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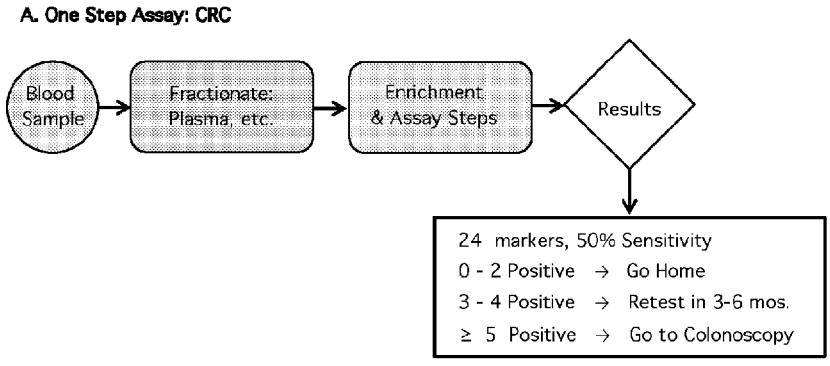
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Figures 1A

(57) Abstract: The present invention relates to methods for identifying and/or quantifying low abundance, nucleotide base mutations, insertions, deletions, translocations, splice variants, miRNA variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicings, exon insertions, exon deletions, intron insertions, or other rearrangement at the genome level and/or methylated nucleotide bases.

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MARKERS FOR IDENTIFYING AND QUANTIFYING OF NUCLEIC ACID SEQUENCE MUTATION, EXPRESSION, SPLICE VARIANT, TRANSLOCATION, COPY NUMBER, OR METHYLATION CHANGES

[0001] This application claims benefit of U.S. Provisional Patent Application Serial No. 62/843,032, filed May 3, 2019, which is hereby incorporated by reference in its entirety.

[0002] This invention was made with government support under grant number P41 EB020594 awarded by National Institutes of Health. The government has certain rights in the invention.

FIELD

[0003] The present application relates to methods and markers for identifying and quantifying nucleic acid sequence, mutation, expression, splice variant, translocation, copy number, and/or methylation changes using combinations of bisulfite treatment, nuclease, ligation, and polymerase reactions with carryover prevention.

BACKGROUND

[0004] Cancer is the leading cause of death in developed countries and the second leading cause of death in developing countries. Cancer kills 580,000 patients annually in the US, 1.3 million in Europe, and 2.8 million in China (Siegel et al., "Cancer Statistics, 2016," *CA Cancer J. Clin.* 66(1):7-30 (2016)). Cancer is now the biggest cause of mortality worldwide, with an estimated 8.2 million deaths from cancer in 2012 (Torre et al., "Global Cancer Statistics, 2012," *CA Cancer J. Clin.* 65(2):87-108 (2015)). Cancer cases worldwide are forecast to rise by 75% and reach close to 25 million over the next two decades. The lifetime risk of a woman dying from an invasive cancer is 19%, for a man it is 23%. With total annual costs of cancer care in the U.S. exceeding $400 billion, there is no other medical issue that so urgently needs intelligent solutions.

[0005] In the U.S., new cancer cases among men are dominated by prostate (21%), lung (14%), colorectal (8%), urinary bladder (7%), melanoma (6%), non-Hodgkin lymphoma (5%), renal (5%), head and neck (4%), leukemia (4%), and liver and bile cancer (3%). Among women, most of the newly diagnosed cancers are breast (29%), lung (13%), colorectal (8%), uterine corpus (7%), thyroid (6%), non-Hodgkin lymphoma (4%), melanoma (3%), leukemia (3%), pancreatic (3%), and renal cancer (3%). The leading causes of cancer deaths are lung cancer (27%), prostate cancer (8%), colorectal cancer (8%), and lung cancer (26%), breast cancer (14%), colorectal cancer (8%), for men and women, respectively. These cancers are driven by

different biological processes, and while there have been exciting advancements in the treatment of some cancers, such as the emergence of targeted therapeutics and immunotherapy, most cancers are found at later stage, where survival is poor. Due to lack of reliable and inexpensive early detection tests, many cancer types are diagnosed at later stages, where survival rates for some cancers drop to below 10%. The current screening technologies are failing due to low patient compliance, high expense, and low sensitivity and specificity rates (Das et al., "Predictive and Prognostic Biomarkers in Colorectal Cancer: A Systematic Review of Recent Advances and Challenges," *Biomedicine & Pharmacotherapy* 87:8-19 (2016)). For example, the high cost, discomfort, and invasiveness of colonoscopy are significant impediments to patient compliance for CRC screening (Beydoun et al., "Predictors of Colorectal Cancer Screening Behaviors Among Average-risk Older Adults in the United States," *Cancer Causes & Control: CCC* 19(4):339-359 (2008)). Likewise, patient distaste for handling feces has limited the success of FOBT/FIT, and eliminated stool-based tests as a remedy for low compliance. In contrast, the current proposal addresses these problems by developing a blood test with the potential to become widely adopted. Increasing patient compliance for CRC testing will lead to earlier detection and, ultimately, increased patient survival.

[0006] Ultimately, there is an urgent need to develop non-invasive, highly sensitive, highly specific, and cost-effective tests which will detect early-stage cancers. Two relatively recent developments in cancer research serve as the guiding principles for these tasks. First, is the use of modern genomic tools (such as genome-wide sequencing, transcriptional, and methylation profiling). Public accessibility to vast databases generated from these studies has accelerated the discovery of a wider list of molecular markers (such as promoter methylation, mutation, copy number, or expression levels of mRNA, microRNA, non-coding RNA (ncRNA), and long non-coding RNA (lncRNA) associated with cancer progression. Second is the discovery that nucleic acids can be released by the cancer cells into the patient's bloodstream. Cancer cells may undergo apoptosis (triggered cell death), which releases cell free DNA (cfDNA) into the patients' blood (Salvi et al., "Cell-free DNA as a Diagnostic Marker for Cancer: Current Insights," *OncoTargets and Therapy* 9:6549-6559 (2016)). The levels of cfDNA in serum from patients with cancer vary from vanishingly small to high, but do not correlate with cancer stage (Perlin et al., "Serum DNA Levels in Patients With Malignant Disease," *American Journal of Clinical Pathology* 58(5):601-602 (1972); Leon et al., "Free DNA in the Serum of Cancer Patients and the Effect of Therapy," *Cancer Res.* 37(3):646-650 (1977)). Moreover, exosomes (lipid vesicles ranging from 30 to 100 nm), which are released into the blood by cancer cells, can contain the same RNA molecules which serve as transcriptional signatures of the tumors. Exosomes, or tumor associated vesicles, shield mRNA,

lncRNA, ncRNA, and even mutant tumor DNA from exogenous nucleases, and, as such, the markers are in a protected state. Other protected states include, but are not limited to, DNA, RNA, and proteins within circulating tumor cells (CTCs), within other non-cellular membrane containing vesicles or particles, within nucleosomes, or within Argonaute or other protein complexes. cfDNA in particular, contains the same molecular aberrations as the solid tumors, such as mutations hyper/hypo methylation, copy number changes, or chromosomal rearrangements (Ignatiadis et al., "Circulating Tumor Cells and Circulating Tumor DNA for Precision Medicine: Dream or Reality?" *Ann. Oncol.* 25(12):2304-2313 (2014)).

[0007] Tumor-specific CpG methylations have been detected in the plasma from patients with a variety of solid tumors (Pratt VM, "Are We Ready for a Blood-Based Test to Detect Colon Cancer?" *Clinical Chemistry* 60(9):1141-1142 (2014); Warton et al., "Methylation of Cell-free Circulating DNA in the Diagnosis of Cancer," *Frontiers in Molecular Biosciences* 2:13 (2015)), through various techniques involving bisulfite conversion of unmethylated cytosines, methylation-sensitive enzymes, or immunoprecipitation of 5-methylcytosines (Jorda et al., "Methods for DNA methylation analysis and applications in colon cancer," *Mutat. Res.* 693(1-2):84-93 (2010)). Methylation signatures have better specificity towards a particular cancer type likely because methylation patterns are highly tissue specific (Issa JP, "DNA Methylation as a Therapeutic Target in Cancer," *Clin. Cancer Res.* 13(6):1634-1637 (2007)). The best studied blood-based methylation markers for CRC detection are located in the promoter region of the SEPT9 gene (Church et al., "Prospective Evaluation of Methylated SEPT9 in Plasma for Detection of Asymptomatic Colorectal Cancer," *Gut* 63(2):317-325 (2014); Lofton-Day et al., "DNA Methylation Biomarkers for Blood-Based Colorectal Cancer Screening," *Clinical Chemistry* 54(2):414-423 (2008); Potter et al., "Validation of a Real-time PCR-based Qualitative Assay for the Detection of Methylated SEPT9 DNA in Human Plasma," *Clinical Chemistry* 60(9):1183-1191 (2014); Ravegnini et al., "Simultaneous Analysis of SEPT9 Promoter Methylation Status, Micronuclei Frequency, and Folate-Related Gene Polymorphisms: The Potential for a Novel Blood-Based Colorectal Cancer Biomarker," *International Journal of Molecular Sciences* 16(12):28486-28497 (2015); Toth et al., "Detection of Methylated SEPT9 in Plasma is a Reliable Screening Method for Both Left- and Right-sided Colon Cancers," *PloS One* 7(9):e46000 (2002); Toth et al., "Detection of Methylated Septin 9 in Tissue and Plasma of Colorectal Patients with Neoplasia and the Relationship to the Amount of Circulating Cell-Free DNA," *PloS One* 9(12):e115415 (2014); Warren et al., "Septin 9 Methylated DNA is a Sensitive and Specific Blood Test for Colorectal Cancer," *BMC Medicine* 9:133 (2011)), and other potential markers for CRC diagnostics include CpG sites on promoter regions of THBD (Lange et al., "Genome-scale Discovery of DNA-methylation Biomarkers for Blood-Based

Detection of Colorectal Cancer," *PloS One* 7(11):e50266 (2012)), C9orf50 (Lange et al., "Genome-scale Discovery of DNA-methylation Biomarkers for Blood-Based Detection of Colorectal Cancer," *PloS One* 7(11):e50266 (2012)), ZNF154 (Margolin et al., "Robust Detection of DNA Hypermethylation of ZNF154 as a Pan-Cancer Locus with in Silico Modeling for Blood-Based Diagnostic Development," *The Journal of Molecular Diagnostics* 18(2):283-298 (2016)), and AGBL4, FLI1 and TWIST1 (Lin et al., "Clinical Relevance of Plasma DNA Methylation in Colorectal Cancer Patients Identified by Using a Genome-Wide High-Resolution Array," *Ann. Surg. Oncol.* 22 Suppl 3:S1419-1427 (2015)). In breast cancer, methylation at promoter regions of tumor suppressor genes (including ATM, BRCA1, RASSF1, APC, and RARβ) has been detected in patients' cfDNAs (Tang et al., "Blood-based DNA Methylation as Biomarker for Breast Cancer: a Systematic Review," *Clinical Epigenetics* 8:115 (2016)). A caveat for using methylation markers is that bisulfite conversion tends to destroy DNA, and thus decreases the overall signal that can be detected. Methylation detection techniques may also lead to false-positive signals due to incomplete conversion of unmethylated cytosines. As described herein, an extensive bioinformatics analysis of public databases has been performed to identify CRC-specific, and tissues-specific methylation markers suitable for detection of cancer in the plasma. The methylation marker detection assays enable a higher level of multiplexing with single-molecule detection capabilities, which are predicted to allow for higher sensitivity and specificity across a broad spectrum of cancers.

[0008] The challenge to develop reliable diagnostic and screening tests is to distinguish those markers emanating from the tumor that are indicative of disease (*e.g.*, early cancer) vs. presence of the same markers emanating from normal tissue (which would lead to a false-positive signal). There is also a need to balance the number of markers examined and the cost of the test, with the specificity and sensitivity of the assay. Comprehensive molecular profiling (mRNA, methylation, copy number, miRNA, mutations) of thousands of tumors by The Cancer Genome Atlas Consortium (TCGA), has revealed that colorectal tumors are as different from each other as they are from breast, prostrate, or other epithelial cancers (TCGA "Comprehensive Molecular Characterization of Human Colon and Rectal Cancer *Nature* 487:330-337 (2014)). Further, those few markers they share in common are also present in multiple cancer types, hindering the ability to pinpoint the tissue of origin. BRAF mutations frequently occur in melanoma (42%) and thyroid cancer (41%), while KRAS is also highly mutated in pancreatic (55%) and lung (16%) cancers (Forbes et al., "COSMIC: Exploring the World's Knowledge of Somatic Mutations in Human Cancer," *Nucleic Acids Res.* 43(Database issue):D805-811 (2015)). In general, CRC mutation markers such as those of *KRAS* and *BRAF* are found in late-stage primary cancers and metastases (Spindler et al., "Circulating free DNA as Biomarker and Source

for Mutation Detection in Metastatic Colorectal Cancer," *PloS One* 10(4):e0108247 (2015); Gonzalez-Cao et al., "BRAF Mutation Analysis in Circulating Free Tumor DNA of Melanoma Patients Treated with BRAF Inhibitors," *Melanoma Res.* 25(6):486-495 (2015); Sakai et al., "Extended RAS and BRAF Mutation Analysis Using Next-Generation Sequencing," *PloS One* 10(5):e0121891 (2015)). For early cancer detection, the nucleic acid assay should serve primarily as a screening tool, requiring the availability of secondary diagnostic follow-up (*e.g.*, colonoscopy for colorectal cancer).

[0009] Compounding the biological problem is the need to reliably quantify mutation, CpG methylation, or DNA or RNA copy number from either a very small number of initial cells (*i.e.* from CTCs), or when the cancer signal is from cell-free DNA (cfDNA) in the blood and diluted by an excess of nucleic acid arising from normal cells, or inadvertently released from normal blood cells during sample processing (Mateo et al., "The Promise of Circulating Tumor Cell Analysis in Cancer Management," *Genome Biol.* 15:448 (2014); Haque et al., "Challenges in Using ctDNA to Achieve Early Detection of Cancer," *BioRxiv.* 237578 (2017)).

[0010] Some cancer IVD companies have developed commercially available methylation detection tests. The aforementioned SEPT9 methylation is the basis for Epi proColon test, a CRC-detection assay by Epigenomics (Lofton-Day et al., "DNA Methylation Biomarkers for Blood-based Colorectal Cancer Screening," *Clinical Chemistry* 54(2):414-423 (2008)). While initial results on smaller sample sets showed promise, large-scale studies with 1,544 plasma samples showed a sensitivity of 64% for stage I-III CRC, and a specificity of 78%-82%, effectively sending 180 to 220 out of 1,000 individuals to unnecessary colonoscopies (Potter et al., "Validation of a Real-time PCR-based Qualitative Assay for the Detection of Methylated SEPT9 DNA in Human Plasma," *Clinical Chemistry* 60(9):1183-1191 (2014)). Clinical Genomics is currently developing blood based CRC detection test based on the methylation of the BCAT1 and IKZF1 genes (Pedersen et al., "Evaluation of an Assay for Methylated BCAT1 and IKZF1 in Pasma for Detection of Colorectal Neoplasia," *BMC Cancer* 15:654 (2015)]. Large-scale studies using 2,105 plasma samples of this two-marker test showed an overall sensitivity of 66%, with 38% for stage I CRC, and an impressive specificity of 94% (Young et al, "A Cross-sectional Study Comparing a Blood Test for Methylated BCAT1 and IKZF1 Tumor-derived DNA with CEA for Detection of Recurrent Colorectal Cancer," *Cancer Medicine* 5(10): 2763-2772 (2016)). Exact Sciences and collaborators have slightly improved the sensitivity of CRC fecal tests (Bosch et al., "Analytical Sensitivity and Stability of DNA Methylation Testing in Stool Samples for Colorectal Cancer Detection," *Cell Oncol. (Dordr)* 35(4):309-315 (2012); Hong et al., "DNA Methylation Biomarkers of Stool and Blood for Early Detection of Colon Cancer," *Genet. Test. Mol. Biomarkers* 17(5):401-406 (2013); Imperiale et al., "Multitarget Stool

DNA Testing for Colorectal-Cancer Screening," *N. Engl. J. Med.* 370(14):1287-1297 (2014); Xiao et al., "Validation of Methylation-Sensitive High-Resolution Melting (MS-HRM) for the Detection of Stool DNA Methylation in Colorectal Neoplasms," *Clin. Chim. Acta* 431:154-163 (2014); Yang et al., "Diagnostic Value of Stool DNA Testing for Multiple Markers of Colorectal Cancer and Advanced Adenoma: a Meta-Analysis," *Can. J. Gastroenterol.* 27(8):467-475 (2013)), by adding K-ras mutation as well as BMP3 and NDRG4 methylation markers (Lidgard et al., "Clinical Performance of an Automated Stool DNA Assay for Detection of Colorectal Neoplasia," *Clin. Gastroenterol. Hepatol.* 11(10):1313-1318 (2013)). Large-scale studies on 12,500 stool samples claims 93% sensitivity, yet specificity is still only 85%, essentially sending 150 out of 1,000 individuals to unnecessary colonoscopies. Despite logistical issues in handling feces, Exact Sciences recently sold their millionth test. The Cologuard website states the test result has both false-positives and false-negatives, and the test should not be used if the patient has hemorrhoids, menstrual period, or blood in the stool. The Cologuard website also warns that the test is not for use by patients with Ulcerative Colitis (UC), Crohn's disease (CD), Inflammatory Bowel Disease (IBD), or with a family history of cancer. In other words, Exact Sciences excludes the very patients who would most benefit from an accurate CRC detection test. More recently, Laboratory for Advanced Medicine (based in Irvine, CA with ties to various Chinese academic institutions) demonstrated the potential of interrogating the methylation status of a single CpG site (cg10673833) for blood-based detection of colorectal cancer (Luo et al., "Circulating Tumor DNA Methylation Profiles Enable Early Diagnosis, Prognosis Prediction, and Screening for Colorectal Cancer," *Science Translational Medicine* 12:(524) (2020)).

A continuum of diagnostic needs will require a continuum of diagnostic tests.

[0011] The majority of current molecular diagnostics efforts in cancer have centered on: (i) prognostic and predictive genomics, *e.g.*, identifying inherited mutations in cancer predisposition genes, such as BrCA1, BrCA2, (Ford et al., *Am. J. Hum. Genet.* 62:676-689 (1998)) (ii) individualized treatment, *e.g.*, mutations in the EGFR gene guiding personalized medicine (Sequist and Lynch, *Ann. Rev. Med*, 59:429-442 (2008)), and (iii) recurrence monitoring, *e.g.*, detecting emerging KRAS mutations in patients developing resistance to drug treatments (Hiley et al., *Genome Biol.* 15: 453 (2014); Amado et al., *J. Clin. Oncol.* 26:1626-1634 (2008)). Yet, this misses major opportunities in the cancer molecular diagnostics continuum: (i) more frequent screening of those with a family history, (ii) screening for detection of early disease, and (iii) monitoring treatment efficacy. To address these three unmet needs, a new metric for blood-based detection termed "cancer marker load", analogous to viral load is herein proposed.

[0012] DNA sequencing provides the ultimate ability to distinguish all nucleic acid changes associated with disease. However, the process still requires multiple up-front sample and template preparation, and consequently, DNA sequencing is not always cost-effective. DNA microarrays can provide substantial information about multiple sequence variants, such as SNPs or different RNA expression levels, and are less costly then sequencing; however, they are less suited for obtaining highly quantitative results, nor for detecting low abundance mutations. On the other end of the spectrum is the TaqMan™ reaction, which provides real-time quantification of a known gene, but is less suitable for distinguishing multiple sequence variants or low abundance mutations.

[0013] NGS requires substantial up-front sample preparation to polish ends and append linkers, and the current error rates of 0.7% are too high to identify 2-3 molecules of mutant sequence in a 10,000-fold excess of wild-tye molecules. "Deep sequencing" protocols have been developed to overcome this deficiency by appending unique molecular identifiers to both strands of an individual fragment. These approaches are known as: Tam-Seq & CAPP-Seq (Roche), Circle-Seq (Guardant Health), Safe-SeqS (Personal Genome Diagnostics), ThruPlex (Rubicon Genomics), NEBNext (New England Biolabs), QIAseq (Qiagen), Oncomine (ThermoFisher), Duplex Barcoding (Schmitt), SMRT (Pacific Biosciences), SiMSen-Seq (Stahlberg), and smMIP (Shendure). However, these methods require a 30 to 100-fold depth per mutant strand to verify each mutation and distinguish from other types of sequencing errors. Recent work from MSKCC demonstrates that 60,000-fold coverage is required to accurately identify mutations in plasma from metastatic cancer patients (91% sensitivity, 508-gene panel, 60,000x). Compounding the challenge, a recent paper from NEB has called into question the quality of the most widely used databases for rare variant and somatic mutations (Chen et al., "DNA Damage is a Pervasive Cause of Sequencing Errors, Directly Confounding Variant Identification," *Science* 355(6326):752-756 (2017)).

[0014] It is critical to match each unmet diagnostic need with the appropriate diagnostic test – one that combines the divergent goals of achieving both high sensitivity (*i.e.*, low false-negatives) and high specificity (*i.e.*, low false-positives) at a low cost. For example, direct sequencing of EGFR exons from a tumor biopsy to determine treatment for non-small cell lung cancer (NSCLC) is significantly more accurate and cost effective than designing TaqMan™ probes for the over 180 known mutations whose drug response is already catalogued (Jia et al., *Genome Res.* 23:1434-1445 (2013)). The most sensitive technique for detecting point mutations, such as "BEAMing" (Dressman et al., *Proc. Natl. Acad. Sci. USA* 100: 8817-8822 (2003)), rely on prior knowledge of which mutations to look for, and thus are best suited for monitoring for disease recurrence, rather than for early detection. Likewise, to monitor blood levels of Bcr-Abl

translocations when treating CML patients with Gleevec (Jabbour et al., *Cancer* 112: 2112-2118 (2008)), a simple quantitative reverse-transcription PCR assay is far preferable to sequencing the entire genomic DNA in 1 ml of blood (9 million cells x 3 GB = 27 million Gb of raw data).

[0015] Sequencing 2.1 Gb each of cell-free DNA (cfDNA) isolated from NSCLC patients was used to provide 10,000-fold coverage on 125 kb of targeted DNA (Kandoth et al., *Nature* 502**:** 333-339 (2013)). This approach correctly identified mutations present in matched tumors, although only 50% of stage 1 tumors were covered. The approach has promise for NSCLC, where samples average 5 to 20 mutations/Mb, however targeted NGS would not be cost effective for other cancers such as breast and ovarian, that average less than 1 to 2 mutations per Mb. Current up-front ligation, amplification, and/or capture steps required for highly accurate targeted deep sequencing are still more complex than multiplexed PCR-TaqMan™ or PCR-LDR assays.

[0016] Deep sequencing of cfDNAs for 58 cancer-related genes at 30,000-fold coverage is capable of detecting Stage 1 or 2 cancer at moderately high sensitivity but missed 29% of CRC, 41% of breast, 41% of lung, and 32% of ovarian cancer, respectively (Phallen et al., "Direct Detection of Early-stage Cancers Using Circulating Tumor DNA," *Science Translational Medicine* 9(403) (2017)). An alternative strategy relied on targeted sequencing of an average of 30 bases in 61 segments to interrogate "hot-spot" mutations in 16 genes including TP53, KRAS, APC, PIK3CA, PTEN, missed more early cancers (Cohen et al., "Detection and Localization of Surgically Resectable Cancers with a Multi-analyte Blood Test," *Science* (2018). To extend the sensitivity of mutation sequencing, the Hopkins team very recently combined NGS with quantitation of serum protein markers (such as CA-125, CA19-9, CEA, HGF, Myeloperoxidase, OPN, Prolactin, TIMP-1) and improved detection of five cancer types (ovary, liver, stomach, pancreas, and esophagus) at sensitivities ranging from 69% to 98% (Cohen et al. "Detection and Localization of Surgically Resectable Cancers with a Multi-analyte Blood Test," *Science* (2018). One caveat of using these protein markers is that prior large-scale studies with age-matched controls (n= 22,000) have not shown clinical utility (Jacobs et al., "Prevalence Screening for Ovarian Cancer in Postmenopausal Women by CA 125 Measurement and Ultrasonography," *BMJ* 306(6884):1030-1034 (1993)). Thus, in a 2018 JAMA report, "The USPSTF recommends against [CA-125] screening for ovarian cancer in asymptomatic women. This recommendation applies to asymptomatic women who are not known to have a high-risk hereditary cancer syndrome" (USPSTF et al., "Screening for Ovarian Cancer: US Preventive Services Task Force Recommendation Statement" *JAMA* 319(6):588-594 (2018)). Another caveat of using these protein markers is that they reflect tissue damage and are likely to also appear in patients with inflammatory diseases such as arthritis (Kaiser, "'Liquid Biopsy for Cancer Promises Early

Detection," *Science* 359(6373):259 (2018)). With the growing obesity epidemic and an aging population in the U.S., the risk of false-positives from protein markers increases with obesity and age-driven inflammation.

[0017] More recently, the NGS sequencing companies (Grail, Guardant Health, Natera, Freenome) have moved aggressively to expand their targeted sequencing panels to also now include whole genome sequencing (WGS) and whole genome bisulfite sequencing (Bis-WGS). The recent results from Grail, abstract published at ASCO 2018 (Klein et al., "Development of a Comprehensive Cell-free DNA (cfDNA) Assay for Early Detection of Multiple Tumor Types: The Circulating Cell-free Genome Atlas (CCGA) Study," ASCO Annual Meeting 2018, Chicago, Il; Abstract 12021#134)) reveal that while sensitivity claims of detecting "early" CRC are at 63%, that is based on only 27 samples, most of which are Stage III. Even mutation rich lung cancer gives sensitivity at 50%, again with most samples at Stage III. When most of the samples are Stage I & II, such as prostate cancer, the sensitivity for "early cancer" detection drops to < 5%. When attempting to detect the most common form of breast cancer (HR+/HER2), the sensitivity drops to < 13%. Worse, those breast cancers diagnosed by screening gave sensitivities of < 11%. In short, the NGS approach fails by consistently missing 30% to 80% of early-stage cancers (i.e. stage I & II). In a research initially reported in 2019 ASCO meeting (Liu et al., "Simultaneous Multi-cancer Detection and Tissue of Origin (TOO) Localization Using Targeted Bisulfite Sequencing Plasma Cell-free DNA (cfDNA)," ASCO Breakthrough Presentation 2019)), and subsequently published in 2020 (Liu et al., "Sensitive and Specific Multi-cancer Detection and Localization Using Methylation Signatures in Cell-free DNA," *Annals of Oncology*; In Press (2020)), GRAIL indicated that their Multi-Cancer Early Detection Test exhibited an Overall Detection Rate (12 deadly cancer types) of 76 % (99.3 % specificity). A combined analysis of this group of cancers showed robust detection across all stages with detection rates of 39 percent (27-52%), 69 percent (56-80%), 83 percent (75-90%), and 92 percent (86-96%) at stages I (n=62), II (n=62), III (n=102), and IV (n=130), respectively. In another conference, GRAIL and collaborators (Oxnard et al., "Simultaneous Multi-cancer Detection and Tissue of Origin (TOO) Localization Using Targeted Bisulfite Sequencing of Plasma Cell-free DNA (cfDNA)," ESMO Congress (2019)), reported the results from their analysis of cell-free DNA (DNA that had once been confined to cells but had entered the bloodstream upon the cells' death) in 3,583 blood samples, including 1,530 from patients diagnosed with cancer and 2,053 from people without cancer. The patient samples comprised more than 20 types of cancer, including hormone receptor-negative breast, colorectal, esophageal, gallbladder, gastric, head and neck, lung, lymphoid leukemia, multiple myeloma, ovarian, and pancreatic cancer. The overall specificity was 99.4%, meaning only 0.6% of the

results incorrectly indicated that cancer was present. The sensitivity of the assay for detecting a pre-specified high mortality cancer (the percent of blood samples from these patients that tested positive for cancer) was 76%. Within this group, the sensitivity was 32% for patients with stage I cancer; 76% for those with stage II; 85% for stage III; and 93% for stage IV. Sensitivity across all cancer types was 55%, with similar increases in detection by stage. For the 97% of samples that returned a tissue of origin result, the test correctly identified the organ or tissue of origin in 89% of cases. However, another 2019 study (reported by GRAIL and collaborators) questioned the validity of the aforementioned reports (Razavi et al., "High-intensity Sequencing Reveals the Sources of Plasma Circulating Cell-free DNA Variants," *Nat Med* 25(12):1928-1937 (2019)). Through a 2 Mb, 508-gene panel sequencing (60,000x depth), the authors demonstrated the vast majority of cell-free DNA mutations in both non-cancer controls and cancer patients had features consistent with clonal hematopoiesis, a process whereby white blood cells progressively accumulate somatic alterations without necessarily producing a hematological condition or malignancy. Indeed. mutations appeared in 93.6 percent of the white blood cells from individuals without cancer and 99.1 percent of those with cancer. In a recently held conference, GRAIL and their collaborators reported that their blood-based test can detect multiple GI cancers at sensitivity of under 50% for Stage I, and 73% for Stage 1-III (Wolpin et al., "Performance of a Blood-based Test for the Detection of Multiple Cancer Types," In: Gastrointestinal Cancers Symposium 2020 (2020)). As for Freenome, a recent ASCO presentation indicated that their platform (plasma analysis by whole-genome sequencing, bisulfite sequencing, and protein quantification methods) was able to achieve a mean sensitivity of 92% in early-stage (n = 17) and 84% in late-stage (n = 11) at a specificity of 90% for colorectal adenocarcinoma detection. Across all CRC pathological subtypes, the Freenome test achieved a specificity of 90%, and sensitivities of 80% and 83% for early-stage (n = 19) and late-stage (n = 12), respectively. Private discussion with Imran Haque, who just resigned as CSO of Freenome – where he had a $70 million budget and 30 scientists to sequence the plasma of 817 CRC and matched control patients – confirmed that Freenome (as well as GRAIL) were overcalling the data, and that none of them had a cogent approach to achieve cost-effective true early cancer detection (Wan et al., "Machine Learning Enables Detection of Early-stage Colorectal Cancer by Whole-genome Sequencing of Plasma Cell-free DNA," *BioRxiv* 478065 (2018)).

[0018] A comprehensive data analysis of over 600 colorectal cancer samples that takes into account tumor heterogeneity, tumor clusters, and biological/technical false-positives ranging from 3% to 10% per individual marker showed that the optimal early detection screen for colorectal cancer would require at least 5 to 6 positive markers out of 24 markers tested (Bacolod et al,. *Cancer Res.* 69:723-727 (2009); Tsafrir et al., *Cancer Res.* 66: 2129-2137 (2006);

Weinstein et al., *Nat. Genet.* 45: 1113-1120 (2013); Navin N.E. *Genome Biol.* 15: 452 (2014); Hiley et al., *Genome Biol* 15:453 (2014)); Esserman et al. *Lancet Oncol* 15:e234-242 (2014)). Further, marker distribution is biased into different tumor clades, *e.g.*, some tumors are heavily methylated, while others are barely methylated, and indistinguishable from age-related methylation of adjacent tissue. Consequently, a multidimensional approach using combinations of 3-5 sets of mutation, methylation, miRNA, ncRNA, lncRNA, mRNA, copy-variation, alternative splicing, or translocation markers is needed to obtain sufficient coverage of all different tumor clades. Analogous to non-invasive prenatal screening for trisomy, based on sequencing or performing ligation detection on random fragments of cfDNA (Benn et al., *Ultrasound Obstet. Gynecol.* 42(1):15-33 (2013); Chiu et al., *Proc. Natl. Acad. Sci. USA* 105: 20458 – 20463 (2008); Juneau et al., *Fetal Diagn. Ther.* 36(4) (2014)), the actual markers scored in a cancer screen are secondary to accurate quantification of those positive markers in the plasma.

[0019] As ponted out above, cancer-specific RNA markers (including microRNAs, lncRNAs, and mRNAs) may also be present in blood, either free of any compartment (Souza et al., "Circulating mRNAs and miRNAs as Candidate Markers for the Diagnosis and Prognosis of Prostate Cancer," *PloS One* 12(9):e0184094 (2017)), or contained in exosomes (Nedaeinia et al., "Circulating Exosomes and Exosomal microRNAs as Biomarkers in Gastrointestinal Cancer," *Cancer Gene Ther* 24(2):48-56 (2017); Lai et al., "A microRNA Signature in Circulating Exosomes is Superior to Exosomal Glypican-1 Levels for Diagnosing Pancreatic Cancer," *Cancer Lett* 39:86-93 (2017)) or circulating tumor cells ("CTCs"), and have been tagged as potential indicators of early- stage cancers. Challenges abound regarding the use of plasma-derived nucleic markers in early cancer detection, including the minuscule amount of these markers in blood relative to those derived from surrounding cells. Indeed, these limitations make it appear that these "early" detection assays are more likely to detect late-stage primary and metastatic cancers (Pantel "Blood-Based Analysis of Circulating Cell-Free DNA and Tumor Cells for Early Cancer Detection," *PLoS Med* 13(12):e1002205 (2016)).

Technical Challenges of Cancer Diagnostic Test Development.

[0020] Diagnostic tests that aim to find very rare or low-abundance mutant sequences face potential false-positive signal arising from: (i) polymerase error in replicating wild-type target, (ii) DNA sequencing error, (iii) mis-ligation on wild-type target, (iii) target independent PCR product, and (iv) carryover contamination of PCR products arising from a previous positive

sample. The profound clinical implications of a positive test result when screening for cancer demand that such a test use all means possible to virtually eliminate false-positives.

[0021] Central to the concept of nucleic acid detection is the selective amplification or purification of the desired cancer-specific markers away from the same or closely similar markers from normal cells. These approaches include: (i) multiple primer binding regions for orthogonal amplification and detection, (ii) affinity selection of CTC's or exosomes, and (iii) spatial dilution of the sample.

[0022] The success of PCR-LDR, which uses 4 primer-binding regions to assure sensitivity and specificity, has previously been demonstrated. Desired regions are amplified using pairs or even tandem pairs of PCR primers, followed by orthogonal nested LDR primer pairs for detection. One advantage of using PCR-LDR is the ability to perform proportional PCR amplification of multiple fragments to enrich for low copy targets, and then use quantitative LDR to directly identify cancer-specific mutations. Biofire/bioMerieux has developed a similar technology termed "film array"; wherein initial multiplexed PCR reaction products are redistributed into individual wells, and then nested real-time PCR performed with SYBR Green Dye detection.

[0023] Affinity purification of CTC's using antibody or aptamer capture has been demonstrated (Adams et al., *J. Am. Chem. Soc.* 130: 8633-8641 (2008); Dharmasiri et al., *Electrophoresis* 30:3289-3300 (2009); Soper *et al. Biosens. Bioelectron.* 21:1932-1942 (2006)). Peptide affinity capture of exosomes has been reported in the literature. Enrichment of these tumor-specific fractions from the blood enables copy number quantification, as well as simplifying screening and verification assays.

[0024] The last approach, spatial dilution of the sample, is employed in digital PCR as well as its close cousin known as BEAMing (Vogelstein and Kinzler, *Proc. Natl. Acad. Sci. U S A.* 96(16):9236-41 (1999); Dressman et al., *Proc. Natl. Acad. Sci. USA* 100:8817-8822 (2003)). The rational for digital PCR is to overcome the limit of enzymatic discrimination when the sample comprises very few target molecules containing a known mutation in a 1,000 to 10,000-fold excess of wild-type DNA. By diluting input DNA into 20,000 or more droplets or beads to distribute less than one molecule of target per droplet, the DNA may be amplified via PCR, and then detected via probe hybridization or TaqMan™ reaction, giving in essence a 0/1 digital score. The approach is currently the most sensitive for finding point mutations in plasma, but it does require prior knowledge of the mutations being scored, as well as a separate digital dilution for each mutation, which would deplete the entire sample to score just a few mutations (Alcaide et al., "A Novel Multiplex Droplet Digital PCR Assay to Identify and Quantify KRAS Mutations in Clinical Specimens," *J. Mol. Diagn.* 21:28-33 (2019); Guibert et al., "Liquid Biopsy of Fine-

Needle Aspiration Supernatant for Lung Cancer Genotyping," *Lung Cancer* 1768:193-207 (2018); Yoshida et al., "Highly Sensitive Detection of ALK Resistance Mutations in Plasma Using Droplet Digital PCR," *BMC Cancer* 18:1136 (2018)).

[0025] When developing multiplexed assays, there is a tricky balance between performing enough preliminary cycles of PCR or other amplification techniques to generate sufficient copies of each mutant or methylated region such that when diluting into uniplex qPCR, multiplex qPCR, uniplex droplet PCR or multiplexed droplet PCR there are sufficient copies to get a signal if true positive; and performing too many PCR cycles such that some markers over-amplify while others are suppressed, or relative quantification is lost.

[0026] The present application is directed at overcoming these and other deficiencies in the art.

SUMMARY

[0027] A first aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences of other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules are then provided. One or more primary oligonucleotide primer sets are also provided. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The sample, the one or more first primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase extension reaction mixtures and for carrying out one or

more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising nucleotide sequences complementary to the target nucleotide sequence. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more second primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixtures, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures. The method further comprises subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more first polymerase chain reaction products comprising the target nucleotide sequence or a complement thereof. One or more oligonucleotide probe sets are then provided. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on a complementary target nucleotide sequence of a secondary extension product. The one or more first polymerase chain reaction products are blended with a ligase, and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures. The one or more ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets are ligated together, when hybridized to complementary sequences, to form ligated product sequences in the ligation reaction mixtures wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion. The method further includes providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable

for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more second polymerase chain reaction products. The method further comprises detecting and distinguishing the one or more second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

[0028] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. One or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, one or more nucleases capable of digesting nucleic acid molecules not comprising modified nucleotides, and one or more first primary oligonucleotide primer(s) are provided. The one or more first primary oligonucleotide primer(s) comprise a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence. The sample, the one or more first primary oligonucleotide primers, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix that comprises one or more modified nucleotides that protect extension products but not target DNA from nuclease digestion, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase extension reaction mixture and for carrying out one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the target nucleotide sequence. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer

having a first 5' primer-specific portion and a 3' portion that is complementary to a portion of a primary extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a second 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary oligonucleotide primer sets, the one or more nucleases, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting nucleic acid molecules present in the first polymerase chain reaction mixtures, but not primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more first polymerase chain reaction products comprising the first 5' primer-specific portion, a target-specific nucleotide sequence or a complement thereof, and a complement of the second 5' primer-specific portion. One or more tertiary oligonucleotide primer sets are provided. Each tertiary oligonucleotide primer set comprises (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the first 5' primer-specific portion of the one or more first polymerase chain reaction products and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the one or more first polymerase chain reaction products. The one or more first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures, and the one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU) containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more second polymerase chain reaction products. The method further involves detecting and distinguishing the one or more second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

[0029] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences of other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules and one or more nucleases capable of digesting nucleic acid molecules present not comprising modified nucleotides are provided. The method also involves providing one or more primary oligonucleotide primer sets. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The sample, the one or more first primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix that comprises one or more modified nucleotides that protect extension product but not target DNA from nuclease digestion, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase extension reaction mixtures and for carrying out one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the target nucleotide sequence. The method further comprises blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more second primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more nucleases, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures. The one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting nucleic acid molecules present in the polymerase chain reaction mixtures, but not primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension

treatment, thereby forming first polymerase chain reaction products comprising the target nucleotide sequence or a complement thereof. One or more secondary oligonucleotide primer sets are then provided. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer. The first polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further comprises detecting and distinguishing the second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

[0030] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues and subjecting the nucleic acid molecules in the sample to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules are then provided. The method further involves providing one or more primary oligonucleotide primer sets. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer

that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The bisulfite-treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite-treated target nucleotide sequence. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures. The one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising the bisulfite-treated target nucleotide sequence or a complement thereof. The method further involves providing one or more oligonucleotide probe sets. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion, and (b) a second oligonucleotide probe having a 5' bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion and a 3' primer-specific portion, and wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on a complementary nucleotide sequence of a first polymerase chain reaction product. The first polymerase chain reaction products are blended with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures. The one or more ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets are ligated together, when hybridized to complementary sequences, to form ligated product sequences in the ligation reaction mixture wherein each ligated product sequence comprises the 5' primer-specific portion, the bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portions, and the 3' primer-specific portion. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set

comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures, and the one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming a second polymerase chain reaction products. The method further involves detecting and distinguishing the second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

[0031] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues. The nucleic acid molecules in the sample are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules are provided, and one or more first primary oligonucleotide primer(s) are provided. Each first primary oligonucleotide primer comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue. The bisulfite-treated sample, the one or more first primary oligonucleotide primers, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, to form primary extension products

comprising the complement of the bisulfite-treated target nucleotide sequence. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of the polymerase extension reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting nucleic acid molecules present in the first polymerase chain reaction mixtures, but not primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reactions products comprising a 5' primer-specific portion of the first secondary oligonucleotide primer, the bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion, and a complement of the 5' primer-specific portion of the second secondary oligonucleotide primer. The method further involves providing one or more tertiary oligonucleotide primer sets. Each tertiary oligonucleotide primer set comprises (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reactions product sequence. The first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain reaction products. The method further involves detecting and distinguishing the secondary polymerase chain reactions products in the one or

more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

[0032] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues. The nucleic acid molecules in the sample are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample are provided, and one or more primary oligonucleotide primer sets are provided. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The bisulfite-treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures. The one or more polymerase extension reaction mixtures to are subjected to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite treated target nucleotide sequence. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixture, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures. The one or more first polymerase chain reaction mixtures are sujected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization

treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising the bisulfite- treated target nucleotide sequence or a complement thereof. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of a first polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a first polymerase chain reaction product formed from the first secondary oligonucleotide primer. The first polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further involves detecting and distinguishing the second polymerase chain reactions products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

[0033] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues, and subjecting the nucleic acid molecules in the sample to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample are provided. One or more primary oligonucleotide primer sets are also provided. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to

the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The bisulfite treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures. The one or more polymerase extension reaction mixtures are subjected to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite treated target nucleotide sequence. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixture, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures. The method further comprises subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reactions products comprising the bisulfite-treated target nucleotide sequence or a complement thereof. One or more secondary oligonucleotide primer sets are provided. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products or their complements and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction products or their complements. The primary polymerase chain reaction product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment

thereby forming second polymerase chain reaction products. The method further involves detecting and distinguishing the second polymerase chain reactions products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

[0034] Another aspect of the present application is directed to a method for identifying in a sample, one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level. The method involves providing a sample containing one or more parent ribonucleic acid molecules containing a target ribonucleic acid molecule potentially differing in sequence from other parent ribonucleic acid molecules and providing one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU containing nucleic acid molecules potentially present in the sample. One or more primary oligonucleotide primer sets are then provided. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to the RNA sequence in the parent ribonucleic acid molecule adjacent to the target ribonucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of the cDNA extension product formed from the first primary oligonucleotide primer. The contacted sample, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix including dUTP, a reverse transcriptase, and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target ribonucleic nucleic acid and to carry out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse transcription/polymerase products. The method further comprises providing one or more oligonucleotide probe sets. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to

hybridize, in a base specific manner, on complementary portions of a reverse transcriptase/polymerase product corresponding to the target ribonucleic acid molecule sequence. The reverse transcriptase/polymerase products are contacted with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures, and the one or more ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form ligated product sequences in the ligase reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion. The method further involves providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences, the one or more secondary oligonucleotide primer sets with one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming first polymerase chain reaction products. The method further comprises detecting and distinguishing the first polymerase chain reaction products, thereby identifying the presence of one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level.

[0035] Another aspect of the present application is directed to a method for identifying in a sample, one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level. The method involves providing a sample containing one or more parent ribonucleic acid molecules

containing a target ribonucleic acid molecule potentially differing in sequence from other parent ribonucleic acid molecules and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU containing nucleic acid molecules potentially present in the sample. The method further involves providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to the RNA sequence in the parent ribonucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of the cDNA extension product formed from the first primary oligonucleotide primer. The contacted sample, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures, and the one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target RNA and to carry out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse-transcription/primary polymerase chain reaction products. The method futher comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of a reverse-transcription/primary polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a reverse-transcription/primary polymerase chain reaction product formed from the first secondary oligonucleotide primer. The reverse-transcription/primary polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming first polymerase chain reaction products. The method further involves detecting and distinguishing the first polymerase chain reaction

products, thereby identifying the presence of one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequences differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level.

[0036] Another aspect of the present application is directed to a method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases. The method involves providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample, and the contacted sample is blended with a ligase and one or more first oligonucleotide preliminary probes comprising a 5' phosphate, a 5' stem-loop portion, an internal primer-specific portion within the loop region, a blocking group, and a 3' nucleotide sequence that is complementary to a 3' portion of the target miRNA molecule sequence to form one or more first ligation reaction mixtures. The method further comprises ligating, in the one or more first ligation reaction mixtures, the one or more target miRNA molecules at their 3'end to the 5' phosphate of the one or more first oligonucleotide preliminary probes to generate chimeric nucleic acid molecules comprising the target miRNA molecule sequence, if present in the sample, appended to the one or more first oligonucleotide preliminary probes. One or more primary oligonucleotide primer sets are then provided. Each primer set comprises (a) a first primary oligonucleotide primer comprising a nucleotide sequence that is complementary to the internal primer-specific portion of the first oligonucleotide preliminary probe, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets. The one or more first ligation reaction mixtures comprising chimeric nucleic acid molecules, the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the sample, a deoxynucleotide mix including dUTP, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic

acid molecules present in the reverse-transcription/polymerase chain reaction mixtures to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the chimeric nucleic acid molecules, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different primary reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion, a nucleotide sequence corresponding to the target miRNA molecule sequence, and the complement of the internal primer-specific portion, and complements thereof. The method further involves providing one or more oligonucleotide probe sets. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion, a portion complementary to a primary extension product, and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on complementary portions of a primary reverse-transcription/polymerase chain reaction product corresponding to the target miRNA molecule sequence, or complement thereof. The primary reverse-transcription/polymerase chain reaction products are contacted with a ligase and the one or more oligonucleotide probe sets to form one or more second ligation reaction mixtures, and the one or more second ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form ligated product sequences in the ligation reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion. The method further involves providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences and the one or more secondary oligonucleotide primer sets are blended with one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain

reaction products. The method further comprises detecting and distinguishing the secondary polymerase chain reaction products in the one or more reactions thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

[0037] Another aspect of the present application is directed to a method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases. The method involves providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample, and the contacted sample is blended with a ligase and one or more first oligonucleotide probes comprising a 5' phosphate, a 5' stem-loop portion, an internal primer-specific portion within the loop region, a blocking group, and a 3' nucleotide sequence that is complementary to a 3' portion of the target miRNA molecule sequence to form one or more ligation reaction mixtures. The method further involves ligating, in the one or more ligation reaction mixtures, the one or more target miRNA molecules at their 3'end to the 5' phosphate of the one or more first oligonucleotide probes to generate chimeric nucleic acid molecules comprising the target miRNA molecule sequence, if present in the sample, appended to the one or more first oligonucleotide probes. One or more primary oligonucleotide primer sets are then provided. Each primer set comprises (a) a first primary oligonucleotide primer comprising a nucleotide sequence that is complementary to the internal primer-specific portion of the first oligonucleotide probe, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets. The one or more ligation reaction mixtures comprising chimeric nucleic acid molecules, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the chimeric nucleic acid molecules, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different primary reverse-transcription/polymerase chain reaction products comprising the 5'

primer-specific portion, a nucleotide sequence corresponding to the target miRNA molecule sequence, and the complement of the internal primer-specific portion, and complements thereof. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer. The primary reverse-transcription/polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising a 5' primer-specific portion of the first secondary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence or a complement thereof, and a complement of the other 5' primer-specific portion second secondary oligonucleotide primer. The method further involves providing one or more tertiary oligonucleotide primer sets. Each tertiary oligonucleotide primer set comprises (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products or their complements and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction products or their complements. The first polymerase chain reaction process products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures, and the one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU) containing nucleic acid molecules present in the secondpolymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further comprises detecting and distinguishing the second polymerase chain reaction products, thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

[0038] Another aspect of the present application is directed to a method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases. The method involves providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample. The contacted sample is blended with ATP and a Poly(A) polymerase to form a Poly(A) polymerase reaction mixture, and the Poly(A) polymerase reaction mixture is subjected to conditions suitable for appending homopolymer A to the 3' ends of the one or more target miRNA molecules potentially present in the sample. The method further involves providing one or more primary oligonucleotide primer sets. Each primer set comprises (a) a first primary oligonucleotide primer comprising a 5' primer-specific portion, an internal poly dT portion, and a 3' portion comprising from 1 to 10 bases complementary to the 3' end of the target miRNA, wherein the first primary oligonucleotide primer may be the same or may differ from other first primary oligonucleotide primers in other sets, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets. The Poly(A) polymerase reaction mixture, the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the sample, a deoxynucleotide mix including dUTP, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the reverse-transcription/polymerase chain reaction mixtures, then to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target miRNA sequences with 3' polyA tails, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion of the second primary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence, a poly dA region, and the complement of the 5' primer-specific portion of the first primary oligonucleotide primer, and complements thereof. The method further comprises providing one or more oligonucleotide probe sets. Each

probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion, a portion complementary to the one or more reverse-transcription/polymerase chain reaction products, and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, to complementary portions of the one or more reverse-transcription/polymerase chain reaction products corresponding to the target miRNA molecule sequence, or complement thereof. The one or more reverse-transcription/polymerase chain reaction products are contacted with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures, and the one or more ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form ligated product sequences in the ligation reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion. The method further involves providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences and the one or more secondary oligonucleotide primer sets are blended with one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain reaction products. The method further comprises detecting and distinguishing the secondary polymerase chain reaction products, thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

[0039] Another aspect of the present application is directed to a method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases. The method involves providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases and providing one or

more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample. The contacted sample is blended with ATP and a Poly(A) polymerase to form a Poly(A) polymerase reaction mixture, and the Poly(A) polymerase reaction mixture is subjected to conditions suitable for appending a homopolymer A to the 3' ends of the one or more target miRNA molecules potentially present in the sample. The method further involves providing one or more primary oligonucleotide primer sets. Each primer set comprises (a) a first primary oligonucleotide primer comprising a 5' primer-specific portion, an internal poly dT portion, and a 3' portion comprising from 1 to 10 bases complementary to the 3' end of the target miRNA, wherein the first primary oligonucleotide primer may be the same or may differ from other first primary oligonucleotide primers in other sets, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets. The Poly(A) polymerase reaction mixture potentially comprising target miRNA sequences is blended with 3' polyA tails, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target miRNA sequences with 3' polyA tails, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more different reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion of the second primary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence, a poly dA region, and the complement of the 5' primer-specific portion of the first primary oligonucleotide primer, and complements thereof. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of a reverse-transcription/polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a reverse-transcription/polymerase chain reaction product formed from the first secondary oligonucleotide primer. The reverse-transcription/polymerase chain reaction products, the one or more

secondary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising a 5' primer-specific portion, a nucleotide sequence corresponding to the target miRNA molecule sequence or a complement thereof, and a complement of the other 5' primer-specific portion. The method further involves providing one or more tertiary oligonucleotide primer sets. Each tertiary oligonucleotide primer set comprises (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction product sequence and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction product sequence. The first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures, and one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further comprises detecting and distinguishing the second polymerase chain reaction products in the one or more reactions thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

[0040] Another aspect of the present application is directed to a method of diagnosing or prognosing a disease state of cells or tissue based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual. The plurality of markers is in a set comprising from 6-12 markers, 12-24 markers, 24-36 markers, 36-48 markers, 48-72 markers, 72-96 markers, or > 96 markers. Each marker in a given set is selected by having any one or more of the following criteria: present, or above a cutoff level, in > 50% of biological samples of the disease cells or tissue from individuals diagnosed with the disease state; absent, or below a cutoff level, in > 95% of biological samples of the normal cells or tissue from individuals without the disease state; present, or above a cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or

fractions thereof, from individuals diagnosed with the disease state; absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without the disease state; present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with the disease state. At least 50% of the markers in a set each comprise one or more methylated residues, and/or at least 50% of the markers in a set that are present, or above a cutoff level, or present with a z-value of > 1.65, comprise of one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with the disease state. The method involves obtaining a biological sample. The biological sample includes cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, and the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof. The sample is fractionated into one or more fractions, wherein at least one fraction comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein. Nucleic acid molecules in the one or more fractions are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. At least two enrichment steps are carried out for 50% or more disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step. The method further involves performing one or more assays to detect and distinguish the plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with the disease state if a minimum of 2 or 3 markers are present or are above a cutoff level in a marker set comprising from 6-12 markers; or a minimum of 3, 4, or 5 markers are present or are above a cutoff level in a marker set comprising from 12-24 markers; or a minimum of 3, 4, 5, or 6 markers are present or are above a cutoff level in a marker set comprising from 24-36 markers; or a minimum of 4, 5, 6, 7, or 8 markers are present or are above a cutoff level in a marker set comprising from 36-48 markers; or a minimum of 6, 7, 8, 9, 10, 11, or 12 markers are present or are above a cutoff level in a marker set comprising from 48-72 markers, or a minimum of 7, 8, 9, 10, 11, 12 or 13 markers are present or are above a cutoff level in a marker set comprising from 72-96 markers, or a minimum of 8, 9, 10, 11, 12, 13 or "n"/12 markers are

present or are above a cutoff level in a marker set comprising 96 – "n" markers, when "n" > 168 markers.

[0041] Another aspect of the present application is directed to a method of diagnosing or prognosing a disease state of a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual. The plurality of markers is in a set comprising from 48-72 total cancer markers, 72-96 total cancer markers or ≥ 96 total cancer markers, wherein on average greater than one quarter such markers in a given set cover each of the aforementioned major cancers being tested. Each marker in a given set for a given solid tissue cancer is selected by having any one or more of the following criteria for that solid tissue cancer: present, or above a cutoff level, in > 50% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer; present, or above a cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer; present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer. At least 50% of the markers in a set each comprise one or more methylated residues, and/or at least 50% of the markers in a set that are present, or above a cutoff level, or present with a z-value of > 1.65, comprise of one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with a given solid tissue cancer. The method involves obtaining a biological sample including cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma,

amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof. The sample is fractionated into one or more fractions, wherein at least one fraction comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein. The nucleic acid molecules in the one or more fractions are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. At least two enrichment steps are carried out for 50% or more of the given solid tissue cancer-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step. The method further involves performing one or more assays to detect and distinguish the plurality of cancer -specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with the a solid-tissue cancer if a minimum of 4 markers are present or are above a cutoff level in a marker set comprising from 48-72 total cancer markers; or a minimum of 5 markers are present or are above a cutoff level in a marker set comprising from 72-96 total cancer markers; or a minimum of 6 or "n"/18 markers are present or are above a cutoff level in a marker set comprising 96 to "n" total cancer markers, when "n" > 96 total cancer markers.

[0042] Another aspect of the present application is directed to a method of diagnosing or prognosing a disease state of and identifying the most likely specific tissue(s) of origin of a solid tissue cancer in the following groups: Group 1 (colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma); Group 2 (breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma); Group 3 (lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma); Group 4 (prostate adenocarcinoma, invasive urothelial bladder cancer); and/or Group 5 (liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma) based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual, wherein the plurality of markers is in a set comprising from 36-48 group-specific cancer markers, 48-64 group-specific cancer markers, or ≥ 64 group-specific cancer markers, wherein on average greater than one third of such markers in a given set cover each of the aforementioned cancers being tested within that group. Each marker in a given set for a given solid tissue cancer is selected by having any one or more of the following criteria for that solid tissue cancer: present, or above a cutoff level, in > 50% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer; present, or above a

cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer; present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer. At least 50% of the markers in a set each comprise one or more methylated residues, and/or at least 50% of the markers in a set that are present, or above a cutoff level, or present with a z-value of > 1.65 comprise one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with a given solid tissue cancer. The method involves obtaining the biological sample. The biological sample includes cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof. The sample is fractionated into one or more fractions, wherein at least one fraction comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein. The nucleic acid molecules in the one or more fractions are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. At least two enrichment steps are carried out for 50% or more of the given solid tissue cancer-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step. The method further involves performing one or more assays to detect and distinguish the plurality of cancer-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with a solid-tissue cancer if a minimum of 4 markers are present or are above a cutoff level in a marker set comprising from 36-48 group-specific cancer markers; or a minimum of 5 markers are present or are above a cutoff level in a marker set comprising from 48-64 group-specific cancer markers; or a minimum of 6 or "n"/12 markers are present or are above a cutoff level in a marker set comprising 64 to "n" total cancer markers, when "n" > 64 group-specific cancer markers.

[0043] The present application describes a number of approaches for detecting mutations, expression, splice variant, translocation, copy number, and/or methylation changes in target

nucleic acid molecules using nuclease, ligase, and polymerase reactions. The present application solves the problems of carry over prevention, as well as allowing for spatial multiplexing to provide relative quantification, similar to digital PCR. Such technology may be utilized for non-invasive early detection of cancer, non-invasive prognosis of cancer, and monitoring for cancer recurrence from plasma or serum samples.

[0044] The present application provides a comprehensive roadmap of nucleic acid methylation, miRNA, lncRNA, ncRNA, mRNA Exons, as well as cancer-associated protein markers that are specific for solid-tissue cancers and matched normal tissues. The present application teaches the art of selecting the desired number of markers and types of markers for both pan-oncology and specific cancers (*i.e.* colorectal cancer) to guide the physician to improve the treatment of the patient. Details on primer design and optimized primer sequences are provided to enable rapid validation of these tests for both pan-oncology and specific cancers. The two-step procedure is designed to cast a wide net to initially identify most of the individuals harboring an early cancer, followed by a more stringent second step to improve specificity and narrow the patients to those most likely to harbor a hidden cancer, who are then sent for imaging and followup. The advantage of this 2-step approach is that it not only identifies the potential tissue of origin, but it is designed to provide the highest positive predictive value (PPV). Thus, when a result for a rare cancer comes back as presumptive positive (*i.e.* early ovarian cancer) the physician can focus her attention on providing imaging and followup to those patients who need it the most, while the test minimizes the false-positives that create unnecessary anxiety and unwanted invasive procedures.

[0045] The present application provides robust approaches for detecting markers of cancer (mutations, expression, splice variant, translocation, copy number, and/or methylation changes) using either qPCR or dPCR readout using protocols that are amenable to automation and work on readily available commercial instruments. The approach provides advantages in being integrated and convenient for laboratory setup, allowing for cost reduction, scalability, and fit with medical and laboratory flow in a CLIA-compatible automated setting. The benefit in lives saved world-wide would be of incalculable value.

BRIEF DESCRIPTION OF THE DRAWINGS

[0046] Figure 1A-B illustrates a conditional logic tree for an early detection colorectal cancer test based on analysis of a patient's blood sample. Figure 1A illustrates a one-step colorectal cancer assay using 24 markers at average sensitivity of 50%. Figure 1B illustrates a two-step colorectal cancer assay using 24 markers in the first step at average sensitivity of 50%,

and 48 markers in a second step. Figures 1C-D illustrate a conditional logic tree for a two-step assay for early detection pan-oncology pan-oncology cancer test based on analysis of a patient's blood sample. Figure 1C illustrates a two-step pan-oncology assay using 96 group-specific markers at average sensitivity of 50% in the first step, followed by 1 or 2 groups of 64 type-specific markers each at average sensitivity of 50% in the second step. Figure 1D illustrates a two-step pan-oncology assay using 96 group-specific markers at average sensitivity of 66% in the first step, followed by 1 or 2 groups of 64 type-specific markers each at average sensitivity of 66% in the second step.

[0047] Figure 2 illustrates exPCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify target(s) and/or low-level mutations.

[0048] Figure 3 illustrates exPCR-LDR-qPCR carryover prevention reaction with UniTaq detection to identify or relatively quantify target(s) and/or low-level mutations.

[0049] Figure 4 illustrates exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify target(s) and/or low-level mutations.

[0050] Figure 5 illustrates exPCR-qPCR carryover prevention reaction with UniTaq detection to identify or relatively quantify target(s) and/or low-level mutations.

[0051] Figure 6 illustrates a variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify target(s) and/or low-level mutations.

[0052] Figure 7 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify target(s) and/or low-level mutations.

[0053] Figure 8 illustrates a variation of exPCR-qPCR carryover prevention reaction with UniTaq detection to identify or relatively quantify target(s) and/or low-level mutations.

[0054] Figure 9 illustrates exPCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0055] Figure 10 illustrates a variation of exPCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0056] Figure 11 illustrates exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0057] Figure 12 illustrates a variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0058] Figure 13 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0059] Figure 14 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0060] Figure 15 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0061] Figure 16 illustrates another variation of exPCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0062] Figure 17 illustrates another variation of exPCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0063] Figure 18 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0064] Figure 19 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0065] Figure 20 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0066] Figure 21 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0067] Figure 22 illustrates another variation of exPCR-qPCR carryover prevention reaction with Taqman™ detection to identify or relatively quantify low-level methylation.

[0068] Figure 23 illustrates RT-PCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate translocation events at the mRNA level.

[0069] Figure 24 illustrates RT-PCR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate translocation events at the mRNA level.

[0070] Figure 25 illustrates RT-PCR-PCR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate translocation events at the mRNA level.

[0071] Figure 26 illustrates RT-PCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate RNA copy number.

[0072] Figure 27 illustrates RT-PCR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate RNA copy number.

[0073] Figure 28 illustrates RT-PCR-PCR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate RNA copy number.

[0074] Figure 29 illustrates Ligation-RT-PCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate miRNA.

[0075] Figure 30 illustrates Ligation-RT-PCR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate miRNA.

[0076] Figure 31 illustrates RT-PCR-LDR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate miRNA.

[0077] Figure 32 illustrates RT-PCR-qPCR carryover prevention reaction with Taqman™ detection to detect and enumerate miRNA.

[0078] Figures 33A-B illustrate results for calculated overall Sensitivity and Specificity for a 24-marker assay, where the average individual marker sensitivity is 50% (Figure 33A), and the average individual marker false-positive rate is from 2% to 5% (Figure 33B).

[0079] Figures 34A-B illustrate results for calculated overall Sensitivity and Specificity for a 36-marker assay, where the average individual marker sensitivity is 50% (Figure 34A), and the average individual marker false-positive rate is from 2% to 5% (Figure 34B).

[0080] Figures 35A-B illustrate results for calculated overall Sensitivity and Specificity for a 48-marker assay, where the average individual marker sensitivity is 50% (Figure 35A), and the average individual marker false-positive rate is from 2% to 5% (Figure 35B).

[0081] Figures 36A-B illustrate results for calculated overall Sensitivity and Specificity for a 96-marker assay, where the average individual marker sensitivity is 50% (Figure 36A), and the average individual marker false-positive rate is from 2% to 5% (Figure 36B).

[0082] Figures 37A-B illustrate the ROC curve for a 48-marker assay, where the average individual marker sensitivity is 50%, as well as the calculated AUC, when the average number of molecules per marker in the blood ranges from 150 to 600 molecules. For Figures 37A and 37B, the calculations are based on an average individual marker false-positive rate of 2% and 3%, respectively.

[0083] Figures 38A-B illustrate the ROC curve for a 48-marker assay, where the average individual marker sensitivity is 50%, as well as the calculated AUC, when the average number of molecules per marker in the blood ranges from 150 to 600 molecules. For Figures 38A and 38B, the calculations are based on an average individual marker false-positive rate of 4% and 5%, respectively.

[0084] Figures 39A-B provide a list of blood-based, colon cancer-specific microRNA markers derived through analysis of TCGA microRNA datasets, which may be present in exosomes or other protected state in the blood.

[0085] Figures 40A-X provide a list of blood-based, colon cancer-specific ncRNA and lncRNA markers, which may be present in exosomes or other protected state in the blood.

[0086] Figures 41A-C provide a list of candidate blood-based colon cancer-specific exon transcripts that may be enriched in in exosomes or other protected state in the blood.

[0087] Figures 42A-J provide a list of cancer proteins markers, identified through, mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from Colorectal tumors, which may be identified in

the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0088] Figure 43 provides a list of protein markers that can be secreted by Colorectal tumors into the blood.

[0089] Figures 44A-Y provide a list of primary CpG sites that are Colorectal Cancer and Colon-tissue specific markers, that may be used to identify the presence of colorectal cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0090] Figures 45A-P provide a list of chromosomal regions or sub-regions within which are primary CpG sites that are Colorectal Cancer and Colon-tissue specific markers, that may be used to identify the presence of colorectal cancer from cfDNA, or DNA within exosomes, or DNA in other protected state (such as within CTCs) within the blood.

[0091] Figures 46A-B illustrate results for calculated overall Sensitivity and Specificity for a 24-marker assay, where the average individual marker sensitivity is 50%, and the average individual marker false-positive rate is from 2% to 5%; including one marker with a sensitivity at 90% (Figure 46A) and a 10% (Figure 46B) false-positive rate.

[0092] Figures 47A-B illustrate results for calculated overall Sensitivity and Specificity for a 24-marker assay, where the average individual marker sensitivity is 50%, and the average individual marker false-positive rate is from 2% to 5%; including two markers with a sensitivity at 90% (Figure 47A) and a 10% (Figure 47B) false-positive rate.

[0093] Figures 48A-B illustrate results for calculated overall Sensitivity and Specificity for a 48-marker assay, where the average individual marker sensitivity is 50%, and the average individual marker false-positive rate is from 2% to 5%; including one marker with a sensitivity at 90% (Figure 48A) and a 10% (Figure 48B) false-positive rate.

[0094] Figures 49A-B illustrate results for calculated overall Sensitivity and Specificity for a 48-marker assay, where the average individual marker sensitivity is 50%, and the average individual marker false-positive rate is from 2% to 5%; including two markers with a sensitivity at 90% (Figure 49A) and a 10% (Figure 49B) false-positive rate.

[0095] Figures 50A-B illustrate results for calculated overall Sensitivity and Specificity for a 24-marker assay, where the average individual marker sensitivity is 66% (Figure 50A), and the average individual marker false-positive rate is from 2% to 5% (Figure 50B).

[0096] Figures 51A-B illustrate results for calculated overall Sensitivity and Specificity for a 36-marker assay, where the average individual marker sensitivity is 66% (Figure 51A), and the average individual marker false-positive rate is from 2% to 5% (Figure 51B).

[0097] Figures 52A-B illustrate results for calculated overall Sensitivity and Specificity for a 48-marker assay, where the average individual marker sensitivity is 66% (Figure 52A), and the average individual marker false-positive rate is from 2% to 5% (Figure 52B).

[0098] Figure 53 provides a list of blood-based, solid tumor-specific ncRNA and lncRNA markers, which may be present in exosomes or other protected state in the blood.

[0099] Figures 54A-F provide a list of candidate blood-based solid tumor-specific exon transcripts that may be enriched in in exosomes or other protected state in the blood.

[0100] Figures 55A-H provide a list of cancer proteins markers, identified through, mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from solid tumors, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0101] Figures 56A-S provide a list of primary CpG sites that are solid-tumor and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0102] Figures 57A-J provide a list of chromosomal regions or sub-regions within which are primary CpG sites that are solid-tumor and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0103] Figures 58 provide a list of cancer proteins markers, identified through, mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0104] Figures 59A-S provide a list of primary CpG sites that are colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0105] Figures 60A-J provide a list of chromosomal regions or sub-regions within which are primary CpG sites that are colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0106] Figures 61A-C provide a list of primary CpG sites that are breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, or uterine carcinosarcoma and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0107] Figures 62A-B provide a list of chromosomal regions or sub-regions within which are primary CpG sites that are breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, or uterine carcinosarcoma and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0108] Figure 63 provides a list of primary CpG sites that are lung adenocarcinoma, lung squamous cell carcinoma, or head & neck squamous cell carcinoma and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0109] Figure 64 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are lung adenocarcinoma, lung squamous cell carcinoma, or head & neck squamous cell carcinoma and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0110] Figure 65 provides a list of primary CpG sites that are prostate adenocarcinoma or invasive urothelial bladder cancer and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0111] Figure 66 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are prostate adenocarcinoma or invasive urothelial bladder cancer and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0112] Figure 67 provides a list of blood-based, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma-specific ncRNA and lncRNA markers, which may be present in exosomes or other protected state in the blood.

[0113] Figures 68A-E provide a list of candidate blood-based liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma-specific exon transcripts that may be enriched in exosomes or other protected state in the blood.

[0114] Figures 69A-B provide a list of cancer proteins markers, identified through, mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0115] Figures 70A-E provide a list of primary CpG sites that are liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0116] Figures 71A-C provide a list of chromosomal regions or sub-regions within which are primary CpG sites that are liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma and tissue-specific markers, that may be used to identify the presence of solid-tumor cancer from cfDNA, or DNA within exosomes, or DNA in another protected state (such as within CTCs) within the blood.

[0117] Figure 72 illustrates the real-time PCR amplification plots obtained in the pixel Bisulfite-PCR-LDR-qPCR experiments to enumerate single molecules of methylated DNA in the presence of an excess of unmethylated DNA (Roche DNA).

[0118] Figure 73 illustrates the real-time PCR amplification plots obtained in a multiplexed detection of 10 CRC methylation markers by Bisulfite-PCR-LDR-qPCR, using HT29 cell line DNA, with an average of 20 molecules each marker in 10,000 molecules of normal, *e.g.* unmethylated DNA (Roche DNA).

[0119] Figure 74 illustrates the real-time PCR amplification plots obtained in a multiplexed detection of 7 CRC methylation markers by Bisulfite-exPCR-LDR-qPCR, using HT29 cell line DNA, with an average of 30 molecules each marker in 3,000 molecules of normal, *e.g.* unmethylated DNA (Roche DNA).

[0120] Figures 75A-B illustrate the real-time PCR amplification plots obtained in a multiplexed detection of 7 CRC methylation markers by Bisulfite-exPCR-LDR-qPCR, using cfDNA isolated from CRC (Figure 75A) and Normal (Figure 75B) plasma.

[0121] Figures 76A-B illustrate the real-time PCR amplification plots obtained in a multiplexed detection of 7 CRC methylation markers by Bisulfite-exPCR-LDR-qPCR, using cfDNA isolated from CRC (Figure 76A) and Normal (Figure 76B) plasma.

[0122] Figures 77A-B illustrate the real-time PCR amplification plots obtained in a multiplexed detection of 7 CRC methylation markers by Bisulfite-exPCR-LDR-qPCR, using cfDNA isolated from CRC (Figure 77A) and Normal (Figure 77B) plasma.

[0123] Figures 78A-B illustrate the real-time PCR amplification plots obtained in a multiplexed detection of 20 CRC methylation markers by Bisulfite-exPCR-LDR-qPCR, using HT29 cell line DNA, with 1,500 genome equivalents of HT29 cell line DNA in 7,500 genome equivalents of normal, *e.g.* unmethylated DNA (Roche DNA; Figure 78A) compared with 7,500 genome equivalents of normal, *e.g.* unmethylated DNA (Figure 78B).

[0124] Figures 79A-B illustrate the real-time PCR amplification plots obtained in a multiplexed detection of 20 CRC methylation markers by Bisulfite-exPCR-LDR-qPCR, using reverse primers with tails, using HT29 cell line DNA, with 200 genome equivalents of HT29 cell line DNA in 7,500 genome equivalents of normal, *e.g.* unmethylated DNA (Roche DNA; Figure 79A) compared with 7,500 genome equivalents of normal, *e.g.* unmethylated DNA (Figure 79B).

[0125] Figures 80A-B illustrate the real-time PCR amplification plots obtained in a multiplexed detection of 20 CRC methylation markers by Bisulfite-exPCR-LDR-qPCR, using reverse primers without tails, using HT29 cell line DNA, with 200 genome equivalents of HT29 cell line DNA in 7,500 genome equivalents of normal, *e.g.* unmethylated DNA (Roche DNA; Figure 80A) compared with 7,500 genome equivalents of normal, *e.g.* unmethylated DNA (Figure 80B).

DETAILED DESCRIPTION

A Universal Design for Early Detection of Cancer Using "Cancer Marker Load"

[0126] The most cost-effective early cancer detection test may combine an initial multiplexed coupled amplification and ligation assay to determine "cancer load". For cancer detection, this would achieve > 95% sensitivity for all cancers (pan-oncology), at > 97% specificity.

[0127] Several flow charts for a cancer tumor load assay is illustrated in Figure 1. In its simplest form, the assay would be a one-step assay to identify individuals with early colorectal cancer (CRC). A blood sample is fractionated into plasma and other components as needed, a set of 24 markers with average sensitivity of 50% are assayed, and the results are recorded (Figure 1A). For example, an initial multiplexed PCR/LDR screening assay scoring for mutation, methylation, miRNA, mRNA, alternative splicing, and/or translocations identifies those samples with positive results. The physician is not concerned with which specific markers are positive but gives a simple directive. Those patients with 0-2 markers positive are told not to worry, go home, you are cancer-free. Those patients with ≥ 5 of 24 markers positive are directed to get a

colonoscopy. Those patients with an intermediate number of positive markers (3-4) are instructed to come back in 3-6 months for retesting. Thus, the test is based on the overall cancer marker load and not dependent on the specific markers that teat positive.

[0128] In an advanced version of the test, a two-step assay would be performed to identify if the patient has colorectal cancer. The rationale for a two-step test is to initially cast a wide net to maximize sensitivity in identifying the most individuals with potential cancer, followed by a second step only on the positive samples (which contain both true and false-positives) to maximize specificity, eliminate virtually all the false-positives, and hone in on those individuals most likely to have cancer. In the first step, a blood sample is fractionated into plasma and other components as needed, followed by an assay to interrogate an initial set of 24 markers with an average sensitivity of 50% (Figure 1B). The first step assay can employ multiplexed PCR/LDR, or digital PCR screening to score for mutation, methylation, miRNA, mRNA, alternative splicing, and/or translocations events. As in the one-step assay, patients with 0-2 markers positive are presumed to be cancer-free. On the other hand, patients with ≥ 3 markers positive will undergo a second step, wherein 48 (new) markers are assayed and scored as follows: 0-3 positive markers are considered cancer-free; 4-5 positive markers are advised to come back in 3-6 months for retesting; ≥ 6 positive markers are directed to go get a colonoscopy.

[0129] In a pan-oncology version of the test, in the first step the assay would screen 96 markers, wherein on average ≥ 36 such markers would exhibit an average sensitivity of 50% for most major cancers (see Figure 1C). These cancers would cluster to certain groups, which include: Group 1 (Colorectal, Stomach, Esophagus); Group 2 (Breast, Endometrial, Ovarian, Cervical, Uterine); Group 3 (Lung, Head & Neck); Group 4 (Prostate, Bladder), and Group 5 (Liver, Pancreatic, Gall Bladder). Patients with 0-4 markers positive are presumed to be cancer-free, while patients with ≥ 5 markers positive will undergo a second step. Presumptive positive samples are then assayed in the second step, testing 1 or 2 groups, using 64 markers per group, wherein on average ≥ 36 such markers would exhibit an average sensitivity of 50% for each specific type of cancer within that group, including using tissue-specific markers to validate the initial result and to identify tissue of origin. Results are scored as follows: 0-3 positive markers are considered cancer-free; 4 positive markers are advised to come back in 3-6 months for retesting; ≥ 5 positive markers are directed to go to imaging that matches the type(s) of cancer most likely to be the tissue of origin. For higher sensitivities, both the initial 96 markers in the first step, and the group-specific markers in the second step would have average sensitivity of 66% (Figure 1D). The physician may then order targeted sequencing to further guide treatment decisions for the patient.

[0130] The present application is directed to a universal diagnostic approach that seeks to combine the best features of digital polymerase chain reaction (PCR), or quantitative polymerase chain reaction (qPCR), with bisulfite conversion, ligation detection reaction (LDR), and quantitative detection of multiple disease markers, *e.g.*, cancer markers.

Multiplexing, avoiding False-Positives, and Carryover Protection

[0131] There is a technical challenge of distinguishing true signal generated from the desired disease-specific nucleic acid differences vs. false signal generated from normal nucleic acids present in the sample vs. false signal generated in the absence of the disease-specific nucleic acid differences (*i.e.* somatic mutations).

[0132] A number of solutions to these challenges are presented below, but they share some common themes.

[0133] The first theme is multiplexing. PCR works best when primer concentration is relatively high, from 50nM to 500nM, limiting multiplexing. Further, the more PCR primer pairs added, the chances of amplifying incorrect products or creating primer-dimers increase exponentially. In contrast, for LDR probes, low concentrations on the order of 4 nM to 20 nM are used, and probe-dimers are limited by the requirement for adjacent hybridization on the target to allow for a ligation event. Use of low concentrations of gene-specific PCR primers or LDR probes containing universal primer sequence "tails" allows for subsequent addition of higher concentrations of universal primers to achieve proportional amplification of the initial PCR or LDR products. Another way to avoid or minimize false PCR amplicons or primer dimers is to use PCR primers containing a few extra bases and a blocking group, which is liberated to form a free 3'OH by cleavage with a nuclease only when hybridized to the target, *e.g.*, a ribonucleotide base as the blocking group and RNase H2 as the cleaving nuclease.

[0134] The second theme is fluctuations in signal due to low input target nucleic acids. Often, the target nucleic acid originated from a few cells, either captured as CTCs, or from tumor cells that underwent apoptosis and released their DNA as small fragments (140 – 160 bp) in the serum. Under such conditions, it is preferable to perform some level of proportional amplification to avoid missing the signal altogether or reporting inaccurate copy number due to fluctuations when distributing small numbers of starting molecules into individual wells (for real-time, or droplet PCR quantification). As long as these initial amplifications are kept at a reasonable level (approximately 12 to 20 cycles), the risk of carryover contamination during opening of the tube and distributing amplicons for subsequent detection/quantification (using real-time, or droplet PCR) is minimized. Other schemes use even lower amounts of limited amplifications, (approximately 8 to 12 cycles).

[0135] The third theme is target-independent signal, also known as "No Template Control" (NTC). This arises from either polymerase or ligase reactions that occur in the absence of the correct target. Some of this signal may be minimized by judicious primer design. For ligation reactions, the 5'→ 3' nuclease activity of polymerase may be used to liberate the 5' phosphate of the downstream ligation primer (only when hybridized to the target), so it is suitable for ligation. Further specificity for distinguishing presence of a low-level mutation using LDR may be achieved by: (i) using upstream mutation-specific LDR probes containing a mismatch in the 2nd or 3rd position from the 3'OH base, (ii) using LNA or PNA probes to wild-type sequence that would reduce hybridization of mutation-specific LDR probes to wild-type sequences, (iii) using LDR probes to wild-type sequence that (optionally) ligate but do not undergo additional amplification, and (iv) using upstream LDR probes containing a few extra bases and a blocking group, which is liberated to form a free 3'OH by cleavage with a nuclease only when hybridized to the complementary target (*e.g.*, RNase H2 and a ribonucleotide base). Similar approaches for improving the specificity for distinguishing presence of a low-level mutation using PCR may be achieved by: (i) using mutation-specific PCR Primers containing a mismatch in the 2nd or 3rd position from the 3'OH base, (ii) using LNA or PNA probes to wild-type sequence that would reduce hybridization of mutation-specific PCR primers to wild-type sequences, (iii) using PCR primers to wild-type sequence that are blocked and do not undergo additional amplification, and (iv) using upstream PCR primers containing a few extra bases and a blocking group, which is liberated to form a free 3'OH by cleavage with a nuclease only when hybridized to the complementary target (*e.g.*, RNase H2 and a ribonucleotide base).

[0136] The fourth theme is either suppressed (reduced) amplification or incorrect (false) amplification due to unused primers in the reaction. One approach to eliminate such unused primers is to capture genomic or target or amplified target DNA on a solid support, allow ligation probes to hybridize and ligate, and then remove probes or products that are not hybridized. Alternative solutions include pre-amplification, followed by subsequent nested LDR and/or PCR steps, such that there is a second level of selection in the process.

[0137] The fifth theme is carryover prevention. Carryover signal may be eliminated by standard uracil incorporation during the universal PCR amplification step, and by using UDG (and optionally AP endonuclease) in the pre-amplification workup procedure. Incorporation of carryover prevention is central to the methods of the present application as described in more detail below. The initial PCR amplification is performed using incorporation of uracil. The LDR reaction is performed with LDR probes lacking uracil. Thus, when the LDR products are subjected to real-time PCR quantification, addition of UDG destroys the initial PCR products, but not the LDR products. Further, since LDR is a linear process and the tag primers use

sequences absent from the human genome, accidental carryover of LDR products back to the original PCR will not cause template-independent amplification. Additional schemes to provide carryover prevention with methylated targets include use of restriction endonucleases to destroy unmethylated DNA prior to PCTR amplification, or capturing and enriching methylated DNA using methyl-specific DNA binding proteins or antibodies.

[0138] The sixth theme is achieving even amplification of many mutation-specific or methylation-specific targets in the multiplexed reaction. One approach, as already described above, is to perform limited initial PCR amplifications (8 to 12, or 12 to 20 cycles). However, sometimes different products amplify at different rates, especially when using mutation-or methylation-specific primers, or when using blocking LNA or PNA probes or other means to suppress amplification of wild-type DNA. This is because a regular PCR reaction has both forward and reverse primers working simultaneously. Although there may be preferential amplification using as an example a forward methylation-specific primer (*i.e.* after bisulfite treatment), the reverse primer will amplify both methylated and un-methylated DNA (again, after bisulfite treatment), and thus will magnify differences in initial rates of forward primer amplification. Further, and this also holds when using mutation-specific forward primers, the use of non-selecting reverse primers means that initial amplification products still contain substantial amounts of wild-type DNA sequence, which may lead to undesired false-positives in subsequent amplification steps. One approach is to perform an initial single-sided linear amplification, using primers that amplify only one strand of target DNA. This is particularly useful when amplifying bisulfite-treated DNA, where the two resultant strands are no longer complementary to each other. An important variation of this theme destroys the initial target DNA after the linear amplification step. This may be achieved by incorporating one or more modified nucleotides, such as α-thio-dNTPs, that protect the initial extension products (but not the original cfDNA or genomic DNA) from exonuclease I digestion. When using bisulfite converted DNA, after performing the initial single-sided linear amplification (*i.e.,* the polymerase extension reaction) with regular dNTP's (*i.e.* NO dUTP), the original bisulfite-converted DNA may be destroyed using UDG.

Methods of Identifying Cancer Markers

[0139] A first aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences of other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. The method involves providing a sample containing one or more

parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules are then provided. One or more primary oligonucleotide primer sets are also provided. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The sample, the one or more first primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase extension reaction mixtures and for carrying out one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising nucleotide sequences complementary to the target nucleotide sequence. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more second primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixtures, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures. The method further comprises subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more first polymerase chain reaction products comprising the target nucleotide sequence or a complement thereof. One or more oligonucleotide probe sets are then provided. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on a complementary target nucleotide sequence of a secondary extension product. The one or more first polymerase

chain reaction products are blended with a ligase, and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures. The one or more ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets are ligated together, when hybridized to complementary sequences, to form ligated product sequences in the ligation reaction mixtures wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion. The method further includes providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more second polymerase chain reaction products. The method further comprises detecting and distinguishing the one or more second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

[0140] Figures 2 and 3 illustrate various embodiments of this aspect of the present application, abbreviated as exPCR-LDR-qPCR carryover prevention reaction to detect low-level mutations (exPCR is an abbreviation for one-sided extension using primers to one strand of a locus, followed by PCR – using either the same primers in the initial extension, or additional primers for the PCR step). Genomic or cfDNA is isolated (Figure 2, step A), and the isolated DNA sample is treated with UDG to digest dU containing nucleic acid molecules that may be present in the sample (Figure 2, step B). Suitable enzymes include, without limitation, *E. coli* uracil DNA glycosylase (UDG), Antarctic Thermolabile UDG, or Human single-strand-selective monofunctional uracil-DNA Glycosylase (hSMUG1). The regions of interest are selectively

extended using locus-specific upstream primers, a blocking LNA or PNA probe comprising wild-type sequence, and a deoxynucleotide mix that includes dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the mutation, and suitable for polymerase extension (Figure 2 or 3, step B; *see e.g.*, Dobosy et. al. "RNase H-Dependent PCR (rhPCR): Improved Specificity and Single Nucleotide Polymorphism Detection Using Blocked Cleavable Primers," *BMC Biotechnology* 11(80):1011 (2011), which is hereby incorporated by reference in its entirety). A blocking LNA or PNA probe comprising wild-type sequence that partially overlaps with the upstream PCR primer will preferentially compete in binding to wild-type sequence over the upstream primer, but not as much to mutant DNA, and thus suppresses extension of wild-type DNA during each round of primer extension. Sample is optionally aliquoted into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the locus-specific downstream primers are added, followed by limited (8 to 20 cycles) or full (20-40 cycles) PCR. Optionally, the downstream primers contain identical 8-11 base tails to prevent primer dimers. Further, such tails provide the option for asymmetric PCR at the end of the PCR cycles, by raising the hybridization temperature above that for the forward primers, but at or below that for the reverse primers – which at 8-11 bases longer will have higher Tm values. This generates more bottom strand products, which are suitable substrates for the subsequent LDR step. In an alternative embodiment, the initial extension products incorporate one or more modified nucleotides, such as α-thio-dNTPs, that protect the initial extension products (but not the original cfDNA or genomic DNA) from exonuclease I digestion. After exonuclease I digestion, the down-stream locus-specific primers (optionally containing identical 8-11 base tails) are added, again followed by limited (8 to 20 cycles) or full (20-40 cycles) PCR. The amplified products contain dU as shown in Figure 2 or 3, step D, which allows for subsequent treatment with UDG or a similar enzyme for carryover prevention.

[0141] As shown in Figure 2 step E, target-specific oligonucleotide probes are hybridized to the amplified products and ligase (filled circle) covalently seals the two oligonucleotides together when hybridized to their complementary sequence. In this embodiment, the upstream oligonucleotide probe having a sequence specific for detecting the mutation of interest further contains a 5' primer-specific portion (Ai) to facilitate subsequent detection of the ligation product. Once again, the presence of blocking LNA or PNA probe comprising wild-type sequence suppresses ligation to wild-type target sequence if present after the enrichment of mutant sequence during the PCR amplification step. The downstream oligonucleotide probe,

having a sequence common to both mutant and wild-type sequences contains a 3' primer-specific portion (Ci') that, together with the 5' primer specific portion (Ai) of the upstream probe having a sequence specific for detecting the mutation, permit subsequent amplification and detection of only mutant ligation products. As illustrated in step E of Figure 2, another layer of specificity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream ligation probe. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to generate a ligation competent 3'OH group (Figure 2, step D).

[0142] As shown in Figure 2, step F, target-specific oligonucleotide probes are hybridized to the amplified products and ligase (filled circle) covalently seals the two oligonucleotides together when hybridized to their complementary sequence. The upstream oligonucleotide probe contains a 5' primer-specific portion (Ai) and the downstream oligonucleotide probe contains a 3' primer-specific portion (Ci') that permits subsequent amplification of the ligation product. Following ligation, the ligation products are aliquoted into separate wells, micro-pores or droplets containing one or more tag-specific primer pairs, each pair comprising matched primers Ai and Ci, treated with UDG or similar enzyme to remove dU containing amplification products or contaminants, PCR amplified, and detected. As shown in Figures 2, steps G & H, detection of the ligation product can be carried out using traditional TaqMan™ detection assay (*see* U.S. Patent No. 6,270,967 to Whitcombe et al., and U.S. Patent No. 7,601,821 to Anderson et al., which are hereby incorporated by reference in their entirety). For detection using TaqMan™ an oligonucleotide probe spanning the ligation junction is used in conjunction with primers suitable for hybridization on the primer-specific portions of the ligation products for amplification and detection. The TaqMan™ probe contains a fluorescent reporter group on one end (F1) and a quencher molecule (Q) on the other end that are in close enough proximity to each other in the intact probe that the quencher molecule quenches fluorescence of the reporter group. During amplification, the TaqMan™ probe and upstream primer hybridize to their complementary regions of the ligation product. The 5'→ 3' nuclease activity of the polymerase extends the hybridized primer and liberates the fluorescent group of the TaqMan™ probe to generate a detectable signal (Figure 2, step H). In a preferred embodiment, the Taqman probe contains a second quencher group (ZEN) about 9 bases in from the fluorescent reporter group, and the probe is designed such that the ZEN group is at or adjacent to the mutant base. Use of dUTP during the amplification reaction generates products containing dU, which can subsequently be destroyed using UDG for carryover prevention.

[0143] As shown in Figure 3 step D, target-specific oligonucleotide probes are hybridized to the amplified products and ligase (filled circle) covalently seals the two

oligonucleotides together when hybridized to their complementary sequence. In this embodiment, the upstream oligonucleotide probe having a sequence specific for detecting the mutation of interest further contains a 5' primer-specific portion (Ai) to facilitate subsequent detection of the ligation product. Once again, the presence of blocking LNA or PNA probe comprising wild-type sequence suppresses ligation to wild-type target sequence if present after the enrichment of mutant sequence during the PCR amplification step. The downstream oligonucleotide probe, having a sequence common to both mutant and wild-type sequences contains a 3' primer-specific portion (Bi-Ci') that, together with the 5' primer specific portion (Ai) of the upstream probe having a sequence specific for detecting the mutation, permit subsequent amplification and detection of only mutant ligation products. As illustrated in step D of Figure 3, another layer of specificity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream ligation probe. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to generate a ligation competent 3'OH group (Figure 3, step D).

[0144] In this embodiment, the ligation probes are designed to contain UniTaq primer and tag sequences to facilitate detections. The UniTaq system is fully described in U.S. Patent Application Publication No. 2011/0212846 to Spier, which is hereby incorporated by reference in its entirety. The UniTaq system involves the use of three unique "tag" sequences, where at least one of the unique tag sequences (Ai) is present in the first oligonucleotide probe, and the second and third unique tag portions (Bi' and Ci') are in the second oligonucleotide probe sequence as shown in Figure 3, step D & E. Upon ligation of oligonucleotide probes in a probe set, the resulting ligation product will contain the Ai sequence—target specific sequences—Bi' sequence—Ci' sequence. The essence of the UniTaq approach is that both oligonucleotide probes of a ligation probe set need to be correct in order to get a positive signal, which allows for highly multiplexed nucleic acid detection. For example, and as described herein, this is achieved by requiring hybridization of two parts, *i.e.,* two of the tags, to each other.

[0145] Prior to detecting the ligation product, the sample is treated with UDG to destroy original target amplicons allowing only authentic ligation products to be detected. Following ligation, the ligation products are aliquoted into separate wells, micro-pores or droplets containing one or more tag-specific primer pairs. For the detection step, the ligation product containing Ai (a first primer-specific portion), Bi' (a UniTaq detection portion), and Ci' (a second primer-specific portion) is primed on both strands using a first oligonucleotide primer having the same nucleotide sequence as Ai, and a second oligonucleotide primer that is complementary to Ci' (*i.e.,* Ci). The first oligonucleotide primer also includes a UniTaq detection probe (Bi) that has a detectable label F1 on one end and a quencher molecule (Q) on

the other end (F1-Bi-Q-Ai). Optionally positioned proximal to the quencher is a polymerase-blocking unit, *e.g.*, HEG, THF, Sp-18, ZEN, or any other blocker known in the art that is sufficient to stop polymerase extension. In another embodiment, a ZEN quencher group is also positioned about 9 bases from the fluorescent reporter group to assure more complete quenching. PCR amplification results in the formation of double stranded products as shown in Figure 3, step G). In this example, a polymerase-blocking unit prevents a polymerase from copying the 5' portion (Bi) of the first universal primer, such that the bottom strand of product cannot form a hairpin when it becomes single-stranded. Formation of such a hairpin would result in the 3' end of the stem annealing to the amplicon such that polymerase extension of this 3' end would terminate the PCR reaction.

[0146] The double stranded PCR products are denatured, and when the temperature is subsequently decreased, the upper strand of product forms a hairpin having a stem between the 5' portion (Bi) of the first oligonucleotide primer and portion Bi' at the opposite end of the strand (Figure 3, step H). Also, during this step, the second oligonucleotide primer anneals to the 5'-primer specific portion (Ci') of the hairpinned product. Upon extension of the second universal primer in step H, 5' nuclease activity of the polymerase cleaves the detectable label D1 or the quencher molecule from the 5' end of the amplicon, thereby increasing the distance between the label and the quencher and permitting detection of the label.

[0147] The ligation reaction used in the methods of the present application is well known in the art. Ligases suitable for ligating oligonucleotide probes of a probe set together (optionally following cleavage of a 3' ribose and blocking group on the first oligonucleotide probe, or the 5' flap on the second oligonucleotide probe) include, without limitation *Thermus aquaticus* ligase, *E. coli* ligase, T4 DNA ligase, T4 RNA ligase, *Taq* ligase, 9 N ligase, and *Pyrococcus* ligase, or any other thermostable ligase known in the art. In accordance with the present application, the nuclease-ligation process of the present application can be carried out by employing an oligonucleotide ligation assay (OLA) reaction (*see* Landegren, et al., "A Ligase-Mediated Gene Detection Technique," *Science* 241:1077-80 (1988); Landegren, et al., "DNA Diagnostics -- Molecular Techniques and Automation," *Science* 242:229-37 (1988); and U.S. Patent No. 4,988,617 to Landegren, et al., which are hereby incorporated by reference in their entirety), a ligation detection reaction (LDR) that utilizes one set of complementary oligonucleotide probes (*see e.g.*, WO 90/17239 to Barany et al, which is hereby incorporated by reference in its entirety), or a ligation chain reaction (LCR) that utilizes two sets of complementary oligonucleotide probes *see e.g.*, WO 90/17239 to Barany et al, which is hereby incorporated by reference in its entirety).

[0148] The oligonucleotide probes of a probe sets can be in the form of ribonucleotides, deoxynucleotides, modified ribonucleotides, modified deoxyribonucleotides, peptide nucleotide analogues, modified peptide nucleotide analogues, modified phosphate-sugar-backbone oligonucleotides, nucleotide analogs, and mixtures thereof.

[0149] The hybridization step in the ligase detection reaction, which is preferably a thermal hybridization treatment, discriminates between nucleotide sequences based on a distinguishing nucleotide at the ligation junctions. The difference between the target nucleotide sequences can be, for example, a single nucleic acid base difference, a nucleic acid deletion, a nucleic acid insertion, or rearrangement. Such sequence differences involving more than one base can also be detected. Preferably, the oligonucleotide probe sets have substantially the same length so that they hybridize to target nucleotide sequences at substantially similar hybridization conditions.

[0150] Ligase discrimination can be further enhanced by employing various probe design features. For example, an intentional mismatch or nucleotide analogue (*e.g.*, Inosine, Nitroindole, or Nitropyrrole) can be incorporated into the first oligonucleotide probe at the 2nd or 3rd base from the 3' junction end to slightly destabilize hybridization of the 3' end if it is perfectly matched at the 3' end, but significantly destabilize hybridization of the 3' end if it is mis-matched at the 3' end. This design reduces inappropriate misligations when mutant probes hybridize to wild-type target. Alternatively, RNA bases that are cleaved by RNases can be incorporated into the oligonucleotide probes to ensure template-dependent product formation. For example, Dobosy et. al. "RNase H-Dependent PCR (rhPCR): Improved Specificity and Single Nucleotide Polymorphism Detection Using Blocked Cleavable Primers," *BMC Biotechnology* 11(80): 1011 (2011), which is hereby incorporated by reference in its entirety, describes using an RNA-base close to the 3' end of an oligonucleotide probe with 3'-blocked end, and cutting it with RNase H2 generating a PCR-extendable and ligatable 3'-OH. This approach can be used to generate either ligation-competent 3'OH (for standard DNA ligases) or 5'-P, or both, in the latter case, provided a ligase that can ligate 5'-RNA base is utilized.

[0151] Other possible modifications included abasic sites, e.g., internal abasic furan or oxo-G. These abnormal "bases" are removed by specific enzymes to generate ligation-competent 3'-OH or 5'P sites. Endonuclease IV, Tth EndoIV (NEB) will remove abasic residues after the ligation oligonucleotides anneal to the target nucleic acid, but not from a single-stranded DNA. Similarly, one can use oxo-G with Fpg or inosine/uracil with EndoV or Thymine glycol with EndoVIII.

[0152] Ligation discrimination can also be enhanced by using the coupled nuclease-ligase reaction described in WO2013/123220 to Barany et al. or U.S. Patent Application

Publication No. 2006/0234252 to Anderson et al., which are hereby incorporated by reference in their entirety. In this embodiment, the first oligonucleotide probe bears a ligation competent 3' OH group while the second oligonucleotide probe bears a ligation incompetent 5' end (*i.e.*, an oligonucleotide probe without a 5' phosphate). The oligonucleotide probes of a probe set are designed such that the 3'-most base of the first oligonucleotide probe is overlapped by the immediately flanking 5'-most base of the second oligonucleotide probe that is complementary to the target nucleic acid molecule. The overlapping nucleotide is referred to as a "flap". When the overlapping flap nucleotide of the second oligonucleotide probe is complementary to the target nucleic acid molecule sequence and the same sequence as the terminating 3' nucleotide of the first oligonucleotide probe, the phosphodiester bond immediately upstream of the flap nucleotide of the second oligonucleotide probe is discriminatingly cleaved by an enzyme having flap endonuclease (FEN) or 5' nuclease activity. That specific FEN activity produces a novel ligation competent 5' phosphate end on the second oligonucleotide probe that is precisely positioned alongside the adjacent 3' OH of the first oligonucleotide probe to allow ligation of the two probes to occur. In accordance with this embodiment, flap endonucleases or 5' nucleases that are suitable for cleaving the 5' flap of the second oligonucleotide probe prior to ligation include, without limitation, polymerases with 5' nuclease activity such as *E.coli* DNA polymerase and polymerases from *Taq* and *T. thermophilus*, as well as T4 RNase H and *Taq*Exo. In another embodiment, the second probe of the probe set has a 3′ primer-specific portion, a target specific portion, and a 5′ nucleotide sequence, where the 5′ nucleotide sequence is complementary to at least a portion of the 3′ primer-specific portion, and where the 5′ nucleotide sequence hybridizes to its complementary portion of the 3′ primer-specific portion to form a hair-pinned second oligonucleotide probe when the second probe is not hybridized to a target nucleotide sequence.

[0153] For insertions or deletions, incorporation of a matched base or nucleotide analogues (*e.g.*, -amino-dA or 5-propynyl-dC) in the first oligonucleotide probe at the 2nd or 3rd position from the junction improves stability and may improve discrimination of such frameshift mutations from wild-type sequences. For insertions, use of one or more thiophosphate-modified nucleotides downstream from the desired scissile phosphate bond of the second oligonucleotide probe will prevent inappropriate cleavage by the 5' nuclease enzyme when the probes are hybridized to wild-type DNA, and thus reduce false-positive ligation on wild-type target. Likewise, for deletions, use of one or more thiophosphate-modified nucleotides upstream from the desired scissile phosphate bond of the second oligonucleotide probe will prevent inappropriate cleavage by the 5' nuclease enzyme when the probes are hybridized to wild-type DNA, and thus reduce false-positive ligation on wild-type target.

[0154] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. One or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, one or more nucleases capable of digesting nucleic acid molecules not comprising modified nucleotides, and one or more first primary oligonucleotide primer(s) are provided. The one or more first primary oligonucleotide primer(s) comprise a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence. The sample, the one or more first primary oligonucleotide primers, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix that comprises one or more modified nucleotides that protect extension products but not target DNA from nuclease digestion, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase extension reaction mixture and for carrying out one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the target nucleotide sequence. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a first 5' primer-specific portion and a 3' portion that is complementary to a portion of a primary extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a second 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary oligonucleotide primer sets, the one or more nucleases, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting nucleic acid molecules present in the first polymerase chain reaction mixtures, but not

primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more first polymerase chain reaction products comprising the first 5' primer-specific portion, a target-specific nucleotide sequence or a complement thereof, and a complement of the second 5' primer-specific portion. One or more tertiary oligonucleotide primer sets are provided. Each tertiary oligonucleotide primer set comprises (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the first 5' primer-specific portion of the one or more first polymerase chain reaction products and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the one or more first polymerase chain reaction products. The one or more first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures, and the one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU) containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more second polymerase chain reaction products. The method further involves detecting and distinguishing the one or more second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

[0155] Figures 4-8 illustrate various embodiments of this aspect of the present application.

[0156] Figure 4 illustrates an exemplary exPCR-qPCR carryover prevention reaction to detect low-level mutations. Genomic or cfDNA is isolated (Figure 4, step A), and the isolated DNA sample is treated with UDG to digest dU containing nucleic acid molecules that may be present in the sample (Figure 4, step A). The sample is then subject to a linear amplification reaction, *e.g.*, one or more polymerase extension reactions to generate complementary copies of mutation containing regions of interest. The regions of interest are selectively extended using locus-specific upstream primers, a blocking LNA or PNA probe comprising wild-type sequence, and a deoxynucleotide mix that includes one of more modified nucleotides. In this embodiment,

another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the mutation, and suitable for polymerase extension (Figure 4, step B). A blocking LNA or PNA probe comprising wild-type sequence that partially overlaps with the upstream PCR primer will preferentially compete in binding to wild-type sequence over the upstream primer, but not as much to mutant DNA, and thus suppresses extension of wild-type DNA during each round of primer extension. Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step.

[0157] The initial extension products incorporate one or more modified nucleotides, such as α-thio-dNTPs, that protect the initial extension products (but not the original cfDNA or genomic DNA) from exonuclease I digestion (Figure 4, step C). Using just upstream locus-specific primers in the presence of blocking LNA or PNA probes enriches for extension of mutation-containing products with each extension cycle. The exonuclease digestion destroys wild-type DNA present in the original genomic or cfDNA sample, and thus the enriched extension products will not be diluted by subsequent extension or amplification off original wild-type DNA (see step D below).

[0158] As shown in Figure 4 step D, mutation-specific and locus-specific oligonucleotide primers are added to then perform limited cycle nested PCR to amplify the mutation-containing sequence, if present in the sample. In this embodiment, the upstream mutation-specific primer having a sequence specific for detecting the mutation of interest further contains a 5' primer-specific portion (Ai) to facilitate subsequent detection of the nested PCR product. Once again, the presence of blocking LNA or PNA probe comprising wild-type sequence suppresses extension of wild-type target sequence if present after the enrichment of mutant sequence during the initial extension step. The reverse locus-specific primer, having a sequence common to both mutant and wild-type sequences contains a 5' primer-specific portion (Ci) that, together with the 5' primer specific portion (Ai) of the upstream primer having a sequence specific for detecting the mutation, permit subsequent amplification and detection of only mutant PCR products. As illustrated in step D of Figure 4, another layer of specificity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the mutation-specific and locus-specific primers. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to generate a polymerase extension competent 3'OH group (Figure 4, step D). In the initial primer extension (step B) the liberated 3'OH base is a few bases upstream from the mutation position, and thus would extend both wild-type and mutant sequences if cleaved (although the blocking LNA or PNA should limit cleavage of primer

hybridized to wild-type sequence). In contrast, in the nested PCR (step D), the mutation-specific base of the primer is at the 3'OH base, such that extension on wild-type sequence would be less likely, since the base is mismatched. The specificity for polymerase extension of mutant over wild-type sequence may be further improved by: (i) using mutation-specific PCR Primers containing a mismatch in the 2nd or 3rd position from the 3'OH base, (ii) using LNA or PNA probes to wild-type sequence that would reduce hybridization of mutation-specific PCR primers to wild-type sequences, (iii) using PCR primers to wild-type sequence that are blocked and do not undergo additional amplification, and (iv) avoiding G:T or T:G mismatches between primer and wild-type sequence at the 3'OH base.

[0159] As shown in Figure 4 step E, nested PCR products comprise a 5' primer-specific portion (Ai) target-specific sequence, and a 3' primer-specific portion (Ci') that permits subsequent amplification of the nested PCR product. Following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing one or more tag-specific primer pairs, each pair comprising matched primers Ai and Ci, treated with UDG or similar enzyme to remove dU containing amplification products or contaminants, PCR amplified, and detected. As shown in Figures 4, steps F & G, detection of the ligation product can be carried out using traditional TaqMan™ detection assay (*see* U.S. Patent No. 6,270,967 to Whitcombe et al., and U.S. Patent No. 7,601,821 to Anderson et al., which are hereby incorporated by reference in their entirety). For detection using TaqMan™ an oligonucleotide probe spanning the mutation-specific region is used in conjunction with primers suitable for hybridization on the primer-specific portions of the nested PCR products for amplification and detection. The TaqMan™ probe contains a fluorescent reporter group on one end (F1) and a quencher molecule (Q) on the other end that are in close enough proximity to each other in the intact probe that the quencher molecule quenches fluorescence of the reporter group. During amplification, the TaqMan™ probe and upstream primer hybridize to their complementary regions of the nested PCR product. The 5'→ 3' nuclease activity of the polymerase extends the hybridized primer and liberates the fluorescent group of the TaqMan™ probe to generate a detectable signal (Figure 4, step G). In a preferred embodiment, the TaqMan™ probe contains a second quencher group (ZEN) about 9 bases in from the fluorescent reporter group, and the probe is designed such that the ZEN group is at or adjacent to the mutant base. Use of dUTP during the amplification reaction generates products containing dU, which can subsequently be destroyed using UDG for carryover prevention.

[0160] Figure 5 illustrates an another exPCR-qPCR carryover prevention reaction to detect low-level mutations. Genomic or cfDNA is isolated (Figure 5, step A), and the isolated DNA sample is treated with UDG to digest dU containing nucleic acid molecules that may be

present in the sample (Figure 5, step A). The sample is then subject to a linear amplification reaction, *e.g.*, one or more polymerase extension reactions to generate complementary copies of mutation containing regions of interest. The regions of interest are selectively extended using locus-specific upstream primers, a blocking LNA or PNA probe comprising wild-type sequence, and a deoxynucleotide mix that includes one of more modified nucleotides. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the mutation, and suitable for polymerase extension (Figure 5, step B). A blocking LNA or PNA probe comprising wild-type sequence that partially overlaps with the upstream PCR primer will preferentially compete in binding to wild-type sequence over the upstream primer, but not as much to mutant DNA, and thus suppresses extension of wild-type DNA during each round of primer extension. Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step.

[0161] The initial extension products incorporate one or more modified nucleotides, such as α-thio-dNTPs, that protect the initial extension products (but not the original cfDNA or genomic DNA) from exonuclease I digestion (Figure 5, step C). Using just upstream locus-specific primers in the presence of blocking LNA or PNA probes enriches for extension of mutation-containing products with each extension cycle. The exonuclease digestion destroys wild-type DNA present in the original genomic or cfDNA sample, and thus the enriched extension products will not be diluted by subsequent extension or amplification off original wild-type DNA (see step D below).

[0162] As shown in Figure 5 step D, mutation-specific and locus-specific oligonucleotide primers are added to then perform limited cycle nested PCR to amplify the mutation-containing sequence, if present in the sample. In this embodiment, the upstream mutation-specific primer having a sequence specific for detecting the mutation of interest further contains a 5' primer-specific portion (Ai) to facilitate subsequent detection of the nested PCR product. Once again, the presence of blocking LNA or PNA probe comprising wild-type sequence suppresses extension of wild-type target sequence if present after the enrichment of mutant sequence during the initial extension step. The reverse locus-specific primer, having a sequence common to both mutant and wild-type sequences contains a 3' primer-specific portion (Bi-Ci) that, together with the 5' primer specific portion (Ai) of the upstream primer having a sequence specific for detecting the mutation, permit subsequent amplification and detection of only mutant PCR products. As illustrated in step D of this Figure, another layer of specificity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA

base (r), in the mutation-specific and locus-specific primers. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to generate a polymerase extension competent 3'OH group (Figure 5, step D). In the initial primer extension (step B) the liberated 3'OH base is a few bases upstream from the mutation position, and thus would extend both wild-type and mutant sequences if cleaved (although the blocking LNA or PNA should limit cleavage of primer hybridized to wild-type sequence). In contrast, in the nested PCR (step D), the mutation-specific base of the primer is at the 3'OH base, such that extension on wild-type sequence would be less likely, since the base is mismatched. The specificity for polymerase extension of mutant over wild-type sequence may be further improved by: (i) using mutation-specific PCR Primers containing a mismatch in the 2nd or 3rd position from the 3'OH base, (ii) using LNA or PNA probes to wild-type sequence that would reduce hybridization of mutation-specific PCR primers to wild-type sequences, (iii) using PCR primers to wild-type sequence that are blocked and do not undergo additional amplification, and (iv) avoiding G:T or T:G mismatches between primer and wild-type sequence at the 3'OH base.

[0163] As shown in Figure 5 step E, nested PCR products comprise a 5' primer-specific portion (Ai) target-specific sequence, and a 3' primer-specific portion (Bi'-Ci') that permits subsequent amplification of the nested PCR product. Following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing one or more tag-specific primer pairs, each pair comprising matched primers F1-Bi-Q-Ai and Ci, treated with UDG or similar enzyme to remove dU containing amplification products or contaminants, PCR amplified (Figure 5, step F), and detected. PCR amplification results in the formation of double stranded products as shown in Figure 5, step G. In this example, a polymerase-blocking unit prevents a polymerase from copying the 5' portion (Bi) of the first universal primer, such that the bottom strand of product cannot form a hairpin when it becomes single-stranded. Formation of such a hairpin would result in the 3' end of the stem annealing to the amplicon such that polymerase extension of this 3' end would terminate the PCR reaction.

[0164] The double stranded PCR products are denatured, and when the temperature is subsequently decreased, the upper strand of product forms a hairpin having a stem between the 5' portion (Bi) of the first oligonucleotide primer and portion Bi' at the opposite end of the strand (Figure 5, step H). Also, during this step, the second oligonucleotide primer anneals to the 5'-primer specific portion (Ci') of the hairpinned product. Upon extension of the second universal primer in step H, 5' nuclease activity of the polymerase cleaves the detectable label D1 or the quencher molecule from the 5' end of the amplicon, thereby increasing the distance between the label and the quencher and permitting detection of the label.

[0165] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences of other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules and one or more nucleases capable of digesting nucleic acid molecules present not comprising modified nucleotides are provided. The method also involves providing one or more primary oligonucleotide primer sets. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The sample, the one or more first primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix that comprises one or more modified nucleotides that protect extension product but not target DNA from nuclease digestion, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase extension reaction mixtures and for carrying out one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the target nucleotide sequence. The method further comprises blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more second primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more nucleases, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures. The one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting nucleic acid molecules present in the polymerase chain reaction mixtures, but not primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension

treatment, thereby forming first polymerase chain reaction products comprising the target nucleotide sequence or a complement thereof. One or more secondary oligonucleotide primer sets are then provided. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer. The first polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further comprises detecting and distinguishing the second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

[0166] Figures 6, 7, and 8 illustrate various embodiments of this aspect of the present application.

[0167] Figure 6 illustrates another exemplary exPCR-qPCR carryover prevention reaction to detect low-level mutations. Genomic or cfDNA is isolated (Figure 6, step A), and the isolated DNA sample is treated with UDG to digest dU containing nucleic acid molecules that may be present in the sample (Figure 6, step A). The sample is then subject to a linear amplification reaction, *e.g.*, one or more polymerase extension reactions to generate complementary copies of mutation containing regions of interest. The regions of interest are selectively extended using locus-specific upstream primers, a blocking LNA or PNA probe comprising wild-type sequence, and a deoxynucleotide mix that includes one of more modified nucleotides. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the mutation, and suitable for

polymerase extension (Figure 6, step B). A blocking LNA or PNA probe comprising wild-type sequence that partially overlaps with the upstream PCR primer will preferentially compete in binding to wild-type sequence over the upstream primer, but not as much to mutant DNA, and thus suppresses extension of wild-type DNA during each round of primer extension. The initial extension products incorporate one or more modified nucleotides, such as α-thio-dNTPs, that protect the initial extension products (but not the original cfDNA or genomic DNA) from exonuclease I digestion (Figure 6, step B). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step.

[0168] Subsequently, the locus-specific downstream primers are added, followed by limited cycle PCR (8 to 12 cycles, Figure 6, step C). In the preferred embodiment, the locus-specific downstream primers are approximately 20 to 40 bases downstream from the locus-specific upstream primers. Optionally, the downstream primers contain identical 8-11 base tails to prevent primer dimers.

[0169] Following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes, mutation-specific, and locus-specific primers, to amplify the mutation-containing sequence, if present in the sample (Figure 6, step D). Once again, the presence of blocking LNA or PNA probe comprising wild-type sequence suppresses extension of wild-type target sequence if present after the enrichment of mutant sequence during the initial extension-amplification steps (Figure 6, steps B & C). As illustrated in step D of this Figure 6, another layer of specificity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the mutation-specific and locus-specific primers. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to generate a polymerase extension competent 3'OH group (Figure 6, step D). In the initial primer extension (step B), the liberated 3'OH is a few bases upstream from the mutation position, and thus would extend both wild-type and mutant sequences if cleaved (although the blocking LNA or PNA should limit cleavage of primer hybridized to wild-type sequence). In contrast, in the nested PCR (step D), the mutation-specific base of the primer is at the 3'OH, such that extension on wild-type sequence would be less likely, since the base is mismatched. The specificity for polymerase extension of mutant over wild-type sequence may be further improved by: (i) using mutation-specific PCR Primers containing a mismatch in the 2nd or 3rd position from the 3'OH base, (ii) using LNA or PNA probes to wild-type sequence that would reduce hybridization of mutation-specific PCR primers to wild-type sequences, (iii) using PCR primers to wild-type sequence that are blocked and do not undergo additional amplification, and (iv) avoiding G:T or T:G mismatches between primer and wild-type sequence at the 3'OH base. The TaqMan™ probe spans the mutation region and

contains a fluorescent reporter group on one end (F1) and a quencher molecule (Q) on the other end that are in close enough proximity to each other in the intact probe that the quencher molecule quenches fluorescence of the reporter group. During amplification, the TaqMan™ probe and upstream primer hybridize to their complementary regions of the initial PCR product. The 5'→ 3' nuclease activity of the polymerase extends the hybridized primer and liberates the fluorescent group of the TaqMan™ probe to generate a detectable signal (Figure 6, step E). In a preferred embodiment, the Taqman™ probe contains a second quencher group (ZEN) about 9 bases in from the fluorescent reporter group, and the probe is designed such that the ZEN group is at or adjacent to the mutant base. Use of dUTP during the amplification reaction generates products containing dU, which can subsequently be destroyed using UDG for carryover prevention

[0170] Figures 7 and 8 illustrate additional exemplary exPCR-qPCR carryover prevention reaction to detect low-level mutations. Genomic or cfDNA is isolated (Figure 7 and 8, step A), and the isolated DNA sample is treated with UDG to digest dU containing nucleic acid molecules that may be present in the sample (Figure 7, step A). The regions of interest are selectively extended using locus-specific upstream primers, a blocking LNA or PNA probe comprising wild-type sequence, and a deoxynucleotide mix that includes one of more modified nucleotides. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the mutation, and suitable for polymerase extension (Figure 7 and 8, step B). A blocking LNA or PNA probe comprising wild-type sequence that partially overlaps with the upstream PCR primer will preferentially compete in binding to wild-type sequence over the upstream primer, but not as much to mutant DNA, and thus suppresses extension of wild-type DNA during each round of primer extension. The initial extension products incorporate one or more modified nucleotides, such as α-thio-dNTPs, that protect the initial extension products (but not the original cfDNA or genomic DNA) from exonuclease I digestion (Figure 7 and 8, step B). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the locus-specific downstream primers are added, followed by limited cycle PCR (8 to 12 cycles, Figure 7 and 8, step B).

[0171] For the protocol illustrated in Figure 7, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes, mutation-specific primers comprising 5' primer-specific portions (Ai), locus-specific primers comprising 5' primer-specific portions (Ci) and matching primers Ai and Ci. These primers combine to amplify the mutation-containing sequence, if present in the sample (Figure 7, step C).

Once again, the presence of blocking LNA or PNA probe comprising wild-type sequence suppresses extension of wild-type target sequence if present after the enrichment of mutant sequence during the initial extension-amplification steps (Figure 7, step B). As illustrated in step C of this Figure, another layer of specificity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the mutation-specific and locus-specific primers. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to generate a polymerase extension competent 3'OH group (Figure 7, step C). In the initial primer extension (step B) the liberated 3'OH is a few bases upstream from the mutation position, and thus would extend both wild-type and mutant sequences if cleaved (although the blocking LNA or PNA should limit cleavage of primer hybridized to wild-type sequence). In contrast, in the combined Taqman™ – universal tag PCR amplification (steps C-F), the mutation-specific base of the upstream primer is at the 3'OH, such that extension on wild-type sequence would be less likely, since the base is mismatched. Following the mutation-specific and locus-specific extensions to generate products comprising the Ai tag sequence, target-specific sequence and Ci' tag sequence (Figure 7, step D), the products can be detected by the pairs of matched primers Ai and Ci, and TaqMan™ probes that span the ligation junction as described *supra* for Figure 4 steps F-G (see Figure 7, steps E & F), or using other suitable means known in the art. The specificity for polymerase extension of mutant over wild-type sequence may be further improved by: (i) using mutation-specific PCR Primers containing a mismatch in the 2nd or 3rd position from the 3'OH base, (ii) using LNA or PNA probes to wild-type sequence that would reduce hybridization of mutation-specific PCR primers to wild-type sequences, (iii) using PCR primers to wild-type sequence that are blocked and do not undergo additional amplification, and (iv) avoiding G:T or T:G mismatches between primer and wild-type sequence at the 3'OH base. Further, the longer target-specific primers are at a significantly lower concentration than the Taqman™ probe and tag-specific primers (Ai, Ci), such that the longer mutation-specific primers are depleted, allowing the Taqman™ probe and tag-specific primers to hybridize and enable target-dependent detection.

[0172] For the protocol illustrated in Figure 8, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores, or droplets containing Taqman™ probes, mutation-specific primers comprising 5' primer-specific portions (Ai), locus-specific primers comprising 5' primer-specific portions (Bi-Ci) and matching UniTaq primers F1-Bi-Q-Ai and Ci. These primers combine to amplify the mutation-containing sequence, if present in the sample (Figure 8, step C). Once again, the presence of blocking LNA or PNA probe comprising wild-type sequence suppresses extension of wild-type target sequence if present after the enrichment of mutant sequence during the initial extension-amplification steps (Figure 8, step B). As

illustrated in step C of this Figure, another layer of specificity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the mutation-specific and locus-specific primers. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to generate a polymerase extension competent 3'OH group (Figure 8, step C). In the initial primer extension (step B) the liberated 3'OH is a few bases upstream from the mutation position, and thus would extend both wild-type and mutant sequences if cleaved (although the blocking LNA or PNA should limit cleavage of primer hybridized to wild-type sequence). In contrast, in the combined Taqman™ – UniTaq PCR amplification (steps C-G), the mutation-specific base of the upstream primer is at the 3'OH, such that extension on wild-type sequence would be less likely, since the base is mismatched. Following the mutation-specific and locus-specific extensions to generate products comprising the Ai tag sequence, target-specific sequence and Bi'-Ci' tag sequence (Figure 8, step D), the products can be detected by the pairs of matched UniTaq primers (*i.e.* F1-Bi-Q-Ai and Ci), as described *supra* for Figure 5 steps F-H (see Figure 8, steps E-G), or using other suitable means known in the art. The specificity for polymerase extension of mutant over wild-type sequence may be further improved by: (i) using mutation-specific PCR Primers containing a mismatch in the 2nd or 3rd position from the 3'OH base, (ii) using LNA or PNA probes to wild-type sequence that would reduce hybridization of mutation-specific PCR primers to wild-type sequences, (iii) using PCR primers to wild-type sequence that are blocked and do not undergo additional amplification, and (iv) avoiding G:T or T:G mismatches between primer and wild-type sequence at the 3'OH base. Further, the longer target-specific primers are at a significantly lower concentration than the UniTaq primers (F1-Bi-Q-Ai, Ci), such that the longer mutation-specific primers are depleted, allowing the UniTaq primers to hybridize and enable target-dependent detection.

[0173] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues and subjecting the nucleic acid molecules in the sample to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules are then provided. The method further involves providing one or more primary oligonucleotide primer sets. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide

primer that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The bisulfite-treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite-treated target nucleotide sequence. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures. The one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising the bisulfite-treated target nucleotide sequence or a complement thereof. The method further involves providing one or more oligonucleotide probe sets. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion, and (b) a second oligonucleotide probe having a 5' bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion and a 3' primer-specific portion, and wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on a complementary nucleotide sequence of a first polymerase chain reaction product. The first polymerase chain reaction products are blended with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures. The one or more ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets are ligated together, when hybridized to complementary sequences, to form ligated product sequences in the ligation reaction mixture wherein each ligated product sequence comprises the 5' primer-specific

portion, the bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portions, and the 3' primer-specific portion. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures, and the one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming a second polymerase chain reaction products. The method further involves detecting and distinguishing the second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

[0174] Figures 9 and 10 illustrate exPCR-LDR-qPCR carryover prevention reaction to detect low-level methylation in accordance with this aspect of the present application. The steps are similar to those steps described for Figure 2, with two key differentiators. First, after isolating the genomic or cfDNA, it is optionally treated with a DNA repair kit prior to bisulfite conversion (Figures 9 and 10, Step A). Bisulfite converts unmethylated cytosines, but not 5-methyl cytosines (5meC) nor 5-hydroxymethyl cytosine (5hmC) into a uracil base, which base-pairs with A. Thus, after a single cycle of PCR amplification, unmethylated Cm but not 5meC nor 5hmC is converted to a "T" base, thus allowing for both modified forms of cytosine to be distinguished from unmodified cytosine. Second, the regions of interest are selectively extended using locus-specific downstream primers (with optional identical 8-11 base tails), and a deoxynucleotide mix that does NOT include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 9, step B). Add UDG, which destroys the bisulfite

converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the locus-specific upstream primers are added, followed by limited (8 to 20 cycles) or full (20-40 cycles) PCR using a deoxynucleotide mix that includes dUTP (Figure 9, step C). Upon target-specific hybridization, RNase H removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 9, step C). A blocking LNA or PNA probe comprising the bisulfite converted unmethylated sequence (or its complement) that partially overlaps with the upstream PCR primer will preferentially compete for binding to the bisulfite converted unmethylated sequence over the upstream primer, thus suppressing amplification of bisulfite converted unmethylated sequence DNA during each round of PCR. Optionally, the downstream primers contain identical 8-11 base tails to prevent primer dimers. Further, such tails provide the option for asymmetric PCR at the end of the PCR cycles, by raising the hybridization temperature above that for the forward primers, but at or below that for the reverse primers – which at 8-11 bases longer will have higher Tm values. This generates more bottom strand products, which are suitable substrates for the subsequent LDR step. The amplified products contain dU as shown in Figure 9, step D, which allows for subsequent treatment with UDG or a similar enzyme for carryover prevention.

[0175] Alternatively, as shown in Figure 10, the regions of interest are selectively extended using locus-specific upstream primers, a blocking LNA or PNA probe comprising bisulfite converted unmethylated sequence (or its complement), and a deoxynucleotide mix that does NOT include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 10, step B). A blocking LNA or PNA probe comprising the bisulfite converted unmethylated sequence (or its complement) that partially overlaps with the upstream PCR primer will preferentially compete for binding to the bisulfite converted unmethylated sequence over the upstream primer, thus suppressing amplification of bisulfite converted unmethylated sequence DNA during each round of PCR. Add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the locus-specific downstream primers are added, followed by limited (8 to 20 cycles) or full (20-40 cycles) PCR using a deoxynucleotide mix that includes dUTP (Figure 14, step C). Optionally, the downstream primers contain identical 8-11 base tails to prevent primer dimers.

[0176] For Figures 9 and 10, methylation-specific upstream and locus-specific downstream probes containing tails (Ai, Ci') enable formation of a ligation product in the presence of bisulfite converted methylated base-containing PCR products. Following ligation, the ligation products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the ligation junction as described *supra* for Figure 2 (see Figure 9, steps E-H), or using other suitable means known in the art.

[0177] Alternatively, methylation-specific upstream and locus-specific downstream probes containing tails (Ai, Bi'-Ci') enable formation of a ligation product in the presence of bisulfite converted methylated base-containing PCR products. Following ligation, the ligation products are amplified using UniTaq-specific primers (*i.e.*, F1-Bi-Q-Ai, Ci) and detected as described *supra* for Figure 3, or using other suitable means known in the art.

[0178] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues. The nucleic acid molecules in the sample are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules are provided, and one or more first primary oligonucleotide primer(s) are provided. Each first primary oligonucleotide primer comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue. The bisulfite-treated sample, the one or more first primary oligonucleotide primers, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures, and the one or more polymerase extension reaction mixtures are subjected to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, to form primary extension products comprising the complement of the bisulfite-treated target nucleotide sequence. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of the polymerase extension reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that

comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting nucleic acid molecules present in the first polymerase chain reaction mixtures, but not primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reactions products comprising a 5' primer-specific portion of the first secondary oligonucleotide primer, the bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion, and a complement of the 5' primer-specific portion of the second secondary oligonucleotide primer. The method further involves providing one or more tertiary oligonucleotide primer sets. Each tertiary oligonucleotide primer set comprises (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reactions product sequence. The first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain reaction products. The method further involves detecting and distinguishing the secondary polymerase chain reactions products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

[0179] Figures 11, 12, 18, and 19 illustrate various embodiments of this aspect of the present application.

[0180] Figure 11 illustrates an exemplary exPCR-qPCR carryover prevention reaction to detect low-level methylations. Genomic or cfDNA is isolated and is optionally treated with a DNA repair kit prior to bisulfite conversion (Figure 11, Step A). The regions of interest are selectively extended using locus-specific downstream primers (with optional identical 8-11 base tails), and a deoxynucleotide mix that does NOT include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 11, step B). If the locus-specific downstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. After the extension cycles, add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step.

[0181] Alternatively, as shown in Figure 12, regions of interest are selectively extended using locus-specific upstream primers, a blocking LNA or PNA probe comprising bisulfite converted unmethylated sequence (or its complement), and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 12, step B). A blocking LNA or PNA probe comprising the bisulfite converted unmethylated sequence (or its complement) that partially overlaps with the upstream PCR primer will preferentially compete for binding to the bisulfite converted unmethylated sequence over the upstream primer, thus suppressing amplification of bisulfite converted unmethylated sequence DNA during each round of PCR. Add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step.

[0182] As shown in Figures 11 and 12 step C, bisulfite converted methylation base-specific primers (comprising 5' primer-specific portions Ai) and bisulfite converted locus-specific primers (comprising 5' primer-specific portions Ci) are added to then perform limited cycle nested PCR to amplify the bisulfite converted methylation-containing sequence, if present in the sample. Blocking LNA or PNA probes comprising the bisulfite converted unmethylated sequence (or its complement) enables amplification of originally methylated but not originally

un-methylated allele. Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the bisulfite-converted methylation target regions as described *supra* for Figure 4 (see Figures 11 and 12, steps D-F), or using other suitable means known in the art.

[0183] Alternatively, bisulfite converted methylation base-specific primers (comprising 5' primer-specific portions Ai) and bisulfite converted locus-specific primers (comprising 5' primer-specific portions Bi-Ci) are added to then perform limited cycle nested PCR to amplify the bisulfite converted methylation-containing sequence, if present in the sample. Blocking LNA or PNA probes comprising the bisulfite converted unmethylated sequence (or its complement) enables amplification of originally methylated but not originally un-methylated alleles. Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products are amplified using UniTaq-specific primers (*i.e.*, F1-Bi-Q-Ai, Ci) and detected as described *supra* for Figure 5, or using other suitable means known in the art.

[0184] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues. The nucleic acid molecules in the sample are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample are provided, and one or more primary oligonucleotide primer sets are provided. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The bisulfite-treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures. The one or more polymerase extension reaction mixtures to are subjected to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the

bisulfite treated target nucleotide sequence. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixture, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures. The one or more first polymerase chain reaction mixtures are sujected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising the bisulfite- treated target nucleotide sequence or a complement thereof. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of a first polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a first polymerase chain reaction product formed from the first secondary oligonucleotide primer. The first polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further involves detecting and distinguishing the second polymerase chain reactions products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

[0185] Figures 13-15, 20, and 21 illustrate various embodiments of this aspect of the present application.

[0186] Figure 13 illustrates another exemplary exPCR-qPCR carryover prevention reaction to detect low-level methylation. Genomic or cfDNA is isolated, and optionally treated

with a DNA repair kit prior to bisulfite conversion (Figure 13, Step A). The regions of interest are selectively extended using locus-specific downstream primers (with optional identical 8-11 base tails), and a deoxynucleotide mix that does NOT include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 13, step B). If the locus-specific downstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. After the extension cycles, add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the regions of interest are selectively amplified in a limited cycle PCR (8-20 cycles) using locus-specific upstream primers, a blocking LNA or PNA probe comprising bisulfite converted unmethylated sequence (or its complement), and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 13, step C). A blocking LNA or PNA probe comprising the bisulfite converted unmethylated sequence (or its complement) that partially overlaps with the upstream PCR primer will preferentially compete for binding to the bisulfite converted unmethylated sequence over the upstream primer, thus suppressing amplification of bisulfite converted unmethylated sequence DNA during each round of PCR.

[0187] Following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes, bisulfite-converted, methylation base-specific, and bisulfite converted locus-specific primers, to amplify the bisulfite converted methylation-containing sequence, if present in the sample (Figure 13, step D). The bisulfite converted methylation-containing products are amplified and detected as described *supra* for Figure 6 (see Figure 13, steps D-E), or using other suitable means known in the art.

[0188] Figures 14 and 15 illustrate additional exemplary exPCR-qPCR carryover prevention reaction to detect low-level methylation. Genomic or cfDNA is isolated, and optionally treated with a DNA repair kit prior to bisulfite conversion (Figures 14 and 15, Step A). The regions of interest are selectively extended using locus-specific downstream primers (with optional identical 8-11 base tails), and a deoxynucleotide mix that does NOT include

dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 14, step B). If the locus-specific downstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. After the extension cycles, add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the regions of interest are selectively amplified in a limited cycle PCR (8-20 cycles) using locus-specific upstream primers, a blocking LNA or PNA probe comprising bisulfite converted unmethylated sequence (or its complement), and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 14, step C). A blocking LNA or PNA probe comprising the bisulfite converted unmethylated sequence (or its complement) that partially overlaps with the upstream PCR primer will preferentially compete for binding to the bisulfite converted unmethylated sequence over the upstream primer, thus suppressing amplification of bisulfite converted unmethylated sequence DNA during each round of PCR.

[0189] Alternatively, as shown in Figure 15, the regions of interest are selectively extended using locus-specific upstream primers, a blocking LNA or PNA probe comprising bisulfite converted unmethylated sequence (or its complement), and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 15, step B). A blocking LNA or PNA probe comprising the bisulfite converted unmethylated sequence (or its complement) that partially overlaps with the upstream PCR primer will preferentially compete for binding to the bisulfite converted unmethylated sequence over the upstream primer, thus suppressing amplification of bisulfite converted unmethylated sequence DNA during each round of PCR. Add UDG, which destroys the bisulfite converted DNA (but not the primer extension

products). Subsequently, the locus-specific downstream primers are added, followed by limited cycle PCR (8 to 12 cycles, Figure 15, step C). If the locus-specific downstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step.

[0190] For the protocol illustrated in Figures 14 and 15, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes, bisulfite converted methylation base-specific primers comprising 5' primer-specific portions (Ai), bisulfite converted locus-specific primers comprising 5' primer-specific portions (Ci) and matching primers Ai and Ci. These primers combine to amplify the bisulfite converted methylation-containing sequence, if present in the sample (Figures 14 and 15, step D). Blocking LNA or PNA probes comprising the bisulfite converted unmethylated sequence (or its complement) enables amplification of originally methylated but not originally un-methylated allele. Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the bisulfite-converted methylation target regions as described *supra* for Figure 4 (see Figure 14, steps E-G), or using other suitable means known in the art.

[0191] Alternatively, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes, bisulfite converted methylation base-specific primers comprising 5' primer-specific portions (Ai), bisulfite converted locus-specific primers comprising 5' primer-specific portions (Bi-Ci) and matching UniTaq primers F1-Bi-Q-Ai and Ci. Blocking LNA or PNA probes comprising the bisulfite converted unmethylated sequence (or its complement) enables amplification of originally methylated but not originally un-methylated alleles. Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products are amplified using UniTaq-specific primers (*i.e.*, F1-Bi-Q-Ai, Ci) and detected as described *supra* for Figure 5, or using other suitable means known in the art.

[0192] Figures 16 and 17 illustrate additional exemplary exPCR-LDR-qPCR carryover prevention reactions to detect low-level methylation. Genomic or cfDNA is isolated and then either treated with: (i) methyl-sensitive restriction endonucleases, *e.g.*, Bsh1236I (CG^CG), to completely digest unmethylated DNA and prevent carryover, or (ii) capture and enrich for methylated DNA, (iii) followed by a DNA repair kit (Figures 16 and 17, step A). The DNA is bisulfite-treated to convert unmethylated residues to uracil thereby rendering the double stranded DNA non-complementary. The regions of interest are selectively extended using locus-specific downstream primers (with optional identical 8-11 base tails), and a deoxynucleotide mix that

does NOT include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 16, step B). If the locus-specific downstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. After the extension cycles, add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the regions of interest are selectively amplified in a limited cycle PCR (8-20 cycles) using locus-specific upstream primers, and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 16, step C). If the locus-specific upstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence.

[0193] Alternatively, as shown in Figure 17, the regions of interest are selectively extended using locus-specific upstream primers for bisulfite converted DNA, and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 17, step B). If the locus-specific upstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. Add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Subsequently, the locus-specific downstream primers are added, followed by limited (8 to 20 cycles) or full (20-40 cycles) PCR using a deoxynucleotide mix that includes dUTP (Figure 17, step C). Optionally, the downstream primers contain identical 8-11 base tails to prevent primer dimers. Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. The amplified

products contain dU as shown in Figure 17, step D, which allows for subsequent treatment with UDG or a similar enzyme for carryover prevention.

[0194] For Figure 16, methylation-specific upstream and locus-specific downstream probes containing tails (Ai, Ci') enable formation of a ligation product in the presence of bisulfite converted methylated base-containing PCR products. Following ligation, the ligation products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the ligation junction as described *supra* for Figure 2 (see Figure 16, steps E-H), or using other suitable means known in the art.

[0195] Alternatively, methylation-specific upstream and locus-specific downstream probes containing tails (Ai, Bi'-Ci') enable formation of a ligation product in the presence of bisulfite converted methylated base-containing PCR products. Following ligation, the ligation products are amplified using UniTaq-specific primers (*i.e.*, F1-Bi-Q-Ai, Ci) and detected as described *supra* for Figure 3, or using other suitable means known in the art.

[0196] Figures 18 and 19 illustrate additional exemplary exPCR-LDR-qPCR carryover prevention reactions to detect low-level methylation. Genomic or cfDNA is isolated, and then either treated with: (i) methyl-sensitive restriction endonucleases, *e.g.*, Bsh1236I (CG^CG), to completely digest unmethylated DNA and prevent carryover, or (ii) capture and enrich for methylated DNA, (iii) followed by a DNA repair kit (Figures 18 and 19, step A). The DNA is bisulfite-treated to convert unmethylated residues to uracil thereby rendering the double stranded DNA non-complementary. The regions of interest are selectively extended using locus-specific downstream primers (with optional identical 8-11 base tails), and a deoxynucleotide mix that does NOT include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 18, step B). If the locus-specific downstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. After the extension cycles, add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step.

[0197] Alternatively, as shown in Figure 19, the regions of interest are selectively extended using locus-specific upstream primers for bisulfite converted DNA, and a deoxynucleotide mix that does not include dUTP. Blocking LNA or PNA probes comprising the bisulfite converted unmethylated sequence (or its complement) enables amplification of originally methylated but not originally un-methylated alleles. In this embodiment, another layer

of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 19, step B). Add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step.

[0198] As shown in Figure 18, step C, bisulfite converted methylation base-specific primers (comprising 5' primer-specific portions Ai) and bisulfite converted locus-specific primers (comprising 5' primer-specific portions Ci) are added to then perform limited cycle nested PCR to amplify the bisulfite converted methylation-containing sequence, if present in the sample. Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the bisulfite-converted methylation target regions as described *supra* for Figure 4 (see Figure 18, steps D-F), or using other suitable means known in the art.

[0199] Alternatively, bisulfite converted methylation base-specific primers (comprising 5' primer-specific portions Ai) and bisulfite converted locus-specific primers (comprising 5' primer-specific portions Bi-Ci) are added to then perform limited cycle nested PCR to amplify the bisulfite converted methylation-containing sequence, if present in the sample. Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products are amplified using UniTaq-specific primers (*i.e.*, F1-Bi-Q-Ai, Ci) and detected as described *supra* for Figure 5, or using other suitable means known in the art.

[0200] Figure 20 illustrates another exemplary exPCR-qPCR carryover prevention reaction to detect low-level methylation. Genomic or cfDNA is isolated and then either treated with: (i) methyl-sensitive restriction endonucleases, *e.g.*, Bsh1236I (CG^CG), to completely digest unmethylated DNA and prevent carryover, or (ii) capture and enrich for methylated DNA, (iii) followed by a DNA repair kit (Figure 20, step A). The DNA is bisulfite-treated to convert unmethylated residues to uracil thereby rendering the double stranded DNA non-complementary. The regions of interest are selectively extended using locus-specific downstream primers (with optional identical 8-11 base tails), and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 20, step B). If the locus-specific downstream primer covers one or more methylation sites, another layer of

specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. After the extension cycles, add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the regions of interest are selectively amplified in a limited cycle PCR (8-20 cycles) using locus-specific upstream primers, and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figures 20, step C). If the locus-specific upstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence.

[0201] Following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes, bisulfite-converted, methylation base-specific, and bisulfite converted locus-specific primers, to amplify the bisulfite converted methylation-containing sequence, if present in the sample (Figure 20, step D). The bisulfite converted methylation-containing products are amplified and detected as described *supra* for Figure 6 (see Figure 20, steps D-E), or using other suitable means known in the art.

[0202] Figure 21 illustrate additional exemplary exPCR-qPCR carryover prevention reaction to detect low-level methylation. Genomic or cfDNA is isolated, and then either treated with: (i) methyl-sensitive restriction endonucleases, *e.g.*, Bsh1236I (CG^CG), to completely digest unmethylated DNA and prevent carryover, or (ii) capture and enrich for methylated DNA, (iii) followed by a DNA repair kit (Figure 21, step A). The DNA is bisulfite-treated to convert unmethylated residues to uracil thereby rendering the double stranded DNA non-complementary. The regions of interest are selectively extended using locus-specific downstream primers (with optional identical 8-11 base tails), and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 21, step B). If the locus-specific downstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. After the extension cycles, UDG, which destroys the bisulfite converted DNA (but not the primer extension products) is added. Optionally, samples are

aliquoted into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the regions of interest are selectively amplified in a limited cycle PCR (8-20 cycles) using locus-specific upstream primers, and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 21, step C). If the locus-specific upstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence.

[0203] For the protocol illustrated in Figure 21, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes, bisulfite converted methylation base-specific primers comprising 5' primer-specific portions (Ai), bisulfite converted locus-specific primers comprising 5' primer-specific portions (Ci) and matching primers Ai and Ci. These primers combine to amplify the bisulfite converted methylation-containing sequence, if present in the sample (Figure 21, step D). Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the bisulfite-converted methylation target regions as described *supra* for Figure 4 (see Figure 21, steps E-G), or using other suitable means known in the art.

[0204] Alternatively, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes, bisulfite converted methylation base-specific primers comprising 5' primer-specific portions (Ai), bisulfite converted locus-specific primers comprising 5' primer-specific portions (Bi-Ci) and matching UniTaq primers F1-Bi-Q-Ai and Ci. Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products are amplified using UniTaq-specific primers (*i.e.*, F1-Bi-Q-Ai, Ci) and detected as described *supra* for Figure 5, or using other suitable means known in the art.

[0205] Another aspect of the present application is directed to a method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues. The method involves providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues, and subjecting the nucleic acid molecules in the sample to a bisulfite treatment under

conditions suitable to convert unmethylated cytosine residues to uracil residues. One or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample are provided. One or more primary oligonucleotide primer sets are also provided. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer. The bisulfite treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more polymerase extension reaction mixtures. The one or more polymerase extension reaction mixtures are subjected to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite treated target nucleotide sequence. The one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixture, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures. The method further comprises subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reactions products comprising the bisulfite-treated target nucleotide sequence or a complement thereof. One or more secondary oligonucleotide primer sets are provided. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products or their complements and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction products or their complements. The primary polymerase chain reaction product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil

(dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further involves detecting and distinguishing the second polymerase chain reactions products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

[0206] Figure 22 illustrates an embodiment of this aspect of the present application.

[0207] Figure 22 illustrates an additional exemplary exPCR-qPCR carryover prevention reaction to detect low-level methylation. Genomic or cfDNA is isolated, and then either treated with: (i) methyl-sensitive restriction endonucleases, *e.g.*, Bsh1236I (CG^CG), to completely digest unmethylated DNA and prevent carryover, or (ii) capture and enrich for methylated DNA, (iii) followed by a DNA repair kit (Figure 22, step A). The DNA is bisulfite-treated to convert unmethylated residues to uracil thereby rendering the double stranded DNA non-complementary. The regions of interest are selectively extended using bisulfite converted locus-specific downstream primers comprising 5' primer-specific portions (Ci for Figure 22), and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the downstream primer. Upon target-specific hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is suitable for polymerase extension (Figure 22, step B). In this embodiment, the locus-specific downstream primer covers one or more methylation sites, and another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence. After the extension cycles, add UDG, which destroys the bisulfite converted DNA (but not the primer extension products). Optionally aliquot sample into 12, 24, 36, 48, or 96 wells prior to the initial extension step. Subsequently, the regions of interest are selectively amplified in a limited cycle PCR (8-20 cycles) using bisulfite converted methylation base-specific upstream primers comprising 5' primer-specific portions (Ai), and a deoxynucleotide mix that does not include dUTP. In this embodiment, another layer of selectivity can be incorporated into the method by including a 3' cleavable blocking group (Blk 3', *e.g.* C3 spacer), and an RNA base (r), in the upstream primer. Upon target-specific

hybridization, RNase H (star symbol) removes the RNA base to liberate a 3'OH group which is a few bases upstream of the bisulfite converted methylated target base, and suitable for polymerase extension (Figure 22, step C). Since the methylation base-specific upstream primer covers one or more methylation sites, another layer of specificity may be added by using blocking primers whose sequence corresponds to the bisulfite-converted unmethylated sequence.

[0208] As shown in Figure 22 step D, the limited cycle PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence, and are distributed into wells, micro-pores, or droplets for Taqman™ reactions. Following PCR, the products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the bisulfite-converted methylation target regions as described *supra* for Figure 4 (see Figure 22, steps D-F), or using other suitable means known in the art.

[0209] Alternatively, the limited cycle PCR products comprise of Ai tag sequence, methylation-specific sequence, and Bi'-Ci' tag sequence, and are distributed into wells, micro-pores, or droplets for Taqman™ reactions. Following PCR, the products are amplified using UniTaq-specific primers (*i.e.*, F1-Bi-Q-Ai, Ci) and detected as described *supra* for Figure 5, or using other suitable means known in the art.

[0210] In one embodiment, the method further comprises contacting the sample with DNA repair enzymes to repair damaged DNA, abasic sites, oxidized bases, or nicks in the DNA.

[0211] In another embodiment, the method further comprises contacting the sample with at least a first methylation sensitive enzyme to form a restriction enzyme reaction mixture prior to, or concurrent with, said blending to form one or more polymerase extension reaction mixtures, wherein said first methylation sensitive enzyme cleaves nucleic acid molecules in the sample that contain one or more unmethylated residues within at least one methylation sensitive enzyme recognition sequence, and whereby said detecting involves detection of one or more parent nucleic acid molecules containing the target nucleotide sequence, wherein said parent nucleic acid molecules originally contained one or more methylated residues.

[0212] In accordance with this and all aspects of the present invention, a "methylation sensitive enzyme" is an endonuclease that will not cleave or has reduced cleavage efficiency of its cognate recognition sequence in a nucleic acid molecule when the recognition sequence contains a methylated residue (*i.e.*, it is sensitive to the presence of a methylated residue within its recognition sequence). A "methylation sensitive enzyme recognition sequence" is the cognate recognition sequence for a methylation sensitive enzyme. In some embodiments, the methylated residue is a 5-methyl-C, within the sequence CpG (*i.e.*, 5-methyl-CpG). A non-limiting list of methylation sensitive restriction endonuclease enzymes that are suitable for use in the methods

of the present invention include, without limitation, AciI, HinP1I, Hpy99I, HpyCH4IV, BstUI, HpaII, HhaI, or any combination thereof.

[0213] In a further embodiment, the method further comprises contacting the sample with an immobilized methylated nucleic acid binding protein or antibody to selectively bind and enrich for methylated nucleic acid in the sample.

[0214] In one embodiment, the primers from the one or more primary or secondary oligonucleotide primer sets comprise a portion that has no or one nucleotide sequence mismatch when hybridized in a base-specific manner to the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof, but have one or more additional nucleotide sequence mismatches that interferes with polymerase extension when primers from said one or more primary or secondary oligonucleotide primer sets hybridize in a base-specific manner to a corresponding nucleotide sequence portion in wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof.

[0215] One or both primary oligonucleotide primers of the primary oligonucleotide primer set and/or one or both secondary oligonucleotide primers of the secondary oligonucleotide primer set may have a 3' portion comprising a cleavable nucleotide or nucleotide analogue and a blocking group, such that the 3' end of said primer or primers is unsuitable for polymerase extension. This embodiment of the method further comprises cleaving the cleavable nucleotide or nucleotide analog of one or both oligonucleotide primers during said hybridization treatment, thereby liberating free 3'OH ends on one or both oligonucleotide primers prior to said extension treatment. In one embodiment, the cleavable nucleotide comprises one or more RNA bases.

[0216] In another embodiment, primers from the one or more primary or secondary oligonucleotide primer sets comprise a sequence that differs from the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof, said difference is located two or three nucleotide bases from the liberated free 3'OH end.

[0217] In another embodiment, the method further comprises providing one or more blocking oligonucleotide primers comprising one or more mismatched bases at the 3' end or one or more nucleotide analogs and a blocking group at the 3' end, such that the 3' end of said blocking oligonucleotide primer is unsuitable for polymerase extension when hybridized in a base-specific manner to wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof, wherein said blocking oligonucleotide primer comprises a portion having a nucleotide sequence that is the same as a nucleotide sequence portion in the wildtype nucleic acid sequence or bisulfite-converted unmethylated

nucleic acid sequence or complement sequence thereof to which the blocking oligonucleotide primer hybridizes but has one or more nucleotide sequence mismatches to a corresponding nucleotide sequence portion in the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof. The one or more blocking oligonucleotide primers are blended with the sample or subsequent products prior to a polymerase extension reaction, polymerase chain reaction, or ligation reaction, whereby during hybridization the one or more blocking oligonucleotide primers preferentially hybridize in a base-specific manner to a wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof, thereby interfering with polymerase extension or ligation during reaction of a primer or probes hybridized in a base-specific manner to the wildtype sequence or bisulfite-converted unmethylated sequence or complement sequence thereof.

[0218] In certain embodiments, the first secondary oligonucleotide primer has a 5' primer-specific portion and the second secondary oligonucleotide primer has a 5' primer-specific portion, said one or more secondary oligonucleotide primer sets further comprising a third secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first secondary oligonucleotide primer and (d) a fourth secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the second secondary oligonucleotide primer.

[0219] Another aspect of the present application is directed to a method for identifying in a sample, one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level. The method involves providing a sample containing one or more parent ribonucleic acid molecules containing a target ribonucleic acid molecule potentially differing in sequence from other parent ribonucleic acid molecules and providing one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU containing nucleic acid molecules potentially present in the sample. One or more primary oligonucleotide primer sets are then provided. Each primary oligonucleotide primer set comprises (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to the RNA sequence in the parent ribonucleic acid molecule adjacent to the target ribonucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a

portion of the cDNA extension product formed from the first primary oligonucleotide primer. The contacted sample, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix including dUTP, a reverse transcriptase, and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target ribonucleic nucleic acid and to carry out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse transcription/polymerase products. The method further comprises providing one or more oligonucleotide probe sets. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on complementary portions of a reverse transcriptase/polymerase product corresponding to the target ribonucleic acid molecule sequence. The reverse transcriptase/polymerase products are contacted with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures, and the one or more ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form ligated product sequences in the ligase reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion. The method further involves providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences, the one or more secondary oligonucleotide primer sets with one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension

treatment thereby forming first polymerase chain reaction products. The method further comprises detecting and distinguishing the first polymerase chain reaction products, thereby identifying the presence of one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level.

[0220] Figures 23 and 26 illustrate embodiments of this aspect of the present application.

[0221] Figure 23 illustrates an exemplary RT-PCR-LDR-qPCR carryover prevention reaction to detect translocations at the mRNA level. Such fusion mRNA may be isolated from circulating tumor cells, exosomes or from other plasma fractions. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. In this embodiment, mRNA is isolated (Figure 23, step A), and treated with UDG for carryover prevention (Figure 23, step B). cDNA is generated using 3' transcript-specific primers and reverse-transcriptase in the presence of dUTP. Suitable reverse transcriptases include but are not limited to Moloney Murine Leukemia Virus Reverse Transcriptase (M-MLV RT, New England Biolabs), or Superscript II or III Reverse Transcriptase (Life Technologies). Taq polymerase is activated to perform limited cycle PCR amplification (12-20) to maintain relative ratios of different amplicons (Figure 23, step B). The primers contain identical 8-11 base tails to prevent primer dimers. PCR products incorporate dUTP, allowing for carryover prevention (Figure 23, step C).

[0222] As shown in Figure 23, step D, exon junction-specific ligation oligonucleotide probes containing primer-specific portions (Ai, Ci') suitable for subsequent PCR amplification, hybridize to their corresponding target sequence in a base-specific manner. Ligase covalently seals the two oligonucleotides together (Figure 23, step D), and ligation products are aliquoted into separate wells for detection using tag-primers (Ai, Ci) and TaqMan™ probe (F1-Q) which spans the ligation junction (Figure 23, step E). Treat samples with UDG for carryover prevention, which also destroys original target amplicons (Figure 23, step E). Only authentic LDR products will amplify, when using PCR in the presence of dUTP. Copy number of fusion transcripts is determined based on signal from wells where original distribution was one copy/well. Neither original PCR primers nor LDR probes amplify LDR products, providing additional carryover protection.

[0223] Another aspect of the present application is directed to a method for identifying in a sample, one or more parent ribonucleic acid molecules containing a target ribonucleic acid

sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level. The method involves providing a sample containing one or more parent ribonucleic acid molecules containing a target ribonucleic acid molecule potentially differing in sequence from other parent ribonucleic acid molecules and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU containing nucleic acid molecules potentially present in the sample. The method further involves providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to the RNA sequence in the parent ribonucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of the cDNA extension product formed from the first primary oligonucleotide primer. The contacted sample, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures, and the one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target RNA and to carry out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse-transcription/primary polymerase chain reaction products. The method futher comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of a reverse-transcription/primary polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a reverse-transcription/primary polymerase chain reaction product formed from the first secondary oligonucleotide primer. The reverse-transcription/primary polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction

mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming first polymerase chain reaction products. The method further involves detecting and distinguishing the first polymerase chain reaction products, thereby identifying the presence of one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequences differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level.

[0224] Figures 24, 25, 27, and 28 illustrate various embodiments of this aspect of the present application.

[0225] Figures 24 and 25 illustrate additional exemplary RT-PCR-LDR-qPCR carryover prevention reaction to detect translocations at the mRNA level. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. In this embodiment, mRNA is isolated (Figures 24 and 25, step A), and treated with UDG for carryover prevention (Figures 24 and 25, step B). cDNA is generated using 3' transcript-specific primers and reverse-transcriptase. Taq polymerase is activated to perform limited cycle PCR amplification (8-20) to maintain relative ratios of different amplicons (Figures 24 and 25, step B). The primers contain identical 8-11 base tails to prevent primer dimers and are added only to wells with the anticipated low-copy dilution.

[0226] For the protocol illustrated in Figure 24, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes across the cDNA fusion junction, cDNA-specific primers, to amplify the junction sequence, if present in the sample (Figure 24, step C). The fusion cDNA products are amplified and detected as described *supra* for Figure 6 (see Figure 24, steps C-D), or using other suitable means known in the art.

[0227] For the protocol illustrated in Figure 25, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes across the cDNA fusion junction, cDNA-specific (forward) primers comprising 5' primer-specific portions (Ai), cDNA-specific (reverse) primers comprising 5' primer-specific portions (Ci) and matching primers Ai and Ci. These primers combine to amplify the fusion cDNA sequence, if present in the sample (Figure 25, step C). Primers are unblocked with

RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the fusion cDNA regions as described *supra* for Figure 4 (see Figure 25, steps D-F), or using other suitable means known in the art.

[0228] Figure 26 illustrates an exemplary RT-PCR-LDR-qPCR carryover prevention reaction to enumerate mRNA, ncRNA, or lncRNA copy number. RNA is isolated from whole blood cells, exosomes, CTCs or other plasma fractions. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. In this embodiment, mRNA is isolated (Figure 26, step A), and treated with UDG for carryover prevention (Figure 26, step B). cDNA is generated using 3' transcript-specific primers and reverse-transcriptase in the presence of dUTP. Taq polymerase is activated to perform limited cycle PCR amplification (12-20) to maintain relative ratios of different amplicons (Figure 26, step B). The primers contain identical 8-11 base tails to prevent primer dimers. PCR products incorporate dUTP, allowing for carryover prevention (Figure 26, step C).

[0229] As shown in Figure 26, step D, exon junction-specific ligation oligonucleotide probes containing primer-specific portions (Ai, Ci') suitable for subsequent PCR amplification, hybridize to their corresponding target sequence in a base-specific manner. Ligase covalently seals the two oligonucleotides together (Figure 26, step D), and ligation products are aliquoted into separate wells for detection using tag-primers (Ai, Ci) and TaqMan™ probe (F1-Q) which spans the ligation junction (Figure 26, step E). Treat samples with UDG for carryover prevention, which also destroys original target amplicons (Figure 26, step E). Only authentic LDR products will amplify, when using PCR in presence of dUTP. Determine copy number of mRNA, ncRNA, or lncRNA transcripts based on signal from wells where original distribution was one copy/well. Neither original PCR primers nor LDR probes amplify LDR products, providing additional carryover protection.

[0230] Figures 27 illustrates an exemplary RT-PCR-LDR-qPCR carryover prevention reaction to enumerate mRNA, ncRNA, or lncRNA copy number. Isolate RNA from whole blood cells, exosomes, CTCs or other plasma fractions. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. In this embodiment, mRNA is isolated (Figures 27, step A), and treated with UDG for carryover prevention (Figures 27, step B). cDNA is generated using 3' transcript-specific primers and reverse-transcriptase in the presence of dUTP. Taq polymerase is activated to perform limited cycle PCR amplification (8-20) to maintain relative ratios of different amplicons (Figures 27, step B). The primers

contain identical 8-11 base tails to prevent primer dimers. PCR products incorporate dUTP, allowing for carryover prevention (Figures 27, step C).

[0231] For the protocol illustrated in Figure 27, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes across the cDNA target region, cDNA-specific primers, to amplify the target sequence, if present in the sample (Figure 27, step C). The cDNA target products are amplified and detected as described *supra* for Figure 6 (see Figure 27, steps C-D), or using other suitable means known in the art.

[0232] For the protocol illustrated in Figure 28, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes across the cDNA target region, cDNA-specific (forward) primers comprising 5' primer-specific portions (Ai), cDNA-specific (reverse) primers comprising 5' primer-specific portions (Ci) and matching primers Ai and Ci. These primers combine to amplify the target cDNA sequence, if present in the sample (Figure 28, step C). Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the target cDNA regions as described *supra* for Figure 4 (see Figure 28, steps D-F), or using other suitable means known in the art.

[0233] Alternatively, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes across the cDNA target region, cDNA-specific (forward) primers comprising 5' primer-specific portions (Ai), cDNA-specific (reverse) primers comprising 5' primer-specific portions (Bi-Ci) and matching primers F1-Bi-Q-Ai and Ci (Figure not shown). These primers combine to amplify the target cDNA sequence, if present in the sample. Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of matched primers F1-Bi-Q-Ai and Ci, and TaqMan™ probes that span the target cDNA regions as described *supra* for Figure 4, or using other suitable means known in the art.

[0234] Another aspect of the present application is directed to a method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases. The method involves providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample, and the contacted sample

is blended with a ligase and one or more first oligonucleotide preliminary probes comprising a 5' phosphate, a 5' stem-loop portion, an internal primer-specific portion within the loop region, a blocking group, and a 3' nucleotide sequence that is complementary to a 3' portion of the target miRNA molecule sequence to form one or more first ligation reaction mixtures. The method further comprises ligating, in the one or more first ligation reaction mixtures, the one or more target miRNA molecules at their 3'end to the 5' phosphate of the one or more first oligonucleotide preliminary probes to generate chimeric nucleic acid molecules comprising the target miRNA molecule sequence, if present in the sample, appended to the one or more first oligonucleotide preliminary probes. One or more primary oligonucleotide primer sets are then provided. Each primer set comprises (a) a first primary oligonucleotide primer comprising a nucleotide sequence that is complementary to the internal primer-specific portion of the first oligonucleotide preliminary probe, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets. The one or more first ligation reaction mixtures comprising chimeric nucleic acid molecules, the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the sample, a deoxynucleotide mix including dUTP, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the reverse-transcription/polymerase chain reaction mixtures to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the chimeric nucleic acid molecules, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different primary reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion, a nucleotide sequence corresponding to the target miRNA molecule sequence, and the complement of the internal primer-specific portion, and complements thereof. The method further involves providing one or more oligonucleotide probe sets. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion, a portion complementary to a primary extension product, and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on complementary portions of a primary reverse-transcription/polymerase chain reaction product corresponding to the target

miRNA molecule sequence, or complement thereof. The primary reverse-transcription/polymerase chain reaction products are contacted with a ligase and the one or more oligonucleotide probe sets to form one or more second ligation reaction mixtures, and the one or more second ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form ligated product sequences in the ligation reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion. The method further involves providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences and the one or more secondary oligonucleotide primer sets are blended with one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain reaction products. The method further comprises detecting and distinguishing the secondary polymerase chain reaction products in the one or more reactions thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

[0235] Figure 29 illustrates an embodiment of this aspect of the present application.

[0236] Figure 29 illustrates an exemplary Ligation-RT-PCR-LDR-qPCR carryover prevention reaction to quantify miRNA. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. This method involves isolating miRNA from exosomes and treating with UDG for carryover prevention (Figure 29, step B). An oligonucleotide probe having a portion that is complementary to the 3' end of the target miRNA, and containing a stem-loop, tag (Di'), and blocking group (filled circle) is ligated at its 5' end to the 3' end of the target miRNA. The ligation product comprises the miRNA, Di' tag, the blocking group, and a sequence complementary to the 3' portion of the miRNA (Figure 29, step

B). A reverse transcriptase such as Moloney Murine Leukemia Virus Reverse Transcriptase (M-MLV RT, New England Biolabs), or Superscript II or III Reverse Transcriptase (Life Technologies) extends primer (Di) to make a full-length copy of the target, and appends three C bases to the 3' end of extended target sequence (Figure 29, step B). Tag oligonucleotide (Ei) having the 3' rGrG+G (+G is the symbol for LNA) hybridizes to the three C bases of the extended target sequence as shown in Figure 29, step B. The reverse transcriptase undergoes strand switching and copies the Ei tag sequence. Activate Taq polymerase and perform limited cycle PCR amplification (12-20), using dUTP, using tag primers (Di, Ei), to maintain relative ratios of different amplicons. The PCR products incorporate dU, allowing for carryover prevention (Figure 29, step C).

[0237] As shown in Figure 29, step D miRNA sequence-specific ligation probes containing primer-specific portions (Ai, Ci') suitable for subsequent PCR amplification, hybridize to their corresponding target sequence in a base-specific manner. Following ligation, the ligation products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the ligation junction as described *supra* for Figure 2 (see Figure 29, steps D-F), or using other suitable means known in the art.

[0238] Another aspect of the present application is directed to a method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases. The method involves providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample, and the contacted sample is blended with a ligase and one or more first oligonucleotide probes comprising a 5' phosphate, a 5' stem-loop portion, an internal primer-specific portion within the loop region, a blocking group, and a 3' nucleotide sequence that is complementary to a 3' portion of the target miRNA molecule sequence to form one or more ligation reaction mixtures. The method further involves ligating, in the one or more ligation reaction mixtures, the one or more target miRNA molecules at their 3'end to the 5' phosphate of the one or more first oligonucleotide probes to generate chimeric nucleic acid molecules comprising the target miRNA molecule sequence, if present in the sample, appended to the one or more first oligonucleotide probes. One or more primary oligonucleotide primer sets are then provided. Each primer set comprises (a) a first primary oligonucleotide primer comprising a nucleotide sequence that is complementary to the internal primer-specific portion of the first oligonucleotide probe, and (b) a second primary

oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets. The one or more ligation reaction mixtures comprising chimeric nucleic acid molecules, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the chimeric nucleic acid molecules, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different primary reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion, a nucleotide sequence corresponding to the target miRNA molecule sequence, and the complement of the internal primer-specific portion, and complements thereof. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer. The primary reverse-transcription/polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising a 5' primer-specific portion of the first secondary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence or a complement thereof, and a complement of the other 5' primer-specific portion second secondary oligonucleotide primer. The method further involves providing one or more tertiary oligonucleotide primer sets. Each tertiary oligonucleotide primer set comprises (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products or their complements and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction

products or their complements. The first polymerase chain reaction process products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures, and the one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU) containing nucleic acid molecules present in the secondpolymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further comprises detecting and distinguishing the second polymerase chain reaction products, thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

[0239] Figure 30 illustrates an embodiment of this aspect of the present application.

[0240] Figure 30 illustrates an exemplary Ligation-RT-PCR-qPCR carryover prevention reaction to quantify miRNA. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. This method involves isolating miRNA from exosomes and treating with UDG for carryover prevention (Figure 30, step B). An oligonucleotide probe having a portion that is complementary to the 3' end of the target miRNA, and containing a stem-loop, tag (Di'), and blocking group (filled circle) is ligated at its 5' end to the 3' end of the target miRNA. The ligation product comprises the miRNA, Di' tag, the blocking group, and a sequence complementary to the 3' portion of the miRNA (Figure 30, step B). A reverse transcriptase extends primer (Di) to make a full-length copy of the target and appends three C bases to the 3' end of extended target sequence (Figure 30, step B). Tag oligonucleotide (Ei) having the 3' rGrG+G hybridizes to the three C bases of the extended target sequence as shown in Figure 30, step B. The reverse transcriptase undergoes strand switching and copies the Ei tag sequence. Activate Taq polymerase and perform limited cycle PCR amplification (8-20), using tag primers (Di, Ei), to maintain relative ratios of different amplicons.

[0241] As shown in Figure 30, step C, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes across the miRNA target region, miRNA-specific (forward) primers comprising 5' primer-specific portions (Ai), cDNA-specific (reverse) primers comprising 5' primer-specific portions (Ci) and matching primers Ai and Ci. These primers combine to amplify the target miRNA sequence, if present in the sample (Figure 30, step C). Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of

matched primers Ai and Ci, and TaqMan™ probes that span the target miRNA regions as described *supra* for Figure 4 (see Figure 30, steps C-F), or using other suitable means known in the art.

[0242] In one embodiment, the 3' portion of the second primary oligonucleotide primer comprises ribo-G and/or G nucleotide analogue, wherein the reverse transcriptase appends two or three cytosine nucleotides to the 3' end of the complementary deoxyribonucleic acid products of the target miRNAs, enabling transient hybridization to the 3' end of the second primary oligonucleotide primer, enabling the reverse transcriptase to undergo strand switching and to extend the complementary deoxyribonucleic acid products to include the complementary sequence of the 5' primer-specific portion of the second primary oligonucleotide primer to form the one or more different first polymerase chain reaction products comprising a 5' primer-specific portion, a nucleotide sequence portion corresponding to the target miRNA molecule sequence or a complement thereof, a further portion, and a complement of the other 5' primer-specific portion.

[0243] In another embodiment, the 3' portion of the second primary oligonucleotide primers contains from 6 to 14 bases comprising, from 5' to 3', three ribo-G or G bases, followed by additional bases that are the same as the 5' end of the target miRNA sequences, wherein the reverse transcriptase appends two or three cytosine residues to the 3' end of the initial complementary deoxyribonucleic acid extension products of the target miRNAs, and wherein subsequent to when the denaturation treatment of the polymerase chain reaction is initiated the conditions are adjusted to enable transient hybridization to the 3' end of the second primary oligonucleotide primers to the 3' end of the complementary deoxyribonucleic acid extension products, allowing for extension of either one or both the second primary oligonucleotide primers and the complementary deoxyribonucleic acid extension products to form the one or more different primary reverse-transcription/polymerase chain reaction products comprising a 5' primer-specific portion, a nucleotide sequence portion corresponding to the target miRNA molecule sequence or a complement thereof, a further portion, and a complement of the other 5' primer-specific portion.

[0244] In certain embdiments, the second oligonucleotide probe of the oligonucleotide probe set further comprises a unitaq detection portion, thereby forming ligated product sequences comprising the 5' primer-specific portion, the target-specific portions, the unitaq detection portion, and the 3' primer-specific portion. In accorandce with this embodiment, one or more unitaq detection probes are provided, wherein each unitaq detection probe hybridizes to a complementary unitaq detection portion and said detection probe comprises a quencher molecule and a detectable label separated from the quencher molecule. The one or more unitaq detection

probes are added to the second polymerase chain reaction mixture, and the one or more unitaq detection probes are hybridized to complementary unitaq detection portions on the ligated product sequence or complement thereof during said subjecting the second polymerase chain reaction mixture to conditions suitable for one or more polymerase chain reaction cycles, wherein the quencher molecule and the detectable label are cleaved from the one or more unitaq detection probes during the extension treatment and said detecting involves the detection of the cleaved detectable label.

[0245] In another embodiment, one primary oligonucleotide primer or one secondary oligonucleotide primer further comprises a unitaq detection portion, thereby forming extension product sequences comprising the 5' primer-specific portion, the target-specific portions, the unitaq detection portion, and the complement of the other 5' primer-specific portion, and complements thereof. In accordance with this embodiment, one or more unitaq detection probes are provided, wherein each unitaq detection probe hybridizes to a complementary unitaq detection portion and said detection probe comprises a quencher molecule and a detectable label separated from the quencher molecule. The one or more unitaq detection probes are added to the one or more first or second polymerase chain reaction mixtures, and the one or more unitaq detection probes are hybridized to complementary unitaq detection portions on the ligated product sequence or complement thereof during polymerase chain reaction cycles after the first polymerization chain reaction, wherein the quencher molecule and the detectable label are cleaved from the one or more unitaq detection probes during the extension treatment and said detecting involves the detection of the cleaved detectable label.

[0246] In another embodiment, one or both oligonucleotide probes of the oligonucleotide probe set comprises a portion that has no or one nucleotide sequence mismatch when hybridized in a base-specific manner to the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof, but have one or more additional nucleotide sequence mismatches that interferes with ligation when said oligonucleotide probe hybridizes in a base-specific manner to a corresponding nucleotide sequence portion in the wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof.

[0247] In one embodiment, the 3' portion of the first oligonucleotide probe of the oligonucleotide probe set comprises a cleavable nucleotide or nucleotide analogue and a blocking group, such that the 3' end is unsuitable for polymerase extension or ligation. The cleavable nucleotide or nucleotide analog of the first oligonucleotide probe is cleaved when said probe is hybridized to it complementary target nucleotide sequence of the primary extension product, thereby liberating a 3'OH on the first oligonucleotide probe prior to the ligating.

[0248] The one or more first oligonucleotide probe of the oligonucleotide probe set may comprises a sequence that differs from the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof, said difference is located two or three nucleotide bases from the liberated free 3'OH end.

[0249] In a further embodiment, the second oligonucleotide probe has, at its 5' end, an overlapping identical nucleotide with the 3' end of the first oligonucleotide probe, and, upon hybridization of the first and second oligonucleotide probes of a probe set at adjacent positions on a complementary target nucleotide sequence of a primary extension product to form a junction, the overlapping identical nucleotide of the second oligonucleotide probe forms a flap at the junction with the first oligonucleotide probe. This embodiment further comprises cleaving the overlapping identical nucleotide of the second oligonucleotide probe with an enzyme having 5' nuclease activity thereby liberating a phosphate at the 5' end of the second oligonucleotide probe prior to said ligating.

[0250] In other embodiments, the one or more oligonucleotide probe sets further comprise a third oligonucleotide probe having a target-specific portion, wherein the second and third oligonucleotide probes of a probe set are configured to hybridize adjacent to one another on the target nucleotide sequence with a junction between them to allow ligation between the second and third oligonucleotide probes to form a ligated product sequence comprising the first, second, and third oligonucleotide probes of a probe set.

[0251] In certain embodiments, the sample is selected from the group consisting of tissue, cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, cell-free circulating nucleic acids, cell-free circulating tumor nucleic acids, cell-free circulating fetal nucleic acids in pregnant woman, circulating tumor cells, tumor, tumor biopsy, and exosomes.

[0252] The one or more target nucleotide sequences may be low-abundance nucleic acid molecules comprising one or more nucleotide base mutations, insertions, deletions, translocations, splice variants, miRNA variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicings, exon insertions, exon deletions, intron insertions, or other rearrangement at the genome level and/or methylated nucleotide bases.

[0253] As used herein "low abundance nucleic acid molecule" refers to a target nucleic acid molecule that is present at levels as low as 1% to 0.01% of the sample. In other words, a low abundance nucleic acid molecule with one or more nucleotide base mutations, insertions, deletions, translocations, splice variants, miRNA variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicings, exon

insertions, exon deletions, intron insertions, other rearrangement at the genome level, and/or methylated nucleotide bases can be distinguished from a 100 to 10,000-fold excess of nucleic acid molecules in the sample (i.e., high abundance nucleic acid molecules) having a similar nucleotide sequence as the low abundance nucleic acid molecules but without the one or more nucleotide base mutations, insertions, deletions, translocations, splice variants, miRNA variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicings, exon insertions, exon deletions, intron insertions, other rearrangement at the genome level, and/or methylated nucleotide bases.

[0254] In some embodiments of the present invention, the copy number of one or more low abundance target nucleotide sequences are quantified relative to the copy number of high abundance nucleic acid molecules in the sample having a similar nucleotide sequence as the low abundance nucleic acid molecules. In other embodiments of the present invention, the one or more target nucleotide sequences are quantified relative to other nucleotide sequences in the sample. In other embodiments of the present invention, the relative copy number of one or more target nucleotide sequences is quantified. Methods of relative and absolute (i.e., copy number) quantitation are well known in the art.

[0255] The low abundance target nucleic acid molecules to be detected can be present in any biological sample, including, without limitation, tissue, cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, cell-free circulating nucleic acids, cell-free circulating tumor nucleic acids, cell-free circulating fetal nucleic acids in pregnant woman, circulating tumor cells, tumor, tumor biopsy, and exosomes.

[0256] The methods of the present invention are suitable for diagnosing or prognosing a disease state and/or distinguishing a genotype or disease predisposition.

[0257] Another aspect of the present application is directed to a method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases. The method involves providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample. The contacted sample is blended with ATP and a Poly(A) polymerase to form a Poly(A) polymerase reaction mixture, and the Poly(A) polymerase reaction mixture is subjected to conditions suitable for appending homopolymer A to the 3' ends of the one or more target miRNA molecules potentially present in the sample. The method further involves providing one or more primary oligonucleotide primer

sets. Each primer set comprises (a) a first primary oligonucleotide primer comprising a 5' primer-specific portion, an internal poly dT portion, and a 3' portion comprising from 1 to 10 bases complementary to the 3' end of the target miRNA, wherein the first primary oligonucleotide primer may be the same or may differ from other first primary oligonucleotide primers in other sets, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets. The Poly(A) polymerase reaction mixture, the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the sample, a deoxynucleotide mix including dUTP, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity are blended to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the reverse-transcription/polymerase chain reaction mixtures, then to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target miRNA sequences with 3' polyA tails, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion of the second primary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence, a poly dA region, and the complement of the 5' primer-specific portion of the first primary oligonucleotide primer, and complements thereof. The method further comprises providing one or more oligonucleotide probe sets. Each probe set comprises (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion, a portion complementary to the one or more reverse-transcription/polymerase chain reaction products, and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, to complementary portions of the one or more reverse-transcription/polymerase chain reaction products corresponding to the target miRNA molecule sequence, or complement thereof. The one or more reverse-transcription/polymerase chain reaction products are contacted with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures, and the one or more ligation reaction mixtures are subjected to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form

ligated product sequences in the ligation reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion. The method further involves providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence. The ligated product sequences and the one or more secondary oligonucleotide primer sets are blended with one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain reaction products. The method further comprises detecting and distinguishing the secondary polymerase chain reaction products, thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

[0258] Figure 31 illustrates an embodiment of this aspect of the present application.

[0259] Figure 31 illustrates an exemplary RT-PCR-LDR-qPCR carryover prevention reaction to quantify miRNA. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. This method involves isolating miRNA from exosomes and treating with UDG for carryover prevention (Figure 31, step B). PolyA tail miRNA with *E. coli* Poly(A) polymerase. A reverse transcriptase extends primer comprising a Tag Di on the 5' end, and a T30VN sequence at the 3' end, to make a full-length copy of the target, and appends three C bases to the 3' end of extended target sequence (Figure 31, step B). Tag oligonucleotide (Ei) having the 3' rGrG+G hybridizes to the three C bases of the extended target sequence as shown in Figure 31, step B. The reverse transcriptase undergoes strand switching and copies the Ei tag sequence. Activate Taq polymerase and perform limited cycle PCR amplification (12-20), using dUTP, using tag primers (Di, Ei), to maintain relative ratios of different amplicons. The PCR products incorporate dU, allowing for carryover prevention (Figure 31, step C).

[0260] As shown in Figure 31, step D miRNA sequence-specific ligation probes containing primer-specific portions (Ai, Ci') suitable for subsequent PCR amplification,

hybridize to their corresponding target sequence in a base-specific manner. Following ligation, the ligation products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the ligation junction as described *supra* for Figure 2 (see Figure 31, steps D-F), or using other suitable means known in the art.

[0261] Another aspect of the present application is directed to a method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases. The method involves providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases and providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample. The sample is contacted with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample. The contacted sample is blended with ATP and a Poly(A) polymerase to form a Poly(A) polymerase reaction mixture, and the Poly(A) polymerase reaction mixture is subjected to conditions suitable for appending a homopolymer A to the 3' ends of the one or more target miRNA molecules potentially present in the sample. The method further involves providing one or more primary oligonucleotide primer sets. Each primer set comprises (a) a first primary oligonucleotide primer comprising a 5' primer-specific portion, an internal poly dT portion, and a 3' portion comprising from 1 to 10 bases complementary to the 3' end of the target miRNA, wherein the first primary oligonucleotide primer may be the same or may differ from other first primary oligonucleotide primers in other sets, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets. The Poly(A) polymerase reaction mixture potentially comprising target miRNA sequences is blended with 3' polyA tails, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity to form one or more reverse-transcription/polymerase chain reaction mixtures. The one or more reverse-transcription/polymerase chain reaction mixtures are subjected to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target miRNA sequences with 3' polyA tails, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more different reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion of the second primary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence, a poly dA region, and the complement of the 5' primer-specific portion of the first

primary oligonucleotide primer, and complements thereof. The method further comprises providing one or more secondary oligonucleotide primer sets. Each secondary oligonucleotide primer set comprises (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of a reverse-transcription/polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a reverse-transcription/polymerase chain reaction product formed from the first secondary oligonucleotide primer. The reverse-transcription/polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase are blended to form one or more first polymerase chain reaction mixtures, and the one or more first polymerase chain reaction mixtures are subjected to conditions suitable for two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising a 5' primer-specific portion, a nucleotide sequence corresponding to the target miRNA molecule sequence or a complement thereof, and a complement of the other 5' primer-specific portion. The method further involves providing one or more tertiary oligonucleotide primer sets. Each tertiary oligonucleotide primer set comprises (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction product sequence and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction product sequence. The first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase are blended to form one or more second polymerase chain reaction mixtures. The one or more second polymerase chain reaction mixtures are subjected to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures, and one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products. The method further comprises detecting and distinguishing the second polymerase chain reaction products in the one or more reactions thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

[0262] Figure 32 illustrates an embodiment of this aspect of the present application.

[0263] Figure 32 illustrates an exemplary RT-PCR-qPCR carryover prevention reaction to quantify miRNA. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. This method involves isolating miRNA from exosomes and treating with UDG for carryover prevention (Figure 32, step B). PolyA tail miRNA with *E. coli* Poly(A) polymerase. A reverse transcriptase extends primer comprising a Tag Di on the 5' end, and a T30VN sequence at the 3' end, to make a full-length copy of the target, and appends three C bases to the 3' end of extended target sequence (Figure 32, step B). Tag oligonucleotide (Ei) having the 3' rGrG+G hybridizes to the three C bases of the extended target sequence as shown in Figure 32, step B. The reverse transcriptase undergoes strand switching and copies the Ei tag sequence. Activate Taq polymerase and perform limited cycle PCR amplification (8-20), using dUTP, using tag primers (Di, Ei), to maintain relative ratios of different amplicons.

[00124] As shown in Figure 32, step C, following the limited cycle PCR, the PCR products are aliquoted into separate wells, micro-pores or droplets containing Taqman™ probes across the miRNA target region, miRNA-specific (forward) primers comprising 5' primer-specific portions (Ai), cDNA-specific (reverse) primers comprising 5' primer-specific portions (Ci) and matching primers Ai and Ci. These primers combine to amplify the target miRNA sequence, if present in the sample (Figure 32, step C). Primers are unblocked with RNaseH2 only when bound to correct target. Following PCR, the products can be detected using pairs of matched primers Ai and Ci, and TaqMan™ probes that span the target miRNA regions as described *supra* for Figure 6 (see Figure 32, steps C-F), or using other suitable means known in the art.

[0264] Another aspect of the present application is directed to a method of diagnosing or prognosing a disease state of cells or tissue based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual. The plurality of markers is in a set comprising from 6-12 markers, 12-24 markers, 24-36 markers, 36-48 markers, 48-72 markers, 72-96 markers, or > 96 markers. Each marker in a given set is selected by having any one or more of the following criteria: present, or above a cutoff level, in > 50% of biological samples of the disease cells or tissue from individuals diagnosed with the disease state; absent, or below a cutoff level, in > 95% of biological samples of the normal cells or tissue from individuals without the disease state; present, or above a cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with the disease state; absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid,

sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without the disease state; present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with the disease state. At least 50% of the markers in a set each comprise one or more methylated residues, and/or at least 50% of the markers in a set that are present, or above a cutoff level, or present with a z-value of > 1.65, comprise of one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with the disease state. The method involves obtaining a biological sample. The biological sample includes cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, and the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof. The sample is fractionated into one or more fractions, wherein at least one fraction comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein. Nucleic acid molecules in the one or more fractions are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. At least two enrichment steps are carried out for 50% or more disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step. The method further involves performing one or more assays to detect and distinguish the plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with the disease state if a minimum of 2 or 3 markers are present or are above a cutoff level in a marker set comprising from 6-12 markers; or a minimum of 3, 4, or 5 markers are present or are above a cutoff level in a marker set comprising from 12-24 markers; or a minimum of 3, 4, 5, or 6 markers are present or are above a cutoff level in a marker set comprising from 24-36 markers; or a minimum of 4, 5, 6, 7, or 8 markers are present or are above a cutoff level in a marker set comprising from 36-48 markers; or a minimum of 6, 7, 8, 9, 10, 11, or 12 markers are present or are above a cutoff level in a marker set comprising from 48-72 markers, or a minimum of 7, 8, 9, 10, 11, 12 or 13 markers are present or are above a cutoff level in a marker set comprising from 72-96 markers, or a minimum of 8, 9, 10, 11, 12, 13 or "n"/12 markers are present or are above a cutoff level in a marker set comprising 96 – "n" markers, when "n" > 168 markers.

[0265] Another aspect of the present application is directed to a method of diagnosing or prognosing a disease state of a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual. The plurality of markers is in a set comprising from 48-72 total cancer markers, 72-96 total cancer markers or ≥ 96 total cancer markers, wherein on average greater than one quarter such markers in a given set cover each of the aforementioned major cancers being tested. Each marker in a given set for a given solid tissue cancer is selected by having any one or more of the following criteria for that solid tissue cancer: present, or above a cutoff level, in > 50% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer; present, or above a cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer; present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer. At least 50% of the markers in a set each comprise one or more methylated residues, and/or at least 50% of the markers in a set that are present, or above a cutoff level, or present with a z-value of > 1.65, comprise of one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with a given solid tissue cancer. The method involves obtaining a biological sample including cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof. The sample is fractionated into one or more fractions, wherein at least one fraction

comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein. The nucleic acid molecules in the one or more fractions are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. At least two enrichment steps are carried out for 50% or more of the given solid tissue cancer-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step. The method further involves performing one or more assays to detect and distinguish the plurality of cancer -specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with the a solid-tissue cancer if a minimum of 4 markers are present or are above a cutoff level in a marker set comprising from 48-72 total cancer markers; or a minimum of 5 markers are present or are above a cutoff level in a marker set comprising from 72-96 total cancer markers; or a minimum of 6 or "n"/18 markers are present or are above a cutoff level in a marker set comprising 96 to "n" total cancer markers, when "n" > 96 total cancer markers.

[0266] In accoradance with this aspect of the present application, each marker in a given set for a given solid tissue cancer may be selected by having any one or more of the following criteria for that solid tissue cancer: present, or above a cutoff level, in > 66% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer; present, or above a cutoff level, in > 66% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer; present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer.

[0267] Another aspect of the present application is directed to a method of diagnosing or prognosing a disease state of and identifying the most likely specific tissue(s) of origin of a solid tissue cancer in the following groups: Group 1 (colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma); Group 2 (breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma); Group 3 (lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma); Group 4 (prostate

adenocarcinoma, invasive urothelial bladder cancer); and/or Group 5 (liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma) based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual, wherein the plurality of markers is in a set comprising from 36-48 group-specific cancer markers, 48-64 group-specific cancer markers, or ≥ 64 group-specific cancer markers, wherein on average greater than one third of such markers in a given set cover each of the aforementioned cancers being tested within that group. Each marker in a given set for a given solid tissue cancer is selected by having any one or more of the following criteria for that solid tissue cancer: present, or above a cutoff level, in > 50% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer; present, or above a cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer; present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer. At least 50% of the markers in a set each comprise one or more methylated residues, and/or at least 50% of the markers in a set that are present, or above a cutoff level, or present with a z-value of > 1.65 comprise one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with a given solid tissue cancer. The method involves obtaining the biological sample. The biological sample includes cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof. The sample is fractionated into one or more fractions, wherein at least one fraction comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein. The nucleic acid molecules in the one or more fractions are subjected to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues. At least two enrichment steps are carried out for 50% or more of the given solid tissue cancer-specific and/or

cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step. The method further involves performing one or more assays to detect and distinguish the plurality of cancer -specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with a solid-tissue cancer if a minimum of 4 markers are present or are above a cutoff level in a marker set comprising from 36-48 group-specific cancer markers; or a minimum of 5 markers are present or are above a cutoff level in a marker set comprising from 48-64 group-specific cancer markers; or a minimum of 6 or "n"/12 markers are present or are above a cutoff level in a marker set comprising 64 to "n" total cancer markers, when "n" > 64 group-specific cancer markers.

[0268] In accordance with this aspect of the present application, each marker in a given set for a given solid tissue cancer may be selected by having any one or more of the following criteria for that solid tissue cancer: present, or above a cutoff level, in > 66% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer; present, or above a cutoff level, in > 66% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer; absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer; present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer.

[0269] In one embodiment, the at least two enrichment steps comprise of two or more of the following steps: capturing or separating exosomes or extracellular vesicles or markers in other protected states; capturing or separating a platelet fraction; capturing or separating circulating tumor cells; capturing or separating RNA-containing complexes; capturing or separating cfDNA-nucleosome or differentially modified cfDNA-histone complexes; capturing or separating protein targets or protein target complexes; capturing or separating auto-antibodies; capturing or separating cytokines; capturing or separating methylated cfDNA; capturing or separating marker specific DNA, cDNA, miRNA, lncRNA, ncRNA, or mRNA, or amplified complements, by hybridization to complementary capture probes in solution, on magnetic beads, or on a microarray; amplifying miRNA markers, non-coding RNA markers (lncRNA & ncRNA markers), mRNA markers, exon markers, splice-variant markers, translocation markers, or copy

number variation markers in a linear or exponential manner via a polymerase extension reaction, polymerase chain reaction, bisulfite-methyl-specific polymerase chain reaction, reverse-transcription reaction, bisulfite-methyl-specific ligation reaction, and/or ligation reaction, using DNA polymerase, reverse transcriptase, DNA ligase, RNA ligase, DNA repair enzyme, RNase, RNaseH2, endonuclease, restriction endonuclease, exonuclease, CRISPR, DNA glycosylase or combinations thereof; selectively amplifying one or more target regions containing mutation markers or bisulfite-converted DNA methylation markers, while suppressing amplification of the target regions containing wild-type sequence or bisulfite-converted unmethylated sequence or complement sequence thereof, in a linear or exponential manner via a polymerase extension reaction, polymerase chain reaction, bisulfite-methyl-specific polymerase chain reaction, reverse-transcription reaction, bisulfite-methyl-specific ligation reaction, and/or ligation reaction, using DNA polymerase, reverse transcriptase, DNA ligase, RNA ligase, DNA repair enzyme, RNase, RNaseH2, endonuclease, restriction endonuclease, exonuclease, CRISPR, DNA glycosylase or combinations thereof; preferentially extending, ligating, or amplifying one or more primers or probes whose 3'-OH end has been liberated in an enzyme and sequence-dependent process; using one or more blocking oligonucleotide primers comprising one or more mismatched bases at the 3' end or comprising one or more nucleotide analogs and a blocking group at the 3' end under conditions that interfere with polymerase extension or ligation during said reaction of target-specific primer or probes hybridized in a base-specific manner to wildtype sequence or bisulfite-converted unmethylated sequence or complement sequence thereof.

[0270] In certain embodiment, the one or more assays to detect and distinguish the plurality of disease-specific and/or cell/tissue-specific DNA, RNA, or protein markers comprise one or more of the following: a quantitative real-time PCR method (qPCR); a reverse transcriptase-polymerase chain reaction (RTPCR) method; a bisulfite qPCR method; a digital PCR method (dPCR); a bisulfite dPCR method; a ligation detection method, a ligase chain reaction, a restriction endonuclease cleavage method; a DNA or RNA nuclease cleavage method; a micro-array hybridization method; a peptide-array binding method; an antibody-array method; a mass spectrometry method; a liquid chromatography-tandem mass spectrometry (LC-MS/MS) method; a capillary or gel electrophoresis method; a chemiluminescence method; a fluorescence method; a DNA sequencing method; a bisulfite conversion-DNA sequencing method; an RNA sequencing method; a proximity ligation method; a proximity PCR method; a method comprising immobilizing an antibody-target complex; a method comprising immobilizing an aptamer-target complex; an immunoassay method; a method comprising a Western blot assay; a method comprising an enzyme linked immunosorbent assay (ELISA); a method comprising a high-throughput microarray-based enzyme-linked immunosorbent assay (ELISA); or a method

comprising a high-throughput flow-cytometry-based enzyme-linked immunosorbent assay (ELISA).

[0271] In certain embodiments, the one or more cutoff levels of the one or more assays to detect and distinguish the plurality of disease-specific and/or cell/tissue-specific DNA, RNA, or protein markers comprise one or more of the following calculations, comparisons, or determinations, in the one or more marker assays comparing samples from the disease vs. normal individual: marker ΔCt value is > 2; marker ΔCt value is > 4; ratio of detected marker-specific signal is > 1.5; ratio of detected marker-specific signal is > 3; ratio of marker concentrations is > 1.5; ratio of marker concentrations is > 3; enumerated marker-specific signals differ by > 20%; enumerated marker-specific signals differ by > 50%; marker-specific signal from a given disease sample is > 85%; > 90%; > 95%; > 96%; > 97%; or > 98% of the same marker-specific signals from a set of normal samples; or marker-specific signal from a given disease sample has a z-score of > 1.03; > 1.28; > 1.65; > 1.75; > 1.88; or > 2.05 compared to the same marker-specific signals from a set of normal samples.

[0272] Another aspect of the present application relates to a two-step method of diagnosing or prognosing a disease state of cells or tissue based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual. The method involves obtaining a biological sample, the biological sample including exosomes, tumor-associated vesicles, markers within other protected states, cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof. A first step is applied to the biological samples with an overall sensitivity of > 80% and an overall specificity of > 90% or an overall Z-score of > 1.28 to identify individuals more likely to be diagnosed or prognosed with the disease state. A second step is then applied to biological samples from those individuals identified in the first step with an overall specificity of > 95% or an overall Z-score of > 1.65 to diagnose or prognose individuals with the disease state. The first step and the second step are carried out using a method of the present application. The first step uses markers to cover many cancers, where the aim is to obtain high sensitivity for early cancers where the number of marker molecules in the blood may be limited. The second step then would score for additional markers to verify that the initial result was a true positive, as well as to identify the likely tissue of origin. The second step may include the methods described herein, and/or additional methods such as next-generation sequencing.

[0273] **Fluorescent labeling**. Consider an instrument that can detect 5 fluorescent signals, F1, F2, F3, F4, and F5, respectively. As an example, in the case of colon cancer, the highest frequency mutations will be found for K-ras, p53, APC and BRAF. Mutations in these four genes could be detected with a single fluorescent signal; F1, F2, F3, F4. If the scale is 1000 FU, then primer would be added using ratios of labeled and unlabeled UniTaq primers, such that amplification of LDR products on mutant target of these genes yields about 300 FU at the plateau. For the controls, the F5 would be calibrated to give a signal of 100 FU for a 1:1,000 dilution quantification control, and an additional 300 FU for ligation of mutant probe on wild-type control (should give no or low background signal).

[0274] For the other genes commonly mutated in colon cancer as shown below, (or even lower abundance mutations in the p53 gene,) the following coding system may be used: Two fluorescent signals in equimolar amount at the 5' end of the same UniTaq, with unlabeled primer titrated in, such that both fluorescent signals plateau at 100 FU. If fluorescent signals are F1, F2, F3, F4, then that gives the ability to detect mutations in 4 genes using a single fluorescent signal, and in mutations in 6 genes using combinations of fluorescent signal:

Gene 1 = F1 (300 FU) (p53, Hot Spots)
Gene 2 = F2 (300 FU) (KRAS)
Gene 3 = F3 (300 FU) (APC)
Gene 4 = F4 (300 FU) (BRAF)
Gene 5 = F1 (100 FU), F2 (100 FU) (PIK3CA)
Gene 6 = F1 (100 FU), F3 (100 FU) (FBXW7)
Gene 7 = F1 (100 FU), F4 (100 FU) (SMAD4)
Gene 8 = F2 (100 FU), F3 (100 FU) (p53, additional)
Gene 9 = F2 (100 FU), F4 (100 FU) (CTNNB1)
Gene 10= F3 (100 FU), F4 (100 FU) (NRAS)

[0275] Suppose there is a second mutation, combined with a mutation in one of the top genes. This is easy to distinguish, since the top gene will always give more signal, independent if it is overlapping with the other fluorescent signals or not. For example, if the fluorescent signal is F1 100 FU, and F2 400 FU, that would correspond to mutations in Gene 2 and Gene 5.

[0276] If there are two mutations from the less commonly mutated genes (Gene 5- Gene 10) then the results will appear either as an overlap in fluorescent signals, *i.e.* F1 200 FU, F2 100 FU, F4 100 FU, or all 4 fluorescent signals. If the fluorescent signals are in the ratio of 2:1:1, then it is rather straightforward to figure out the 2 mutations: in the above example, F1 200 FU, F2 100 FU, F4 100 FU, would correspond to mutations in Gene 5 and Gene 7. A similar approach for multiplexing different colors has been described by the Kartalov group (Rajagopal et al., "Supercolor Coding Methods for Large-Scale Multiplexing of Biochemical Assays," *Anal.*

Chem. 85(16):7629-36 (2013); U.S. Patent Application Publication No. 20140213471A1, which are hereby incorporated by reference in their entirety).

[0277] More recently, digital droplet PCR (ddPCR) has been used to provide accurate quantification of the number of mutant or methylated molecules in a clinical sample. In general, amplification in a droplet implies at least a single molecule of the target was present in that droplet. Thus, when using a sufficient number of droplets that way exceed the number of initial targets, it is assumed that a given droplet only had a single molecule of the target. Thus, end-point PCR is often used to monitor the number of products.

[0278] Consider an instrument that can detect 5 fluorescent signals, F1, F2, F3, F4, and F5, respectively. Methylation in the promoter region of some genes often methylated in colon cancer could be used for the first four channels, for example F1 = VIM, F2 = SEPT9, F3 = CLIP4, and F4 = GSG1L. The last channel, F5 would be used as a control to assure a given droplet contained proper reagents, etc. Once again, combinations of fluorescent signal may be used to simultaneously detect methylation at 10 different promoter regions.

Gene 1 = F1	(VIM)
Gene 2 = F2	(SEPT9)
Gene 3 = F3	(CLIP4
Gene 4 = F4	(GSG1L)
Gene 5 = F1 + F2	(PP1R16B)
Gene 6 = F1 + F3	(KCNA3)
Gene 7 = F1 + F4	(GDF6)
Gene 8 = F2 + F3	(ZNF677)
Gene 9 = F2 + F4	(CCNA1)
Gene 10= F3 + F4	(STK32B)

[0279] For simplicity, consider how ddPCR may be used to accurately enumerate the number of original methylated molecules at 4 promoter regions using exPCR-ddPCR (see for example, Figures 4, 5, 6, 7, 8, 11, 12, 13, 14, 15, 18, 19, 20, 21 & 22). The approach also works using PCR-LDR-qPCR or exPCR-LDR-qPCR (see Figures 2, 3, 9, 10, 16 & 17). For this illustration, consider a total of 48 methylated regions are being detected, with 4 promoter regions in a single ddPCR reaction comprising 10,000 droplets or micro-pores or micro-wells. Consider a sample with 2, 4, 5, and 1 molecule(s) of methylated promoter regions for VIM, SEPT9, CLIP4, and GSG1L, respectively. Assume the initial one-sided primer extension with blocking primer has an efficiency of 50%, so after 20 cycles, there are = 20; 40; 50; and 10 extension products of methylated promoter regions for VIM, SEPT9, CLIP4, and GSG1L, respectively. Also, with blocking primer for the top strand, again, assuming a general efficiency of 50%, after 10 cycles of PCR, there are (1.5 to the 10th = 57 x number of initial extension products) = 1,140;

2,280; 2,850; and 570 copies of the PCR products for methylated VIM, SEPT9, CLIP4, and GSG1L respectively. When such products are then diluted into 12 ddPCR reactions, on average, a given chamber will comprise of 95; 190; 237; and 48 copies of the PCR products for methylated VIM, SEPT9, CLIP4, and GSG1L, respectively. This is a total of about 570 of molecules that would be amplified with primers for the total PCR products for methylated VIM, SEPT9, CLIP4, and GSG1L. If the ddPCR comprises 10,000 droplets or micro-pores or micro-wells, on average, only 1 in 20 will actually comprise a PCR reaction; the chances of a given droplet having two amplicons that would compete with each other for resources would be about 1 in 400, or about 25 droplets would comprise 2 amplicons, which would be only 5% of the total number of droplets with only a single amplicon. Since there are 6 combinations of 2 different amplicons, on average, less than 2% of the droplets would contain two amplicons. In other words, the rare droplet comprising 2 or 3 or 4 colors would not need to be de-convoluted, they could simply be ignored as they represent approximately 4-6 droplets compared to about 48 droplets arising from a single molecule in the original sample. While it may be a bit difficult to distinguish 190 from 237 droplets, *i.e.* starting with 4 or 5 molecules of a given methylated target, it should be relatively straightforward to distinguish 95; 190; and 48 copies, corresponding to 2, 4, and 1 target molecules in the original sample.

[0280] For distinguishing and enumerating 10 methylation markers simultaneously in a single ddPCR reaction, consider a total of 50 methylated regions are being detected, with 10 promoter regions in a single ddPCR reaction comprising 10,000 droplets or micro-pores or micro-wells. Consider a sample with 2, 4, 5, 1, 0, 1, 3, 2, 0, and 1 molecule(s) of methylated promoter regions for VIM, SEPT9, CLIP4, GSG1L, PP1R16B, KCNA3, GDF6, ZNF677, CCNA1, and STK32B, respectively. Assume the initial one-sided primer extension with blocking primer has an efficiency of 50%, so after 20 cycles, there are = 20; 40; 50; 10; 0; 10; 30; 20; 0; and 10 extension products of methylated promoter regions for VIM, SEPT9, CLIP4, GSG1L, PP1R16B, KCNA3, GDF6, ZNF677, CCNA1, and STK32B, respectively. Also, with blocking primer for the top strand, again, assuming a general efficiency of 50%, after 6 cycles of PCR, there are (1.5 to the 6th = 11 x number of initial extension products) = 220; 440; 550; 110; 0; 110; 330; 220; 0; and 110 copies of the PCR products for methylated VIM, SEPT9, CLIP4, GSG1L, PP1R16B, KCNA3, GDF6, ZNF677, CCNA1, and STK32B, respectively. When such products are then diluted into 5 ddPCR reactions, on average, a given chamber will comprise of 44; 88; 110; 22; 0; 22; 66; 44; 0; and 22 copies of the PCR products for methylated VIM, SEPT9, CLIP4, GSG1L, PP1R16B, KCNA3, GDF6, ZNF677, CCNA1, and STK32B, respectively. This is a total of about 418 of molecules that would be amplified with primers for the total PCR products for methylated VIM, SEPT9, CLIP4, GSG1L, PP1R16B, KCNA3,

GDF6, ZNF677, CCNA1, and STK32B. If the ddPCR comprises 10,000 droplets or micro-pores or micro-wells, on average, only 1 in 25 will actually comprise a PCR reaction; the chances of a given droplet having two amplicons that would compete with each other for resources would be about 1 in 625, or about 16 droplets would comprise 2 amplicons, which would be only 4% of the total number of droplets with only a single amplicon. Since there are 45 combinations of 2 different amplicons, on average, less than 0.1% of the droplets would contain a given two amplicons. In other words, the rare droplet comprising 2 or 3 or 4 colors would not need to be de-convoluted, they could simply be ignored as they represent one or two droplets compared to about 22 droplets arising from a single molecule in the original sample. While it may be a bit difficult to distinguish 88 from 110 droplets, *i.e.* starting with 4 or 5 molecules of a given methylated target, it should be relatively straightforward to distinguish 44, 88, and 22 copies, corresponding to 2, 4, and 1 target molecules in the original sample.

[0281] The above approach would also work for accurately enumerating mRNA, miRNA, ncRNA, or lncRNA target molecules. Instead of aliquoting the sample as described in step B of Figures 23-32, the sample is used directly for subsequent ddPCR enumeration. For distinguishing and enumerating 10 mRNA, ncRNA, or lncRNA markers simultaneously in a single ddPCR reaction, consider a total of 50 mRNA, ncRNA, or lncRNA regions are being detected in a single ddPCR reaction comprising 10,000 droplets or micro-pores or micro-wells. Once again, combinations of fluorescent signal may be used to simultaneously detect 10 mRNA or lncRNA markers.

Gene 1 = F1	(mRNA1)
Gene 2 = F2	(mRNA2)
Gene 3 = F3	(mRNA3)
Gene 4 = F4	(mRNA4)
Gene 5 = F1 + F2	(ncRNA5)
Gene 6 = F1 + F3	(ncRNA6)
Gene 7 = F1 + F4	(ncRNA7)
Gene 8 = F2 + F3	(ncRNA8)
Gene 9 = F2 + F4	(ncRNA9)
Gene 10= F3 + F4	(ncRNA10)

[0282] Consider a sample with 2, 4, 15, 1, 0, 10, 3, 20, 0, and 1 molecule(s) of mRNA1-4 and ncRNA5-10, respectively, using essentially the RT-PCR-PCR-qPCR example illustrated in Figure 28. Six cycles of RT-PCR will generate 64 cDNA copies of each transcript generating = 128; 256; 960; 64; 0; 640; 192; 1280; 0; and 64 copies of mRNA1-4 and ncRNA5-10, respectively. When such products are then diluted into 5 ddPCR reactions, on average, a given chamber will comprise of 25; 51; 192; 13; 0; 128; 28; 256; 0; and 13 copies of the PCR products

for mRNA1-4 and ncRNA5-10, respectively. This is a total of about 706 of molecules that would be amplified with primers for the total PCR products for methylated mRNA1-4 and ncRNA5-10. If the ddPCR comprises 10,000 droplets or micro-pores or micro-wells, on average, only 1 in 14 will actually comprise a PCR reaction. The two most common RNA's in this example; mRNA 3 and ncRNA5 would be present on average of 1 in 52 and 1 in 39, thus the chances of a given droplet having these two amplicons that would compete with each other for resources would be about 1 in 2028, or about 5 droplets would comprise 2 amplicons, which is still less than for a single molecule after amplification – which will generate 13 copies. In other words, the rare droplet comprising 2 or 3 or 4 colors would not need to be de-convoluted, they could simply be ignored as they represent from 1 to 5 droplets compared to at least 13 droplets arising from a single molecule in the original sample. If some RNA molecules are present in higher amounts, one can still de-convolute multiple signals arising from 2 amplicons in a given droplet, using the same approach of different color probes at different levels of FU (*i.e.* 300 FU for products with a single color; 100 FU each for products using 2 colors) as articulated earlier.

[0283] Another aspect of the present application relates to the ability to distinguish cancer at the earliest stages when analyzing markers within a blood sample. The average body contains about 6 liters (6,000 ml) of blood. A 10 ml sample will then comprise 1/600th of the sample. While some cancers (*i.e.* lung cancer, melanoma) have a high mutational load, other cancers (*i.e.* breast, ovarian) have few mutations, and even fewer at the earliest stages. In contrast, methylation changes in promoter regions (*i.e.* methylation markers) appear to be early events. For the purposes of the calculations below, assume that if a marker is present in the sample, it can be detected down to the single molecule level, independent of the technology that is being used to identify the marker.

[0284] On a practical level, different cancers have different frequencies for different mutational markers. For example, the mutation rate for gene K-ras is ~30% and > 90% for colorectal cancer and pancreatic cancer, respectively. While p53 is found mutated in about 50% of all cancers, more often than not, such a mutation is manifested in late-stage tumors. As a benchmark, a given cancer during its earliest stage, generates at least one detectable mutation. Suppose that at any given time, there are 200 mutated molecules circulating in the plasma of the patient. Given the total volume, if there is a 10 ml sample, taken, then there is about a $1/3^{rd}$ chance that the sample will contain at least 1 mutated molecule. A more accurate prediction would be based on the Poisson distribution. If there are 200 objects (*i.e.* mutated molecules) distributed into 600 bins (*i.e.* 600 aliquots of 10 ml representing the total blood volume of a patient), Poisson calculation would indicate that: 72% of wells will have 0 objects, 23.7% will

have 1 object, 3.9% will have 2 objects, 0.4% will have 3 objects, etc. In other words, 28.1% of the aliquots would have at least one mutated molecule. If the assay is capable of detecting every single mutated molecule, then its sensitivity would be 28.1%. Likewise, if there were 300 objects (*i.e.* mutated molecules) distributed into 600 bins (*i.e.* 600 aliquots of 10 ml), then: 61% of wells will have 0 objects, 30.3% will have 1 object, 7.6% will have 2 objects, 1.3% will have 3 objects, etc. In other words, 39.4% of the aliquots would have at least one mutated molecule. If the assay is capable of detecting every single mutated molecule, its sensitivity is at 39.4%. Likewise, if there were 400 objects (*i.e.* mutated molecules) distributed into 600 bins (*i.e.* 600 aliquots of 10 ml), then: 51% of wells will have 0 objects, 34.3% will have 1 object, 11.5% will have 2 objects, 2.5% will have 3 objects, etc. In other words, 49% of the aliquots would have at least one mutated molecule. If the assay detects every single mutated molecule, its sensitivity would be 49%. Likewise, if there were 600 objects (*i.e.* mutated molecules) distributed into 600 bins (*i.e.* 600 aliquots of 10 ml), then the Poisson calculation would be: 36.8% of wells will have 0 objects, 36.8% will have 1 object, 18.3% will have 2 objects, 6.1% will have 3 objects, etc. In other words, 63.2% of the aliquots would have at least one mutated molecule. If the assay detects every single mutated molecule, then its sensitivity will be 63.2%. Nevertheless, on a practical level, even with a detectable marker load as high as 600 molecules, the assay would still miss 36.8% of early cancers for that type of tumor. Recent literature results have argued what constitutes "early cancer", with some groups claiming stage I & II cancers are early cancer, while others claiming that stages I, II, and III cancers are early cancer, both the definition and type varies, but general when scoring form mutations the results have reported sensitivities ranging from around 20% to around 70% -- which translates into missing 30% to 80% of early cancers (Klein et al., "Development of a Comprehensive Cell-free DNA (cfDNA) Assay for Early Detection of Multiple Tumor Types: The Circulating Cell-free Genome Atlas (CCGA) Study," *Journal of Clinical Oncology* 36(15):12021-12021 (2018); Liu et al., "Breast Cancer Cell-free DNA (cfDNA) Profiles Reflect Underlying Tumor Biology: The Circulating Cell-free Genome Atlas (CCGA) Study," *Journal of Clinical Oncology* 36(15):536-536 (2018), which are hereby incorporated by reference in their entirety).

[0285] The above calculations are performed based on the assumption that detecting even a single mutation is sufficient to call a patient positive. Initial work identifying mutations in the blood from patients with metastatic disease revealed an average of 5 mutations not only in the patients, but also in age-matched controls (Razavi, et al., "Cell-free DNA (cfDNA) Mutations From Clonal Hematopoiesis: Implications for Interpretation of Liquid Biopsy Tests," *Journal of Clinical Oncology* 35(15):11526-11526 (2017), which is hereby incorporated by reference in its entirety). This phenomenon, known as clonal hematopoiesis, results from accumulation of

mutations in white-blood cells, that then undergo clonal expansion. Once the presence of such mutations are accounted for (by sequencing an aliquot of WBC DNA from the same individual), the accuracy or specificity of these tests has been set at 98%. For some cancers like ovarian cancer, which exhibit low mutation load, an estimated 60% of the disease at its early stage would be missed. To put these number in perspective, there were 20,240 new cases of ovarian cancer in the U.S. in 2018. Thus, about 55 million women (over the age of 50) should be tested for the disease. Such test would identify 8,096 women with ovarian cancer. However, there would be about 1.1 million false-positives. The positive predictive value of such a test would be around 0.74%. In other words, only one in 136 women who tested positive would actually have ovarian cancer, the rest would be false-positives.

[0286] For a multi-marker test of the present application, two or more markers need to be deemed positive in order the overall screening result to be deemed positive. By increasing the total number of individual markers used, as well as the number of markers required to call the overall screening test positive, both sensitivity and specificity for detecting early cancer may be improved. The overall early cancer detection sensitivity is a function of the average number of each marker in the blood, the average number of markers positive, the minimum number of markers required to call the sample positive, and the total number of markers scored. For example, if the test uses 12 methylation markers, that on average are methylated in > 50% of tumors for that cancer type, then on average, about 6 markers will be methylated for a given sample. If on average there are 600 methylated molecules in the blood for each marker, then on average a total of 600 x 600 = 3,600 objects (*i.e.* methylated molecules) are distributed into 600 bins (*i.e.* 600 aliquots of 10 ml). As an approximate calculation based on the Poisson calculation, the distribution would be: 0.2% of wells will have 0 objects, 1.5% will have 1 object, 4.5% will have 2 objects, 8.9% will have 3 objects, 13.3% will have 4 objects, 16.0% will have 5 objects, 16.0% will have 6 objects, 13.8% will have 7 objects, 10.3% will have 8 objects etc. Suppose that at least two markers need to be called positive. In this case, 1.7% (= 0.2% + 1.5%) of the aliquots with either 0 or 1 object (*i.e.* methylation markers) would be called negative . Thus, if a minimum of two markers are required to call the sample positive, then the sensitivity of the assay would be 100% - 1.7% = 98.3% sensitivity. Suppose that at least three markers need to be called positive. In this case, aliquots with either 0, 1 or 2 objects (*i.e.* methylation markers) would be called negative = 0.2% + 1.5% + 4.5% = 6.2%. Thus, if a minimum of two markers are required to call the sample positive, then the sensitivity of the assay would be 100% - 6.2% = 93.8% sensitivity. It is understood that a small fraction of aliquots with 3 markers positive will contain 2 molecules of one marker, and one molecule of a second marker, and thus not contain

the minimum of three *different* markers positive, nevertheless, this is a small deviation from the approximate calculation above.

[0287] The overall early cancer detection specificity is a function of the average number of markers positive, the false-positive rate for each individual marker, the minimum number of markers required to call the sample positive, and the total number of markers scored. To estimate the overall false-positive rate, a formula is used based on the probability of binning different color socks into a number of drawers. Consider the percentage of false positives for each marker = "%FP"; the total number of markers "m", and the minimum number of markers required to call the sample positive "n". Then the formula for calculating overall false positive would be: $(\%FP)^n$ x [m!/(m-n)!n!]. Suppose that percentage of false positives for each marker = "%FP" is at 4%; the total number of markers "m" is 12, and the minimum number of markers required to call the sample positive "n" is 3. Then the above formula for overall false-positives would be $(4\%)^3$ x [12!/9!3!] = $(4\%)^3$ x [12 x 11 x 10 / 6] = 1.4%. Thus, the overall specificity would be [100% - 1.4%] = 98.6%. The actual individual false-positive rate may differ for different markers. Further, it may depend on the source of the false-positive signal. If for example, age-related methylation is due to clonal hematopoiesis, *i.e.* results from accumulation of methylations in white-blood cells, that then undergo clonal expansion. This type of false-positive may be mitigated by also scoring for methylation changes in white blood cells from the same patient. On the other hand, if the source of the false-positive signal is due to release of DNA into the plasma due to tissue inflammation, or for example breakdown of muscle tissue from weight-lifting, then mitigating that signal may require sampling the blood at a different time when the body is rested, or a month later when inflammation has subsided.

[0288] Figures 33, 34, 35, and 36 illustrate results for calculated overall Sensitivity and Specificity for 24, 36, 48, and 96 markers, respectively. These graphs are based on the assumption that the average individual marker sensitivity is 50%, and the average individual marker false-positive rate is from 2% to 5%. The sensitivity curves provide overall sensitivity as a function of the average number of molecules in the blood for each marker, with separate curves for each minimum number of markers needed to call a sample as positive. The specificity curves provide overall specificity as a function of individual marker false-positive rates, again with separate curves for each minimum number of markers needed to call a sample as positive. The calculated numbers for overall Sensitivity and Specificity for 12, 24, 36, 48, 72 and 96 markers, respectively are provided in the tables below.

Table 1

12 Markers Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity			
Average Number of Molecules in Blood	Mutation, 1 Positive	12 markers, Minimum 2 Positive	12 markers, Minimum 3 Positive
150	22.1%	44.2%	19.1%
200	28.1%	59.4%	32.3%
240	33.0%	69.2%	43.0%
300	39.4%	80.1%	57.7%
400	48.8%	90.8%	76.2%
480	55.1%	95.2%	85.7%
600	63.2%	98.3%	93.8%

Table 2.

12 Marker Specificity		
Individual marker FP rate	Minimum 2 Markers Positive	Minimum 3 Markers Positive
2%	97.4%	99.9%
3%	94.1%	99.5%
4%	89.4%	98.6%
5%		97.2%

Table 3.

24 Markers Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity				
Average Number of Molecules in Blood	Mutation, 1 Positive	24 markers, Minimum 3 Positive	24 markers, Minimum 4 Positive	24 markers, Minimum 5 Positive
150	22.1%	57.7%	35.3%	18.5%
200	28.1%	76.2%	56.7%	37.1%
240	33.0%	85.7%	70.6%	52.4%
300	39.4%	93.8%	84.9%	71.5%
400	48.8%	98.6%	95.8%	90.0%
480	55.1%	99.6%	98.6%	96.2%
600	63.2%	99.9%	99.8%	99.2%

Table 4.

24 Marker Specificity			
Individual marker FP rate	Minimum 3 Markers Positive	Minimum 4 Markers Positive	Minimum 5 Markers Positive
2%	98.4%	99.8%	99.9%
3%	94.6%	99.1%	99.9%
4%	87.1%	97.3%	99.6%
5%		93.4%	98.7%

Table 5.

36 Marker Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity					
Average Number of Molecules in Blood	Mutation, 1 Positive	36 markers, Minimum 3 Positive	36 markers, Minimum 4 Positive	36 markers, Minimum 5 Positive	36 markers, Minimum 6 Positive
150	22.1%	82.6%	65.8%	46.8%	29.7%
200	28.1%	93.8%	84.9%	71.5%	55.4%
240	33.0%	97.5%	92.8%	84.4%	72.4%
300	39.4%	99.4%	97.9%	94.5%	88.4%
400	48.8%	99.9%	99.8%	99.2%	98.0%
480	55.1%	100.0%	100.0%	99.9%	99.6%
600	63.2%	100.0%	100.0%	100.0%	100.0%

Table 6.

36 Marker Specificity				
Individual marker FP rate	Minimum 3 Markers Positive	Minimum 4 Markers Positive	Minimum 5 Markers Positive	Minimum 6 Markers Positive
2%	94.3%	99.1%	99.9%	100.0%
3%	80.7%	95.2%	99.1%	99.9%
4%		84.9%	96.1%	99.2%
5%			88.2%	97.0%

Table 7.

48 Marker Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity						
Average Number of Molecules in Blood	Mutation, 1 Positive	48 markers, Minimum 4 Positive	48 markers, Minimum 5 Positive	48 markers, Minimum 6 Positive	48 markers, Minimum 7 Positive	48 markers, Minimum 8 Positive
150	22.1%	84.9%	71.6%	55.6%	39.6%	25.8%
200	28.1%	95.8%	90.1%	80.9%	68.7%	54.8%
240	33.0%	99.1%	97.2%	93.4%	87.1%	78.1%
300	39.4%	99.8%	99.3%	98.1%	95.6%	92.3%
400	48.8%	99.9%	99.9%	99.8%	99.7%	99.1%
480	55.1%	99.9%	99.9%	99.9%	99.9%	99.9%
600	63.2%	99.9%	99.9%	99.9%	99.9%	99.9%

Table 8.

48 Marker Specificity					
Individual marker FP rate	Minimum 4 Markers Positive	Minimum 5 Markers Positive	Minimum 6 Markers Positive	Minimum 7 Markers Positive	Minimum 8 Markers Positive
2%	96.9%	99.4%	99.9%	99.9%	99.9%
3%	84.3%	95.8%	99.1%	99.8%	99.9%
4%		82.5%	95.0%	98.8%	99.8%
5%				94.3%	98.6%

Table 9.

72 Marker Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity								
Average Number of Molecules in Blood	Mutation, 1 Positive	72 markers, Minimum 6 Positive	72 markers, Minimum 7 Positive	72 markers, Minimum 8 Positive	72 markers, Minimum 9 Positive	72 markers, Minimum 10 Positive	72 markers, Minimum 11 Positive	72 markers, Minimum 12 Positive
150	22.1%	88.4%	79.3%	67.6%	54.4%	41.3%	29.4%	19.7%
200	28.1%	98.0%	95.4%	91.0%	84.5%	75.8%	65.3%	53.8%
240	33.0%	99.6%	98.9%	97.5%	94.9%	90.8%	84.9%	77.2%
300	39.4%	99.9%	99.9%	99.7%	99.3%	98.5%	97.0%	94.5%
400	48.8%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%	99.7%
480	55.1%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%
600	63.2%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%

Table 10.

72 Marker Specificity							
Individual marker FP rate	Minimum 6 Markers Positive	Minimum 7 Markers Positive	Minimum 8 Markers Positive	Minimum 9 Markers Positive	Minimum 10 Markers Positive	Minimum 11 Markers Positive	Minimum 12 Markers Positive
2%	99.0%	99.8%	100.0%	100.0%	100.0%	100.0%	100.0%
3%	88.6%	96.8%	99.2%	99.8%	100.0%	100.0%	100.0%
4%			92.2%	97.8%	99.4%	99.9%	100.0%
5%				83.4%	94.8%	98.5%	99.6%

Table 11.

96 Marker Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity								
Average Number of Molecules in Blood	Mutation, 1 Positive	96 markers, Minimum 7 Positive	96 markers, Minimum 8 Positive	96 markers, Minimum 9 Positive	96 markers, Minimum 10 Positive	96 markers, Minimum 11 Positive	96 markers, Minimum 12 Positive	96 markers, Minimum 13 Positive
150	22.1%	95.4%	91.0%	84.5%	75.8%	65.3%	53.8%	42.4%
200	28.1%	99.6%	99.0%	97.8%	95.7%	92.3%	87.3%	80.7%
240	33.0%	99.9%	99.9%	99.7%	99.2%	98.3%	96.8%	94.4%
300	39.4%	99.9%	99.9%	99.9%	99.9%	99.9%	99.7%	99.5%
400	48.8%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%
480	55.1%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%
600	63.2%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%	99.9%

Table 12.

96 Marker Specificity							
Individual marker FP rate	Minimum 7 Markers Positive	Minimum 8 Markers Positive	Minimum 9 Markers Positive	Minimum 10 Markers Positive	Minimum 11 Markers Positive	Minimum 12 Markers Positive	Minimum 13 Markers Positive
2%	98.5%	99.7%	99.9%	100.0%	100.0%	100.0%	100.0%
3%		91.3%	97.4%	99.3%	99.8%	100.0%	100.0%
4%				88.2%	96.3%	99.0%	99.7%
5%						84.7%	95.1%

[0289] From the above tables, the receiver operating characteristic (ROC) curves may be calculated by plotting Sensitivity vs. 1-Specificity. Since these are theoretical calculations, the curves were generated for different levels of average marker false-positive rates of 2%, 3%, 4%, and 5%. To assist in visualizing the graphs and calculating the AUC (Area under curve), the edges were set at 100% and 0%, respectively. The ROC curves for 24 marker, 3% & 4% average marker false-positives, 36 marker, 3% & 4% average marker false-positives, and 48 marker, 2%, 3%, 4% & 5% average marker false-positives are provided in Table 13 below and

for 48 Markers illustrated in Figures 37 and 38, respectively. Generally, the closer the AUC is to 1, the more accurate the test – values of >95% are desirable, and values >99% are superb. Using the benchmark of an average of 300 molecules in the blood for early cancer (Stage I & II), AUC values are at 95% with 24 markers, improve to 99% with 36 markers, and range from 98% to >99% with 48 markers, depending on average marker false-positive values. These graphs provide an illustration of the power of multiple marker assays for achieving good sensitivities and specificities.

Table 13.

24, 36, & 48 Marker AUC Values from ROC Curves; Avg. Indiv. Mkr,: 50% Sensitivity							
Total Markers: Individual marker FP rate	150 Molecules	200 Molecules	240 Molecules	300 Molecules	400 Molecules	480 Molecules	600 Molecules
24 Mkrs: 3%	77%	87%		96%	99%	>99%	>99%
24 Mkrs: 4%	74%	85%		95%	99%	>99%	>99%
36 Mkrs: 3%	87%	95%	98%	99%	>99%	>99%	>99%
36 Mkrs: 4%	78%	89%	95%	98%	99%	>99%	>99%
48 Mkrs: 2%	92%	98%	99%	>99%	>99%	>99%	>99%
48 Mkrs: 3%	89%	97%	99%	>99%	>99%	>99%	>99%
48 Mkrs: 4%	81%	93%	98%	99%	>99%	>99%	>99%
48 Mkrs: 5%	71%	86%	94%	98%	99%	99%	99%

[0290] How would the above markers work in a one-step cancer assay? To illustrate the challenges of developing an early cancer detection screen, consider the challenge of screening 107 million adults in the U.S. over the age of 50 for colorectal cancer – of which there are about 135,000 new cases that are diagnosed a year. In this example, if there is an average of 300 molecules in the blood for early cancer (Stage I & II), and taking the best-case scenario of individual marker FP rate is 2%, then if there is a 3-marker minimum, then overall FP rate is 1.6% for 24 markers, for a specificity of 98.4% (See Figure 33B). At 3 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), the test would miss 6.2%; *i.e.* for Stage I & II cancer the overall sensitivity would be 93.8% (See Figure 33A), *e.g.* the test would correctly identify 93.8% of individuals with disease, which would be 126,630 individuals (out of 135,000 new cases). At a specificity of 98.4%, for 107 million individuals screened, the test would also generate 1.6% x 107,000,000 = 1,712,000 false positives. Thus, the positive predictive value would be 126,630/(126,630 + 1,712,000) = around 6.8%, in other words, only

one in 14 individuals who tested positive would actually have colorectal cancer, the rest would be false-positives.

[0291] However, if the individual marker FP rate is more realistic, say 4%, then more markers will be required to achieve better than 98% specificity, and this will be at the cost of sensitivity. If individual marker FP rate is 4%, then if there is a 5-marker minimum, then overall FP rate is 0.4% for 24 markers, for a specificity of 99.6% (See Figure 33B). At 5 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), the test would miss 28.5%; *i.e.* for Stage I & II cancer the overall sensitivity would be 71.5% (See Figure 33A), *e.g.* the test would correctly identify 71.5% of individuals with disease, which would be 90,540 individuals (out of 135,000 new cases). At a specificity of 99.6%, for 107 million individuals screened, the test would also generate 0.4% x 107,000,000 = 428,000 false positives. Thus, the positive predictive value would be 90,540/(90,540 + 428,000) = around 17.5%, in other words, only one in 5.7 individuals who tested positive would actually have colorectal cancer, the rest would be false-positives. A PPV of 17.5% is quite respectable, however, it would be achieved at the cost of missing 28.5% of early cancer.

[0292] Another aspect of the present application relates to a two-step method of diagnosing or prognosing a disease state of cells or tissue based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual. The method involves obtaining a biological sample, the biological sample including exosomes, tumor-associated vesicles, markers within other protected states, cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof. A first step is applied to the biological samples with an overall sensitivity of > 80% and an overall specificity of > 85% or an overall Z-score of > 1.03 to identify individuals more likely to be diagnosed or prognosed with the disease state. A second step is the applied to biological samples from those individuals identified in the first step with an overall specificity of > 95% or an overall Z-score of > 1.65 to diagnose or prognose individuals with the disease state. The first step and the second step are carried out using a method of the present application. The first step uses markers to cover many cancers, where the aim is to obtain high sensitivity for early cancers where the number of marker molecules in the blood may be limited. The second step then would score for additional markers to verify that the initial result was a true positive, as well as to identify the likely tissue of origin. The second step may include the methods described herein, and/or additional methods such as next-generation sequencing. The first step uses markers to cover many cancers, where the aim is

to obtain high sensitivity for early cancers where the number of marker molecules in the blood may be limited. The second step then would score for additional markers to verify that the initial result was a true positive, as well as to identify the likely tissue of origin. The second step may include the methods described herein, and/or additional methods such as next-generation sequencing.

[0293] To illustrate one embodiment of how such a two-step cancer test may be designed, consider again the challenge of identifying patients with early colorectal cancer. In 2017, there were an estimated 95,520 new cases of colon cancer and 39,910 cases of rectal cancer diagnosed in the U.S. – for a total of about 135,000 new cases. Consider an initial test using 24 markers. In this example, if there is an average of 300 molecules in the blood for early cancer (Stage I & II), and if that would cover at least one mutation, then the sensitivity for identifying such a cancer by next generation sequencing would be 39.4% (See Figure 33A). If the individual marker FP rate is 3%, then if there is a 3-marker minimum, then overall FP rate is 5.4% for 24 markers, for a specificity of 94.6% (See Figure 33B). At 3 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), the test would miss 6.2%; *i.e.* for Stage I & II cancer the overall sensitivity would be 93.8% (See Figure 33A). Note that these levels of sensitivity and specificity are better than the current tests on the market. However, if the individual marker FP rate is 5%, then if there is a 4-marker minimum, then overall FP rate is 6.6% for 24 markers, for a specificity of 93.4% (See Figure 33B). At 4 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), the test would miss 15.1%; *i.e.* for Stage I & II cancer the sensitivity would be 84.9% (See Figure 33A). These graphs illustrate a basic conflict of most diagnostic tests – improve the sensitivity of a test (*i.e.* less false-negatives), but sacrifice the test specificity (*i.e.* more false-positives), or improve the specificity of a test (less false-positives) at the risk of losing the test sensitivity (*i.e.* more false-negatives).

[0294] By using a two-step cancer test, the parameters may be adjusted to improve BOTH sensitivity and specificity. For example, the aforementioned 24 marker test, using 3 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), the overall sensitivity would be 93.8%. Those samples that are scored as positives in the first step (24-markers specific to GI cancers) – including the false-positives would be retested in the second step with an expanded panel of 48 markers to provide coverage of colorectal cancers. If the individual marker FP rate is 3%, then if there is a 5-marker minimum, then overall FP rate is 4.2% for 48 markers, for a specificity of 95.8% (See Figure 35B). At 5 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), the test would miss 0.7%; *i.e.* for Stage I & 2 cancer the sensitivity would be 99.3% (See Figure 35A). Technically, since

the samples were already culled in the first step, the overall sensitivity is 93.8% x 99.3% = 93.1%. If the individual marker FP rate is 3%, then if there is a 6-marker minimum, then overall FP rate is <1% for 48 markers, for a specificity of 99.1% (See Figure 35B). At 6 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), the test would miss 1.9%; *i.e.* for Stage I & II cancer the sensitivity would be 98.1% (See Figure 35A). Since the samples were already culled in the first step, the overall sensitivity is 93.8% x 98.1% = 92.0%. If the individual marker FP rate is 3%, then if there is a 7-marker minimum, then overall FP rate is <0.2% for 48 markers, for a specificity of 99.8% (See Figure 35B). At 7 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), the test would miss 4.4%; *i.e.* for Stage I & II cancer the sensitivity would be 95.6% (See Figure 35A). Since the samples were already culled in the first step, the overall sensitivity is 93.8% x 95.6% = 89.7%.

[0295] Returning to the example of colorectal cancer, in particular the cases of microsatellite stable tumors (MSS) where the mutation load is low, for these calculations when relying on NGS sequencing alone (assuming 300 molecules with one mutation in the blood), an estimated 60% of early colorectal cancer would be missed. Again, to put these number in perspective, in the U.S., about 135,000 new cases of colorectal cancer are predicted in 2018. About 107 million individuals in the U.S. are over the age of 50 and should be tested for colorectal cancer. With the assumption of these samples containing at least 300 molecules with one mutation in the blood, such a test would find 54,000 men and women (out of 135,000 new cases) with colorectal cancer. However, with a specificity for sequencing at 98%, there would be about 2.1 million false-positives. The positive predictive value of such a test would be around 2.6%, in other words, only one in 39 individuals who tested positive would actually have colorectal cancer, the rest would be false-positives. In contrast, consider the two-step methylation marker test described above, wherein the first step has 24 methylation markers specific to GI cancers, while the second step has 48 methylation markers specific to colorectal cancer. In this example, as above, the calculations are done with the anticipation of an average of 300 methylated molecules per positive marker in the blood. Assuming individual marker false-positive rates of 3%, and with the first step requiring a minimum of 3 markers positive, then with an overall specificity of 94.6%, the first step would identify 5,778,000 individuals (out of 107,000,000 total adults over 50 in the U.S.) which would include at 93.8% sensitivity about 126,630 individuals with Stage I & II colorectal cancer (out of 135,000 total). However, those 5,778,000 presumptive positive individuals would be evaluated in the second step of 48 markers requiring a minimum of 6 markers positive, then the two-step test would identify 98.1% x 93.8% = 92.0% = 124,200 individuals (out of 135,000 new cases) with colorectal cancer. With a

specificity of 99.1%, the second test would also generate 5,778,000 x 0.9% = 52,000 false-positives. The positive predictive value of such a test would be 124,200/176,200 = 70.5%, in other words, 2 in 3 individuals who tested positive would actually have colorectal cancer, an extraordinarily successful screen to focus on those patients who would most benefit from follow-up colonoscopy. The benefit in lives saved would be of incalculable value.

[0296] While the foregoing discussion has focused on methylation markers, with an average sensitivity of 50%, and individual marker false-positives ranging from 2%-5%, there are many other markers of cancer with varying sensitivities and specificities. In general, protein markers (with the exception of PSA and PSMA) have been of limited clinical utility for detection of early cancer because the false-positives are so high, resulting in very low positive predictive value. Cancer markers from bodily fluids (*i.e.* plasma, urine) include, but are not limited to plasma microRNAs (miRNA); mutations or methylation in cfDNA; exosomes with surface cancer-specific protein markers, or internal miRNA, ncRNA, lncRNA, mRNA, DNA; circulating cytokines, circulating proteins, or circulating antibodies against cancer-antigens; or nucleic-acid markers in whole blood (for review, see Nikolaou et al., "Systematic Review of Blood Diagnostic Markers in Colorectal Cancer," *Techniques in Coloproctology* (2018), which is hereby incorporated by reference in its entirety). Several methods have been reported for detecting cancer-specific miRNAs in the serum or plasma of patients with colorectal (or others) cancers; these miRNAs include, but are not limited to: miR-1290; miR-21; miR-24; miR-320a; miR-423-5p; miR-29a; miR-125b; miR-17-3p; miR-92a; miR-19a; miR-19b; miR-15b; mir23a; miR-150; miR-223; miR-1229; miR-1246; miR-612; miR-1296; miR-933; miR-937; miR-1207; miR-31; miR-141; miR-224-3p; miR-576-5p; miR-885-5p; miR-200c; miR-203 (Imaoka et al., "Circulating MicroRNA-1290 as a Novel Diagnostic and Prognostic Biomarker in Human Colorectal Cancer," *Ann. Oncol.* 27(10):1879–1886 (2016); Zhang et al., "Diagnostic and Prognostic Value of MicroRNA-21 in Colorectal Cancer: an Original Study and Individual Participant Data Meta-Analysis," *Cancer Epidemiol. Biomark. Prev.* 23(12):2783–2792 (2016); Fang et al., "Plasma Levels of MicroRNA-24, MicroRNA-320a, and Micro-RNA-423-5p are Potential Biomarkers for Colorectal Carcinoma," *J. Exp. Clin. Cancer Res.*34:86 (2015); Toiyama et al., "MicroRNAs as Potential Liquid Biopsy Biomarkers in Colorectal Cancer: A Systematic Review," *Biochim. Biophys. Acta.* pii: S0304-419X(18)30067-2 (2018); Nagy et al., "Comparison of Circulating miRNAs Expression Alterations in Matched Tissue and Plasma Samples During Colorectal Cancer Progression," *Pathol. Oncol. Res.* doi: 10.1007/s12253-017-0308-1 (2017); Wang et al., "Novel Circulating MicroRNAs Expression Profile in Colon Cancer: a Pilot Study," *Eur. J. Med. Res.* 22(1):51 (2017); United States Patent No. 9,689,036 to Getts, et al.; United States Patent No. 9,708,643 to Duttagupta, et al.; United States Patent No. 9,868,992

to Goel, et al; United States Patent No. 9,926,603 to Sozzi et al., which are hereby incorporated by reference in their entirety). Additional approaches for detecting low abundance miRNA are described in WO2016057832A2, which is hereby incorporated by reference in its entirety, or using other suitable means known in the art. Figure 39 provides a list of blood-based, colon cancer-specific microRNA markers derived through analysis of TCGA microRNA datasets, which may be present in exosomes, tumor-associated vesicles, Argonaute complexes, or other protected states in the blood.

[0297] Several methods have been reported for detecting cancer-specific ncRNA or lncRNAs in the serum, plasma, or exosomes of patients with colorectal (and other) cancers; these ncRNAs include but are not limited to: NEAT_v1; NEAT_v2; CCAT1; HOTAIR; CRNDE-h; H19; MALAT1; 91H; GAS5 (Wu et al., "Nuclear-enriched Abundant Transcript 1 as a Diagnostic and Prognostic Biomarker in Colorectal Cancer," *Mol. Cancer* 14:191 (2015); Zhao et al., "Combined Identification of Long Non-Coding RNA CCAT1 and HOTAIR in Serum as an Effective Screening for Colorectal Carcinoma," *Int. J. Clin. Exp. Pathol.* 8(11):14131-40 (2015); Liu et al., "Exosomal Long Noncoding RNA CRNDE-h as a Novel Serum-Based Biomarker for Diagnosis and Prognosis of Colorectal Cancer," *Oncotarget* 7(51):85551-85563 (2016); Slaby O, "Non-coding RNAs as Biomarkers for Colorectal Cancer Screening and Early Detection," *Adv Exp Med Biol.* 937:153-70 (2016); Gao et al., "Exosomal lncRNA 91H is Associated With Poor Development in Colorectal Cancer by Modifying HNRNPK Expression," *Cancer Cell Int.* 23;18:11 (2018); Liu et al., "Prognostic and Predictive Value of Long Non-Coding RNA GAS5 and MicroRNA-221 in Colorectal Cancer and Their Effects on Colorectal Cancer Cell Proliferation, Migration and Invasion," *Cancer Biomark.* 22(2):283-299 (2018); United States Patent No. 9,410,206 to Hoon, et al.; United States Patent No. 9,921,223 to Kalluri et al., which are hereby incorporated by reference in their entirety). Additional approaches for detecting low abundance lncRNA, ncRNA, mRNA translocations, splice variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicing, exon insertions, exon deletions, and intron insertions are described in WO2016057832A2, which is hereby incorporated by reference in its entirety, or using other suitable means known in the art. Figure 40 provides a list of blood-based, colon cancer-specific ncRNA and lncRNA markers derived through analysis of various publicly available Affymetrix Exon ST CEL data, which were aligned to GENCODE annotations to generate ncRNA and lncRNA transriptome datasets. Comparative analyses across these datasets (various cancer types, along with normal tissues, and peripheral blood) were conducted to generate the ncRNA and lncRNA markers list (Figure 40). Such lncRNA and ncRNA may be enriched in exosomes or other protected states in the blood. In addition, Figure 41 provides a list of blood-based colon

cancer-specific exon transcripts that may be enriched in exosomes, tumor-associated vesicles, or other protected states in the blood.

[0298] The most common protein marker for colorectal cancer is based on detecting hemoglobin from blood in the stool and is known as the FOBT or FIT test. Sensitivity and specificities (Sens.: Spec.) for these tests have been reported as: OC-Light iFOB Test (also called OC Light S FIT), manufactured by Polymedco (78.6%-97.0% : 88.0%-92.8%); QuickVue iFOB, manufactured by Quidel (91.9% : 74.9%); Hemosure One-Step iFOB Test, manufactured by Hemosure, Inc. (54.5% : 90.5%); InSure FIT, manufactured by ClinicalGenomics (75.0% : 96.6%); Hemoccult-ICT, manufactured by Beckman Coulter (23.2%-81.8% : 95.8%-96.9%); Cologuard – stool FIT-DNA, manufactured by Exact Sciences (92.3%; 84.4%). The large ranges and differences in sensitivities and specificities may reflect the range from early to late cancer, as well as differences in methodology, number of samples collected, and clinical study size. Cut-off values for FIT tests may range from 10 ug protein/gram stool to 300 ug protein/gram stool (*See* Robertson et al., "Recommendations on Fecal Immunochemical Testing to Screen for Colorectal Neoplasia: a Consensus Statement by the US Multi-Society Task Force on Colorectal Cancer," *Gastrointest. Endosc.* 85(1):2-21 (2017), which is hereby incorporated by reference in its entirety).

[0299] A number of tumor-associated antigens elicit an immune response within the patient, and these may be identified as autoantibodies, or indirectly as increased cytokines in the serum. Some tumor antigens may be detected directly within the serum, or on the surface of cancer-associated exosomes or extracellular vesicles, while others may be detected indirectly, for example by an increase in mRNA within cancer-associated exosomes or extracellular vesicles. These cancer-specific proteins markers may be identified through, mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product, and these markers include but are not limited to: RPH3AL; RPL36; SLP2; TP53; Survivin; ANAXA4; SEC61B; CCCAP; NYCO16; NMDAR; PLSCR1; HDAC5; MDM2; STOML2; SEC61-beta; IL8; TFF3; CA11-19; IGFBP2; DKK3; PKM2; DC-SIGN; DC-SIGNR; GDF-15; AREG; FasL; Flt3L; IMPDH2; MAGEA4; BAG4; IL6ST; VWF; EGFR; CD44; CEA; NSE; CA 19-9, CA 125; NMMT; PSA; proGRP; DPPIV/seprase complex; TFAP2A; E2F5; CLIC4; CLIC1; TPM1; TPM2; TPM3; TPM4; CTSD-30; PRDX-6; LRG1; TTR; CLU; KLKB1; C1R; KLK3; KLK2; HOXB13; GHRL2; FOXA1 (Fan et al., "Development of a Multiplexed Tumor-Associated Autoantibody-Based Blood Test for the Detection of Colorectal Cancer," *Clin. Chim. Acta.* 475:157-163 (2017); Xia et al., "Prognostic Value, Clinicopathologic Features and Diagnostic Accuracy of Interleukin-8 in Colorectal Cancer: a Meta-Analysis," *PLoS One* 10(4):e0123484 (2015); Li et al., "Serum Trefoil Factor 3 as a Protein Biomarker for the

Diagnosis of Colorectal Cancer," *Technol. Cancer. Res. Treat.* 16(4):440-445 (2017); Overholt et al., "CA11-19: a Tumor Marker for the Detection of Colorectal Cancer," *Gastrointest. Endosc.* 83(3):545–551 (2016); Fung et al., "Blood-based Protein Biomarker Panel for the Detection of Colorectal Cancer," *PLoS One* 10(3): e0120425 (2015); Jiang et al., "The Clinical Significance of DC-SIGN and DC-SIGNR, Which are Novel Markers Expressed in Human Colon Cancer," *PLoS One* 9(12):e11474 (2014).; Chen et al., "Development and Validation of a Panel of Five Proteins as Blood Biomarkers for Early Detection of Colorectal Cancer," *Clin. Epidemiol.* 9:517-526 (2017); Chen et al., "Prospective Evaluation of 64 Serum Autoantibodies as Biomarkers for Early Detection of Colorectal Cancer in a True Screening Setting," *Oncotarget* 7(13):16420-32 (2016); Rho et al., "Protein and Glycomic Plasma Markers for Early Detection of Adenoma and Colon Cancer," *Gut* 67(3):473-484 (2018); United States Patent No. 9,518,990 to Wild et al.; United States Patent No. 9,835,636 to Chan et al.; United States Patent No. 9,885,718 to Man et al.; United States Patent No. 9,889,135 to Andy Koff et al.; United States Patent No. 9,903,870 to Speicher et al.; United States Patent No. 9,914,974 to Bajic et al.; United States Patent No. 9,983,208 to Choi et al; United States Patent No. 10,030,271 to Scher et al., which are hereby incorporated by reference in their entirety). Additional approaches for detecting low abundance mRNA translocations, splice variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicing, exon insertions, exon deletions, and intron insertions are described in WO2016057832A2, which is hereby incorporated by reference in its entirety, or using other suitable means known in the art, Figure 42 provides a list of cancer protein markers, identified through mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from Colorectal tumors, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma. Figure 43 provides protein markers that can be secreted by Colorectal tumors into the blood. A comparative analysis was performed across various TCGA datasets (tumors, normals), followed by an additional bioinformatics filter (Meinken et al., "Computational Prediction of Protein Subcellular Locations in Eukaryotes: an Experience Report," *Computational Molecular Biology* 2(1):1-7 (2012), which is hereby incorporated by reference in its entirety), which predicts the likelihood that the translated protein is secreted by the cells.

[0300] The distribution of mutations in colorectal cancers are available in the public COSMIC database, with the 20 most commonly altered genes listed as: APC; TP53; KRAS; FAT4; LRP1B; PIK3CA; TGFBR2; ACVR2A; BRAF; ZFHX3; KMT2C; KMT2D; FBXW7; SMAD4; ARID1A; TRRAP; RNF43: FAT1; TCF7L2; PREX2 (Forbes et al., "COSMIC: Exploring the World's Knowledge of Somatic Mutations in Human Cancer," *Nucleic Acids Res.*

43(Database issue):D805-811 (2015), which is hereby incorporated by reference in its entirety). Analysis of TCGA COADREAD mutational dataset, however indicate the following genes have at least 10% mutation rate among colorectal cancer primary tumors: APC, TP53, KRAS, TTN, SYNE1, PIK3CA, FAT4, MUC16, FBXW7, LRP1B, LRP2, DNAH5, DMD, ANK2, RYR2, FLG, HMCN1, FAT2, TCF7L2, CSMD3, USH2A, SDK1, CSMD1, COL6A3, DNAH2, SMAD4, PKHD1, FAM123B, ATM, ACVR2A, MDN1, DCHS2, ZFHX4, CUBN, CSMD2, FREM2, RYR1, TGFBR2, RYR3, SACS, DNAH10, ABCA12, BRAF, ODZ1, PCDH9, MACF1, AHNAK2. In addition to the approaches described herein, there are several approaches for enriching for and detecting low-abundance mutations either at the DNA or mRNA level (for example, mRNA within exosomes), including but not limited to next generation sequencing, allele-specific PCR, ARMS, primer-extension PCR, using blocking primers, full-COLD-PCR, fast-COLD-PCR, ice-COLD-PCR, E-ice-COLD-PCR, TT-COLD-PCR, etc. (Mauger et al., "COLD-PCR Technologies in the Area of Personalized Medicine: Methodology and Applications," *Mol. Diagn Ther.* (3):269-283 (2017); Sefrioui et al., "Comparison of the Quantification of KRAS Mutations by Digital PCR and E-ice-COLD-PCR in Circulating-Cell-Free DNA From Metastatic Colorectal Cancer Patients," *Clin. Chim. Acta.* 465:1-4 (2017); United States Patent No. 9,062,350 to Platica; United States Patent No. 9,598,735 to Song et al., which are hereby incorporated by reference in their entirety). Additional approaches for detecting low abundance mutations are described in WO2016057832A2, which is hereby incorporated by reference in its entirety, or using other suitable means known in the art.

[0301] The best studied blood-based methylation markers for CRC detection are located in the promoter region of the SEPT9 gene (Church et al., "Prospective Evaluation of Methylated SEPT9 in Plasma for Detection of Asymptomatic Colorectal Cancer," *Gut* 63(2):317-325 (2014); Lofton-Day et al., "DNA Methylation Biomarkers for Blood-Based Colorectal Cancer Screening," *Clinical Chemistry* 54(2):414-423 (2008); Potter et al., "Validation of a Real-time PCR-based Qualitative Assay for the Detection of Methylated SEPT9 DNA in Human Plasma," *Clinical Chemistry* 60(9):1183-1191 (2014); Ravegnini et al., "Simultaneous Analysis of SEPT9 Promoter Methylation Status, Micronuclei Frequency, and Folate-Related Gene Polymorphisms: The Potential for a Novel Blood-Based Colorectal Cancer Biomarker," *International Journal of Molecular Sciences* 16(12):28486-28497 (2015); Toth et al., "Detection of Methylated SEPT9 in Plasma is a Reliable Screening Method for Both Left- and Right-sided Colon Cancers," *PloS One* 7(9):e46000 (2012); Toth et al., "Detection of Methylated Septin 9 in Tissue and Plasma of Colorectal Patients With Neoplasia and the Relationship to the Amount of Circulating Cell-free DNA," *PloS One* 9(12):e115415 (2014); Warren et al., "Septin 9 Methylated DNA is a Sensitive and Specific Blood Test for Colorectal Cancer," *BMC Medicine* 9:133 (2011), which are hereby

incorporated by reference in their entirety), and other potential markers for CRC diagnostics include CpG sites on promoter regions of THBD, C9orf50, ZNF154, AGBL4, FLI1, and TWIST1 (Lange et al., "Genome-scale Discovery of DNA-methylation Biomarkers for Blood-based Detection of Colorectal Cancer," *PloS One* 7(11):e50266 (2012); Margolin et al., "Robust Detection of DNA Hypermethylation of ZNF154 as a Pan-Cancer Locus with in Silico Modeling for Blood-Based Diagnostic Development," *The Journal of Molecular Diagnostics: JMD* 18(2):283-298 (2016); Lin et al., "Clinical Relevance of Plasma DNA Methylation in Colorectal Cancer Patients Identified by Using a Genome-Wide High-Resolution Array," *Ann. Surg. Oncol.* 22 Suppl 3:S1419-1427 (2015), which are hereby incorporated by reference in their entirety).

[0302] *SEPT9* methylation is the basis for Epi proColon test, a CRC-detection assay by Epigenomics (Lofton-Day et al:, "DNA Methylation Biomarkers for Blood-based Colorectal Cancer Screening," *Clinical Chemistry* 54(2):414-423 (2008), which is hereby incorporated by reference in its entirety). While initial results on smaller sample sets showed promise, large-scale studies with 1,544 plasma samples showed a sensitivity of 64% for stage I-III CRC, and a specificity of 78%-82%, effectively sending 180 to 220 out of 1,000 individuals to unnecessary colonoscopies (Potter et al., "Validation of a Real-time PCR-based Qualitative Assay for the Detection of Methylated SEPT9 DNA in Human Plasma," *Clinical Chemistry* 60(9):1183-1191 (2014), which is hereby incorporated by reference in its entirety). ClinicalGenomics is currently developing blood-based CRC detection test based on the methylation of the *BCAT1* and *IKZF1* genes (Pedersen et al., "Evaluation of an Assay for Methylated BCAT1 and IKZF1 in Plasma for Detection of Colorectal Neoplasia," *BMC Cancer* 15:654 (2015), which is hereby incorporated by reference in its entirety). Large-scale studies using 2,105 plasma samples of this two-marker test showed an overall sensitivity of 66%, with 38% for stage I CRC, and an impressive specificity of 94%. Exact Sciences and collaborators have slightly improved the sensitivity of CRC fecal tests (Bosch et al., "Analytical Sensitivity and Stability of DNA Methylation Testing in Stool Samples for Colorectal Cancer Detection," *Cell Oncol. (Dordr)* 35(4):309-315 (2012); Hong et al., "DNA Methylation Biomarkers of Stool and Blood for Early Detection of Colon Cancer," *Genet. Test. Mol. Biomarkers* 17(5):401-406 (2013); Imperiale et al., "Multitarget Stool DNA Testing for Colorectal-Cancer Screening," *N. Engl. J. Med.* 370(14):1287-1297 (2014); Xiao et al., "Validation of Methylation-Sensitive High-resolution Melting (MS-HRM) for the Detection of Stool DNA Methylation in Colorectal Neoplasms," *Clin. Chim. Acta.* 431:154-163 (2014); Yang et al., "Diagnostic Value of Stool DNA Testing for Multiple Markers of Colorectal Cancer and Advanced Adenoma: a Meta-Analysis," *Can. J. Gastroenterol.* 27(8):467-475 (2013), which are hereby incorporated by reference in their entirety), by adding K-ras mutation as well as BMP3 and NDRG4 methylation markers (Lidgard

et al., "Clinical Performance of an Automated Stool DNA Assay for Detection of Colorectal Neoplasia," *Clin. Gastroenterol. Hepatol.* 11(10):1313-1318 (2013), which is hereby incorporated by reference in its entirety). Epigenetic changes may mark not only the DNA (as methylation or hydroxy-methylation of promoter CpG sites) but also by appending methyl or acetyl groups on the histone proteins that bind to these promoters. These different epigenetic marks may be detected in circulating nucleosomes of colorectal cancer patients (Rahier et al., "Circulating Nucleosomes as New Blood-based Biomarkers for Detection of Colorectal Cancer," *Clin Epigenetics* 9:53 (2017), which is hereby incorporated by reference in its entirety). The identification of blood-based, cancer-specific methylation markers has employed the entire TCGA Illumina 450K methylation datasets (consisting of primary tumors, matching normal for 33 types of cancer including CRC), along with additional methylation datasets (primary tumors, normal tissues, cell lines, peripheral blood, immune cells) from the Gene Expression Omnibus (GEO). In order to identify CRC-specific methylation markers, comparative statistical analyses of these datasets were used to identify candidate methylation markers with the following characteristics: highly methylated in CRC tissues and cell lines, unmethylated in normal colon, unmethylated in peripheral blood and immune infiltrates, unmethylated in most other cancer types. Validating the bioinformatic scheme, these methodologies also identified CpG sites previously reported to be hypermethylated in plasma from CRC patients (Church et al., "Prospective Evaluation of Methylated SEPT9 in Plasma for Detection of Asymptomatic Colorectal Cancer," *Gut* 63(2):317-325 (2014); Lofton-Day et al., "DNA Methylation Biomarkers for Blood-based Colorectal Cancer Screening," *Clinical Chemistry* 54(2):414-423 (2008); Toth et al., "Detection of Methylated SEPT9 in Plasma is a Reliable Screening Method for Both Left- and Right-sided Colon Cancers," *PloS One* 7(9):e46000 (2012); Warren et al., "Septin 9 Methylated DNA is a Sensitive and Specific Blood Test for Colorectal Cancer," *BMC Medicine* 9:133 (2011); Lange et al., "Genome-scale Discovery of DNA-methylation Biomarkers for Blood-based Detection of Colorectal Cancer," *PloS One* 7(11):e50266 (2012); Margolin et al., "Robust Detection of DNA Hypermethylation of ZNF154 as a Pan-Cancer Locus with in Silico Modeling for Blood-Based Diagnostic Development," *The Journal of Molecular Diagnostics : JMD* 18(2):283-298 (2016); Lin et al., "Clinical Relevance of Plasma DNA Methylation in Colorectal Cancer Patients Identified by Using a Genome-Wide High-Resolution Array," *Ann. Surg. Oncol.* 22 Suppl 3:S1419-1427 (2015); Pedersen et al., "Evaluation of an Assay for Methylated BCAT1 and IKZF1 in Plasma for Detection of Colorectal Neoplasia," *BMC Cancer* 15:654 (2015), which are hereby incorporated by reference in their entirety). To ensure that these methylation sites were specific to CRC and not a result of aging-related methylation (McClay et al., "A Methylome-wide Study of Aging Using Massively Parallel

Sequencing of the Methyl-CpG-enriched Genomic Fraction From Blood in Over 700 subjects," *Hum. Mol. Genet.* 23(5):1175-1185 (2014), which is hereby incorporated by reference in its entirety), the Pearson correlation was calculated between levels of methylation and patient age. Furthermore, hypermethylation of these sites did not significantly correlate with MSI status, implying that markers have been identified for all CRC subtypes. Overall, ~ 10,000 tissue samples, > 4 billion datapoints (datapoint = CpG percentage methylation per sample) were analyzed to identify an initial list of few hundred CRC-specific markers. CpG markers consistently show up in many types of cancer and are labeled as potential Pan-Oncology markers. Additional approaches for detecting low abundance 5mC (or 5hmC) are described in WO2016057832A2, which is hereby incorporated by reference in its entirety, or using other suitable means known in the art. Figure 44 provides a list of primary CpG sites that are Colorectal Cancer and Colon-tissue specific markers, that may be used to identify the presence of colorectal cancer from cfDNA, or DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. Figure 45 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are Colorectal Cancer and Colon-tissue specific markers, that may be used to identify the presence of colorectal cancer from cfDNA, or DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood.

[0303] Mutation or methylation status may give a clear analytical cut-off, *i.e.* the assay either records a mutation or CpG methylation event, and false-positives are a consequence of biology, for example from age-related methylation. With other markers there may be a greater overlap between marker level for individuals with cancer and their matched normal controls, especially in attempting to identify cancer at the earliest stages. In such cases, cut-offs may be defined by "Z-score", 2 standard deviations above normal values, or by setting the false-positive rate at an arbitrary level, *i.e.* 5% when evaluating a suitable set of age-matched normal samples. Generally, the set of age-matched normal should be suitably large enough to set cut-off of the marker-specific signal from a given disease sample at > 85%; > 90%; > 95%; > 96%; > 97%; or > 98% of the same marker-specific signals from the set of normal samples. The "Z-score" may be calculated using the formula: $Z = (X - \mu)/\sigma$; where Z = Z-score, X = each value in the dataset, μ = mean of all values in the dataset, and σ = standard deviation of a sample. Likewise, when using the Z-score, the cut-off for marker-specific signal from a given disease sample may be set at a z-score of > 1.03; > 1.28; > 1.65; > 1.75; > 1.88; or > 2.05 compared to the same marker-specific signals from the set of normal samples In some assays, marker levels, (*i.e.* DNA methylation levels for several gene promoter regions in plasma, or miRNA levels in urine) are quantified in relation to another marker, either internal or externally added in a qPCR reaction, where the cut-off is determined as a ΔCt value in the assay (Fackler et al., "Novel Methylated

Biomarkers and a Robust Assay to Detect Circulating Tumor DNA in Metastatic Breast Cancer," *Cancer Res.* 74(8):2160-70 (2014); United States Patent No. 9,416,404 to Sukumar et al., which are hereby incorporated by reference in their entirety). Methylation status at defined promoter regions may also be determined using digital bisulfite genomic sequencing and digital MethyLight assays; using bisulfite conversion and preferential amplification of converted methylated sequences by blocking primers that interfere with amplification of converted unmethylated sequences; or depletion of unmethylated DNA using methyl-sensitive restriction endonucleases, followed by PCR (see U.S. Patent No. 9,290,803 to Laird et al.; U.S. Patent No. 9,476,100 to Frumkin, et al.; U.S. Patent No. 9,765,397 to McEvoy et al.; U.S. Patent No. 9,896,732 to Tabori et al.; U.S. Patent No. 9,957,575 to Kottwitz et al., which are hereby incorporated by reference in their entirety).

[0304] The genome-wide methylation profile of cfDNA (known as the methylome) can be determined using next-generation sequencing, and the methylation pattern may be used to identify the presence of fetal, tumor, or other tissue DNA in the plasma (Sun et al., "Plasma DNA Tissue Mapping by Genome-wide Methylation Sequencing for Noninvasive Prenatal, Cancer, and Transplantation Assessments," *Proc. Natl. Acad. Sci. USA* 112(40):E5503-12 (2015); Lehmann-Werman et al., "Identification of Tissue-specific Cell Death Using Methylation Patterns of Circulating DNA," *Proc. Natl. Acad. Sci. USA* 113(13):E1826-34 (2016); U.S. Patent No. 9,732,390 to Lo et al.; U.S. Patent No. 9,984,201 to Zhang et al., which are hereby incorporated by reference in their entirety).

[0305] The aforementioned two-step screening assay sensitivities and specificities were calculated based on having an initial screen (with fewer markers) that cast a wide net to maximize sensitivity, followed by a second test (with more markers) on the initial presumptive positive samples, but the second test not only maintains the sensitivity, but also achieves high specificity to obtain a respectable positive predictive value. While colorectal cancer is a more frequent cancer, other cancers are less common, so achieving a good positive predictive value is critical to avoiding unnecessary follow-up diagnostic procedures. These initial calculations (in Figures 33-37) focused on methylation markers, with an average sensitivity of 50%, and individual marker false-positives ranging from 2%-5%, and average number of molecules in the blood set at 300 molecules. As a benchmark for these initial calculations, a mutation marker would give an overall sensitivity of 40%. In order to explore the influence of combining the methylation markers with other markers that may differ in both these values, additional calculations were performed, with an emphasis on potentially identifying the earliest (*i.e.* Stage I cancers), where the average number of molecules in the blood may be as low as 150 molecules. Four types of calculations were performed: (A) Average individual markers at 50% sensitivity

and 2%-5% false-positives, with one marker (*i.e.* a protein marker) at 90% sensitivity but with 10% false-positives; (B) Average individual markers at 50% sensitivity and 2%-5% false-positives, with one marker at 80% sensitivity but with 15% false-positives; (C) Average individual markers at 50% sensitivity and 2%-5% false-positives, with two markers at 90% sensitivity each but with 10% false-positives each; and (D) Average individual markers at 50% sensitivity and 2%-5% false-positives, with two markers at 80% sensitivity each but with 15% false-positives each.

[0306] Figures 47-48 illustrate results for calculated overall Sensitivity and Specificity for 24 markers using conditions (A) and (C). The sensitivity curves provide overall sensitivity as a function of the average number of molecules in the blood for each marker, with separate curves for each minimum number of markers needed to call a sample as positive. The specificity curves provide overall specificity as a function of individual marker false-positive rates, again with separate curves for each minimum number of markers needed to call a sample as positive. The calculated numbers for overall Sensitivity and Specificity for 24 markers using the above 4 conditions are provided in the tables below.

Table 14.

24 Markers Sensitivity Avg. Indiv. Mkr,:50% Sensitivity; One Mkr 90% Sensitivity				
Average Number of Molecules in Blood	Mutation, 1 Positive	24 markers, Minimum 3 Positive	24 markers, Minimum 4 Positive	24 markers, Minimum 5 Positive
150	22.1%	88.2%	68.9%	44.4%
200	28.1%	95.7%	86.4%	69.2%
300	39.4%	99.3%	97.6%	93.3%
400	48.8%	99.9%	99.5%	98.6%
480	55.1%	100.0%	99.9%	99.5%
600	63.2%	100.0%	100.0%	99.9%

Table 15.

24 Markers Specificity Avg. Indiv. Mkr,: 2%-5% FP; One Mkr.: 10% FP			
Individual marker FP rate	Minimum 3 Markers Positive	Minimum 4 Markers Positive	Minimum 5 Markers Positive
2%	91.9%	99.1%	99.9%
3%	81.8%	97.1%	99.7%
4%		93.2%	98.9%
5%		86.7%	97.3%

Table 16.

24 Markers Sensitivity Avg. Indiv. Mkr: 50% Sensitivity; One Mkr: 80% Sensitivity				
Average Number of Molecules in Blood	Mutation, 1 Positive	24 markers, Minimum 3 Positive	24 markers, Minimum 4 Positive	24 markers, Minimum 5 Positive
150	22.1%	84.8%	65.1%	41.5%
200	28.1%	93.5%	83.1%	65.6%
300	39.4%	98.7%	96.2%	90.9%
400	48.8%	99.7%	99.1%	97.7%
480	55.1%	99.9%	99.7%	99.2%
600	63.2%	100.0%	100.0%	99.8%

Table 17.

24 Markers Specificity Avg. Indiv. Mkr,: 2%-5% FP; One Mkr.: 15% FP			
Individual marker FP rate	Minimum 3 Markers Positive	Minimum 4 Markers Positive	Minimum 5 Markers Positive
2%	87.9%	98.7%	99.9%
3%		95.7%	99.5%
4%		89.8%	98.4%
5%		80.1%	96.0%

Table 18.

24 Markers Sensitivity Avg. Indiv. Mkr: 50% Sensitivity; Two Mkrs: 90% Sensitivity				
Average Number of Molecules in Blood	Mutation, 1 Positive	24 markers, Minimum 3 Positive	24 markers, Minimum 4 Positive	24 markers, Minimum 5 Positive
150	22.1%	90.9%	71.9%	46.7%
200	28.1%	97.4%	89.0%	72.0%
300	39.4%	99.8%	98.8%	95.2%
400	48.8%	100.0%	99.9%	99.4%
480	55.1%	100.0%	100.0%	99.8%
600	63.2%	100.0%	100.0%	100.0%

Table 19.

24 Markers Specificity Avg. Indiv. Mkr,: 2%-5% FP; Two Mkrs.: 10% FP			
Individual marker FP rate	Minimum 3 Markers Positive	Minimum 4 Markers Positive	Minimum 5 Markers Positive
2%		95.7%	99.7%
3%		90.4%	98.9%
4%		83.0%	97.3%
5%			94.7%

Table 20.

24 Markers Sensitivity Avg. Indiv. Mkr,:50% Sensitivity; Two Mkrs: 80% Sensitivity				
Average Number of Molecules in Blood	Mutation, 1 Positive	24 markers, Minimum 3 Positive	24 markers, Minimum 4 Positive	24 markers, Minimum 5 Positive
150	22.1%	90.2%	71.1%	46.1%
200	28.1%	97.0%	88.4%	71.3%
300	39.4%	99.6%	98.5%	94.7%
400	48.8%	99.9%	99.8%	99.2%
480	55.1%	100.0%	99.9%	99.8%
600	63.2%	100.0%	100.0%	100.0%

Table 21.

24 Markers Specificity Avg. Indiv. Mkr,: 2%-5% FP; Two Mkrs.: 15% FP			
Individual marker FP rate	Minimum 3 Markers Positive	Minimum 4 Markers Positive	Minimum 5 Markers Positive
2%		90.4%	99.2%
3%			97.4%
4%			93.9%
5%			88.0%

[0307] Before evaluating what advantages, if any, there are to combining protein (or other markers) with methylation markers, an analysis is performed on the original 24 marker set, with average individual marker sensitivity at 50%, and 2%-5% false positive rates. In this example, if there is an average of 150 molecules in the blood for the earliest cancer (Stage I), and if that would cover at least one mutation, then the sensitivity for identifying such a cancer by next generation sequencing would be 22.1% (See Figure 33A). If the individual marker FP rate is 3%, then if there is a 3-marker minimum, then overall FP rate is 5.4% for 24 markers, for a specificity of 94.6% (See Figure 33B). At 3 markers, for Stage I cancer (at about 150 molecules of each positive marker in the blood), the test would miss 42.3%; *i.e.* for Stage I cancer the overall sensitivity would be 57.7% (See Figure 33A). However, if the individual marker FP rate is 5%, then if there is a 4-marker minimum, then overall FP rate is 6.6% for 24 markers, for a specificity of 93.4% (See Figure 33B). At 4 markers, for Stage I cancer (at about 150 molecules of each positive marker in the blood), the test would miss 64.7%; *i.e.* for Stage I cancer the sensitivity would be 35.3% (See Figure 33A). While the specificity is reasonable, limiting the number of samples that would need to be retested in the second step of the assay, the assay would miss 2/3rd of the earliest cancers.

[0308] The above numbers are compared to the graph in Figure 46, *i.e.* condition (A), with average individual markers at 50% sensitivity and 2%-5% false-positives, with one marker (*i.e.* a protein marker) at 90% sensitivity but with 10% false-positives. While use of only 3 markers positive out of 24 markers provides a sensitivity of 88.2%, even with an individual marker 2% FP rate, the specificity would be 91.9%, if the FP rate were 3%, the overall specificity drops to 81.8%. This is the negative influence of the single marker with the high FP rate of 10%. Use of 4 markers positive out of 24 markers provides a sensitivity of 68.9% - still better than the original number of 57.7%, but now specificity improves to 97.1% with individual marker FP rates of 3%.

[0309] For condition (B), the average individual markers are at 50% sensitivity and 2%-5% false-positives, with one marker at 80% sensitivity but with 15% false-positives. Under these conditions, the specificity for 3 markers positive out of 24 markers would be at 87.9%, and thus would most likely not be used. Use of 4 markers positive out of 24 markers provides a sensitivity of 65.1% - still better than the original number of 57.7%, but now specificity improves to 95.7% with individual marker FP rates of 3%.

[0310] What if there are two markers with higher sensitivity (as well as higher FP rates)? For condition (C), with average individual markers at 50% sensitivity and 2%-5% false-positives, with two markers at 90% sensitivity each but with 10% false-positives each, see graph in Figure 47. Under these conditions, the specificity for 3 markers positive out of 24 markers would be below 80%, and thus would not be used. Use of 4 markers positive out of 24 markers provides a sensitivity of 71.9% - still better than the original number of 57.7%, but now specificity is at 95.7% with individual marker FP rates of 2%. Should the individual marker FP rates rise to 3%, then overall specificity drops to 90.4%.

[0311] For condition (D), the average individual markers are at 50% sensitivity and 2%-5% false-positives, with two markers at 80% sensitivity each but with 15% false-positives each. Under these conditions, the specificity for 3 markers positive out of 24 markers would be below 80%, and thus would not be used. Use of 4 markers positive out of 24 markers provides a sensitivity of 71.1% - still better than the original number of 57.7%, but now specificity is at 90.4% with individual marker FP rates of 2%. Should the individual marker FP rates rise to 3%, then 5 markers would be required, and while overall specificity would rise to 97.4%, the sensitivity would drop to 46.1%, which is worse than the original number of 57.7%.

[0312] Thus, from analysis of the above 4 conditions (A-D), condition (C) provided the best improvement in overall sensitivity (71.9%) for detecting Stage I cancer, while still keeping overall specificity reasonable (95.7%) for the initial 24 marker screen, should it now include two markers with higher sensitivity (90%), but worse FP rate of 10% for each of these markers.

[0313] The calculated numbers for two of the overall Sensitivity and Specificity for 36 markers using two of the aforementioned 4 conditions: (A) Average individual markers at 50% sensitivity and 2%-5% false-positives, with one marker (*i.e.* a protein marker) at 90% sensitivity but with 10% false-positives; and (C) Average individual markers at 50% sensitivity and 2%-5% false-positives, with two markers at 90% sensitivity each but with 10% false-positives each – are provided in the tables below. The sensitivity curves provide overall sensitivity as a function of the average number of molecules in the blood for each marker, with separate curves for each minimum number of markers needed to call a sample as positive. The specificity curves provide overall specificity as a function of individual marker false-positive rates, again with separate curves for each minimum number of markers needed to call a sample as positive. The calculated numbers for overall Sensitivity and Specificity for 36 markers using the above two conditions are provided in the tables below.

Table 22.

36 Markers Sensitivity Avg. Indiv. Mkr,: 50% Sensitivity; One Mkr 90% Sensitivity					
Average Number of Molecules in Blood	Mutation, 1 Positive	36 markers, Minimum 3 Positive	36 markers, Minimum 4 Positive	36 markers, Minimum 5 Positive	36 markers, Minimum 6 Positive
150	22.1%	96.8%	89.2%	73.9%	53.2%
200	28.1%	99.3%	97.6%	93.3%	84.1%
240	33.0%	99.7%	99.1%	97.4%	93.4%
300	39.4%	99.9%	99.8%	99.3%	98.3%
400	48.8%	100.0%	100.0%	99.9%	99.8%
480	55.1%	100.0%	100.0%	100.0%	100.0%
600	63.2%	100.0%	100.0%	100.0%	100.0%

Table 23.

36 Markers Specificity Avg. Indiv. Mkr,: 2%-5% FP; One Mkr.: 10% FP				
Individual marker FP rate	Minimum 4 Markers Positive	Minimum 5 Markers Positive	Minimum 6 Markers Positive	Minimum 7 Markers Positive
2%	95.3%	99.4%	99.9%	100.0%
3%	84.1%	96.9%	99.5%	99.9%
4%		90.3%	98.0%	99.7%
5%			93.9%	98.7%

Table 24.

36 Markers Sensitivity Avg. Indiv. Mkr: 50% Sensitivity; Two Mkrs: 90% Sensitivity					
Average Number of Molecules in Blood	Mutation, 1 Positive	36 markers, Minimum 3 Positive	36 markers, Minimum 4 Positive	36 markers, Minimum 5 Positive	36 markers, Minimum 6 Positive
150	22.1%	98.1%	91.3%	76.3%	55.3%
200	28.1%	99.8%	98.8%	95.2%	86.7%
240	33.0%	99.9%	99.7%	98.6%	95.2%
300	39.4%	100.0%	99.9%	99.8%	99.1%
400	48.8%	100.0%	100.0%	100.0%	99.9%
480	55.1%	100.0%	100.0%	100.0%	100.0%
600	63.2%	100.0%	100.0%	100.0%	100.0%

Table 25.

36 Markers Specificity Avg. Indiv. Mkr,: 2%-5% FP; Two Mkrs.: 10% FP				
Individual marker FP rate	Minimum 4 Markers Positive	Minimum 5 Markers Positive	Minimum 6 Markers Positive	Minimum 7 Markers Positive
2%		97.0%	99.7%	100.0%
3%		89.8%	98.4%	99.8%
4%			95.0%	99.1%
5%			87.8%	97.4%

[0314] What are the advantages to combining protein (or other markers) with methylation markers, an analysis is performed on the original 36 marker set, with average individual marker sensitivity at 50%, and 2%-5% false positive rates? In this example, if there is an average of 150 molecules in the blood for the earliest cancer (Stage I), and if that would cover at least one mutation, then the sensitivity for identifying such a cancer by next generation sequencing would be 22.1% (See Figure 34A). If the individual marker FP rate is 2%, then if there is a 3-marker minimum, then overall FP rate is 5.7% for 36 markers, for a specificity of 94.3% (See Figure 34B). At 3 markers, for Stage I cancer (at about 150 molecules of each positive marker in the blood), the test would miss 17.4%; *i.e.* for Stage I cancer the overall sensitivity would be 82.6% (See Figure 34A). However, if the individual marker FP rate is 3%, then if there is a 4-marker minimum, then overall FP rate is 4.8% for 36 markers, for a specificity of 95.2% (See Figure 34B). At 4 markers, for Stage I cancer (at about 150 molecules of each positive marker in the blood), the test would miss 34.1%; *i.e.* for Stage I cancer the sensitivity would be 65.8% (See Figure 34A). While the specificity is reasonable, limiting the

number of samples that would need to be retested in the second step of the assay, the assay would miss 1/3rd of the earliest cancers.

[0315] The above numbers are compared to the results for condition (A), with average individual markers at 50% sensitivity and 2%-5% false-positives, with one marker (*i.e.* a protein marker) at 90% sensitivity but with 10% false-positives. Use of only 4 markers positive out of 36 markers provides a sensitivity of 89.2%, and with an individual marker 2% FP rate, the specificity would be 95.3%. If the FP rate were 3%, this would require use of 5 markers positive out of 36 markers to provide a sensitivity of 73.9% - still better than the original number of 65.8%, but now specificity improves to 96.9% with individual marker FP rates of 3%.

[0316] For condition (C), the average individual markers are at 50% sensitivity and 2%-5% false-positives, with two markers at 90% sensitivity each but with 10% false-positives each. Under these conditions, use of 5 markers positive out of 36 markers provides a sensitivity of 76.3% - still better than the original number of 65.8%, but now specificity is at 97.0% with individual marker FP rates of 2%. Should the individual marker FP rates rise to 3%, then overall specificity drops to 89.8%.

[0317] Thus, from analysis of the above conditions (A, C), condition (C) provided the best improvement in overall sensitivity (76.3%) for detecting Stage I cancer, while still keeping overall specificity reasonable (97.0%) for the initial 36 marker screen, should it now include two markers with higher sensitivity (90%), but worse FP rate of 10% for each of these markers.

[0318] Figures 49-50 illustrate results for calculated overall Sensitivity and Specificity for 48 markers using the aforementioned 2 conditions: (A) Average individual markers at 50% sensitivity and 2%-5% false-positives, with one marker (i.e. a protein marker) at 90% sensitivity but with 10% false-positives; and (C) Average individual markers at 50% sensitivity and 2%-5% false-positives, with two markers at 90% sensitivity each but with 10% false-positives each. The sensitivity curves provide overall sensitivity as a function of the average number of molecules in the blood for each marker, with separate curves for each minimum number of markers needed to call a sample as positive. The specificity curves provide overall specificity as a function of individual marker false-positive rates, again with separate curves for each minimum number of markers needed to call a sample as positive. The calculated numbers for overall Sensitivity and Specificity for 48 markers using the above two conditions are provided in the tables below.

Table 26.

48 Markers Sensitivity Avg. Indiv. Mkr,: 50% Sensitivity; One Mkr 90% Sensitivity						
Average Number of Molecules in Blood	Mutation, 1 Positive	48 markers, Minimum 4 Positive	48 markers, Minimum 5 Positive	48 markers, Minimum 6 Positive	48 markers, Minimum 7 Positive	48 markers, Minimum 8 Positive
150	22.1%	97.6%	93.3%	84.2%	69.7%	52.1%
200	28.1%	99.5%	98.6%	96.4%	91.5%	82.7%
240	33.0%	99.9%	99.7%	99.1%	97.7%	94.7%
300	39.4%	100.0%	99.9%	99.8%	99.5%	98.9%
400	48.8%	100.0%	100.0%	100.0%	100.0%	99.9%
480	55.1%	100.0%	100.0%	100.0%	100.0%	100.0%
600	63.2%	100.0%	100.0%	100.0%	100.0%	100.0%

Table 27.

48 Markers Specificity Avg. Indiv. Mkr,: 2%-5% FP; One Mkr.: 10% FP				
Individual marker FP rate	Minimum 5 Markers Positive	Minimum 6 Markers Positive	Minimum 7 Markers Positive	Minimum 8 Markers Positive
2%	97.3%	99.6%	100.0%	100.0%
3%	86.1%	97.0%	99.5%	99.9%
4%		87.4%	97.0%	99.4%
5%			88.5%	97.1%

Table 28.

48 Markers Sensitivity Avg. Indiv. Mkr: 50% Sensitivity; Two Mkrs: 90% Sensitivity						
Average Number of Molecules in Blood	Mutation, 1 Positive	48 markers, Minimum 4 Positive	48 markers, Minimum 5 Positive	48 markers, Minimum 6 Positive	48 markers, Minimum 7 Positive	48 markers, Minimum 8 Positive
150	22.1%	98.8%	95.2%	86.7%	72.4%	54.4%
200	28.1%	99.9%	99.4%	97.8%	93.5%	85.2%
240	33.0%	100.0%	99.9%	99.6%	98.7%	96.2%
300	39.4%	100.0%	100.0%	99.9%	99.8%	99.5%
400	48.8%	100.0%	100.0%	100.0%	100.0%	100.0%
480	55.1%	100.0%	100.0%	100.0%	100.0%	100.0%
600	63.2%	100.0%	100.0%	100.0%	100.0%	100.0%

Table 29.

48 Markers Specificity Avg. Indiv. Mkr,: 2%-5% FP; Two Mkrs.: 10% FP				
Individual marker FP rate	Minimum 5 Markers Positive	Minimum 6 Markers Positive	Minimum 7 Markers Positive	Minimum 8 Markers Positive
2%	86.3%	98.0%	99.8%	100.0%
3%		90.1%	98.2%	99.7%
4%			92.5%	98.5%
5%				94.1%

[0319] What are the advantages to combining protein (or other markers) with methylation markers, an analysis is performed on the original 48-marker set, with average individual marker sensitivity at 50%, and 2%-5% false positive rates? In this example, if there is an average of 150 molecules in the blood for the earliest cancer (Stage I), and if that would cover at least one mutation, then the sensitivity for identifying such a cancer by next generation sequencing would be 22.1% (See Figure 35A). If the individual marker FP rate is 2%, then if there is a 4-marker minimum, then overall FP rate is 3.1% for 48 markers, for a specificity of 96.9% (See Figure 35B). At 4 markers, for Stage I cancer (at about 150 molecules of each positive marker in the blood), the test would miss 15.1%; *i.e.* for Stage I cancer the overall sensitivity would be 84.9% (See Figure 35A). However, if the individual marker FP rate is 3%, then if there is a 5-marker minimum, then overall FP rate is 4.2% for 48 markers, for a specificity of 95.8% (See Figure 35B). At 5 markers minimum, for Stage I cancer (at about 150 molecules of each positive marker in the blood), the test would miss 28.4%; *i.e.* for Stage I cancer the sensitivity would be 71.6% (See Figure 35A). While the specificity is reasonable, limiting the number of samples that would need to be retested in the second step of the assay, the assay would miss a little over $1/4^{th}$ of the earliest cancers.

[0320] The above numbers are compared to the graph in Figure 48, *i.e.* condition (A), with average individual markers at 50% sensitivity and 2%-5% false-positives, with one marker (*i.e.* a protein marker) at 90% sensitivity but with 10% false-positives. Use of only 5 markers positive out of 48 markers provides a sensitivity of 93.3%, and with an individual marker 2% FP rate, the specificity would be 97.3%. If the FP rate were 3%, this would require use of 6 markers positive out of 48 markers to provide a sensitivity of 84.2% - still better than the original number of 71.6%, but now specificity improves to 97.0% with individual marker FP rates of 3%.

[0321] For condition (C), with average individual markers at 50% sensitivity and 2%-5% false-positives, with two markers at 90% sensitivity each but with 10% false-positives each, see graph in Figure 49. Under these conditions, use of 5 markers positive out of 48 markers provides

a sensitivity of 90.9% - still better than the original number of 71.6%, but now specificity is at 97.0% with individual marker FP rates of 2%. If the FP rate were 3%, this would require use of 6 markers positive out of 48 markers to provide a sensitivity of 81.0% - still better than the original number of 71.6%, but now specificity changes to 95.5% with individual marker FP rates of 3%.

[0322] From the above charts, the receiver operating characteristic (ROC) curves may be calculated by plotting Sensitivity vs. 1-Specificity. Since these are theoretical calculations, the curves were generated for different levels of average marker false-positive rates of 2%, 3%, 4%, and 5%. The AUC (Area under curve) were calculated for ROC curves for 24 markers, with average individual marker at 50% Sensitivity with 2%-3% FP, and one marker at 90% Sensitivity with 10% FP; 24 markers, with average individual marker at 50% Sensitivity with 2%-3% FP, and two markers at 90% Sensitivity with 10% FP; 36 markers, with average individual marker at 50% Sensitivity with 2%-3% FP, and one marker at 90% Sensitivity with 10% FP; 36 markers, with average individual marker at 50% Sensitivity with 2%-3% FP, and two markers at 90% Sensitivity with 10% FP; 48 markers, with average individual marker at 50% Sensitivity with 2%-3% FP, and one marker at 90% Sensitivity with 10% FP; and 48 markers, with average individual marker at 50% Sensitivity with 2%-3% FP, and two markers at 90% Sensitivity with 10% FP; and are provided in the Tables below. Using the benchmark of an average of 150 molecules in the blood for the earliest cancer (Stage I), and looking only at the 3% individual marker FP rate AUC values are at 77% with 24 markers (average individual marker at 50% Sensitivity), improve to 91% with 24 markers (average individual marker at 50% Sensitivity, and one marker at 90% Sensitivity with 10% FP), but decrease to 83% with 24 markers (average individual marker at 50% Sensitivity, and two markers at 90% Sensitivity with 10% FP); AUC values are at 87% with 36 markers (average individual marker at 50% Sensitivity), improve to 91% with 36 markers (average individual marker at 50% Sensitivity, and one marker at 90% Sensitivity with 10% FP), but decrease to 85% with 36 markers (average individual marker at 50% Sensitivity, and two markers at 90% Sensitivity with 10% FP); and AUC values are at 89% with 48 markers (average individual marker at 50% Sensitivity), improve to 91% with 48 markers (average individual marker at 50% Sensitivity, and one marker at 90% Sensitivity with 10% FP), and improve slightly to 92% with 48 markers (average individual marker at 50% Sensitivity, and two markers at 90% Sensitivity with 10% FP). These results illustrate that for multiple marker assays achieving good sensitivities and specificities for the earliest cancers is aided by having a single marker with above average sensitivities (*i.e.* 90%), even at the cost of a higher false-positive rate (*i.e.* 10%). There is no major benefit in sensitivity increasing the number of markers in the first step of the assay from 24 to 36 to 48

markers – but the increase in markers does improve specificity, which is important in limiting the number of samples that undergo the second step of the test.

Table 30.

24, 36, & 48 Marker AUC Values from ROC Curves; Avg. Indiv. Mkr,: 50% Sensitivity; One Mkr: 90% Sensitivity at 10% FP							
Total Markers: Individual marker FP rate	150 Molecules	200 Molecules	240 Molecules	300 Molecules	400 Molecules	480 Molecules	600 Molecules
24 Mkrs: 2%	93%	97%		99%	>99%	>99%	>99%
24 Mkrs: 3%	91%	96%		99%	>99%	>99%	>99%
36 Mkrs: 2%	94%	99%	>99%	>99%	>99%	>99%	>99%
36 Mkrs: 3%	91%	96%	99%	>99%	>99%	>99%	>99%
48 Mkrs: 2%	96%	99%	>99%	>99%	>99%	>99%	>99%
48 Mkrs: 3%	91%	96%	99%	>99%	>99%	>99%	>99%

Table 31.

24, 36, & 48 Marker AUC Values from ROC Curves; Avg. Indiv. Mkr,: 50% Sensitivity; Two Mkrs: 90% Sensitivity at 10% FP							
Total Markers: Individual marker FP rate	150 Molecules	200 Molecules	240 Molecules	300 Molecules	400 Molecules	480 Molecules	600 Molecules
24 Mkrs: 2%	85%	94%		99%	>99%	>99%	>99%
24 Mkrs: 3%	83%	93%		99%	>99%	>99%	>99%
36 Mkrs: 2%	87%	97%	99%	>99%	>99%	>99%	>99%
36 Mkrs: 3%	85%	96%	98%	99%	>99%	>99%	>99%
48 Mkrs: 2%	96%	99%	>99%	>99%	>99%	>99%	>99%
48 Mkrs: 3%	92%	98%	>99%	>99%	>99%	>99%	>99%

[0323] While the above calculations are based on increasing the sensitivity of one or two markers, what if the average sensitivity of individual markers was increased from 50% to 66%? Figures 51-53 illustrate results for calculated overall Sensitivity and Specificity for 24, 36, and 48 markers, respectively. These graphs are based on the assumption that the average individual marker sensitivity is 66%, and the average individual marker false-positive rate is from 2% to 5%. The sensitivity curves provide overall sensitivity as a function of the average number of molecules in the blood for each marker, with separate curves for each minimum number of markers needed to call a sample as positive. The specificity curves provide overall specificity as a function of individual marker false-positive rates, again with separate curves for each minimum number of markers needed to call a sample as positive. The calculated numbers for overall Sensitivity and Specificity for 24, 36, and 48 markers, respectively, where the average individual marker sensitivity is 50% (as described previously) or 66% are provided in the tables below.

Table 32.

24 Markers Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity				
Average Number of Molecules in Blood	Mutation, 1 Positive	24 markers, Minimum 3 Positive	24 markers, Minimum 4 Positive	24 markers, Minimum 5 Positive
150	22.1%	57.7%	35.3%	18.5%
200	28.1%	76.2%	56.7%	37.1%
240	33.0%	85.7%	70.6%	52.4%
300	39.4%	93.8%	84.9%	71.5%
400	48.8%	98.6%	95.8%	90.0%
480	55.1%	99.6%	98.6%	96.2%
600	63.2%	99.9%	99.8%	99.2%

Table 33.

24 Markers Sensitivity; Avg. Indiv. Mkr,: 66% Sensitivity				
Average Number of Molecules in Blood	Mutation, 1 Positive	24 markers, Minimum 3 Positive	24 markers, Minimum 4 Positive	24 markers, Minimum 5 Positive
150	22.1%	76.2%	56.7%	37.1%
200	28.1%	89.8%	77.5%	61.0%
240	33.0%	95.4%	88.1%	76.5%
300	39.4%	98.6%	95.8%	90.0%
400	48.8%	99.8%	99.3%	98.0%
480	55.1%	100.0%	99.9%	99.6%
600	63.2%	100.0%	100.0%	100.0%

Table 34.

24 Marker Specificity			
Individual marker FP rate	Minimum 3 Markers Positive	Minimum 4 Markers Positive	Minimum 5 Markers Positive
2%	98.4%	99.8%	99.9%
3%	94.6%	99.1%	99.9%
4%	87.1%	97.3%	99.6%
5%		93.4%	98.7%

Table 35.

36 Marker Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity					
Average Number of Molecules in Blood	Mutation, 1 Positive	36 markers, Minimum 3 Positive	36 markers, Minimum 4 Positive	36 markers, Minimum 5 Positive	36 markers, Minimum 6 Positive
150	22.1%	82.6%	65.8%	46.8%	29.7%
200	28.1%	93.8%	84.9%	71.5%	55.4%
240	33.0%	97.5%	92.8%	84.4%	72.4%
300	39.4%	99.4%	97.9%	94.5%	88.4%
400	48.8%	99.9%	99.8%	99.2%	98.0%
480	55.1%	100.0%	100.0%	99.9%	99.6%
600	63.2%	100.0%	100.0%	100.0%	100.0%

Table 36.

36 Marker Sensitivity; Avg. Indiv. Mkr,: 66% Sensitivity					
Average Number of Molecules in Blood	Mutation, 1 Positive	36 markers, Minimum 3 Positive	36 markers, Minimum 4 Positive	36 markers, Minimum 5 Positive	36 markers, Minimum 6 Positive
150	22.1%	93.8%	84.9%	71.5%	55.4%
200	28.1%	98.6%	95.8%	90.0%	80.9%
240	33.0%	99.6%	98.6%	96.2%	91.6%
300	39.4%	99.9%	99.8%	99.2%	98.0%
400	48.8%	100.0%	100.0%	100.0%	99.9%
480	55.1%	100.0%	100.0%	100.0%	100.0%
600	63.2%	100.0%	100.0%	100.0%	100.0%

Table 37.

36 Marker Specificity				
Individual marker FP rate	Minimum 3 Markers Positive	Minimum 4 Markers Positive	Minimum 5 Markers Positive	Minimum 6 Markers Positive
2%	94.3%	99.1%	99.9%	100.0%
3%	80.7%	95.2%	99.1%	99.9%
4%		84.9%	96.1%	99.2%
5%			88.2%	97.0%

Table 38.

48 Marker Sensitivity; Avg. Indiv. Mkr,: 50% Sensitivity						
Average Number of Molecules in Blood	Mutation, 1 Positive	48 markers, Minimum 4 Positive	48 markers, Minimum 5 Positive	48 markers, Minimum 6 Positive	48 markers, Minimum 7 Positive	48 markers, Minimum 8 Positive
150	22.1%	84.9%	71.6%	55.6%	39.6%	25.8%
200	28.1%	95.8%	90.1%	80.9%	68.7%	54.8%
240	33.0%	99.1%	97.2%	93.4%	87.1%	78.1%
300	39.4%	99.8%	99.3%	98.1%	95.6%	92.3%
400	48.8%	99.9%	99.9%	99.8%	99.7%	99.1%
480	55.1%	99.9%	99.9%	99.9%	99.9%	99.9%
600	63.2%	99.9%	99.9%	99.9%	99.9%	99.9%

Table 39.

48 Marker Sensitivity; Avg. Indiv. Mkr,: 66% Sensitivity						
Average Number of Molecules in Blood	Mutation, 1 Positive	48 markers, Minimum 4 Positive	48 markers, Minimum 5 Positive	48 markers, Minimum 6 Positive	48 markers, Minimum 7 Positive	48 markers, Minimum 8 Positive
150	22.1%	95.8%	90.0%	80.9%	68.7%	54.7%
200	28.1%	99.3%	98.0%	95.2%	90.3%	82.9%
240	33.0%	99.9%	99.6%	98.8%	97.1%	94.0%
300	39.4%	100.0%	100.0%	99.9%	99.6%	99.0%
400	48.8%	100.0%	100.0%	100.0%	100.0%	100.0%
480	55.1%	100.0%	100.0%	100.0%	100.0%	100.0%
600	63.2%	100.0%	100.0%	100.0%	100.0%	100.0%

Table 40.

48 Marker Specificity					
Individual marker FP rate	Minimum 4 Markers Positive	Minimum 5 Markers Positive	Minimum 6 Markers Positive	Minimum 7 Markers Positive	Minimum 8 Markers Positive
2%	96.9%	99.4%	99.9%	99.9%	99.9%
3%	84.3%	95.8%	99.1%	99.8%	99.9%
4%		82.5%	95.0%	98.8%	99.8%
5%				94.3%	98.6%

[0324] The above tables, and Figures 51-53, as well as Figures 33-35, allow for a direct comparison in the overall improvement in sensitivity when the average individual marker sensitivity improves from 50% to 66%. In this example, if there is an average of 150 molecules in the blood for the earliest cancer (Stage I), and if that would cover at least one mutation, then

the sensitivity for identifying such a cancer by next generation sequencing would be 22.1% (See any of the aforementioned figures). For 24 markers, with a minimum of 3 markers positive and a 3% FP rate, overall sensitivity improves from 57.7% to 76.2%, when the average individual marker sensitivity improves from 50% to 66%, for detecting Stage I cancer (at about 150 molecules of each positive marker in the blood, see Figures 33A and 51A, orange line). If the individual marker FP rate is 3%, then if there is a 3-marker minimum, then overall FP rate is 5.4% for 24 markers, for a specificity of 94.6% (See Figures 33B or 51B). However, if the individual marker FP rate is 5%, then if there is a 4-marker minimum, then overall FP rate is 6.6% for 24 markers, for a specificity of 93.4% (See Figure 33B). At 4 markers, for Stage I cancer (at about 150 molecules of each positive marker in the blood), overall sensitivity improves from 35.3% to 56.7%, when the average individual marker sensitivity improves from 50% to 66% (See Figure 33A and Figure 50A). For 36 markers, with a minimum of 3 markers positive and a 2% FP rate, overall sensitivity improves from 82.6% to 93.8%, when the average individual marker sensitivity improves from 50% to 66%, for detecting Stage I cancer (at about 150 molecules of each positive marker in the blood, see Figures 34A and 52A). If the individual marker FP rate is 2%, then if there is a 3-marker minimum, then overall FP rate is 5.7% for 36 markers, for a specificity of 94.3% (See Figure 34B or 52B). However, if the individual marker FP rate is 3%, then the assay requires a 4-marker minimum, then overall FP rate is 4.8% for 36 markers, for a specificity of 95.2% (See Figure 34B). At 4 markers, for Stage I cancer (at about 150 molecules of each positive marker in the blood), overall sensitivity improves from 65.8% to 84.9%, when the average individual marker sensitivity improves from 50% to 66% (See Figure 34A and Figure 51A). For 48 markers, with a minimum of 4 markers positive and a 2% FP rate, overall sensitivity improves from 84.9% to 95.8%, when the average individual marker sensitivity improves from 50% to 66%, for detecting Stage I cancer (at about 150 molecules of each positive marker in the blood, see Figures 35A and 52A). If the individual marker FP rate is 2%, then if there is a 4-marker minimum, then overall FP rate is 3.1% for 48 markers, for a specificity of 96.9% (See Figures 35B or 52B). However, if the individual marker FP rate is 3%, then the assay requires a 5-marker minimum, then overall FP rate is 4.2% for 48 markers, for a specificity of 95.8% (See Figure 35B). At 5 markers, for Stage I cancer (at about 150 molecules of each positive marker in the blood), overall sensitivity improves from 71.6% to 90.0%, when the average individual marker sensitivity improves from 50% to 66% (See Figure 35A and Figure 52A).

[0325] From the above charts, the receiver operating characteristic (ROC) curves may be calculated by plotting Sensitivity vs. 1-Specificity. Since these are theoretical calculations, the curves were generated for different levels of average marker false-positive rates of 2%, 3%, 4%,

and 5%. The AUC values, calculated for ROC curves for 24 markers, with average individual marker at 66% Sensitivity with 2%-3% FP; 36 markers, with average individual marker at 66% Sensitivity with 2%-3% FP; and 48 markers, with average individual marker at 66% Sensitivity with 2%-3% FP; are provided in the Table below. Using the benchmark of an average of 150 molecules in the blood for the earliest cancer (Stage I), and looking only at the 3% individual marker FP rate AUC values are at 77% with 24 markers (average individual marker at 50% Sensitivity), improve to 87% with 24 markers (average individual marker at 66% Sensitivity); AUC values are at 87% with 36 markers (average individual marker at 50% Sensitivity), improve to 95% with 36 markers (average individual marker at 66% Sensitivity); and AUC values are at 89% with 48 markers (average individual marker at 50% Sensitivity), improve to 97% with 48 markers (average individual marker at 66% Sensitivity). These results illustrate that for multiple marker assays achieving good sensitivities and specificities for the earliest cancers is aided when the average individual marker sensitivity improves from 50% to 66%.

Table 41.

24, 36, & 48 Marker AUC Values from ROC Curves; Avg. Indiv. Mkr,: 66% Sensitivity							
Total Markers: Individual marker FP rate	150 Molecules	200 Molecules	240 Molecules	300 Molecules	400 Molecules	480 Molecules	600 Molecules
24 Mkrs: 2%	88%	95%	98%	>99%	>99%	>99%	>99%
24 Mkrs: 3%	87%	94%	97%	99%	>99%	>99%	>99%
36 Mkrs: 2%	96%	99%	>99%	>99%	>99%	>99%	>99%
36 Mkrs: 3%	95%	99%	>99%	>99%	>99%	>99%	>99%
48 Mkrs: 2%	98%	>99%	>99%	>99%	>99%	>99%	>99%
48 Mkrs: 3%	97%	99%	>99%	>99%	>99%	>99%	>99%

[0326] How would increasing the average individual marker sensitivity from 50% sensitivity to 66% sensitivity improve upon a one-step cancer assay? To review: the challenge is to screen 107 million adults in the U.S. over the age of 50 for colorectal cancer – of which there are about 135,000 new cases that are diagnosed a year. In this example, if there is an average of 300 molecules in the blood for early cancer (Stage I & II), and taking the best-case scenario of individual marker FP rate is 2%, then if there is a 3-marker minimum, then overall FP rate is 1.6% for 24 markers, for a specificity of 98.4% (See Figure 33B or 50B). At 3 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), for average marker sensitivity of 50%, the test would miss 6.2%; *i.e.* for Stage I & II cancer the overall sensitivity would be 93.8% (See Figure 33A), *e.g.* the test would correctly identify 93.8% of individuals with disease, which would be 126,630 individuals (out of 135,000 new cases). At a

specificity of 98.4%, for 107 million individuals screened, the test would also generate 1.6% x 107,000,000 = 1,712,000 false positives. Thus, the positive predictive value would be 126,630/(126,630 + 1,712,000) = around 6.8%, in other words, only one in 14 individuals who tested positive would actually have colorectal cancer, the rest would be false-positives. At 3 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), for average marker sensitivity of 66%, the test would miss 1.4%; *i.e.* for Stage I & II cancer the overall sensitivity would be 98.6% (See Figure 50A), *e.g.* the test would correctly identify 98.6% of individuals with disease, which would be 133,110 individuals (out of 135,000 new cases). At a specificity of 98.4%, for 107 million individuals screened, the test would also generate 1.6% x 107,000,000 = 1,712,000 false positives. Thus, the positive predictive value would be 133,110/(133,110 + 1,712,000) = around 7.2%, in other words, only one in 14 individuals who tested positive would actually have colorectal cancer, the rest would be false-positives. Thus, if the FP is low, *i.e.* 2%, then there is marginal benefit in going from an average marker sensitivity of 50% to an average marker sensitivity of 66%.

[0327] However, if the individual marker FP rate is more realistic, say 4%, then more markers will be required to achieve better than 98% specificity, and this will be at the cost of sensitivity. If individual marker FP rate is 4%, then if there is a 5-marker minimum, then overall FP rate is 0.4% for 24 markers, for a specificity of 99.6% (See Figure 33B). At 5 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), at an average marker sensitivity of 50%, the test would miss 28.5%; *i.e.* for Stage I & II cancer the overall sensitivity would be 71.5% (See Figure 33A), e.g. the test would correctly identify 71.5% of individuals with disease, which would be 90,540 individuals (out of 135,000 new cases). At a specificity of 99.6%, for 107 million individuals screened, the test would also generate 0.4% x 107,000,000 = 428,000 false positives. Thus, the positive predictive value would be 90,540/(90,540 + 428,000) = around 17.5%, in other words, one in 5.7 individuals who tested positive would actually have colorectal cancer, the rest would be false-positives. A PPV of 17.5% is quite respectable, however, it would be achieved at the cost of missing 28.5% of early cancer. At 3 markers, for Stage I & II cancer (at about 300 molecules of each positive marker in the blood), for average marker sensitivity of 66%, the test would miss 10.0%; i.e. for Stage I & II cancer the overall sensitivity would be 90.0% (See Figure 50A), *e.g.* the test would correctly identify 90.0% of individuals with disease, which would be 121,500 individuals (out of 135,000 new cases). At a specificity of 99.6%, for 107 million individuals screened, the test would also generate 0.4% x 107,000,000 = 428,000 false positives. Thus, the positive predictive value would be 121,500/(121,500 + 428,000) = around 22.1 %, in other words, one in 4.5 individuals who tested positive would actually have colorectal cancer, the rest would be false-positives. A

PPV of 22.1% is excellent, and further, it would be achieved at the cost of missing only 10% of early cancer. Thus, if the FP is more realistic *i.e.* 4%, then there is a significant benefit in going from an average marker sensitivity of 50% to an average marker sensitivity of 66%.

[0328] Returning to the example of colorectal cancer, in particular the cases of microsatellite stable tumors (MSS) where the mutation load is low, for these calculations when relying on NGS sequencing alone (assuming 150 molecules with one mutation in the blood), an estimated 78% of early colorectal cancer would be missed. Again, to put these number in perspective, in the U.S., about 135,000 new cases of colorectal cancer were diagnosed in 2018, of which about 60% is late cancer (*i.e.* Stage III & IV). About 107 million individuals in the U.S. are over the age of 50 and should be tested for colorectal cancer. While it cannot be predicted how many individuals have a hidden cancer (*i.e.* Stage I) within them, who are non-compliant to testing, for the purposes of this calculation, assume that the average late cancer was once the average early cancer, and thus individuals with Stage I cancer would be about 40,500 individuals. With the assumption of these samples containing at least 150 molecules with one mutation in the blood, such a test would find 8,910 individuals (out of 40,500 individuals with Stage I cancer) with colorectal cancer. However, with a specificity for sequencing at 98%, there would be about 2.1 million false-positives. The positive predictive value of such a test would be around 0.4%, in other words, only one in 236 individuals who tested positive would actually have Stage I colorectal cancer, the rest would be false-positives. In contrast, consider the two-step methylation marker test described above, wherein the first step has 24 methylation markers specific to GI cancers, while the second step has 48 methylation markers specific to colorectal cancer. In this example, the average individual marker sensitivity is set at 66%. In this example, as above, the calculations are done with the anticipation of an average of 150 methylated molecules per positive marker in the blood. Assuming individual marker false-positive rates of 3%, and with the first step requiring a minimum of 3 markers positive, then with an overall specificity of 94.6%, the first step would identify 5,778,000 individuals (out of 107,000,000 total adults over 50 in the U.S.) which would include at 76.2% sensitivity or about 30,861 individuals with Stage I colorectal cancer (out of 40,500 individuals with Stage I cancer). However, those 5,778,000 presumptive positive individuals would be evaluated in the second step of 48 markers requiring a minimum of 5 markers positive, then the two-step test would identify 76.2% x 90.0% = 68.6% = 27,775 individuals (out of 40,500 individuals with Stage I cancer) with colorectal cancer. With a specificity of 95.8%, the second test would also generate 5,778,000 x 4.2% = 242,676 false-positives. The positive predictive value of such a test would be 27,775/270,451 = 10.3%, in other words, 1 in 10 individuals who tested positive would actually have Stage I colorectal cancer, an extraordinarily successful screen to focus on those patients who would most

benefit from follow-up colonoscopy. Since >90% of individuals identified with Stage I colon cancer have long-term survival after just surgery, the benefit in lives saved would be of incalculable value.

[0329] How would the above numbers change if the initial test in the two-step assay uses 36 markers? In this example, as above, the calculations are done with the anticipation of an average of 150 methylated molecules per positive marker in the blood. Assuming individual marker false-positive rates of 3%, and with the first step requiring a minimum of 4 markers positive, then with an overall specificity of 95.2%, the first step would identify 5,136,000 individuals (out of 107,000,000 total adults over 50 in the U.S.) which would include at 84.9% sensitivity or about 34,385 individuals with Stage I colorectal cancer (out of 40,500 individuals with Stage I cancer). However, those 5,136,000 presumptive positive individuals would be evaluated in the second step of 48 markers requiring a minimum of 5 markers positive, then the two-step test would identify 84.9% x 90.0% = 76.4% = 30,946 individuals (out of 40,500 individuals with Stage I cancer) with colorectal cancer. With a specificity of 95.8%, the second test would also generate 5,136,000 x 4.2% = 215,712 false-positives. The positive predictive value of such a test would be 30,946/246,658 = 12.5%, in other words, 1 in 8 individuals who tested positive would actually have Stage I colorectal cancer. In reality, one would need to also include the success for identifying Stage 2 and higher cancers. In expanding this example, the calculations are done with the anticipation that Stage I CRC has an average of 150 methylated molecules per positive marker in the blood, Stage II CRC has an average of 200 methylated molecules per positive marker, and the higher stages (III & IV) have at least an average of 300 methylated molecules per positive marker, and the higher stages. Also, to be consistent with the idea that as the test is used repeatedly, more of early and less of late CRC will be detected, then an estimate of 40,500 individuals with Stage I cancer, 40,500 individuals with Stage II cancer, and the remaining 54,000 individuals have late-stage cancer = 135,000 total individuals with colorectal cancer identified per year in the U.S. The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 95.8% would be identified in the first step, of which 95.8% x 98.0% = 93.9% = 38,023 individuals with Stage II cancer would be verified in the second step. For Stage III and IV cancer, 99.8% would be identified in the first step, of which 99.8% x (100%) = 53,892 individuals with late cancer would be identified. This brings the total identified at 30,946 + 38,023 + 53,892 = 122,861 individuals out of 135,000 with colorectal cancer. Overall, the positive predictive value of such a test would be 122,861/369,519 = 33.2%, in other words, 1 in 3 individuals who tested positive would actually have colorectal cancer, and this test would identify 68,969/81,000 or 85% of those individuals with early cancer – which would be unprecedented in diagnostic approaches to detect early cancer.

[0330] The ultimate goal is to develop a high-throughput scalable test to detect the majority of cancers that occur worldwide. The solid tumor cancers have been grouped into the following subclasses, as listed below in Tables 42, 43, and 44 for both sexes, for men, and for women.

Table 42

Global cancer incidence; Both Sexes (Numbers in thousands; most common cancers have incidence above 100,000 per year)				
	Incidence	% Group	% total All	(Total)
Group 1:				
Colorectal (1,801)	1801	52.9%	12.9%	13981
Stomach (1,033)	1033	30.3%	7.4%	13981
Esophagus (572)	572	16.8%	4.1%	13981
Total, Group 1:	3406			
Group 2:				
Breast (2,089)	2089	62.6%	14.9%	13981
Endometrial & Cervical (570)	570	17.1%	4.1%	13981
Uterine (382)	382	11.5%	2.7%	13981
Ovarian (295)	295	8.8%	2.1%	13981
Total, Group 2:	3336			
Group 3:				
Lung (2,093)	2093	59.9%	15.0%	13981
Head & Neck (832)	832	23.8%	6.0%	13981
Thyroid (567)	567	16.2%	4.1%	13981
Total, Group 3:	3492			
Group 4:				
Prostate (1,276)	1276	57.3%	9.1%	13981
Bladder (549)	549	24.6%	3.9%	13981
Kidney (403)	403	18.1%	2.9%	13981
Total, Group 4:	2228			
Group 5:				
Liver (841)	841	55.4%	6.0%	13981
Pancreas (459)	459	30.2%	3.3%	13981
Gallbladder (219)	219	14.4%	1.6%	13981
Total, Group 5:	1519			
Total	13981			

Table 43

Global cancer incidence; Male (Numbers in thousands; most common cancers have incidence above 100,000 per year)				
	Incidence	% Group	% total All	(Total)
Group 1:				
Colorectal (1,801)	1006	48.2%	14.1%	7114
Stomach (1,033)	683	32.7%	9.6%	7114
Esophagus (572)	400	19.1%	5.6%	7114
Total, Group 1:	2089			
Group 2:				
Breast (2,089)				
Endometrial & Cervical (570)				
Uterine (382)				
Ovarian (295)				
Total, Group 2:				
Group 3:				
Lung (2,093)	1368	64.1%	19.2%	7114
Head & Neck (832)	635	29.8%	8.9%	7114
Thyroid (567)	131	6.1%	1.8%	7114
Total, Group 3:	2134			
Group 4:				
Prostate (1,276)	1276	65.3%	17.9%	7114
Bladder (549)	424	21.7%	6.0%	7114
Kidney (403)	254	13.0%	3.6%	7114
Total, Group 4:	1954			
Group 5:				
Liver (841)	597	63.7%	8.4%	7114
Pancreas (459)	243	25.9%	3.4%	7114
Gallbladder (219)	97	10.4%	1.4%	7114
Total, Group 5:	937			
Total	7114			

Table 44

Global cancer incidence; Female (Numbers in thousands; most common cancers have incidence above 100,000 per year)				
	Incidence	% Group	% total All	(Total)
Group 1:				
Colorectal (1,801)	795	60.4%	11.5%	6930
Stomach (1,033)	350	26.6%	5.1%	6930
Esophagus (572)	172	13.1%	2.5%	6930
Total, Group 1:	1317			
Group 2:				
Breast (2,089)	2089	62.6%	30.1%	6930
Endometrial & Cervical (570)	570	17.1%	8.2%	6930
Uterine (382)	382	11.5%	5.5%	6930
Ovarian (295)	295	8.8%	4.3%	6930
Total, Group 2:	3336			
Group 3:				
Lung (2,093)	725	53.4%	10.5%	6930
Head & Neck (832)	196	14.4%	2.8%	6930
Thyroid (567)	436	32.1%	6.3%	6930
Total, Group 3:	1357			
Group 4:				
Prostate (1,276)	0	0.0%	0.0%	6930
Bladder (549)	216	63.9%	3.1%	6930
Kidney (403)	122	36.1%	1.8%	6930
Total, Group 4:	338			
Group 5:				
Liver (841)	244	41.9%	3.5%	6930
Pancreas (459)	216	37.1%	3.1%	6930
Gallbladder (219)	122	21.0%	1.8%	6930
Total, Group 5:	582			
Total	6930			

[0331] The above list does not include liquid cancers, nor some of the less common solid tumors. Worldwide incidence (numbers in thousands) of liquid tumors include Non-Hodgkin lymphoma (225), leukemia (187), multiple myeloma (70), and Hodgkin lymphoma (33). These would be detected in a separate test not discussed herein. Further, the list excludes melanoma

(287) and brain tumors (134). Testing for these would be done with separate sets of markers, optimized as described above for colorectal cancer. In addition, while some cancers listed in the tables above are of extreme medical importance (*e.g.*, mesothelioma, thyroid cancer, and the three different subcategories of kidney cancer), their biology is sufficiently different as to usually merit a separate set of markers, again, optimized as described above for colorectal cancer.

[0332] Thus, for the present application, a Pan-Oncology test is developed that would include the following major cancers by the following groupings: Group 1 (colorectal, stomach, and esophagus); Group 2 (breast, endometrial, ovarian, cervical, and uterine); Group 3 (lung and head & neck); Group 4 (prostate and bladder); and Group 5 (liver, pancreatic, or gall bladder). Note that some cancers within Group 3 may be tested as a sputum sample, and cancers in Group 4 may be tested as a urine sample.

[0333] Careful analysis of the TCGA methylation database revealed a general commonality in methylation patterns among cancers within these 5 separate goups. Further, there are some methylation markers that are common among several cancers, while absent in normal white blood cells. The following strategy was used to design a multi-step pan-oncology test.

[0334] The first step is to identify markers that cover multiple cancers in one or more of the above groups. The markers should be sufficiently diverse as to cover cancers in all 5 groups. For example, a first step of the assay would use a set of 96 markers that on average comprise of at least 36 markers with 50% sensitivity that covers each of the aforementioned 16 types of solid tumors (covered in the 5 Groups; see Figure 1E; for 66% sensitivity, see Figure 1C). If at least 5 markers are positive, the assay would then move to a second step that would be used to verify the initial results and identify the most probable tissue of origin. In most cases, more than 5 markers would be positive, and then pattern of distribution of these methylation markers would guide the choice of which groups to test in the second step. The second step of the assay would test, on average, 2 or more sets of the group-specific markers. For example, the second step of the assay would use 2 or more sets of 64 group-specific markers that, on average, comprise of at least 36 markers with 50% sensitivity that covers each of the aforementioned types of solid tumors that may be present in that group (for 66% sensitivity, see Figure 1D). By scoring the markers that are positive and comparing to predicted positives for each cancer type within the group tested, the physician can identify the most probable tissue of origin, and subsequently send the patient to the appropriate imaging.

[0335] A close evaluation of the TCGA database reveals pan-oncology markers that meet the criteria for use in a set of 96 markers that on average comprise of at least 36 markers with 50% sensitivity that covers each of the aforementioned 16 types of solid tumors. These pan-

oncology markers include but are not limited to cancer-specific microRNA markers, cancer-specific ncRNA and lncRNA markers, cancer-specific exon transcripts, tumor-associated antigens, cancer protein markers, protein markers that can be secreted by solid tumors into the blood, common mutations, primary CpG sites that are solid tumor and tissue specific markers, chromosomal regions or sub-regions within which are primary CpG sites that are solid tumor and tissue specific markers, and primary and flanking CpG sites that are solid tumor and tissue specific markers. Methods for detecting said markers have been discussed *supra*, and these markers are listed below and in accompanying figures.

[0336] Blood-based, solid tumor-specific microRNA markers derived through analysis of TCGA microRNA datasets, includes, but is not limited to, the following markers: (mir ID , Gene ID); hsa-mir-21, MIR21; hsa-mir-182, MIR182; hsa-mir-454, MIR454; hsa-mir-96, MIR96; hsa-mir-183, MIR183; hsa-mir-549, MIR549; hsa-mir-301^{a}, MIR301A; hsa-mir-548f-1, MIR548F1; hsa-mir-301b, MIR301B; hsa-mir-103-1, MIR1031; hsa-mir-18^{a}, MIR18A; hsa-mir-147b, MIR147B; hsa-mir-4326, MIR4326; and hsa-mir-573, MIR573. These markers may be present in exosomes, tumor-associated vesicles, Argonaute complexes, or other protected states in the blood.

[0337] Figure 53 provides a list of blood-based, solid tumor-specific ncRNA and lncRNA markers derived through analysis of various publicly available Affymetrix Exon ST CEL data, which were aligned to GENCODE annotations to generate ncRNA and lncRNA transcriptome datasets. Comparative analyses across these datasets (various cancer types, along with normal tissues, and peripheral blood) were conducted to generate the ncRNA and lncRNA markers list. Such lncRNA and ncRNA may be enriched in exosomes or other protected states in the blood.

[0338] In addition, Figure 54 provides a list of blood-based solid tumor-specific exon transcripts that may be enriched in exosomes, tumor-associated vesicles, or other protected states in the blood. Overexpressed oncogene transcripts, or transcripts of mutant oncogenes may be enriched in exosomes, as they may drive spread of the cancer.

[0339] Figure 55 provides a list of cancer protein markers, identified through mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from solid tumors, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0340] Protein markers that can be secreted by solid tumors into the blood include, but are not limited to: (Protein name, UniProt ID); Uncharacterized protein C19orf48, Q6RUI8; Protein FAM72B, Q86X60; Protein FAM72D, Q6L9T8; Hydroxyacylglutathione hydrolase-like protein, Q6PII5; Putative methyltransferase NSUN5, Q96P11; RNA pseudouridylate synthase

domain-containing protein 1, Q9UJJ7; Collagen triple helix repeat-containing protein 1, Q96CG8; Interleukin-11, P20809; Stromelysin-2, P09238; Matrix metalloproteinase-9, P14780; Podocan-like protein 1, Q6PEZ8;Putative peptide YY-2, Q9NRI6; Osteopontin, P10451; Sulfhydryl oxidase 2, Q6ZRP7; Glypican-2, Q8N158; Macrophage migration inhibitory factor, P14174; Peptidyl-prolyl cis-trans isomerase A, P62937; Calreticulin, P27797. A comparative analysis was performed across various TCGA datasets (tumors, normals), followed by an additional bioinformatics filter (Meinken et al., "Computational Prediction of Protein Subcellular Locations in Eukaryotes: an Experience Report," *Computational Molecular Biology* 2(1):1-7 (2012), which is hereby incorporated by reference in its entirety), which predicts the likelihood that the translated protein is secreted by the cells.

[0341] The distribution of mutations in solid tumors are available in the public COSMIC database, with the commonly altered genes across solid tumors listed as: TP53 (tumor protein p53), TTN (titin), MUC16 (mucin 16), and KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog).

[0342] A deep analysis of the TCGA database of methylation markers that are absent in blood but on average are present in solid tumor types at 50% sensitivity show three general categories of clusters: (i) Markers that are present in colorectal cancers, and related GI cancer (stomach & esophagus), (ii) Markers that are present in colorectal cancers, and related GI cancer (stomach & esophagus), as well as other tumors, and (iii) Markers that are mostly absent in colorectal cancers, but present in other tumors. Second, while for some tumor types one could readily identify markers that were unique to that group, such as Group 2 (breast, endometrial, ovarian, cervical, and uterine), for other tumor types such as lung cancer or pancreatic cancer, it was difficult to identify methylation markers that were unique to that cancer. Consequently, to assemble a set of 96 markers that satisfied the criteria of at least 36 markers with 50% sensitivity that covers each of the aforementioned 16 types of solid tumors, the first 48 markers comprised of about 12 markers that were strongly represented in Group 2 tumors, about 12 markers that were strongly represented in Group 3 tumors, about 12 markers that were strongly represented in Group 4 tumors, and about 12 markers that were strongly represented in Group 5 tumors. The remaining 48 markers comprised of about 12 markers that were strongly represented in Groups 1 & 2 tumors, about 12 markers that were strongly represented in Groups 1 & 3 tumors, about 12 markers that were strongly represented in Groups 1 & 4 tumors, and about 12 markers that were strongly represented in Groups 1 & 5 tumors.

[0343] Figure 56 provides a list of primary CpG sites that are solid tumors and tissue-specific markers, that may be used to identify the presence of solid tumors from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood.

Figure 57 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are solid tumors and tissue-specific markers, that may be used to identify the presence of solid tumors from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. These lists contain preferred primary CpG sites and their flanking sites, as well as alternative markers that are low to no-CRC, and alternative markers that are high is CRC, with or without being high for other cancers as well. Primer sets for exemplary prefered and alternate methylation markers are listed in Table 46 in the experimental section.

[0344] Table 47, in the experimental section, provides simulations of the 96-marker assay, with average sensitivities of 50%, for identifying most probably group for tissue of origin, for both sexes. A set of 96 markers was assembled as above and the percentage of samples positive within each of the cancer patients in the TCGA and GEO databases was assessed. The total number of patients for each cancer analyzed are: Group 1 (colorectal, CRC-PT = 395; stomach, ST-Pt = 260; esophagus, ES-Pt = 185); Group 2 (breast, BR-Pt = 668; endometrial, END-Pt = 431; ovarian, OV-Pt = 79; cervical, CERV-Pt = 307; uterine, UTCS-Pt = 57); Group 3 (lung adenocarcinoma, LUAD = 450; lung squamous cell carcinoma, LUSC = 372; head & neck, HNSC-Pt = 528); Group 4 (prostate, PROS-Pt = 192; bladder, BLAD-Pt = 412); and Group 5 (liver, LIV-Pt = 377; pancreatic, PANC-Pt = 184; and gall bladder, BILE-Pt = 36). The columns reflect the total percent patients positive for each of the markers divided by the total number of markers used – for the first row of all cancers, that would be 96 markers. Thus, on average, of the 96 markers chosen, the number of average sensitivity scores are: Group 1 (colorectal = 44, stomach = 45, esophagus = 40); Group 2 (breast = 38, endometrial = 40, ovarian = 22, cervical = 39, uterine = 33); Group 3 (lung adenocarcinoma = 31, lung squamous cell carcinoma = 31, head & neck = 33); Group 4 (prostate = 45, bladder = 36); and Group 5 (liver = 38, pancreatic = 27, gall bladder = 47). This translates into the following number of marker equivalents with average sensitivities of 50% (= 96 x score/50); (colorectal = 85 marker equivalents; stomach = 86 marker equivalents; esophagus = 78 marker equivalents); Group 2 (breast = 74 marker equivalents; endometrial = 76 marker equivalents; ovarian = 42 marker equivalents; cervical = 75 marker equivalents; uterine = 64 marker equivalents); Group 3 (lung adenocarcinoma = 60 marker equivalents; lung squamous cell carcinoma = 59 marker equivalents; head & neck = 64 marker equivalents); Group 4 (prostate = 86 marker equivalents; bladder = 70 marker equivalents); and Group 5 (liver = 74 marker equivalents; pancreatic = 51 marker equivalents; gall bladder = 91 marker equivalents). Thus, cancers were well represented, ranging from 42 to 91 marker equivalents for the different cancer types, and all well above the minimum of 36 markers with average sensitivities of 50%.

[0345] The above numbers translate into the following number of marker equivalents with average sensitivities of 66% (= 96 x score/66); (colorectal = 65 marker equivalents; stomach = 65 marker equivalents; esophagus = 59 marker equivalents); Group 2 (breast = 56 marker equivalents; endometrial = 58 marker equivalents; ovarian = 32 marker equivalents; cervical = 57 marker equivalents; uterine = 48 marker equivalents); Group 3 (lung adenocarcinoma = 45 marker equivalents; lung squamous cell carcinoma = 45 marker equivalents; head & neck = 48 marker equivalents); Group 4 (prostate = 65 marker equivalents; bladder = 53 marker equivalents); and Group 5 (liver = 56 marker equivalents; pancreatic = 39 marker equivalents; gall bladder = 69 marker equivalents). Thus, cancers were well represented, ranging from 32 to 69 marker equivalents for the different cancer types, and with the exception of ovarian cancer at 32, the other cancer types are above the minimum of 36 markers with average sensitivities of 66%.

[0346] The aforementioned markers were then re-ordered for each of the above cancer types such that the most prevalent markers were listed first. For example, with CRC, of the 96 markers, 54 markers gave scores above 55 (*i.e.* were positive in greater than 55% of the 395 patients) and 9 gave scores of between 25 and 54 (*i.e.* were positive for from 25% to 54% of the 395 patients). Half of the higher, and a third of the lower set, for a total of 30 markers were distributed into two marker test sets, designated "CRC1" and "CRC2" (Table 47, rows 2 & 3). These marker sets would reflect an ideal result if half the markers with the potential to be positive are detected in the assay. This does not account for the chances that earlier stage tumors would have a lower number of marker molecules in the plasma, and thus consequently the actual number of markers positive would be less than the ideal result in this simulation. The percent of patients positive for each of the cancers were recorded and then divided by the total number of markers used for that cancer type. As anticipated, when selecting markers for a given tumor type, those markers should give a higher score than the average, *i.e.* 66 for CRC in each of the two sets of selected 30 markers, compared with a score of 44 for the unselected 96 markers. These markers form a diagonal across Table 47 and are highlighted in bold and light grey background.

[0347] For each column, marker sets that are in the same range or higher than the number of positive markers for that cancer type are also shown with a light grey background. For example, a patient with colorectal, stomach, or esophageal cancer will be scored as potentially positive with stomach cancer. This makes sense as the markers for these three cancers ovelap (*i.e.*, they all bin to Group 1). They could be distinguished in step 2 of the assay on the group 1 markers, where these markers are more cancer types specific and tease out the most probable cancer type. Evaluation of the ST-Pt column shows that simulations for one of the two LUAD,

BLAD, and both PANC also gave scores that might be interpreted as stomach cancer. Thus, the first step is not always able to pinpoint what Groups should be tested in the second step of the assay. However, most of the ambiguity is within group members (*i.e.* Group 2), and this makes sense, since the markers were chosen to maximize the ability to chose which groups to test in the second step.

[0348] Tables 48 and 49 (see prophetic experimental section) takes the aforementioned results in the simulations in Table 47 and multiplies them by the percent incidence of the given cancer type for that gender (see tables 37 and 38 respectively), and the result is adjusted to the same order of magnitude (multiple by 10). The concept is for the physician to take into account that a lower score for a high incidence cancer (such as CRC) may be a more common tissue of origin for a higher score for a low incidence cancer (such as lung squamous cell carcinoma). Tables 48 and 49 show the level of ambiguity in identifying tissue of origin is higher among female patients than among male patients, as indicated by the number of cells highlighted in grey that are not on the diagonal. In all cases, the physician will need to incorporate all data, such as smoking history, and not just molecular data to determine the most likely tissue of origin before sending the patient to confirmatory imaging.

[0349] Tables 50, 51, and 52 (see prophetic experimental section) takes the aforementioned results in the simulations in Tables 47, 48, and 49 and determines the percent deviation from the neutral result by taking the percentage of (= score specific cancer type simulation /score all cancer for that type – 1). Thus, the first row of each of these tables should be 0%. Again, those percentages that are higher than, or in the same range as, the percentages across the diagonal are highlighted in light gray. While this set of marker selection may be less than ideal for distinguishing esophageal or gall bladder cancers as the tissue of origin, they are nevertheless quite informative for guiding the physician to which groups of the Step 2 assays should be tested. This simple scoring may be augmented by using AI approaches based on a database of results with clinical samples using the aforementioned 96-marker set.

[0350] For the second step of the assay, one, two, or more of the following groups will be tested, each group with a set of 64 markers that on average comprise of at least 36 markers with 50% sensitivity that covers each of the aforementioned 16 types of solid tumors in the following groups: Group 1 (colorectal, stomach, and esophagus); Group 2 (breast, endometrial, ovarian, cervical, and uterine); Group 3 (lung and head & neck); Group 4 (prostate and bladder); and Group 5 (liver, pancreatic, or gall bladder). These Group-specific and cancer type-specific markers include, but are not limited to, cancer-specific microRNA markers, cancer-specific ncRNA and lncRNA markers, cancer-specific exon transcripts, tumor-associated antigens, cancer protein markers, protein markers that can be secreted by solid tumors into the blood, common

mutations, primary CpG sites that are solid tumor and tissue specific markers, chromosomal regions or sub-regions within which are primary CpG sites that are solid tumor and tissue specific markers, and primary and flanking CpG sites that are solid tumor and tissue specific markers. Methods for detecting said markers have been discussed *supra*, and listing of these markers are described for each of the groups below as well as in the corresponding figures.

[0351] Group 1 (colorectal, stomach, and esophagus): Blood-based, colorectal, stomach, and esophageal cancer-specific microRNA markers that may be used to distinguish group 1 from other groups include, but are not limited to: (mir ID, Gene ID): hsa-mir-624, MIR624. This miRNA was identified through analysis of TCGA microRNA datasets, and may be present in exosomes, tumor-associated vesicles, Argonaute complexes, or other protected states in the blood.

[0352] Blood-based, colorectal, stomach, and esophageal cancer-specific ncRNA and lncRNA markers that may be used to distinguish group 1 from other groups include, but are not limited to: [Gene ID, Coordinate (GRCh38)], ENSEMBL ID: LINC01558, chr6:167784537-167796859, ENSG00000146521.8. This ncRNA was identified through comparative analysis of various publicly available Affymetrix Exon ST CEL data, which were aligned to GENCODE annotations to generate ncRNA and lncRNA transcriptome datasets. Such lncRNA and ncRNA may be enriched in exosomes or other protected states in the blood.

[0353] In addition, Figure 58 provides a list of blood-based colorectal, stomach, and esophageal cancer-specific exon transcripts that may be enriched in exosomes, tumor-associated vesicles, or other protected states in the blood.

[0354] Colorectal, stomach, and esophageal cancer protein encoding markers that may be used to distinguish group 1 from other groups include, but are not limited to: (Gene Symbol , Chromosome Band, Gene Title, UniProt ID): SELE, 1q22-q25, selectin E, P16581; OTUD4, 4q31.21, OTU domain containing 4, Q01804; BPI, 20q11.23, bactericidal/permeability-increasing protein, P17213; ASB4, 7q21-q22, ankyrin repeat and SOCS box containing 4, Q9Y574; C6orf123, 6q27, chromosome 6 open reading frame 123, Q9Y6Z2; KPNA3, 13q14.3, karyopherin alpha 3 (importin alpha 4), O00505; NUP98, 11p15, nucleoporin 98kDa , P52948, identified through mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from colorectal, stomach, and esophageal cancers, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0355] Protein markers that can be secreted by colorectal, stomach, and esophageal cancer into the blood, and may be used to distinguish group 1 from other groups include, but are not limited to: (Protein name, UniProt ID); Bactericidal permeability-increasing protein (BPI)

(CAP 57), P1721. A comparative analysis was performed across various TCGA datasets (tumors, normals), followed by an additional bioinformatics filter (Meinken et al., "Computational Prediction of Protein Subcellular Locations in Eukaryotes: an Experience Report," *Computational Molecular Biology* 2(1):1-7 (2012), which is hereby incorporated by reference in its entirety), which predicts the likelihood that the translated protein is secreted by the cells.

[0356] The distribution of mutations in colorectal, stomach, and esophageal cancer are available in the public COSMIC database, with the most common being: APC (APC regulator of WNT signaling pathway), ATM (ATM serine/threonine kinase), CSMD1 (CUB and Sushi multiple domains 1), DNAH11 (dynein axonemal heavy chain 11), DST (dystonin), EP400 (E1A binding protein p400), FAT3 (FAT atypical cadherin 3), FAT4 (FAT atypical cadherin 4), FLG (filaggrin), GLI3 (GLI family zinc finger 3), KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog), LRP1B (LDL receptor related protein 1B), MUC16 (mucin 16, cell surface associated), OBSCN (obscurin, cytoskeletal calmodulin and titin-interacting RhoGEF), PCLO (piccolo presynaptic cytomatrix protein), PIK3CA (phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha), RYR2 (ryanodine receptor 2), SYNE1 (spectrin repeat containing nuclear envelope protein 1), TP53 (tumor protein p53), TTN (titin), and UNC13C (unc-13 homolog C).

[0357] Figure 59 provides a list of primary CpG sites that are colorectal, stomach, and esophageal cancer and tissue-specific markers, that may be used to identify the presence of colorectal, stomach, and esophageal cancer from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. Figure 60 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are colorectal, stomach, and esophageal cancer and tissue-specific markers, that may be used to identify the presence of colorectal, stomach, and esophageal cancer from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. These lists contain preferred primary CpG sites and their flanking sites, as well as alternative markers that are high in CRC, and alternative markers that are low to no-CRC, but high in stomach and/or esophageal cancers. Primer sets for exemplary preferred and alternate methylation markers are listed in Table 53 in the experimental section. A selection of 64 of these markers with average sensitivities of 50% gave the following scores for Group 1: (colorectal = 48, stomach = 51, esophagus = 43), which would translate into the following number of marker equivalents with average sensitivities of 50% (= 64 x score/50); (colorectal = 62 marker equivalents; stomach = 65 marker equivalents; esophagus = 55 marker equivalents) and thus all were well above the average 36-marker equivalents minimum. The marker equivalents with average sensitivities of

66% gave the following scores (= 64 x score/66); (colorectal = 47 marker equivalents; stomach = 50 marker equivalents; esophagus = 42 marker equivalents). Thus, all were well above the average 36-marker equivalents minimum.

[0358] Group 2 (breast, endometrial, ovarian, cervical, and uterine): Blood-based, breast, endometrial, ovarian, cervical, and uterine cancer-specific microRNA markers that may be used to distinguish group 2 from other groups include, but are not limited to: (mir ID, Gene ID): hsa-mir-1265, MIR1265. This marker was identified through analysis of TCGA microRNA datasets, which may be present in exosomes, tumor-associated vesicles, Argonaute complexes, or other protected states in the blood.

[0359] Blood-based breast, endometrial, ovarian, cervical, and uterine cancer-specific exon transcripts that may be used to distinguish group 2 from other groups include, but are not limited to: (mir ID, Gene ID): hsa-mir-1265, MIR1265. This marker was identified through analysis of TCGA microRNA datasets, which may be present in exosomes, tumor-associated vesicles, Argonaute complexes, or other protected states in the blood.

[0360] Breast, endometrial, ovarian, cervical, and uterine cancer protein markers that may be used to distinguish group 2 from other groups include, but are not limited to: (Gene Symbol, Chromosome Band, Gene Title, UniProt ID): RSPO2, 8q23.1, R-spondin 2, Q6UXX9; KLC4 , 6p21.1, kinesin light chain 4, Q9NSK0; GLRX, 5q14, glutaredoxin (thioltransferase), P35754. These markers may be identified through mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from breast, endometrial, ovarian, cervical, and uterine cancers, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0361] Protein markers that can be secreted by breast, endometrial, ovarian, cervical, and uterine cancer into the blood that may be used to distinguish group 2 from other groups include, but are not limited to: (Protein name, UniProt ID); R-spondin-2 (Roof plate-specific spondin-2) (hRspo2), Q6UXX9. A comparative analysis was performed across various TCGA datasets (tumors, normals), followed by an additional bioinformatics filter (Meinken et al., 2012, as described above), which predicts the likelihood that the translated protein is secreted by the cells.

[0362] The distribution of mutations in breast, endometrial, ovarian, cervical, and uterine cancer are available in the public COSMIC database, with the most common being: PIK3CA (phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha), and TTN (titin).

[0363] Figure 61 provides a list of primary CpG sites that are breast, endometrial, ovarian, cervical, and uterine cancer and tissue-specific markers, that may be used to identify the presence of breast, endometrial, ovarian, cervical, and uterine cancer from cfDNA, DNA within

exosomes, or DNA in other protected states (such as within CTCs) within the blood. Figure 62 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are breast, endometrial, ovarian, cervical, and uterine cancer and tissue-specific markers, that may be used to identify the presence of breast, endometrial, ovarian, cervical, and uterine cancer from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. These lists contain preferred primary CpG sites and their flanking sites, as well as alternative markers that may be used to distinguish breast, endometrial, ovarian, cervical, and uterine cancers. Primer sets for exemplary preferred and alternate methylation markers are listed in Table 54 in the experimental section. A selection of 64 of these markers with average sensitivities of 50% gave the following scores for Group 2: (breast = 36, endometrial = 49, ovarian = 32, cervical = 33, uterine = 47), which would translate into the following number of marker equivalents with average sensitivities of 50% (= 64 x score/50); (breast = 47 marker equivalents; endometrial = 63 marker equivalents; ovarian = 41 marker equivalents; cervical = 42 marker equivalents; uterine = 61 marker equivalents). Thus, all were well above the average 36-marker equivalents minimum. The marker equivalents with average sensitivities of 66% gave the following scores (= 64 x score/66); (breast = 35 marker equivalents; endometrial = 48 marker equivalents; ovarian = 31 marker equivalents; cervical = 32 marker equivalents; uterine = 46 marker equivalents). Thus three markers are below and two markers are above the average 36-marker equivalents minimum. However, such scores may be improved by selection of different markers.

[0364] Group 3 (lung adenocarcinoma, lung squamous cell carcinoma, and head & neck): Blood-based, lung, head, and neck cancer-specific microRNA markers that may be used to distinguish group 3 from other groups include, but are not limited to: (mir ID, Gene ID): hsa-mir-28, MIR28. This marker was identified through analysis of TCGA microRNA datasets, and may be present in exosomes, tumor-associated vesicles, Argonaute complexes, or other protected states in the blood.

[0365] Blood-based lung, head, and neck cancer-specific exon transcripts that may be used to distinguish group 3 from other groups include, but are not limited to: (Exon location, Gene); chr2: chr1:93307721-93309752:-, FAM69A; chr1:93312740-93312916:-, FAM69A; chr1:93316405-93316512:-, FAM69A; chr1:93341853-93342152:-, FAM69A; chr1:93426933-93427079:-, FAM69A; chr7:40221554-40221627:+, C7orf10; chr7:40234539-40234659:+, C7orf10;chr8:22265823-22266009:+, SLC39A14; chr8:22272293-22272415:+, SLC39A14; chr14:39509936-39510091:-, SEC23A; chr14:39511990-39512076:-, SEC23A, and may be enriched in exosomes, tumor-associated vesicles, or other protected states in the blood.

[00125] Lung, head, and neck cancer protein encoding markers that may be used to distinguish group 3 from other groups include, but are not limited to: (Gene Symbol, Chromosome Band, Gene Title, UniProt ID): STRN3, 14q13-q21, striatin, calmodulin binding protein 3, Q13033; LRRC17, 7q22.1, leucine rich repeat containing 17, Q8N6Y2; FAM69A, 1p22, family with sequence similarity 69, member A, Q5T7M9; ATF2 , 2q32, activating transcription factor 2, P15336; BHMT, 5q14.1, betaine--homocysteine S-methyltransferase, Q93088; ODZ3/TENM3, 4q34.3-q35.1, teneurin transmembrane protein 3, Q9P273; ZFHX4, 8q21.11, zinc finger homeobox 4, Q86UP3. These markers may be identified through mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from lung, head, and neck cancers, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0366] Protein markers that can be secreted by lung, head, and neck cancer into the blood may be used to distinguish group 3 from other groups include, but are not limited to: (Protein name, UniProt ID); Leucine-rich repeat-containing protein 17 (p37NB), Q8N6Y2. A comparative analysis was performed across various TCGA datasets (tumors, normals), followed by an additional bioinformatics filter (Meinken et al., "Computational Prediction of Protein Subcellular Locations in Eukaryotes: an Experience Report," *Computational Molecular Biology* 2(1):1-7 (2012), which is hereby incorporated by reference in its entirety), which predicts the likelihood that the translated protein is secreted by the cells.

[0367] The distribution of mutations in lung, head, and neck cancer are available in the public COSMIC database, with the common being: CSMD3 (CUB and Sushi multiple domains 3), DNAH5 (dynein axonemal heavy chain 5), FAT1 (FAT atypical cadherin 1), FLG (filaggrin), KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog), LRP1B (LDL receptor related protein 1B), MUC16 (mucin 16, cell surface associated), PCLO (piccolo presynaptic cytomatrix protein), PKHD1L1 (PKHD1 like 1), RELN (reelin), RYR2 (ryanodine receptor 2), SI (sucrase-isomaltase), SYNE1 (spectrin repeat containing nuclear envelope protein 1), TP53 (tumor protein p53), TTN (titin), USH2A (usherin), and XIRP2 (xin actin binding repeat containing 2).

[0368] Figure 63 provides a list of primary CpG sites that are lung, head, and neck cancer and tissue-specific markers, that may be used to identify the presence of lung, head, and neck cancer from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. Figure 64 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are lung, head, and neck cancer and tissue-specific markers, that may be used to identify the presence of lung, head, and neck from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. These lists

contain preferred primary CpG sites and their flanking sites that may be used to distinguish lung, head, and neck cancers. Primer sets for exemplary methylation markers are listed in Table 55 in the experimental section. A selection of 64 of these markers with average sensitivities of 50% gave the following scores for Group 3: (lung adenocarcinoma = 41, lung squamous cell carcinoma = 49, head & neck = 53), which would translate into the following number of marker equivalents with average sensitivities of 50% (= 64 x score/50); (lung adenocarcinoma = 52 marker equivalents; lung squamous cell carcinoma = 62 marker equivalents; head & neck = 67 marker equivalents). Thus, all were well above the average 36-marker equivalents minimum. The marker equivalents with average sensitivities of 66% gave the following scores (= 64 x score/66); (lung adenocarcinoma = 40 marker equivalents; lung squamous cell carcinoma = 47 marker equivalents; head & neck = 51 marker equivalents). Thus all were well above the average 36-marker equivalents minimum.

[0369] Group 4 (prostate and bladder): Blood or urine-based, prostate and bladder cancer-specific microRNA markers may be used to distinguish group 4 from other groups include, but are not limited to: (mir ID, Gene ID): hsa-mir-491, MIR491; hsa-mir-1468, MIR1468. These markers were identified through analysis of TCGA microRNA datasets, and may be present in exosomes, tumor-associated vesicles, Argonaute complexes, or other protected states in the blood or urine.

[0370] Blood or urine-based, prostate and bladder cancer-specific ncRNA and lncRNA markers that may be used to distinguish group 4 from other groups include, but are not limited to: [Gene ID, Coordinate (GRCh38), ENSEMBL ID]: AC007383.3, chr2:206084605-206086564, ENSG00000227946.1; LINC00324, chr17:8220642-8224043, ENSG00000178977.3. These markers were identified through comparative analysis of various publicly available Affymetrix Exon ST CEL data, which were aligned to GENCODE annotations to generate ncRNA and lncRNA transcriptome datasets. Such lncRNA and ncRNA may be enriched in exosomes or other protected states in the blood or urine.

[0371] Blood or urine-based prostate and bladder cancer-specific exon transcripts that may may be used to distinguish group 4 from other groups include, but are not limited to: (Exon location, Gene); chr21:45555942-45556055:+ , C21orf33 and may be enriched in exosomes, tumor-associated vesicles, or other protected states in the blood or urine.

[0372] Prostate and bladder cancer protein markers that may be used to distinguish group 4 from other groups include, but are not limited to: (Gene Symbol, Chromosome Band, Gene Title, UniProt ID): PMM1, 22q13, phosphomannomutase 1, Q92871. This marker may be identified through mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from lung, head, and neck cancers,

which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma, or within the urine.

[0373] The distribution of mutations in prostate and bladder cancer are available in the public COSMIC database, with the most common being BAGE2 (BAGE family member 2), DNM1P47 (dynamin 1 pseudogene 47), FRG1BP (region gene 1 family member B, pseudogene), KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog), RP11-156P1.3, TTN (titin), and TUBB8P7 (tubulin beta 8 class VIII pseudogene 7).

[0374] Figure 65 provides a list of primary CpG sites that are prostate and bladder cancer-specific markers, that may be used to identify the presence of prostate and bladder cancer from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood or urine. Figure 66 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are prostate and bladder cancer specific markers, that may be used to identify the presence of prostate and bladder from cfDNA, or DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood or urine. These lists contain preferred primary CpG sites and their flanking sites that may be used to distinguish prostate and bladder cancers. Primer sets for exemplary methylation markers are listed in Table 56 in the experimental section. A selection of 48 of these markers with average sensitivities of 50% gave the following scores for Group 4: (prostate = 48, bladder = 22), which would translate into the following number of marker equivalents with average sensitivities of 50% (= 48 x score/50); (prostate = 46 marker equivalents; bladder = 21 marker equivalents). Thus, bladder was below the average 36-marker equivalents minimum. Likewise, the marker equivalents with average sensitivities of 66% gave the following scores (= 48 x score/60); (prostate = 35 marker equivalents; bladder = 16 marker equivalents). Thus, bladder was well below the average 36-marker equivalents minimum. However, a different selection of markers, for example by increasing from 48 to 64 markers and including markers that were positive for both prostate and bladder, would rectify this situation. The markers were limited to those that were not methylated in normal prostate, bladder, or kidney tissue to minimize false-positive results from urine samples.

[0375] Group 5 (liver, pancreatic and gall-bladder): Blood-based, liver, pancreatic and gall-bladder cancer-specific microRNA markers that may be used to distinguish group 5 from other groups include, but are not limited to: (mir ID, Gene ID): hsa-mir-132, MIR132. This marker was identified through analysis of TCGA microRNA datasets, which may be present in exosomes, tumor-associated vesicles, Argonaute complexes, or other protected states in the blood.

[0376] Figure 67 provides a list of blood-based, liver, pancreatic and gall-bladder cancer-specific ncRNA and lncRNA markers derived through comparative analysis of various publicly available Affymetrix Exon ST CEL data, which were aligned to GENCODE annotations to generate ncRNA and lncRNA transcriptome datasets. Such lncRNA and ncRNA may be enriched in exosomes or other protected states in the blood.

[0377] In addition, Figure 68 provides a list of blood-based liver, pancreatic and gall-bladder cancer-specific exon transcripts that may be enriched in exosomes, tumor-associated vesicles, or other protected states in the blood.

[0378] Figure 69 provides a list of liver, pancreatic and gall-bladder cancer protein markers, identified through mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product arising from liver, pancreatic and gall-bladder cancers, which may be identified in the blood, either within exosomes, other protected states, tumor-associated vesicles, or free within the plasma.

[0379] Protein markers that can be secreted by liver, pancreatic and gall-bladder cancer into the blood that may be used to distinguish group 5 from other groups include, but are not limited to: (Protein name, UniProt ID); Gelsolin (AGEL) (Actin-depolymerizing factor) (ADF) (Brevin), P06396; Pro-neuregulin-2, O14511; CD59 glycoprotein (1F5 antigen) (20 kDa homologous restriction factor) (HRF-20) (HRF20) (MAC-inhibitory protein) (MAC-IP) (MEM43 antigen) (Membrane attack complex inhibition factor) (MACIF) (Membrane inhibitor of reactive lysis) (MIRL) (Protectin) (CD antigen CD59), P13987; Divergent protein kinase domain 2B (Deleted in autism-related protein 1), Q9H7Y0. A comparative analysis was performed across various TCGA datasets (tumors, normals), followed by an additional bioinformatics filter (Meinken et al., "Computational Prediction of Protein Subcellular Locations in Eukaryotes: an Experience Report," *Computational Molecular Biology* 2(1):1-7 (2012), which is hereby incorporated by reference in its entirety), which predicts the likelihood that the translated protein is secreted by the cells.

[0380] The distribution of mutations in liver, pancreatic and gall-bladder cancer are available in the public COSMIC database, with the most common being: KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog), MUC16 (mucin 16, cell surface associated), MUC4 (mucin 4, cell surface associated), TP53 (tumor protein p53), and TTN (titin).

[0381] Figure 70 provides a list of primary CpG sites that are liver, pancreatic and gall-bladder cancer and tissue-specific markers, that may be used to identify the presence of lung, head, and neck cancer from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. Figure 71 provides a list of chromosomal regions or sub-regions within which are primary CpG sites that are liver, pancreatic and gall-bladder cancer

and tissue-specific markers, that may be used to identify the presence of liver, pancreatic and gall-bladder from cfDNA, DNA within exosomes, or DNA in other protected states (such as within CTCs) within the blood. These lists contain preferred primary CpG sites and their flanking sites, as well as alternative markers that may be used to distinguish liver, pancreatic and gall-bladder cancers. Primer sets for exemplary prefered and alternate methylation markers are listed in Table 57 in the experimental section. A selection of 64 of these markers with average sensitivities of 50% gave the following scores for Group 5: (liver = 57, pancreatic = 30, gall bladder = 60), which would translate into the following number of marker equivalents with average sensitivities of 50% (= 64 x score/50); (liver = 73 marker equivalents; pancreatic = 38 marker equivalents; gall bladder = 77 marker equivalents). Thus, all were above the average 36-marker equivalents minimum. The marker equivalents with average sensitivities of 66% gave the following scores (= 64 x score/66); (liver = 56 marker equivalents; pancreatic = 29 marker equivalents; gall bladder = 58 marker equivalents). Thus, liver and gall bladder were above the average 36-marker equivalents minimum, while pancreatic was below.

[0382] Consider the first strategy using the 96 pan-oncology markers of detecting early colorectal cancer (Figure 1C). The calculations are done with the anticipation of an average of 150 methylated molecules per positive marker in the blood. As described *supra*, for the example of colorectal cancer, in particular the cases of microsatellite stable tumors (MSS) where the mutation load is low, for these calculations when relying on NGS sequencing alone (assuming 150 molecules with one mutation in the blood), an estimated 78% of early colorectal cancer would be missed. Again, to put these number in perspective, in the U.S., about 135,000 new cases of colorectal cancer were diagnosed in 2018, of which about 60% is late cancer (*i.e.* Stage III & IV). About 107 million individuals in the U.S. are over the age of 50 and should be tested for colorectal cancer. While it cannot be predicted how many individuals have a hidden cancer (*i.e.* Stage I) within them, who are non-compliant to testing, for the purposes of this calculation, assume that the average late cancer was once the average early cancer, and thus individuals with Stage I cancer would be about 40,500 individuals. Assuming individual marker false-positive rates of 3%, and with the first step using 96 markers (48 markers for CRC) with average sensitivities of 50%, requiring a minimum of 5 markers positive, then with an overall specificity of 95.8%, the first step would identify 4,494,000 individuals (out of 107,000,000 total adults over 50 in the U.S.) which would include at 71.6% sensitivity or about 28,998 individuals with Stage I colorectal cancer (out of 40,500 individuals with Stage I cancer). However, those 4,494,000 presumptive positive individuals would be evaluated in a second step of 64 markers (48 markers for CRC) with average sensitivities of 50%, requiring a minimum of 5 markers positive, then the two-step test would identify 71.6% x 71.6% = 51.2% = 20,762 individuals (out

of 40,500 individuals with Stage I cancer) with colorectal cancer. With a specificity of 95.8%, the second test would also generate 4,494,000 x 4.2% = 188,748 false-positives. The positive predictive value of such a test would be 20,762 / (188,748 + 20,762) = 9.9%, in other words, 1 in 10 individuals who tested positive would actually have Stage I colorectal cancer. In reality, one would need to also include the success for identifying Stage 2 and higher cancers. In expanding this example, the calculations are done with the anticipation that Stage I CRC has an average of 150 methylated molecules per positive marker in the blood, Stage II CRC has an average of 200 methylated molecules per positive marker, and the higher stages (III & IV) have at least an average of 300 methylated molecules per positive marker, and the higher stages. Also, to be consistent with the idea that as the test is used repeatedly, more of early and less of late CRC will be detected, than an estimate of 40,500 individuals with Stage I cancer, 40,500 individuals with Stage II cancer, and the remaining 54,000 individuals have late-stage cancer = 135,000 total individuals with colorectal cancer identified per year in the U.S. The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 90.1% would be identified in the first step, of which 90.1% x 90.1% = 81.0% = 32,877 individuals with Stage II cancer would be verified in the second step. For Stage III and IV cancer, 99.3 % would be identified in the first step, of which 99.3% x 99.3% = 98.6% = 53,246 individuals with late cancer would be identified. This brings the total identified at 20,762 + 32,877 + 53,246 = 106,885 individuals out of 135,000 with colorectal cancer, for an overall sensitivity of 79%. Overall, the positive predictive value of such a test would be 106,885/(188,748 + 106,885) = 36.1%, in other words, 1 in 3 individuals who tested positive would actually have colorectal cancer, and this test would identify 53,639 / 81,000 or 66% of those individuals with early cancer, compared with the current rate of 40%.

[0383] How would these results vary for using this strategy (Figure 1C) for detection of early colorectal cancer using 50% average marker sensitivities, with the anticipation of Stage I CRC having an average of 200 methylated molecules per positive marker in the blood, Stage II CRC having an average of 240 methylated molecules per positive marker, and the higher stages (III & IV) having at least an average of 300 methylated molecules per positive marker?

[0384] Assuming individual marker false-positive rates of 3%, the first step using 96 markers (48 markers for CRC) with average sensitivities of 50%, requiring a minimum of 5 markers positive, and an overall specificity of 95.8%, the first step would identify 4,494,000 individuals (out of 107,000,000 total adults over 50 in the U.S.). This would include, at 90.1% sensitivity, or about 36,490 individuals with Stage I colorectal cancer (out of 40,500 individuals with Stage I cancer). However, those 4,494,000 presumptive positive individuals would be evaluated in a second step of 64 markers (48 markers for CRC) with average sensitivities of

50%, requiring a minimum of 5 markers positive. The two-step test would identify 90.1% x 90.1% = 81.2% = 32,877 individuals (out of 40,500 individuals with Stage I cancer) with colorectal cancer. With a specificity of 95.8%, the second test would also generate 4,494,000 x 4.2% = 188,748 false-positives. The positive predictive value of such a test would be 32,877 / (188,748 + 32,877) = 14.8%. In other words, 1 in 6.7 individuals who tested positive would actually have Stage I colorectal cancer. In reality, one would need to also include the success for identifying Stage 2 and higher cancers. To be consistent with the idea that, as the test is used repeatedly, more of early and less of late CRC will be detected, an estimated 40,500 individuals with Stage I cancer, 40,500 individuals with Stage II cancer, and 54,000 individuals with late-stage cancer (135,000 total individuals with colorectal cancer) would be identified per year in the U.S. The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 90.1% would be identified in the first step, of which 97.2% x 97.2% = 94.5% = 38,263 individuals with Stage II cancer would be verified in the second step. For Stage III and IV cancer, 99.3 % would be identified in the first step, of which 99.3% x 99.3% = 98.6% = 53,246 individuals with late cancer would be identified. This brings the total identified to 32,877 + 38,263 + 53,246 = 124,386 individuals out of 135,000 with colorectal cancer, for an overall sensitivity of 92.1%. Overall, the positive predictive value of such a test would be 124,386 /(188,748 + 124,386) = 39.7%. In other words, 1 in 2.5 individuals who tested positive would actually have colorectal cancer, and this test would identify 71,104 / 81,000 or 87.7% of those individuals with early cancer, compared with the current rate of 40%.

[0385] How would these results vary when using this strategy (Figure 1C) for detection of early ovarian cancer, with the anticipation of an average of 150 methylated molecules per positive marker in the blood? When relying on NGS sequencing alone (assuming 150 molecules with one mutation in the blood), an estimated 78% of early ovarian cancer would be missed. Again, to put these number in perspective, in the U.S., about 22,000 new cases of ovarian cancer were diagnosed in 2018, of which about 85% was late cancer (*i.e.* Stage III & IV). About 54 million women in the U.S. are between the ages of 50 and 79 and should be tested for ovarian cancer. While it cannot be predicted how many individuals have a hidden cancer (*i.e.* Stage I), for the purposes of this calculation, assume that the stages are evenly divided. Thus, the number of individuals with Stage I ovarian cancer would be about 5,500 individuals. Assuming individual marker false-positive rates of 3%, the first step using 96 markers (36 markers for ovarian) with average sensitivities of 50%, requiring a minimum of 5 markers positive, and an overall specificity of 99.1%, the first step would identify 486,000 individuals (out of 54,000,000 total women ages 50-79 in the U.S.) with ovarian cancer. This would include, at 46.8% sensitivity, or about 2,574 individuals with Stage I ovarian cancer (out of 5,500 individuals with

Stage I ovarian cancer). However, those 486,000 presumptive positive individuals would be evaluated in a second step of 64 markers (36 markers for ovarian cancer) with average sensitivities of 50%, requiring a minimum of 5 markers positive. The two-step test would identify 46.8% x 46.8% = 21.9% = 1,204 individuals (out of 5,500 individuals with Stage I ovarian cancer) with ovarian cancer. With a specificity of 99.1%, the second test would also generate 486,000 x 0.9% = 4,374 false-positives. The positive predictive value of such a test would be 1,204 / (4,374 + 1,204) = 21.6%. In other words, 1 in 4.6 individuals who tested positive would actually have Stage I ovarian cancer. In reality, one would need to also include the success for identifying Stage 2 and higher ovarian cancers. In expanding this example, the calculations are done with the anticipation that Stage I ovarian cancer has an average of 150 methylated molecules per positive marker in the blood, Stage II ovarian cancer has an average of 200 methylated molecules per positive marker, and the higher stages (III & IV) have at least an average of 300 methylated molecules per positive marker. To be consistent with the idea that, as the test is used repeatedly, more cancer will be detected and all four stages are at 5,500, then 5,500 x 4 = 22,000 total individuals with ovarian cancer would be identified per year in the U.S. The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 71.5% would be identified in the first step, of which 71.5% x 71.5% = 51.1% = 2,810 individuals with Stage II ovarian cancer would be verified in the second step. For Stage III and IV ovarian cancer, 94.5% would be identified in the first step, of which 94.5% x 94.5% = 89.3% = 9,823 individuals with late ovarian cancer would be identified. This brings the total identified at 1,204 + 2,810 + 9,823 = 13,837 individuals out of 22,000 with ovarian cancer, for an overall sensitivity of 62.9%. Overall, the positive predictive value of such a test would be 13,837 /(13,837 + 4,374) = 76.0%. In other words, 3 in 4 women who tested positive would actually have ovarian cancer, and this test would identify 4,014 / 11,000, or 36.5%, of those individuals with early cancer, compared with the current rate of 15%.

[0386] How would these results vary for using this strategy (Figure 1C) for detection of early ovarian cancer using 50% average marker sensitivities, with the anticipation that Stage I ovarian cancer has an average of 200 methylated molecules per positive marker in the blood, Stage II ovarian cancer has an average of 240 methylated molecules per positive marker, and the higher stages (III & IV) have at least an average of 300 methylated molecules per positive marker?

[0387] Assuming individual marker false-positive rates of 3%, the first step using 96 markers (36 markers for ovarian) with average sensitivities of 50%, and requiring a minimum of 5 markers positive, with an overall specificity of 99.1%, the first step would identify 486,000 individuals (out of 54,000,000 total women ages 50-79 in the U.S.) with ovarian cancer. This

would include at, 71.5% sensitivity, about 3,932 individuals with Stage I ovarian cancer (out of 5,500 individuals with Stage I ovarian cancer). However, those 486,000 presumptive positive individuals would be evaluated in a second step of 64 markers (36 markers for ovarian cancer) with average sensitivities of 50%, requiring a minimum of 5 markers positive. The two-step test would identify 71.5% x 71.5% = 51.1% = 2,810 individuals (out of 5,500 individuals with Stage I ovarian cancer) with ovarian cancer. With a specificity of 99.1%, the second test would also generate 486,000 x 0.9% = 4,374 false-positives. The positive predictive value of such a test would be 2,810 / (4,374 + 2,810) = 39.1%. In other words, 1 in 2.5 individuals who tested positive would actually have Stage I ovarian cancer. In reality, one would need to also include the success for identifying Stage 2 and higher ovarian cancers. As the test is used repeatedly, assume all four stages are at 5,500, and, therefore, 5,500 x 4 = 22,000 total individuals with ovarian cancer would be identified per year in the U.S. The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 84.4% would be identified in the first step, of which 84.4% x 84.4% = 71.2% = 3,916 individuals with Stage II ovarian cancer would be verified in the second step. For Stage III and IV ovarian cancer, 94.5% would be identified in the first step, of which 94.5% x 94.5% = 89.3% = 9,823 individuals with late ovarian cancer would be identified. This brings the total identified to 2,810 + 3,916 + 9,823 = 16,549 individuals out of 22,000 with ovarian cancer, for an overall sensitivity of 75.2%. Overall, the positive predictive value of such a test would be 16,549 /(16,549 + 4,374) = 79.0%. In other words, 4 in 5 women who tested positive would actually have ovarian cancer. This test would identify 6,006 / 11,000 or 54.6% of those individuals with early cancer, compared with the current rate of 15%.

[0388] The above calculations worked under the assumption of limiting at least one set of markers to an average of 50% sensitivity. How would the results improve if the average of 50% sensitivity was improved to 66% sensitivity?

[0389] Consider the strategy of using the 96 pan-oncology markers for detecting early colorectal cancer (Figure 1D). The calculations are done with the anticipation of an average of 150 methylated molecules per positive marker in the blood. As described *supra*, assume that the average late cancer was once the average early cancer, and thus individuals with Stage I cancer would be about 40,500 individuals. Assuming individual marker false-positive rates of 3%, the first step using 96 markers (48 markers for CRC) with average sensitivities of 66%, requiring a minimum of 5 markers positive, and an overall specificity of 95.8%, the first step would identify 4,494,000 individuals (out of 107,000,000 total adults over 50 in the U.S.) with Stage I colorectal cancer. This would include, at 90.0% sensitivity, about 36,450 individuals with Stage I colorectal cancer (out of 40,500 individuals with Stage I cancer). However, those 4,494,000

presumptive positive individuals would be evaluated in a second step of 64 markers (48 markers for CRC) with average sensitivities of 66%, and requiring a minimum of 5 markers positive. The two-step test would identify 90.0% x 90.0% = 89.0% = 32,805 individuals (out of 40,500 individuals with Stage I cancer) with colorectal cancer. With a specificity of 95.8%, the second test would also generate 4,494,000 x 4.2% = 188,748 false-positives. The positive predictive value of such a test would be 32,805 / (188,748 + 32,805) = 14.8%. In other words, 1 in 7 individuals who tested positive would actually have Stage I colorectal cancer. In reality, one would need to also include the success for identifying Stage 2 and higher cancers. In expanding this example, the calculations are done with the anticipation that Stage I CRC has an average of 150 methylated molecules per positive marker in the blood, Stage II CRC has an average of 200 methylated molecules per positive marker, and the higher stages (III & IV) have at least an average of 300 methylated molecules per positive marker. Also, to be consistent with the idea that as the test is used repeatedly, more of early and less of late CRC will be detected, an estimate of 40,500 individuals would be identified with Stage I cancer, 40,500 individuals would be identified with Stage II cancer, and the remaining 54,000 individuals would be identified with late-stage cancer. This equals 135,000 total individuals with colorectal cancer identified per year in the U.S. The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 98.0% would be identified in the first step, of which 98.0% x 98.0% = 96.0% = 38,896 individuals with Stage II cancer would be verified in the second step. For Stage III and IV cancer, 99.6 % would be identified in the first step, of which 99.6% x 99.6% = 99.2% = 53,568 individuals with late cancer would be identified. This brings the total identified to 32,805 + 38,896 + 53,568 = 125,269 individuals out of 135,000 with colorectal cancer, for an overall sensitivity of 92.7%. Overall, the positive predictive value of such a test would be 125,269 / (188,748 + 125,269) = 39.9%. In other words, 1 in 2.5 individuals who tested positive would actually have colorectal cancer. This test would identify 71,701 / 81,000, or 88%, of those individuals with early cancer, compared with the current rate of 40%.

[0390] How would these results vary for using this strategy (Figure 1D) for detection of early colorectal cancer using 66% average marker sensitivities, with the anticipation of Stage I CRC having an average of 200 methylated molecules per positive marker in the blood, Stage II CRC having an average of 240 methylated molecules per positive marker in the blood, and the higher stages (III & IV) having at least an average of 300 methylated molecules per positive marker in the blood?

[0391] Assuming individual marker false-positive rates of 3%, the first step using 96 markers (48 markers for CRC) with average sensitivities of 66%, and requiring a minimum of 5 markers positive, then, with an overall specificity of 95.8%, the first step would identify

4,494,000 individuals with colorectal cancer (out of 107,000,000 total adults over 50 in the U.S.). This would include, at 98.0% sensitivity, about 39,690 individuals with Stage I colorectal cancer (out of 40,500 individuals with Stage I cancer). However, those 4,494,000 presumptive positive individuals would be evaluated in a second step of 64 markers (48 markers for CRC) with average sensitivities of 66%, requiring a minimum of 5 markers positive. The two-step test would identify 98.0% x 98.0% = 96.0% = 38,896 individuals (out of 40,500 individuals with Stage I cancer) with colorectal cancer. With a specificity of 95.8%, the second test would also generate 4,494,000 x 4.2% = 188,748 false-positives. The positive predictive value of such a test would be 38,896 / (188,748 + 38,896) = 17.81%. In other words, 1 in 6 individuals who tested positive would actually have Stage I colorectal cancer. In reality, one would need to also include the success for identifying Stage 2 and higher cancers. To be consistent with the idea that, as the test is used repeatedly, more of early and less of late CRC will be detected, an estimate of 40,500 individuals with Stage I cancer would be identified, 40,500 individuals with Stage II cancer would be identified, and the remaining 54,000 individuals would be identified as having late-stage cancer (*i.e.,* 135,000 total individuals with colorectal cancer identified per year in the U.S.). The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 98.0% would be identified in the first step, of which 99.6% x 99.6% = 99.2% = 40,176 individuals with Stage II cancer would be verified in the second step. For Stage III and IV cancer, 99.9 % would be identified in the first step, of which 99.9% x 99.9% = 99.8% = 53,568 individuals with late cancer would be identified. This brings the total identified to 38,896 + 40,176 + 53,892 = 132,964 individuals out of 135,000 with colorectal cancer, for an overall sensitivity of 98.5%. Overall, the positive predictive value of such a test would be 132,964 / (188,748 + 132,964) = 41.3%. In other words, 1 in 2.5 individuals who tested positive would actually have colorectal cancer, and this test would identify 79,072 / 81,000 or 97.6% of those individuals with early cancer, compared with the current rate of 40%.

[0392] How would these results vary for using the first strategy (Figure 1D) for detection of early ovarian cancer, with the anticipation of an average of 150 methylated molecules per positive marker in the blood? Again, assume that the stages are evenly divided, and thus, individuals with Stage I ovarian cancer would be about 5,500 individuals. Assuming individual marker false-positive rates of 3%, the first step using 96 markers (36 markers for ovarian) with average sensitivities of 66%, and requiring a minimum of 5 markers positive, then, with an overall specificity of 99.1%, the first step would identify 486,000 individuals (out of 54,000,000 total women ages 50-79 in the U.S.). This would include, at 71.5% sensitivity, about 3,932 individuals with Stage I ovarian cancer (out of 5,500 individuals with Stage I ovarian cancer). However, those 486,000 presumptive positive individuals would be evaluated in a second step of

64 markers (36 markers for ovarian cancer) with average sensitivities of 66%, and requiring a minimum of 5 markers positive. The two-step test would identify 71.5% x 71.5% = 51.1% = 2,810 individuals (out of 5,500 individuals with Stage I ovarian cancer) with ovarian cancer. With a specificity of 99.1%, the second test would also generate 486,000 x 0.9% = 4,374 false-positives. The positive predictive value of such a test would be 2,810 / (4,374 + 2,810) = 39.1%. In other words, 1 in 2.5 individuals who tested positive would actually have Stage I ovarian cancer. In reality, one would need to also include the success for identifying Stage 2 and higher ovarian cancers. In expanding this example, the calculations are done with the anticipation that Stage I ovarian cancer has an average of 150 methylated molecules per positive marker in the blood, Stage II ovarian cancer has an average of 200 methylated molecules per positive marker, and the higher stages (III & IV) have at least an average of 300 methylated molecules per positive marker. Also, assuming all four stages are at 5,500, then 5,500 x 4 = 22,000 total individuals with ovarian cancer would be identified per year in the U.S. The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 90.0% would be identified in the first step, of which 90.0% x 90.0% = 81.0% = 4,485 individuals with Stage II ovarian cancer would be verified in the second step. For Stage III and IV ovarian cancer, 99.2% would be identified in the first step, of which 99.2% x 99.2% = 98.4% = 10,824 individuals with late ovarian cancer would be identified. This brings the total identified at 2,810 + 4,485 + 10,824 = 18,119 individuals out of 22,000 with ovarian cancer, for an overall sensitivity of 82.4%. Overall, the positive predictive value of such a test would be 18,119 /(18,119 + 4,374) = 80.5%. In other words, 4 in 5 women who tested positive would actually have ovarian cancer, and this test would identify 7,295 / 11,000 or 66.3% of those individuals with early cancer, compared with the current rate of 15%.

[0393] How would these results vary for using this strategy (Figure 1D) for detection of early ovarian cancer using 66% average marker sensitivities, with the anticipation that Stage I ovarian cancer has an average of 200 methylated molecules per positive marker in the blood, Stage II ovarian cancer has an average of 240 methylated molecules per positive marker, and the higher stages (III & IV) have at least an average of 300 methylated molecules per positive marker?

[0394] Assuming individual marker false-positive rates of 3%, the first step using 96 markers (36 markers for ovarian) with average sensitivities of 66%, and requiring a minimum of 5 markers positive, then, with an overall specificity of 99.1%, the first step would identify 486,000 individuals (out of 54,000,000 total women ages 50-79 in the U.S). This would include, at 90.0% sensitivity, about 4,950 individuals with Stage I ovarian cancer (out of 5,500 individuals with Stage I ovarian cancer). However, those 486,000 presumptive positive

individuals would be evaluated in a second step of 64 markers (36 markers for ovarian cancer) with average sensitivities of 66%, requiring a minimum of 5 markers positive. The two-step test would identify 90.0% x 90.0% = 81.0% = 4,895 individuals (out of 5,500 individuals with Stage I ovarian cancer) with ovarian cancer. With a specificity of 99.1%, the second test would also generate 486,000 x 0.9% = 4,374 false-positives. The positive predictive value of such a test would be 4,895 / (4,374 + 4,895) = 52.8%. In other words, 1 in 2 individuals who tested positive would actually have Stage I ovarian cancer. In reality, one would need to also include the success for identifying Stage 2 and higher ovarian cancers. Assuming all four stages are at 5,500, then 5,500 x 4 = 22,000 total individuals with ovarian cancer would be identified per year in the U.S. The above calculation already provided the false-positive rate for the early cancer. For Stage II cancer, 96.2% would be identified in the first step, of which 96.2% x 96.2% = 92.5% = 5,087 individuals with Stage II ovarian cancer would be verified in the second step. For Stage III and IV ovarian cancer, 99.2% would be identified in the first step, of which 99.2% x 99.2% = 98.4% = 10,824 individuals with late ovarian cancer would be identified. This brings the total identified at 4,895 + 5087 + 10,824 = 20,806 individuals out of 22,000 with ovarian cancer, for an overall sensitivity of 94.6%. Overall, the positive predictive value of such a test would be 20,806 /(20,806 + 4,374) = 87.4%. In other words, 7 in 8 women who tested positive would actually have ovarian cancer, and this test would identify 9,982 / 11,000 or 90.1% of those individuals with early cancer, compared with the current rate of 15%.

[0395] The biology of each cancer is different, and thus the observed sensitivity and specificity for detecting early cancer, monitoring treatment, and detecting early recurrence may be higher or lower from the idealized calculations described herein.

EXAMPLES

Examples: Multiplex PCR-LDR-qPCR Detection of Cancer-Related Methylation Markers

General Methods for Examples 1-5

[0396] HT-29 colon adenocarcinoma cells were seeded in 60 cm^2 culture dishes in McCoy's 5A medium containing 4.5 g/l glucose, supplemented with 10% fetal calf serum, and kept in a humidified atmosphere containing 5% CO_2. Once cells reached 80-90% confluence, they were washed in Phosphate Buffered Saline (x3), and collected by centrifugation (500xg). Genomic DNA was isolated using the DNeasy Blood & Tissue Kit (Qiagen; Valencia, Calif.), and its concentration measured using Quant-iT Pico green Assay (Life Technologies/Thermo-Fisher; Waltham, Mass.).

[0397] High molecular weight (>50 kb) genomic DNA (0.2 mg/ml) isolated from human blood (buffy coat) (Roche human genomic DNA) was purchased from Roche (Indianapolis, Ind.). Its concentration was similarly determined using Quant-iT PicoGreen dsDNA Assay Kit.

[0398] Cell free DNA was isolated from 5 ml plasma sample (with K_2EDTA additive as anti-coagulant) using the QIA amp Circulating Nucleic Acid Kit according to manufacturer's instructions, and quantified using Quant-iT Pico Green Assay (Life Technologies/ThermoFisher; Waltham, Mass.).

[0399] 0.5-1.0 µg HT29 cell line genomic DNA was digested with 10 units of the restriction enzyme Bsh1236I in 20 µl of reaction solution containing 1xCutSmart buffer (50 mM Potassium Acetate, 20 mM Tris-Acetate, 10 mM Magnesium Acetate, 100 µg/ml BSA, pH 7.9 at 25°C). The digestion reaction was carried out at 37°C for 1 hour, followed by enzyme inactivation at 80°C for 20 min. Alternatively, genomic DNAs can be fragmented through non random sonication method, using Covaris ultra sonicator (Woburn, Massachusetts). After shearing, the quality of the resulting DNA fragments (length ranged from 50 to 1 kb base pairs) was assessed with Agilent Bioanalyzer system. This is followed by an enrichment step wherein the DNA fragments containing methylated CpGs are then captured by methylation-specific antibodies, using the EpiMark® Methylated DNA Enrichment Kit according to manufacturer's instructions (New England Biolabs; Ipswich, MA).

[0400] **PCR primers, LDR probes, and LNA or PNA blocking primers.** All primers used for the one or two-step assay to detect colorectal cancer are listed in Table 45 below. All primers were purchased from Integrated DNA Technologies Inc. (IDT) (Coralville, Iowa), except for LNA1 and LNA2, which were purchased from Exiqon Inc. (Woburn, Mass.), and PNA, which was purchased from PNA Bio (Thousand Oaks, Calif.). Primers designed for use in Step 1 of the 96-marker assay, with average sensitivities of 50%, detect solid tumors are listed in Table 46 below. Primers designed for use in Step 2 of the Group 1- 64-marker assay, with average sensitivities of 50%, to detect and identify colorectal, stomach, and esophageal cancers are listed in Table 53 below. Primers designed for use in Step 2 of the Group 2- 48-64-marker assay, with average sensitivities of 50%, to detect and identify breast, endometrial, ovarian, cervical, and uterine cancers are listed in Table 54 below. Primers designed for use in Step 2 of the Group 3- 48-64-marker assay, with average sensitivities of 50%, to detect and identify lung adenocarcinomas, lung squamous cell carcinoma, and head & neck cancers are listed in Table 55 below. Primers designed for use in Step 2 of the Group 4- 36-48-marker assay, with average sensitivities of 50%, to detect and identify prostate and bladder cancers are listed in Table 56 below. Primers designed for use in Step 2 of the Group 5- 48-64-marker assay, with average

sensitivities of 50%, to detect and identify liver, pancreatic, and gall-bladder cancers are listed in Table 57 below.

Table 45.

Primers for use in one or two-step assay to detect colorectal cancer.

Site	Primer Name	Sequence	Length	Seq. Id. No.
VIM-S3				
Forward PCR Primer	iCDx-2031-VIM-S3-FP	GAACTCCAACCGAAACTACGTAArCTACA/3SpC3/	28	1
Reverse PCR Primer	iCDx-2032A-VIM-S3-RP	GGTGTCGTGGACGAGGCGTAGAGGTTGCrGGTTA/3SpC3/	33	2
Upstream LDR	iCDx-2033A-VIM-S3-Up	TAGACACGAGCGAGGTCACAACTCCAACCGAAACTACGTAACTGCGrUCCGT/3SpC3/	51	3
Downstream LDR	iCDx-2034A-VIM-S3-Dn	/5Phos/TCCACCCGCACCTACAACCTAAACAACGCGTGCAAAATTCAGGCTGTGCA	50	4
Real-Time Probe	iCDx-2035B-VIM-S3-RT-Pb	/5HEX/TTTAACTGC/ZEN/GTCCACCCGCACCTAC/3IABkFQ/	25	5
Tag Forward Primer	iCDx-2036-VIM-S3-RT-FP	TAGACACGAGCGAGGTCAC	19	6
Tag Reverse Primer	iCDx-2037-VIM-S3-RT-RP	TGCACAGCCTGAATTTTGCAC	21	7
CLIP4-G1				
Forward PCR Primer	iCDx-2371-CLIP4-R-S1-FP	CGCGAGGTTGAGGGTTGTGrAAGGT/3SpC3/	24	8
Reverse PCR Primer	iCDx-2372-CLIP4-R-S1-RP	GGTGTCGTGGGTCTACGAAATATCGCAATATTACCTCCrCCCGT/3SpC3/	43	9
Upstream LDR	iCDx-2373-CLIP4-R-S1-Up	TTCAGAGCACCTGCGTACCGAGGTTGAGGGTTGTGAAAGCrGgtAA/3SpC3/	43	10
Downstream LDR	iCDx-2374-CLIP4-R-S1-Dn	/5Phos/GGTGGGTACGTACGGCGTGTCGGGTTCTTCGGCTGGCTCAA	41	11
Real-Time Probe	iCDx-2375-CLIP4-R-S1-RT-Pb	/56-FAM/CCTTGTGAA/ZEN/AGCGGTGGGTACGTAC/3IABkFQ/	25	12
Tag Forward Primer	iCDx-2376-CLIP4-R-S1-RT-FP	TTCAGAGCACCTGCGTACC	19	13
Tag Reverse Primer	iCDx-2377-CLIP4-R-S1-RT-RP	TTGAGCCAGCCGAAGAACC	19	14
GSG1L-G3				
Forward PCR Primer	iCDx-2391-1-GSG1L-F-S1-FP,	GAAAAAACCGAAACCGAACTAACCrGCCGT/3SpC3/	29	15
Reverse PCR Primer	iCDx-2392-1-GSG1L-F-S1-RP,	GGTGTCGTGGTTTTTATATCGGTATTTGGTATTCGTGCrGAGGG/3SpC3/	33	16
Upstream LDR	iCDx-2393-GSG1L-F-S1-Up	TTCGTCCCTGCACGCTAACCGAACTAACCGCCGTCCrGCGTA/3SpC3/	41	17
Downstream LDR	iCDx-2394-GSG1L-F-S1-Dn	/5Phos/GCGCGCACTCACCAAACCCGGTTCCATCACCGTTAGGCCA	40	18
Real-Time Probe	iCDx-2395-GSG1L-F-S1-RT-Pb	/56-FAM/TTCCGTCCG/ZEN/CGCGCA/3IABkFQ/	15	19
Tag Forward Primer	iCDx-2396-GSG1L-F-S1-RT-FP	TTCGTCCCTGCACGCTAAC	19	20
Tag Reverse Primer	iCDx-2397-GSG1L-F-S1-RT-	TGGCCTAACGGTGATGGAAC	20	21

	RP			
LNA Blocking Probe	iCDx-2398-GSG1L-F-S1-LNA	cC+AcCAccC+AC+AC+AcactcacC/3SpC3/	11	22
PP1R16B-1				
Forward PCR Primer	iCDx-2531-1-PP1R16B-F-S1-FP,	GTTAAAAAAACTATTCCGAAACCTAACCrACGTT/3SpC3/	33	23
Reverse PCR Primer	iCDx-2532-PP1R16B-F-S1-RP	GGTGTCGTGGAGGTGGGCGCGTTTAATTTTATTCrGGTTC/3SpC3/	39	24
Upstream LDR	iCDx-2533-PP1R16B-F-S1-Up	TTCAGCAGCCTGGCATCACGAAACCTAACCACGTCCCAGCCrGAtCC/3SpC3/	45	25
Downstream LDR	iCDx-2534-PP1R16B-F-S1-Dn	/5Phos/GATTTCAACTTCCTACAACTCAAAAAAAAAATCCCCACCGTGGAGCGCTAAGGTTGCA	58	26
Real-Time Probe	iCDx-2535-PP1R16B-F-S1-RT-Pb	/56-FAM/TTCGTCCCA/ZEN/GCCGATTTCAACTTCCTACAA/3IABkFQ/	30	27
Tag Forward Primer	iCDx-2536-PP1R16B-F-S1-RT-FP	TTCAGCAGCCTGGCATCAC	19	28
Tag Reverse Primer	iCDx-2537-PP1R16B-F-S1-RT-RP	TGCAACCTTAGCGCTCCAC	19	29
KCNA3-1				
Forward PCR Primer	iCDx-2551-KCNA3-F-S1-FP	GACTCGTAACGATCGCAACCGrCCGCT/3SpC3/	26	30
Reverse PCR Primer	iCDx-2552-KCNA3-F-S1-RP	GGTGTCGTGGCGGTTACGCGGAGTTCGTCrGTAGA/3SpC3/	34	31
Upstream LDR	iCDx-2553-KCNA3-F-S1-Up	TCACAGAGACTTGCCGATCACGATCGCAACCGCCACCrGCCGT/3SpC3/	42	32
Downstream LDR	iCDx-2554-KCNA3-F-S1-Dn	/5Phos/GCCACAACCGCCTTAAAACGAAACCCGTGTGTAGCTTAGACATGGCCA	48	33
Real-Time Probe	iCDx-2555-KCNA3-F-S1-RT-Pb	/56-FAM/TTCGCCACC/ZEN/GCCACAACC/3IABkFQ/	18	34
Tag Forward Primer	iCDx-2556-KCNA3-F-S1-RT-FP	TCACAGAGACTTGCCGATCAC	21	35
Tag Reverse Primer	iCDx-2557-KCNA3-F-S1-RT-RP	TGGCCATGTCTAAGCTACACAC	22	36
LONFR2-G5				
Forward PCR Primer	iCDx-2301-LONFR2-R-S1-FP	GCGAAAGAGTTCGGTTGTTATTTCGrUAGTT/3SpC3/	30	37
Reverse PCR Primer	iCDx-2302-LONFR2-R-S1-RP	GGTGTCGTGGCAATATCCTAACTACGACCGCGCrGAAAT/3SpC3/	38	38
Upstream LDR	iCDx-2303-LONFR2-R-S1-Up	TCCAAACGATTAGGAGCGTCAACGAAAGAGTTCGGTTGTTATTTCGTAATCrGTTTA/3SpC	56	39

		3/		
Downstream LDR	iCDx-2304-LONFR2-R-S1-Dn	/5Phos/GTTCGCGCGGAAGGTTTCGTCGTTTTGGACAGAGGTATACGCCCA	45	40
Real-Time Probe	iCDx-2305-LONFR2-R-S1-RT-Pb	/56-FAM/AAATTTCGT/ZEN/AATCGTTCGCGCGGA/3IABkFQ/	24	41
Tag Forward Primer	iCDx-2306-LONFR2-R-S1-RT-FP	TCCAAACGATTAGGAGCGTCAA	22	42
Tag Reverse Primer	iCDx-2307-LONFR2-R-S1-RT-RP	TGGGCGTATACCTCTGTCCAA	21	43
TBC1D10C				
Forward PCR Primer	iCDx-2931-TBC1D10C-F-S1-FP	CCCCGAAAAAACCCAAACCACrCGCGG/3SpC3/	26	44
Reverse PCR Primer	iCDx-2932-TBC1D10C-F-S1-RP	GGTGTCGTGGGGTTTCGGGTTCGAGTTAGGArGTTTG/3SpC3/	36	45
Upstream LDR	iCDx-2933-TBC1D10C-F-S1-Up	TAGCAGCTGAACAACCCAACAAAACCCAAACCACCGCAACrGAAGG/3SpC3/	45	46
Downstream LDR	iCDx-2934-TBC1D10C-F-S1-Dn	/5Phos/GAAAACCCCAAACCCGCGACAAAACTTGTTGTATGGTCGGCATGCTA	47	47
Real-Time Probe	iCDx-2935-TBC1D10C-F-S1-RT-Pb	/56-FAM/TTCCGCAAC/ZEN/GAAAACCCCAAACCC/3IABkFQ/	24	48
Tag Forward Primer	iCDx-2936-TBC1D10C-F-S1-RT-FP	TAGCAGCTGAACAACCCAAC	20	49
Tag Reverse Primer	iCDx-2937-TBC1D10C-F-S1-RT-RP	TAGCATGCCGACCATACAAC	20	50
GDF6-1				
Forward PCR Primer	iCDx-2641-GDF6-F-S1-FP	CGACGCCAAAACCAAAAAACTACrcCGaT/3SpC3/	26	51
Reverse PCR Primer	iCDx-2642-GDF6-F-S1-RP	GGTGTCGTGGGAGATAGTTTTGGAAAGTTTTGGGTAAAGTCrggtac/3SpC3/	41	52
Upstream LDR	iCDx-2643-GDF6-F-S1-Up	TCTTACGCCCAGGGAATGTAACCGCCAAAACCAAAAAACTACCCAACrGcCAT/3SpC3/	51	53
Downstream LDR	iCDx-2644-GDF6-F-S1-Dn	/5Phos/GCCGCTCGCGAACTAATTCCTCAAACTATAAAACGTTGTCCGGCTGTGGTTACA	54	54
Real-Time Probe	iCDx-2645-GDF6-F-S1-RT-Pb	/56-FAM/TGTACCCAA/ZEN/CGCCGCTCGC/3IABkFQ/	19	55
Tag Forward Primer	iCDx-2646-GDF6-F-S1-RT-FP	TCTTACGCCCAGGGAATGTAAC	22	56
Tag Reverse Primer	iCDx-2647-GDF6-F-S1-RT-RP	TGTAACCACAGCCGGACAAC	20	57

CLIP4-G3				
Forward PCR Primer	iCDx-2361-CLIP4-F-S2-FP	CGCGAAACTAAAAACTATAAAAACGATAAACArCGcaT/3SpC3/	35	58
Reverse PCR Primer	iCDx-2362-CLIP4-F-S2-RP	GGTGTCGTGGTGCGGGGTGTCGTAGTGTTA rUTTTC/3SpC3/	35	59
Upstream LDR	iCDx-2363-CLIP4-F-S2-Up	TTTCGCTCGACGCATACCAAAACTAAAAACTATAAAAACGATAAACACGTACrGACAC/3SpC3/	57	60
Downstream LDR	iCDx-2364-CLIP4-F-S2-Dn	/5Phos/GACGTACCGAAACCGCCTAACTTCGCTGGCGCGGCTACTGTAAAA	45	61
Real-Time Probe	iCDx-2365-CLIP4-F-S2-RT-Pb	/5HEX/CCAACACGT/ZEN/ACGACGTACCGAAACC/3IABkFQ/	25	62
Tag Forward Primer	iCDx-2366-CLIP4-F-S2-RT-FP	TTTCGCTCGACGCATACCA	19	63
Tag Reverse Primer	iCDx-2367-CLIP4-F-S2-RT-RP	TTTTACAGTAGCCGCGCCA	19	64
SEPT9-G1				
Forward PCR Primer ver A	iCDx-2801-SEPT9-F-S1-FP	TCCTCCGACGACTAACTCTACACrUACAG/3SpC3/	28	65
Forward PCR Primer ver B	iCDx-2801A-SEPT9-F-S1-FP	CTCCGACGACTAACTCTACACTACAAArAACGT/3SpC3/	32	66
Reverse PCR Primer	iCDx-2802-SEPT9-F-S1-RP	GGTGTCGTGAGGTAGCGGCGAGGAAGCrGTTTC/3SpC3/	32	67
Upstream LDR	iCDx-2803-SEPT9-F-S1-Up	TAGGAACACGGAGGACATCAACGACGACTAACTCTACACTACAAAAATGCrGAATA/3SpC3/	55	68
Downstream LDR	iCDx-2804-SEPT9-F-S1-Dn	/5Phos/GAACGCGACGCCCCAACCAAC TTGTGGGTGGGTATAGGTCAGA	43	69
Real-Time Probe	iCDx-2805-SEPT9-F-S1-RT-Pb	/56-FAM/TTAAAATGC/ZEN/GAACGCGACGCCC/3IABkFQ/	22	70
Tag Forward Primer	iCDx-2806-SEPT9-F-S1-RT-FP	TAGGAACACGGAGGACATCAA	21	71
Tag Reverse Primer	iCDx-2807-SEPT9-F-S1-RT-RP	TCTGACCTATACCCACCCACAA	22	72
VIM-1				
Forward PCR Primer	AcDx-5001-VIM-S1-FP	CGAGTCGGTCGAGTTTTAGTCrGGAGC/3SpC3/	26	73
Reverse PCR Primer	AcDx-5002-VIM-S1-RP	GGTGTCGTGGCCCGAAAACGAAACGTAAAAACTACrGACTG/3SpC3/	40	74

Upstream LDR	AcDx-5003-VIM-S1-Up	TCTCATACCAGACGCGGTAACTCGAGTTTTAGTCGGAGTTACGTGATCACrGTTCG/3SpC3/	55	75
Downstream LDR	AcDx-5004-VIM-S1-Dn	/5Phos/GTTTATTCGTATTTATAGTTTGGGTAGCGCGTTGCGGTTCGTGTCGCTGTGCTTA	55	76
Real-Time Probe	AcDx-5005-VIM-S1-RT-Pb	/56-FAM/AATGATCAC/ZEN/GTTTATTCGTATTTATAGTTTGGGTAGCG/3IABkFQ/	38	77
Tag Forward Primer	AcDx-5006-VIM-S1-RT-FP	TCTCATACCAGACGCGGTAAC	21	78
Tag Reverse Primer	AcDx-5007-VIM-S1-RT-RP	TAAGCACAGCGACACGAAC	19	79
Downstream PCR Primer	AcDx-5008-VIM-S1-PCR-V	TAAGCACAGCGACACGAACCGAAACGTAAAAACTACGACTAATACTAAAATGrCAACA/3SpC3/	57	80
VIM-2				
Forward PCR Primer	AcDx-5011-VIM-S2-FP	ATTTTTTTTTTATCGCGCGATTTCrGTTTC/3SpC3/	29	81
Reverse PCR Primer	AcDx-5012-VIM-S2-RP	GGTGTCGTGGCCTACTCCGAAACCTAAAACTAAACrATAAC/3SpC3/	40	82
Upstream LDR	AcDx-5013-VIM-S2-Up	TCTGCCCTTCGCTTCGAACTTTTTTTAGTATTTTAGGGTGAGTTTAGTTTAGATTATTACTCrGGAAG/3SpC3/	67	83
Downstream LDR	AcDx-5014-VIM-S2-Dn	/5Phos/GGAAAGTTTTTAAAAGTTTTAGTTTAGCGTTGAAGTAACGGGGTTGTATGGTCGGCATGCTA	62	84
Real-Time Probe	AcDx-5015-VIM-S2-RT-Pb	/56-FAM/CCATTACTC/ZEN/GGAAAGTTTTTAAAAGTTTTAGTTTAGCGTTGA/3IABkFQ/	42	85
Tag Forward Primer	AcDx-5016-VIM-S2-RT-FP	TCTGCCCTTCGCTTCGAAC	19	86
Tag Reverse Primer	AcDx-5017-VIM-S2-RT-RP	TAGCATGCCGACCATACAAC	20	87
Downstream PCR Primer	AcDx-5018-VIM-S2-PCR-V	TAGCATGCCGACCATACAACCCGAAACCTAAAACTAAACATAATCCTGrUTACC/3SpC3/	53	88
CLIP4-1				
Forward PCR Primer	AcDx-5021-CLIP4-S1-FP	GGTTGAGGGTTGTGAAGGCrGGTGA/3SpC3/	24	89
Reverse PCR Primer	AcDx-5022-CLIP4-S1-RP	GGTGTCGTGGCGTCTACGAAATATCGCAATATTACCrUCCCT/3SpC3/	41	90
Upstream LDR	AcDx-5023-CLIP4-S1-Up	TCCAAACAAGCTGATCCGTACAGGTTGTGAAGGCGGTGGGCACrGTATA/3SpC3/	48	91
Downstream LDR	AcDx-5024-CLIP4-S1-Dn	/5Phos/GTACGGCGTGTCGGAGTCGTTTGGTGTGTCGGAGCGGTTACTA	43	92
Real-Time Probe	AcDx-5025-CLIP4-S1-RT-Pb	/56-FAM/AATGGGCAC/ZEN/GTACGGCGTGT/3IABkFQ/	20	93
Tag Forward Primer	AcDx-5026-CLIP4-S1-RT-FP	TCCAAACAAGCTGATCCGTACA	22	94
Tag Reverse Primer	AcDx-5027-CLIP4-S1-RT-RP	TAGTAACCGCTCCGACACA	19	95
Downstream PCR Primer	AcDx-5028-CLIP4-S1-PCR-V	TAGTAACCGCTCCGACACACGCCGCGAAACCAAATGrACCCT/3SpC3/	41	96

CLIP4-2				
Forward PCR Primer	AcDx-5031-CLIP4-S2-FP	GGTTGAGGGTTGTGAAGGCrGGTGA/3SpC3/	24	97
Reverse PCR Primer	AcDx-5032-CLIP4-S2-RP	GGTGTCGTGGCTAACAACGTCTACGAAATATCGCrAATAC/3SpC3/	39	98
Upstream LDR	AcDx-5033-CLIP4-S2-Up	TCCAAACAAGCTGATCCGTACATGAAGGCGGTGGGTACGCACrGGCAC/3SpC3/	47	99
Downstream LDR	AcDx-5034-CLIP4-S2-Dn	/5Phos/GGCGTGTCGGAGTCGTTTGGTTTCGTGTGTCGGAGCGGTTACTA	44	100
Real-Time Probe	AcDx-5035-CLIP4-S2-RT-Pb	/56-FAM/AATACGCAC/ZEN/GGCGTGTCGGAG/3IABkFQ/	21	101
Tag Forward Primer	AcDx-5036-CLIP4-S2-RT-FP	TCCAAACAAGCTGATCCGTACA	22	102
Tag Reverse Primer	AcDx-5037-CLIP4-S2-RT-RP	TAGTAACCGCTCCGACACA	19	103
Downstream PCR Primer	AcDx-5038-CLIP4-S2-PCR-V	TAGTAACCGCTCCGACACACGCCGCGAAACCAAATGrACCCT/3SpC3/	41	104
CLIP4-3				
Forward PCR Primer	AcDx-5041-CLIP4-S3-FP	GTTAGAGACGTGAGGTCGCrGAGGC/3SpC3/	24	105
Reverse PCR Primer	AcDx-5042-CLIP4-S3-RP	GGTGTCGTGGCTACGAAATATCGCAATATTACCTCCrCCCGT/3SpC3/	41	106
Upstream LDR	AcDx-5043-CLIP4-S3-Up	TCCAAACAAGCTGATCCGTACAGAGGTTGAGGGTTGTGAAAGCrGGTAA/3SpC3/	48	107
Downstream LDR	AcDx-5044-CLIP4-S3-Dn	/5Phos/GGTGGGTACGTACGGCGTGTCGTGTGTCGGAGCGGTTACTA	41	108
Real-Time Probe	AcDx-5045-CLIP4-S3-RT-Pb	/56-FAM/AATGAAAGC/ZEN/GGTGGGTACGTACGG/3IABkFQ/	24	109
Tag Forward Primer	AcDx-5046-CLIP4-S3-RT-FP	TCCAAACAAGCTGATCCGTACA	22	110
Tag Reverse Primer	AcDx-5047-CLIP4-S3-RT-RP	TAGTAACCGCTCCGACACA	19	111
Downstream PCR Primer	AcDx-5048-CLIP4-S3-PCR-V	TAGTAACCGCTCCGACACACGCCGCGAAACCAAATGrACCCT/3SpC3/	41	112
GSG1L				
Forward PCR Primer	AcDx-5051-GSG1L-S1-FP	AGTCGGAGTCGAGTTGGTCrGTCGC/3SpC3/	24	113
Reverse PCR Primer	AcDx-5052-GSG1L-S1-RP	GGTGTCGTGGAAAAATTTCCACACCGACATCTAArUACTT/3SpC3/	39	114
Upstream LDR	AcDx-5053-GSG1L-S1-Up	TCTGCCAGAACACCGACACGGAGTCGAGTTGGTCGTCGCTCrGCGTA/3SpC3/	46	115
Downstream LDR	AcDx-5054-GSG1L-S1-Dn	/5Phos/GCGCGTATTTATTAAGTTCGTTGAGTTTTTTTTCGTACGGTGTGTTGGCGTACGGTGA	58	116
Real-Time Probe	AcDx-5055-GSG1L-S1-RT-Pb	/56-FAM/AAGTCGCTC/ZEN/GCGCGTATTTATTAAGTTCGT/3IABkFQ/	30	117
Tag Forward Primer	AcDx-5056-GSG1L-S1-RT-FP	TCTGCCAGAACACCGACAC	19	118

Tag Reverse Primer	AcDx-5057-GSG1L-S1-RT-RP	TCACCGTACGCCAACACAC	19	119
Downstream PCR Primer	AcDx-5058-GSG1L-S1-PCR-V	TCACCGTACGCCAACACACCACACCGACATCTAATACTCGTATGrAAAAG/3SpC3/	49	120
PP1R16B-2				
Forward PCR Primer	AcDx-5061-PP1R16B-S1-FP	GGGTTTTTATTCGAGAGCGTCrGGGAC/3SpC3/	26	121
Reverse PCR Primer	AcDx-5062-PP1R16B-S1-RP	GGTGTCGTGGCCCAAAACGAAACCTAAACTCCrUAAAG/3SpC3/	37	122
Upstream LDR	AcDx-5063-PP1R16B-S1-Up	TTCGTGGGCACACAAGCAACGAGAGCGTCGGGATTTTGTCTCrGCGCC/3SpC3/	47	123
Downstream LDR	AcDx-5064-PP1R16B-S1-Dn	/5Phos/GCGTTGTTTTTTAAGTCGGATGGAGTTGAGCTTGCTTGGCTTGATCTACCTGA	53	124
Real-Time Probe	AcDx-5065-PP1R16B-S1-RT-Pb	/56-FAM/AATTGTCTC/ZEN/GCGTTGTTTTTTAAGTCGGATG/3IABkFQ/	31	125
Tag Forward Primer	AcDx-5066-PP1R16B-S1-RT-FP	TTCGTGGGCACACAAGCAA	19	126
Tag Reverse Primer	AcDx-5067-PP1R16B-S1-RT-RP	TCAGGTAGATCAAGCCAAGCAA	22	127
Downstream PCR Primer	AcDx-5068-PP1R16B-S1-PCR-V	TCAGGTAGATCAAGCCAAGCAAACCTAAACTCCTAAAACTAAAATAAACGTGrCTCAG/3SpC3/	57	128
KCNA3-2				
Forward PCR Primer	AcDx-5071-KCNA3-S1-FP	GCGCGCGTTTCGTTTTCrGGGAA/3SpC3/	22	129
Reverse PCR Primer	AcDx-5072-KCNA3-S1-RP	GGTGTCGTGGCGCCGAAATACAACATAAAAACTCrUTTCA/3SpC3/	39	130
Upstream LDR	AcDx-5073-KCNA3-S1-Up	TTTCAGGCCCTAACCACCACGCGTTTCGTTTTCGGAGGTAATCrGTCAA/3SpC3/	48	131
Downstream LDR	AcDx-5074-KCNA3-S1-Dn	/5Phos/GTCGGGTTTGTATTTTTTGTAGTTTTTAAGGTTTTTCGGTGTGGGATTAAGGGCGATGGA	60	132
Real-Time Probe	AcDx-5075-KCNA3-S1-RT-Pb	/56-FAM/AAGGTAATC/ZEN/GTCGGGTTTGTATTTTTTGTAGTTTTTAAGG/3IABkFQ/	40	133
Tag Forward Primer	AcDx-5076-KCNA3-S1-RT-FP	TTTCAGGCCCTAACCACCAC	20	134
Tag Reverse Primer	AcDx-5077-KCNA3-S1-RT-RP	TCCATCGCCCTTAATCCCAC	20	135
Downstream PCR Primer	AcDx-5078-KCNA3-S1-PCR-V	TCCATCGCCCTTAATCCCACCAACATAAAAACTCTTTCGCTAACACTGrAAAAG/3SpC3/	53	136
GDF6-2				
Forward PCR Primer	AcDx-5081-GDF6-S1-FP	GGTTGCGTTTTTTTAGGAGGCrGGTGA/3SpC3/	26	137
Reverse PCR Primer	AcDx-5082-GDF6-S1-RP	GGTGTCGTGGACCCCGACCGCTATCCrAACCA/3SpC3/	31	138

Upstream LDR	AcDx-5083-GDF6-S1-Up	TCACTATCGGCGTAGTCACCAGAGGCGGTGGCAGCrGGCAC/3SpC3/	40	139
Downstream LDR	AcDx-5084-GDF6-S1-Dn	/5Phos/GGCGTAGGACGCGCGGG TGGTGACTTTACCCGGAGGA	37	140
Real-Time Probe	AcDx-5085-GDF6-S1-RT-Pb	/56-FAM/AATGGCAGC/ZEN/GGCGTAGGACG/3IABkFQ/	20	141
Tag Forward Primer	AcDx-5086-GDF6-S1-RT-FP	TCACTATCGGCGTAGTCACCA	21	142
Tag Reverse Primer	AcDx-5087-GDF6-S1-RT-RP	TCCTCCGGGTAAAGTCACCA	20	143
Downstream PCR Primer	AcDx-5088-GDF6-S1-PCR-V	TCCTCCGGGTAAAGTCACCAAACCGCTCCGTACCCTGrCGCGC/3SpC3/	42	144
SEPT9-5				
Forward PCR Primer	AcDx-5091-SEPT9-S1-FP	TTTTTTGTAGAAGGATTTTGCGTTCrGGGAG/3SpC3/	30	145
Reverse PCR Primer	AcDx-5092-SEPT9-S1-RP	GGTGTCGTGGCCGAACGCCCCGCTArCGACT/3SpC3/	30	146
Upstream LDR	AcDx-5093-SEPT9-S1-Up	TAGCGATAGTACCGACAGTCACGGGAGGAATTTTTTTTTTTTTGGGCGCTCrGTTCC/3SpC3/	56	147
Downstream LDR	AcDx-5094-SEPT9-S1-Dn	/5Phos/GTTTTTTTCGTTATGGTTCGGTTTTTATATTCGTTTATATTTGGTCGTGCGGAAACCTATCGTCGA	66	148
Real-Time Probe	AcDx-5095-SEPT9-S1-RT-Pb	/56-FAM/AAGGCGCTC/ZEN/GTTTTTTTCGTTATGGTTCG/3IABkFQ/	29	149
Tag Forward Primer	AcDx-5096-SEPT9-S1-RT-FP	TAGCGATAGTACCGACAGTCAC	22	150
Tag Reverse Primer	AcDx-5097-SEPT9-S1-RT-RP	TCGACGATAGGTTTCCGCAC	20	151
Downstream PCR Primer	AcDx-5098-SEPT9-S1-PCR-V	TCGACGATAGGTTTCCGCACCCCCGCTACGACCAAATATAAATGrAATAC/3SpC3/	49	152
ADHFE1				
Forward PCR Primer	AcDx-5101-ADHFE1-S1-FP	GGTGCGAGCGTCGTTrGGGAC/3SpC3/	20	153
Reverse PCR Primer	AcDx-5102-ADHFE1-S1-RP	GGTGTCGTGGGCCTACCCACCCGCrUTCGT/3SpC3/	29	154
Upstream LDR	AcDx-5103-ADHFE1-S1-Up	TTGATTGGGATCGTTCGCACGGGTAGTTGGCGTTTTGGTTTTTATCTCrGTGAA/3SpC3/	53	155
Downstream LDR	AcDx-5104-ADHFE1-S1-Dn	/5Phos/GTGGGAAAATGGTTTTGAGTTCGATTGGTTTGAGGTGGCTCAATAACGGGCAGA	54	156
Real-Time Probe	AcDx-5105-ADHFE1-S1-RT-Pb	/56-FAM/AATTATCTC/ZEN/GTGGGAAAATGGTTTTGAGTTCGA/3IABkFQ/	33	157
Tag Forward Primer	AcDx-5106-ADHFE1-S1-RT-FP	TTGATTGGGATCGTTCGCAC	20	158
Tag Reverse Primer	AcDx-5107-ADHFE1-S1-RT-RP	TCTGCCCGTTATTGAGCCAC	20	159
Downstream PCR	AcDx-5108-ADHFE1-S1-PCR-	TCTGCCCGTTATTGAGCCACCCCACCCGCTTCGTG rAAATT/3SpC3/	40	160

Primer	V			
KRBA1				
Forward PCR Primer	AcDx-5111-KRBA1-S1-FP	TTTTGGGAAAGATGGGTTTTTTTTTArUTTTC/3SpC3/	31	161
Reverse PCR Primer	AcDx-5112-KRBA1-S1-RP	GGTGTCGTGGTACAAACCCGCGAATACGCrGCCGT/3SpC3/	34	162
Upstream LDR	AcDx-5113-KRBA1-S1-Up	TGTGCACTAGTCCACGTGAAACGGTTTTTTTTTATTTTTATTATTAGTTTTTGGTATATGGAACrGCGCC/3SpC3/	69	163
Downstream LDR	AcDx-5114-KRBA1-S1-Dn	/5Phos/GCGTTGAGTGGTGGTTGCGGGGTTTCCGCGATCTTTGCATTCA	43	164
Real-Time Probe	AcDx-5115-KRBA1-S1-RT-Pb	/56-FAM/CCATGGAAC/ZEN/GCGTTGAGTGGTGGT/3IABkFQ/	24	165
Tag Forward Primer	AcDx-5116-KRBA1-S1-RT-FP	TGTGCACTAGTCCACGTGAAAC	22	166
Tag Reverse Primer	AcDx-5117-KRBA1-S1-RT-RP	TGAATGCAAAGATCGCGGAAAC	22	167
Downstream PCR Primer	AcDx-5118-KRBA1-S1-PCR-V	TGAATGCAAAGATCGCGGAAACCCGCTTATCACTCATTCATTCCTGrCAACT/3SpC3/	51	168
GATA5-1				
Forward PCR Primer	AcDx-5121-GATA5-S1-FP	CGGTAGGGTACGGCGrGTGGT/3SpC3/	20	169
Reverse PCR Primer	AcDx-5122-GATA5-S1-RP	GGTGTCGTGGACTACGAAACCTCAACGACCrCGAAT/3SpC3/	35	170
Upstream LDR	AcDx-5123-GATA5-S1-Up	TCCTAGTACCTACAGTGGGCAACGGTGGCGGTGGATCrGGCAG/3SpC3/	42	171
Downstream LDR	AcDx-5124-GATA5-S1-Dn	/5Phos/GGCGATCGGCGGGTCGAAGATTGACCGCTGTTATACGTTGCA	42	172
Real-Time Probe	AcDx-5125-GATA5-S1-RT-Pb	/56-FAM/AAGTGGATC/ZEN/GGCGATCGGCGG/3IABkFQ/	21	173
Tag Forward Primer	AcDx-5126-GATA5-S1-RT-FP	TCCTAGTACCTACAGTGGGCAA	22	174
Tag Reverse Primer	AcDx-5127-GATA5-S1-RT-RP	TGCAACGTATAACAGCGGTCAA	22	175
Downstream PCR Primer	AcDx-5128-GATA5-S1-PCR-V	TGCAACGTATAACAGCGGTCAACGACCCGAACTTCCAATCTTTGrACCCA/3SpC3/	49	176
CCDC48				
Forward PCR Primer	AcDx-5131-CCDC48-S1-FP	GGTCGTTTTTGGAAGTGGCrGGGAT/3SpC3/	24	177
Reverse PCR Primer	AcDx-5132-CCDC48-S1-RP	GGTGTCGTGGCGCCCGTACTAACCGACrUCCAC/3SpC3/	32	178
Upstream LDR	AcDx-5133-CCDC48-S1-Up	TAGACACGAGCGAGGTCACGGCGGAGCGAAGGGTTCrGGAAA/3SpC3/	41	179
Downstream LDR	AcDx-5134-CCDC48-S1-Dn	/5Phos/GGAGGAGGGATCGGAGGAGCGAGTGCAAAATTCAGGCTGTGCA	43	180
Real-Time Probe	AcDx-5135-CCDC48-S1-RT-Pb	/56-FAM/TTAGGGTTC/ZEN/GGAGGAGGGATCGG/3IABkFQ/	23	181

Tag Forward Primer	AcDx-5136-CCDC48-S1-RT-FP	TAGACACGAGCGAGGTCAC	19	182
Tag Reverse Primer	AcDx-5137-CCDC48-S1-RT-RP	TGCACAGCCTGAATTTTGCAC	21	183
Downstream PCR Primer	AcDx-5138-CCDC48-S1-PCR-V	TGCACAGCCTGAATTTTGCACACTAACCGACTCCATCGCTATGrCCGCA/3SpC3/	48	184
SEPT_9-RC				
Forward PCR Primer	AcDx-5261-SEPT-9(RC)-S2-FP	GTAGATGTTTTTTATTAAATGTTAAGAGTAATrATTTC/3SpC3/	37	185
Reverse PCR Primer	AcDx-5262-SEPT-9(RC)-S2-RP	GGTGTCGTGGCACATTCCTACTCCCCAACCrCTTTT/3SpC3/	35	186
Upstream LDR	AcDx-5263-SEPT-9(RC)-S2-Up	TAATCTCCAGACCTCCGAACCGTTAAGAGTAATATTTTTTTTTATTTTTTGTGAGTTAGCrGGTTA/3SpC3/	65	187
Downstream LDR	AcDx-5264-SEPT-9(RC)-S2-Dn	/5Phos/GGTCGCGTTTTTGTATGGAGGTGCGGGTGTAAGGATTGAACGGGACA	47	188
Real-Time Probe	AcDx-5265-SEPT-9(RC)-S2-RT-Pb	/56-FAM/AAAGTTAGC/ZEN/GGTCGCGTTTTTGTATGGG/3IABkFQ/	28	189
Tag Forward Primer	AcDx-5266-SEPT-9(RC)-S2-RT-FP	TAATCTCCAGACCTCCGAACC	21	190
Tag Reverse Primer	AcDx-5267-SEPT-9(RC)-S2-RT-RP	TGTCCCGTTCAATCCTTACACC	22	191
Downstream PCR Primer	AcDx-5268-SEPT-9(RC)-S2-PCR-V	TGTCCCGTTCAATCCTTACACCTCCCCAACCCTTTCCTCTGrCACCT/3SpC3/	46	192
BCAT1_S1				
Forward PCR Primer	AcDx-5271-BCAT-S1-S1-FP	TAGTTTCGTGGTTTTGTTAATTATAGACrGTATG/3SpC3/	33	193
Reverse PCR Primer	AcDx-5272-BCAT-S1-S1-RP	GGTGTCGTGGTTAAAATATAAAATAAATACTACGACATCGAArAAACT/3SpC3/	47	194
Upstream LDR	AcDx-5273-BCAT-S1-S1-Up	TACGAATCACCCGAGAGTTCAATGTTAATTATAGACGTATAATAGTAAGTTTGTAAAGGGCACrGGGAG/3SpC3/	68	195
Downstream LDR	AcDx-5274-BCAT-S1-S1-Dn	/5Phos/GGAGACGGGCGTGAATTATTTTTTTTATTAGTAGGGTTGTGGGTGGGTATAGGTCAGA	58	196
Real-Time Probe	AcDx-5275-BCAT-S1-S1-RT-Pb	/56-FAM/TTAGGGCAC/ZEN/GGAGACGGGC/3IABkFQ/	19	197
Tag Forward Primer	AcDx-5276-BCAT-S1-S1-RT-FP	TACGAATCACCCGAGAGTTCAA	22	198

Tag Reverse Primer	AcDx-5277-BCAT-S1-S1-RT-RP	TCTGACCTATACCCACCCACAA	22	199
Downstream PCR Primer	AcDx-5278-BCAT-S1-S1-PCR-V	TCTGACCTATACCCACCCACAATCGAAAAACCCTACTAATAAAAAAAATAATTCATGrCCCGC/3SpC3/	62	200
BCAT1_S2				
Forward PCR Primer	AcDx-5281-BCAT-S2-S1-FP	TGTTGTAAGTAAATGATACGGTTATTTTCrGAATC/3SpC3/	34	201
Reverse PCR Primer	AcDx-5282-BCAT-S2-S1-RP	GGTGTCGTGGCCTCTAATCTATTTATTTCCTCCTCCTTArACGTT/3SpC3/	44	202
Upstream LDR	AcDx-5283-BCAT-S2-S1-Up	TGCTTACCCACGATGCACCTTTCGAATTAGTTATTGTGGGTGCGTACTCrGAGCA/3SpC3/	54	203
Downstream LDR	AcDx-5284-BCAT-S2-S1-Dn	/5Phos/GAGTGTGGAGATGTTCGTGTAATATTTATATGGAGACGTGGTCGTATGACTTGCTCGCA	59	204
Real-Time Probe	AcDx-5285-BCAT-S2-S1-RT-Pb	/56-FAM/AACGTACTC/ZEN/GAGTGTGGAGATGTTCGTGT/3IABkFQ/	29	205
Tag Forward Primer	AcDx-5286-BCAT-S2-S1-RT-FP	TGCTTACCCACGATGCACC	19	206
Tag Reverse Primer	AcDx-5287-BCAT-S2-S1-RT-RP	TGCGAGCAAGTCATACGACC	20	207
Downstream PCR Primer	AcDx-5288-BCAT-S2-S1-PCR-V	TGCGAGCAAGTCATACGACCCCTCTAATCTATTTATTTCCTCCTCCTTAATGrUCTCT/3SpC3/	57	208
IKZF1				
Forward PCR Primer	AcDx-5291-IKZF1-S1-FP	CGGGGACGGGACGACrGTATC/3SpC3/	20	209
Reverse PCR Primer	AcDx-5292-IKZF1-S1-RP	GGTGTCGTGGGCTACTCCGATACAAAAAACGAArACGCA/3SpC3/	38	210
Upstream LDR	AcDx-5293-IKZF1-S1-Up	TGGACACTTCGCCCTTCTTAACGGGACGGGACGACGTATTTTTCTCrGTGCC/3SpC3/	51	211
Downstream LDR	AcDx-5294-IKZF1-S1-Dn	/5Phos/GTGTTTCGTTTTGCGTTTTTTTGCGCGTGTTTGGGATCTGGGCATCACA	49	212
Real-Time Probe	AcDx-5295-IKZF1-S1-RT-Pb	/56-FAM/CCTTTTCTC/ZEN/GTGTTTCGTTTTGCGTTTTTTTGC/3IABkFQ/	33	213
Tag Forward Primer	AcDx-5296-IKZF1-S1-RT-FP	TGGACACTTCGCCCTTCTTAAC	22	214
Tag Reverse Primer	AcDx-5297-IKZF1-S1-RT-RP	TGTGATGCCCAGATCCCAAAC	21	215
Downstream PCR Primer	AcDx-5298-IKZF1-S1-PCR-V	TGTGATGCCCAGATCCCAAACGCTACTCCGATACAAAAAACGAAATGrCGCAG/3SpC3/	52	216
NPY-1				
Forward PCR Primer	AcDx-5301-NPY-S1-FP	CGAGGAAGTTTTATAAAAGTTTTGTCrGCGAC/3SpC3/	31	217

Reverse PCR Primer	AcDx-5302-NPY-S1-RP	GGTGTCGTGGCTAAACGACAAATACTATACTATCGAACrGAACA/3SpC3/	43	218
Upstream LDR	AcDx-5303-NPY-S1-Up	TCAAACAAAGGCGACCACAACGAGGAAGTTTTATAAAAGTTTTGTCGCGACTCrGTTCC/3SpC3/	58	219
Downstream LDR	AcDx-5304-NPY-S1-Dn	/5Phos/GTTTTTTGTATTTTATTCGTTGGTTTTTATTTTTCGGAGACGTTGTTGTCGCATAGGCAGTTCATA	66	220
Real-Time Probe	AcDx-5305-NPY-S1-RT-Pb	/56-FAM/AAGCGACTC/ZEN/GTTTTTTGTATTTTATTCGTTGGTTTTTAT/3IABkFQ/	39	221
Tag Forward Primer	AcDx-5306-NPY-S1-RT-FP	TCAAACAAAGGCGACCACAAC	21	222
Tag Reverse Primer	AcDx-5307-NPY-S1-RT-RP	TATGAACTGCCTATGCGACAAC	22	223
Downstream PCR Primer	AcDx-5308-NPY-S1-PCR-V	TATGAACTGCCTATGCGACAACATACTATACTATCGAACGAACGTCTCTGrAAAAG/3SpC3/	55	224
VIPR2-1				
Forward PCR Primer	AcDx-5311-VIPR2-S1-FP	CGTATTTCGAGTTTAGCGTGTCrGGGAA/3SpC3/	27	225
Reverse PCR Primer	AcDx-5312-VIPR2-S1-RP	GGTGTCGTGGCCGACCCATACTAAAAACGACrGAAAT/3SpC3/	36	226
Upstream LDR	AcDx-5313-VIPR2-S1-Up	TCGCAACGTGCCGAATACACGTGTCGGAGGTCGTTTGGCrGTTTA/3SpC3/	44	227
Downstream LDR	AcDx-5314-VIPR2-S1-Dn	/5Phos/GTTCGTCGTTTTCGTTTTAGGTTTTCGCGGTTGTTGCACGGTCGAGCTAA	50	228
Real-Time Probe	AcDx-5315-VIPR2-S1-RT-Pb	/56-FAM/AAGTTTGGC/ZEN/GTTCGTCGTTTTCGTTTTAGG/3IABkFQ/	30	229
Tag Forward Primer	AcDx-5316-VIPR2-S1-RT-FP	TCGCAACGTGCCGAATACA	19	230
Tag Reverse Primer	AcDx-5317-VIPR2-S1-RT-RP	TTAGCTCGACCGTGCAACA	19	231
Downstream PCR Primer	AcDx-5318-VIPR2-S1-PCR-V	TTAGCTCGACCGTGCAACACATACTAAAAACGACGAAACCGTGrAAAAT/3SpC3/	48	232
THBD				
Forward PCR Primer	AcDx-5331-THBD-S1-FP	TATAGGACGTCGATGGCGATArGTTTC/3SpC3/	26	233
Reverse PCR Primer	AcDx-5332-THBD-S1-RP	GGTGTCGTGGCGATCCGCATATCAAAAACTACCrUCGCG/3SpC3/	38	234
Upstream LDR	AcDx-5333-THBD-S1-Up	TTCAGAGCACCTGCGTACCACGTCGATGGCGATAGTTTTTTTTGCTCrGTTCC/3SpC3/	52	235
Downstream LDR	AcDx-5334-THBD-S1-Dn	/5Phos/GTTTTAGTTTAGATATTTTTTGTCGTTGCGCGTAGTTTTTGGGTTCTTCGGCTGGCTCAA	60	236
Real-Time Probe	AcDx-5335-THBD-S1-RT-Pb	/56-FAM/AATTTGCTC/ZEN/GTTTTAGTTTAGATATTTTTTGTCGTTGCG/3IABkFQ/	39	237
Tag Forward Primer	AcDx-5336-THBD-S1-RT-FP	TTCAGAGCACCTGCGTACC	19	238
Tag Reverse Primer	AcDx-5337-THBD-S1-RT-RP	TTGAGCCAGCCGAAGAACC	19	239
Downstream PCR Primer	AcDx-5338-THBD-S1-PCR-V	TTGAGCCAGCCGAAGAACCCATATCAAAAACTACCTCGCAAAAACTATGrCGCAG/3SpC3/	54	240

SEPT9				
Forward PCR Primer	AcDx-5351-SEPT9-S4-FP	GTGGGTGTTGGGTTGGTrUGTTA/3SpC3/	22	241
Reverse PCR Primer	AcDx-5352-SEPT9-S4-RP	GGTGTCGTGGCAAACCCACCCGCAAAArUCCTT/3SpC3/	32	242
Upstream LDR	AcDx-5353-SEPT9-S4-Up	TACACGTGGATATCTCCGACCGGGTGTTGGGTTGGTTGTCGCrGGTTA/3SpC3/	47	243
Downstream LDR	AcDx-5354-SEPT9-S4-Dn	/5Phos/GGTCGCGGACGTGTTGGAGAGGGGTGCTAGTCACACAGTTCCA	43	244
Real-Time Probe	AcDx-5355-SEPT9-S4-RT-Pb	/56-FAM/TATTGTCGC/ZEN/GGTCGCGGACG/3IABkFQ/	20	245
Tag Forward Primer	AcDx-5356-SEPT9-S4-RT-FP	TACACGTGGATATCTCCGACC	21	246
Tag Reverse Primer	AcDx-5357-SEPT9-S4-RT-RP	TGGAACTGTGTGACTAGCACC	21	247
Downstream PCR Primer	AcDx-5358-SEPT9-S4-PCR-V	TGGAACTGTGTGACTAGCACCCCGCAAAATCCTCTCCAACATGrUCCGT/3SpC3/	48	248
GATA5-2				
Forward PCR Primer	AcDx-5361-GATA5-S1-FP	GGTAGGGAGGGCGGTArGGGTG/3SpC3/	21	249
Reverse PCR Primer	AcDx-5362-GATA5-S1-RP	GGTGTCGTGGCGCGAAAAAAACTACGAAACCrUCAAT/3SpC3/	36	250
Upstream LDR	AcDx-5363-GATA5-S1-Up	TCCGGGTATACACTGTCCCATGGGTCGGCGATCAGCrGGGCT/3SpC3/	41	251
Downstream LDR	AcDx-5364-GATA5-S1-Dn	/5Phos/GGGTCGAAGATTGGAAGTTCGGGTCGTGGTTAACAGAGGACAGGCCA	47	252
Real-Time Probe	AcDx-5365-GATA5-S1-RT-Pb	/56-FAM/TTGATCAGC/ZEN/GGGTCGAAGATTGGAAGT/3IABkFQ/	27	253
Tag Forward Primer	AcDx-5366-GATA5-S1-RT-FP	TCCGGGTATACACTGTCCCA	20	254
Tag Reverse Primer	AcDx-5367-GATA5-S1-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	255
Downstream PCR Primer	AcDx-5368-GATA5-S1-PCR-V	TGGCCTGTCCTCTGTTAACCAGCGAAAAAAACTACGAAACCTCAATGrACCCA/3SpC3/	52	256
SPG20-1				
Forward PCR Primer	AcDx-5371-SPG20-S1-FP	GGGTTTAGGGCGTTTATTTCrGTTTG/3SpC3/	25	257
Reverse PCR Primer	AcDx-5372-SPG20-S1-RP	GGTGTCGTGGCGAAACGCCCGAAATCTAAArAAACA/3SpC3/	35	258
Upstream LDR	AcDx-5373-SPG20-S1-Up	TTCAGCAGCCTGGCATCACAGGGCGTTTATTTCGTTTATTTAAGGGTTCrGCGCG/3SpC3/	54	259
Downstream LDR	AcDx-5374-SPG20-S1-Dn	/5Phos/GCGTAGGAGTGAATTTTTTCGCGTTTTATTTGCGTGGAGCGCTAAGGTTGCA	52	260
Real-Time Probe	AcDx-5375-SPG20-S1-RT-Pb	/56-FAM/CCAGGGTTC/ZEN/GCGTAGGAGTGAATTTTTTCG/3IABkFQ/	30	261
Tag Forward Primer	AcDx-5376-SPG20-S1-RT-FP	TTCAGCAGCCTGGCATCAC	19	262
Tag Reverse Primer	AcDx-5377-SPG20-S1-RT-RP	TGCAACCTTAGCGCTCCAC	19	263
Downstream PCR	AcDx-5378-SPG20-S1-PCR-V	TGCAACCTTAGCGCTCCACCCCGAAATCTAAAAAACGCAAATAAAATGrCGAAG/3SpC3/	53	264

Primer				
TRPS1				
Forward PCR Primer	AcDx-5391-TRPS1-S1-FP	ATGTTTTATCGTTGTTAGGTATTTAATTATCrGGTTG/3SpC3/	36	265
Reverse PCR Primer	AcDx-5392-TRPS1-S1-RP	GGTGTCGTGGCCGTAAAAACTAAAAAAAAAACAAACTTCrCTCTG/3SpC3/	44	266
Upstream LDR	AcDx-5393-TRPS1-S1-Up	TCAGACGCACTAAACAGGCAATGTTAGGTATTTAATTATCGGTTAGTGTTTTTTGACTCrG CGTA/3SpC3/	64	267
Downstream LDR	AcDx-5394-TRPS1-S1-Dn	/5Phos/GCGCGATATATGGCGTATTAATCGTATCGTAGAGGTTGCGGATCGTCGTGTGAA	54	268
Real-Time Probe	AcDx-5395-TRPS1-S1-RT-Pb	/56-FAM/AATTGACTC/ZEN/GCGCGATATATGGCGTATTAATC/3IABkFQ/	32	269
Tag Forward Primer	AcDx-5396-TRPS1-S1-RT-FP	TCAGACGCACTAAACAGGCAA	21	270
Tag Reverse Primer	AcDx-5397-TRPS1-S1-RT-RP	TTCACACGACGATCCGCAA	19	271
Downstream PCR Primer	AcDx-5398-TRPS1-S1-PCR-V	TTCACACGACGATCCGCAACGTAAAAACTAAAAAAAAAACAAACTTCCTCTATGrATACA/ 3SpC3/	59	272
SEMA3B				
Forward PCR Primer	AcDx-5401-SEMA3B-S1-FP	CGTCGCGTGTTAGGGTTCrGGAAA/3SpC3/	23	273
Reverse PCR Primer	AcDx-5402-SEMA3B-S1-RP	GGTGTCGTGGCGATACGCTCCTCTACCAACrACCTG/3SpC3/	35	274
Upstream LDR	AcDx-5403-SEMA3B-S1-Up	TCCTGCTCTGAAAACCTACACCCGTGTTAGGGTTCGGAAGTTTTGTTCTCrGGTCT/3SpC3/	55	275
Downstream LDR	AcDx-5404-SEMA3B-S1-Dn	/5Phos/GGTTCGATATTTTCGTTTTACGTTGTTTTTTGTTCGTAGGGGTTACATAGGCGGCT TAGACA	62	276
Real-Time Probe	AcDx-5405-SEMA3B-S1-RT-Pb	/56-FAM/AATGTTCTC/ZEN/GGTTCGATATTTTCGTTTTACGTTGT/3IABkFQ/	35	277
Tag Forward Primer	AcDx-5406-SEMA3B-S1-RT-FP	TCCTGCTCTGAAAACCTACACC	22	278
Tag Reverse Primer	AcDx-5407-SEMA3B-S1-RT-RP	TGTCTAAGCCGCCTATGTAACC	22	279
Downstream PCR Primer	AcDx-5408-SEMA3B-S1-PCR-V	TGTCTAAGCCGCCTATGTAACCCGCTCCTCTACCAACACCTATGrAACAG/3SpC3/	49	280
VIPR2-2				
Forward PCR Primer	AcDx-5411-VIPR2-S2-FP	GTAGTCGAGCGTTCGAGCrGCGGA/3SpC3/	23	281
Reverse PCR Primer	AcDx-5412-VIPR2-S2-RP	GGTGTCGTGGCGCGTAAAAAACGAACGTACArUTAAG/3SpC3/	36	282
Upstream LDR	AcDx-5413-VIPR2-S2-Up	TCCTCGAGCCGATGACACAGCGTTCGAGCGCAGGGATCrGTTCC/3SpC3/	43	283

Downstream LDR	AcDx-5414-VIPR2-S2-Dn	/5Phos/GTTTTTTTCGTTTCGGTAGTTGGAGCGCGTGTAACGTCCGTGGGCTAA	48	284
Real-Time Probe	AcDx-5415-VIPR2-S2-RT-Pb	/56-FAM/TTAGGGATC/ZEN/GTTTTTTTCGTTTCGGTAGTTGG/3IABkFQ/	32	285
Tag Forward Primer	AcDx-5416-VIPR2-S2-RT-FP	TCCTCGAGCCGATGACACA	19	286
Tag Reverse Primer	AcDx-5417-VIPR2-S2-RT-RP	TTAGCCCACGGACGTTACA	19	287
Downstream PCR Primer	AcDx-5418-VIPR2-S2-PCR-V	TTAGCCCACGGACGTTACACGCGTAAAAAACGAACGTACATTAAATGrCGCTT/3SpC3/	52	288
GATA5-3				
Forward PCR Primer	AcDx-5421-GATA5-S1-FP	CGCGGTCGTAGGACGTArGGGTC/3SpC3/	22	289
Reverse PCR Primer	AcDx-5422-GATA5-S1-RP	GGTGTCGTGGTCCAACCCGAACTACAACCrGCGCA/3SpC3/	34	290
Upstream LDR	AcDx-5423-GATA5-S1-Up	TTGTCTCTGCGACCCATCAAGTAGGACGTAGGGTTTGGAGGGCrGGGAC/3SpC3/	48	291
Downstream LDR	AcDx-5424-GATA5-S1-Dn	/5Phos/GGGATTTCGTCGCGTTGGGAGGGTTGGTACACGTTCGGCACA	42	292
Real-Time Probe	AcDx-5425-GATA5-S1-RT-Pb	/56-FAM/AAGGAGGGC/ZEN/GGGATTTCGTCGC/3IABkFQ/	22	293
Tag Forward Primer	AcDx-5426-GATA5-S1-RT-FP	TTGTCTCTGCGACCCATCAA	20	294
Tag Reverse Primer	AcDx-5427-GATA5-S1-RT-RP	TGTGCCGAACGTGTACCAA	19	295
Downstream PCR Primer	AcDx-5428-GATA5-S1-PCR-V	TGTGCCGAACGTGTACCAACCCGACCCCTCCCAATGrCGACA/3SpC3/	41	296
ZNF542				
Forward PCR Primer	AcDx-5431-ZNF542-S1-FP	CGTTTTTGTATTTCGGTTATTGGGArGCGGA/3SpC3/	30	297
Reverse PCR Primer	AcDx-5432-ZNF542-S1-RP	GGTGTCGTGGACGCCCGAATAATTTCTAAAAATAAACrGAAAG/3SpC3/	42	298
Upstream LDR	AcDx-5433-ZNF542-S1-Up	TTTCGCTCGACGCATACCACGGTTATTGGGAGCGGGATCrGTGAA/3SpC3/	44	299
Downstream LDR	AcDx-5434-ZNF542-S1-Dn	/5Phos/GTGGGAGTTGTATATGCGTATTGCGAGTTTTCTGGCGCGGCTACTGTAAAA	51	300
Real-Time Probe	AcDx-5435-ZNF542-S1-RT-Pb	/56-FAM/TTCGGGATC/ZEN/GTGGGAGTTGTATATGCG/3IABkFQ/	27	301
Tag Forward Primer	AcDx-5436-ZNF542-S1-RT-FP	TTTCGCTCGACGCATACCA	19	302
Tag Reverse Primer	AcDx-5437-ZNF542-S1-RT-RP	TTTTACAGTAGCCGCGCCA	19	303
Downstream PCR Primer	AcDx-5438-ZNF542-S1-PCR-V	TTTTACAGTAGCCGCGCCACCCGAATAATTTCTAAAAATAAACGAAAACTTGrCAATG/3SpC3/	57	304
RCN3				
Forward PCR Primer	AcDx-5441-RCN3-S1-FP	CGTGAGGCGTTGTGATTAGAATArGTTGA/3SpC3/	28	305
Reverse PCR Primer	AcDx-5442-RCN3-S1-RP	GGTGTCGTGGTAACGCGACCGAAAAAAACTACrAACTT/3SpC3/	37	306

Upstream LDR	AcDx-5443-RCN3-S1-Up	TTGCACGTTGTCCTGCACCCGTTGTGATTAGAATAGTTGGAGGTGAACrGGTGA/3SpC3/	53	307
Downstream LDR	AcDx-5444-RCN3-S1-Dn	/5Phos/GGTAGAGTGTCGCGACGATTGTTAGGAGTGGTAGTTTCCCATGACGGCA	49	308
Real-Time Probe	AcDx-5445-RCN3-S1-RT-Pb	/56-FAM/TTGGTGAAC/ZEN/GGTAGAGTGTCGCGAC/3IABkFQ/	25	309
Tag Forward Primer	AcDx-5446-RCN3-S1-RT-FP	TTGCACGTTGTCCTGCACC	19	310
Tag Reverse Primer	AcDx-5447-RCN3-S1-RT-RP	TGCCGTCATGGGAAACTACC	20	311
Downstream PCR Primer	AcDx-5448-RCN3-S1-PCR-V	TGCCGTCATGGGAAACTACCACCGAAAAAAACTACAACTCCTAACAATTGrUCGCA/3SpC3/	55	312
MYO15B				
Forward PCR Primer	AcDx-5451-MYO15B-S1-FP	TTTAGGAGTTTTAATGGAGATACGTCrGGGTA/3SpC3/	31	313
Reverse PCR Primer	AcDx-5452-MYO15B-S1-RP	GGTGTCGTGGCCGAACTATACCGCGCTAACrUACCA/3SpC3/	35	314
Upstream LDR	AcDx-5453-MYO15B-S1-Up	TTAGCCGCCAAACGTACCATGGGAACGGAGGTAGTTTTTGCTCrGGACG/3SpC3/	48	315
Downstream LDR	AcDx-5454-MYO15B-S1-Dn	/5Phos/GGATAGCGAAATTCGCGAGGTTTAGGAGAGTGGGCAGGAACACGATAGTA	50	316
Real-Time Probe	AcDx-5455-MYO15B-S1-RT-Pb	/56-FAM/CCTTTGCTC/ZEN/GGATAGCGAAATTCGCGA/3IABkFQ/	27	317
Tag Forward Primer	AcDx-5456-MYO15B-S1-RT-FP	TTAGCCGCCAAACGTACCA	19	318
Tag Reverse Primer	AcDx-5457-MYO15B-S1-RT-RP	TACTATCGTGTTCCTGCCCA	20	319
Downstream PCR Primer	AcDx-5458-MYO15B-S1-PCR-V	TACTATCGTGTTCCTGCCCACGAACTATACCGCGCTAACTACTGrCTCTT/3SpC3/	49	320
ANKRD13B				
Forward PCR Primer	AcDx-5461-ANKRD13B-S1-FP	CGAGTAGTTGCGGTTGGCrGATGA/3SpC3/	23	321
Reverse PCR Primer	AcDx-5462-ANKRD13B-S1-RP	GGTGTCGTGGCCAACTCCTCCTCCTCCTAArCGCGT/3SpC3/	35	322
Upstream LDR	AcDx-5463-ANKRD13B-S1-Up	TTCGTACCTCGGCACACCAGCGGTTGGCGATGGAATTATCrGGCAC/3SpC3/	45	323
Downstream LDR	AcDx-5464-ANKRD13B-S1-Dn	/5Phos/GGCGTAGGAGTAGGAGGAGAGGCGTGGCTCCGTTACTCTGTCGA	44	324
Real-Time Probe	AcDx-5465-ANKRD13B-S1-RT-Pb	/56-FAM/CCAATTATC/ZEN/GGCGTAGGAGTAGGAGGAGAGG/3IABkFQ/	31	325
Tag Forward Primer	AcDx-5466-ANKRD13B-S1-RT-FP	TTCGTACCTCGGCACACCA	19	326

Tag Reverse Primer	AcDx-5467-ANKRD13B-S1-RT-RP	TCGACAGAGTAACGGAGCCA	20	327
Downstream PCR Primer	AcDx-5468-ANKRD13B-S1-PCR-V	TCGACAGAGTAACGGAGCCACCAACTCCTCCTCCTCCTAATGrCGCGT/3SpC3/	47	328
FAM155A				
Forward PCR Primer	AcDx-5471-FAM115A-S1-FP	AGGTTGGTGTTGGTGGTCrGGCGA/3SpC3/	23	329
Reverse PCR Primer	AcDx-5472-FAM115A-S1-RP	GGTGTCGTGGCGCTAACAATACCTAAATAACCGAAACrCGCGT/3SpC3/	42	330
Upstream LDR	AcDx-5473-FAM115A-S1-Up	TTTGCCTCTTGTAGGTGCCAGAGGTTGGGTGTAGGGAGCrGATAA/3SpC3/	44	331
Downstream LDR	AcDx-5474-FAM115A-S1-Dn	/5Phos/GATGGTGGAGGTGATAGGGTGGTTGGTGGGCAACGCGGATATTCA	45	332
Real-Time Probe	AcDx-5475-FAM115A-S1-RT-Pb	/56-FAM/AAAGGGAGC/ZEN/GATGGTGGAGGTGA/3IABkFQ/	23	333
Tag Forward Primer	AcDx-5476-FAM115A-S1-RT-FP	TTTGCCTCTTGTAGGTGCCA	20	334
Tag Reverse Primer	AcDx-5477-FAM115A-S1-RT-RP	TGAATATCCGCGTTGCCCA	19	335
Downstream PCR Primer	AcDx-5478-FAM115A-S1-PCR-V	TGAATATCCGCGTTGCCCATAACAATACCTAAATAACCGAAACCGTGrCCAAT/3SpC3/	52	336
FLI1-1				
Forward PCR Primer	AcDx-5481-FLI1-S1-FP	GATTTGGTTAAATGGACGGGATTArUTAAA/3SpC3/	29	337
Reverse PCR Primer	AcDx-5482-FLI1-S1-RP	GGTGTCGTGGTCGCCTCCCCGACCrGATCT/3SpC3/	29	338
Upstream LDR	AcDx-5483-FLI1-S1-Up	TCGTAGACTCGCTATCGCCAGGACGGGATTATTAAGGTAAGCAGCrGGGAC/3SpC3/	50	339
Downstream LDR	AcDx-5484-FLI1-S1-Dn	/5Phos/GGAGTAACGGACGCGGGCGGTGGTGAGCAGGGATGAGCA	39	340
Real-Time Probe	AcDx-5485-FLI1-S1-RT-Pb	/56-FAM/ACAAGCAGC/ZEN/GGAGTAACGGACGC/3IABkFQ/	23	341
Tag Forward Primer	AcDx-5486-FLI1-S1-RT-FP	TCGTAGACTCGCTATCGCCA	20	342
Tag Reverse Primer	AcDx-5487-FLI1-S1-RT-RP	TGCTCATCCCTGCTCACCA	19	343
Downstream PCR Primer	AcDx-5488-FLI1-S1-PCR-V	TGCTCATCCCTGCTCACCATCGCCTCCCCGACTGrATCCT/3SpC3/	39	344
RGS10				
Forward PCR Primer	AcDx-5491-RGS10-S1-FP	CGTTCGTAGCGGAGGCrGGAGG/3SpC3/	21	345
Reverse PCR Primer	AcDx-5492-RGS10-S1-RP	GGTGTCGTGGAAAAACGCCCCAAATCTCCrAAACG/3SpC3/	34	346

Upstream LDR	AcDx-5493-RGS10-S1-Up	TCCCTCGTCATCTCCCTTACCCGGAGGGAGAAGTTCGTGCrGTCAC/3SpC3/	45	347
Downstream LDR	AcDx-5494-RGS10-S1-Dn	/5Phos/GTCGTTTCGTTTTCGGAATTTGGAGTTTTATGTTATTTTGGTCTTGGTGATGGAGCGA	58	348
Real-Time Probe	AcDx-5495-RGS10-S1-RT-Pb	/56-FAM/CCTTCGTGC/ZEN/GTCGTTTCGTTTTCGGA/3IABkFQ/	26	349
Tag Forward Primer	AcDx-5496-RGS10-S1-RT-FP	TCCCTCGTCATCTCCCTTACC	21	350
Tag Reverse Primer	AcDx-5497-RGS10-S1-RT-RP	TCGCTCCATCACCAAGACC	19	351
Downstream PCR Primer	AcDx-5498-RGS10-S1-PCR-V	TCGCTCCATCACCAAGACCCCAAACTTTAAAAATAACATAAAACTCCAAATTCTGrAAAAT/3SpC3/	60	352
HCG4				
Forward PCR Primer	AcDx-5501-HCG4-S1-FP	GTCGGAATATTGGGAAGAGGArGATAA/3SpC3/	26	353
Reverse PCR Primer	AcDx-5502-HCG4-S1-RP	GGTGTCGTGGCCTCACTCTAATTATAATAACCGCTCrAAAAC/3SpC3/	41	354
Upstream LDR	AcDx-5503-HCG4-S1-Up	TTCTAGATACCACGGACGCACCGGAATATTGGGAAGAGGAGATAGGGTTCrGTTGG/3SpC3/	55	355
Downstream LDR	AcDx-5504-HCG4-S1-Dn	/5Phos/GTTAAGGTTAAAGTATAGTTTTATCGAGTGAATTTGCGGATTTTGTGTTGGTGTGCAAAGCTGA	64	356
Real-Time Probe	AcDx-5505-HCG4-S1-RT-Pb	/56-FAM/CCAGGGTTC/ZEN/GTTAAGGTTAAAGTATAGTTTTATCGAGTGA/3IABkFQ/	40	357
Tag Forward Primer	AcDx-5506-HCG4-S1-RT-FP	TTCTAGATACCACGGACGCAC	21	358
Tag Reverse Primer	AcDx-5507-HCG4-S1-RT-RP	TCAGCTTTGCACACCAACAC	20	359
Downstream PCR Primer	AcDx-5508-HCG4-S1-PCR-V	TCAGCTTTGCACACCAACACCTCACTCTAATTATAATAACCGCTCAAAATCTGrCAAAC/3SpC3/	58	360
STK32B				
Forward PCR Primer	AcDx-5511-STK32B-S1-FP	CGCGCGCGGTTATAATTCrGGATC/3SpC3/	23	361
Reverse PCR Primer	AcDx-5512-STK32B-S1-RP	GGTGTCGTGGTACATACCCGACCGCTAAAATACrCGAAC/3SpC3/	38	362
Upstream LDR	AcDx-5513-STK32B-S1-Up	TCGCTCTTCAGCCTCCTACATTATAATTCGGATTGGGCGCGTTCTCrGGTGC/3SpC3/	51	363
Downstream LDR	AcDx-5514-STK32B-S1-Dn	/5Phos/GGTATTTCGTATTTTTGCGCGCGTTTTATATTTCGTATTCTGTTCTGGGAATTATTGCCGGA	62	364
Real-Time Probe	AcDx-5515-STK32B-S1-RT-Pb	/56-FAM/TTCGTTCTC/ZEN/GGTATTTCGTATTTTTGCGCG/3IABkFQ/	30	365
Tag Forward Primer	AcDx-5516-STK32B-S1-RT-FP	TCGCTCTTCAGCCTCCTACA	20	366
Tag Reverse Primer	AcDx-5517-STK32B-S1-RT-RP	TCCGGCAATAATTCCCAGAACA	22	367
Downstream PCR Primer	AcDx-5518-STK32B-S1-PCR-V	TCCGGCAATAATTCCCAGAACACCGCTAAAATACCGAATACGAAATATAAAATGrCGCGT/3SpC3/	59	368

CNRIP1				
Forward PCR Primer	AcDx-5521-CNRIP1-S1-FP	TCGTTATTAGGTTGGATGCGTArGCGCA/3SpC3/	27	369
Reverse PCR Primer	AcDx-5522-CNRIP1-S1-RP	GGTGTCGTGGTCCGCCCCCCGAAArCCGCT/3SpC3/	29	370
Upstream LDR	AcDx-5523-CNRIP1-S1-Up	TACCCTCCTAGCTCCGTACACGTAGCGCGATGGAGAAGCrGTATA/3SpC3/	44	371
Downstream LDR	AcDx-5524-CNRIP1-S1-Dn	/5Phos/GTACGAGGTTCGGTAGGTTTTTTATGTTTGGGCTGTGTTGTCTGGTGGTGCA	52	372
Real-Time Probe	AcDx-5525-CNRIP1-S1-RT-Pb	/56-FAM/AAGAGAAGC/ZEN/GTACGAGGTTCGGTAGGT/3IABkFQ/	27	373
Tag Forward Primer	AcDx-5526-CNRIP1-S1-RT-FP	TACCCTCCTAGCTCCGTACA	20	374
Tag Reverse Primer	AcDx-5527-CNRIP1-S1-RT-RP	TGCACCACCAGACAACACA	19	375
Downstream PCR Primer	AcDx-5528-CNRIP1-S1-PCR-V	TGCACCACCAGACAACACACCGCGCCAAACCCTTGrCCCAG/3SpC3/	40	376
CYP1B1				
Forward PCR Primer	AcDx-5531-CYP1B1-S1-FP	AAGTTGCGGTTGAGTTGTTCrGAATC/3SpC3/	25	377
Reverse PCR Primer	AcDx-5532-CYP1B1-S1-RP	GGTGTCGTGGCTAATAAACGTAATACCCTAACTACAATACrUTCCT/3SpC3/	45	378
Upstream LDR	AcDx-5533-CYP1B1-S1-Up	TTGGCAACTCTCCACCCAAGCGGTTGAGTTGTTCGAATTTGCrGGAGG/3SpC3/	47	379
Downstream LDR	AcDx-5534-CYP1B1-S1-Dn	/5Phos/GGAAAACGGTGCGTATCGGGTTGGGTTGTATTGCGCCAGGATAGCA	46	380
Real-Time Probe	AcDx-5535-CYP1B1-S1-RT-Pb	/56-FAM/ACAATTTGC/ZEN/GGAAAACGGTGCGTATCG/3IABkFQ/	27	381
Tag Forward Primer	AcDx-5536-CYP1B1-S1-RT-FP	TTGGCAACTCTCCACCCAA	19	382
Tag Reverse Primer	AcDx-5537-CYP1B1-S1-RT-RP	TGCTATCCTGGCGCAATACAA	21	383
Downstream PCR Primer	AcDx-5538-CYP1B1-S1-PCR-V	TGCTATCCTGGCGCAATACAACCTAACTACAATACTTCCCCAACCTGrATACA/3SpC3/	52	384
CHST2				
Forward PCR Primer	AcDx-5541-CHST2-S1-FP	GGGCGTTTAGGGTCGTTTTCrGGTCA/3SpC3/	25	385
Reverse PCR Primer	AcDx-5542-CHST2-S1-RP	GGTGTCGTGGGCCCCGACGACGACAACGAAArAAACA/3SpC3/	36	386
Upstream LDR	AcDx-5543-CHST2-S1-Up	TTGCAAACCACCCGGACAAGGCGTTTAGGGTCGTTTTCGATCrGGGCT/3SpC3/	47	387
Downstream LDR	AcDx-5544-CHST2-S1-Dn	/5Phos/GGGTCGTTTCGTGTTTATGTTCGTTTGGATTTTCGTTGGTCAGCATCGACTCCTA	55	388
Real-Time Probe	AcDx-5545-CHST2-S1-RT-Pb	/56-FAM/CCTTCGATC/ZEN/GGGTCGTTTCGTGTTTATGT/3IABkFQ/	29	389
Tag Forward Primer	AcDx-5546-CHST2-S1-RT-FP	TTGCAAACCACCCGGACAA	19	390
Tag Reverse Primer	AcDx-5547-CHST2-S1-RT-RP	TAGGAGTCGATGCTGACCAA	20	391
Downstream PCR	AcDx-5548-CHST2-S1-PCR-V	TAGGAGTCGATGCTGACCAAGAAAAAACGATAAAAAATACGAAAATCCAAATGrAACAC/	58	392

Primer		3SpC3/		
SPOCK1				
Forward PCR Primer	AcDx-5551-SPOCK1-S1-FP	GCGTTACGAATTCGAGGTTGTCrGTTTG/3SpC3/	27	393
Reverse PCR Primer	AcDx-5552-SPOCK1-S1-RP	GGTGTCGTGGCGCTTCTAAAAAAAAATACCAAACCCrCTAAC/3SpC3/	41	394
Upstream LDR	AcDx-5553-SPOCK1-S1-Up	TCCGACTTTAGTGCGTCACAATCGAGGTTGTCGTTTAATTATAGAGGACTCrGTTCT/3SpC3/	56	395
Downstream LDR	AcDx-5554-SPOCK1-S1-Dn	/5Phos/GTTTCGTTTATTTTTCGAGTGTTTTATAGATGTCGTTTTTTTATTAGGGTTGTGGGTCTCGCTCGTATA	69	396
Real-Time Probe	AcDx-5555-SPOCK1-S1-RT-Pb	/56-FAM/TTGGGACTC/ZEN/GTTTCGTTTATTTTTCGAGTGTTTTATAGA/3IABkFQ/	39	397
Tag Forward Primer	AcDx-5556-SPOCK1-S1-RT-FP	TCCGACTTTAGTGCGTCACAA	21	398
Tag Reverse Primer	AcDx-5557-SPOCK1-S1-RT-RP	TATACGAGCGAGACCCACAA	20	399
Downstream PCR Primer	AcDx-5558-SPOCK1-S1-PCR-V	TATACGAGCGAGACCCACAATCTAAAAAAAAATACCAAACCCCTAATAAAAAAATGrACATT/3SpC3/	61	400
IRF4				
Forward PCR Primer	AcDx-5561-IRF4-S1-FP	GTTGCGGTAACGGGAAGTTTCrGTTAA/3SpC3/	26	401
Reverse PCR Primer	AcDx-5562-IRF4-S1-RP	GGTGTCGTGGAATACGAAAAATACTCTTCTCCTCGTTCrUCCCG/3SpC3/	43	402
Upstream LDR	AcDx-5563-IRF4-S1-Up	TTCTAGGCGACACGACAACAACGGGAAGTTTCGTTAGTGGTTGGTCrGATCG/3SpC3/	51	403
Downstream LDR	AcDx-5564-IRF4-S1-Dn	/5Phos/GATTAGATCGATAGCGGTAAGTATTTCGGGTTGGTGTGTGGGTACTGTCCGTGGA	55	404
Real-Time Probe	AcDx-5565-IRF4-S1-RT-Pb	/56-FAM/CCGTTGGTC/ZEN/GATTAGATCGATAGCGGTAAGTATTTCG/3IABkFQ/	37	405
Tag Forward Primer	AcDx-5566-IRF4-S1-RT-FP	TTCTAGGCGACACGACAACA	20	406
Tag Reverse Primer	AcDx-5567-IRF4-S1-RT-RP	TCCACGGACAGTACCCACA	19	407
Downstream PCR Primer	AcDx-5568-IRF4-S1-PCR-V	TCCACGGACAGTACCCACACGTTCTCCCACACCAACCTGrAAATG/3SpC3/	44	408
TWIST1				
Forward PCR Primer	AcDx-5571-TWIST1-S1-FP	GTCGGTGTTGTAGAGTTCGCrGAGGC/3SpC3/	25	409
Reverse PCR Primer	AcDx-5572-TWIST1-S1-RP	GGTGTCGTGGCCAATAACACTACTACCCCCAAACrUTTCT/3SpC3/	39	410
Upstream LDR	AcDx-5573-TWIST1-S1-Up	TCTGCCCAAAATACTGCACAACGAGGTGTTTGGGAGTTGAGCrGAGGA/3SpC3/	47	411

Downstream LDR	AcDx-5574-TWIST1-S1-Dn	/5Phos/GAGAGTTGTAGATTTGGAGGTTTTTATATTTTCGTGTAGGCGTTGAAACTGAGGCGGTGTTCA	63	412
Real-Time Probe	AcDx-5575-TWIST1-S1-RT-Pb	/56-FAM/TTGTTGAGC/ZEN/GAGAGTTGTAGATTTGGAGGTTT/3IABkFQ/	32	413
Tag Forward Primer	AcDx-5576-TWIST1-S1-RT-FP	TCTGCCCAAAATACTGCACAA	21	414
Tag Reverse Primer	AcDx-5577-TWIST1-S1-RT-RP	TGAACACCGCCTCAGTTTCAA	21	415
Downstream PCR Primer	AcDx-5578-TWIST1-S1-PCR-V	TGAACACCGCCTCAGTTTCAACCCAAACTTTCCGCCTACATGrAAAAC/3SpC3/	47	416
AEBP1				
Forward PCR Primer	AcDx-5581-AEBP1-S1-FP	GGAGACGGTTATTCGCGCrGGGAA/3SpC3/	23	417
Reverse PCR Primer	AcDx-5582-AEBP1-S1-RP	GGTGTCGTGGAAACTCCGAAACCAAAAAAACTCrAAAAG/3SpC3/	38	418
Upstream LDR	AcDx-5583-AEBP1-S1-Up	TTGCATTTCGTTAGCGACACAAGTGCGTTACGCGGGATCrGGAAT/3SpC3/	44	419
Downstream LDR	AcDx-5584-AEBP1-S1-Dn	/5Phos/GGAGCGTTTATTAGTCGTTAGGATTTCGGAGCGTGTGAGTCGATCTACCCGCA	53	420
Real-Time Probe	AcDx-5585-AEBP1-S1-RT-Pb	/56-FAM/TTCGGGATC/ZEN/GGAGCGTTTATTAGTCGTTAGG/3IABkFQ/	31	421
Tag Forward Primer	AcDx-5586-AEBP1-S1-RT-FP	TTGCATTTCGTTAGCGACACA	21	422
Tag Reverse Primer	AcDx-5587-AEBP1-S1-RT-RP	TGCGGGTAGATCGACTCACA	20	423
Downstream PCR Primer	AcDx-5588-AEBP1-S1-PCR-V	TGCGGGTAGATCGACTCACAAAAAACTCAAAAATAATCGAAACGCTCTGrAAATT/3SpC3/	54	424
TDH				
Forward PCR Primer	AcDx-5591-TDH-S1-FP	GAGGTTTGGTTGCGCrGGGAC/3SpC3/	20	425
Reverse PCR Primer	AcDx-5592-TDH-S1-RP	GGTGTCGTGGCCCACACTAACCTTCCTACGrCACCT/3SpC3/	35	426
Upstream LDR	AcDx-5593-TDH-S1-Up	TCTCGGGACCACAATACGAACGGTTGCGCGAGGTTCGCrGGGCG/3SpC3/	43	427
Downstream LDR	AcDx-5594-TDH-S1-Dn	/5Phos/GGGTAGTTTTAATTTTGGGTTCGTAGTTTGCGTTGGGTTACGCTAAGCTGGTGCCA	56	428
Real-Time Probe	AcDx-5595-TDH-S1-RT-Pb	/56-FAM/TTGGTTCGC/ZEN/GGGTAGTTTTAATTTTGGGTTC/3IABkFQ/	31	429
Tag Forward Primer	AcDx-5596-TDH-S1-RT-FP	TCTCGGGACCACAATACGAAC	21	430
Tag Reverse Primer	AcDx-5597-TDH-S1-RT-RP	TGGCACCAGCTTAGCGTAAC	20	431
Downstream PCR Primer	AcDx-5598-TDH-S1-PCR-V	TGGCACCAGCTTAGCGTAACACCTTCCTACGCACCCAATGrCAAAT/3SpC3/	45	432
C9orf50				
Forward PCR Primer	AcDx-5601-C9orf50-S1-FP	AGTTGGTGTAGGAATTTACGGAArUTTGC/3SpC3/	28	433

Reverse PCR Primer	AcDx-5602-C9orf50-S1-RP	GGTGTCGTGGCCTAAAAAACGCGAACGCCrCCCGG/3SpC3/	34	434
Upstream LDR	AcDx-5603-C9orf50-S1-Up	TCACGCACGTAGGGTCTAAACTTGAGGAGGAGTTTTTTTAGGAAAGCrGTTCG/3SpC3/	52	435
Downstream LDR	AcDx-5604-C9orf50-S1-Dn	/5Phos/GTTTAAGAAGTCGAGGTTTTTTTTGGTTACGCGTGTTGTCCGGCTGTGGTTACA	54	436
Real-Time Probe	AcDx-5605-C9orf50-S1-RT-Pb	/56-FAM/ATGGAAAGC/ZEN/GTTTAAGAAGTCGAGGTTTTTTTTGG/3IABkFQ/	35	437
Tag Forward Primer	AcDx-5606-C9orf50-S1-RT-FP	TCACGCACGTAGGGTCTAAAC	21	438
Tag Reverse Primer	AcDx-5607-C9orf50-S1-RT-RP	TGTAACCACAGCCGGACAAC	20	439
Downstream PCR Primer	AcDx-5608-C9orf50-S1-PCR-V	TGTAACCACAGCCGGACAACCGCCCCCGAAAACGTGrUAACT/3SpC3/	41	440
SFMBT2				
Forward PCR Primer	AcDx-5611-SFMBT2-S1-FP	GTTTTTAAGTTAGTGTCGTTTGTTTTCrGGTTC/3SpC3/	32	441
Reverse PCR Primer	AcDx-5612-SFMBT2-S1-RP	GGTGTCGTGGATATTAACCAAATTTATAATTAACGCAACArACGTG/3SpC3/	45	442
Upstream LDR	AcDx-5613-SFMBT2-S1-Up	TAACGGGATTGAGAGTGGACATCGGTTTTTATTTATTGATAGTACGGCGCTCrGAGCA/3SpC3/	57	443
Downstream LDR	AcDx-5614-SFMBT2-S1-Dn	/5Phos/GAGTGATTTTGTTTAGTTTCGTTTTGCGTTTTTGTAAATTACGTTGTCTGCCGCCCTTACTAA	63	444
Real-Time Probe	AcDx-5615-SFMBT2-S1-RT-Pb	/56-FAM/TTGGCGCTC/ZEN/GAGTGATTTTGTTTAGTTTCGT/3IABkFQ/	31	445
Tag Forward Primer	AcDx-5616-SFMBT2-S1-RT-FP	TAACGGGATTGAGAGTGGACA	21	446
Tag Reverse Primer	AcDx-5617-SFMBT2-S1-RT-RP	TTAGTAAGGGCGGCAGACA	19	447
Downstream PCR Primer	AcDx-5618-SFMBT2-S1-PCR-V	TTAGTAAGGGCGGCAGACAAATTAACGCAACAACGTAATTTACAAAAATGrCAAAG/3SpC3/	55	448
NCAM1				
Forward PCR Primer	AcDx-5621-NCAM1-S1-FP	GCGGGCGGTATAAGAGTAGrCGTTT/3SpC3/	24	449
Reverse PCR Primer	AcDx-5622-NCAM1-S1-RP	GGTGTCGTGGCGCCCAACCAACTTCGCrACTAG/3SpC3/	32	450
Upstream LDR	AcDx-5623-NCAM1-S1-Up	TTGAAGGAGGAAATCGGCACAGCGGTATAAGAGTAGCGTTCGATCrGTCAC/3SpC3/	50	451
Downstream LDR	AcDx-5624-NCAM1-S1-Dn	/5Phos/GTCGTTTTTAGTTAATTCGGGTTTTTTTTACGGCGATTAATTAGTGTCGAACCGTTTTAGGACTGA	66	452

Real-Time Probe	AcDx-5625-NCAM1-S1-RT-Pb	/56-FAM/TCTTCGATC/ZEN/GTCGTTTTTAGTTAATTCGGGTTTTT/3IABkFQ/	35	453
Tag Forward Primer	AcDx-5626-NCAM1-S1-RT-FP	TTGAAGGAGGAAATCGGCACA	21	454
Tag Reverse Primer	AcDx-5627-NCAM1-S1-RT-RP	TCAGTCCTAAAACGGTTCGACA	22	455
Downstream PCR Primer	AcDx-5628-NCAM1-S1-PCR-V	TCAGTCCTAAAACGGTTCGACACCAACTTCGCACTAATTAATCGCTGrUAAAG/3SpC3/	52	456
NOS1				
Forward PCR Primer	AcDx-5631-NOS1-S1-FP	AGGAATTTATTTGACGTCGAGTCrGGGTG/3SpC3/	28	457
Reverse PCR Primer	AcDx-5632-NOS1-S1-RP	GGTGTCGTGGCGCGAACGCCCGAAArAAAAG/3SpC3/	30	458
Upstream LDR	AcDx-5633-NOS1-S1-Up	TACAGATACGGACGGGAATCAACGTCGAGTCGGGTAGCATCrGAGAC/3SpC3/	46	459
Downstream LDR	AcDx-5634-NOS1-S1-Dn	/5Phos/GAGGTTGTTTTTTCGCGTTGCGTATTCGTTCGTTGTTTACATCCTCCTGCGTCA	54	460
Real-Time Probe	AcDx-5635-NOS1-S1-RT-Pb	/56-FAM/TTTAGCATC/ZEN/GAGGTTGTTTTTTCGCGTTG/3IABkFQ/	29	461
Tag Forward Primer	AcDx-5636-NOS1-S1-RT-FP	TACAGATACGGACGGGAATCAA	22	462
Tag Reverse Primer	AcDx-5637-NOS1-S1-RT-RP	TGACGCAGGAGGATGTAAACAA	22	463
Downstream PCR Primer	AcDx-5638-NOS1-S1-PCR-V	TGACGCAGGAGGATGTAAACAAAAAAAAAATACGAAACGAACGAACGAATATGrCAACA/3SpC3/	57	464
FGF14				
Forward PCR Primer	AcDx-5641-FGF14-S1-FP	GGAAAGGTTGCGTTTAGTTTTTTCrGTTAA/3SpC3/	29	465
Reverse PCR Primer	AcDx-5642-FGF14-S1-RP	GGTGTCGTGGATTAAAAACGACCGCGAAACTATAArCGAAT/3SpC3/	40	466
Upstream LDR	AcDx-5643-FGF14-S1-Up	TGAACGCTCAAACACGTGAACGGTTGCGTTTAGTTTTTTCGTTAGCATCrGTCAC/3SpC3/	54	467
Downstream LDR	AcDx-5644-FGF14-S1-Dn	/5Phos/GTCGTTATTATTATGTAGTTTTTTAGTTTGTTTTTTTTTGGGTTGTTCGGTTGGCCTGTAAGCGTTCCA	68	468
Real-Time Probe	AcDx-5645-FGF14-S1-RT-Pb	/56-FAM/AATAGCATC/ZEN/GTCGTTATTATTATGTAGTTTTTTAGTTTGTTTTTT/3IABkFQ/	45	469
Tag Forward Primer	AcDx-5646-FGF14-S1-RT-FP	TGAACGCTCAAACACGTGAAC	21	470
Tag Reverse Primer	AcDx-5647-FGF14-S1-RT-RP	TGGAACGCTTACAGGCCAAC	20	471
Downstream PCR Primer	AcDx-5648-FGF14-S1-PCR-V	TGGAACGCTTACAGGCCAACATTAAAAACGACCGCGAAACTATAAATGrAACAG/3SpC3/	53	472
COL4A2,COL4A1				
Forward PCR Primer	AcDx-5651-COL4A2/1-S1-FP	GGGTGAAGGCGTTTAGTTGTCrGAGTG/3SpC3/	26	473
Reverse PCR Primer	AcDx-5652-COL4A2/1-S1-RP	GGTGTCGTGGCAACCCGCGCTATAACGAArATAAG/3SpC3/	34	474

Upstream LDR	AcDx-5653-COL4A2/1-S1-Up	TGCGACTCTATTCACGTCCAAGGTGAAGGCGTTTAGTTGTCGAGTATACTCrGCGGT/3SpC3/	56	475
Downstream LDR	AcDx-5654-COL4A2/1-S1-Dn	/5Phos/GCGACGCGGGTTTAAATTTCGTCGGTTTGCTATTTGGTGTACCGCCA	47	476
Real-Time Probe	AcDx-5655-COL4A2/1-S1-RT-Pb	/56-FAM/CCTATACTC/ZEN/GCGACGCGGGTTTAAATTTCG/3IABkFQ/	30	477
Tag Forward Primer	AcDx-5656-COL4A2/1-S1-RT-FP	TGCGACTCTATTCACGTCCAA	21	478
Tag Reverse Primer	AcDx-5657-COL4A2/1-S1-RT-RP	TGGCGGTACACCAAATAGCAA	21	479
Downstream PCR Primer	AcDx-5658-COL4A2/1-S1-PCR-V	TGGCGGTACACCAAATAGCAAGCGCTATAACGAAATAAAAATACCTAAAAACTGrACGAG/3SpC3/	59	480
VSX1				
Forward PCR Primer	AcDx-5661-VSX1-S1-FP	GGTACGTTCGTTAGGAGTAGGTArGGGTA/3SpC3/	28	481
Reverse PCR Primer	AcDx-5662-VSX1-S1-RP	GGTGTCGTGGCCTACCGCTAAAACTCGACCrUCCTT/3SpC3/	35	482
Upstream LDR	AcDx-5663-VSX1-S1-Up	TGCCCTATCGAAAAGGACAACAAGGAGTAGGTAGGGTGTTCGGGCrGGTTA/3SpC3/	50	483
Downstream LDR	AcDx-5664-VSX1-S1-Dn	/5Phos/GGTCGTCGGCGGTTGCGTGTGTTTGCGGCTGTCTATGACA	40	484
Real-Time Probe	AcDx-5665-VSX1-S1-RT-Pb	/56-FAM/CATTCGGGC/ZEN/GGTCGTCGGC/3IABkFQ/	19	485
Tag Forward Primer	AcDx-5666-VSX1-S1-RT-FP	TGCCCTATCGAAAAGGACAACA	22	486
Tag Reverse Primer	AcDx-5667-VSX1-S1-RT-RP	TGTCATAGACAGCCGCAAACA	21	487
Downstream PCR Primer	AcDx-5668-VSX1-S1-PCR-V	TGTCATAGACAGCCGCAAACACGACCTCCTCTATAACTTCGACATGrCAACT/3SpC3/	51	488
PTPRT				
Forward PCR Primer	AcDx-5671-PTPRT-S1-FP	CGCGTATTTTTTTCGGGGTCrGATTG/3SpC3/	25	489
Reverse PCR Primer	AcDx-5672-PTPRT-S1-RP	GGTGTCGTGGAAACGAAAAAACGAAAACCCTACCrCGACA/3SpC3/	39	490
Upstream LDR	AcDx-5673-PTPRT-S1-Up	TTCTTGCGGTTCTGGAACACTCGGGGTCGATTAGTTCGGATCrGTTAA/3SpC3/	47	491
Downstream LDR	AcDx-5674-PTPRT-S1-Dn	/5Phos/GTTGGTATAGTTACGCGCGTTTATATATTATTTTCGTATTTATACGTCGGTGATGCTCCGTTGTTGCTAA	70	492
Real-Time Probe	AcDx-5675-PTPRT-S1-RT-Pb	/56-FAM/AATCGGATC/ZEN/GTTGGTATAGTTACGCGCG/3IABkFQ/	28	493
Tag Forward Primer	AcDx-5676-PTPRT-S1-RT-FP	TTCTTGCGGTTCTGGAACAC	20	494
Tag Reverse Primer	AcDx-5677-PTPRT-S1-RT-RP	TTAGCAACAACGGAGCATCAC	21	495
Downstream PCR	AcDx-5678-PTPRT-S1-PCR-V	TTAGCAACAACGGAGCATCACCGAAAACCCTACCCGACGTATAAATATGrAAAAC/3SpC3	54	496

Primer		/		
MIR124-3				
Forward PCR Primer	AcDx-5681-MIR124-3-S1-FP	GTAGCGGCGGGGTCrGGTGC/3SpC3/	19	497
Reverse PCR Primer	AcDx-5682-MIR124-3-S1-RP	GGTGTCGTGGCGTATCCGCGCTCCGrCTCCA/3SpC3/	30	498
Upstream LDR	AcDx-5683-MIR124-3-S1-Up	TGTGCCTTACGGAAAACCCAGGGTCGGTGTTCGGGTAACrGTTGC/3SpC3/	44	499
Downstream LDR	AcDx-5684-MIR124-3-S1-Dn	/5Phos/GTTATCGCGCGTTTTAGTGATAATCGGTCGGTTGGAGCTAGTTCGGCGACA	51	500
Real-Time Probe	AcDx-5685-MIR124-3-S1-RT-Pb	/56-FAM/AAGGGTAAC/ZEN/GTTATCGCGCGTTTTAGTGATA/3IABkFQ/	31	501
Tag Forward Primer	AcDx-5686-MIR124-3-S1-RT-FP	TGTGCCTTACGGAAAACCCA	20	502
Tag Reverse Primer	AcDx-5687-MIR124-3-S1-RT-RP	TGTCGCCGAACTAGCTCCA	19	503
Downstream PCR Primer	AcDx-5688-MIR124-3-S1-PCR-V	TGTCGCCGAACTAGCTCCACGCTCCGACACCGACTGrATTAC/3SpC3/	41	504
ZNF677				
Forward PCR Primer	AcDx-5691-ZNF677-S1-FP	GTTTTAATTTATAGGGCGATTTTAAAATTCrGATAA/3SpC3/	35	505
Reverse PCR Primer	AcDx-5692-ZNF677-S1-RP	GGTGTCGTGGCGAATAAATCTCCGTCTTTCGAATTCrAAACT/3SpC3/	41	506
Upstream LDR	AcDx-5693-ZNF677-S1-Up	TTGCAAACCACCCGGACAAGGCGATTTTAAAATTCGATAGCGGCGCrGTGAA/3SpC3/	51	507
Downstream LDR	AcDx-5694-ZNF677-S1-Dn	/5Phos/GTGGGAATTTGTGGTTCGCGAGGCTTGGTCAGCATCGACTCCTA	44	508
Real-Time Probe	AcDx-5695-ZNF677-S1-RT-Pb	/56-FAM/ATGCGGCGC/ZEN/GTGGGAATTTGTG/3IABkFQ/	22	509
Tag Forward Primer	AcDx-5696-ZNF677-S1-RT-FP	TTGCAAACCACCCGGACAA	19	510
Tag Reverse Primer	AcDx-5697-ZNF677-S1-RT-RP	TAGGAGTCGATGCTGACCAA	20	511
Downstream PCR Primer	AcDx-5698-ZNF677-S1-PCR-V	TAGGAGTCGATGCTGACCAAGAATTCAAACCTACGCCTCGTGrAACCG/3SpC3/	47	512
EVC				
Forward PCR Primer	AcDx-5701-EVC-S1-FP	TTGAAAGTTTTGAGCGGTGATTTArGGTTC/3SpC3/	29	513
Reverse PCR Primer	AcDx-5702-EVC-S1-RP	GGTGTCGTGGGCTCCGCAAACTTTCTAACCrUATAT/3SpC3/	35	514
Upstream LDR	AcDx-5703-EVC-S1-Up	TTCGCCTACCGCAGTGAACGTGATTTAGGTTTTTTTTTTCGGTTCAGCrGAAAC/3SpC3/	53	515
Downstream LDR	AcDx-5704-EVC-S1-Dn	/5Phos/GAAGTAGGGAAGAGGAGAGAAGTAGGAGTCGGTTGAGACATGGGCTCGCA	50	516
Real-Time Probe	AcDx-5705-EVC-S1-RT-Pb	/56-FAM/TTGTTCAGC/ZEN/GAAGTAGGGAAGAGGAGAGAAGTA/3IABkFQ/	33	517

Tag Forward Primer	AcDx-5706-EVC-S1-RT-FP	TTCGCCTACCGCAGTGAAC	19	518
Tag Reverse Primer	AcDx-5707-EVC-S1-RT-RP	TGCGAGCCCATGTCTCAAC	19	519
Downstream PCR Primer	AcDx-5708-EVC-S1-PCR-V	TGCGAGCCCATGTCTCAACCGCAAACTTTCTAACCTATACAATCTCCTGrACTCT/3SpC3/	54	520
JAM2				
Forward PCR Primer	AcDx-5711-JAM2-S1-FP	GAAAGATGGTATTATTGGGAGGCrGGGCA/3SpC3/	28	521
Reverse PCR Primer	AcDx-5712-JAM2-S1-RP	GGTGTCGTGGGACTCAACTAAAACGCTACAAAACCrCAAAT/3SpC3/	40	522
Upstream LDR	AcDx-5713-JAM2-S1-Up	TTGCATTTCGTTAGCGACACACGGGCGTCGGTTATATTTTGAGTTCTCrGGGAG/3SpC3/	53	523
Downstream LDR	AcDx-5714-JAM2-S1-Dn	/5Phos/GAGGAGGAGGTAGCGTTCGCGTGTGAGTCGATCTACCCGCA/3IABkFQ/	41	524
Real-Time Probe	AcDx-5715-JAM2-S1-RT-Pb	/56-FAM/CCAGTTCTC/ZEN/GAGGAGGAGGTAGCGTTCG	28	525
Tag Forward Primer	AcDx-5716-JAM2-S1-RT-FP	TTGCATTTCGTTAGCGACACA	21	526
Tag Reverse Primer	AcDx-5717-JAM2-S1-RT-RP	TGCGGGTAGATCGACTCACA	20	527
Downstream PCR Primer	AcDx-5718-JAM2-S1-PCR-V	TGCGGGTAGATCGACTCACAACGCTACAAAACCCAAACCATGrCGAAT/3SpC3/	47	528
GFRA1				
Forward PCR Primer	AcDx-5721-GFRA1-S1-FP	GAAGGGTCGAGTTGGGTTArGGACA/3SpC3/	24	529
Reverse PCR Primer	AcDx-5722-GFRA1-S1-RP	GGTGTCGTGGGCCGCTTCCAATAACCACrUAACG/3SpC3/	33	530
Upstream LDR	AcDx-5723-GFRA1-S1-Up	TAACGGGATTGAGAGTGGACAGGTCGAGTTGGGTTAGGACGATTCTCrGAGCG/3SpC3/	52	531
Downstream LDR	AcDx-5724-GFRA1-S1-Dn	/5Phos/GAGTAGAGTTTTCGGTTCGGATGTTCGTTAGGGTGTCTGCCGCCCTTACTAA	52	532
Real-Time Probe	AcDx-5725-GFRA1-S1-RT-Pb	/56-FAM/TTGATTCTC/ZEN/GAGTAGAGTTTTCGGTTCGGATGTTC/3IABkFQ/	35	533
Tag Forward Primer	AcDx-5726-GFRA1-S1-RT-FP	TAACGGGATTGAGAGTGGACA	21	534
Tag Reverse Primer	AcDx-5727-GFRA1-S1-RT-RP	TTAGTAAGGGCGGCAGACA	19	535
Downstream PCR Primer	AcDx-5728-GFRA1-S1-PCR-V	TTAGTAAGGGCGGCAGACACTTCCAATAACCACTAACATCCCTAATGrAACAC/3SpC3/	52	536
FLI1-2				
Forward PCR Primer	AcDx-5731-FLI1-S1-FP	GGGTTAATTCGAAGAGGTTGCrGAGGC/3SpC3/	26	537
Reverse PCR Primer	AcDx-5732-FLI1-S1-RP	GGTGTCGTGGCCTTAATAATCCCGTCCATTTAACCrAAATT/3SpC3/	40	538
Upstream LDR	AcDx-5733-FLI1-S1-Up	TTCGTGCGTCGTGTAGCAAGAGGTTGCGAGGTTAGGTTGTAGTCrGGGCC/3SpC3/	49	539
Downstream LDR	AcDx-5734-FLI1-S1-Dn	/5Phos/GGGTTAATGTGTGGAATATTGGGAGGTTCGGTTGCCCATTTTCTGCACCCA	51	540

Real-Time Probe	AcDx-5735-FLI1-S1-RT-Pb	/56-FAM/AATGTAGTC/ZEN/GGGTTAATGTGTGGAATATTGGGA/3IABkFQ/	33	541
Tag Forward Primer	AcDx-5736-FLI1-S1-RT-FP	TTCGTGCGTCGTGTAGCAA	19	542
Tag Reverse Primer	AcDx-5737-FLI1-S1-RT-RP	TGGGTGCAGAAAATGGGCAA	20	543
Downstream PCR Primer	AcDx-5738-FLI1-S1-PCR-V	TGGGTGCAGAAAATGGGCAACCGTCCATTTAACCAAATCTACAACTGrAACCT/3SpC3/	52	544
GNAO1,DKFZP434H168				
Forward PCR Primer	AcDx-5741-GNAO1-S1-FP	GTTTAGTTCGGCGACGTTCrGGTTG/3SpC3/	24	545
Reverse PCR Primer	AcDx-5742-GNAO1-S1-RP	GGTGTCGTGGAAACACCGACTCGACTCCrGAACG/3SpC3/	33	546
Upstream LDR	AcDx-5743-GNAO1-S1-Up	TCCAAACGATTAGGAGCGTCAAACGTTCGGTTAGGTTCGGCGCrGGTCC/3SpC3/	48	547
Downstream LDR	AcDx-5744-GNAO1-S1-Dn	/5Phos/GGTTTGGGCGCGAGGTTCGGTTGGACAGAGGTATACGCCCA	41	548
Real-Time Probe	AcDx-5745-GNAO1-S1-RT-Pb	/56-FAM/AATCGGCGC/ZEN/GGTTTGGGCG/3IABkFQ/	19	549
Tag Forward Primer	AcDx-5746-GNAO1-S1-RT-FP	TCCAAACGATTAGGAGCGTCAA	22	550
Tag Reverse Primer	AcDx-5747-GNAO1-S1-RT-RP	TGGGCGTATACCTCTGTCCAA	21	551
Downstream PCR Primer	AcDx-5748-GNAO1-S1-PCR-V	TGGGCGTATACCTCTGTCCAACGACTCCGAACACCGCTGrACTCT/3SpC3/	44	552
CoCaNCR2				
Forward PCR Primer	AcDx-5751-CoCaNCR2-S1-FP	GAAGGGTAGTTTCGGGTAGTACrGGATC/3SpC3/	27	553
Reverse PCR Primer	AcDx-5752-CoCaNCR2-S1-RP	GGTGTCGTGGCGACTTTAACGCTAACATATCCCrUCTCT/3SpC3/	38	554
Upstream LDR	AcDx-5753-CoCaNCR2-S1-Up	TGATGCTGGCAAACCCTAGAACGGGTAGTACGGATTAGATATTGTTATGAAGGACrGTTCC/3SpC3/	60	555
Downstream LDR	AcDx-5754-CoCaNCR2-S1-Dn	/5Phos/GTTTTCGATAAGGGAAGGTTAGGGTAGGTGGGGTTCCATCACCGTTAGGCCA	52	556
Real-Time Probe	AcDx-5755-CoCaNCR2-S1-RT-Pb	/56-FAM/CCGAAGGAC/ZEN/GTTTTCGATAAGGGAAGGTTAGGG/3IABkFQ/	33	557
Tag Forward Primer	AcDx-5756-CoCaNCR2-S1-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	558
Tag Reverse Primer	AcDx-5757-CoCaNCR2-S1-RT-RP	TGGCCTAACGGTGATGGAAC	20	559
Downstream PCR Primer	AcDx-5758-CoCaNCR2-S1-PCR-V	TGGCCTAACGGTGATGGAACCGCTAACATATCCCTCTCCCCGCrCTCCT/3SpC3/	48	560

CoCaNCR9				
Forward PCR Primer	AcDx-5761-CoCaNCR9-S1-FP	GGGTTGGTTTTCGTCGAGArUTTTA/3SpC3/	24	561
Reverse PCR Primer	AcDx-5762-CoCaNCR9-S1-RP	GGTGTCGTGGGCGACAACGAAACGCArUCACT/3SpC3/	31	562
Upstream LDR	AcDx-5763-CoCaNCR9-S1-Up	TTCGCTGCCCGGTTAAACAGTTTTCGTCGAGATTTTGTTGGGCGCrGGGCC/3SpC3/	50	563
Downstream LDR	AcDx-5764-CoCaNCR9-S1-Dn	/5Phos/GGGTTTTGGTTTTACGGTGAGTTCGGTTTTAGTTGTTATCGGACCTAGCTCGACA	55	564
Real-Time Probe	AcDx-5765-CoCaNCR9-S1-RT-Pb	/56-FAM/AATGGGCGC/ZEN/GGGTTTTGGTTTTAC/3IABkFQ/	24	565
Tag Forward Primer	AcDx-5766-CoCaNCR9-S1-RT-FP	TTCGCTGCCCGGTTAAACA	19	566
Tag Reverse Primer	AcDx-5767-CoCaNCR9-S1-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	567
Downstream PCR Primer	AcDx-5768-CoCaNCR9-S1-PCR-V	TGTCGAGCTAGGTCCGATAACAAACGCATCACCGAAACTAAAACTGrAACTT/3SpC3/	51	568
CoCaNCR10				
Forward PCR Primer	AcDx-5771-CoCaNCR10-S1-FP	TTAGAGTAGGAGTCGGTGTGArGTTGA/3SpC3/	26	569
Reverse PCR Primer	AcDx-5772-CoCaNCR10-S1-RP	GGTGTCGTGGCGACTTCGAACCTACTCTCTAATTCrUATAT/3SpC3/	40	570
Upstream LDR	AcDx-5773-CoCaNCR10-S1-Up	TCACAGAGACTTGCCGATCACGGTGTGAGTTGGGAGGATGCrGGGCC/3SpC3/	46	571
Downstream LDR	AcDx-5774-CoCaNCR10-S1-Dn	/5Phos/GGGTTGTAGACGTGATCGGCGATAGATAGATTATCGGTGTGTAGCTTAGACATGGCCA	58	572
Real-Time Probe	AcDx-5775-CoCaNCR10-S1-RT-Pb	/56-FAM/TTAGGATGC/ZEN/GGGTTGTAGACGTGATCG/3IABkFQ/	27	573
Tag Forward Primer	AcDx-5776-CoCaNCR10-S1-RT-FP	TCACAGAGACTTGCCGATCAC	21	574
Tag Reverse Primer	AcDx-5777-CoCaNCR10-S1-RT-RP	TGGCCATGTCTAAGCTACACAC	22	575
Downstream PCR Primer	AcDx-5778-CoCaNCR10-S1-PCR-V	TGGCCATGTCTAAGCTACACACAATTCTATACATCCGATAATCTATCTATCGCTGrATCAT/3SpC3/	60	576
CoCaNCR8				
Forward PCR Primer	AcDx-5781-CoCaNCR8-S1-FP	TCGGGGAGCGGTTGTArGGGCA/3SpC3/	21	577

Reverse PCR Primer	AcDx-5782-CoCaNCR8-S1-RP	GGTGTCGTGGCAAACGAAAAACTCTTTTACTCCTCCrCAAAG/3SpC3/	41	578
Upstream LDR	AcDx-5783-CoCaNCR8-S1-Up	TCGACGAATCTGCTCAGACAAGGTTGTAGGGCGGCAGCrGGCAA/3SpC3/	43	579
Downstream LDR	AcDx-5784-CoCaNCR8-S1-Dn	/5Phos/GGCGGTCGGGTTATTAGAAGTATAGTGGAGTCGTTGAAGCAGCGTCTGAGCA	52	580
Real-Time Probe	AcDx-5785-CoCaNCR8-S1-RT-Pb	/56-FAM/TTCGGCAGC/ZEN/GGCGGTCG/3IABkFQ/	17	581
Tag Forward Primer	AcDx-5786-CoCaNCR8-S1-RT-FP	TCGACGAATCTGCTCAGACAA	21	582
Tag Reverse Primer	AcDx-5787-CoCaNCR8-S1-RT-RP	TGCTCAGACGCTGCTTCAA	19	583
Downstream PCR Primer	AcDx-5788-CoCaNCR8-S1-PCR-V	TGCTCAGACGCTGCTTCAACTCTTTTACTCCTCCCAAAACAAAAAATGrACTCT/3SpC3/	53	584
NPY-2				
Forward PCR Primer	AcDx-5791-NPY-S1-FP	CGCGATTCGTTTTTTGTATTTTATTCrGTTGA/3SpC3/	31	585
Reverse PCR Primer	AcDx-5792-NPY-S1-RP	GGTGTCGTGGAATCGTAACACTCACGATAACTAACrGCGCA/3SpC3/	40	586
Upstream LDR	AcDx-5793-NPY-S1-Up	TCGATGGTCAATGAGCTTCACATGTATTTTATTCGTTGGTTTTTATTTTTCGGAAACrGTTTA /3SpC3/	62	587
Downstream LDR	AcDx-5794-NPY-S1-Dn	/5Phos/GTTCGTTCGATAGTATAGTATTTGTCGTTTAGTTACGTTCGCTGTTACGTGATCTC CCTCTCCA	64	588
Real-Time Probe	AcDx-5795-NPY-S1-RT-Pb	/56-FAM/AACGGAAAC/ZEN/GTTCGTTCGATAGTATAGTATTTGTCGTTTAG/3IABkFQ/	41	589
Tag Forward Primer	AcDx-5796-NPY-S1-RT-FP	TCGATGGTCAATGAGCTTCACA	22	590
Tag Reverse Primer	AcDx-5797-NPY-S1-RT-RP	TGGAGAGGGAGATCACGTAACA	22	591
Downstream PCR Primer	AcDx-5798-NPY-S1-PCR-V	TGGAGAGGGAGATCACGTAACATCACGATAACTAACGCGCGAATGrUAACC/3SpC3/	50	592
SLC18A3,CHAT				
Forward PCR Primer	AcDx-5801-SLC18A3-S1-FP	GCGGGAGTGTGTGTGTAArGGGAC/3SpC3/	23	593
Reverse PCR Primer	AcDx-5802-SLC18A3-S1-RP	GGTGTCGTGGCAACTAAATTATAAAACCGCGTCCrGAAAG/3SpC3/	39	594
Upstream LDR	AcDx-5803-SLC18A3-S1-Up	TACGAATCACCCGAGAGTTCAAGGGTTAGGAGGCGGAATCrGGAAG/3SpC3/	45	595
Downstream LDR	AcDx-5804-SLC18A3-S1-Dn	/5Phos/GGAGAAAGGGTTGGAGGTGTTCGTTTGTGGGTGGGTATAGGTCAGA	46	596
Real-Time Probe	AcDx-5805-SLC18A3-S1-RT-Pb	/56-FAM/TTCGGAATC/ZEN/GGAGAAAGGGTTGGAGGT/3IABkFQ/	27	597
Tag Forward Primer	AcDx-5806-SLC18A3-S1-RT-	TACGAATCACCCGAGAGTTCAA	22	598

	FP			
Tag Reverse Primer	AcDx-5807-SLC18A3-S1-RT-RP	TCTGACCTATACCCACCCACAA	22	599
Downstream PCR Primer	AcDx-5808-SLC18A3-S1-PCR-V	TCTGACCTATACCCACCCACAACGTCCGAAAAAACGCTAAACATGrAACAT/3SpC3/	50	600
LIFR				
Forward PCR Primer	AcDx-5811-LIFR-S1-FP	GTTTCGCGTCGCGTTTATTCrGTTTC/3SpC3/	25	601
Reverse PCR Primer	AcDx-5812-LIFR-S1-RP	GGTGTCGTGGCCGAAACGACGACCGAAACrUACAG/3SpC3/	34	602
Upstream LDR	AcDx-5813-LIFR-S1-Up	TCCGGCCTTTGACGATACCGCGTCGCGTTTATTCGTTTTTAGGATTCACrGGTGT/3SpC3/	54	603
Downstream LDR	AcDx-5814-LIFR-S1-Dn	/5Phos/GGTACGTTTTCGCGTCGTTATTTTGTTATTTTTTGTCGGGTAATTCACTCGAACGGAGCA	60	604
Real-Time Probe	AcDx-5815-LIFR-S1-RT-Pb	/56-FAM/TTGATTCAC/ZEN/GGTACGTTTTCGCGTCGT/3IABkFQ/	27	605
Tag Forward Primer	AcDx-5816-LIFR-S1-RT-FP	TCCGGCCTTTGACGATACC	19	606
Tag Reverse Primer	AcDx-5817-LIFR-S1-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	607
Downstream PCR Primer	AcDx-5818-LIFR-S1-PCR-V	TGCTCCGTTCGAGTGAATTACCCGACCGAAACTACAAAACCGATGrACAAG/3SpC3/	50	608
LONRF2-1				
Forward PCR Primer	AcDx-5821-LONRF2-S1-FP	GTTAGTATGGAGCGAAAGAGTTCrGGTTA/3SpC3/	28	609
Reverse PCR Primer	AcDx-5822-LONRF2-S1-RP	GGTGTCGTGGCCTAACTACGACCGCGCrGAAAT/3SpC3/	32	610
Upstream LDR	AcDx-5823-LONRF2-S1-Up	TAGCCGATGGCGTAAAACCGCGAAAGAGTTCGGTTGTTATTTCGTAATCrGTTTA/3SpC3/	54	611
Downstream LDR	AcDx-5824-LONRF2-S1-Dn	/5Phos/GTTCGCGCGGAAGGTTTCGTCGTGGTCGTATGACTTGCTCGCA	43	612
Real-Time Probe	AcDx-5825-LONRF2-S1-RT-Pb	/56-FAM/AACGTAATC/ZEN/GTTCGCGCGGAAGGTTTC/3IABkFQ/	27	613
Tag Forward Primer	AcDx-5826-LONRF2-S1-RT-FP	TAGCCGATGGCGTAAAACC	19	614
Tag Reverse Primer	AcDx-5827-LONRF2-S1-RT-RP	TGCGAGCAAGTCATACGACC	20	615
Downstream PCR Primer	AcDx-5828-LONRF2-S1-PCR-V	TGCGAGCAAGTCATACGACCACCGCGCGAAACCGATTGrCCCAG/3SpC3/	43	616
LONRF2-2				

Forward PCR Primer	AcDx-5831-LONRF2-S2-FP	TTTCGTAGGGTTGTAATCGTTTCrGGGAC/3SpC3/	28	617
Reverse PCR Primer	AcDx-5832-LONRF2-S2-RP	GGTGTCGTGGATTACCTAAACCGACGAAACTAAArAAACT/3SpC3/	39	618
Upstream LDR	AcDx-5833-LONRF2-S2-Up	TTGCTGTGCGCGGTAGAACCGTTTCGAGGTGAGGATCrGGGTC/3SpC3/	42	619
Downstream LDR	AcDx-5834-LONRF2-S2-Dn	/5Phos/GGGATAGGTATCGCGGGCGTAATTTGAGTGTTACGCTAAGCTGGTGCCA	49	620
Real-Time Probe	AcDx-5835-LONRF2-S2-RT-Pb	/56-FAM/AAGAGGATC/ZEN/GGGATAGGTATCGCGGG/3IABkFQ/	26	621
Tag Forward Primer	AcDx-5836-LONRF2-S2-RT-FP	TTGCTGTGCGCGGTAGAAC	19	622
Tag Reverse Primer	AcDx-5837-LONRF2-S2-RT-RP	TGGCACCAGCTTAGCGTAAC	20	623
Downstream PCR Primer	AcDx-5838-LONRF2-S2-PCR-V	TGGCACCAGCTTAGCGTAACGACGAAACTAAAAAACCGCTACGTGrUAAAT/3SpC3/	50	624
DCLK1				
Forward PCR Primer	AcDx-5841-DCLK1-S1-FP	CGTTTCGGTTGCGGTTATTArGTGTG/3SpC3/	25	625
Reverse PCR Primer	AcDx-5842-DCLK1-S1-RP	GGTGTCGTGGGCCGCCCTCCTCTAAArAAAAG/3SpC3/	31	626
Upstream LDR	AcDx-5843-DCLK1-S1-Up	TTGCAGCGGGTCACAACAATTGCGGTTATTAGTGTAGGGATGTGTCTCrGGTCT/3SpC3/	53	627
Downstream LDR	AcDx-5844-DCLK1-S1-Dn	/5Phos/GGTTCGTTTTTATTGAAATGCGGCGTTTCGTATAGTTTGGACAGAGGTATACGCCCA	57	628
Real-Time Probe	AcDx-5845-DCLK1-S1-RT-Pb	/56-FAM/CCGTGTCTC/ZEN/GGTTCGTTTTTATTGAAATGCGG/3IABkFQ/	32	629
Tag Forward Primer	AcDx-5846-DCLK1-S1-RT-FP	TTGCAGCGGGTCACAACAA	19	630
Tag Reverse Primer	AcDx-5847-DCLK1-S1-RT-RP	TGGGCGTATACCTCTGTCCAA	21	631
Downstream PCR Primer	AcDx-5848-DCLK1-S1-PCR-V	TGGGCGTATACCTCTGTCCAAGCCCTCCTCTAAAAAAAAAAACTAAAATAAAACTATATGrAAACA/3SpC3/	65	632
SPG20-2				
Forward PCR Primer	AcDx-5851-SPG20-S1-FP	GTAAATTCGTTAAATTATGATATAATAAGGTTAATTTCrGAGGG/3SpC3/	43	633
Reverse PCR Primer	AcDx-5852-SPG20-S1-RP	GGTGTCGTGGGCAACATTATTACGTAACTCTAAAACTAACrCCAAC/3SpC3/	45	634
Upstream LDR	AcDx-5853-SPG20-S1-Up	TCTTACGCCCAGGGAATGTAACTCGTTAAATTATGATATAATAAGGTTAATTTCGAGGATTCTCrGGTAA/3SpC3/	69	635
Downstream LDR	AcDx-5854-SPG20-S1-Dn	/5Phos/GGTGGATTATTAGCGTATTTTTTTATAATTAGAAAGTGAAAAATTGGGTTGTCCGGCTGTGGTTACA	67	636
Real-Time Probe	AcDx-5855-SPG20-S1-RT-Pb	/56-	46	637

		FAM/TTGATTCTC/ZEN/GGTGGATTATTAGCGTATTTTTTTATAATTAGAAAGT/3IABkFQ/		
Tag Forward Primer	AcDx-5856-SPG20-S1-RT-FP	TCTTACGCCCAGGGAATGTAAC	22	638
Tag Reverse Primer	AcDx-5857-SPG20-S1-RT-RP	TGTAACCACAGCCGGACAAC	20	639
ZIK1				
Forward PCR Primer	AcDx-5861-ZIK1-S1-FP	GGGAGCGTTTTGTTTGGCrGATAA/3SpC3/	23	640
Reverse PCR Primer	AcDx-5862-ZIK1-S1-RP	GGTGTCGTGGCCGCCGACCCTTAAAACCrCCCAT/3SpC3/	33	641
Upstream LDR	AcDx-5863-ZIK1-S1-Up	TATAGTCACGCAGGACCACATGTTTGGCGATAGTCGGAAAGCrGCGGA/3SpC3/	47	642
Downstream LDR	AcDx-5864-ZIK1-S1-Dn	/5Phos/GCGAGAGGAGAGGTTTTTTCGTTGAATAGTGTTTGCGGCTGTCTATGACA	50	643
Real-Time Probe	AcDx-5865-ZIK1-S1-RT-Pb	/56-FAM/AAGGAAAGC/ZEN/GCGAGAGGAGAGGTTTT/3IABkFQ/	26	644
Tag Forward Primer	AcDx-5866-ZIK1-S1-RT-FP	TATAGTCACGCAGGACCACA	20	645
Tag Reverse Primer	AcDx-5867-ZIK1-S1-RT-RP	TGTCATAGACAGCCGCAAACA	21	646
Downstream PCR Primer	AcDx-5868-ZIK1-S1-PCR-V	TGTCATAGACAGCCGCAAACAACCCTTAAAACCCCCACTATTCAATGrAAAAG/3SpC3/	52	647
ZNF154				
Forward PCR Primer	AcDx-5871-ZNF154-S1-FP	GTCGTTAAGGTTTAGACGTTTTCrGTGTG/3SpC3/	28	648
Reverse PCR Primer	AcDx-5872-ZNF154-S1-RP	GGTGTCGTGGCGACCATTTTAACTTCTCTAAAATATATTCrACCGG/3SpC3/	45	649
Upstream LDR	AcDx-5873-ZNF154-S1-Up	TTCAACGATCGCGCAGACACGTGTAGGAGGGACGACGATTTTTTTCACrGTTCC/3SpC3/	53	650
Downstream LDR	AcDx-5874-ZNF154-S1-Dn	/5Phos/GTTTTCGTGGTTTTAATTCGGCGTTTTGTTATTTTTGATTCTGTTCTGGGAATTATTGCCGGA	63	651
Real-Time Probe	AcDx-5875-ZNF154-S1-RT-Pb	/56-FAM/CCTTTTCAC/ZEN/GTTTTCGTGGTTTTAATTCGGCG/3IABkFQ/	32	652
Tag Forward Primer	AcDx-5876-ZNF154-S1-RT-FP	TTCAACGATCGCGCAGACA	19	653
Tag Reverse Primer	AcDx-5877-ZNF154-S1-RT-RP	TCCGGCAATAATTCCCAGAACA	22	654
Downstream PCR Primer	AcDx-5878-ZNF154-S1-PCR-V	TCCGGCAATAATTCCCAGAACACGACCATTTTAACTTCTCTAAAATATATTCACTGrAATCG/3SpC3/	61	655
GPR88				
Forward PCR Primer	AcDx-5881-GPR88-S1-FP	GGTTGTTGGGTTTCGGATTTACrGGTGC/3SpC3/	27	656
Reverse PCR Primer	AcDx-5882-GPR88-S1-RP	GGTGTCGTGGCCGCCGTATAACGCCTCrUAATG/3SpC3/	32	657
Upstream LDR	AcDx-5883-GPR88-S1-Up	TGGATAAACTAAGTCCGCCCACGTGTTTTTTTTTTTTTATTGTTTCGTGGTTTTGAGTCrGTTGC/3SpC3/	64	658

Downstream LDR	AcDx-5884-GPR88-S1-Dn	/5Phos/GTTATTTGTTTATTATTCGGGCGTTCGTTATTTATTAGGCGTGTGACTGAGCGACGTCTAACA	63	659
Real-Time Probe	AcDx-5885-GPR88-S1-RT-Pb	/56-FAM/CCTTGAGTC/ZEN/GTTATTTGTTTATTATTCGGGCGTTCG/3IABkFQ/	36	660
Tag Forward Primer	AcDx-5886-GPR88-S1-RT-FP	TGGATAAACTAAGTCCGCCCAC	22	661
Tag Reverse Primer	AcDx-5887-GPR88-S1-RT-RP	TGTTAGACGTCGCTCAGTCAC	21	662
Downstream PCR Primer	AcDx-5888-GPR88-S1-PCR-V	TGTTAGACGTCGCTCAGTCACCGCCGTATAACGCCTCTAATACAATGrCCTAG/3SpC3/	52	663
SPSB4				
Forward PCR Primer	AcDx-5891-SPS84-S1-FP	TTATTTTAGGTTCGATTTTCGGCrGTTTA/3SpC3/	28	664
Reverse PCR Primer	AcDx-5892-SPS84-S1-RP	GGTGTCGTGGTAATACCCGACAACAACGCCrCCCTT/3SpC3/	35	665
Upstream LDR	AcDx-5893-SPS84-S1-Up	TACTATCGTATCACGCCGACAGGTTCGATTTTCGGCGTTTGGGCrGTTGG/3SpC3/	49	666
Downstream LDR	AcDx-5894-SPS84-S1-Dn	/5Phos/GTTAATTTCGTTAAGAACGTTTTTAATTTTAGGTTATTTTGTAGCGTAGTGTAACGTCCGTGGGCTAA	68	667
Real-Time Probe	AcDx-5895-SPS84-S1-RT-Pb	/56-FAM/AATTTGGGC/ZEN/GTTAATTTCGTTAAGAACGTTTTTAATTTTAG/3IABkFQ/	41	668
Tag Forward Primer	AcDx-5896-SPS84-S1-RT-FP	TACTATCGTATCACGCCGACA	21	669
Tag Reverse Primer	AcDx-5897-SPS84-S1-RT-RP	TTAGCCCACGGACGTTACA	19	670
Downstream PCR Primer	AcDx-5898-SPS84-S1-PCR-V	TTAGCCCACGGACGTTACAACAACAACGCCCCCTCTATGrCTACG/3SpC3/	44	671
GUCY1B3				
Forward PCR Primer	AcDx-5901-GUCY1B3-S1-FP	GGGAGAATTTATTAGGGATTGGGrGGGTC/3SpC3/	28	672
Reverse PCR Primer	AcDx-5902-GUCY1B3-S1-RP	GGTGTCGTGGTCACCCCGCCCCTAArCTCCC/3SpC3/	30	673
Upstream LDR	AcDx-5903-GUCY1B3-S1-Up	TGCTATGCCGCATTCAACCAGGAGAATTTATTAGGGATTGGGAGGTCGCrGTTCC/3SpC3/	54	674
Downstream LDR	AcDx-5904-GUCY1B3-S1-Dn	/5Phos/GTTTTTACGATTCGATTTTTTTAGTGTATAGTATTTTTAAGGGAGAGGAGTGGAGCTAGTTCGGCGACA	69	675
Real-Time Probe	AcDx-5905-GUCY1B3-S1-RT-Pb	/56-FAM/TTGGGTCGC/ZEN/GTTTTTACGATTCGATTTTTTTAGTG/3IABkFQ/	35	676
Tag Forward Primer	AcDx-5906-GUCY1B3-S1-RT-FP	TGCTATGCCGCATTCAACCA	20	677
Tag Reverse Primer	AcDx-5907-GUCY1B3-S1-RT-RP	TGTCGCCGAACTAGCTCCA	19	678

SYNE1				
Forward PCR Primer	AcDx-5911-SYNE1-S1-FP	GGCGGTCGCGGTTCrGGCGC/3SpC3/	19	679
Reverse PCR Primer	AcDx-5912-SYNE1-S1-RP	GGTGTCGTGGCGAAAAACGCGACTAAACAACCrCGACA/3SpC3/	37	680
Upstream LDR	AcDx-5913-SYNE1-S1-Up	TGGAGGCCGGAGAAATTAAACGTCGCGGTTCGGCGCTCrGCGCC/3SpC3/	43	681
Downstream LDR	AcDx-5914-SYNE1-S1-Dn	/5Phos/GCGTTTATTTTGGTTGGTTCGTTTATATTTATTTTCGCGTGTTTGGGATCTGGGCATCACA	61	682
Real-Time Probe	AcDx-5915-SYNE1-S1-RT-Pb	/56-FAM/AAGGCGCTC/ZEN/GCGTTTATTTTGGTTGG/3IABkFQ/	26	683
Tag Forward Primer	AcDx-5916-SYNE1-S1-RT-FP	TGGAGGCCGGAGAAATTAAAC	21	684
Tag Reverse Primer	AcDx-5917-SYNE1-S1-RT-RP	TGTGATGCCCAGATCCCAAAC	21	685
Downstream PCR Primer	AcDx-5918-SYNE1-S1-PCR-V	TGTGATGCCCAGATCCCAAACCAACCCGACGCGAAAATAAATATAAATGrAACCG/3SpC3/	54	686
AMPH				
Forward PCR Primer	AcDx-5921-AMPH-S1-FP	TTCGTAGCGCGTCGTATTTCrGAGGC/3SpC3/	25	687
Reverse PCR Primer	AcDx-5922-AMPH-S1-RP	GGTGTCGTGGCGACCGAAACTAAAACCGAAAACrGCCAG/3SpC3/	38	688
Upstream LDR	AcDx-5923-AMPH-S1-Up	TAGTTTGTCGAAAGTCCCACACCGTCGTATTTCGAGGTTCGGGTCTCrGTACG/3SpC3/	52	689
Downstream LDR	AcDx-5924-AMPH-S1-Dn	/5Phos/GTATAGTTGTAGTCGGTGTTTTTCGGTTAAGTTTTCGTTTGTGCAAAATTCAGGCTGTGCA	61	690
Real-Time Probe	AcDx-5925-AMPH-S1-RT-Pb	/56-FAM/AAGGGTCTC/ZEN/GTATAGTTGTAGTCGGTGTTTTTCG/3IABkFQ/	34	691
Tag Forward Primer	AcDx-5926-AMPH-S1-RT-FP	TAGTTTGTCGAAAGTCCCACAC	22	692
Tag Reverse Primer	AcDx-5927-AMPH-S1-RT-RP	TGCACAGCCTGAATTTTGCAC	21	693
Downstream PCR Primer	AcDx-5928-AMPH-S1-PCR-V	TGCACAGCCTGAATTTTGCACGAAACTAAAACCGAAAACGCCAAATGrAAAAT/3SpC3/	52	694
RIMS2				
Forward PCR Primer	AcDx-5931-RIMS2-S1-FP	CGCGTCGGTGTCGTCrGTTGC/3SpC3/	20	695
Reverse PCR Primer	AcDx-5932-RIMS2-S1-RP	GGTGTCGTGGGACCCCGAAACCCGACrAAAAG/3SpC3/	31	696
Upstream LDR	AcDx-5933-RIMS2-S1-Up	TACCACTCATCTTCTGCGACATCGTCGTTGTTAGTGGAGTTGTTTTCTCrGTTCC/3SpC3/	54	697
Downstream LDR	AcDx-5934-RIMS2-S1-Dn	/5Phos/GTTTTTTTAGGGTGGTTCGGTTTTATTAAATATGTCGGTTTTTGTGAGTCGATCTACCCGCA	62	698
Real-Time Probe	AcDx-5935-RIMS2-S1-RT-Pb	/56-FAM/CCTTTTCTC/ZEN/GTTTTTTTAGGGTGGTTCGGTTTTATTAAATATG/3IABkFQ/	43	699
Tag Forward Primer	AcDx-5936-RIMS2-S1-RT-FP	TACCACTCATCTTCTGCGACA	21	700

Tag Reverse Primer	AcDx-5937-RIMS2-S1-RT-RP	TGCGGGTAGATCGACTCACA	20	701
Downstream PCR Primer	AcDx-5938-RIMS2-S1-PCR-V	TGCGGGTAGATCGACTCACACCCGAAACCCGACAAAAACTGrACATG/3SpC3/	46	702
DAB2IP				
Forward PCR Primer	AcDx-5941-DAB2IP-S1-FP	CGTGCGTAGGGTTTTTTTAGTCrGTCGC/3SpC3/	27	703
Reverse PCR Primer	AcDx-5942-DAB2IP-S1-RP	GGTGTCGTGGCGACGCTCCGAAACCCrCGACG/3SpC3/	31	704
Upstream LDR	AcDx-5943-DAB2IP-S1-Up	TCCTGAGGGACAAATACACACCTTTTTTAGTCGTCGTTTTAAGGGTTTTATTAGGCrGTACC /3SpC3/	61	705
Downstream LDR	AcDx-5944-DAB2IP-S1-Dn	/5Phos/GTATTAAGAGTTAGTTTAAGTTGGATCGTAATTATAGTTTTCGTTATATTTTGTCG GTAGGTAAGGAAGTCACGCA	76	706
Real-Time Probe	AcDx-5945-DAB2IP-S1-RT-Pb	/56-FAM/CCATTAGGC/ZEN/GTATTAAGAGTTAGTTTAAGTTGGATCGTAATTATAG/3IABkFQ/	46	707
Tag Forward Primer	AcDx-5946-DAB2IP-S1-RT-FP	TCCTGAGGGACAAATACACACC	22	708
Tag Reverse Primer	AcDx-5947-DAB2IP-S1-RT-RP	TGCGTGACTTCCTTACCTACC	21	709
Downstream PCR Primer	AcDx-5948-DAB2IP-S1-PCR-V	TGCGTGACTTCCTTACCTACCCTCCGAAACCCCGACAAAATATAATGrAAAAT/3SpC3/	52	710
GLB1L3				
Forward PCR Primer	AcDx-5951-GLB1L3-S1-FP	GATCGTTCGTTGTGTTTCGCrGTTGT/3SpC3/	25	711
Reverse PCR Primer	AcDx-5952-GLB1L3-S1-RP	GGTGTCGTGGCGACCTCGAACTTCCTAACCrUAAAC/3SpC3/	35	712
Upstream LDR	AcDx-5953-GLB1L3-S1-Up	TTGTTCGCCCGTTGGTCACTTGTGTTTCGCGTTGCGTTTTTCTCrGTTCC/3SpC3/	49	713
Downstream LDR	AcDx-5954-GLB1L3-S1-Dn	/5Phos/GTTTTTTTTTCGCGTAGGTTTTTTTGTGTTAGTCGGTGTGGAGCGCTAAGGTTGCA-	56	714
Real-Time Probe	AcDx-5955-GLB1L3-S1-RT-Pb	/56-FAM/CCTTTTCTC/ZEN/GTTTTTTTTTCGCGTAGGTTTTTTTGT/3IABkFQ/	36	715
Tag Forward Primer	AcDx-5956-GLB1L3-S1-RT-FP	TTGTTCGCCCGTTGGTCAC	19	716
Tag Reverse Primer	AcDx-5957-GLB1L3-S1-RT-RP	TGCAACCTTAGCGCTCCAC	19	717
Downstream PCR Primer	AcDx-5958-GLB1L3-S1-PCR-V	TGCAACCTTAGCGCTCCACGAACTTCCTAACCTAAATCGACAACTGrACTAG/3SpC3/	51	718
DBX2				
Forward PCR Primer	AcDx-5961-DBX2-S1-FP	GCGGCGGATTTGGAACrGATAC/3SpC3/	21	719

Reverse PCR Primer	AcDx-5962-DBX2-S1-RP	GGTGTCGTGGGACCAAAAAAACTCACAACTTCCrCCCTT/3SpC3/	38	720
Upstream LDR	AcDx-5963-DBX2-S1-Up	TAGGAACACGGAGGACATCAACGGCGGATTTGGAACGATATTATTTATTCGCrGTTAA/3SpC3/	57	721
Downstream LDR	AcDx-5964-DBX2-S1-Dn	/5Phos/GTTGGGCGTTTTTTTAAGGGATGGGAGGATTGTGGGTGGGTATAGGTCAGA	51	722
Real-Time Probe	AcDx-5965-DBX2-S1-RT-Pb	/56-FAM/CCTATTCGC/ZEN/GTTGGGCGTTTTTTTAAGGG/3IABkFQ/	29	723
Tag Forward Primer	AcDx-5966-DBX2-S1-RT-FP	TAGGAACACGGAGGACATCAA	21	724
Tag Reverse Primer	AcDx-5967-DBX2-S1-RT-RP	TCTGACCTATACCCACCCACAA	22	725
DPY19L2				
Forward PCR Primer	AcDx-5971-DPY19L2-S1-FP	ATATGCGTAGTTTTTATTTTGCGCr/3SpC3/	24	726
Reverse PCR Primer	AcDx-5972-DPY19L2-S1-RP	GGTGTCGTGGTACACTCCCAATAAAACGCGTAArAAAAT/3SpC3/	38	727
Upstream LDR	AcDx-5973-DPY19L2-S1-Up	TTCGGCAGGCTACGGTACAGTAGTTTTTATTTTGCGCGTAGTTTTATGTATAGTCTCrGGATA/3SpC3/	62	728
Downstream LDR	AcDx-5974-DPY19L2-S1-Dn	/5Phos/GGACGGGTTTTTAGTTTTATTGGCGTAGAATTTATTATCGTTGTCGAACCGTTTTAGGACTGA	63	729
Real-Time Probe	AcDx-5975-DPY19L2-S1-RT-Pb	/56-FAM/CCTAGTCTC/ZEN/GGACGGGTTTTTAGTTTTATTGGC/3IABkFQ/	33	730
Tag Forward Primer	AcDx-5976-DPY19L2-S1-RT-FP	TTCGGCAGGCTACGGTACA	19	731
Tag Reverse Primer	AcDx-5977-DPY19L2-S1-RT-RP	TCAGTCCTAAAACGGTTCGACA	22	732
Downstream PCR Primer	AcDx-5978-DPY19L2-S1-PCR-V	TCAGTCCTAAAACGGTTCGACATCCCAATAAAACGCGTAAAAAACAAATGrATAAC/3SpC3/	55	733
ZNF829,ZNF568				
Forward PCR Primer	AcDx-5981-ZNF829,ZNF568-S1-FP	GGAGGGATGGTTCGGCrGTTTC/3SpC3/	21	734
Reverse PCR Primer	AcDx-5982-ZNF829,ZNF568-S1-RP	GGTGTCGTGGACCCGAATATTCATCCCGCrGCGCG/3SpC3/	34	735
Upstream LDR	AcDx-5983-ZNF829,ZNF568-S1-Up	TAAACAATGAGACCCGCTGAACGGGATGGTTCGGCGTTTTAAGCrGTTCA/3SpC3/	49	736
Downstream LDR	AcDx-5984-ZNF829,ZNF568-S1-Dn	/5Phos/GTTTGTTATAGATTTATTTGCGGGTCGTTTTTATTTAGTATTTTAGAAATGTTGTCGCATAGGCAGTTCATA	72	737
Real-Time Probe	AcDx-5985-ZNF829,ZNF568-	/56-FAM/CCTTTAAGC/ZEN/GTTTGTTATAGATTTATTTGCGGGTCG/3IABkFQ/	36	738

	S1-RT-Pb			
Tag Forward Primer	AcDx-5986-ZNF829,ZNF568-S1-RT-FP	TAAACAATGAGACCCGCTGAAC	22	739
Tag Reverse Primer	AcDx-5987-ZNF829,ZNF568-S1-RT-RP	TATGAACTGCCTATGCGACAAC	22	740
Downstream PCR Primer	AcDx-5988-ZNF829,ZNF568-S1-PCR-V	TATGAACTGCCTATGCGACAACCCGAATATTCATCCCGCGTGrCAATC/3SpC3/	47	741
ZNF334				
Forward PCR Primer	AcDx-5991-ZNF334-S1-FP	CGTTTAGGTAAAAAATAGGAATAGTATATGCrGTAGG/3SpC3/	36	742
Reverse PCR Primer	AcDx-5992-ZNF334-S1-RP	GGTGTCGTGGCGACGCCTTCTAAAAACTATAATCCrGAAAG/3SpC3/	40	743
Upstream LDR	AcDx-5993-ZNF334-S1-Up	TAAGACGTATGCTAGCGCCAAGGTAAAAAATAGGAATAGTATATGCGTAGAAAGGTTCTCrGGGCT/3SpC3/	65	744
Downstream LDR	AcDx-5994-ZNF334-S1-Dn	/5Phos/GGGTCGTTTTTTTCGAGGTTTTTTCGGGTTGGTCAGCATCGACTCCTA	48	745
Real-Time Probe	AcDx-5995-ZNF334-S1-RT-Pb	/56-FAM/TTGGTTCTC/ZEN/GGGTCGTTTTTTTCGAGGTTTTTTC/3IABkFQ/	34	746
Tag Forward Primer	AcDx-5996-ZNF334-S1-RT-FP	TAAGACGTATGCTAGCGCCAA	21	747
Tag Reverse Primer	AcDx-5997-ZNF334-S1-RT-RP	TAGGAGTCGATGCTGACCAA	20	748
Downstream PCR Primer	AcDx-5998-ZNF334-S1-PCR-V	TAGGAGTCGATGCTGACCAACCTTCTAAAAACTATAATCCGAAAAACCGATGrAAAAG/3SpC3/	57	749
CoCaNCR1				
Forward PCR Primer	AcDx-6001-CoCaNCR1-S1-FP	ATTGAGTTTTGATACGTAGGATTAGCrGTTAC/3SpC3/	31	750
Reverse PCR Primer	AcDx-6002-CoCaNCR1-S1-RP	GGTGTCGTGGCAATAAAAATAACCCAAACAAACTCCrCCAAG/3SpC3/	41	751
Upstream LDR	AcDx-6003-CoCaNCR1-S1-Up	TTTCCGCCGCTACAACCAATGATACGTAGGATTAGCGTTATTAATAGATTTGTTTCTCrGGTCC/3SpC3/	63	752
Downstream LDR	AcDx-6004-CoCaNCR1-S1-Dn	/5Phos/GGTTTAGTTTTTTTTAGGGTTTTTGTTGGAAATCGATTTTATTTGGTTGAAGCAGCGTCTGAGCA	65	753
Real-Time Probe	AcDx-6005-CoCaNCR1-S1-RT-Pb	/56-FAM/CCGTTTCTC/ZEN/GGTTTAGTTTTTTTTAGGGTTTTTGTTGGAA/3IABkFQ/	40	754
Tag Forward Primer	AcDx-6006-CoCaNCR1-S1-RT-FP	TTTCCGCCGCTACAACCAA	19	755
Tag Reverse Primer	AcDx-6007-CoCaNCR1-S1-RT-RP	TGCTCAGACGCTGCTTCAA	19	756

Downstream PCR Primer	AcDx-6008-CoCaNCR1-S1-PCR-V	TGCTCAGACGCTGCTTCAACCCAAACAAACTCCCCAAATAAAATTGrATTTT/3SpC3/	51	757
CoCaNCR3				
Forward PCR Primer	AcDx-6011-CoCaNCR3-S1-FP	GGTTTTGCGATTTTTAGGTAAGACrGTAGC/3SpC3/	29	758
Reverse PCR Primer	AcDx-6012-CoCaNCR3-S1-RP	GGTGTCGTGGAAACGATAACGAAAAAAACAAAAAAAArACTTG/3SpC3/	42	759
Upstream LDR	AcDx-6013-CoCaNCR3-S1-Up	TAGCATTCGAGAACGCACCAGACGTAGTTTAGGAGGTTTTTTTTCGAATCrGCGCC/3SpC3/	55	760
Downstream LDR	AcDx-6014-CoCaNCR3-S1-Dn	/5Phos/GCGTTTCGGATTACGTCGTAATGTTTCGTTATTTTAGTAATAAGGGTGCTAGTCACACAGTTCCA	65	761
Real-Time Probe	AcDx-6015-CoCaNCR3-S1-RT-Pb	/56-FAM/AATCGAATC/ZEN/GCGTTTCGGATTACGTCGTA/3IABkFQ/	29	762
Tag Forward Primer	AcDx-6016-CoCaNCR3-S1-RT-FP	TAGCATTCGAGAACGCACC	19	763
Tag Reverse Primer	AcDx-6017-CoCaNCR3-S1-RT-RP	TGGAACTGTGTGACTAGCACC	21	764
Downstream PCR Primer	AcDx-6018-CoCaNCR3-S1-PCR-V	TGGAACTGTGTGACTAGCACCGATAACGAAAAAAACAAAAAAAAACTTATTACTAAAATAATGrAAACG/3SpC3/	68	765
CoCaNCR4				
Forward PCR Primer	AcDx-6021-CoCaNCR4-S1-FP	GTAAGTTTTAGATAAGGGATTTTTAATTATAGATArAAAGA/3SpC3/	40	766
Reverse PCR Primer	AcDx-6022-CoCaNCR4-S1-RP	GGTGTCGTGGCACCCCCCAACGTTCTArUTTCT/3SpC3/	32	767
Upstream LDR	AcDx-6023-CoCaNCR4-S1-Up	TTCGTCCCTGCACGCTAACGTTTTAGATAAGGGATTTTTAATTATAGATAAAAGGTAAATAAAGTCrGTACG/3SpC3/	71	768
Downstream LDR	AcDx-6024-CoCaNCR4-S1-Dn	/5Phos/GTATATTATATTTTTTACGTGTTAGTTGACGGTCGCGATAAAAGGGTTCCATCACCGTTAGGCCA	65	769
Real-Time Probe	AcDx-6025-CoCaNCR4-S1-RT-Pb	/56-FAM/CCTAAAGTC/ZEN/GTATATTATATTTTTTACGTGTTAGTTGACGGTCG/3IABkFQ/	44	770
Tag Forward Primer	AcDx-6026-CoCaNCR4-S1-RT-FP	TTCGTCCCTGCACGCTAAC	19	771
Tag Reverse Primer	AcDx-6027-CoCaNCR4-S1-RT-RP	TGGCCTAACGGTGATGGAAC	20	772
Downstream PCR Primer	AcDx-6028-CoCaNCR4-S1-PCR-V	TGGCCTAACGGTGATGGAACCCCCCAACGTTCTATTTCCTTTTATTGrCGACT/3SpC3/	52	773

CoCaNCR6				
Forward PCR Primer	AcDx-6031-CoCaNCR6-S1-FP	TTACGTTTTTTTTTCGGTTCGTTArGTTGC/3SpC3/	29	774
Reverse PCR Primer	AcDx-6032-CoCaNCR6-S1-RP	GGTGTCGTGGCCCTCAACCGCCGAATCrUCCCA/3SpC3/	32	775
Upstream LDR	AcDx-6033-CoCaNCR6-S1-Up	TTGGTACGAGGAGGGCACATTTTTCGGTTCGTTAGTTGTTTTTTAGCrGTTAC/3SpC3/	52	776
Downstream LDR	AcDx-6034-CoCaNCR6-S1-Dn	/5Phos/GTTGTTTTTAGATTTGTTGCGTTTCGATTTGGAGGTGTCTGCCGCCCTTACTAA	54	777
Real-Time Probe	AcDx-6035-CoCaNCR6-S1-RT-Pb	/56-FAM/CCTTTTAGC/ZEN/GTTGTTTTTAGATTTGTTGCGTTTCGA/3IABkFQ/	36	778
Tag Forward Primer	AcDx-6036-CoCaNCR6-S1-RT-FP	TTGGTACGAGGAGGGCACA	19	779
Tag Reverse Primer	AcDx-6037-CoCaNCR6-S1-RT-RP	TTAGTAAGGGCGGCAGACA	19	780
Downstream PCR Primer	AcDx-6038-CoCaNCR6-S1-PCR-V	TTAGTAAGGGCGGCAGACATCAACCGCCGAATCTCCTGrAACCG/3SpC3/	43	781
CoCaNCR7				
Forward PCR Primer	AcDx-6041-CoCaNCR7-S1-FP	TTCGGGTTTGGGTCGCrGGTTC/3SpC3/	21	782
Reverse PCR Primer	AcDx-6042-CoCaNCR7-S1-RP	GGTGTCGTGGCAATCTCAAAACCCGACAACCrGCCCA/3SpC3/	36	783
Upstream LDR	AcDx-6043-CoCaNCR7-S1-Up	TCTCAATGTCGAGCCGTACCGGTTTGGGTCGCGGTTTTTTTTTTCTCrGGCAC/3SpC3/	52	784
Downstream LDR	AcDx-6044-CoCaNCR7-S1-Dn	/5Phos/GGCGTTTCGTTTTGAGCGGAGTAATAATTAGAAGGTTCTTCGGCTGGCTCAA	52	785
Real-Time Probe	AcDx-6045-CoCaNCR7-S1-RT-Pb	/56-FAM/CCTTTTCTC/ZEN/GGCGTTTCGTTTTGAGCGG/3IABkFQ/	28	786
Tag Forward Primer	AcDx-6046-CoCaNCR7-S1-RT-FP	TCTCAATGTCGAGCCGTACC	20	787
Tag Reverse Primer	AcDx-6047-CoCaNCR7-S1-RT-RP	TTGAGCCAGCCGAAGAACC	19	788
Downstream PCR Primer	AcDx-6048-CoCaNCR7-S1-PCR-V	TTGAGCCAGCCGAAGAACCCCCGACAACCGCCTGrAAATC/3SpC3/	39	789

IDT Abbreviation	Modifications
/5Phos/	5' Phosphorylation
rX (X=A,C,G,U)	RNA Base

/3spC3/	3' C3 DNA Spacer
/56-FAM/	5' 6-FAM™ Fluorescent Tag
/Zen/	Internal Quencher
/3IABkFQ/	3' Iowa Black® FQ Quencher

Table 46. Primers for use in Step 1 of the 96-marker assay, with average sensitivities of 50%, detect solid tumors.

Site	Primer Name	Sequence	Length	Seq. ID No.
Preferred Solid Tumor Markers				
DLGAP1-S1				
Forward PCR Primer	AcDx-7001-DLGAP1-S1-FP	GGAGATGTAGATTTCGATGTTTTCrGGCGC/3SpC3/	29	790
Reverse PCR Primer	AcDx-7002-DLGAP1-S1-RP	GGTGTCGTGGAAAACCCGCAAACGCCrUAATG/3SpC3/	31	791
Upstream LDR	AcDx-7003-DLGAP1-S1-Up	TCCCTTAGAGAGAACGCCCAGTTTTCGGTGGTCGAGTTTTAGCrGGATA/3SpC3/	48	792
Downstream LDR	AcDx-7004-DLGAP1-S1-Dn	/5Phos/GGACGGACGCGGCGCGTGGTGACGTACGAGTGTTCTTA	38	793
Real-Time Probe	AcDx-7005-DLGAP1-S1-RT-Pb	/56-FAM/CCTTTTAGC/ZEN/GGACGGACGCGGC/3IABkFQ/	22	794
Tag Forward Primer	AcDx-7006-DLGAP1-S1-RT-FP	TCCCTTAGAGAGAACGCCCA	20	795
Tag Reverse Primer	AcDx-7007-DLGAP1-S1-RT-RP	TAAGAACACTCGTACGTCACCA	22	796
Downstream PCR Primer	AcDx-7008-DLGAP1-S1-PCR-V	TAAGAACACTCGTACGTCACCACCCGCAAACGCCTAATAACTGrCCAAG/3SpC3/	48	797
VAMP5				
Forward PCR Primer	AcDx-7011-VAMP5-FP	TCGGGAGGGTTCGATTTTACrGGATC/3SpC3/	25	798
Reverse PCR Primer	AcDx-7012-VAMP5-RP	GGTGTCGTGGCGAAAAACGCGCCTTCCrGCGAG/3SpC3/	32	799
Upstream LDR	AcDx-7013-VAMP5-Up	TCTCATACCAGACGCGGTAACGGAGGGTTCGATTTTACGGATTTAGATCrGTTAC/3SpC3/	54	800

Downstream LDR	AcDx-7014-VAMP5-Dn	/5Phos/GTTGTGGTTTATCGTTTTCGATTTGATTTGGTTTTTGTCGGTTCGTGTCGCTGTG CTTA	59	801
Real-Time Probe	AcDx-7015-VAMP5-RT-Pb	/56-FAM/CCTTAGATC/ZEN/GTTGTGGTTTATCGTTTTCGATTTGATTTG/3IABkFQ/	39	802
Tag Forward Primer	AcDx-7016-VAMP5-RT-FP	TCTCATACCAGACGCGGTAAC	21	803
Tag Reverse Primer	AcDx-7017-VAMP5-RT-RP	TAAGCACAGCGACACGAAC	19	804
Downstream PCR Primer	AcDx-7018-VAMP5-PCR-V	TAAGCACAGCGACACGAACCGCCTTCCGCGAATAATGrACAAG/3SpC3/	42	805
CaNCR20				
Forward PCR Primer	AcDx-7021-CaNCR20-FP	AGTTGCGGGTCGGGTArGTGAC/3SpC3/	21	806
Reverse PCR Primer	AcDx-7022-CaNCR20-RP	GGTGTCGTGGCTCTAAAATAAAATACGCAATAAACAACCrAAACA/3SpC3/	44	807
Upstream LDR	AcDx-7023-CaNCR20-Up	TTTTCGGCGGCAGCTAAACCGGGTCGGGTAGTGATTGATAGATCrGGGCG/3SpC3/	49	808
Downstream LDR	AcDx-7024-CaNCR20-Dn	/5Phos/GGGAATAGGGTTTTTCGTTTTTTTTTCGTTTGGGTGTTCGTGTCGCTGTGCTTA	54	809
Real-Time Probe	AcDx-7025-CaNCR20-RT-Pb	/56-FAM/CCATAGATC/ZEN/GGGAATAGGGTTTTTCGTTTTTTTTTCG/3IABkFQ/	37	810
Tag Forward Primer	AcDx-7026-CaNCR20-RT-FP	TTTTCGGCGGCAGCTAAAC	19	811
Tag Reverse Primer	AcDx-7027-CaNCR20-RT-RP	TAAGCACAGCGACACGAAC	19	812
Downstream PCR Primer	AcDx-7028-CaNCR20-PCR-V	TAAGCACAGCGACACGAACCTCTAAAATAAAATACGCAATAAACAACCAAATGrCCCCT /3SpC3/	58	813
ATP6V1B1				
Forward PCR Primer	AcDx-7031-ATP6V1B1-FP	TTTTTCGTTCGATTTTTTTTTTCGCrGTTTC/3SpC3/	30	814
Reverse PCR Primer	AcDx-7032-ATP6V1B1-RP	GGTGTCGTGGACATAAAAATACAAATACTCCCGTCAArUATAG/3SpC3/	42	815
Upstream LDR	AcDx-7033-ATP6V1B1-Up	TGATGCTGGCAAACCCTAGAACTCGATTTTTTTTTTCGCGTTTTATGTTTTTTTCTCrGATC C/3SpC3/	62	816
Downstream LDR	AcDx-7034-ATP6V1B1-Dn	/5Phos/GATTTTCGGTGTTGCGGAAGAATTGAAGGTTGGTTCCATCACCGTTAGGCCA	52	817
Real-Time Probe	AcDx-7035-ATP6V1B1-RT-Pb	/56-FAM/CCTTTTCTC/ZEN/GATTTTCGGTGTTGCGGAAG/3IABkFQ/	29	818
Tag Forward Primer	AcDx-7036-ATP6V1B1-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	819

Tag Reverse Primer	AcDx-7037-ATP6V1B1-RT-RP	TGGCCTAACGGTGATGGAAC	20	820
Downstream PCR Primer	AcDx-7038-ATP6V1B1-PCR-V	TGGCCTAACGGTGATGGAACCAAATACTCCCGTCAAAATAAAACATTGrCAACT/3SpC3/	53	821
CaNCR21				
Forward PCR Primer	AcDx-7041-CaNCR21-FP	AAATTCGTTCGGTTTATTTAAGTTTCrGTTGC/3SpC3/	31	822
Reverse PCR Primer	AcDx-7042-CaNCR21-RP	GGTGTCGTGGCCTTTAACTTCCTCCTCGATTCCrCCCAG/3SpC3/	38	823
Upstream LDR	AcDx-7043-CaNCR21-Up	TTAGCCGCCAAACGTACCACGTTTTTTAGGGTCGTTTTTGGGCACrGGGTA/3SpC3/	50	824
Downstream LDR	AcDx-7044-CaNCR21-Dn	/5Phos/GGGCGGATTTTTCGTTAATATTTTGTTTGTAAGATTTTTTATTGTGGGCAGGAACACGATAGTA	64	825
Real-Time Probe	AcDx-7045-CaNCR21-RT-Pb	/56-FAM/AATGGGCAC/ZEN/GGGCGGATTTTTCGTTA/3IABkFQ/	26	826
Tag Forward Primer	AcDx-7046-CaNCR21-RT-FP	TTAGCCGCCAAACGTACCA	19	827
Tag Reverse Primer	AcDx-7047-CaNCR21-RT-RP	TACTATCGTGTTCCTGCCCA	20	828
UBTF				
Forward PCR Primer	AcDx-7051-UBTF-FP	GGCGTTTTCGTCGGCrGGGTG/3SpC3/	20	829
Reverse PCR Primer	AcDx-7052-UBTF-RP	GGTGTCGTGGTCGGTTGTTGGGCGTrAAAAT/3SpC3/	30	830
Upstream LDR	AcDx-7053-UBTF-Up	TAGCAGCTGAACAACCCAACGGTGTAGATGTTTTTCGTTTGAGGCTCrGTTCC/3SpC3/	52	831
Downstream LDR	AcDx-7054-UBTF-Dn	/5Phos/GTTTTCGTTTGGTTTGCGGCGTTTAGTTGATAGGTTGTATGGTCGGCATGCTA	53	832
Real-Time Probe	AcDx-7055-UBTF-RT-Pb	/56-FAM/AAGAGGCTC/ZEN/GTTTTCGTTTGGTTTGCG/3IABkFQ/	27	833
Tag Forward Primer	AcDx-7056-UBTF-RT-FP	TAGCAGCTGAACAACCCAAC	20	834
Tag Reverse Primer	AcDx-7057-UBTF-RT-RP	TAGCATGCCGACCATACAAC	20	835
Downstream PCR Primer	AcDx-7058-UBTF-PCR-V	TAGCATGCCGACCATACAACACGCCCAACAACCGAAAAATGrAAAAG/3SpC3/	46	836
LOC284100				
Forward PCR Primer	AcDx-7061-LOC284100-FP	TTAATTTTCGTTTAGGTTTTCGTTTCrGGGTC/3SpC3/	31	837

Reverse PCR Primer	AcDx-7062-LOC284100-RP	GGTGTCGTGGCTAAACAATCAAACGAACTAAAATACGArUTCCC/3SpC3/	43	838
Upstream LDR	AcDx-7063-LOC284100-Up	TCTGCCCTTCGCTTCGAACGTTTTCGAGGGTTTATTTTTTAGTTTTGAGGCrGGTGG/3SpC3/	56	839
Downstream LDR	AcDx-7064-LOC284100-Dn	/5Phos/GGTAAACGTTTTATTTTTAGTGGCGAGTGCGAGGTTGTATGGTCGGCATGCTA	53	840
Real-Time Probe	AcDx-7065-LOC284100-RT-Pb	/56-FAM/AATTGAGGC/ZEN/GGTAAACGTTTTATTTTTAGTGGCG/3IABkFQ/	34	841
Tag Forward Primer	AcDx-7066-LOC284100-RT-FP	TCTGCCCTTCGCTTCGAAC	19	842
Tag Reverse Primer	AcDx-7067-LOC284100-RT-RP	TAGCATGCCGACCATACAAC	20	843
Downstream PCR Primer	AcDx-7068-LOC284100-PCR-V	TAGCATGCCGACCATACAACCAATCAAACGAACTAAAATACGATTCCTTGrCACTT/3SpC3/	55	844
HOXA9-S1				
Forward PCR Primer	AcDx-7071-HOXA9-S1-FP	GTTAGCGTCGTCGTTTGTCrGGGAG/3SpC3/	24	845
Reverse PCR Primer	AcDx-7072-HOXA9-S1-RP	GGTGTCGTGGGTAAACTCGTTCCTACTAAACGCrCGACA/3SpC3/	38	846
Upstream LDR	AcDx-7073-HOXA9-S1-Up	TTGCAAACCACCCGGACAAGAGGAGGTTGGTTTAGGGTTCTCrGGCAC/3SpC3/	47	847
Downstream LDR	AcDx-7074-HOXA9-S1-Dn	/5Phos/GGCGTATAGCGGTTAACGTTTAGTTTATTCGCGGTTGGTCAGCATCGACTCCTA	54	848
Real-Time Probe	AcDx-7075-HOXA9-S1-RT-Pb	/56-FAM/TTGGTTCTC/ZEN/GGCGTATAGCGGTTAACGTTTAGT/3IABkFQ/	33	849
Tag Forward Primer	AcDx-7076-HOXA9-S1-RT-FP	TTGCAAACCACCCGGACAA	19	850
Tag Reverse Primer	AcDx-7077-HOXA9-S1-RT-RP	TAGGAGTCGATGCTGACCAA	20	851
Downstream PCR Primer	AcDx-7078-HOXA9-S1-PCR-V	TAGGAGTCGATGCTGACCAACCTACTAAACGCCGACGCTGrCGAAC/3SpC3/	45	852
WNT7B-S1				
Forward PCR Primer	AcDx-7081-WNT7B-S1-FP	CGTGTATGTCGGTGTTTGTACrGAGGA/3SpC3/	26	853
Reverse PCR Primer	AcDx-7082-WNT7B-S1-RP	GGTGTCGTGGTCCTAAACCAACGAAAAACCCrCTCCT/3SpC3/	36	854

Upstream LDR	AcDx-7083-WNT7B-S1-Up	TAAGACGTATGCTAGCGCCAAGAGCGGGTGTGTGAGCGCrGGTCC/3SpC3/	44	855
Downstream LDR	AcDx-7084-WNT7B-S1-Dn	/5Phos/GGTTTTTTTAAGTGTGGTATGGTATTGCGCGTCGTTGGTCAGCATCGACTCCTA	54	856
Real-Time Probe	AcDx-7085-WNT7B-S1-RT-Pb	/56-FAM/AATGAGCGC/ZEN/GGTTTTTTTAAGTGTGGTATGGT/3IABkFQ/	32	857
Tag Forward Primer	AcDx-7086-WNT7B-S1-RT-FP	TAAGACGTATGCTAGCGCCAA	21	858
Tag Reverse Primer	AcDx-7087-WNT7B-S1-RT-RP	TAGGAGTCGATGCTGACCAA	20	859
Downstream PCR Primer	AcDx-7088-WNT7B-S1-PCR-V	TAGGAGTCGATGCTGACCAACATATTAACCCACGCGACGTGrCAATG/3SpC3/	46	860
HOXD9-S1				
Forward PCR Primer	AcDx-7091-HOXD9-S1-FP	GATTACGTGGGTCGCGCrGATTG/3SpC3/	22	861
Reverse PCR Primer	AcDx-7092-HOXD9-S1-RP	GGTGTCGTGGACATTTTAAAACGTCCCGCACrUCCCG/3SpC3/	36	862
Upstream LDR	AcDx-7093-HOXD9-S1-Up	TACATGCCATCCCACGACATCGGTGGTTCGGGCATCrGGCAA/3SpC3/	41	863
Downstream LDR	AcDx-7094-HOXD9-S1-Dn	/5Phos/GGCGAGGAGTTGTTCGGCGGTGTGTCGGAGCGGTTACTA	39	864
Real-Time Probe	AcDx-7095-HOXD9-S1-RT-Pb	/56-FAM/AAGAGCATC/ZEN/GGCGAGGAGTTG/3IABkFQ/	21	865
Tag Forward Primer	AcDx-7096-HOXD9-S1-RT-FP	TACATGCCATCCCACGACA	19	866
Tag Reverse Primer	AcDx-7097-HOXD9-S1-RT-RP	TAGTAACCGCTCCGACACA	19	867
Downstream PCR Primer	AcDx-7098-HOXD9-S1-PCR-V	TAGTAACCGCTCCGACACAACATTACACTATCCGCCGCTGrAACAG/3SpC3/	45	868
NRN1				
Forward PCR Primer	AcDx-7101-NRN1-FP	GGTAGTTTTTTGGCGGTTGCrGTTTA/3SpC3/	25	869
Reverse PCR Primer	AcDx-7102-NRN1-RP	GGTGTCGTGGCTTCGACGTCTAACCCGArCGCTT/3SpC3/	33	870
Upstream LDR	AcDx-7103-NRN1-Up	TAGGGCGACAGTTACCACAAGTGTTTCGGGAGGATCGGATATTTTAATTTCTCrGGTCC/3SpC3/	58	871
Downstream LDR	AcDx-7104-NRN1-Dn	/5Phos/GGTTTTTAACGCGGGCGTTTGTTCGCGTTGTGGGTCTCGCTCGTATA	47	872

Real-Time Probe	AcDx-7105-NRN1-RT-Pb	/56-FAM/CCATTTCTC/ZEN/GGTTTTTAACGCGGGCGTTTGT/3IABkFQ/	31	873
Tag Forward Primer	AcDx-7106-NRN1-RT-FP	TAGGGCGACAGTTACCACAA	20	874
Tag Reverse Primer	AcDx-7107-NRN1-RT-RP	TATACGAGCGAGACCCACAA	20	875
Downstream PCR Primer	AcDx-7108-NRN1-PCR-V	TATACGAGCGAGACCCACAATCTAACCCGACGCTCGTGrAACAG/3SpC3/	43	876
CaNCR23				
Forward PCR Primer	AcDx-7111-CaNCR23-FP	ATTTGTTATTCGCGTGCGTCrGTTTC/3SpC3/	25	877
Reverse PCR Primer	AcDx-7112-CaNCR23-RP	GGTGTCGTGGATTCGAAACACTACTCTAATACGATCCrUAATG/3SpC3/	42	878
Upstream LDR	AcDx-7113-CaNCR23-Up	TCCGACTTTAGTGCGTCACAAATTTTGGTTATATTTTTTTTTTATTTGCGGTGTTTTATCTCrGCGCG/3SpC3/	66	879
Downstream LDR	AcDx-7114-CaNCR23-Dn	/5Phos/GCGTAAGATGCGTTGATAGAGGTTATTTTAAACGAATTTTTGTGGGTCTCGCTCGTATA	59	880
Real-Time Probe	AcDx-7115-CaNCR23-RT-Pb	/56-FAM/AATTATCTC/ZEN/GCGTAAGATGCGTTGATAGAGGTTATT/3IABkFQ/	36	881
Tag Forward Primer	AcDx-7116-CaNCR23-RT-FP	TCCGACTTTAGTGCGTCACAA	21	882
Tag Reverse Primer	AcDx-7117-CaNCR23-RT-RP	TATACGAGCGAGACCCACAA	20	883
Downstream PCR Primer	AcDx-7118-CaNCR23-PCR-V	TATACGAGCGAGACCCACAAAACACTACTCTAATACGATCCTAATAAAAAATAAATTTGrUTTAG/3SpC3/	64	884
PRRX1				
Forward PCR Primer	AcDx-7121-PRRX1-FP	GGATGTAAATATAAAATAGCGACGCrGGGAG/3SpC3/	30	885
Reverse PCR Primer	AcDx-7122-PRRX1-RP	GGTGTCGTGGGGCGAAAAAAAATTAACGAAACAAAATCrCCGAG/3SpC3/	43	886
Upstream LDR	AcDx-7123-PRRX1-Up	TCAAACAAAGGCGACCACAACATCGATTTGTTATAAAGGGAGAGGTGTAGACrGTAAT/3SpC3/	57	887
Downstream LDR	AcDx-7124-PRRX1-Dn	/5Phos/GTAGCGTAAAGGAATTGTTTGTTTAATTTATTAGTTATATTTTTTCGGGTTGTCGCATAGGCAGTTCATA	70	888
Real-Time Probe	AcDx-7125-PRRX1-RT-Pb	/56-FAM/AATGTAGAC/ZEN/GTAGCGTAAAGGAATTGTTTGTTTAATTTATT/3IABkFQ/	41	889
Tag Forward Primer	AcDx-7126-PRRX1-RT-FP	TCAAACAAAGGCGACCACAAC	21	890

Tag Reverse Primer	AcDx-7127-PRRX1-RT-RP	TATGAACTGCCTATGCGACAAC	22	891
Downstream PCR Primer	AcDx-7128-PRRX1-PCR-V	TATGAACTGCCTATGCGACAACGCGAAAAAAAATTAACGAAACAAAATCCTGrAAAG/3SpC3/	56	892
CaNCR24				
Forward PCR Primer	AcDx-7131-CaNCR24-FP	CGAGTTGTAAAGTTGTTGTCGCrGGCGC/3SpC3/	27	893
Reverse PCR Primer	AcDx-7132-CaNCR24-RP	GGTGTCGTGGCGAACGAAAAAAAACCCCGACrCAAAT/3SpC3/	36	894
Upstream LDR	AcDx-7133-CaNCR24-Up	TAAACAATGAGACCCGCTGAACGGCGTCGGGAACGGCGCrGCGCC/3SpC3/	44	895
Downstream LDR	AcDx-7134-CaNCR24-Dn	/5Phos/GCGTTTAATTTTTAGCGGGAGTCGTTAGGTTTGGTTGTCGCATAGGCAGTTCATA	55	896
Real-Time Probe	AcDx-7135-CaNCR24-RT-Pb	/56-FAM/TTACGGCGC/ZEN/GCGTTTAATTTTTAGCGG/3IABkFQ/	27	897
Tag Forward Primer	AcDx-7136-CaNCR24-RT-FP	TAAACAATGAGACCCGCTGAAC	22	898
Tag Reverse Primer	AcDx-7137-CaNCR24-RT-RP	TATGAACTGCCTATGCGACAAC	22	899
Downstream PCR Primer	AcDx-7138-CaNCR24-PCR-V	TATGAACTGCCTATGCGACAACCCCGACCAAACCAAACCTAATGrACTT/3SpC3/	48	900
ISL2-S1				
Forward PCR Primer	AcDx-7141-ISL2-S1-FP	GTGGTCGTCGTTTTGTTTTTTTACrGTTTC/3SpC3/	29	901
Reverse PCR Primer	AcDx-7142-ISL2-S1-RP	GGTGTCGTGGCCCGAACGAAACCGCCrAAAAG/3SpC3/	31	902
Upstream LDR	AcDx-7143-ISL2-S1-Up	TCCGGATCAAAGCAGCCACTTTTATTTCGGTCGGTTCGGAGCTCrGTTCG/3SpC3/	49	903
Downstream LDR	AcDx-7144-ISL2-S1-Dn	/5Phos/GTTTACGCGGATTTTCGTTTTGTTTTAGTTTAGCGTGTGTGTTGGCGTACGGTGA	55	904
Real-Time Probe	AcDx-7145-ISL2-S1-RT-Pb	/56-FAM/AAGGAGCTC/ZEN/GTTTACGCGGATTTTCGTTTTG/3IABkFQ/	31	905
Tag Forward Primer	AcDx-7146-ISL2-S1-RT-FP	TCCGGATCAAAGCAGCCAC	19	906
Tag Reverse Primer	AcDx-7147-ISL2-S1-RT-RP	TCACCGTACGCCAACACAC	19	907
Downstream PCR Primer	AcDx-7148-ISL2-S1-PCR-V	TCACCGTACGCCAACACACCGAACGAAACCGCCAAAAAATGrCTAAG/3SpC3/	46	908

NR5A2-S1				
Forward PCR Primer	AcDx-7151-NR5A2-S1-FP	ATGTGCGGGTCGGCrGGGTC/3SpC3/	19	909
Reverse PCR Primer	AcDx-7152-NR5A2-S1-RP	GGTGTCGTGGTCTACTCTCAACACCTCCCAArUCCTC/3SpC3/	36	910
Upstream LDR	AcDx-7153-NR5A2-S1-Up	TCATCGCCCTCAGATCTTCCAGGTCGGCGGGTTTGTGATCTCrGGAGT/3SpC3/	47	911
Downstream LDR	AcDx-7154-NR5A2-S1-Dn	/5Phos/GGAACGTTTTTTGTTATTTTTTTGCGCGAATTTGAAAGTGGAGGATAGATTGGAGGGCA	59	912
Real-Time Probe	AcDx-7155-NR5A2-S1-RT-Pb	/56-FAM/AATGATCTC/ZEN/GGAACGTTTTTTGTTATTTTTTTGCG/3IABkFQ/	35	913
Tag Forward Primer	AcDx-7156-NR5A2-S1-RT-FP	TCATCGCCCTCAGATCTTCCA	21	914
Tag Reverse Primer	AcDx-7157-NR5A2-S1-RT-RP	TGCCCTCCAATCTATCCTCCA	21	915
Downstream PCR Primer	AcDx-7158-NR5A2-S1-PCR-V	TGCCCTCCAATCTATCCTCCACACCTCCCAATCCTTTCAAATTTGrCGCAG/3SpC3/	50	916
SIGIRR				
Forward PCR Primer	AcDx-7161-SIGIRR-FP	GTGGCGGTGTTGGCrGTAGC/3SpC3/	19	917
Reverse PCR Primer	AcDx-7162-SIGIRR-RP	GGTGTCGTGGTCTTCATCACCTTCGAAAACCrAAAAG/3SpC3/	36	918
Upstream LDR	AcDx-7163-SIGIRR-Up	TTCGTGGGCACACAAGCAAGCGGTGTTGGCGTAGTAAGCrGGAAT/3SpC3/	44	919
Downstream LDR	AcDx-7164-SIGIRR-Dn	/5Phos/GGAGCGTCGGGTGCGCGTTGCTTGGCTTGATCTACCTGA	39	920
Real-Time Probe	AcDx-7165-SIGIRR-RT-Pb	/56-FAM/AAAGTAAGC/ZEN/GGAGCGTCGGGTGC/3IABkFQ/	23	921
Tag Forward Primer	AcDx-7166-SIGIRR-RT-FP	TTCGTGGGCACACAAGCAA	19	922
Tag Reverse Primer	AcDx-7167-SIGIRR-RT-RP	TCAGGTAGATCAAGCCAAGCAA	22	923
Downstream PCR Primer	AcDx-7168-SIGIRR-PCR-V	TCAGGTAGATCAAGCCAAGCAAAACCAAAAACGCGACCCTGrCGCAT/3SpC3/	46	924
RASSF1-S1				
Forward PCR Primer	AcDx-7171-RASSF1-S1-FP	GGTACGTTTTAGTCGGGTGCrGGTTC/3SpC3/	25	925
Reverse PCR Primer	AcDx-7172-RASSF1-S1-RP	GGTGTCGTGGACACGAACCCAACCGAACrCATAC/3SpC3/	33	926
Upstream LDR	AcDx-7173-RASSF1-S1-Up	TTGAAGGAGGAAATCGGCACAGCGGTTTTTTTTAGCGCGTTTGGCrGGGCA/3SpC3/	50	927
Downstream LDR	AcDx-7174-RASSF1-S1-Dn	/5Phos/GGGTGTTAGTTTTCGTAGTTTAATGAGTTTAGGTTTTTTCGTGTCGAACCGTTTTAGGACTGA	63	928

Real-Time Probe	AcDx-7175-RASSF1-S1-RT-Pb	/56-FAM/TTGTTTGGC/ZEN/GGGTGTTAGTTTTCGTAGTTTAATGAG/3IABkFQ/	36	929
Tag Forward Primer	AcDx-7176-RASSF1-S1-RT-FP	TTGAAGGAGGAAATCGGCACA	21	930
Tag Reverse Primer	AcDx-7177-RASSF1-S1-RT-RP	TCAGTCCTAAAACGGTTCGACA	22	931
Downstream PCR Primer	AcDx-7178-RASSF1-S1-PCR-V	TCAGTCCTAAAACGGTTCGACACGAACCCAACCGAACCATATTGrAAAAG/3SpC3/	49	932
CCDC151-S1				
Forward PCR Primer	AcDx-7181-CCDC151-S1-FP	TTTGTTCGTTTTCGTTTTTGAGCrGTTTC/3SpC3/	28	933
Reverse PCR Primer	AcDx-7182-CCDC151-S1-RP	GGTGTCGTGGCTAACTCTAACTAAAACTAAATAACCCGrCAAGT/3SpC3/	43	934
Upstream LDR	AcDx-7183-CCDC151-S1-Up	TTCGGCAGGCTACGGTACACGTTTTTAATTGCGCGAAGGTATCrGTTTT/3SpC3/	48	935
Downstream LDR	AcDx-7184-CCDC151-S1-Dn	/5Phos/GTTTTGGGTTAGGAATCGTCGTATTAACGATTGTGTCGAACCGTTTTAGGACTGA	55	936
Real-Time Probe	AcDx-7185-CCDC151-S1-RT-Pb	/56-FAM/TTAGGTATC/ZEN/GTTTTGGGTTAGGAATCGTCGTAT/3IABkFQ/	33	937
Tag Forward Primer	AcDx-7186-CCDC151-S1-RT-FP	TTCGGCAGGCTACGGTACA	19	938
Tag Reverse Primer	AcDx-7187-CCDC151-S1-RT-RP	TCAGTCCTAAAACGGTTCGACA	22	939
Downstream PCR Primer	AcDx-7188-CCDC151-S1-PCR-V	TCAGTCCTAAAACGGTTCGACAACTCTAACTAAAACTAAATAACCCGCAATTGrUTAAC/3SpC3/	58	940
PDE4D				
Forward PCR Primer	AcDx-7191-PDE4D-FP	TGTAGACGAGGAGGCGCrGGGCA/3SpC3/	22	941
Reverse PCR Primer	AcDx-7192-PDE4D-RP	GGTGTCGTGGGAACGCGACCCCGAArCGACA/3SpC3/	30	942
Upstream LDR	AcDx-7193-PDE4D-Up	TCTCGACGATGAAAAGCAACAGATTGTTTGTGTTAGGAGAGCAGCrGGAGG/3SpC3/	50	943
Downstream LDR	AcDx-7194-PDE4D-Dn	/5Phos/GGAAAATTCGGCGTGGAGCGTTTGTAGTTTGTGGGTACTGTCCGTGGA	48	944

Real-Time Probe	AcDx-7195-PDE4D-RT-Pb	/56-FAM/TTGAGCAGC/ZEN/GGAAAATTCGGCGTGGA/3IABkFQ/	26	945
Tag Forward Primer	AcDx-7196-PDE4D-RT-FP	TCTCGACGATGAAAAGCAACA	21	946
Tag Reverse Primer	AcDx-7197-PDE4D-RT-RP	TCCACGGACAGTACCCACA	19	947
Downstream PCR Primer	AcDx-7198-PDE4D-PCR-V	TCCACGGACAGTACCCACACCCGAACGACGAACTACAAATGrCTCCG/3SpC3/	46	948
TNFRSF1B				
Forward PCR Primer	AcDx-7201-TNFRSF1B-FP	GAGGGTATAGTTGGAGGGCrGAGTC/3SpC3/	24	949
Reverse PCR Primer	AcDx-7202-TNFRSF1B-RP	GGTGTCGTGGAAACCTCCCTCGCTTACCrCTATT/3SpC3/	33	950
Upstream LDR	AcDx-7203-TNFRSF1B-Up	TTCTAGGCGACACGACAACAGTTGTTTGTTTGTTTTTTTCGATGCGCTCrGGGAA/3SpC3/	54	951
Downstream LDR	AcDx-7204-TNFRSF1B-Dn	/5Phos/GGGAGTTATTGTAGAGTTTGTTTTTTGCGGTTTTGTGGGTACTGTCCGTGGA	52	952
Real-Time Probe	AcDx-7205-TNFRSF1B-RT-Pb	/56-FAM/TTTGCGCTC/ZEN/GGGAGTTATTGTAGAGTTTG/3IABkFQ/	29	953
Tag Forward Primer	AcDx-7206-TNFRSF1B-RT-FP	TTCTAGGCGACACGACAACA	20	954
Tag Reverse Primer	AcDx-7207-TNFRSF1B-RT-RP	TCCACGGACAGTACCCACA	19	955
Downstream PCR Primer	AcDx-7208-TNFRSF1B-PCR-V	TCCACGGACAGTACCCACAGCTTACCCTATCCCCAAAAACTGrCAAAG/3SpC3/	47	956
KCP				
Forward PCR Primer	AcDx-7211-KCP-FP	GTAGGGATCGTCGGGCrGGGAG/3SpC3/	21	957
Reverse PCR Primer	AcDx-7212-KCP-RP	GGTGTCGTGGCCTTTAATACGCTATCTCGCCrGCAAT/3SpC3/	36	958
Upstream LDR	AcDx-7213-KCP-Up	TTTCAGGCCCTAACCACCACGAGAAGTATTTTTGGTGGCGAGCrGTGAA/3SpC3/	48	959
Downstream LDR	AcDx-7214-KCP-Dn	/5Phos/GTGGGTCGCGTCGGGTCGTGGGTGGGATTAAGGGCGATGGA	41	960
Real-Time Probe	AcDx-7215-KCP-RT-Pb	/56-FAM/AAGGCGAGC/ZEN/GTGGGTCGCG/3IABkFQ/	19	961
Tag Forward Primer	AcDx-7216-KCP-RT-FP	TTTCAGGCCCTAACCACCAC	20	962
Tag Reverse Primer	AcDx-7217-KCP-RT-RP	TCCATCGCCCTTAATCCCAC	20	963
Downstream PCR Primer	AcDx-7218-KCP-PCR-V	TCCATCGCCCTTAATCCCACCCGACTACCCACGACCTGrACGCA/3SpC3/	43	964

CaNCR25				
Forward PCR Primer	AcDx-7221-CaNCR25-FP	GTTTTTTTTAGTTTGGAGTTTTGGTTCrGGGTC/3SpC3/	32	965
Reverse PCR Primer	AcDx-7222-CaNCR25-RP	GGTGTCGTGGAAACAAAAAAACCCAAAAACAACGrCCCGT/3SpC3/	39	966
Upstream LDR	AcDx-7223-CaNCR25-Up	TATCTCCTAAAAGAAGCCGCACTAGTTTGGAGTTTTGGTTCGGGTTCTCrGGGCC/3SpC3/	54	967
Downstream LDR	AcDx-7224-CaNCR25-Dn	/5Phos/GGGTTTTAATATTTTTTCGTTGAGATCGCGGGGTGGGATTAAGGGCGATGGA	52	968
Real-Time Probe	AcDx-7225-CaNCR25-RT-Pb	/56-FAM/AAGGTTCTC/ZEN/GGGTTTTAATATTTTTTCGTTGAGATCGC/3IABkFQ/	38	969
Tag Forward Primer	AcDx-7226-CaNCR25-RT-FP	TATCTCCTAAAAGAAGCCGCAC	22	970
Tag Reverse Primer	AcDx-7227-CaNCR25-RT-RP	TCCATCGCCCTTAATCCCAC	20	971
Downstream PCR Primer	AcDx-7228-CaNCR25-PCR-V	TCCATCGCCCTTAATCCCACCAAAAAAACCCAAAAACAACGCCTGrCGATT/3SpC3/	50	972
HOXA10				
Forward PCR Primer	AcDx-7231-HOXA10-FP	GGTAAGATCGAGGCGCrGTTTG/3SpC3/	21	973
Reverse PCR Primer	AcDx-7232-HOXA10-RP	GGTGTCGTGGCGCTAAACGACAAACGCAArUAAAG/3SpC3/	34	974
Upstream LDR	AcDx-7233-HOXA10-Up	TCGCTCTTCAGCCTCCTACAGAGGGTTCGTAGTCGTGCGTCTCrGGGCC/3SpC3/	48	975
Downstream LDR	AcDx-7234-HOXA10-Dn	/5Phos/GGGATTTAGATTTTCGTTATCGTTATCGTTGTTCGGCTGTTCTGGGAATTATTGCCGGA	59	976
Real-Time Probe	AcDx-7235-HOXA10-RT-Pb	/56-FAM/AAGCGTCTC/ZEN/GGGATTTAGATTTTCGTTATCGTTATCG/3IABkFQ/	37	977
Tag Forward Primer	AcDx-7236-HOXA10-RT-FP	TCGCTCTTCAGCCTCCTACA	20	978
Tag Reverse Primer	AcDx-7237-HOXA10-RT-RP	TCCGGCAATAATTCCCAGAACA	22	979
Downstream PCR Primer	AcDx-7238-HOXA10-PCR-V	TCCGGCAATAATTCCCAGAACAACGCAATAAAACAACGTCGCTGrAACAG/3SpC3/	49	980
ABHD8				
Forward PCR Primer	AcDx-7241-ABHD8-FP	GACGGAAGCGGAGAGCrGGAAC/3SpC3/	21	981
Reverse PCR Primer	AcDx-7242-ABHD8-RP	GGTGTCGTGGGTTCGCTCCGATAAACGAAACrCAAAC/3SpC3/	36	982
Upstream LDR	AcDx-7243-ABHD8-Up	TTCAACGATCGCGCAGACAGTTCGTTGGGTTTTTTCGGATGTCACrGTTCT/3SpC3/	50	983

Downstream LDR	AcDx-7244-ABHD8-Dn	/5Phos/GTTTCGTTTAGGTAGTTTGGAGGCGTTTTTTTGTGTTCTGGGAATTATTGCCGGA	55	984
Real-Time Probe	AcDx-7245-ABHD8-RT-Pb	/56-FAM/TTATGTCAC/ZEN/GTTTCGTTTAGGTAGTTTGGAGG/3IABkFQ/	32	985
Tag Forward Primer	AcDx-7246-ABHD8-RT-FP	TTCAACGATCGCGCAGACA	19	986
Tag Reverse Primer	AcDx-7247-ABHD8-RT-RP	TCCGGCAATAATTCCCAGAACA	22	987
Downstream PCR Primer	AcDx-7248-ABHD8-PCR-V	TCCGGCAATAATTCCCAGAACACTCCGATAAACGAAACCAAATCAAAAAAATGrCCCCA/3SpC3/	58	988
HOXD8-S1				
Forward PCR Primer	AcDx-7251-HOXD8-S1-FP	TTTAGAGTCGAGGTTTGTAAATCrGAGGC/3SpC3/	28	989
Reverse PCR Primer	AcDx-7252-HOXD8-S1-RP	GGTGTCGTGGACGACCTACCCCGCTACrCTCCG/3SpC3/	32	990
Upstream LDR	AcDx-7253-HOXD8-S1-Up	TCACTATCGGCGTAGTCACCAGTTAGAGTGTTTTCGTGGGTCGGGCrGTACC/3SpC3/	51	991
Downstream LDR	AcDx-7254-HOXD8-S1-Dn	/5Phos/GTATTTTTTTTTTGTTCGGGTGCGTTTAGTTATTGGTGTGGTGACTTTACCCGGAGGA	58	992
Real-Time Probe	AcDx-7255-HOXD8-S1-RT-Pb	/56-FAM/TTGTCGGGC/ZEN/GTATTTTTTTTTTGTTCGGGTG/3IABkFQ/	31	993
Tag Forward Primer	AcDx-7256-HOXD8-S1-RT-FP	TCACTATCGGCGTAGTCACCA	21	994
Tag Reverse Primer	AcDx-7257-HOXD8-S1-RT-RP	TCCTCCGGGTAAAGTCACCA	20	995
Downstream PCR Primer	AcDx-7258-HOXD8-S1-PCR-V	TCCTCCGGGTAAAGTCACCAACTATTTCCTCTCAAACACCAATAACTAAATGrCACCT/3SpC3/	57	996
CaNCR26				
Forward PCR Primer	AcDx-7261-CaNCR26-FP	CGGGACGGGTTTTTTTGCrGGATC/3SpC3/	23	997
Reverse PCR Primer	AcDx-7262-CaNCR26-RP	GGTGTCGTGGACACCTAAAACAATAACAACCGCrCCGAT/3SpC3/	38	998
Upstream LDR	AcDx-7263-CaNCR26-Up	TCATCTGTTCGTCAGGGTCCAGATTTTTTGAAATGAAATAATGTGATGTACGTTGCrGATGG/3SpC3/	61	999
Downstream LDR	AcDx-7264-CaNCR26-Dn	/5Phos/GATAAGGGTCGGTTTGTAATGAGGTTTAGGTCGTGGTGACTTTACCCGGAGGA	53	1000
Real-Time Probe	AcDx-7265-CaNCR26-RT-	/56-FAM/AAACGTTGC/ZEN/GATAAGGGTCGGTTTGTAATGAGG/3IABkFQ/	33	1001

	Pb			
Tag Forward Primer	AcDx-7266-CaNCR26-RT-FP	TCATCTGTTCGTCAGGGTCCA	21	1002
Tag Reverse Primer	AcDx-7267-CaNCR26-RT-RP	TCCTCCGGGTAAAGTCACCA	20	1003
Downstream PCR Primer	AcDx-7268-CaNCR26-PCR-V	TCCTCCGGGTAAAGTCACCACTAAAACAATAACAACCGCCCGATGrACCTG/3SpC3/	50	1004
CaNCR27				
Forward PCR Primer	AcDx-7271-CaNCR27-FP	TAGTTTTTCGGTGGCGGCrGGCGC/3SpC3/	23	1005
Reverse PCR Primer	AcDx-7272-CaNCR27-RP	GGTGTCGTGGCGACCATAACCCCGACACrUAAAG/3SpC3/	33	1006
Upstream LDR	AcDx-7273-CaNCR27-Up	TTCGTACCTCGGCACACCAGGCGGCGTTAGTTGGTTTGCACrGAAGT/3SpC3/	46	1007
Downstream LDR	AcDx-7274-CaNCR27-Dn	/5Phos/GAAACGTTTTTTTTTTCGTTTCGTGTGTTTTATTTTTTATTTGGCTCCGTTACTCTGTCGA	61	1008
Real-Time Probe	AcDx-7275-CaNCR27-RT-Pb	/56-FAM/CCTTTGCAC/ZEN/GAAACGTTTTTTTTTTCGTTTCGTGTGTTTTA/3IABkFQ/	41	1009
Tag Forward Primer	AcDx-7276-CaNCR27-RT-FP	TTCGTACCTCGGCACACCA	19	1010
Tag Reverse Primer	AcDx-7277-CaNCR27-RT-RP	TCGACAGAGTAACGGAGCCA	20	1011
Downstream PCR Primer	AcDx-7278-CaNCR27-PCR-V	TCGACAGAGTAACGGAGCCAGACCATAACCCCGACACTAAAAAATGrUAAAG/3SpC3/	51	1012
USP2-S1				
Forward PCR Primer	AcDx-7281-USP2-S1-FP	TAAATGTAAACGTCGAGGGTACrGGGAT/3SpC3/	27	1013
Reverse PCR Primer	AcDx-7282-USP2-S1-RP	GGTGTCGTGGATCAAAATCTAAAACAAAAAACCGAACrUTTCT/3SpC3/	42	1014
Upstream LDR	AcDx-7283-USP2-S1-Up	TCTACAGCTAGATGCGGCCAGGTACGAGGCGATGTTGGTTTCACrGTTCG/3SpC3/	49	1015
Downstream LDR	AcDx-7284-USP2-S1-Dn	/5Phos/GTTTAGGCGATTTTTTTTGTCGGGTTATAGTTTAGGTGGCTCCGTTACTCTGTCGA	56	1016
Real-Time Probe	AcDx-7285-USP2-S1-RT-Pb	/56-FAM/AAGTTTCAC/ZEN/GTTTAGGCGATTTTTTTTGTCGGGT/3IABkFQ/	34	1017
Tag Forward Primer	AcDx-7286-USP2-S1-RT-FP	TCTACAGCTAGATGCGGCCA	20	1018

Tag Reverse Primer	AcDx-7287-USP2-S1-RT-RP	TCGACAGAGTAACGGAGCCA	20	1019
POU4F1				
Forward PCR Primer	AcDx-7291-POU4F1-FP	GTCGTTTCGAGGAGTTTTCrGCGAA/3SpC3/	24	1020
Reverse PCR Primer	AcDx-7292-POU4F1-RP	GGTGTCGTGGCACGAAACCGCCGAAArAAAAG/3SpC3/	31	1021
Upstream LDR	AcDx-7293-POU4F1-Up	TAGCGATAGTACCGACAGTCACTCGCGAGAGTTCGCGGTTTTGTCrGCGCC/3SpC3/	50	1022
Downstream LDR	AcDx-7294-POU4F1-Dn	/5Phos/GCGTTGATAGGTATTAGTTGTTTTCGTTTGTTTGACGGTGCGGAAACCTATCGTCGA	57	1023
Real-Time Probe	AcDx-7295-POU4F1-RT-Pb	/56-FAM/TTTTGTCGC/ZEN/GTTGATAGGTATTAGTTGTTTTCGTTTG/3IABkFQ/	37	1024
Tag Forward Primer	AcDx-7296-POU4F1-RT-FP	TAGCGATAGTACCGACAGTCAC	22	1025
Tag Reverse Primer	AcDx-7297-POU4F1-RT-RP	TCGACGATAGGTTTCCGCAC	20	1026
Downstream PCR Primer	AcDx-7298-POU4F1-PCR-V	TCGACGATAGGTTTCCGCACCGCCGAAAAAAAAAACGCGTGrUCAAG/3SpC3/	46	1027
FAM59B-S1				
Forward PCR Primer	AcDx-7301-FAM59B-S1-FP	TCGAGTTTTGGGCGGCrGGAGC/3SpC3/	21	1028
Reverse PCR Primer	AcDx-7302-FAM59B-S1-RP	GGTGTCGTGGCGCTCCCCCTCGTACTArACCTT/3SpC3/	32	1029
Upstream LDR	AcDx-7303-FAM59B-S1-Up	TCCAGGGTATTTGGCGCACGCGGAGTAGGATAGGGTGCrGGGCA/3SpC3/	43	1030
Downstream LDR	AcDx-7304-FAM59B-S1-Dn	/5Phos/GGGTGGGCGCGGTTTTCGGGTGCGGAAACCTATCGTCGA	39	1031
Real-Time Probe	AcDx-7305-FAM59B-S1-RT-Pb	/56-FAM/CCAGGGTGC/ZEN/GGGTGGGCG/3IABkFQ/	18	1032
Tag Forward Primer	AcDx-7306-FAM59B-S1-RT-FP	TCCAGGGTATTTGGCGCAC	19	1033
Tag Reverse Primer	AcDx-7307-FAM59B-S1-RT-RP	TCGACGATAGGTTTCCGCAC	20	1034
Downstream PCR Primer	AcDx-7308-FAM59B-S1-PCR-V	TCGACGATAGGTTTCCGCACTCGTACTAACCTCCCGAAAACTGrCGCCT/3SpC3/	48	1035
SHH				

Forward PCR Primer	AcDx-7311-SHH-FP	GATTCGGAGGATGGATTAGCrGTTGC/3SpC3/	25	1036
Reverse PCR Primer	AcDx-7312-SHH-RP	GGTGTCGTGGACGCCCCCTACGCAArAACCC/3SpC3/	30	1037
Upstream LDR	AcDx-7313-SHH-Up	TCCCTCGTCATCTCCCTTACCTGGGAGGAGGTTTTCGGAGATTCTCrGTTGA/3SpC3/	51	1038
Downstream LDR	AcDx-7314-SHH-Dn	/5Phos/GTTAGGAGGATTTCGCGGGTAGGGAGTCGGTCTTGGTGATGGAGCGA	47	1039
Real-Time Probe	AcDx-7315-SHH-RT-Pb	/56-FAM/TTGATTCTC/ZEN/GTTAGGAGGATTTCGCGGG/3IABkFQ/	28	1040
Tag Forward Primer	AcDx-7316-SHH-RT-FP	TCCCTCGTCATCTCCCTTACC	21	1041
Tag Reverse Primer	AcDx-7317-SHH-RT-RP	TCGCTCCATCACCAAGACC	19	1042
Downstream PCR Primer	AcDx-7318-SHH-PCR-V	TCGCTCCATCACCAAGACCCGCAAAACCTCCTCCCTGrACTCT/3SpC3/	42	1043
CYB5R2				
Forward PCR Primer	AcDx-7321-CYB5R2-FP	GAGGCGGGTGTTTGCrGGGAC/3SpC3/	20	1044
Reverse PCR Primer	AcDx-7322-CYB5R2-RP	GGTGTCGTGGCTCTCTCCACCCAACGAATArAATAG/3SpC3/	35	1045
Upstream LDR	AcDx-7323-CYB5R2-Up	TCATAATGTTGTCAGCCCGACCGTCGAAGTAGAGGCGTCATCrGAGCC/3SpC3/	47	1046
Downstream LDR	AcDx-7324-CYB5R2-Dn	/5Phos/GAGTTGAAGACGTGTATTTCGAGCGTTTTTGCGGGTCTTGGTGATGGAGCGA	52	1047
Real-Time Probe	AcDx-7325-CYB5R2-RT-Pb	/56-FAM/TTGGGCATC/ZEN/GTCGAGTTGAAGACGTGTATTTC/3IABkFQ/	32	1048
Tag Forward Primer	AcDx-7326-CYB5R2-RT-FP	TCATAATGTTGTCAGCCCGACC	22	1049
Tag Reverse Primer	AcDx-7327-CYB5R2-RT-RP	TCGCTCCATCACCAAGACC	19	1050
Downstream PCR Primer	AcDx-7328-CYB5R2-PCR-V	TCGCTCCATCACCAAGACCACCCAACGAATAAATAAACGCAAAAATGrCTCGG/3SpC3/	52	1051
TRH-S1				
Forward PCR Primer	AcDx-7331-TRH-S1-FP	GGGTCGGTTGTCGTTAGCrGTTTC/3SpC3/	23	1052
Reverse PCR Primer	AcDx-7332-TRH-S1-RP	GGTGTCGTGGCTTCAAATAAACCGCCGCTTArUATCC/3SpC3/	36	1053
Upstream LDR	AcDx-7333-TRH-S1-Up	TACGAATCACCCGAGAGTTCAAGTTAGCGTTTTTTTTCGGCGATCrGCGGC/3SpC3/	50	1054
Downstream LDR	AcDx-7334-TRH-S1-Dn	/5Phos/GCGATTTTTTTTCGTTGATTTTATTCGAGTCGTCGTTTGTTGTGGGTGGGTATA GGTCAGA	61	1055
Real-Time Probe	AcDx-7335-TRH-S1-RT-Pb	/56-FAM/AAGGCGATC/ZEN/GCGATTTTTTTTCGTTGATTTTATTCG/3IABkFQ/	36	1056
Tag Forward Primer	AcDx-7336-TRH-S1-RT-FP	TACGAATCACCCGAGAGTTCAA	22	1057
Tag Reverse Primer	AcDx-7337-TRH-S1-RT-RP	TCTGACCTATACCCACCCACAA	22	1058
Downstream PCR Primer	AcDx-7338-TRH-S1-PCR-V	TCTGACCTATACCCACCCACAACGCTTATATCTACGCCAAACGATGrACTCA/3SpC3/	51	1059

ALDH1A3				
Forward PCR Primer	AcDx-7341-ALDH1A3-FP	GCGATGATTTTTAGGGTGGACrGAGTT/3SpC3/	26	1060
Reverse PCR Primer	AcDx-7342-ALDH1A3-RP	GGTGTCGTGGCCAACTACAACTCCGCCATCrUACCC/3SpC3/	35	1061
Upstream LDR	AcDx-7343-ALDH1A3-Up	TACGAATCACCCGAGAGTTCAAGACGAGTTTGGTTTTGAGAGGATCrGGAAA/3SpC3/	51	1062
Downstream LDR	AcDx-7344-ALDH1A3-Dn	/5Phos/GGAGGGTTCGAGAGCGTTAAGAGAGAGTTCGTTGTGGGTGGGTATAGGTCAGA	53	1063
Real-Time Probe	AcDx-7345-ALDH1A3-RT-Pb	/56-FAM/TTGAGGATC/ZEN/GGAGGGTTCGAGAGCGTTAAG/3IABkFQ/	30	1064
Tag Forward Primer	AcDx-7346-ALDH1A3-RT-FP	TACGAATCACCCGAGAGTTCAA	22	1065
Tag Reverse Primer	AcDx-7347-ALDH1A3-RT-RP	TCTGACCTATACCCACCCACAA	22	1066
Downstream PCR Primer	AcDx-7348-ALDH1A3-PCR-V	TCTGACCTATACCCACCCACAACTCCGCCATCTACCTCCTGrAACTT/3SpC3/	46	1067
CaNCR28				
Forward PCR Primer	AcDx-7351-CaNCR28-FP	TTAGTTCGCGGAAGTTAGGTTCrGGGAA/3SpC3/	27	1068
Reverse PCR Primer	AcDx-7352-CaNCR28-RP	GGTGTCGTGGTTCTTTCCAAATAAACGCTTTACArGCTTC/3SpC3/	39	1069
Upstream LDR	AcDx-7353-CaNCR28-Up	TCTCATAAACACTCCGGCCACAGTTAGGTTCGGGAGACAGCrGGGAC/3SpC3/	46	1070
Downstream LDR	AcDx-7354-CaNCR28-Dn	/5Phos/GGAGTTTATTGTGTTTTCGGATTTATATTTTGTTCGAGGGTGGCTCAATAACGGGCAGA	59	1071
Real-Time Probe	AcDx-7355-CaNCR28-RT-Pb	/56-FAM/TTAGACAGC/ZEN/GGAGTTTATTGTGTTTTCGGA/3IABkFQ/	30	1072
Tag Forward Primer	AcDx-7356-CaNCR28-RT-FP	TCTCATAAACACTCCGGCCAC	21	1073
Tag Reverse Primer	AcDx-7357-CaNCR28-RT-RP	TCTGCCCGTTATTGAGCCAC	20	1074
Downstream PCR Primer	AcDx-7358-CaNCR28-PCR-V	TCTGCCCGTTATTGAGCCACGCTTTACAAAAATCCGACCCCTGrAACAG/3SpC3/	48	1075

GHSR				
Forward PCR Primer	AcDx-7361-GHSR-FP	GGAAGAGTTTGTAGAGGAGGTCrGTCGG/3SpC3/	27	1076
Reverse PCR Primer	AcDx-7362-GHSR-RP	GGTGTCGTGGAACATAACCTTCTCCGATCTACTCrATCTC/3SpC3/	39	1077
Upstream LDR	AcDx-7363-GHSR-Up	TTGATTGGGATCGTTCGCACGGAGGTCGTCGAAGTTTTAGGATCrGGTGC/3SpC3/	49	1078
Downstream LDR	AcDx-7364-GHSR-Dn	/5Phos/GGTATTGTTAGAGGCGAACGAGGTTTAGAGGTATGGTGGCTCAATAACGGGCAGA	55	1079
Real-Time Probe	AcDx-7365-GHSR-RT-Pb	/56-FAM/TTTAGGATC/ZEN/GGTATTGTTAGAGGCGAACGAGG/3IABkFQ/	32	1080
Tag Forward Primer	AcDx-7366-GHSR-RT-FP	TTGATTGGGATCGTTCGCAC	20	1081
Tag Reverse Primer	AcDx-7367-GHSR-RT-RP	TCTGCCCGTTATTGAGCCAC	20	1082
CDO1-S1				
Forward PCR Primer	AcDx-7371-CDO1-S1-FP	GTTTATTTTTTCGGGTTTTTTTTAAGCrGTTTA/3SpC3/	32	1083
Reverse PCR Primer	AcDx-7372-CDO1-S1-RP	GGTGTCGTGGGCAAAATCTCCCCGCCrUCCGT/3SpC3/	31	1084
Upstream LDR	AcDx-7373-CDO1-S1-Up	TCTGCCCAAAATACTGCACAAGTTTGGAGTTATTAGGAATGTATTAACGGTTCTCrGGAAA/3SpC3/	60	1085
Downstream LDR	AcDx-7374-CDO1-S1-Dn	/5Phos/GGAGGGAGGACGAGGCGGAGAGTTATTTAAGTTGAAACTGAGGCGGTGTTCA	52	1086
Real-Time Probe	AcDx-7375-CDO1-S1-RT-Pb	/56-FAM/TTGGTTCTC/ZEN/GGAGGGAGGACGAGGC/3IABkFQ/	25	1087
Tag Forward Primer	AcDx-7376-CDO1-S1-RT-FP	TCTGCCCAAAATACTGCACAA	21	1088
Tag Reverse Primer	AcDx-7377-CDO1-S1-RT-RP	TGAACACCGCCTCAGTTTCAA	21	1089
Downstream PCR Primer	AcDx-7378-CDO1-S1-PCR-V	TGAACACCGCCTCAGTTTCAACAAAATCTCCCCGCCTCTGrCCACT/3SpC3/	45	1090
RASAL1				
Forward PCR Primer	AcDx-7381-RASAL1-FP	TAGCGGGTGGGAGGCrGATTG/3SpC3/	20	1091
Reverse PCR Primer	AcDx-7382-RASAL1-RP	GGTGTCGTGGTAAAACCCCACCGCGATAArAAAAT/3SpC3/	34	1092
Upstream LDR	AcDx-7383-RASAL1-Up	TATGGACTGTACCAGCCCAAGGGAGGCGATTAGGGAGCrGGGCA/3SpC3/	43	1093
Downstream LDR	AcDx-7384-RASAL1-Dn	/5Phos/GGGTGGGAATAGAGGGACGTTTTTCGTTTGAAACTGAGGCGGTGTTCA	48	1094

Real-Time Probe	AcDx-7385-RASAL1-RT-Pb	/56-FAM/AAAGGGAGC/ZEN/GGGTGGGAATAGAGG/3IABkFQ/	24	1095
Tag Forward Primer	AcDx-7386-RASAL1-RT-FP	TATGGACTGTACCAGCCCAA	20	1096
Tag Reverse Primer	AcDx-7387-RASAL1-RT-RP	TGAACACCGCCTCAGTTTCAA	21	1097
Downstream PCR Primer	AcDx-7388-RASAL1-PCR-V	TGAACACCGCCTCAGTTTCAACCACCGCGATAAAAAACGAAAAATGrUCCCC/3SpC3/	51	1098
CaNCR29				
Forward PCR Primer	AcDx-7391-CaNCR29-FP	CGTTTTTGGGAGTGTATGTGCrGTTGC/3SpC3/	26	1099
Reverse PCR Primer	AcDx-7392-CaNCR29-RP	GGTGTCGTGGCGCCGCGAAACCGAArAACTG/3SpC3/	30	1100
Upstream LDR	AcDx-7393-CaNCR29-Up	TTTGCCTCTTGTAGGTGCCAGGAGTGTATGTGCGTTGTTGGGCrGTGAC/3SpC3/	48	1101
Downstream LDR	AcDx-7394-CaNCR29-Dn	/5Phos/GTGGTTTGAGTTCGGTTAGCGGTCGGTGGGCAACGCGGATATTCA	45	1102
Real-Time Probe	AcDx-7395-CaNCR29-RT-Pb	/56-FAM/AAGTTGGGC/ZEN/GTGGTTTGAGTTCGGTTAGC/3IABkFQ/	29	1103
Tag Forward Primer	AcDx-7396-CaNCR29-RT-FP	TTTGCCTCTTGTAGGTGCCA	20	1104
Tag Reverse Primer	AcDx-7397-CaNCR29-RT-RP	TGAATATCCGCGTTGCCCA	19	1105
Downstream PCR Primer	AcDx-7398-CaNCR29-PCR-V	TGAATATCCGCGTTGCCCAGAAAACTACCACCGACGCCTGrACCGT/3SpC3/	45	1106
GYPC				
Forward PCR Primer	AcDx-7401-GYPC-FP	TTCGGTTTGGTTAGTTTTCGCrGGTTC/3SpC3/	26	1107
Reverse PCR Primer	AcDx-7402-GYPC-RP	GGTGTCGTGGCGACGAATACTCACCGAAACrUAAAG/3SpC3/	35	1108
Upstream LDR	AcDx-7403-GYPC-Up	TGTCGCCCGGTAGCAATAAACCGCGGTTTTTGTTCGGGTTAACrGTTCG/3SpC3/	48	1109
Downstream LDR	AcDx-7404-GYPC-Dn	/5Phos/GTTTAGGAATGTGGTCGACGAGAAGTTTTAATAGTACGGTTTCCGCGATCTTTGCATTCA	60	1110
Real-Time Probe	AcDx-7405-GYPC-RT-Pb	/56-FAM/TTGGTTAAC/ZEN/GTTTAGGAATGTGGTCGACGAGA/3IABkFQ/	32	1111
Tag Forward Primer	AcDx-7406-GYPC-RT-FP	TGTCGCCCGGTAGCAATAAAC	21	1112
Tag Reverse Primer	AcDx-7407-GYPC-RT-RP	TGAATGCAAAGATCGCGGAAAC	22	1113
Downstream PCR Primer	AcDx-7408-GYPC-PCR-V	TGAATGCAAAGATCGCGGAAACACCGAAACTAAAAAACCACGCTGrUACTG/3SpC3/	50	1114

CaNCR30				
Forward PCR Primer	AcDx-7411-CaNCR30-FP	GCGGTGTATAAAATATACGTTGTTTCrGGGAG/3SpC3/	31	1115
Reverse PCR Primer	AcDx-7412-CaNCR30-RP	GGTGTCGTGGATTTAAAAACCCAAACGACCCTACrCTAAG/3SpC3/	39	1116
Upstream LDR	AcDx-7413-CaNCR30-Up	TGTGCACTAGTCCACGTGAAACCGGGAATTGCGAGAGGAACrGCGAC/3SpC3/	46	1117
Downstream LDR	AcDx-7414-CaNCR30-Dn	/5Phos/GCGGTGGGAGATATTCGAGGATAGAAGGGTTTCCGCGATCTTTGCATTCA	50	1118
Real-Time Probe	AcDx-7415-CaNCR30-RT-Pb	/56-FAM/TTGAGGAAC/ZEN/GCGGTGGGAGATATTCG/3IABkFQ/	26	1119
Tag Forward Primer	AcDx-7416-CaNCR30-RT-FP	TGTGCACTAGTCCACGTGAAAC	22	1120
Tag Reverse Primer	AcDx-7417-CaNCR30-RT-RP	TGAATGCAAAGATCGCGGAAAC	22	1121
Downstream PCR Primer	AcDx-7418-CaNCR30-PCR-V	TGAATGCAAAGATCGCGGAAACCCCAAACGACCCTACCTAAAATGrACAAG/3SpC3/	50	1122
DMRTA2-S1				
Forward PCR Primer	AcDx-7421-DMRTA2-S1-FP	TTACGTATGAGATCGTTAAAGGCrGTAGC/3SpC3/	28	1123
Reverse PCR Primer	AcDx-7422-DMRTA2-S1-RP	GGTGTCGTGGACGCACAACCGCGATCrUAACT/3SpC3/	31	1124
Upstream LDR	AcDx-7423-DMRTA2-S1-Up	TACAGATACGGACGGGAATCAAGTTTATGGGTGGGCGGAAATCrGAGTA/3SpC3/	48	1125
Downstream LDR	AcDx-7424-DMRTA2-S1-Dn	/5Phos/GAGCGTGGGTATTAAGTCGGTAGTGGAGTAGTTGTTTACATCCTCCTGCGTCA	53	1126
Real-Time Probe	AcDx-7425-DMRTA2-S1-RT-Pb	/56-FAM/AAGGAAATC/ZEN/GAGCGTGGGTATTAAGTCGGTAG/3IABkFQ/	32	1127
Tag Forward Primer	AcDx-7426-DMRTA2-S1-RT-FP	TACAGATACGGACGGGAATCAA	22	1128
Tag Reverse Primer	AcDx-7427-DMRTA2-S1-RT-RP	TGACGCAGGAGGATGTAAACAA	22	1129
Downstream PCR Primer	AcDx-7428-DMRTA2-S1-PCR-V	TGACGCAGGAGGATGTAAACAAACAACCGCGATCTAACCTTCATAATGrCCCTG/3SpC3/	53	1130

CPAMD8				
Forward PCR Primer	AcDx-7431-CPAMD8-FP	GTTTCGAAGGTTACGGGACrGAGGA/3SpC3/	24	1131
Reverse PCR Primer	AcDx-7432-CPAMD8-RP	GGTGTCGTGGTACGACTACTACGAACCCGATArAACCT/3SpC3/	37	1132
Upstream LDR	AcDx-7433-CPAMD8-Up	TTGGCGCAACGGTTTCCAAGCGGGTGATTAAGTGTAGGTGCrGACAA/3SpC3/	46	1133
Downstream LDR	AcDx-7434-CPAMD8-Dn	/5Phos/GACGGGTTAGGGATCGGGTCGGGTTGTTTACATCCTCCTGCGTCA	45	1134
Real-Time Probe	AcDx-7435-CPAMD8-RT-Pb	/56-FAM/AATAGGTGC/ZEN/GACGGGTTAGGGATCGG/3IABkFQ/	26	1135
Tag Forward Primer	AcDx-7436-CPAMD8-RT-FP	TTGGCGCAACGGTTTCCAA	19	1136
Tag Reverse Primer	AcDx-7437-CPAMD8-RT-RP	TGACGCAGGAGGATGTAAACAA	22	1137
Downstream PCR Primer	AcDx-7438-CPAMD8-PCR-V	TGACGCAGGAGGATGTAAACAACGCACCCCCGACCTGrACCCA/3SpC3/	42	1138
ZNF529				
Forward PCR Primer	AcDx-7441-ZNF529-FP	GGGTATTGGGAGTTTCGTTTCrGGGAA/3SpC3/	26	1139
Reverse PCR Primer	AcDx-7442-ZNF529-RP	GGTGTCGTGGCCTCAAAATTTCCAAACCCGACrUCGAG/3SpC3/	37	1140
Upstream LDR	AcDx-7443-ZNF529-Up	TTTGCCTCTTGTAGGTGCCATCGGGAGAGTTGTTTAAGGATGCrGGAGC/3SpC3/	48	1141
Downstream LDR	AcDx-7444-ZNF529-Dn	/5Phos/GGAATCGGGTGAGGAGGTTAGGGAGTGGGCAACGCGGATATTCA	44	1142
Real-Time Probe	AcDx-7445-ZNF529-RT-Pb	/56-FAM/CCAGGATGC/ZEN/GGAATCGGGTGAGG/3IABkFQ/	23	1143
Tag Forward Primer	AcDx-7446-ZNF529-RT-FP	TTTGCCTCTTGTAGGTGCCA	20	1144
Tag Reverse Primer	AcDx-7447-ZNF529-RT-RP	TGAATATCCGCGTTGCCCA	19	1145
Downstream PCR Primer	AcDx-7448-ZNF529-PCR-V	TGAATATCCGCGTTGCCCACCTCAAAATTTCCAAACCCGACTTGrAACTT/3SpC3/	49	1146
CaNCR32				
Forward PCR Primer	AcDx-7451-CaNCR32-FP	GAGGATTGTAGGGTGCGCrGGGTG/3SpC3/	23	1147
Reverse PCR Primer	AcDx-7452-CaNCR32-RP	GGTGTCGTGGACAAACCCCACGAATATAACAArAAACG/3SpC3/	37	1148
Upstream LDR	AcDx-7453-CaNCR32-Up	TATCGCATCAAATGGAGAGCAAGATTGTAGGGTGCGCGGGTATCTCrGGGCG/3SpC3/	51	1149
Downstream LDR	AcDx-7454-CaNCR32-Dn	/5Phos/GGGTAGTCGTTAGCGGAGGAAGCGGTTGACCGCTGTTATACGTTGCA	47	1150
Real-Time Probe	AcDx-7455-CaNCR32-RT-Pb	/56-FAM/TTGTATCTC/ZEN/GGGTAGTCGTTAGCGGAGGAA/3IABkFQ/	30	1151

Tag Forward Primer	AcDx-7456-CaNCR32-RT-FP	TATCGCATCAAATGGAGAGCAA	22	1152
Tag Reverse Primer	AcDx-7457-CaNCR32-RT-RP	TGCAACGTATAACAGCGGTCAA	22	1153
Downstream PCR Primer	AcDx-7458-CaNCR32-PCR-V	TGCAACGTATAACAGCGGTCAACAAAAACAACTTCCGAAAAAAACGAAACTGrCTTCT/3SpC3/	57	1154
CaNCR33				
Forward PCR Primer	AcDx-7461-CaNCR33-FP	TGAGTTTGTGAATACGGAGACrGGTCA/3SpC3/	26	1155
Reverse PCR Primer	AcDx-7462-CaNCR33-RP	GGTGTCGTGGCTTCATTTATTCCCGCGAATAACrAAACG/3SpC3/	38	1156
Upstream LDR	AcDx-7463-CaNCR33-Up	TCCTAGTACCTACAGTGGGCAAGAATACGGAGACGGTCGAGGACTCrGAGTC/3SpC3/	51	1157
Downstream LDR	AcDx-7464-CaNCR33-Dn	/5Phos/GAGATAAGGAGTTCGTTCGTTCGGTTTTTTTCGGTTGACCGCTGTTATACGTTGCA	56	1158
Real-Time Probe	AcDx-7465-CaNCR33-RT-Pb	/56-FAM/TTAGGACTC/ZEN/GAGATAAGGAGTTCGTTCGTTCG/3IABkFQ/	32	1159
Tag Forward Primer	AcDx-7466-CaNCR33-RT-FP	TCCTAGTACCTACAGTGGGCAA	22	1160
Tag Reverse Primer	AcDx-7467-CaNCR33-RT-RP	TGCAACGTATAACAGCGGTCAA	22	1161
Downstream PCR Primer	AcDx-7468-CaNCR33-PCR-V	TGCAACGTATAACAGCGGTCAACCGCGAATAACAAACACAACCTGrAAAAG/3SpC3/	50	1162
KRT7				
Forward PCR Primer	AcDx-7471-KRT7-FP	GGCGCGGAGTGTTTTCrGAGGC/3SpC3/	21	1163
Reverse PCR Primer	AcDx-7472-KRT7-RP	GGTGTCGTGGCGCGAAATAAATACCGAAAAACTAAArATAAG/3SpC3/	41	1164
Upstream LDR	AcDx-7473-KRT7-Up	TGTGCACTAGTCCACGTGAAACAGTGTTTTCGAGGTTAGCGAGCGCrGCGCC/3SpC3/	51	1165
Downstream LDR	AcDx-7474-KRT7-Dn	/5Phos/GCGTTTTTTTTCGTTCGTCGTTAGGTTTATTTCGGGTTTCCGCGATCTTTGCATTCA	57	1166
Real-Time Probe	AcDx-7475-KRT7-RT-Pb	/56-FAM/TTCGAGCGC/ZEN/GCGTTTTTTTTCGTTCGT/3IABkFQ/	27	1167
Tag Forward Primer	AcDx-7476-KRT7-RT-FP	TGTGCACTAGTCCACGTGAAAC	22	1168
Tag Reverse Primer	AcDx-7477-KRT7-RT-RP	TGAATGCAAAGATCGCGGAAAC	22	1169

Downstream PCR Primer	AcDx-7478-KRT7-PCR-V	TGAATGCAAAGATCGCGGAAACAATAAATAAACATAATAACTAAACCGAAATAAACCTAATGrACGAG/3SpC3/	67	1170
TRPS1				
Forward PCR Primer	AcDx-7481-TRPS1-FP	ATGTTTTATCGTTGTTAGGTATTTAATTATCrGGTTG/3SpC3/	36	1171
Reverse PCR Primer	AcDx-7482-TRPS1-RP	GGTGTCGTGGCCGTAAAAACTAAAAAAAAAACAAACTTCrCTCTG/3SpC3/	44	1172
Upstream LDR	AcDx-7483-TRPS1-Up	TCAGACGCACTAAACAGGCAATGTTAGGTATTTAATTATCGGTTAGTGTTTTTTGACTCrGCGTA/3SpC3/	64	1173
Downstream LDR	AcDx-7484-TRPS1-Dn	/5Phos/GCGCGATATATGGCGTATTAATCGTATCGTAGAGGTTGCGGATCGTCGTGTGAA	54	1174
Real-Time Probe	AcDx-7485-TRPS1-RT-Pb	/56-FAM/AATTGACTC/ZEN/GCGCGATATATGGCGTATTAATC/3IABkFQ/	32	1175
Tag Forward Primer	AcDx-7486-TRPS1-RT-FP	TCAGACGCACTAAACAGGCAA	21	1176
Tag Reverse Primer	AcDx-7487-TRPS1-RT-RP	TTCACACGACGATCCGCAA	19	1177
Downstream PCR Primer	AcDx-7488-TRPS1-PCR-V	TTCACACGACGATCCGCAACGTAAAAACTAAAAAAAAAACAAACTTCCTCTATGrATACA/3SpC3/	59	1178
CaNCR34				
Forward PCR Primer	AcDx-7491-CaNCR34-FP	GTAACGGAGTCGCGTTTTTCrGGGAC/3SpC3/	25	1179
Reverse PCR Primer	AcDx-7492-CaNCR34-RP	GGTGTCGTGGCGCATCCGACCCGAArACGCG/3SpC3/	30	1180
Upstream LDR	AcDx-7493-CaNCR34-Up	TTCTTCACAGTACCGCCACACGAGGTGGTTATCGGGTCGCrGTGAC/3SpC3/	45	1181
Downstream LDR	AcDx-7494-CaNCR34-Dn	/5Phos/GTGGTTTTCGGTCGGTTTTTTGTATTCGTTGTTTTTGTGTGTTGTCTGGTGGTGCA	56	1182
Real-Time Probe	AcDx-7495-CaNCR34-RT-Pb	/56-FAM/AAGGGTCGC/ZEN/GTGGTTTTCGGTCG/3IABkFQ/	23	1183
Tag Forward Primer	AcDx-7496-CaNCR34-RT-FP	TTCTTCACAGTACCGCCACA	20	1184
Tag Reverse Primer	AcDx-7497-CaNCR34-RT-RP	TGCACCACCAGACAACACA	19	1185
Downstream PCR Primer	AcDx-7498-CaNCR34-PCR-V	TGCACCACCAGACAACACAACCCGAAACGCACAAAAACAATGrAATAT/3SpC3/	47	1186

TLX2				
Forward PCR Primer	AcDx-7501-TLX2-FP	GTGGTAGTCGGGCGCrGTGAA/3SpC3/	20	1187
Reverse PCR Primer	AcDx-7502-TLX2-RP	GGTGTCGTGGCGCCTCTAAACTAACCGTACATrCTACA/3SpC3/	37	1188
Upstream LDR	AcDx-7503-TLX2-Up	TACCCTCCTAGCTCCGTACAGCGGTAGGTTTTGAGGTTGTCATCrGAGAA/3SpC3/	49	1189
Downstream LDR	AcDx-7504-TLX2-Dn	/5Phos/GAGGGTTTTTTTATTATCGGTCGGTTTTTAAGTTAGCGTGTGTTGTCTGGTGGTGCA	57	1190
Real-Time Probe	AcDx-7505-TLX2-RT-Pb	/56-FAM/AATGTCATC/ZEN/GAGGGTTTTTTTATTATCGGTCGGTT/3IABkFQ/	35	1191
Tag Forward Primer	AcDx-7506-TLX2-RT-FP	TACCCTCCTAGCTCCGTACA	20	1192
Tag Reverse Primer	AcDx-7507-TLX2-RT-RP	TGCACCACCAGACAACACA	19	1193
Downstream PCR Primer	AcDx-7508-TLX2-PCR-V	TGCACCACCAGACAACACACTAAACTAACCGTACATCTACGCAATGrCTAAT/3SpC3/	51	1194
CRHR2				
Forward PCR Primer	AcDx-7511-CRHR2-FP	CGGTGGGTTGGAGAGCrGTGGA/3SpC3/	21	1195
Reverse PCR Primer	AcDx-7512-CRHR2-RP	GGTGTCGTGGCTCCTCGAACAAACGAACAAArUAAAT/3SpC3/	36	1196
Upstream LDR	AcDx-7513-CRHR2-Up	TGAACGCTCAAACACGTGAACTGAGGTGACGGAATGTTTTGCGCrGGGAG/3SpC3/	49	1197
Downstream LDR	AcDx-7514-CRHR2-Dn	/5Phos/GAGGATCGTAGGTTTTCGAGTTGTAGAGGGGTTGGCCTGTAAGCGTTCCA	50	1198
Real-Time Probe	AcDx-7515-CRHR2-RT-Pb	/56-FAM/CCTTTGCGC/ZEN/GAGGATCGTAGGTTTTCGAG/3IABkFQ/	29	1199
Tag Forward Primer	AcDx-7516-CRHR2-RT-FP	TGAACGCTCAAACACGTGAAC	21	1200
Tag Reverse Primer	AcDx-7517-CRHR2-RT-RP	TGGAACGCTTACAGGCCAAC	20	1201
Downstream PCR Primer	AcDx-7518-CRHR2-PCR-V	TGGAACGCTTACAGGCCAACCTCGAACAAACGAACAAATAAACGTGrCTTTG/3SpC3/	51	1202
IGF2AS				
Forward PCR Primer	AcDx-7521-IGF2AS-FP	CGAACGTTTTGTGGTAGGCrGGTGA/3SpC3/	24	1203
Reverse PCR Primer	AcDx-7522-IGF2AS-RP	GGTGTCGTGGCCCGAATCTTCCAACGAACrUAAAT/3SpC3/	34	1204
Upstream LDR	AcDx-7523-IGF2AS-Up	TCCTAGTACCTACAGTGGGCAAGTGGACGTTGTTGAAGGTGGGCrGAGGC/3SpC3/	49	1205
Downstream LDR	AcDx-7524-IGF2AS-Dn	/5Phos/GAGATTTCGAGGTTAGTTTTTAGTGTTCGGGAGTAACTTGACCGCTGTTATACGTTGCA	59	1206
Real-Time Probe	AcDx-7525-IGF2AS-RT-Pb	/56-FAM/TTGGTGGGC/ZEN/GAGATTTCGAGGTTAGTTTTTAG/3IABkFQ/	32	1207
Tag Forward Primer	AcDx-7526-IGF2AS-RT-FP	TCCTAGTACCTACAGTGGGCAA	22	1208

Tag Reverse Primer	AcDx-7527-IGF2AS-RT-RP	TGCAACGTATAACAGCGGTCAA	22	1209
Downstream PCR Primer	AcDx-7528-IGF2AS-PCR-V	TGCAACGTATAACAGCGGTCAATCCAACGAACTAAACGTTACTCCTGrAACAT/3SpC3/	52	1210
MYO15B-S1				
Forward PCR Primer	AcDx-7531-MYO15B-S1-FP	ATACGTCGGGTGAGGACrGGAGA/3SpC3/	22	1211
Reverse PCR Primer	AcDx-7532-MYO15B-S1-RP	GGTGTCGTGGCCGAACTATACCGCGCTArACTAT/3SpC3/	33	1212
Upstream LDR	AcDx-7533-MYO15B-S1-Up	TCGCACCGGAATTCTGACCGTGAGGACGGAGGTAGTTTTTGCTCrGGACG/3SpC3/	49	1213
Downstream LDR	AcDx-7534-MYO15B-S1-Dn	/5Phos/GGATAGCGAAATTCGCGAGGTTTAGGAGAGCGGTAGTTTCCCATGACGGCA	51	1214
Real-Time Probe	AcDx-7535-MYO15B-S1-RT-Pb	/56-FAM/AATTTGCTC/ZEN/GGATAGCGAAATTCGCGAGG/3IABkFQ/	29	1215
Tag Forward Primer	AcDx-7536-MYO15B-S1-RT-FP	TCGCACCGGAATTCTGACC	19	1216
Tag Reverse Primer	AcDx-7537-MYO15B-S1-RT-RP	TGCCGTCATGGGAAACTACC	20	1217
Downstream PCR Primer	AcDx-7538-MYO15B-S1-PCR-V	TGCCGTCATGGGAAACTACCCGAACTATACCGCGCTAACTACTGrCTCTT/3SpC3/	49	1218
EVX1-S1				
Forward PCR Primer	AcDx-7541-EVX1-S1-FP	GTTTTTTATTAGATAAGAGGTCGTCrGGTTA/3SpC3/	30	1219
Reverse PCR Primer	AcDx-7542-EVX1-S1-RP	GGTGTCGTGGCCTAAAAACGAAAAACGAAAAACGAArAAACA/3SpC3/	41	1220
Upstream LDR	AcDx-7543-EVX1-S1-Up	TTGCACGTTGTCCTGCACCCGTCGGTTGGTTGTCGGTTTTATAACrGTTTA/3SpC3/	50	1221
Downstream LDR	AcDx-7544-EVX1-S1-Dn	/5Phos/GTTCGTGGAGTTATTTTTCGGTTTCGGGATTTAGGGTAGTTTCCCATGACGGCA	54	1222
Real-Time Probe	AcDx-7545-EVX1-S1-RT-Pb	/56-FAM/AATTATAAC/ZEN/GTTCGTGGAGTTATTTTTCGGTTTCG/3IABkFQ/	35	1223
Tag Forward Primer	AcDx-7546-EVX1-S1-RT-FP	TTGCACGTTGTCCTGCACC	19	1224
Tag Reverse Primer	AcDx-7547-EVX1-S1-RT-RP	TGCCGTCATGGGAAACTACC	20	1225
Downstream PCR Primer	AcDx-7548-EVX1-S1-PCR-V	TGCCGTCATGGGAAACTACCCTAAAAACGAAAAACGAAAAACGAAAAATGrAAACC/3SpC3/	55	1226

EFCAB4A				
Forward PCR Primer	AcDx-7551-EFCAB4A-FP	TACGAGGAGACGGAGTAGTTTCrGGGAA/3SpC3/	26	1227
Reverse PCR Primer	AcDx-7552-EFCAB4A-RP	GGTGTCGTGGCCAACGAAATAAAATTAAACCTAACGArAAAAG/3SpC3/	42	1228
Upstream LDR	AcDx-7553-EFCAB4A-Up	TGCTTACCCACGATGCACCACGGAGTAGTTTCGGGAGTAGAATCrGGCAC/3SpC3/	49	1229
Downstream LDR	AcDx-7554-EFCAB4A-Dn	/5Phos/GGCGTTCGTCGAGTTAGGTGGGTTTCGGTCGTATGACTTGCTCGCA	46	1230
Real-Time Probe	AcDx-7555-EFCAB4A-RT-Pb	/56-FAM/TTTAGAATC/ZEN/GGCGTTCGTCGAGTTAGGTGG/3IABkFQ/	30	1231
Tag Forward Primer	AcDx-7556-EFCAB4A-RT-FP	TGCTTACCCACGATGCACC	19	1232
Tag Reverse Primer	AcDx-7557-EFCAB4A-RT-RP	TGCGAGCAAGTCATACGACC	20	1233
Downstream PCR Primer	AcDx-7558-EFCAB4A-PCR-V	TGCGAGCAAGTCATACGACCAAAATTAAACCTAACGAAAAAAACGAAACCTGrAAACT/3SpC3/	57	1234
GRASP				
Forward PCR Primer	AcDx-7561-GRASP-FP	TTGGGTGTTTCGATTTTTCGCrGTTTC/3SpC3/	26	1235
Reverse PCR Primer	AcDx-7562-GRASP-RP	GGTGTCGTGGAAAAACGTACATAAAACGCGAArATTAG/3SpC3/	37	1236
Upstream LDR	AcDx-7563-GRASP-Up	TAGCCGATGGCGTAAAACCGCGTTTTTCGTTGTTGCGAAGATCrGTGGC/3SpC3/	48	1237
Downstream LDR	AcDx-7564-GRASP-Dn	/5Phos/GTGGTTTTCGTTTGTATATCGCGTTTAGGTTCGGGTCGTATGACTTGCTCGCA	53	1238
Real-Time Probe	AcDx-7565-GRASP-RT-Pb	/56-FAM/TTGAAGATC/ZEN/GTGGTTTTCGTTTGTATATCGCGTTT/3IABkFQ/	35	1239
Tag Forward Primer	AcDx-7566-GRASP-RT-FP	TAGCCGATGGCGTAAAACC	19	1240
Tag Reverse Primer	AcDx-7567-GRASP-RT-RP	TGCGAGCAAGTCATACGACC	20	1241
Downstream PCR Primer	AcDx-7568-GRASP-PCR-V	TGCGAGCAAGTCATACGACCGTACATAAAACGCGAAATTAAAAACCACTGrAACCC/3SpC3/	55	1242
SND1				
Forward PCR Primer	AcDx-7571-SND1-FP	CGAGGTTAGGGCGAGCrGGGCA/3SpC3/	21	1243
Reverse PCR Primer	AcDx-7572-SND1-RP	GGTGTCGTGGCCTCCGACCGACCCAArAATAG/3SpC3/	31	1244
Upstream LDR	AcDx-7573-SND1-Up	TACGAATCACCCGAGAGTTCAACGAGCGAGTCGGAGTGGGCrGTTGG/3SpC3/	46	1245
Downstream LDR	AcDx-7574-SND1-Dn	/5Phos/GTTAAGTGAGAGGTCGCGCGTAAGTCGTTGTGGGTGGGTATAGGTCAGA	49	1246

Real-Time Probe	AcDx-7575-SND1-RT-Pb	/56-FAM/TTAGTGGGC/ZEN/GTTAAGTGAGAGGTCGC/3IABkFQ/	26	1247
Tag Forward Primer	AcDx-7576-SND1-RT-FP	TACGAATCACCCGAGAGTTCAA	22	1248
Tag Reverse Primer	AcDx-7577-SND1-RT-RP	TCTGACCTATACCCACCCACAA	22	1249
Downstream PCR Primer	AcDx-7578-SND1-PCR-V	TCTGACCTATACCCACCCACAAGACCCAAAATAACTACGAACGCCTATGrACTTG/3SpC3/	54	1250
SEPT9-S2				
Forward PCR Primer	AcDx-7581-SEPT9-S2-FP	CGTCGTTTTTTGGGCGCrGGGTC/3SpC3/	22	1251
Reverse PCR Primer	AcDx-7582-SEPT9-S2-RP	GGTGTCGTGGCTAAAACCCGAATAACCGCTAArAAAAT/3SpC3/	37	1252
Upstream LDR	AcDx-7583-SEPT9-S2-Up	TTCGCCTACCGCAGTGAACGGGCGCGGGTTAAGCrGGGAG/3SpC3/	39	1253
Downstream LDR	AcDx-7584-SEPT9-S2-Dn	/5Phos/GGAGAGGAGGGAGGCGTTTCGTTGAGACATGGGCTCGCA	39	1254
Real-Time Probe	AcDx-7585-SEPT9-S2-RT-Pb	/56-FAM/TTGTTAAGC/ZEN/GGAGAGGAGGGAGG/3IABkFQ/	23	1255
Tag Forward Primer	AcDx-7586-SEPT9-S2-RT-FP	TTCGCCTACCGCAGTGAAC	19	1256
Tag Reverse Primer	AcDx-7587-SEPT9-S2-RT-RP	TGCGAGCCCATGTCTCAAC	19	1257
Downstream PCR Primer	AcDx-7588-SEPT9-S2-PCR-V	TGCGAGCCCATGTCTCAACAATCCTAAACACACGACCGAAATGrCCCCT/3SpC3/	48	1258
CaNCR36				
Forward PCR Primer	AcDx-7591-CaNCR36-FP	GCGAATTAGGTCGTTGGCrGGAGA/3SpC3/	23	1259
Reverse PCR Primer	AcDx-7592-CaNCR36-RP	GGTGTCGTGGCACCTACAAAACGAACAACGACrCGATC/3SpC3/	37	1260
Upstream LDR	AcDx-7593-CaNCR36-Up	TTGCATTTCGTTAGCGACACATTTAGGAGGAGCGTAGAAGCrGACAA/3SpC3/	46	1261
Downstream LDR	AcDx-7594-CaNCR36-Dn	/5Phos/GACGGTTTTTATGTCGGCGGTTAATATGATGGAGAATGTGAGTCGATCTACCCGCA	56	1262
Real-Time Probe	AcDx-7595-CaNCR36-RT-Pb	/56-FAM/TTTAGAAGC/ZEN/GACGGTTTTTATGTCGGCGGTTA/3IABkFQ/	32	1263
Tag Forward Primer	AcDx-7596-CaNCR36-RT-FP	TTGCATTTCGTTAGCGACACA	21	1264
Tag Reverse Primer	AcDx-7597-CaNCR36-RT-	TGCGGGTAGATCGACTCACA	20	1265

	RP			
Downstream PCR Primer	AcDx-7598-CaNCR36-PCR-V	TGCGGGTAGATCGACTCACACACCTACAAAACGAACAACGACTGrATTCC/3SpC3/	49	1266
HOXC4-S1				
Forward PCR Primer	AcDx-7601-HOXC4-S1-FP	GAATAAAGCGATTCGGTTTTTTATTCrGGGAG/3SpC3/	31	1267
Reverse PCR Primer	AcDx-7602-HOXC4-S1-RP	GGTGTCGTGGCCTCCTACTAACTAACGACCCTArUAAAG/3SpC3/	38	1268
Upstream LDR	AcDx-7603-HOXC4-S1-Up	TTGCATTTCGTTAGCGACACAGTTTTTTATTCGAGGATTGGGTTGTCTCrGTGCA/3SpC3/	54	1269
Downstream LDR	AcDx-7604-HOXC4-S1-Dn	/5Phos/GTGTGATTGGTCGGAGGAGTTATATGGTGAAAGTGTGAGTCGATCTACCCGCA	53	1270
Real-Time Probe	AcDx-7605-HOXC4-S1-RT-Pb	/56-FAM/TTTTGTCTC/ZEN/GTGTGATTGGTCGGAGGAGTTATAT/3IABkFQ/	34	1271
Tag Forward Primer	AcDx-7606-HOXC4-S1-RT-FP	TTGCATTTCGTTAGCGACACA	21	1272
Tag Reverse Primer	AcDx-7607-HOXC4-S1-RT-RP	TGCGGGTAGATCGACTCACA	20	1273
LOC100128239				
Forward PCR Primer	AcDx-7611-LOC100128239-FP	GAGGAAGGGAAAGGTGGTACrGGATC/3SpC3/	25	1274
Reverse PCR Primer	AcDx-7612-LOC100128239-RP	GGTGTCGTGGCTTAACGATAAAAATAAAAACGAAAACGCrGCGAT/3SpC3/	44	1275
Upstream LDR	AcDx-7613-LOC100128239-Up	TACCACTCATCTTCTGCGACAGTGGTACGGATTTTAAGGGAGGGACTCrGGAAC/3SpC3/	53	1276
Downstream LDR	AcDx-7614-LOC100128239-Dn	/5Phos/GGAGTTAATAGATTTGATTTTTTCGAGTTTGTAGTTTTTAGTCGTGTGAGTCGATCTACCCGCA	64	1277
Real-Time Probe	AcDx-7615-LOC100128239-RT-Pb	/56-FAM/TTGGGACTC/ZEN/GGAGTTAATAGATTTGATTTTTTCGAGTTTG/3IABkFQ/	40	1278
Tag Forward Primer	AcDx-7616-LOC100128239-RT-FP	TACCACTCATCTTCTGCGACA	21	1279
Tag Reverse Primer	AcDx-7617-	TGCGGGTAGATCGACTCACA	20	1280

	LOC100128239-RT-RP			
Downstream PCR Primer	AcDx-7618-LOC100128239-PCR-V	TGCGGGTAGATCGACTCACACGATAAAAATAAAAACGAAAACGCGTGrACTAG/3SpC3/	52	1281
RBP1				
Forward PCR Primer	AcDx-7621-RBP1-FP	TGGGAGGTTTAATTACGGGCrGAGAG/3SpC3/	25	1282
Reverse PCR Primer	AcDx-7622-RBP1-RP	GGTGTCGTGGCCCAAAATACTAAAATTACAAACGTAAArCCTCT/3SpC3/	43	1283
Upstream LDR	AcDx-7623-RBP1-Up	TCTCATGGGCGCTAGTATCAACGGGCGAGAAAATTGAGGTATTTTTCACrGTTCC/3SpC3/	54	1284
Downstream LDR	AcDx-7624-RBP1-Dn	/5Phos/GTTTTTTTCGAAAATAAAAGAAAGGATGTCGGGTACGGGTTTCCCTGATTGATACCCGCA	60	1285
Real-Time Probe	AcDx-7625-RBP1-RT-Pb	/56-FAM/CCTTTTCAC/ZEN/GTTTTTTTCGAAAATAAAAGAAAGGATGTCGG/3IABkFQ/	41	1286
Tag Forward Primer	AcDx-7626-RBP1-RT-FP	TCTCATGGGCGCTAGTATCAAC	22	1287
Tag Reverse Primer	AcDx-7627-RBP1-RT-RP	TGCGGGTATCAATCAGGGAAAC	22	1288
Downstream PCR Primer	AcDx-7628-RBP1-PCR-V	TGCGGGTATCAATCAGGGAAACAATACTAAAATTACAAACGTAAACCGCTGrUACCT/3SpC3/	56	1289
CLDN10				
Forward PCR Primer	AcDx-7631-CLDN10-FP	TGGAGAATTCGGAGGGCrGATCA/3SpC3/	22	1290
Reverse PCR Primer	AcDx-7632-CLDN10-RP	GGTGTCGTGGCCTCCGCCCACCGTCrCCCGT/3SpC3/	30	1291
Upstream LDR	AcDx-7633-CLDN10-Up	TGGATCGAGACGGAATGCAACTGTGAGAGGTCGCGCGCrGTTCC/3SpC3/	43	1292
Downstream LDR	AcDx-7634-CLDN10-Dn	/5Phos/GTTTTTTGGCGTCGGGTTCGTTGGGGTTTCCCTGATTGATACCCGCA	47	1293
Real-Time Probe	AcDx-7635-CLDN10-RT-Pb	/56-FAM/AACGCGCGC/ZEN/GTTTTTTGGCGTC/3IABkFQ/	22	1294
Tag Forward Primer	AcDx-7636-CLDN10-RT-FP	TGGATCGAGACGGAATGCAAC	21	1295
Tag Reverse Primer	AcDx-7637-CLDN10-RT-RP	TGCGGGTATCAATCAGGGAAAC	22	1296
Downstream PCR Primer	AcDx-7638-CLDN10-PCR-V	TGCGGGTATCAATCAGGGAAACCCACCCCGCCCAATGrAACCT/3SpC3/	42	1297
GNB4				
Forward PCR Primer	AcDx-7641-GNB4-FP	GTTTTGTTGGGTAGTTTCGAATATTCrGTTAC/3SpC3/	31	1298

Reverse PCR Primer	AcDx-7642-GNB4-RP	GGTGTCGTGGCCCCCCTAATTTCTCGTCACrUCCCT/3SpC3/	35	1299
Upstream LDR	AcDx-7643-GNB4-Up	TCCTGAGGGACAAATACACACCAGTTTCGAATATTCGTTATTTTAGGGATACrGTATA/3SpC3/	57	1300
Downstream LDR	AcDx-7644-GNB4-Dn	/5Phos/GTACGGGTTCGTGTTTTGAGTTTTTGGAAGGGGTAGGTAAGGAAGTCACGCA	52	1301
Real-Time Probe	AcDx-7645-GNB4-RT-Pb	/56-FAM/AAGGGATAC/ZEN/GTACGGGTTCGTGTTTTGAGTTT/3IABkFQ/	32	1302
Tag Forward Primer	AcDx-7646-GNB4-RT-FP	TCCTGAGGGACAAATACACACC	22	1303
Tag Reverse Primer	AcDx-7647-GNB4-RT-RP	TGCGTGACTTCCTTACCTACC	21	1304
Downstream PCR Primer	AcDx-7648-GNB4-PCR-V	TGCGTGACTTCCTTACCTACCCCCTAATTTCTCGTCACTCCCTGrAAACT/3SpC3/	49	1305
CaNCR37				
Forward PCR Primer	AcDx-7651-CaNCR37-FP	GTAGACGTTTTTTTTTTAGGAGGTCrGGAAC/3SpC3/	30	1306
Reverse PCR Primer	AcDx-7652-CaNCR37-RP	GGTGTCGTGGCTCACCGACCCTCGCArACGAC/3SpC3/	31	1307
Upstream LDR	AcDx-7653-CaNCR37-Up	TTGCAACAGGCTACCGACCCGGAATTTTTGTTTTTATTTTTTATCGGGATTCrGGTCA/3SpC3/	57	1308
Downstream LDR	AcDx-7654-CaNCR37-Dn	/5Phos/GGTTGGTTAGAGGTAAGTTTCGAGATTTTTTATTAATTATTATTATCGGGTAGGTAAGGAAGTCACGCA	69	1309
Real-Time Probe	AcDx-7655-CaNCR37-RT-Pb	/56-FAM/AAGGGATTC/ZEN/GGTTGGTTAGAGGTAAGTTTCGAGAT/3IABkFQ/	35	1310
Tag Forward Primer	AcDx-7656-CaNCR37-RT-FP	TTGCAACAGGCTACCGACC	19	1311
Tag Reverse Primer	AcDx-7657-CaNCR37-RT-RP	TGCGTGACTTCCTTACCTACC	21	1312
Downstream PCR Primer	AcDx-7658-CaNCR37-PCR-V	TGCGTGACTTCCTTACCTACCCTCACCGACCCTCGCAATGrATAAC/3SpC3/	45	1313
ZMIZ1-S1				
Forward PCR Primer	AcDx-7661-ZMIZ1-S1-FP	GGTTTTTCGTTCGAGGAATTTCrGGGAA/3SpC3/	27	1314
Reverse PCR Primer	AcDx-7662-ZMIZ1-S1-RP	GGTGTCGTGGAACTAAACATCCAAATTAAATCTCGArUTTAG/3SpC3/	41	1315
Upstream LDR	AcDx-7663-ZMIZ1-S1-Up	TAAGCCTGCTTTTCCGAAACAAGGTTAGGGAAGTAAGATGTCGGATCrGTTCG/3SpC3/	52	1316
Downstream LDR	AcDx-7664-ZMIZ1-S1-Dn	/5Phos/TTTAAAAAATTTTCGAAACGAACTACGAAATAAAAAAAATAAACTTGTATTGCGCCAGGATAGCA	65	1317

Real-Time Probe	AcDx-7665-ZMIZ1-S1-RT-Pb	/56-FAM/AATCGGATC/ZEN/GTTTATTTTTTTTATTTCGTAGTTCGTTTCGAA/3IABkFQ/	42	1318
Tag Forward Primer	AcDx-7666-ZMIZ1-S1-RT-FP	TAAGCCTGCTTTTCCGAAACAA	22	1319
Tag Reverse Primer	AcDx-7667-ZMIZ1-S1-RT-RP	TGCTATCCTGGCGCAATACAA	21	1320
KCNA3-S1				
Forward PCR Primer	AcDx-7671-KCNA3-S1-FP	AGGTTAGGTCGTTTTTTTTCGCrGTTTC/3SpC3/	27	1321
Reverse PCR Primer	AcDx-7672-KCNA3-S1-RP	GGTGTCGTGGCGCGCCAAACCCAAACrAAAAT/3SpC3/	31	1322
Upstream LDR	AcDx-7673-KCNA3-S1-Up	TTGGCAACTCTCCACCCAATTTTTTCGCGTTTTTCGTTTTTTCATCrGTTCC/3SpC3/	51	1323
Downstream LDR	AcDx-7674-KCNA3-S1-Dn	/5Phos/GTTTTCGTTTTCGAGTCGAGTTTATCGTTTGTTGTAGTTGTATTGCGCCAGGATAGCA	58	1324
Real-Time Probe	AcDx-7675-KCNA3-S1-RT-Pb	/56-FAM/AATTTCATC/ZEN/GTTTTCGTTTTCGAGTCGAGTTTATCG/3IABkFQ/	36	1325
Tag Forward Primer	AcDx-7676-KCNA3-S1-RT-FP	TTGGCAACTCTCCACCCAA	19	1326
Tag Reverse Primer	AcDx-7677-KCNA3-S1-RT-RP	TGCTATCCTGGCGCAATACAA	21	1327
Downstream PCR Primer	AcDx-7678-KCNA3-S1-PCR-V	TGCTATCCTGGCGCAATACAACCAAACCCAAACAAAACATCGTGrACTTC/3SpC3/	49	1328
ESR1				
Forward PCR Primer	AcDx-7681-ESR1-FP	GAGTTGGCGGAGGGCrGTTCA/3SpC3/	20	1329
Reverse PCR Primer	AcDx-7682-ESR1-RP	GGTGTCGTGGGCGAACTCTAACCCCGACrCCTAT/3SpC3/	33	1330
Upstream LDR	AcDx-7683-ESR1-Up	TCGACGAATCTGCTCAGACAAGGGATTGTATTTGTTTTCGTCGGATCrGTTTA/3SpC3/	52	1331
Downstream LDR	AcDx-7684-ESR1-Dn	/5Phos/GTTCGGTTTTATCGGATTCGTAGGTTTTCGAGGTTGAAGCAGCGTCTGAGCA	52	1332
Real-Time Probe	AcDx-7685-ESR1-RT-Pb	/56-FAM/TTTCGGATC/ZEN/GTTCGGTTTTATCGGATTCGTAGG/3IABkFQ/	33	1333
Tag Forward Primer	AcDx-7686-ESR1-RT-FP	TCGACGAATCTGCTCAGACAA	21	1334
Tag Reverse Primer	AcDx-7687-ESR1-RT-RP	TGCTCAGACGCTGCTTCAA	19	1335
Downstream PCR Primer	AcDx-7688-ESR1-PCR-V	TGCTCAGACGCTGCTTCAAAACCCCGACCCTACCCTGrAAAAT/3SpC3/	42	1336

A1BG				
Forward PCR Primer	AcDx-7691-A1BG-FP	GGGTATTTAGGAGCGGTAGCrGGTAC/3SpC3/	25	1337
Reverse PCR Primer	AcDx-7692-A1BG-RP	GGTGTCGTGGATCCGCACCCCCGAArACCGT/3SpC3/	30	1338
Upstream LDR	AcDx-7693-A1BG-Up	TCGACGAATCTGCTCAGACAAGTAGCGGTATTTGTAGTTGTCAGCrGTGCC/3SpC3/	50	1339
Downstream LDR	AcDx-7694-A1BG-Dn	/5Phos/GTGTTGGAGTTTTACGAAGATTAGTTCGAGGTTCTTGAAGCAGCGTCTGAGCA	53	1340
Real-Time Probe	AcDx-7695-A1BG-RT-Pb	/56-FAM/AATGTCAGC/ZEN/GTGTTGGAGTTTTACGAAGATTAGTTC/3IABkFQ/	36	1341
Tag Forward Primer	AcDx-7696-A1BG-RT-FP	TCGACGAATCTGCTCAGACAA	21	1342
Tag Reverse Primer	AcDx-7697-A1BG-RT-RP	TGCTCAGACGCTGCTTCAA	19	1343
Downstream PCR Primer	AcDx-7698-A1BG-PCR-V	TGCTCAGACGCTGCTTCAACCCCGAAACCGCGATGrAACCC/3SpC3/	40	1344
BEND4				
Forward PCR Primer	AcDx-7701-BEND4-FP	GCGGAGAGTTGGTATTTGCrGTTGG/3SpC3/	24	1345
Reverse PCR Primer	AcDx-7702-BEND4-RP	GGTGTCGTGGGTTCGCCGCTATCGTCAArATATG/3SpC3/	33	1346
Upstream LDR	AcDx-7703-BEND4-Up	TTTCCGCCGCTACAACCAAGAGTTTTAGGTTGGCGTTGTCGTTACTCrGTGCT/3SpC3/	52	1347
Downstream LDR	AcDx-7704-BEND4-Dn	/5Phos/GTGTCGTCGGTGTCGGCGGTCTTGAAGCAGCGTCTGAGCA	40	1348
Real-Time Probe	AcDx-7705-BEND4-RT-Pb	/56-FAM/AAGTTACTC/ZEN/GTGTCGTCGGTGTCGGC/3IABkFQ/	26	1349
Tag Forward Primer	AcDx-7706-BEND4-RT-FP	TTTCCGCCGCTACAACCAA	19	1350
Tag Reverse Primer	AcDx-7707-BEND4-RT-RP	TGCTCAGACGCTGCTTCAA	19	1351
Downstream PCR Primer	AcDx-7708-BEND4-PCR-V	TGCTCAGACGCTGCTTCAATCAAATATAACCCAAACGCGACGATGrACCGT/3SpC3/	50	1352
SARM1				
Forward PCR Primer	AcDx-7711-SARM1-FP	CGTCGAGCGGTTGGCrGGTGC/3SpC3/	20	1353
Reverse PCR Primer	AcDx-7712-SARM1-RP	GGTGTCGTGGCGACGACACTTTCGCGAArCCCGG/3SpC3/	33	1354
Upstream LDR	AcDx-7713-SARM1-Up	TCAGTGAAAACACATCCACCCAGGTTAGATGGAGGCGGTGGCACrGGGCC/3SpC3/	49	1355
Downstream LDR	AcDx-7714-SARM1-Dn	/5Phos/GGGTTTATGGTGGGTTGCGGGTGGTGGTGAGCAGGGATGAGCA	43	1356
Real-Time Probe	AcDx-7715-SARM1-RT-Pb	/56-FAM/AAGTGGCAC/ZEN/GGGTTTATGGTGGGTTG/3IABkFQ/	26	1357
Tag Forward Primer	AcDx-7716-SARM1-RT-FP	TCAGTGAAAACACATCCACCCA	22	1358
Tag Reverse Primer	AcDx-7717-SARM1-RT-RP	TGCTCATCCCTGCTCACCA	19	1359

Downstream PCR Primer	AcDx-7718-SARM1-PCR-V	TGCTCATCCCTGCTCACCAGCGAACCCGAACCACCTGrCAACT/3SpC3/	42	1360
CR1L				
Forward PCR Primer	AcDx-7721-CR1L-FP	GGTTTCGTTTCGCGGTTCrGTCGG/3SpC3/	23	1361
Reverse PCR Primer	AcDx-7722-CR1L-RP	GGTGTCGTGGCACAATTTCCAAAACCGCATAArCAAAC/3SpC3/	37	1362
Upstream LDR	AcDx-7723-CR1L-Up	TCCGGCCTTTGACGATACCGGTTCGTCGATTGGTTTTAGTCGTCTCrGGCAC/3SpC3/	51	1363
Downstream LDR	AcDx-7724-CR1L-Dn	/5Phos/GGCGTTAAGATTTAATTTTATATTACGCGTTTAGGTTACGTTTATTTGGGTAAT TCACTCGAACGGAGCA	70	1364
Real-Time Probe	AcDx-7725-CR1L-RT-Pb	/56-FAM/TTTCGTCTC/ZEN/GGCGTTAAGATTTAATTTTATATTACGCGTTTAGG/3IABkFQ/	44	1365
Tag Forward Primer	AcDx-7726-CR1L-RT-FP	TCCGGCCTTTGACGATACC	19	1366
Tag Reverse Primer	AcDx-7727-CR1L-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	1367
Downstream PCR Primer	AcDx-7728-CR1L-PCR-V	TGCTCCGTTCGAGTGAATTACCACAATTTCCAAAACCGCATAACAAATAAATGrUAACT/ 3SpC3/	58	1368
ANKLE1				
Forward PCR Primer	AcDx-7731-ANKLE1-FP	GTTGTTGCGTTGCGGCrGCGGG/3SpC3/	21	1369
Reverse PCR Primer	AcDx-7732-ANKLE1-RP	GGTGTCGTGGACCCCGAAACAACGCAArACCGT/3SpC3/	32	1370
Upstream LDR	AcDx-7733-ANKLE1-Up	TTGCACCCGCGACATAACCGGATTTTAATTTGGTGTTAGAGGACAGCrGTAAT/3SpC3/	52	1371
Downstream LDR	AcDx-7734-ANKLE1-Dn	/5Phos/GTAGCGGTTGTGTATTTGGCGGTCGGGGTAATTCACTCGAACGGAGCA	48	1372
Real-Time Probe	AcDx-7735-ANKLE1-RT-Pb	/56-FAM/TTGGACAGC/ZEN/GTAGCGGTTGTGTATTTGGC/3IABkFQ/	29	1373
Tag Forward Primer	AcDx-7736-ANKLE1-RT-FP	TTGCACCCGCGACATAACC	19	1374
Tag Reverse Primer	AcDx-7737-ANKLE1-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	1375
Downstream PCR Primer	AcDx-7738-ANKLE1-PCR-V	TGCTCCGTTCGAGTGAATTACCCGCGCGAATACCGAACTCTGrACCGT/3SpC3/	47	1376
WNT6-S1				
Forward PCR Primer	AcDx-7741-WNT6-S1-FP	TTTGTTGTTCGTCGTTCGTACrGTTTG/3SpC3/	26	1377
Reverse PCR Primer	AcDx-7742-WNT6-S1-RP	GGTGTCGTGGCCGAAAAAACCGATACGTCGArUTAAG/3SpC3/	36	1378
Upstream LDR	AcDx-7743-WNT6-S1-Up	TCCGGCCTTTGACGATACCCGTTTAAGTCGTCGGGTCGGGCrGGACC/3SpC3/	46	1379

Downstream LDR	AcDx-7744-WNT6-S1-Dn	/5Phos/GGATTTTTTTTACGTCGTCGATTCGTTCGATTTTTGCGGGTAATTCACTCGAACGGAGCA	60	1380
Real-Time Probe	AcDx-7745-WNT6-S1-RT-Pb	/56-FAM/TTGTCGGGC/ZEN/GGATTTTTTTTACGTCGTCGATTC/3IABkFQ/	33	1381
Tag Forward Primer	AcDx-7746-WNT6-S1-RT-FP	TCCGGCCTTTGACGATACC	19	1382
Tag Reverse Primer	AcDx-7747-WNT6-S1-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	1383
Downstream PCR Primer	AcDx-7748-WNT6-S1-PCR-V	TGCTCCGTTCGAGTGAATTACCCGAAAAAACCGATACGTCGATTAAAAATGrCAAAG/3SpC3/	56	1384
SGIP1				
Forward PCR Primer	AcDx-7751-SGIP1-FP	GTTTTTGATAATTAATTTCGGGTATTTAGTCrGTTTC/3SpC3/	36	1385
Reverse PCR Primer	AcDx-7752-SGIP1-RP	GGTGTCGTGGCATCGCCTCCCGCTTATCrACCAG/3SpC3/	33	1386
Upstream LDR	AcDx-7753-SGIP1-Up	TCCGGCCTTTGACGATACCGTATTTAGTCGTTTTTGTAAGTTTAAGGAGACAACrGAGAG/3SpC3/	59	1387
Downstream LDR	AcDx-7754-SGIP1-Dn	/5Phos/GAGGAGGAGCGGAGGAAGTGGTAATTCACTCGAACGGAGCA	41	1388
Real-Time Probe	AcDx-7755-SGIP1-RT-Pb	/56-FAM/TTAGACAAC/ZEN/GAGGAGGAGCGGG/3IABkFQ/	22	1389
Tag Forward Primer	AcDx-7756-SGIP1-RT-FP	TCCGGCCTTTGACGATACC	19	1390
Tag Reverse Primer	AcDx-7757-SGIP1-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	1391
Downstream PCR Primer	AcDx-7758-SGIP1-PCR-V	TGCTCCGTTCGAGTGAATTACCCAACCCCTTCCCCACCTAATAATGrCACTC/3SpC3/	51	1392
CaNCR38				
Forward PCR Primer	AcDx-7761-CaNCR38-FP	CGGATCGTAGGTTGGGCrGGTTC/3SpC3/	22	1393
Reverse PCR Primer	AcDx-7762-CaNCR38-RP	GGTGTCGTGGCCGCGACCTAAAAACGCrUCCCT/3SpC3/	32	1394
Upstream LDR	AcDx-7763-CaNCR38-Up	TCGCGGAAAGTCCCAGTAACGTTGGGCGGTTTTTGATTTTTTGCrGTTCC/3SpC3/	49	1395
Downstream LDR	AcDx-7764-CaNCR38-Dn	/5Phos/GTTTTTCGTTTATAGTCGGAGTTCGGTAGTTGGAAGTGTTGGCCTGTAAGCGTTCCA	57	1396
Real-Time Probe	AcDx-7765-CaNCR38-RT-Pb	/56-FAM/CCTTTTTGC/ZEN/GTTTTTCGTTTATAGTCGGAGTTCGGTAGTTG/3IABkFQ/	41	1397
Tag Forward Primer	AcDx-7766-CaNCR38-RT-	TCGCGGAAAGTCCCAGTAAC	20	1398

	FP			
Tag Reverse Primer	AcDx-7767-CaNCR38-RT-RP	TGGAACGCTTACAGGCCAAC	20	1399
Downstream PCR Primer	AcDx-7768-CaNCR38-PCR-V	TGGAACGCTTACAGGCCAACCCTAAAAACGCTCCCCGAAATAATGrCAACC/3SpC3/	50	1400
TBX1-S1				
Forward PCR Primer	AcDx-7771-TBX1-S1-FP	TTTCGTTTCGGTTTTGTATAGTTTCrGAAGC/3SpC3/	30	1401
Reverse PCR Primer	AcDx-7772-TBX1-S1-RP	GGTGTCGTGGAAAAAAAAACCGCGATAACCCrCTAAG/3SpC3/	36	1402
Upstream LDR	AcDx-7773-TBX1-S1-Up	TGAACGCTCAAACACGTGAACTGTATAGTTTCGAAGTTCGTCGGGCTCrGGTGC/3SpC3/	53	1403
Downstream LDR	AcDx-7774-TBX1-S1-Dn	/5Phos/GGTTATTTTGTTTTAAGGGTAAGTAAGGAATACGTTTTTTTAGTTTTAGGTTGGCCTGTAAGCGTTCCA	69	1404
Real-Time Probe	AcDx-7775-TBX1-S1-RT-Pb	/56-FAM/AACGGGCTC/ZEN/GGTTATTTTGTTTTAAGGGTAAGTAAGGA/3IABkFQ/	38	1405
Tag Forward Primer	AcDx-7776-TBX1-S1-RT-FP	TGAACGCTCAAACACGTGAAC	21	1406
Tag Reverse Primer	AcDx-7777-TBX1-S1-RT-RP	TGGAACGCTTACAGGCCAAC	20	1407
Alternate Solid Tumor Markers				
CaNCR39				
Forward PCR Primer	AcDx-7781-CaNCR39-FP	GATAAGAGGATGATTTTAAAGGGACrGTAGC/3SpC3/	30	1408
Reverse PCR Primer	AcDx-7782-CaNCR39-RP	GGTGTCGTGGCAACGCCATCGCGTAACrCAACC/3SpC3/	32	1409
Upstream LDR	AcDx-7783-CaNCR39-Up	TACACGTGGATATCTCCGACCGGGACGTAGTGGTTTCGTTTATCATCrGTTCC/3SpC3/	52	1410
Downstream LDR	AcDx-7784-CaNCR39-Dn	/5Phos/GTTTTTATAGGGAGGGAGTCGCGGTTTCGGGTGCTAGTCACACAGTTCCA	50	1411
Real-Time Probe	AcDx-7785-CaNCR39-RT-Pb	/56-FAM/AATATCATC/ZEN/GTTTTTATAGGGAGGGAGTCGCGG/3IABkFQ/	33	1412
Tag Forward Primer	AcDx-7786-CaNCR39-RT-FP	TACACGTGGATATCTCCGACC	21	1413
Tag Reverse Primer	AcDx-7787-CaNCR39-RT-RP	TGGAACTGTGTGACTAGCACC	21	1414

Downstream PCR Primer	AcDx-7788-CaNCR39-PCR-V	TGGAACTGTGTGACTAGCACCCGCCATCGCGTAACCAACTTGrAAACT/3SpC3/	47	1415
PTGER4				
Forward PCR Primer	AcDx-7791-PTGER4-FP	GTTTATTTTCGAGGTTAATTCGTTCrGTTTC/3SpC3/	30	1416
Reverse PCR Primer	AcDx-7792-PTGER4-RP	GGTGTCGTGGTTACCCACCACCCCGAArAATAG/3SpC3/	32	1417
Upstream LDR	AcDx-7793-PTGER4-Up	TAGCATTCGAGAACGCACCGAGGTTAATTCGTTCGTTTTTTTGAGTCTCrGATTA/3SpC3/	54	1418
Downstream LDR	AcDx-7794-PTGER4-Dn	/5Phos/GATCGGTTGAATAGTTTAGTGATTATTTCGGCGGTGGGTGCTAGTCACACAGTTCCA	57	1419
Real-Time Probe	AcDx-7795-PTGER4-RT-Pb	/56-FAM/AAGAGTCTC/ZEN/GATCGGTTGAATAGTTTAGTGATTATTTCGG/3IABkFQ/	40	1420
Tag Forward Primer	AcDx-7796-PTGER4-RT-FP	TAGCATTCGAGAACGCACC	19	1421
Tag Reverse Primer	AcDx-7797-PTGER4-RT-RP	TGGAACTGTGTGACTAGCACC	21	1422
Downstream PCR Primer	AcDx-7798-PTGER4-PCR-V	TGGAACTGTGTGACTAGCACCCCACCCCGAAAATAAACATCACTGrCCGAG/3SpC3/	50	1423
CaNCR40				
Forward PCR Primer	AcDx-7801-CaNCR40-FP	GTTTGTGTAGGGCGAGGACrGGGAT/3SpC3/	24	1424
Reverse PCR Primer	AcDx-7802-CaNCR40-RP	GGTGTCGTGGACGAATCTTCTACATCCGACArACAAC/3SpC3/	36	1425
Upstream LDR	AcDx-7803-CaNCR40-Up	TAACCGGGCCTAAAGTGACACGAGGCGGTTTTGCGCTCrGGTCG/3SpC3/	43	1426
Downstream LDR	AcDx-7804-CaNCR40-Dn	/5Phos/GGTTAGGTTGGTTTTCGAGGATTTAGTCGTTTTTAATTTTTTATGTTACGTGATCTCCCTCTCCA	65	1427
Real-Time Probe	AcDx-7805-CaNCR40-RT-Pb	/56-FAM/AATGCGCTC/ZEN/GGTTAGGTTGGTTTTCGAG/3IABkFQ/	28	1428
Tag Forward Primer	AcDx-7806-CaNCR40-RT-FP	TAACCGGGCCTAAAGTGACA	20	1429
Tag Reverse Primer	AcDx-7807-CaNCR40-RT-RP	TGGAGAGGGAGATCACGTAACA	22	1430
Downstream PCR Primer	AcDx-7808-CaNCR40-PCR-V	TGGAGAGGGAGATCACGTAACATCTACATCCGACAACAATTTAAAAAATTAAAAATGrACTAG/3SpC3/	62	1431

SHF				
Forward PCR Primer	AcDx-7811-SHF-FP	GGGTTTCGATTCGAATAAGGCrGGTGC/3SpC3/	26	1432
Reverse PCR Primer	AcDx-7812-SHF-RP	GGTGTCGTGGTCCGAAAAATCGTCCGACTCrCGCCA/3SpC3/	35	1433
Upstream LDR	AcDx-7813-SHF-Up	TCGATGGTCAATGAGCTTCACAGTGGAGGAGAGTCGTAGCrGCGAA/3SpC3/	45	1434
Downstream LDR	AcDx-7814-SHF-Dn	/5Phos/GCGGGAGCGGCGTGGTGTTACGTGATCTCCCTCTCCA	37	1435
Real-Time Probe	AcDx-7815-SHF-RT-Pb	/56-FAM/TTTCGTAGC/ZEN/GCGGGAGCGG/3IABkFQ/	19	1436
Tag Forward Primer	AcDx-7816-SHF-RT-FP	TCGATGGTCAATGAGCTTCACA	22	1437
Tag Reverse Primer	AcDx-7817-SHF-RT-RP	TGGAGAGGGAGATCACGTAACA	22	1438
Downstream PCR Primer	AcDx-7818-SHF-PCR-V	TGGAGAGGGAGATCACGTAACACACCCCTATCGCCCCTGrCACCT/3SpC3/	44	1439
PON3-S1				
Forward PCR Primer	AcDx-7821-PON3-S1-FP	GTCGTAGTAGGGCGTTGACrGAGTC/3SpC3/	24	1440
Reverse PCR Primer	AcDx-7822-PON3-S1-RP	GGTGTCGTGGTCTCCGTTAAACCTTAACCTCTArCCCAG/3SpC3/	38	1441
Upstream LDR	AcDx-7823-PON3-S1-Up	TCGATGGTCAATGAGCTTCACAGCGTTGACGAGTTTCGTCGAGTCTCrGTCAC/3SpC3/	52	1442
Downstream LDR	AcDx-7824-PON3-S1-Dn	/5Phos/GTCGTTCGGGTTTAAGGTCGTTTTTACGTTTACGTGTTACGTGATCTCCCTCTCCA	56	1443
Real-Time Probe	AcDx-7825-PON3-S1-RT-Pb	/56-FAM/AAGAGTCTC/ZEN/GTCGTTCGGGTTTAAGGTCG/3IABkFQ/	29	1444
Tag Forward Primer	AcDx-7826-PON3-S1-RT-FP	TCGATGGTCAATGAGCTTCACA	22	1445
Tag Reverse Primer	AcDx-7827-PON3-S1-RT-RP	TGGAGAGGGAGATCACGTAACA	22	1446
Downstream PCR Primer	AcDx-7828-PON3-S1-PCR-V	TGGAGAGGGAGATCACGTAACACCTCTACCCAAAAAACAAAAAATCGTAAATGrUAAAG/3SpC3/	58	1447
PAX2-S1				
Forward PCR Primer	AcDx-7831-PAX2-S1-FP	TTTGGAGGGTGCGTAGTCrGGGCA/3SpC3/	23	1448
Reverse PCR Primer	AcDx-7832-PAX2-S1-RP	GGTGTCGTGGAAACCTTAACTAAACAATAAAACCAATTACrCCCGG/3SpC3/	45	1449
Upstream LDR	AcDx-7833-PAX2-S1-Up	TCTCGGGACCACAATACGAACGTGCGTAGTCGGGCGTTATTAGGTCrGGGCC/3SpC3/	51	1450
Downstream LDR	AcDx-7834-PAX2-S1-Dn	/5Phos/GGGTTTTTTTTATGTTTCGTGAACGTAATTATTTTTCGAGGGTTACGCTAAGCTGGTGCCA	61	1451

Real-Time Probe	AcDx-7835-PAX2-S1-RT-Pb	/56-FAM/CCTTAGGTC/ZEN/GGGTTTTTTTTATGTTTCGTGAACGTA/3IABkFQ/	36	1452
Tag Forward Primer	AcDx-7836-PAX2-S1-RT-FP	TCTCGGGACCACAATACGAAC	21	1453
Tag Reverse Primer	AcDx-7837-PAX2-S1-RT-RP	TGGCACCAGCTTAGCGTAAC	20	1454
Downstream PCR Primer	AcDx-7838-PAX2-S1-PCR-V	TGGCACCAGCTTAGCGTAACCCTTAACTAAACAATAAAACCAATTACCCTGrAAAAG/3SpC3/	56	1455
LRRFIP1				
Forward PCR Primer	AcDx-7841-LRRFIP1-FP	TAGTTTGGACGGTGTGGATTTCrGGGTC/3SpC3/	27	1456
Reverse PCR Primer	AcDx-7842-LRRFIP1-RP	GGTGTCGTGGCGATACGACGACCCGCrAAAAG/3SpC3/	31	1457
Upstream LDR	AcDx-7843-LRRFIP1-Up	TTGCTGTGCGCGGTAGAACTGTGGATTTCGGGTTTTGCAACrGTTGA/3SpC3/	46	1458
Downstream LDR	AcDx-7844-LRRFIP1-Dn	/5Phos/GTTAGATCGGTTTTAGCGGTTCGTATTCGTATTTTGTAAGGTTACGCTAAGCTGGTGCCA	60	1459
Real-Time Probe	AcDx-7845-LRRFIP1-RT-Pb	/56-FAM/AATTGCAAC/ZEN/GTTAGATCGGTTTTAGCGGTTCG/3IABkFQ/	32	1460
Tag Forward Primer	AcDx-7846-LRRFIP1-RT-FP	TTGCTGTGCGCGGTAGAAC	19	1461
Tag Reverse Primer	AcDx-7847-LRRFIP1-RT-RP	TGGCACCAGCTTAGCGTAAC	20	1462
Downstream PCR Primer	AcDx-7848-LRRFIP1-PCR-V	TGGCACCAGCTTAGCGTAACCCGCAAAAAAAAACGAACTTACAAAATATGrAATAT/3SpC3/	55	1463
ITPKA-S1				
Forward PCR Primer	AcDx-7851-ITPKA-S1-FP	AAGTTTTATAGAGTAGGAATATTTTTCGTCrGTTAA/3SpC3/	35	1464
Reverse PCR Primer	AcDx-7852-ITPKA-S1-RP	GGTGTCGTGGAAACGATCCTAAATCCGAAACTArCTCTG/3SpC3/	38	1465
Upstream LDR	AcDx-7853-ITPKA-S1-Up	TCTCGATTACGCTCCGCACTTCGTCGTTAGGTGTTGGGCGCrGTCAA/3SpC3/	46	1466
Downstream LDR	AcDx-7854-ITPKA-S1-Dn	/5Phos/GTCGGTTCGGTTATTAGTTTGTCGTTTTTTTTTTTTTTTCGGTGTGTAGCTTAGACATGGCCA	63	1467
Real-Time Probe	AcDx-7855-ITPKA-S1-RT-Pb	/56-FAM/AATGGGCGC/ZEN/GTCGGTTCGGTTATTAG/3IABkFQ/	26	1468
Tag Forward Primer	AcDx-7856-ITPKA-S1-RT-FP	TCTCGATTACGCTCCGCAC	19	1469
Tag Reverse Primer	AcDx-7857-ITPKA-S1-RT-RP	TGGCCATGTCTAAGCTACACAC	22	1470
Downstream PCR Primer	AcDx-7858-ITPKA-S1-PCR-	TGGCCATGTCTAAGCTACACACCCTAAATCCGAAACTACTCTAACCACTGrAAAAG/3Sp	55	1471

	V	C3/		
CaNCR41				
Forward PCR Primer	AcDx-7861-CaNCR41-FP	GGATATGGTGCGGTGGCrGGTAA/3SpC3/	22	1472
Reverse PCR Primer	AcDx-7862-CaNCR41-RP	GGTGTCGTGGCGCCTTAACCGCGAACTCrCCTCT/3SpC3/	33	1473
Upstream LDR	AcDx-7863-CaNCR41-Up	TCACAGAGACTTGCCGATCACGCGGGCGGTTGGATTTTAAATCrGGCAC/3SpC3/	48	1474
Downstream LDR	AcDx-7864-CaNCR41-Dn	/5Phos/GGCGTTGCGTTTTATATGACGGTTCGCGGTGTGTAGCTTAGACATGGCCA	50	1475
Real-Time Probe	AcDx-7865-CaNCR41-RT-Pb	/56-FAM/AATTAAATC/ZEN/GGCGTTGCGTTTTATATGACGGTTC/3IABkFQ/	34	1476
Tag Forward Primer	AcDx-7866-CaNCR41-RT-FP	TCACAGAGACTTGCCGATCAC	21	1477
Tag Reverse Primer	AcDx-7867-CaNCR41-RT-RP	TGGCCATGTCTAAGCTACACAC	22	1478
Downstream PCR Primer	AcDx-7868-CaNCR41-PCR-V	TGGCCATGTCTAAGCTACACACCTCCCTCCCCGAACCTGrCGAAT/3SpC3/	44	1479
ASCL2				
Forward PCR Primer	AcDx-7871-ASCL2-FP	AGGTTTAGGTTTTCGAGGCrGTTTC/3SpC3/	24	1480
Reverse PCR Primer	AcDx-7872-ASCL2-RP	GGTGTCGTGGCCCAAAACCCTCAAACCGArAAACA/3SpC3/	34	1481
Upstream LDR	AcDx-7873-ASCL2-Up	TCACAGAGACTTGCCGATCACGGGCGTTTTTTTTAATTCGTTTCGTTTTTCTCrGTTCC/3SpC3/	58	1482
Downstream LDR	AcDx-7874-ASCL2-Dn	/5Phos/GTTTTTTTACGCGTATTTTGTTTGTGGTTTTCGTGCGGTGTGTAGCTTAGACATGGCCA	59	1483
Real-Time Probe	AcDx-7875-ASCL2-RT-Pb	/56-FAM/AATTTTCTC/ZEN/GTTTTTTTACGCGTATTTTGTTTGTGGTTTTC/3IABkFQ/	41	1484
Tag Forward Primer	AcDx-7876-ASCL2-RT-FP	TCACAGAGACTTGCCGATCAC	21	1485
Tag Reverse Primer	AcDx-7877-ASCL2-RT-RP	TGGCCATGTCTAAGCTACACAC	22	1486
Downstream PCR Primer	AcDx-7878-ASCL2-PCR-V	TGGCCATGTCTAAGCTACACACCCCAAAACCCTCAAACCGAAAATGrCACTG/3SpC3/	51	1487
CaNCR42				
Forward PCR Primer	AcDx-7881-CaNCR42-FP	GTCGTTATTCGGTCGTGATTATAATCrGAGGC/3SpC3/	31	1488

Reverse PCR Primer	AcDx-7882-CaNCR42-RP	GGTGTCGTGGACTTTCCTCTACGACTCAAATAAArAATTG/3SpC3/	39	1489
Upstream LDR	AcDx-7883-CaNCR42-Up	TTCGTCCCTGCACGCTAACTTTTGTTTGTAGAGTAATATATTAGGTTTAAATTTATCGTTC TCrGGTGA/3SpC3/	68	1490
Downstream LDR	AcDx-7884-CaNCR42-Dn	/5Phos/GGTAGTCGTTTTACGCGGGTTTTTTTTCGGTAGGTTCCATCACCGTTAGGCCA	53	1491
Real-Time Probe	AcDx-7885-CaNCR42-RT-Pb	/56-FAM/CCCGTTCTC/ZEN/GGTAGTCGTTTTACGCGGG/3IABkFQ/	28	1492
Tag Forward Primer	AcDx-7886-CaNCR42-RT-FP	TTCGTCCCTGCACGCTAAC	19	1493
Tag Reverse Primer	AcDx-7887-CaNCR42-RT-RP	TGGCCTAACGGTGATGGAAC	20	1494
Downstream PCR Primer	AcDx-7888-CaNCR42-PCR-V	TGGCCTAACGGTGATGGAACCCTCTACGACTCAAATAAAAATTAAAAAATCTACTGrAA AG/3SpC3/	60	1495
HLF				
Forward PCR Primer	AcDx-7891-HLF-FP	GATTTAGTTTGAGGTTATAAAGGTTTTTACrGTTTC/3SpC3/	35	1496
Reverse PCR Primer	AcDx-7892-HLF-RP	GGTGTCGTGGCCTTACTAAAACCCTAATCTTCGArUCTAG/3SpC3/	39	1497
Upstream LDR	AcDx-7893-HLF-Up	TGATGCTGGCAAACCCTAGAACGAGGTTATAAAGGTTTTTACGTTTTATTGAAAGACrG GTCC/3SpC3/	62	1498
Downstream LDR	AcDx-7894-HLF-Dn	/5Phos/GGTTTTTTTTTATTTGTGTTTATTCGTTTTTGTAGTTTTTTTTTTTAGGTTCCATCAC CGTTAGGCCA	67	1499
Real-Time Probe	AcDx-7895-HLF-RT-Pb	/56-FAM/AAGAAAGAC/ZEN/GGTTTTTTTTTATTTGTGTTTATTCGTTTTTGTAG/3IABkFQ/	44	1500
Tag Forward Primer	AcDx-7896-HLF-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	1501
Tag Reverse Primer	AcDx-7897-HLF-RT-RP	TGGCCTAACGGTGATGGAAC	20	1502
CCDC151-S2				
Forward PCR Primer	AcDx-7901-CCDC151-S2-FP	GTCGCGTTTTTTAGTTTTATAGGATTCrGTTTA/3SpC3/	32	1503
Reverse PCR Primer	AcDx-7902-CCDC151-S2-RP	GGTGTCGTGGAACTAATCAACCAAAAAAAAATCTCGAArAACAG/3SpC3/	43	1504
Upstream LDR	AcDx-7903-CCDC151-S2-Up	TGATGCTGGCAAACCCTAGAACGTTTGTATTTTTTTTGGTTTTTTTTTTCGTGGCTCrGGG CC/3SpC3/	62	1505

Downstream LDR	AcDx-7904-CCDC151-S2-Dn	/5Phos/GGGTTTATTTTTGGTTCGTTAAATTTCGTTCGTTGGTTCCATCACCGTTAGGCCA	55	1506
Real-Time Probe	AcDx-7905-CCDC151-S2-RT-Pb	/56-FAM/AAGTGGCTC/ZEN/GGGTTTATTTTTGGTTCGTTAAATTTCG/3IABkFQ/	37	1507
Tag Forward Primer	AcDx-7906-CCDC151-S2-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	1508
Tag Reverse Primer	AcDx-7907-CCDC151-S2-RT-RP	TGGCCTAACGGTGATGGAAC	20	1509
Downstream PCR Primer	AcDx-7908-CCDC151-S2-PCR-V	TGGCCTAACGGTGATGGAACACTAATCAACCAAAAAAAAATCTCGAAAACAATGrAACGG/3SpC3/	59	1510
CHRD				
Forward PCR Primer	AcDx-7911-CHRD-FP	GGTTCGTGGTGACGGTTATACrGGGTG/3SpC3/	26	1511
Reverse PCR Primer	AcDx-7912-CHRD-RP	GGTGTCGTGGAATAACCTAAACACTATCAAACGCrCGATC/3SpC3/	39	1512
Upstream LDR	AcDx-7913-CHRD-Up	TTTTCGGCATCCGCTTCCAGGTAGGAGGTTGGCGAATGGCrGGGAC/3SpC3/	45	1513
Downstream LDR	AcDx-7914-CHRD-Dn	/5Phos/GGAGTTGTGGTTGAGGTTG*GGTTATTAAAGCGGTGGTTAACAGAGGACAGGCCA	54	1514
Real-Time Probe	AcDx-7915-CHRD-RT-Pb	/56-FAM/TTGAATGGC/ZEN/GGAGTTGTGGTTGAGGTTG/3IABkFQ/	28	1515
Tag Forward Primer	AcDx-7916-CHRD-RT-FP	TTTTCGGCATCCGCTTCCA	19	1516
Tag Reverse Primer	AcDx-7917-CHRD-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	1517
Downstream PCR Primer	AcDx-7918-CHRD-PCR-V	TGGCCTGTCCTCTGTTAACCAAAAAAATAACCTAAACACTATCAAACGCTGrCTTTG/3SpC3/	56	1518
GLI3-S1				
Forward PCR Primer	AcDx-7921-GLI3-S1-FP	GTCGGGTTCGCGTAGCrGTTGC/3SpC3/	21	1519
Reverse PCR Primer	AcDx-7922-GLI3-S1-RP	GGTGTCGTGGCGCGCCGAAACCGAArAAAAT/3SpC3/	30	1520
Upstream LDR	AcDx-7923-GLI3-S1-Up	TCCGGGTATACACTGTCCCACGTTGTTTGTAGTCGCGTCATCrGTACC/3SpC3/	47	1521
Downstream LDR	AcDx-7924-GLI3-S1-Dn	/5Phos/GTATTTATTATTATTATATTAGCGCGAGGAAGTTTACGTCGTGGTTAACAGAGGACAGGCCA	62	1522
Real-Time Probe	AcDx-7925-GLI3-S1-RT-Pb	/56-FAM/TTCGTCATC/ZEN/GTATTTATTATTATTATATTAGCGCGAGGAAGTT/3IABkFQ/	43	1523

Tag Forward Primer	AcDx-7926-GLI3-S1-RT-FP	TCCGGGTATACACTGTCCCA	20	1524
Tag Reverse Primer	AcDx-7927-GLI3-S1-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	1525
Downstream PCR Primer	AcDx-7928-GLI3-S1-PCR-V	TGGCCTGTCCTCTGTTAACCAGCCGAAACCGAAAAAACTCGATGrUAAAT/3SpC3/	49	1526
SORCS3				
Forward PCR Primer	AcDx-7931-SORCS3-FP	TTTTCGAGATTTTTGTATTTTTTTCGCrGCGTC/3SpC3/	32	1527
Reverse PCR Primer	AcDx-7932-SORCS3-RP	GGTGTCGTGGCCGACGAAAAATTAAAAACCGCrUTCTG/3SpC3/	37	1528
Upstream LDR	AcDx-7933-SORCS3-Up	TGCGACTCTATTCACGTCCAAGATTTTTGTATTTTTTTCGCGCGTTATTTCTCrGTTCC/3SpC3/	58	1529
Downstream LDR	AcDx-7934-SORCS3-Dn	/5Phos/GTTTTTTATAATAGGTTCGAAGAGTAAATTTTTAAACGCGTTTAGTTGCTATTTGGTGTACCGCCA	66	1530
Real-Time Probe	AcDx-7935-SORCS3-RT-Pb	/56-FAM/AAATTTCTC/ZEN/GTTTTTTATAATAGGTTCGAAGAGTAAATTTTTAAACG/3IABkFQ/	47	1531
Tag Forward Primer	AcDx-7936-SORCS3-RT-FP	TGCGACTCTATTCACGTCCAA	21	1532
Tag Reverse Primer	AcDx-7937-SORCS3-RT-RP	TGGCGGTACACCAAATAGCAA	21	1533
Downstream PCR Primer	AcDx-7938-SORCS3-PCR-V	TGGCGGTACACCAAATAGCAACGACGAAAAATTAAAAACCGCTTCTAAATGrCGTTC/3SpC3/	56	1534
ZNF334				
Forward PCR Primer	AcDx-7941-ZNF334-FP	GGCGAATCGGAAGGGCrGTTTG/3SpC3/	21	1535
Reverse PCR Primer	AcDx-7942-ZNF334-RP	GGTGTCGTGGATAATCCGAAAAACCGACGAArAAACT/3SpC3/	36	1536
Upstream LDR	AcDx-7943-ZNF334-Up	TTGTGCAGAGCGAACAACAAGGGCGTTTAGGTAAAAAATAGGAATAGTATACGCrGTAAG/3SpC3/	59	1537
Downstream LDR	AcDx-7944-ZNF334-Dn	/5Phos/GTAGAAAGGTTTTCGGGTCGTTTTTTTCGAGGTTGCTATTTGGTGTACCGCCA	53	1538
Real-Time Probe	AcDx-7945-ZNF334-RT-Pb	/56-FAM/TTTATACGC/ZEN/GTAGAAAGGTTTTCGGGTCGTTTTTTTCG/3IABkFQ/	38	1539
Tag Forward Primer	AcDx-7946-ZNF334-RT-FP	TTGTGCAGAGCGAACAACAA	20	1540
Tag Reverse Primer	AcDx-7947-ZNF334-RT-RP	TGGCGGTACACCAAATAGCAA	21	1541
Downstream PCR Primer	AcDx-7948-ZNF334-PCR-V	TGGCGGTACACCAAATAGCAACCGAAAAACCGACGAAAAACCTGrAAAAG/3SpC3/	49	1542

CaNCR43				
Forward PCR Primer	AcDx-7951-CaNCR43-FP	AGATTTTAGGGTCGTAGTTTTTTTTGCrGTATG/3SpC3/	32	1543
Reverse PCR Primer	AcDx-7952-CaNCR43-RP	GGTGTCGTGGCCAACAACCCCGAATATTTCCrCAAAC/3SpC3/	36	1544
Upstream LDR	AcDx-7953-CaNCR43-Up	TCCAAACGATTAGGAGCGTCAAGATTTTAGGGTCGTAGTTTTTTTTGCGTATATGTCrGAGCT/3SpC3/	62	1545
Downstream LDR	AcDx-7954-CaNCR43-Dn	/5Phos/GAGTCGGGATTTTGTTTCGATTATTCGTCGGGTTGGACAGAGGTATACGCCCA	53	1546
Real-Time Probe	AcDx-7955-CaNCR43-RT-Pb	/56-FAM/AAATATGTC/ZEN/GAGTCGGGATTTTGTTTCGATTATTCGT/3IABkFQ/	37	1547
Tag Forward Primer	AcDx-7956-CaNCR43-RT-FP	TCCAAACGATTAGGAGCGTCAA	22	1548
Tag Reverse Primer	AcDx-7957-CaNCR43-RT-RP	TGGGCGTATACCTCTGTCCAA	21	1549
Downstream PCR Primer	AcDx-7958-CaNCR43-PCR-V	TGGGCGTATACCTCTGTCCAACCGAATATTTCCCAAATTATTCAAAAATACCTGrACGAG/3SpC3/	59	1550
ITGA8				
Forward PCR Primer	AcDx-7961-ITGA8-FP	GGGTTGGTGGAATTTGGCrGGTTC/3SpC3/	23	1551
Reverse PCR Primer	AcDx-7962-ITGA8-RP	GGTGTCGTGGCCGAACACTATCACCACGAArUACGT/3SpC3/	35	1552
Upstream LDR	AcDx-7963-ITGA8-Up	TCCAAACGATTAGGAGCGTCAATGGAATTTGGCGGTTTTTAGTTGCTCrGTGCC/3SpC3/	53	1553
Downstream LDR	AcDx-7964-ITGA8-Dn	/5Phos/GTGTTTCGGGTCGGTGCGTTCGTTGGACAGAGGTATACGCCCA	43	1554
Real-Time Probe	AcDx-7965-ITGA8-RT-Pb	/56-FAM/AAGTTGCTC/ZEN/GTGTTTCGGGTCGGTG/3IABkFQ/	25	1555
Tag Forward Primer	AcDx-7966-ITGA8-RT-FP	TCCAAACGATTAGGAGCGTCAA	22	1556
Tag Reverse Primer	AcDx-7967-ITGA8-RT-RP	TGGGCGTATACCTCTGTCCAA	21	1557
Downstream PCR Primer	AcDx-7968-ITGA8-PCR-V	TGGGCGTATACCTCTGTCCAACTATCACCACGAATACGCCGAATGrCACCA/3SpC3/	50	1558
BVES-S1				
Forward PCR Primer	AcDx-7971-BVES-S1-FP	TCGAGGGTTCGAGGACrGTTGA/3SpC3/	21	1559
Reverse PCR Primer	AcDx-7972-BVES-S1-RP	GGTGTCGTGGGCCCTTCTCCCCGAArUCTAG/3SpC3/	30	1560
Upstream LDR	AcDx-7973-BVES-S1-Up	TTGCAGCGGGTCACAACAAGACGTTGAGGTTGTTTGATTTGGGCrGGCAA/3SpC3/	49	1561
Downstream LDR	AcDx-7974-BVES-S1-Dn	/5Phos/CAAAAACCCTCAACGATCAACGACCGCCTTGGACAGAGGTATACGCCCA	49	1562

Real-Time Probe	AcDx-7975-BVES-S1-RT-Pb	/56-FAM/CCTTTGGGC/ZEN/GGCGGTCGTTGATC/3IABkFQ/	23	1563
Tag Forward Primer	AcDx-7976-BVES-S1-RT-FP	TTGCAGCGGGTCACAACAA	19	1564
Tag Reverse Primer	AcDx-7977-BVES-S1-RT-RP	TGGGCGTATACCTCTGTCCAA	21	1565
Downstream PCR Primer	AcDx-7978-BVES-S1-PCR-V	TGGGCGTATACCTCTGTCCAACTTCTCCCCGAATCTAAACGCTATGrACCAT/3SpC3/	51	1566
TBX5				
Forward PCR Primer	AcDx-7981-TBX5-FP	GTTTCGTTAGTCGCGGCrGGTTC/3SpC3/	22	1567
Reverse PCR Primer	AcDx-7982-TBX5-RP	GGTGTCGTGGACGACGAAAATAACGAATTCCAACrCCGAT/3SpC3/	39	1568
Upstream LDR	AcDx-7983-TBX5-Up	TCGTCCCGGTCAGTAGTCAACGGTTTTTTAGATTTTATTTGTCGAGGAGTCTCrGAGCT/3SpC3/	58	1569
Downstream LDR	AcDx-7984-TBX5-Dn	/5Phos/GAGTCGCGTAAAATTTTTGTAAGTTGTGTTTTGTAGAATGTTGCCCATTTTCTGCACCCA	60	1570
Real-Time Probe	AcDx-7985-TBX5-RT-Pb	/56-FAM/TTGAGTCTC/ZEN/GAGTCGCGTAAAATTTTTGTAAGTTGTG/3IABkFQ/	37	1571
Tag Forward Primer	AcDx-7986-TBX5-RT-FP	TCGTCCCGGTCAGTAGTCAA	20	1572
Tag Reverse Primer	AcDx-7987-TBX5-RT-RP	TGGGTGCAGAAAATGGGCAA	20	1573
Downstream PCR Primer	AcDx-7988-TBX5-PCR-V	TGGGTGCAGAAAATGGGCAACGACGAAAATAACGAATTCCAACCTGrACTTC/3SpC3/	51	1574
EFS				
Forward PCR Primer	AcDx-7991-EFS-FP	GTCGTGAGGTTGGGTCrGTTGA/3SpC3/	21	1575
Reverse PCR Primer	AcDx-7992-EFS-RP	GGTGTCGTGGCGCTAAATAAATACGCGCGTAAArAACCA/3SpC3/	38	1576
Upstream LDR	AcDx-7993-EFS-Up	TTCGTGCGTCGTGTAGCAAGGTTGGGTCGTTGGAGTGATCrGTTCT/3SpC3/	45	1577
Downstream LDR	AcDx-7994-EFS-Dn	/5Phos/GTTTCGGTTTAGATATTCGTATTTTCGGCGGTGCTTGCCCATTTTCTGCACCCA	54	1578
Real-Time Probe	AcDx-7995-EFS-RT-Pb	/56-FAM/TTAGTGATC/ZEN/GTTTCGGTTTAGATATTCGTATTTTCGGCG/3IABkFQ/	39	1579
Tag Forward Primer	AcDx-7996-EFS-RT-FP	TTCGTGCGTCGTGTAGCAA	19	1580
Tag Reverse Primer	AcDx-7997-EFS-RT-RP	TGGGTGCAGAAAATGGGCAA	20	1581
Downstream PCR Primer	AcDx-7998-EFS-PCR-V	TGGGTGCAGAAAATGGGCAACGCGCGTAAAAACCGCTGrCACCA/3SpC3/	43	1582
GRIA4				
Forward PCR Primer	AcDx-8001-GRIA4-FP	GCGAGTTGGAGAGCGCrGTGTA/3SpC3/	21	1583

Reverse PCR Primer	AcDx-8002-GRIA4-RP	GGTGTCGTGGCACTCGCACTCGCGCrUCGCT/3SpC3/	30	1584
Upstream LDR	AcDx-8003-GRIA4-Up	TTCGTGCGTCGTGTAGCAAGCGCGTGTGGCGATCrGCGAT/3SpC3/	39	1585
Downstream LDR	AcDx-8004-GRIA4-Dn	/5Phos/GCGGCGTTAGTGTTTGTGTGTATGTGAGAAAGTTGCCCATTTTCTGCACCCA	52	1586
Real-Time Probe	AcDx-8005-GRIA4-RT-Pb	/56-FAM/AAGGCGATC/ZEN/GCGGCGTTAGTGTTTGT/3IABkFQ/	26	1587
Tag Forward Primer	AcDx-8006-GRIA4-RT-FP	TTCGTGCGTCGTGTAGCAA	19	1588
Tag Reverse Primer	AcDx-8007-GRIA4-RT-RP	TGGGTGCAGAAAATGGGCAA	20	1589
Downstream PCR Primer	AcDx-8008-GRIA4-PCR-V	TGGGTGCAGAAAATGGGCAACTCGCACTCGCGCTTGrCCCCC/3SpC3/	41	1590
ELMO1-S1				
Forward PCR Primer	AcDx-8011-ELMO1-S1-FP	CGGTTTTTTTTTTTGTCGCGTCrGAGGC/3SpC3/	27	1591
Reverse PCR Primer	AcDx-8012-ELMO1-S1-RP	GGTGTCGTGGTTACCGCTACTATCCTACGACCrCCAAG/3SpC3/	37	1592
Upstream LDR	AcDx-8013-ELMO1-S1-Up	TGGCGTAAGGACACTCTGAAACCGCGTCGAGGTTAGCGAATCrGGGAT/3SpC3/	47	1593
Downstream LDR	AcDx-8014-ELMO1-S1-Dn	/5Phos/GGAGCGCGGCGTTAGTTTAGGAAATTTTACGTTTCCTCTGTGCGGACCA	49	1594
Real-Time Probe	AcDx-8015-ELMO1-S1-RT-Pb	/56-FAM/ATGCGAATC/ZEN/GGAGCGCGGC/3IABkFQ/	19	1595
Tag Forward Primer	AcDx-8016-ELMO1-S1-RT-FP	TGGCGTAAGGACACTCTGAAAC	22	1596
Tag Reverse Primer	AcDx-8017-ELMO1-S1-RT-RP	TGGTCCGCACAGAGGAAAC	19	1597
Downstream PCR Primer	AcDx-8018-ELMO1-S1-PCR-V	TGGTCCGCACAGAGGAAACCTATCCTACGACCCCAAACAAATTTGrUAAAG/3SpC3/	50	1598
CTNND2-S1				
Forward PCR Primer	AcDx-8021-CTNND2-S1-FP	TTTGAGCGCGGTCGCrGGGAC/3SpC3/	20	1599
Reverse PCR Primer	AcDx-8022-CTNND2-S1-RP	GGTGTCGTGGGAAAACCGCCTCTCGCrCCGCT/3SpC3/	31	1600
Upstream LDR	AcDx-8023-CTNND2-S1-Up	TCACGCACGTAGGGTCTAAACGATAAGGGATGTTGGCGAGCrGGTGA/3SpC3/	46	1601
Downstream LDR	AcDx-8024-CTNND2-S1-Dn	/5Phos/GGTAGGAGCGAGCGTCGCGTTGTCCGGCTGTGGTTACA	38	1602
Real-Time Probe	AcDx-8025-CTNND2-S1-RT-Pb	/56-FAM/AAGGCGAGC/ZEN/GGTAGGAGCG/3IABkFQ/	19	1603

Tag Forward Primer	AcDx-8026-CTNND2-S1-RT-FP	TCACGCACGTAGGGTCTAAAC	21	1604
Tag Reverse Primer	AcDx-8027-CTNND2-S1-RT-RP	TGTAACCACAGCCGGACAAC	20	1605
Downstream PCR Primer	AcDx-8028-CTNND2-S1-PCR-V	TGTAACCACAGCCGGACAACCCTCTCGCCCGCGATGrCTCGT/3SpC3/	41	1606
ADCYAP1				
Forward PCR Primer	AcDx-8031-ADCYAP1-FP	CGAGTTTCGGTAAACGAGTTTCrGTAGC/3SpC3/	27	1607
Reverse PCR Primer	AcDx-8032-ADCYAP1-RP	GGTGTCGTGGCCACCCGAAAAACATCGCrCGTCC/3SpC3/	33	1608
Upstream LDR	AcDx-8033-ADCYAP1-Up	TCTTACGCCCAGGGAATGTAACGTAAACGAGTTTCGTAGTTTTTTTTGTTGTTCTCrGTTAA/3SpC3/	61	1609
Downstream LDR	AcDx-8034-ADCYAP1-Dn	/5Phos/GTTGGTTTTTGCGGTTTTTGTTTAGATATTAACGTTAGACGGTTGTCCGGCTGTGGTTACA	61	1610
Real-Time Probe	AcDx-8035-ADCYAP1-RT-Pb	/56-FAM/AATGTTCTC/ZEN/GTTGGTTTTTGCGGTTTTTGTTTAGAT/3IABkFQ/	36	1611
Tag Forward Primer	AcDx-8036-ADCYAP1-RT-FP	TCTTACGCCCAGGGAATGTAAC	22	1612
Tag Reverse Primer	AcDx-8037-ADCYAP1-RT-RP	TGTAACCACAGCCGGACAAC	20	1613
Downstream PCR Primer	AcDx-8038-ADCYAP1-PCR-V	TGTAACCACAGCCGGACAACCCGAAAAACATCGCCGTCTAATGrUTAAC/3SpC3/	48	1614
CaNCR44				
Forward PCR Primer	AcDx-8041-CaNCR44-FP	GCGTCGAATTTTTTAGTATGAGCrGAATC/3SpC3/	28	1615
Reverse PCR Primer	AcDx-8042-CaNCR44-RP	GGTGTCGTGGCCCCGAAATCCGAACCTCrUCTAG/3SpC3/	33	1616
Upstream LDR	AcDx-8043-CaNCR44-Up	TGAGCAAAATCTTCGTCGACCGAATTTTTTAGTATGAGCGAATTTTAAGTTATTGTTCTCrGCGCC/3SpC3/	65	1617
Downstream LDR	AcDx-8044-CaNCR44-Dn	/5Phos/GCGTTTGTCGAGCGAATTTTTAAGGTTTGTAGGTCGGTTCGATGCCTTCCGTACA	55	1618
Real-Time Probe	AcDx-8045-CaNCR44-RT-Pb	/56-FAM/CCTGTTCTC/ZEN/GCGTTTGTCGAGCGAATTTTTAAGG/3IABkFQ/	34	1619

Tag Forward Primer	AcDx-8046-CaNCR44-RT-FP	TGAGCAAAATCTTCGTCGACC	21	1620
Tag Reverse Primer	AcDx-8047-CaNCR44-RT-RP	TGTACGGAAGGCATCGAACC	20	1621
Downstream PCR Primer	AcDx-8048-CaNCR44-PCR-V	TGTACGGAAGGCATCGAACCGAAATCCGAACCTCTCTAACTCCTAAAAATGrACCTG/3SpC3/	56	1622
PTGDR-S1				
Forward PCR Primer	AcDx-8051-PTGDR-S1-FP	GTAATTGTGAGTTTTCGGGTTTCrGAGGC/3SpC3/	28	1623
Reverse PCR Primer	AcDx-8052-PTGDR-S1-RP	GGTGTCGTGGCCATCCCGATCCGCTCrCACCT/3SpC3/	31	1624
Upstream LDR	AcDx-8053-PTGDR-S1-Up	TGGCACATGAGAGTAGTTGACCGTTTCGAGGTAGTAGGGTATTGAGATTGCTCrGGTTA/3SpC3/	58	1625
Downstream LDR	AcDx-8054-PTGDR-S1-Dn	/5Phos/GGTCGCGGATGCGGAGCGGTTCGATGCCTTCCGTACA	37	1626
Real-Time Probe	AcDx-8055-PTGDR-S1-RT-Pb	/56-FAM/TTATTGCTC/ZEN/GGTCGCGGATGCG/3IABkFQ/	22	1627
Tag Forward Primer	AcDx-8056-PTGDR-S1-RT-FP	TGGCACATGAGAGTAGTTGACC	22	1628
Tag Reverse Primer	AcDx-8057-PTGDR-S1-RT-RP	TGTACGGAAGGCATCGAACC	20	1629
Downstream PCR Primer	AcDx-8058-PTGDR-S1-PCR-V	TGTACGGAAGGCATCGAACCCCGCTCCACCCTTCCTGrCCCCA/3SpC3/	42	1630
CaNCR45				
Forward PCR Primer	AcDx-8061-CaNCR45-FP	TTTGGGTTAACGGAGGCrGAGGA/3SpC3/	22	1631
Reverse PCR Primer	AcDx-8062-CaNCR45-RP	GGTGTCGTGGGCGCCAAAAAACCGCCrCAACG/3SpC3/	31	1632
Upstream LDR	AcDx-8063-CaNCR45-Up	TATAGTCACGCAGGACCACAGGAGCGAGGGAGTTGCrGTAAG/3SpC3/	41	1633
Downstream LDR	AcDx-8064-CaNCR45-Dn	/5Phos/GTAGAGGGTAGGTTTCGATTTTTTTTCGTCGTATTTTGTGGTGTTTGCGGCTGTCTATGACA	62	1634
Real-Time Probe	AcDx-8065-CaNCR45-RT-Pb	/56-FAM/TTGAGTTGC/ZEN/GTAGAGGGTAGGTTTCGATTTTTTTTCG/3IABkFQ/	37	1635
Tag Forward Primer	AcDx-8066-CaNCR45-RT-FP	TATAGTCACGCAGGACCACA	20	1636

Tag Reverse Primer	AcDx-8067-CaNCR45-RT-RP	TGTCATAGACAGCCGCAAACA	21	1637
Downstream PCR Primer	AcDx-8068-CaNCR45-PCR-V	TGTCATAGACAGCCGCAAACAAACACCCGACCCACAAAATATGrACGAG/3SpC3/	48	1638
TLR5				
Forward PCR Primer	AcDx-8071-TLR5-FP	TCGGTTATTCGTGCGGTCrGGATG/3SpC3/	23	1639
Reverse PCR Primer	AcDx-8072-TLR5-RP	GGTGTCGTGGTTCCTCTCCCAATAACGCTACrUAAAC/3SpC3/	36	1640
Upstream LDR	AcDx-8073-TLR5-Up	TGCCCTATCGAAAAGGACAACAGGTCGGATATCGTGTTAGGTTTGCrGAGAG/3SpC3/	51	1641
Downstream LDR	AcDx-8074-TLR5-Dn	/5Phos/GAGGAGGGTTTTGTCGTAGTTTCGGAGAATAGTGTTTGCGGCTGTCTATGACA	53	1642
Real-Time Probe	AcDx-8075-TLR5-RT-Pb	/56-FAM/ATGGTTTGC/ZEN/GAGGAGGGTTTTGTCGTAGTTT/3IABkFQ/	31	1643
Tag Forward Primer	AcDx-8076-TLR5-RT-FP	TGCCCTATCGAAAAGGACAACA	22	1644
Tag Reverse Primer	AcDx-8077-TLR5-RT-RP	TGTCATAGACAGCCGCAAACA	21	1645
Downstream PCR Primer	AcDx-8078-TLR5-PCR-V	TGTCATAGACAGCCGCAAACACCCAATAACGCTACTAAATACTATTCCCTGrAAACC/3SpC3/	56	1646
CaNCR46				
Forward PCR Primer	AcDx-8081-CaNCR46-FP	GGGTTCGGGAAGTTCGCrGGAAG/3SpC3/	22	1647
Reverse PCR Primer	AcDx-8082-CaNCR46-RP	GGTGTCGTGGACCTAAACTAAAACAAAACTCCGArAAATG/3SpC3/	39	1648
Upstream LDR	AcDx-8083-CaNCR46-Up	TAATCTCCAGACCTCCGAACCGTTCGCGGAAACGTAGGAAGCrGGTCG/3SpC3/	47	1649
Downstream LDR	AcDx-8084-CaNCR46-Dn	/5Phos/GGTTAGGAGAGGTAGCGTTACGTATATTTTTTTTATTTGGTGTAAGGATTGAACGGGACA	60	1650
Real-Time Probe	AcDx-8085-CaNCR46-RT-Pb	/56-FAM/AAAGGAAGC/ZEN/GGTTAGGAGAGGTAGCGTTAC/3IABkFQ/	30	1651
Tag Forward Primer	AcDx-8086-CaNCR46-RT-FP	TAATCTCCAGACCTCCGAACC	21	1652
Tag Reverse Primer	AcDx-8087-CaNCR46-RT-RP	TGTCCCGTTCAATCCTTACATC	22	1653
CaNCR47				

Forward PCR Primer	AcDx-8091-CaNCR47-FP	TTTCGTTTTTGTCGGCGGTAGCrGATTT/3SpC3/	27	1654
Reverse PCR Primer	AcDx-8092-CaNCR47-RP	GGTGTCGTGGACTCAATCCGCGCGCrCCAAT/3SpC3/	30	1655
Upstream LDR	AcDx-8093-CaNCR47-Up	TTTTCCGCGTCAGAGCACAGGCGGTAGCGATTCGGATTCTCrGTTCC/3SpC3/	46	1656
Downstream LDR	AcDx-8094-CaNCR47-Dn	/5Phos/GTTTTCGATAAAGTTTTAGTTTCGTAGTAGTATTCGGCGCGTGTTATCGGACCTAGCTCGACA	63	1657
Real-Time Probe	AcDx-8095-CaNCR47-RT-Pb	/56-FAM/TTGATTCTC/ZEN/GTTTTCGATAAAGTTTTAGTTTCGTAGTAGTATTCGG/3IABkFQ/	46	1658
Tag Forward Primer	AcDx-8096-CaNCR47-RT-FP	TTTTCCGCGTCAGAGCACA	19	1659
Tag Reverse Primer	AcDx-8097-CaNCR47-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	1660
Downstream PCR Primer	AcDx-8098-CaNCR47-PCR-V	TGTCGAGCTAGGTCCGATAACACGCCCAACGAATCCGTGrCCGAG/3SpC3/	44	1661
CaNCR48				
Forward PCR Primer	AcDx-8101-CaNCR48-FP	GAGTATAGAGTATAGTAAATCGGGATTTTCrGGCGA/3SpC3/	35	1662
Reverse PCR Primer	AcDx-8102-CaNCR48-RP	GGTGTCGTGGGACCGCCCAACCCGArCGCGG/3SpC3/	30	1663
Upstream LDR	AcDx-8103-CaNCR48-Up	TTCGCTGCCCGGTTAAACAGGATTTTCGGCGGACAGCrGTTCC/3SpC3/	42	1664
Downstream LDR	AcDx-8104-CaNCR48-Dn	/5Phos/GTTTTTCGTTCGTTTGTTCGTTATGTTGGAGAGTGTTATCGGACCTAGCTCGACA	55	1665
Real-Time Probe	AcDx-8105-CaNCR48-RT-Pb	/56-FAM/TTGGACAGC/ZEN/GTTTTTCGTTCGTTTGTTCGTTATG/3IABkFQ/	34	1666
Tag Forward Primer	AcDx-8106-CaNCR48-RT-FP	TTCGCTGCCCGGTTAAACA	19	1667
Tag Reverse Primer	AcDx-8107-CaNCR48-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	1668
Downstream PCR Primer	AcDx-8108-CaNCR48-PCR-V	TGTCGAGCTAGGTCCGATAACAGCCCAACCCGACGTGrAAAAG/3SpC3/	42	1669
CaNCR49				
Forward PCR Primer	AcDx-8111-CaNCR49-FP	CGACGTGTTTCGTTGAAAGCrGGGTG/3SpC3/	25	1670

Reverse PCR Primer	AcDx-8112-CaNCR49-RP	GGTGTCGTGGCGCCGAAACCCGACAArACCGG/3SpC3/	31	1671
Upstream LDR	AcDx-8113-CaNCR49-Up	TTCGCTGCCCGGTTAAACATGAAAGCGGGTAGGTGGTATCrGGGTA/3SpC3/	45	1672
Downstream LDR	AcDx-8114-CaNCR49-Dn	/5Phos/GGGCGGAGTTGCGTTGAGGTGTTATCGGACCTAGCTCGACA	41	1673
Real-Time Probe	AcDx-8115-CaNCR49-RT-Pb	/56-FAM/TTTGGTATC/ZEN/GGGCGGAGTTGCG/3IABkFQ/	22	1674
Tag Forward Primer	AcDx-8116-CaNCR49-RT-FP	TTCGCTGCCCGGTTAAACA	19	1675
Tag Reverse Primer	AcDx-8117-CaNCR49-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	1676
Downstream PCR Primer	AcDx-8118-CaNCR49-PCR-V	TGTCGAGCTAGGTCCGATAACAGAAACCCGACAAACCGACAAATATGrAAAAG/3SpC3/	52	1677
ALOX5-S1				
Forward PCR Primer	AcDx-8121-ALOX5-S1-FP	TTTGCGGTTAGGTGAAGGCrGTAGG/3SpC3/	24	1678
Reverse PCR Primer	AcDx-8122-ALOX5-S1-RP	GGTGTCGTGGCCCCGCTTTCTCTCTCGArCCTCC/3SpC3/	33	1679
Upstream LDR	AcDx-8123-ALOX5-S1-Up	TGCTATGCCGCATTCAACCATAGAGGTAGGGAGGTCGAGCrGAGTA/3SpC3/	45	1680
Downstream LDR	AcDx-8124-ALOX5-S1-Dn	/5Phos/GAGCGTAGGCGGGCGGAGAGTAGTGGAGCTAGTTCGGCGACA	42	1681
Real-Time Probe	AcDx-8125-ALOX5-S1-RT-Pb	/56-FAM/TTGTCGAGC/ZEN/GAGCGTAGGC/3IABkFQ/	19	1682
Tag Forward Primer	AcDx-8126-ALOX5-S1-RT-FP	TGCTATGCCGCATTCAACCA	20	1683
Tag Reverse Primer	AcDx-8127-ALOX5-S1-RT-RP	TGTCGCCGAACTAGCTCCA	19	1684
Downstream PCR Primer	AcDx-8128-ALOX5-S1-PCR-V	TGTCGCCGAACTAGCTCCACCTCTTCCGCGTAATCTACTCTCTGrCCCGT/3SpC3/	49	1685
PAX9				
Forward PCR Primer	AcDx-8131-PAX9-FP	GCGTATTAATTTTTTATCGTTTCGTTCrGTTGC/3SpC3/	32	1686
Reverse PCR Primer	AcDx-8132-PAX9-RP	GGTGTCGTGGCTCACTCACTACGCGTTTCCrUCTAG/3SpC3/	35	1687
Upstream LDR	AcDx-8133-PAX9-Up	TGTGCCTTACGGAAAACCCAGTTCGTTGTTTAGGTGCGAGGACTCrGTTGA/3SpC3/	50	1688
Downstream LDR	AcDx-8134-PAX9-Dn	/5Phos/GTTAGACGTTATAGGATTGGGAGCGATTCGTAAGGTGGAGCTAGTTCGGCGA	54	1689

		CA		
Real-Time Probe	AcDx-8135-PAX9-RT-Pb	/56-FAM/TTAGGACTC/ZEN/GTTAGACGTTATAGGATTGGGAG/3IABkFQ/	32	1690
Tag Forward Primer	AcDx-8136-PAX9-RT-FP	TGTGCCTTACGGAAAACCCA	20	1691
Tag Reverse Primer	AcDx-8137-PAX9-RT-RP	TGTCGCCGAACTAGCTCCA	19	1692
Downstream PCR Primer	AcDx-8138-PAX9-PCR-V	TGTCGCCGAACTAGCTCCAGTTTCCTCTAAATAACATAATACCCTTACGAATTGrCCCCT/3SpC3/	59	1693
RNF220-S1				
Forward PCR Primer	AcDx-8141-RNF220-S1-FP	GTAGAAGTGATTCGGGTTGTCrGTTTC/3SpC3/	26	1694
Reverse PCR Primer	AcDx-8142-RNF220-S1-RP	GGTGTCGTGGCGCCCCCTCCCCCAArAAAAC/3SpC3/	30	1695
Upstream LDR	AcDx-8143-RNF220-S1-Up	TCCTGCTCTGAAAACCTACACCGTGATTCGGGTTGTCGTTTTTTTGCrGGTTA/3SpC3/	52	1696
Downstream LDR	AcDx-8144-RNF220-S1-Dn	/5Phos/GGTCGTTCGTTAGTTTTTCGTCGTTGTCGGTAGGGTTACATAGGCGGCTTAGACA	55	1697
Real-Time Probe	AcDx-8145-RNF220-S1-RT-Pb	/56-FAM/AATTTTTGC/ZEN/GGTCGTTCGTTAGTTTTTCGTCG/3IABkFQ/	32	1698
Tag Forward Primer	AcDx-8146-RNF220-S1-RT-FP	TCCTGCTCTGAAAACCTACACC	22	1699
Tag Reverse Primer	AcDx-8147-RNF220-S1-RT-RP	TGTCTAAGCCGCCTATGTAACC	22	1700
Downstream PCR Primer	AcDx-8148-RNF220-S1-PCR-V	TGTCTAAGCCGCCTATGTAACCAATCAATTACCAACCACCCTTCTACTGrACAAT/3SpC3/	54	1701
CaNCR50				
Forward PCR Primer	AcDx-8151-CaNCR50-FP	GGAGGGCGTTTTTCGCrGTTGC/3SpC3/	21	1702
Reverse PCR Primer	AcDx-8152-CaNCR50-RP	GGTGTCGTGGCGCCCCCGAACCCTArATCCA/3SpC3/	30	1703
Upstream LDR	AcDx-8153-CaNCR50-Up	TTTTTACGCACAGCACCACCCGTTTTTCGCGTTGTTTTTGATTCGCTCrGGTCC/3SpC3/	53	1704
Downstream LDR	AcDx-8154-CaNCR50-Dn	/5Phos/GGTTTTGCGTTTGTTTGAAGTTTTTCGTTTCGTATTTGGTTACATAGGCGGCTTAGACA	59	1705
Real-Time Probe	AcDx-8155-CaNCR50-RT-Pb	/56-FAM/CCTTCGCTC/ZEN/GGTTTTGCGTTTGTTTGAAGTTTTTC/3IABkFQ/	35	1706
Tag Forward Primer	AcDx-8156-CaNCR50-RT-	TTTTTACGCACAGCACCACC	20	1707

	FP			
Tag Reverse Primer	AcDx-8157-CaNCR50-RT-RP	TGTCTAAGCCGCCTATGTAACC	22	1708
Downstream PCR Primer	AcDx-8158-CaNCR50-PCR-V	TGTCTAAGCCGCCTATGTAACCCCCCGAACCCTAATCCGAAAATATGrAAACA/3SpC3/	52	1709
GALR3				
Forward PCR Primer	AcDx-8161-GALR3-FP	GTACGGTCGTTTCGTTTTTAGTTCrGGTTG/3SpC3/	29	1710
Reverse PCR Primer	AcDx-8162-GALR3-RP	GGTGTCGTGGGCGAAACGAACGCGTAArACGAG/3SpC3/	32	1711
Upstream LDR	AcDx-8163-GALR3-Up	TGGACACTTCGCCCTTCTTAACGTTTCGTTTTTAGTTCGGTTATTTACGTTTATCrGTTCA/3SpC3/	60	1712
Downstream LDR	AcDx-8164-GALR3-Dn	/5Phos/GTTTGGTTTTATATTGTTTGGTTTACGTTAATTTTTGTTTTAATTCGTTTGGGATCTGGGCATCACA	67	1713
Real-Time Probe	AcDx-8165-GALR3-RT-Pb	/56-FAM/TTGTTTATC/ZEN/GTTTGGTTTTATATTGTTTGGTTTACGTTAATTTTTG/3IABkFQ/	46	1714
Tag Forward Primer	AcDx-8166-GALR3-RT-FP	TGGACACTTCGCCCTTCTTAAC	22	1715
Tag Reverse Primer	AcDx-8167-GALR3-RT-RP	TGTGATGCCCAGATCCCAAAC	21	1716
Downstream PCR Primer	AcDx-8168-GALR3-PCR-V	TGTGATGCCCAGATCCCAAACCGAAACGAACGCGTAAACGAATGrAATTG/3SpC3/	49	1717
CaNCR51				
Forward PCR Primer	AcDx-8171-CaNCR51-FP	GGGATTGCGGGACGCrGGGCA/3SpC3/	20	1718
Reverse PCR Primer	AcDx-8172-CaNCR51-RP	GGTGTCGTGGCCGCAACCCCTAAAACGAArAATAG/3SpC3/	34	1719
Upstream LDR	AcDx-8173-CaNCR51-Up	TGGAGGCCGGAGAAATTAAACGAGCGAGAGTTTTGTGGGTTGTTAAATCrGGCAT/3SpC3/	54	1720
Downstream LDR	AcDx-8174-CaNCR51-Dn	/5Phos/GGCGCGGTCGCGTCGCGTTTGGGATCTGGGCATCACA	37	1721
Real-Time Probe	AcDx-8175-CaNCR51-RT-Pb	/56-FAM/AATTAAATC/ZEN/GGCGCGGTCGCGTC/3IABkFQ/	23	1722
Tag Forward Primer	AcDx-8176-CaNCR51-RT-FP	TGGAGGCCGGAGAAATTAAAC	21	1723
Tag Reverse Primer	AcDx-8177-CaNCR51-RT-	TGTGATGCCCAGATCCCAAAC	21	1724

	RP			
Downstream PCR Primer	AcDx-8178-CaNCR51-PCR-V	TGTGATGCCCAGATCCCAAACCTAAAACGAAAATAACAAAAAAAACGACTACTGrCGACA/3SpC3/	59	1725
CaNCR52				
Forward PCR Primer	AcDx-8181-CaNCR52-FP	GTTTTTTATTGTAAGCGGCGTTATCrGGATC/3SpC3/	30	1726
Reverse PCR Primer	AcDx-8182-CaNCR52-RP	GGTGTCGTGGCTAATTAACATACGACGCCGATTArCCCGG/3SpC3/	39	1727
Upstream LDR	AcDx-8183-CaNCR52-Up	TTGTCTCTGCGACCCATCAAGGATTTTATTTGTAAATTTATTAGTGTGCGTTTGCrGGGAG/3SpC3/	60	1728
Downstream LDR	AcDx-8184-CaNCR52-Dn	/5Phos/GGAGAGGAGGTTCGGGAATTCGTTGGTACACGTTCGGCACA	41	1729
Real-Time Probe	AcDx-8185-CaNCR52-RT-Pb	/56-FAM/AACGTTTGC/ZEN/GGAGAGGAGGTTCG/3IABkFQ/	23	1730
Tag Forward Primer	AcDx-8186-CaNCR52-RT-FP	TTGTCTCTGCGACCCATCAA	20	1731
Tag Reverse Primer	AcDx-8187-CaNCR52-RT-RP	TGTGCCGAACGTGTACCAA	19	1732
Downstream PCR Primer	AcDx-8188-CaNCR52-PCR-V	TGTGCCGAACGTGTACCAACGCCGATTACCCGAACCTGrAATTT/3SpC3/	43	1733
PRKCB				
Forward PCR Primer	AcDx-8191-PRKCB-FP	TTAAGCGTAGTTGGACGAGCrGGTAA/3SpC3/	25	1734
Reverse PCR Primer	AcDx-8192-PRKCB-RP	GGTGTCGTGGTCCCCTACGCCGACTCrUAACA/3SpC3/	31	1735
Upstream LDR	AcDx-8193-PRKCB-Up	TTCGCCTACCGCAGTGAACACGAGCGGTAGTAGTTGAGCrGAGCA/3SpC3/	44	1736
Downstream LDR	AcDx-8194-PRKCB-Dn	/5Phos/GAGTGATAGTTTCGGTTTCGCGCGTCGGTTGAGACATGGGCTCGCA	46	1737
Real-Time Probe	AcDx-8195-PRKCB-RT-Pb	/56-FAM/ATGTTGAGC/ZEN/GAGTGATAGTTTCGGTTTCGC/3IABkFQ/	30	1738
Tag Forward Primer	AcDx-8196-PRKCB-RT-FP	TTCGCCTACCGCAGTGAAC	19	1739
Tag Reverse Primer	AcDx-8197-PRKCB-RT-RP	TGCGAGCCCATGTCTCAAC	19	1740
Downstream PCR Primer	AcDx-8198-PRKCB-PCR-V	TGCGAGCCCATGTCTCAACCGCCGACTCTAACGACTGrCGACA/3SpC3/	42	1741
BNC1				

Forward PCR Primer	AcDx-8201-BNC1-FP	TTTCGGAGTTTTATTGTCGCrGGAAA/3SpC3/	25	1742
Reverse PCR Primer	AcDx-8202-BNC1-RP	GGTGTCGTGGCATCCCAAAACGCTCGTCrCCGCG/3SpC3/	33	1743
Upstream LDR	AcDx-8203-BNC1-Up	TCCTGAATTGGCCACACCACGTCGCGGAGGAAGAAGGTGGCrGTCAC/3SpC3/	46	1744
Downstream LDR	AcDx-8204-BNC1-Dn	/5Phos/GTCGTTTTCGTTTGTAGAAATTTTATTTTCGAGAAAGTGCGGGTGACTGAGCG ACGTCTAACA	63	1745
Real-Time Probe	AcDx-8205-BNC1-RT-Pb	/56-FAM/TTAGGTGGC/ZEN/GTCGTTTTCGTTTGTAGAAATT/3IABkFQ/	31	1746
Tag Forward Primer	AcDx-8206-BNC1-RT-FP	TCCTGAATTGGCCACACCAC	20	1747
Tag Reverse Primer	AcDx-8207-BNC1-RT-RP	TGTTAGACGTCGCTCAGTCAC	21	1748
Downstream PCR Primer	AcDx-8208-BNC1-PCR-V	TGTTAGACGTCGCTCAGTCACGCTCGTCCCGCACTTTCTTGrAAAAC/3SpC3/	46	1749
MIR129-2				
Forward PCR Primer	AcDx-8211-MIR129-2-FP	GGAGATACGAGTTTAGAGGCrGCGAG/3SpC3/	25	1750
Reverse PCR Primer	AcDx-8212-MIR129-2-RP	GGTGTCGTGGCTTCAACCCAAAATATCTCCGAArCCCTG/3SpC3/	38	1751
Upstream LDR	AcDx-8213-MIR129-2-Up	TTCTTGCGGTTCTGGAACACCGGAGTGGTGAGATTGAGTTGCrGATAA/3SpC3/	47	1752
Downstream LDR	AcDx-8214-MIR129-2-Dn	/5Phos/GATGGAACGCGTTGAGGAGATTTAGTTTGTTCGTGATGCTCCGTTGTTGCTAA	53	1753
Real-Time Probe	AcDx-8215-MIR129-2-RT-Pb	/56-FAM/AAGAGTTGC/ZEN/GATGGAACGCGTTGAG/3IABkFQ/	25	1754
Tag Forward Primer	AcDx-8216-MIR129-2-RT-FP	TTCTTGCGGTTCTGGAACAC	20	1755
Tag Reverse Primer	AcDx-8217-MIR129-2-RT-RP	TTAGCAACAACGGAGCATCAC	21	1756
Downstream PCR Primer	AcDx-8218-MIR129-2-PCR-V	TTAGCAACAACGGAGCATCACCCAAAATATCTCCGAACCCTAAAACTGrAACAG/3SpC3/	53	1757
ADCY4-S1				
Forward PCR Primer	AcDx-8221-ADCY4-S1-FP	CGGAGGTTGAAGAGGCrGGGTC/3SpC3/	21	1758
Reverse PCR Primer	AcDx-8222-ADCY4-S1-RP	GGTGTCGTGGACCCGCCGAACCGAArAAAAG/3SpC3/	30	1759
Upstream LDR	AcDx-8223-ADCY4-S1-Up	TATGGTAAAATGTCAGCGGCACGAAGAGGCGGGTTATGATTTTTTTAGTCTCrGAGCC/3SpC3/	57	1760
Downstream LDR	AcDx-8224-ADCY4-S1-Dn	/5Phos/GAGTTTCGAGGTTGGTTAGGGTCGGGTGATGCTCCGTTGTTGCTAA	46	1761
Real-Time Probe	AcDx-8225-ADCY4-S1-RT-	/56-FAM/AATAGTCTC/ZEN/GAGTTTCGAGGTTGGTTAGGG/3IABkFQ/	30	1762

	Pb			
Tag Forward Primer	AcDx-8226-ADCY4-S1-RT-FP	TATGGTAAAATGTCAGCGGCAC	22	1763
Tag Reverse Primer	AcDx-8227-ADCY4-S1-RT-RP	TTAGCAACAACGGAGCATCAC	21	1764
Downstream PCR Primer	AcDx-8228-ADCY4-S1-PCR-V	TTAGCAACAACGGAGCATCACCGAAAAAAATAACCCGACGCCTGrACCCC/3SpC3/	49	1765
CaNCR53				
Forward PCR Primer	AcDx-8231-CaNCR53-FP	TTTTTAGTCGTGTTCGGTTTTCrGTCGC/3SpC3/	27	1766
Reverse PCR Primer	AcDx-8232-CaNCR53-RP	GGTGTCGTGGCCGCTCTCCCCGATCrUACCT/3SpC3/	30	1767
Upstream LDR	AcDx-8233-CaNCR53-Up	TCCTCGAGCCGATGACACACGTGTTCGGTTTTCGTCGTTTTTTTATTTCACrGTGAC/3SpC3/	56	1768
Downstream LDR	AcDx-8234-CaNCR53-Dn	/5Phos/GTGGTGGAATTTTTCGCGTTTTTTATAGTCGTCGTGTAACGTCCGTGGGCTAA	53	1769
Real-Time Probe	AcDx-8235-CaNCR53-RT-Pb	/56-FAM/CCATTTCAC/ZEN/GTGGTGGAATTTTTCGCGTTTTT/3IABkFQ/	32	1770
Tag Forward Primer	AcDx-8236-CaNCR53-RT-FP	TCCTCGAGCCGATGACACA	19	1771
Tag Reverse Primer	AcDx-8237-CaNCR53-RT-RP	TTAGCCCACGGACGTTACA	19	1772
Downstream PCR Primer	AcDx-8238-CaNCR53-PCR-V	TTAGCCCACGGACGTTACACCCCGATCTACCCATTAATTCGATGrACTAC/3SpC3/	49	1773
CaNCR54				
Forward PCR Primer	AcDx-8241-CaNCR54-FP	GAAATTTTTTTGCGTTATTAGATTGCrGTTTG/3SpC3/	31	1774
Reverse PCR Primer	AcDx-8242-CaNCR54-RP	GGTGTCGTGGAAAAACTTATAAATACTTTCCGTCGAArCTCAG/3SpC3/	42	1775
Upstream LDR	AcDx-8243-CaNCR54-Up	TACTATCGTATCACGCCGACAGTTTAGTATAGTAGATGGTACGAGTACGCGCrGGCAC/3SpC3/	57	1776
Downstream LDR	AcDx-8244-CaNCR54-Dn	/5Phos/GGCGTTTAGTTTAGGTTTTCGGAGGTAGTTGAGTGTAACGTCCGTGGGCTAA	52	1777
Real-Time Probe	AcDx-8245-CaNCR54-RT-Pb	/56-FAM/TTTACGCGC/ZEN/GGCGTTTAGTTTAGGTTTT/3IABkFQ/	28	1778

Tag Forward Primer	AcDx-8246-CaNCR54-RT-FP	TACTATCGTATCACGCCGACA	21	1779
Tag Reverse Primer	AcDx-8247-CaNCR54-RT-RP	TTAGCCCACGGACGTTACA	19	1780
Downstream PCR Primer	AcDx-8248-CaNCR54-PCR-V	TTAGCCCACGGACGTTACAGTCGAAATTTCCAACTCAACTACCTCTGrAAAAT/3SpC3/	52	1781
HIST1H4F-S1				
Forward PCR Primer	AcDx-8251-HIST1H4F-S1-FP	GTGTAGAGGACGTTTAGTAAGTTACrGGAAG/3SpC3/	30	1782
Reverse PCR Primer	AcDx-8252-HIST1H4F-S1-RP	GGTGTCGTGGACCCCTTATATATCAATTATACGAACCrUTTTC/3SpC3/	42	1783
Upstream LDR	AcDx-8253-HIST1H4F-S1-Up	TTGAGACCGCTGACCGACAGGACGTTTAGTAAGTTACGGAAAAAGCrGGAAG/3SpC3/	51	1784
Downstream LDR	AcDx-8254-HIST1H4F-S1-Dn	/5Phos/GGAGATAGAGGTTTCGTTTTCGTTTTTTTAATTTAGTTTTTAAAATGTTGCACG GTCGAGCTAA	64	1785
Real-Time Probe	AcDx-8255-HIST1H4F-S1-RT-Pb	/56-FAM/CCAAAAAGC/ZEN/GGAGATAGAGGTTTCGTTTTCGTTTTTT/3IABkFQ/	37	1786
Tag Forward Primer	AcDx-8256-HIST1H4F-S1-RT-FP	TTGAGACCGCTGACCGACA	19	1787
Tag Reverse Primer	AcDx-8257-HIST1H4F-S1-RT-RP	TTAGCTCGACCGTGCAACA	19	1788
TDRD10-S1				
Forward PCR Primer	AcDx-8261-TDRD10-S1-FP	TAGTTCGCGTTTGTATCGAGTCrGGTTC/3SpC3/	27	1789
Reverse PCR Primer	AcDx-8262-TDRD10-S1-RP	GGTGTCGTGGACGAAAAACTTCCTTCCCGAArAATAG/3SpC3/	36	1790
Upstream LDR	AcDx-8263-TDRD10-S1-Up	TCGCAACGTGCCGAATACAGAGTCGGTTTCGGTCGAGCrGTTAC/3SpC3/	43	1791
Downstream LDR	AcDx-8264-TDRD10-S1-Dn	/5Phos/GTTGTTTTTATACGCGTTTAGGAGTGTTACGTGCGTGTTGCACGGTCGAGCTA A	54	1792
Real-Time Probe	AcDx-8265-TDRD10-S1-RT-Pb	/56-FAM/TTGTCGAGC/ZEN/GTTGTTTTTATACGCGTTTAGGAGTG/3IABkFQ/	35	1793
Tag Forward Primer	AcDx-8266-TDRD10-S1-	TCGCAACGTGCCGAATACA	19	1794

	RT-FP			
Tag Reverse Primer	AcDx-8267-TDRD10-S1-RT-RP	TTAGCTCGACCGTGCAACA	19	1795
Downstream PCR Primer	AcDx-8268-TDRD10-S1-PCR-V	TTAGCTCGACCGTGCAACACTTCCCGAAAATAAACGACGCATGrUAACG/3SpC3/	48	1796
HEPACAM				
Forward PCR Primer	AcDx-8271-HEPACAM-FP	GTTTTATTATATTAATATTGTCGTTTTCGCrGTATC/3SpC3/	35	1797
Reverse PCR Primer	AcDx-8272-HEPACAM-RP	GGTGTCGTGGGCTAAAAACGAAAAAAAATCCCGArAAAAG/3SpC3/	39	1798
Upstream LDR	AcDx-8273-HEPACAM-Up	TAACGGGATTGAGAGTGGACAAATATTGTCGTTTTCGCGTATTCGTTCTCrGTTCA/3SpC3/	55	1799
Downstream LDR	AcDx-8274-HEPACAM-Dn	/5Phos/GTTTGCGTATGTTTATATACGTTTATATTCGAGATATTAGCGTTTTTGTCTGCCGCCCTTACTAA	65	1800
Real-Time Probe	AcDx-8275-HEPACAM-RT-Pb	/56-FAM/AACGTTCTC/ZEN/GTTTGCGTATGTTTATATACGTTTATATTCGAG/3IABkFQ/	42	1801
Tag Forward Primer	AcDx-8276-HEPACAM-RT-FP	TAACGGGATTGAGAGTGGACA	21	1802
Tag Reverse Primer	AcDx-8277-HEPACAM-RT-RP	TTAGTAAGGGCGGCAGACA	19	1803
Downstream PCR Primer	AcDx-8278-HEPACAM-PCR-V	TTAGTAAGGGCGGCAGACAGCTAAAAACGAAAAAAAATCCCGAAAAAATGrCTAAC/3SpC3/	55	1804
TRIM15-S1				
Forward PCR Primer	AcDx-8281-TRIM15-S1-FP	TCGTTGTTGATGTTTGCGCrGTTTC/3SpC3/	24	1805
Reverse PCR Primer	AcDx-8282-TRIM15-S1-RP	GGTGTCGTGGTCTAACAAACTCTTCTTCTACCGAArUATAT/3SpC3/	40	1806
Upstream LDR	AcDx-8283-TRIM15-S1-Up	TAACGGGATTGAGAGTGGACATTTTAGAGGTTATTATTTTGGATTTTTAGATCGTTAATCrGGAAC/3SpC3/	65	1807
Downstream LDR	AcDx-8284-TRIM15-S1-Dn	/5Phos/GGAGTTTGGTTTTTTCGGAAGATAGGAAGTTAGTGAGTGTCTGCCGCCCTTACTAA	56	1808
Real-Time Probe	AcDx-8285-TRIM15-S1-RT-Pb	/56-FAM/AAGTTAATC/ZEN/GGAGTTTGGTTTTTTCGGAAGATAGGAAG/3IABkFQ/	38	1809
Tag Forward Primer	AcDx-8286-TRIM15-S1-RT-	TAACGGGATTGAGAGTGGACA	21	1810

	FP				
Tag Reverse Primer	AcDx-8287-TRIM15-S1-RT-RP	TTAGTAAGGGCGGCAGACA		19	1811

IDT Abbreviation	Modifications
/5Phos/	5' Phosphorylation
rX (X=A,C,G,U)	RNA Base
/3spC3/	3' C3 DNA Spacer
/56-FAM/	5' 6-FAM™ Fluorescent Tag
/Zen/	Internal Quencher
/3IABkFQ/	3' Iowa Black® FQ Quencher

Table 47. Simulation of 96-marker assay, with average sensitivities of 50%, for identifying most probably group for tissue of origin, for both sexes.

		CRC-Pt	ST-Pt	ES-Pt	BR-Pt	END-Pt	OV-Pt	CERV-Pt	UTCS-Pt	LUAD-Pt	LUSC-Pt	HNSC-Pt	PROS-Pt	BLAD-Pt	LIV-Pt	PANC-Pt	BILE-Pt
All	All Cancer	44	45	40	38	40	22	39	33	31	31	33	45	36	38	27	47
CRC1	Total	**66**	57	51	39	37	15	48	30	35	34	39	37	42	42	31	51
CRC2	Total	**66**	53	47	35	43	22	42	31	28	28	35	46	39	35	28	46
ST1	Total	56	**55**	46	36	34	17	40	26	37	31	33	39	37	46	31	53
ST2	Total	56	**54**	50	39	41	20	48	30	33	36	42	42	43	37	30	51
ES1	Total	57	54	**52**	38	47	21	51	38	38	40	42	36	38	28	26	45
ES2	Total	58	56	**50**	39	34	19	41	23	31	30	39	45	41	48	34	55
BR1	Total	47	49	47	**50**	47	26	45	35	33	35	41	50	41	38	30	51
BR2	Total	40	39	37	**49**	50	31	41	41	36	33	32	49	37	34	26	49
ENDO1	Total	44	49	50	50	**62**	41	48	54	37	43	44	50	43	34	29	53
ENDO2	Total	42	39	39	41	**61**	30	47	50	31	33	36	39	33	26	24	38
OV1	Total	35	41	43	58	68	**58**	49	59	38	40	38	49	36	23	27	44
OV2	Total	37	40	40	43	71	**56**	48	66	31	36	37	33	34	23	21	40
CERV1	Total	40	47	53	41	47	23	**57**	38	37	49	53	49	43	26	25	44
CERV2	Total	56	52	50	40	53	32	**57**	43	32	35	44	34	40	32	27	46
UTCS1	Total	25	30	33	47	66	39	42	**60**	31	34	32	53	34	27	21	37
UTCS2	Total	46	46	47	40	58	38	51	**59**	28	40	42	38	36	26	24	47
LUAD1	Total	50	55	53	47	43	24	49	31	**47**	44	44	42	41	37	30	54
LUAD2	Total	49	50	47	44	49	28	49	39	**46**	39	40	51	41	39	31	52
LUSC1	Total	46	50	55	40	48	30	51	41	40	**53**	52	40	42	24	24	45
LUSC2	Total	44	49	53	50	52	27	54	43	43	**51**	52	56	44	33	29	54
HNSC1	Total	47	54	58	44	49	28	55	39	39	49	**53**	43	50	34	29	52
HNSC2	Total	56	51	52	44	51	23	58	40	37	43	**52**	47	40	30	27	44
PROS1	Total	42	43	41	41	40	21	38	35	36	34	35	**63**	39	42	29	52
PROS2	Total	42	40	37	42	41	21	39	34	28	30	33	**62**	39	35	23	45
BLAD1	Total	48	47	45	38	42	20	47	36	35	39	39	53	**49**	31	25	48
BLAD2	Total	58	54	51	44	43	21	46	34	35	35	43	42	**48**	41	31	49
LIV1	Total	46	52	41	37	25	12	30	18	34	27	28	41	36	**61**	34	62

LIV2	Total	49	48	38	38	33	20	32	27	33	23	27	47	36	**60**	33	58
PANC1	Total	52	57	47	38	32	16	40	21	34	29	35	46	36	58	**41**	60
PANC2	Total	53	54	46	39	35	24	36	33	36	33	33	41	45	44	**40**	57
BILE1	Total	50	51	45	40	37	22	40	27	39	33	35	47	39	44	32	**58**
BILE2	Total	48	50	43	37	35	17	40	28	29	31	34	42	37	50	30	**57**

Table 48. Simulation of 96-marker assay, with average sensitivities of 50%, for identifying most probably group for tissue of origin, for male cancers.

		CRC-Pt	ST-Pt	ES-Pt	BR-Pt	END-Pt	OV-Pt	CERV-Pt	UTCS-Pt	LUAD-Pt	LUSC-Pt	HNSC-Pt	PROS-Pt	BLAD-Pt	LIV-Pt	PANC-Pt	BILE-Pt
All	Male Score	63	43	23	0	0	0	0	0	51	9	29	80	22	32	9	7
CRC1	Male Score	**94**	54	29	0	0	0	0	0	57	10	35	67	25	35	11	7
CRC2	Male Score	**93**	51	26	0	0	0	0	0	46	8	31	82	23	30	9	6
ST1	Male Score	79	**53**	26	0	0	0	0	0	61	9	30	70	22	38	10	7
ST2	Male Score	79	**52**	28	0	0	0	0	0	54	10	38	74	26	31	10	7
ES1	Male Score	80	52	**29**	0	0	0	0	0	62	11	37	64	23	24	9	6
ES2	Male Score	82	54	**28**	0	0	0	0	0	50	9	34	80	25	40	12	8
BR1	Male Score	66	47	26	0	0	0	0	0	54	10	36	89	25	32	10	7
BR2	Male Score	56	37	21	0	0	0	0	0	59	10	29	88	22	29	9	7
ENDO1	Male Score	62	47	28	0	0	0	0	0	60	13	40	89	26	28	10	7
ENDO2	Male Score	59	37	22	0	0	0	0	0	50	10	32	70	20	22	8	5
OV1	Male Score	50	39	24	0	0	0	0	0	62	12	34	87	22	19	9	6
OV2	Male Score	52	38	23	0	0	0	0	0	51	10	33	59	20	20	7	6
CERV1	Male Score	57	45	30	0	0	0	0	0	60	14	47	88	26	22	8	6
CERV2	Male Score	79	50	28	0	0	0	0	0	53	10	39	62	24	27	9	6
UTCS1	Male Score	36	29	19	0	0	0	0	0	50	10	29	96	20	23	7	5
UTCS2	Male Score	65	44	26	0	0	0	0	0	46	12	38	69	22	22	8	7
LUAD1	Male Score	71	53	30	0	0	0	0	0	**77**	13	40	75	25	31	10	8
LUAD2	Male Score	70	48	26	0	0	0	0	0	**74**	11	36	92	25	33	10	7
LUSC1	Male Score	65	48	31	0	0	0	0	0	65	**15**	47	71	25	20	8	6
LUSC2	Male Score	61	48	30	0	0	0	0	0	70	**15**	47	100	26	28	10	8
HNSC1	Male Score	67	52	33	0	0	0	0	0	63	14	**47**	77	30	28	10	7

HNSC2	Male Score	79	49	29	0	0	0	0	0	60	12	**46**	85	24	25	9	6
PROS1	Male Score	59	42	23	0	0	0	0	0	59	10	31	**113**	23	35	10	7
PROS2	Male Score	59	38	21	0	0	0	0	0	46	9	29	**111**	24	29	8	6
BLAD1	Male Score	67	46	25	0	0	0	0	0	57	11	35	96	**29**	26	9	7
BLAD2	Male Score	81	51	28	0	0	0	0	0	57	10	38	75	**29**	35	10	7
LIV1	Male Score	64	50	23	0	0	0	0	0	55	8	25	73	21	**51**	12	9
LIV2	Male Score	69	46	21	0	0	0	0	0	53	7	24	85	21	**50**	11	8
PANC1	Male Score	73	55	27	0	0	0	0	0	56	9	31	82	22	48	**14**	8
PANC2	Male Score	75	52	26	0	0	0	0	0	58	10	29	74	27	37	**14**	8
BILE1	Male Score	71	49	25	0	0	0	0	0	63	10	31	84	23	37	11	**8**
BILE2	Male Score	67	48	24	0	0	0	0	0	48	9	30	76	22	42	10	**8**

Table 49. Simulation of 96-marker assay, with average sensitivities of 50%, for identifying most probably group for tissue of origin, for female cancers.

		CRC-Pt	ST-Pt	ES-Pt	BR-Pt	END-Pt	OV-Pt	CERV-Pt	UTCS-Pt	LUAD-Pt	LUSC-Pt	HNSC-Pt	PROS-Pt	BLAD-Pt	LIV-Pt	PANC-Pt	BILE-Pt
All	Female Score	46	20	9	103	19	8	29	16	25	4	27	0	20	30	8	6
CRC1	Female Score	**68**	25	11	105	18	6	36	15	28	5	32	0	23	32	10	7
CRC2	Female Score	**68**	24	10	94	21	8	31	15	22	4	28	0	22	27	9	6
ST1	Female Score	58	**25**	10	98	17	7	29	13	29	4	27	0	20	35	9	7
ST2	Female Score	58	**24**	11	105	20	8	35	15	26	5	35	0	23	28	9	7
ES1	Female Score	58	24	**11**	101	23	8	38	19	30	6	34	0	21	22	8	6
ES2	Female Score	60	25	**11**	105	17	7	31	11	24	4	32	0	23	37	11	7
BR1	Female Score	48	22	10	**134**	23	10	34	17	26	5	33	0	23	30	9	7
BR2	Female Score	41	18	8	**132**	24	12	30	20	29	5	27	0	20	26	8	6
ENDO1	Female Score	46	22	11	134	**30**	16	36	27	29	6	36	0	24	26	9	7
ENDO2	Female Score	43	18	9	111	**30**	11	35	25	24	5	29	0	18	20	7	5
OV1	Female Score	36	18	9	155	34	**22**	36	29	30	6	32	0	20	18	8	6
OV2	Female Score	38	18	9	115	35	**21**	36	32	25	5	30	0	19	18	6	5
CERV1	Female Score	42	21	12	111	23	9	**42**	19	29	7	43	0	24	20	8	6
CERV2	Female Score	58	23	11	108	26	12	**42**	21	26	5	36	0	22	24	8	6

UTCS1	Female Score	26	14	7	125	33	15	31	**30**	24	5	27	0	19	21	6	5
UTCS2	Female Score	48	21	10	107	29	15	37	**29**	22	6	35	0	20	20	7	6
LUAD1	Female Score	52	25	12	127	21	9	36	15	**37**	6	36	0	23	29	9	7
LUAD2	Female Score	51	23	10	117	24	11	36	19	**36**	5	33	0	23	30	9	7
LUSC1	Female Score	48	22	12	108	23	11	38	20	32	**7**	43	0	23	18	7	6
LUSC2	Female Score	45	22	12	136	26	10	40	21	34	**7**	43	0	24	26	9	7
HNSC1	Female Score	49	25	13	117	24	11	41	19	31	7	**43**	0	27	26	9	7
HNSC2	Female Score	57	23	12	118	25	9	43	19	29	6	**43**	0	22	23	8	6
PROS1	Female Score	43	19	9	110	19	8	28	17	29	5	28	0	21	32	9	7
PROS2	Female Score	43	18	8	114	20	8	29	17	22	4	27	0	22	27	7	6
BLAD1	Female Score	49	21	10	103	21	8	35	18	28	5	32	0	**27**	24	8	6
BLAD2	Female Score	59	24	11	117	21	8	34	17	28	5	35	0	**26**	32	10	6
LIV1	Female Score	47	23	9	99	12	4	22	9	27	4	23	0	20	**47**	11	8
LIV2	Female Score	50	21	8	102	16	8	24	13	26	3	22	0	20	**46**	10	7
PANC1	Female Score	53	26	10	102	16	6	30	10	27	4	29	0	20	44	**13**	8
PANC2	Female Score	55	24	10	105	17	9	26	16	28	5	27	0	25	34	**12**	7
BILE1	Female Score	52	23	10	108	18	8	30	13	31	5	29	0	22	33	10	**8**
BILE2	Female Score	49	23	10	99	17	6	29	14	23	4	28	0	20	38	9	**7**

Table 50. Simulation of 96-marker assay, with average sensitivities of 50%, showing percent deviation from neutral result, for identifying most probably group for tissue of origin, for both sexes.

		CRC-Pt	ST-Pt	ES-Pt	BR-Pt	END-Pt	OV-Pt	CERV-Pt	UTCS-Pt	LUAD-Pt	LUSC-Pt	HNSC-Pt	PROS-Pt	BLAD-Pt	LIV-Pt	PANC-Pt	BILE-Pt
All	All Cancer	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
CRC1	Total	**50%**	27%	27%	2%	-6%	-31%	23%	-10%	12%	9%	17%	-16%	16%	9%	16%	8%
CRC2	Total	**49%**	19%	16%	-9%	9%	2%	8%	-6%	-9%	-8%	5%	3%	8%	-8%	3%	-3%
ST1	Total	26%	**23%**	14%	-6%	-14%	-22%	2%	-22%	19%	1%	0%	-13%	2%	20%	16%	12%
ST2	Total	26%	**21%**	24%	2%	3%	-8%	22%	-10%	6%	17%	27%	-6%	19%	-4%	12%	8%
ES1	Total	28%	21%	**29%**	-1%	18%	-4%	30%	14%	22%	30%	27%	-19%	5%	-27%	-3%	-5%
ES2	Total	31%	26%	**25%**	2%	-15%	-12%	5%	-30%	-1%	-2%	17%	0%	13%	24%	27%	17%

BR1	Total	6%	10%	16%	**30%**	18%	19%	15%	5%	6%	14%	24%	12%	13%	-1%	12%	8%
BR2	Total	-11%	-13%	-8%	**28%**	25%	44%	4%	24%	16%	8%	-2%	11%	1%	-10%	-3%	4%
ENDO1	Total	-1%	10%	24%	30%	**56%**	88%	22%	63%	19%	40%	33%	12%	19%	-11%	8%	12%
ENDO2	Total	-6%	-13%	-4%	7%	**53%**	37%	20%	52%	-1%	7%	8%	-13%	-8%	-31%	-12%	-21%
OV1	Total	-21%	-8%	6%	51%	71%	**166%**	25%	78%	22%	30%	15%	10%	-1%	-40%	1%	-7%
OV2	Total	-17%	-11%	0%	12%	79%	**157%**	22%	98%	0%	16%	12%	-27%	-7%	-39%	-22%	-15%
CERV1	Total	-9%	5%	32%	8%	18%	7%	**46%**	14%	18%	61%	59%	10%	18%	-32%	-8%	-7%
CERV2	Total	27%	16%	24%	4%	34%	46%	**44%**	29%	4%	13%	31%	-23%	9%	-17%	1%	-4%
UTCS1	Total	-44%	-33%	-18%	23%	66%	79%	7%	**81%**	0%	10%	-3%	19%	-6%	-30%	-22%	-22%
UTCS2	Total	4%	4%	16%	4%	47%	77%	29%	**78%**	-10%	31%	28%	-14%	0%	-33%	-10%	0%
LUAD1	Total	13%	23%	31%	23%	8%	10%	25%	-7%	**51%**	43%	33%	-6%	13%	-4%	12%	14%
LUAD2	Total	11%	13%	15%	14%	23%	28%	24%	16%	**47%**	27%	22%	15%	14%	2%	14%	11%
LUSC1	Total	4%	12%	36%	4%	21%	38%	30%	23%	29%	**72%**	57%	-10%	16%	-37%	-10%	-5%
LUSC2	Total	-2%	11%	31%	31%	31%	25%	38%	28%	38%	**67%**	58%	25%	21%	-13%	9%	14%
HNSC1	Total	6%	21%	44%	15%	23%	29%	40%	17%	25%	59%	**60%**	-4%	38%	-11%	8%	10%
HNSC2	Total	26%	15%	30%	14%	30%	7%	47%	19%	18%	39%	**57%**	6%	12%	-22%	2%	-8%
PROS1	Total	-6%	-3%	0%	7%	0%	-2%	-3%	5%	16%	12%	5%	**42%**	8%	9%	7%	9%
PROS2	Total	-7%	-10%	-7%	10%	4%	-5%	0%	1%	-9%	-1%	-2%	**39%**	8%	-9%	-14%	-5%
BLAD1	Total	7%	6%	10%	0%	6%	-6%	20%	9%	12%	27%	19%	20%	**36%**	-18%	-5%	2%
BLAD2	Total	29%	20%	26%	14%	8%	-2%	18%	4%	13%	14%	30%	-6%	**32%**	8%	15%	3%
LIV1	Total	4%	17%	2%	-4%	-37%	-45%	-24%	-46%	9%	-12%	-16%	-8%	-1%	**59%**	27%	31%
LIV2	Total	10%	7%	-7%	-1%	-17%	-9%	-19%	-19%	5%	-24%	-18%	6%	-2%	**56%**	22%	21%
PANC1	Total	16%	29%	18%	-1%	-19%	-27%	3%	-38%	11%	-4%	6%	3%	0%	50%	**54%**	27%
PANC2	Total	20%	21%	13%	1%	-12%	10%	-9%	-1%	15%	8%	0%	-7%	26%	16%	**50%**	19%
BILE1	Total	13%	14%	11%	5%	-7%	-1%	2%	-19%	24%	8%	6%	5%	8%	13%	20%	**23%**
BILE2	Total	8%	12%	7%	-4%	-13%	-24%	1%	-16%	-6%	-1%	2%	-5%	2%	29%	12%	**21%**

Table 51. Simulation of 96-marker assay, with average sensitivities of 50%, showing percent deviation from neutral result, for identifying most probably group for tissue of origin, for male cancers.

		CRC-Pt	ST-Pt	ES-Pt	BR-Pt	END-Pt	OV-Pt	CERV-Pt	UTCS-Pt	LUAD-Pt	LUSC-Pt	HNSC-Pt	PROS-Pt	BLAD-Pt	LIV-Pt	PANC-Pt	BILE-Pt
All	Male Score	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
CRC1	Male Score	**50%**	26%	28%	0%	0%	0%	0%	0%	13%	12%	19%	-16%	15%	9%	21%	6%
CRC2	Male Score	**49%**	19%	16%	0%	0%	0%	0%	0%	-9%	-8%	5%	3%	8%	-8%	3%	-3%
ST1	Male Score	26%	**24%**	15%	0%	0%	0%	0%	0%	20%	0%	1%	-12%	1%	19%	14%	12%
ST2	Male Score	27%	**22%**	24%	0%	0%	0%	0%	0%	7%	17%	28%	-7%	18%	-4%	13%	8%
ES1	Male Score	27%	21%	**29%**	0%	0%	0%	0%	0%	22%	29%	26%	-19%	6%	-26%	-2%	-4%
ES2	Male Score	31%	26%	**25%**	0%	0%	0%	0%	0%	-1%	-2%	17%	0%	13%	24%	27%	17%
BR1	Male Score	6%	11%	16%	0%	0%	0%	0%	0%	7%	13%	23%	12%	14%	0%	12%	7%
BR2	Male Score	-11%	-13%	-8%	0%	0%	0%	0%	0%	16%	8%	-2%	11%	1%	-10%	-3%	4%
ENDO1	Male Score	0%	9%	25%	0%	0%	0%	0%	0%	19%	41%	34%	11%	19%	-12%	10%	12%
ENDO2	Male Score	-6%	-13%	-4%	0%	0%	0%	0%	0%	-1%	7%	8%	-13%	-8%	-31%	-12%	-21%
OV1	Male Score	-21%	-9%	6%	0%	0%	0%	0%	0%	22%	31%	16%	9%	-1%	-40%	0%	-7%
OV2	Male Score	-17%	-11%	0%	0%	0%	0%	0%	0%	0%	16%	12%	-27%	-7%	-39%	-22%	-15%
CERV1	Male Score	-9%	5%	32%	0%	0%	0%	0%	0%	18%	61%	59%	10%	18%	-32%	-8%	-7%
CERV2	Male Score	27%	16%	24%	0%	0%	0%	0%	0%	4%	13%	31%	-23%	9%	-17%	1%	-4%
UTCS1	Male Score	-43%	-33%	-18%	0%	0%	0%	0%	0%	-2%	10%	-2%	20%	-6%	-28%	-23%	-23%
UTCS2	Male Score	4%	4%	16%	0%	0%	0%	0%	0%	-10%	31%	28%	-14%	0%	-33%	-10%	0%
LUAD1	Male Score	13%	24%	31%	0%	0%	0%	0%	0%	**51%**	43%	34%	-6%	14%	-2%	10%	14%
LUAD2	Male Score	11%	13%	15%	0%	0%	0%	0%	0%	**47%**	27%	22%	15%	14%	2%	14%	11%
LUSC1	Male Score	4%	12%	36%	0%	0%	0%	0%	0%	29%	**71%**	58%	-11%	15%	-38%	-11%	-4%
LUSC2	Male Score	-2%	11%	31%	0%	0%	0%	0%	0%	38%	**67%**	58%	25%	21%	-13%	9%	14%
HNSC1	Male Score	7%	22%	44%	0%	0%	0%	0%	0%	25%	59%	**59%**	-4%	38%	-12%	7%	11%
HNSC2	Male Score	26%	15%	30%	0%	0%	0%	0%	0%	18%	39%	**57%**	6%	12%	-22%	2%	-8%
PROS1	Male Score	-6%	-3%	0%	0%	0%	0%	0%	0%	16%	12%	5%	**42%**	8%	9%	7%	9%
PROS2	Male Score	-7%	-10%	-7%	0%	0%	0%	0%	0%	-9%	-1%	-2%	**39%**	8%	-9%	-14%	-5%
BLAD1	Male Score	7%	6%	10%	0%	0%	0%	0%	0%	12%	27%	19%	20%	**36%**	-18%	-5%	2%
BLAD2	Male Score	29%	20%	26%	0%	0%	0%	0%	0%	13%	14%	30%	-6%	**32%**	8%	15%	3%
LIV1	Male Score	3%	17%	1%	0%	0%	0%	0%	0%	9%	-11%	-16%	-9%	-1%	**59%**	27%	30%

LIV2	Male Score	10%	7%	-7%	0%	0%	0%	0%	0%	5%	-24%	-18%	6%	-2%	**56%**	22%	21%
PANC1	Male Score	16%	29%	18%	0%	0%	0%	0%	0%	11%	-4%	6%	3%	0%	50%	**54%**	27%
PANC2	Male Score	20%	21%	13%	0%	0%	0%	0%	0%	15%	8%	0%	-7%	26%	16%	**50%**	19%
BILE1	Male Score	13%	14%	11%	0%	0%	0%	0%	0%	24%	8%	6%	5%	8%	13%	20%	**23%**
BILE2	Male Score	8%	12%	7%	0%	0%	0%	0%	0%	-6%	-1%	2%	-5%	2%	29%	12%	**21%**

Table 52. Simulation of 96-marker assay, with average sensitivities of 50%, showing percent deviation from neutral result, for identifying most probably group for tissue of origin, for female cancers.

		CRC-Pt	ST-Pt	ES-Pt	BR-Pt	END-Pt	OV-Pt	CERV-Pt	UTCS-Pt	LUAD-Pt	LUSC-Pt	HNSC-Pt	PROS-Pt	BLAD-Pt	LIV-Pt	PANC-Pt	BILE-Pt
All	Female Score	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%	0%
CRC1	Female Score	**50%**	27%	27%	2%	-6%	-31%	23%	-10%	12%	9%	17%	0%	16%	9%	16%	8%
CRC2	Female Score	**49%**	19%	16%	-9%	9%	2%	8%	-6%	-9%	-8%	5%	0%	8%	-8%	3%	-3%
ST1	Female Score	26%	**24%**	15%	-5%	-13%	-20%	1%	-22%	20%	0%	1%	0%	1%	19%	14%	12%
ST2	Female Score	27%	**22%**	24%	2%	4%	-9%	22%	-9%	7%	17%	28%	0%	18%	-4%	13%	8%
ES1	Female Score	27%	21%	**29%**	-2%	19%	-3%	31%	14%	22%	29%	26%	0%	6%	-26%	-2%	-4%
ES2	Female Score	31%	26%	**25%**	2%	-15%	-12%	5%	-30%	-1%	-2%	17%	0%	13%	24%	27%	17%
BR1	Female Score	6%	11%	16%	**30%**	18%	17%	15%	6%	7%	13%	23%	0%	14%	0%	12%	7%
BR2	Female Score	-11%	-13%	-8%	**28%**	25%	44%	4%	24%	16%	8%	-2%	0%	1%	-10%	-3%	4%
ENDO1	Female Score	0%	9%	25%	30%	**56%**	89%	23%	63%	19%	41%	34%	0%	19%	-12%	10%	12%
ENDO2	Female Score	-6%	-13%	-4%	7%	**53%**	37%	20%	52%	-1%	7%	8%	0%	-8%	-31%	-12%	-21%
OV1	Female Score	-21%	-9%	6%	50%	72%	**168%**	24%	76%	22%	31%	16%	0%	-1%	-40%	0%	-7%
OV2	Female Score	-17%	-11%	0%	12%	79%	**157%**	22%	98%	0%	16%	12%	0%	-7%	-39%	-22%	-15%
CERV1	Female Score	-9%	5%	32%	8%	18%	7%	**46%**	14%	18%	61%	59%	0%	18%	-32%	-8%	-7%
CERV2	Female Score	27%	16%	24%	4%	34%	46%	**44%**	29%	4%	13%	31%	0%	9%	-17%	1%	-4%
UTCS1	Female Score	-43%	-33%	-18%	21%	67%	80%	7%	**82%**	-2%	10%	-2%	0%	-6%	-28%	-23%	-23%
UTCS2	Female Score	4%	4%	16%	4%	47%	77%	29%	**78%**	-10%	31%	28%	0%	0%	-33%	-10%	0%
LUAD1	Female Score	13%	24%	31%	23%	8%	11%	24%	-7%	**51%**	43%	34%	0%	14%	-2%	10%	14%
LUAD2	Female Score	11%	13%	15%	14%	23%	28%	24%	16%	**47%**	27%	22%	0%	14%	2%	14%	11%
LUSC1	Female Score	4%	12%	36%	4%	20%	37%	31%	22%	29%	**71%**	58%	0%	15%	-38%	-11%	-4%
LUSC2	Female Score	-2%	11%	31%	31%	31%	25%	38%	28%	38%	**67%**	58%	0%	21%	-13%	9%	14%

HNSC1	Female Score	7%	22%	44%	14%	23%	28%	40%	18%	25%	59%	**59%**	0%	38%	-12%	7%	11%
HNSC2	Female Score	26%	15%	30%	14%	30%	7%	47%	19%	18%	39%	**57%**	0%	12%	-22%	2%	-8%
PROS1	Female Score	-6%	-3%	0%	7%	0%	-2%	-3%	5%	16%	12%	5%	0%	8%	9%	7%	9%
PROS2	Female Score	-7%	-10%	-7%	10%	4%	-5%	0%	1%	-9%	-1%	-2%	0%	8%	-9%	-14%	-5%
BLAD1	Female Score	7%	6%	10%	0%	6%	-6%	20%	9%	12%	27%	19%	0%	**36%**	-18%	-5%	2%
BLAD2	Female Score	29%	20%	26%	14%	8%	-2%	18%	4%	13%	14%	30%	0%	**32%**	8%	15%	3%
LIV1	Female Score	3%	17%	1%	-4%	-36%	-46%	-24%	-47%	9%	-11%	-16%	0%	-1%	**59%**	27%	30%
LIV2	Female Score	10%	7%	-7%	-1%	-17%	-9%	-19%	-19%	5%	-24%	-18%	0%	-2%	**56%**	22%	21%
PANC1	Female Score	16%	29%	18%	-1%	-19%	-27%	3%	-38%	11%	-4%	6%	0%	0%	50%	**54%**	27%
PANC2	Female Score	20%	21%	13%	1%	-12%	10%	-9%	-1%	15%	8%	0%	0%	26%	16%	**50%**	19%
BILE1	Female Score	13%	14%	11%	5%	-7%	-1%	2%	-19%	24%	8%	6%	0%	8%	13%	20%	**23%**
BILE2	Female Score	8%	12%	7%	-4%	-13%	-24%	1%	-16%	-6%	-1%	2%	0%	2%	29%	12%	**21%**

Table 53. Primers for use in Step 2 of the Group 1- 64-marker assay, with average sensitivities of 50%, to detect and identify colorectal, stomach, and esophageal cancers.

Site	Primer Name	Sequence	Length	Seq. ID No.
Prefered Group 1 Markers				
ISL2-S1				
Forward PCR Primer	AcDx-7141-ISL2-S1-FP	GTGGTCGTCGTTTTGTTTTTTTACrGTTTC/3SpC3/	29	1812
Reverse PCR Primer	AcDx-7142-ISL2-S1-RP	GGTGTCGTGGCCCGAACGAAACCGCCrAAAAG/3SpC3/	31	1813
Upstream LDR	AcDx-7143-ISL2-S1-Up	TCCGGATCAAAGCAGCCACTTTTATTTCGGTCGGTTCGGAGCTCrGTTCG/3SpC3/	49	1814
Downstream LDR	AcDx-7144-ISL2-S1-Dn	/5Phos/GTTTACGCGGATTTTCGTTTTGTTTTAGTTTAGCGTGTGTGTTGGCGTACGGTGA	55	1815
Real-Time Probe	AcDx-7145-ISL2-S1-RT-Pb	/56-FAM/AAGGAGCTC/ZEN/GTTTACGCGGATTTTCGTTTTG/3IABkFQ/	31	1816
Tag Forward Primer	AcDx-7146-ISL2-S1-RT-FP	TCCGGATCAAAGCAGCCAC	19	1817
Tag Reverse Primer	AcDx-7147-ISL2-S1-RT-RP	TCACCGTACGCCAACACAC	19	1818
Downstream PCR	AcDx-7148-ISL2-S1-PCR-V	TCACCGTACGCCAACACACCGAACGAAACCGCCAAAAAATGrCTAAG/3SpC3/	46	1819

Primer				
HOXD8-S1				
Forward PCR Primer	AcDx-7251-HOXD8-S1-FP	TTTAGAGTCGAGGTTTGTAAATCrGAGGC/3SpC3/	28	1820
Reverse PCR Primer	AcDx-7252-HOXD8-S1-RP	GGTGTCGTGGACGACCTACCCCGCTACrCTCCG/3SpC3/	32	1821
Upstream LDR	AcDx-7253-HOXD8-S1-Up	TCACTATCGGCGTAGTCACCAGTTAGAGTGTTTTCGTGGGTCGGGCrGTACC/3SpC3/	51	1822
Downstream LDR	AcDx-7254-HOXD8-S1-Dn	/5Phos/GTATTTTTTTTTTGTTCGGGTGCGTTTAGTTATTGGTGTGGTGACTTTACCCG GAGGA	58	1823
Real-Time Probe	AcDx-7255-HOXD8-S1-RT-Pb	/56-FAM/TTGTCGGGC/ZEN/GTATTTTTTTTTTGTTCGGGTG/3IABkFQ/	31	1824
Tag Forward Primer	AcDx-7256-HOXD8-S1-RT-FP	TCACTATCGGCGTAGTCACCA	21	1825
Tag Reverse Primer	AcDx-7257-HOXD8-S1-RT-RP	TCCTCCGGGTAAAGTCACCA	20	1826
Downstream PCR Primer	AcDx-7258-HOXD8-S1-PCR-V	TCCTCCGGGTAAAGTCACCAACTATTTCCTCTCAAACACCAATAACTAAATGrCACCT/ 3SpC3/	57	1827
PTGDR-S1				
Forward PCR Primer	AcDx-8051-PTGDR-S1-FP	GTAATTGTGAGTTTTCGGGTTTCrGAGGC/3SpC3/	28	1828
Reverse PCR Primer	AcDx-8052-PTGDR-S1-RP	GGTGTCGTGGCCATCCCGATCCGCTCrCACCT/3SpC3/	31	1829
Upstream LDR	AcDx-8053-PTGDR-S1-Up	TGGCACATGAGAGTAGTTGACCGTTTCGAGGTAGTAGGGTATTGAGATTGCTCrGGT TA/3SpC3/	58	1830
Downstream LDR	AcDx-8054-PTGDR-S1-Dn	/5Phos/GGTCGCGGATGCGGAGCGGTTCGATGCCTTCCGTACA	37	1831
Real-Time Probe	AcDx-8055-PTGDR-S1-RT-Pb	/56-FAM/TTATTGCTC/ZEN/GGTCGCGGATGCG/3IABkFQ/	22	1832
Tag Forward Primer	AcDx-8056-PTGDR-S1-RT-FP	TGGCACATGAGAGTAGTTGACC	22	1833
Tag Reverse Primer	AcDx-8057-PTGDR-S1-RT-RP	TGTACGGAAGGCATCGAACC	20	1834
Downstream PCR	AcDx-8058-PTGDR-S1-PCR-	TGTACGGAAGGCATCGAACCCCGCTCCACCCTTCCTGrCCCCA/3SpC3/	42	1835

Primer	V			
LIFR				
Forward PCR Primer	AcDx-8291-LIFR-FP	GTTTCGCGTCGCGTTTATTCrGTTTC/3SpC3/	25	1836
Reverse PCR Primer	AcDx-8292-LIFR-RP	GGTGTCGTGGCCGAAACGACGACCGAAACrUACAG/3SpC3/	34	1837
Upstream LDR	AcDx-8293-LIFR-Up	TCCGGCCTTTGACGATACCGCGTCGCGTTTATTCGTTTTTAGGATTCACrGGTGT/3SpC3/	54	1838
Downstream LDR	AcDx-8294-LIFR-Dn	/5Phos/GGTACGTTTTCGCGTCGTTATTTTGTTATTTTTTGTCGGGTAATTCACTCGAACGGAGCA	60	1839
Real-Time Probe	AcDx-8295-LIFR-RT-Pb	/56-FAM/TTGATTCAC/ZEN/GGTACGTTTTCGCGTCGT/3IABkFQ/	27	1840
Tag Forward Primer	AcDx-8296-LIFR-RT-FP	TCCGGCCTTTGACGATACC	19	1841
Tag Reverse Primer	AcDx-8297-LIFR-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	1842
Downstream PCR Primer	AcDx-8298-LIFR-PCR-V	TGCTCCGTTCGAGTGAATTACCCGACCGAAACTACAAAACCGATGrACAAG/3SpC3/	50	1843
ZNF304-S1				
Forward PCR Primer	AcDx-8301-ZNF304-S1-FP	GTTTTGGGTTGTAGAGGCrGAGAC/3SpC3/	23	1844
Reverse PCR Primer	AcDx-8302-ZNF304-S1-RP	GGTGTCGTGGCGACCCGATCTCTAAAACTCArACCTC/3SpC3/	36	1845
Upstream LDR	AcDx-8303-ZNF304-S1-Up	TCAGACGCACTAAACAGGCAATAGAGGCGAGATTTTTGGCATCrGTCAC/3SpC3/	48	1846
Downstream LDR	AcDx-8304-ZNF304-S1-Dn	/5Phos/GTCGTCGTGACGTATTTTTTTTATGTTCGGTTCGTGTTGCGGATCGTCGTGTGAA	55	1847
Real-Time Probe	AcDx-8305-ZNF304-S1-RT-Pb	/56-FAM/AATGGCATC/ZEN/GTCGTCGTGACGTATTTTTTTTATGTTC/3IABkFQ/	37	1848
Tag Forward Primer	AcDx-8306-ZNF304-S1-RT-FP	TCAGACGCACTAAACAGGCAA	21	1849
Tag Reverse Primer	AcDx-8307-ZNF304-S1-RT-RP	TTCACACGACGATCCGCAA	19	1850
Downstream PCR Primer	AcDx-8308-ZNF304-S1-PCR-V	TTCACACGACGATCCGCAAACTCAACCTTCACAACCAAAATACATGrAACCA/3SpC3/	51	1851

KRBA1				
Forward PCR Primer	AcDx-8311-KRBA1-FP	GGGATTTTTTCGTAATTTCGGGATCrGGTTA/3SpC3/	30	1852
Reverse PCR Primer	AcDx-8312-KRBA1-RP	GGTGTCGTGGAATAAAAACTAAACCGTAAATACCGArCGACA/3SpC3/	41	1853
Upstream LDR	AcDx-8313-KRBA1-Up	TTCCATCGAGCGCCAACAAGTAGGTTTGATGTTTAGAGGTTCGTATCrGGGCG/3SpC3/	52	1854
Downstream LDR	AcDx-8314-KRBA1-Dn	/5Phos/GGGAAGTTTAGGTTTTCGGATAATTTTTTTTTAGTTTTTCGTCTTGCGGATCGTCGTGTGAA	62	1855
Real-Time Probe	AcDx-8315-KRBA1-RT-Pb	/56-FAM/AATCGTATC/ZEN/GGGAAGTTTAGGTTTTCGGATAATTTTTTTTTAG/3IABkFQ/	43	1856
Tag Forward Primer	AcDx-8316-KRBA1-RT-FP	TTCCATCGAGCGCCAACAA	19	1857
Tag Reverse Primer	AcDx-8317-KRBA1-RT-RP	TTCACACGACGATCCGCAA	19	1858
Downstream PCR Primer	AcDx-8318-KRBA1-PCR-V	TTCACACGACGATCCGCAATAAAAACTAAACCGTAAATACCGACGATGrAAAAG/3SpC3/	53	1859
GATM				
Forward PCR Primer	AcDx-8321-GATM-FP	ATTAGAAGGGATATTTTCGTGTAATTTCrGGTTG/3SpC3/	33	1860
Reverse PCR Primer	AcDx-8322-GATM-RP	GGTGTCGTGGGTAAAACACTCGCTCGCTCrCCTAT/3SpC3/	34	1861
Upstream LDR	AcDx-8323-GATM-Up	TTCAGAGCACCTGCGTACCGATATTTTCGTGTAATTTCGGTTAGGGAAGGCrGAGAA/3SpC3/	56	1862
Downstream LDR	AcDx-8324-GATM-Dn	/5Phos/GAGGGAAGGTGGCGGTTTCGGGGGTTCTTCGGCTGGCTCAA	41	1863
Real-Time Probe	AcDx-8325-GATM-RT-Pb	/56-FAM/TTGGAAGGC/ZEN/GAGGGAAGGTGGC/3IABkFQ/	22	1864
Tag Forward Primer	AcDx-8326-GATM-RT-FP	TTCAGAGCACCTGCGTACC	19	1865
Tag Reverse Primer	AcDx-8327-GATM-RT-RP	TTGAGCCAGCCGAAGAACC	19	1866
Downstream PCR Primer	AcDx-8328-GATM-PCR-V	TTGAGCCAGCCGAAGAACCTCGCTCGCTCCCTACCTGrAAACT/3SpC3/	42	1867
DFNB31				

Forward PCR Primer	AcDx-8331-DFNB31-FP	TTGGATTTTAGGAGGTCGCrGGAGG/3SpC3/	24	1868
Reverse PCR Primer	AcDx-8332-DFNB31-RP	GGTGTCGTGGCGAATTCCAAATTCGCGACCrCCCGT/3SpC3/	35	1869
Upstream LDR	AcDx-8333-DFNB31-Up	TCTCAATGTCGAGCCGTACCGGATTTTAGGAGGTCGCGGAGGTCrGTTAC/3SpC3/	49	1870
Downstream LDR	AcDx-8334-DFNB31-Dn	/5Phos/GTTGTTAGAGTTTCGGAGGCGCGAAGAGGTTCTTCGGCTGGCTCAA	46	1871
Real-Time Probe	AcDx-8335-DFNB31-RT-Pb	/56-FAM/TTGGAGGTC/ZEN/GTTGTTAGAGTTTCGGAGGC/3IABkFQ/	29	1872
Tag Forward Primer	AcDx-8336-DFNB31-RT-FP	TCTCAATGTCGAGCCGTACC	20	1873
Tag Reverse Primer	AcDx-8337-DFNB31-RT-RP	TTGAGCCAGCCGAAGAACC	19	1874
Downstream PCR Primer	AcDx-8338-DFNB31-PCR-V	TTGAGCCAGCCGAAGAACCACCCCCGCCGTCTTTGrCGCCC/3SpC3/	40	1875
LONRF2				
Forward PCR Primer	AcDx-8341-LONRF2-FP	GTTAGTATGGAGCGAAAGAGTTCrGGTTA/3SpC3/	28	1876
Reverse PCR Primer	AcDx-8342-LONRF2-RP	GGTGTCGTGGCCTAACTACGACCGCGCrGAAAT/3SpC3/	32	1877
Upstream LDR	AcDx-8343-LONRF2-Up	TTTCGCTCGACGCATACCAGCGAAAGAGTTCGGTTGTTATTTCGTAATCrGTTTA/3SpC3/	54	1878
Downstream LDR	AcDx-8344-LONRF2-Dn	/5Phos/GTTCGCGCGGAAGGTTTCGTCGTTGGCGCGGCTACTGTAAAA	42	1879
Real-Time Probe	AcDx-8345-LONRF2-RT-Pb	/56-FAM/AACGTAATC/ZEN/GTTCGCGCGGAAGGTTTC/3IABkFQ/	27	1880
Tag Forward Primer	AcDx-8346-LONRF2-RT-FP	TTTCGCTCGACGCATACCA	19	1881
Tag Reverse Primer	AcDx-8347-LONRF2-RT-RP	TTTTACAGTAGCCGCGCCA	19	1882
Downstream PCR Primer	AcDx-8348-LONRF2-PCR-V	TTTTACAGTAGCCGCGCCAACCGCGCGAAACCGATTGrCCCAG/3SpC3/	42	1883
ST3GAL4-S1				
Forward PCR Primer	AcDx-8351-ST3GAL4-S1-FP	GAGTAAGCGGGATAGTTTTGCrGGAAG/3SpC3/	26	1884
Reverse PCR Primer	AcDx-8352-ST3GAL4-S1-RP	GGTGTCGTGGAAAAACCATCGCTCGAAAATACrUAAAG/3SpC3/	37	1885
Upstream LDR	AcDx-8353-ST3GAL4-S1-Up	TTCTTGCCCGCTTGTTCCAGCGGGATAGTTTTGCGGAAAGTTCTCrGTTCC/3SpC3/	50	1886
Downstream LDR	AcDx-8354-ST3GAL4-S1-Dn	/5Phos/GTTTTTAATTTTTTAGTTTTGCGTTCGGATTGAAGCGGCTGGCGCGGCTACTGTAAAA	58	1887
Real-Time Probe	AcDx-8355-ST3GAL4-S1-	/56-FAM/TTAGTTCTC/ZEN/GTTTTTAATTTTTTAGTTTTGCGTTCGGA/3IABkFQ/	38	1888

	RT-Pb			
Tag Forward Primer	AcDx-8356-ST3GAL4-S1-RT-FP	TTCTTGCCCGCTTGTTCCA	19	1889
Tag Reverse Primer	AcDx-8357-ST3GAL4-S1-RT-RP	TTTTACAGTAGCCGCGCCA	19	1890
Downstream PCR Primer	AcDx-8358-ST3GAL4-S1-PCR-V	TTTTACAGTAGCCGCGCCAGCTCGAAAATACTAAAAATAAAAACCGCTGrCTTCG/3SpC3/	54	1891
CLIP4				
Forward PCR Primer	AcDx-8361-CLIP4-FP	GAGGTGTGGAGCGGCrGCGGT/3SpC3/	20	1892
Reverse PCR Primer	AcDx-8362-CLIP4-RP	GGTGTCGTGGAAAACCGAAAAACGCGCTCrCCGCT/3SpC3/	34	1893
Upstream LDR	AcDx-8363-CLIP4-Up	TCCCTTAGAGAGAACGCCCAGCGGGAGGTTTCGTGAGCrGGTCG/3SpC3/	43	1894
Downstream LDR	AcDx-8364-CLIP4-Dn	/5Phos/GGTTACGGGAGATAGCGTCGGCGTGGTGACGTACGAGTGTTCTTA	45	1895
Real-Time Probe	AcDx-8365-CLIP4-RT-Pb	/56-FAM/AACGTGAGC/ZEN/GGTTACGGGAGATAGCG/3IABkFQ/	26	1896
Tag Forward Primer	AcDx-8366-CLIP4-RT-FP	TCCCTTAGAGAGAACGCCCA	20	1897
Tag Reverse Primer	AcDx-8367-CLIP4-RT-RP	TAAGAACACTCGTACGTCACCA	22	1898
Downstream PCR Primer	AcDx-8368-CLIP4-PCR-V	TAAGAACACTCGTACGTCACCAAACGCGCTCCCGCTGrACGCC/3SpC3/	42	1899
THBD-S1				
Forward PCR Primer	AcDx-8371-THBD-S1-FP	GGAGTTTTATTGAGGTCGAGTCrGTGTC/3SpC3/	27	1900
Reverse PCR Primer	AcDx-8372-THBD-S1-RP	GGTGTCGTGGCAACTACCACCCGACTACGArCGACT/3SpC3/	35	1901
Upstream LDR	AcDx-8373-THBD-S1-Up	TCTCATACCAGACGCGGTAACGGTGTTGTTGTTTTTCGTAATTTATTGGAAATCrGCGCG/3SpC3/	59	1902
Downstream LDR	AcDx-8374-THBD-S1-Dn	/5Phos/GCGTAGGAGTTCGAGGCGTTTGGTTCGTGTCGCTGTGCTTA	41	1903
Real-Time Probe	AcDx-8375-THBD-S1-RT-Pb	/56-FAM/AAGGAAATC/ZEN/GCGTAGGAGTTCGAGGC/3IABkFQ/	26	1904
Tag Forward Primer	AcDx-8376-THBD-S1-RT-FP	TCTCATACCAGACGCGGTAAC	21	1905
Tag Reverse Primer	AcDx-8377-THBD-S1-RT-RP	TAAGCACAGCGACACGAAC	19	1906
Downstream PCR	AcDx-8378-THBD-S1-PCR-V	TAAGCACAGCGACACGAACCAACTACCACCCGACTACGATGrGCCTT/3SpC3/	46	1907

Primer				
CANCR55				
Forward PCR Primer	AcDx-8381-CANCR55-FP	GGTTTTGGTGGAGGTTGCrGGTTA/3SpC3/	23	1908
Reverse PCR Primer	AcDx-8382-CANCR55-RP	GGTGTCGTGGTAAACGCCTCTACTCCGAAATCrGAAAT/3SpC3/	37	1909
Upstream LDR	AcDx-8383-CANCR55-Up	TTTTCGGCGGCAGCTAAACGGTGGAGGTTGCGGTTGTCTCrGGTCT/3SpC3/	45	1910
Downstream LDR	AcDx-8384-CANCR55-Dn	/5Phos/GGTTCGCGGGAGCGGGGTTCGTGTCGCTGTGCTTA	35	1911
Real-Time Probe	AcDx-8385-CANCR55-RT-Pb	/56-FAM/CCTTGTCTC/ZEN/GGTTCGCGGGAGC/3IABkFQ/	22	1912
Tag Forward Primer	AcDx-8386-CANCR55-RT-FP	TTTTCGGCGGCAGCTAAAC	19	1913
Tag Reverse Primer	AcDx-8387-CANCR55-RT-RP	TAAGCACAGCGACACGAAC	19	1914
Downstream PCR Primer	AcDx-8388-CANCR55-PCR-V	TAAGCACAGCGACACGAACTAAACGCCTCTACTCCGAAATTGrAAACT/3SpC3/	47	1915
SLFN11				
Forward PCR Primer	AcDx-8391-SLFN11-FP	GTTGAGGTAGGTTTCGTAGATTTCrGAGTC/3SpC3/	29	1916
Reverse PCR Primer	AcDx-8392-SLFN11-RP	GGTGTCGTGGTTTTCATATCACTAACAACGTAACGArUTACT/3SpC3/	41	1917
Upstream LDR	AcDx-8393-SLFN11-Up	TCCGGACCTTCATCCTCCATTCGAGTTAGAGTGGGATTTAACAGCrGGTTA/3SpC3/	50	1918
Downstream LDR	AcDx-8394-SLFN11-Dn	/5Phos/GGTCGGTGCGTTGTGTTTGGTTAATTTCGGTGGGCAGGAACACGATAGTA	50	1919
Real-Time Probe	AcDx-8395-SLFN11-RT-Pb	/56-FAM/AATAACAGC/ZEN/GGTCGGTGCGTTGTGT/3IABkFQ/	25	1920
Tag Forward Primer	AcDx-8396-SLFN11-RT-FP	TCCGGACCTTCATCCTCCA	19	1921
Tag Reverse Primer	AcDx-8397-SLFN11-RT-RP	TACTATCGTGTTCCTGCCCA	20	1922
Downstream PCR Primer	AcDx-8398-SLFN11-PCR-V	TACTATCGTGTTCCTGCCCATTTTCATATCACTAACAACGTAACGATTACTGrAAATC/3SpC3/	57	1923
CANCR56				

Forward PCR Primer	AcDx-8401-CANCR56-FP	GTTAGGGTAGTTCGGTCGATTTCrGAGAA/3SpC3/	28	1924
Reverse PCR Primer	AcDx-8402-CANCR56-RP	GGTGTCGTGGTCTCCCGCGCCCAAArCCCCA/3SpC3/	30	1925
Upstream LDR	AcDx-8403-CANCR56-Up	TTAGCCGCCAAACGTACCAGTTCGGTCGATTTCGAGAGGTTCTCrGCGCC/3SpC3/	49	1926
Downstream LDR	AcDx-8404-CANCR56-Dn	/5Phos/GCGTTTTTTTTTCGTTTTGTTTAGGAGTGGAGTGTGGGCAGGAACACGATAGTA	54	1927
Real-Time Probe	AcDx-8405-CANCR56-RT-Pb	/56-FAM/TTGGTTCTC/ZEN/GCGTTTTTTTTTCGTTTTGTTTAGGAG/3IABkFQ/	36	1928
Tag Forward Primer	AcDx-8406-CANCR56-RT-FP	TTAGCCGCCAAACGTACCA	19	1929
Tag Reverse Primer	AcDx-8407-CANCR56-RT-RP	TACTATCGTGTTCCTGCCCA	20	1930
Downstream PCR Primer	AcDx-8408-CANCR56-PCR-V	TACTATCGTGTTCCTGCCCACGCGCCCAAACCCTGrACACT/3SpC3/	40	1931
CANCR57				
Forward PCR Primer	AcDx-8411-CANCR57-FP	CGTAGTTTTCGAGAGGTTTCrGGTTG/3SpC3/	25	1932
Reverse PCR Primer	AcDx-8412-CANCR57-RP	GGTGTCGTGGAATACTCTCTACCTTCGCCCrCCAAG/3SpC3/	35	1933
Upstream LDR	AcDx-8413-CANCR57-Up	TAGCAGCTGAACAACCCAACGGGTTTCGGTTAGGGCGTTTTTTACACrGTTTA/3SpC3/	52	1934
Downstream LDR	AcDx-8414-CANCR57-Dn	/5Phos/GTTCGGTTATTTGTAGGAGTTCGTTATTGTAAGTCGGTTGTATGGTCGGCATGCTA	56	1935
Real-Time Probe	AcDx-8415-CANCR57-RT-Pb	/56-FAM/AATTTACAC/ZEN/GTTCGGTTATTTGTAGGAGTTCGTTATTG/3IABkFQ/	38	1936
Tag Forward Primer	AcDx-8416-CANCR57-RT-FP	TAGCAGCTGAACAACCCAAC	20	1937
Tag Reverse Primer	AcDx-8417-CANCR57-RT-RP	TAGCATGCCGACCATACAAC	20	1938
Downstream PCR Primer	AcDx-8418-CANCR57-PCR-V	TAGCATGCCGACCATACAACGCCCCCAAACTTCTACTAACCTGrACTTG/3SpC3/	48	1939
CANCR58				

Forward PCR Primer	AcDx-8421-CANCR58-FP	GTTCGCGTTTTTTTATCGTTTTTTTCrGATTC/3SpC3/	31	1940
Reverse PCR Primer	AcDx-8422-CANCR58-RP	GGTGTCGTGGCGCCTCCCTCTACCACCrGATAT/3SpC3/	32	1941
Upstream LDR	AcDx-8423-CANCR58-Up	TCTGCCCTTCGCTTCGAACTCGTTTTTTTCGATTTTTTTAAGTTTTGAGCrGGAGC/3SpC3/	55	1942
Downstream LDR	AcDx-8424-CANCR58-Dn	/5Phos/GGAATTTGATTTTGTTTTATCGGTTTTAGGTATTTTAGGATTTGAGGTTGTATGGTCGGCATGCTA	66	1943
Real-Time Probe	AcDx-8425-CANCR58-RT-Pb	/56-FAM/AATTTGAGC/ZEN/GGAATTTGATTTTGTTTTATCGGTTTTAGGTATTTTAG/3IABkFQ/	47	1944
Tag Forward Primer	AcDx-8426-CANCR58-RT-FP	TCTGCCCTTCGCTTCGAAC	19	1945
Tag Reverse Primer	AcDx-8427-CANCR58-RT-RP	TAGCATGCCGACCATACAAC	20	1946
Downstream PCR Primer	AcDx-8428-CANCR58-PCR-V	TAGCATGCCGACCATACAACCGCCTCCCTCTACCACTGrATACT/3SpC3/	43	1947
GHR				
Forward PCR Primer	AcDx-8431-GHR-FP	TGTTATAGTGGCGGTGGCrGGCGA/3SpC3/	23	1948
Reverse PCR Primer	AcDx-8432-GHR-RP	GGTGTCGTGGCTCCGACCCGCGTAArCCCCG/3SpC3/	30	1949
Upstream LDR	AcDx-8433-GHR-Up	TTGCAAACCACCCGGACAAGCGGCGGTTGTTGTTGAGCTCrGGGTA/3SpC3/	45	1950
Downstream LDR	AcDx-8434-GHR-Dn	/5Phos/GGGCGGCGGCGGAGATTTGGTCAGCATCGACTCCTA	36	1951
Real-Time Probe	AcDx-8435-GHR-RT-Pb	/56-FAM/AATGAGCTC/ZEN/GGGCGGCGG/3IABkFQ/	18	1952
Tag Forward Primer	AcDx-8436-GHR-RT-FP	TTGCAAACCACCCGGACAA	19	1953
Tag Reverse Primer	AcDx-8437-GHR-RT-RP	TAGGAGTCGATGCTGACCAA	20	1954
Downstream PCR Primer	AcDx-8438-GHR-PCR-V	TAGGAGTCGATGCTGACCAACCCCAACCCGAAATCCCTGrCCGCT/3SpC3/	44	1955
P2RY1				
Forward PCR Primer	AcDx-8441-P2RY1-FP	GTTCGTTTTGGAGGAATAGTACrGGTCA/3SpC3/	27	1956
Reverse PCR Primer	AcDx-8442-P2RY1-RP	GGTGTCGTGGTATAAACAACCGACAAATAATAAAACTAAArAACCT/3SpC3/	45	1957

Upstream LDR	AcDx-8443-P2RY1-Up	TTGCAAACCACCCGGACAAGGAGAATAGTACGGTCGTTTTTATTATCrGTCAC/3SpC3/	52	1958
Downstream LDR	AcDx-8444-P2RY1-Dn	/5Phos/GTCGTTTTTTCGTCGTTTAAATGCGTTTTGATTAAGACTTGGTCAGCATCGACTCCTA	58	1959
Real-Time Probe	AcDx-8445-P2RY1-RT-Pb	/56-FAM/AATATTATC/ZEN/GTCGTTTTTTCGTCGTTTAAATGCGTTTT/3IABkFQ/	38	1960
Tag Forward Primer	AcDx-8446-P2RY1-RT-FP	TTGCAAACCACCCGGACAA	19	1961
Tag Reverse Primer	AcDx-8447-P2RY1-RT-RP	TAGGAGTCGATGCTGACCAA	20	1962
Downstream PCR Primer	AcDx-8448-P2RY1-PCR-V	TAGGAGTCGATGCTGACCAAAAACAACCGACAAATAATAAAACTAAAAACCTGrUCTTG/3SpC3/	58	1963
CANCR59				
Forward PCR Primer	AcDx-8451-CANCR59-FP	GTAGAAAGATTAAGCGCGGTATTTCrGTGAC/3SpC3/	30	1964
Reverse PCR Primer	AcDx-8452-CANCR59-RP	GGTGTCGTGGCGCTTACTCTATAATTTTCCCGCrACCAG/3SpC3/	38	1965
Upstream LDR	AcDx-8453-CANCR59-Up	TAAGACGTATGCTAGCGCCAACGGTATTTCGTGATAGGATGGGATAGATTTCTCrGGGCC/3SpC3/	59	1966
Downstream LDR	AcDx-8454-CANCR59-Dn	/5Phos/GGGTTTCGAGGTTTTAGGATGGCGTAGGGTTGGTCAGCATCGACTCCTA	49	1967
Real-Time Probe	AcDx-8455-CANCR59-RT-Pb	/56-FAM/TTATTTCTC/ZEN/GGGTTTCGAGGTTTTAGGATGGC/3IABkFQ/	32	1968
Tag Forward Primer	AcDx-8456-CANCR59-RT-FP	TAAGACGTATGCTAGCGCCAA	21	1969
Tag Reverse Primer	AcDx-8457-CANCR59-RT-RP	TAGGAGTCGATGCTGACCAA	20	1970
Downstream PCR Primer	AcDx-8458-CANCR59-PCR-V	TAGGAGTCGATGCTGACCAACTATAATTTTCCCGCACCAACCTGrCTACT/3SpC3/	49	1971
OPRD1				
Forward PCR Primer	AcDx-8461-OPRD1-FP	GGTTTTTTTCGTCGGCGTCrGAGTC/3SpC3/	24	1972
Reverse PCR Primer	AcDx-8462-OPRD1-RP	GGTGTCGTGGCGCATTAACGCCAACGCrUAAAG/3SpC3/	32	1973
Upstream LDR	AcDx-8463-OPRD1-Up	TACATGCCATCCCACGACACGTCGGCGTCGAGTTGTAGTTCTCrGTTCC/3SpC3/	48	1974
Downstream LDR	AcDx-8464-OPRD1-Dn	/5Phos/GTTTTTCGTTAACGTTTCGGACGTTTATTTTAGCGTTTGTGTGTCGGAGCGG	58	1975

		TTACTA		
Real-Time Probe	AcDx-8465-OPRD1-RT-Pb	/56-FAM/AAAGTTCTC/ZEN/GTTTTTCGTTAACGTTTCGGACGTTTATTTTA/3IABkFQ/	41	1976
Tag Forward Primer	AcDx-8466-OPRD1-RT-FP	TACATGCCATCCCACGACA	19	1977
Tag Reverse Primer	AcDx-8467-OPRD1-RT-RP	TAGTAACCGCTCCGACACA	19	1978
Downstream PCR Primer	AcDx-8468-OPRD1-PCR-V	TAGTAACCGCTCCGACACACATTAACGCCAACGCTAAAACAAATGrCTAAG/3SpC3/	50	1979
APC-S1				
Forward PCR Primer	AcDx-8471-APC-S1-FP	TGGAGATTGAGGTCGCrGAGGA/3SpC3/	21	1980
Reverse PCR Primer	AcDx-8472-APC-S1-RP	GGTGTCGTGGCAAAACCCCGCCCAACCrGCACG/3SpC3/	32	1981
Upstream LDR	AcDx-8473-APC-S1-Up	TCCAAACAAGCTGATCCGTACAATTGGAGTCGCGAGGGTATATTCTCrGAGAA/3SpC3/	52	1982
Downstream LDR	AcDx-8474-APC-S1-Dn	/5Phos/GAGGAGTACGGAGTTAGGGTTAGGTAGGTGTGTCGGAGCGGTTACTA	47	1983
Real-Time Probe	AcDx-8475-APC-S1-RT-Pb	/56-FAM/CCTATTCTC/ZEN/GAGGAGTACGGAGTTAGGGTTAG/3IABkFQ/	32	1984
Tag Forward Primer	AcDx-8476-APC-S1-RT-FP	TCCAAACAAGCTGATCCGTACA	22	1985
Tag Reverse Primer	AcDx-8477-APC-S1-RT-RP	TAGTAACCGCTCCGACACA	19	1986
Downstream PCR Primer	AcDx-8478-APC-S1-PCR-V	TAGTAACCGCTCCGACACACAAAACCCCGCCCAACTGrCACAG/3SpC3/	42	1987
RUNDC3B				
Forward PCR Primer	AcDx-8481-RUNDC3B-FP	GGATCGAGGTCGCGTCrGTTTC/3SpC3/	21	1988
Reverse PCR Primer	AcDx-8482-RUNDC3B-RP	GGTGTCGTGGAAACTAACGACCCGCCTAArAAAAG/3SpC3/	34	1989
Upstream LDR	AcDx-8483-RUNDC3B-Up	TAGGGCGACAGTTACCACAAGTTTTTGGGTTTATTATTTTTAGATTCGGATTTTCGATCTCrGGAAC/3SpC3/	66	1990
Downstream LDR	AcDx-8484-RUNDC3B-Dn	/5Phos/GGAGTTACGGTTTAGGACGCGAAAAGATTTTTTGTGGGTCTCGCTCGTATA	51	1991
Real-Time Probe	AcDx-8485-RUNDC3B-RT-Pb	/56-FAM/AACGATCTC/ZEN/GGAGTTACGGTTTAGGACGCG/3IABkFQ/	30	1992
Tag Forward Primer	AcDx-8486-RUNDC3B-RT-FP	TAGGGCGACAGTTACCACAA	20	1993

Tag Reverse Primer	AcDx-8487-RUNDC3B-RT-RP	TATACGAGCGAGACCCACAA	20	1994
Downstream PCR Primer	AcDx-8488-RUNDC3B-PCR-V	TATACGAGCGAGACCCACAAAACTAACGACCCGCCTAAAAAAAAAATGrUAATG/3SpC3/	52	1995
CACNB2				
Forward PCR Primer	AcDx-8491-CACNB2-FP	GAATTGTTAGAGAACGTGGTTTTCrGCGGA/3SpC3/	29	1996
Reverse PCR Primer	AcDx-8492-CACNB2-RP	GGTGTCGTGGCCCTACCCGACGACTCArCAAAG/3SpC3/	32	1997
Upstream LDR	AcDx-8493-CACNB2-Up	TCCGACTTTAGTGCGTCACAAGTGGTTTTCGCGGGAGCrGTTTA/3SpC3/	43	1998
Downstream LDR	AcDx-8494-CACNB2-Dn	/5Phos/GTTCGGAGTCGTCGTATAGGTAGCGAGAGCTTGTGGGTCTCGCTCGTATA	50	1999
Real-Time Probe	AcDx-8495-CACNB2-RT-Pb	/56-FAM/TTCGGGAGC/ZEN/GTTCGGAGTCGTCGTATAG/3IABkFQ/	28	2000
Tag Forward Primer	AcDx-8496-CACNB2-RT-FP	TCCGACTTTAGTGCGTCACAA	21	2001
Tag Reverse Primer	AcDx-8497-CACNB2-RT-RP	TATACGAGCGAGACCCACAA	20	2002
Downstream PCR Primer	AcDx-8498-CACNB2-PCR-V	TATACGAGCGAGACCCACAAAAAAACGCCGCGCTCTTGrCTACT/3SpC3/	43	2003
ST8SIA6				
Forward PCR Primer	AcDx-8501-ST8SIA6-FP	GGTTGGCGAGTAGGGCrGAGTG/3SpC3/	21	2004
Reverse PCR Primer	AcDx-8502-ST8SIA6-RP	GGTGTCGTGGGCCCCGACAACAACGArUAACA/3SpC3/	31	2005
Upstream LDR	AcDx-8503-ST8SIA6-Up	TCAAACAAAGGCGACCACAACGCGAGTAGGGCGAGTAGCGCrGTTCC/3SpC3/	46	2006
Downstream LDR	AcDx-8504-ST8SIA6-Dn	/5Phos/GTTTTTCGGTCGTATTTTGAGTTATAGGCGTCGTCGGTTGTCGCATAGGCAGTTCATA	58	2007
Real-Time Probe	AcDx-8505-ST8SIA6-RT-Pb	/56-FAM/TTGTAGCGC/ZEN/GTTTTTCGGTCGTATTTTGAGTTATAGG/3IABkFQ/	37	2008
Tag Forward Primer	AcDx-8506-ST8SIA6-RT-FP	TCAAACAAAGGCGACCACAAC	21	2009
Tag Reverse Primer	AcDx-8507-ST8SIA6-RT-RP	TATGAACTGCCTATGCGACAAC	22	2010
Downstream PCR Primer	AcDx-8508-ST8SIA6-PCR-V	TATGAACTGCCTATGCGACAACCGACAACAACGATAACGACGATGrCCTAC/3SpC3/	50	2011

HOXD13-S1				
Forward PCR Primer	AcDx-8511-HOXD13-S1-FP	GAATTTCGTTTTTATAAACGTTTCGCrGATGG/3SpC3/	31	2012
Reverse PCR Primer	AcDx-8512-HOXD13-S1-RP	GGTGTCGTGGTTTCTCTCTAATTCGCTCGCrUCCCA/3SpC3/	35	2013
Upstream LDR	AcDx-8513-HOXD13-S1-Up	TAAACAATGAGACCCGCTGAACGTTGGAGGGTAGGCGGATCrGGAAA/3SpC3/	46	2014
Downstream LDR	AcDx-8514-HOXD13-S1-Dn	/5Phos/GGAGGCGGGAGGTTTATAGAGGGAGAGGTTGTCGCATAGGCAGTTCATA	49	2015
Real-Time Probe	AcDx-8515-HOXD13-S1-RT-Pb	/56-FAM/TTGCGGATC/ZEN/GGAGGCGGGAGG/3IABkFQ/	21	2016
Tag Forward Primer	AcDx-8516-HOXD13-S1-RT-FP	TAAACAATGAGACCCGCTGAAC	22	2017
Tag Reverse Primer	AcDx-8517-HOXD13-S1-RT-RP	TATGAACTGCCTATGCGACAAC	22	2018
Downstream PCR Primer	AcDx-8518-HOXD13-S1-PCR-V	TATGAACTGCCTATGCGACAACCTCTCTAATTCGCTCGCTCCTGrCCCTT/3SpC3/	49	2019
ISM1				
Forward PCR Primer	AcDx-8521-ISM1-FP	CGGTGTTACGGATCGATATAGTTCrGTTTC/3SpC3/	29	2020
Reverse PCR Primer	AcDx-8522-ISM1-RP	GGTGTCGTGGTCCCGCGTACGACTCArUACCC/3SpC3/	31	2021
Upstream LDR	AcDx-8523-ISM1-Up	TCTGCCAGAACACCGACACGTTCGTTTTAGTATTTGACGTTCGGATCTCrGCGCC/3SpC3/	54	2022
Downstream LDR	AcDx-8524-ISM1-Dn	/5Phos/GCGTTTTTAGTCGGATTTTAGAGTTTTTTTATTCGTTAATCGGTGTGTTGGCGTACGGTGA	61	2023
Real-Time Probe	AcDx-8525-ISM1-RT-Pb	/56-FAM/AAGGATCTC/ZEN/GCGTTTTTAGTCGGATTTTAGAGTTTT/3IABkFQ/	36	2024
Tag Forward Primer	AcDx-8526-ISM1-RT-FP	TCTGCCAGAACACCGACAC	19	2025
Tag Reverse Primer	AcDx-8527-ISM1-RT-RP	TCACCGTACGCCAACACAC	19	2026
Downstream PCR Primer	AcDx-8528-ISM1-PCR-V	TCACCGTACGCCAACACACTACGACTCATACCTTACAATAAACGATTAATGrAATAG/3SpC3/	56	2027
CTNND2-S2				
Forward PCR Primer	AcDx-8531-CTNND2-S2-FP	TTTGAGCGCGGTCGCrGGGAC/3SpC3/	20	2028
Reverse PCR Primer	AcDx-8532-CTNND2-S2-RP	GGTGTCGTGGCGAAAACCGCCTCTCGCrCCGCA/3SpC3/	32	2029

Upstream LDR	AcDx-8533-CTNND2-S2-Up	TCCGGATCAAAGCAGCCACGCGGGATAAGGGATGTTAGCrGGGTA/3SpC3/	44	2030
Downstream LDR	AcDx-8534-CTNND2-S2-Dn	/5Phos/GGGCGGTAGGAGCGAGCGTGTGTTGGCGTACGGTGA	36	2031
Real-Time Probe	AcDx-8535-CTNND2-S2-RT-Pb	/56-FAM/TTTGTTAGC/ZEN/GGGCGGTAGGAGC/3IABkFQ/	22	2032
Tag Forward Primer	AcDx-8536-CTNND2-S2-RT-FP	TCCGGATCAAAGCAGCCAC	19	2033
Tag Reverse Primer	AcDx-8537-CTNND2-S2-RT-RP	TCACCGTACGCCAACACAC	19	2034
Downstream PCR Primer	AcDx-8538-CTNND2-S2-PCR-V	TCACCGTACGCCAACACACCCTCTCGCCCGCGATGrCTCGT/3SpC3/	40	2035
GNAO1				
Forward PCR Primer	AcDx-8541-GNAO1-FP	CGGAGAAGGCGGGTTACrGGGCA/3SpC3/	22	2036
Reverse PCR Primer	AcDx-8542-GNAO1-RP	GGTGTCGTGGGAACTACCCGCAAAACGATCrAAATG/3SpC3/	35	2037
Upstream LDR	AcDx-8543-GNAO1-Up	TTCTAGATACCACGGACGCACGTTACGGGCGATAAGATGGAGAATCrGGTCT/3SpC3/	51	2038
Downstream LDR	AcDx-8544-GNAO1-Dn	/5Phos/GGTTCGAGAGGGATAGAGGTCGCGTTGGTGTTGGTGTGCAAAGCTGA	47	2039
Real-Time Probe	AcDx-8545-GNAO1-RT-Pb	/56-FAM/AAGAGAATC/ZEN/GGTTCGAGAGGGATAGAGGTCG/3IABkFQ/	31	2040
Tag Forward Primer	AcDx-8546-GNAO1-RT-FP	TTCTAGATACCACGGACGCAC	21	2041
Tag Reverse Primer	AcDx-8547-GNAO1-RT-RP	TCAGCTTTGCACACCAACAC	20	2042
Downstream PCR Primer	AcDx-8548-GNAO1-PCR-V	TCAGCTTTGCACACCAACACCGATCAAATAAACCCCCCAACTTGrCTACT/3SpC3/	49	2043
SIM2-S1				
Forward PCR Primer	AcDx-8551-SIM2-S1-FP	GTATTCGAGTTTCGTCGTTATTTTTTCrGTTCA/3SpC3/	32	2044
Reverse PCR Primer	AcDx-8552-SIM2-S1-RP	GGTGTCGTGGCTCCTAACGCGAACGACCrAACGG/3SpC3/	33	2045
Upstream LDR	AcDx-8553-SIM2-S1-Up	TATGCGGACCGATGACTCAATATTTTTTCGTTCGGCGCGTTTTTATCrGTACC/3SpC3/	52	2046
Downstream LDR	AcDx-8554-SIM2-S1-Dn	/5Phos/GTATTATTTGGGCGTTTCGGTTATTATTATTAACGGGAGGTTGCTTGGCTTGATCTACCTGA	62	2047
Real-Time Probe	AcDx-8555-SIM2-S1-RT-Pb	/56-	44	2048

		FAM/AATTTTATC/ZEN/GTATTATTTGGGCGTTTCGGTTATTATTATTAACG/3IABkFQ/		
Tag Forward Primer	AcDx-8556-SIM2-S1-RT-FP	TATGCGGACCGATGACTCAA	20	2049
Tag Reverse Primer	AcDx-8557-SIM2-S1-RT-RP	TCAGGTAGATCAAGCCAAGCAA	22	2050
Downstream PCR Primer	AcDx-8558-SIM2-S1-PCR-V	TCAGGTAGATCAAGCCAAGCAACTCCTAACGCGAACGACCAATGrCGTTG/3SpC3/	49	2051
GREM1-S1				
Forward PCR Primer	AcDx-8561-GREM1-S1-FP	GTGAGCGCGGACGTATTCrGGTAA/3SpC3/	23	2052
Reverse PCR Primer	AcDx-8562-GREM1-S1-RP	GGTGTCGTGGGAAAAACTACCGCGCGTAAArACTAG/3SpC3/	35	2053
Upstream LDR	AcDx-8563-GREM1-S1-Up	TTCGTGGGCACACAAGCAACGGTAGGGATGTGAGTGAGCrGGAAA/3SpC3/	44	2054
Downstream LDR	AcDx-8564-GREM1-S1-Dn	/5Phos/GGAGGGAAGAGGGTCGTAAATTAATTTAGGATTCGTTGCTTGGCTTGATCTACCTGA	57	2055
Real-Time Probe	AcDx-8565-GREM1-S1-RT-Pb	/56-FAM/AAAGTGAGC/ZEN/GGAGGGAAGAGGGTCGTAAATTAATTTAG/3IABkFQ/	38	2056
Tag Forward Primer	AcDx-8566-GREM1-S1-RT-FP	TTCGTGGGCACACAAGCAA	19	2057
Tag Reverse Primer	AcDx-8567-GREM1-S1-RT-RP	TCAGGTAGATCAAGCCAAGCAA	22	2058
Downstream PCR Primer	AcDx-8568-GREM1-S1-PCR-V	TCAGGTAGATCAAGCCAAGCAAAAACTACCGCGCGTAAAACTAAATGrAATCT/3SpC3/	52	2059
ZNF470				
Forward PCR Primer	AcDx-8571-ZNF470-FP	CGTTAGGTTTGGAGAGGAGTCrGTTTC/3SpC3/	26	2060
Reverse PCR Primer	AcDx-8572-ZNF470-RP	GGTGTCGTGGATAAACGTATCCTCCGCTACArATAAT/3SpC3/	36	2061
Upstream LDR	AcDx-8573-ZNF470-Up	TTGAAGGAGGAAATCGGCACAGAGAGGAGTCGTTTTTGGTAAAGTTGGCrGTTCC/3SpC3/	54	2062
Downstream LDR	AcDx-8574-ZNF470-Dn	/5Phos/GTTTTATTTTTGAACGTCGTTTTCGTTTCGGATTATATTTTTTAGTGTCGAACCGTTTTAGGACTGA	67	2063
Real-Time Probe	AcDx-8575-ZNF470-RT-Pb	/56-FAM/TTAGTTGGC/ZEN/GTTTTATTTTTGAACGTCGTTTTCGTTTC/3IABkFQ/	38	2064

Tag Forward Primer	AcDx-8576-ZNF470-RT-FP	TTGAAGGAGGAAATCGGCACA	21	2065
Tag Reverse Primer	AcDx-8577-ZNF470-RT-RP	TCAGTCCTAAAACGGTTCGACA	22	2066
Downstream PCR Primer	AcDx-8578-ZNF470-PCR-V	TCAGTCCTAAAACGGTTCGACACCGCTACAATAACCCCTAAAAAATATAATCTGrAAACA/3SpC3/	59	2067
FMN2				
Forward PCR Primer	AcDx-8581-FMN2-FP	GGTGATGTTTTGTACGAAGGCrGGCGA/3SpC3/	26	2068
Reverse PCR Primer	AcDx-8582-FMN2-RP	GGTGTCGTGGTTACCGTACTTACCTAACGCCrUTCTC/3SpC3/	36	2069
Upstream LDR	AcDx-8583-FMN2-Up	TTCGGCAGGCTACGGTACAGAAGGCGGCGGTAGCrGTCAG/3SpC3/	39	2070
Downstream LDR	AcDx-8584-FMN2-Dn	/5Phos/GTCGAGGATGCGTTGGAGTTTAGGGTGTCGAACCGTTTTAGGACTGA	47	2071
Real-Time Probe	AcDx-8585-FMN2-RT-Pb	/56-FAM/TTCGGTAGC/ZEN/GTCGAGGATGCGTTGG/3IABkFQ/	25	2072
Tag Forward Primer	AcDx-8586-FMN2-RT-FP	TTCGGCAGGCTACGGTACA	19	2073
Tag Reverse Primer	AcDx-8587-FMN2-RT-RP	TCAGTCCTAAAACGGTTCGACA	22	2074
Downstream PCR Primer	AcDx-8588-FMN2-PCR-V	TCAGTCCTAAAACGGTTCGACACCTAACGCCTTCTTACCCCTGrCTCCT/3SpC3/	48	2075
T				
Forward PCR Primer	AcDx-8591-T-FP	GGATTTGTTGTCGAGGTTAGCrGCGCA/3SpC3/	26	2076
Reverse PCR Primer	AcDx-8592-T-RP	GGTGTCGTGGCCTTCTCCCACCTTCCGTArAAAAT/3SpC3/	34	2077
Upstream LDR	AcDx-8593-T-Up	TCTCGACGATGAAAAGCAACAGTCGAGGTTAGCGCGCGTGTCrGAGCA/3SpC3/	47	2078
Downstream LDR	AcDx-8594-T-Dn	/5Phos/GAGTGTGTTAGGCGTGTGCGTGGTTTTTGTGTGGGTACTGTCCGTGGA	48	2079
Real-Time Probe	AcDx-8595-T-RT-Pb	/56-FAM/TTGCGTGTC/ZEN/GAGTGTGTTAGGCGTGTG/3IABkFQ/	27	2080
Tag Forward Primer	AcDx-8596-T-RT-FP	TCTCGACGATGAAAAGCAACA	21	2081
Tag Reverse Primer	AcDx-8597-T-RT-RP	TCCACGGACAGTACCCACA	19	2082
Downstream PCR Primer	AcDx-8598-T-PCR-V	TCCACGGACAGTACCCACACGTAAAAACAATAACACAACAAAAACCATGrCACAT/3SpC3/	54	2083

ZNF665				
Forward PCR Primer	AcDx-8601-ZNF665-FP	CGGGTTTGTAAGAAATTGTTCGCrGTAAC/3SpC3/	28	2084
Reverse PCR Primer	AcDx-8602-ZNF665-RP	GGTGTCGTGGCCAAAATTCAAACCCCGCCrCCGAT/3SpC3/	34	2085
Upstream LDR	AcDx-8603-ZNF665-Up	TTCTAGGCGACACGACAACAGTAGGTAAAGAGGCGGGTCGCrGGTAA/3SpC3/	46	2086
Downstream LDR	AcDx-8604-ZNF665-Dn	/5Phos/GGTGGAGAAAGGTTAGGCGTTGGGTGTGGGTACTGTCCGTGGA	43	2087
Real-Time Probe	AcDx-8605-ZNF665-RT-Pb	/56-FAM/CCGGGTCGC/ZEN/GGTGGAGAAAGGTTAG/3IABkFQ/	25	2088
Tag Forward Primer	AcDx-8606-ZNF665-RT-FP	TTCTAGGCGACACGACAACA	20	2089
Tag Reverse Primer	AcDx-8607-ZNF665-RT-RP	TCCACGGACAGTACCCACA	19	2090
Downstream PCR Primer	AcDx-8608-ZNF665-PCR-V	TCCACGGACAGTACCCACACCCTATACTCTCAATCCCTCCCAATGrCCTAG/3SpC3/	50	2091
NKX1-2				
Forward PCR Primer	AcDx-8611-NKX1-2-FP	GGTGAGCGCGTTAGGTTCrGGTAA/3SpC3/	23	2092
Reverse PCR Primer	AcDx-8612-NKX1-2-RP	GGTGTCGTGGCTAAAAAATTCCGAAACGAAAAAAAAAArAAAAG/3SpC3/	42	2093
Upstream LDR	AcDx-8613-NKX1-2-Up	TTTCAGGCCCTAACCACCACGCGTTAGGTTCGGTAGTAAGTGCrGTAAC/3SpC3/	48	2094
Downstream LDR	AcDx-8614-NKX1-2-Dn	/5Phos/GTAGTTCGTTTTCGTAGTCGCGGTTTTTTCGGGTGGGATTAAGGGCGATGGA	52	2095
Real-Time Probe	AcDx-8615-NKX1-2-RT-Pb	/56-FAM/TTTAAGTGC/ZEN/GTAGTTCGTTTTCGTAGTCGCG/3IABkFQ/	31	2096
Tag Forward Primer	AcDx-8616-NKX1-2-RT-FP	TTTCAGGCCCTAACCACCAC	20	2097
Tag Reverse Primer	AcDx-8617-NKX1-2-RT-RP	TCCATCGCCCTTAATCCCAC	20	2098
Downstream PCR Primer	AcDx-8618-NKX1-2-PCR-V	TCCATCGCCCTTAATCCCACCGAAACGAAAAAAAAAAAAAATACGAAAAATCTGrAAAAG/3SpC3/	59	2099
ZNF781-S1				
Forward PCR Primer	AcDx-8621-ZNF781-S1-FP	CGTTTAAGCGTAATTTTTTTATTTTAAATCrGTTAA/3SpC3/	35	2100
Reverse PCR Primer	AcDx-8622-ZNF781-S1-RP	GGTGTCGTGGCCCGAAAACGTAAAAACGTTCrUCTTG/3SpC3/	36	2101
Upstream LDR	AcDx-8623-ZNF781-S1-Up	TATCTCCTAAAAGAAGCCGCACCGTTAGGAATTAGTGGTTGGATTTATTGCrGTTGC/3SpC3/	56	2102
Downstream LDR	AcDx-8624-ZNF781-S1-Dn	/5Phos/GTTATAAAAGTATAAATTGTGATTCGATGAGTTCGCGTATTCGGTGGGATT	63	2103

		AAGGGCGATGGA		
Real-Time Probe	AcDx-8625-ZNF781-S1-RT-Pb	/56-FAM/TTTTATTGC/ZEN/GTTATAAAAGTATAAATTGTGATTCGATGAGTTCG/3IABkFQ/	44	2104
Tag Forward Primer	AcDx-8626-ZNF781-S1-RT-FP	TATCTCCTAAAAGAAGCCGCAC	22	2105
Tag Reverse Primer	AcDx-8627-ZNF781-S1-RT-RP	TCCATCGCCCTTAATCCCAC	20	2106
Downstream PCR Primer	AcDx-8628-ZNF781-S1-PCR-V	TCCATCGCCCTTAATCCCACCGTAAAAACGTTCTCTTATTTTCCGAATATGrAGAAT/3SpC3/	56	2107
SOST-S1				
Forward PCR Primer	AcDx-8631-SOST-S1-FP	GGGTTTTAGTAGGCGTTTTTTAGTTCrGGTTC/3SpC3/	31	2108
Reverse PCR Primer	AcDx-8632-SOST-S1-RP	GGTGTCGTGGAAACCGCTCGACCGCrAAAAG/3SpC3/	30	2109
Upstream LDR	AcDx-8633-SOST-S1-Up	TCGCTCTTCAGCCTCCTACACGGTTTGGTTGGTTTTGGCGTCTCrGGGTA/3SpC3/	49	2110
Downstream LDR	AcDx-8634-SOST-S1-Dn	/5Phos/GGGCGCGGAGTCGCGTGTTCTGGGAATTATTGCCGGA	37	2111
Real-Time Probe	AcDx-8635-SOST-S1-RT-Pb	/56-FAM/AAGCGTCTC/ZEN/GGGCGCGGG/3IABkFQ/	18	2112
Tag Forward Primer	AcDx-8636-SOST-S1-RT-FP	TCGCTCTTCAGCCTCCTACA	20	2113
Tag Reverse Primer	AcDx-8637-SOST-S1-RT-RP	TCCGGCAATAATTCCCAGAACA	22	2114
Downstream PCR Primer	AcDx-8638-SOST-S1-PCR-V	TCCGGCAATAATTCCCAGAACACGCAAAAAAACCGAAAACCGTGrACCCT/3SpC3/	49	2115
DMRTA2-S2				
Forward PCR Primer	AcDx-8641-DMRTA2-S2-FP	GCGTTATGAAGGTTAGATCGCrGGTTA/3SpC3/	26	2116
Reverse PCR Primer	AcDx-8642-DMRTA2-S2-RP	GGTGTCGTGGTACCCGCTAAACGCGCrCGCTT/3SpC3/	31	2117
Upstream LDR	AcDx-8643-DMRTA2-S2-Up	TTCAACGATCGCGCAGACAGGTTAGATCGCGGTTGTGCATCrGTCAC/3SpC3/	46	2118
Downstream LDR	AcDx-8644-DMRTA2-S2-Dn	/5Phos/GTCGTCGCGGAGTAGGTTAGGCGTAGTGTTCTGGGAATTATTGCCGGA	48	2119
Real-Time Probe	AcDx-8645-DMRTA2-S2-RT-Pb	/56-FAM/AAGTGCATC/ZEN/GTCGTCGCGGAGTAGG/3IABkFQ/	25	2120

Tag Forward Primer	AcDx-8646-DMRTA2-S2-RT-FP	TTCAACGATCGCGCAGACA	19	2121
Tag Reverse Primer	AcDx-8647-DMRTA2-S2-RT-RP	TCCGGCAATAATTCCCAGAACA	22	2122
Downstream PCR Primer	AcDx-8648-DMRTA2-S2-PCR-V	TCCGGCAATAATTCCCAGAACACTCGACCTCAACCCCCTATGrCCTAG/3SpC3/	47	2123
ZNF582				
Forward PCR Primer	AcDx-8651-ZNF582-FP	GGCGCGTTTTTATTACGGTATCrGGTGA/3SpC3/	27	2124
Reverse PCR Primer	AcDx-8652-ZNF582-RP	GGTGTCGTGGCCGACCCTATCATAACGCArACATT/3SpC3/	34	2125
Upstream LDR	AcDx-8653-ZNF582-Up	TCACTATCGGCGTAGTCACCAGGTATCGGTGGATTCGTCGCGCrGTAAC/3SpC3/	48	2126
Downstream LDR	AcDx-8654-ZNF582-Dn	/5Phos/GTAGTCGGAAGATGGCGTAGACGTATAAAGTATATCGATGTGGTGACTTTACCCGGAGGA	60	2127
Real-Time Probe	AcDx-8655-ZNF582-RT-Pb	/56-FAM/AAGTCGCGC/ZEN/GTAGTCGGAAGATGGC/3IABkFQ/	25	2128
Tag Forward Primer	AcDx-8656-ZNF582-RT-FP	TCACTATCGGCGTAGTCACCA	21	2129
Tag Reverse Primer	AcDx-8657-ZNF582-RT-RP	TCCTCCGGGTAAAGTCACCA	20	2130
Downstream PCR Primer	AcDx-8658-ZNF582-PCR-V	TCCTCCGGGTAAAGTCACCACCCTATCATAACGCAACATCGATATACTTTATATGrUCTAT/3SpC3/	60	2131
TLX3				
Forward PCR Primer	AcDx-8661-TLX3-FP	TTCGGGCGTAAAGCGCrGGGTC/3SpC3/	21	2132
Reverse PCR Primer	AcDx-8662-TLX3-RP	GGTGTCGTGGCCCGACCCTCCCGAArAACCA/3SpC3/	30	2133
Upstream LDR	AcDx-8663-TLX3-Up	TCATCTGTTCGTCAGGGTCCACGCGGGTTGGGAGGTAAACrGGGCC/3SpC3/	45	2134
Downstream LDR	AcDx-8664-TLX3-Dn	/5Phos/GGGTTTTGCGTTTCGAGGTTTTCGGATGGTGGTGACTTTACCCGGAGGA	49	2135
Real-Time Probe	AcDx-8665-TLX3-RT-Pb	/56-FAM/TTGGTAAAC/ZEN/GGGTTTTGCGTTTCGAGGTTTTC/3IABkFQ/	32	2136
Tag Forward Primer	AcDx-8666-TLX3-RT-FP	TCATCTGTTCGTCAGGGTCCA	21	2137
Tag Reverse Primer	AcDx-8667-TLX3-RT-RP	TCCTCCGGGTAAAGTCACCA	20	2138
Downstream PCR Primer	AcDx-8668-TLX3-PCR-V	TCCTCCGGGTAAAGTCACCACCGACCCTCCCGAAAACTGrAAACT/3SpC3/	44	2139

RNF220-S2				
Forward PCR Primer	AcDx-8671-RNF220-S2-FP	GGGTTCGTCGCGTTTTTCrGTTTC/3SpC3/	23	2140
Reverse PCR Primer	AcDx-8672-RNF220-S2-RP	GGTGTCGTGGTCTCTTCTACTCAAACGACGTArAAAAG/3SpC3/	37	2141
Upstream LDR	AcDx-8673-RNF220-S2-Up	TTCGTACCTCGGCACACCACGTTTTTCGTTTTGTTTCGTTTCGTTTTTCACrGTTCC/3SpC3/	56	2142
Downstream LDR	AcDx-8674-RNF220-S2-Dn	/5Phos/GTTTTTGTATTTTTAAGTCGTATTAAGAATTTAGTTTTCGATCGGTTTTTTTGGCTCCGTTACTCTGTCGA	71	2143
Real-Time Probe	AcDx-8675-RNF220-S2-RT-Pb	/56-FAM/AATTTTCAC/ZEN/GTTTTTGTATTTTTAAGTCGTATTAAGAATTTAGTTTTCG/3IABkFQ/	49	2144
Tag Forward Primer	AcDx-8676-RNF220-S2-RT-FP	TTCGTACCTCGGCACACCA	19	2145
Tag Reverse Primer	AcDx-8677-RNF220-S2-RT-RP	TCGACAGAGTAACGGAGCCA	20	2146
Downstream PCR Primer	AcDx-8678-RNF220-S2-PCR-V	TCGACAGAGTAACGGAGCCACTTCTACTCAAACGACGTAAAAAAAACTGrATCGG/3SpC3/	54	2147
ZNF568				
Forward PCR Primer	AcDx-8681-ZNF568-FP	GGAGGGATGGTTCGGCrGTTTC/3SpC3/	21	2148
Reverse PCR Primer	AcDx-8682-ZNF568-RP	GGTGTCGTGGACCCGAATATTCATCCCGCrGCGCG/3SpC3/	34	2149
Upstream LDR	AcDx-8683-ZNF568-Up	TCTACAGCTAGATGCGGCCAGGGATGGTTCGGCGTTTTAAGCrGTTCA/3SpC3/	47	2150
Downstream LDR	AcDx-8684-ZNF568-Dn	/5Phos/GTTTGTTATAGATTTATTTGCGGGTCGTTTTTATTTAGTATTTTAGAAATTGGCTCCGTTACTCTGTCGA	70	2151
Real-Time Probe	AcDx-8685-ZNF568-RT-Pb	/56-FAM/CCTTTAAGC/ZEN/GTTTGTTATAGATTTATTTGCGGGTCG/3IABkFQ/	36	2152
Tag Forward Primer	AcDx-8686-ZNF568-RT-FP	TCTACAGCTAGATGCGGCCA	20	2153
Tag Reverse Primer	AcDx-8687-ZNF568-RT-RP	TCGACAGAGTAACGGAGCCA	20	2154
Downstream PCR Primer	AcDx-8688-ZNF568-PCR-V	TCGACAGAGTAACGGAGCCACCGAATATTCATCCCGCGTGrCAATC/3SpC3/	45	2155

TFPI2				
Forward PCR Primer	AcDx-8691-TFPI2-FP	AGGTTTGGAGGGCGTTTATACrGGGAC/3SpC3/	26	2156
Reverse PCR Primer	AcDx-8692-TFPI2-RP	GGTGTCGTGGCTAAACGATACTACTCAAAAACCAACrAAATG/3SpC3/	41	2157
Upstream LDR	AcDx-8693-TFPI2-Up	TAGCGATAGTACCGACAGTCACTTATACGGAGTTTTATGGTTCGTCGCrGTTCC/3SpC3/	53	2158
Downstream LDR	AcDx-8694-TFPI2-Dn	/5Phos/GTTTTTTTCGTCGGTTGGAGAGAGAAGTTTTTGGGTGCGGAAACCTATCGTCGA	54	2159
Real-Time Probe	AcDx-8695-TFPI2-RT-Pb	/56-FAM/AATCGTCGC/ZEN/GTTTTTTTCGTCGGTTGG/3IABkFQ/	27	2160
Tag Forward Primer	AcDx-8696-TFPI2-RT-FP	TAGCGATAGTACCGACAGTCAC	22	2161
Tag Reverse Primer	AcDx-8697-TFPI2-RT-RP	TCGACGATAGGTTTCCGCAC	20	2162
Downstream PCR Primer	AcDx-8698-TFPI2-PCR-V	TCGACGATAGGTTTCCGCACCGATACTACTCAAAAACCAACAAATATCTAACTGrCTCCG/3SpC3/	59	2163
CANCR60				
Forward PCR Primer	AcDx-8701-CANCR60-FP	GTTTAGGTTTCGGGTGGCrGTATC/3SpC3/	23	2164
Reverse PCR Primer	AcDx-8702-CANCR60-RP	GGTGTCGTGGACGTTCCAACCCTTTACTTTCrCAACG/3SpC3/	36	2165
Upstream LDR	AcDx-8703-CANCR60-Up	TCCAGGGTATTTGGCGCACATTTTTGGATTATATTTTTTAGGAGGTTTTGCrGGTCG/3SpC3/	56	2166
Downstream LDR	AcDx-8704-CANCR60-Dn	/5Phos/GGTTAATTTTTTTCGGTCGAAGAAGGTTAGTTATTTTTTTTTGGGTGCGGAAACCTATCGTCGA	64	2167
Real-Time Probe	AcDx-8705-CANCR60-RT-Pb	/56-FAM/TTGTTTTGC/ZEN/GGTTAATTTTTTTCGGTCGAAGAAGGTTAG/3IABkFQ/	39	2168
Tag Forward Primer	AcDx-8706-CANCR60-RT-FP	TCCAGGGTATTTGGCGCAC	19	2169
Tag Reverse Primer	AcDx-8707-CANCR60-RT-RP	TCGACGATAGGTTTCCGCAC	20	2170
Downstream PCR Primer	AcDx-8708-CANCR60-PCR-V	TCGACGATAGGTTTCCGCACCAACCCTTTACTTTCCAACACCTGrCCAAG/3SpC3/	49	2171

ARHGEF4				
Forward PCR Primer	AcDx-8711-ARHGEF4-FP	GTAATTTGTTTTTCGGCGTTTAGCrGCGTA/3SpC3/	29	2172
Reverse PCR Primer	AcDx-8712-ARHGEF4-RP	GGTGTCGTGGGCGACCGCTCAACGAArAAACT/3SpC3/	31	2173
Upstream LDR	AcDx-8713-ARHGEF4-Up	TCCCTCGTCATCTCCCTTACCCGTTTAGCGCGTGGTGGTTCTCrGTCAC/3SpC3/	48	2174
Downstream LDR	AcDx-8714-ARHGEF4-Dn	/5Phos/GTCGTGTTGGCGTTTTTGGTTTTTATTAACGTCGGGTCTTGGTGATGGAGCGA	53	2175
Real-Time Probe	AcDx-8715-ARHGEF4-RT-Pb	/56-FAM/AAGGTTCTC/ZEN/GTCGTGTTGGCGTTTTTGG/3IABkFQ/	28	2176
Tag Forward Primer	AcDx-8716-ARHGEF4-RT-FP	TCCCTCGTCATCTCCCTTACC	21	2177
Tag Reverse Primer	AcDx-8717-ARHGEF4-RT-RP	TCGCTCCATCACCAAGACC	19	2178
Downstream PCR Primer	AcDx-8718-ARHGEF4-PCR-V	TCGCTCCATCACCAAGACCCGAAAAACCAACAAAAAAATCAATACGATGrUTAAC/3SpC3/	54	2179
ELMO1-S2				
Forward PCR Primer	AcDx-8721-ELMO1-S2-FP	TCGTTTTTTTCGGTTTCGGCrGGAGG/3SpC3/	25	2180
Reverse PCR Primer	AcDx-8722-ELMO1-S2-RP	GGTGTCGTGGCCAAAAAAACCGAAAAAAACGATCrUAAAT/3SpC3/	39	2181
Upstream LDR	AcDx-8723-ELMO1-S2-Up	TCATAATGTTGTCAGCCCGACCTTTCGGTTTCGGCGGAGATCTCrGGTAC/3SpC3/	49	2182
Downstream LDR	AcDx-8724-ELMO1-S2-Dn	/5Phos/GGTGTGTTTTCGGGAGGAGAGTATTTATTAATTAGAATTTTAGGGTCTTGGTGATGGAGCGA	62	2183
Real-Time Probe	AcDx-8725-ELMO1-S2-RT-Pb	/56-FAM/TTAGATCTC/ZEN/GGTGTGTTTTCGGGAGGAGAGTATTTATTAATTAG/3IABkFQ/	44	2184
Tag Forward Primer	AcDx-8726-ELMO1-S2-RT-FP	TCATAATGTTGTCAGCCCGACC	22	2185
Tag Reverse Primer	AcDx-8727-ELMO1-S2-RT-RP	TCGCTCCATCACCAAGACC	19	2186
Downstream PCR Primer	AcDx-8728-ELMO1-S2-PCR-V	TCGCTCCATCACCAAGACCAAAACCGAAAAAAACGATCTAAACAACTGrAAAAT/3SpC3/	53	2187

VSTM2A				
Forward PCR Primer	AcDx-8731-VSTM2A-FP	TTTTTGTTTCGAGTTTTTCGTTAGCrGTCGC/3SpC3/	30	2188
Reverse PCR Primer	AcDx-8732-VSTM2A-RP	GGTGTCGTGGTCTCTCCCCCATCGAATATCrUTTCC/3SpC3/	35	2189
Upstream LDR	AcDx-8733-VSTM2A-Up	TACGAATCACCCGAGAGTTCAAGTTTCGAGTTTTTCGTTAGCGTCGTAATCTCrGTTGC/3SpC3/	58	2190
Downstream LDR	AcDx-8734-VSTM2A-Dn	/5Phos/GTTATTGTATTGTTTTTTTCGGTATCGAGCGATTTGATTCGTTGTGGGTGGGTATAGGTCAGA	63	2191
Real-Time Probe	AcDx-8735-VSTM2A-RT-Pb	/56-FAM/TTTAATCTC/ZEN/GTTATTGTATTGTTTTTTTCGGTATCGAGCG/3IABkFQ/	40	2192
Tag Forward Primer	AcDx-8736-VSTM2A-RT-FP	TACGAATCACCCGAGAGTTCAA	22	2193
Tag Reverse Primer	AcDx-8737-VSTM2A-RT-RP	TCTGACCTATACCCACCCACAA	22	2194
Downstream PCR Primer	AcDx-8738-VSTM2A-PCR-V	TCTGACCTATACCCACCCACAACCCCATCGAATATCTTTCTAACTCGATGrAATCG/3SpC3/	55	2195
ZNF542-S1				
Forward PCR Primer	AcDx-8741-ZNF542-S1-FP	CGTTTTTTAGTTATGTACGTTTTTGTATTTCrGGTTG/3SpC3/	36	2196
Reverse PCR Primer	AcDx-8742-ZNF542-S1-RP	GGTGTCGTGGCGCGTATACGCCCGAATAArUTTCC/3SpC3/	34	2197
Upstream LDR	AcDx-8743-ZNF542-S1-Up	TACGAATCACCCGAGAGTTCAACGGTTATTGGGAGCGGGATCrGTGAA/3SpC3/	47	2198
Downstream LDR	AcDx-8744-ZNF542-S1-Dn	/5Phos/GTGGAGGTTGTATATGCGTATTGCGAGTTTTCTTGTGGGTGGGTATAGGTCAGA	54	2199
Real-Time Probe	AcDx-8745-ZNF542-S1-RT-Pb	/56-FAM/TTCGGGATC/ZEN/GTGGAGGTTGTATATGCGTATTG/3IABkFQ/	32	2200
Tag Forward Primer	AcDx-8746-ZNF542-S1-RT-FP	TACGAATCACCCGAGAGTTCAA	22	2201
Tag Reverse Primer	AcDx-8747-ZNF542-S1-RT-RP	TCTGACCTATACCCACCCACAA	22	2202
Downstream PCR Primer	AcDx-8748-ZNF542-S1-PCR-V	TCTGACCTATACCCACCCACAAGCCCGAATAATTTCTAAAAATAAACGAAAACTTGrCAATG/3SpC3/	61	2203
ST6GALNAC5				
Forward PCR Primer	AcDx-8751-ST6GALNAC5-	TAGGCGGCGGTAGGCrGGTAA/3SpC3/	20	2204

	FP			
Reverse PCR Primer	AcDx-8752-ST6GALNAC5-RP	GGTGTCGTGGAAACCCGAAAAACGCGACTArUTACG/3SpC3/	35	2205
Upstream LDR	AcDx-8753-ST6GALNAC5-Up	TAGGAACACGGAGGACATCAACGGTAGGCGGTAGTTGTGTAGGTCrGTTAG/3SpC3/	50	2206
Downstream LDR	AcDx-8754-ST6GALNAC5-Dn	/5Phos/GTTGAGAGATTACGAGGGTTCGGTTTAGTTTTAATTTTGTTGTGGGTGGGTATAGGTCAGA	61	2207
Real-Time Probe	AcDx-8755-ST6GALNAC5-RT-Pb	/56-FAM/AAGTAGGTC/ZEN/GTTGAGAGATTACGAGGGTTCGG/3IABkFQ/	32	2208
Tag Forward Primer	AcDx-8756-ST6GALNAC5-RT-FP	TAGGAACACGGAGGACATCAA	21	2209
Tag Reverse Primer	AcDx-8757-ST6GALNAC5-RT-RP	TCTGACCTATACCCACCCACAA	22	2210
ZFP82				
Forward PCR Primer	AcDx-8761-ZFP82-FP	GTTTTTTTGCGATATTGTAGTGGTTTCrGTTTG/3SpC3/	32	2211
Reverse PCR Primer	AcDx-8762-ZFP82-RP	GGTGTCGTGGGAATTCGCGCCCCGAATArAACAG/3SpC3/	33	2212
Upstream LDR	AcDx-8763-ZFP82-Up	TCTCATAAACACTCCGGCCACGTGGTTTCGTTTATTTTTTTTTATTTTTTGGATCrGTAAT/3SpC3/	60	2213
Downstream LDR	AcDx-8764-ZFP82-Dn	/5Phos/GTAGCGTTTTTTTTATAGATTATTAGGATCGACGTTTCGGGTGGCTCAATAACGGGCAGA	60	2214
Real-Time Probe	AcDx-8765-ZFP82-RT-Pb	/56-FAM/CCTTGGATC/ZEN/GTAGCGTTTTTTTTATAGATTATTAGGATCGACG/3IABkFQ/	43	2215
Tag Forward Primer	AcDx-8766-ZFP82-RT-FP	TCTCATAAACACTCCGGCCAC	21	2216
Tag Reverse Primer	AcDx-8767-ZFP82-RT-RP	TCTGCCCGTTATTGAGCCAC	20	2217
Downstream PCR Primer	AcDx-8768-ZFP82-PCR-V	TCTGCCCGTTATTGAGCCACCCCCGAATAAACAATCCTAACCTGrAAACA/3SpC3/	49	2218
ADAMTS16-S1				

Forward PCR Primer	AcDx-8771-ADAMTS16-S1-FP	GTAAATTTTAGAGCGAGGTACGCrGTTTC/3SpC3/	28	2219
Reverse PCR Primer	AcDx-8772-ADAMTS16-S1-RP	GGTGTCGTGGCGCCTCTATACCCGAACArCAACC/3SpC3/	33	2220
Upstream LDR	AcDx-8773-ADAMTS16-S1-Up	TTGATTGGGATCGTTCGCACCGAGGTACGCGTTTTTAAAGGTAGGCTCrGCGAC/3SpC3/	53	2221
Downstream LDR	AcDx-8774-ADAMTS16-S1-Dn	/5Phos/GCGGTTTTTTTACGTTTTGGCGTTCGGTTTTTCGTGGCTCAATAACGGGCAGA	53	2222
Real-Time Probe	AcDx-8775-ADAMTS16-S1-RT-Pb	/56-FAM/TTTAGGCTC/ZEN/GCGGTTTTTTTACGTTTTGGCG/3IABkFQ/	31	2223
Tag Forward Primer	AcDx-8776-ADAMTS16-S1-RT-FP	TTGATTGGGATCGTTCGCAC	20	2224
Tag Reverse Primer	AcDx-8777-ADAMTS16-S1-RT-RP	TCTGCCCGTTATTGAGCCAC	20	2225
Downstream PCR Primer	AcDx-8778-ADAMTS16-S1-PCR-V	TCTGCCCGTTATTGAGCCACGAACACAACTATAAAAACACTAAATACGAAAAACTGrAACGT/3SpC3/	61	2226
EDNRB				
Forward PCR Primer	AcDx-8781-EDNRB-FP	TAGCGCGTTTAGGAGTGCrGTCGA/3SpC3/	23	2227
Reverse PCR Primer	AcDx-8782-EDNRB-RP	GGTGTCGTGGAAATATTTAAAACCGTTTAAAAAAAACAACrGATTC/3SpC3/	45	2228
Upstream LDR	AcDx-8783-EDNRB-Up	TCTGCCCAAAATACTGCACAACGTTTAGGAGTGCGTCGGAGACTCrGGAGG/3SpC3/	50	2229
Downstream LDR	AcDx-8784-EDNRB-Dn	/5Phos/GGAAATTCGTAGAGATTTTTTAAGTTAGTAGGAATTTGGAAATTTGAAACTGAGGCGGTGTTCA	64	2230
Real-Time Probe	AcDx-8785-EDNRB-RT-Pb	/56-FAM/TTGAGACTC/ZEN/GGAAATTCGTAGAGATTTTTTAAGTTAGTAGGAATT/3IABkFQ/	45	2231
Tag Forward Primer	AcDx-8786-EDNRB-RT-FP	TCTGCCCAAAATACTGCACAA	21	2232
Tag Reverse Primer	AcDx-8787-EDNRB-RT-RP	TGAACACCGCCTCAGTTTCAA	21	2233
EPB41L3				

Forward PCR Primer	AcDx-8791-EPB41L3-FP	ATTTAAAATATGGCGTTTCGGGCrGGGAA/3SpC3/	28	2234
Reverse PCR Primer	AcDx-8792-EPB41L3-RP	GGTGTCGTGGCGACCTCCGACCTAAAAACCrUCCCC/3SpC3/	35	2235
Upstream LDR	AcDx-8793-EPB41L3-Up	TATGGACTGTACCAGCCCAAGCGAGGGATTTGTGTAAATTAGCrGGAAG/3SpC3/	48	2236
Downstream LDR	AcDx-8794-EPB41L3-Dn	/5Phos/GGAGAAGGGACGCGAGGATCGTTGAAACTGAGGCGGTGTTCA	42	2237
Real-Time Probe	AcDx-8795-EPB41L3-RT-Pb	/56-FAM/CCAATTAGC/ZEN/GGAGAAGGGACGCGGG/3IABkFQ/	25	2238
Tag Forward Primer	AcDx-8796-EPB41L3-RT-FP	TATGGACTGTACCAGCCCAA	20	2239
Tag Reverse Primer	AcDx-8797-EPB41L3-RT-RP	TGAACACCGCCTCAGTTTCAA	21	2240
Downstream PCR Primer	AcDx-8798-EPB41L3-PCR-V	TGAACACCGCCTCAGTTTCAACTAAAATCCCTAACCGAAAAAACGACTTGrATCCT/3SpC3/	55	2241
FLI1-S1				
Forward PCR Primer	AcDx-8801-FLI1-S1-FP	GTTTCGTCGTTTTTTCGGCrGGGAG/3SpC3/	23	2242
Reverse PCR Primer	AcDx-8802-FLI1-S1-RP	GGTGTCGTGGTAAAACCGACGCACGCCrUAAAT/3SpC3/	32	2243
Upstream LDR	AcDx-8803-FLI1-S1-Up	TTTGCCTCTTGTAGGTGCCATTTTTTCGGCGGAGAATTTTTTTTAGTTCTCrGTTTA/3SpC3/	56	2244
Downstream LDR	AcDx-8804-FLI1-S1-Dn	/5Phos/GTTCGTATAGATTTTTAGCGTTTCGAGTTTTCGTTTTTCGTGGGCAACGCGGATATTCA	59	2245
Real-Time Probe	AcDx-8805-FLI1-S1-RT-Pb	/56-FAM/AAAGTTCTC/ZEN/GTTCGTATAGATTTTTAGCGTTTCGAGTT/3IABkFQ/	38	2246
Tag Forward Primer	AcDx-8806-FLI1-S1-RT-FP	TTTGCCTCTTGTAGGTGCCA	20	2247
Tag Reverse Primer	AcDx-8807-FLI1-S1-RT-RP	TGAATATCCGCGTTGCCCA	19	2248
Downstream PCR Primer	AcDx-8808-FLI1-S1-PCR-V	TGAATATCCGCGTTGCCCAGCACGCCTAAACGCGAAAAATGrAAAAT/3SpC3/	46	2249
BHLHE23-S1				
Forward PCR Primer	AcDx-8811-BHLHE23-S1-FP	CGGTCGTTTCGAGGATTTCrGTTAC/3SpC3/	24	2250
Reverse PCR Primer	AcDx-8812-BHLHE23-S1-RP	GGTGTCGTGGTATCGCCAAAAACTCTTACGCrUAAAG/3SpC3/	36	2251
Upstream LDR	AcDx-8813-BHLHE23-S1-Up	TGTCGCCCGGTAGCAATAAACCGTTTCGAGGATTTCGTTATTAAACGCACrGCGCC/3SpC3/	55	2252
Downstream LDR	AcDx-8814-BHLHE23-S1-	/5Phos/GCGTTTTAGGTGGGTATTCGTTTCGGTTCGGTTTCCGCGATCTTTGCATTCA	52	2253

	Dn			
Real-Time Probe	AcDx-8815-BHLHE23-S1-RT-Pb	/56-FAM/CCAACGCAC/ZEN/GCGTTTTAGGTGGGTATTCG/3IABkFQ/	29	2254
Tag Forward Primer	AcDx-8816-BHLHE23-S1-RT-FP	TGTCGCCCGGTAGCAATAAAC	21	2255
Tag Reverse Primer	AcDx-8817-BHLHE23-S1-RT-RP	TGAATGCAAAGATCGCGGAAAC	22	2256
Downstream PCR Primer	AcDx-8818-BHLHE23-S1-PCR-V	TGAATGCAAAGATCGCGGAAACCGCTAAAATCAAACTCAAAAACGAACTGrAAACA/3SpC3/	55	2257
FOXC2				
Forward PCR Primer	AcDx-8821-FOXC2-FP	AATTGTTCGTAGCGACGCrGGATA/3SpC3/	23	2258
Reverse PCR Primer	AcDx-8822-FOXC2-RP	GGTGTCGTGGAAAACCAAACAACGAAATACATAAAArAAAAG/3SpC3/	41	2259
Upstream LDR	AcDx-8823-FOXC2-Up	TGTGCACTAGTCCACGTGAAACGCGACGCGGATGATTTATCGAATTTCTCrGCGGG/3SpC3/	55	2260
Downstream LDR	AcDx-8824-FOXC2-Dn	/5Phos/GCGAATTCGGAGGATTAAGTTGTTAGTTAGTACGTTGGTTTCCGCGATCTTTGCATTCA	59	2261
Real-Time Probe	AcDx-8825-FOXC2-RT-Pb	/56-FAM/TTATTTCTC/ZEN/GCGAATTCGGAGGATTAAGTTGTTAGTTAG/3IABkFQ/	39	2262
Tag Forward Primer	AcDx-8826-FOXC2-RT-FP	TGTGCACTAGTCCACGTGAAAC	22	2263
Tag Reverse Primer	AcDx-8827-FOXC2-RT-RP	TGAATGCAAAGATCGCGGAAAC	22	2264
Downstream PCR Primer	AcDx-8828-FOXC2-PCR-V	TGAATGCAAAGATCGCGGAAACCCAAACAACGAAATACATAAAAAAAAAAATAACAATGrUACTG/3SpC3/	64	2265
Alternate Group 1 Markers				
ZNF304-S2				
Forward PCR Primer	AcDx-8831-ZNF304-S2-FP	CGAGATTTTTGGCGTCGTCrGTCGC/3SpC3/	24	2266
Reverse PCR Primer	AcDx-8832-ZNF304-S2-RP	GGTGTCGTGGCGTAAAAAACCGACCCGATCrUCTAG/3SpC3/	35	2267
Upstream LDR	AcDx-8833-ZNF304-S2-Up	TACAGATACGGACGGGAATCAATTTTGGCGTCGTCGTCGTAACrGTACC/3SpC3/	48	2268

Downstream LDR	AcDx-8834-ZNF304-S2-Dn	/5Phos/GTATTTTTTTTATGTTCGGTTCGTGTATTTTGGTTGTGAAGGTTGTTTACATCCTCCTGCGTCA	64	2269
Real-Time Probe	AcDx-8835-ZNF304-S2-RT-Pb	/56-FAM/TTTCGTAAC/ZEN/GTATTTTTTTTATGTTCGGTTCGTGTATTTTG/3IABkFQ/	41	2270
Tag Forward Primer	AcDx-8836-ZNF304-S2-RT-FP	TACAGATACGGACGGGAATCAA	22	2271
Tag Reverse Primer	AcDx-8837-ZNF304-S2-RT-RP	TGACGCAGGAGGATGTAAACAA	22	2272
AMOTL1				
Forward PCR Primer	AcDx-8841-AMOTL1-FP	GTGGAGGGTAAAGTTGCGTCrGGGAG/3SpC3/	25	2273
Reverse PCR Primer	AcDx-8842-AMOTL1-RP	GGTGTCGTGGCGTAAAAAACTCGAAAACGCCrCTCCT/3SpC3/	36	2274
Upstream LDR	AcDx-8843-AMOTL1-Up	TTGGCGCAACGGTTTCCAACGTCGAGGAATTTGTGAGTTCGCrGGTAG/3SpC3/	47	2275
Downstream LDR	AcDx-8844-AMOTL1-Dn	/5Phos/GGTGAAAGGTAATTAGTTTTTATTCGAGGTGTCGGGAGTTGTTTACATCCTCCTGCGTCA	60	2276
Real-Time Probe	AcDx-8845-AMOTL1-RT-Pb	/56-FAM/CCAGTTCGC/ZEN/GGTGAAAGGTAATTAGTTTTTATTCGAGG/3IABkFQ/	38	2277
Tag Forward Primer	AcDx-8846-AMOTL1-RT-FP	TTGGCGCAACGGTTTCCAA	19	2278
Tag Reverse Primer	AcDx-8847-AMOTL1-RT-RP	TGACGCAGGAGGATGTAAACAA	22	2279
Downstream PCR Primer	AcDx-8848-AMOTL1-PCR-V	TGACGCAGGAGGATGTAAACAAAACTCGAAAACGCCCTCCTGrACACT/3SpC3/	47	2280
ST3GAL4-S2				
Forward PCR Primer	AcDx-8851-ST3GAL4-S2-FP	GGATAGTTTTGCGGAAAGTTTTCrGTTTC/3SpC3/	28	2281
Reverse PCR Primer	AcDx-8852-ST3GAL4-S2-RP	GGTGTCGTGGCCTAACCGAAATAATTATAAAAAAAACCATCrGCTCA/3SpC3/	46	2282
Upstream LDR	AcDx-8853-ST3GAL4-S2-Up	TTCAGCAGCCTGGCATCACGAAAGTTTTCGTTTTTAATTTTTTAGTTTTGCGCTCrGGACC/3SpC3/	60	2283
Downstream LDR	AcDx-8854-ST3GAL4-S2-Dn	/5Phos/GGATTGAAGCGGCGGTTTTTATTTTTAGTATTTTCGAGGTGGAGCGCTAAGGTTGCA	57	2284
Real-Time Probe	AcDx-8855-ST3GAL4-S2-	/56-FAM/AATGCGCTC/ZEN/GGATTGAAGCGGC/3IABkFQ/	22	2285

	RT-Pb			
Tag Forward Primer	AcDx-8856-ST3GAL4-S2-RT-FP	TTCAGCAGCCTGGCATCAC	19	2286
Tag Reverse Primer	AcDx-8857-ST3GAL4-S2-RT-RP	TGCAACCTTAGCGCTCCAC	19	2287
Downstream PCR Primer	AcDx-8858-ST3GAL4-S2-PCR-V	TGCAACCTTAGCGCTCCACAACCGAAATAATTATAAAAAAAACCATCGCTTGrAAAAC/3SpC3/	57	2288
THBD-S2				
Forward PCR Primer	AcDx-8861-THBD-S2-FP	TATAGGACGTCGATGGCGATArGTTTC/3SpC3/	26	2289
Reverse PCR Primer	AcDx-8862-THBD-S2-RP	GGTGTCGTGGCGATCCGCATATCAAAAACTACCrUCGCG/3SpC3/	38	2290
Upstream LDR	AcDx-8863-THBD-S2-Up	TTCAGAGCACCTGCGTACCACGTCGATGGCGATAGTTTTTTTTGCTCrGTTCC/3SpC3/	52	2291
Downstream LDR	AcDx-8864-THBD-S2-Dn	/5Phos/GTTTTAGTTTAGATATTTTTTGTCGTTGCGCGTAGTTTTTGGGTTCTTCGGCTGGCTCAA	60	2292
Real-Time Probe	AcDx-8865-THBD-S2-RT-Pb	/56-FAM/AATTTGCTC/ZEN/GTTTTAGTTTAGATATTTTTTGTCGTTGCG/3IABkFQ/	39	2293
Tag Forward Primer	AcDx-8866-THBD-S2-RT-FP	TTCAGAGCACCTGCGTACC	19	2294
Tag Reverse Primer	AcDx-8867-THBD-S2-RT-RP	TTGAGCCAGCCGAAGAACC	19	2295
Downstream PCR Primer	AcDx-8868-THBD-S2-PCR-V	TTGAGCCAGCCGAAGAACCCATATCAAAAACTACCTCGCAAAAACTATGrCGCAG/3SpC3/	54	2296
RNLS-S1				
Forward PCR Primer	AcDx-8871-RNLS-S1-FP	GTCGGAGTTGAGAGGTTTTTTCrGTTTC/3SpC3/	27	2297
Reverse PCR Primer	AcDx-8872-RNLS-S1-RP	GGTGTCGTGGCTTTTCTAAACGAAAAACAAAACGCCrCGACT/3SpC3/	41	2298
Upstream LDR	AcDx-8873-RNLS-S1-Up	TCCTAGTACCTACAGTGGGCAAGAGTAAATTTGAGGTTGCGGAGAGTCrGGGAC/3SpC3/	53	2299
Downstream LDR	AcDx-8874-RNLS-S1-Dn	/5Phos/GGAGTTGTTTGTTTTTTCGTTTTGGAGGTCTTGACCGCTGTTATACGTTGCA	52	2300
Real-Time Probe	AcDx-8875-RNLS-S1-RT-Pb	/56-FAM/TTGAGAGTC/ZEN/GGAGTTGTTTGTTTTTTCGTTTTGGG/3IABkFQ/	35	2301
Tag Forward Primer	AcDx-8876-RNLS-S1-RT-FP	TCCTAGTACCTACAGTGGGCAA	22	2302
Tag Reverse Primer	AcDx-8877-RNLS-S1-RT-RP	TGCAACGTATAACAGCGGTCAA	22	2303

Downstream PCR Primer	AcDx-8878-RNLS-S1-PCR-V	TGCAACGTATAACAGCGGTCAACTTTTCTAAACGAAAAACAAAACGCCTGrACCCT/3SpC3/	55	2304
NKAIN2				
Forward PCR Primer	AcDx-8881-NKAIN2-FP	GGAGTCGCGGGTTGCrGTAGA/3SpC3/	20	2305
Reverse PCR Primer	AcDx-8882-NKAIN2-RP	GGTGTCGTGGTCGCCCTACGCTCGCrCGCGT/3SpC3/	30	2306
Upstream LDR	AcDx-8883-NKAIN2-Up	TAGTTTGTCGAAAGTCCCACACCGTAGGGAGGCGGGTGGCrGGTCC/3SpC3/	45	2307
Downstream LDR	AcDx-8884-NKAIN2-Dn	/5Phos/GGTTTTCGCGCGTTTTTGTCGTTTTCGGGTGCAAAATTCAGGCTGTGCA	49	2308
Real-Time Probe	AcDx-8885-NKAIN2-RT-Pb	/56-FAM/TTGGGTGGC/ZEN/GGTTTTCGCGCG/3IABkFQ/	21	2309
Tag Forward Primer	AcDx-8886-NKAIN2-RT-FP	TAGTTTGTCGAAAGTCCCACAC	22	2310
Tag Reverse Primer	AcDx-8887-NKAIN2-RT-RP	TGCACAGCCTGAATTTTGCAC	21	2311
Downstream PCR Primer	AcDx-8888-NKAIN2-PCR-V	TGCACAGCCTGAATTTTGCACGCGCCGACTCCGAAAATGrACAAG/3SpC3/	44	2312
RNLS-S2				
Forward PCR Primer	AcDx-8891-RNLS-S2-FP	TAGTAAGGTATAAGGGATCGGACrGTTTA/3SpC3/	28	2313
Reverse PCR Primer	AcDx-8892-RNLS-S2-RP	GGTGTCGTGGCGCTACTCCCTCTCGCCrATAAT/3SpC3/	32	2314
Upstream LDR	AcDx-8893-RNLS-S2-Up	TAGACACGAGCGAGGTCACCGGACGTTTGTTTTTTTAGTAGCGTGGCrGTACG/3SpC3/	52	2315
Downstream LDR	AcDx-8894-RNLS-S2-Dn	/5Phos/GTATAAGTTTTTTGTTATTTCGGCGTTTACGATTAGTATTTGCGGTGCAAAATTCAGGCTGTGCA	65	2316
Real-Time Probe	AcDx-8895-RNLS-S2-RT-Pb	/56-FAM/CCGCGTGGC/ZEN/GTATAAGTTTTTTGTTATTTCGGCGTTTA/3IABkFQ/	38	2317
Tag Forward Primer	AcDx-8896-RNLS-S2-RT-FP	TAGACACGAGCGAGGTCAC	19	2318
Tag Reverse Primer	AcDx-8897-RNLS-S2-RT-RP	TGCACAGCCTGAATTTTGCAC	21	2319
Downstream PCR Primer	AcDx-8898-RNLS-S2-PCR-V	TGCACAGCCTGAATTTTGCACCTCTCGCCATAACGCAAATACTAATTGrUAAAT/3SpC3/	53	2320
RNLS-S3				

Forward PCR Primer	AcDx-8901-RNLS-S3-FP	GAATCGGAGTTGTTTGTTTTTTCrGTTTC/3SpC3/	28	2321
Reverse PCR Primer	AcDx-8902-RNLS-S3-RP	GGTGTCGTGGACTCTATTACGAAACGAATTCTCCrCAACG/3SpC3/	39	2322
Upstream LDR	AcDx-8903-RNLS-S3-Up	TTCTTCACAGTACCGCCACATCGTTTTGGAGGTCGAGCrGTTCC/3SpC3/	43	2323
Downstream LDR	AcDx-8904-RNLS-S3-Dn	/5Phos/GTTTTGTTTTTCGTTTAGAAAAGTTTGGAAATGGTGTGGGTGTGTTGTCTGGTGGTGCA	59	2324
Real-Time Probe	AcDx-8905-RNLS-S3-RT-Pb	/56-FAM/TTGTCGAGC/ZEN/GTTTTGTTTTTCGTTTAGAAAAGTTTGG/3IABkFQ/	37	2325
Tag Forward Primer	AcDx-8906-RNLS-S3-RT-FP	TTCTTCACAGTACCGCCACA	20	2326
Tag Reverse Primer	AcDx-8907-RNLS-S3-RT-RP	TGCACCACCAGACAACACA	19	2327
RNLS-S4				
Forward PCR Primer	AcDx-8911-RNLS-S4-FP	GAATCGGAGTTGTTTGTTTTTTCrGTTTC/3SpC3/	28	2328
Reverse PCR Primer	AcDx-8912-RNLS-S4-RP	GGTGTCGTGGCTATTACGAAACGAATTCTCCCAACrAATCG/3SpC3/	40	2329
Upstream LDR	AcDx-8913-RNLS-S4-Up	TACCCTCCTAGCTCCGTACAGGTTGTTTGTTTTTTCGTTTTGGGAGTCrGGGTA/3SpC3/	53	2330
Downstream LDR	AcDx-8914-RNLS-S4-Dn	/5Phos/GGGCGTTTTGTTTTTCGTTTAGAAAAGTTTGGAAATGGTGTGTTGTCTGGTGGTGCA	57	2331
Real-Time Probe	AcDx-8915-RNLS-S4-RT-Pb	/56-FAM/AAGGGAGTC/ZEN/GGGCGTTTTGTTTTTCGTTTAGAAAAG/3IABkFQ/	36	2332
Tag Forward Primer	AcDx-8916-RNLS-S4-RT-FP	TACCCTCCTAGCTCCGTACA	20	2333
Tag Reverse Primer	AcDx-8917-RNLS-S4-RT-RP	TGCACCACCAGACAACACA	19	2334
RNLS-S5				
Forward PCR Primer	AcDx-8921-RNLS-S5-FP	TTTGGAGTTGCGGAGAATCrGGGAC/3SpC3/	24	2335
Reverse PCR Primer	AcDx-8922-RNLS-S5-RP	GGTGTCGTGGCATTTCCAAACTTTTCTAAACGAAArAACAG/3SpC3/	40	2336
Upstream LDR	AcDx-8923-RNLS-S5-Up	TCATCGCCCTCAGATCTTCCACGGAGAATCGGAGTTGTTTGTTTTCTCrGTTCC/3SpC3/	53	2337
Downstream LDR	AcDx-8924-RNLS-S5-Dn	/5Phos/GTTTTGGAGGTCGGGCGTTTTGTTGGAGGATAGATTGGAGGGCA	44	2338
Real-Time Probe	AcDx-8925-RNLS-S5-RT-Pb	/56-FAM/AATTTTCTC/ZEN/GTTTTGGAGGTCGGG/3IABkFQ/	24	2339

Tag Forward Primer	AcDx-8926-RNLS-S5-RT-FP	TCATCGCCCTCAGATCTTCCA	21	2340
Tag Reverse Primer	AcDx-8927-RNLS-S5-RT-RP	TGCCCTCCAATCTATCCTCCA	21	2341
Downstream PCR Primer	AcDx-8928-RNLS-S5-PCR-V	TGCCCTCCAATCTATCCTCCACATTTCCAAACTTTTCTAAACGAAAAACAAAATGrCCCGG/3SpC3/	60	2342
APC-S2				
Forward PCR Primer	AcDx-8931-APC-S2-FP	TGAGGATTGAGGTCGCrGAGGA/3SpC3/	21	2343
Reverse PCR Primer	AcDx-8932-APC-S2-RP	GGTGTCGTGGCACAAAACCCCGCCCArACCGT/3SpC3/	31	2344
Upstream LDR	AcDx-8933-APC-S2-Up	TAACCAGTTACCACCGCCAGATTGAGGTCGCGAGGGTATATTCTCrGAGAA/3SpC3/	50	2345
Downstream LDR	AcDx-8934-APC-S2-Dn	/5Phos/GAGGAGTACGGAGTTAGGGTTAGGTAGTGGAGGATAGATTGGAGGGCA	48	2346
Real-Time Probe	AcDx-8935-APC-S2-RT-Pb	/56-FAM/CCTATTCTC/ZEN/GAGGAGTACGGAGTTAGGGTTAG/3IABkFQ/	32	2347
Tag Forward Primer	AcDx-8936-APC-S2-RT-FP	TAACCAGTTACCACCGCCA	19	2348
Tag Reverse Primer	AcDx-8937-APC-S2-RT-RP	TGCCCTCCAATCTATCCTCCA	21	2349
Downstream PCR Primer	AcDx-8938-APC-S2-PCR-V	TGCCCTCCAATCTATCCTCCACAAAACCCCGCCCAACTGrCACAG/3SpC3/	44	2350
RNLS-S6				
Forward PCR Primer	AcDx-8941-RNLS-S6-FP	GTCGGAGTTGAGAGGTTTTTTCrGTTTC/3SpC3/	27	2351
Reverse PCR Primer	AcDx-8942-RNLS-S6-RP	GGTGTCGTGGTCTAAACGAAAAACAAAACGCCrCGACT/3SpC3/	37	2352
Upstream LDR	AcDx-8943-RNLS-S6-Up	TCGCACCGGAATTCTGACCGGAGTAAATTTGAGGTTGCGGAGAGTCrGGGAC/3SpC3/	51	2353
Downstream LDR	AcDx-8944-RNLS-S6-Dn	/5Phos/GGAGTTGTTTGTTTTTTCGTTTTGGAGGTCGGGTAGTTTCCCATGACGGCA	51	2354
Real-Time Probe	AcDx-8945-RNLS-S6-RT-Pb	/56-FAM/TTGAGAGTC/ZEN/GGAGTTGTTTGTTTTTTCGTTTTGG/3IABkFQ/	34	2355
Tag Forward Primer	AcDx-8946-RNLS-S6-RT-FP	TCGCACCGGAATTCTGACC	19	2356
Tag Reverse Primer	AcDx-8947-RNLS-S6-RT-RP	TGCCGTCATGGGAAACTACC	20	2357
Downstream PCR Primer	AcDx-8948-RNLS-S6-PCR-V	TGCCGTCATGGGAAACTACCTCTAAACGAAAAACAAAACGCCTGrACCCT/3SpC3/	49	2358

CANCR61				
Forward PCR Primer	AcDx-8951-CANCR61-FP	GCGGTGTTTTTGTGTGCrGGTGA/3SpC3/	22	2359
Reverse PCR Primer	AcDx-8952-CANCR61-RP	GGTGTCGTGGCCTAACCCCGACCCGAArCCCGT/3SpC3/	32	2360
Upstream LDR	AcDx-8953-CANCR61-Up	TTGCACGTTGTCCTGCACCGTGTGCGGTGGGTGCrGGCAG/3SpC3/	39	2361
Downstream LDR	AcDx-8954-CANCR61-Dn	/5Phos/GGCGATGGGTTGTTTCGGTTCGTAGTTCGGTAGTTTCCCATGACGGCA	48	2362
Real-Time Probe	AcDx-8955-CANCR61-RT-Pb	/56-FAM/TTTGGGTGC/ZEN/GGCGATGGGTTG/3IABkFQ/	21	2363
Tag Forward Primer	AcDx-8956-CANCR61-RT-FP	TTGCACGTTGTCCTGCACC	19	2364
Tag Reverse Primer	AcDx-8957-CANCR61-RT-RP	TGCCGTCATGGGAAACTACC	20	2365
Downstream PCR Primer	AcDx-8958-CANCR61-PCR-V	TGCCGTCATGGGAAACTACCTCCCCTCGAACTACGAACTGrAAACG/3SpC3/	45	2366
APC-S3				
Forward PCR Primer	AcDx-8961-APC-S3-FP	AGGGTTAGGTAGGTTGTGCrGGTTA/3SpC3/	24	2367
Reverse PCR Primer	AcDx-8962-APC-S3-RP	GGTGTCGTGGTCTCTCCGCTTCCCGACrCCGCG/3SpC3/	32	2368
Upstream LDR	AcDx-8963-APC-S3-Up	TGCTTACCCACGATGCACCGTAGGTTGTGCGGTTGAGCrGGGAC/3SpC3/	43	2369
Downstream LDR	AcDx-8964-APC-S3-Dn	/5Phos/GGAGTTTTGTGTTTTATTGCGGAGTGCGGGTCGTATGACTTGCTCGCA	48	2370
Real-Time Probe	AcDx-8965-APC-S3-RT-Pb	/56-FAM/TTGTTGAGC/ZEN/GGAGTTTTGTGTTTTATTGCGG/3IABkFQ/	31	2371
Tag Forward Primer	AcDx-8966-APC-S3-RT-FP	TGCTTACCCACGATGCACC	19	2372
Tag Reverse Primer	AcDx-8967-APC-S3-RT-RP	TGCGAGCAAGTCATACGACC	20	2373
Downstream PCR Primer	AcDx-8968-APC-S3-PCR-V	TGCGAGCAAGTCATACGACCCTCCGCTTCCCGACCTGrCACTT/3SpC3/	42	2374
APC-S4				
Forward PCR Primer	AcDx-8971-APC-S4-FP	AGGGTATATTTTCGAGAGGTACrGGGAC/3SpC3/	27	2375
Reverse PCR Primer	AcDx-8972-APC-S4-RP	GGTGTCGTGGCTCTCCGCTTCCCGACrCCGCG/3SpC3/	31	2376
Upstream LDR	AcDx-8973-APC-S4-Up	TTACAGGCCGCATAGCAACGTAGGTTGTGCGGTTGAGCrGGGAC/3SpC3/	43	2377

Downstream LDR	AcDx-8974-APC-S4-Dn	/5Phos/GGAGTTTTGTGTTTTATTGCGGAGTGCGGTTGAGACATGGGCTCGCA	47	2378
Real-Time Probe	AcDx-8975-APC-S4-RT-Pb	/56-FAM/TTGTTGAGC/ZEN/GGAGTTTTGTGTTTTATTGCGGA/3IABkFQ/	32	2379
Tag Forward Primer	AcDx-8976-APC-S4-RT-FP	TTACAGGCCGCATAGCAAC	19	2380
Tag Reverse Primer	AcDx-8977-APC-S4-RT-RP	TGCGAGCCCATGTCTCAAC	19	2381
Downstream PCR Primer	AcDx-8978-APC-S4-PCR-V	TGCGAGCCCATGTCTCAACCTCCGCTTCCCGACCTGrCACTT/3SpC3/	41	2382
APC-S5				
Forward PCR Primer	AcDx-8981-APC-S5-FP	GTTGTGCGGTTGGGCrGGGAC/3SpC3/	20	2383
Reverse PCR Primer	AcDx-8982-APC-S5-RP	GGTGTCGTGGCTAATCCGCATCCAACGAATTACrACAAT/3SpC3/	38	2384
Upstream LDR	AcDx-8983-APC-S5-Up	TTCGCCTACCGCAGTGAACGGCGGAGTTTTGTGTTTTATCGCrGGAAC/3SpC3/	47	2385
Downstream LDR	AcDx-8984-APC-S5-Dn	/5Phos/GGAGTGCGGGTCGGGAAGCGTTGAGACATGGGCTCGCA	38	2386
Real-Time Probe	AcDx-8985-APC-S5-RT-Pb	/56-FAM/AATTATCGC/ZEN/GGAGTGCGGGTCGG/3IABkFQ/	23	2387
Tag Forward Primer	AcDx-8986-APC-S5-RT-FP	TTCGCCTACCGCAGTGAAC	19	2388
Tag Reverse Primer	AcDx-8987-APC-S5-RT-RP	TGCGAGCCCATGTCTCAAC	19	2389
Downstream PCR Primer	AcDx-8988-APC-S5-PCR-V	TGCGAGCCCATGTCTCAACACGAATTACACAACTACTTCTCTCTCTGrCTTCT/3SpC3/	52	2390
GPX7				
Forward PCR Primer	AcDx-8991-GPX7-FP	GCGTAGTAGGAGTAGGATTTTTACrGATTC/3SpC3/	29	2391
Reverse PCR Primer	AcDx-8992-GPX7-RP	GGTGTCGTGGCGCGCACTCACCGATCrCGCGG/3SpC3/	31	2392
Upstream LDR	AcDx-8993-GPX7-Up	TTCGCCTACCGCAGTGAACGTAGGATTTTTACGATTTTAAGGCGGTTAATACTCrGGGAC/3SpC3/	59	2393
Downstream LDR	AcDx-8994-GPX7-Dn	/5Phos/GGAGTAAATTGGTGTCGTTGGAGAAGTATCGCGTTGAGACATGGGCTCGCA	51	2394
Real-Time Probe	AcDx-8995-GPX7-RT-Pb	/56-FAM/AAAATACTC/ZEN/GGAGTAAATTGGTGTCGTTGGAGA/3IABkFQ/	33	2395
Tag Forward Primer	AcDx-8996-GPX7-RT-FP	TTCGCCTACCGCAGTGAAC	19	2396
Tag Reverse Primer	AcDx-8997-GPX7-RT-RP	TGCGAGCCCATGTCTCAAC	19	2397
Downstream PCR	AcDx-8998-GPX7-PCR-V	TGCGAGCCCATGTCTCAACGCACTCACCGATCCGTGrATACC/3SpC3/	41	2398

Primer				
SPSB4				
Forward PCR Primer	AcDx-9001-SPSB4-FP	GTTGGTCGTGGCGGTCrGGCGT/3SpC3/	21	2399
Reverse PCR Primer	AcDx-9002-SPSB4-RP	GGTGTCGTGGCCAACGCCCTAAACGCArCGCAG/3SpC3/	32	2400
Upstream LDR	AcDx-9003-SPSB4-Up	TTGCATTTCGTTAGCGACACAGTTTTAGAGGAGGGCGATGAATATATTATATCTCrGGGCC/3SpC3/	60	2401
Downstream LDR	AcDx-9004-SPSB4-Dn	/5Phos/GGGTTCGGGTTTTAGTTGTTGTTATCGTTGCGTGTGAGTCGATCTACCCGCA	52	2402
Real-Time Probe	AcDx-9005-SPSB4-RT-Pb	/56-FAM/AAATATCTC/ZEN/GGGTTCGGGTTTTAGTTGTTGTTATCG/3IABkFQ/	36	2403
Tag Forward Primer	AcDx-9006-SPSB4-RT-FP	TTGCATTTCGTTAGCGACACA	21	2404
Tag Reverse Primer	AcDx-9007-SPSB4-RT-RP	TGCGGGTAGATCGACTCACA	20	2405
Downstream PCR Primer	AcDx-9008-SPSB4-PCR-V	TGCGGGTAGATCGACTCACACCAACGCCCTAAACGCATGrCAACA/3SpC3/	44	2406
CHRNA3				
Forward PCR Primer	AcDx-9011-CHRNA3-FP	GTTAGAGGTAGCGAGAGCrGGGTC/3SpC3/	23	2407
Reverse PCR Primer	AcDx-9012-CHRNA3-RP	GGTGTCGTGGACCGTCTCTACGACCGCrCGCGT/3SpC3/	32	2408
Upstream LDR	AcDx-9013-CHRNA3-Up	TTGCATTTCGTTAGCGACACACGGGTTAGAGTTTATGGTTGGTGATCrGGGCC/3SpC3/	52	2409
Downstream LDR	AcDx-9014-CHRNA3-Dn	/5Phos/GGGTTGGTCGCGGATTCGGACGTGTGAGTCGATCTACCCGCA	42	2410
Real-Time Probe	AcDx-9015-CHRNA3-RT-Pb	/56-FAM/AAGGTGATC/ZEN/GGGTTGGTCGCGGATT/3IABkFQ/	25	2411
Tag Forward Primer	AcDx-9016-CHRNA3-RT-FP	TTGCATTTCGTTAGCGACACA	21	2412
Tag Reverse Primer	AcDx-9017-CHRNA3-RT-RP	TGCGGGTAGATCGACTCACA	20	2413
Downstream PCR Primer	AcDx-9018-CHRNA3-PCR-V	TGCGGGTAGATCGACTCACACGCTCCCGACCGTCTGrAATCT/3SpC3/	41	2414
CSMD2				
Forward PCR Primer	AcDx-9021-CSMD2-FP	TTTCGCGAGGTTCGCrGTCGA/3SpC3/	20	2415

Reverse PCR Primer	AcDx-9022-CSMD2-RP	GGTGTCGTGGTTTATCCTTAACTACGCCTCCArCCCCT/3SpC3/	37	2416
Upstream LDR	AcDx-9023-CSMD2-Up	TACCACTCATCTTCTGCGACAGGCGGGAGGAGGTGAATTATTCTCrGGACC/3SpC3/	50	2417
Downstream LDR	AcDx-9024-CSMD2-Dn	/5Phos/GGATTAATTATTTCGGAGTCGTTTTTTCGTCGTTTTAAATGTTGTGTGAGTCGATCTACCCGCA	64	2418
Real-Time Probe	AcDx-9025-CSMD2-RT-Pb	/56-FAM/CCTATTCTC/ZEN/GGATTAATTATTTCGGAGTCGTTTTTTCGT/3IABkFQ/	39	2419
Tag Forward Primer	AcDx-9026-CSMD2-RT-FP	TACCACTCATCTTCTGCGACA	21	2420
Tag Reverse Primer	AcDx-9027-CSMD2-RT-RP	TGCGGGTAGATCGACTCACA	20	2421
Downstream PCR Primer	AcDx-9028-CSMD2-PCR-V	TGCGGGTAGATCGACTCACACCTCCACCCCCAACATTTAAAATGrACGAG/3SpC3/	49	2422
FOXI3				
Forward PCR Primer	AcDx-9031-FOXI3-FP	GTGTCGGCGGTAGGCrGGTGA/3SpC3/	20	2423
Reverse PCR Primer	AcDx-9032-FOXI3-RP	GGTGTCGTGGCCGCCGCGTACCTAArCGCTT/3SpC3/	30	2424
Upstream LDR	AcDx-9033-FOXI3-Up	TCTCATGGGCGCTAGTATCAACGGCGGTGGTTGTAGAAAGGGCTCrGGTTA/3SpC3/	50	2425
Downstream LDR	AcDx-9034-FOXI3-Dn	/5Phos/GGTCGCGGTTTCGGGAGGCGTTTCCCTGATTGATACCCGCA	41	2426
Real-Time Probe	AcDx-9035-FOXI3-RT-Pb	/56-FAM/TTAGGGCTC/ZEN/GGTCGCGGTTTC/3IABkFQ/	21	2427
Tag Forward Primer	AcDx-9036-FOXI3-RT-FP	TCTCATGGGCGCTAGTATCAAC	22	2428
Tag Reverse Primer	AcDx-9037-FOXI3-RT-RP	TGCGGGTATCAATCAGGGAAAC	22	2429
Downstream PCR Primer	AcDx-9038-FOXI3-PCR-V	TGCGGGTATCAATCAGGGAAACCGTACCTAAACGCTCCGCTGrCCGCT/3SpC3/	47	2430
EPHA6				
Forward PCR Primer	AcDx-9041-EPHA6-FP	GGTTTCGTAAGTGTTTATTTGAGTCrGGGAA/3SpC3/	30	2431
Reverse PCR Primer	AcDx-9042-EPHA6-RP	GGTGTCGTGGCCTCACCGCGAAAACGAArAACGG/3SpC3/	33	2432
Upstream LDR	AcDx-9043-EPHA6-Up	TGGATCGAGACGGAATGCAACGTGTTTATTTGAGTCGGGAGAGGTTCTCrGGGCT/3SpC3/	54	2433
Downstream LDR	AcDx-9044-EPHA6-Dn	/5Phos/GGGTCGTTTTAATTGTTCGTTTTATTTGGTAGTTCGTTTTCGTTTCCCTGATTGATACCCGCA	63	2434
Real-Time Probe	AcDx-9045-EPHA6-RT-Pb	/56-FAM/TTGGTTCTC/ZEN/GGGTCGTTTTAATTGTTCGTTTTATTTGG/3IABkFQ/	38	2435

Tag Forward Primer	AcDx-9046-EPHA6-RT-FP	TGGATCGAGACGGAATGCAAC	21	2436
Tag Reverse Primer	AcDx-9047-EPHA6-RT-RP	TGCGGGTATCAATCAGGGAAAC	22	2437
Downstream PCR Primer	AcDx-9048-EPHA6-PCR-V	TGCGGGTATCAATCAGGGAAACACCGCGAAAACGAAAACGAAAATGrAACTG/3SpC3/	51	2438
TSHZ3				
Forward PCR Primer	AcDx-9051-TSHZ3-FP	GGTTTATTTACGGGCGGCrGGTGG/3SpC3/	23	2439
Reverse PCR Primer	AcDx-9052-TSHZ3-RP	GGTGTCGTGGTCCGACTCCCGCTCAArCCGCT/3SpC3/	31	2440
Upstream LDR	AcDx-9053-TSHZ3-Up	TCCTGAGGGACAAATACACACCCGGGTGGGTCGGAGTTTACGCrGGGTA/3SpC3/	48	2441
Downstream LDR	AcDx-9054-TSHZ3-Dn	/5Phos/GGGCGATTTTCGGGCGGCGGGTAGGTAAGGAAGTCACGCA	40	2442
Real-Time Probe	AcDx-9055-TSHZ3-RT-Pb	/56-FAM/CCTTTACGC/ZEN/GGGCGATTTTCGGGC/3IABkFQ/	24	2443
Tag Forward Primer	AcDx-9056-TSHZ3-RT-FP	TCCTGAGGGACAAATACACACC	22	2444
Tag Reverse Primer	AcDx-9057-TSHZ3-RT-RP	TGCGTGACTTCCTTACCTACC	21	2445
Downstream PCR Primer	AcDx-9058-TSHZ3-PCR-V	TGCGTGACTTCCTTACCTACCTCCCGCTCAACCGCTGrCCGCT/3SpC3/	42	2446
TRIM9				
Forward PCR Primer	AcDx-9061-TRIM9-FP	TCGGTTATTTAGGATGAGGTTGCrGGTGG/3SpC3/	28	2447
Reverse PCR Primer	AcDx-9062-TRIM9-RP	GGTGTCGTGGGCGTATTTCCCCCGACTArUACCA/3SpC3/	33	2448
Upstream LDR	AcDx-9063-TRIM9-Up	TTGCAACAGGCTACCGACCGGTGATATAGGAGTTGCAGGGTGTCrGGGAC/3SpC3/	49	2449
Downstream LDR	AcDx-9064-TRIM9-Dn	/5Phos/GGAGTTAGGGTCGGTGATAAGTGGGTGGGTAGGTAAGGAAGTCACGCA	48	2450
Real-Time Probe	AcDx-9065-TRIM9-RT-Pb	/56-FAM/TTGGGTGTC/ZEN/GGAGTTAGGGTCGG/3IABkFQ/	23	2451
Tag Forward Primer	AcDx-9066-TRIM9-RT-FP	TTGCAACAGGCTACCGACC	19	2452
Tag Reverse Primer	AcDx-9067-TRIM9-RT-RP	TGCGTGACTTCCTTACCTACC	21	2453
Downstream PCR Primer	AcDx-9068-TRIM9-PCR-V	TGCGTGACTTCCTTACCTACCCCCGACTATACCGCCACTGrACCAT/3SpC3/	45	2454

PHF21B				
Forward PCR Primer	AcDx-9071-PHF21B-FP	GCGAAGAGGAAGATAGGTTTTCrGGGCA/3SpC3/	27	2455
Reverse PCR Primer	AcDx-9072-PHF21B-RP	GGTGTCGTGGCCGAAAACCCGACGCAArCTTAT/3SpC3/	32	2456
Upstream LDR	AcDx-9073-PHF21B-Up	TAAGCCTGCTTTTCCGAAACAAGATAGGTTTTCGGGCGTTGCrGGCAC/3SpC3/	47	2457
Downstream LDR	AcDx-9074-PHF21B-Dn	/5Phos/GGCGTCGAGTTTTCGGTTGTTTTAATTTTTTAGGTTGTTGTATTGCGCCAGG ATAGCA	58	2458
Real-Time Probe	AcDx-9075-PHF21B-RT-Pb	/56-FAM/TTGCGTTGC/ZEN/GGCGTCGAGTTTTCG/3IABkFQ/	24	2459
Tag Forward Primer	AcDx-9076-PHF21B-RT-FP	TAAGCCTGCTTTTCCGAAACAA	22	2460
Tag Reverse Primer	AcDx-9077-PHF21B-RT-RP	TGCTATCCTGGCGCAATACAA	21	2461
Downstream PCR Primer	AcDx-9078-PHF21B-PCR-V	TGCTATCCTGGCGCAATACAAGAAAACCCGACGCAACTTACTGrAACAG/3SpC3/	48	2462
NTNG1				
Forward PCR Primer	AcDx-9081-NTNG1-FP	GAGGTTTTTTTCGGAGGCrGGAGA/3SpC3/	23	2463
Reverse PCR Primer	AcDx-9082-NTNG1-RP	GGTGTCGTGGAAAAAACCCCGAAAACCGAArAACCT/3SpC3/	35	2464
Upstream LDR	AcDx-9083-NTNG1-Up	TTGGCAACTCTCCACCCAAGAGGCGAAGGTAATTAAGCAGCrGGGAG/3SpC3/	46	2465
Downstream LDR	AcDx-9084-NTNG1-Dn	/5Phos/GGAGAGTGGCGGGTCGGTTGTATTGCGCCAGGATAGCA	38	2466
Real-Time Probe	AcDx-9085-NTNG1-RT-Pb	/56-FAM/CCAAGCAGC/ZEN/GGAGAGTGGCG/3IABkFQ/	20	2467
Tag Forward Primer	AcDx-9086-NTNG1-RT-FP	TTGGCAACTCTCCACCCAA	19	2468
Tag Reverse Primer	AcDx-9087-NTNG1-RT-RP	TGCTATCCTGGCGCAATACAA	21	2469
Downstream PCR Primer	AcDx-9088-NTNG1-PCR-V	TGCTATCCTGGCGCAATACAAAAAACCGAAAACCCAACGCTGrACCCA/3SpC3/	47	2470
LOC389634				
Forward PCR Primer	AcDx-9091-LOC389634-FP	TAGGTTACGAGGTTTTTCGTCrGTTTC/3SpC3/	26	2471
Reverse PCR Primer	AcDx-9092-LOC389634-RP	GGTGTCGTGGCCGACACCCAAACAAATAACGArCGACG/3SpC3/	37	2472
Upstream LDR	AcDx-9093-LOC389634-Up	TCGACGAATCTGCTCAGACAACGTCGTTTTTATTTTGCGTTTTCGGTATCTCrGGGCC/ 3SpC3/	57	2473
Downstream LDR	AcDx-9094-LOC389634-Dn	/5Phos/GGGATTTTCGCGTTCGCGTTATTTATTTTTTTGTCGTTGAAGCAGCGTCTGA	55	2474

		GCA		
Real-Time Probe	AcDx-9095-LOC389634-RT-Pb	/56-FAM/TTGTATCTC/ZEN/GGGATTTTCGCGTTCGCGTTATTTA/3IABkFQ/	34	2475
Tag Forward Primer	AcDx-9096-LOC389634-RT-FP	TCGACGAATCTGCTCAGACAA	21	2476
Tag Reverse Primer	AcDx-9097-LOC389634-RT-RP	TGCTCAGACGCTGCTTCAA	19	2477
Downstream PCR Primer	AcDx-9098-LOC389634-PCR-V	TGCTCAGACGCTGCTTCAACCGACACCCAAACAAATAACGATGrACAAG/3SpC3/	48	2478
ELMO1-S3				
Forward PCR Primer	AcDx-9101-ELMO1-S3-FP	TTTAGCGTGAGGTCGCrGGCGA/3SpC3/	21	2479
Reverse PCR Primer	AcDx-9102-ELMO1-S3-RP	GGTGTCGTGGAACGACAACCGACGATACAACrAACCA/3SpC3/	36	2480
Upstream LDR	AcDx-9103-ELMO1-S3-Up	TCGACGAATCTGCTCAGACAAGTTTTGTTTTTATTTTGTGTTTATTTTCGTTCGTTCTCrGTTTA/3SpC3/	64	2481
Downstream LDR	AcDx-9104-ELMO1-S3-Dn	/5Phos/GTTCGTTTTAGTATACGTATTTATATTTTGGGTCGGTCGGTTGAAGCAGCGTCTGAGCA	59	2482
Real-Time Probe	AcDx-9105-ELMO1-S3-RT-Pb	/56-FAM/AACGTTCTC/ZEN/GTTCGTTTTAGTATACGTATTTATATTTTGGGTCG/3IABkFQ/	44	2483
Tag Forward Primer	AcDx-9106-ELMO1-S3-RT-FP	TCGACGAATCTGCTCAGACAA	21	2484
Tag Reverse Primer	AcDx-9107-ELMO1-S3-RT-RP	TGCTCAGACGCTGCTTCAA	19	2485
Downstream PCR Primer	AcDx-9108-ELMO1-S3-PCR-V	TGCTCAGACGCTGCTTCAAGACAACCGACGATACAACAACTGrACCGG/3SpC3/	47	2486
BVES-S2				
Forward PCR Primer	AcDx-9111-BVES-S2-FP	CGGTTGTTCGGGCGCrGTAGG/3SpC3/	20	2487
Reverse PCR Primer	AcDx-9112-BVES-S2-RP	GGTGTCGTGGCGCTCCCCTAACTAAATCGAAAArACGAG/3SpC3/	38	2488
Upstream LDR	AcDx-9113-BVES-S2-Up	TTTCCGCCGCTACAACCAACGGGCGCGTAGAATAAGTAGTTATCrGTTGA/3SpC3/	49	2489

Downstream LDR	AcDx-9114-BVES-S2-Dn	/5Phos/GTTAGGGTCGATAAAGTTGTTTTTCGAGGAGTTGAGATTTTGAAGCAGCGTCTGAGCA	58	2490
Real-Time Probe	AcDx-9115-BVES-S2-RT-Pb	/56-FAM/AAAGTTATC/ZEN/GTTAGGGTCGATAAAGTTGTTTTTCGAGG/3IABkFQ/	38	2491
Tag Forward Primer	AcDx-9116-BVES-S2-RT-FP	TTTCCGCCGCTACAACCAA	19	2492
Tag Reverse Primer	AcDx-9117-BVES-S2-RT-RP	TGCTCAGACGCTGCTTCAA	19	2493
Downstream PCR Primer	AcDx-9118-BVES-S2-PCR-V	TGCTCAGACGCTGCTTCAACGCTCCCCTAACTAAATCGAAAAATGrAAAAC/3SpC3/	50	2494
SIM2-S2				
Forward PCR Primer	AcDx-9121-SIM2-S2-FP	GGTGGTTGCGGTCGCrGTTTC/3SpC3/	20	2495
Reverse PCR Primer	AcDx-9122-SIM2-S2-RP	GGTGTCGTGGCGCCGAACCTACGCArCGACA/3SpC3/	30	2496
Upstream LDR	AcDx-9123-SIM2-S2-Up	TCAGTGAAAACACATCCACCCAGCGGTCGCGTTTTTTGTTGTTTTTTGACrGTGCT/3SpC3/	55	2497
Downstream LDR	AcDx-9124-SIM2-S2-Dn	/5Phos/GTGTCGTTTGTTTTCGTAGCGTTCGTCGTCTGGTGAGCAGGGATGAGCA	49	2498
Real-Time Probe	AcDx-9125-SIM2-S2-RT-Pb	/56-FAM/AATTTTGAC/ZEN/GTGTCGTTTGTTTTCGTAGCGTTCG/3IABkFQ/	34	2499
Tag Forward Primer	AcDx-9126-SIM2-S2-RT-FP	TCAGTGAAAACACATCCACCCA	22	2500
Tag Reverse Primer	AcDx-9127-SIM2-S2-RT-RP	TGCTCATCCCTGCTCACCA	19	2501
Downstream PCR Primer	AcDx-9128-SIM2-S2-PCR-V	TGCTCATCCCTGCTCACCACGAACCTACGCACGACGATGrACGAG/3SpC3/	44	2502
ISL2-S2				
Forward PCR Primer	AcDx-9131-ISL2-S2-FP	GGTTTGTTTTTAGTTGGCGGTCrGGTTC/3SpC3/	27	2503
Reverse PCR Primer	AcDx-9132-ISL2-S2-RP	GGTGTCGTGGAACGACGAAAACACCGAAArAAAAG/3SpC3/	34	2504
Upstream LDR	AcDx-9133-ISL2-S2-Up	TCGTAGACTCGCTATCGCCAGGCGGTCGGTTTTTAAGGGATATTTCTCrGATCT/3SpC3/	53	2505
Downstream LDR	AcDx-9134-ISL2-S2-Dn	/5Phos/GATTCGGAGTACGCGGTTTTGGAGTATTAGTTCTGGTGAGCAGGGATGAGCA	52	2506
Real-Time Probe	AcDx-9135-ISL2-S2-RT-Pb	/56-FAM/CCATTTCTC/ZEN/GATTCGGAGTACGCGGTTTTGG/3IABkFQ/	31	2507
Tag Forward Primer	AcDx-9136-ISL2-S2-RT-FP	TCGTAGACTCGCTATCGCCA	20	2508

Tag Reverse Primer	AcDx-9137-ISL2-S2-RT-RP	TGCTCATCCCTGCTCACCA	19	2509
Downstream PCR Primer	AcDx-9138-ISL2-S2-PCR-V	TGCTCATCCCTGCTCACCACACCGAAAAAAAAACAAATAAAAAAAACACGTGrAACTG/3SpC3/	55	2510
SOST-S2				
Forward PCR Primer	AcDx-9141-SOST-S2-FP	GGTGAGGCGTTTGTATTTGTACrGAGGC/3SpC3/	27	2511
Reverse PCR Primer	AcDx-9142-SOST-S2-RP	GGTGTCGTGGCGCGTACAACTACTATATCCCGArUAATG/3SpC3/	38	2512
Upstream LDR	AcDx-9143-SOST-S2-Up	TTGCACCCGCGACATAACCGCGTTTGTATTTGTACGAGGTTATTAAGCrGTACC/3SpC3/	53	2513
Downstream LDR	AcDx-9144-SOST-S2-Dn	/5Phos/GTATTTTGCGCGCGCGCGGGGTAATTCACTCGAACGGAGCA	41	2514
Real-Time Probe	AcDx-9145-SOST-S2-RT-Pb	/56-FAM/CCATTAAGC/ZEN/GTATTTTGCGCGCGCGC/3IABkFQ/	26	2515
Tag Forward Primer	AcDx-9146-SOST-S2-RT-FP	TTGCACCCGCGACATAACC	19	2516
Tag Reverse Primer	AcDx-9147-SOST-S2-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	2517
Downstream PCR Primer	AcDx-9148-SOST-S2-PCR-V	TGCTCCGTTCGAGTGAATTACCCGCGTACAACTACTATATCCCGATAATAAAATGrCCGCA/3SpC3/	60	2518
ZNF781-S2				
Forward PCR Primer	AcDx-9151-ZNF781-S2-FP	CGTTTGTGGGTTGGTAGTTCrGGAAC/3SpC3/	25	2519
Reverse PCR Primer	AcDx-9152-ZNF781-S2-RP	GGTGTCGTGGCTCCGCTACCGCGTCrCTCCG/3SpC3/	30	2520
Upstream LDR	AcDx-9153-ZNF781-S2-Up	TCCGGCCTTTGACGATACCCGGAATATATTTTTTAGAGGTTTTCGCGATCrGACAC/3SpC3/	55	2521
Downstream LDR	AcDx-9154-ZNF781-S2-Dn	/5Phos/GACGTGTTTCGCGTAGGAACGTAGTCGTTTTTCGGTAATTCACTCGAACGGAGCA	55	2522
Real-Time Probe	AcDx-9155-ZNF781-S2-RT-Pb	/56-FAM/AACGCGATC/ZEN/GACGTGTTTCGCGTAGG/3IABkFQ/	26	2523
Tag Forward Primer	AcDx-9156-ZNF781-S2-RT-FP	TCCGGCCTTTGACGATACC	19	2524
Tag Reverse Primer	AcDx-9157-ZNF781-S2-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	2525
Downstream PCR	AcDx-9158-ZNF781-S2-	TGCTCCGTTCGAGTGAATTACCCGTCCTCCAATCGAAAAACGACTATGrUTCCC/3SpC	53	2526

Primer	PCR-V	3/		
ISL2-S3				
Forward PCR Primer	AcDx-9161-ISL2-S3-FP	GTTTAGCGAGTTTGTTTTTTAGAGCrGATTC/3SpC3/	30	2527
Reverse PCR Primer	AcDx-9162-ISL2-S3-RP	GGTGTCGTGGAAATCCCCAACCCGCAAArATCCT/3SpC3/	33	2528
Upstream LDR	AcDx-9163-ISL2-S3-Up	TCCGGCCTTTGACGATACCCGAGTTTGTTTTTTAGAGCGATTTGGATTAACTCrGTTCC /3SpC3/	58	2529
Downstream LDR	AcDx-9164-ISL2-S3-Dn	/5Phos/GTTTTTTAATAGTTGGTGAGGTTTTGTTTTATTCGTTTCGATTTCGGTAATTC ACTCGAACGGAGCA	67	2530
Real-Time Probe	AcDx-9165-ISL2-S3-RT-Pb	/56-FAM/TTTTAACTC/ZEN/GTTTTTTAATAGTTGGTGAGGTTTTGTTTTATTCGT/3IABkFQ/	45	2531
Tag Forward Primer	AcDx-9166-ISL2-S3-RT-FP	TCCGGCCTTTGACGATACC	19	2532
Tag Reverse Primer	AcDx-9167-ISL2-S3-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	2533
Downstream PCR Primer	AcDx-9168-ISL2-S3-PCR-V	TGCTCCGTTCGAGTGAATTACCCAACCCGCAAAATCCCGAAATTGrAAACA/3SpC3/	50	2534
PAX7				
Forward PCR Primer	AcDx-9171-PAX7-FP	GAGGTAGATATTTTTATATAGTCGGGATTTCrGAGAA/3SpC3/	36	2535
Reverse PCR Primer	AcDx-9172-PAX7-RP	GGTGTCGTGGTCCTTTAATAAATAAACCGAATCCGCrCGAAC/3SpC3/	41	2536
Upstream LDR	AcDx-9173-PAX7-Up	TCGCGGAAAGTCCCAGTAACGATATTTTTATATAGTCGGGATTTCGAGAGTAATCrGT GAA/3SpC3/	60	2537
Downstream LDR	AcDx-9174-PAX7-Dn	/5Phos/GTGGGCGATTTGGGAGTCGGGGTTGGCCTGTAAGCGTTCCA	41	2538
Real-Time Probe	AcDx-9175-PAX7-RT-Pb	/56-FAM/TTAGTAATC/ZEN/GTGGGCGATTTGGGAGTC/3IABkFQ/	27	2539
Tag Forward Primer	AcDx-9176-PAX7-RT-FP	TCGCGGAAAGTCCCAGTAAC	20	2540
Tag Reverse Primer	AcDx-9177-PAX7-RT-RP	TGGAACGCTTACAGGCCAAC	20	2541
Downstream PCR Primer	AcDx-9178-PAX7-PCR-V	TGGAACGCTTACAGGCCAACTCCTTTAATAAATAAACCGAATCCGCTGrAATCT/3SpC 3/	53	2542

BARX1				
Forward PCR Primer	AcDx-9181-BARX1-FP	CGTTTTTTTAGGTTTATTAGTTGTAGTTCrGGTGG/3SpC3/	34	2543
Reverse PCR Primer	AcDx-9182-BARX1-RP	GGTGTCGTGGTACAACTCCGCGAAAAACTAAArAACGG/3SpC3/	37	2544
Upstream LDR	AcDx-9183-BARX1-Up	TGAACGCTCAAACACGTGAACGGTTTATTAGTTGTAGTTCGGTGAATATAGCGCrGGTCC/3SpC3/	59	2545
Downstream LDR	AcDx-9184-BARX1-Dn	/5Phos/GGTTTCGACGTTTTTTTTTGGTTTTGGTGTTTGGGTTGGCCTGTAAGCGTTCCA	54	2546
Real-Time Probe	AcDx-9185-BARX1-RT-Pb	/56-FAM/CCATAGCGC/ZEN/GGTTTCGACGTTTTTTTTTGGTTTTG/3IABkFQ/	35	2547
Tag Forward Primer	AcDx-9186-BARX1-RT-FP	TGAACGCTCAAACACGTGAAC	21	2548
Tag Reverse Primer	AcDx-9187-BARX1-RT-RP	TGGAACGCTTACAGGCCAAC	20	2549
Downstream PCR Primer	AcDx-9188-BARX1-PCR-V	TGGAACGCTTACAGGCCAACTACAACTCCGCGAAAAACTAAAAATGrACAAG/3SpC3/	51	2550
HOXC13				
Forward PCR Primer	AcDx-9191-HOXC13-FP	GTTTATTATTAAAGAGAAGCGTCGGCrGTATC/3SpC3/	31	2551
Reverse PCR Primer	AcDx-9192-HOXC13-RP	GGTGTCGTGGACCTTCTTCTCTTTAACCCGCrCGATC/3SpC3/	36	2552
Upstream LDR	AcDx-9193-HOXC13-Up	TACACGTGGATATCTCCGACCGAGAAGCGTCGGCGTATTTTCGTTATCACrGAACC/3SpC3/	55	2553
Downstream LDR	AcDx-9194-HOXC13-Dn	/5Phos/GAATTTTTTTGAGCGTTAGGTAATTATTTGGTTTTAGAATCGGGTGCTAGTCACACAGTTCCA	63	2554
Real-Time Probe	AcDx-9195-HOXC13-RT-Pb	/56-FAM/TTTTATCAC/ZEN/GAATTTTTTTGAGCGTTAGGTAATTATTTGGTTTTA/3IABkFQ/	45	2555
Tag Forward Primer	AcDx-9196-HOXC13-RT-FP	TACACGTGGATATCTCCGACC	21	2556
Tag Reverse Primer	AcDx-9197-HOXC13-RT-RP	TGGAACTGTGTGACTAGCACC	21	2557
Downstream PCR Primer	AcDx-9198-HOXC13-PCR-V	TGGAACTGTGTGACTAGCACCACCTTCTTCTCTTTAACCCGCTGrATTCC/3SpC3/	49	2558
TRPV3				

Forward PCR Primer	AcDx-9201-TRPV3-FP	GTTTTTTTGGCGTGCGCrGTTGG/3SpC3/	22	2559
Reverse PCR Primer	AcDx-9202-TRPV3-RP	GGTGTCGTGGCGCCATCGAACGACGACrAAAAG/3SpC3/	32	2560
Upstream LDR	AcDx-9203-TRPV3-Up	TAGCATTCGAGAACGCACCGCGTGCGCGTTGACATCrGGCAC/3SpC3/	41	2561
Downstream LDR	AcDx-9204-TRPV3-Dn	/5Phos/GGCGTCGGCGGCGATGAGTAGGGTGCTAGTCACACAGTTCCA	42	2562
Real-Time Probe	AcDx-9205-TRPV3-RT-Pb	/56-FAM/AATGACATC/ZEN/GGCGTCGGCGG/3IABkFQ/	20	2563
Tag Forward Primer	AcDx-9206-TRPV3-RT-FP	TAGCATTCGAGAACGCACC	19	2564
Tag Reverse Primer	AcDx-9207-TRPV3-RT-RP	TGGAACTGTGTGACTAGCACC	21	2565
Downstream PCR Primer	AcDx-9208-TRPV3-PCR-V	TGGAACTGTGTGACTAGCACCCAAAAAAACATCGCAACCCTACTCATTGrCCGCT/3SpC3/	54	2566
KIAA1239				
Forward PCR Primer	AcDx-9211-KIAA1239-FP	CGTCGGTCGTAGCGTTCrGGGTC/3SpC3/	22	2567
Reverse PCR Primer	AcDx-9212-KIAA1239-RP	GGTGTCGTGGAACGCTAACGAACAAAAAAACGArACGAG/3SpC3/	38	2568
Upstream LDR	AcDx-9213-KIAA1239-Up	TAACCGGGCCTAAAGTGACAGTCGTAGCGTTCGGGTTTTTATTGGCrGTTGG/3SpC3/	51	2569
Downstream LDR	AcDx-9214-KIAA1239-Dn	/5Phos/GTTAATTTTGAAGGTACGTTTTTCGTTCGGGTTTGTTGTTACGTGATCTCCCTCTCCA	58	2570
Real-Time Probe	AcDx-9215-KIAA1239-RT-Pb	/56-FAM/AATATTGGC/ZEN/GTTAATTTTGAAGGTACGTTTTTCGTTCG/3IABkFQ/	38	2571
Tag Forward Primer	AcDx-9216-KIAA1239-RT-FP	TAACCGGGCCTAAAGTGACA	20	2572
Tag Reverse Primer	AcDx-9217-KIAA1239-RT-RP	TGGAGAGGGAGATCACGTAACA	22	2573
Downstream PCR Primer	AcDx-9218-KIAA1239-PCR-V	TGGAGAGGGAGATCACGTAACAAACGCTAACGAACAAAAAAACGAATGrAACAG/3SpC3/	53	2574
CANCR62				
Forward PCR Primer	AcDx-9221-CANCR62-FP	CGAGGTAGATAATGGCGCrGGAGA/3SpC3/	23	2575
Reverse PCR Primer	AcDx-9222-CANCR62-RP	GGTGTCGTGGACATCGAAATAAACGAACGCCrCCGCA/3SpC3/	36	2576
Upstream LDR	AcDx-9223-CANCR62-Up	TCGATGGTCAATGAGCTTCACAATAATGGCGCGGAGGAGCGCrGGAGC/3SpC3/	47	2577

Downstream LDR	AcDx-9224-CANCR62-Dn	/5Phos/GGAATTTGGGATTAAATTTGGCGTTGTTTTTGTAGGGTGTTACGTGATCTCCCTCTCCA	59	2578
Real-Time Probe	AcDx-9225-CANCR62-RT-Pb	/56-FAM/TTGGAGCGC/ZEN/GGAATTTGGGATTAAATTTGGC/3IABkFQ/	31	2579
Tag Forward Primer	AcDx-9226-CANCR62-RT-FP	TCGATGGTCAATGAGCTTCACA	22	2580
Tag Reverse Primer	AcDx-9227-CANCR62-RT-RP	TGGAGAGGGAGATCACGTAACA	22	2581
Downstream PCR Primer	AcDx-9228-CANCR62-PCR-V	TGGAGAGGGAGATCACGTAACAGAAATAAACGAACGCCCCGTGrCCCTG/3SpC3/	48	2582
CANCR63				
Forward PCR Primer	AcDx-9231-CANCR63-FP	TGTGAGTTGGTGTGTTTAGGTCrGGGTC/3SpC3/	27	2583
Reverse PCR Primer	AcDx-9232-CANCR63-RP	GGTGTCGTGGTTTACAACGCCTTAAAATATAAACGTTrUCCCT/3SpC3/	42	2584
Upstream LDR	AcDx-9233-CANCR63-Up	TCGATGGTCAATGAGCTTCACAAGGTCGGGTTGGGTTGACGCrGGACG/3SpC3/	47	2585
Downstream LDR	AcDx-9234-CANCR63-Dn	/5Phos/GGATACGGGAGGTAGTAGGTATTCGGAGTGTTACGTGATCTCCCTCTCCA	50	2586
Real-Time Probe	AcDx-9235-CANCR63-RT-Pb	/56-FAM/TTTTGACGC/ZEN/GGATACGGGAGGTAGTAGGTATTC/3IABkFQ/	33	2587
Tag Forward Primer	AcDx-9236-CANCR63-RT-FP	TCGATGGTCAATGAGCTTCACA	22	2588
Tag Reverse Primer	AcDx-9237-CANCR63-RT-RP	TGGAGAGGGAGATCACGTAACA	22	2589
OTX2OS1				
Forward PCR Primer	AcDx-9241-OTX2OS1-FP	TTATTTTTTGGTAAGTGTGGCGCrGGGAC/3SpC3/	28	2590
Reverse PCR Primer	AcDx-9242-OTX2OS1-RP	GGTGTCGTGGGAAACGACGCAACTCGAArACCCA/3SpC3/	33	2591
Upstream LDR	AcDx-9243-OTX2OS1-Up	TCTCGGGACCACAATACGAACGGCGCGAGGTTGAACrGTTCC/3SpC3/	41	2592
Downstream LDR	AcDx-9244-OTX2OS1-Dn	/5Phos/GTTTTTGTCGGCGGGTCGTCGGGTTACGCTAAGCTGGTGCCA	42	2593
Real-Time Probe	AcDx-9245-OTX2OS1-RT-	/56-FAM/TTGTTGAAC/ZEN/GTTTTTGTCGGCGGGTCG/3IABkFQ/	27	2594

	Pb			
Tag Forward Primer	AcDx-9246-OTX2OS1-RT-FP	TCTCGGGACCACAATACGAAC	21	2595
Tag Reverse Primer	AcDx-9247-OTX2OS1-RT-RP	TGGCACCAGCTTAGCGTAAC	20	2596
Downstream PCR Primer	AcDx-9248-OTX2OS1-PCR-V	TGGCACCAGCTTAGCGTAACCGACGCAACTCGAAACCTGrACGAT/3SpC3/	44	2597
CANCR64				
Forward PCR Primer	AcDx-9251-CANCR64-FP	CGTTTTAGTTTTGTTGTAATTTTGTTTTCrGTTAA/3SpC3/	34	2598
Reverse PCR Primer	AcDx-9252-CANCR64-RP	GGTGTCGTGGCTCCAAACGCGAAATAACGCrUAAAG/3SpC3/	35	2599
Upstream LDR	AcDx-9253-CANCR64-Up	TTGCTGTGCGCGGTAGAACGTTGTAATTTTGTTTTCGTTAGATAGATTAGTTGGAGTCrGTTGA/3SpC3/	63	2600
Downstream LDR	AcDx-9254-CANCR64-Dn	/5Phos/GTTAGTTTTAGTTTGGAGTTTCGTTTTACGGAGTATTTTTGTTACGCTAAGCTGGTGCCA	60	2601
Real-Time Probe	AcDx-9255-CANCR64-RT-Pb	/56-FAM/AATGGAGTC/ZEN/GTTAGTTTTAGTTTGGAGTTTCGTTTTACG/3IABkFQ/	39	2602
Tag Forward Primer	AcDx-9256-CANCR64-RT-FP	TTGCTGTGCGCGGTAGAAC	19	2603
Tag Reverse Primer	AcDx-9257-CANCR64-RT-RP	TGGCACCAGCTTAGCGTAAC	20	2604
VAX1				
Forward PCR Primer	AcDx-9261-VAX1-FP	TAGTCGGTAGCGGTAGTAGTCrGTAGC/3SpC3/	26	2605
Reverse PCR Primer	AcDx-9262-VAX1-RP	GGTGTCGTGGACCTAATACCCTAACCCCGTArCCCAG/3SpC3/	36	2606
Upstream LDR	AcDx-9263-VAX1-Up	TCTCGATTACGCTCCGCACGTAGCGGTAGTAGTCGTAGTAGTTGCrGGACC/3SpC3/	50	2607
Downstream LDR	AcDx-9264-VAX1-Dn	/5Phos/GGATTTAGAGTAAACGTTTTGTGTTTTGGAGTTAGAATTAGTTTTTTGTGTGTAGCTTAGACATGGCCA	69	2608
Real-Time Probe	AcDx-9265-VAX1-RT-Pb	/56-	44	2609

		FAM/TTTAGTTGC/ZEN/GGATTTAGAGTAAACGTTTTGTGTTTTGGAGTTAG/3IABkFQ/		
Tag Forward Primer	AcDx-9266-VAX1-RT-FP	TCTCGATTACGCTCCGCAC	19	2610
Tag Reverse Primer	AcDx-9267-VAX1-RT-RP	TGGCCATGTCTAAGCTACACAC	22	2611
CANCR65				
Forward PCR Primer	AcDx-9271-CANCR65-FP	ATTTTTTTAGGTTTCGCGGGCrGTTTC/3SpC3/	26	2612
Reverse PCR Primer	AcDx-9272-CANCR65-RP	GGTGTCGTGGGCCCGCCTTCCCAAACrUAAAG/3SpC3/	31	2613
Upstream LDR	AcDx-9273-CANCR65-Up	TCACAGAGACTTGCCGATCACGCGTTTTTTGGGTGGGTTTTGCrGGTCC/3SpC3/	48	2614
Downstream LDR	AcDx-9274-CANCR65-Dn	/5Phos/GGTTTTTTTAATTTTGTTTAGCGTGTGTGCGCGGGTGTGTAGCTTAGACATGGCCA	56	2615
Real-Time Probe	AcDx-9275-CANCR65-RT-Pb	/56-FAM/TTGTTTTGC/ZEN/GGTTTTTTTAATTTTGTTTAGCGTGTGTG/3IABkFQ/	38	2616
Tag Forward Primer	AcDx-9276-CANCR65-RT-FP	TCACAGAGACTTGCCGATCAC	21	2617
Tag Reverse Primer	AcDx-9277-CANCR65-RT-RP	TGGCCATGTCTAAGCTACACAC	22	2618
Downstream PCR Primer	AcDx-9278-CANCR65-PCR-V	TGGCCATGTCTAAGCTACACACTCCCAAACTAAAAAAAAAAAATAACTAAACCGTGrCACAT/3SpC3/	60	2619
ZNF781-S3				
Forward PCR Primer	AcDx-9281-ZNF781-S3-FP	GTTCGGAATATATTTTTTAGAGGTTTTCrGCGGC/3SpC3/	33	2620
Reverse PCR Primer	AcDx-9282-ZNF781-S3-RP	GGTGTCGTGGCTCGCTTTAAACAACTCCGCTArCCGCA/3SpC3/	37	2621
Upstream LDR	AcDx-9283-ZNF781-S3-Up	TCACAGAGACTTGCCGATCACTATTTTTTAGAGGTTTTCGCGGTCAACrGTGCC/3SpC3/	53	2622
Downstream LDR	AcDx-9284-ZNF781-S3-Dn	/5Phos/GTGTTTCGCGTAGGAACGTAGTCGTTTTTCGGTGTGTAGCTTAGACATGGCCA	53	2623
Real-Time Probe	AcDx-9285-ZNF781-S3-RT-Pb	/56-FAM/TTGGTCAAC/ZEN/GTGTTTCGCGTAGGAACGTAG/3IABkFQ/	30	2624

Tag Forward Primer	AcDx-9286-ZNF781-S3-RT-FP	TCACAGAGACTTGCCGATCAC	21	2625
Tag Reverse Primer	AcDx-9287-ZNF781-S3-RT-RP	TGGCCATGTCTAAGCTACACAC	22	2626
Downstream PCR Primer	AcDx-9288-ZNF781-S3-PCR-V	TGGCCATGTCTAAGCTACACACCGCGTCCTCCAATCGAAAAATGrACTAT/3SpC3/	49	2627
ZNF397OS				
Forward PCR Primer	AcDx-9291-ZNF397OS-FP	GTTTTTTTAGGAGTTCGGAGGATCrGTAGC/3SpC3/	29	2628
Reverse PCR Primer	AcDx-9292-ZNF397OS-RP	GGTGTCGTGGTACGATACCCCCGCCAArATCTG/3SpC3/	32	2629
Upstream LDR	AcDx-9293-ZNF397OS-Up	TTCGTCCCTGCACGCTAACGATCGTAGTTTTGTGGTAGGTGCrGGGCT/3SpC3/	47	2630
Downstream LDR	AcDx-9294-ZNF397OS-Dn	/5Phos/GGGTCGTGTTTCGTAGGAGGAGCGGTTCCATCACCGTTAGGCCA	44	2631
Real-Time Probe	AcDx-9295-ZNF397OS-RT-Pb	/56-FAM/TTTAGGTGC/ZEN/GGGTCGTGTTTCGTAGGAG/3IABkFQ/	28	2632
Tag Forward Primer	AcDx-9296-ZNF397OS-RT-FP	TTCGTCCCTGCACGCTAAC	19	2633
Tag Reverse Primer	AcDx-9297-ZNF397OS-RT-RP	TGGCCTAACGGTGATGGAAC	20	2634
Downstream PCR Primer	AcDx-9298-ZNF397OS-PCR-V	TGGCCTAACGGTGATGGAACCCAAATCTAAACCGAACCCGTGrCCCCC/3SpC3/	47	2635
CANCR66				
Forward PCR Primer	AcDx-9301-CANCR66-FP	TTAGTTATTTTATTTAACGTCGTTTTTATTTCrGTTTC/3SpC3/	37	2636
Reverse PCR Primer	AcDx-9302-CANCR66-RP	GGTGTCGTGGAATAAACGCGCCCTACGCrGCCGT/3SpC3/	33	2637
Upstream LDR	AcDx-9303-CANCR66-Up	TGATGCTGGCAAACCCTAGAACCGTCGTTTTTATTTCGTTTTCGTTTTAATAATGCTCrGGCAG/3SpC3/	63	2638
Downstream LDR	AcDx-9304-CANCR66-Dn	/5Phos/GGCGATTTTCGGTTTTGCGTTTTATTTTTGCGGGTTCCATCACCGTTAGGCCA	53	2639
Real-Time Probe	AcDx-9305-CANCR66-RT-Pb	/56-FAM/CCAATGCTC/ZEN/GGCGATTTTCGGTTTTGCG/3IABkFQ/	28	2640
Tag Forward Primer	AcDx-9306-CANCR66-RT-	TGATGCTGGCAAACCCTAGAAC	22	2641

	FP			
Tag Reverse Primer	AcDx-9307-CANCR66-RT-RP	TGGCCTAACGGTGATGGAAC	20	2642
Downstream PCR Primer	AcDx-9308-CANCR66-PCR-V	TGGCCTAACGGTGATGGAACCGCTCGACTTCTACCGCTGrCAAAG/3SpC3/	44	2643
ZNF542-S2				
Forward PCR Primer	AcDx-9311-ZNF542-S2-FP	GAATTTTTTTTATTAGGTTTCGCGTCrGAGGC/3SpC3/	31	2644
Reverse PCR Primer	AcDx-9312-ZNF542-S2-RP	GGTGTCGTGGAATAACCGAAATACAAAAACGTACArUAACC/3SpC3/	40	2645
Upstream LDR	AcDx-9313-ZNF542-S2-Up	TGATGCTGGCAAACCCTAGAACGCGTCGAGGTTTTACGGAGCrGTTGG/3SpC3/	47	2646
Downstream LDR	AcDx-9314-ZNF542-S2-Dn	/5Phos/GTTAATACGTACGCGTATTTTGTGTTTTAGGTAGTTCGTTTTGTTCCATCACC GTTAGGCCA	62	2647
Real-Time Probe	AcDx-9315-ZNF542-S2-RT-Pb	/56-FAM/AAACGGAGC/ZEN/GTTAATACGTACGCGTATTTTGTGTTTTAG/3IABkFQ/	39	2648
Tag Forward Primer	AcDx-9316-ZNF542-S2-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	2649
Tag Reverse Primer	AcDx-9317-ZNF542-S2-RT-RP	TGGCCTAACGGTGATGGAAC	20	2650
Downstream PCR Primer	AcDx-9318-ZNF542-S2-PCR-V	TGGCCTAACGGTGATGGAACCCGAAATACAAAAACGTACATAACTAAAAAATGrAAC TG/3SpC3/	58	2651
ADAMTS16-S2				
Forward PCR Primer	AcDx-9321-ADAMTS16-S2-FP	GCGTTTTTAAAGGTAGGTTCGCrGGTTC/3SpC3/	27	2652
Reverse PCR Primer	AcDx-9322-ADAMTS16-S2-RP	GGTGTCGTGGGATCTAACCGCGCCTCTATACrCCGAG/3SpC3/	36	2653
Upstream LDR	AcDx-9323-ADAMTS16-S2-Up	TTTTCGGCATCCGCTTCCACGTTTTTAAAGGTAGGTTCGCGGTTTTTTCACrGTTCC/3SpC3/	56	2654
Downstream LDR	AcDx-9324-ADAMTS16-S2-Dn	/5Phos/GTTTTGGCGTTCGGTTTTTCGTATTTAGTGTTTTTATAGTTGTGTGGTTAACA GAGGACAGGCCA	65	2655
Real-Time Probe	AcDx-9325-ADAMTS16-S2-	/56-FAM/AATTTTCAC/ZEN/GTTTTGGCGTTCGGTTTTTCGTATTTAGTG/3IABkFQ/	39	2656

	RT-Pb			
Tag Forward Primer	AcDx-9326-ADAMTS16-S2-RT-FP	TTTTCGGCATCCGCTTCCA	19	2657
Tag Reverse Primer	AcDx-9327-ADAMTS16-S2-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	2658
Downstream PCR Primer	AcDx-9328-ADAMTS16-S2-PCR-V	TGGCCTGTCCTCTGTTAACCAATCTAACCGCGCCTCTATACCTGrAACAT/3SpC3/	49	2659
LHX5				
Forward PCR Primer	AcDx-9331-LHX5-FP	CGGTGAATTTGAGGTTGTTCrGCGGT/3SpC3/	25	2660
Reverse PCR Primer	AcDx-9332-LHX5-RP	GGTGTCGTGGCTAACGACCTCTAACCCCGArCTCGG/3SpC3/	35	2661
Upstream LDR	AcDx-9333-LHX5-Up	TCCGGGTATACACTGTCCCAGGTTGTTCGCGGCGCGCrGGGCT/3SpC3/	42	2662
Downstream LDR	AcDx-9334-LHX5-Dn	/5Phos/GGGTCGGCGAGCGGCGTGGTTAACAGAGGACAGGCCA	37	2663
Real-Time Probe	AcDx-9335-LHX5-RT-Pb	/56-FAM/TTGGCGCGC/ZEN/GGGTCGGC/3IABkFQ/	17	2664
Tag Forward Primer	AcDx-9336-LHX5-RT-FP	TCCGGGTATACACTGTCCCA	20	2665
Tag Reverse Primer	AcDx-9337-LHX5-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	2666
Downstream PCR Primer	AcDx-9338-LHX5-PCR-V	TGGCCTGTCCTCTGTTAACCACGACTCGACGCCGCTAAAAATGrCTAAG/3SpC3/	48	2667
CANCR67				
Forward PCR Primer	AcDx-9341-CANCR67-FP	GGGTCGAGGTAGGATTTCrGTTTT/3SpC3/	23	2668
Reverse PCR Primer	AcDx-9342-CANCR67-RP	GGTGTCGTGGCAAAACCCTAACCCTCGCCrCCTCA/3SpC3/	34	2669
Upstream LDR	AcDx-9343-CANCR67-Up	TGCGACTCTATTCACGTCCAAGTAGGATTTCGTTTCGGGAGGCrGTAAA/3SpC3/	48	2670
Downstream LDR	AcDx-9344-CANCR67-Dn	/5Phos/GTAGGTGCGGCGTGAGGATCGCTTGCTATTTGGTGTACCGCCA	43	2671
Real-Time Probe	AcDx-9345-CANCR67-RT-Pb	/56-FAM/AAGGGAGGC/ZEN/GTAGGTGCGGC/3IABkFQ/	20	2672
Tag Forward Primer	AcDx-9346-CANCR67-RT-FP	TGCGACTCTATTCACGTCCAA	21	2673
Tag Reverse Primer	AcDx-9347-CANCR67-RT-RP	TGGCGGTACACCAAATAGCAA	21	2674

Downstream PCR Primer	AcDx-9348-CANCR67-PCR-V	TGGCGGTACACCAAATAGCAACCCTCCCTCCTCAAAACGTGrATCCC/3SpC3/	46	2675
ANK1				
Forward PCR Primer	AcDx-9351-ANK1-FP	GGCGAAGGTAGTTTCGGACrGCGGG/3SpC3/	24	2676
Reverse PCR Primer	AcDx-9352-ANK1-RP	GGTGTCGTGGCCGCGACGACAACGCrCAGCT/3SpC3/	30	2677
Upstream LDR	AcDx-9353-ANK1-Up	TTGTGCAGAGCGAACAACAAGTTTCGGACGCGGAGAGCrGAGAA/3SpC3/	43	2678
Downstream LDR	AcDx-9354-ANK1-Dn	/5Phos/GAGGAGCGGTTGTTCGCGGTTTGCTATTTGGTGTACCGCCA	41	2679
Real-Time Probe	AcDx-9355-ANK1-RT-Pb	/56-FAM/TTGGAGAGC/ZEN/GAGGAGCGGTTGTTC/3IABkFQ/	24	2680
Tag Forward Primer	AcDx-9356-ANK1-RT-FP	TTGTGCAGAGCGAACAACAA	20	2681
Tag Reverse Primer	AcDx-9357-ANK1-RT-RP	TGGCGGTACACCAAATAGCAA	21	2682
Downstream PCR Primer	AcDx-9358-ANK1-PCR-V	TGGCGGTACACCAAATAGCAAACAACGCCATCCCGAACTGrCGAAT/3SpC3/	45	2683
C17orf46-S1				
Forward PCR Primer	AcDx-9361-C17orf46-S1-FP	GGTAGGGAGTCGTATTTAGGTTTCrGGGCC/3SpC3/	29	2684
Reverse PCR Primer	AcDx-9362-C17orf46-S1-RP	GGTGTCGTGGCGAAAAAAAAAAAATTCTAACTTCGAACrCCGCG/3SpC3/	43	2685
Upstream LDR	AcDx-9363-C17orf46-S1-Up	TCCAAACGATTAGGAGCGTCAACGGACGAGGGACGGCTCrGAGCA/3SpC3/	44	2686
Downstream LDR	AcDx-9364-C17orf46-S1-Dn	/5Phos/GAGTGAGGCGTTTTTCGAAGGTTGCGTTGGACAGAGGTATACGCCCA	47	2687
Real-Time Probe	AcDx-9365-C17orf46-S1-RT-Pb	/56-FAM/TTACGGCTC/ZEN/GAGTGAGGCGTTTTTCG/3IABkFQ/	26	2688
Tag Forward Primer	AcDx-9366-C17orf46-S1-RT-FP	TCCAAACGATTAGGAGCGTCAA	22	2689
Tag Reverse Primer	AcDx-9367-C17orf46-S1-RT-RP	TGGGCGTATACCTCTGTCCAA	21	2690
Downstream PCR Primer	AcDx-9368-C17orf46-S1-PCR-V	TGGGCGTATACCTCTGTCCAACGAAAAAAAAAAAATTCTAACTTCGAACCTGrCAACT/3SpC3/	57	2691

IDT Abbreviation	Modifications
/5Phos/	5' Phosphorylation
rX (X=A,C,G,U)	RNA Base
/3spC3/	3' C3 DNA Spacer
/56-FAM/	5' 6-FAM™ Fluorescent Tag
/Zen/	Internal Quencher
/3IABkFQ/	3' Iowa Black® FQ Quencher

Table 54. Primers for use in Step 2 of the Group 2- 64-marker assay, with average sensitivities of 50%, to detect and identify breast, endometrial, ovarian, cervical, and uterine cancers.

Site	Primer Name	Sequence	Length	Seq. ID No.
Prefered Group 2 Markers				
DLGAP1-S1				
Forward PCR Primer	AcDx-7001-DLGAP1-S1-FP	GGAGATGTAGATTTCGATGTTTTCrGGCGC/3SpC3/	29	2692
Reverse PCR Primer	AcDx-7002-DLGAP1-S1-RP	GGTGTCGTGGAAAACCCGCAAACGCCrUAATG/3SpC3/	31	2693
Upstream LDR	AcDx-7003-DLGAP1-S1-Up	TCCCTTAGAGAGAACGCCCAGTTTTCGGTGGTCGAGTTTTAGCrGGATA/3SpC3/	48	2694
Downstream LDR	AcDx-7004-DLGAP1-S1-Dn	/5Phos/GGACGGACGCGGCGCGTGGTGACGTACGAGTGTTCTTA	38	2695
Real-Time Probe	AcDx-7005-DLGAP1-S1-RT-Pb	/56-FAM/CCTTTTAGC/ZEN/GGACGGACGCGGC/3IABkFQ/	22	2696
Tag Forward Primer	AcDx-7006-DLGAP1-S1-RT-FP	TCCCTTAGAGAGAACGCCCA	20	2697
Tag Reverse Primer	AcDx-7007-DLGAP1-S1-RT-RP	TAAGAACACTCGTACGTCACCA	22	2698
Downstream PCR Primer	AcDx-7008-DLGAP1-S1-PCR-V	TAAGAACACTCGTACGTCACCACCCGCAAACGCCTAATAACTGrCCAAG/3SpC3/	48	2699

VAMP5				
Forward PCR Primer	AcDx-7011-VAMP5-FP	TCGGGAGGGTTCGATTTTACrGGATC/3SpC3/	25	2700
Reverse PCR Primer	AcDx-7012-VAMP5-RP	GGTGTCGTGGCGAAAAACGCGCCTTCCrGCGAG/3SpC3/	32	2701
Upstream LDR	AcDx-7013-VAMP5-Up	TCTCATACCAGACGCGGTAACGGAGGGTTCGATTTTACGGATTTAGATCrGTTAC/3SpC3/	54	2702
Downstream LDR	AcDx-7014-VAMP5-Dn	/5Phos/GTTGTGGTTTATCGTTTTCGATTTGATTTGGTTTTTGTCGGTTCGTGTCGCTGTGCTTA	59	2703
Real-Time Probe	AcDx-7015-VAMP5-RT-Pb	/56-FAM/CCTTAGATC/ZEN/GTTGTGGTTTATCGTTTTCGATTTGATTTG/3IABkFQ/	39	2704
Tag Forward Primer	AcDx-7016-VAMP5-RT-FP	TCTCATACCAGACGCGGTAAC	21	2705
Tag Reverse Primer	AcDx-7017-VAMP5-RT-RP	TAAGCACAGCGACACGAAC	19	2706
Downstream PCR Primer	AcDx-7018-VAMP5-PCR-V	TAAGCACAGCGACACGAACCGCCTTCCGCGAATAATGrACAAG/3SpC3/	42	2707
CaNCR20				
Forward PCR Primer	AcDx-7021-CaNCR20-FP	AGTTGCGGGTCGGGTArGTGAC/3SpC3/	21	2708
Reverse PCR Primer	AcDx-7022-CaNCR20-RP	GGTGTCGTGGCTCTAAAATAAAATACGCAATAAACAACCrAAACA/3SpC3/	44	2709
Upstream LDR	AcDx-7023-CaNCR20-Up	TTTTCGGCGGCAGCTAAACCGGGTCGGGTAGTGATTGATAGATCrGGGCG/3SpC3/	49	2710
Downstream LDR	AcDx-7024-CaNCR20-Dn	/5Phos/GGGAATAGGGTTTTTCGTTTTTTTTTCGTTTGGGTGTTCGTGTCGCTGTGCTTA	54	2711
Real-Time Probe	AcDx-7025-CaNCR20-RT-Pb	/56-FAM/CCATAGATC/ZEN/GGGAATAGGGTTTTTCGTTTTTTTTTCG/3IABkFQ/	37	2712
Tag Forward Primer	AcDx-7026-CaNCR20-RT-FP	TTTTCGGCGGCAGCTAAAC	19	2713
Tag Reverse Primer	AcDx-7027-CaNCR20-RT-RP	TAAGCACAGCGACACGAAC	19	2714
Downstream PCR Primer	AcDx-7028-CaNCR20-PCR-V	TAAGCACAGCGACACGAACCTCTAAAATAAAATACGCAATAAACAACCAAATGrCCCCT/3SpC3/	58	2715
ATP6V1B1				
Forward PCR Primer	AcDx-7031-ATP6V1B1-FP	TTTTTCGTTCGATTTTTTTTTTCGCrGTTTC/3SpC3/	30	2716

Reverse PCR Primer	AcDx-7032-ATP6V1B1-RP	GGTGTCGTGGACATAAAAATACAAATACTCCCGTCAArUATAG/3SpC3/	42	2717
Upstream LDR	AcDx-7033-ATP6V1B1-Up	TGATGCTGGCAAACCCTAGAACTCGATTTTTTTTTTCGCGTTTTATGTTTTTTTCTCrGATCC/3SpC3/	62	2718
Downstream LDR	AcDx-7034-ATP6V1B1-Dn	/5Phos/GATTTTCGGTGTTGCGGAAGAATTGAAGGTTGGTTCCATCACCGTTAGGCCA	52	2719
Real-Time Probe	AcDx-7035-ATP6V1B1-RT-Pb	/56-FAM/CCTTTTCTC/ZEN/GATTTTCGGTGTTGCGGAAG/3IABkFQ/	29	2720
Tag Forward Primer	AcDx-7036-ATP6V1B1-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	2721
Tag Reverse Primer	AcDx-7037-ATP6V1B1-RT-RP	TGGCCTAACGGTGATGGAAC	20	2722
Downstream PCR Primer	AcDx-7038-ATP6V1B1-PCR-V	TGGCCTAACGGTGATGGAACCAAATACTCCCGTCAAAATAAAACATTGrCAACT/3SpC3/	53	2723
CaNCR21				
Forward PCR Primer	AcDx-7041-CaNCR21-FP	AAATTCGTTCGGTTTATTTAAGTTTCrGTTGC/3SpC3/	31	2724
Reverse PCR Primer	AcDx-7042-CaNCR21-RP	GGTGTCGTGGCCTTTAACTTCCTCCTCGATTCCrCCCAG/3SpC3/	38	2725
Upstream LDR	AcDx-7043-CaNCR21-Up	TTAGCCGCCAAACGTACCACGTTTTTTAGGGTCGTTTTTGGGCACrGGGTA/3SpC3/	50	2726
Downstream LDR	AcDx-7044-CaNCR21-Dn	/5Phos/GGGCGGATTTTTCGTTAATATTTTGTTTGTAAGATTTTTTATTGTGGGCAGGAACACGATAGTA	64	2727
Real-Time Probe	AcDx-7045-CaNCR21-RT-Pb	/56-FAM/AATGGGCAC/ZEN/GGGCGGATTTTTCGTTA/3IABkFQ/	26	2728
Tag Forward Primer	AcDx-7046-CaNCR21-RT-FP	TTAGCCGCCAAACGTACCA	19	2729
Tag Reverse Primer	AcDx-7047-CaNCR21-RT-RP	TACTATCGTGTTCCTGCCCA	20	2730
UBTF				
Forward PCR Primer	AcDx-7051-UBTF-FP	GGCGTTTTCGTCGGCrGGGTG/3SpC3/	20	2731
Reverse PCR Primer	AcDx-7052-UBTF-RP	GGTGTCGTGGTCGGTTGTTGGGCGTrAAAAT/3SpC3/	30	2732
Upstream LDR	AcDx-7053-UBTF-Up	TAGCAGCTGAACAACCCAACGGTGTAGATGTTTTTCGTTTGAGGCTCrGTTCC/3SpC3/	52	2733
Downstream LDR	AcDx-7054-UBTF-Dn	/5Phos/GTTTTCGTTTGGTTTGCGGCGTTTAGTTGATAGGTTGTATGGTCGGCATGCTA	53	2734

Real-Time Probe	AcDx-7055-UBTF-RT-Pb	/56-FAM/AAGAGGCTC/ZEN/GTTTTCGTTTGGTTTGCG/3IABkFQ/	27	2735
Tag Forward Primer	AcDx-7056-UBTF-RT-FP	TAGCAGCTGAACAACCCAAC	20	2736
Tag Reverse Primer	AcDx-7057-UBTF-RT-RP	TAGCATGCCGACCATACAAC	20	2737
Downstream PCR Primer	AcDx-7058-UBTF-PCR-V	TAGCATGCCGACCATACAACACGCCCAACAACCGAAAAATGrAAAAG/3SpC3/	46	2738
LOC284100				
Forward PCR Primer	AcDx-7061-LOC284100-FP	TTAATTTTCGTTTAGGTTTTCGTTTCrGGGTC/3SpC3/	31	2739
Reverse PCR Primer	AcDx-7062-LOC284100-RP	GGTGTCGTGGCTAAACAATCAAACGAACTAAAATACGArUTCCC/3SpC3/	43	2740
Upstream LDR	AcDx-7063-LOC284100-Up	TCTGCCCTTCGCTTCGAACGTTTTCGAGGGTTTATTTTTTAGTTTTGAGGCrGGTGG/3SpC3/	56	2741
Downstream LDR	AcDx-7064-LOC284100-Dn	/5Phos/GGTAAACGTTTTATTTTTAGTGGCGAGTGCGAGGTTGTATGGTCGGCATGCTA	53	2742
Real-Time Probe	AcDx-7065-LOC284100-RT-Pb	/56-FAM/AATTGAGGC/ZEN/GGTAAACGTTTTATTTTTAGTGGCG/3IABkFQ/	34	2743
Tag Forward Primer	AcDx-7066-LOC284100-RT-FP	TCTGCCCTTCGCTTCGAAC	19	2744
Tag Reverse Primer	AcDx-7067-LOC284100-RT-RP	TAGCATGCCGACCATACAAC	20	2745
Downstream PCR Primer	AcDx-7068-LOC284100-PCR-V	TAGCATGCCGACCATACAACCAATCAAACGAACTAAAATACGATTCCTTGrCACTT/3SpC3/	55	2746
WNT6-S1				
Forward PCR Primer	AcDx-7741-WNT6-S1-FP	TTTGTTGTTCGTCGTTCGTACrGTTTG/3SpC3/	26	2747
Reverse PCR Primer	AcDx-7742-WNT6-S1-RP	GGTGTCGTGGCCGAAAAAACCGATACGTCGArUTAAG/3SpC3/	36	2748
Upstream LDR	AcDx-7743-WNT6-S1-Up	TCCGGCCTTTGACGATACCCGTTTAAGTCGTCGGGTCGGGCrGGACC/3SpC3/	46	2749
Downstream LDR	AcDx-7744-WNT6-S1-Dn	/5Phos/GGATTTTTTTTACGTCGTCGATTCGTTCGATTTTTGCGGGTAATTCACTCGAACGGAGCA	60	2750
Real-Time Probe	AcDx-7745-WNT6-S1-RT-Pb	/56-FAM/TTGTCGGGC/ZEN/GGATTTTTTTTACGTCGTCGATTC/3IABkFQ/	33	2751
Tag Forward Primer	AcDx-7746-WNT6-S1-RT-FP	TCCGGCCTTTGACGATACC	19	2752

Tag Reverse Primer	AcDx-7747-WNT6-S1-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	2753
Downstream PCR Primer	AcDx-7748-WNT6-S1-PCR-V	TGCTCCGTTCGAGTGAATTACCCGAAAAAACCGATACGTCGATTAAAAATGrCAAAG/3SpC3/	56	2754
SGIP1				
Forward PCR Primer	AcDx-7751-SGIP1-FP	GTTTTTGATAATTAATTTCGGGTATTTAGTCrGTTTC/3SpC3/	36	2755
Reverse PCR Primer	AcDx-7752-SGIP1-RP	GGTGTCGTGGCATCGCCTCCCGCTTATCrACCAG/3SpC3/	33	2756
Upstream LDR	AcDx-7753-SGIP1-Up	TCCGGCCTTTGACGATACCGTATTTAGTCGTTTTTGTAAGTTTAAGGAGACAACrGAGAG/3SpC3/	59	2757
Downstream LDR	AcDx-7754-SGIP1-Dn	/5Phos/GAGGAGGAGCGGAGGAAGTGGTAATTCACTCGAACGGAGCA	41	2758
Real-Time Probe	AcDx-7755-SGIP1-RT-Pb	/56-FAM/TTAGACAAC/ZEN/GAGGAGGAGCGGG/3IABkFQ/	22	2759
Tag Forward Primer	AcDx-7756-SGIP1-RT-FP	TCCGGCCTTTGACGATACC	19	2760
Tag Reverse Primer	AcDx-7757-SGIP1-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	2761
Downstream PCR Primer	AcDx-7758-SGIP1-PCR-V	TGCTCCGTTCGAGTGAATTACCCAACCCCTTCCCCACCTAATAATGrCACTC/3SpC3/	51	2762
PON3-S1				
Forward PCR Primer	AcDx-7821-PON3-S1-FP	GTCGTAGTAGGGCGTTGACrGAGTC/3SpC3/	24	2763
Reverse PCR Primer	AcDx-7822-PON3-S1-RP	GGTGTCGTGGTCTCCGTTAAACCTTAACCTCTArCCCAG/3SpC3/	38	2764
Upstream LDR	AcDx-7823-PON3-S1-Up	TCGATGGTCAATGAGCTTCACAGCGTTGACGAGTTTCGTCGAGTCTCrGTCAC/3SpC3/	52	2765
Downstream LDR	AcDx-7824-PON3-S1-Dn	/5Phos/GTCGTTCGGGTTTAAGGTCGTTTTTACGTTTACGTGTTACGTGATCTCCCTCTCCA	56	2766
Real-Time Probe	AcDx-7825-PON3-S1-RT-Pb	/56-FAM/AAGAGTCTC/ZEN/GTCGTTCGGGTTTAAGGTCG/3IABkFQ/	29	2767
Tag Forward Primer	AcDx-7826-PON3-S1-RT-FP	TCGATGGTCAATGAGCTTCACA	22	2768
Tag Reverse Primer	AcDx-7827-PON3-S1-RT-RP	TGGAGAGGGAGATCACGTAACA	22	2769
Downstream PCR Primer	AcDx-7828-PON3-S1-PCR-V	TGGAGAGGGAGATCACGTAACACCTCTACCCAAAAAACAAAAAATCGTAAATGrUAAAG/3SpC3/	58	2770

ITPKA-S1				
Forward PCR Primer	AcDx-7851-ITPKA-S1-FP	AAGTTTTATAGAGTAGGAATATTTTTCGTCrGTTAA/3SpC3/	35	2771
Reverse PCR Primer	AcDx-7852-ITPKA-S1-RP	GGTGTCGTGGAAACGATCCTAAATCCGAAACTArCTCTG/3SpC3/	38	2772
Upstream LDR	AcDx-7853-ITPKA-S1-Up	TCTCGATTACGCTCCGCACTTCGTCGTTAGGTGTTGGGCGCrGTCAA/3SpC3/	46	2773
Downstream LDR	AcDx-7854-ITPKA-S1-Dn	/5Phos/GTCGGTTCGGTTATTAGTTTGTCGTTTTTTTTTTTTTTTCGGTGTGTAGCTTAG ACATGGCCA	63	2774
Real-Time Probe	AcDx-7855-ITPKA-S1-RT-Pb	/56-FAM/AATGGGCGC/ZEN/GTCGGTTCGGTTATTAG/3IABkFQ/	26	2775
Tag Forward Primer	AcDx-7856-ITPKA-S1-RT-FP	TCTCGATTACGCTCCGCAC	19	2776
Tag Reverse Primer	AcDx-7857-ITPKA-S1-RT-RP	TGGCCATGTCTAAGCTACACAC	22	2777
Downstream PCR Primer	AcDx-7858-ITPKA-S1-PCR-V	TGGCCATGTCTAAGCTACACACCCTAAATCCGAAACTACTCTAACCACTGrAAAAG/3SpC3/	55	2778
CaNCR41				
Forward PCR Primer	AcDx-7861-CaNCR41-FP	GGATATGGTGCGGTGGCrGGTAA/3SpC3/	22	2779
Reverse PCR Primer	AcDx-7862-CaNCR41-RP	GGTGTCGTGGCGCCTTAACCGCGAACTCrCCTCT/3SpC3/	33	2780
Upstream LDR	AcDx-7863-CaNCR41-Up	TCACAGAGACTTGCCGATCACGCGGGCGGTTGGATTTTAAATCrGGCAC/3SpC3/	48	2781
Downstream LDR	AcDx-7864-CaNCR41-Dn	/5Phos/GGCGTTGCGTTTTATATGACGGTTCGCGGTGTGTAGCTTAGACATGGCCA	50	2782
Real-Time Probe	AcDx-7865-CaNCR41-RT-Pb	/56-FAM/AATTAAATC/ZEN/GGCGTTGCGTTTTATATGACGGTTC/3IABkFQ/	34	2783
Tag Forward Primer	AcDx-7866-CaNCR41-RT-FP	TCACAGAGACTTGCCGATCAC	21	2784
Tag Reverse Primer	AcDx-7867-CaNCR41-RT-RP	TGGCCATGTCTAAGCTACACAC	22	2785
Downstream PCR Primer	AcDx-7868-CaNCR41-PCR-V	TGGCCATGTCTAAGCTACACACCTCCCTCCCCGAACCTGrCGAAT/3SpC3/	44	2786
ASCL2				
Forward PCR Primer	AcDx-7871-ASCL2-FP	AGGTTTAGGTTTTCGAGGCrGTTTC/3SpC3/	24	2787
Reverse PCR Primer	AcDx-7872-ASCL2-RP	GGTGTCGTGGCCCAAAACCCTCAAACCGArAAACA/3SpC3/	34	2788
Upstream LDR	AcDx-7873-ASCL2-Up	TCACAGAGACTTGCCGATCACGGGCGTTTTTTTTAATTCGTTTCGTTTTTCTCrGTTCC/3SpC3/	58	2789

Downstream LDR	AcDx-7874-ASCL2-Dn	/5Phos/GTTTTTTTACGCGTATTTTGTTTGTGGTTTTCGTGCGGTGTGTAGCTTAGACA TGGCCA	59	2790
Real-Time Probe	AcDx-7875-ASCL2-RT-Pb	/56-FAM/AATTTTCTC/ZEN/GTTTTTTTACGCGTATTTTGTTTGTGGTTTTC/3IABkFQ/	41	2791
Tag Forward Primer	AcDx-7876-ASCL2-RT-FP	TCACAGAGACTTGCCGATCAC	21	2792
Tag Reverse Primer	AcDx-7877-ASCL2-RT-RP	TGGCCATGTCTAAGCTACACAC	22	2793
Downstream PCR Primer	AcDx-7878-ASCL2-PCR-V	TGGCCATGTCTAAGCTACACACCCCAAAACCCTCAAACCGAAAATGrCACTG/3SpC3/	51	2794
CACNB2				
Forward PCR Primer	AcDx-8491-CACNB2-FP	GAATTGTTAGAGAACGTGGTTTTCrGCGGA/3SpC3/	29	2795
Reverse PCR Primer	AcDx-8492-CACNB2-RP	GGTGTCGTGGCCCTACCCGACGACTCArCAAAG/3SpC3/	32	2796
Upstream LDR	AcDx-8493-CACNB2-Up	TCCGACTTTAGTGCGTCACAAGTGGTTTTCGCGGGAGCrGTTTA/3SpC3/	43	2797
Downstream LDR	AcDx-8494-CACNB2-Dn	/5Phos/GTTCGGAGTCGTCGTATAGGTAGCGAGAGCTTGTGGGTCTCGCTCGTATA	50	2798
Real-Time Probe	AcDx-8495-CACNB2-RT-Pb	/56-FAM/TTCGGGAGC/ZEN/GTTCGGAGTCGTCGTATAG/3IABkFQ/	28	2799
Tag Forward Primer	AcDx-8496-CACNB2-RT-FP	TCCGACTTTAGTGCGTCACAA	21	2800
Tag Reverse Primer	AcDx-8497-CACNB2-RT-RP	TATACGAGCGAGACCCACAA	20	2801
Downstream PCR Primer	AcDx-8498-CACNB2-PCR-V	TATACGAGCGAGACCCACAAAAAAACGCCGCGCTCTTGrCTACT/3SpC3/	43	2802
CANCR68				
Forward PCR Primer	AcDx-9371-CANCR68-FP	GCGTTTTTGTTTTCGTTTTTTAGCrGTAGA/3SpC3/	29	2803
Reverse PCR Primer	AcDx-9372-CANCR68-RP	GGTGTCGTGGTTCTAACCTCGAAAACGAAACCrAAACA/3SpC3/	37	2804
Upstream LDR	AcDx-9373-CANCR68-Up	TTGCAGCGGGTCACAACAAAGCGTAGGTTGGGTGGGATCrGATAA/3SpC3/	44	2805
Downstream LDR	AcDx-9374-CANCR68-Dn	/5Phos/GATGGTTATTAAAGTTTCGTTTTTCGCGGTTTATCGTCGTTGGACAGAGGTAT ACGCCCA	60	2806
Real-Time Probe	AcDx-9375-CANCR68-RT-Pb	/56-FAM/TTTGGGATC/ZEN/GATGGTTATTAAAGTTTCGTTTTTCGCG/3IABkFQ/	37	2807
Tag Forward Primer	AcDx-9376-CANCR68-RT-FP	TTGCAGCGGGTCACAACAA	19	2808
Tag Reverse Primer	AcDx-9377-CANCR68-RT-RP	TGGGCGTATACCTCTGTCCAA	21	2809
Downstream PCR	AcDx-9378-CANCR68-PCR-V	TGGGCGTATACCTCTGTCCAACCTCGAAAACGAAACCAAACGATGrATAAG/3SpC3/	50	2810

Primer				
EFR3B				
Forward PCR Primer	AcDx-9381-EFR3B-FP	AAGATTATTAAGGAGTACGAGGAGCrGTATA/3SpC3/	30	2811
Reverse PCR Primer	AcDx-9382-EFR3B-RP	GGTGTCGTGGACTCCGCGCCTCGACrCCCGT/3SpC3/	30	2812
Upstream LDR	AcDx-9383-EFR3B-Up	TCGTCCCGGTCAGTAGTCAAGAGCGTATGTTTTAGGAGGTCGTTATTAAGGTCrGTGAA/3SpC3/	58	2813
Downstream LDR	AcDx-9384-EFR3B-Dn	/5Phos/GTGGGTGCGGCGCGGGTTGCCCATTTTCTGCACCCA	36	2814
Real-Time Probe	AcDx-9385-EFR3B-RT-Pb	/56-FAM/CCTAAGGTC/ZEN/GTGGGTGCGGCGC/3IABkFQ/	22	2815
Tag Forward Primer	AcDx-9386-EFR3B-RT-FP	TCGTCCCGGTCAGTAGTCAA	20	2816
Tag Reverse Primer	AcDx-9387-EFR3B-RT-RP	TGGGTGCAGAAAATGGGCAA	20	2817
Downstream PCR Primer	AcDx-9388-EFR3B-PCR-V	TGGGTGCAGAAAATGGGCAACGCGCCTCGACCCTGrCCCCA/3SpC3/	40	2818
CANCR105				
Forward PCR Primer	AcDx-9391-CANCR105-FP	CGCGGGAGGTTATGGCrGGGAA/3SpC3/	21	2819
Reverse PCR Primer	AcDx-9392-CANCR105-RP	GGTGTCGTGGACAAAAACTAATATAAAAAATTTTCAACGCrCGAAC/3SpC3/	45	2820
Upstream LDR	AcDx-9393-CANCR105-Up	TTCGTGCGTCGTGTAGCAAGGAATTTTTGTGAGTTTCGCAGCrGGAAA/3SpC3/	47	2821
Downstream LDR	AcDx-9394-CANCR105-Dn	/5Phos/GGAGGTTAGAATATCGTTGTGGTTGCGGGATTTTTATTCTTGCCCATTTTCTGCACCCA	59	2822
Real-Time Probe	AcDx-9395-CANCR105-RT-Pb	/56-FAM/AATCGCAGC/ZEN/GGAGGTTAGAATATCGTTGTGG/3IABkFQ/	31	2823
Tag Forward Primer	AcDx-9396-CANCR105-RT-FP	TTCGTGCGTCGTGTAGCAA	19	2824
Tag Reverse Primer	AcDx-9397-CANCR105-RT-RP	TGGGTGCAGAAAATGGGCAA	20	2825
Downstream PCR Primer	AcDx-9398-CANCR105-PCR-V	TGGGTGCAGAAAATGGGCAAATTTTCAACGCCGAATAAAAATCCTGrCAACT/3SpC3/	51	2826

C7orf51-S1				
Forward PCR Primer	AcDx-9401-C7orf51-S1-FP	TTGTTGGAGGTGATCGAGCrGTAAA/3SpC3/	24	2827
Reverse PCR Primer	AcDx-9402-C7orf51-S1-RP	GGTGTCGTGGCTCCTACTTACAAAAACCTCGATCrCGAAT/3SpC3/	39	2828
Upstream LDR	AcDx-9403-C7orf51-S1-Up	TTCGTGCGTCGTGTAGCAAGGTGATCGAGCGTAAGCGTCGCrGTGCA/3SpC3/	46	2829
Downstream LDR	AcDx-9404-C7orf51-S1-Dn	/5Phos/GTGTGTAAGGAGATTAAGGCGCGTTATCGTTCGTTGCCCATTTTCTGCACCCA	53	2830
Real-Time Probe	AcDx-9405-C7orf51-S1-RT-Pb	/56-FAM/TTGCGTCGC/ZEN/GTGTGTAAGGAGATTAAGGC/3IABkFQ/	29	2831
Tag Forward Primer	AcDx-9406-C7orf51-S1-RT-FP	TTCGTGCGTCGTGTAGCAA	19	2832
Tag Reverse Primer	AcDx-9407-C7orf51-S1-RT-RP	TGGGTGCAGAAAATGGGCAA	20	2833
Downstream PCR Primer	AcDx-9408-C7orf51-S1-PCR-V	TGGGTGCAGAAAATGGGCAATTACAAAAACCTCGATCCGAACGATAATGrCGCCC/3SpC3/	54	2834
OXT				
Forward PCR Primer	AcDx-9421-OXT-FP	CGAGGAGGGAGGGATTCrGTAGC/3SpC3/	22	2835
Reverse PCR Primer	AcDx-9422-OXT-RP	GGTGTCGTGGGCGATCAAAAACGAAACGTCArAAATT/3SpC3/	36	2836
Upstream LDR	AcDx-9423-OXT-Up	TCACGCACGTAGGGTCTAAACGGAGGGATTCGTAGTTATAGGAGCrGCGCC/3SpC3/	50	2837
Downstream LDR	AcDx-9424-OXT-Dn	/5Phos/GCGTTTCGTTTCGGTTTCGTTTGAGAATTTTAGGGTTGTCCGGCTGTGGTTACA	54	2838
Real-Time Probe	AcDx-9425-OXT-RT-Pb	/56-FAM/CCTAGGAGC/ZEN/GCGTTTCGTTTCGGTTTCG/3IABkFQ/	28	2839
Tag Forward Primer	AcDx-9426-OXT-RT-FP	TCACGCACGTAGGGTCTAAAC	21	2840
Tag Reverse Primer	AcDx-9427-OXT-RT-RP	TGTAACCACAGCCGGACAAC	20	2841
Downstream PCR Primer	AcDx-9428-OXT-PCR-V	TGTAACCACAGCCGGACAACCGATCAAAAACGAAACGTCAAAATCTGrCTCAG/3SpC3/	52	2842
TMEM101-S1				
Forward PCR Primer	AcDx-9431-TMEM101-S1-FP	GAGGATGGGATACGTAGTTTTCrGGGTC/3SpC3/	27	2843

Reverse PCR Primer	AcDx-9432-TMEM101-S1-RP	GGTGTCGTGGAACTTCGCAACTAAATCACCTTAAArUCCGG/3SpC3/	40	2844
Upstream LDR	AcDx-9433-TMEM101-S1-Up	TCTTACGCCCAGGGAATGTAACATGGGATACGTAGTTTTCGGGTTAGGCrGTTCC/3SpC3/	54	2845
Downstream LDR	AcDx-9434-TMEM101-S1-Dn	/5Phos/GTTTTTTTTTTTTTTTTTCGGAAATAACGGTGTTATTTTTTAGTTTCGTTGTCCGGCTGTGGTTACA	67	2846
Real-Time Probe	AcDx-9435-TMEM101-S1-RT-Pb	/56-FAM/TTGTTAGGC/ZEN/GTTTTTTTTTTTTTTTTTCGGAAATAACGGTGTTAT/3IABkFQ/	45	2847
Tag Forward Primer	AcDx-9436-TMEM101-S1-RT-FP	TCTTACGCCCAGGGAATGTAAC	22	2848
Tag Reverse Primer	AcDx-9437-TMEM101-S1-RT-RP	TGTAACCACAGCCGGACAAC	20	2849
Downstream PCR Primer	AcDx-9438-TMEM101-S1-PCR-V	TGTAACCACAGCCGGACAACAACTTCGCAACTAAATCACCTTAAATCTGrAAACC/3SpC3/	54	2850
CANCR69				
Forward PCR Primer	AcDx-9441-CANCR69-FP	TTTCGTTTCGGAGCGAGTCrGCGAC/3SpC3/	24	2851
Reverse PCR Primer	AcDx-9442-CANCR69-RP	GGTGTCGTGGAAAACTATAAAACGAAATTTACTCAATAAArCAATC/3SpC3/	45	2852
Upstream LDR	AcDx-9443-CANCR69-Up	TGAGCAAAATCTTCGTCGACCGAGTCGCGGTTCGGAGTGGCrGTTGC/3SpC3/	46	2853
Downstream LDR	AcDx-9444-CANCR69-Dn	/5Phos/GTTATTCGAAAGTTATCGGAGATATCGAGAGAGTTTTTTAATTGGGTTCGATGCCTTCCGTACA	64	2854
Real-Time Probe	AcDx-9445-CANCR69-RT-Pb	/56-FAM/TTGAGTGGC/ZEN/GTTATTCGAAAGTTATCGGAGATATCGAG/3IABkFQ/	38	2855
Tag Forward Primer	AcDx-9446-CANCR69-RT-FP	TGAGCAAAATCTTCGTCGACC	21	2856
Tag Reverse Primer	AcDx-9447-CANCR69-RT-RP	TGTACGGAAGGCATCGAACC	20	2857
PYY				
Forward PCR Primer	AcDx-9451-PYY-FP	CGGCGGGTTTGTGTTACrGCGCC/3SpC3/	22	2858
Reverse PCR Primer	AcDx-9452-PYY-RP	GGTGTCGTGGCAAAACGAACAAAATAACACGCTCrUAAAG/3SpC3/	39	2859

Upstream LDR	AcDx-9453-PYY-Up	TGGCACATGAGAGTAGTTGACCGTTTGTGTTACGCGTTGTCGAATCrGGTCC/3SpC3/	51	2860
Downstream LDR	AcDx-9454-PYY-Dn	/5Phos/GGTTTTTTATGTTAGGTGAGTGTTCGTTTTGTTCGTTTAGGGGTTCGATGCCTTCCGTACA	61	2861
Real-Time Probe	AcDx-9455-PYY-RT-Pb	/56-FAM/AATCGAATC/ZEN/GGTTTTTTATGTTAGGTGAGTGTTCGTTTTG/3IABkFQ/	40	2862
Tag Forward Primer	AcDx-9456-PYY-RT-FP	TGGCACATGAGAGTAGTTGACC	22	2863
Tag Reverse Primer	AcDx-9457-PYY-RT-RP	TGTACGGAAGGCATCGAACC	20	2864
Downstream PCR Primer	AcDx-9458-PYY-PCR-V	TGTACGGAAGGCATCGAACCATAACACGCTCTAAAAACGCCTAAATGrAACAG/3SpC3/	52	2865
LOC389333-S1				
Forward PCR Primer	AcDx-9461-LOC389333-S1-FP	TTTCGGGCGATCGTGTCrGAAGA/3SpC3/	22	2866
Reverse PCR Primer	AcDx-9462-LOC389333-S1-RP	GGTGTCGTGGAAAACCCCCCGCCCCrGACTT/3SpC3/	30	2867
Upstream LDR	AcDx-9463-LOC389333-S1-Up	TATAGTCACGCAGGACCACAGTCGAAGGACGGTTTAGGAGGCTCrGTAAA/3SpC3/	49	2868
Downstream LDR	AcDx-9464-LOC389333-S1-Dn	/5Phos/GTAGGAGTGCGGTATTATCGGTAGAAAATTTTTGATGTGTTTGCGGCTGTCTATGACA	58	2869
Real-Time Probe	AcDx-9465-LOC389333-S1-RT-Pb	/56-FAM/TTGAGGCTC/ZEN/GTAGGAGTGCGGTATTATCG/3IABkFQ/	29	2870
Tag Forward Primer	AcDx-9466-LOC389333-S1-RT-FP	TATAGTCACGCAGGACCACA	20	2871
Tag Reverse Primer	AcDx-9467-LOC389333-S1-RT-RP	TGTCATAGACAGCCGCAAACA	21	2872
Downstream PCR Primer	AcDx-9468-LOC389333-S1-PCR-V	TGTCATAGACAGCCGCAAACACCCCGACTCACTACACTTCTGrUTTTT/3SpC3/	47	2873
TXNRD1-S1				
Forward PCR Primer	AcDx-9471-TXNRD1-S1-FP	TGCGATATGGGTTGCGTCrGAGGA/3SpC3/	23	2874
Reverse PCR Primer	AcDx-9472-TXNRD1-S1-RP	GGTGTCGTGGCGTACCTAACCTACGACGACrCATCA/3SpC3/	35	2875

Upstream LDR	AcDx-9473-TXNRD1-S1-Up	TGCCCTATCGAAAAGGACAACACGTCGAGGGTAAGGTAGTAGCrGGCAA/3SpC3/	48	2876
Downstream LDR	AcDx-9474-TXNRD1-S1-Dn	/5Phos/GGCGGTCGTTTTAACGGAGTTGTAGACGAAAGTGTTTGCGGCTGTCTATGACA	53	2877
Real-Time Probe	AcDx-9475-TXNRD1-S1-RT-Pb	/56-FAM/TTTAGTAGC/ZEN/GGCGGTCGTTTTAACGGAGTTG/3IABkFQ/	31	2878
Tag Forward Primer	AcDx-9476-TXNRD1-S1-RT-FP	TGCCCTATCGAAAAGGACAACA	22	2879
Tag Reverse Primer	AcDx-9477-TXNRD1-S1-RT-RP	TGTCATAGACAGCCGCAAACA	21	2880
Downstream PCR Primer	AcDx-9478-TXNRD1-S1-PCR-V	TGTCATAGACAGCCGCAAACACCTACGACGACCATCGCTGrUTCTC/3SpC3/	45	2881
CARD11				
Forward PCR Primer	AcDx-9481-CARD11-FP	TTATTTGAGGCGTTTATGTTTCrGTTAA/3SpC3/	27	2882
Reverse PCR Primer	AcDx-9482-CARD11-RP	GGTGTCGTGGCCGAACCCGAAAAAACGAArAAACT/3SpC3/	34	2883
Upstream LDR	AcDx-9483-CARD11-Up	TAATCTCCAGACCTCCGAACCGTTAGTTAGTTGCGCGGAGCrGTTCC/3SpC3/	46	2884
Downstream LDR	AcDx-9484-CARD11-Dn	/5Phos/GTTTTTTTTACGTAGGTTTTCGGTTTTTTTTAGGCGATTAGGGTGTAAGGATTGAACGGGACA	63	2885
Real-Time Probe	AcDx-9485-CARD11-RT-Pb	/56-FAM/TTGCGGAGC/ZEN/GTTTTTTTTACGTAGGTTTTCGG/3IABkFQ/	32	2886
Tag Forward Primer	AcDx-9486-CARD11-RT-FP	TAATCTCCAGACCTCCGAACC	21	2887
Tag Reverse Primer	AcDx-9487-CARD11-RT-RP	TGTCCCGTTCAATCCTTACACC	22	2888
Downstream PCR Primer	AcDx-9488-CARD11-PCR-V	TGTCCCGTTCAATCCTTACACCGAACCCGAAAAAACGAAAAACCTAATTGrCCTAG/3SpC3/	55	2889
LYPLAL1				
Forward PCR Primer	AcDx-9491-LYPLAL1-FP	TTTAGTGGGTGGTTTTTTCGTCrGTTAA/3SpC3/	27	2890
Reverse PCR Primer	AcDx-9492-LYPLAL1-RP	GGTGTCGTGGTAACCGCCCGATAAAAAAAAAAAAACrUAACA/3SpC3/	41	2891
Upstream LDR	AcDx-9493-LYPLAL1-Up	TTTTCCGCGTCAGAGCACAGTGGGTGGTTTTTTCGTCGTTAGTCTCrGGTTA/3SpC3/	51	2892
Downstream LDR	AcDx-9494-LYPLAL1-Dn	/5Phos/GGTTGTAGATGTACGGGTAGCGTTATTTGTTTTTAGTTTTTGTTATCGGACCTAGCTCGACA	62	2893

Real-Time Probe	AcDx-9495-LYPLAL1-RT-Pb	/56-FAM/CCTAGTCTC/ZEN/GGTTGTAGATGTACGGGTAGCG/3IABkFQ/	31	2894
Tag Forward Primer	AcDx-9496-LYPLAL1-RT-FP	TTTTCCGCGTCAGAGCACA	19	2895
Tag Reverse Primer	AcDx-9497-LYPLAL1-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	2896
Downstream PCR Primer	AcDx-9498-LYPLAL1-PCR-V	TGTCGAGCTAGGTCCGATAACATAACCGCCCGATAAAAAAAAAAAAACTAATGrAAAAG/3SpC3/	58	2897
RREB1				
Forward PCR Primer	AcDx-9501-RREB1-FP	TTTGAAGTTTTGGAGGGCGACrGTGGG/3SpC3/	26	2898
Reverse PCR Primer	AcDx-9502-RREB1-RP	GGTGTCGTGGCGAAAAACTACCGTAACTGCrCAAAG/3SpC3/	35	2899
Upstream LDR	AcDx-9503-RREB1-Up	TTCGCTGCCCGGTTAAACACGACGTGGATGATATAGTATTAGTTATTTTATCGCrGGGAC/3SpC3/	59	2900
Downstream LDR	AcDx-9504-RREB1-Dn	/5Phos/GGAGTGGAGATATAGTGGCGGTAGGTGTTATCGGACCTAGCTCGACA	47	2901
Real-Time Probe	AcDx-9505-RREB1-RT-Pb	/56-FAM/AATTATTGC/ZEN/GGAGTGGAGATATAGTGGCG/3IABkFQ/	29	2902
Tag Forward Primer	AcDx-9506-RREB1-RT-FP	TTCGCTGCCCGGTTAAACA	19	2903
Tag Reverse Primer	AcDx-9507-RREB1-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	2904
Downstream PCR Primer	AcDx-9508-RREB1-PCR-V	TGTCGAGCTAGGTCCGATAACACTACCGTAACCGCCAAAAACTAAAAATGrATTCT/3SpC3/	55	2905
LRRC32				
Forward PCR Primer	AcDx-9511-LRRC32-FP	CGCGGTGGTTTTAGGAGTCrGTAGA/3SpC3/	24	2906
Reverse PCR Primer	AcDx-9512-LRRC32-RP	GGTGTCGTGGCCTATTTACCCCCGCGTTAArATACA/3SpC3/	35	2907
Upstream LDR	AcDx-9513-LRRC32-Up	TTCGCTGCCCGGTTAAACAGTTTTAGGAGTCGTAGGTTTATAGAGGAAACTCrGGTCC/3SpC3/	57	2908
Downstream LDR	AcDx-9514-LRRC32-Dn	/5Phos/GGTTTGGTTTCGATTTTTTAGTTGTTGGCGTCGTGTTATCGGACCTAGCTCGACA	55	2909
Real-Time Probe	AcDx-9515-LRRC32-RT-Pb	/56-FAM/TTGAAACTC/ZEN/GGTTTGGTTTCGATTTTTTAGTTGTTGGCG/3IABkFQ/	39	2910
Tag Forward Primer	AcDx-9516-LRRC32-RT-FP	TTCGCTGCCCGGTTAAACA	19	2911
Tag Reverse Primer	AcDx-9517-LRRC32-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	2912
Downstream PCR	AcDx-9518-LRRC32-PCR-V	TGTCGAGCTAGGTCCGATAACATTACCCCCGCGTTAAATACGATGrACGCT/3SpC3/	50	2913

Primer				
EMX2OS				
Forward PCR Primer	AcDx-9521-EMX2OS-FP	GAAAAGTTAATATTGAATTAATGTTGTTTTTGTTCrGCGTT/3SpC3/	40	2914
Reverse PCR Primer	AcDx-9522-EMX2OS-RP	GGTGTCGTGGTAACCCCGACCGCCAACrAACTC/3SpC3/	32	2915
Upstream LDR	AcDx-9523-EMX2OS-Up	TGCTATGCCGCATTCAACCAGTTGTTTTTGTTCGCGTCGTCGCrGATAC/3SpC3/	48	2916
Downstream LDR	AcDx-9524-EMX2OS-Dn	/5Phos/GATGTTATTTTTTGTTGCGTACGTCGTTTTCGGTAAGTGGAGCTAGTTCGGCGACA	56	2917
Real-Time Probe	AcDx-9525-EMX2OS-RT-Pb	/56-FAM/TTTCGTCGC/ZEN/GATGTTATTTTTTGTTGCGTACGTC/3IABkFQ/	34	2918
Tag Forward Primer	AcDx-9526-EMX2OS-RT-FP	TGCTATGCCGCATTCAACCA	20	2919
Tag Reverse Primer	AcDx-9527-EMX2OS-RT-RP	TGTCGCCGAACTAGCTCCA	19	2920
Downstream PCR Primer	AcDx-9528-EMX2OS-PCR-V	TGTCGCCGAACTAGCTCCACCGACCGCCAACAACTTACTGrAAAAT/3SpC3/	45	2921
PTGDR-S2				
Forward PCR Primer	AcDx-9531-PTGDR-S2-FP	CGTTTTTTAATGTTAGCGTTAGGCrGTTTG/3SpC3/	29	2922
Reverse PCR Primer	AcDx-9532-PTGDR-S2-RP	GGTGTCGTGGCGAAATATTCCCCACGACAAAAACrCTCCC/3SpC3/	39	2923
Upstream LDR	AcDx-9533-PTGDR-S2-Up	TGTGCCTTACGGAAAACCCAGTTAGCGTTAGGCGTTTATTTTGTAGGGCrGTTCT/3SpC3/	54	2924
Downstream LDR	AcDx-9534-PTGDR-S2-Dn	/5Phos/GTTTCGTTTTTTAAAGAGGAGTGTGATTCGCGAGTGGAGCTAGTTCGGCGACA	53	2925
Real-Time Probe	AcDx-9535-PTGDR-S2-RT-Pb	/56-FAM/AAGTAGGGC/ZEN/GTTTCGTTTTTTAAAGAGGAGTGT/3IABkFQ/	33	2926
Tag Forward Primer	AcDx-9536-PTGDR-S2-RT-FP	TGTGCCTTACGGAAAACCCA	20	2927
Tag Reverse Primer	AcDx-9537-PTGDR-S2-RT-RP	TGTCGCCGAACTAGCTCCA	19	2928
Downstream PCR Primer	AcDx-9538-PTGDR-S2-PCR-V	TGTCGCCGAACTAGCTCCACGACAAAAACCTCCTATCTAAACTCGTGrAAGCG/3SpC3/	52	2929

CANCR70				
Forward PCR Primer	AcDx-9541-CANCR70-FP	TTGGAGGCGTTCGGCrGTGGA/3SpC3/	20	2930
Reverse PCR Primer	AcDx-9542-CANCR70-RP	GGTGTCGTGGCTCGCTACGCCGCGAATAAArUTCGG/3SpC3/	35	2931
Upstream LDR	AcDx-9543-CANCR70-Up	TCCTGCTCTGAAAACCTACACCCGTTCGGCGTGAGGTTTTCACrGGCAG/3SpC3/	48	2932
Downstream LDR	AcDx-9544-CANCR70-Dn	/5Phos/GGCGATTTTTTTTCGGTCGTTTTTTTATCGTAGTTCGAGGTTACATAGGCGGC TTAGACA	60	2933
Real-Time Probe	AcDx-9545-CANCR70-RT-Pb	/56-FAM/TTTTTTCAC/ZEN/GGCGATTTTTTTTCGGTCGTTTTTTTATCG/3IABkFQ/	39	2934
Tag Forward Primer	AcDx-9546-CANCR70-RT-FP	TCCTGCTCTGAAAACCTACACC	22	2935
Tag Reverse Primer	AcDx-9547-CANCR70-RT-RP	TGTCTAAGCCGCCTATGTAACC	22	2936
Downstream PCR Primer	AcDx-9548-CANCR70-PCR-V	TGTCTAAGCCGCCTATGTAACCCTCGCTACGCCGCGAATAAATTTGrAACTG/3SpC3/	51	2937
NPHS2-S1				
Forward PCR Primer	AcDx-9551-NPHS2-S1-FP	GTAGTTTTTGTGGAGTCGTTGCrGGGTC/3SpC3/	27	2938
Reverse PCR Primer	AcDx-9552-NPHS2-S1-RP	GGTGTCGTGGCCGATATCCCGCCCCTCrGCCCC/3SpC3/	32	2939
Upstream LDR	AcDx-9553-NPHS2-S1-Up	TTTTTACGCACAGCACCACCGCGGGTTCGCGTAGCrGTGCC/3SpC3/	40	2940
Downstream LDR	AcDx-9554-NPHS2-S1-Dn	/5Phos/GTGTTCGGGAGATTTTAAAGATCGTCGGGTGGGTTACATAGGCGGCTTAGA CA	53	2941
Real-Time Probe	AcDx-9555-NPHS2-S1-RT-Pb	/56-FAM/AAGCGTAGC/ZEN/GTGTTCGGGAGATTTTAAAGATC/3IABkFQ/	32	2942
Tag Forward Primer	AcDx-9556-NPHS2-S1-RT-FP	TTTTTACGCACAGCACCACC	20	2943
Tag Reverse Primer	AcDx-9557-NPHS2-S1-RT-RP	TGTCTAAGCCGCCTATGTAACC	22	2944
Downstream PCR Primer	AcDx-9558-NPHS2-S1-PCR-V	TGTCTAAGCCGCCTATGTAACCCCCTCAACCCCCACCTGrACGAC/3SpC3/	44	2945
LOC645323				
Forward PCR Primer	AcDx-9561-LOC645323-FP	TAGGTAGCGGTTGGCGTCrGGGTG/3SpC3/	23	2946
Reverse PCR Primer	AcDx-9562-LOC645323-RP	GGTGTCGTGGTTTCACTCCCTAACGACAATCTCrAAAAT/3SpC3/	38	2947

Upstream LDR	AcDx-9563-LOC645323-Up	TGGACACTTCGCCCTTCTTAACGGTTGGCGTCGGGTATACGAGTCrGGTTA/3SpC3/	50	2948
Downstream LDR	AcDx-9564-LOC645323-Dn	/5Phos/GGTCGAGTTTTTTTGTTAGTTAGTTTCGGGTATTTTTAGTGGTTTGGGATCTGGGCATCACA	62	2949
Real-Time Probe	AcDx-9565-LOC645323-RT-Pb	/56-FAM/CCACGAGTC/ZEN/GGTCGAGTTTTTTTGTTAGTTAGTTTCG/3IABkFQ/	37	2950
Tag Forward Primer	AcDx-9566-LOC645323-RT-FP	TGGACACTTCGCCCTTCTTAAC	22	2951
Tag Reverse Primer	AcDx-9567-LOC645323-RT-RP	TGTGATGCCCAGATCCCAAAC	21	2952
Downstream PCR Primer	AcDx-9568-LOC645323-PCR-V	TGTGATGCCCAGATCCCAAACTTTCACTCCCTAACGACAATCTCAAAATGrCACTG/3SpC3/	55	2953
CANCR71				
Forward PCR Primer	AcDx-9571-CANCR71-FP	GTGTTTATAAAGTAGGAAGCGAAGAACrGTTTC/3SpC3/	32	2954
Reverse PCR Primer	AcDx-9572-CANCR71-RP	GGTGTCGTGGGCAACGAAACCCGAATCCrCTACC/3SpC3/	33	2955
Upstream LDR	AcDx-9573-CANCR71-Up	TGGAGGCCGGAGAAATTAAACGGAAGCGAAGAACGTTTTTTTTATTTAGGGCrGTAAC/3SpC3/	57	2956
Downstream LDR	AcDx-9574-CANCR71-Dn	/5Phos/GTAGTCGTTTTGCGGAAGGTATAGTTCGGGTTTCGTTTGGGATCTGGGCATCACA	55	2957
Real-Time Probe	AcDx-9575-CANCR71-RT-Pb	/56-FAM/CCTTAGGGC/ZEN/GTAGTCGTTTTGCGGAAGGTATAG/3IABkFQ/	33	2958
Tag Forward Primer	AcDx-9576-CANCR71-RT-FP	TGGAGGCCGGAGAAATTAAAC	21	2959
Tag Reverse Primer	AcDx-9577-CANCR71-RT-RP	TGTGATGCCCAGATCCCAAAC	21	2960
Downstream PCR Primer	AcDx-9578-CANCR71-PCR-V	TGTGATGCCCAGATCCCAAACCCCTACTTAACTCCCGAAACCTGrAACTG/3SpC3/	49	2961
ZNF506				
Forward PCR Primer	AcDx-9581-ZNF506-FP	GTTGACGGTATAGAGTAGTGAAGACrGATAA/3SpC3/	30	2962
Reverse PCR Primer	AcDx-9582-ZNF506-RP	GGTGTCGTGGCGCAACTATAACGAACAAAACGArUTTCT/3SpC3/	38	2963
Upstream LDR	AcDx-9583-ZNF506-Up	TTGTCTCTGCGACCCATCAAGTAGTGAAGACGATAGTTGGATTTTTAGCrGTTGA/3SpC3/	54	2964

Downstream LDR	AcDx-9584-ZNF506-Dn	/5Phos/GTTAGCGAGAGATAATGGGTTCGTTAAAGTCGGAAGTCTTGGTACACGTTCGGCACA	57	2965
Real-Time Probe	AcDx-9585-ZNF506-RT-Pb	/56-FAM/CCTTTTAGC/ZEN/GTTAGCGAGAGATAATGGGTTCGTT/3IABkFQ/	34	2966
Tag Forward Primer	AcDx-9586-ZNF506-RT-FP	TTGTCTCTGCGACCCATCAA	20	2967
Tag Reverse Primer	AcDx-9587-ZNF506-RT-RP	TGTGCCGAACGTGTACCAA	19	2968
Downstream PCR Primer	AcDx-9588-ZNF506-PCR-V	TGTGCCGAACGTGTACCAACGAACAAAACGATTTCCGACTTCTGrACTTC/3SpC3/	49	2969
SOX14				
Forward PCR Primer	AcDx-9591-SOX14-FP	ATAGATTTTTATTCGTTAGTTTACGTTACrGGTTG/3SpC3/	34	2970
Reverse PCR Primer	AcDx-9592-SOX14-RP	GGTGTCGTGGAACGTCTACAAACCGAAACTAAArCTCAG/3SpC3/	38	2971
Upstream LDR	AcDx-9593-SOX14-Up	TGGATAAACTAAGTCCGCCCACCGTTACGGTTATGTAATTTTTAGTTCGGCTCrGGACC/3SpC3/	58	2972
Downstream LDR	AcDx-9594-SOX14-Dn	/5Phos/GGATTTGAGGCGTGGTTTGAAAGTCGGGGTGACTGAGCGACGTCTAACA	49	2973
Real-Time Probe	AcDx-9595-SOX14-RT-Pb	/56-FAM/AATCGGCTC/ZEN/GGATTTGAGGCGTGG/3IABkFQ/	24	2974
Tag Forward Primer	AcDx-9596-SOX14-RT-FP	TGGATAAACTAAGTCCGCCCAC	22	2975
Tag Reverse Primer	AcDx-9597-SOX14-RT-RP	TGTTAGACGTCGCTCAGTCAC	21	2976
Downstream PCR Primer	AcDx-9598-SOX14-PCR-V	TGTTAGACGTCGCTCAGTCACGAAACTAAACTCAAAAAACAAAATACAAACCTGrACTTC/3SpC3/	59	2977
PRKCG				
Forward PCR Primer	AcDx-9601-PRKCG-FP	TGATAAGTTTTTTACGCGGGCrGGCGC/3SpC3/	26	2978
Reverse PCR Primer	AcDx-9602-PRKCG-RP	GGTGTCGTGGTCCGAATACACGAAATCGAAArUTCAT/3SpC3/	36	2979
Upstream LDR	AcDx-9603-PRKCG-Up	TCCTGAATTGGCCACACCACGCGGCGTTAGCGTTGATTTTTTTAGGTCrGTTAG/3SpC3/	53	2980
Downstream LDR	AcDx-9604-PRKCG-Dn	/5Phos/GTTTAGTTTTGGTTAGTATCGATTAGGTCGATTTTTAGGGTTTTGTGACTGAGCGACGTCTAACA	65	2981
Real-Time Probe	AcDx-9605-PRKCG-RT-Pb	/56-FAM/AATTAGGTC/ZEN/GTTTAGTTTTGGTTAGTATCGATTAGGTCGA/3IABkFQ/	40	2982
Tag Forward Primer	AcDx-9606-PRKCG-RT-FP	TCCTGAATTGGCCACACCAC	20	2983

Tag Reverse Primer	AcDx-9607-PRKCG-RT-RP	TGTTAGACGTCGCTCAGTCAC	21	2984
Downstream PCR Primer	AcDx-9608-PRKCG-PCR-V	TGTTAGACGTCGCTCAGTCACTCCGAATACACGAAATCGAAATTCATGrUAAAC/3SpC3/	53	2985
CANCR72				
Forward PCR Primer	AcDx-9611-CANCR72-FP	GGGAGGGTTTGTTTCGTTTCrGGTTC/3SpC3/	25	2986
Reverse PCR Primer	AcDx-9612-CANCR72-RP	GGTGTCGTGGAACGCCCTCACTTCGAAArAAACG/3SpC3/	33	2987
Upstream LDR	AcDx-9613-CANCR72-Up	TTCTTGCGGTTCTGGAACACCGGTTTAGGATTTTTGTTTTCGGTTGAGGCrGTAAT/3SpC3/	55	2988
Downstream LDR	AcDx-9614-CANCR72-Dn	/5Phos/GTAGCGTTTTTAGTCGTTCGTAGGAGGGATCGACGTGATGCTCCGTTGTTGCTAA	55	2989
Real-Time Probe	AcDx-9615-CANCR72-RT-Pb	/56-FAM/TTTTGAGGC/ZEN/GTAGCGTTTTTAGTCGTTCGTAGG/3IABkFQ/	33	2990
Tag Forward Primer	AcDx-9616-CANCR72-RT-FP	TTCTTGCGGTTCTGGAACAC	20	2991
Tag Reverse Primer	AcDx-9617-CANCR72-RT-RP	TTAGCAACAACGGAGCATCAC	21	2992
Downstream PCR Primer	AcDx-9618-CANCR72-PCR-V	TTAGCAACAACGGAGCATCACCCCTCACTTCGAAAAAACAAAAACGTTGrATCCT/3SpC3/	54	2993
PCOLCE				
Forward PCR Primer	AcDx-9621-PCOLCE-FP	GTTAGGTTATCGCGTTGATTTTCrGAGAG/3SpC3/	28	2994
Reverse PCR Primer	AcDx-9622-PCOLCE-RP	GGTGTCGTGGAACCTCCGAAAATCGTCGCrUCACA/3SpC3/	34	2995
Upstream LDR	AcDx-9623-PCOLCE-Up	TATGGTAAAATGTCAGCGGCACGAAGTTTGATTTGGAGTCGGATATTTATTATCrGTTGC/3SpC3/	59	2996
Downstream LDR	AcDx-9624-PCOLCE-Dn	/5Phos/GTTATGATTCGGTTAGCGTGTTTAACGGAGTCGTGGTGATGCTCCGTTGTTGCTAA	56	2997
Real-Time Probe	AcDx-9625-PCOLCE-RT-Pb	/56-FAM/AATATTATC/ZEN/GTTATGATTCGGTTAGCGTGTTTAACGGAG/3IABkFQ/	39	2998
Tag Forward Primer	AcDx-9626-PCOLCE-RT-FP	TATGGTAAAATGTCAGCGGCAC	22	2999
Tag Reverse Primer	AcDx-9627-PCOLCE-RT-RP	TTAGCAACAACGGAGCATCAC	21	3000
Downstream PCR Primer	AcDx-9628-PCOLCE-PCR-V	TTAGCAACAACGGAGCATCACCTCCGAAAATCGTCGCTCATGrACTCT/3SpC3/	47	3001

C3orf55				
Forward PCR Primer	AcDx-9631-C3orf55-FP	GTTTTTTGGGAGGTGGTGTTCrGTAAG/3SpC3/	26	3002
Reverse PCR Primer	AcDx-9632-C3orf55-RP	GGTGTCGTGGCCTAAACACGCGAAACGAAArACCGG/3SpC3/	35	3003
Upstream LDR	AcDx-9633-C3orf55-Up	TCCTCGAGCCGATGACACATTGGTAGTTGATTGGGTGGAGCrGTGCC/3SpC3/	46	3004
Downstream LDR	AcDx-9634-C3orf55-Dn	/5Phos/GTGTTTTTTTTGGTAGTCGCGGAGGAGAAGTGTAACGTCCGTGGGCTAA	49	3005
Real-Time Probe	AcDx-9635-C3orf55-RT-Pb	/56-FAM/TTGTGGAGC/ZEN/GTGTTTTTTTTGGTAGTCGCGG/3IABkFQ/	31	3006
Tag Forward Primer	AcDx-9636-C3orf55-RT-FP	TCCTCGAGCCGATGACACA	19	3007
Tag Reverse Primer	AcDx-9637-C3orf55-RT-RP	TTAGCCCACGGACGTTACA	19	3008
Downstream PCR Primer	AcDx-9638-C3orf55-PCR-V	TTAGCCCACGGACGTTACACCTAAACACGCGAAACGAAAACTGrAAGCT/3SpC3/	48	3009
SLC25A39				
Forward PCR Primer	AcDx-10171-SLC25A39-FP	GCGGTAGTGAGAATGAGTTTTTCrGAGAC/3SpC3/	28	3010
Reverse PCR Primer	AcDx-10172-SLC25A39-RP	GGTGTCGTGGAAAATCGCCGCGAAATTTTCrAAATT/3SpC3/	35	3011
Upstream LDR	AcDx-10173-SLC25A39-Up	TCTGCCCAAAATACTGCACAAGGTAGTGAGAATGAGTTTTTCGAGATCATCrGGGCA/3SpC3/	56	3012
Downstream LDR	AcDx-10174-SLC25A39-Dn	/5Phos/GGGAGATTATATAATAACGACGAAGTTATTTGTCGGGTTATTCGTTGAAACTGAGGCGGTGTTCA	65	3013
Real-Time Probe	AcDx-10175-SLC25A39-RT-Pb	/56-FAM/TTGATCATC/ZEN/GGGAGATTATATAATAACGACGAAGTTATTTGTCG/3IABkFQ/	44	3014
Tag Forward Primer	AcDx-10176-SLC25A39-RT-FP	TCTGCCCAAAATACTGCACAA	21	3015
Tag Reverse Primer	AcDx-10177-SLC25A39-RT-RP	TGAACACCGCCTCAGTTTCAA	21	3016
Downstream PCR Primer	AcDx-10178-SLC25A39-PCR-V	TGAACACCGCCTCAGTTTCAAGCCGCGAAATTTTCAAATCTCGATGrAATAG/3SpC3/	51	3017
Alternate Group 2				

Markers				
FOXI3				
Forward PCR Primer	AcDx-9031-FOXI3-FP	GTGTCGGCGGTAGGCrGGTGA/3SpC3/	20	3018
Reverse PCR Primer	AcDx-9032-FOXI3-RP	GGTGTCGTGGCCGCCGCGTACCTAArCGCTT/3SpC3/	30	3019
Upstream LDR	AcDx-9033-FOXI3-Up	TCTCATGGGCGCTAGTATCAACGGCGGTGGTTGTAGAAAGGGCTCrGGTTA/3SpC3/	50	3020
Downstream LDR	AcDx-9034-FOXI3-Dn	/5Phos/GGTCGCGGTTTCGGGAGGCGTTTCCCTGATTGATACCCGCA	41	3021
Real-Time Probe	AcDx-9035-FOXI3-RT-Pb	/56-FAM/TTAGGGCTC/ZEN/GGTCGCGGTTTC/3IABkFQ/	21	3022
Tag Forward Primer	AcDx-9036-FOXI3-RT-FP	TCTCATGGGCGCTAGTATCAAC	22	3023
Tag Reverse Primer	AcDx-9037-FOXI3-RT-RP	TGCGGGTATCAATCAGGGAAAC	22	3024
Downstream PCR Primer	AcDx-9038-FOXI3-PCR-V	TGCGGGTATCAATCAGGGAAACCGTACCTAAACGCTCCGCTGrCCGCT/3SpC3/	47	3025
C7orf51-S2				
Forward PCR Primer	AcDx-9641-C7orf51-S2-FP	GTTATGTTGAGGTTTTCGATTTTATTCrGCGTC/3SpC3/	32	3026
Reverse PCR Primer	AcDx-9642-C7orf51-S2-RP	GGTGTCGTGGCGAACCGACTAAAACGACAAArAACAG/3SpC3/	36	3027
Upstream LDR	AcDx-9643-C7orf51-S2-Up	TACTATCGTATCACGCCGACAGTTTTCGATTTTATTCGCGTTTTTTTTTTTTTCTCrGTTTA/3SpC3/	61	3028
Downstream LDR	AcDx-9644-C7orf51-S2-Dn	/5Phos/GTTCGCGGTATAGTTTTATTTCGCGTTGTCGTTGTGTAACGTCCGTGGGCTAA	53	3029
Real-Time Probe	AcDx-9645-C7orf51-S2-RT-Pb	/56-FAM/CCTTTTCTC/ZEN/GTTCGCGGTATAGTTTTATTTCGCGTTG/3IABkFQ/	37	3030
Tag Forward Primer	AcDx-9646-C7orf51-S2-RT-FP	TACTATCGTATCACGCCGACA	21	3031
Tag Reverse Primer	AcDx-9647-C7orf51-S2-RT-RP	TTAGCCCACGGACGTTACA	19	3032
Downstream PCR Primer	AcDx-9648-C7orf51-S2-PCR-V	TTAGCCCACGGACGTTACACGACTAAAACGACAAAAACAAAAACAATGrACAAT/3SpC3/	53	3033
C7orf51-S3				

Forward PCR Primer	AcDx-9651-C7orf51-S3-FP	GGATTTGTTTTTGAGAGGAGACrGTGGA/3SpC3/	27	3034
Reverse PCR Primer	AcDx-9652-C7orf51-S3-RP	GGTGTCGTGGCGAAAAAAAAAAAAAAACGCGAATAAAArUCGAG/3SpC3/	43	3035
Upstream LDR	AcDx-9653-C7orf51-S3-Up	TTGAGACCGCTGACCGACATTTTTGAGAGGAGACGTGGGTCTCrGGTGA/3SpC3/	48	3036
Downstream LDR	AcDx-9654-C7orf51-S3-Dn	/5Phos/GGTAGTGCGGATTCGTTCGGTTATGTTGAGGTGTTGCACGGTCGAGCTAA	50	3037
Real-Time Probe	AcDx-9655-C7orf51-S3-RT-Pb	/56-FAM/AAGGGTCTC/ZEN/GGTAGTGCGGATTCGTTC/3IABkFQ/	27	3038
Tag Forward Primer	AcDx-9656-C7orf51-S3-RT-FP	TTGAGACCGCTGACCGACA	19	3039
Tag Reverse Primer	AcDx-9657-C7orf51-S3-RT-RP	TTAGCTCGACCGTGCAACA	19	3040
Downstream PCR Primer	AcDx-9658-C7orf51-S3-PCR-V	TTAGCTCGACCGTGCAACACGAAAAAAAAAAAAAAACGCGAATAAAATTGrAAAAT/3SpC3/	55	3041
GP1BB-S2				
Forward PCR Primer	AcDx-9661-GP1BB-S2-FP	GTTAGGTCGTAGTATTGTGGTCrGGTGT/3SpC3/	27	3042
Reverse PCR Primer	AcDx-9662-GP1BB-S2-RP	GGTGTCGTGGCGACAACCCGACTTACTCTACrAACGG/3SpC3/	36	3043
Upstream LDR	AcDx-9663-GP1BB-S2-Up	TCGCAACGTGCCGAATACAGTGGTAGAGGCGGAGCrGGTCC/3SpC3/	40	3044
Downstream LDR	AcDx-9664-GP1BB-S2-Dn	/5Phos/GGTTTTATCGTTCGGTTTTTTCGTTTACGTTTTTCGTGTTGCACGGTCGAGCTAA	55	3045
Real-Time Probe	AcDx-9665-GP1BB-S2-RT-Pb	/56-FAM/TTGCGGAGC/ZEN/GGTTTTATCGTTCGGTTTTTTCG/3IABkFQ/	32	3046
Tag Forward Primer	AcDx-9666-GP1BB-S2-RT-FP	TCGCAACGTGCCGAATACA	19	3047
Tag Reverse Primer	AcDx-9667-GP1BB-S2-RT-RP	TTAGCTCGACCGTGCAACA	19	3048
Downstream PCR Primer	AcDx-9668-GP1BB-S2-PCR-V	TTAGCTCGACCGTGCAACACCCGACTTACTCTACAACGAAAAATGrUAAAT/3SpC3/	50	3049
CANCR73				
Forward PCR Primer	AcDx-9671-CANCR73-FP	GTTGATAACGTATGTGCGAAAGAGACrGCGGA/3SpC3/	31	3050
Reverse PCR Primer	AcDx-9672-CANCR73-RP	GGTGTCGTGGACTAAACGACCGACAAAAACGTArUATCT/3SpC3/	38	3051
Upstream LDR	AcDx-9673-CANCR73-Up	TAACGGGATTGAGAGTGGACAGCGAGGTTTAGTTAGTTTTTTTCGTATTTAAACrGTA	59	3052

		TA/3SpC3/		
Downstream LDR	AcDx-9674-CANCR73-Dn	/5Phos/GTACGTTTTAATTGTCGGGCGTTTTTATTTGATACGGTGTCTGCCGCCCTTACTAA	56	3053
Real-Time Probe	AcDx-9675-CANCR73-RT-Pb	/56-FAM/CCTTTAAAC/ZEN/GTACGTTTTAATTGTCGGGCGTTTTTATTT/3IABkFQ/	39	3054
Tag Forward Primer	AcDx-9676-CANCR73-RT-FP	TAACGGGATTGAGAGTGGACA	21	3055
Tag Reverse Primer	AcDx-9677-CANCR73-RT-RP	TTAGTAAGGGCGGCAGACA	19	3056
Downstream PCR Primer	AcDx-9678-CANCR73-PCR-V	TTAGTAAGGGCGGCAGACACTAAACGACCGACAAAAACGTATATCTGrUATCG/3SpC3/	52	3057
VWCE				
Forward PCR Primer	AcDx-9681-VWCE-FP	TTTGGGAGTCGGTTTTTTGCrGAAGT/3SpC3/	25	3058
Reverse PCR Primer	AcDx-9682-VWCE-RP	GGTGTCGTGGGAAACGCGCAACCTCTAATArCTAAT/3SpC3/	35	3059
Upstream LDR	AcDx-9683-VWCE-Up	TAACGGGATTGAGAGTGGACATTTGCGAAGCGTTGATTGAAAGCrGTTGA/3SpC3/	49	3060
Downstream LDR	AcDx-9684-VWCE-Dn	/5Phos/GTTAGATCGAGGTTTCGGGAGGAAATAGATTTCGGTGTCTGCCGCCCTTACTAA	54	3061
Real-Time Probe	AcDx-9685-VWCE-RT-Pb	/56-FAM/CCTGAAAGC/ZEN/GTTAGATCGAGGTTTCGGGAGG/3IABkFQ/	31	3062
Tag Forward Primer	AcDx-9686-VWCE-RT-FP	TAACGGGATTGAGAGTGGACA	21	3063
Tag Reverse Primer	AcDx-9687-VWCE-RT-RP	TTAGTAAGGGCGGCAGACA	19	3064
Downstream PCR Primer	AcDx-9688-VWCE-PCR-V	TTAGTAAGGGCGGCAGACACGCAACCTCTAATACTAACCTTATTCCTGrAAATT/3SpC3/	53	3065
FAM59B-S2				
Forward PCR Primer	AcDx-9691-FAM59B-S2-FP	GAGGCGGTGAGTGAGCrGCGTC/3SpC3/	21	3066
Reverse PCR Primer	AcDx-9692-FAM59B-S2-RP	GGTGTCGTGGCCACATACTCCGAAACCGAAArCCCAG/3SpC3/	36	3067
Upstream LDR	AcDx-9693-FAM59B-S2-Up	TTGGTACGAGGAGGGCACATGAGCGCGTTGGGAGTCrGAGCC/3SpC3/	41	3068
Downstream LDR	AcDx-9694-FAM59B-S2-Dn	/5Phos/GAGTTTCGGGTTTAGTTAGGGTTTAAAGTTCGTTTTTTTGTGTCTGCCGCCCTTACTAA	59	3069
Real-Time Probe	AcDx-9695-FAM59B-S2-RT-Pb	/56-FAM/AAGGGAGTC/ZEN/GAGTTTCGGGTTTAGTTAGGGTTTAAAG/3IABkFQ/	37	3070

Tag Forward Primer	AcDx-9696-FAM59B-S2-RT-FP	TTGGTACGAGGAGGGCACA	19	3071
Tag Reverse Primer	AcDx-9697-FAM59B-S2-RT-RP	TTAGTAAGGGCGGCAGACA	19	3072
Downstream PCR Primer	AcDx-9698-FAM59B-S2-PCR-V	TTAGTAAGGGCGGCAGACACCGAAACCGAAACCCAAAAAAATGrAACTC/3SpC3/	48	3073
FAM59B-S3				
Forward PCR Primer	AcDx-9701-FAM59B-S3-FP	TGTTATAAGTTGGTTAGTATTATTTTTAAGACrGGTGA/3SpC3/	37	3074
Reverse PCR Primer	AcDx-9702-FAM59B-S3-RP	GGTGTCGTGGCGCGACGTATCCGTAAACArACAAG/3SpC3/	34	3075
Upstream LDR	AcDx-9703-FAM59B-S3-Up	TCAGACGCACTAAACAGGCAAGGTGGTGTTGAGGTTAGCrGTTAT/3SpC3/	44	3076
Downstream LDR	AcDx-9704-FAM59B-S3-Dn	/5Phos/GTTGCGTCGCGAGGGTTCGGTTGCGGATCGTCGTGTGAA	39	3077
Real-Time Probe	AcDx-9705-FAM59B-S3-RT-Pb	/56-FAM/TTGGTTAGC/ZEN/GTTGCGTCGCGAGGG/3IABkFQ/	24	3078
Tag Forward Primer	AcDx-9706-FAM59B-S3-RT-FP	TCAGACGCACTAAACAGGCAA	21	3079
Tag Reverse Primer	AcDx-9707-FAM59B-S3-RT-RP	TTCACACGACGATCCGCAA	19	3080
Downstream PCR Primer	AcDx-9708-FAM59B-S3-PCR-V	TTCACACGACGATCCGCAAGTAAACAACAAAAAATACAACGACGCTGrAACCT/3SpC3/	52	3081
CANCR74				
Forward PCR Primer	AcDx-9711-CANCR74-FP	GTTGTAGTTTTAGTTTTTTTTTTAGATGTCrGGTAA/3SpC3/	35	3082
Reverse PCR Primer	AcDx-9712-CANCR74-RP	GGTGTCGTGGCCCTAAAAACCGTACTCCGTArUCCGG/3SpC3/	36	3083
Upstream LDR	AcDx-9713-CANCR74-Up	TTCCATCGAGCGCCAACAAGTAGTATTAGTTTTTGTTTGTATGAGGTTGCrGAGAC/3SpC3/	55	3084
Downstream LDR	AcDx-9714-CANCR74-Dn	/5Phos/GAGGTTTGTAGTGATATTTTTCGAGTTTTTTGATTTGTTTCGTTGCGGATCGTCGTGTGAA	61	3085
Real-Time Probe	AcDx-9715-CANCR74-RT-Pb	/56-FAM/AAGGGTTGC/ZEN/GAGGTTTGTAGTGATATTTTTCGAGTTTTTTG/3IABkFQ/	41	3086

Tag Forward Primer	AcDx-9716-CANCR74-RT-FP	TTCCATCGAGCGCCAACAA	19	3087
Tag Reverse Primer	AcDx-9717-CANCR74-RT-RP	TTCACACGACGATCCGCAA	19	3088
Downstream PCR Primer	AcDx-9718-CANCR74-PCR-V	TTCACACGACGATCCGCAACCCTAAAAACCGTACTCCGTATCTGrAAACG/3SpC3/	49	3089
MAST1				
Forward PCR Primer	AcDx-9721-MAST1-FP	GTAGTAAGCGATTTTTCGCGTTCrGAGGC/3SpC3/	28	3090
Reverse PCR Primer	AcDx-9722-MAST1-RP	GGTGTCGTGGATCCTCGTCCTCTTAAAAAACGCrUAAAG/3SpC3/	38	3091
Upstream LDR	AcDx-9723-MAST1-Up	TTCAGAGCACCTGCGTACCGAGGTTAGTTTTTTTTTGGGCGTTCATCrGTCAC/3SpC3/	52	3092
Downstream LDR	AcDx-9724-MAST1-Dn	/5Phos/GTCGTTTTTCGGCGTTGTTGGAGTTTAGTCGGGTTCTTCGGCTGGCTCAA	50	3093
Real-Time Probe	AcDx-9725-MAST1-RT-Pb	/56-FAM/TTGTTCATC/ZEN/GTCGTTTTTCGGCGTTGTTGGAG/3IABkFQ/	32	3094
Tag Forward Primer	AcDx-9726-MAST1-RT-FP	TTCAGAGCACCTGCGTACC	19	3095
Tag Reverse Primer	AcDx-9727-MAST1-RT-RP	TTGAGCCAGCCGAAGAACC	19	3096
Downstream PCR Primer	AcDx-9728-MAST1-PCR-V	TTGAGCCAGCCGAAGAACCATCCTCGTCCTCTTAAAAAACGCTAAAATGrACTAG/3SpC3/	54	3097
LMX1B				
Forward PCR Primer	AcDx-9731-LMX1B-FP	GTTTTCGTCGTCGTTCGTTCrGGTTT/3SpC3/	25	3098
Reverse PCR Primer	AcDx-9732-LMX1B-RP	GGTGTCGTGGTATTTTAAAACAAAAAAAAAACCTATTAATAAAAArUCCAG/3SpC3/	49	3099
Upstream LDR	AcDx-9733-LMX1B-Up	TCCGGCCTTTGACGATACCGTTCGTTCGGTTCGCGATCrGTCAC/3SpC3/	43	3100
Downstream LDR	AcDx-9734-LMX1B-Dn	/5Phos/GTCGTATTAAAGTAGATGTTTGTAGTTGGTTTGAGTTGGAGGTAATTCACTCGAACGGAGCA	62	3101
Real-Time Probe	AcDx-9735-LMX1B-RT-Pb	/56-FAM/AACGCGATC/ZEN/GTCGTATTAAAGTAGATGTTTGTAGTT/3IABkFQ/	36	3102
Tag Forward Primer	AcDx-9736-LMX1B-RT-FP	TCCGGCCTTTGACGATACC	19	3103
Tag Reverse Primer	AcDx-9737-LMX1B-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	3104

CANCR75				
Forward PCR Primer	AcDx-9741-CANCR75-FP	CGTTCGTGGGTGGAGTTCrGTTTC/3SpC3/	23	3105
Reverse PCR Primer	AcDx-9742-CANCR75-RP	GGTGTCGTGGGCAAATCCAAAACCTCCGAArCGCTG/3SpC3/	35	3106
Upstream LDR	AcDx-9743-CANCR75-Up	TTTCGCTCGACGCATACCAGGGTGGAGTTCGTTTTTTGGTTGGTCrGTACA/3SpC3/	50	3107
Downstream LDR	AcDx-9744-CANCR75-Dn	/5Phos/GTATGGTGTTCGTACGGTTTTTTTTTTTTTTTTAGCGTTTAGTGGCGCGGCTACTGTAAAA	61	3108
Real-Time Probe	AcDx-9745-CANCR75-RT-Pb	/56-FAM/AAGTTGGTC/ZEN/GTATGGTGTTCGTACGGTTTTTTTTTTTTTTTTAG/3IABkFQ/	44	3109
Tag Forward Primer	AcDx-9746-CANCR75-RT-FP	TTTCGCTCGACGCATACCA	19	3110
Tag Reverse Primer	AcDx-9747-CANCR75-RT-RP	TTTTACAGTAGCCGCGCCA	19	3111
Downstream PCR Primer	AcDx-9748-CANCR75-PCR-V	TTTTACAGTAGCCGCGCCATCCAAAACCTCCGAACGCTAAATGrCTAAG/3SpC3/	48	3112
PON3-S2				
Forward PCR Primer	AcDx-9751-PON3-S2-FP	GGGTGTCGTAGTAGGGCrGTTGG/3SpC3/	22	3113
Reverse PCR Primer	AcDx-9752-PON3-S2-RP	GGTGTCGTGGCCTCTACCCAAAAAACAAAAAATCGTArAACGC/3SpC3/	42	3114
Upstream LDR	AcDx-9753-PON3-S2-Up	TTCTTGCCCGCTTGTTCCAGTAGGGCGTTGACGAGTTTCATCrGAGCC/3SpC3/	47	3115
Downstream LDR	AcDx-9754-PON3-S2-Dn	/5Phos/GAGTTTCGTCGTTCGGGTTTAAGGTCGTTTTTACGTGGCGCGGCTACTGTAAAA	54	3116
Real-Time Probe	AcDx-9755-PON3-S2-RT-Pb	/56-FAM/CCTTTCATC/ZEN/GAGTTTCGTCGTTCGGGTTTAAGGTC/3IABkFQ/	35	3117
Tag Forward Primer	AcDx-9756-PON3-S2-RT-FP	TTCTTGCCCGCTTGTTCCA	19	3118
Tag Reverse Primer	AcDx-9757-PON3-S2-RT-RP	TTTTACAGTAGCCGCGCCA	19	3119
Downstream PCR Primer	AcDx-9758-PON3-S2-PCR-V	TTTTACAGTAGCCGCGCCACCTCTACCCAAAAAACAAAAAATCGTAAATGrUAAAG/3SpC3/	55	3120
TMEM101-S2				
Forward PCR Primer	AcDx-9761-TMEM101-S2-FP	TTTTGGGTATTGGTTTTTGATTTTTCrGTTAC/3SpC3/	31	3121

Reverse PCR Primer	AcDx-9762-TMEM101-S2-RP	GGTGTCGTGGCCATTAACACCTACGCCGCrCTACG/3SpC3/	34	3122
Upstream LDR	AcDx-9763-TMEM101-S2-Up	TCCCTTAGAGAGAACGCCCAGGAAGATTTTTTTGAGGTCGAAGCrGTTAT/3SpC3/	49	3123
Downstream LDR	AcDx-9764-TMEM101-S2-Dn	/5Phos/GTTTATTTTTAGTTAGTTTTCGTAGTGGACGTAGTTTTCGATGTAGTGGTGACGTACGAGTGTTCTTA	68	3124
Real-Time Probe	AcDx-9765-TMEM101-S2-RT-Pb	/56-FAM/AATCGAAGC/ZEN/GTTTATTTTTAGTTAGTTTTCGTAGTGGACG/3IABkFQ/	40	3125
Tag Forward Primer	AcDx-9766-TMEM101-S2-RT-FP	TCCCTTAGAGAGAACGCCCA	20	3126
Tag Reverse Primer	AcDx-9767-TMEM101-S2-RT-RP	TAAGAACACTCGTACGTCACCA	22	3127
Downstream PCR Primer	AcDx-9768-TMEM101-S2-PCR-V	TAAGAACACTCGTACGTCACCACACCTACGCCGCCTACATTGrAAAAT/3SpC3/	47	3128
TMEM101-S3				
Forward PCR Primer	AcDx-9771-TMEM101-S3-FP	CGTATAGTACGGTTGTTTTTATGTCrGAAAC/3SpC3/	30	3129
Reverse PCR Primer	AcDx-9772-TMEM101-S3-RP	GGTGTCGTGGACTTCTTCCCCCACCGCrUTCAC/3SpC3/	32	3130
Upstream LDR	AcDx-9773-TMEM101-S3-Up	TCTCATACCAGACGCGGTAACGGTATTGGGATGTCGGGTTTTTCGCrGAGCT/3SpC3/	51	3131
Downstream LDR	AcDx-9774-TMEM101-S3-Dn	/5Phos/GAGTCGGGAGTGGAGAAGTAACGAGGATAGGTTCGTGTCGCTGTGCTTA	49	3132
Real-Time Probe	AcDx-9775-TMEM101-S3-RT-Pb	/56-FAM/AATTTTCGC/ZEN/GAGTCGGGAGTGGAG/3IABkFQ/	24	3133
Tag Forward Primer	AcDx-9776-TMEM101-S3-RT-FP	TCTCATACCAGACGCGGTAAC	21	3134
Tag Reverse Primer	AcDx-9777-TMEM101-S3-RT-RP	TAAGCACAGCGACACGAAC	19	3135
Downstream PCR Primer	AcDx-9778-TMEM101-S3-PCR-V	TAAGCACAGCGACACGAACCCCACCGCTTCATTCTATCCTTGrUTACC/3SpC3/	47	3136

CANCR76				
Forward PCR Primer	AcDx-9781-CANCR76-FP	ATTAGGTTTATTTGACGTTTTAGGTCrGATAA/3SpC3/	31	3137
Reverse PCR Primer	AcDx-9782-CANCR76-RP	GGTGTCGTGGCTAATCCCGAAACAAAAAACGCrCTAAG/3SpC3/	37	3138
Upstream LDR	AcDx-9783-CANCR76-Up	TTTTCGGCGGCAGCTAAACGACGTTTTAGGTCGATAGTAAATTTCGCGCrGTTCT/3SpC3/	54	3139
Downstream LDR	AcDx-9784-CANCR76-Dn	/5Phos/GTTTCGTTGGTTCGGTTACGTTCGTAGTTTGTTTAGGTTCGTGTCGCTGTGCTTA	55	3140
Real-Time Probe	AcDx-9785-CANCR76-RT-Pb	/56-FAM/CCTTCGCGC/ZEN/GTTTCGTTGGTTCGG/3IABkFQ/	24	3141
Tag Forward Primer	AcDx-9786-CANCR76-RT-FP	TTTTCGGCGGCAGCTAAAC	19	3142
Tag Reverse Primer	AcDx-9787-CANCR76-RT-RP	TAAGCACAGCGACACGAAC	19	3143
Downstream PCR Primer	AcDx-9788-CANCR76-PCR-V	TAAGCACAGCGACACGAACCAAAAAACGCCTAAACAAACTACGAATGrUAACT/3SpC3/	52	3144
TXNRD1-S2				
Forward PCR Primer	AcDx-9791-TXNRD1-S2-FP	TTTGAAGAAGATATACGGGTTATGATTTCrGTTGC/3SpC3/	34	3145
Reverse PCR Primer	AcDx-9792-TXNRD1-S2-RP	GGTGTCGTGGCGAACCCCCTACCGACrCGCGG/3SpC3/	31	3146
Upstream LDR	AcDx-9793-TXNRD1-S2-Up	TCCGGACCTTCATCCTCCAGAAGATATACGGGTTATGATTTCGTTGTTGTTGTCrGAGTA/3SpC3/	59	3147
Downstream LDR	AcDx-9794-TXNRD1-S2-Dn	/5Phos/GAGCGTTTCGTTTATCGCGTTTTTCGATTCGCTGGGCAGGAACACGATAGTA	52	3148
Real-Time Probe	AcDx-9795-TXNRD1-S2-RT-Pb	/56-FAM/AATGTTGTC/ZEN/GAGCGTTTCGTTTATCGCG/3IABkFQ/	28	3149
Tag Forward Primer	AcDx-9796-TXNRD1-S2-RT-FP	TCCGGACCTTCATCCTCCA	19	3150
Tag Reverse Primer	AcDx-9797-TXNRD1-S2-RT-RP	TACTATCGTGTTCCTGCCCA	20	3151
Downstream PCR Primer	AcDx-9798-TXNRD1-S2-PCR-V	TACTATCGTGTTCCTGCCCACCCTACCGACCGCGAATTGrAAAAG/3SpC3/	44	3152
LOC389333-S2				

Forward PCR Primer	AcDx-9801-LOC389333-S2-FP	GAAGGACGGTTTAGGAGGTTCrGTAGA/3SpC3/	26	3153
Reverse PCR Primer	AcDx-9802-LOC389333-S2-RP	GGTGTCGTGGAAAACCGCCAACACTAAAACCrCCCCA/3SpC3/	36	3154
Upstream LDR	AcDx-9803-LOC389333-S2-Up	TTAGCCGCCAAACGTACCAGGTTCGTAGAGGTGCGGTATTGTCrGGTGA/3SpC3/	48	3155
Downstream LDR	AcDx-9804-LOC389333-S2-Dn	/5Phos/GGTAGAAAATTTTTGATGAAAACGGAAGTGTAGTGAGTCTGGGCAGGAACACGATAGTA	59	3156
Real-Time Probe	AcDx-9805-LOC389333-S2-RT-Pb	/56-FAM/TTTATTGTC/ZEN/GGTAGAAAATTTTTGATGAAAACGGAAGTGTA/3IABkFQ/	41	3157
Tag Forward Primer	AcDx-9806-LOC389333-S2-RT-FP	TTAGCCGCCAAACGTACCA	19	3158
Tag Reverse Primer	AcDx-9807-LOC389333-S2-RT-RP	TACTATCGTGTTCCTGCCCA	20	3159
Downstream PCR Primer	AcDx-9808-LOC389333-S2-PCR-V	TACTATCGTGTTCCTGCCCAGCCAACACTAAAACCCCCTGrCCCCA/3SpC3/	45	3160
NPHS2-S2				
Forward PCR Primer	AcDx-9811-NPHS2-S2-FP	TCGAGGGATTTTAAAGATCGTCrGGGTC/3SpC3/	27	3161
Reverse PCR Primer	AcDx-9812-NPHS2-S2-RP	GGTGTCGTGGAACTATCGAAATTCCGAATACGCrCCCGT/3SpC3/	38	3162
Upstream LDR	AcDx-9813-NPHS2-S2-Up	TAGCAGCTGAACAACCCAACGGTTGAGGGCGAGGAGCrGGGCC/3SpC3/	42	3163
Downstream LDR	AcDx-9814-NPHS2-S2-Dn	/5Phos/GGGATATCGGAGTCGCGGGCGTTGTATGGTCGGCATGCTA	40	3164
Real-Time Probe	AcDx-9815-NPHS2-S2-RT-Pb	/56-FAM/TTGAGGAGC/ZEN/GGGATATCGGAGTCG/3IABkFQ/	24	3165
Tag Forward Primer	AcDx-9816-NPHS2-S2-RT-FP	TAGCAGCTGAACAACCCAAC	20	3166
Tag Reverse Primer	AcDx-9817-NPHS2-S2-RT-RP	TAGCATGCCGACCATACAAC	20	3167
Downstream PCR Primer	AcDx-9818-NPHS2-S2-PCR-V	TAGCATGCCGACCATACAACCTATCGAAATTCCGAATACGCCCTGrCCCGT/3SpC3/	50	3168
CANCR77				

Forward PCR Primer	AcDx-9821-CANCR77-FP	CGGGAAAGCGAAATTCGTTCrGGTTC/3SpC3/	25	3169
Reverse PCR Primer	AcDx-9822-CANCR77-RP	GGTGTCGTGGAAAAATCTTACAAACAAAATATTAACGAAAArATCCA/3SpC3/	46	3170
Upstream LDR	AcDx-9823-CANCR77-Up	TCTGCCCTTCGCTTCGAACTTCGGTTTATTTAAGTTTCGTTGTTTAGCrGTTCC/3SpC3/	53	3171
Downstream LDR	AcDx-9824-CANCR77-Dn	/5Phos/GTTTTTTAGGGTCGTTTTTGGGAACGGGCGGTTGTATGGTCGGCATGCTA	50	3172
Real-Time Probe	AcDx-9825-CANCR77-RT-Pb	/56-FAM/AAGTTTAGC/ZEN/GTTTTTTAGGGTCGTTTTTGGGAACG/3IABkFQ/	35	3173
Tag Forward Primer	AcDx-9826-CANCR77-RT-FP	TCTGCCCTTCGCTTCGAAC	19	3174
Tag Reverse Primer	AcDx-9827-CANCR77-RT-RP	TAGCATGCCGACCATACAAC	20	3175
Downstream PCR Primer	AcDx-9828-CANCR77-PCR-V	TAGCATGCCGACCATACAACAAAAATCTTACAAACAAAATATTAACGAAAAATCTGrCCCGC/3SpC3/	61	3176

IDT Abbreviation	Modifications
/5Phos/	5' Phosphorylation
rX (X=A,C,G,U)	RNA Base
/3spC3/	3' C3 DNA Spacer
/56-FAM/	5' 6-FAM™ Fluorescent Tag
/Zen/	Internal Quencher
/3IABkFQ/	3' Iowa Black® FQ Quencher

Table 55. Primers for use in Step 2 of the Group 3- 60-64-marker assay, with average sensitivities of 50%, to detect and identify lung adenocarcinomas, lung squamous cell carcinoma, and head & neck cancers.

Site	Primer Name	Sequence	Length	Seq. ID No.
Prefered Group 3 Markers				
HOXA9-S1				
Forward PCR Primer	AcDx-7071-HOXA9-S1-FP	GTTAGCGTCGTCGTTTGTCrGGGAG/3SpC3/	24	3177
Reverse PCR Primer	AcDx-7072-HOXA9-S1-RP	GGTGTCGTGGGTAAACTCGTTCCTACTAAACGCrCGACA/3SpC3/	38	3178

Upstream LDR	AcDx-7073-HOXA9-S1-Up	TTGCAAACCACCCGGACAAGAGGAGGTTGGTTTAGGGTTCTCrGGCAC/3SpC3/	47	3179
Downstream LDR	AcDx-7074-HOXA9-S1-Dn	/5Phos/GGCGTATAGCGGTTAACGTTTAGTTTATTCGCGGTTGGTCAGCATCGACTCCTA	54	3180
Real-Time Probe	AcDx-7075-HOXA9-S1-RT-Pb	/56-FAM/TTGGTTCTC/ZEN/GGCGTATAGCGGTTAACGTTTAGT/3IABkFQ/	33	3181
Tag Forward Primer	AcDx-7076-HOXA9-S1-RT-FP	TTGCAAACCACCCGGACAA	19	3182
Tag Reverse Primer	AcDx-7077-HOXA9-S1-RT-RP	TAGGAGTCGATGCTGACCAA	20	3183
Downstream PCR Primer	AcDx-7078-HOXA9-S1-PCR-V	TAGGAGTCGATGCTGACCAACCTACTAAACGCCGACGCTGrCGAAC/3SpC3/	45	3184
WNT7B-S1				
Forward PCR Primer	AcDx-7081-WNT7B-S1-FP	CGTGTATGTCGGTGTTTGTACrGAGGA/3SpC3/	26	3185
Reverse PCR Primer	AcDx-7082-WNT7B-S1-RP	GGTGTCGTGGTCCTAAACCAACGAAAAACCCrCTCCT/3SpC3/	36	3186
Upstream LDR	AcDx-7083-WNT7B-S1-Up	TAAGACGTATGCTAGCGCCAAGAGCGGGTGTGTGAGCGCrGGTCC/3SpC3/	44	3187
Downstream LDR	AcDx-7084-WNT7B-S1-Dn	/5Phos/GGTTTTTTTAAGTGTGGTATGGTATTGCGCGTCGTTGGTCAGCATCGACTCCTA	54	3188
Real-Time Probe	AcDx-7085-WNT7B-S1-RT-Pb	/56-FAM/AATGAGCGC/ZEN/GGTTTTTTTAAGTGTGGTATGGT/3IABkFQ/	32	3189
Tag Forward Primer	AcDx-7086-WNT7B-S1-RT-FP	TAAGACGTATGCTAGCGCCAA	21	3190
Tag Reverse Primer	AcDx-7087-WNT7B-S1-RT-RP	TAGGAGTCGATGCTGACCAA	20	3191
Downstream PCR Primer	AcDx-7088-WNT7B-S1-PCR-V	TAGGAGTCGATGCTGACCAACATATTAACCCACGCGACGTGrCAATG/3SpC3/	46	3192
HOXD9-S1				
Forward PCR Primer	AcDx-7091-HOXD9-S1-FP	GATTACGTGGGTCGCGCrGATTG/3SpC3/	22	3193
Reverse PCR Primer	AcDx-7092-HOXD9-S1-RP	GGTGTCGTGGACATTTTAAAACGTCCCGCACrUCCCG/3SpC3/	36	3194
Upstream LDR	AcDx-7093-HOXD9-S1-	TACATGCCATCCCACGACATCGGTGGTTCGGGCATCrGGCAA/3SpC3/	41	3195

	Up			
Downstream LDR	AcDx-7094-HOXD9-S1-Dn	/5Phos/GGCGAGGAGTTGTTCGGCGGTGTGTCGGAGCGGTTACTA	39	3196
Real-Time Probe	AcDx-7095-HOXD9-S1-RT-Pb	/56-FAM/AAGAGCATC/ZEN/GGCGAGGAGTTG/3IABkFQ/	21	3197
Tag Forward Primer	AcDx-7096-HOXD9-S1-RT-FP	TACATGCCATCCCACGACA	19	3198
Tag Reverse Primer	AcDx-7097-HOXD9-S1-RT-RP	TAGTAACCGCTCCGACACA	19	3199
Downstream PCR Primer	AcDx-7098-HOXD9-S1-PCR-V	TAGTAACCGCTCCGACACAACATTACACTATCCGCCGCTGrAACAG/3SpC3/	45	3200
NRN1				
Forward PCR Primer	AcDx-7101-NRN1-FP	GGTAGTTTTTTGGCGGTTGCrGTTTA/3SpC3/	25	3201
Reverse PCR Primer	AcDx-7102-NRN1-RP	GGTGTCGTGGCTTCGACGTCTAACCCGArCGCTT/3SpC3/	33	3202
Upstream LDR	AcDx-7103-NRN1-Up	TAGGGCGACAGTTACCACAAGTGTTTCGGGAGGATCGGATATTTTAATTTCTCrGGTCC/3SpC3/	58	3203
Downstream LDR	AcDx-7104-NRN1-Dn	/5Phos/GGTTTTTAACGCGGGCGTTTGTTCGCGTTGTGGGTCTCGCTCGTATA	47	3204
Real-Time Probe	AcDx-7105-NRN1-RT-Pb	/56-FAM/CCATTTCTC/ZEN/GGTTTTTAACGCGGGCGTTTGT/3IABkFQ/	31	3205
Tag Forward Primer	AcDx-7106-NRN1-RT-FP	TAGGGCGACAGTTACCACAA	20	3206
Tag Reverse Primer	AcDx-7107-NRN1-RT-RP	TATACGAGCGAGACCCACAA	20	3207
Downstream PCR Primer	AcDx-7108-NRN1-PCR-V	TATACGAGCGAGACCCACAATCTAACCCGACGCTCGTGrAACAG/3SpC3/	43	3208
CaNCR23				
Forward PCR Primer	AcDx-7111-CaNCR23-FP	ATTTGTTATTCGCGTGCGTCrGTTTC/3SpC3/	25	3209
Reverse PCR Primer	AcDx-7112-CaNCR23-RP	GGTGTCGTGGATTCGAAACACTACTCTAATACGATCCrUAATG/3SpC3/	42	3210
Upstream LDR	AcDx-7113-CaNCR23-Up	TCCGACTTTAGTGCGTCACAAATTTTGGTTATATTTTTTTTTATTTGCGGTGTTTTATCTCrGCGCG/3SpC3/	66	3211
Downstream LDR	AcDx-7114-CaNCR23-Dn	/5Phos/GCGTAAGATGCGTTGATAGAGGTTATTTTAAACGAATTTTTGTGGGTCTCGCTCGTATA	59	3212

Real-Time Probe	AcDx-7115-CaNCR23-RT-Pb	/56-FAM/AATTATCTC/ZEN/GCGTAAGATGCGTTGATAGAGGTTATT/3IABkFQ/	36	3213
Tag Forward Primer	AcDx-7116-CaNCR23-RT-FP	TCCGACTTTAGTGCGTCACAA	21	3214
Tag Reverse Primer	AcDx-7117-CaNCR23-RT-RP	TATACGAGCGAGACCCACAA	20	3215
Downstream PCR Primer	AcDx-7118-CaNCR23-PCR-V	TATACGAGCGAGACCCACAAAACACTACTCTAATACGATCCTAATAAAAAATAAATTTGrUTTAG/3SpC3/	64	3216
PRRX1				
Forward PCR Primer	AcDx-7121-PRRX1-FP	GGATGTAAATATAAAATAGCGACGCrGGGAG/3SpC3/	30	3217
Reverse PCR Primer	AcDx-7122-PRRX1-RP	GGTGTCGTGGGGCGAAAAAAAATTAACGAAACAAAATCrCCGAG/3SpC3/	43	3218
Upstream LDR	AcDx-7123-PRRX1-Up	TCAAACAAAGGCGACCACAACATCGATTTGTTATAAAGGGAGAGGTGTAGACrGTAAT/3SpC3/	57	3219
Downstream LDR	AcDx-7124-PRRX1-Dn	/5Phos/GTAGCGTAAAGGAATTGTTTGTTTAATTTATTAGTTATATTTTTTCGGGTTGTCGCATAGGCAGTTCATA	70	3220
Real-Time Probe	AcDx-7125-PRRX1-RT-Pb	/56-FAM/AATGTAGAC/ZEN/GTAGCGTAAAGGAATTGTTTGTTTAATTTATT/3IABkFQ/	41	3221
Tag Forward Primer	AcDx-7126-PRRX1-RT-FP	TCAAACAAAGGCGACCACAAC	21	3222
Tag Reverse Primer	AcDx-7127-PRRX1-RT-RP	TATGAACTGCCTATGCGACAAC	22	3223
Downstream PCR Primer	AcDx-7128-PRRX1-PCR-V	TATGAACTGCCTATGCGACAACGCGAAAAAAAATTAACGAAACAAAATCCTGrAAAG/3SpC3/	56	3224
CaNCR24				
Forward PCR Primer	AcDx-7131-CaNCR24-FP	CGAGTTGTAAAGTTGTTGTCGCrGGCGC/3SpC3/	27	3225
Reverse PCR Primer	AcDx-7132-CaNCR24-RP	GGTGTCGTGGCGAACGAAAAAAAACCCCGACrCAAAT/3SpC3/	36	3226
Upstream LDR	AcDx-7133-CaNCR24-Up	TAAACAATGAGACCCGCTGAACGGCGTCGGGAACGGCGCrGCGCC/3SpC3/	44	3227
Downstream LDR	AcDx-7134-CaNCR24-Dn	/5Phos/GCGTTTAATTTTTAGCGGGAGTCGTTAGGTTTGGTTGTCGCATAGGCAGTTCATA	55	3228
Real-Time Probe	AcDx-7135-CaNCR24-RT-Pb	/56-FAM/TTACGGCGC/ZEN/GCGTTTAATTTTTAGCGG/3IABkFQ/	27	3229
Tag Forward Primer	AcDx-7136-CaNCR24-RT-FP	TAAACAATGAGACCCGCTGAAC	22	3230

Tag Reverse Primer	AcDx-7137-CaNCR24-RT-RP	TATGAACTGCCTATGCGACAAC	22	3231
Downstream PCR Primer	AcDx-7138-CaNCR24-PCR-V	TATGAACTGCCTATGCGACAACCCCGACCAAACCAAACCTAATGrACTT/3SpC3/	48	3232
NR5A2-S1				
Forward PCR Primer	AcDx-7151-NR5A2-S1-FP	ATGTGCGGGTCGGCrGGGTC/3SpC3/	19	3233
Reverse PCR Primer	AcDx-7152-NR5A2-S1-RP	GGTGTCGTGGTCTACTCTCAACACCTCCCAArUCCTC/3SpC3/	36	3234
Upstream LDR	AcDx-7153-NR5A2-S1-Up	TCATCGCCCTCAGATCTTCCAGGTCGGCGGGTTTGTGATCTCrGGAGT/3SpC3/	47	3235
Downstream LDR	AcDx-7154-NR5A2-S1-Dn	/5Phos/GGAACGTTTTTTGTTATTTTTTTGCGCGAATTTGAAAGTGGAGGATAGATTGGAGGGCA	59	3236
Real-Time Probe	AcDx-7155-NR5A2-S1-RT-Pb	/56-FAM/AATGATCTC/ZEN/GGAACGTTTTTTGTTATTTTTTTGCG/3IABkFQ/	35	3237
Tag Forward Primer	AcDx-7156-NR5A2-S1-RT-FP	TCATCGCCCTCAGATCTTCCA	21	3238
Tag Reverse Primer	AcDx-7157-NR5A2-S1-RT-RP	TGCCCTCCAATCTATCCTCCA	21	3239
Downstream PCR Primer	AcDx-7158-NR5A2-S1-PCR-V	TGCCCTCCAATCTATCCTCCACACCTCCCAATCCTTTCAAATTTGrCGCAG/3SpC3/	50	3240
HOXA10				
Forward PCR Primer	AcDx-7231-HOXA10-FP	GGTAAGATCGAGGCGCrGTTTG/3SpC3/	21	3241
Reverse PCR Primer	AcDx-7232-HOXA10-RP	GGTGTCGTGGCGCTAAACGACAAACGCAArUAAAG/3SpC3/	34	3242
Upstream LDR	AcDx-7233-HOXA10-Up	TCGCTCTTCAGCCTCCTACAGAGGGTTCGTAGTCGTGCGTCTCrGGGCC/3SpC3/	48	3243
Downstream LDR	AcDx-7234-HOXA10-Dn	/5Phos/GGGATTTAGATTTTCGTTATCGTTATCGTTGTTCGGCTGTTCTGGGAATTATTGCCGGA	59	3244
Real-Time Probe	AcDx-7235-HOXA10-RT-Pb	/56-FAM/AAGCGTCTC/ZEN/GGGATTTAGATTTTCGTTATCGTTATCG/3IABkFQ/	37	3245
Tag Forward Primer	AcDx-7236-HOXA10-RT-FP	TCGCTCTTCAGCCTCCTACA	20	3246
Tag Reverse Primer	AcDx-7237-HOXA10-RT-RP	TCCGGCAATAATTCCCAGAACA	22	3247

Downstream PCR Primer	AcDx-7238-HOXA10-PCR-V	TCCGGCAATAATTCCCAGAACAACGCAATAAAACAACGTCGCTGrAACAG/3SpC3/	49	3248
CaNCR26				
Forward PCR Primer	AcDx-7261-CaNCR26-FP	CGGGACGGGTTTTTTTGCrGGATC/3SpC3/	23	3249
Reverse PCR Primer	AcDx-7262-CaNCR26-RP	GGTGTCGTGGACACCTAAAACAATAACAACCGCrCCGAT/3SpC3/	38	3250
Upstream LDR	AcDx-7263-CaNCR26-Up	TCATCTGTTCGTCAGGGTCCAGATTTTTTGAAATGAAATAATGTGATGTACGTTGCrGATGG/3SpC3/	61	3251
Downstream LDR	AcDx-7264-CaNCR26-Dn	/5Phos/GATAAGGGTCGGTTTGTAATGAGGTTTAGGTCGTGGTGACTTTACCCGGAGGA	53	3252
Real-Time Probe	AcDx-7265-CaNCR26-RT-Pb	/56-FAM/AAACGTTGC/ZEN/GATAAGGGTCGGTTTGTAATGAGG/3IABkFQ/	33	3253
Tag Forward Primer	AcDx-7266-CaNCR26-RT-FP	TCATCTGTTCGTCAGGGTCCA	21	3254
Tag Reverse Primer	AcDx-7267-CaNCR26-RT-RP	TCCTCCGGGTAAAGTCACCA	20	3255
Downstream PCR Primer	AcDx-7268-CaNCR26-PCR-V	TCCTCCGGGTAAAGTCACCACTAAAACAATAACAACCGCCCGATGrACCTG/3SpC3/	50	3256
CaNCR38				
Forward PCR Primer	AcDx-7761-CaNCR38-FP	CGGATCGTAGGTTGGGCrGGTTC/3SpC3/	22	3257
Reverse PCR Primer	AcDx-7762-CaNCR38-RP	GGTGTCGTGGCCGCGACCTAAAAACGCrUCCCT/3SpC3/	32	3258
Upstream LDR	AcDx-7763-CaNCR38-Up	TCGCGGAAAGTCCCAGTAACGTTGGGCGGTTTTTGATTTTTTGCrGTTCC/3SpC3/	49	3259
Downstream LDR	AcDx-7764-CaNCR38-Dn	/5Phos/GTTTTTCGTTTATAGTCGGAGTTCGGTAGTTGGAAGTGTTGGCCTGTAAGCGTTCCA	57	3260
Real-Time Probe	AcDx-7765-CaNCR38-RT-Pb	/56-FAM/CCTTTTTGC/ZEN/GTTTTTCGTTTATAGTCGGAGTTCGGTAGTTG/3IABkFQ/	41	3261
Tag Forward Primer	AcDx-7766-CaNCR38-RT-FP	TCGCGGAAAGTCCCAGTAAC	20	3262
Tag Reverse Primer	AcDx-7767-CaNCR38-RT-RP	TGGAACGCTTACAGGCCAAC	20	3263
Downstream PCR Primer	AcDx-7768-CaNCR38-PCR-V	TGGAACGCTTACAGGCCAACCCTAAAAACGCTCCCCGAAATAATGrCAACC/3SpC3/	50	3264

CaNCR39				
Forward PCR Primer	AcDx-7781-CaNCR39-FP	GATAAGAGGATGATTTTAAAGGGACrGTAGC/3SpC3/	30	3265
Reverse PCR Primer	AcDx-7782-CaNCR39-RP	GGTGTCGTGGCAACGCCATCGCGTAACrCAACC/3SpC3/	32	3266
Upstream LDR	AcDx-7783-CaNCR39-Up	TACACGTGGATATCTCCGACCGGGACGTAGTGGTTTCGTTTATCATCrGTTCC/3SpC3/	52	3267
Downstream LDR	AcDx-7784-CaNCR39-Dn	/5Phos/GTTTTTATAGGGAGGGAGTCGCGGTTTCGGGTGCTAGTCACACAGTTCCA	50	3268
Real-Time Probe	AcDx-7785-CaNCR39-RT-Pb	/56-FAM/AATATCATC/ZEN/GTTTTTATAGGGAGGGAGTCGCGG/3IABkFQ/	33	3269
Tag Forward Primer	AcDx-7786-CaNCR39-RT-FP	TACACGTGGATATCTCCGACC	21	3270
Tag Reverse Primer	AcDx-7787-CaNCR39-RT-RP	TGGAACTGTGTGACTAGCACC	21	3271
Downstream PCR Primer	AcDx-7788-CaNCR39-PCR-V	TGGAACTGTGTGACTAGCACCCGCCATCGCGTAACCAACTTGrAAACT/3SpC3/	47	3272
PTGER4				
Forward PCR Primer	AcDx-7791-PTGER4-FP	GTTTATTTTCGAGGTTAATTCGTTCrGTTTC/3SpC3/	30	3273
Reverse PCR Primer	AcDx-7792-PTGER4-RP	GGTGTCGTGGTTACCCACCACCCCGAArAATAG/3SpC3/	32	3274
Upstream LDR	AcDx-7793-PTGER4-Up	TAGCATTCGAGAACGCACCGAGGTTAATTCGTTCGTTTTTTTGAGTCTCrGATTA/3SpC3/	54	3275
Downstream LDR	AcDx-7794-PTGER4-Dn	/5Phos/GATCGGTTGAATAGTTTAGTGATTATTTCGGCGGTGGGTGCTAGTCACACAGTTCCA	57	3276
Real-Time Probe	AcDx-7795-PTGER4-RT-Pb	/56-FAM/AAGAGTCTC/ZEN/GATCGGTTGAATAGTTTAGTGATTATTTCGG/3IABkFQ/	40	3277
Tag Forward Primer	AcDx-7796-PTGER4-RT-FP	TAGCATTCGAGAACGCACC	19	3278
Tag Reverse Primer	AcDx-7797-PTGER4-RT-RP	TGGAACTGTGTGACTAGCACC	21	3279
Downstream PCR Primer	AcDx-7798-PTGER4-PCR-V	TGGAACTGTGTGACTAGCACCCCACCCCGAAAATAAACATCACTGrCCGAG/3SpC3/	50	3280

CaNCR40				
Forward PCR Primer	AcDx-7801-CaNCR40-FP	GTTTGTGTAGGGCGAGGACrGGGAT/3SpC3/	24	3281
Reverse PCR Primer	AcDx-7802-CaNCR40-RP	GGTGTCGTGGACGAATCTTCTACATCCGACArACAAC/3SpC3/	36	3282
Upstream LDR	AcDx-7803-CaNCR40-Up	TAACCGGGCCTAAAGTGACACGAGGCGGTTTTGCGCTCrGGTCG/3SpC3/	43	3283
Downstream LDR	AcDx-7804-CaNCR40-Dn	/5Phos/GGTTAGGTTGGTTTTCGAGGATTTAGTCGTTTTTAATTTTTTATGTTACGTGATCTCC CTCTCCA	65	3284
Real-Time Probe	AcDx-7805-CaNCR40-RT-Pb	/56-FAM/AATGCGCTC/ZEN/GGTTAGGTTGGTTTTCGAG/3IABkFQ/	28	3285
Tag Forward Primer	AcDx-7806-CaNCR40-RT-FP	TAACCGGGCCTAAAGTGACA	20	3286
Tag Reverse Primer	AcDx-7807-CaNCR40-RT-RP	TGGAGAGGGAGATCACGTAACA	22	3287
Downstream PCR Primer	AcDx-7808-CaNCR40-PCR-V	TGGAGAGGGAGATCACGTAACATCTACATCCGACAACAATTTAAAAAATTAAAAATGrACTA G/3SpC3/	62	3288
SHF				
Forward PCR Primer	AcDx-7811-SHF-FP	GGGTTTCGATTCGAATAAGGCrGGTGC/3SpC3/	26	3289
Reverse PCR Primer	AcDx-7812-SHF-RP	GGTGTCGTGGTCCGAAAAATCGTCCGACTCrCGCCA/3SpC3/	35	3290
Upstream LDR	AcDx-7813-SHF-Up	TCGATGGTCAATGAGCTTCACAGTGGAGGAGAGTCGTAGCrGCGAA/3SpC3/	45	3291
Downstream LDR	AcDx-7814-SHF-Dn	/5Phos/GCGGGAGCGGCGTGGTGTTACGTGATCTCCCTCTCCA	37	3292
Real-Time Probe	AcDx-7815-SHF-RT-Pb	/56-FAM/TTTCGTAGC/ZEN/GCGGGAGCGG/3IABkFQ/	19	3293
Tag Forward Primer	AcDx-7816-SHF-RT-FP	TCGATGGTCAATGAGCTTCACA	22	3294
Tag Reverse Primer	AcDx-7817-SHF-RT-RP	TGGAGAGGGAGATCACGTAACA	22	3295
Downstream PCR Primer	AcDx-7818-SHF-PCR-V	TGGAGAGGGAGATCACGTAACACACCCCTATCGCCCCTGrCACCT/3SpC3/	44	3296
CaNCR42				
Forward PCR Primer	AcDx-7881-CaNCR42-FP	GTCGTTATTCGGTCGTGATTATAATCrGAGGC/3SpC3/	31	3297
Reverse PCR Primer	AcDx-7882-CaNCR42-RP	GGTGTCGTGGACTTTCCTCTACGACTCAAATAAArAATTG/3SpC3/	39	3298
Upstream LDR	AcDx-7883-CaNCR42-Up	TTCGTCCCTGCACGCTAACTTTTGTTTGTAGAGTAATATATTAGGTTTAAATTTATCGTTCTCrG GTGA/3SpC3/	68	3299

Downstream LDR	AcDx-7884-CaNCR42-Dn	/5Phos/GGTAGTCGTTTTACGCGGGTTTTTTTTCGGTAGGTTCCATCACCGTTAGGCCA	53	3300
Real-Time Probe	AcDx-7885-CaNCR42-RT-Pb	/56-FAM/CCCGTTCTC/ZEN/GGTAGTCGTTTTACGCGGG/3IABkFQ/	28	3301
Tag Forward Primer	AcDx-7886-CaNCR42-RT-FP	TTCGTCCCTGCACGCTAAC	19	3302
Tag Reverse Primer	AcDx-7887-CaNCR42-RT-RP	TGGCCTAACGGTGATGGAAC	20	3303
Downstream PCR Primer	AcDx-7888-CaNCR42-PCR-V	TGGCCTAACGGTGATGGAACCCTCTACGACTCAAATAAAAATTAAAAAATCTACTGrAAAG/3SpC3/	60	3304
HLF				
Forward PCR Primer	AcDx-7891-HLF-FP	GATTTAGTTTGAGGTTATAAAGGTTTTTACrGTTTC/3SpC3/	35	3305
Reverse PCR Primer	AcDx-7892-HLF-RP	GGTGTCGTGGCCTTACTAAAACCCTAATCTTCGArUCTAG/3SpC3/	39	3306
Upstream LDR	AcDx-7893-HLF-Up	TGATGCTGGCAAACCCTAGAACGAGGTTATAAAGGTTTTTACGTTTTATTGAAAGACrGGTCC/3SpC3/	62	3307
Downstream LDR	AcDx-7894-HLF-Dn	/5Phos/GGTTTTTTTTTATTTGTGTTTATTCGTTTTTGTAGTTTTTTTTTTAGGTTCCATCACCGTTAGGCCA	67	3308
Real-Time Probe	AcDx-7895-HLF-RT-Pb	/56-FAM/AAGAAAGAC/ZEN/GGTTTTTTTTTATTTGTGTTTATTCGTTTTTGTAG/3IABkFQ/	44	3309
Tag Forward Primer	AcDx-7896-HLF-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	3310
Tag Reverse Primer	AcDx-7897-HLF-RT-RP	TGGCCTAACGGTGATGGAAC	20	3311
GLI3-S1				
Forward PCR Primer	AcDx-7921-GLI3-S1-FP	GTCGGGTTCGCGTAGCrGTTGC/3SpC3/	21	3312
Reverse PCR Primer	AcDx-7922-GLI3-S1-RP	GGTGTCGTGGCGCGCCGAAACCGAArAAAAT/3SpC3/	30	3313
Upstream LDR	AcDx-7923-GLI3-S1-Up	TCCGGGTATACACTGTCCCACGTTGTTTGTAGTCGCGTCATCrGTACC/3SpC3/	47	3314
Downstream LDR	AcDx-7924-GLI3-S1-Dn	/5Phos/GTATTTATTATTATTATATTAGCGCGAGGAAGTTTACGTCGTGGTTAACAGAGGACAGGCCA	62	3315
Real-Time Probe	AcDx-7925-GLI3-S1-RT-Pb	/56-FAM/TTCGTCATC/ZEN/GTATTTATTATTATTATATTAGCGCGAGGAAGTT/3IABkFQ/	43	3316
Tag Forward Primer	AcDx-7926-GLI3-S1-RT-	TCCGGGTATACACTGTCCCA	20	3317

	FP			
Tag Reverse Primer	AcDx-7927-GLI3-S1-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	3318
Downstream PCR Primer	AcDx-7928-GLI3-S1-PCR-V	TGGCCTGTCCTCTGTTAACCAGCCGAAACCGAAAAAACTCGATGrUAAAT/3SpC3/	49	3319
CaNCR45				
Forward PCR Primer	AcDx-8061-CaNCR45-FP	TTTGGGTTAACGGAGGCrGAGGA/3SpC3/	22	3320
Reverse PCR Primer	AcDx-8062-CaNCR45-RP	GGTGTCGTGGGCGCCAAAAAACCGCCrCAACG/3SpC3/	31	3321
Upstream LDR	AcDx-8063-CaNCR45-Up	TATAGTCACGCAGGACCACAGGAGCGAGGGAGTTGCrGTAAG/3SpC3/	41	3322
Downstream LDR	AcDx-8064-CaNCR45-Dn	/5Phos/GTAGAGGGTAGGTTTCGATTTTTTTTCGTCGTATTTTGTGGTGTTTGCGGCTGTCTATGACA	62	3323
Real-Time Probe	AcDx-8065-CaNCR45-RT-Pb	/56-FAM/TTGAGTTGC/ZEN/GTAGAGGGTAGGTTTCGATTTTTTTTCG/3IABkFQ/	37	3324
Tag Forward Primer	AcDx-8066-CaNCR45-RT-FP	TATAGTCACGCAGGACCACA	20	3325
Tag Reverse Primer	AcDx-8067-CaNCR45-RT-RP	TGTCATAGACAGCCGCAAACA	21	3326
Downstream PCR Primer	AcDx-8068-CaNCR45-PCR-V	TGTCATAGACAGCCGCAAACAAACACCCGACCCACAAAATATGrACGAG/3SpC3/	48	3327
TLR5				
Forward PCR Primer	AcDx-8071-TLR5-FP	TCGGTTATTCGTGCGGTCrGGATG/3SpC3/	23	3328
Reverse PCR Primer	AcDx-8072-TLR5-RP	GGTGTCGTGGTTCCTCTCCCAATAACGCTACrUAAAC/3SpC3/	36	3329
Upstream LDR	AcDx-8073-TLR5-Up	TGCCCTATCGAAAAGGACAACAGGTCGGATATCGTGTTAGGTTTGCrGAGAG/3SpC3/	51	3330
Downstream LDR	AcDx-8074-TLR5-Dn	/5Phos/GAGGAGGGTTTTGTCGTAGTTTCGGAGAATAGTGTTTGCGGCTGTCTATGACA	53	3331
Real-Time Probe	AcDx-8075-TLR5-RT-Pb	/56-FAM/ATGGTTTGC/ZEN/GAGGAGGGTTTTGTCGTAGTTT/3IABkFQ/	31	3332
Tag Forward Primer	AcDx-8076-TLR5-RT-FP	TGCCCTATCGAAAAGGACAACA	22	3333
Tag Reverse Primer	AcDx-8077-TLR5-RT-RP	TGTCATAGACAGCCGCAAACA	21	3334
Downstream PCR Primer	AcDx-8078-TLR5-PCR-V	TGTCATAGACAGCCGCAAACACCCAATAACGCTACTAAATACTATTCCCTGrAAACC/3SpC3/	56	3335

CaNCR46				
Forward PCR Primer	AcDx-8081-CaNCR46-FP	GGGTTCGGGAAGTTCGCrGGAAG/3SpC3/	22	3336
Reverse PCR Primer	AcDx-8082-CaNCR46-RP	GGTGTCGTGGACCTAAACTAAAACAAAACTCCGArAAATG/3SpC3/	39	3337
Upstream LDR	AcDx-8083-CaNCR46-Up	TAATCTCCAGACCTCCGAACCGTTCGCGGAAACGTAGGAAGCrGGTCG/3SpC3/	47	3338
Downstream LDR	AcDx-8084-CaNCR46-Dn	/5Phos/GGTTAGGAGAGGTAGCGTTACGTATATTTTTTTTATTTGGTGTAAGGATTGAACGGGACA	60	3339
Real-Time Probe	AcDx-8085-CaNCR46-RT-Pb	/56-FAM/AAAGGAAGC/ZEN/GGTTAGGAGAGGTAGCGTTAC/3IABkFQ/	30	3340
Tag Forward Primer	AcDx-8086-CaNCR46-RT-FP	TAATCTCCAGACCTCCGAACC	21	3341
Tag Reverse Primer	AcDx-8087-CaNCR46-RT-RP	TGTCCCGTTCAATCCTTACATC	22	3342
CaNCR47				
Forward PCR Primer	AcDx-8091-CaNCR47-FP	TTTCGTTTTTGTCGGCGGTAGCrGATTT/3SpC3/	27	3343
Reverse PCR Primer	AcDx-8092-CaNCR47-RP	GGTGTCGTGGACTCAATCCGCGCGCrCCAAT/3SpC3/	30	3344
Upstream LDR	AcDx-8093-CaNCR47-Up	TTTTCCGCGTCAGAGCACAGGCGGTAGCGATTCGGATTCTCrGTTCC/3SpC3/	46	3345
Downstream LDR	AcDx-8094-CaNCR47-Dn	/5Phos/GTTTTCGATAAAGTTTTAGTTTCGTAGTAGTATTCGGCGCGTGTTATCGGACCTAGCTCGACA	63	3346
Real-Time Probe	AcDx-8095-CaNCR47-RT-Pb	/56-FAM/TTGATTCTC/ZEN/GTTTTCGATAAAGTTTTAGTTTCGTAGTAGTATTCGG/3IABkFQ/	46	3347
Tag Forward Primer	AcDx-8096-CaNCR47-RT-FP	TTTTCCGCGTCAGAGCACA	19	3348
Tag Reverse Primer	AcDx-8097-CaNCR47-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	3349
Downstream PCR Primer	AcDx-8098-CaNCR47-PCR-V	TGTCGAGCTAGGTCCGATAACACGCCCAACGAATCCGTGrCCGAG/3SpC3/	44	3350

CaNCR48				
Forward PCR Primer	AcDx-8101-CaNCR48-FP	GAGTATAGAGTATAGTAAATCGGGATTTTCrGGCGA/3SpC3/	35	3351
Reverse PCR Primer	AcDx-8102-CaNCR48-RP	GGTGTCGTGGGACCGCCCAACCCGArCGCGG/3SpC3/	30	3352
Upstream LDR	AcDx-8103-CaNCR48-Up	TTCGCTGCCCGGTTAAACAGGATTTTCGGCGGACAGCrGTTCC/3SpC3/	42	3353
Downstream LDR	AcDx-8104-CaNCR48-Dn	/5Phos/GTTTTTCGTTCGTTTGTTCGTTATGTTGGAGAGTGTTATCGGACCTAGCTCGACA	55	3354
Real-Time Probe	AcDx-8105-CaNCR48-RT-Pb	/56-FAM/TTGGACAGC/ZEN/GTTTTTCGTTCGTTTGTTCGTTATG/3IABkFQ/	34	3355
Tag Forward Primer	AcDx-8106-CaNCR48-RT-FP	TTCGCTGCCCGGTTAAACA	19	3356
Tag Reverse Primer	AcDx-8107-CaNCR48-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	3357
Downstream PCR Primer	AcDx-8108-CaNCR48-PCR-V	TGTCGAGCTAGGTCCGATAACAGCCCAACCCGACGTGrAAAAG/3SpC3/	42	3358
CaNCR49				
Forward PCR Primer	AcDx-8111-CaNCR49-FP	CGACGTGTTTCGTTGAAAGCrGGGTG/3SpC3/	25	3359
Reverse PCR Primer	AcDx-8112-CaNCR49-RP	GGTGTCGTGGCGCCGAAACCCGACAArACCGG/3SpC3/	31	3360
Upstream LDR	AcDx-8113-CaNCR49-Up	TTCGCTGCCCGGTTAAACATGAAAGCGGGTAGGTGGTATCrGGGTA/3SpC3/	45	3361
Downstream LDR	AcDx-8114-CaNCR49-Dn	/5Phos/GGGCGGAGTTGCGTTGAGGTGTTATCGGACCTAGCTCGACA	41	3362
Real-Time Probe	AcDx-8115-CaNCR49-RT-Pb	/56-FAM/TTTGGTATC/ZEN/GGGCGGAGTTGCG/3IABkFQ/	22	3363
Tag Forward Primer	AcDx-8116-CaNCR49-RT-FP	TTCGCTGCCCGGTTAAACA	19	3364
Tag Reverse Primer	AcDx-8117-CaNCR49-RT-RP	TGTCGAGCTAGGTCCGATAACA	22	3365
Downstream PCR Primer	AcDx-8118-CaNCR49-PCR-V	TGTCGAGCTAGGTCCGATAACAGAAACCCGACAAACCGACAAATATGrAAAAG/3SpC3/	52	3366
RNF220-S1				
Forward PCR Primer	AcDx-8141-RNF220-S1-FP	GTAGAAGTGATTCGGGTTGTCrGTTTC/3SpC3/	26	3367

Reverse PCR Primer	AcDx-8142-RNF220-S1-RP	GGTGTCGTGGCGCCCCCTCCCCCAArAAAAC/3SpC3/	30	3368
Upstream LDR	AcDx-8143-RNF220-S1-Up	TCCTGCTCTGAAAACCTACACCGTGATTCGGGTTGTCGTTTTTTTGCrGGTTA/3SpC3/	52	3369
Downstream LDR	AcDx-8144-RNF220-S1-Dn	/5Phos/GGTCGTTCGTTAGTTTTTCGTCGTTGTCGGTAGGGTTACATAGGCGGCTTAGACA	55	3370
Real-Time Probe	AcDx-8145-RNF220-S1-RT-Pb	/56-FAM/AATTTTTGC/ZEN/GGTCGTTCGTTAGTTTTTCGTCG/3IABkFQ/	32	3371
Tag Forward Primer	AcDx-8146-RNF220-S1-RT-FP	TCCTGCTCTGAAAACCTACACC	22	3372
Tag Reverse Primer	AcDx-8147-RNF220-S1-RT-RP	TGTCTAAGCCGCCTATGTAACC	22	3373
Downstream PCR Primer	AcDx-8148-RNF220-S1-PCR-V	TGTCTAAGCCGCCTATGTAACCAATCAATTACCAACCACCCTTCTACTGrACAAT/3SpC3/	54	3374
PRKCB				
Forward PCR Primer	AcDx-8191-PRKCB-FP	TTAAGCGTAGTTGGACGAGCrGGTAA/3SpC3/	25	3375
Reverse PCR Primer	AcDx-8192-PRKCB-RP	GGTGTCGTGGTCCCCTACGCCGACTCrUAACA/3SpC3/	31	3376
Upstream LDR	AcDx-8193-PRKCB-Up	TTCGCCTACCGCAGTGAACACGAGCGGTAGTAGTTGAGCrGAGCA/3SpC3/	44	3377
Downstream LDR	AcDx-8194-PRKCB-Dn	/5Phos/GAGTGATAGTTTCGGTTTCGCGCGTCGGTTGAGACATGGGCTCGCA	46	3378
Real-Time Probe	AcDx-8195-PRKCB-RT-Pb	/56-FAM/ATGTTGAGC/ZEN/GAGTGATAGTTTCGGTTTCGC/3IABkFQ/	30	3379
Tag Forward Primer	AcDx-8196-PRKCB-RT-FP	TTCGCCTACCGCAGTGAAC	19	3380
Tag Reverse Primer	AcDx-8197-PRKCB-RT-RP	TGCGAGCCCATGTCTCAAC	19	3381
Downstream PCR Primer	AcDx-8198-PRKCB-PCR-V	TGCGAGCCCATGTCTCAACCGCCGACTCTAACGACTGrCGACA/3SpC3/	42	3382
CaNCR53				
Forward PCR Primer	AcDx-8231-CaNCR53-FP	TTTTTAGTCGTGTTCGGTTTTCrGTCGC/3SpC3/	27	3383
Reverse PCR Primer	AcDx-8232-CaNCR53-RP	GGTGTCGTGGCCGCTCTCCCCGATCrUACCT/3SpC3/	30	3384
Upstream LDR	AcDx-8233-CaNCR53-Up	TCCTCGAGCCGATGACACACGTGTTCGGTTTTCGTCGTTTTTTTATTTCACrGTGAC/3SpC3/	56	3385
Downstream LDR	AcDx-8234-CaNCR53-Dn	/5Phos/GTGGTGGAATTTTTCGCGTTTTTTATAGTCGTCGTGTAACGTCCGTGGGCTAA	53	3386

Real-Time Probe	AcDx-8235-CaNCR53-RT-Pb	/56-FAM/CCATTTCAC/ZEN/GTGGTGGAATTTTTCGCGTTTTT/3IABkFQ/	32	3387
Tag Forward Primer	AcDx-8236-CaNCR53-RT-FP	TCCTCGAGCCGATGACACA	19	3388
Tag Reverse Primer	AcDx-8237-CaNCR53-RT-RP	TTAGCCCACGGACGTTACA	19	3389
Downstream PCR Primer	AcDx-8238-CaNCR53-PCR-V	TTAGCCCACGGACGTTACACCCCGATCTACCCATTAATTCGATGrACTAC/3SpC3/	49	3390
CaNCR54				
Forward PCR Primer	AcDx-8241-CaNCR54-FP	GAAATTTTTTTGCGTTATTAGATTGCrGTTTG/3SpC3/	31	3391
Reverse PCR Primer	AcDx-8242-CaNCR54-RP	GGTGTCGTGGAAAAACTTATAAATACTTTCCGTCGAArCTCAG/3SpC3/	42	3392
Upstream LDR	AcDx-8243-CaNCR54-Up	TACTATCGTATCACGCCGACAGTTTAGTATAGTAGATGGTACGAGTACGCGCrGGCAC/3SpC3/	57	3393
Downstream LDR	AcDx-8244-CaNCR54-Dn	/5Phos/GGCGTTTAGTTTAGGTTTTCGGAGGTAGTTGAGTGTAACGTCCGTGGGCTAA	52	3394
Real-Time Probe	AcDx-8245-CaNCR54-RT-Pb	/56-FAM/TTTACGCGC/ZEN/GGCGTTTAGTTTAGGTTTT/3IABkFQ/	28	3395
Tag Forward Primer	AcDx-8246-CaNCR54-RT-FP	TACTATCGTATCACGCCGACA	21	3396
Tag Reverse Primer	AcDx-8247-CaNCR54-RT-RP	TTAGCCCACGGACGTTACA	19	3397
Downstream PCR Primer	AcDx-8248-CaNCR54-PCR-V	TTAGCCCACGGACGTTACAGTCGAAATTTCCAACTCAACTACCTCTGrAAAAT/3SpC3/	52	3398
ISL2-S2				
Forward PCR Primer	AcDx-9131-ISL2-S2-FP	GGTTTGTTTTTAGTTGGCGGTCrGGTTC/3SpC3/	27	3399
Reverse PCR Primer	AcDx-9132-ISL2-S2-RP	GGTGTCGTGGAACGACGAAAACACCGAAArAAAAG/3SpC3/	34	3400
Upstream LDR	AcDx-9133-ISL2-S2-Up	TCGTAGACTCGCTATCGCCAGGCGGTCGGTTTTTAAGGGATATTTCTCrGATCT/3SpC3/	53	3401
Downstream LDR	AcDx-9134-ISL2-S2-Dn	/5Phos/GATTCGGAGTACGCGGTTTTGGAGTATTAGTTCTGGTGAGCAGGGATGAGCA	52	3402
Real-Time Probe	AcDx-9135-ISL2-S2-RT-Pb	/56-FAM/CCATTTCTC/ZEN/GATTCGGAGTACGCGGTTTTGG/3IABkFQ/	31	3403
Tag Forward Primer	AcDx-9136-ISL2-S2-RT-FP	TCGTAGACTCGCTATCGCCA	20	3404

Tag Reverse Primer	AcDx-9137-ISL2-S2-RT-RP	TGCTCATCCCTGCTCACCA	19	3405
Downstream PCR Primer	AcDx-9138-ISL2-S2-PCR-V	TGCTCATCCCTGCTCACCACACCGAAAAAAAACAAATAAAAAAACACGTGrAACTG/3SpC3/	55	3406
TRPV3				
Forward PCR Primer	AcDx-9201-TRPV3-FP	GTTTTTTTGGCGTGCGCrGTTGG/3SpC3/	22	3407
Reverse PCR Primer	AcDx-9202-TRPV3-RP	GGTGTCGTGGCGCCATCGAACGACGACrAAAAG/3SpC3/	32	3408
Upstream LDR	AcDx-9203-TRPV3-Up	TAGCATTCGAGAACGCACCGCGTGCGCGTTGACATCrGGCAC/3SpC3/	41	3409
Downstream LDR	AcDx-9204-TRPV3-Dn	/5Phos/GGCGTCGGCGGCGATGAGTAGGGTGCTAGTCACACAGTTCCA	42	3410
Real-Time Probe	AcDx-9205-TRPV3-RT-Pb	/56-FAM/AATGACATC/ZEN/GGCGTCGGCGG/3IABkFQ/	20	3411
Tag Forward Primer	AcDx-9206-TRPV3-RT-FP	TAGCATTCGAGAACGCACC	19	3412
Tag Reverse Primer	AcDx-9207-TRPV3-RT-RP	TGGAACTGTGTGACTAGCACC	21	3413
Downstream PCR Primer	AcDx-9208-TRPV3-PCR-V	TGGAACTGTGTGACTAGCACCCAAAAAAACATCGCAACCCTACTCATTGrCCGCT/3SpC3/	54	3414
NXPH4				
Forward PCR Primer	AcDx-9831-NXPH4-FP	GCGCGCGTTAAAAAGATTTTCrGGTTA/3SpC3/	26	3415
Reverse PCR Primer	AcDx-9832-NXPH4-RP	GGTGTCGTGGACACACTAAAAATACCGTTCACATAArUCCAT/3SpC3/	41	3416
Upstream LDR	AcDx-9833-NXPH4-Up	TTGCAAACCACCCGGACAAGATTTTCGGTTGGAGGGATTTTTATTCTCrGGGCA/3SpC3/	53	3417
Downstream LDR	AcDx-9834-NXPH4-Dn	/5Phos/GGGTGTATATTTTTAAGTTTTCGTTGTTGGTGATCGGTAAGATCTTGGTCAGCATCGACTCCTA	64	3418
Real-Time Probe	AcDx-9835-NXPH4-RT-Pb	/56-FAM/AATATTCTC/ZEN/GGGTGTATATTTTTAAGTTTTCGTTGTTGGTG/3IABkFQ/	41	3419
Tag Forward Primer	AcDx-9836-NXPH4-RT-FP	TTGCAAACCACCCGGACAA	19	3420
Tag Reverse Primer	AcDx-9837-NXPH4-RT-RP	TAGGAGTCGATGCTGACCAA	20	3421
Downstream PCR Primer	AcDx-9838-NXPH4-PCR-V	TAGGAGTCGATGCTGACCAACCGTTCACATAATCCACGATCTTACTGrATCAT/3SpC3/	52	3422
CANCR78				
Forward PCR Primer	AcDx-9841-CANCR78-FP	TTTTCGATTGAGGTTTGTATATATAGTCrGTTTC/3SpC3/	33	3423

Reverse PCR Primer	AcDx-9842-CANCR78-RP	GGTGTCGTGGGCCTCCGCAACCGATAAArACAAT/3SpC3/	33	3424
Upstream LDR	AcDx-9843-CANCR78-Up	TTGCAAACCACCCGGACAATAGTCGTTTTTTTTTTCGTTATCGTCAGCrGGTGC/3SpC3/	53	3425
Downstream LDR	AcDx-9844-CANCR78-Dn	/5Phos/GGTATATATTCGGTTGGGTTTTCGTTTTGTAAGGGAAGTTGGTCAGCATCGACTCCTA	58	3426
Real-Time Probe	AcDx-9845-CANCR78-RT-Pb	/56-FAM/CCCGTCAGC/ZEN/GGTATATATTCGGTTGGGTTTTCGTTTTG/3IABkFQ/	38	3427
Tag Forward Primer	AcDx-9846-CANCR78-RT-FP	TTGCAAACCACCCGGACAA	19	3428
Tag Reverse Primer	AcDx-9847-CANCR78-RT-RP	TAGGAGTCGATGCTGACCAA	20	3429
TGFB1I1				
Forward PCR Primer	AcDx-9851-TGFB1I1-FP	GAGGGTCGTTTTTATTGTCGTCrGGGAC/3SpC3/	27	3430
Reverse PCR Primer	AcDx-9852-TGFB1I1-RP	GGTGTCGTGGCCAAAACGCGCTAAACGCrCGAAG/3SpC3/	33	3431
Upstream LDR	AcDx-9853-TGFB1I1-Up	TAAGACGTATGCTAGCGCCAAATTGTCGTCGGGATTTTTTGTAGTTGCTCrGTTCT/3SpC3/	55	3432
Downstream LDR	AcDx-9854-TGFB1I1-Dn	/5Phos/GTTTCGCGTTGTTAGGGTTGTTAGGGTTTTATTTTGTTGGTCAGCATCGACTCCTA	56	3433
Real-Time Probe	AcDx-9855-TGFB1I1-RT-Pb	/56-FAM/CCGTTGCTC/ZEN/GTTTCGCGTTGTTAGGGTTGTTAG/3IABkFQ/	33	3434
Tag Forward Primer	AcDx-9856-TGFB1I1-RT-FP	TAAGACGTATGCTAGCGCCAA	21	3435
Tag Reverse Primer	AcDx-9857-TGFB1I1-RT-RP	TAGGAGTCGATGCTGACCAA	20	3436
Downstream PCR Primer	AcDx-9858-TGFB1I1-PCR-V	TAGGAGTCGATGCTGACCAACCAAAACGCGCTAAACGCTGrAAATG/3SpC3/	45	3437
TBX1-S2				
Forward PCR Primer	AcDx-9861-TBX1-S2-FP	GTTCGGTTTGCGTGGTTACrGGTTG/3SpC3/	24	3438
Reverse PCR Primer	AcDx-9862-TBX1-S2-RP	GGTGTCGTGGCTACGACAACGACGACGArCGACT/3SpC3/	33	3439
Upstream LDR	AcDx-9863-TBX1-S2-Up	TACATGCCATCCCACGACACGTGGTTACGGTTATTATTCGTATGCrGTACC/3SpC3/	50	3440
Downstream LDR	AcDx-9864-TBX1-S2-Dn	/5Phos/GTATTCGTATTATTATTATTATTTCGTGAGTTTAGTCGTCGCGGTCTGTGTCGGAGC	65	3441

		GGTTACTA		
Real-Time Probe	AcDx-9865-TBX1-S2-RT-Pb	/56-FAM/AACGTATGC/ZEN/GTATTCGTATTATTATTATTATTTCGTGAGTTTAGTCGTC/3IABkFQ/	49	3442
Tag Forward Primer	AcDx-9866-TBX1-S2-RT-FP	TACATGCCATCCCACGACA	19	3443
Tag Reverse Primer	AcDx-9867-TBX1-S2-RT-RP	TAGTAACCGCTCCGACACA	19	3444
Downstream PCR Primer	AcDx-9868-TBX1-S2-PCR-V	TAGTAACCGCTCCGACACAGACAACGACGACGACGACTGrCGACA/3SpC3/	44	3445
CANCR79				
Forward PCR Primer	AcDx-9871-CANCR79-FP	GTTCGGTTTTTGGAGGATAGCrGGGAC/3SpC3/	26	3446
Reverse PCR Primer	AcDx-9872-CANCR79-RP	GGTGTCGTGGCACAATACACTTCCCGACAACArUAAAG/3SpC3/	37	3447
Upstream LDR	AcDx-9873-CANCR79-Up	TCCAAACAAGCTGATCCGTACAAGGATAGCGGGATGGGAGAGTCTCrGTTCG/3SpC3/	51	3448
Downstream LDR	AcDx-9874-CANCR79-Dn	/5Phos/GTTTATATTTAGTTTCGTAGAGTCGGATAGAAACGTTGTTATTGTCGTGTGTCGGAGCGGTTACTA	66	3449
Real-Time Probe	AcDx-9875-CANCR79-RT-Pb	/56-FAM/TTGAGTCTC/ZEN/GTTTATATTTAGTTTCGTAGAGTCGGATAGAAACG/3IABkFQ/	44	3450
Tag Forward Primer	AcDx-9876-CANCR79-RT-FP	TCCAAACAAGCTGATCCGTACA	22	3451
Tag Reverse Primer	AcDx-9877-CANCR79-RT-RP	TAGTAACCGCTCCGACACA	19	3452
Downstream PCR Primer	AcDx-9878-CANCR79-PCR-V	TAGTAACCGCTCCGACACACACAATACACTTCCCGACAACATAAAATTGrACAAC/3SpC3/	54	3453
CANCR80				
Forward PCR Primer	AcDx-9881-CANCR80-FP	GGGAATAAAACGCGTTGGTAAACrGCGGT/3SpC3/	28	3454
Reverse PCR Primer	AcDx-9882-CANCR80-RP	GGTGTCGTGGCCGATTACCTCTCGATATCGACrCCCCA/3SpC3/	37	3455
Upstream LDR	AcDx-9883-CANCR80-Up	TAGGGCGACAGTTACCACAAGGCGACGTATAGGGAGTGCrGAGTA/3SpC3/	44	3456
Downstream LDR	AcDx-9884-CANCR80-Dn	/5Phos/GAGCGAGGGAGGGAGCGTCGTTGTGGGTCTCGCTCGTATA	40	3457

Real-Time Probe	AcDx-9885-CANCR80-RT-Pb	/56-FAM/TTGGAGTGC/ZEN/GAGCGAGGGAGGG/3IABkFQ/	22	3458
Tag Forward Primer	AcDx-9886-CANCR80-RT-FP	TAGGGCGACAGTTACCACAA	20	3459
Tag Reverse Primer	AcDx-9887-CANCR80-RT-RP	TATACGAGCGAGACCCACAA	20	3460
Downstream PCR Primer	AcDx-9888-CANCR80-PCR-V	TATACGAGCGAGACCCACAAGATTACCTCTCGATATCGACCCCTGrCGCTT/3SpC3/	50	3461
CANCR81				
Forward PCR Primer	AcDx-9891-CANCR81-FP	GTTATTCGTTAGGTGTCGGCrGTTTA/3SpC3/	25	3462
Reverse PCR Primer	AcDx-9892-CANCR81-RP	GGTGTCGTGGAAACATTACTACCACCGAAAAACrUATCT/3SpC3/	38	3463
Upstream LDR	AcDx-9893-CANCR81-Up	TCCGACTTTAGTGCGTCACAACGGCGTTTGTTGAGTTTTAGCATCrGTTGA/3SpC3/	50	3464
Downstream LDR	AcDx-9894-CANCR81-Dn	/5Phos/GTTAGATTTTTGTTAGCGAGGTTTAGAGTAGTGTAGAGATTTAGCGTTGTGGGTCTCGCTCGTATA	66	3465
Real-Time Probe	AcDx-9895-CANCR81-RT-Pb	/56-FAM/AATAGCATC/ZEN/GTTAGATTTTTGTTAGCGAGGTTTAGAGTAGTG/3IABkFQ/	42	3466
Tag Forward Primer	AcDx-9896-CANCR81-RT-FP	TCCGACTTTAGTGCGTCACAA	21	3467
Tag Reverse Primer	AcDx-9897-CANCR81-RT-RP	TATACGAGCGAGACCCACAA	20	3468
Downstream PCR Primer	AcDx-9898-CANCR81-PCR-V	TATACGAGCGAGACCCACAAAAACATTACTACCACCGAAAAACTATCTGrCTAAG/3SpC3/	54	3469
IRF8				
Forward PCR Primer	AcDx-9901-IRF8-FP	GGGCGGGAAAGTGATTTTTCrGGAAG/3SpC3/	25	3470
Reverse PCR Primer	AcDx-9902-IRF8-RP	GGTGTCGTGGCGATACGCAAAACGACGACrCACAT/3SpC3/	34	3471
Upstream LDR	AcDx-9903-IRF8-Up	TCAAACAAAGGCGACCACAACGTAGAGTATTTCGAAGAAGGCGGATCrGCGTA/3SpC3/	52	3472
Downstream LDR	AcDx-9904-IRF8-Dn	/5Phos/GCGCGAGTTAAGTTGACGTTATTGGTCGGGTTGTCGCATAGGCAGTTCATA	51	3473
Real-Time Probe	AcDx-9905-IRF8-RT-Pb	/56-FAM/TTGCGGATC/ZEN/GCGCGAGTTAAGTTGACG/3IABkFQ/	27	3474
Tag Forward Primer	AcDx-9906-IRF8-RT-FP	TCAAACAAAGGCGACCACAAC	21	3475

Tag Reverse Primer	AcDx-9907-IRF8-RT-RP	TATGAACTGCCTATGCGACAAC	22	3476
Downstream PCR Primer	AcDx-9908-IRF8-PCR-V	TATGAACTGCCTATGCGACAACGCAAAACGACGACCACACTGrACCAG/3SpC3/	47	3477
CANCR82				
Forward PCR Primer	AcDx-9911-CANCR82-FP	GTTCGGTGGGTGGGCrGTATA/3SpC3/	20	3478
Reverse PCR Primer	AcDx-9912-CANCR82-RP	GGTGTCGTGGATATAATATATAATAAAATAAAATTAAACCTCCGArAAAAG/3SpC3/	50	3479
Upstream LDR	AcDx-9913-CANCR82-Up	TAAACAATGAGACCCGCTGAACGGGCGTATGCGTATAGGATTTTCATCrGTTGA/3SpC3/	53	3480
Downstream LDR	AcDx-9914-CANCR82-Dn	/5Phos/GTTAGGTATTTGCGGAATATATTTGGAAATCGTTTTTTGTTGTCGCATAGGCAGTTCATA	60	3481
Real-Time Probe	AcDx-9915-CANCR82-RT-Pb	/56-FAM/CCTTTCATC/ZEN/GTTAGGTATTTGCGGAATATATTTGGAAATCG/3IABkFQ/	41	3482
Tag Forward Primer	AcDx-9916-CANCR82-RT-FP	TAAACAATGAGACCCGCTGAAC	22	3483
Tag Reverse Primer	AcDx-9917-CANCR82-RT-RP	TATGAACTGCCTATGCGACAAC	22	3484
CANCR83				
Forward PCR Primer	AcDx-9921-CANCR83-FP	TTTTTTTGTAGACGAGGAGGCrGTAGA/3SpC3/	26	3485
Reverse PCR Primer	AcDx-9922-CANCR83-RP	GGTGTCGTGGCCGCCGACCCAACTAACrUCAAG/3SpC3/	32	3486
Upstream LDR	AcDx-9923-CANCR83-Up	TCTGCCAGAACACCGACACCGAGGAGGCGTAGGAGCrGAGCA/3SpC3/	41	3487
Downstream LDR	AcDx-9924-CANCR83-Dn	/5Phos/GAGTGGAGAGGGCGTTTTCGGTGGTGTGTTGGCGTACGGTGA	42	3488
Real-Time Probe	AcDx-9925-CANCR83-RT-Pb	/56-FAM/TTTAGGAGC/ZEN/GAGTGGAGAGGGC/3IABkFQ/	22	3489
Tag Forward Primer	AcDx-9926-CANCR83-RT-FP	TCTGCCAGAACACCGACAC	19	3490
Tag Reverse Primer	AcDx-9927-CANCR83-RT-RP	TCACCGTACGCCAACACAC	19	3491
Downstream PCR Primer	AcDx-9928-CANCR83-PCR-V	TCACCGTACGCCAACACACCGACCCAACTAACTCAAAAAAATATTCGTGrCGCTT/3SpC3/	54	3492

CANCR84				
Forward PCR Primer	AcDx-9931-CANCR84-FP	AGTTTGTGTCGTGGAGTAGCrGGCGA/3SpC3/	25	3493
Reverse PCR Primer	AcDx-9932-CANCR84-RP	GGTGTCGTGGCCGCTATCCCCCGATACrCTCCG/3SpC3/	32	3494
Upstream LDR	AcDx-9933-CANCR84-Up	TCCGGATCAAAGCAGCCACGTGGAGTAGCGGCGGGCACrGGGAA/3SpC3/	43	3495
Downstream LDR	AcDx-9934-CANCR84-Dn	/5Phos/GGGAGATATTGGGAGGAGTAGCGGGTGTGTTGGCGTACGGTGA	43	3496
Real-Time Probe	AcDx-9935-CANCR84-RT-Pb	/56-FAM/TTCGGGCAC/ZEN/GGGAGATATTGGG/3IABkFQ/	22	3497
Tag Forward Primer	AcDx-9936-CANCR84-RT-FP	TCCGGATCAAAGCAGCCAC	19	3498
Tag Reverse Primer	AcDx-9937-CANCR84-RT-RP	TCACCGTACGCCAACACAC	19	3499
Downstream PCR Primer	AcDx-9938-CANCR84-PCR-V	TCACCGTACGCCAACACACCGATACCTCCAAATACCCGCTGrCTACC/3SpC3/	46	3500
NR5A2-S2				
Forward PCR Primer	AcDx-9941-NR5A2-S2-FP	CGTAGATTTGCGGGTGTTCrGTTTG/3SpC3/	24	3501
Reverse PCR Primer	AcDx-9942-NR5A2-S2-RP	GGTGTCGTGGCCCCAAAAAATACCTAATCCGArUATTC/3SpC3/	37	3502
Upstream LDR	AcDx-9943-NR5A2-S2-Up	TTCTAGATACCACGGACGCACCGGGTGTTCGTTTATTCGCGTCTCrGCGCC/3SpC3/	50	3503
Downstream LDR	AcDx-9944-NR5A2-S2-Dn	/5Phos/GCGTTTTTATATTTGATTTGTGTGGATGTGGAGTTTTCGGGTGTTGGTGTGCAAAGCTGA	60	3504
Real-Time Probe	AcDx-9945-NR5A2-S2-RT-Pb	/56-FAM/AAGCGTCTC/ZEN/GCGTTTTTATATTTGATTTGTGTGGATGT/3IABkFQ/	38	3505
Tag Forward Primer	AcDx-9946-NR5A2-S2-RT-FP	TTCTAGATACCACGGACGCAC	21	3506
Tag Reverse Primer	AcDx-9947-NR5A2-S2-RT-RP	TCAGCTTTGCACACCAACAC	20	3507
Downstream PCR Primer	AcDx-9948-NR5A2-S2-PCR-V	TCAGCTTTGCACACCAACACCCCAAAAAATACCTAATCCGATATTTCCTGrAAAAT/3SpC3/	55	3508

PROM1-S1				
Forward PCR Primer	AcDx-9951-PROM1-S1-FP	GGATTTAGGTATGAGGTCGTCrGTAAC/3SpC3/	26	3509
Reverse PCR Primer	AcDx-9952-PROM1-S1-RP	GGTGTCGTGGAACTCGAATTTCGCGATCTTTAAArUAACC/3SpC3/	39	3510
Upstream LDR	AcDx-9953-PROM1-S1-Up	TATGCGGACCGATGACTCAAGTAATGGAAAGGATTTTTTAAATATATTTATCGCAGCrGGGCA/3SpC3/	62	3511
Downstream LDR	AcDx-9954-PROM1-S1-Dn	/5Phos/GGGAGAGTTGAGAGTATGGTTAGGTGTCGCGTTGCTTGGCTTGATCTACCTGA	53	3512
Real-Time Probe	AcDx-9955-PROM1-S1-RT-Pb	/56-FAM/CCTCGCAGC/ZEN/GGGAGAGTTGAGAGTATGG/3IABkFQ/	28	3513
Tag Forward Primer	AcDx-9956-PROM1-S1-RT-FP	TATGCGGACCGATGACTCAA	20	3514
Tag Reverse Primer	AcDx-9957-PROM1-S1-RT-RP	TCAGGTAGATCAAGCCAAGCAA	22	3515
Downstream PCR Primer	AcDx-9958-PROM1-S1-PCR-V	TCAGGTAGATCAAGCCAAGCAACTTTAAATAACTAAAACAAATCCCCACGTGrACACT/3SpC3/	57	3516
HOXB4-S1				
Forward PCR Primer	AcDx-9961-HOXB4-S1-FP	GTTTCGGGAGGAGGGTTTCrGGCGA/3SpC3/	24	3517
Reverse PCR Primer	AcDx-9962-HOXB4-S1-RP	GGTGTCGTGGTAAACCCCCGCCGCCrUCCGT/3SpC3/	30	3518
Upstream LDR	AcDx-9963-HOXB4-S1-Up	TTCGTGGGCACACAAGCAAGGTTTCGGCGGGTAGCrGGCAC/3SpC3/	40	3519
Downstream LDR	AcDx-9964-HOXB4-S1-Dn	/5Phos/GGCGTAGGAGTTCGAGGAGATAGATCGGTTGCTTGGCTTGATCTACCTGA	50	3520
Real-Time Probe	AcDx-9965-HOXB4-S1-RT-Pb	/56-FAM/TTGGGTAGC/ZEN/GGCGTAGGAGTTCGAGG/3IABkFQ/	26	3521
Tag Forward Primer	AcDx-9966-HOXB4-S1-RT-FP	TTCGTGGGCACACAAGCAA	19	3522
Tag Reverse Primer	AcDx-9967-HOXB4-S1-RT-RP	TCAGGTAGATCAAGCCAAGCAA	22	3523
Downstream PCR Primer	AcDx-9968-HOXB4-S1-PCR-V	TCAGGTAGATCAAGCCAAGCAACCGCCACCACCCCTGrCCGCT/3SpC3/	42	3524

CANCR85				
Forward PCR Primer	AcDx-9971-CANCR85-FP	ATTTTTATTTTTATTCGCGTTTTTCGGArUTGGC/3SpC3/	33	3525
Reverse PCR Primer	AcDx-9972-CANCR85-RP	GGTGTCGTGGGATACGAAAATTACCCGTAACGTCrAAAAC/3SpC3/	39	3526
Upstream LDR	AcDx-9973-CANCR85-Up	TTGAAGGAGGAAATCGGCACAGGTAGGTTTTTGGAAGTTTAGGTTGCrGGCAC/3SpC3/	52	3527
Downstream LDR	AcDx-9974-CANCR85-Dn	/5Phos/GGCGTTTATCGTAAAATTTTTTTTCGTCGTAGTTTCGTGATGTCGAACCGTTTTAGGACTGA	62	3528
Real-Time Probe	AcDx-9975-CANCR85-RT-Pb	/56-FAM/AAAGGTTGC/ZEN/GGCGTTTATCGTAAAATTTTTTTTCGTC/3IABkFQ/	37	3529
Tag Forward Primer	AcDx-9976-CANCR85-RT-FP	TTGAAGGAGGAAATCGGCACA	21	3530
Tag Reverse Primer	AcDx-9977-CANCR85-RT-RP	TCAGTCCTAAAACGGTTCGACA	22	3531
Downstream PCR Primer	AcDx-9978-CANCR85-PCR-V	TCAGTCCTAAAACGGTTCGACAGAAAATTACCCGTAACGTCAAAATCATGrAAACC/3SpC3/	55	3532
CANCR86				
Forward PCR Primer	AcDx-9981-CANCR86-FP	GGGATCGTTGGTTGTAGGCrGTTTC/3SpC3/	24	3533
Reverse PCR Primer	AcDx-9982-CANCR86-RP	GGTGTCGTGGCCACTCCCTCATCTACGACrCCCGC/3SpC3/	34	3534
Upstream LDR	AcDx-9983-CANCR86-Up	TTCGGCAGGCTACGGTACAGGTTGTAGGCGTTTTGTTGGAGCTCrGGAGT/3SpC3/	49	3535
Downstream LDR	AcDx-9984-CANCR86-Dn	/5Phos/GGAACGAGGGTGGTTAGGGTCGTTTTTTTACGTGTCGAACCGTTTTAGGACTGA	54	3536
Real-Time Probe	AcDx-9985-CANCR86-RT-Pb	/56-FAM/AAGGAGCTC/ZEN/GGAACGAGGGTGGTTAGG/3IABkFQ/	27	3537
Tag Forward Primer	AcDx-9986-CANCR86-RT-FP	TTCGGCAGGCTACGGTACA	19	3538
Tag Reverse Primer	AcDx-9987-CANCR86-RT-RP	TCAGTCCTAAAACGGTTCGACA	22	3539
Downstream PCR Primer	AcDx-9988-CANCR86-PCR-V	TCAGTCCTAAAACGGTTCGACAACTCCCTCATCTACGACCCTGrUAAAG/3SpC3/	48	3540
HOXA7				
Forward PCR Primer	AcDx-9991-HOXA7-FP	TGGCGAGGTTATTGTAGAGTTCrGGGCG/3SpC3/	27	3541

Reverse PCR Primer	AcDx-9992-HOXA7-RP	GGTGTCGTGGCCTTTACGTCCGACTACGACrCTAAG/3SpC3/	35	3542
Upstream LDR	AcDx-9993-HOXA7-Up	TCTCGACGATGAAAAGCAACAGGGATGTTTTGGTCGTAGGAAGCrGTAAA/3SpC3/	49	3543
Downstream LDR	AcDx-9994-HOXA7-Dn	/5Phos/GTAGGGTAGGTTGTCGTAGGCGTCGGTGTGGGTACTGTCCGTGGA	45	3544
Real-Time Probe	AcDx-9995-HOXA7-RT-Pb	/56-FAM/AAAGGAAGC/ZEN/GTAGGGTAGGTTGTCGTAGG/3IABkFQ/	29	3545
Tag Forward Primer	AcDx-9996-HOXA7-RT-FP	TCTCGACGATGAAAAGCAACA	21	3546
Tag Reverse Primer	AcDx-9997-HOXA7-RT-RP	TCCACGGACAGTACCCACA	19	3547
Downstream PCR Primer	AcDx-9998-HOXA7-PCR-V	TCCACGGACAGTACCCACACCGACTACGACCTAAACGCTGrACGCT/3SpC3/	45	3548
CANCR87				
Forward PCR Primer	AcDx-10001-CANCR87-FP	GCGGTCGTTATTGGTTGCrGGTTC/3SpC3/	23	3549
Reverse PCR Primer	AcDx-10002-CANCR87-RP	GGTGTCGTGGAAAATAAAACTCTAAAAAAACAAAATACGAAAArAAAAC/3SpC3/	48	3550
Upstream LDR	AcDx-10003-CANCR87-Up	TTCTAGGCGACACGACAACAGGTTGGTTATGTGTAGGCGTCGCTCrGTTCC/3SpC3/	50	3551
Downstream LDR	AcDx-10004-CANCR87-Dn	/5Phos/GTTTTTACGGTTTTTTTTTTGATTTTTTCGGCGAATTGTGGGTACTGTCCGTGGA	55	3552
Real-Time Probe	AcDx-10005-CANCR87-RT-Pb	/56-FAM/TTGTCGCTC/ZEN/GTTTTTACGGTTTTTTTTTTGATTTTTTCG/3IABkFQ/	39	3553
Tag Forward Primer	AcDx-10006-CANCR87-RT-FP	TTCTAGGCGACACGACAACA	20	3554
Tag Reverse Primer	AcDx-10007-CANCR87-RT-RP	TCCACGGACAGTACCCACA	19	3555
Downstream PCR Primer	AcDx-10008-CANCR87-PCR-V	TCCACGGACAGTACCCACAAATAAAACTCTAAAAAAACAAAATACGAAAAAAAATTTGrCCGAG/3SpC3/	63	3556
CANCR88				
Forward PCR Primer	AcDx-10011-CANCR88-FP	GGAGACGTCGTTCGTGTCrGTGTC/3SpC3/	23	3557
Reverse PCR Primer	AcDx-10012-CANCR88-RP	GGTGTCGTGGCTACTATTCCGATAACTAAAAACGAAArAACAT/3SpC3/	42	3558
Upstream LDR	AcDx-10013-CANCR88-	TTTCAGGCCCTAACCACCACGTCGTGTTAGGTTGTTTTCGAGCGCrGTTAC/3SpC3/	50	3559

	Up			
Downstream LDR	AcDx-10014-CANCR88-Dn	/5Phos/GTTTGTCGTGAGTTTTTGTCGCGTGATTTATTCGTGGTGGGATTAAGGGCGATGGA	56	3560
Real-Time Probe	AcDx-10015-CANCR88-RT-Pb	/56-FAM/AACGAGCGC/ZEN/GTTTGTCGTGAGTTTTTGTCG/3IABkFQ/	30	3561
Tag Forward Primer	AcDx-10016-CANCR88-RT-FP	TTTCAGGCCCTAACCACCAC	20	3562
Tag Reverse Primer	AcDx-10017-CANCR88-RT-RP	TCCATCGCCCTTAATCCCAC	20	3563
Downstream PCR Primer	AcDx-10018-CANCR88-PCR-V	TCCATCGCCCTTAATCCCACGATAACTAAAAACGAAAAACACGAATAAATCATGrCGACG/3SpC3/	59	3564
GREM1-S2				
Forward PCR Primer	AcDx-10021-GREM1-S2-FP	TGGTAGACGTTTTGGCGTCrGGGCG/3SpC3/	24	3565
Reverse PCR Primer	AcDx-10022-GREM1-S2-RP	GGTGTCGTGGATAAAACGCTACAAAACGAAACTArAAACT/3SpC3/	39	3566
Upstream LDR	AcDx-10023-GREM1-S2-Up	TATCTCCTAAAAGAAGCCGCACGGCGTCGGGTAGCGGTTGTCrGGTCA/3SpC3/	47	3567
Downstream LDR	AcDx-10024-GREM1-S2-Dn	/5Phos/GGTTGGTTTTTATTTTCGCGCGTATTTTTTAAATTGTGGTGGGATTAAGGGCGATGGA	58	3568
Real-Time Probe	AcDx-10025-GREM1-S2-RT-Pb	/56-FAM/TTGGTTGTC/ZEN/GGTTGGTTTTTATTTTCGCGCG/3IABkFQ/	31	3569
Tag Forward Primer	AcDx-10026-GREM1-S2-RT-FP	TATCTCCTAAAAGAAGCCGCAC	22	3570
Tag Reverse Primer	AcDx-10027-GREM1-S2-RT-RP	TCCATCGCCCTTAATCCCAC	20	3571
Alternate Group 3 Markers				
TBX1-S1				

Forward PCR Primer	AcDx-7771-TBX1-S1-FP	TTTCGTTTCGGTTTTGTATAGTTTCrGAAGC/3SpC3/	30	3572
Reverse PCR Primer	AcDx-7772-TBX1-S1-RP	GGTGTCGTGGAAAAAAAAACCGCGATAACCCrCTAAG/3SpC3/	36	3573
Upstream LDR	AcDx-7773-TBX1-S1-Up	TGAACGCTCAAACACGTGAACTGTATAGTTTCGAAGTTCGTCGGGCTCrGGTGC/3SpC3/	53	3574
Downstream LDR	AcDx-7774-TBX1-S1-Dn	/5Phos/GGTTATTTTGTTTTAAGGGTAAGTAAGGAATACGTTTTTTTAGTTTTAGGTTGGCCT GTAAGCGTTCCA	69	3575
Real-Time Probe	AcDx-7775-TBX1-S1-RT-Pb	/56-FAM/AACGGGCTC/ZEN/GGTTATTTTGTTTTAAGGGTAAGTAAGGA/3IABkFQ/	38	3576
Tag Forward Primer	AcDx-7776-TBX1-S1-RT-FP	TGAACGCTCAAACACGTGAAC	21	3577
Tag Reverse Primer	AcDx-7777-TBX1-S1-RT-RP	TGGAACGCTTACAGGCCAAC	20	3578
PTPRU-S2				
Forward PCR Primer	AcDx-10031-PTPRU-S2-FP	GCGGTAGAGCGAGGCrGTTGA/3SpC3/	20	3579
Reverse PCR Primer	AcDx-10032-PTPRU-S2-RP	GGTGTCGTGGACTCACGACAACCAACGAAArAAATG/3SpC3/	35	3580
Upstream LDR	AcDx-10033-PTPRU-S2-Up	TCGCTCTTCAGCCTCCTACACGTTGGTGTCGGCAGCrGGGTA/3SpC3/	41	3581
Downstream LDR	AcDx-10034-PTPRU-S2-Dn	/5Phos/GGGCGTGCGGTATATTAGTTATCGGCGTGTTCTGGGAATTATTGCCGGA	49	3582
Real-Time Probe	AcDx-10035-PTPRU-S2-RT-Pb	/56-FAM/AACGGCAGC/ZEN/GGGCGTGC/3IABkFQ/	17	3583
Tag Forward Primer	AcDx-10036-PTPRU-S2-RT-FP	TCGCTCTTCAGCCTCCTACA	20	3584
Tag Reverse Primer	AcDx-10037-PTPRU-S2-RT-RP	TCCGGCAATAATTCCCAGAACA	22	3585
Downstream PCR Primer	AcDx-10038-PTPRU-S2-PCR-V	TCCGGCAATAATTCCCAGAACACCAACGAAAAAATAACCAAAAAACGCTGrATAAT/3SpC3/	55	3586
HOXB4-S2				
Forward PCR Primer	AcDx-10041-HOXB4-S2-	GGTCGTCGGCGTAGTATTCrGGGCA/3SpC3/	24	3587

	FP			
Reverse PCR Primer	AcDx-10042-HOXB4-S2-RP	GGTGTCGTGGTCAAACTATATCGACCCCAAATTCrCCTCT/3SpC3/	39	3588
Upstream LDR	AcDx-10043-HOXB4-S2-Up	TTCAACGATCGCGCAGACACGTAGTATTCGGGCGAGTGATCrGTTAA/3SpC3/	46	3589
Downstream LDR	AcDx-10044-HOXB4-S2-Dn	/5Phos/GTTGGGTAGGTAATCGTTTTGTGAATATTTTTCGTATGGAGTGTTCTGGGAATTATTGCCGGA	63	3590
Real-Time Probe	AcDx-10045-HOXB4-S2-RT-Pb	/56-FAM/TTAGTGATC/ZEN/GTTGGGTAGGTAATCGTTTTGTGAAT/3IABkFQ/	35	3591
Tag Forward Primer	AcDx-10046-HOXB4-S2-RT-FP	TTCAACGATCGCGCAGACA	19	3592
Tag Reverse Primer	AcDx-10047-HOXB4-S2-RT-RP	TCCGGCAATAATTCCCAGAACA	22	3593
Downstream PCR Primer	AcDx-10048-HOXB4-S2-PCR-V	TCCGGCAATAATTCCCAGAACACGACCCCAAATTCCCTCCATATGrAAAAG/3SpC3/	50	3594
HOXB4-S3				
Forward PCR Primer	AcDx-10051-HOXB4-S3-FP	GTTAAGTGAATAAAGTTAGGCGTTTACrGTGAC/3SpC3/	32	3595
Reverse PCR Primer	AcDx-10052-HOXB4-S3-RP	GGTGTCGTGGAACCTCTTTAACTAAAATAAAATCCGAArUAAAT/3SpC3/	43	3596
Upstream LDR	AcDx-10053-HOXB4-S3-Up	TCACTATCGGCGTAGTCACCACGTTTACGTGATTTTTCGAGTTAATGATCrGTTCT/3SpC3/	55	3597
Downstream LDR	AcDx-10054-HOXB4-S3-Dn	/5Phos/GTTTCGTTTGCGATTTTCGGATAAGGAAATTTGTTTGGTGACTTTACCCGGAGGA	55	3598
Real-Time Probe	AcDx-10055-HOXB4-S3-RT-Pb	/56-FAM/AAAATGATC/ZEN/GTTTCGTTTGCGATTTTCGGATAAGG/3IABkFQ/	35	3599
Tag Forward Primer	AcDx-10056-HOXB4-S3-RT-FP	TCACTATCGGCGTAGTCACCA	21	3600
Tag Reverse Primer	AcDx-10057-HOXB4-S3-RT-RP	TCCTCCGGGTAAAGTCACCA	20	3601

HOXA9-S2				
Forward PCR Primer	AcDx-10061-HOXA9-S2-FP	GTTATGAGGATTTTTGTGGTTCrGGGTT/3SpC3/	27	3602
Reverse PCR Primer	AcDx-10062-HOXA9-S2-RP	GGTGTCGTGGCGACCGATTCCTTCCACTTCrUTACT/3SpC3/	35	3603
Upstream LDR	AcDx-10063-HOXA9-S2-Up	TCATCTGTTCGTCAGGGTCCAGGATTTTTGTGGTTCGGGTCGCTCrGTAAC/3SpC3/	50	3604
Downstream LDR	AcDx-10064-HOXA9-S2-Dn	/5Phos/GTAGTTTTATTTTGGTAGGATTTACGTCGTTATTGGTCGAAGTGGTGACTTTACCCGGAGGA	62	3605
Real-Time Probe	AcDx-10065-HOXA9-S2-RT-Pb	/56-FAM/TTGTCGCTC/ZEN/GTAGTTTTATTTTGGTAGGATTTACGTCG/3IABkFQ/	38	3606
Tag Forward Primer	AcDx-10066-HOXA9-S2-RT-FP	TCATCTGTTCGTCAGGGTCCA	21	3607
Tag Reverse Primer	AcDx-10067-HOXA9-S2-RT-RP	TCCTCCGGGTAAAGTCACCA	20	3608
Downstream PCR Primer	AcDx-10068-HOXA9-S2-PCR-V	TCCTCCGGGTAAAGTCACCACCGATTCCTTCCACTTCTTACCTTTGrACCAG/3SpC3/	51	3609
HOXA9-S3				
Forward PCR Primer	AcDx-10071-HOXA9-S3-FP	GCGTTCGCGTTTTTATTGGTCrGTGCA/3SpC3/	26	3610
Reverse PCR Primer	AcDx-10072-HOXA9-S3-RP	GGTGTCGTGGATAACTACAAAACATCGAACCATTArATAAT/3SpC3/	40	3611
Upstream LDR	AcDx-10073-HOXA9-S3-Up	TTCGTACCTCGGCACACCAGTTTTTATTGGTCGTGCGCGTCACrGTGCC/3SpC3/	48	3612
Downstream LDR	AcDx-10074-HOXA9-S3-Dn	/5Phos/GTGTTCGTTTAGTAGAATAATAACGCGTAAATTATTTCGTACGTGGCTCCGTTACTCTGTCGA	63	3613
Real-Time Probe	AcDx-10075-HOXA9-S3-RT-Pb	/56-FAM/TTGCGTCAC/ZEN/GTGTTCGTTTAGTAGAATAATAACGCGTAAAT/3IABkFQ/	41	3614
Tag Forward Primer	AcDx-10076-HOXA9-S3-RT-FP	TTCGTACCTCGGCACACCA	19	3615
Tag Reverse Primer	AcDx-10077-HOXA9-S3-RT-RP	TCGACAGAGTAACGGAGCCA	20	3616

Downstream PCR Primer	AcDx-10078-HOXA9-S3-PCR-V	TCGACAGAGTAACGGAGCCAAACTACAAAACATCGAACCATTAATAACGTATGrAAATG/3SpC3/	58	3617
HOXA9-S4				
Forward PCR Primer	AcDx-10081-HOXA9-S4-FP	GTTTTTATTGGTCGTGCGCrGTTAT/3SpC3/	24	3618
Reverse PCR Primer	AcDx-10082-HOXA9-S4-RP	GGTGTCGTGGGCTAAAAATTATAACTACAAAACATCGAArCCATC/3SpC3/	44	3619
Upstream LDR	AcDx-10083-HOXA9-S4-Up	TCTACAGCTAGATGCGGCCAGTCGTGCGCGTTACGTGCTCrGTTCG/3SpC3/	45	3620
Downstream LDR	AcDx-10084-HOXA9-S4-Dn	/5Phos/GTTTAGTAGAATAATAACGCGTAAATTATTTCGTACGTTATTAATGGTGGCTCCGTTACTCTGTCGA	67	3621
Real-Time Probe	AcDx-10085-HOXA9-S4-RT-Pb	/56-FAM/CCCGTGCTC/ZEN/GTTTAGTAGAATAATAACGCGTAAATTATTTCGTACG/3IABkFQ/	46	3622
Tag Forward Primer	AcDx-10086-HOXA9-S4-RT-FP	TCTACAGCTAGATGCGGCCA	20	3623
Tag Reverse Primer	AcDx-10087-HOXA9-S4-RT-RP	TCGACAGAGTAACGGAGCCA	20	3624
WNT7B-S2				
Forward PCR Primer	AcDx-10091-WNT7B-S2-FP	GTTCGGATTTTGTAATTATGTTTTATAGTTCrGGGAG/3SpC3/	36	3625
Reverse PCR Primer	AcDx-10092-WNT7B-S2-RP	GGTGTCGTGGCCTTCTATAAATAATCGCGCATCTTAArAACAG/3SpC3/	42	3626
Upstream LDR	AcDx-10093-WNT7B-S2-Up	TCCAGGGTATTTGGCGCACGATTTTGTAATTATGTTTTATAGTTCGGGAAATTGCrGGGCC/3SpC3/	60	3627
Downstream LDR	AcDx-10094-WNT7B-S2-Dn	/5Phos/GGGTTTCGTAGTATTTGGTTGCGGTTGCGGTGCGGAAACCTATCGTCGA	49	3628
Real-Time Probe	AcDx-10095-WNT7B-S2-RT-Pb	/56-FAM/TTAAATTGC/ZEN/GGGTTTCGTAGTATTTGGTTGCG/3IABkFQ/	32	3629
Tag Forward Primer	AcDx-10096-WNT7B-S2-	TCCAGGGTATTTGGCGCAC	19	3630

	RT-FP			
Tag Reverse Primer	AcDx-10097-WNT7B-S2-RT-RP	TCGACGATAGGTTTCCGCAC	20	3631
Downstream PCR Primer	AcDx-10098-WNT7B-S2-PCR-V	TCGACGATAGGTTTCCGCACTAAATAATCGCGCATCTTAAAACAAAAATTTGrCAACT/3SpC3/	57	3632
CANCR89				
Forward PCR Primer	AcDx-10101-CANCR89-FP	GCGTTTCGTTTAAAGGTTGTTTCrGATTC/3SpC3/	28	3633
Reverse PCR Primer	AcDx-10102-CANCR89-RP	GGTGTCGTGGCAAAAAAAATTAAAATATAATAATAAAAAAACCGAArUTTCT/3SpC3/	51	3634
Upstream LDR	AcDx-10103-CANCR89-Up	TCCCTCGTCATCTCCCTTACCGTTGATTAGTGATTTGTGTAAAAGAAAATTATTCGGTCrGTTAC/3SpC3/	64	3635
Downstream LDR	AcDx-10104-CANCR89-Dn	/5Phos/GTTGTTTGTTTTTTTAGCGTTTTTTTCGGTTGCGGGTCTTGGTGATGGAGCGA	53	3636
Real-Time Probe	AcDx-10105-CANCR89-RT-Pb	/56-FAM/CCTTCGGTC/ZEN/GTTGTTTGTTTTTTTAGCGTTTTTTTCG/3IABkFQ/	37	3637
Tag Forward Primer	AcDx-10106-CANCR89-RT-FP	TCCCTCGTCATCTCCCTTACC	21	3638
Tag Reverse Primer	AcDx-10107-CANCR89-RT-RP	TCGCTCCATCACCAAGACC	19	3639
Downstream PCR Primer	AcDx-10108-CANCR89-PCR-V	TCGCTCCATCACCAAGACCCAAAAAAAATTAAAATATAATAATAAAAAAACCGAATTTCTGrCAACT/3SpC3/	66	3640
CANCR90				
Forward PCR Primer	AcDx-10111-CANCR90-FP	CGGGTTTAGTTTATTGGTTGTAGTCrGAGGA/3SpC3/	30	3641
Reverse PCR Primer	AcDx-10112-CANCR90-RP	GGTGTCGTGGATACCCCGAACCACCAACrCCATT/3SpC3/	33	3642
Upstream LDR	AcDx-10113-CANCR90-Up	TCATAATGTTGTCAGCCCGACCGTTTATTGGTTGTAGTCGAGAGGTTTGTCrGAGAA/3SpC3/	56	3643
Downstream LDR	AcDx-10114-CANCR90-Dn	/5Phos/GAGGGTAGCGGTTTTTGATAGGATTTGTTTTTTTAATTTTTTGGATAGGGTCTTGGTGATGGAGCGA	67	3644
Real-Time Probe	AcDx-10115-CANCR90-	/56-FAM/TTGTTTGTC/ZEN/GAGGGTAGCGGTTTTTGATAGGATTTG/3IABkFQ/	36	3645

	RT-Pb			
Tag Forward Primer	AcDx-10116-CANCR90-RT-FP	TCATAATGTTGTCAGCCCGACC	22	3646
Tag Reverse Primer	AcDx-10117-CANCR90-RT-RP	TCGCTCCATCACCAAGACC	19	3647
Downstream PCR Primer	AcDx-10118-CANCR90-PCR-V	TCGCTCCATCACCAAGACCCGAACCACCAACCCATCCTGrCAAAG/3SpC3/	44	3648
CANCR91				
Forward PCR Primer	AcDx-10121-CANCR91-FP	TGTTTGGGAAGGAAGTTTGCrGGGTC/3SpC3/	25	3649
Reverse PCR Primer	AcDx-10122-CANCR91-RP	GGTGTCGTGGCACGCAACTAATACGAAAAAAATAAATAArCTAAG/3SpC3/	44	3650
Upstream LDR	AcDx-10123-CANCR91-Up	TACGAATCACCCGAGAGTTCAAGGGAAGGAAGTTTGCGGGTTTTCTCrGGGCC/3SpC3/	52	3651
Downstream LDR	AcDx-10124-CANCR91-Dn	/5Phos/GGGTTTCGTTGTAGTTTAGTTAGGTTAATTAATTAGTAGTCGTTTAGTTGTGGGTGGGTATAGGTCAGA	69	3652
Real-Time Probe	AcDx-10125-CANCR91-RT-Pb	/56-FAM/TTTTTTCTC/ZEN/GGGTTTCGTTGTAGTTTAGTTAGGTTAATT/3IABkFQ/	39	3653
Tag Forward Primer	AcDx-10126-CANCR91-RT-FP	TACGAATCACCCGAGAGTTCAA	22	3654
Tag Reverse Primer	AcDx-10127-CANCR91-RT-RP	TCTGACCTATACCCACCCACAA	22	3655
Downstream PCR Primer	AcDx-10128-CANCR91-PCR-V	TCTGACCTATACCCACCCACAACACGCAACTAATACGAAAAAAATAAATAACTAAATGrACTAT/3SpC3/	63	3656
HOXD9-S2				
Forward PCR Primer	AcDx-10131-HOXD9-S2-FP	GAGGTTGTAGTTTGCGAATTAGTCrGGTGA/3SpC3/	29	3657
Reverse PCR Primer	AcDx-10132-HOXD9-S2-RP	GGTGTCGTGGATTTTAAAACGTCCCGCACTCrCCACT/3SpC3/	36	3658
Upstream LDR	AcDx-10133-HOXD9-S2-Up	TACGAATCACCCGAGAGTTCAAGAATTAGTCGGTGGTTCGAGCrGTCAA/3SpC3/	48	3659

Downstream LDR	AcDx-10134-HOXD9-S2-Dn	/5Phos/GTCGGCGGAGAGTTGTTCGGTTGTGGGTGGGTATAGGTCAGA	42	3660
Real-Time Probe	AcDx-10135-HOXD9-S2-RT-Pb	/56-FAM/CCTTCGAGC/ZEN/GTCGGCGGAGAG/3IABkFQ/	21	3661
Tag Forward Primer	AcDx-10136-HOXD9-S2-RT-FP	TACGAATCACCCGAGAGTTCAA	22	3662
Tag Reverse Primer	AcDx-10137-HOXD9-S2-RT-RP	TCTGACCTATACCCACCCACAA	22	3663
Downstream PCR Primer	AcDx-10138-HOXD9-S2-PCR-V	TCTGACCTATACCCACCCACAACAACATTACACTATCCGCCGCTGrAACAG/3SpC3/	50	3664

IDT Abbreviation	Modifications
/5Phos/	5' Phosphorylation
rX (X=A,C,G,U)	RNA Base
/3spC3/	3' C3 DNA Spacer
/56-FAM/	5' 6-FAM™ Fluorescent Tag
/Zen/	Internal Quencher
/3IABkFQ/	3' Iowa Black® FQ Quencher

Table 56. Primers for use in Step 2 of the Group 4- 44-48-marker assay, with average sensitivities of 50%, to detect and identify prostate and bladder cancers.

Site	Primer Name	Sequence	Length	Seq. ID No.
Prefered Group 4 Markers				
TMEM106A-S1				
Forward PCR Primer	AcDx-10141-TMEM106A-S1-	ATTTTTATTCGGATTGGTTAGTTTTTGCrGGAAA/3SpC3/	33	3665

	FP			
Reverse PCR Primer	AcDx-10142-TMEM106A-S1-RP	GGTGTCGTGGCTTCGCCACGCAACAACrCTAAG/3SpC3/	32	3666
Upstream LDR	AcDx-10143-TMEM106A-S1-Up	TAGGAACACGGAGGACATCAAGGTTAGTTTTTGCGGAAGTAGCGCTCrGGGCC/3SpC3/	52	3667
Downstream LDR	AcDx-10144-TMEM106A-S1-Dn	/5Phos/GGGTTTTCGGGTAACGTTTGAAGAGTTGGTAGTTGTGGGTGGGTATAGGTCAGA	54	3668
Real-Time Probe	AcDx-10145-TMEM106A-S1-RT-Pb	/56-FAM/AAAGCGCTC/ZEN/GGGTTTTCGGGTAACG/3IABkFQ/	25	3669
Tag Forward Primer	AcDx-10146-TMEM106A-S1-RT-FP	TAGGAACACGGAGGACATCAA	21	3670
Tag Reverse Primer	AcDx-10147-TMEM106A-S1-RT-RP	TCTGACCTATACCCACCCACAA	22	3671
Downstream PCR Primer	AcDx-10148-TMEM106A-S1-PCR-V	TCTGACCTATACCCACCCACAACGCCACGCAACAACCTAAAATGrCTACT/3SpC3/	49	3672
CANCR92				
Forward PCR Primer	AcDx-10151-CANCR92-FP	GGGCGGAAAGCGATACrGGTTA/3SpC3/	21	3673
Reverse PCR Primer	AcDx-10152-CANCR92-RP	GGTGTCGTGGTCCAAAACCTCTCTTCGATTTATCrUCCAT/3SpC3/	39	3674
Upstream LDR	AcDx-10153-CANCR92-Up	TCTCATAAACACTCCGGCCACGAAAGCGATACGGTTGGGAGACrGATCG/3SpC3/	48	3675
Downstream LDR	AcDx-10154-CANCR92-Dn	/5Phos/GATTACGTGTTTTTTTAATTTTTTTTTCGTTGTTAGTAAGGAAGTGGGTGGCTCAATAACGGGCAGA	67	3676
Real-Time Probe	AcDx-10155-CANCR92-RT-Pb	/56-FAM/AAGGGAGAC/ZEN/GATTACGTGTTTTTTTAATTTTTTTTTCGTTGTTA/3IABkFQ/	44	3677
Tag Forward Primer	AcDx-10156-CANCR92-RT-FP	TCTCATAAACACTCCGGCCAC	21	3678
Tag Reverse Primer	AcDx-10157-CANCR92-RT-RP	TCTGCCCGTTATTGAGCCAC	20	3679
HLA-F-S1				
Forward PCR Primer	AcDx-10161-HLA-F-S1-FP	ATTTTTTAGGGAATGAATGGTTGCrGATAC/3SpC3/	29	3680

Reverse PCR Primer	AcDx-10162-HLA-F-S1-RP	GGTGTCGTGGAAACGCAAATCCTCGTTCAAArAAAAC/3SpC3/	36	3681
Upstream LDR	AcDx-10163-HLA-F-S1-Up	TTGATTGGGATCGTTCGCACGCGATATGAGGTTCGACGAACrGTTCC/3SpC3/	46	3682
Downstream LDR	AcDx-10164-HLA-F-S1-Dn	/5Phos/GTTTTTTTCGCGGGTATTATTAGTACGCGTACGACGTGGCTCAATAACGGGCAGA	55	3683
Real-Time Probe	AcDx-10165-HLA-F-S1-RT-Pb	/56-FAM/AAGACGAAC/ZEN/GTTTTTTTCGCGGGTATTATTAGTACGC/3IABkFQ/	37	3684
Tag Forward Primer	AcDx-10166-HLA-F-S1-RT-FP	TTGATTGGGATCGTTCGCAC	20	3685
Tag Reverse Primer	AcDx-10167-HLA-F-S1-RT-RP	TCTGCCCGTTATTGAGCCAC	20	3686
Downstream PCR Primer	AcDx-10168-HLA-F-S1-PCR-V	TCTGCCCGTTATTGAGCCACCCTCGTTCAAAAAAATATAATCCTTACCGTTGrUACGT/3SpC3/	57	3687
SLC25A39				
Forward PCR Primer	AcDx-10171-SLC25A39-FP	GCGGTAGTGAGAATGAGTTTTTCrGAGAC/3SpC3/	28	3688
Reverse PCR Primer	AcDx-10172-SLC25A39-RP	GGTGTCGTGGAAAATCGCCGCGAAATTTTCrAAATT/3SpC3/	35	3689
Upstream LDR	AcDx-10173-SLC25A39-Up	TCTGCCCAAAATACTGCACAAGGTAGTGAGAATGAGTTTTTCGAGATCATCrGGGCA/3SpC3/	56	3690
Downstream LDR	AcDx-10174-SLC25A39-Dn	/5Phos/GGGAGATTATATAATAACGACGAAGTTATTTGTCGGGTTATTCGTTGAAACTGAGGCGGTGTTCA	65	3691
Real-Time Probe	AcDx-10175-SLC25A39-RT-Pb	/56-FAM/TTGATCATC/ZEN/GGGAGATTATATAATAACGACGAAGTTATTTGTCG/3IABkFQ/	44	3692
Tag Forward Primer	AcDx-10176-SLC25A39-RT-FP	TCTGCCCAAAATACTGCACAA	21	3693
Tag Reverse Primer	AcDx-10177-SLC25A39-RT-RP	TGAACACCGCCTCAGTTTCAA	21	3694
Downstream PCR Primer	AcDx-10178-SLC25A39-PCR-V	TGAACACCGCCTCAGTTTCAAGCCGCGAAATTTTCAAATCTCGATGrAATAG/3SpC3/	51	3695
EIF5A2				
Forward PCR Primer	AcDx-10181-EIF5A2-FP	CGTTGAGGGTGAGGATATTCrGTTTC/3SpC3/	25	3696
Reverse PCR Primer	AcDx-10182-EIF5A2-RP	GGTGTCGTGGAAAAATCGCCGACTTCTACGArUAAAG/3SpC3/	36	3697
Upstream LDR	AcDx-10183-EIF5A2-Up	TATGGACTGTACCAGCCCAAGGGTGAGGATATTCGTTTTGAGCrGTTCC/3SpC3/	48	3698
Downstream LDR	AcDx-10184-EIF5A2-Dn	/5Phos/GTTTTTTCGTGTTGGAAATAGTAGGTTGCGGTTTTTATCGTTGAAACTGAGGCG	61	3699

		GTGTTCA		
Real-Time Probe	AcDx-10185-EIF5A2-RT-Pb	/56-FAM/CCTTTGAGC/ZEN/GTTTTTTCGTGTTGGAAATAGTAGGTTGC/3IABkFQ/	38	3700
Tag Forward Primer	AcDx-10186-EIF5A2-RT-FP	TATGGACTGTACCAGCCCAA	20	3701
Tag Reverse Primer	AcDx-10187-EIF5A2-RT-RP	TGAACACCGCCTCAGTTTCAA	21	3702
Downstream PCR Primer	AcDx-10188-EIF5A2-PCR-V	TGAACACCGCCTCAGTTTCAAAAATCGCCGACTTCTACGATAAAATGrATAAG/3SpC3/	52	3703
ICAM4				
Forward PCR Primer	AcDx-10191-ICAM4-FP	GTGGAGTCGAGGAGGCrGGGAC/3SpC3/	21	3704
Reverse PCR Primer	AcDx-10192-ICAM4-RP	GGTGTCGTGGACAACGACAAAAAAAACAAAAACCrCCATG/3SpC3/	39	3705
Upstream LDR	AcDx-10193-ICAM4-Up	TTTGCCTCTTGTAGGTGCCAGAGGTTGCGGGTTTTTTTATTTTTAGAATCrGGTCC/3SpC3/	55	3706
Downstream LDR	AcDx-10194-ICAM4-Dn	/5Phos/GGTTTTGGTTTTTTGGCGCGGAGTTTTTTAGTGGGCAACGCGGATATTCA	50	3707
Real-Time Probe	AcDx-10195-ICAM4-RT-Pb	/56-FAM/AATAGAATC/ZEN/GGTTTTGGTTTTTTGGCGCGG/3IABkFQ/	30	3708
Tag Forward Primer	AcDx-10196-ICAM4-RT-FP	TTTGCCTCTTGTAGGTGCCA	20	3709
Tag Reverse Primer	AcDx-10197-ICAM4-RT-RP	TGAATATCCGCGTTGCCCA	19	3710
Downstream PCR Primer	AcDx-10198-ICAM4-PCR-V	TGAATATCCGCGTTGCCCAAAACAAAAACCCCATAACAAAAAACCTGrAACTG/3SpC3/	52	3711
GSTP1				
Forward PCR Primer	AcDx-10201-GSTP1-FP	TAGGACGTTTTTAGTGTCGTTAGCrGGTTC/3SpC3/	29	3712
Reverse PCR Primer	AcDx-10202-GSTP1-RP	GGTGTCGTGGCCAAAACGCGATAAACGCAArUCTAT/3SpC3/	35	3713
Upstream LDR	AcDx-10203-GSTP1-Up	TGTCGCCCGGTAGCAATAAACTCGTTAGCGGTTTTTAAGGGATTCrGGAAT/3SpC3/	50	3714
Downstream LDR	AcDx-10204-GSTP1-Dn	/5Phos/GGAGCGTTTCGGAGAGGGATGGGTTTCCGCGATCTTTGCATTCA	44	3715
Real-Time Probe	AcDx-10205-GSTP1-RT-Pb	/56-FAM/TTGGGATTC/ZEN/GGAGCGTTTCGGAGA/3IABkFQ/	24	3716
Tag Forward Primer	AcDx-10206-GSTP1-RT-FP	TGTCGCCCGGTAGCAATAAAC	21	3717
Tag Reverse Primer	AcDx-10207-GSTP1-RT-RP	TGAATGCAAAGATCGCGGAAAC	22	3718

SERPINB9-S1				
Forward PCR Primer	AcDx-10211-SERPINB9-S1-FP	GATTTTCGAGGCGTAGGATTCrGGTTC/3SpC3/	26	3719
Reverse PCR Primer	AcDx-10212-SERPINB9-S1-RP	GGTGTCGTGGTAAAAAACGCCCGACCCTArCGATG/3SpC3/	34	3720
Upstream LDR	AcDx-10213-SERPINB9-S1-Up	TGTGCACTAGTCCACGTGAAACTTCGGTTTTGTTGTTGCGTTCATCrGCGAG/3SpC3/	51	3721
Downstream LDR	AcDx-10214-SERPINB9-S1-Dn	/5Phos/GCGGATTTTCGTTAGGGTTTTTGTAGGTATCGTTGTTTCCGCGATCTTTGCATTCA	56	3722
Real-Time Probe	AcDx-10215-SERPINB9-S1-RT-Pb	/56-FAM/TTGTTCATC/ZEN/GCGGATTTTCGTTAGGGTTTTTGTA/3IABkFQ/	34	3723
Tag Forward Primer	AcDx-10216-SERPINB9-S1-RT-FP	TGTGCACTAGTCCACGTGAAAC	22	3724
Tag Reverse Primer	AcDx-10217-SERPINB9-S1-RT-RP	TGAATGCAAAGATCGCGGAAAC	22	3725
Downstream PCR Primer	AcDx-10218-SERPINB9-S1-PCR-V	TGAATGCAAAGATCGCGGAAACCGCCCGACCCTACGATATAAAAATGrATACT/3SpC3/	52	3726
NFATC4				
Forward PCR Primer	AcDx-10221-NFATC4-FP	GAGTTGGTTTTTGGCGGATCrGAGTC/3SpC3/	25	3727
Reverse PCR Primer	AcDx-10222-NFATC4-RP	GGTGTCGTGGACGCCCCAACGAAACCrUAACG/3SpC3/	31	3728
Upstream LDR	AcDx-10223-NFATC4-Up	TACAGATACGGACGGGAATCAAGTTTTTGGCGGATCGAGTTTTGGCTCrGGTTA/3SpC3/	53	3729
Downstream LDR	AcDx-10224-NFATC4-Dn	/5Phos/GGTCGATTGGACGTTCGGAGTTGGTTGTTTACATCCTCCTGCGTCA	46	3730
Real-Time Probe	AcDx-10225-NFATC4-RT-Pb	/56-FAM/AATTGGCTC/ZEN/GGTCGATTGGACGTTCG/3IABkFQ/	26	3731
Tag Forward Primer	AcDx-10226-NFATC4-RT-FP	TACAGATACGGACGGGAATCAA	22	3732
Tag Reverse Primer	AcDx-10227-NFATC4-RT-RP	TGACGCAGGAGGATGTAAACAA	22	3733
Downstream PCR Primer	AcDx-10228-NFATC4-PCR-V	TGACGCAGGAGGATGTAAACAAGCCCCAACGAAACCTAACAATGrCCCCT/3SpC3/	49	3734
CANCR93				

Forward PCR Primer	AcDx-10231-CANCR93-FP	CGGGTTGTATAGAGAGGTCrGGATA/3SpC3/	24	3735
Reverse PCR Primer	AcDx-10232-CANCR93-RP	GGTGTCGTGGAACCCCTCTCCCGCCrACCCA/3SpC3/	30	3736
Upstream LDR	AcDx-10233-CANCR93-Up	TTGGCGCAACGGTTTCCAAAGAGGTCGGATGTGGAGCrGAGCC/3SpC3/	42	3737
Downstream LDR	AcDx-10234-CANCR93-Dn	/5Phos/GAGTTGGGTAGAGGAAGAGGAATCGTAGATGTCGTTGTTTACATCCTCCTGCG TCA	56	3738
Real-Time Probe	AcDx-10235-CANCR93-RT-Pb	/56-FAM/CCGTGGAGC/ZEN/GAGTTGGGTAGAGGAAGAGG/3IABkFQ/	29	3739
Tag Forward Primer	AcDx-10236-CANCR93-RT-FP	TTGGCGCAACGGTTTCCAA	19	3740
Tag Reverse Primer	AcDx-10237-CANCR93-RT-RP	TGACGCAGGAGGATGTAAACAA	22	3741
Downstream PCR Primer	AcDx-10238-CANCR93-PCR-V	TGACGCAGGAGGATGTAAACAACCGCCACCCGACATCTATGrATTCT/3SpC3/	46	3742
HLA-J-S1				
Forward PCR Primer	AcDx-10241-HLA-J-S1-FP	TTAAGATCGATTTCGAGGTTGCrGGGAC/3SpC3/	27	3743
Reverse PCR Primer	AcDx-10242-HLA-J-S1-RP	GGTGTCGTGGCCTAACAACCTACGAAAATTTTAACCrUCAAT/3SpC3/	41	3744
Upstream LDR	AcDx-10243-HLA-J-S1-Up	TTCAGCAGCCTGGCATCACCGAGGTTGCGGGATTTGTAGAGATTCTCrGATCT/3SpC3/	52	3745
Downstream LDR	AcDx-10244-HLA-J-S1-Dn	/5Phos/GATTCGGGAGAGTTTTAGGCGTTTTTATTTGGTTTTAGTGGAGCGCTAAGGTTG CA	56	3746
Real-Time Probe	AcDx-10245-HLA-J-S1-RT-Pb	/56-FAM/TTGATTCTC/ZEN/GATTCGGGAGAGTTTTAGGCGTTTTTA/3IABkFQ/	36	3747
Tag Forward Primer	AcDx-10246-HLA-J-S1-RT-FP	TTCAGCAGCCTGGCATCAC	19	3748
Tag Reverse Primer	AcDx-10247-HLA-J-S1-RT-RP	TGCAACCTTAGCGCTCCAC	19	3749
SPOCK2				
Forward PCR Primer	AcDx-10251-SPOCK2-FP	GTTAGGAGTAGTCGGCGTCrGTTTC/3SpC3/	24	3750
Reverse PCR Primer	AcDx-10252-SPOCK2-RP	GGTGTCGTGGCAAAAATAAAACAAAACGAACGAAAArATATG/3SpC3/	41	3751
Upstream LDR	AcDx-10253-SPOCK2-Up	TTGTTCGCCCGTTGGTCACGTAGTCGGCGTCGTTTTTAGCrGTACC/3SpC3/	45	3752
Downstream LDR	AcDx-10254-SPOCK2-Dn	/5Phos/GTATTCGGGTTTATTTTTTATAGTATTTTTAACGATTTCGAGGTTTAGGTGGAGC GCTAAGGTTGCA	67	3753

Real-Time Probe	AcDx-10255-SPOCK2-RT-Pb	/56-FAM/CCTTTTAGC/ZEN/GTATTCGGGTTTATTTTTTATAGTATTTTTAACGATTT/3IABkFQ/	47	3754
Tag Forward Primer	AcDx-10256-SPOCK2-RT-FP	TTGTTCGCCCGTTGGTCAC	19	3755
Tag Reverse Primer	AcDx-10257-SPOCK2-RT-RP	TGCAACCTTAGCGCTCCAC	19	3756
Downstream PCR Primer	AcDx-10258-SPOCK2-PCR-V	TGCAACCTTAGCGCTCCACCAAAACGAACGAAAAATATAACTAAACCCTGrAAATT/3SpC3/	55	3757
HTRA3				
Forward PCR Primer	AcDx-10261-HTRA3-FP	GTTAGGCGAGGTTTTGTCrGGGAC/3SpC3/	23	3758
Reverse PCR Primer	AcDx-10262-HTRA3-RP	GGTGTCGTGGGCCCTCGCTAAAAAAAACGACrCCGAT/3SpC3/	36	3759
Upstream LDR	AcDx-10263-HTRA3-Up	TATCGCATCAAATGGAGAGCAAGGTTTTGTCGAGGTTTGAGCrGGGAC/3SpC3/	47	3760
Downstream LDR	AcDx-10264-HTRA3-Dn	/5Phos/GGAGTTGGCGGAGGAGTCGTTGACCGCTGTTATACGTTGCA	41	3761
Real-Time Probe	AcDx-10265-HTRA3-RT-Pb	/56-FAM/CCTTTGAGC/ZEN/GGAGTTGGCGGAGG/3IABkFQ/	23	3762
Tag Forward Primer	AcDx-10266-HTRA3-RT-FP	TATCGCATCAAATGGAGAGCAA	22	3763
Tag Reverse Primer	AcDx-10267-HTRA3-RT-RP	TGCAACGTATAACAGCGGTCAA	22	3764
Downstream PCR Primer	AcDx-10268-HTRA3-PCR-V	TGCAACGTATAACAGCGGTCAACCCTCGCTAAAAAAAACGACCTGrACCCT/3SpC3/	50	3765
CANCR94				
Forward PCR Primer	AcDx-10271-CANCR94-FP	GGGTGATAGTATTCGGTGGCrGTTAG/3SpC3/	25	3766
Reverse PCR Primer	AcDx-10272-CANCR94-RP	GGTGTCGTGGCGAACCTTTACGCTTTACGCrUCCGC/3SpC3/	35	3767
Upstream LDR	AcDx-10273-CANCR94-Up	TCCTAGTACCTACAGTGGGCAAGGTGGCGTTAAGGGATTTAGCrGGGAA/3SpC3/	48	3768
Downstream LDR	AcDx-10274-CANCR94-Dn	/5Phos/GGGAGTTAGGTCGGAGGGAGGGTTGACCGCTGTTATACGTTGCA	44	3769
Real-Time Probe	AcDx-10275-CANCR94-RT-Pb	/56-FAM/TTATTTAGC/ZEN/GGGAGTTAGGTCGGAGGGAG/3IABkFQ/	29	3770
Tag Forward Primer	AcDx-10276-CANCR94-RT-FP	TCCTAGTACCTACAGTGGGCAA	22	3771
Tag Reverse Primer	AcDx-10277-CANCR94-RT-RP	TGCAACGTATAACAGCGGTCAA	22	3772
Downstream PCR	AcDx-10278-CANCR94-PCR-	TGCAACGTATAACAGCGGTCAAGAACCTTTACGCTTTACGCTCTGrUAACT/3SpC3/	50	3773

Primer	V			
CANCR95				
Forward PCR Primer	AcDx-10281-CANCR95-FP	CGCGAGTTTTTTTGGAGTTTCrGGTCA/3SpC3/	26	3774
Reverse PCR Primer	AcDx-10282-CANCR95-RP	GGTGTCGTGGAAACACCAAAAAAAACTAAAACGAAACrACTAG/3SpC3/	42	3775
Upstream LDR	AcDx-10283-CANCR95-Up	TAGTTTGTCGAAAGTCCCACACCGAGTTTTTTTGGAGTTTCGGTCGTCTCrGTTCG/3SpC3/	55	3776
Downstream LDR	AcDx-10284-CANCR95-Dn	/5Phos/GTTTAGTGGTTTTGTGTAGCGTTTTTCGTTCGGAGGTGCAAAATTCAGGCTGTGCA	56	3777
Real-Time Probe	AcDx-10285-CANCR95-RT-Pb	/56-FAM/TTTCGTCTC/ZEN/GTTTAGTGGTTTTGTGTAGCGTT/3IABkFQ/	32	3778
Tag Forward Primer	AcDx-10286-CANCR95-RT-FP	TAGTTTGTCGAAAGTCCCACAC	22	3779
Tag Reverse Primer	AcDx-10287-CANCR95-RT-RP	TGCACAGCCTGAATTTTGCAC	21	3780
Downstream PCR Primer	AcDx-10288-CANCR95-PCR-V	TGCACAGCCTGAATTTTGCACACTAAAACGAAACACTAAATTCTCCGAATGrAAAAG/3SpC3/	56	3781
TPM4				
Forward PCR Primer	AcDx-10291-TPM4-FP	CGCGTAGTTTTCGGGTCrGTTTC/3SpC3/	22	3782
Reverse PCR Primer	AcDx-10292-TPM4-RP	GGTGTCGTGGCCCAAAAAAAAAAACGACGAAAAAAATrAAAAG/3SpC3/	42	3783
Upstream LDR	AcDx-10293-TPM4-Up	TAGACACGAGCGAGGTCACCGGGTCGTTTTTTTACGTTTTCGAACrGTCAG/3SpC3/	50	3784
Downstream LDR	AcDx-10294-TPM4-Dn	/5Phos/GTCGATCGTCGATTTATATTTTTGCGTTCGTAGTTAGGGTGCAAAATTCAGGCTGTGCA	59	3785
Real-Time Probe	AcDx-10295-TPM4-RT-Pb	/56-FAM/AATTCGAAC/ZEN/GTCGATCGTCGATTTATATTTTTGCGTT/3IABkFQ/	37	3786
Tag Forward Primer	AcDx-10296-TPM4-RT-FP	TAGACACGAGCGAGGTCAC	19	3787
Tag Reverse Primer	AcDx-10297-TPM4-RT-RP	TGCACAGCCTGAATTTTGCAC	21	3788
Downstream PCR Primer	AcDx-10298-TPM4-PCR-V	TGCACAGCCTGAATTTTGCACAAAAACGACGAAAAAAATAAAAAATCCTAACTATGrAACGT/3SpC3/	61	3789

CANCR96				
Forward PCR Primer	AcDx-10301-CANCR96-FP	TTTGGCGGGCGAGTATCrGTTTC/3SpC3/	22	3790
Reverse PCR Primer	AcDx-10302-CANCR96-RP	GGTGTCGTGGTCTATTTCTCGTTCGCGATCTAAAArUCGCT/3SpC3/	40	3791
Upstream LDR	AcDx-10303-CANCR96-Up	TTCTTCACAGTACCGCCACAGCGGGCGAGTATCGTTTTTATATAAATTATTGGCrGTTGA/3SpC3/	59	3792
Downstream LDR	AcDx-10304-CANCR96-Dn	/5Phos/GTTAGAGTTTCGGGAAAAGGCGTATTTTTTATTGTTGGTGTGTTGTCTGGTGGTGCA	57	3793
Real-Time Probe	AcDx-10305-CANCR96-RT-Pb	/56-FAM/CCTATTGGC/ZEN/GTTAGAGTTTCGGGAAAAGGCG/3IABkFQ/	31	3794
Tag Forward Primer	AcDx-10306-CANCR96-RT-FP	TTCTTCACAGTACCGCCACA	20	3795
Tag Reverse Primer	AcDx-10307-CANCR96-RT-RP	TGCACCACCAGACAACACA	19	3796
Downstream PCR Primer	AcDx-10308-CANCR96-PCR-V	TGCACCACCAGACAACACATCTATTTCTCGTTCGCGATCTAAAATTGrCCCTG/3SpC3/	52	3797
PFKP				
Forward PCR Primer	AcDx-10311-PFKP-FP	GGTCGAGGAAGTCGAGTTTCrGAGGC/3SpC3/	25	3798
Reverse PCR Primer	AcDx-10312-PFKP-RP	GGTGTCGTGGCCGTTATAAAAACCGAACCCGArCCGCA/3SpC3/	37	3799
Upstream LDR	AcDx-10313-PFKP-Up	TACCCTCCTAGCTCCGTACAGAGGAAGTCGAGTTTCGAGGTTTTTGGCrGTCAC/3SpC3/	53	3800
Downstream LDR	AcDx-10314-PFKP-Dn	/5Phos/GTCGTTTTAGGAGGAATTTTGCGGGTTATTAGAAGTTGTGTGTTGTCTGGTGGTGCA	57	3801
Real-Time Probe	AcDx-10315-PFKP-RT-Pb	/56-FAM/AATTTTGGC/ZEN/GTCGTTTTAGGAGGAATTTTGCG/3IABkFQ/	32	3802
Tag Forward Primer	AcDx-10316-PFKP-RT-FP	TACCCTCCTAGCTCCGTACA	20	3803
Tag Reverse Primer	AcDx-10317-PFKP-RT-RP	TGCACCACCAGACAACACA	19	3804
Downstream PCR Primer	AcDx-10318-PFKP-PCR-V	TGCACCACCAGACAACACACCGTTATAAAAACCGAACCCGACTGrCGCGT/3SpC3/	49	3805
FUT2				
Forward PCR Primer	AcDx-10321-FUT2-FP	TAGTGAGGTGCGATTGAGTCrGGATA/3SpC3/	25	3806
Reverse PCR Primer	AcDx-10322-FUT2-RP	GGTGTCGTGGCTCCGCTCACCCGAAACrCCCGC/3SpC3/	32	3807

Upstream LDR	AcDx-10323-FUT2-Up	TCATCGCCCTCAGATCTTCCAGATGTGTTGAGTTGTTCGGGATACrGGGAT/3SpC3/	50	3808
Downstream LDR	AcDx-10324-FUT2-Dn	/5Phos/GGAGCGGTTGGAGTAGAGAAAGGTTTTTAGTGGAGGATAGATTGGAGGGCA	51	3809
Real-Time Probe	AcDx-10325-FUT2-RT-Pb	/56-FAM/AAGGGACAC/ZEN/GGAGCGGTTGGAGT/3IABkFQ/	23	3810
Tag Forward Primer	AcDx-10326-FUT2-RT-FP	TCATCGCCCTCAGATCTTCCA	21	3811
Tag Reverse Primer	AcDx-10327-FUT2-RT-RP	TGCCCTCCAATCTATCCTCCA	21	3812
Downstream PCR Primer	AcDx-10328-FUT2-PCR-V	TGCCCTCCAATCTATCCTCCACGCTCACCCGAAACCCTGrUCCTG/3SpC3/	44	3813
KLF16				
Forward PCR Primer	AcDx-10331-KLF16-FP	GTGGGTGAGGATAACGCrGGGAA/3SpC3/	22	3814
Reverse PCR Primer	AcDx-10332-KLF16-RP	GGTGTCGTGGCTACCTATTTCCGAAACTATCCTAArACTCG/3SpC3/	40	3815
Upstream LDR	AcDx-10333-KLF16-Up	TAACCAGTTACCACCGCCACGGGAGGGTAGTGTGAGCrGGGAC/3SpC3/	42	3816
Downstream LDR	AcDx-10334-KLF16-Dn	/5Phos/GGAGTCGCGGTGGATTTGATTTTTTGATTTAGGTGGAGGATAGATTGGAGGGCA	54	3817
Real-Time Probe	AcDx-10335-KLF16-RT-Pb	/56-FAM/CCTGTGAGC/ZEN/GGAGTCGCGGT/3IABkFQ/	20	3818
Tag Forward Primer	AcDx-10336-KLF16-RT-FP	TAACCAGTTACCACCGCCA	19	3819
Tag Reverse Primer	AcDx-10337-KLF16-RT-RP	TGCCCTCCAATCTATCCTCCA	21	3820
Downstream PCR Primer	AcDx-10338-KLF16-PCR-V	TGCCCTCCAATCTATCCTCCACTATTTCCGAAACTATCCTAAACTCACCTGrAACCT/3SpC3/	56	3821
RHCG				
Forward PCR Primer	AcDx-10341-RHCG-FP	GGTAGCGGGCGGTTCrGGACA/3SpC3/	20	3822
Reverse PCR Primer	AcDx-10342-RHCG-RP	GGTGTCGTGGGTACCCCACGTAACCGCrCCCGT/3SpC3/	32	3823
Upstream LDR	AcDx-10343-RHCG-Up	TCGCACCGGAATTCTGACCCGGGCGGTTCGGACGCTCrGGAAA/3SpC3/	42	3824
Downstream LDR	AcDx-10344-RHCG-Dn	/5Phos/GGAGGTCGGCGAGGTTTTTGAGATTCGGTAGTTTCCCATGACGGCA	46	3825
Real-Time Probe	AcDx-10345-RHCG-RT-Pb	/56-FAM/TTGACGCTC/ZEN/GGAGGTCGGCGG/3IABkFQ/	21	3826
Tag Forward Primer	AcDx-10346-RHCG-RT-FP	TCGCACCGGAATTCTGACC	19	3827
Tag Reverse Primer	AcDx-10347-RHCG-RT-RP	TGCCGTCATGGGAAACTACC	20	3828
Downstream PCR Primer	AcDx-10348-RHCG-PCR-V	TGCCGTCATGGGAAACTACCCCCACGTAACCGCCCTGrCCCCT/3SpC3/	42	3829

TMEM106A-S2				
Forward PCR Primer	AcDx-10351-TMEM106A-S2-FP	GTTTGAGAGGAGTGTTCGTTTTCrGTATC/3SpC3/	28	3830
Reverse PCR Primer	AcDx-10352-TMEM106A-S2-RP	GGTGTCGTGGATCTAAAAAAAACCTTACCCGCGArACGAT/3SpC3/	39	3831
Upstream LDR	AcDx-10353-TMEM106A-S2-Up	TTGCACGTTGTCCTGCACCGTTTGAGAGGAGTGTTCGTTTTCGTATTCTCrGATCC/3SpC3/	55	3832
Downstream LDR	AcDx-10354-TMEM106A-S2-Dn	/5Phos/GATTTTATTTTTTTTCGTTTTTGTCGTTGTAGTCGTGGTAGTTTCCCATGACGGCA	56	3833
Real-Time Probe	AcDx-10355-TMEM106A-S2-RT-Pb	/56-FAM/TTTATTCTC/ZEN/GATTTTATTTTTTTTCGTTTTTGTCGTTGTAGT/3IABkFQ/	42	3834
Tag Forward Primer	AcDx-10356-TMEM106A-S2-RT-FP	TTGCACGTTGTCCTGCACC	19	3835
Tag Reverse Primer	AcDx-10357-TMEM106A-S2-RT-RP	TGCCGTCATGGGAAACTACC	20	3836
Downstream PCR Primer	AcDx-10358-TMEM106A-S2-PCR-V	TGCCGTCATGGGAAACTACCATCTAAAAAAAACCTTACCCGCGAATGrACTAT/3SpC3/	52	3837
NEU1				
Forward PCR Primer	AcDx-10361-NEU1-FP	GGTTTTTTATTGAGTCGTAATAGAGATCrGGTTG/3SpC3/	33	3838
Reverse PCR Primer	AcDx-10362-NEU1-RP	GGTGTCGTGGACTTAAATTCCAAATCCCGCAAArCAACC/3SpC3/	38	3839
Upstream LDR	AcDx-10363-NEU1-Up	TGCTTACCCACGATGCACCATTGAGTCGTAATAGAGATCGGTTAGGGCrGGACC/3SpC3/	53	3840
Downstream LDR	AcDx-10364-NEU1-Dn	/5Phos/GGATTTTGGAGTTAGGAAATCGAGGATTGGGGTCGTATGACTTGCTCGCA	50	3841
Real-Time Probe	AcDx-10365-NEU1-RT-Pb	/56-FAM/TTTTAGGGC/ZEN/GGATTTTGAGGTTAGGAAATCG/3IABkFQ/	31	3842
Tag Forward Primer	AcDx-10366-NEU1-RT-FP	TGCTTACCCACGATGCACC	19	3843
Tag Reverse Primer	AcDx-10367-NEU1-RT-RP	TGCGAGCAAGTCATACGACC	20	3844

SEPT9-S3				
Forward PCR Primer	AcDx-10371-SEPT9-S3-FP	TTTATCGAGTTTTTTTTTTTAGGTAGTCrGAGGC/3SpC3/	33	3845
Reverse PCR Primer	AcDx-10372-SEPT9-S3-RP	GGTGTCGTGGCATAACTCAAAACCGAACAAAAAATCrAAAAG/3SpC3/	41	3846
Upstream LDR	AcDx-10373-SEPT9-S3-Up	TAGCCGATGGCGTAAAACCTTAGGTAGTCGAGGTTTTTTTTTATTTTTGTCTCrGCGCC/3SpC3/	58	3847
Downstream LDR	AcDx-10374-SEPT9-S3-Dn	/5Phos/GCGTTTTGGGAGGTTTTTTGTTTCGCGATTATAAAGGGTCGTATGACTTGCTCGCA	56	3848
Real-Time Probe	AcDx-10375-SEPT9-S3-RT-Pb	/56-FAM/CCTTGTCTC/ZEN/GCGTTTTGGGAGGTTTTTTGTTTCG/3IABkFQ/	34	3849
Tag Forward Primer	AcDx-10376-SEPT9-S3-RT-FP	TAGCCGATGGCGTAAAACC	19	3850
Tag Reverse Primer	AcDx-10377-SEPT9-S3-RT-RP	TGCGAGCAAGTCATACGACC	20	3851
Downstream PCR Primer	AcDx-10378-SEPT9-S3-PCR-V	TGCGAGCAAGTCATACGACCCAAAACCGAACAAAAAATCAAAAAAACTTTATAATTGrCGAAG/3SpC3/	62	3852
AIM1-S1				
Forward PCR Primer	AcDx-10381-AIM1-S1-FP	TTTATTGGTTTGTTTTGCGTTTTTTTTCrGTTTC/3SpC3/	33	3853
Reverse PCR Primer	AcDx-10382-AIM1-S1-RP	GGTGTCGTGGACGCTCCCAAAAAAAAAAAAAAACCrUAAAT/3SpC3/	39	3854
Upstream LDR	AcDx-10383-AIM1-S1-Up	TTACAGGCCGCATAGCAACGTTTGTTTTGCGTTTTTTTTCGTTTTTTAGCrGCGAA/3SpC3/	55	3855
Downstream LDR	AcDx-10384-AIM1-S1-Dn	/5Phos/GCGGGTATTGGATGTTAGTCGTTTTGTTTTAGGAGGTTGAGACATGGGCTCGCA	54	3856
Real-Time Probe	AcDx-10385-AIM1-S1-RT-Pb	/56-FAM/AATTTTAGC/ZEN/GCGGGTATTGGATGTTAGTCGTT/3IABkFQ/	32	3857
Tag Forward Primer	AcDx-10386-AIM1-S1-RT-FP	TTACAGGCCGCATAGCAAC	19	3858
Tag Reverse Primer	AcDx-10387-AIM1-S1-RT-RP	TGCGAGCCCATGTCTCAAC	19	3859
CANCR97				
Forward PCR Primer	AcDx-10391-CANCR97-FP	GGATTTATTGACGCGGGCrGTATG/3SpC3/	23	3860
Reverse PCR Primer	AcDx-10392-CANCR97-RP	GGTGTCGTGGATTTCGATAATTTCTTCAAAAAAAAATArCCACT/3SpC3/	43	3861
Upstream LDR	AcDx-10393-CANCR97-Up	TTCGCCTACCGCAGTGAACCGCGGGCGTATAGAGATTAGGTGCTCrGGGCC/3SpC3/	50	3862
Downstream LDR	AcDx-10394-CANCR97-Dn	/5Phos/GGGTTTTACGGTTTTTTATTTTTTGTTTTTATTAAAATTGTAGGTGGGTTGAGACATGGGCTCGCA	66	3863

Real-Time Probe	AcDx-10395-CANCR97-RT-Pb	/56-FAM/CCGGTGCTC/ZEN/GGGTTTTACGGTTTTTTATTTTTTGTTTTTATTAAAATTG/3IABkFQ/	49	3864
Tag Forward Primer	AcDx-10396-CANCR97-RT-FP	TTCGCCTACCGCAGTGAAC	19	3865
Tag Reverse Primer	AcDx-10397-CANCR97-RT-RP	TGCGAGCCCATGTCTCAAC	19	3866
CANCR98				
Forward PCR Primer	AcDx-10401-CANCR98-FP	TTTTTAAATGTTAGAAAATTGTGTTATTTATTCrGTTAC/3SpC3/	38	3867
Reverse PCR Primer	AcDx-10402-CANCR98-RP	GGTGTCGTGGCGCCCCCTCCTCCTACrUACCT/3SpC3/	31	3868
Upstream LDR	AcDx-10403-CANCR98-Up	TTCGCCTACCGCAGTGAACGAAAATTGTGTTATTTATTCGTTATGTTGAGTATTGTTATACTCrGGAAG/3SpC3/	68	3869
Downstream LDR	AcDx-10404-CANCR98-Dn	/5Phos/GGAGATTTCGGAGATTTTCGGGTAGGTTTTAATGGTTGAGACATGGGCTCGCA	53	3870
Real-Time Probe	AcDx-10405-CANCR98-RT-Pb	/56-FAM/AATATACTC/ZEN/GGAGATTTCGGAGATTTTCGGGTAGG/3IABkFQ/	35	3871
Tag Forward Primer	AcDx-10406-CANCR98-RT-FP	TTCGCCTACCGCAGTGAAC	19	3872
Tag Reverse Primer	AcDx-10407-CANCR98-RT-RP	TGCGAGCCCATGTCTCAAC	19	3873
Downstream PCR Primer	AcDx-10408-CANCR98-PCR-V	TGCGAGCCCATGTCTCAACCCCTCCTCCTACTACCCGTGrACCCT/3SpC3/	44	3874
CANCR99				
Forward PCR Primer	AcDx-10411-CANCR99-FP	GTTTTCGGGCGTTTCGTTTCrGTTTC/3SpC3/	25	3875
Reverse PCR Primer	AcDx-10412-CANCR99-RP	GGTGTCGTGGCCCCGACGACCGACCrUCCGG/3SpC3/	30	3876
Upstream LDR	AcDx-10413-CANCR99-Up	TTGCATTTCGTTAGCGACACATGTTTTTTTATTTTGATTTTCGGGTTTTTGTTCTCrGATCC/3SpC3/	61	3877
Downstream LDR	AcDx-10414-CANCR99-Dn	/5Phos/GATTTTAGTTTTTTAGGAGGCGGTTTTAGTTTAGTTAGGGATGTGAGTCGATCTACCCGCA	61	3878
Real-Time Probe	AcDx-10415-CANCR99-RT-	/56-FAM/AATGTTCTC/ZEN/GATTTTAGTTTTTTAGGAGGCGGTTTTAGTT/3IABkFQ/	40	3879

	Pb			
Tag Forward Primer	AcDx-10416-CANCR99-RT-FP	TTGCATTTCGTTAGCGACACA	21	3880
Tag Reverse Primer	AcDx-10417-CANCR99-RT-RP	TGCGGGTAGATCGACTCACA	20	3881
Downstream PCR Primer	AcDx-10418-CANCR99-PCR-V	TGCGGGTAGATCGACTCACACCGACGACCGACCTCTGrAAAAG/3SpC3/	42	3882
HOXC4-S2				
Forward PCR Primer	AcDx-10421-HOXC4-S2-FP	AATTATAGATCGATTAGGTAAAGGTAAArGGGAC/3SpC3/	33	3883
Reverse PCR Primer	AcDx-10422-HOXC4-S2-RP	GGTGTCGTGGAAAATACCTCACTAAATCATAAACGArUAAAG/3SpC3/	41	3884
Upstream LDR	AcDx-10423-HOXC4-S2-Up	TTGCATTTCGTTAGCGACACAATGAATTTTTATTTTATTAATTTTTTTTTATTTTGTTATTTCATCrGGGAA/3SpC3/	71	3885
Downstream LDR	AcDx-10424-HOXC4-S2-Dn	/5Phos/GGGAGTTAGGACGTTTTTTTTAACGTCGTTTTTAATTTTATGTGAGTCGATCTACCCGCA	60	3886
Real-Time Probe	AcDx-10425-HOXC4-S2-RT-Pb	/56-FAM/CCTTTCATC/ZEN/GGGAGTTAGGACGTTTTTTTTAACG/3IABkFQ/	34	3887
Tag Forward Primer	AcDx-10426-HOXC4-S2-RT-FP	TTGCATTTCGTTAGCGACACA	21	3888
Tag Reverse Primer	AcDx-10427-HOXC4-S2-RT-RP	TGCGGGTAGATCGACTCACA	20	3889
Downstream PCR Primer	AcDx-10428-HOXC4-S2-PCR-V	TGCGGGTAGATCGACTCACAAAAATACCTCACTAAATCATAAACGATAAAATTAAAAATGrACGTC/3SpC3/	65	3890
LDLR				
Forward PCR Primer	AcDx-10431-LDLR-FP	GCGTTTTTAATTTTGAGGAGGCrGTTAA/3SpC3/	27	3891
Reverse PCR Primer	AcDx-10432-LDLR-RP	GGTGTCGTGGCTAATTCCCACTCCAATCCTTCGArAAATG/3SpC3/	39	3892
Upstream LDR	AcDx-10433-LDLR-Up	TACCACTCATCTTCTGCGACACGTTTTTAATTTTGAGGAGGCGTTAGTTTTTTGTCrGGAAG/3SpC3/	61	3893
Downstream LDR	AcDx-10434-LDLR-Dn	/5Phos/GGAGATTTAAATATAATAAATTAAGTCGTTTGTTTTGGCGATATTTTGTGAGTCGATCTACCCGCA	66	3894
Real-Time Probe	AcDx-10435-LDLR-RT-Pb	/56-	47	3895

		FAM/AATTTTGTC/ZEN/GGAGATTTAAATATAATAAATTAAGTCGTTTGTTTTGG/3IABkFQ/		
Tag Forward Primer	AcDx-10436-LDLR-RT-FP	TACCACTCATCTTCTGCGACA	21	3896
Tag Reverse Primer	AcDx-10437-LDLR-RT-RP	TGCGGGTAGATCGACTCACA	20	3897
Downstream PCR Primer	AcDx-10438-LDLR-PCR-V	TGCGGGTAGATCGACTCACACTAATTCCCACTCCAATCCTTCGAAAATATTGrCCAAG/3SpC3/	57	3898
CANCR100				
Forward PCR Primer	AcDx-10441-CANCR100-FP	GGAGTCGAGGTCGGCrGTGGA/3SpC3/	20	3899
Reverse PCR Primer	AcDx-10442-CANCR100-RP	GGTGTCGTGGCCCCTCGTCGCGTATCrCCGAG/3SpC3/	31	3900
Upstream LDR	AcDx-10443-CANCR100-Up	TCTCATGGGCGCTAGTATCAACGCGTGAGGTGGGAGCrGTTGA/3SpC3/	42	3901
Downstream LDR	AcDx-10444-CANCR100-Dn	/5Phos/GTTAGGGAGGCGGGCGGGTTTCCCTGATTGATACCCGCA	39	3902
Real-Time Probe	AcDx-10445-CANCR100-RT-Pb	/56-FAM/TTTGGGAGC/ZEN/GTTAGGGAGGCG/3IABkFQ/	21	3903
Tag Forward Primer	AcDx-10446-CANCR100-RT-FP	TCTCATGGGCGCTAGTATCAAC	22	3904
Tag Reverse Primer	AcDx-10447-CANCR100-RT-RP	TGCGGGTATCAATCAGGGAAAC	22	3905
Downstream PCR Primer	AcDx-10448-CANCR100-PCR-V	TGCGGGTATCAATCAGGGAAACCCCTCGTCGCGTATCCTGrAACTT/3SpC3/	45	3906
OXR1-S1				
Forward PCR Primer	AcDx-10451-OXR1-S1-FP	GGTTAGTTTTATTGGCGTTAGGCrGGGTA/3SpC3/	28	3907
Reverse PCR Primer	AcDx-10452-OXR1-S1-RP	GGTGTCGTGGATATATTCTTCCTAAAACCGATACGArAATCG/3SpC3/	41	3908
Upstream LDR	AcDx-10453-OXR1-S1-Up	TGGATCGAGACGGAATGCAACGCGGGTGAGGGAACGCrGTTAG/3SpC3/	42	3909
Downstream LDR	AcDx-10454-OXR1-S1-Dn	/5Phos/GTTGAGAGGATTCGTTTAGTTTGTTTAAAAGTTATTAGTTTTTTGATTGTTTCCCTGATTGATACCCGCA	70	3910
Real-Time Probe	AcDx-10455-OXR1-S1-RT-Pb	/56-FAM/TTGGAACGC/ZEN/GTTGAGAGGATTCGTTTAGTTTGTTTAAAAG/3IABkFQ/	40	3911
Tag Forward Primer	AcDx-10456-OXR1-S1-RT-FP	TGGATCGAGACGGAATGCAAC	21	3912
Tag Reverse Primer	AcDx-10457-OXR1-S1-RT-RP	TGCGGGTATCAATCAGGGAAAC	22	3913

VSX1				
Forward PCR Primer	AcDx-11141-VSX1-FP	GGTACGTTCGTTAGGAGTAGGTArGGGTA/3SpC3/	28	3914
Reverse PCR Primer	AcDx-11142-VSX1-RP	GGTGTCGTGGCCTACCGCTAAAACTCGACCrUCCTT/3SpC3/	35	3915
Upstream LDR	AcDx-11143-VSX1-Up	TGCCCTATCGAAAAGGACAACAAGGAGTAGGTAGGGTGTTCGGGCrGGTTA/3SpC3/	50	3916
Downstream LDR	AcDx-11144-VSX1-Dn	/5Phos/GGTCGTCGGCGGTTGCGTGTGTTTGCGGCTGTCTATGACA	40	3917
Real-Time Probe	AcDx-11145-VSX1-RT-Pb	/56-FAM/CATTCGGGC/ZEN/GGTCGTCGGC/3IABkFQ/	19	3918
Tag Forward Primer	AcDx-11146-VSX1-RT-FP	TGCCCTATCGAAAAGGACAACA	22	3919
Tag Reverse Primer	AcDx-11147-VSX1-RT-RP	TGTCATAGACAGCCGCAAACA	21	3920
Downstream PCR Primer	AcDx-11148-VSX1-PCR-V	TGTCATAGACAGCCGCAAACACGACCTCCTCTATAACTTCGACATGrCAACT/3SpC3/	51	3921
Alternate Group 4 Markers				
HLA-J-S2				
Forward PCR Primer	AcDx-10461-HLA-J-S2-FP	TTTCGTAGGTTGTTAGAGGTCrGGGTC/3SpC3/	26	3922
Reverse PCR Primer	AcDx-10462-HLA-J-S2-RP	GGTGTCGTGGCGTCGAACCCCAAATCGCrAACCG/3SpC3/	33	3923
Upstream LDR	AcDx-10463-HLA-J-S2-Up	TCCTGAGGGACAAATACACACCGGGTTAGGGTTCGGTGAGCrGGGAC/3SpC3/	46	3924
Downstream LDR	AcDx-10464-HLA-J-S2-Dn	/5Phos/GGAGTTGATCGCGGGAATTGGGTTAGGGTAGGTAAGGAAGTCACGCA	47	3925
Real-Time Probe	AcDx-10465-HLA-J-S2-RT-Pb	/56-FAM/AAGGTGAGC/ZEN/GGAGTTGATCGCG/3IABkFQ/	22	3926
Tag Forward Primer	AcDx-10466-HLA-J-S2-RT-FP	TCCTGAGGGACAAATACACACC	22	3927
Tag Reverse Primer	AcDx-10467-HLA-J-S2-RT-RP	TGCGTGACTTCCTTACCTACC	21	3928
MFI2				
Forward PCR Primer	AcDx-10471-MFI2-FP	CGGCGGTCGTCGTTCrGCGAC/3SpC3/	20	3929
Reverse PCR Primer	AcDx-10472-MFI2-RP	GGTGTCGTGGCCCGAAAAATCAAAACCCGArAATAG/3SpC3/	35	3930
Upstream LDR	AcDx-10473-MFI2-Up	TTGCAACAGGCTACCGACCCGGTCGTCGTTCGCGATTCTCrGCGGA/3SpC3/	45	3931
Downstream LDR	AcDx-10474-MFI2-Dn	/5Phos/GCGAGTCGTTTTTCGAGGTTTCGTTTTTCGTGGTAGGTAAGGAAGTCACGCA	52	3932

Real-Time Probe	AcDx-10475-MFI2-RT-Pb	/56-FAM/TTGATTCTC/ZEN/GCGAGTCGTTTTTCGAGGTTTCG/3IABkFQ/	32	3933
Tag Forward Primer	AcDx-10476-MFI2-RT-FP	TTGCAACAGGCTACCGACC	19	3934
Tag Reverse Primer	AcDx-10477-MFI2-RT-RP	TGCGTGACTTCCTTACCTACC	21	3935
Downstream PCR Primer	AcDx-10478-MFI2-PCR-V	TGCGTGACTTCCTTACCTACCCCGAAAAATCAAAACCCGAAATAATAAAATGrAAAAG/3SpC3/	57	3936
HLA-J-S3				
Forward PCR Primer	AcDx-10481-HLA-J-S3-FP	GTTTTTTTTTTTTTTTTTTAGTTTGCGACrGGGTC/3SpC3/	35	3937
Reverse PCR Primer	AcDx-10482-HLA-J-S3-RP	GGTGTCGTGGACCGAAACCGCGCGArCGCTG/3SpC3/	30	3938
Upstream LDR	AcDx-10483-HLA-J-S3-Up	TAAGCCTGCTTTTCCGAAACAACGACGGGTTTTTTTTTTTGGATATTTACAACrGCGAG/3SpC3/	58	3939
Downstream LDR	AcDx-10484-HLA-J-S3-Dn	/5Phos/GCGGATTTAGTTTTTATTTTTATTGAGTGTCGGGTTTTTAGGTTGTATTGCGCCAGGATAGCA	63	3940
Real-Time Probe	AcDx-10485-HLA-J-S3-RT-Pb	/56-FAM/CCTTACAAC/ZEN/GCGGATTTAGTTTTTATTTTTATTGAGTGTCGG/3IABkFQ/	42	3941
Tag Forward Primer	AcDx-10486-HLA-J-S3-RT-FP	TAAGCCTGCTTTTCCGAAACAA	22	3942
Tag Reverse Primer	AcDx-10487-HLA-J-S3-RT-RP	TGCTATCCTGGCGCAATACAA	21	3943
Downstream PCR Primer	AcDx-10488-HLA-J-S3-PCR-V	TGCTATCCTGGCGCAATACAACCGAAACCGCGCGATGrCTAAC/3SpC3/	42	3944
ACSF2				
Forward PCR Primer	AcDx-10491-ACSF2-FP	GTTTTGGTTGTTTTTTTAATTTCGCrGGGAC/3SpC3/	30	3945
Reverse PCR Primer	AcDx-10492-ACSF2-RP	GGTGTCGTGGCCCCTCGAATCACGTACTArACCCT/3SpC3/	34	3946
Upstream LDR	AcDx-10493-ACSF2-Up	TTGGCAACTCTCCACCCAATTAGTTGGTATGTTTTAAGGACGTTTTGCrGTTCA/3SpC3/	53	3947
Downstream LDR	AcDx-10494-ACSF2-Dn	/5Phos/GTTTGTTTTTTTGGTTTTCGGTTCGTAAGTAGCGGGTTGTATTGCGCCAGGATAGCA	57	3948
Real-Time Probe	AcDx-10495-ACSF2-RT-Pb	/56-FAM/TTGTTTTGC/ZEN/GTTTGTTTTTTTGGTTTTCGGTTCGTAAGTAG/3IABkFQ/	41	3949
Tag Forward Primer	AcDx-10496-ACSF2-RT-FP	TTGGCAACTCTCCACCCAA	19	3950
Tag Reverse Primer	AcDx-10497-ACSF2-RT-RP	TGCTATCCTGGCGCAATACAA	21	3951
Downstream PCR Primer	AcDx-10498-ACSF2-PCR-V	TGCTATCCTGGCGCAATACAACCTCGAATCACGTACTAACCCTGrCTACC/3SpC3/	49	3952

HCG4P6				
Forward PCR Primer	AcDx-10501-HCG4P6-FP	TATTTCGAGGTTGCGGGATTCrGTTAA/3SpC3/	26	3953
Reverse PCR Primer	AcDx-10502-HCG4P6-RP	GGTGTCGTGGACCAACCCGCGAAAATTTTAACrCTAAG/3SpC3/	37	3954
Upstream LDR	AcDx-10503-HCG4P6-Up	TCGACGAATCTGCTCAGACAACGAGGTTGCGGGATTCGTTAGATTCTCrGATCT/3SpC3/	53	3955
Downstream LDR	AcDx-10504-HCG4P6-Dn	/5Phos/GATTCGGGAGAGGTTTAGGCGTTTTTATTAGGTTTTTTGAAGCAGCGTCTGAGCA	55	3956
Real-Time Probe	AcDx-10505-HCG4P6-RT-Pb	/56-FAM/CCGATTCTC/ZEN/GATTCGGGAGAGGTTTAGGCGTTTTTAT/3IABkFQ/	37	3957
Tag Forward Primer	AcDx-10506-HCG4P6-RT-FP	TCGACGAATCTGCTCAGACAA	21	3958
Tag Reverse Primer	AcDx-10507-HCG4P6-RT-RP	TGCTCAGACGCTGCTTCAA	19	3959
Downstream PCR Primer	AcDx-10508-HCG4P6-PCR-V	TGCTCAGACGCTGCTTCAACCAACCCGCGAAAATTTTAACCTAAATGrAAAAG/3SpC3/	52	3960
SOCS3				
Forward PCR Primer	AcDx-10511-SOCS3-FP	CGAAGGTTTTAGTCGGAGTATTTCrGGGAG/3SpC3/	29	3961
Reverse PCR Primer	AcDx-10512-SOCS3-RP	GGTGTCGTGGCCTCCCCTTCCCGAATCArUTCCT/3SpC3/	33	3962
Upstream LDR	AcDx-10513-SOCS3-Up	TCGACGAATCTGCTCAGACAATCGGAGTATTTCGGGAATTTGGGATTCTCrGGTCC/3SpC3/	55	3963
Downstream LDR	AcDx-10514-SOCS3-Dn	/5Phos/GGTTTTTATTCGTTTTGTTTTTATTTCGTTATTTTTTTGTCGGGTTGAAGCAGCGTCTGAGCA	63	3964
Real-Time Probe	AcDx-10515-SOCS3-RT-Pb	/56-FAM/TTGATTCTC/ZEN/GGTTTTTATTCGTTTTGTTTTTATTTCGTTATTTTTTTG/3IABkFQ/	48	3965
Tag Forward Primer	AcDx-10516-SOCS3-RT-FP	TCGACGAATCTGCTCAGACAA	21	3966
Tag Reverse Primer	AcDx-10517-SOCS3-RT-RP	TGCTCAGACGCTGCTTCAA	19	3967
Downstream PCR Primer	AcDx-10518-SOCS3-PCR-V	TGCTCAGACGCTGCTTCAACCCCTTCCCGAATCATTCCTGrACAAG/3SpC3/	45	3968
SERPINB9-S2				
Forward PCR Primer	AcDx-10521-SERPINB9-S2-	GGATTCGGTTTTGTTGTTGCrGTTCA/3SpC3/	25	3969

	FP			
Reverse PCR Primer	AcDx-10522-SERPINB9-S2-RP	GGTGTCGTGGACGTAAAAAACGCCCGACrCCTAT/3SpC3/	33	3970
Upstream LDR	AcDx-10523-SERPINB9-S2-Up	TTTCCGCCGCTACAACCAAGTTTTGTTGTTGCGTTCGTTGCrGGACC/3SpC3/	46	3971
Downstream LDR	AcDx-10524-SERPINB9-S2-Dn	/5Phos/GGATTTTCGTTAGGGTTTTTGTAGGTATCGTTTTTATATCGTAGTTGAAGCAGC GTCTGAGCA	63	3972
Real-Time Probe	AcDx-10525-SERPINB9-S2-RT-Pb	/56-FAM/AATCGTTGC/ZEN/GGATTTTCGTTAGGGTTTTTGTAGG/3IABkFQ/	34	3973
Tag Forward Primer	AcDx-10526-SERPINB9-S2-RT-FP	TTTCCGCCGCTACAACCAA	19	3974
Tag Reverse Primer	AcDx-10527-SERPINB9-S2-RT-RP	TGCTCAGACGCTGCTTCAA	19	3975
Downstream PCR Primer	AcDx-10528-SERPINB9-S2-PCR-V	TGCTCAGACGCTGCTTCAACGCCCGACCCTACGATATAAAAATGrATACT/3SpC3/	49	3976
CANCR101				
Forward PCR Primer	AcDx-10531-CANCR101-FP	TTAGTTCGTAGTAATTGCGTAGCrGGGAA/3SpC3/	28	3977
Reverse PCR Primer	AcDx-10532-CANCR101-RP	GGTGTCGTGGGCGTCCTCCTCGCAArCCTCC/3SpC3/	30	3978
Upstream LDR	AcDx-10533-CANCR101-Up	TCAGTGAAAACACATCCACCCAGTAGCGGAGGGAATTTAGGGTCTCrGCGAC/3SpC3/	51	3979
Downstream LDR	AcDx-10534-CANCR101-Dn	/5Phos/GCGGTATTCGTAGGTTTTAGTTATTGTTTCGAGGTTTTCGTGGTGAGCAGGGAT GAGCA	59	3980
Real-Time Probe	AcDx-10535-CANCR101-RT-Pb	/56-FAM/CCGGGTCTC/ZEN/GCGGTATTCGTAGGTTTTAGTTATTGTTT/3IABkFQ/	38	3981
Tag Forward Primer	AcDx-10536-CANCR101-RT-FP	TCAGTGAAAACACATCCACCCA	22	3982
Tag Reverse Primer	AcDx-10537-CANCR101-RT-RP	TGCTCATCCCTGCTCACCA	19	3983
Downstream PCR Primer	AcDx-10538-CANCR101-PCR-V	TGCTCATCCCTGCTCACCACAACCTCTCTTTAATCCGAAAACCTTGrAAACG/3SpC3/	51	3984
CANCR102				

Forward PCR Primer	AcDx-10541-CANCR102-FP	TTTATTATTTTTCGTTTATTTTTGTTTTTTTCrGGGTA/3SpC3/	37	3985
Reverse PCR Primer	AcDx-10542-CANCR102-RP	GGTGTCGTGGAAACTCATCCAAAAACGCCTTAArAAACC/3SpC3/	38	3986
Upstream LDR	AcDx-10543-CANCR102-Up	TCGTAGACTCGCTATCGCCACGTTTATTTTTGTTTTTTTCGGGTGGGACACrGTGGA/3SpC3/	56	3987
Downstream LDR	AcDx-10544-CANCR102-Dn	/5Phos/GTGAGTTGTTTTGTTGTTCGGATGGAGGTTTTAGGTGGTGAGCAGGGATGAGCA	54	3988
Real-Time Probe	AcDx-10545-CANCR102-RT-Pb	/56-FAM/AAGGGACAC/ZEN/GTGAGTTGTTTTGTTGTTCGGATG/3IABkFQ/	33	3989
Tag Forward Primer	AcDx-10546-CANCR102-RT-FP	TCGTAGACTCGCTATCGCCA	20	3990
Tag Reverse Primer	AcDx-10547-CANCR102-RT-RP	TGCTCATCCCTGCTCACCA	19	3991
Downstream PCR Primer	AcDx-10548-CANCR102-PCR-V	TGCTCATCCCTGCTCACCACTCATCCAAAAACGCCTTAAAAACTATGrAAACT/3SpC3/	52	3992
HLA-J-S4				
Forward PCR Primer	AcDx-10551-HLA-J-S4-FP	CGGGTCGATAGTGACGTCrGTGAA/3SpC3/	23	3993
Reverse PCR Primer	AcDx-10552-HLA-J-S4-RP	GGTGTCGTGGCCTAAACCTTAACGCCCAArUATCC/3SpC3/	34	3994
Upstream LDR	AcDx-10553-HLA-J-S4-Up	TTGCACCCGCGACATAACCGTCGTGAGTTTGAGGATGAAGATGCrGGGTA/3SpC3/	49	3995
Downstream LDR	AcDx-10554-HLA-J-S4-Dn	/5Phos/GGGCGCGGTGGGTGGAGTAGGGTAATTCACTCGAACGGAGCA	42	3996
Real-Time Probe	AcDx-10555-HLA-J-S4-RT-Pb	/56-FAM/AAAAGATGC/ZEN/GGGCGCGGTGG/3IABkFQ/	20	3997
Tag Forward Primer	AcDx-10556-HLA-J-S4-RT-FP	TTGCACCCGCGACATAACC	19	3998
Tag Reverse Primer	AcDx-10557-HLA-J-S4-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	3999
Downstream PCR Primer	AcDx-10558-HLA-J-S4-PCR-V	TGCTCCGTTCGAGTGAATTACCCGCCCAATATCTATAAATCCCAATACTCTGrACCCT/3SpC3/	57	4000
SEPT9-S4				
Forward PCR Primer	AcDx-10561-SEPT9-S4-FP	CGAGGTTTTTTTTTATTTTTGTTTCGCrGTTTC/3SpC3/	32	4001
Reverse PCR Primer	AcDx-10562-SEPT9-S4-RP	GGTGTCGTGGAAAACCGCGACCCGCrCCACT/3SpC3/	30	4002
Upstream LDR	AcDx-10563-SEPT9-S4-Up	TCCGGCCTTTGACGATACCCGTTTTGGGAGGTTTTTTGTTTTGCrGATCG/3SpC3/	49	4003
Downstream LDR	AcDx-10564-SEPT9-S4-Dn	/5Phos/GATTATAAAGTTTTTTTGATTTTTTGTTCGGTTTTGAGTTATGTGATTCGGGTAA	72	4004

		TTCACTCGAACGGAGCA		
Real-Time Probe	AcDx-10565-SEPT9-S4-RT-Pb	/56-FAM/AAGTTTTGC/ZEN/GATTATAAAGTTTTTTTGATTTTTTGTTCGGTTTTGAG/3IABkFQ/	47	4005
Tag Forward Primer	AcDx-10566-SEPT9-S4-RT-FP	TCCGGCCTTTGACGATACC	19	4006
Tag Reverse Primer	AcDx-10567-SEPT9-S4-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	4007
Downstream PCR Primer	AcDx-10568-SEPT9-S4-PCR-V	TGCTCCGTTCGAGTGAATTACCGCGACCCGCCCACTGrAATCG/3SpC3/	42	4008
AIM1-S2				
Forward PCR Primer	AcDx-10571-AIM1-S2-FP	TTTTTTTTGGGAGCGTTGCrGGATC/3SpC3/	24	4009
Reverse PCR Primer	AcDx-10572-AIM1-S2-RP	GGTGTCGTGGGCACGACTAAAACCCCAAArAACCA/3SpC3/	34	4010
Upstream LDR	AcDx-10573-AIM1-S2-Up	TCCGGCCTTTGACGATACCTTTTGGGAGCGTTGCGGATTACTCrGTAAA/3SpC3/	48	4011
Downstream LDR	AcDx-10574-AIM1-S2-Dn	/5Phos/GTAGGCGGTTCGGTATTTTCGTATAGGTGGGGGTAATTCACTCGAACGGAGCA	53	4012
Real-Time Probe	AcDx-10575-AIM1-S2-RT-Pb	/56-FAM/TTATTACTC/ZEN/GTAGGCGGTTCGGTATTTTCGTATAGG/3IABkFQ/	36	4013
Tag Forward Primer	AcDx-10576-AIM1-S2-RT-FP	TCCGGCCTTTGACGATACC	19	4014
Tag Reverse Primer	AcDx-10577-AIM1-S2-RT-RP	TGCTCCGTTCGAGTGAATTACC	22	4015
Downstream PCR Primer	AcDx-10578-AIM1-S2-PCR-V	TGCTCCGTTCGAGTGAATTACCGCACGACTAAAACCCCAAAAACTGrACCCG/3SpC3/	51	4016
AIM1-S3				
Forward PCR Primer	AcDx-10581-AIM1-S3-FP	TCGTGCGATTTCGGAATTATACrGTGTA/3SpC3/	27	4017
Reverse PCR Primer	AcDx-10582-AIM1-S3-RP	GGTGTCGTGGGCCCCCTAAAACCGAAAAAArATATG/3SpC3/	34	4018
Upstream LDR	AcDx-10583-AIM1-S3-Up	TCGCGGAAAGTCCCAGTAACCGGAATTATACGTGTGGCGAGGGACrGGGCC/3SpC3/	50	4019
Downstream LDR	AcDx-10584-AIM1-S3-Dn	/5Phos/GGGTTTCGGAGCGGAGTAGGTAGAGGTTGGCCTGTAAGCGTTCCA	45	4020
Real-Time Probe	AcDx-10585-AIM1-S3-RT-Pb	/56-FAM/TTGAGGGAC/ZEN/GGGTTTCGGAGCG/3IABkFQ/	22	4021
Tag Forward Primer	AcDx-10586-AIM1-S3-RT-FP	TCGCGGAAAGTCCCAGTAAC	20	4022
Tag Reverse Primer	AcDx-10587-AIM1-S3-RT-RP	TGGAACGCTTACAGGCCAAC	20	4023
Downstream PCR Primer	AcDx-10588-AIM1-S3-PCR-V	TGGAACGCTTACAGGCCAACCCCCCTAAAACCGAAAAAATATAACTAAAAAATATGrCTTAG/3SpC3/	61	4024

SLC5A10				
Forward PCR Primer	AcDx-10591-SLC5A10-FP	CGTGTCGTTCGAGGCrGGGAC/3SpC3/	20	4025
Reverse PCR Primer	AcDx-10592-SLC5A10-RP	GGTGTCGTGGCTAATATTTTCGAAATTCGAACACCArAAAAG/3SpC3/	41	4026
Upstream LDR	AcDx-10593-SLC5A10-Up	TGAACGCTCAAACACGTGAACGCGAGGTCGGAAGTTGATTTATTTCTCrGATGA/3SpC3/	53	4027
Downstream LDR	AcDx-10594-SLC5A10-Dn	/5Phos/GATAGATAGATAGATTTGGAGCGGACGTATTGTTTACGGTTGGCCTGTAAGCG TTCCA	58	4028
Real-Time Probe	AcDx-10595-SLC5A10-RT-Pb	/56-FAM/AAATTTCTC/ZEN/GATAGATAGATAGATTTGGAGCGGACGTATTG/3IABkFQ/	41	4029
Tag Forward Primer	AcDx-10596-SLC5A10-RT-FP	TGAACGCTCAAACACGTGAAC	21	4030
Tag Reverse Primer	AcDx-10597-SLC5A10-RT-RP	TGGAACGCTTACAGGCCAAC	20	4031
Downstream PCR Primer	AcDx-10598-SLC5A10-PCR-V	TGGAACGCTTACAGGCCAACCTAATATTTTCGAAATTCGAACACCAAAAAACTGrUAAAT /3SpC3/	59	4032
OXR1-S2				
Forward PCR Primer	AcDx-10601-OXR1-S2-FP	TTATTTTTTTTTTTGTAGTTTTGAAAGTTCrGGTTT/3SpC3/	35	4033
Reverse PCR Primer	AcDx-10602-OXR1-S2-RP	GGTGTCGTGGCTAACCTAACCCGCCCAArCCCCC/3SpC3/	33	4034
Upstream LDR	AcDx-10603-OXR1-S2-Up	TACACGTGGATATCTCCGACCTGTAGTTTTGAAAGTTCGGTTCGGGCrGCGAG/3SpC3/	52	4035
Downstream LDR	AcDx-10604-OXR1-S2-Dn	/5Phos/GCGGACGGGACGGAGAGGGTGCTAGTCACACAGTTCCA	38	4036
Real-Time Probe	AcDx-10605-OXR1-S2-RT-Pb	/56-FAM/TTTTCGGGC/ZEN/GCGGACGGG/3IABkFQ/	18	4037
Tag Forward Primer	AcDx-10606-OXR1-S2-RT-FP	TACACGTGGATATCTCCGACC	21	4038
Tag Reverse Primer	AcDx-10607-OXR1-S2-RT-RP	TGGAACTGTGTGACTAGCACC	21	4039
Downstream PCR Primer	AcDx-10608-OXR1-S2-PCR-V	TGGAACTGTGTGACTAGCACCGCCCAACCCCTCCCTGrCCCCA/3SpC3/	42	4040
PTPRU-S3				
Forward PCR Primer	AcDx-10611-PTPRU-S3-FP	CGGTTTTAGGAATTTATGTTCGCrGCGGC/3SpC3/	28	4041
Reverse PCR Primer	AcDx-10612-PTPRU-S3-RP	GGTGTCGTGGCAAAAACGCCTCGAAACGAAArAATCT/3SpC3/	36	4042
Upstream LDR	AcDx-10613-PTPRU-S3-Up	TAGCATTCGAGAACGCACCCGCGCGGTGTAGTAGGAACrGGTCG/3SpC3/	43	4043
Downstream LDR	AcDx-10614-PTPRU-S3-Dn	/5Phos/GGTTAAATGGAGTTCGGGTTAGAGCGAGATCGGTGCTAGTCACACAGTTCCA	52	4044

Real-Time Probe	AcDx-10615-PTPRU-S3-RT-Pb	/56-FAM/TTTAGGAAC/ZEN/GGTTAAATGGAGTTCGGGTTAGAG/3IABkFQ/	33	4045
Tag Forward Primer	AcDx-10616-PTPRU-S3-RT-FP	TAGCATTCGAGAACGCACC	19	4046
Tag Reverse Primer	AcDx-10617-PTPRU-S3-RT-RP	TGGAACTGTGTGACTAGCACC	21	4047
Downstream PCR Primer	AcDx-10618-PTPRU-S3-PCR-V	TGGAACTGTGTGACTAGCACCCTCGAAACGAAAAATCCCGATCTTGrCTCTG/3SpC3/	51	4048

IDT Abbreviation	Modifications
/5Phos/	5' Phosphorylation
rX (X=A,C,G,U)	RNA Base
/3spC3/	3' C3 DNA Spacer
/56-FAM/	5' 6-FAM™ Fluorescent Tag
/Zen/	Internal Quencher
/3IABkFQ/	3' Iowa Black® FQ Quencher

Table 57. Primers for use in Step 2 of the Group 5- 64-marker assay, with average sensitivities of 50%, to detect and identify liver, pancreatic, and gall-bladder cancers.

Site	Primer Name	Sequence	Length	Seq. ID No.
Prefered Group 5 Markers				
CaNCR25				
Forward PCR Primer	AcDx-7221-CaNCR25-FP	GTTTTTTTTAGTTTGGAGTTTTGGTTCrGGGTC/3SpC3/	32	4049
Reverse PCR Primer	AcDx-7222-CaNCR25-RP	GGTGTCGTGGAAACAAAAAAACCCAAAAACAACGrCCCGT/3SpC3/	39	4050
Upstream LDR	AcDx-7223-CaNCR25-	TATCTCCTAAAAGAAGCCGCACTAGTTTGGAGTTTTGGTTCGGGTTCTCrGGGCC/3SpC3/	54	4051

	Up			
Downstream LDR	AcDx-7224-CaNCR25-Dn	/5Phos/GGGTTTTAATATTTTTTCGTTGAGATCGCGGGGTGGGATTAAGGGCGATGGA	52	4052
Real-Time Probe	AcDx-7225-CaNCR25-RT-Pb	/56-FAM/AAGGTTCTC/ZEN/GGGTTTTAATATTTTTTCGTTGAGATCGC/3IABkFQ/	38	4053
Tag Forward Primer	AcDx-7226-CaNCR25-RT-FP	TATCTCCTAAAAGAAGCCGCAC	22	4054
Tag Reverse Primer	AcDx-7227-CaNCR25-RT-RP	TCCATCGCCCTTAATCCCAC	20	4055
Downstream PCR Primer	AcDx-7228-CaNCR25-PCR-V	TCCATCGCCCTTAATCCCACCAAAAAAACCCAAAAACAACGCCTGrCGATT/3SpC3/	50	4056
HOXA10				
Forward PCR Primer	AcDx-7231-HOXA10-FP	GGTAAGATCGAGGCGCrGTTTG/3SpC3/	21	4057
Reverse PCR Primer	AcDx-7232-HOXA10-RP	GGTGTCGTGGCGCTAAACGACAAACGCAArUAAAG/3SpC3/	34	4058
Upstream LDR	AcDx-7233-HOXA10-Up	TCGCTCTTCAGCCTCCTACAGAGGGTTCGTAGTCGTGCGTCTCrGGGCC/3SpC3/	48	4059
Downstream LDR	AcDx-7234-HOXA10-Dn	/5Phos/GGGATTTAGATTTTCGTTATCGTTATCGTTGTTCGGCTGTTCTGGGAATTATTGCCGGA	59	4060
Real-Time Probe	AcDx-7235-HOXA10-RT-Pb	/56-FAM/AAGCGTCTC/ZEN/GGGATTTAGATTTTCGTTATCGTTATCG/3IABkFQ/	37	4061
Tag Forward Primer	AcDx-7236-HOXA10-RT-FP	TCGCTCTTCAGCCTCCTACA	20	4062
Tag Reverse Primer	AcDx-7237-HOXA10-RT-RP	TCCGGCAATAATTCCCAGAACA	22	4063
Downstream PCR Primer	AcDx-7238-HOXA10-PCR-V	TCCGGCAATAATTCCCAGAACAACGCAATAAAACAACGTCGCTGrAACAG/3SpC3/	49	4064
ABHD8				
Forward PCR Primer	AcDx-7241-ABHD8-FP	GACGGAAGCGGAGAGCrGGAAC/3SpC3/	21	4065
Reverse PCR Primer	AcDx-7242-ABHD8-RP	GGTGTCGTGGGTTCGCTCCGATAAACGAAACrCAAAC/3SpC3/	36	4066
Upstream LDR	AcDx-7243-ABHD8-Up	TTCAACGATCGCGCAGACAGTTCGTTGGGTTTTTTCGGATGTCACrGTTCT/3SpC3/	50	4067
Downstream LDR	AcDx-7244-ABHD8-Dn	/5Phos/GTTTCGTTTAGGTAGTTTGGAGGCGTTTTTTTGTGTTCTGGGAATTATTGCCGGA	55	4068
Real-Time Probe	AcDx-7245-ABHD8-RT-	/56-FAM/TTATGTCAC/ZEN/GTTTCGTTTAGGTAGTTTGGAGG/3IABkFQ/	32	4069

	Pb			
Tag Forward Primer	AcDx-7246-ABHD8-RT-FP	TTCAACGATCGCGCAGACA	19	4070
Tag Reverse Primer	AcDx-7247-ABHD8-RT-RP	TCCGGCAATAATTCCCAGAACA	22	4071
Downstream PCR Primer	AcDx-7248-ABHD8-PCR-V	TCCGGCAATAATTCCCAGAACACTCCGATAAACGAAACCAAATCAAAAAAATGrCCCCA/3SpC3/	58	4072
HOXD8-S1				
Forward PCR Primer	AcDx-7251-HOXD8-S1-FP	TTTAGAGTCGAGGTTTGTAAATCrGAGGC/3SpC3/	28	4073
Reverse PCR Primer	AcDx-7252-HOXD8-S1-RP	GGTGTCGTGGACGACCTACCCCGCTACrCTCCG/3SpC3/	32	4074
Upstream LDR	AcDx-7253-HOXD8-S1-Up	TCACTATCGGCGTAGTCACCAGTTAGAGTGTTTTCGTGGGTCGGGCrGTACC/3SpC3/	51	4075
Downstream LDR	AcDx-7254-HOXD8-S1-Dn	/5Phos/GTATTTTTTTTTTGTTCGGGTGCGTTTAGTTATTGGTGTGGTGACTTTACCCGGAGGA	58	4076
Real-Time Probe	AcDx-7255-HOXD8-S1-RT-Pb	/56-FAM/TTGTCGGGC/ZEN/GTATTTTTTTTTTGTTCGGGTG/3IABkFQ/	31	4077
Tag Forward Primer	AcDx-7256-HOXD8-S1-RT-FP	TCACTATCGGCGTAGTCACCA	21	4078
Tag Reverse Primer	AcDx-7257-HOXD8-S1-RT-RP	TCCTCCGGGTAAAGTCACCA	20	4079
Downstream PCR Primer	AcDx-7258-HOXD8-S1-PCR-V	TCCTCCGGGTAAAGTCACCAACTATTTCCTCTCAAACACCAATAACTAAATGrCACCT/3SpC3/	57	4080
CaNCR26				
Forward PCR Primer	AcDx-7261-CaNCR26-FP	CGGGACGGGTTTTTTTGCrGGATC/3SpC3/	23	4081
Reverse PCR Primer	AcDx-7262-CaNCR26-RP	GGTGTCGTGGACACCTAAAACAATAACAACCGCrCCGAT/3SpC3/	38	4082
Upstream LDR	AcDx-7263-CaNCR26-Up	TCATCTGTTCGTCAGGGTCCAGATTTTTTGAAATGAAATAATGTGATGTACGTTGCrGATGG/3SpC3/	61	4083
Downstream LDR	AcDx-7264-CaNCR26-Dn	/5Phos/GATAAGGGTCGGTTTGTAATGAGGTTTAGGTCGTGGTGACTTTACCCGGAGGA	53	4084

Real-Time Probe	AcDx-7265-CaNCR26-RT-Pb	/56-FAM/AAACGTTGC/ZEN/GATAAGGGTCGGTTTGTAATGAGG/3IABkFQ/	33	4085
Tag Forward Primer	AcDx-7266-CaNCR26-RT-FP	TCATCTGTTCGTCAGGGTCCA	21	4086
Tag Reverse Primer	AcDx-7267-CaNCR26-RT-RP	TCCTCCGGGTAAAGTCACCA	20	4087
Downstream PCR Primer	AcDx-7268-CaNCR26-PCR-V	TCCTCCGGGTAAAGTCACCACTAAAACAATAACAACCGCCCGATGrACCTG/3SpC3/	50	4088
CaNCR27				
Forward PCR Primer	AcDx-7271-CaNCR27-FP	TAGTTTTTCGGTGGCGGCrGGCGC/3SpC3/	23	4089
Reverse PCR Primer	AcDx-7272-CaNCR27-RP	GGTGTCGTGGCGACCATAACCCCGACACrUAAAG/3SpC3/	33	4090
Upstream LDR	AcDx-7273-CaNCR27-Up	TTCGTACCTCGGCACACCAGGCGGCGTTAGTTGGTTTGCACrGAAGT/3SpC3/	46	4091
Downstream LDR	AcDx-7274-CaNCR27-Dn	/5Phos/GAAACGTTTTTTTTTTCGTTTCGTGTGTTTTATTTTTTATTTGGCTCCGTTACTCTGTCGA	61	4092
Real-Time Probe	AcDx-7275-CaNCR27-RT-Pb	/56-FAM/CCTTTGCAC/ZEN/GAAACGTTTTTTTTTTCGTTTCGTGTGTTTTA/3IABkFQ/	41	4093
Tag Forward Primer	AcDx-7276-CaNCR27-RT-FP	TTCGTACCTCGGCACACCA	19	4094
Tag Reverse Primer	AcDx-7277-CaNCR27-RT-RP	TCGACAGAGTAACGGAGCCA	20	4095
Downstream PCR Primer	AcDx-7278-CaNCR27-PCR-V	TCGACAGAGTAACGGAGCCAGACCATAACCCCGACACTAAAAAATGrUAAAG/3SpC3/	51	4096
USP2-S1				
Forward PCR Primer	AcDx-7281-USP2-S1-FP	TAAATGTAAACGTCGAGGGTACrGGGAT/3SpC3/	27	4097
Reverse PCR Primer	AcDx-7282-USP2-S1-RP	GGTGTCGTGGATCAAAATCTAAAACAAAAAACCGAACrUTTCT/3SpC3/	42	4098
Upstream LDR	AcDx-7283-USP2-S1-Up	TCTACAGCTAGATGCGGCCAGGTACGAGGCGATGTTGGTTTCACrGTTCG/3SpC3/	49	4099
Downstream LDR	AcDx-7284-USP2-S1-Dn	/5Phos/GTTTAGGCGATTTTTTTTGTCGGGTTATAGTTTAGGTGGCTCCGTTACTCTGTCGA	56	4100
Real-Time Probe	AcDx-7285-USP2-S1-RT-Pb	/56-FAM/AAGTTTCAC/ZEN/GTTTAGGCGATTTTTTTTGTCGGGT/3IABkFQ/	34	4101

Tag Forward Primer	AcDx-7286-USP2-S1-RT-FP	TCTACAGCTAGATGCGGCCA	20	4102
Tag Reverse Primer	AcDx-7287-USP2-S1-RT-RP	TCGACAGAGTAACGGAGCCA	20	4103
POU4F1				
Forward PCR Primer	AcDx-7291-POU4F1-FP	GTCGTTTCGAGGAGTTTTCrGCGAA/3SpC3/	24	4104
Reverse PCR Primer	AcDx-7292-POU4F1-RP	GGTGTCGTGGCACGAAACCGCCGAAArAAAAG/3SpC3/	31	4105
Upstream LDR	AcDx-7293-POU4F1-Up	TAGCGATAGTACCGACAGTCACTCGCGAGAGTTCGCGGTTTTGTCrGCGCC/3SpC3/	50	4106
Downstream LDR	AcDx-7294-POU4F1-Dn	/5Phos/GCGTTGATAGGTATTAGTTGTTTTCGTTTGTTTGACGGTGCGGAAACCTATCGTCGA	57	4107
Real-Time Probe	AcDx-7295-POU4F1-RT-Pb	/56-FAM/TTTTGTCGC/ZEN/GTTGATAGGTATTAGTTGTTTTCGTTTG/3IABkFQ/	37	4108
Tag Forward Primer	AcDx-7296-POU4F1-RT-FP	TAGCGATAGTACCGACAGTCAC	22	4109
Tag Reverse Primer	AcDx-7297-POU4F1-RT-RP	TCGACGATAGGTTTCCGCAC	20	4110
Downstream PCR Primer	AcDx-7298-POU4F1-PCR-V	TCGACGATAGGTTTCCGCACCGCCGAAAAAAAAAACGCGTGrUCAAG/3SpC3/	46	4111
FAM59B-S1				
Forward PCR Primer	AcDx-7301-FAM59B-S1-FP	TCGAGTTTTGGGCGGCrGGAGC/3SpC3/	21	4112
Reverse PCR Primer	AcDx-7302-FAM59B-S1-RP	GGTGTCGTGGCGCTCCCCCTCGTACTArACCTT/3SpC3/	32	4113
Upstream LDR	AcDx-7303-FAM59B-S1-Up	TCCAGGGTATTTGGCGCACGCGGAGTAGGATAGGGTGCrGGGCA/3SpC3/	43	4114
Downstream LDR	AcDx-7304-FAM59B-S1-Dn	/5Phos/GGGTGGGCGCGGTTTTCGGGTGCGGAAACCTATCGTCGA	39	4115
Real-Time Probe	AcDx-7305-FAM59B-S1-RT-Pb	/56-FAM/CCAGGGTGC/ZEN/GGGTGGGCG/3IABkFQ/	18	4116
Tag Forward Primer	AcDx-7306-FAM59B-S1-	TCCAGGGTATTTGGCGCAC	19	4117

	RT-FP			
Tag Reverse Primer	AcDx-7307-FAM59B-S1-RT-RP	TCGACGATAGGTTTCCGCAC	20	4118
Downstream PCR Primer	AcDx-7308-FAM59B-S1-PCR-V	TCGACGATAGGTTTCCGCACTCGTACTAACCTCCCGAAAACTGrCGCCT/3SpC3/	48	4119
SHH				
Forward PCR Primer	AcDx-7311-SHH-FP	GATTCGGAGGATGGATTAGCrGTTGC/3SpC3/	25	4120
Reverse PCR Primer	AcDx-7312-SHH-RP	GGTGTCGTGGACGCCCCCTACGCAArAACCC/3SpC3/	30	4121
Upstream LDR	AcDx-7313-SHH-Up	TCCCTCGTCATCTCCCTTACCTGGGAGGAGGTTTTCGGAGATTCTCrGTTGA/3SpC3/	51	4122
Downstream LDR	AcDx-7314-SHH-Dn	/5Phos/GTTAGGAGGATTTCGCGGGTAGGGAGTCGGTCTTGGTGATGGAGCGA	47	4123
Real-Time Probe	AcDx-7315-SHH-RT-Pb	/56-FAM/TTGATTCTC/ZEN/GTTAGGAGGATTTCGCGGG/3IABkFQ/	28	4124
Tag Forward Primer	AcDx-7316-SHH-RT-FP	TCCCTCGTCATCTCCCTTACC	21	4125
Tag Reverse Primer	AcDx-7317-SHH-RT-RP	TCGCTCCATCACCAAGACC	19	4126
Downstream PCR Primer	AcDx-7318-SHH-PCR-V	TCGCTCCATCACCAAGACCCGCAAAACCTCCTCCCTGrACTCT/3SpC3/	42	4127
CYB5R2				
Forward PCR Primer	AcDx-7321-CYB5R2-FP	GAGGCGGGTGTTTGCrGGGAC/3SpC3/	20	4128
Reverse PCR Primer	AcDx-7322-CYB5R2-RP	GGTGTCGTGGCTCTCTCCACCCAACGAATArAATAG/3SpC3/	35	4129
Upstream LDR	AcDx-7323-CYB5R2-Up	TCATAATGTTGTCAGCCCGACCGTCGAAGTAGAGGCGTCATCrGAGCC/3SpC3/	47	4130
Downstream LDR	AcDx-7324-CYB5R2-Dn	/5Phos/GAGTTGAAGACGTGTATTTCGAGCGTTTTTGCGGGTCTTGGTGATGGAGCGA	52	4131
Real-Time Probe	AcDx-7325-CYB5R2-RT-Pb	/56-FAM/TTGGGCATC/ZEN/GTCGAGTTGAAGACGTGTATTTC/3IABkFQ/	32	4132
Tag Forward Primer	AcDx-7326-CYB5R2-RT-FP	TCATAATGTTGTCAGCCCGACC	22	4133
Tag Reverse Primer	AcDx-7327-CYB5R2-RT-RP	TCGCTCCATCACCAAGACC	19	4134
Downstream PCR Primer	AcDx-7328-CYB5R2-PCR-V	TCGCTCCATCACCAAGACCACCCAACGAATAAATAAACGCAAAAATGrCTCGG/3SpC3/	52	4135

GNB4				
Forward PCR Primer	AcDx-7641-GNB4-FP	GTTTTGTTGGGTAGTTTCGAATATTCrGTTAC/3SpC3/	31	4136
Reverse PCR Primer	AcDx-7642-GNB4-RP	GGTGTCGTGGCCCCCCTAATTTCTCGTCACrUCCCT/3SpC3/	35	4137
Upstream LDR	AcDx-7643-GNB4-Up	TCCTGAGGGACAAATACACACCAGTTTCGAATATTCGTTATTTTAGGGATACrGTATA/3SpC3/	57	4138
Downstream LDR	AcDx-7644-GNB4-Dn	/5Phos/GTACGGGTTCGTGTTTTGAGTTTTTGGAAGGGGTAGGTAAGGAAGTCACGCA	52	4139
Real-Time Probe	AcDx-7645-GNB4-RT-Pb	/56-FAM/AAGGGATAC/ZEN/GTACGGGTTCGTGTTTTGAGTTT/3IABkFQ/	32	4140
Tag Forward Primer	AcDx-7646-GNB4-RT-FP	TCCTGAGGGACAAATACACACC	22	4141
Tag Reverse Primer	AcDx-7647-GNB4-RT-RP	TGCGTGACTTCCTTACCTACC	21	4142
Downstream PCR Primer	AcDx-7648-GNB4-PCR-V	TGCGTGACTTCCTTACCTACCCCCTAATTTCTCGTCACTCCCTGrAAACT/3SpC3/	49	4143
CaNCR37				
Forward PCR Primer	AcDx-7651-CaNCR37-FP	GTAGACGTTTTTTTTTTAGGAGGTCrGGAAC/3SpC3/	30	4144
Reverse PCR Primer	AcDx-7652-CaNCR37-RP	GGTGTCGTGGCTCACCGACCCTCGCArACGAC/3SpC3/	31	4145
Upstream LDR	AcDx-7653-CaNCR37-Up	TTGCAACAGGCTACCGACCCGGAATTTTTGTTTTTATTTTTTATCGGGATTCrGGTCA/3SpC3/	57	4146
Downstream LDR	AcDx-7654-CaNCR37-Dn	/5Phos/GGTTGGTTAGAGGTAAGTTTCGAGATTTTTTATTAATTATTATTATCGGGTAGGTAAGGAAGTCACGCA	69	4147
Real-Time Probe	AcDx-7655-CaNCR37-RT-Pb	/56-FAM/AAGGGATTC/ZEN/GGTTGGTTAGAGGTAAGTTTCGAGAT/3IABkFQ/	35	4148
Tag Forward Primer	AcDx-7656-CaNCR37-RT-FP	TTGCAACAGGCTACCGACC	19	4149
Tag Reverse Primer	AcDx-7657-CaNCR37-RT-RP	TGCGTGACTTCCTTACCTACC	21	4150
Downstream PCR Primer	AcDx-7658-CaNCR37-PCR-V	TGCGTGACTTCCTTACCTACCCTCACCGACCCTCGCAATGrATAAC/3SpC3/	45	4151
ZMIZ1-S1				
Forward PCR Primer	AcDx-7661-ZMIZ1-S1-FP	GGTTTTTCGTTCGAGGAATTTCrGGGAA/3SpC3/	27	4152
Reverse PCR Primer	AcDx-7662-ZMIZ1-S1-RP	GGTGTCGTGGAACTAAACATCCAAATTAAATCTCGArUTTAG/3SpC3/	41	4153

Upstream LDR	AcDx-7663-ZMIZ1-S1-Up	TAAGCCTGCTTTTCCGAAACAAGGTTAGGGAAGTAAGATGTCGGATCrGTTCG/3SpC3/	52	4154
Downstream LDR	AcDx-7664-ZMIZ1-S1-Dn	/5Phos/TTTAAAAAATTTTCGAAACGAACTACGAAATAAAAAAAATAAACTTGTATTGCGCCAGGATAGCA	65	4155
Real-Time Probe	AcDx-7665-ZMIZ1-S1-RT-Pb	/56-FAM/AATCGGATC/ZEN/GTTTATTTTTTTTATTTCGTAGTTCGTTTCGAA/3IABkFQ/	42	4156
Tag Forward Primer	AcDx-7666-ZMIZ1-S1-RT-FP	TAAGCCTGCTTTTCCGAAACAA	22	4157
Tag Reverse Primer	AcDx-7667-ZMIZ1-S1-RT-RP	TGCTATCCTGGCGCAATACAA	21	4158
LRRFIP1				
Forward PCR Primer	AcDx-7841-LRRFIP1-FP	TAGTTTGGACGGTGTGGATTTCrGGGTC/3SpC3/	27	4159
Reverse PCR Primer	AcDx-7842-LRRFIP1-RP	GGTGTCGTGGCGATACGACGACCCGCrAAAAG/3SpC3/	31	4160
Upstream LDR	AcDx-7843-LRRFIP1-Up	TTGCTGTGCGCGGTAGAACTGTGGATTTCGGGTTTTGCAACrGTTGA/3SpC3/	46	4161
Downstream LDR	AcDx-7844-LRRFIP1-Dn	/5Phos/GTTAGATCGGTTTTAGCGGTTCGTATTCGTATTTTGTAAGGTTACGCTAAGCTGGTGCCA	60	4162
Real-Time Probe	AcDx-7845-LRRFIP1-RT-Pb	/56-FAM/AATTGCAAC/ZEN/GTTAGATCGGTTTTAGCGGTTCG/3IABkFQ/	32	4163
Tag Forward Primer	AcDx-7846-LRRFIP1-RT-FP	TTGCTGTGCGCGGTAGAAC	19	4164
Tag Reverse Primer	AcDx-7847-LRRFIP1-RT-RP	TGGCACCAGCTTAGCGTAAC	20	4165
Downstream PCR Primer	AcDx-7848-LRRFIP1-PCR-V	TGGCACCAGCTTAGCGTAACCCGCAAAAAAAAACGAACTTACAAAATATGrAATAT/3SpC3/	55	4166
ASCL2				
Forward PCR Primer	AcDx-7871-ASCL2-FP	AGGTTTAGGTTTTCGAGGCrGTTTC/3SpC3/	24	4167
Reverse PCR Primer	AcDx-7872-ASCL2-RP	GGTGTCGTGGCCCAAAACCCTCAAACCGArAAACA/3SpC3/	34	4168
Upstream LDR	AcDx-7873-ASCL2-Up	TCACAGAGACTTGCCGATCACGGGCGTTTTTTTTAATTCGTTTCGTTTTTCTCrGTTCC/3SpC3/	58	4169

Downstream LDR	AcDx-7874-ASCL2-Dn	/5Phos/GTTTTTTTACGCGTATTTTGTTTGTGGTTTTCGTGCGGTGTGTAGCTTAGACATGGCCA	59	4170
Real-Time Probe	AcDx-7875-ASCL2-RT-Pb	/56-FAM/AATTTTCTC/ZEN/GTTTTTTTACGCGTATTTTGTTTGTGGTTTTC/3IABkFQ/	41	4171
Tag Forward Primer	AcDx-7876-ASCL2-RT-FP	TCACAGAGACTTGCCGATCAC	21	4172
Tag Reverse Primer	AcDx-7877-ASCL2-RT-RP	TGGCCATGTCTAAGCTACACAC	22	4173
Downstream PCR Primer	AcDx-7878-ASCL2-PCR-V	TGGCCATGTCTAAGCTACACACCCCAAAACCCTCAAACCGAAAATGrCACTG/3SpC3/	51	4174
CCDC151-S2				
Forward PCR Primer	AcDx-7901-CCDC151-S2-FP	GTCGCGTTTTTTAGTTTTATAGGATTCrGTTTA/3SpC3/	32	4175
Reverse PCR Primer	AcDx-7902-CCDC151-S2-RP	GGTGTCGTGGAACTAATCAACCAAAAAAAAATCTCGAArAACAG/3SpC3/	43	4176
Upstream LDR	AcDx-7903-CCDC151-S2-Up	TGATGCTGGCAAACCCTAGAACGTTTGTATTTTTTTTGGTTTTTTTTTTCGTGGCTCrGGGCC/3SpC3/	62	4177
Downstream LDR	AcDx-7904-CCDC151-S2-Dn	/5Phos/GGGTTTATTTTTGGTTCGTTAAATTTCGTTCGTTGGTTCCATCACCGTTAGGCCA	55	4178
Real-Time Probe	AcDx-7905-CCDC151-S2-RT-Pb	/56-FAM/AAGTGGCTC/ZEN/GGGTTTATTTTTGGTTCGTTAAATTTCG/3IABkFQ/	37	4179
Tag Forward Primer	AcDx-7906-CCDC151-S2-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	4180
Tag Reverse Primer	AcDx-7907-CCDC151-S2-RT-RP	TGGCCTAACGGTGATGGAAC	20	4181
Downstream PCR Primer	AcDx-7908-CCDC151-S2-PCR-V	TGGCCTAACGGTGATGGAACACTAATCAACCAAAAAAAAATCTCGAAAACAATGrAACGG/3SpC3/	59	4182
CHRD				
Forward PCR Primer	AcDx-7911-CHRD-FP	GGTTCGTGGTGACGGTTATACrGGGTG/3SpC3/	26	4183
Reverse PCR Primer	AcDx-7912-CHRD-RP	GGTGTCGTGGAATAACCTAAACACTATCAAACGCrCGATC/3SpC3/	39	4184
Upstream LDR	AcDx-7913-CHRD-Up	TTTTCGGCATCCGCTTCCAGGTAGGAGGTTGGCGAATGGCrGGGAC/3SpC3/	45	4185
Downstream LDR	AcDx-7914-CHRD-Dn	/5Phos/GGAGTTGTGGTTGAGGTTG*GGTTATTAAAGCGGTGGTTAACAGAGGACAGGCCA	54	4186

Real-Time Probe	AcDx-7915-CHRD-RT-Pb	/56-FAM/TTGAATGGC/ZEN/GGAGTTGTGGTTGAGGTTG/3IABkFQ/	28	4187
Tag Forward Primer	AcDx-7916-CHRD-RT-FP	TTTTCGGCATCCGCTTCCA	19	4188
Tag Reverse Primer	AcDx-7917-CHRD-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	4189
Downstream PCR Primer	AcDx-7918-CHRD-PCR-V	TGGCCTGTCCTCTGTTAACCAAAAAAATAACCTAAACACTATCAAACGCTGrCTTTG/3SpC3/	56	4190
CaNCR50				
Forward PCR Primer	AcDx-8151-CaNCR50-FP	GGAGGGCGTTTTTCGCrGTTGC/3SpC3/	21	4191
Reverse PCR Primer	AcDx-8152-CaNCR50-RP	GGTGTCGTGGCGCCCCCGAACCCTArATCCA/3SpC3/	30	4192
Upstream LDR	AcDx-8153-CaNCR50-Up	TTTTTACGCACAGCACCACCCGTTTTTCGCGTTGTTTTTGATTCGCTCrGGTCC/3SpC3/	53	4193
Downstream LDR	AcDx-8154-CaNCR50-Dn	/5Phos/GGTTTTGCGTTTGTTTGAAGTTTTTCGTTTCGTATTTGGTTACATAGGCGGCTTAGACA	59	4194
Real-Time Probe	AcDx-8155-CaNCR50-RT-Pb	/56-FAM/CCTTCGCTC/ZEN/GGTTTTGCGTTTGTTTGAAGTTTTTC/3IABkFQ/	35	4195
Tag Forward Primer	AcDx-8156-CaNCR50-RT-FP	TTTTTACGCACAGCACCACC	20	4196
Tag Reverse Primer	AcDx-8157-CaNCR50-RT-RP	TGTCTAAGCCGCCTATGTAACC	22	4197
Downstream PCR Primer	AcDx-8158-CaNCR50-PCR-V	TGTCTAAGCCGCCTATGTAACCCCCCGAACCCTAATCCGAAAATATGrAAACA/3SpC3/	52	4198
GALR3				
Forward PCR Primer	AcDx-8161-GALR3-FP	GTACGGTCGTTTCGTTTTTAGTTCrGGTTG/3SpC3/	29	4199
Reverse PCR Primer	AcDx-8162-GALR3-RP	GGTGTCGTGGGCGAAACGAACGCGTAArACGAG/3SpC3/	32	4200
Upstream LDR	AcDx-8163-GALR3-Up	TGGACACTTCGCCCTTCTTAACGTTTCGTTTTTAGTTCGGTTATTTACGTTTATCrGTTCA/3SpC3/	60	4201
Downstream LDR	AcDx-8164-GALR3-Dn	/5Phos/GTTTGGTTTTATATTGTTTGGTTTACGTTAATTTTTGTTTTAATTCGTTTGGGATCTGGGCATCACA	67	4202
Real-Time Probe	AcDx-8165-GALR3-RT-Pb	/56-FAM/TTGTTTATC/ZEN/GTTTGGTTTTATATTGTTTGGTTTACGTTAATTTTTG/3IABkFQ/	46	4203
Tag Forward Primer	AcDx-8166-GALR3-RT-	TGGACACTTCGCCCTTCTTAAC	22	4204

	FP			
Tag Reverse Primer	AcDx-8167-GALR3-RT-RP	TGTGATGCCCAGATCCCAAAC	21	4205
Downstream PCR Primer	AcDx-8168-GALR3-PCR-V	TGTGATGCCCAGATCCCAAACCGAAACGAACGCGTAAACGAATGrAATTG/3SpC3/	49	4206
CaNCR51				
Forward PCR Primer	AcDx-8171-CaNCR51-FP	GGGATTGCGGGACGCrGGGCA/3SpC3/	20	4207
Reverse PCR Primer	AcDx-8172-CaNCR51-RP	GGTGTCGTGGCCGCAACCCCTAAAACGAArAATAG/3SpC3/	34	4208
Upstream LDR	AcDx-8173-CaNCR51-Up	TGGAGGCCGGAGAAATTAAACGAGCGAGAGTTTTGTGGGTTGTTAAATCrGGCAT/3SpC3/	54	4209
Downstream LDR	AcDx-8174-CaNCR51-Dn	/5Phos/GGCGCGGTCGCGTCGCGTTTGGGATCTGGGCATCACA	37	4210
Real-Time Probe	AcDx-8175-CaNCR51-RT-Pb	/56-FAM/AATTAAATC/ZEN/GGCGCGGTCGCGTC/3IABkFQ/	23	4211
Tag Forward Primer	AcDx-8176-CaNCR51-RT-FP	TGGAGGCCGGAGAAATTAAAC	21	4212
Tag Reverse Primer	AcDx-8177-CaNCR51-RT-RP	TGTGATGCCCAGATCCCAAAC	21	4213
Downstream PCR Primer	AcDx-8178-CaNCR51-PCR-V	TGTGATGCCCAGATCCCAAACCTAAAACGAAAATAACAAAAAAAACGACTACTGrCGACA/3SpC3/	59	4214
CaNCR52				
Forward PCR Primer	AcDx-8181-CaNCR52-FP	GTTTTTTATTGTAAGCGGCGTTATCrGGATC/3SpC3/	30	4215
Reverse PCR Primer	AcDx-8182-CaNCR52-RP	GGTGTCGTGGCTAATTAACATACGACGCCGATTArCCCGG/3SpC3/	39	4216
Upstream LDR	AcDx-8183-CaNCR52-Up	TTGTCTCTGCGACCCATCAAGGATTTTATTTGTAAATTTATTAGTGTGCGTTTGCrGGGAG/3SpC3/	60	4217
Downstream LDR	AcDx-8184-CaNCR52-Dn	/5Phos/GGAGAGGAGGTTCGGGAATTCGTTGGTACACGTTCGGCACA	41	4218
Real-Time Probe	AcDx-8185-CaNCR52-RT-Pb	/56-FAM/AACGTTTGC/ZEN/GGAGAGGAGGTTCG/3IABkFQ/	23	4219
Tag Forward Primer	AcDx-8186-CaNCR52-RT-FP	TTGTCTCTGCGACCCATCAA	20	4220

Tag Reverse Primer	AcDx-8187-CaNCR52-RT-RP	TGTGCCGAACGTGTACCAA	19	4221
Downstream PCR Primer	AcDx-8188-CaNCR52-PCR-V	TGTGCCGAACGTGTACCAACGCCGATTACCCGAACCTGrAATTT/3SpC3/	43	4222
PRKCB				
Forward PCR Primer	AcDx-8191-PRKCB-FP	TTAAGCGTAGTTGGACGAGCrGGTAA/3SpC3/	25	4223
Reverse PCR Primer	AcDx-8192-PRKCB-RP	GGTGTCGTGGTCCCCTACGCCGACTCrUAACA/3SpC3/	31	4224
Upstream LDR	AcDx-8193-PRKCB-Up	TTCGCCTACCGCAGTGAACACGAGCGGTAGTAGTTGAGCrGAGCA/3SpC3/	44	4225
Downstream LDR	AcDx-8194-PRKCB-Dn	/5Phos/GAGTGATAGTTTCGGTTTCGCGCGTCGGTTGAGACATGGGCTCGCA	46	4226
Real-Time Probe	AcDx-8195-PRKCB-RT-Pb	/56-FAM/ATGTTGAGC/ZEN/GAGTGATAGTTTCGGTTTCGC/3IABkFQ/	30	4227
Tag Forward Primer	AcDx-8196-PRKCB-RT-FP	TTCGCCTACCGCAGTGAAC	19	4228
Tag Reverse Primer	AcDx-8197-PRKCB-RT-RP	TGCGAGCCCATGTCTCAAC	19	4229
Downstream PCR Primer	AcDx-8198-PRKCB-PCR-V	TGCGAGCCCATGTCTCAACCGCCGACTCTAACGACTGrCGACA/3SpC3/	42	4230
TDRD10-S1				
Forward PCR Primer	AcDx-8261-TDRD10-S1-FP	TAGTTCGCGTTTGTATCGAGTCrGGTTC/3SpC3/	27	4231
Reverse PCR Primer	AcDx-8262-TDRD10-S1-RP	GGTGTCGTGGACGAAAAACTTCCTTCCCGAArAATAG/3SpC3/	36	4232
Upstream LDR	AcDx-8263-TDRD10-S1-Up	TCGCAACGTGCCGAATACAGAGTCGGTTTCGGTCGAGCrGTTAC/3SpC3/	43	4233
Downstream LDR	AcDx-8264-TDRD10-S1-Dn	/5Phos/GTTGTTTTTATACGCGTTTAGGAGTGTTACGTGCGTGTTGCACGGTCGAGCTAA	54	4234
Real-Time Probe	AcDx-8265-TDRD10-S1-RT-Pb	/56-FAM/TTGTCGAGC/ZEN/GTTGTTTTTATACGCGTTTAGGAGTG/3IABkFQ/	35	4235
Tag Forward Primer	AcDx-8266-TDRD10-S1-RT-FP	TCGCAACGTGCCGAATACA	19	4236

Tag Reverse Primer	AcDx-8267-TDRD10-S1-RT-RP	TTAGCTCGACCGTGCAACA	19	4237
Downstream PCR Primer	AcDx-8268-TDRD10-S1-PCR-V	TTAGCTCGACCGTGCAACACTTCCCGAAAATAAACGACGCATGrUAACG/3SpC3/	48	4238
HEPACAM				
Forward PCR Primer	AcDx-8271-HEPACAM-FP	GTTTTATTATATTAATATTGTCGTTTTCGCrGTATC/3SpC3/	35	4239
Reverse PCR Primer	AcDx-8272-HEPACAM-RP	GGTGTCGTGGGCTAAAAACGAAAAAAAATCCCGArAAAAG/3SpC3/	39	4240
Upstream LDR	AcDx-8273-HEPACAM-Up	TAACGGGATTGAGAGTGGACAAATATTGTCGTTTTCGCGTATTCGTTCTCrGTTCA/3SpC3/	55	4241
Downstream LDR	AcDx-8274-HEPACAM-Dn	/5Phos/GTTTGCGTATGTTTATATACGTTTATATTCGAGATATTAGCGTTTTTGTCTGCCGCCCTTACTAA	65	4242
Real-Time Probe	AcDx-8275-HEPACAM-RT-Pb	/56-FAM/AACGTTCTC/ZEN/GTTTGCGTATGTTTATATACGTTTATATTCGAG/3IABkFQ/	42	4243
Tag Forward Primer	AcDx-8276-HEPACAM-RT-FP	TAACGGGATTGAGAGTGGACA	21	4244
Tag Reverse Primer	AcDx-8277-HEPACAM-RT-RP	TTAGTAAGGGCGGCAGACA	19	4245
Downstream PCR Primer	AcDx-8278-HEPACAM-PCR-V	TTAGTAAGGGCGGCAGACAGCTAAAAACGAAAAAAAATCCCGAAAAAATGrCTAAC/3SpC3/	55	4246
TRIM15-S1				
Forward PCR Primer	AcDx-8281-TRIM15-S1-FP	TCGTTGTTGATGTTTGCGCrGTTTC/3SpC3/	24	4247
Reverse PCR Primer	AcDx-8282-TRIM15-S1-RP	GGTGTCGTGGTCTAACAAACTCTTCTTCTACCGAArUATAT/3SpC3/	40	4248
Upstream LDR	AcDx-8283-TRIM15-S1-Up	TAACGGGATTGAGAGTGGACATTTTAGAGGTTATTATTTTGGATTTTTAGATCGTTAATCrGGAAC/3SpC3/	65	4249
Downstream LDR	AcDx-8284-TRIM15-S1-Dn	/5Phos/GGAGTTTGGTTTTTTCGGAAGATAGGAAGTTAGTGAGTGTCTGCCGCCCTTACTAA	56	4250

Real-Time Probe	AcDx-8285-TRIM15-S1-RT-Pb	/56-FAM/AAGTTAATC/ZEN/GGAGTTTGGTTTTTTCGGAAGATAGGAAG/3IABkFQ/	38	4251
Tag Forward Primer	AcDx-8286-TRIM15-S1-RT-FP	TAACGGGATTGAGAGTGGACA	21	4252
Tag Reverse Primer	AcDx-8287-TRIM15-S1-RT-RP	TTAGTAAGGGCGGCAGACA	19	4253
CANCR103				
Forward PCR Primer	AcDx-10621-CANCR103-FP	GGCGGAGATAAAATTAATAAGCGCrGGTTC/3SpC3/	29	4254
Reverse PCR Primer	AcDx-10622-CANCR103-RP	GGTGTCGTGGACTCAATTTAAACGACCTCGCrUCGAG/3SpC3/	36	4255
Upstream LDR	AcDx-10623-CANCR103-Up	TAACCGGGCCTAAAGTGACAAATAAGCGCGGTTTATTTTTTAGGTGCrGAGCG/3SpC3/	52	4256
Downstream LDR	AcDx-10624-CANCR103-Dn	/5Phos/GAGTATTAATTTTAGTCGTTTATTTTTTTTTTAGAGTTAATTTAGAGTTATTGTATTTTG TTACGTGATCTCCCTCTCCA	80	4257
Real-Time Probe	AcDx-10625-CANCR103-RT-Pb	/56-FAM/AATAGGTGC/ZEN/GAGTATTAATTTTAGTCGTTTATTTTTTTTTTAGAGTTAATT/3IABkFQ/	51	4258
Tag Forward Primer	AcDx-10626-CANCR103-RT-FP	TAACCGGGCCTAAAGTGACA	20	4259
Tag Reverse Primer	AcDx-10627-CANCR103-RT-RP	TGGAGAGGGAGATCACGTAACA	22	4260
Downstream PCR Primer	AcDx-10628-CANCR103-PCR-V	TGGAGAGGGAGATCACGTAACAACTCAATTTAAACGACCTCGCTTGrAAATG/3SpC3/	51	4261
BACH2				
Forward PCR Primer	AcDx-10631-BACH2-FP	CGAGAATTAGGGTGAGGCrGATCC/3SpC3/	23	4262
Reverse PCR Primer	AcDx-10632-BACH2-RP	GGTGTCGTGGAACTAACATAAAAAAAACTAAACGTAAAAAArCTACT/3SpC3/	46	4263
Upstream LDR	AcDx-10633-BACH2-Up	TCGATGGTCAATGAGCTTCACATAGGGTGAGGCGATTTTATATTTTAAGTAAAACrGTTTA/3SpC3/	60	4264

Downstream LDR	AcDx-10634-BACH2-Dn	/5Phos/GTTCGTTGTTTTATTAAAATAGTTTTTTTCGTTAAAGGAGGAGTAGTGTTACGTGATCTCCCTCTCCA	68	4265
Real-Time Probe	AcDx-10635-BACH2-RT-Pb	/56-FAM/TTGTAAAAC/ZEN/GTTCGTTGTTTTATTAAAATAGTTTTTTTCGTTAAAGGAGG/3IABkFQ/	50	4266
Tag Forward Primer	AcDx-10636-BACH2-RT-FP	TCGATGGTCAATGAGCTTCACA	22	4267
Tag Reverse Primer	AcDx-10637-BACH2-RT-RP	TGGAGAGGGAGATCACGTAACA	22	4268
CANCR104				
Forward PCR Primer	AcDx-10641-CANCR104-FP	CGGGTTGGGATTACGGTTTCrGGTAA/3SpC3/	25	4269
Reverse PCR Primer	AcDx-10642-CANCR104-RP	GGTGTCGTGGATAAACGCCTAACAACACTCCrCTAAC/3SpC3/	36	4270
Upstream LDR	AcDx-10643-CANCR104-Up	TCGATGGTCAATGAGCTTCACACGGTTTCGGTAGAGAGGGTGTTTAGATAGTCrGATGA/3SpC3/	58	4271
Downstream LDR	AcDx-10644-CANCR104-Dn	/5Phos/GATAGCGTTTTTTTTATTTTTATTATTGTTCGATTTATTTCGGGTTGGTGTTACGTGATCTCCCTCTCCA	70	4272
Real-Time Probe	AcDx-10645-CANCR104-RT-Pb	/56-FAM/CCGATAGTC/ZEN/GATAGCGTTTTTTTTATTTTTATTATTGTTCGATTTATTT/3IABkFQ/	49	4273
Tag Forward Primer	AcDx-10646-CANCR104-RT-FP	TCGATGGTCAATGAGCTTCACA	22	4274
Tag Reverse Primer	AcDx-10647-CANCR104-RT-RP	TGGAGAGGGAGATCACGTAACA	22	4275
Downstream PCR Primer	AcDx-10648-CANCR104-PCR-V	TGGAGAGGGAGATCACGTAACACTAACAACACTCCCTAATACCAACCTGrAAATG/3SpC3/	54	4276
TFR2-S1				
Forward PCR Primer	AcDx-10651-TFR2-S1-FP	TTTTTTCGAGTTGTAGATTTTTTGTCrGTAGC/3SpC3/	31	4277
Reverse PCR Primer	AcDx-10652-TFR2-S1-RP	GGTGTCGTGGCCCAAACCCGCGAACTAArCCCTG/3SpC3/	33	4278

Upstream LDR	AcDx-10653-TFR2-S1-Up	TCTCGGGACCACAATACGAACCGAGTTGTAGATTTTTTGTCGTAGTTTTTTCATCrGTTTA/3SpC3/	60	4279
Downstream LDR	AcDx-10654-TFR2-S1-Dn	/5Phos/GTTCGGATGTAGTTTTTTCGCGTCGAGTATATTTATTGTAGGGTTACGCTAAGCTGGTGCCA	62	4280
Real-Time Probe	AcDx-10655-TFR2-S1-RT-Pb	/56-FAM/AATTTCATC/ZEN/GTTCGGATGTAGTTTTTTCGCGT/3IABkFQ/	32	4281
Tag Forward Primer	AcDx-10656-TFR2-S1-RT-FP	TCTCGGGACCACAATACGAAC	21	4282
Tag Reverse Primer	AcDx-10657-TFR2-S1-RT-RP	TGGCACCAGCTTAGCGTAAC	20	4283
Downstream PCR Primer	AcDx-10658-TFR2-S1-PCR-V	TGGCACCAGCTTAGCGTAACCGCGAACTAACCCTACAATAAATATACTTGrACGCA/3SpC3/	55	4284
USP2-S2				
Forward PCR Primer	AcDx-10661-USP2-S2-FP	GTAGGCGTCGTTGGTGCrGTATC/3SpC3/	22	4285
Reverse PCR Primer	AcDx-10662-USP2-S2-RP	GGTGTCGTGGGACGAAAAATCTCCCGATCCrUACCT/3SpC3/	35	4286
Upstream LDR	AcDx-10663-USP2-S2-Up	TTGCTGTGCGCGGTAGAACGGAGGATATGGGAGTGACGTCGCrGGAAG/3SpC3/	47	4287
Downstream LDR	AcDx-10664-USP2-S2-Dn	/5Phos/GGAGAGAGCGGGAAGTTAGTTCGGAGGTTACGCTAAGCTGGTGCCA	46	4288
Real-Time Probe	AcDx-10665-USP2-S2-RT-Pb	/56-FAM/AAACGTCGC/ZEN/GGAGAGAGCGGG/3IABkFQ/	21	4289
Tag Forward Primer	AcDx-10666-USP2-S2-RT-FP	TTGCTGTGCGCGGTAGAAC	19	4290
Tag Reverse Primer	AcDx-10667-USP2-S2-RT-RP	TGGCACCAGCTTAGCGTAAC	20	4291
Downstream PCR Primer	AcDx-10668-USP2-S2-PCR-V	TGGCACCAGCTTAGCGTAACCCGATCCTACCCGCTCTGrAACTG/3SpC3/	43	4292
HIST1H3G				
Forward PCR Primer	AcDx-10671-HIST1H3G-	GGCGCGTTTTTTCGAGTCrGTTTC/3SpC3/	23	4293

	FP			
Reverse PCR Primer	AcDx-10672-HIST1H3G-RP	GGTGTCGTGGCTAAAATAACCCGCACCAAACrAAACC/3SpC3/	36	4294
Upstream LDR	AcDx-10673-HIST1H3G-Up	TCTCGATTACGCTCCGCACGTCGTTTTAGTGGTTAGTTGTTTGCrGGCAC/3SpC3/	49	4295
Downstream LDR	AcDx-10674-HIST1H3G-Dn	/5Phos/GGCGTTTTGTTATCGGTGGATTTGCGTGTAGGTGTGTAGCTTAGACATGGCCA	53	4296
Real-Time Probe	AcDx-10675-HIST1H3G-RT-Pb	/56-FAM/AATTTGTGC/ZEN/GGCGTTTTGTTATCGGTGGA/3IABkFQ/	29	4297
Tag Forward Primer	AcDx-10676-HIST1H3G-RT-FP	TCTCGATTACGCTCCGCAC	19	4298
Tag Reverse Primer	AcDx-10677-HIST1H3G-RT-RP	TGGCCATGTCTAAGCTACACAC	22	4299
Downstream PCR Primer	AcDx-10678-HIST1H3G-PCR-V	TGGCCATGTCTAAGCTACACACCCCGCACCAAACAAACTACATGrCAAAC/3SpC3/	49	4300
HIST1H2BH				
Forward PCR Primer	AcDx-10681-HIST1H2BH-FP	CGGTGAGGTTTTTTTACGTTATCrGGTGA/3SpC3/	28	4301
Reverse PCR Primer	AcDx-10682-HIST1H2BH-RP	GGTGTCGTGGCGTAAATCCACTAACGACAAAACCrCCGCA/3SpC3/	39	4302
Upstream LDR	AcDx-10683-HIST1H2BH-Up	TCACAGAGACTTGCCGATCACGGTTTTTTTACGTTATCGGTGGTTAGCrGCGCC/3SpC3/	53	4303
Downstream LDR	AcDx-10684-HIST1H2BH-Dn	/5Phos/GCGTTTTTGCGAGTCGTTTTAGTGGTTAGTTGTTTGGTGTGTAGCTTAGACATGGCCA	58	4304
Real-Time Probe	AcDx-10685-HIST1H2BH-RT-Pb	/56-FAM/AAGGTTAGC/ZEN/GCGTTTTTGCGAGTCGTTTTAGTG/3IABkFQ/	33	4305
Tag Forward Primer	AcDx-10686-HIST1H2BH-RT-FP	TCACAGAGACTTGCCGATCAC	21	4306
Tag Reverse Primer	AcDx-10687-HIST1H2BH-RT-RP	TGGCCATGTCTAAGCTACACAC	22	4307
Downstream PCR Primer	AcDx-10688-HIST1H2BH-PCR-V	TGGCCATGTCTAAGCTACACACCACTAACGACAAAACCCCGTGrCAAAT/3SpC3/	48	4308

FLJ26850				
Forward PCR Primer	AcDx-10691-FLJ26850-FP	GTTTTTTTTTATGGTAATTGTTTACGTCrGTAGA/3SpC3/	33	4309
Reverse PCR Primer	AcDx-10692-FLJ26850-RP	GGTGTCGTGGCGTTTTCCACCAAATTTTACGArCCCGG/3SpC3/	37	4310
Upstream LDR	AcDx-10693-FLJ26850-Up	TCACAGAGACTTGCCGATCACCGTAGGTTGGAGTTGTTTATTATCGGAGCTCrGTAGA/3SpC3/	57	4311
Downstream LDR	AcDx-10694-FLJ26850-Dn	/5Phos/GTAAGTAGTATGGGTGTTGGATAAATAGCGTGATCGGGGTGTGTAGCTTAGACATGGCCA	60	4312
Real-Time Probe	AcDx-10695-FLJ26850-RT-Pb	/56-FAM/AAGGAGCTC/ZEN/GTAAGTAGTATGGGTGTTGGATAAATAGCG/3IABkFQ/	39	4313
Tag Forward Primer	AcDx-10696-FLJ26850-RT-FP	TCACAGAGACTTGCCGATCAC	21	4314
Tag Reverse Primer	AcDx-10697-FLJ26850-RT-RP	TGGCCATGTCTAAGCTACACAC	22	4315
Downstream PCR Primer	AcDx-10698-FLJ26850-PCR-V	TGGCCATGTCTAAGCTACACACCGTTTTCCACCAAATTTTACGACCTGrATCAT/3SpC3/	53	4316
SLC2A9				
Forward PCR Primer	AcDx-10701-SLC2A9-FP	GGGTGGAGGTATTTATTATCGCrGATAA/3SpC3/	26	4317
Reverse PCR Primer	AcDx-10702-SLC2A9-RP	GGTGTCGTGGTAATCCTACTCGCTCCTCGTAACrCTCCT/3SpC3/	38	4318
Upstream LDR	AcDx-10703-SLC2A9-Up	TTCGTCCCTGCACGCTAACGTGAGGGTATTTATTATCGATAGGTTGTAATCrGTAAG/3SpC3/	56	4319
Downstream LDR	AcDx-10704-SLC2A9-Dn	/5Phos/GTAGAGGAAGGAGGAGTCGAAGGCGTTCGTTCCATCACCGTTAGGCCA	48	4320
Real-Time Probe	AcDx-10705-SLC2A9-RT-Pb	/56-FAM/AATGTAATC/ZEN/GTAGAGGAAGGAGGAGTCGAAGGC/3IABkFQ/	33	4321
Tag Forward Primer	AcDx-10706-SLC2A9-RT-FP	TTCGTCCCTGCACGCTAAC	19	4322
Tag Reverse Primer	AcDx-10707-SLC2A9-RT-RP	TGGCCTAACGGTGATGGAAC	20	4323
Downstream PCR Primer	AcDx-10708-SLC2A9-PCR-V	TGGCCTAACGGTGATGGAACCCTCGTAACCTCCCTCGTGrAACGT/3SpC3/	44	4324

PLTP				
Forward PCR Primer	AcDx-10711-PLTP-FP	GAGGCGATTTTAATTTCGTTCrGTTTC/3SpC3/	26	4325
Reverse PCR Primer	AcDx-10712-PLTP-RP	GGTGTCGTGGCAAACTACAAAATCCGCGTCACrCTCCG/3SpC3/	37	4326
Upstream LDR	AcDx-10713-PLTP-Up	TGATGCTGGCAAACCCTAGAACGGCGATTTTAATTTCGTTCGTTTTCGCTCrGTTCC/3SpC3/	56	4327
Downstream LDR	AcDx-10714-PLTP-Dn	/5Phos/GTTTTTGTTTGTTTTTGCGTTTTATTTAGTTTTAGCGTTTTGGTTCCATCACCGTTAGGCCA	62	4328
Real-Time Probe	AcDx-10715-PLTP-RT-Pb	/56-FAM/AATTCGCTC/ZEN/GTTTTTGTTTGTTTTTGCGTTTTATT/3IABkFQ/	35	4329
Tag Forward Primer	AcDx-10716-PLTP-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	4330
Tag Reverse Primer	AcDx-10717-PLTP-RT-RP	TGGCCTAACGGTGATGGAAC	20	4331
Downstream PCR Primer	AcDx-10718-PLTP-PCR-V	TGGCCTAACGGTGATGGAACATCCGCGTCACCTCCAAAATGrCTAAG/3SpC3/	46	4332
IGF1R				
Forward PCR Primer	AcDx-10721-IGF1R-FP	GGTTTTGGTTTTGTTGTGGTCrGGCGC/3SpC3/	26	4333
Reverse PCR Primer	AcDx-10722-IGF1R-RP	GGTGTCGTGGCAAAAAACAACGAATATAAAACGACACrUACTC/3SpC3/	42	4334
Upstream LDR	AcDx-10723-IGF1R-Up	TGATGCTGGCAAACCCTAGAACGGTTTTGTTGTGGTCGGCATCrGTGCT/3SpC3/	48	4335
Downstream LDR	AcDx-10724-IGF1R-Dn	/5Phos/GTGTCGTTTTTTTTTTTGTCGTTTCGTATTTTTTTTGTACGGTTCCATCACCGTTAGGCCA	61	4336
Real-Time Probe	AcDx-10725-IGF1R-RT-Pb	/56-FAM/AACGGCATC/ZEN/GTGTCGTTTTTTTTTTTGTCGTTTCG/3IABkFQ/	35	4337
Tag Forward Primer	AcDx-10726-IGF1R-RT-FP	TGATGCTGGCAAACCCTAGAAC	22	4338
Tag Reverse Primer	AcDx-10727-IGF1R-RT-RP	TGGCCTAACGGTGATGGAAC	20	4339
Downstream PCR Primer	AcDx-10728-IGF1R-PCR-V	TGGCCTAACGGTGATGGAACAAAACAACGAATATAAAACGACACTACTTATTAACTATGrUACAG/3SpC3/	64	4340
PTK2B				
Forward PCR Primer	AcDx-10731-PTK2B-FP	TTTAATTTGTTAGTTTTTTTATTTAGTTATAGTTTTCrGGAGC/3SpC3/	42	4341

Reverse PCR Primer	AcDx-10732-PTK2B-RP	GGTGTCGTGGCACCGCCAAATAAACCGAArACGAT/3SpC3/	34	4342
Upstream LDR	AcDx-10733-PTK2B-Up	TTTTCGGCATCCGCTTCCAAATTTGTTAGTTTTTTTATTTAGTTATAGTTTTCGGAATCrGTTAC/3SpC3/	64	4343
Downstream LDR	AcDx-10734-PTK2B-Dn	/5Phos/GTTGTATATTTATTTGTTCGGTCGATTTATTTGTATTTGTCGTCGTGGTTAACAGAGGACAGGCCA	66	4344
Real-Time Probe	AcDx-10735-PTK2B-RT-Pb	/56-FAM/AACGGAATC/ZEN/GTTGTATATTTATTTGTTCGGTCGATTTATTTGTA/3IABkFQ/	44	4345
Tag Forward Primer	AcDx-10736-PTK2B-RT-FP	TTTTCGGCATCCGCTTCCA	19	4346
Tag Reverse Primer	AcDx-10737-PTK2B-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	4347
Downstream PCR Primer	AcDx-10738-PTK2B-PCR-V	TGGCCTGTCCTCTGTTAACCACCGCCAAATAAACCGAAACGATGrACAAG/3SpC3/	49	4348
HIST1H2BI				
Forward PCR Primer	AcDx-10741-HIST1H2BI-FP	GGTTGTGCGTTTGTTGTTATTCrGGGAG/3SpC3/	27	4349
Reverse PCR Primer	AcDx-10742-HIST1H2BI-RP	GGTGTCGTGGTAAAAAAACCTTTAAATCGTTAACGCrUTTTG/3SpC3/	41	4350
Upstream LDR	AcDx-10743-HIST1H2BI-Up	TCCGGGTATACACTGTCCCAGTTTGTTGTTATTCGAGGAGTTGGTTAAACACrGCGAC/3SpC3/	57	4351
Downstream LDR	AcDx-10744-HIST1H2BI-Dn	/5Phos/GCGGTGTCGGAGGGTATTAAGGCGTGGTTAACAGAGGACAGGCCA	45	4352
Real-Time Probe	AcDx-10745-HIST1H2BI-RT-Pb	/56-FAM/AATAAACAC/ZEN/GCGGTGTCGGAGGGTA/3IABkFQ/	25	4353
Tag Forward Primer	AcDx-10746-HIST1H2BI-RT-FP	TCCGGGTATACACTGTCCCA	20	4354
Tag Reverse Primer	AcDx-10747-HIST1H2BI-RT-RP	TGGCCTGTCCTCTGTTAACCA	21	4355
Downstream PCR Primer	AcDx-10748-HIST1H2BI-PCR-V	TGGCCTGTCCTCTGTTAACCACCCAAATAACTAATTTACTTAAAACTAATATACTTAATAACTGrCCTTG/3SpC3/	69	4356

PACS2				
Forward PCR Primer	AcDx-10751-PACS2-FP	GGTGGGTGGCGGTTCrGGGTA/3SpC3/	20	4357
Reverse PCR Primer	AcDx-10752-PACS2-RP	GGTGTCGTGGAACATACATAATAAAAAAACCTCGTCCTCrUTACT/3SpC3/	44	4358
Upstream LDR	AcDx-10753-PACS2-Up	TGCGACTCTATTCACGTCCAAGGCGGTGGTTTGGGCGCrGCGAC/3SpC3/	43	4359
Downstream LDR	AcDx-10754-PACS2-Dn	/5Phos/GCGGTGGGCGGTATTATTTAGGTTTGTTGCGTTGCTATTTGGTGTACCGCCA	52	4360
Real-Time Probe	AcDx-10755-PACS2-RT-Pb	/56-FAM/AATGGGCGC/ZEN/GCGGTGGG/3IABkFQ/	17	4361
Tag Forward Primer	AcDx-10756-PACS2-RT-FP	TGCGACTCTATTCACGTCCAA	21	4362
Tag Reverse Primer	AcDx-10757-PACS2-RT-RP	TGGCGGTACACCAAATAGCAA	21	4363
Downstream PCR Primer	AcDx-10758-PACS2-PCR-V	TGGCGGTACACCAAATAGCAACCAACATCGCAACAAACCTAAATAATACTGrCCCAT/3SpC3/	56	4364
ATL1				
Forward PCR Primer	AcDx-10761-ATL1-FP	GGCGAGATTTAGTATGTTTATATCGCrGAGTA/3SpC3/	31	4365
Reverse PCR Primer	AcDx-10762-ATL1-RP	GGTGTCGTGGCGAACGAAACCGCGATTTCrUTCCA/3SpC3/	34	4366
Upstream LDR	AcDx-10763-ATL1-Up	TTGTGCAGAGCGAACAACAAGAGTGGGTGGGCGGTGTTCACrGTAAA/3SpC3/	46	4367
Downstream LDR	AcDx-10764-ATL1-Dn	/5Phos/GTAGGCGGAGAAGAACGGGCGTAGTTGCTATTTGGTGTACCGCCA	45	4368
Real-Time Probe	AcDx-10765-ATL1-RT-Pb	/56-FAM/TTTGTTCAC/ZEN/GTAGGCGGAGAAGAACGG/3IABkFQ/	27	4369
Tag Forward Primer	AcDx-10766-ATL1-RT-FP	TTGTGCAGAGCGAACAACAA	20	4370
Tag Reverse Primer	AcDx-10767-ATL1-RT-RP	TGGCGGTACACCAAATAGCAA	21	4371
Downstream PCR Primer	AcDx-10768-ATL1-PCR-V	TGGCGGTACACCAAATAGCAACGATTTCTTCCGCATCGCTATGrCCCGC/3SpC3/	48	4372
RASSF10				
Forward PCR Primer	AcDx-10771-RASSF10-FP	GGGTATTTTGGGTAGAGTTAGAGCrGGCGA/3SpC3/	29	4373
Reverse PCR Primer	AcDx-10772-RASSF10-RP	GGTGTCGTGGCCCGAACGACGAACGCrUCCCA/3SpC3/	31	4374
Upstream LDR	AcDx-10773-RASSF10-	TCCAAACGATTAGGAGCGTCAAGAGTTAGAGCGGCGGGAATCrGGTCC/3SpC3/	47	4375

	Up			
Downstream LDR	AcDx-10774-RASSF10-Dn	/5Phos/GGTTTTGGGCGCGTTGTTTCGGGTTGGACAGAGGTATACGCCCA	44	4376
Real-Time Probe	AcDx-10775-RASSF10-RT-Pb	/56-FAM/TTGGGAATC/ZEN/GGTTTTGGGCGCGTTG/3IABkFQ/	25	4377
Tag Forward Primer	AcDx-10776-RASSF10-RT-FP	TCCAAACGATTAGGAGCGTCAA	22	4378
Tag Reverse Primer	AcDx-10777-RASSF10-RT-RP	TGGGCGTATACCTCTGTCCAA	21	4379
Downstream PCR Primer	AcDx-10778-RASSF10-PCR-V	TGGGCGTATACCTCTGTCCAACGAACGACGAACGCTCCTGrAAACG/3SpC3/	45	4380
VANGL2				
Forward PCR Primer	AcDx-10781-VANGL2-FP	GTAGGGTTGGAGGAGCrGGCGG/3SpC3/	21	4381
Reverse PCR Primer	AcDx-10782-VANGL2-RP	GGTGTCGTGGCGACTATTCCCACAACCGCrCGCCT/3SpC3/	34	4382
Upstream LDR	AcDx-10783-VANGL2-Up	TCCAAACGATTAGGAGCGTCAATTGGAGGAGCGGCGATCTCrGGGCG/3SpC3/	46	4383
Downstream LDR	AcDx-10784-VANGL2-Dn	/5Phos/GGGAAGAGCGGAGTCGTAGATAATGGACTTGGACAGAGGTATACGCCCA	49	4384
Real-Time Probe	AcDx-10785-VANGL2-RT-Pb	/56-FAM/TTCGATCTC/ZEN/GGGAAGAGCGGAGTC/3IABkFQ/	24	4385
Tag Forward Primer	AcDx-10786-VANGL2-RT-FP	TCCAAACGATTAGGAGCGTCAA	22	4386
Tag Reverse Primer	AcDx-10787-VANGL2-RT-RP	TGGGCGTATACCTCTGTCCAA	21	4387
Downstream PCR Primer	AcDx-10788-VANGL2-PCR-V	TGGGCGTATACCTCTGTCCAAGCCGCCCCTCCTACTGrCCGCA/3SpC3/	42	4388
EFCAB4B				
Forward PCR Primer	AcDx-10791-EFCAB4B-FP	GTTTTTCGTGGATTTGTTCGCrGTCGC/3SpC3/	26	4389

Reverse PCR Primer	AcDx-10792-EFCAB4B-RP	GGTGTCGTGGCGCGCGACTCGAAACrGAAAG/3SpC3/	30	4390
Upstream LDR	AcDx-10793-EFCAB4B-Up	TTGCAGCGGGTCACAACAAGGATTTGTTCGCGTCGTTCGCTCrGGTCC/3SpC3/	47	4391
Downstream LDR	AcDx-10794-EFCAB4B-Dn	/5Phos/GGTTTTTAGTTGTTAAGTAGTCGTTTGTTTCGTTCGATTTTTTTCTTGGACAGAGGTATACGCCCA	66	4392
Real-Time Probe	AcDx-10795-EFCAB4B-RT-Pb	/56-FAM/TTTTCGCTC/ZEN/GGTTTTTAGTTGTTAAGTAGTCGTTTGTTTCG/3IABkFQ/	41	4393
Tag Forward Primer	AcDx-10796-EFCAB4B-RT-FP	TTGCAGCGGGTCACAACAA	19	4394
Tag Reverse Primer	AcDx-10797-EFCAB4B-RT-RP	TGGGCGTATACCTCTGTCCAA	21	4395
Downstream PCR Primer	AcDx-10798-EFCAB4B-PCR-V	TGGGCGTATACCTCTGTCCAAGCGCGACTCGAAACGAAAAAAATTGrAACGG/3SpC3/	51	4396
KCNK10				
Forward PCR Primer	AcDx-10801-KCNK10-FP	GGAGAAGTTTATTGTAGTGTTATTTTAGTTTTCrGAAGA/3SpC3/	38	4397
Reverse PCR Primer	AcDx-10802-KCNK10-RP	GGTGTCGTGGACTAACATACCCGCCCAAArCGCGT/3SpC3/	34	4398
Upstream LDR	AcDx-10803-KCNK10-Up	TTCGTGCGTCGTGTAGCAATGTTATTTTAGTTTTCGAAGGCGTGCGCrGTACG/3SpC3/	52	4399
Downstream LDR	AcDx-10804-KCNK10-Dn	/5Phos/GTATATATTTTCGTATATATATATTCGTATATATATTTTCGGGCGCGCGTTGCCCATTTTCTGCACCCA	69	4400
Real-Time Probe	AcDx-10805-KCNK10-RT-Pb	/56-FAM/TTCGTGCGC/ZEN/GTATATATTTTCGTATATATATATTCGTATATATATTTTCGGG/3IABkFQ/	52	4401
Tag Forward Primer	AcDx-10806-KCNK10-RT-FP	TTCGTGCGTCGTGTAGCAA	19	4402
Tag Reverse Primer	AcDx-10807-KCNK10-RT-RP	TGGGTGCAGAAAATGGGCAA	20	4403
Downstream PCR Primer	AcDx-10808-KCNK10-PCR-V	TGGGTGCAGAAAATGGGCAACCGCCCAAACGCGTGrCCCGG/3SpC3/	40	4404

SLC16A1-S1				
Forward PCR Primer	AcDx-10811-SLC16A1-S1-FP	TGTTTCGTTTTTTTCGTAAAGTTTATTCrGAATC/3SpC3/	33	4405
Reverse PCR Primer	AcDx-10812-SLC16A1-S1-RP	GGTGTCGTGGCGATCCTTACCCCACCTAATAACrUTACA/3SpC3/	38	4406
Upstream LDR	AcDx-10813-SLC16A1-S1-Up	TCCCTTAGAGAGAACGCCCACGTTTTTTTCGTAAAGTTTATTCGAATTTTGTAGTCACrGTTCC/3SpC3/	63	4407
Downstream LDR	AcDx-10814-SLC16A1-S1-Dn	/5Phos/GTTTTGGTTGTGGAGAATGCGGTTTTTCGTGGTGACGTACGAGTGTTCTTA	51	4408
Real-Time Probe	AcDx-10815-SLC16A1-S1-RT-Pb	/56-FAM/AATAGTCAC/ZEN/GTTTTGGTTGTGGAGAATGCG/3IABkFQ/	30	4409
Tag Forward Primer	AcDx-10816-SLC16A1-S1-RT-FP	TCCCTTAGAGAGAACGCCCA	20	4410
Tag Reverse Primer	AcDx-10817-SLC16A1-S1-RT-RP	TAAGAACACTCGTACGTCACCA	22	4411
Downstream PCR Primer	AcDx-10818-SLC16A1-S1-PCR-V	TAAGAACACTCGTACGTCACCACCTTACCCCACCTAATAACTTACGAAAATAATATGrAAAAG/3SpC3/	62	4412
SLC16A1-S2				
Forward PCR Primer	AcDx-10821-SLC16A1-S2-FP	CGTTTTAAGTTTATGAGTAGAAGGGCrGTTGT/3SpC3/	31	4413
Reverse PCR Primer	AcDx-10822-SLC16A1-S2-RP	GGTGTCGTGGAAACGAAACAACCCGCAAACrUCCTG/3SpC3/	35	4414
Upstream LDR	AcDx-10823-SLC16A1-S2-Up	TCTCATACCAGACGCGGTAACGGCGTTGCGGCGAACrGTTAA/3SpC3/	41	4415
Downstream LDR	AcDx-10824-SLC16A1-S2-Dn	/5Phos/GTTGGTAGTTATAGGTTGAATGATAAAGAGTTTTCGTTAGTATTATTTTTAGTTCGTGTCGCTGTGCTTA	70	4416
Real-Time Probe	AcDx-10825-SLC16A1-S2-RT-Pb	/56-FAM/TTGGCGAAC/ZEN/GTTGGTAGTTATAGGTTGAATGATAAAGAGT/3IABkFQ/	40	4417
Tag Forward Primer	AcDx-10826-SLC16A1-S2-RT-FP	TCTCATACCAGACGCGGTAAC	21	4418
Tag Reverse Primer	AcDx-10827-SLC16A1-S2-RT-RP	TAAGCACAGCGACACGAAC	19	4419

Downstream PCR Primer	AcDx-10828-SLC16A1-S2-PCR-V	TAAGCACAGCGACACGAACAACCCGCAAACTCCTAAAAATAATACTAATGrAAAAT/3SpC3/	55	4420
Alternate Group 5 Markers				
TFR2-S2				
Forward PCR Primer	AcDx-10831-TFR2-S2-FP	GGGTGAGTTTTGAGGAGCrGGGAC/3SpC3/	23	4421
Reverse PCR Primer	AcDx-10832-TFR2-S2-RP	GGTGTCGTGGCGCGCGCGAACGAAArAACGT/3SpC3/	30	4422
Upstream LDR	AcDx-10833-TFR2-S2-Up	TTTTCGGCGGCAGCTAAACGGAGCGAGGTTGGTGGGCrGTTCT/3SpC3/	42	4423
Downstream LDR	AcDx-10834-TFR2-S2-Dn	/5Phos/GTTTCGAGTCGCGTTTTTTTTTTCGCGCGGTTCGTGTCGCTGTGCTTA	48	4424
Real-Time Probe	AcDx-10835-TFR2-S2-RT-Pb	/56-FAM/AAGGTGGGC/ZEN/GTTTCGAGTCGCG/3IABkFQ/	22	4425
Tag Forward Primer	AcDx-10836-TFR2-S2-RT-FP	TTTTCGGCGGCAGCTAAAC	19	4426
Tag Reverse Primer	AcDx-10837-TFR2-S2-RT-RP	TAAGCACAGCGACACGAAC	19	4427
Downstream PCR Primer	AcDx-10838-TFR2-S2-PCR-V	TAAGCACAGCGACACGAACGCGCGAACGAAAAACGTGrCGAAG/3SpC3/	42	4428
FXYD7-S1				
Forward PCR Primer	AcDx-10841-FXYD7-S1-FP	GATTTTTAATAAAACGTGCGTTTTTTTCrGATAA/3SpC3/	33	4429
Reverse PCR Primer	AcDx-10842-FXYD7-S1-RP	GGTGTCGTGGCCTTCCTCTAAAATAAAACGCCCrCCTAT/3SpC3/	38	4430
Upstream LDR	AcDx-10843-FXYD7-S1-Up	TCCGGACCTTCATCCTCCAGCGTTTTTTTCGATAGTATTTTGTCGGTCTCrGGTCC/3SpC3/	55	4431
Downstream LDR	AcDx-10844-FXYD7-S1-Dn	/5Phos/GGTTTTTTAGCGCGAAACGTTAGCGTTATTGGGTGGGCAGGAACACGATAGTA	53	4432
Real-Time Probe	AcDx-10845-FXYD7-S1-	/56-FAM/AACGGTCTC/ZEN/GGTTTTTTAGCGCGAAACGTTAG/3IABkFQ/	32	4433

	RT-Pb			
Tag Forward Primer	AcDx-10846-FXYD7-S1-RT-FP	TCCGGACCTTCATCCTCCA	19	4434
Tag Reverse Primer	AcDx-10847-FXYD7-S1-RT-RP	TACTATCGTGTTCCTGCCCA	20	4435
Downstream PCR Primer	AcDx-10848-FXYD7-S1-PCR-V	TACTATCGTGTTCCTGCCCACGCCCCCTACTAAAACCCAATAATGrCTGGT/3SpC3/	50	4436
FXYD7-S2				
Forward PCR Primer	AcDx-10851-FXYD7-S2-FP	GGCGGTAGAAATATTTGGAGCrGATGA/3SpC3/	26	4437
Reverse PCR Primer	AcDx-10852-FXYD7-S2-RP	GGTGTCGTGGCCTCCCTCTCCCGCCrUTTCT/3SpC3/	30	4438
Upstream LDR	AcDx-10853-FXYD7-S2-Up	TTAGCCGCCAAACGTACCACGGTAGAAATATTTGGAGCGATGGAACTCrGGTCG/3SpC3/	53	4439
Downstream LDR	AcDx-10854-FXYD7-S2-Dn	/5Phos/GGTTAGGTGTTGTAGTTTTGATACGGCGGTGGTGGGCAGGAACACGATAGTA	52	4440
Real-Time Probe	AcDx-10855-FXYD7-S2-RT-Pb	/56-FAM/TAGGAACTC/ZEN/GGTTAGGTGTTGTAGTTTTGATACGG/3IABkFQ/	35	4441
Tag Forward Primer	AcDx-10856-FXYD7-S2-RT-FP	TTAGCCGCCAAACGTACCA	19	4442
Tag Reverse Primer	AcDx-10857-FXYD7-S2-RT-RP	TACTATCGTGTTCCTGCCCA	20	4443
Downstream PCR Primer	AcDx-10858-FXYD7-S2-PCR-V	TACTATCGTGTTCCTGCCCACCTCCTTCCCTCCCACTGrCCGTG/3SpC3/	43	4444
LDHB				
Forward PCR Primer	AcDx-10861-LDHB-FP	GTAAATGTAATAAGTTTTTTCGTTGAGTCrGTAGC/3SpC3/	34	4445
Reverse PCR Primer	AcDx-10862-LDHB-RP	GGTGTCGTGGTTAACTCTAAACATCCGACCGCrGAACT/3SpC3/	37	4446
Upstream LDR	AcDx-10863-LDHB-Up	TAGCAGCTGAACAACCCAACGTTATTTTTTGTTTTTTGTTTTTTCGTTAGGGCACrGGTCC/3SpC3/	60	4447
Downstream LDR	AcDx-10864-LDHB-Dn	/5Phos/GGTTTTTTTTGCGTTTTAAATTGAGCGGTAAAGTTAGGGTTGTATGGTCGGCATGCTA	58	4448

Real-Time Probe	AcDx-10865-LDHB-RT-Pb	/56-FAM/AAAGGGCAC/ZEN/GGTTTTTTTTGCGTTTTAAATTGAGC/3IABkFQ/	35	4449
Tag Forward Primer	AcDx-10866-LDHB-RT-FP	TAGCAGCTGAACAACCCAAC	20	4450
Tag Reverse Primer	AcDx-10867-LDHB-RT-RP	TAGCATGCCGACCATACAAC	20	4451
Downstream PCR Primer	AcDx-10868-LDHB-PCR-V	TAGCATGCCGACCATACAACTTAACTCTAAACATCCGACCGTGrAACCT/3SpC3/	48	4452
C20orf195				
Forward PCR Primer	AcDx-10871-C20orf195-FP	GGGTTTTTTAGGAAGCGAAGCrGGATA/3SpC3/	26	4453
Reverse PCR Primer	AcDx-10872-C20orf195-RP	GGTGTCGTGGCTTTATAACGCTACGACCCGAArAACGG/3SpC3/	37	4454
Upstream LDR	AcDx-10873-C20orf195-Up	TCTGCCCTTCGCTTCGAACGGAAGCGAAGCGGATGGAAGATTACTCrGGTCT/3SpC3/	51	4455
Downstream LDR	AcDx-10874-C20orf195-Dn	/5Phos/GGTTCGGAGTTTCGGGTTTTTATTTTCGTTTCGGTTGTATGGTCGGCATGCTA	53	4456
Real-Time Probe	AcDx-10875-C20orf195-RT-Pb	/56-FAM/TTATTACTC/ZEN/GGTTCGGAGTTTCGGGTTTTTATTTTC/3IABkFQ/	36	4457
Tag Forward Primer	AcDx-10876-C20orf195-RT-FP	TCTGCCCTTCGCTTCGAAC	19	4458
Tag Reverse Primer	AcDx-10877-C20orf195-RT-RP	TAGCATGCCGACCATACAAC	20	4459
Downstream PCR Primer	AcDx-10878-C20orf195-PCR-V	TAGCATGCCGACCATACAACCTTTATAACGCTACGACCCGAAAATGrAAACA/3SpC3/	51	4460
DUOX1				
Forward PCR Primer	AcDx-10881-DUOX1-FP	CGTTCGTTGGAAGTATTTTCGCrGTTTC/3SpC3/	27	4461
Reverse PCR Primer	AcDx-10882-DUOX1-RP	GGTGTCGTGGACACGACCCGACTCGArCTCAG/3SpC3/	31	4462
Upstream LDR	AcDx-10883-DUOX1-Up	TTGCAAACCACCCGGACAACGTTGGAAGTATTTTCGCGTTTTTTTAAGCrGTTGA/3SpC3/	54	4463
Downstream LDR	AcDx-10884-DUOX1-Dn	/5Phos/GTTAGATTTTATTTCGTTTTATTGCGTTTCGTTTCGTGGTTGTTGGTCAGCATCGACTCC	62	4464

		TA		
Real-Time Probe	AcDx-10885-DUOX1-RT-Pb	/56-FAM/AATTTAAGC/ZEN/GTTAGATTTTATTTCGTTTTATTGCGTTTCGTTT/3IABkFQ/	43	4465
Tag Forward Primer	AcDx-10886-DUOX1-RT-FP	TTGCAAACCACCCGGACAA	19	4466
Tag Reverse Primer	AcDx-10887-DUOX1-RT-RP	TAGGAGTCGATGCTGACCAA	20	4467
Downstream PCR Primer	AcDx-10888-DUOX1-PCR-V	TAGGAGTCGATGCTGACCAACCCGACTCGACTCAACCATGrAAACA/3SpC3/	45	4468
ZNF783				
Forward PCR Primer	AcDx-10891-ZNF783-FP	GTTAATGAAAAGTTGGTATTGGGTCrGGAGA/3SpC3/	30	4469
Reverse PCR Primer	AcDx-10892-ZNF783-RP	GGTGTCGTGGCGCCCACATCCCGAAACrCAAAC/3SpC3/	32	4470
Upstream LDR	AcDx-10893-ZNF783-Up	TTGCAAACCACCCGGACAAGAAAAGTTGGTATTGGGTCGGAAGCrGTTGA/3SpC3/	49	4471
Downstream LDR	AcDx-10894-ZNF783-Dn	/5Phos/GTTAGTTAAGTGGGAGGCGGAGTTTTTATTATCGGTTTTGGTCAGCATCGACTCCTA	57	4472
Real-Time Probe	AcDx-10895-ZNF783-RT-Pb	/56-FAM/AACGGAAGC/ZEN/GTTAGTTAAGTGGGAGGC/3IABkFQ/	27	4473
Tag Forward Primer	AcDx-10896-ZNF783-RT-FP	TTGCAAACCACCCGGACAA	19	4474
Tag Reverse Primer	AcDx-10897-ZNF783-RT-RP	TAGGAGTCGATGCTGACCAA	20	4475
Downstream PCR Primer	AcDx-10898-ZNF783-PCR-V	TAGGAGTCGATGCTGACCAACCGAAACCAAATCCCCATTAACTGrATAAC/3SpC3/	49	4476
MYADM				
Forward PCR Primer	AcDx-10901-MYADM-FP	GGGACGATTGTAGAGCGCrGGTGG/3SpC3/	23	4477
Reverse PCR Primer	AcDx-10902-MYADM-RP	GGTGTCGTGGGAAATATCGAACCCCCGAAArAAAAG/3SpC3/	35	4478
Upstream LDR	AcDx-10903-MYADM-Up	TAAGACGTATGCTAGCGCCAAGATTGTAGAGCGCGGTGAGTGGACrGCGAC/3SpC3/	50	4479
Downstream LDR	AcDx-10904-MYADM-Dn	/5Phos/GCGGTTAGAGGGAGGGATTTAGGCGTTTTAGTTGGTCAGCATCGACTCCTA	51	4480

Real-Time Probe	AcDx-10905-MYADM-RT-Pb	/56-FAM/AAAGTGGAC/ZEN/GCGGTTAGAGGGAGGG/3IABkFQ/	25	4481
Tag Forward Primer	AcDx-10906-MYADM-RT-FP	TAAGACGTATGCTAGCGCCAA	21	4482
Tag Reverse Primer	AcDx-10907-MYADM-RT-RP	TAGGAGTCGATGCTGACCAA	20	4483
Downstream PCR Primer	AcDx-10908-MYADM-PCR-V	TAGGAGTCGATGCTGACCAACCCGAAAAAAAACAAAACGAAAACCTAAAATGrCCTAG/3SpC3/	57	4484
CDKL2-S1				
Forward PCR Primer	AcDx-10911-CDKL2-S1-FP	GGTTTTTTATTCGTTTCGAGAGCrGTTAA/3SpC3/	28	4485
Reverse PCR Primer	AcDx-10912-CDKL2-S1-RP	GGTGTCGTGGAAAAACAAAACCTAAACCGAACCrCTAAG/3SpC3/	38	4486
Upstream LDR	AcDx-10913-CDKL2-S1-Up	TACATGCCATCCCACGACAGTTAGGTTTTTTTTTTTTTGATTGGTCGAATCrGAGCT/3SpC3/	56	4487
Downstream LDR	AcDx-10914-CDKL2-S1-Dn	/5Phos/GAGTCGTTACGAGTTATGATTGGTTTAGGGTTAATTATTTCGTGTGTCGGAGCGGTTACTA	61	4488
Real-Time Probe	AcDx-10915-CDKL2-S1-RT-Pb	/56-FAM/TTTCGAATC/ZEN/GAGTCGTTACGAGTTATGATTGGTTTAGG/3IABkFQ/	38	4489
Tag Forward Primer	AcDx-10916-CDKL2-S1-RT-FP	TACATGCCATCCCACGACA	19	4490
Tag Reverse Primer	AcDx-10917-CDKL2-S1-RT-RP	TAGTAACCGCTCCGACACA	19	4491
Downstream PCR Primer	AcDx-10918-CDKL2-S1-PCR-V	TAGTAACCGCTCCGACACACAAAACCTAAACCGAACCCTAAATAAAAAAATGrAAATG/3SpC3/	57	4492
CDKL2-S2				
Forward PCR Primer	AcDx-10921-CDKL2-S2-FP	TTAATTATTTCGTTTTTTATTTAGGGTTCrGGTTC/3SpC3/	35	4493
Reverse PCR Primer	AcDx-10922-CDKL2-S2-RP	GGTGTCGTGGCTCCTCCCGCTAAAAAAAACArAAAAG/3SpC3/	36	4494

Upstream LDR	AcDx-10923-CDKL2-S2-Up	TCCAAACAAGCTGATCCGTACAGGTTTTGTTTTTTGATACGTAGAGGTTTCGCrGTTCC/3SpC3/	58	4495
Downstream LDR	AcDx-10924-CDKL2-S2-Dn	/5Phos/GTTTTTATACGTTTTGGTTTTCGTTAGGAGGCGTTTTTTGTGTCGGAGCGGTTACTA	57	4496
Real-Time Probe	AcDx-10925-CDKL2-S2-RT-Pb	/56-FAM/TTGTTTCGC/ZEN/GTTTTTATACGTTTTGGTTTTCGTTAGGAG/3IABkFQ/	39	4497
Tag Forward Primer	AcDx-10926-CDKL2-S2-RT-FP	TCCAAACAAGCTGATCCGTACA	22	4498
Tag Reverse Primer	AcDx-10927-CDKL2-S2-RT-RP	TAGTAACCGCTCCGACACA	19	4499
Downstream PCR Primer	AcDx-10928-CDKL2-S2-PCR-V	TAGTAACCGCTCCGACACATCCTCCCGCTAAAAAAAACAAAAAAATGrCCTCT/3SpC3/	52	4500
HIST3H2BB				
Forward PCR Primer	AcDx-10931-HIST3H2BB-FP	GGGAATTGTAGTTTCGCGCrGCGAT/3SpC3/	24	4501
Reverse PCR Primer	AcDx-10932-HIST3H2BB-RP	GGTGTCGTGGCTCGCTTTTCGATTACCGTTATCrUTTTC/3SpC3/	38	4502
Upstream LDR	AcDx-10933-HIST3H2BB-Up	TAGGGCGACAGTTACCACAAGAGCGCGATTTAGTTTTGGTGCrGCGCC/3SpC3/	47	4503
Downstream LDR	AcDx-10934-HIST3H2BB-Dn	/5Phos/GCGTTTTGTTATTTTGTTTATTACGATCGGATATTTTCGAGTTAAGTTGTGGGTCTCGC TCGTATA	66	4504
Real-Time Probe	AcDx-10935-HIST3H2BB-RT-Pb	/56-FAM/AATTGGTGC/ZEN/GCGTTTTGTTATTTTGTTTATTACGATCG/3IABkFQ/	38	4505
Tag Forward Primer	AcDx-10936-HIST3H2BB-RT-FP	TAGGGCGACAGTTACCACAA	20	4506
Tag Reverse Primer	AcDx-10937-HIST3H2BB-RT-RP	TATACGAGCGAGACCCACAA	20	4507
Downstream PCR Primer	AcDx-10938-HIST3H2BB-PCR-V	TATACGAGCGAGACCCACAATTCGATTACCGTTATCTTTTTTCCTTAACTTGrAAAAC/3SpC3/	57	4508
SLC22A20				

Forward PCR Primer	AcDx-10941-SLC22A20-FP	GACTCCATAAAAAAATAACCCCACCrGATTT/3SpC3/	30	4509
Reverse PCR Primer	AcDx-10942-SLC22A20-RP	GGTGTCGTGGCGAATACGATTAACAAATCCCGArCGTAT/3SpC3/	38	4510
Upstream LDR	AcDx-10943-SLC22A20-Up	TCCGACTTTAGTGCGTCACAAGGGTTATTTTTTTATGGAGTCGATTCGACTCrGTTCG/3SpC3/	57	4511
Downstream LDR	AcDx-10944-SLC22A20-Dn	/5Phos/GTTTAAAGTTGTTTCGGGAATCGTAGGTTTGGTTCGTTGTGGGTCTCGCTCGTATA	56	4512
Real-Time Probe	AcDx-10945-SLC22A20-RT-Pb	/56-FAM/CCTCGACTC/ZEN/GTTTAAAGTTGTTTCGGGAATCGTAGG/3IABkFQ/	36	4513
Tag Forward Primer	AcDx-10946-SLC22A20-RT-FP	TCCGACTTTAGTGCGTCACAA	21	4514
Tag Reverse Primer	AcDx-10947-SLC22A20-RT-RP	TATACGAGCGAGACCCACAA	20	4515
Downstream PCR Primer	AcDx-10948-SLC22A20-PCR-V	TATACGAGCGAGACCCACAAGATTAACAAATCCCGACGTACGTGrCCTAT/3SpC3/	49	4516
RNASE4				
Forward PCR Primer	AcDx-10951-RNASE4-FP	GAGGCGCGGTTTTAATTACrGTAGG/3SpC3/	24	4517
Reverse PCR Primer	AcDx-10952-RNASE4-RP	GGTGTCGTGGCCTACTTTACATCTAAATAAAATTAACGTCrCTTAC/3SpC3/	45	4518
Upstream LDR	AcDx-10953-RNASE4-Up	TCAAACAAAGGCGACCACAACCGGTTTTAATTACGTAGAGGTTGGTACATCrGATCC/3SpC3/	56	4519
Downstream LDR	AcDx-10954-RNASE4-Dn	/5Phos/GATTTTTTATATTTTATTACGTTTTTATTTTCGTTCGTGTATATATATTTATATAAGGTTGTCGCATAGGCAGTTCATA	79	4520
Real-Time Probe	AcDx-10955-RNASE4-RT-Pb	/56-FAM/AAGTACATC/ZEN/GATTTTTTATATTTTATTACGTTTTTATTTTCGTTCGTGTATATA/3IABkFQ/	54	4521
Tag Forward Primer	AcDx-10956-RNASE4-RT-FP	TCAAACAAAGGCGACCACAAC	21	4522
Tag Reverse Primer	AcDx-10957-RNASE4-RT-RP	TATGAACTGCCTATGCGACAAC	22	4523

NFIA				
Forward PCR Primer	AcDx-10961-NFIA-FP	TGGTTAATGGCGTTTTTTTATTAATGCrGTTAG/3SpC3/	32	4524
Reverse PCR Primer	AcDx-10962-NFIA-RP	GGTGTCGTGGCGCCGAAACACGTACGAArATATT/3SpC3/	33	4525
Upstream LDR	AcDx-10963-NFIA-Up	TAAACAATGAGACCCGCTGAACGGTTAATGGCGTTTTTTTATTAATGCGTTAATTAATGTCrGGGAT/3SpC3/	66	4526
Downstream LDR	AcDx-10964-NFIA-Dn	/5Phos/GGAGCGACGGATTGCGGAATAGTAGGTTGTCGCATAGGCAGTTCATA	47	4527
Real-Time Probe	AcDx-10965-NFIA-RT-Pb	/56-FAM/CCTAATGTC/ZEN/GGAGCGACGGATTGCG/3IABkFQ/	25	4528
Tag Forward Primer	AcDx-10966-NFIA-RT-FP	TAAACAATGAGACCCGCTGAAC	22	4529
Tag Reverse Primer	AcDx-10967-NFIA-RT-RP	TATGAACTGCCTATGCGACAAC	22	4530
Downstream PCR Primer	AcDx-10968-NFIA-PCR-V	TATGAACTGCCTATGCGACAACAACACGTACGAAATATCTGrAAAT/3SpC3/	45	4531

IDT Abbreviation	Modifications
/5Phos/	5' Phosphorylation
rX (X=A,C,G,U)	RNA Base
/3spC3/	3' C3 DNA Spacer
/56-FAM/	5' 6-FAM™ Fluorescent Tag
/Zen/	Internal Quencher
/3IABkFQ/	3' Iowa Black® FQ Quencher

Example 1: Detection of VIM Promoter Methylation in HT29 Colorectal Cancer Cell Line Using exPCR-LDR-qPCR at the Single Molecule Level

[0401] 500 ng HT29 cell line genomic DNA was digested with 10 units of restriction enzyme Bsh1236I in 20 µl of reaction solution containing 1xCutSmart buffer (50 mM Potassium Acetate, 20 mM Tris-Acetate, 10 mM Magnesium Acetate, 100 µg/ml BSA, pH 7.9 at 25°C). The digestion reactions were carried out at 37°C for 1 hour, followed by enzyme inactivation at 80°C for 20 min.

[0402] The digests were then bisulfite converted using EZ DNA Methylation-Lightning kit from Zymo Research Corporation (Irvine, CA). In this reaction, 130 µl of Lightning Conversion Reagent was added to 20 µl of digested genomic DNA. The reaction was incubated at 98°C for 8 minutes, 54°C for one hour, and stopped at 4°C.

[0403] 600 µl of M-Binding Buffer was added to a Zymo-Spin TM Column and the column inserted into a collection tube. The digested DNA (150 µl) from previous step was then loaded into the column. Capped, the column was inverted several times to mix the solution, then centrifuged at full speed (> 10,000xg) for 30 sec. After discarding the flow through, 100 µl of M-washing buffer was added to the column, followed by another round of centrifugation at full speed for 30 second. The resulting flow through was then discarded. 200 µl of L-Desulphonation Buffer was then added to the column, and the column allowed to stand at room temperature for 15-20 minutes. After the incubation, the column was centrifuged at full speed for 30 seconds. 200 µl of M-Wash Buffer was then added to the column, followed by centrifugation at full speed for 30 second. The flow through was discarded. This wash step was repeated once. The column was then inserted into a 1.5 ml micro centrifuge tube, and 10 µl of M-Elution buffer was added to the column matrix. Finally, the column was centrifuged at full speed for 30 second to elute the DNA.

[0404] All the necessary primers (listed in Table 42) were purchased from Integrated DNA Technologies Inc. (IDT) (Coralville, IA), except for LNA1 and LNA2, which was purchased from Exiqon Inc. (Woburn, MA), and PNA, which was purchased from PNA Bio (Thousand Oaks, CA).

[0405] A 130 µl-volume PCR reaction was set up as follows: 23.14 µl of nuclease free water (IDT), 26 µl of Gotaq Flexi buffer 5x without Magnesium (Pro- mega, Madison, Wis.), 10.4 µl of $MgC1_2$ at 25 mM (Pro- mega, Madison, Wis.), 2.6 µl of dNTPs (10 mM each of dATP, dCTP, dGTP and dUTP) (Promega, Madison, Wis.), 3.25 µl of iCDx-2031-VIM-S3-FP forward primer at 2 µM, 3.25 µl of iCDx-2032-VIM-S3-PR reverse primer at 2 µM, 16.25 µl of iCDx-VIM-S3-LNA2 blocking primer at 2 µM, 3.25 µl of RNAseH2 (IDT) at 20 mU/µl , 2.86 µl

of Klentaql polymerase (DNA Polymerase Technology, St. Louis, Mo.) mixed with Plantinim Taq Antibody (Invitrogen, Carlsbad, Calif.) (the mixture is prepared by adding 0.3 µl of Klentaql polymerase at 50 U/µl to 3 µl of Platinum Taq Antibody at 5 U/µl), and 39 µl of corresponding template. 39 µl of templates contains: (1) 0.070 ng (20 copy of Genome Equivalent GE) HT-29 DNA mixed with 9 ng (2500 GE) Roche hgDNA. (2) 0.035 ng (10 GE) HT-29 cell line DNA mixed with 9 ng (2500 GE) Roche hgDNA. (3) 9 ng (2500 GE) Roche DNA, (4) nuclease free water for the Non Template Control (NTC).

[0406] Each 130 µl PCR mixture was divided into 12 tubes, 10 µl each, and then the PCR reactions were run in a Proflex PCR system thermo-cycler (Applied Biosystems/ ThermoFisher; Waltham, Mass.) with the following program: 2 min at 95 °C, 35 cycles of (10 sec at 94 °C, 30 sec at 60 °C, and 30 sec at 72° C), 10 min at 99.5° C, and a final hold at 4° C.

[0407] The LDR step was performed in a 10 µl reaction prepared by adding: 5.82 µl of nuclease free water (IDT), 1 µl of 10X AK16D ligase reaction buffer [1X buffer contains 20 mM Tris-HCl at pH 8.5 (Bio-Rad, Hercules, CA), 5 mM $MgCl_2$ (Sigma-Aldrich, St. Louis, MO), 50 mM KCl (Sigma-Aldrich), 10 mM DTT (Sigma-Aldrich) and 20 µg/ml of BSA (Sigma-Aldrich), 0.25 µl of DTT (Sigma-Aldrich) at 40 mM, 0.25 µl of NAD^+ (Sigma-Aldrich) at 40 mM, 0.25 µl of RNAseH2 (IDT) at 20 mU/µl, 0.2 µl of iCDx-2033-Vim-S3-Up probe at 500 nM, 0.2 µl of iCDx-2034A-Vim-S3-Dn probe at 500 nM, 0.028 µl of purified AK16D ligase at 8.8 µM, and 2 µl of PCR reaction. LDR reactions were run in a Proflex PCR system thermocycler (Applied Biosystems) with the following program: 20 cycles of (10 sec at 94°C, and 4 min at 60°C) followed by a final infinite hold at 4°C.

[0408] The qPCR step was performed in a 10 µl of reaction mixture prepared by adding: 1.5 µl of nuclease free water (IDT), 5 µl of 2X TaqMan® Fast Universal PCR Master Mix (consists of Amplitaq, UDG and dUTP) from Applied Biosystems (Life Technologies, Grand Island, NY), 1 µl of iCDx-2036-Vim-S3-RT-FP forward primer at 2.5 µM, 1 µl of iCDx-2037-Vim-S3-RT-RP reverse primer at 2.5 µM, 0.5 µl of iCDx-2035-Vim-S3-RT-Pb Taqman™ probe at 5 µM, and 1 µl of LDR reaction products. The qPCR reactions were run in a ViiA7 real-time thermo-cycler from Applied Biosystems (Life Technologies, Grand Island, NY), using MicroAmp® Fast-96-Well Reaction 0.1 ml plates sealed with MicroAmp™ Optical adhesive film (Applied Biosystems), with the following settings: fast block, Standard curve as experiment type, ROX as passive reference, Ct as quantification method (automatic threshold, but adjusted to 0.05 when needed), TAMRA as reporter, and NFQ-MGB as quencher. The program was set at: 2 min at 50°C, and 40 cycles of (1 sec at 95°C, and 20 sec at 60°C).

[0409] The pixel experiment results are shown in Figure 72, while the resulting Ct values for different conditions are listed in Table 43. The following primer sequences from Table 58 were used in this Example: SEQ ID NOs.:1-10.

Table 58. Results of Pixel Experiments to Detect Methylation in HT-29 DNA in the Background of Roche Human Genomic DNA. Ct Values in the Different Conditions.

Tem-plates	GE per 130 µl of PCR	1	2	3	4	5	6	7	8	9	10	11	12	Total No. Amplif.
HT29+ Roche hgDNA	20 GE +2500 GE	21.5	17.5	14.5	16.0	15.9	14.4	15.0	14.5	14.3	14.2	16.1	13.5	12
HT29+ Roche hgDNA	10 GE+2500 GE	No Ct	15.2	No Ct	16.2	No Ct	15.8	16.3	No Ct	20.3	16.1	No Ct	No Ct	6
Roche hgDNA	2500 GE	No Ct	No Ct	No Ct	No Ct	No Ct	38.5	No Ct	36.8	No Ct	No Ct	37.7	36.6	0
NTC	No Ct	No Ct	37.6	No Ct	37.9	38.3	No Ct	No Ct	No Ct	36.8	38.0	39.5	No Ct	0

Example 2: Multiplexed Detection of 10 CRC Methylation Markers by PCR-LDR-qPCR on Bisulfite Converted HT29 Cell Line DNA

[0410] The experiment started with 500 ng of HT29 cell line genomic DNA digested with 12 units of the restriction enzyme Bsh1236I, in 20 µl of reaction solution containing 1xCutSmart buffer (50 mM Potassium Acetate, 20 mM Tris-Acetate, 10 mM Magnesium Acetate, 100 µg/ml BSA, pH 7.9 at 25 °C). The digestion reactions were carried out at 37°C for 1 hour, followed by enzyme inactivation at 80°C for 20 min.

[0411] For the bisulfite conversion reaction, 130 µl of Lightning Conversion Reagent (EZ DNA Methylation-Lightning kit from Zymo Research Corporation; Irvine, CA) was first added to 20 µl of digested genomic DNA. This is followed by incubation at 98°C for 8 minutes, 54°C for one hour, and stopped at 4°C. 600 µl of M-Binding Buffer was then added to a Zymo-Spin TM column placed inside a collection tube. 150 µl of digested DNA reaction mixture was then loaded into the Zymo-Spin IC column (containing the M-Binding Buffer), capped, and the solution mixed by inverting the column several times. After centrifugation at full speed (> 10,000xg) for 30 sec, the flow through was discarded. 100 µl of M-Wash buffer was added to the column, followed by another round of centrifugation (full speed for 30 seconds). After discarding the flow through, 200 µl of L-Desulfonation Buffer was added into the column, which was allowed to stand at room temperature for 15-20 minutes, prior to centrifugation at full speed

for 30 seconds. 200 µl of M-Wash Buffer was added anew, followed by centrifugation at full speed for 30 seconds, and discarding the flow through. This wash step cycle was repeated once. The column was placed into a 1.5 ml micro centrifuge tube, to which 10 µl of M-Elution buffer was added, followed by full speed (30 s) centrifugation to elute the DNA.

[0412] Most of the primers used in this particular experiment were purchased from Integrated DNA Technologies Inc. (IDT; Coralville, IA). The primers LNA1 and LNA2 were purchased from Exiqon Inc. (Woburn, MA), while the primer PNA was purchased from PNA Bio (Thousand Oaks, CA). The following primer sequences from Table 45 were used in PCR-LDR-qPCR multiplex methylation assays for 10 genes using HT29 cell line DNA as template: SEQ ID NOs.:1, 2, and 6-75.

[0413] **PCR reaction**. The PCR reaction was performed by mixing the following (in a 10 µl volume): 2 µl of Gotaq Flexi buffer 5x without Magnesium (Promega, Madison, Wis.), 0.8 µl of $MgCl_2$ at 25 mM (Promega, Madison, Wis.), 0.2 µl of dNTPs (with dATP, dCTP, dGTP and dUTP; 10 mM each) (Promega, Madison, Wis.), 0.125 µl of 10 specific forward and reverse primers at 4 µM each. 0.625 µl of Blocker at 4 µM (if available), 0.25 µl of RNAseH2 (IDT) at 20 mU/µl (diluted in RNAseH2 dilution buffer from IDT original RNAseH2 at 2 U/µl), 0.22 µl of Klentaql polymerase (DNA Polymerase Technology, St. Louis, Mo.) mixed with Platinum Taq Antibody (Invitrogen/Thermo Fisher, Waltham, Mass.) (the mixture is prepared by adding 0.02 µl of Klentaql polymerase at 50 U/µl to 0.2 µl of Platinum Taq Antibody at 5 U/µl), DNA template: (1) 10 ng of bisulfite converted HT29 DNA, (2) 0.07ng of bisulfite converted HT29 DNA and 10 ng bisulfite converted normal DNA, (3) 10 ng bisulfite converted normal DNA. PCR reactions were carried out in a ProFlex PCR system thermocycler (Applied Bio-systems/ThermoFisher, Waltham, Mass.) and run with the following program: 94°C for 2 min, 40 cycles of (20 sec at 94° C., 40 sec at 60° C. and 30 sec at 72° C.), 10 min at 99.5° C to inactivate Klan Taq Polymerase, and a final hold at 4° C.

[0414] **LDR step.** The LDR step was performed in a 10 µl reaction prepared by adding: 2.02 µl of nuclease free water (IDT), 1 µl of 10X AK16D ligase reaction buffer [lX buffer contains 20 mM Tris-HCI, pH 8.5 (Bio-Rad, Hercules, Calif.), 5 mM $MgCl_2$ (Sigma-Aldrich, St. Louis, Mo.), 50 mM KCl (Sigma-Aldrich, St. Louis, Mo.), 10 mM DTT (Sigma-Aldrich, St. Louis, Mo.) and 20 µg/ml of BSA (Sigma- Aldrich, St. Louis, Mo.)], 0.25 µl of DTT (Sigma-Aldrich, St. Louis, Mo.) at 40 mM, 0.2 µl of NAD+ (Sigma-Aldrich, St. Louis, Mo.) at 50 mM, 0.25 µl of RNAseH2 (IDT) at 20 mU/µl, 0.2 µl of corresponding 10 gene LDR upstream probes at 500 nM, 0.2 µl of corresponding 10 gene LDR downstream probes at 500 nM, 0.284 µl of purified AK16D ligase at 0.88 µM, and 2 µl of PCR reaction products from previous step. LDR reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/ThermoFisher;

Waltham, Mass.) using the following program: 20 cycles of (10 sec at 94° C., and 4 min at 60° C.) followed by a hold at 4° C.

[0415] **qPCR step.** The qPCR reactions were performed as uniplex, wherein each reaction mixture contains only one set of gene-specific primers and probe. A 10 µl reaction mixture was prepared by mixing: 1.5 µl of nuclease free water (IDT), 5 µl of 2x TaqMan® Fast Universal PCR Master Mix (Fast amplitaq, UDG and dUTP) from Applied Biosystems (Applied Biosystems/ThermoFisher; Waltham, Mass.), 1 µl of TaqMan™ Assay forward primer at 2.5 µM, 1 µl of Taqman™ reverse primer at 2.5µM, 0.5 µl of Taqman™ probe at 5 µM, and 1 µl of LDR reaction products. qPCR reactions were run in a ViiA7 real-time thermo-cycler from Applied Biosystems (Applied Biosystems/Thermo-Fisher; Waltham, Mass.), using MicroAmp® Fast-96-Well Reaction 0.1 ml plates sealed with MicroAmp™ Optical adhesive film (Applied Biosystems/ThermoFisher; Waltham, Mass.), with the following setting: fast block, Standard curve as experiment type, ROX as passive reference, Ct as quantification method (automatic threshold, but adjusted to 0.05 when needed), TAMRA as reporter, and NFQ-MGB as quencher. The specific program used was the following: 2 min at 50° C, and 45 cycles of (1 sec at 95° C, and 20 sec at 60° C). Results are shown in Figure 73 and Ct Values are shown in Table 59.

Table 59. Ct Values for Genes Assayed in Example 2.

Ct Value	CLIP4-R-S1	GSG1L-F-S1	PP1R16B	KCNA3	VIM-S3	LONFR2-R-S1	TBC1D10C-F-S1	GDF6-F-S1	CLIP4-F-S2	SEPT9-F-S1
10 ng HT29 DNA	7.2	9.1	15.8	19.6	19.7	25.1	14.2	17.3	20.9	11.2
0.07 ng HT29 DNA +10 ng Roche	10.2	18.8	23.2	27.8	27.4	31.9	15.6	26.6	27.9	17.4
10 ng Roche DNA	No Ct	No Ct	35.6	35.8	No Ct	38.4	36.7	No Ct	No Ct	No Ct
NTC_PCR	No Ct	No Ct	36.1	36.7	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct
NTC_LDR	No Ct	38.2	36.0	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	37.7
NTC_Taq Man™	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct

Note: NTC: No Template Control

Example 3: Multiplexed Detection of 7 CRC Methylation Markers Using ex-PCR-LDR-qPCR on Bisulfite Converted HT29 Cell line DNA

[0416] In a 20 µl reaction volume, 500 ng HT29 cell line genomic DNA was mixed with 12 units of restriction enzyme Bsh1236I, in 1xCutSmart buffer (50 mM Potassium Acetate, 20

mM Tris-Acetate, 10 mM Magnesium Acetate, 100 µg/ml BSA, pH7.9@25°C). This digestion reactions were carried out at 37°C for 1 hour, followed by enzyme inactivation at 80°C for 20 min. Bisulfite conversion was carried out using the Cells-to-CpG Bisulfite Conversion kit from Applied Biosystem Corporation (Carlsbad, Calif.). For this reaction, 5 µl of Denaturation Reagent was added to 45 µl of restriction digested or methyl enriched genomic DNA, and the mixture was incubated at 50°C for 10 min (to denature the DNA). 100 µl of Conversion Reagent was added to the mixture, followed by incubation in a thermal cycler with the following cycling conditions : 65°C 30 min, 90°C 30 sec, 65°C 30 min, 90°C 30 sec, 65°C 30 min. 150 µl of the bisulfite converted DNA mixture was then mixed with 600 µl of binding buffer in the binding Column. The column was centrifuged at 10,000 rpm for 1 min, and the flow through discarded thereafter. The column was then washed with 600 µl of washing buffer. This was followed with addition of 200 µl of Desulfonation Reagent, and the column incubation at room temperature for 15 min. After spinning, the column was washed again with 400 µl of washing buffer. 50 µl of Elution Buffer was then added to the column to elute the DNA. Since the eluted bisulfite converted DNA was mostly single stranded DNA, it was quantified using Quant-iT Oli Green and Pico Green kits (Life Technologies/ThermoFisher; Waltham, Mass.).

[0417] The necessary primers were mostly purchased from Integrated DNA Technologies Inc. (IDT; Coralville, IA). The LNA1 and LNA2 primers were purchased from Exiqon Inc. (Woburn, MA), while the PNA primer, which was purchased from PNA Bio (Thousand Oaks, CA). The following primer sequences from Table 45 were used in Example 3: SEQ ID NOs.:1, 2, 6-39, 54-60, and 68-75.

[0418] **Linear Amplification step.** The Linear Amplification step was performed in a 25 µl of reaction mixture with: 5 µl of Gotaq Flexi buffer 5X without Magnesium (Promega, Madison, Wis.), 2.5 µl of $MgCl_2$ at 25 mM (Promega, Madison, Wis.), 0.5 µl of dNTPs (with dATP, dCTP, dGTP and dTTP, 10 mM each) (Promega, Madison, Wis.), 1.25 µl of 7 plex gene specific one direction primers (concentration of primer for each gene is 2 µM), 0.625 µl of RNAseH2 (IDT) at 20 mU/µl (diluted in RNAseH2 dilution buffer from IDT), and 0.55 µl of Klentaql polymerase (DNA Polymerase Technology, St. Louis, Mo.) mixed with Platinum Taq Antibody (Invitrogen/Thermo Fisher, Waltham, Mass.) (the mixture is prepared by adding 0.02 µl of Klentaql polymerase at 50 U/µl to 0.2 µl of Platinum Taq Antibody at 5 U/µl), and 14.5 µl of corresponding bisulfite converted genomic DNA (out of 50 µl of eluted DNA after bisulfite conversion.) DNA templates were: (1) 10 ng of bisulfite converted HT29 DNA, (2) 0.1ng bisulfite converted HT29 mixed with 10 ng bisulfite converted Normal DNA from Roche, (3) 10 ng bisulfite converted Normal DNA from Roche. Linear Amplification reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/ThermoFisher, Waltham, Mass.) and

run with the following program: 2 min at 94 °C, 40 cycles of (20 sec at 94 °C, 40 sec at 60 °C. and 30 sec at 72 °C.), a final hold at 4 °C. After the reaction, Platinum Taq antibodies were added in the reaction mixture to inhibit the Klentaq DNA polymerase.

[0419] PCR reaction. The linear amplification products were equally divided into two parts (4-plex for 4 CpG markers, and 3-plex for 3 CpG markers). The PCR step was performed in a 20 µl reaction volume by prepared by mixing: 2 µl of 5X GoTaq Flexi buffer without Magnesium (Promega, Madison, Wis.), 1 µl of $MgCl_2$ at 25 mM (Promega, Madison, Wis.), 0.4 µl of dNTPs (with 10 mM each for dATP, dCTP, dGTP and dUTP) (Promega, Madison, Wis.), 1 µ1 of each of the opposite-strand PCR primer (2 µM concentration), 0.4 µl of Antarctic Thermolabile UDG (1u/µl)(New England Biolab, Ipswich, MA), 0.25 µl of RNAseH2 (IDT) at 20 mU/µl , 0.44 µl of Klentaql polymerase (DNA Polymerase Technology, St. Louis, Mo.) mixed with Platinum Taq Antibody (Invitrogen/Thermo Fisher, Waltham, Mass.) (the mixture is prepared by adding 0.02 µl of Klentaql polymerase at 50 U/µl to 0.2 µI of Platinum Taq Antibody at 5 U/µl), and 12 µl of corresponding linear amplification products . PCR reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/ThermoFisher, Waltham, Mass.) with the following program: 10 min at 37 °C, 40 cycles of (20 sec at 94 °C, 40 sec at 60 °C and 30 sec at 72 °C), 10 min at 99.5 °C, and a final hold at 4° C.

[0420] LDR step. The LDR step was performed in a 20 µl of reaction volume by mixing: 11.6 µl of nuclease free water (IDT), 2 µl of 10X AK16D ligase reaction buffer [lX buffer contains 20 mM Tris-HCI pH 8.5 (Bio-Rad, Hercules, Calif.), 5 mM $MgCl_2$ (Sigma-Aldrich, St. Louis, Mo.), 50 mM KCl (Sigma-Aldrich, St. Louis, Mo.), 10 mM DTT (Sigma-Aldrich, St. Louis, Mo.) and 20 µg/ml of BSA (Sigma- Aldrich, St. Louis, Mo.), 0.5 µl of DTT (Sigma-Aldrich, St. Louis, Mo.) at 40 mM, 0.4 µl of NAD+ (Sigma-Aldrich, St. Louis, Mo.) at 50 mM, 0.5 µl of RNAseH2 (IDT) at 20 mu/µl, 0.4 µl of corresponding 4 plex LDR upstream probes at 500 nM, 0.4 µl of corresponding 4 plex LDR downstream probes at 500 nM, (3 plex for the other LDR reaction), 0.57 µl of purified AK16D ligase at 0.88 µM, and 4 µl of corresponding PCR reaction products from previous step. LDR reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/ThermoFisher; Waltham, Mass.) using the following program: 20 cycles of (10 sec at 94 °C, and 4 min at 60 °C.) followed by a final hold at 4 °C.

[0421] qPCR step. Uni-plex qPCR reactions (*i.e.* each reaction tube contains only the primers and probe specific to a unique CpG site) were performed in a 10 µl reaction mixture containing: 3 µl of nuclease free water (IDT), 5 µl of 2x TaqMan® Fast Universal PCR Master Mix (Fast Amplitaq, UDG and dUTP) from Applied Biosystems (Applied Biosystems/ThermoFisher; Waltham, Mass.), 0.5 µl of mixture of TaqMan™ Assay forward and

reverse primers at 5 μM each, 0.5 μl of Taqman™ probe at 5 μM, and 1 μl of corresponding LDR reaction products. The reactions were run in a ViiA7 real-time thermo-cycler from Applied Biosystems (Applied Biosystems/ThermoFisher; Waltham, Mass.), using MicroAmp® Fast-96-Well Reaction 0.1 ml plates sealed with MicroAmp™ Optical adhesive film (Applied Biosystems/ThermoFisher; Waltham, Mass.), with the following setting: fast block, Standard curve as experiment type, ROX as passive reference, Ct as quantification method (automatic threshold, but adjusted to 0.05 when needed), TAMRA as reporter, and NFQ-MGB as quencher. The thermocycler was programmed as follows: 2 min at 50 °C, and 45 cycles of (1 sec at 95 °C, and 20 sec at 60 °C). The Ct plots and values are shown in Figure 74 and Ct Table 60, respectively.

Table 60. Ct Values for 7 Genes in Example 3.

CT Values	10 ng HT29	01 ng HT29+10 ng Roche Normal	10 ng Roche Normal	NTC_PCR	NTC_LDR	NTC_TaqMan™
VIM-S3	5.9	9.1	No Ct	No Ct	No Ct	No Ct
CLIP4-R-S1	6.7	7.0	31.8	31.9	31.5	No Ct
GSG1L-F-S1	4.9	5.6	No Ct	No Ct	No Ct	No Ct
PP1R16B-F-S1	4.7	8.7	34.3	35.7	36.0	No Ct
KCNA3-F-S1	5.4	5.5	25.0	30.5	37.1	No Ct
GDF6-F-S1	6.8	8.2	7.3	No Ct	No Ct	No Ct
SEPT9-F-S1	5.3	5.5	No Ct	No Ct	No Ct	No Ct

Example 4: Multiplexed Detection of 7 CRC Methylation Markers Using ex-PCR-LDR-qPCR on Bisulfite Converted Cell Free DNA in Tumor Plasma

[0422] Human plasma (with K2-EDTA as an anti-coagulant) samples were purchesed from a vendor. Cell free DNA was isolated from individual plasma samples (5 mL) using the QIA amp Circulating Nucleic Acid Kit (Qiagen) according to manufacturer's instructions, and quantified with Quant-iT Pico Green Assay Life Technologies/Thermo-Fisher; Waltham, Mass.). CpG Methylated cell free DNA fragments were enriched by antibodies containing methyl-CpG binding domain. A series of wash steps, magnetic capture, and incubation at 65°C, were employed prior to elution of methylation-enriched cell free DNA.

[0423] Bisulfite conversion of enriched methylated cell free DNA was carried out using the innuCONVERT bisulfite Body Fluids kit from Analytic Jena Corporation (Jena, Germany). Conversion reaction was carried out in 150 μl of mixture by adding 50 μl of enriched cell free DNA, 70 μl of Conversion Reagent, 30 μl of Conversion Buffer. The mixture was incubated in a thermal mixer at 85 °C for 45 min, with shaking at 800 rpm speed. After incubation, 700 μl of

binding buffer was added to the reaction mixture, prior to being loaded into a spin column, then centrifuged. 200 µl of washing solution BS was then added to the column, prior to centrifugation. This is followed by addition of 700 µl of desulfonation buffer to the column, and incubation at room temperature for 10 minutes. The column was then subjected to a series of wash steps: 500 µl of washing solution C, 650 µl of washing solution BS, 650 µl of ethanol (twice). The column was then incubated at 60°C for 10 min to remove residue ethanol. The DNA was finally eluted with 50 µl of elution buffer.

[0424] The primers used for the example described above were identical to those used in Example 3. The linear amplification, PCR, LDR and qPCR steps were carried out as described in Example 3. Three sets of cell free DNA samples (a set consists of CRC patient/normal pair) were tested using this protocol. The results using cfDNA isolated from the plasma of 3 different CRC patients and 3 different normal controls are shown in Figures 75, 76, and 77.

Example 5: Multiplexed Detection of 20 CRCM Markers Using ex-PCR-LDR-qPCR on Bisulfite Converted HT29 Cell Line DNA

[0425] The template for this particular example was also the genomic DNA extracted from HT29, and fragmented though Non Random Sonication method using ultra-sonicator from Covaris (Woburn, Massachusetts). After shearing, DNA quality was assessed with an Agilent Bioanalyzer system. The length of DNA ranged from 50 bp to 1 kb. DNA in the elution buffer was quantified using Pico Green kit (Life Technologies/Thermo-Fisher; Waltham, Mass.).

[0426] DNA fragments containing methylated CpG sites were enriched by binding to the antibodies containing methyl-CpG binding domain. After a series of wash steps followed by magnetic capture, the enriched DNA sample is eluted in a small volume of water by incubation at 65°C.

[0427] Bisulfite conversion was then carried out using the Cells-to-CpG Bisulfite Conversion kit from Applied Biosystem division of ThermoFisher (Carlsbad, Calif.). 5 µl of Denaturation Reagent was added to 45 µl of methyl enriched genomic DNA, followed by the mixture's incubation at 50 °C for 10 min. This is followed by addition of 100 µl of Conversion Reagent, and incubation in a thermal cycler with the following program: 65 °C 30 min, 90°C 30 sec, 65°C 30 min, 90°C 30 sec, 65°C 30 min. 150 µl of converted DNA mixture was mixed with 600 µl of binding buffer in the binding Column. The column was centrifuged at 10,000 rpm for 1 min, followed by discarding the flow through. The column was washed with 600 µl of washing buffer. 200 µl of Desulfonation Reagent was added to the column, followed by incubation at room temperature for 15 min. After spinning, the column was washed again with 400 µl of washing buffer. 50 µl of Elution Buffer was then added to the column to elute the

bound DNA. The mostly single stranded, bisulfite converted DNA was quantified with both Quant-iT Oli Green and Pico Green kit (Life Technologies/ThermoFisher; Waltham, Mass.).

[0428] All primers used in the preceding example were purchased from Integrated DNA Technologies Inc. (IDT; Coralville, IA). The following primer sequences from Table 45 were used in Example 5: SEQ ID NOs.: 76-235.

[0429] **Linear Amplification Step**. In a 25 µl of reaction volume, the linear amplification step was performed by mixing: 5 µl of 5x GoTaq Flexi buffer (no Magnesium) (Promega, Madison, Wis.), 2.5 µl of 25 mM $MgCl_2$ (Promega, Madison, Wis.), 0.5 µl of 10 mM dNTPs (dATP, dCTP, dGTP and dTTP) (Promega, Madison, Wis.), 2.5 µl of 20 plex gene specific reverse primers (concentration of each primer is 1 µM), 0.625 µl of 20 mU/µl RNAseH2 (diluted in RNAseH2 dilution buffer from IDT) (IDT), and 0.55 µl of Klentaql polymerase (DNA Polymerase Technology, St. Louis, Mo.) mixed with Platinum Taq Antibody (Invitrogen/Thermo Fisher, Waltham, Mass.) (the mixture is prepared by adding 0.02 µl of Klentaql polymerase at 50 U/µl to 0.2 µl of Platinum Taq Antibody at 5 U/µl), and 5.0 µl of corresponding bisulfite converted genomic DNA (out of 50 µl of eluted DNA after bisulfite conversion.) The template was either: 1) 1.0 µg of Normal Human genomic DNA (purchased from Roche) mixed with 66.0 ng of HT29 colorectal cell line genomic DNA, or 2) just 1.0 µg of Normal Human genomic DNA (normal control). The template was either digested with restriction enzyme Bsh1236I, or enriched in methylated DNA, bisulfite converted, and eluted into 50 µl of elution buffer. 5 µl of elution buffer was used in the linear amplification reaction. The reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/ThermoFisher, Waltham, Mass.) using the following program: 2 min at 94 °C, 40 cycles of (20 sec at 94°C, 40 sec at 60 °C, and 30 sec at 72 °C.), and a final hold at 4° C. After the reaction, Platinum Taq Antibodies were added in the reaction mixture to inhibit the Klentaq DNA polymerase.

[0430] **PCR reaction.** The linear amplification products were equally divided into two parts. In the 1st part, first 10 plex (out of 20 plex) gene specific forward primers and other reagents were added, in the 2nd part, the other 10 plex (out of 20 plex) gene specific forward primers and other reagents were added. Two 10 plex PCR reactions were carried out. The PCR step was performed in a 20 µl of reaction mixture prepared by adding: 2 µl of GoTaq Flexi buffer 5x without Magnesium (Promega, Madison, Wis.), 1 µl of MgCl2 at 25 mM (Promega, Madison, Wis.), 0.4 µl of dNTPs (with dATP, dCTP, dGTP and dUTP, 10 mM each) (Promega, Madison, Wis.), 2 µ1 of 10 plex (out of 20 plex in linear amplification step) gene specific forward primers at 0.5 µM each. 0.4 µl of Antarctic Thermolabile UDG (1u/µl) (New England Biolab, Ipswich, MA), 0.25 µI of RNAseH2 (IDT) at 20 mU/µl , 0.44 µl of Klentaql polymerase

(DNA Polymerase Technology, St. Louis, Mo.) mixed with Platinum Taq Antibody (Invitrogen/Thermo Fisher, Waltham, Mass.) (The mixture is prepared by adding 0.02 µl of Klentaql polymerase at 50 U/µl to 0.2 µl of Platinum Taq Antibody at 5 U/µl), and 10 µl of corresponding linear amplification products . PCR reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/ThermoFisher, Waltham, Mass.) and using the following program: 10 min at 37 °C, 40 cycles of (20 sec at 94 °C, 40 sec at 60 °C. and 30 sec at 72 °C), 10 min at 99.5 °C, and a final hold at 4 °C.

[0431] LDR step. The LDR step was performed in a 20 µl reaction prepared by combining: 5.82 µl of nuclease free water (IDT), 2 µl of 10X AK16D ligase reaction buffer [lx buffer contains 20 mM Tris-HCI pH 8.5 (Bio-Rad, Hercules, Calif.), 5 mM $MgCl_2$ (Sigma-Aldrich, St. Louis, Mo.), 50 mM KCl (Sigma-Aldrich, St. Louis, Mo.), 10 mM DTT (Sigma-Aldrich, St. Louis, Mo.) and 20 µg/ml of BSA (Sigma-Aldrich, St. Louis, Mo.), 0.5 µl of 40 mM DTT (Sigma-Aldrich, St. Louis, Mo.), 0.25 µl of 40 mM NAD+ (Sigma-Aldrich, St. Louis, Mo.), 0.5 µl of 20 mU/µl RNAseH2 (IDT), 0.4 µl of corresponding 10 plex LDR upstream probes at 500 nM each, 0.4 µl of corresponding 10 plex LDR downstream probes at 500 nM each, 0.57 µl of purified AK16D ligase (at 0.88 µM), and 4 µl of PCR reaction products from previous step. LDR reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/Thermo-Fisher; Waltham, Mass.) using the following program: 20 cycles of (10 sec at 94 °C, and 4 min at 60 °C) followed by a final hold at 4° C.

[0432] qPCR step. The qPCR step was performed in a 10 µl of reaction volume by combining: 1.5 µl of nuclease free water (IDT), 5 µl of 2x TaqMan® Fast Universal PCR Master Mix (Fast amplitaq, UDG and dUTP) from Applied Biosystems (Applied Biosystems/ThermoFisher; Waltham, Mass.), 1 µl of 2.5 µM TaqMan™ Assay forward primer, 1 µl of 2.5µM Taqman™ reverse primer , 0.5 µl of 5 µM Taqman™ probe, and 1 µl of LDR reaction products. qPCR reactions were run in a ViiA7 real-time thermo-cycler from Applied Biosystems (Applied Biosystems/Thermo-Fisher; Waltham, Mass.), using MicroAmp® Fast-96-Well Reaction 0.1 ml plates sealed with MicroAmp™ Optical adhesive film (Applied Biosystems/ThermoFisher; Waltham, Mass.), with the following setting: fast block, Standard curve as experiment type, ROX as passive reference, Ct as quantification method (automatic threshold, but adjusted to 0.05 when needed), TAMRA as reporter, and NFQ-MGB as quencher. The program employed was: 2 min at 50 °C, and 45 cycles of (1 sec at 95 °C, and 20 sec at 60 °C). Results are shown in Figure 78, and Table 61 below.

Table 61. Ct values for each gene in Example 5.

Ct Values	1	2	3	4	5	6	7	8	9	10
Gene	GATA5-S2	ZNF542-S1	RCN-3-S1	MYO15B-S1	ANKRD13B-S1	FAM115A-S1	RGS10-S1	HCG4-S1	STK32B-S1	CNRIP1-S1
starting number for primer	5421	5431	5441	5451	5461	5471	5491	5501	5511	5521
1,000 GE of HT29 DNA + 7,500 GE of Roche DNA	17.0	12.6	12.6	10.0	12.9	16.5	8.8	10.9	6.6	11.0
7,500 GE of Roche DNA	34.2	39.4	39.8	No Ct	37.5	24.7	No Ct	35.7	31.4	35.7
1st Linaer Amp_NTC	34.4	39.6	39.6	36.3	No Ct	No Ct	No Ct	37.0	31.6	38.4
2nd Linear Amp_NTC	32.6	36.6	37.9	38.3	No Ct	No Ct	No Ct	36.4	32.2	33.5
LDR_NTC	34.2	44.8	35.7	37.9	37.1	No Ct	37.6	35.5	31.1	32.4
TaqMan™_NTC	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct
	11	12	13	14	15	16	17	18	19	20
Gene	VIM-S1	CLIP4-S1	GSG1L-S1	PP1R16B-S1	KCNA3	GDF6-S1	ADHFE1-S1	THBD-S1	SEPT9-S4	SEMA3B-S1
starting number for primer	5001	5021	5051	5061	5071	5081	5101	5331	5351	5401
1,000 GE of HT29 DNA + 7,500 GE of Roche DNA	10.4	5.7	7.7	17.9	11.2	14.2	17.5	30.0	7.2	9.2
7,500 GE of Roche DNA	17.6	No Ct	36.6	36.5	36.8	32.7	36.7	No Ct	31.2	No Ct
1st Linaer Amp_NTC	No Ct	No Ct	36.8	36.3	No Ct	32.8	35.1	No Ct	31.5	41.4
2nd Linear Amp_NTC	No Ct	No Ct	No Ct	35.1	No Ct	33.3	36.8	No Ct	31.2	No Ct
LDR_NTC	No Ct	No Ct	No Ct	36.7	No Ct	32.6	38.3	No Ct	30.7	39.8
TaqMan™_NTC	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct	No Ct

Example 6: Multiplexed Detection of 20 CRCM Markers Using ex-PCR-LDR-qPCR on Bisulfite Converted HT29 Cell Line DNA

[0433] General Methods: HT-29 colon adenocarcinoma cells were seeded in 60 cm2 culture dishes in McCoy's 5A medium containing 4.5 g/l glucose, supplemented with 10% fetal calf serum, and kept in a humidified atmosphere containing 5% CO_2. Once cells reached 80-90% confluence, they were washed in Phosphate Buffered Saline (x3), and collected by centrifugation (500xg). Genomic DNA was isolated using the DNeasy Blood & Tissue Kit frpm QIAGEN (Qiagen, Valencia, Calif.), and its concentration was measured using Quant-iT Pico green ds DNA Assay kit (Thermo-Fisher, Waltham, MA.). High molecular weight (>50 kb) genomic DNA (0.2 mg/ml) isolated from normal human blood (buffy coat) (Roche human genomic DNA) was purchased from Roche (Indianapolis, Ind.). Its concentration was similarly determined using Quant-iT PicoGreen dsDNA Assay Kit (Thermo-Fisher, Waltham, MA.). 1.0 µg HT29 cell line genomic DNA or Roche Normal DNA was fragmented through non random sonication method, using Covaris ultra sonicator E220 (Covaris, Woburn, MA). After shearing, the quality of the resulting DNA fragments (length ranged from 50 to 1 kb base pairs) was assessed with Agilent Bioanalyzer system 2100 (Agilent, Santa Clara, CA).

[0434] Enrichment of methylated DNA: The DNA fragments containing methylated CpGs was captured by methylation-specific antibodies, using the EpiMark® Methylated DNA Enrichment Kit from New England Biolabs, according to manufacturer's instructions (New England Biolabs, Ipswich, MA). DNA fragments containing methylated CpG sites were enriched by binding to the antibodies containing methyl-CpG binding domain. After a series of wash steps followed by magnetic capture, the enriched DNA sample was eluted in a small volume of water by incubation at 65°C.

[0435] Bisulfite conversion of DNA: Bisulfite conversion of cytosine bases in DNA was then carried out using the Cells-to-CpG Bisulfite Conversion kit from Applied Biosystem division of ThermoFisher (ThermoFisher , Carlsbad, Calif.). 5 µl of Denaturation Reagent was added to 45 µl of methyl enriched genomic DNA, followed by the mixture's incubation at 50 °C for 10 min. After adding 100 µl of Conversion Reagent, the mixture was incubated in a thermal cycler with the following program: 65 °C 30 min, 90°C 30 sec, 65°C 30 min, 90°C 30 sec, 65°C 30 min. 150 µl of converted DNA mixture was mixed with 600 µl of binding buffer in the binding Column. The column was centrifuged at 10,000 rpm for 1 min, followed by discarding the flow through. The column was washed with 600 µl of washing buffer. 200 µl of Desulfonation Reagent was added to the column, followed by incubation at room temperature for 15 min. After spinning, the column was washed again with 400 µl of washing buffer. 50 µl of Elution Buffer was then added to the column to elute the bound DNA. The mostly single

stranded, bisulfite converted DNA was quantified with both Quant-iT Oli Green ss DNA kit and Pico Green ds DNA kit (ThermoFisher, Waltham, Mass.).

[0436] PCR primers, LDR probes, and LNA or PNA blocking primers: All primers used are listed in Table 45 above. All primers were purchased from Integrated DNA Technologies Inc. (IDT) (Coralville, Iowa). All primers used in the preceding example were purchased from Integrated DNA Technologies Inc. (IDT; Coralville, IA). The primer number ended with 02A has the same sequences of the primer end with 02, but it has no short (10-mer) 5' end tail sequences.

[0437] Templates preparation: **Template "A"**: 1 ug of sonicated and methylation enriched Roche Normal DNA was mixed with 6.6 ng of sonicated HT29 DNA in 50 µl, 10 ng human genomic DNA equals to 3,000 genomic equivalent (GE). The yield of methylation enrichment is about 50%, and the bisulfite conversion is about 50%. So the template A contains 75,000 GE of Roche DNA and 2,000 GE of HT29 DNA in 50 µl. 5 ul of DNA template A was used in the linear amplification reaction, it contains 7,500 GE of Roche DNA and 200 GE of HT29 DNA, **Template "B"**: 1 µg of sonicated and methylation enriched Roche Normal DNA was bisulfite converted and it has 75,000 GE of Roche DNA in 50 µl.
In the following experiment, 5 µl of HT29 DNA will be used, and it contains 1,000 GE of HT29 and 7,500 GE of Roche DNA. 5 µl of DNA template B was used in the linear amplification reaction, it contains 7,500 GE of Roche normal DNA.

[0438] Linear Amplification Step: For condition A, In a 25 µl of reaction volume, the linear amplification step was performed by mixing: 5 µl of 5x Gotaq Flexi buffer (no Magnesium) (Promega, Madison, Wis.), 3.5 µl of 25 mM MgCl2 (Promega, Madison, Wis.), 0.5 µl of 10 mM dNTPs (dATP, dCTP, dGTP and dTTP) (Promega, Madison, Wis.), 2.5 µl of second set of 20 plex marker-specific reverse primers with 10-mer short tail (concentration of each primer is 0.5 µM, primer number ends with 02 has a short tail, a universal sequences), 0.5 µl of tween 20 (5%), 0.9 µl of 20 mU/µl RNAseH2 (diluted in RNAseH2 dilution buffer from IDT) (IDT), and 0.5 µl of Klentaql polymerase (50 U/µl)(DNA Polymerase Technology, St. Louis, Mo.) mixed with Platinum Taq Antibody (Invitrogen/Thermo Fisher, Waltham, Mass.) (The ratio of mixing of Klentaql polymerase with Antibody is 1:10, and the final concentration of Klentaql polymerase is 5U/ul), and 5.0 µl of corresponding bisulfite converted methylated enriched genomic DNA (out of 50 µl of eluted DNA after bisulfite conversion.). **For condition B**, in the other 25 ul of reaction mixture, all the reagents were the same except 2.5 µl of second set of 20 plex marker-specific reverse primers with no tail (Primer number ending with 02A). The template was either: 1) 5 ul of template A it contains 200 GE of HT29 DNA and 7,500 GE of Roche DNA. 2) 5 ul of template B, it contains 7,500 GE of Roche DNA. The reactions were

run in a ProFlex PCR system thermocycler (Applied Biosystems/ ThermoFisher, Waltham, Mass.) using the following program: 2 min at 94 °C, 40 cycles of (20 sec at 94°C, 40 sec at 60 °C, and 30 sec at 72 °C.), and a final hold at 4° C. After the reaction, 0.5 µl of Platinum Taq antibodies were added in the reaction mixture to inhibit the Klentaq DNA polymerase.

[0439] PCR reaction: The linear amplification products were equally divided into two parts with two 10-plex reaction being carried out. In the 1st part, first 10 plex (marker number 21 to number 30) marker-specific forward primers and other reagents were added, in the 2nd part, the second 10 plex (marker number 31 to number 40) marker specific forward primers and other reagents were added. The PCR step was performed in a 20 µl of reaction mixture prepared by adding: 2 µl of Gotaq Flexi buffer 5x without Magnesium (Promega, Madison, Wis.), 1.4 µl of $MgCl_2$ at 25 mM (Promega, Madison, Wis.), 0.4 µl of dNTPs (with dATP, dCTP, dGTP and dUTP, 10 mM each) (Promega, Madison, Wis.), 0.2 µl of tween 20 (5%), 2 µl of 10 plex (1st 10-plex and 2nd 10-plex) marker-specific forward primers at 0.25 µM each. 0.4 µl of Antarctic Thermolabile UDG (1u/µl)(New England Biolab, Ipswich, MA), 0.29 µI of RNAseH2 (IDT) at 20 mU/µl , 1.6 µl of Klentaql polymerase (DNA Polymerase Technology, St. Louis, Mo.) mixed with Platinum Taq Antibody (Invitrogen/Thermo Fisher, Waltham, Mass.) (The ratio of mixing of Klentaql polymerase with Antibody is 1:10, and the final concentration of Klentaql polymerase is 5U/ul), 10 µl of corresponding linear amplification products and 1.7 µl water. PCR reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/ThermoFisher, Waltham, Mass.) and using the following program: 10 min at 37 °C, 50 cycles of (94 ° C 10s, 60 ° C 30s ,72 ° C 20s,), 10 min at 99.5 °C., and a final hold at 4 °C.

[0440] LDR step: The LDR step was performed in a 20 µl of reaction mixture prepared by combining: 11.6 µl of nuclease free water (IDT), 2 µl of 10X AK16D ligase reaction buffer [1x buffer contains 20 mM Tris-HCI pH 8.5 (Bio-Rad, Hercules, Calif.), 5 mM $MgCl_2$ (Sigma-Aldrich, St. Louis, Mo.), 50 mM KCl (Sigma-Aldrich, St. Louis, Mo.), 10 mM DTT (Sigma-Aldrich, St. Louis, Mo.) and 20 µg/ml of BSA (Sigma Aldrich, St. Louis, Mo.)], 0.5 µl of 40 mM DTT (Sigma-Aldrich, St. Louis, Mo.), 0.4 µl of 50 mM NAD+ (Sigma-Aldrich, St. Louis, Mo.), 0.5 µl of 20 mU/µl RNAseH2 (IDT), 0.4 µl of corresponding 10 plex LDR upstream probes at 500 nM each, 0.4 µl of corresponding 10 plex LDR downstream probes at 500 nM each, 0.57 µl of purified AK16D ligase (at 0.88 µM), and 4 µl of PCR reaction products from previous step. LDR reactions were run in a ProFlex PCR system thermocycler (Applied Biosystems/Thermo-Fisher; Waltham, Mass.) using the following program: 20 cycles of (10 sec at 94 °C, and 4 min at 60 °C) followed by a final hold at 4° C.

[0441] qPCR step: The qPCR reaction is run uni-plex with the qPCR step performed in a 10 µl of reaction volume by combining: 3 µl of nuclease free water (IDT), 5 µl of 2x TaqMan®

Fast Universal PCR Master Mix (Fast amplitaq, UDG and dUTP) from Applied Biosystems (Applied Biosystems/ThermoFisher, Waltham, MA), 0.5 µl of a mixture of 5 µM TaqMan™ Assay one marker-specific forward primer and 5 µM of Taqman™ corresponding marker-specific reverse primer , 0.5 µl of 5 µM Taqman™ one marker-specific probe, and 1 µl of LDR 10-plex reaction products. qPCR reactions were carried out in a ViiA7 real-time thermo-cycler from Applied Biosystems (Applied Biosystems/Thermo-Fisher; Waltham, Mass.), using MicroAmp® Fast-96-Well Reaction 0.1 ml plates sealed with MicroAmp™ Optical adhesive film (Applied Biosystems/ThermoFisher; Waltham, Mass.), with the following setting: fast block, Standard curve as experiment type, ROX as passive reference, Ct as quantification method (automatic threshold, but adjusted to 0.05 when needed), TAMRA as reporter, and NFQ-MGB as quencher. The program employed was: 2 min at 50 °C, and 45 cycles of (1 sec at 95 °C, and 20 sec at 60 °C.). Results are shown in Figures 79 and 80 and Table 62 below.

Table 62. Ct values for each gene in Example 6.

CRC-Marker		1	2	3	4	5	6	7	8	9	10
Gene		IRF4	TWIST1	TDH	NCAM1	COL4A2/1	VSX1	PTPRT	MIR124-3	ZNF677	EVC
Starting number for primer	RP02 Status	5561	5571	5591	5621	5651	5661	5671	5681	5691	5701
200GE of HT29 + 7,500 GE of Roche	RP02 with Short Tail	30.2	35.9	31.0	10.3	17.1	17.7	31.6	33.6	30.4	21.2
7,500 GE of Roche DNA		No Ct	No Ct	32.3	32.8	34.5	40.3	33.9	33.9	35.8	30.2
1st Linear Amplification_NTC		No Ct	39.0	32.5	33.2	35.5	No Ct	33.1	33.1	35.2	30.6
2nd PCR_NTC		No Ct	37.5	32.3	33.1	34.2	No Ct	32.4	33.3	37.0	31.0
200GE of HT29 + 7,500 GE of Roche	RP02A without Tail	9.1	11.8	32.6	23.5	10.7	9.7	10.9	7.3	19.1	23.0
7,500 GE of Roche DNA		No Ct	38.5	32.6	33.1	34.7	No Ct	8.3	33.2	36.3	24.9
1st Linear Amplification_NTC		No Ct	No Ct	32.5	32.6	36.3	No Ct	32.8	33.8	34.8	30.8
2nd PCR_NTC		No Ct	35.7	32.2	33.6	35.4	No Ct	33.1	32.9	37.0	31.2

Table 62 continued. Ct values for each gene in Example 6.

CRC-Marker		11	12	13	14	15	16	17	18	19	20
Gene		JAM2	GFRA1	FLI1	GNAO1	CoCaNCR2	CoCaNCR9	CoCaNCR10	CoCaNCR8	SLC18A3,CHAT	LIFR
starting number for primer	RP02 Status	5711	5721	5731	5741	5751	5761	5771	5781	5801	5811
200GE of HT29 + 7,500 GE of Roche	RP02 With Short Tail	35.0	23.8	16.5	No Ct	20.5	14.4	16.1	19.1	35.8	16.9
7,500 GE of Roche DNA		No Ct	38.4	32.7	No Ct	38.2	36.0	No Ct	41.2	35.9	No Ct
1st Linear Amplification_NTC		No Ct	38.6	32.4	No Ct	41.6	35.6	No Ct	38.9	35.5	No Ct
2nd PCR_NTC		No Ct	38.0	32.0	No Ct	40.6	37.4	No Ct	35.7	36.1	No Ct
200GE of HT29 + 7,500 GE of Roche	RP02A Without Tail	16.1	10.9	10.7	21.5	18.2	6.9	29.7	9.2	29.9	9.5
7,500 GE of Roche DNA		No Ct	21.6	32.7	11.7	37.8	7.8	No Ct	No Ct	41.5	No Ct
!st Linear Amplification_NTC		No Ct	39.2	32.6	No Ct	39.7	39.4	No Ct	No Ct	35.4	No Ct
2nd PCR_NTC		No Ct	35.3	32.8	No Ct	37.9	38.3	No Ct	37.3	36.6	No Ct

[00126] Although preferred embodiments have been depicted and described in detail herein, it will be apparent to those skilled in the relevant art that various modifications, additions, substitutions, and the like can be made without departing from the spirit of the present application and these are therefore considered to be within the scope of the present application as defined in the claims which follow.

WHAT IS CLAIMED:

1. A method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences of other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues, said method comprising:

providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules;

providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer;

blending the sample, the one or more first primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix, and a DNA polymerase to form one or more polymerase extension reaction mixtures;

subjecting the one or more polymerase extension reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase extension reaction mixtures and for carrying out one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising nucleotide sequences complementary to the target nucleotide sequence;

blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more second primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixtures, a deoxynucleotide mix

including dUTP, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more first polymerase chain reaction products comprising the target nucleotide sequence or a complement thereof;

providing one or more oligonucleotide probe sets, each probe set comprising (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on a complementary target nucleotide sequence of a secondary extension product;

blending the one or more first polymerase chain reaction products with a ligase, and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures;

subjecting the one or more ligation reaction mixtures to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets are ligated together, when hybridized to their complementary sequence, to form ligated product sequences in the ligation reaction mixtures wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence;

blending the ligated product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an

extension treatment thereby forming one or more second polymerase chain reaction products; and

detecting and distinguishing the one or more second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

2. A method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues, said method comprising:

providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules;

providing one or more nucleases capable of digesting nucleic acid molecules not comprising modified nucleotides;

providing one or more first primary oligonucleotide primer(s) that comprises a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence;

blending the sample, the one or more first primary oligonucleotide primers, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix that comprises one or more modified nucleotides that protect extension products but not target DNA from nuclease digestion, and a DNA polymerase to form one or more polymerase extension reaction mixtures;

subjecting the one or more polymerase extension reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase extension reaction mixture and for carrying out one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension

treatment, thereby forming primary extension products comprising the complement of the target nucleotide sequence;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer having a first 5' primer-specific portion and a 3' portion that is complementary to a portion of a primary extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a second 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer;

blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary oligonucleotide primer sets, the one or more nucleases, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting nucleic acid molecules present in the first polymerase chain reaction mixtures, but not primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more first polymerase chain reaction products comprising the first 5' primer-specific portion, a target-specific nucleotide sequence or a complement thereof, and a complement of the second 5' primer-specific portion;

providing one or more tertiary oligonucleotide primer sets, each tertiary oligonucleotide primer set comprising (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the first 5' primer-specific portion of the one or more first polymerase chain reaction products and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the one or more first polymerase chain reaction products;

blending the one or more first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU) containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an

extension treatment, thereby forming one or more second polymerase chain reaction products; and

detecting and distinguishing the one or more second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

3. A method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences of other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues, said method comprising:

providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences of other parent nucleic acid molecules by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules;

providing one or more nucleases capable of digesting nucleic acid molecules present not comprising modified nucleotides;

providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the parent nucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer;

blending the sample, the one or more first primary oligonucleotide primers of the primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix that comprises one or more modified nucleotides that protect extension product but not target DNA from nuclease digestion, and a DNA polymerase to form one or more polymerase extension reaction mixtures;

subjecting the one or more polymerase extension reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the

polymerase extension reaction mixtures and for carrying out one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the target nucleotide sequence;

blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more second primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more nucleases, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting nucleic acid molecules present in the polymerase chain reaction mixtures, but not primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising the target nucleotide sequence or a complement thereof;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer;

blending the first polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products; and

detecting and distinguishing the second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more nucleotides, one or more copy numbers, one or more transcript sequences, and/or one or more methylated residues.

4. A method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues, said method comprising:

providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues;

subjecting the nucleic acid molecules in the sample to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules;

providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer;

blending the bisulfite-treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase to form one or more polymerase extension reaction mixtures;

subjecting the one or more polymerase extension reaction mixtures to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite-treated target nucleotide sequence;

blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first

polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising the bisulfite-treated target nucleotide sequence or a complement thereof;

providing one or more oligonucleotide probe sets, each probe set comprising (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion, and (b) a second oligonucleotide probe having a 5' bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion and a 3' primer-specific portion, and wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on a complementary nucleotide sequence of a first polymerase chain reaction product;

blending the first polymerase chain reaction products with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures;

subjecting the one or more ligation reaction mixtures to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets are ligated together, when hybridized to complementary sequences, to form ligated product sequences in the ligation reaction mixture wherein each ligated product sequence comprises the 5' primer-specific portion, the bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portions, and the 3' primer-specific portion;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence;

blending the ligated product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming a second polymerase chain reaction products; and

detecting and distinguishing the second polymerase chain reaction products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more

nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

5. A method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more more methylated residues, said method comprising:

providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues;

subjecting the nucleic acid molecules in the sample to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules;

providing one or more first primary oligonucleotide primer(s) that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue;

blending the bisulfite-treated sample, the one or more first primary oligonucleotide primers, a deoxynucleotide mix, and a DNA polymerase to form one or more polymerase extension reaction mixtures;

subjecting the one or more polymerase extension reaction mixtures to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite-treated target nucleotide sequence;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of the polymerase extension reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer;

blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting nucleic acid molecules present in the first polymerase chain reaction mixtures, but not primary extension products comprising modified nucleotides and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reactions products comprising a 5' primer-specific portion of the first secondary oligonucleotide primer, the bisulfite-treated target nucleotide sequence-specific or complement sequence-specific portion, and a complement of the 5' primer-specific portion of the second secondary oligonucleotide primer;

providing one or more tertiary oligonucleotide primer sets, each tertiary oligonucleotide primer set comprising (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reactions product sequence;

blending the first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain reaction products; and

detecting and distinguishing the secondary polymerase chain reactions products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

6. A method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues, said method comprising:

providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues;

subjecting the nucleic acid molecules in the sample to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample;

providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer;

blending the bisulfite-treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase to form one or more polymerase extension reaction mixtures;

subjecting the one or more polymerase extension reaction mixtures to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite treated target nucleotide sequence;

blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixture, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction

cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising the bisulfite-treated target nucleotide sequence or a complement thereof;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of a first polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a first polymerase chain reaction product formed from the first secondary oligonucleotide primer;

blending the first polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products; and

detecting and distinguishing the second polymerase chain reactions products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

7. A method for identifying, in a sample, one or more parent nucleic acid molecules containing a target nucleotide sequence differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues, said method comprising:

providing a sample containing one or more parent nucleic acid molecules potentially containing the target nucleotide sequence differing from the nucleotide sequences in other parent nucleic acid molecules by one or more methylated residues;

subjecting the nucleic acid molecules in the sample to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample;

providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a sequence in the bisulfite-treated parent nucleic acid molecules adjacent to the bisulfite-treated target nucleotide sequence containing the one or more methylated residue and (b) a second primary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer;

blending the bisulfite treated sample, the one or more first primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase to form one or more polymerase extension reaction mixtures;

subjecting the one or more polymerase extension reaction mixtures to conditions suitable for one or more polymerase extension reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming primary extension products comprising the complement of the bisulfite treated target nucleotide sequence;

blending the one or more polymerase extension reaction mixtures comprising the primary extension products, the one or more secondary primary oligonucleotide primers of the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the reaction mixture, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reactions products comprising the bisulfite-treated target nucleotide sequence or a complement thereof;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products or their complements and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction products or their complements;

blending the primary polymerase chain reaction product sequences, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products; and

detecting and distinguishing the second polymerase chain reactions products in the one or more second polymerase chain reaction mixtures to identify the presence of one or more parent nucleic acid molecules containing target nucleotide sequences differing from nucleotide sequences in other parent nucleic acid molecules in the sample by one or more methylated residues.

8. The method of any one of claims 1 through 7 further comprising:

contacting the sample with DNA repair enzymes to repair damaged DNA, abasic sites, oxidized bases, or nicks in the DNA.

9. The method of any one of claims 4 through 7 further comprising:

contacting the sample with at least a first methylation sensitive enzyme to form a restriction enzyme reaction mixture prior to, or concurrent with, said blending to form one or more polymerase extension reaction mixtures, wherein said first methylation sensitive enzyme cleaves nucleic acid molecules in the sample that contain one or more unmethylated residues within at least one methylation sensitive enzyme recognition sequence, and whereby said detecting involves detection of one or more parent nucleic acid molecules containing the target nucleotide sequence, wherein said parent nucleic acid molecules originally contained one or more methylated residues.

10. The method of any one of claims 4 through 7 further comprising:

contacting the sample with an immobilized methylated nucleic acid binding protein or antibody to selectively bind and enrich for methylated nucleic acid in the sample.

11. The method of any one of claims 1 through 7, wherein primers from said one or more primary or secondary oligonucleotide primer sets comprise a portion that has no or

one nucleotide sequence mismatch when hybridized in a base-specific manner to the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof, but have one or more additional nucleotide sequence mismatches that interferes with polymerase extension when primers from said one or more primary or secondary oligonucleotide primer sets hybridize in a base-specific manner to a corresponding nucleotide sequence portion in wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof.

12. The method of any one of claims 1 through 7, wherein one or both primary oligonucleotide primers of the primary oligonucleotide primer set and/or one or both secondary oligonucleotide primers of the secondary oligonucleotide primer set have a 3' portion comprising a cleavable nucleotide or nucleotide analogue and a blocking group, such that the 3' end of said primer or primers is unsuitable for polymerase extension, said method further comprising:

cleaving the cleavable nucleotide or nucleotide analog of one or both oligonucleotide primers during said hybridization treatment, thereby liberating free 3'OH ends on one or both oligonucleotide primers prior to said extension treatment.

13. The method of claim 12, wherein primers from said one or more primary or secondary oligonucleotide primer sets comprise a sequence that differs from the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof, said difference is located two or three nucleotide bases from the liberated free 3'OH end.

14. The method of claim 12, wherein the cleavable nucleotide comprises one or more RNA bases.

15. The method of any one of claims 1 through 7 further comprising;

providing one or more blocking oligonucleotide primers comprising one or more mismatched bases at the 3' end or one or more nucleotide analogs and a blocking group at the 3' end, such that the 3' end of said blocking oligonucleotide primer is unsuitable for polymerase extension when hybridized in a base-specific manner to wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof, wherein said blocking oligonucleotide primer comprises a portion having a nucleotide sequence that is the same as a nucleotide sequence portion in the wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof to

which the blocking oligonucleotide primer hybridizes but has one or more nucleotide sequence mismatches to a corresponding nucleotide sequence portion in the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof and

blending the one or more blocking oligonucleotide primers with the sample or subsequent products prior to a polymerase extension reaction, polymerase chain reaction, or ligation reaction, whereby during hybridization said one or more blocking oligonucleotide primers preferentially hybridize in a base-specific manner to a wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof, thereby interfering with polymerase extension or ligation during reaction of a primer or probes hybridized in a base-specific manner to the wildtype sequence or bisulfite-converted unmethylated sequence or complement sequence thereof.

16. The method of any one of claims 3 or 6, wherein the first secondary oligonucleotide primer has a 5' primer-specific portion and the second secondary oligonucleotide primer has a 5' primer-specific portion, said one or more secondary oligonucleotide primer sets further comprising a third secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first secondary oligonucleotide primer and (d) a fourth secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the second secondary oligonucleotide primer.

17. A method for identifying in a sample, one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level, said method comprising:

providing a sample containing one or more parent ribonucleic acid molecules containing a target ribonucleic acid molecule potentially differing in sequence from other parent ribonucleic acid molecules;

providing one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules present in the sample;

contacting the sample with one or more enzymes capable of digesting dU containing nucleic acid molecules potentially present in the sample;

providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer that comprises a

nucleotide sequence that is complementary to the RNA sequence in the parent ribonucleic acid molecule adjacent to the target ribonucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of the cDNA extension product formed from the first primary oligonucleotide primer;

blending the contacted sample, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix including dUTP, a reverse transcriptase, and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity to form one or more reverse-transcription/polymerase chain reaction mixtures;

subjecting the one or more reverse-transcription/polymerase chain reaction mixtures to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target ribonucleic nucleic acid and to carry out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse transcription/polymerase products;

providing one or more oligonucleotide probe sets, each probe set comprising (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on complementary portions of a reverse transcriptase/polymerase product corresponding to the target ribonucleic acid molecule sequence;

contacting the reverse transcriptase/polymerase products with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures;

subjecting the one or more ligation reaction mixtures to one or more ligation reaction cycles whereby the first and second probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form ligated product sequences in the ligase reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence;

blending the ligated product sequences, the one or more secondary oligonucleotide primer sets with one or more enzymes capable of digesting deoxyuracil

(dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming first polymerase chain reaction products; and

detecting and distinguishing the first polymerase chain reaction products, thereby identifying the presence of one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level.

18. A method for identifying in a sample, one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequence differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level, said method comprising:

providing a sample containing one or more parent ribonucleic acid molecules containing a target ribonucleic acid molecule potentially differing in sequence from other parent ribonucleic acid molecules;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample;

contacting the sample with one or more enzymes capable of digesting dU containing nucleic acid molecules potentially present in the sample;

providing one or more primary oligonucleotide primer sets, each primary oligonucleotide primer set comprising (a) a first primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to the RNA sequence in the parent ribonucleic acid molecule adjacent to the target nucleotide sequence and (b) a second primary oligonucleotide primer that comprises a nucleotide sequence that is complementary to a portion of the cDNA extension product formed from the first primary oligonucleotide primer;

blending the contacted sample, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, a reverse transcriptase and a DNA polymerase or a DNA

polymerase with reverse-transcriptase activity to form one or more reverse-transcription/polymerase chain reaction mixtures;

subjecting the one or more reverse-transcription/polymerase chain reaction mixtures to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target RNA and to carry out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse-transcription/primary polymerase chain reaction products;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer having a 3' portion that is complementary to a portion of a reverse-transcription/primary polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a reverse-transcription/primary polymerase chain reaction product formed from the first secondary oligonucleotide primer;

blending the reverse-transcription/primary polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming first polymerase chain reaction products; and

detecting and distinguishing the first polymerase chain reaction products, thereby identifying the presence of one or more parent ribonucleic acid molecules containing a target ribonucleic acid sequences differing from ribonucleic acid sequences of other parent ribonucleic acid molecules in the sample due to alternative splicing, alternative transcript, alternative start site, alternative coding sequence, alternative non-coding sequence, exon insertion, exon deletion, intron insertion, translocation, mutation, or other rearrangement at the genome level.

19. A method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases, said method comprising:

providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample;

contacting the sample with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample;

blending the contacted sample with a ligase and one or more first oligonucleotide preliminary probes comprising a 5' phosphate, a 5' stem-loop portion, an internal primer-specific portion within the loop region, a blocking group, and a 3' nucleotide sequence that is complementary to a 3' portion of the target miRNA molecule sequence to form one or more first ligation reaction mixtures;

ligating, in the one or more first ligation reaction mixtures, the one or more target miRNA molecules at their 3'end to the 5' phosphate of the one or more first oligonucleotide preliminary probes to generate chimeric nucleic acid molecules comprising the target miRNA molecule sequence, if present in the sample, appended to the one or more first oligonucleotide preliminary probes;

providing one or more primary oligonucleotide primer sets, each primer set comprising (a) a first primary oligonucleotide primer comprising a nucleotide sequence that is complementary to the internal primer-specific portion of the first oligonucleotide preliminary probe, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets;

blending the one or more first ligation reaction mixtures comprising chimeric nucleic acid molecules, the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the sample, a deoxynucleotide mix including dUTP, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity to form one or more reverse-transcription/polymerase chain reaction mixtures,

subjecting the one or more reverse-transcription/polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the reverse-transcription/polymerase chain reaction mixtures to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the chimeric nucleic acid molecules, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different primary reverse-transcription/polymerase chain reaction products comprising the 5'

primer-specific portion, a nucleotide sequence corresponding to the target miRNA molecule sequence, and the complement of the internal primer-specific portion, and complements thereof;

providing one or more oligonucleotide probe sets, each probe set comprising (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion, a portion complementary to a primary extension product, and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, on complementary portions of a primary reverse-transcription/polymerase chain reaction product corresponding to the target miRNA molecule sequence, or complement thereof;

contacting the primary reverse-transcription/polymerase chain reaction products with a ligase and the one or more oligonucleotide probe sets to form one or more second ligation reaction mixtures;

subjecting the one or more second ligation reaction mixtures to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form ligated product sequences in the ligation reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence;

blending the ligated product sequences and the one or more secondary oligonucleotide primer sets, with one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the second polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain reaction products; and

detecting and distinguishing the secondary polymerase chain reaction products in the one or more reactions thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

20. A method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases, said method comprising:

providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample;

contacting the sample with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample;

blending the contacted sample with a ligase and one or more first oligonucleotide probes comprising a 5' phosphate, a 5' stem-loop portion, an internal primer-specific portion within the loop region, a blocking group, and a 3' nucleotide sequence that is complementary to a 3' portion of the target miRNA molecule sequence to form one or more ligation reaction mixtures;

ligating, in the one or more ligation reaction mixtures, the one or more target miRNA molecules at their 3'end to the 5' phosphate of the one or more first oligonucleotide probes to generate chimeric nucleic acid molecules comprising the target miRNA molecule sequence, if present in the sample, appended to the one or more first oligonucleotide probes;

providing one or more primary oligonucleotide primer sets, each primer set comprising (a) a first primary oligonucleotide primer comprising a nucleotide sequence that is complementary to the internal primer-specific portion of the first oligonucleotide probe, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets;

blending the one or more ligation reaction mixtures comprising chimeric nucleic acid molecules, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity to form one or more reverse-transcription/polymerase chain reaction mixtures,

subjecting the one or more reverse-transcription/polymerase chain reaction mixtures to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the chimeric nucleic acid molecules, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different primary reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion, a nucleotide sequence corresponding

to the target miRNA molecule sequence, and the complement of the internal primer-specific portion, and complements thereof;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of an extension product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of an extension product formed from the first secondary oligonucleotide primer;

blending the primary reverse-transcription/polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising a 5' primer-specific portion of the first secondary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence or a complement thereof, and a complement of the other 5' primer-specific portion second secondary oligonucleotide primer;

providing one or more tertiary oligonucleotide primer sets, each tertiary oligonucleotide primer set comprising (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction products or their complements and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction products or their complements;

blending the first polymerase chain reaction process products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU) containing nucleic acid molecules present in the secondpolymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products; and

detecting and distinguishing the second polymerase chain reaction products, thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

21. A method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases, said method comprising:

providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample;

contacting the sample with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample;

blending the contacted sample with ATP and a Poly(A) polymerase to form a Poly(A) polymerase reaction mixture;

subjecting the Poly(A) polymerase reaction mixture to conditions suitable for appending homopolymer A to the 3' ends of the one or more target miRNA molecules potentially present in the sample;

providing one or more primary oligonucleotide primer sets, each primer set comprising (a) a first primary oligonucleotide primer comprising a 5' primer-specific portion, an internal poly dT portion, and a 3' portion comprising from 1 to 10 bases complementary to the 3' end of the target miRNA, wherein the first primary oligonucleotide primer may be the same or may differ from other first primary oligonucleotide primers in other sets, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets;

blending the Poly(A) polymerase reaction mixture, the one or more primary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules in the sample, a deoxynucleotide mix including dUTP, and a reverse transcriptase and a DNA polymerase or a DNA polymerase with reverse-transcriptase activity to form one or more reverse-transcription/polymerase chain reaction mixtures;

subjecting the one or more reverse-transcription/polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the reverse-transcription/polymerase chain reaction mixtures, then to conditions

suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target miRNA sequences with 3' polyA tails, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming one or more different reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion of the second primary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence, a poly dA region, and the complement of the 5' primer-specific portion of the first primary oligonucleotide primer, and complements thereof;

providing one or more oligonucleotide probe sets, each probe set comprising (a) a first oligonucleotide probe having a 5' primer-specific portion and a 3' target sequence-specific portion, and (b) a second oligonucleotide probe having a 5' target sequence-specific portion, a portion complementary to the one or more reverse-transcription/polymerase chain reaction products, and a 3' primer-specific portion, wherein the first and second oligonucleotide probes of a probe set are configured to hybridize, in a base specific manner, to complementary portions of the one or more reverse-transcription/polymerase chain reaction products corresponding to the target miRNA molecule sequence, or complement thereof;

contacting the one or more reverse-transcription/polymerase chain reaction products with a ligase and the one or more oligonucleotide probe sets to form one or more ligation reaction mixtures;

subjecting the one or more ligation reaction mixtures to one or more ligation reaction cycles whereby the first and second oligonucleotide probes of the one or more oligonucleotide probe sets, when hybridized to their complement, are ligated together to form ligated product sequences in the ligation reaction mixture, wherein each ligated product sequence comprises the 5' primer-specific portion, the target-specific portions, and the 3' primer-specific portion;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the ligated product sequence and (b) a second secondary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the ligated product sequence;

blending the ligated product sequences and the one or more secondary oligonucleotide primer sets, with one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures and for carrying out one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming secondary polymerase chain reaction products; and

detecting and distinguishing the secondary polymerase chain reaction products, thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

22. A method for identifying, in a sample, one or more target micro-ribonucleic acid (miRNA) molecules differing in sequence from other miRNA molecules in the sample by one or more bases, said method comprising:

providing a sample containing one or more target miRNA molecules potentially differing in sequence from other miRNA molecules in the sample by one or more bases;

providing one or more enzymes capable of digesting deoxyuracil (dU)-containing nucleic acid molecules present in the sample;

contacting the sample with one or more enzymes capable of digesting dU-containing nucleic acid molecules potentially present in the sample;

blending the contacted sample with ATP and a Poly(A) polymerase to form a Poly(A) polymerase reaction mixture;

subjecting the Poly(A) polymerase reaction mixture to conditions suitable for appending a homopolymer A to the 3' ends of the one or more target miRNA molecules potentially present in the sample;

providing one or more primary oligonucleotide primer sets, each primer set comprising (a) a first primary oligonucleotide primer comprising a 5' primer-specific portion, an internal poly dT portion, and a 3' portion comprising from 1 to 10 bases complementary to the 3' end of the target miRNA, wherein the first primary oligonucleotide primer may be the same or may differ from other first primary oligonucleotide primers in other sets, and (b) a second primary oligonucleotide primer comprising a 5' primer-specific portion and a 3' portion, wherein the second primary oligonucleotide primer may be the same or may differ from other second primary oligonucleotide primers in other sets;

blending the Poly(A) polymerase reaction mixture potentially comprising target miRNA sequences with 3' polyA tails, the one or more primary oligonucleotide primer sets, a deoxynucleotide mix, and a reverse transcriptase and a DNA polymerase or a DNA polymerase

with reverse-transcriptase activity to form one or more reverse-transcription/polymerase chain reaction mixtures;

subjecting the one or more reverse-transcription/polymerase chain reaction mixtures to conditions suitable for generating complementary deoxyribonucleic acid (cDNA) molecules to the target miRNA sequences with 3' polyA tails, and to one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming one or more different reverse-transcription/polymerase chain reaction products comprising the 5' primer-specific portion of the second primary oligonucleotide primer, a nucleotide sequence corresponding to the target miRNA molecule sequence, a poly dA region, and the complement of the 5' primer-specific portion of the first primary oligonucleotide primer, and complements thereof;

providing one or more secondary oligonucleotide primer sets, each secondary oligonucleotide primer set comprising (a) a first secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that is complementary to a portion of a reverse-transcription/polymerase chain reaction product formed from the first primary oligonucleotide primer and (b) a second secondary oligonucleotide primer having a 5' primer-specific portion and a 3' portion that comprises a nucleotide sequence that is complementary to a portion of a reverse-transcription/polymerase chain reaction product formed from the first secondary oligonucleotide primer;

blending the reverse-transcription/polymerase chain reaction products, the one or more secondary oligonucleotide primer sets, a deoxynucleotide mix, and a DNA polymerase to form one or more first polymerase chain reaction mixtures;

subjecting the one or more first polymerase chain reaction mixtures to conditions suitable for two or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment, thereby forming first polymerase chain reaction products comprising a 5' primer-specific portion, a nucleotide sequence corresponding to the target miRNA molecule sequence or a complement thereof, and a complement of the other 5' primer-specific portion;

providing one or more tertiary oligonucleotide primer sets, each tertiary oligonucleotide primer set comprising (a) a first tertiary oligonucleotide primer comprising the same nucleotide sequence as the 5' primer-specific portion of the first polymerase chain reaction product sequence and (b) a second tertiary oligonucleotide primer comprising a nucleotide sequence that is complementary to the 3' primer-specific portion of the first polymerase chain reaction product sequence;

blending the first polymerase chain reaction products, the one or more tertiary oligonucleotide primer sets, the one or more enzymes capable of digesting deoxyuracil (dU) containing nucleic acid molecules, a deoxynucleotide mix including dUTP, and a DNA polymerase to form one or more second polymerase chain reaction mixtures;

subjecting the one or more second polymerase chain reaction mixtures to conditions suitable for digesting deoxyuracil (dU)-containing nucleic acid molecules present in the first polymerase chain reaction mixtures, and one or more polymerase chain reaction cycles comprising a denaturation treatment, a hybridization treatment, and an extension treatment thereby forming second polymerase chain reaction products; and

detecting and distinguishing the second polymerase chain reaction products in the one or more reactions thereby identifying one or more target miRNA molecules differing in sequence from other miRNA molecules in the sample by one or more bases.

23. The method of any one of claims 19 through 22 wherein the 3' portion of the second primary oligonucleotide primer comprises ribo-G and/or G nucleotide analogue, wherein the reverse transcriptase appends two or three cytosine nucleotides to the 3' end of the complementary deoxyribonucleic acid products of the target miRNAs, enabling transient hybridization to the 3' end of the second primary oligonucleotide primer, enabling the reverse transcriptase to undergo strand switching and to extend the complementary deoxyribonucleic acid products to include the complementary sequence of the 5' primer-specific portion of the second primary oligonucleotide primer to form the one or more different first polymerase chain reaction products comprising a 5' primer-specific portion, a nucleotide sequence portion corresponding to the target miRNA molecule sequence or a complement thereof, a further portion, and a complement of the other 5' primer-specific portion.

24. The method of any one of claims 19 through 22 wherein the 3' portion of the second primary oligonucleotide primers contains from 6 to 14 bases comprising, from 5' to 3', three ribo-G or G bases, followed by additional bases that are the same as the 5' end of the target miRNA sequences, wherein the reverse transcriptase appends two or three cytosine residues to the 3' end of the initial complementary deoxyribonucleic acid extension products of the target miRNAs, and wherein subsequent to when the denaturation treatment of the polymerase chain reaction is initiated the conditions are adjusted to enable transient hybridization to the 3' end of the second primary oligonucleotide primers to the 3' end of the complementary deoxyribonucleic acid extension products, allowing for extension of either one or both the second primary oligonucleotide primers and the complementary deoxyribonucleic acid

extension products to form the one or more different primary reverse-transcription/polymerase chain reaction products comprising a 5' primer-specific portion, a nucleotide sequence portion corresponding to the target miRNA molecule sequence or a complement thereof, a further portion, and a complement of the other 5' primer-specific portion.

25. The method of any one of claims 1, 4, or 17, wherein the second oligonucleotide probe of the oligonucleotide probe set further comprises a unitaq detection portion, thereby forming ligated product sequences comprising the 5' primer-specific portion, the target-specific portions, the unitaq detection portion, and the 3' primer-specific portion, said method further comprising:

providing one or more unitaq detection probes, wherein each unitaq detection probe hybridizes to a complementary unitaq detection portion and said detection probe comprises a quencher molecule and a detectable label separated from the quencher molecule;

adding the one or more unitaq detection probes to the second polymerase chain reaction mixture; and

hybridizing the one or more unitaq detection probes to complementary unitaq detection portions on the ligated product sequence or complement thereof during said subjecting the second polymerase chain reaction mixture to conditions suitable for one or more polymerase chain reaction cycles, wherein the quencher molecule and the detectable label are cleaved from the one or more unitaq detection probes during the extension treatment and said detecting involves the detection of the cleaved detectable label.

26. The method of any one of claims 2, 3, 5, 6, 7, or 18, wherein one primary oligonucleotide primer or one secondary oligonucleotide primer further comprises a unitaq detection portion, thereby forming extension product sequences comprising the 5' primer-specific portion, the target-specific portions, the unitaq detection portion, and the complement of the other 5' primer-specific portion, and complements thereof, said method further comprising:

providing one or more unitaq detection probes, wherein each unitaq detection probe hybridizes to a complementary unitaq detection portion and said detection probe comprises a quencher molecule and a detectable label separated from the quencher molecule;

adding the one or more unitaq detection probes to the one or more first or second polymerase chain reaction mixtures; and

hybridizing the one or more unitaq detection probes to complementary unitaq detection portions on the ligated product sequence or complement thereof during polymerase chain reaction cycles after the first polymerization chain reaction, wherein the quencher

molecule and the detectable label are cleaved from the one or more unitaq detection probes during the extension treatment and said detecting involves the detection of the cleaved detectable label.

27. The method of any one of claims 1, 4, 17, 19, or 21, wherein one or both oligonucleotide probes of the oligonucleotide probe set comprises a portion that has no or one nucleotide sequence mismatch when hybridized in a base-specific manner to the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof, but have one or more additional nucleotide sequence mismatches that interferes with ligation when said oligonucleotide probe hybridizes in a base-specific manner to a corresponding nucleotide sequence portion in the wildtype nucleic acid sequence or bisulfite-converted unmethylated nucleic acid sequence or complement sequence thereof.

28. The method of any one of claims 1, 4, or 17, wherein the 3' portion of the first oligonucleotide probe of the oligonucleotide probe set comprises a cleavable nucleotide or nucleotide analogue and a blocking group, such that the 3' end is unsuitable for polymerase extension or ligation, said method further comprising;

cleaving the cleavable nucleotide or nucleotide analog of the first oligonucleotide probe when said probe is hybridized to it complementary target nucleotide sequence of the primary extension product, thereby liberating a 3'OH on the first oligonucleotide probe prior to said ligating.

29. The method of claim 28, wherein the one or more first oligonucleotide probe of the oligonucleotide probe set comprises a sequence that differs from the target nucleic acid sequence or bisulfite-converted methylated nucleic acid sequence or complement sequence thereof, said difference is located two or three nucleotide bases from the liberated free 3'OH end.

30. The method of any one of claims 1, 4, or 17, wherein the second oligonucleotide probe has, at its 5' end, an overlapping identical nucleotide with the 3' end of the first oligonucleotide probe, and, upon hybridization of the first and second oligonucleotide probes of a probe set at adjacent positions on a complementary target nucleotide sequence of a primary extension product to form a junction, the overlapping identical nucleotide of the second oligonucleotide probe forms a flap at the junction with the first oligonucleotide probe, said method further comprising:

cleaving the overlapping identical nucleotide of the second oligonucleotide probe with an enzyme having 5' nuclease activity thereby liberating a phosphate at the 5' end of the second oligonucleotide probe prior to said ligating.

31. The method of any one of claims 1, 4, or 17, wherein the one or more oligonucleotide probe sets further comprise a third oligonucleotide probe having a target-specific portion, wherein the second and third oligonucleotide probes of a probe set are configured to hybridize adjacent to one another on the target nucleotide sequence with a junction between them to allow ligation between the second and third oligonucleotide probes to form a ligated product sequence comprising the first, second, and third oligonucleotide probes of a probe set.

32. The method of any one of claims 1 through 31, wherein the sample is selected from the group consisting of tissue, cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, cell-free circulating nucleic acids, cell-free circulating tumor nucleic acids, cell-free circulating fetal nucleic acids in pregnant woman, circulating tumor cells, tumor, tumor biopsy, and exosomes.

33. The method of any one of claims 1 through 31, wherein the one or more target nucleotide sequences are low-abundance nucleic acid molecules comprising one or more nucleotide base mutations, insertions, deletions, translocations, splice variants, miRNA variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicings, exon insertions, exon deletions, intron insertions, or other rearrangement at the genome level and/or methylated nucleotide bases.

34. The method of claim 33, wherein the low-abundance nucleic acid molecules with one or more nucleotide base mutations, insertions, deletions, translocations, splice variants, miRNA variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicings, exon insertions, exon deletions, intron insertions, or other rearrangement at the genome level, and/or methylated nucleotide bases are identified and distinguished from a high-abundance of nucleic acid molecules in the sample having a similar nucleotide sequence as the low abundance nucleic acid molecules but without the one or more nucleotide base mutations, insertions, deletions, translocations, splice variants, miRNA variants, alternative transcripts, alternative start sites, alternative coding sequences, alternative non-coding sequences, alternative splicings, exon

insertions, exon deletions, intron insertions, or other rearrangement at the genome level, and/or methylated nucleotide bases.

35. The method of claim 34, wherein the copy number of one or more low-abundance target nucleotide sequences are quantified relative to the copy number of the high-abundance nucleic acid molecules in the sample.

36. The method of any one of claims 1 through 31, wherein the one or more target nucleotide sequences are quantified or enumerated.

37. The method of claim 36, wherein the one or more target nucleotide sequences are quantified or enumerated relative to other nucleotide sequences in the sample or other samples undergoing the identical subsequent steps.

38. The method of claim 37, wherein the relative copy number of one or more target nucleotide sequences are quantified or enumerated.

39. The method of any one of claims 1 through 31 further comprising:

diagnosing or prognosing a disease state based on said identifying.

40. The method of any one of claims 1 through 31 further comprising:

distinguishing a genotype or disease predisposition based on said identifying.

41. A method of diagnosing or prognosing a disease state of cells or tissue based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual, wherein the plurality of markers is in a set comprising from 6-12 markers, 12-24 markers, 24-36 markers, 36-48 markers, 48-72 markers, 72-96 markers, or > 96 markers, wherein each marker in a given set is selected by having any one or more of the following criteria:

present, or above a cutoff level, in > 50% of biological samples of the disease cells or tissue from individuals diagnosed with the disease state;

absent, or below a cutoff level, in > 95% of biological samples of the normal cells or tissue from individuals without the disease state;

present, or above a cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with the disease state;

absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without the disease state;

present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with the disease state;

and, wherein at least 50% of the markers in a set each comprise one or more methylated residues, and/or wherein at least 50% of the markers in a set that are present, or above a cutoff level, or present with a z-value of > 1.65, comprise of one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with the disease state, said method comprising:

obtaining a biological sample including cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof;

fractionating the sample into one or more fractions, wherein at least one fraction comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein;

subjecting nucleic acid molecules in the one or more fractions to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues;

carrying out at least two enrichment steps for 50% or more disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step; and

performing one or more assays to detect and distinguish the plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with the disease state if a minimum of 2 or 3 markers are present or are above a cutoff level in a marker set comprising from 6-12 markers; or a minimum of 3, 4, or 5 markers are present or are above a cutoff level in a marker set comprising from 12-24 markers; or a minimum of 3, 4, 5, or 6 markers are present or are above a cutoff level in a marker set comprising from 24-36 markers; or a minimum of 4, 5, 6, 7, or 8 markers are present or are above a cutoff level in a marker set

comprising from 36-48 markers; or a minimum of 6, 7, 8, 9, 10, 11, or 12 markers are present or are above a cutoff level in a marker set comprising from 48-72 markers, or a minimum of 7, 8, 9, 10, 11, 12 or 13 markers are present or are above a cutoff level in a marker set comprising from 72-96 markers, or a minimum of 8, 9, 10, 11, 12, 13 or "n"/12 markers are present or are above a cutoff level in a marker set comprising 96 – "n" markers, when "n" > 168 markers.

42. A method of diagnosing or prognosing a disease state of a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual, wherein the plurality of markers is in a set comprising from 48-72 total cancer markers, 72-96 total cancer markers or ≥ 96 total cancer markers, wherein on average greater than one quarter such markers in a given set cover each of the aforementioned major cancers being tested, wherein each marker in a given set for a given solid tissue cancer is selected by having any one or more of the following criteria for that solid tissue cancer:

present, or above a cutoff level, in > 50% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer;

absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer;

present, or above a cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer;

absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer;

present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer;

and, wherein at least 50% of the markers in a set each comprise one or more methylated residues, and/or wherein at least 50% of the markers in a set that are present, or

above a cutoff level, or present with a z-value of > 1.65, comprise of one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with a given solid tissue cancer, said method comprising:

obtaining a biological sample including cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof;

fractionating the sample into one or more fractions, wherein at least one fraction comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein;

subjecting the nucleic acid molecules in the one or more fractions to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues;

carrying out at least two enrichment steps for 50% or more of the given solid tissue cancer-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step; and

performing one or more assays to detect and distinguish the plurality of cancer - specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with the a solid-tissue cancer if a minimum of 4 markers are present or are above a cutoff level in a marker set comprising from 48-72 total cancer markers; or a minimum of 5 markers are present or are above a cutoff level in a marker set comprising from 72-96 total cancer markers; or a minimum of 6 or "n"/18 markers are present or are above a cutoff level in a marker set comprising 96 to "n" total cancer markers, when "n" > 96 total cancer markers.

43. The method of claim 42, wherein each marker in a given set for a given solid tissue cancer is selected by having any one or more of the following criteria for that solid tissue cancer:

present, or above a cutoff level, in > 66% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer;

absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer;

present, or above a cutoff level, in > 66% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer;

absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer;

present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer.

44. A method of diagnosing or prognosing a disease state of and identifying the most likely specific tissue(s) of origin of a solid tissue cancer in the following groups: Group 1 (colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma); Group 2 (breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma); Group 3 (lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma); Group 4 (prostate adenocarcinoma, invasive urothelial bladder cancer); and/or Group 5 (liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma) based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual, wherein the plurality of markers is in a set comprising from 36-48 group-specific cancer markers, 48-64 group-specific cancer markers, or ≥ 64 group-specific cancer markers, wherein on average greater than one third of such markers in a given set cover each of the aforementioned cancers being tested within that group, wherein each marker in a given set for a given solid tissue cancer is selected by having any one or more of the following criteria for that solid tissue cancer:

present, or above a cutoff level, in > 50% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer;

absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer;

present, or above a cutoff level, in > 50% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer;

absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer;

present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer;

and, wherein at least 50% of the markers in a set each comprise one or more methylated residues, and/or wherein at least 50% of the markers in a set that are present, or above a cutoff level, or present with a z-value of > 1.65 comprise one or more methylated residues, in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from at least 50% of individuals diagnosed with a given solid tissue cancer, said method comprising:

obtaining the biological sample, the biological sample including cell-free DNA, RNA, and/or protein originating from the cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof;

fractionating the sample into one or more fractions, wherein at least one fraction comprises exosomes, tumor-associated vesicles, other protected states, or cell-free DNA, RNA, and/or protein;

subjecting the nucleic acid molecules in the one or more fractions to a bisulfite treatment under conditions suitable to convert unmethylated cytosine residues to uracil residues;

carrying out at least two enrichment steps for 50% or more of the given solid tissue cancer-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers during either said fractionating and/or by carrying out a nucleic acid amplification step; and

performing one or more assays to detect and distinguish the plurality of cancer - specific and/or cell/tissue-specific DNA, RNA, and/or protein markers, thereby identifying their presence or levels in the sample, wherein individuals are diagnosed or prognosed with a solid-tissue cancer if a minimum of 4 markers are present or are above a cutoff level in a marker set comprising from 36-48 group-specific cancer markers; or a minimum of 5 markers are present or are above a cutoff level in a marker set comprising from 48-64 group-specific cancer markers; or a minimum of 6 or "n"/12 markers are present or are above a cutoff level in a marker set comprising 64 to "n" total cancer markers, when "n" > 64 group-specific cancer markers.

45. The method of claim 44, wherein each marker in a given set for a given solid tissue cancer is selected by having any one or more of the following criteria for that solid tissue cancer:

present, or above a cutoff level, in > 66% of biological samples of a given cancer tissue from individuals diagnosed with a given solid tissue cancer;

absent, or below a cutoff level, in > 95% of biological samples of the normal tissue from individuals without that given solid tissue cancer;

present, or above a cutoff level, in > 66% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer;

absent, or below a cutoff level, in > 95% of biological samples comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals without that given solid tissue cancer;

present with a z-value of > 1.65 in the biological sample comprising cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, bodily excretions, or fractions thereof, from individuals diagnosed with a given solid tissue cancer.

46. The method of any one of claims 41 through 45, wherein the at least two enrichment steps comprise of two or more of the following steps:

capturing or separating exosomes or extracellular vesicles or markers in other protected states; capturing or separating a platelet fraction; capturing or separating circulating tumor cells; capturing or separating RNA-containing complexes; capturing or separating cfDNA-nucleosome or differentially modified cfDNA-histone complexes; capturing or separating protein targets or protein target complexes; capturing or separating auto-antibodies; capturing or separating cytokines; capturing or separating methylated cfDNA; capturing or separating marker specific DNA, cDNA, miRNA, lncRNA, ncRNA, or mRNA, or amplified complements, by hybridization to complementary capture probes in solution, on magnetic beads, or on a microarray; amplifying miRNA markers, non-coding RNA markers (lncRNA & ncRNA markers), mRNA markers, exon markers, splice-variant markers, translocation markers, or copy number variation markers in a linear or exponential manner via a polymerase extension reaction, polymerase chain reaction, bisulfite-methyl-specific polymerase chain reaction, reverse-transcription reaction, bisulfite-methyl-specific ligation reaction, and/or ligation reaction, using DNA polymerase, reverse transcriptase, DNA ligase, RNA ligase, DNA repair enzyme, RNase, RNaseH2, endonuclease, restriction endonuclease, exonuclease, CRISPR, DNA glycosylase or combinations thereof; selectively amplifying one or more target regions containing mutation markers or bisulfite-converted DNA methylation markers, while suppressing amplification of the target regions containing wild-type sequence or bisulfite-converted unmethylated sequence or complement sequence thereof, in a linear or exponential manner via a polymerase extension

reaction, polymerase chain reaction, bisulfite-methyl-specific polymerase chain reaction, reverse-transcription reaction, bisulfite-methyl-specific ligation reaction, and/or ligation reaction, using DNA polymerase, reverse transcriptase, DNA ligase, RNA ligase, DNA repair enzyme, RNase, RNaseH2, endonuclease, restriction endonuclease, exonuclease, CRISPR, DNA glycosylase or combinations thereof; preferentially extending, ligating, or amplifying one or more primers or probes whose 3'-OH end has been liberated in an enzyme and sequence-dependent process; using one or more blocking oligonucleotide primers comprising one or more mismatched bases at the 3' end or comprising one or more nucleotide analogs and a blocking group at the 3' end under conditions that interfere with polymerase extension or ligation during said reaction of target-specific primer or probes hybridized in a base-specific manner to wildtype sequence or bisulfite-converted unmethylated sequence or complement sequence thereof.

47. The method of any one of claims 41 through 46, wherein the one or more assays to detect and distinguish the plurality of disease-specific and/or cell/tissue-specific DNA, RNA, or protein markers comprise one or more of the following:

a quantitative real-time PCR method (qPCR); a reverse transcriptase-polymerase chain reaction (RTPCR) method; a bisulfite qPCR method; a digital PCR method (dPCR); a bisulfite dPCR method; a ligation detection method, a ligase chain reaction, a restriction endonuclease cleavage method; a DNA or RNA nuclease cleavage method; a micro-array hybridization method; a peptide-array binding method; an antibody-array method; a mass spectrometry method; a liquid chromatography-tandem mass spectrometry (LC-MS/MS) method; a capillary or gel electrophoresis method; a chemiluminescence method; a fluorescence method; a DNA sequencing method; a bisulfite conversion-DNA sequencing method; an RNA sequencing method; a proximity ligation method; a proximity PCR method; a method comprising immobilizing an antibody-target complex; a method comprising immobilizing an aptamer-target complex; an immunoassay method; a method comprising a Western blot assay; a method comprising an enzyme linked immunosorbent assay (ELISA); a method comprising a high-throughput microarray-based enzyme-linked immunosorbent assay (ELISA); or a method comprising a high-throughput flow-cytometry-based enzyme-linked immunosorbent assay (ELISA).

48. The method of any one of claims 41 through 47, wherein the one or more cutoff levels of the one or more assays to detect and distinguish the plurality of disease-specific and/or cell/tissue-specific DNA, RNA, or protein markers comprise one or more of the following

calculations, comparisons, or determinations, in the one or more marker assays comparing samples from the disease vs. normal individual:

marker ΔCt value is > 2; marker ΔCt value is > 4; ratio of detected marker-specific signal is > 1.5; ratio of detected marker-specific signal is > 3; ratio of marker concentrations is > 1.5; ratio of marker concentrations is > 3; enumerated marker-specific signals differ by > 20%; enumerated marker-specific signals differ by > 50%; marker-specific signal from a given disease sample is > 85%; > 90%; > 95%; > 96%; > 97%; or > 98% of the same marker-specific signals from a set of normal samples; or marker-specific signal from a given disease sample has a z-score of > 1.03; > 1.28; > 1.65; > 1.75; > 1.88; or > 2.05 compared to the same marker-specific signals from a set of normal samples.

49. A two-step method of diagnosing or prognosing a disease state of cells or tissue based on identifying the presence or level of a plurality of disease-specific and/or cell/tissue-specific DNA, RNA, and/or protein markers in a biological sample of an individual, said two-step method comprising:

obtaining a biological sample, the biological sample including exosomes, tumor-associated vesicles, markers within other protected states, cell-free DNA, RNA, and/or protein originating from the potentially disease state cells or tissue and from one or more other tissues or cells, wherein the biological sample is selected from the group consisting of cells, serum, blood, plasma, amniotic fluid, sputum, urine, bodily fluids, bodily secretions, and bodily excretions, or fractions thereof;

applying a first step to the biological samples with an overall sensitivity of > 80% and an overall specificity of > 90% or an overall Z-score of > 1.28 to identify individuals more likely to be diagnosed or prognosed with the disease state; and

applying a second step to biological samples from those individuals identified in the first step with an overall specificity of > 95% or an overall Z-score of > 1.65 to diagnose or prognose individuals with the disease state, wherein said applying the first step and/or said applying the second step is carried out using the method of one of claims 41-44.

50. The method of any one of claims 41 through 49, wherein the disease state is a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder

cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein at least 50% of the markers in a set each comprise one or more methylated cytosine residues of a CpG sequence, or the complement of one or more methylated cytosine residues of a CpG sequence selected from the list in Figure 56.

51. The method of one of claims 41 through 49, wherein the disease state is a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein at least 50% of the markers in a set each comprise of one or more methylated residues of one or more chromosomal sub-regions selected from the list in Figure 57.

52. The method of one of claims 41 through 49, wherein the disease state is a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein the one or more markers in a set comprise of one or more miRNA sequences (mir ID, Gene ID) selected from the group consisting of: hsa-mir-21, MIR21; hsa-mir-182, MIR182; hsa-mir-454, MIR454; hsa-mir-96, MIR96; hsa-mir-183, MIR183; hsa-mir-549, MIR549; hsa-mir-301[a], MIR301A; hsa-mir-548f-1, MIR548F1; hsa-mir-301b, MIR301B; hsa-mir-103-1, MIR1031; hsa-mir-18[a], MIR18A; hsa-mir-147b, MIR147B; hsa-mir-4326, MIR4326; and hsa-mir-573, MIR573, or one or more lncRNA or ncRNA sequences selected from the list in Figure 53.

53. The method of one of claims 41 through 49, wherein the disease state is a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous

cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein the one or more markers in a set comprise of one or more Exon RNA sequences selected from the list in Figure 54.

54. The method of one of claims 41 through 49, wherein the disease state is a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein the one or more markers in a set comprise of one or more mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product selected from the list in Figure 55 or from the group consisting of (Protein name, UniProt ID): Uncharacterized protein C19orf48, Q6RUI8; Protein FAM72B, Q86X60; Protein FAM72D, Q6L9T8; Hydroxyacylglutathione hydrolase-like protein, Q6PII5; Putative methyltransferase NSUN5, Q96P11; RNA pseudouridylate synthase domain-containing protein 1, Q9UJJ7; Collagen triple helix repeat-containing protein 1, Q96CG8; Interleukin-11. P20809; Stromelysin-2, P09238; Matrix metalloproteinase-9 , P14780; Podocan-like protein 1, Q6PEZ8; Putative peptide YY-2, Q9NRI6; Osteopontin, P10451; Sulfhydryl oxidase 2, Q6ZRP7; Glypican-2, Q8N158; Macrophage migration inhibitory factor, P14174; Peptidyl-prolyl cis-trans isomerase A, P62937; and Calreticulin, P27797.

55. The method of one of claims 41 through 49, wherein the disease state is a solid tissue cancer including colorectal adenocarcinoma, stomach adenocarcinoma, esophageal carcinoma, breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, uterine carcinosarcoma, lung adenocarcinoma, lung squamous cell carcinoma, head & neck squamous cell carcinoma, prostate adenocarcinoma, invasive urothelial bladder cancer, liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein the one or more markers in a set comprise of one or more mutations, insertions, deletions, copy number changes, or expression changes in a gene selected from the group consisting of TP53 (tumor protein p53), TTN (titin), MUC16 (mucin 16), and KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog).

56. The method of one of claims 41 through 49, wherein the disease state is colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma, wherein at least 50% of the markers in a set each comprise one or more methylated cytosine residues of a CpG sequence, or the complement of one or more methylated cytosine residues of a CpG sequence selected from the list in Figure 44 or in Figure 59.

57. The method of one of claims 41 through 49, wherein the disease state is colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma, wherein at least 50% of the markers in a set each comprise of one or more methylated residues of one or more chromosomal sub-regions selected from the list in Figure 45 or in Figure 60.

58. The method of one of claims 41 through 49, wherein the disease state is colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma, wherein the one or more markers in a set comprise of one or more miRNA sequences selected from the list in Figure 39; hsa-mir-624, MIR624; or one or more lncRNA or ncRNA sequences selected from the list in Figure 40 or the group consisting of [Gene ID, Coordinate (GRCh38)]: ENSEMBL ID: LINC01558, chr6:167784537-167796859, and ENSG00000146521.8.

59. The method of one of claims 41 through 49, wherein the disease state is colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma, wherein the one or more markers in a set comprise of one or more Exon RNA sequences selected from the list in Figure 41 or in Figure 58.

60. The method of one of claims 41 through 49, wherein the disease state is colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma, wherein the one or more markers in a set comprise of one or more mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product selected from the list in Figure 42, Figure 43, or from the group consisting of (Gene Symbol, Chromosome Band, Gene Title, UniProt ID): SELE, 1q22-q25, selectin E, P16581; OTUD4, 4q31.21, OTU domain containing 4, Q01804; BPI, 20q11.23, bactericidal/permeability-increasing protein, P17213; ASB4, 7q21-q22, ankyrin repeat and SOCS box containing 4, Q9Y574; C6orf123, 6q27, chromosome 6 open reading frame 123,

Q9Y6Z2; KPNA3, 13q14.3, karyopherin alpha 3 (importin alpha 4), O00505; and NUP98, 11p15, nucleoporin 98kDa, P52948; or (Protein name, UniProt ID) Bactericidal permeability-increasing protein (BPI) (CAP 57), P17213.

61. The method of one of claims 41 through 49, wherein the disease state is colon adenocarcinoma, rectal adenocarcinoma, stomach adenocarcinoma, or esophageal carcinoma, wherein the one or more markers in a set comprise of one or more mutations, insertions, deletions, copy number changes, or expression changes in a gene selected from the group consisting of APC (APC regulator of WNT signaling pathway), ATM (ATM serine/threonine kinase), CSMD1 (CUB and Sushi multiple domains 1), DNAH11 (dynein axonemal heavy chain 11), DST (dystonin), EP400 (E1A binding protein p400), FAT3 (FAT atypical cadherin 3), FAT4 (FAT atypical cadherin 4), FLG (filaggrin), GLI3 (GLI family zinc finger 3), KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog), LRP1B (LDL receptor related protein 1B), MUC16 (mucin 16, cell surface associated), OBSCN (obscurin, cytoskeletal calmodulin and titin-interacting RhoGEF), PCLO (piccolo presynaptic cytomatrix protein), PIK3CA (phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha), RYR2 (ryanodine receptor 2), SYNE1 (spectrin repeat containing nuclear envelope protein 1), TP53 (tumor protein p53), TTN (titin), and UNC13C (unc-13 homolog C).

62. The method of one of claims 41 through 49, wherein the disease state is breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, or uterine carcinosarcoma, wherein at least 50% of the markers in a set each comprise one or more methylated cytosine residues of a CpG sequence, or the complement of one or more methylated cytosine residues of a CpG sequence selected from the list in Figure 61.

63. The method of one of claims 41 through 49, wherein the disease state is breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, or uterine carcinosarcoma, wherein at least 50% of the markers in a set each comprise of one or more methylated residues of one or more chromosomal sub-regions selected from the list in Figure 62.

64. The method of one of claims 41 through 49, wherein the disease state is breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, or uterine

carcinosarcoma, wherein the one or more markers in a set comprise of one or more miRNA sequences selected from the group consisting of (mir ID, Gene ID): hsa-mir-1265, MIR1265.

65. The method of one of claims 41 through 49, wherein the disease state is breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, or uterine carcinosarcoma, wherein the one or more markers in a set comprise of one or more Exon RNA sequences (Exon location, Gene) selected from the group consisting of: chr2:179209013-179209087:+ , OSBPL6; chr2:179251788-179251866:+ , OSBPL6; and chr2:179253736-179253880:+ , OSBPL6.

66. The method of one of claims 41 through 49, wherein the disease state is breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, or uterine carcinosarcoma, wherein the one or more markers in a set comprise of one or more mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product selected from the group consisting of (Gene Symbol, Chromosome Band , Gene Title, UniProt ID): RSPO2, 8q23.1, R-spondin 2, Q6UXX9; KLC4, 6p21.1, kinesin light chain 4, Q9NSK0; and GLRX, 5q14, glutaredoxin (thioltransferase), P35754; or (Protein name, UniProt ID) R-spondin-2 (Roof plate-specific spondin-2) (hRspo2), Q6UXX9.

67. The method of one of claims 41 through 49, wherein the disease state is breast lobular and ductal carcinoma, uterine corpus endometrial carcinoma, ovarian serous cystadenocarcinoma, cervical squamous cell carcinoma and adenocarcinoma, or uterine carcinosarcoma, wherein the one or more markers in a set comprise of one or more mutations, insertions, deletions, copy number changes, or expression changes in a gene selected from the group consisting of PIK3CA (phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit alpha), and TTN (titin).

68. The method of one of claims 41 through 49, wherein the disease state is lung adenocarcinoma, lung squamous cell carcinoma, or head & neck squamous cell carcinoma, wherein at least 50% of the markers in a set each comprise one or more methylated cytosine residues of a CpG sequence, or the complement of one or more methylated cytosine residues of a CpG sequence selected from the list in Figure 63.

69. The method of one of claims 41 through 49, wherein the disease state is lung adenocarcinoma, lung squamous cell carcinoma, or head & neck squamous cell carcinoma, wherein at least 50% of the markers in a set each comprise of one or more methylated residues of one or more chromosomal sub-regions selected from the list in Figure 64.

70. The method of one of claims 41 through 49, wherein the disease state is lung adenocarcinoma, lung squamous cell carcinoma, or head & neck squamous cell carcinoma, wherein the one or more markers in a set comprise of one or more miRNA sequences selected from (mir ID, Gene ID): hsa-mir-28, MIR28.

71 The method of one of claims 41 through 49, wherein the disease state is lung adenocarcinoma, lung squamous cell carcinoma, or head & neck squamous cell carcinoma, wherein the one or more markers in a set comprise of one or more Exon RNA sequences (Exon location, Gene) selected from the group consisting of: chr2: chr1:93307721-93309752:- , FAM69A; chr1:93312740-93312916:- , FAM69A; chr1:93316405-93316512:- , FAM69A; chr1:93341853-93342152:- , FAM69A; chr1:93426933-93427079:- , FAM69A; chr7:40221554-40221627:+ , C7orf10; chr7:40234539-40234659:+ , C7orf10; chr8:22265823-22266009:+ , SLC39A14; chr8:22272293-22272415:+ , SLC39A14; chr14:39509936-39510091:- , SEC23A; and chr14:39511990-39512076:- , SEC23A.

72. The method of one of claims 41 through 49, wherein the disease state is lung adenocarcinoma, lung squamous cell carcinoma, or head & neck squamous cell carcinoma, wherein the one or more markers in a set comprise of one or more mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product selected from the group consisting of (Gene Symbol, Chromosome Band, Gene Title, UniProt ID): STRN3, 14q13-q21, striatin, calmodulin binding protein 3, Q13033; LRRC17, 7q22.1, leucine rich repeat containing 17, Q8N6Y2; FAM69A, 1p22, family with sequence similarity 69, member A, Q5T7M9; ATF2, 2q32, activating transcription factor 2, P15336; BHMT, 5q14.1, betaine--homocysteine S-methyltransferase, Q93088; ODZ3/TENM3, 4q34.3-q35.1, teneurin transmembrane protein 3, Q9P273; and ZFHX4, 8q21.11, zinc finger homeobox 4, Q86UP3; or (Protein name , UniProt ID): Leucine-rich repeat-containing protein 17 (p37NB), Q8N6Y2.

73. The method of one of claims 41 through 49, wherein the disease state is lung adenocarcinoma, lung squamous cell carcinoma, or head & neck squamous cell carcinoma,

wherein the one or more markers in a set comprise of one or more mutations, insertions, deletions, copy number changes, or expression changes in a gene selected from the group consisting of CSMD3 (CUB and Sushi multiple domains 3), DNAH5 (dynein axonemal heavy chain 5), FAT1 (FAT atypical cadherin 1), FLG (filaggrin), KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog), LRP1B (LDL receptor related protein 1B), MUC16 (mucin 16, cell surface associated), PCLO (piccolo presynaptic cytomatrix protein), PKHD1L1 (PKHD1 like 1), RELN (reelin), RYR2 (ryanodine receptor 2), SI (sucrase-isomaltase), SYNE1 (spectrin repeat containing nuclear envelope protein 1), TP53 (tumor protein p53), TTN (titin), USH2A (usherin), and XIRP2 (xin actin binding repeat containing 2).

74. The method of one of claims 41 through 49, wherein the disease state is prostate adenocarcinoma or invasive urothelial bladder cancer, wherein at least 50% of the markers in a set each comprise one or more methylated cytosine residues of a CpG sequence, or the complement of one or more methylated cytosine residues of a CpG sequence selected from the list in Figure 65.

75. The method of one of claims 41 through 49, wherein the disease state is prostate adenocarcinoma or invasive urothelial bladder cancer, wherein at least 50% of the markers in a set each comprise of one or more methylated residues of one or more chromosomal sub-regions selected from the list in Figure 66.

76. The method of one of claims 41 through 49, wherein the disease state is prostate adenocarcinoma or invasive urothelial bladder cancer, wherein the one or more markers in a set comprise of one or more miRNA sequences selected from the the group consisting of (mir ID, Gene ID): hsa-mir-491, MIR491; and hsa-mir-1468, MIR1468, or one or more lncRNA or ncRNA sequences selected from the group consisting of [Gene ID, Coordinate (GRCh38), ENSEMBL ID]: AC007383.3, chr2:206084605-206086564, ENSG00000227946.1; and LINC00324, chr17:8220642-8224043, ENSG00000178977.3.

77. The method of one of claims 41 through 49, wherein the disease state is prostate adenocarcinoma or invasive urothelial bladder cancer, wherein the one or more markers in a set comprise of one or more Exon RNA sequences selected from (Exon location, Gene); chr21:45555942-45556055:+ , C21orf33.

78. The method of one of claims 41 through 49, wherein the disease state is prostate adenocarcinoma or invasive urothelial bladder cancer, wherein the one or more markers in a set comprise of one or more mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product selected from (Gene Symbol, Chromosome Band, Gene Title , UniProt ID): PMM1, 22q13, phosphomannomutase 1, Q92871.

79. The method of one of claims 41 through 49, wherein the disease state is prostate adenocarcinoma or invasive urothelial bladder cancer, wherein the one or more markers in a set comprise of one or more mutations, insertions, deletions, copy number changes, or expression changes in a gene selected from the group consisting of BAGE2 (BAGE family member 2), DNM1P47 (dynamin 1 pseudogene 47), FRG1BP (region gene 1 family member B, pseudogene), KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog), RP11-156P1.3, TTN (titin), and TUBB8P7 (tubulin beta 8 class VIII pseudogene 7).

80. The method of one of claims 41 through 49, wherein the disease state is liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein at least 50% of the markers in a set each comprise one or more methylated cytosine residues of a CpG sequence, or the complement of one or more methylated cytosine residues of a CpG sequence selected from the list in Figure 70.

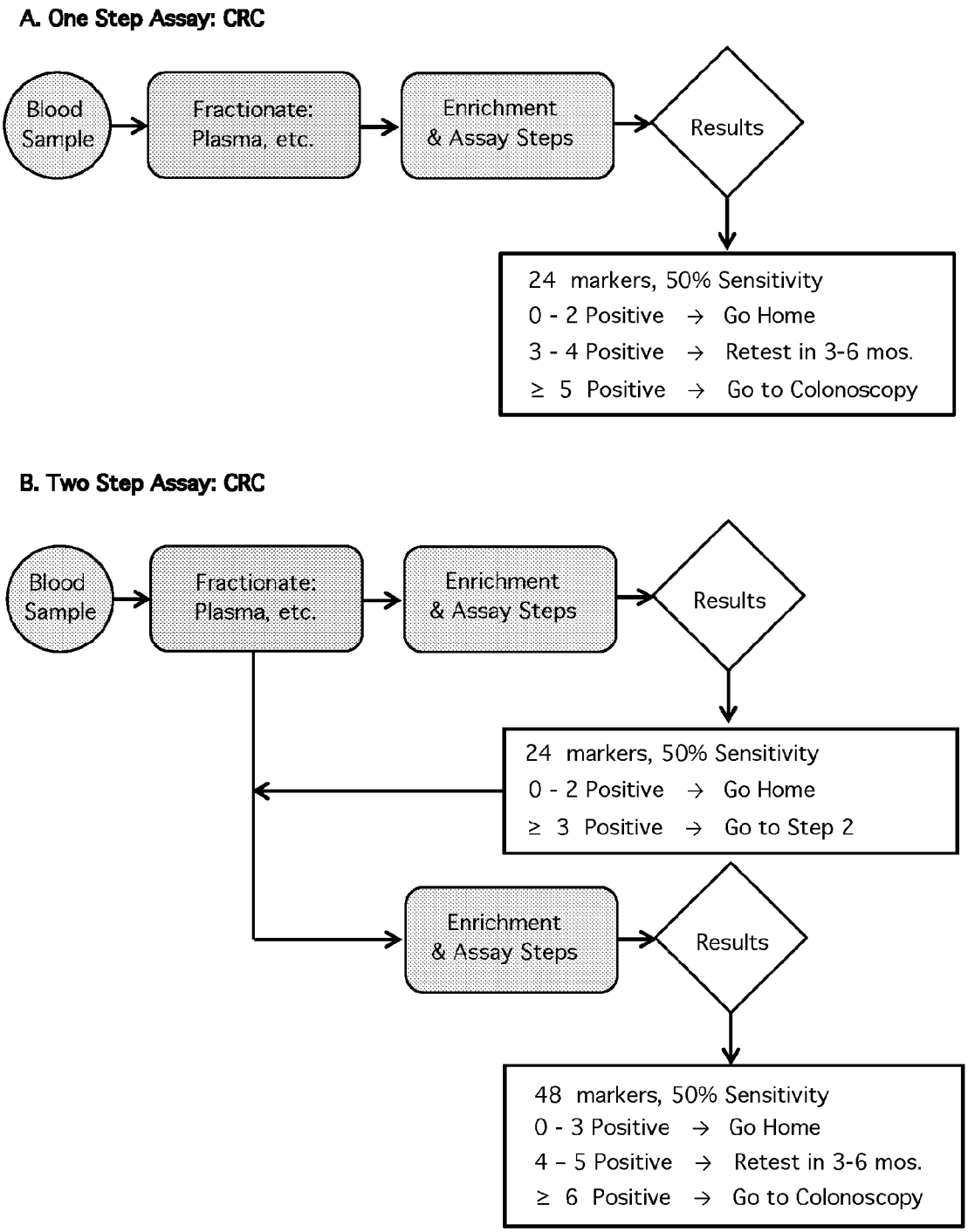
81. The method of one of claims 41 through 49, wherein the disease state is liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein at least 50% of the markers in a set each comprise of one or more methylated residues of one or more chromosomal sub-regions selected from the list in Figure 71.

82. The method of one of claims 41 through 49, wherein the disease state is liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein the one or more markers in a set comprise of one or more miRNA sequences selected from (mir ID, Gene ID): hsa-mir-132, MIR132, or one or more lncRNA or ncRNA sequences selected from the list in Figure 67.

83. The method of one of claims 41 through 49, wherein the disease state is liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein the one or more markers in a set comprise of one or more Exon RNA sequences selected from the list in Figure 68.

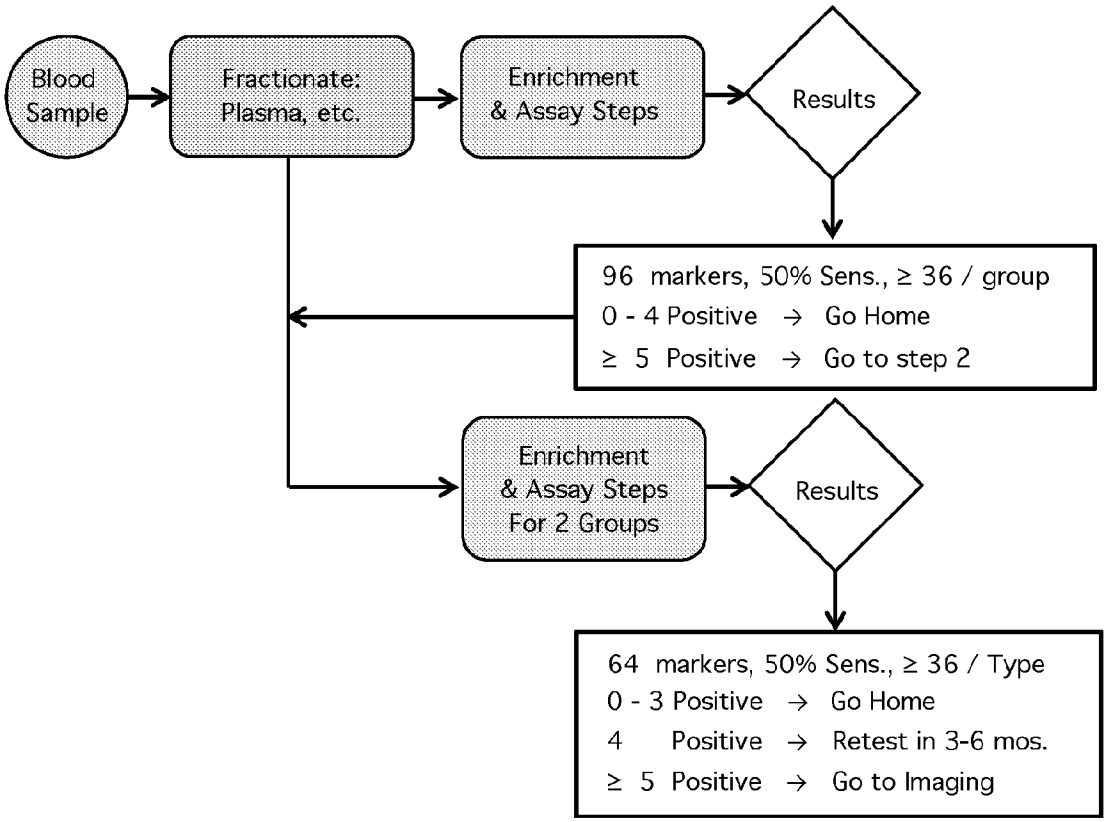
84. The method of one of claims 41 through 49, wherein the disease state is liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein the one or more markers in a set comprise of one or more mRNA sequences, protein expression levels, protein product concentrations, cytokines, or autoantibody to the protein product selected from the list in Figure 69 or from the group consisting of (Protein name, UniProt ID); Gelsolin (AGEL) (Actin-depolymerizing factor) (ADF) (Brevin), P06396; Pro-neuregulin-2, O14511; CD59 glycoprotein (1F5 antigen) (20 kDa homologous restriction factor) (HRF-20) (HRF20) (MAC-inhibitory protein) (MAC-IP) (MEM43 antigen) (Membrane attack complex inhibition factor) (MACIF) (Membrane inhibitor of reactive lysis) (MIRL) (Protectin) (CD antigen CD59), P13987; and Divergent protein kinase domain 2B (Deleted in autism-related protein 1), Q9H7Y0.

85. The method of one of claims 41 through 49, wherein the disease state is liver hepatoceullular carcinoma, pancreatic ductal adenocarcinoma, or gallbladder adenocarcinoma, wherein the one or more markers in a set comprise of one or more mutations, insertions, deletions, copy number changes, or expression changes in a gene selected from the group consisting of KRAS (Ki-ras2 Kirsten rat sarcoma viral oncogene homolog), MUC16 (mucin 16, cell surface associated), MUC4 (mucin 4, cell surface associated), TP53 (tumor protein p53), and TTN (titin).


A. One Step Assay: CRC
Blood Sample
Fractionate: Plasma, etc.
Enrichment & Assay Steps
Results
24 markers, 50% Sensitivity
0 - 2 Positive → Go Home
3 - 4 Positive → Retest in 3-6 mos.
≥ 5 Positive → Go to Colonoscopy
B. Two Step Assay: CRC
Blood Sample
Fractionate: Plasma, etc.
Enrichment & Assay Steps
Results
24 markers, 50% Sensitivity
0 - 2 Positive → Go Home
≥ 3 Positive → Go to Step 2
Enrichment & Assay Steps
Results
48 markers, 50% Sensitivity
0 - 3 Positive → Go Home
4 – 5 Positive → Retest in 3-6 mos.
≥ 6 Positive → Go to Colonoscopy

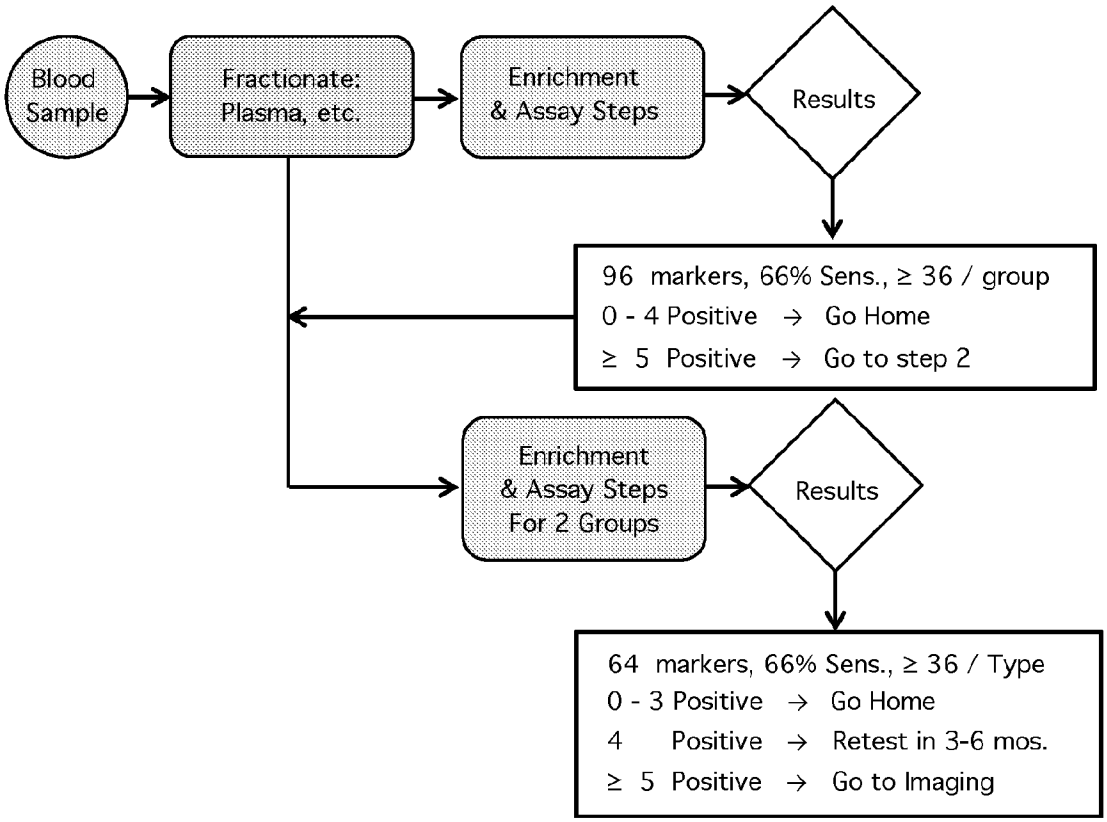
Figures 1A-B

C. Two Step Assay: Pan Oncology, Detection of Early Cancer



Figures 1C

D. Two Step Assay: Pan Oncology, Detection of Early Cancer



Figures 1D

A. exPCR-LDR-qPCR carryover prevention reaction to detect low-level mutation. Isolate genomic or cfDNA. Treat with UDG for carryover prevention.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA Wt probes facilitate preferential amplification of mutation containing regions. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Add downstream primers, (optionally, containing identical 8-11 base tails to prevent primer dimers), treat with UDG, and perform PCR (and dUTP).

D. PCR products incorporate dU, allowing for carryover prevention.

E. Mutation-specific ligation oligonucleotides (Mt) contain tags (Ai, Ci') for subsequent PCR amplification. Blocking LNA or PNA Wt probes suppress ligation to Wt sequence. Upstream ligation primer contains block, which may be removed by cleaving ribose base with RNaseH2 only when hybridized to target.

F. Ligase covalently seals the unblocked mutation-specific upstream and phosphorylated downstream oligonucleotides together. Aliquot into separate wells, micro-pores, or droplets for detection.

G. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci'). Each Taqman probe spans the ligation junction.

H. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

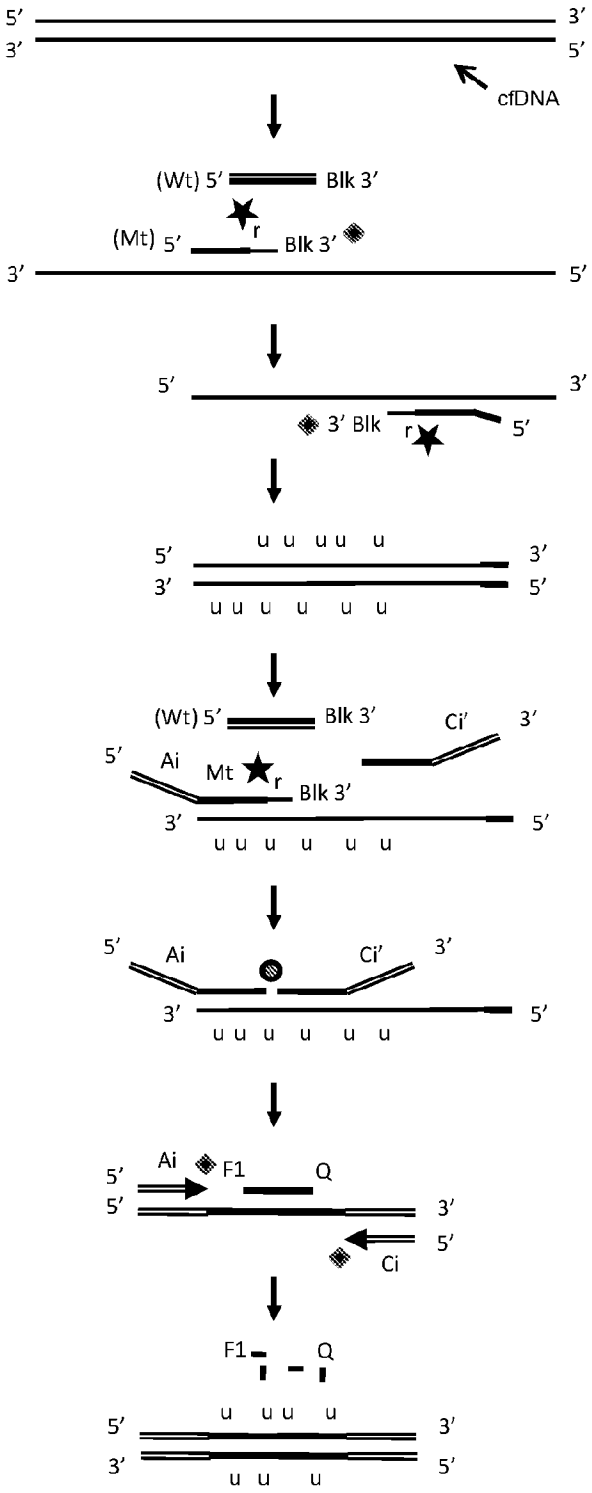


Figure 2

A. exPCR-LDR-qPCR carryover prevention reaction to detect low-level mutation. Isolate genomic or cfDNA. Treat with UDG for carryover prevention.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA Wt probes facilitate preferential amplification of mutation containing regions. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Add downstream primers, (optionally, containing identical 8-11 base tails to prevent primer dimers), treat with UDG, and perform PCR (and dUTP). PCR products incorporate dU, allowing for carryover prevention.

D. Mutation-specific ligation oligonucleotides (Mt) contain tags (Ai, Bi', Ci') for subsequent PCR amplification. Blocking LNA or PNA Wt probes suppress ligation to Wt sequence. Upstream ligation primer contains block, which may be removed by cleaving ribose base with RNaseH2 only when hybridized to target.

E. Ligase covalently seals the unblocked mutation-specific upstream and phosphorylated downstream oligonucleotides together. Aliquot into separate wells, micro-pores, or droplets for detection.

F. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with Unitaq-specific primers (F1-Bi-Q-Ai, Ci).

G. Double-stranded DNA products of PCR; the polymerase blocker stops extension of the bottom strand.

H. After the denaturation step, as the temperature decreases, a hairpin forms between Bi and Bi'. The 5'->3' nuclease activity of Taq polymerase extends primer Ci and liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

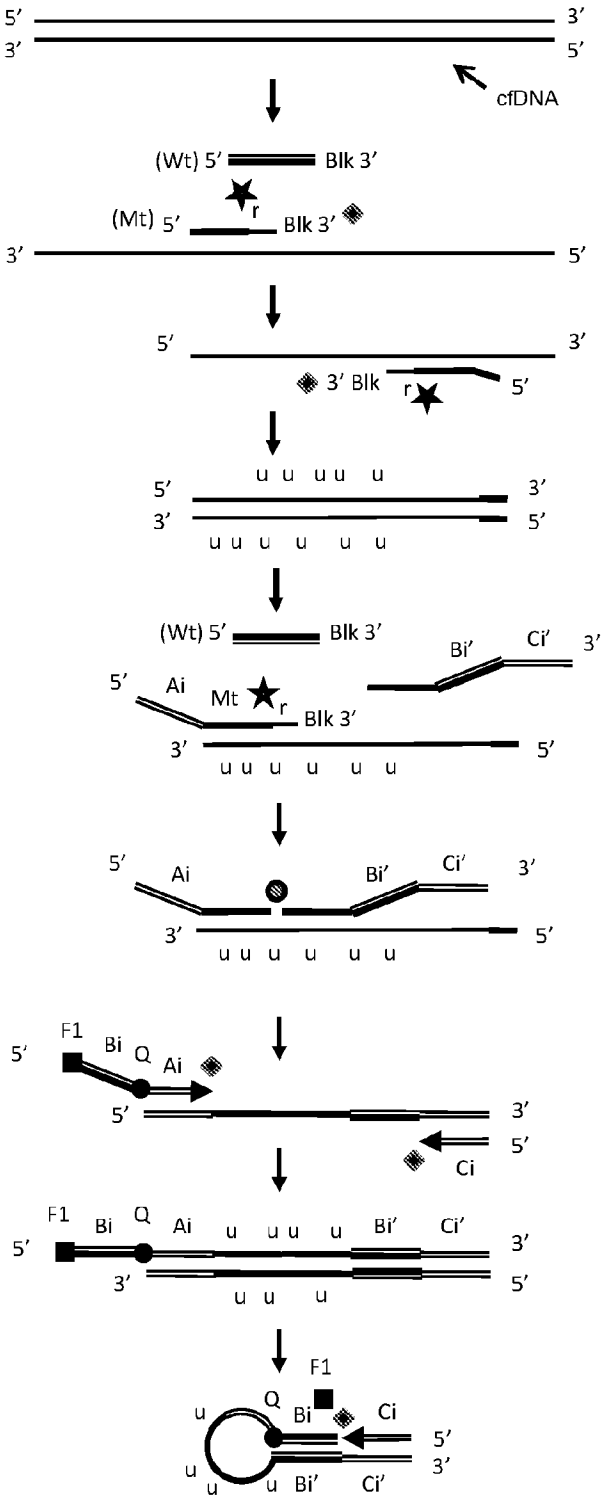


Figure 3

A. exPCR-qPCR carryover prevention reaction to detect low-level mutation. Isolate genomic or cfDNA. Treat with UDG for carryover prevention.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA Wt probes facilitate preferential amplification of mutation containing regions. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Extension products incorporate modified nucleotides (i.e. one or more αSdNTP's) that protect products but not cfDNA from Exonuclease I digestion.

D. Perform limited cycle nested PCR using mutation-specific and locus-specific primers with Universal tag tails. Blocking LNA or PNA Wt probes enables amplification of mutant but not Wt allele. Primers are unblocked with RNaseH2 only when bound to correct target.

E. PCR products comprise of Ai tag sequence, mutation-specific sequence, and Ci' tag sequence. Distribute products into wells, micro-pores, or droplets for Taqman reactions.

F. Amplify authentic mutation-specific products with Tag-specific primers (Ai and Ci). Each Taqman probe spans a unique mutation-specific sequence.

G. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

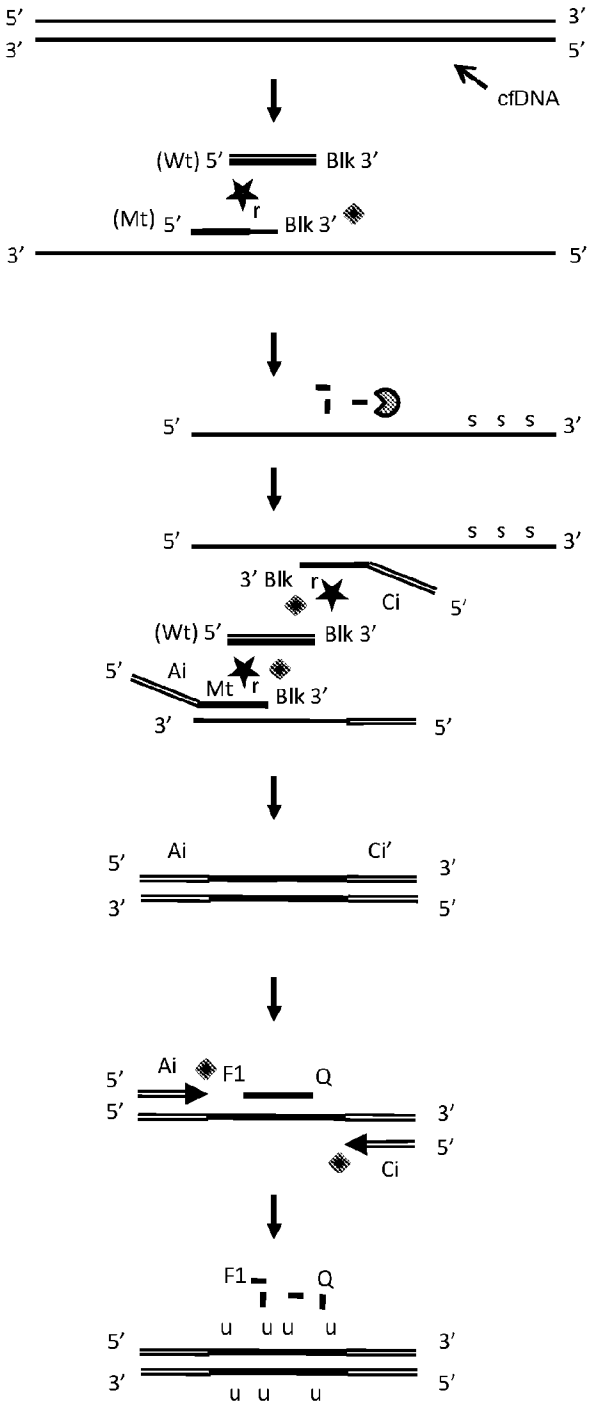


Figure 4

A. exPCR-qPCR carryover prevention reaction to detect low-level mutation. Isolate genomic or cfDNA. Treat with UDG for carryover prevention.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA Wt probes facilitate preferential amplification of mutation containing regions. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Extension products incorporate modified nucleotides (i.e. one or more αSdNTP's) that protect products but not cfDNA from Exonuclease I digestion.

D. Perform limited cycle nested PCR using mutation-specific (Mt) and locus-specific primers with UniTaq tails. Blocking LNA or PNA Wt probes enables amplification of mutant but not Wt allele. Primers are unblocked with RNaseH2 only when bound to correct target.

E. PCR products comprise of Ai tag sequence, mutation-specific sequence, Bi', and Ci' tag sequence. Distribute products into wells, micro-pores, or droplets for Taqman reactions.

F. Amplify authentic mutation-specific products with Tag-specific primers (F1-Bi-Q-Ai and Ci).

G. Double-stranded DNA products of PCR; the polymerase blocker stops extension of the bottom strand.

H. After the denaturation step, as the temperature decreases, a hairpin forms between Bi and Bi'. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

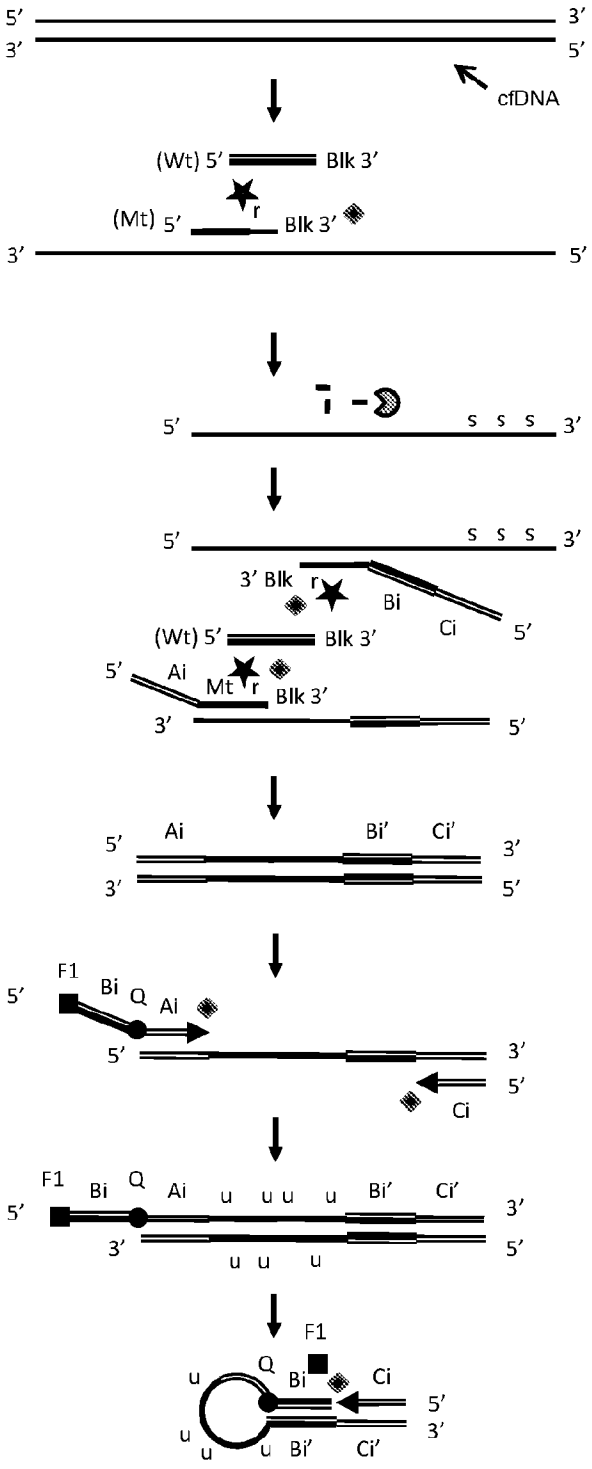


Figure 5

A. exPCR-qPCR carryover prevention reaction to detect low-level mutation. Isolate genomic or cfDNA. Treat with UDG for carryover prevention.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA Wt probes facilitate preferential amplification of mutation containing regions. Extension products incorporate modified nucleotides (i.e. one or more αSdNTP's) that protect products but not cfDNA from Exonuclease I digestion. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Add downstream primers and perform limited cycle PCR.

D. Distribute products into wells, micro-pores, or droplets for Taqman reactions using mutation-specific and locus-specific primers. Blocking LNA or PNA Wt probes enables amplification of mutant but not Wt allele. Primers are unblocked with RNaseH2 only when bound to correct target.

E. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

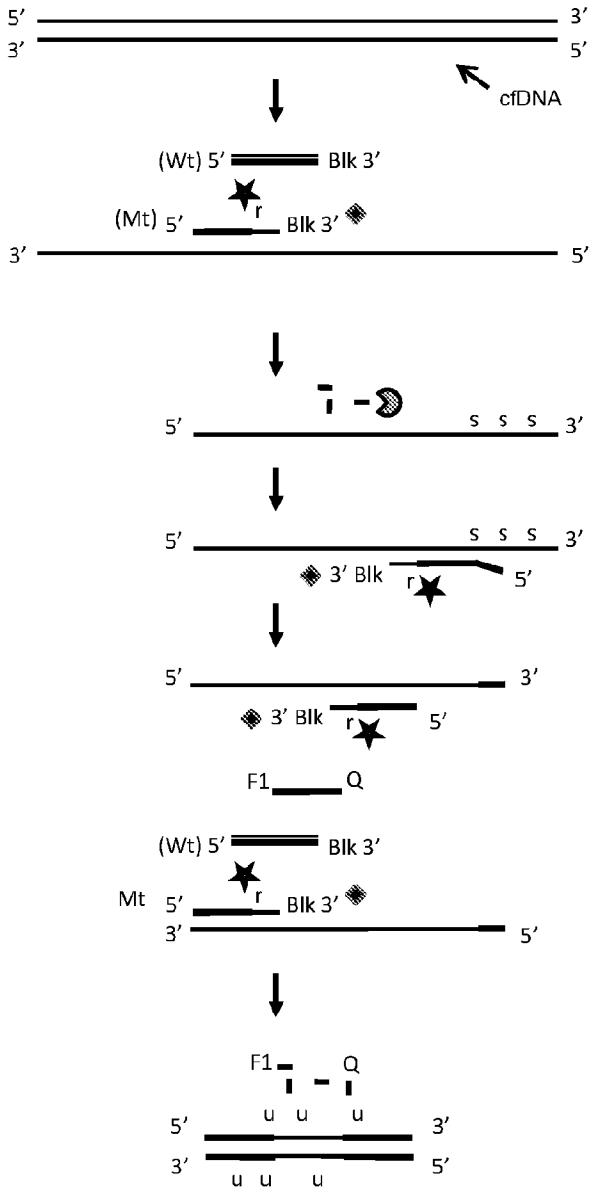


Figure 6

A. exPCR-qPCR carryover prevention reaction to detect low-level mutation. Isolate genomic or cfDNA. Treat with UDG for carryover prevention.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA Wt probes facilitate preferential amplification of mutation containing regions. Extension products incorporate modified nucleotides (i.e. one or more αSdNTP's) that protect products but not cfDNA from Exonuclease I digestion. Add downstream primers and perform limited cycle PCR. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Distribute products into wells, micro-pores, or droplets for Taqman reactions using mutation-specific and locus-specific primers with Universal tag tails, including tag-specific primers Ai and Ci. Blocking LNA or PNA Wt probes enables amplification of mutant but not Wt allele. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, mutation-specific sequence, and Ci' tag sequence.

E. Tag-specific primers (Ai and Ci) then amplify authentic mutation-specific products. Each Taqman probe spans a unique mutation-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

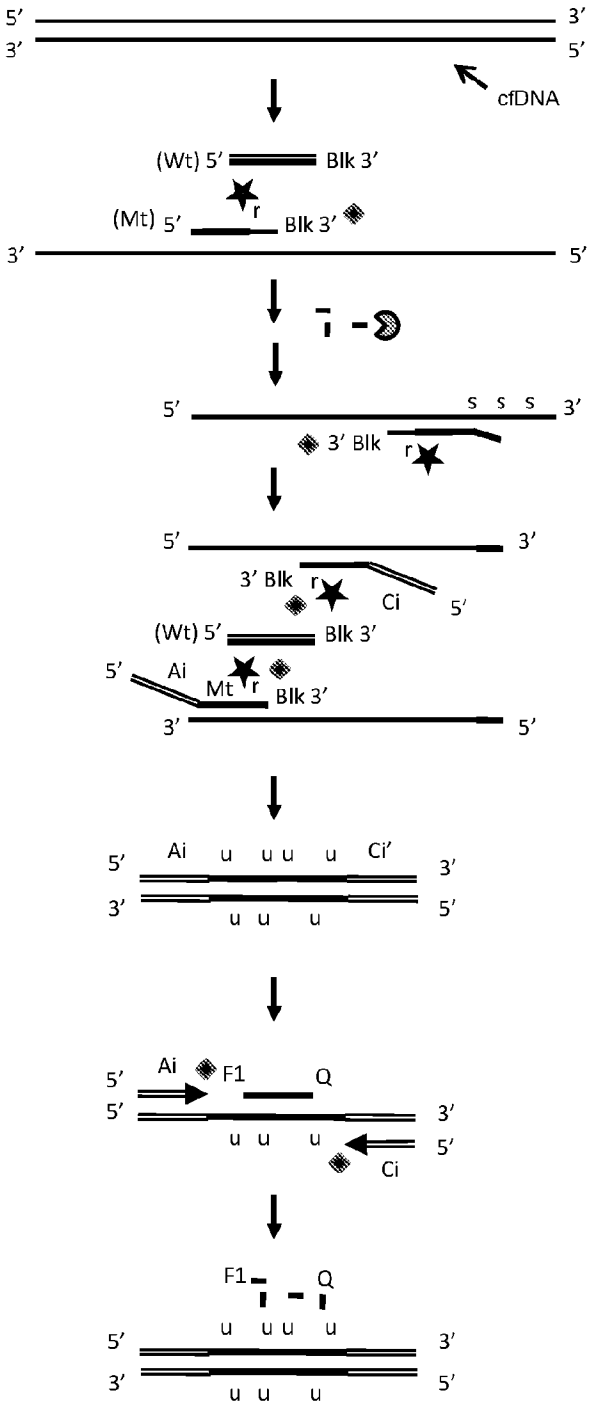


Figure 7

A. exPCR-qPCR carryover prevention reaction to detect low-level mutation. Isolate genomic or cfDNA. Treat with UDG for carryover prevention.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA Wt probes facilitate preferential amplification of mutation containing regions. Extension products incorporate modified nucleotides (i.e. one or more αSdNTP's) that protect products but not cfDNA from Exonuclease I digestion. Add downstream primers and perform limited cycle PCR. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Distribute products into wells, micro-pores, or droplets for Taqman reactions using mutation-specific and locus-specific primers with UniTaq tails, including tag-specific primers F1-Bi-Q-Ai and Ci. Blocking LNA or PNA Wt probes enables amplification of mutant but not Wt allele. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, mutation-specific sequence, Bi', and Ci' tag sequence.

E. Tag-specific primers (F1-Bi-Q-Ai and Ci) then amplify authentic mutation-specific products.

F. Double-stranded DNA products of PCR; the polymerase blocker stops extension of the bottom strand.

G. After the denaturation step, as the temperature decreases, a hairpin forms between Bi and Bi'. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

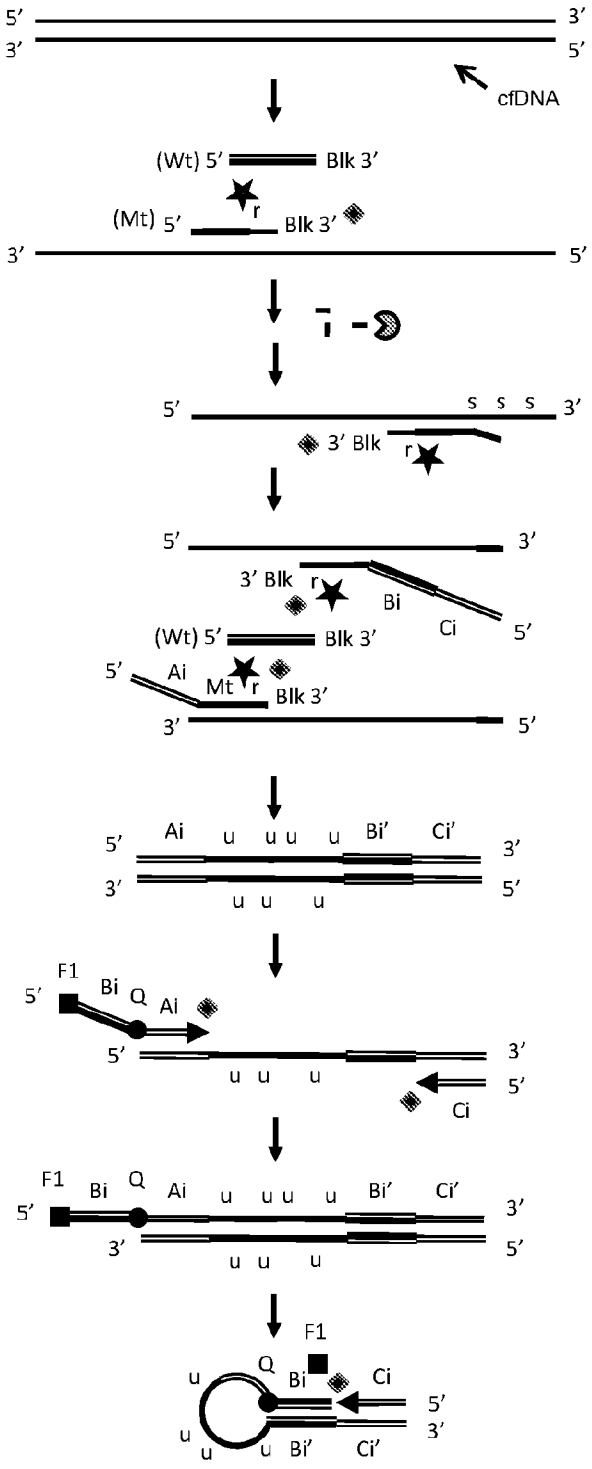


Figure 8

A. exPCR-LDR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Optional, treat with a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers that contain identical 8-11 base tails and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Add upstream primers, which also contain ribose bases (r), and perform PCR (using dUTP). Blocking LNA or PNA probes may be used to limit amplification of originally un-methylated DNA.

D. PCR products incorporate dU, allowing for carryover prevention.

E. Methyl region-specific ligation oligonucleotides (m) contain tags (Ai, Ci') for subsequent PCR amplification. Blocking LNA or PNA probes suppress ligation to originally un-methylated DNA sequence. Upstream ligation primer contains block, which may be removed by cleaving ribose base with RNaseH2 only when hybridized to target.

F. Ligase covalently seals the unblocked methyl region-specific upstream and phosphorylated downstream oligonucleotides together. Aliquot into separate wells, micro-pores, or droplets for detection.

G. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci'). Each Taqman probe spans the ligation junction.

H. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

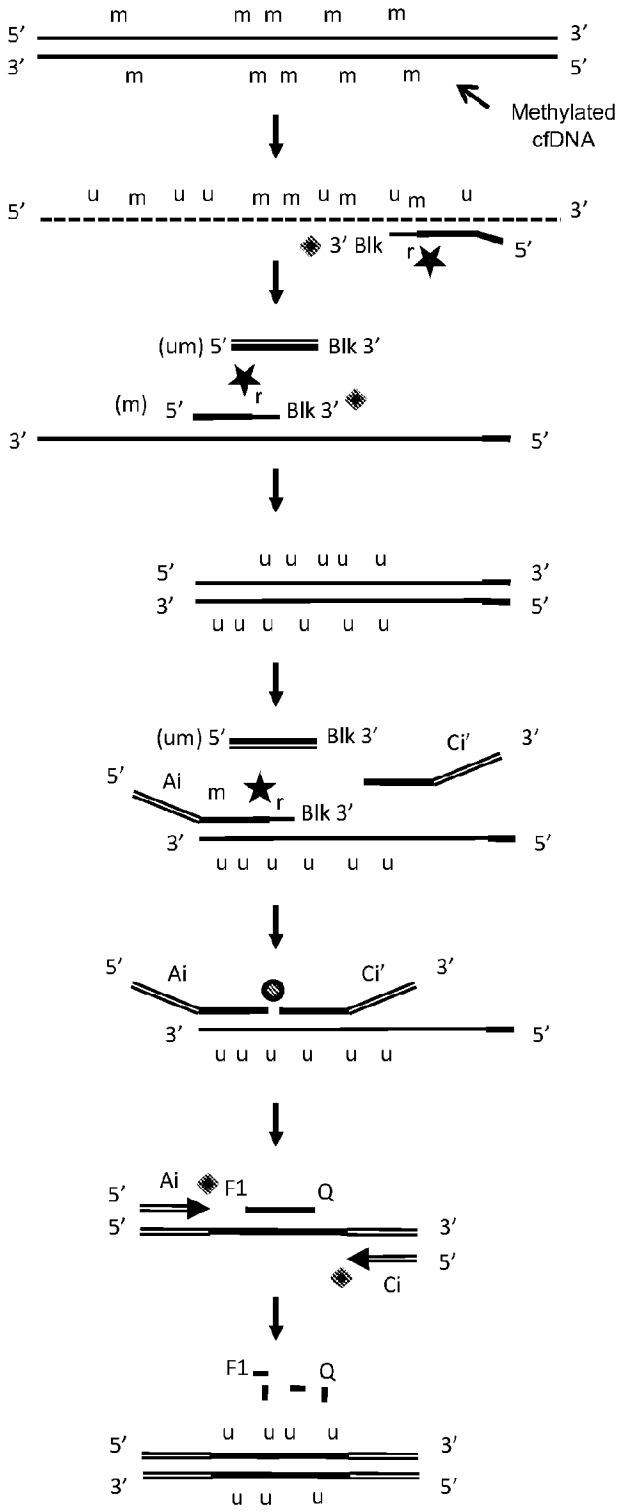


Figure 9

A. exPCR-LDR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Optional, treat with a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA probes may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Add downstream primers, (contain identical 8-11 base tails to prevent primer dimers) and perform PCR (and dUTP).

D. PCR products incorporate dU, allowing for carryover prevention.

E. Methyl region-specific ligation oligonucleotides (m) contain tags (Ai, Ci') for subsequent PCR amplification. Blocking LNA or PNA probes suppress ligation to originally un-methylated DNA sequence. Upstream ligation primer contains block, which may be removed by cleaving ribose base with RNaseH2 only when hybridized to target.

F. Ligase covalently seals the unblocked methyl region-specific upstream and phosphorylated downstream oligonucleotides together. Aliquot into separate wells, micro-pores, or droplets for detection.

G. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci'). Each Taqman probe spans the ligation junction.

H. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

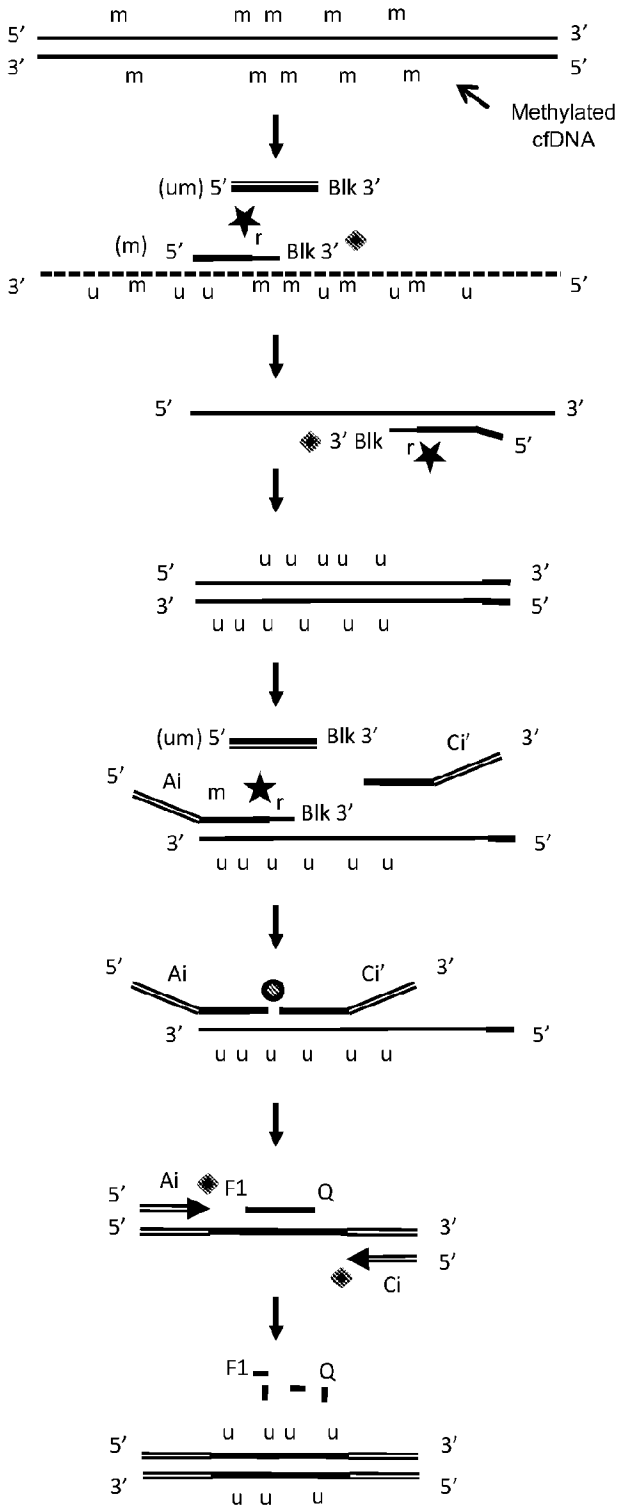


Figure 10

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Optional, treat with a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers that contain identical 8-11 base tails and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Perform limited cycle nested PCR using methylation-specific (m) and locus-specific primers with Universal tag tails. Blocking LNA or PNA Wt probes enables amplification of originally methylated but not originally un-methylated allele. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence. Distribute products into wells, micro-pores, or droplets for Taqman reactions.

E. Amplify authentic methylation-specific products with Tag-specific primers (Ai and Ci). Each Taqman probe spans a unique methylation-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

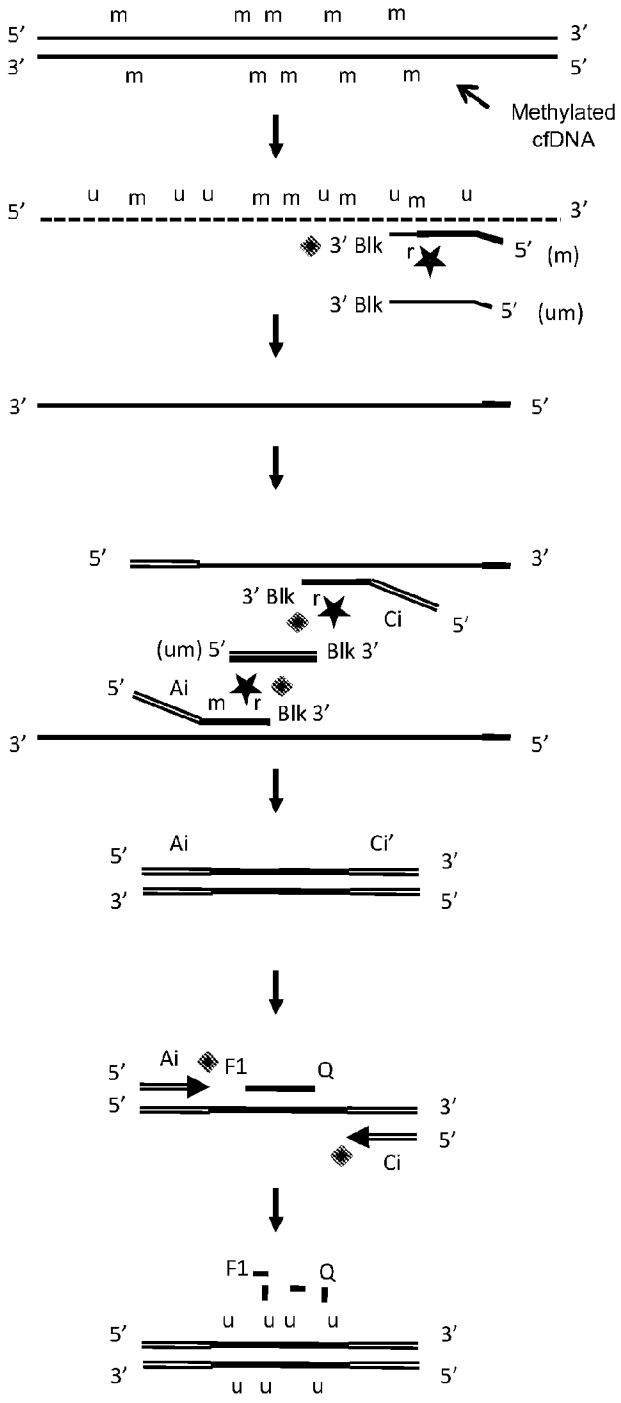


Figure 11

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Optional, treat with a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA probes may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Perform limited cycle nested PCR using methylation-specific (m) and locus-specific primers with Universal tag tails. Blocking LNA or PNA Wt probes enables amplification of originally methylated but not originally un-methylated allele. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence. Distribute products into wells, micro-pores, or droplets for Taqman reactions.

E. Amplify authentic methylation-specific products with Tag-specific primers (Ai and Ci). Each Taqman probe spans a unique methylation-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

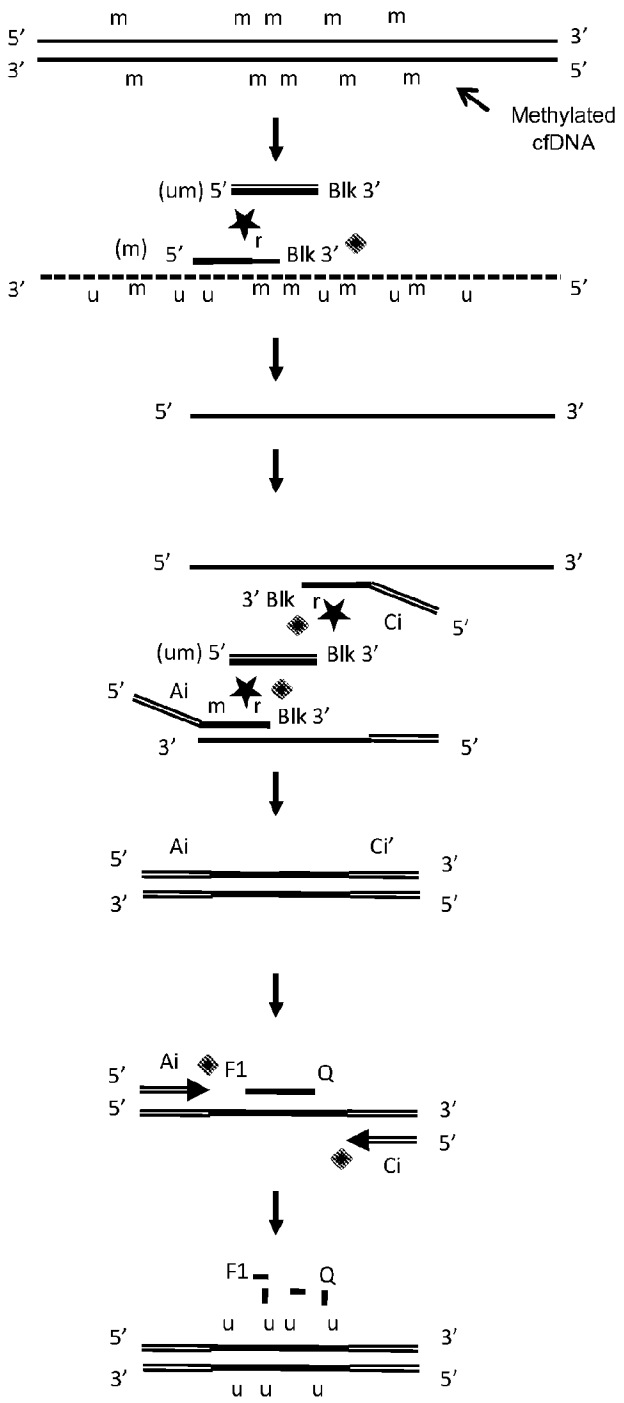


Figure 12

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Optional, treat with a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers that contain identical 8-11 base tails and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Add upstream primers, which also contain ribose bases (r), and perform PCR. Blocking LNA or PNA probes may be used to limit amplification of originally un-methylated DNA.

D. Distribute products into wells, micro-pores, or droplets for Taqman reactions using methylation-specific and locus-specific primers. Blocking LNA or PNA Wt probes enables amplification of originally methylated but not originally un-methylated allele. Primers are unblocked with RNaseH2 only when bound to correct target.

E. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

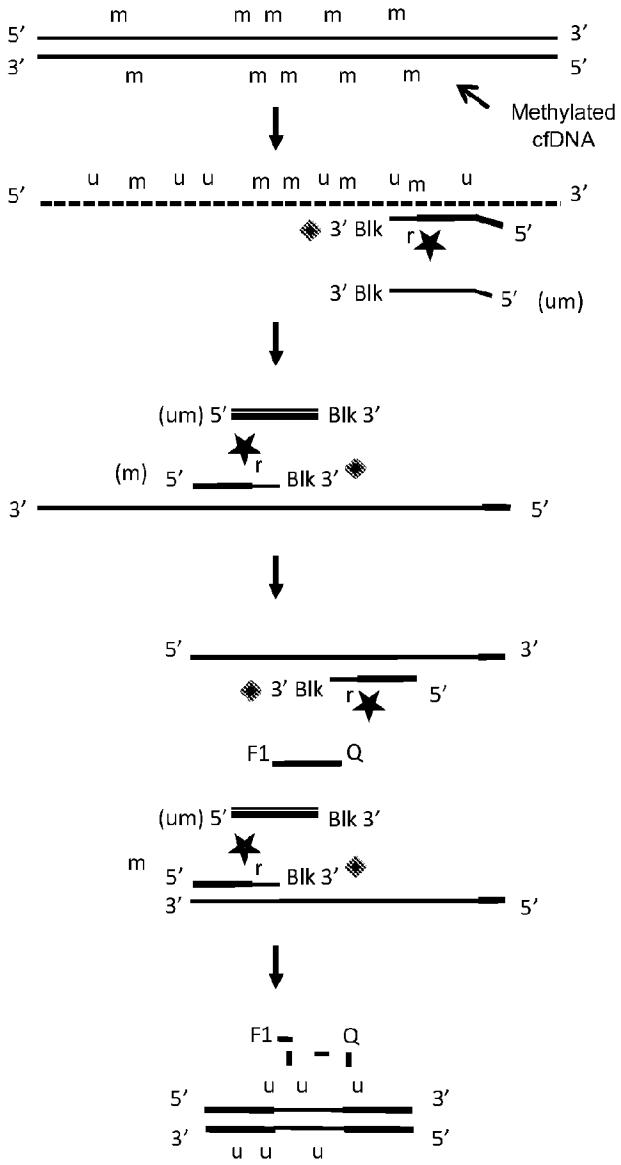


Figure 13

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Optional, treat with a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers that contain identical 8-11 base tails and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Add upstream primers, which also contain ribose bases (r), and perform PCR. Blocking LNA or PNA probes may be used to limit amplification of originally un-methylated DNA.

D. Distribute products into wells, micro-pores, or droplets for Taqman reactions using methylation-specific and locus-specific primers with Universal tag tails, including tag-specific primers Ai and Ci. Blocking LNA or PNA Wt probes enables amplification of originally methylated but not originally un-methylated allele. Primers are unblocked with RNaseH2 only when bound to correct target.

E. PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence.

F. Tag-specific primers (Ai and Ci) then amplify authentic methylation-specific products. Each Taqman probe spans a unique methylation-specific sequence.

G. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

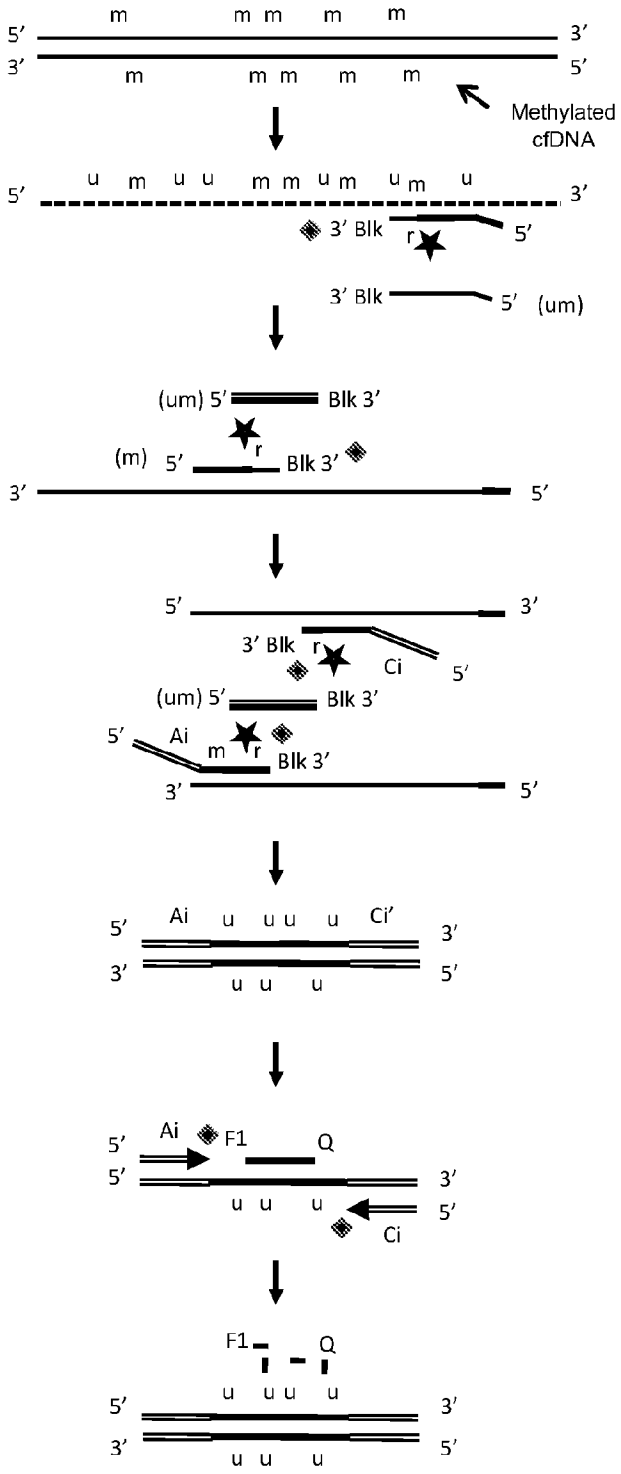


Figure 14

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Optional, treat with a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extension with primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA probes may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Add downstream primers, which also contain ribose bases (r), and perform limited cycle PCR. Blocking primers may be used to limit amplification of originally un-methylated DNA.

D. Distribute products into wells, micro-pores, or droplets for Taqman reactions using methylation-specific and locus-specific primers with Universal tag tails, including tag-specific primers Ai and Ci. Blocking LNA or PNA Wt probes enables amplification of originally methylated but not originally un-methylated allele. Primers are unblocked with RNaseH2 only when bound to correct target.

E. PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence.

F. Tag-specific primers (Ai and Ci) then amplify authentic methylation-specific products. Each Taqman probe spans a unique methylation-specific sequence.

G. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

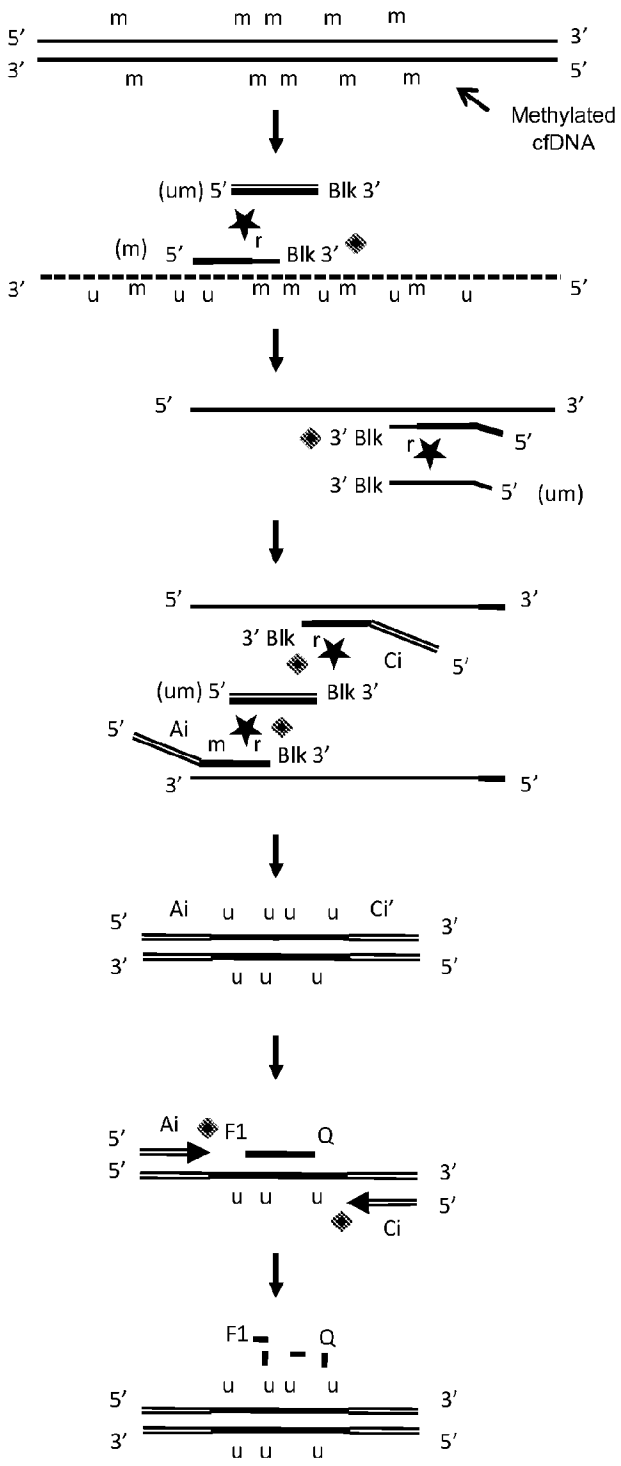


Figure 15

A. exPCR-LDR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Treat with (i) Bsh1236I (CG^CG) to completely digest un-methylated DNA, or (ii) capture and enrich methylated DNA, followed by a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers that contain identical 8-11 base tails and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Add upstream primers, which also contain ribose bases (r), and perform PCR (using dUTP). Blocking primers may be used to limit amplification of originally un-methylated DNA.

D. PCR products incorporate dU, allowing for carryover prevention.

E. Methyl region-specific ligation oligonucleotides (m) contain tags (Ai, Ci') for subsequent PCR amplification. Upstream ligation primer contains block, which may be removed by cleaving ribose base with RNaseH2 only when hybridized to target.

F. Ligase covalently seals the unblocked methyl region-specific upstream and phosphorylated downstream oligonucleotides together. Aliquot into separate wells, micro-pores, or droplets for detection.

G. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci'). Each Taqman probe spans the ligation junction.

H. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

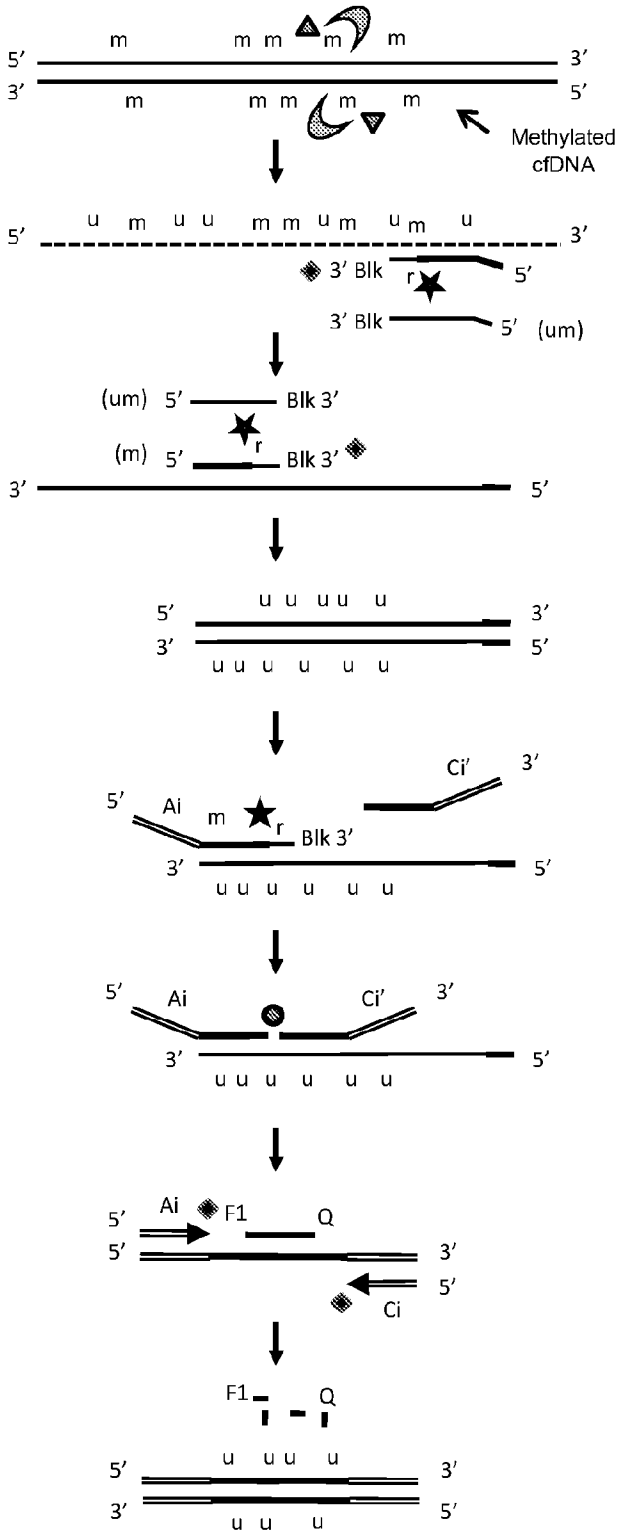


Figure 16

A. exPCR-LDR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Treat with (i) Bsh1236I (CG^CG) to completely digest un-methylated DNA, or (ii) capture and enrich methylated DNA, followed by a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Add downstream primers, (contain identical 8-11 base tails to prevent primer dimers) and perform PCR (and dUTP).

D. PCR products incorporate dU, allowing for carryover prevention.

E. Methyl region-specific ligation oligonucleotides (m) contain tags (Ai, Ci') for subsequent PCR amplification. Upstream ligation primer contains block, which may be removed by cleaving ribose base with RNaseH2 only when hybridized to target.

F. Ligase covalently seals the unblocked methyl region-specific upstream and phosphorylated downstream oligonucleotides together. Aliquot into separate wells, micro-pores, or droplets for detection.

G. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci'). Each Taqman probe spans the ligation junction.

H. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

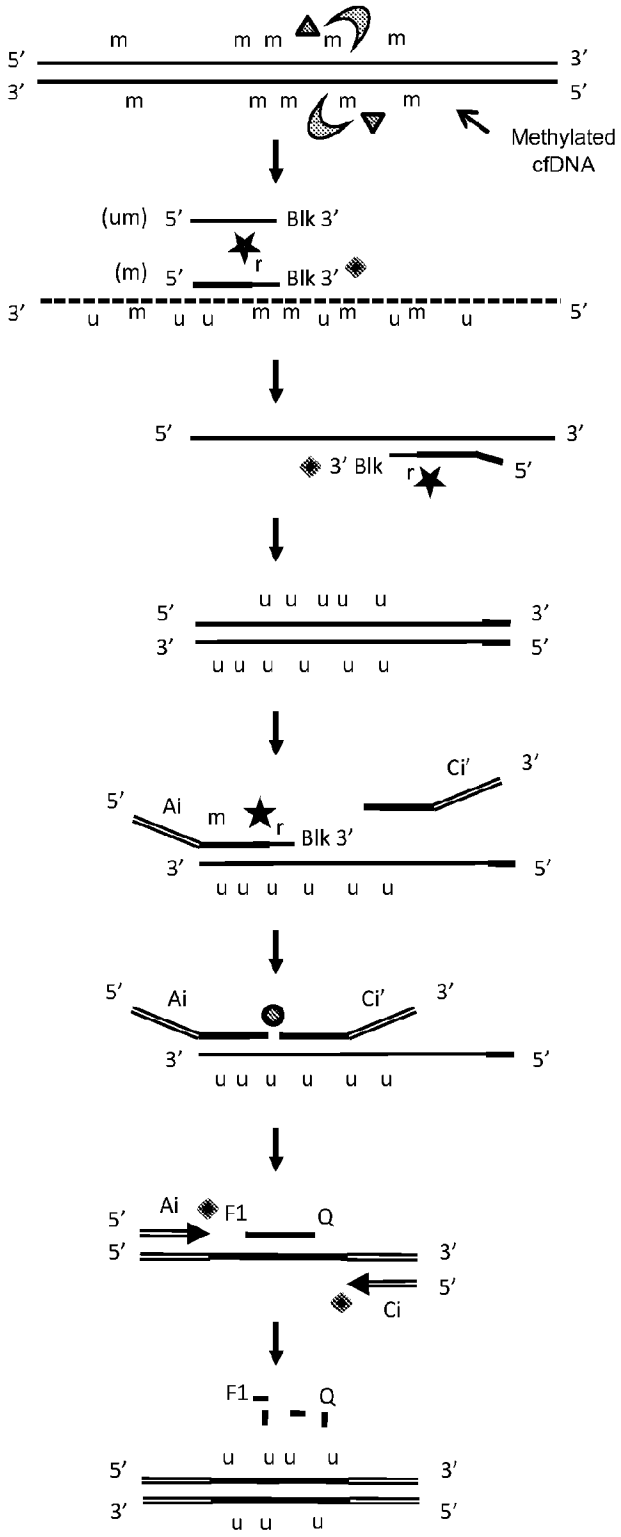


Figure 17

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Treat with (i) Bsh1236I (CG^CG) to completely digest un-methylated DNA, or (ii) capture and enrich methylated DNA, followed by a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers that contain identical 8-11 base tails and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Perform limited cycle nested PCR using methylation-specific (m) and locus-specific primers with Universal tag tails. Primers are unblocked with RNaseH2 only when bound to correct target. Blocking primers may be used to limit amplification of originally un-methylated DNA.

D. PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence. Distribute products into wells, micro-pores, or droplets for Taqman reactions.

E. Amplify authentic methylation-specific products with Tag-specific primers (Ai and Ci). Each Taqman probe spans a unique methylation-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

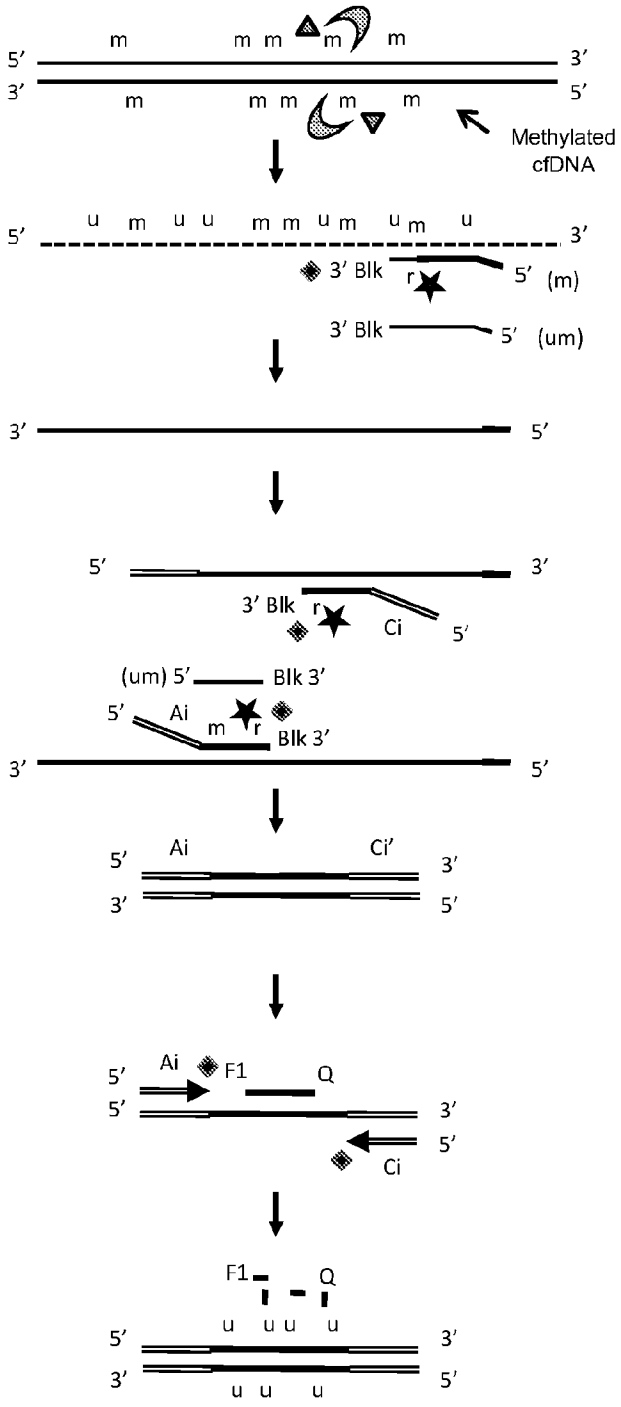


Figure 18

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Treat with (i) Bsh1236I (CG^CG) to completely digest un-methylated DNA, or (ii) capture and enrich methylated DNA, followed by a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with upstream primers that contain ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking LNA or PNA probes may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to extension.)

C. Perform limited cycle nested PCR using methylation-specific (m) and locus-specific primers with Universal tag tails. Blocking LNA or PNA Wt probes enables amplification of originally methylated but not originally un-methylated allele. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence. Distribute products into wells, micro-pores, or droplets for Taqman reactions.

E. Amplify authentic methylation-specific products with Tag-specific primers (Ai and Ci). Each Taqman probe spans a unique methylation-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

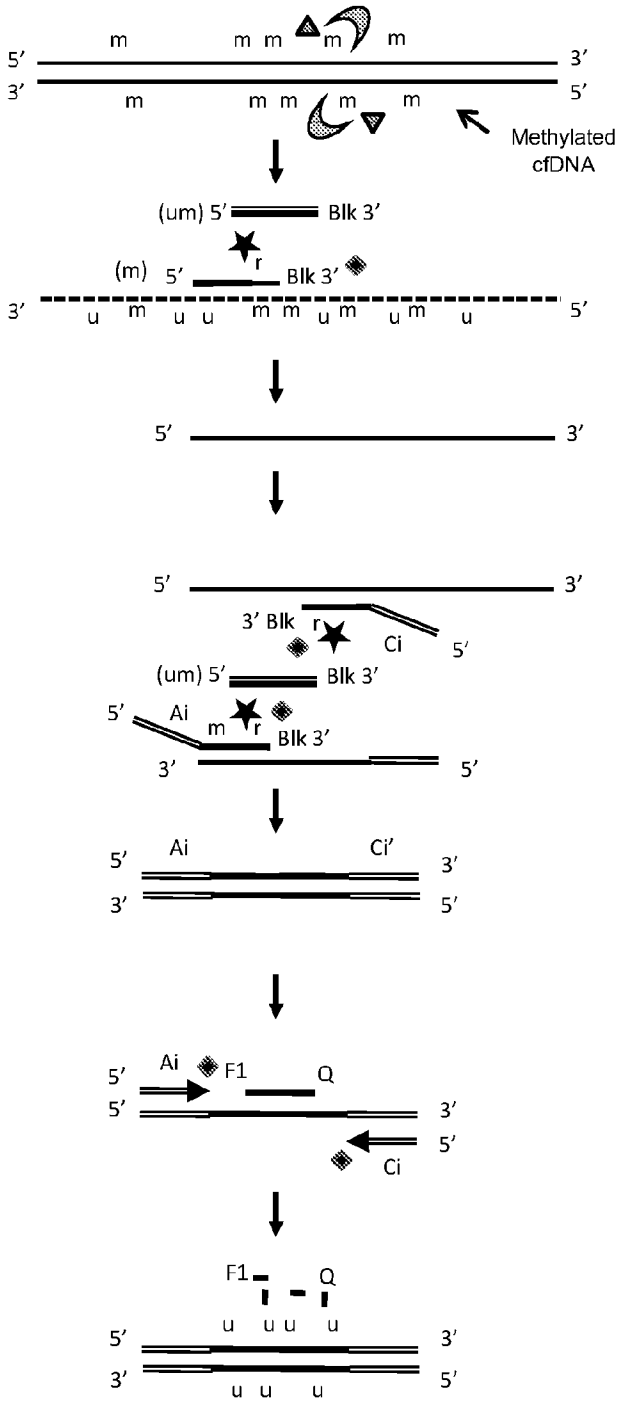


Figure 19

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Treat with (i) Bsh1236I (CG^CG) to completely digest un-methylated DNA, or (ii) capture and enrich methylated DNA, followed by a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers that contain identical 8-11 base tails and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Add upstream primers, which also contain ribose bases (r), and perform PCR. Blocking primer may be used to limit amplification of originally un-methylated DNA.

D. Distribute products into wells, micro-pores, or droplets for Taqman reactions using methylation-specific and locus-specific primers. Primers are unblocked with RNaseH2 only when bound to correct target.

E. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

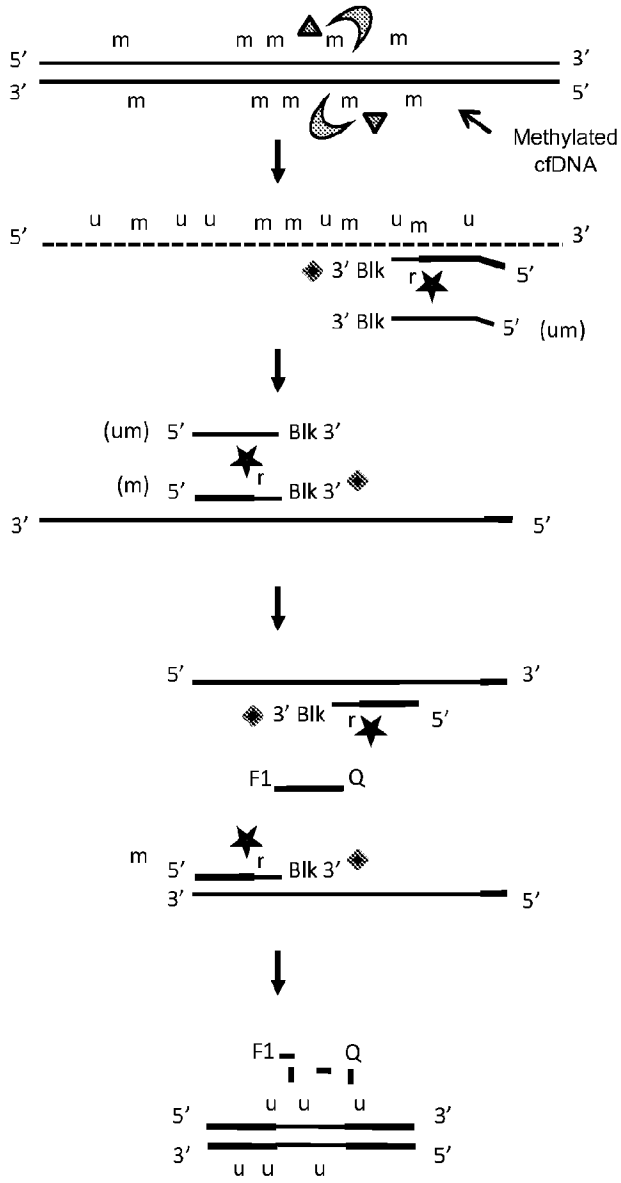


Figure 20

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Treat with (i) Bsh1236I (CG^CG) to completely digest un-methylated DNA, or (ii) capture and enrich methylated DNA, followed by a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers that contain identical 8-11 base tails and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Add upstream primers, which also contain ribose bases (r), and perform PCR. Blocking primer may be used to limit amplification of originally un-methylated DNA.

D. Distribute products into wells, micro-pores, or droplets for Taqman reactions using methylation-specific and locus-specific primers with Universal tag tails, including tag-specific primers Ai and Ci. Primers are unblocked with RNaseH2 only when bound to correct target.

E. PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence.

F. Tag-specific primers (Ai and Ci) then amplify authentic methylation-specific products. Each Taqman probe spans a unique methylation-specific sequence.

G. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

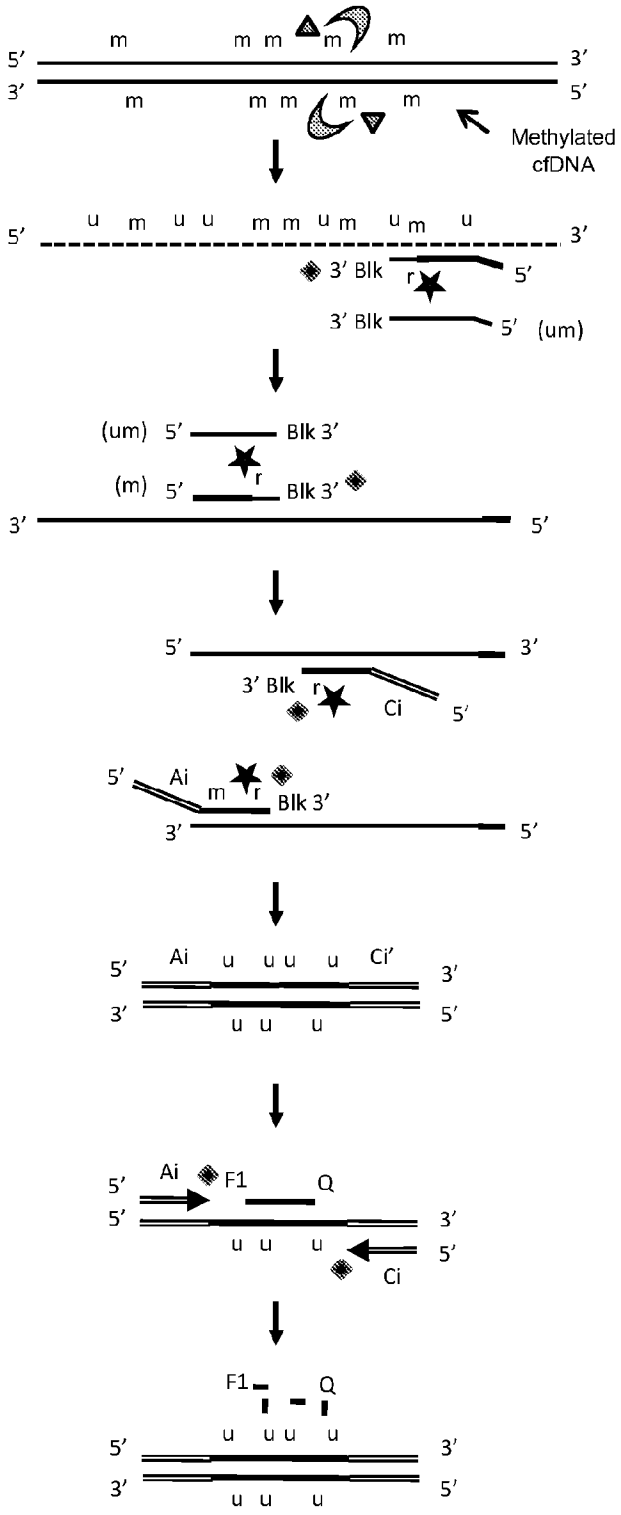


Figure 21

A. exPCR-qPCR carryover prevention reaction to detect low-level methylation. Isolate genomic or cfDNA. Treat with (i) Bsh1236I (CG^CG) to completely digest un-methylated DNA, or (ii) capture and enrich methylated DNA, followed by a DNA repair kit. Treat with bisulfite, which renders the strands non-complementary.

B. Perform multiple rounds of one-sided locus primer extensions with downstream primers comprising 5' universal tails (Ci) and ribose bases (r), which are removed with RNaseH2 only when bound to target, thus unblocking primer. Blocking primers may be used to limit amplification of originally un-methylated DNA. Add UDG to destroy bisulfite converted DNA. (Optional: aliquot into 12, 24, 36, 48, or 96 wells prior to initial extension.)

C. Add upstream primers, comprising 5' universal tails (Ai) and ribose bases (r), and perform PCR. Blocking primer may be used to limit amplification of originally un-methylated DNA.

D. PCR products comprise of Ai tag sequence, methylation-specific sequence, and Ci' tag sequence. Distribute products into wells, micro-pores, or droplets for Taqman reactions.

E. Amplify authentic methylation-specific products with Tag-specific primers (Ai and Ci). Each Taqman probe spans a unique methylation-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. (PCR products incorporate dU, allowing for carryover prevention.)

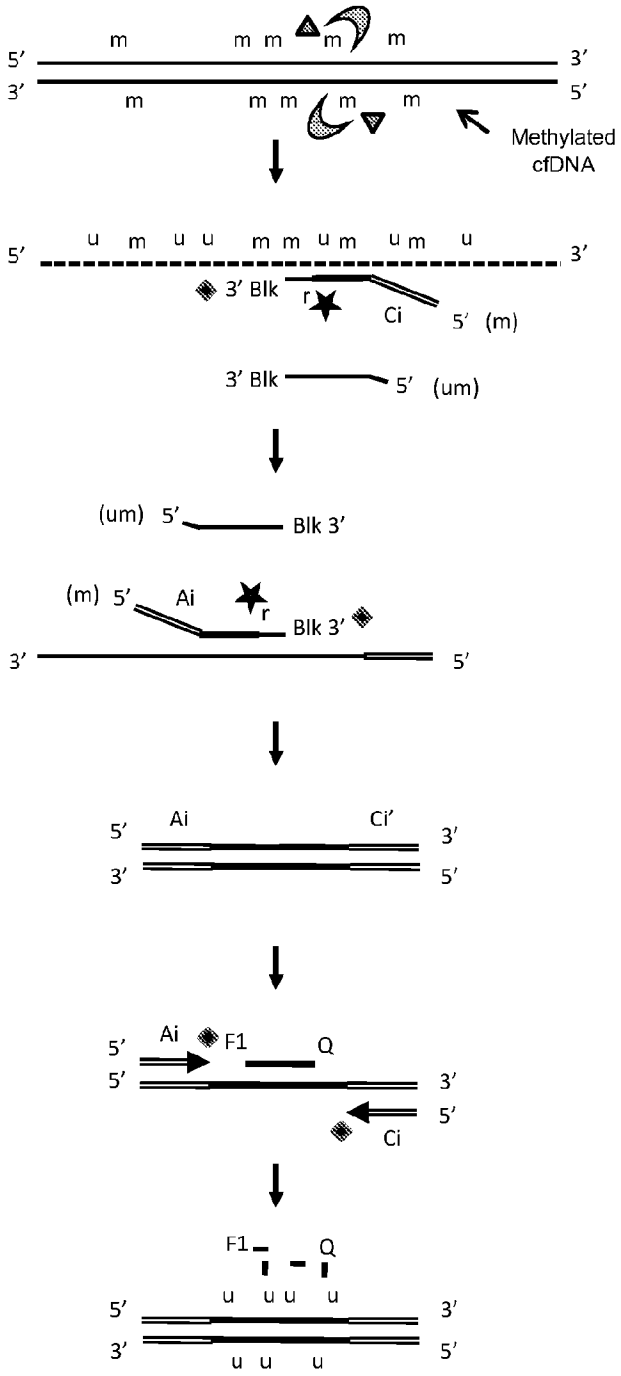


Figure 22

A. RT-PCR-LDR-qPCR carryover prevention reaction to detect and enumerate translocation at the mRNA level. Isolate mRNA.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. Use reverse-transcriptase to make cDNA copy (using dUTP) with 3' transcript-specific primers. Activate Taq polymerase and perform limited cycle PCR amplification with exon-specific primers (12-20) to maintain relative ratios of different amplicons. Primers contain identical 8-11 base tails to prevent primer dimers, and are added only to wells with the anticipated low-copy dilution.

C. PCR products incorporate dU, allowing for carryover prevention.

D. Exon junction-specific ligation oligonucleotides contain tags (Ai, Ci') for subsequent PCR amplification. Ligase covalently seals the two oligonucleotides together. Aliquot into separate wells for detection. (LDR products are not amplified by either original PCR or LDR primers, providing additional protection.)

E. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci). Each Taqman probe spans the ligation junction.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine copy number of fusion transcripts based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

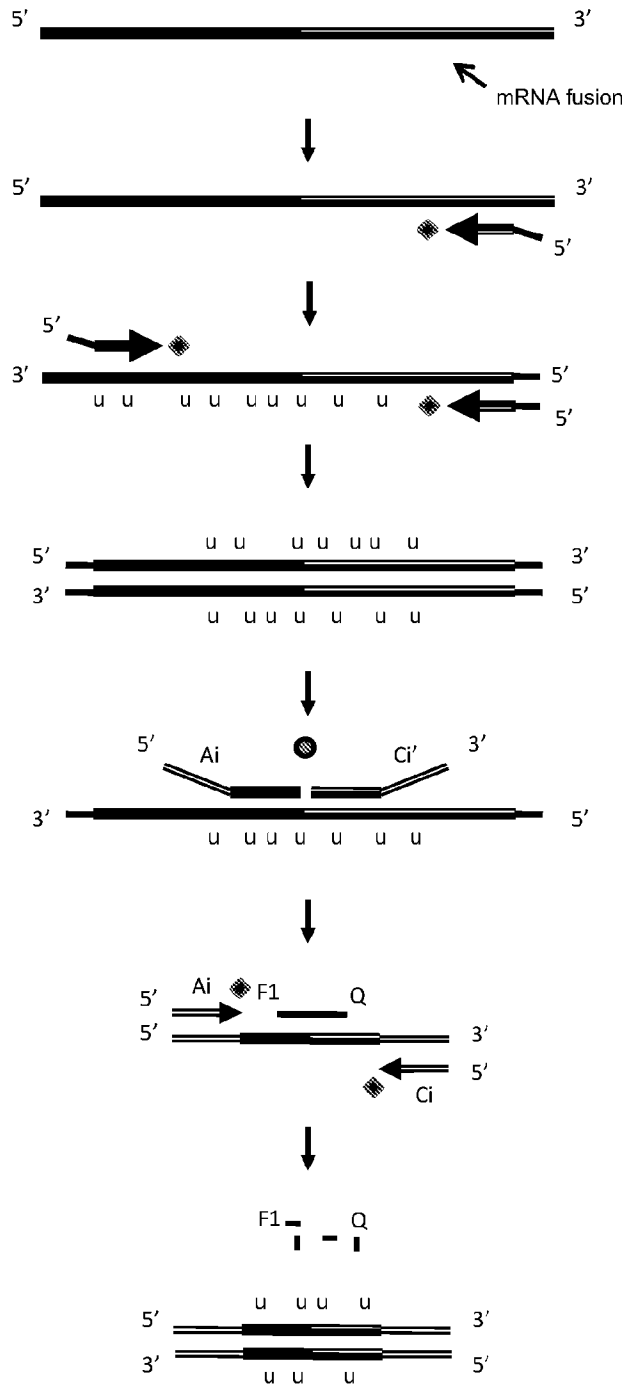


Figure 23

A. RT-PCR-qPCR carryover prevention reaction to detect and enumerate translocation at the mRNA level. Isolate mRNA.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. Use reverse-transcriptase to make cDNA copy with 3' transcript-specific primers. Activate Taq polymerase and perform limited cycle PCR amplification with exon-specific primers (8-12) to maintain relative ratios of different amplicons. Primers contain identical 8-11 base tails to prevent primer dimers, and are added only to wells with the anticipated low-copy dilution.

C. Distribute products into wells, micro-pores, or droplets for Taqman reactions using nested transcript-specific primers. Primers are unblocked with RNaseH2 only when bound to correct target.

D. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine copy number of fusion transcripts based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

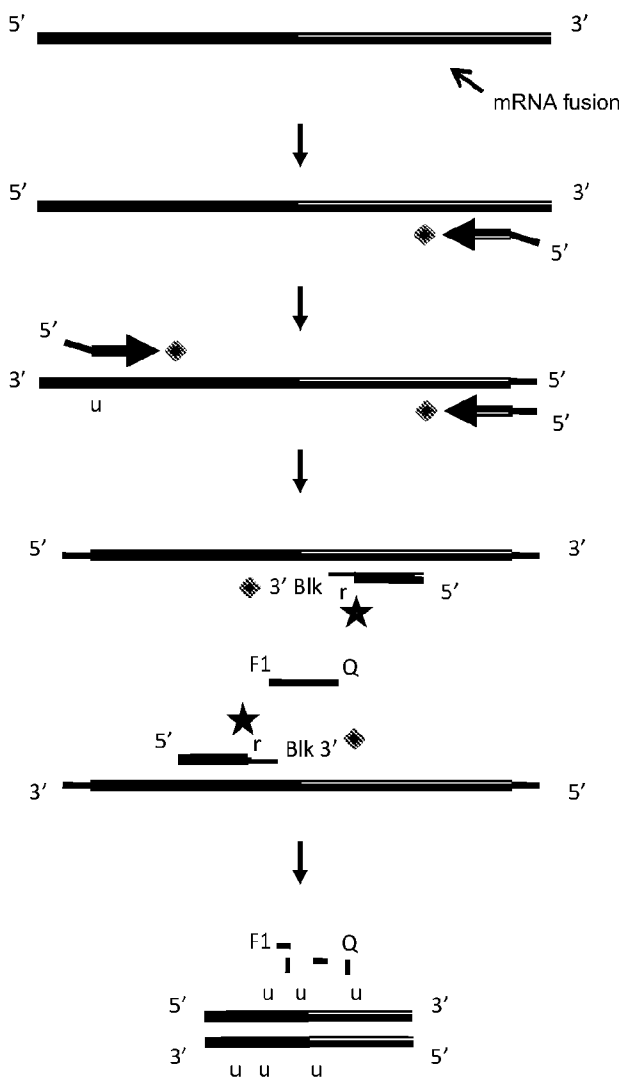


Figure 24

A. RT-PCR-PCR-qPCR carryover prevention reaction to detect and enumerate translocation at the mRNA level. Isolate mRNA.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. Use reverse-transcriptase to make cDNA copy with 3' transcript-specific primers. Activate Taq polymerase and perform limited cycle PCR amplification with exon-specific primers (8-12) to maintain relative ratios of different amplicons. Primers contain identical 8-11 base tails to prevent primer dimers, and are added only to wells with the anticipated low-copy dilution.

C. Distribute products into wells, micro-pores, or droplets for Taqman reactions using nested transcript-specific primers with Universal tag tails, including tag-specific primers Ai and Ci. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, transcript-specific sequence, and Ci' tag sequence.

E. Tag-specific primers (Ai and Ci) then amplify authentic transcript-specific products. Each Taqman probe spans a unique methylation-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine copy number of fusion transcripts based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

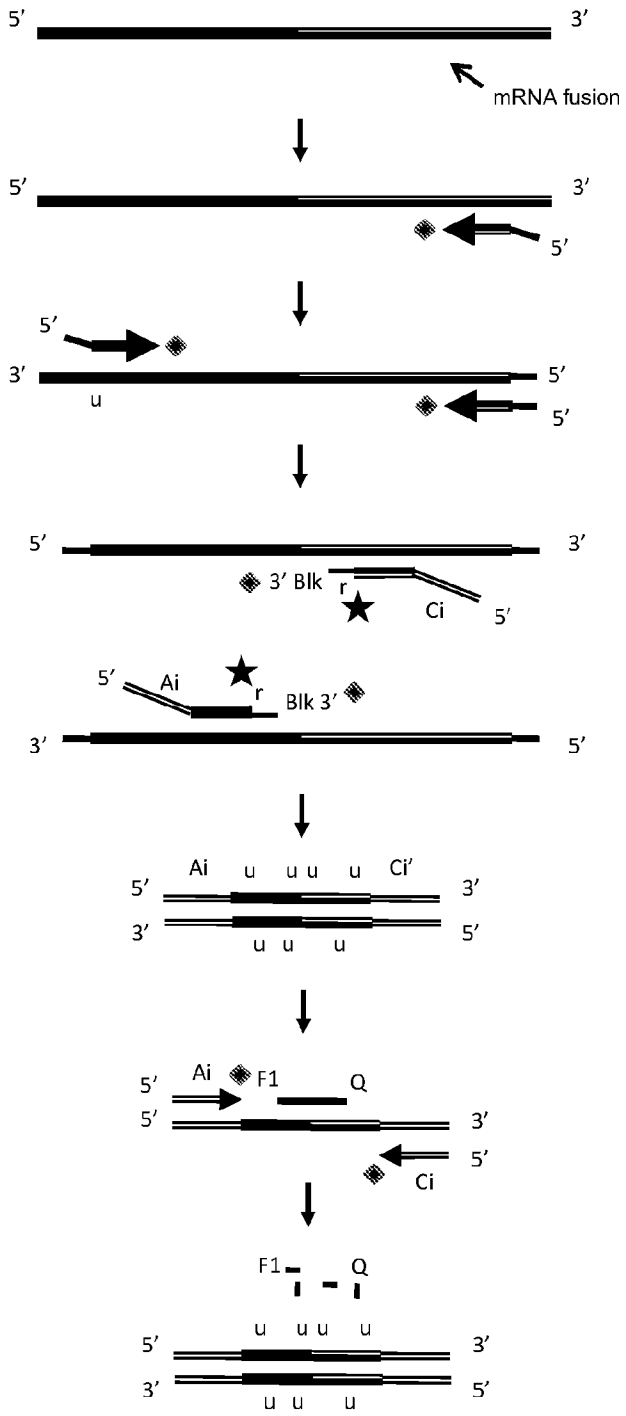


Figure 25

A. RT-PCR-LDR-qPCR carryover prevention reaction to enumerate RNA copy number. Isolate RNA from whole blood cells, exosomes, or CTCs.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. Use reverse-transcriptase to make cDNA copy (using dUTP) with 3′ transcript-specific primers. Activate Taq polymerase and perform limited cycle PCR amplification (12-20) to maintain relative ratios of different amplicons. Primers contain identical 8-11 base tails to prevent primer dimers, and are added only to wells with the anticipated low-copy dilution.

C. PCR products incorporate dU, allowing for carryover prevention.

D. Locus-specific ligation oligonucleotides contain tags (Ai, Ci′) for subsequent PCR amplification. Ligase covalently seals the two oligonucleotides together. Distribute into separate wells, micro-pores, or droplets for detection.

E. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci). Each Taqman probe spans the ligation junction.

F. The 5′->3′ nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine copy number based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

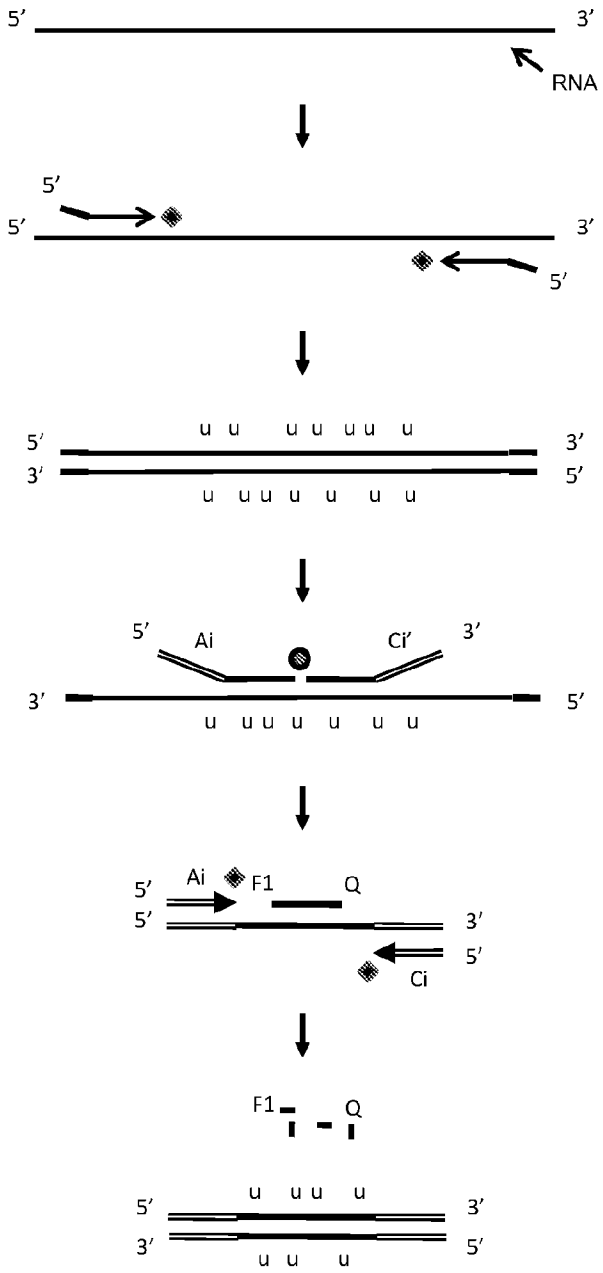


Figure 26

A. RT-PCR-qPCR carryover prevention reaction to enumerate RNA copy number. Isolate RNA from whole blood cells, exosomes, or CTCs.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. Use reverse-transcriptase to make cDNA copy with 3′ transcript-specific primers. Activate Taq polymerase and perform limited cycle PCR amplification (8-12) to maintain relative ratios of different amplicons. Primers contain identical 8-11 base tails to prevent primer dimers, and are added only to wells with the anticipated low-copy dilution.

C. Distribute products into wells, micro-pores, or droplets for Taqman reactions using nested transcript-specific primers. Primers are unblocked with RNaseH2 only when bound to correct target.

D. The 5′->3′ nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine copy number based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

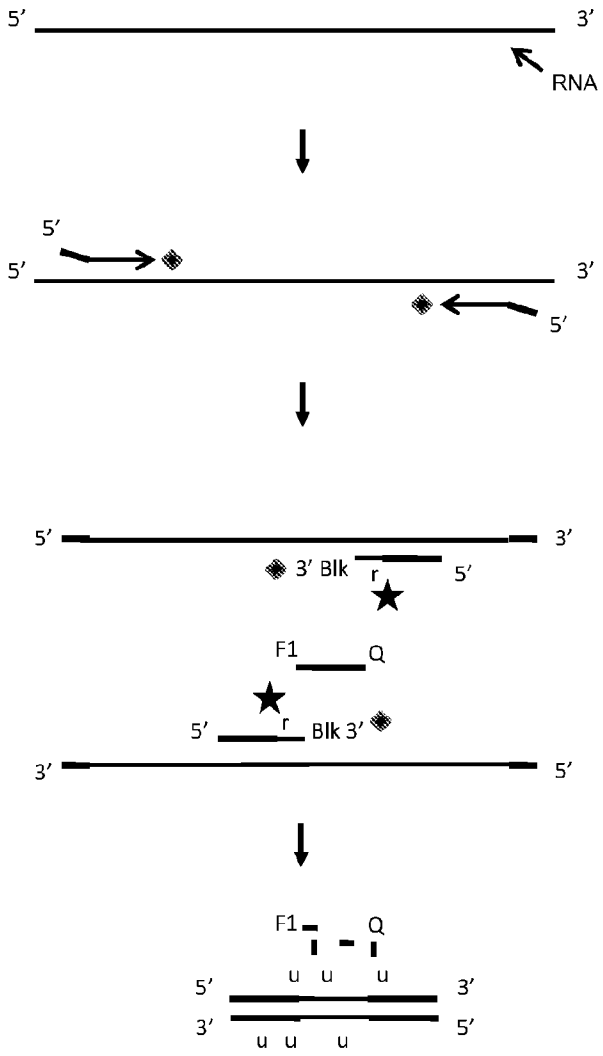


Figure 27

A. RT-PCR-PCR-qPCR carryover prevention reaction to enumerate RNA copy number. Isolate RNA from whole blood cells, exosomes, or CTCs.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. Use reverse-transcriptase to make cDNA copy with 3′ transcript-specific primers. Activate Taq polymerase and perform limited cycle PCR amplification (8-12) to maintain relative ratios of different amplicons. Primers contain identical 8-11 base tails to prevent primer dimers, and are added only to wells with the anticipated low-copy dilution.

C. Distribute products into wells, micro-pores, or droplets for Taqman reactions using nested transcript-specific primers with Universal tag tails, including tag-specific primers Ai and Ci. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, transcript-specific sequence, and Ci' tag sequence.

E. Tag-specific primers (Ai and Ci) then amplify authentic transcript-specific products. Each Taqman probe spans a unique methylation-specific sequence.

F. The 5′->3′ nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine copy number based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

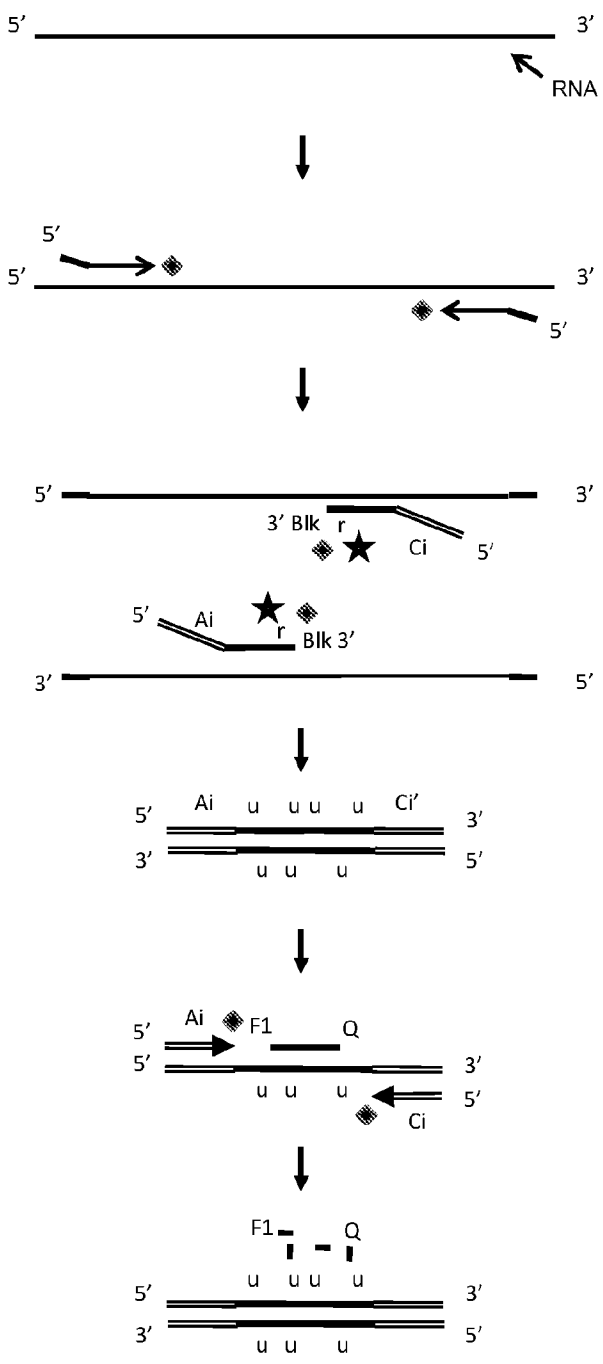


Figure 28

A. Ligation-RT-PCR-LDR-qPCR carryover prevention reaction to detect and enumerate miRNA. Isolate miRNA from exosomes.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. Ligate probe complementary to 3' end of target miRNA, containing a stem-loop, Tag (Di'), and blocking group. Extend with primer Di and reverse transcriptase, which appends C3 on 3' end. Hybridize Tag (Ei) with rGrG+G on 3' end (+G is symbol for LNA). Reverse transcriptase undergoes strand switching and copies Ei. Activate Taq polymerase and perform limited cycle PCR amplification (12-20), using dUTP, using tag primers (Di, Ei), to maintain relative ratios of different amplicons.

C. PCR products incorporate dU, allowing for carryover prevention.

D. miRNA-specific ligation oligonucleotides contain tags (Ai, Ci') for subsequent PCR amplification. Ligase covalently seals the two oligonucleotides together. Distribute into separate wells, micro-pores, or droplets for detection.

E. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci). Each Taqman probe spans the miRNA-ligation junction.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine miRNA copy number based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

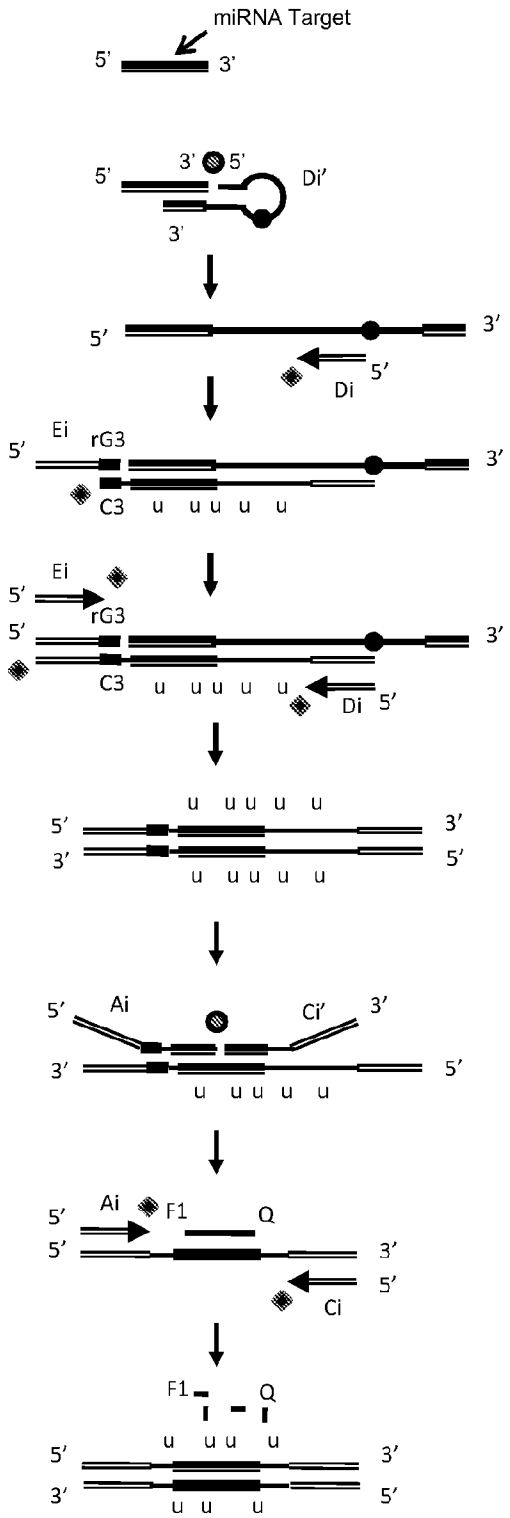


Figure 29

A. Ligation-RT-PCR-qPCR carryover prevention reaction to detect and enumerate miRNA. Isolate miRNA from exosomes.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. Ligate probe complementary to 3' end of target miRNA, containing a stem-loop, Tag (Di'), and blocking group. Extend with primer Di and reverse transcriptase, which appends C3 on 3' end. Hybridize Tag (Ei) with rGrG+G on 3' end (+G is symbol for LNA). Reverse transcriptase undergoes strand switching and copies Ei. Activate Taq polymerase and perform limited cycle PCR amplification (8-12), using tag primers (Di, Ei), to maintain relative ratios of different amplicons.

C. Distribute products into wells, micro-pores, or droplets for Taqman reactions using nested miRNA-specific primers with Universal tag tails, including tag-specific primers Ai and Ci. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, a portion of Ei, G3, miRNA-specific sequence, a portion of the stem-loop sequence, and Ci' tag sequence.

E. Tag-specific primers (Ai and Ci) then amplify authentic transcript-specific products. Each Taqman probe spans a unique miRNA-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine miRNA copy number based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

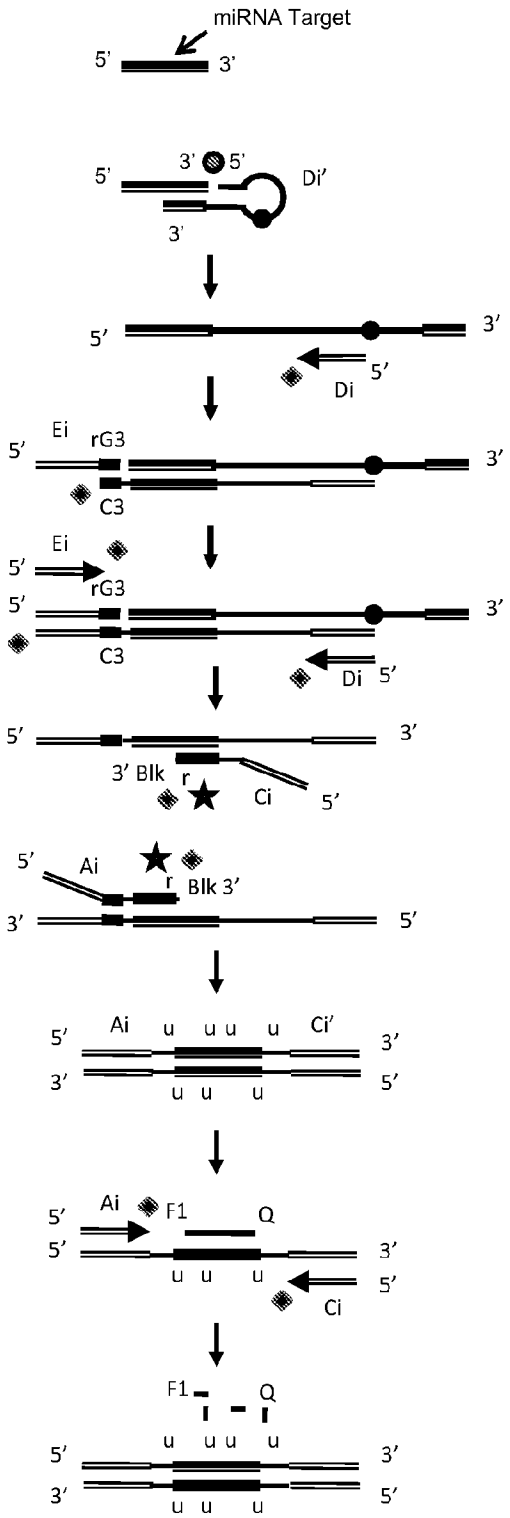


Figure 30

A. RT-PCR-LDR-qPCR carryover prevention reaction to detect and enumerate miRNA. Isolate miRNA from exosomes.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. PolyA tail miRNA with Poly(A) Polymerase. Extend with primer comprising a Tag Di on the 5' end, and a T30VN sequence at the 3' end, using reverse transcriptase, which appends C3 on 3' end. Hybridize Tag (Ei) with rGrG+G on 3' end (+G is symbol for LNA). Reverse transcriptase undergoes strand switching and copies Ei. Activate Taq polymerase and perform limited cycle PCR amplification (12-20), using dUTP, using tag primers (Di, Ei), to maintain relative ratios of different amplicons.

C. PCR products incorporate dU, allowing for carryover prevention.

D. miRNA-specific ligation oligonucleotides contain tags (Ai, Ci') for subsequent PCR amplification. Ligase covalently seals the two oligonucleotides together. Distribute into separate wells, micro-pores, or droplets for detection.

E. Treat with UDG for carryover prevention, which also destroys original target amplicons. Only authentic LDR products will amplify, when using PCR (and dUTP) with tag-specific primers (Ai, Ci). Each Taqman probe spans the miRNA-ligation junction.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine miRNA copy number based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

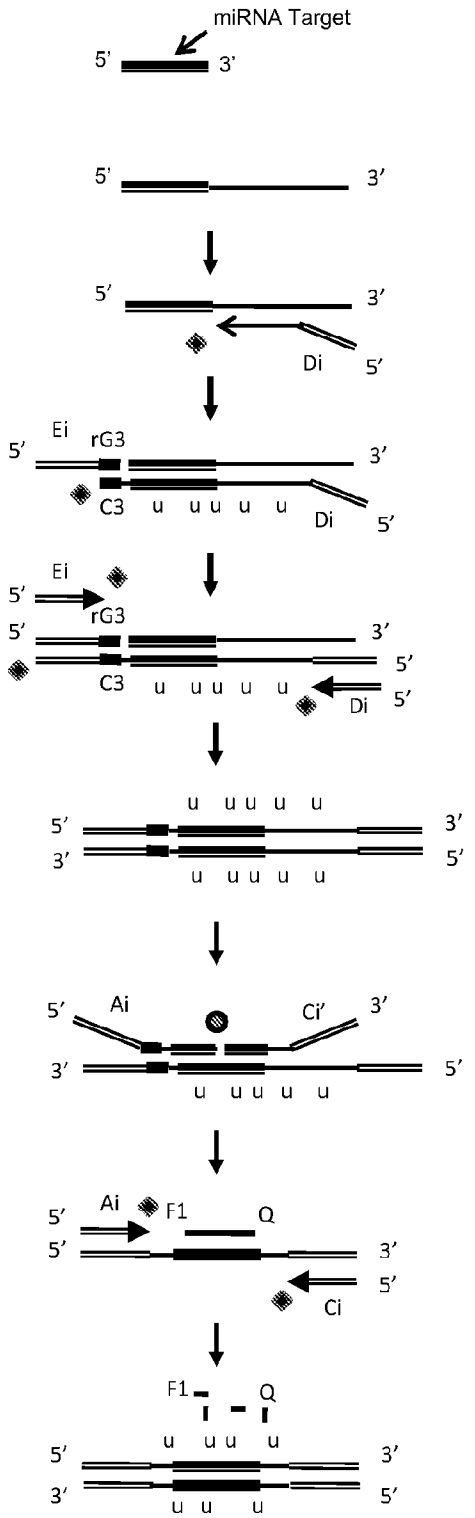


Figure 31

A. RT-PCR-qPCR carryover prevention reaction to detect and enumerate miRNA. Isolate miRNA from exosomes.

B. For accurate enumeration, aliquot into 12, 24, 36, or 48 wells prior to PCR. For higher copy number, distribute equally in 13 wells, dilute last well equally into 13 additional wells, and repeat for remaining wells. Treat with UDG (uracil DNA glycosylase) for carryover prevention, heat kill. PolyA tail miRNA with Poly(A) Polymerase. Extend with primer comprising a Tag Di on the 5' end, and a T30VN sequence at the 3' end, using reverse transcriptase, which appends C3 on 3' end. Hybridize Tag (Ei) with rGrG+G on 3' end (+G is symbol for LNA). Reverse transcriptase undergoes strand switching and copies Ei. Activate Taq polymerase and perform limited cycle PCR amplification (8-12), using tag primers (Di, Ei), to maintain relative ratios of different amplicons.

C. Distribute products into wells, micro-pores, or droplets for Taqman reactions using nested miRNA-specific primers with Universal tag tails, including tag-specific primers Ai and Ci. Primers are unblocked with RNaseH2 only when bound to correct target.

D. PCR products comprise of Ai tag sequence, a portion of Ei, G3, miRNA-specific sequence, a portion of the stem-loop sequence, and Ci' tag sequence.

E. Tag-specific primers (Ai and Ci) then amplify authentic transcript-specific products. Each Taqman probe spans a unique miRNA-specific sequence.

F. The 5'->3' nuclease activity of Taq polymerase liberates the fluorescent group to generate signal. Determine miRNA copy number based on signal from wells where original distribution was one copy/well. (PCR products incorporate dU, allowing for carryover prevention.)

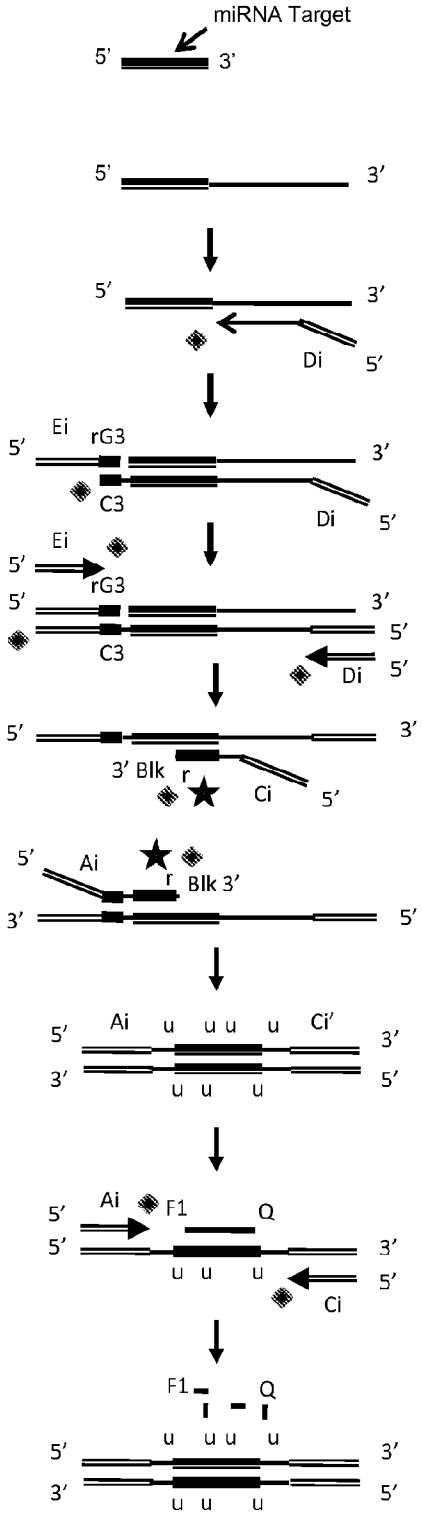
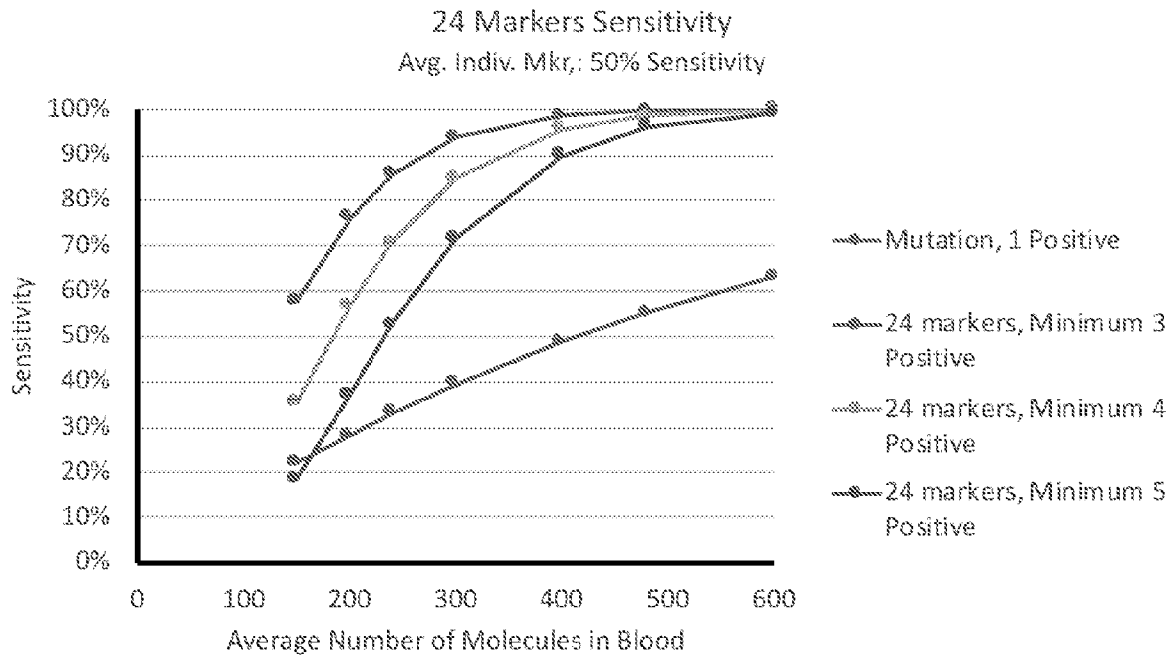
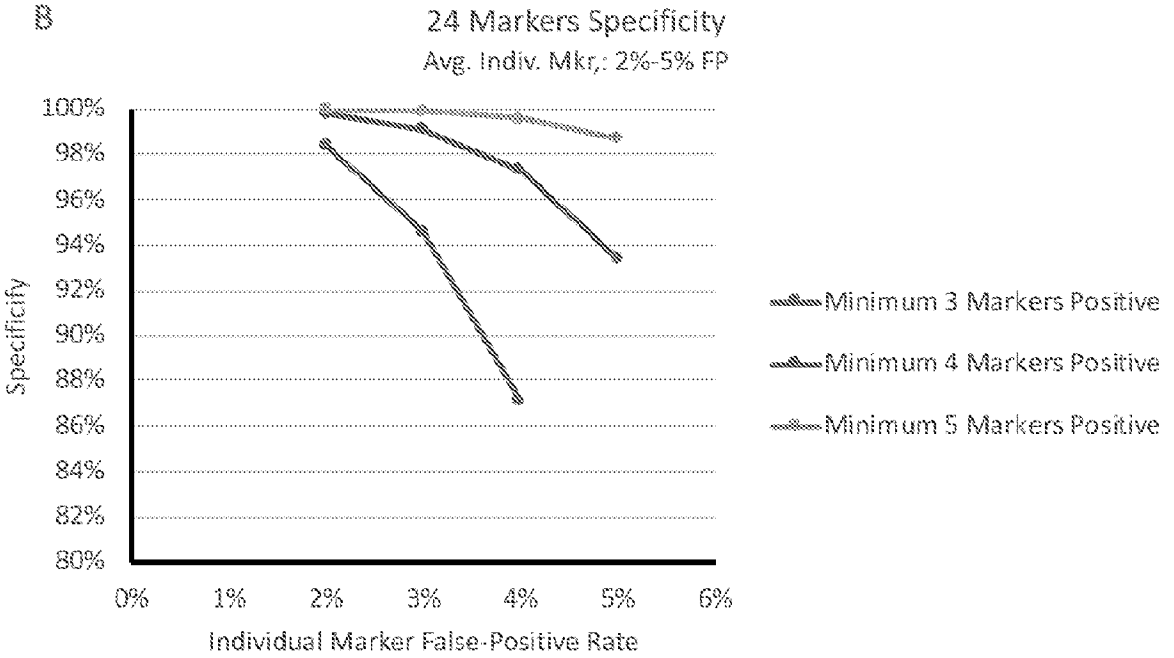


Figure 32

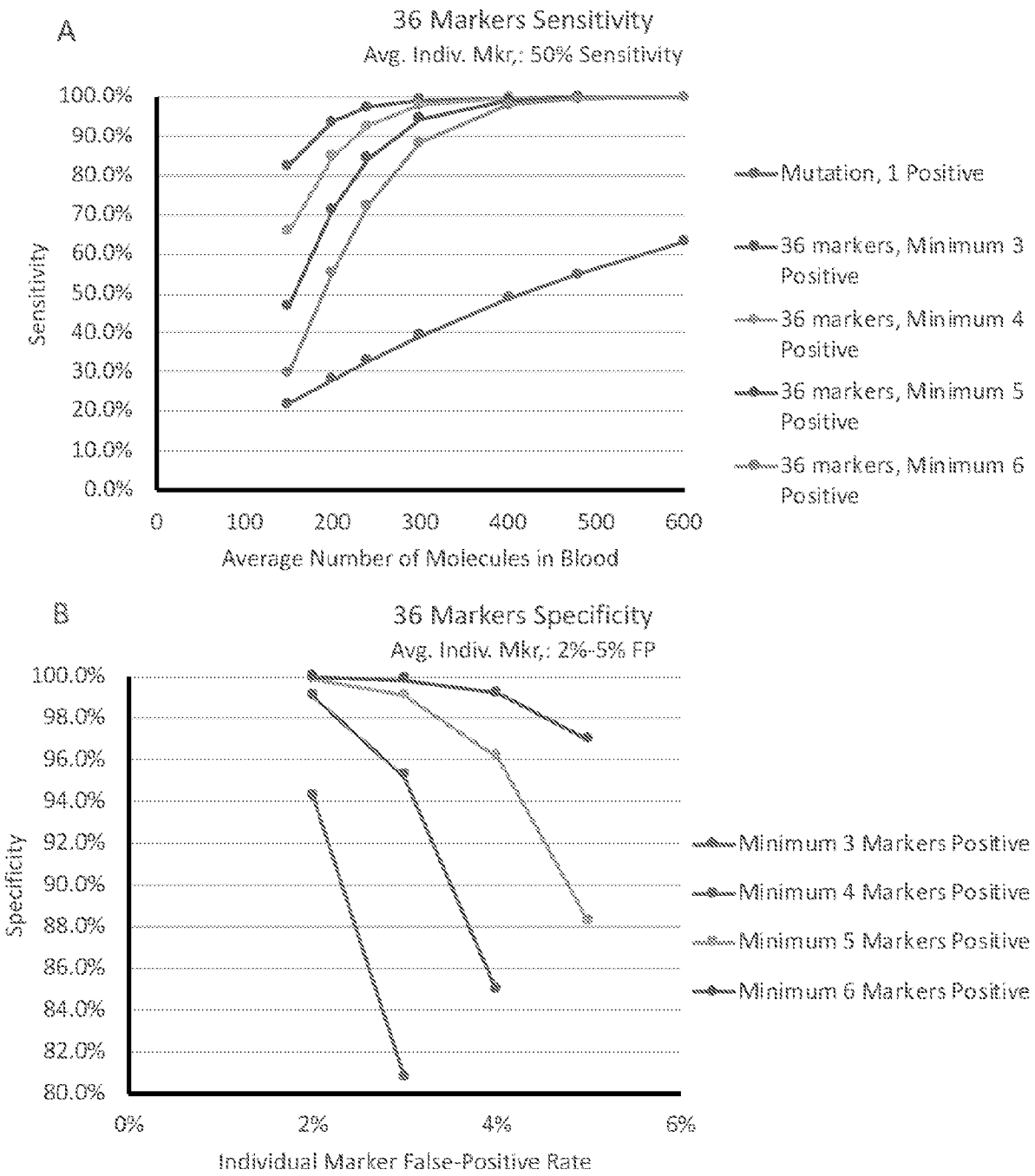
A



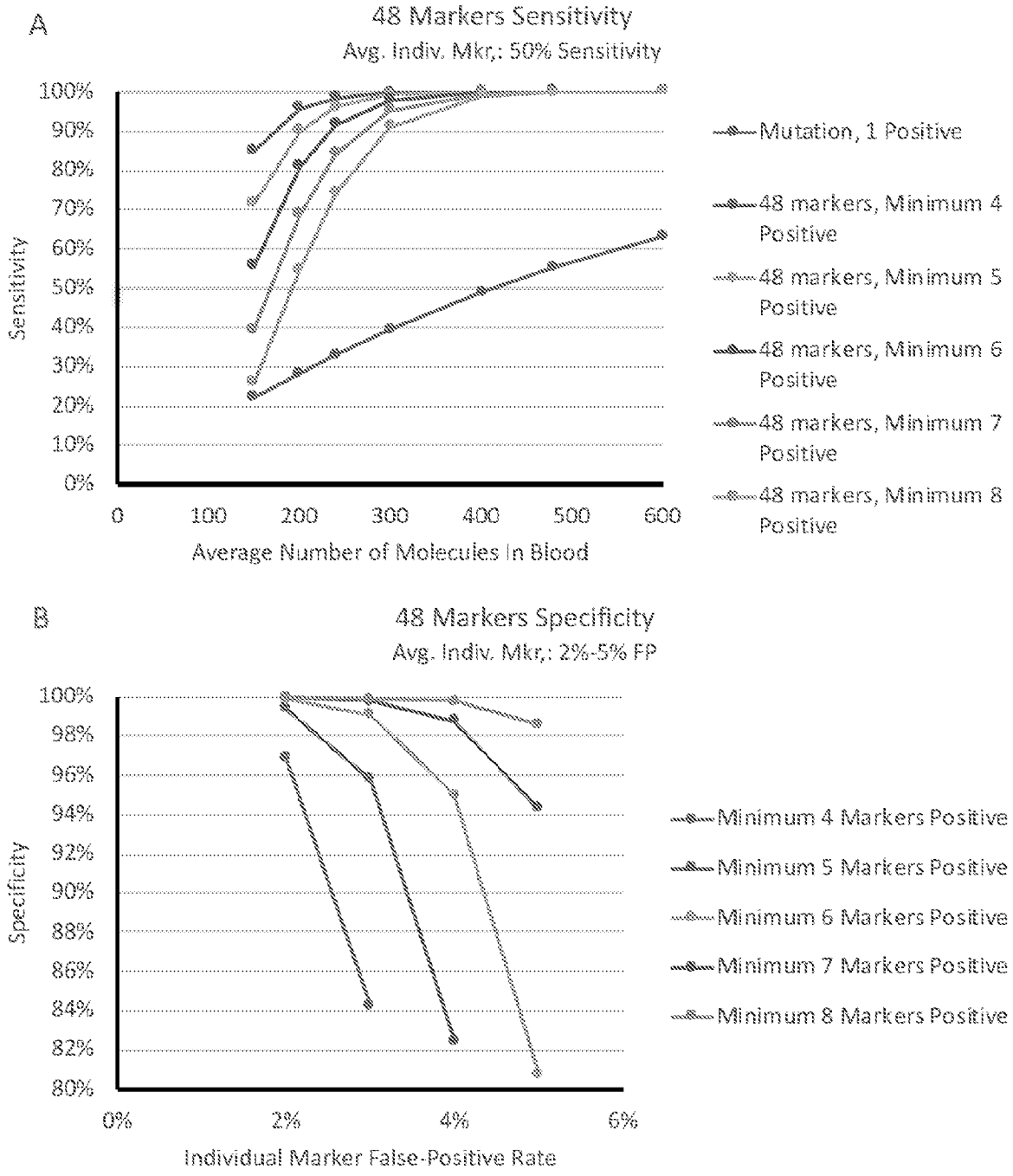
B



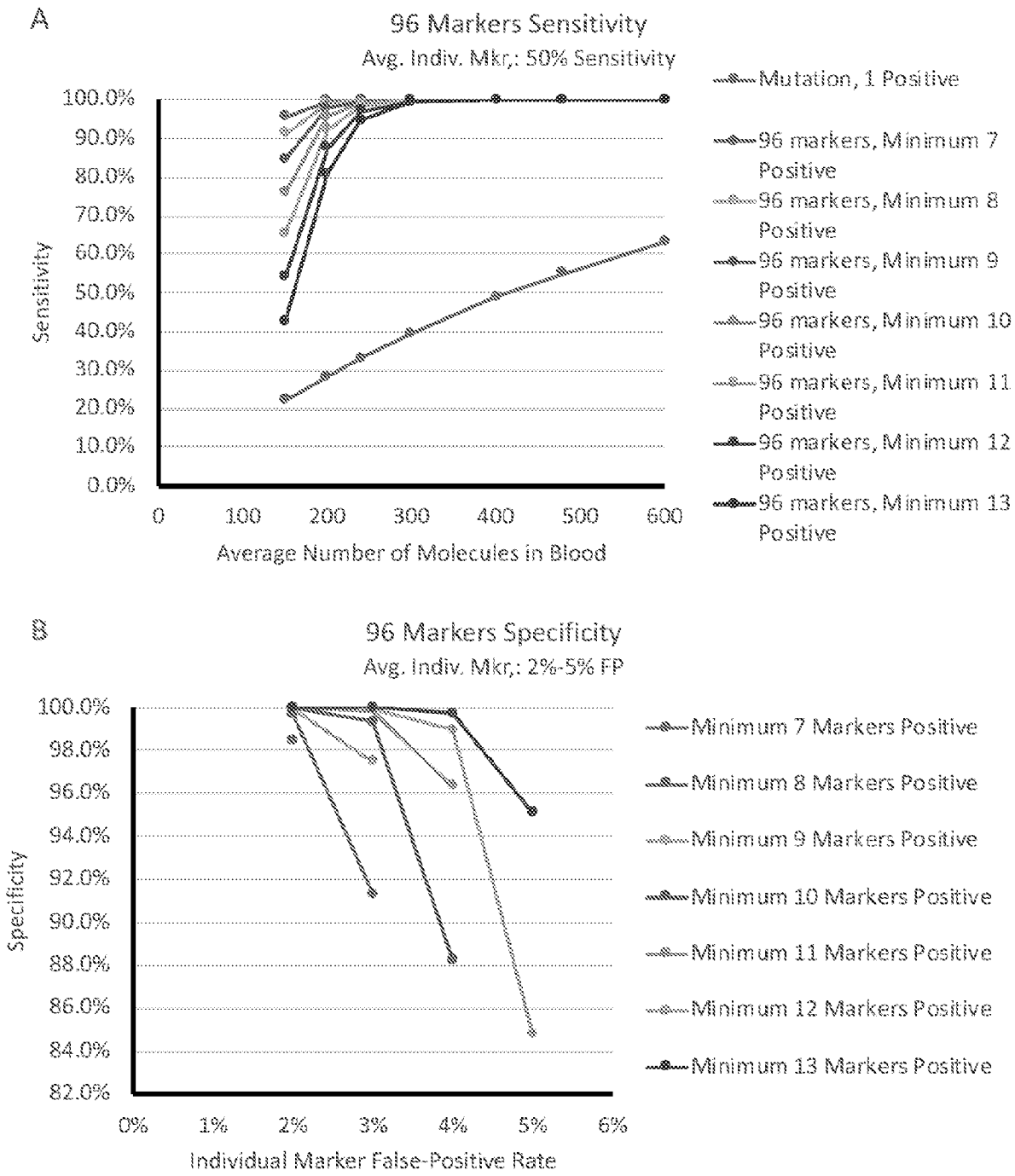
Figures 33A-B



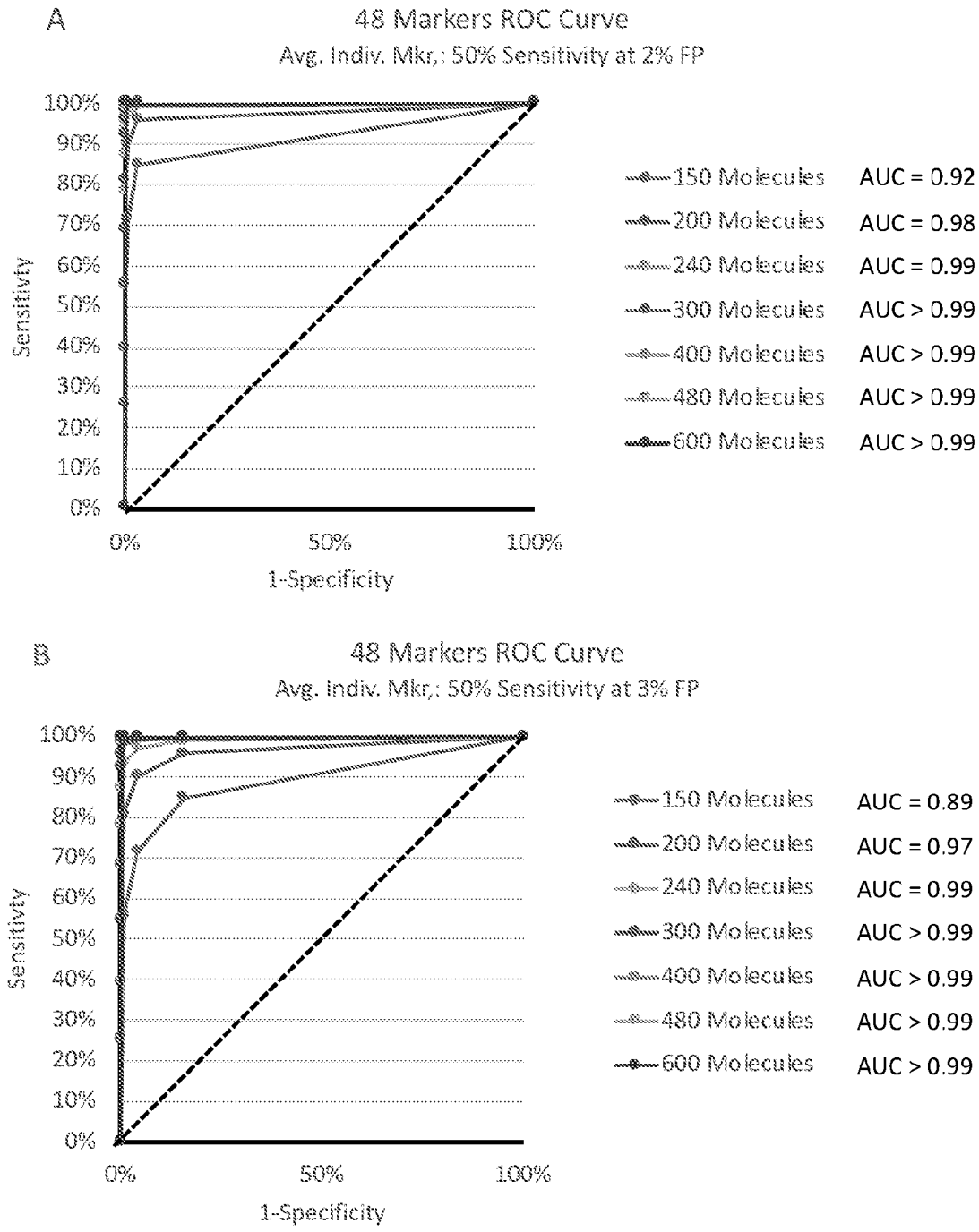
Figures 34A-B



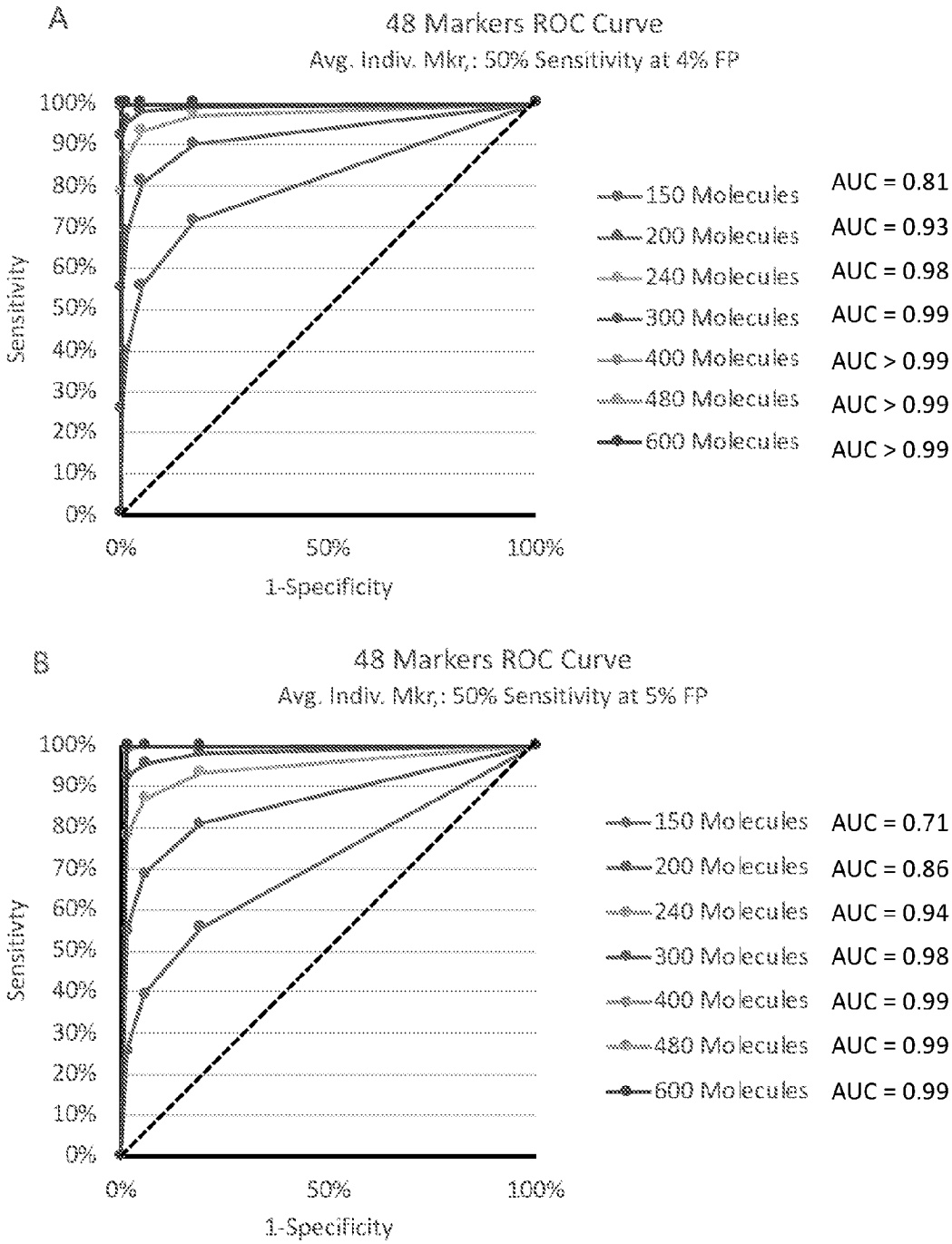
Figures 35A-B



Figures 36A-B



Figures 37A-B



Figures 38A-B

CRC miRNA markers

Order	Id
1	hsa-let-7f-1
2	hsa-let-7f-2
3	hsa-let-7g
4	hsa-mir-101-1
5	hsa-mir-101-2
6	hsa-mir-107
7	hsa-mir-10a
8	hsa-mir-1-2
9	hsa-mir-1207
10	hsa-mir-1229
11	hsa-mir-1245
12	hsa-mir-1246
13	hsa-mir-125b
14	hsa-mir-126
15	hsa-mir-1290
16	hsa-mir-1296
17	hsa-mir-135b
18	hsa-mir-136
19	hsa-mir-141
20	hsa-mir-142
21	hsa-mir-143
22	hsa-mir-144
23	hsa-mir-148a
24	hsa-mir-150
25	hsa-mir-151
26	hsa-mir-152
27	hsa-mir-153-1
28	hsa-mir-153-2
29	hsa-mir-15a
30	hsa-mir-15b
31	hsa-mir-16-1

Order	Id
32	hsa-mir-16-2
33	hsa-mir-17
34	hsa-mir-17-3p
35	hsa-mir-181d
36	hsa-mir-182
37	hsa-mir-183
38	hsa-mir-185
39	hsa-mir-196a-1
40	hsa-mir-199a-2
41	hsa-mir-199b
42	hsa-mir-19a
43	hsa-mir-19b
44	hsa-mir-19b-1
45	hsa-mir-19b-2
46	hsa-mir-200c
47	hsa-mir-203
48	hsa-mir-20a
49	hsa-mir-21
50	hsa-mir-215
51	hsa-mir-217
52	hsa-mir-218-2
53	hsa-mir-223
54	hsa-mir-224
55	hsa-mir-224-3p
56	hsa-mir-23a
57	hsa-mir-24
58	hsa-mir-24-2
59	hsa-mir-26a-2
60	hsa-mir-26b
61	hsa-mir-27a
62	hsa-mir-29a

Figure 39A

Order	Id
63	hsa-mir-29b-1
64	hsa-mir-29b-2
65	hsa-mir-29c
66	hsa-mir-301a
67	hsa-mir-30b
68	hsa-mir-30e
69	hsa-mir-31
70	hsa-mir-32
71	hsa-mir-320a
72	hsa-mir-335
73	hsa-mir-337
74	hsa-mir-33a
75	hsa-mir-340
76	hsa-mir-34a
77	hsa-mir-369
78	hsa-mir-374a
79	hsa-mir-374b
80	hsa-mir-376a-1
81	hsa-mir-376c
82	hsa-mir-379
83	hsa-mir-380
84	hsa-mir-411
85	hsa-mir-421
86	hsa-mir-423-5p
87	hsa-mir-424
88	hsa-mir-429
89	hsa-mir-450a-1
90	hsa-mir-450a-2
91	hsa-mir-450b
92	hsa-mir-451
93	hsa-mir-452

Order	Id
94	hsa-mir-454
95	hsa-mir-494
96	hsa-mir-495
97	hsa-mir-496
98	hsa-mir-542
99	hsa-mir-552
100	hsa-mir-556
101	hsa-mir-576-5p
102	hsa-mir-577
103	hsa-mir-582
104	hsa-mir-590
105	hsa-mir-592
106	hsa-mir-598
107	hsa-mir-612
108	hsa-mir-628
109	hsa-mir-653
110	hsa-mir-655
111	hsa-mir-660
112	hsa-mir-708
113	hsa-mir-7-1
114	hsa-mir-7-2
115	hsa-mir-7-3
116	hsa-mir-885-5p
117	hsa-mir-889
118	hsa-mir-92a
119	hsa-mir-933
120	hsa-mir-937
121	hsa-mir-95
122	hsa-mir-96
123	hsa-mir-98

Figure 39B

CRC ncRNA markers

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
1	LINC00115	chr1:826206-827522	ENSG00000225880.5
2	RP11-54O7.10	chr1:1008076-1008229	ENSG00000231702.2
3	RP11-465B22.3	chr1:1059734-1069355	ENSG00000217801.8
4	RP11-465B22.8	chr1:1169357-1170343	ENSG00000272141.1
5	RP4-758J18.13	chr1:1409096-1410618	ENSG00000272455.1
6	RP11-547D24.1	chr1:2013213-2015530	ENSG00000233542.1
7	RP11-22L13.1	chr1:3306636-3310096	ENSG00000272235.1
8	RP11-338N10.2	chr1:7698303-7698872	ENSG00000270035.1
9	RP11-338N10.3	chr1:7700704-7700970	ENSG00000269978.1
10	RP11-5P18.10	chr1:16035178-16040029	ENSG00000232456.1
11	EIF1AXP1	chr1:16685621-16686055	ENSG00000236698.1
12	RP5-1056L3.3	chr1:19608114-19608568	ENSG00000226396.1
13	LINC01355	chr1:23281309-23286752	ENSG00000261326.2
14	NPM1P39	chr1:27206930-27207796	ENSG00000225159.1
15	RP11-4K3__A.5	chr1:27325329-27325796	ENSG00000241547.1
16	SNHG3	chr1:28505980-28510892	ENSG00000242125.3
17	SNORA73B	chr1:28508559-28508762	ENSG00000200087.1
18	PRDX3P2	chr1:28526318-28527227	ENSG00000231468.2
19	SNHG12	chr1:28578538-28582983	ENSG00000197989.12
20	SNORD99	chr1:28578749-28578822	ENSG00000221539.1
21	RP4-604A21.1	chr1:28982278-28982608	ENSG00000225616.2
22	MATN1-AS1	chr1:30718504-30726827	ENSG00000186056.8
23	RP1-118J21.25	chr1:39788976-39790171	ENSG00000261798.1
24	RP1-228H13.5	chr1:40464319-40466767	ENSG00000260920.2
25	NFYC-AS1	chr1:40690380-40692066	ENSG00000272145.1
26	RP1-92O14.6	chr1:43385113-43389155	ENSG00000229431.1
27	KDM4A-AS1	chr1:43699765-43708138	ENSG00000236200.4
28	RP11-570P14.1	chr1:44122153-44122512	ENSG00000233514.1
29	RP11-269F19.2	chr1:44759037-44775810	ENSG00000225721.4
30	SNORD46	chr1:44776490-44776593	ENSG00000200913.1
31	SNORD38A	chr1:44777843-44777912	ENSG00000202031.1
32	CYP4A22-AS1	chr1:47096653-47179271	ENSG00000225506.2
33	RP11-296A18.5	chr1:51250603-51251472	ENSG00000233406.5
34	RPS13P2	chr1:52772194-52772648	ENSG00000228929.1
35	RP11-776H12.1	chr1:60540249-60640491	ENSG00000226476.2
36	RP11-436K8.1	chr1:60659631-60825584	ENSG00000231252.1
37	LINC01555	chr1:84628230-84635020	ENSG00000180869.4
38	RP11-384B12.3	chr1:87045875-87046700	ENSG00000225568.1
39	HSP90B3P	chr1:91642516-91644082	ENSG00000203914.4
40	GAPDHP29	chr1:94302038-94303022	ENSG00000178596.10
41	RP11-421L21.3	chr1:101025878-101087268	ENSG00000233184.5
42	RP4-575N6.2	chr1:101256274-101256616	ENSG00000228399.1
43	RP11-270C12.3	chr1:107505203-107505448	ENSG00000224550.1
44	RP11-552M11.4	chr1:111438638-111441364	ENSG00000243960.1
45	LINC01356	chr1:112820170-112850643	ENSG00000215866.6
46	RP3-522D1.1	chr1:112850028-112877871	ENSG00000224167.1
47	SLC16A1-AS1	chr1:112956415-112999496	ENSG00000226419.5
48	RP11-389O22.4	chr1:113168994-113169403	ENSG00000233839.1
49	PKMP1	chr1:114535995-114537840	ENSG00000236480.1
50	NAP1L4P1	chr1:116532936-116534092	ENSG00000177173.5

Figure 40A

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
51	U1	chr1:145465617-145465780	ENSG00000270722.1
52	LINC01138	chr1:148362995-148460150	ENSG00000274020.1
53	RP11-196G18.22	chr1:149844498-149849024	ENSG00000261716.1
54	BX470102.3	chr1:153533603-153535115	ENSG00000238279.1
55	RP11-422P24.9	chr1:153995632-153995960	ENSG00000231416.1
56	GBAP1	chr1:155213821-155227422	ENSG00000160766.13
57	RUSC1-AS1	chr1:155316863-155324176	ENSG00000225855.5
58	MSTO2P	chr1:155745829-155750137	ENSG00000203761.5
59	RP11-284F21.10	chr1:156641656-156644887	ENSG00000272405.1
60	RP11-404O13.1	chr1:158132044-158140639	ENSG00000231666.4
61	RP1-79C4.4	chr1:170667381-170669425	ENSG00000271811.1
62	GAS5	chr1:173863900-173868882	ENSG00000234741.6
63	RP1-35C21.2	chr1:177393287-177597709	ENSG00000227579.4
64	RP11-568K15.1	chr1:178006136-178037950	ENSG00000242193.8
65	RP11-568K15.2	chr1:178017127-178017343	ENSG00000276563.1
66	RASAL2-AS1	chr1:178091508-178093984	ENSG00000224687.1
67	RP11-533E19.7	chr1:179881607-179882595	ENSG00000272906.1
68	SMG7-AS1	chr1:183460874-183472265	ENSG00000232860.6
69	ERVMER61-1	chr1:187641226-187643860	ENSG00000230426.2
70	RP11-510N19.5	chr1:202011370-202015657	ENSG00000249007.1
71	PCAT6	chr1:202810954-202812156	ENSG00000228288.5
72	RP11-480I12.5	chr1:202851828-202861620	ENSG00000214796.7
73	SNRPGP10	chr1:205351247-205351471	ENSG00000235363.1
74	LEMD1-AS1	chr1:205373252-205387440	ENSG00000226235.1
75	MIR135B	chr1:205448291-205448403	ENSG00000199059.2
76	SNX25P1	chr1:211417025-211417934	ENSG00000236809.2
77	FDPSP8	chr1:211660189-211661277	ENSG00000224763.1
78	PROX1-AS1	chr1:213817751-213988508	ENSG00000230461.7
79	RP11-478J18.2	chr1:213983793-213986419	ENSG00000274895.1
80	RP11-191N8.2	chr1:221827666-221840666	ENSG00000227925.1
81	RNU6-403P	chr1:221837334-221837437	ENSG00000200033.1
82	RP11-815M8.1	chr1:221880981-221978523	ENSG00000238042.4
83	RP11-400N13.2	chr1:221966341-221984964	ENSG00000228437.4
84	RP11-400N13.3	chr1:222041705-222064763	ENSG00000232679.1
85	RP11-504P24.8	chr1:223951388-223953439	ENSG00000272645.2
86	RP11-285F7.2	chr1:225840883-225846522	ENSG00000242861.1
87	RP11-396C23.2	chr1:226083639-226090292	ENSG00000225518.2
88	DUSP5P1	chr1:228650241-228651379	ENSG00000183929.7
89	RP5-1061H20.4	chr1:229258281-229271028	ENSG00000177788.5
90	RP5-858B6.3	chr1:230874846-230879015	ENSG00000223393.1
91	LINC01132	chr1:234724042-234731643	ENSG00000227630.2
92	LINC01348	chr1:235065479-235074220	ENSG00000280587.1
93	RP11-467I20.3	chr1:240636599-240637052	ENSG00000228844.1
94	RP11-278H7.5	chr1:244064330-244067909	ENSG00000279774.1
95	RP11-278H7.4	chr1:244068820-244093026	ENSG00000229960.1
96	AC144450.2	chr2:1620510-1625419	ENSG00000203635.2
97	AC018685.1	chr2:2641989-2656630	ENSG00000234929.1
98	TSSC1-IT1	chr2:3298341-3301465	ENSG00000224885.1
99	RNASEH1-AS1	chr2:3558492-3561745	ENSG00000234171.2
100	RP11-734K21.2	chr2:9103440-9105114	ENSG00000242136.1

Figure 40B

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
101	RP11-734K21.5	chr2:9106593-9109865	ENSG00000261104.1
102	RP11-95D17.1	chr2:9936360-9939590	ENSG00000269973.1
103	RP11-254F7.1	chr2:10021578-10022825	ENSG00000260476.1
104	AC079145.4	chr2:19990217-20004210	ENSG00000227210.1
105	RP11-443B20.1	chr2:24825610-24826717	ENSG00000271936.1
106	TRMT112P6	chr2:26028208-26028612	ENSG00000268412.2
107	GTF3C2-AS1	chr2:27335535-27342599	ENSG00000234945.6
108	AC074117.10	chr2:27356246-27367622	ENSG00000234072.1
109	DDX50P1	chr2:32201600-32203999	ENSG00000229816.1
110	AC016735.1	chr2:43001355-43006060	ENSG00000224739.2
111	RP11-89K21.1	chr2:44921077-44939199	ENSG00000259439.2
112	AC093732.1	chr2:47035279-47040524	ENSG00000233845.1
113	AC073283.7	chr2:47067822-47071204	ENSG00000225187.1
114	AC010967.2	chr2:52722677-52910020	ENSG00000228033.4
115	RP11-416L21.2	chr2:60682873-60683244	ENSG00000271134.1
116	AHSA2	chr2:61177418-61191203	ENSG00000173209.21
117	AC107081.5	chr2:61868432-61886082	ENSG00000236498.1
118	RP11-474G23.3	chr2:68252870-68253848	ENSG00000273064.1
119	RP11-44N22.3	chr2:73113018-73115907	ENSG00000272702.1
120	HMGA1P8	chr2:74418122-74418442	ENSG00000233155.1
121	AC007099.1	chr2:75524068-75542706	ENSG00000231172.2
122	AC105399.2	chr2:77788382-77793087	ENSG00000237077.1
123	SNORD94	chr2:86135870-86136006	ENSG00000208772.1
124	AC133644.2	chr2:87439523-87459044	ENSG00000280721.1
125	LINC00152	chr2:87455368-87606805	ENSG00000222041.9
126	AC093616.4	chr2:87700984-87738105	ENSG00000234231.3
127	AC012671.2	chr2:88428078-88428615	ENSG00000224291.1
128	RP11-332H14.2	chr2:105334027-105337475	ENSG00000272994.1
129	AC023672.2	chr2:108049200-108052755	ENSG00000231221.1
130	AC019100.3	chr2:108167748-108217841	ENSG00000225328.1
131	RP11-443K8.1	chr2:108317725-108318611	ENSG00000261209.1
132	RP11-465O11.2	chr2:108367116-108367938	ENSG00000271070.1
133	MIR4435-1HG	chr2:111196350-111495100	ENSG00000172965.13
134	AC108463.2	chr2:111203964-111206215	ENSG00000227992.1
135	RP11-803D5.1	chr2:111321433-111324812	ENSG00000279191.1
136	Y_RNA	chr2:112579484-112579584	ENSG00000207383.1
137	AC016745.3	chr2:113424495-113425324	ENSG00000234997.1
138	AC009404.2	chr2:117833937-117841658	ENSG00000236255.1
139	AC012363.4	chr2:120174885-120216544	ENSG00000224789.1
140	MTND4P26	chr2:120213631-120214989	ENSG00000234065.2
141	MTND5P28	chr2:120215181-120217279	ENSG00000223549.1
142	LINC01101	chr2:120464335-120466218	ENSG00000280409.1
143	AC133785.1	chr2:130830978-130836994	ENSG00000233221.5
144	AC073869.20	chr2:131442644-131444587	ENSG00000217950.4
145	AC093838.4	chr2:131492813-131521573	ENSG00000152117.16
146	C2orf27A	chr2:131722375-131767404	ENSG00000197927.11
147	AC093724.2	chr2:131723253-131724279	ENSG00000213222.3
148	AC064850.4	chr2:135510546-135511943	ENSG00000235651.1
149	DARS-AS1	chr2:135985176-136022593	ENSG00000231890.6
150	AC092484.1	chr2:145569294-145588024	ENSG00000225107.1

Figure 40C

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
151	AC012066.1	chr2:152389937-152390630	ENSG00000213197.3
152	AC012501.3	chr2:153420677-153421957	ENSG00000242575.1
153	AC012501.2	chr2:153421616-153593288	ENSG00000227400.2
154	AC005042.4	chr2:158658337-158735002	ENSG00000204380.3
155	RP11-452N17.1	chr2:164573741-164574054	ENSG00000277998.1
156	AC007277.3	chr2:170618628-170695374	ENSG00000213981.7
157	LINC01124	chr2:170712451-170714567	ENSG00000222033.1
158	EIF2S2P4	chr2:170751805-170752788	ENSG00000128692.8
159	AC007405.6	chr2:170771113-170778148	ENSG00000239467.4
160	AC007405.8	chr2:170816293-170818037	ENSG00000235934.1
161	AC007969.5	chr2:171517270-171517716	ENSG00000233762.3
162	AC104088.1	chr2:172315287-172323737	ENSG00000232555.1
163	MLK7-AS1	chr2:173166446-173282036	ENSG00000238133.5
164	AC092573.2	chr2:173338219-173356208	ENSG00000236391.3
165	RP11-796E10.1	chr2:173811076-173811392	ENSG00000270460.1
166	RP11-394I13.2	chr2:173968351-173969418	ENSG00000271151.1
167	RP11-438L19.1	chr2:183214319-183215400	ENSG00000272800.1
168	LINC01473	chr2:186032891-186083233	ENSG00000237877.5
169	AC013439.4	chr2:189486480-189487992	ENSG00000233996.1
170	AC092614.2	chr2:191229165-191246172	ENSG00000227542.1
171	AC064834.1	chr2:195448532-195478925	ENSG00000224099.1
172	PSMA2P3	chr2:202179297-202179971	ENSG00000227890.1
173	SNORD51	chr2:206161878-206161957	ENSG00000207047.2
174	PPP1R14BP2	chr2:207681590-207682033	ENSG00000213082.3
175	AC093850.2	chr2:215718043-215719424	ENSG00000230838.1
176	HSPA9P1	chr2:221961737-221963765	ENSG00000226666.1
177	AC012512.1	chr2:223704621-223705338	ENSG00000226539.1
178	SPATA3-AS1	chr2:230984368-230996032	ENSG00000238062.4
179	AC010148.1	chr2:234834315-234913384	ENSG00000235726.4
180	AC112721.1	chr2:237421420-237425276	ENSG00000222022.1
181	AC112721.2	chr2:237428920-237434822	ENSG00000222032.1
182	AC079612.1	chr2:239578301-239586094	ENSG00000196758.3
183	CAPN10-AS1	chr2:240582700-240586699	ENSG00000260942.1
184	RP11-97C16.1	chr3:3152942-3153435	ENSG00000271870.1
185	PNPT1P1	chr3:3981981-3984295	ENSG00000229241.1
186	LMCD1-AS1	chr3:7952805-8611924	ENSG00000227110.5
187	THUMPD3-AS1	chr3:9349689-9398579	ENSG00000206573.7
188	PRRT3-AS1	chr3:9947404-9954787	ENSG00000230082.1
189	EMC3-AS1	chr3:9986893-10006990	ENSG00000180385.7
190	TPRXL	chr3:13937256-14082811	ENSG00000180438.13
191	LINC01267	chr3:14348451-14352568	ENSG00000251576.1
192	COL6A4P1	chr3:15151833-15205959	ENSG00000230524.7
193	AC090945.1	chr3:15878047-15879571	ENSG00000224728.1
194	AC104297.1	chr3:17871729-17872598	ENSG00000213383.2
195	RP11-384L8.1	chr3:32236688-32238578	ENSG00000261572.1
196	KRT18P15	chr3:32258940-32260230	ENSG00000234737.1
197	RPL30P4	chr3:32635222-32635554	ENSG00000237676.1
198	AC018359.1	chr3:34159334-34562877	ENSG00000226320.4
199	AC123023.1	chr3:34203244-34268811	ENSG00000236452.1
200	HSPD1P6	chr3:36767117-36784167	ENSG00000230067.3

Figure 40D

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
201	C3orf35	chr3:37386269-37435497	ENSG00000198590.10
202	HNRNPA1P21	chr3:39334979-39335939	ENSG00000228168.1
203	RP11-697K23.3	chr3:45749627-45750651	ENSG00000279017.1
204	LRRC2-AS1	chr3:46557398-46559694	ENSG00000268324.2
205	NRADDP	chr3:47011542-47013146	ENSG00000236409.1
206	RP11-447D11.3	chr3:47346950-47349073	ENSG00000280173.1
207	GPX1	chr3:49357176-49358600	ENSG00000233276.3
208	U73166.2	chr3:50260303-50263358	ENSG00000230454.1
209	RP11-168J18.6	chr3:52373652-52374882	ENSG00000239557.1
210	SNORD19	chr3:52689248-52689315	ENSG00000212493.1
211	SNORD19	chr3:52691378-52691453	ENSG00000222345.1
212	SNORD69	chr3:52692736-52692812	ENSG00000212452.1
213	RP11-884K10.7	chr3:53858994-53861576	ENSG00000271976.1
214	RP11-157F20.3	chr3:57078943-57080101	ENSG00000272202.1
215	RPL10AP6	chr3:61742455-61743106	ENSG00000226360.5
216	LINC00698	chr3:62950430-63125062	ENSG00000244342.4
217	RP11-383I23.2	chr3:99802699-99806058	ENSG00000273374.1
218	RP11-234A1.1	chr3:101576489-101576947	ENSG00000242299.1
219	RP11-18H7.1	chr3:120094895-120136783	ENSG00000242622.1
220	RP11-174O3.3	chr3:120349510-120367998	ENSG00000240661.1
221	NAP1L1P3	chr3:120805866-120806745	ENSG00000213371.4
222	FAM86JP	chr3:125916620-125930024	ENSG00000171084.14
223	RP11-390G14.1	chr3:126624792-126626061	ENSG00000248557.1
224	GATA2-AS1	chr3:128489212-128502348	ENSG00000244300.2
225	RP11-723O4.7	chr3:128914833-128915060	ENSG00000279507.1
226	RPL32P3	chr3:129382922-129399655	ENSG00000251474.5
227	TMCC1-AS1	chr3:129893871-129918575	ENSG00000271270.4
228	RP11-93K22.6	chr3:129954105-129969294	ENSG00000250643.1
229	FAM86HP	chr3:130099092-130111472	ENSG00000253540.4
230	LINC01210	chr3:137771949-137780878	ENSG00000239513.4
231	RP11-2A4.4	chr3:137791973-137796678	ENSG00000261146.1
232	RP11-343B5.1	chr3:142043623-142044198	ENSG00000233597.3
233	RP11-372E1.1	chr3:142827096-142827728	ENSG00000240950.1
234	TM4SF1-AS1	chr3:149377778-149386583	ENSG00000240541.2
235	RP11-278L15.2	chr3:149384179-149385800	ENSG00000243885.1
236	RP11-167H9.4	chr3:150039214-150213726	ENSG00000243944.4
237	RP11-674E16.4	chr3:155742142-155743726	ENSG00000214919.3
238	SETP14	chr3:155987304-155988176	ENSG00000240489.1
239	PA2G4P4	chr3:156809551-156810732	ENSG00000230457.3
240	KRT8P12	chr3:160565447-160569248	ENSG00000229320.3
241	RP11-373E16.1	chr3:170653846-170654259	ENSG00000213174.3
242	ATP5G1P4	chr3:172828736-172829176	ENSG00000227440.1
243	RP11-393N4.2	chr3:174377244-174378005	ENSG00000213169.3
244	KCNMB2-AS1	chr3:178526505-178860405	ENSG00000237978.4
245	YEATS2-AS1	chr3:183806457-183810783	ENSG00000233885.6
246	RP11-778D9.4	chr3:184149704-184150435	ENSG00000228205.1
247	RPL4P4	chr3:185417495-185418778	ENSG00000229638.1
248	RP11-573D15.2	chr3:186810880-186825521	ENSG00000232233.1
249	RPS20P14	chr3:186900198-186900557	ENSG00000223803.1
250	RP11-30O15.1	chr3:187796187-187805258	ENSG00000224049.1

Figure 40E

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
251	RP11-197K6.1	chr3:191557324-191590356	ENSG00000223812.4
252	RP11-480A16.1	chr3:195949188-195952695	ENSG00000260261.1
253	MFI2-AS1	chr3:196999460-197004744	ENSG00000228109.1
254	RP11-440L14.1	chr4:762387-781849	ENSG00000249592.4
255	RP11-20I20.4	chr4:1167778-1168174	ENSG00000273179.1
256	RP11-572O17.1	chr4:1712821-1713622	ENSG00000270195.1
257	STX18-AS1	chr4:4542131-4710938	ENSG00000247708.6
258	RP11-539L10.2	chr4:6687448-6690519	ENSG00000246526.2
259	GMPSP1	chr4:8174421-8174999	ENSG00000250471.2
260	RNPS1P1	chr4:11371975-11372892	ENSG00000250896.1
261	RP11-333E13.2	chr4:39973128-39973442	ENSG00000250568.1
262	PABPC1P1	chr4:39973444-39974338	ENSG00000231707.4
263	SNORA26	chr4:52713249-52713370	ENSG00000212588.1
264	RPS15AP17	chr4:62105660-62106049	ENSG00000225405.3
265	HMGA1P2	chr4:73098822-73099145	ENSG00000248641.1
266	AC142293.3	chr4:74552584-74589481	ENSG00000249942.1
267	TXNP6	chr4:76958860-76959169	ENSG00000234036.4
268	RP11-127B20.3	chr4:82374301-82384027	ENSG00000272677.1
269	RP11-791G16.2	chr4:82494786-82496278	ENSG00000227304.3
270	THAP9-AS1	chr4:82893009-82900960	ENSG00000251022.5
271	RP11-742B18.1	chr4:87568035-87733956	ENSG00000249001.4
272	HSP90AB3P	chr4:87891843-87894015	ENSG00000183199.6
273	RP11-84C13.2	chr4:89119284-89119871	ENSG00000270720.1
274	RP11-428L21.2	chr4:92821986-92822918	ENSG00000249239.2
275	RP11-328K4.1	chr4:103256159-103453658	ENSG00000248740.4
276	LEF1-AS1	chr4:108167525-108256836	ENSG00000232021.5
277	AC113617.1	chr4:117360625-117372852	ENSG00000228358.4
278	KLHL2P1	chr4:119334329-119378233	ENSG00000250412.1
279	TECRP2	chr4:124462863-124463790	ENSG00000248565.2
280	RP11-420A23.1	chr4:128292751-128519394	ENSG00000251432.5
281	RP11-173E2.1	chr4:138560704-138561941	ENSG00000248795.1
282	RP11-223C24.2	chr4:142514446-142518983	ENSG00000251248.1
283	RP11-364L4.1	chr4:143349275-143350117	ENSG00000228981.3
284	HSPD1P5	chr4:144845625-144847344	ENSG00000249193.1
285	RP11-142A22.4	chr4:145335263-145340421	ENSG00000272727.1
286	RP11-752L20.3	chr4:147487971-147496062	ENSG00000280219.1
287	GTF2F2P1	chr4:147506091-147506765	ENSG00000251254.1
288	DKFZP434I0714	chr4:152536264-152539263	ENSG00000268471.4
289	RANP6	chr4:173633728-173634371	ENSG00000213370.3
290	RN7SL28P	chr4:184278936-184279233	ENSG00000244512.3
291	ADAM20P3	chr4:187746094-187748199	ENSG00000249162.1
292	RP11-310P5.1	chr5:602620-612210	ENSG00000249650.1
293	RP11-43F13.4	chr5:1005039-1006623	ENSG00000272347.1
294	LINC01511	chr5:1363582-1380067	ENSG00000250584.2
295	CTD-2256P15.1	chr5:10248325-10249915	ENSG00000248968.1
296	CCT6P2	chr5:14639426-14641018	ENSG00000250526.1
297	RP11-454P21.1	chr5:17684584-17955857	ENSG00000249937.4
298	HSPD1P1	chr5:21882585-21884310	ENSG00000213430.6
299	CTD-2108O9.4	chr5:38429892-38430160	ENSG00000248234.1
300	SNORD72	chr5:40832656-40832735	ENSG00000212296.1

Figure 40F

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
301	RP11-289K10.1	chr5:53413504-53414260	ENSG00000240052.1
302	RP11-506H20.1	chr5:55233934-55295201	ENSG00000251307.1
303	SNRPCP2	chr5:76376675-76377149	ENSG00000251235.3
304	NPM1P27	chr5:93682838-93683667	ENSG00000249353.2
305	RP11-138J23.1	chr5:104079911-104105403	ENSG00000251026.1
306	RN7SL255P	chr5:104260531-104260826	ENSG00000239808.3
307	RP11-313L6.2	chr5:104434772-104435499	ENSG00000278958.1
308	CCT5P1	chr5:115512077-115513694	ENSG00000250444.1
309	RP11-436H11.1	chr5:124808828-124809233	ENSG00000242814.2
310	KDELC1P1	chr5:128109486-128110994	ENSG00000250329.1
311	RP11-485M7.2	chr5:133052769-133052971	ENSG00000279691.1
312	C5orf66	chr5:135033280-135358219	ENSG00000224186.7
313	SNHG4	chr5:139274102-139283244	ENSG00000281398.1
314	CTB-35F21.2	chr5:139741349-139746223	ENSG00000249131.1
315	AC005592.2	chr5:142325293-142672001	ENSG00000231185.5
316	CTC-327F10.5	chr5:147886086-147886878	ENSG00000248362.1
317	CTC-327F10.4	chr5:147887112-147887704	ENSG00000251320.1
318	CTC-529P8.1	chr5:149063317-149109787	ENSG00000250072.4
319	GAPDHP40	chr5:159950493-159951498	ENSG00000248626.1
320	CTC-231O11.1	chr5:160468268-160487426	ENSG00000253522.3
321	RP11-67M9.1	chr5:166128498-166155439	ENSG00000254171.1
322	RP11-117L6.1	chr5:171365595-171367693	ENSG00000254256.1
323	SNORA70	chr5:171366118-171366252	ENSG00000206909.1
324	CTC-308K20.3	chr5:172975511-172976374	ENSG00000253785.1
325	CTB-43E15.2	chr5:173689459-173705849	ENSG00000253428.1
326	LINC01484	chr5:173707614-173746211	ENSG00000253686.1
327	LINC01485	chr5:173786790-173809039	ENSG00000254211.4
328	LINC01411	chr5:174336295-174532457	ENSG00000249306.4
329	GAPDHP71	chr5:174513305-174514307	ENSG00000213376.4
330	RP11-267A15.3	chr5:174618164-174625775	ENSG00000250256.1
331	CTD-2532K18.2	chr5:174820027-174826324	ENSG00000251144.1
332	PRMT1P1	chr5:177265580-177266588	ENSG00000270706.1
333	PRR7-AS1	chr5:177438503-177447699	ENSG00000246334.2
334	RP11-423H2.1	chr5:177809407-177950732	ENSG00000170089.14
335	RP11-889L3.1	chr5:178055604-178056194	ENSG00000218227.3
336	RP11-443C10.1	chr5:179377531-179378761	ENSG00000253163.1
337	RP1-140K8.5	chr6:3904920-3911979	ENSG00000260604.2
338	RP3-380B8.4	chr6:5851506-5870220	ENSG00000233064.2
339	PKMP5	chr6:5972412-5974012	ENSG00000215057.4
340	RP11-69L16.5	chr6:7338839-7339568	ENSG00000220472.1
341	OFCC1	chr6:9596110-10211608	ENSG00000181355.19
342	TFAP2A-AS1	chr6:10409340-10416446	ENSG00000229950.1
343	BTN2A3P	chr6:26421391-26432383	ENSG00000124549.13
344	RP1-15D7.1	chr6:27652602-27652825	ENSG00000216676.2
345	LINC01012	chr6:27694035-27710222	ENSG00000281706.1
346	XXbac-BPG308K3.5	chr6:28837869-28839006	ENSG00000225173.1
347	ZNRD1-AS1	chr6:30001011-30061640	ENSG00000204623.7
348	HCG18	chr6:30287397-30327150	ENSG00000231074.7
349	RANP1	chr6:30485940-30486590	ENSG00000236603.2
350	RNU6-850P	chr6:31756951-31757053	ENSG00000252743.1

Figure 40G

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
351	Y_RNA	chr6:31778817-31778905	ENSG00000201555.1
352	C6orf48	chr6:31834608-31839766	ENSG00000204387.11
353	SNORD48	chr6:31835263-31835326	ENSG00000201823.1
354	C2-AS1	chr6:31934474-31941724	ENSG00000281756.1
355	MIR6835	chr6:34240673-34240736	ENSG00000276404.1
356	MIR7111	chr6:35470508-35470579	ENSG00000276712.1
357	RP1-179N16.6	chr6:36146698-36197205	ENSG00000246982.5
358	RP1-278E11.3	chr6:39958414-39958833	ENSG00000180211.5
359	TREML3P	chr6:41208713-41217947	ENSG00000184106.7
360	FOXP4-AS1	chr6:41494853-41548621	ENSG00000234753.4
361	RP3-475N16.1	chr6:42940364-42948360	ENSG00000231113.2
362	RP11-480N24.3	chr6:43310231-43310357	ENSG00000219222.1
363	RP3-337H4.9	chr6:43519180-43519724	ENSG00000271754.1
364	RP3-337H4.6	chr6:43538822-43539703	ENSG00000219470.1
365	RP5-1120P11.1	chr6:43995723-44074652	ENSG00000237686.5
366	RP5-1120P11.3	chr6:44073913-44077952	ENSG00000231881.1
367	RP1-152L7.5	chr6:52665148-52669155	ENSG00000216775.2
368	RP11-59D5__B.2	chr6:63806836-63822642	ENSG00000236345.1
369	RP11-134K13.2	chr6:70608234-70609334	ENSG00000232389.1
370	PRDX2P4	chr6:100136064-100136663	ENSG00000213177.3
371	RP1-191J18.66	chr6:107957413-107959986	ENSG00000272476.1
372	RPL3P7	chr6:108004357-108005568	ENSG00000225093.1
373	RP5-1112D6.7	chr6:111297126-111298510	ENSG00000271789.1
374	RP11-346D19.1	chr6:113531118-113543793	ENSG00000236347.1
375	RP3-399L15.3	chr6:113969701-114471705	ENSG00000228624.6
376	RP3-486I3.4	chr6:116258493-116259115	ENSG00000233558.1
377	CBX3P9	chr6:116453014-116453565	ENSG00000217241.1
378	RPL5P18	chr6:127362513-127363403	ENSG00000220744.1
379	SNORD101	chr6:132815307-132815379	ENSG00000206754.1
380	SNORD100	chr6:132816802-132816877	ENSG00000221500.1
381	SNORA33	chr6:132817219-132817348	ENSG00000200534.1
382	RP3-325F22.5	chr6:136550661-136552554	ENSG00000272189.1
383	RP11-95M15.2	chr6:137705423-137707201	ENSG00000220412.1
384	RP11-390P2.4	chr6:138692548-138697288	ENSG00000225177.4
385	RP3-468K18.6	chr6:143858052-143858689	ENSG00000270890.1
386	RP11-350J20.12	chr6:149852462-149853192	ENSG00000273132.1
387	RAET1K	chr6:149998019-150005157	ENSG00000218358.2
388	RP1-276N6.2	chr6:160272617-160276130	ENSG00000230234.1
389	LPAL2	chr6:160453428-160511124	ENSG00000213071.9
390	RP11-351J23.1	chr6:167679626-167683787	ENSG00000203688.4
391	RP11-417E7.1	chr6:169158092-169162924	ENSG00000223485.1
392	RP11-417E7.2	chr6:169175304-169182740	ENSG00000261039.2
393	RP1-266L20.2	chr6:169725091-169725854	ENSG00000232640.1
394	AC091729.9	chr7:1160374-1165267	ENSG00000229043.2
395	ELFN1-AS1	chr7:1738630-1742291	ENSG00000236081.1
396	RNF216P1	chr7:4973988-5040675	ENSG00000196204.10
397	AC093620.5	chr7:5419827-5420767	ENSG00000241269.1
398	AC092171.2	chr7:5419846-5423122	ENSG00000188365.3
399	RP11-1275H24.1	chr7:5426277-5428927	ENSG00000234432.4
400	AC006483.5	chr7:5560743-5561055	ENSG00000228974.5

Figure 40H

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
401	PMS2CL	chr7:6710128-6753862	ENSG00000187953.9
402	RP5-1159O4.1	chr7:7552462-7566996	ENSG00000272894.4
403	AC007128.1	chr7:8262264-8344516	ENSG00000229970.3
404	RP4-594A5.1	chr7:8303741-8341343	ENSG00000272328.1
405	AC073333.8	chr7:16695871-16719898	ENSG00000235837.1
406	RP11-511H23.2	chr7:17940503-17942922	ENSG00000279048.1
407	PCMTD1P3	chr7:23721311-23721782	ENSG00000234800.2
408	CTD-2227E11.1	chr7:25948657-25949403	ENSG00000270933.1
409	RP1-170O19.17	chr7:27239243-27241228	ENSG00000253308.2
410	EVX1-AS	chr7:27241429-27247229	ENSG00000253405.1
411	AC004593.3	chr7:29199540-29208970	ENSG00000235669.1
412	AC018641.7	chr7:32456963-32457758	ENSG00000226468.2
413	DPY19L1P1	chr7:32580949-32761787	ENSG00000229358.3
414	RP9P	chr7:32916815-32943176	ENSG00000205763.12
415	AC009262.2	chr7:34209526-34256343	ENSG00000236212.1
416	NPSR1-AS1	chr7:34346512-34871582	ENSG00000197085.10
417	NCAPD2P1	chr7:34750253-34752053	ENSG00000230356.1
418	RP11-609L3.1	chr7:35971147-35971567	ENSG00000270889.1
419	AC011290.5	chr7:39569376-39570198	ENSG00000236015.1
420	LINC00265	chr7:39733632-39793092	ENSG00000188185.10
421	INHBA-AS1	chr7:41693916-41779388	ENSG00000224116.5
422	AC004453.8	chr7:44467897-44468304	ENSG00000146677.7
423	SNHG15	chr7:44983023-44986961	ENSG00000232956.7
424	RP4-725G10.3	chr7:56163145-56163533	ENSG00000230191.1
425	RP11-700P18.1	chr7:56288230-56291425	ENSG00000223559.1
426	CCT6P3	chr7:65038354-65074713	ENSG00000234585.5
427	LINC00174	chr7:66376044-66401338	ENSG00000179406.6
428	GS1-124K5.2	chr7:66409143-66490059	ENSG00000230189.5
429	GS1-124K5.4	chr7:66493705-66495474	ENSG00000237310.1
430	RP11-458F8.4	chr7:66902857-66906297	ENSG00000273142.1
431	NSUN5P2	chr7:72947581-72954790	ENSG00000106133.16
432	CTA-315H11.2	chr7:73609262-73611502	ENSG00000274080.1
433	NSUN5P1	chr7:75410322-75416787	ENSG00000223705.8
434	PMS2P3	chr7:75502930-75528148	ENSG00000127957.15
435	AC004980.7	chr7:76549360-76627982	ENSG00000205485.12
436	STEAP2-AS1	chr7:89882353-90211635	ENSG00000227646.6
437	OR7E38P	chr7:97966090-97967074	ENSG00000183444.10
438	AC004967.7	chr7:97969005-97972254	ENSG00000243554.1
439	MYH16	chr7:99238794-99311130	ENSG00000002079.11
440	AZGP1P1	chr7:99980762-99987535	ENSG00000214313.7
441	MIR25	chr7:100093560-100093643	ENSG00000207547.1
442	PMS2P1	chr7:100320992-100341908	ENSG00000078319.8
443	STAG3L5P	chr7:100336079-100351900	ENSG00000242294.5
444	STAG3L5P-PVRIG2P-PILRB	chr7:100336104-100367831	ENSG00000272752.4
445	RP11-126L15.4	chr7:100837314-100852616	ENSG00000236305.1
446	AZGP1P2	chr7:101287482-101289771	ENSG00000214252.4
447	RP11-132A1.4	chr7:101308346-101310985	ENSG00000232445.1
448	RP11-401L13.4	chr7:103141349-103141939	ENSG00000224415.1
449	S100A11P1	chr7:103262000-103262311	ENSG00000237632.3
450	CTA-276O3.4	chr7:103348601-103348771	ENSG00000235354.1

Figure 40I

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
451	RP4-778K6.1	chr7:105244652-105244970	ENSG00000213361.2
452	RP5-884M6.1	chr7:106775011-106795564	ENSG00000228742.8
453	EIF3IP1	chr7:109959218-109960184	ENSG00000237064.1
454	ST7-OT4	chr7:116953899-117098806	ENSG00000214188.8
455	RP11-3L10.3	chr7:121440834-121441261	ENSG00000234985.2
456	FEZF1-AS1	chr7:122303658-122310077	ENSG00000230316.5
457	RP11-332K15.1	chr7:123456629-123459856	ENSG00000232524.1
458	AC000123.3	chr7:127349833-127350293	ENSG00000279265.1
459	RP11-20L24.1	chr7:127857852-127858488	ENSG00000240579.1
460	RP11-448A19.1	chr7:129604548-129611630	ENSG00000273329.1
461	RP11-2E11.9	chr7:130507660-130508282	ENSG00000270953.1
462	AC058791.1	chr7:130876809-130913310	ENSG00000226380.6
463	LINC-PINT	chr7:130941760-131110176	ENSG00000231721.5
464	AC009362.2	chr7:131242590-131242995	ENSG00000233287.1
465	RP11-634H22.1	chr7:139359032-139359566	ENSG00000273391.1
466	NDUFB2-AS1	chr7:140695336-140697077	ENSG00000240889.1
467	RP5-894A10.2	chr7:141652381-141656810	ENSG00000244701.1
468	RP4-800G7.2	chr7:149285281-149297312	ENSG00000244560.5
469	ZNF767P	chr7:149547154-149624752	ENSG00000133624.12
470	RP11-148K1.12	chr7:151074742-151076530	ENSG00000244151.1
471	PAXIP1-AS1	chr7:155003448-155005703	ENSG00000273344.1
472	AC144652.1	chr7:155295918-155297541	ENSG00000273117.1
473	AC009403.2	chr7:155611231-155645205	ENSG00000216895.7
474	AC021218.2	chr7:155962632-155966343	ENSG00000204876.4
475	AC005534.8	chr7:156654185-156657693	ENSG00000224903.1
476	MNX1-AS1	chr7:157010805-157016426	ENSG00000243479.3
477	PAICSP4	chr8:4787332-4788584	ENSG00000254244.1
478	RP11-10A14.9	chr8:9158063-9158621	ENSG00000279949.1
479	RP11-10A14.5	chr8:9189011-9202854	ENSG00000248538.5
480	RP11-589N15.2	chr8:11846154-11846391	ENSG00000269899.1
481	RP11-875O11.3	chr8:23071377-23074488	ENSG00000253616.4
482	RP11-1149O23.3	chr8:23224471-23230926	ENSG00000246582.2
483	EXTL3-AS1	chr8:28696198-28701464	ENSG00000246339.5
484	RP11-527N22.2	chr8:37405439-37406724	ENSG00000253746.1
485	RP11-150O12.1	chr8:37421341-37554183	ENSG00000253161.4
486	RP11-150O12.6	chr8:37516410-37517021	ENSG00000253414.2
487	RP11-150O12.3	chr8:37597480-37599858	ENSG00000254290.1
488	RP11-546K22.1	chr8:51961458-52022974	ENSG00000253844.1
489	RP11-1112C15.1	chr8:57978358-57984126	ENSG00000205293.3
490	PTTG3P	chr8:66767400-66768005	ENSG00000213005.3
491	SNHG6	chr8:66921684-66926398	ENSG00000245910.7
492	RP11-152C15.1	chr8:70103798-70104179	ENSG00000253173.3
493	RP11-434I12.3	chr8:73358913-73362788	ENSG00000253339.1
494	RP13-923O23.7	chr8:81696368-81698694	ENSG00000280193.1
495	RP11-219B4.3	chr8:85177522-85178150	ENSG00000254208.1
496	RP11-37B2.1	chr8:89609409-89757727	ENSG00000251136.7
497	RBM12B-AS1	chr8:93740121-93740773	ENSG00000279331.1
498	RP3-388N13.5	chr8:93915734-93916682	ENSG00000264448.3
499	KB-1208A12.3	chr8:98041726-98044121	ENSG00000245970.2
500	GAPDHP62	chr8:100550724-100551713	ENSG00000251013.1

Figure 40J

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
501	NACAP1	chr8:101361794-101372707	ENSG00000228224.3
502	TAGLN2P1	chr8:106697427-106698013	ENSG00000253676.1
503	RP11-85I17.2	chr8:119838736-119840385	ENSG00000279347.1
504	CTC-820M8.1	chr8:120635558-120638204	ENSG00000250290.1
505	RP11-713M15.2	chr8:120812219-120813359	ENSG00000272502.1
506	RNF139-AS1	chr8:124462485-124474576	ENSG00000245149.3
507	PCAT1	chr8:126847055-127021014	ENSG00000253438.2
508	CASC19	chr8:127185635-127203222	ENSG00000254166.1
509	CASC8	chr8:127289817-127482139	ENSG00000246228.5
510	CCAT2	chr8:127400399-127402150	ENSG00000280997.1
511	PVT1	chr8:127794533-128101253	ENSG00000249859.6
512	TMEM75	chr8:127946559-127948723	ENSG00000280055.1
513	RNU1-106P	chr8:127999131-127999294	ENSG00000207110.1
514	RP11-629O1.2	chr8:133573183-133573861	ENSG00000261220.2
515	RP11-642A1.2	chr8:140572142-140572812	ENSG00000279766.1
516	ERICD	chr8:140636281-140638283	ENSG00000280303.2
517	RP11-10J21.4	chr8:141252286-141253292	ENSG00000253307.1
518	CTD-3064M3.4	chr8:141389939-141392574	ENSG00000244998.1
519	CTD-3064M3.7	chr8:141434545-141437954	ENSG00000271959.1
520	RP13-582O9.5	chr8:143280161-143281690	ENSG00000253716.4
521	RHPN1-AS1	chr8:143366631-143368548	ENSG00000254389.3
522	RP11-661A12.9	chr8:143573490-143577397	ENSG00000255050.1
523	RP11-661A12.8	chr8:143578358-143583304	ENSG00000279605.1
524	RP11-429J17.5	chr8:143718246-143718891	ENSG00000254548.1
525	RP11-429J17.7	chr8:143758153-143771822	ENSG00000254973.1
526	CTD-3065J16.6	chr8:144049129-144051522	ENSG00000204791.7
527	CTD-3065J16.9	chr8:144078002-144079265	ENSG00000255224.1
528	CTD-2517M22.14	chr8:144495458-144505444	ENSG00000255182.2
529	RP11-307I14.2	chr9:4781475-4782078	ENSG00000220105.2
530	RPL4P5	chr9:7477045-7478320	ENSG00000230207.1
531	RP11-109M17.2	chr9:10948372-10948481	ENSG00000270372.1
532	AKAP8P1	chr9:11011748-11013096	ENSG00000234146.1
533	LDHAP4	chr9:14921337-14922334	ENSG00000214110.3
534	RP11-146N23.1	chr9:19200335-19201046	ENSG00000232176.1
535	RP11-378I6.1	chr9:19926094-19929937	ENSG00000261402.1
536	IFNWP19	chr9:21455484-21456049	ENSG00000238271.2
537	MIR31HG	chr9:21455642-21559669	ENSG00000171889.3
538	RP11-70L8.4	chr9:21858910-21861926	ENSG00000265194.1
539	RBMXP2	chr9:30689105-30690272	ENSG00000215210.3
540	SLC25A6P2	chr9:31253901-31254777	ENSG00000232203.3
541	TCEA1P4	chr9:32979560-32980403	ENSG00000236184.1
542	LAGE3P1	chr9:33019682-33020165	ENSG00000225693.1
543	RP11-571F15.3	chr9:34223984-34224628	ENSG00000233668.1
544	RP11-112J3.16	chr9:35772163-35790432	ENSG00000227388.2
545	RP11-613M10.6	chr9:37509150-37510299	ENSG00000234160.1
546	FKBP4P6	chr9:42384805-42386136	ENSG00000268234.1
547	CNN2P4	chr9:42972385-42973289	ENSG00000213816.3
548	RPL35AP21	chr9:71589601-71589933	ENSG00000229814.1
549	RP11-82L18.2	chr9:89088604-89109934	ENSG00000224945.1
550	RP13-93L13.2	chr9:89449558-89450754	ENSG00000231799.2

Figure 40K

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
551	RP11-435O5.4	chr9:95514045-95514520	ENSG00000271659.1
552	RPS2P35	chr9:100116755-100117651	ENSG00000238172.2
553	RP11-220D10.1	chr9:108626833-108627150	ENSG00000169253.3
554	YBX1P6	chr9:109532830-109534332	ENSG00000213539.4
555	RP11-356B19.11	chr9:127366770-127367397	ENSG00000271833.1
556	SNORA65	chr9:127448501-127448630	ENSG00000201302.1
557	PTGES2-AS1	chr9:128128529-128130628	ENSG00000232850.3
558	LINC00963	chr9:129483451-129513683	ENSG00000204054.10
559	RP11-492E3.2	chr9:129575409-129584556	ENSG00000227619.1
560	RP11-426A6.5	chr9:135503273-135506447	ENSG00000226706.1
561	C9orf163	chr9:136483495-136486067	ENSG00000196366.3
562	SNHG7	chr9:136721366-136728184	ENSG00000233016.5
563	RP11-216L13.21	chr9:136728953-136729855	ENSG00000274356.1
564	RP11-216L13.19	chr9:136799223-136810042	ENSG00000273066.4
565	RP11-229P13.15	chr9:137052662-137053375	ENSG00000236394.2
566	RP13-463N16.6	chr10:5594991-5596118	ENSG00000242147.1
567	RP11-536K7.3	chr10:5934270-5945900	ENSG00000232807.2
568	RP11-186N15.3	chr10:12244751-12247845	ENSG00000228302.2
569	STAM-AS1	chr10:17641284-17643878	ENSG00000260589.1
570	RP11-305E6.4	chr10:30302826-30306066	ENSG00000259994.1
571	HSD17B7P2	chr10:38356380-38378505	ENSG00000099251.13
572	ZNF37BP	chr10:42513510-42552822	ENSG00000234420.6
573	RP11-351D16.3	chr10:43136824-43138334	ENSG00000273008.1
574	RP11-445N18.3	chr10:45073146-45073541	ENSG00000228462.1
575	PARPG1	chr10:45854093-45972154	ENSG00000239883.7
576	RP11-170M17.1	chr10:63664664-63990568	ENSG00000228566.1
577	RP11-170M17.2	chr10:63902451-63903245	ENSG00000213609.3
578	RP11-9E13.4	chr10:68424560-68424997	ENSG00000234806.1
579	RP11-227H15.5	chr10:69265342-69268148	ENSG00000231748.1
580	RP11-152N13.16	chr10:73124573-73125532	ENSG00000272599.2
581	RP11-574K11.24	chr10:73742995-73744230	ENSG00000279088.1
582	LINC00858	chr10:84279980-84294659	ENSG00000229404.4
583	RP11-77P6.2	chr10:86521945-86525101	ENSG00000227896.2
584	RP11-380G5.3	chr10:87945502-87946024	ENSG00000213613.2
585	NHP2P1	chr10:92216332-92216793	ENSG00000105988.6
586	RP11-108L7.15	chr10:101060029-101061005	ENSG00000273162.1
587	RP11-190J1.3	chr10:101704652-101708401	ENSG00000224817.1
588	RPARP-AS1	chr10:102449817-102461106	ENSG00000269609.4
589	SH3PXD2A-AS1	chr10:103745966-103755423	ENSG00000280693.1
590	ADD3-AS1	chr10:109940104-110008381	ENSG00000203876.8
591	CTAGE7P	chr10:130106046-130108481	ENSG00000233122.2
592	RP11-326C3.2	chr11:287305-288987	ENSG00000255026.1
593	RP11-326C3.13	chr11:325703-326294	ENSG00000270030.1
594	RP11-496I9.1	chr11:557595-560107	ENSG00000254815.4
595	AP006621.5	chr11:777578-784297	ENSG00000255284.1
596	AC068580.7	chr11:1776930-1778388	ENSG00000230834.1
597	H19	chr11:1995163-2001470	ENSG00000130600.14
598	KCNQ1OT1	chr11:2608328-2699994	ENSG00000269821.1
599	SNORA45B	chr11:8685439-8685569	ENSG00000212607.1
600	CTD-2371O3.3	chr11:9459556-9460702	ENSG00000268403.2

Figure 40L

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
601	RP11-466H18.1	chr11:16974693-16975013	ENSG00000244398.1
602	RP1-239B22.5	chr11:17380649-17383531	ENSG00000260196.1
603	BBOX1-AS1	chr11:27047186-27220113	ENSG00000254560.4
604	HSP90AA2P	chr11:27888838-27891033	ENSG00000224411.3
605	RP1-89D4.1	chr11:33237008-33237409	ENSG00000241255.1
606	RP1-68D18.4	chr11:35210343-35214985	ENSG00000255443.1
607	AP001258.4	chr11:59130133-59143015	ENSG00000245571.5
608	AP000442.1	chr11:59616429-59639861	ENSG00000255139.1
609	RP11-794G24.1	chr11:61539516-61542259	ENSG00000256443.1
610	RPLP0P2	chr11:61615036-61639449	ENSG00000243742.4
611	RP11-727F15.9	chr11:62832234-62834043	ENSG00000256690.1
612	SNHG1	chr11:62851988-62855914	ENSG00000255717.5
613	RP11-783K16.5	chr11:64245964-64248217	ENSG00000256940.1
614	RP11-783K16.13	chr11:64246939-64249494	ENSG00000257086.1
615	MALAT1	chr11:65497762-65506516	ENSG00000251562.6
616	SSSCA1-AS1	chr11:65568482-65570423	ENSG00000260233.3
617	RP11-658F2.3	chr11:66761575-66762399	ENSG00000213409.4
618	RPL37P2	chr11:67682772-67683058	ENSG00000239559.2
619	FAM86C2P	chr11:67791648-67805336	ENSG00000160172.9
620	IFITM9P	chr11:69303412-69303807	ENSG00000213275.2
621	RP11-211G23.2	chr11:69371463-69372511	ENSG00000260877.1
622	AP000487.5	chr11:70372246-70398488	ENSG00000246889.2
623	RP11-660L16.2	chr11:71448674-71452157	ENSG00000254682.1
624	SNORD15B	chr11:75404421-75404566	ENSG00000207445.1
625	RP11-535A19.1	chr11:75800877-75803415	ENSG00000254814.1
626	RP11-727A23.5	chr11:83184491-83193794	ENSG00000247137.7
627	GAPDHP70	chr11:88408179-88409223	ENSG00000249489.1
628	SCARNA9	chr11:93721513-93721865	ENSG00000254911.3
629	RP11-178H8.7	chr11:93726654-93729805	ENSG00000279696.1
630	RP11-732A21.3	chr11:102109827-102110457	ENSG00000277459.1
631	RP11-817J15.2	chr11:102606916-102628070	ENSG00000256916.1
632	RP11-832N8.1	chr11:107908420-107908764	ENSG00000240652.1
633	RPL37AP8	chr11:111889199-111889474	ENSG00000176343.5
634	SNORD14E	chr11:123058077-123058161	ENSG00000200879.1
635	RP11-598F7.5	chr12:164664-166321	ENSG00000256694.1
636	RP11-598F7.6	chr12:166856-182399	ENSG00000256540.1
637	RP4-816N1.7	chr12:2797136-2803938	ENSG00000258092.1
638	RP11-253E3.3	chr12:3041437-3044950	ENSG00000250899.3
639	DSTNP2	chr12:6884682-6885786	ENSG00000248593.3
640	SCARNA12	chr12:6967337-6967606	ENSG00000238795.1
641	C1RL-AS1	chr12:7108052-7122501	ENSG00000205885.6
642	RP11-273B20.1	chr12:7129079-7131198	ENSG00000256967.1
643	PTMAP4	chr12:9239986-9240331	ENSG00000231503.4
644	RP11-22B23.1	chr12:9277235-9313241	ENSG00000111788.10
645	DDX12P	chr12:9418673-9448229	ENSG00000214826.5
646	RP11-267J23.4	chr12:12111163-12111489	ENSG00000198134.3
647	RP11-180M15.7	chr12:12718973-12719521	ENSG00000275560.1
648	RP11-392P7.6	chr12:12927726-12984645	ENSG00000247498.8
649	PDCD5P1	chr12:19413153-19413461	ENSG00000255909.1
650	RP11-424C20.2	chr12:20551590-20553012	ENSG00000256663.1

Figure 40M

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
651	ARNTL2-AS1	chr12:27389789-27446625	ENSG00000245311.2
652	LINC00941	chr12:30795681-30802711	ENSG00000235884.3
653	DDX11-AS1	chr12:31020763-31073847	ENSG00000245614.3
654	RP11-843B15.4	chr12:32109076-32109602	ENSG00000277342.1
655	RP11-817I4.1	chr12:32339368-32340724	ENSG00000274964.1
656	RP11-817I4.2	chr12:32352349-32354144	ENSG00000276115.1
657	RP11-278C7.1	chr12:32726383-32727387	ENSG00000257511.1
658	RP11-351C21.2	chr12:42361267-42361703	ENSG00000274943.1
659	PLEKHA8P1	chr12:45173054-45216041	ENSG00000134297.6
660	RP11-493L12.3	chr12:47237734-47279021	ENSG00000257925.1
661	RP11-493L12.6	chr12:47341614-47342089	ENSG00000258116.1
662	RP1-90J4.1	chr12:47415008-47420179	ENSG00000257906.1
663	RP1-228P16.1	chr12:48054813-48055591	ENSG00000240399.1
664	RP11-386G11.5	chr12:48998367-49019235	ENSG00000257913.2
665	RP11-386G11.10	chr12:49127782-49147869	ENSG00000258017.1
666	RP11-133N21.7	chr12:49595148-49595688	ENSG00000257243.1
667	RP11-469H8.6	chr12:49951512-49962924	ENSG00000257588.1
668	LINC00592	chr12:52210930-52223804	ENSG00000258279.2
669	RP11-680A11.5	chr12:53298655-53300314	ENSG00000257605.2
670	RP11-968A15.2	chr12:54262615-54279063	ENSG00000257596.1
671	RP11-153M3.1	chr12:56511002-56512703	ENSG00000257576.1
672	AGAP2-AS1	chr12:57726271-57728356	ENSG00000255737.2
673	RP11-631N16.4	chr12:62603909-62604399	ENSG00000275180.1
674	RPSAP52	chr12:65758020-65826997	ENSG00000241749.4
675	RP11-366L20.2	chr12:65851340-65882167	ENSG00000197301.6
676	RP11-1143G9.5	chr12:69326574-69331882	ENSG00000274979.1
677	RP11-785D18.3	chr12:70570969-70571440	ENSG00000277247.1
678	RP11-578B16.1	chr12:71448405-71448850	ENSG00000277223.1
679	RP11-186F10.2	chr12:71582293-71583852	ENSG00000257761.1
680	RP11-81H3.2	chr12:74133166-74402535	ENSG00000251138.5
681	RP11-585P4.5	chr12:75483454-75489820	ENSG00000257497.2
682	RP11-114H23.1	chr12:75563202-75984015	ENSG00000258077.2
683	RP11-114H23.3	chr12:75688794-75689096	ENSG00000258245.1
684	SNORA70	chr12:75689152-75689304	ENSG00000251893.2
685	RP11-114H23.2	chr12:75694010-75698816	ENSG00000258088.1
686	RP11-290L1.5	chr12:75964440-75967062	ENSG00000257329.1
687	RP11-290L1.3	chr12:76030494-76031378	ENSG00000257453.1
688	RP11-54A9.1	chr12:76259839-76305131	ENSG00000257219.4
689	RP11-390N6.1	chr12:78960258-79045644	ENSG00000257191.1
690	RP11-734K2.4	chr12:89525654-89548005	ENSG00000270344.2
691	TMPO-AS1	chr12:98512973-98516422	ENSG00000257167.2
692	HELLPAR	chr12:102197585-102402596	ENSG00000281344.1
693	FAM222A-AS1	chr12:109734166-109773508	ENSG00000255650.4
694	RP11-686G8.1	chr12:111581379-111581773	ENSG00000238168.5
695	LINC01234	chr12:113679459-113773683	ENSG00000249550.5
696	RP11-103B5.4	chr12:117002463-117003152	ENSG00000277840.1
697	NME2P1	chr12:120282303-120282716	ENSG00000123009.4
698	DYNLL1-AS1	chr12:120490328-120495940	ENSG00000248008.2
699	RP11-728G15.1	chr12:120629511-120641591	ENSG00000256008.2
700	RP13-942N8.1	chr12:123363868-123366113	ENSG00000256092.2

Figure 40N

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
701	RP11-972P1.10	chr12:123515275-123515513	ENSG00000274427.1
702	RP11-972P1.11	chr12:123519390-123519856	ENSG00000278112.1
703	RP11-486O12.2	chr12:123575891-123585115	ENSG00000247373.3
704	RP11-158L12.4	chr12:125138245-125141711	ENSG00000279233.1
705	RP11-474D1.4	chr12:130024493-130044956	ENSG00000261650.1
706	RP11-474D1.3	chr12:130033454-130045057	ENSG00000214039.7
707	RP11-474D1.1	chr12:130047132-130048376	ENSG00000256298.1
708	RP11-143E21.2	chr12:130121925-130122974	ENSG00000279193.1
709	RP11-143E21.3	chr12:130138693-130140768	ENSG00000276122.1
710	RP11-143E21.6	chr12:130249579-130266725	ENSG00000259862.1
711	RP11-200K12.3	chr12:130350996-130351707	ENSG00000279343.1
712	RP11-417L19.6	chr12:131934642-131934928	ENSG00000273568.1
713	EP400NL	chr12:132084283-132131639	ENSG00000185684.11
714	RPS11P5	chr12:132825701-132826184	ENSG00000232888.4
715	MRPL3P1	chr13:19304593-19305625	ENSG00000215349.2
716	RP11-172H24.4	chr13:20699307-20703718	ENSG00000278291.1
717	RP11-261P13.5	chr13:20712431-20714146	ENSG00000279135.1
718	RP11-261P13.6	chr13:20768876-20769375	ENSG00000275485.1
719	MIPEPP3	chr13:21298139-21306373	ENSG00000233325.3
720	ZDHHC20-IT1	chr13:21376977-21377874	ENSG00000236953.1
721	ANKRD20A19P	chr13:23939557-23946782	ENSG00000196593.8
722	IPO7P2	chr13:24122065-24123814	ENSG00000225674.1
723	PDX1-AS1	chr13:27833444-27921404	ENSG00000247381.2
724	ATP5EP2	chr13:27945206-27945590	ENSG00000180389.6
725	PAN3-AS1	chr13:28136843-28138193	ENSG00000261485.1
726	RP11-142E9.1	chr13:33846190-33850825	ENSG00000276672.1
727	MIR621	chr13:40810766-40810861	ENSG00000207652.1
728	LINC01050	chr13:42810366-42812562	ENSG00000271216.1
729	DGKZP1	chr13:43968424-43971199	ENSG00000179611.3
730	SNORA31	chr13:45337480-45337609	ENSG00000199477.1
731	COX4I1P2	chr13:45671845-45672391	ENSG00000228869.1
732	NAP1L4P3	chr13:47763460-47764606	ENSG00000234145.1
733	DLEU2	chr13:49982552-50125720	ENSG00000231607.7
734	RP11-34F20.7	chr13:50125816-50128463	ENSG00000274270.1
735	MRPS31P5	chr13:52167709-52194465	ENSG00000243406.5
736	RP11-93H24.3	chr13:52651305-52652279	ENSG00000273723.1
737	LMO7-AS1	chr13:75604700-75635994	ENSG00000261105.4
738	RP11-173B14.4	chr13:75631271-75632135	ENSG00000228444.2
739	RBM26-AS1	chr13:79406309-79424328	ENSG00000227354.5
740	PTMAP5	chr13:81689911-81691072	ENSG00000214182.5
741	MIR17HG	chr13:91347820-91354579	ENSG00000215417.9
742	CLDN10-AS1	chr13:95479444-95533910	ENSG00000223392.1
743	DNAJC3-AS1	chr13:95648733-95676925	ENSG00000247400.3
744	UBAC2-AS1	chr13:99196377-99200710	ENSG00000228889.5
745	H2AFZP3	chr13:99215372-99215758	ENSG00000218502.3
746	LINC01232	chr13:99486962-99499306	ENSG00000280734.1
747	COX5BP6	chr13:100623875-100624429	ENSG00000237082.1
748	LINC00460	chr13:106376563-106378217	ENSG00000233532.4
749	RP11-54H7.4	chr13:109269534-109273838	ENSG00000275216.1
750	LINC00346	chr13:110863987-110870251	ENSG00000255874.2

Figure 40O

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
751	RP11-88E10.5	chr13:112686769-112689815	ENSG00000274922.1
752	MCF2L-AS1	chr13:112967484-112968824	ENSG00000235280.2
753	GAS6-AS1	chr13:113815630-113845744	ENSG00000233695.2
754	DUXAP10	chr14:19062316-19115270	ENSG00000225210.4
755	LINC01296	chr14:19284653-19337730	ENSG00000244306.8
756	PRMT5-AS1	chr14:22918947-22926900	ENSG00000237054.8
757	Y_RNA	chr14:31244761-31244861	ENSG00000202402.1
758	RP11-159D23.2	chr14:31925801-31930146	ENSG00000257472.1
759	RPS19P3	chr14:34568689-34569129	ENSG00000240463.1
760	HNRNPUP1	chr14:43300263-43300595	ENSG00000259051.1
761	RP11-326E7.1	chr14:49373966-49374383	ENSG00000258401.1
762	AL163953.2	chr14:53599832-53614026	ENSG00000225680.1
763	AL162759.1	chr14:53768942-53850882	ENSG00000235269.1
764	KTN1-AS1	chr14:55499278-55580110	ENSG00000186615.9
765	RPL7AP4	chr14:55543401-55544169	ENSG00000213641.4
766	RP11-307P22.1	chr14:60779978-60780595	ENSG00000258445.3
767	HSPE1P2	chr14:63601509-63601814	ENSG00000258645.2
768	RP11-544I20.2	chr14:64329431-64338599	ENSG00000214770.2
769	PTBP1P	chr14:65279267-65281166	ENSG00000259078.2
770	RP11-72M17.1	chr14:66212810-66509394	ENSG00000258561.1
771	MAGOH3P	chr14:68861762-68862525	ENSG00000270975.1
772	RP6-65G23.3	chr14:70810205-70815403	ENSG00000259153.1
773	RP1-146I3.1	chr14:72929612-72929930	ENSG00000176043.5
774	CTD-3035D6.1	chr14:90766531-90766989	ENSG00000213315.5
775	RP11-529H20.3	chr14:92026566-92026887	ENSG00000259020.3
776	DICER1-AS1	chr14:95157645-95179933	ENSG00000235706.6
777	SNHG10	chr14:95532297-95534872	ENSG00000247092.5
778	SCARNA13	chr14:95533355-95533629	ENSG00000252481.1
779	LINC00618	chr14:96943579-96945394	ENSG00000225163.3
780	RP11-1029J19.4	chr14:101628984-101632530	ENSG00000258919.1
781	RP11-1029J19.5	chr14:101634454-101731108	ENSG00000258404.1
782	RP11-73M18.8	chr14:103696353-103697163	ENSG00000269958.1
783	RP11-521B24.4	chr14:105370418-105372918	ENSG00000279495.1
784	RPS20P33	chr14:105374618-105374971	ENSG00000242085.1
785	ARHGAP11B	chr15:30624494-30772993	ENSG00000187951.9
786	RP11-1008C21.1	chr15:37984766-38062340	ENSG00000259225.5
787	C15orf54	chr15:39250684-39254845	ENSG00000175746.6
788	FAM92A1P1	chr15:41163162-41164374	ENSG00000157021.9
789	RP11-23P13.6	chr15:41892793-41898575	ENSG00000174171.5
790	SORD2P	chr15:44826371-44884694	ENSG00000259479.5
791	AHCYP7	chr15:50417385-50418692	ENSG00000259465.2
792	RP11-108K3.1	chr15:51037488-51293912	ENSG00000259240.1
793	RP11-108K3.2	chr15:51315841-51321996	ENSG00000259306.2
794	CTD-2184D3.1	chr15:52095295-52095747	ENSG00000242327.1
795	LINC01169	chr15:66582190-66685794	ENSG00000259471.1
796	RPS24P16	chr15:67232446-67232837	ENSG00000242634.1
797	RP11-253M7.1	chr15:69396904-69415029	ENSG00000259426.4
798	RP11-106M3.3	chr15:72278867-72351794	ENSG00000261460.1
799	ANP32BP1	chr15:75321397-75322693	ENSG00000259790.1
800	CTD-2562G15.3	chr15:75452964-75453947	ENSG00000276744.1

Figure 40P

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
801	SCARNA15	chr15:82752884-82757208	ENSG00000252690.3
802	GOLGA2P7	chr15:84199311-84230136	ENSG00000225151.9
803	HMGB1P8	chr15:89135547-89136495	ENSG00000259699.2
804	RP11-661P17.1	chr15:91408708-91605211	ENSG00000258551.4
805	CTD-2147F2.1	chr15:97295881-97432094	ENSG00000259485.1
806	CTD-2147F2.2	chr15:97370371-97521811	ENSG00000259664.2
807	RP11-20G13.3	chr15:99395179-99396593	ENSG00000259590.1
808	WASIR2	chr16:22910-25123	ENSG00000231439.4
809	Z97634.3	chr16:382097-392960	ENSG00000236829.8
810	LA16c-313D11.12	chr16:689001-692554	ENSG00000261659.2
811	LA16c-306A4.1	chr16:883780-885090	ENSG00000260022.1
812	TPSP2	chr16:1286351-1288837	ENSG00000260284.1
813	RP11-616M22.7	chr16:1294551-1299166	ENSG00000260710.1
814	LA16c-358B7.3	chr16:1317891-1322845	ENSG00000261505.1
815	LA16c-390E6.4	chr16:1451760-1452653	ENSG00000260051.1
816	LA16c-380F5.3	chr16:1553655-1554130	ENSG00000280231.1
817	LA16c-431H6.7	chr16:1688355-1690536	ENSG00000278987.1
818	LA16c-329F2.2	chr16:1707252-1707973	ENSG00000275092.1
819	AC005363.11	chr16:1971655-1971896	ENSG00000277602.1
820	RP11-304L19.1	chr16:2091436-2095433	ENSG00000259933.5
821	RP11-304L19.3	chr16:2094830-2097026	ENSG00000261123.1
822	SNORD60	chr16:2155023-2155105	ENSG00000206630.1
823	AC141586.5	chr16:2603350-2630494	ENSG00000215154.5
824	PRSS30P	chr16:2839568-2842744	ENSG00000172460.13
825	LA16c-325D7.1	chr16:2857898-2859726	ENSG00000263325.1
826	MMP25-AS1	chr16:3051096-3059370	ENSG00000261971.5
827	RP11-473M20.9	chr16:3076911-3087100	ENSG00000262370.4
828	ZNF213-AS1	chr16:3116042-3134882	ENSG00000263072.4
829	LA16c-360H6.1	chr16:3292879-3293403	ENSG00000279031.1
830	RP11-461A8.4	chr16:3650636-3651703	ENSG00000263235.1
831	LINC01569	chr16:4245825-4253789	ENSG00000262468.4
832	RP11-295D4.1	chr16:4335870-4337818	ENSG00000262712.1
833	RP11-295D4.3	chr16:4346694-4348648	ENSG00000280063.1
834	RP11-77H9.2	chr16:8848105-8860456	ENSG00000260276.2
835	RP11-485G7.6	chr16:11348143-11349321	ENSG00000262703.1
836	RP11-166B2.3	chr16:11881075-11882569	ENSG00000261560.1
837	NPIPP1	chr16:15104312-15123498	ENSG00000188599.16
838	CTB-193M12.5	chr16:15701237-15702118	ENSG00000280206.1
839	CRYM-AS1	chr16:21300849-21318591	ENSG00000189149.10
840	RRN3P1	chr16:21796106-21820410	ENSG00000248124.6
841	RP11-645C24.4	chr16:21938865-21945130	ENSG00000257408.1
842	CTD-2649C14.2	chr16:21950218-21951708	ENSG00000274460.1
843	CTD-2196E14.6	chr16:23568673-23569696	ENSG00000260751.2
844	CTD-2547G23.4	chr16:25257952-25261066	ENSG00000274925.1
845	AC002331.1	chr16:26584755-26594813	ENSG00000236481.1
846	RP11-57A19.2	chr16:28284885-28292064	ENSG00000246465.1
847	RP11-1348G14.8	chr16:28822431-28823969	ENSG00000275807.1
848	ATP2A1-AS1	chr16:28878957-28879920	ENSG00000260442.4
849	RP11-264B17.4	chr16:28973962-28978824	ENSG00000260367.2
850	RP11-426C22.4	chr16:29217170-29220031	ENSG00000259807.1

Figure 40Q

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
851	SLC7A5P1	chr16:29613104-29613640	ENSG00000260727.1
852	CDIPT-AS1	chr16:29863593-29868053	ENSG00000214725.6
853	FBXL19-AS1	chr16:30919319-30923269	ENSG00000260852.1
854	RP11-1072A3.3	chr16:30984630-30988270	ENSG00000279196.1
855	CLUHP3	chr16:31700590-31711986	ENSG00000131797.11
856	RP11-401P9.1	chr16:50551824-50553940	ENSG00000260029.2
857	RP11-401P9.7	chr16:50555355-50558719	ENSG00000278909.1
858	CRNDE	chr16:54918863-54929189	ENSG00000245694.7
859	RP11-322D14.2	chr16:57052505-57058497	ENSG00000260145.1
860	CTD-2600O9.1	chr16:57798186-57816946	ENSG00000187185.4
861	LINC00922	chr16:65284499-65576300	ENSG00000261742.4
862	LINC00920	chr16:66408524-66412135	ENSG00000246898.1
863	RP11-403P17.6	chr16:66565620-66566001	ENSG00000277978.1
864	RP11-615I2.2	chr16:68644248-68646168	ENSG00000260577.2
865	RP11-615I2.3	chr16:68651056-68651948	ENSG00000260860.2
866	HSPE1P5	chr16:68686399-68686670	ENSG00000261395.3
867	CTD-2033A16.3	chr16:69727013-69742563	ENSG00000262136.1
868	CTD-2033A16.2	chr16:69756536-69757937	ENSG00000260290.2
869	SMG1P7	chr16:70219574-70245616	ENSG00000261556.7
870	VAC14-AS1	chr16:70755098-70773251	ENSG00000214353.6
871	LINC01572	chr16:72283301-72665009	ENSG00000261008.5
872	RP5-991G20.1	chr16:72665170-72791624	ENSG00000259768.5
873	RPL18P13	chr16:76235055-76235623	ENSG00000244485.1
874	RP11-303E16.2	chr16:81030770-81031485	ENSG00000261061.1
875	RP11-568J23.1	chr16:85815873-85816536	ENSG00000214259.3
876	CTD-2015G9.2	chr16:86722091-86741059	ENSG00000261175.4
877	RP11-106D4.2	chr16:87470370-87474370	ENSG00000276337.1
878	RP11-863P13.3	chr16:88177298-88178941	ENSG00000261327.4
879	RP5-1142A6.9	chr16:88718615-88720459	ENSG00000260121.1
880	AC137932.4	chr16:89268104-89273044	ENSG00000268218.1
881	VPS9D1-AS1	chr16:89711856-89718165	ENSG00000261373.1
882	RP11-566K11.7	chr16:89906157-89918233	ENSG00000267048.1
883	AFG3L1P	chr16:89972586-90002161	ENSG00000223959.7
884	URAHP	chr16:90039761-90047773	ENSG00000222019.6
885	RP11-235E17.6	chr17:3655621-3658092	ENSG00000262903.1
886	RYKP1	chr17:4222091-4223828	ENSG00000263219.1
887	AC118754.4	chr17:4497394-4499674	ENSG00000229782.1
888	RP11-141J13.5	chr17:4610449-4617650	ENSG00000261863.1
889	CTC-524C5.2	chr17:5425139-5432876	ENSG00000263272.1
890	RP11-186B7.4	chr17:7572706-7582024	ENSG00000264772.4
891	RPL29P2	chr17:7754320-7754976	ENSG00000240480.1
892	RP11-849F2.9	chr17:8277763-8278436	ENSG00000269947.1
893	UBE2SP1	chr17:15704232-15704900	ENSG00000233966.1
894	RP11-416I2.1	chr17:17011914-17014990	ENSG00000260328.1
895	RPL13P12	chr17:17383377-17384012	ENSG00000215030.5
896	UBE2SP2	chr17:18677261-18677757	ENSG00000224126.2
897	SNORD3B-2	chr17:19063346-19064136	ENSG00000262074.5
898	SCDP1	chr17:20784645-20785725	ENSG00000226549.3
899	RP11-663N22.1	chr17:27237859-27241661	ENSG00000265246.1
900	RPL34P31	chr17:27274107-27274477	ENSG00000239223.3

Figure 40R

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
901	RP11-173M1.5	chr17:27351858-27352584	ENSG00000265683.1
902	RPS7P1	chr17:28467822-28468406	ENSG00000263266.2
903	AC010761.8	chr17:28721487-28722877	ENSG00000264577.1
904	RP11-68I3.7	chr17:29591703-29592241	ENSG00000264647.1
905	SUZ12P1	chr17:30709299-30790908	ENSG00000264538.5
906	CTD-2349P21.5	chr17:30729469-30731202	ENSG00000263603.1
907	RP1-41C23.4	chr17:31560132-31560573	ENSG00000278546.1
908	CTD-2095E4.3	chr17:32159311-32160371	ENSG00000280033.1
909	RP11-749I16.3	chr17:40113215-40115442	ENSG00000279199.1
910	RP11-58O9.2	chr17:40473554-40475289	ENSG00000279806.1
911	AC003958.2	chr17:41402416-41425049	ENSG00000234859.4
912	KRT41P	chr17:41406512-41411775	ENSG00000225438.1
913	RP11-156E6.1	chr17:41848518-41851447	ENSG00000259623.1
914	RP11-242D8.1	chr17:43164183-43170403	ENSG00000267002.1
915	RP13-890H12.2	chr17:45168800-45171584	ENSG00000267288.2
916	CRHR1-IT1	chr17:45620328-45648216	ENSG00000204650.12
917	SP2-AS1	chr17:47897330-47941410	ENSG00000234494.6
918	RP11-501C14.9	chr17:48899131-48899748	ENSG00000270781.1
919	RP11-481C4.2	chr17:51183191-51186403	ENSG00000279089.1
920	RP11-159D12.2	chr17:57989039-57994850	ENSG00000264112.1
921	BZRAP1-AS1	chr17:58325450-58415766	ENSG00000265148.4
922	SETP3	chr17:58476138-58476950	ENSG00000266648.1
923	CTD-2510F5.4	chr17:59202677-59203829	ENSG00000265415.1
924	RP11-329L6.2	chr17:64145970-64146476	ENSG00000266402.3
925	SNORD104	chr17:64146083-64146152	ENSG00000199753.1
926	RP13-104F24.2	chr17:64749663-64781707	ENSG00000215769.7
927	RP13-104F24.3	chr17:64750420-64751311	ENSG00000265298.1
928	RP11-927P21.4	chr17:64778842-64778937	ENSG00000265982.1
929	PLEKHM1P	chr17:64779259-64837154	ENSG00000214176.8
930	CTD-2535L24.3	chr17:65457541-65458408	ENSG00000263520.1
931	RP11-57A1.1	chr17:72425939-72428852	ENSG00000263680.2
932	AC011933.2	chr17:75338827-75339147	ENSG00000228007.1
933	RP11-552F3.10	chr17:75897060-75900148	ENSG00000267342.1
934	SNHG16	chr17:76557766-76565348	ENSG00000163597.13
935	SNHG20	chr17:77086716-77094990	ENSG00000234912.8
936	EIF5AP2	chr17:78157403-78159135	ENSG00000267679.1
937	RP11-559N14.5	chr17:78484882-78503056	ENSG00000267432.4
938	CTD-2357A8.3	chr17:78617389-78632057	ENSG00000267123.4
939	RP11-353N14.1	chr17:79800598-79802529	ENSG00000262768.2
940	RP11-353N14.2	chr17:79823452-79827704	ENSG00000262772.1
941	RP11-353N14.5	chr17:79915252-79926725	ENSG00000262585.1
942	RP11-353N14.4	chr17:79919357-79925462	ENSG00000262188.1
943	RP11-353N14.7	chr17:79952663-79952992	ENSG00000275516.1
944	CTD-2529O21.1	chr17:79991944-79992841	ENSG00000262172.1
945	RP11-334C17.3	chr17:80147250-80148596	ENSG00000279259.1
946	RP11-334C17.5	chr17:80200673-80205949	ENSG00000262580.4
947	CTD-2047H16.2	chr17:80315651-80316633	ENSG00000262979.1
948	AC027601.1	chr17:81228707-81233983	ENSG00000260005.5
949	TMEM105	chr17:81311270-81330674	ENSG00000185332.5
950	RP11-1055B8.1	chr17:81395609-81397144	ENSG00000279692.1

Figure 40S

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
951	RP11-498C9.4	chr17:81862778-81863102	ENSG00000278876.1
952	MAFG-AS1	chr17:81927829-81930753	ENSG00000265688.1
953	AC137723.5	chr17:81941859-81947601	ENSG00000235296.1
954	RP13-516M14.2	chr17:82214227-82217352	ENSG00000264548.1
955	RP13-638C3.3	chr17:82587313-82588411	ENSG00000262147.1
956	BOLA2P1	chr18:832238-832469	ENSG00000266575.1
957	RP11-672L10.5	chr18:856526-857045	ENSG00000264179.1
958	RP11-161I6.2	chr18:1883524-2240847	ENSG00000263745.4
959	DLGAP1-AS1	chr18:3593732-3598352	ENSG00000177337.6
960	DLGAP1-AS2	chr18:3603000-3608336	ENSG00000262001.1
961	RP11-874J12.4	chr18:3653410-3656282	ENSG00000266401.1
962	RP11-419J16.1	chr18:10125340-10144406	ENSG00000265554.1
963	RP11-703I16.3	chr18:12040101-12040300	ENSG00000266995.1
964	RP11-815J4.7	chr18:12067173-12068417	ENSG00000267533.1
965	FAM60BP	chr18:26255698-26256355	ENSG00000263829.1
966	DSG1-AS1	chr18:31343368-31426988	ENSG00000266729.4
967	SNHG22	chr18:49814023-49851059	ENSG00000267322.2
968	SNRPGP2	chr18:54406889-54407114	ENSG00000264350.1
969	CTD-2171N6.1	chr18:55105904-55124306	ENSG00000267013.4
970	RP11-126O1.4	chr18:58672613-58752730	ENSG00000267476.1
971	OACYLP	chr18:58996734-59069338	ENSG00000224367.5
972	NFE2L3P1	chr18:59969438-59971396	ENSG00000267497.1
973	ATP5G1P6	chr18:63496989-63497400	ENSG00000214318.3
974	AC006273.5	chr19:782755-785080	ENSG00000267530.2
975	CTB-25B13.9	chr19:1508375-1508963	ENSG00000267092.2
976	AC005256.1	chr19:1748056-1748745	ENSG00000267073.1
977	CTB-31O20.2	chr19:1874871-1876169	ENSG00000261526.2
978	AC005264.2	chr19:3141576-3155175	ENSG00000267551.3
979	CTB-50L17.5	chr19:4429689-4430934	ENSG00000267255.1
980	SNRPEP4	chr19:5576660-5576938	ENSG00000233270.1
981	CTD-3128G10.6	chr19:6748293-6751467	ENSG00000269680.1
982	CTD-3214H19.6	chr19:7633766-7636990	ENSG00000268204.1
983	CTD-3193O13.12	chr19:7898804-7903542	ENSG00000214248.2
984	C3P1	chr19:10037803-10074135	ENSG00000167798.15
985	CTD-2369P2.4	chr19:10259109-10260045	ENSG00000267105.1
986	CTD-2006C1.6	chr19:11648364-11652877	ENSG00000267500.1
987	CTD-2006C1.2	chr19:11987617-12046275	ENSG00000219665.7
988	CTD-2666L21.3	chr19:12260126-12260537	ENSG00000213293.4
989	AD000092.3	chr19:12894133-12894880	ENSG00000240616.1
990	CTD-3252C9.4	chr19:13834516-13836289	ENSG00000267519.3
991	EEF1DP1	chr19:14070342-14071237	ENSG00000228887.3
992	AC005785.2	chr19:15379076-15381194	ENSG00000268189.2
993	AC006128.2	chr19:15398696-15400356	ENSG00000279716.1
994	CTB-187L3.1	chr19:15515342-15516836	ENSG00000269765.2
995	CYP4F23P	chr19:15564074-15584709	ENSG00000269516.5
996	UCA1	chr19:15828961-15836320	ENSG00000214049.6
997	CTD-2231E14.2	chr19:16074293-16077395	ENSG00000279198.1
998	CTD-3222D19.12	chr19:16542746-16544814	ENSG00000269399.2
999	CTD-3222D19.8	chr19:16572624-16575340	ENSG00000279529.1
1000	CTD-2521M24.6	chr19:17414257-17422324	ENSG00000269481.1

Figure 40T

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
1001	CTD-3131K8.2	chr19:17488990-17511889	ENSG00000269439.4
1002	MIR3189	chr19:18386562-18386634	ENSG00000264175.1
1003	AC005253.2	chr19:18557775-18561560	ENSG00000268030.1
1004	AC079466.1	chr19:28606688-28615229	ENSG00000266976.1
1005	AC007773.2	chr19:32390050-32405560	ENSG00000267213.4
1006	CTC-523E23.3	chr19:34949038-34951205	ENSG00000280106.1
1007	CTD-2527I21.14	chr19:35014961-35025335	ENSG00000271032.1
1008	TMEM147-AS1	chr19:35540738-35546029	ENSG00000236144.5
1009	AC002116.7	chr19:36014508-36045972	ENSG00000267698.1
1010	RPL31P61	chr19:36901742-36902117	ENSG00000243297.2
1011	CTD-2540F13.2	chr19:38738284-38739863	ENSG00000267892.1
1012	IFNL3P1	chr19:39236433-39237293	ENSG00000268510.1
1013	RPL36P16	chr19:41169851-41170166	ENSG00000224476.3
1014	CTC-435M10.12	chr19:41373971-41374419	ENSG00000277744.1
1015	MIR6797	chr19:41869627-41869698	ENSG00000276926.1
1016	AC092066.1	chr19:44664131-44666158	ENSG00000279095.1
1017	AC007191.4	chr19:45714387-45717381	ENSG00000279407.1
1018	AC006262.5	chr19:46189166-46203083	ENSG00000268621.4
1019	IGFL1P1	chr19:46196238-46197323	ENSG00000268879.1
1020	CTC-453G23.5	chr19:48118432-48127706	ENSG00000269534.4
1021	AC018766.6	chr19:49852887-49854967	ENSG00000268047.1
1022	PTOV1-AS2	chr19:49856970-49859289	ENSG00000269352.1
1023	RPL37P23	chr19:52143043-52143336	ENSG00000243680.1
1024	TPM3P9	chr19:53431984-53444670	ENSG00000241015.2
1025	AC012314.8	chr19:54119511-54125343	ENSG00000237017.1
1026	AC008746.12	chr19:54438665-54439544	ENSG00000267838.2
1027	LENG8-AS1	chr19:54444813-54449045	ENSG00000226696.4
1028	CTD-2537I9.12	chr19:55661901-55674715	ENSG00000267523.1
1029	CTC-444N24.8	chr19:57261354-57262738	ENSG00000268713.1
1030	CTC-444N24.11	chr19:57304305-57308562	ENSG00000268205.1
1031	AC004076.5	chr19:57449689-57453011	ENSG00000276449.1
1032	CTD-2583A14.8	chr19:57867038-57868172	ENSG00000269867.1
1033	CTD-2583A14.11	chr19:57867885-57868834	ENSG00000270804.1
1034	AC010642.1	chr19:58278966-58315197	ENSG00000267216.1
1035	CTD-2619J13.13	chr19:58400221-58400679	ENSG00000268307.1
1036	CTD-2619J13.19	chr19:58440448-58445849	ENSG00000269473.1
1037	RP11-128M1.1	chr20:2207217-2213151	ENSG00000226644.4
1038	RP5-1009E24.8	chr20:3808357-3812434	ENSG00000275491.1
1039	LINC01433	chr20:4193065-4195943	ENSG00000230176.2
1040	LINC00658	chr20:5426895-5471094	ENSG00000226995.6
1041	RPS18P1	chr20:5532878-5533335	ENSG00000225831.1
1042	EIF4EP1	chr20:5548840-5549481	ENSG00000233578.1
1043	RP5-967N21.11	chr20:6000418-6000941	ENSG00000275632.1
1044	RP4-775C13.1	chr20:6213699-6215381	ENSG00000234241.1
1045	PA2G4P2	chr20:12380056-12381255	ENSG00000235698.1
1046	SNORD17	chr20:17962710-17962946	ENSG00000212232.1
1047	RPS19P1	chr20:18504633-18505066	ENSG00000214612.3
1048	RP1-198K11.5	chr20:21302731-21303704	ENSG00000275457.1
1049	RP3-322G13.7	chr20:23346941-23351486	ENSG00000234832.1
1050	XXyac-YX60D10.2	chr20:23772785-23776169	ENSG00000277975.1

Figure 40U

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
1051	ZNF337-AS1	chr20:25624045-25678074	ENSG00000213742.5
1052	RP4-694B14.8	chr20:25680781-25681246	ENSG00000278383.1
1053	RP4-760C5.3	chr20:26008791-26010531	ENSG00000231081.1
1054	HM13-IT1	chr20:31563166-31564076	ENSG00000235313.1
1055	ABALON	chr20:31721507-31723409	ENSG00000281376.1
1056	RP5-836N17.4	chr20:32116171-32116629	ENSG00000275576.1
1057	RP5-1085F17.3	chr20:32856621-32858751	ENSG00000260257.2
1058	RPL31P2	chr20:33752167-33752543	ENSG00000230837.1
1059	RP4-785G19.5	chr20:34234840-34281173	ENSG00000250917.1
1060	CTD-3216D2.5	chr20:34281632-34286466	ENSG00000272945.1
1061	RP11-42O4.2	chr20:35174355-35174919	ENSG00000278367.1
1062	FAM83C-AS1	chr20:35285251-35285756	ENSG00000235214.1
1063	FER1L4	chr20:35558737-35607562	ENSG00000088340.14
1064	RPL37P1	chr20:35588324-35588607	ENSG00000227401.1
1065	LINC00657	chr20:36045622-36050960	ENSG00000260032.1
1066	DLGAP4-AS1	chr20:36507702-36573391	ENSG00000232907.6
1067	RP4-564F22.5	chr20:38406011-38416797	ENSG00000224635.1
1068	SNHG17	chr20:38420588-38435353	ENSG00000196756.10
1069	SNORA71B	chr20:38425083-38425354	ENSG00000235408.4
1070	SNORA71A	chr20:38427309-38427442	ENSG00000225091.3
1071	SNORA71C	chr20:38429670-38429803	ENSG00000201512.1
1072	SNORA71D	chr20:38433865-38433998	ENSG00000200354.1
1073	SNHG11	chr20:38446578-38450921	ENSG00000174365.18
1074	SNORA60	chr20:38449370-38449503	ENSG00000199266.1
1075	RP4-616B8.5	chr20:38955910-38956547	ENSG00000274825.1
1076	RP4-616B8.6	chr20:38962472-38962993	ENSG00000277581.1
1077	RP5-881L22.5	chr20:44347552-44355185	ENSG00000226812.2
1078	RP3-453C12.14	chr20:45345115-45345823	ENSG00000275894.1
1079	RP11-465L10.10	chr20:46013500-46022073	ENSG00000204044.6
1080	RPL13P2	chr20:46099518-46100176	ENSG00000213820.3
1081	RP1-101A2.1	chr20:46681676-46682375	ENSG00000273893.1
1082	ZFAS1	chr20:49278178-49295738	ENSG00000177410.11
1083	SNORD12B	chr20:49280319-49280409	ENSG00000222365.1
1084	SNRPFP1	chr20:49729591-49729842	ENSG00000231878.1
1085	KRT18P4	chr20:49956745-49958032	ENSG00000229222.1
1086	LINC01272	chr20:50267486-50279795	ENSG00000224397.4
1087	LINC01270	chr20:50292720-50314922	ENSG00000203999.7
1088	LINC01429	chr20:51831797-51862912	ENSG00000227964.1
1089	RPS4XP3	chr20:56601701-56602113	ENSG00000214541.3
1090	RP5-1059L7.1	chr20:57710183-57712780	ENSG00000278709.1
1091	RP5-907D15.4	chr20:58634772-58635738	ENSG00000268941.2
1092	RP11-157P1.4	chr20:62305432-62306325	ENSG00000226332.2
1093	RP5-908M14.9	chr20:62386303-62386970	ENSG00000273619.1
1094	RP5-908M14.10	chr20:62402236-62405935	ENSG00000275437.1
1095	RP5-908M14.5	chr20:62427827-62447621	ENSG00000233017.2
1096	WI2-87327B8.2	chr20:62596732-62603115	ENSG00000273812.1
1097	SLCO4A1-AS1	chr20:62663019-62666724	ENSG00000232803.1
1098	LINC00659	chr20:62774128-62775412	ENSG00000228705.1
1099	RP4-563E14.1	chr20:62928621-62929297	ENSG00000273759.1
1100	RP5-963E22.6	chr20:63218041-63218502	ENSG00000273821.1

Figure 40V

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
1101	RP4-697K14.15	chr20:63543411-63543738	ENSG00000275812.1
1102	CTD-3184A7.4	chr20:63627227-63628824	ENSG00000232442.1
1103	RP4-591C20.9	chr20:63861212-63864293	ENSG00000268858.2
1104	MIR647	chr20:63942631-63942726	ENSG00000207554.1
1105	UCKL1-AS1	chr20:63953384-63956985	ENSG00000280213.1
1106	AF127577.8	chr21:14918534-14947096	ENSG00000235277.1
1107	LINC01549	chr21:17438890-17449185	ENSG00000232560.5
1108	RPL39P40	chr21:17500679-17500824	ENSG00000226580.1
1109	AP000695.4	chr21:36445731-36532408	ENSG00000233818.1
1110	AP000696.2	chr21:36632681-36637033	ENSG00000231324.1
1111	AP000697.6	chr21:36698773-36701564	ENSG00000224269.1
1112	DSCR9	chr21:37208503-37221736	ENSG00000230366.8
1113	SPATA20P1	chr21:38238227-38238664	ENSG00000231123.1
1114	AF064858.6	chr21:38863676-38956467	ENSG00000205622.8
1115	PCBP2P1	chr21:39171130-39172106	ENSG00000235701.1
1116	PLAC4	chr21:41175231-41185239	ENSG00000280109.1
1117	AP001628.6	chr21:42831040-42836477	ENSG00000225218.1
1118	AP001631.9	chr21:43140523-43141092	ENSG00000236663.1
1119	TRPM2-AS	chr21:44414588-44425272	ENSG00000230061.2
1120	TSPEAR-AS2	chr21:44517216-44525952	ENSG00000182912.6
1121	MCM3AP-AS1	chr21:46229217-46259390	ENSG00000215424.8
1122	AP001469.9	chr21:46251549-46254133	ENSG00000239415.1
1123	DUXAP8	chr22:15784959-15829984	ENSG00000206195.9
1124	LL22NC03-N14H11.1	chr22:15823197-15823890	ENSG00000272872.1
1125	AC004471.9	chr22:19121529-19124503	ENSG00000223461.1
1126	LINC01311	chr22:19171395-19172839	ENSG00000260924.2
1127	AC000067.1	chr22:19667023-19667555	ENSG00000225007.1
1128	LINC00896	chr22:20206397-20208524	ENSG00000236499.2
1129	TMEM191A	chr22:20701114-20704606	ENSG00000226287.6
1130	KB-1440D3.13	chr22:21661934-21662363	ENSG00000272954.1
1131	FBXW4P1	chr22:23262767-23265005	ENSG00000230701.2
1132	CRYBB2P1	chr22:25448105-25520854	ENSG00000100058.11
1133	CTA-445C9.14	chr22:26512537-26514568	ENSG00000261188.1
1134	RP1-40G4P.1	chr22:26886909-26887548	ENSG00000231369.1
1135	TTC28-AS1	chr22:27919376-28008581	ENSG00000235954.5
1136	RP1-102K2.8	chr22:30246205-30246998	ENSG00000268812.3
1137	CTA-963H5.5	chr22:30653877-30654814	ENSG00000250318.1
1138	RP1-90G24.10	chr22:32205115-32269666	ENSG00000242082.1
1139	CPSF1P1	chr22:32269381-32273110	ENSG00000214076.3
1140	RP5-1014D13.2	chr22:37876148-37895563	ENSG00000279738.1
1141	CTA-228A9.3	chr22:38090127-38091559	ENSG00000272720.1
1142	RP5-821D11.7	chr22:41831215-41834665	ENSG00000184068.2
1143	CYP2D7	chr22:42140203-42144577	ENSG00000205702.9
1144	CYP2D8P	chr22:42149886-42155001	ENSG00000226450.2
1145	OGFRP1	chr22:42269753-42275196	ENSG00000182057.4
1146	LINC01315	chr22:42364400-42369236	ENSG00000229891.1
1147	CTA-126B4.7	chr22:42438023-42446195	ENSG00000230107.1
1148	RRP7B	chr22:42555223-42582038	ENSG00000182841.11
1149	CTA-414D7.1	chr22:44102779-44116232	ENSG00000279175.1
1150	NUP50-AS1	chr22:45133020-45163781	ENSG00000226328.5

Figure 40W

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
1151	CTA-217C2.2	chr22:45155539-45156011	ENSG00000273243.1
1152	CTA-941F9.10	chr22:45657019-45680130	ENSG00000280383.1
1153	MIRLET7BHG	chr22:46053859-46113928	ENSG00000197182.11
1154	RP3-439F8.1	chr22:46541495-46548196	ENSG00000234869.1
1155	CTA-29F11.1	chr22:46761894-46762563	ENSG00000260708.1
1156	CTA-384D8.36	chr22:50523926-50524780	ENSG00000272821.1
1157	MT-TM	chrM:4402-4469	ENSG00000210112.1
1158	LINC00106	chrX:1396427-1399402	ENSG00000236871.5
1159	ASMTL-AS1	chrX:1401769-1414028	ENSG00000236017.6
1160	CD99P1	chrX:2609348-2657229	ENSG00000223773.5
1161	RP11-459A10.1	chrX:6422092-6422701	ENSG00000237730.1
1162	RP11-120D5.1	chrX:10847877-11111141	ENSG00000234129.6
1163	CA5BP1	chrX:15674932-15703724	ENSG00000186312.9
1164	snoU2_19	chrX:20136306-20136385	ENSG00000201592.1
1165	RP11-298C3.2	chrX:25645722-25646327	ENSG00000228519.3
1166	HMGA1P1	chrX:26748029-26749090	ENSG00000236683.3
1167	RP11-320G24.1	chrX:40262917-40287720	ENSG00000236393.1
1168	RP6-218J18.2	chrX:44029332-44030921	ENSG00000214019.2
1169	FDPSP5	chrX:44476027-44477422	ENSG00000215035.2
1170	RP5-1158E12.3	chrX:45848074-45851490	ENSG00000231566.1
1171	LINC01186	chrX:46325924-46327645	ENSG00000236751.1
1172	GAPDHP65	chrX:46439709-46440714	ENSG00000235587.2
1173	AF196972.3	chrX:48433201-48441619	ENSG00000235350.1
1174	RP11-1148L6.9	chrX:48568014-48574860	ENSG00000204620.3
1175	LL0XNC01-7P3.1	chrX:49273054-49275768	ENSG00000270012.1
1176	RP11-485B17.5	chrX:52448587-52451138	ENSG00000250084.1
1177	RP11-3D23.1	chrX:62779250-62780326	ENSG00000236852.1
1178	HNRNPDP1	chrX:64044305-64045058	ENSG00000232794.1
1179	RPS23P8	chrX:70962964-70963293	ENSG00000230629.2
1180	AL590762.11	chrX:71420083-71420514	ENSG00000213018.2
1181	SEPHS1P4	chrX:73769248-73770423	ENSG00000230146.1
1182	DDX3P1	chrX:74121012-74131923	ENSG00000232928.2
1183	AC004386.4	chrX:74376125-74376885	ENSG00000213269.2
1184	PABPC1P3	chrX:74583088-74583546	ENSG00000230673.3
1185	STIP1P3	chrX:86084716-86086339	ENSG00000225536.2
1186	HNRNPA1P26	chrX:100855288-100856379	ENSG00000239345.2
1187	HNRNPA1P27	chrX:100887636-100888590	ENSG00000233680.4
1188	LL0XNC01-131B10.2	chrX:100910712-100911489	ENSG00000179031.8
1189	DNAJA1P3	chrX:107351650-107352843	ENSG00000215007.3
1190	TDGF1P3	chrX:110520312-110523021	ENSG00000225366.4
1191	SNORA69	chrX:119787353-119787484	ENSG00000206622.1
1192	PA2G4P1	chrX:120868611-120869807	ENSG00000237828.1
1193	RP5-1052M9.1	chrX:124326175-124326258	ENSG00000226532.1
1194	FIRRE	chrX:131702650-131830643	ENSG00000213468.3
1195	FAM195CP	chrX:131795657-131796167	ENSG00000241418.1
1196	MIR503HG	chrX:134543337-134546632	ENSG00000223749.6
1197	RP11-1114A5.4	chrX:136840931-136847797	ENSG00000232611.1
1198	RP11-308D16.2	chrX:136909395-137021630	ENSG00000234062.6
1199	RP11-332E4.1	chrX:137324972-137325049	ENSG00000214748.2
1200	RP13-150K15.1	chrX:145630218-145632375	ENSG00000231093.1
1201	LINC00894	chrX:149938628-150224580	ENSG00000235703.4
1202	RP11-49C9.2	chrX:150114378-150116520	ENSG00000183171.5
1203	XX-FW83563B9.5	chrX:154396878-154398816	ENSG00000280195.1

Figure 40X

CRC Exon RNA sequence markers

Order	Exon	*Gene*
1	chr1:53711212-53712728:-	*LRP8*
2	chr1:92511089-92511217:+	*EPHX4*
3	chr1:92515874-92515977:+	*EPHX4*
4	chr1:92518067-92518215:+	*EPHX4*
5	chr1:92528612-92529093:+	*EPHX4*
6	chr1:150956757-150956870:+	*ANXA9*
7	chr1:150957062-150957152:+	*ANXA9*
8	chr1:150958812-150958891:+	*ANXA9*
9	chr1:150960348-150960432:+	*ANXA9*
10	chr1:150960567-150960662:+	*ANXA9*
11	chr1:150960759-150960817:+	*ANXA9*
12	chr2:113332424-113334727:+	*POLR1B*
13	chr2:174130778-174132736:+	*ZAK*
14	chr2:196599607-196602425:+	*SLC39A10*
15	chr3:190023503-190026228:-	*CLDN1*
16	chr3:190027958-190028042:-	*CLDN1*
17	chr3:190030661-190030825:-	*CLDN1*
18	chr3:190039773-190040215:-	*CLDN1*
19	chr4:57259531-57261714:-	*PPAT*
20	chr5:54273696-54275263:-	*ESM1*
21	chr5:54277825-54277974:-	*ESM1*
22	chr5:54281045-54281414:-	*ESM1*
23	chr5:73923235-73925817:-	*ENC1*
24	chr5:135398875-135399506:+	*TGFBI*
25	chr5:176882725-176883283:+	*PRR7*
26	chr6:1312675-1314992:+	*FOXQ1*
27	chr6:41617357-41617581:+	*MDFI*
28	chr6:41621057-41621981:+	*MDFI*
29	chr6:44220782-44221115:+	*HSP90AB1*
30	chr6:44221226-44221614:+	*HSP90AB1*
31	chr6:151203898-151204022:+	*MTHFD1L*
32	chr6:151206770-151206870:+	*MTHFD1L*
33	chr6:151208978-151209117:+	*MTHFD1L*
34	chr6:151247258-151247431:+	*MTHFD1L*
35	chr6:151281411-151281551:+	*MTHFD1L*
36	chr6:151330955-151331094:+	*MTHFD1L*
37	chr6:151336016-151336116:+	*MTHFD1L*
38	chr6:151336652-151336829:+	*MTHFD1L*
39	chr6:151355629-151355736:+	*MTHFD1L*
40	chr6:151358101-151358253:+	*MTHFD1L*
41	chr6:151413603-151413723:+	*MTHFD1L*
42	chr6:151422662-151423020:+	*MTHFD1L*
43	chr7:24836165-24839898:-	*OSBPL3*
44	chr7:24846395-24846521:-	*OSBPL3*
45	chr7:24849426-24849570:-	*OSBPL3*

Order	Exon	*Gene*
46	chr7:26217563-26217742:+	*NFE2L3*
47	chr7:26223321-26223404:+	*NFE2L3*
48	chr7:26224153-26226754:+	*NFE2L3*
49	chr7:36492086-36493398:+	*ANLN*
50	chr7:44608502-44608604:-	*DDX56*
51	chr7:56130706-56131682:+	*CCT6A*
52	chr7:73933739-73934049:+	*GTF2IRD1*
53	chr7:73935538-73935627:+	*GTF2IRD1*
54	chr7:73938401-73938484:+	*GTF2IRD1*
55	chr7:73944064-73944247:+	*GTF2IRD1*
56	chr7:73950497-73950605:+	*GTF2IRD1*
57	chr7:73953010-73953090:+	*GTF2IRD1*
58	chr7:73954209-73954298:+	*GTF2IRD1*
59	chr7:73959085-73959132:+	*GTF2IRD1*
60	chr7:73960074-73960139:+	*GTF2IRD1*
61	chr7:73961433-73961616:+	*GTF2IRD1*
62	chr7:73969504-73969553:+	*GTF2IRD1*
63	chr7:73969723-73969824:+	*GTF2IRD1*
64	chr7:73971969-73972052:+	*GTF2IRD1*
65	chr7:73973191-73973374:+	*GTF2IRD1*
66	chr7:73973957-73973985:+	*GTF2IRD1*
67	chr7:74004180-74004257:+	*GTF2IRD1*
68	chr7:74005154-74005346:+	*GTF2IRD1*
69	chr7:74009341-74009378:+	*GTF2IRD1*
70	chr7:74015371-74016918:+	*GTF2IRD1*
71	chr7:102957277-102957461:-	*DNAJC2*
72	chr7:105096962-105098373:-	*PUS7*
73	chr7:116435941-116438439:+	*MET*
74	chr7:138516129-138522905:-	*KIAA1549*
75	chr7:138524878-138525023:-	*KIAA1549*
76	chr7:138529062-138529219:-	*KIAA1549*
77	chr7:138545885-138546202:-	*KIAA1549*
78	chr7:138552721-138552874:-	*KIAA1549*
79	chr7:138554284-138554507:-	*KIAA1549*
80	chr7:138874067-138874550:+	*TTC26*
81	chr8:128902835-128903244:+	*PVT1*
82	chr8:129001408-129001537:+	*PVT1*
83	chr8:129108764-129108900:+	*PVT1*
84	chr8:129113226-129113498:+	*PVT1*
85	chr8:144826030-144828507:+	*LOC100128338*
86	chr10:1063015-1063704:+	*GTPBP4*
87	chr10:102752947-102754158:+	*C10orf2*
88	chr11:45945705-45945828:+	*GYLTL1B*
89	chr11:45946057-45946228:+	*GYLTL1B*
90	chr11:45946336-45946440:+	*GYLTL1B*

Figure 41A

Order	Exon	*Gene*
91	chr11:45947590-45947702:+	*GYLTL1B*
92	chr11:45947773-45947898:+	*GYLTL1B*
93	chr11:45947914-45948148:+	*GYLTL1B*
94	chr11:45948262-45948422:+	*GYLTL1B*
95	chr11:45948866-45949144:+	*GYLTL1B*
96	chr11:45949471-45949620:+	*GYLTL1B*
97	chr11:45949728-45949923:+	*GYLTL1B*
98	chr11:45950181-45950647:+	*GYLTL1B*
99	chr11:60702724-60704630:+	*TMEM132A*
100	chr11:66452721-66453911:-	*SPTBN2*
101	chr11:66454465-66454638:-	*SPTBN2*
102	chr11:66454898-66455118:-	*SPTBN2*
103	chr11:66455475-66455550:-	*SPTBN2*
104	chr11:66455640-66455782:-	*SPTBN2*
105	chr11:66456124-66456320:-	*SPTBN2*
106	chr11:66456567-66456651:-	*SPTBN2*
107	chr11:66457276-66457414:-	*SPTBN2*
108	chr11:66457510-66457754:-	*SPTBN2*
109	chr11:66458755-66459129:-	*SPTBN2*
110	chr11:66460007-66460211:-	*SPTBN2*
111	chr11:66460441-66460571:-	*SPTBN2*
112	chr11:66460657-66460917:-	*SPTBN2*
113	chr11:66461610-66461834:-	*SPTBN2*
114	chr11:66463748-66464011:-	*SPTBN2*
115	chr12:3131010-3131183:+	*TEAD4*
116	chr12:3147134-3147274:+	*TEAD4*
117	chr12:3149273-3149425:+	*TEAD4*
118	chr12:3149531-3149841:+	*TEAD4*
119	chr12:52562781-52565341:-	*KRT80*
120	chr12:52565442-52565497:-	*KRT80*
121	chr12:52566822-52566947:-	*KRT80*
122	chr12:52567384-52567548:-	*KRT80*
123	chr12:52574297-52574747:-	*KRT80*
124	chr12:52579163-52579533:-	*KRT80*
125	chr12:52585387-52585784:-	*KRT80*
126	chr12:63037763-63042416:-	*PPM1H*
127	chr12:64841885-64842461:+	*XPOT*
128	chr13:28498393-28500451:+	*PDX1*
129	chr14:23440412-23442777:-	*AJUBA*
130	chr14:23443256-23443324:-	*AJUBA*
131	chr14:23444048-23444099:-	*AJUBA*
132	chr14:23444183-23444313:-	*AJUBA*
133	chr14:23445665-23445727:-	*AJUBA*
134	chr15:81166205-81166314:+	*KIAA1199*
135	chr15:81171062-81171208:+	*KIAA1199*

Order	Exon	*Gene*
136	chr15:81172057-81172195:+	*KIAA1199*
137	chr15:81173241-81173477:+	*KIAA1199*
138	chr15:81176516-81176695:+	*KIAA1199*
139	chr15:81180035-81180105:+	*KIAA1199*
140	chr15:81181042-81181137:+	*KIAA1199*
141	chr15:81181812-81181933:+	*KIAA1199*
142	chr15:81187331-81187463:+	*KIAA1199*
143	chr15:81188210-81188401:+	*KIAA1199*
144	chr15:81199004-81199179:+	*KIAA1199*
145	chr15:81201438-81201647:+	*KIAA1199*
146	chr15:81212435-81212640:+	*KIAA1199*
147	chr15:81213373-81213442:+	*KIAA1199*
148	chr15:81214350-81214478:+	*KIAA1199*
149	chr15:81216962-81217047:+	*KIAA1199*
150	chr15:81217965-81218096:+	*KIAA1199*
151	chr15:81221243-81221278:+	*KIAA1199*
152	chr15:81221360-81221515:+	*KIAA1199*
153	chr15:81224200-81224380:+	*KIAA1199*
154	chr15:81225586-81225801:+	*KIAA1199*
155	chr15:81229015-81229226:+	*KIAA1199*
156	chr15:81230135-81230320:+	*KIAA1199*
157	chr15:81234190-81234394:+	*KIAA1199*
158	chr15:81234592-81234678:+	*KIAA1199*
159	chr15:81235286-81235443:+	*KIAA1199*
160	chr15:81239306-81239406:+	*KIAA1199*
161	chr15:81241138-81243999:+	*KIAA1199*
162	chr16:4312537-4312702:-	*TFAP4*
163	chr16:67132613-67134956:+	*CBFB*
164	chr16:68678151-68679327:+	*CDH3*
165	chr16:68679539-68679653:+	*CDH3*
166	chr16:68710288-68710373:+	*CDH3*
167	chr16:68712037-68712180:+	*CDH3*
168	chr16:68712404-68712559:+	*CDH3*
169	chr16:68712665-68712809:+	*CDH3*
170	chr16:68713702-68713877:+	*CDH3*
171	chr16:68714871-68714999:+	*CDH3*
172	chr16:68716205-68716390:+	*CDH3*
173	chr16:68718486-68718727:+	*CDH3*
174	chr16:68719108-68719253:+	*CDH3*
175	chr16:68721415-68721639:+	*CDH3*
176	chr16:68725623-68725829:+	*CDH3*
177	chr16:68729158-68729288:+	*CDH3*
178	chr16:68729680-68729826:+	*CDH3*
179	chr16:68732094-68732957:+	*CDH3*
180	chr16:87863630-87866631:-	*SLC7A5*

Figure 41B

Order	Exon	*Gene*
181	chr16:89643947-89644129:+	*CPNE7*
182	chr16:89651872-89651961:+	*CPNE7*
183	chr16:89652095-89652193:+	*CPNE7*
184	chr16:89653078-89653164:+	*CPNE7*
185	chr16:89653451-89653510:+	*CPNE7*
186	chr16:89656305-89656359:+	*CPNE7*
187	chr16:89657415-89657466:+	*CPNE7*
188	chr17:36827961-36831187:-	*C17orf96*
189	chr17:41605212-41606111:-	*ETV4*
190	chr17:41606503-41606604:-	*ETV4*
191	chr17:41606872-41607044:-	*ETV4*
192	chr17:41607252-41607320:-	*ETV4*
193	chr17:41607475-41607802:-	*ETV4*
194	chr17:41610042-41610307:-	*ETV4*
195	chr17:41610555-41610716:-	*ETV4*
196	chr17:41611227-41611353:-	*ETV4*
197	chr17:41613794-41613847:-	*ETV4*
198	chr17:41622343-41622390:-	*ETV4*
199	chr17:41622642-41622754:-	*ETV4*
200	chr17:41622926-41623081:-	*ETV4*
201	chr17:77768176-77769357:-	*CBX8*
202	chr17:79887352-79888628:+	*MAFG-AS1*
203	chr19:7963546-7966908:+	*LRRC8E*
204	chr19:48907991-48908610:+	*GRIN2D*
205	chr19:48917248-48917362:+	*GRIN2D*
206	chr19:48917630-48917841:+	*GRIN2D*
207	chr19:48918121-48918289:+	*GRIN2D*
208	chr19:48919259-48919412:+	*GRIN2D*
209	chr19:48922491-48922616:+	*GRIN2D*
210	chr19:48922842-48923071:+	*GRIN2D*
211	chr19:48925042-48925202:+	*GRIN2D*
212	chr19:48945026-48945213:+	*GRIN2D*
213	chr19:48945407-48945639:+	*GRIN2D*
214	chr19:48945857-48948187:+	*GRIN2D*
215	chr20:368655-368945:+	*TRIB3*
216	chr20:371931-372223:+	*TRIB3*
217	chr20:376842-378201:+	*TRIB3*
218	chr20:43570772-43571854:-	*TOMM34*
219	chr20:43572094-43572220:-	*TOMM34*
220	chr20:47711269-47711500:+	*CSE1L*
221	chr20:61287711-61288602:+	*SLCO4A1*
222	chr20:61290029-61290119:+	*SLCO4A1*
223	chr20:61291764-61291885:+	*SLCO4A1*
224	chr20:61292416-61292527:+	*SLCO4A1*
225	chr20:61296276-61296849:-	*LOC100127888*

Order	Exon	*Gene*
226	chr20:61296286-61296440:+	*SLCO4A1*
227	chr20:61297732-61297927:+	*SLCO4A1*
228	chr20:61297737-61297973:-	*LOC100127888*
229	chr20:61299097-61299262:+	*SLCO4A1*
230	chr20:61299364-61299536:+	*SLCO4A1*
231	chr20:61299829-61299893:+	*SLCO4A1*
232	chr20:61300282-61300430:+	*SLCO4A1*
233	chr20:61303102-61303647:+	*SLCO4A1*
234	chr20:61429939-61430020:+	*C20orf20*
235	chr20:61430808-61431943:+	*C20orf20*
236	chr21:38071991-38072221:+	*SIM2*
237	chr21:38084833-38084922:+	*SIM2*
238	chr21:38092122-38092230:+	*SIM2*
239	chr21:38095346-38095431:+	*SIM2*
240	chr21:38098420-38098619:+	*SIM2*
241	chr21:38103346-38103452:+	*SIM2*
242	chr21:38114018-38114165:+	*SIM2*
243	chr21:38115688-38115856:+	*SIM2*
244	chr21:38117029-38118218:+	*SIM2*
245	chr22:24125598-24126502:+	*MMP11*
246	chrX:118722307-118725272:-	*NKRF*
247	chrX:154003470-154003548:+	*DKC1*
248	chrX:154004462-154004599:+	*DKC1*
249	chrX:154005074-154005963:+	*DKC1*

Figure 41C

CRC mRNA protein encoding markers

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
1	ACAN	15q26.1	aggrecan	NP_037359	E7EX88|E7ENV9
2	ACTR3B	7q36.1	ARP3 actin-related protein 3 homolog B (yeast)	NP_065178	Q9P1U1|Q59GD5
3	ADAM12	10q26	ADAM metallopeptidase domain 12	NP_067673	O43184|U5NE98|A8K6G4
4	ADAMTS6	5q13	ADAM metallopeptidase with thrombospondin type 1 motif, 6	NP_922932	Q9UKP5|Q5IR90
5	AMPD2	1p13.3	adenosine monophosphate deaminase 2	NP_981949	Q01433|A0A024R098|B2RB47
6	ANKRD13B	17q11.2	ankyrin repeat domain 13B	NP_689558	Q86YJ7|A0A024QZ29|B3KS27
7	ANLN	7p15-p14	anillin, actin binding protein	NP_061155	Q9NQW6|A0A024RA49|A8K5D9
8	ANXA4	2p13	annexin A4	NP_001144	P09525|Q6LES2
9	APLN	Xq25	Apelin	NP_059109	Q9ULZ1
10	AREG	4q13.3	amphiregulin	NP_001648	P15514
11	ARNTL2	12p12.2-p11.2	aryl hydrocarbon receptor nuclear translocator-like 2	NP_064568	Q8WYA1
12	ASPHD1	16p11.2	aspartate beta-hydroxylase domain containing 1	NP_859069	Q5U4P2
13	ATAD2	8q24.13	ATPase family, AAA domain containing 2	NP_054828	Q6PL18|A0A024R9G7
14	ATG9B	7q36	ATG9 autophagy related 9 homolog B (S. cerevisiae)	NP_775952	Q674R7
15	ATIC	2q35	5-aminoimidazole-4-carboxamide ribonucleotide formyltransferase/IMP cyclohydrolase	NP_004035	P31939|V9HWH7
16	ATP11A	13q34	ATPase, class VI, type 11A	NP_115565	P98196|Q6PJ25|E9PEJ6|Q659C3
17	BAG4	8p11.23	BCL2-associated athanogene 4	NP_004865	O95429
18	BFSP1	20p12.1	beaded filament structural protein 1, filensin	NP_001265537	Q12934|B7ZAD2|B7Z999
19	BIRC5	17q25.3	baculoviral inhibitor of apoptosis repeat-containing 5 or survivin	NP_001159	O15392|A0A0B4J1S3|H3BLT4
20	BYSL	6p21.1	bystin-like	NP_004044	Q13895
21	C10orf2	10q24	chromosome 10 open reading frame 2	NP_068602	Q96RR1|Q9H6V3|E5KSY5
22	C11orf84	11q13.1	chromosome 11 open reading frame 84	NP_612480	Q9BUA3
23	C12orf11 or INTS13	12p11.23	integrator complex subunit 13		
24	C16orf46	16q23.2	chromosome 16 open reading frame 46	NP_689550	Q6P387
25	C17orf96	17q12	chromosome 17 open reading frame 96	NP_001124149	A6NHQ4
26	C19orf28 or MFSD12	19p13.3	major facilitator superfamily domain containing 12	NP_778148	Q6NUT3
27	C19orf48	19q13.33	chromosome 19 open reading frame 48	NP_954858	Q6RUI8|A0A024R4G9
28	C1orf135 or AUNIP	1p36.11	aurora kinase A and ninein interacting protein	NP_076942	Q9H7T9
29	C1orf170	1p36.33	chromosome 1 open reading frame 170		
30	C1R	12p13	complement component 1, r subcomponent	NP_001724	P00736
31	C20orf20 or MARGBP	20q13.33	MRG domain binding protein		

Figure 42A

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
32	C2CD4A	15q22.2	C2 calcium-dependent domain containing 4A	NP_997205	Q8NCU7
33	C2orf70	2p23.3	chromosome 2 open reading frame 70	NP_001098989	A6NJV1
34	C5orf34	5p12	chromosome 5 open reading frame 34	NP_940968	Q96MH7
35	C6orf223	6p21.1	chromosome 6 open reading frame 223	NP_694978	Q8N319
36	C7orf40 or SNHG15	7p13	small nucleolar RNA host gene 15		
37	C7orf68 or HILPDA	7q32.1	hypoxia inducible lipid droplet associated	NP_037464	Q9Y5L2
38	CBFB	16q22.1	core-binding factor, beta subunit	NP_074036	Q13951|A0A024R6X2
39	CBX2	17q25.3	chromobox homolog 2		
40	CBX3	7p15.2	chromobox homolog 3	NP_057671	Q13185|A4D177|B8ZZ43
41	CBX4	17q25.3	chromobox homolog 4	NP_003646	O00257
42	CBX8	17q25.3	chromobox homolog 8	NP_065700	Q9HC52
43	CCDC150	2q33.1	coiled-coil domain containing 150	NP_001074008	Q8NCX0|B8ZZI4
44	CCDC19	1q23.2	coiled-coil domain containing 19		
45	CCDC78	16p13.3	coiled-coil domain containing 78	NP_001026907	A2IDD5
46	CCDC86	11q12.2	coiled-coil domain containing 86	NP_077003	Q9H6F5
47	CCND1	11q13	cyclin D1		
48	CCT3	1q23	chaperonin containing TCP1, subunit 3 (gamma)	NP_005989	B3KX11|Q59H77|P49368
49	CCT6A	7p11.2	chaperonin containing TCP1, subunit 6A (zeta 1)	NP_001753	A0A024RDL1|P40227
50	CD209	19p13.2	CD209 molecule	NP_066978	Q9NNX6|M0R0P0|X6RB12|B4E2A8|B2R907
51	CD3EAP	19q13.3	CD3e molecule, epsilon associated protein	NP_036231	O15446
52	CD44	11p13	CD44 molecule (Indian blood group)	NP_001189486	P16070
53	CDC25B	20p13	cell division cycle 25 homolog B (S. pombe)	NP_068659	P30305|B4DIG0|B3KS38|G8JLH2|E1YMX4|Q6MZW8|B4DRC3|B4DQZ3
54	CDH3	16q22.1	cadherin 3, type 1, P-cadherin (placental)	NP_001784	P22223
55	CEACAM5	19q13.2	carcinoembryonic antigen related cell adhesion molecule	NP_004354	P06731|A0A024R0K5|Q53G30
56	CELSR3	3p21.31	cadherin, EGF LAG seven-pass G-type receptor 3 (flamingo homolog, Drosophila)	NP_001398	Q9NYQ7
57	CEP72	5p15.33	centrosomal protein 72kDa	NP_060610	Q9P209
58	CGREF1	2p23.3	cell growth regulator with EF-hand domain 1		
59	CKAP2	13q14	cytoskeleton associated protein 2	NP_060674	Q8WWK9
60	CLDN1	3q28-q29	claudin 1	NP_066924	O95832|A5JSJ9
61	CLEC4M	19p13.2	C-type lectin domain family 4 member M	NP_055072	Q9H2X3|H7BXU4|B4E2Z5
62	CLIC1	6p21.3	chloride intracellular channel 1	NP_001279	O00299|Q5SRT3|Q53FB0

Figure 42B

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
63	CLIC4	1p36.11	chloride intracellular channel 4	NP_039234	Q9Y696|Q6FIC5
64	CLU	8p21-p12	clusterin	NP_001822	P10909
65	CNTD2	19q13.2	cyclin N-terminal domain containing 2	NP_079153	Q9H8S5
66	COL10A1	6q21-q22	collagen, type X, alpha 1	NP_000484	Q03692
67	COL11A1	1p21	collagen, type XI, alpha 1	NP_542197	P12107|Q59HB5
68	COL7A1	3p21.1	collagen, type VII, alpha 1	NP_000085	Q02388|Q59F16
69	CPNE7	16q24.3	copine VII	NP_705900	Q9UBL6
70	CSE1L	20q13	CSE1 chromosome segregation 1-like (yeast)	NP_001307	P55060
71	CST1	20p11.2	cystatin SN	NP_001889	P01037
72	CST4	20p11.2	cystatin S	NP_001890	P01036
73	CTHRC1	8q22.3	collagen triple helix repeat containing 1	NP_612464	Q96CG8
74	CTSD	11p15.5	cathepsin D	NP_001900	P07339|V9HWI3
75	CXCL8	4q13.3	C-X-C motif chemokine ligand 8	NP_000575	P10145|A0A024RDA5
76	CYP27B1	12q13.3-q14	cytochrome P450, family 27, subfamily B, polypeptide 1	NP_000776	O15528
77	DCUN1D5	11q22.3	DCN1, defective in cullin neddylation 1, domain containing 5 (S. cerevisiae)	NP_115675	Q9BTE7|A0A024R3A2|B4DP84
78	DDN	12q13	dendrin	NP_055901	O94850
79	DDX10	11q22-q23	DEAD (Asp-Glu-Ala-Asp) box polypeptide 10	NP_004389	Q13206
80	DDX31	9q34.2	DEAD (Asp-Glu-Ala-Asp) box polypeptide 31	NP_619526	Q9H8H2
81	DDX55	12q24.31	DEAD (Asp-Glu-Ala-Asp) box polypeptide 55	NP_065987	Q8NHQ9|A0A024RBS0|Q8TEC9
82	DDX56	7p13	DEAD (Asp-Glu-Ala-Asp) box polypeptide 56	NP_061955	Q9NY93
83	DKC1	Xq28	dyskeratosis congenita 1, dyskerin	NP_001354	O60832
84	DKK3	11p15.2	dickkopf WNT signaling pathway inhibitor 3	NP_056965	Q9UBP4
85	DNAJC2	7q22-q32	DnaJ (Hsp40) homolog, subfamily C, member 2	NP_055192	Q99543
86	DPP4	2q24.2	dipeptidyl peptidase 4	NP_001926	P27487
87	DSCC1	8q24.12	defective in sister chromatid cohesion 1 homolog (S. cerevisiae)	NP_076999	Q9BVC3
88	E2F5	8q21.2	E2F transcription factor 5, p130-binding	NP_001942	Q15329
89	ECT2	3q26.1-q26.2	Epithelial cell transforming sequence 2 oncogene	NP_060568	Q9H8V3
90	EGFL6	Xp22	EGF-like-domain, multiple 6	NP_056322	Q8IUX8
91	EGFR	7p12	epidermal growth factor receptor	NP_958441	P00533
92	EIF2S2	20q11.2	eukaryotic translation initiation factor 2, subunit 2 beta, 38kDa	NP_003899	P20042|Q6IBR8
93	EIF3B	7p22	eukaryotic translation initiation factor 3, subunit B	NP_003742	P55884|A0A024R821

Figure 42C

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
94	ENC1	5q13	ectodermal-neural cortex 1 (with BTB-like domain)	NP_003624	O14682|Q53XS2
95	ENO2 or NSE	12p13.31	enolase 2		P09104
96	EPHX4	1p22.1	epoxide hydrolase 4	NP_775838	Q8IUS5
97	ESM1	5q11	endothelial cell-specific molecule 1	NP_008967	Q9NQ30
98	ETV4	17q21	ets variant 4	NP_001977	P43268
99	FAM176A or EVA1A	2p12	eva-1 homolog A, regulator of programmed cell death	NP_115557	Q9H8M9
100	FAM40B or STRIP2	7q32.1	striatin interacting protein 2	NP_065755	Q9ULQ0
101	FAM89A	1q42.2	family with sequence similarity 89, member A	NP_940954	Q96GI7
102	FANCB	Xp22.2	Fanconi anemia, complementation group B	NP_689846	Q8NB91|A0A024RBW1
103	FAP	2q24.2	fibroblast activation protein alpha	NP_004451	B4DLR2|Q12884
104	FASLG	1q24.3	Fas ligand	NP_001289675	Q53ZZ1|P48023
105	FJX1	11p13	four jointed box 1 (Drosophila)	NP_055159	Q86VR8
106	FLT3LG	19q13.33	fms related tyrosine kinase 3 ligand	NP_001450	P49771|B7ZLY4
107	FOXA1	14q21.1	forkhead box A1	NP_004487	P55317
108	FOXQ1	6p25	forkhead box Q1	NP_150285	Q9C009
109	FTSJ1	Xp11.23	FtsJ homolog 1 (E. coli)	NP_803188	B7Z4K4|A0A024QYX5|B3KN91|Q9UET6
110	FTSJ2	7p22	FtsJ homolog 2 (E. coli)	NP_037525	Q9UI43|V9HWJ9
111	FUT1	19q13.33	fucosyltransferase 1 (galactoside 2-alpha-L-fucosyltransferase, H blood group)	NP_000139	P19526|Q6IZA2
112	FXYD5	19q13.12	FXYD domain containing ion transport regulator 5	NP_659003	Q96DB9|F5H4X8
113	GABRD	1p36.3	gamma-aminobutyric acid (GABA) A receptor, delta	NP_000806	O14764|A8K496
114	GAD1	2q31	glutamate decarboxylase 1 (brain, 67kDa)	NP_038473	Q99259|Q8IVA8
115	GARS	7p15	glycyl-tRNA synthetase	NP_002038	P41250
116	GDF15	19p13.11	growth differentiation factor 15	NP_004855	Q99988
117	GLS2	12q13	glutaminase 2 (liver, mitochondrial)	NP_037399	Q9UI32|A0A087X004
118	GMPS	3q25.31	guanine monphosphate synthetase	NP_003866	P49915
119	GRHL1	2p25.2	grainyhead-like 1 (Drosophila)	NP_937825	Q9NZI5
120	GRHL2	8q22.3	grainyhead like transcription factor 2	NP_079191	Q6ISB3
121	GRHL3	1p36	grainyhead-like 3 (Drosophila)	NP_937817	Q8TE85
122	GRIN1	9q34.3	glutamate ionotropic receptor NMDA type subunit 1	NP_067544	Q05586|Q5VSF9|Q59GW0
123	GRIN2A	16p13.2	glutamate ionotropic receptor NMDA type subunit 2A	NP_001127880	Q12879|Q547U9|Q59EW6
124	GRIN2B	12p13.1	glutamate ionotropic receptor NMDA type subunit 2B	NP_000825	Q13224

Figure 42D

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
125	GRIN2C	17q25.1	glutamate ionotropic receptor NMDA type subunit 2C	NP_001265482	Q14957|H0Y2V8|Q8IW23|O15398
126	GRIN2D	19q13.33	glutamate ionotropic receptor NMDA type subunit 2D	NP_000827	O15399|Q59G17
127	GRP	18q21.32	gastrin releasing peptide	NP_002082	P07492
128	GTF2IRD1	7q11.23	GTF2I repeat domain containing 1	NP_057412	Q9UHL9|E9PFE2
129	GTPBP4	10p15-p14	GTP binding protein 4	NP_036473	Q9BZE4|D2CFK9
130	GYLTL1B	11p11.12	glycosyltransferase-like 1B	NP_689525	Q8N3Y3|E9PIZ2|B3KP69
131	HAGHL	16p13.3	hydroxyacylglutathione hydrolase-like		
132	HAUS6	9p22.1	HAUS augmin-like complex, subunit 6	NP_060115	Q7Z4H7
133	HDAC5	17q21	histone deacetylase 5	NP_005465	Q9UQL6
134	HOXB13	17q21.2	homeobox B13	NP_006352	Q92826|Q4KR72
135	HSP90AB1	6p12	heat shock protein 90kDa alpha (cytosolic), class B member 1	NP_031381	P08238|A0A024RD80|Q6PK50|B4DGL0
136	HSPD1	2q33.1	heat shock 60kDa protein 1 (chaperonin)	NP_955472	P10809|A0A024R3X4
137	IGFBP2	2q35	insulin-like growth factor binding protein 2, 36kDa	NP_000588	P18065
138	IL23A	12q13.13	interleukin 23, alpha subunit p19	NP_057668	Q9NPF7
139	IL6ST	5q11.2	interleukin 6 signal transducer	NP_786943	P40189|Q17RA0|A0A0A0N0L2
140	IMPDH1	7q31.3-q32	IMP (inosine 5'-monophosphate) dehydrogenase 1	NP_899066	P20839|B3KRZ3
141	IMPDH2	3p21.2	IMP (inosine 5'-monophosphate) dehydrogenase 2	NP_000875	P12268
142	INHBA	7p15-p13	inhibin, beta A	NP_002183	P08476|A4D1W7
143	JUB or AJUBA	14q11.2	ajuba LIM protein	NP_932352	Q96IF1
144	KAT2A	17q12-q21	K(lysine) acetyltransferase 2A	NP_066564	Q92830
145	KCNJ14	19q13	potassium inwardly-rectifying channel, subfamily J, member 14	NP_037480	Q9UNX9
146	KIAA1199	15q25.1	KIAA1199		
147	KIAA1257	3q21.3	KIAA1257	NP_065792	Q9ULG3
148	KIAA1549	7q34	KIAA1549	NP_065961	Q9HCM3
149	KIF14	1q32.1	kinesin family member 14	NP_055690	Q15058
150	KIF26B	1q44	kinesin family member 26B	NP_060482	Q2KJY2
151	KLHL35	11q13.4	kelch-like 35 (Drosophila)	NP_001034637	Q6PF15
152	KLK2	19q13.41	kallikrein-related peptidase 2	NP_005542	P20151|A0A024R4N3|A0A024R4J4|B4DU77
153	KLK3 or PSA	19q13.41	kallikrein-related peptidase 3		
154	KLK6	19q13.3	kallikrein-related peptidase 6	NP_002765	Q92876|A0A024R4J8
155	KLKB1	4q35	kallikrein B, plasma (Fletcher factor) 1	NP_000883	P03952
156	KRT23	17q21.2	keratin 23 (histone deacetylase inducible)		

Figure 42E

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
157	KRT80	12q13.13	keratin 80	NP_872313	Q6KB66
158	LEMD1	1q32.1	LEM domain containing 1		
159	LOC646851 or FAM227A	22q13.1	family with sequence similarity 227 member A	NP_001277959	F5H4B4
160	LOC92659 or MAFG-DT	17q25.3	MAFG divergent transcript		
161	LRG1	19p13.3	leucine-rich alpha-2-glycoprotein 1	NP_443204	P02750|Q68CK4
162	LRP8	1p32.3	low density lipoprotein receptor-related protein 8, apolipoprotein e receptor		
163	LRRC8E	19p13.2	leucine rich repeat containing 8 family, member E	NP_079337	Q6NSJ5
164	LYAR	4p16.3	Ly1 antibody reactive homolog (mouse)	NP_060286	Q9NX58
165	MAGEA4	Xq28	melanoma antigen family A4	NP_002353	P43358|A0A024RC12
166	MC1R	16q24.3	melanocortin 1 receptor (alpha melanocyte stimulating hormone receptor)	NP_002377	Q01726|Q1JUL4
167	MDFI	6p21	MyoD family inhibitor	NP_005577	Q99750|A0A024RCY2|B1AKB6
168	MDM2	12q14.3-q15	MDM2 proto-oncogene, E3 ubiquitin protein ligase	NP_002383	Q00987|G3XA89|A7UKX8|A0A0A8KB75|Q96DS0|A7UKX9|A7UKX7
169	MET	7q31	met proto-oncogene (hepatocyte growth factor receptor)	NP_001120972	P08581|A0A024R728|A0A024R759|B4DLF5
170	MEX3A	1q22	mex-3 homolog A (C. elegans)	NP_001087194	A1L020
171	MGC14436 or FAM222A-AS1	12q24.11	FAM222A antisense RNA 1		
172	MKI67IP or NIFK	2q14.3	nucleolar protein interacting with the FHA domain of MKI67	NP_115766	Q9BYG3
173	MMP11	22q11.23	matrix metallopeptidase 11 (stromelysin 3)	NP_005931	P24347|B3KQS8
174	MMP7	11q21-q22	matrix metallopeptidase 7 (matrilysin, uterine)	NP_002414	P09237
175	MSX1	4p16.2	Msh homeobox 1	NP_002439	P28360
176	MSX2	5q35.2	msh homeobox 2	NP_002440	P35548
177	MTBP	8q24.1-q24.2	Mdm2, transformed 3T3 cell double minute 2, p53 binding protein (mouse) binding protein, 104kDa	NP_071328	Q96DY7
178	MTHFD1L	6q25.1	methylenetetrahydrofolate dehydrogenase (NADP+ dependent) 1-like	NP_056255	Q6UB35|B7ZM99|A0A087WVM4|B2RD24
179	MUC16	19p13.2	mucin 16, cell surface associated	NP_078966	Q8WXI7|B5ME49|B3KY81
180	NAA25	12q24.13	N(alpha)-acetyltransferase 25, NatB auxiliary subunit	NP_079229	Q14CX7
181	NAT10	11p13	N-acetyltransferase 10 (GCN5-related)	NP_078938	Q9H0A0
182	NEBL	10p12	nebulette	NP_998734	O76041|Q59FZ8
183	NFE2L3	7p15.2	nuclear factor (erythroid-derived 2)-like 3	NP_004280	Q9Y4A8
184	NFXL1	4p12	nuclear transcription factor, X-box binding-like 1		
185	NKD2	5p15.3	naked cuticle homolog 2 (Drosophila)	NP_149111	Q969F2
186	NKRF	Xq24	NFKB repressing factor	NP_060014	O15226|A3F769|A3F768
187	NMB	15q25.2-q25.3	neuromedin B	NP_995580	P08949

Figure 42F

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
188	NNMT	11q23.2	nicotinamide N-methyltransferase	NP_006160	P40261|Q6FH49
189	NOB1	16q22.1	NIN1/RPN12 binding protein 1 homolog (S. cerevisiae)	NP_054781	Q9ULX3
190	NOL8	9q22.32	nucleolar protein 8	NP_060418	Q76FK4|A6H8Z4
191	NOLC1	10q24.32	nucleolar and coiled-body phosphoprotein 1	NP_004732	Q14978|Q96J17|B2RAU8
192	NONO	Xq13.1	non-POU domain containing, octamer-binding	NP_031389	Q15233
193	NOP2	12p13	NOP2 nucleolar protein homolog (yeast)	NP_006161	P46087
194	NOP58	2q33.1	NOP58 ribonucleoprotein homolog (yeast)	NP_057018	Q9Y2X3
195	NOTUM	17q25.3	notum pectinacetylesterase homolog (Drosophila)	NP_848588	Q6P988
196	NUFIP1	13q14	nuclear fragile X mental retardation protein interacting protein 1	NP_036477	Q9UHK0
197	NYCO16 or UTP14A	Xq26.1	UTP14A, small subunit processome component	NP_006640	Q9BVJ6
198	OSBPL3	7p15.3	oxysterol binding protein-like 3	NP_663162	Q9H4L5
199	OTX1	2p15	orthodenticle homeobox 1	NP_055377	P32242|B4DM62
200	OXTR	3p25	oxytocin receptor	NP_000907	P30559|B2R9L7
201	PAICS	4q12	phosphoribosylaminoimidazole carboxylase, phosphoribosylaminoimidazole succinocarboxamide synthetase		
202	PALB2	16p12.1	partner and localizer of BRCA2	NP_078951	Q86YC2
203	PAQR4	16p13	progestin and adipoQ receptor family member IV	NP_689554	Q8N4S7|I3L1A2|I3L265
204	PDCD2L	19q13.11	programmed cell death 2-like	NP_115722	Q9BRP1
205	PDX1	13q12.1	pancreatic and duodenal homeobox 1	NP_000200	P52945
206	PHF16 or JADE3	Xp11.3	jade family PHD finger 3	NP_055550	Q92613|A0A024R1A2
207	PKM	15q23	pyruvate kinase M1/2	NP_872271	P14618|B4DNK4|V9HWB8|A0A024R5Z9
208	PLEKHA9 or PLEKHA8P1	12q12	pleckstrin homology domain containing A8		
209	PLEKHN1	1p36.33	pleckstrin homology domain containing, family N member 1	NP_115505	Q494U1
210	PLSCR1	3q23	phospholipid scramblase 1	NP_066928	O15162
211	POLR1B	2q13	polymerase (RNA) I polypeptide B, 128kDa	NP_061887	Q9H9Y6|B7Z1W6
212	POU5F1B	8q24.21	POU class 5 homeobox 1B	NP_001153014	Q06416
213	PPAT	4q12	Phosphoribosyl pyrophosphate amidotransferase	NP_002694	Q06203|A8K4H7|Q59G63
214	PPM1H	12q14.1	protein phosphatase, Mg2+/Mn2+ dependent, 1H	NP_065751	Q9ULR3
215	PRDM12	9q33-q34	PR domain containing 12	NP_067632	Q9H4Q4
216	PRDX6	1q25.1	peroxiredoxin 6	NP_004896	P30041|V9HWC7
217	PRPF4	9q31-q33	PRP4 pre-mRNA processing factor 4 homolog (yeast)	NP_004688	O43172|Q5T1M7
218	PRR19	19q13.2	proline rich 19	NP_954979	A6NJB7

Figure 42G

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
219	PRR7	5q35.3	proline rich 7 (synaptic)	NP_085044	Q8TB68|A0A024R7R8
220	PRSS22	16p13.3	protease, serine, 22	NP_071402	Q9GZN4
221	PUS7	7q22.3	pseudouridylate synthase 7 homolog (S. cerevisiae)	NP_061915	Q96PZ0|B3KY42|B3KRB2
222	PVT1	8q24	Pvt1 oncogene (non-protein coding)		
223	QSOX2	9q34.3	quiescin Q6 sulfhydryl oxidase 2	NP_859052	Q6ZRP7
224	RAD54B	8q22.1	RAD54 homolog B (S. cerevisiae)	NP_036547	O95073|Q9Y620|A0A087X0H2
225	RAE1	20q13.31	RAE1 RNA export 1 homolog (S. pombe)	NP_003601	P78406
226	RBM28	7q32.2	RNA binding motif protein 28	NP_060547	Q9NW13|A0A024R753
227	RFC3	13q13.2	replication factor C (activator 1) 3, 38kDa	NP_853536	P40938|A0A024RDQ8
228	RHPN1	8q24.3	rhophilin, Rho GTPase binding protein 1	NP_443156	Q8TCX5
229	RIPK2	8q21	receptor-interacting serine-threonine kinase 2	NP_003812	O43353
230	RPGRIP1L	16q12.2	RPGRIP1-like	NP_056087	Q68CZ1|H3BV03|A0A087WX34
231	RPH3AL	17p13.3	rabphilin 3A-like (without C2 domains)	NP_008918	Q9UNE2|A8K7D5
232	RPL36	19p13.3	ribosomal protein L36		
233	RUVBL1	3q21	RuvB-like 1 (E. coli)	NP_003698	Q9Y265
234	S100A11	1q21	S100 calcium binding protein A11	NP_005611	V9HWH9|P31949
235	S100A2	1q21	S100 calcium binding protein A2	NP_005969	P29034
236	SALL4	20q13.2	Sal-like 4 (Drosophila)	NP_065169	Q9UJQ4
237	SCRN1	7p14.3	secernin 1	NP_055581	A0A090N7T9|A0A024RA21|Q12765
238	SEC61B	9q22.33	Sec61 translocon beta subunit	NP_006799	P60468
239	SET	9q34	SET nuclear oncogene	NP_003002	A0A024R895|Q5VXV3|A0A0C4DFV9|Q01105
240	SFTA2	6p21.3	surfactant associated 2	NP_995326	Q6UW10
241	SH3TC2	5q32	SH3 domain and tetratricopeptide repeats 2	NP_078853	Q8TF17
242	SHISA2	13q12.13	shisa homolog 2 (Xenopus laevis)	NP_001007539	Q6UWI4
243	SIM2	21q22.2	single-minded homolog 2 (Drosophila)	NP_033664	Q14190
244	SLC39A10	2q33.1	solute carrier family 39 (zinc transporter), member 10	NP_065075	A0A024R3W5|Q05C42|Q9ULF5
245	SLC4A11	20p13	solute carrier family 4, sodium borate transporter, member 11	NP_114423	Q8NBS3
246	SLC6A6	3p25.1	solute carrier family 6 (neurotransmitter transporter, taurine), member 6	NP_003034	Q59GD7|A0A087WY96|B4E140|P31641
247	SLC7A5	16q24.3	solute carrier family 7 (cationic amino acid transporter, y+ system), member 5	NP_003477	Q01650
248	SLCO4A1	20q13.1	Solute carrier organic anion transporter family, member 4A1	NP_057438	Q96BD0
249	SLMO1	18p11.21	slowmo homolog 1 (Drosophila)	NP_006544	Q96N28

Figure 42H

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
250	SMOX	20p13	spermine oxidase	NP_787036	Q9NWM0
251	SMYD5	2p12	SMYD family member 5	NP_006053	Q6GMV2
252	SND1	7q31.3	staphylococcal nuclease and tudor domain containing 1	NP_055205	Q7KZF4
253	SNORD1C	17q25.1	small nucleolar RNA, C/D box 1C		
254	SNTB1	8q23-q24	syntrophin, beta 1 (dystrophin-associated protein A1, 59kDa, basic component 1)	NP_066301	Q13884
255	SOX4	6p22.3	SRY (sex determining region Y)-box 4	NP_003098	Q06945
256	SPATA12	3p21.2	spermatogenesis associated 12	NP_859078	Q7Z6I5
257	SPERT	13q14.13	spermatid associated	NP_689932	Q8NA61
258	SPTBN2	11q13.2	spectrin, beta, non-erythrocytic 2	NP_008877	O15020
259	SSB	2q31.1	Sjogren syndrome antigen B (autoantigen La)	NP_003133	P05455
260	STOML2 or SLP2	9p13.3	stomatin like 2	NP_038470	A0A087WYB4|Q9UJZ1
261	STRA6	15q24.1	stimulated by retinoic acid gene 6 homolog (mouse)	NP_071764	B3KPB8|Q9BX79
262	TAF1A	1q42	TATA box binding protein (TBP)-associated factor, RNA polymerase I, A, 48kDa	NP_647603	B4DS21|A8K4K5|Q15573
263	TAF1D	11q21	TATA box binding protein (TBP)-associated factor, RNA polymerase I, D, 41kDa	NP_077021	A0A024R3A9|Q9H5J8
264	TEAD4	12p13.3-p13.2	TEA domain family member 4	NP_958851	Q15561
265	TESC	12q24.22	Tescalcin	NP_060369	Q96BS2
266	TEX10	9q31.1	testis expressed 10	NP_060216	A0A024R169|Q9NXF1
267	TFAP2A	6p24	transcription factor AP-2 alpha (activating enhancer binding protein 2 alpha)	NP_003211	P05549
268	TFAP4	16p13	transcription factor AP-4 (activating enhancer binding protein 4)	NP_003214	Q01664
269	TFF3	21q22.3	trefoil factor 3 (intestinal)	NP_003217	A0A0A6YYJ4|Q07654
270	TGFBI	5q31	transforming growth factor, beta-induced, 68kDa	NP_000349	Q15582
271	TGIF2	20q11.23	TGFB-induced factor homeobox 2	NP_068581	Q9GZN2
272	THUMPD2	2p22.2	THUMP domain containing 2	NP_079540	D6W593|Q969Z2|Q9BTF0
273	TIGD1	2q37.1	tigger transposable element derived 1	NP_663748	Q96MW7
274	TMEM132A	11q12.2	transmembrane protein 132A	NP_821174	Q24JP5
275	TMEM206	1q32.3	transmembrane protein 206	NP_060722	B4DHQ5|Q9H813
276	TOMM34	20q12-q13.1	translocase of outer mitochondrial membrane 34	NP_006800	Q15785
277	TP53	17p13.1	tumor protein p53	NP_001263690	P04637|K7PPA8|A0A087X1Q1|A0A087WT22|A0A087WXZ1|H2EHT1|Q53GA5
278	TPM1	15q22.1	tropomyosin 1 (alpha)	NP_001288218	B7Z596|H7BYY1|Q6ZN40|B7Z722|O15513|D9YZV5|A0A024R5W6|F5H7S3|D9YZV4|D9YZV7|P09493
279	TPM2	9p13	tropomyosin 2 (beta)	NP_998839	A7XZE4|Q5TCU3|V9HW25|P07951
280	TPM3	1q21.2	tropomyosin 3	NP_705935	A0A087WWU8|J3KN67|B4DQ80|P06753

Figure 42I

Order	Gene Symbol	Chromosome Band	Gene Title	Protein Accession (representative)	UniProt IDs
281	TPM4	19p13.1	tropomyosin 4	NP_003281	V9HW56|P67936
282	TPX2	20q11.2	TPX2, microtubule-associated, homolog (Xenopus laevis)	NP_036244	Q643R0|Q9ULW0
283	TRIB3	20p13-p12.2	tribbles homolog 3 (Drosophila)	NP_066981	J3KR25|B4DMM9|Q96RU7
284	TRIP13	5p15	thyroid hormone receptor interactor 13	NP_004228	Q15645
285	TTC26	7q34	tetratricopeptide repeat domain 26	NP_079202	A0AVF1|B7Z2T3|B7Z1H0
286	TTR	18q12.1	transthyretin	NP_000362	E9KL36|P02766
287	UBE2I	16p13.3	ubiquitin-conjugating enzyme E2I (UBC9 homolog, yeast)	NP_919237	B0QYN7|A8K503|P63279
288	UCN2	3p21.3	urocortin 2	NP_149976	Q96RP3
289	ULBP1	6q25	UL16 binding protein 1		
290	ULBP2	6q25	UL16 binding protein 2	NP_079493	Q9BZM5
291	UTP14A	Xq26.1	UTP14, U3 small nucleolar ribonucleoprotein, homolog A (yeast)	NP_006640	Q9BVJ6
292	VWA2	10q25.3	von Willebrand factor A domain containing 2	NP_001258975	Q5GFL6
293	VWF	12p13.3	von Willebrand factor	NP_000543	P04275
294	WDR3	1p12	WD repeat domain 3	NP_006775	Q5TDG3|Q9UNX4
295	WDR43	2p23.3	WD repeat domain 43	NP_055946	Q15061
296	WDR66	12q24.31	WD repeat domain 66	NP_653269	Q8TBY9
297	WDR74	11q12.3	WD repeat domain 74	NP_060563	Q6RFH5
298	WDR75	2q32.2	WD repeat domain 75	NP_115544	Q8IWA0
299	WNT2	7q31	wingless-type MMTV integration site family member 2	NP_003382	P09544
300	XPO5	6p21.1	exportin 5		
301	XPOT	12q14.1	exportin, tRNA (nuclear export receptor for tRNAs)	NP_009166	A8KA19|O43592
302	XRCC2	7q36	X-ray repair complementing defective repair in Chinese hamster cells 2	NP_005422	O43543
303	YTHDF1	20q13.33	YTH domain family, member 1	NP_060268	Q9BYJ9
304	ZBTB12	6p21.31	zinc finger and BTB domain containing 12	NP_862825	Q9Y330
305	ZC3H8	2q13	zinc finger CCCH-type containing 8	NP_115883	Q8N5P1
306	ZC3HAV1L	7q34	zinc finger CCCH-type, antiviral 1-like	NP_542391	Q96H79
307	ZMYND19	9q34.3	zinc finger, MYND-type containing 19	NP_612471	Q96E35
308	ZNF280C	Xq25	zinc finger protein 280C	NP_060136	Q8ND82
309	ZNF473	19q13.33	zinc finger protein 473	NP_056243	A0A024QZI1|Q8WTR7
310	ZNF643 or ZFP69B	1p34.2	ZFP69 zinc finger protein B	NP_075558	D3DPV3|Q9UJL9
311	ZSWIM4	19p13.13	zinc finger, SWIM-type containing 4	NP_075560	Q9HA55|Q9H7M6

Figure 42J

CRC secreted Protein markers

Order	Protein	UniProt ID
1	A disintegrin and metalloproteinase with thrombospondin motifs 6	Q9UKP5
2	Aggrecan core protein	E7ENV9
3	Aggrecan core protein	E7EX88
4	Alpha(1,2)fucosyltransferase	Q6IZA2
5	Apelin	Q9ULZ1
6	Brain-specific serine protease 4	Q9GZN4
7	CA-11-19	
8	CA-125	
9	Carcinoembryonic antigen-related cell adhesion molecule 5 variant (Fragment)	Q53G30
10	Cathepsin D	P07339
11	CD209 antigen	Q9NNX6
12	cDNA FLJ31267 fis, clone KIDNE2006053, highly similar to Glycosyltransferase-like protein LARGE2 (EC 2.4.-.-)	B3KP69
13	cDNA FLJ77564	A8K6G4
14	Clusterin	P10909
15	Collagen alpha-1(VII) chain	Q02388
16	Collagen alpha-1(X) chain	Q03692
17	Collagen alpha-1(XI) chain	P12107
18	Collagen triple helix repeat-containing protein 1	Q96CG8
19	Complement C1r subcomponent	P00736
20	C-type lectin domain family 4 member M	Q9H2X3
21	Cystatin-S	P01036
22	Cystatin-SN	P01037
23	Dickkopf-related protein 3	Q9UBP4
24	Disintegrin and metalloproteinase domain-containing protein 12	O43184
25	Endothelial cell-specific molecule 1	Q9NQ30
26	Epidermal growth factor receptor	P00533
27	Epidermal growth factor-like protein 6	Q8IUX8
28	Epididymis tissue sperm binding protein Li 4a	E9KL36
29	Fms-related tyrosine kinase 3 ligand	P49771
30	Four-jointed box protein 1	Q86VR8
31	Gastrin-releasing peptide	P07492
32	Glycosyltransferase-like protein LARGE2	E9PIZ2
33	Growth/differentiation factor 15	Q99988

Order	Protein	UniProt ID
34	Hepatocyte growth factor receptor	P08581
35	Hypoxia-inducible lipid droplet-associated protein	Q9Y5L2
36	Inhibin beta A chain	P08476
37	Inhibin, beta A (Activin A, activin AB alpha polypeptide)	A4D1W7
38	Insulin-like growth factor-binding protein 2	P18065
39	Interleukin-23 subunit alpha	Q9NPF7
40	Interleukin-6 receptor subunit beta	P40189
41	Interleukin-8	P10145
42	Kallikrein-2	P20151
43	Kallikrein-6	Q92876
44	Leucine-rich alpha-2-glycoprotein	Q68CK4
45	Leucine-rich alpha-2-glycoprotein	P02750
46	Matrilysin	P09237
47	Matrix metallopeptidase 11 (Stromelysin 3), isoform CRA_b	B3KQS8
48	Mucin-16	Q8WXI7
49	Neuromedin-B	P08949
50	NKG2D ligand 2	Q9BZM5
51	Phosphoribosyl pyrophosphate amidotransferase proprotein variant (Fragment)	Q59G63
52	Plasma kallikrein	P03952
53	Protein notum homolog	Q6P988
54	Protein Wnt-2	P09544
55	Seprase	B4DLR2
56	SLC39A10 protein (Fragment)	Q05C42
57	Stromelysin-3	P24347
58	Sulfhydryl oxidase 2	Q6ZRP7
59	Surfactant-associated protein 2	Q6UW10
60	Transforming growth factor-beta-induced protein ig-h3	Q15582
61	Transthyretin	P02766
62	Trefoil factor 3	Q07654
63	Urocortin-2	Q96RP3
64	von Willebrand factor	P04275
65	von Willebrand factor A domain-containing protein 2	Q5GFL6

Figure 43

CRC Methylation markers, 50% sensitivity, CpG Regions

Order	Coordinate of CpG (hg19)	Gene
1	Chr1:_969257_969258	*AGRN*
2	Chr1:_975551_975552	*AGRN*
3	Chr1:_1144658_1144659	*Non Coding Region*
4	Chr1:_1306946_1306947	*Non Coding Region*
5	Chr1:_1565856_1565857	*MIB2*
6	Chr1:_1566699_1566700	*MMP23B,MMP23A*
7	Chr1:_2222253_2222254	*SKI*
8	Chr1:_2232166_2232167	*SKI*
9	Chr1:_2984869_2984870	*PRDM16,FLJ42875*
10	Chr1:_2987645_2987646	*PRDM16*
11	Chr1:_3100343_3100344	*PRDM16*
12	Chr1:_3663531_3663532	*KIAA0495*
13	Chr1:_6088550_6088551	*KCNAB2*
14	Chr1:_6520354_6520355	*ESPN*
15	Chr1:_6526051_6526052	*TNFRSF25*
16	Chr1:_7692367_7692368	*CAMTA1*
17	Chr1:_8457758_8457759	*RERE*
18	Chr1:_9486780_9486781	*Non Coding Region*
19	Chr1:_9689803_9689804	*Non Coding Region*
20	Chr1:_12123262_12123263	*TNFRSF8*
21	Chr1:_12123313_12123314	*TNFRSF8*
22	Chr1:_13910796_13910797	*PDPN*
23	Chr1:_16023706_16023707	*PLEKHM2*
24	Chr1:_18957020_18957021	*PAX7*
25	Chr1:_18957632_18957633	*PAX7*
26	Chr1:_18959268_18959269	*PAX7*
27	Chr1:_19254031_19254032	*IFFO2*
28	Chr1:_19254731_19254732	*IFFO2*
29	Chr1:_19803932_19803933	*CAPZB*
30	Chr1:_25349006_25349007	*Non Coding Region*
31	Chr1:_26377899_26377900	*TRIM63*
32	Chr1:_27070873_27070874	*ARID1A*
33	Chr1:_27687074_27687075	*MAP3K6*
34	Chr1:_28213800_28213801	*Non Coding Region*
35	Chr1:_29585898_29585899	*PTPRU*
36	Chr1:_31230601_31230602	*LAPTM5*
37	Chr1:_31230846_31230847	*LAPTM5*
38	Chr1:_32716773_32716774	*LCK*
39	Chr1:_34629400_34629401	*CSMD2*
40	Chr1:_34630153_34630154	*CSMD2*
41	Chr1:_34630944_34630945	*CSMD2*
42	Chr1:_35351340_35351341	*DLGAP3*
43	Chr1:_35395663_35395664	*Non Coding Region*
44	Chr1:_35395821_35395822	*Non Coding Region*
45	Chr1:_36043002_36043003	*TFAP2E*

Order	Coordinate of CpG (hg19)	Gene
46	Chr1:_36043014_36043015	*TFAP2E*
47	Chr1:_36043084_36043085	*TFAP2E*
48	Chr1:_36043633_36043634	*TFAP2E*
49	Chr1:_37499547_37499548	*GRIK3*
50	Chr1:_38100811_38100812	*RSPO1*
51	Chr1:_38230779_38230780	*EPHA10*
52	Chr1:_39025249_39025250	*Non Coding Region*
53	Chr1:_39270149_39270150	*Non Coding Region*
54	Chr1:_42217077_42217078	*HIVEP3*
55	Chr1:_44031300_44031301	*PTPRF*
56	Chr1:_44873064_44873065	*RNF220*
57	Chr1:_44873229_44873230	*RNF220*
58	Chr1:_46951318_46951319	*Non Coding Region*
59	Chr1:_46951354_46951355	*Non Coding Region*
60	Chr1:_47701206_47701207	*Non Coding Region*
61	Chr1:_47911449_47911450	*Non Coding Region*
62	Chr1:_48058985_48058986	*Non Coding Region*
63	Chr1:_48191031_48191032	*Non Coding Region*
64	Chr1:_48450020_48450021	*Non Coding Region*
65	Chr1:_49242513_49242514	*AGBL4,BEND5*
66	Chr1:_49242519_49242520	*AGBL4,BEND5*
67	Chr1:_50882006_50882007	*Non Coding Region*
68	Chr1:_50884480_50884481	*DMRTA2*
69	Chr1:_50886782_50886783	*DMRTA2*
70	Chr1:_53099053_53099054	*FAM159A*
71	Chr1:_53527665_53527666	*PODN*
72	Chr1:_53527693_53527694	*PODN*
73	Chr1:_57889035_57889036	*DAB1*
74	Chr1:_57889604_57889605	*DAB1*
75	Chr1:_58715539_58715540	*DAB1*
76	Chr1:_58715553_58715554	*DAB1*
77	Chr1:_65731432_65731433	*DNAJC6*
78	Chr1:_65731738_65731739	*DNAJC6*
79	Chr1:_67217826_67217827	*TCTEX1D1*
80	Chr1:_67217853_67217854	*TCTEX1D1*
81	Chr1:_67218165_67218166	*TCTEX1D1*
82	Chr1:_67773719_67773720	*IL12RB2*
83	Chr1:_70032907_70032908	*Non Coding Region*
84	Chr1:_70034475_70034476	*Non Coding Region*
85	Chr1:_70034491_70034492	*Non Coding Region*
86	Chr1:_72748124_72748125	*NEGR1*
87	Chr1:_76080727_76080728	*Non Coding Region*
88	Chr1:_76081962_76081963	*Non Coding Region*
89	Chr1:_77333138_77333139	*ST6GALNAC5*
90	Chr1:_77333159_77333160	*ST6GALNAC5*

Figure 44A

Order	Coordinate of CpG (hg19)	Gene
91	Chr1:_77334207_77334208	*ST6GALNAC5*
92	Chr1:_77747875_77747876	*AK5*
93	Chr1:_77748129_77748130	*AK5*
94	Chr1:_86622628_86622629	*COL24A1*
95	Chr1:_87598069_87598070	*LOC339524*
96	Chr1:_87617350_87617351	*LOC339524*
97	Chr1:_88921473_88921474	*Non Coding Region*
98	Chr1:_91191001_91191002	*Non Coding Region*
99	Chr1:_91196332_91196333	*Non Coding Region*
100	Chr1:_91300215_91300216	*Non Coding Region*
101	Chr1:_91870362_91870363	*HFM1*
102	Chr1:_91870382_91870383	*HFM1*
103	Chr1:_98511258_98511259	*Non Coding Region*
104	Chr1:_98511789_98511790	*MIR137*
105	Chr1:_101004638_101004639	*GPR88*
106	Chr1:_101004934_101004935	*GPR88*
107	Chr1:_101702602_101702603	*S1PR1*
108	Chr1:_107683725_107683726	*NTNG1*
109	Chr1:_107683775_107683776	*NTNG1*
110	Chr1:_107684339_107684340	*NTNG1*
111	Chr1:_107684425_107684426	*NTNG1*
112	Chr1:_108507468_108507469	*VAV3*
113	Chr1:_108508185_108508186	*VAV3*
114	Chr1:_108508207_108508208	*VAV3*
115	Chr1:_111098150_111098151	*Non Coding Region*
116	Chr1:_111217194_111217195	*KCNA3*
117	Chr1:_111217497_111217498	*KCNA3*
118	Chr1:_111217712_111217713	*KCNA3*
119	Chr1:_111217715_111217716	*KCNA3*
120	Chr1:_114414340_114414341	*PTPN22*
121	Chr1:_115880865_115880866	*NGF*
122	Chr1:_117057968_117057969	*CD58*
123	Chr1:_119530600_119530601	*TBX15*
124	Chr1:_119532195_119532196	*TBX15*
125	Chr1:_119532925_119532926	*TBX15*
126	Chr1:_147790806_147790807	*Non Coding Region*
127	Chr1:_150123734_150123735	*PLEKHO1*
128	Chr1:_150254280_150254281	*C1orf51*
129	Chr1:_150254546_150254547	*C1orf51*
130	Chr1:_150254869_150254870	*C1orf51*
131	Chr1:_151129298_151129299	*TNFAIP8L2*
132	Chr1:_151129844_151129845	*TNFAIP8L2*
133	Chr1:_151810887_151810888	*LOC100132111,C2CD4D*
134	Chr1:_151810893_151810894	*LOC100132111,C2CD4D*
135	Chr1:_151811364_151811365	*LOC100132111,C2CD4D*

Order	Coordinate of CpG (hg19)	Gene
136	Chr1:_153518384_153518385	*S100A4*
137	Chr1:_154298335_154298336	*ATP8B2*
138	Chr1:_154475173_154475174	*TDRD10,SHE*
139	Chr1:_156066986_156066987	*Non Coding Region*
140	Chr1:_156358231_156358232	*Non Coding Region*
141	Chr1:_156405924_156405925	*Non Coding Region*
142	Chr1:_156626749_156626750	*BCAN*
143	Chr1:_156829691_156829692	*NTRK1,INSRR*
144	Chr1:_158150649_158150650	*CD1D*
145	Chr1:_160807694_160807695	*CD244*
146	Chr1:_161039681_161039682	*ARHGAP30*
147	Chr1:_161039690_161039691	*ARHGAP30*
148	Chr1:_161039695_161039696	*ARHGAP30*
149	Chr1:_161228674_161228675	*PCP4L1*
150	Chr1:_165414544_165414545	*RXRG*
151	Chr1:_167598958_167598959	*RCSD1*
152	Chr1:_167601275_167601276	*RCSD1*
153	Chr1:_167682922_167682923	*Non Coding Region*
154	Chr1:_169396635_169396636	*C1orf114*
155	Chr1:_169396637_169396638	*C1orf114*
156	Chr1:_169396706_169396707	*C1orf114*
157	Chr1:_169396785_169396786	*C1orf114*
158	Chr1:_169396858_169396859	*C1orf114*
159	Chr1:_170630558_170630559	*Non Coding Region*
160	Chr1:_171810376_171810377	*DNM3*
161	Chr1:_181452044_181452045	*CACNA1E*
162	Chr1:_190447290_190447291	*FAM5C*
163	Chr1:_200008026_200008027	*NR5A2*
164	Chr1:_202128682_202128683	*PTPN7*
165	Chr1:_202129204_202129205	*PTPN7*
166	Chr1:_202129255_202129256	*PTPN7*
167	Chr1:_203598761_203598762	*ATP2B4*
168	Chr1:_203643084_203643085	*ATP2B4*
169	Chr1:_204328843_204328844	*PLEKHA6*
170	Chr1:_204653629_204653630	*LRRN2*
171	Chr1:_206732715_206732716	*RASSF5*
172	Chr1:_206748850_206748851	*RASSF5*
173	Chr1:_207818395_207818396	*CR1L*
174	Chr1:_208084456_208084457	*CD34*
175	Chr1:_209929546_209929547	*TRAF3IP3*
176	Chr1:_209929622_209929623	*TRAF3IP3*
177	Chr1:_214153294_214153295	*Non Coding Region*
178	Chr1:_218520792_218520793	*TGFB2*
179	Chr1:_220702225_220702226	*MARK1*
180	Chr1:_221050459_221050460	*Non Coding Region*

Figure 44B

Order	Coordinate of CpG (hg19)	Gene
181	Chr1:_221051758_221051759	*HLX*
182	Chr1:_221886639_221886640	*DUSP10*
183	Chr1:_226865321_226865322	*ITPKB*
184	Chr1:_226924846_226924847	*ITPKB*
185	Chr1:_228604240_228604241	*TRIM17*
186	Chr1:_228652478_228652479	*Non Coding Region*
187	Chr1:_228652528_228652529	*Non Coding Region*
188	Chr1:_229543112_229543113	*Non Coding Region*
189	Chr1:_229568252_229568253	*ACTA1*
190	Chr1:_232862025_232862026	*Non Coding Region*
191	Chr1:_234350051_234350052	*SLC35F3*
192	Chr1:_237205174_237205175	*RYR2*
193	Chr1:_237205999_237206000	*RYR2*
194	Chr1:_240255486_240255487	*FMN2*
195	Chr1:_241587609_241587610	*Non Coding Region*
196	Chr1:_244013006_244013007	*Non Coding Region*
197	Chr1:_247712237_247712238	*C1orf150*
198	Chr1:_247712239_247712240	*C1orf150*
199	Chr2:_468179_468180	*Non Coding Region*
200	Chr2:_5836111_5836112	*SOX11*
201	Chr2:_5836256_5836257	*SOX11*
202	Chr2:_7058390_7058391	*RNF144A*
203	Chr2:_7058432_7058433	*RNF144A*
204	Chr2:_10220886_10220887	*CYS1*
205	Chr2:_20865651_20865652	*GDF7*
206	Chr2:_25439110_25439111	*Non Coding Region*
207	Chr2:_25499764_25499765	*DNMT3A*
208	Chr2:_29337988_29337989	*CLIP4*
209	Chr2:_29338100_29338101	*CLIP4*
210	Chr2:_29338109_29338110	*CLIP4*
211	Chr2:_29338113_29338114	*CLIP4*
212	Chr2:_29338121_29338122	*CLIP4*
213	Chr2:_29338258_29338259	*CLIP4*
214	Chr2:_29338432_29338433	*CLIP4*
215	Chr2:_29339076_29339077	*CLIP4*
216	Chr2:_29339091_29339092	*CLIP4*
217	Chr2:_30144579_30144580	*ALK*
218	Chr2:_31360693_31360694	*GALNT14*
219	Chr2:_31360817_31360818	*GALNT14*
220	Chr2:_31361104_31361105	*GALNT14*
221	Chr2:_31457043_31457044	*EHD3*
222	Chr2:_31806042_31806043	*SRD5A2*
223	Chr2:_37875836_37875837	*CDC42EP3*
224	Chr2:_38301756_38301757	*CYP1B1*
225	Chr2:_39893078_39893079	*TMEM178*
226	Chr2:_39893080_39893081	*TMEM178*
227	Chr2:_40678691_40678692	*SLC8A1*
228	Chr2:_45155991_45155992	*Non Coding Region*
229	Chr2:_45169548_45169549	*SIX3*
230	Chr2:_45169562_45169563	*SIX3*
231	Chr2:_45233528_45233529	*SIX2*
232	Chr2:_45233773_45233774	*SIX2*
233	Chr2:_45238442_45238443	*Non Coding Region*
234	Chr2:_47797488_47797489	*KCNK12*
235	Chr2:_47797963_47797964	*KCNK12*
236	Chr2:_56411534_56411535	*CCDC85A*
237	Chr2:_58656064_58656065	*Non Coding Region*
238	Chr2:_63275509_63275510	*LOC100132215*
239	Chr2:_63283013_63283014	*OTX1*
240	Chr2:_63283967_63283968	*OTX1*
241	Chr2:_63284132_63284133	*OTX1*
242	Chr2:_66653254_66653255	*Non Coding Region*
243	Chr2:_66660874_66660875	*Non Coding Region*
244	Chr2:_66673316_66673317	*MEIS1*
245	Chr2:_66803033_66803034	*Non Coding Region*
246	Chr2:_66810606_66810607	*Non Coding Region*
247	Chr2:_68546467_68546468	*CNRIP1*
248	Chr2:_68546507_68546508	*CNRIP1*
249	Chr2:_68546579_68546580	*CNRIP1*
250	Chr2:_68546899_68546900	*CNRIP1*
251	Chr2:_68547052_68547053	*CNRIP1*
252	Chr2:_68547088_68547089	*CNRIP1*
253	Chr2:_68961987_68961988	*ARHGAP25*
254	Chr2:_68962012_68962013	*ARHGAP25*
255	Chr2:_69001774_69001775	*ARHGAP25*
256	Chr2:_69001799_69001800	*ARHGAP25*
257	Chr2:_69001854_69001855	*ARHGAP25*
258	Chr2:_69001870_69001871	*ARHGAP25*
259	Chr2:_69002111_69002112	*ARHGAP25*
260	Chr2:_70313417_70313418	*PCBP1*
261	Chr2:_71298920_71298921	*NAGK*
262	Chr2:_73151595_73151596	*EMX1*
263	Chr2:_74743437_74743438	*TLX2*
264	Chr2:_85362030_85362031	*TCF7L1*
265	Chr2:_87755371_87755372	*NCRNA00152*
266	Chr2:_95691255_95691256	*MAL*
267	Chr2:_97193289_97193290	*Non Coding Region*
268	Chr2:_97193524_97193525	*Non Coding Region*
269	Chr2:_99553442_99553443	*C2orf55*
270	Chr2:_100938444_100938445	*LONRF2*

Figure 44C

Order	Coordinate of CpG (hg19)	Gene
271	Chr2:_100938813_100938814	*LONRF2*
272	Chr2:_101034257_101034258	*CHST10*
273	Chr2:_102090444_102090445	*REX8*
274	Chr2:_105470561_105470562	*POU3F3*
275	Chr2:_105479054_105479055	*Non Coding Region*
276	Chr2:_105760890_105760891	*Non Coding Region*
277	Chr2:_106415145_106415146	*NCK2*
278	Chr2:_106776775_106776776	*UXS1*
279	Chr2:_106777038_106777039	*UXS1*
280	Chr2:_109647956_109647957	*Non Coding Region*
281	Chr2:_109648931_109648932	*Non Coding Region*
282	Chr2:_113874068_113874069	*IL1RN*
283	Chr2:_113931518_113931519	*LOC440839,PSD4*
284	Chr2:_113931525_113931526	*LOC440839,PSD4*
285	Chr2:_115920612_115920613	*DPP10*
286	Chr2:_119600002_119600003	*EN1*
287	Chr2:_119606746_119606747	*EN1*
288	Chr2:_119916017_119916018	*C1QL2*
289	Chr2:_121104400_121104401	*INHBB*
290	Chr2:_127413831_127413832	*GYPC*
291	Chr2:_127414381_127414382	*GYPC*
292	Chr2:_127414827_127414828	*GYPC*
293	Chr2:_127644075_127644076	*Non Coding Region*
294	Chr2:_127783168_127783169	*Non Coding Region*
295	Chr2:_127839539_127839540	*BIN1*
296	Chr2:_128433373_128433374	*LIMS2*
297	Chr2:_131792772_131792773	*ARHGEF4*
298	Chr2:_139537197_139537198	*NXPH2*
299	Chr2:_143886326_143886327	*ARHGAP15*
300	Chr2:_144694405_144694406	*Non Coding Region*
301	Chr2:_144695257_144695258	*Non Coding Region*
302	Chr2:_145281942_145281943	*Non Coding Region*
303	Chr2:_149633226_149633227	*KIF5C*
304	Chr2:_149634141_149634142	*KIF5C*
305	Chr2:_158300475_158300476	*CYTIP*
306	Chr2:_158300485_158300486	*CYTIP*
307	Chr2:_161264607_161264608	*RBMS1*
308	Chr2:_164593221_164593222	*FIGN*
309	Chr2:_171573223_171573224	*SP5*
310	Chr2:_171674855_171674856	*GAD1*
311	Chr2:_172945274_172945275	*MAP1D*
312	Chr2:_172972840_172972841	*Non Coding Region*
313	Chr2:_175191045_175191046	*Non Coding Region*
314	Chr2:_175208588_175208589	*Non Coding Region*
315	Chr2:_175208605_175208606	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
316	Chr2:_175546916_175546917	*WIPF1*
317	Chr2:_175547056_175547057	*WIPF1*
318	Chr2:_176956678_176956679	*HOXD13*
319	Chr2:_177030150_177030151	*HOXD3*
320	Chr2:_182321373_182321374	*ITGA4*
321	Chr2:_182321786_182321787	*ITGA4*
322	Chr2:_182322268_182322269	*ITGA4*
323	Chr2:_182322749_182322750	*ITGA4*
324	Chr2:_182548807_182548808	*Non Coding Region*
325	Chr2:_185462408_185462409	*ZNF804A*
326	Chr2:_187713902_187713903	*ZSWIM2*
327	Chr2:_187713986_187713987	*ZSWIM2*
328	Chr2:_193059230_193059231	*TMEFF2*
329	Chr2:_193059645_193059646	*TMEFF2*
330	Chr2:_193059717_193059718	*TMEFF2*
331	Chr2:_198063653_198063654	*ANKRD44*
332	Chr2:_198063759_198063760	*ANKRD44*
333	Chr2:_198063781_198063782	*ANKRD44*
334	Chr2:_198120525_198120526	*Non Coding Region*
335	Chr2:_202125212_202125213	*CASP8*
336	Chr2:_206551364_206551365	*NRP2*
337	Chr2:_207507096_207507097	*LOC200726*
338	Chr2:_208989248_208989249	*CRYGD*
339	Chr2:_208989382_208989383	*CRYGD*
340	Chr2:_219847136_219847137	*FEV*
341	Chr2:_220112465_220112466	*STK16*
342	Chr2:_220174201_220174202	*PTPRN*
343	Chr2:_220299740_220299741	*SPEG*
344	Chr2:_220307073_220307074	*SPEG*
345	Chr2:_220307207_220307208	*SPEG*
346	Chr2:_220361467_220361468	*Non Coding Region*
347	Chr2:_220417030_220417031	*OBSL1*
348	Chr2:_222435214_222435215	*EPHA4*
349	Chr2:_222435351_222435352	*EPHA4*
350	Chr2:_223162881_223162882	*CCDC140,PAX3*
351	Chr2:_223163809_223163810	*CCDC140,PAX3*
352	Chr2:_223167400_223167401	*CCDC140*
353	Chr2:_227288518_227288519	*Non Coding Region*
354	Chr2:_230578216_230578217	*DNER*
355	Chr2:_233180415_233180416	*DIS3L2*
356	Chr2:_233499070_233499071	*EFHD1*
357	Chr2:_238535910_238535911	*LRRFIP1*
358	Chr2:_242743588_242743589	*GAL3ST2*
359	Chr3:_238392_238393	*CHL1*
360	Chr3:_9821816_9821817	*TADA3*

Figure 44D

Order	Coordinate of CpG (hg19)	Gene
361	Chr3:_16555379_16555380	*RFTN1*
362	Chr3:_27771766_27771767	*Non Coding Region*
363	Chr3:_32859438_32859439	*TRIM71*
364	Chr3:_37034473_37034474	*EPM2AIP1,MLH1*
365	Chr3:_42947690_42947691	*ZNF662*
366	Chr3:_43431343_43431344	*ANO10*
367	Chr3:_44037128_44037129	*Non Coding Region*
368	Chr3:_44063593_44063594	*Non Coding Region*
369	Chr3:_44063832_44063833	*Non Coding Region*
370	Chr3:_44626538_44626539	*ZNF660*
371	Chr3:_44727069_44727070	*Non Coding Region*
372	Chr3:_49843809_49843810	*UBA7*
373	Chr3:_50313021_50313022	*SEMA3B*
374	Chr3:_50313693_50313694	*SEMA3B*
375	Chr3:_51990575_51990576	*GPR62*
376	Chr3:_55517853_55517854	*WNT5A*
377	Chr3:_56789110_56789111	*ARHGEF3*
378	Chr3:_62356962_62356963	*FEZF2*
379	Chr3:_68981866_68981867	*FAM19A4*
380	Chr3:_87040286_87040287	*VGLL3*
381	Chr3:_96532043_96532044	*EPHA6*
382	Chr3:_96533290_96533291	*EPHA6*
383	Chr3:_111852156_111852157	*GCET2*
384	Chr3:_111852227_111852228	*GCET2*
385	Chr3:_112931194_112931195	*BOC*
386	Chr3:_118955844_118955845	*B4GALT4*
387	Chr3:_119528656_119528657	*NR1I2*
388	Chr3:_119528956_119528957	*NR1I2*
389	Chr3:_120004186_120004187	*Non Coding Region*
390	Chr3:_121379745_121379746	*HCLS1*
391	Chr3:_121379777_121379778	*HCLS1*
392	Chr3:_121379791_121379792	*HCLS1*
393	Chr3:_121903520_121903521	*CASR*
394	Chr3:_124860686_124860687	*SLC12A8*
395	Chr3:_128211686_128211687	*GATA2*
396	Chr3:_128212273_128212274	*GATA2*
397	Chr3:_128720433_128720434	*CCDC48*
398	Chr3:_128720884_128720885	*CCDC48*
399	Chr3:_129693370_129693371	*TRH*
400	Chr3:_129693385_129693386	*TRH*
401	Chr3:_129693586_129693587	*TRH*
402	Chr3:_132757087_132757088	*TMEM108*
403	Chr3:_132757106_132757107	*TMEM108*
404	Chr3:_134369939_134369940	*KY*
405	Chr3:_138658554_138658555	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
406	Chr3:_138665984_138665985	*FOXL2,C3orf72*
407	Chr3:_138666054_138666055	*FOXL2,C3orf72*
408	Chr3:_138666144_138666145	*FOXL2,C3orf72*
409	Chr3:_138679512_138679513	*Non Coding Region*
410	Chr3:_138679578_138679579	*Non Coding Region*
411	Chr3:_139258939_139258940	*RBP1*
412	Chr3:_139653872_139653873	*CLSTN2*
413	Chr3:_140770549_140770550	*SPSB4*
414	Chr3:_140770588_140770589	*SPSB4*
415	Chr3:_140770599_140770600	*SPSB4*
416	Chr3:_140770608_140770609	*SPSB4*
417	Chr3:_140771786_140771787	*SPSB4*
418	Chr3:_142682378_142682379	*PAQR9*
419	Chr3:_142682652_142682653	*PAQR9*
420	Chr3:_142839022_142839023	*CHST2*
421	Chr3:_142839837_142839838	*CHST2*
422	Chr3:_142839991_142839992	*CHST2*
423	Chr3:_150102779_150102780	*Non Coding Region*
424	Chr3:_150803669_150803670	*MED12L*
425	Chr3:_150804063_150804064	*MED12L*
426	Chr3:_159756793_159756794	*Non Coding Region*
427	Chr3:_169384519_169384520	*Non Coding Region*
428	Chr3:_172165884_172165885	*GHSR*
429	Chr3:_172166517_172166518	*GHSR*
430	Chr3:_179168677_179168678	*GNB4*
431	Chr3:_179168760_179168761	*GNB4*
432	Chr3:_179754239_179754240	*PEX5L*
433	Chr3:_179754533_179754534	*PEX5L*
434	Chr3:_183273245_183273246	*KLHL6*
435	Chr3:_183273442_183273443	*KLHL6*
436	Chr3:_184301730_184301731	*Non Coding Region*
437	Chr3:_185080816_185080817	*MAP3K13*
438	Chr3:_185080905_185080906	*MAP3K13*
439	Chr3:_186079142_186079143	*DGKG*
440	Chr3:_192126849_192126850	*FGF12*
441	Chr3:_192126935_192126936	*FGF12*
442	Chr3:_194117679_194117680	*GP5*
443	Chr3:_194118985_194118986	*GP5*
444	Chr3:_194408516_194408517	*FAM43A*
445	Chr3:_194408845_194408846	*FAM43A*
446	Chr3:_195488725_195488726	*MUC4*
447	Chr3:_195623347_195623348	*TNK2*
448	Chr4:_570361_570362	*Non Coding Region*
449	Chr4:_1195318_1195319	*LOC100130872, LOC100130872-SPON2*
450	Chr4:_2788986_2788987	*Non Coding Region*

Figure 44E

Order	Coordinate of CpG (hg19)	Gene
451	Chr4:_2933715_2933716	*MFSD10*
452	Chr4:_4387917_4387918	*D4S234E*
453	Chr4:_4388505_4388506	*D4S234E*
454	Chr4:_5052526_5052527	*STK32B*
455	Chr4:_5053504_5053505	*STK32B*
456	Chr4:_5710319_5710320	*EVC2*
457	Chr4:_5710407_5710408	*EVC2*
458	Chr4:_5710411_5710412	*EVC2*
459	Chr4:_5712937_5712938	*EVC*
460	Chr4:_5892072_5892073	*CRMP1*
461	Chr4:_6201430_6201431	*JAKMIP1*
462	Chr4:_6202558_6202559	*JAKMIP1*
463	Chr4:_7195206_7195207	*SORCS2*
464	Chr4:_8037325_8037326	*ABLIM2*
465	Chr4:_10459219_10459220	*ZNF518B*
466	Chr4:_10686413_10686414	*CLNK*
467	Chr4:_13524143_13524144	*Non Coding Region*
468	Chr4:_13530269_13530270	*Non Coding Region*
469	Chr4:_13540946_13540947	*Non Coding Region*
470	Chr4:_13546151_13546152	*NKX3-2*
471	Chr4:_13546157_13546158	*NKX3-2*
472	Chr4:_15780306_15780307	*CD38*
473	Chr4:_24802340_24802341	*SOD3*
474	Chr4:_37246827_37246828	*KIAA1239*
475	Chr4:_40198392_40198393	*RHOH*
476	Chr4:_40198595_40198596	*RHOH*
477	Chr4:_40858984_40858985	*APBB2*
478	Chr4:_40859251_40859252	*APBB2*
479	Chr4:_40859280_40859281	*APBB2*
480	Chr4:_41258910_41258911	*UCHL1*
481	Chr4:_41883890_41883891	*Non Coding Region*
482	Chr4:_42399851_42399852	*SHISA3*
483	Chr4:_54965863_54965864	*GSX2*
484	Chr4:_55097576_55097577	*PDGFRA*
485	Chr4:_55991782_55991783	*KDR*
486	Chr4:_55991850_55991851	*KDR*
487	Chr4:_55991860_55991861	*KDR*
488	Chr4:_57522493_57522494	*HOPX*
489	Chr4:_57522632_57522633	*HOPX*
490	Chr4:_71860600_71860601	*DCK*
491	Chr4:_73434856_73434857	*ADAMTS3*
492	Chr4:_81106414_81106415	*PRDM8*
493	Chr4:_81187798_81187799	*FGF5*
494	Chr4:_84034390_84034391	*PLAC8*
495	Chr4:_89378894_89378895	*HERC5*

Order	Coordinate of CpG (hg19)	Gene
496	Chr4:_90758797_90758798	*SNCA*
497	Chr4:_90815973_90815974	*MMRN1*
498	Chr4:_96471105_96471106	*UNC5C*
499	Chr4:_101111261_101111262	*DDIT4L*
500	Chr4:_111555440_111555441	*PITX2*
501	Chr4:_122301816_122301817	*QRFPR*
502	Chr4:_122302226_122302227	*QRFPR*
503	Chr4:_122686432_122686433	*TMEM155, LOC100192379*
504	Chr4:_122686453_122686454	*TMEM155, LOC100192379*
505	Chr4:_122686456_122686457	*TMEM155, LOC100192379*
506	Chr4:_122686493_122686494	*TMEM155, LOC100192379*
507	Chr4:_123748386_123748387	*FGF2*
508	Chr4:_126237371_126237372	*FAT4*
509	Chr4:_126237373_126237374	*FAT4*
510	Chr4:_134067789_134067790	*Non Coding Region*
511	Chr4:_134067896_134067897	*Non Coding Region*
512	Chr4:_134068777_134068778	*Non Coding Region*
513	Chr4:_134070369_134070370	*PCDH10*
514	Chr4:_134070416_134070417	*PCDH10*
515	Chr4:_142054254_142054255	*RNF150*
516	Chr4:_144621898_144621899	*FREM3*
517	Chr4:_144621945_144621946	*FREM3*
518	Chr4:_144621952_144621953	*FREM3*
519	Chr4:_147569938_147569939	*Non Coding Region*
520	Chr4:_154681197_154681198	*RNF175*
521	Chr4:_154709878_154709879	*SFRP2*
522	Chr4:_154711070_154711071	*SFRP2*
523	Chr4:_154712422_154712423	*Non Coding Region*
524	Chr4:_155254497_155254498	*DCHS2*
525	Chr4:_156297854_156297855	*MAP9*
526	Chr4:_156680400_156680401	*GUCY1B3*
527	Chr4:_156680769_156680770	*GUCY1B3*
528	Chr4:_166794786_166794787	*TLL1*
529	Chr4:_166795524_166795525	*TLL1*
530	Chr4:_172734276_172734277	*GALNTL6*
531	Chr4:_174450408_174450409	*NBLA00301,HAND2*
532	Chr4:_174451443_174451444	*NBLA00301,HAND2*
533	Chr4:_175750637_175750638	*GLRA3*
534	Chr4:_176922779_176922780	*GPM6A*
535	Chr4:_176986950_176986951	*WDR17*
536	Chr4:_176987174_176987175	*WDR17*
537	Chr4:_183062301_183062302	*MGC45800*
538	Chr4:_183063459_183063460	*MGC45800*
539	Chr4:_183369754_183369755	*ODZ3*
540	Chr4:_184827754_184827755	*STOX2*

Figure 44F

Order	Coordinate of CpG (hg19)	Gene
541	Chr4:_185456125_185456126	*Non Coding Region*
542	Chr4:_185941601_185941602	*HELT*
543	Chr4:_186559412_186559413	*SORBS2*
544	Chr5:_528580_528581	*Non Coding Region*
545	Chr5:_1445567_1445568	*SLC6A3*
546	Chr5:_1667643_1667644	*Non Coding Region*
547	Chr5:_1875886_1875887	*Non Coding Region*
548	Chr5:_1883005_1883006	*IRX4*
549	Chr5:_1931620_1931621	*Non Coding Region*
550	Chr5:_2038899_2038900	*Non Coding Region*
551	Chr5:_2038922_2038923	*Non Coding Region*
552	Chr5:_3595728_3595729	*IRX1*
553	Chr5:_3596704_3596705	*IRX1*
554	Chr5:_3597487_3597488	*IRX1*
555	Chr5:_5139853_5139854	*ADAMTS16*
556	Chr5:_5139866_5139867	*ADAMTS16*
557	Chr5:_7850070_7850071	*C5orf49*
558	Chr5:_7850203_7850204	*C5orf49*
559	Chr5:_11904110_11904111	*CTNND2*
560	Chr5:_11904114_11904115	*CTNND2*
561	Chr5:_17218278_17218279	*BASP1,LOC285696*
562	Chr5:_31854651_31854652	*PDZD2*
563	Chr5:_31855185_31855186	*PDZD2*
564	Chr5:_32709200_32709201	*Non Coding Region*
565	Chr5:_32709301_32709302	*Non Coding Region*
566	Chr5:_32711429_32711430	*NPR3*
567	Chr5:_33936232_33936233	*RXFP3*
568	Chr5:_33936253_33936254	*RXFP3*
569	Chr5:_36690259_36690260	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
586	Chr5:_54516805_54516806	*Non Coding Region*
587	Chr5:_63802491_63802492	*RGS7BP*
588	Chr5:_63986392_63986393	*FAM159B*
589	Chr5:_63986656_63986657	*FAM159B*
590	Chr5:_63986793_63986794	*FAM159B*
591	Chr5:_71015523_71015524	*CARTPT*
592	Chr5:_71404593_71404594	*MAP1B*
593	Chr5:_72528296_72528297	*Non Coding Region*
594	Chr5:_72595685_72595686	*Non Coding Region*
595	Chr5:_72595741_72595742	*Non Coding Region*
596	Chr5:_76010658_76010659	*F2R*
597	Chr5:_82768488_82768489	*VCAN*
598	Chr5:_88180146_88180147	*MEF2C*
599	Chr5:_92909429_92909430	*FLJ42709*
600	Chr5:_92930813_92930814	*Non Coding Region*
601	Chr5:_94956017_94956018	*GPR150*
602	Chr5:_100236841_100236842	*ST8SIA4*
603	Chr5:_101632310_101632311	*SLCO4C1*
604	Chr5:_101632314_101632315	*SLCO4C1*
605	Chr5:_101632327_101632328	*SLCO4C1*
606	Chr5:_101632341_101632342	*SLCO4C1*
607	Chr5:_115152485_115152486	*CDO1*
608	Chr5:_127873980_127873981	*FBN2*
609	Chr5:_131792825_131792826	*C5orf56*
610	Chr5:_131991993_131991994	*Non Coding Region*
611	Chr5:_134363324_134363325	*Non Coding Region*
612	Chr5:_136834464_136834465	*SPOCK1*
613	Chr5:_136834492_136834493	*SPOCK1*
614	Chr5:_136834877_136834878	*SPOCK1*

Figure 44G

Order	Coordinate of CpG (hg19)	Gene	Order	Coordinate of CpG (hg19)	Gene
570	Chr5:_36690657_36690658	*Non Coding Region*	615	Chr5:_140787504_140787505	*PCDHGB1,PCDHGB3,PCDHGB2,PCDHGA8,PCDHGA9,PCDHGB6,PCDHGB5,PCDHGB4,PCDHGA2,PCDHGA3,PCDHGA1,PCDHGA6,PCDHGA7,PCDHGA4,PCDHGA5*
571	Chr5:_37834958_37834959	*GDNF*	616	Chr5:_140787507_140787508	*PCDHGB1,PCDHGB3,PCDHGB2,PCDHGA8,PCDHGA9,PCDHGB6,PCDHGB5,PCDHGB4,PCDHGA2,PCDHGA3,PCDHGA1,PCDHGA6,PCDHGA7,PCDHGA4,PCDHGA5*
572	Chr5:_37836748_37836749	*GDNF*	617	Chr5:_140797172_140797173	*PCDHGA10,PCDHGA8,PCDHGB3,PCDHGB2,PCDHGB1,PCDHGA9,PCDHGB7,PCDHGB6,PCDHGB5,PCDHGB4,PCDHGA2,PCDHGA3,PCDHGA1,PCDHGA6,PCDHGA7,PCDHGA4,PCDHGA5*
573	Chr5:_37839829_37839830	*GDNF*	618	Chr5:_140855562_140855563	*PCDHGC3,PCDHGA8,PCDHGA10,PCDHGA11,PCDHGA12,PCDHGB3,PCDHGB2,PCDHGB1,PCDHGA9,PCDHGB7,PCDHGB6,PCDHGB5,PCDHGB4,PCDHGA2,PCDHGA3,PCDHGA1,PCDHGA6,PCDHGA7,PCDHGA4,PCDHGA5*
574	Chr5:_37840075_37840076	*GDNF*	619	Chr5:_140855582_140855583	*PCDHGC3,PCDHGA8,PCDHGA10,PCDHGA11,PCDHGA12,PCDHGB3,PCDHGB2,PCDHGB1,PCDHGA9,PCDHGB7,PCDHGB6,PCDHGB5,PCDHGB4,PCDHGA2,PCDHGA3,PCDHGA1,PCDHGA6,PCDHGA7,PCDHGA4,PCDHGA5*
575	Chr5:_38258333_38258334	*EGFLAM*	620	Chr5:_141262801_141262802	*Non Coding Region*
576	Chr5:_38258884_38258885	*EGFLAM*	621	Chr5:_141395447_141395448	*Non Coding Region*
577	Chr5:_38556435_38556436	*LIFR*	622	Chr5:_145720024_145720025	*POU4F3*
578	Chr5:_42424525_42424526	*GHR*	623	Chr5:_146258546_146258547	*PPP2R2B*
579	Chr5:_42951192_42951193	*Non Coding Region*	624	Chr5:_150593684_150593685	*CCDC69*
580	Chr5:_42994709_42994710	*Non Coding Region*	625	Chr5:_156680241_156680242	*ITK*
581	Chr5:_44389406_44389407	*FGF10*	626	Chr5:_156696351_156696352	*CYFIP2*
582	Chr5:_49737182_49737183	*EMB*	627	Chr5:_156698854_156698855	*CYFIP2*
583	Chr5:_50265443_50265444	*Non Coding Region*	628	Chr5:_159933856_159933857	*Non Coding Region*
584	Chr5:_54397592_54397593	*GZMA*	629	Chr5:_169064024_169064025	*DOCK2*
585	Chr5:_54516487_54516488	*Non Coding Region*	630	Chr5:_169064451_169064452	*DOCK2*

Figure 44H

Order	Coordinate of CpG (hg19)	Gene
631	Chr5:_169533347_169533348	*FOXI1*
632	Chr5:_169724590_169724591	*LCP2*
633	Chr5:_169724768_169724769	*LCP2*
634	Chr5:_170738274_170738275	*TLX3*
635	Chr5:_170739179_170739180	*Non Coding Region*
636	Chr5:_170740870_170740871	*Non Coding Region*
637	Chr5:_171605778_171605779	*STK10*
638	Chr5:_172659730_172659731	*NKX2-5*
639	Chr5:_174159039_174159040	*Non Coding Region*
640	Chr5:_175223982_175223983	*CPLX2*
641	Chr5:_175792498_175792499	*ARL10*
642	Chr5:_175793049_175793050	*ARL10*
643	Chr5:_176799366_176799367	*RGS14*
644	Chr5:_177412718_177412719	*Non Coding Region*
645	Chr5:_178771776_178771777	*ADAMTS2*
646	Chr5:_180017349_180017350	*SCGB3A1*
647	Chr5:_180230767_180230768	*MGAT1*
648	Chr5:_180230959_180230960	*MGAT1*
649	Chr5:_180231026_180231027	*MGAT1*
650	Chr5:_180231058_180231059	*MGAT1*
651	Chr5:_180231084_180231085	*MGAT1*
652	Chr6:_391208_391209	*IRF4*
653	Chr6:_391764_391765	*IRF4*
654	Chr6:_391936_391937	*IRF4*
655	Chr6:_393239_393240	*IRF4*
656	Chr6:_393616_393617	*IRF4*
657	Chr6:_475074_475075	*Non Coding Region*
658	Chr6:_1379694_1379695	*Non Coding Region*
659	Chr6:_1391265_1391266	*FOXF2*
660	Chr6:_1625111_1625112	*GMDS*
661	Chr6:_6004207_6004208	*NRN1*
662	Chr6:_10382828_10382829	*Non Coding Region*
663	Chr6:_10391412_10391413	*Non Coding Region*
664	Chr6:_10422322_10422323	*Non Coding Region*
665	Chr6:_10882240_10882241	*GCM2*
666	Chr6:_11382553_11382554	*NEDD9*
667	Chr6:_11382556_11382557	*NEDD9*
668	Chr6:_11382769_11382770	*NEDD9*
669	Chr6:_13428522_13428523	*GFOD1*
670	Chr6:_13428657_13428658	*GFOD1*
671	Chr6:_13455261_13455262	*GFOD1*
672	Chr6:_14735175_14735176	*Non Coding Region*
673	Chr6:_25042090_25042091	*Non Coding Region*
674	Chr6:_25042495_25042496	*Non Coding Region*
675	Chr6:_25042497_25042498	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
676	Chr6:_26240528_26240529	*HIST1H4F*
677	Chr6:_26240920_26240921	*HIST1H4F*
678	Chr6:_26240939_26240940	*HIST1H4F*
679	Chr6:_27173753_27173754	*Non Coding Region*
680	Chr6:_27173991_27173992	*Non Coding Region*
681	Chr6:_27463347_27463348	*Non Coding Region*
682	Chr6:_28493661_28493662	*GPX5*
683	Chr6:_28510339_28510340	*Non Coding Region*
684	Chr6:_28603033_28603034	*Non Coding Region*
685	Chr6:_28603059_28603060	*Non Coding Region*
686	Chr6:_28603230_28603231	*Non Coding Region*
687	Chr6:_28603779_28603780	*Non Coding Region*
688	Chr6:_28956247_28956248	*Non Coding Region*
689	Chr6:_28956268_28956269	*Non Coding Region*
690	Chr6:_29521602_29521603	*Non Coding Region*
691	Chr6:_29521751_29521752	*Non Coding Region*
692	Chr6:_29521756_29521757	*Non Coding Region*
693	Chr6:_29595653_29595654	*GABBR1*
694	Chr6:_29760164_29760165	*HCG4*
695	Chr6:_29760410_29760411	*HCG4*
696	Chr6:_29760505_29760506	*HCG4*
697	Chr6:_29795815_29795816	*HLA-G*
698	Chr6:_29943480_29943481	*HCG9*
699	Chr6:_30095728_30095729	*Non Coding Region*
700	Chr6:_30139634_30139635	*TRIM15*
701	Chr6:_30139903_30139904	*TRIM15*
702	Chr6:_30227883_30227884	*HLA-L*
703	Chr6:_30418941_30418942	*Non Coding Region*
704	Chr6:_30418985_30418986	*Non Coding Region*
705	Chr6:_30419070_30419071	*Non Coding Region*
706	Chr6:_30649909_30649910	*KIAA1949*
707	Chr6:_30650844_30650845	*KIAA1949*
708	Chr6:_30652202_30652203	*KIAA1949*
709	Chr6:_30652376_30652377	*KIAA1949*
710	Chr6:_30652396_30652397	*KIAA1949*
711	Chr6:_30652399_30652400	*KIAA1949*
712	Chr6:_30652688_30652689	*KIAA1949*
713	Chr6:_31527889_31527890	*Non Coding Region*
714	Chr6:_31527893_31527894	*Non Coding Region*
715	Chr6:_31527920_31527921	*Non Coding Region*
716	Chr6:_31528154_31528155	*Non Coding Region*
717	Chr6:_31554199_31554200	*LST1*
718	Chr6:_31554548_31554549	*LST1*
719	Chr6:_31783322_31783323	*HSPA1A,HSPA1L*
720	Chr6:_31783455_31783456	*HSPA1A,HSPA1L*

Figure 44I

Order	Coordinate of CpG (hg19)	Gene
721	Chr6:_31783466_31783467	*HSPA1A,HSPA1L*
722	Chr6:_31783468_31783469	*HSPA1A,HSPA1L*
723	Chr6:_31783482_31783483	*HSPA1A,HSPA1L*
724	Chr6:_31831489_31831490	*SLC44A4,NEU1*
725	Chr6:_32813084_32813085	*TAP1,PSMB8*
726	Chr6:_32813441_32813442	*TAP1,PSMB8*
727	Chr6:_32813448_32813449	*TAP1,PSMB8*
728	Chr6:_32813522_32813523	*TAP1,PSMB8*
729	Chr6:_32813531_32813532	*TAP1,PSMB8*
730	Chr6:_32813544_32813545	*TAP1,PSMB8*
731	Chr6:_32813582_32813583	*TAP1,PSMB8*
732	Chr6:_34112818_34112819	*Non Coding Region*
733	Chr6:_39282393_39282394	*KCNK17*
734	Chr6:_41341601_41341602	*Non Coding Region*
735	Chr6:_41528395_41528396	*FOXP4*
736	Chr6:_41528502_41528503	*FOXP4*
737	Chr6:_41528531_41528532	*FOXP4*
738	Chr6:_41528750_41528751	*FOXP4*
739	Chr6:_41528785_41528786	*FOXP4*
740	Chr6:_41606439_41606440	*MDFI*
741	Chr6:_42072147_42072148	*C6orf132*
742	Chr6:_42738967_42738968	*Non Coding Region*
743	Chr6:_42739021_42739022	*Non Coding Region*
744	Chr6:_42739049_42739050	*Non Coding Region*
745	Chr6:_43253097_43253098	*TTBK1*
746	Chr6:_45387917_45387918	*RUNX2*
747	Chr6:_45387932_45387933	*RUNX2*
748	Chr6:_55444061_55444062	*HMGCLL1*
749	Chr6:_56819429_56819430	*BEND6,DST*
750	Chr6:_71666682_71666683	*B3GAT2*
751	Chr6:_73331114_73331115	*KCNQ5*
752	Chr6:_73331238_73331239	*KCNQ5*
753	Chr6:_73331405_73331406	*KCNQ5*
754	Chr6:_73332073_73332074	*KCNQ5*
755	Chr6:_84562892_84562893	*RIPPLY2*
756	Chr6:_85474028_85474029	*TBX18*
757	Chr6:_91321019_91321020	*Non Coding Region*
758	Chr6:_91321150_91321151	*Non Coding Region*
759	Chr6:_99282348_99282349	*POU3F2*
760	Chr6:_99284027_99284028	*POU3F2*
761	Chr6:_100912940_100912941	*SIM1*
762	Chr6:_100912946_100912947	*SIM1*
763	Chr6:_105389599_105389600	*Non Coding Region*
764	Chr6:_105584551_105584552	*BVES*
765	Chr6:_105584709_105584710	*BVES*

Order	Coordinate of CpG (hg19)	Gene
766	Chr6:_106429316_106429317	*Non Coding Region*
767	Chr6:_106429443_106429444	*Non Coding Region*
768	Chr6:_108488335_108488336	*NR2E1*
769	Chr6:_108489977_108489978	*NR2E1*
770	Chr6:_110679107_110679108	*C6orf186*
771	Chr6:_117086850_117086851	*FAM162B*
772	Chr6:_117086931_117086932	*FAM162B*
773	Chr6:_118228078_118228079	*SLC35F1*
774	Chr6:_119131733_119131734	*Non Coding Region*
775	Chr6:_124124275_124124276	*NKAIN2*
776	Chr6:_127440000_127440001	*RSPO3*
777	Chr6:_127440510_127440511	*RSPO3*
778	Chr6:_133562056_133562057	*EYA4*
779	Chr6:_134638871_134638872	*SGK1*
780	Chr6:_137814728_137814729	*OLIG3*
781	Chr6:_138145161_138145162	*Non Coding Region*
782	Chr6:_149803206_149803207	*ZC3H12D*
783	Chr6:_149803426_149803427	*ZC3H12D*
784	Chr6:_149803444_149803445	*ZC3H12D*
785	Chr6:_151562026_151562027	*AKAP12*
786	Chr6:_152129388_152129389	*ESR1*
787	Chr6:_152957910_152957911	*SYNE1*
788	Chr6:_152957999_152958000	*SYNE1*
789	Chr6:_153451251_153451252	*RGS17*
790	Chr6:_159590059_159590060	*FNDC1*
791	Chr6:_166582310_166582311	*T*
792	Chr6:_166582393_166582394	*T*
793	Chr6:_166666838_166666839	*Non Coding Region*
794	Chr6:_166721155_166721156	*PRR18*
795	Chr6:_167275809_167275810	*RPS6KA2*
796	Chr6:_168768014_168768015	*Non Coding Region*
797	Chr6:_168768113_168768114	*Non Coding Region*
798	Chr6:_168939666_168939667	*SMOC2*
799	Chr6:_170463397_170463398	*Non Coding Region*
800	Chr7:_641337_641338	*PRKAR1B*
801	Chr7:_644219_644220	*PRKAR1B*
802	Chr7:_644302_644303	*PRKAR1B*
803	Chr7:_1270558_1270559	*Non Coding Region*
804	Chr7:_1705861_1705862	*Non Coding Region*
805	Chr7:_1709580_1709581	*Non Coding Region*
806	Chr7:_1959989_1959990	*MAD1L1*
807	Chr7:_1960073_1960074	*MAD1L1*
808	Chr7:_2445727_2445728	*CHST12*
809	Chr7:_2445843_2445844	*CHST12*
810	Chr7:_3134557_3134558	*Non Coding Region*

Figure 44J

Order	Coordinate of CpG (hg19)	Gene
811	Chr7:_4848934_4848935	*RADIL*
812	Chr7:_4848939_4848940	*RADIL*
813	Chr7:_4922708_4922709	*RADIL*
814	Chr7:_6576152_6576153	*GRID2IP*
815	Chr7:_6576353_6576354	*GRID2IP*
816	Chr7:_8482614_8482615	*NXPH1*
817	Chr7:_19146032_19146033	*Non Coding Region*
818	Chr7:_19147157_19147158	*Non Coding Region*
819	Chr7:_19156086_19156087	*TWIST1*
820	Chr7:_19157193_19157194	*TWIST1*
821	Chr7:_19157263_19157264	*TWIST1*
822	Chr7:_19157296_19157297	*TWIST1*
823	Chr7:_19158647_19158648	*TWIST1*
824	Chr7:_19158664_19158665	*TWIST1*
825	Chr7:_19158747_19158748	*TWIST1*
826	Chr7:_19812570_19812571	*TMEM196*
827	Chr7:_21583304_21583305	*DNAH11*
828	Chr7:_21583438_21583439	*DNAH11*
829	Chr7:_24323799_24323800	*NPY*
830	Chr7:_24323840_24323841	*NPY*
831	Chr7:_27208285_27208286	*Non Coding Region*
832	Chr7:_30722114_30722115	*CRHR2*
833	Chr7:_30722320_30722321	*CRHR2*
834	Chr7:_30722327_30722328	*CRHR2*
835	Chr7:_32338150_32338151	*Non Coding Region*
836	Chr7:_32802765_32802766	*Non Coding Region*
837	Chr7:_35293759_35293760	*TBX20*
838	Chr7:_35296968_35296969	*Non Coding Region*
839	Chr7:_36764019_36764020	*AOAH*
840	Chr7:_36764140_36764141	*AOAH*
841	Chr7:_37487633_37487634	*ELMO1*
842	Chr7:_37488253_37488254	*ELMO1*
843	Chr7:_38670412_38670413	*AMPH*
844	Chr7:_38670804_38670805	*AMPH*
845	Chr7:_38670985_38670986	*AMPH*
846	Chr7:_38670987_38670988	*AMPH*
847	Chr7:_38671001_38671002	*AMPH*
848	Chr7:_41739742_41739743	*INHBA,LOC285954*
849	Chr7:_42267547_42267548	*GLI3*
850	Chr7:_42267719_42267720	*GLI3*
851	Chr7:_42267747_42267748	*GLI3*
852	Chr7:_44143993_44143994	*AEBP1*
853	Chr7:_45018658_45018659	*MYO1G*
854	Chr7:_45018789_45018790	*MYO1G*
855	Chr7:_45018849_45018850	*MYO1G*

Order	Coordinate of CpG (hg19)	Gene
856	Chr7:_45019005_45019006	*MYO1G*
857	Chr7:_45066738_45066739	*CCM2*
858	Chr7:_45066841_45066842	*CCM2*
859	Chr7:_45066858_45066859	*CCM2*
860	Chr7:_45066895_45066896	*CCM2*
861	Chr7:_45067057_45067058	*CCM2*
862	Chr7:_45067319_45067320	*CCM2*
863	Chr7:_45197441_45197442	*RAMP3*
864	Chr7:_49813031_49813032	*VWC2*
865	Chr7:_49813033_49813034	*VWC2*
866	Chr7:_49814435_49814436	*VWC2*
867	Chr7:_49815365_49815366	*VWC2*
868	Chr7:_50343883_50343884	*IKZF1*
869	Chr7:_50727932_50727933	*GRB10*
870	Chr7:_54609776_54609777	*VSTM2A*
871	Chr7:_66096806_66096807	*KCTD7*
872	Chr7:_69064093_69064094	*AUTS2*
873	Chr7:_69064801_69064802	*AUTS2*
874	Chr7:_70597058_70597059	*WBSCR17*
875	Chr7:_71800987_71800988	*CALN1*
876	Chr7:_71801793_71801794	*CALN1*
877	Chr7:_75443174_75443175	*CCL24*
878	Chr7:_86273718_86273719	*GRM3*
879	Chr7:_89748294_89748295	*DPY19L2P4*
880	Chr7:_93205136_93205137	*CALCR*
881	Chr7:_93519473_93519474	*TFPI2*
882	Chr7:_93519855_93519856	*TFPI2*
883	Chr7:_93520012_93520013	*TFPI2*
884	Chr7:_93520024_93520025	*TFPI2*
885	Chr7:_96650668_96650669	*DLX5*
886	Chr7:_96652115_96652116	*DLX5*
887	Chr7:_98246001_98246002	*NPTX2*
888	Chr7:_98467380_98467381	*TMEM130*
889	Chr7:_100488942_100488943	*ACHE*
890	Chr7:_101006063_101006064	*EMID2*
891	Chr7:_102066852_102066853	*PRKRIP1*
892	Chr7:_102067162_102067163	*Non Coding Region*
893	Chr7:_106505772_106505773	*PIK3CG*
894	Chr7:_108095719_108095720	*NRCAM*
895	Chr7:_112726538_112726539	*GPR85*
896	Chr7:_116964012_116964013	*WNT2*
897	Chr7:_127672458_127672459	*LRRC4,SND1*
898	Chr7:_129913310_129913311	*CPA2*
899	Chr7:_132262353_132262354	*PLXNA4*
900	Chr7:_134143971_134143972	*AKR1B1*

Figure 44K

Order	Coordinate of CpG (hg19)	Gene		Order	Coordinate of CpG (hg19)	Gene
901	Chr7:_142494773_142494774	*Non Coding Region*		946	Chr8:_23563925_23563926	*NKX2-6*
902	Chr7:_142494914_142494915	*Non Coding Region*		947	Chr8:_23564025_23564026	*NKX2-6*
903	Chr7:_142494953_142494954	*Non Coding Region*		948	Chr8:_23564156_23564157	*NKX2-6*
904	Chr7:_142494985_142494986	*Non Coding Region*		949	Chr8:_23564193_23564194	*NKX2-6*
905	Chr7:_143579951_143579952	*FAM115A*		950	Chr8:_24814137_24814138	*NEFL*
906	Chr7:_143580147_143580148	*FAM115A*		951	Chr8:_25901017_25901018	*EBF2*
907	Chr7:_145813008_145813009	*CNTNAP2*		952	Chr8:_25909326_25909327	*Non Coding Region*
908	Chr7:_149411652_149411653	*KRBA1*		953	Chr8:_25909599_25909600	*Non Coding Region*
909	Chr7:_149487338_149487339	*SSPO*		954	Chr8:_26723584_26723585	*ADRA1A*
910	Chr7:_149917263_149917264	*Non Coding Region*		955	Chr8:_30890617_30890618	*WRN,PURG*
911	Chr7:_150147922_150147923	*GIMAP8*		956	Chr8:_30890620_30890621	*WRN,PURG*
912	Chr7:_150147988_150147989	*GIMAP8*		957	Chr8:_30890632_30890633	*WRN,PURG*
913	Chr7:_150329449_150329450	*GIMAP6*		958	Chr8:_32405792_32405793	*NRG1*
914	Chr7:_150413801_150413802	*GIMAP1*		959	Chr8:_32406928_32406929	*NRG1*
915	Chr7:_150653001_150653002	*KCNH2*		960	Chr8:_37823409_37823410	*ADRB3*
916	Chr7:_151107257_151107258	*WDR86*		961	Chr8:_38578802_38578803	*Non Coding Region*
917	Chr7:_151107261_151107262	*WDR86*		962	Chr8:_38586066_38586067	*TACC1*
918	Chr7:_151107285_151107286	*WDR86*		963	Chr8:_38627889_38627890	*TACC1*
919	Chr7:_152622455_152622456	*Non Coding Region*		964	Chr8:_38628105_38628106	*TACC1*
920	Chr7:_155166290_155166291	*Non Coding Region*		965	Chr8:_38645408_38645409	*TACC1*
921	Chr7:_155167038_155167039	*Non Coding Region*		966	Chr8:_41167107_41167108	*SFRP1*
922	Chr7:_156798258_156798259	*MNX1*		967	Chr8:_41424982_41424983	*Non Coding Region*
923	Chr7:_157461310_157461311	*PTPRN2*		968	Chr8:_41754181_41754182	*ANK1*
924	Chr7:_157461368_157461369	*PTPRN2*		969	Chr8:_41755140_41755141	*ANK1*
925	Chr7:_158937494_158937495	*VIPR2*		970	Chr8:_48100410_48100411	*Non Coding Region*
926	Chr7:_158937610_158937611	*VIPR2*		971	Chr8:_49468828_49468829	*Non Coding Region*
927	Chr8:_494721_494722	*C8orf42*		972	Chr8:_49468988_49468989	*Non Coding Region*
928	Chr8:_688417_688418	*Non Coding Region*		973	Chr8:_53851156_53851157	*NPBWR1*
929	Chr8:_9760877_9760878	*LOC157627*		974	Chr8:_53852030_53852031	*NPBWR1*
930	Chr8:_9760880_9760881	*LOC157627*		975	Chr8:_53852184_53852185	*NPBWR1*
931	Chr8:_9761171_9761172	*MIR124-1,LOC157627*		976	Chr8:_53852422_53852423	*NPBWR1*
932	Chr8:_9762100_9762101	*MIR124-1,LOC157627*		977	Chr8:_53852446_53852447	*NPBWR1*
933	Chr8:_10587604_10587605	*SOX7*		978	Chr8:_54164161_54164162	*OPRK1*
934	Chr8:_11204681_11204682	*TDH*		979	Chr8:_54792627_54792628	*RGS20*
935	Chr8:_11205166_11205167	*TDH*		980	Chr8:_55367611_55367612	*Non Coding Region*
936	Chr8:_15397729_15397730	*TUSC3*		981	Chr8:_55367627_55367628	*Non Coding Region*
937	Chr8:_21645923_21645924	*GFRA2*		982	Chr8:_55372111_55372112	*SOX17*
938	Chr8:_21646662_21646663	*GFRA2*		983	Chr8:_55379248_55379249	*Non Coding Region*
939	Chr8:_21771252_21771253	*DOK2*		984	Chr8:_56852290_56852291	*LYN*
940	Chr8:_21771258_21771259	*DOK2*		985	Chr8:_57025662_57025663	*MOS*
941	Chr8:_21771278_21771279	*DOK2*		986	Chr8:_57069688_57069689	*Non Coding Region*
942	Chr8:_22960385_22960386	*TNFRSF10C*		987	Chr8:_57069907_57069908	*Non Coding Region*
943	Chr8:_22960572_22960573	*TNFRSF10C*		988	Chr8:_57070013_57070014	*Non Coding Region*
944	Chr8:_22960820_22960821	*TNFRSF10C*		989	Chr8:_58907821_58907822	*FAM110B*
945	Chr8:_22961088_22961089	*TNFRSF10C*		990	Chr8:_59058585_59058586	*FAM110B*

Figure 44L

Order	Coordinate of CpG (hg19)	Gene
991	Chr8:_59058656_59058657	*FAM110B*
992	Chr8:_59058660_59058661	*FAM110B*
993	Chr8:_65281496_65281497	*Non Coding Region*
994	Chr8:_65291513_65291514	*MIR124-2*
995	Chr8:_65492792_65492793	*BHLHE22*
996	Chr8:_65494451_65494452	*BHLHE22*
997	Chr8:_67344553_67344554	*ADHFE1*
998	Chr8:_67344556_67344557	*ADHFE1*
999	Chr8:_67344588_67344589	*ADHFE1*
1000	Chr8:_67344642_67344643	*ADHFE1*
1001	Chr8:_67344665_67344666	*ADHFE1*
1002	Chr8:_67344720_67344721	*ADHFE1*
1003	Chr8:_67344936_67344937	*ADHFE1*
1004	Chr8:_67345006_67345007	*ADHFE1*
1005	Chr8:_67875033_67875034	*Non Coding Region*
1006	Chr8:_67875261_67875262	*Non Coding Region*
1007	Chr8:_67940950_67940951	*LRRC67*
1008	Chr8:_69244510_69244511	*Non Coding Region*
1009	Chr8:_70982039_70982040	*PRDM14*
1010	Chr8:_72469507_72469508	*Non Coding Region*
1011	Chr8:_72754953_72754954	*MSC*
1012	Chr8:_72756155_72756156	*MSC*
1013	Chr8:_72756341_72756342	*MSC*
1014	Chr8:_72917350_72917351	*Non Coding Region*
1015	Chr8:_75897297_75897298	*CRISPLD1*
1016	Chr8:_77595321_77595322	*ZFHX4,LOC100192378*
1017	Chr8:_85097056_85097057	*RALYL*
1018	Chr8:_89339622_89339623	*MMP16*
1019	Chr8:_95229872_95229873	*CDH17*
1020	Chr8:_97158015_97158016	*GDF6*
1021	Chr8:_97158052_97158053	*GDF6*
1022	Chr8:_97170666_97170667	*GDF6*
1023	Chr8:_97171827_97171828	*GDF6*
1024	Chr8:_97172012_97172013	*GDF6*
1025	Chr8:_97172961_97172962	*GDF6*
1026	Chr8:_97173526_97173527	*GDF6*
1027	Chr8:_97505764_97505765	*SDC2*
1028	Chr8:_97506251_97506252	*SDC2*
1029	Chr8:_102149838_102149839	*Non Coding Region*
1030	Chr8:_104512858_104512859	*RIMS2*
1031	Chr8:_104513083_104513084	*RIMS2*
1032	Chr8:_104513909_104513910	*RIMS2*
1033	Chr8:_105235606_105235607	*RIMS2*
1034	Chr8:_105235943_105235944	*RIMS2*
1035	Chr8:_116660586_116660587	*TRPS1*

Order	Coordinate of CpG (hg19)	Gene
1036	Chr8:_116679763_116679764	*TRPS1*
1037	Chr8:_121137288_121137289	*COL14A1*
1038	Chr8:_129005526_129005527	*PVT1*
1039	Chr8:_129005567_129005568	*PVT1*
1040	Chr8:_132054555_132054556	*Non Coding Region*
1041	Chr8:_134072526_134072527	*TG,SLA*
1042	Chr8:_134072611_134072612	*TG,SLA*
1043	Chr8:_140715540_140715541	*KCNK9*
1044	Chr8:_140716991_140716992	*Non Coding Region*
1045	Chr8:_143532613_143532614	*Non Coding Region*
1046	Chr8:_143533499_143533500	*Non Coding Region*
1047	Chr8:_143533532_143533533	*Non Coding Region*
1048	Chr8:_143858636_143858637	*LYNX1*
1049	Chr8:_145106299_145106300	*OPLAH*
1050	Chr8:_145106438_145106439	*OPLAH*
1051	Chr8:_145106582_145106583	*OPLAH*
1052	Chr9:_6645686_6645687	*GLDC*
1053	Chr9:_37027384_37027385	*PAX5*
1054	Chr9:_77112896_77112897	*RORB*
1055	Chr9:_79633737_79633738	*FOXB2*
1056	Chr9:_80524371_80524372	*GNAQ*
1057	Chr9:_89763370_89763371	*C9orf170*
1058	Chr9:_92137791_92137792	*Non Coding Region*
1059	Chr9:_100611516_100611517	*Non Coding Region*
1060	Chr9:_100616469_100616470	*FOXE1*
1061	Chr9:_112810402_112810403	*PALM2-AKAP2,AKAP2*
1062	Chr9:_124461599_124461600	*DAB2IP*
1063	Chr9:_124461640_124461641	*DAB2IP*
1064	Chr9:_124462172_124462173	*DAB2IP*
1065	Chr9:_124462182_124462183	*DAB2IP*
1066	Chr9:_124889019_124889020	*Non Coding Region*
1067	Chr9:_130540941_130540942	*SH2D3C*
1068	Chr9:_132382433_132382434	*C9orf50*
1069	Chr9:_132650648_132650649	*FNBP1*
1070	Chr9:_132652889_132652890	*FNBP1*
1071	Chr9:_133536345_133536346	*Non Coding Region*
1072	Chr9:_134148648_134148649	*FAM78A*
1073	Chr9:_134151519_134151520	*FAM78A*
1074	Chr9:_135037323_135037324	*NTNG2*
1075	Chr9:_137160473_137160474	*Non Coding Region*
1076	Chr10:_1778844_1778845	*ADARB2*
1077	Chr10:_1779835_1779836	*ADARB2*
1078	Chr10:_7450355_7450356	*SFMBT2*
1079	Chr10:_7451822_7451823	*SFMBT2*
1080	Chr10:_7452598_7452599	*SFMBT2*

Figure 44M

Order	Coordinate of CpG (hg19)	Gene
1081	Chr10:_7708670_7708671	*ITIH5*
1082	Chr10:_8078038_8078039	*Non Coding Region*
1083	Chr10:_8096650_8096651	*FLJ45983,GATA3*
1084	Chr10:_8096991_8096992	*GATA3*
1085	Chr10:_8097689_8097690	*GATA3*
1086	Chr10:_8102583_8102584	*GATA3*
1087	Chr10:_11206221_11206222	*CUGBP2*
1088	Chr10:_11206320_11206321	*CUGBP2*
1089	Chr10:_11206792_11206793	*CUGBP2*
1090	Chr10:_11206813_11206814	*CUGBP2*
1091	Chr10:_11206868_11206869	*CUGBP2*
1092	Chr10:_11206870_11206871	*CUGBP2*
1093	Chr10:_11207437_11207438	*CUGBP2*
1094	Chr10:_11208339_11208340	*CUGBP2*
1095	Chr10:_14701815_14701816	*FAM107B*
1096	Chr10:_15761854_15761855	*ITGA8*
1097	Chr10:_17271006_17271007	*VIM*
1098	Chr10:_17271519_17271520	*VIM*
1099	Chr10:_22542024_22542025	*Non Coding Region*
1100	Chr10:_22625465_22625466	*Non Coding Region*
1101	Chr10:_23463094_23463095	*Non Coding Region*
1102	Chr10:_23982480_23982481	*KIAA1217*
1103	Chr10:_26727318_26727319	*APBB1IP*
1104	Chr10:_27548180_27548181	*Non Coding Region*
1105	Chr10:_29096857_29096858	*Non Coding Region*
1106	Chr10:_30338246_30338247	*KIAA1462*
1107	Chr10:_43947405_43947406	*ZNF487*
1108	Chr10:_44788812_44788813	*Non Coding Region*
1109	Chr10:_44880545_44880546	*CXCL12*
1110	Chr10:_48438724_48438725	*GDF10*
1111	Chr10:_48439229_48439230	*GDF10*
1112	Chr10:_50716478_50716479	*ERCC6*
1113	Chr10:_50817810_50817811	*SLC18A3,CHAT*
1114	Chr10:_52833909_52833910	*PRKG1*
1115	Chr10:_57390306_57390307	*Non Coding Region*
1116	Chr10:_64575442_64575443	*EGR2*
1117	Chr10:_70847399_70847400	*SRGN*
1118	Chr10:_70849854_70849855	*SRGN*
1119	Chr10:_73496014_73496015	*CDH23,C10orf105*
1120	Chr10:_81002218_81002219	*ZMIZ1*
1121	Chr10:_81002480_81002481	*ZMIZ1*
1122	Chr10:_82116571_82116572	*DYDC2,DYDC1*
1123	Chr10:_83636249_83636250	*NRG3*
1124	Chr10:_88123453_88123454	*GRID1*
1125	Chr10:_88126089_88126090	*GRID1*

Order	Coordinate of CpG (hg19)	Gene
1126	Chr10:_88126291_88126292	*GRID1*
1127	Chr10:_88126853_88126854	*GRID1*
1128	Chr10:_91295299_91295300	*SLC16A12*
1129	Chr10:_93393030_93393031	*PPP1R3C*
1130	Chr10:_94822908_94822909	*CYP26C1*
1131	Chr10:_100993583_100993584	*HPSE2*
1132	Chr10:_100993587_100993588	*HPSE2*
1133	Chr10:_101292766_101292767	*NKX2-3*
1134	Chr10:_102419245_102419246	*Non Coding Region*
1135	Chr10:_102484232_102484233	*Non Coding Region*
1136	Chr10:_102497666_102497667	*Non Coding Region*
1137	Chr10:_102508286_102508287	*PAX2*
1138	Chr10:_102893980_102893981	*TLX1*
1139	Chr10:_102894120_102894121	*TLX1*
1140	Chr10:_102894148_102894149	*TLX1*
1141	Chr10:_102899285_102899286	*Non Coding Region*
1142	Chr10:_102900365_102900366	*Non Coding Region*
1143	Chr10:_103603563_103603564	*KCNIP2*
1144	Chr10:_103603722_103603723	*KCNIP2*
1145	Chr10:_103603810_103603811	*KCNIP2*
1146	Chr10:_103603812_103603813	*KCNIP2*
1147	Chr10:_105036611_105036612	*INA*
1148	Chr10:_105036727_105036728	*INA*
1149	Chr10:_105452352_105452353	*SH3PXD2A*
1150	Chr10:_106088702_106088703	*ITPRIP*
1151	Chr10:_106088711_106088712	*ITPRIP*
1152	Chr10:_106089003_106089004	*ITPRIP*
1153	Chr10:_106398816_106398817	*Non Coding Region*
1154	Chr10:_106400667_106400668	*SORCS3*
1155	Chr10:_106400686_106400687	*SORCS3*
1156	Chr10:_106400702_106400703	*SORCS3*
1157	Chr10:_108924366_108924367	*SORCS1*
1158	Chr10:_118030970_118030971	*GFRA1*
1159	Chr10:_118031864_118031865	*GFRA1*
1160	Chr10:_118031870_118031871	*GFRA1*
1161	Chr10:_118032879_118032880	*GFRA1*
1162	Chr10:_118032892_118032893	*GFRA1*
1163	Chr10:_118032905_118032906	*GFRA1*
1164	Chr10:_118033115_118033116	*GFRA1*
1165	Chr10:_118033370_118033371	*GFRA1*
1166	Chr10:_118892581_118892582	*VAX1*
1167	Chr10:_118922887_118922888	*Non Coding Region*
1168	Chr10:_119000927_119000928	*SLC18A2*
1169	Chr10:_119292062_119292063	*EMX2OS*
1170	Chr10:_119295770_119295771	*EMX2OS*

Figure 44N

Order	Coordinate of CpG (hg19)	Gene
1171	Chr10:_119304487_119304488	*EMX2OS,EMX2*
1172	Chr10:_119310288_119310289	*Non Coding Region*
1173	Chr10:_121277385_121277386	*RGS10*
1174	Chr10:_121301765_121301766	*RGS10*
1175	Chr10:_123922970_123922971	*TACC2*
1176	Chr10:_123923066_123923067	*TACC2*
1177	Chr10:_123923081_123923082	*TACC2*
1178	Chr10:_123923273_123923274	*TACC2*
1179	Chr10:_123923881_123923882	*TACC2*
1180	Chr10:_131768928_131768929	*Non Coding Region*
1181	Chr10:_131769581_131769582	*Non Coding Region*
1182	Chr10:_133110244_133110245	*TCERG1L*
1183	Chr10:_133110349_133110350	*TCERG1L*
1184	Chr10:_134901294_134901295	*GPR123*
1185	Chr10:_134901297_134901298	*GPR123*
1186	Chr10:_135043492_135043493	*UTF1*
1187	Chr10:_135043506_135043507	*UTF1*
1188	Chr10:_135139100_135139101	*CALY*
1189	Chr11:_627152_627153	*SCT*
1190	Chr11:_627175_627176	*SCT*
1191	Chr11:_637170_637171	*DRD4*
1192	Chr11:_637173_637174	*DRD4*
1193	Chr11:_637175_637176	*DRD4*
1194	Chr11:_2161473_2161474	*IGF2,IGF2AS,INS-IGF2*
1195	Chr11:_2162183_2162184	*IGF2,IGF2AS,INS-IGF2*
1196	Chr11:_2162211_2162212	*IGF2,IGF2AS,INS-IGF2*
1197	Chr11:_2321770_2321771	*TSPAN32,C11orf21*
1198	Chr11:_2321780_2321781	*TSPAN32,C11orf21*
1199	Chr11:_2321800_2321801	*TSPAN32,C11orf21*
1200	Chr11:_2322642_2322643	*TSPAN32,C11orf21*
1201	Chr11:_2322781_2322782	*TSPAN32,C11orf21*
1202	Chr11:_2554562_2554563	*KCNQ1*
1203	Chr11:_6440065_6440066	*APBB1*
1204	Chr11:_6668214_6668215	*DCHS1*
1205	Chr11:_8040551_8040552	*Non Coding Region*
1206	Chr11:_8103017_8103018	*TUB*
1207	Chr11:_9623178_9623179	*Non Coding Region*
1208	Chr11:_10327870_10327871	*ADM*
1209	Chr11:_12030289_12030290	*DKK3*
1210	Chr11:_14926738_14926739	*Non Coding Region*
1211	Chr11:_15136456_15136457	*INSC*
1212	Chr11:_18813544_18813545	*PTPN5*
1213	Chr11:_19366714_19366715	*Non Coding Region*
1214	Chr11:_20185267_20185268	*Non Coding Region*
1215	Chr11:_20229517_20229518	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
1216	Chr11:_20618399_20618400	*Non Coding Region*
1217	Chr11:_20691126_20691127	*NELL1*
1218	Chr11:_20691161_20691162	*NELL1*
1219	Chr11:_20691341_20691342	*NELL1*
1220	Chr11:_30607068_30607069	*MPPED2*
1221	Chr11:_31821388_31821389	*PAX6*
1222	Chr11:_31827084_31827085	*PAX6*
1223	Chr11:_32355022_32355023	*Non Coding Region*
1224	Chr11:_32456912_32456913	*WT1,WIT1*
1225	Chr11:_32457124_32457125	*WT1,WIT1*
1226	Chr11:_32457127_32457128	*WT1,WIT1*
1227	Chr11:_32457158_32457159	*WT1,WIT1*
1228	Chr11:_32457162_32457163	*WT1,WIT1*
1229	Chr11:_33890371_33890372	*LMO2*
1230	Chr11:_35441088_35441089	*SLC1A2*
1231	Chr11:_43602847_43602848	*MIR129-2*
1232	Chr11:_43602879_43602880	*MIR129-2*
1233	Chr11:_43963907_43963908	*LOC387763*
1234	Chr11:_44332653_44332654	*ALX4*
1235	Chr11:_44332664_44332665	*ALX4*
1236	Chr11:_46382725_46382726	*DGKZ*
1237	Chr11:_46383066_46383067	*DGKZ*
1238	Chr11:_46383209_46383210	*DGKZ*
1239	Chr11:_46414626_46414627	*Non Coding Region*
1240	Chr11:_47416487_47416488	*Non Coding Region*
1241	Chr11:_47416535_47416536	*Non Coding Region*
1242	Chr11:_57250469_57250470	*Non Coding Region*
1243	Chr11:_58343318_58343319	*LPXN*
1244	Chr11:_60738995_60738996	*CD6*
1245	Chr11:_60739005_60739006	*CD6*
1246	Chr11:_60739019_60739020	*CD6*
1247	Chr11:_60739172_60739173	*CD6*
1248	Chr11:_60739178_60739179	*CD6*
1249	Chr11:_60739183_60739184	*CD6*
1250	Chr11:_61283968_61283969	*SYT7*
1251	Chr11:_61322922_61322923	*SYT7*
1252	Chr11:_61595807_61595808	*FADS2*
1253	Chr11:_61596307_61596308	*FADS2*
1254	Chr11:_63321562_63321563	*HRASLS2*
1255	Chr11:_63768032_63768033	*MACROD1*
1256	Chr11:_63828713_63828714	*MACROD1*
1257	Chr11:_63974860_63974861	*FERMT3*
1258	Chr11:_63974955_63974956	*FERMT3*
1259	Chr11:_64059054_64059055	*KCNK4*
1260	Chr11:_64059307_64059308	*KCNK4*

Figure 44O

Order	Coordinate of CpG (hg19)	Gene
1261	Chr11:_64108241_64108242	*CCDC88B*
1262	Chr11:_64621386_64621387	*EHD1*
1263	Chr11:_65407734_65407735	*SIPA1*
1264	Chr11:_65408445_65408446	*SIPA1*
1265	Chr11:_65408496_65408497	*SIPA1*
1266	Chr11:_65601328_65601329	*SNX32*
1267	Chr11:_65661516_65661517	*FOSL1*
1268	Chr11:_67039119_67039120	*ADRBK1*
1269	Chr11:_67171191_67171192	*TBC1D10C*
1270	Chr11:_67171200_67171201	*TBC1D10C*
1271	Chr11:_67171476_67171477	*TBC1D10C*
1272	Chr11:_67171585_67171586	*TBC1D10C*
1273	Chr11:_67177034_67177035	*TBC1D10C*
1274	Chr11:_67177103_67177104	*TBC1D10C*
1275	Chr11:_69632335_69632336	*FGF3*
1276	Chr11:_69634372_69634373	*FGF3*
1277	Chr11:_69704700_69704701	*Non Coding Region*
1278	Chr11:_69830611_69830612	*Non Coding Region*
1279	Chr11:_69919990_69919991	*Non Coding Region*
1280	Chr11:_70211301_70211302	*PPFIA1*
1281	Chr11:_70211531_70211532	*PPFIA1*
1282	Chr11:_70961872_70961873	*Non Coding Region*
1283	Chr11:_71955332_71955333	*PHOX2A*
1284	Chr11:_71955599_71955600	*PHOX2A*
1285	Chr11:_72295535_72295536	*PDE2A*
1286	Chr11:_75379864_75379865	*MAP6*
1287	Chr11:_78673073_78673074	*ODZ4*
1288	Chr11:_78673100_78673101	*ODZ4*
1289	Chr11:_91958458_91958459	*Non Coding Region*
1290	Chr11:_91958799_91958800	*Non Coding Region*
1291	Chr11:_94501384_94501385	*AMOTL1*
1292	Chr11:_94501387_94501388	*AMOTL1*
1293	Chr11:_94501461_94501462	*AMOTL1*
1294	Chr11:_94501467_94501468	*AMOTL1*
1295	Chr11:_94501718_94501719	*AMOTL1*
1296	Chr11:_104034619_104034620	*PDGFD*
1297	Chr11:_104034754_104034755	*PDGFD*
1298	Chr11:_105481319_105481320	*GRIA4*
1299	Chr11:_105481322_105481323	*GRIA4*
1300	Chr11:_105481406_105481407	*GRIA4*
1301	Chr11:_106889334_106889335	*GUCY1A2*
1302	Chr11:_106889364_106889365	*GUCY1A2*
1303	Chr11:_107462437_107462438	*ELMOD1,LOC643923*
1304	Chr11:_111383515_111383516	*BTG4,MIR34C,MIR34B*
1305	Chr11:_111809582_111809583	*DIXDC1*

Order	Coordinate of CpG (hg19)	Gene
1306	Chr11:_112833773_112833774	*NCAM1*
1307	Chr11:_115531033_115531034	*Non Coding Region*
1308	Chr11:_115630889_115630890	*Non Coding Region*
1309	Chr11:_115631116_115631117	*Non Coding Region*
1310	Chr11:_117801492_117801493	*TMPRSS13*
1311	Chr11:_119293385_119293386	*THY1*
1312	Chr11:_119293601_119293602	*THY1*
1313	Chr11:_119293869_119293870	*THY1*
1314	Chr11:_120435056_120435057	*Non Coding Region*
1315	Chr11:_122848098_122848099	*Non Coding Region*
1316	Chr11:_123066560_123066561	*ASAM*
1317	Chr11:_123301171_123301172	*Non Coding Region*
1318	Chr11:_124629580_124629581	*ESAM*
1319	Chr11:_124735105_124735106	*ROBO3*
1320	Chr11:_124790875_124790876	*HEPACAM*
1321	Chr11:_125036385_125036386	*PKNOX2*
1322	Chr11:_125365004_125365005	*FEZ1*
1323	Chr11:_125774082_125774083	*PUS3,DDX25*
1324	Chr11:_126226007_126226008	*ST3GAL4*
1325	Chr11:_128563027_128563028	*FLI1*
1326	Chr11:_128563685_128563686	*FLI1*
1327	Chr11:_128564106_128564107	*FLI1*
1328	Chr11:_128564180_128564181	*FLI1*
1329	Chr11:_128564756_128564757	*FLI1*
1330	Chr11:_128564874_128564875	*FLI1*
1331	Chr11:_132934554_132934555	*OPCML*
1332	Chr11:_133825519_133825520	*IGSF9B*
1333	Chr11:_133906693_133906694	*LOC100128239*
1334	Chr11:_134146253_134146254	*GLB1L3*
1335	Chr11:_134146676_134146677	*GLB1L3*
1336	Chr12:_5540116_5540117	*NTF3*
1337	Chr12:_6571347_6571348	*TAPBPL*
1338	Chr12:_6664872_6664873	*IFFO1*
1339	Chr12:_6665108_6665109	*IFFO1*
1340	Chr12:_6665288_6665289	*IFFO1*
1341	Chr12:_6665330_6665331	*IFFO1*
1342	Chr12:_6665370_6665371	*IFFO1*
1343	Chr12:_6665424_6665425	*IFFO1*
1344	Chr12:_6881595_6881596	*LAG3*
1345	Chr12:_6881601_6881602	*LAG3*
1346	Chr12:_6881624_6881625	*LAG3*
1347	Chr12:_6881629_6881630	*LAG3*
1348	Chr12:_7060187_7060188	*PTPN6*
1349	Chr12:_7060263_7060264	*PTPN6*
1350	Chr12:_7060386_7060387	*PTPN6*

Figure 44P

Order	Coordinate of CpG (hg19)	Gene
1351	Chr12:_7060569_7060570	*PTPN6*
1352	Chr12:_8542839_8542840	*LOC389634*
1353	Chr12:_15113093_15113094	*ARHGDIB*
1354	Chr12:_15374303_15374304	*RERG*
1355	Chr12:_15374457_15374458	*RERG*
1356	Chr12:_15374560_15374561	*RERG*
1357	Chr12:_15374577_15374578	*RERG*
1358	Chr12:_15475342_15475343	*PTPRO*
1359	Chr12:_15475690_15475691	*PTPRO*
1360	Chr12:_24715478_24715479	*SOX5*
1361	Chr12:_24715864_24715865	*SOX5*
1362	Chr12:_25055262_25055263	*BCAT1*
1363	Chr12:_25055967_25055968	*BCAT1*
1364	Chr12:_29936174_29936175	*TMTC1*
1365	Chr12:_39299326_39299327	*CPNE8*
1366	Chr12:_39299364_39299365	*CPNE8*
1367	Chr12:_41086365_41086366	*CNTN1*
1368	Chr12:_45444895_45444896	*DBX2*
1369	Chr12:_45445115_45445116	*DBX2*
1370	Chr12:_46609063_46609064	*SLC38A1*
1371	Chr12:_47609357_47609358	*LOC100233209, FAM113B*
1372	Chr12:_47610220_47610221	*LOC100233209, FAM113B*
1373	Chr12:_48207862_48207863	*HDAC7*
1374	Chr12:_48398056_48398057	*COL2A1*
1375	Chr12:_48398370_48398371	*COL2A1*
1376	Chr12:_48398374_48398375	*COL2A1*
1377	Chr12:_48398380_48398381	*COL2A1*
1378	Chr12:_48591994_48591995	*Non Coding Region*
1379	Chr12:_49484169_49484170	*DHH*
1380	Chr12:_50297581_50297582	*FAIM2*
1381	Chr12:_50297777_50297778	*FAIM2*
1382	Chr12:_50297793_50297794	*FAIM2*
1383	Chr12:_50297945_50297946	*FAIM2*
1384	Chr12:_51717865_51717866	*BIN2*
1385	Chr12:_51717960_51717961	*BIN2*
1386	Chr12:_51717978_51717979	*BIN2*
1387	Chr12:_51718011_51718012	*BIN2*
1388	Chr12:_51718088_51718089	*BIN2*
1389	Chr12:_51718108_51718109	*BIN2*
1390	Chr12:_51718112_51718113	*BIN2*
1391	Chr12:_52400907_52400908	*GRASP*
1392	Chr12:_52401214_52401215	*GRASP*
1393	Chr12:_52627047_52627048	*KRT7*
1394	Chr12:_54088972_54088973	*Non Coding Region*
1395	Chr12:_54345862_54345863	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
1396	Chr12:_54393460_54393461	*HOXC9*
1397	Chr12:_54408664_54408665	*Non Coding Region*
1398	Chr12:_54424679_54424680	*HOXC5,HOXC4*
1399	Chr12:_54447584_54447585	*HOXC4*
1400	Chr12:_54447873_54447874	*HOXC4*
1401	Chr12:_54812000_54812001	*ITGA5*
1402	Chr12:_54812085_54812086	*ITGA5*
1403	Chr12:_54812606_54812607	*ITGA5*
1404	Chr12:_54891491_54891492	*NCKAP1L*
1405	Chr12:_54891634_54891635	*NCKAP1L*
1406	Chr12:_54891636_54891637	*NCKAP1L*
1407	Chr12:_56415271_56415272	*IKZF4*
1408	Chr12:_57873704_57873705	*ARHGAP9*
1409	Chr12:_57998762_57998763	*DTX3*
1410	Chr12:_57998786_57998787	*DTX3*
1411	Chr12:_58021917_58021918	*B4GALNT1*
1412	Chr12:_58026010_58026011	*B4GALNT1*
1413	Chr12:_62585031_62585032	*FAM19A2*
1414	Chr12:_62585204_62585205	*FAM19A2*
1415	Chr12:_62585877_62585878	*FAM19A2*
1416	Chr12:_62586267_62586268	*FAM19A2*
1417	Chr12:_64062498_64062499	*DPY19L2*
1418	Chr12:_64062564_64062565	*DPY19L2*
1419	Chr12:_64062724_64062725	*DPY19L2*
1420	Chr12:_64784483_64784484	*C12orf56*
1421	Chr12:_65515276_65515277	*WIF1*
1422	Chr12:_65515290_65515291	*WIF1*
1423	Chr12:_65515470_65515471	*WIF1*
1424	Chr12:_66123127_66123128	*Non Coding Region*
1425	Chr12:_66275988_66275989	*HMGA2*
1426	Chr12:_72667398_72667399	*LOC283392,TRHDE*
1427	Chr12:_72667493_72667494	*LOC283392,TRHDE*
1428	Chr12:_81471754_81471755	*ACSS3*
1429	Chr12:_81471884_81471885	*ACSS3*
1430	Chr12:_85305408_85305409	*SLC6A15*
1431	Chr12:_85667353_85667354	*Non Coding Region*
1432	Chr12:_95941988_95941989	*USP44*
1433	Chr12:_103889772_103889773	*C12orf42*
1434	Chr12:_103889789_103889790	*C12orf42*
1435	Chr12:_104444006_104444007	*GLT8D2*
1436	Chr12:_104850326_104850327	*CHST11*
1437	Chr12:_104850759_104850760	*CHST11*
1438	Chr12:_104850767_104850768	*CHST11*
1439	Chr12:_104851909_104851910	*CHST11*
1440	Chr12:_104852439_104852440	*CHST11*

Figure 44Q

Order	Coordinate of CpG (hg19)	Gene
1441	Chr12:_105114399_105114400	*CHST11*
1442	Chr12:_109028385_109028386	*SELPLG*
1443	Chr12:_111471192_111471193	*CUX2*
1444	Chr12:_111472183_111472184	*CUX2*
1445	Chr12:_113013048_113013049	*Non Coding Region*
1446	Chr12:_113229374_113229375	*RPH3A*
1447	Chr12:_113494422_113494423	*DTX1*
1448	Chr12:_113541915_113541916	*RASAL1*
1449	Chr12:_113901210_113901211	*LHX5*
1450	Chr12:_113901529_113901530	*LHX5*
1451	Chr12:_114075881_114075882	*Non Coding Region*
1452	Chr12:_114843296_114843297	*TBX5*
1453	Chr12:_115103960_115103961	*Non Coding Region*
1454	Chr12:_115105290_115105291	*Non Coding Region*
1455	Chr12:_117798954_117798955	*NOS1*
1456	Chr12:_120032374_120032375	*TMEM233*
1457	Chr12:_120799725_120799726	*MSI1*
1458	Chr12:_122096667_122096668	*MORN3*
1459	Chr12:_122227872_122227873	*RHOF*
1460	Chr12:_124941423_124941424	*NCOR2*
1461	Chr12:_124941871_124941872	*NCOR2*
1462	Chr12:_124942016_124942017	*NCOR2*
1463	Chr12:_124942176_124942177	*NCOR2*
1464	Chr12:_124950784_124950785	*NCOR2*
1465	Chr12:_124950938_124950939	*NCOR2*
1466	Chr12:_130389010_130389011	*TMEM132D*
1467	Chr12:_133464327_133464328	*CHFR*
1468	Chr12:_133464857_133464858	*CHFR*
1469	Chr12:_133481390_133481391	*Non Coding Region*
1470	Chr12:_133484853_133484854	*Non Coding Region*
1471	Chr13:_20768560_20768561	*GJB2*
1472	Chr13:_20806732_20806733	*GJB6*
1473	Chr13:_23733862_23733863	*Non Coding Region*
1474	Chr13:_25621027_25621028	*Non Coding Region*
1475	Chr13:_25745784_25745785	*FAM123A*
1476	Chr13:_28366759_28366760	*GSX1*
1477	Chr13:_28497117_28497118	*PDX1*
1478	Chr13:_28498384_28498385	*PDX1*
1479	Chr13:_28500882_28500883	*Non Coding Region*
1480	Chr13:_28503373_28503374	*Non Coding Region*
1481	Chr13:_28674451_28674452	*FLT3*
1482	Chr13:_29107069_29107070	*Non Coding Region*
1483	Chr13:_36705446_36705447	*DCLK1*
1484	Chr13:_36919674_36919675	*SPG20*
1485	Chr13:_36919819_36919820	*SPG20*

Order	Coordinate of CpG (hg19)	Gene
1486	Chr13:_36920332_36920333	*SPG20*
1487	Chr13:_37004617_37004618	*CCNA1*
1488	Chr13:_37005501_37005502	*CCNA1*
1489	Chr13:_37005570_37005571	*CCNA1*
1490	Chr13:_37006107_37006108	*CCNA1*
1491	Chr13:_37006127_37006128	*CCNA1*
1492	Chr13:_37006611_37006612	*CCNA1*
1493	Chr13:_43148935_43148936	*TNFSF11*
1494	Chr13:_44360259_44360260	*ENOX1*
1495	Chr13:_46425508_46425509	*SIAH3*
1496	Chr13:_46756209_46756210	*LCP1*
1497	Chr13:_47468154_47468155	*HTR2A*
1498	Chr13:_50070550_50070551	*PHF11*
1499	Chr13:_53313174_53313175	*LECT1*
1500	Chr13:_53313529_53313530	*LECT1*
1501	Chr13:_53420425_53420426	*PCDH8*
1502	Chr13:_58204224_58204225	*Non Coding Region*
1503	Chr13:_78493066_78493067	*EDNRB*
1504	Chr13:_78493100_78493101	*EDNRB*
1505	Chr13:_78493305_78493306	*EDNRB*
1506	Chr13:_78493313_78493314	*EDNRB*
1507	Chr13:_78493564_78493565	*EDNRB*
1508	Chr13:_79170146_79170147	*Non Coding Region*
1509	Chr13:_79170303_79170304	*Non Coding Region*
1510	Chr13:_84456171_84456172	*SLITRK1*
1511	Chr13:_84456258_84456259	*SLITRK1*
1512	Chr13:_88325384_88325385	*SLITRK5*
1513	Chr13:_92051400_92051401	*GPC5*
1514	Chr13:_95364675_95364676	*SOX21*
1515	Chr13:_96204854_96204855	*CLDN10*
1516	Chr13:_99880947_99880948	*UBAC2*
1517	Chr13:_99910587_99910588	*UBAC2,GPR18*
1518	Chr13:_100310841_100310842	*CLYBL*
1519	Chr13:_102569815_102569816	*FGF14*
1520	Chr13:_102569876_102569877	*FGF14*
1521	Chr13:_103046930_103046931	*FGF14*
1522	Chr13:_103047053_103047054	*FGF14*
1523	Chr13:_103047225_103047226	*FGF14*
1524	Chr13:_103052828_103052829	*FGF14*
1525	Chr13:_108519318_108519319	*FAM155A*
1526	Chr13:_109149011_109149012	*Non Coding Region*
1527	Chr13:_110958977_110958978	*COL4A2,COL4A1*
1528	Chr13:_110993493_110993494	*COL4A2*
1529	Chr13:_112758491_112758492	*Non Coding Region*
1530	Chr13:_114896683_114896684	*RASA3*

Figure 44R

Order	Coordinate of CpG (hg19)	Gene
1531	Chr14:_22004993_22004994	*SALL2*
1532	Chr14:_22694547_22694548	*Non Coding Region*
1533	Chr14:_23834977_23834978	*EFS*
1534	Chr14:_24803873_24803874	*ADCY4*
1535	Chr14:_24803903_24803904	*ADCY4*
1536	Chr14:_24803917_24803918	*ADCY4*
1537	Chr14:_24803925_24803926	*ADCY4*
1538	Chr14:_24804339_24804340	*ADCY4*
1539	Chr14:_29254680_29254681	*C14orf23*
1540	Chr14:_36003741_36003742	*INSM2*
1541	Chr14:_36989550_36989551	*NKX2-1*
1542	Chr14:_48145108_48145109	*MDGA2*
1543	Chr14:_51561293_51561294	*TRIM9*
1544	Chr14:_52536436_52536437	*NID2*
1545	Chr14:_54687160_54687161	*Non Coding Region*
1546	Chr14:_54687314_54687315	*Non Coding Region*
1547	Chr14:_57283436_57283437	*OTX2OS1*
1548	Chr14:_60975964_60975965	*SIX6*
1549	Chr14:_60976285_60976286	*SIX6*
1550	Chr14:_60978161_60978162	*SIX6*
1551	Chr14:_70014873_70014874	*Non Coding Region*
1552	Chr14:_70041283_70041284	*Non Coding Region*
1553	Chr14:_70653719_70653720	*SLC8A3*
1554	Chr14:_70653919_70653920	*SLC8A3*
1555	Chr14:_70653964_70653965	*SLC8A3*
1556	Chr14:_75800941_75800942	*Non Coding Region*
1557	Chr14:_75801023_75801024	*Non Coding Region*
1558	Chr14:_75988800_75988801	*BATF*
1559	Chr14:_75988820_75988821	*BATF*
1560	Chr14:_77504561_77504562	*Non Coding Region*
1561	Chr14:_77736904_77736905	*NGB*
1562	Chr14:_77737146_77737147	*NGB*
1563	Chr14:_85996202_85996203	*FLRT2*
1564	Chr14:_92413706_92413707	*FBLN5*
1565	Chr14:_93532563_93532564	*ITPK1,NCRNA00203*
1566	Chr14:_93532632_93532633	*ITPK1,NCRNA00203*
1567	Chr14:_94254726_94254727	*PRIMA1*
1568	Chr14:_95235026_95235027	*GSC*
1569	Chr14:_95235125_95235126	*GSC*
1570	Chr14:_95235127_95235128	*GSC*
1571	Chr14:_95236123_95236124	*GSC*
1572	Chr14:_96342250_96342251	*Non Coding Region*
1573	Chr14:_97685386_97685387	*Non Coding Region*
1574	Chr14:_97685576_97685577	*Non Coding Region*
1575	Chr14:_100259341_100259342	*EML1*
1576	Chr14:_100438647_100438648	*Non Coding Region*
1577	Chr14:_100438737_100438738	*Non Coding Region*
1578	Chr14:_100438957_100438958	*Non Coding Region*
1579	Chr14:_100531591_100531592	*EVL*
1580	Chr14:_101924596_101924597	*Non Coding Region*
1581	Chr14:_105310421_105310422	*Non Coding Region*
1582	Chr14:_105393171_105393172	*PLD4*
1583	Chr15:_26108263_26108264	*ATP10A*
1584	Chr15:_26108796_26108797	*ATP10A*
1585	Chr15:_27216088_27216089	*GABRG3*
1586	Chr15:_28342183_28342184	*OCA2*
1587	Chr15:_28352098_28352099	*Non Coding Region*
1588	Chr15:_28983117_28983118	*WHAMML2*
1589	Chr15:_29077493_29077494	*Non Coding Region*
1590	Chr15:_34806592_34806593	*Non Coding Region*
1591	Chr15:_34807050_34807051	*Non Coding Region*
1592	Chr15:_34807143_34807144	*Non Coding Region*
1593	Chr15:_37170381_37170382	*LOC145845*
1594	Chr15:_37170454_37170455	*LOC145845*
1595	Chr15:_37172509_37172510	*LOC145845*
1596	Chr15:_37387438_37387439	*MEIS2*
1597	Chr15:_37390284_37390285	*MEIS2*
1598	Chr15:_38988755_38988756	*C15orf53*
1599	Chr15:_41793881_41793882	*ITPKA*
1600	Chr15:_41794412_41794413	*ITPKA*
1601	Chr15:_45670865_45670866	*GATM,LOC145663*
1602	Chr15:_48938347_48938348	*FBN1*
1603	Chr15:_51385891_51385892	*TNFAIP8L3*
1604	Chr15:_53083532_53083533	*ONECUT1*
1605	Chr15:_53097777_53097778	*Non Coding Region*
1606	Chr15:_55881044_55881045	*PYGO1*
1607	Chr15:_60296996_60296997	*FOXB1*
1608	Chr15:_61520873_61520874	*RORA*
1609	Chr15:_65689298_65689299	*IGDCC4*
1610	Chr15:_68122139_68122140	*LBXCOR1*
1611	Chr15:_70828097_70828098	*Non Coding Region*
1612	Chr15:_73661652_73661653	*HCN4*
1613	Chr15:_74421535_74421536	*ISLR2,LOC283731*
1614	Chr15:_74726561_74726562	*SEMA7A*
1615	Chr15:_77287656_77287657	*PSTPIP1*
1616	Chr15:_78632572_78632573	*CRABP1*
1617	Chr15:_79383167_79383168	*RASGRF1*
1618	Chr15:_79383924_79383925	*RASGRF1*
1619	Chr15:_79575048_79575049	*ANKRD34C*
1620	Chr15:_79576216_79576217	*ANKRD34C*

Figure 44S

Order	Coordinate of CpG (hg19)	Gene
1621	Chr15:_81586571_81586572	*IL16*
1622	Chr15:_81589248_81589249	*IL16*
1623	Chr15:_83316640_83316641	*CPEB1*
1624	Chr15:_83776420_83776421	*TM6SF1*
1625	Chr15:_83776422_83776423	*TM6SF1*
1626	Chr15:_83877070_83877071	*HDGFRP3*
1627	Chr15:_83953744_83953745	*BNC1*
1628	Chr15:_84115811_84115812	*SH3GL3*
1629	Chr15:_84115895_84115896	*SH3GL3*
1630	Chr15:_84115969_84115970	*SH3GL3*
1631	Chr15:_86233220_86233221	*AKAP13*
1632	Chr15:_86233236_86233237	*AKAP13*
1633	Chr15:_88800567_88800568	*NTRK3*
1634	Chr15:_89922328_89922329	*LOC254559*
1635	Chr15:_89952499_89952500	*Non Coding Region*
1636	Chr15:_92398112_92398113	*SLCO3A1*
1637	Chr15:_92937735_92937736	*ST8SIA2*
1638	Chr15:_92937992_92937993	*ST8SIA2*
1639	Chr15:_93631754_93631755	*RGMA*
1640	Chr15:_96895341_96895342	*Non Coding Region*
1641	Chr15:_98836248_98836249	*Non Coding Region*
1642	Chr15:_101420636_101420637	*ALDH1A3*
1643	Chr16:_215960_215961	*HBM*
1644	Chr16:_2041512_2041513	*SYNGR3*
1645	Chr16:_2737356_2737357	*KCTD5*
1646	Chr16:_2765843_2765844	*PRSS27*
1647	Chr16:_3237991_3237992	*Non Coding Region*
1648	Chr16:_3627497_3627498	*NLRC3*
1649	Chr16:_3628412_3628413	*NLRC3*
1650	Chr16:_10133501_10133502	*GRIN2A*
1651	Chr16:_11327141_11327142	*Non Coding Region*
1652	Chr16:_12996046_12996047	*SHISA9*
1653	Chr16:_12996234_12996235	*SHISA9*
1654	Chr16:_16156039_16156040	*ABCC1*
1655	Chr16:_21294947_21294948	*CRYM*
1656	Chr16:_23846893_23846894	*PRKCB*
1657	Chr16:_23847325_23847326	*PRKCB*
1658	Chr16:_23848439_23848440	*PRKCB*
1659	Chr16:_25703528_25703529	*HS3ST4*
1660	Chr16:_25704505_25704506	*HS3ST4*
1661	Chr16:_25704507_25704508	*HS3ST4*
1662	Chr16:_28074384_28074385	*GSG1L*
1663	Chr16:_28074388_28074389	*GSG1L*
1664	Chr16:_28074462_28074463	*GSG1L*
1665	Chr16:_28074670_28074671	*GSG1L*
1666	Chr16:_28074980_28074981	*GSG1L*
1667	Chr16:_28075269_28075270	*GSG1L*
1668	Chr16:_28995882_28995883	*LAT*
1669	Chr16:_28996053_28996054	*LAT*
1670	Chr16:_28996329_28996330	*LAT*
1671	Chr16:_28996362_28996363	*LAT*
1672	Chr16:_28996449_28996450	*LAT*
1673	Chr16:_28996533_28996534	*LAT*
1674	Chr16:_29086271_29086272	*RRN3P2*
1675	Chr16:_29674618_29674619	*SPN*
1676	Chr16:_29674959_29674960	*SPN*
1677	Chr16:_29674972_29674973	*SPN*
1678	Chr16:_29675160_29675161	*SPN*
1679	Chr16:_29675846_29675847	*SPN*
1680	Chr16:_29676003_29676004	*SPN*
1681	Chr16:_29676071_29676072	*SPN*
1682	Chr16:_29756951_29756952	*C16orf54*
1683	Chr16:_29757318_29757319	*C16orf54*
1684	Chr16:_29757323_29757324	*C16orf54*
1685	Chr16:_29757334_29757335	*C16orf54*
1686	Chr16:_29757344_29757345	*C16orf54*
1687	Chr16:_29757375_29757376	*C16orf54*
1688	Chr16:_30197211_30197212	*CORO1A*
1689	Chr16:_30393854_30393855	*SEPT_1*
1690	Chr16:_30904573_30904574	*BCL7C,MIR762*
1691	Chr16:_31021933_31021934	*STX1B*
1692	Chr16:_31228059_31228060	*TRIM72,PYDC1*
1693	Chr16:_31228202_31228203	*TRIM72,PYDC1*
1694	Chr16:_49312033_49312034	*Non Coding Region*
1695	Chr16:_49312238_49312239	*CBLN1*
1696	Chr16:_49475615_49475616	*Non Coding Region*
1697	Chr16:_51184379_51184380	*SALL1*
1698	Chr16:_51184562_51184563	*SALL1*
1699	Chr16:_51184786_51184787	*SALL1*
1700	Chr16:_51186260_51186261	*SALL1*
1701	Chr16:_54407990_54407991	*Non Coding Region*
1702	Chr16:_54408014_54408015	*Non Coding Region*
1703	Chr16:_54970523_54970524	*Non Coding Region*
1704	Chr16:_55690130_55690131	*SLC6A2*
1705	Chr16:_55690381_55690382	*SLC6A2*
1706	Chr16:_56224901_56224902	*GNAO1,LOC283856*
1707	Chr16:_56228442_56228443	*GNAO1,DKFZP434H168*
1708	Chr16:_56228467_56228468	*GNAO1,DKFZP434H168*
1709	Chr16:_56228498_56228499	*GNAO1,DKFZP434H168*
1710	Chr16:_56697229_56697230	*Non Coding Region*

Figure 44T

Order	Coordinate of CpG (hg19)	Gene
1711	Chr16:_58019123_58019124	*TEPP*
1712	Chr16:_66612955_66612956	*CMTM2,CMTM1*
1713	Chr16:_66613266_66613267	*CMTM2*
1714	Chr16:_66638293_66638294	*CMTM3*
1715	Chr16:_66638412_66638413	*CMTM3*
1716	Chr16:_66638433_66638434	*CMTM3*
1717	Chr16:_66638438_66638439	*CMTM3*
1718	Chr16:_67197186_67197187	*FBXL8,HSF4*
1719	Chr16:_67428898_67428899	*ZDHHC1,TPPP3*
1720	Chr16:_77822419_77822420	*VAT1L*
1721	Chr16:_86541480_86541481	*Non Coding Region*
1722	Chr16:_86544339_86544340	*FOXF1*
1723	Chr16:_86599745_86599746	*FOXC2*
1724	Chr16:_86600326_86600327	*FOXC2*
1725	Chr16:_86601133_86601134	*FOXC2*
1726	Chr16:_86613067_86613068	*FOXL1*
1727	Chr16:_86962747_86962748	*Non Coding Region*
1728	Chr16:_87736062_87736063	*LOC100129637*
1729	Chr16:_87736388_87736389	*LOC100129637*
1730	Chr16:_88767261_88767262	*RNF166*
1731	Chr16:_88769854_88769855	*RNF166*
1732	Chr16:_88769970_88769971	*RNF166*
1733	Chr16:_88770558_88770559	*RNF166*
1734	Chr16:_90092755_90092756	*GAS8*
1735	Chr17:_1626575_1626576	*WDR81*
1736	Chr17:_4618322_4618323	*ARRB2*
1737	Chr17:_4802735_4802736	*C17orf107,CHRNE*
1738	Chr17:_4847601_4847602	*RNF167*
1739	Chr17:_5403879_5403880	*LOC728392*
1740	Chr17:_5404508_5404509	*LOC728392*
1741	Chr17:_5487376_5487377	*NLRP1*
1742	Chr17:_6616867_6616868	*SLC13A5*
1743	Chr17:_6616883_6616884	*SLC13A5*
1744	Chr17:_7240223_7240224	*ACAP1*
1745	Chr17:_7554681_7554682	*ATP1B2*
1746	Chr17:_7976456_7976457	*ALOX12B*
1747	Chr17:_8230350_8230351	*Non Coding Region*
1748	Chr17:_8533218_8533219	*MYH10*
1749	Chr17:_8533391_8533392	*MYH10*
1750	Chr17:_10024297_10024298	*GAS7*
1751	Chr17:_10101581_10101582	*GAS7*
1752	Chr17:_13505558_13505559	*HS3ST3A1*
1753	Chr17:_14212480_14212481	*HS3ST3B1*
1754	Chr17:_19483367_19483368	*Non Coding Region*
1755	Chr17:_19881632_19881633	*AKAP10*
1756	Chr17:_27038886_27038887	*PROCA1*
1757	Chr17:_27039032_27039033	*PROCA1*
1758	Chr17:_27940509_27940510	*ANKRD13B*
1759	Chr17:_32484259_32484260	*ACCN1*
1760	Chr17:_33701321_33701322	*SLFN11*
1761	Chr17:_33701350_33701351	*SLFN11*
1762	Chr17:_33867795_33867796	*Non Coding Region*
1763	Chr17:_35293930_35293931	*LHX1*
1764	Chr17:_35294476_35294477	*LHX1*
1765	Chr17:_35294481_35294482	*LHX1*
1766	Chr17:_35294491_35294492	*LHX1*
1767	Chr17:_37321490_37321491	*ARL5C*
1768	Chr17:_37914349_37914350	*Non Coding Region*
1769	Chr17:_38347603_38347604	*RAPGEFL1*
1770	Chr17:_38347816_38347817	*RAPGEFL1*
1771	Chr17:_38721675_38721676	*CCR7*
1772	Chr17:_42467307_42467308	*ITGA2B*
1773	Chr17:_43047733_43047734	*Non Coding Region*
1774	Chr17:_43299826_43299827	*FMNL1*
1775	Chr17:_43339223_43339224	*C17orf46, LOC100133991*
1776	Chr17:_43339328_43339329	*C17orf46, LOC100133991*
1777	Chr17:_43339497_43339498	*C17orf46, LOC100133991*
1778	Chr17:_43861104_43861105	*CRHR1*
1779	Chr17:_43862032_43862033	*CRHR1*
1780	Chr17:_44897431_44897432	*WNT3*
1781	Chr17:_44929245_44929246	*WNT9B*
1782	Chr17:_45810610_45810611	*TBX21*
1783	Chr17:_45810865_45810866	*TBX21*
1784	Chr17:_45867446_45867447	*Non Coding Region*
1785	Chr17:_46802888_46802889	*HOXB13*
1786	Chr17:_47074576_47074577	*IGF2BP1*
1787	Chr17:_47653306_47653307	*NXPH3*
1788	Chr17:_48070891_48070892	*DLX3*
1789	Chr17:_54675137_54675138	*Non Coding Region*
1790	Chr17:_55213563_55213564	*Non Coding Region*
1791	Chr17:_55952063_55952064	*CUEDC1*
1792	Chr17:_56408197_56408198	*Non Coding Region*
1793	Chr17:_56408688_56408689	*MIR142*
1794	Chr17:_56408694_56408695	*MIR142*
1795	Chr17:_56408804_56408805	*MIR142*
1796	Chr17:_56409518_56409519	*MIR142*
1797	Chr17:_59478068_59478069	*TBX2*
1798	Chr17:_59534998_59534999	*TBX4*
1799	Chr17:_66597500_66597501	*FAM20A*
1800	Chr17:_70216332_70216333	*Non Coding Region*

Figure 44U

Order	Coordinate of CpG (hg19)	Gene
1801	Chr17:_70723386_70723387	*SLC39A11*
1802	Chr17:_71161258_71161259	*SSTR2*
1803	Chr17:_72352897_72352898	*BTBD17*
1804	Chr17:_72588412_72588413	*CD300LD,C17orf77*
1805	Chr17:_72755647_72755648	*SLC9A3R1*
1806	Chr17:_72889549_72889550	*FADS6*
1807	Chr17:_73584068_73584069	*MYO15B*
1808	Chr17:_73584070_73584071	*MYO15B*
1809	Chr17:_73584360_73584361	*MYO15B*
1810	Chr17:_73584617_73584618	*MYO15B*
1811	Chr17:_73749654_73749655	*ITGB4*
1812	Chr17:_73749694_73749695	*ITGB4*
1813	Chr17:_74511719_74511720	*Non Coding Region*
1814	Chr17:_74533969_74533970	*CYGB*
1815	Chr17:_74533976_74533977	*CYGB*
1816	Chr17:_75369091_75369092	*SEPT_9*
1817	Chr17:_75369484_75369485	*SEPT_9*
1818	Chr17:_75369657_75369658	*SEPT_9*
1819	Chr17:_75401367_75401368	*SEPT_9*
1820	Chr17:_75441392_75441393	*SEPT_9*
1821	Chr17:_75471308_75471309	*SEPT_9*
1822	Chr17:_76126301_76126302	*TMC6,TMC8*
1823	Chr17:_76126702_76126703	*TMC6,TMC8*
1824	Chr17:_76127217_76127218	*TMC6,TMC8*
1825	Chr17:_76355202_76355203	*SOCS3*
1826	Chr17:_76361166_76361167	*Non Coding Region*
1827	Chr17:_76755745_76755746	*CYTH1*
1828	Chr17:_76775543_76775544	*CYTH1*
1829	Chr17:_76921845_76921846	*TIMP2*
1830	Chr17:_77179709_77179710	*HRNBP3*
1831	Chr17:_77721631_77721632	*Non Coding Region*
1832	Chr17:_78452048_78452049	*Non Coding Region*
1833	Chr17:_79259283_79259284	*SLC38A10*
1834	Chr17:_79259383_79259384	*SLC38A10*
1835	Chr17:_79259536_79259537	*SLC38A10*
1836	Chr17:_79259699_79259700	*SLC38A10*
1837	Chr17:_80260329_80260330	*Non Coding Region*
1838	Chr17:_80358850_80358851	*C17orf101*
1839	Chr17:_80358876_80358877	*C17orf101*
1840	Chr17:_80407031_80407032	*C17orf62*
1841	Chr17:_80407590_80407591	*C17orf62*
1842	Chr17:_80549621_80549622	*FOXK2*
1843	Chr18:_904851_904852	*ADCYAP1*
1844	Chr18:_904920_904921	*ADCYAP1*
1845	Chr18:_5197327_5197328	*LOC642597*

Order	Coordinate of CpG (hg19)	Gene
1846	Chr18:_5543271_5543272	*EPB41L3*
1847	Chr18:_5543548_5543549	*EPB41L3*
1848	Chr18:_5544169_5544170	*EPB41L3*
1849	Chr18:_5544231_5544232	*EPB41L3*
1850	Chr18:_7116977_7116978	*LAMA1*
1851	Chr18:_7118077_7118078	*LAMA1*
1852	Chr18:_11752089_11752090	*GNAL*
1853	Chr18:_18821913_18821914	*GREB1L*
1854	Chr18:_20811408_20811409	*CABLES1*
1855	Chr18:_22929205_22929206	*ZNF521*
1856	Chr18:_22930271_22930272	*ZNF521*
1857	Chr18:_22930283_22930284	*ZNF521*
1858	Chr18:_25755942_25755943	*CDH2*
1859	Chr18:_25756510_25756511	*CDH2*
1860	Chr18:_34833938_34833939	*BRUNOL4*
1861	Chr18:_35146766_35146767	*BRUNOL4*
1862	Chr18:_44337922_44337923	*ST8SIA5*
1863	Chr18:_53257019_53257020	*TCF4*
1864	Chr18:_53446528_53446529	*Non Coding Region*
1865	Chr18:_53446784_53446785	*Non Coding Region*
1866	Chr18:_54789058_54789059	*Non Coding Region*
1867	Chr18:_56887271_56887272	*GRP*
1868	Chr18:_56935199_56935200	*RAX*
1869	Chr18:_56941246_56941247	*RAX*
1870	Chr18:_65184398_65184399	*DSEL*
1871	Chr18:_67067867_67067868	*DOK6*
1872	Chr18:_67067893_67067894	*DOK6*
1873	Chr18:_73167671_73167672	*Non Coding Region*
1874	Chr18:_74961737_74961738	*GALR1*
1875	Chr18:_74961968_74961969	*GALR1*
1876	Chr18:_76738999_76739000	*SALL3*
1877	Chr19:_1401310_1401311	*GAMT*
1878	Chr19:_1401767_1401768	*GAMT*
1879	Chr19:_1401775_1401776	*GAMT*
1880	Chr19:_1401781_1401782	*GAMT*
1881	Chr19:_1467952_1467953	*APC2*
1882	Chr19:_2085297_2085298	*MOBKL2A*
1883	Chr19:_2085301_2085302	*MOBKL2A*
1884	Chr19:_2085483_2085484	*MOBKL2A*
1885	Chr19:_2253398_2253399	*JSRP1*
1886	Chr19:_3178844_3178845	*S1PR4*
1887	Chr19:_3178955_3178956	*S1PR4*
1888	Chr19:_3179364_3179365	*S1PR4*
1889	Chr19:_3179828_3179829	*S1PR4*
1890	Chr19:_3180035_3180036	*S1PR4*

Figure 44V

Order	Coordinate of CpG (hg19)	Gene
1891	Chr19:_4327106_4327107	*STAP2*
1892	Chr19:_4327350_4327351	*STAP2*
1893	Chr19:_4328759_4328760	*STAP2*
1894	Chr19:_4328818_4328819	*STAP2*
1895	Chr19:_4916593_4916594	*UHRF1*
1896	Chr19:_4916783_4916784	*UHRF1*
1897	Chr19:_4916983_4916984	*UHRF1*
1898	Chr19:_5339092_5339093	*PTPRS*
1899	Chr19:_6773074_6773075	*VAV1*
1900	Chr19:_11532175_11532176	*CCDC151*
1901	Chr19:_11689715_11689716	*ACP5*
1902	Chr19:_12203349_12203350	*ZNF788*
1903	Chr19:_12724912_12724913	*ZNF791*
1904	Chr19:_13123217_13123218	*NFIX*
1905	Chr19:_13124604_13124605	*NFIX*
1906	Chr19:_13125084_13125085	*NFIX*
1907	Chr19:_13207602_13207603	*NFIX*
1908	Chr19:_13215292_13215293	*Non Coding Region*
1909	Chr19:_13617091_13617092	*CACNA1A*
1910	Chr19:_13617584_13617585	*CACNA1A*
1911	Chr19:_14550846_14550847	*PKN1*
1912	Chr19:_14550855_14550856	*PKN1*
1913	Chr19:_14550997_14550998	*PKN1*
1914	Chr19:_15090872_15090873	*Non Coding Region*
1915	Chr19:_15218713_15218714	*SYDE1*
1916	Chr19:_15580721_15580722	*PGLYRP2*
1917	Chr19:_17008650_17008651	*CPAMD8*
1918	Chr19:_17392770_17392771	*ANKLE1*
1919	Chr19:_17392923_17392924	*ANKLE1*
1920	Chr19:_17957106_17957107	*JAK3*
1921	Chr19:_18113604_18113605	*ARRDC2*
1922	Chr19:_18118666_18118667	*ARRDC2*
1923	Chr19:_18540208_18540209	*SSBP4*
1924	Chr19:_18901666_18901667	*COMP*
1925	Chr19:_19322778_19322779	*NCAN*
1926	Chr19:_21106053_21106054	*ZNF85*
1927	Chr19:_22018958_22018959	*ZNF43*
1928	Chr19:_22034523_22034524	*Non Coding Region*
1929	Chr19:_30016147_30016148	*VSTM2B*
1930	Chr19:_30019529_30019530	*VSTM2B*
1931	Chr19:_31841555_31841556	*TSHZ3*
1932	Chr19:_31841663_31841664	*TSHZ3*
1933	Chr19:_31841952_31841953	*Non Coding Region*
1934	Chr19:_31842873_31842874	*Non Coding Region*
1935	Chr19:_31847970_31847971	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
1936	Chr19:_31848008_31848009	*Non Coding Region*
1937	Chr19:_31848130_31848131	*Non Coding Region*
1938	Chr19:_34113398_34113399	*CHST8*
1939	Chr19:_36233324_36233325	*TMEM149*
1940	Chr19:_36233380_36233381	*TMEM149*
1941	Chr19:_36233447_36233448	*U2AF1L4,TMEM149*
1942	Chr19:_36523584_36523585	*CLIP3*
1943	Chr19:_36909413_36909414	*ZFP82*
1944	Chr19:_37096148_37096149	*ZNF529,ZNF382*
1945	Chr19:_37096321_37096322	*ZNF529,ZNF382*
1946	Chr19:_37096323_37096324	*ZNF529,ZNF382*
1947	Chr19:_37096329_37096330	*ZNF529,ZNF382*
1948	Chr19:_37157565_37157566	*ZNF461*
1949	Chr19:_37407123_37407124	*ZNF829,ZNF568*
1950	Chr19:_37407284_37407285	*ZNF829,ZNF568*
1951	Chr19:_37464066_37464067	*Non Coding Region*
1952	Chr19:_37997471_37997472	*ZNF793*
1953	Chr19:_37997730_37997731	*ZNF793*
1954	Chr19:_37997817_37997818	*ZNF793*
1955	Chr19:_37997867_37997868	*ZNF793*
1956	Chr19:_37997991_37997992	*ZNF793*
1957	Chr19:_38747201_38747202	*PPP1R14A*
1958	Chr19:_39108609_39108610	*MAP4K1,EIF3K*
1959	Chr19:_39108672_39108673	*MAP4K1,EIF3K*
1960	Chr19:_41595975_41595976	*CYP2A13*
1961	Chr19:_42636817_42636818	*POU2F2*
1962	Chr19:_42703609_42703610	*DEDD2*
1963	Chr19:_42703763_42703764	*DEDD2*
1964	Chr19:_42703852_42703853	*DEDD2*
1965	Chr19:_42828156_42828157	*TMEM145*
1966	Chr19:_46996516_46996517	*PNMAL2*
1967	Chr19:_48021503_48021504	*Non Coding Region*
1968	Chr19:_48752990_48752991	*CARD8*
1969	Chr19:_48918116_48918117	*GRIN2D*
1970	Chr19:_48918205_48918206	*GRIN2D*
1971	Chr19:_48984071_48984072	*CYTH2*
1972	Chr19:_49838777_49838778	*CD37*
1973	Chr19:_50031017_50031018	*RCN3*
1974	Chr19:_50031246_50031247	*RCN3*
1975	Chr19:_50037661_50037662	*RCN3*
1976	Chr19:_51142933_51142934	*SYT3*
1977	Chr19:_51162508_51162509	*Non Coding Region*
1978	Chr19:_51830960_51830961	*IGLON5*
1979	Chr19:_51925128_51925129	*Non Coding Region*
1980	Chr19:_52873106_52873107	*ZNF880*

Figure 44W

Order	Coordinate of CpG (hg19)	Gene
1981	Chr19:_52873125_52873126	*ZNF880*
1982	Chr19:_53636108_53636109	*ZNF415*
1983	Chr19:_53758031_53758032	*ZNF677*
1984	Chr19:_54024110_54024111	*ZNF331*
1985	Chr19:_54412993_54412994	*Non Coding Region*
1986	Chr19:_54926431_54926432	*TTYH1*
1987	Chr19:_54926805_54926806	*TTYH1*
1988	Chr19:_55593580_55593581	*EPS8L1*
1989	Chr19:_55766853_55766854	*SAPS1*
1990	Chr19:_55836752_55836753	*TMEM150B*
1991	Chr19:_56879554_56879555	*ZNF542*
1992	Chr19:_56879645_56879646	*ZNF542*
1993	Chr19:_56904965_56904966	*ZNF582*
1994	Chr19:_56989543_56989544	*ZNF667*
1995	Chr19:_57049695_57049696	*ZFP28*
1996	Chr19:_58095595_58095596	*ZIK1*
1997	Chr19:_58095659_58095660	*ZIK1*
1998	Chr19:_58220370_58220371	*ZNF154*
1999	Chr19:_58220494_58220495	*ZNF154*
2000	Chr19:_58238850_58238851	*ZNF671*
2001	Chr19:_58238928_58238929	*ZNF671*
2002	Chr19:_58238987_58238988	*ZNF671*
2003	Chr19:_58239012_58239013	*ZNF671*
2004	Chr19:_58458769_58458770	*ZNF256*
2005	Chr19:_58458917_58458918	*ZNF256*
2006	Chr19:_58609361_58609362	*ZSCAN18*
2007	Chr19:_58951599_58951600	*ZNF132*
2008	Chr19:_58951602_58951603	*ZNF132*
2009	Chr20:_2539552_2539553	*TMC2*
2010	Chr20:_2781262_2781263	*CPXM1*
2011	Chr20:_3052692_3052693	*OXT*
2012	Chr20:_3063238_3063239	*AVP*
2013	Chr20:_3662948_3662949	*ADAM33*
2014	Chr20:_3713345_3713346	*HSPA12B*
2015	Chr20:_4795886_4795887	*RASSF2*
2016	Chr20:_4804052_4804053	*RASSF2*
2017	Chr20:_6033205_6033206	*LRRN4*
2018	Chr20:_8112754_8112755	*PLCB1*
2019	Chr20:_8112972_8112973	*PLCB1*
2020	Chr20:_8113573_8113574	*PLCB1*
2021	Chr20:_10199302_10199303	*SNAP25*
2022	Chr20:_10199536_10199537	*SNAP25*
2023	Chr20:_21492914_21492915	*NKX2-2*
2024	Chr20:_21682362_21682363	*Non Coding Region*
2025	Chr20:_23029806_23029807	*THBD*

Order	Coordinate of CpG (hg19)	Gene
2026	Chr20:_23030250_23030251	*THBD*
2027	Chr20:_23066944_23066945	*CD93*
2028	Chr20:_23066999_23067000	*CD93*
2029	Chr20:_25058429_25058430	*VSX1*
2030	Chr20:_25062447_25062448	*VSX1*
2031	Chr20:_30640256_30640257	*HCK*
2032	Chr20:_34356200_34356201	*Non Coding Region*
2033	Chr20:_34894648_34894649	*Non Coding Region*
2034	Chr20:_35258570_35258571	*SLA2*
2035	Chr20:_35274570_35274571	*SLA2*
2036	Chr20:_35274595_35274596	*SLA2*
2037	Chr20:_35274627_35274628	*SLA2*
2038	Chr20:_35274639_35274640	*SLA2*
2039	Chr20:_36012946_36012947	*SRC*
2040	Chr20:_37353126_37353127	*SLC32A1*
2041	Chr20:_37433745_37433746	*PPP1R16B*
2042	Chr20:_37433765_37433766	*PPP1R16B*
2043	Chr20:_37433803_37433804	*PPP1R16B*
2044	Chr20:_37434158_37434159	*PPP1R16B*
2045	Chr20:_37434167_37434168	*PPP1R16B*
2046	Chr20:_37434229_37434230	*PPP1R16B*
2047	Chr20:_37434950_37434951	*PPP1R16B*
2048	Chr20:_41817903_41817904	*PTPRT*
2049	Chr20:_41818358_41818359	*PTPRT*
2050	Chr20:_41818756_41818757	*PTPRT*
2051	Chr20:_41818770_41818771	*PTPRT*
2052	Chr20:_42875779_42875780	*GDAP1L1*
2053	Chr20:_42875931_42875932	*GDAP1L1*
2054	Chr20:_43439387_43439388	*RIMS4*
2055	Chr20:_43439400_43439401	*RIMS4*
2056	Chr20:_44937426_44937427	*Non Coding Region*
2057	Chr20:_44941542_44941543	*Non Coding Region*
2058	Chr20:_45142244_45142245	*ZNF334*
2059	Chr20:_45142258_45142259	*ZNF334*
2060	Chr20:_47935683_47935684	*Non Coding Region*
2061	Chr20:_50720908_50720909	*ZFP64*
2062	Chr20:_53092233_53092234	*DOK5*
2063	Chr20:_53092334_53092335	*DOK5*
2064	Chr20:_55500670_55500671	*Non Coding Region*
2065	Chr20:_55964998_55964999	*RBM38*
2066	Chr20:_55965077_55965078	*RBM38*
2067	Chr20:_55965245_55965246	*RBM38*
2068	Chr20:_57224919_57224920	*STX16*
2069	Chr20:_57225195_57225196	*STX16*
2070	Chr20:_58179847_58179848	*PHACTR3*

Figure 44X

Order	Coordinate of CpG (hg19)	Gene
2071	Chr20:_61050914_61050915	*GATA5*
2072	Chr20:_61051021_61051022	*GATA5*
2073	Chr20:_61051029_61051030	*GATA5*
2074	Chr20:_61051032_61051033	*GATA5*
2075	Chr20:_61051036_61051037	*GATA5*
2076	Chr20:_61051039_61051040	*GATA5*
2077	Chr20:_61051341_61051342	*GATA5*
2078	Chr20:_61051348_61051349	*GATA5*
2079	Chr20:_61051432_61051433	*GATA5*
2080	Chr20:_61051561_61051562	*GATA5*
2081	Chr20:_61638149_61638150	*BHLHE23*
2082	Chr20:_61638574_61638575	*BHLHE23*
2083	Chr20:_61638588_61638589	*BHLHE23*
2084	Chr20:_61808888_61808889	*MIR124-3*
2085	Chr20:_61810624_61810625	*Non Coding Region*
2086	Chr20:_61886079_61886080	*NKAIN4,FLJ16779*
2087	Chr20:_62103400_62103401	*KCNQ2*
2088	Chr20:_62198872_62198873	*PRIC285*
2089	Chr20:_62199156_62199157	*PRIC285*
2090	Chr20:_62199181_62199182	*PRIC285*
2091	Chr20:_62673890_62673891	*Non Coding Region*
2092	Chr20:_62733901_62733902	*Non Coding Region*
2093	Chr21:_27011788_27011789	*JAM2*
2094	Chr21:_27012615_27012616	*JAM2*
2095	Chr21:_27945125_27945126	*CYYR1*
2096	Chr21:_28339442_28339443	*ADAMTS5*
2097	Chr21:_28339907_28339908	*ADAMTS5*
2098	Chr21:_32716090_32716091	*TIAM1*
2099	Chr21:_34398199_34398200	*OLIG2*
2100	Chr21:_34398222_34398223	*OLIG2*
2101	Chr21:_34398226_34398227	*OLIG2*
2102	Chr21:_34442360_34442361	*OLIG1*
2103	Chr21:_36421467_36421468	*RUNX1*
2104	Chr21:_36421472_36421473	*RUNX1*
2105	Chr21:_36421503_36421504	*RUNX1*
2106	Chr21:_38078767_38078768	*SIM2*
2107	Chr21:_38083149_38083150	*SIM2*
2108	Chr21:_39870704_39870705	*ERG*
2109	Chr21:_45336673_45336674	*AGPAT3*
2110	Chr21:_45336903_45336904	*AGPAT3*
2111	Chr21:_46352084_46352085	*Non Coding Region*
2112	Chr21:_47518747_47518748	*COL6A2*
2113	Chr22:_18217141_18217142	*BID*
2114	Chr22:_19754125_19754126	*TBX1*
2115	Chr22:_20790985_20790986	*SCARF2*
2116	Chr22:_23480087_23480088	*RTDR1*
2117	Chr22:_24038523_24038524	*LOC91316,RGL4*
2118	Chr22:_24823514_24823515	*ADORA2A*
2119	Chr22:_25678988_25678989	*Non Coding Region*
2120	Chr22:_28196834_28196835	*MN1*
2121	Chr22:_30661949_30661950	*OSM*
2122	Chr22:_30662987_30662988	*OSM*
2123	Chr22:_30662994_30662995	*OSM*
2124	Chr22:_30663007_30663008	*OSM*
2125	Chr22:_30663316_30663317	*OSM*
2126	Chr22:_37882078_37882079	*MFNG*
2127	Chr22:_37962569_37962570	*CDC42EP1*
2128	Chr22:_39784481_39784482	*Non Coding Region*
2129	Chr22:_39853885_39853886	*MGAT3*
2130	Chr22:_44258572_44258573	*SULT4A1*
2131	Chr22:_45064471_45064472	*PRR5*
2132	Chr22:_45065052_45065053	*PRR5*
2133	Chr22:_45405061_45405062	*PHF21B*
2134	Chr22:_45405627_45405628	*PHF21B*
2135	Chr22:_46263207_46263208	*Non Coding Region*
2136	Chr22:_47187343_47187344	*TBC1D22A*
2137	Chr22:_50453437_50453438	*Non Coding Region*
2138	Chr22:_50623687_50623688	*TRABD*
2139	Chr22:_50623692_50623693	*TRABD*
2140	ChrX:_6145730_6145731	*NLGN4X*

Figure 44Y

CRC Methylation markers, 50% sensitivity, Chromosomal Regions

Order	Coordinates of Region (hg19)	Gene
1	Chr1:_966966-971238	*AGRN*
2	Chr1:_974496-981305	*AGRN*
3	Chr1:_1144568-1146878	*Non Coding Region*
4	Chr1:_1308140-1312039	*Non Coding Region*
5	Chr1:_1563421-1571006	*MIB2*
6	Chr1:_2221198-2223569	*SKI*
7	Chr1:_2231310-2233617	*SKI*
8	Chr1:_2982925-2988962	*PRDM16,FLJ42875*
9	Chr1:_3101540-3104352	*PRDM16*
10	Chr1:_3661963-3665085	*KIAA0495*
11	Chr1:_6085244-6087494	*KCNAB2*
12	Chr1:_6519584-6522955	*ESPN*
13	Chr1:_6524996-6527228	*TNFRSF25*
14	Chr1:_7691231-7693503	*CAMTA1*
15	Chr1:_8456795-8458721	*RERE*
16	Chr1:_9487937-9490249	*Non Coding Region*
17	Chr1:_9686125-9688367	*Non Coding Region*
18	Chr1:_12122488-12125148	*TNFRSF8*
19	Chr1:_13909137-13911868	*PDPN*
20	Chr1:_16022570-16024842	*PLEKHM2*
21	Chr1:_18955895-18960829	*PAX7*
22	Chr1:_19253055-19255007	*IFFO2*
23	Chr1:_19802883-19804981	*CAPZB*
24	Chr1:_25348052-25349960	*Non Coding Region*
25	Chr1:_26376903-26378895	*TRIM63*
26	Chr1:_27069755-27071991	*ARID1A*
27	Chr1:_27686024-27688743	*MAP3K6*
28	Chr1:_28212887-28214713	*Non Coding Region*
29	Chr1:_29584897-29587598	*PTPRU*
30	Chr1:_31229615-31231587	*LAPTM5*
31	Chr1:_32712918-32715186	*LCK*
32	Chr1:_34627783-34631976	*CSMD2*
33	Chr1:_35349878-35352854	*DLGAP3*
34	Chr1:_35393747-35397206	*Non Coding Region*
35	Chr1:_36041432-36044444	*TFAP2E*
36	Chr1:_37497377-37501624	*GRIK3*
37	Chr1:_38098677-38101864	*RSPO1*
38	Chr1:_38228838-38231888	*EPHA10*
39	Chr1:_39024260-39026238	*Non Coding Region*
40	Chr1:_39268729-39271251	*Non Coding Region*
41	Chr1:_42216025-42218129	*HIVEP3*
42	Chr1:_44030286-44032853	*PTPRF*
43	Chr1:_44870109-44875047	*RNF220*
44	Chr1:_46950168-46952792	*Non Coding Region*
45	Chr1:_47695504-47699197	*Non Coding Region*

Order	Coordinates of Region (hg19)	Gene
46	Chr1:_47908712-47912020	*Non Coding Region*
47	Chr1:_48057793-48060230	*Non Coding Region*
48	Chr1:_48189756-48191992	*Non Coding Region*
49	Chr1:_48448870-48451144	*Non Coding Region*
50	Chr1:_49241371-49243810	*AGBL4,BEND5*
51	Chr1:_50880884-50883103	*Non Coding Region*
52	Chr1:_50883228-50892471	*DMRTA2*
53	Chr1:_53097629-53100303	*FAM159A*
54	Chr1:_53526572-53529974	*PODN*
55	Chr1:_57886963-57891637	*DAB1*
56	Chr1:_58714272-58717095	*DAB1*
57	Chr1:_65730411-65732849	*DNAJC6*
58	Chr1:_67217079-67219293	*TCTEX1D1*
59	Chr1:_67772329-67774767	*IL12RB2*
60	Chr1:_70031967-70035495	*Non Coding Region*
61	Chr1:_72747471-72750736	*NEGR1*
62	Chr1:_76079454-76081808	*Non Coding Region*
63	Chr1:_76080378-76083365	*Non Coding Region*
64	Chr1:_77332111-77335534	*ST6GALNAC5*
65	Chr1:_77746314-77749224	*AK5*
66	Chr1:_86620278-86623871	*COL24A1*
67	Chr1:_87596543-87599515	*LOC339524*
68	Chr1:_88920477-88922469	*Non Coding Region*
69	Chr1:_91189489-91193804	*Non Coding Region*
70	Chr1:_91299979-91302891	*Non Coding Region*
71	Chr1:_91868876-91871390	*HFM1*
72	Chr1:_98509966-98512710	*Non Coding Region*
73	Chr1:_101003471-101006885	*GPR88*
74	Chr1:_101701445-101703745	*S1PR1*
75	Chr1:_107681889-107685463	*NTNG1*
76	Chr1:_108506251-108508841	*VAV3*
77	Chr1:_111097066-111099362	*Non Coding Region*
78	Chr1:_111215244-111218937	*KCNA3*
79	Chr1:_114413363-114415317	*PTPN22*
80	Chr1:_115879167-115882332	*NGF*
81	Chr1:_117057046-117058890	*CD58*
82	Chr1:_119528819-119533196	*TBX15*
83	Chr1:_147788652-147791094	*Non Coding Region*
84	Chr1:_150120695-150124078	*PLEKHO1*
85	Chr1:_150253365-150255637	*C1orf51*
86	Chr1:_151128241-151130355	*TNFAIP8L2*
87	Chr1:_151809460-151812684	*LOC100132111,C2CD4D*
88	Chr1:_153517261-153519507	*S100A4*
89	Chr1:_154297205-154299544	*ATP8B2*
90	Chr1:_154473107-154476699	*TDRD10,SHE*

Figure 45A

Order	Coordinates of Region (hg19)	Gene
91	Chr1:_156065916-156068056	*Non Coding Region*
92	Chr1:_156357050-156359252	*Non Coding Region*
93	Chr1:_156404160-156407350	*Non Coding Region*
94	Chr1:_156626342-156628576	*BCAN*
95	Chr1:_156827804-156832102	*NTRK1,INSRR*
96	Chr1:_158149620-158152503	*CD1D*
97	Chr1:_160806686-160808702	*CD244*
98	Chr1:_161038658-161040704	*ARHGAP30*
99	Chr1:_161227477-161230028	*PCP4L1*
100	Chr1:_165413452-165415636	*RXRG*
101	Chr1:_167598464-167600839	*RCSD1*
102	Chr1:_167682477-167684917	*Non Coding Region*
103	Chr1:_169395621-169397869	*C1orf114*
104	Chr1:_170629456-170631851	*Non Coding Region*
105	Chr1:_171809467-171812325	*DNM3*
106	Chr1:_181450311-181453049	*CACNA1E*
107	Chr1:_190446200-190448380	*FAM5C*
108	Chr1:_200007392-200010047	*NR5A2*
109	Chr1:_202128104-202130304	*PTPN7*
110	Chr1:_203597471-203644187	*ATP2B4*
111	Chr1:_204327717-204329969	*PLEKHA6*
112	Chr1:_204653132-204655885	*LRRN2*
113	Chr1:_206729397-206731908	*RASSF5*
114	Chr1:_206747772-206749928	*RASSF5*
115	Chr1:_207817389-207819952	*CR1L*
116	Chr1:_208083098-208085513	*CD34*
117	Chr1:_209928577-209930515	*TRAF3IP3*
118	Chr1:_214152214-214154668	*Non Coding Region*
119	Chr1:_218518057-218520674	*TGFB2*
120	Chr1:_220700507-220703348	*MARK1*
121	Chr1:_221049448-221051864	*Non Coding Region*
122	Chr1:_221050966-221054673	*HLX*
123	Chr1:_221885725-221887553	*DUSP10*
124	Chr1:_226923560-226927553	*ITPKB*
125	Chr1:_228602706-228605567	*TRIM17*
126	Chr1:_228650447-228653529	*Non Coding Region*
127	Chr1:_229541943-229544770	*Non Coding Region*
128	Chr1:_229566206-229571684	*ACTA1*
129	Chr1:_232861119-232862931	*Non Coding Region*
130	Chr1:_234348884-234351471	*SLC35F3*
131	Chr1:_237204126-237207644	*RYR2*
132	Chr1:_240253959-240258063	*FMN2*
133	Chr1:_241586601-241588883	*Non Coding Region*
134	Chr1:_244011712-244014245	*Non Coding Region*
135	Chr1:_247711325-247713149	*C1orf150*

Order	Coordinates of Region (hg19)	Gene
136	Chr2:_466849-469659	*Non Coding Region*
137	Chr2:_5835068-5838643	*SOX11*
138	Chr2:_7056530-7059433	*RNF144A*
139	Chr2:_10218802-10222016	*CYS1*
140	Chr2:_20864289-20868589	*GDF7*
141	Chr2:_25437725-25440276	*Non Coding Region*
142	Chr2:_25498763-25501429	*DNMT3A*
143	Chr2:_29336983-29339909	*CLIP4*
144	Chr2:_30141831-30145624	*ALK*
145	Chr2:_31359321-31362821	*GALNT14*
146	Chr2:_31455740-31458653	*EHD3*
147	Chr2:_31804293-31807403	*SRD5A2*
148	Chr2:_37874753-37876919	*CDC42EP3*
149	Chr2:_38300276-38305518	*CYP1B1*
150	Chr2:_39892121-39894496	*TMEM178*
151	Chr2:_40677477-40679717	*SLC8A1*
152	Chr2:_45154195-45158049	*Non Coding Region*
153	Chr2:_45168505-45172884	*SIX3*
154	Chr2:_45234511-45238792	*SIX2*
155	Chr2:_45239372-45242579	*Non Coding Region*
156	Chr2:_47795923-47800166	*KCNK12*
157	Chr2:_56409865-56413539	*CCDC85A*
158	Chr2:_58654977-58657151	*Non Coding Region*
159	Chr2:_63273475-63280430	*LOC100132215*
160	Chr2:_63281514-63284122	*OTX1*
161	Chr2:_63282936-63285147	*OTX1*
162	Chr2:_66651691-66655218	*Non Coding Region*
163	Chr2:_66660118-66662657	*Non Coding Region*
164	Chr2:_66671431-66674636	*MEIS1*
165	Chr2:_66801890-66805831	*Non Coding Region*
166	Chr2:_66807568-66810404	*Non Coding Region*
167	Chr2:_68545339-68548111	*CNRIP1*
168	Chr2:_68961019-69002868	*ARHGAP25*
169	Chr2:_70312200-70316490	*PCBP1*
170	Chr2:_71294160-71297002	*NAGK*
171	Chr2:_73150200-73153060	*EMX1*
172	Chr2:_74739455-74744795	*TLX2*
173	Chr2:_85358956-85363593	*TCF7L1*
174	Chr2:_87754380-87756362	*NCRNA00152*
175	Chr2:_95689857-95693431	*MAL*
176	Chr2:_97191977-97194383	*Non Coding Region*
177	Chr2:_99551185-99554831	*C2orf55*
178	Chr2:_100936779-100940059	*LONRF2*
179	Chr2:_101032606-101035296	*CHST10*
180	Chr2:_102089370-102092130	*RFX8*

Figure 45B

Order	Coordinates of Region (hg19)	Gene
181	Chr2:_105467851-105474488	*POU3F3*
182	Chr2:_105477600-105480188	*Non Coding Region*
183	Chr2:_105759109-105762018	*Non Coding Region*
184	Chr2:_106414105-106416185	*NCK2*
185	Chr2:_106775741-106777809	*UXS1*
186	Chr2:_109646955-109649230	*Non Coding Region*
187	Chr2:_113873016-113875120	*IL1RN*
188	Chr2:_113930379-113932430	*LOC440839,PSD4*
189	Chr2:_115917737-115921765	*DPP10*
190	Chr2:_119598458-119601966	*EN1*
191	Chr2:_119913126-119917663	*C1QL2*
192	Chr2:_121100800-121105534	*INHBB*
193	Chr2:_127412696-127415171	*GYPC*
194	Chr2:_127642800-127645104	*Non Coding Region*
195	Chr2:_127782081-127784311	*Non Coding Region*
196	Chr2:_127838615-127840463	*BIN1*
197	Chr2:_128432010-128434490	*LIMS2*
198	Chr2:_131791237-131794189	*ARHGEF4*
199	Chr2:_139536692-139539650	*NXPH2*
200	Chr2:_143885389-143887263	*ARHGAP15*
201	Chr2:_144693666-144696180	*Non Coding Region*
202	Chr2:_145280736-145283269	*Non Coding Region*
203	Chr2:_149631682-149634882	*KIF5C*
204	Chr2:_158299349-158301601	*CYTIP*
205	Chr2:_161262871-161265460	*RBMS1*
206	Chr2:_164591917-164594511	*FIGN*
207	Chr2:_171568877-171574904	*SP5*
208	Chr2:_171671310-171676447	*GAD1*
209	Chr2:_172943988-172946384	*MAP1D*
210	Chr2:_172971804-172974242	*Non Coding Region*
211	Chr2:_175189866-175193123	*Non Coding Region*
212	Chr2:_175207415-175210076	*Non Coding Region*
213	Chr2:_175545862-175548880	*WIPF1*
214	Chr2:_176955504-176957707	*HOXD13*
215	Chr2:_177028413-177030941	*HOXD3*
216	Chr2:_182320761-182324029	*ITGA4*
217	Chr2:_182546873-182550177	*Non Coding Region*
218	Chr2:_185462208-185464589	*ZNF804A*
219	Chr2:_187712846-187715069	*ZSWIM2*
220	Chr2:_193057995-193061812	*TMEFF2*
221	Chr2:_198062660-198064646	*ANKRD44*
222	Chr2:_198119444-198121606	*Non Coding Region*
223	Chr2:_202124077-202126347	*CASP8*
224	Chr2:_206548599-206552316	*NRP2*
225	Chr2:_207505774-207508422	*LOC200726*

Order	Coordinates of Region (hg19)	Gene
226	Chr2:_208987834-208990269	*CRYGD*
227	Chr2:_219845291-219848137	*FEV*
228	Chr2:_220108984-220111488	*STK16*
229	Chr2:_220172870-220175283	*PTPRN*
230	Chr2:_220298483-220301243	*SPEG*
231	Chr2:_220305855-220308096	*SPEG*
232	Chr2:_220360462-220364254	*Non Coding Region*
233	Chr2:_220415863-220418869	*OBSL1*
234	Chr2:_222435034-222439941	*EPHA4*
235	Chr2:_223161946-223164912	*CCDC140,PAX3*
236	Chr2:_223166205-223168560	*CCDC140*
237	Chr2:_227287523-227289513	*Non Coding Region*
238	Chr2:_230577062-230580981	*DNER*
239	Chr2:_233179385-233181445	*DIS3L2*
240	Chr2:_233496826-233500727	*EFHD1*
241	Chr2:_238534561-238537840	*LRRFIP1*
242	Chr2:_242741598-242744758	*GAL3ST2*
243	Chr3:_237391-241140	*CHL1*
244	Chr3:_9820903-9822729	*TADA3*
245	Chr3:_16553327-16556413	*RFTN1*
246	Chr3:_27770638-27772942	*Non Coding Region*
247	Chr3:_32857194-32861506	*TRIM71*
248	Chr3:_37033228-37036356	*EPM2AIP1,MLH1*
249	Chr3:_42946480-42949170	*ZNF662*
250	Chr3:_43430249-43432437	*ANO10*
251	Chr3:_44035407-44042104	*Non Coding Region*
252	Chr3:_44062314-44064837	*Non Coding Region*
253	Chr3:_44625325-44627794	*ZNF660*
254	Chr3:_44725928-44728237	*Non Coding Region*
255	Chr3:_49839103-49842191	*UBA7*
256	Chr3:_50311687-50315419	*SEMA3B*
257	Chr3:_51988763-51991639	*GPR62*
258	Chr3:_55516658-55518939	*WNT5A*
259	Chr3:_56787961-56790259	*ARHGEF3*
260	Chr3:_62355773-62358315	*FEZF2*
261	Chr3:_68979783-68983035	*FAM19A4*
262	Chr3:_87038610-87041516	*VGLL3*
263	Chr3:_96531023-96534625	*EPHA6*
264	Chr3:_111851137-111853175	*GCET2*
265	Chr3:_112929437-112932506	*BOC*
266	Chr3:_118958339-118960934	*B4GALT4*
267	Chr3:_119527920-119530159	*NR1I2*
268	Chr3:_120002883-120005426	*Non Coding Region*
269	Chr3:_121378698-121380792	*HCLS1*
270	Chr3:_121901742-121904645	*CASR*

Figure 45C

Order	Coordinates of Region (hg19)	Gene
271	Chr3:_124859569-124862019	*SLC12A8*
272	Chr3:_128204495-128213274	*GATA2*
273	Chr3:_128718865-128722245	*CCDC48*
274	Chr3:_129692127-129695841	*TRH*
275	Chr3:_132755985-132758895	*TMEM108*
276	Chr3:_134368464-134371242	*KY*
277	Chr3:_138655627-138660107	*Non Coding Region*
278	Chr3:_138662718-138667346	*FOXL2,C3orf72*
279	Chr3:_138678244-138680579	*Non Coding Region*
280	Chr3:_139256712-139258949	*RBP1*
281	Chr3:_139652430-139656115	*CLSTN2*
282	Chr3:_140768886-140772080	*SPSB4*
283	Chr3:_142680137-142684268	*PAQR9*
284	Chr3:_142836885-142841838	*CHST2*
285	Chr3:_150101829-150103729	*Non Coding Region*
286	Chr3:_150801996-150806168	*MED12L*
287	Chr3:_159755632-159757997	*Non Coding Region*
288	Chr3:_169385268-169388187	*Non Coding Region*
289	Chr3:_172164372-172167738	*GHSR*
290	Chr3:_179167735-179170593	*GNB4*
291	Chr3:_179753520-179756245	*PEX5L*
292	Chr3:_183272151-183274339	*KLHL6*
293	Chr3:_184300117-184303134	*Non Coding Region*
294	Chr3:_185079679-185081953	*MAP3K13*
295	Chr3:_186077710-186081111	*DGKG*
296	Chr3:_192124821-192128994	*FGF12*
297	Chr3:_194116601-194119988	*GP5*
298	Chr3:_194405440-194410045	*FAM43A*
299	Chr3:_195487595-195490381	*MUC4*
300	Chr3:_195621187-195624033	*TNK2*
301	Chr4:_567513-571488	*Non Coding Region*
302	Chr4:_1194231-1196405	*LOC100130872, LOC100130872-SPON2*
303	Chr4:_2787882-2790090	*Non Coding Region*
304	Chr4:_2932644-2934879	*MFSD10*
305	Chr4:_4386593-4390919	*D4S234E*
306	Chr4:_5052230-5055092	*STK32B*
307	Chr4:_5708985-5711495	*EVC2*
308	Chr4:_5712035-5714451	*EVC*
309	Chr4:_5890981-5893365	*CRMP1*
310	Chr4:_6199912-6203724	*JAKMIP1*
311	Chr4:_7193537-7196467	*SORCS2*
312	Chr4:_8036339-8038311	*ABLIM2*
313	Chr4:_10457129-10460353	*ZNF518B*
314	Chr4:_10685328-10687498	*CLNK*
315	Chr4:_13523062-13527083	*Non Coding Region*

Order	Coordinates of Region (hg19)	Gene
316	Chr4:_13529847-13532244	*Non Coding Region*
317	Chr4:_13542562-13547494	*Non Coding Region*
318	Chr4:_15778998-15781729	*CD38*
319	Chr4:_24800109-24802902	*SOD3*
320	Chr4:_37244250-37247979	*KIAA1239*
321	Chr4:_40197316-40199468	*RHOH*
322	Chr4:_40857884-40860162	*APBB2*
323	Chr4:_41257759-41260867	*UCHL1*
324	Chr4:_41882492-41885570	*Non Coding Region*
325	Chr4:_42398152-42401802	*SHISA3*
326	Chr4:_54965163-54969063	*GSX2*
327	Chr4:_55095185-55101331	*PDGFRA*
328	Chr4:_55990402-55993171	*KDR*
329	Chr4:_57520621-57523703	*HOPX*
330	Chr4:_71858158-71860884	*DCK*
331	Chr4:_73433855-73436321	*ADAMTS3*
332	Chr4:_81103761-81106527	*PRDM8*
333	Chr4:_81186494-81188913	*FGF5*
334	Chr4:_84029630-84032309	*PLAC8*
335	Chr4:_89377223-89379948	*HERC5*
336	Chr4:_90757008-90759870	*SNCA*
337	Chr4:_90814904-90817042	*MMRN1*
338	Chr4:_96468456-96470667	*UNC5C*
339	Chr4:_101110146-101112497	*DDIT4L*
340	Chr4:_111553965-111556504	*PITX2*
341	Chr4:_122300567-122303290	*QRFPR*
342	Chr4:_122684860-122687565	*TMEM155, LOC100192379*
343	Chr4:_123746740-123749980	*FGF2*
344	Chr4:_126234895-126239930	*FAT4*
345	Chr4:_134068162-134071442	*Non Coding Region*
346	Chr4:_142052327-142055601	*RNF150*
347	Chr4:_144619822-144623218	*FREM3*
348	Chr4:_147569029-147570847	*Non Coding Region*
349	Chr4:_154679774-154682377	*RNF175*
350	Chr4:_154708512-154711827	*SFRP2*
351	Chr4:_154711073-154713706	*SFRP2*
352	Chr4:_155253130-155255603	*DCHS2*
353	Chr4:_156296601-156299094	*MAP9*
354	Chr4:_156679095-156682386	*GUCY1B3*
355	Chr4:_166793545-166796341	*TLL1*
356	Chr4:_172732734-172736118	*GALNTL6*
357	Chr4:_174449046-174452469	*NBLA00301,HAND2*
358	Chr4:_175749330-175751584	*GLRA3*
359	Chr4:_176921473-176923820	*GPM6A*
360	Chr4:_176985921-176988360	*WDR17*

Figure 45D

Order	Coordinates of Region (hg19)	Gene
361	Chr4:_183061278-183063481	*MGC45800*
362	Chr4:_183368276-183371459	*ODZ3*
363	Chr4:_184825253-184829177	*STOX2*
364	Chr4:_185455221-185457029	*Non Coding Region*
365	Chr4:_185938222-185943747	*HELT*
366	Chr4:_186559031-186561361	*SORBS2*
367	Chr5:_527383-529982	*Non Coding Region*
368	Chr5:_1443678-1447648	*SLC6A3*
369	Chr5:_1666187-1669081	*Non Coding Region*
370	Chr5:_1873907-1880032	*Non Coding Region*
371	Chr5:_1880924-1888743	*IRX4*
372	Chr5:_1929754-1932986	*Non Coding Region*

Order	Coordinates of Region (hg19)	Gene
406	Chr5:_94954630-94958244	*GPR150*
407	Chr5:_100235567-100237955	*ST8SIA4*
408	Chr5:_101631049-101633373	*SLCO4C1*
409	Chr5:_115150348-115153713	*CDO1*
410	Chr5:_127871563-127875945	*FBN2*
411	Chr5:_131791870-131793780	*C5orf56*
412	Chr5:_131990554-131993304	*Non Coding Region*
413	Chr5:_134362092-134366146	*Non Coding Region*
414	Chr5:_136833016-136836146	*SPOCK1*
415	Chr5:_140786447-140789044	*PCDHGB1,PCDHGB3,PCDHGB2,PCDHGA8,PCDHGA9,PCDHGB6,PCDHGB5,PCDHGB4,PCDHGA2,PCDHGA3,PCDHGA1,PCDHGA6,PCDHGA7,PCDHGA4,PCDHGA5*
416	Chr5:_140796162-140798701	*PCDHGA10,PCDHGA8,PCDHGB3,PCDHGB2,PCDHGB1,PCDHGA9,PCDHGB7,PCDHGB6,PCDHGB5,PCDHGB4,PCDHGA2,PCDHGA3,PCDHGA1,PCDHGA6,PCDHGA7,PCDHGA4,PCDHGA5*
417	Chr5:_140854386-140857620	*PCDHGC3,PCDHGA8,PCDHGA10,PCDHGA11,PCDHGA12,PCDHGB3,PCDHGB2,PCDHGB1,PCDHGA9,PCDHGB7,PCDHGB6,PCDHGB5,PCDHGB4,PCDHGA2,PCDHGA3,PCDHGA1,PCDHGA6,PCDHGA7,PCDHGA4,PCDHGA5*

Figure 45E

Order	Coordinates of Region (hg19)	Gene
373	Chr5:_2037527-2039949	*Non Coding Region*
374	Chr5:_3593467-3604054	*IRX1*
375	Chr5:_5138441-5141941	*ADAMTS16*
376	Chr5:_7848945-7851439	*C5orf49*
377	Chr5:_11902550-11905703	*CTNND2*
378	Chr5:_17216788-17220021	*BASP1,LOC285696*
379	Chr5:_31854003-31856426	*PDZD2*
380	Chr5:_32708910-32715450	*Non Coding Region*
381	Chr5:_33935168-33939309	*RXFP3*
382	Chr5:_36689207-36691658	*Non Coding Region*
383	Chr5:_37833671-37836128	*GDNF*
384	Chr5:_37835747-37841726	*GDNF*
385	Chr5:_38256825-38260136	*EGFLAM*
386	Chr5:_38555222-38558563	*LIFR*
387	Chr5:_42423338-42426047	*GHR*
388	Chr5:_42950076-42953410	*Non Coding Region*
389	Chr5:_42993626-42995936	*Non Coding Region*
390	Chr5:_44388468-44390344	*FGF10*
391	Chr5:_49735607-49738300	*EMB*
392	Chr5:_50264085-50266737	*Non Coding Region*
393	Chr5:_54396616-54398568	*GZMA*
394	Chr5:_54515267-54517919	*Non Coding Region*
395	Chr5:_63800781-63803605	*RGS7BP*
396	Chr5:_63985189-63987974	*FAM159B*
397	Chr5:_71013917-71016715	*CARTPT*
398	Chr5:_71401877-71405868	*MAP1B*
399	Chr5:_72528099-72530976	*Non Coding Region*
400	Chr5:_72593147-72596808	*Non Coding Region*
401	Chr5:_76010120-76013292	*F2R*
402	Chr5:_82767387-82770268	*VCAN*
403	Chr5:_88178866-88181258	*MEF2C*
404	Chr5:_92905239-92909875	*FLJ42709*
405	Chr5:_92929529-92931830	*Non Coding Region*

Order	Coordinates of Region (hg19)	Gene
418	Chr5:_141261988-141264195	*Non Coding Region*
419	Chr5:_141391360-141393587	*Non Coding Region*
420	Chr5:_145717289-145721095	*POU4F3*
421	Chr5:_146256346-146259575	*PPP2R2B*
422	Chr5:_150592778-150594590	*CCDC69*
423	Chr5:_156679149-156681333	*ITK*
424	Chr5:_156691812-156694749	*CYFIP2*
425	Chr5:_156697931-156699777	*CYFIP2*
426	Chr5:_159932904-159934808	*Non Coding Region*
427	Chr5:_169063270-169065702	*DOCK2*
428	Chr5:_169532080-169534534	*FOXI1*
429	Chr5:_169723640-169725540	*LCP2*
430	Chr5:_170734169-170740863	*TLX3*
431	Chr5:_170740603-170743751	*Non Coding Region*
432	Chr5:_171604653-171606903	*STK10*
433	Chr5:_172658049-172661277	*NKX2-5*
434	Chr5:_174157680-174160729	*Non Coding Region*
435	Chr5:_175222609-175225679	*CPLX2*
436	Chr5:_175791355-175794505	*ARL10*
437	Chr5:_176798454-176800278	*RGS14*
438	Chr5:_177411534-177413753	*Non Coding Region*
439	Chr5:_178769724-178773794	*ADAMTS2*
440	Chr5:_180016099-180020062	*SCGB3A1*
441	Chr5:_180228375-180231147	*MGAT1*
442	Chr6:_390188-394790	*IRF4*
443	Chr6:_473935-476213	*Non Coding Region*
444	Chr6:_1378693-1381014	*Non Coding Region*
445	Chr6:_1388139-1392393	*FOXF2*
446	Chr6:_1623185-1626468	*GMDS*
447	Chr6:_6001471-6006125	*NRN1*
448	Chr6:_10382525-10385114	*Non Coding Region*
449	Chr6:_10389038-10391565	*Non Coding Region*
450	Chr6:_10421321-10423648	*Non Coding Region*

Figure 45F

Order	Coordinates of Region (hg19)	Gene
451	Chr6:_10881926-10884149	*GCM2*
452	Chr6:_11381416-11383690	*NEDD9*
453	Chr6:_13427509-13429535	*GFOD1*
454	Chr6:_14734212-14736138	*Non Coding Region*
455	Chr6:_25041098-25043082	*Non Coding Region*
456	Chr6:_25041458-25043532	*Non Coding Region*
457	Chr6:_25041498-25043496	*Non Coding Region*
458	Chr6:_26239697-26241951	*HIST1H4F*
459	Chr6:_27172690-27174816	*Non Coding Region*
460	Chr6:_27172849-27175133	*Non Coding Region*
461	Chr6:_27462176-27464379	*Non Coding Region*
462	Chr6:_28492600-28494722	*GPX5*
463	Chr6:_28509432-28511246	*Non Coding Region*
464	Chr6:_28601853-28604295	*Non Coding Region*
465	Chr6:_28955106-28957578	*Non Coding Region*
466	Chr6:_29520110-29522833	*Non Coding Region*
467	Chr6:_29594298-29596795	*GABBR1*
468	Chr6:_29758946-29761903	*HCG4*
469	Chr6:_29794553-29797594	*HLA-G*
470	Chr6:_29943402-29946169	*HCG9*
471	Chr6:_30094173-30096610	*Non Coding Region*
472	Chr6:_30138718-30141263	*TRIM15*
473	Chr6:_30226320-30229255	*HLA-L*
474	Chr6:_30417844-30420630	*Non Coding Region*
475	Chr6:_30648925-30655934	*KIAA1949*
476	Chr6:_31526790-31529050	*Non Coding Region*
477	Chr6:_31526879-31528907	*Non Coding Region*
478	Chr6:_31526931-31528847	*Non Coding Region*
479	Chr6:_31527167-31529141	*Non Coding Region*
480	Chr6:_31553288-31555110	*LST1*
481	Chr6:_31781872-31786190	*HSPA1A,HSPA1L*
482	Chr6:_31829299-31831948	*SLC44A4,NEU1*
483	Chr6:_32810494-32812839	*TAP1,PSMB8*
484	Chr6:_34110735-34114944	*Non Coding Region*
485	Chr6:_39280135-39283215	*KCNK17*
486	Chr6:_41339667-41343543	*Non Coding Region*
487	Chr6:_41527266-41529900	*FOXP4*
488	Chr6:_41603583-41607663	*MDFI*
489	Chr6:_42071032-42073701	*C6orf132*
490	Chr6:_42737915-42740127	*Non Coding Region*
491	Chr6:_42737936-42739998	*Non Coding Region*
492	Chr6:_42738126-42739972	*Non Coding Region*
493	Chr6:_43251734-43254476	*TTBK1*
494	Chr6:_45386346-45389164	*RUNX2*
495	Chr6:_55442732-55445092	*HMGCLL1*

Order	Coordinates of Region (hg19)	Gene
496	Chr6:_56817873-56821308	*BEND6,DST*
497	Chr6:_71664360-71668132	*B3GAT2*
498	Chr6:_73329942-73334109	*KCNQ5*
499	Chr6:_84561820-84564758	*RIPPLY2*
500	Chr6:_85471702-85475132	*TBX18*
501	Chr6:_91319890-91322219	*Non Coding Region*
502	Chr6:_99278317-99284842	*POU3F2*
503	Chr6:_100911071-100914337	*SIM1*
504	Chr6:_105387190-105390545	*Non Coding Region*
505	Chr6:_105583148-105586621	*BVES*
506	Chr6:_106428111-106430772	*Non Coding Region*
507	Chr6:_108484671-108491539	*NR2E1*
508	Chr6:_110677854-110680108	*C6orf186*
509	Chr6:_117084739-117087942	*FAM162B*
510	Chr6:_118227042-118230811	*SLC35F1*
511	Chr6:_119130644-119132822	*Non Coding Region*
512	Chr6:_124123177-124126309	*NKAIN2*
513	Chr6:_127438265-127442072	*RSPO3*
514	Chr6:_133561086-133564586	*EYA4*
515	Chr6:_134637797-134640021	*SGK1*
516	Chr6:_137813355-137816202	*OLIG3*
517	Chr6:_138144094-138146228	*Non Coding Region*
518	Chr6:_149802279-149804133	*ZC3H12D*
519	Chr6:_149802353-149804535	*ZC3H12D*
520	Chr6:_149802447-149804405	*ZC3H12D*
521	Chr6:_151560283-151563550	*AKAP12*
522	Chr6:_152127822-152130771	*ESR1*
523	Chr6:_152956810-152959472	*SYNE1*
524	Chr6:_153449992-153453792	*RGS17*
525	Chr6:_159588636-159592319	*FNDC1*
526	Chr6:_166578973-166584423	*T*
527	Chr6:_166665837-166668541	*Non Coding Region*
528	Chr6:_166719239-166723275	*PRR18*
529	Chr6:_167274720-167277626	*RPS6KA2*
530	Chr6:_168764814-168767041	*Non Coding Region*
531	Chr6:_168770137-168772640	*Non Coding Region*
532	Chr6:_168935452-168937664	*SMOC2*
533	Chr6:_170462269-170464525	*Non Coding Region*
534	Chr7:_643181-645461	*PRKAR1B*
535	Chr7:_1268165-1283864	*Non Coding Region*
536	Chr7:_1702861-1711517	*Non Coding Region*
537	Chr7:_1958868-1961110	*MAD1L1*
538	Chr7:_2441792-2445011	*CHST12*
539	Chr7:_3133415-3135699	*Non Coding Region*
540	Chr7:_4847251-4850204	*RADIL*

Figure 45G

Order	Coordinates of Region (hg19)	Gene
541	Chr7:_4921707-4924576	*RADIL*
542	Chr7:_6574923-6577389	*GRID2IP*
543	Chr7:_8480974-8483762	*NXPH1*
544	Chr7:_19144872-19147256	*Non Coding Region*
545	Chr7:_19145923-19148308	*Non Coding Region*
546	Chr7:_19155050-19159042	*TWIST1*
547	Chr7:_19811569-19814005	*TMEM196*
548	Chr7:_21581443-21584313	*DNAH11*
549	Chr7:_24322558-24326080	*NPY*
550	Chr7:_27207871-27210616	*Non Coding Region*
551	Chr7:_30720372-30723445	*CRHR2*
552	Chr7:_32336983-32339725	*Non Coding Region*
553	Chr7:_32800484-32802972	*Non Coding Region*
554	Chr7:_35291985-35295674	*TBX20*
555	Chr7:_35295921-35299218	*Non Coding Region*
556	Chr7:_36763117-36764921	*AOAH*
557	Chr7:_37486354-37489672	*ELMO1*
558	Chr7:_38669439-38672008	*AMPH*
559	Chr7:_41738732-41740752	*INHBA,LOC285954*
560	Chr7:_42266546-42268823	*GLI3*
561	Chr7:_44142887-44145099	*AEBP1*
562	Chr7:_45017515-45019801	*MYO1G*
563	Chr7:_45065757-45067719	*CCM2*
564	Chr7:_45196181-45198807	*RAMP3*
565	Chr7:_49812008-49816752	*VWC2*
566	Chr7:_50342757-50345519	*IKZF1*
567	Chr7:_50726951-50728913	*GRB10*
568	Chr7:_54608734-54611140	*VSTM2A*
569	Chr7:_66092348-66095174	*KCTD7*
570	Chr7:_69061374-69066037	*AUTS2*
571	Chr7:_70595228-70599382	*WBSCR17*
572	Chr7:_71799757-71803768	*CALN1*
573	Chr7:_75442228-75444120	*CCL24*
574	Chr7:_86272798-86274638	*GRM3*
575	Chr7:_89746892-89750036	*DPY19L2P4*
576	Chr7:_93203085-93205381	*CALCR*
577	Chr7:_93518366-93521184	*TFPI2*
578	Chr7:_96649221-96652551	*DLX5*
579	Chr7:_98244805-98248759	*NPTX2*
580	Chr7:_98466023-98469058	*TMEM130*
581	Chr7:_100488981-100491365	*ACHE*
582	Chr7:_101004899-101008443	*EMID2*
583	Chr7:_102065702-102068002	*PRKRIP1*
584	Chr7:_102066220-102068104	*Non Coding Region*
585	Chr7:_106507057-106509733	*PIK3CG*

Order	Coordinates of Region (hg19)	Gene
586	Chr7:_108094401-108098206	*NRCAM*
587	Chr7:_112725548-112727528	*GPR85*
588	Chr7:_116961841-116965803	*WNT2*
589	Chr7:_127670158-127673853	*LRRC4,SND1*
590	Chr7:_129912246-129914374	*CPA2*
591	Chr7:_132259474-132263527	*PLXNA4*
592	Chr7:_134142115-134145063	*AKR1B1*
593	Chr7:_142493563-142496248	*Non Coding Region*
594	Chr7:_143581125-143583610	*FAM115A*
595	Chr7:_145812030-145815084	*CNTNAP2*
596	Chr7:_149409858-149413351	*KRBA1*
597	Chr7:_149486283-149488769	*SSPO*
598	Chr7:_149915640-149919264	*Non Coding Region*
599	Chr7:_150146886-150148958	*GIMAP8*
600	Chr7:_150328343-150330555	*GIMAP6*
601	Chr7:_150416414-150419018	*GIMAP1*
602	Chr7:_150651807-150654080	*KCNH2*
603	Chr7:_151105244-151108808	*WDR86*
604	Chr7:_152621343-152624305	*Non Coding Region*
605	Chr7:_155163557-155168854	*Non Coding Region*
606	Chr7:_156794355-156800394	*MNX1*
607	Chr7:_157460391-157462229	*PTPRN2*
608	Chr7:_158935507-158939492	*VIPR2*
609	Chr8:_493155-497083	*C8orf42*
610	Chr8:_685830-690496	*Non Coding Region*
611	Chr8:_9759750-9762643	*LOC157627*
612	Chr8:_9761661-9765748	*MIR124-1,LOC157627*
613	Chr8:_10586304-10592109	*SOX7*
614	Chr8:_11203825-11206689	*TDH*
615	Chr8:_15396636-15399287	*TUSC3*
616	Chr8:_21643908-21648845	*GFRA2*
617	Chr8:_21768694-21771080	*DOK2*
618	Chr8:_22959384-22961927	*TNFRSF10C*
619	Chr8:_23561475-23566175	*NKX2-6*
620	Chr8:_24811946-24815299	*NEFL*
621	Chr8:_25899562-25906842	*EBF2*
622	Chr8:_25908292-25910686	*Non Coding Region*
623	Chr8:_26720642-26725566	*ADRA1A*
624	Chr8:_30889630-30891604	*WRN,PURG*
625	Chr8:_32404407-32407901	*NRG1*
626	Chr8:_37821486-37825008	*ADRB3*
627	Chr8:_38577687-38579917	*Non Coding Region*
628	Chr8:_38585064-38587068	*TACC1*
629	Chr8:_38643473-38646760	*TACC1*
630	Chr8:_41164852-41168140	*SFRP1*

Figure 45H

Order	Coordinates of Region (hg19)	Gene
631	Chr8:_41423341-41426300	*Non Coding Region*
632	Chr8:_41752340-41756250	*ANK1*
633	Chr8:_48099163-48101914	*Non Coding Region*
634	Chr8:_49467683-49469959	*Non Coding Region*
635	Chr8:_53850701-53855426	*NPBWR1*
636	Chr8:_54162303-54165443	*OPRK1*
637	Chr8:_54790845-54796141	*RGS20*
638	Chr8:_55365180-55368628	*Non Coding Region*
639	Chr8:_55369170-55373525	*SOX17*
640	Chr8:_55377928-55381186	*Non Coding Region*
641	Chr8:_56851276-56853304	*LYN*
642	Chr8:_57024546-57027543	*MOS*
643	Chr8:_57068687-57071047	*Non Coding Region*
644	Chr8:_58906001-58908822	*FAM110B*
645	Chr8:_59057167-59060414	*FAM110B*
646	Chr8:_65280903-65284043	*Non Coding Region*
647	Chr8:_65289108-65291946	*MIR124-2*
648	Chr8:_65491935-65495452	*BHLHE22*
649	Chr8:_67343497-67345989	*ADHFE1*
650	Chr8:_67872388-67876600	*Non Coding Region*
651	Chr8:_67939691-67941997	*LRRC67*
652	Chr8:_69241920-69244803	*Non Coding Region*
653	Chr8:_70980873-70985888	*PRDM14*
654	Chr8:_72467560-72470561	*Non Coding Region*
655	Chr8:_72754783-72757667	*MSC*
656	Chr8:_72916305-72918641	*Non Coding Region*
657	Chr8:_75895528-75898116	*CRISPLD1*
658	Chr8:_77593255-77595860	*ZFHX4,LOC100192378*
659	Chr8:_85095759-85098247	*RALYL*
660	Chr8:_89338965-89341481	*MMP16*
661	Chr8:_95228915-95230829	*CDH17*
662	Chr8:_97155763-97159030	*GDF6*
663	Chr8:_97170805-97173022	*GDF6*
664	Chr8:_97171634-97174880	*GDF6*
665	Chr8:_97504747-97508607	*SDC2*
666	Chr8:_102148868-102150808	*Non Coding Region*
667	Chr8:_104509870-104514913	*RIMS2*
668	Chr8:_105234384-105236979	*RIMS2*
669	Chr8:_116659432-116661747	*TRPS1*
670	Chr8:_116678698-116680936	*TRPS1*
671	Chr8:_121136099-121138778	*COL14A1*
672	Chr8:_129004430-129006622	*PVT1*
673	Chr8:_132051203-132055749	*Non Coding Region*
674	Chr8:_134071619-134073433	*TG,SLA*
675	Chr8:_140713585-140719259	*KCNK9*

Order	Coordinates of Region (hg19)	Gene
676	Chr8:_143530118-143535495	*Non Coding Region*
677	Chr8:_143857279-143860411	*LYNX1*
678	Chr8:_145102285-145109027	*OPLAH*
679	Chr9:_6644080-6646975	*GLDC*
680	Chr9:_37025222-37029014	*PAX5*
681	Chr9:_77111712-77114583	*RORB*
682	Chr9:_79632326-79637030	*FOXB2*
683	Chr9:_80523239-80525503	*GNAQ*
684	Chr9:_89762336-89764767	*C9orf170*
685	Chr9:_92136795-92138787	*Non Coding Region*
686	Chr9:_100609696-100612517	*Non Coding Region*
687	Chr9:_100614234-100618510	*FOXE1*
688	Chr9:_112809858-112812128	*PALM2-AKAP2,AKAP2*
689	Chr9:_124460797-124463190	*DAB2IP*
690	Chr9:_124887949-124890382	*Non Coding Region*
691	Chr9:_130539846-130542036	*SH2D3C*
692	Chr9:_132381432-132384004	*C9orf50*
693	Chr9:_132651350-132653715	*FNBP1*
694	Chr9:_133533534-133543394	*Non Coding Region*
695	Chr9:_134150853-134154015	*FAM78A*
696	Chr9:_135035713-135039933	*NTNG2*
697	Chr9:_137159400-137161546	*Non Coding Region*
698	Chr10:_1777784-1781018	*ADARB2*
699	Chr10:_7448376-7456339	*SFMBT2*
700	Chr10:_7707524-7709816	*ITIH5*
701	Chr10:_8076829-8079378	*Non Coding Region*
702	Chr10:_8090374-8099329	*FLJ45983,GATA3*
703	Chr10:_8099384-8101768	*GATA3*
704	Chr10:_11206178-11208980	*CUGBP2*
705	Chr10:_14700680-14702950	*FAM107B*
706	Chr10:_15760423-15763101	*ITGA8*
707	Chr10:_17269430-17273617	*VIM*
708	Chr10:_22539707-22543739	*Non Coding Region*
709	Chr10:_22622350-22626875	*Non Coding Region*
710	Chr10:_23461224-23464889	*Non Coding Region*
711	Chr10:_23981382-23983589	*KIAA1217*
712	Chr10:_26726061-26728992	*APBB1IP*
713	Chr10:_27546668-27549402	*Non Coding Region*
714	Chr10:_29095724-29097990	*Non Coding Region*
715	Chr10:_30337216-30339276	*KIAA1462*
716	Chr10:_43949884-43952437	*ZNF487*
717	Chr10:_44787811-44790120	*Non Coding Region*
718	Chr10:_44878714-44883391	*CXCL12*
719	Chr10:_48437411-48440320	*GDF10*
720	Chr10:_50715559-50717397	*ERCC6*

Figure 45I

Order	Coordinates of Region (hg19)	Gene
721	Chr10:_50816601-50821356	*SLC18A3,CHAT*
722	Chr10:_52832539-52835752	*PRKG1*
723	Chr10:_57389343-57392215	*Non Coding Region*
724	Chr10:_64573660-64579778	*EGR2*
725	Chr10:_70846498-70848300	*SRGN*
726	Chr10:_73494938-73497090	*CDH23,C10orf105*
727	Chr10:_81001109-81004687	*ZMIZ1*
728	Chr10:_82115202-82118120	*DYDC2,DYDC1*
729	Chr10:_83632820-83636729	*NRG3*
730	Chr10:_88121924-88128364	*GRID1*
731	Chr10:_91293947-91296925	*SLC16A12*
732	Chr10:_93391667-93394147	*PPP1R3C*
733	Chr10:_94819026-94824252	*CYP26C1*
734	Chr10:_100992820-100995188	*HPSE2*
735	Chr10:_101291436-101293851	*NKX2-3*
736	Chr10:_102418147-102420668	*Non Coding Region*
737	Chr10:_102483200-102485476	*Non Coding Region*
738	Chr10:_102496154-102498675	*Non Coding Region*
739	Chr10:_102506482-102510646	*PAX2*
740	Chr10:_102892660-102896059	*TLX1*
741	Chr10:_102898177-102900489	*Non Coding Region*
742	Chr10:_103602166-103604642	*KCNIP2*
743	Chr10:_105035628-105039084	*INA*
744	Chr10:_105451338-105454230	*SH3PXD2A*
745	Chr10:_106087658-106089746	*ITPRIP*
746	Chr10:_106398567-106403812	*Non Coding Region*
747	Chr10:_108922780-108925805	*SORCS1*
748	Chr10:_118029732-118035230	*GFRA1*
749	Chr10:_118891161-118893639	*VAX1*
750	Chr10:_118921351-118924520	*Non Coding Region*
751	Chr10:_118999435-119002530	*SLC18A2*
752	Chr10:_119292917-119298675	*EMX2OS*
753	Chr10:_119300321-119306368	*EMX2OS,EMX2*
754	Chr10:_119310204-119313104	*Non Coding Region*
755	Chr10:_121276453-121278317	*RGS10*
756	Chr10:_121300478-121304169	*RGS10*
757	Chr10:_123921850-123924542	*TACC2*
758	Chr10:_131760169-131772696	*Non Coding Region*
759	Chr10:_133108193-133112052	*TCERG1L*
760	Chr10:_134900198-134903619	*GPR123*
761	Chr10:_135042221-135045560	*UTF1*
762	Chr10:_135137682-135140717	*CALY*
763	Chr11:_625728-629037	*SCT*
764	Chr11:_635906-641628	*DRD4*
765	Chr11:_2157951-2163484	*IGF2,IGF2AS,INS-IGF2*

Order	Coordinates of Region (hg19)	Gene
766	Chr11:_2320706-2322834	*TSPAN32,C11orf21*
767	Chr11:_2554405-2556618	*KCNQ1*
768	Chr11:_6438863-6441732	*APBB1*
769	Chr11:_6667255-6669173	*DCHS1*
770	Chr11:_8039409-8042558	*Non Coding Region*
771	Chr11:_8101358-8103913	*TUB*
772	Chr11:_9622178-9624178	*Non Coding Region*
773	Chr11:_10326013-10330831	*ADM*
774	Chr11:_12028737-12031841	*DKK3*
775	Chr11:_14925471-14928770	*Non Coding Region*
776	Chr11:_15135058-15137545	*INSC*
777	Chr11:_18811572-18815079	*PTPN5*
778	Chr11:_19365653-19369234	*Non Coding Region*
779	Chr11:_20183570-20186497	*Non Coding Region*
780	Chr11:_20228502-20230532	*Non Coding Region*
781	Chr11:_20617197-20620920	*Non Coding Region*
782	Chr11:_20689579-20692845	*NELL1*
783	Chr11:_30605004-30609128	*MPPED2*
784	Chr11:_31819060-31822416	*PAX6*
785	Chr11:_31826696-31828921	*PAX6*
786	Chr11:_32353760-32356484	*Non Coding Region*
787	Chr11:_32453874-32458311	*WT1,WIT1*
788	Chr11:_33889357-33892495	*LMO2*
789	Chr11:_35439375-35442882	*SLC1A2*
790	Chr11:_43601545-43604215	*MIR129-2*
791	Chr11:_43962626-43967048	*LOC387763*
792	Chr11:_44331314-44333757	*ALX4*
793	Chr11:_46381712-46383738	*DGKZ*
794	Chr11:_46409921-46415687	*Non Coding Region*
795	Chr11:_47415357-47417598	*Non Coding Region*
796	Chr11:_57248946-57251848	*Non Coding Region*
797	Chr11:_58344673-58348321	*LPXN*
798	Chr11:_60737928-60740062	*CD6*
799	Chr11:_61282864-61285147	*SYT7*
800	Chr11:_61321732-61324285	*SYT7*
801	Chr11:_61593996-61597710	*FADS2*
802	Chr11:_63320554-63322570	*HRASLS2*
803	Chr11:_63765058-63829748	*MACROD1*
804	Chr11:_63973829-63976048	*FERMT3*
805	Chr11:_64057808-64061118	*KCNK4*
806	Chr11:_64108822-64111228	*CCDC88B*
807	Chr11:_64620207-64623246	*EHD1*
808	Chr11:_65407344-65409631	*SIPA1*
809	Chr11:_65600069-65602551	*SNX32*
810	Chr11:_65656874-65659970	*FOSL1*

Figure 45J

Order	Coordinates of Region (hg19)	Gene
811	Chr11:_67038074-67040164	*ADRBK1*
812	Chr11:_67167616-67170098	*TBC1D10C*
813	Chr11:_67175945-67178169	*TBC1D10C*
814	Chr11:_69631033-69635710	*FGF3*
815	Chr11:_69705778-69708017	*Non Coding Region*
816	Chr11:_69830571-69833484	*Non Coding Region*
817	Chr11:_69918916-69921064	*Non Coding Region*
818	Chr11:_70210277-70212325	*PPFIA1*
819	Chr11:_70210569-70212493	*PPFIA1*
820	Chr11:_70961173-70965161	*Non Coding Region*
821	Chr11:_71953816-71956659	*PHOX2A*
822	Chr11:_72294167-72296875	*PDE2A*
823	Chr11:_75377416-75380882	*MAP6*
824	Chr11:_78672007-78674213	*ODZ4*
825	Chr11:_91956635-91961290	*Non Coding Region*
826	Chr11:_94500366-94503696	*AMOTL1*
827	Chr11:_104033536-104036058	*PDGFD*
828	Chr11:_105480126-105482422	*GRIA4*
829	Chr11:_106887323-106891203	*GUCY1A2*
830	Chr11:_107460743-107463648	*ELMOD1,LOC643923*
831	Chr11:_111382168-111384892	*BTG4,MIR34C,MIR34B*
832	Chr11:_111806607-111809203	*DIXDC1*
833	Chr11:_112831524-112835490	*NCAM1*
834	Chr11:_115528997-115531548	*Non Coding Region*
835	Chr11:_115629398-115632117	*Non Coding Region*
836	Chr11:_117800500-117802484	*TMPRSS13*
837	Chr11:_119292320-119294943	*THY1*
838	Chr11:_120433822-120436940	*Non Coding Region*
839	Chr11:_122846494-122849608	*Non Coding Region*
840	Chr11:_123065517-123067986	*ASAM*
841	Chr11:_123300049-123303149	*Non Coding Region*
842	Chr11:_124627367-124630590	*ESAM*
843	Chr11:_124734069-124737465	*ROBO3*
844	Chr11:_124789814-124792354	*HEPACAM*
845	Chr11:_125033896-125037426	*PKNOX2*
846	Chr11:_125364137-125367319	*FEZ1*
847	Chr11:_125773292-125775584	*PUS3,DDX25*
848	Chr11:_126224355-126227073	*ST3GAL4*
849	Chr11:_128561671-128566011	*FLI1*
850	Chr11:_132933059-132935291	*OPCML*
851	Chr11:_133824337-133828457	*IGSF9B*
852	Chr11:_133905735-133908359	*LOC100128239*
853	Chr11:_134144559-134148180	*GLB1L3*
854	Chr12:_5538895-5541703	*NTF3*
855	Chr12:_6570272-6572422	*TAPBPL*

Order	Coordinates of Region (hg19)	Gene
856	Chr12:_6663425-6666336	*IFFO1*
857	Chr12:_6881855-6884184	*LAG3*
858	Chr12:_7059274-7061100	*PTPN6*
859	Chr12:_8541555-8544334	*LOC389634*
860	Chr12:_15112174-15114012	*ARHGDIB*
861	Chr12:_15372986-15375514	*RERG*
862	Chr12:_15474318-15476901	*PTPRO*
863	Chr12:_24713956-24717243	*SOX5*
864	Chr12:_25054599-25057246	*BCAT1*
865	Chr12:_29934995-29938433	*TMTC1*
866	Chr12:_39298248-39300629	*CPNE8*
867	Chr12:_41085522-41088102	*CNTN1*
868	Chr12:_45443202-45446386	*DBX2*
869	Chr12:_46608101-46610025	*SLC38A1*
870	Chr12:_47608271-47610443	*LOC100233209, FAM113B*
871	Chr12:_48205721-48208126	*HDAC7*
872	Chr12:_48396889-48399731	*COL2A1*
873	Chr12:_48590977-48593785	*Non Coding Region*
874	Chr12:_49482601-49485255	*DHH*
875	Chr12:_50296580-50298988	*FAIM2*
876	Chr12:_51716888-51718842	*BIN2*
877	Chr12:_52399457-52402696	*GRASP*
878	Chr12:_52625793-52628577	*KRT7*
879	Chr12:_54088211-54090413	*Non Coding Region*
880	Chr12:_54344563-54346916	*Non Coding Region*
881	Chr12:_54392374-54395648	*HOXC9*
882	Chr12:_54407426-54409713	*Non Coding Region*
883	Chr12:_54423610-54426173	*HOXC5,HOXC4*
884	Chr12:_54446744-54449091	*HOXC4*
885	Chr12:_54810981-54813202	*ITGA5*
886	Chr12:_54890426-54892556	*NCKAP1L*
887	Chr12:_56414209-56416333	*IKZF4*
888	Chr12:_57868075-57871182	*ARHGAP9*
889	Chr12:_57997861-57999663	*DTX3*
890	Chr12:_58020294-58023037	*B4GALNT1*
891	Chr12:_58024661-58028056	*B4GALNT1*
892	Chr12:_62583768-62587268	*FAM19A2*
893	Chr12:_64060836-64063774	*DPY19L2*
894	Chr12:_64783010-64785664	*C12orf56*
895	Chr12:_65513878-65516863	*WIF1*
896	Chr12:_66121788-66124328	*Non Coding Region*
897	Chr12:_66275006-66276970	*HMGA2*
898	Chr12:_72664683-72668551	*LOC283392,TRHDE*
899	Chr12:_81470569-81473119	*ACSS3*
900	Chr12:_85304200-85306422	*SLC6A15*

Figure 45K

Order	Coordinates of Region (hg19)	Gene
901	Chr12:_85666340-85668366	*Non Coding Region*
902	Chr12:_95940906-95943979	*USP44*
903	Chr12:_103888148-103890843	*C12orf42*
904	Chr12:_104443100-104444912	*GLT8D2*
905	Chr12:_104849253-105115323	*CHST11*
906	Chr12:_109027341-109029429	*SELPLG*
907	Chr12:_111470061-111474546	*CUX2*
908	Chr12:_113012099-113014529	*Non Coding Region*
909	Chr12:_113228353-113230395	*RPH3A*
910	Chr12:_113493389-113496534	*DTX1*
911	Chr12:_113540697-113543086	*RASAL1*
912	Chr12:_113899750-113907442	*LHX5*
913	Chr12:_114074847-114076915	*Non Coding Region*
914	Chr12:_114842022-114844610	*TBX5*
915	Chr12:_115101687-115104982	*Non Coding Region*
916	Chr12:_115103848-115106548	*Non Coding Region*
917	Chr12:_117797076-117800448	*NOS1*
918	Chr12:_120030494-120034212	*TMEM233*
919	Chr12:_120798372-120800912	*MSI1*
920	Chr12:_122095559-122097775	*MORN3*
921	Chr12:_122229903-122233359	*RHOF*
922	Chr12:_124940314-124942584	*NCOR2*
923	Chr12:_124949705-124951939	*NCOR2*
924	Chr12:_130386609-130390139	*TMEM132D*
925	Chr12:_133462807-133465858	*CHFR*
926	Chr12:_133483658-133486739	*Non Coding Region*
927	Chr13:_20765208-20768779	*GJB2*
928	Chr13:_20805016-20807867	*GJB6*
929	Chr13:_23732808-23735816	*Non Coding Region*
930	Chr13:_25619937-25622554	*Non Coding Region*
931	Chr13:_25742998-25747127	*FAM123A*
932	Chr13:_28365549-28369505	*GSX1*
933	Chr13:_28497226-28500046	*PDX1*
934	Chr13:_28500859-28503090	*Non Coding Region*
935	Chr13:_28502057-28504374	*Non Coding Region*
936	Chr13:_28673227-28676170	*FLT3*
937	Chr13:_29105576-29108254	*Non Coding Region*
938	Chr13:_36704040-36706707	*DCLK1*
939	Chr13:_36918737-36922004	*SPG20*
940	Chr13:_37004581-37007453	*CCNA1*
941	Chr13:_43147277-43150282	*TNFSF11*
942	Chr13:_44358858-44362696	*ENOX1*
943	Chr13:_46424374-46426642	*SIAH3*
944	Chr13:_46755071-46757347	*LCP1*
945	Chr13:_47467229-47469079	*HTR2A*

Order	Coordinates of Region (hg19)	Gene
946	Chr13:_50069022-50071719	*PHF11*
947	Chr13:_53312127-53315045	*LECT1*
948	Chr13:_53418897-53423872	*PCDH8*
949	Chr13:_58202586-58205322	*Non Coding Region*
950	Chr13:_78491425-78494382	*EDNRB*
951	Chr13:_79169114-79172231	*Non Coding Region*
952	Chr13:_84452664-84454897	*SLITRK1*
953	Chr13:_88325600-88327919	*SLITRK5*
954	Chr13:_92050153-92052716	*GPC5*
955	Chr13:_95362152-95367051	*SOX21*
956	Chr13:_96203691-96206496	*CLDN10*
957	Chr13:_99879931-99881963	*UBAC2*
958	Chr13:_99909500-99911674	*UBAC2,GPR18*
959	Chr13:_100309204-100311446	*CLYBL*
960	Chr13:_102567425-102570495	*FGF14*
961	Chr13:_108518292-108522063	*FAM155A*
962	Chr13:_109146798-109150019	*Non Coding Region*
963	Chr13:_110957891-110961590	*COL4A2,COL4A1*
964	Chr13:_110992508-110994478	*COL4A2*
965	Chr13:_112757598-112761491	*Non Coding Region*
966	Chr13:_114896358-114899516	*RASA3*
967	Chr14:_22003937-22006049	*SALL2*
968	Chr14:_22693463-22695631	*Non Coding Region*
969	Chr14:_23833435-23836947	*EFS*
970	Chr14:_24802678-24805353	*ADCY4*
971	Chr14:_29253365-29256069	*C14orf23*
972	Chr14:_36001598-36006013	*INSM2*
973	Chr14:_36985362-36991576	*NKX2-1*
974	Chr14:_48142433-48146589	*MDGA2*
975	Chr14:_51559116-51563487	*TRIM9*
976	Chr14:_52533581-52537722	*NID2*
977	Chr14:_54686134-54688186	*Non Coding Region*
978	Chr14:_54686313-54688315	*Non Coding Region*
979	Chr14:_57282967-57285558	*OTX2OS1*
980	Chr14:_60974732-60979180	*SIX6*
981	Chr14:_70013543-70015993	*Non Coding Region*
982	Chr14:_70040213-70042662	*Non Coding Region*
983	Chr14:_70654095-70657077	*SLC8A3*
984	Chr14:_75799836-75802046	*Non Coding Region*
985	Chr14:_75799929-75802117	*Non Coding Region*
986	Chr14:_75987889-75989733	*BATF*
987	Chr14:_77503622-77505500	*Non Coding Region*
988	Chr14:_77735733-77738772	*NGB*
989	Chr14:_85995494-85997958	*FLRT2*
990	Chr14:_92412581-92415324	*FBLN5*

Figure 45L

Order	Coordinates of Region (hg19)	Gene
991	Chr14:_93531457-93533669	*ITPK1,NCRNA00203*
992	Chr14:_94252952-94256733	*PRIMA1*
993	Chr14:_95233612-95237569	*GSC*
994	Chr14:_96341209-96344570	*Non Coding Region*
995	Chr14:_97684059-97686946	*Non Coding Region*
996	Chr14:_100258160-100260542	*EML1*
997	Chr14:_100436381-100439863	*Non Coding Region*
998	Chr14:_100530519-100532663	*EVL*
999	Chr14:_101922575-101926995	*Non Coding Region*
1000	Chr14:_105308690-105311500	*Non Coding Region*
1001	Chr14:_105392036-105394306	*PLD4*
1002	Chr15:_26106503-26109818	*ATP10A*
1003	Chr15:_27214951-27217856	*GABRG3*
1004	Chr15:_28338774-28343442	*OCA2*
1005	Chr15:_28350606-28354102	*Non Coding Region*
1006	Chr15:_28981669-28984497	*WHAMML2*
1007	Chr15:_29076251-29078684	*Non Coding Region*
1008	Chr15:_34805490-34808382	*Non Coding Region*
1009	Chr15:_37171473-37174018	*LOC145845*
1010	Chr15:_37386386-37388614	*MEIS2*
1011	Chr15:_38987823-38989687	*C15orf53*
1012	Chr15:_41792562-41796305	*ITPKA*
1013	Chr15:_45668970-45672029	*GATM,LOC145663*
1014	Chr15:_48935810-48939577	*FBN1*
1015	Chr15:_51384861-51388203	*TNFAIP8L3*
1016	Chr15:_53079458-53084699	*ONECUT1*
1017	Chr15:_53096561-53099476	*Non Coding Region*
1018	Chr15:_55878792-55882228	*PYGO1*
1019	Chr15:_60295135-60299520	*FOXB1*
1020	Chr15:_61519423-61522716	*RORA*
1021	Chr15:_65688142-65690362	*IGDCC4*
1022	Chr15:_68114485-68123575	*LBXCOR1*
1023	Chr15:_70827082-70829112	*Non Coding Region*
1024	Chr15:_73658860-73663015	*HCN4*
1025	Chr15:_74418870-74424044	*ISLR2,LOC283731*
1026	Chr15:_74724323-74727487	*SEMA7A*
1027	Chr15:_77286731-77288581	*PSTPIP1*
1028	Chr15:_78631669-78634108	*CRABP1*
1029	Chr15:_79380804-79385209	*RASGRF1*
1030	Chr15:_79575059-79577270	*ANKRD34C*
1031	Chr15:_81588232-81590264	*IL16*
1032	Chr15:_83314116-83318541	*CPEB1*
1033	Chr15:_83774861-83777922	*TM6SF1*
1034	Chr15:_83874648-83878079	*HDGFRP3*
1035	Chr15:_83950980-83954930	*BNC1*

Order	Coordinates of Region (hg19)	Gene
1036	Chr15:_84114762-84117921	*SH3GL3*
1037	Chr15:_86232136-86234304	*AKAP13*
1038	Chr15:_88798522-88802017	*NTRK3*
1039	Chr15:_89919793-89923768	*LOC254559*
1040	Chr15:_89951271-89954061	*Non Coding Region*
1041	Chr15:_92395013-92398682	*SLCO3A1*
1042	Chr15:_92935677-92939521	*ST8SIA2*
1043	Chr15:_93630428-93634173	*RGMA*
1044	Chr15:_96894306-96896729	*Non Coding Region*
1045	Chr15:_98835137-98837359	*Non Coding Region*
1046	Chr15:_101418261-101422133	*ALDH1A3*
1047	Chr16:_213343-217720	*HBM*
1048	Chr16:_2040825-2044026	*SYNGR3*
1049	Chr16:_2736416-2738296	*KCTD5*
1050	Chr16:_2764619-2766855	*PRSS27*
1051	Chr16:_3237805-3240492	*Non Coding Region*
1052	Chr16:_3626433-3628561	*NLRC3*
1053	Chr16:_10132364-10134638	*GRIN2A*
1054	Chr16:_11326008-11328234	*Non Coding Region*
1055	Chr16:_12993574-12998690	*SHISA9*
1056	Chr16:_16155017-16157061	*ABCC1*
1057	Chr16:_21293980-21296414	*CRYM*
1058	Chr16:_23845941-23849102	*PRKCB*
1059	Chr16:_25701955-25705631	*HS3ST4*
1060	Chr16:_28072817-28076296	*GSG1L*
1061	Chr16:_28994976-28996788	*LAT*
1062	Chr16:_29085220-29087434	*RRN3P2*
1063	Chr16:_29674845-29677120	*SPN*
1064	Chr16:_29755968-29757934	*C16orf54*
1065	Chr16:_30193747-30196357	*CORO1A*
1066	Chr16:_30388035-30391631	*SEPT_1*
1067	Chr16:_30904549-30906783	*BCL7C,MIR762*
1068	Chr16:_31020986-31022880	*STX1B*
1069	Chr16:_31224956-31229264	*TRIM72,PYDC1*
1070	Chr16:_49310413-49313308	*Non Coding Region*
1071	Chr16:_49474558-49476672	*Non Coding Region*
1072	Chr16:_51182699-51189763	*SALL1*
1073	Chr16:_54406885-54409143	*Non Coding Region*
1074	Chr16:_54407062-54408918	*Non Coding Region*
1075	Chr16:_54969301-54973846	*Non Coding Region*
1076	Chr16:_55688598-55691895	*SLC6A2*
1077	Chr16:_56223731-56225980	*GNAO1,LOC283856*
1078	Chr16:_56695892-56698293	*Non Coding Region*
1079	Chr16:_58017400-58020471	*TEPP*
1080	Chr16:_66611749-66614412	*CMTM2,CMTM1*

Figure 45M

Order	Coordinates of Region (hg19)	Gene
1081	Chr16:_66637254-66640561	*CMTM3*
1082	Chr16:_67195792-67200988	*FBXL8,HSF4*
1083	Chr16:_67426284-67429950	*ZDHHC1,TPPP3*
1084	Chr16:_77821375-77824134	*VAT1L*
1085	Chr16:_86540430-86546228	*Non Coding Region*
1086	Chr16:_86598355-86603649	*FOXC2*
1087	Chr16:_86611188-86614821	*FOXL1*
1088	Chr16:_86964271-86966496	*Non Coding Region*
1089	Chr16:_87735652-87737907	*LOC100129637*
1090	Chr16:_88764041-88766399	*RNF166*
1091	Chr16:_88771237-88774195	*RNF166*
1092	Chr16:_90087915-90090413	*GAS8*
1093	Chr17:_1626745-1630897	*WDR81*
1094	Chr17:_4612400-4615423	*ARRB2*
1095	Chr17:_4801265-4806402	*C17orf107,CHRNE*
1096	Chr17:_4849942-4854907	*RNF167*
1097	Chr17:_5402131-5405602	*LOC728392*
1098	Chr17:_5486298-5488454	*NLRP1*
1099	Chr17:_6615422-6618471	*SLC13A5*
1100	Chr17:_7239301-7241145	*ACAP1*
1101	Chr17:_7553139-7556338	*ATP1B2*
1102	Chr17:_7975342-7977570	*ALOX12B*
1103	Chr17:_8229195-8231941	*Non Coding Region*
1104	Chr17:_8532004-8535933	*MYH10*
1105	Chr17:_10023321-10103440	*GAS7*
1106	Chr17:_13502829-13506559	*HS3ST3A1*
1107	Chr17:_14211363-14213788	*HS3ST3B1*
1108	Chr17:_19482305-19484687	*Non Coding Region*
1109	Chr17:_19879676-19882633	*AKAP10*
1110	Chr17:_27037084-27039919	*PROCA1*
1111	Chr17:_27938298-27941770	*ANKRD13B*
1112	Chr17:_32483007-32485280	*ACCN1*
1113	Chr17:_33699989-33702657	*SLFN11*
1114	Chr17:_33866708-33868882	*Non Coding Region*
1115	Chr17:_35290899-35301875	*LHX1*
1116	Chr17:_37320482-37323099	*ARL5C*
1117	Chr17:_37909088-37912190	*Non Coding Region*
1118	Chr17:_38346533-38348765	*RAPGEFL1*
1119	Chr17:_38720636-38722714	*CCR7*
1120	Chr17:_42461135-42464579	*ITGA2B*
1121	Chr17:_43046547-43048954	*Non Coding Region*
1122	Chr17:_43297725-43300640	*FMNL1*
1123	Chr17:_43338124-43340832	*C17orf46,LOC100133991*
1124	Chr17:_43859684-43863928	*CRHR1*
1125	Chr17:_44894800-44898025	*WNT3*

Order	Coordinates of Region (hg19)	Gene
1126	Chr17:_44927287-44930690	*WNT9B*
1127	Chr17:_45809589-45812388	*TBX21*
1128	Chr17:_45866103-45868744	*Non Coding Region*
1129	Chr17:_46801790-46804076	*HOXB13*
1130	Chr17:_47071820-47077042	*IGF2BP1*
1131	Chr17:_47652211-47655369	*NXPH3*
1132	Chr17:_48069698-48073654	*DLX3*
1133	Chr17:_54673873-54676232	*Non Coding Region*
1134	Chr17:_55212599-55214527	*Non Coding Region*
1135	Chr17:_55950987-55953139	*CUEDC1*
1136	Chr17:_56407245-56409149	*Non Coding Region*
1137	Chr17:_59472060-59484266	*TBX2*
1138	Chr17:_59530723-59536254	*TBX4*
1139	Chr17:_66595072-66598578	*FAM20A*
1140	Chr17:_70215346-70217318	*Non Coding Region*
1141	Chr17:_70722349-70724423	*SLC39A11*
1142	Chr17:_71159922-71163350	*SSTR2*
1143	Chr17:_72351827-72354798	*BTBD17*
1144	Chr17:_72587360-72589464	*CD300LD,C17orf77*
1145	Chr17:_72754558-72756736	*SLC9A3R1*
1146	Chr17:_72888349-72891060	*FADS6*
1147	Chr17:_73582838-73587337	*MYO15B*
1148	Chr17:_73748618-73751178	*ITGB4*
1149	Chr17:_74510783-74512655	*Non Coding Region*
1150	Chr17:_74532281-74535566	*CYGB*
1151	Chr17:_75367688-75371506	*SEPT_9*
1152	Chr17:_75397284-75399527	*SEPT_9*
1153	Chr17:_75440383-75442401	*SEPT_9*
1154	Chr17:_76126521-76129406	*TMC6,TMC8*
1155	Chr17:_76353818-76358038	*SOCS3*
1156	Chr17:_76360065-76362267	*Non Coding Region*
1157	Chr17:_76754808-76779390	*CYTH1*
1158	Chr17:_76919684-76922961	*TIMP2*
1159	Chr17:_77177828-77181229	*HRNBP3*
1160	Chr17:_77720246-77723497	*Non Coding Region*
1161	Chr17:_78448507-78453783	*Non Coding Region*
1162	Chr17:_79258282-79260537	*SLC38A10*
1163	Chr17:_80259390-80261268	*Non Coding Region*
1164	Chr17:_80360614-80363151	*C17orf101*
1165	Chr17:_80407294-80409879	*C17orf62*
1166	Chr17:_80544433-80546658	*FOXK2*
1167	Chr18:_903578-910574	*ADCYAP1*
1168	Chr18:_5195243-5198389	*LOC642597*
1169	Chr18:_5542437-5545241	*EPB41L3*
1170	Chr18:_7115852-7119241	*LAMA1*

Figure 45N

Order	Coordinates of Region (hg19)	Gene
1171	Chr18:_11749953-11753756	*GNAL*
1172	Chr18:_18820383-18824349	*GREB1L*
1173	Chr18:_20810280-20812536	*CABLES1*
1174	Chr18:_22929018-22931904	*ZNF521*
1175	Chr18:_25754414-25759175	*CDH2*
1176	Chr18:_34832554-34835606	*BRUNOL4*
1177	Chr18:_35143907-35148628	*BRUNOL4*
1178	Chr18:_44336510-44339100	*ST8SIA5*
1179	Chr18:_53256018-53258309	*TCF4*
1180	Chr18:_53446074-53448842	*Non Coding Region*
1181	Chr18:_54787959-54790194	*Non Coding Region*
1182	Chr18:_56886091-56888665	*GRP*
1183	Chr18:_56933833-56937827	*RAX*
1184	Chr18:_56938624-56942540	*RAX*
1185	Chr18:_65182237-65185014	*DSEL*
1186	Chr18:_67066509-67070168	*DOK6*
1187	Chr18:_73166402-73168920	*Non Coding Region*
1188	Chr18:_74960556-74964822	*GALR1*
1189	Chr18:_76736005-76742244	*SALL3*
1190	Chr19:_1400092-1402784	*GAMT*
1191	Chr19:_1464206-1472241	*APC2*
1192	Chr19:_2084223-2086379	*MOBKL2A*
1193	Chr19:_2084577-2086389	*MOBKL2A*
1194	Chr19:_2249560-2254959	*JSRP1*
1195	Chr19:_3177741-3180986	*S1PR4*
1196	Chr19:_4325878-4328380	*STAP2*
1197	Chr19:_4327658-4329865	*STAP2*
1198	Chr19:_4915560-4917784	*UHRF1*
1199	Chr19:_5337788-5340130	*PTPRS*
1200	Chr19:_6772045-6774103	*VAV1*
1201	Chr19:_11532198-11534619	*CCDC151*
1202	Chr19:_11688417-11690768	*ACP5*
1203	Chr19:_12202028-12204503	*ZNF788*
1204	Chr19:_12720532-12723116	*ZNF791*
1205	Chr19:_13119986-13125368	*NFIX*
1206	Chr19:_13214244-13216543	*Non Coding Region*
1207	Chr19:_13615752-13618267	*CACNA1A*
1208	Chr19:_14550998-14553255	*PKN1*
1209	Chr19:_15089016-15091873	*Non Coding Region*
1210	Chr19:_15216951-15219617	*SYDE1*
1211	Chr19:_15579382-15581863	*PGLYRP2*
1212	Chr19:_17006101-17009857	*CPAMD8*
1213	Chr19:_17391383-17394775	*ANKLE1*
1214	Chr19:_17957490-17959808	*JAK3*
1215	Chr19:_18110566-18113481	*ARRDC2*

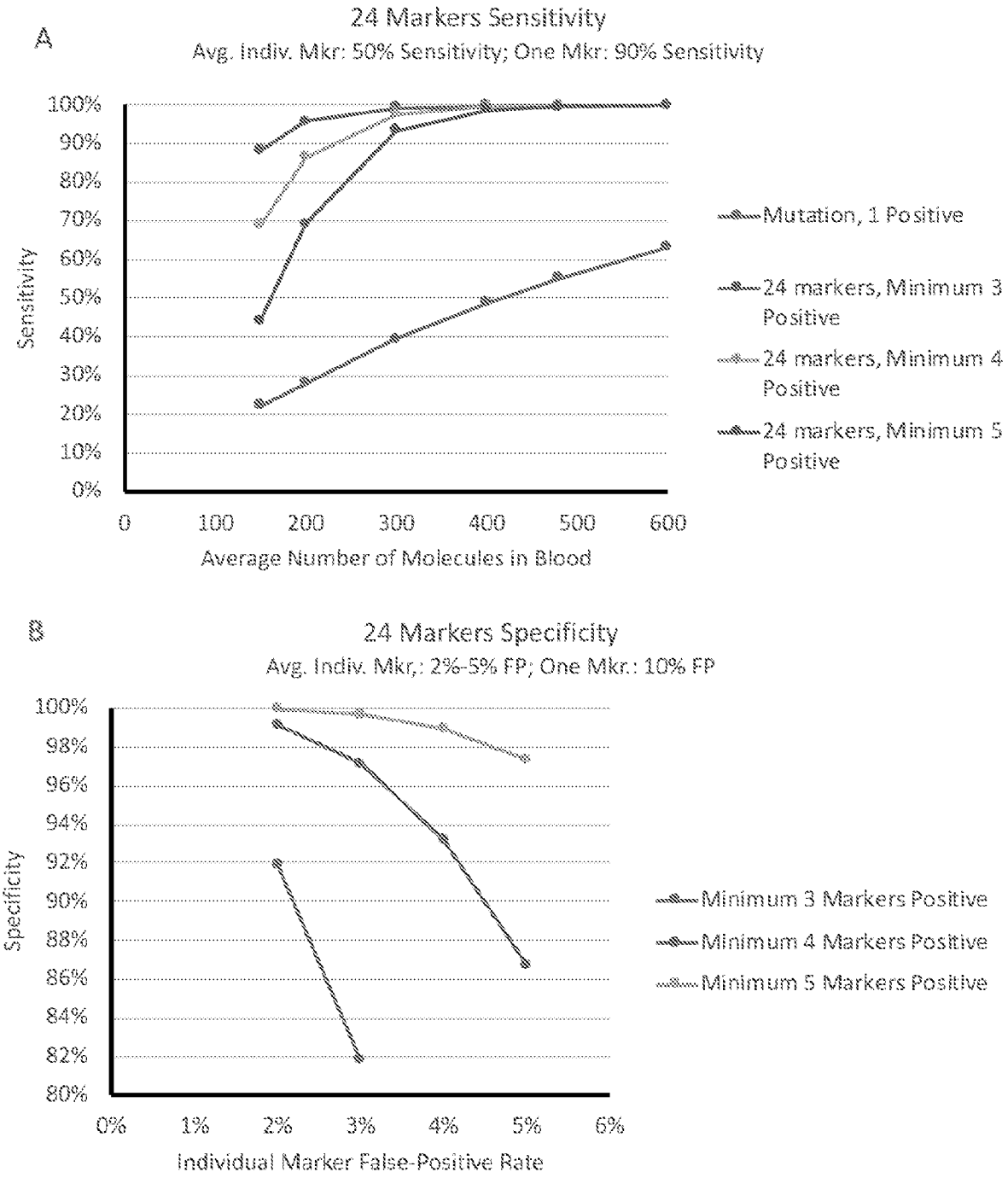
Order	Coordinates of Region (hg19)	Gene
1216	Chr19:_18117741-18120553	*ARRDC2*
1217	Chr19:_18538781-18541341	*SSBP4*
1218	Chr19:_18898037-18903284	*COMP*
1219	Chr19:_19321675-19323916	*NCAN*
1220	Chr19:_21104962-21107144	*ZNF85*
1221	Chr19:_22017594-22019827	*ZNF43*
1222	Chr19:_22033394-22035943	*Non Coding Region*
1223	Chr19:_30014781-30022367	*VSTM2B*
1224	Chr19:_31838635-31844049	*TSHZ3*
1225	Chr19:_31846946-31849209	*Non Coding Region*
1226	Chr19:_34111279-34115353	*CHST8*
1227	Chr19:_36230186-36233219	*TMEM149*
1228	Chr19:_36522391-36524887	*CLIP3*
1229	Chr19:_36908281-36910854	*ZFP82*
1230	Chr19:_37094680-37097589	*ZNF529,ZNF382*
1231	Chr19:_37156632-37159119	*ZNF461*
1232	Chr19:_37405931-37408463	*ZNF829,ZNF568*
1233	Chr19:_37462991-37465606	*Non Coding Region*
1234	Chr19:_37996790-37999125	*ZNF793*
1235	Chr19:_38745638-38748379	*PPP1R14A*
1236	Chr19:_39108432-39110862	*MAP4K1,EIF3K*
1237	Chr19:_41594895-41597140	*CYP2A13*
1238	Chr19:_42636277-42638684	*POU2F2*
1239	Chr19:_42702604-42704976	*DEDD2*
1240	Chr19:_42826743-42830149	*TMEM145*
1241	Chr19:_46995327-46999437	*PNMAL2*
1242	Chr19:_48016717-48019782	*Non Coding Region*
1243	Chr19:_48751899-48754081	*CARD8*
1244	Chr19:_48917115-48919340	*GRIN2D*
1245	Chr19:_48982562-48985175	*CYTH2*
1246	Chr19:_49840187-49842628	*CD37*
1247	Chr19:_50029981-50032300	*RCN3*
1248	Chr19:_50036078-50038670	*RCN3*
1249	Chr19:_51141284-51143986	*SYT3*
1250	Chr19:_51160923-51163514	*Non Coding Region*
1251	Chr19:_51829064-51832145	*IGLON5*
1252	Chr19:_51923999-51926413	*Non Coding Region*
1253	Chr19:_52872084-52874572	*ZNF880*
1254	Chr19:_53634625-53637230	*ZNF415*
1255	Chr19:_53756819-53759148	*ZNF677*
1256	Chr19:_54022868-54025560	*ZNF331*
1257	Chr19:_54411710-54414087	*Non Coding Region*
1258	Chr19:_54926902-54929225	*TTYH1*
1259	Chr19:_55590905-55594980	*EPS8L1*
1260	Chr19:_55768269-55771875	*SAPS1*

Figure 45O

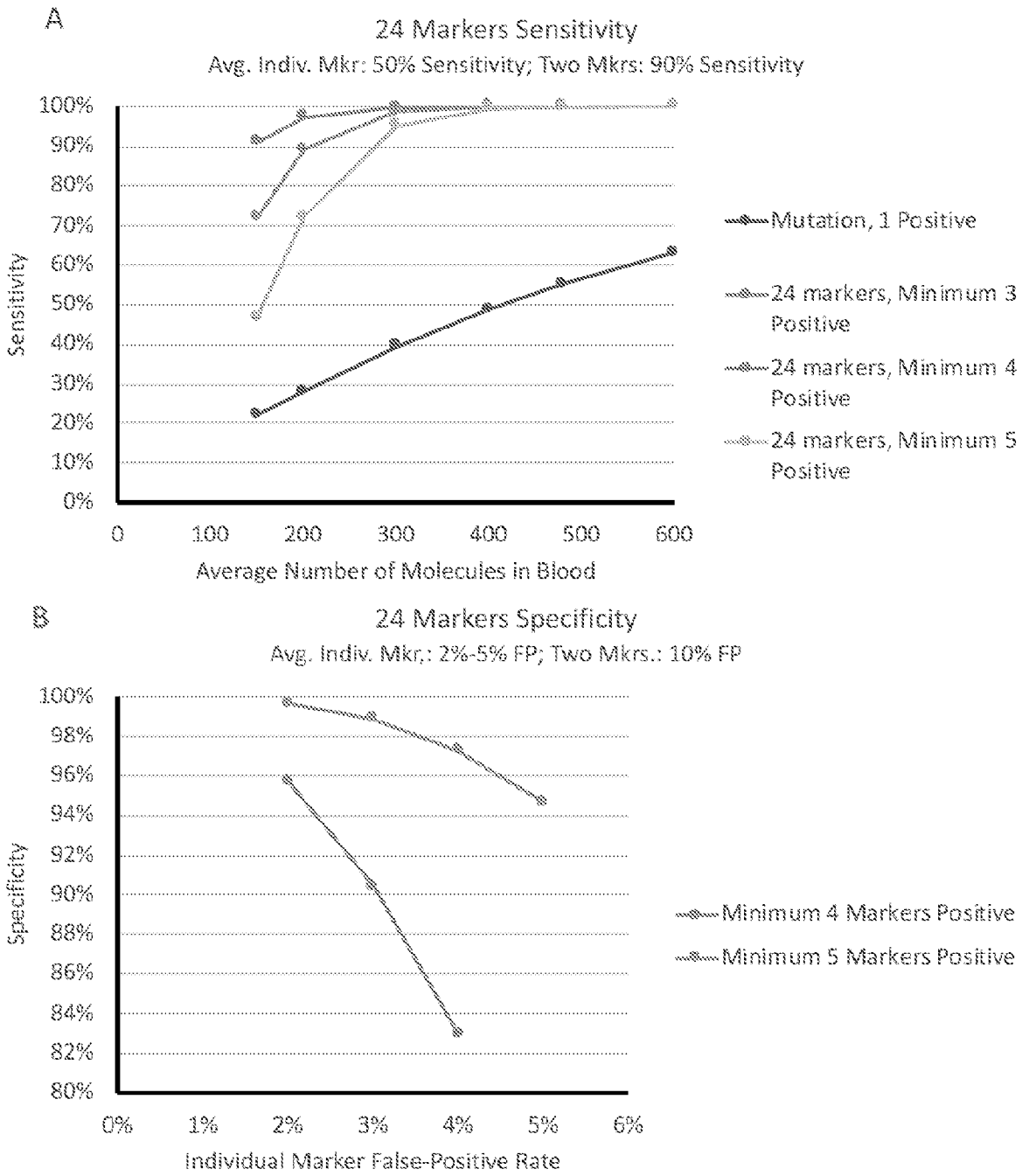
Order	Coordinates of Region (hg19)	Gene
1261	Chr19:_55835823-55837681	*TMEM150B*
1262	Chr19:_56878417-56880995	*ZNF542*
1263	Chr19:_56903636-56906355	*ZNF582*
1264	Chr19:_56987313-56990741	*ZNF667*
1265	Chr19:_57048675-57051646	*ZFP28*
1266	Chr19:_58093739-58096764	*ZIK1*
1267	Chr19:_58219189-58221517	*ZNF154*
1268	Chr19:_58237585-58240028	*ZNF671*
1269	Chr19:_58457686-58460219	*ZNF256*
1270	Chr19:_58608338-58610988	*ZSCAN18*
1271	Chr19:_58950214-58953250	*ZNF132*
1272	Chr20:_2538133-2540877	*TMC2*
1273	Chr20:_2779978-2782497	*CPXM1*
1274	Chr20:_3051097-3054103	*OXT*
1275	Chr20:_3062005-3065146	*AVP*
1276	Chr20:_3661310-3664187	*ADAM33*
1277	Chr20:_3712159-3714515	*HSPA12B*
1278	Chr20:_4794773-4796999	*RASSF2*
1279	Chr20:_4802013-4805146	*RASSF2*
1280	Chr20:_6031675-6034517	*LRRN4*
1281	Chr20:_8111884-8114592	*PLCB1*
1282	Chr20:_10199088-10201384	*SNAP25*
1283	Chr20:_21484932-21497714	*NKX2-2*
1284	Chr20:_21682744-21685420	*Non Coding Region*
1285	Chr20:_23027403-23033218	*THBD*
1286	Chr20:_23065842-23068046	*CD93*
1287	Chr20:_25057288-25059726	*VSX1*
1288	Chr20:_25060761-25063928	*VSX1*
1289	Chr20:_30638908-30641786	*HCK*
1290	Chr20:_34358673-34361250	*Non Coding Region*
1291	Chr20:_34892891-34896214	*Non Coding Region*
1292	Chr20:_36011595-36014439	*SRC*
1293	Chr20:_37351130-37358372	*SLC32A1*
1294	Chr20:_37433206-37436592	*PPP1R16B*
1295	Chr20:_41816475-41820212	*PTPRT*
1296	Chr20:_42874764-42877184	*GDAP1L1*
1297	Chr20:_43437737-43440546	*RIMS4*
1298	Chr20:_44934932-44938310	*Non Coding Region*
1299	Chr20:_44940342-44942569	*Non Coding Region*
1300	Chr20:_45141000-45143337	*ZNF334*
1301	Chr20:_47933839-47936684	*Non Coding Region*
1302	Chr20:_50720310-50722912	*ZFP64*
1303	Chr20:_53090733-53093993	*DOK5*
1304	Chr20:_55499347-55502102	*Non Coding Region*
1305	Chr20:_55963916-55966271	*RBM38*

Order	Coordinates of Region (hg19)	Gene
1306	Chr20:_57223695-57227322	*STX16*
1307	Chr20:_58178808-58181787	*PHACTR3*
1308	Chr20:_61048361-61052897	*GATA5*
1309	Chr20:_61635171-61640055	*BHLHE23*
1310	Chr20:_61805254-61811867	*MIR124-3*
1311	Chr20:_61883644-61887387	*NKAIN4,FLJ16779*
1312	Chr20:_62101541-62104995	*KCNQ2*
1313	Chr20:_62192966-62199985	*PRIC285*
1314	Chr20:_62199085-62201320	*PRIC285*
1315	Chr20:_62672792-62675131	*Non Coding Region*
1316	Chr20:_62732463-62735322	*Non Coding Region*
1317	Chr21:_27010624-27013398	*JAM2*
1318	Chr21:_27944010-27946646	*CYYR1*
1319	Chr21:_28336856-28341237	*ADAMTS5*
1320	Chr21:_32715044-32717485	*TIAM1*
1321	Chr21:_34394128-34401245	*OLIG2*
1322	Chr21:_34441206-34445543	*OLIG1*
1323	Chr21:_36420527-36422479	*RUNX1*
1324	Chr21:_38078941-38082833	*SIM2*
1325	Chr21:_39869594-39871814	*ERG*
1326	Chr21:_45335990-45337816	*AGPAT3*
1327	Chr21:_46350328-46353911	*Non Coding Region*
1328	Chr21:_47516651-47519999	*COL6A2*
1329	Chr22:_18216056-18218226	*BID*
1330	Chr22:_19752312-19756013	*TBX1*
1331	Chr22:_20789638-20793665	*SCARF2*
1332	Chr22:_23482997-23485309	*RTDR1*
1333	Chr22:_24037489-24039557	*LOC91316,RGL4*
1334	Chr22:_24818537-24821810	*ADORA2A*
1335	Chr22:_25677576-25680273	*Non Coding Region*
1336	Chr22:_28191793-28199592	*MN1*
1337	Chr22:_30661898-30664076	*OSM*
1338	Chr22:_37881061-37883095	*MFNG*
1339	Chr22:_37961481-37963819	*CDC42EP1*
1340	Chr22:_39783354-39786104	*Non Coding Region*
1341	Chr22:_39852323-39855065	*MGAT3*
1342	Chr22:_44256942-44259612	*SULT4A1*
1343	Chr22:_45063159-45066092	*PRR5*
1344	Chr22:_45402036-45407372	*PHF21B*
1345	Chr22:_46261065-46264747	*Non Coding Region*
1346	Chr22:_47188484-47190707	*TBC1D22A*
1347	Chr22:_50452056-50455137	*Non Coding Region*
1348	Chr22:_50622164-50626226	*TRABD*
1349	ChrX:_6143120-6146906	*NLGN4X*

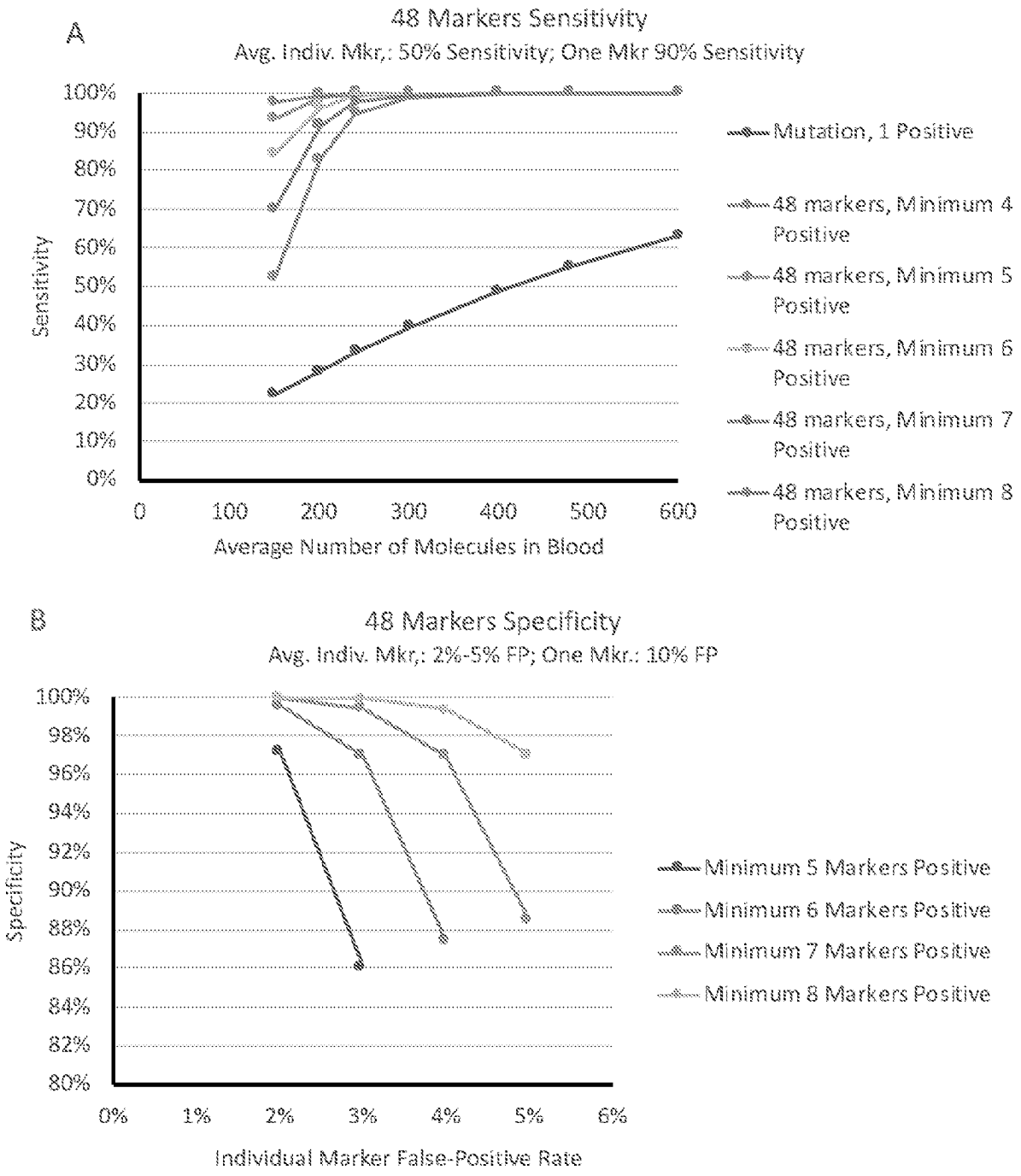
Figure 45P



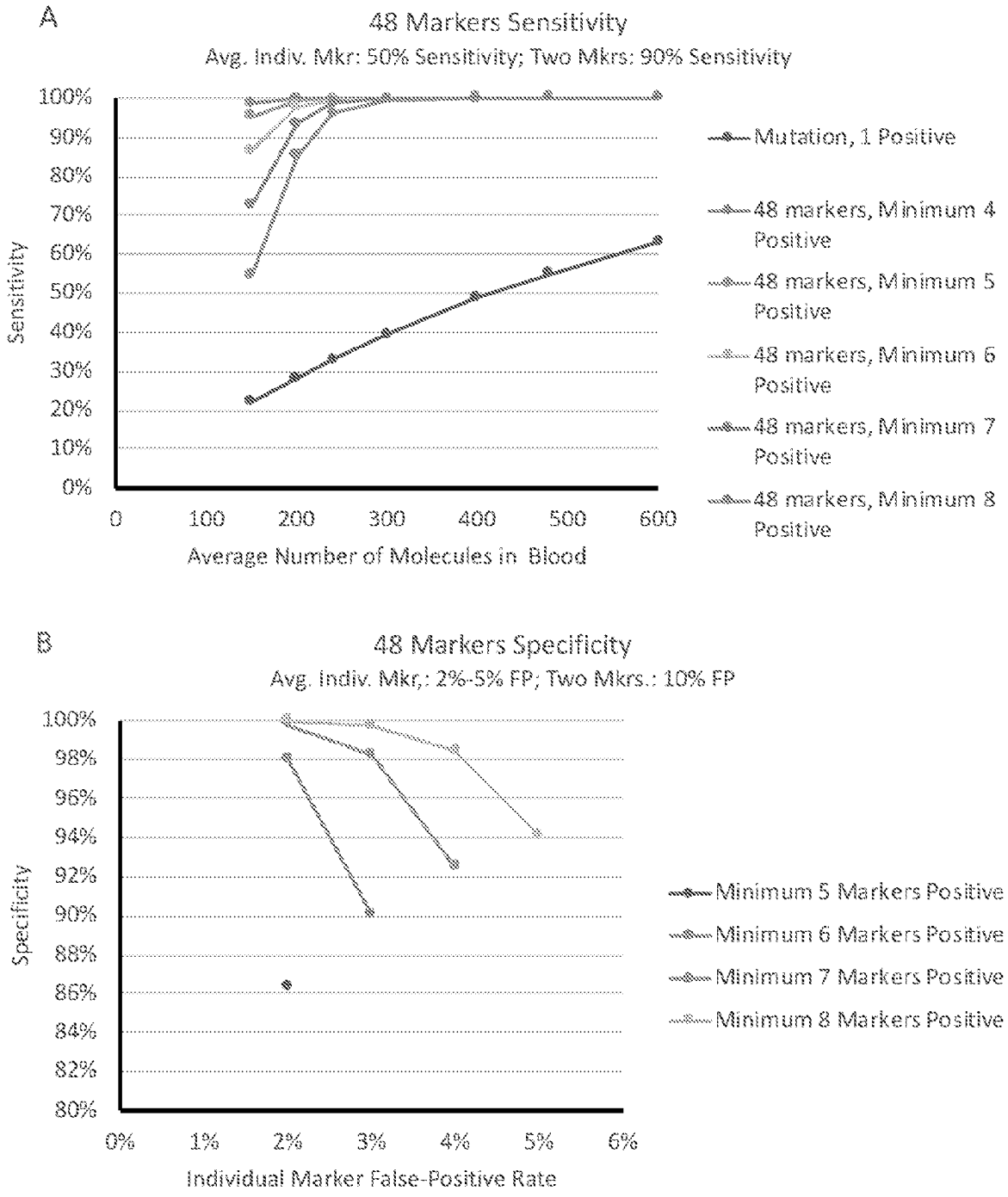
Figures 46A-B



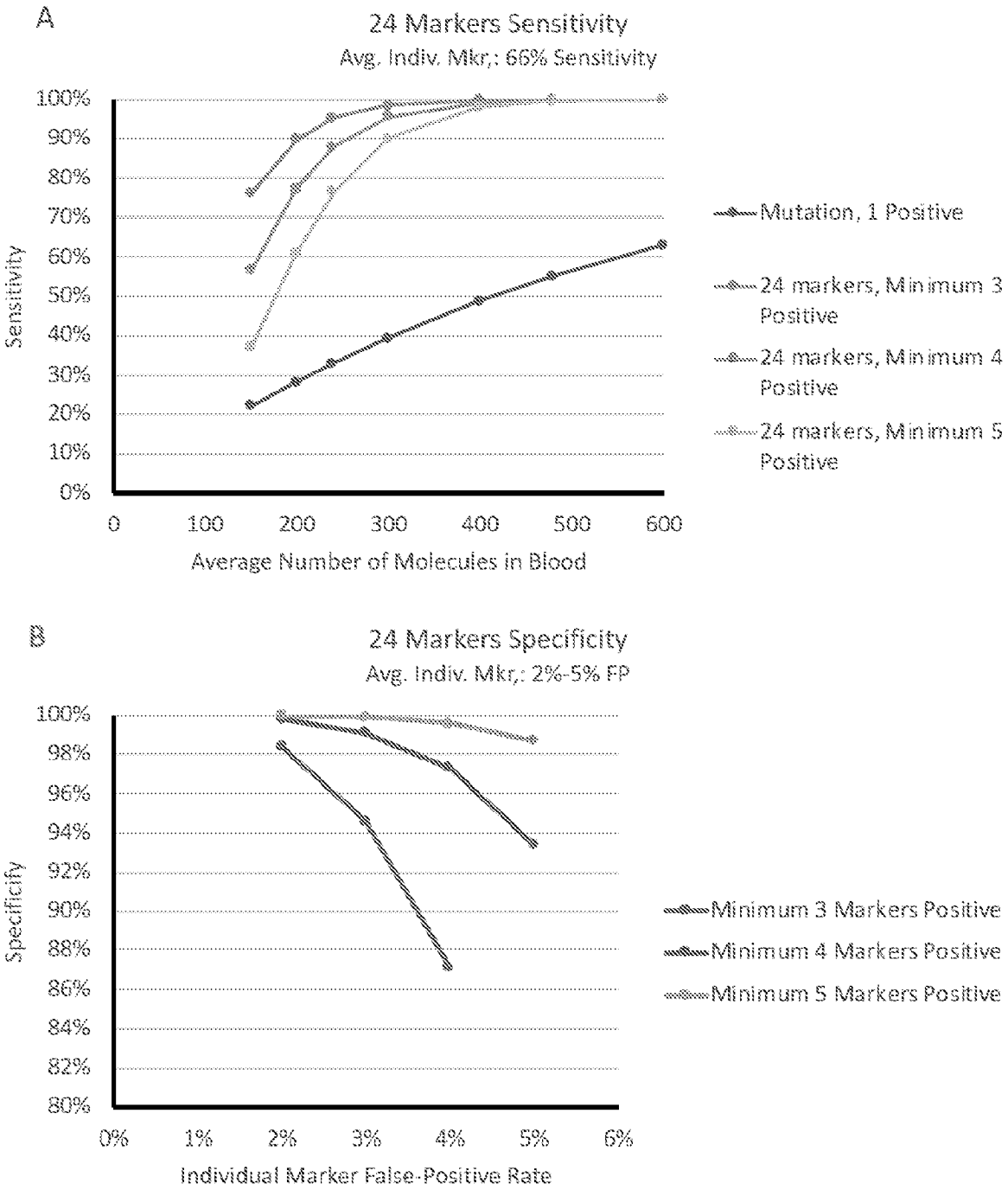
Figures 47A-B



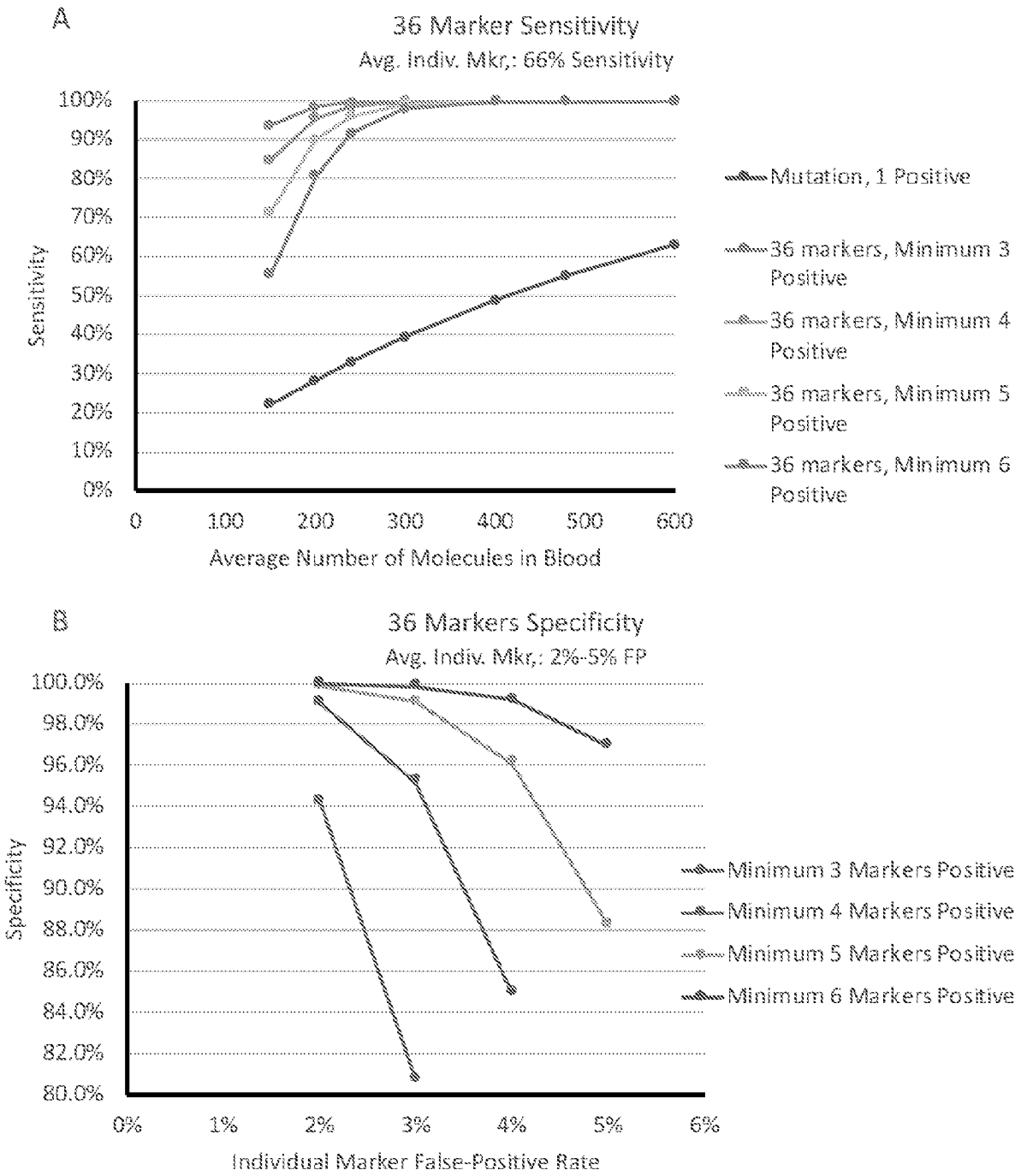
Figures 48A-B



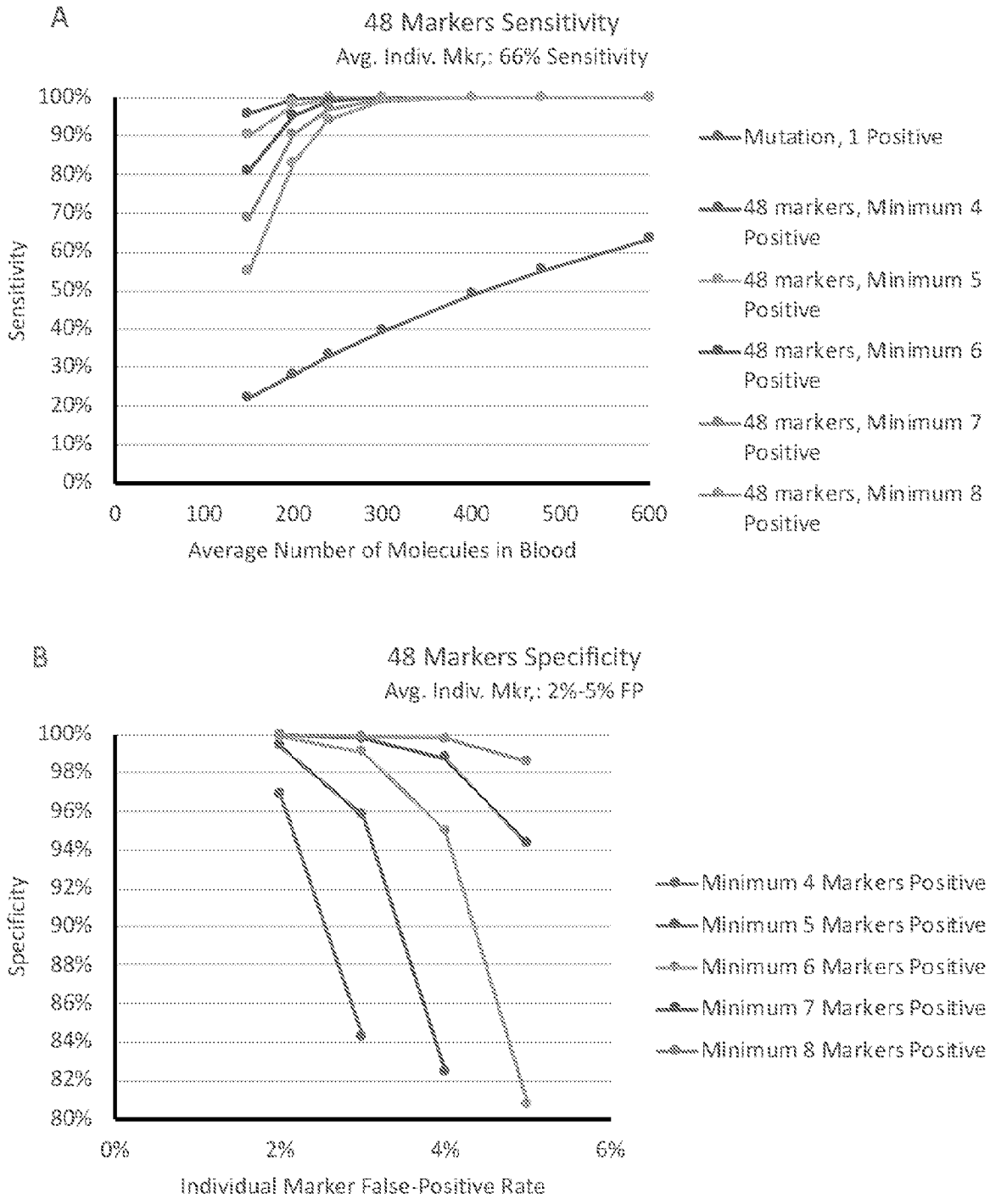
Figures 49A-B



Figures 50A-B



Figures 51A-B



Figures 52A-B

Solid tumors ncRNA markers

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
1	RP11-54O7.18	chr1:1062208-1063288	ENSG00000273443.1
2	RP4-758J18.13	chr1:1409096-1410618	ENSG00000272455.1
3	SNORD99	chr1:28578749-28578822	ENSG00000221539.1
4	RP1-228H13.5	chr1:40464319-40466767	ENSG00000260920.2
5	CYP4A22-AS1	chr1:47096653-47179271	ENSG00000225506.2
6	RP11-196G18.22	chr1:149844498-149849024	ENSG00000261716.1
7	SNORA2	chr2:10155072-10155206	ENSG00000206647.1
8	RP11-443B20.1	chr2:24825610-24826717	ENSG00000271936.1
9	AC074117.10	chr2:27356246-27367622	ENSG00000234072.1
10	YEATS2-AS1	chr3:183806457-183810783	ENSG00000233885.6
11	RP11-480A16.1	chr3:195949188-195952695	ENSG00000260261.1
12	RP11-572O17.1	chr4:1712821-1713622	ENSG00000270195.1
13	LEF1-AS1	chr4:108167525-108256836	ENSG00000232021.5
14	RP11-138J23.1	chr5:104079911-104105403	ENSG00000251026.1
15	TFAP2A-AS1	chr6:10409340-10416446	ENSG00000229950.1
16	RP3-337H4.9	chr6:43519180-43519724	ENSG00000271754.1
17	RAET1K	chr6:149998019-150005157	ENSG00000218358.2
18	AC007128.1	chr7:8262264-8344516	ENSG00000229970.3
19	PMS2P3	chr7:75502930-75528148	ENSG00000127957.15
20	RNF139-AS1	chr8:124462485-124474576	ENSG00000245149.3
21	PVT1	chr8:127794533-128101253	ENSG00000249859.6
22	CTD-3065J16.9	chr8:144078002-144079265	ENSG00000255224.1
23	RP11-435O5.2	chr9:95414834-95426796	ENSG00000237857.2
24	RP11-152N13.16	chr10:73124573-73125532	ENSG00000272599.2
25	RP11-564D11.3	chr10:122879851-122898024	ENSG00000255624.2
26	CTD-2371O3.3	chr11:9459556-9460702	ENSG00000268403.2
27	RP11-540A21.2	chr11:9754770-9759533	ENSG00000245522.2
28	RPLP0P2	chr11:61615036-61639449	ENSG00000243742.4
29	DDX11-AS1	chr12:31020763-31073847	ENSG00000245614.3
30	RP11-680A11.5	chr12:53298655-53300314	ENSG00000257605.2
31	RP11-968A15.2	chr12:54262615-54279063	ENSG00000257596.1
32	MRPL40P1	chr12:67351436-67352039	ENSG00000256037.1
33	RP11-34F20.7	chr13:50125816-50128463	ENSG00000274270.1
34	RP11-93H24.3	chr13:52651305-52652279	ENSG00000273723.1
35	SNHG10	chr14:95532297-95534872	ENSG00000247092.5
36	ARHGAP11B	chr15:30624494-30772993	ENSG00000187951.9
37	RP11-932O9.10	chr15:30648797-30649529	ENSG00000269974.1
38	CTD-2574D22.4	chr16:29919634-29921905	ENSG00000260114.2
39	RP11-199F11.2	chr17:7685260-7686371	ENSG00000262251.1
40	UBE2SP1	chr17:15704232-15704900	ENSG00000233966.1
41	RP11-1381I.3	chr17:16440479-16440952	ENSG00000266651.1
42	UBE2SP2	chr17:18677261-18677757	ENSG00000224126.2
43	SNORD7	chr17:35573657-35573753	ENSG00000207297.1
44	RP5-890E16.2	chr17:48045141-48048073	ENSG00000263412.1
45	RP11-353N14.1	chr17:79800598-79802529	ENSG00000262768.2
46	RP5-836N17.4	chr20:32116171-32116629	ENSG00000275576.1
47	RP5-908M14.10	chr20:62402236-62405935	ENSG00000275437.1
48	LINC01311	chr22:19171395-19172839	ENSG00000260924.2
49	RP3-323A16.1	chr22:35160947-35215025	ENSG00000279184.1

Figure 53

Solid Tumor Exon RNA sequence markers

Order	Exon	Gene
1	chr1:1455928-1455997:+	ATAD3A
2	chr1:38167426-38167490:+	CDCA8
3	chr1:38173994-38175389:+	CDCA8
4	chr1:43630314-43630476:-	EBNA1BP2
5	chr1:43824692-43825067:+	CDC20
6	chr1:43825161-43825309:+	CDC20
7	chr1:43825396-43825492:+	CDC20
8	chr1:43825640-43825768:+	CDC20
9	chr1:43825864-43826060:+	CDC20
10	chr1:43826170-43826264:+	CDC20
11	chr1:43826404-43826632:+	CDC20
12	chr1:43827866-43827983:+	CDC20
13	chr1:43828622-43828871:+	CDC20
14	chr1:45228018-45228123:+	KIF2C
15	chr1:45228217-45228330:+	KIF2C
16	chr1:45232498-45232621:+	KIF2C
17	chr1:45232769-45233436:+	KIF2C
18	chr1:46739810-46739888:+	RAD54L
19	chr1:46740210-46740389:+	RAD54L
20	chr1:47715811-47717594:-	STIL
21	chr1:47728575-47728788:-	STIL
22	chr1:47737748-47737913:-	STIL
23	chr1:145764595-145765434:-	GPR89A
24	chr1:154959942-154961487:+	FLAD1
25	chr1:154962036-154962183:+	FLAD1
26	chr1:155182176-155182358:+	MTX1
27	chr1:155216997-155217746:-	FAM189B
28	chr1:156281832-156282012:-	CCT3
29	chr1:156286957-156287038:-	CCT3
30	chr1:156287206-156287338:-	CCT3
31	chr1:156495198-156496391:-	IQGAP3
32	chr1:156497744-156497852:-	IQGAP3
33	chr1:156498301-156498402:-	IQGAP3
34	chr1:156498708-156498874:-	IQGAP3
35	chr1:156499897-156500109:-	IQGAP3
36	chr1:156500952-156501039:-	IQGAP3
37	chr1:156502772-156502898:-	IQGAP3
38	chr1:156503565-156503667:-	IQGAP3
39	chr1:156503801-156503933:-	IQGAP3
40	chr1:156504293-156504525:-	IQGAP3
41	chr1:156504955-156505039:-	IQGAP3
42	chr1:156506973-156507113:-	IQGAP3
43	chr1:156509630-156509782:-	IQGAP3
44	chr1:156510500-156510708:-	IQGAP3
45	chr1:156521497-156521660:-	IQGAP3

Order	Exon	Gene
46	chr1:156532926-156533104:-	IQGAP3
47	chr1:161142017-161142121:-	B4GALT3
48	chr1:161143395-161143867:-	B4GALT3
49	chr1:163317553-163317728:+	NUF2
50	chr1:163318735-163318870:+	NUF2
51	chr1:165865427-165865569:+	UCK2
52	chr1:165872419-165872516:+	UCK2
53	chr1:165875158-165875206:+	UCK2
54	chr1:165876921-165877337:+	UCK2,MIR3658
55	chr1:173825804-173825879:+	DARS2
56	chr1:197053258-197053556:-	ASPM
57	chr1:197055933-197056102:-	ASPM
58	chr1:197057250-197057562:-	ASPM
59	chr1:197059060-197059214:-	ASPM
60	chr1:197059326-197059518:-	ASPM
61	chr1:197059980-197060171:-	ASPM
62	chr1:202300786-202301088:-	UBE2T
63	chr1:202302138-202302221:-	UBE2T
64	chr1:203830727-203830871:+	SNRPE
65	chr1:203831310-203831350:+	SNRPE
66	chr1:203832791-203832853:+	SNRPE
67	chr1:211836115-211836994:-	NEK2
68	chr1:211842455-211842674:-	NEK2
69	chr1:211848726-211848967:-	NEK2
70	chr1:214817900-214820743:+	CENPF
71	chr1:214822018-214822170:+	CENPF
72	chr1:214825053-214825229:+	CENPF
73	chr1:214826171-214826332:+	CENPF
74	chr1:214828584-214828746:+	CENPF
75	chr1:214830276-214830752:+	CENPF
76	chr1:214832193-214832371:+	CENPF
77	chr1:214836934-214837912:+	CENPF
78	chr1:225152181-225152311:+	DNAH14
79	chr1:242048616-242048809:+	EXO1
80	chr1:245927343-245927451:-	SMYD3
81	chr1:247162650-247162742:-	ZNF695,ZNF670-ZNF695
82	chr1:247163214-247163376:-	ZNF695,ZNF670-ZNF695
83	chr1:247171208-247171355:-	ZNF695
84	chr2:10729169-10729416:-	NOL10
85	chr2:27014999-27015108:+	CENPA
86	chr2:70516482-70516504:-	SNRPG
87	chr2:111398884-111399041:-	BUB1
88	chr2:113495444-113496625:-	CKAP2L
89	chr2:169727402-169728065:-	SPC25
90	chr2:169730095-169730193:-	SPC25

Figure 54A

Order	Exon	Gene
91	chr2:169732582-169732686:-	SPC25
92	chr2:169733748-169733894:-	SPC25
93	chr2:169745736-169745801:-	SPC25
94	chr2:169745897-169746043:-	SPC25
95	chr2:169746828-169746944:-	SPC25
96	chr2:198351310-198351922:-	HSPD1
97	chr2:198352582-198352760:-	HSPD1
98	chr2:198353041-198353215:-	HSPD1
99	chr2:198353726-198353971:-	HSPD1
100	chr2:198364721-198365163:+	HSPE1-MOB4,HSPE1
101	chr2:198367763-198367852:+	HSPE1
102	chr2:201440034-201440184:+	SGOL2
103	chr2:203748332-203748464:-	WDR12
104	chr2:234745487-234746298:-	HJURP
105	chr2:234749255-234750851:-	HJURP
106	chr2:234756043-234756107:-	HJURP
107	chr2:234763030-234763212:-	HJURP
108	chr2:242625184-242625292:-	DTYMK
109	chr3:48198668-48200575:-	CDC25A
110	chr3:48673902-48675665:-	CELSR3
111	chr3:48680987-48681363:-	CELSR3
112	chr3:48685274-48685429:-	CELSR3
113	chr3:52109250-52110001:-	POC1A
114	chr3:52130585-52130728:-	POC1A
115	chr3:52156395-52156493:-	POC1A
116	chr3:52159129-52159197:-	POC1A
117	chr3:52172185-52172318:-	POC1A
118	chr3:52179862-52179977:-	POC1A
119	chr3:52183226-52183405:-	POC1A
120	chr3:122078437-122078766:-	CCDC58
121	chr3:122081815-122081874:-	CCDC58
122	chr3:122087023-122087169:-	CCDC58
123	chr3:122090522-122090647:-	CCDC58
124	chr3:124462762-124464038:+	UMPS
125	chr3:127816143-127816341:-	RUVBL1
126	chr3:139077586-139077654:-	COPB2
127	chr3:155652705-155652835:+	GMPS
128	chr3:155654127-155654299:+	GMPS
129	chr3:172537938-172539263:+	ECT2
130	chr3:183960117-183960464:-	ALG3
131	chr3:183960601-183960745:-	ALG3
132	chr3:183961342-183961418:-	ALG3
133	chr3:183961579-183961784:-	ALG3
134	chr3:183962389-183962509:-	ALG3
135	chr3:183962566-183963146:-	ALG3

Order	Exon	Gene
136	chr3:183963256-183963403:-	ALG3
137	chr3:183967445-183967678:+	ECE2
138	chr3:183975261-183975544:+	ECE2
139	chr3:183976076-183976546:+	ECE2
140	chr3:184082754-184082837:+	POLR2H
141	chr3:184082921-184083014:+	POLR2H
142	chr3:184084505-184084588:+	POLR2H
143	chr3:184085965-184086363:+	POLR2H
144	chr3:186301654-186301765:+	DNAJB11
145	chr3:186302219-186302378:+	DNAJB11
146	chr3:196311004-196315928:+	FBXO45
147	chr4:1976363-1976486:+	SCARNA22
148	chr4:104035612-104035709:-	CENPE
149	chr4:104037734-104037846:-	CENPE
150	chr4:104053850-104054053:-	CENPE
151	chr4:122739199-122739332:-	CCNA2
152	chr4:154318371-154318485:+	MND1
153	chr5:896780-896909:+	TRIP13
154	chr5:901456-901546:+	TRIP13
155	chr5:904263-904335:+	TRIP13
156	chr5:907245-907308:+	TRIP13
157	chr5:908103-908189:+	TRIP13
158	chr5:908470-909047:+	TRIP13
159	chr5:911958-912111:+	TRIP13
160	chr5:914580-914692:+	TRIP13
161	chr5:916019-916088:+	TRIP13
162	chr5:917123-918158:+	TRIP13
163	chr5:1320731-1320846:-	CLPTM1L
164	chr5:1798500-1799061:-	MRPL36
165	chr5:10260904-10261023:+	CCT5
166	chr5:10261672-10262312:+	CCT5
167	chr5:10263246-10263426:+	CCT5
168	chr5:10264768-10266500:+	CCT5
169	chr5:34925331-34925785:+	BRIX1
170	chr5:59892740-59893741:-	DEPDC1B
171	chr5:59894902-59895087:-	DEPDC1B
172	chr5:59899218-59899394:-	DEPDC1B
173	chr5:68467097-68467279:+	CCNB1
174	chr5:68470078-68470236:+	CCNB1
175	chr5:68470704-68470940:+	CCNB1
176	chr5:68471224-68471364:+	CCNB1
177	chr5:68473066-68473183:+	CCNB1
178	chr5:68473351-68474066:+	CCNB1
179	chr5:137515349-137515534:+	KIF20A
180	chr5:137517298-137517417:+	KIF20A

Figure 54B

Order	Exon	Gene
181	chr5:137518012-137518150:+	KIF20A
182	chr5:137518275-137518462:+	KIF20A
183	chr5:137518550-137518679:+	KIF20A
184	chr5:137518858-137519052:+	KIF20A
185	chr5:137519159-137519270:+	KIF20A
186	chr5:137519377-137519445:+	KIF20A
187	chr5:137519651-137519794:+	KIF20A
188	chr5:137519928-137520093:+	KIF20A
189	chr5:137520201-137520365:+	KIF20A
190	chr5:137520496-137520635:+	KIF20A
191	chr5:137521201-137521399:+	KIF20A
192	chr5:137521650-137521741:+	KIF20A
193	chr5:137521983-137522120:+	KIF20A
194	chr5:137522785-137523403:+	KIF20A
195	chr5:137620959-137621530:-	CDC25C
196	chr5:137621711-137621822:-	CDC25C
197	chr5:137622186-137622319:-	CDC25C
198	chr5:137622858-137622956:-	CDC25C
199	chr5:137625190-137625252:-	CDC25C
200	chr5:137626328-137626429:-	CDC25C
201	chr5:137627659-137627805:-	CDC25C
202	chr5:137666676-137666907:-	CDC25C
203	chr5:137667284-137667516:-	CDC25C
204	chr5:159849215-159849410:+	PTTG1
205	chr5:159849716-159849900:+	PTTG1
206	chr5:159851244-159851337:+	PTTG1
207	chr5:159854722-159854880:+	PTTG1
208	chr5:162898191-162898277:+	HMMR
209	chr5:162918118-162918949:+	HMMR
210	chr5:170833401-170833729:+	NPM1
211	chr5:176022803-176026878:-	GPRIN1
212	chr6:10414300-10415284:+	LOC100130275
213	chr6:33371506-33371906:+	KIFC1
214	chr6:33374369-33374442:+	KIFC1
215	chr6:33374577-33374652:+	KIFC1
216	chr6:33377423-33377697:+	KIFC1
217	chr6:34393573-34393792:-	RPS10
218	chr6:41899134-41899294:+	BYSL
219	chr6:80746192-80746316:+	TTK
220	chr6:80749413-80749589:+	TTK
221	chr6:80749913-80749997:+	TTK
222	chr6:80750323-80750420:+	TTK
223	chr6:80751836-80752237:+	TTK
224	chr7:1606970-1607486:-	PSMG3
225	chr7:1608760-1609629:-	PSMG3

Order	Exon	Gene
226	chr7:5528809-5529843:-	
227	chr7:6061880-6064432:-	EIF2AK1
228	chr7:44154287-44154544:-	POLD2
229	chr7:44154894-44154995:-	POLD2
230	chr7:44155365-44155492:-	POLD2
231	chr7:44155714-44155871:-	POLD2
232	chr7:44156029-44156109:-	POLD2
233	chr7:54823472-54823574:-	SEC61G
234	chr7:54825188-54825287:-	SEC61G
235	chr7:56020872-56021011:+	MRPS17
236	chr7:56022602-56023031:+	MRPS17
237	chr7:72983279-72985302:-	TBL2
238	chr7:76057955-76058950:+	ZP3
239	chr7:76062787-76062964:+	ZP3
240	chr7:96324110-96324203:-	SHFM1
241	chr7:96339000-96339203:-	SHFM1
242	chr7:97923386-97923423:-	BAIAP2L1
243	chr7:97933508-97933688:-	BAIAP2L1
244	chr7:97935751-97935828:-	BAIAP2L1
245	chr7:97937001-97937208:-	BAIAP2L1
246	chr7:97939757-97939904:-	BAIAP2L1
247	chr7:100304444-100305122:+	POP7
248	chr7:100797686-100797798:+	AP1S1
249	chr7:100802340-100802477:+	AP1S1
250	chr7:100803800-100804556:+	AP1S1
251	chr7:158439152-158439255:-	NCAPG2
252	chr7:158443524-158443664:-	NCAPG2
253	chr8:74872000-74872053:-	TCEB1
254	chr8:104442840-104442917:+	DCAF13
255	chr8:104453683-104453846:+	DCAF13
256	chr8:144679049-144679310:-	EEF1D
257	chr8:144686083-144688052:-	PYCRL
258	chr8:144689123-144689302:-	PYCRL
259	chr8:144690232-144690296:-	PYCRL
260	chr8:144806104-144807177:-	FAM83H
261	chr8:144807180-144807326:-	FAM83H
262	chr8:144826030-144828507:+	LOC100128338
263	chr8:145267897-145268366:+	HEATR7A
264	chr8:145582844-145583083:+	SLC52A2
265	chr8:145654165-145654719:-	TONSL
266	chr8:145655787-145655920:-	TONSL
267	chr8:145656461-145656534:-	TONSL
268	chr8:145660400-145660553:-	TONSL
269	chr8:145660551-145660823:+	
270	chr8:145661041-145661801:-	TONSL

Figure 54C

Order	Exon	Gene
271	chr8:145662134-145662378:+	
272	chr8:145662140-145662303:-	TONSL
273	chr8:145665734-145665859:-	TONSL
274	chr8:145666101-145666253:-	TONSL
275	chr8:145666349-145666494:-	TONSL
276	chr9:36633099-36633197:+	MELK
277	chr9:36642994-36643080:+	MELK
278	chr9:36651743-36651874:+	MELK
279	chr9:36657238-36657360:+	MELK
280	chr9:36665347-36665578:+	MELK
281	chr9:36669307-36669403:+	MELK
282	chr9:36670995-36671163:+	MELK
283	chr9:36674831-36674934:+	MELK
284	chr9:36677157-36677678:+	MELK
285	chr9:91926113-91926266:+	CKS2
286	chr9:140483111-140483170:-	ZMYND19
287	chr9:140484665-140484937:-	ZMYND19
288	chr10:58117199-58118220:-	ZWINT
289	chr10:58118317-58118485:-	ZWINT
290	chr10:58118566-58118708:-	ZWINT
291	chr10:62538220-62538426:+	CDK1
292	chr10:62553635-62554604:+	CDK1
293	chr10:94410158-94410274:+	KIF11
294	chr10:95287707-95288847:+	CEP55
295	chr10:129899522-129899965:-	MKI67
296	chr11:6701616-6703633:-	MRPL17
297	chr11:6703978-6704046:-	MRPL17
298	chr11:62623518-62623853:+	SLC3A2
299	chr11:64844927-64846659:-	CDCA5
300	chr11:68517658-68518167:-	MTL5
301	chr11:77811988-77812241:-	ALG8
302	chr12:2966850-2968829:-	FOXM1
303	chr12:2973486-2973661:-	FOXM1
304	chr12:2973849-2973921:-	FOXM1
305	chr12:2974521-2974565:-	FOXM1
306	chr12:2975559-2975687:-	FOXM1
307	chr12:2977729-2977920:-	FOXM1
308	chr12:2981262-2981413:-	FOXM1
309	chr12:2983143-2983691:-	FOXM1
310	chr12:6957972-6958362:-	CDCA3
311	chr12:6958487-6958593:-	CDCA3
312	chr12:6958729-6959022:-	CDCA3
313	chr12:6959631-6959760:-	CDCA3
314	chr12:6959997-6960173:-	CDCA3
315	chr12:49716971-49717132:+	TROAP

Order	Exon	Gene
316	chr12:49717387-49717535:+	TROAP
317	chr12:49717628-49717820:+	TROAP
318	chr12:49717938-49718169:+	TROAP
319	chr12:49719530-49719667:+	TROAP
320	chr12:49719859-49719941:+	TROAP
321	chr12:49720992-49721113:+	TROAP
322	chr12:49722710-49722838:+	TROAP
323	chr12:49723928-49725190:+	TROAP
324	chr12:50382945-50384126:-	RACGAP1
325	chr12:56504966-56505093:+	PA2G4
326	chr12:56704742-56704838:-	CNPY2
327	chr12:56704995-56705198:-	CNPY2
328	chr12:112847215-112847443:-	RPL6
329	chr12:122690865-122692083:+	B3GNT4
330	chr13:21727735-21729289:-	SKA3
331	chr13:21729832-21729950:-	SKA3
332	chr13:21732061-21732264:-	SKA3
333	chr13:21735929-21736014:-	SKA3
334	chr13:21742127-21742538:-	SKA3
335	chr13:21746478-21746643:-	SKA3
336	chr13:100634319-100635393:+	ZIC2
337	chr13:100637200-100637363:+	ZIC2
338	chr13:100637577-100639018:+	ZIC2
339	chr14:50116903-50117159:-	POLE2
340	chr14:54875463-54875507:+	CDKN3
341	chr14:54878202-54878424:+	CDKN3
342	chr14:54882617-54882648:+	CDKN3
343	chr14:54884566-54884669:+	CDKN3
344	chr14:54886710-54886932:+	CDKN3
345	chr14:55614836-55615191:-	DLGAP5
346	chr14:55617517-55617566:-	DLGAP5
347	chr14:55618413-55618659:-	DLGAP5
348	chr14:55619308-55619365:-	DLGAP5
349	chr14:55621335-55621524:-	DLGAP5
350	chr14:55625240-55625459:-	DLGAP5
351	chr14:55629689-55629793:-	DLGAP5
352	chr14:55636117-55636277:-	DLGAP5
353	chr14:55637420-55637505:-	DLGAP5
354	chr14:55642064-55642190:-	DLGAP5
355	chr14:55642612-55642736:-	DLGAP5
356	chr15:40477366-40477580:+	BUB1B
357	chr15:40477752-40477843:+	BUB1B
358	chr15:40488746-40488975:+	BUB1B
359	chr15:40685394-40686488:+	C15orf23
360	chr15:41021703-41021832:+	RAD51

Figure 54D

Order	Exon	Gene
361	chr15:41022051-41022172:+	RAD51
362	chr15:41023253-41024354:+	RAD51
363	chr15:41601468-41602047:-	OIP5
364	chr15:44058926-44059108:+	PDIA3
365	chr15:44060687-44060795:+	PDIA3
366	chr15:44062448-44062527:+	PDIA3
367	chr15:44062721-44062778:+	PDIA3
368	chr15:44063303-44064803:+	PDIA3
369	chr15:59397320-59397491:+	CCNB2
370	chr15:59399521-59399649:+	CCNB2
371	chr15:59406917-59407075:+	CCNB2
372	chr15:59409427-59409567:+	CCNB2
373	chr15:59415716-59415826:+	CCNB2
374	chr15:64657213-64658274:-	KIAA0101
375	chr15:64668942-64669104:-	KIAA0101
376	chr15:64673157-64673237:-	KIAA0101
377	chr15:89858513-89858620:+	FANCI
378	chr15:91509274-91510432:-	PRC1
379	chr15:91512677-91512853:-	PRC1
380	chr15:91513634-91513744:-	PRC1
381	chr15:91517366-91517476:-	PRC1
382	chr15:91517815-91517961:-	PRC1
383	chr15:91519906-91520001:-	PRC1
384	chr15:91522388-91522524:-	PRC1
385	chr15:91523472-91523619:-	PRC1
386	chr15:91537637-91537804:-	PRC1
387	chr16:101558-101645:-	POLR3K
388	chr16:103476-103625:-	POLR3K
389	chr16:2510817-2511225:+	C16orf59
390	chr16:3022794-3023065:-	PKMYT1
391	chr16:3024551-3024657:-	PKMYT1
392	chr16:3026665-3027032:-	PKMYT1
393	chr16:3029649-3029937:-	PKMYT1
394	chr16:3030287-3030474:-	PKMYT1
395	chr16:8941581-8943512:+	PMM2
396	chr16:23690201-23690661:+	PLK1
397	chr16:23691405-23691573:+	PLK1
398	chr16:23692235-23692379:+	PLK1
399	chr16:23693385-23693478:+	PLK1
400	chr16:23695191-23695410:+	PLK1
401	chr16:23698790-23698945:+	PLK1
402	chr16:23699989-23700066:+	PLK1
403	chr16:23700559-23700713:+	PLK1
404	chr16:23700815-23700997:+	PLK1
405	chr16:23701181-23701688:+	PLK1

Order	Exon	Gene
406	chr16:53734551-53734642:-	RPGRIP1L
407	chr16:81045535-81045715:+	CENPN
408	chr16:81047741-81047786:+	CENPN
409	chr16:81050931-81050990:+	CENPN
410	chr16:81051363-81051439:+	CENPN
411	chr16:81053705-81053881:+	CENPN
412	chr16:81056360-81056486:+	CENPN
413	chr16:88874523-88875665:+	CDT1
414	chr17:6348357-6348724:+	FAM64A
415	chr17:26905391-26905546:-	SPAG5
416	chr17:26905690-26905770:-	SPAG5
417	chr17:26910871-26911004:-	SPAG5
418	chr17:26911168-26911270:-	SPAG5
419	chr17:26911351-26911498:-	SPAG5
420	chr17:26911670-26911763:-	SPAG5
421	chr17:26912337-26912429:-	SPAG5
422	chr17:26912882-26913016:-	SPAG5
423	chr17:26913108-26913200:-	SPAG5
424	chr17:34858937-34859040:-	MYO19
425	chr17:34869472-34869568:-	MYO19
426	chr17:38152436-38152591:+	PSMD3
427	chr17:38153583-38153633:+	PSMD3
428	chr17:38457080-38457282:+	CDC6
429	chr17:38457720-38457860:+	CDC6
430	chr17:38544798-38545899:-	TOP2A
431	chr17:38546217-38546416:-	TOP2A
432	chr17:38547758-38547892:-	TOP2A
433	chr17:38552535-38552717:-	TOP2A
434	chr17:38554810-38554893:-	TOP2A
435	chr17:49230920-49231023:+	NME1-NME2,NME1
436	chr17:76181136-76181246:-	TK1
437	chr17:76210277-76210508:+	BIRC5
438	chr17:76210321-76210508:-	
439	chr17:76210761-76210870:-	
440	chr17:76210761-76211064:+	BIRC5
441	chr17:76212745-76212862:+	BIRC5
442	chr17:76212745-76212862:-	
443	chr17:76219545-76220783:-	
444	chr17:76219546-76221715:+	BIRC5
445	chr18:2578926-2579028:+	NDC80
446	chr18:19192260-19192404:+	SNRPD1
447	chr18:19202686-19202762:+	SNRPD1
448	chr19:11257831-11257965:-	SPC24
449	chr19:11258494-11258570:-	SPC24
450	chr19:11258689-11258793:-	SPC24

Figure 54E

Order	Exon	Gene
451	chr19:11259770-11260027:-	SPC24
452	chr19:12921131-12921218:+	RNASEH2A
453	chr19:18423414-18423512:-	LSM4
454	chr19:19035042-19035117:+	DDX49
455	chr19:33073101-33073138:+	PDCD5
456	chr19:33075856-33075917:+	PDCD5
457	chr19:34900066-34900415:+	PDCD2L
458	chr19:34912424-34912572:+	PDCD2L
459	chr19:34916895-34917070:+	PDCD2L
460	chr19:36581347-36581422:+	WDR62
461	chr19:39422973-39423657:+	MRPS12
462	chr19:45395650-45395717:+	TOMM40
463	chr19:45396094-45396186:+	TOMM40
464	chr19:45397037-45397138:+	TOMM40
465	chr19:45397218-45397323:+	TOMM40
466	chr19:45403987-45404109:+	TOMM40
467	chr19:46192491-46192605:-	SNRPD2
468	chr19:55912650-55913130:-	UBE2S
469	chr19:55915656-55915846:-	UBE2S
470	chr19:55918183-55918330:-	UBE2S
471	chr19:55918955-55919325:-	UBE2S
472	chr20:25206135-25207360:+	ENTPD6
473	chr20:30326904-30327424:+	TPX2
474	chr20:30345210-30345385:+	TPX2
475	chr20:30347860-30347982:+	TPX2
476	chr20:30354359-30354485:+	TPX2
477	chr20:30358146-30358274:+	TPX2
478	chr20:30359363-30359485:+	TPX2
479	chr20:30363670-30363791:+	TPX2
480	chr20:30365290-30365441:+	TPX2
481	chr20:30366616-30366787:+	TPX2
482	chr20:30370052-30370193:+	TPX2
483	chr20:30371508-30371724:+	TPX2
484	chr20:30380538-30380633:+	TPX2
485	chr20:30381651-30381827:+	TPX2
486	chr20:30382205-30382351:+	TPX2
487	chr20:30385207-30385318:+	TPX2
488	chr20:30386168-30386355:+	TPX2
489	chr20:30388773-30389603:+	TPX2
490	chr20:30753098-30755061:+	TM9SF4
491	chr20:32263293-32264785:-	E2F1
492	chr20:32676117-32677711:-	EIF2S2
493	chr20:44441255-44442103:+	UBE2C
494	chr20:44443023-44443109:+	UBE2C
495	chr20:44444029-44444384:+	UBE2C

Order	Exon	Gene
496	chr20:44444493-44444552:+	UBE2C
497	chr20:44445348-44445594:+	UBE2C
498	chr20:54944445-54945396:-	AURKA
499	chr20:54945541-54945715:-	AURKA
500	chr20:54948464-54948612:-	AURKA
501	chr22:19471385-19471528:+	CDC45
502	chr22:19486624-19486674:+	CDC45
503	chr22:19492885-19493004:+	CDC45
504	chr22:19494909-19495040:+	CDC45
505	chr22:19495289-19495387:+	CDC45
506	chr22:19496053-19496214:+	CDC45
507	chr22:19502272-19502571:+	CDC45
508	chr22:19504050-19504168:+	CDC45
509	chr22:19504337-19504416:+	CDC45
510	chr22:19506367-19506432:+	CDC45
511	chr22:19508004-19508134:+	CDC45
512	chr22:41652025-41652109:-	RANGAP1
513	chr22:41652195-41652294:-	RANGAP1
514	chr22:41676937-41678518:-	RANGAP1
515	chr22:50927469-50927578:+	MIOX
516	chrX:69521750-69521916:+	KIF4A
517	chrX:69572450-69572506:+	KIF4A
518	chrX:69573472-69573657:+	KIF4A
519	chrX:69594001-69594104:+	KIF4A
520	chrX:69595054-69595198:+	KIF4A
521	chrX:69595950-69596068:+	KIF4A
522	chrX:69606468-69606551:+	KIF4A
523	chrX:69607034-69607147:+	KIF4A
524	chrX:69615521-69615676:+	KIF4A
525	chrX:69615799-69615899:+	KIF4A
526	chrX:69622416-69622545:+	KIF4A
527	chrX:69623714-69623884:+	KIF4A
528	chrX:69624565-69624672:+	KIF4A
529	chrX:69625678-69625755:+	KIF4A
530	chrX:69626109-69626204:+	KIF4A
531	chrX:69626743-69626925:+	KIF4A
532	chrX:69637738-69637854:+	KIF4A
533	chrX:69639511-69639633:+	KIF4A
534	chrX:69639912-69640735:+	KIF4A
535	chrX:119744866-119744933:+	MCTS1
536	chrX:150151893-150152064:+	
537	chrX:48760889-48762759:-	

Figure 54F

Solid Tumor mRNA protein coding markers

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
1	ACLY	17q21.2	ATP citrate lyase	P53396
2	ACTL6A	3q26.33	actin-like 6A	O96019
3	ACYP1	14q24.3	acylphosphatase 1, erythrocyte (common) type	P07311
4	ANLN	7p15-p14	anillin, actin binding protein	Q9NQW6
5	AP1S1	7q22.1	adaptor-related protein complex 1, sigma 1 subunit	P61966
6	ARHGAP11A	15q13.3	Rho GTPase activating protein 11A	Q6P4F7
7	ASF1B	19p13.12	anti-silencing function 1B histone chaperone	Q9NVP2
8	ASPM	1q31	asp (abnormal spindle) homolog, microcephaly associated (Drosophila)	Q8IZT6
9	ATAD3B	1p36.33	ATPase family, AAA domain containing 3B	Q5T9A4
10	ATXN2L	16p11	ataxin 2-like	Q8WWM7
11	AURKA	20q13	aurora kinase A	O14965
12	AURKB	17p13.1	aurora kinase B	AIM1
13	AZI1	17q25.3	5-azacytidine induced 1	Q9UPN4
14	B3GNT4	12q24	UDP-GlcNAc:betaGal beta-1,3-N-acetylglucosaminyltransferase 4	Q9C0J1
15	BIRC5	17q25.3	baculoviral IAP repeat containing 5	O15392
16	BLM	15q26.1	Bloom syndrome, RecQ helicase-like	P54132
17	BOLA2	16p11.2	bolA family member 2	BOLA2B
18	BOP1	8q24.3	block of proliferation 1	Q14137
19	BRIX1	5p13.2	BRX1, biogenesis of ribosomes, homolog (S. cerevisiae)	Q8TDN6
20	BUB1	2q13	BUB1 mitotic checkpoint serine/threonine kinase	O43683
21	BUB1B	15q15	BUB1 mitotic checkpoint serine/threonine kinase B	O60566
22	BUB3	10q24	BUB3 mitotic checkpoint protein	O43684
23	BYSL	6p21.1	bystin-like	Q13895
24	C15orf42/TICRR	15q26.1	TOPBP1 interacting checkpoint and replication regulator	Q7Z2Z1
25	C16orf59	16p13.3	chromosome 16 open reading frame 59	Q7L2K0
26	C16orf88/KNOP1	16p12.3	lysine rich nucleolar protein 1	Q1ED39
27	C19orf40	19q13.11	chromosome 19 open reading frame 40	Q9BTP7
28	C19orf48	19q13.33	chromosome 19 open reading frame 48	Q6RUI8
29	C1orf159	1p36.33	chromosome 1 open reading frame 159	Q96HA4
30	C1orf35	1q42.13	chromosome 1 open reading frame 35	Q9BU76
31	C20orf20/MRGBP	20q13.33	MRG domain binding protein	Q9NV56
32	C4orf21	4q25	chromosome 4 open reading frame 21	Q86YA3
33	C4orf46	4q32.1	chromosome 4 open reading frame 46	Q504U0
34	CABLES2	20q13.33	Cdk5 and Abl enzyme substrate 2	Q9BTV7
35	CALR	19p13.3-p13.2	calreticulin	P27797
36	CBX2	17q25.3	chromobox homolog 2	Q14781
37	CBX3	7p15.2	chromobox homolog 3	Q13185
38	CBX8	17q25.3	chromobox homolog 8	Q9HC52
39	CCDC77	12p13.33	coiled-coil domain containing 77	Q9BR77
40	CCDC86	11q12.2	coiled-coil domain containing 86	Q9H6F5
41	CCDC99/SPDL1	5q35.1	spindle apparatus coiled-coil protein 1	Q96EA4
42	CCNA2	4q27	cyclin A2	P20248
43	CCNB1	5q12	cyclin B1	P14635
44	CCNB2	15q21.3	cyclin B2	O95067
45	CCNE1	19q12	cyclin E1	P24864

Figure 55A

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
46	CCNE2	8q22.1	cyclin E2	O96020
47	CCT2	12q14	chaperonin containing TCP1, subunit 2 (beta)	P78371
48	CCT3	1q23	chaperonin containing TCP1, subunit 3 (gamma)	P49368
49	CCT5	5p15.2	chaperonin containing TCP1, subunit 5 (epsilon)	P48643
50	CCT6A	7p11.2	chaperonin containing TCP1, subunit 6A (zeta 1)	P40227
51	CD3EAP	19q13.3	CD3e molecule, epsilon associated protein	CAST
52	CDC20	1p34.1	cell division cycle 20	Q12834
53	CDC25A	3p21	cell division cycle 25A	P30304
54	CDC25C	5q31	cell division cycle 25C	P30307
55	CDC45	22q11.21	cell division cycle 45	O75419
56	CDC6	17q21.3	cell division cycle 6	Q99741
57	CDCA2	8p21.2	cell division cycle associated 2	Q69YH5
58	CDCA3	12p13.31	cell division cycle associated 3	Q99618
59	CDCA5	11q13.1	cell division cycle associated 5	Q96FF9
60	CDCA8	1p34.3	cell division cycle associated 8	Q53HL2
61	CDH24	14q11.2	cadherin 24, type 2	Q86UP0
62	CDK1	10q21.2	cyclin-dependent kinase 1	P06493
63	CDK4	12q13	cyclin-dependent kinase 4	P11802
64	CDKN3	14q22	cyclin-dependent kinase inhibitor 3	CDK3
65	CDT1	16q24.3	chromatin licensing and DNA replication factor 1	Q9H211
66	CENPA	2p23.3	centromere protein A	P49450
67	CENPE	4q24-q25	centromere protein E, 312kDa	Q02224
68	CENPF	1q41	centromere protein F, 350/400kDa	P49454
69	CENPH	5p15.2	centromere protein H	Q9H3R5
70	CENPI	Xq22.1	centromere protein I	Q92674
71	CENPK	5q12.3	centromere protein K	Q9BS16
72	CENPL	1q25.1	centromere protein L	Q8N0S6
73	CENPO	2p23.3	centromere protein O	Q9BU64
74	CEP55	10q24.1	centrosomal protein 55kDa	Q53EZ4
75	CEP72	5p15.33	centrosomal protein 72kDa	Q9P209
76	CHAF1A	19p13.3	chromatin assembly factor 1, subunit A (p150)	Q13111
77	CHAF1B	21q22.2	chromatin assembly factor 1, subunit B (p60)	MPP7
78	CHRNA5	15q24	cholinergic receptor, nicotinic, alpha 5 (neuronal)	P30532
79	CHTF18	16p13.3	CTF18, chromosome transmission fidelity factor 18 homolog (S. cerevisiae)	Q8WVB6
80	CKAP2L	2q13	cytoskeleton associated protein 2-like	Q8IYA6
81	CKS2	9q22	CDC28 protein kinase regulatory subunit 2	P33552
82	CLSPN	1p34.3	claspin	Q9HAW4
83	CPNE7	16q24.3	copine VII	Q9UBL6
84	CPSF3	2p25.1	cleavage and polyadenylation specific factor 3, 73kDa	Q9UKF6
85	CTHRC1	8q22.3	collagen triple helix repeat containing 1	Q96CG8
86	CYP27B1	12q13.3-q14	cytochrome P450, family 27, subfamily B, polypeptide 1	O15528
87	DAP3	1q22	death associated protein 3	P51398
88	DARS2	1q25.1	aspartyl-tRNA synthetase 2, mitochondrial	Q6PI48
89	DAZAP1	19p13.3	DAZ associated protein 1	Q96EP5
90	DBF4	7q21.3	DBF4 homolog (S. cerevisiae)	Q9UBU7

Figure 55B

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
91	DCAF13	8q22.3	DDB1 and CUL4 associated factor 13	Q9NV06
92	DDX27	20q13.13	DEAD (Asp-Glu-Ala-Asp) box polypeptide 27	Q96GQ7
93	DDX31	9q34.2	DEAD (Asp-Glu-Ala-Asp) box polypeptide 31	Q9H8H2
94	DDX55	12q24.31	DEAD (Asp-Glu-Ala-Asp) box polypeptide 55	Q8NHQ9
95	DDX56	7p13	DEAD (Asp-Glu-Ala-Asp) box helicase 56	
96	DENR	12q24.31	density-regulated protein	O43583
97	DEPDC1B	5q12	DEP domain containing 1B	Q8WUY9
98	DHX34	19q13.3	DEAH (Asp-Glu-Ala-His) box polypeptide 34	Q14147
99	DHX37	12q24.31	DEAH (Asp-Glu-Ala-His) box polypeptide 37	Q8IY37
100	DKC1	Xq28	dyskeratosis congenita 1, dyskerin	O60832
101	DLGAP5	14q22.3	discs, large (Drosophila) homolog-associated protein 5	Q15398
102	DMBX1	1p34.1	diencephalon/mesencephalon homeobox 1	Q8NFW5
103	DNA2	10q21.3-q22.1	DNA replication helicase/nuclease 2	P51530
104	DNAJB11	3q27	DnaJ (Hsp40) homolog, subfamily B, member 11	Q9UBS4
105	DNAJC2	7q22-q32	DnaJ (Hsp40) homolog, subfamily C, member 2	Q99543
106	DNAJC9	10q22.3	DnaJ (Hsp40) homolog, subfamily C, member 9	Q8WXX5
107	DNMT3B	20q11.2	DNA (cytosine-5-)-methyltransferase 3 beta	Q9UBC3
108	DPF1	19q13.12	D4, zinc and double PHD fingers family 1	Q92782
109	DPH2	1p34	DPH2 homolog (S. cerevisiae)	Q9BQC3
110	DSCC1	8q24.12	DNA replication and sister chromatid cohesion 1	Q9BVC3
111	DTL	1q32	denticleless E3 ubiquitin protein ligase homolog (Drosophila)	Q9NZJ0
112	DTYMK	2q37	deoxythymidylate kinase (thymidylate kinase)	P23919
113	E2F1	20q11	E2F transcription factor 1	Q01094
114	E2F3	6p22	E2F transcription factor 3	O00716
115	EBNA1BP2	1p35-p33	EBNA1 binding protein 2	Q99848
116	ECT2	3q26.1-q26.2	epithelial cell transforming sequence 2 oncogene	Q9H8V3
117	EIF2S2	20q11.2	eukaryotic translation initiation factor 2, subunit 2 beta, 38kDa	P20042
118	EIF3B	7p22	eukaryotic translation initiation factor 3, subunit B	P55884
119	ELAVL1	19p13.2	ELAV like RNA binding protein 1	Q15717
120	EME1	17q21.33	essential meiotic structure-specific endonuclease 1	Q96AY2
121	ERCC6L	Xq13.1	excision repair cross-complementing rodent repair deficiency, complementation group 6-like	Q2NKX8
122	ESCO2	8p21.1	establishment of sister chromatid cohesion N-acetyltransferase 2	Q56NI9
123	EXO1	1q42-q43	exonuclease 1	Q9UQ84
124	EZH2	7q35-q36	enhancer of zeste homolog 2 (Drosophila)	Q15910
125	FAM111B	11q12.1	family with sequence similarity 111, member B	Q6SJ93
126	FAM54A/MTFR2	6q23.3	mitochondrial fission regulator 2	Q6P444
127	FAM64A	17p13.2	family with sequence similarity 64, member A	Q9BSJ6
128	FAM72A	1q32.1	family with sequence similarity 72, member A	Q5TYM5
129	FAM72B	1p12	family with sequence similarity 72, member B	Q86X60
130	FAM72D	1q21.1	family with sequence similarity 72, member D	Q6L9T8
131	FAM83H	8q24.3	family with sequence similarity 83, member H	Q6ZRV2
132	FANCA	16q24.3	Fanconi anemia, complementation group A	O15360
133	FANCD2	3p25.3	Fanconi anemia, complementation group D2	Q9BXW9
134	FANCG	9p13	Fanconi anemia, complementation group G	O15287
135	FANCI	15q26.1	Fanconi anemia, complementation group I	Q9NVI1

Figure 55C

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
136	FBXL19	16p11.2	F-box and leucine-rich repeat protein 19	Q6PCT2
137	FBXL6	8q24.3	F-box and leucine-rich repeat protein 6	Q8N531
138	FBXO43	8q22.3	F-box protein 43	Q4G163
139	FEN1	11q12	flap structure-specific endonuclease 1	P39748
140	FLAD1	1q22	flavin adenine dinucleotide synthetase 1	Q8NFF5
141	FOXM1	12p13	forkhead box M1	MPP2
142	GAD1	2q31	glutamate decarboxylase 1 (brain, 67kDa)	Q99259
143	GINS2	16q24.1	GINS complex subunit 2 (Psf2 homolog)	Q9Y248
144	GINS4	8p11.21	GINS complex subunit 4 (Sld5 homolog)	Q9BRT9
145	GPC2	7q22.1	glypican 2	Q8N158
146	GPR19	12p12.3	G protein-coupled receptor 19	Q15760
147	GPRIN1	5q35.2	G protein regulated inducer of neurite outgrowth 1	Q7Z2K8
148	GSG2	17p13	germ cell associated 2 (haspin)	Q8TF76
149	GTPBP3	19p13.2	GTP binding protein 3 (mitochondrial)	Q969Y2
150	GTPBP4	10p15-p14	GTP binding protein 4	Q9BZE4
151	GTSE1	22q13.2-q13.3	G-2 and S-phase expressed 1	Q9NYZ3
152	H2AFX	11q23.3	H2A histone family, member X	P16104
153	HAGHL	16p13.3	hydroxyacylglutathione hydrolase-like	Q6PII5
154	HAUS5	19q13.12	HAUS augmin-like complex, subunit 5	O94927
155	HDGF	1q21-q23	hepatoma-derived growth factor	P51858
156	HGS	17q25	hepatocyte growth factor-regulated tyrosine kinase substrate	O14964
157	HIST1H2BL	6p22.1	histone cluster 1, H2bl	Q99880
158	HIST1H2BO	6p22.1	histone cluster 1, H2bo	P23527
159	HIST1H3J	6p22.1	histone cluster 1, H3j	
160	HIST1H4I	6p22.1	histone cluster 1, H4i	
161	HIST2H2BF	1q21.2	histone cluster 2, H2bf	Q5QNW6
162	HIST2H3D	1q21.2	histone cluster 2, H3d	HIST2H3C
163	HJURP	2q37.1	Holliday junction recognition protein	Q8NCD3
164	HMMR	5q34	hyaluronan-mediated motility receptor (RHAMM)	O75330
165	HNRNPA2B1	7p15	heterogeneous nuclear ribonucleoprotein A2/B1	P22626
166	HOXC13	12q13.13	homeobox C13	P31276
167	HSF2BP	21q22.3	heat shock transcription factor 2 binding protein	O75031
168	IL11	19q13.3-q13.4	interleukin 11	P20809
169	ILF2	1q21.3	interleukin enhancer binding factor 2	Q12905
170	ILF3	19p13.2	interleukin enhancer binding factor 3, 90kDa	Q12906
171	INTS8	8q22.1	integrator complex subunit 8	Q75QN2
172	IQGAP3	1q21.3	IQ motif containing GTPase activating protein 3	Q86VI3
173	IRAK1	Xq28	interleukin-1 receptor-associated kinase 1	P51617
174	KCNK9	8q24.3	potassium channel, subfamily K, member 9	Q9NPC2
175	KIAA0101	15q22	KIAA0101	Q15004
176	KIF11	10q24.1	kinesin family member 11	P52732
177	KIF14	1q32.1	kinesin family member 14	Q15058
178	KIF15	3p21.31	kinesin family member 15	Q9NS87
179	KIF18B	17q21.31	kinesin family member 18B	Q86Y91
180	KIF20A	5q31	kinesin family member 20A	O95235

Figure 55D

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
181	KIF23	15q23	kinesin family member 23	Q02241
182	KIF2C	1p34.1	kinesin family member 2C	Q99661
183	KIF4A	Xq13.1	kinesin family member 4A	O95239
184	KIF4B	5q33.2	kinesin family member 4B	Q2VIQ3
185	KIFC1	6p21.32	kinesin family member C1	Q9BW19
186	KIFC2	8q24.3	kinesin family member C2	Q96AC6
187	KLHL17	1p36	kelch-like family member 17	Q6TDP4
188	KPNA2	17q24.2	karyopherin alpha 2 (RAG cohort 1, importin alpha 1)	P52292
189	KPNB1	17q21.32	karyopherin (importin) beta 1	Q14974
190	KRI1	19p13.2	KRI1 homolog (S. cerevisiae)	Q8N9T8
191	LMNB1	5q23.2	lamin B1	P20700
192	LMNB2	19p13.3	lamin B2	Q03252
193	MAD2L1	4q27	MAD2 mitotic arrest deficient-like 1 (yeast)	Q13257
194	MCM10	10p13	minichromosome maintenance complex component 10	Q7L590
195	MCM7	7q21.3-q22.1	minichromosome maintenance complex component 7	MCM2
196	MELK	9p13.1	maternal embryonic leucine zipper kinase	Q14680
197	METTL1	12q13	methyltransferase like 1	Q9UBP6
198	MEX3A	1q22	mex-3 RNA binding family member A	A1L020
199	MIF	22q11.23	macrophage migration inhibitory factor (glycosylation-inhibiting factor)	P14174
200	MKI67	10q26.2	marker of proliferation Ki-67	P46013
201	MLF1IP	4q35.1	MLF1 interacting protein	Q71F23
202	MMP10	11q22.3	matrix metallopeptidase 10 (stromelysin 2)	P09238
203	MMP9	20q12-q13	matrix metallopeptidase 9 (gelatinase B, 92kDa gelatinase, 92kDa type IV collagenase)	P14780
204	MND1	4q31.3	meiotic nuclear divisions 1 homolog (S. cerevisiae)	Q9BWT6
205	MORC2	22q12.2	MORC family CW-type zinc finger 2	Q9Y6X9
206	MRPL17	11p15.5-p15.4	mitochondrial ribosomal protein L17	Q9NRX2
207	MSTO1	1q22	misato 1, mitochondrial distribution and morphology regulator	Q9BUK6
208	MTL5	11q13.2-q13.3	metallothionein-like 5, testis-specific (tesmin)	Q9Y4I5
209	MYBL2	20q13.1	v-myb avian myeloblastosis viral oncogene homolog-like 2	P10244
210	MYO19	17q21.1	myosin XIX	Q96H55
211	NAA10	Xq28	N(alpha)-acetyltransferase 10, NatA catalytic subunit	P41227
212	NACC1	19p13.13	nucleus accumbens associated 1, BEN and BTB (POZ) domain containing	Q96RE7
213	NCAPG	4p15.32	non-SMC condensin I complex, subunit G	CAPG
214	NCAPG2	7q36.3	non-SMC condensin II complex, subunit G2	Q86XI2
215	NCAPH	2q11.2	non-SMC condensin I complex, subunit H	Q15003
216	NCBP2	3q29	nuclear cap binding protein subunit 2, 20kDa	P52298
217	NEIL3	4q34	nei endonuclease VIII-like 3 (E. coli)	Q8TAT5
218	NEK2	1q32-q42	NIMA-related kinase 2	P51955
219	NFE2L3	7p15.2	nuclear factor, erythroid 2-like 3	Q9Y4A8
220	NOL10	2p25.1	nucleolar protein 10	Q9BSC4
221	NOL11	17q24.2	nucleolar protein 11	Q9H8H0
222	NOP2	12p13	NOP2 nucleolar protein	P46087
223	NOP56	20p13	NOP56 ribonucleoprotein	O00567
224	NOP58	2q33.1	NOP58 ribonucleoprotein	Q9Y2X3
225	NR2C2AP	19p13.11	nuclear receptor 2C2-associated protein	Q86WQ0

Figure 55E

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
226	NSUN5	7q11.23	NOP2/Sun domain family, member 5	Q96P11
227	NUDT1	7p22	nudix (nucleoside diphosphate linked moiety X)-type motif 1	P36639
228	NUF2	1q23.3	NUF2, NDC80 kinetochore complex component	Q9BZD4
229	NUP37	12q23	nucleoporin 37kDa	Q8NFH4
230	NUP85	17q25	nucleoporin 85kDa	Q9BW27
231	NUP93	16q13	nucleoporin 93kDa	Q8N1F7
232	NUSAP1	15q14	nucleolar and spindle associated protein 1	Q9BXS6
233	OBFC2B/NABP2	12q13.3	nucleic acid binding protein 2	Q9BQ15
234	OIP5	15q14	Opa interacting protein 5	O43482
235	OR2B6	6p22.1	olfactory receptor, family 2, subfamily B, member 6	P58173
236	PAFAH1B3	19q13.1	platelet-activating factor acetylhydrolase 1b, catalytic subunit 3 (29kDa)	Q15102
237	PAQR4	16p13	progestin and adipoQ receptor family member IV	Q8N4S7
238	PASK	2q37.3	PAS domain containing serine/threonine kinase	Q96RG2
239	PDAP1	7q	PDGFA associated protein 1	Q13442
240	PKMYT1	16p13.3	protein kinase, membrane associated tyrosine/threonine 1	MYT1
241	PLK1	16p	polo-like kinase 1	P53350
242	PLXNA3	Xq28	plexin A3	P51805
243	PMS2	7p22.1	PMS2 postmeiotic segregation increased 2 (S. cerevisiae)	P54278
244	PODNL1	19p13.12	podocan-like 1	Q6PEZ8
245	POLD1	19q13.3	polymerase (DNA directed), delta 1, catalytic subunit	P28340
246	POLQ	3q13.3	polymerase (DNA directed), theta	POLH
247	POLR2H	3q28	polymerase (RNA) II (DNA directed) polypeptide H	P52434
248	POLR3K	16p13.3	polymerase (RNA) III (DNA directed) polypeptide K, 12.3 kDa	Q9Y2Y1
249	POP1	8q22.2	processing of precursor 1, ribonuclease P/MRP subunit (S. cerevisiae)	Q99575
250	PPEF1	Xp22	protein phosphatase, EF-hand calcium binding domain 1	O14829
251	PPIA	7p13	peptidylprolyl isomerase A (cyclophilin A)	P62937
252	PPIH	1p34.1	peptidylprolyl isomerase H (cyclophilin H)	O43447
253	PPM1G	2p23.3	protein phosphatase, Mg2+/Mn2+ dependent, 1G	O15355
254	PPP1R14B	11q13	protein phosphatase 1, regulatory (inhibitor) subunit 14B	Q96C90
255	PRAME	22q11.22	preferentially expressed antigen in melanoma	P78395
256	PRC1	15q26.1	protein regulator of cytokinesis 1	O43663
257	PRDM12	9q33-q34	PR domain containing 12	Q9H4Q4
258	PRPF19	11q12.2	pre-mRNA processing factor 19	Q9UMS4
259	PRR19	19q13.2	proline rich 19	A6NJB7
260	PSMG3	7p22.3	proteasome (prosome, macropain) assembly chaperone 3	Q9BT73
261	PTBP1	19p13.3	polypyrimidine tract binding protein 1	P26599
262	PTTG1	5q35.1	pituitary tumor-transforming 1	O95997
263	PUS7	7q22.3	pseudouridylate synthase 7 homolog (S. cerevisiae)	Q96PZ0
264	QSOX2	9q34.3	quiescin Q6 sulfhydryl oxidase 2	Q6ZRP7
265	R3HDM1	2q21.3	R3H domain containing 1	Q15032
266	RACGAP1	12q13	Rac GTPase activating protein 1	Q9H0H5
267	RAD51	15q15.1	RAD51 recombinase	Q06609
268	RAD51AP1	12p13.2-p13.1	RAD51 associated protein 1	Q96B01
269	RAD54L	1p32	RAD54-like (S. cerevisiae)	Q92698
270	RAN	12q24.33	RAN, member RAS oncogene family	P62826

Figure 55F

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
271	RANGAP1	22q13	Ran GTPase activating protein 1	P46060
272	RCC2	1p36.13	regulator of chromosome condensation 2	Q9P258
273	RDM1	17q11.2	RAD52 motif 1	Q8NG50
274	RELL2	5q31.3	RELT-like 2	Q8NC24
275	RFC2	7q11.23	replication factor C (activator 1) 2, 40kDa	P35250
276	RFC3	13q13.2	replication factor C (activator 1) 3, 38kDa	P40938
277	RFC4	3q27	replication factor C (activator 1) 4, 37kDa	P35249
278	RHEBL1	12q13.12	Ras homolog enriched in brain like 1	Q8TAI7
279	RNASEH2A	19p13.13	ribonuclease H2, subunit A	O75792
280	RNFT2	12q24.22	ring finger protein, transmembrane 2	Q96EX2
281	RPUSD1	16p13.3	RNA pseudouridylate synthase domain containing 1	Q9UJJ7
282	RRM2	2p25-p24	ribonucleotide reductase M2	P31350
283	RUVBL1	3q21	RuvB-like AAA ATPase 1	Q9Y265
284	SAAL1	11p15.1	serum amyloid A-like 1	Q96ER3
285	SASS6	1p21.3	spindle assembly 6 homolog (C. elegans)	Q6UVJ0
286	SGOL1	3p24.3	shugoshin-like 1 (S. pombe)	Q5FBB7
287	SGOL2	2q33.2	shugoshin-like 2 (S. pombe)	Q562F6
288	SHCBP1	16q11	SHC SH2-domain binding protein 1	Q8NEM2
289	SKA1	18q21.1	spindle and kinetochore associated complex subunit 1	Q96BD8
290	SKA3	13q11	spindle and kinetochore associated complex subunit 3	Q8IX90
291	SLC25A39	17q12	solute carrier family 25, member 39	Q9BZJ4
292	SMC1B	22q13	structural maintenance of chromosomes 1B	Q8NDV3
293	SMPD4	2q21.1	sphingomyelin phosphodiesterase 4, neutral membrane (neutral sphingomyelinase-3)	Q9NXE4
294	SNRPA1	15q26.3	small nuclear ribonucleoprotein polypeptide A'	P09661
295	SNRPB	20p13	small nuclear ribonucleoprotein polypeptides B and B1	P14678
296	SNRPC	6p21	small nuclear ribonucleoprotein polypeptide C	P09234
297	SNRPD1	18q11.2	small nuclear ribonucleoprotein D1 polypeptide 16kDa	P62314
298	SPC24	19p13.2	SPC24, NDC80 kinetochore complex component	Q8NBT2
299	SPC25	2q31.1	SPC25, NDC80 kinetochore complex component	SPCS2
300	SPP1	4q22.1	secreted phosphoprotein 1	P10451
301	SRRT	7q21	serrate RNA effector molecule homolog (Arabidopsis)	Q9BXP5
302	STIL	1p32	SCL/TAL1 interrupting locus	Q15468
303	SUV420H2	19q13.42	suppressor of variegation 4-20 homolog 2 (Drosophila)	Q86Y97
304	TACC3	4p16.3	transforming, acidic coiled-coil containing protein 3	Q9Y6A5
305	TBRG4	7p13	transforming growth factor beta regulator 4	Q969Z0
306	TCF3	19p13.3	transcription factor 3	TCF7L1
307	TELO2	16p13.3	telomere maintenance 2	Q9Y4R8
308	TERT	5p15.33	telomerase reverse transcriptase	O14746
309	TEX19	17q25.3	testis expressed 19	Q8NA77
310	TH1L/NELFCD	20q13.32	negative elongation factor complex member C/D	Q8IXH7
311	THOC6	16p13.3	THO complex 6 homolog (Drosophila)	Q86W42
312	TIGD1	2q37.1	tigger transposable element derived 1	Q96MW7
313	TIGD5	8q24.3	tigger transposable element derived 5	Q53EQ6
314	TIMELESS	12q13.3	timeless circadian clock	Q9UNS1
315	TIPIN	15q22.31	TIMELESS interacting protein	Q9BVW5

Figure 55G

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
316	TK1	17q23.2-q25.3	thymidine kinase 1, soluble	P04183
317	TMEM206	1q32.3	transmembrane protein 206	Q9H813
318	TMEM69	1p34.1	transmembrane protein 69	Q5SWH9
319	TOMM40	19q13	translocase of outer mitochondrial membrane 40 homolog (yeast)	O96008
320	TOP2A	17q21-q22	topoisomerase (DNA) II alpha 170kDa	P11388
321	TPX2	20q11.2	TPX2, microtubule-associated	Q9ULW0
322	TRAF2	9q34	TNF receptor-associated factor 2	Q12933
323	TRAIP	3p21.31	TRAF interacting protein	Q9BWF2
324	TRIM11	1q42.13	tripartite motif containing 11	Q96F44
325	TRIM28	19q13.4	tripartite motif containing 28	Q13263
326	TRIM59	3q26	tripartite motif containing 59	Q8IWR1
327	TRIP13	5p15	thyroid hormone receptor interactor 13	Q15645
328	TROAP	12q13.12	trophinin associated protein	Q12815
329	TTK	6q13-q21	TTK protein kinase	P33981
330	TTYH3	7p22	tweety family member 3	Q9C0H2
331	TUBB3	16q24.3	tubulin, beta 3 class III	Q13509
332	UBE2C	20q13.12	ubiquitin-conjugating enzyme E2C	O00762
333	UBE2O	17q25.1	ubiquitin-conjugating enzyme E2O	Q9C0C9
334	UBE2S	19q13.43	ubiquitin-conjugating enzyme E2S	Q16763
335	UBE2T	1q32.1	ubiquitin-conjugating enzyme E2T (putative)	Q9NPD8
336	UCK2	1p32	uridine-cytidine kinase 2	Q9BZX2
337	UHRF1	19p13.3	ubiquitin-like with PHD and ring finger domains 1	Q96T88
338	UTP18	17q21.33	UTP18 small subunit (SSU) processome component homolog (yeast)	Q9Y5J1
339	WDR4	21q22.3	WD repeat domain 4	P57081
340	WDR67/TBC1D31	8q24.13	TBC1 domain family member 31	Q96DN5
341	WDR75	2q32.2	WD repeat domain 75	Q8IWA0
342	WDR85/DPH7	9q34.3	diphthamide biosynthesis 7	Q9BTV6
343	WDR90	16p13.3	WD repeat domain 90	Q96KV7
344	WHSC1	4p16.3	Wolf-Hirschhorn syndrome candidate 1	O96028
345	XPO5	6p21.1	exportin 5	Q9HAV4
346	XPO6	16p11.2	exportin 6	Q96QU8
347	XPOT	12q14.1	exportin, tRNA	O43592
348	XRCC2	7q36	X-ray repair complementing defective repair in Chinese hamster cells 2	O43543
349	YDJC	22q11.21	YdjC homolog (bacterial)	A8MPS7
350	ZBTB9	6p21.31	zinc finger and BTB domain containing 9	Q96C00
351	ZC3H3	8q24.3	zinc finger CCCH-type containing 3	Q8IXZ2
352	ZIC2	13q32	Zic family member 2	O95409
353	ZNF200	16p13.3	zinc finger protein 200	P98182
354	ZNF643/ZFP69B	1p34.2	ZFP69 zinc finger protein B	Q9UJL9
355	ZNF695	1q44	zinc finger protein 695	Q8IW36
356	ZNF707	8q24.3	zinc finger protein 707	Q96C28
357	ZWILCH	15q22.31	zwilch kinetochore protein	Q9H900
358	ZWINT	10q21-q22	ZW10 interacting kinetochore protein	O95229

Figure 55H

Solid Tumor Methylation markers, 50% sensitivity, CpG Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Solid Tumor Markers	
1	Chr01: 110611465 110611466	*ALX3*
2	Chr01: 111217497 111217498	*KCNA3*
3	Chr01: 12227825 12227826	*TNFRSF1B*
4	Chr01: 154475139 154475140	*TDRD10,SHE*
5	Chr01: 155290607 155290608	*RUSC1,C1orf104*
6	Chr01: 156405438 156405439	*Non Coding Region*
7	Chr01: 169396635 169396636	*C1orf114*
8	Chr01: 170630558 170630559	*Non Coding Region*
9	Chr01: 170637723 170637724	*PRRX1*
10	Chr01: 200009830 200009831	*NR5A2*
11	Chr01: 207818395 207818396	*CR1L*
12	Chr01: 213090203 213090204	*Non Coding Region*
13	Chr01: 219347410 219347411	*LYPLAL1*
14	Chr01: 223302861 223302862	*TLR5*
15	Chr01: 231113419 231113420	*ARV1,TTC13*
16	Chr01: 29586418 29586419	*PTPRU*
17	Chr01: 44883362 44883363	*RNF220*
18	Chr01: 47999163 47999164	*Non Coding Region*
19	Chr01: 50884480 50884481	*DMRTA2*
20	Chr01: 61623366 61623367	*NFIA*
21	Chr01: 66999588 66999589	*SGIP1*
22	Chr01: 77334207 77334208	*ST6GALNAC5*
23	Chr01: 91196332 91196333	*Non Coding Region*
24	Chr02: 119591144 119591145	*Non Coding Region*
25	Chr02: 127413831 127413832	*GYPC*
26	Chr02: 175208605 175208606	*Non Coding Region*
27	Chr02: 176987465 176987466	*HOXD9*
28	Chr02: 176994665 176994666	*HOXD8*
29	Chr02: 177021826 177021827	*Non Coding Region*
30	Chr02: 219738314 219738315	*WNT6*
31	Chr02: 233368543 233368544	*Non Coding Region*
32	Chr02: 238600061 238600062	*LRRFIP1*
33	Chr02: 26395448 26395449	*FAM59B*
34	Chr02: 45240563 45240564	*Non Coding Region*
35	Chr02: 71192445 71192446	*ATP6V1B1*
36	Chr02: 74743437 74743438	*TLX2*
37	Chr02: 85811432 85811433	*VAMP5*
38	Chr02: 97203640 97203641	*ARID5A*
39	Chr03: 121903520 121903521	*CASR*
40	Chr03: 129693489 129693490	*TRH*
41	Chr03: 139258939 139258940	*RBP1*
42	Chr03: 172165884 172165885	*GHSR*
43	Chr03: 179168760 179168761	*GNB4*
44	Chr03: 181421427 181421428	*SOX2OT*
45	Chr03: 184099432 184099433	*CHRD*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Solid Tumor Markers	
46	Chr03: 44063593 44063594	*Non Coding Region*
47	Chr03: 50378191 50378192	*RASSF1*
48	Chr04: 42153708 42153709	*BEND4*
49	Chr05: 115152413 115152414	*CDO1*
50	Chr05: 11904114 11904115	*CTNND2*
51	Chr05: 134827512 134827513	*Non Coding Region*
52	Chr05: 171538557 171538558	*STK10*
53	Chr05: 40681137 40681138	*PTGER4*
54	Chr05: 43018282 43018283	*Non Coding Region*
55	Chr05: 58335681 58335682	*PDE4D*
56	Chr05: 67547334 67547335	*PIK3R1*
57	Chr05: 72595741 72595742	*Non Coding Region*
58	Chr05: 72732486 72732487	*Non Coding Region*
59	Chr06: 105584551 105584552	*BVES*
60	Chr06: 152128865 152128866	*ESR1*
61	Chr06: 26240528 26240529	*HIST1H4F*
62	Chr06: 27107019 27107020	*HIST1H4I,HIST1H2BK*
63	Chr06: 28175446 28175447	*Non Coding Region*
64	Chr06: 28956609 28956610	*Non Coding Region*
65	Chr06: 30139641 30139642	*TRIM15*
66	Chr06: 41978288 41978289	*CCND3*
67	Chr06: 6004207 6004208	*NRN1*
68	Chr06: 75794800 75794801	*COL12A1*
69	Chr06: 75917560 75917561	*Non Coding Region*
70	Chr07: 100488942 100488943	*ACHE*
71	Chr07: 113722758 113722759	*Non Coding Region*
72	Chr07: 127672458 127672459	*LRRC4,SND1*
73	Chr07: 128531106 128531107	*KCP*
74	Chr07: 143580147 143580148	*FAM115A*
75	Chr07: 149917263 149917264	*Non Coding Region*
76	Chr07: 155597842 155597843	*SHH*
77	Chr07: 27192061 27192062	*Non Coding Region*
78	Chr07: 27204981 27204982	*HOXA9*
79	Chr07: 27208285 27208286	*Non Coding Region*
80	Chr07: 27213610 27213611	*HOXA10*
81	Chr07: 27225058 27225059	*HOXA11AS,HOXA11*
82	Chr07: 27286075 27286076	*EVX1*
83	Chr07: 30722320 30722321	*CRHR2*
84	Chr07: 37488253 37488254	*ELMO1*
85	Chr07: 42267747 42267748	*GLI3*
86	Chr07: 95025836 95025837	*PON3*
87	Chr07: 96650171 96650172	*DLX5*
88	Chr08: 101822108 101822109	*Non Coding Region*
89	Chr08: 104512858 104512859	*RIMS2*
90	Chr08: 116679763 116679764	*TRPS1*

Figure 56A

Order	Coordinate of CpG (hg19)	Gene
91	Chr08: 134511683 134511684	*ST3GAL1*
92	Chr08: 55367627 55367628	*Non Coding Region*
93	Chr08: 72917350 72917351	*Non Coding Region*
94	Chr08: 9760880 9760881	*LOC157627*
95	Chr10: 101605988 101605989	*ABCC2*
96	Chr10: 102587110 102587111	*PAX2*
97	Chr10: 104434010 104434011	*ARL3*
98	Chr10: 106400565 106400566	*SORCS3*
99	Chr10: 119590608 119590609	*Non Coding Region*
100	Chr10: 122708532 122708533	*Non Coding Region*
101	Chr10: 129845660 129845661	*PTPRE*
102	Chr10: 15761854 15761855	*ITGA8*
103	Chr10: 22625465 22625466	*Non Coding Region*
104	Chr10: 45914688 45914689	*ALOX5*
105	Chr10: 57390306 57390307	*Non Coding Region*
106	Chr10: 81002218 81002219	*ZMIZ1*
107	Chr11: 10477847 10477848	*AMPD3*
108	Chr11: 105481317 105481318	*GRIA4*
109	Chr11: 119227328 119227329	*USP2*
110	Chr11: 120435056 120435057	*Non Coding Region*
111	Chr11: 123301490 123301491	*Non Coding Region*
112	Chr11: 124790875 124790876	*HEPACAM*
113	Chr11: 133906693 133906694	*LOC100128239*
114	Chr11: 14926738 14926739	*Non Coding Region*
115	Chr11: 2162183 2162184	*IGF2,IGF2AS,INS-IGF2*
116	Chr11: 2292691 2292692	*ASCL2*
117	Chr11: 406901 406902	*SIGIRR*
118	Chr11: 43602847 43602848	*MIR129-2*
119	Chr11: 57250469 57250470	*Non Coding Region*
120	Chr11: 7695528 7695529	*CYB5R2*
121	Chr11: 830320 830321	*EFCAB4A*
122	Chr12: 113541915 113541916	*RASAL1*
123	Chr12: 114843296 114843297	*TBX5*
124	Chr12: 122519674 122519675	*MLXIP*
125	Chr12: 52401214 52401215	*GRASP*
126	Chr12: 52627047 52627048	*KRT7*
127	Chr12: 52652377 52652378	*Non Coding Region*
128	Chr12: 54421033 54421034	*HOXC5,HOXC4,HOXC6*
129	Chr12: 54447584 54447585	*HOXC4*
130	Chr12: 8088818 8088819	*SLC2A3*
131	Chr13: 41548815 41548816	*ELF1*
132	Chr13: 53312966 53312967	*LECT1*
133	Chr13: 79177782 79177783	*POU4F1*
134	Chr13: 96204856 96204857	*CLDN10*
135	Chr14: 105512306 105512307	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
136	Chr14: 23834891 23834892	*EFS*
137	Chr14: 24803873 24803874	*ADCY4*
138	Chr14: 29254530 29254531	*C14orf23*
139	Chr14: 37125531 37125532	*PAX9*
140	Chr14: 38067622 38067623	*Non Coding Region*
141	Chr14: 52735421 52735422	*PTGDR*
142	Chr14: 60952945 60952946	*C14orf39*
143	Chr15: 101420636 101420637	*ALDH1A3*
144	Chr15: 41787780 41787781	*ITPKA*
145	Chr15: 45479755 45479756	*SHF*
146	Chr15: 75251491 75251492	*Non Coding Region*
147	Chr15: 76634516 76634517	*ISL2*
148	Chr15: 83953880 83953881	*BNC1*
149	Chr15: 96515411 96515412	*Non Coding Region*
150	Chr16: 23847325 23847326	*PRKCB*
151	Chr16: 49312033 49312034	*Non Coding Region*
152	Chr16: 88700818 88700819	*Non Coding Region*
153	Chr17: 26699169 26699170	*SARM1*
154	Chr17: 36204489 36204490	*LOC284100*
155	Chr17: 42287971 42287972	*UBTF*
156	Chr17: 53341243 53341244	*HLF*
157	Chr17: 73584617 73584618	*MYO15B*
158	Chr17: 75369219 75369220	*SEPTIN9*
159	Chr18: 3499253 3499254	*DLGAP1*
160	Chr18: 54789058 54789059	*Non Coding Region*
161	Chr18: 904920 904921	*ADCYAP1*
162	Chr19: 11532175 11532176	*CCDC151*
163	Chr19: 11533499 11533500	*CCDC151*
164	Chr19: 13124776 13124777	*NFIX*
165	Chr19: 16999614 16999615	*F2RL3*
166	Chr19: 17008650 17008651	*CPAMD8*
167	Chr19: 17392923 17392924	*ANKLE1*
168	Chr19: 17403313 17403314	*ABHD8*
169	Chr19: 22034418 22034419	*Non Coding Region*
170	Chr19: 37096323 37096324	*ZNF529,ZNF382*
171	Chr19: 50004294 50004295	*MIR150*
172	Chr19: 58858806 58858807	*A1BG*
173	Chr20: 30639305 30639306	*HCK*
174	Chr20: 36932553 36932554	*BPI*
175	Chr20: 45142244 45142245	*ZNF334*
176	Chr20: 62369445 62369446	*LIME1*
177	Chr22: 19754448 19754449	*TBX1*
178	Chr22: 38221187 38221188	*GALR3*
179	Chr22: 46367617 46367618	*WNT7B*

Figure 56B

Order	Coordinate of CpG (hg19)	Gene
	Alternative markers: Low to no CRC	
180	Chr01: 1072902 1072903	*Non Coding Region*
181	Chr01: 1079561 1079562	*Non Coding Region*
182	Chr01: 1079622 1079623	*Non Coding Region*
183	Chr01: 110610899 110610900	*ALX3*
184	Chr01: 115642150 115642151	*Non Coding Region*
185	Chr01: 12227825 12227826	*TNFRSF1B*
186	Chr01: 14057606 14057607	*PRDM2*
187	Chr01: 149719536 149719537	*Non Coding Region*
188	Chr01: 151693261 151693262	*C1orf230*
189	Chr01:_151812421_151812422	*LOC100132111,C2CD4D*
190	Chr01:_151812435_151812436	*LOC100132111,C2CD4D*
191	Chr01: 153330200 153330201	*S100A9*
192	Chr01: 154377091 154377092	*IL6R*
193	Chr01:_155290607_155290608	*RUSC1,C1orf104*
194	Chr01:_156405438_156405439	*Non Coding Region*
195	Chr01: 159880327 159880328	*Non Coding Region*
196	Chr01: 16489048 16489049	*Non Coding Region*
197	Chr01: 165323692 165323693	*LMX1A*
198	Chr01: 167417043 167417044	*CD247*
199	Chr01: 17027832 17027833	*ESPNP*
200	Chr01: 170637723 170637724	*PRRX1*
201	Chr01: 170640675 170640676	*PRRX1*
202	Chr01: 17577455 17577456	*PADI3*
203	Chr01: 200009830 200009831	*NR5A2*
204	Chr01: 2064691 2064692	*PRKCZ*
205	Chr01: 2064819 2064820	*PRKCZ*
206	Chr01: 207999258 207999259	*Non Coding Region*
207	Chr01: 211500971 211500972	*TRAF5*
208	Chr01: 213090203 213090204	*Non Coding Region*
209	Chr01: 214159131 214159132	*Non Coding Region*
210	Chr01: 217309794 217309795	*ESRRG*
211	Chr01: 219347410 219347411	*LYPLAL1*
212	Chr01: 219347458 219347459	*LYPLAL1*
213	Chr01: 219834537 219834538	*Non Coding Region*
214	Chr01: 2222419 2222420	*SKI*
215	Chr01: 223302861 223302862	*TLR5*
216	Chr01: 226288430 226288431	*Non Coding Region*
217	Chr01: 231113419 231113420	*ARV1,TTC13*
218	Chr01: 23851471 23851472	*E2F2*
219	Chr01: 240255486 240255487	*FMN2*
220	Chr01: 246733057 246733058	*CNST*
221	Chr01: 248020632 248020633	*TRIM58*
222	Chr01: 248020812 248020813	*TRIM58*
223	Chr01: 26611453 26611454	*UBXN11*

Order	Coordinate of CpG (hg19)	Gene
224	Chr01:_27028974_27028975	*ARID1A*
225	Chr01: 27668325 27668326	*SYTL1*
226	Chr01: 28996561 28996562	*GMEB1*
227	Chr01: 29586414 29586415	*PTPRU*
228	Chr01: 29586418 29586419	*PTPRU*
229	Chr01: 32170929 32170930	*COL16A1*
230	Chr01: 32171050 32171051	*COL16A1*
231	Chr01: 32180356 32180357	*Non Coding Region*
232	Chr01: 32410211 32410212	*Non Coding Region*
233	Chr01: 42611386 42611387	*Non Coding Region*
234	Chr01:_44883362_44883363	*RNF220*
235	Chr01:_46958801_46958802	*Non Coding Region*
236	Chr01: 47695138 47695139	*TAL1*
237	Chr01: 47999163 47999164	*Non Coding Region*
238	Chr01:_5526684_5526685	*Non Coding Region*
239	Chr01: 6086448 6086449	*KCNAB2*
240	Chr01: 6086483 6086484	*KCNAB2*
241	Chr01: 61623366 61623367	*NFIA*
242	Chr01: 6508028 6508029	*ESPN*
243	Chr01: 66999588 66999589	*SGIP1*
244	Chr01: 7023625 7023626	*CAMTA1*
245	Chr01: 76540574 76540575	*ST6GALNAC3*
246	Chr01: 848379 848380	*Non Coding Region*
247	Chr01: 90309410 90309411	*LRRC8D*
248	Chr01: 90309559 90309560	*LRRC8D*
249	Chr01: 91196332 91196333	*Non Coding Region*
250	Chr01: 92150549 92150550	*TGFBR3*
251	Chr01: 976168 976169	*AGRN*
252	Chr02: 105470561 105470562	*POU3F3*
253	Chr02: 111875790 111875791	*ACOXL*
254	Chr02: 111875934 111875935	*Non Coding Region*
255	Chr02: 111877065 111877066	*BCL2L11*
256	Chr02: 113450279 113450280	*Non Coding Region*
257	Chr02: 119594696 119594697	*Non Coding Region*
258	Chr02: 119606746 119606747	*EN1*
259	Chr02: 119607885 119607886	*Non Coding Region*
260	Chr02: 127783168 127783169	*Non Coding Region*
261	Chr02: 134325873 134325874	*NCKAP5*
262	Chr02: 160760825 160760826	*LY75*
263	Chr02: 160761244 160761245	*LY75*
264	Chr02: 160761556 160761557	*LY75*
265	Chr02: 160761622 160761623	*LY75*
266	Chr02: 162283705 162283706	*Non Coding Region*
267	Chr02: 173327921 173327922	*ITGA6*
268	Chr02: 175193377 175193378	*Non Coding Region*

Figure 56C

Order	Coordinate of CpG (hg19)	Gene
269	Chr02:_175208588_175208589	*Non Coding Region*
270	Chr02:_175208605_175208606	*Non Coding Region*
271	Chr02:_176976802_176976803	*Non Coding Region*
272	Chr02:_176987462_176987463	*HOXD9*
273	Chr02:_176987465_176987466	*HOXD9*
274	Chr02:_176993632_176993633	*HOXD8*
275	Chr02:_176994665_176994666	*HOXD8*
276	Chr02:_177021826_177021827	*Non Coding Region*
277	Chr02:_177023716_177023717	*Non Coding Region*
278	Chr02:_177029921_177029922	*HOXD3*
279	Chr02:_177039862_177039863	*Non Coding Region*
280	Chr02:_177043255_177043256	*Non Coding Region*
281	Chr02:_192711481_192711482	*SDPR*
282	Chr02:_19561612_19561613	*Non Coding Region*
283	Chr02:_200334137_200334138	*FLJ32063*
284	Chr02:_208491939_208491940	*Non Coding Region*
285	Chr02:_217217837_217217838	*MARCHF4*
286	Chr02:_219735876_219735877	*WNT6*
287	Chr02:_219736549_219736550	*WNT6*
288	Chr02:_219738314_219738315	*WNT6*
289	Chr02:_220117599_220117600	*TUBA4A,TUBA4B*
290	Chr02:_223162881_223162882	*CCDC140,PAX3*
291	Chr02:_227288327_227288328	*Non Coding Region*
292	Chr02:_233368543_233368544	*Non Coding Region*
293	Chr02:_238582364_238582365	*LRRFIP1*
294	Chr02:_238600061_238600062	*LRRFIP1*
295	Chr02:_25142229_25142230	*ADCY3*
296	Chr02:_25518238_25518239	*DNMT3A*
297	Chr02:_25585619_25585620	*Non Coding Region*
298	Chr02:_26395448_26395449	*FAM59B*
299	Chr02:_26395458_26395459	*FAM59B*
300	Chr02:_29537776_29537777	*ALK*
301	Chr02:_43422121_43422122	*Non Coding Region*
302	Chr02:_45162036_45162037	*Non Coding Region*
303	Chr02:_45169548_45169549	*SIX3*
304	Chr02:_45169562_45169563	*SIX3*
305	Chr02:_45169681_45169682	*SIX3*
306	Chr02:_45240563_45240564	*Non Coding Region*
307	Chr02:_62798365_62798366	*Non Coding Region*
308	Chr02:_63284066_63284067	*OTX1*
309	Chr02:_63284574_63284575	*Non Coding Region*
310	Chr02:_71192118_71192119	*ATP6V1B1*
311	Chr02:_71192445_71192446	*ATP6V1B1*
312	Chr02:_73151595_73151596	*EMX1*
313	Chr02:_74782684_74782685	*DOK1*

Order	Coordinate of CpG (hg19)	Gene
314	Chr02:_80530255_80530256	*LRRTM1,CTNNA2*
315	Chr02:_85107320_85107321	*C2orf89*
316	Chr02:_85811432_85811433	*VAMP5*
317	Chr02:_85980500_85980501	*ATOH8*
318	Chr02:_85980533_85980534	*ATOH8*
319	Chr02:_96990762_96990763	*ITPRIPL1*
320	Chr02:_97203640_97203641	*ARID5A*
321	Chr03:_107318032_107318033	*BBX*
322	Chr03:_111261010_111261011	*CD96*
323	Chr03:_113252640_113252641	*SIDT1*
324	Chr03:_122296369_122296370	*PARP15*
325	Chr03:_138658554_138658555	*Non Coding Region*
326	Chr03:_138658704_138658705	*Non Coding Region*
327	Chr03:_147124417_147124418	*ZIC4*
328	Chr03:_157821407_157821408	*SHOX2*
329	Chr03:_16523047_16523048	*RFTN1*
330	Chr03:_181421427_181421428	*SOX2OT*
331	Chr03:_181437299_181437300	*SOX2OT*
332	Chr03:_184099432_184099433	*CHRD*
333	Chr03:_193506005_193506006	*Non Coding Region*
334	Chr03:_194024145_194024146	*LOC100131551*
335	Chr03:_46448579_46448580	*CCRL2*
336	Chr03:_46448631_46448632	*CCRL2*
337	Chr03:_46448671_46448672	*CCRL2*
338	Chr03:_46448753_46448754	*CCRL2*
339	Chr03:_46448868_46448869	*CCRL2*
340	Chr03:_46448920_46448921	*CCRL2*
341	Chr03:_46448963_46448964	*CCRL2*
342	Chr03:_47029335_47029336	*NBEAL2*
343	Chr03:_49057884_49057885	*MIR425,DALRD3,NDUFAF3*
344	Chr03:_5018817_5018818	*Non Coding Region*
345	Chr03:_50377988_50377989	*RASSF1*
346	Chr03:_50378191_50378192	*RASSF1*
347	Chr03:_50378292_50378293	*RASSF1*
348	Chr03:_50378407_50378408	*RASSF1*
349	Chr03:_50378413_50378414	*RASSF1*
350	Chr03:_50378423_50378424	*RASSF1*
351	Chr03:_50378425_50378426	*RASSF1*
352	Chr03:_50378431_50378432	*RASSF1*
353	Chr03:_50378527_50378528	*RASSF1*
354	Chr03:_50378611_50378612	*RASSF1,ZMYND10*
355	Chr03:_53190593_53190594	*Non Coding Region*
356	Chr03:_5470909_5470910	*Non Coding Region*
357	Chr03:_65582925_65582926	*MAGI1*
358	Chr03:_73620925_73620926	*PDZRN3*

Figure 56D

Order	Coordinate of CpG (hg19)	Gene
359	Chr03: 9904411 9904412	*Non Coding Region*
360	Chr04: 166414666 166414667	*CPE*
361	Chr04: 174254281 174254282	*HMGB2*
362	Chr04: 185747981 185747982	*ACSL1*
363	Chr04: 186434993 186434994	*PDLIM3*
364	Chr04: 54960232 54960233	*Non Coding Region*
365	Chr04: 6690702 6690703	*Non Coding Region*
366	Chr04: 6944071 6944072	*TBC1D14*
367	Chr04: 76555980 76555981	*CDKL2*
368	Chr04: 81124600 81124601	*PRDM8*
369	Chr04: 85402870 85402871	*Non Coding Region*
370	Chr04: 85417612 85417613	*NKX6-1*
371	Chr05: 100240048 100240049	*ST8SIA4*
372	Chr05: 101632310 101632311	*SLCO4C1*
373	Chr05: 101632314 101632315	*SLCO4C1*
374	Chr05: 101632321 101632322	*SLCO4C1*
375	Chr05: 101632327 101632328	*SLCO4C1*
376	Chr05: 112073433 112073434	*APC*
377	Chr05: 112073570 112073571	*APC*
378	Chr05: 134827355 134827356	*Non Coding Region*
379	Chr05: 134827512 134827513	*Non Coding Region*
380	Chr05: 138676787 138676788	*PAIP2*
381	Chr05: 139080952 139080953	*Non Coding Region*
382	Chr05: 139081063 139081064	*Non Coding Region*
383	Chr05: 141931455 141931456	*Non Coding Region*
384	Chr05: 153862394 153862395	*Non Coding Region*
385	Chr05: 170739179 170739180	*Non Coding Region*
386	Chr05: 17119012 17119013	*Non Coding Region*
387	Chr05: 171538334 171538335	*STK10*
388	Chr05: 171538557 171538558	*STK10*
389	Chr05: 172655925 172655926	*Non Coding Region*
390	Chr05: 175961105 175961106	*RNF44*
391	Chr05: 176024005 176024006	*GPRIN1*
392	Chr05: 176326365 176326366	*HK3*
393	Chr05: 176784688 176784689	*RGS14*
394	Chr05: 176784719 176784720	*RGS14*
395	Chr05: 37835168 37835169	*GDNF*
396	Chr05: 38257541 38257542	*EGFLAM*
397	Chr05: 40681137 40681138	*PTGER4*
398	Chr05: 40681893 40681894	*PTGER4*
399	Chr05: 40682069 40682070	*PTGER4*
400	Chr05: 42951192 42951193	*Non Coding Region*
401	Chr05: 42952113 42952114	*Non Coding Region*
402	Chr05: 42992774 42992775	*Non Coding Region*
403	Chr05: 42995203 42995204	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
404	Chr05: 43017561 43017562	*Non Coding Region*
405	Chr05: 43017685 43017686	*Non Coding Region*
406	Chr05: 43018282 43018283	*Non Coding Region*
407	Chr05: 43018443 43018444	*Non Coding Region*
408	Chr05: 55219492 55219493	*Non Coding Region*
409	Chr05: 58334676 58334677	*PDE4D*
410	Chr05: 58335681 58335682	*PDE4D*
411	Chr05: 67547334 67547335	*PIK3R1*
412	Chr05: 72732486 72732487	*Non Coding Region*
413	Chr05: 74231171 74231172	*Non Coding Region*
414	Chr05: 76145809 76145810	*S100Z*
415	Chr05: 95159365 95159366	*GLRX*
416	Chr06: 10385903 10385904	*Non Coding Region*
417	Chr06: 10422322 10422323	*Non Coding Region*
418	Chr06: 108145420 108145421	*SCML4*
419	Chr06: 134499419 134499420	*SGK1*
420	Chr06: 136359046 136359047	*PDE7B*
421	Chr06: 1384491 1384492	*Non Coding Region*
422	Chr06: 1386087 1386088	*Non Coding Region*
423	Chr06: 151188081 151188082	*MTHFD1L*
424	Chr06: 152623094 152623095	*SYNE1*
425	Chr06: 1606394 1606395	*Non Coding Region*
426	Chr06: 164088958 164088959	*Non Coding Region*
427	Chr06: 166582206 166582207	*T*
428	Chr06: 168197410 168197411	*C6orf123*
429	Chr06: 25138551 25138552	*CMAH*
430	Chr06: 25962567 25962568	*TRIM38*
431	Chr06: 26614649 26614650	*Non Coding Region*
432	Chr06: 27101763 27101764	*HIST1H2BJ*
433	Chr06: 27107002 27107003	*HIST1H4I,HIST1H2BK*
434	Chr06: 27107019 27107020	*HIST1H4I,HIST1H2BK*
435	Chr06: 27782484 27782485	*HIST1H2BM,HIST1H2AJ*
436	Chr06: 27841230 27841231	*HIST1H3I,HIST1H4L*
437	Chr06: 27858545 27858546	*HIST1H3J*
438	Chr06: 28175446 28175447	*Non Coding Region*
439	Chr06: 28557400 28557401	*Non Coding Region*
440	Chr06: 28557411 28557412	*Non Coding Region*
441	Chr06: 28557518 28557519	*Non Coding Region*
442	Chr06: 28956609 28956610	*Non Coding Region*
443	Chr06: 29894737 29894738	*HCG4P6*
444	Chr06: 29894820 29894821	*HCG4P6*
445	Chr06: 29895074 29895075	*Non Coding Region*
446	Chr06: 30458158 30458159	*HLA-E*
447	Chr06: 31705409 31705410	*CLIC1*
448	Chr06: 32160455 32160456	*GPSM3*

Figure 56E

Order	Coordinate of CpG (hg19)	Gene
449	Chr06: 32407838 32407839	*HLA-DRA*
450	Chr06: 35265341 35265342	*DEF6*
451	Chr06: 35265379 35265380	*DEF6*
452	Chr06: 35265411 35265412	*DEF6*
453	Chr06: 35265544 35265545	*DEF6*
454	Chr06: 35265555 35265556	*DEF6*
455	Chr06: 35265616 35265617	*DEF6*
456	Chr06: 35265624 35265625	*DEF6*
457	Chr06: 35265643 35265644	*DEF6*
458	Chr06: 35265759 35265760	*DEF6*
459	Chr06: 35420754 35420755	*FANCE*
460	Chr06: 36973448 36973449	*FGD2*
461	Chr06: 41472382 41472383	*Non Coding Region*
462	Chr06: 41908262 41908263	*CCND3*
463	Chr06: 41978288 41978289	*CCND3*
464	Chr06: 45645497 45645498	*Non Coding Region*
465	Chr06: 50808699 50808700	*TFAP2B*
466	Chr06: 52439807 52439808	*TRAM2*
467	Chr06: 6004207 6004208	*NRN1*
468	Chr06: 6007670 6007671	*NRN1*
469	Chr06: 7107108 7107109	*RREB1*
470	Chr06: 7137908 7137909	*RREB1*
471	Chr06: 73330208 73330209	*KCNQ5*
472	Chr06: 75794800 75794801	*COL12A1*
473	Chr06: 75917560 75917561	*Non Coding Region*
474	Chr06: 91004452 91004453	*BACH2*
475	Chr07: 100042467 100042468	*Non Coding Region*
476	Chr07: 100203506 100203507	*PCOLCE*
477	Chr07: 113722758 113722759	*Non Coding Region*
478	Chr07: 113723162 113723163	*Non Coding Region*
479	Chr07: 113723569 113723570	*Non Coding Region*
480	Chr07: 127671193 127671194	*LRRC4,SND1*
481	Chr07: 128531106 128531107	*KCP*
482	Chr07: 128809228 128809229	*Non Coding Region*
483	Chr07: 130126024 130126025	*MEST*
484	Chr07: 151078646 151078647	*WDR86*
485	Chr07: 151078750 151078751	*WDR86*
486	Chr07: 151330230 151330231	*PRKAG2*
487	Chr07: 155597842 155597843	*SHH*
488	Chr07: 158620335 158620336	*ESYT2*
489	Chr07: 23508189 23508190	*IGF2BP3*
490	Chr07: 2551513 2551514	*LFNG*
491	Chr07: 26193109 26193110	*NFE2L3*
492	Chr07: 27192061 27192062	*Non Coding Region*
493	Chr07: 27192246 27192247	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
494	Chr07: 27196286 27196287	*HOXA7*
495	Chr07: 27196302 27196303	*HOXA7*
496	Chr07: 27197614 27197615	*HOXA7*
497	Chr07: 27204005 27204006	*HOXA9*
498	Chr07: 27204349 27204350	*HOXA9*
499	Chr07: 27204981 27204982	*HOXA9*
500	Chr07: 27205217 27205218	*HOXA9*
501	Chr07: 27205224 27205225	*HOXA9*
502	Chr07: 27205230 27205231	*HOXA9*
503	Chr07: 27208285 27208286	*Non Coding Region*
504	Chr07: 27213610 27213611	*HOXA10*
505	Chr07: 27225058 27225059	*HOXA11AS,HOXA11*
506	Chr07: 27225523 27225524	*HOXA11AS,HOXA11*
507	Chr07: 27225528 27225529	*HOXA11AS,HOXA11*
508	Chr07: 27240758 27240759	*HOXA13*
509	Chr07: 27242044 27242045	*Non Coding Region*
510	Chr07: 30951432 30951433	*AQP1*
511	Chr07: 32802694 32802695	*Non Coding Region*
512	Chr07: 33080615 33080616	*NT5C3*
513	Chr07: 37960317 37960318	*EPDR1*
514	Chr07: 45025620 45025621	*SNORA9,C7orf40*
515	Chr07: 64349133 64349134	*Non Coding Region*
516	Chr07: 64349557 64349558	*Non Coding Region*
517	Chr07: 64349706 64349707	*Non Coding Region*
518	Chr07: 73417264 73417265	*Non Coding Region*
519	Chr07: 76828885 76828886	*CCDC146,FGL2*
520	Chr07: 95025836 95025837	*PON3*
521	Chr07: 96650171 96650172	*DLX5*
522	Chr07: 96654782 96654783	*DLX5*
523	Chr07: 98311382 98311383	*Non Coding Region*
524	Chr07: 99595276 99595277	*Non Coding Region*
525	Chr08: 101822108 101822109	*Non Coding Region*
526	Chr08: 124173385 124173386	*Non Coding Region*
527	Chr08: 125721998 125721999	*MTSS1*
528	Chr08: 134511683 134511684	*ST3GAL1*
529	Chr08: 134581043 134581044	*ST3GAL1*
530	Chr08: 1414952 1414953	*Non Coding Region*
531	Chr08: 145013249 145013250	*PLEC1*
532	Chr08: 145013478 145013479	*PLEC1*
533	Chr08: 2023251 2023252	*MYOM2*
534	Chr08: 2023317 2023318	*MYOM2*
535	Chr08: 24151625 24151626	*ADAM28*
536	Chr08: 49612450 49612451	*Non Coding Region*
537	Chr08: 61917230 61917231	*Non Coding Region*
538	Chr08: 82192519 82192520	*FABP5*

Figure 56F

539	Chr08: 99951730 99951731	*Non Coding Region*
540	Chr08: 99952047 99952048	*Non Coding Region*
541	Chr09: 112810402 112810403	*PALM2-AKAP2,AKAP2*
542	Chr09: 112812363 112812364	*PALM2-AKAP2,AKAP2*
543	Chr09: 126777673 126777674	*LHX2*
544	Chr09: 136603716 136603717	*SARDH*
545	Chr09: 140172089 140172090	*C9orf167*
546	Chr09: 27528300 27528301	*MOBKL2B*
547	Chr09: 34591041 34591042	*CNTFR*
548	Chr09: 38339335 38339336	*Non Coding Region*
549	Chr09: 79627216 79627217	*Non Coding Region*
550	Chr09: 96721468 96721469	*Non Coding Region*
551	Chr10: 100206094 100206095	*HPS1*
552	Chr10: 101605988 101605989	*ABCC2*
553	Chr10: 101606026 101606027	*ABCC2*
554	Chr10: 102586999 102587000	*PAX2*
555	Chr10: 102587110 102587111	*PAX2*
556	Chr10: 102589102 102589103	*PAX2*
557	Chr10: 104434010 104434011	*ARL3*
558	Chr10: 112624555 112624556	*Non Coding Region*
559	Chr10: 114136053 114136054	*ACSL5*
560	Chr10: 114136073 114136074	*ACSL5*
561	Chr10: 119304487 119304488	*EMX2OS,EMX2*
562	Chr10: 119590450 119590451	*Non Coding Region*
563	Chr10: 119590608 119590609	*Non Coding Region*
564	Chr10: 122708532 122708533	*Non Coding Region*
565	Chr10: 122708739 122708740	*Non Coding Region*
566	Chr10: 129845660 129845661	*PTPRE*
567	Chr10: 133484610 133484611	*Non Coding Region*
568	Chr10: 134872255 134872256	*Non Coding Region*
569	Chr10: 16933505 16933506	*CUBN*
570	Chr10: 22625465 22625466	*Non Coding Region*
571	Chr10: 45914688 45914689	*ALOX5*
572	Chr10: 5671107 5671108	*Non Coding Region*
573	Chr10: 63510947 63510948	*C10orf107*
574	Chr10: 79687289 79687290	*LOC100128292,DLG5*
575	Chr10: 88730946 88730947	*AGAP11*
576	Chr10: 99798430 99798431	*Non Coding Region*
577	Chr11: 10477399 10477400	*AMPD3*
578	Chr11: 10477452 10477453	*AMPD3*
579	Chr11: 10477461 10477462	*AMPD3*
580	Chr11: 10477847 10477848	*AMPD3*
581	Chr11: 104916075 104916076	*CARD16,CASP1*
582	Chr11: 118481025 118481026	*PHLDB1*
583	Chr11: 119227328 119227329	*USP2*

584	Chr11: 120435056 120435057	*Non Coding Region*
585	Chr11: 2292691 2292692	*ASCL2*
586	Chr11: 2548282 2548283	*KCNQ1*
587	Chr11: 268976 268977	*Non Coding Region*
588	Chr11: 31841524 31841525	*Non Coding Region*
589	Chr11: 406829 406830	*SIGIRR*
590	Chr11: 406901 406902	*SIGIRR*
591	Chr11: 415088 415089	*SIGIRR*
592	Chr11: 46370729 46370730	*DGKZ*
593	Chr11: 46389307 46389308	*DGKZ*
594	Chr11: 47290613 47290614	*MADD*
595	Chr11: 61062910 61062911	*VWCE*
596	Chr11: 627152 627153	*SCT*
597	Chr11: 67184596 67184597	*ATPGD1*
598	Chr11: 67351608 67351609	*GSTP1*
599	Chr11: 68622739 68622740	*Non Coding Region*
600	Chr11: 720859 720860	*EPS8L2*
601	Chr11: 76382149 76382150	*LRRC32*
602	Chr11: 7695528 7695529	*CYB5R2*
603	Chr12: 100751051 100751052	*SLC17A8*
604	Chr12: 111844676 111844677	*SH2B3*
605	Chr12: 113416061 113416062	*OAS2*
606	Chr12: 113416518 113416519	*OAS2*
607	Chr12: 114836014 114836015	*TBX5*
608	Chr12: 116997079 116997080	*MAP1LC3B2*
609	Chr12: 116997535 116997536	*MAP1LC3B2*
610	Chr12: 122277360 122277361	*Non Coding Region*
611	Chr12: 122519674 122519675	*MLXIP*
612	Chr12: 130529951 130529952	*Non Coding Region*
613	Chr12: 131996019 131996020	*Non Coding Region*
614	Chr12: 132657819 132657820	*Non Coding Region*
615	Chr12: 278159 278160	*IQSEC3*
616	Chr12: 3309495 3309496	*TSPAN9*
617	Chr12: 3309708 3309709	*TSPAN9*
618	Chr12: 3862575 3862576	*EFCAB4B*
619	Chr12: 3862597 3862598	*EFCAB4B*
620	Chr12: 46663660 46663661	*SLC38A1*
621	Chr12: 49391363 49391364	*DDN*
622	Chr12: 52317302 52317303	*Non Coding Region*
623	Chr12: 52317475 52317476	*Non Coding Region*
624	Chr12: 52652377 52652378	*Non Coding Region*
625	Chr12: 53601015 53601016	*ITGB7*
626	Chr12: 53601050 53601051	*ITGB7*
627	Chr12: 54071111 54071112	*ATP5G2*
628	Chr12: 54132568 54132569	*Non Coding Region*

Figure 56G

Order	Coordinate of CpG (hg19)	Gene
629	Chr12: 54357530 54357531	*HOTAIR*
630	Chr12: 54389891 54389892	*Non Coding Region*
631	Chr12: 54398809 54398810	*Non Coding Region*
632	Chr12: 54408664 54408665	*Non Coding Region*
633	Chr12: 54409321 54409322	*HOXC5,HOXC4,HOXC6*
634	Chr12: 54409491 54409492	*HOXC5,HOXC4,HOXC6*
635	Chr12: 54421033 54421034	*HOXC5,HOXC4,HOXC6*
636	Chr12: 54446944 54446945	*HOXC4*
637	Chr12: 57618943 57618944	*NXPH4*
638	Chr12: 58013458 58013459	*SLC26A10*
639	Chr12: 58013475 58013476	*SLC26A10*
640	Chr12: 58013636 58013637	*SLC26A10*
641	Chr12: 58013645 58013646	*SLC26A10*
642	Chr12: 58021309 58021310	*B4GALNT1*
643	Chr12: 58135833 58135834	*AGAP2*
644	Chr12: 6184355 6184356	*VWF*
645	Chr12: 70083096 70083097	*BEST3*
646	Chr12: 8088818 8088819	*SLC2A3*
647	Chr12: 98793604 98793605	*Non Coding Region*
648	Chr13: 100608263 100608264	*Non Coding Region*
649	Chr13: 100641867 100641868	*Non Coding Region*
650	Chr13: 100641900 100641901	*Non Coding Region*
651	Chr13: 100643312 100643313	*Non Coding Region*
652	Chr13: 108922274 108922275	*TNFSF13B*
653	Chr13: 113623300 113623301	*MCF2L*
654	Chr13: 20767968 20767969	*GJB2*
655	Chr13: 20768138 20768139	*GJB2*
656	Chr13: 24844846 24844847	*SPATA13*
657	Chr13: 24844852 24844853	*SPATA13*
658	Chr13: 24844896 24844897	*SPATA13*
659	Chr13: 28491615 28491616	*Non Coding Region*
660	Chr13: 28527255 28527256	*Non Coding Region*
661	Chr13: 28544760 28544761	*CDX2*
662	Chr13: 41548815 41548816	*ELF1*
663	Chr13: 43566618 43566619	*EPSTI1*
664	Chr13: 43566642 43566643	*EPSTI1*
665	Chr13: 50202547 50202548	*ARL11*
666	Chr13: 50202570 50202571	*ARL11*
667	Chr13: 79176272 79176273	*POU4F1*
668	Chr13: 79176572 79176573	*POU4F1*
669	Chr13: 79177467 79177468	*POU4F1*
670	Chr13: 79177782 79177783	*POU4F1*
671	Chr13: 98794470 98794471	*FARP1*
672	Chr14: 100613518 100613519	*DEGS2*
673	Chr14: 105512306 105512307	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
674	Chr14: 24808787 24808788	*RIPK3*
675	Chr14: 24808987 24808988	*RIPK3*
676	Chr14: 24809068 24809069	*RIPK3*
677	Chr14: 24809073 24809074	*RIPK3*
678	Chr14: 24809226 24809227	*RIPK3*
679	Chr14: 24809297 24809298	*RIPK3*
680	Chr14: 24809442 24809443	*RIPK3*
681	Chr14: 37125531 37125532	*PAX9*
682	Chr14: 50527872 50527873	*Non Coding Region*
683	Chr14: 52734525 52734526	*PTGDR*
684	Chr14: 52735421 52735422	*PTGDR*
685	Chr14: 54423807 54423808	*BMP4*
686	Chr14: 57283436 57283437	*OTX2OS1*
687	Chr14: 60618622 60618623	*DHRS7*
688	Chr14: 61104967 61104968	*Non Coding Region*
689	Chr14: 62035475 62035476	*Non Coding Region*
690	Chr14: 62218076 62218077	*Non Coding Region*
691	Chr14: 76009633 76009634	*BATF*
692	Chr15: 31727289 31727290	*Non Coding Region*
693	Chr15: 33011166 33011167	*GREM1*
694	Chr15: 41787780 41787781	*ITPKA*
695	Chr15: 42262356 42262357	*EHD4*
696	Chr15: 43414974 43414975	*Non Coding Region*
697	Chr15: 45479755 45479756	*SHF*
698	Chr15: 55543264 55543265	*RAB27A*
699	Chr15: 60690826 60690827	*ANXA2*
700	Chr15: 70785399 70785400	*Non Coding Region*
701	Chr15: 75076251 75076252	*CSK*
702	Chr15: 75082885 75082886	*CSK*
703	Chr15: 75251491 75251492	*Non Coding Region*
704	Chr15: 76627666 76627667	*ISL2*
705	Chr15: 76630095 76630096	*ISL2*
706	Chr15: 76633627 76633628	*ISL2*
707	Chr15: 76633981 76633982	*ISL2*
708	Chr15: 76634516 76634517	*ISL2*
709	Chr15: 81292171 81292172	*MESDC1*
710	Chr15: 89949776 89949777	*Non Coding Region*
711	Chr15: 93617030 93617031	*RGMA*
712	Chr15: 96515411 96515412	*Non Coding Region*
713	Chr16: 19980811 19980812	*Non Coding Region*
714	Chr16: 23847325 23847326	*PRKCB*
715	Chr16: 27378982 27378983	*Non Coding Region*
716	Chr16: 27461445 27461446	*IL21R*
717	Chr16: 29086428 29086429	*RRN3P2*
718	Chr16: 31488238 31488239	*TGFB1I1*

Figure 56H

Order	Coordinate of CpG (hg19)	Gene
719	Chr16: 4363832 4363833	*Non Coding Region*
720	Chr16: 50300322 50300323	*Non Coding Region*
721	Chr16: 54316049 54316050	*Non Coding Region*
722	Chr16: 56672387 56672388	*MT1A*
723	Chr16: 69597925 69597926	*NFAT5*
724	Chr16: 72821498 72821499	*ZFHX3*
725	Chr16: 85784062 85784063	*C16orf74*
726	Chr16: 85932666 85932667	*IRF8*
727	Chr16: 86232457 86232458	*Non Coding Region*
728	Chr16: 88700818 88700819	*Non Coding Region*
729	Chr16: 88717134 88717135	*CYBA*
730	Chr16: 88717374 88717375	*CYBA*
731	Chr16: 88717456 88717457	*CYBA*
732	Chr16: 88717472 88717473	*CYBA*
733	Chr16: 88717646 88717647	*CYBA*
734	Chr16: 89006367 89006368	*CBFA2T3*
735	Chr16: 89006455 89006456	*CBFA2T3*
736	Chr16: 89778301 89778302	*C16orf7*
737	Chr16: 89778660 89778661	*C16orf7*
738	Chr17: 1992954 1992955	*SMG6*
739	Chr17: 19976820 19976821	*Non Coding Region*
740	Chr17: 27893407 27893408	*ABHD15*
741	Chr17: 3438918 3438919	*TRPV3*
742	Chr17: 3438933 3438934	*TRPV3*
743	Chr17: 35078971 35078972	*Non Coding Region*
744	Chr17: 35293930 35293931	*LHX1*
745	Chr17: 35294476 35294477	*LHX1*
746	Chr17: 35294481 35294482	*LHX1*
747	Chr17: 35294491 35294492	*LHX1*
748	Chr17: 36105337 36105338	*HNF1B*
749	Chr17: 36204489 36204490	*LOC284100*
750	Chr17: 36715074 36715075	*SRCIN1*
751	Chr17: 36715953 36715954	*SRCIN1*
752	Chr17: 36728456 36728457	*SRCIN1*
753	Chr17: 37028650 37028651	*LASP1*
754	Chr17: 3819880 3819881	*P2RX1*
755	Chr17: 40089267 40089268	*TTC25*
756	Chr17: 42287971 42287972	*UBTF*
757	Chr17: 43306802 43306803	*FMNL1*
758	Chr17: 43392165 43392166	*MAP3K14*
759	Chr17: 44896877 44896878	*WNT3*
760	Chr17: 46797042 46797043	*Non Coding Region*
761	Chr17: 48050274 48050275	*DLX4*
762	Chr17: 48351821 48351822	*TMEM92*
763	Chr17: 53341243 53341244	*HLF*

Order	Coordinate of CpG (hg19)	Gene
764	Chr17: 56406294 56406295	*BZRAP1*
765	Chr17: 56414613 56414614	*Non Coding Region*
766	Chr17: 56415478 56415479	*Non Coding Region*
767	Chr17: 57719682 57719683	*CLTC*
768	Chr17: 59480674 59480675	*TBX2*
769	Chr17: 59480730 59480731	*TBX2*
770	Chr17: 62777648 62777649	*LOC146880*
771	Chr17: 62777693 62777694	*LOC146880*
772	Chr17: 62777761 62777762	*LOC146880*
773	Chr17: 62777777 62777778	*LOC146880*
774	Chr17: 63225392 63225393	*Non Coding Region*
775	Chr17: 66255427 66255428	*ARSG*
776	Chr17: 7239821 7239822	*ACAP1*
777	Chr17: 75567635 75567636	*Non Coding Region*
778	Chr17: 75877811 75877812	*FLJ45079*
779	Chr17: 76128481 76128482	*TMC6,TMC8*
780	Chr17: 76128522 76128523	*TMC6,TMC8*
781	Chr17: 76128556 76128557	*TMC6,TMC8*
782	Chr17: 7621545 7621546	*DNAH2*
783	Chr17: 7621575 7621576	*DNAH2*
784	Chr17: 76662056 76662057	*Non Coding Region*
785	Chr17: 80186266 80186267	*SLC16A3*
786	Chr17: 80186273 80186274	*SLC16A3*
787	Chr17: 80186275 80186276	*SLC16A3*
788	Chr17: 80186336 80186337	*SLC16A3*
789	Chr17: 80255419 80255420	*Non Coding Region*
790	Chr17: 80255457 80255458	*Non Coding Region*
791	Chr17: 9143174 9143175	*NTN1*
792	Chr18: 2906032 2906033	*EMILIN2*
793	Chr18: 29265842 29265843	*B4GALT6*
794	Chr18: 3499093 3499094	*DLGAP1*
795	Chr18: 3499253 3499254	*DLGAP1*
796	Chr18: 56976186 56976187	*CPLX4*
797	Chr19: 1074727 1074728	*HMHA1*
798	Chr19: 1132256 1132257	*SBNO2*
799	Chr19: 1132294 1132295	*SBNO2*
800	Chr19: 1132313 1132314	*SBNO2*
801	Chr19: 1132339 1132340	*SBNO2*
802	Chr19: 1132372 1132373	*SBNO2*
803	Chr19: 1132389 1132390	*SBNO2*
804	Chr19: 1132537 1132538	*SBNO2*
805	Chr19: 11532175 11532176	*CCDC151*
806	Chr19: 11533499 11533500	*CCDC151*
807	Chr19: 13124776 13124777	*NFIX*
808	Chr19: 13275288 13275289	*Non Coding Region*

Figure 56I

Order	Coordinate of CpG (hg19)	Gene
809	Chr19: 14168210 14168211	*PALM3*
810	Chr19: 14169966 14169967	*PALM3*
811	Chr19: 14785849 14785850	*EMR3*
812	Chr19: 16181372 16181373	*TPM4*
813	Chr19: 16189360 16189361	*TPM4*
814	Chr19: 16999576 16999577	*F2RL3*
815	Chr19: 16999614 16999615	*F2RL3*
816	Chr19: 17346367 17346368	*NR2F6*
817	Chr19: 17346507 17346508	*NR2F6*
818	Chr19: 17403176 17403177	*ABHD8*
819	Chr19: 17403313 17403314	*ABHD8*
820	Chr19: 17403529 17403530	*ABHD8*
821	Chr19: 17958736 17958737	*JAK3*
822	Chr19: 17958825 17958826	*JAK3*
823	Chr19: 1857004 1857005	*KLF16*
824	Chr19: 3136076 3136077	*GNA15*
825	Chr19: 35786821 35786822	*MAG*
826	Chr19: 36132627 36132628	*ETV2*
827	Chr19: 38183259 38183260	*ZNF781*
828	Chr19: 38183262 38183263	*ZNF781*
829	Chr19: 3822189 3822190	*ZFR2*
830	Chr19: 38701994 38701995	*DPF1*
831	Chr19: 42927074 42927075	*LIPE*
832	Chr19: 50004294 50004295	*MIR150*
833	Chr19: 50004444 50004445	*MIR150*
834	Chr19: 51670432 51670433	*SIGLECP3*
835	Chr19: 908894 908895	*C19orf22*
836	Chr20: 21081901 21081902	*Non Coding Region*
837	Chr20: 23123952 23123953	*Non Coding Region*
838	Chr20: 3053037 3053038	*OXT*
839	Chr20: 30639305 30639306	*HCK*
840	Chr20: 36932553 36932554	*BPI*
841	Chr20: 42544728 42544729	*TOX2*
842	Chr20: 48061990 48061991	*KCNB1*
843	Chr20: 55200146 55200147	*Non Coding Region*
844	Chr20: 56025683 56025684	*Non Coding Region*
845	Chr20: 62369445 62369446	*LIME1*
846	Chr20: 62369462 62369463	*LIME1*
847	Chr21: 36399146 36399147	*RUNX1*
848	Chr21: 36399226 36399227	*RUNX1*
849	Chr21: 36399258 36399259	*RUNX1*
850	Chr21: 38120350 38120351	*SIM2*
851	Chr21: 39878866 39878867	*ERG*
852	Chr21: 42733825 42733826	*MX2*
853	Chr21: 42733894 42733895	*MX2*

Order	Coordinate of CpG (hg19)	Gene
854	Chr21: 42880714 42880715	*TMPRSS2*
855	Chr21: 46340905 46340906	*ITGB2*
856	Chr21: 46898137 46898138	*COL18A1*
857	Chr22: 19711007 19711008	*SEPT5,GP1BB*
858	Chr22: 19753364 19753365	*TBX1*
859	Chr22: 19754251 19754252	*TBX1*
860	Chr22: 19754448 19754449	*TBX1*
861	Chr22: 19879176 19879177	*TXNRD2*
862	Chr22: 37813098 37813099	*ELFN2*
863	Chr22: 38221187 38221188	*GALR3*
864	Chr22: 39493114 39493115	*APOBEC3H*
865	Chr22: 46366859 46366860	*WNT7B*
866	Chr22: 46367617 46367618	*WNT7B*
867	Chr22: 46454012 46454013	*LOC150381*
868	Chr22: 46770195 46770196	*CELSR1*
869	Chr22: 46770253 46770254	*CELSR1*
870	ChrX: 106161451 106161452	*CLDN2*
871	ChrX: 109936239 109936240	*CHRDL1*
	Alternative markers: High in CRC, other cancers	
872	Chr01: 110611465 110611466	*ALX3*
873	Chr01: 111217194 111217195	*KCNA3*
874	Chr01: 111217406 111217407	*KCNA3*
875	Chr01: 111217497 111217498	*KCNA3*
876	Chr01: 111217691 111217692	*KCNA3*
877	Chr01: 111217712 111217713	*KCNA3*
878	Chr01: 114697113 114697114	*SYT6*
879	Chr01: 119532195 119532196	*TBX15*
880	Chr01: 119543336 119543337	*Non Coding Region*
881	Chr01: 119549263 119549264	*Non Coding Region*
882	Chr01: 119550803 119550804	*Non Coding Region*
883	Chr01: 1475675 1475676	*C1orf70*
884	Chr01: 151810893 151810894	*LOC100132111,C2CD4D*
885	Chr01: 154475139 154475140	*TDRD10,SHE*
886	Chr01: 154475173 154475174	*TDRD10,SHE*
887	Chr01: 156130826 156130827	*SEMA4A*
888	Chr01: 1566699 1566700	*MMP23B,MMP23A*
889	Chr01: 169396635 169396636	*C1orf114*
890	Chr01: 169396637 169396638	*C1orf114*
891	Chr01: 169396706 169396707	*C1orf114*
892	Chr01: 169396712 169396713	*C1orf114*
893	Chr01: 169396785 169396786	*C1orf114*
894	Chr01: 169396834 169396835	*C1orf114*
895	Chr01: 169396858 169396859	*C1orf114*
896	Chr01: 169396868 169396869	*C1orf114*
897	Chr01: 170630558 170630559	*Non Coding Region*

Figure 56J

Order	Coordinate of CpG (hg19)	Gene
898	Chr01: 18959268 18959269	*PAX7*
899	Chr01: 190447290 190447291	*FAM5C*
900	Chr01: 200008026 200008027	*NR5A2*
901	Chr01: 207818395 207818396	*CR1L*
902	Chr01: 221050491 221050492	*Non Coding Region*
903	Chr01: 228652338 228652339	*Non Coding Region*
904	Chr01: 228652478 228652479	*Non Coding Region*
905	Chr01: 228652528 228652529	*Non Coding Region*
906	Chr01: 229543112 229543113	*Non Coding Region*
907	Chr01: 229568252 229568253	*ACTA1*
908	Chr01: 237205174 237205175	*RYR2*
909	Chr01: 237205409 237205410	*RYR2*
910	Chr01: 243646318 243646319	*SDCCAG8*
911	Chr01: 2987645 2987646	*PRDM16*
912	Chr01: 36043352 36043353	*TFAP2E*
913	Chr01: 44031300 44031301	*PTPRF*
914	Chr01: 44873064 44873065	*RNF220*
915	Chr01: 44873229 44873230	*RNF220*
916	Chr01: 4714974 4714975	*AJAP1*
917	Chr01: 47911449 47911450	*Non Coding Region*
918	Chr01: 47911508 47911509	*Non Coding Region*
919	Chr01: 48058985 48058986	*Non Coding Region*
920	Chr01: 48059173 48059174	*Non Coding Region*
921	Chr01: 50882006 50882007	*Non Coding Region*
922	Chr01: 50884480 50884481	*DMRTA2*
923	Chr01: 50886782 50886783	*DMRTA2*
924	Chr01: 50886949 50886950	*DMRTA2*
925	Chr01: 58715539 58715540	*DAB1*
926	Chr01: 63785544 63785545	*Non Coding Region*
927	Chr01: 6526325 6526326	*TNFRSF25,PLEKHG5*
928	Chr01: 67217853 67217854	*TCTEX1D1*
929	Chr01: 67217886 67217887	*TCTEX1D1*
930	Chr01: 67218135 67218136	*TCTEX1D1*
931	Chr01: 67218165 67218166	*TCTEX1D1*
932	Chr01: 77334207 77334208	*ST6GALNAC5*
933	Chr01: 91183051 91183052	*BARHL2*
934	Chr01: 91300215 91300216	*Non Coding Region*
935	Chr01: 9712928 9712929	*C1orf200,PIK3CD*
936	Chr01: 98511258 98511259	*Non Coding Region*
937	Chr01: 98511523 98511524	*Non Coding Region*
938	Chr01: 98511789 98511790	*MIR137*
939	Chr01: 99470129 99470130	*LPPR5*
940	Chr02: 105459164 105459165	*Non Coding Region*
941	Chr02: 105461096 105461097	*Non Coding Region*
942	Chr02: 119591144 119591145	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
943	Chr02: 119599748 119599749	*EN1*
944	Chr02: 119600002 119600003	*EN1*
945	Chr02: 127413831 127413832	*GYPC*
946	Chr02: 127414108 127414109	*GYPC*
947	Chr02: 127414381 127414382	*GYPC*
948	Chr02: 131721692 131721693	*ARHGEF4*
949	Chr02: 139537197 139537198	*NXPH2*
950	Chr02: 171678954 171678955	*GAD1*
951	Chr02: 172972840 172972841	*Non Coding Region*
952	Chr02: 176956678 176956679	*HOXD13*
953	Chr02: 177030150 177030151	*HOXD3*
954	Chr02: 182321489 182321490	*ITGA4*
955	Chr02: 182321496 182321497	*ITGA4*
956	Chr02: 185462408 185462409	*ZNF804A*
957	Chr02: 206551364 206551365	*NRP2*
958	Chr02: 208989248 208989249	*CRYGD*
959	Chr02: 208989382 208989383	*CRYGD*
960	Chr02: 220299740 220299741	*SPEG*
961	Chr02: 220361467 220361468	*Non Coding Region*
962	Chr02: 220361609 220361610	*Non Coding Region*
963	Chr02: 222435214 222435215	*EPHA4*
964	Chr02: 223161771 223161772	*CCDC140,PAX3*
965	Chr02: 223162049 223162050	*CCDC140,PAX3*
966	Chr02: 228736253 228736254	*WDR69*
967	Chr02: 233499070 233499071	*EFHD1*
968	Chr02: 237416338 237416339	*IQCA1*
969	Chr02: 238864632 238864633	*Non Coding Region*
970	Chr02: 239140318 239140319	*LOC643387,LOC151174*
971	Chr02: 242743588 242743589	*GAL3ST2*
972	Chr02: 25439110 25439111	*Non Coding Region*
973	Chr02: 45029004 45029005	*Non Coding Region*
974	Chr02: 45155991 45155992	*Non Coding Region*
975	Chr02: 45231382 45231383	*Non Coding Region*
976	Chr02: 468179 468180	*Non Coding Region*
977	Chr02: 63281139 63281140	*OTX1*
978	Chr02: 63283967 63283968	*OTX1*
979	Chr02: 66667101 66667102	*MEIS1*
980	Chr02: 66667433 66667434	*MEIS1*
981	Chr02: 66809923 66809924	*Non Coding Region*
982	Chr02: 66810606 66810607	*Non Coding Region*
983	Chr02: 74743437 74743438	*TLX2*
984	Chr02: 86263270 86263271	*POLR1A*
985	Chr02: 97193289 97193290	*Non Coding Region*
986	Chr03: 119528956 119528957	*NR1I2*
987	Chr03: 119529158 119529159	*NR1I2*

Figure 56K

Order	Coordinate of CpG (hg19)	Gene
988	Chr03: 121903520 121903521	*CASR*
989	Chr03: 128202176 128202177	*GATA2*
990	Chr03: 128720884 128720885	*CCDC48*
991	Chr03: 129693385 129693386	*TRH*
992	Chr03: 129693489 129693490	*TRH*
993	Chr03: 129693586 129693587	*TRH*
994	Chr03: 129694057 129694058	*TRH*
995	Chr03: 129694487 129694488	*TRH*
996	Chr03: 133393454 133393455	*Non Coding Region*
997	Chr03: 138679512 138679513	*Non Coding Region*
998	Chr03: 138679578 138679579	*Non Coding Region*
999	Chr03: 139258939 139258940	*RBP1*
1000	Chr03: 141516271 141516272	*GRK7*
1001	Chr03: 141516291 141516292	*GRK7*
1002	Chr03: 169378764 169378765	*MECOM*
1003	Chr03: 172165884 172165885	*GHSR*
1004	Chr03: 172166517 172166518	*GHSR*
1005	Chr03: 179168677 179168678	*GNB4*
1006	Chr03: 179168760 179168761	*GNB4*
1007	Chr03: 184301468 184301469	*Non Coding Region*
1008	Chr03: 184301730 184301731	*Non Coding Region*
1009	Chr03: 185080816 185080817	*MAP3K13*
1010	Chr03: 192126165 192126166	*FGF12*
1011	Chr03: 194408845 194408846	*FAM43A*
1012	Chr03: 27765232 27765233	*EOMES*
1013	Chr03: 27765409 27765410	*Non Coding Region*
1014	Chr03: 32859377 32859378	*TRIM71*
1015	Chr03: 38035642 38035643	*VILL*
1016	Chr03: 44063593 44063594	*Non Coding Region*
1017	Chr03: 44063832 44063833	*Non Coding Region*
1018	Chr03: 62356962 62356963	*FEZF2*
1019	Chr04: 10459219 10459220	*ZNF518B*
1020	Chr04: 111555440 111555441	*PITX2*
1021	Chr04: 13524143 13524144	*Non Coding Region*
1022	Chr04: 13530269 13530270	*Non Coding Region*
1023	Chr04: 13532951 13532952	*Non Coding Region*
1024	Chr04: 13540946 13540947	*Non Coding Region*
1025	Chr04: 13543985 13543986	*NKX3-2*
1026	Chr04: 1396593 1396594	*Non Coding Region*
1027	Chr04: 140656749 140656750	*MAML3*
1028	Chr04: 140656794 140656795	*MAML3*
1029	Chr04: 140656926 140656927	*MAML3*
1030	Chr04: 147561775 147561776	*POU4F2*
1031	Chr04: 147569938 147569939	*Non Coding Region*
1032	Chr04: 155254497 155254498	*DCHS2*

Order	Coordinate of CpG (hg19)	Gene
1033	Chr04: 155254602 155254603	*DCHS2*
1034	Chr04: 169799087 169799088	*PALLD*
1035	Chr04: 172734276 172734277	*GALNTL6*
1036	Chr04: 174430614 174430615	*Non Coding Region*
1037	Chr04: 174450408 174450409	*NBLA00301,HAND2*
1038	Chr04: 175133151 175133152	*Non Coding Region*
1039	Chr04: 42153708 42153709	*BEND4*
1040	Chr04: 4870486 4870487	*Non Coding Region*
1041	Chr04: 57522493 57522494	*HOPX*
1042	Chr04: 57522632 57522633	*HOPX*
1043	Chr04: 57522801 57522802	*HOPX*
1044	Chr04: 779568 779569	*CPLX1*
1045	Chr04: 779691 779692	*CPLX1*
1046	Chr04: 81189927 81189928	*FGF5*
1047	Chr05: 100239919 100239920	*ST8SIA4*
1048	Chr05: 100240059 100240060	*ST8SIA4*
1049	Chr05: 110406506 110406507	*TSLP*
1050	Chr05: 115152413 115152414	*CDO1*
1051	Chr05: 115152431 115152432	*CDO1*
1052	Chr05: 115152485 115152486	*CDO1*
1053	Chr05: 115152492 115152493	*CDO1*
1054	Chr05: 115152494 115152495	*CDO1*
1055	Chr05: 11904114 11904115	*CTNND2*
1056	Chr05: 134363324 134363325	*Non Coding Region*
1057	Chr05: 134825791 134825792	*Non Coding Region*
1058	Chr05: 135266135 135266136	*FBXL21*
1059	Chr05: 137225191 137225192	*PKD2L2*
1060	Chr05: 140787504 140787505	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
1061	Chr05: 140787507 140787508	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
1062	Chr05: 140797172 140797173	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
1063	Chr05: 140797234 140797235	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
1064	Chr05: 140797280 140797281	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
1065	Chr05: 140800586 140800587	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
1066	Chr05: 140800929 140800930	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
1067	Chr05: 140864733 140864734	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
1068	Chr05: 145720024 145720025	*POU4F3*
1069	Chr05: 149535650 149535651	*PDGFRB*
1070	Chr05: 151066484 151066485	*SPARC*
1071	Chr05: 151066486 151066487	*SPARC*
1072	Chr05: 158532401 158532402	*Non Coding Region*
1073	Chr05: 16180055 16180056	*MARCHF11*
1074	Chr05: 169064024 169064025	*DOCK2*
1075	Chr05: 169064451 169064452	*DOCK2*
1076	Chr05: 170740870 170740871	*Non Coding Region*
1077	Chr05: 172659730 172659731	*NKX2-5*

Figure 56L

Order	Coordinate of CpG (hg19)	Gene
1078	Chr05: 176107427 176107428	*Non Coding Region*
1079	Chr05: 177412718 177412719	*Non Coding Region*
1080	Chr05: 178594097 178594098	*ADAMTS2*
1081	Chr05: 179220795 179220796	*LTC4S*
1082	Chr05: 1875886 1875887	*Non Coding Region*
1083	Chr05: 1876168 1876169	*Non Coding Region*
1084	Chr05: 1876337 1876338	*Non Coding Region*
1085	Chr05: 2756113 2756114	*Non Coding Region*
1086	Chr05: 2757244 2757245	*Non Coding Region*
1087	Chr05: 3596704 3596705	*IRX1*
1088	Chr05: 3597487 3597488	*IRX1*
1089	Chr05: 37834672 37834673	*GDNF*
1090	Chr05: 37834958 37834959	*GDNF*
1091	Chr05: 42994709 42994710	*Non Coding Region*
1092	Chr05: 42994776 42994777	*Non Coding Region*
1093	Chr05: 528580 528581	*Non Coding Region*
1094	Chr05: 528621 528622	*Non Coding Region*
1095	Chr05: 54516487 54516488	*Non Coding Region*
1096	Chr05: 54516805 54516806	*Non Coding Region*
1097	Chr05: 72528111 72528112	*Non Coding Region*
1098	Chr05: 72594461 72594462	*Non Coding Region*
1099	Chr05: 72595685 72595686	*Non Coding Region*
1100	Chr05: 72595741 72595742	*Non Coding Region*
1101	Chr05: 87967845 87967846	*LOC645323*
1102	Chr06: 100912940 100912941	*SIM1*
1103	Chr06: 100912946 100912947	*SIM1*
1104	Chr06: 10382828 10382829	*Non Coding Region*
1105	Chr06: 10391412 10391413	*Non Coding Region*
1106	Chr06: 105389599 105389600	*Non Coding Region*
1107	Chr06: 105584551 105584552	*BVES*
1108	Chr06: 106429316 106429317	*Non Coding Region*
1109	Chr06: 106429443 106429444	*Non Coding Region*
1110	Chr06: 108488335 108488336	*NR2E1*
1111	Chr06: 108495678 108495679	*NR2E1*
1112	Chr06: 10881925 10881926	*GCM2*
1113	Chr06: 10882240 10882241	*GCM2*
1114	Chr06: 10882336 10882337	*GCM2*
1115	Chr06: 134210307 134210308	*TCF21*
1116	Chr06: 137809529 137809530	*Non Coding Region*
1117	Chr06: 137814728 137814729	*OLIG3*
1118	Chr06: 1379694 1379695	*Non Coding Region*
1119	Chr06: 1384591 1384592	*Non Coding Region*
1120	Chr06: 150286508 150286509	*ULBP1*
1121	Chr06: 152128471 152128472	*ESR1*
1122	Chr06: 152128865 152128866	*ESR1*

Order	Coordinate of CpG (hg19)	Gene
1123	Chr06: 152129388 152129389	*ESR1*
1124	Chr06: 159590059 159590060	*FNDC1*
1125	Chr06: 1624978 1624979	*GMDS*
1126	Chr06: 1625111 1625112	*GMDS*
1127	Chr06: 166582393 166582394	*T*
1128	Chr06: 26240528 26240529	*HIST1H4F*
1129	Chr06: 26240579 26240580	*HIST1H4F*
1130	Chr06: 26240782 26240783	*HIST1H4F*
1131	Chr06: 26240920 26240921	*HIST1H4F*
1132	Chr06: 26240939 26240940	*HIST1H4F*
1133	Chr06: 26745662 26745663	*Non Coding Region*
1134	Chr06: 27173633 27173634	*Non Coding Region*
1135	Chr06: 27173753 27173754	*Non Coding Region*
1136	Chr06: 27173991 27173992	*Non Coding Region*
1137	Chr06: 27463217 27463218	*Non Coding Region*
1138	Chr06: 27463347 27463348	*Non Coding Region*
1139	Chr06: 27513092 27513093	*Non Coding Region*
1140	Chr06: 27513479 27513480	*Non Coding Region*
1141	Chr06: 27647713 27647714	*Non Coding Region*
1142	Chr06: 28411287 28411288	*ZSCAN23*
1143	Chr06: 28458175 28458176	*Non Coding Region*
1144	Chr06: 28603230 28603231	*Non Coding Region*
1145	Chr06: 28603779 28603780	*Non Coding Region*
1146	Chr06: 28778226 28778227	*Non Coding Region*
1147	Chr06: 28956247 28956248	*Non Coding Region*
1148	Chr06: 28956268 28956269	*Non Coding Region*
1149	Chr06: 28956731 28956732	*Non Coding Region*
1150	Chr06: 29521143 29521144	*Non Coding Region*
1151	Chr06: 29521162 29521163	*Non Coding Region*
1152	Chr06: 29521568 29521569	*Non Coding Region*
1153	Chr06: 29521602 29521603	*Non Coding Region*
1154	Chr06: 29521695 29521696	*Non Coding Region*
1155	Chr06: 29521751 29521752	*Non Coding Region*
1156	Chr06: 29521756 29521757	*Non Coding Region*
1157	Chr06: 29760164 29760165	*HCG4*
1158	Chr06: 29795815 29795816	*HLA-G*
1159	Chr06: 29943414 29943415	*HCG9*
1160	Chr06: 29943480 29943481	*HCG9*
1161	Chr06: 30095728 30095729	*Non Coding Region*
1162	Chr06: 30139634 30139635	*TRIM15*
1163	Chr06: 30139641 30139642	*TRIM15*
1164	Chr06: 30139880 30139881	*TRIM15*
1165	Chr06: 30139903 30139904	*TRIM15*
1166	Chr06: 30140018 30140019	*TRIM15*
1167	Chr06: 32116905 32116906	*PRRT1*

Figure 56M

Order	Coordinate of CpG (hg19)	Gene
1168	Chr06: 35108921 35108922	*TCP11*
1169	Chr06: 391764 391765	*IRF4*
1170	Chr06: 391936 391937	*IRF4*
1171	Chr06: 41341601 41341602	*Non Coding Region*
1172	Chr06: 42738967 42738968	*Non Coding Region*
1173	Chr06: 42739021 42739022	*Non Coding Region*
1174	Chr06: 43253097 43253098	*TTBK1*
1175	Chr06: 50692252 50692253	*TFAP2D*
1176	Chr06: 5994956 5994957	*Non Coding Region*
1177	Chr06: 5994982 5994983	*Non Coding Region*
1178	Chr06: 78172337 78172338	*HTR1B*
1179	Chr06: 85474028 85474029	*TBX18*
1180	Chr07: 100073936 100073937	*TSC22D4*
1181	Chr07: 100488942 100488943	*ACHE*
1182	Chr07: 1270558 1270559	*Non Coding Region*
1183	Chr07: 127672458 127672459	*LRRC4,SND1*
1184	Chr07: 127672473 127672474	*LRRC4,SND1*
1185	Chr07: 127672658 127672659	*SND1*
1186	Chr07: 143579951 143579952	*FAM115A*
1187	Chr07: 143580147 143580148	*FAM115A*
1188	Chr07: 149487338 149487339	*SSPO*
1189	Chr07: 149917263 149917264	*Non Coding Region*
1190	Chr07: 155167038 155167039	*Non Coding Region*
1191	Chr07: 156797242 156797243	*Non Coding Region*
1192	Chr07: 156798258 156798259	*MNX1*
1193	Chr07: 157481628 157481629	*PTPRN2*
1194	Chr07: 158937494 158937495	*VIPR2*
1195	Chr07: 19146032 19146033	*Non Coding Region*
1196	Chr07: 19158647 19158648	*TWIST1*
1197	Chr07: 19158664 19158665	*TWIST1*
1198	Chr07: 19158692 19158693	*TWIST1*
1199	Chr07: 19158747 19158748	*TWIST1*
1200	Chr07: 19183732 19183733	*Non Coding Region*
1201	Chr07: 24323799 24323800	*NPY*
1202	Chr07: 24323840 24323841	*NPY*
1203	Chr07: 27286075 27286076	*EVX1*
1204	Chr07: 30722320 30722321	*CRHR2*
1205	Chr07: 30722327 30722328	*CRHR2*
1206	Chr07: 35293759 35293760	*TBX20*
1207	Chr07: 35296968 35296969	*Non Coding Region*
1208	Chr07: 37488253 37488254	*ELMO1*
1209	Chr07: 42267719 42267720	*GLI3*
1210	Chr07: 42267747 42267748	*GLI3*
1211	Chr07: 49813033 49813034	*VWC2*
1212	Chr07: 49813065 49813066	*VWC2*

Order	Coordinate of CpG (hg19)	Gene
1213	Chr07: 54612475 54612476	*VSTM2A*
1214	Chr07: 6703803 6703804	*Non Coding Region*
1215	Chr07: 70597058 70597059	*WBSCR17*
1216	Chr07: 8482614 8482615	*NXPH1*
1217	Chr07: 96650323 96650324	*DLX5*
1218	Chr07: 96650668 96650669	*DLX5*
1219	Chr07: 96654788 96654789	*DLX5*
1220	Chr07: 98246001 98246002	*NPTX2*
1221	Chr08: 104512858 104512859	*RIMS2*
1222	Chr08: 11561724 11561725	*GATA4*
1223	Chr08: 116660586 116660587	*TRPS1*
1224	Chr08: 116679763 116679764	*TRPS1*
1225	Chr08: 132054555 132054556	*Non Coding Region*
1226	Chr08: 140715540 140715541	*KCNK9*
1227	Chr08: 143545789 143545790	*BAI1*
1228	Chr08: 144650730 144650731	*C8orf73*
1229	Chr08: 23563925 23563926	*NKX2-6*
1230	Chr08: 23564025 23564026	*NKX2-6*
1231	Chr08: 23564156 23564157	*NKX2-6*
1232	Chr08: 23564193 23564194	*NKX2-6*
1233	Chr08: 23564591 23564592	*NKX2-6*
1234	Chr08: 23567310 23567311	*Non Coding Region*
1235	Chr08: 24771125 24771126	*NEFM*
1236	Chr08: 38586066 38586067	*TACC1*
1237	Chr08: 49468828 49468829	*Non Coding Region*
1238	Chr08: 53851156 53851157	*NPBWR1*
1239	Chr08: 53852030 53852031	*NPBWR1*
1240	Chr08: 55367627 55367628	*Non Coding Region*
1241	Chr08: 55379248 55379249	*Non Coding Region*
1242	Chr08: 56852290 56852291	*LYN*
1243	Chr08: 57025662 57025663	*MOS*
1244	Chr08: 57069907 57069908	*Non Coding Region*
1245	Chr08: 57070013 57070014	*Non Coding Region*
1246	Chr08: 65281496 65281497	*Non Coding Region*
1247	Chr08: 65283263 65283264	*Non Coding Region*
1248	Chr08: 65291513 65291514	*MIR124-2*
1249	Chr08: 67873799 67873800	*Non Coding Region*
1250	Chr08: 70982039 70982040	*PRDM14*
1251	Chr08: 72469507 72469508	*Non Coding Region*
1252	Chr08: 72754469 72754470	*MSC*
1253	Chr08: 72754953 72754954	*MSC*
1254	Chr08: 72917350 72917351	*Non Coding Region*
1255	Chr08: 77595321 77595322	*ZFHX4,LOC100192378*
1256	Chr08: 85097056 85097057	*RALYL*
1257	Chr08: 86350889 86350890	*CA3*

Figure 56N

Order	Coordinate of CpG (hg19)	Gene
1258	Chr08: 86350919 86350920	*CA3*
1259	Chr08: 97157856 97157857	*GDF6*
1260	Chr08: 97170666 97170667	*GDF6*
1261	Chr08: 97171827 97171828	*GDF6*
1262	Chr08: 9760869 9760870	*LOC157627*
1263	Chr08: 9760877 9760878	*LOC157627*
1264	Chr08: 9760880 9760881	*LOC157627*
1265	Chr08: 98290310 98290311	*TSPYL5*
1266	Chr09: 100611516 100611517	*Non Coding Region*
1267	Chr09: 117150495 117150496	*AKNA*
1268	Chr09: 124461599 124461600	*DAB2IP*
1269	Chr09: 124461640 124461641	*DAB2IP*
1270	Chr09: 124461998 124461999	*DAB2IP*
1271	Chr09: 124462172 124462173	*DAB2IP*
1272	Chr09: 124462182 124462183	*DAB2IP*
1273	Chr09: 126776180 126776181	*LHX2*
1274	Chr09: 37002685 37002686	*PAX5*
1275	Chr09: 79633737 79633738	*FOXB2*
1276	Chr09: 91849710 91849711	*Non Coding Region*
1277	Chr10: 100993583 100993584	*HPSE2*
1278	Chr10: 100993587 100993588	*HPSE2*
1279	Chr10: 100993589 100993590	*HPSE2*
1280	Chr10: 100993597 100993598	*HPSE2*
1281	Chr10: 102419245 102419246	*Non Coding Region*
1282	Chr10: 102497354 102497355	*Non Coding Region*
1283	Chr10: 102893980 102893981	*TLX1*
1284	Chr10: 102894120 102894121	*TLX1*
1285	Chr10: 102894148 102894149	*TLX1*
1286	Chr10: 102899285 102899286	*Non Coding Region*
1287	Chr10: 102900130 102900131	*Non Coding Region*
1288	Chr10: 102900365 102900366	*Non Coding Region*
1289	Chr10: 102900491 102900492	*Non Coding Region*
1290	Chr10: 103603722 103603723	*KCNIP2*
1291	Chr10: 103603812 103603813	*KCNIP2*
1292	Chr10: 105037503 105037504	*INA*
1293	Chr10: 105452352 105452353	*SH3PXD2A*
1294	Chr10: 106398816 106398817	*Non Coding Region*
1295	Chr10: 106400565 106400566	*SORCS3*
1296	Chr10: 106400667 106400668	*SORCS3*
1297	Chr10: 106400686 106400687	*SORCS3*
1298	Chr10: 106400702 106400703	*SORCS3*
1299	Chr10: 108924366 108924367	*SORCS1*
1300	Chr10: 11207527 11207528	*CUGBP2*
1301	Chr10: 118030970 118030971	*GFRA1*
1302	Chr10: 118892581 118892582	*VAX1*

Order	Coordinate of CpG (hg19)	Gene
1303	Chr10: 123923081 123923082	*TACC2*
1304	Chr10: 131758786 131758787	*EBF3*
1305	Chr10: 131768928 131768929	*Non Coding Region*
1306	Chr10: 133110244 133110245	*TCERG1L*
1307	Chr10: 133110349 133110350	*TCERG1L*
1308	Chr10: 15761854 15761855	*ITGA8*
1309	Chr10: 22542024 22542025	*Non Coding Region*
1310	Chr10: 22542145 22542146	*Non Coding Region*
1311	Chr10: 22634432 22634433	*SPAG6*
1312	Chr10: 22634439 22634440	*SPAG6*
1313	Chr10: 26727318 26727319	*APBB1IP*
1314	Chr10: 44788812 44788813	*Non Coding Region*
1315	Chr10: 50605613 50605614	*Non Coding Region*
1316	Chr10: 57389779 57389780	*Non Coding Region*
1317	Chr10: 57390306 57390307	*Non Coding Region*
1318	Chr10: 57390704 57390705	*Non Coding Region*
1319	Chr10: 8097331 8097332	*GATA3*
1320	Chr10: 8097689 8097690	*GATA3*
1321	Chr10: 8098005 8098006	*GATA3*
1322	Chr10: 81001957 81001958	*ZMIZ1*
1323	Chr10: 81002218 81002219	*ZMIZ1*
1324	Chr10: 81002480 81002481	*ZMIZ1*
1325	Chr10: 81002615 81002616	*ZMIZ1*
1326	Chr10: 88123453 88123454	*GRID1*
1327	Chr10: 94822908 94822909	*CYP26C1*
1328	Chr11: 105481317 105481318	*GRIA4*
1329	Chr11: 105481319 105481320	*GRIA4*
1330	Chr11: 105481322 105481323	*GRIA4*
1331	Chr11: 105481406 105481407	*GRIA4*
1332	Chr11: 115531033 115531034	*Non Coding Region*
1333	Chr11: 119293863 119293864	*THY1*
1334	Chr11: 119293869 119293870	*THY1*
1335	Chr11: 123066560 123066561	*ASAM*
1336	Chr11: 123301171 123301172	*Non Coding Region*
1337	Chr11: 123301490 123301491	*Non Coding Region*
1338	Chr11: 124790875 124790876	*HEPACAM*
1339	Chr11: 125036385 125036386	*PKNOX2*
1340	Chr11: 125774082 125774083	*PUS3,DDX25*
1341	Chr11: 133906693 133906694	*LOC100128239*
1342	Chr11: 14926738 14926739	*Non Coding Region*
1343	Chr11: 20229517 20229518	*Non Coding Region*
1344	Chr11: 20618399 20618400	*Non Coding Region*
1345	Chr11: 2162183 2162184	*IGF2,IGF2AS,INS-IGF2*
1346	Chr11: 31821388 31821389	*PAX6*
1347	Chr11: 31827084 31827085	*PAX6*

Figure 47O

Order	Coordinate of CpG (hg19)	Gene
1348	Chr11: 31848649 31848650	*Non Coding Region*
1349	Chr11: 32456912 32456913	*WT1,WIT1*
1350	Chr11: 32459428 32459429	*WIT1*
1351	Chr11: 32459626 32459627	*WIT1*
1352	Chr11: 43602845 43602846	*MIR129-2*
1353	Chr11: 43602847 43602848	*MIR129-2*
1354	Chr11: 43602857 43602858	*MIR129-2*
1355	Chr11: 43602879 43602880	*MIR129-2*
1356	Chr11: 43602920 43602921	*MIR129-2*
1357	Chr11: 43602965 43602966	*MIR129-2*
1358	Chr11: 44332653 44332654	*ALX4*
1359	Chr11: 44332664 44332665	*ALX4*
1360	Chr11: 57250469 57250470	*Non Coding Region*
1361	Chr11: 61322922 61322923	*SYT7*
1362	Chr11: 637170 637171	*DRD4*
1363	Chr11: 637173 637174	*DRD4*
1364	Chr11: 637175 637176	*DRD4*
1365	Chr11: 63768032 63768033	*MACROD1*
1366	Chr11: 63828346 63828347	*MACROD1*
1367	Chr11: 69704700 69704701	*Non Coding Region*
1368	Chr11: 71955322 71955323	*PHOX2A*
1369	Chr11: 71955332 71955333	*PHOX2A*
1370	Chr11: 71955599 71955600	*PHOX2A*
1371	Chr11: 78673073 78673074	*ODZ4*
1372	Chr11: 79152821 79152822	*ODZ4*
1373	Chr11: 830320 830321	*EFCAB4A*
1374	Chr12: 113541915 113541916	*RASAL1*
1375	Chr12: 114843296 114843297	*TBX5*
1376	Chr12: 114847438 114847439	*TBX5*
1377	Chr12: 115102726 115102727	*Non Coding Region*
1378	Chr12: 115103351 115103352	*Non Coding Region*
1379	Chr12: 120799725 120799726	*MSI1*
1380	Chr12: 133481390 133481391	*Non Coding Region*
1381	Chr12: 25055967 25055968	*BCAT1*
1382	Chr12: 5019933 5019934	*KCNA1*
1383	Chr12: 50297581 50297582	*FAIM2*
1384	Chr12: 50297588 50297589	*FAIM2*
1385	Chr12: 50297777 50297778	*FAIM2*
1386	Chr12: 50297793 50297794	*FAIM2*
1387	Chr12: 50297945 50297946	*FAIM2*
1388	Chr12: 52401214 52401215	*GRASP*
1389	Chr12: 52627047 52627048	*KRT7*
1390	Chr12: 54088972 54088973	*Non Coding Region*
1391	Chr12: 54345862 54345863	*Non Coding Region*
1392	Chr12: 54428171 54428172	*HOXC5,HOXC4*

Order	Coordinate of CpG (hg19)	Gene
1393	Chr12: 54447584 54447585	*HOXC4*
1394	Chr12: 54447873 54447874	*HOXC4*
1395	Chr12: 54812000 54812001	*ITGA5*
1396	Chr12: 54812085 54812086	*ITGA5*
1397	Chr12: 58021917 58021918	*B4GALNT1*
1398	Chr12: 62585031 62585032	*FAM19A2*
1399	Chr12: 62585204 62585205	*FAM19A2*
1400	Chr12: 62585877 62585878	*FAM19A2*
1401	Chr12: 65515470 65515471	*WIF1*
1402	Chr12: 66275988 66275989	*HMGA2*
1403	Chr12: 85667353 85667354	*Non Coding Region*
1404	Chr12: 95941988 95941989	*USP44*
1405	Chr13: 109149011 109149012	*Non Coding Region*
1406	Chr13: 112758491 112758492	*Non Coding Region*
1407	Chr13: 21520468 21520469	*Non Coding Region*
1408	Chr13: 25320133 25320134	*Non Coding Region*
1409	Chr13: 25745784 25745785	*FAM123A*
1410	Chr13: 28498384 28498385	*PDX1*
1411	Chr13: 28500882 28500883	*Non Coding Region*
1412	Chr13: 28503373 28503374	*Non Coding Region*
1413	Chr13: 29107069 29107070	*Non Coding Region*
1414	Chr13: 53312966 53312967	*LECT1*
1415	Chr13: 53313174 53313175	*LECT1*
1416	Chr13: 53313529 53313530	*LECT1*
1417	Chr13: 53420386 53420387	*PCDH8*
1418	Chr13: 53422777 53422778	*PCDH8*
1419	Chr13: 53422780 53422781	*PCDH8*
1420	Chr13: 58204224 58204225	*Non Coding Region*
1421	Chr13: 79170146 79170147	*Non Coding Region*
1422	Chr13: 79170303 79170304	*Non Coding Region*
1423	Chr13: 79171053 79171054	*Non Coding Region*
1424	Chr13: 84456171 84456172	*SLITRK1*
1425	Chr13: 84456258 84456259	*SLITRK1*
1426	Chr13: 96204856 96204857	*CLDN10*
1427	Chr13: 96205184 96205185	*CLDN10*
1428	Chr14: 101924596 101924597	*Non Coding Region*
1429	Chr14: 102554977 102554978	*HSP90AA1*
1430	Chr14: 23834891 23834892	*EFS*
1431	Chr14: 23834977 23834978	*EFS*
1432	Chr14: 24045549 24045550	*JPH4*
1433	Chr14: 24803873 24803874	*ADCY4*
1434	Chr14: 24803917 24803918	*ADCY4*
1435	Chr14: 24803925 24803926	*ADCY4*
1436	Chr14: 24804339 24804340	*ADCY4*
1437	Chr14: 29254530 29254531	*C14orf23*

Figure 56P

Order	Coordinate of CpG (hg19)	Gene
1438	Chr14:_29254680_29254681	*C14orf23*
1439	Chr14:_38067622_38067623	*Non Coding Region*
1440	Chr14:_48145108_48145109	*MDGA2*
1441	Chr14:_52536436_52536437	*NID2*
1442	Chr14:_54687160_54687161	*Non Coding Region*
1443	Chr14:_57264287_57264288	*Non Coding Region*
1444	Chr14:_60952196_60952197	*C14orf39*
1445	Chr14:_60952945_60952946	*C14orf39*
1446	Chr14:_60974039_60974040	*Non Coding Region*
1447	Chr14:_60975964_60975965	*SIX6*
1448	Chr14:_60976285_60976286	*SIX6*
1449	Chr14:_60977300_60977301	*SIX6*
1450	Chr14:_60978161_60978162	*SIX6*
1451	Chr14:_70014873_70014874	*Non Coding Region*
1452	Chr14:_70653719_70653720	*SLC8A3*
1453	Chr14:_70653919_70653920	*SLC8A3*
1454	Chr14:_70653964_70653965	*SLC8A3*
1455	Chr14:_95235125_95235126	*GSC*
1456	Chr15:_101420636_101420637	*ALDH1A3*
1457	Chr15:_29077493_29077494	*Non Coding Region*
1458	Chr15:_30517589_30517590	*Non Coding Region*
1459	Chr15:_37390284_37390285	*MEIS2*
1460	Chr15:_41794747_41794748	*ITPKA*
1461	Chr15:_51385891_51385892	*TNFAIP8L3*
1462	Chr15:_53083532_53083533	*ONECUT1*
1463	Chr15:_65688530_65688531	*IGDCC4*
1464	Chr15:_68122139_68122140	*LBXCOR1*
1465	Chr15:_83953744_83953745	*BNC1*
1466	Chr15:_83953780_83953781	*BNC1*
1467	Chr15:_83953880_83953881	*BNC1*
1468	Chr15:_86233236_86233237	*AKAP13*
1469	Chr15:_89922328_89922329	*LOC254559*
1470	Chr15:_89952499_89952500	*Non Coding Region*
1471	Chr16:_2041512_2041513	*SYNGR3*
1472	Chr16:_21294947_21294948	*CRYM*
1473	Chr16:_2765843_2765844	*PRSS27*
1474	Chr16:_30904573_30904574	*BCL7C,MIR762*
1475	Chr16:_31580299_31580300	*CSDAP1*
1476	Chr16:_3237991_3237992	*Non Coding Region*
1477	Chr16:_49312033_49312034	*Non Coding Region*
1478	Chr16:_49312238_49312239	*CBLN1*
1479	Chr16:_50699167_50699168	*Non Coding Region*
1480	Chr16:_50699197_50699198	*Non Coding Region*
1481	Chr16:_51184379_51184380	*SALL1*
1482	Chr16:_55363058_55363059	*IRX6*

Order	Coordinate of CpG (hg19)	Gene
1483	Chr16:_55512822_55512823	*MMP2*
1484	Chr16:_55690381_55690382	*SLC6A2*
1485	Chr16:_66612955_66612956	*CMTM2,CMTM1*
1486	Chr16:_66613278_66613279	*CMTM2*
1487	Chr16:_86541480_86541481	*Non Coding Region*
1488	Chr16:_86600326_86600327	*FOXC2*
1489	Chr16:_86613067_86613068	*FOXL1*
1490	Chr16:_86963079_86963080	*Non Coding Region*
1491	Chr17:_17626790_17626791	*RAI1*
1492	Chr17:_17627194_17627195	*RAI1*
1493	Chr17:_19881632_19881633	*AKAP10*
1494	Chr17:_26699169_26699170	*SARM1*
1495	Chr17:_27038861_27038862	*PROCA1*
1496	Chr17:_27038886_27038887	*PROCA1*
1497	Chr17:_27220101_27220102	*FLOT2*
1498	Chr17:_27940509_27940510	*ANKRD13B*
1499	Chr17:_32484259_32484260	*ACCN1*
1500	Chr17:_36734911_36734912	*SRCIN1*
1501	Chr17:_37321490_37321491	*ARL5C*
1502	Chr17:_38347603_38347604	*RAPGEFL1*
1503	Chr17:_38347816_38347817	*RAPGEFL1*
1504	Chr17:_40825980_40825981	*PLEKHH3*
1505	Chr17:_43339223_43339224	*C17orf46,LOC100133991*
1506	Chr17:_43339328_43339329	*C17orf46,LOC100133991*
1507	Chr17:_43339497_43339498	*C17orf46,LOC100133991*
1508	Chr17:_43339512_43339513	*C17orf46,LOC100133991*
1509	Chr17:_46802871_46802872	*HOXB13*
1510	Chr17:_46802888_46802889	*HOXB13*
1511	Chr17:_59529618_59529619	*Non Coding Region*
1512	Chr17:_59534998_59534999	*TBX4*
1513	Chr17:_61778366_61778367	*LIMD2*
1514	Chr17:_70216332_70216333	*Non Coding Region*
1515	Chr17:_7287740_7287741	*TNK1*
1516	Chr17:_73584068_73584069	*MYO15B*
1517	Chr17:_73584070_73584071	*MYO15B*
1518	Chr17:_73584108_73584109	*MYO15B*
1519	Chr17:_73584360_73584361	*MYO15B*
1520	Chr17:_73584617_73584618	*MYO15B*
1521	Chr17:_75369091_75369092	*SEPTIN9*
1522	Chr17:_75369219_75369220	*SEPTIN9*
1523	Chr17:_75369484_75369485	*SEPTIN9*
1524	Chr17:_75369657_75369658	*SEPTIN9*
1525	Chr17:_76355202_76355203	*SOCS3*
1526	Chr17:_76921845_76921846	*TIMP2*
1527	Chr17:_7976456_7976457	*ALOX12B*

Figure 56Q

Order	Coordinate of CpG (hg19)	Gene
1528	Chr17: 8868834 8868835	*PIK3R5*
1529	Chr18: 11752089 11752090	*GNAL*
1530	Chr18: 22929205 22929206	*ZNF521*
1531	Chr18: 31739207 31739208	*NOL4*
1532	Chr18: 43355414 43355415	*Non Coding Region*
1533	Chr18: 44337853 44337854	*ST8SIA5*
1534	Chr18: 44337922 44337923	*ST8SIA5*
1535	Chr18: 53446784 53446785	*Non Coding Region*
1536	Chr18: 54789058 54789059	*Non Coding Region*
1537	Chr18: 56941246 56941247	*RAX*
1538	Chr18: 73167671 73167672	*Non Coding Region*
1539	Chr18: 74961968 74961969	*GALR1*
1540	Chr18: 76738999 76739000	*SALL3*
1541	Chr18: 904851 904852	*ADCYAP1*
1542	Chr18: 904920 904921	*ADCYAP1*
1543	Chr19: 13617091 13617092	*CACNA1A*
1544	Chr19: 15218713 15218714	*SYDE1*
1545	Chr19: 15580721 15580722	*PGLYRP2*
1546	Chr19: 16394638 16394639	*Non Coding Region*
1547	Chr19: 17008650 17008651	*CPAMD8*
1548	Chr19: 17392923 17392924	*ANKLE1*
1549	Chr19: 18540330 18540331	*SSBP4*
1550	Chr19: 20349275 20349276	*Non Coding Region*
1551	Chr19: 22034418 22034419	*Non Coding Region*
1552	Chr19: 24154607 24154608	*Non Coding Region*
1553	Chr19: 31843948 31843949	*Non Coding Region*
1554	Chr19: 31847970 31847971	*Non Coding Region*
1555	Chr19: 31848310 31848311	*Non Coding Region*
1556	Chr19: 36523584 36523585	*CLIP3*
1557	Chr19: 37096148 37096149	*ZNF529,ZNF382*
1558	Chr19: 37096321 37096322	*ZNF529,ZNF382*
1559	Chr19: 37096323 37096324	*ZNF529,ZNF382*
1560	Chr19: 37096329 37096330	*ZNF529,ZNF382*
1561	Chr19: 3786583 3786584	*MATK*
1562	Chr19: 39108609 39108610	*MAP4K1,EIF3K*
1563	Chr19: 39108672 39108673	*MAP4K1,EIF3K*
1564	Chr19: 42828156 42828157	*TMEM145*
1565	Chr19: 4328745 4328746	*STAP2*
1566	Chr19: 46996516 46996517	*PNMAL2*
1567	Chr19: 48918116 48918117	*GRIN2D*
1568	Chr19: 48918205 48918206	*GRIN2D*
1569	Chr19: 48984071 48984072	*CYTH2*
1570	Chr19: 49250439 49250440	*IZUMO1*
1571	Chr19: 50031017 50031018	*RCN3*
1572	Chr19: 50031246 50031247	*RCN3*

Order	Coordinate of CpG (hg19)	Gene
1573	Chr19: 50031273 50031274	*RCN3*
1574	Chr19: 52956771 52956772	*ZNF578*
1575	Chr19: 53758031 53758032	*ZNF677*
1576	Chr19: 54412993 54412994	*Non Coding Region*
1577	Chr19: 54926431 54926432	*TTYH1*
1578	Chr19: 54926514 54926515	*TTYH1*
1579	Chr19: 54926805 54926806	*TTYH1*
1580	Chr19: 56905152 56905153	*ZNF582*
1581	Chr19: 58095595 58095596	*ZIK1*
1582	Chr19: 58095659 58095660	*ZIK1*
1583	Chr19: 58220370 58220371	*ZNF154*
1584	Chr19: 58220494 58220495	*ZNF154*
1585	Chr19: 58238850 58238851	*ZNF671*
1586	Chr19: 58858806 58858807	*A1BG*
1587	Chr19: 58951599 58951600	*ZNF132*
1588	Chr19: 58951778 58951779	*ZNF132*
1589	Chr19: 58951885 58951886	*ZNF132*
1590	Chr20: 21492914 21492915	*NKX2-2*
1591	Chr20: 21682362 21682363	*Non Coding Region*
1592	Chr20: 25062447 25062448	*VSX1*
1593	Chr20: 2781262 2781263	*CPXM1*
1594	Chr20: 36012946 36012947	*SRC*
1595	Chr20: 37353096 37353097	*SLC32A1*
1596	Chr20: 37353126 37353127	*SLC32A1*
1597	Chr20: 37434167 37434168	*PPP1R16B*
1598	Chr20: 37434229 37434230	*PPP1R16B*
1599	Chr20: 42875779 42875780	*GDAP1L1*
1600	Chr20: 45142244 45142245	*ZNF334*
1601	Chr20: 50720731 50720732	*ZFP64*
1602	Chr20: 50720908 50720909	*ZFP64*
1603	Chr20: 57224919 57224920	*STX16*
1604	Chr20: 57225195 57225196	*STX16*
1605	Chr20: 59827212 59827213	*CDH4*
1606	Chr20: 59828528 59828529	*CDH4*
1607	Chr20: 61585924 61585925	*SLC17A9*
1608	Chr20: 61638574 61638575	*BHLHE23*
1609	Chr20: 61886079 61886080	*NKAIN4,FLJ16779*
1610	Chr21: 27945125 27945126	*CYYR1*
1611	Chr21: 34398199 34398200	*OLIG2*
1612	Chr21: 34398222 34398223	*OLIG2*
1613	Chr21: 34398226 34398227	*OLIG2*
1614	Chr21: 34398532 34398533	*OLIG2*
1615	Chr21: 38069950 38069951	*Non Coding Region*
1616	Chr21: 38076869 38076870	*SIM2*
1617	Chr21: 38077673 38077674	*SIM2*

Figure 56R

Order	Coordinate of CpG (hg19)	*Gene*
1618	Chr21:_38078767_38078768	*SIM2*
1619	Chr21:_38079615_38079616	*SIM2*
1620	Chr21:_38080526_38080527	*SIM2*
1621	Chr21:_38083149_38083150	*SIM2*
1622	Chr22:_25678988_25678989	*Non Coding Region*
1623	Chr22:_30720496_30720497	*TBC1D10A*
1624	Chr22:_50623687_50623688	*TRABD*
1625	Chr22:_50623692_50623693	*TRABD*

Figure 56S

Solid Tumor Methylation markers, 50% sensitivity, Chromosomal Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Solid Tumor Markers	
1	chr01:110610265-110613303	*ALX3*
2	chr01:111216244-111217937	*KCNA3*
3	chr01:12226876-12227712	*TNFRSF1B*
4	chr01:154474107-154475699	*TDRD10,SHE*
5	chr01:155290606-155291001	*RUSC1,C1orf104*
6	chr01:156405160-156406350	*Non Coding Region*
7	chr01:169396621-169396869	*C1orf114*
8	chr01:170630456-170630851	*Non Coding Region*
9	chr01:170633274-170634178	*PRRX1*
10	chr01:200009807-200010036	*NR5A2*
11	chr01:207818389-207818952	*CR1L*
12	chr01:219347109-219347572	*LYPLAL1*
13	chr01:231113871-231114873	*ARV1,TTC13*
14	chr01:29585897-29586598	*PTPRU*
15	chr01:44883136-44884272	*RNF220*
16	chr01:47998899-47999517	*Non Coding Region*
17	chr01:50884228-50891471	*DMRTA2*
18	chr01:66998728-66999060	*SGIP1*
19	chr01:77333111-77334534	*ST6GALNAC5*
20	chr01:91190489-91192804	*Non Coding Region*
21	chr02:119592602-119593845	*Non Coding Region*
22	chr02:127413696-127414171	*GYPC*
23	chr02:175208415-175209076	*Non Coding Region*
24	chr02:176986424-176988291	*HOXD9*
25	chr02:176993479-176995557	*HOXD8*
26	chr02:177024501-177025692	*Non Coding Region*
27	chr02:219738081-219738788	*WNT6*
28	chr02:233367780-233368577	*Non Coding Region*
29	chr02:238599857-238601430	*LRRFIP1*
30	chr02:26395416-26395620	*FAM59B*
31	chr02:45240372-45241579	*Non Coding Region*
32	chr02:71192094-71192495	*ATP6V1B1*
33	chr02:74740455-74743795	*TLX2*
34	chr02:85811340-85811855	*VAMP5*
35	chr02:97202313-97203787	*ARID5A*
36	chr03:121902742-121903645	*CASR*
37	chr03:129693127-129694841	*TRH*
38	chr03:139257712-139257949	*RBP1*
39	chr03:172165372-172166738	*GHSR*
40	chr03:179168735-179169593	*GNB4*
41	chr03:184099095-184099582	*CHRD*
42	chr03:44063314-44063837	*Non Coding Region*
43	chr03:50377803-50378540	*RASSF1*
44	chr04:42152766-42154984	*BEND4*
45	chr05:115151348-115152713	*CDO1*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Solid Tumor Markers	
46	chr05:11903550-11904703	*CTNND2*
47	chr05:134827285-134827644	*Non Coding Region*
48	chr05:40679502-40682081	*PTGER4*
49	chr05:43017968-43018668	*Non Coding Region*
50	chr05:58334836-58335881	*PDE4D*
51	chr05:72594147-72595808	*Non Coding Region*
52	chr05:72732365-72733732	*Non Coding Region*
53	chr06:105584148-105585621	*BVES*
54	chr06:152128822-152129771	*ESR1*
55	chr06:26240597-26240951	*HIST1H4F*
56	chr06:27107138-27107394	*HIST1H4I,HIST1H2BK*
57	chr06:28175282-28175605	*Non Coding Region*
58	chr06:28956106-28956578	*Non Coding Region*
59	chr06:30139718-30140263	*TRIM15*
60	chr06:6002471-6005125	*NRN1*
61	chr06:75794880-75795088	*COL12A1*
62	chr06:75914705-75916387	*Non Coding Region*
63	chr07:100489981-100490365	*ACHE*
64	chr07:113724924-113727795	*Non Coding Region*
65	chr07:127671158-127672853	*LRRC4,SND1*
66	chr07:128530629-128531356	*KCP*
67	chr07:143582125-143582610	*FAM115A*
68	chr07:149916640-149918264	*Non Coding Region*
69	chr07:155595692-155599414	*SHH*
70	chr07:27190274-27191115	*Non Coding Region*
71	chr07:27203915-27206462	*HOXA9*
72	chr07:27208871-27209616	*Non Coding Region*
73	chr07:27212416-27214396	*HOXA10*
74	chr07:27225050-27225629	*HOXA11AS,HOXA11*
75	chr07:27284639-27286237	*EVX1*
76	chr07:30721372-30722445	*CRHR2*
77	chr07:37487354-37488672	*ELMO1*
78	chr07:42267546-42267823	*GLI3*
79	chr07:95025559-95026122	*PON3*
80	chr07:96650221-96651551	*DLX5*
81	chr08:104510870-104513913	*RIMS2*
82	chr08:116679698-116679936	*TRPS1*
83	chr08:55366180-55367628	*Non Coding Region*
84	chr08:72917305-72917641	*Non Coding Region*
85	chr08:9760750-9761643	*LOC157627*
86	chr10:102586125-102588109	*PAX2*
87	chr10:106399567-106402812	*SORCS3*
88	chr10:122708510-122708899	*Non Coding Region*
89	chr10:15761423-15762101	*ITGA8*
90	chr10:22623350-22625875	*Non Coding Region*

Figure 57A

Order	Coordinate of CpG (hg19)	Gene
91	chr10:45914374-45914883	*ALOX5*
92	chr10:57390343-57391215	*Non Coding Region*
93	chr10:81002109-81003687	*ZMIZ1*
94	chr11:105481126-105481422	*GRIA4*
95	chr11:119227096-119227758	*USP2*
96	chr11:120434822-120435940	*Non Coding Region*
97	chr11:123301049-123302149	*Non Coding Region*
98	chr11:124790814-124791354	*HEPACAM*
99	chr11:133906735-133907359	*LOC100128239*
100	chr11:14926471-14927770	*Non Coding Region*
101	chr11:2158951-2162484	*IGF2,IGF2AS,INS-IGF2*
102	chr11:2290104-2292932	*ASCL2*
103	chr11:406491-407871	*SIGIRR*
104	chr11:43602545-43603215	*MIR129-2*
105	chr11:57249946-57250848	*Non Coding Region*
106	chr11:7694711-7695685	*CYB5R2*
107	chr11:829284-833558	*EFCAB4A*
108	chr12:113541697-113542086	*RASAL1*
109	chr12:114843022-114843610	*TBX5*
110	chr12:122516193-122517207	*MLXIP*
111	chr12:52400467-52401696	*GRASP*
112	chr12:52626793-52627577	*KRT7*
113	chr12:52652018-52652743	*Non Coding Region*
114	chr12:54423427-54423712	*HOXC5,HOXC4,HOXC6*
115	chr12:54447744-54448091	*HOXC4*
116	chr13:53313127-53314045	*LECT1*
117	chr13:79175610-79177985	*POU4F1*
118	chr13:96204691-96205496	*CLDN10*
119	chr14:105512055-105512307	*Non Coding Region*
120	chr14:23834435-23835947	*EFS*
121	chr14:24803678-24804353	*ADCY4*
122	chr14:29254365-29255069	*C14orf23*
123	chr14:37125219-37125661	*PAX9*
124	chr14:38067446-38069207	*Non Coding Region*
125	chr14:52734207-52735486	*PTGDR*
126	chr14:60951976-60952946	*C14orf39*
127	chr15:101419261-101421133	*ALDH1A3*
128	chr15:41785616-41788539	*ITPKA*
129	chr15:45479509-45480498	*SHF*
130	chr15:75251467-75251773	*Non Coding Region*
131	chr15:76632262-76635515	*ISL2*
132	chr15:83951980-83953930	*BNC1*
133	chr16:23846941-23848102	*PRKCB*
134	chr16:49311413-49312308	*Non Coding Region*
135	chr16:88700455-88701573	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
136	chr17:26698359-26699557	*SARM1*
137	chr17:42287692-42288392	*UBTF*
138	chr17:53342198-53343061	*HLF*
139	chr17:73583838-73586337	*MYO15B*
140	chr17:75368688-75370506	*SEPTIN9*
141	chr18:3498941-3499358	*DLGAP1*
142	chr18:54788959-54789194	*Non Coding Region*
143	chr18:904578-909574	*ADCYAP1*
144	chr19:11533198-11533619	*CCDC151*
145	chr19:13124569-13124788	*NFIX*
146	chr19:17000627-17001398	*F2RL3*
147	chr19:17007101-17008857	*CPAMD8*
148	chr19:17392383-17393775	*ANKLE1*
149	chr19:17403039-17403570	*ABHD8*
150	chr19:22034394-22034943	*Non Coding Region*
151	chr19:37095680-37096589	*ZNF529,ZNF382*
152	chr19:58858453-58859223	*A1BG*
153	chr20:30639908-30640786	*HCK*
154	chr20:45142000-45142337	*ZNF334*
155	chr20:62368955-62371962	*LIME1*
156	chr22:19753312-19755013	*TBX1*
157	chr22:38220589-38221619	*GALR3*
158	chr22:46366726-46368726	*WNT7B*
	Alternative markers: Low to no CRC	
159	chr01:1072369-1072847	*Non Coding Region*
160	chr01:149719314-149719640	*Non Coding Region*
161	chr01:151693991-151694282	*C1orf230*
162	chr01:151812253-151812525	*LOC100132111,C2CD4D*
163	chr01:154377670-154378445	*IL6R*
164	chr01:165323486-165323811	*LMX1A*
165	chr01:17025923-17027942	*ESPNP*
166	chr01:2064628-2064855	*PRKCZ*
167	chr01:211500151-211500636	*TRAF5*
168	chr01:214160798-214161034	*Non Coding Region*
169	chr01:217310749-217311178	*ESRRG*
170	chr01:2222198-2222569	*SKI*
171	chr01:240254959-240257063	*FMN2*
172	chr01:246729207-246730276	*CNST*
173	chr01:248020330-248021252	*TRIM58*
174	chr01:32169537-32169869	*COL16A1*
175	chr01:32180131-32180487	*Non Coding Region*
176	chr01:32410188-32410630	*Non Coding Region*
177	chr01:42611485-42611691	*Non Coding Region*
178	chr01:46954793-46957121	*Non Coding Region*
179	chr01:47696504-47698197	*TAL1*

Figure 57B

Order	Coordinate of CpG (hg19)	Gene
180	chr01:6086244-6086494	*KCNAB2*
181	chr01:6507207-6509186	*ESPN*
182	chr01:7022001-7022308	*CAMTA1*
183	chr01:76540147-76540653	*ST6GALNAC3*
184	chr01:844299-845883	*Non Coding Region*
185	chr01:90308839-90309606	*LRRC8D*
186	chr01:975496-980305	*AGRN*
187	chr02:105468851-105473488	*POU3F3*
188	chr02:111875206-111880965	*ACOXL*
189	chr02:119606569-119606826	*EN1*
190	chr02:119607378-119607910	*Non Coding Region*
191	chr02:127783081-127783311	*Non Coding Region*
192	chr02:160760604-160761452	*LY75*
193	chr02:162283581-162284677	*Non Coding Region*
194	chr02:175193398-175193764	*Non Coding Region*
195	chr02:176977284-176977540	*Non Coding Region*
196	chr02:177029413-177029941	*HOXD3*
197	chr02:177039551-177039951	*Non Coding Region*
198	chr02:177042751-177043444	*Non Coding Region*
199	chr02:19560963-19561650	*Non Coding Region*
200	chr02:200333687-200334172	*FLJ32063*
201	chr02:208489428-208491030	*Non Coding Region*
202	chr02:219736132-219736592	*WNT6*
203	chr02:220117937-220119334	*TUBA4A,TUBA4B*
204	chr02:223162946-223163912	*CCDC140,PAX3*
205	chr02:25142472-25143689	*ADCY3*
206	chr02:45161427-45163058	*Non Coding Region*
207	chr02:45169505-45171884	*SIX3*
208	chr02:63283936-63284147	*OTX1*
209	chr02:63285949-63287097	*Non Coding Region*
210	chr02:73151200-73152060	*EMX1*
211	chr02:74781494-74782685	*DOK1*
212	chr02:80529677-80530846	*LRRTM1,CTNNA2*
213	chr02:85107731-85108430	*C2orf89*
214	chr02:85980499-85982198	*ATOH8*
215	chr02:96990857-96991283	*ITPRIPL1*
216	chr03:113251699-113252290	*SIDT1*
217	chr03:122296612-122296828	*PARP15*
218	chr03:138656627-138659107	*Non Coding Region*
219	chr03:147126988-147128999	*ZIC4*
220	chr03:157821232-157821604	*SHOX2*
221	chr03:181437184-181437478	*SOX2OT*
222	chr03:49058325-49059986	*MIR425,DALRD3,NDUFAF3*
223	chr03:5019876-5022223	*Non Coding Region*
224	chr03:53190509-53190764	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
225	chr03:9904331-9904811	*Non Coding Region*
226	chr04:174254683-174256538	*HMGB2*
227	chr04:185746557-185747827	*ACSL1*
228	chr04:54957979-54958542	*Non Coding Region*
229	chr04:76555366-76556079	*CDKL2*
230	chr04:81124468-81124845	*PRDM8*
231	chr04:85402764-85403175	*Non Coding Region*
232	chr04:85417659-85420799	*NKX6-1*
233	chr05:100238722-100239138	*ST8SIA4*
234	chr05:101632049-101632373	*SLCO4C1*
235	chr05:138677333-138678248	*PAIP2*
236	chr05:139081001-139081202	*Non Coding Region*
237	chr05:153862142-153862451	*Non Coding Region*
238	chr05:170735169-170739863	*Non Coding Region*
239	chr05:172655840-172656202	*Non Coding Region*
240	chr05:175963498-175965116	*RNF44*
241	chr05:176023777-176024651	*GPRIN1*
242	chr05:37836747-37840726	*GDNF*
243	chr05:38257825-38259136	*EGFLAM*
244	chr05:42951076-42952410	*Non Coding Region*
245	chr05:42992033-42992955	*Non Coding Region*
246	chr05:42995122-42995415	*Non Coding Region*
247	chr06:10383525-10384114	*Non Coding Region*
248	chr06:10422321-10422648	*Non Coding Region*
249	chr06:134497536-134497756	*SGK1*
250	chr06:1381743-1385211	*Non Coding Region*
251	chr06:151186747-151188112	*MTHFD1L*
252	chr06:152623279-152623480	*SYNE1*
253	chr06:1604606-1615866	*Non Coding Region*
254	chr06:164092861-164093113	*Non Coding Region*
255	chr06:166579973-166583423	*T*
256	chr06:168197142-168198164	*C6orf123*
257	chr06:25139920-25140246	*CMAH*
258	chr06:26614013-26614851	*Non Coding Region*
259	chr06:27100174-27101124	*HIST1H2BJ*
260	chr06:27782247-27782485	*HIST1H2BM,HIST1H2AJ*
261	chr06:27841001-27841244	*HIST1H3I,HIST1H4L*
262	chr06:27858166-27858564	*HIST1H3J*
263	chr06:28554863-28555124	*Non Coding Region*
264	chr06:29894140-29895117	*HCG4P6*
265	chr06:30457369-30458175	*HLA-E*
266	chr06:31707489-31708336	*CLIC1*
267	chr06:32163292-32164383	*GPSM3*
268	chr06:35419879-35420766	*FANCE*
269	chr06:41472816-41473291	*Non Coding Region*

Figure 57C

Order	Coordinate of CpG (hg19)	Gene
270	chr06:41908745-41909711	*CCND3*
271	chr06:50810642-50810994	*TFAP2B*
272	chr06:52441269-52441793	*TRAM2*
273	chr06:6007387-6007797	*NRN1*
274	chr06:7107075-7109854	*RREB1*
275	chr06:7141036-7141576	*RREB1*
276	chr06:73330942-73333109	*KCNQ5*
277	chr06:91004795-91006944	*BACH2*
278	chr07:100201614-100203652	*PCOLCE*
279	chr07:128809089-128809393	*Non Coding Region*
280	chr07:130126017-130126801	*MEST*
281	chr07:151078462-151079536	*WDR86*
282	chr07:158621005-158622747	*ESYT2*
283	chr07:23508184-23509712	*IGF2BP3*
284	chr07:26191794-26192757	*NFE2L3*
285	chr07:27195601-27196567	*HOXA7*
286	chr07:27198182-27198514	*HOXA7*
287	chr07:27238690-27240311	*HOXA13*
288	chr07:32801484-32801972	*Non Coding Region*
289	chr07:37960316-37961046	*EPDR1*
290	chr07:45025958-45026626	*SNORA9,C7orf40*
291	chr07:64349424-64350151	*Non Coding Region*
292	chr07:96653467-96654199	*DLX5*
293	chr07:99595206-99595505	*Non Coding Region*
294	chr08:124172800-124173541	*Non Coding Region*
295	chr08:134581969-134584875	*ST3GAL1*
296	chr08:145016417-145016916	*PLEC1*
297	chr08:82192378-82193685	*FABP5*
298	chr08:99952020-99954686	*Non Coding Region*
299	chr09:112810858-112811128	*PALM2-AKAP2,AKAP2*
300	chr09:126773246-126780953	*LHX2*
301	chr09:140172613-140176105	*C9orf167*
302	chr09:27528357-27528725	*MOBKL2B*
303	chr09:34589113-34591978	*CNTFR*
304	chr09:79628910-79629393	*Non Coding Region*
305	chr09:96720586-96723189	*Non Coding Region*
306	chr10:102590122-102590402	*PAX2*
307	chr10:119301321-119305368	*EMX2OS,EMX2*
308	chr10:79685719-79686507	*LOC100128292,DLG5*
309	chr10:88730554-88731632	*AGAP11*
310	chr11:118478235-118481896	*PHLDB1*
311	chr11:31841315-31842003	*Non Coding Region*
312	chr11:416419-417650	*SIGIRR*
313	chr11:46368650-46369660	*DGKZ*
314	chr11:46389074-46389378	*DGKZ*

Order	Coordinate of CpG (hg19)	Gene
315	chr11:47291200-47291858	*MADD*
316	chr11:61061751-61063063	*VWCE*
317	chr11:626728-628037	*SCT*
318	chr11:67182965-67183393	*ATPGD1*
319	chr11:67350928-67351953	*GSTP1*
320	chr11:68622722-68623252	*Non Coding Region*
321	chr11:720561-721369	*EPS8L2*
322	chr11:76381449-76382295	*LRRC32*
323	chr12:111842938-111844392	*SH2B3*
324	chr12:114833911-114834210	*TBX5*
325	chr12:122277301-122277539	*Non Coding Region*
326	chr12:130526915-130527117	*Non Coding Region*
327	chr12:3308812-3310270	*TSPAN9*
328	chr12:3862068-3862606	*EFCAB4B*
329	chr12:46661054-46663408	*SLC38A1*
330	chr12:49390617-49392441	*DDN*
331	chr12:54071053-54071265	*ATP5G2*
332	chr12:54132148-54132353	*Non Coding Region*
333	chr12:54359658-54359906	*HOTAIR*
334	chr12:54387825-54388732	*Non Coding Region*
335	chr12:54399412-54399654	*Non Coding Region*
336	chr12:54408426-54408713	*Non Coding Region*
337	chr12:57618769-57619402	*NXPH4*
338	chr12:58013171-58013652	*SLC26A10*
339	chr12:58021294-58022037	*B4GALNT1*
340	chr12:58130870-58132047	*AGAP2*
341	chr13:100608122-100609686	*Non Coding Region*
342	chr13:100641334-100642188	*Non Coding Region*
343	chr13:113622737-113623660	*MCF2L*
344	chr13:20766208-20767779	*GJB2*
345	chr13:28491488-28492518	*Non Coding Region*
346	chr13:28527755-28528996	*Non Coding Region*
347	chr13:28544524-28544890	*CDX2*
348	chr13:43566074-43566508	*EPSTI1*
349	chr13:98794398-98796241	*FARP1*
350	chr14:100615540-100615946	*DEGS2*
351	chr14:24808635-24808992	*RIPK3*
352	chr14:54420184-54422958	*BMP4*
353	chr14:57283967-57284558	*OTX2OS1*
354	chr14:61103978-61104663	*Non Coding Region*
355	chr14:62217725-62218077	*Non Coding Region*
356	chr14:76012830-76013204	*BATF*
357	chr15:33009530-33011696	*GREM1*
358	chr15:42264156-42264861	*EHD4*
359	chr15:60690708-60690930	*ANXA2*

Figure 57D

Order	Coordinate of CpG (hg19)	Gene
360	chr15:75074425-75075146	*CSK*
361	chr15:76627523-76627838	*ISL2*
362	chr15:76630029-76630970	*ISL2*
363	chr15:81292793-81295609	*MESDC1*
364	chr15:89949373-89951130	*Non Coding Region*
365	chr15:93615622-93617185	*RGMA*
366	chr16:29086220-29086434	*RRN3P2*
367	chr16:31487152-31488967	*TGFB1I1*
368	chr16:4363935-4364151	*Non Coding Region*
369	chr16:54315602-54316104	*Non Coding Region*
370	chr16:56671937-56672530	*MT1A*
371	chr16:69599437-69600736	*NFAT5*
372	chr16:72821140-72821592	*ZFHX3*
373	chr16:85783862-85785131	*C16orf74*
374	chr16:85932121-85932942	*IRF8*
375	chr16:88716989-88717606	*CYBA*
376	chr16:89006334-89008600	*CBFA2T3*
377	chr16:89777018-89778789	*C16orf7*
378	chr17:27893086-27896078	*ABHD15*
379	chr17:3438841-3439046	*TRPV3*
380	chr17:35078631-35078972	*Non Coding Region*
381	chr17:35291899-35300875	*LHX1*
382	chr17:36105334-36105583	*HNF1B*
383	chr17:36715691-36715995	*SRCIN1*
384	chr17:36728633-36729284	*SRCIN1*
385	chr17:37025737-37027104	*LASP1*
386	chr17:40086751-40087098	*TTC25*
387	chr17:43393891-43394938	*MAP3K14*
388	chr17:44895800-44897025	*WNT3*
389	chr17:46796234-46797292	*Non Coding Region*
390	chr17:48048953-48050616	*DLX4*
391	chr17:48350422-48350832	*TMEM92*
392	chr17:56401728-56402343	*BZRAP1*
393	chr17:59473060-59483266	*TBX2*
394	chr17:62774366-62777797	*LOC146880*
395	chr17:66255192-66255556	*ARSG*
396	chr17:76127521-76128406	*TMC6,TMC8*
397	chr17:7620616-7621119	*DNAH2*
398	chr17:80186135-80189990	*SLC16A3*
399	chr17:80251357-80251673	*Non Coding Region*
400	chr17:9142980-9143839	*NTN1*
401	chr18:2905950-2907348	*EMILIN2*
402	chr18:29264703-29265504	*B4GALT6*
403	chr18:56975842-56976160	*CPLX4*
404	chr19:1074726-1075071	*HMHA1*

Order	Coordinate of CpG (hg19)	Gene
405	chr19:13273652-13274116	*Non Coding Region*
406	chr19:14168142-14168512	*PALM3*
407	chr19:16178396-16178609	*TPM4*
408	chr19:16186789-16188275	*TPM4*
409	chr19:17346165-17346886	*NR2F6*
410	chr19:17958490-17958808	*JAK3*
411	chr19:1856725-1857443	*KLF16*
412	chr19:35786551-35786962	*MAG*
413	chr19:36134153-36135781	*ETV2*
414	chr19:38182793-38183327	*ZNF781*
415	chr19:3821974-3822278	*ZFR2*
416	chr19:38700333-38700577	*DPF1*
417	chr19:42927285-42928598	*LIPE*
418	chr19:911689-911996	*C19orf22*
419	chr20:21080929-21082824	*Non Coding Region*
420	chr20:3052097-3053103	*OXT*
421	chr20:42543097-42545137	*TOX2*
422	chr20:55200279-55206789	*Non Coding Region*
423	chr21:36398960-36399259	*RUNX1*
424	chr21:38119793-38120742	*SIM2*
425	chr21:42878751-42880674	*TMPRSS2*
426	chr21:46897841-46898158	*COL18A1*
427	chr22:19709081-19712106	*SEPT5,GP1BB*
428	chr22:37815803-37816477	*ELFN2*
429	chr22:46457095-46458028	*LOC150381*
430	chr22:46770433-46770871	*CELSR1*
	Alternative markers: High in CRC, other cancers	
431	chr01:114696886-114697185	*SYT6*
432	chr01:119531991-119532196	*TBX15*
433	chr01:119543056-119543454	*Non Coding Region*
434	chr01:119549144-119551320	*Non Coding Region*
435	chr01:1476093-1476669	*C1orf70*
436	chr01:151810460-151811684	*LOC100132111,C2CD4D*
437	chr01:1564421-1570006	*MMP23B,MMP23A*
438	chr01:18956895-18959829	*PAX7*
439	chr01:200008392-200009047	*NR5A2*
440	chr01:221050448-221050864	*Non Coding Region*
441	chr01:228651447-228652529	*Non Coding Region*
442	chr01:229542943-229543770	*Non Coding Region*
443	chr01:229567206-229570684	*ACTA1*
444	chr01:237205126-237206644	*RYR2*
445	chr01:243646394-243646888	*SDCCAG8*
446	chr01:2983925-2987962	*PRDM16*
447	chr01:36042432-36043444	*TFAP2E*
448	chr01:44031286-44031853	*PTPRF*

Figure 57E

Order	Coordinate of CpG (hg19)	Gene
449	chr01:44871109-44874047	*RNF220*
450	chr01:4713989-4716555	*AJAP1*
451	chr01:47909712-47911020	*Non Coding Region*
452	chr01:48058793-48059230	*Non Coding Region*
453	chr01:50881884-50882103	*Non Coding Region*
454	chr01:58715272-58716095	*DAB1*
455	chr01:63782394-63790471	*Non Coding Region*
456	chr01:6525996-6526228	*TNFRSF25,PLEKHG5*
457	chr01:67218079-67218293	*TCTEX1D1*
458	chr01:91183240-91184540	*BARHL2*
459	chr01:91300979-91301891	*Non Coding Region*
460	chr01:9711780-9713001	*C1orf200,PIK3CD*
461	chr01:98510966-98511710	*Non Coding Region*
462	chr01:99469932-99470968	*LPPR5*
463	chr02:105459127-105461770	*Non Coding Region*
464	chr02:119599458-119600966	*EN1*
465	chr02:131720819-131721867	*ARHGEF4*
466	chr02:139537692-139538650	*NXPH2*
467	chr02:171678546-171680358	*GAD1*
468	chr02:172972804-172973242	*Non Coding Region*
469	chr02:176956504-176956707	*HOXD13*
470	chr02:182321761-182323029	*ITGA4*
471	chr02:185463208-185463589	*ZNF804A*
472	chr02:206549599-206551316	*NRP2*
473	chr02:208988834-208989269	*CRYGD*
474	chr02:220299483-220300243	*SPEG*
475	chr02:220361462-220363254	*Non Coding Region*
476	chr02:222436034-222438941	*EPHA4*
477	chr02:223161531-223161919	*CCDC140,PAX3*
478	chr02:228736230-228736544	*WDR69*
479	chr02:233497826-233499727	*EFHD1*
480	chr02:237416090-237416382	*IQCA1*
481	chr02:238864315-238865170	*Non Coding Region*
482	chr02:239140096-239140659	*LOC643387,LOC151174*
483	chr02:242742598-242743758	*GAL3ST2*
484	chr02:25438725-25439276	*Non Coding Region*
485	chr02:45028955-45029318	*Non Coding Region*
486	chr02:45155195-45157049	*Non Coding Region*
487	chr02:45231211-45231482	*Non Coding Region*
488	chr02:467849-468659	*Non Coding Region*
489	chr02:63281034-63281347	*OTX1*
490	chr02:66808568-66809404	*Non Coding Region*
491	chr02:97192977-97193383	*Non Coding Region*
492	chr03:119528920-119529159	*NR1I2*
493	chr03:128205495-128212274	*GATA2*

Order	Coordinate of CpG (hg19)	Gene
494	chr03:128719865-128721245	*CCDC48*
495	chr03:133393118-133393657	*Non Coding Region*
496	chr03:138679244-138679579	*Non Coding Region*
497	chr03:141516055-141516639	*GRK7*
498	chr03:169379437-169381056	*MECOM*
499	chr03:184301117-184302134	*Non Coding Region*
500	chr03:192125821-192127994	*FGF12*
501	chr03:194406440-194409045	*FAM43A*
502	chr03:27765196-27765675	*EOMES*
503	chr03:32858194-32860506	*TRIM71*
504	chr03:38035701-38036000	*VILL*
505	chr03:62356773-62357315	*FEZF2*
506	chr04:10458129-10459353	*ZNF518B*
507	chr04:111554965-111555504	*PITX2*
508	chr04:13524062-13526083	*Non Coding Region*
509	chr04:13530847-13531244	*Non Coding Region*
510	chr04:13543562-13546494	*Non Coding Region*
511	chr04:1396291-1401730	*Non Coding Region*
512	chr04:140655962-140657135	*MAML3*
513	chr04:147559205-147561901	*POU4F2*
514	chr04:155254130-155254603	*DCHS2*
515	chr04:169799086-169799625	*PALLD*
516	chr04:172733734-172735118	*GALNTL6*
517	chr04:174430386-174430861	*Non Coding Region*
518	chr04:174450046-174451469	*NBLA00301,HAND2*
519	chr04:175135259-175135496	*Non Coding Region*
520	chr04:4868440-4869173	*Non Coding Region*
521	chr04:57521621-57522703	*HOPX*
522	chr04:778661-780592	*CPLX1*
523	chr04:81187494-81187913	*FGF5*
524	chr05:110408704-110409039	*TSLP*
525	chr05:134363092-134365146	*Non Coding Region*
526	chr05:134825398-134826496	*Non Coding Region*
527	chr05:135265848-135266517	*FBXL21*
528	chr05:137224986-137225477	*PKD2L2*
529	chr05:140787447-140788044	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
530	chr05:140797162-140797701	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
531	chr05:140800760-140801072	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
532	chr05:140864527-140864748	*PCDHGB1,PCDHGB3, PCDHGB2, etc.*
533	chr05:145718289-145720095	*POU4F3*
534	chr05:158532294-158532502	*Non Coding Region*
535	chr05:16179064-16180420	*MARCHF11*
536	chr05:169064270-169064702	*DOCK2*
537	chr05:170741603-170742751	*Non Coding Region*
538	chr05:172659049-172660277	*NKX2-5*

Figure 57F

Order	Coordinate of CpG (hg19)	Gene
539	chr05:176107068-176107521	*Non Coding Region*
540	chr05:177412534-177412753	*Non Coding Region*
541	chr05:178593903-178594656	*ADAMTS2*
542	chr05:179222608-179223825	*LTC4S*
543	chr05:1874907-1879032	*Non Coding Region*
544	chr05:2748368-2757024	*Non Coding Region*
545	chr05:3594467-3603054	*IRX1*
546	chr05:37834671-37835128	*GDNF*
547	chr05:42994626-42994936	*Non Coding Region*
548	chr05:528383-528982	*Non Coding Region*
549	chr05:54516267-54516919	*Non Coding Region*
550	chr05:72529099-72529976	*Non Coding Region*
551	chr05:87968635-87968907	*LOC645323*
552	chr06:100912071-100913337	*SIM1*
553	chr06:10390038-10390565	*Non Coding Region*
554	chr06:105388190-105389545	*Non Coding Region*
555	chr06:106429111-106429772	*Non Coding Region*
556	chr06:108485671-108490539	*NR2E1*
557	chr06:108495654-108495986	*NR2E1*
558	chr06:10881846-10882051	*GCM2*
559	chr06:10882926-10883149	*GCM2*
560	chr06:134210639-134211218	*TCF21*
561	chr06:137809342-137810204	*Non Coding Region*
562	chr06:137814355-137815202	*OLIG3*
563	chr06:1379693-1380014	*Non Coding Region*
564	chr06:150284682-150286515	*ULBP1*
565	chr06:159589636-159591319	*FNDC1*
566	chr06:1624185-1625468	*GMDS*
567	chr06:27463176-27463379	*Non Coding Region*
568	chr06:27647793-27648005	*Non Coding Region*
569	chr06:28457568-28457926	*Non Coding Region*
570	chr06:28602853-28603295	*Non Coding Region*
571	chr06:28781857-28782302	*Non Coding Region*
572	chr06:29521110-29521833	*Non Coding Region*
573	chr06:29759946-29760903	*HCG4*
574	chr06:29795553-29796594	*HLA-G*
575	chr06:29944402-29945169	*HCG9*
576	chr06:30095173-30095610	*Non Coding Region*
577	chr06:32116590-32117229	*PRRT1*
578	chr06:35108801-35109499	*TCP11*
579	chr06:391188-393790	*IRF4*
580	chr06:41340667-41342543	*Non Coding Region*
581	chr06:43252734-43253476	*TTBK1*
582	chr06:50692149-50692375	*TFAP2D*
583	chr06:5994909-5995203	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
584	chr06:78172231-78174088	*HTR1B*
585	chr06:85472702-85474132	*TBX18*
586	chr07:100075303-100075551	*TSC22D4*
587	chr07:1269165-1282864	*Non Coding Region*
588	chr07:149487283-149487769	*SSPO*
589	chr07:155164557-155167854	*Non Coding Region*
590	chr07:156795355-156799394	*Non Coding Region*
591	chr07:157476886-157486719	*PTPRN2*
592	chr07:158936507-158938492	*VIPR2*
593	chr07:19145872-19146256	*Non Coding Region*
594	chr07:19156050-19158042	*TWIST1*
595	chr07:19184221-19184686	*Non Coding Region*
596	chr07:24323558-24325080	*NPY*
597	chr07:35292985-35294674	*TBX20*
598	chr07:35296921-35298218	*Non Coding Region*
599	chr07:49813008-49815752	*VWC2*
600	chr07:54612324-54612558	*VSTM2A*
601	chr07:6703503-6704075	*Non Coding Region*
602	chr07:70596228-70598382	*WBSCR17*
603	chr07:8481974-8482762	*NXPH1*
604	chr07:98245805-98247759	*NPTX2*
605	chr08:11559596-11562956	*GATA4*
606	chr08:116660432-116660747	*TRPS1*
607	chr08:132052203-132054749	*Non Coding Region*
608	chr08:140714585-140718259	*KCNK9*
609	chr08:143545445-143546178	*BAI1*
610	chr08:144651572-144652288	*C8orf73*
611	chr08:23562475-23565175	*NKX2-6*
612	chr08:23567180-23567678	*Non Coding Region*
613	chr08:24770908-24772547	*NEFM*
614	chr08:49468683-49468959	*Non Coding Region*
615	chr08:53851701-53854426	*NPBWR1*
616	chr08:55378928-55380186	*Non Coding Region*
617	chr08:57025546-57026543	*MOS*
618	chr08:57069687-57070047	*Non Coding Region*
619	chr08:65281903-65283043	*Non Coding Region*
620	chr08:65290108-65290946	*MIR124-2*
621	chr08:67873388-67875600	*Non Coding Region*
622	chr08:70981873-70984888	*PRDM14*
623	chr08:72468560-72469561	*Non Coding Region*
624	chr08:72753874-72754755	*MSC*
625	chr08:72755783-72756667	*MSC*
626	chr08:77594255-77594860	*ZFHX4,LOC100192378*
627	chr08:85096759-85097247	*RALYL*
628	chr08:86350765-86351196	*CA3*

Figure 57G

Order	Coordinate of CpG (hg19)	Gene
629	chr08:97156763-97158030	*GDF6*
630	chr08:97171805-97172022	*GDF6*
631	chr08:98289604-98290404	*TSPYL5*
632	chr09:100610696-100611517	*Non Coding Region*
633	chr09:124461797-124462190	*DAB2IP*
634	chr09:37002489-37002957	*PAX5*
635	chr09:79633326-79636030	*FOXB2*
636	chr10:100993820-100994188	*HPSE2*
637	chr10:102419147-102419668	*Non Coding Region*
638	chr10:102497154-102497675	*Non Coding Region*
639	chr10:102893660-102895059	*TLX1*
640	chr10:102899177-102899489	*Non Coding Region*
641	chr10:102899822-102900263	*Non Coding Region*
642	chr10:103603166-103603642	*KCNIP2*
643	chr10:105036628-105038084	*INA*
644	chr10:105452338-105453230	*SH3PXD2A*
645	chr10:108923780-108924805	*SORCS1*
646	chr10:11207178-11207980	*CUGBP2*
647	chr10:118030732-118034230	*GFRA1*
648	chr10:118892161-118892639	*VAX1*
649	chr10:123922850-123923542	*TACC2*
650	chr10:131756690-131758787	*EBF3*
651	chr10:131761169-131771696	*Non Coding Region*
652	chr10:133109193-133111052	*TCERG1L*
653	chr10:22540707-22542739	*Non Coding Region*
654	chr10:22634000-22634862	*SPAG6*
655	chr10:26727061-26727992	*APBB1IP*
656	chr10:44788811-44789120	*Non Coding Region*
657	chr10:50602989-50606783	*Non Coding Region*
658	chr10:8091374-8098329	*GATA3*
659	chr10:88122924-88127364	*GRID1*
660	chr10:94820026-94823252	*CYP26C1*
661	chr11:115529997-115530548	*Non Coding Region*
662	chr11:119293320-119293943	*THY1*
663	chr11:123066517-123066986	*ASAM*
664	chr11:125034896-125036426	*PKNOX2*
665	chr11:125774292-125774584	*PUS3,DDX25*
666	chr11:20618197-20619920	*Non Coding Region*
667	chr11:31820060-31821416	*PAX6*
668	chr11:31827696-31827921	*PAX6*
669	chr11:31848487-31848776	*Non Coding Region*
670	chr11:32454874-32457311	*WT1,WIT1*
671	chr11:32459422-32459878	*WIT1*
672	chr11:44332314-44332757	*ALX4*
673	chr11:61322732-61323285	*SYT7*

Order	Coordinate of CpG (hg19)	Gene
674	chr11:636906-640628	*DRD4*
675	chr11:63756058-63768666	*MACROD1*
676	chr11:69706778-69707017	*Non Coding Region*
677	chr11:71954816-71955659	*PHOX2A*
678	chr11:78673007-78673213	*ODZ4*
679	chr11:79148358-79152200	*ODZ4*
680	chr12:114845861-114847650	*TBX5*
681	chr12:115102687-115103982	*Non Coding Region*
682	chr12:120799372-120799912	*MSI1*
683	chr12:133484658-133485739	*Non Coding Region*
684	chr12:25055599-25056246	*BCAT1*
685	chr12:5018585-5021171	*KCNA1*
686	chr12:50297580-50297988	*FAIM2*
687	chr12:54089211-54089413	*Non Coding Region*
688	chr12:54345563-54345916	*Non Coding Region*
689	chr12:54427025-54428709	*HOXC5,HOXC4*
690	chr12:54811981-54812202	*ITGA5*
691	chr12:62584768-62586268	*FAM19A2*
692	chr12:65514878-65515863	*WIF1*
693	chr12:95941906-95942979	*USP44*
694	chr13:109147798-109149019	*Non Coding Region*
695	chr13:112758598-112760491	*Non Coding Region*
696	chr13:21520328-21520564	*Non Coding Region*
697	chr13:25319668-25322255	*Non Coding Region*
698	chr13:25743998-25746127	*FAM123A*
699	chr13:28498226-28499046	*PDX1*
700	chr13:28501859-28502090	*Non Coding Region*
701	chr13:28503057-28503374	*Non Coding Region*
702	chr13:29106576-29107254	*Non Coding Region*
703	chr13:53419897-53422872	*PCDH8*
704	chr13:58203586-58204322	*Non Coding Region*
705	chr13:79170114-79171231	*Non Coding Region*
706	chr13:84453664-84453897	*SLITRK1*
707	chr14:101923575-101925995	*Non Coding Region*
708	chr14:102552721-102554068	*HSP90AA1*
709	chr14:24044886-24046760	*JPH4*
710	chr14:48143433-48145589	*MDGA2*
711	chr14:52534581-52536722	*NID2*
712	chr14:57264638-57265561	*Non Coding Region*
713	chr14:60973772-60974123	*Non Coding Region*
714	chr14:60975732-60978180	*SIX6*
715	chr14:70014543-70014993	*Non Coding Region*
716	chr14:70655095-70656077	*SLC8A3*
717	chr14:95234612-95236569	*GSC*
718	chr15:29077251-29077684	*Non Coding Region*

Figure 57H

Order	Coordinate of CpG (hg19)	Gene
719	chr15:30515266-30518445	*Non Coding Region*
720	chr15:37390175-37390380	*MEIS2*
721	chr15:41793562-41795305	*ITPKA*
722	chr15:51385861-51387203	*TNFAIP8L3*
723	chr15:53080458-53083699	*ONECUT1*
724	chr15:65688039-65688539	*IGDCC4*
725	chr15:68115485-68122575	*LBXCOR1*
726	chr15:89920793-89922768	*LOC254559*
727	chr15:89952271-89953061	*Non Coding Region*
728	chr16:2041825-2043026	*SYNGR3*
729	chr16:21294980-21295414	*CRYM*
730	chr16:2765619-2765855	*PRSS27*
731	chr16:30905549-30905783	*BCL7C,MIR762*
732	chr16:31580070-31580317	*CSDAP1*
733	chr16:3238805-3239492	*Non Coding Region*
734	chr16:51183699-51188763	*SALL1*
735	chr16:55362737-55363287	*IRX6*
736	chr16:55513220-55513526	*MMP2*
737	chr16:55689598-55690895	*SLC6A2*
738	chr16:66612749-66613412	*CMTM2,CMTM1*
739	chr16:86541430-86545228	*Non Coding Region*
740	chr16:86599355-86602649	*FOXC2*
741	chr16:86612188-86613821	*FOXL1*
742	chr16:86965271-86965496	*Non Coding Region*
743	chr17:17625605-17628500	*RAI1*
744	chr17:19880676-19881633	*AKAP10*
745	chr17:27038084-27038919	*PROCA1*
746	chr17:27224005-27224873	*FLOT2*
747	chr17:27939298-27940770	*ANKRD13B*
748	chr17:32484007-32484280	*ACCN1*
749	chr17:36734659-36735246	*SRCIN1*
750	chr17:37321482-37322099	*ARL5C*
751	chr17:38347533-38347765	*RAPGEFL1*
752	chr17:40827975-40828659	*PLEKHH3*
753	chr17:43339124-43339832	*C17orf46,LOC100133991*
754	chr17:46802790-46803076	*HOXB13*
755	chr17:59528979-59530266	*Non Coding Region*
756	chr17:59531723-59535254	*TBX4*
757	chr17:61776890-61778733	*LIMD2*
758	chr17:7287198-7287940	*TNK1*
759	chr17:76354818-76357038	*SOCS3*
760	chr17:76920684-76921961	*TIMP2*
761	chr17:8868469-8869372	*PIK3R5*
762	chr18:11750953-11752756	*GNAL*
763	chr18:22930018-22930904	*ZNF521*

Order	Coordinate of CpG (hg19)	Gene
764	chr18:31739047-31739522	*NOL4*
765	chr18:43355318-43355698	*Non Coding Region*
766	chr18:44337510-44338100	*ST8SIA5*
767	chr18:53447074-53447842	*Non Coding Region*
768	chr18:56939624-56941540	*RAX*
769	chr18:73167402-73167920	*Non Coding Region*
770	chr18:74961556-74963822	*GALR1*
771	chr18:76737005-76741244	*SALL3*
772	chr19:13616752-13617267	*CACNA1A*
773	chr19:15217951-15218617	*SYDE1*
774	chr19:15580382-15580863	*PGLYRP2*
775	chr19:16394450-16394662	*Non Coding Region*
776	chr19:18539781-18540341	*SSBP4*
777	chr19:24154457-24154759	*Non Coding Region*
778	chr19:31843880-31844183	*Non Coding Region*
779	chr19:31847946-31848209	*Non Coding Region*
780	chr19:36523391-36523887	*CLIP3*
781	chr19:3785511-3786653	*MATK*
782	chr19:39109432-39109862	*MAP4K1,EIF3K*
783	chr19:42827743-42829149	*TMEM145*
784	chr19:4328658-4328865	*STAP2*
785	chr19:46996327-46998437	*PNMAL2*
786	chr19:48918115-48918340	*GRIN2D*
787	chr19:48983562-48984175	*CYTH2*
788	chr19:49253688-49254221	*IZUMO1*
789	chr19:50030981-50031300	*RCN3*
790	chr19:52956656-52957245	*ZNF578*
791	chr19:53757819-53758148	*ZNF677*
792	chr19:54412710-54413087	*Non Coding Region*
793	chr19:54927902-54928225	*TTYH1*
794	chr19:56904636-56905355	*ZNF582*
795	chr19:58094739-58095764	*ZIK1*
796	chr19:58220189-58220517	*ZNF154*
797	chr19:58238585-58239028	*ZNF671*
798	chr19:58951214-58952250	*ZNF132*
799	chr20:21485932-21496714	*NKX2-2*
800	chr20:21683744-21684420	*Non Coding Region*
801	chr20:25061761-25062928	*VSX1*
802	chr20:2780978-2781497	*CPXM1*
803	chr20:36012595-36013439	*SRC*
804	chr20:37352130-37357372	*SLC32A1*
805	chr20:37434206-37435592	*PPP1R16B*
806	chr20:42875764-42876184	*GDAP1L1*
807	chr20:50721310-50721912	*ZFP64*
808	chr20:57224695-57226322	*STX16*

Figure 57I

Order	Coordinate of CpG (hg19)	*Gene*
809	chr20:59826977-59828978	*CDH4*
810	chr20:61583891-61584222	*SLC17A9*
811	chr20:61636171-61639055	*BHLHE23*
812	chr20:61884644-61886387	*NKAIN4,FLJ16779*
813	chr21:27945010-27945646	*CYYR1*
814	chr21:34395128-34400245	*OLIG2*
815	chr21:38068193-38073891	*Non Coding Region*
816	chr21:38076762-38077685	*SIM2*
817	chr21:38079941-38081833	*SIM2*
818	chr22:25678576-25679273	*Non Coding Region*
819	chr22:30722572-30723218	*TBC1D10A*
820	chr22:50623164-50625226	*TRABD*

Figure 57J

Group 1 Exon RNA sequence markers

Order	Exon	Gene
1	chr3:46568916-46569071:-	LRRC2
2	chr4:74962764-74963488:-	CXCL2
3	chr4:74964318-74964401:-	CXCL2
4	chr4:74964516-74964639:-	CXCL2
5	chr4:74964738-74964997:-	CXCL2
6	chr4:110661850-110662266:-	CFI
7	chr4:110663647-110663751:-	CFI
8	chr4:110667378-110667658:-	CFI
9	chr4:110670374-110670477:-	CFI
10	chr4:110670655-110670758:-	CFI
11	chr4:110681426-110681536:-	CFI
12	chr4:110681679-110681792:-	CFI
13	chr4:110682673-110683747:-	CFI
14	chr6:90374206-90374347:-	MDN1
15	chr6:90377733-90377870:-	MDN1
16	chr6:90380638-90380799:-	MDN1
17	chr6:90381919-90382082:-	MDN1
18	chr6:90382266-90382421:-	MDN1
19	chr6:90382955-90383181:-	MDN1
20	chr6:90383823-90384308:-	MDN1
21	chr6:90387289-90387415:-	MDN1
22	chr6:90388318-90388464:-	MDN1
23	chr6:90394595-90394704:-	MDN1
24	chr6:90395546-90395692:-	MDN1
25	chr6:90396621-90396742:-	MDN1
26	chr6:90399589-90399862:-	MDN1
27	chr6:168185221-168186176:-	C6orf123
28	chr13:41340882-41341169:-	MRPS31
29	chr13:42140962-42142441:-	KIAA0564

Figure 58

Group 1 Methylation markers, 50% sensitivity, CpG Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 1 Markers	
1	Chr01: 107683775 107683776	*NTNG1*
2	Chr01: 121261602 121261603	*LOC647121*
3	Chr01: 18957020 18957021	*PAX7*
4	Chr01: 240255486 240255487	*FMN2*
5	Chr01: 29138936 29138937	*OPRD1*
6	Chr01: 34630944 34630945	*CSMD2*
7	Chr01: 44883697 44883698	*RNF220*
8	Chr01: 50884544 50884545	*DMRTA2*
9	Chr01: 53068181 53068182	*GPX7*
10	Chr01: 77333138 77333139	*ST6GALNAC5*
11	Chr02: 100938444 100938445	*LONRF2*
12	Chr02: 131792772 131792773	*ARHGEF4*
13	Chr02: 175193377 175193378	*Non Coding Region*
14	Chr02: 176957462 176957463	*HOXD13*
15	Chr02: 176976802 176976803	*Non Coding Region*
16	Chr02: 176994665 176994666	*HOXD8*
17	Chr02: 176995556 176995557	*HOXD8*
18	Chr02: 182548807 182548808	*Non Coding Region*
19	Chr02: 29338432 29338433	*CLIP4*
20	Chr02: 29339076 29339077	*CLIP4*
21	Chr02: 66803033 66803034	*Non Coding Region*
22	Chr02: 88751786 88751787	*FOXI3*
23	Chr03: 140770345 140770346	*SPSB4*
24	Chr03: 152553673 152553674	*P2RY1*
25	Chr03: 181421427 181421428	*SOX2OT*
26	Chr03: 69592090 69592091	*Non Coding Region*
27	Chr03: 96532043 96532044	*EPHA6*
28	Chr03: 96533290 96533291	*EPHA6*
29	Chr04: 37246827 37246828	*KIAA1239*
30	Chr05: 112073348 112073349	*APC*
31	Chr05: 112073350 112073351	*APC*
32	Chr05: 112073398 112073399	*APC*
33	Chr05: 112073406 112073407	*APC*
34	Chr05: 112073426 112073427	*APC*
35	Chr05: 112073613 112073614	*APC*
36	Chr05: 11904110 11904111	*CTNND2*
37	Chr05: 170738274 170738275	*TLX3*
38	Chr05: 170739179 170739180	*Non Coding Region*
39	Chr05: 38556435 38556436	*LIFR*
40	Chr05: 42423947 42423948	*GHR*
41	Chr05: 5139853 5139854	*ADAMTS16*
42	Chr05: 5139866 5139867	*ADAMTS16*
43	Chr06: 105584709 105584710	*BVES*
44	Chr06: 124124875 124124876	*NKAIN2*
45	Chr06: 1384491 1384492	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 1 Markers	
46	Chr06: 166582206 166582207	*T*
47	Chr07: 149411652 149411653	*KRBA1*
48	Chr07: 149412290 149412291	*KRBA1*
49	Chr07: 20817530 20817531	*Non Coding Region*
50	Chr07: 20817859 20817860	*Non Coding Region*
51	Chr07: 37487633 37487634	*ELMO1*
52	Chr07: 37488428 37488429	*ELMO1*
53	Chr07: 37960317 37960318	*EPDR1*
54	Chr07: 54609776 54609777	*VSTM2A*
55	Chr07: 87257074 87257075	*ABCB1,RUNDC3B*
56	Chr07: 93519855 93519856	*TFPI2*
57	Chr08: 41754181 41754182	*ANK1*
58	Chr08: 65492792 65492793	*BHLHE22*
59	Chr09: 117267242 117267243	*DFNB31*
60	Chr09: 124889019 124889020	*Non Coding Region*
61	Chr09: 96715256 96715257	*BARX1*
62	Chr10: 118893174 118893175	*VAX1*
63	Chr10: 126136607 126136608	*NKX1-2*
64	Chr10: 17496239 17496240	*ST8SIA6*
65	Chr10: 18429760 18429761	*CACNB2*
66	Chr10: 90342738 90342739	*RNLS*
67	Chr10: 90342899 90342900	*RNLS*
68	Chr10: 90343166 90343167	*RNLS*
69	Chr10: 90343174 90343175	*RNLS*
70	Chr10: 90343192 90343193	*RNLS*
71	Chr10: 90343204 90343205	*RNLS*
72	Chr10: 90343208 90343209	*RNLS*
73	Chr11: 126226007 126226008	*ST3GAL4*
74	Chr11: 126226033 126226034	*ST3GAL4*
75	Chr11: 128564756 128564757	*FLI1*
76	Chr11: 128564874 128564875	*FLI1*
77	Chr11: 94501718 94501719	*AMOTL1*
78	Chr11: 94502388 94502389	*AMOTL1*
79	Chr12: 113901210 113901211	*LHX5*
80	Chr12: 54338937 54338938	*HOXC13*
81	Chr12: 8542839 8542840	*LOC389634*
82	Chr13: 78493305 78493306	*EDNRB*
83	Chr13: 95620306 95620307	*Non Coding Region*
84	Chr14: 51561293 51561294	*TRIM9*
85	Chr14: 52735421 52735422	*PTGDR*
86	Chr14: 57283436 57283437	*OTX2OS1*
87	Chr15: 33010399 33010400	*GREM1*
88	Chr15: 45670478 45670479	*GATM,LOC145663*
89	Chr15: 76633627 76633628	*ISL2*
90	Chr15: 76633981 76633982	*ISL2*

Figure 59A

Order	Coordinate of CpG (hg19)	Gene
91	Chr15: 76634516 76634517	*ISL2*
92	Chr15: 78913147 78913148	*CHRNA3*
93	Chr16: 56224901 56224902	*GNAO1,LOC283856*
94	Chr16: 86599745 86599746	*FOXC2*
95	Chr17: 33701353 33701354	*SLFN11*
96	Chr17: 3438933 3438934	*TRPV3*
97	Chr17: 41832753 41832754	*SOST*
98	Chr17: 41832873 41832874	*SOST*
99	Chr17: 43339589 43339590	*C17orf46,LOC100133991*
100	Chr18: 32847566 32847567	*ZNF397OS*
101	Chr18: 5543548 5543549	*EPB41L3*
102	Chr19: 31841663 31841664	*TSHZ3*
103	Chr19: 36909413 36909414	*ZFP82*
104	Chr19: 37407284 37407285	*ZNF829,ZNF568*
105	Chr19: 37464066 37464067	*Non Coding Region*
106	Chr19: 38183055 38183056	*ZNF781*
107	Chr19: 38183259 38183260	*ZNF781*
108	Chr19: 38183262 38183263	*ZNF781*
109	Chr19: 51925128 51925129	*Non Coding Region*
110	Chr19: 53696649 53696650	*ZNF665*
111	Chr19: 56879559 56879560	*ZNF542*
112	Chr19: 56879645 56879646	*ZNF542*
113	Chr19: 56904901 56904902	*ZNF582*
114	Chr19: 56989543 56989544	*ZNF667*
115	Chr19: 57078765 57078766	*ZNF470*
116	Chr19: 57862627 57862628	*ZNF304*
117	Chr19: 57862638 57862639	*ZNF304*
118	Chr20: 13200980 13200981	*ISM1*
119	Chr20: 23029640 23029641	*THBD*
120	Chr20: 23029806 23029807	*THBD*
121	Chr20: 23030250 23030251	*THBD*
122	Chr20: 61638588 61638589	*BHLHE23*
123	Chr21: 38119946 38119947	*SIM2*
124	Chr21: 38120350 38120351	*SIM2*
125	Chr22: 45405627 45405628	*PHF21B*
	Alternative markers: High in CRC for Group 1 set	
126	Chr01: 101004638 101004639	*GPR88*
127	Chr01: 101004934 101004935	*GPR88*
128	Chr01: 101702602 101702603	*S1PR1*
129	Chr01: 107683532 107683533	*NTNG1*
130	Chr01: 108507078 108507079	*VAV3*
131	Chr01: 108507468 108507469	*VAV3*
132	Chr01: 108508067 108508068	*VAV3*
133	Chr01: 108508185 108508186	*VAV3*
134	Chr01: 108508207 108508208	*VAV3*

Order	Coordinate of CpG (hg19)	Gene
135	Chr01: 111217575 111217576	*KCNA3*
136	Chr01: 111217715 111217716	*KCNA3*
137	Chr01: 114696350 114696351	*SYT6*
138	Chr01: 115880865 115880866	*NGF*
139	Chr01: 115881044 115881045	*NGF*
140	Chr01: 119530600 119530601	*TBX15*
141	Chr01: 119530677 119530678	*TBX15*
142	Chr01: 12123262 12123263	*TNFRSF8*
143	Chr01: 12123308 12123309	*TNFRSF8*
144	Chr01: 12123313 12123314	*TNFRSF8*
145	Chr01: 121261404 121261405	*LOC647121*
146	Chr01: 121261602 121261603	*LOC647121*
147	Chr01: 13910209 13910210	*PDPN*
148	Chr01: 13910569 13910570	*PDPN*
149	Chr01: 13910698 13910699	*PDPN*
150	Chr01: 13910796 13910797	*PDPN*
151	Chr01: 150254869 150254870	*C1orf51*
152	Chr01: 154298335 154298336	*ATP8B2*
153	Chr01: 156626749 156626750	*BCAN*
154	Chr01: 156627099 156627100	*BCAN*
155	Chr01: 159141440 159141441	*CADM3*
156	Chr01: 16085371 16085372	*FBLIM1*
157	Chr01: 16085642 16085643	*FBLIM1*
158	Chr01: 165414379 165414380	*RXRG*
159	Chr01: 165414544 165414545	*RXRG*
160	Chr01: 171810376 171810377	*DNM3*
161	Chr01: 179712591 179712592	*FAM163A*
162	Chr01: 181287803 181287804	*Non Coding Region*
163	Chr01: 18958924 18958925	*PAX7*
164	Chr01: 196577709 196577710	*KCNT2*
165	Chr01: 207669716 207669717	*CR1*
166	Chr01: 208084415 208084416	*CD34*
167	Chr01: 208084436 208084437	*CD34*
168	Chr01: 208084456 208084457	*CD34*
169	Chr01: 224805287 224805288	*CNIH3*
170	Chr01: 228651908 228651909	*Non Coding Region*
171	Chr01: 229543554 229543555	*Non Coding Region*
172	Chr01: 231297103 231297104	*Non Coding Region*
173	Chr01: 234040833 234040834	*SLC35F3*
174	Chr01: 234350051 234350052	*SLC35F3*
175	Chr01: 237205999 237206000	*RYR2*
176	Chr01: 246952508 246952509	*LOC149134*
177	Chr01: 33219593 33219594	*KIAA1522*
178	Chr01: 33219687 33219688	*KIAA1522*
179	Chr01: 35395541 35395542	*Non Coding Region*

Figure 59B

Order	Coordinate of CpG (hg19)	Gene
180	Chr01: 35395663 35395664	*Non Coding Region*
181	Chr01: 35395821 35395822	*Non Coding Region*
182	Chr01: 3663531 3663532	*KIAA0495*
183	Chr01: 3663832 3663833	*KIAA0495*
184	Chr01: 37499547 37499548	*GRIK3*
185	Chr01: 38100811 38100812	*RSPO1*
186	Chr01: 38230779 38230780	*EPHA10*
187	Chr01: 38510933 38510934	*POU3F1*
188	Chr01: 39270149 39270150	*Non Coding Region*
189	Chr01: 44873057 44873058	*RNF220*
190	Chr01: 46951318 46951319	*Non Coding Region*
191	Chr01: 46951354 46951355	*Non Coding Region*
192	Chr01: 48190979 48190980	*Non Coding Region*
193	Chr01: 48191031 48191032	*Non Coding Region*
194	Chr01: 49242513 49242514	*AGBL4,BEND5*
195	Chr01: 49242519 49242520	*AGBL4,BEND5*
196	Chr01: 53099053 53099054	*FAM159A*
197	Chr01: 53099248 53099249	*FAM159A*
198	Chr01: 57888796 57888797	*DAB1*
199	Chr01: 57889035 57889036	*DAB1*
200	Chr01: 57889475 57889476	*DAB1*
201	Chr01: 57889604 57889605	*DAB1*
202	Chr01: 57890317 57890318	*DAB1*
203	Chr01: 58715499 58715500	*DAB1*
204	Chr01: 58715532 58715533	*DAB1*
205	Chr01: 58715553 58715554	*DAB1*
206	Chr01: 58716033 58716034	*DAB1*
207	Chr01: 67773719 67773720	*IL12RB2*
208	Chr01: 70034475 70034476	*Non Coding Region*
209	Chr01: 70034491 70034492	*Non Coding Region*
210	Chr01: 75601120 75601121	*LHX8*
211	Chr01: 76080727 76080728	*Non Coding Region*
212	Chr01: 77333138 77333139	*ST6GALNAC5*
213	Chr01: 77333159 77333160	*ST6GALNAC5*
214	Chr01: 77333198 77333199	*ST6GALNAC5*
215	Chr01: 77334045 77334046	*ST6GALNAC5*
216	Chr01: 86622628 86622629	*COL24A1*
217	Chr01: 98511629 98511630	*MIR137*
218	Chr01: 99470207 99470208	*LPPR5*
219	Chr01: 99470618 99470619	*LPPR5*
220	Chr02: 100938444 100938445	*LONRF2*
221	Chr02: 100938799 100938800	*LONRF2*
222	Chr02: 10220886 10220887	*CYS1*
223	Chr02: 105461368 105461369	*Non Coding Region*
224	Chr02: 108602906 108602907	*SLC5A7*

Order	Coordinate of CpG (hg19)	Gene
225	Chr02: 11052558 11052559	*KCNF1*
226	Chr02: 115920612 115920613	*DPP10*
227	Chr02: 119916017 119916018	*C1QL2*
228	Chr02: 120281813 120281814	*SCTR*
229	Chr02: 131721099 131721100	*ARHGEF4*
230	Chr02: 131792772 131792773	*ARHGEF4*
231	Chr02: 137522960 137522961	*Non Coding Region*
232	Chr02: 144694405 144694406	*Non Coding Region*
233	Chr02: 144695078 144695079	*Non Coding Region*
234	Chr02: 144695179 144695180	*Non Coding Region*
235	Chr02: 144695257 144695258	*Non Coding Region*
236	Chr02: 145274201 145274202	*ZEB2*
237	Chr02: 145274441 145274442	*ZEB2*
238	Chr02: 145275322 145275323	*ZEB2*
239	Chr02: 145277596 145277597	*ZEB2*
240	Chr02: 145281942 145281943	*Non Coding Region*
241	Chr02: 154728210 154728211	*GALNT13*
242	Chr02: 154729917 154729918	*GALNT13*
243	Chr02: 164593221 164593222	*FIGN*
244	Chr02: 171674855 171674856	*GAD1*
245	Chr02: 1748337 1748338	*PXDN*
246	Chr02: 177054306 177054307	*HOXD1*
247	Chr02: 182321786 182321787	*ITGA4*
248	Chr02: 182322268 182322269	*ITGA4*
249	Chr02: 182322279 182322280	*ITGA4*
250	Chr02: 182322501 182322502	*ITGA4*
251	Chr02: 182322749 182322750	*ITGA4*
252	Chr02: 182548807 182548808	*Non Coding Region*
253	Chr02: 185462397 185462398	*ZNF804A*
254	Chr02: 185462493 185462494	*ZNF804A*
255	Chr02: 187713902 187713903	*ZSWIM2*
256	Chr02: 187713986 187713987	*ZSWIM2*
257	Chr02: 193059230 193059231	*TMEFF2*
258	Chr02: 193059405 193059406	*TMEFF2*
259	Chr02: 193059645 193059646	*TMEFF2*
260	Chr02: 193059717 193059718	*TMEFF2*
261	Chr02: 207507096 207507097	*LOC200726*
262	Chr02: 210636398 210636399	*UNC80*
263	Chr02: 217236040 217236041	*MARCHF4*
264	Chr02: 217559867 217559868	*IGFBP5*
265	Chr02: 218621320 218621321	*DIRC3*
266	Chr02: 220174201 220174202	*PTPRN*
267	Chr02: 220299900 220299901	*SPEG*
268	Chr02: 223167400 223167401	*CCDC140*
269	Chr02: 223168453 223168454	*CCDC140*

Figure 59C

Order	Coordinate of CpG (hg19)	Gene
270	Chr02: 230578216 230578217	*DNER*
271	Chr02: 238535910 238535911	*LRRFIP1*
272	Chr02: 242743214 242743215	*GAL3ST2*
273	Chr02: 242743476 242743477	*GAL3ST2*
274	Chr02: 242743702 242743703	*Non Coding Region*
275	Chr02: 29337988 29337989	*CLIP4*
276	Chr02: 29338100 29338101	*CLIP4*
277	Chr02: 29338109 29338110	*CLIP4*
278	Chr02: 29338113 29338114	*CLIP4*
279	Chr02: 29338121 29338122	*CLIP4*
280	Chr02: 29338258 29338259	*CLIP4*
281	Chr02: 29338432 29338433	*CLIP4*
282	Chr02: 29339076 29339077	*CLIP4*
283	Chr02: 29339091 29339092	*CLIP4*
284	Chr02: 30144152 30144153	*ALK*
285	Chr02: 30144579 30144580	*ALK*
286	Chr02: 31360693 31360694	*GALNT14*
287	Chr02: 31360817 31360818	*GALNT14*
288	Chr02: 31457043 31457044	*EHD3*
289	Chr02: 31806042 31806043	*SRD5A2*
290	Chr02: 40678691 40678692	*SLC8A1*
291	Chr02: 40678760 40678761	*SLC8A1*
292	Chr02: 40679519 40679520	*SLC8A1*
293	Chr02: 40679582 40679583	*SLC8A1*
294	Chr02: 47797488 47797489	*KCNK12*
295	Chr02: 50574624 50574625	*NRXN1*
296	Chr02: 56150925 56150926	*EFEMP1*
297	Chr02: 5836111 5836112	*SOX11*
298	Chr02: 5836256 5836257	*SOX11*
299	Chr02: 66653254 66653255	*Non Coding Region*
300	Chr02: 66803033 66803034	*Non Coding Region*
301	Chr02: 68546467 68546468	*CNRIP1*
302	Chr02: 68546507 68546508	*CNRIP1*
303	Chr02: 68546579 68546580	*CNRIP1*
304	Chr02: 68546899 68546900	*CNRIP1*
305	Chr02: 68547052 68547053	*CNRIP1*
306	Chr02: 68547088 68547089	*CNRIP1*
307	Chr02: 7058390 7058391	*RNF144A*
308	Chr02: 7058432 7058433	*RNF144A*
309	Chr02: 71693389 71693390	*DYSF*
310	Chr02: 73430374 73430375	*NOTO*
311	Chr02: 87017591 87017592	*CD8A*
312	Chr02: 946648 946649	*SNTG2*
313	Chr02: 95691255 95691256	*MAL*
314	Chr02: 97193174 97193175	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
315	Chr02: 97193524 97193525	*Non Coding Region*
316	Chr02: 98962900 98962901	*CNGA3*
317	Chr02: 98963134 98963135	*CNGA3*
318	Chr02: 98963768 98963769	*CNGA3*
319	Chr03: 10857798 10857799	*SLC6A11*
320	Chr03: 10858003 10858004	*SLC6A11*
321	Chr03: 120169783 120169784	*FSTL1*
322	Chr03: 120169821 120169822	*FSTL1*
323	Chr03: 128210000 128210001	*GATA2*
324	Chr03: 128210888 128210889	*GATA2*
325	Chr03: 128211096 128211097	*GATA2*
326	Chr03: 128211540 128211541	*GATA2*
327	Chr03: 128211686 128211687	*GATA2*
328	Chr03: 128212269 128212270	*GATA2*
329	Chr03: 128212273 128212274	*GATA2*
330	Chr03: 128720433 128720434	*CCDC48*
331	Chr03: 13114803 13114804	*IQSEC1*
332	Chr03: 132757087 132757088	*TMEM108*
333	Chr03: 132757090 132757091	*TMEM108*
334	Chr03: 132757102 132757103	*TMEM108*
335	Chr03: 132757106 132757107	*TMEM108*
336	Chr03: 132757267 132757268	*TMEM108*
337	Chr03: 13324132 13324133	*Non Coding Region*
338	Chr03: 134369919 134369920	*KY*
339	Chr03: 134369939 134369940	*KY*
340	Chr03: 134515856 134515857	*EPHB1*
341	Chr03: 137480102 137480103	*Non Coding Region*
342	Chr03: 137486265 137486266	*Non Coding Region*
343	Chr03: 137487932 137487933	*Non Coding Region*
344	Chr03: 140770549 140770550	*SPSB4*
345	Chr03: 140770588 140770589	*SPSB4*
346	Chr03: 140770599 140770600	*SPSB4*
347	Chr03: 140770608 140770609	*SPSB4*
348	Chr03: 140771786 140771787	*SPSB4*
349	Chr03: 142838847 142838848	*CHST2*
350	Chr03: 142838938 142838939	*CHST2*
351	Chr03: 142839022 142839023	*CHST2*
352	Chr03: 142839578 142839579	*CHST2*
353	Chr03: 142839837 142839838	*CHST2*
354	Chr03: 142839903 142839904	*CHST2*
355	Chr03: 142839991 142839992	*CHST2*
356	Chr03: 147127143 147127144	*ZIC1*
357	Chr03: 14852659 14852660	*Non Coding Region*
358	Chr03: 159756793 159756794	*Non Coding Region*
359	Chr03: 159756929 159756930	*Non Coding Region*

Figure 59D

Order	Coordinate of CpG (hg19)	Gene
360	Chr03:_173303193_173303194	*NLGN1*
361	Chr03:_179754239_179754240	*PEX5L*
362	Chr03:_179754521_179754522	*PEX5L*
363	Chr03:_179754533_179754534	*PEX5L*
364	Chr03:_179754613_179754614	*PEX5L*
365	Chr03:_179755086_179755087	*PEX5L*
366	Chr03:_185080905_185080906	*MAP3K13*
367	Chr03:_185080997_185080998	*MAP3K13*
368	Chr03:_186079142_186079143	*DGKG*
369	Chr03:_192126742_192126743	*FGF12*
370	Chr03:_192126849_192126850	*FGF12*
371	Chr03:_192126935_192126936	*FGF12*
372	Chr03:_192126960_192126961	*FGF12*
373	Chr03:_194408516_194408517	*FAM43A*
374	Chr03:_238392_238393	*CHL1*
375	Chr03:_27771766_27771767	*Non Coding Region*
376	Chr03:_28616682_28616683	*Non Coding Region*
377	Chr03:_32859407_32859408	*TRIM71*
378	Chr03:_32859417_32859418	*TRIM71*
379	Chr03:_32859438_32859439	*TRIM71*
380	Chr03:_33259889_33259890	*SUSD5*
381	Chr03:_36422209_36422210	*STAC*
382	Chr03:_38036034_38036035	*VILL*
383	Chr03:_44037128_44037129	*Non Coding Region*
384	Chr03:_44037189_44037190	*Non Coding Region*
385	Chr03:_44626538_44626539	*ZNF660*
386	Chr03:_46506292_46506293	*LTF*
387	Chr03:_46506481_46506482	*LTF*
388	Chr03:_46506483_46506484	*LTF*
389	Chr03:_46923937_46923938	*PTH1R*
390	Chr03:_48693882_48693883	*CELSR3*
391	Chr03:_48694018_48694019	*CELSR3*
392	Chr03:_48699012_48699013	*CELSR3*
393	Chr03:_62359392_62359393	*FEZF2*
394	Chr03:_68981011_68981012	*FAM19A4*
395	Chr03:_68981852_68981853	*FAM19A4*
396	Chr03:_68981854_68981855	*FAM19A4*
397	Chr03:_68981866_68981867	*FAM19A4*
398	Chr03:_68981877_68981878	*FAM19A4*
399	Chr03:_68981890_68981891	*FAM19A4*
400	Chr03:_69591803_69591804	*Non Coding Region*
401	Chr03:_69592049_69592050	*Non Coding Region*
402	Chr03:_69592090_69592091	*Non Coding Region*
403	Chr03:_71802594_71802595	*EIF4E3,GPR27*
404	Chr03:_79816949_79816950	*ROBO1*

Order	Coordinate of CpG (hg19)	Gene
405	Chr04:_107955762_107955763	*DKK2*
406	Chr04:_107956955_107956956	*DKK2*
407	Chr04:_107957362_107957363	*DKK2*
408	Chr04:_110223967_110223968	*COL25A1*
409	Chr04:_110223969_110223970	*COL25A1*
410	Chr04:_111562545_111562546	*Non Coding Region*
411	Chr04:_122301740_122301741	*ORFPR*
412	Chr04:_122301816_122301817	*ORFPR*
413	Chr04:_122302226_122302227	*ORFPR*
414	Chr04:_122686038_122686039	*TMEM155,LOC100192379*
415	Chr04:_122686269_122686270	*TMEM155,LOC100192379*
416	Chr04:_122686432_122686433	*TMEM155,LOC100192379*
417	Chr04:_122686453_122686454	*TMEM155,LOC100192379*
418	Chr04:_122686456_122686457	*TMEM155,LOC100192379*
419	Chr04:_122686493_122686494	*TMEM155,LOC100192379*
420	Chr04:_122686564_122686565	*TMEM155,LOC100192379*
421	Chr04:_126237371_126237372	*FAT4*
422	Chr04:_126237373_126237374	*FAT4*
423	Chr04:_134067789_134067790	*Non Coding Region*
424	Chr04:_134068777_134068778	*Non Coding Region*
425	Chr04:_134069670_134069671	*PCDH10*
426	Chr04:_134070369_134070370	*PCDH10*
427	Chr04:_134070389_134070390	*PCDH10*
428	Chr04:_134070416_134070417	*PCDH10*
429	Chr04:_134070437_134070438	*PCDH10*
430	Chr04:_13537872_13537873	*Non Coding Region*
431	Chr04:_142053623_142053624	*RNF150*
432	Chr04:_142054254_142054255	*RNF150*
433	Chr04:_144621898_144621899	*FREM3*
434	Chr04:_144621945_144621946	*FREM3*
435	Chr04:_144621952_144621953	*FREM3*
436	Chr04:_144621971_144621972	*FREM3*
437	Chr04:_148402574_148402575	*EDNRA*
438	Chr04:_154680909_154680910	*RNF175*
439	Chr04:_154681197_154681198	*RNF175*
440	Chr04:_154710475_154710476	*SFRP2*
441	Chr04:_154710598_154710599	*SFRP2*
442	Chr04:_154712422_154712423	*Non Coding Region*
443	Chr04:_156297854_156297855	*MAP9*
444	Chr04:_156297942_156297943	*MAP9*
445	Chr04:_156680400_156680401	*GUCY1B3*
446	Chr04:_156680769_156680770	*GUCY1B3*
447	Chr04:_156681047_156681048	*GUCY1B3*
448	Chr04:_157997526_157997527	*GLRB*
449	Chr04:_166794971_166794972	*TLL1*

Figure 59E

Order	Coordinate of CpG (hg19)	Gene
450	Chr04:_166795260_166795261	*TLL1*
451	Chr04:_166795524_166795525	*TLL1*
452	Chr04:_168155219_168155220	*SPOCK3*
453	Chr04:_175139081_175139082	*Non Coding Region*
454	Chr04:_175750637_175750638	*GLRA3*
455	Chr04:_176922779_176922780	*GPM6A*
456	Chr04:_176986950_176986951	*WDR17*
457	Chr04:_176987020_176987021	*WDR17*
458	Chr04:_176987174_176987175	*WDR17*
459	Chr04:_176987313_176987314	*WDR17*
460	Chr04:_17783294_17783295	*FAM184B*
461	Chr04:_17783322_17783323	*FAM184B*
462	Chr04:_17783331_17783332	*FAM184B*
463	Chr04:_184826704_184826705	*STOX2*
464	Chr04:_184827754_184827755	*STOX2*
465	Chr04:_185941601_185941602	*HELT*
466	Chr04:_40858984_40858985	*APBB2*
467	Chr04:_40859249_40859250	*APBB2*
468	Chr04:_40859251_40859252	*APBB2*
469	Chr04:_41883164_41883165	*Non Coding Region*
470	Chr04:_41883879_41883880	*Non Coding Region*
471	Chr04:_41883890_41883891	*Non Coding Region*
472	Chr04:_42154140_42154141	*BEND4*
473	Chr04:_42154840_42154841	*BEND4*
474	Chr04:_4388505_4388506	*D4S234E*
475	Chr04:_5052526_5052527	*STK32B*
476	Chr04:_5053341_5053342	*STK32B*
477	Chr04:_5053496_5053497	*STK32B*
478	Chr04:_5053504_5053505	*STK32B*
479	Chr04:_55097476_55097477	*PDGFRA*
480	Chr04:_55097576_55097577	*PDGFRA*
481	Chr04:_55098359_55098360	*PDGFRA*
482	Chr04:_55991782_55991783	*KDR*
483	Chr04:_55991850_55991851	*KDR*
484	Chr04:_55991860_55991861	*KDR*
485	Chr04:_55991943_55991944	*KDR*
486	Chr04:_570361_570362	*Non Coding Region*
487	Chr04:_5710319_5710320	*EVC2*
488	Chr04:_5710407_5710408	*EVC2*
489	Chr04:_5710411_5710412	*EVC2*
490	Chr04:_5712937_5712938	*EVC*
491	Chr04:_5713100_5713101	*EVC*
492	Chr04:_5892072_5892073	*CRMP1*
493	Chr04:_5892208_5892209	*CRMP1*
494	Chr04:_62065809_62065810	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
495	Chr04:_7195206_7195207	*SORCS2*
496	Chr04:_81186901_81186902	*FGF5*
497	Chr04:_81187125_81187126	*FGF5*
498	Chr04:_81187798_81187799	*FGF5*
499	Chr04:_81187906_81187907	*FGF5*
500	Chr04:_96470887_96470888	*UNC5C*
501	Chr04:_96471105_96471106	*UNC5C*
502	Chr05:_100238819_100238820	*ST8SIA4*
503	Chr05:_127873980_127873981	*FBN2*
504	Chr05:_128300708_128300709	*SLC27A6*
505	Chr05:_128301083_128301084	*SLC27A6*
506	Chr05:_135265931_135265932	*FBXL21*
507	Chr05:_136834464_136834465	*SPOCK1*
508	Chr05:_136834492_136834493	*SPOCK1*
509	Chr05:_136834877_136834878	*SPOCK1*
510	Chr05:_140306208_140306209	*PCDHA2,PCDHA3,PCDHA1, etc.*
511	Chr05:_140892370_140892371	*PCDHGB3,PCDHGB2,PCDHGB1, etc.*
512	Chr05:_141262801_141262802	*Non Coding Region*
513	Chr05:_1445561_1445562	*SLC6A3*
514	Chr05:_1445567_1445568	*SLC6A3*
515	Chr05:_146257484_146257485	*PPP2R2B*
516	Chr05:_146258463_146258464	*PPP2R2B*
517	Chr05:_146258512_146258513	*PPP2R2B*
518	Chr05:_146258522_146258523	*PPP2R2B*
519	Chr05:_146258546_146258547	*PPP2R2B*
520	Chr05:_1667643_1667644	*Non Coding Region*
521	Chr05:_168728270_168728271	*SLIT3*
522	Chr05:_172661859_172661860	*NKX2-5*
523	Chr05:_172670879_172670880	*Non Coding Region*
524	Chr05:_172671526_172671527	*Non Coding Region*
525	Chr05:_174115696_174115697	*Non Coding Region*
526	Chr05:_174871289_174871290	*DRD1*
527	Chr05:_175792973_175792974	*ARL10*
528	Chr05:_178017571_178017572	*COL23A1*
529	Chr05:_178017578_178017579	*COL23A1*
530	Chr05:_178771776_178771777	*ADAMTS2*
531	Chr05:_178771781_178771782	*ADAMTS2*
532	Chr05:_178772203_178772204	*ADAMTS2*
533	Chr05:_180017349_180017350	*SCGB3A1*
534	Chr05:_180100963_180100964	*Non Coding Region*
535	Chr05:_1883005_1883006	*IRX4*
536	Chr05:_1883214_1883215	*IRX4*
537	Chr05:_1883940_1883941	*IRX4*
538	Chr05:_1931620_1931621	*Non Coding Region*
539	Chr05:_19988288_19988289	*CDH18*

Figure 59F

Order	Coordinate of CpG (hg19)	Gene
540	Chr05: 19988366 19988367	*CDH18*
541	Chr05: 19988368 19988369	*CDH18*
542	Chr05: 3197730 3197731	*Non Coding Region*
543	Chr05: 32711429 32711430	*NPR3*
544	Chr05: 32713290 32713291	*NPR3*
545	Chr05: 32713723 32713724	*NPR3*
546	Chr05: 33892223 33892224	*ADAMTS12*
547	Chr05: 33936232 33936233	*RXFP3*
548	Chr05: 33936253 33936254	*RXFP3*
549	Chr05: 33936307 33936308	*RXFP3*
550	Chr05: 33936322 33936323	*RXFP3*
551	Chr05: 33936362 33936363	*RXFP3*
552	Chr05: 3595728 3595729	*IRX1*
553	Chr05: 36066972 36066973	*UGT3A2*
554	Chr05: 37836748 37836749	*GDNF*
555	Chr05: 37838563 37838564	*GDNF*
556	Chr05: 37839829 37839830	*GDNF*
557	Chr05: 37840075 37840076	*GDNF*
558	Chr05: 38258333 38258334	*EGFLAM*
559	Chr05: 38258884 38258885	*EGFLAM*
560	Chr05: 38556435 38556436	*LIFR*
561	Chr05: 44389406 44389407	*FGF10*
562	Chr05: 44389848 44389849	*FGF10*
563	Chr05: 49736988 49736989	*EMB*
564	Chr05: 49737182 49737183	*EMB*
565	Chr05: 50265443 50265444	*Non Coding Region*
566	Chr05: 5139853 5139854	*ADAMTS16*
567	Chr05: 5139855 5139856	*ADAMTS16*
568	Chr05: 5139866 5139867	*ADAMTS16*
569	Chr05: 63802280 63802281	*RGS7BP*
570	Chr05: 63802491 63802492	*RGS7BP*
571	Chr05: 63986392 63986393	*FAM159B*
572	Chr05: 63986656 63986657	*FAM159B*
573	Chr05: 63986793 63986794	*FAM159B*
574	Chr05: 71015523 71015524	*CARTPT*
575	Chr05: 76249776 76249777	*CRHBP*
576	Chr05: 82768488 82768489	*VCAN*
577	Chr06: 108486007 108486008	*NR2E1*
578	Chr06: 10887185 10887186	*SYCP2L*
579	Chr06: 10887587 10887588	*SYCP2L*
580	Chr06: 11044106 11044107	*ELOVL2*
581	Chr06: 110679107 110679108	*C6orf186*
582	Chr06: 110797233 110797234	*SLC22A16*
583	Chr06: 110797497 110797498	*SLC22A16*
584	Chr06: 110797506 110797507	*SLC22A16*

Order	Coordinate of CpG (hg19)	Gene
585	Chr06: 117086479 117086480	*FAM162B*
586	Chr06: 117086850 117086851	*FAM162B*
587	Chr06: 117086931 117086932	*FAM162B*
588	Chr06: 118228869 118228870	*SLC35F1*
589	Chr06: 127440000 127440001	*RSPO3*
590	Chr06: 127440510 127440511	*RSPO3*
591	Chr06: 133562035 133562036	*EYA4*
592	Chr06: 133562056 133562057	*EYA4*
593	Chr06: 133562193 133562194	*EYA4*
594	Chr06: 133562246 133562247	*EYA4*
595	Chr06: 133562461 133562462	*EYA4*
596	Chr06: 134211114 134211115	*TCF21*
597	Chr06: 134638871 134638872	*SGK1*
598	Chr06: 1391265 1391266	*FOXF2*
599	Chr06: 151562026 151562027	*AKAP12*
600	Chr06: 152128537 152128538	*ESR1*
601	Chr06: 152129400 152129401	*ESR1*
602	Chr06: 152957910 152957911	*SYNE1*
603	Chr06: 152957999 152958000	*SYNE1*
604	Chr06: 159590231 159590232	*FNDC1*
605	Chr06: 1625238 1625239	*GMDS*
606	Chr06: 1625457 1625458	*GMDS*
607	Chr06: 163836718 163836719	*OK1*
608	Chr06: 166580952 166580953	*T*
609	Chr06: 166666838 166666839	*Non Coding Region*
610	Chr06: 166667504 166667505	*Non Coding Region*
611	Chr06: 28411030 28411031	*ZSCAN23*
612	Chr06: 28411240 28411241	*ZSCAN23*
613	Chr06: 28411271 28411272	*ZSCAN23*
614	Chr06: 28411423 28411424	*ZSCAN23*
615	Chr06: 28510339 28510340	*Non Coding Region*
616	Chr06: 28603059 28603060	*Non Coding Region*
617	Chr06: 28677272 28677273	*Non Coding Region*
618	Chr06: 28793219 28793220	*Non Coding Region*
619	Chr06: 29760410 29760411	*HCG4*
620	Chr06: 29760505 29760506	*HCG4*
621	Chr06: 29760832 29760833	*HCG4*
622	Chr06: 30095762 30095763	*Non Coding Region*
623	Chr06: 30418789 30418790	*Non Coding Region*
624	Chr06: 30419070 30419071	*Non Coding Region*
625	Chr06: 31783455 31783456	*HSPA1A,HSPA1L*
626	Chr06: 34112818 34112819	*Non Coding Region*
627	Chr06: 3752849 3752850	*C6orf145*
628	Chr06: 391189 391190	*IRF4*
629	Chr06: 391208 391209	*IRF4*

Figure 59G

Order	Coordinate of CpG (hg19)	Gene
630	Chr06: 391743 391744	*IRF4*
631	Chr06: 393239 393240	*IRF4*
632	Chr06: 393616 393617	*IRF4*
633	Chr06: 41606317 41606318	*MDFI*
634	Chr06: 41606770 41606771	*MDFI*
635	Chr06: 44529081 44529082	*Non Coding Region*
636	Chr06: 56716884 56716885	*DST*
637	Chr06: 62995523 62995524	*KHDRBS2*
638	Chr06: 62996130 62996131	*KHDRBS2*
639	Chr06: 62996183 62996184	*KHDRBS2*
640	Chr06: 73331114 73331115	*KCNQ5*
641	Chr06: 73331238 73331239	*KCNQ5*
642	Chr06: 73331290 73331291	*KCNQ5*
643	Chr06: 73331405 73331406	*KCNQ5*
644	Chr06: 73332073 73332074	*KCNQ5*
645	Chr06: 84417933 84417934	*SNAP91*
646	Chr06: 84418724 84418725	*SNAP91*
647	Chr06: 84419202 84419203	*SNAP91*
648	Chr06: 84419329 84419330	*SNAP91*
649	Chr06: 84562892 84562893	*RIPPLY2*
650	Chr06: 85473773 85473774	*TBX18*
651	Chr06: 85484717 85484718	*Non Coding Region*
652	Chr06: 87647370 87647371	*HTR1E*
653	Chr06: 91321019 91321020	*Non Coding Region*
654	Chr06: 94127427 94127428	*EPHA7*
655	Chr07: 101006052 101006053	*EMID2*
656	Chr07: 101006063 101006064	*EMID2*
657	Chr07: 103630086 103630087	*RELN*
658	Chr07: 108097436 108097437	*NRCAM*
659	Chr07: 112726538 112726539	*GPR85*
660	Chr07: 1272512 1272513	*UNCX*
661	Chr07: 1272710 1272711	*UNCX*
662	Chr07: 127744553 127744554	*Non Coding Region*
663	Chr07: 142494595 142494596	*Non Coding Region*
664	Chr07: 142494773 142494774	*Non Coding Region*
665	Chr07: 142494914 142494915	*Non Coding Region*
666	Chr07: 142494953 142494954	*Non Coding Region*
667	Chr07: 142494985 142494986	*Non Coding Region*
668	Chr07: 149411652 149411653	*KRBA1*
669	Chr07: 149412290 149412291	*KRBA1*
670	Chr07: 150653001 150653002	*KCNH2*
671	Chr07: 150710730 150710731	*ATG9B,NOS3*
672	Chr07: 151107257 151107258	*WDR86*
673	Chr07: 151107261 151107262	*WDR86*
674	Chr07: 151107285 151107286	*WDR86*

Order	Coordinate of CpG (hg19)	Gene
675	Chr07: 151107400 151107401	*WDR86*
676	Chr07: 151107637 151107638	*WDR86*
677	Chr07: 152622455 152622456	*Non Coding Region*
678	Chr07: 152622815 152622816	*Non Coding Region*
679	Chr07: 155166290 155166291	*Non Coding Region*
680	Chr07: 156871234 156871235	*Non Coding Region*
681	Chr07: 157482508 157482509	*PTPRN2*
682	Chr07: 157485230 157485231	*PTPRN2*
683	Chr07: 158937610 158937611	*VIPR2*
684	Chr07: 158938060 158938061	*VIPR2*
685	Chr07: 19147157 19147158	*Non Coding Region*
686	Chr07: 19156086 19156087	*TWIST1*
687	Chr07: 19157193 19157194	*TWIST1*
688	Chr07: 19157263 19157264	*TWIST1*
689	Chr07: 19157296 19157297	*TWIST1*
690	Chr07: 19157304 19157305	*TWIST1*
691	Chr07: 19157634 19157635	*TWIST1*
692	Chr07: 19157938 19157939	*TWIST1*
693	Chr07: 19158001 19158002	*TWIST1*
694	Chr07: 19812570 19812571	*TMEM196*
695	Chr07: 21583304 21583305	*DNAH11*
696	Chr07: 24323675 24323676	*NPY*
697	Chr07: 24323764 24323765	*NPY*
698	Chr07: 28449847 28449848	*CREB5*
699	Chr07: 30721888 30721889	*CRHR2*
700	Chr07: 30722114 30722115	*CRHR2*
701	Chr07: 32338150 32338151	*Non Coding Region*
702	Chr07: 37487633 37487634	*ELMO1*
703	Chr07: 38670412 38670413	*AMPH*
704	Chr07: 38670804 38670805	*AMPH*
705	Chr07: 38670957 38670958	*AMPH*
706	Chr07: 38670985 38670986	*AMPH*
707	Chr07: 38670987 38670988	*AMPH*
708	Chr07: 38671001 38671002	*AMPH*
709	Chr07: 41739742 41739743	*INHBA,LOC285954*
710	Chr07: 43153223 43153224	*HECW1*
711	Chr07: 43153595 43153596	*HECW1*
712	Chr07: 44143993 44143994	*AEBP1*
713	Chr07: 44143998 44143999	*AEBP1*
714	Chr07: 44144434 44144435	*AEBP1*
715	Chr07: 45197441 45197442	*RAMP3*
716	Chr07: 49813031 49813032	*VWC2*
717	Chr07: 49815365 49815366	*VWC2*
718	Chr07: 49815502 49815503	*VWC2*
719	Chr07: 4998470 4998471	*MMD2*

Figure 59H

Order	Coordinate of CpG (hg19)	Gene
720	Chr07: 50343361 50343362	*IKZF1*
721	Chr07: 50343883 50343884	*IKZF1*
722	Chr07: 54609776 54609777	*VSTM2A*
723	Chr07: 69064801 69064802	*AUTS2*
724	Chr07: 71800987 71800988	*CALN1*
725	Chr07: 71801793 71801794	*CALN1*
726	Chr07: 752149 752150	*PRKAR1B*
727	Chr07: 89748294 89748295	*DPY19L2P4*
728	Chr07: 90226284 90226285	*Non Coding Region*
729	Chr07: 93519473 93519474	*TFPI2*
730	Chr07: 93519855 93519856	*TFPI2*
731	Chr07: 93520012 93520013	*TFPI2*
732	Chr07: 93520024 93520025	*TFPI2*
733	Chr07: 93520183 93520184	*TFPI2*
734	Chr07: 93520269 93520270	*TFPI2*
735	Chr07: 93520275 93520276	*TFPI2*
736	Chr07: 93520288 93520289	*TFPI2*
737	Chr07: 98467380 98467381	*TMEM130*
738	Chr08: 104513083 104513084	*RIMS2*
739	Chr08: 104513909 104513910	*RIMS2*
740	Chr08: 105235606 105235607	*RIMS2*
741	Chr08: 105235943 105235944	*RIMS2*
742	Chr08: 109094581 109094582	*RSPO2*
743	Chr08: 109094880 109094881	*RSPO2*
744	Chr08: 109095782 109095783	*RSPO2*
745	Chr08: 11204681 11204682	*TDH*
746	Chr08: 11205166 11205167	*TDH*
747	Chr08: 116660697 116660698	*TRPS1*
748	Chr08: 121137288 121137289	*COL14A1*
749	Chr08: 140715557 140715558	*KCNK9*
750	Chr08: 140715646 140715647	*KCNK9*
751	Chr08: 143533499 143533500	*Non Coding Region*
752	Chr08: 143533532 143533533	*Non Coding Region*
753	Chr08: 143591819 143591820	*BAI1*
754	Chr08: 143858612 143858613	*LYNX1*
755	Chr08: 143858636 143858637	*LYNX1*
756	Chr08: 144241434 144241435	*LY6H*
757	Chr08: 15397729 15397730	*TUSC3*
758	Chr08: 16884364 16884365	*EFHA2*
759	Chr08: 16885270 16885271	*EFHA2*
760	Chr08: 17270604 17270605	*MTMR7*
761	Chr08: 22960820 22960821	*TNFRSF10C*
762	Chr08: 23572029 23572030	*Non Coding Region*
763	Chr08: 24770342 24770343	*NEFM*
764	Chr08: 24814137 24814138	*NEFL*

Order	Coordinate of CpG (hg19)	Gene
765	Chr08: 25907218 25907219	*Non Coding Region*
766	Chr08: 25909599 25909600	*Non Coding Region*
767	Chr08: 26723584 26723585	*ADRA1A*
768	Chr08: 32405792 32405793	*NRG1*
769	Chr08: 32406928 32406929	*NRG1*
770	Chr08: 37823409 37823410	*ADRB3*
771	Chr08: 38645408 38645409	*TACC1*
772	Chr08: 41167107 41167108	*SFRP1*
773	Chr08: 41754172 41754173	*ANK1*
774	Chr08: 41754181 41754182	*ANK1*
775	Chr08: 494721 494722	*C8orf42*
776	Chr08: 53852184 53852185	*NPBWR1*
777	Chr08: 53852422 53852423	*NPBWR1*
778	Chr08: 53852443 53852444	*NPBWR1*
779	Chr08: 53852446 53852447	*NPBWR1*
780	Chr08: 53853604 53853605	*Non Coding Region*
781	Chr08: 54164161 54164162	*OPRK1*
782	Chr08: 54164310 54164311	*OPRK1*
783	Chr08: 55367466 55367467	*Non Coding Region*
784	Chr08: 55367611 55367612	*Non Coding Region*
785	Chr08: 55372111 55372112	*SOX17*
786	Chr08: 55379034 55379035	*Non Coding Region*
787	Chr08: 57069688 57069689	*Non Coding Region*
788	Chr08: 58907821 58907822	*FAM110B*
789	Chr08: 60032607 60032608	*TOX*
790	Chr08: 65711369 65711370	*CYP7B1*
791	Chr08: 67344553 67344554	*ADHFE1*
792	Chr08: 67344556 67344557	*ADHFE1*
793	Chr08: 67344588 67344589	*ADHFE1*
794	Chr08: 67344642 67344643	*ADHFE1*
795	Chr08: 67344665 67344666	*ADHFE1*
796	Chr08: 67344720 67344721	*ADHFE1*
797	Chr08: 67345006 67345007	*ADHFE1*
798	Chr08: 67875033 67875034	*Non Coding Region*
799	Chr08: 67875261 67875262	*Non Coding Region*
800	Chr08: 67875441 67875442	*Non Coding Region*
801	Chr08: 687384 687385	*Non Coding Region*
802	Chr08: 688018 688019	*Non Coding Region*
803	Chr08: 688417 688418	*Non Coding Region*
804	Chr08: 69243510 69243511	*Non Coding Region*
805	Chr08: 69244510 69244511	*Non Coding Region*
806	Chr08: 72755784 72755785	*MSC*
807	Chr08: 72756058 72756059	*MSC*
808	Chr08: 72756155 72756156	*MSC*
809	Chr08: 72756341 72756342	*MSC*

Figure 59I

Order	Coordinate of CpG (hg19)	Gene
810	Chr08: 72756812 72756813	*MSC*
811	Chr08: 72756839 72756840	*MSC*
812	Chr08: 72756863 72756864	*MSC*
813	Chr08: 73163696 73163697	*Non Coding Region*
814	Chr08: 75897297 75897298	*CRISPLD1*
815	Chr08: 77594256 77594257	*ZFHX4,LOC100192378*
816	Chr08: 77594595 77594596	*ZFHX4,LOC100192378*
817	Chr08: 85095498 85095499	*RALYL*
818	Chr08: 91997810 91997811	*Non Coding Region*
819	Chr08: 97157756 97157757	*GDF6*
820	Chr08: 97172961 97172962	*GDF6*
821	Chr08: 97173526 97173527	*GDF6*
822	Chr08: 97505764 97505765	*SDC2*
823	Chr08: 97506251 97506252	*SDC2*
824	Chr08: 9761171 9761172	*MIR124-1,LOC157627*
825	Chr08: 9762716 9762717	*Non Coding Region*
826	Chr08: 9762850 9762851	*Non Coding Region*
827	Chr08: 9763558 9763559	*Non Coding Region*
828	Chr08: 9763739 9763740	*Non Coding Region*
829	Chr08: 9763877 9763878	*Non Coding Region*
830	Chr09: 100615201 100615202	*FOXE1*
831	Chr09: 100616469 100616470	*FOXE1*
832	Chr09: 101706617 101706618	*COL15A1*
833	Chr09: 112403301 112403302	*PALM2*
834	Chr09: 117267242 117267243	*DFNB31*
835	Chr09: 129485542 129485543	*Non Coding Region*
836	Chr09: 132382433 132382434	*C9orf50*
837	Chr09: 132382812 132382813	*C9orf50*
838	Chr09: 133536345 133536346	*Non Coding Region*
839	Chr09: 137299285 137299286	*RXRA*
840	Chr09: 139090991 139090992	*LHX3*
841	Chr09: 37027384 37027385	*PAX5*
842	Chr09: 71788748 71788749	*TJP2*
843	Chr09: 71788759 71788760	*TJP2*
844	Chr09: 77112896 77112897	*RORB*
845	Chr09: 841850 841851	*DMRT1*
846	Chr09: 88137909 88137910	*Non Coding Region*
847	Chr10: 100992656 100992657	*HPSE2*
848	Chr10: 100993826 100993827	*HPSE2*
849	Chr10: 102484232 102484233	*Non Coding Region*
850	Chr10: 102504039 102504040	*PAX2*
851	Chr10: 102504226 102504227	*PAX2*
852	Chr10: 102508286 102508287	*PAX2*
853	Chr10: 103603869 103603870	*KCNIP2*
854	Chr10: 105036611 105036612	*INA*

Order	Coordinate of CpG (hg19)	Gene
855	Chr10: 105036701 105036702	*INA*
856	Chr10: 105036727 105036728	*INA*
857	Chr10: 111216426 111216427	*Non Coding Region*
858	Chr10: 11206792 11206793	*CUGBP2*
859	Chr10: 11206813 11206814	*CUGBP2*
860	Chr10: 11206838 11206839	*CUGBP2*
861	Chr10: 11206892 11206893	*CUGBP2*
862	Chr10: 11207437 11207438	*CUGBP2*
863	Chr10: 11207907 11207908	*CUGBP2*
864	Chr10: 11207969 11207970	*CUGBP2*
865	Chr10: 118031654 118031655	*GFRA1*
866	Chr10: 118031864 118031865	*GFRA1*
867	Chr10: 118031870 118031871	*GFRA1*
868	Chr10: 118031950 118031951	*GFRA1*
869	Chr10: 118032081 118032082	*GFRA1*
870	Chr10: 118032330 118032331	*GFRA1*
871	Chr10: 118032626 118032627	*GFRA1*
872	Chr10: 118032879 118032880	*GFRA1*
873	Chr10: 118032892 118032893	*GFRA1*
874	Chr10: 118032905 118032906	*GFRA1*
875	Chr10: 118034031 118034032	*GFRA1*
876	Chr10: 118892423 118892424	*VAX1*
877	Chr10: 119000927 119000928	*SLC18A2*
878	Chr10: 124895886 124895887	*HMX3*
879	Chr10: 129535786 129535787	*FOXI2*
880	Chr10: 131768098 131768099	*Non Coding Region*
881	Chr10: 133796164 133796165	*BNIP3*
882	Chr10: 134000124 134000125	*DPYSL4*
883	Chr10: 134901294 134901295	*GPR123*
884	Chr10: 134901297 134901298	*GPR123*
885	Chr10: 135043492 135043493	*UTF1*
886	Chr10: 135043506 135043507	*UTF1*
887	Chr10: 13933996 13933997	*FRMD4A*
888	Chr10: 17271051 17271052	*VIM*
889	Chr10: 1778844 1778845	*ADARB2*
890	Chr10: 1779835 1779836	*ADARB2*
891	Chr10: 23216695 23216696	*ARMC3*
892	Chr10: 23463094 23463095	*Non Coding Region*
893	Chr10: 23488710 23488711	*Non Coding Region*
894	Chr10: 23982480 23982481	*KIAA1217*
895	Chr10: 23983990 23983991	*KIAA1217*
896	Chr10: 26504969 26504970	*GAD2*
897	Chr10: 26506353 26506354	*GAD2*
898	Chr10: 26727261 26727262	*APBB1IP*
899	Chr10: 28034797 28034798	*MKX*

Figure 59J

Order	Coordinate of CpG (hg19)	Gene
900	Chr10: 43600497 43600498	*RET*
901	Chr10: 50817810 50817811	*SLC18A3,CHAT*
902	Chr10: 50818299 50818300	*SLC18A3,CHAT*
903	Chr10: 50818707 50818708	*SLC18A3,CHAT*
904	Chr10: 64578469 64578470	*EGR2*
905	Chr10: 72043661 72043662	*Non Coding Region*
906	Chr10: 74094697 74094698	*DNAJB12*
907	Chr10: 7450355 7450356	*SFMBT2*
908	Chr10: 7451822 7451823	*SFMBT2*
909	Chr10: 7452598 7452599	*SFMBT2*
910	Chr10: 7452950 7452951	*SFMBT2*
911	Chr10: 7453329 7453330	*SFMBT2*
912	Chr10: 7708670 7708671	*ITIH5*
913	Chr10: 7708869 7708870	*ITIH5*
914	Chr10: 8078038 8078039	*Non Coding Region*
915	Chr10: 82116571 82116572	*DYDC2,DYDC1*
916	Chr10: 83636249 83636250	*NRG3*
917	Chr10: 83636325 83636326	*NRG3*
918	Chr10: 88126853 88126854	*GRID1*
919	Chr10: 91295045 91295046	*SLC16A12*
920	Chr10: 91295299 91295300	*SLC16A12*
921	Chr10: 91295317 91295318	*SLC16A12*
922	Chr11: 101454683 101454684	*TRPC6*
923	Chr11: 104034619 104034620	*PDGFD*
924	Chr11: 104034754 104034755	*PDGFD*
925	Chr11: 111383515 111383516	*BTG4,MIR34C,MIR34B*
926	Chr11: 112833773 112833774	*NCAM1*
927	Chr11: 115630889 115630890	*Non Coding Region*
928	Chr11: 119293385 119293386	*THY1*
929	Chr11: 119293601 119293602	*THY1*
930	Chr11: 122848098 122848099	*Non Coding Region*
931	Chr11: 122855029 122855030	*Non Coding Region*
932	Chr11: 122855292 122855293	*Non Coding Region*
933	Chr11: 124790815 124790816	*HEPACAM*
934	Chr11: 125757328 125757329	*HYLS1*
935	Chr11: 126226007 126226008	*ST3GAL4*
936	Chr11: 126226033 126226034	*ST3GAL4*
937	Chr11: 128563027 128563028	*FLI1*
938	Chr11: 128563685 128563686	*FLI1*
939	Chr11: 128563903 128563904	*FLI1*
940	Chr11: 128564106 128564107	*FLI1*
941	Chr11: 128564180 128564181	*FLI1*
942	Chr11: 128564756 128564757	*FLI1*
943	Chr11: 128564874 128564875	*FLI1*
944	Chr11: 131780353 131780354	*NTM*

Order	Coordinate of CpG (hg19)	Gene
945	Chr11: 132934139 132934140	*OPCML*
946	Chr11: 133825519 133825520	*IGSF9B*
947	Chr11: 134146132 134146133	*GLB1L3*
948	Chr11: 134146253 134146254	*GLB1L3*
949	Chr11: 134146676 134146677	*GLB1L3*
950	Chr11: 134281742 134281743	*B3GAT1*
951	Chr11: 1410385 1410386	*BRSK2*
952	Chr11: 1770066 1770067	*HCCA2,LOC402778*
953	Chr11: 17741070 17741071	*MYOD1*
954	Chr11: 18813544 18813545	*PTPN5*
955	Chr11: 20180700 20180701	*DBX1*
956	Chr11: 20185077 20185078	*Non Coding Region*
957	Chr11: 20185267 20185268	*Non Coding Region*
958	Chr11: 20690628 20690629	*NELL1*
959	Chr11: 20690682 20690683	*NELL1*
960	Chr11: 20690720 20690721	*NELL1*
961	Chr11: 20690807 20690808	*NELL1*
962	Chr11: 20691126 20691127	*NELL1*
963	Chr11: 20691161 20691162	*NELL1*
964	Chr11: 20691341 20691342	*NELL1*
965	Chr11: 20691429 20691430	*NELL1*
966	Chr11: 2162211 2162212	*IGF2,IGF2AS,INS-IGF2*
967	Chr11: 30607068 30607069	*MPPED2*
968	Chr11: 32457124 32457125	*WT1,WIT1*
969	Chr11: 32457127 32457128	*WT1,WIT1*
970	Chr11: 32457158 32457159	*WT1,WIT1*
971	Chr11: 32457162 32457163	*WT1,WIT1*
972	Chr11: 33890371 33890372	*LMO2*
973	Chr11: 44330958 44330959	*ALX4*
974	Chr11: 46317036 46317037	*CREB3L1*
975	Chr11: 46317154 46317155	*CREB3L1*
976	Chr11: 46317577 46317578	*CREB3L1*
977	Chr11: 57194562 57194563	*SLC43A3*
978	Chr11: 63768040 63768041	*MACROD1*
979	Chr11: 69632335 69632336	*FGF3*
980	Chr11: 69634372 69634373	*FGF3*
981	Chr11: 8190572 8190573	*RIC3*
982	Chr11: 8289974 8289975	*Non Coding Region*
983	Chr11: 830190 830191	*EFCAB4A*
984	Chr11: 91958458 91958459	*Non Coding Region*
985	Chr11: 91958799 91958800	*Non Coding Region*
986	Chr11: 94134580 94134581	*GPR83*
987	Chr11: 94134739 94134740	*GPR83*
988	Chr11: 94134756 94134757	*GPR83*
989	Chr11: 94501718 94501719	*AMOTL1*

Figure 59K

Order	Coordinate of CpG (hg19)	Gene
990	Chr11: 94502388 94502389	*AMOTL1*
991	Chr12: 103352294 103352295	*ASCL1*
992	Chr12: 103352326 103352327	*ASCL1*
993	Chr12: 103889772 103889773	*C12orf42*
994	Chr12: 103889789 103889790	*C12orf42*
995	Chr12: 104852355 104852356	*CHST11*
996	Chr12: 104852439 104852440	*CHST11*
997	Chr12: 1059459 1059460	*RAD52*
998	Chr12: 111471489 111471490	*CUX2*
999	Chr12: 111472183 111472184	*CUX2*
1000	Chr12: 113013048 113013049	*Non Coding Region*
1001	Chr12: 113494422 113494423	*DTX1*
1002	Chr12: 113494429 113494430	*DTX1*
1003	Chr12: 113901210 113901211	*LHX5*
1004	Chr12: 113901529 113901530	*LHX5*
1005	Chr12: 114075881 114075882	*Non Coding Region*
1006	Chr12: 114833912 114833913	*TBX5*
1007	Chr12: 117798954 117798955	*NOS1*
1008	Chr12: 117799083 117799084	*NOS1*
1009	Chr12: 119419329 119419330	*SRRM4*
1010	Chr12: 119419331 119419332	*SRRM4*
1011	Chr12: 120032374 120032375	*TMEM233*
1012	Chr12: 128850630 128850631	*TMEM132C*
1013	Chr12: 129338484 129338485	*GLT1D1*
1014	Chr12: 130389010 130389011	*TMEM132D*
1015	Chr12: 130647004 130647005	*FZD10*
1016	Chr12: 132906010 132906011	*GALNT9*
1017	Chr12: 15475690 15475691	*PTPRO*
1018	Chr12: 22487459 22487460	*ST8SIA1*
1019	Chr12: 25056083 25056084	*BCAT1*
1020	Chr12: 25102072 25102073	*BCAT1*
1021	Chr12: 25102274 25102275	*BCAT1*
1022	Chr12: 29936174 29936175	*TMTC1*
1023	Chr12: 41086227 41086228	*CNTN1*
1024	Chr12: 41086365 41086366	*CNTN1*
1025	Chr12: 41086800 41086801	*CNTN1*
1026	Chr12: 45444623 45444624	*DBX2*
1027	Chr12: 45444895 45444896	*DBX2*
1028	Chr12: 45445115 45445116	*DBX2*
1029	Chr12: 5018715 5018716	*KCNA1*
1030	Chr12: 5018798 5018799	*KCNA1*
1031	Chr12: 5018805 5018806	*KCNA1*
1032	Chr12: 5019017 5019018	*KCNA1*
1033	Chr12: 5019482 5019483	*KCNA1*
1034	Chr12: 50355307 50355308	*AQP5*

Order	Coordinate of CpG (hg19)	Gene
1035	Chr12: 5153174 5153175	*KCNA5*
1036	Chr12: 52400907 52400908	*GRASP*
1037	Chr12: 54321653 54321654	*Non Coding Region*
1038	Chr12: 54393445 54393446	*HOXC9*
1039	Chr12: 54394116 54394117	*HOXC9*
1040	Chr12: 5540116 5540117	*NTF3*
1041	Chr12: 62586003 62586004	*FAM19A2*
1042	Chr12: 62586267 62586268	*FAM19A2*
1043	Chr12: 64062498 64062499	*DPY19L2*
1044	Chr12: 64062526 64062527	*DPY19L2*
1045	Chr12: 64062564 64062565	*DPY19L2*
1046	Chr12: 64062724 64062725	*DPY19L2*
1047	Chr12: 64784483 64784484	*C12orf56*
1048	Chr12: 65515276 65515277	*WIF1*
1049	Chr12: 65515290 65515291	*WIF1*
1050	Chr12: 66123127 66123128	*Non Coding Region*
1051	Chr12: 72667150 72667151	*LOC283392,TRHDE*
1052	Chr12: 72667386 72667387	*LOC283392,TRHDE*
1053	Chr12: 72667398 72667399	*LOC283392,TRHDE*
1054	Chr12: 72667493 72667494	*LOC283392,TRHDE*
1055	Chr12: 81471754 81471755	*ACSS3*
1056	Chr12: 81471757 81471758	*ACSS3*
1057	Chr12: 81471884 81471885	*ACSS3*
1058	Chr12: 81471938 81471939	*ACSS3*
1059	Chr12: 8171463 8171464	*Non Coding Region*
1060	Chr12: 85306021 85306022	*SLC6A15*
1061	Chr12: 85306797 85306798	*SLC6A15*
1062	Chr12: 8548978 8548979	*Non Coding Region*
1063	Chr13: 102068530 102068531	*NALCN*
1064	Chr13: 102069132 102069133	*NALCN*
1065	Chr13: 102069169 102069170	*NALCN*
1066	Chr13: 102569815 102569816	*FGF14*
1067	Chr13: 102569876 102569877	*FGF14*
1068	Chr13: 102569894 102569895	*FGF14*
1069	Chr13: 103046930 103046931	*FGF14*
1070	Chr13: 103047053 103047054	*FGF14*
1071	Chr13: 103047225 103047226	*FGF14*
1072	Chr13: 103052362 103052363	*FGF14*
1073	Chr13: 103052468 103052469	*FGF14*
1074	Chr13: 103052828 103052829	*FGF14*
1075	Chr13: 108519318 108519319	*FAM155A*
1076	Chr13: 108520945 108520946	*FAM155A*
1077	Chr13: 109149171 109149172	*Non Coding Region*
1078	Chr13: 110958977 110958978	*COL4A2,COL4A1*
1079	Chr13: 112758379 112758380	*Non Coding Region*

Figure 59L

Order	Coordinate of CpG (hg19)	Gene
1080	Chr13:_112758599_112758600	*Non Coding Region*
1081	Chr13:_23733862_23733863	*Non Coding Region*
1082	Chr13:_23734304_23734305	*Non Coding Region*
1083	Chr13:_23734763_23734764	*Non Coding Region*
1084	Chr13:_25621027_25621028	*Non Coding Region*
1085	Chr13:_26043165_26043166	*ATP8A2*
1086	Chr13:_26625396_26625397	*SHISA2*
1087	Chr13:_26625766_26625767	*SHISA2*
1088	Chr13:_28366759_28366760	*GSX1*
1089	Chr13:_28371000_28371001	*Non Coding Region*
1090	Chr13:_28497117_28497118	*PDX1*
1091	Chr13:_29106422_29106423	*Non Coding Region*
1092	Chr13:_36704955_36704956	*DCLK1*
1093	Chr13:_36705606_36705607	*DCLK1*
1094	Chr13:_36919344_36919345	*SPG20*
1095	Chr13:_36919409_36919410	*SPG20*
1096	Chr13:_36919674_36919675	*SPG20*
1097	Chr13:_36919819_36919820	*SPG20*
1098	Chr13:_36920332_36920333	*SPG20*
1099	Chr13:_37004617_37004618	*CCNA1*
1100	Chr13:_37005570_37005571	*CCNA1*
1101	Chr13:_37006107_37006108	*CCNA1*
1102	Chr13:_37006127_37006128	*CCNA1*
1103	Chr13:_37006133_37006134	*CCNA1*
1104	Chr13:_37006611_37006612	*CCNA1*
1105	Chr13:_37248078_37248079	*C13orf36*
1106	Chr13:_37248305_37248306	*C13orf36*
1107	Chr13:_43148684_43148685	*TNFSF11*
1108	Chr13:_43149234_43149235	*TNFSF11*
1109	Chr13:_46425508_46425509	*SIAH3*
1110	Chr13:_49794635_49794636	*MLNR*
1111	Chr13:_53420425_53420426	*PCDH8*
1112	Chr13:_53422808_53422809	*PCDH8*
1113	Chr13:_58205045_58205046	*PCDH17*
1114	Chr13:_78492922_78492923	*EDNRB*
1115	Chr13:_78493012_78493013	*EDNRB*
1116	Chr13:_78493066_78493067	*EDNRB*
1117	Chr13:_78493100_78493101	*EDNRB*
1118	Chr13:_78493126_78493127	*EDNRB*
1119	Chr13:_78493128_78493129	*EDNRB*
1120	Chr13:_78493205_78493206	*EDNRB*
1121	Chr13:_78493229_78493230	*EDNRB*
1122	Chr13:_78493282_78493283	*EDNRB*
1123	Chr13:_78493294_78493295	*EDNRB*
1124	Chr13:_78493305_78493306	*EDNRB*

Order	Coordinate of CpG (hg19)	Gene
1125	Chr13:_78493313_78493314	*EDNRB*
1126	Chr13:_78493349_78493350	*EDNRB*
1127	Chr13:_78493365_78493366	*EDNRB*
1128	Chr13:_78493561_78493562	*EDNRB*
1129	Chr13:_78493564_78493565	*EDNRB*
1130	Chr13:_78493657_78493658	*EDNRB*
1131	Chr13:_78493671_78493672	*EDNRB*
1132	Chr13:_88324193_88324194	*SLITRK5*
1133	Chr13:_88324597_88324598	*SLITRK5*
1134	Chr13:_88325384_88325385	*SLITRK5*
1135	Chr13:_93880842_93880843	*GPC6*
1136	Chr13:_95359718_95359719	*Non Coding Region*
1137	Chr13:_95364675_95364676	*SOX21*
1138	Chr13:_95620306_95620307	*Non Coding Region*
1139	Chr13:_96204854_96204855	*CLDN10*
1140	Chr13:_96204873_96204874	*CLDN10*
1141	Chr14:_100438221_100438222	*Non Coding Region*
1142	Chr14:_100438647_100438648	*Non Coding Region*
1143	Chr14:_100438737_100438738	*Non Coding Region*
1144	Chr14:_100438957_100438958	*Non Coding Region*
1145	Chr14:_24803903_24803904	*ADCY4*
1146	Chr14:_27066404_27066405	*NOVA1*
1147	Chr14:_54687314_54687315	*Non Coding Region*
1148	Chr14:_60975846_60975847	*SIX6*
1149	Chr14:_62583919_62583920	*FLJ43390*
1150	Chr14:_70041283_70041284	*Non Coding Region*
1151	Chr14:_74707911_74707912	*VSX2*
1152	Chr14:_76605056_76605057	*Non Coding Region*
1153	Chr14:_77736904_77736905	*NGB*
1154	Chr14:_77737146_77737147	*NGB*
1155	Chr14:_85996202_85996203	*FLRT2*
1156	Chr14:_85996751_85996752	*FLRT2*
1157	Chr14:_96342250_96342251	*Non Coding Region*
1158	Chr14:_96342747_96342748	*Non Coding Region*
1159	Chr14:_96343119_96343120	*Non Coding Region*
1160	Chr14:_97685060_97685061	*Non Coding Region*
1161	Chr14:_97685274_97685275	*Non Coding Region*
1162	Chr14:_97685386_97685387	*Non Coding Region*
1163	Chr14:_97685576_97685577	*Non Coding Region*
1164	Chr14:_97685823_97685824	*Non Coding Region*
1165	Chr14:_97685945_97685946	*Non Coding Region*
1166	Chr15:_26108263_26108264	*ATP10A*
1167	Chr15:_26108663_26108664	*ATP10A*
1168	Chr15:_26108674_26108675	*ATP10A*
1169	Chr15:_26108683_26108684	*ATP10A*

Figure 59M

Order	Coordinate of CpG (hg19)	Gene
1170	Chr15: 26108796 26108797	*ATP10A*
1171	Chr15: 26108817 26108818	*ATP10A*
1172	Chr15: 27216083 27216084	*GABRG3*
1173	Chr15: 27216088 27216089	*GABRG3*
1174	Chr15: 28342183 28342184	*OCA2*
1175	Chr15: 28352098 28352099	*Non Coding Region*
1176	Chr15: 28352347 28352348	*Non Coding Region*
1177	Chr15: 28352485 28352486	*Non Coding Region*
1178	Chr15: 28352558 28352559	*Non Coding Region*
1179	Chr15: 30484033 30484034	*Non Coding Region*
1180	Chr15: 33603312 33603313	*RYR3*
1181	Chr15: 34786594 34786595	*Non Coding Region*
1182	Chr15: 35047189 35047190	*GJD2*
1183	Chr15: 45670478 45670479	*GATM,LOC145663*
1184	Chr15: 45670865 45670866	*GATM,LOC145663*
1185	Chr15: 45670872 45670873	*GATM,LOC145663*
1186	Chr15: 48937213 48937214	*FBN1*
1187	Chr15: 48937856 48937857	*FBN1*
1188	Chr15: 48938347 48938348	*FBN1*
1189	Chr15: 48938370 48938371	*FBN1*
1190	Chr15: 51385916 51385917	*TNFAIP8L3*
1191	Chr15: 55881044 55881045	*PYGO1*
1192	Chr15: 74421535 74421536	*ISLR2,LOC283731*
1193	Chr15: 79381867 79381868	*RASGRF1*
1194	Chr15: 79382998 79382999	*RASGRF1*
1195	Chr15: 79383167 79383168	*RASGRF1*
1196	Chr15: 79575048 79575049	*ANKRD34C*
1197	Chr15: 79576216 79576217	*ANKRD34C*
1198	Chr15: 83316640 83316641	*CPEB1*
1199	Chr15: 83316734 83316735	*CPEB1*
1200	Chr15: 83349420 83349421	*AP3B2*
1201	Chr15: 83776269 83776270	*TM6SF1*
1202	Chr15: 83776271 83776272	*TM6SF1*
1203	Chr15: 83776420 83776421	*TM6SF1*
1204	Chr15: 83776446 83776447	*TM6SF1*
1205	Chr15: 83952345 83952346	*BNC1*
1206	Chr15: 83953068 83953069	*BNC1*
1207	Chr15: 84115811 84115812	*SH3GL3*
1208	Chr15:_84115895_84115896	*SH3GL3*
1209	Chr15:_84115969_84115970	*SH3GL3*
1210	Chr15:_84116151_84116152	*SH3GL3*
1211	Chr15:_88798582_88798583	*NTRK3*
1212	Chr15:_88798666_88798667	*NTRK3*
1213	Chr15: 88798877 88798878	*NTRK3*
1214	Chr15: 88799707 88799708	*NTRK3*

Order	Coordinate of CpG (hg19)	Gene
1215	Chr15: 88800567 88800568	*NTRK3*
1216	Chr15: 88801004 88801005	*NTRK3*
1217	Chr15: 92937735 92937736	*ST8SIA2*
1218	Chr15: 93632730 93632731	*RGMA*
1219	Chr15: 98836248 98836249	*Non Coding Region*
1220	Chr16: 10133501 10133502	*GRIN2A*
1221	Chr16: 10276580 10276581	*GRIN2A*
1222	Chr16: 10276799 10276800	*GRIN2A*
1223	Chr16: 10276984 10276985	*GRIN2A*
1224	Chr16: 1203997 1203998	*CACNA1H*
1225	Chr16: 12996046 12996047	*SHISA9*
1226	Chr16: 12996234 12996235	*SHISA9*
1227	Chr16: 23847675 23847676	*PRKCB*
1228	Chr16: 25703528 25703529	*HS3ST4*
1229	Chr16: 25704505 25704506	*HS3ST4*
1230	Chr16: 25704507 25704508	*HS3ST4*
1231	Chr16: 28074384 28074385	*GSG1L*
1232	Chr16: 28074388 28074389	*GSG1L*
1233	Chr16: 28074462 28074463	*GSG1L*
1234	Chr16: 28075269 28075270	*GSG1L*
1235	Chr16: 31021933 31021934	*STX1B*
1236	Chr16: 31228059 31228060	*TRIM72,PYDC1*
1237	Chr16: 31228202 31228203	*TRIM72,PYDC1*
1238	Chr16: 3233068 3233069	*Non Coding Region*
1239	Chr16: 3237964 3237965	*Non Coding Region*
1240	Chr16: 51184562 51184563	*SALL1*
1241	Chr16: 51184786 51184787	*SALL1*
1242	Chr16: 51185001 51185002	*SALL1*
1243	Chr16: 51185082 51185083	*SALL1*
1244	Chr16: 51186211 51186212	*SALL1*
1245	Chr16: 51186260 51186261	*SALL1*
1246	Chr16: 51190201 51190202	*Non Coding Region*
1247	Chr16: 55358458 55358459	*IRX6*
1248	Chr16: 55512870 55512871	*MMP2*
1249	Chr16: 55513480 55513481	*MMP2*
1250	Chr16: 55689901 55689902	*SLC6A2*
1251	Chr16: 55690130 55690131	*SLC6A2*
1252	Chr16: 56224793 56224794	*GNAO1,LOC283856*
1253	Chr16:_56228442_56228443	*GNAO1,DKFZP434H168*
1254	Chr16:_56228467_56228468	*GNAO1,DKFZP434H168*
1255	Chr16:_56228495_56228496	*GNAO1,DKFZP434H168*
1256	Chr16:_56228498_56228499	*GNAO1,DKFZP434H168*
1257	Chr16:_56228505_56228506	*GNAO1,DKFZP434H168*
1258	Chr16: 58019123 58019124	*TEPP*
1259	Chr16: 66638210 66638211	*CMTM3*

Figure 59N

Order	Coordinate of CpG (hg19)	Gene
1260	Chr16: 66638293 66638294	*CMTM3*
1261	Chr16: 66638320 66638321	*CMTM3*
1262	Chr16: 66638433 66638434	*CMTM3*
1263	Chr16: 66638438 66638439	*CMTM3*
1264	Chr16: 67197129 67197130	*FBXL8,HSF4*
1265	Chr16: 71264579 71264580	*HYDIN*
1266	Chr16: 77468893 77468894	*ADAMTS18*
1267	Chr16: 77822419 77822420	*VAT1L*
1268	Chr16: 86529961 86529962	*Non Coding Region*
1269	Chr16: 86599745 86599746	*FOXC2*
1270	Chr16: 86601133 86601134	*FOXC2*
1271	Chr16: 86602083 86602084	*FOXC2*
1272	Chr17: 10101581 10101582	*GAS7*
1273	Chr17: 10102558 10102559	*GAS7*
1274	Chr17: 13505558 13505559	*HS3ST3A1*
1275	Chr17: 14212480 14212481	*HS3ST3B1*
1276	Chr17: 26699187 26699188	*SARM1*
1277	Chr17: 27940404 27940405	*ANKRD13B*
1278	Chr17: 32905966 32905967	*C17orf102*
1279	Chr17: 32907705 32907706	*TMEM132E,C17orf102*
1280	Chr17: 32907919 32907920	*TMEM132E*
1281	Chr17: 32908285 32908286	*TMEM132E*
1282	Chr17: 33701350 33701351	*SLFN11*
1283	Chr17: 33701353 33701354	*SLFN11*
1284	Chr17: 33814163 33814164	*SLFN12L*
1285	Chr17: 35165439 35165440	*Non Coding Region*
1286	Chr17: 35297146 35297147	*LHX1*
1287	Chr17: 38347659 38347660	*RAPGEFL1*
1288	Chr17: 38347710 38347711	*RAPGEFL1*
1289	Chr17:_43339358_43339359	*C17orf46,LOC100133991*
1290	Chr17:_43339589_43339590	*C17orf46,LOC100133991*
1291	Chr17:_43339594_43339595	*C17orf46,LOC100133991*
1292	Chr17: 45929657 45929658	*SP6*
1293	Chr17: 46811527 46811528	*Non Coding Region*
1294	Chr17: 48071845 48071846	*DLX3*
1295	Chr17: 54675137 54675138	*Non Coding Region*
1296	Chr17: 54675154 54675155	*Non Coding Region*
1297	Chr17: 56234476 56234477	*MSX2P1*
1298	Chr17: 64826184 64826185	*Non Coding Region*
1299	Chr17: 6616867 6616868	*SLC13A5*
1300	Chr17: 6616883 6616884	*SLC13A5*
1301	Chr17: 72348160 72348161	*KIF19*
1302	Chr17: 75369939 75369940	*SEPTIN9*
1303	Chr17: 7554681 7554682	*ATP1B2*
1304	Chr17: 76227996 76227997	*LOC283999*

Order	Coordinate of CpG (hg19)	Gene
1305	Chr17: 76228121 76228122	*LOC283999*
1306	Chr17: 77178924 77178925	*HRNBP3*
1307	Chr17: 77179455 77179456	*HRNBP3*
1308	Chr17: 77721631 77721632	*Non Coding Region*
1309	Chr17: 77721989 77721990	*Non Coding Region*
1310	Chr17: 78451563 78451564	*NPTX1*
1311	Chr17: 78452048 78452049	*Non Coding Region*
1312	Chr17: 8533391 8533392	*MYH10*
1313	Chr17: 8869213 8869214	*PIK3R5*
1314	Chr17: 8906601 8906602	*Non Coding Region*
1315	Chr17: 8907213 8907214	*Non Coding Region*
1316	Chr17: 8907561 8907562	*Non Coding Region*
1317	Chr18: 11752592 11752593	*GNAL*
1318	Chr18: 12254175 12254176	*CIDEA*
1319	Chr18: 22930271 22930272	*ZNF521*
1320	Chr18: 22930283 22930284	*ZNF521*
1321	Chr18: 25755942 25755943	*CDH2*
1322	Chr18: 31020511 31020512	*C18orf34*
1323	Chr18: 35104868 35104869	*BRUNOL4*
1324	Chr18: 35147532 35147533	*Non Coding Region*
1325	Chr18: 53447211 53447212	*Non Coding Region*
1326	Chr18: 55108852 55108853	*ONECUT2*
1327	Chr18: 5543548 5543549	*EPB41L3*
1328	Chr18: 56887271 56887272	*GRP*
1329	Chr18: 56887279 56887280	*GRP*
1330	Chr18: 56935199 56935200	*RAX*
1331	Chr18: 56940693 56940694	*RAX*
1332	Chr18: 59000800 59000801	*Non Coding Region*
1333	Chr18: 67067867 67067868	*DOK6*
1334	Chr18:_67067893_67067894	*DOK6*
1335	Chr18:_7116977_7116978	*LAMA1*
1336	Chr18:_7117321_7117322	*LAMA1*
1337	Chr18: 7118077 7118078	*LAMA1*
1338	Chr18: 76740258 76740259	*SALL3*
1339	Chr18: 904885 904886	*ADCYAP1*
1340	Chr19: 12203349 12203350	*ZNF788*
1341	Chr19: 12267308 12267309	*ZNF625*
1342	Chr19: 12267676 12267677	*ZNF625*
1343	Chr19: 13617094 13617095	*CACNA1A*
1344	Chr19: 1467952 1467953	*APC2*
1345	Chr19: 15090242 15090243	*Non Coding Region*
1346	Chr19: 15090872 15090873	*Non Coding Region*
1347	Chr19: 18118666 18118667	*ARRDC2*
1348	Chr19: 19322778 19322779	*NCAN*
1349	Chr19: 19372387 19372388	*HAPLN4*

Figure 59O

Order	Coordinate of CpG (hg19)	Gene
1350	Chr19: 21106053 21106054	*ZNF85*
1351	Chr19: 21646447 21646448	*Non Coding Region*
1352	Chr19: 21658001 21658002	*Non Coding Region*
1353	Chr19: 22034523 22034524	*Non Coding Region*
1354	Chr19: 22605186 22605187	*ZNF98*
1355	Chr19: 30016147 30016148	*VSTM2B*
1356	Chr19: 30017829 30017830	*VSTM2B*
1357	Chr19: 30866192 30866193	*ZNF536*
1358	Chr19: 34113398 34113399	*CHST8*
1359	Chr19: 34973644 34973645	*WTIP*
1360	Chr19: 36822442 36822443	*Non Coding Region*
1361	Chr19: 36909413 36909414	*ZFP82*
1362	Chr19: 36909831 36909832	*ZFP82*
1363	Chr19: 37096010 37096011	*ZNF529,ZNF382*
1364	Chr19: 37096487 37096488	*ZNF529,ZNF382*
1365	Chr19: 37407152 37407153	*ZNF829,ZNF568*
1366	Chr19: 37407284 37407285	*ZNF829,ZNF568*
1367	Chr19: 37407374 37407375	*ZNF829,ZNF568*
1368	Chr19: 37464066 37464067	*Non Coding Region*
1369	Chr19: 37997471 37997472	*ZNF793*
1370	Chr19: 37997867 37997868	*ZNF793*
1371	Chr19: 37997991 37997992	*ZNF793*
1372	Chr19: 41018880 41018881	*SPTBN4*
1373	Chr19: 41595975 41595976	*CYP2A13*
1374	Chr19: 44952572 44952573	*ZNF229*
1375	Chr19: 44952618 44952619	*ZNF229*
1376	Chr19: 44952665 44952666	*ZNF229*
1377	Chr19: 50037018 50037019	*RCN3*
1378	Chr19: 51111388 51111389	*Non Coding Region*
1379	Chr19: 51830477 51830478	*IGLON5*
1380	Chr19: 51830960 51830961	*IGLON5*
1381	Chr19: 51924572 51924573	*Non Coding Region*
1382	Chr19: 51925128 51925129	*Non Coding Region*
1383	Chr19: 52873106 52873107	*ZNF880*
1384	Chr19: 52873125 52873126	*ZNF880*
1385	Chr19: 52900974 52900975	*ZNF528*
1386	Chr19: 52901124 52901125	*ZNF528*
1387	Chr19: 53636108 53636109	*ZNF415*
1388	Chr19: 53811940 53811941	*Non Coding Region*
1389	Chr19: 54024110 54024111	*ZNF331*
1390	Chr19: 54411773 54411774	*Non Coding Region*
1391	Chr19: 56879559 56879560	*ZNF542*
1392	Chr19: 56879645 56879646	*ZNF542*
1393	Chr19: 56879933 56879934	*ZNF542*
1394	Chr19: 56905032 56905033	*ZNF582*

Order	Coordinate of CpG (hg19)	Gene
1395	Chr19: 56989543 56989544	*ZNF667*
1396	Chr19: 57683712 57683713	*Non Coding Region*
1397	Chr19: 57683885 57683886	*Non Coding Region*
1398	Chr19: 57862627 57862628	*ZNF304*
1399	Chr19: 57862638 57862639	*ZNF304*
1400	Chr19: 58238928 58238929	*ZNF671*
1401	Chr19: 58238987 58238988	*ZNF671*
1402	Chr19: 58239012 58239013	*ZNF671*
1403	Chr19: 58609361 58609362	*ZSCAN18*
1404	Chr19: 58951602 58951603	*ZNF132*
1405	Chr19: 7795619 7795620	*CLEC4G*
1406	Chr19: 7853544 7853545	*CLEC4GP1*
1407	Chr20: 10199302 10199303	*SNAP25*
1408	Chr20: 10199536 10199537	*SNAP25*
1409	Chr20: 21695334 21695335	*PAX1*
1410	Chr20: 23029640 23029641	*THBD*
1411	Chr20: 23029806 23029807	*THBD*
1412	Chr20: 23030250 23030251	*THBD*
1413	Chr20: 25058429 25058430	*VSX1*
1414	Chr20: 2781122 2781123	*CPXM1*
1415	Chr20: 2781241 2781242	*CPXM1*
1416	Chr20: 30063249 30063250	*REM1*
1417	Chr20: 30582838 30582839	*XKR7*
1418	Chr20: 30640256 30640257	*HCK*
1419	Chr20: 34188940 34188941	*FER1L4*
1420	Chr20: 34894648 34894649	*Non Coding Region*
1421	Chr20: 37302636 37302637	*Non Coding Region*
1422	Chr20: 37353032 37353033	*SLC32A1*
1423	Chr20: 37353117 37353118	*SLC32A1*
1424	Chr20: 37355965 37355966	*SLC32A1*
1425	Chr20: 37434231 37434232	*PPP1R16B*
1426	Chr20: 37434552 37434553	*PPP1R16B*
1427	Chr20: 37435119 37435120	*PPP1R16B*
1428	Chr20: 39317034 39317035	*MAFB*
1429	Chr20: 41817903 41817904	*PTPRT*
1430	Chr20: 41818358 41818359	*PTPRT*
1431	Chr20: 41818756 41818757	*PTPRT*
1432	Chr20: 41818770 41818771	*PTPRT*
1433	Chr20: 41818788 41818789	*PTPRT*
1434	Chr20: 41818905 41818906	*PTPRT*
1435	Chr20: 42875931 42875932	*GDAP1L1*
1436	Chr20: 44941542 44941543	*Non Coding Region*
1437	Chr20: 45142258 45142259	*ZNF334*
1438	Chr20: 47934987 47934988	*Non Coding Region*
1439	Chr20: 47935243 47935244	*Non Coding Region*

Figure 59P

Order	Coordinate of CpG (hg19)	Gene
1440	Chr20:_47935683_47935684	*Non Coding Region*
1441	Chr20:_49639134_49639135	*KCNG1*
1442	Chr20:_53092233_53092234	*DOK5*
1443	Chr20:_53092334_53092335	*DOK5*
1444	Chr20:_53092781_53092782	*DOK5*
1445	Chr20:_55500670_55500671	*Non Coding Region*
1446	Chr20:_57090013_57090014	*APCDD1L*
1447	Chr20:_58180241_58180242	*PHACTR3*
1448	Chr20:_61050914_61050915	*GATA5*
1449	Chr20:_61051021_61051022	*GATA5*
1450	Chr20:_61051029_61051030	*GATA5*
1451	Chr20:_61051032_61051033	*GATA5*
1452	Chr20:_61051036_61051037	*GATA5*
1453	Chr20:_61051039_61051040	*GATA5*
1454	Chr20:_61051317_61051318	*GATA5*
1455	Chr20:_61051341_61051342	*GATA5*
1456	Chr20:_61051348_61051349	*GATA5*
1457	Chr20:_61051423_61051424	*GATA5*
1458	Chr20:_61051432_61051433	*GATA5*
1459	Chr20:_61051438_61051439	*GATA5*
1460	Chr20:_61051561_61051562	*GATA5*
1461	Chr20:_61638149_61638150	*BHLHE23*
1462	Chr20:_61638420_61638421	*BHLHE23*
1463	Chr20:_61638433_61638434	*BHLHE23*
1464	Chr20:_61638518_61638519	*BHLHE23*
1465	Chr20:_61638588_61638589	*BHLHE23*
1466	Chr20:_61806651_61806652	*Non Coding Region*
1467	Chr20:_61808888_61808889	*MIR124-3*
1468	Chr20:_61809557_61809558	*MIR124-3*
1469	Chr20:_61809827_61809828	*MIR124-3*
1470	Chr20:_61809932_61809933	*MIR124-3*
1471	Chr20:_61810624_61810625	*Non Coding Region*
1472	Chr20:_61885291_61885292	*NKAIN4,FLJ16779*
1473	Chr20:_61886082_61886083	*NKAIN4,FLJ16779*
1474	Chr20:_62103400_62103401	*KCNQ2*
1475	Chr20:_8112972_8112973	*PLCB1*
1476	Chr20:_8113573_8113574	*PLCB1*
1477	Chr21:_19617873_19617874	*CHODL*
1478	Chr21:_27011963_27011964	*JAM2*
1479	Chr21:_27012176_27012177	*JAM2*
1480	Chr21:_27012615_27012616	*JAM2*
1481	Chr21:_28339442_28339443	*ADAMTS5*
1482	Chr21:_28339907_28339908	*ADAMTS5*
1483	Chr21:_34399260_34399261	*OLIG2*
1484	Chr21:_47518747_47518748	*COL6A2*

Order	Coordinate of CpG (hg19)	Gene
1485	Chr22:_19138209_19138210	*GSC2*
1486	Chr22:_19511987_19511988	*CLDN5*
1487	Chr22:_25678615_25678616	*Non Coding Region*
1488	Chr22:_39853885_39853886	*MGAT3*
1489	Chr22:_44258572_44258573	*SULT4A1*
1490	Chr22:_45405061_45405062	*PHF21B*
1491	ChrX:_6144633_6144634	*NLGN4X*
1492	ChrX:_6145346_6145347	*NLGN4X*
1493	ChrX:_6145730_6145731	*NLGN4X*
1494	ChrX:_8699141_8699142	*KAL1*
	Alternative Markers: Lower in CRC for Group 1 set	
1495	Chr01:_107683775_107683776	*NTNG1*
1496	Chr01:_148854315_148854316	*Non Coding Region*
1497	Chr01:_152488093_152488094	*CRCT1*
1498	Chr01:_156405924_156405925	*Non Coding Region*
1499	Chr01:_181452044_181452045	*CACNA1E*
1500	Chr01:_18957020_18957021	*PAX7*
1501	Chr01:_224804222_224804223	*CNIH3*
1502	Chr01:_29138936_29138937	*OPRD1*
1503	Chr01:_34630944_34630945	*CSMD2*
1504	Chr01:_44883697_44883698	*RNF220*
1505	Chr01:_50799316_50799317	*Non Coding Region*
1506	Chr01:_50884544_50884545	*DMRTA2*
1507	Chr01:_53068181_53068182	*GPX7*
1508	Chr01:_65731738_65731739	*DNAJC6*
1509	Chr01:_91191001_91191002	*Non Coding Region*
1510	Chr02:_127644075_127644076	*Non Coding Region*
1511	Chr02:_149633226_149633227	*KIF5C*
1512	Chr02:_176957462_176957463	*HOXD13*
1513	Chr02:_176995556_176995557	*HOXD8*
1514	Chr02:_45241110_45241111	*Non Coding Region*
1515	Chr02:_47797963_47797964	*KCNK12*
1516	Chr02:_88751786_88751787	*FOXI3*
1517	Chr02:_88752056_88752057	*FOXI3*
1518	Chr03:_140770345_140770346	*SPSB4*
1519	Chr03:_142682652_142682653	*PAQR9*
1520	Chr03:_152553673_152553674	*P2RY1*
1521	Chr03:_16554910_16554911	*RFTN1*
1522	Chr03:_42947690_42947691	*ZNF662*
1523	Chr03:_96532043_96532044	*EPHA6*
1524	Chr03:_96533290_96533291	*EPHA6*
1525	Chr04:_123748386_123748387	*FGF2*
1526	Chr04:_154709878_154709879	*SFRP2*
1527	Chr04:_37246827_37246828	*KIAA1239*
1528	Chr04:_81952024_81952025	*BMP3*

Figure 59Q

Order	Coordinate of CpG (hg19)	Gene
1529	Chr05: 101632341 101632342	*SLCO4C1*
1530	Chr05: 112073348 112073349	*APC*
1531	Chr05: 112073350 112073351	*APC*
1532	Chr05: 112073373 112073374	*APC*
1533	Chr05: 112073398 112073399	*APC*
1534	Chr05: 112073406 112073407	*APC*
1535	Chr05: 112073426 112073427	*APC*
1536	Chr05: 112073438 112073439	*APC*
1537	Chr05: 112073538 112073539	*APC*
1538	Chr05: 112073613 112073614	*APC*
1539	Chr05: 11904110 11904111	*CTNND2*
1540	Chr05: 15500833 15500834	*FBXL7*
1541	Chr05: 170738274 170738275	*TLX3*
1542	Chr05: 42423947 42423948	*GHR*
1543	Chr05: 42424525 42424526	*GHR*
1544	Chr06: 100051116 100051117	*Non Coding Region*
1545	Chr06: 100061307 100061308	*PRDM13*
1546	Chr06: 105584709 105584710	*BVES*
1547	Chr06: 117591684 117591685	*VGLL2*
1548	Chr06: 124124275 124124276	*NKAIN2*
1549	Chr06: 124124875 124124876	*NKAIN2*
1550	Chr06: 125284359 125284360	*STL*
1551	Chr06: 1378868 1378869	*Non Coding Region*
1552	Chr06: 153451251 153451252	*RGS17*
1553	Chr06: 153452724 153452725	*RGS17*
1554	Chr06: 88876741 88876742	*CNR1*
1555	Chr06: 94129234 94129235	*EPHA7*
1556	Chr07: 140773905 140773906	*Non Coding Region*
1557	Chr07: 20817530 20817531	*Non Coding Region*
1558	Chr07: 20817859 20817860	*Non Coding Region*
1559	Chr07: 27196302 27196303	*HOXA7*
1560	Chr07: 37488428 37488429	*ELMO1*
1561	Chr07: 4922708 4922709	*RADIL*
1562	Chr07: 49814435 49814436	*VWC2*
1563	Chr07: 87230344 87230345	*ABCB1*
1564	Chr07: 87257074 87257075	*ABCB1,RUNDC3B*
1565	Chr08: 65492792 65492793	*BHLHE22*
1566	Chr08: 72460135 72460136	*Non Coding Region*
1567	Chr08: 99439685 99439686	*KCNS2*
1568	Chr09: 124889019 124889020	*Non Coding Region*
1569	Chr09: 96715256 96715257	*BARX1*
1570	Chr10: 101292766 101292767	*NKX2-3*
1571	Chr10: 102497666 102497667	*Non Coding Region*
1572	Chr10: 118893174 118893175	*VAX1*
1573	Chr10: 118897847 118897848	*VAX1*

Order	Coordinate of CpG (hg19)	Gene
1574	Chr10: 118922841 118922842	*Non Coding Region*
1575	Chr10: 118922887 118922888	*Non Coding Region*
1576	Chr10: 119302053 119302054	*EMX2OS,EMX2*
1577	Chr10: 124896232 124896233	*HMX3*
1578	Chr10: 126136607 126136608	*NKX1-2*
1579	Chr10: 17496239 17496240	*ST8SIA6*
1580	Chr10: 18429749 18429750	*CACNB2*
1581	Chr10: 18429760 18429761	*CACNB2*
1582	Chr10: 23481385 23481386	*PTF1A*
1583	Chr10: 48438724 48438725	*GDF10*
1584	Chr10: 48439229 48439230	*GDF10*
1585	Chr10: 90342738 90342739	*RNLS*
1586	Chr10: 90342899 90342900	*RNLS*
1587	Chr10: 90343166 90343167	*RNLS*
1588	Chr10: 90343174 90343175	*RNLS*
1589	Chr10: 90343192 90343193	*RNLS*
1590	Chr10: 90343204 90343205	*RNLS*
1591	Chr10: 90343208 90343209	*RNLS*
1592	Chr10: 94834201 94834202	*CYP26A1*
1593	Chr11: 106889364 106889365	*GUCY1A2*
1594	Chr11: 58672888 58672889	*Non Coding Region*
1595	Chr11: 58673167 58673168	*Non Coding Region*
1596	Chr12: 133484853 133484854	*Non Coding Region*
1597	Chr12: 39299364 39299365	*CPNE8*
1598	Chr12: 47224954 47224955	*Non Coding Region*
1599	Chr12: 54338937 54338938	*HOXC13*
1600	Chr12: 8542839 8542840	*LOC389634*
1601	Chr13: 112727521 112727522	*Non Coding Region*
1602	Chr13: 28674451 28674452	*FLT3*
1603	Chr13: 79177516 79177517	*POU4F1*
1604	Chr13: 79177700 79177701	*POU4F1*
1605	Chr13: 88323607 88323608	*SLITRK5*
1606	Chr14: 36003741 36003742	*INSM2*
1607	Chr14: 36995009 36995010	*Non Coding Region*
1608	Chr14: 51561293 51561294	*TRIM9*
1609	Chr14: 52535973 52535974	*NID2*
1610	Chr14: 52536066 52536067	*NID2*
1611	Chr15: 33010399 33010400	*GREM1*
1612	Chr15: 60296996 60296997	*FOXB1*
1613	Chr15: 78913147 78913148	*CHRNA3*
1614	Chr15: 83875703 83875704	*HDGFRP3*
1615	Chr15: 93631754 93631755	*RGMA*
1616	Chr16: 28074980 28074981	*GSG1L*
1617	Chr16: 56224901 56224902	*GNAO1,LOC283856*
1618	Chr16: 86544339 86544340	*FOXF1*

Figure 59R

Order	Coordinate of CpG (hg19)	*Gene*
1619	Chr17:_41832753_41832754	*SOST*
1620	Chr17:_41832873_41832874	*SOST*
1621	Chr17:_77179709_77179710	*HRNBP3*
1622	Chr17:_79448527_79448528	*Non Coding Region*
1623	Chr18:_32847566_32847567	*ZNF397OS*
1624	Chr18:_5197327_5197328	*LOC642597*
1625	Chr18:_5543271_5543272	*EPB41L3*
1626	Chr18:_5544169_5544170	*EPB41L3*
1627	Chr18:_74961727_74961728	*GALR1*
1628	Chr18:_74961737_74961738	*GALR1*
1629	Chr19:_30019529_30019530	*VSTM2B*
1630	Chr19:_30019647_30019648	*VSTM2B*
1631	Chr19:_31839734_31839735	*TSHZ3*
1632	Chr19:_31841555_31841556	*TSHZ3*
1633	Chr19:_31841663_31841664	*TSHZ3*
1634	Chr19:_31841952_31841953	*Non Coding Region*
1635	Chr19:_38183055_38183056	*ZNF781*
1636	Chr19:_53696649_53696650	*ZNF665*
1637	Chr19:_56904901_56904902	*ZNF582*
1638	Chr19:_57078765_57078766	*ZNF470*
1639	Chr20:_13200980_13200981	*ISM1*
1640	Chr20:_43439387_43439388	*RIMS4*
1641	Chr20:_43439400_43439401	*RIMS4*
1642	Chr20:_56803964_56803965	*Non Coding Region*
1643	Chr21:_34442360_34442361	*OLIG1*
1644	Chr21:_34442377_34442378	*OLIG1*
1645	Chr21:_38119946_38119947	*SIM2*
1646	Chr22:_45405627_45405628	*PHF21B*

Figure 59S

Grp 1 Methylation markers, 50% sensitivity, Chromosomal Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 1 Markers	
1	chr01:107682889-107684463	*NTNG1*
2	chr01:121260857-121261525	*LOC647121*
3	chr01:18956895-18959829	*PAX7*
4	chr01:240254959-240257063	*FMN2*
5	chr01:29138501-29139348	*OPRD1*
6	chr01:34628783-34630976	*CSMD2*
7	chr01:44883136-44884272	*RNF220*
8	chr01:50884228-50891471	*DMRTA2*
9	chr01:53067880-53068608	*GPX7*
10	chr01:77333111-77334534	*ST6GALNAC5*
11	chr02:100937779-100939059	*LONRF2*
12	chr02:131792237-131793189	*ARHGEF4*
13	chr02:175193398-175193764	*Non Coding Region*
14	chr02:176957054-176958279	*HOXD13*
15	chr02:176977284-176977540	*Non Coding Region*
16	chr02:176993479-176995557	*HOXD8*
17	chr02:182547873-182549177	*Non Coding Region*
18	chr02:29337983-29338909	*CLIP4*
19	chr02:66802890-66804831	*Non Coding Region*
20	chr02:88751335-88752865	*FOXI3*
21	chr03:140769886-140771080	*SPSB4*
22	chr03:152552602-152553712	*P2RY1*
23	chr03:69591011-69592050	*Non Coding Region*
24	chr03:96532023-96533625	*EPHA6*
25	chr04:37245250-37246979	*KIAA1239*
26	chr05:11903550-11904703	*CTNND2*
27	chr05:170735169-170739863	*TLX3*
28	chr05:38556222-38557563	*LIFR*
29	chr05:42424338-42425047	*GHR*
30	chr05:5139441-5140941	*ADAMTS16*
31	chr06:105584148-105585621	*BVES*
32	chr06:124124177-124125309	*NKAIN2*
33	chr06:1381743-1385211	*Non Coding Region*
34	chr06:166579973-166583423	*T*
35	chr07:149410858-149412351	*KRBA1*
36	chr07:20817455-20818227	*Non Coding Region*
37	chr07:37487354-37488672	*ELMO1*
38	chr07:37960316-37961046	*EPDR1*
39	chr07:54609734-54610140	*VSTM2A*
40	chr07:87256958-87258444	*ABCB1,RUNDC3B*
41	chr07:93519366-93520184	*TFPI2*
42	chr08:41753340-41755250	*ANK1*
43	chr08:65492935-65494452	*BHLHE22*
44	chr09:117266468-117268094	*DFNB31*
45	chr09:124888949-124889382	*Non Coding Region*
46	chr09:96713326-96718186	*BARX1*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 1 Markers	
47	chr10:118893527-118894432	*VAX1*
48	chr10:126135809-126138896	*NKX1-2*
49	chr10:17495407-17496721	*ST8SIA6*
50	chr10:18429125-18430234	*CACNB2*
51	chr10:90342528-90343221	*RNLS*
52	chr11:126225355-126226073	*ST3GAL4*
53	chr11:128562671-128565011	*FLI1*
54	chr11:94501366-94502696	*AMOTL1*
55	chr12:113900750-113906442	*LHX5*
56	chr12:54338761-54339168	*HOXC13*
57	chr12:8542555-8543334	*LOC389634*
58	chr13:78492425-78493382	*EDNRB*
59	chr13:95619883-95620977	*Non Coding Region*
60	chr14:51560116-51562487	*TRIM9*
61	chr14:52734207-52735486	*PTGDR*
62	chr14:57283967-57284558	*OTX2OS1*
63	chr15:33009530-33011696	*GREM1*
64	chr15:45669970-45671029	*GATM,LOC145663*
65	chr15:76632262-76635515	*ISL2*
66	chr15:78912246-78913616	*CHRNA3*
67	chr16:56224731-56224980	*GNAO1,LOC283856*
68	chr16:86599355-86602649	*FOXC2*
69	chr17:33700989-33701657	*SLFN11*
70	chr17:3438841-3439046	*TRPV3*
71	chr17:41832717-41833195	*SOST*
72	chr17:43339124-43339832	*C17orf46,LOC100133991*
73	chr18:32847284-32848130	*ZNF397OS*
74	chr18:5543437-5544241	*EPB41L3*
75	chr19:31839635-31843049	*TSHZ3*
76	chr19:36909281-36909854	*ZFP82*
77	chr19:37406931-37407463	*ZNF829,ZNF568*
78	chr19:37463991-37464606	*Non Coding Region*
79	chr19:38182793-38183327	*ZNF781*
80	chr19:51924999-51925413	*Non Coding Region*
81	chr19:53696029-53696650	*ZNF665*
82	chr19:56879417-56879995	*ZNF542*
83	chr19:56904636-56905355	*ZNF582*
84	chr19:56988313-56989741	*ZNF667*
85	chr19:57078365-57079301	*ZNF470*
86	chr19:57862441-57863236	*ZNF304*
87	chr20:13200670-13202616	*ISM1*
88	chr20:23028403-23032218	*THBD*
89	chr20:61636171-61639055	*BHLHE23*
90	chr21:38119793-38120742	*SIM2*
91	chr22:45403036-45406372	*PHF21B*

Figure 60A

Order	Coordinate of CpG (hg19)	Gene
	Alternative markers: High in CRC for Group 1 set	
92	chr01:101004471-101005885	*GPR88*
93	chr01:101702445-101702745	*S1PR1*
94	chr01:108507251-108507841	*VAV3*
95	chr01:111216244-111217937	*KCNA3*
96	chr01:114695136-114696672	*SYT6*
97	chr01:115880167-115881332	*NGF*
98	chr01:119529819-119530712	*TBX15*
99	chr01:12123488-12124148	*TNFRSF8*
100	chr01:13910137-13910868	*PDPN*
101	chr01:150254365-150254637	*C1orf51*
102	chr01:154298205-154298544	*ATP8B2*
103	chr01:156627342-156627576	*BCAN*
104	chr01:159141202-159141718	*CADM3*
105	chr01:16085147-16085862	*FBLIM1*
106	chr01:171810467-171811325	*DNM3*
107	chr01:179711947-179713951	*FAM163A*
108	chr01:181287300-181287873	*Non Coding Region*
109	chr01:196577708-196578011	*KCNT2*
110	chr01:207669637-207670015	*CR1*
111	chr01:208084098-208084513	*CD34*
112	chr01:224803637-224805314	*CNIH3*
113	chr01:228651447-228652529	*Non Coding Region*
114	chr01:229542943-229543770	*Non Coding Region*
115	chr01:231296559-231297345	*Non Coding Region*
116	chr01:234040298-234041457	*SLC35F3*
117	chr01:234349884-234350471	*SLC35F3*
118	chr01:237205126-237206644	*RYR2*
119	chr01:246952214-246952511	*LOC149134*
120	chr01:33219427-33220028	*KIAA1522*
121	chr01:35394747-35396206	*Non Coding Region*
122	chr01:3662963-3664085	*KIAA0495*
123	chr01:37498377-37500624	*GRIK3*
124	chr01:38099677-38100864	*RSPO1*
125	chr01:38229838-38230888	*EPHA10*
126	chr01:38510102-38513642	*POU3F1*
127	chr01:39269729-39270251	*Non Coding Region*
128	chr01:44871109-44874047	*RNF220*
129	chr01:46951168-46951792	*Non Coding Region*
130	chr01:48190756-48190992	*Non Coding Region*
131	chr01:49242371-49242810	*AGBL4,BEND5*
132	chr01:53098629-53099303	*FAM159A*
133	chr01:57887963-57890637	*DAB1*
134	chr01:58715272-58716095	*DAB1*
135	chr01:67773329-67773767	*IL12RB2*

Order	Coordinate of CpG (hg19)	Gene
136	chr01:70032967-70034495	*Non Coding Region*
137	chr01:75600674-75601415	*LHX8*
138	chr01:76080454-76080808	*Non Coding Region*
139	chr01:86621278-86622871	*COL24A1*
140	chr01:98510966-98511710	*MIR137*
141	chr01:99469932-99470968	*LPPR5*
142	chr02:10219802-10221016	*CYS1*
143	chr02:105459127-105461770	*Non Coding Region*
144	chr02:108602824-108603467	*SLC5A7*
145	chr02:11051858-11053476	*KCNF1*
146	chr02:115918737-115920765	*DPP10*
147	chr02:119914126-119916663	*C1QL2*
148	chr02:120281661-120282211	*SCTR*
149	chr02:131720819-131721867	*ARHGEF4*
150	chr02:137522460-137523696	*Non Coding Region*
151	chr02:144694666-144695180	*Non Coding Region*
152	chr02:145273456-145275378	*ZEB2*
153	chr02:145281736-145282269	*Non Coding Region*
154	chr02:154727906-154728271	*GALNT13*
155	chr02:154729613-154729918	*GALNT13*
156	chr02:164592917-164593511	*FIGN*
157	chr02:171672310-171675447	*GAD1*
158	chr02:1746833-1748971	*PXDN*
159	chr02:177052957-177054350	*HOXD1*
160	chr02:182321761-182323029	*ITGA4*
161	chr02:185463208-185463589	*ZNF804A*
162	chr02:187713846-187714069	*ZSWIM2*
163	chr02:193058995-193060812	*TMEFF2*
164	chr02:207506774-207507422	*LOC200726*
165	chr02:210636344-210636935	*UNC80*
166	chr02:217236025-217237129	*MARCHF4*
167	chr02:217559039-217559484	*IGFBP5*
168	chr02:218621279-218621498	*DIRC3*
169	chr02:220173870-220174283	*PTPRN*
170	chr02:220299483-220300243	*SPEG*
171	chr02:223167205-223167560	*CCDC140*
172	chr02:223168653-223169008	*CCDC140*
173	chr02:230578062-230579981	*DNER*
174	chr02:238535561-238536840	*LRRFIP1*
175	chr02:242742598-242743758	*GAL3ST2*
176	chr02:30142831-30144624	*ALK*
177	chr02:31360321-31361821	*GALNT14*
178	chr02:31456740-31457653	*EHD3*
179	chr02:31805293-31806403	*SRD5A2*
180	chr02:40678477-40678717	*SLC8A1*

Figure 60B

Order	Coordinate of CpG (hg19)	Gene
181	chr02:47796923-47799166	*KCNK12*
182	chr02:50574045-50574817	*NRXN1*
183	chr02:56150340-56151180	*EFEMP1*
184	chr02:5836068-5837643	*SOX11*
185	chr02:66652691-66654218	*Non Coding Region*
186	chr02:68546339-68547111	*CNRIP1*
187	chr02:7057530-7058433	*RNF144A*
188	chr02:71693164-71694127	*DYSF*
189	chr02:73429283-73430647	*NOTO*
190	chr02:87015974-87018182	*CD8A*
191	chr02:945686-946774	*SNTG2*
192	chr02:95690857-95692431	*MAL*
193	chr02:97192977-97193383	*Non Coding Region*
194	chr02:98962873-98964187	*CNGA3*
195	chr03:10857687-10858447	*SLC6A11*
196	chr03:120169202-120170519	*FSTL1*
197	chr03:128205495-128212274	*GATA2*
198	chr03:128719865-128721245	*CCDC48*
199	chr03:13114627-13115245	*IQSEC1*
200	chr03:132756985-132757895	*TMEM108*
201	chr03:13323438-13324929	*Non Coding Region*
202	chr03:134369464-134370242	*KY*
203	chr03:134514085-134515980	*EPHB1*
204	chr03:137479995-137480197	*Non Coding Region*
205	chr03:137487429-137488021	*Non Coding Region*
206	chr03:142837885-142840838	*CHST2*
207	chr03:147126988-147128999	*ZIC1*
208	chr03:14851829-14853094	*Non Coding Region*
209	chr03:159756632-159756997	*Non Coding Region*
210	chr03:179754520-179755245	*PEX5L*
211	chr03:186078710-186080111	*DGKG*
212	chr03:192125821-192127994	*FGF12*
213	chr03:194406440-194409045	*FAM43A*
214	chr03:238391-240140	*CHL1*
215	chr03:27771638-27771942	*Non Coding Region*
216	chr03:28616663-28617970	*Non Coding Region*
217	chr03:32858194-32860506	*TRIM71*
218	chr03:33259919-33260763	*SUSD5*
219	chr03:36421702-36422210	*STAC*
220	chr03:38035701-38036000	*VILL*
221	chr03:44036407-44041104	*Non Coding Region*
222	chr03:44626325-44626794	*ZNF660*
223	chr03:46506161-46506416	*LTF*
224	chr03:46923265-46925182	*PTH1R*
225	chr03:48693118-48694768	*CELSR3*

Order	Coordinate of CpG (hg19)	Gene
226	chr03:48698335-48701667	*CELSR3*
227	chr03:62357639-62359774	*FEZF2*
228	chr03:68980783-68982035	*FAM19A4*
229	chr03:71802560-71804209	*EIF4E3,GPR27*
230	chr03:79815638-79815900	*ROBO1*
231	chr04:107956555-107957453	*DKK2*
232	chr04:110222970-110224257	*COL25A1*
233	chr04:111560563-111560808	*Non Coding Region*
234	chr04:122301567-122302290	*ORFPR*
235	chr04:122685860-122686565	*TMEM155,LOC100192379*
236	chr04:126235895-126238930	*FAT4*
237	chr04:134069162-134070442	*Non Coding Region*
238	chr04:13537646-13537973	*Non Coding Region*
239	chr04:142053327-142054601	*RNF150*
240	chr04:144620822-144622218	*FREM3*
241	chr04:148402198-148402760	*EDNRA*
242	chr04:154680774-154681377	*RNF175*
243	chr04:154709512-154710827	*SFRP2*
244	chr04:154712073-154712706	*Non Coding Region*
245	chr04:156297601-156298094	*MAP9*
246	chr04:156680095-156681386	*GUCY1B3*
247	chr04:157997166-157997686	*GLRB*
248	chr04:166794545-166795341	*TLL1*
249	chr04:168155005-168155765	*SPOCK3*
250	chr04:175135259-175135496	*Non Coding Region*
251	chr04:175750330-175750584	*GLRA3*
252	chr04:176922473-176922820	*GPM6A*
253	chr04:176986921-176987360	*WDR17*
254	chr04:17782642-17783624	*FAM184B*
255	chr04:184826253-184828177	*STOX2*
256	chr04:185939222-185942747	*HELT*
257	chr04:40858884-40859162	*APBB2*
258	chr04:41883492-41884570	*Non Coding Region*
259	chr04:42152766-42154984	*BEND4*
260	chr04:4387593-4389919	*D4S234E*
261	chr04:5053230-5054092	*STK32B*
262	chr04:55096185-55100331	*PDGFRA*
263	chr04:55991402-55992171	*KDR*
264	chr04:568513-570488	*Non Coding Region*
265	chr04:5709985-5710495	*EVC2*
266	chr04:5713035-5713451	*EVC*
267	chr04:5891981-5892365	*CRMP1*
268	chr04:62065883-62068801	*Non Coding Region*
269	chr04:7194537-7195467	*SORCS2*
270	chr04:81187494-81187913	*FGF5*

Figure 60C

Order	Coordinate of CpG (hg19)	Gene
271	chr04:96469456-96469667	*UNC5C*
272	chr04:96469859-96470994	*UNC5C*
273	chr05:100238722-100239138	*ST8SIA4*
274	chr05:127872563-127874945	*FBN2*
275	chr05:128300800-128301329	*SLC27A6*
276	chr05:135265848-135266517	*FBXL21*
277	chr05:136834016-136835146	*SPOCK1*
278	chr05:140305712-140307193	*PCDHA2,PCDHA3,PCDHA1, etc*
279	chr05:140892913-140893189	*PCDHGB3,PCDHGB2,PCDHGB1, etc.*
280	chr05:141262988-141263195	*Non Coding Region*
281	chr05:1444678-1446648	*SLC6A3*
282	chr05:146257346-146258575	*PPP2R2B*
283	chr05:1667187-1668081	*Non Coding Region*
284	chr05:168727429-168728275	*SLIT3*
285	chr05:172661486-172662228	*NKX2-5*
286	chr05:172670829-172671824	*Non Coding Region*
287	chr05:174870753-174872345	*DRD1*
288	chr05:175792355-175793505	*ARL10*
289	chr05:178016558-178017670	*COL23A1*
290	chr05:178770724-178772794	*ADAMTS2*
291	chr05:180017099-180019062	*SCGB3A1*
292	chr05:180100700-180101209	*Non Coding Region*
293	chr05:1881924-1887743	*IRX4*
294	chr05:1930754-1931986	*Non Coding Region*
295	chr05:32709910-32714450	*NPR3*
296	chr05:33892191-33892403	*ADAMTS12*
297	chr05:33936168-33938309	*RXFP3*
298	chr05:3594467-3603054	*IRX1*
299	chr05:37836747-37840726	*GDNF*
300	chr05:38257825-38259136	*EGFLAM*
301	chr05:49736607-49737300	*EMB*
302	chr05:50265085-50265737	*Non Coding Region*
303	chr05:63801781-63802605	*RGS7BP*
304	chr05:63986189-63986974	*FAM159B*
305	chr05:71014917-71015715	*CARTPT*
306	chr05:76249435-76250528	*CRHBP*
307	chr05:82768387-82769268	*VCAN*
308	chr06:108485671-108490539	*NR2E1*
309	chr06:10887184-10887622	*SYCP2L*
310	chr06:11043913-11045206	*ELOVL2*
311	chr06:110678854-110679108	*C6orf186*
312	chr06:110797297-110798201	*SLC22A16*
313	chr06:117085739-117086942	*FAM162B*
314	chr06:118228042-118229811	*SLC35F1*
315	chr06:127439265-127441072	*RSPO3*

Order	Coordinate of CpG (hg19)	Gene
316	chr06:133562086-133563586	*EYA4*
317	chr06:134210639-134211218	*TCF21*
318	chr06:134638797-134639021	*SGK1*
319	chr06:1389139-1391393	*FOXF2*
320	chr06:151561283-151562550	*AKAP12*
321	chr06:152128822-152129771	*ESR1*
322	chr06:152957810-152958472	*SYNE1*
323	chr06:159589636-159591319	*FNDC1*
324	chr06:1624185-1625468	*GMDS*
325	chr06:166666837-166667541	*Non Coding Region*
326	chr06:28602853-28603295	*Non Coding Region*
327	chr06:29759946-29760903	*HCG4*
328	chr06:30095173-30095610	*Non Coding Region*
329	chr06:30418844-30419630	*Non Coding Region*
330	chr06:31782872-31785190	*HSPA1A,HSPA1L*
331	chr06:34111735-34113944	*Non Coding Region*
332	chr06:3750788-3752767	*C6orf145*
333	chr06:391188-393790	*IRF4*
334	chr06:41604583-41606663	*MDF1*
335	chr06:41606994-41607238	*MDF1*
336	chr06:62995855-62996228	*KHDRBS2*
337	chr06:73330942-73333109	*KCNQ5*
338	chr06:84417597-84419378	*SNAP91*
339	chr06:84562820-84563758	*RIPPLY2*
340	chr06:85472702-85474132	*TBX18*
341	chr06:85482569-85484718	*Non Coding Region*
342	chr06:87647253-87647707	*HTR1E*
343	chr06:91320890-91321219	*Non Coding Region*
344	chr06:94126272-94129677	*EPHA7*
345	chr07:101005899-101007443	*EMID2*
346	chr07:103629525-103630652	*RELN*
347	chr07:108095401-108097206	*NRCAM*
348	chr07:1269165-1282864	*UNCX*
349	chr07:127744194-127744554	*Non Coding Region*
350	chr07:142494563-142495248	*Non Coding Region*
351	chr07:150652807-150653080	*KCNH2*
352	chr07:150710349-150711333	*ATG9B,NOS3*
353	chr07:151106244-151107808	*WDR86*
354	chr07:152622343-152623305	*Non Coding Region*
355	chr07:155164557-155167854	*Non Coding Region*
356	chr07:156871054-156871297	*Non Coding Region*
357	chr07:157476886-157486719	*PTPRN2*
358	chr07:158936507-158938492	*VIPR2*
359	chr07:19146923-19147308	*Non Coding Region*
360	chr07:19156050-19158042	*TWIST1*

Figure 60D

Order	Coordinate of CpG (hg19)	Gene
361	chr07:19812569-19813005	*TMEM196*
362	chr07:21582443-21583313	*DNAH11*
363	chr07:24323558-24325080	*NPY*
364	chr07:28448716-28450028	*CREB5*
365	chr07:30721372-30722445	*CRHR2*
366	chr07:32337983-32338725	*Non Coding Region*
367	chr07:38670439-38671008	*AMPH*
368	chr07:43152020-43153340	*HECW1*
369	chr07:45197181-45197807	*RAMP3*
370	chr07:49813008-49815752	*VWC2*
371	chr07:4998405-4998950	*MMD2*
372	chr07:50342895-50343456	*IKZF1*
373	chr07:50343757-50344519	*IKZF1*
374	chr07:69062374-69065037	*AUTS2*
375	chr07:71800757-71802768	*CALN1*
376	chr07:751712-752150	*PRKAR1B*
377	chr07:89747892-89749036	*DPY19L2P4*
378	chr07:90225082-90226364	*Non Coding Region*
379	chr07:98467023-98468058	*TMEM130*
380	chr08:104510870-104513913	*RIMS2*
381	chr08:105235384-105235979	*RIMS2*
382	chr08:109094485-109095849	*RSPO2*
383	chr08:11204825-11205689	*TDH*
384	chr08:116660432-116660747	*TRPS1*
385	chr08:121137099-121137778	*COL14A1*
386	chr08:140714585-140718259	*KCNK9*
387	chr08:143531118-143534495	*Non Coding Region*
388	chr08:143591567-143592674	*BA11*
389	chr08:143858279-143859411	*LYNX1*
390	chr08:144240761-144242113	*LY6H*
391	chr08:15397636-15398287	*TUSC3*
392	chr08:16884363-16885339	*EFHA2*
393	chr08:17270603-17271120	*MTMR7*
394	chr08:22960384-22960927	*TNFRSF10C*
395	chr08:23571922-23572208	*Non Coding Region*
396	chr08:24770908-24772547	*NEFM*
397	chr08:24812946-24814299	*NEFL*
398	chr08:25900562-25905842	*Non Coding Region*
399	chr08:25909292-25909686	*Non Coding Region*
400	chr08:26721642-26724566	*ADRA1A*
401	chr08:32405407-32406901	*NRG1*
402	chr08:37822486-37824008	*ADRB3*
403	chr08:38644473-38645760	*TACC1*
404	chr08:41165852-41167140	*SFRP1*
405	chr08:494155-496083	*C8orf42*

Order	Coordinate of CpG (hg19)	Gene
406	chr08:53851701-53854426	*NPBWR1*
407	chr08:54163303-54164443	*OPRK1*
408	chr08:55366180-55367628	*Non Coding Region*
409	chr08:55370170-55372525	*SOX17*
410	chr08:55378928-55380186	*Non Coding Region*
411	chr08:57069687-57070047	*Non Coding Region*
412	chr08:58907001-58907822	*FAM110B*
413	chr08:60032532-60032746	*TOX*
414	chr08:65710990-65711722	*CYP7B1*
415	chr08:67344497-67344989	*ADHFE1*
416	chr08:67873388-67875600	*Non Coding Region*
417	chr08:686830-689496	*Non Coding Region*
418	chr08:69242920-69243803	*Non Coding Region*
419	chr08:72755783-72756667	*MSC*
420	chr08:73163618-73164191	*Non Coding Region*
421	chr08:75896528-75897116	*CRISPLD1*
422	chr08:77594255-77594860	*ZFHX4,LOC100192378*
423	chr08:85096759-85097247	*RALYL*
424	chr08:91997230-91997906	*Non Coding Region*
425	chr08:97156763-97158030	*GDF6*
426	chr08:97172634-97173880	*GDF6*
427	chr08:97505747-97507607	*SDC2*
428	chr08:9760750-9761643	*MIR124-1,LOC157627*
429	chr08:9762661-9764748	*Non Coding Region*
430	chr09:100615234-100617510	*FOXE1*
431	chr09:101705907-101706682	*COL15A1*
432	chr09:112402767-112403349	*PALM2*
433	chr09:129484841-129485887	*Non Coding Region*
434	chr09:132382432-132383004	*C9orf50*
435	chr09:133534534-133542394	*Non Coding Region*
436	chr09:137299190-137299437	*RXRA*
437	chr09:139090420-139095472	*LHX3*
438	chr09:37026222-37028014	*PAX5*
439	chr09:71788715-71789542	*TJP2*
440	chr09:77112712-77113583	*RORB*
441	chr09:841374-843364	*DMRT1*
442	chr09:88137609-88137985	*Non Coding Region*
443	chr10:100992156-100992687	*HPSE2*
444	chr10:100993820-100994188	*HPSE2*
445	chr10:102484200-102484476	*Non Coding Region*
446	chr10:102503913-102506146	*PAX2*
447	chr10:102507482-102509646	*PAX2*
448	chr10:103603166-103603642	*KCNIP2*
449	chr10:105036628-105038084	*INA*
450	chr10:111216604-111217083	*Non Coding Region*

Figure 60E

Order	Coordinate of CpG (hg19)	Gene
451	chr10:11207178-11207980	*CUGBP2*
452	chr10:118030732-118034230	*GFRA1*
453	chr10:118892161-118892639	*VAX1*
454	chr10:119000435-119001530	*SLC18A2*
455	chr10:124894075-124897366	*HMX3*
456	chr10:129534410-129537366	*FOXI2*
457	chr10:131761169-131771696	*Non Coding Region*
458	chr10:133794898-133796598	*BNIP3*
459	chr10:133998912-134001500	*DPYSL4*
460	chr10:134901198-134902619	*GPR123*
461	chr10:135043221-135044560	*UTF1*
462	chr10:13933482-13934184	*FRMD4A*
463	chr10:17270430-17272617	*VIM*
464	chr10:1778784-1780018	*ADARB2*
465	chr10:23462224-23463889	*Non Coding Region*
466	chr10:23487757-23488543	*Non Coding Region*
467	chr10:23982382-23982589	*KIAA1217*
468	chr10:23983366-23984978	*KIAA1217*
469	chr10:26504383-26507434	*GAD2*
470	chr10:26727061-26727992	*APBB1IP*
471	chr10:28030182-28035211	*MKX*
472	chr10:43600146-43601165	*RET*
473	chr10:50817601-50820356	*SLC18A3,CHAT*
474	chr10:64574660-64578778	*EGR2*
475	chr10:72043446-72043886	*Non Coding Region*
476	chr10:7449376-7455339	*SFMBT2*
477	chr10:8077829-8078378	*Non Coding Region*
478	chr10:82116202-82117120	*DYDC2,DYDC1*
479	chr10:83633820-83635729	*NRG3*
480	chr10:88122924-88127364	*GRID1*
481	chr10:91294947-91295925	*SLC16A12*
482	chr11:101453357-101454684	*TRPC6*
483	chr11:104034536-104035058	*PDGFD*
484	chr11:111383168-111383892	*BTG4,MIR34C,MIR34B*
485	chr11:112832524-112834490	*NCAM1*
486	chr11:115630398-115631117	*Non Coding Region*
487	chr11:119293320-119293943	*THY1*
488	chr11:122847494-122848608	*Non Coding Region*
489	chr11:122854930-122855582	*Non Coding Region*
490	chr11:124790814-124791354	*HEPACAM*
491	chr11:125757055-125758107	*HYLS1*
492	chr11:131780328-131781532	*NTM*
493	chr11:132934059-132934291	*OPCML*
494	chr11:133825337-133827457	*IGSF9B*
495	chr11:134145559-134147180	*GLB1L3*

Order	Coordinate of CpG (hg19)	Gene
496	chr11:134281380-134282919	*B3GAT1*
497	chr11:1410119-1412346	*BRSK2*
498	chr11:1768979-1770712	*HCCA2,LOC402778*
499	chr11:17740789-17743779	*MYOD1*
500	chr11:18812572-18814079	*PTPN5*
501	chr11:20181200-20182325	*DBX1*
502	chr11:20184570-20185497	*Non Coding Region*
503	chr11:20690579-20691845	*NELL1*
504	chr11:2158951-2162484	*IGF2,IGF2AS,INS-IGF2*
505	chr11:30606004-30608128	*MPPED2*
506	chr11:32454874-32457311	*WT1,WIT1*
507	chr11:33890357-33891495	*LMO2*
508	chr11:44330542-44331761	*ALX4*
509	chr11:46316875-46317485	*CREB3L1*
510	chr11:57194340-57194650	*SLC43A3*
511	chr11:63766058-63768666	*MACROD1*
512	chr11:69632033-69634710	*FGF3*
513	chr11:8190226-8190671	*RIC3*
514	chr11:8289532-8290322	*Non Coding Region*
515	chr11:829284-833558	*EFCAB4A*
516	chr11:91957635-91960290	*Non Coding Region*
517	chr11:94134226-94134848	*GPR83*
518	chr12:103351579-103352695	*ASCL1*
519	chr12:103889148-103889843	*C12orf42*
520	chr12:104850253-104852395	*CHST11*
521	chr12:1058481-1059430	*RAD52*
522	chr12:111471061-111471578	*CUX2*
523	chr12:111471960-111473546	*CUX2*
524	chr12:113013099-113013529	*Non Coding Region*
525	chr12:113494389-113495534	*DTX1*
526	chr12:114833911-114834210	*TBX5*
527	chr12:117798076-117799448	*NOS1*
528	chr12:119419105-119419810	*SRRM4*
529	chr12:120031494-120033212	*TMEM233*
530	chr12:128850549-128850755	*TMEM132C*
531	chr12:129337870-129338653	*GLT1D1*
532	chr12:130387609-130389139	*TMEM132D*
533	chr12:130645327-130648988	*FZD10*
534	chr12:132905449-132906206	*GALNT9*
535	chr12:15475318-15475901	*PTPRO*
536	chr12:22486835-22488666	*ST8SIA1*
537	chr12:25055599-25056246	*BCAT1*
538	chr12:25101607-25102073	*BCAT1*
539	chr12:29935995-29937433	*TMTC1*
540	chr12:41086522-41087102	*CNTN1*

Figure 60F

Order	Coordinate of CpG (hg19)	Gene
541	chr12:45444202-45445386	*DBX2*
542	chr12:5018585-5021171	*KCNA1*
543	chr12:50354839-50356163	*AQP5*
544	chr12:5153012-5154346	*KCNA5*
545	chr12:52400467-52401696	*GRASP*
546	chr12:54321301-54321721	*Non Coding Region*
547	chr12:54393374-54394648	*HOXC9*
548	chr12:5539895-5540703	*NTF3*
549	chr12:62584768-62586268	*FAM19A2*
550	chr12:64061836-64062774	*DPY19L2*
551	chr12:64784010-64784664	*C12orf56*
552	chr12:65514878-65515863	*WIF1*
553	chr12:66122788-66123328	*Non Coding Region*
554	chr12:72665683-72667551	*LOC283392,TRHDE*
555	chr12:81471569-81472119	*ACSS3*
556	chr12:8171336-8171790	*Non Coding Region*
557	chr12:85306282-85306829	*SLC6A15*
558	chr12:8548855-8549512	*Non Coding Region*
559	chr13:102068116-102069258	*NALCN*
560	chr13:102568425-102569495	*FGF14*
561	chr13:103046680-103047226	*FGF14*
562	chr13:103052361-103052944	*FGF14*
563	chr13:108519292-108521063	*FAM155A*
564	chr13:109147798-109149019	*Non Coding Region*
565	chr13:110958891-110960590	*COL4A2,COL4A1*
566	chr13:112758598-112760491	*Non Coding Region*
567	chr13:23733808-23734816	*Non Coding Region*
568	chr13:25620937-25621554	*Non Coding Region*
569	chr13:26042693-26043486	*ATP8A2*
570	chr13:26624725-26626265	*SHISA2*
571	chr13:28366549-28368505	*GSX1*
572	chr13:28371509-28371768	*Non Coding Region*
573	chr13:28498226-28499046	*PDX1*
574	chr13:29106576-29107254	*Non Coding Region*
575	chr13:36705040-36705707	*DCLK1*
576	chr13:36919737-36921004	*SPG20*
577	chr13:37005581-37006453	*CCNA1*
578	chr13:37247949-37248463	*C13orf36*
579	chr13:43148277-43149282	*TNFSF11*
580	chr13:49794125-49795542	*MLNR*
581	chr13:53419897-53422872	*PCDH8*
582	chr13:58206526-58208930	*PCDH17*
583	chr13:88323569-88324640	*SLITRK5*
584	chr13:88326600-88326919	*SLITRK5*
585	chr13:93879245-93880877	*GPC6*

Order	Coordinate of CpG (hg19)	Gene
586	chr13:95359641-95359874	*Non Coding Region*
587	chr13:95363152-95366051	*SOX21*
588	chr13:96204691-96205496	*CLDN10*
589	chr14:100437381-100438863	*Non Coding Region*
590	chr14:24803678-24804353	*ADCY4*
591	chr14:27066313-27066578	*NOVA1*
592	chr14:60975732-60978180	*SIX6*
593	chr14:62583679-62584279	*FLJ43390*
594	chr14:70041213-70041662	*Non Coding Region*
595	chr14:74706188-74708192	*VSX2*
596	chr14:76604999-76605268	*Non Coding Region*
597	chr14:77736733-77737772	*NGB*
598	chr14:85996494-85996958	*FLRT2*
599	chr14:96342209-96343570	*Non Coding Region*
600	chr14:97685059-97685946	*Non Coding Region*
601	chr15:26107503-26108818	*ATP10A*
602	chr15:27215951-27216856	*GABRG3*
603	chr15:28339774-28342442	*OCA2*
604	chr15:28351606-28353102	*Non Coding Region*
605	chr15:30483597-30484928	*Non Coding Region*
606	chr15:33602816-33604003	*RYR3*
607	chr15:34786492-34788138	*Non Coding Region*
608	chr15:35046443-35047480	*GJD2*
609	chr15:48936810-48938577	*FBN1*
610	chr15:51385861-51387203	*TNFAIP8L3*
611	chr15:55879792-55881228	*PYGO1*
612	chr15:74419870-74423044	*ISLR2,LOC283731*
613	chr15:79381804-79384209	*RASGRF1*
614	chr15:79574830-79575211	*ANKRD34C*
615	chr15:79576059-79576270	*ANKRD34C*
616	chr15:83315116-83317541	*CPEB1*
617	chr15:83348839-83349865	*AP3B2*
618	chr15:83775861-83776922	*TM6SF1*
619	chr15:83951980-83953930	*BNC1*
620	chr15:84115762-84116921	*SH3GL3*
621	chr15:88799522-88801017	*NTRK3*
622	chr15:92936677-92938521	*ST8SIA2*
623	chr15:93631428-93633173	*RGMA*
624	chr16:10273943-10277424	*GRIN2A*
625	chr16:1202414-1204624	*CACNA1H*
626	chr16:12994574-12997690	*SHISA9*
627	chr16:23846941-23848102	*PRKCB*
628	chr16:25702955-25704631	*HS3ST4*
629	chr16:28073817-28075296	*GSG1L*
630	chr16:31225956-31228264	*TRIM72,PYDC1*

Figure 60G

Order	Coordinate of CpG (hg19)	Gene
631	chr16:3232835-3234048	*Non Coding Region*
632	chr16:3238805-3239492	*Non Coding Region*
633	chr16:51183699-51188763	*SALL1*
634	chr16:51189800-51190260	*Non Coding Region*
635	chr16:55357521-55359040	*IRX6*
636	chr16:55513220-55513526	*MMP2*
637	chr16:55689598-55690895	*SLC6A2*
638	chr16:58018400-58019471	*TEPP*
639	chr16:66638254-66639561	*CMTM3*
640	chr16:67196792-67199988	*FBXL8,HSF4*
641	chr16:71264360-71264659	*HYDIN*
642	chr16:77468309-77469074	*ADAMTS18*
643	chr16:77822375-77823134	*VAT1L*
644	chr16:86530747-86532994	*Non Coding Region*
645	chr17:10101034-10102440	*GAS7*
646	chr17:13503829-13505559	*HS3ST3A1*
647	chr17:14212363-14212788	*HS3ST3B1*
648	chr17:26698359-26699557	*SARM1*
649	chr17:27939298-27940770	*ANKRD13B*
650	chr17:32905965-32908951	*C17orf102*
651	chr17:33814234-33814947	*SLFN12L*
652	chr17:35165323-35165983	*Non Coding Region*
653	chr17:35291899-35300875	*LHX1*
654	chr17:38347533-38347765	*RAPGEFL1*
655	chr17:45928211-45928710	*SP6*
656	chr17:46810609-46810945	*Non Coding Region*
657	chr17:48070698-48072654	*DLX3*
658	chr17:54674873-54675232	*Non Coding Region*
659	chr17:56234321-56234827	*MSX2P1*
660	chr17:6616422-6617471	*SLC13A5*
661	chr17:72347924-72348322	*KIF19*
662	chr17:75368688-75370506	*SEPTIN9*
663	chr17:7554139-7555338	*ATP1B2*
664	chr17:76228110-76228380	*LOC283999*
665	chr17:77178828-77180229	*HRNBP3*
666	chr17:77721246-77722497	*Non Coding Region*
667	chr17:78449507-78452783	*NPTX1*
668	chr17:8533004-8534933	*MYH10*
669	chr17:8868469-8869372	*PIK3R5*
670	chr17:8906236-8908080	*Non Coding Region*
671	chr18:11750953-11752756	*GNAL*
672	chr18:12254147-12255089	*CIDEA*
673	chr18:22930018-22930904	*ZNF521*
674	chr18:25755414-25758175	*CDH2*
675	chr18:31020510-31020847	*C18orf34*

Order	Coordinate of CpG (hg19)	Gene
676	chr18:35104533-35104942	*BRUNOL4*
677	chr18:35144907-35147628	*Non Coding Region*
678	chr18:53447074-53447842	*Non Coding Region*
679	chr18:55103154-55108853	*ONECUT2*
680	chr18:56887091-56887665	*GRP*
681	chr18:56934833-56936827	*RAX*
682	chr18:56939624-56941540	*RAX*
683	chr18:59000683-59001692	*Non Coding Region*
684	chr18:67067509-67069168	*DOK6*
685	chr18:7116852-7118241	*LAMA1*
686	chr18:76737005-76741244	*SALL3*
687	chr18:904578-909574	*ADCYAP1*
688	chr19:12203028-12203503	*ZNF788*
689	chr19:12266998-12267686	*ZNF625*
690	chr19:13616752-13617267	*CACNA1A*
691	chr19:1465206-1471241	*APC2*
692	chr19:15090016-15090873	*Non Coding Region*
693	chr19:18118741-18119553	*ARRDC2*
694	chr19:19322675-19322916	*NCAN*
695	chr19:19371675-19372393	*HAPLN4*
696	chr19:21646328-21646536	*Non Coding Region*
697	chr19:21657569-21658035	*Non Coding Region*
698	chr19:22034394-22034943	*Non Coding Region*
699	chr19:30015781-30021367	*VSTM2B*
700	chr19:30865683-30866490	*ZNF536*
701	chr19:34112279-34114353	*CHST8*
702	chr19:34972054-34973645	*WTIP*
703	chr19:36822395-36822690	*Non Coding Region*
704	chr19:37095680-37096589	*ZNF529,ZNF382*
705	chr19:37997790-37998125	*ZNF793*
706	chr19:41018525-41019486	*SPTBN4*
707	chr19:41595895-41596140	*CYP2A13*
708	chr19:44952416-44952809	*ZNF229*
709	chr19:50037078-50037670	*RCN3*
710	chr19:51111184-51111519	*Non Coding Region*
711	chr19:51830064-51831145	*IGLON5*
712	chr19:52873084-52873572	*ZNF880*
713	chr19:52900955-52901158	*ZNF528*
714	chr19:53635625-53636230	*ZNF415*
715	chr19:53811620-53812011	*Non Coding Region*
716	chr19:54023868-54024560	*ZNF331*
717	chr19:54411376-54411968	*Non Coding Region*
718	chr19:57683596-57683850	*Non Coding Region*
719	chr19:58238585-58239028	*ZNF671*
720	chr19:58609338-58609988	*ZSCAN18*

Figure 60H

Order	Coordinate of CpG (hg19)	Gene
721	chr19:58951214-58952250	*ZNF132*
722	chr19:7794793-7796410	*CLEC4G*
723	chr19:7852932-7854557	*CLEC4GP1*
724	chr20:10200088-10200384	*SNAP25*
725	chr20:21694472-21695344	*PAX1*
726	chr20:25058288-25058726	*VSX1*
727	chr20:2780978-2781497	*CPXM1*
728	chr20:30062969-30063281	*REM1*
729	chr20:30582788-30583099	*XKR7*
730	chr20:30639908-30640786	*HCK*
731	chr20:34188719-34190267	*FER1L4*
732	chr20:34893891-34895214	*Non Coding Region*
733	chr20:37302813-37303388	*Non Coding Region*
734	chr20:37352130-37357372	*SLC32A1*
735	chr20:37434206-37435592	*PPP1R16B*
736	chr20:39316550-39319987	*MAFB*
737	chr20:41817475-41819212	*PTPRT*
738	chr20:42875764-42876184	*GDAP1L1*
739	chr20:44941342-44941569	*Non Coding Region*
740	chr20:45142000-45142337	*ZNF334*
741	chr20:47934839-47935684	*Non Coding Region*
742	chr20:49639071-49640212	*KCNG1*
743	chr20:53091733-53092993	*DOK5*
744	chr20:55500347-55501102	*Non Coding Region*
745	chr20:57089460-57090237	*APCDD1L*
746	chr20:58179808-58180787	*PHACTR3*
747	chr20:61049361-61051897	*GATA5*
748	chr20:61806254-61810867	*Non Coding Region*
749	chr20:61884644-61886387	*NKAIN4,FLJ16779*
750	chr20:62102541-62103995	*KCNQ2*
751	chr20:8112884-8113592	*PLCB1*
752	chr21:19617098-19617874	*CHODL*
753	chr21:27011624-27012398	*JAM2*
754	chr21:28337856-28340237	*ADAMTS5*
755	chr21:34395128-34400245	*OLIG2*
756	chr21:47517651-47518999	*COL6A2*
757	chr22:19136293-19138512	*GSC2*
758	chr22:19510871-19512254	*CLDN5*
759	chr22:25678576-25679273	*Non Coding Region*
760	chr22:39853323-39854065	*MGAT3*
761	chr22:44257942-44258612	*SULT4A1*
762	chrX:6144120-6145906	*NLGN4X*
763	chrX:8698857-8700500	*KAL1*
	Alternative Markers: Lower in CRC for Group 1 set	
764	chr01:148853996-148855761	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
765	chr01:152487978-152488270	*CRCT1*
766	chr01:156405160-156406350	*Non Coding Region*
767	chr01:181451311-181452049	*CACNA1E*
768	chr01:50798667-50799536	*Non Coding Region*
769	chr01:65731411-65731849	*DNAJC6*
770	chr01:91190489-91192804	*Non Coding Region*
771	chr02:127643800-127644104	*Non Coding Region*
772	chr02:149632682-149633882	*KIF5C*
773	chr02:45240372-45241579	*Non Coding Region*
774	chr03:142681137-142683268	*PAQR9*
775	chr03:16554327-16555413	*RFTN1*
776	chr03:42947480-42948170	*ZNF662*
777	chr04:123747740-123748980	*FGF2*
778	chr04:81951941-81952808	*BMP3*
779	chr05:101632049-101632373	*SLCO4C1*
780	chr05:15500076-15500923	*FBXL7*
781	chr06:100051018-100051972	*Non Coding Region*
782	chr06:100060979-100062657	*PRDM13*
783	chr06:117591533-117592279	*VGLL2*
784	chr06:125283124-125284389	*STL*
785	chr06:1378445-1379318	*Non Coding Region*
786	chr06:153450992-153452792	*RGS17*
787	chr06:88875397-88877449	*CNR1*
788	chr07:140772358-140774560	*Non Coding Region*
789	chr07:27195601-27196567	*HOXA7*
790	chr07:4922707-4923576	*RADIL*
791	chr07:87229551-87229890	*ABCB1*
792	chr08:72459564-72460523	*Non Coding Region*
793	chr08:99438692-99440425	*KCNS2*
794	chr10:101292436-101292851	*NKX2-3*
795	chr10:102497154-102497675	*Non Coding Region*
796	chr10:118895963-118898037	*VAX1*
797	chr10:118922351-118923520	*Non Coding Region*
798	chr10:119301321-119305368	*EMX2OS,EMX2*
799	chr10:23480697-23482455	*PTF1A*
800	chr10:48438411-48439320	*GDF10*
801	chr10:94833272-94835256	*CYP26A1*
802	chr11:106888323-106890203	*GUCY1A2*
803	chr11:58672851-58673288	*Non Coding Region*
804	chr12:133484658-133485739	*Non Coding Region*
805	chr12:39299248-39299629	*CPNE8*
806	chr12:47224912-47225664	*Non Coding Region*
807	chr13:112726281-112728419	*Non Coding Region*
808	chr13:28674227-28675170	*FLT3*
809	chr13:79175610-79177985	*POU4F1*

Figure 60I

Order	Coordinate of CpG (hg19)	*Gene*
810	chr14:36002598-36005013	*INSM2*
811	chr14:36993488-36994488	*Non Coding Region*
812	chr14:52534581-52536722	*NID2*
813	chr15:60296135-60298520	*FOXB1*
814	chr15:83875648-83877079	*HDGFRP3*
815	chr16:86541430-86545228	*FOXF1*
816	chr17:79448218-79448827	*Non Coding Region*
817	chr18:5196243-5197389	*LOC642597*
818	chr18:74961556-74963822	*GALR1*
819	chr20:43438737-43439546	*RIMS4*
820	chr20:56803282-56804112	*Non Coding Region*
821	chr21:34442206-34444543	*OLIG1*

Figure 60J

Group 2 Methylation markers, 50% sensitivity, CpG Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 2 Markers	
1	Chr01: 12227700 12227701	*TNFRSF1B*
2	Chr01: 151694320 151694321	*C1orf230*
3	Chr01: 155043561 155043562	*Non Coding Region*
4	Chr01: 155043744 155043745	*Non Coding Region*
5	Chr01: 179545091 179545092	*NPHS2*
6	Chr01: 179545134 179545135	*NPHS2*
7	Chr01: 184633273 184633274	*Non Coding Region*
8	Chr01: 219347021 219347022	*LYPLAL1*
9	Chr01: 219347458 219347459	*LYPLAL1*
10	Chr01: 219834537 219834538	*Non Coding Region*
11	Chr01: 223937043 223937044	*CAPN2*
12	Chr01: 66999588 66999589	*SGIP1*
13	Chr01: 997858 997859	*Non Coding Region*
14	Chr02: 134325873 134325874	*NCKAP5*
15	Chr02: 219738314 219738315	*WNT6*
16	Chr02: 25354776 25354777	*EFR3B*
17	Chr02: 26407628 26407629	*FAM59B*
18	Chr02: 26408167 26408168	*FAM59B*
19	Chr02: 71192445 71192446	*ATP6V1B1*
20	Chr02: 85811432 85811433	*VAMP5*
21	Chr03: 10207069 10207070	*IRAK2*
22	Chr03: 137484364 137484365	*SOX14*
23	Chr03: 157261086 157261087	*C3orf55*
24	Chr03: 9904557 9904558	*Non Coding Region*
25	Chr04: 46391728 46391729	*GABRA2*
26	Chr05: 134827512 134827513	*Non Coding Region*
27	Chr05: 138728685 138728686	*LOC389333*
28	Chr05: 138728704 138728705	*LOC389333*
29	Chr05: 43019873 43019874	*Non Coding Region*
30	Chr05: 43020451 43020452	*Non Coding Region*
31	Chr05: 87976902 87976903	*LOC645323*
32	Chr06: 137810767 137810768	*Non Coding Region*
33	Chr06: 168197667 168197668	*C6orf123*
34	Chr06: 26044220 26044221	*HIST1H3C,HIST1H2BB*
35	Chr06: 27064769 27064770	*Non Coding Region*
36	Chr06: 28175578 28175579	*Non Coding Region*
37	Chr06: 3232246 3232247	*Non Coding Region*
38	Chr06: 7107108 7107109	*RREB1*
39	Chr07: 100091181 100091182	*C7orf51*
40	Chr07: 100091255 100091256	*C7orf51*
41	Chr07: 100091364 100091365	*C7orf51*
42	Chr07: 100203350 100203351	*PCOLCE*
43	Chr07: 23514347 23514348	*Non Coding Region*
44	Chr07: 3083338 3083339	*CARD11*
45	Chr07: 64349450 64349451	*Non Coding Region*
46	Chr07: 95025829 95025830	*PON3*
47	Chr07: 95025836 95025837	*PON3*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 2 Markers	
48	Chr08: 23021690 23021691	*TNFRSF10D*
49	Chr09: 129388668 129388669	*LMX1B*
50	Chr09: 129445599 129445600	*LMX1B*
51	Chr10: 119294173 119294174	*EMX2OS*
52	Chr10: 119296297 119296298	*EMX2OS*
53	Chr10: 18429760 18429761	*CACNB2*
54	Chr11: 2292691 2292692	*ASCL2*
55	Chr11: 61062902 61062903	*VWCE*
56	Chr11: 71952356 71952357	*PHOX2A*
57	Chr11: 76033391 76033392	*Non Coding Region*
58	Chr11: 76382149 76382150	*LRRC32*
59	Chr12: 104609476 104609477	*TXNRD1*
60	Chr12: 104609609 104609610	*TXNRD1*
61	Chr12: 116946163 116946164	*Non Coding Region*
62	Chr12: 116946354 116946355	*Non Coding Region*
63	Chr12: 130529951 130529952	*Non Coding Region*
64	Chr12: 14134486 14134487	*GRIN2B*
65	Chr12: 52652377 52652378	*Non Coding Region*
66	Chr13: 24844880 24844881	*SPATA13*
67	Chr14: 105512071 105512072	*Non Coding Region*
68	Chr14: 105512268 105512269	*Non Coding Region*
69	Chr14: 105512306 105512307	*Non Coding Region*
70	Chr14: 52734325 52734326	*PTGDR*
71	Chr14: 55348681 55348682	*GCH1*
72	Chr15: 41787780 41787781	*ITPKA*
73	Chr15: 79502127 79502128	*MIR184*
74	Chr17: 36204489 36204490	*LOC284100*
75	Chr17: 42083264 42083265	*PYY,NAGS*
76	Chr17: 42091713 42091714	*TMEM101*
77	Chr17: 42091909 42091910	*TMEM101*
78	Chr17: 42092403 42092404	*TMEM101*
79	Chr17: 42287971 42287972	*UBTF*
80	Chr17: 58498871 58498872	*C17orf64*
81	Chr17: 58498977 58498978	*C17orf64*
82	Chr18: 19745458 19745459	*Non Coding Region*
83	Chr18: 3499253 3499254	*DLGAP1*
84	Chr18: 3499253 3499254	*DLGAP1*
85	Chr19: 11354132 11354133	*DOCK6*
86	Chr19: 12978611 12978612	*MAST1*
87	Chr19: 13209548 13209549	*NFIX*
88	Chr19: 19932543 19932544	*ZNF506*
89	Chr19: 54410030 54410031	*PRKCG*
90	Chr19: 55592342 55592343	*EPS8L1*
91	Chr20: 3052483 3052484	*OXT*
92	Chr22: 19711023 19711024	*SEPT5,GP1BB*
93	Chr22: 19711036 19711037	*SEPT5,GP1BB*

Figure 61A

Order	Coordinate of CpG (hg19)	Gene
	Alternative markers for Group 2 set	
94	Chr01: 12227700 12227701	*TNFRSF1B*
95	Chr01: 151694320 151694321	*C1orf230*
96	Chr01: 155043561 155043562	*Non Coding Region*
97	Chr01: 155043744 155043745	*Non Coding Region*
98	Chr01: 156815274 156815275	*NTRK1,INSRR*
99	Chr01: 165322199 165322200	*LMX1A*
100	Chr01: 179545091 179545092	*NPHS2*
101	Chr01: 179545134 179545135	*NPHS2*
102	Chr01: 184633273 184633274	*Non Coding Region*
103	Chr01: 200011858 200011859	*NR5A2*
104	Chr01: 219347021 219347022	*LYPLAL1*
105	Chr01: 219347410 219347411	*LYPLAL1*
106	Chr01: 219347458 219347459	*LYPLAL1*
107	Chr01: 219834537 219834538	*Non Coding Region*
108	Chr01: 223937043 223937044	*CAPN2*
109	Chr01: 232940990 232940991	*KIAA1383*
110	Chr01: 232941176 232941177	*KIAA1383*
111	Chr01: 232941253 232941254	*KIAA1383*
112	Chr01: 241520598 241520599	*RGS7*
113	Chr01: 29101952 29101953	*Non Coding Region*
114	Chr01: 38600119 38600120	*Non Coding Region*
115	Chr01: 76540168 76540169	*ST6GALNAC3*
116	Chr01: 76540222 76540223	*ST6GALNAC3*
117	Chr01: 76540438 76540439	*ST6GALNAC3*
118	Chr01: 997858 997859	*Non Coding Region*
119	Chr02: 134325873 134325874	*NCKAP5*
120	Chr02: 155554688 155554689	*KCNJ3*
121	Chr02: 173099597 173099598	*Non Coding Region*
122	Chr02: 173100097 173100098	*Non Coding Region*
123	Chr02: 176988580 176988581	*HOXD9*
124	Chr02: 219735876 219735877	*WNT6*
125	Chr02: 219738314 219738315	*WNT6*
126	Chr02: 25354776 25354777	*EFR3B*
127	Chr02: 26407628 26407629	*FAM59B*
128	Chr02: 26408167 26408168	*FAM59B*
129	Chr02: 43020169 43020170	*HAAO*
130	Chr02: 74741136 74741137	*TLX2*
131	Chr02: 79739940 79739941	*CTNNA2*
132	Chr02: 85804732 85804733	*VAMP8*
133	Chr03: 10207069 10207070	*IRAK2*
134	Chr03: 137484364 137484365	*SOX14*
135	Chr03: 147140754 147140755	*Non Coding Region*
136	Chr03: 157261086 157261087	*C3orf55*
137	Chr03: 157261106 157261107	*C3orf55*

Order	Coordinate of CpG (hg19)	Gene
138	Chr03: 169482358 169482359	*Non Coding Region*
139	Chr03: 170137240 170137241	*CLDN11*
140	Chr03: 62861142 62861143	*CADPS*
141	Chr03: 62861145 62861146	*CADPS*
142	Chr03: 9904557 9904558	*Non Coding Region*
143	Chr04: 174431058 174431059	*Non Coding Region*
144	Chr04: 46391728 46391729	*GABRA2*
145	Chr05: 113697330 113697331	*KCNN2*
146	Chr05: 113697897 113697898	*KCNN2*
147	Chr05: 137475002 137475003	*NME5*
148	Chr05: 138728685 138728686	*LOC389333*
149	Chr05: 138728704 138728705	*LOC389333*
150	Chr05: 138729068 138729069	*LOC389333*
151	Chr05: 153862183 153862184	*Non Coding Region*
152	Chr05: 2751854 2751855	*C5orf38,IRX2*
153	Chr05: 43019873 43019874	*Non Coding Region*
154	Chr05: 43020451 43020452	*Non Coding Region*
155	Chr05: 57878390 57878391	*RAB3C*
156	Chr05: 57878953 57878954	*RAB3C*
157	Chr05: 87976889 87976890	*LOC645323*
158	Chr05: 87976902 87976903	*LOC645323*
159	Chr06: 137810767 137810768	*Non Coding Region*
160	Chr06: 166401464 166401465	*C6orf176,LOC441177*
161	Chr06: 168197667 168197668	*C6orf123*
162	Chr06: 26044220 26044221	*HIST1H3C,HIST1H2BB*
163	Chr06: 26045532 26045533	*HIST1H3C*
164	Chr06: 26045534 26045535	*HIST1H3C*
165	Chr06: 27064769 27064770	*Non Coding Region*
166	Chr06: 27858380 27858381	*HIST1H3J*
167	Chr06: 28175578 28175579	*Non Coding Region*
168	Chr06: 28367749 28367750	*ZSCAN12*
169	Chr06: 29692009 29692010	*HLA-F*
170	Chr06: 29692011 29692012	*HLA-F*
171	Chr06: 3232246 3232247	*Non Coding Region*
172	Chr06: 32920962 32920963	*HLA-DMA*
173	Chr06: 6320710 6320711	*F13A1*
174	Chr06: 7107108 7107109	*RREB1*
175	Chr06: 75794800 75794801	*COL12A1*
176	Chr06: 78176085 78176086	*Non Coding Region*
177	Chr07: 100091181 100091182	*C7orf51*
178	Chr07: 100091255 100091256	*C7orf51*
179	Chr07: 100091364 100091365	*C7orf51*
180	Chr07: 100202882 100202883	*PCOLCE*
181	Chr07: 100203350 100203351	*PCOLCE*
182	Chr07: 154862021 154862022	*HTR5A*

Figure 61B

Order	Coordinate of CpG (hg19)	Gene
183	Chr07: 23514347 23514348	*Non Coding Region*
184	Chr07: 3083338 3083339	*CARD11*
185	Chr07: 37960317 37960318	*EPDR1*
186	Chr07: 64349450 64349451	*Non Coding Region*
187	Chr07: 95025829 95025830	*PON3*
188	Chr08: 23021690 23021691	*TNFRSF10D*
189	Chr09: 129388668 129388669	*LMX1B*
190	Chr09: 129445599 129445600	*LMX1B*
191	Chr09: 136930919 136930920	*BRD3*
192	Chr10: 119294173 119294174	*EMX2OS*
193	Chr10: 119296297 119296298	*EMX2OS*
194	Chr10: 18429760 18429761	*CACNB2*
195	Chr11: 61062902 61062903	*VWCE*
196	Chr11: 63656090 63656091	*MARK2*
197	Chr11: 71952356 71952357	*PHOX2A*
198	Chr11: 76033391 76033392	*Non Coding Region*
199	Chr11: 76382149 76382150	*LRRC32*
200	Chr12: 104609476 104609477	*TXNRD1*
201	Chr12: 104609609 104609610	*TXNRD1*
202	Chr12: 116946163 116946164	*Non Coding Region*
203	Chr12: 116946354 116946355	*Non Coding Region*
204	Chr12: 130529951 130529952	*Non Coding Region*
205	Chr12: 14134486 14134487	*GRIN2B*
206	Chr12: 93966317 93966318	*SOCS2*
207	Chr12: 93966485 93966486	*SOCS2*
208	Chr13: 24844880 24844881	*SPATA13*
209	Chr13: 28491590 28491591	*Non Coding Region*
210	Chr13: 28492425 28492426	*Non Coding Region*
211	Chr14: 103655424 103655425	*Non Coding Region*
212	Chr14: 103655568 103655569	*Non Coding Region*
213	Chr14: 105512071 105512072	*Non Coding Region*
214	Chr14: 105512213 105512214	*Non Coding Region*
215	Chr14: 105512268 105512269	*Non Coding Region*
216	Chr14: 105512306 105512307	*Non Coding Region*
217	Chr14: 21101007 21101008	*Non Coding Region*
218	Chr14: 52734325 52734326	*PTGDR*
219	Chr14: 52734397 52734398	*PTGDR*
220	Chr14: 52735485 52735486	*PTGDR*
221	Chr14: 55348681 55348682	*GCH1*
222	Chr15: 33011166 33011167	*GREM1*
223	Chr15: 79502127 79502128	*MIR184*
224	Chr16: 72821566 72821567	*ZFHX3*
225	Chr17: 33673036 33673037	*Non Coding Region*
226	Chr17: 36105239 36105240	*HNF1B*
227	Chr17: 42083264 42083265	*PYY,NAGS*
228	Chr17: 42091713 42091714	*TMEM101*

Order	Coordinate of CpG (hg19)	Gene
229	Chr17: 42091909 42091910	*TMEM101*
230	Chr17: 42092187 42092188	*TMEM101*
231	Chr17: 42092247 42092248	*TMEM101*
232	Chr17: 42092315 42092316	*TMEM101*
233	Chr17: 42092370 42092371	*TMEM101*
234	Chr17: 42092403 42092404	*TMEM101*
235	Chr17: 42092431 42092432	*TMEM101*
236	Chr17: 42092450 42092451	*TMEM101*
237	Chr17: 42092472 42092473	*TMEM101*
238	Chr17: 42092488 42092489	*TMEM101*
239	Chr17: 46714130 46714131	*Non Coding Region*
240	Chr17: 58498871 58498872	*C17orf64*
241	Chr17: 58498977 58498978	*C17orf64*
242	Chr17: 59531696 59531697	*Non Coding Region*
243	Chr17: 9143754 9143755	*NTN1*
244	Chr18: 19745458 19745459	*Non Coding Region*
245	Chr18: 3499253 3499254	*DLGAP1*
246	Chr18: 5238589 5238590	*C18orf18,LOC339290*
247	Chr19: 11354132 11354133	*DOCK6*
248	Chr19: 12978611 12978612	*MAST1*
249	Chr19: 13209548 13209549	*NFIX*
250	Chr19: 14542965 14542966	*PKN1*
251	Chr19: 19932543 19932544	*ZNF506*
252	Chr19: 20278144 20278145	*ZNF486*
253	Chr19: 35531859 35531860	*HPN*
254	Chr19: 46283080 46283081	*DMPK*
255	Chr19: 50554451 50554452	*FLJ26850*
256	Chr19: 50836391 50836392	*Non Coding Region*
257	Chr19: 50836561 50836562	*Non Coding Region*
258	Chr19: 50836910 50836911	*Non Coding Region*
259	Chr19: 54410030 54410031	*PRKCG*
260	Chr19: 54485494 54485495	*CACNG8,MIR935*
261	Chr19: 54486058 54486059	*CACNG8*
262	Chr19: 55592342 55592343	*EPS8L1*
263	Chr19: 56915653 56915654	*ZNF583*
264	Chr19: 56915658 56915659	*ZNF583*
265	Chr19: 56915732 56915733	*ZNF583*
266	Chr19: 58011309 58011310	*ZNF773*
267	Chr19: 6482062 6482063	*DENND1C*
268	Chr20: 21503898 21503899	*Non Coding Region*
269	Chr20: 3052483 3052484	*OXT*
270	Chr22: 19711023 19711024	*SEPT5,GP1BB*
271	Chr22: 19711036 19711037	*SEPT5,GP1BB*
272	Chr22: 39417451 39417452	*APOBEC3D*
273	ChrX: 114957471 114957472	*Non Coding Region*

Figure 61C

Group 2 Methylation markers, 50% sensitivity, Chromosomal Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 2 Markers	
1	chr01:12226876-12227712	*TNFRSF1B*
2	chr01:151693991-151694282	*C1orf230*
3	chr01:155043412-155043922	*Non Coding Region*
4	chr01:179544720-179545307	*NPHS2*
5	chr01:184633223-184633663	*Non Coding Region*
6	chr01:219347109-219347572	*LYPLAL1*
7	chr01:223936342-223937044	*CAPN2*
8	chr01:66998728-66999060	*SGIP1*
9	chr01:999678-999911	*Non Coding Region*
10	chr02:219738081-219738788	*WNT6*
11	chr02:25354283-25354777	*EFR3B*
12	chr02:26407422-26408253	*FAM59B*
13	chr02:71192094-71192495	*ATP6V1B1*
14	chr02:85811340-85811855	*VAMP5*
15	chr03:10206449-10207024	*IRAK2*
16	chr03:137482964-137484454	*SOX14*
17	chr03:157260666-157261224	*C3orf55*
18	chr03:9904331-9904811	*Non Coding Region*
19	chr04:46391903-46392572	*GABRA2*
20	chr05:134827285-134827644	*Non Coding Region*
21	chr05:138727710-138730911	*LOC389333*
22	chr05:43017968-43018668	*Non Coding Region*
23	chr05:87976094-87976546	*LOC645323*
24	chr06:137809342-137810204	*Non Coding Region*
25	chr06:168197142-168198164	*C6orf123*
26	chr06:26044203-26044469	*HIST1H3C,HIST1H2BB*
27	chr06:27064628-27065004	*Non Coding Region*
28	chr06:28175282-28175605	*Non Coding Region*
29	chr06:3231595-3232289	*Non Coding Region*
30	chr06:7107075-7109854	*RREB1*
31	chr07:100091180-100091598	*C7orf51*
32	chr07:100201614-100203652	*PCOLCE*
33	chr07:23513534-23514412	*Non Coding Region*
34	chr07:64349424-64350151	*Non Coding Region*
35	chr07:95025559-95026122	*PON3*
36	chr08:23021056-23021588	*TNFRSF10D*
37	chr09:129386112-129389231	*LMX1B*
38	chr09:129445270-129445548	*LMX1B*
39	chr10:119293917-119297675	*EMX2OS*
40	chr10:18429125-18430234	*CACNB2*
41	chr11:2290104-2292932	*ASCL2*
42	chr11:61061751-61063063	*VWCE*
43	chr11:71952112-71952528	*PHOX2A*
44	chr11:76032911-76033392	*Non Coding Region*
45	chr11:76381449-76382295	*LRRC32*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 2 Markers	
46	chr12:104609397-104610172	*TXNRD1*
47	chr12:116946195-116946607	*Non Coding Region*
48	chr12:130526915-130527117	*Non Coding Region*
49	chr12:14134626-14135242	*GRIN2B*
50	chr12:52652018-52652743	*Non Coding Region*
51	chr14:105512055-105512307	*Non Coding Region*
52	chr14:52734207-52735486	*PTGDR*
53	chr15:41785616-41788539	*ITPKA*
54	chr17:42082027-42084972	*PYY,NAGS*
55	chr17:42091712-42091948	*TMEM101*
56	chr17:42092143-42092432	*TMEM101*
57	chr17:42287692-42288392	*UBTF*
58	chr17:58498711-58499332	*C17orf64*
59	chr18:19744936-19752363	*Non Coding Region*
60	chr18:3498941-3499358	*DLGAP1*
61	chr19:11353960-11354362	*DOCK6*
62	chr19:12978359-12978785	*MAST1*
63	chr19:13208343-13210525	*NFIX*
64	chr19:54409966-54410200	*PRKCG*
65	chr19:55591905-55593980	*EPS8L1*
66	chr20:3052097-3053103	*OXT*
67	chr22:19709081-19712106	*SEPT5,GP1BB*
	Alternative markers for Group 2 set	
68	chr01:156814881-156815792	*NTRK1,INSRR*
69	chr01:165321703-165322318	*LMX1A*
70	chr01:200011400-200012055	*NR5A2*
71	chr01:232941054-232941707	*KIAA1383*
72	chr01:241520103-241520790	*RGS7*
73	chr01:29101790-29102069	*Non Coding Region*
74	chr01:76540147-76540653	*ST6GALNAC3*
75	chr02:155554005-155556069	*KCNJ3*
76	chr02:173099482-173100098	*Non Coding Region*
77	chr02:176989337-176989587	*HOXD9*
78	chr02:219736132-219736592	*WNT6*
79	chr02:43019665-43020509	*HAAO*
80	chr02:74740455-74743795	*TLX2*
81	chr02:79739696-79740243	*CTNNA2*
82	chr03:147142181-147142391	*Non Coding Region*
83	chr03:169482338-169483052	*Non Coding Region*
84	chr03:170136242-170137886	*CLDN11*
85	chr03:62859617-62861190	*CADPS*
86	chr04:174430386-174430861	*Non Coding Region*
87	chr05:113696516-113699195	*KCNN2*
88	chr05:153862142-153862451	*Non Coding Region*
89	chr05:2748368-2757024	*C5orf38,IRX2*

Figure 62A

Order	Coordinate of CpG (hg19)	*Gene*
90	chr05:57878725-57879177	*RAB3C*
91	chr06:166401526-166402659	*C6orf176,LOC441177*
92	chr06:26045644-26046033	*HIST1H3C*
93	chr06:27858166-27858564	*HIST1H3J*
94	chr06:28367124-28367651	*ZSCAN12*
95	chr06:29691106-29692093	*HLA-F*
96	chr06:75794880-75795088	*COL12A1*
97	chr06:78172231-78174088	*Non Coding Region*
98	chr07:154861797-154862074	*HTR5A*
99	chr07:37960316-37961046	*EPDR1*
100	chr12:93963963-93967395	*SOCS2*
101	chr13:28491488-28492518	*Non Coding Region*
102	chr14:103655241-103655928	*Non Coding Region*
103	chr14:21100838-21101043	*Non Coding Region*
104	chr15:33009530-33011696	*GREM1*
105	chr16:72821140-72821592	*ZFHX3*
106	chr17:36105334-36105583	*HNF1B*
107	chr17:46713926-46714131	*Non Coding Region*
108	chr17:59531723-59535254	*Non Coding Region*
109	chr17:9142980-9143839	*NTN1*
110	chr18:5237723-5238674	*C18orf18,LOC339290*
111	chr19:14542768-14543102	*PKN1*
112	chr19:35531780-35531991	*HPN*
113	chr19:46282544-46283127	*DMPK*
114	chr19:50554012-50554492	*FLJ26850*
115	chr19:50836390-50837447	*Non Coding Region*
116	chr19:54485303-54486322	*CACNG8,MIR935*
117	chr19:56915356-56915856	*ZNF583*
118	chr19:58011124-58011743	*ZNF773*
119	chr20:21501961-21503931	*Non Coding Region*
120	chrX:114959730-115005442	*Non Coding Region*

Figure 62B

Group 3 Methylation markers, 50% sensitivity, CpG Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 3 Markers	
1	Chr01: 110611465 110611466	*ALX3*
2	Chr01: 156405438 156405439	*Non Coding Region*
3	Chr01: 170630558 170630559	*Non Coding Region*
4	Chr01: 170637723 170637724	*PRRX1*
5	Chr01: 200009830 200009831	*NR5A2*
6	Chr01: 200011858 200011859	*NR5A2*
7	Chr01: 223302861 223302862	*TLR5*
8	Chr01: 226288430 226288431	*Non Coding Region*
9	Chr01: 29586414 29586415	*PTPRU*
10	Chr01: 29586418 29586419	*PTPRU*
11	Chr01: 44883362 44883363	*RNF220*
12	Chr01: 47999163 47999164	*Non Coding Region*
13	Chr02: 173099597 173099598	*Non Coding Region*
14	Chr02: 175208605 175208606	*Non Coding Region*
15	Chr02: 176987462 176987463	*HOXD9*
16	Chr02: 176987465 176987466	*HOXD9*
17	Chr02: 177001263 177001264	*Non Coding Region*
18	Chr02: 177021826 177021827	*Non Coding Region*
19	Chr02: 177039862 177039863	*Non Coding Region*
20	Chr02: 45240563 45240564	*Non Coding Region*
21	Chr02: 62798365 62798366	*Non Coding Region*
22	Chr03: 107318032 107318033	*BBX*
23	Chr03: 181421427 181421428	*SOX2OT*
24	Chr04: 16085606 16085607	*PROM1*
25	Chr04: 56813898 56813899	*CEP135*
26	Chr05: 153862183 153862184	*Non Coding Region*
27	Chr05: 40681137 40681138	*PTGER4*
28	Chr05: 40681893 40681894	*PTGER4*
29	Chr05: 42995203 42995204	*Non Coding Region*
30	Chr05: 72595741 72595742	*Non Coding Region*
31	Chr05: 72732486 72732487	*Non Coding Region*
32	Chr06: 6004207 6004208	*NRN1*
33	Chr07: 113722758 113722759	*Non Coding Region*
34	Chr07: 113723162 113723163	*Non Coding Region*
35	Chr07: 113723569 113723570	*Non Coding Region*
36	Chr07: 27192061 27192062	*Non Coding Region*
37	Chr07: 27192246 27192247	*Non Coding Region*
38	Chr07: 27195918 27195919	*HOXA7*
39	Chr07: 27204005 27204006	*HOXA9*
40	Chr07: 27204349 27204350	*HOXA9*
41	Chr07: 27204981 27204982	*HOXA9*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 3 Markers	
42	Chr07: 27205224 27205225	*HOXA9*
43	Chr07: 27205230 27205231	*HOXA9*
44	Chr07: 27208285 27208286	*Non Coding Region*
45	Chr07: 27213610 27213611	*HOXA10*
46	Chr07: 27225058 27225059	*HOXA11AS,HOXA11*
47	Chr07: 27225523 27225524	*HOXA11AS,HOXA11*
48	Chr07: 27225528 27225529	*HOXA11AS,HOXA11*
49	Chr07: 42267747 42267748	*GLI3*
50	Chr07: 64349133 64349134	*Non Coding Region*
51	Chr07: 64349557 64349558	*Non Coding Region*
52	Chr07: 96650171 96650172	*DLX5*
53	Chr08: 101822108 101822109	*Non Coding Region*
54	Chr08: 124173231 124173232	*Non Coding Region*
55	Chr10: 122708532 122708533	*Non Coding Region*
56	Chr10: 122708739 122708740	*Non Coding Region*
57	Chr11: 68622739 68622740	*Non Coding Region*
58	Chr11: 75527994 75527995	*UVRAG*
59	Chr12: 54409491 54409492	*HOXC5,HOXC4,HOXC6*
60	Chr12: 57618943 57618944	*NXPH4*
61	Chr15: 33011166 33011167	*GREM1*
62	Chr15: 45479755 45479756	*SHF*
63	Chr15: 75251491 75251492	*Non Coding Region*
64	Chr15: 76633981 76633982	*ISL2*
65	Chr16: 23847325 23847326	*PRKCB*
66	Chr16: 31488238 31488239	*TGFB1I1*
67	Chr16: 56672387 56672388	*MT1A*
68	Chr16: 85932666 85932667	*IRF8*
69	Chr17: 3438933 3438934	*TRPV3*
70	Chr17: 46655394 46655395	*HOXB4*
71	Chr17: 46655588 46655589	*HOXB4*
72	Chr17: 46655818 46655819	*HOXB4*
73	Chr17: 46655829 46655830	*HOXB4*
74	Chr17: 46655836 46655837	*HOXB4*
75	Chr17: 53341243 53341244	*HLF*
76	Chr19: 6482062 6482063	*DENND1C*
77	Chr20: 21081901 21081902	*Non Coding Region*
78	Chr20: 55200146 55200147	*Non Coding Region*
79	Chr22: 19754251 19754252	*TBX1*
80	Chr22: 19754448 19754449	*TBX1*
81	Chr22: 46366859 46366860	*WNT7B*
82	Chr22: 46367617 46367618	*WNT7B*

Figure 63

Group 3 Methylation markers, 50% sensitivity, Chromosomal Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 3 Markers	
1	chr01:110610265-110613303	*ALX3*
2	chr01:156405160-156406350	*Non Coding Region*
3	chr01:170630456-170630851	*Non Coding Region*
4	chr01:170633274-170634178	*PRRX1*
5	chr01:200009807-200010036	*NR5A2*
6	chr01:200011400-200012055	*NR5A2*
7	chr01:29585897-29586598	*PTPRU*
8	chr01:44883136-44884272	*RNF220*
9	chr01:47998899-47999517	*Non Coding Region*
10	chr02:173099482-173100098	*Non Coding Region*
11	chr02:175208415-175209076	*Non Coding Region*
12	chr02:176986424-176988291	*HOXD9*
13	chr02:177001221-177001783	*Non Coding Region*
14	chr02:177024501-177025692	*Non Coding Region*
15	chr02:177039551-177039951	*Non Coding Region*
16	chr02:45240372-45241579	*Non Coding Region*
17	chr04:16084195-16085735	*PROM1*
18	chr04:56814800-56815511	*CEP135*
19	chr05:153862142-153862451	*Non Coding Region*
20	chr05:40679502-40682081	*PTGER4*
21	chr05:42995122-42995415	*Non Coding Region*
22	chr05:72594147-72595808	*Non Coding Region*
23	chr05:72732365-72733732	*Non Coding Region*
24	chr06:6002471-6005125	*NRN1*
25	chr07:113724924-113727795	*Non Coding Region*
26	chr07:27190274-27191115	*Non Coding Region*
27	chr07:27195601-27196567	*HOXA7*
28	chr07:27203915-27206462	*HOXA9*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 3 Markers	
29	chr07:27208871-27209616	*Non Coding Region*
30	chr07:27212416-27214396	*HOXA10*
31	chr07:27225050-27225629	*HOXA11AS,HOXA11*
32	chr07:42267546-42267823	*GLI3*
33	chr07:64349424-64350151	*Non Coding Region*
34	chr07:96650221-96651551	*DLX5*
35	chr08:124172800-124173541	*Non Coding Region*
36	chr10:122708510-122708899	*Non Coding Region*
37	chr11:68622722-68623252	*Non Coding Region*
38	chr11:75526396-75526926	*UVRAG*
39	chr12:54408426-54408713	*HOXC5,HOXC4,HOXC6*
40	chr12:57618769-57619402	*NXPH4*
41	chr15:33009530-33011696	*GREM1*
42	chr15:45479509-45480498	*SHF*
43	chr15:75251467-75251773	*Non Coding Region*
44	chr15:76632262-76635515	*ISL2*
45	chr16:23846941-23848102	*PRKCB*
46	chr16:31487152-31488967	*TGFB1I1*
47	chr16:56671937-56672530	*MT1A*
48	chr16:85932121-85932942	*IRF8*
49	chr17:3438841-3439046	*TRPV3*
50	chr17:46654053-46654369	*HOXB4*
51	chr17:46655215-46655604	*HOXB4*
52	chr17:53342198-53343061	*HLF*
53	chr20:21080929-21082824	*Non Coding Region*
54	chr20:55200279-55206789	*Non Coding Region*
55	chr22:19753312-19755013	*TBX1*
56	chr22:46366726-46368726	*WNT7B*

Figure 64

Group 4 Methylation markers, 50% sensitivity, CpG Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 4 Markers	
1	Chr01:_1072642_1072643	*Non Coding Region*
2	Chr01:_244893503_244893504	*Non Coding Region*
3	Chr01:_24648984_24648985	*GRHL3*
4	Chr01:_29586299_29586300	*PTPRU*
5	Chr01:_41848436_41848437	*Non Coding Region*
6	Chr02:_21022587_21022588	*C2orf43*
7	Chr02:_232479675_232479676	*Non Coding Region*
8	Chr03:_170625872_170625873	*EIF5A2*
9	Chr03:_196755973_196755974	*MFI2*
10	Chr04:_8271391_8271392	*HTRA3*
11	Chr04:_87281281_87281282	*MAPK10*
12	Chr05:_131800430_131800431	*Non Coding Region*
13	Chr05:_150536851_150536852	*ANXA6*
14	Chr05:_50263586_50263587	*Non Coding Region*
15	Chr05:_52778745_52778746	*FST*
16	Chr06:_106958303_106958304	*AIM1*
17	Chr06:_106958388_106958389	*AIM1*
18	Chr06:_106958474_106958475	*AIM1*
19	Chr06:_2903533_2903534	*SERPINB9*
20	Chr06:_2903535_2903536	*SERPINB9*
21	Chr06:_29692000_29692001	*HLA-F*
22	Chr06:_29894831_29894832	*HCG4P6*
23	Chr06:_29974251_29974252	*HLA-J,NCRNA00171*
24	Chr06:_29974717_29974718	*HLA-J,NCRNA00171*
25	Chr06:_29974971_29974972	*HLA-J,NCRNA00171*
26	Chr06:_29975068_29975069	*HLA-J,NCRNA00171*
27	Chr06:_31831553_31831554	*SLC44A4,NEU1*
28	Chr07:_141211627_141211628	*Non Coding Region*
29	Chr07:_141211956_141211957	*Non Coding Region*
30	Chr07:_3018391_3018392	*CARD11*
31	Chr08:_107669783_107669784	*OXR1*
32	Chr08:_107669856_107669857	*OXR1*
33	Chr10:_101280209_101280210	*Non Coding Region*
34	Chr10:_3109053_3109054	*PFKP*
35	Chr10:_43429459_43429460	*Non Coding Region*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 4 Markers	
36	Chr10:_45914276_45914277	*ALOX5*
37	Chr10:_73846858_73846859	*SPOCK2*
38	Chr11:_47208959_47208960	*PACSIN3*
39	Chr11:_67351490_67351491	*GSTP1*
40	Chr12:_54089497_54089498	*Non Coding Region*
41	Chr12:_54422350_54422351	*HOXC5,HOXC4,HOXC 6*
42	Chr12:_54427636_54427637	*HOXC5,HOXC4*
43	Chr12:_99139571_99139572	*ANKS1B*
44	Chr13:_113623233_113623234	*MCF2L*
45	Chr13:_28492425_28492426	*Non Coding Region*
46	Chr14:_24838008_24838009	*NFATC4*
47	Chr15:_70878049_70878050	*Non Coding Region*
48	Chr15:_90039822_90039823	*RHCG*
49	Chr15:_96888959_96888960	*Non Coding Region*
50	Chr16:_68118754_68118755	*NFATC3*
51	Chr17:_1131781_1131782	*Non Coding Region*
52	Chr17:_18908220_18908221	*SLC5A10,FAM83G*
53	Chr17:_18908235_18908236	*SLC5A10,FAM83G*
54	Chr17:_18908235_18908236	*SLC5A10,FAM83G*
55	Chr17:_18908242_18908243	*SLC5A10,FAM83G*
56	Chr17:_41363741_41363742	*TMEM106A*
57	Chr17:_41363899_41363900	*TMEM106A*
58	Chr17:_42402882_42402883	*SLC25A39*
59	Chr17:_48503163_48503164	*ACSF2*
60	Chr17:_74965708_74965709	*Non Coding Region*
61	Chr17:_75447478_75447479	*SEPTIN9*
62	Chr17:_75447504_75447505	*SEPTIN9*
63	Chr17:_76355701_76355702	*SOCS3*
64	Chr19:_10397642_10397643	*ICAM4*
65	Chr19:_11199917_11199918	*LDLR*
66	Chr19:_16187631_16187632	*TPM4*
67	Chr19:_17958647_17958648	*JAK3*
68	Chr19:_1857434_1857435	*KLF16*
69	Chr19:_49200058_49200059	*FUT2*
70	Chr20:_50158057_50158058	*NFATC2*

Figure 65

Group 4 Methylation markers, 50% sensitivity, Chromosomal Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 4 Markers	
1	chr01:1072369-1072847	*Non Coding Region*
2	chr01:244893441-244894366	*Non Coding Region*
3	chr01:24648202-24648985	*GRHL3*
4	chr01:29585897-29586598	*PTPRU*
5	chr01:41847264-41849204	*Non Coding Region*
6	chr02:21022564-21022934	*C2orf43*
7	chr02:232478359-232479925	*Non Coding Region*
8	chr03:170625831-170626841	*EIF5A2*
9	chr03:196755677-196757374	*MFI2*
10	chr04:8271214-8272091	*HTRA3*
11	chr05:150537019-150537418	*ANXA6*
12	chr05:50265085-50265737	*Non Coding Region*
13	chr05:52777788-52777996	*FST*
14	chr06:106958232-106958718	*AIM1*
15	chr06:2903175-2903906	*SERPINB9*
16	chr06:29691106-29692093	*HLA-F*
17	chr06:29894140-29895117	*HCG4P6*
18	chr06:29974220-29975369	*HLA-J,NCRNA00171*
19	chr06:31830299-31830948	*SLC44A4,NEU1*
20	chr08:107669777-107670591	*OXR1*
21	chr10:101279941-101280382	*Non Coding Region*
22	chr10:3108650-3111419	*PFKP*
23	chr10:43428167-43429460	*Non Coding Region*
24	chr10:45914374-45914883	*ALOX5*
25	chr10:73846806-73848233	*SPOCK2*
26	chr11:47207188-47209176	*PACSIN3*
27	chr11:67350928-67351953	*GSTP1*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 4 Markers	
28	chr12:54089211-54089413	*Non Coding Region*
29	chr12:54423427-54423712	*HOXC5,HOXC4,HOXC 6*
30	chr12:54427025-54428709	*HOXC5,HOXC4*
31	chr12:99139386-99139769	*ANKS1B*
32	chr13:113622737-113623660	*MCF2L*
33	chr13:28491488-28492518	*Non Coding Region*
34	chr14:24837872-24838324	*NFATC4*
35	chr15:70877380-70877958	*Non Coding Region*
36	chr15:90039464-90039984	*RHCG*
37	chr15:96888958-96889417	*Non Coding Region*
38	chr16:68118564-68119986	*NFATC3*
39	chr17:1131877-1133367	*Non Coding Region*
40	chr17:18906909-18908243	*SLC5A10,FAM83G*
41	chr17:41363727-41364273	*TMEM106A*
42	chr17:42402787-42403266	*SLC25A39*
43	chr17:48503056-48503887	*ACSF2*
44	chr17:74965089-74965303	*Non Coding Region*
45	chr17:75447477-75447821	*SEPTIN9*
46	chr17:76354818-76357038	*SOCS3*
47	chr19:10397603-10398703	*ICAM4*
48	chr19:11200919-11201686	*LDLR*
49	chr19:16186789-16188275	*TPM4*
50	chr19:17958490-17958808	*JAK3*
51	chr19:1856725-1857443	*KLF16*
52	chr19:49199964-49200184	*FUT2*
53	chr20:50158904-50159509	*NFATC2*

Figure 66

Group 5 ncRNA markers

Order	Gene ID	Coordinate (GRCh38)	ENSEMBL ID
1	AL365181.2	chr1:156597173-156599410	ENSG00000280316.1
2	SERTAD4-AS1	chr1:210231456-210234047	ENSG00000203706.7
3	RP11-333I13.1	chr2:46777783-46780245	ENSG00000260977.1
4	RP11-427M20.1	chr4:133871246-134010690	ENSG00000251388.1
5	CTB-35F21.4	chr5:139848290-139856389	ENSG00000250692.1
6	AC093668.2	chr7:102480080-102480165	ENSG00000271745.1
7	LINC00702	chr10:4201141-4243912	ENSG00000233117.2
8	RP11-455B2.9	chr10:15171036-15171278	ENSG00000228181.1
9	RP11-57H14.4	chr10:112823490-112827726	ENSG00000260917.1
10	RP11-288A5.2	chr10:127734562-127736996	ENSG00000279725.1
11	RP11-890B15.3	chr11:130866254-130870247	ENSG00000255455.2
12	RP11-131L12.4	chr12:118430147-118430699	ENSG00000275409.1
13	SACS-AS1	chr13:23418971-23428869	ENSG00000229558.2
14	RP11-227D13.1	chr15:48645951-48652016	ENSG00000259705.1
15	RP11-463O9.9	chr16:86520383-86523897	ENSG00000270020.1
16	CTC-429P9.1	chr19:16630743-16643942	ENSG00000269427.1
17	CTC-429P9.5	chr19:16639967-16640668	ENSG00000267904.1
18	AL022476.2	chr22:43038585-43052366	ENSG00000230319.1

Figure 67

Group 5 Exon RNA sequence markers

Order	Exon	Gene
1	chr1:26487703-26489118:+	FAM110D
2	chr1:38275239-38281238:-	MTF1
3	chr1:41512079-41512270:-	SCMH1
4	chr1:89578155-89578367:-	GBP2
5	chr1:89579699-89579979:-	GBP2
6	chr1:89582675-89582917:-	GBP2
7	chr1:89583260-89583456:-	GBP2
8	chr1:89585862-89585971:-	GBP2
9	chr1:89586826-89586953:-	GBP2
10	chr1:89587460-89587666:-	GBP2
11	chr1:89591546-89591799:-	GBP2
12	chr1:95362509-95363639:-	CNN3
13	chr1:112256830-112259310:+	
14	chr1:115259538-115260837:-	CSDE1
15	chr1:115261234-115261366:-	CSDE1
16	chr1:154143889-154143964:-	TPM3
17	chr1:154841508-154842754:-	KCNN3
18	chr1:154918048-154919279:-	PBXIP1
19	chr1:155291479-155292921:+	RUSC1
20	chr1:156924684-156924713:-	ARHGEF11
21	chr1:156933312-156933386:-	ARHGEF11
22	chr1:208059883-208061463:-	CD34
23	chr1:208062027-208062191:-	CD34
24	chr1:208062506-208062558:-	CD34
25	chr1:208062810-208062966:-	CD34
26	chr1:208070838-208070918:-	CD34
27	chr1:208072318-208072571:-	CD34
28	chr1:208073166-208073348:-	CD34
29	chr1:208084347-208084683:-	CD34
30	chr1:219254319-219255285:-	LOC643723
31	chr1:223175727-223179335:+	DISP1
32	chr2:24369800-24369956:+	LOC375190
33	chr2:24384376-24384483:+	LOC375190
34	chr2:24387068-24387204:+	LOC375190
35	chr2:24392225-24392506:+	LOC375190
36	chr2:27848511-27850784:-	CCDC121
37	chr2:30573784-30575295:+	
38	chr2:42946128-42946214:+	
39	chr2:42950029-42950175:+	
40	chr2:56611401-56613308:+	CCDC85A
41	chr2:66795799-66799890:+	MEIS1
42	chr2:100625274-100625394:-	AFF3
43	chr2:122407877-122407957:+	LOC254128
44	chr2:149267629-149267704:+	MBD5
45	chr2:149270439-149271042:+	MBD5

Order	Exon	Gene
46	chr2:152973316-152977316:-	STAM2
47	chr2:202241931-202245941:-	TRAK2
48	chr2:238678586-238678635:+	LRRFIP1
49	chr2:238680724-238680841:+	LRRFIP1
50	chr2:238681509-238681588:+	LRRFIP1
51	chr2:238683001-238683105:+	LRRFIP1
52	chr2:238688065-238690288:+	LRRFIP1
53	chr2:239969865-239974832:-	HDAC4
54	chr3:15111580-15116437:-	ZFYVE20
55	chr3:16298569-16302336:-	DPH3
56	chr3:42705729-42709072:+	ZBTB47
57	chr3:43618184-43618753:-	ANO10
58	chr3:44619303-44619329:+	
59	chr3:45959396-45963332:-	FYCO1
60	chr3:49765536-49765711:-	IP6K1
61	chr3:49770224-49770405:-	IP6K1
62	chr3:49775645-49775855:-	IP6K1
63	chr3:49785251-49785601:-	IP6K1
64	chr3:50595463-50597168:-	C3orf18
65	chr3:50604894-50605182:-	C3orf18
66	chr3:52283190-52284614:+	PPM1M
67	chr3:52489524-52489750:+	NISCH
68	chr3:52510463-52510615:+	NISCH
69	chr3:52512113-52512298:+	NISCH
70	chr3:52512465-52512593:+	NISCH
71	chr3:52513765-52513878:+	NISCH
72	chr3:52521212-52522621:+	NISCH
73	chr3:52523352-52523694:+	NISCH
74	chr3:52524048-52524247:+	NISCH
75	chr3:52524721-52524849:+	NISCH
76	chr3:52525368-52525529:+	NISCH
77	chr3:113005522-113006303:+	BOC
78	chr3:119360908-119361408:-	POPDC2
79	chr3:119367107-119367515:-	POPDC2
80	chr3:133651542-133653674:-	SLCO2A1
81	chr3:133654618-133654741:-	SLCO2A1
82	chr3:133657273-133657337:-	SLCO2A1
83	chr3:133661157-133661612:-	SLCO2A1
84	chr3:138117311-138117410:+	MRAS
85	chr3:138119376-138119455:+	MRAS
86	chr3:138121012-138124376:+	MRAS
87	chr3:156241616-156241736:+	KCNAB1
88	chr3:159570742-159571246:+	
89	chr4:5016316-5016961:-	CYTL1
90	chr4:17629629-17629791:+	

Figure 68A

Order	Exon	Gene
91	chr4:25199090-25202440:+	LOC285540
92	chr4:84406139-84406290:-	FAM175A
93	chr4:88394490-88394955:-	SPARCL1
94	chr4:88400582-88400730:-	SPARCL1
95	chr4:88401524-88401672:-	SPARCL1
96	chr4:88403576-88403712:-	SPARCL1
97	chr4:88411425-88411545:-	SPARCL1
98	chr4:88411912-88412030:-	SPARCL1
99	chr4:88412770-88412842:-	SPARCL1
100	chr4:88414716-88415750:-	SPARCL1
101	chr4:88416133-88416279:-	SPARCL1
102	chr4:88450193-88450655:-	SPARCL1
103	chr5:15936559-15939900:+	FBXL7
104	chr5:40711680-40716600:-	TTC33
105	chr5:88014058-88018742:-	MEF2C
106	chr5:92953432-92956835:-	FAM172A,MIR2277
107	chr5:102421704-102423876:-	GIN1
108	chr5:118456666-118456834:+	DMXL1
109	chr5:133496678-133496821:-	SKP1
110	chr5:137273642-137276037:-	FAM13B
111	chr5:138747381-138749961:-	DNAJC18
112	chr5:138837342-138837392:-	ECSCR
113	chr5:138837724-138837768:-	ECSCR
114	chr5:138842195-138842320:-	ECSCR
115	chr5:139231189-139231416:-	NRG2
116	chr5:139232017-139232144:-	NRG2
117	chr5:139235261-139235363:-	NRG2
118	chr5:139266925-139267096:-	NRG2
119	chr5:139505521-139508391:+	IGIP
120	chr5:140027384-140027509:+	IK,MIR3655
121	chr5:140081591-140081714:+	ZMAT2
122	chr5:158122924-158126150:-	EBF1
123	chr5:158134987-158135181:-	EBF1
124	chr5:158139162-158139341:-	EBF1
125	chr5:158139978-158140155:-	EBF1
126	chr5:158250184-158250325:-	EBF1
127	chr5:158523982-158524138:-	EBF1
128	chr5:158526353-158526788:-	EBF1
129	chr5:158527690-158527846:+	
130	chr5:158542120-158544486:+	
131	chr5:169805167-169805977:-	KCNMB1
132	chr5:176918405-176918421:-	PDLIM7
133	chr6:292101-292560:+	DUSP22
134	chr6:348103-348274:+	DUSP22
135	chr6:17463227-17463304:+	CAP2

Order	Exon	Gene
136	chr6:17507872-17507957:+	CAP2
137	chr6:17514080-17514185:+	CAP2
138	chr6:17539500-17539689:+	CAP2
139	chr6:17541204-17541379:+	CAP2
140	chr6:17543068-17543191:+	CAP2
141	chr6:17543292-17543374:+	CAP2
142	chr6:17551695-17551835:+	CAP2
143	chr6:17556590-17558021:+	CAP2
144	chr6:31367561-31368540:+	MICA
145	chr6:90340152-90343402:+	ANKRD6
146	chr6:117086310-117086418:-	FAM162B
147	chr6:132778663-132781989:-	STX7
148	chr7:11414173-11415505:-	THSD7A
149	chr7:28992976-28997497:-	TRIL
150	chr7:44078406-44078607:-	RASA4P
151	chr7:102133563-102133732:-	
152	chr7:102133563-102133758:-	
153	chr7:102135725-102135837:-	
154	chr7:102135770-102135837:-	
155	chr7:102256970-102257205:-	RASA4
156	chr7:150936050-150936242:-	SMARCD3
157	chr7:150936710-150936832:-	SMARCD3
158	chr7:150937198-150937333:-	SMARCD3
159	chr7:150937511-150939064:-	SMARCD3
160	chr8:59571858-59573985:+	
161	chr9:35684243-35684312:-	TPM2
162	chr9:35684485-35684547:-	TPM2
163	chr9:35684988-35685336:-	TPM2
164	chr9:35685431-35685548:-	TPM2
165	chr9:35685644-35685777:-	TPM2
166	chr9:35689143-35689268:-	TPM2
167	chr9:35689701-35690054:-	TPM2
168	chr9:35810880-35811054:-	SPAG8
169	chr9:35829513-35829952:+	TMEM8B
170	chr9:35841131-35841264:+	TMEM8B
171	chr9:35841523-35841791:+	TMEM8B
172	chr9:35842389-35842714:+	TMEM8B
173	chr9:35852824-35852970:+	TMEM8B
174	chr9:35853138-35853254:+	TMEM8B
175	chr9:35853502-35854843:+	TMEM8B
176	chr9:129265456-129269319:+	FAM125B
177	chr9:132649466-132655393:-	FNBP1
178	chr10:16632619-16635495:-	RSU1
179	chr10:28339923-28343173:-	MPP7
180	chr10:29756169-29758044:+	

Figure 68B

Order	Exon	Gene
181	chr10:29771762-29774635:+	
182	chr10:34398518-34399205:-	PARD3
183	chr10:34398809-34399990:+	
184	chr10:75196564-75198178:-	PPP3CB
185	chr10:93220184-93220322:+	HECTD2
186	chr10:93221011-93221113:+	HECTD2
187	chr10:96290959-96291217:+	TBC1D12
188	chr10:102307850-102313680:+	HIF1AN
189	chr10:112657786-112659763:+	PDCD4,MIR4680
190	chr10:114583255-114587482:+	
191	chr11:6416355-6416931:-	APBB1
192	chr11:6417009-6417192:-	APBB1
193	chr11:6417330-6417804:-	APBB1
194	chr11:6422219-6423212:-	APBB1
195	chr11:6423312-6424014:-	APBB1
196	chr11:10594638-10598122:-	MRVI1
197	chr11:58391507-58393203:+	ZFP91-CNTF,CNTF
198	chr11:77926343-77930461:-	GAB2
199	chr11:104776175-104776284:-	LOC643733
200	chr11:104779562-104779735:-	LOC643733
201	chr11:117075683-117078178:+	
202	chr11:117075789-117077072:-	PCSK7
203	chr11:118938493-118938913:+	VPS11
204	chr11:118944510-118944661:+	VPS11
205	chr11:118947607-118947793:+	VPS11
206	chr11:134019041-134021649:+	JAM3
207	chr12:2934515-2936465:-	NRIP2
208	chr12:2936791-2936843:-	NRIP2
209	chr12:2937092-2937213:-	NRIP2
210	chr12:2937498-2937580:-	NRIP2
211	chr12:2943808-2944221:-	NRIP2
212	chr12:6105168-6105388:-	VWF
213	chr12:6125920-6126036:-	VWF
214	chr12:46119810-46121530:-	LOC400027
215	chr12:48134425-48134606:-	RAPGEF3
216	chr12:48137313-48137459:-	RAPGEF3
217	chr12:48541575-48543781:-	ASB8
218	chr12:52308223-52308369:+	ACVRL1
219	chr12:52309009-52309284:+	ACVRL1
220	chr12:53473095-53473138:-	SPRYD3
221	chr12:56086628-56086759:-	ITGA7
222	chr12:56086925-56087101:-	ITGA7
223	chr12:56087805-56087907:-	ITGA7
224	chr12:56088040-56088114:-	ITGA7
225	chr12:56088227-56088306:-	ITGA7

Order	Exon	Gene
226	chr12:56088399-56088479:-	ITGA7
227	chr12:56088562-56088754:-	ITGA7
228	chr12:56089306-56089421:-	ITGA7
229	chr12:56090067-56090216:-	ITGA7
230	chr12:56090683-56090852:-	ITGA7
231	chr12:56091000-56091061:-	ITGA7
232	chr12:56091235-56091330:-	ITGA7
233	chr12:56092167-56092360:-	ITGA7
234	chr12:56101261-56101686:-	ITGA7
235	chr12:56385881-56388478:+	RAB5B
236	chr12:108960966-108963158:+	ISCU
237	chr12:109915065-109915155:-	KCTD10
238	chr13:113633655-113634072:+	
239	chr14:21558205-21559299:-	ZNF219
240	chr14:21559570-21559701:-	ZNF219
241	chr14:21560024-21561449:-	ZNF219
242	chr14:21668240-21672514:-	LOC283624
243	chr14:21989234-21993788:-	SALL2
244	chr14:24792550-24792660:-	ADCY4
245	chr14:55004440-55004554:+	CGRRF1
246	chr14:55004781-55005332:+	CGRRF1
247	chr14:74500731-74500874:+	C14orf45
248	chr14:74507268-74507348:+	C14orf45
249	chr15:42705022-42709558:-	ZFP106
250	chr15:43484941-43485533:+	CCNDBP1
251	chr15:43486925-43488228:+	CCNDBP1
252	chr15:45490970-45491318:-	SHF
253	chr15:52100617-52108556:+	TMOD2
254	chr15:63893495-63894618:+	FBXL22
255	chr15:64429767-64436431:+	SNX1
256	chr15:65236849-65236944:+	ANKDD1A
257	chr15:65242062-65242193:+	ANKDD1A
258	chr15:65345675-65347612:-	RASL12
259	chr15:65350765-65350955:-	RASL12
260	chr15:65351683-65351756:-	RASL12
261	chr15:74890843-74891878:+	
262	chr15:74893933-74894236:+	
263	chr15:74895508-74895587:+	
264	chr15:74897274-74898074:+	
265	chr15:89060013-89060204:-	DET1
266	chr15:90443832-90445035:-	C15orf38
267	chr16:2156092-2156305:-	PKD1
268	chr16:2156399-2156678:-	PKD1
269	chr16:2157884-2158033:-	PKD1
270	chr16:12618559-12618698:+	SNX29

Figure 68C

Order	Exon	Gene
271	chr16:49524522-49525215:-	ZNF423
272	chr16:67942583-67943991:+	PSKH1
273	chr17:6012927-6013086:+	WSCD1
274	chr17:6014091-6014255:+	WSCD1
275	chr17:6021308-6021508:+	WSCD1
276	chr17:17083921-17083983:+	MPRIP
277	chr17:18149622-18149776:-	FLII
278	chr17:18149908-18150142:-	FLII
279	chr17:21142186-21146742:-	C17orf103
280	chr17:40852294-40854610:-	EZH1
281	chr17:40854895-40854979:-	EZH1
282	chr17:40869994-40870085:-	EZH1
283	chr17:40870472-40870635:-	EZH1
284	chr17:40914358-40914481:+	RAMP2
285	chr17:40914615-40915057:+	RAMP2
286	chr17:41168324-41168565:-	VAT1
287	chr17:41843490-41847182:-	DUSP3
288	chr17:42158131-42158250:-	HDAC5
289	chr17:42160133-42160188:-	HDAC5
290	chr17:42160913-42161020:-	HDAC5
291	chr17:42161173-42161222:-	HDAC5
292	chr17:42161923-42162043:-	HDAC5
293	chr17:42162390-42162723:-	HDAC5
294	chr17:42163940-42164105:-	HDAC5
295	chr17:42164780-42165064:-	HDAC5
296	chr17:42165755-42165966:-	HDAC5
297	chr17:42168638-42168860:-	HDAC5
298	chr17:42170943-42171202:-	HDAC5
299	chr17:45129892-45131988:-	
300	chr17:58233921-58234076:+	CA4
301	chr17:58234788-58234933:+	CA4
302	chr17:58235051-58235149:+	CA4
303	chr17:58235644-58235807:+	CA4
304	chr17:58236591-58236904:+	CA4
305	chr17:74084852-74085004:-	EXOC7
306	chr17:74708914-74712804:-	JMJD6
307	chr18:11883474-11884626:-	MPPE1
308	chr18:11884878-11885042:-	MPPE1
309	chr18:32259432-32259556:+	DTNA
310	chr18:32335940-32336007:+	DTNA
311	chr18:32345925-32346005:+	DTNA
312	chr18:32374001-32374214:+	DTNA
313	chr18:32386183-32386268:+	DTNA
314	chr18:32391923-32392077:+	DTNA
315	chr18:32395873-32395978:+	DTNA

Order	Exon	Gene
316	chr18:32397926-32398294:+	DTNA
317	chr18:32400755-32400900:+	DTNA
318	chr18:32407557-32407640:+	DTNA
319	chr18:32428260-32428347:+	DTNA
320	chr18:32431795-32431892:+	DTNA
321	chr18:32438249-32438362:+	DTNA
322	chr18:32443930-32444026:+	DTNA
323	chr18:32446075-32448774:+	DTNA
324	chr18:32455203-32455362:+	DTNA
325	chr18:32459515-32459683:+	DTNA
326	chr18:32462033-32462165:+	DTNA
327	chr18:32467730-32471808:+	DTNA
328	chr18:32885940-32890729:+	ZNF271
329	chr18:34801980-34805287:+	KIAA1328
330	chr18:77119354-77123359:+	ATP9B
331	chr18:77133898-77134197:+	ATP9B
332	chr19:8483421-8483691:+	MAR.-2
333	chr19:8486673-8486900:+	MAR.-2
334	chr19:8491493-8491688:+	MAR.-2
335	chr19:8495542-8495751:+	MAR.-2
336	chr19:8503272-8503897:+	MAR.-2
337	chr19:9923907-9923950:-	
338	chr19:10908056-10908194:+	DNM2
339	chr19:12222061-12225491:+	ZNF788
340	chr19:15547617-15547940:-	WIZ
341	chr19:15558914-15559178:-	WIZ
342	chr19:18474201-18480763:+	PGPEP1
343	chr19:40519572-40523514:+	ZNF546
344	chr19:40899672-40904723:-	PRX
345	chr19:42875502-42875634:+	MEGF8
346	chr19:44038489-44038653:+	ZNF575
347	chr19:48578024-48578131:-	PLA2G4C
348	chr19:48581028-48581081:-	PLA2G4C
349	chr19:48588316-48588369:-	PLA2G4C
350	chr19:48591943-48591969:-	PLA2G4C
351	chr19:48593621-48593674:-	PLA2G4C
352	chr19:48598721-48598861:-	PLA2G4C
353	chr19:48601052-48601516:-	PLA2G4C
354	chr19:48602928-48603117:-	PLA2G4C
355	chr19:48607845-48607981:-	PLA2G4C
356	chr19:48608590-48608708:-	PLA2G4C
357	chr19:48613744-48614109:-	PLA2G4C
358	chr19:50380829-50381001:+	TBC1D17
359	chr19:50385298-50385409:+	TBC1D17
360	chr19:50385499-50385657:+	TBC1D17

Figure 68D

Order	Exon	Gene
361	chr19:50386021-50386149:+	TBC1D17
362	chr19:50386230-50386333:+	TBC1D17
363	chr20:420892-421033:-	TBC1D20
364	chr20:422232-422687:-	TBC1D20
365	chr21:27838529-27840953:-	CYYR1
366	chr21:34100426-34102972:+	GCFC1-AS1
367	chr21:34821089-34823170:-	TMEM50B
368	chr21:34961650-34962227:-	CRYZL1
369	chr21:34962147-34962227:-	CRYZL1
370	chr21:34963468-34963517:-	CRYZL1
371	chr21:34963468-34963677:-	CRYZL1
372	chr21:34968037-34968142:-	CRYZL1
373	chr21:34969586-34969707:-	CRYZL1
374	chr21:48055527-48055675:+	PRMT2
375	chr21:48063198-48063551:+	PRMT2
376	chr21:48064218-48064400:+	PRMT2
377	chr21:48068370-48068531:+	PRMT2
378	chr21:48069487-48069651:+	PRMT2
379	chr22:19428417-19431913:-	C22orf39
380	chr22:38142241-38142427:+	TRIOBP
381	chr22:38153620-38154145:+	TRIOBP
382	chr22:38155161-38155271:+	TRIOBP
383	chr22:38161677-38161824:+	TRIOBP
384	chr22:38164081-38164183:+	TRIOBP
385	chr22:38165035-38165194:+	TRIOBP
386	chr22:38165269-38165382:+	TRIOBP
387	chr22:38169790-38172562:+	TRIOBP
388	chr22:41791749-41795328:+	TEF
389	chrX:62854848-62858089:-	ARHGEF9
390	chrX:62898220-62898452:-	ARHGEF9
391	chrX:107169901-107174865:+	MID2
392	chrX:150868978-150870013:+	PRRG3
393	chr5:138784245-138784597:-	
394	chr5:138784767-138784865:-	

Figure 68E

Group 5 mRNA protein encoding markers

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
1	CA4	17q23.1	carbonic anhydrase IV	P22748
2	GPD1L	3p22.3	glycerol-3-phosphate dehydrogenase 1-like	Q8N335
3	PLCE1	10q23	phospholipase C, epsilon 1	Q9P212
4	GSN	9q33	gelsolin	P06396
5	MAPK3	16p11.2	mitogen-activated protein kinase 3	P27361
6	NRG2	5q23-q33	neuregulin 2	O14511
7	PAQR8	6p12	progestin and adipoQ receptor family member VIII	Q8TEZ7
8	CAPN2	1q41-q42	calpain 2, (m/II) large subunit	P17655
9	ZNF219	14q11	zinc finger protein 219	Q9P2Y4
10	PLCD3	17q21	phospholipase C, delta 3	Q8N3E9
11	PLXNA2	1q32.2	plexin A2	O75051
12	APBB1	11p15	amyloid beta (A4) precursor protein-binding, family B, member 1 (Fe65)	O00213
13	PDE1C	7p15.1-p14.3	phosphodiesterase 1C, calmodulin-dependent 70kDa	Q14123
14	KCTD2	17q25.2	potassium channel tetramerization domain containing 2	Q14681
15	KCNMB1	5q34	potassium large conductance calcium-activated channel, subfamily M, beta member 1	Q16558
16	ACTG2	2p13.1	actin, gamma 2, smooth muscle, enteric	P63267
17	TMEM108	3q21	transmembrane protein 108	Q6UXF1
18	JPH2	20q12-q13.11	junctophilin 2	Q9BR39
19	TRIOBP	22q13.1	TRIO and F-actin binding protein	Q9H2D6
20	TMBIM1	2q35	transmembrane BAX inhibitor motif containing 1	Q969X1
21	PRRG3	Xq28	proline rich Gla (G-carboxyglutamic acid) 3 (transmembrane)	Q9BZD7
22	IP6K1	3p21.31	inositol hexakisphosphate kinase 1	Q92551
23	DTNA	18q12	dystrobrevin, alpha	Q9Y4J8
24	C6orf89	6p21.31	chromosome 6 open reading frame 89	BRAP
25	FLNC	7q32-q35	filamin C, gamma	Q14315
26	RASL12	15q11.2-q22.33	RAS-like, family 12	Q9NYN1
27	SMARCD3	7q35-q36	SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily d, member 3	Q6STE5
28	DACH2	Xq21.3	dachshund homolog 2 (Drosophila)	Q96NX9
29	BOC	3q13.2	BOC cell adhesion associated, oncogene regulated	Q9BWV1
30	KCTD10	12q24.12	potassium channel tetramerization domain containing 10	Q9H3F6
31	PRX	19q13.2	periaxin	Q9BXM0
32	CD59	11p13	CD59 molecule, complement regulatory protein	P13987
33	GAB2	11q13.4-q13.5	GRB2-associated binding protein 2	Q9UQC2
34	VPS39	15q14	vacuolar protein sorting 39 homolog (S. cerevisiae)	Q96JC1
35	POPDC2	3q13.33	popeye domain containing 2	Q9HBU9
36	HIF1AN	10q24	hypoxia inducible factor 1, alpha subunit inhibitor	Q9NWT6
37	IL17B	5q33.1	interleukin 17B	
38	HECTD2	10q23.32	HECT domain containing E3 ubiquitin protein ligase 2	Q5U5R9
39	NOV	8q24.12	nephroblastoma overexpressed	PLXNA1
40	TACC2	10q26	transforming, acidic coiled-coil containing protein 2	O95359

Figure 69A

Order	Gene Symbol	Chromosome Band	Gene Title	UniProt ID
41	HDAC4	2q37.3	histone deacetylase 4	P56524
42	TPM2	9p13	tropomyosin 2 (beta)	P07951
43	DUSP22	6p25.3	dual specificity phosphatase 22	Q9NRW4
44	RGS5	1q23.1	regulator of G-protein signaling 5	O15539
45	TBC1D20	20p13	TBC1 domain family, member 20	Q96BZ9
46	MAP1A	15q15.3	microtubule-associated protein 1A	P78559
47	CD34	1q32	CD34 molecule	P28906
48	NRIP2	12p13.33	nuclear receptor interacting protein 2	Q9BQI9
49	EZH1	17q21.1-q21.3	enhancer of zeste homolog 1 (Drosophila)	Q92800
50	OPHN1	Xq12	oligophrenin 1	O60890
51	CXorf36	Xp11.3-p11.23	chromosome X open reading frame 36	Q9H7Y0
52	SLU7	5q33.3	SLU7 splicing factor homolog (S. cerevisiae)	O95391
53	SKP1	5q31	S-phase kinase-associated protein 1	P63208
54	DNAJC18	5q31.2	DnaJ (Hsp40) homolog, subfamily C, member 18	Q9H819
55	PKD1	16p13.3	polycystic kidney disease 1 (autosomal dominant)	P98161
56	LIMS3	2q13	LIM and senescent cell antigen-like domains 3	P0CW19
57	APOBEC2	6p21	apolipoprotein B mRNA editing enzyme, catalytic polypeptide-like 2	Q9Y235
58	FAM175A	4q21.23	family with sequence similarity 175, member A	Q6UWZ7
59	ANKDD1A	15q22.31	ankyrin repeat and death domain containing 1A	Q495B1
60	MECP2	Xq28	methyl CpG binding protein 2 (Rett syndrome)	P51608
61	RUFY2	10q22.2	RUN and FYVE domain containing 2	Q8WXA3

Figure 69B

Group 5 Methylation markers, 50% sensitivity, CpG Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 5 Markers	
1	Chr01: 113497999 113498000	*SLC16A1*
2	Chr01: 113498106 113498107	*SLC16A1*
3	Chr01: 14057606 14057607	*PRDM2*
4	Chr01: 154475139 154475140	*TDRD10,SHE*
5	Chr01: 160370208 160370209	*VANGL2*
6	Chr01: 203596940 203596941	*ATP2B4*
7	Chr01: 203597085 203597086	*ATP2B4*
8	Chr01: 228645486 228645487	*HIST3H2BB,HIST3H2A*
9	Chr01: 47999163 47999164	*Non Coding Region*
10	Chr01: 61548783 61548784	*NFIA*
11	Chr02: 176994665 176994666	*HOXD8*
12	Chr02: 233368543 233368544	*Non Coding Region*
13	Chr02: 238600061 238600062	*LRRFIP1*
14	Chr02: 26395448 26395449	*FAM59B*
15	Chr02: 74425580 74425581	*MTHFD2*
16	Chr03: 179168760 179168761	*GNB4*
17	Chr03: 184099432 184099433	*CHRD*
18	Chr04: 10020634 10020635	*SLC2A9*
19	Chr04: 76555782 76555783	*CDKL2*
20	Chr04: 76555856 76555857	*CDKL2*
21	Chr04: 89079587 89079588	*ABCG2*
22	Chr05: 43018282 43018283	*Non Coding Region*
23	Chr06: 26031939 26031940	*HIST1H3B*
24	Chr06: 26250744 26250745	*HIST1H3F,HIST1H2BH*
25	Chr06: 26250936 26250937	*HIST1H3F,HIST1H2BH*
26	Chr06: 26252265 26252266	*HIST1H3F,HIST1H2BH*
27	Chr06: 26271587 26271588	*HIST1H3G*
28	Chr06: 26271718 26271719	*HIST1H3G,HIST1H2BI*
29	Chr06: 26273556 26273557	*HIST1H2BI*
30	Chr06: 27840105 27840106	*HIST1H3I*
31	Chr06: 30139641 30139642	*TRIM15*
32	Chr06: 41978288 41978289	*CCND3*
33	Chr06: 91004452 91004453	*BACH2*
34	Chr07: 100224470 100224471	*TFR2*
35	Chr07: 100230985 100230986	*TFR2*
36	Chr07: 100231278 100231279	*TFR2*
37	Chr07: 148982284 148982285	*ZNF783*
38	Chr07: 155597842 155597843	*SHH*
39	Chr07: 27213610 27213611	*HOXA10*
40	Chr08: 101661918 101661919	*SNX31*
41	Chr08: 27183097 27183098	*PTK2B*
42	Chr08: 27183873 27183874	*PTK2B*
43	Chr10: 133484610 133484611	*Non Coding Region*
44	Chr10: 21807211 21807212	*C10orf140*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 5 Markers	
45	Chr10: 22625465 22625466	*Non Coding Region*
46	Chr10: 72218642 72218643	*Non Coding Region*
47	Chr10: 81002218 81002219	*ZMIZ1*
48	Chr11: 119227328 119227329	*USP2*
49	Chr11: 119227328 119227329	*USP2*
50	Chr11: 119227631 119227632	*USP2*
51	Chr11: 120435056 120435057	*Non Coding Region*
52	Chr11: 124790875 124790876	*HEPACAM*
53	Chr11: 13030873 13030874	*RASSF10*
54	Chr11: 2292691 2292692	*ASCL2*
55	Chr11: 57250469 57250470	*Non Coding Region*
56	Chr11: 64993281 64993282	*SLC22A20*
57	Chr11: 7695528 7695529	*CYB5R2*
58	Chr12: 102872708 102872709	*IGF1*
59	Chr12: 21810450 21810451	*LDHB*
60	Chr12: 21810489 21810490	*LDHB*
61	Chr12: 21810960 21810961	*LDHB*
62	Chr12: 3862225 3862226	*EFCAB4B*
63	Chr13: 79177782 79177783	*POU4F1*
64	Chr14: 105830859 105830860	*PACS2*
65	Chr14: 21152735 21152736	*ANG,RNASE4*
66	Chr14: 51027861 51027862	*ATL1*
67	Chr14: 88789549 88789550	*KCNK10*
68	Chr15: 45422095 45422096	*DUOX1,DUOXA1*
69	Chr15: 93617030 93617031	*RGMA*
70	Chr15: 99194021 99194022	*IGF1R*
71	Chr16: 23847325 23847326	*PRKCB*
72	Chr16: 86232457 86232458	*Non Coding Region*
73	Chr16: 88700818 88700819	*Non Coding Region*
74	Chr17: 76732854 76732855	*CYTH1*
75	Chr19: 11532175 11532176	*CCDC151*
76	Chr19: 13124776 13124777	*NFIX*
77	Chr19: 17403313 17403314	*ABHD8*
78	Chr19: 35633669 35633670	*FXYD7,FXYD1*
79	Chr19: 35633794 35633795	*FXYD7,FXYD1*
80	Chr19: 35633972 35633973	*FXYD7*
81	Chr19: 41316883 41316884	*Non Coding Region*
82	Chr19: 50554451 50554452	*FLJ26850*
83	Chr19: 54369681 54369682	*MYADM*
84	Chr20: 44539966 44539967	*PLTP*
85	Chr20: 62185401 62185402	*C20orf195*
86	Chr20: 825240 825241	*FAM110A*
87	Chr22: 38221187 38221188	*GALR3*

Figure 70A

Order	Coordinate of CpG (hg19)	Gene
	Alternative markers for Group 5 set	
88	Chr01: 113497999 113498000	*SLC16A1*
89	Chr01: 113498106 113498107	*SLC16A1*
90	Chr01: 12227700 12227701	*TNFRSF1B*
91	Chr01: 145395607 145395608	*Non Coding Region*
92	Chr01: 145395846 145395847	*Non Coding Region*
93	Chr01: 145455384 145455385	*Non Coding Region*
94	Chr01: 147245485 147245486	*GJA5*
95	Chr01: 151812710 151812711	*LOC100132111,C2CD4D*
96	Chr01: 154300117 154300118	*ATP8B2*
97	Chr01: 160370208 160370209	*VANGL2*
98	Chr01: 183188297 183188298	*LAMC2*
99	Chr01: 203259772 203259773	*Non Coding Region*
100	Chr01: 203596940 203596941	*ATP2B4*
101	Chr01: 203597085 203597086	*ATP2B4*
102	Chr01: 203598330 203598331	*ATP2B4*
103	Chr01: 203598594 203598595	*ATP2B4*
104	Chr01: 203598954 203598955	*ATP2B4*
105	Chr01: 203599089 203599090	*ATP2B4*
106	Chr01: 2144244 2144245	*Non Coding Region*
107	Chr01: 21616619 21616620	*ECE1*
108	Chr01: 228645486 228645487	*HIST3H2BB,HIST3H2A*
109	Chr01: 230560318 230560319	*Non Coding Region*
110	Chr01: 26686587 26686588	*Non Coding Region*
111	Chr01: 26686703 26686704	*Non Coding Region*
112	Chr01: 32410413 32410414	*Non Coding Region*
113	Chr01: 32714308 32714309	*FAM167B*
114	Chr01: 41250062 41250063	*KCNQ4*
115	Chr01: 41250071 41250072	*KCNQ4*
116	Chr01: 41250117 41250118	*KCNQ4*
117	Chr01: 55008319 55008320	*Non Coding Region*
118	Chr01: 55008516 55008517	*Non Coding Region*
119	Chr01: 61548783 61548784	*NFIA*
120	Chr01: 62761846 62761847	*KANK4*
121	Chr02: 128180829 128180830	*PROC*
122	Chr02: 128180959 128180960	*PROC*
123	Chr02: 128181325 128181326	*PROC*
124	Chr02: 131129567 131129568	*PTPN18*
125	Chr02: 131129811 131129812	*PTPN18*
126	Chr02: 131130131 131130132	*PTPN18*
127	Chr02: 131130468 131130469	*PTPN18*
128	Chr02: 160761413 160761414	*LY75*
129	Chr02: 177001263 177001264	*Non Coding Region*
130	Chr02: 233793088 233793089	*NGEF*
131	Chr02: 233793120 233793121	*NGEF*

Order	Coordinate of CpG (hg19)	Gene
132	Chr02: 24300158 24300159	*SF3B14*
133	Chr02: 24300242 24300243	*SF3B14*
134	Chr02: 264120 264121	*SH3YL1,ACP1*
135	Chr02: 264164 264165	*SH3YL1,ACP1*
136	Chr02: 264178 264179	*SH3YL1,ACP1*
137	Chr02: 264199 264200	*SH3YL1,ACP1*
138	Chr02: 264204 264205	*SH3YL1,ACP1*
139	Chr02: 42328740 42328741	*Non Coding Region*
140	Chr02: 44059094 44059095	*ABCG5*
141	Chr02: 74425523 74425524	*MTHFD2*
142	Chr02: 74425580 74425581	*MTHFD2*
143	Chr02: 85134468 85134469	*Non Coding Region*
144	Chr02: 8724060 8724061	*Non Coding Region*
145	Chr02: 9144629 9144630	*MBOAT2*
146	Chr02: 96055142 96055143	*Non Coding Region*
147	Chr03: 157824150 157824151	*SHOX2*
148	Chr03: 183542750 183542751	*MAP6D1*
149	Chr03: 183542914 183542915	*MAP6D1*
150	Chr03: 183543564 183543565	*MAP6D1*
151	Chr03: 184099259 184099260	*CHRD*
152	Chr03: 197281468 197281469	*BDH1*
153	Chr03: 197281934 197281935	*BDH1*
154	Chr03: 197282691 197282692	*BDH1*
155	Chr03: 197282902 197282903	*BDH1*
156	Chr03: 197283111 197283112	*BDH1*
157	Chr03: 197283127 197283128	*BDH1*
158	Chr03: 38080635 38080636	*DLEC1*
159	Chr03: 46671334 46671335	*Non Coding Region*
160	Chr03: 49756736 49756737	*AMIGO3,RNF123*
161	Chr03: 49757016 49757017	*AMIGO3,RNF123*
162	Chr03: 49757239 49757240	*AMIGO3,RNF123*
163	Chr03: 52567130 52567131	*NT5DC2*
164	Chr03: 55524048 55524049	*Non Coding Region*
165	Chr03: 72338583 72338584	*Non Coding Region*
166	Chr04: 10020555 10020556	*SLC2A9*
167	Chr04: 10020634 10020635	*SLC2A9*
168	Chr04: 113445029 113445030	*Non Coding Region*
169	Chr04: 41884631 41884632	*Non Coding Region*
170	Chr04: 76555634 76555635	*CDKL2*
171	Chr04: 76555772 76555773	*CDKL2*
172	Chr04: 76555777 76555778	*CDKL2*
173	Chr04: 76555782 76555783	*CDKL2*
174	Chr04: 76555856 76555857	*CDKL2*
175	Chr04: 76556042 76556043	*CDKL2*
176	Chr04: 89079587 89079588	*ABCG2*

Figure 70B

Order	Coordinate of CpG (hg19)	Gene
177	Chr05: 138861847 138861848	*TMEM173*
178	Chr05: 176784914 176784915	*RGS14*
179	Chr05: 176829639 176829640	*F12*
180	Chr05: 176830358 176830359	*F12*
181	Chr05: 176830667 176830668	*F12*
182	Chr05: 176831297 176831298	*F12*
183	Chr05: 176831638 176831639	*F12*
184	Chr05: 58571903 58571904	*PDE4D*
185	Chr06: 24360304 24360305	*Non Coding Region*
186	Chr06: 26031939 26031940	*HIST1H3B*
187	Chr06: 26235290 26235291	*HIST1H1D*
188	Chr06: 26250685 26250686	*HIST1H3F,HIST1H2BH*
189	Chr06: 26250744 26250745	*HIST1H3F,HIST1H2BH*
190	Chr06: 26250936 26250937	*HIST1H3F,HIST1H2BH*
191	Chr06: 26252265 26252266	*HIST1H3F,HIST1H2BH*
192	Chr06: 26271587 26271588	*HIST1H3G*
193	Chr06: 26271718 26271719	*HIST1H3G,HIST1H2BI*
194	Chr06: 26271827 26271828	*HIST1H3G,HIST1H2BI*
195	Chr06: 26272695 26272696	*HIST1H3G,HIST1H2BI*
196	Chr06: 26273556 26273557	*HIST1H2BI*
197	Chr06: 27774865 27774866	*HIST1H2AI*
198	Chr06: 27840105 27840106	*HIST1H3I*
199	Chr06: 27841352 27841353	*HIST1H3I,HIST1H4L*
200	Chr06: 27858380 27858381	*HIST1H3J*
201	Chr06: 30923241 30923242	*Non Coding Region*
202	Chr06: 30923243 30923244	*Non Coding Region*
203	Chr06: 30923306 30923307	*Non Coding Region*
204	Chr06: 30923327 30923328	*Non Coding Region*
205	Chr06: 30923370 30923371	*Non Coding Region*
206	Chr06: 30923545 30923546	*Non Coding Region*
207	Chr06: 30923672 30923673	*Non Coding Region*
208	Chr06: 33244976 33244977	*B3GALT4*
209	Chr06: 33245128 33245129	*B3GALT4*
210	Chr06: 33245163 33245164	*B3GALT4*
211	Chr06: 33245181 33245182	*B3GALT4*
212	Chr06: 33245328 33245329	*B3GALT4*
213	Chr06: 33245342 33245343	*B3GALT4*
214	Chr06: 33245359 33245360	*B3GALT4*
215	Chr06: 34203564 34203565	*HMGA1*
216	Chr06: 43044771 43044772	*PTK7*
217	Chr06: 52226698 52226699	*PAQR8*
218	Chr07: 100182075 100182076	*LRCH4*
219	Chr07: 100224470 100224471	*TFR2*
220	Chr07: 100230781 100230782	*TFR2*
221	Chr07: 100230985 100230986	*TFR2*

Order	Coordinate of CpG (hg19)	Gene
222	Chr07: 100231278 100231279	*TFR2*
223	Chr07: 102074615 102074616	*ORAI2*
224	Chr07: 112430772 112430773	*TMEM168*
225	Chr07: 112430779 112430780	*TMEM168*
226	Chr07: 12755396 12755397	*Non Coding Region*
227	Chr07: 138720989 138720990	*ZC3HAV1L*
228	Chr07: 139929714 139929715	*Non Coding Region*
229	Chr07: 143042635 143042636	*CLCN1*
230	Chr07: 143042741 143042742	*CLCN1*
231	Chr07: 148982284 148982285	*ZNF783*
232	Chr07: 151106022 151106023	*WDR86*
233	Chr07: 151330008 151330009	*PRKAG2*
234	Chr07: 155283817 155283818	*Non Coding Region*
235	Chr07: 155283886 155283887	*Non Coding Region*
236	Chr07: 155597490 155597491	*SHH*
237	Chr07: 25896490 25896491	*Non Coding Region*
238	Chr07: 27136424 27136425	*HOXA1*
239	Chr07: 27195918 27195919	*HOXA7*
240	Chr07: 27196153 27196154	*HOXA7*
241	Chr07: 27209350 27209351	*MIR196B*
242	Chr07: 27209582 27209583	*MIR196B*
243	Chr07: 27213984 27213985	*HOXA10*
244	Chr07: 855365 855366	*UNC84A*
245	Chr07: 87848292 87848293	*SRI*
246	Chr07: 87848358 87848359	*SRI*
247	Chr07: 99595437 99595438	*Non Coding Region*
248	Chr08: 101661918 101661919	*SNX31*
249	Chr08: 101661921 101661922	*SNX31*
250	Chr08: 11558334 11558335	*Non Coding Region*
251	Chr08: 11559156 11559157	*Non Coding Region*
252	Chr08: 142216031 142216032	*Non Coding Region*
253	Chr08: 145013728 145013729	*PLEC1*
254	Chr08: 145013910 145013911	*PLEC1*
255	Chr08: 145013932 145013933	*PLEC1*
256	Chr08: 22457446 22457447	*C8orf58*
257	Chr08: 27182871 27182872	*PTK2B*
258	Chr08: 27182889 27182890	*PTK2B*
259	Chr08: 27182909 27182910	*PTK2B*
260	Chr08: 27182930 27182931	*PTK2B*
261	Chr08: 27182965 27182966	*PTK2B*
262	Chr08: 27183097 27183098	*PTK2B*
263	Chr08: 27183342 27183343	*PTK2B*
264	Chr08: 27183873 27183874	*PTK2B*
265	Chr08: 37554832 37554833	*ZNF703*
266	Chr09: 103235181 103235182	*TMEFF1*

Figure 70C

Order	Coordinate of CpG (hg19)	Gene
312	Chr12: 109729809 109729810	*FOXN4*
313	Chr12: 110511112 110511113	*Non Coding Region*
314	Chr12: 122277439 122277440	*HPD*
315	Chr12: 122277851 122277852	*HPD*
316	Chr12: 12868983 12868984	*CDKN1B*
317	Chr12: 133065906 133065907	*FBRSL1*
318	Chr12: 133065912 133065913	*FBRSL1*
319	Chr12: 133065924 133065925	*FBRSL1*
320	Chr12: 133065928 133065929	*FBRSL1*
321	Chr12: 133065939 133065940	*FBRSL1*
322	Chr12: 133065941 133065942	*FBRSL1*
323	Chr12: 15114494 15114495	*ARHGDIB*
324	Chr12: 21809973 21809974	*LDHB*
325	Chr12: 21810092 21810093	*LDHB*
326	Chr12: 21810139 21810140	*LDHB*
327	Chr12: 21810450 21810451	*LDHB*
328	Chr12: 21810489 21810490	*LDHB*
329	Chr12: 21810960 21810961	*LDHB*
330	Chr12: 3862221 3862222	*EFCAB4B*
331	Chr12: 3862225 3862226	*EFCAB4B*
332	Chr12: 3862417 3862418	*EFCAB4B*
333	Chr12: 3862423 3862424	*EFCAB4B*
334	Chr12: 3862428 3862429	*EFCAB4B*
335	Chr12: 3862444 3862445	*EFCAB4B*
336	Chr12: 3862457 3862458	*EFCAB4B*
337	Chr12: 3862497 3862498	*EFCAB4B*
338	Chr12: 47474262 47474263	*AMIGO2*
339	Chr12: 47474763 47474764	*AMIGO2*
340	Chr12: 48207063 48207064	*HDAC7*
341	Chr12: 54070931 54070932	*ATP5G2*
342	Chr12: 54427884 54427885	*HOXC5,HOXC4*
343	Chr12: 54785003 54785004	*ZNF385A*
344	Chr12: 54785171 54785172	*ZNF385A*
345	Chr12: 54785286 54785287	*ZNF385A*
346	Chr12: 54785320 54785321	*ZNF385A*
347	Chr12: 56123961 56123962	*CD63*
348	Chr12: 56123972 56123973	*CD63*
349	Chr12: 98897216 98897217	*Non Coding Region*
350	Chr14: 105830859 105830860	*PACS2*
351	Chr14: 105940377 105940378	*CRIP2*
352	Chr14: 21152735 21152736	*ANG,RNASE4*
353	Chr14: 51027861 51027862	*ATL1*
354	Chr14: 54423433 54423434	*BMP4*
355	Chr14: 60794590 60794591	*Non Coding Region*
356	Chr14: 61188239 61188240	*SIX4*

Order	Coordinate of CpG (hg19)	Gene
357	Chr14: 62217806 62217807	*Non Coding Region*
358	Chr14: 65016591 65016592	*C14orf50*
359	Chr14: 65016595 65016596	*C14orf50*
360	Chr14: 65170209 65170210	*PLEKHG3*
361	Chr14: 74210521 74210522	*C14orf43*
362	Chr14: 75039090 75039091	*LTBP2*
363	Chr14: 88789549 88789550	*KCNK10*
364	Chr15: 31684836 31684837	*Non Coding Region*
365	Chr15: 41805273 41805274	*LTK*
366	Chr15: 45002662 45002663	*B2M*
367	Chr15: 45422095 45422096	*DUOX1,DUOXA1*
368	Chr15: 65341736 65341737	*OSTBETA*
369	Chr15: 65341868 65341869	*OSTBETA*
370	Chr15: 65341902 65341903	*OSTBETA*
371	Chr15: 65341995 65341996	*OSTBETA*
372	Chr15: 78556935 78556936	*DNAJA4*
373	Chr15: 85278581 85278582	*Non Coding Region*
374	Chr15: 90208810 90208811	*PLIN1*
375	Chr15: 90208915 90208916	*PLIN1*
376	Chr15: 96596349 96596350	*Non Coding Region*
377	Chr15: 99193929 99193930	*IGF1R*
378	Chr15: 99193993 99193994	*IGF1R*
379	Chr15: 99194021 99194022	*IGF1R*
380	Chr15: 99194431 99194432	*IGF1R*
381	Chr16: 29625235 29625236	*SLC7A5P1*
382	Chr16: 433504 433505	*LOC100134368*
383	Chr16: 87943182 87943183	*CA5A*
384	Chr16: 89641360 89641361	*CPNE7*
385	Chr17: 1390182 1390183	*MYO1C*
386	Chr17: 29297757 29297758	*RNF135*
387	Chr17: 29297873 29297874	*RNF135*
388	Chr17: 29298184 29298185	*RNF135*
389	Chr17: 34091901 34091902	*C17orf50*
390	Chr17: 36572011 36572012	*Non Coding Region*
391	Chr17: 41985074 41985075	*MPP2*
392	Chr17: 41985183 41985184	*MPP2*
393	Chr17: 42084612 42084613	*NAGS*
394	Chr17: 43325791 43325792	*LOC100133991*
395	Chr17: 46655394 46655395	*HOXB4*
396	Chr17: 46655588 46655589	*HOXB4*
397	Chr17: 46655818 46655819	*HOXB4*
398	Chr17: 46655829 46655830	*HOXB4*
399	Chr17: 46655836 46655837	*HOXB4*
400	Chr17: 61524073 61524074	*CYB561*
401	Chr17: 61524292 61524293	*CYB561*

Figure 70D

Order	Coordinate of CpG (hg19)	Gene
402	Chr17: 61524304 61524305	*CYB561*
403	Chr17: 62777744 62777745	*LOC146880*
404	Chr17: 6659189 6659190	*XAF1*
405	Chr17: 6659375 6659376	*XAF1*
406	Chr17: 6659562 6659563	*XAF1*
407	Chr17: 73083692 73083693	*SLC16A5*
408	Chr17: 73083876 73083877	*SLC16A5*
409	Chr17: 73083881 73083882	*SLC16A5*
410	Chr17: 73083887 73083888	*SLC16A5*
411	Chr17: 73893206 73893207	*TRIM65*
412	Chr17: 75315563 75315564	*SEPTIN9*
413	Chr17: 76732854 76732855	*CYTH1*
414	Chr17: 78992323 78992324	*Non Coding Region*
415	Chr17: 79482181 79482182	*Non Coding Region*
416	Chr17: 80291688 80291689	*SECTM1*
417	Chr17: 80291775 80291776	*SECTM1*
418	Chr18: 55862627 55862628	*NEDD4L*
419	Chr18: 55862678 55862679	*NEDD4L*
420	Chr19: 12978611 12978612	*MAST1*
421	Chr19: 12978624 12978625	*MAST1*
422	Chr19: 13124215 13124216	*NFIX*
423	Chr19: 18210017 18210018	*MAST3*
424	Chr19: 18499309 18499310	*GDF15*
425	Chr19: 19221316 19221317	*SLC25A42*
426	Chr19: 3434930 3434931	*NFIC*
427	Chr19: 3434945 3434946	*NFIC*
428	Chr19: 3435350 3435351	*NFIC*
429	Chr19: 35633553 35633554	*FXYD7,FXYD1*
430	Chr19: 35633669 35633670	*FXYD7,FXYD1*
431	Chr19: 35633794 35633795	*FXYD7,FXYD1*
432	Chr19: 35633972 35633973	*FXYD7*
433	Chr19: 35634229 35634230	*FXYD7*
434	Chr19: 35646150 35646151	*FXYD5*
435	Chr19: 36389795 36389796	*NFKBID*
436	Chr19: 36389833 36389834	*NFKBID*
437	Chr19: 39440462 39440463	*FBXO17*
438	Chr19: 39894767 39894768	*Non Coding Region*
439	Chr19: 41316883 41316884	*Non Coding Region*
440	Chr19: 43979464 43979465	*PHLDB3*
441	Chr19: 48232988 48232989	*EHD2*
442	Chr19: 48828835 48828836	*EMP3*
443	Chr19: 49255779 49255780	*FUT1*
444	Chr19: 50096610 50096611	*PRR12*
445	Chr19: 50400885 50400886	*IL4I1*
446	Chr19: 50554451 50554452	*FLJ26850*

Order	Coordinate of CpG (hg19)	Gene
447	Chr19: 54369436 54369437	*MYADM*
448	Chr19: 54369447 54369448	*MYADM*
449	Chr19: 54369471 54369472	*MYADM*
450	Chr19: 54369556 54369557	*MYADM*
451	Chr19: 54369571 54369572	*MYADM*
452	Chr19: 54369576 54369577	*MYADM*
453	Chr19: 54369681 54369682	*MYADM*
454	Chr19: 54370796 54370797	*MYADM*
455	Chr19: 54486058 54486059	*CACNG8*
456	Chr19: 55866412 55866413	*FAM71E2,COX6B2*
457	Chr19: 6740812 6740813	*TRIP10*
458	Chr19: 6740888 6740889	*TRIP10*
459	Chr19: 6740952 6740953	*TRIP10*
460	Chr19: 9896955 9896956	*Non Coding Region*
461	Chr19: 9896993 9896994	*Non Coding Region*
462	Chr20: 33759753 33759754	*PROCR*
463	Chr20: 42137055 42137056	*Non Coding Region*
464	Chr20: 44539966 44539967	*PLTP*
465	Chr20: 62185401 62185402	*C20orf195*
466	Chr20: 62285239 62285240	*STMN3*
467	Chr20: 62711729 62711730	*OPRL1,RGS19*
468	Chr20: 825052 825053	*FAM110A*
469	Chr20: 825240 825241	*FAM110A*
470	Chr20: 825408 825409	*FAM110A*
471	Chr21: 26934197 26934198	*MIR155HG*
472	Chr21: 26934576 26934577	*MIR155HG*
473	Chr21: 26934682 26934683	*MIR155HG*
474	Chr21: 26934885 26934886	*MIR155HG*
475	Chr22: 31644282 31644283	*LIMK2*
476	Chr22: 39410396 39410397	*APOBEC3C*
477	Chr22: 40390957 40390958	*FAM83F*
478	Chr22: 42679577 42679578	*Non Coding Region*

Figure 70E

Group 5 Methylation markers, 50% sensitivity, Chromosomal Regions

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 5 Markers	
1	chr01:113497986-113498206	*SLC16A1*
2	chr01:154474107-154475699	*TDRD10,SHE*
3	chr01:160370112-160370658	*VANGL2*
4	chr01:203598471-203598853	*ATP2B4*
5	chr01:228645196-228646434	*HIST3H2BB,HIST3H2A*
6	chr01:47998899-47999517	*Non Coding Region*
7	chr01:61548753-61549564	*NFIA*
8	chr02:176993479-176995557	*HOXD8*
9	chr02:233367780-233368577	*Non Coding Region*
10	chr02:238599857-238601430	*LRRFIP1*
11	chr02:26395416-26395620	*FAM59B*
12	chr02:74425444-74426423	*MTHFD2*
13	chr03:179168735-179169593	*GNB4*
14	chr03:184099095-184099582	*CHRD*
15	chr04:10020481-10021026	*SLC2A9*
16	chr04:76555366-76556079	*CDKL2*
17	chr04:89079756-89080454	*ABCG2*
18	chr05:43017968-43018668	*Non Coding Region*
19	chr06:26031880-26032264	*HIST1H3B*
20	chr06:26250436-26250827	*HIST1H3F,HIST1H2BH*
21	chr06:26251898-26252233	*HIST1H3F,HIST1H2BH*
22	chr06:26271345-26271588	*HIST1H3G*
23	chr06:26272470-26272696	*HIST1H3G,HIST1H2BI*
24	chr06:26273290-26273557	*HIST1H2BI*
25	chr06:27841001-27841244	*HIST1H3I*
26	chr06:30139718-30140263	*TRIM15*
27	chr06:91004795-91006944	*BACH2*
28	chr07:100224270-100224558	*TFR2*
29	chr07:100230649-100231320	*TFR2*
30	chr07:148982079-148982675	*ZNF783*
31	chr07:155595692-155599414	*SHH*
32	chr07:27212416-27214396	*HOXA10*
33	chr08:101661671-101662022	*SNX31*
34	chr08:27183090-27183390	*PTK2B*
35	chr10:21805197-21805459	*C10orf140*
36	chr10:22623350-22625875	*Non Coding Region*
37	chr10:72218130-72218484	*Non Coding Region*
38	chr10:81002109-81003687	*ZMIZ1*
39	chr11:119227096-119227758	*USP2*
40	chr11:120434822-120435940	*Non Coding Region*
41	chr11:124790814-124791354	*HEPACAM*
42	chr11:13030330-13032584	*RASSF10*
43	chr11:2290104-2292932	*ASCL2*
44	chr11:57249946-57250848	*Non Coding Region*
45	chr11:64993280-64993484	*SLC22A20*

Order	Coordinate of CpG (hg19)	Gene
	Preferred Group 5 Markers	
46	chr11:7694711-7695685	*CYB5R2*
47	chr12:21810488-21810766	*LDHB*
48	chr12:3862068-3862606	*EFCAB4B*
49	chr13:79175610-79177985	*POU4F1*
50	chr14:105830605-105830846	*PACS2*
51	chr14:21152483-21152740	*ANG,RNASE4*
52	chr14:51026872-51027570	*ATL1*
53	chr14:88789500-88790493	*KCNK10*
54	chr15:45421236-45422394	*DUOX1,DUOXA1*
55	chr15:93615622-93617185	*RGMA*
56	chr15:99190446-99194559	*IGF1R*
57	chr16:23846941-23848102	*PRKCB*
58	chr16:88700455-88701573	*Non Coding Region*
59	chr19:11533198-11533619	*CCDC151*
60	chr19:13124569-13124788	*NFIX*
61	chr19:17403039-17403570	*ABHD8*
62	chr19:35633409-35633697	*FXYD7,FXYD1*
63	chr19:35633847-35634629	*FXYD7,FXYD1*
64	chr19:41316543-41317318	*Non Coding Region*
65	chr19:50554012-50554492	*FLJ26850*
66	chr19:54369387-54369809	*MYADM*
67	chr20:44539729-44540099	*PLTP*
68	chr20:62185073-62185572	*C20orf195*
69	chr20:825267-826276	*FAM110A*
70	chr22:38220589-38221619	*GALR3*
	Alternative markers for Group 5 set	
71	chr01:12226876-12227712	*TNFRSF1B*
72	chr01:151810460-151811684	*LOC100132111,C2CD4D*
73	chr01:154300975-154301528	*ATP8B2*
74	chr01:2144199-2144497	*Non Coding Region*
75	chr01:21616380-21617101	*ECE1*
76	chr01:230561103-230562702	*Non Coding Region*
77	chr01:26686516-26687281	*Non Coding Region*
78	chr01:32410188-32410630	*Non Coding Region*
79	chr01:32713918-32714186	*FAM167B*
80	chr01:41249090-41250161	*KCNQ4*
81	chr01:55007704-55008448	*Non Coding Region*
82	chr02:128180294-128181381	*PROC*
83	chr02:131129718-131130511	*PTPN18*
84	chr02:160760604-160761452	*LY75*
85	chr02:177001221-177001783	*Non Coding Region*
86	chr02:233791783-233793922	*NGEF*
87	chr02:24300060-24300294	*SF3B14*
88	chr02:263400-265238	*SH3YL1,ACP1*
89	chr02:42328300-42328827	*Non Coding Region*

Figure 71A

Order	Coordinate of CpG (hg19)	Gene
90	chr02:44058765-44059232	*ABCG5*
91	chr02:85132338-85133083	*Non Coding Region*
92	chr02:8723855-8724093	*Non Coding Region*
93	chr02:9143127-9144630	*MBOAT2*
94	chr02:96054894-96055143	*Non Coding Region*
95	chr03:157822973-157823836	*SHOX2*
96	chr03:183542496-183543804	*MAP6D1*
97	chr03:197281605-197283128	*BDH1*
98	chr03:38080628-38081187	*DLEC1*
99	chr03:49755763-49757317	*AMIGO3,RNF123*
100	chr03:52567009-52568807	*NT5DC2*
101	chr03:55520226-55522344	*Non Coding Region*
102	chr04:113445022-113445350	*Non Coding Region*
103	chr04:41882450-41882964	*Non Coding Region*
104	chr05:176829339-176829756	*F12*
105	chr05:176830275-176831639	*F12*
106	chr06:24360029-24360411	*Non Coding Region*
107	chr06:27775329-27776258	*HIST1H2AI*
108	chr06:27839698-27840069	*HIST1H3I,HIST1H4L*
109	chr06:27858166-27858564	*HIST1H3J*
110	chr06:30923208-30923624	*Non Coding Region*
111	chr06:33244677-33245554	*B3GALT4*
112	chr06:34202567-34206193	*HMGA1*
113	chr06:43044044-43045057	*PTK7*
114	chr06:52226753-52228006	*PAQR8*
115	chr07:100183527-100184625	*LRCH4*
116	chr07:102073605-102074334	*ORAI2*
117	chr07:112429980-112430566	*TMEM168*
118	chr07:138720044-138721019	*ZC3HAV1L*
119	chr07:139930065-139930356	*Non Coding Region*
120	chr07:143042508-143042867	*CLCN1*
121	chr07:151106244-151107808	*WDR86*
122	chr07:155284018-155284356	*Non Coding Region*
123	chr07:25896713-25896925	*Non Coding Region*
124	chr07:27135342-27136736	*HOXA1*
125	chr07:27195601-27196567	*HOXA7*
126	chr07:27208871-27209616	*MIR196B*
127	chr07:855308-856747	*UNC84A*
128	chr07:87849003-87849594	*SRI*
129	chr07:99595206-99595505	*Non Coding Region*
130	chr08:11559596-11562956	*Non Coding Region*
131	chr08:142215691-142216299	*Non Coding Region*
132	chr08:145016417-145016916	*PLEC1*
133	chr08:22457123-22457753	*C8orf58*
134	chr08:37551944-37555922	*ZNF703*
135	chr09:103235171-103236627	*TMEFF1*

Order	Coordinate of CpG (hg19)	Gene
136	chr09:112810858-112811128	*PALM2-AKAP2,AKAP2*
137	chr09:79073907-79074561	*GCNT1*
138	chr10:102882977-102883551	*TLX1NB*
139	chr10:103879633-103880628	*LDB1*
140	chr10:113943283-113943657	*GPAM*
141	chr10:135089969-135090491	*ADAM8*
142	chr10:17270430-17272617	*VIM*
143	chr10:76584625-76586746	*Non Coding Region*
144	chr10:88730554-88731632	*AGAP11*
145	chr11:2158951-2162484	*IGF2,IGF2AS,INS-IGF2*
146	chr11:66623620-66626614	*PC,LRFN4*
147	chr11:68778569-68778851	*MRGPRF*
148	chr11:69258149-69258544	*Non Coding Region*
149	chr11:73693702-73694802	*UCP2*
150	chr11:75917271-75917926	*WNT11*
151	chr11:77122736-77123088	*PAK1*
152	chr12:109729572-109729826	*FOXN4*
153	chr12:122277301-122277539	*HPD*
154	chr12:12869798-12871248	*CDKN1B*
155	chr12:133065843-133066393	*FBRSL1*
156	chr12:47473177-47474120	*AMIGO2*
157	chr12:48206721-48207126	*HDAC7*
158	chr12:54071053-54071265	*ATP5G2*
159	chr12:54427025-54428709	*HOXC5,HOXC4*
160	chr12:54784900-54785238	*ZNF385A*
161	chr12:56122017-56123186	*CD63*
162	chr12:98897604-98897849	*Non Coding Region*
163	chr14:105940325-105941900	*CRIP2*
164	chr14:54420184-54422958	*BMP4*
165	chr14:60794576-60794867	*Non Coding Region*
166	chr14:61188116-61190772	*SIX4*
167	chr14:62217725-62218077	*Non Coding Region*
168	chr14:65016516-65016909	*C14orf50*
169	chr14:65170876-65172007	*PLEKHG3*
170	chr15:31683984-31685824	*Non Coding Region*
171	chr15:41803443-41805531	*LTK*
172	chr15:45003460-45004345	*B2M*
173	chr15:78556405-78557542	*DNAJA4*
174	chr15:90208787-90209279	*PLIN1*
175	chr16:29624387-29625391	*SLC7A5P1*
176	chr16:431189-432810	*LOC100134368*
177	chr16:89641053-89643415	*CPNE7*
178	chr17:1390456-1390786	*MYO1C*
179	chr17:29298046-29298606	*RNF135*
180	chr17:34091136-34091919	*C17orf50*
181	chr17:36575499-36575782	*Non Coding Region*

Figure 71B

Order	Coordinate of CpG (hg19)	*Gene*
182	chr17:41984148-41985012	*MPP2*
183	chr17:42082027-42084972	*NAGS*
184	chr17:43324689-43325729	*LOC100133991*
185	chr17:46654053-46654369	*HOXB4*
186	chr17:46655215-46655604	*HOXB4*
187	chr17:61523000-61524470	*CYB561*
188	chr17:62774366-62777797	*LOC146880*
189	chr17:73083866-73084495	*SLC16A5*
190	chr17:73892496-73893448	*TRIM65*
191	chr17:79478235-79482426	*Non Coding Region*
192	chr17:80291020-80292394	*SECTM1*
193	chr18:55862653-55862873	*NEDD4L*
194	chr19:12978359-12978785	*MAST1*
195	chr19:13120986-13124368	*NFIX*
196	chr19:18209911-18210268	*MAST3*
197	chr19:18499097-18499697	*GDF15*
198	chr19:19221196-19221575	*SLC25A42*
199	chr19:3434929-3435417	*NFIC*
200	chr19:35645597-35646047	*FXYD5*
201	chr19:36389545-36389967	*NFKBID*
202	chr19:39440404-39441011	*FBXO17*
203	chr19:39894558-39894768	*Non Coding Region*
204	chr19:43979359-43979684	*PHLDB3*
205	chr19:48229072-48229507	*EHD2*
206	chr19:48824529-48825493	*EMP3*
207	chr19:49255778-49256495	*FUT1*
208	chr19:50096609-50096912	*PRR12*
209	chr19:54371854-54372990	*MYADM*
210	chr19:54485303-54486322	*CACNG8*
211	chr19:55865016-55865434	*FAM71E2,COX6B2*
212	chr19:6740669-6741203	*TRIP10*
213	chr19:9896563-9896994	*Non Coding Region*
214	chr20:33762403-33762774	*PROCR*
215	chr20:42135793-42137062	*Non Coding Region*
216	chr20:62283516-62284835	*STMN3*
217	chr20:62710769-62711745	*OPRL1,RGS19*
218	chr21:26934423-26934805	*MIR155HG*
219	chr22:40390484-40391490	*FAM83F*
220	chr22:42679333-42679893	*Non Coding Region*

Figure 71C

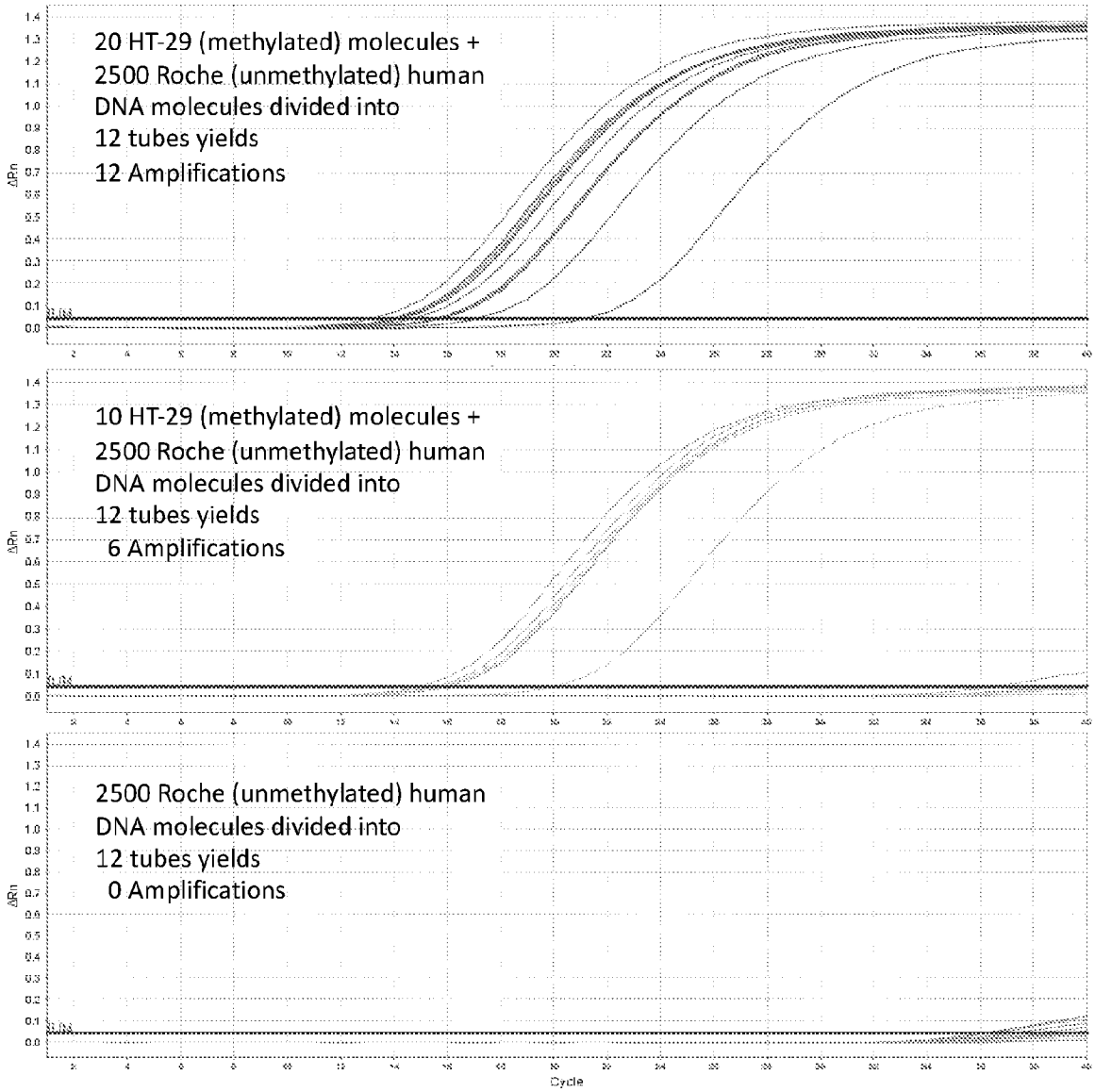

20 HT-29 (methylated) molecules +
2500 Roche (unmethylated) human
DNA molecules divided into
12 tubes yields
12 Amplifications
10 HT-29 (methylated) molecules +
2500 Roche (unmethylated) human
DNA molecules divided into
12 tubes yields
6 Amplifications
2500 Roche (unmethylated) human
DNA molecules divided into
12 tubes yields
0 Amplifications
ΔRn
Cycle

Figure 72

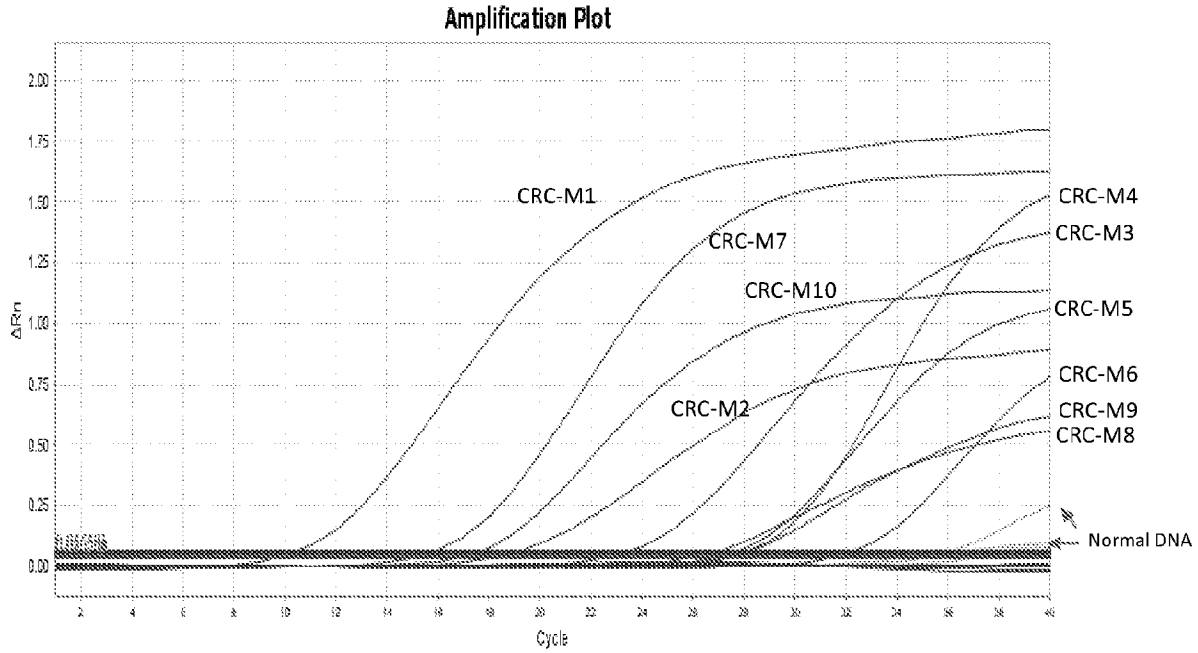

Amplification Plot
ΔRn
Cycle
CRC-M1
CRC-M7
CRC-M10
CRC-M2
CRC-M4
CRC-M3
CRC-M5
CRC-M6
CRC-M9
CRC-M8
Normal DNA

Figure 73

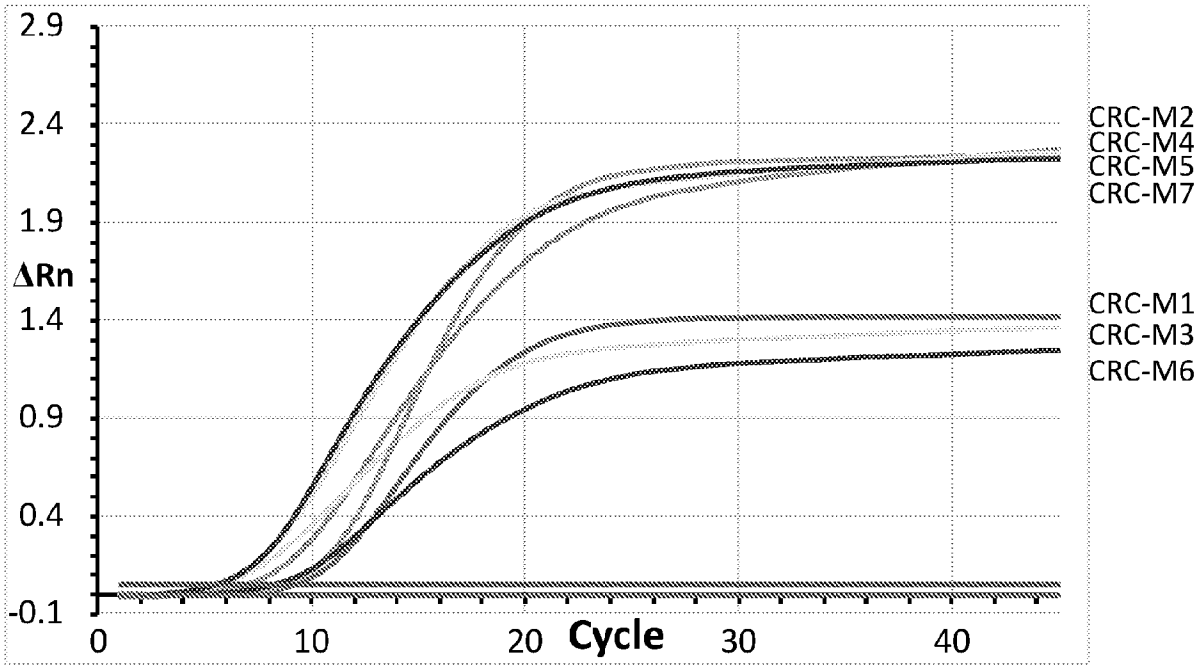
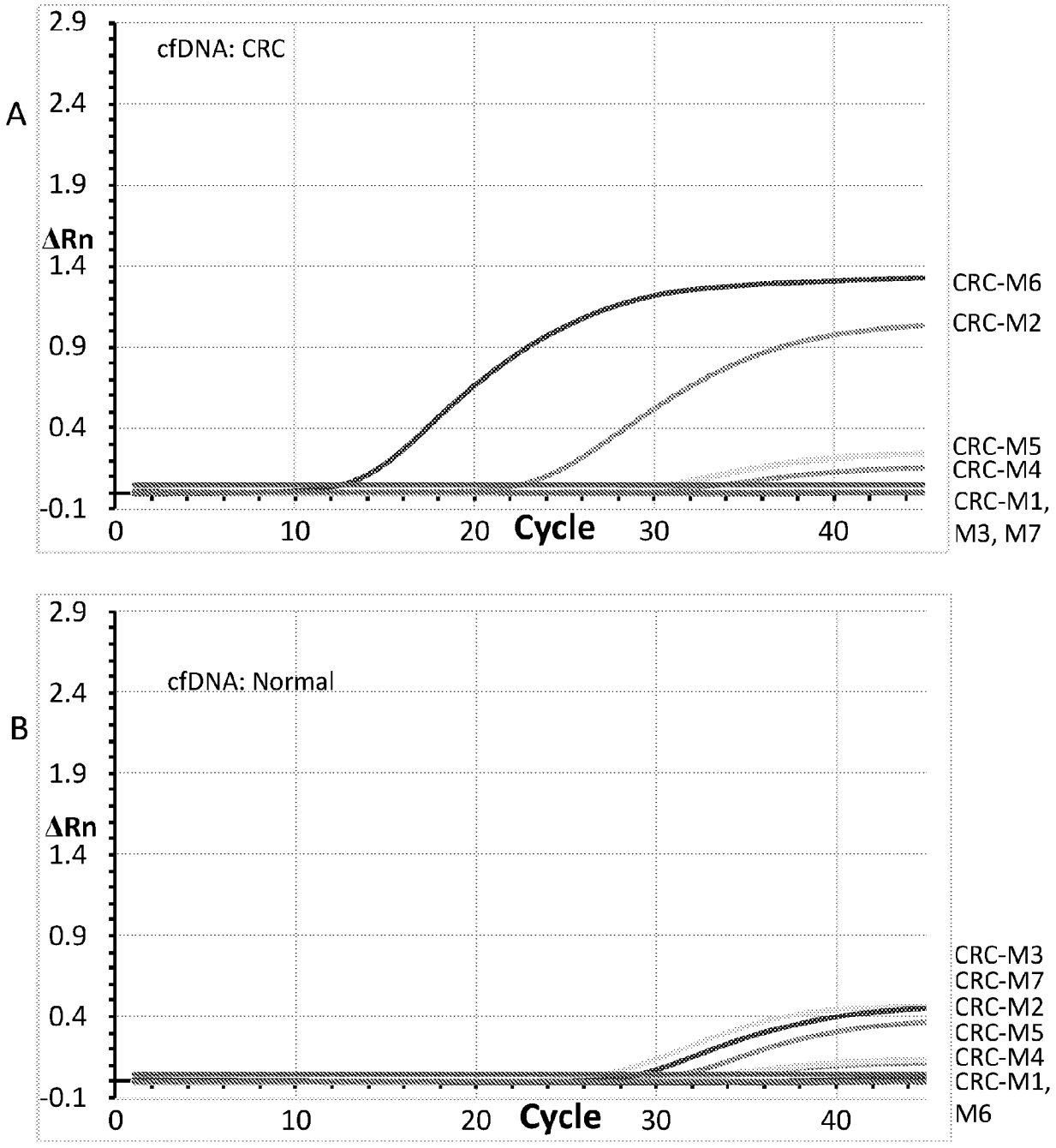
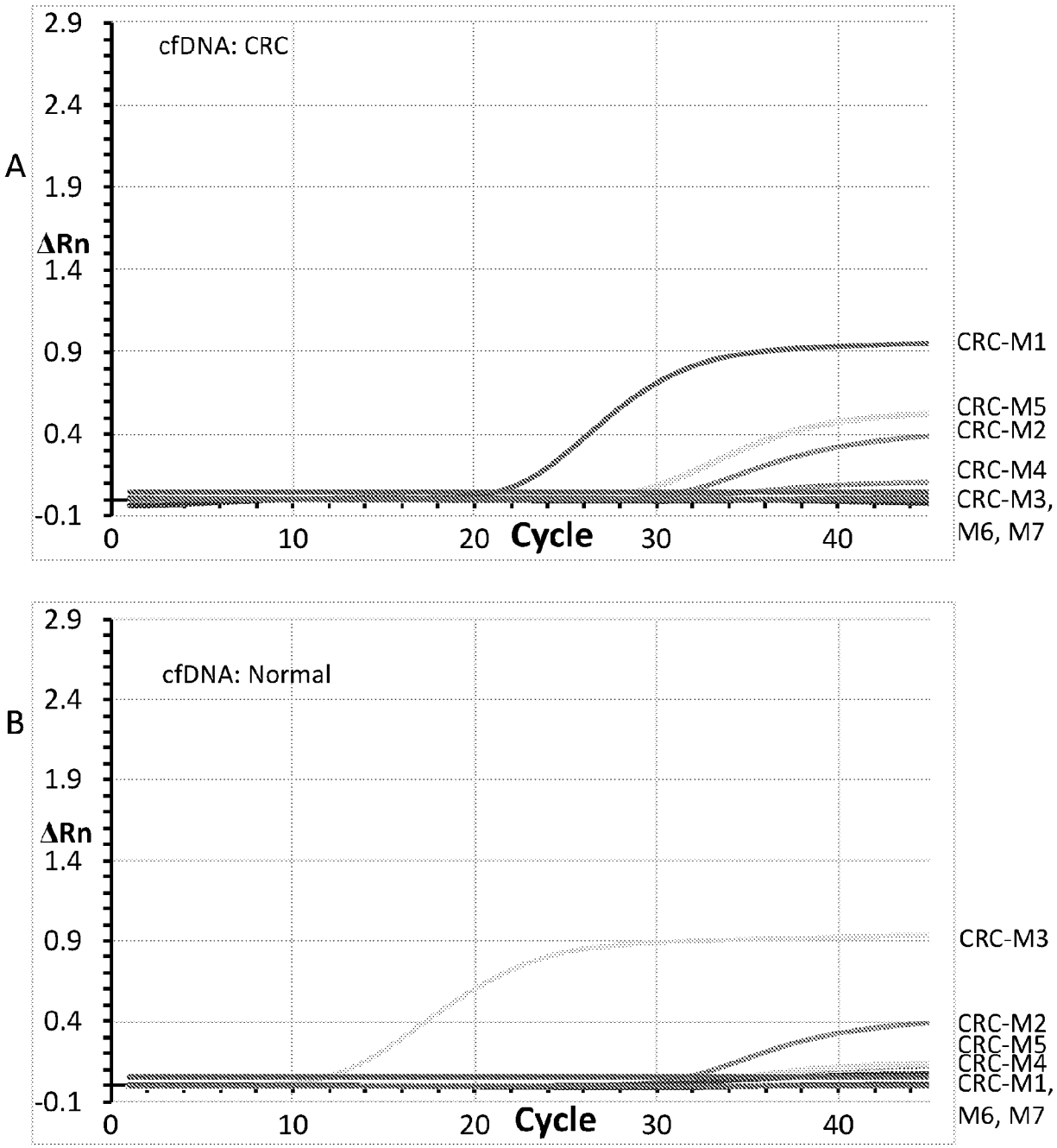

2.9
2.4
1.9
ΔRn
1.4
0.9
0.4
-0.1
0
10
20
Cycle
30
40
CRC-M2
CRC-M4
CRC-M5
CRC-M7
CRC-M1
CRC-M3
CRC-M6

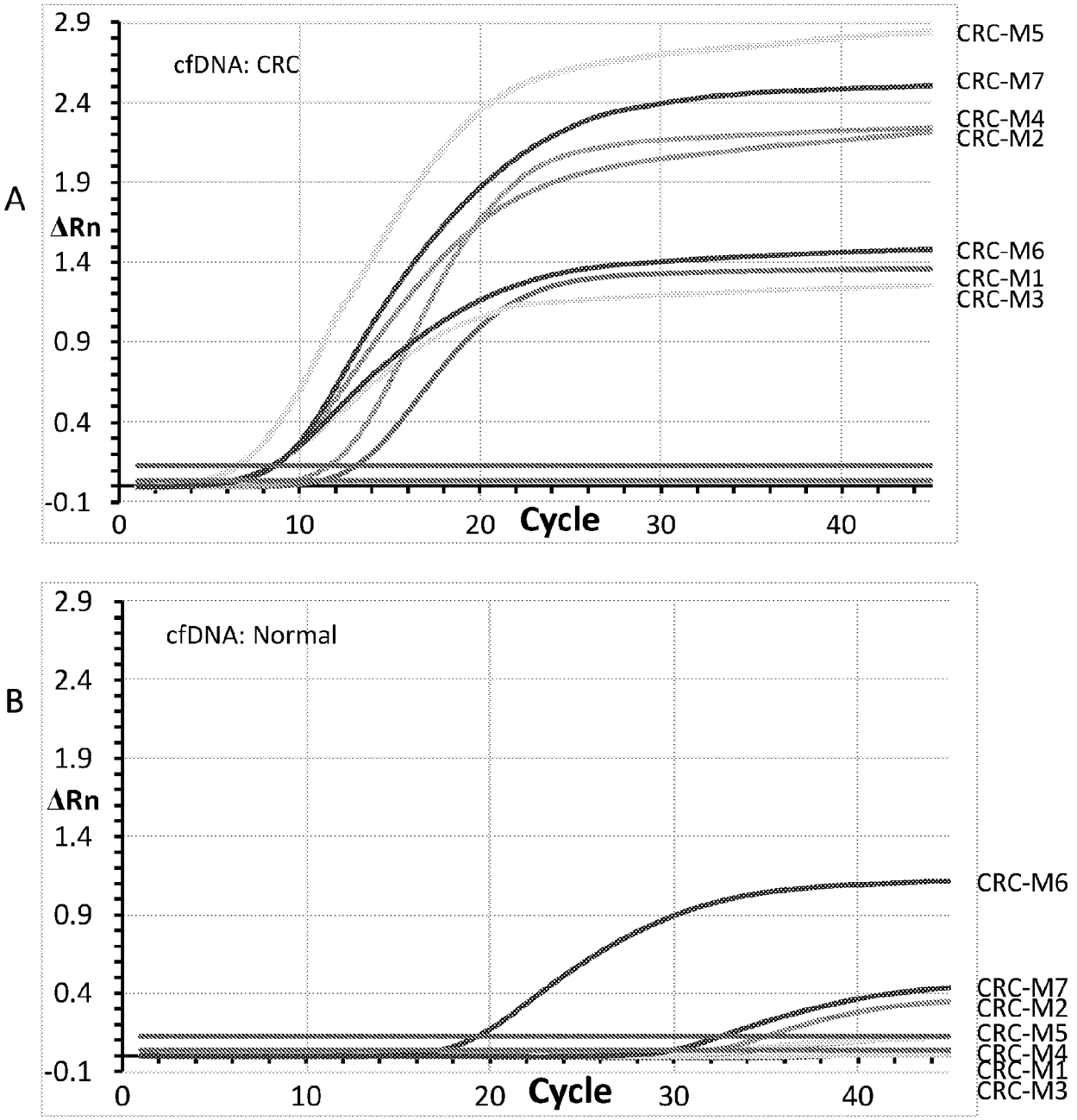
Figure 74



Figures 75A-B

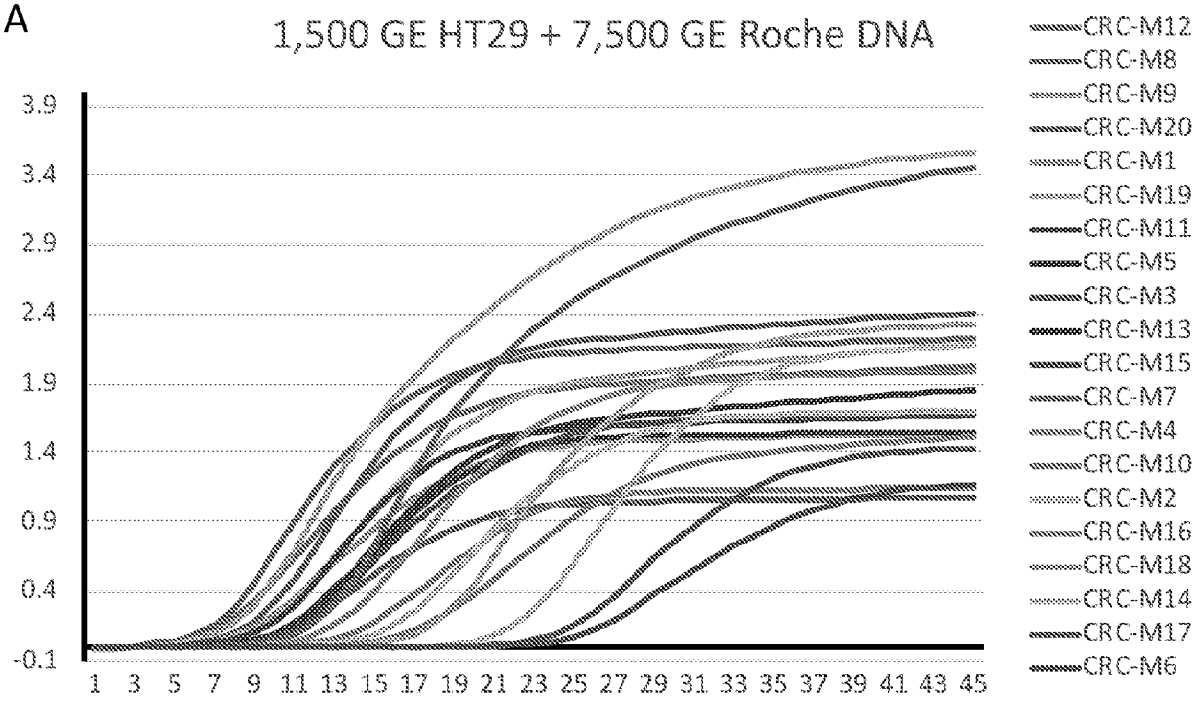


Figures 76A-B

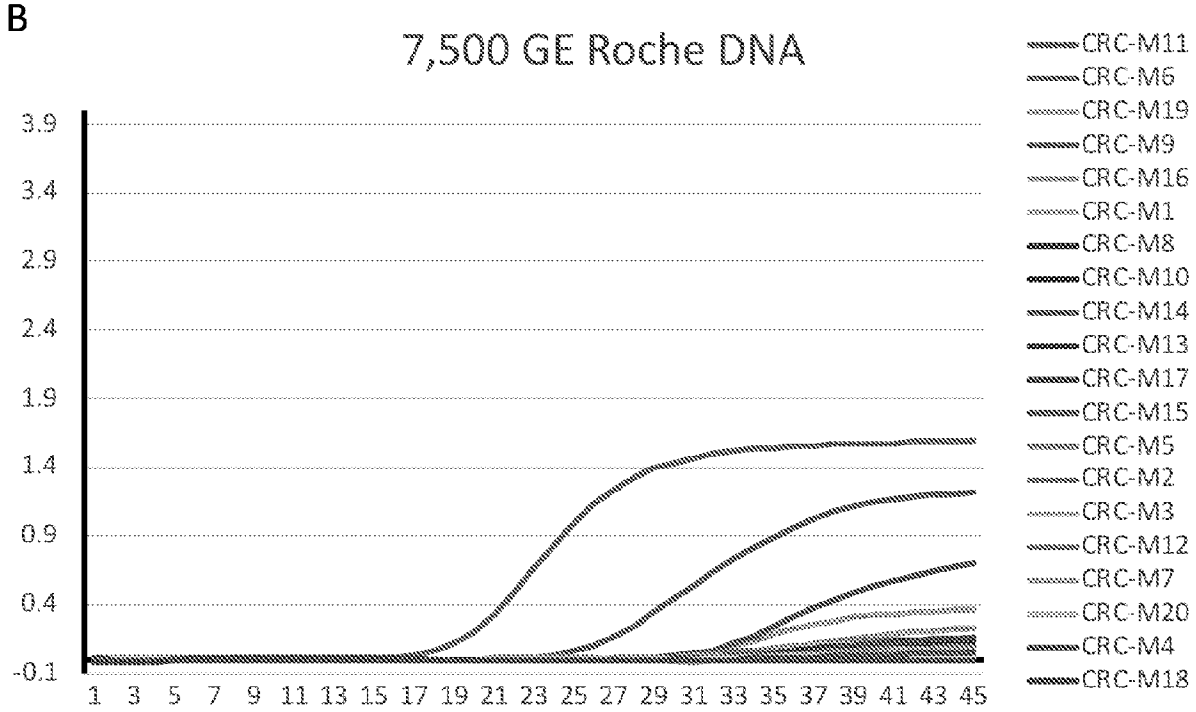


Figures 77A-B

A



B



Figures 78A-B

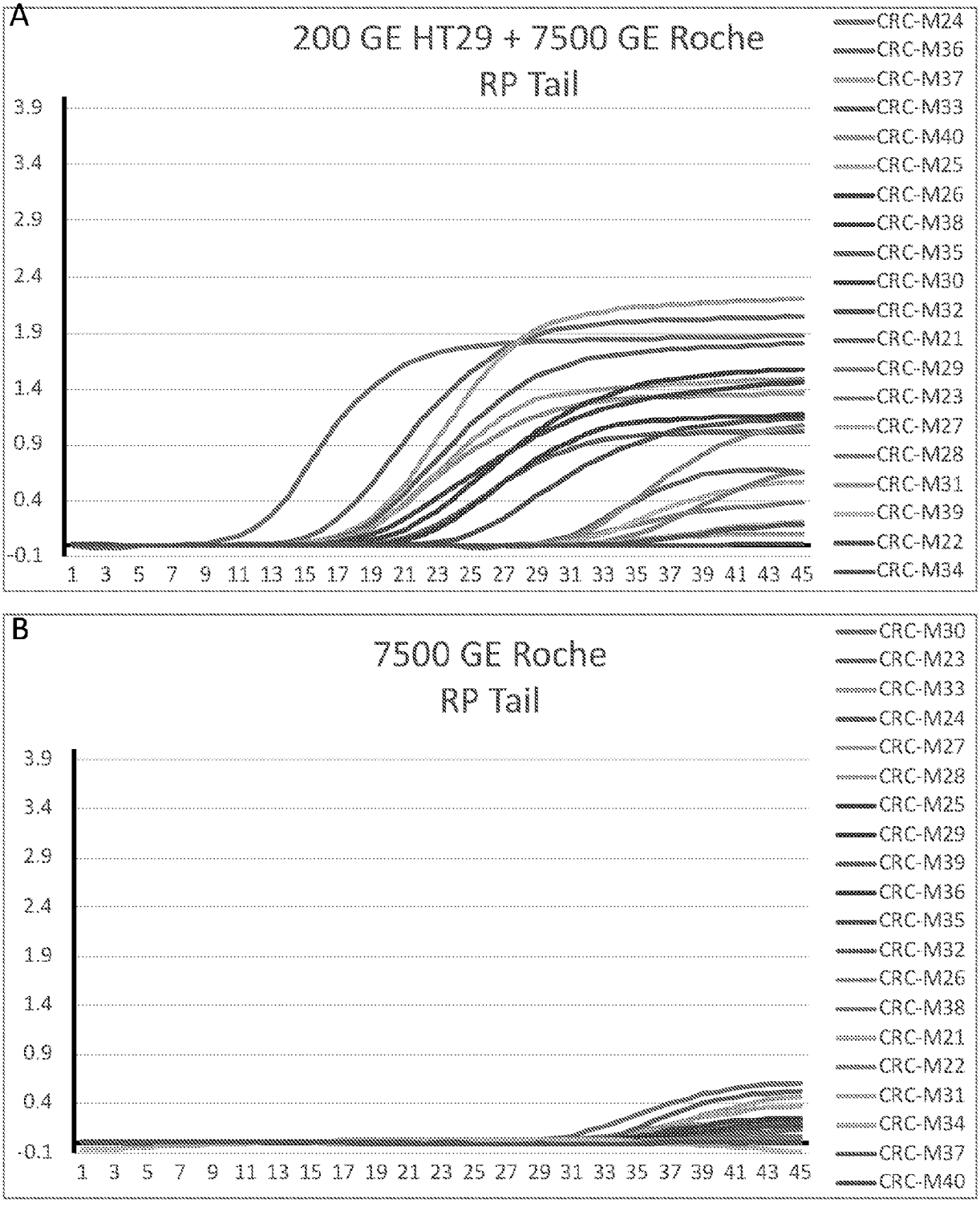
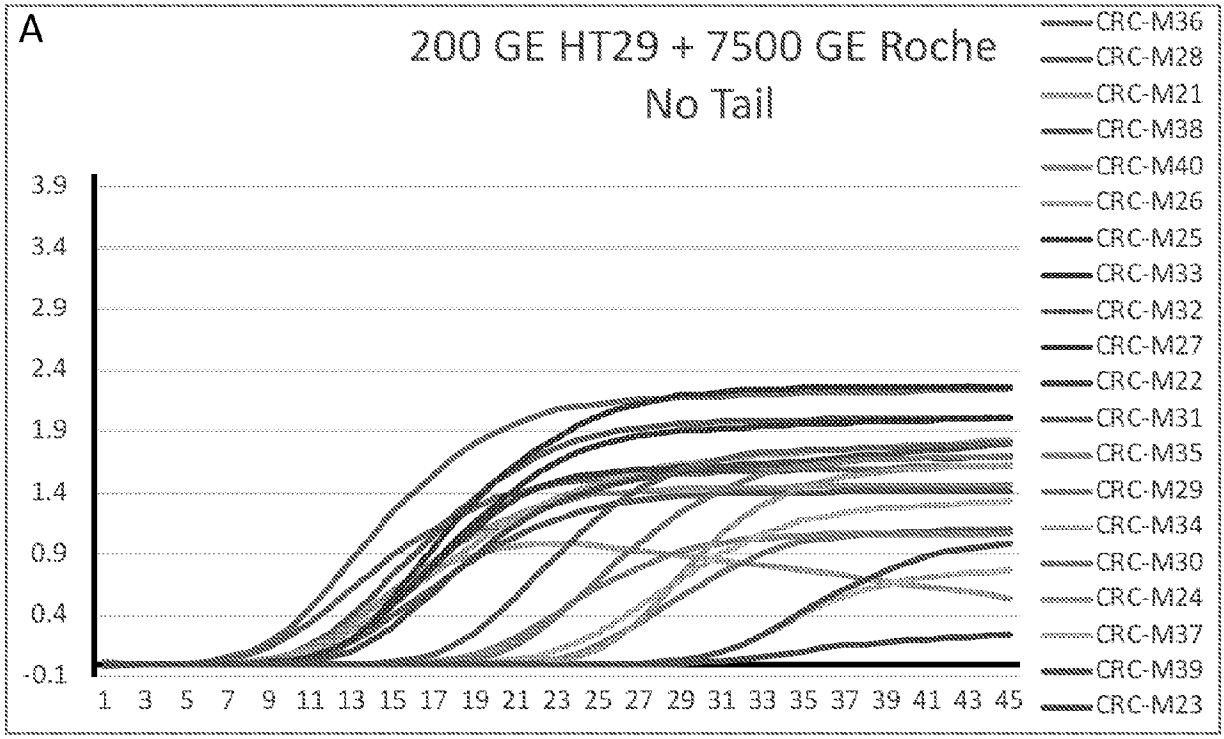

A
200 GE HT29 + 7500 GE Roche
RP Tail
3.9
3.4
2.9
2.4
1.9
1.4
0.9
0.4
-0.1
1 3 5 7 9 11 13 15 17 19 21 23 25 27 29 31 33 35 37 39 41 43 45
CRC-M24
CRC-M36
CRC-M37
CRC-M33
CRC-M40
CRC-M25
CRC-M26
CRC-M38
CRC-M35
CRC-M30
CRC-M32
CRC-M21
CRC-M29
CRC-M23
CRC-M27
CRC-M28
CRC-M31
CRC-M39
CRC-M22
CRC-M34
B
7500 GE Roche
RP Tail
3.9
3.4
2.9
2.4
1.9
1.4
0.9
0.4
-0.1
1 3 5 7 9 11 13 15 17 19 21 23 25 27 29 31 33 35 37 39 41 43 45
CRC-M30
CRC-M23
CRC-M33
CRC-M24
CRC-M27
CRC-M28
CRC-M25
CRC-M29
CRC-M39
CRC-M36
CRC-M35
CRC-M32
CRC-M26
CRC-M38
CRC-M21
CRC-M22
CRC-M31
CRC-M34
CRC-M37
CRC-M40

Figure 79A-B


A
200 GE HT29 + 7500 GE Roche
No Tail
3.9
3.4
2.9
2.4
1.9
1.4
0.9
0.4
-0.1
1 3 5 7 9 11 13 15 17 19 21 23 25 27 29 31 33 35 37 39 41 43 45
CRC-M36
CRC-M28
CRC-M21
CRC-M38
CRC-M40
CRC-M26
CRC-M25
CRC-M33
CRC-M32
CRC-M27
CRC-M22
CRC-M31
CRC-M35
CRC-M29
CRC-M34
CRC-M30
CRC-M24
CRC-M37
CRC-M39
CRC-M23

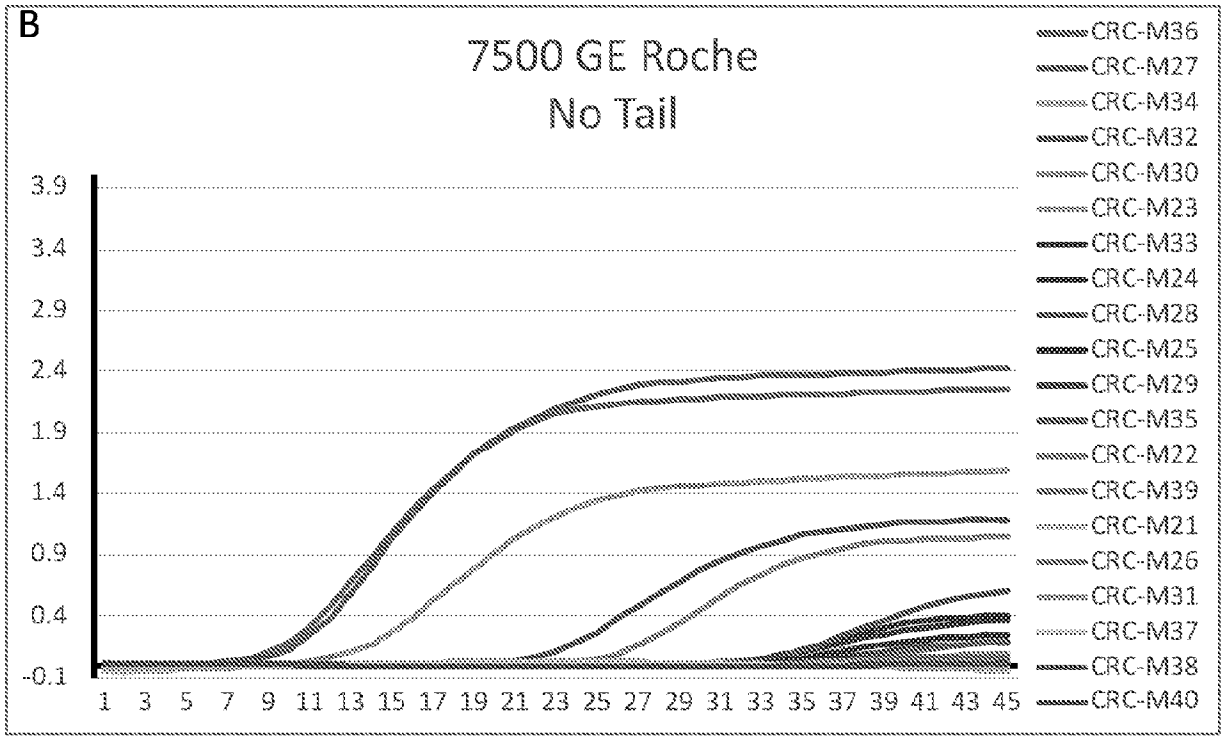

B
7500 GE Roche
No Tail
3.9
3.4
2.9
2.4
1.9
1.4
0.9
0.4
-0.1
1 3 5 7 9 11 13 15 17 19 21 23 25 27 29 31 33 35 37 39 41 43 45
CRC-M36
CRC-M27
CRC-M34
CRC-M32
CRC-M30
CRC-M23
CRC-M33
CRC-M24
CRC-M28
CRC-M25
CRC-M29
CRC-M35
CRC-M22
CRC-M39
CRC-M21
CRC-M26
CRC-M31
CRC-M37
CRC-M38
CRC-M40

Figure 80A-B