub>O<sub>7</sub>P (coenzym A); og

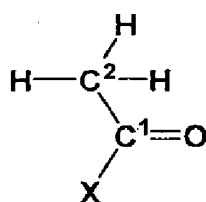
- (iii) et enzym, der er i stand til at katalysere dannelsen af en covalent binding mellem carbonatomet i oxogruppen (dvs. C=O-gruppen) i acetone og det carbonatom (C<sup>2</sup>), der svarer til methylgruppen i den under (ii) definerede

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forbindelse, idet enzymet er et enzym med en HMG-CoA-syntases (EC 2.3.3.10) enzymatiske aktivitet.

12. Anvendelse af et enzym ifølge krav 1 til fremstilling af 3-hydroxy-3-methyl-  
5 smørsyre.

13. Anvendelse af acetone til fremstilling af 3-hydroxy-3-methylsmørsyre omfattende enzymatisk omdannelse af acetone og en forbindelse, som tilvejebringer en aktiveret acetylgruppe og er **kendetegnet ved** følgende formel I:



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hvor X er S-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH<sub>2</sub>-CH<sub>2</sub>-NH-CO-CH(OH)-C(CH<sub>3</sub>)<sub>2</sub>-CH<sub>2</sub>-O-PO<sub>2</sub>H-O-PO<sub>2</sub>H-C<sub>10</sub>H<sub>13</sub>N<sub>5</sub>O<sub>7</sub>P (coenzym A), idet den enzymatiske omdannelse effektueres af et enzym med en HMG-CoA-syntases (EC 2.3.3.10) enzymatiske aktivitet.

## DRAWINGS

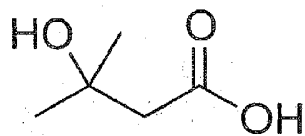


Figure 1

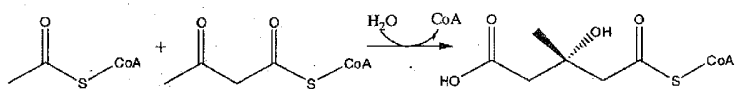


Figure 2

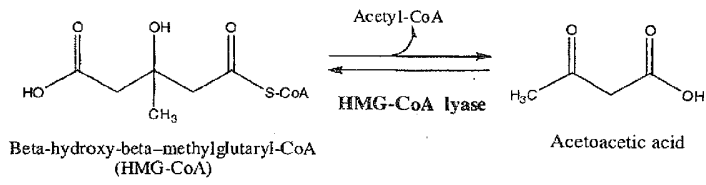


Figure 3

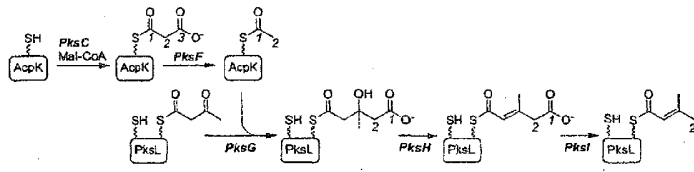


Figure 4

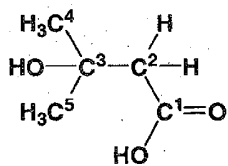
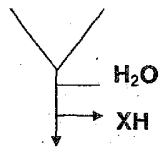
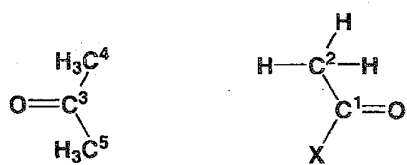


Figure 5

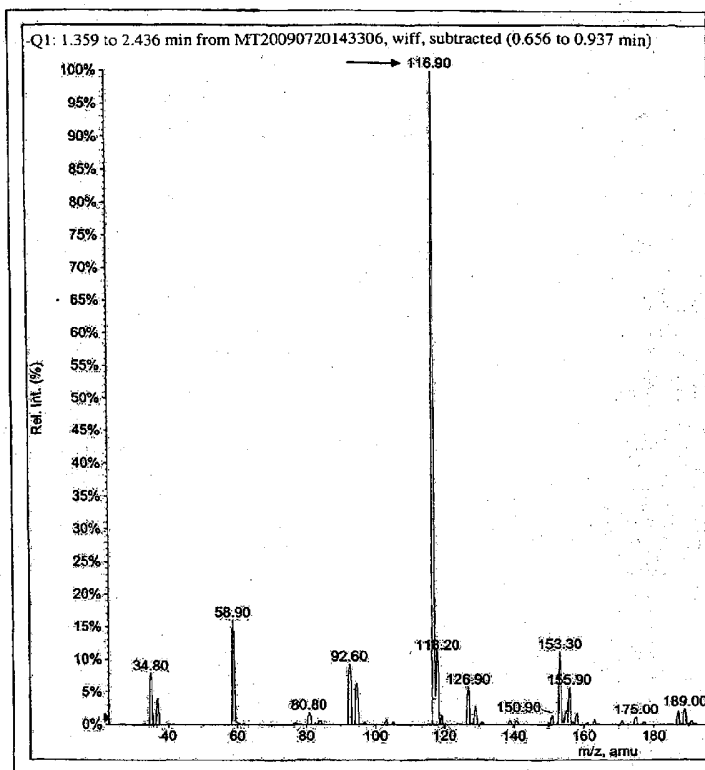


Figure 6

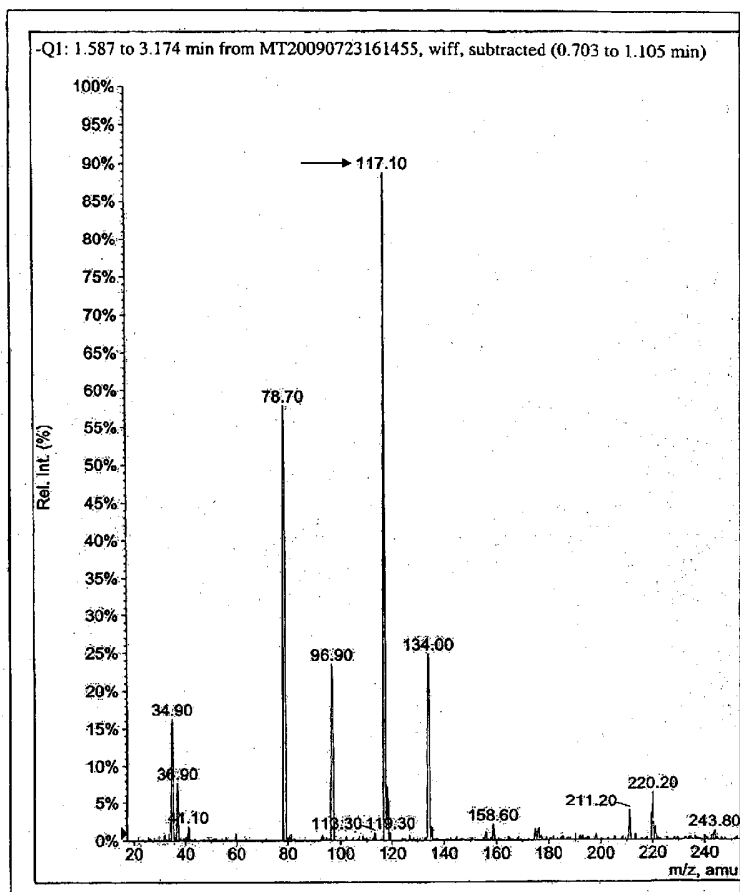


Figure 7

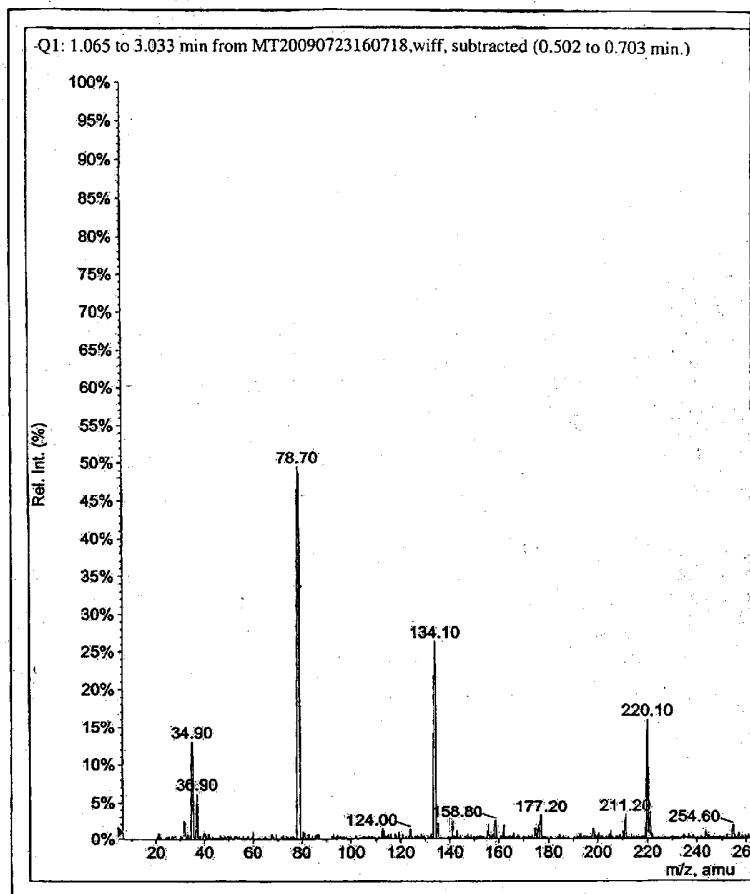


Figure 8

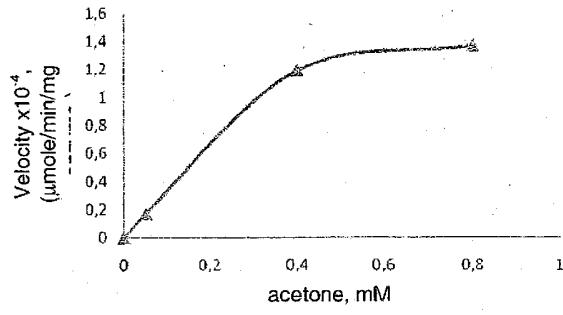


Figure 9