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DESCRIPTION

[0001] This application was filed electronically via the USPTO EFS-WEB server, as authorized and set forth in MPEP §1730 II.B.2.(a)(A), and this electronic filing includes an electronically submitted sequence (SEQ ID) listing; the entire content of this sequence listing is herein incorporated for all purposes. The sequence listing is identified on the electronically filed .txt file as follows:

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FIELD OF THE INVENTION

[0002] This invention relates generally to enzymes, polynucleotides encoding the enzymes, the use of such polynucleotides and polypeptides, the enzymes having xylanase activity, e.g., endoxylanase activity, and/or catalyzing hydrolysis of internal β -1,4-xylosidic linkages or endo- β -1,4-glucanase linkages; and/or degrading a linear polysaccharide beta-1,4-xylan into xylose. Thus, the invention provides methods and processes for breaking down hemicellulose, which is a major component of the cell wall of plants, including methods and processes for hydrolyzing hemicelluloses in any organic compound, plant or wood or wood product or byproduct, wood waste, paper pulp, paper product or paper waste or byproduct. The invention further provides methods and processes for breaking down plant matter containing cellulose and/or hemicellulose into simple sugars using the "cocktails" of the invention.

BACKGROUND

[0003] Xylanases (e.g., endo-1,4-beta-xylanase, EC 3.2.1.8) hydrolyze internal β -1,4-xylosidic linkages in xylan to produce smaller molecular weight xylose and xylo-oligomers. Xylans are polysaccharides formed from 1,4- β -glycoside-linked D-xylopyranoses. Xylanases are of considerable commercial value, being used in the food industry, for baking and fruit and vegetable processing, breakdown of agricultural waste, in the manufacture of animal feed and in pulp and paper production. Xylanases are formed by fungi and bacteria.

[0004] Arabinoxylans are major non-starch polysaccharides of cereals representing 2.5 - 7.1% w/w depending on variety and growth conditions. The physicochemical properties of this polysaccharide are such that it gives rise to viscous solutions or even gels under oxidative conditions. In addition, arabinoxylans have high water-binding capacity and may have a role in protein foam stability. All of these characteristics present problems for several industries including brewing, baking, animal nutrition and paper manufacturing. In brewing applications, the presence of xylan results in wort filterability and haze formation issues. In baking applications (especially for cookies and crackers), these arabinoxylans create sticky doughs that are difficult to machine and reduce biscuit size. In addition, this carbohydrate is implicated in rapid rehydration of the baked product resulting in loss of crispiness and reduced shelf-life. For monogastric animal feed applications with cereal diets, arabinoxylan is a major contributing factor to viscosity of gut contents and thereby adversely affects the digestibility of the feed and animal growth rate. For ruminant animals, these polysaccharides represent substantial components of fiber intake and more complete digestion of arabinoxylans would facilitate higher feed conversion efficiencies.

[0005] There remains a need in the art for xylanases to be used in the paper and pulp industry, for example, where the enzyme is active in the temperature range of 65°C to 75°C and at a pH of approximately 10. Additionally, an enzyme useful in the paper and pulp industry would decrease the need for bleaching chemicals, such as chlorine dioxide.

[0006] Additionally, there remains a need to provide efficient, low cost processes and compositions for producing bioalcohols, biofuels and/or biofuel- (e.g., bioethanol-, propanol-, butanol- and/or methanol-) by conversion of biomass. An enzyme or enzyme "cocktail" could provide a route to convert biomass into sugars that could then be fermented into biofuels.

SUMMARY OF THE INVENTION

[0007] The invention provides enzymes having: xylanase activity, e.g., endoxylanase activity, and/or catalyzing hydrolysis of internal β -1,4-xylosidic linkages or endo- β -1,4-glucanase linkages; and, nucleic acids encoding them, vectors and cells comprising them, probes for amplifying and identifying these xylanase-encoding nucleic acids, and methods for making and using these polypeptides and peptides.

[0008] For example, the invention provides enzymes having xylanase (e.g., endoxylanase activity), and compositions and methods comprising them, for hydrolyzing internal β -1,4-xylosidic linkages or endo- β -1,4-glucanase linkages, or hemicelluloses, in a wood, wood product, paper pulp, paper product or paper waste. In one aspect, the xylanase activity comprises catalyzing hydrolysis of xylan, e.g., degrading a linear polysaccharide beta-1,4-xylan into a xylose. Thus, the invention provides methods and processes for breaking down a xylan-comprising composition and/or a hemicellulose, which is a major component of the cell wall of plants.

[0009] In one aspect, the xylanase activity comprises hydrolyzing a glucan or other polysaccharide to produce a smaller molecular weight polysaccharide or oligomer. In one aspect, the glucan comprises a beta-glucan, such as a water soluble beta-glucan. The invention provides enzymes, compositions, methods and processes for hydrolyzing hemicelluloses in any organic matter, including cells, plants and/or wood or wood products, wood waste, paper pulp, paper products or paper waste or byproducts. The invention further provides methods and processes for breaking down plant matter containing cellulose and/or hemicellulose into simple sugars using the "cocktails" of the invention.

[0010] The invention provides enzymes for the bioconversion of any biomass, e.g., a lignocellulosic residue, into fermentable sugars or polysaccharides; and these sugars or polysaccharides can be used as a chemical feedstock for the production of alcohols such as ethanol, propanol, butanol and/or methanol, production of fuels, e.g., biofuels such as synthetic liquids or gases, such as syngas, and the production of other fermentation products, e.g. succinic acid, lactic acid, or acetic acid. Enzymes of the invention can be added to bioconversion and other industrial processes continuously, in batches or by fed-batch methods. In another aspect, enzymes of the invention can be recycled in bioconversion and other industrial processes, thereby lowering enzyme requirements.

[0011] In one aspect, the enzymes of the invention have an increased catalytic rate to improve the process of substrate (e.g., a lignocellulosic residue, cellulose, bagasse) hydrolysis. This increased efficiency in catalytic rate leads to an increased efficiency in producing sugars or polysaccharides, which can be useful in industrial, agricultural or medical applications, e.g., to make a biofuel or an alcohol such as ethanol, propanol, butanol and/or methanol. In one aspect, sugars produced by hydrolysis using enzymes of this invention can be used by microorganisms for alcohol (e.g., ethanol, propanol, butanol

and/or methanol) production and/or fuel (e.g., biofuel) production. Additionally, the sugars produced by hydrolysis using the enzymes of the invention can be used by microorganisms for the production of other fermentation products, e.g. succinic acid, lactic acid, or acetic acid.

[0012] The invention provides industrial, agricultural or medical applications: e.g., biomass to biofuel, e.g., ethanol, propanol, butanol and/or methanol, using enzymes of the invention having decreased enzyme costs, e.g., decreased costs in biomass to biofuel conversion processes. Thus, the invention provides efficient processes for producing bioalcohols, biofuels and/or biofuel- (e.g., bioethanol-, propanol-, butanol- and/or methanol-) comprising compositions, including synthetic, liquid or gas fuels comprising a bioalcohol, from any biomass.

[0013] In one aspect, enzymes of the invention, including the enzyme "cocktails" of the invention ("cocktails" meaning mixtures of enzymes comprising at least one enzyme of this invention), are used to hydrolyze the major components of a lignocellulosic biomass, or any composition comprising cellulose and/or hemicellulose (lignocellulosic biomass also comprises lignin), e.g., seeds, grains, tubers, plant waste (such as a hay or straw, e.g., a rice straw or a wheat straw, or any the dry stalk of any cereal plant) or byproducts of food processing or industrial processing (e.g., stalks), corn (including cobs, stover, and the like), grasses (e.g., Indian grass, such as *Sorghastrum nutans*; or, switch grass, e.g., *Panicum* species, such as *Panicum virgatum*), wood (including wood chips, processing waste, such as wood waste), paper, pulp, recycled paper (e.g., newspaper); also including a monocot or a dicot, or a monocot corn, sugarcane or parts thereof (e.g., cane tops), rice, wheat, barley, switchgrass or *Miscanthus*; or a dicot oilseed crop, soy, canola, rapeseed, flax, cotton, palm oil, sugar beet, peanut, tree, poplar or lupine; or, woods or wood processing byproducts, such as wood waste, e.g., in the wood processing, pulp and/or paper industry, in textile manufacture and in household and industrial cleaning agents, and/or in biomass waste processing.

[0014] In one aspect, enzymes of the invention are used to hydrolyze cellulose comprising a linear chain of β -1,4-linked glucose moieties, and/or hemicellulose as a complex structure that varies from plant to plant. In one aspect, enzymes of the invention are used to hydrolyze hemicelluloses containing a backbone of β -1,4 linked xylose molecules with intermittent branches of arabinose, galactose, glucuronic acid and/or mannose. In one aspect, enzymes of the invention are used to hydrolyze hemicellulose containing non-carbohydrate constituents such as acetyl groups on xylose and ferulic acid esters on arabinose. In one aspect, enzymes of the invention are used to hydrolyze hemicelluloses covalently linked to lignin and/or coupled to other hemicellulose strands via diferulate crosslinks.

[0015] In one aspect, the compositions and methods of the invention are used in the enzymatic digestion of biomass and can comprise use of many different enzymes, including the cellulases and hemicellulases. Lignocellulosic enzymes used to practice the invention can digest cellulose to monomeric sugars, including glucose. In one aspect, compositions used to practice the invention can include mixtures of enzymes, e.g., glycosyl hydrolases, glucose oxidases, xylanases, xylosidases (e.g., β -xylosidases), cellobiohydrolases, and/or arabinofuranosidases or other enzymes that can digest hemicellulose to monomer sugars. Mixtures of the invention can comprise, or consist of, only enzymes of this invention, or can include at least one enzyme of this invention and another enzyme, which can also be a lignocellulosic enzyme and/or any other enzyme.

[0016] In alternative embodiments, the invention provides polypeptides (and the nucleic acids that encode them) having at least one conservative amino acid substitution and retaining its xylanase activity; or, wherein the at least one conservative amino acid substitution comprises substituting an amino acid with another amino acid of like characteristics; or, a conservative substitution comprises: replacement of an aliphatic amino acid with another aliphatic amino acid; replacement of a Serine with a Threonine or vice versa; replacement of an acidic residue with another acidic residue; replacement of a residue bearing an

amide group with another residue bearing an amide group; exchange of a basic residue with another basic residue; or replacement of an aromatic residue with another aromatic residue;

[0017] In alternative embodiments, the invention provides polypeptides (and the nucleic acids that encode them) having a xylanase (e.g., an endoxylanase) activity but lacking a signal sequence, a prepro domain, a dockerin domain, and/or a carbohydrate binding module (CBM); and in one aspect, the carbohydrate binding module (CBM) comprises, or consists of, a xylan binding module, a cellulose binding module, a lignin binding module, a xylose binding module, a mannanase binding module, a xyloglucan-specific module and/or an arabinofuranosidase binding module.

[0018] In alternative embodiments, the invention provides polypeptides (and the nucleic acids that encode them) having a xylanase (e.g., an endoxylanase) activity further comprising a heterologous sequence; and in one aspect, the heterologous sequence comprises, or consists of a sequence encoding: (i) a heterologous signal sequence, a heterologous carbohydrate binding module, a heterologous dockerin domain, a heterologous catalytic domain (CD), or a combination thereof; (ii) the sequence of (i), wherein the heterologous signal sequence, carbohydrate binding module or catalytic domain (CD) is derived from a heterologous enzyme; or, (iii) a tag, an epitope, a targeting peptide, a cleavable sequence, a detectable moiety or an enzyme; and in one aspect, the heterologous carbohydrate binding module (CBM) comprises, or consists of, a xylan binding module, a cellulose binding module, a lignin binding module, a xylose binding module, a mannanase binding module, a xyloglucan-specific module and/or an arabinofuranosidase binding module; and in one aspect, the heterologous signal sequence targets the encoded protein to a vacuole, the endoplasmic reticulum, a chloroplast or a starch granule.

[0019] The invention provides isolated, synthetic or recombinant nucleic acids comprising a nucleic acid (polynucleotide) encoding at least one polypeptide having a xylanase activity, wherein the nucleic acid comprises a sequence having at least 95%, 96%, 97%, 98%, 99%, or more or complete (100%) sequence identity to a sequence which is (i) the nucleic acid (polynucleotide) sequence of SEQ ID NO:1 wherein the nucleic acid codon of SEQ ID NO:1 encoding residue 38 of SEQ ID NO:2 is changed from CGT to CAC and amino acid 38 R is changed to H and wherein the polypeptide has increased xylanase activity as measured by increased xylan hydrolysis as compared to SEQ ID NO:2, as set out in the accompanying claims. The nucleic acid may have at least one of the following further nucleotide residue changes: the codon encoding amino acid residue 4 changed from ACC to AAC; the codon encoding amino acid residue 4 changed from ACC to CGC; the codon encoding amino acid residue 4 changed from ACC to CAC; the codon encoding amino acid residue 9 changed from CCC to GAC; the codon encoding amino acid residue 17 changed from TTC to GTC; the codon encoding amino acid residue 21 changed from TTC to TAC; the codon encoding amino acid residue 33 changed from CTG to GCG; the codon encoding amino acid residue 44 changed from AGC to ACG; the codon encoding amino acid residue 63 changed from ATC to GTC; the codon encoding amino acid residue 73 changed from GGC to TAC; the codon encoding amino acid residue 73 changed from GGC to GAG; the codon encoding amino acid residue 73 changed from GGC to GTC; the codon encoding amino acid residue 108 changed from TTC to AAG; the codon encoding amino acid residue 125 changed from CAG to TAC; the codon encoding amino acid residue 150 changed from GTA to GCC; the codon encoding amino acid residue 188 changed from AGC to GAG; and/or, the codon encoding amino acid residue 189 changed from TCC to CAG.

[0020] The sequence identities may be determined: (A) by analysis with a sequence comparison algorithm or by a visual inspection, or (B) over a region of at least about 20, 30, 40, 50, 75, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1050, 1100, 1150 or more residues, or over the full length of a cDNA, transcript (mRNA) or gene.

[0021] The sequence comparison algorithm may be a BLAST version 2.2.2 algorithm where a filtering setting is set to blastall -p blastp -d "nr pataa" -F F, and all other options are set to default.

[0022] A nucleic acid (polynucleotide) encoding at least one polypeptide or peptide, may comprise a sequence that hybridizes under stringent conditions to a nucleic acid comprising the nucleic acid (polynucleotide) sequence of SEQ ID NO:1 having one or more nucleotide residue changes (or the equivalent thereof) as set forth above, wherein the polypeptide or peptide has a xylanase activity or is capable of generating a xylanase specific antibody (a polypeptide or peptide that acts as an epitope or immunogen),
and the stringent conditions comprise a wash step comprising a wash in 0.2X SSC at a temperature of about 65°C for about 15 minutes.

[0023] Fragments of any nucleic acid (polynucleotide) mentioned above may have a length of at least about 20, 25, 30, 50, 75, 100, 125, 150, 175, 200, 225, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1050, 1100, 1150 or more nucleotide residues, or the full length of a gene or a transcript.

[0024] The invention provides a nucleic acid (polynucleotide) encoding at least one polypeptide having a xylanase activity, wherein the polypeptide comprises the sequence of SEQ ID NO:2, and has the amino acid residue changes as set out herein.

[0025] These nucleic acids (polynucleotide) of the invention may encode a polypeptide having at least one conservative amino acid substitution and retaining its xylanase activity. The at least one conservative amino acid substitution may comprise substituting an amino acid with another amino acid of like characteristics; or, a conservative substitution comprises: replacement of an aliphatic amino acid with another aliphatic amino acid; replacement of a Serine with a Threonine or vice versa; replacement of an acidic residue with another acidic residue; replacement of a residue bearing an amide group with another residue bearing an amide group; exchange of a basic residue with another basic residue; or replacement of an aromatic residue with another aromatic residue.

[0026] The nucleic acid (polynucleotide) may encode a polypeptide having a xylanase activity but lacking a signal sequence, a prepro domain, a dockerin domain, and/or a carbohydrate binding module (CBM).

[0027] The carbohydrate binding module (CBM) may comprise, or consist of, a xylan binding module, a cellulose binding module, a lignin binding module, a xylose binding module, a mannanase binding module, a xyloglucan-specific module and/or a arabinofuranosidase binding module.

[0028] The nucleic acids (polynucleotide) of the invention may encode a polypeptide having a xylanase activity further comprising a heterologous sequence. The heterologous sequence may comprise, or consist of a sequence encoding: (A) a heterologous signal sequence, a heterologous carbohydrate binding module, a heterologous dockerin domain, a heterologous catalytic domain (CD), or a combination thereof; (B) the sequence of (1), wherein the heterologous signal sequence, carbohydrate binding module or catalytic domain (CD) is derived from a heterologous enzyme; or, (C) a tag, an epitope, a targeting peptide, a cleavable sequence, a detectable moiety or an enzyme.

[0029] The heterologous carbohydrate binding module (CBM) may comprise, or consist of, a xylan binding module, a cellulose binding module, a lignin binding module, a xylose binding module, a mannanase binding module, a xyloglucan-specific module and/or a arabinofuranosidase binding module.

[0030] The heterologous signal sequence may target the encoded protein to a vacuole, the endoplasmic

reticulum, a chloroplast or a starch granule.

[0031] The invention provides a nucleic acid sequence (polynucleotide) fully (completely) complementary to the sequences of the invention described herein.

[0032] The invention provides isolated, synthetic or recombinant nucleic acids comprising a nucleic acid encoding at least one polypeptide having a xylanase (e.g., an endoxylanase) activity, wherein the polypeptide has a sequence as set forth in SEQ ID NO:2 having one or more changes as described herein and in Table 1.

[0033] The invention provides isolated, synthetic or recombinant nucleic acids comprising sequences completely complementary to all of these nucleic acid sequences of the invention (complementary (non-coding) and coding sequences also hereinafter collectively referred to as nucleic acid sequences of the invention).

[0034] In one aspect, the invention also provides enzyme-encoding nucleic acids with a common novelty in that they encode a novel subset of xylanases, or a clade, comprising the "X14 module" (J Bacteriol. 2002 August; 184(15): 4124-4133). In one aspect, the invention also provides enzyme-encoding nucleic acids with a common novelty in that they encode a novel subset of xylanases, or a clade, comprising the "X14 module". Thus, in one aspect, the invention provides a novel genus of xylanases comprising xylanase members of SEQ ID NO:2 having one or more mutations as described herein, e.g., in Table 1.

[0035] In one aspect (optionally), the xylanase activity of the sequences comprises catalyzing hydrolysis of internal β -1,4-xylosidic linkages; comprises an endo-1,4-beta-xylanase activity; comprises hydrolyzing a xylan or an arabinoxylan to produce a smaller molecular weight xylose and xylo-oligomer; comprises hydrolyzing a polysaccharide comprising a 1,4- β -glycoside-linked D-xylopyranose; comprises hydrolyzing a cellulose or a hemicellulose; comprises hydrolyzing a cellulose or a hemicellulose in a wood, wood product, paper pulp, paper product or paper waste; comprises catalyzing hydrolysis of a xylan or an arabinoxylan in a feed or a food product; or, comprises catalyzing hydrolysis of a xylan or an arabinoxylan in a microbial cell or a plant cell. In one aspect, the xylanase activity comprises hydrolyzing polysaccharides comprising 1,4- β -glycoside-linked D-xylopyranoses or hydrolyzing hemicelluloses, e.g., hydrolyzing hemicelluloses in a wood, wood product, paper pulp, paper product or paper waste. In one aspect, the arabinoxylan is a cereal arabinoxylan, such as a wheat arabinoxylan.

[0036] In one aspect, the xylanase activity comprises catalyzing hydrolysis of polysaccharides, e.g., mannans or xylans, in a feed or a food product, such as a cereal-based animal feed, a wort or a beer, a milk or a milk product, a fruit or a vegetable. In one aspect, the xylanase activity comprises catalyzing hydrolysis of polysaccharides, e.g., mannans or xylans, in a microbial cell or a plant cell.

[0037] In one aspect, the xylanase activity is thermostable, e.g., wherein the polypeptide retains a xylanase activity under conditions comprising a temperature range from about -100°C to about -80°C, about -80°C to about -40°C, about -40°C to about -20°C, about -20°C to about 0°C, about 0°C to about 5°C, about 5°C to about 15°C, about 15°C to about 25°C, about 25°C to about 37°C, about 37°C to about 45°C, about 45°C to about 55°C, about 55°C to about 70°C, about 70°C to about 75°C, about 75°C to about 85°C, about 85°C to about 90°C, about 90°C to about 95°C, about 95°C to about 100°C, about 100°C to about 105°C, about 105°C to about 110°C, about 110°C to about 120°C, or 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 104°C, 105°C, 106°C, 107°C, 108°C, 109°C, 110°C, 111°C, 112°C, 113°C, 114°C, 115°C or more. In some embodiments, the thermostable polypeptides according to the invention retains activity at a temperature in the ranges described above, at about pH 3.0, about pH

3.5, about pH 4.0, about pH 4.5, about pH 5.0, about pH 5.5, about pH 6.0, about pH 6.5, about pH 7.0, about pH 7.5, about pH 8.0, about pH 8.5, about pH 9.0, about pH 9.5, about pH 10.0, about pH 10.5, about pH 11.0, about pH 11.5, about pH 12.0 or more.

[0038] In one aspect, the xylanase activity is thermotolerant, e.g., wherein the polypeptide retains a xylanase activity after exposure to a temperature in the range from about -100°C to about -80°C, about -80°C to about -40°C, about -40°C to about -20°C, about -20°C to about 0°C, about 0°C to about 5°C, about 5°C to about 15°C, about 15°C to about 25°C, about 25°C to about 37°C, about 37°C to about 45°C, about 45°C to about 55°C, about 55°C to about 70°C, about 70°C to about 75°C, about 75°C to about 85°C, about 85°C to about 90°C, about 90°C to about 95°C, about 95°C to about 100°C, about 100°C to about 105°C, about 105°C to about 110°C, about 110°C to about 120°C, or 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 104°C, 105°C, 106°C, 107°C, 108°C, 109°C, 110°C, 111°C, 112°C, 113°C, 114°C, 115°C or more. The thermotolerant polypeptides according to the invention can retain activity after exposure to a temperature in the range from about -100°C to about -80°C, about -80°C to about -40°C, about -40°C to about -20°C, about -20°C to about 0°C, about 0°C to about 5°C, about 5°C to about 15°C, about 15°C to about 25°C, about 25°C to about 37°C, about 37°C to about 45°C, about 45°C to about 55°C, about 55°C to about 70°C, about 70°C to about 75°C, about 75°C to about 85°C, about 85°C to about 90°C, about 90°C to about 95°C, about 95°C to about 100°C, about 100°C to about 105°C, about 105°C to about 110°C, about 110°C to about 120°C, or 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 104°C, 105°C, 106°C, 107°C, 108°C, 109°C, 110°C, 111°C, 112°C, 113°C, 114°C, 115°C or more. In some embodiments, the thermotolerant polypeptides according to the invention retains activity after exposure to a temperature in the ranges described above, at about pH 3.0, about pH 3.5, about pH 4.0, about pH 4.5, about pH 5.0, about pH 5.5, about pH 6.0, about pH 6.5, about pH 7.0, about pH 7.5, about pH 8.0, about pH 8.5, about pH 9.0, about pH 9.5, about pH 10.0, about pH 10.5, about pH 11.0, about pH 11.5, about pH 12.0 or more.

[0039] In one aspect, the polypeptides encoded by nucleic acids of the invention retain activity under acidic conditions comprising about pH 6.5, pH 6, pH 5.5, pH 5, pH 4.5, pH 4.0, pH 3.5, pH 3.0 or less (more acidic) pH, or, retain activity after exposure to acidic conditions comprising about pH 6.5, pH 6, pH 5.5, pH 5, pH 4.5, pH 4.0, pH 3.5, pH 3.0 or less (more acidic) pH; or, retain activity under basic conditions comprising about pH 7, pH 7.5 pH 8.0, pH 8.5, pH 9, pH 9.5, pH 10, pH 10.5, pH 11, pH 11.5, pH 12, pH 12.5 or more (more basic) or, retain activity after exposure to basic conditions comprising about pH 7, pH 7.5 pH 8.0, pH 8.5, pH 9, pH 9.5, pH 10, pH 10.5, pH 11, pH 11.5, pH 12, pH 12.5 or more (more basic). In one aspect, xylanase activity of polypeptides encoded by nucleic acids of the invention retain activity at a temperature of at least about 80°C, 81°C, 82°C, 83°C, 84°C, 85°C, 86°C, 87°C, 88°C, 89°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 7.5 pH 8.0, pH 8.5, pH 9, pH 9.5, pH 10, pH 10.5, pH 11, pH 11.5, pH 12, pH 12.5 or more (more basic).

[0040] The invention provides expression cassettes, cloning vehicles, or a vector (e.g., expression vectors) comprising a nucleic acid comprising a sequence of the invention. The cloning vehicle can comprise a viral vector, a plasmid, a phage, a phagemid, a cosmid, a fosmid, a bacteriophage or an artificial chromosome. The viral vector can comprise an adenovirus vector, a retroviral vector or an adeno-associated viral vector. The cloning vehicle can comprise an artificial chromosome comprising a bacterial artificial chromosome (BAC), a bacteriophage P1-derived vector (PAC), a yeast artificial chromosome (YAC), or a mammalian artificial chromosome (MAC).

[0041] Probes for identifying a nucleic acid encoding a polypeptide with a xylanase activity may comprise at least about 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 100, 125, 150, 175, 200, 225, 250, 275, 300 or more consecutive bases of a nucleic acid comprising an exemplary sequence of the invention,

or, any sequence of the invention (as defined herein), wherein in one aspect (optionally) the probe comprises an oligonucleotide comprising between at least about 10 to 300, about 25 to 250, about 10 to 50, about 20 to 60, about 30 to 70, about 40 to 80, about 60 to 100, or about 50 to 150 or more consecutive bases.

[0042] An amplification primer pair for amplifying a nucleic acid encoding a polypeptide having a xylanase activity, may be capable of amplifying a nucleic acid comprising an exemplary sequence of the invention, or, any sequence of the invention (as defined herein), or a subsequence thereof, wherein optionally a member of the amplification primer sequence pair comprises an oligonucleotide comprising at least about 10 to 50 consecutive bases of the sequence, or, about 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35 or more consecutive bases of the sequence. The primer pair may comprise a first member having a sequence as set forth by about the first (the 5') 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35 or more residues of an exemplary sequence of the invention, or, any sequence of the invention (as defined herein), and a second member having a sequence as set forth by about the first (the 5') 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35 or more residues of the complementary strand of the first member.

[0043] Xylanase- encoding nucleic acids may be generated by amplification of a polynucleotide using such an amplification primer pair. Optionally the amplification is by polymerase chain reaction (PCR). The nucleic acid may be generated by amplification of a gene library, which may be an environmental library. Isolated, synthetic or recombinant xylanases may be encoded by a xylanase- encoding nucleic acid generated by amplification of a polynucleotide using such an amplification primer pair. Methods of amplifying a nucleic acid encoding a polypeptide having a xylanase activity comprises the step of amplification of a template nucleic acid with an amplification primer sequence pair capable of amplifying an exemplary sequence of the invention, or, any sequence of the invention (as defined herein), or a subsequence thereof.

[0044] The invention provides expression cassette, a vector or a cloning vehicle comprising a nucleic acid comprising a sequence of the invention, wherein optionally the cloning vehicle comprises a viral vector, a plasmid, a phage, a phagemid, a cosmid, a fosmid, a bacteriophage or an artificial chromosome. The viral vector can comprise an adenovirus vector, a retroviral vector or an adeno-associated viral vector, or, the artificial chromosome comprises a bacterial artificial chromosome (BAC), a bacteriophage P1-derived vector (PAC), a yeast artificial chromosome (YAC), or a mammalian artificial chromosome (MAC).

[0045] The invention provides transformed cells comprising a nucleic acid or vector of the invention, or an expression cassette or cloning vehicle of the invention. The transformed cell can be a bacterial cell, a mammalian cell, a fungal cell, a yeast cell, an insect cell or a plant cell.

[0046] The invention provides transgenic non-human animals comprising a sequence of the invention. The transgenic non-human animal can be a mouse, a rat, a rabbit, a sheep, a pig, a chicken, a goat, a fish, a dog, or a cow. The invention provides transgenic plants comprising a sequence of the invention, e.g., wherein the plant is a corn plant, a sorghum plant, a potato plant, a tomato plant, a wheat plant, an oilseed plant, a rapeseed plant, a soybean plant, a rice plant, a barley plant, a grass, or a tobacco plant. The invention provides transgenic seeds comprising a sequence of the invention, e.g., wherein the seed is a corn seed, a wheat kernel, an oilseed, a rapeseed, a soybean seed, a palm kernel, a sunflower seed, a sesame seed, a rice, a barley, a peanut or a tobacco plant seed.

[0047] Antisense oligonucleotides may comprise a nucleic acid sequence complementary to or capable of

hybridizing under stringent conditions to a sequence of the invention (including, e.g., exemplary sequences of the invention), or a subsequence thereof, wherein optionally the antisense oligonucleotide is between about 10 to 50, about 20 to 60, about 30 to 70, about 40 to 80, or about 60 to 100 bases in length, and in one aspect (optionally) the stringent conditions comprise a wash step comprising a wash in 0.2X SSC at a temperature of about 65°C for about 15 minutes.

[0048] Methods of inhibiting the translation of a xylanase message in a cell comprise administering to the cell or expressing in the cell an antisense oligonucleotide comprising a nucleic acid sequence complementary to or capable of hybridizing under stringent conditions to a sequence of the invention (including, e.g., exemplary sequences of the invention).

[0049] Double-stranded inhibitory RNA (RNAi) molecules may comprise a subsequence of a sequence of the invention (including, e.g., exemplary sequences of the invention). The double-stranded inhibitory RNA (RNAi) molecule can be about 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, or 30 or more duplex nucleotides in length. Methods of inhibiting the expression of a xylanase in a cell comprise administering to the cell or expressing in the cell a double-stranded inhibitory RNA (iRNA), wherein the RNA comprises a subsequence of a sequence of the invention (including, e.g., exemplary sequences of the invention).

[0050] The invention provides isolated, synthetic or recombinant polypeptides having a xylanase activity, or polypeptides capable of generating an immune response specific for a xylanase (e.g., an endoxylanase); and in alternative aspects peptide and polypeptide of the invention comprise a sequence comprising an amino acid sequence having at least 95%, 96%, 97%, 98%, 99%, or more, or has 100% (complete) sequence identity to a sequence which is:

1. (i) the amino acid sequence of SEQ ID NO:2, and having a change at amino acid residue 38 from R to H and wherein the polypeptide has increased xylanase activity as measured by increased xylan hydrolysis as compared to SEQ ID NO:2, as set out in the accompanying claims. The polypeptide may have at least one of the following further amino acid residue changes (or the equivalent thereof): amino acid residue 4 is changed from a T (or thr, or threonine) to an N (or asn, or asparagine); amino acid residue 4 is changed from a T (or thr, or threonine) to an R (or arg, or arginine); amino acid residue 4 is changed from a T (or thr, or threonine) to an H (or his, or histidine); amino acid residue 9 is changed from a P (or pro, or proline) to an D (or asp, or aspartic acid); amino acid residue 17 is changed from a F (or phe, or phenylalanine) to an V (or val, or valine); amino acid residue 21 is changed from a F (or phe, or phenylalanine) to an Y (or tyr, or tyrosine); amino acid residue 33 is changed from a L (or leu, or leucine) to an A (or ala, or alanine); amino acid residue 44 is changed from a S (or ser, or serine) to a T (or thr, or threonine); amino acid residue 63 is changed from a I (or ile, or isoleucine) to an V (or val, or valine); amino acid residue 73 is change from a G (or gly, or glycine) to an Y (or tyr, or tyrosine); amino acid residue 73 is changed from a G (or gly, or glycine) to an V (or val, or valine); amino acid residue 73 is changed from a G (or gly, or glycine) to an E (or glu, or glutamic acid); amino acid residue 108 is changed from a F (or phe, or phenylalaine) to an K (or lys, or lysine); amino acid residue 125 is change from a Q (or gln, or glutamine) to an Y (or tyr, or tyrosine); amino acid residue 150 is change from a V (or val, or valine) to an A (or ala, or alanine); amino acid residue 188 is changed from a S (or ser, or serine) to an E (or glu, or glutamic acid); and/or amino acid residue 189 is changed from a S (or ser, or serine) to an Q (or gln, or glutamine), or
2. (ii) the amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22 or SEQ ID NO:24;

wherein the polypeptide or peptide of (i) or (ii) has a xylanase activity, or the polypeptide or peptide is capable of generating a xylanase specific antibody (a polypeptide or peptide that acts as an epitope or immunogen).

[0051] The sequence identities may be determined: (A) by analysis with a sequence comparison algorithm or by a visual inspection, or (B) over a region of at least about 20, 25, 30, 35, 40, 45, 50, 55, 60, 75, 100, 150, 200, 250, 300 or more amino acid residues, or over the full length of the polypeptide or peptide or enzyme, and/or enzymatically active subsequences (fragments) thereof. The sequence identities may be determined by analysis with a sequence comparison algorithm or by a visual inspection. Optionally the sequence comparison algorithm is a BLAST version 2.2.2 algorithm where a filtering setting is set to blastall -p blastp -d "nr pataa" -F F, and all other options are set to default.

[0052] The invention provides an amino acid sequence encoded by the nucleic acid of claim 1, wherein the polypeptide has (i) a xylanase activity, or, (ii) has immunogenic activity in that it is capable of generating an antibody that specifically binds to a polypeptide having a sequence as set out above, and/or enzymatically active subsequences (fragments) thereof.

[0053] The amino acid sequence of the invention may comprise at least one amino acid residue conservative substitution, and the polypeptide or peptide retains xylanase activity.

[0054] The conservative substitution may comprise replacement of an aliphatic amino acid with another aliphatic amino acid; replacement of a serine with a threonine or vice versa; replacement of an acidic residue with another acidic residue; replacement of a residue bearing an amide group with another residue bearing an amide group; exchange of a basic residue with another basic residue; or, replacement of an aromatic residue with another aromatic residue, or a combination thereof.

[0055] The aliphatic residue may comprise Alanine, Valine, Leucine, Isoleucine or a synthetic equivalent thereof; the acidic residue comprises Aspartic acid, Glutamic acid or a synthetic equivalent thereof; the residue comprising an amide group comprises Aspartic acid, Glutamic acid or a synthetic equivalent thereof; the basic residue comprises Lysine, Arginine or a synthetic equivalent thereof; or, the aromatic residue comprises Phenylalanine, Tyrosine or a synthetic equivalent thereof.

[0056] The polypeptide of the invention may lack a signal sequence, a prepro domain, a dockerin domain, and/or a carbohydrate binding module (CBM). The carbohydrate binding module (CBM) may comprise or consist of, a xylan binding module, a cellulose binding module, a lignin binding module, a xylose binding module, a mannanase binding module, a xyloglucan-specific module and/or a arabinofuranosidase binding module.

[0057] The polypeptide of the invention may further comprise a heterologous sequence. The heterologous sequence may comprise, or consist of: (A) a heterologous signal sequence, a heterologous carbohydrate binding module, a heterologous dockerin domain, a heterologous catalytic domain (CD), or a combination thereof; (B) the sequence of (A), wherein the heterologous signal sequence, carbohydrate binding module or catalytic domain (CD) is derived from a heterologous lignocellulosic enzyme; and/or, (C) a tag, an epitope, a targeting peptide, a cleavable sequence, a detectable moiety or an enzyme.

[0058] The heterologous sequence or the heterologous carbohydrate binding module (CBM) may comprise, or consist of, a xylan binding module, a cellulose binding module, a lignin binding module, a xylose binding module, a mannan binding module, a xyloglucan-specific module and/or a arabinofuranosidase binding module.

[0059] The heterologous signal sequence may target the encoded protein to a vacuole, the endoplasmic reticulum, a chloroplast or a starch granule.

[0060] The isolated, synthetic or recombinant peptides of the invention have a xylanase activity, e.g., wherein the xylanase activity comprises catalyzing hydrolysis of internal β -1,4-xylosidic linkages; comprises an endo-1,4-beta-xylanase activity; comprises hydrolyzing a xylan or an arabinoxylan to produce a smaller molecular weight xylose and xylo-oligomer; comprises hydrolyzing a polysaccharide comprising a 1,4- β -glycoside-linked D-xylopyranose; comprises hydrolyzing a cellulose or a hemicellulose; comprises hydrolyzing a cellulose or a hemicellulose in a wood, wood product, paper pulp, paper product or paper waste; comprises catalyzing hydrolysis of a xylan or an arabinoxylan in a feed or a food product; or, comprises catalyzing hydrolysis of a xylan or an arabinoxylan in a microbial cell or a plant cell. The xylan can comprises an arabinoxylan, e.g., a water soluble arabinoxylan, e.g., a water soluble arabinoxylan in a dough or a bread product.

[0061] In one aspect, the xylanase activity comprises hydrolyzing polysaccharides, for example, comprising 1,4- β -glycoside-linked D-xylopyranoses, or hydrolyzing hemicelluloses, e.g., hydrolyzing hemicelluloses in a wood, wood product, paper pulp, paper product or paper waste.

[0062] In one aspect, the xylanase activity comprises catalyzing hydrolysis of polysaccharides, e.g., xylans, in a feed or a food product, such as a cereal-based animal feed, a wort or a beer, a milk or a milk product, a fruit or a vegetable. In one aspect, the xylanase activity comprises catalyzing hydrolysis of xylans in a microbial cell or a plant cell.

[0063] The invention provides isolated, synthetic or recombinant polypeptides comprising a polypeptide of the invention and lacking a signal sequence or a prepro sequence. The invention provides isolated, synthetic or recombinant polypeptides comprising a polypeptide of the invention and having a heterologous signal sequence or a heterologous prepro sequence.

[0064] In one aspect, a polypeptide of the invention has xylanase activity comprising a specific activity at about 37°C in the range from about 100 to about 1000 units per milligram of protein, from about 500 to about 750 units per milligram of protein, from about 500 to about 1200 units per milligram of protein, or from about 750 to about 1000 units per milligram of protein. In one aspect, units are defined as 0.1 to 20 units/g of pulp, where a unit equals μmol of xylose released per minute per mg of enzyme, using arabinoxylan as a substrate as described in the Nelson Somogyi assay, described in detail below. In alternative aspects, polypeptides of the invention have xylanase activity in the range of between about 0.05 to 20 units per gram of pulp, or 0.05, 0.10, 0.20, 0.30, 0.40, 0.50, 0.60, 0.70, 0.80, 0.90, 1.0, 1.5, 2.0, 2.5, 3.0, 3.5, 4.0, 4.5, 5.0, 5.5, 6.0, 6.5, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 or 20 or more units per gram of pulp (where a unit equals μmol of xylose released per minute per mg of enzyme, using arabinoxylan as a substrate as described in the Nelson Somogyi assay).

[0065] In one aspect, the thermotolerance comprises retention of at least half of the specific activity of the xylanase at 37°C after being heated to an elevated temperature, such as a temperature from about 0°C to about 20°C, about 20°C to about 37°C, about 37°C to about 50°C, about 50°C to about 70°C, about 70°C to about 75°C, about 75°C to about 80°C, about 80°C to about 85°C, about 85°C to about 90°C, about 90°C to about 95°C, about 95°C to about 100°C, about 100°C to about 110°C, or higher. The thermotolerance can comprise retention of specific activity at 37°C in the range from about 500 to about 1200 units per milligram of protein after being heated to an elevated temperature, such as a temperature from about 0°C to about 20°C, about 20°C to about 37°C, about 37°C to about 50°C, about 50°C to about 70°C, about 70°C to about 75°C, about 75°C to about 80°C, about 80°C to about 85°C, about 85°C to

about 90°C, about 90°C to about 95°C, about 95°C to about 100°C, about 100°C to about 110°C, or higher.

[0066] In one aspect, the polypeptides of the invention comprise at least one glycosylation site or further comprises a polysaccharide. The glycosylation can be an N-linked glycosylation, e.g., wherein the polypeptide is glycosylated after being expressed in a *P. pastoris* or a *S. pombe*.

[0067] In one aspect, the xylanase activity of polypeptides of the invention retain activity under acidic conditions comprising about pH 6.5, pH 6, pH 5.5, pH 5, pH 4.5 or pH 4 or less (more acidic), or, retain a xylanase activity after exposure to acidic conditions comprising about pH 6.5, pH 6, pH 5.5, pH 5, pH 4.5 or pH 4 or less (more acidic); or, retain activity under basic conditions comprising about pH 7, pH 7.5 pH 8.0, pH 8.5, pH 9, pH 9.5, pH 10, pH 10.5, pH 11, pH 11.5, pH 12, pH 12.5 or more (more basic) or, retain a xylanase activity after exposure to basic conditions comprising about pH 7, pH 7.5 pH 8.0, pH 8.5, pH 9, pH 9.5, pH 10, pH 10.5, pH 11, pH 11.5, pH 12, pH 12.5 or more (more basic). In one aspect, xylanase activity of polypeptides of the invention retain activity at a temperature of at least about 80°C, 81°C, 82°C, 83°C, 84°C, 85°C, 86°C, 87°C, 88°C, 89°C or 90°C, and a basic pH of at least about pH 7.5 pH 8.0, pH 8.5, pH 9, pH 9.5, pH 10, pH 10.5, pH 11, pH 11.5, pH 12, pH 12.5 or more (more basic).

[0068] The invention provides protein preparation comprising a polypeptide of the invention, wherein the protein preparation comprises a liquid, a slurry, a solid or a gel. Heterodimers may comprise a polypeptide of the invention and a second domain. The second domain can be a polypeptide and the heterodimer is a fusion protein. The second domain can be an epitope or a tag. Homodimers or heterodimers may comprise a polypeptide of the invention.

[0069] Immobilized polypeptides comprising a sequence of the invention, or a subsequence thereof, or a polypeptide encoded by a nucleic acid of the invention, or a polypeptide comprising a polypeptide of the invention and a second domain may be immobilized on or inside a cell, a vesicle, a liposome, a film, a membrane, a metal, a resin, a polymer, a ceramic, a glass, a microelectrode, a graphitic particle, a bead, a gel, a plate, an array, a capillary tube, a crystal, a tablet, a pill, a capsule, a powder, an agglomerate, a surface, a porous structure, or materials such as wood chips, brownstock, pulp, paper, and materials deriving therefrom.

[0070] The xylanases of the invention can be used or formulated alone or as mixture (a "cocktail") of xylanases and glucanases, and other hydrolytic enzymes such as cellulases, mannanases, proteases, lipases, amylases, or redox enzymes such as laccases, peroxidases, catalases, oxidases, or reductases. They can be used formulated in a solid form such as a powder, a lyophilized preparation, a granule, a tablet, a bar, a crystal, a capsule, a pill, a pellet, or in a liquid form such as in an aqueous solution, an aerosol, a gel, a paste, a slurry, an aqueous/oil emulsion, a cream, a capsule, or in a vesicular or micellar suspension. The formulations of the invention can comprise any or a combination of the following ingredients: polyols such as a polyethylene glycol, a polyvinylalcohol, a glycerol, a sugar such as a sucrose, a sorbitol, a trehalose, a glucose, a fructose, a maltose, a mannose, a gelling agent such as a guar gum, a carageenan, an alginate, a dextrans, a cellulosic derivative, a pectin, a salt such as a sodium chloride, a sodium sulfate, an ammonium sulfate, a calcium chloride, a magnesium chloride, a zinc chloride, a zinc sulfate, a salt of a fatty acid and a fatty acid derivative, a metal chelator such as an EDTA, an EGTA, a sodium citrate, an antimicrobial agent such as a fatty acid or a fatty acid derivative, a paraben, a sorbate, a benzoate, an additional modulating compound to block the impact of an enzyme such as a protease, a bulk proteins such as a BSA, a wheat hydrolysate, a borate compound, an amino acid or a peptide, an appropriate pH or temperature modulating compound, an emulsifier such as a non-ionic and/or an ionic detergent, a redox agent such as a cystine/cysteine, a glutathione, an oxidized glutathione, a reduced or an antioxidant compound such as an ascorbic acid, or a dispersant. Cross-linking and protein

modification such as pegylation, fatty acid modification, glycosylation can also be used to improve enzyme stability.

[0071] Arrays may comprise immobilized polypeptide(s) and/or nucleic acids of the invention, or an immobilized oligonucleotide of the invention. The enzymes, fragments thereof and nucleic acids which encode the enzymes, or probes therefor, and fragments thereof, can be affixed to a solid support; and these can be economical and efficient in the use of enzymes and nucleic acids of the invention in industrial, medical, research, pharmaceutical, food and feed and food and feed supplement processing and other applications and processes. For example, a consortium or cocktail of enzymes (or active fragments thereof), which are used in a specific chemical reaction, can be attached to a solid support and dunked into a process vat. The enzymatic reaction can occur. Then, the solid support can be taken out of the vat, along with the enzymes affixed thereto, for repeated use. The isolated nucleic acid may be affixed to a solid support, which may be selected from the group of a gel, a resin, a polymer, a ceramic, a glass, a microelectrode and any combination thereof.

[0072] For example useful solid supports include gels. Some examples of gels include sepharose, gelatin, glutaraldehyde, chitosan-treated glutaraldehyde, albumin-glutaraldehyde, chitosan-Xanthan, toyopearl gel (polymer gel), alginate, alginate-polylysine, carrageenan, agarose, glyoxyl agarose, magnetic agarose, dextran-agarose, poly(Carbamoyl Sulfonate) hydrogel, BSA-PEG hydrogel, phosphorylated polyvinyl alcohol (PVA), monoaminoethyl-N-aminoethyl (MANA), amino, or any combination thereof. Also useful are resins or polymers. Some examples of resins or polymers include cellulose, acrylamide, nylon, rayon, polyester, anion-exchange resin, AMBERLITE™ XAD-7, AMBERLITE™ XAD-8, AMBERLITE™ IRA-94, AMBERLITE™ IRC-50, polyvinyl, polyacrylic, polymethacrylate, or any combination thereof. Another useful type of solid support is ceramic. Some examples include non-porous ceramic, porous ceramic, SiO₂, Al₂O₃. Glass supports are useful, including non-porous glass, porous glass, aminopropyl glass or any combination thereof. Another type of solid support which can be used is a microelectrode. An example is a polyethyleneimine-coated magnetite. Graphitic particles can be used as a solid support. Another example of a solid support is a cell, such as a red blood cell.

[0073] There are many methods which would be known to one of skill in the art for immobilizing enzymes or fragments thereof, or nucleic acids, onto a solid support. Some examples of such methods include electrostatic droplet generation, electrochemical means, via adsorption, via covalent binding, via cross-linking, via a chemical reaction or process, via encapsulation, via entrapment, via calcium alginate, or via poly (2-hydroxyethyl methacrylate). Like methods are described in *Methods in Enzymology, Immobilized Enzymes and Cells*, Part C. 1987. Academic Press. Edited by S. P. Colowick and N. O. Kaplan. Volume 136; and *Immobilization of Enzymes and Cells*. 1997. Humana Press. Edited by G. F. Bickerstaff. Series: *Methods in Biotechnology*, Edited by J. M. Walker.

[0074] The invention provides isolated, synthetic or recombinant antibodies that specifically binds to a polypeptide of the invention. The antibody can be a monoclonal or a polyclonal antibody, or is a single chained antibody. Hybridomas may comprise an antibody that specifically binds to a polypeptide of the invention.

[0075] Methods of isolating or identifying a polypeptide with a xylanase activity comprise the steps of: (a) providing an antibody of the invention; (b) providing a sample comprising polypeptides; and (c) contacting the sample of step (b) with the antibody of step (a) under conditions wherein the antibody can specifically bind to the polypeptide, thereby isolating or identifying a polypeptide having a xylanase activity. The invention provides methods of making an anti-xylanase antibody comprising administering to a non-human animal a nucleic acid of the invention or a subsequence thereof in an amount sufficient to generate a

humoral immune response, thereby making an anti-xylanase antibody. The invention provides methods of making an anti-xylanase antibody comprising administering to a non-human animal a polypeptide of the invention or a subsequence thereof in an amount sufficient to generate a humoral immune response, thereby making an anti-xylanase antibody.

[0076] Methods of producing a recombinant polypeptide comprise the steps of: (a) providing a nucleic acid operably linked to a promoter, wherein the nucleic acid comprises a sequence of the invention; and (b) expressing the nucleic acid of step (a) under conditions that allow expression of the polypeptide, thereby producing a recombinant polypeptide. The method can further comprise transforming a host cell with the nucleic acid of step (a) followed by expressing the nucleic acid of step (a), thereby producing a recombinant polypeptide in a transformed cell.

[0077] Methods for identifying a polypeptide having a xylanase activity comprise: (a) providing a polypeptide of the invention; (b) providing a xylanase substrate; and (c) contacting the polypeptide with the substrate of step (b) and detecting a decrease in the amount of substrate or an increase in the amount of a reaction product, wherein a decrease in the amount of the substrate or an increase in the amount of the reaction product detects a polypeptide having a xylanase activity.

[0078] Methods for identifying a xylanase substrate comprise: (a) providing a polypeptide of the invention; (b) providing a test substrate; and (c) contacting the polypeptide of step (a) with the test substrate of step (b) and detecting a decrease in the amount of substrate or an increase in the amount of reaction product, wherein a decrease in the amount of the substrate or an increase in the amount of a reaction product identifies the test substrate as a xylanase substrate.

[0079] Methods of determining whether a test compound specifically binds to a polypeptide comprise: (a) expressing a nucleic acid or a vector comprising the nucleic acid under conditions permissive for translation of the nucleic acid to a polypeptide, wherein the nucleic acid has a sequence of the invention; (b) providing a test compound; (c) contacting the polypeptide with the test compound; and (d) determining whether the test compound of step (b) specifically binds to the polypeptide.

[0080] Methods of determining whether a test compound specifically binds to a polypeptide comprise: (a) providing a polypeptide of the invention; (b) providing a test compound; (c) contacting the polypeptide with the test compound; and (d) determining whether the test compound of step (b) specifically binds to the polypeptide.

[0081] Methods for identifying a modulator of a xylanase activity comprising: (a) providing a polypeptide of the invention; (b) providing a test compound; (c) contacting the polypeptide of step (a) with the test compound of step (b) and measuring an activity of the xylanase wherein a change in the xylanase activity measured in the presence of the test compound compared to the activity in the absence of the test compound provides a determination that the test compound modulates the xylanase activity. The xylanase activity can be measured by providing a xylanase substrate and detecting a decrease in the amount of the substrate or an increase in the amount of a reaction product, or, an increase in the amount of the substrate or a decrease in the amount of a reaction product. A decrease in the amount of the substrate or an increase in the amount of the reaction product with the test compound as compared to the amount of substrate or reaction product without the test compound identifies the test compound as an activator of a xylanase activity. In one aspect, an increase in the amount of the substrate or a decrease in the amount of the reaction product with the test compound as compared to the amount of substrate or reaction product without the test compound identifies the test compound as an inhibitor of a xylanase activity.

[0082] Computer systems may comprise a processor and a data storage device wherein said data storage

device has stored thereon a polypeptide sequence or a nucleic acid sequence, wherein the polypeptide sequence comprises sequence of the invention, a polypeptide encoded by a nucleic acid of the invention. The computer systems can further comprise a sequence comparison algorithm and a data storage device having at least one reference sequence stored thereon. The sequence comparison algorithm may comprise a computer program that indicates polymorphisms. The computer system can further comprise an identifier that identifies one or more features in said sequence. Computer readable media may have stored thereon a polypeptide sequence or a nucleic acid sequence of the invention. Methods for identifying a feature in a sequence comprise the steps of: (a) reading the sequence using a computer program which identifies one or more features in a sequence, wherein the sequence comprises a polypeptide sequence or a nucleic acid sequence of the invention; and (b) identifying one or more features in the sequence with the computer program. Methods for comparing a first sequence to a second sequence comprise the steps of: (a) reading the first sequence and the second sequence through use of a computer program which compares sequences, wherein the first sequence comprises a polypeptide sequence or a nucleic acid sequence of the invention; and (b) determining differences between the first sequence and the second sequence with the computer program. The step of determining differences between the first sequence and the second sequence can further comprise the step of identifying polymorphisms. The method can further comprise an identifier that identifies one or more features in a sequence. The method can comprise reading the first sequence using a computer program and identifying one or more features in the sequence.

[0083] Methods for isolating or recovering a nucleic acid encoding a polypeptide having a xylanase activity from an environmental sample comprise the steps of: (a) providing an amplification primer sequence pair for amplifying a nucleic acid encoding a polypeptide having a xylanase activity, wherein the primer pair is capable of amplifying a nucleic acid of the invention; (b) isolating a nucleic acid from the environmental sample or treating the environmental sample such that nucleic acid in the sample is accessible for hybridization to the amplification primer pair; and, (c) combining the nucleic acid of step (b) with the amplification primer pair of step (a) and amplifying nucleic acid from the environmental sample, thereby isolating or recovering a nucleic acid encoding a polypeptide having a xylanase activity from an environmental sample. One or each member of the amplification primer sequence pair can comprise an oligonucleotide comprising at least about 10 to 50 consecutive bases of a sequence of the invention. The amplification primer sequence pair may be an amplification pair as disclosed herein.

[0084] Methods for isolating or recovering a nucleic acid encoding a polypeptide having a xylanase activity from an environmental sample comprise the steps of: (a) providing a polynucleotide probe comprising a nucleic acid of the invention or a subsequence thereof; (b) isolating a nucleic acid from the environmental sample or treating the environmental sample such that nucleic acid in the sample is accessible for hybridization to a polynucleotide probe of step (a); (c) combining the isolated nucleic acid or the treated environmental sample of step (b) with the polynucleotide probe of step (a); and (d) isolating a nucleic acid that specifically hybridizes with the polynucleotide probe of step (a), thereby isolating or recovering a nucleic acid encoding a polypeptide having a xylanase activity from an environmental sample. The environmental sample can comprise a water sample, a liquid sample, a soil sample, an air sample or a biological sample. In one aspect, the biological sample can be derived from a bacterial cell, a protozoan cell, an insect cell, a yeast cell, a plant cell, a fungal cell or a mammalian cell.

[0085] Methods of generating a variant of a nucleic acid encoding a polypeptide having a xylanase activity comprise the steps of: (a) providing a template nucleic acid comprising a nucleic acid of the invention; and (b) modifying, deleting or adding one or more nucleotides in the template sequence, or a combination thereof, to generate a variant of the template nucleic acid. The method can further comprise expressing the variant nucleic acid to generate a variant xylanase polypeptide. The modifications, additions or

deletions can be introduced by a method comprising error-prone PCR, shuffling, oligonucleotide-directed mutagenesis, assembly PCR, sexual PCR mutagenesis, *in vivo* mutagenesis, cassette mutagenesis, recursive ensemble mutagenesis, exponential ensemble mutagenesis, site-specific mutagenesis, gene reassembly (e.g., GeneReassembly, see, e.g., U.S. Patent No. 6,537,776), Gene Site Saturation Mutagenesis (GSSM), synthetic ligation reassembly (SLR) or a combination thereof. The modifications, additions or deletions may be introduced by a method comprising recombination, recursive sequence recombination, phosphothioate-modified DNA mutagenesis, uracil-containing template mutagenesis, gapped duplex mutagenesis, point mismatch repair mutagenesis, repair-deficient host strain mutagenesis, chemical mutagenesis, radiogenic mutagenesis, deletion mutagenesis, restriction-selection mutagenesis, restriction-purification mutagenesis, artificial gene synthesis, ensemble mutagenesis, chimeric nucleic acid multimer creation and a combination thereof.

[0086] The method can be iteratively repeated until a xylanase having an altered or different activity or an altered or different stability from that of a polypeptide encoded by the template nucleic acid is produced. The variant xylanase polypeptide may be thermotolerant, and retain some activity after being exposed to an elevated temperature. The variant xylanase polypeptide may have increased glycosylation as compared to the xylanase encoded by a template nucleic acid. Alternatively, the variant xylanase polypeptide has a xylanase activity under a high temperature, wherein the xylanase encoded by the template nucleic acid is not active under the high temperature. The method can be iteratively repeated until a xylanase coding sequence having an altered codon usage from that of the template nucleic acid is produced. The method can be iteratively repeated until a xylanase gene having higher or lower level of message expression or stability from that of the template nucleic acid is produced. Formulation of the final xylanase product may enable an increase or modulation of the performance of the xylanase in the product.

[0087] Methods for modifying codons in a nucleic acid encoding a polypeptide having a xylanase activity to increase its expression in a host cell comprise: (a) providing a nucleic acid of the invention encoding a polypeptide having a xylanase activity; and, (b) identifying a non-preferred or a less preferred codon in the nucleic acid of step (a) and replacing it with a preferred or neutrally used codon encoding the same amino acid as the replaced codon, wherein a preferred codon is a codon over-represented in coding sequences in genes in the host cell and a non-preferred or less preferred codon is a codon under-represented in coding sequences in genes in the host cell, thereby modifying the nucleic acid to increase its expression in a host cell.

[0088] Methods for modifying codons in a nucleic acid encoding a polypeptide having a xylanase activity comprise: (a) providing a nucleic acid of the invention; and, (b) identifying a codon in the nucleic acid of step (a) and replacing it with a different codon encoding the same amino acid as the replaced codon, thereby modifying codons in a nucleic acid encoding a xylanase.

[0089] Methods for modifying codons in a nucleic acid encoding a polypeptide having a xylanase activity to increase its expression in a host cell comprise: (a) providing a nucleic acid of the invention encoding a xylanase polypeptide; and, (b) identifying a non-preferred or a less preferred codon in the nucleic acid of step (a) and replacing it with a preferred or neutrally used codon encoding the same amino acid as the replaced codon, wherein a preferred codon is a codon over-represented in coding sequences in genes in the host cell and a non-preferred or less preferred codon is a codon under-represented in coding sequences in genes in the host cell, thereby modifying the nucleic acid to increase its expression in a host cell.

[0090] Methods for modifying a codon in a nucleic acid encoding a polypeptide having a xylanase activity to decrease its expression in a host cell comprise: (a) providing a nucleic acid of the invention; and (b) identifying at least one preferred codon in the nucleic acid of step (a) and replacing it with a non-preferred

or less preferred codon encoding the same amino acid as the replaced codon, wherein a preferred codon is a codon over-represented in coding sequences in genes in a host cell and a non-preferred or less preferred codon is a codon under-represented in coding sequences in genes in the host cell, thereby modifying the nucleic acid to decrease its expression in a host cell. The host cell can be a bacterial cell, a fungal cell, an insect cell, a yeast cell, a plant cell or a mammalian cell.

[0091] Methods for producing a library of nucleic acids encoding a plurality of modified xylanase active sites or substrate binding sites, wherein the modified active sites or substrate binding sites are derived from a first nucleic acid comprising a sequence encoding a first active site or a first substrate binding site, comprise: (a) providing a first nucleic acid encoding a first active site or first substrate binding site, wherein the first nucleic acid sequence comprises a sequence that hybridizes under stringent conditions to a sequence of the invention, or a subsequence thereof, and the nucleic acid encodes a xylanase active site or a xylanase substrate binding site; (b) providing a set of mutagenic oligonucleotides that encode naturally-occurring amino acid variants at a plurality of targeted codons in the first nucleic acid; and, (c) using the set of mutagenic oligonucleotides to generate a set of active site-encoding or substrate binding site-encoding variant nucleic acids encoding a range of amino acid variations at each amino acid codon that was mutagenized, thereby producing a library of nucleic acids encoding a plurality of modified xylanase active sites or substrate binding sites. The method may comprise mutagenizing the first nucleic acid of step (a) by a method comprising an optimized directed evolution system, Gene Site Saturation Mutagenesis (GSSM), or a synthetic ligation reassembly (SLR). The method may comprise mutagenizing the first nucleic acid of step (a) or variants by a method comprising error-prone PCR, shuffling, oligonucleotide-directed mutagenesis, assembly PCR, sexual PCR mutagenesis, in vivo mutagenesis, cassette mutagenesis, recursive ensemble mutagenesis, exponential ensemble mutagenesis, site-specific mutagenesis, gene reassembly (GeneReassembly, U.S. Patent No. 6,537,776), Gene Site Saturation Mutagenesis (GSSM), synthetic ligation reassembly (SLR) and a combination thereof. The method may comprise mutagenizing the first nucleic acid of step (a) or variants by a method comprising recombination, recursive sequence recombination, phosphothioate-modified DNA mutagenesis, uracil-containing template mutagenesis, gapped duplex mutagenesis, point mismatch repair mutagenesis, repair-deficient host strain mutagenesis, chemical mutagenesis, radiogenic mutagenesis, deletion mutagenesis, restriction-selection mutagenesis, restriction-purification mutagenesis, artificial gene synthesis, ensemble mutagenesis, chimeric nucleic acid multimer creation and a combination thereof.

[0092] Methods for making a small molecule comprise: (a) providing a plurality of biosynthetic enzymes capable of synthesizing or modifying a small molecule, wherein one of the enzymes comprises a xylanase enzyme encoded by a nucleic acid of the invention; (b) providing a substrate for at least one of the enzymes of step (a); and (c) reacting the substrate of step (b) with the enzymes under conditions that facilitate a plurality of biocatalytic reactions to generate a small molecule by a series of biocatalytic reactions. Methods for modifying a small molecule comprise: (a) providing a xylanase enzyme, wherein the enzyme comprises a polypeptide of the invention, or, a polypeptide encoded by a nucleic acid of the invention, or a subsequence thereof; (b) providing a small molecule; and (c) reacting the enzyme of step (a) with the small molecule of step (b) under conditions that facilitate an enzymatic reaction catalyzed by the xylanase enzyme, thereby modifying a small molecule by a xylanase enzymatic reaction. The method can comprise a plurality of small molecule substrates for the enzyme of step (a), thereby generating a library of modified small molecules produced by at least one enzymatic reaction catalyzed by the xylanase enzyme. The method can comprise a plurality of additional enzymes under conditions that facilitate a plurality of biocatalytic reactions by the enzymes to form a library of modified small molecules produced by the plurality of enzymatic reactions. The method can further comprise the step of testing the library to determine if a particular modified small molecule that exhibits a desired activity is present within the library. The step of testing the library can further comprise the steps of systematically eliminating all but one of the

biocatalytic reactions used to produce a portion of the plurality of the modified small molecules within the library by testing the portion of the modified small molecule for the presence or absence of the particular modified small molecule with a desired activity, and identifying at least one specific biocatalytic reaction that produces the particular modified small molecule of desired activity.

[0093] Methods for determining a functional fragment of a xylanase enzyme comprise the steps of: (a) providing a xylanase enzyme, wherein the enzyme comprises a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention, or a subsequence thereof; and (b) deleting a plurality of amino acid residues from the sequence of step (a) and testing the remaining subsequence for a xylanase activity, thereby determining a functional fragment of a xylanase enzyme. In one aspect, the xylanase activity is measured by providing a xylanase substrate and detecting a decrease in the amount of the substrate or an increase in the amount of a reaction product.

[0094] Methods for whole cell engineering of new or modified phenotypes by using real-time metabolic flux analysis comprise: (a) making a modified cell by modifying the genetic composition of a cell, wherein the genetic composition is modified by addition to the cell of a nucleic acid of the invention; (b) culturing the modified cell to generate a plurality of modified cells; (c) measuring at least one metabolic parameter of the cell by monitoring the cell culture of step (b) in real time; and, (d) analyzing the data of step (c) to determine if the measured parameter differs from a comparable measurement in an unmodified cell under similar conditions, thereby identifying an engineered phenotype in the cell using real-time metabolic flux analysis. The genetic composition of the cell can be modified by a method comprising deletion of a sequence or modification of a sequence in the cell, or, knocking out the expression of a gene. The method can further comprise selecting a cell comprising a newly engineered phenotype. The method can comprise culturing the selected cell, thereby generating a new cell strain comprising a newly engineered phenotype.

[0095] Isolated, synthetic or recombinant signal sequences may consist of, or comprise, a sequence as set forth in residues 1 to 12, 1 to 13, 1 to 14, 1 to 15, 1 to 16, 1 to 17, 1 to 18, 1 to 19, 1 to 20, 1 to 21, 1 to 22, 1 to 23, 1 to 24, 1 to 25, 1 to 26, 1 to 27, 1 to 28, 1 to 28, 1 to 30, 1 to 31, 1 to 32, 1 to 33, 1 to 34, 1 to 35, 1 to 36, 1 to 37, 1 to 38, 1 to 40, 1 to 41, 1 to 42, 1 to 43 or 1 to 44, of a polypeptide of the invention, including exemplary polypeptide sequences of the invention.

[0096] Chimeric polypeptides may comprise at least a first domain comprising a signal peptide (SP) and at least a second domain comprising a heterologous polypeptide or peptide comprising a sequence of the invention, or a subsequence thereof, wherein the heterologous polypeptide or peptide is not naturally associated with the signal peptide (SP). The signal peptide (SP) may not be derived from a xylanase. The heterologous polypeptide or peptide can be amino terminal to, carboxy terminal to or on both ends of the signal peptide (SP) or a xylanase catalytic domain (CD). Isolated, synthetic or recombinant nucleic acids encoding a chimeric polypeptide may comprise at least a first domain comprising signal peptide (SP) and at least a second domain comprising a heterologous polypeptide or peptide comprising a sequence of the invention, or a subsequence thereof, wherein the heterologous polypeptide or peptide is not naturally associated with the signal peptide (SP).

[0097] Methods of increasing thermotolerance or thermostability of a xylanase polypeptide comprising glycosylating a xylanase polypeptide, wherein the polypeptide comprises at least thirty contiguous amino acids of a polypeptide of the invention; or a polypeptide encoded by a nucleic acid sequence of the invention, thereby increasing the thermotolerance or thermostability of the xylanase polypeptide. In one aspect, the xylanase specific activity can be thermostable or thermotolerant at a temperature in the range from greater than about 0°C to about 20°C, about 20°C to about 37°C, about 37°C to about 50°C, about 50°C to about 70°C, about 70°C to about 75°C, about 75°C to about 80°C, about 80°C to about 85°C, about 85°C to about 90°C, about 90°C to about 95°C, about 95°C to about 100°C, about 100°C to about

110°C, or higher.

[0098] Methods for overexpressing a recombinant xylanase polypeptide in a cell comprise expressing a vector comprising a nucleic acid comprising a nucleic acid of the invention or a nucleic acid sequence of the invention, wherein the sequence identities are determined by analysis with a sequence comparison algorithm or by visual inspection, wherein overexpression is effected by use of a high activity promoter, a dicistronic vector or by gene amplification of the vector.

[0099] Methods of making a transgenic plant and seeds comprise: (a) introducing a heterologous nucleic acid sequence into the cell, wherein the heterologous nucleic sequence comprises a nucleic acid sequence of the invention, thereby producing a transformed plant or seed cell; and (b) producing a transgenic plant from the transformed cell or seed. The step (a) can further comprise introducing the heterologous nucleic acid sequence by electroporation or microinjection of plant cell protoplasts. The step (a) can further comprise introducing the heterologous nucleic acid sequence directly to plant tissue by DNA particle bombardment. Alternatively, the step (a) can further comprise introducing the heterologous nucleic acid sequence into the plant cell DNA using an *Agrobacterium tumefaciens* host. The plant cell can be a potato, corn, rice, wheat, tobacco, or barley cell.

[0100] Methods of expressing a heterologous nucleic acid sequence in a plant cell comprise: (a) transforming the plant cell with a heterologous nucleic acid sequence operably linked to a promoter, wherein the heterologous nucleic sequence comprises a nucleic acid of the invention; (b) growing the plant under conditions wherein the heterologous nucleic acids sequence is expressed in the plant cell. Methods of expressing a heterologous nucleic acid sequence in a plant cell comprise: (a) transforming the plant cell with a heterologous nucleic acid sequence operably linked to a promoter, wherein the heterologous nucleic sequence comprises a sequence of the invention; (b) growing the plant under conditions wherein the heterologous nucleic acids sequence is expressed in the plant cell.

[0101] The invention provides methods for hydrolyzing, breaking up or disrupting a xylan-comprising composition comprising: (a) providing a polypeptide of the invention having a xylanase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a composition comprising a xylan; and (c) contacting the polypeptide of step (a) with the composition of step (b) under conditions wherein the xylanase hydrolyzes, breaks up or disrupts the xylan-comprising composition. In one aspect, the composition comprises a plant cell, a bacterial cell, a yeast cell, an insect cell, or an animal cell. Thus, the composition can comprise any plant or plant part, any xylan-containing food or feed, a waste product and the like.

[0102] The invention provides methods for liquefying or removing a xylan-comprising composition comprising: (a) providing a polypeptide of the invention having a xylanase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a composition comprising a xylan; and (c) contacting the polypeptide of step (a) with the composition of step (b) under conditions wherein the xylanase removes, softens or liquefies the xylan-comprising composition.

[0103] Detergent compositions may comprise a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention. The said xylanase can be a nonsurface-active xylanase or a surface-active xylanase. The xylanase can be formulated in a non-aqueous liquid composition, a cast solid, a granular form, a particulate form, a compressed tablet, a gel form, a paste or a slurry form. Methods for washing an object comprise: (a) providing a composition comprising a polypeptide of the invention having a xylanase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing an object; and (c) contacting the polypeptide of step (a) and the object of step (b) under conditions wherein the composition

can wash the object.

[0104] Textiles or fabrics, including, e.g., threads, may comprise a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention. The textiles or fabrics may comprise xylan-containing fibers. Methods for treating a textile or fabric (e.g., removing a stain from a composition) comprise: (a) providing a composition comprising a polypeptide of the invention having a xylanase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a textile or fabric comprising a xylan; and (c) contacting the polypeptide of step (a) and the composition of step (b) under conditions wherein the xylanase can treat the textile or fabric (e.g., remove the stain). The invention provides methods for improving the finish of a fabric comprising: (a) providing a composition comprising a polypeptide of the invention having a xylanase activity, or a polypeptide encoded by a nucleic acid of the invention; (b) providing a fabric; and (c) contacting the polypeptide of step (a) and the fabric of step (b) under conditions wherein the polypeptide can treat the fabric thereby improving the finish of the fabric. The fabric may be wool or a silk, or the fabric is a cellulosic fiber or a blend of a natural fiber and a synthetic fiber.

[0105] Feeds or foods may comprise a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention. Methods for hydrolyzing xylans in a feed or a food prior to consumption by an animal comprise: (a) obtaining a feed material comprising a xylanase of the invention, or a xylanase encoded by a nucleic acid of the invention; and (b) adding the polypeptide of step (a) to the feed or food material in an amount sufficient for a sufficient time period to cause hydrolysis of the xylan and formation of a treated food or feed, thereby hydrolyzing the xylans in the food or the feed prior to consumption by the animal. Methods for hydrolyzing xylans in a feed or a food after consumption by an animal comprise: (a) obtaining a feed material comprising a xylanase of the invention, or a xylanase encoded by a nucleic acid of the invention; (b) adding the polypeptide of step (a) to the feed or food material; and (c) administering the feed or food material to the animal, wherein after consumption, the xylanase causes hydrolysis of xylans in the feed or food in the digestive tract of the animal. The food or the feed can be, e.g., a cereal, a grain, a corn and the like.

[0106] Dough or bread products may comprise a polypeptide having a xylanase activity, wherein the polypeptide comprises a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof. Methods of dough conditioning comprise contacting a dough or a bread product with at least one polypeptide having a xylanase activity, wherein the polypeptide comprises a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof, under conditions sufficient for conditioning the dough.

[0107] Beverages may comprise a polypeptide having a xylanase activity, wherein the polypeptide comprises a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention. Methods of beverage production comprise administration of at least one polypeptide having a xylanase activity, wherein the polypeptide comprises a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof, to a beverage or a beverage precursor under conditions sufficient for decreasing the viscosity of the beverage, wherein in one aspect (optionally) the beverage or beverage precursor is a wort or a beer.

[0108] Food or nutritional supplements for an animal may comprise a polypeptide of the invention, e.g., a polypeptide encoded by the nucleic acid of the invention. The polypeptide in the food or nutritional supplement can be glycosylated. Edible enzyme delivery matrices may comprise a polypeptide of the invention, e.g., a polypeptide encoded by the nucleic acid of the invention. The delivery matrix may

comprise a pellet. The polypeptide can be glycosylated. The xylanase activity may be thermotolerant or thermostable.

[0109] A food, a feed or a nutritional supplement may comprise a polypeptide of the invention. Methods for utilizing a xylanase as a nutritional supplement in an animal diet comprise: preparing a nutritional supplement containing a xylanase enzyme comprising at least thirty contiguous amino acids of a polypeptide of the invention; and administering the nutritional supplement to an animal to increase utilization of a xylan contained in a feed or a food ingested by the animal. The animal can be a human, a ruminant or a monogastric animal. The xylanase enzyme can be prepared by expression of a polynucleotide encoding the xylanase in an organism selected from the group consisting of a bacterium, a yeast, a plant, an insect, a fungus and an animal. The organism can be selected from the group consisting of an *S. pombe*, *S. cerevisiae*, *Pichia pastoris*, *Pseudomonas* sp., *E. coli*, *Streptomyces* sp., *Bacillus* sp. and *Lactobacillus* sp.

[0110] Edible enzyme delivery matrix may comprise a thermostable recombinant xylanase enzyme of the invention. Methods for delivering a xylanase supplement to an animal comprise: preparing an edible enzyme delivery matrix in the form of pellets comprising a granulate edible carrier and a thermostable recombinant xylanase enzyme, wherein the pellets readily disperse the xylanase enzyme contained therein into aqueous media, and administering the edible enzyme delivery matrix to the animal. The recombinant xylanase enzyme can comprise a polypeptide of the invention. The granulate edible carrier can comprise a carrier selected from the group consisting of a grain germ, a grain germ that is spent of oil, a hay, an alfalfa, a timothy, a soy hull, a sunflower seed meal and a wheat midd. The edible carrier can comprise grain germ that is spent of oil. The xylanase enzyme can be glycosylated to provide thermostability at pelletizing conditions. The delivery matrix can be formed by pelletizing a mixture comprising a grain germ and a xylanase. The pelletizing conditions can include application of steam. The pelletizing conditions can comprise application of a temperature in excess of about 80°C for about 5 minutes and the enzyme retains a specific activity of at least 350 to about 900 units per milligram of enzyme.

[0111] Methods for improving texture and flavor of a dairy product comprise: (a) providing a polypeptide of the invention having a xylanase activity, or a xylanase encoded by a nucleic acid of the invention; (b) providing a dairy product; and (c) contacting the polypeptide of step (a) and the dairy product of step (b) under conditions wherein the xylanase can improve the texture or flavor of the dairy product. The dairy product may comprise a cheese or a yogurt. Dairy products may comprise a xylanase of the invention, or is encoded by a nucleic acid of the invention.

[0112] Methods for improving the extraction of oil from an oil-rich plant material comprise: (a) providing a polypeptide of the invention having a xylanase activity, or a xylanase encoded by a nucleic acid of the invention; (b) providing an oil-rich plant material; and (c) contacting the polypeptide of step (a) and the oil-rich plant material. The oil-rich plant material may comprise an oil-rich seed. The oil can be a soybean oil, an olive oil, a rapeseed (canola) oil or a sunflower oil.

[0113] Methods for preparing a fruit or vegetable juice, syrup, puree or extract comprise: (a) providing a polypeptide of the invention having a xylanase activity, or a xylanase encoded by a nucleic acid of the invention; (b) providing a composition or a liquid comprising a fruit or vegetable material; and (c) contacting the polypeptide of step (a) and the composition, thereby preparing the fruit or vegetable juice, syrup, puree or extract.

[0114] Papers or paper products or paper pulp may comprise a xylanase of the invention, or a

polypeptide encoded by a nucleic acid of the invention. Methods for treating a biomass, e.g., any paper or a paper or wood pulp comprise: (a) providing a polypeptide of the invention having a xylanase activity, or a xylanase encoded by a nucleic acid of the invention; (b) providing a composition, e.g., a biomass, comprising a paper or a paper or wood pulp; and (c) contacting the polypeptide of step (a) and the composition of step (b) under conditions wherein the xylanase can treat the paper or paper or wood pulp.

[0115] Methods for reducing the amount of lignin (delignification), or solubilizing a lignin, in a paper or paper product, a paper waste, a wood, wood pulp or wood product, or a wood or paper recycling composition, comprise contacting the paper or paper product, wood, wood pulp or wood product, or wood or paper recycling composition with a polypeptide of the invention, or an enzymatically active fragment thereof.

[0116] Methods for hydrolyzing hemicelluloses in a wood, wood product, paper pulp, paper product or paper waste comprise contacting the wood, wood product, paper pulp, paper product or paper waste with a polypeptide of the invention, or an enzymatically active fragment thereof.

[0117] Methods for enzymatic decoloring (e.g., bleaching) of paper, hemp or flax pulp comprise contacting the paper, hemp or flax pulp with a xylanase and a decoloring (e.g., bleaching) agent, wherein the xylanase comprises a polypeptide of the invention, or an enzymatically active fragment thereof. The decoloring (e.g., bleaching) agent can comprise oxygen or hydrogen peroxide.

[0118] Methods for of decoloring (e.g., bleaching) a lignocellulose pulp comprise contacting the lignocellulose pulp with a xylanase, comprising a polypeptide of the invention, or an enzymatically active fragment thereof.

[0119] Methods for enzymatic deinking of paper, paper waste, paper recycled product, deinking toner from non-contact printed wastepaper or mixtures of non-contact and contact printed wastepaper, comprise contacting the paper, paper waste, paper recycled product, non-contact printed wastepaper or contact printed wastepaper with a xylanase comprising a polypeptide of the invention, or an enzymatically active fragment thereof.

[0120] Methods for decoloring (e.g., bleaching) a thread, fabric, yarn, cloth or textile comprise contacting the fabric, yarn, cloth or textile with a xylanase under conditions suitable to produce a whitening of the textile, wherein the xylanase comprises a polypeptide of the invention, or an enzymatically active fragment thereof. The thread, fabric, yarn, cloth or textile can comprise a non-cotton cellulosic thread, fabric, yarn, cloth or textile. Fabrics, yarns, cloths or textiles may comprise a polypeptide having a sequence of the invention, or a polypeptide encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof, wherein in one aspect (optionally) the fabric, yarn, cloth or textile comprises a non-cotton cellulosic fabric, yarn, cloth or textile.

[0121] Methods for decoloring (e.g., bleaching) or deinking newspaper comprise contacting the newspaper, wherein the xylanase comprises a polypeptide of the invention, or an enzymatically active fragment thereof.

[0122] Wood, wood chips, wood pulp, wood products, paper pulps, paper products, newspapers or paper waste may comprise a polypeptide of the invention, or an enzymatically active fragment thereof. Thread, fabric, yarn, cloth or textile may comprise a polypeptide of the invention, or an enzymatically active fragment thereof.

[0123] Methods for reducing lignin in a wood or wood product comprise contacting the wood or wood

product with a polypeptide having a xylanase activity, wherein the polypeptide has a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof.

[0124] Methods for reducing a lignin in a biomass, e.g., in a wood, a wood pulp, a Kraft pulp, a paper, a paper product or a paper pulp under high temperature and basic pH conditions comprise: (a) providing at least one polypeptide having a xylanase activity, wherein the polypeptide retains xylanase activity under conditions comprising a temperature of at least about 80°C, 85°C, 90°C or more, and a basic pH of at least about pH 10.5, pH 11, pH 12, pH 12.5 or more (basic) wherein the polypeptide comprises a xylanase having a sequence of the invention, or the xylanase is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof; (b) providing a lignin-comprising biomass, e.g., a lignin-comprising wood, wood pulp, Kraft pulp, paper, paper product or paper pulp; and (c) contacting the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp with the polypeptide of step (a) under conditions comprising a temperature of at least about 80°C, 85°C, 90°C or more, and a basic pH of at least about pH 10.5, pH 11, pH 12, pH 12.5 or more (basic), wherein the polypeptide reduces the lignin-comprising biomass, e.g., the lignin in the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp.

[0125] Methods for treating a lignin-comprising biomass, e.g., a wood, a wood pulp, a Kraft pulp, a paper product, a paper or a paper pulp under high temperature and basic pH conditions, comprise: (a) providing at least one polypeptide having a xylanase activity, wherein the polypeptide retains xylanase activity under conditions comprising a temperature of at least about 80°C, 85°C, 90°C or more, and a basic pH of at least about pH 10.5, pH 11, pH 12, pH 12.5 or more (basic), wherein the polypeptide comprises a xylanase having a sequence of the invention, or the xylanase is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof; (b) providing a lignin-comprising biomass, e.g., a wood, a wood pulp, a Kraft pulp, a paper, a paper product or a paper pulp; and (c) contacting the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp with the polypeptide of step (a) under conditions comprising a temperature of at least about 80°C, 85°C, 90°C or more, and a basic pH of at least about pH 10.5, pH 11, pH 12, pH 12.5 or more (basic), wherein the polypeptide catalyzes hydrolysis of compounds in the lignin-comprising biomass, e.g., wood, wood pulp, Kraft pulp, paper, paper product or paper pulp, and wherein in one aspect (optionally) the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp comprises a softwood and hardwood, or the wood, wood pulp, Kraft pulp, paper or paper pulp is derived from a softwood and hardwood; and wherein optionally after the treatment the pulp has a consistency of at least about 10%, or at least about 32%.

[0126] Methods for decoloring a biomass, e.g., a wood, a wood pulp, a Kraft pulp, a paper, a paper product or a paper pulp under high temperature and basic pH conditions, comprise: (a) providing at least one polypeptide having a xylanase activity, wherein the polypeptide retains xylanase activity under conditions comprising a temperature of at least about 80°C, 85°C, 90°C or more, and a basic pH of at least about pH 10.5, pH 11, pH 12, pH 12.5 or more (basic), wherein the polypeptide comprises a xylanase having a sequence of the invention, or the xylanase is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof; (b) providing a biomass, e.g., a wood, a wood pulp, a Kraft pulp, a paper, a paper product or a paper pulp; and (c) contacting the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp with the polypeptide of step (a) under conditions comprising a temperature of at least about 80°C, 85°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 9.5, pH 10.0, pH 10.5, pH 11, pH 12, pH 12.5 or more (basic), wherein the polypeptide catalyzes hydrolysis of compounds in the biomass, e.g., a wood, wood pulp, Kraft pulp, paper, paper product or paper pulp, thereby decoloring (e.g., bleaching) the biomass, e.g., a wood, wood pulp,

Kraft pulp, paper, paper product or paper pulp.

[0127] Methods for reducing the use of decoloring (e.g., bleaching) chemicals in a biomass, e.g., a wood, a wood pulp, a Kraft pulp, a paper, a paper product or a paper pulp decoloring (e.g., bleaching) process under high temperature and basic pH conditions, comprise: (a) providing at least one polypeptide having a xylanase activity, wherein the polypeptide retains xylanase activity under conditions comprising a temperature of at least about 80°C, 85°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 10.5, pH 11, pH 12, pH 12.5 or more (basic), wherein the polypeptide comprises a xylanase having a sequence of the invention, or the xylanase is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof; (b) providing a biomass, e.g., a wood, a wood pulp, a Kraft pulp, a paper, a paper product or a paper pulp; and (c) contacting the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp with the polypeptide of step (a) under conditions comprising a temperature of at least about 80°C, 85°C, 86°C, 87°C, 88°C, 89°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 10.5, pH 11, pH 12, pH 12.5 or more (basic), wherein the polypeptide catalyzes hydrolysis of compounds in the biomass, e.g., wood, wood pulp, Kraft pulp, paper, paper product or paper pulp, thereby biobleaching the biomass, e.g., wood, wood pulp, Kraft pulp, paper, paper product or paper pulp and reducing the use of decoloring (e.g., bleaching) chemicals in the decoloring (e.g., bleaching) process; wherein optionally the decoloring (e.g., bleaching) chemical comprises a chlorine, a chlorine dioxide, a caustic, a peroxide, or any combination thereof.

[0128] Methods for paper or pulp deinking under high temperature and basic pH conditions, comprise: (a) providing at least one polypeptide having a xylanase activity, wherein the polypeptide retains xylanase activity under conditions comprising a temperature of at least about 80°C, 85°C, 86°C, 87°C, 88°C, 89°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 11, wherein the polypeptide comprises a xylanase having a sequence of the invention, or the xylanase is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof; (b) providing an ink-comprising biomass, e.g., a wood, wood pulp, Kraft pulp, paper, a paper product or paper pulp; and (c) contacting the biomass, e.g., wood, wood pulp, Kraft pulp, paper, paper product or paper pulp with the polypeptide of step (a) under conditions comprising a temperature of at least about 85°C and a basic pH of at least about pH 11, wherein the polypeptide catalyzes hydrolysis of compounds in the biomass, e.g., wood, wood pulp, Kraft pulp, paper or paper pulp, thereby facilitating deinking of the biomass, e.g., wood, wood pulp, Kraft pulp, paper, paper product or paper pulp.

[0129] Methods for releasing a lignin from a biomass, e.g., a wood, a wood pulp, a Kraft pulp, a paper, a paper product or a paper pulp under high temperature and basic pH conditions, comprise: (a) providing at least one polypeptide having a xylanase activity, wherein the polypeptide retains xylanase activity under conditions comprising a temperature of at least about 80°C, 85°C, 86°C, 87°C, 88°C, 89°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 11, wherein the polypeptide comprises a xylanase having a sequence of the invention, or the xylanase is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof; (b) providing a lignin-comprising biomass, e.g., wood, wood pulp, Kraft pulp, paper, paper product or paper pulp; and (c) contacting the wood, wood pulp, Kraft pulp, paper, paper product or a paper pulp of step (b) with the polypeptide of step (a) under conditions comprising a temperature of at least about 80°C, 85°C, 86°C, 87°C, 88°C, 89°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about

pH 11, wherein the polypeptide catalyzes hydrolysis of compounds in the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp, thereby facilitating release of lignin from the biomass, e.g., wood, wood pulp, Kraft pulp, paper, paper product or paper pulp; wherein in one aspect (optionally) after the treatment the pulp has a consistency of about 10%.

[0130] Compositions may comprise a biomass, e.g., wood, a wood pulp, a Kraft pulp, a paper, a paper product or a paper pulp comprising a polypeptide having a xylanase activity, wherein the polypeptide has a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof. Optionally the biomass, e.g., wood, wood pulp, Kraft pulp, paper, paper product or paper pulp comprises a softwood and hardwood, or the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp derived from a softwood and hardwood.

[0131] Methods for making ethanol comprise contacting a starch-comprising composition with a polypeptide having a xylanase activity, wherein the polypeptide has a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof. Compositions may comprise an ethanol and a polypeptide having a xylanase activity, wherein the polypeptide has a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof. Methods for making ethanol under high temperature and basic pH conditions, comprise: (a) providing at least one polypeptide having a xylanase activity, wherein the polypeptide retains xylanase activity under conditions comprising a temperature of at least about 80°C, 85°C, 86°C, 87°C, 88°C, 89°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 11, wherein the polypeptide comprises a xylanase having a sequence of the invention, or the xylanase is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof; (b) providing a starch-comprising composition comprising a wood, wood pulp, Kraft pulp, paper, a paper product or paper pulp; and (c) contacting the composition of step (b) with the polypeptide of step (a) under conditions comprising a temperature of at least about 80°C, 85°C, 86°C, 87°C, 88°C, 89°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 11, wherein the polypeptide catalyzes hydrolysis of compounds in the wood, wood pulp, Kraft pulp, paper or paper pulp, thereby generating ethanol from the wood, wood pulp, Kraft pulp, paper, paper product or paper pulp.

[0132] Pharmaceutical compositions may comprise a polypeptide having a xylanase activity, wherein the polypeptide comprises a sequence of the invention, or the polypeptide is encoded by a nucleic acid comprising a sequence of the invention, or an enzymatically active fragment thereof. Methods for eliminating or protecting animals from a microorganism comprising a xylan comprise administering a polypeptide of the invention. The microorganism can be a bacterium comprising a xylan, e.g., *Salmonella*.

[0133] The pharmaceutical composition may act as a digestive aid or an anti-microbial (e.g., against *Salmonella*). The treatment may be prophylactic. Oral care products may comprise a polypeptide of the invention having a xylanase activity, or a xylanase encoded by a nucleic acid of the invention. The oral care product can comprise a toothpaste, a dental cream, a gel or a tooth powder, an odontic, a mouth wash, a pre- or post brushing rinse formulation, a chewing gum, a lozenge or a candy. Contact lens cleaning compositions may comprise a polypeptide of the invention having a xylanase activity, or a xylanase encoded by a nucleic acid of the invention.

[0134] Chimeric xylanases may comprise a polypeptide sequence of the invention and at least one heterologous carbohydrate-binding module (CBM), wherein optionally the CBM comprises a CBM3a,

CBM3b, CBM4, CBM6, CBM22 or X14, or a CBM as listed and discussed, below. Chimeric xylanases may comprise at least one heterologous carbohydrate-binding module (CBM), wherein the CBM comprises a carbohydrate-binding subsequence of a xylanase sequence of the invention, or a carbohydrate-binding subsequence comprising a X14. Methods for designing a chimeric xylanase having a new carbohydrate-binding specificity or an enhanced carbohydrate-binding specificity, comprise inserting a heterologous or an additional endogenous carbohydrate-binding module (CBM) into a xylanase, wherein the CBM comprises a carbohydrate-binding subsequence of a xylanase sequence of the invention, or a carbohydrate-binding subsequence comprising a X14, or alternatively a heterologous CBM, or an additional endogenous CBM, is inserted into a xylanase sequence of the invention.

[0135] The invention provides enzyme mixtures, or "cocktails" comprising at least one enzyme of the invention and one or more other enzyme(s), which can be another xylanase, a mannanase or glucanase, or any other enzyme; for example, the "cocktails" of the invention, in addition to at least one enzyme of this invention, can comprise any other enzyme, such as xylanase (not of this invention), cellulases, lipases, esterases, proteases, or endoglycosidases, endo-beta.-1,4-glucanases, beta-glucanases, endo-beta-1,3(4)-glucanases, cutinases, peroxidases, catalases, laccases, amylases, glucoamylases, pectinases, reductases, oxidases, phenoloxidases, ligninases, pullulanases, arabinanases, hemicellulases, mannanases, xyloglucanases, xylanases, pectin acetyl esterases, rhamnogalacturonan acetyl esterases, polygalacturonases, rhamnogalacturonases, galactanases, pectin lyases, pectin methylesterases, cellobiohydrolases and/or transglutaminases, to name just a few examples. In alternative embodiments, these enzyme mixtures, or "cocktails" comprising at least one enzyme of the invention can be used in any process or method described herein, or composition of the disclosure, e.g., in foods or feeds, food or feed supplements, textiles, papers, processed woods, etc. and methods for making them, and in compositions and methods for treating paper, pulp, wood, paper, pulp or wood waste or by-products, and the like, and in the final products thereof.

[0136] Aspects of the invention are set out in the accompanying claims.

[0137] The details of one or more embodiments of the invention are set forth in the accompanying drawings and the description below. Other features, objects, and advantages of the invention will be apparent from the description and drawings, and from the claims.

BRIEF DESCRIPTION OF THE DRAWINGS

[0138] The following drawings are illustrative of aspects of the invention and are not meant to limit the scope of the invention as encompassed by the claims.

[0139] The patent or application file contains at least one drawing executed in color. Copies of this patent or patent application publication with color drawing(s) will be provided by the Office upon request and payment of the necessary fee.

Figure 1 is a block diagram of a computer system.

Figure 2 is a flow diagram illustrating one aspect of a process for comparing a new nucleotide or protein sequence with a database of sequences in order to determine the homology levels between the new sequence and the sequences in the database.

Figure 3 is a flow diagram illustrating one aspect of a process in a computer for determining whether two sequences are homologous.

Figure 4 is a flow diagram illustrating one aspect of an identifier process 300 for detecting the presence of a feature in a sequence.

Figure 5 is a schematic flow diagram of an exemplary routine screening protocol to determine whether a xylanase of the invention is useful in pretreating paper pulp, as described in detail in Example 3, below.

Figure 6 illustrates a biobleaching industrial process of the invention, as described in detail in Example 5, below.

[0140] Like reference symbols in the various drawings indicate like elements.

DETAILED DESCRIPTION OF THE INVENTION

[0141] The invention provides xylanases (e.g., endoxylanases) and polynucleotides encoding them and methods of making and using them. Xylanase activity of the polypeptides of the invention encompasses enzymes capable of hydrolyzing glycosidic linkages in a polysaccharide, for example a glycosidic linkage present in xylan, e.g., catalyzing hydrolysis of internal β -1,4-xylosidic linkages. The xylanases of the invention can be used to make and/or process foods, feeds, nutritional supplements, textiles, detergents and the like. The xylanases of the invention can be used in pharmaceutical compositions and dietary aids.

[0142] Xylanases of the invention are particularly useful in baking, animal feed, beverage and wood, wood pulp, Kraft pulp, paper, paper product or paper pulp processes. In one aspect, an enzyme of the invention is thermotolerant and/or tolerant of high and/or low pH conditions. For example, in one aspect, a xylanase of the invention retains activity under conditions comprising a temperature of at least about 80°C, 85°C, 86°C, 87°C, 88°C, 89°C, 90°C, 91°C, 92°C, 93°C, 94°C, 95°C, 96°C, 97°C, 98°C, 99°C, 100°C, 101°C, 102°C, 103°C, 103.5°C, 104°C, 105°C, 107°C, 108°C, 109°C or 110°C, or more, and a basic pH of at least about pH 11, or more.

[0143] The invention provides isolated, synthetic or recombinant nucleic acids comprising a nucleic acid encoding at least one polypeptide having a xylanase (e.g., an endoxylanase) activity, or other activity as described herein, wherein the nucleic acid comprises a sequence having at least 95% to 99%, or more, or complete (100%) sequence identity (homology) to a sequence which is SEQ ID NO:1 having one or more nucleotide residue changes (modifications, mutations) a sequences which is as described herein. Nucleic acids of the invention includes those encoding a polypeptide of this invention, which includes, e.g., SEQ ID NO:2 having one or more amino acid residue changes (mutations) as described herein.

Table 1:

Clone Name (Xyl 111 and Xyl 11 mutants)	Residue	Original Codon	Changed To	Other Codons for same AA change	original AA	changed to AA	Average A560	Average WT-A560	Average % of WT
Xyl 11	WT							100	100
Xyl 11 mutant 1	108	TTC	AAG	AAA	F	K	0.655	0.740	87.5
Xyl 11 mutant 2	21	TTC	TAC	TAT	F	Y	0.815	0.813	100.3

Clone Name (Xyl 111 and Xyl 11 mutants)	Residue	Original Codon	Changed To	Other Codons for same AA change	original AA	changed to AA	Average A560	Average WT-A560	Average % of WT
18	4	ACC	AAC	AAT	T	N	1.092	0.881	123.7

[0144] Variants of polynucleotides or polypeptides of the invention, may comprise sequences modified at one or more base pairs, codons, introns, exons, or amino acid residues (respectively) yet still retain the biological activity of a xylanase of the invention. Variants can be produced by any number of means included methods such as, for example, error-prone PCR, shuffling, oligonucleotide-directed mutagenesis, assembly PCR, sexual PCR mutagenesis, *in vivo* mutagenesis, cassette mutagenesis, recursive ensemble mutagenesis, exponential ensemble mutagenesis, site-specific mutagenesis, gene reassembly (e.g., GeneReassembly, see, e.g., U.S. Patent No. 6,537,776), GSSM and any combination thereof.

[0145] The term "saturation mutagenesis", "gene site saturation mutagenesis" or "GSSM" includes a method that uses degenerate oligonucleotide primers to introduce point mutations into a polynucleotide, as described in detail, below.

[0146] The term "optimized directed evolution system" or "optimized directed evolution" includes a method for reassembling fragments of related nucleic acid sequences, e.g., related genes, and explained in detail, below.

[0147] The term "synthetic ligation reassembly" or "SLR" includes a method of ligating oligonucleotide fragments in a non-stochastic fashion, and explained in detail, below.

Generating and Manipulating Nucleic Acids

[0148] The invention provides nucleic acids as disclosed herein. The disclosure also includes methods for discovering new xylanase sequences using the nucleic acids of the invention. Methods for inhibiting the expression of xylanase genes, transcripts and polypeptides may use the nucleic acids of the invention. Methods for modifying the nucleic acids of the invention may be by, e.g., synthetic ligation reassembly, optimized directed evolution system and/or saturation mutagenesis.

[0149] The nucleic acids of the invention can be made, isolated and/or manipulated by, e.g., cloning and expression of cDNA libraries, amplification of message or genomic DNA by PCR, and the like.

[0150] Homologous genes can be modified by manipulating a template nucleic acid, as described herein. This can be practiced in conjunction with any method or protocol or device known in the art, which are well described in the scientific and patent literature.

[0151] One aspect of the invention is an isolated nucleic acid as disclosed herein. The isolated nucleic acids may comprise DNA, including cDNA, genomic DNA and synthetic DNA. The DNA may be double-stranded or single-stranded and if single stranded may be the coding strand or non-coding (anti-sense) strand. Alternatively, the isolated nucleic acids may comprise RNA.

[0152] Accordingly, another aspect of the invention is an isolated nucleic acid which encodes one of the polypeptides of the invention, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids of one of the polypeptides of the invention. The coding sequences of these nucleic acids may be identical to one of the coding sequences of one of the nucleic acids of the invention, or a fragment thereof or may be different coding sequences which encode one of the polypeptides of the invention, sequences substantially identical thereto and fragments having at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids of one of the polypeptides of the invention, as a result of the redundancy or degeneracy of the genetic code. The genetic code is well known to those of skill in the art and can be obtained, for example, on page 214 of B. Lewin, *Genes VI*, Oxford University Press, 1997.

[0153] The isolated nucleic acid which encodes one of the polypeptides of the invention and sequences substantially identical thereto, may include, but is not limited to: only the coding sequence of a nucleic acid of the invention and sequences substantially identical thereto and additional coding sequences, such as leader sequences or proprotein sequences and non-coding sequences, such as introns or non-coding sequences 5' and/or 3' of the coding sequence. Thus, as used herein, the term "polynucleotide encoding a polypeptide" encompasses a polynucleotide which includes only the coding sequence for the polypeptide as well as a polynucleotide which includes additional coding and/or non-coding sequence.

[0154] Alternatively, the nucleic acid sequences of the invention and sequences substantially identical thereto, may be mutagenized using conventional techniques, such as site directed mutagenesis, or other techniques familiar to those skilled in the art, to introduce silent changes into the polynucleotides of the invention and sequences substantially identical thereto. As used herein, "silent changes" include, for example, changes which do not alter the amino acid sequence encoded by the polynucleotide. Such changes may be desirable in order to increase the level of the polypeptide produced by host cells containing a vector encoding the polypeptide by introducing codons or codon pairs which occur frequently in the host organism.

[0155] The invention also relates to polynucleotides which have nucleotide changes which result in amino acid substitutions, additions, deletions, fusions and truncations in the polypeptides of the invention and sequences substantially identical thereto. Such nucleotide changes may be introduced using techniques such as site directed mutagenesis, random chemical mutagenesis, exonuclease III deletion and other recombinant DNA techniques. Alternatively, such nucleotide changes may be naturally occurring allelic variants which are isolated by identifying nucleic acids which specifically hybridize to probes comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive bases of one of the sequences of The invention and sequences substantially identical thereto (or the sequences complementary thereto) under conditions of high, moderate, or low stringency as provided herein.

General Techniques

[0156] The nucleic acids used to practice this invention, whether RNA, iRNA, antisense nucleic acid, cDNA, genomic DNA, vectors, viruses or hybrids thereof, may be isolated from a variety of sources, genetically engineered, amplified, and/or expressed/ generated recombinantly. Recombinant polypeptides (e.g., glycosyl hydrolases of the invention) generated from these nucleic acids can be individually isolated or cloned and tested for a desired activity. Any recombinant expression system can be used, including bacterial, mammalian, yeast, insect or plant cell expression systems.

[0157] Alternatively, these nucleic acids can be synthesized *in vitro* by well-known chemical synthesis techniques, as described in, e.g., Adams (1983) *J. Am. Chem. Soc.* 105:661; Belousov (1997) *Nucleic*

Acids Res. 25:3440-3444; Frenkel (1995) Free Radic. Biol. Med. 19:373-380; Blommers (1994) Biochemistry 33:7886-7896; Narang (1979) Meth. Enzymol. 68:90; Brown (1979) Meth. Enzymol. 68:109; Beaucage (1981) Tetra. Lett. 22:1859; U.S. Patent No. 4,458,066.

[0158] Techniques for the manipulation of nucleic acids, such as, e.g., subcloning, labeling probes (e.g., random-primer labeling using Klenow polymerase, nick translation, amplification), sequencing, hybridization and the like are well described in the scientific and patent literature, see, e.g., Sambrook, ed., MOLECULAR CLONING: A LABORATORY MANUAL (2ND ED.), Vols. 1-3, Cold Spring Harbor Laboratory, (1989); CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, Ausubel, ed. John Wiley & Sons, Inc., New York (1997); LABORATORY TECHNIQUES IN BIOCHEMISTRY AND MOLECULAR BIOLOGY: HYBRIDIZATION WITH NUCLEIC ACID PROBES, Part I. Theory and Nucleic Acid Preparation, Tijssen, ed. Elsevier, N.Y. (1993).

[0159] Another useful means of obtaining and manipulating nucleic acids used to practice the methods of the invention is to clone from genomic samples, and, if desired, screen and re-clone inserts isolated or amplified from, e.g., genomic clones or cDNA clones. Sources of nucleic acid used in the methods of the invention include genomic or cDNA libraries contained in, e.g., mammalian artificial chromosomes (MACs), see, e.g., U.S. Patent Nos. 5,721,118; 6,025,155; human artificial chromosomes, see, e.g., Rosenfeld (1997) Nat. Genet. 15:333-335; yeast artificial chromosomes (YAC); bacterial artificial chromosomes (BAC); P1 artificial chromosomes, see, e.g., Woon (1998) Genomics 50:306-316; P1-derived vectors (PACs), see, e.g., Kern (1997) Biotechniques 23:120-124; cosmids, recombinant viruses, phages or plasmids.

[0160] In one aspect, a nucleic acid encoding a polypeptide of the invention is assembled in appropriate phase with a leader sequence capable of directing secretion of the translated polypeptide or fragment thereof.

[0161] The invention provides fusion proteins and nucleic acids encoding them. A polypeptide of the invention can be fused to a heterologous peptide or polypeptide, such as N-terminal identification peptides which impart desired characteristics, such as increased stability or simplified purification. Peptides and polypeptides of the invention can also be synthesized and expressed as fusion proteins with one or more additional domains linked thereto for, e.g., producing a more immunogenic peptide, to more readily isolate a recombinantly synthesized peptide, to identify and isolate antibodies and antibody-expressing B cells, and the like. Detection and purification facilitating domains include, e.g., metal chelating peptides such as polyhistidine tracts and histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp, Seattle WA). The inclusion of a cleavable linker sequences such as Factor Xa or enterokinase (Invitrogen, San Diego CA) between a purification domain and the motif-comprising peptide or polypeptide to facilitate purification. For example, an expression vector can include an epitope-encoding nucleic acid sequence linked to six histidine residues followed by a thioredoxin and an enterokinase cleavage site (see e.g., Williams (1995) Biochemistry 34:1787-1797; Dobeli (1998) Protein Expr. Purif. 12:404-414). The histidine residues facilitate detection and purification while the enterokinase cleavage site provides a means for purifying the epitope from the remainder of the fusion protein. Technology pertaining to vectors encoding fusion proteins and application of fusion proteins are well described in the scientific and patent literature, see e.g., Kroll (1993) DNA Cell. Biol., 12:441-53.

[0162] The phrases "nucleic acid" or "nucleic acid sequence" as used herein refer to an oligonucleotide, nucleotide, polynucleotide, or to a fragment of any of these, to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent a sense or antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material, natural or synthetic in origin. The

phrases "nucleic acid" or "nucleic acid sequence" includes oligonucleotide, nucleotide, polynucleotide, or to a fragment of any of these, to DNA or RNA (e.g., mRNA, rRNA, tRNA, iRNA) of genomic or synthetic origin which may be single-stranded or double-stranded and may represent a sense or antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material, natural or synthetic in origin, including, e.g., iRNA, ribonucleoproteins (e.g., e.g., double stranded iRNAs, e.g., iRNPs). The term encompasses nucleic acids, i.e., oligonucleotides, containing known analogues of natural nucleotides. The term also encompasses nucleic-acid-like structures with synthetic backbones, see e.g., Mata (1997) *Toxicol. Appl. Pharmacol.* 144:189-197; Strauss-Soukup (1997) *Biochemistry* 36:8692-8698; Samstag (1996) *Antisense Nucleic Acid Drug Dev* 6:153-156. "Oligonucleotide" includes either a single stranded polydeoxynucleotide or two complementary polydeoxynucleotide strands that may be chemically synthesized. Such synthetic oligonucleotides have no 5' phosphate and thus will not ligate to another oligonucleotide without adding a phosphate with an ATP in the presence of a kinase. A synthetic oligonucleotide can ligate to a fragment that has not been dephosphorylated.

[0163] A "coding sequence of" or a "nucleotide sequence encoding" a particular polypeptide or protein, is a nucleic acid sequence which is transcribed and translated into a polypeptide or protein when placed under the control of appropriate regulatory sequences.

[0164] The term "gene" means the segment of DNA involved in producing a polypeptide chain; it includes regions preceding and following the coding region (leader and trailer) as well as, where applicable, intervening sequences (introns) between individual coding segments (exons). "Operably linked" as used herein refers to a functional relationship between two or more nucleic acid (e.g., DNA) segments. Typically, it refers to the functional relationship of transcriptional regulatory sequence to a transcribed sequence. For example, a promoter is operably linked to a coding sequence, such as a nucleic acid of the invention, if it stimulates or modulates the transcription of the coding sequence in an appropriate host cell or other expression system. Generally, promoter transcriptional regulatory sequences that are operably linked to a transcribed sequence are physically contiguous to the transcribed sequence, i.e., they are cis-acting. However, some transcriptional regulatory sequences, such as enhancers, need not be physically contiguous or located in close proximity to the coding sequences whose transcription they enhance.

[0165] The term "expression cassette" as used herein refers to a nucleotide sequence which is capable of affecting expression of a structural gene (i.e., a protein coding sequence, such as a xylanase of the invention) in a host compatible with such sequences. Expression cassettes include at least a promoter operably linked with the polypeptide coding sequence; and, in one aspect, with other sequences, e.g., transcription

termination signals. Additional factors necessary or helpful in effecting expression may also be used, e.g., enhancers. Thus, expression cassettes also include plasmids, expression vectors, recombinant viruses, any form of recombinant "naked DNA" vector, and the like. A "vector" comprises a nucleic acid that can infect, transfect, transiently or permanently transduce a cell. It will be recognized that a vector can be a naked nucleic acid, or a nucleic acid complexed with protein or lipid. The vector in one aspect comprises viral or bacterial nucleic acids and/or proteins, and/or membranes (e.g., a cell membrane, a viral lipid envelope, etc.). Vectors include, but are not limited to replicons (e.g., RNA replicons, bacteriophages) to which fragments of DNA may be attached and become replicated. Vectors thus include, but are not limited to RNA, autonomous self-replicating circular or linear DNA or RNA (e.g., plasmids, viruses, and the like, see, e.g., U.S. Patent No. 5,217,879), and include both the expression and non-expression plasmids. Where a recombinant microorganism or cell culture is described as hosting an "expression vector" this includes both extra-chromosomal circular and linear DNA and DNA that has been incorporated into the host chromosome(s). Where a vector is being maintained by a host cell, the vector may either be stably replicated by the cells during mitosis as an autonomous structure, or is incorporated within the host's

genome.

[0166] As used herein, the term "promoter" includes all sequences capable of driving transcription of a coding sequence in a cell, e.g., a plant cell. Thus, promoters used in the constructs of the invention include cis-acting transcriptional control elements and regulatory sequences that are involved in regulating or modulating the timing and/or rate of transcription of a gene. For example, a promoter can be a cis-acting transcriptional control element, including an enhancer, a promoter, a transcription terminator, an origin of replication, a chromosomal integration sequence, 5' and 3' untranslated regions, or an intronic sequence, which are involved in transcriptional regulation. These cis-acting sequences typically interact with proteins or other biomolecules to carry out (turn on/off, regulate, modulate, etc.) transcription. "Constitutive" promoters are those that drive expression continuously under most environmental conditions and states of development or cell differentiation. "Inducible" or "regulatable" promoters direct expression of the nucleic acid of the invention under the influence of environmental conditions or developmental conditions. Examples of environmental conditions that may affect transcription by inducible promoters include anaerobic conditions, elevated temperature, drought, or the presence of light.

[0167] "Tissue-specific" promoters are transcriptional control elements that are only active in particular cells or tissues or organs, e.g., in plants or animals. Tissue-specific regulation may be achieved by certain intrinsic factors that ensure that genes encoding proteins specific to a given tissue are expressed. Such factors are known to exist in mammals and plants so as to allow for specific tissues to develop.

[0168] As used herein, the term "isolated" means that the material (e.g., a nucleic acid, a polypeptide, a cell) is removed from its original environment (e.g., the natural environment if it is naturally occurring). For example, a naturally-occurring polynucleotide or polypeptide present in a living animal is not isolated, but the same polynucleotide or polypeptide, separated from some or all of the coexisting materials in the natural system, is isolated. Such polynucleotides could be part of a vector and/or such polynucleotides or polypeptides could be part of a composition and still be isolated in that such vector or composition is not part of its natural environment. As used herein, the term "purified" does not require absolute purity; rather, it is intended as a relative definition. Individual nucleic acids obtained from a library have been conventionally purified to electrophoretic homogeneity. The sequences obtained from these clones could not be obtained directly either from the library or from total human DNA. The purified nucleic acids of the invention have been purified from the remainder of the genomic DNA in the organism by at least 10^4 - 10^6 fold. However, the term "purified" also includes nucleic acids that have been purified from the remainder of the genomic DNA or from other sequences in a library or other environment by at least one order of magnitude, typically two or three orders and more typically four or five orders of magnitude.

[0169] As used herein, the term "recombinant" means that the nucleic acid is adjacent to a "backbone" nucleic acid to which it is not adjacent in its natural environment. Additionally, to be "enriched" the nucleic acids will represent 5% or more of the number of nucleic acid inserts in a population of nucleic acid backbone molecules. Backbone molecules according to the invention include nucleic acids such as expression vectors, self-replicating nucleic acids, viruses, integrating nucleic acids and other vectors or nucleic acids used to maintain or manipulate a nucleic acid insert of interest. Typically, the enriched nucleic acids represent 15% or more of the number of nucleic acid inserts in the population of recombinant backbone molecules. More typically, the enriched nucleic acids represent 50% or more of the number of nucleic acid inserts in the population of recombinant backbone molecules. In a one aspect, the enriched nucleic acids represent 90% or more of the number of nucleic acid inserts in the population of recombinant backbone molecules.

[0170] "Plasmids" are designated by a lower case "p" preceded and/or followed by capital letters and/or

numbers. The starting plasmids herein are either commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accord with published procedures. In addition, equivalent plasmids to those described herein are known in the art and will be apparent to the ordinarily skilled artisan. "Plasmids" can be commercially available, publicly available on an unrestricted basis, or can be constructed from available plasmids in accord with published procedures. Equivalent plasmids to those described herein are known in the art and will be apparent to the ordinarily skilled artisan.

[0171] "Digestion" of DNA refers to catalytic cleavage of the DNA with a restriction enzyme that acts only at certain sequences in the DNA. The various restriction enzymes used herein are commercially available and their reaction conditions, cofactors and other requirements were used as would be known to the ordinarily skilled artisan. For analytical purposes, typically 1 µg of plasmid or DNA fragment is used with about 2 units of enzyme in about 20 µl of buffer solution. For the purpose of isolating DNA fragments for plasmid construction, typically 5 to 50 µg of DNA are digested with 20 to 250 units of enzyme in a larger volume. Appropriate buffers and substrate amounts for particular restriction enzymes are specified by the manufacturer. Incubation times of about 1 hour at 37°C are ordinarily used, but may vary in accordance with the supplier's instructions. After digestion, gel electrophoresis may be performed to isolate the desired fragment.

[0172] "Hybridization" refers to the process by which a nucleic acid strand joins with a complementary strand through base pairing. Hybridization reactions can be sensitive and selective so that a particular sequence of interest can be identified even in samples in which it is present at low concentrations. Suitably stringent conditions can be defined by, for example, the concentrations of salt or formamide in the prehybridization and hybridization solutions, or by the hybridization temperature and are well known in the art. In particular, stringency can be increased by reducing the concentration of salt, increasing the concentration of formamide, or raising the hybridization temperature. In alternative aspects, nucleic acids of the invention are defined by their ability to hybridize under various stringency conditions (e.g., high, medium, and low), as set forth herein.

[0173] For example, hybridization under high stringency conditions could occur in about 50% formamide at about 37°C to 42°C. Hybridization could occur under reduced stringency conditions in about 35% to 25% formamide at about 30°C to 35°C. In particular, hybridization could occur under high stringency conditions at 42°C in 50% formamide, 5X SSPE, 0.3% SDS and 200 µg/ml sheared and denatured salmon sperm DNA. Hybridization could occur under reduced stringency conditions as described above, but in 35% formamide at a reduced temperature of 35°C. The temperature range corresponding to a particular level of stringency can be further narrowed by calculating the purine to pyrimidine ratio of the nucleic acid of interest and adjusting the temperature accordingly. Variations on the above ranges and conditions are well known in the art.

Transcriptional and translational control sequences

[0174] The invention provides nucleic acid (e.g., DNA) sequences of the invention operatively linked to expression (e.g., transcriptional or translational) control sequence(s), e.g., promoters or enhancers, to direct or modulate RNA synthesis/ expression. The expression control sequence can be in an expression vector. Exemplary bacterial promoters include *lacI*, *lacZ*, T3, T7, *gpt*, *lambda PR*, *PL* and *trp*. Exemplary eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus, and mouse metallothionein I. A promoter sequence is "operably linked to" a coding sequence when RNA polymerase which initiates transcription at the promoter will transcribe the coding sequence into

mRNA. Promoters suitable for expressing a polypeptide in bacteria include the *E. coli* lac or trp promoters, the lacI promoter, the lacZ promoter, the T3 promoter, the T7 promoter, the gpt promoter, the lambda PR promoter, the lambda PL promoter, promoters from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK), and the acid phosphatase promoter. Eukaryotic promoters include the CMV immediate early promoter, the HSV thymidine kinase promoter, heat shock promoters, the early and late SV40 promoter, LTRs from retroviruses, and the mouse metallothionein-I promoter. Other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses may also be used. Promoters suitable for expressing the polypeptide or fragment thereof in bacteria include the *E. coli* lac or trp promoters, the lacI promoter, the lacZ promoter, the T3 promoter, the T7 promoter, the gpt promoter, the lambda P_R promoter, the lambda P_L promoter, promoters from operons encoding glycolytic enzymes such as 3-phosphoglycerate kinase (PGK) and the acid phosphatase promoter. Fungal promoters include the ν factor promoter. Eukaryotic promoters include the CMV immediate early promoter, the HSV thymidine kinase promoter, heat shock promoters, the early and late SV40 promoter, LTRs from retroviruses and the mouse metallothionein-I promoter. Other promoters known to control expression of genes in prokaryotic or eukaryotic cells or their viruses may also be used.

Tissue-Specific Plant Promoters

[0175] The invention provides expression cassettes that can be expressed in a tissue-specific manner, e.g., that can express a xylanase of the invention in a tissue-specific manner. The invention also provides plants or seeds that express a xylanase of the invention in a tissue-specific manner. The tissue-specificity can be seed specific, stem specific, leaf specific, root specific, fruit specific and the like.

[0176] In one aspect, a constitutive promoter such as the CaMV 35S promoter can be used for expression in specific parts of the plant or seed or throughout the plant. For example, for overexpression, a plant promoter fragment can be employed which will direct expression of a nucleic acid in some or all tissues of a plant, e.g., a regenerated plant. Such promoters are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the cauliflower mosaic virus (CaMV) 35S transcription initiation region, the 1'- or 2'- promoter derived from T-DNA of *Agrobacterium tumefaciens*, and other transcription initiation regions from various plant genes known to those of skill. Such genes include, e.g., *ACT11* from *Arabidopsis* (Huang (1996) Plant Mol. Biol. 33:125-139); *Cat3* from *Arabidopsis* (GenBank No. U43147, Zhong (1996) Mol. Gen. Genet. 251:196-203); the gene encoding stearyl-acyl carrier protein desaturase from *Brassica napus* (Genbank No. X74782, Solomon (1994) Plant Physiol. 104:1167-1176); *Gpc1* from maize (GenBank No. X15596; Martinez (1989) J. Mol. Biol. 208:551-565); the *Gpc2* from maize (GenBank No. U45855, Manjunath (1997) Plant Mol. Biol. 33:97-112); plant promoters described in U.S. Patent Nos. 4,962,028; 5,633,440.

[0177] The invention uses tissue-specific or constitutive promoters derived from viruses which can include, e.g., the tobamovirus subgenomic promoter (Kumagai (1995) Proc. Natl. Acad. Sci. USA 92:1679-1683; the rice tungro bacilliform virus (RTBV), which replicates only in phloem cells in infected rice plants, with its promoter which drives strong phloem-specific reporter gene expression; the cassava vein mosaic virus (CVMV) promoter, with highest activity in vascular elements, in leaf mesophyll cells, and in root tips (Verdaguer (1996) Plant Mol. Biol. 31:1129-1139).

[0178] Alternatively, the plant promoter may direct expression of xylanase-expressing nucleic acid in a specific tissue, organ or cell type (*i.e.* tissue-specific promoters) or may be otherwise under more precise environmental or developmental control or under the control of an inducible promoter. Examples of

environmental conditions that may affect transcription include anaerobic conditions, elevated temperature, the presence of light, or sprayed with chemicals/hormones. For example, the invention incorporates the drought-inducible promoter of maize (Busk (1997) *supra*); the cold, drought, and high salt inducible promoter from potato (Kirch (1997) *Plant Mol. Biol.* 33:897-909).

[0179] Tissue-specific promoters can promote transcription only within a certain time frame of developmental stage within that tissue. See, e.g., Blazquez (1998) *Plant Cell* 10:791-800, characterizing the *Arabidopsis* LEAFY gene promoter. See also Cardon (1997) *Plant J* 12:367-77, describing the transcription factor SPL3, which recognizes a conserved sequence motif in the promoter region of the *A. thaliana* floral meristem identity gene AP1; and Mandel (1995) *Plant Molecular Biology*, Vol. 29, pp 995-1004, describing the meristem promoter eIF4. Tissue specific promoters which are active throughout the life cycle of a particular tissue can be used. In one aspect, the nucleic acids of the invention are operably linked to a promoter active primarily only in cotton fiber cells. In one aspect, the nucleic acids of the invention are operably linked to a promoter active primarily during the stages of cotton fiber cell elongation, e.g., as described by Rinehart (1996) *supra*. The nucleic acids can be operably linked to the FbI2A gene promoter to be preferentially expressed in cotton fiber cells (*Ibid*). See also, John (1997) *Proc. Natl. Acad. Sci. USA* 89:5769-5773; John, et al., U.S. Patent Nos. 5,608,148 and 5,602,321, describing cotton fiber-specific promoters and methods for the construction of transgenic cotton plants. Root-specific promoters may also be used to express the nucleic acids of the invention. Examples of root-specific promoters include the promoter from the alcohol dehydrogenase gene (DeLisle (1990) *Int. Rev. Cytol.* 123:39-60). Other promoters that can be used to express the nucleic acids of the invention include, e.g., ovule-specific, embryo-specific, endosperm-specific, integument-specific, seed coat-specific promoters, or some combination thereof; a leaf-specific promoter (see, e.g., Busk (1997) *Plant J.* 11:1285-1295, describing a leaf-specific promoter in maize); the ORF13 promoter from *Agrobacterium rhizogenes* (which exhibits high activity in roots, see, e.g., Hansen (1997) *supra*); a maize pollen specific promoter (see, e.g., Guerrero (1990) *Mol. Gen. Genet.* 224:161-168); a tomato promoter active during fruit ripening, senescence and abscission of leaves and, to a lesser extent, of flowers can be used (see, e.g., Blume (1997) *Plant J.* 12:731-746); a pistil-specific promoter from the potato SK2 gene (see, e.g., Ficker (1997) *Plant Mol. Biol.* 35:425-431); the Blec4 gene from pea, which is active in epidermal tissue of vegetative and floral shoot apices of transgenic alfalfa making it a useful tool to target the expression of foreign genes to the epidermal layer of actively growing shoots or fibers; the ovule-specific BEL1 gene (see, e.g., Reiser (1995) *Cell* 83:735-742, GenBank No. U39944); and/or, the promoter in Klee, U.S. Patent No. 5,589,583, describing a plant promoter region is capable of conferring high levels of transcription in meristematic tissue and/or rapidly dividing cells.

[0180] Alternatively, plant promoters which are inducible upon exposure to plant hormones, such as auxins, are used to express the nucleic acids of the invention. For example, the invention can use the auxin-response elements E1 promoter fragment (AuxREs) in the soybean (*Glycine max* L.) (Liu (1997) *Plant Physiol.* 115:397-407); the auxin-responsive *Arabidopsis* GST6 promoter (also responsive to salicylic acid and hydrogen peroxide) (Chen (1996) *Plant J.* 10: 955-966); the auxin-inducible parC promoter from tobacco (Sakai (1996) 37:906-913); a plant biotin response element (Streit (1997) *Mol. Plant Microbe Interact.* 10:933-937); and, the promoter responsive to the stress hormone abscisic acid (Sheen (1996) *Science* 274:1900-1902).

[0181] The nucleic acids of the invention can also be operably linked to plant promoters which are inducible upon exposure to chemicals reagents which can be applied to the plant, such as herbicides or antibiotics. For example, the maize In2-2 promoter, activated by benzenesulfonamide herbicide safeners, can be used (De Veylder (1997) *Plant Cell Physiol.* 38:568-577); application of different herbicide safeners induces distinct gene expression patterns, including expression in the root, hydathodes, and the shoot apical meristem. Coding sequence can be under the control of, e.g., a tetracycline-inducible promoter,

e.g., as described with transgenic tobacco plants containing the *Avena sativa* L. (oat) arginine decarboxylase gene (Masgrau (1997) Plant J. 11:465-473); or, a salicylic acid-responsive element (Stange (1997) Plant J. 11:1315-1324). Using chemically- (e.g., hormone- or pesticide-) induced promoters, i.e., promoter responsive to a chemical which can be applied to the transgenic plant in the field, expression of a polypeptide of the invention can be induced at a particular stage of development of the plant. Thus, the invention also provides for transgenic plants containing an inducible gene encoding for polypeptides of the invention whose host range is limited to target plant species, such as corn, rice, barley, wheat, potato or other crops, inducible at any stage of development of the crop.

[0182] One of skill will recognize that a tissue-specific plant promoter may drive expression of operably linked sequences in tissues other than the target tissue. Thus, a tissue-specific promoter is one that drives expression preferentially in the target tissue or cell type, but may also lead to some expression in other tissues as well.

[0183] The nucleic acids of the invention can also be operably linked to plant promoters which are inducible upon exposure to chemicals reagents. These reagents include, e.g., herbicides, synthetic auxins, or antibiotics which can be applied, e.g., sprayed, onto transgenic plants. Inducible expression of the xylanase- producing nucleic acids of the invention will allow the grower to select plants with the optimal xylanase expression and/or activity. The development of plant parts can thus controlled. In this way the invention provides the means to facilitate the harvesting of plants and plant parts. For example, in various embodiments, the maize In2-2 promoter, activated by benzenesulfonamide herbicide safeners, is used (De Veylder (1997) Plant Cell Physiol. 38:568-577); application of different herbicide safeners induces distinct gene expression patterns, including expression in the root, hydathodes, and the shoot apical meristem. Coding sequences of the invention are also under the control of a tetracycline-inducible promoter, e.g., as described with transgenic tobacco plants containing the *Avena sativa* L. (oat) arginine decarboxylase gene (Masgrau (1997) Plant J. 11:465-473); or, a salicylic acid-responsive element (Stange (1997) Plant J. 11:1315-1324).

[0184] In some aspects, proper polypeptide expression may require polyadenylation region at the 3'-end of the coding region. The polyadenylation region can be derived from the natural gene, from a variety of other plant (or animal or other) genes, or from genes in the *Agrobacterium* T-DNA.

[0185] The term "plant" (e.g., as in a transgenic plant or plant seed of this invention, or plant promoter used in a vector of the invention) includes whole plants, plant parts (e.g., leaves, stems, flowers, roots, etc.), plant protoplasts, seeds and plant cells and progeny of same; the classes of plants that can be used to practice this invention (including compositions and methods) can be as broad as the class of higher plants, including plants amenable to transformation techniques, including angiosperms (monocotyledonous and dicotyledonous plants), as well as gymnosperms; also including plants of a variety of ploidy levels, including polyploid, diploid, haploid and hemizygous states. As used herein, the term "transgenic plant" includes plants or plant cells into which a heterologous nucleic acid sequence has been inserted, e.g., the nucleic acids and various recombinant constructs (e.g., expression cassettes, such as vectors) of the invention. Transgenic plants of the invention are also discussed, below.

Expression vectors and cloning vehicles

[0186] The invention provides expression vectors and cloning vehicles comprising nucleic acids of the invention, e.g., sequences encoding the xylanases of the invention. Expression vectors and cloning vehicles of the invention can comprise viral particles, baculovirus, phage, plasmids, phagemids, cosmids,

fosmids, bacterial artificial chromosomes, viral DNA (e.g., vaccinia, adenovirus, fowl pox virus, pseudorabies and derivatives of SV40), P1-based artificial chromosomes, yeast plasmids, yeast artificial chromosomes, and any other vectors specific for specific hosts of interest (such as bacillus, *Aspergillus* and yeast). Vectors of the invention can include chromosomal, non-chromosomal and synthetic DNA sequences. Large numbers of suitable vectors are known to those of skill in the art, and are commercially available. Exemplary vectors include: bacterial: pQE vectors (Qiagen), pBluescript plasmids, pNH vectors, (lambda-ZAP vectors (Stratagene); ptrc99a, pKK223-3, pDR540, pRIT2T (Pharmacia); Eukaryotic: pXT1, pSG5 (Stratagene), pSVK3, pBPV, pMSG, pSVLSV40 (Pharmacia). However, any other plasmid or other vector may be used so long as they are replicable and viable in the host. Low copy number or high copy number vectors may be employed with the present invention.

[0187] The expression vector can comprise a promoter, a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression. Mammalian expression vectors can comprise an origin of replication, any necessary ribosome binding sites, a polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences, and 5' flanking non-transcribed sequences. In some aspects, DNA sequences derived from the SV40 splice and polyadenylation sites may be used to provide the required non-transcribed genetic elements.

[0188] In one aspect, the expression vectors contain one or more selectable marker genes to permit selection of host cells containing the vector. Such selectable markers include genes encoding dihydrofolate reductase or genes conferring neomycin resistance for eukaryotic cell culture, genes conferring tetracycline or ampicillin resistance in *E. coli*, and the *S. cerevisiae* TRP1 gene. Promoter regions can be selected from any desired gene using chloramphenicol transferase (CAT) vectors or other vectors with selectable markers.

[0189] Vectors for expressing the polypeptide or fragment thereof in eukaryotic cells can also contain enhancers to increase expression levels. Enhancers are cis-acting elements of DNA, usually from about 10 to about 300 bp in length that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin bp 100 to 270, the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and the adenovirus enhancers.

[0190] A nucleic acid sequence can be inserted into a vector by a variety of procedures. In general, the sequence is ligated to the desired position in the vector following digestion of the insert and the vector with appropriate restriction endonucleases. Alternatively, blunt ends in both the insert and the vector may be ligated. A variety of cloning techniques are known in the art, e.g., as described in Ausubel and Sambrook. Such procedures and others are deemed to be within the scope of those skilled in the art.

[0191] The vector can be in the form of a plasmid, a viral particle, or a phage. Other vectors include chromosomal, non-chromosomal and synthetic DNA sequences, derivatives of SV40; bacterial plasmids, phage DNA, baculovirus, yeast plasmids, vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus, and pseudorabies. A variety of cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by, e.g., Sambrook.

[0192] Particular bacterial vectors which can be used include the commercially available plasmids comprising genetic elements of the well known cloning vector pBR322 (ATCC 37017), pKK223-3 (Pharmacia Fine Chemicals, Uppsala, Sweden), GEM1 (Promega Biotec, Madison, WI, USA) pQE70, pQE60, pQE-9 (Qiagen), pD10, psiX174 pBluescript II KS, pNH8A, pNH16a, pNH18A, pNH46A (Stratagene), ptrc99a, pKK223-3, pKK233-3, DR540, pRIT5 (Pharmacia), pKK232-8 and pCM7. Particular eukaryotic vectors include pSV2CAT, pOG44, pXT1, pSG (Stratagene) pSVK3, pBPV, pMSG, and pSVL

(Pharmacia). However, any other vector may be used as long as it is replicable and viable in the host cell.

[0193] The nucleic acids of the invention can be expressed in expression cassettes, vectors or viruses and transiently or stably expressed in plant cells and seeds. One exemplary transient expression system uses episomal expression systems, e.g., cauliflower mosaic virus (CaMV) viral RNA generated in the nucleus by transcription of an episomal mini-chromosome containing supercoiled DNA, see, e.g., Covey (1990) Proc. Natl. Acad. Sci. USA 87:1633-1637. Alternatively, coding sequences, i.e., all or sub-fragments of sequences of the invention can be inserted into a plant host cell genome becoming an integral part of the host chromosomal DNA. Sense or antisense transcripts can be expressed in this manner. A vector comprising the sequences (e.g., promoters or coding regions) from nucleic acids of the invention can comprise a marker gene that confers a selectable phenotype on a plant cell or a seed. For example, the marker may encode biocide resistance, particularly antibiotic resistance, such as resistance to kanamycin, G418, bleomycin, hygromycin, or herbicide resistance, such as resistance to chlorosulfuron or Basta.

[0194] Expression vectors capable of expressing nucleic acids and proteins in plants are well known in the art, and can include, e.g., vectors from *Agrobacterium* spp., potato virus X (see, e.g., Angell (1997) EMBO J. 16:3675-3684), tobacco mosaic virus (see, e.g., Casper (1996) Gene 173:69-73), tomato bushy stunt virus (see, e.g., Hillman (1989) Virology 169:42-50), tobacco etch virus (see, e.g., Dolja (1997) Virology 234:243-252), bean golden mosaic virus (see, e.g., Morinaga (1993) Microbiol Immunol. 37:471-476), cauliflower mosaic virus (see, e.g., Cecchini (1997) Mol. Plant Microbe Interact. 10:1094-1101), maize Ac/Ds transposable element (see, e.g., Rubin (1997) Mol. Cell. Biol. 17:6294-6302; Kunze (1996) Curr. Top. Microbiol. Immunol. 204:161-194), and the maize suppressor-mutator (Spm) transposable element (see, e.g., Schlappi (1996) Plant Mol. Biol. 32:717-725); and derivatives thereof.

[0195] In one aspect, the expression vector can have two replication systems to allow it to be maintained in two organisms, for example in mammalian or insect cells for expression and in a prokaryotic host for cloning and amplification. Furthermore, for integrating expression vectors, the expression vector can contain at least one sequence homologous to the host cell genome. It can contain two homologous sequences which flank the expression construct. The integrating vector can be directed to a specific locus in the host cell by selecting the appropriate homologous sequence for inclusion in the vector. Constructs for integrating vectors are well known in the art.

[0196] Expression vectors of the invention may also include a selectable marker gene to allow for the selection of bacterial strains that have been transformed, e.g., genes which render the bacteria resistant to drugs such as ampicillin, chloramphenicol, erythromycin, kanamycin, neomycin and tetracycline. Selectable markers can also include biosynthetic genes, such as those in the histidine, tryptophan and leucine biosynthetic pathways.

[0197] The DNA sequence in the expression vector is operatively linked to an appropriate expression control sequence(s) (promoter) to direct RNA synthesis. Particular named bacterial promoters include *lacI*, *lacZ*, *T3*, *T7*, *gpt*, *lambda P_R*, *P_L* and *trp*. Eukaryotic promoters include CMV immediate early, HSV thymidine kinase, early and late SV40, LTRs from retrovirus and mouse metallothionein-I. Selection of the appropriate vector and promoter is well within the level of ordinary skill in the art. The expression vector also contains a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression. Promoter regions can be selected from any desired gene using chloramphenicol transferase (CAT) vectors or other vectors with selectable markers. In addition, the expression vectors preferably contain one or more selectable marker genes to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in *E. coli*.

[0198] Mammalian expression vectors may also comprise an origin of replication, any necessary ribosome binding sites, a polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences and 5' flanking nontranscribed sequences. In some aspects, DNA sequences derived from the SV40 splice and polyadenylation sites may be used to provide the required nontranscribed genetic elements.

[0199] Vectors for expressing the polypeptide or fragment thereof in eukaryotic cells may also contain enhancers to increase expression levels. Enhancers are cis-acting elements of DNA, usually from about 10 to about 300 bp in length that act on a promoter to increase its transcription. Examples include the SV40 enhancer on the late side of the replication origin bp 100 to 270, the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin and the adenovirus enhancers.

[0200] In addition, the expression vectors typically contain one or more selectable marker genes to permit selection of host cells containing the vector. Such selectable markers include genes encoding dihydrofolate reductase or genes conferring neomycin resistance for eukaryotic cell culture, genes conferring tetracycline or ampicillin resistance in *E. coli* and the *S. cerevisiae* *TRP1* gene.

[0201] In some aspects, the nucleic acid encoding one of the polypeptides of the invention and sequences substantially identical thereto, or fragments comprising at least about 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof is assembled in appropriate phase with a leader sequence capable of directing secretion of the translated polypeptide or fragment thereof. The nucleic acid can encode a fusion polypeptide in which one of the polypeptides of the invention and sequences substantially identical thereto, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof is fused to heterologous peptides or polypeptides, such as N-terminal identification peptides which impart desired characteristics, such as increased stability or simplified purification.

[0202] The appropriate DNA sequence may be inserted into the vector by a variety of procedures. In general, the DNA sequence is ligated to the desired position in the vector following digestion of the insert and the vector with appropriate restriction endonucleases. Alternatively, blunt ends in both the insert and the vector may be ligated. A variety of cloning techniques are disclosed in Ausubel et al. Current Protocols in Molecular Biology, John Wiley 503 Sons, Inc. 1997 and Sambrook et al., Molecular Cloning: A Laboratory Manual 2nd Ed., Cold Spring Harbor Laboratory Press (1989). Such procedures and others are deemed to be within the scope of those skilled in the art.

[0203] The vector may be, for example, in the form of a plasmid, a viral particle, or a phage. Other vectors include chromosomal, nonchromosomal and synthetic DNA sequences, derivatives of SV40; bacterial plasmids, phage DNA, baculovirus, yeast plasmids, vectors derived from combinations of plasmids and phage DNA, viral DNA such as vaccinia, adenovirus, fowl pox virus and pseudorabies. A variety of cloning and expression vectors for use with prokaryotic and eukaryotic hosts are described by Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, N.Y., (1989).

Host cells and transformed cells

[0204] The invention also provides transformed cells comprising a nucleic acid sequence of the invention or a vector of the invention. The host cell may be any of the host cells familiar to those skilled in the art, including prokaryotic cells, eukaryotic cells, such as bacterial cells, fungal cells, yeast cells, mammalian cells, insect cells, or plant cells. Exemplary bacterial cells include any species within the genera *Escherichia*, *Bacillus*, *Streptomyces*, *Salmonella*, *Pseudomonas* and *Staphylococcus*, including, e.g.,

Escherichia coli, *Lactococcus lactis*, *Bacillus subtilis*, *Bacillus cereus*, *Salmonella typhimurium*, *Pseudomonas fluorescens*. Exemplary fungal cells include any species of *Aspergillus*. Exemplary yeast cells include any species of *Pichia*, *Saccharomyces*, *Schizosaccharomyces*, or *Schwanniomyces*, including *Pichia pastoris*, *Saccharomyces cerevisiae*, or *Schizosaccharomyces pombe*. Exemplary insect cells include any species of *Spodoptera* or *Drosophila*, including *Drosophila S2* and *Spodoptera Sf9*. Exemplary animal cells include CHO, COS or Bowes melanoma or any mouse or human cell line. The selection of an appropriate host is within the abilities of those skilled in the art. Techniques for transforming a wide variety of higher plant species are well known and described in the technical and scientific literature. See, e.g., Weising (1988) Ann. Rev. Genet. 22:421-477; U.S. Patent No. 5,750,870.

[0205] The vector can be introduced into the host cells using any of a variety of techniques, including transformation, transfection, transduction, viral infection, gene guns, or Timediated gene transfer. Particular methods include calcium phosphate transfection, DEAE-Dextran mediated transfection, lipofection, or electroporation (Davis, L., Dibner, M., Battey, I., Basic Methods in Molecular Biology, (1986)).

[0206] In one aspect, the nucleic acids or vectors of the invention are introduced into the cells for screening, thus, the nucleic acids enter the cells in a manner suitable for subsequent expression of the nucleic acid. The method of introduction is largely dictated by the targeted cell type. Exemplary methods include CaPO_4 precipitation, liposome fusion, lipofection (e.g., LIPOFECTIN™), electroporation, viral infection, etc. The candidate nucleic acids may stably integrate into the genome of the host cell (for example, with retroviral introduction) or may exist either transiently or stably in the cytoplasm (i.e. through the use of traditional plasmids, utilizing standard regulatory sequences, selection markers, etc.). As many pharmaceutically important screens require human or model mammalian cell targets, retroviral vectors capable of transfecting such targets are can be used.

[0207] Where appropriate, the engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the genes of the invention. Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter may be induced by appropriate means (e.g., temperature shift or chemical induction) and the cells may be cultured for an additional period to allow them to produce the desired polypeptide or fragment thereof.

[0208] Cells can be harvested by centrifugation, disrupted by physical or chemical means, and the resulting crude extract is retained for further purification. Microbial cells employed for expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents. Such methods are well known to those skilled in the art. The expressed polypeptide or fragment thereof can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the polypeptide. If desired, high performance liquid chromatography (HPLC) can be employed for final purification steps.

[0209] The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Depending upon the host employed in a recombinant production procedure, the polypeptides produced by host cells containing the vector may be glycosylated or may be non-glycosylated. Polypeptides of the invention may or may not also include an initial methionine amino acid residue.

[0210] Cell-free translation systems can also be employed to produce a polypeptide of the invention. Cell-free translation systems can use mRNAs transcribed from a DNA construct comprising a promoter operably linked to a nucleic acid encoding the polypeptide or fragment thereof. In some aspects, the DNA construct may be linearized prior to conducting an in vitro transcription reaction. The transcribed mRNA is then incubated with an appropriate cell-free translation extract, such as a rabbit reticulocyte extract, to produce the desired polypeptide or fragment thereof.

[0211] The expression vectors can contain one or more selectable marker genes to provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture, or such as tetracycline or ampicillin resistance in *E. coli*.

[0212] Host cells containing the polynucleotides of interest, e.g., nucleic acids of the invention, can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying genes. The culture conditions, such as temperature, pH and the like, are those previously used with the host cell selected for expression and will be apparent to the ordinarily skilled artisan. The clones which are identified as having the specified enzyme activity may then be sequenced to identify the polynucleotide sequence encoding an enzyme having the enhanced activity.

[0213] The invention provides a method for overexpressing a recombinant xylanase, in a cell comprising expressing a vector comprising a nucleic acid of the invention, or, a nucleic acid that hybridizes under stringent conditions to a nucleic acid sequence of the invention, or a subsequence thereof. The overexpression can be effected by any means, e.g., use of a high activity promoter, a dicistronic vector or by gene amplification of the vector.

[0214] The nucleic acids of the invention can be expressed, or overexpressed, in any in vitro or in vivo expression system. Any cell culture systems can be employed to express, or overexpress, recombinant protein, including bacterial, insect, yeast, fungal or mammalian cultures. Over-expression can be effected by appropriate choice of promoters, enhancers, vectors (e.g., use of replicon vectors, dicistronic vectors (see, e.g., Gurtu (1996) Biochem. Biophys. Res. Commun. 229:295-8), media, culture systems and the like. In one aspect, gene amplification using selection markers, e.g., glutamine synthetase (see, e.g., Sanders (1987) Dev. Biol. Stand. 66:55-63), in cell systems are used to overexpress the polypeptides of the invention.

[0215] Additional details regarding this approach are in the public literature and/or are known to the skilled artisan. In a particular non-limiting exemplification, such publicly available literature includes EP 0659215 (WO 9403612 A1) (Nevalainen et al.); Lapidot, A., Mechaly, A., Shoham, Y., "Overexpression and single-step purification of a thermostable xylanase from *Bacillus stearothermophilus* T-6," J. Biotechnol. Nov 51:259-64 (1996); Lüthi, E., Jasmat, N.B., Bergquist, P.L., "Xylanase from the extremely thermophilic bacterium *Caldocellum saccharolyticum*: overexpression of the gene in *Escherichia coli* and characterization of the gene product," Appl. Environ. Microbiol. Sep 56:2677-83 (1990); and Sung, W.L., Luk, C.K., Zahab, D.M., Wakarchuk, W., "Overexpression of the *Bacillus subtilis* and *circulans* xylanases in *Escherichia coli*," Protein Expr. Purif. Jun 4:200-6 (1993), although these references do not teach the inventive enzymes of the instant application.

[0216] The host cell may be any of the host cells familiar to those skilled in the art, including prokaryotic cells, eukaryotic cells, mammalian cells, insect cells, or plant cells. As representative examples of appropriate hosts, there may be mentioned: bacterial cells, such as *E. coli*, *Streptomyces*, *Bacillus subtilis*, *Bacillus cereus*, *Salmonella typhimurium* and various species within the genera *Pseudomonas*, *Streptomyces* and *Staphylococcus*, fungal cells, such as *Aspergillus*, yeast such as any species of *Pichia*, *Saccharomyces*, *Schizosaccharomyces*, *Schwanniomyces*, including *Pichia pastoris*, *Saccharomyces*

cerevisiae, or *Schizosaccharomyces pombe*, insect cells such as *Drosophila S2* and *Spodoptera Sf9*, animal cells such as CHO, COS or Bowes melanoma and adenoviruses. The selection of an appropriate host is within the abilities of those skilled in the art.

[0217] The vector may be introduced into the host cells using any of a variety of techniques, including transformation, transfection, transduction, viral infection, gene guns, or Timediated gene transfer. Particular methods include calcium phosphate transfection, DEAE-Dextran mediated transfection, lipofection, or electroporation (Davis, L., Dibner, M., Battey, I., Basic Methods in Molecular Biology, (1986)).

[0218] Where appropriate, the engineered host cells can be cultured in conventional nutrient media modified as appropriate for activating promoters, selecting transformants or amplifying the genes of the invention. Following transformation of a suitable host strain and growth of the host strain to an appropriate cell density, the selected promoter may be induced by appropriate means (e.g., temperature shift or chemical induction) and the cells may be cultured for an additional period to allow them to produce the desired polypeptide or fragment thereof.

[0219] Cells are typically harvested by centrifugation, disrupted by physical or chemical means and the resulting crude extract is retained for further purification. Microbial cells employed for expression of proteins can be disrupted by any convenient method, including freeze-thaw cycling, sonication, mechanical disruption, or use of cell lysing agents. Such methods are well known to those skilled in the art. The expressed polypeptide or fragment thereof can be recovered and purified from recombinant cell cultures by methods including ammonium sulfate or ethanol precipitation, acid extraction, anion or cation exchange chromatography, phosphocellulose chromatography, hydrophobic interaction chromatography, affinity chromatography, hydroxylapatite chromatography and lectin chromatography. Protein refolding steps can be used, as necessary, in completing configuration of the polypeptide. If desired, high performance liquid chromatography (HPLC) can be employed for final purification steps.

[0220] Various mammalian cell culture systems can also be employed to express recombinant protein. Examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts (described by Gluzman, Cell, 23:175, 1981) and other cell lines capable of expressing proteins from a compatible vector, such as the C127, 3T3, CHO, HeLa and BHK cell lines.

[0221] The constructs in host cells can be used in a conventional manner to produce the gene product encoded by the recombinant sequence. Depending upon the host employed in a recombinant production procedure, the polypeptides produced by host cells containing the vector may be glycosylated or may be non-glycosylated. Polypeptides of the invention may or may not also include an initial methionine amino acid residue.

[0222] Alternatively, the polypeptides of amino acid sequences of the invention, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof can be synthetically produced by conventional peptide synthesizers. In other aspects, fragments or portions of the polypeptides may be employed for producing the corresponding full-length polypeptide by peptide synthesis; therefore, the fragments may be employed as intermediates for producing the full-length polypeptides.

[0223] Cell-free translation systems can also be employed to produce one of the polypeptides of amino acid sequences of the invention, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof using mRNAs transcribed from a DNA construct comprising a promoter operably linked to a nucleic acid encoding the polypeptide or fragment thereof. In some aspects, the DNA construct may be linearized prior to conducting an *in vitro* transcription reaction. The transcribed

mRNA is then incubated with an appropriate cell-free translation extract, such as a rabbit reticulocyte extract, to produce the desired polypeptide or fragment thereof.

Amplification of Nucleic Acids

[0224] In practicing the invention, nucleic acids of the invention and nucleic acids encoding the xylanases of the invention, or modified nucleic acids of the invention, can be reproduced by amplification. Amplification can also be used to clone or modify the nucleic acids of the invention. One of skill in the art can design amplification primer sequence pairs for any part of or the full length of these sequences.

[0225] A nucleic acid may be amplified by such a primer pair, e.g., a primer pair as set forth by about the first (the 5') 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 residues of a nucleic acid of the invention, and about the first (the 5') 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 residues of the complementary strand.

[0226] An amplification primer sequence pair for amplifying a nucleic acid encoding a polypeptide having a xylanase activity, may be capable of amplifying a nucleic acid comprising a sequence of the invention, or fragments or subsequences thereof. One or each member of the amplification primer sequence pair can comprise an oligonucleotide comprising at least about 10 to 50 consecutive bases of the sequence, or about 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 consecutive bases of the sequence. The primer pair may comprise a first member having a sequence as set forth by about the first (the 5') 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 residues of a nucleic acid of the invention, and a second member having a sequence as set forth by about the first (the 5') 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, or 25 residues of the complementary strand of the first member. Xylanases may be generated by amplification, e.g., polymerase chain reaction (PCR), using such an amplification primer pair. Methods of making a xylanase by amplification, e.g., polymerase chain reaction (PCR), use such an amplification primer pair of the disclosure. The amplification primer pair may amplify a nucleic acid from a library, e.g., a gene library, such as an environmental library.

[0227] Amplification reactions can also be used to quantify the amount of nucleic acid in a sample (such as the amount of message in a cell sample), label the nucleic acid (e.g., to apply it to an array or a blot), detect the nucleic acid, or quantify the amount of a specific nucleic acid in a sample. Message isolated from a cell or a cDNA library may be amplified.

[0228] The skilled artisan can select and design suitable oligonucleotide amplification primers. Amplification methods are also well known in the art, and include, e.g., polymerase chain reaction, PCR (see, e.g., PCR PROTOCOLS, A GUIDE TO METHODS AND APPLICATIONS, ed. Innis, Academic Press, N.Y. (1990) and PCR STRATEGIES (1995), ed. Innis, Academic Press, Inc., N.Y., ligase chain reaction (LCR) (see, e.g., Wu (1989) Genomics 4:560; Landegren (1988) Science 241:1077; Barringer (1990) Gene 89:117); transcription amplification (see, e.g., Kwoh (1989) Proc. Natl. Acad. Sci. USA 86:1173); and, self-sustained sequence replication (see, e.g., Guatelli (1990) Proc. Natl. Acad. Sci. USA 87:1874); Q Beta replicase amplification (see, e.g., Smith (1997) J. Clin. Microbiol. 35:1477-1491), automated Q-beta replicase amplification assay (see, e.g., Burg (1996) Mol. Cell. Probes 10:257-271) and other RNA polymerase mediated techniques (e.g., NASBA, Cangene, Mississauga, Ontario); see also Berger (1987) Methods Enzymol. 152:307-316; Sambrook; Ausubel; U.S. Patent Nos. 4,683,195 and 4,683,202; Sooknanan (1995) Biotechnology 13:563-564.

Determining the degree of sequence identity

[0229] The invention provides nucleic acids comprising sequences having at least 95%, 96%, 97%, 98%, 99%, or more, or complete (100%) sequence identity to an exemplary nucleic acid of the invention as defined above. The invention provides polypeptides comprising sequences having at least 95%, 96%, 97%, 98%, 99%, or more, or complete (100%) sequence identity to an exemplary polypeptide of the invention. The extent of sequence identity (homology) may be determined using any computer program and associated parameters, including those described herein, such as BLAST 2.2.2. or FASTA version 3.0t78, with the default parameters.

[0230] As used herein, the terms "computer," "computer program" and "processor" are used in their broadest general contexts and incorporate all such devices, as described in detail, below. A "coding sequence of" or a "sequence encodes" a particular polypeptide or protein, is a nucleic acid sequence which is transcribed and translated into a polypeptide or protein when placed under the control of appropriate regulatory sequences.

[0231] The phrase "substantially identical" in the context of two nucleic acids or polypeptides, refers to two or more sequences that have at least 95%, 96%, 97%, 98%, 99%, or more nucleotide or amino acid residue (sequence) identity, when compared and aligned for maximum correspondence, as measured using one of the known sequence comparison algorithms or by visual inspection. Typically, the substantial identity exists over a region of at least about 100 residues and most commonly the sequences are substantially identical over at least about 150-200 residues. In some aspects, the sequences are substantially identical over the entire length of the coding regions.

[0232] Additionally a "substantially identical" amino acid sequence is a sequence that differs from a reference sequence by one or more conservative or non-conservative amino acid substitutions, deletions, or insertions, particularly when such a substitution occurs at a site that is not the active site of the molecule and provided that the polypeptide essentially retains its functional properties. A conservative amino acid substitution, for example, substitutes one amino acid for another of the same class (e.g., substitution of one hydrophobic amino acid, such as isoleucine, valine, leucine, or methionine, for another, or substitution of one polar amino acid for another, such as substitution of arginine for lysine, glutamic acid for aspartic acid or glutamine for asparagine). One or more amino acids can be deleted, for example, from a xylanase polypeptide, resulting in modification of the structure of the polypeptide, without significantly altering its biological activity. For example, amino- or carboxyl-terminal amino acids that are not required for xylanase biological activity can be removed. Modified polypeptide sequences of the invention can be assayed for xylanase biological activity by any number of methods, including contacting the modified polypeptide sequence with a xylanase substrate and determining whether the modified polypeptide decreases the amount of specific substrate in the assay or increases the bioproducts of the enzymatic reaction of a functional xylanase polypeptide with the substrate.

[0233] Nucleic acid sequences of the disclosure can comprise at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive nucleotides of an exemplary sequence of the invention and sequences substantially identical thereto. Nucleic acid sequences of the disclosure can comprise homologous sequences and fragments of nucleic acid sequences and sequences substantially identical thereto, refer to a sequence having at least 50%, 51%, 52%, 53%, 54%, 55%, 56%, 57%, 58%, 59%, 60%, 61%, 62%, 63%, 64%, 65%, 66%, 67%, 68%, 69%, 70%, 71%, 72%, 73%, 74%, 75%, 76%, 77%, 78%, 79%, 80%, 81%, 82%, 83%, 84%, 85%, 86%, 87%, 88%, 89%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or more sequence identity (homology) to these sequences. Homology may be determined using any of the computer programs and parameters described herein, including FASTA version 3.0t78 with the default parameters. Homologous sequences also include RNA sequences in which

uridines replace the thymines in the nucleic acid sequences of the invention. The homologous sequences may be obtained using any of the procedures described herein or may result from the correction of a sequencing error. It will be appreciated that the nucleic acid sequences of the invention and sequences substantially identical thereto, can be represented in the traditional single character format (See the inside back cover of Stryer, Lubert. Biochemistry, 3rd Ed., W. H Freeman & Co., New York.) or in any other format which records the identity of the nucleotides in a sequence.

[0234] Various sequence comparison programs identified elsewhere in this patent specification are particularly contemplated for use in this aspect of the invention. Protein and/or nucleic acid sequence homologies may be evaluated using any of the variety of sequence comparison algorithms and programs known in the art. Such algorithms and programs include, but are by no means limited to, TBLASTN, BLASTP, FASTA, TFASTA and CLUSTALW (Pearson and Lipman, Proc. Natl. Acad. Sci. USA 85(8):2444-2448, 1988; Altschul et al., J. Mol. Biol. 215(3):403-410, 1990; Thompson et al., Nucleic Acids Res. 22(2):4673-4680, 1994; Higgins et al., Methods Enzymol. 266:383-402, 1996; Altschul et al., J. Mol. Biol. 215(3):403-410, 1990; Altschul et al., Nature Genetics 3:266-272, 1993).

[0235] Homology or identity is often measured using sequence analysis software (e.g., Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, WI 53705). Such software matches similar sequences by assigning degrees of homology to various deletions, substitutions and other modifications. The terms "homology" and "identity" in the context of two or more nucleic acids or polypeptide sequences, refer to two or more sequences or subsequences that are the same or have a specified percentage of amino acid residues or nucleotides that are the same when compared and aligned for maximum correspondence over a comparison window or designated region as measured using any number of sequence comparison algorithms or by manual alignment and visual inspection.

[0236] For sequence comparison, typically one sequence acts as a reference sequence, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are entered into a computer, subsequence coordinates are designated, if necessary and sequence algorithm program parameters are designated. Default program parameters can be used, or alternative parameters can be designated. The sequence comparison algorithm then calculates the percent sequence identities for the test sequences relative to the reference sequence, based on the program parameters.

[0237] A "comparison window", as used herein, includes reference to a segment of any one of the number of contiguous positions selected from the group consisting of from 20 to 600, usually about 50 to about 200, more usually about 100 to about 150 in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned. Methods of alignment of sequence for comparison are well-known in the art. Optimal alignment of sequences for comparison can be conducted, e.g., by the local homology algorithm of Smith & Waterman, Adv. Appl. Math. 2:482, 1981, by the homology alignment algorithm of Needleman & Wunsch, J. Mol. Biol. 48:443, 1970, by the search for similarity method of person & Lipman, Proc. Nat'l. Acad. Sci. USA 85:2444, 1988, by computerized implementations of these algorithms (GAP, BESTFIT, FASTA and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by manual alignment and visual inspection. Other algorithms for determining homology or identity include, for example, in addition to a BLAST program (Basic Local Alignment Search Tool at the National Center for Biological Information), ALIGN, AMAS (Analysis of Multiply Aligned Sequences), AMPS (Protein Multiple Sequence Alignment), ASSET (Aligned Segment Statistical Evaluation Tool), BANDS, BESTSCOR, BIOSCAN (Biological Sequence Comparative Analysis Node), BLIMPS (BLOCKS IMPROVED Searcher), FASTA, Intervals & Points, BMB, CLUSTAL V, CLUSTAL W, CONSENSUS, LCONSENSUS,

WCONSENSUS, Smith-Waterman algorithm, DARWIN, Las Vegas algorithm, FNAT (Forced Nucleotide Alignment Tool), Framealign, Framesearch, DYNAMIC, FILTER, FSAP (Fristensky Sequence Analysis Package), GAP (Global Alignment Program), GENAL, GIBBS, GenQuest, ISSC (Sensitive Sequence Comparison), LALIGN (Local Sequence Alignment), LCP (Local Content Program), MACAW (Multiple Alignment Construction & Analysis Workbench), MAP (Multiple Alignment Program), MBLKP, MBLKN, PIMA (Pattern-Induced Multi-sequence Alignment), SAGA (Sequence Alignment by Genetic Algorithm) and WHAT-IF. Such alignment programs can also be used to screen genome databases to identify polynucleotide sequences having substantially identical sequences. A number of genome databases are available, for example, a substantial portion of the human genome is available as part of the Human Genome Sequencing Project. At least twenty-one other genomes have already been sequenced, including, for example, *M. genitalium* (Fraser *et al.*, 1995), *M. jannaschii* (Bult *et al.*, 1996), *H. influenzae* (Fleischmann *et al.*, 1995), *E. coli* (Blattner *et al.*, 1997) and yeast (*S. cerevisiae*) (Mewes *et al.*, 1997) and *D. melanogaster* (Adams *et al.*, 2000). Significant progress has also been made in sequencing the genomes of model organism, such as mouse, *C. elegans* and *Arabidopsis sp.* Several databases containing genomic information annotated with some functional information are maintained by different organization and are accessible via the internet

[0238] One example of a useful algorithm is BLAST and BLAST 2.0 algorithms, which are described in Altschul *et al.*, Nuc. Acids Res. 25:3389-3402, 1997 and Altschul *et al.*, J. Mol. Biol. 215:403-410, 1990, respectively. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul *et al.*, *supra*). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, M=5, N=-4 and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength of 3 and expectations (E) of 10 and the BLOSUM62 scoring matrix (see Henikoff & Henikoff, Proc. Natl. Acad. Sci. USA 89:10915, 1989) alignments (B) of 50, expectation (E) of 10, M=5, N= -4 and a comparison of both strands.

[0239] The BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, *e.g.*, Karlin & Altschul, Proc. Natl. Acad. Sci. USA 90:5873, 1993). One measure of similarity provided by BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a references sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.2, more preferably less than about 0.01 and most preferably less than about 0.001.

[0240] In one aspect, protein and nucleic acid sequence homologies are evaluated using the Basic Local Alignment Search Tool ("BLAST") In particular, five specific BLAST programs are used to perform the following task:

1. (1) BLASTP and BLAST3 compare an amino acid query sequence against a protein sequence database;
2. (2) BLASTN compares a nucleotide query sequence against a nucleotide sequence database;
3. (3) BLASTX compares the six-frame conceptual translation products of a query nucleotide sequence (both strands) against a protein sequence database;
4. (4) TBLASTN compares a query protein sequence against a nucleotide sequence database translated in all six reading frames (both strands); and
5. (5) TBLASTX compares the six-frame translations of a nucleotide query sequence against the six-frame translations of a nucleotide sequence database.

[0241] The BLAST programs identify homologous sequences by identifying similar segments, which are referred to herein as "high-scoring segment pairs," between a query amino or nucleic acid sequence and a test sequence which is preferably obtained from a protein or nucleic acid sequence database. High-scoring segment pairs are preferably identified (*i.e.*, aligned) by means of a scoring matrix, many of which are known in the art. Preferably, the scoring matrix used is the BLOSUM62 matrix (Gonnet et al., Science 256:1443-1445, 1992; Henikoff and Henikoff, Proteins 17:49-61, 1993). Less preferably, the PAM or PAM250 matrices may also be used (see, e.g., Schwartz and Dayhoff, eds., 1978, Matrices for Detecting Distance Relationships: Atlas of Protein Sequence and Structure, Washington: National Biomedical Research Foundation). BLAST programs are accessible through the U.S. National Library of Medicine.

[0242] The parameters used with the above algorithms may be adapted depending on the sequence length and degree of homology studied. In some aspects, the parameters may be the default parameters used by the algorithms in the absence of instructions from the user.

Computer systems and computer program products

[0243] To determine and identify sequence identities, structural homologies, motifs and the like *in silico*, a nucleic acid or polypeptide sequence of the invention can be stored, recorded, and manipulated on any medium which can be read and accessed by a computer.

[0244] A nucleic acid or polypeptide sequence of the invention can be stored, recorded and manipulated on any medium which can be read and accessed by a computer. As used herein, the words "recorded" and "stored" refer to a process for storing information on a computer medium. A skilled artisan can readily adopt any of the presently known methods for recording information on a computer readable medium to generate manufactures comprising one or more of the nucleic acid sequences of the invention and sequences substantially identical thereto, one or more of the polypeptide sequences of the invention and sequences substantially identical thereto.

[0245] A computer readable medium may have recorded thereon one or more of the nucleic acid sequences of the invention and sequences substantially identical thereto. A computer readable medium may have recorded thereon one or more of the polypeptide sequences of the invention and sequences substantially identical thereto. Another aspect of the invention is a computer readable medium having recorded thereon at least 2, 5, 10, 15, or 20 or more of the sequences as set forth above.

[0246] Computer readable media include magnetically readable media, optically readable media, electronically readable media and magnetic/optical media. For example, the computer readable media may be a hard disk, a floppy disk, a magnetic tape, CD-ROM, Digital Versatile Disk (DVD), Random Access

Memory (RAM), or Read Only Memory (ROM) as well as other types of other media known to those skilled in the art.

[0247] One example of a computer system 100 is illustrated in block diagram form in Figure 1. As used herein, "a computer system" refers to the hardware components, software components and data storage components used to analyze a nucleotide sequence of a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence as set forth in the amino acid sequences of the invention. The computer system 100 typically includes a processor for processing, accessing and manipulating the sequence data. The processor 105 can be any well-known type of central processing unit, such as, for example, the Pentium III from Intel Corporation, or similar processor from Sun, Motorola, Compaq, AMD or International Business Machines.

[0248] The computer system 100 is a general purpose system that comprises the processor 105 and one or more internal data storage components 110 for storing data and one or more data retrieving devices for retrieving the data stored on the data storage components. A skilled artisan can readily appreciate that any one of the currently available computer systems are suitable.

[0249] The computer system 100 includes a processor 105 connected to a bus which is connected to a main memory 115 (preferably implemented as RAM) and one or more internal data storage devices 110, such as a hard drive and/or other computer readable media having data recorded thereon. The computer system 100 further includes one or more data retrieving device 118 for reading the data stored on the internal data storage devices 110.

[0250] The data retrieving device 118 may represent, for example, a floppy disk drive, a compact disk drive, a magnetic tape drive, or a modem capable of connection to a remote data storage system (e.g., via the internet) etc. The internal data storage device 110 is a removable computer readable medium such as a floppy disk, a compact disk, a magnetic tape, etc. containing control logic and/or data recorded thereon. The computer system 100 may advantageously include or be programmed by appropriate software for reading the control logic and/or the data from the data storage component once inserted in the data retrieving device.

[0251] The computer system 100 includes a display 120 which is used to display output to a computer user. It should also be noted that the computer system 100 can be linked to other computer systems 125a-c in a network or wide area network to provide centralized access to the computer system 100.

[0252] Software for accessing and processing the nucleotide sequences of a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto, (such as search tools, compare tools and modeling tools etc.) may reside in main memory 115 during execution.

[0253] The computer system 100 may further comprise a sequence comparison algorithm for comparing a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto, stored on a computer readable medium to a reference nucleotide or polypeptide sequence(s) stored on a computer readable medium. A "sequence comparison algorithm" refers to one or more programs which are implemented (locally or remotely) on the computer system 100 to compare a nucleotide sequence with other nucleotide sequences and/or compounds stored within a data storage means. For example, the sequence comparison algorithm may compare the nucleotide sequences of a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences

substantially identical thereto, stored on a computer readable medium to reference sequences stored on a computer readable medium to identify homologies or structural motifs.

[0254] Figure 2 is a flow diagram illustrating one aspect of a process 200 for comparing a new nucleotide or protein sequence with a database of sequences in order to determine the homology levels between the new sequence and the sequences in the database. The database of sequences can be a private database stored within the computer system 100, or a public database such as GENBANK that is available through the Internet.

[0255] The process 200 begins at a start state 201 and then moves to a state 202 wherein the new sequence to be compared is stored to a memory in a computer system 100. As discussed above, the memory could be any type of memory, including RAM or an internal storage device.

[0256] The process 200 then moves to a state 204 wherein a database of sequences is opened for analysis and comparison. The process 200 then moves to a state 206 wherein the first sequence stored in the database is read into a memory on the computer. A comparison is then performed at a state 210 to determine if the first sequence is the same as the second sequence. It is important to note that this step is not limited to performing an exact comparison between the new sequence and the first sequence in the database. Well-known methods are known to those of skill in the art for comparing two nucleotide or protein sequences, even if they are not identical. For example, gaps can be introduced into one sequence in order to raise the homology level between the two tested sequences. The parameters that control whether gaps or other features are introduced into a sequence during comparison are normally entered by the user of the computer system.

[0257] Once a comparison of the two sequences has been performed at the state 210, a determination is made at a decision state 210 whether the two sequences are the same. Of course, the term "same" is not limited to sequences that are absolutely identical. Sequences that are within the homology parameters entered by the user will be marked as "same" in the process 200.

[0258] If a determination is made that the two sequences are the same, the process 200 moves to a state 214 wherein the name of the sequence from the database is displayed to the user. This state notifies the user that the sequence with the displayed name fulfills the homology constraints that were entered. Once the name of the stored sequence is displayed to the user, the process 200 moves to a decision state 218 wherein a determination is made whether more sequences exist in the database. If no more sequences exist in the database, then the process 200 terminates at an end state 220. However, if more sequences do exist in the database, then the process 200 moves to a state 224 wherein a pointer is moved to the next sequence in the database so that it can be compared to the new sequence. In this manner, the new sequence is aligned and compared with every sequence in the database.

[0259] It should be noted that if a determination had been made at the decision state 212 that the sequences were not homologous, then the process 200 would move immediately to the decision state 218 in order to determine if any other sequences were available in the database for comparison.

[0260] A computer system comprises a processor, a data storage device having stored thereon a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto, a data storage device having retrievably stored thereon reference nucleotide sequences or polypeptide sequences to be compared to a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto and a sequence comparer for conducting the comparison. The sequence comparer may indicate a homology level between the sequences compared or

identify structural motifs in the above described nucleic acid code of nucleic acid sequences of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto, or it may identify structural motifs in sequences which are compared to these nucleic acid codes and polypeptide codes. The data storage device may have stored thereon the sequences of at least 2, 5, 10, 15, 20, 25, 30 or 40 or more of the nucleic acid sequences of the invention and sequences substantially identical thereto, or the polypeptide sequences of the invention and sequences substantially identical thereto.

[0261] A method for determining the level of homology between a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto and a reference nucleotide sequence, includes reading the nucleic acid code or the polypeptide code and the reference nucleotide or polypeptide sequence through the use of a computer program which determines homology levels and determining homology between the nucleic acid code or polypeptide code and the reference nucleotide or polypeptide sequence with the computer program. The computer program may be any of a number of computer programs for determining homology levels, including those specifically enumerated herein, (*e.g.*, BLAST2N with the default parameters or with any modified parameters). The method may be implemented using the computer systems described above. The method may also be performed by reading at least 2, 5, 10, 15, 20, 25, 30 or 40 or more of the above described nucleic acid sequences of the invention, or the polypeptide sequences of the invention through use of the computer program and determining homology between the nucleic acid codes or polypeptide codes and reference nucleotide sequences or polypeptide sequences.

[0262] Figure 3 is a flow diagram illustrating one aspect of a process 250 in a computer for determining whether two sequences are homologous. The process 250 begins at a start state 252 and then moves to a state 254 wherein a first sequence to be compared is stored to a memory. The second sequence to be compared is then stored to a memory at a state 256. The process 250 then moves to a state 260 wherein the first character in the first sequence is read and then to a state 262 wherein the first character of the second sequence is read. It should be understood that if the sequence is a nucleotide sequence, then the character would normally be either A, T, C, G or U. If the sequence is a protein sequence, then it is preferably in the single letter amino acid code so that the first and sequence sequences can be easily compared.

[0263] A determination is then made at a decision state 264 whether the two characters are the same. If they are the same, then the process 250 moves to a state 268 wherein the next characters in the first and second sequences are read. A determination is then made whether the next characters are the same. If they are, then the process 250 continues this loop until two characters are not the same. If a determination is made that the next two characters are not the same, the process 250 moves to a decision state 274 to determine whether there are any more characters either sequence to read.

[0264] If there are not any more characters to read, then the process 250 moves to a state 276 wherein the level of homology between the first and second sequences is displayed to the user. The level of homology is determined by calculating the proportion of characters between the sequences that were the same out of the total number of sequences in the first sequence. Thus, if every character in a first 100 nucleotide sequence aligned with a every character in a second sequence, the homology level would be 100%.

[0265] Alternatively, the computer program may be a computer program which compares the nucleotide sequences of a nucleic acid sequence as set forth in the invention, to one or more reference nucleotide sequences in order to determine whether the nucleic acid code of a nucleic acid sequence of the invention

and sequences substantially identical thereto, differs from a reference nucleic acid sequence at one or more positions. In one aspect such a program records the length and identity of inserted, deleted or substituted nucleotides with respect to the sequence of either the reference polynucleotide or a nucleic acid sequence of the invention and sequences substantially identical thereto. In one aspect, the computer program may be a program which determines whether a nucleic acid sequence of the invention and sequences substantially identical thereto, contains a single nucleotide polymorphism (SNP) with respect to a reference nucleotide sequence.

[0266] A method for determining whether a nucleic acid sequence of the invention and sequences substantially identical thereto, differs at one or more nucleotides from a reference nucleotide sequence comprises the steps of reading the nucleic acid code and the reference nucleotide sequence through use of a computer program which identifies differences between nucleic acid sequences and identifying differences between the nucleic acid code and the reference nucleotide sequence with the computer program. The computer program may be a program which identifies single nucleotide polymorphisms. The method may be implemented by the computer systems described above and the method illustrated in Figure 3. The method may also be performed by reading at least 2, 5, 10, 15, 20, 25, 30, or 40 or more of the nucleic acid sequences of the invention and sequences substantially identical thereto and the reference nucleotide sequences through the use of the computer program and identifying differences between the nucleic acid codes and the reference nucleotide sequences with the computer program.

[0267] The computer based system may further comprise an identifier for identifying features within a nucleic acid sequence of the invention or a polypeptide sequence of the invention and sequences substantially identical thereto.

[0268] An "identifier" refers to one or more programs which identifies certain features within a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto. The identifier may comprise a program which identifies an open reading frame in a nucleic acid sequence of the invention and sequences substantially identical thereto.

[0269] Figure 4 is a flow diagram illustrating one aspect of an identifier process 300 for detecting the presence of a feature in a sequence. The process 300 begins at a start state 302 and then moves to a state 304 wherein a first sequence that is to be checked for features is stored to a memory 115 in the computer system 100. The process 300 then moves to a state 306 wherein a database of sequence features is opened. Such a database would include a list of each feature's attributes along with the name of the feature. For example, a feature name could be "Initiation Codon" and the attribute would be "ATG". Another example would be the feature name "TAATAA Box" and the feature attribute would be "TAATAA". An example of such a database is produced by the University of Wisconsin Genetics Computer Group. Alternatively, the features may be structural polypeptide motifs such as alpha helices, beta sheets, or functional polypeptide motifs such as enzymatic active sites, helix-turn-helix motifs or other motifs known to those skilled in the art.

[0270] Once the database of features is opened at the state 306, the process 300 moves to a state 308 wherein the first feature is read from the database. A comparison of the attribute of the first feature with the first sequence is then made at a state 310. A determination is then made at a decision state 316 whether the attribute of the feature was found in the first sequence. If the attribute was found, then the process 300 moves to a state 318 wherein the name of the found feature is displayed to the user.

[0271] The process 300 then moves to a decision state 320 wherein a determination is made whether more features exist in the database. If no more features do exist, then the process 300 terminates at an

end state 324. However, if more features do exist in the database, then the process 300 reads the next sequence feature at a state 326 and loops back to the state 310 wherein the attribute of the next feature is compared against the first sequence. It should be noted, that if the feature attribute is not found in the first sequence at the decision state 316, the process 300 moves directly to the decision state 320 in order to determine if any more features exist in the database.

[0272] A method of identifying a feature within a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto, comprises reading the nucleic acid code(s) or polypeptide code(s) through the use of a computer program which identifies features therein and identifying features within the nucleic acid code(s) with the computer program. The computer program may comprise a computer program which identifies open reading frames. The method may be performed by reading a single sequence or at least 2, 5, 10, 15, 20, 25, 30, or 40 of the nucleic acid sequences of the invention and sequences substantially identical thereto, or the polypeptide sequences of the invention and sequences substantially identical thereto, through the use of the computer program and identifying features within the nucleic acid codes or polypeptide codes with the computer program.

[0273] A nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto, may be stored and manipulated in a variety of data processor programs in a variety of formats. For example, a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto, may be stored as text in a word processing file, such as Microsoft WORD™ or WORDPERFECT™ or as an ASCII file in a variety of database programs familiar to those of skill in the art, such as DB2™, SYBASE™, or ORACLE™. In addition, many computer programs and databases may be used as sequence comparison algorithms, identifiers, or sources of reference nucleotide sequences or polypeptide sequences to be compared to a nucleic acid sequence of the invention and sequences substantially identical thereto, or a polypeptide sequence of the invention and sequences substantially identical thereto. The following list provides guidance to programs and databases which are useful with the nucleic acid sequences of the invention and sequences substantially identical thereto, or the polypeptide sequences of the invention and sequences substantially identical thereto.

[0274] The programs and databases which may be used include, but are not limited to: MacPattern (EMBL), DiscoveryBase (Molecular Applications Group), GeneMine (Molecular Applications Group), Look (Molecular Applications Group), MacLook (Molecular Applications Group), BLAST and BLAST2 (NCBI), BLASTN and BLASTX (Altschul et al, J. Mol. Biol. 215: 403, 1990), FASTA (Pearson and Lipman, Proc. Natl. Acad. Sci. USA, 85: 2444, 1988), FASTDB (Brutlag et al. Comp. App. Biosci. 6:237-245, 1990), Catalyst (Molecular Simulations Inc.), Catalyst/SHAPE (Molecular Simulations Inc.), Cerius².DBAccess (Molecular Simulations Inc.), HypoGen (Molecular Simulations Inc.), Insight II, (Molecular Simulations Inc.), Discover (Molecular Simulations Inc.), CHARMM (Molecular Simulations Inc.), Felix (Molecular Simulations Inc.), DelPhi, (Molecular Simulations Inc.), QuanteMM, (Molecular Simulations Inc.), Homology (Molecular Simulations Inc.), Modeler (Molecular Simulations Inc.), ISIS (Molecular Simulations Inc.), Quanta/Protein Design (Molecular Simulations Inc.), WebLab (Molecular Simulations Inc.), WebLab Diversity Explorer (Molecular Simulations Inc.), Gene Explorer (Molecular Simulations Inc.), SeqFold (Molecular Simulations Inc.), the MDL Available Chemicals Directory database, the MDL Drug Data Report data base, the Comprehensive Medicinal Chemistry database, Derwent's World Drug Index database, the BioByteMasterFile database, the Genbank database and the Genseqn database. Many other programs and data bases would be apparent to one of skill in the art given the present disclosure.

[0275] Motifs which may be detected using the above programs include sequences encoding leucine

zippers, helix-turn-helix motifs, glycosylation sites, ubiquitination sites, alpha helices and beta sheets, signal sequences encoding signal peptides which direct the secretion of the encoded proteins, sequences implicated in transcription regulation such as homeoboxes, acidic stretches, enzymatic active sites, substrate binding sites and enzymatic cleavage sites.

Hybridization of nucleic acids

[0276] The invention provides isolated, synthetic or recombinant nucleic acids that hybridize under stringent conditions to an exemplary sequence of the invention. The stringent conditions can be highly stringent conditions, medium stringent conditions and/or low stringent conditions, including the high and reduced stringency conditions described herein. In one aspect, it is the stringency of the wash conditions that set forth the conditions which determine whether a nucleic acid is within the scope of the invention, as discussed below.

[0277] Nucleic acids defined by their ability to hybridize under stringent conditions can be between about five residues and the full length of nucleic acid of the invention; e.g., they can be at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 55, 60, 65, 70, 75, 80, 90, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, or more, residues in length. Nucleic acids shorter than full length are also included. These nucleic acids can be useful as, e.g., hybridization probes, labeling probes, PCR oligonucleotide probes, iRNA (single or double stranded), antisense or sequences encoding antibody binding peptides (epitopes), motifs, active sites and the like.

[0278] Nucleic acids may be defined by their ability to hybridize under high stringency comprising conditions of about 50% formamide at about 37°C to 42°C. Alternatively, by their ability to hybridize under reduced stringency comprising conditions in about 35% to 25% formamide at about 30°C to 35°C.

[0279] Alternatively, nucleic acids are defined by their ability to hybridize under high stringency comprising conditions at 42°C in 50% formamide, 5X SSPE, 0.3% SDS, and a repetitive sequence blocking nucleic acid, such as cot-1 or salmon sperm DNA (e.g., 200 ug/ml sheared and denatured salmon sperm DNA). Nucleic acids may also be defined by their ability to hybridize under reduced stringency conditions comprising 35% formamide at a reduced temperature of 35°C.

[0280] In nucleic acid hybridization reactions, the conditions used to achieve a particular level of stringency will vary, depending on the nature of the nucleic acids being hybridized. For example, the length, degree of complementarity, nucleotide sequence composition (e.g., GC v. AT content) and nucleic acid type (e.g., RNA v. DNA) of the hybridizing regions of the nucleic acids can be considered in selecting hybridization conditions. An additional consideration is whether one of the nucleic acids is immobilized, for example, on a filter.

[0281] Hybridization may be carried out under conditions of low stringency, moderate stringency or high stringency. As an example of nucleic acid hybridization, a polymer membrane containing immobilized denatured nucleic acids is first prehybridized for 30 minutes at 45°C in a solution consisting of 0.9 M NaCl, 50 mM NaH₂PO₄, pH 7.0, 5.0 mM Na₂EDTA, 0.5% SDS, 10X Denhardt's and 0.5 mg/ml polyriboadenylic acid. Approximately 2×10^7 cpm (specific activity $4-9 \times 10^8$ cpm/ug) of ³²P end-labeled oligonucleotide probe are then added to the solution. After 12-16 hours of incubation, the membrane is washed for 30 minutes at room temperature in 1X SET (150 mM NaCl, 20 mM Tris hydrochloride, pH 7.8, 1 mM Na₂EDTA) containing 0.5% SDS, followed by a 30 minute wash in fresh 1X SET at T_m-10°C for the oligonucleotide probe. The membrane is then exposed to auto-radiographic film for detection of

hybridization signals.

[0282] All of the foregoing hybridizations would be considered to be under conditions of high stringency.

[0283] Following hybridization, a filter can be washed to remove any non-specifically bound detectable probe. The stringency used to wash the filters can also be varied depending on the nature of the nucleic acids being hybridized, the length of the nucleic acids being hybridized, the degree of complementarity, the nucleotide sequence composition (*e.g.*, GC v. AT content) and the nucleic acid type (*e.g.*, RNA v. DNA). Examples of progressively higher stringency condition washes are as follows: 2X SSC, 0.1% SDS at room temperature for 15 minutes (low stringency); 0.1X SSC, 0.5% SDS at room temperature for 30 minutes to 1 hour (moderate stringency); 0.1X SSC, 0.5% SDS for 15 to 30 minutes at between the hybridization temperature and 68°C (high stringency); and 0.15M NaCl for 15 minutes at 72°C (very high stringency). A final low stringency wash can be conducted in 0.1X SSC at room temperature. The examples above are merely illustrative of one set of conditions that can be used to wash filters. One of skill in the art would know that there are numerous recipes for different stringency washes. Some other examples are given below.

[0284] Nucleic acids which have hybridized to the probe are identified by autoradiography or other conventional techniques.

[0285] The above procedure may be modified to identify nucleic acids having decreasing levels of homology to the probe sequence. For example, to obtain nucleic acids of decreasing homology to the detectable probe, less stringent conditions may be used. For example, the hybridization temperature may be decreased in increments of 5°C from 68°C to 42°C in a hybridization buffer having a Na⁺ concentration of approximately 1M. Following hybridization, the filter may be washed with 2X SSC, 0.5% SDS at the temperature of hybridization. These conditions are considered to be "moderate" conditions above 50°C and "low" conditions below 50°C. A specific example of "moderate" hybridization conditions is when the above hybridization is conducted at 55°C. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at 45°C.

[0286] Alternatively, the hybridization may be carried out in buffers, such as 6X SSC, containing formamide at a temperature of 42°C. In this case, the concentration of formamide in the hybridization buffer may be reduced in 5% increments from 50% to 0% to identify clones having decreasing levels of homology to the probe. Following hybridization, the filter may be washed with 6X SSC, 0.5% SDS at 50°C. These conditions are considered to be "moderate" conditions above 25% formamide and "low" conditions below 25% formamide. A specific example of "moderate" hybridization conditions is when the above hybridization is conducted at 30% formamide. A specific example of "low stringency" hybridization conditions is when the above hybridization is conducted at 10% formamide.

[0287] However, the selection of a hybridization format is not critical - it is the stringency of the wash conditions that set forth the conditions which determine whether a nucleic acid is relevant. Wash conditions used to identify relevant nucleic acids include, *e.g.*: a salt concentration of about 0.02 molar at pH 7 and a temperature of at least about 50°C or about 55°C to about 60°C; or, a salt concentration of about 0.15 M NaCl at 72°C for about 15 minutes; or, a salt concentration of about 0.2X SSC at a temperature of at least about 50°C or about 55°C to about 60°C for about 15 to about 20 minutes; or, the hybridization complex is washed twice with a solution with a salt concentration of about 2X SSC containing 0.1% SDS at room temperature for 15 minutes and then washed twice by 0.1X SSC containing 0.1% SDS at 68°C for 15 minutes; or, equivalent conditions. See Sambrook, Tijssen and Ausubel for a description of SSC buffer and equivalent conditions.

[0288] These methods may be used to isolate nucleic acids. For example, the preceding methods may be used to isolate nucleic acids having a sequence with at least about 97%, at least 95%, homology to a nucleic acid sequence selected from the group consisting of one of the sequences of the invention and sequences substantially identical thereto, or fragments comprising at least about 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive bases thereof and the sequences complementary thereto. Homology may be measured using the alignment algorithm. For example, the homologous polynucleotides may have a coding sequence which is a naturally occurring allelic variant of one of the coding sequences described herein. Such allelic variants may have a substitution, deletion or addition of one or more nucleotides when compared to the nucleic acids of the invention or the sequences complementary thereto.

[0289] Additionally, the above procedures may be used to isolate nucleic acids which encode polypeptides having at least about 99%, 95%, homology to a polypeptide having the sequence of one of amino acid sequences of the invention, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof as determined using a sequence alignment algorithm (e.g., such as the FASTA version 3.0t78 algorithm with the default parameters).

Oligonucleotides probes and methods for using them

[0290] Nucleic acid probes can be used, e.g., for identifying nucleic acids encoding a polypeptide with a xylanase activity or fragments thereof or for identifying xylanase genes. The probe may comprise at least 10 consecutive bases of a nucleic acid of the invention. Alternatively, a probe can be at least about 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 110, 120, 130, 150 or about 10 to 50, about 20 to 60 about 30 to 70, consecutive bases of a sequence as set forth in a nucleic acid of the invention. The probes identify a nucleic acid by binding and/or hybridization. The probes can be used in arrays of the invention, see discussion below, including, e.g., capillary arrays. The probes can also be used to isolate other nucleic acids or polypeptides.

[0291] The isolated nucleic acids of the invention and sequences substantially identical thereto, the sequences complementary thereto, or a fragment comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive bases of one of the sequences of the invention and sequences substantially identical thereto, or the sequences complementary thereto may also be used as probes to determine whether a biological sample, such as a soil sample, contains an organism having a nucleic acid sequence of the invention or an organism from which the nucleic acid was obtained. In such procedures, a biological sample potentially harboring the organism from which the nucleic acid was isolated is obtained and nucleic acids are obtained from the sample. The nucleic acids are contacted with the probe under conditions which permit the probe to specifically hybridize to any complementary sequences from which are present therein.

[0292] Where necessary, conditions which permit the probe to specifically hybridize to complementary sequences may be determined by placing the probe in contact with complementary sequences from samples known to contain the complementary sequence as well as control sequences which do not contain the complementary sequence. Hybridization conditions, such as the salt concentration of the hybridization buffer, the formamide concentration of the hybridization buffer, or the hybridization temperature, may be varied to identify conditions which allow the probe to hybridize specifically to complementary nucleic acids.

[0293] If the sample contains the organism from which the nucleic acid was isolated, specific hybridization

of the probe is then detected. Hybridization may be detected by labeling the probe with a detectable agent such as a radioactive isotope, a fluorescent dye or an enzyme capable of catalyzing the formation of a detectable product.

[0294] Many methods for using the labeled probes to detect the presence of complementary nucleic acids in a sample are familiar to those skilled in the art. These include Southern Blots, Northern Blots, colony hybridization procedures and dot blots. Protocols for each of these procedures are provided in Ausubel et al. *Current Protocols in Molecular Biology*, John Wiley 503 Sons, Inc. (1997) and Sambrook et al., *Molecular Cloning: A Laboratory Manual* 2nd Ed., Cold Spring Harbor Laboratory Press (1989).

[0295] Alternatively, more than one probe (at least one of which is capable of specifically hybridizing to any complementary sequences which are present in the nucleic acid sample), may be used in an amplification reaction to determine whether the sample contains an organism containing a nucleic acid sequence of the invention (e.g., an organism from which the nucleic acid was isolated). Typically, the probes comprise oligonucleotides. The amplification reaction may comprise a PCR reaction. PCR protocols are described in Ausubel and Sambrook, *supra*. Alternatively, the amplification may comprise a ligase chain reaction, 3SR, or strand displacement reaction. (See Barany, F., "The Ligase Chain Reaction in a PCR World", *PCR Methods and Applications* 1:5-16, 1991; E. Fahy et al., "Self-sustained Sequence Replication (3SR): An Isothermal Transcription-based Amplification System Alternative to PCR", *PCR Methods and Applications* 1:25-33, 1991; and Walker G.T. et al., "Strand Displacement Amplification-an Isothermal in vitro DNA Amplification Technique", *Nucleic Acid Research* 20:1691-1696, 1992). In such procedures, the nucleic acids in the sample are contacted with the probes, the amplification reaction is performed and any resulting amplification product is detected. The amplification product may be detected by performing gel electrophoresis on the reaction products and staining the gel with an intercalator such as ethidium bromide. Alternatively, one or more of the probes may be labeled with a radioactive isotope and the presence of a radioactive amplification product may be detected by autoradiography after gel electrophoresis.

[0296] Probes derived from sequences near the ends of the sequences of the invention and sequences substantially identical thereto, may also be used in chromosome walking procedures to identify clones containing genomic sequences located adjacent to the sequences of the invention and sequences substantially identical thereto. Such methods allow the isolation of genes which encode additional proteins from the host organism.

[0297] The isolated nucleic acids of the invention and sequences substantially identical thereto, the sequences complementary thereto, or a fragment comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive bases of one of the sequences of the invention and sequences substantially identical thereto, or the sequences complementary thereto may be used as probes to identify and isolate related nucleic acids. In some aspects, the related nucleic acids may be cDNAs or genomic DNAs from organisms other than the one from which the nucleic acid was isolated. For example, the other organisms may be related organisms. In such procedures, a nucleic acid sample is contacted with the probe under conditions which permit the probe to specifically hybridize to related sequences. Hybridization of the probe to nucleic acids from the related organism is then detected using any of the methods described above.

[0298] By varying the stringency of the hybridization conditions used to identify nucleic acids, such as cDNAs or genomic DNAs, which hybridize to the detectable probe, nucleic acids having different levels of homology to the probe can be identified and isolated. Stringency may be varied by conducting the hybridization at varying temperatures below the melting temperatures of the probes. The melting

temperature, T_m , is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly complementary probe. Very stringent conditions are selected to be equal to or about 5°C lower than the T_m for a particular probe. The melting temperature of the probe may be calculated using the following formulas:

[0299] For probes between 14 and 70 nucleotides in length the melting temperature (T_m) is calculated using the formula: $T_m = 81.5 + 16.6(\log [Na^+]) + 0.41(\text{fraction G+C}) - (600/N)$ where N is the length of the probe.

[0300] If the hybridization is carried out in a solution containing formamide, the melting temperature may be calculated using the equation: $T_m = 81.5 + 16.6(\log [Na^+]) + 0.41(\text{fraction G+C}) - (0.63\% \text{ formamide}) - (600/N)$ where N is the length of the probe.

[0301] Prehybridization may be carried out in 6X SSC, 5X Denhardt's reagent, 0.5% SDS, 100 µg/ml denatured fragmented salmon sperm DNA or 6X SSC, 5X Denhardt's reagent, 0.5% SDS, 100 µg/ml denatured fragmented salmon sperm DNA, 50% formamide. The formulas for SSC and Denhardt's solutions are listed in Sambrook *et al.*, *supra*.

[0302] Hybridization is conducted by adding the detectable probe to the prehybridization solutions listed above. Where the probe comprises double stranded DNA, it is denatured before addition to the hybridization solution. The filter is contacted with the hybridization solution for a sufficient period of time to allow the probe to hybridize to cDNAs or genomic DNAs containing sequences complementary thereto or homologous thereto. For probes over 200 nucleotides in length, the hybridization may be carried out at 15-25°C below the T_m . For shorter probes, such as oligonucleotide probes, the hybridization may be conducted at 5-10°C below the T_m . Typically, for hybridizations in 6X SSC, the hybridization is conducted at approximately 68°C. Usually, for hybridizations in 50% formamide containing solutions, the hybridization is conducted at approximately 42°C.

Inhibiting Expression of Glycosyl Hydrolases

[0303] The invention provides nucleic acids complementary to (e.g., antisense sequences to) the nucleic acids of the invention, e.g., xylanase- encoding nucleic acids. Antisense sequences are capable of inhibiting the transport, splicing or transcription of xylanase-encoding genes. The inhibition can be effected through the targeting of genomic DNA or messenger RNA. The transcription or function of targeted nucleic acid can be inhibited, for example, by hybridization and/or cleavage. One particularly useful set of inhibitors provided by the present invention includes oligonucleotides which are able to either bind xylanase gene or message, in either case preventing or inhibiting the production or function of xylanase. The association can be through sequence specific hybridization. Another useful class of inhibitors includes oligonucleotides which cause inactivation or cleavage of xylanase message. The oligonucleotide can have enzyme activity which causes such cleavage, such as ribozymes. The oligonucleotide can be chemically modified or conjugated to an enzyme or composition capable of cleaving the complementary nucleic acid. A pool of many different such oligonucleotides can be screened for those with the desired activity. Thus, the invention provides various compositions for the inhibition of xylanase expression on a nucleic acid and/or protein level, e.g., antisense, iRNA and ribozymes comprising xylanase sequences of the invention and the anti-xylanase antibodies of the invention.

[0304] Inhibition of xylanase expression can have a variety of industrial, medical, pharmaceutical,

research, food and feed and food and feed supplement processing and other applications and processes. For example, inhibition of xylanase expression can slow or prevent spoilage. Spoilage can occur when polysaccharides, e.g., structural polysaccharides, are enzymatically degraded. This can lead to the deterioration, or rot, of fruits and vegetables. Compositions of the invention that inhibit the expression and/or activity of xylanases, e.g., antibodies, antisense oligonucleotides, ribozymes and RNAi, are used to slow or prevent spoilage. Thus, methods comprise application onto a plant or plant product (e.g., a cereal, a grain, a fruit, seed, root, leaf, etc.) antibodies, antisense oligonucleotides, ribozymes and RNAi of compositions of the invention to slow or prevent spoilage. These compositions also can be expressed by the plant (e.g., a transgenic plant) or another organism (e.g., a bacterium or other microorganism transformed with a xylanase gene of the invention).

[0305] The compositions of the invention for the inhibition of xylanase expression (e.g., antisense, iRNA, ribozymes, antibodies) can be used as pharmaceutical compositions, e.g., as anti-pathogen agents or in other therapies, e.g., as anti-microbials for, e.g., *Salmonella*.

Antisense Oligonucleotides

[0306] The invention provides antisense oligonucleotides capable of binding xylanase message which can inhibit xylan hydrolase activity (e.g., catalyzing hydrolysis of internal β -1,4-xylosidic linkages) by targeting mRNA. Strategies for designing antisense oligonucleotides are well described in the scientific and patent literature, and the skilled artisan can design such xylanase oligonucleotides using the novel reagents of the invention. For example, gene walking/ RNA mapping protocols to screen for effective antisense oligonucleotides are well known in the art, see, e.g., Ho (2000) *Methods Enzymol.* 314:168-183, describing an RNA mapping assay, which is based on standard molecular techniques to provide an easy and reliable method for potent antisense sequence selection. See also Smith (2000) *Eur. J. Pharm. Sci.* 11:191-198.

[0307] Naturally occurring nucleic acids are used as antisense oligonucleotides. The antisense oligonucleotides can be of any length; for example, in alternative aspects, the antisense oligonucleotides are between about 5 to 100, about 10 to 80, about 15 to 60, about 18 to 40. The optimal length can be determined by routine screening. The antisense oligonucleotides can be present at any concentration. The optimal concentration can be determined by routine screening. A wide variety of synthetic, non-naturally occurring nucleotide and nucleic acid analogues are known which can address this potential problem. For example, peptide nucleic acids (PNAs) containing non-ionic backbones, such as N-(2-aminoethyl) glycine units can be used. Antisense oligonucleotides having phosphorothioate linkages can also be used, as described in WO 97/03211; WO 96/39154; Mata (1997) *Toxicol Appl Pharmacol* 144:189-197; *Antisense Therapeutics*, ed. Agrawal (Humana Press, Totowa, N.J., 1996). Antisense oligonucleotides having synthetic DNA backbone analogues provided by the invention can also include phosphoro-dithioate, methylphosphonate, phosphoramidate, alkyl phosphotriester, sulfamate, 3'-thioacetal, methylene(methylimino), 3'-N-carbamate, and morpholino carbamate nucleic acids, as described above. Combinatorial chemistry methodology can be used to create vast numbers of oligonucleotides that can be rapidly screened for specific oligonucleotides that have appropriate binding affinities and specificities toward any target, such as the sense and antisense xylanase sequences of the invention (see, e.g., Gold (1995) *J. of Biol. Chem.* 270: 13581-13584).

Inhibitory Ribozymes

[0308] Ribozymes may be capable of binding xylanase message. These ribozymes can inhibit xylanase

activity by, e.g., targeting mRNA. Strategies for designing ribozymes and selecting the xylanase- specific antisense sequence for targeting are well described in the scientific and patent literature, and the skilled artisan can design such ribozymes using the novel reagents of the invention. Ribozymes act by binding to a target RNA through the target RNA binding portion of a ribozyme which is held in close proximity to an enzymatic portion of the RNA that cleaves the target RNA. Thus, the ribozyme recognizes and binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cleave and inactivate the target RNA. Cleavage of a target RNA in such a manner will destroy its ability to direct synthesis of an encoded protein if the cleavage occurs in the coding sequence. After a ribozyme has bound and cleaved its RNA target, it can be released from that RNA to bind and cleave new targets repeatedly.

[0309] In some circumstances, the enzymatic nature of a ribozyme can be advantageous over other technologies, such as antisense technology (where a nucleic acid molecule simply binds to a nucleic acid target to block its transcription, translation or association with another molecule) as the effective concentration of ribozyme necessary to effect a therapeutic treatment can be lower than that of an antisense oligonucleotide. This potential advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of target RNA. In addition, a ribozyme is typically a highly specific inhibitor, with the specificity of inhibition depending not only on the base pairing mechanism of binding, but also on the mechanism by which the molecule inhibits the expression of the RNA to which it binds. That is, the inhibition is caused by cleavage of the RNA target and so specificity is defined as the ratio of the rate of cleavage of the targeted RNA over the rate of cleavage of non-targeted RNA. This cleavage mechanism is dependent upon factors additional to those involved in base pairing. Thus, the specificity of action of a ribozyme can be greater than that of antisense oligonucleotide binding the same RNA site.

[0310] The ribozyme, e.g., an enzymatic ribozyme RNA molecule, can be formed in a hammerhead motif, a hairpin motif, as a hepatitis delta virus motif, a group I intron motif and/or an RNaseP-like RNA in association with an RNA guide sequence. Examples of hammerhead motifs are described by, e.g., Rossi (1992) *Aids Research and Human Retroviruses* 8:183; hairpin motifs by Hampel (1989) *Biochemistry* 28:4929, and Hampel (1990) *Nuc. Acids Res.* 18:299; the hepatitis delta virus motif by Perrotta (1992) *Biochemistry* 31:16; the RNaseP motif by Guerrier-Takada (1983) *Cell* 35:849; and the group I intron by Cech U.S. Pat. No. 4,987,071. The recitation of these specific motifs is not intended to be limiting. Those skilled in the art will recognize that a ribozyme, e.g., an enzymatic RNA molecule, can have a specific substrate binding site complementary to one or more of the target gene RNA regions. A ribozyme can have a nucleotide sequence within or surrounding that substrate binding site which imparts an RNA cleaving activity to the molecule.

RNA interference (RNAi)

[0311] An RNA inhibitory molecule, a so-called "RNAi" molecule may comprise a xylanase enzyme sequence of the invention. The RNAi molecule can comprise a double-stranded RNA (dsRNA) molecule, e.g., siRNA, miRNA and/or short hairpin RNA (shRNA) molecules. The RNAi molecule, e.g., siRNA (small inhibitory RNA) can inhibit expression of a xylanase enzyme gene, and/or miRNA (micro RNA) to inhibit translation of a xylanase message. In one aspect, the RNAi molecule, e.g., siRNA and/or miRNA, is about 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or more duplex nucleotides in length. While not limited by any particular mechanism of action, the RNAi can enter a cell and cause the degradation of a single-stranded RNA (ssRNA) of similar or identical sequences, including endogenous mRNAs. When a cell is exposed to double-stranded RNA (dsRNA), mRNA from the homologous gene is

selectively degraded by a process called RNA interference (RNAi). A possible basic mechanism behind RNAi is the breaking of a double-stranded RNA (dsRNA) matching a specific gene sequence into short pieces called short interfering RNA, which trigger the degradation of mRNA that matches its sequence. The RNAi's may be used in gene-silencing therapeutics, see, e.g., Shuey (2002) Drug Discov. Today 7:1040-1046. Methods to selectively degrade RNA may use the said RNAi's molecules, e.g., siRNA and/or miRNA. The process may be practiced *in vitro*, *ex vivo* or *in vivo*. The RNAi molecules can be used to generate a loss-of-function mutation in a cell, an organ or an animal.

[0312] Intracellular introduction of the RNAi may be by internalization of a target cell specific ligand bonded to an RNA binding protein comprising an RNAi (e.g., microRNA) is adsorbed. The ligand is specific to a unique target cell surface antigen. The ligand can be spontaneously internalized after binding to the cell surface antigen. If the unique cell surface antigen is not naturally internalized after binding to its ligand, internalization can be promoted by the incorporation of an arginine-rich peptide, or other membrane permeable peptide, into the structure of the ligand or RNA binding protein or attachment of such a peptide to the ligand or RNA binding protein. See, e.g., U.S. Patent App. Pub. Nos. 20060030003; 20060025361; 20060019286; 20060019258. Lipid-based formulations may deliver, e.g., introducing nucleic acids of the invention as nucleic acid-lipid particles comprising an RNAi molecule to a cell, see .g., U.S. Patent App. Pub. No. 20060008910.

[0313] Methods for making and using RNAi molecules, e.g., siRNA and/or miRNA, for selectively degrade RNA are well known in the art, see, e.g., U.S. Patent No. 6,506,559; 6,511,824; 6,515,109; 6,489,127.

Modification of Nucleic Acids

[0314] Methods of generating variants of the nucleic acids of the invention, e.g., those encoding a xylanase can be repeated or used in various combinations to generate xylanases having an altered or different activity or an altered or different stability from that of a xylanase encoded by the template nucleic acid. These methods also can be repeated or used in various combinations, e.g., to generate variations in gene/ message expression, message translation or message stability. In another aspect, the genetic composition of a cell is altered by, e.g., modification of a homologous gene *ex vivo*, followed by its reinsertion into the cell.

[0315] A nucleic acid of the invention can be altered by any means. For example, random or stochastic methods, or, non-stochastic, or "directed evolution," methods, see, e.g., U.S.

[0316] Patent No. 6,361,974. Methods for random mutation of genes are well known in the art, see, e.g., U.S. Patent No. 5,830,696. For example, mutagens can be used to randomly mutate a gene. Mutagens include, e.g., ultraviolet light or gamma irradiation, or a chemical mutagen, e.g., mitomycin, nitrous acid, photoactivated psoralens, alone or in combination, to induce DNA breaks amenable to repair by recombination. Other chemical mutagens include, for example, sodium bisulfite, nitrous acid, hydroxylamine, hydrazine or formic acid. Other mutagens are analogues of nucleotide precursors, e.g., nitrosoguanidine, 5-bromouracil, 2-aminopurine, or acridine. These agents can be added to a PCR reaction in place of the nucleotide precursor thereby mutating the sequence. Intercalating agents such as proflavine, acriflavine, quinacrine and the like can also be used.

[0317] Any technique in molecular biology can be used, e.g., random PCR mutagenesis, see, e.g., Rice (1992) Proc. Natl. Acad. Sci. USA 89:5467-5471; or, combinatorial multiple cassette mutagenesis, see, e.g., Crameri (1995) Biotechniques 18:194-196. Alternatively, nucleic acids, e.g., genes, can be

reassembled after random, or "stochastic," fragmentation, see, e.g., U.S. Patent Nos. 6,291,242; 6,287,862; 6,287,861; 5,955,358; 5,830,721; 5,824,514; 5,811,238; 5,605,793. In alternative aspects, modifications, additions or deletions are introduced by error-prone PCR, shuffling, oligonucleotide-directed mutagenesis, assembly PCR, sexual PCR mutagenesis, in vivo mutagenesis, cassette mutagenesis, recursive ensemble mutagenesis, exponential ensemble mutagenesis, site-specific mutagenesis, gene reassembly (e.g., GeneReassembly, see, e.g., U.S. Patent No. 6,537,776), gene site saturation mutagenesis (GSSM), synthetic ligation reassembly (SLR), recombination, recursive sequence recombination, phosphothioate-modified DNA mutagenesis, uracil-containing template mutagenesis, gapped duplex mutagenesis, point mismatch repair mutagenesis, repair-deficient host strain mutagenesis, chemical mutagenesis, radiogenic mutagenesis, deletion mutagenesis, restriction-selection mutagenesis, restriction-purification mutagenesis, artificial gene synthesis, ensemble mutagenesis, chimeric nucleic acid multimer creation, and/or a combination of these and other methods.

[0318] The following publications describe a variety of recursive recombination procedures and/or methods which can be used: Stemmer (1999) "Molecular breeding of viruses for targeting and other clinical properties" *Tumor Targeting* 4:1-4; Ness (1999) *Nature Biotechnology* 17:893-896; Chang (1999) "Evolution of a cytokine using DNA family shuffling" *Nature Biotechnology* 17:793-797; Minshull (1999) "Protein evolution by molecular breeding" *Current Opinion in Chemical Biology* 3:284-290; Christians (1999) "Directed evolution of thymidine kinase for AZT phosphorylation using DNA family shuffling" *Nature Biotechnology* 17:259-264; Cramer (1998) "DNA shuffling of a family of genes from diverse species accelerates directed evolution" *Nature* 391:288-291; Cramer (1997) "Molecular evolution of an arsenate detoxification pathway by DNA shuffling," *Nature Biotechnology* 15:436-438; Zhang (1997) "Directed evolution of an effective fucosidase from a galactosidase by DNA shuffling and screening" *Proc. Natl. Acad. Sci. USA* 94:4504-4509; Patten et al. (1997) "Applications of DNA Shuffling to Pharmaceuticals and Vaccines" *Current Opinion in Biotechnology* 8:724-733; Cramer et al. (1996) "Construction and evolution of antibody-phage libraries by DNA shuffling" *Nature Medicine* 2:100-103; Gates et al. (1996) "Affinity selective isolation of ligands from peptide libraries through display on a lac repressor 'headpiece dimer'" *Journal of Molecular Biology* 255:373-386; Stemmer (1996) "Sexual PCR and Assembly PCR" In: *The Encyclopedia of Molecular Biology*. VCH Publishers, New York. pp.447-457; Cramer and Stemmer (1995) "Combinatorial multiple cassette mutagenesis creates all the permutations of mutant and wildtype cassettes" *BioTechniques* 18:194-195; Stemmer et al. (1995) "Single-step assembly of a gene and entire plasmid from large numbers of oligodeoxyribonucleotides" *Gene*, 164:49-53; Stemmer (1995) "The Evolution of Molecular Computation" *Science* 270: 1510; Stemmer (1995) "Searching Sequence Space" *Bio/Technology* 13:549-553; Stemmer (1994) "Rapid evolution of a protein in vitro by DNA shuffling" *Nature* 370:389-391; and Stemmer (1994) "DNA shuffling by random fragmentation and reassembly: In vitro recombination for molecular evolution." *Proc. Natl. Acad. Sci. USA* 91:10747-10751.

[0319] Mutational methods of generating diversity include, for example, site-directed mutagenesis (Ling et al. (1997) "Approaches to DNA mutagenesis: an overview" *Anal Biochem.* 254(2): 157-178; Dale et al. (1996) "Oligonucleotide-directed random mutagenesis using the phosphorothioate method" *Methods Mol. Biol.* 57:369-374; Smith (1985) "In vitro mutagenesis" *Ann. Rev. Genet.* 19:423-462; Botstein & Shortle (1985) "Strategies and applications of in vitro mutagenesis" *Science* 229:1193-1201; Carter (1986) "Site-directed mutagenesis" *Biochem. J.* 237:1-7; and Kunkel (1987) "The efficiency of oligonucleotide directed mutagenesis" in *Nucleic Acids & Molecular Biology* (Eckstein, F. and Lilley, D. M. J. eds., Springer Verlag, Berlin)); mutagenesis using uracil containing templates (Kunkel (1985) "Rapid and efficient site-specific mutagenesis without phenotypic selection" *Proc. Natl. Acad. Sci. USA* 82:488-492; Kunkel et al. (1987) "Rapid and efficient site-specific mutagenesis without phenotypic selection" *Methods in Enzymol.* 154, 367-382; and Bass et al. (1988) "Mutant Trp repressors with new DNA-binding specificities" *Science* 242:240-245); oligonucleotide-directed mutagenesis (*Methods in Enzymol.* 100: 468-500 (1983); *Methods in Enzymol.* 154: 329-350 (1987); Zoller (1982) "Oligonucleotide-directed mutagenesis using M13-derived

vectors: an efficient and general procedure for the production of point mutations in any DNA fragment" *Nucleic Acids Res.* 10:6487-6500; Zoller & Smith (1983) "Oligonucleotide-directed mutagenesis of DNA fragments cloned into M13 vectors" *Methods in Enzymol.* 100:468-500; and Zoller (1987) "Oligonucleotide-directed mutagenesis: a simple method using two oligonucleotide primers and a single-stranded DNA template" *Methods in Enzymol.* 154:329-350; phosphorothioate-modified DNA mutagenesis (Taylor (1985) "The use of phosphorothioate-modified DNA in restriction enzyme reactions to prepare nicked DNA" *Nucl. Acids Res.* 13: 8749-8764; Taylor (1985) "The rapid generation of oligonucleotide-directed mutations at high frequency using phosphorothioate-modified DNA" *Nucl. Acids Res.* 13: 8765-8787 (1985); Nakamaye (1986) "Inhibition of restriction endonuclease Nci I cleavage by phosphorothioate groups and its application to oligonucleotide-directed mutagenesis" *Nucl. Acids Res.* 14: 9679-9698; Sayers (1988) "Y-T Exonucleases in phosphorothioate-based oligonucleotide-directed mutagenesis" *Nucl. Acids Res.* 16:791-802; and Sayers et al. (1988) "Strand specific cleavage of phosphorothioate-containing DNA by reaction with restriction endonucleases in the presence of ethidium bromide" *Nucl. Acids Res.* 16: 803-814); mutagenesis using gapped duplex DNA (Kramer et al. (1984) "The gapped duplex DNA approach to oligonucleotide-directed mutation construction" *Nucl. Acids Res.* 12: 9441-9456; Kramer & Fritz (1987) *Methods in Enzymol.* "Oligonucleotide-directed construction of mutations via gapped duplex DNA" 154:350-367; Kramer (1988) "Improved enzymatic in vitro reactions in the gapped duplex DNA approach to oligonucleotide-directed construction of mutations" *Nucl. Acids Res.* 16: 7207; and Fritz (1988) "Oligonucleotide-directed construction of mutations: a gapped duplex DNA procedure without enzymatic reactions in vitro" *Nucl. Acids Res.* 16: 6987-6999).

[0320] Additional protocols that can be used include point mismatch repair (Kramer (1984) "Point Mismatch Repair" *Cell* 38:879-887), mutagenesis using repair-deficient host strains (Carter et al. (1985) "Improved oligonucleotide site-directed mutagenesis using M13 vectors" *Nucl. Acids Res.* 13: 4431-4443; and Carter (1987) "Improved oligonucleotide-directed mutagenesis using M13 vectors" *Methods in Enzymol.* 154: 382-403), deletion mutagenesis (Eghtedarzadeh (1986) "Use of oligonucleotides to generate large deletions" *Nucl. Acids Res.* 14: 5115), restriction-selection and restriction-selection and restriction-purification (Wells et al. (1986) "Importance of hydrogen-bond formation in stabilizing the transition state of subtilisin" *Phil. Trans. R. Soc. Lond. A* 317: 415-423), mutagenesis by total gene synthesis (Nambiar et al. (1984) "Total synthesis and cloning of a gene coding for the ribonuclease S protein" *Science* 223: 1299-1301; Sakamar and Khorana (1988) "Total synthesis and expression of a gene for the α -subunit of bovine rod outer segment guanine nucleotide-binding protein (transducin)" *Nucl. Acids Res.* 14: 6361-6372; Wells et al. (1985) "Cassette mutagenesis: an efficient method for generation of multiple mutations at defined sites" *Gene* 34:315-323; and Grundstrom et al. (1985) "Oligonucleotide-directed mutagenesis by microscale 'shot-gun' gene synthesis" *Nucl. Acids Res.* 13: 3305-3316), double-strand break repair (Mandecki (1986); Arnold (1993) "Protein engineering for unusual environments" *Current Opinion in Biotechnology* 4:450-455. "Oligonucleotide-directed double-strand break repair in plasmids of *Escherichia coli*: a method for site-specific mutagenesis" *Proc. Natl. Acad. Sci. USA*, 83:7177-7181). Additional details on many of the above methods can be found in *Methods in Enzymology* Volume 154, which also describes useful controls for trouble-shooting problems with various mutagenesis methods.

[0321] Protocols that can be used are described, e.g., in U.S. Patent Nos. 5,605,793 to Stemmer (Feb. 25, 1997), "Methods for In Vitro Recombination;" U.S. Pat. No. 5,811,238 to Stemmer et al. (Sep. 22, 1998) "Methods for Generating Polynucleotides having Desired Characteristics by Iterative Selection and Recombination;" U.S. Pat. No. 5,830,721 to Stemmer et al. (Nov. 3, 1998), "DNA Mutagenesis by Random Fragmentation and Reassembly;" U.S. Pat. No. 5,834,252 to Stemmer, et al. (Nov. 10, 1998) "End-Complementary Polymerase Reaction;" U.S. Pat. No. 5,837,458 to Minshull, et al. (Nov. 17, 1998), "Methods and Compositions for Cellular and Metabolic Engineering;" WO 95/22625, Stemmer and

Cramer, "Mutagenesis by Random Fragmentation and Reassembly;" WO 96/33207 by Stemmer and Lipschutz "End Complementary Polymerase Chain Reaction;" WO 97/20078 by Stemmer and Cramer "Methods for Generating Polynucleotides having Desired Characteristics by Iterative Selection and Recombination;" WO 97/35966 by Minshull and Stemmer, "Methods and Compositions for Cellular and Metabolic Engineering;" WO 99/41402 by Punnonen et al. "Targeting of Genetic Vaccine Vectors;" WO 99/41383 by Punnonen et al. "Antigen Library Immunization;" WO 99/41369 by Punnonen et al. "Genetic Vaccine Vector Engineering;" WO 99/41368 by Punnonen et al. "Optimization of Immunomodulatory Properties of Genetic Vaccines;" EP 752008 by Stemmer and Cramer, "DNA Mutagenesis by Random Fragmentation and Reassembly;" EP 0932670 by Stemmer "Evolving Cellular DNA Uptake by Recursive Sequence Recombination;" WO 99/23107 by Stemmer et al., "Modification of Virus Tropism and Host Range by Viral Genome Shuffling;" WO 99/21979 by Apt et al., "Human Papillomavirus Vectors;" WO 98/31837 by del Cardayre et al. "Evolution of Whole Cells and Organisms by Recursive Sequence Recombination;" WO 98/27230 by Patten and Stemmer, "Methods and Compositions for Polypeptide Engineering;" WO 98/27230 by Stemmer et al., "Methods for Optimization of Gene Therapy by Recursive Sequence Shuffling and Selection," WO 00/00632, "Methods for Generating Highly Diverse Libraries," WO 00/09679, "Methods for Obtaining in Vitro Recombined Polynucleotide Sequence Banks and Resulting Sequences," WO 98/42832 by Arnold et al., "Recombination of Polynucleotide Sequences Using Random or Defined Primers," WO 99/29902 by Arnold et al., "Method for Creating Polynucleotide and Polypeptide Sequences," WO 98/41653 by Vind, "An in Vitro Method for Construction of a DNA Library," WO 98/41622 by Borchert et al., "Method for Constructing a Library Using DNA Shuffling," and WO 98/42727 by Pati and Zarling, "Sequence Alterations using Homologous Recombination."

[0322] Protocols that can be used (providing details regarding various diversity generating methods) are described, e.g., in U.S. Patent application serial no. (USSN) 09/407,800, "SHUFFLING OF CODON ALTERED GENES" by Patten et al. filed Sep. 28, 1999; "EVOLUTION OF WHOLE CELLS AND ORGANISMS BY RECURSIVE SEQUENCE RECOMBINATION" by del Cardayre et al., United States Patent No. 6,379,964; "OLIGONUCLEOTIDE MEDIATED NUCLEIC ACID RECOMBINATION" by Cramer et al., United States Patent Nos. 6,319,714; 6,368,861; 6,376,246; 6,423,542; 6,426,224 and PCT/US00/01203; "USE OF CODON-VARIED OLIGONUCLEOTIDE SYNTHESIS FOR SYNTHETIC SHUFFLING" by Welch et al., United States Patent No. 6,436,675; "METHODS FOR MAKING CHARACTER STRINGS, POLYNUCLEOTIDES & POLYPEPTIDES HAVING DESIRED CHARACTERISTICS" by Selifonov et al., filed Jan. 18, 2000, (PCT/US00/01202) and, e.g. "METHODS FOR MAKING CHARACTER STRINGS, POLYNUCLEOTIDES & POLYPEPTIDES HAVING DESIRED CHARACTERISTICS" by Selifonov et al., filed Jul. 18, 2000 (U.S. Ser. No. 09/618,579); "METHODS OF POPULATING DATA STRUCTURES FOR USE IN EVOLUTIONARY SIMULATIONS" by Selifonov and Stemmer, filed Jan. 18, 2000 (PCT/US00/01138); and "SINGLE-STRANDED NUCLEIC ACID TEMPLATE-MEDIATED RECOMBINATION AND NUCLEIC ACID FRAGMENT ISOLATION" by Affholter, filed Sep. 6, 2000 (U.S. Ser. No. 09/656,549); and United States Patent Nos. 6,177,263; 6,153,410.

[0323] Non-stochastic, or "directed evolution," methods include, e.g., saturation mutagenesis (GSSM), synthetic ligation reassembly (SLR), or a combination thereof are used to modify the nucleic acids of the invention to generate xylanases with new or altered properties (e.g., activity under highly acidic or alkaline conditions, high or low temperatures, and the like). Polypeptides encoded by the modified nucleic acids can be screened for an activity before testing for xylan hydrolysis or other activity. Any testing modality or protocol can be used, e.g., using a capillary array platform. See, e.g., U.S. Patent Nos. 6,361,974; 6,280,926; 5,939,250.

Gene Site Saturation Mutagenesis, or, GSSM

[0324] Methods for making enzyme using Gene Site Saturation mutagenesis, or, GSSM, are described herein, and also in U.S. Patent Nos. 6,171,820 and 6,579,258. Codon primers containing a degenerate N,N,G/T sequence may be used to introduce point mutations into a polynucleotide, e.g., a xylanase or an antibody of the invention, so as to generate a set of progeny polypeptides in which a full range of single amino acid substitutions is represented at each amino acid position, e.g., an amino acid residue in an enzyme active site or ligand binding site targeted to be modified. These oligonucleotides can comprise a contiguous first homologous sequence, a degenerate N,N,G/T sequence, and, in one aspect, a second homologous sequence. The downstream progeny translational products from the use of such oligonucleotides include all possible amino acid changes at each amino acid site along the polypeptide, because the degeneracy of the N,N,G/T sequence includes codons for all 20 amino acids. One such degenerate oligonucleotide (comprised of, e.g., one degenerate N,N,G/T cassette) may be used for subjecting each original codon in a parental polynucleotide template to a full range of codon substitutions. At least two degenerate cassettes may be used - either in the same oligonucleotide or not, for subjecting at least two original codons in a parental polynucleotide template to a full range of codon substitutions. For example, more than one N,N,G/T sequence can be contained in one oligonucleotide to introduce amino acid mutations at more than one site. This plurality of N,N,G/T sequences can be directly contiguous, or separated by one or more additional nucleotide sequence(s). Oligonucleotides serviceable for introducing additions and deletions can be used either alone or in combination with the codons containing an N,N,G/T sequence, to introduce any combination or permutation of amino acid additions, deletions, and/or substitutions.

[0325] Simultaneous mutagenesis of two or more contiguous amino acid positions may be done using an oligonucleotide that contains contiguous N,N,G/T triplets, i.e. a degenerate (N,N,G/T)_n sequence. Degenerate cassettes having less degeneracy than the N,N,G/T sequence may be used. For example, it may be desirable in some instances to use (e.g. in an oligonucleotide) a degenerate triplet sequence comprised of only one N, where said N can be in the first second or third position of the triplet. Any other bases including any combinations and permutations thereof can be used in the remaining two positions of the triplet. Alternatively, it may be desirable in some instances to use (e.g. in an oligo) a degenerate N,N,N triplet sequence.

[0326] In one aspect, use of degenerate triplets (e.g., N,N,G/T triplets) allows for systematic and easy generation of a full range of possible natural amino acids (for a total of 20 amino acids) into each and every amino acid position in a polypeptide (in alternative aspects, the methods also include generation of less than all possible substitutions per amino acid residue, or codon, position). For example, for a 100 amino acid polypeptide, 2000 distinct species (i.e. 20 possible amino acids per position X 100 amino acid positions) can be generated. Through the use of an oligonucleotide or set of oligonucleotides containing a degenerate N,N,G/T triplet, 32 individual sequences can code for all 20 possible natural amino acids. Thus, in a reaction vessel in which a parental polynucleotide sequence is subjected to saturation mutagenesis using at least one such oligonucleotide, there are generated 32 distinct progeny polynucleotides encoding 20 distinct polypeptides. In contrast, the use of a non-degenerate oligonucleotide in site-directed mutagenesis leads to only one progeny polypeptide product per reaction vessel. Nondegenerate oligonucleotides can be used in combination with degenerate primers disclosed; for example, nondegenerate oligonucleotides can be used to generate specific point mutations in a working polynucleotide. This provides one means to generate specific silent point mutations, point mutations leading to corresponding amino acid changes, and point mutations that cause the generation of stop codons and the corresponding expression of polypeptide fragments.

[0327] Each saturation mutagenesis reaction vessel may contain polynucleotides encoding at least 20 progeny polypeptide molecules such that all 20 natural amino acids are represented at the one specific

amino acid position corresponding to the codon position mutagenized in the parental polynucleotide (other aspects use less than all 20 natural combinations). The 32-fold degenerate progeny polypeptides generated from each saturation mutagenesis reaction vessel can be subjected to clonal amplification (e.g. cloned into a suitable host, e.g., *E. coli* host, using, e.g., an expression vector) and subjected to expression screening. When an individual progeny polypeptide is identified by screening to display a favorable change in property (when compared to the parental polypeptide, such as increased xylan hydrolysis activity under alkaline or acidic conditions), it can be sequenced to identify the correspondingly favorable amino acid substitution contained therein.

[0328] Upon mutagenizing each and every amino acid position in a parental polypeptide using saturation mutagenesis as disclosed herein, favorable amino acid changes may be identified at more than one amino acid position. One or more new progeny molecules can be generated that contain a combination of all or part of these favorable amino acid substitutions. For example, if 2 specific favorable amino acid changes are identified in each of 3 amino acid positions in a polypeptide, the permutations include 3 possibilities at each position (no change from the original amino acid, and each of two favorable changes) and 3 positions. Thus, there are $3 \times 3 \times 3$ or 27 total possibilities, including 7 that were previously examined - 6 single point mutations (i.e. 2 at each of three positions) and no change at any position.

[0329] Site-saturation mutagenesis can be used together with shuffling, chimerization, recombination and other mutagenizing processes, along with screening. This provides for the use of any mutagenizing process(es), including saturation mutagenesis, in an iterative manner. The iterative use of any mutagenizing process(es) may be used in combination with screening.

[0330] Proprietary codon primers (containing a degenerate N,N,N sequence) may be used to introduce point mutations into a polynucleotide, so as to generate a set of progeny polypeptides in which a full range of single amino acid substitutions is represented at each amino acid position (gene site saturation mutagenesis (GSSM)). The oligos used are comprised contiguously of a first homologous sequence, a degenerate N,N,N sequence and preferably but not necessarily a second homologous sequence. The downstream progeny translational products from the use of such oligos include all possible amino acid changes at each amino acid site along the polypeptide, because the degeneracy of the N,N,N sequence includes codons for all 20 amino acids.

[0331] One such degenerate oligo (comprised of one degenerate N,N,N cassette) may be used for subjecting each original codon in a parental polynucleotide template to a full range of codon substitutions. Or, at least two degenerate N,N,N cassettes are used - either in the same oligo or not, for subjecting at least two original codons in a parental polynucleotide template to a full range of codon substitutions. Thus, more than one N,N,N sequence can be contained in one oligo to introduce amino acid mutations at more than one site. This plurality of N,N,N sequences can be directly contiguous, or separated by one or more additional nucleotide sequence(s). Oligos serviceable for introducing additions and deletions can be used either alone or in combination with the codons containing an N,N,N sequence, to introduce any combination or permutation of amino acid additions, deletions and/or substitutions.

[0332] It is possible to simultaneously mutagenize two or more contiguous amino acid positions using an oligo that contains contiguous N,N,N triplets, i.e. a degenerate (N,N,N)_n sequence.

[0333] Degenerate cassettes having less degeneracy than the N,N,N sequence may be used. For example, it may be desirable in some instances to use (e.g. in an oligo) a degenerate triplet sequence comprised of only one N, where the N can be in the first second or third position of the triplet. Any other bases including any combinations and permutations thereof can be used in the remaining two positions of

the triplet. Alternatively, it may be desirable in some instances to use (e.g., in an oligo) a degenerate N,N,N triplet sequence, N,N,G/T, or an N,N, G/C triplet sequence.

[0334] It is appreciated, however, that the use of a degenerate triplet (such as N,N,G/T or an N,N, G/C triplet sequence) as disclosed is advantageous for several reasons. This may provide a means to systematically and fairly easily generate the substitution of the full range of possible amino acids (for a total of 20 amino acids) into each and every amino acid position in a polypeptide. Thus, for a 100 amino acid polypeptide, this provides a way to systematically and fairly easily generate 2000 distinct species (*i.e.*, 20 possible amino acids per position times 100 amino acid positions). It is appreciated that there is provided, through the use of an oligo containing a degenerate N,N,G/T or an N,N, G/C triplet sequence, 32 individual sequences that code for 20 possible amino acids. Thus, in a reaction vessel in which a parental polynucleotide sequence is subjected to saturation mutagenesis using one such oligo, there are generated 32 distinct progeny polynucleotides encoding 20 distinct polypeptides. In contrast, the use of a non-degenerate oligo in site-directed mutagenesis leads to only one progeny polypeptide product per reaction vessel.

[0335] Nondegenerate oligos can be used in combination with degenerate primers disclosed. It is appreciated that in some situations, it is advantageous to use nondegenerate oligos to generate specific point mutations in a working polynucleotide. This provides a means to generate specific silent point mutations, point mutations leading to corresponding amino acid changes and point mutations that cause the generation of stop codons and the corresponding expression of polypeptide fragments. Thus, each saturation mutagenesis reaction vessel contains polynucleotides encoding at least 20 progeny polypeptide molecules such that all 20 amino acids are represented at the one specific amino acid position corresponding to the codon position mutagenized in the parental polynucleotide. The 32-fold degenerate progeny polypeptides generated from each saturation mutagenesis reaction vessel can be subjected to clonal amplification (e.g., cloned into a suitable *E. coli* host using an expression vector) and subjected to expression screening. When an individual progeny polypeptide is identified by screening to display a favorable change in property (when compared to the parental polypeptide), it can be sequenced to identify the correspondingly favorable amino acid substitution contained therein.

[0336] It is appreciated that upon mutagenizing each and every amino acid position in a parental polypeptide using saturation mutagenesis as disclosed herein, favorable amino acid changes may be identified at more than one amino acid position. One or more new progeny molecules can be generated that contain a combination of all or part of these favorable amino acid substitutions. For example, if 2 specific favorable amino acid changes are identified in each of 3 amino acid positions in a polypeptide, the permutations include 3 possibilities at each position (no change from the original amino acid and each of two favorable changes) and 3 positions. Thus, there are $3 \times 3 \times 3$ or 27 total possibilities, including 7 that were previously examined - 6 single point mutations (*i.e.*, 2 at each of three positions) and no change at any position.

[0337] Thus, saturation mutagenesis may be used in combination with additional mutagenization processes, such as process where two or more related polynucleotides are introduced into a suitable host cell such that a hybrid polynucleotide is generated by recombination and reductive reassortment.

[0338] In addition to performing mutagenesis along the entire sequence of a gene, the disclosure provides that mutagenesis can be used to replace each of any number of bases in a polynucleotide sequence, wherein the number of bases to be mutagenized is preferably every integer from 15 to 100,000. Thus, instead of mutagenizing every position along a molecule, one can subject every or a discrete number of bases (preferably a subset totaling from 15 to 100,000) to mutagenesis. Preferably, a separate nucleotide is used for mutagenizing each position or group of positions along a polynucleotide sequence. A group of 3

positions to be mutagenized may be a codon. The mutations are preferably introduced using a mutagenic primer, containing a heterologous cassette, also referred to as a mutagenic cassette. Exemplary cassettes can have from 1 to 500 bases. Each nucleotide position in such heterologous cassettes be N, A, C, G, T, A/C, A/G, A/T, C/G, C/T, G/T, C/G/T, A/G/T, A/C/T, A/C/G, or E, where E is any base that is not A, C, G, or T (E can be referred to as a designer oligo).

[0339] In a general sense, saturation mutagenesis is comprised of mutagenizing a complete set of mutagenic cassettes (wherein each cassette is preferably about 1-500 bases in length) in defined polynucleotide sequence to be mutagenized (wherein the sequence to be mutagenized is preferably from about 15 to 100,000 bases in length). Thus, a group of mutations (ranging from 1 to 100 mutations) is introduced into each cassette to be mutagenized. A grouping of mutations to be introduced into one cassette can be different or the same from a second grouping of mutations to be introduced into a second cassette during the application of one round of saturation mutagenesis. Such groupings are exemplified by deletions, additions, groupings of particular codons and groupings of particular nucleotide cassettes.

[0340] Defined sequences to be mutagenized include a whole gene, pathway, cDNA, an entire open reading frame (ORF) and entire promoter, enhancer, repressor/transactivator, origin of replication, intron, operator, or any polynucleotide functional group. Generally, a "defined sequences" for this purpose may be any polynucleotide that a 15 base-polynucleotide sequence and polynucleotide sequences of lengths between 15 bases and 15,000 bases. Considerations in choosing groupings of codons include types of amino acids encoded by a degenerate mutagenic cassette.

[0341] A grouping of mutations that can be introduced into a mutagenic cassette, includes degenerate codon substitutions (using degenerate oligos) that code for 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19 and 20 amino acids at each position and a library of polypeptides encoded thereby.

Synthetic Ligation Reassembly (SLR)

[0342] A non-stochastic gene modification system termed "synthetic ligation reassembly," or simply "SLR," a "directed evolution process," may be used to generate polypeptides, e.g., xylanases or antibodies of the invention, with new or altered properties.

[0343] SLR is a method of ligating oligonucleotide fragments together non-stochastically. This method differs from stochastic oligonucleotide shuffling in that the nucleic acid building blocks are not shuffled, concatenated or chimerized randomly, but rather are assembled non-stochastically. See, e.g., U.S. Patent Nos. 6,773,900; 6,740,506; 6,713,282; 6,635,449; 6,605,449; 6,537,776. SLR comprises: (a) providing a template polynucleotide, wherein the template polynucleotide comprises sequence encoding a homologous gene; (b) providing a plurality of building block polynucleotides, wherein the building block polynucleotides are designed to cross-over reassemble with the template polynucleotide at a predetermined sequence, and a building block polynucleotide comprises a sequence that is a variant of the homologous gene and a sequence homologous to the template polynucleotide flanking the variant sequence; (c) combining a building block polynucleotide with a template polynucleotide such that the building block polynucleotide cross-over reassembles with the template polynucleotide to generate polynucleotides comprising homologous gene sequence variations.

[0344] SLR does not depend on the presence of high levels of homology between polynucleotides to be rearranged. Thus, this method can be used to non-stochastically generate libraries (or sets) of progeny molecules comprised of over 10^{100} different chimeras. SLR can be used to generate libraries comprised of

over 10^{1000} different progeny chimeras. Non-stochastic methods may produce a set of finalized chimeric nucleic acid molecules having an overall assembly order that is chosen by design. This method includes the steps of generating by design a plurality of specific nucleic acid building blocks having serviceable mutually compatible ligatable ends, and assembling these nucleic acid building blocks, such that a designed overall assembly order is achieved.

[0345] The mutually compatible ligatable ends of the nucleic acid building blocks to be assembled are considered to be "serviceable" for this type of ordered assembly if they enable the building blocks to be coupled in predetermined orders. Thus, the overall assembly order in which the nucleic acid building blocks can be coupled is specified by the design of the ligatable ends. If more than one assembly step is to be used, then the overall assembly order in which the nucleic acid building blocks can be coupled is also specified by the sequential order of the assembly step(s). The annealed building pieces may be treated with an enzyme, such as a ligase (e.g. T4 DNA ligase), to achieve covalent bonding of the building pieces.

[0346] The design of the oligonucleotide building blocks may be obtained by analyzing a set of progenitor nucleic acid sequence templates that serve as a basis for producing a progeny set of finalized chimeric polynucleotides. These parental oligonucleotide templates thus serve as a source of sequence information that aids in the design of the nucleic acid building blocks that are to be mutagenized, e.g., chimerized or shuffled. The sequences of a plurality of parental nucleic acid templates may be aligned in order to select one or more demarcation points. The demarcation points can be located at an area of homology, and are comprised of one or more nucleotides. These demarcation points are preferably shared by at least two of the progenitor templates. The demarcation points can thereby be used to delineate the boundaries of oligonucleotide building blocks to be generated in order to rearrange the parental polynucleotides. The demarcation points identified and selected in the progenitor molecules serve as potential chimerization points in the assembly of the final chimeric progeny molecules. A demarcation point can be an area of homology (comprised of at least one homologous nucleotide base) shared by at least two parental polynucleotide sequences. Alternatively, a demarcation point can be an area of homology that is shared by at least half of the parental polynucleotide sequences, or, it can be an area of homology that is shared by at least two thirds of the parental polynucleotide sequences. Even more preferably a serviceable demarcation points is an area of homology that is shared by at least three fourths of the parental polynucleotide sequences, or, it can be shared by at almost all of the parental polynucleotide sequences. A demarcation point may be an area of homology that is shared by all of the parental polynucleotide sequences.

[0347] A ligation reassembly process may be performed exhaustively in order to generate an exhaustive library of progeny chimeric polynucleotides. In other words, all possible ordered combinations of the nucleic acid building blocks are represented in the set of finalized chimeric nucleic acid molecules. At the same time, in another aspect, the assembly order (i.e. the order of assembly of each building block in the 5' to 3' sequence of each finalized chimeric nucleic acid) in each combination is by design (or non-stochastic) as described above. Because of the non-stochastic nature, the possibility of unwanted side products is greatly reduced.

[0348] The ligation reassembly method may be performed systematically. For example, the method is performed in order to generate a systematically compartmentalized library of progeny molecules, with compartments that can be screened systematically, e.g. one by one. Through the selective and judicious use of specific nucleic acid building blocks, coupled with the selective and judicious use of sequentially stepped assembly reactions, a design can be achieved where specific sets of progeny products are made in each of several reaction vessels. This allows a systematic examination and screening procedure to be performed. Thus, these methods allow a potentially very large number of progeny molecules to be

examined systematically in smaller groups. Because of its ability to perform chimerizations in a manner that is highly flexible yet exhaustive and systematic as well, particularly when there is a low level of homology among the progenitor molecules, these methods provide for the generation of a library (or set) comprised of a large number of progeny molecules. Because of the non-stochastic nature, the progeny molecules generated comprise a library of finalized chimeric nucleic acid molecules having an overall assembly order that is chosen by design. The saturation mutagenesis and optimized directed evolution methods also can be used to generate different progeny molecular species. It is appreciated that the invention provides freedom of choice and control regarding the selection of demarcation points, the size and number of the nucleic acid building blocks, and the size and design of the couplings. It is appreciated, furthermore, that the requirement for intermolecular homology is highly relaxed for the operability of this invention. In fact, demarcation points can even be chosen in areas of little or no intermolecular homology. For example, because of codon wobble, i.e. the degeneracy of codons, nucleotide substitutions can be introduced into nucleic acid building blocks without altering the amino acid originally encoded in the corresponding progenitor template. Alternatively, a codon can be altered such that the coding for an originally amino acid is altered. Such substitutions can be introduced into the nucleic acid building block in order to increase the incidence of intermolecular homologous demarcation points and thus to allow an increased number of couplings to be achieved among the building blocks, which in turn allows a greater number of progeny chimeric molecules to be generated.

Synthetic gene reassembly

[0349] A non-stochastic method termed synthetic gene reassembly (e.g., GeneReassembly, see, e.g., U.S. Patent No. 6,537,776), differs from stochastic shuffling in that the nucleic acid building blocks are not shuffled or concatenated or chimerized randomly, but rather are assembled non-stochastically.

[0350] The synthetic gene reassembly method does not depend on the presence of a high level of homology between polynucleotides to be shuffled. This can be used to non-stochastically generate libraries (or sets) of progeny molecules comprised of over 10^{100} different chimeras. Conceivably, synthetic gene reassembly can even be used to generate libraries comprised of over 10^{1000} different progeny chimeras. A non-stochastic method may produce a set of finalized chimeric nucleic acid molecules having an overall assembly order that is chosen by design, which method is comprised of the steps of generating by design a plurality of specific nucleic acid building blocks having serviceable mutually compatible ligatable ends and assembling these nucleic acid building blocks, such that a designed overall assembly order is achieved.

[0351] Synthetic gene reassembly comprises a method of: 1) preparing a progeny generation of molecule(s) (including a molecule comprising a polynucleotide sequence, e.g., a molecule comprising a polypeptide coding sequence), that is mutagenized to achieve at least one point mutation, addition, deletion, &/or chimerization, from one or more ancestral or parental generation template(s); 2) screening the progeny generation molecule(s), e.g., using a high throughput method, for at least one property of interest (such as an improvement in an enzyme activity); 3) obtaining &/or cataloguing structural &/or and functional information regarding the parental &/or progeny generation molecules; and 4) repeating any of steps 1) to 3). There may be generated (e.g., from a parent polynucleotide template), in what is termed "codon site-saturation mutagenesis," a progeny generation of polynucleotides, each having at least one set of up to three contiguous point mutations (i.e. different bases comprising a new codon), such that every codon (or every family of degenerate codons encoding the same amino acid) is represented at each codon position. Corresponding to, and encoded by, this progeny generation of polynucleotides, there is also generated a set of progeny polypeptides, each having at least one single amino acid point mutation.

There is generated, in what is termed "amino acid site-saturation mutagenesis", one such mutant polypeptide for each of the 19 naturally encoded polypeptide-forming alpha-amino acid substitutions at each and every amino acid position along the polypeptide. This yields, for each and every amino acid position along the parental polypeptide, a total of 20 distinct progeny polypeptides including the original amino acid, or potentially more than 21 distinct progeny polypeptides if additional amino acids are used either instead of or in addition to the 20 naturally encoded amino acids.

[0352] This approach is also serviceable for generating mutants containing, in addition to &/or in combination with the 20 naturally encoded polypeptide-forming alpha-amino acids, other rare &/or not naturally-encoded amino acids and amino acid derivatives. This approach is also serviceable for generating mutants by the use of, in addition to &/or in combination with natural or unaltered codon recognition systems of suitable hosts, altered, mutagenized, &/or designer codon recognition systems (such as in a host cell with one or more altered tRNA molecules).

[0353] Polynucleotides encoding a polypeptide may be prepared by a method of *in vivo* reassortment of polynucleotide sequences containing regions of partial homology, assembling the polynucleotides to form at least one polynucleotide and screening the polynucleotides for the production of polypeptide(s) having a useful property.

[0354] This is serviceable for analyzing and cataloguing, with respect to any molecular property (e.g. an enzymatic activity) or combination of properties allowed by current technology, the effects of any mutational change achieved (including particularly saturation mutagenesis). Thus, a comprehensive method is provided for determining the effect of changing each amino acid in a parental polypeptide into each of at least 19 possible substitutions. This allows each amino acid in a parental polypeptide to be characterized and catalogued according to its spectrum of potential effects on a measurable property of the polypeptide.

[0355] An intron may be introduced into a chimeric progeny molecule by way of a nucleic acid building block. Introns often have consensus sequences at both termini in order to render them operational. In addition to enabling gene splicing, introns may serve an additional purpose by providing sites of homology to other nucleic acids to enable homologous recombination. For this purpose, and potentially others, it may be sometimes desirable to generate a large nucleic acid building block for introducing an intron. If the size is overly large easily generating by direct chemical synthesis of two single stranded oligos, such a specialized nucleic acid building block may also be generated by direct chemical synthesis of more than two single stranded oligos or by using a polymerase-based amplification reaction.

[0356] The mutually compatible ligatable ends of the nucleic acid building blocks to be assembled are considered to be "serviceable" for this type of ordered assembly if they enable the building blocks to be coupled in predetermined orders. Thus, the overall assembly order in which the nucleic acid building blocks can be coupled is specified by the design of the ligatable ends and, if more than one assembly step is to be used, then the overall assembly order in which the nucleic acid building blocks can be coupled is also specified by the sequential order of the assembly step(s). The annealed building pieces may be treated with an enzyme, such as a ligase (e.g., T4 DNA ligase) to achieve covalent bonding of the building pieces.

[0357] Coupling can occur in a manner that does not make use of every nucleotide in a participating overhang. The coupling is particularly lively to survive (e.g. in a transformed host) if the coupling reinforced by treatment with a ligase enzyme to form what may be referred to as a "gap ligation" or a "gapped ligation". This type of coupling can contribute to generation of unwanted background product(s), but it can

also be used advantageously increase the diversity of the progeny library generated by the designed ligation reassembly. Certain overhangs are able to undergo self-coupling to form a palindromic coupling. A coupling is strengthened substantially if it is reinforced by treatment with a ligase enzyme. Lack of 5' phosphates on these overhangs can be used advantageously to prevent this type of palindromic self-ligation. Accordingly, this provides that nucleic acid building blocks can be chemically made (or ordered) that lack a 5' phosphate group. Alternatively, they can be removed, e.g. by treatment with a phosphatase enzyme, such as a calf intestinal alkaline phosphatase (CIAP), in order to prevent palindromic self-ligations in ligation reassembly processes.

[0358] The design of nucleic acid building blocks may be obtained upon analysis of the sequences of a set of progenitor nucleic acid templates that serve as a basis for producing a progeny set of finalized chimeric nucleic acid molecules. These progenitor nucleic acid templates thus serve as a source of sequence information that aids in the design of the nucleic acid building blocks that are to be mutagenized, *i.e.* chimerized or shuffled.

[0359] Chimerization may be of a family of related genes may provide their encoded family of related products. The encoded products may be enzymes. The xylanases of the present invention can be mutagenized in accordance with the methods described herein.

[0360] Thus, the sequences of a plurality of progenitor nucleic acid templates (*e.g.*, polynucleotides of the invention) are aligned in order to select one or more demarcation points, which demarcation points can be located at an area of homology. The demarcation points can be used to delineate the boundaries of nucleic acid building blocks to be generated. Thus, the demarcation points identified and selected in the progenitor molecules serve as potential chimerization points in the assembly of the progeny molecules.

[0361] Typically a serviceable demarcation point is an area of homology (comprised of at least one homologous nucleotide base) shared by at least two progenitor templates, but the demarcation point can be an area of homology that is shared by at least half of the progenitor templates, at least two thirds of the progenitor templates, at least three fourths of the progenitor templates and preferably at almost all of the progenitor templates. Even more preferably still a serviceable demarcation point is an area of homology that is shared by all of the progenitor templates.

[0362] The gene reassembly process may be performed exhaustively in order to generate an exhaustive library. In other words, all possible ordered combinations of the nucleic acid building blocks are represented in the set of finalized chimeric nucleic acid molecules. At the same time, the assembly order (*i.e.* the order of assembly of each building block in the 5' to 3' sequence of each finalized chimeric nucleic acid) in each combination is by design (or non-stochastic). Because of the non-stochastic nature of the method, the possibility of unwanted side products is greatly reduced.

[0363] The gene reassembly process may be performed systematically, for example to generate a systematically compartmentalized library, with compartments that can be screened systematically, *e.g.*, one by one. Through the selective and judicious use of specific nucleic acid building blocks, coupled with the selective and judicious use of sequentially stepped assembly reactions, an experimental design can be achieved where specific sets of progeny products are made in each of several reaction vessels. This allows a systematic examination and screening procedure to be performed. Thus, it allows a potentially very large number of progeny molecules to be examined systematically in smaller groups.

[0364] Because of its ability to perform chimerizations in a manner that is highly flexible yet exhaustive and systematic as well, particularly when there is a low level of homology among the progenitor molecules, a library (or set) comprised of a large number of progeny molecules may be generated. Because of the non-

stochastic nature of the instant gene reassembly, the progeny molecules generated preferably comprise a library of finalized chimeric nucleic acid molecules having an overall assembly order that is chosen by design. In a particularly aspect, such a generated library is comprised of greater than 10^3 to greater than 10^{1000} different progeny molecular species.

[0365] A set of finalized chimeric nucleic acid molecules, produced as described may comprise a polynucleotide encoding a polypeptide. This polynucleotide may be a gene, which may be a man-made gene. Or, this polynucleotide may be a gene pathway, which may be a man-made gene pathway. One or more man-made genes so generated may be incorporated into a man-made gene pathway, such as pathway operable in a eukaryotic organism (including a plant).

[0366] The synthetic nature of the step in which the building blocks are generated allows the design and introduction of nucleotides (*e.g.*, one or more nucleotides, which may be, for example, codons or introns or regulatory sequences) that can later be in one aspect removed in an *in vitro* process (*e.g.*, by mutagenesis) or in an *in vivo* process (*e.g.*, by utilizing the gene splicing ability of a host organism). It is appreciated that in many instances the introduction of these nucleotides may also be desirable for many other reasons in addition to the potential benefit of creating a serviceable demarcation point.

[0367] Thus, a nucleic acid building block can be used to introduce an intron. Functional introns may be introduced into a man-made gene of the invention. Functional introns may be introduced into a man-made gene pathway of the invention. A chimeric polynucleotide may be generated that is a man-made gene containing one (or more) artificially introduced intron(s).

[0368] A chimeric polynucleotide may be a man-made gene pathway containing one (or more) artificially introduced intron(s). Preferably, the artificially introduced intron(s) are functional in one or more host cells for gene splicing much in the way that naturally-occurring introns serve functionally in gene splicing. Man-made intron-containing polynucleotides may be introduced into host organisms for recombination and/or splicing.

[0369] A man-made gene so produced can also serve as a substrate for recombination with another nucleic acid. Likewise, a man-made gene pathway can also serve as a substrate for recombination with another nucleic acid. The recombination may be facilitated by, or occurs at, areas of homology between the man-made, intron-containing gene and a nucleic acid, which serves as a recombination partner. The recombination partner may also be a nucleic acid generated by the methods described herein, including a man-made gene or a man-made gene pathway. Recombination may be facilitated by or may occur at areas of homology that exist at the one (or more) artificially introduced intron(s) in the man-made gene.

[0370] The synthetic gene reassembly method utilizes a plurality of nucleic acid building blocks, each of which preferably has two ligatable ends. The two ligatable ends on each nucleic acid building block may be two blunt ends (*i.e.* each having an overhang of zero nucleotides), or preferably one blunt end and one overhang, or more preferably still two overhangs.

[0371] A useful overhang for this purpose may be a 3' overhang or a 5' overhang. Thus, a nucleic acid building block may have a 3' overhang or alternatively a 5' overhang or alternatively two 3' overhangs or alternatively two 5' overhangs. The overall order in which the nucleic acid building blocks are assembled to form a finalized chimeric nucleic acid molecule is determined by purposeful experimental design and is not random.

[0372] In one aspect, a nucleic acid building block is generated by chemical synthesis of two single-

stranded nucleic acids (also referred to as single-stranded oligos) and contacting them so as to allow them to anneal to form a double-stranded nucleic acid building block.

[0373] A double-stranded nucleic acid building block can be of variable size. The sizes of these building blocks can be small or large. Exemplary sizes for building block range from 1 base pair (not including any overhangs) to 100,000 base pairs (not including any overhangs). Other exemplary size ranges are also provided, which have lower limits of from 1 bp to 10,000 bp (including every integer value in between) and upper limits of from 2 bp to 100,000 bp (including every integer value in between).

[0374] Many methods exist by which a double-stranded nucleic acid building block can be generated that is serviceable; and these are known in the art and can be readily performed by the skilled artisan.

[0375] A double-stranded nucleic acid building block may be generated by first generating two single stranded nucleic acids and allowing them to anneal to form a double-stranded nucleic acid building block. The two strands of a double-stranded nucleic acid building block may be complementary at every nucleotide apart from any that form an overhang; thus containing no mismatches, apart from any overhang(s). Or, the two strands of a double-stranded nucleic acid building block are complementary at fewer than every nucleotide apart from any that form an overhang. Thus, a double-stranded nucleic acid building block can be used to introduce codon degeneracy. The codon degeneracy can be introduced using the site-saturation mutagenesis described herein, using one or more N,N,G/T cassettes or alternatively using one or more N,N,N cassettes.

[0376] The *in vivo* recombination method can be performed blindly on a pool of unknown hybrids or alleles of a specific polynucleotide or sequence. However, it is not necessary to know the actual DNA or RNA sequence of the specific polynucleotide.

[0377] The approach of using recombination within a mixed population of genes can be useful for the generation of any useful proteins, for example, interleukin I, antibodies, tPA and growth hormone. This approach may be used to generate proteins having altered specificity or activity. The approach may also be useful for the generation of hybrid nucleic acid sequences, for example, promoter regions, introns, exons, enhancer sequences, 3' untranslated regions or 5' untranslated regions of genes. Thus this approach may be used to generate genes having increased rates of expression. This approach may also be useful in the study of repetitive DNA sequences. Finally, this approach may be useful to mutate ribozymes or aptamers.

[0378] In one aspect the processes described herein are directed to the use of repeated cycles of reductive reassortment, recombination and selection which allow for the directed molecular evolution of highly complex linear sequences, such as DNA, RNA or proteins through recombination.

Optimized Directed Evolution System

[0379] A non-stochastic gene modification system termed "optimized directed evolution system" may generate polypeptides, e.g., xylanases or antibodies of the invention, with new or altered properties. Optimized directed evolution is directed to the use of repeated cycles of reductive reassortment, recombination and selection that allow for the directed molecular evolution of nucleic acids through recombination. Optimized directed evolution allows generation of a large population of evolved chimeric sequences, wherein the generated population is significantly enriched for sequences that have a predetermined number of crossover events.

[0380] A crossover event is a point in a chimeric sequence where a shift in sequence occurs from one parental variant to another parental variant. Such a point is normally at the juncture of where oligonucleotides from two parents are ligated together to form a single sequence. This method allows calculation of the correct concentrations of oligonucleotide sequences so that the final chimeric population of sequences is enriched for the chosen number of crossover events. This provides more control over choosing chimeric variants having a predetermined number of crossover events.

[0381] In addition, this method provides a convenient means for exploring a tremendous amount of the possible protein variant space in comparison to other systems. Previously, if one generated, for example, 10^{13} chimeric molecules during a reaction, it would be extremely difficult to test such a high number of chimeric variants for a particular activity. Moreover, a significant portion of the progeny population would have a very high number of crossover events which resulted in proteins that were less likely to have increased levels of a particular activity. By using these methods, the population of chimeric molecules can be enriched for those variants that have a particular number of crossover events. Thus, although one can still generate 10^{13} chimeric molecules during a reaction, each of the molecules chosen for further analysis most likely has, for example, only three crossover events. Because the resulting progeny population can be skewed to have a predetermined number of crossover events, the boundaries on the functional variety between the chimeric molecules is reduced. This provides a more manageable number of variables when calculating which oligonucleotide from the original parental polynucleotides might be responsible for affecting a particular trait.

[0382] One method for creating a chimeric progeny polynucleotide sequence is to create oligonucleotides corresponding to fragments or portions of each parental sequence. Each oligonucleotide preferably includes a unique region of overlap so that mixing the oligonucleotides together results in a new variant that has each oligonucleotide fragment assembled in the correct order. Additional information can also be found, e.g., in USSN 09/332,835; U.S. Patent No. 6,361,974.

[0383] The number of oligonucleotides generated for each parental variant bears a relationship to the total number of resulting crossovers in the chimeric molecule that is ultimately created. For example, three parental nucleotide sequence variants might be provided to undergo a ligation reaction in order to find a chimeric variant having, for example, greater activity at high temperature. As one example, a set of 50 oligonucleotide sequences can be generated corresponding to each portions of each parental variant. Accordingly, during the ligation reassembly process there could be up to 50 crossover events within each of the chimeric sequences. The probability that each of the generated chimeric polynucleotides will contain oligonucleotides from each parental variant in alternating order is very low. If each oligonucleotide fragment is present in the ligation reaction in the same molar quantity it is likely that in some positions oligonucleotides from the same parental polynucleotide will ligate next to one another and thus not result in a crossover event. If the concentration of each oligonucleotide from each parent is kept constant during any ligation step in this example, there is a 1/3 chance (assuming 3 parents) that an oligonucleotide from the same parental variant will ligate within the chimeric sequence and produce no crossover.

[0384] Accordingly, a probability density function (PDF) can be determined to predict the population of crossover events that are likely to occur during each step in a ligation reaction given a set number of parental variants, a number of oligonucleotides corresponding to each variant, and the concentrations of each variant during each step in the ligation reaction. The statistics and mathematics behind determining the PDF is described below. By utilizing these methods, one can calculate such a probability density function, and thus enrich the chimeric progeny population for a predetermined number of crossover events resulting from a particular ligation reaction. Moreover, a target number of crossover events can be

predetermined, and the system then programmed to calculate the starting quantities of each parental oligonucleotide during each step in the ligation reaction to result in a probability density function that centers on the predetermined number of crossover events. These methods are directed to the use of repeated cycles of reductive reassortment, recombination and selection that allow for the directed molecular evolution of a nucleic acid encoding a polypeptide through recombination. This system allows generation of a large population of evolved chimeric sequences, wherein the generated population is significantly enriched for sequences that have a predetermined number of crossover events. A crossover event is a point in a chimeric sequence where a shift in sequence occurs from one parental variant to another parental variant. Such a point is normally at the juncture of where oligonucleotides from two parents are ligated together to form a single sequence. The method allows calculation of the correct concentrations of oligonucleotide sequences so that the final chimeric population of sequences is enriched for the chosen number of crossover events. This provides more control over choosing chimeric variants having a predetermined number of crossover events.

[0385] In addition, these methods provide a convenient means for exploring a tremendous amount of the possible protein variant space in comparison to other systems. By using the methods described herein, the population of chimeric molecules can be enriched for those variants that have a particular number of crossover events. Thus, although one can still generate 10^{13} chimeric molecules during a reaction, each of the molecules chosen for further analysis most likely has, for example, only three crossover events. Because the resulting progeny population can be skewed to have a predetermined number of crossover events, the boundaries on the functional variety between the chimeric molecules is reduced. This provides a more manageable number of variables when calculating which oligonucleotide from the original parental polynucleotides might be responsible for affecting a particular trait.

[0386] The method may create a chimeric progeny polynucleotide sequence by creating oligonucleotides corresponding to fragments or portions of each parental sequence. Each oligonucleotide preferably includes a unique region of overlap so that mixing the oligonucleotides together results in a new variant that has each oligonucleotide fragment assembled in the correct order. See also USSN 09/332,835.

Determining Crossover Events

[0387] A system and software may receive a desired crossover probability density function (PDF), the number of parent genes to be reassembled, and the number of fragments in the reassembly as inputs. The output of this program is a "fragment PDF" that can be used to determine a recipe for producing reassembled genes, and the estimated crossover PDF of those genes. The processing described herein is preferably performed in MATLAB™ (The Mathworks, Natick, Massachusetts) a programming language and development environment for technical computing.

Iterative Processes

[0388] The processes described herein can be iteratively repeated. For example, a nucleic acid (or, the nucleic acid) responsible for an altered or new xylanase phenotype is identified, re-isolated, again modified, re-tested for activity. This process can be iteratively repeated until a desired phenotype is engineered. For example, an entire biochemical anabolic or catabolic pathway can be engineered into a cell, including xylanase activity.

[0389] Similarly, if it is determined that a particular oligonucleotide has no affect at all on the desired trait

(e.g., a new xylanase phenotype), it can be removed as a variable by synthesizing larger parental oligonucleotides that include the sequence to be removed. Since incorporating the sequence within a larger sequence prevents any crossover events, there will no longer be any variation of this sequence in the progeny polynucleotides. This iterative practice of determining which oligonucleotides are most related to the desired trait, and which are unrelated, allows more efficient exploration all of the possible protein variants that might be provide a particular trait or activity.

In vivo shuffling

[0390] *In vivo* shuffling of molecules is used in the methods of the disclosure that provide variants of polypeptides of the invention, e.g., antibodies, xylanases, and the like. *In vivo* shuffling can be performed utilizing the natural property of cells to recombine multimers. While recombination *in vivo* has provided the major natural route to molecular diversity, genetic recombination remains a relatively complex process that involves 1) the recognition of homologies; 2) strand cleavage, strand invasion, and metabolic steps leading to the production of recombinant chiasma; and finally 3) the resolution of chiasma into discrete recombined molecules. The formation of the chiasma requires the recognition of homologous sequences.

[0391] A hybrid polynucleotide may be produced from at least a first polynucleotide and a second polynucleotide, by introducing at least a first polynucleotide and a second polynucleotide which share at least one region of partial sequence homology into a suitable host cell. The regions of partial sequence homology promote processes which result in sequence reorganization producing a hybrid polynucleotide. The term "hybrid polynucleotide", as used herein, is any nucleotide sequence which results from the method of the present invention and contains sequence from at least two original polynucleotide sequences. Such hybrid polynucleotides can result from intermolecular recombination events which promote sequence integration between DNA molecules. In addition, such hybrid polynucleotides can result from intramolecular reductive reassortment processes which utilize repeated sequences to alter a nucleotide sequence within a DNA molecule.

[0392] *In vivo* reassortment is focused on "inter-molecular" processes collectively referred to as "recombination" which in bacteria, is generally viewed as a "RecA-dependent" phenomenon. The process can rely on recombination processes of a host cell to recombine and re-assort sequences, or the cells' ability to mediate reductive processes to decrease the complexity of quasi-repeated sequences in the cell by deletion. This process of "reductive reassortment" occurs by an "intra-molecular", RecA-independent process.

[0393] Therefore, novel polynucleotides can be generated by the process of reductive reassortment. The method involves the generation of constructs containing consecutive sequences (original encoding sequences), their insertion into an appropriate vector and their subsequent introduction into an appropriate host cell. The reassortment of the individual molecular identities occurs by combinatorial processes between the consecutive sequences in the construct possessing regions of homology, or between quasi-repeated units. The reassortment process recombines and/or reduces the complexity and extent of the repeated sequences and results in the production of novel molecular species. Various treatments may be applied to enhance the rate of reassortment. These could include treatment with ultraviolet light, or DNA damaging chemicals and/or the use of host cell lines displaying enhanced levels of "genetic instability". Thus the reassortment process may involve homologous recombination or the natural property of quasi-repeated sequences to direct their own evolution.

[0394] Repeated or "quasi-repeated" sequences play a role in genetic instability. "Quasi-repeats" are

repeats that are not restricted to their original unit structure. Quasi-repeated units can be presented as an array of sequences in a construct; consecutive units of similar sequences. Once ligated, the junctions between the consecutive sequences become essentially invisible and the quasi-repetitive nature of the resulting construct is now continuous at the molecular level. The deletion process the cell performs to reduce the complexity of the resulting construct operates between the quasi-repeated sequences. The quasi-repeated units provide a practically limitless repertoire of templates upon which slippage events can occur. The constructs containing the quasi-repeats thus effectively provide sufficient molecular elasticity that deletion (and potentially insertion) events can occur virtually anywhere within the quasi-repetitive units.

[0395] When the quasi-repeated sequences are all ligated in the same orientation, for instance head to tail or vice versa, the cell cannot distinguish individual units. Consequently, the reductive process can occur throughout the sequences. In contrast, when for example, the units are presented head to head, rather than head to tail, the inversion delineates the endpoints of the adjacent unit so that deletion formation will favor the loss of discrete units. Thus, it is preferable with the present method that the sequences are in the same orientation. Random orientation of quasi-repeated sequences will result in the loss of reassortment efficiency, while consistent orientation of the sequences will offer the highest efficiency. However, while having fewer of the contiguous sequences in the same orientation decreases the efficiency, it may still provide sufficient elasticity for the effective recovery of novel molecules. Constructs can be made with the quasi-repeated sequences in the same orientation to allow higher efficiency.

[0396] Sequences can be assembled in a head to tail orientation using any of a variety of methods, including the following:

1. a) Primers that include a poly-A head and poly-T tail which when made single-stranded would provide orientation can be utilized. This is accomplished by having the first few bases of the primers made from RNA and hence easily removed RNaseH.
2. b) Primers that include unique restriction cleavage sites can be utilized. Multiple sites, a battery of unique sequences and repeated synthesis and ligation steps would be required.
3. c) The inner few bases of the primer could be thiolated and an exonuclease used to produce properly tailed molecules.

[0397] The recovery of the re-assorted sequences relies on the identification of cloning vectors with a reduced repetitive index (RI). The re-assorted encoding sequences can then be recovered by amplification. The products are re-cloned and expressed. The recovery of cloning vectors with reduced RI can be affected by:

1. 1) The use of vectors only stably maintained when the construct is reduced in complexity.
2. 2) The physical recovery of shortened vectors by physical procedures. In this case, the cloning vector would be recovered using standard plasmid isolation procedures and size fractionated on either an agarose gel, or column with a low molecular weight cut off utilizing standard procedures.
3. 3) The recovery of vectors containing interrupted genes which can be selected when insert size decreases.
4. 4) The use of direct selection techniques with an expression vector and the appropriate selection.

[0398] Encoding sequences (for example, genes) from related organisms may demonstrate a high degree of homology and encode quite diverse protein products. These types of sequences are particularly useful as quasi-repeats. However, while the examples illustrated below demonstrate the reassortment of nearly

identical original encoding sequences (quasi-repeats), this process is not limited to such nearly identical repeats.

[0399] The following example demonstrates such a method. Encoding nucleic acid sequences (quasi-repeats) derived from three (3) unique species are described. Each sequence encodes a protein with a distinct set of properties. Each of the sequences differs by a single or a few base pairs at a unique position in the sequence. The quasi-repeated sequences are separately or collectively amplified and ligated into random assemblies such that all possible permutations and combinations are available in the population of ligated molecules. The number of quasi-repeat units can be controlled by the assembly conditions. The average number of quasi-repeated units in a construct is defined as the repetitive index (RI).

[0400] Once formed, the constructs may, or may not be size fractionated on an agarose gel according to published protocols, inserted into a cloning vector and transfected into an appropriate host cell. The cells are then propagated and "reductive reassortment" is effected. The rate of the reductive reassortment process may be stimulated by the introduction of DNA damage if desired. Whether the reduction in RI is mediated by deletion formation between repeated sequences by an "intra-molecular" mechanism, or mediated by recombination-like events through "inter-molecular" mechanisms is immaterial. The end result is a reassortment of the molecules into all possible combinations.

[0401] The method may comprise the additional step of screening the library members of the shuffled pool to identify individual shuffled library members having the ability to bind or otherwise interact, or catalyze a particular reaction (e.g., such as catalytic domain of an enzyme) with a predetermined macromolecule, such as for example a proteinaceous receptor, an oligosaccharide, virion, or other predetermined compound or structure.

[0402] The polypeptides that are identified from such libraries can be used for therapeutic, diagnostic, research and related purposes (e.g., catalysts, solutes for increasing osmolarity of an aqueous solution and the like) and/or can be subjected to one or more additional cycles of shuffling and/or selection.

[0403] Prior to or during recombination or reassortment, polynucleotides generated by the method described herein can be subjected to agents or processes which promote the introduction of mutations into the original polynucleotides. The introduction of such mutations would increase the diversity of resulting hybrid polynucleotides and polypeptides encoded therefrom. The agents or processes which promote mutagenesis can include, but are not limited to: (+)-CC-1065, or a synthetic analog such as (+)-CC-1065-(N3-Adenine) (See Sun and Hurley, (1992); an N-acetylated or deacetylated 4'-fluoro-4-aminobiphenyl adduct capable of inhibiting DNA synthesis (See, for example, van de Poll *et al.* (1992)); or a N-acetylated or deacetylated 4-aminobiphenyl adduct capable of inhibiting DNA synthesis (See also, van de Poll *et al.* (1992), pp. 751-758); trivalent chromium, a trivalent chromium salt, a polycyclic aromatic hydrocarbon (PAH) DNA adduct capable of inhibiting DNA replication, such as 7-bromomethyl-benz[*a*]anthracene ("BMA"), tris(2,3-dibromopropyl)phosphate ("Tris-BP"), 1,2-dibromo-3-chloropropane ("DBCP"), 2-bromoacrolein (2BA), benzo[*a*]pyrene-7,8-dihydrodiol-9-10-epoxide ("BPDE"), a platinum(II) halogen salt, N-hydroxy-2-amino-3-methylimidazo[4,5-*f*]quinoline ("N-hydroxy-IQ") and N-hydroxy-2-amino-1-methyl-6-phenylimidazo[4,5-*f*]pyridine ("N-hydroxy-PhIP"). Exemplary means for slowing or halting PCR amplification consist of UV light (+)-CC-1065 and (+)-CC-1065-(N3-Adenine). Particularly encompassed means are DNA adducts or polynucleotides comprising the DNA adducts from the polynucleotides or polynucleotides pool, which can be released or removed by a process including heating the solution comprising the polynucleotides prior to further processing.

[0404] Recombinant proteins having biological activity may be produced by treating a sample comprising double-stranded template polynucleotides encoding a wild-type protein under conditions which provide for

the production of hybrid or re-assorted polynucleotides.

Producing sequence variants

[0405] Additional methods may make sequence variants of the nucleic acid sequences of the invention. Additional methods may isolate xylanases using the nucleic acids and polypeptides of the invention. Variants of a xylanase coding sequence (e.g., a gene, cDNA or message) of the invention, which can be altered by any means, including, e.g., random or stochastic methods, or, non-stochastic, or "directed evolution," methods, as described above.

[0406] The isolated variants may be naturally occurring. Variant can also be created *in vitro*. Variants may be created using genetic engineering techniques such as site directed mutagenesis, random chemical mutagenesis, Exonuclease III deletion procedures, and standard cloning techniques. Alternatively, such variants, fragments, analogs, or derivatives may be created using chemical synthesis or modification procedures. Other methods of making variants are also familiar to those skilled in the art. These include procedures in which nucleic acid sequences obtained from natural isolates are modified to generate new nucleic acids which encode polypeptides having characteristics which enhance their value in industrial, medical, laboratory (research), pharmaceutical, food and feed and food and feed supplement processing and other applications and processes. In such procedures, a large number of variant sequences having one or more nucleotide differences with respect to the sequence obtained from the natural isolate are generated and characterized. These nucleotide differences can result in amino acid changes with respect to the polypeptides encoded by the nucleic acids from the natural isolates.

[0407] For example, variants may be created using error prone PCR. In error prone PCR, PCR is performed under conditions where the copying fidelity of the DNA polymerase is low, such that a high rate of point mutations is obtained along the entire length of the PCR product. Error prone PCR is described, e.g., in Leung, D.W., et al., Technique, 1:11-15, 1989) and Caldwell, R. C. & Joyce G.F., PCR Methods Applic., 2:28-33, 1992. Briefly, in such procedures, nucleic acids to be mutagenized are mixed with PCR primers, reaction buffer, MgCl₂, MnCl₂, Taq polymerase and an appropriate concentration of dNTPs for achieving a high rate of point mutation along the entire length of the PCR product. For example, the reaction may be performed using 20 fmoles of nucleic acid to be mutagenized, 30 pmole of each PCR primer, a reaction buffer comprising 50mM KCl, 10mM Tris HCl (pH 8.3) and 0.01% gelatin, 7mM MgCl₂, 0.5mM MnCl₂, 5 units of Taq polymerase, 0.2mM dGTP, 0.2mM dATP, 1mM dCTP, and 1mM dTTP. PCR may be performed for 30 cycles of 94°C for 1 min, 45°C for 1 min, and 72°C for 1 min. However, it will be appreciated that these parameters may be varied as appropriate. The mutagenized nucleic acids are cloned into an appropriate vector and the activities of the polypeptides encoded by the mutagenized nucleic acids are evaluated.

[0408] Variants may also be created using oligonucleotide directed mutagenesis to generate site-specific mutations in any cloned DNA of interest. Oligonucleotide mutagenesis is described, e.g., in Reidhaar-Olson (1988) Science 241:53-57. Briefly, in such procedures a plurality of double stranded oligonucleotides bearing one or more mutations to be introduced into the cloned DNA are synthesized and inserted into the cloned DNA to be mutagenized. Clones containing the mutagenized DNA are recovered and the activities of the polypeptides they encode are assessed.

[0409] Another method for generating variants is assembly PCR. Assembly PCR involves the assembly of a PCR product from a mixture of small DNA fragments. A large number of different PCR reactions occur in parallel in the same vial, with the products of one reaction priming the products of another reaction.

Assembly PCR is described in, e.g., U.S. Patent No. 5,965,408.

[0410] Still another method of generating variants is sexual PCR mutagenesis. In sexual PCR mutagenesis, forced homologous recombination occurs between DNA molecules of different but highly related DNA sequence *in vitro*, as a result of random fragmentation of the DNA molecule based on sequence homology, followed by fixation of the crossover by primer extension in a PCR reaction. Sexual PCR mutagenesis is described, e.g., in Stemmer (1994) Proc. Natl. Acad. Sci. USA 91:10747-10751. Briefly, in such procedures a plurality of nucleic acids to be recombined are digested with DNase to generate fragments having an average size of 50-200 nucleotides. Fragments of the desired average size are purified and resuspended in a PCR mixture. PCR is conducted under conditions which facilitate recombination between the nucleic acid fragments. For example, PCR may be performed by resuspending the purified fragments at a concentration of 10-30ng/μl in a solution of 0.2mM of each dNTP, 2.2mM MgCl₂, 50mM KCL, 10mM Tris HCl, pH 9.0, and 0.1% Triton X-100. 2.5 units of Taq polymerase per 100:1 of reaction mixture is added and PCR is performed using the following regime: 94°C for 60 seconds, 94°C for 30 seconds, 50-55°C for 30 seconds, 72°C for 30 seconds (30-45 times) and 72°C for 5 minutes. However, it will be appreciated that these parameters may be varied as appropriate. In some aspects, oligonucleotides may be included in the PCR reactions. In other aspects, the Klenow fragment of DNA polymerase I may be used in a first set of PCR reactions and Taq polymerase may be used in a subsequent set of PCR reactions. Recombinant sequences are isolated and the activities of the polypeptides they encode are assessed.

[0411] Variants may also be created by *in vivo* mutagenesis. In some aspects, random mutations in a sequence of interest are generated by propagating the sequence of interest in a bacterial strain, such as an E. coli strain, which carries mutations in one or more of the DNA repair pathways. Such "mutator" strains have a higher random mutation rate than that of a wild-type parent. Propagating the DNA in one of these strains will eventually generate random mutations within the DNA. Mutator strains suitable for use for *in vivo* mutagenesis are described in PCT Publication No. WO 91/16427, published October 31, 1991, entitled "Methods for Phenotype Creation from Multiple Gene Populations".

[0412] Variants may also be generated using cassette mutagenesis. In cassette mutagenesis a small region of a double stranded DNA molecule is replaced with a synthetic oligonucleotide "cassette" that differs from the native sequence. The oligonucleotide often contains completely and/or partially randomized native sequence.

[0413] Recursive ensemble mutagenesis may also be used to generate variants. Recursive ensemble mutagenesis is an algorithm for protein engineering (protein mutagenesis) developed to produce diverse populations of phenotypically related mutants whose members differ in amino acid sequence. This method uses a feedback mechanism to control successive rounds of combinatorial cassette mutagenesis. Recursive ensemble mutagenesis is described in Arkin, A.P. and Youvan, D.C., PNAS, USA, 89:7811-7815, 1992.

[0414] Variants may be created using exponential ensemble mutagenesis. Exponential ensemble mutagenesis is a process for generating combinatorial libraries with a high percentage of unique and functional mutants, wherein small groups of residues are randomized in parallel to identify, at each altered position, amino acids which lead to functional proteins. Exponential ensemble mutagenesis is described in Delegrave, S. and Youvan, D.C., Biotechnology Research, 11:1548-1552, 1993. Random and site-directed mutagenesis are described in Arnold, F.H., Current Opinion in Biotechnology, 4:450-455, 1993.

[0415] In some aspects, the variants are created using shuffling procedures wherein portions of a plurality

of nucleic acids which encode distinct polypeptides are fused together to create chimeric nucleic acid sequences which encode chimeric polypeptides as described in U.S. Patent No. 5,965,408, filed July 9, 1996, entitled, "Method of DNA Reassembly by Interrupting Synthesis" and U.S. Patent No. 5,939,250, filed May 22, 1996, entitled, "Production of Enzymes Having Desired Activities by Mutagenesis.

[0416] The variants of the polypeptides of the invention may be variants in which one or more of the amino acid residues of the polypeptides of the invention are substituted with a conserved or non-conserved amino acid residue (preferably a conserved amino acid residue) and such substituted amino acid residue may or may not be one encoded by the genetic code.

[0417] Conservative substitutions are those that substitute a given amino acid in a polypeptide by another amino acid of like characteristics. Typically seen as conservative substitutions are the following replacements: replacements of an aliphatic amino acid such as Alanine, Valine, Leucine and Isoleucine with another aliphatic amino acid; replacement of a Serine with a Threonine or vice versa; replacement of an acidic residue such as Aspartic acid and Glutamic acid with another acidic residue; replacement of a residue bearing an amide group, such as Asparagine and Glutamine, with another residue bearing an amide group; exchange of a basic residue such as Lysine and Arginine with another basic residue; and replacement of an aromatic residue such as Phenylalanine, Tyrosine with another aromatic residue.

[0418] Other variants are those in which one or more of the amino acid residues of the polypeptides of the invention includes a substituent group.

[0419] Still other variants are those in which the polypeptide is associated with another compound, such as a compound to increase the half-life of the polypeptide (for example, polyethylene glycol).

[0420] Additional variants are those in which additional amino acids are fused to the polypeptide, such as a leader sequence, a secretory sequence, a proprotein sequence or a sequence which facilitates purification, enrichment, or stabilization of the polypeptide.

[0421] The fragments, derivatives and analogs may retain the same biological function or activity as the polypeptides of the invention and sequences substantially identical thereto, or the fragment, derivative, or analog may include a proprotein, such that the fragment, derivative, or analog can be activated by cleavage of the proprotein portion to produce an active polypeptide.

Optimizing codons to achieve high levels of protein expression in host cells

[0422] Methods for modifying xylanase-encoding nucleic acids may modify codon usage. Methods for modifying codons in a nucleic acid encoding a xylanase may increase or decrease its expression in a host cell. The method comprises identifying a "non-preferred" or a "less preferred" codon in xylanase-encoding nucleic acid and replacing one or more of these non-preferred or less preferred codons with a "preferred codon" encoding the same amino acid as the replaced codon and at least one non-preferred or less preferred codon in the nucleic acid has been replaced by a preferred codon encoding the same amino acid. A preferred codon is a codon over-represented in coding sequences in genes in the host cell and a non-preferred or less preferred codon is a codon under-represented in coding sequences in genes in the host cell.

[0423] Host cells for expressing the nucleic acids, expression cassettes and vectors of the invention include bacteria, yeast, fungi, plant cells, insect cells and mammalian cells. Exemplary host cells include

gram negative bacteria, such as *Escherichia coli* and *Pseudomonas fluorescens*; gram positive bacteria, such as *Lactobacillus gasseri*, *Lactococcus lactis*, *Lactococcus cremoris*, *Bacillus subtilis*. Exemplary host cells also include eukaryotic organisms, e.g., various yeast, such as *Saccharomyces* sp., including *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Pichia pastoris*, and *Kluyveromyces lactis*, *Hansenula polymorpha*, *Aspergillus niger*, and mammalian cells and cell lines and insect cells and cell lines. Other exemplary host cells include bacterial cells, such as *E. coli*, *Streptomyces*, *Bacillus subtilis*, *Bacillus cereus*, *Salmonella typhimurium* and various species within the genera *Pseudomonas*, *Streptomyces* and *Staphylococcus*, fungal cells, such as *Aspergillus*, yeast such as any species of *Pichia*, *Saccharomyces*, *Schizosaccharomyces*, *Schwanniomyces*, including *Pichia pastoris*, *Saccharomyces cerevisiae*, or *Schizosaccharomyces pombe*, insect cells such as *Drosophila* S2 and *Spodoptera Sf9*, animal cells such as CHO, COS or Bowes melanoma and adenoviruses. The selection of an appropriate host is within the abilities of those skilled in the art. Thus, the disclosure also includes nucleic acids and polypeptides optimized for expression in these organisms and species.

[0424] For example, the codons of a nucleic acid encoding a xylanase isolated from a bacterial cell are modified such that the nucleic acid is optimally expressed in a bacterial cell different from the bacteria from which the xylanase was derived, a yeast, a fungi, a plant cell, an insect cell or a mammalian cell. Methods for optimizing codons are well known in the art, see, e.g., U.S. Patent No. 5,795,737; Baca (2000) Int. J. Parasitol. 30:113-118; Hale (1998) Protein Expr. Purif. 12:185-188; Narum (2001) Infect. Immun. 69:7250-7253. See also Narum (2001) Infect. Immun. 69:7250-7253, describing optimizing codons in mouse systems; Outchkourov (2002) Protein Expr. Purif. 24:18-24, describing optimizing codons in yeast; Feng (2000) Biochemistry 39:15399-15409, describing optimizing codons in *E. coli*; Humphreys (2000) Protein Expr. Purif. 20:252-264, describing optimizing codon usage that affects secretion in *E. coli*.

Transgenic non-human animals

[0425] The invention provides transgenic non-human animals comprising a nucleic acid, a polypeptide (e.g., a xylanase), an expression cassette or vector or a transfected or transformed cell of the invention.

[0426] The transgenic non-human animals can be, e.g., goats, rabbits, sheep, pigs, cows, rats, horses, dogs, fish and mice, comprising the nucleic acids of the invention. These animals can be used, e.g., as *in vivo* models to study xylanase activity, or, as models to screen for agents that change the xylanase activity *in vivo*. The coding sequences for the polypeptides to be expressed in the transgenic non-human animals can be designed to be constitutive, or, under the control of tissue-specific, developmental-specific or inducible transcriptional regulatory factors. Transgenic non-human animals can be designed and generated using any method known in the art; see, e.g., U.S. Patent Nos. 6,211,428; 6,187,992; 6,156,952; 6,118,044; 6,111,166; 6,107,541; 5,959,171; 5,922,854; 5,892,070; 5,880,327; 5,891,698; 5,639,940; 5,573,933; 5,387,742; 5,087,571, describing making and using transformed cells and eggs and transgenic mice, rats, rabbits, sheep, pigs, chickens, goats, fish and cows. See also, e.g., Pollock (1999) J. Immunol. Methods 231:147-157, describing the production of recombinant proteins in the milk of transgenic dairy animals; Baguisi (1999) Nat. Biotechnol. 17:456-461, demonstrating the production of transgenic goats. U.S. Patent No. 6,211,428, describes making and using transgenic non-human mammals which express in their brains a nucleic acid construct comprising a DNA sequence. U.S. Patent No. 5,387,742, describes injecting cloned recombinant or synthetic DNA sequences into fertilized mouse eggs, implanting the injected eggs in pseudo-pregnant females, and growing to term transgenic mice whose cells express proteins related to the pathology of Alzheimer's disease. U.S. Patent No. 6,187,992, describes making and using a transgenic mouse whose genome comprises a disruption of the gene encoding amyloid precursor protein (APP).

[0427] "Knockout animals" e.g., a "knockout mouse," may be engineered not to express an endogenous gene, which is replaced with a gene expressing a xylanase of the invention, or, a fusion protein comprising a xylanase of the invention.

Transgenic Plants and Seeds

[0428] The invention provides transgenic plants and seeds comprising a nucleic acid, a polypeptide (a xylanase), an expression cassette or vector or a transfected or transformed cell of the invention. The invention also provides plant products or byproducts, e.g., fruits, oils, seeds, leaves, extracts and the like, including any plant part, comprising a nucleic acid and/or a polypeptide (a xylanase) of the invention, e.g., wherein the nucleic acid or polypeptide of the invention is heterologous to the plant, plant part, seed etc. The transgenic plant (which includes plant parts, fruits, seeds etc.) can be dicotyledonous (a dicot) or monocotyledonous (a monocot). The transgenic plant or plant cell expressing a polypeptide of the present invention may be constructed in accordance with any method known in the art. See, for example, U.S. Patent No. 6,309,872.

[0429] Nucleic acids and expression constructs of the invention can be introduced into a plant cell by any means. For example, nucleic acids or expression constructs can be introduced into the genome of a desired plant host, or, the nucleic acids or expression constructs can be episomes. Introduction into the genome of a desired plant can be such that the host's xylanase production is regulated by endogenous transcriptional or translational control elements. The invention also provides "knockout plants" where insertion of gene sequence by, e.g., homologous recombination, has disrupted the expression of the endogenous gene. Means to generate "knockout" plants are well-known in the art, see, e.g., Strepp (1998) Proc Natl. Acad. Sci. USA 95:4368-4373; Miao (1995) Plant J 7:359-365. See discussion on transgenic plants, below.

[0430] The nucleic acids of the invention can be used to confer desired traits on essentially any plant, e.g., on starch-producing plants, such as potato, wheat, rice, barley, and the like. Nucleic acids of the invention can be used to manipulate metabolic pathways of a plant in order to optimize or alter host's expression of xylanase. They can change xylanase activity in a plant. Alternatively, a xylanase of the invention can be used in production of a transgenic plant to produce a compound not naturally produced by that plant. This can lower production costs or create a novel product.

[0431] The first step in production of a transgenic plant involves making an expression construct for expression in a plant cell. These techniques are well known in the art. They can include selecting and cloning a promoter, a coding sequence for facilitating efficient binding of ribosomes to mRNA and selecting the appropriate gene terminator sequences. One exemplary constitutive promoter is CaMV35S, from the cauliflower mosaic virus, which generally results in a high degree of expression in plants. Other promoters are more specific and respond to cues in the plant's internal or external environment. An exemplary light-inducible promoter is the promoter from the cab gene, encoding the major chlorophyll a/b binding protein.

[0432] The nucleic acid may be modified to achieve greater expression in a plant cell. For example, a sequence of the invention is likely to have a higher percentage of A-T nucleotide pairs compared to that seen in a plant, some of which prefer G-C nucleotide pairs. Therefore, A-T nucleotides in the coding sequence can be substituted with G-C nucleotides without significantly changing the amino acid sequence to enhance production of the gene product in plant cells.

[0433] Selectable marker gene can be added to the gene construct in order to identify plant cells or

tissues that have successfully integrated the transgene. This may be necessary because achieving incorporation and expression of genes in plant cells is a rare event, occurring in just a few percent of the targeted tissues or cells. Selectable marker genes encode proteins that provide resistance to agents that are normally toxic to plants, such as antibiotics or herbicides. Only plant cells that have integrated the selectable marker gene will survive when grown on a medium containing the appropriate antibiotic or herbicide. As for other inserted genes, marker genes also require promoter and termination sequences for proper function.

[0434] Making transgenic plants or seeds comprises incorporating sequences of the invention and, in one aspect (optionally), marker genes into a target expression construct (e.g., a plasmid), along with positioning of the promoter and the terminator sequences. This can involve transferring the modified gene into the plant through a suitable method. For example, a construct may be introduced directly into the genomic DNA of the plant cell using techniques such as electroporation and microinjection of plant cell protoplasts, or the constructs can be introduced directly to plant tissue using ballistic methods, such as DNA particle bombardment. For example, see, e.g., Christou (1997) *Plant Mol. Biol.* 35:197-203; Pawlowski (1996) *Mol. Biotechnol.* 6:17-30; Klein (1987) *Nature* 327:70-73; Takumi (1997) *Genes Genet. Syst.* 72:63-69, discussing use of particle bombardment to introduce transgenes into wheat; and Adam (1997) *supra*, for use of particle bombardment to introduce YACs into plant cells. For example, Rinehart (1997) *supra*, used particle bombardment to generate transgenic cotton plants. Apparatus for accelerating particles is described U.S. Pat. No. 5,015,580; and, the commercially available BioRad (Biolistics) PDS-2000 particle acceleration instrument; see also, John, U.S. Patent No. 5,608,148; and Ellis, U.S. Patent No. 5,681,730, describing particle-mediated transformation of gymnosperms.

[0435] Protoplasts can be immobilized and injected with a nucleic acids, e.g., an expression construct. Although plant regeneration from protoplasts is not easy with cereals, plant regeneration is possible in legumes using somatic embryogenesis from protoplast derived callus. Organized tissues can be transformed with naked DNA using gene gun technique, where DNA is coated on tungsten microprojectiles, shot 1/100th the size of cells, which carry the DNA deep into cells and organelles. Transformed tissue is then induced to regenerate, usually by somatic embryogenesis. This technique has been successful in several cereal species including maize and rice.

[0436] Nucleic acids, e.g., expression constructs, can also be introduced in to plant cells using recombinant viruses. Plant cells can be transformed using viral vectors, such as, e.g., tobacco mosaic virus derived vectors (Rouwendal (1997) *Plant Mol. Biol.* 33:989-999), see Porta (1996) "Use of viral replicons for the expression of genes in plants," *Mol. Biotechnol.* 5:209-221.

[0437] Alternatively, nucleic acids, e.g., an expression construct, can be combined with suitable T-DNA flanking regions and introduced into a conventional *Agrobacterium tumefaciens* host vector. The virulence functions of the *Agrobacterium tumefaciens* host will direct the insertion of the construct and adjacent marker into the plant cell DNA when the cell is infected by the bacteria. *Agrobacterium tumefaciens*-mediated transformation techniques, including disarming and use of binary vectors, are well described in the scientific literature. See, e.g., Horsch (1984) *Science* 233:496-498; Fraley (1983) *Proc. Natl. Acad. Sci. USA* 80:4803 (1983); *Gene Transfer to Plants*, Potrykus, ed. (Springer-Verlag, Berlin 1995). The DNA in an *A. tumefaciens* cell is contained in the bacterial chromosome as well as in another structure known as a Ti (tumor-inducing) plasmid. The Ti plasmid contains a stretch of DNA termed T-DNA (~20 kb long) that is transferred to the plant cell in the infection process and a series of vir (virulence) genes that direct the infection process. *A. tumefaciens* can only infect a plant through wounds: when a plant root or stem is wounded it gives off certain chemical signals, in response to which, the vir genes of *A. tumefaciens* become activated and direct a series of events necessary for the transfer of the T-DNA from the Ti plasmid

to the plant's chromosome. The T-DNA then enters the plant cell through the wound. One speculation is that the T-DNA waits until the plant DNA is being replicated or transcribed, then inserts itself into the exposed plant DNA. In order to use *A. tumefaciens* as a transgene vector, the tumor-inducing section of T-DNA have to be removed, while retaining the T-DNA border regions and the vir genes. The transgene is then inserted between the T-DNA border regions, where it is transferred to the plant cell and becomes integrated into the plant's chromosomes.

[0438] The transformation of monocotyledonous plants may use the nucleic acids of the invention, including important cereals, see Hiei (1997) Plant Mol. Biol. 35:205-218. See also, e.g., Horsch, Science (1984) 233:496; Fraley (1983) Proc. Natl. Acad. Sci USA 80:4803; Thykjaer (1997) supra; Park (1996) Plant Mol. Biol. 32:1135-1148, discussing T-DNA integration into genomic DNA. See also D'Halluin, U.S. Patent No. 5,712,135, describing a process for the stable integration of a DNA comprising a gene that is functional in a cell of a cereal, or other monocotyledonous plant.

[0439] The third step can involve selection and regeneration of whole plants capable of transmitting the incorporated target gene to the next generation. Such regeneration techniques rely on manipulation of certain phytohormones in a tissue culture growth medium, typically relying on a biocide and/or herbicide marker that has been introduced together with the desired nucleotide sequences. Plant regeneration from cultured protoplasts is described in Evans et al., Protoplasts Isolation and Culture, Handbook of Plant Cell Culture, pp. 124-176, MacMillan Publishing Company, New York, 1983; and Binding, Regeneration of Plants, Plant Protoplasts, pp. 21-73, CRC Press, Boca Raton, 1985. Regeneration can also be obtained from plant callus, explants, organs, or parts thereof. Such regeneration techniques are described generally in Klee (1987) Ann. Rev. of Plant Phys. 38:467-486. To obtain whole plants from transgenic tissues such as immature embryos, they can be grown under controlled environmental conditions in a series of media containing nutrients and hormones, a process known as tissue culture. Once whole plants are generated and produce seed, evaluation of the progeny begins.

[0440] After the expression cassette is stably incorporated in transgenic plants, it can be introduced into other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed. Since transgenic expression of the nucleic acids of the invention leads to phenotypic changes, plants comprising the recombinant nucleic acids of the invention can be sexually crossed with a second plant to obtain a final product. Thus, the seed of the invention can be derived from a cross between two transgenic plants of the invention, or a cross between a plant of the invention and another plant. The desired effects (e.g., expression of the polypeptides of the invention to produce a plant in which flowering behavior is altered) can be enhanced when both parental plants express the polypeptides (xylanase) of the invention. The desired effects can be passed to future plant generations by standard propagation means.

[0441] The nucleic acids and polypeptides of the invention are expressed in or inserted in any plant or seed. Transgenic plants of the invention can be dicotyledonous or monocotyledonous. Examples of monocot transgenic plants of the invention are grasses, such as meadow grass (blue grass, *Poa*), forage grass such as festuca, lolium, temperate grass, such as *Agrostis*, and cereals, e.g., wheat, oats, rye, barley, rice, sorghum, and maize (corn). Examples of dicot transgenic plants of the invention are tobacco, legumes, such as lupins, potato, sugar beet, pea, bean and soybean, and cruciferous plants (family *Brassicaceae*), such as cauliflower, rape seed, and the closely related model organism *Arabidopsis thaliana*. Thus, the transgenic plants and seeds of the invention include a broad range of plants, including, but not limited to, species from the genera *Anacardium*, *Arachis*, *Asparagus*, *Atropa*, *Avena*, *Brassica*, *Citrus*, *Citrullus*, *Capsicum*, *Carthamus*, *Cocos*, *Coffea*, *Cucumis*, *Cucurbita*, *Daucus*, *Elaeis*, *Fragaria*, *Glycine*, *Gossypium*, *Helianthus*, *Heterocallis*, *Hordeum*, *Hyoscyamus*, *Lactuca*, *Linum*, *Lolium*, *Lupinus*, *Lycopersicon*, *Malus*, *Manihot*, *Majorana*, *Medicago*, *Nicotiana*, *Olea*, *Oryza*, *Panicum*, *Pennisetum*,

Persea, *Phaseolus*, *Pistachia*, *Pisum*, *Pyrus*, *Prunus*, *Raphanus*, *Ricinus*, *Secale*, *Senecio*, *Sinapis*, *Solanum*, *Sorghum*, *Theobromus*, *Trigonella*, *Triticum*, *Vicia*, *Vitis*, *Vigna*, and *Zea*. Transgenic plants and seeds of the invention can be any monocot or dicot, e.g., a monocot corn, sugarcane, rice, wheat, barley, switchgrass or *Miscanthus*; or a dicot oilseed crop, soy, canola, rapeseed, flax, cotton, palm oil, sugar beet, peanut, tree, poplar or lupine.

[0442] In alternative embodiments, the nucleic acids of the invention are expressed in plants (and/or their seeds) which contain fiber cells, including, e.g., cotton, silk cotton tree (Kapok, *Ceiba pentandra*), desert willow, creosote bush, winterfat, balsa, ramie, kenaf, hemp, roselle, jute, sisal abaca and flax. In alternative embodiments, the transgenic plants of the invention can be members of the genus *Gossypium*, including members of any *Gossypium* species, such as *G. arboreum*, *G. herbaceum*, *G. barbadense*, and *G. hirsutum*.

[0443] The invention also provides for transgenic plants (and/or their seeds) to be used for producing large amounts of the polypeptides (xylanase or antibody) of the invention. For example, see Palmgren (1997) Trends Genet. 13:348; Chong (1997) Transgenic Res. 6:289-296 (producing human milk protein beta-casein in transgenic potato plants using an auxin-inducible, bidirectional mannopine synthase (mas1',2') promoter with *Agrobacterium tumefaciens*-mediated leaf disc transformation methods).

[0444] Using known procedures, one of skill can screen for plants (and/or their seeds) of the invention by detecting the increase or decrease of transgene mRNA or protein in transgenic plants. Means for detecting and quantitation of mRNAs or proteins are well known in the art.

Polypeptides and peptides

[0445] In one aspect, the invention provides isolated, synthetic or recombinant polypeptides and peptides having xylanase activity, or polypeptides and peptides capable of generating an antibody that specifically binds to a xylanase, including an enzyme of this invention as described herein. The disclosure includes a genus of polypeptides having various sequence identities based on the exemplary SEQ ID NO:2, SEQ ID NO:4, SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, SEQ ID NO:16, SEQ ID NO:18, SEQ ID NO:20, SEQ ID NO:22 or SEQ ID NO:24; and these exemplary polypeptides have the following enzymatic activity (e.g., the xylanase of SEQ ID NO:2, is encoded e.g., by SEQ ID NO:1; the arabinofuranosidase of SEQ ID NO:14, is encoded e.g., by SEQ ID NO:13, and the like):

SEQ ID NO:	Name	Activity
1, 2	xyl 11	Xylanase
11, 12	control xylanase	Xylanase
13, 14		Arabinofuranosidase
15, 16		Xylanase
17, 18		Oligomerase
19, 20		B-glucosidase
21, 22		Arabinofuranosidase
23, 24		Beta-xylosidase
3, 4		Endoglucanase
5, 6		Oligomerase
7, 8		Cellobiohydrolase

SEQ ID NO:	Name	Activity
9, 10		Cellobiohydrolase

[0446] The disclosure also provides enzyme-encoding nucleic acids with a common novelty in that they encode a subset of xylanases, or a clade, comprising the "X14 module". The disclosure also provides enzyme-encoding nucleic acids with a common novelty in that they encode a clade comprising the "X14 module" (see, e.g., J. Bacteriol. 2002 August; 184(15): 4124-4133). X14-comprising xylanase members include SEQ ID NO:2 having one or more amino acid residue changes (mutations) as set forth in Table 1 and as described herein.

[0447] Chimeric enzymes, including xylanases, glucanases and/or glycosidases, have heterologous carbohydrate-binding modules (CBMs), e.g., for use in various industrial, medical, pharmaceutical, research, food and feed and food and feed supplement processing and other applications. Hydrolases, including glycosyl hydrolases (such as xylanases, glucanases) may comprise one or more CBMs of an enzyme of the invention, including the CBM-like X14 module discussed above. CBMs, e.g., X14 modules, between different enzymes can be swapped; or, alternatively, one or more CBMs of one or more enzymes can be spliced into an enzyme, e.g., a hydrolase, e.g., any glycosyl hydrolase, such as a xylanase.

[0448] Glycosyl hydrolases that utilize insoluble substrates are modular, usually comprising catalytic modules appended to one or more non-catalytic carbohydrate-binding modules (CBMs). In nature, CBMs are thought to promote the interaction of the glycosyl hydrolase with its target substrate polysaccharide. For example, as discussed above, X14 is a xylan binding module. Chimeric enzymes may have heterologous, non-natural substrates; including chimeric enzymes having multiple substrates by nature of their "spliced-in" heterologous CBMs, e.g., a spliced-in X14 module - thus giving the chimeric enzyme new specificity for xylan and galactan, or enhanced binding to xylan and galactan. The heterologous CBMs of the chimeric enzymes can be designed to be modular, i.e., to be appended to a catalytic module or catalytic domain (e.g., an active site), which also can be heterologous or can be homologous to the enzyme.

[0449] Utilization of just the catalytic module of a xylanase or a glucanase has been shown to be effective. Thus, the disclosure provides peptides and polypeptides consisting of, or comprising, modular CBM/ active site modules (e.g., X14), which can be homologously paired or joined as chimeric (heterologous) active site-CBM pairs. Thus, these chimeric polypeptides/ peptides can be used to improve or alter the performance of an individual enzyme, e.g., a xylanase enzyme. A chimeric catalytic module (comprising, e.g., at least one CBM of the disclosure, e.g., X14) can be designed to target the enzyme to particular regions of a substrate, e.g., to particular regions of a pulp. For example, this is achieved by making fusions of the xylanase and various CBMs (either a xylanase of the invention with a heterologous CBM, or, a CBM of the xylanase of the invention with another enzyme, e.g., a hydrolase, such as a xylanase. For example, CBM4, CBM6, and CBM22 are known to bind xylan and may enhance the effectiveness of the xylanase in pulp biobleaching (see, e.g., Czjzek (2001) J. Biol. Chem. 276(51):48580-7, noting that CBM4, CBM6, and CBM22 are related and CBM interact primarily with xylan). Fusion of xylanase and CBM3a or CBM3b, which bind crystalline cellulose, may help the xylanase penetrate the complex polysaccharide matrix of pulp and reach inaccessible xylans. Any CBM can be used, e.g., as reviewed by Boraston (2004) Biochem. J. 382:769-781:

Family	Protein	PDB
		code
CBM1	Cellulase 7A (<i>Trichoderma reesei</i>)	1CBH

Family	Protein	PDB
CBM2	Xylanase 10A (<i>Cellulomonas fimi</i>)	1EXG
	Xylanase 11A (<i>Cellulomonas fimi</i>)	2XBD
	Xylanase 11A (<i>Cellulomonas fimi</i>)	1HEH
CBM3	Scaffoldin (<i>Clostridium cellulolyticum</i>)	1G43
	Scaffoldin (<i>Clostridium thermocellum</i>)	1NBC
	Cellulase 9A (<i>Thermobifida fusca</i>)	1TF4
CBM4	Laminarinase 16A (<i>Thermotoga maritima</i>)	1GUI
	Cellulase 9B (<i>Cellulomonas fimi</i>)	1ULO;
		1GU3
	Cellulase 9B (<i>Cellulomonas fimi</i>)	1CX1
	Xylanase 10A (<i>Rhodothermus marinus</i>)	1K45
CBM5	Cellulase 5A (<i>Erwinia chrysanthemi</i>)	1AIW
	Chitinase B (<i>Serratia marcescens</i>)	1E15
CBM6	Xylanase 11A (<i>Clostridium thermocellum</i>)	1UXX
	Xylanase 11A (<i>Clostridium stercorarium</i>)	1NAE
	Xylanase 11A (<i>Clostridium stercorarium</i>)	1UY4
	Endoglucanase 5A (<i>Cellvibrio mixtus</i>)	1UZ0
CBM9	Xylanase 10A (<i>Thermotoga maritima</i>)	1I8A
CBM10	Xylanase 10A (<i>Cellvibrio japonicus</i>)	1QLD
CBM12	Chitinase Chi1 (<i>Bacillus circulans</i>)	1ED7
CBM13*	Xylanase 10A (<i>Streptomyces olivaceoviridis</i>)	1XYF
	Xylanase 10A (<i>Streptomyces lividans</i>)	1MC9
	Ricin toxin B-chain (<i>Ricinus communis</i>)	2AAI
	Abrin (<i>Abrus precatorius</i>)	1ABR
CBM14	Tachycitin (<i>Tachypleus tridentatus</i>)	1DQC
CBM15	Xylanase 10C (<i>Cellvibrio japonicus</i>)	1GNY
CBM17	Cellulase 5A (<i>Clostridium cellulovorans</i>)	1J83
CBM18*	Agglutinin (<i>Triticum aestivum</i>)	1WGC
	Antimicrobial peptide (<i>Amaranthus caudatus</i>)	1MMC
	Chitinase/agglutinin (<i>Urtica dioica</i>)	1EIS
CBM20*	Glucoamylase (<i>Aspergillus niger</i>)	1AC0
	β -amylase (<i>Bacillus cereus</i>)	1CQY
CBM22	Xylanase 10B (<i>Clostridium thermocellum</i>)	1DY0
CBM27	Mannanase 5A (<i>Thermotoga maritima</i>)	1OF4
CBM28	Cellulase 5A (<i>Bacillus</i> sp. 1139)	1UWW
CBM29	Non-catalytic protein 1 (<i>Pyromyces equi</i>)	1GWK
CBM32	Sialidase 33A (<i>Micromonospora viridifaciens</i>)	1EUU
	Galactose oxidase (<i>Cladobotryum dendroides</i>)	1GOF

Family	Protein	PDB
CBM34*	α -Amylase 13A (<i>Thermoactinomyces vulgaris</i>)	1UH2
	Neopullulanase (<i>Geobacillus stearothermophilus</i>)	1J0H
CBM36	Xylanase 43A (<i>Paenibacillus polymyxa</i>)	1UX7
*These families contain too many structure entries to list them all so only representatives are given.		

[0450] Chimeric hydrolases, e.g., a fusion of a glycosidase with different (e.g., heterologous) CBMs may target the enzyme to particular insoluble polysaccharides to enhance performance in an application. The chimeric glycosidase may comprise an enzyme of the invention. The chimeric enzyme may comprise fusions of different CBMs to enhance pulp biobleaching performance, e.g., to achieve greater percentage reduction of bleaching chemicals. Methods comprise recombining different CBMs with different xylanases and screening the resultant chimerics to find the best combination for a particular application or substrate.

[0451] In other variations, one, two, three, four or five or more residues are removed from the carboxy- or amino- terminal ends of any polypeptide of the invention. Another variation includes modifying any residue to increase or decrease pI of a polypeptide, e.g., removing or modifying (e.g., to another amino acid) a glutamate. This method was used as a general scheme for improving the enzyme's properties without creating regulatory issues since no amino acids are mutated; and this general scheme can be used with any polypeptide of the invention.

[0452] In one aspect, the polypeptide has a xylanase activity; for example, wherein the xylanase activity can comprise hydrolyzing a glycosidic bond in a polysaccharide, e.g., a xylan. In one aspect, the polypeptide has a xylanase activity comprising catalyzing hydrolysis of internal β -1,4-xylosidic linkages. In one aspect, the xylanase activity comprises an endo-1,4-beta-xylanase activity. In one aspect, the xylanase activity comprises hydrolyzing a xylan to produce a smaller molecular weight xylose and xylo-oligomer. In one aspect, the xylan comprises an arabinoxylan, such as a water soluble arabinoxylan.

[0453] In one aspect, the xylanase activity comprises hydrolyzing a polysaccharide to produce a smaller molecular weight polysaccharide or oligomer.

[0454] Any xylanase assay known in the art can be used to determine if a polypeptide has xylanase activity and is within scope of the invention. For example, reducing sugar assays such as the Nelson-Somogyi method or the dinitrosalicylic acid (DNS) method can be used to assay for the product sugars (and thus, xylanase activity). In one aspect, reactions are carried out by mixing and incubating a dilution of the enzyme preparation with a known amount of substrate at a buffered pH and set temperature. Xylanase assays are similar to cellulase assays except that a solution of xylan (e.g., oat spelts or birch) is substituted for CMC or filter paper. The DNS assay is easier to use than the Nelson-Somogyi assay. The DNS assay is satisfactory for cellulase activities, but tends to over estimate xylanase activity. The Somogyi-Nelson procedure is more accurate in the determination of reducing sugars, to measure specific activities and to quantify the total amount of xylanase produced in the optimized growth conditions, see, e.g., Breuil (1985) Comparison of the 3, 5-dinitrosalicylic acid and Nelson-Somogyi methods of assaying for reducing sugars and determining cellulase activity, *Enzyme Microb. Technol.* 7:327-332; Somogyi, M. 1952, Notes on sugar determination, *J. Biol. Chem.* 195:19-23. The disclosure incorporates use of any reducing sugar assay, e.g., by Nelson-Somogyi, e.g., based on references Nelson, N. (1944) *J. Biol. Chem.* 153:375-380, and Somogyi, M. (1952) *J. Biol. Chem.* 195:19-23.

[0455] The polypeptides of the invention include xylanases in an active or inactive form. For example, the polypeptides of the invention include proproteins before "maturation" or processing of prepro sequences, e.g., by a proprotein-processing enzyme, such as a proprotein convertase to generate an "active" mature protein. The polypeptides of the invention include xylanases inactive for other reasons, e.g., before "activation" by a post-translational processing event, e.g., an endo- or exo-peptidase or proteinase action, a phosphorylation event, an amidation, a glycosylation or a sulfation, a dimerization event, and the like. The polypeptides of the invention include all active forms, including active subsequences, e.g., catalytic domains or active sites, of the xylanase.

[0456] Methods for identifying "prepro" domain sequences and signal sequences are well known in the art, see, e.g., Van de Ven (1993) Crit. Rev. Oncog. 4(2):115-136. For example, to identify a prepro sequence, the protein is purified from the extracellular space and the N-terminal protein sequence is determined and compared to the unprocessed form.

[0457] The invention includes polypeptides with or without a signal sequence and/or a prepro sequence. The invention includes polypeptides with heterologous signal sequences and/or prepro sequences. The prepro sequence (including a sequence of the invention used as a heterologous prepro domain) can be located on the amino terminal or the carboxy terminal end of the protein. The invention also includes isolated, synthetic or recombinant signal sequences, prepro sequences and catalytic domains (e.g., "active sites") comprising sequences of the invention.

[0458] The percent sequence identity can be over the full length of the polypeptide, or, the identity can be over a region of at least about 50, 60, 70, 80, 90, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700 or more residues. Polypeptides of the invention can also be shorter than the full length of exemplary polypeptides. In alternative aspects, the invention provides polypeptides (peptides, fragments) ranging in size between about 5 and the full length of a polypeptide exemplary sizes being of about 5, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 100, 125, 150, 175, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, or more residues, e.g., contiguous residues of an exemplary xylanase of the invention.

[0459] Peptides of the invention (e.g., a subsequence of an exemplary polypeptide of the invention) can be useful as, e.g., labeling probes, antigens, toleragens, motifs, xylanase active sites (e.g., "catalytic domains"), signal sequences and/or prepro domains.

[0460] Polypeptides and peptides of the invention can be isolated from natural sources, be synthetic, or be recombinantly generated polypeptides. Peptides and proteins can be recombinantly expressed *in vitro* or *in vivo*. The peptides and polypeptides of the invention can be made and isolated using any method known in the art. Polypeptide and peptides of the invention can also be synthesized, whole or in part, using chemical methods well known in the art. See e.g., Caruthers (1980) Nucleic Acids Res. Symp. Ser. 215-223; Horn (1980) Nucleic Acids Res. Symp. Ser. 225-232; Banga, A.K., Therapeutic Peptides and Proteins, Formulation, Processing and Delivery Systems (1995) Technomic Publishing Co., Lancaster, PA. For example, peptide synthesis can be performed using various solid-phase techniques (see e.g., Roberge (1995) Science 269:202; Merrifield (1997) Methods Enzymol. 289:3-13) and automated synthesis may be achieved, e.g., using the ABI 431A Peptide Synthesizer (Perkin Elmer) in accordance with the instructions provided by the manufacturer.

[0461] The peptides and polypeptides of the invention can also be glycosylated. The glycosylation can be added post-translationally either chemically or by cellular biosynthetic mechanisms, wherein the later incorporates the use of known glycosylation motifs, which can be native to the sequence or can be added as a peptide or added in the nucleic acid coding sequence. The glycosylation can be O-linked or N-linked.

[0462] "Amino acid" or "amino acid sequence" as used herein refer to an oligopeptide, peptide, polypeptide, or protein sequence, or to a fragment, portion, or subunit of any of these and to naturally occurring or synthetic molecules. "Amino acid" or "amino acid sequence" include an oligopeptide, peptide, polypeptide, or protein sequence, or to a fragment, portion, or subunit of any of these, and to naturally occurring or synthetic molecules. The term "polypeptide" as used herein, refers to amino acids joined to each other by peptide bonds or modified peptide bonds, *i.e.*, peptide isosteres and may contain modified amino acids other than the 20 gene-encoded amino acids. The polypeptides may be modified by either natural processes, such as post-translational processing, or by chemical modification techniques that are well known in the art. Modifications can occur anywhere in the polypeptide, including the peptide backbone, the amino acid side-chains and the amino or carboxyl termini. It will be appreciated that the same type of modification may be present in the same or varying degrees at several sites in a given polypeptide. Also a given polypeptide may have many types of modifications. Modifications include acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of a phosphatidylinositol, cross-linking cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cysteine, formation of pyroglutamate, formylation, gamma-carboxylation, glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, pegylation, xylan hydrolase processing, phosphorylation, prenylation, racemization, selenoylation, sulfation and transfer-RNA mediated addition of amino acids to protein such as arginylation. (See Creighton, T.E., *Proteins - Structure and Molecular Properties* 2nd Ed., W.H. Freeman and Company, New York (1993); *Posttranslational Covalent Modification of Proteins*, B.C. Johnson, Ed., Academic Press, New York, pp. 1-12 (1983)). The peptides and polypeptides of the invention also include all "mimetic" and "peptidomimetic" forms, as described in further detail, below.

[0463] "Recombinant" polypeptides or proteins refer to polypeptides or proteins produced by recombinant DNA techniques; *i.e.*, produced from cells transformed by an exogenous DNA construct encoding the desired polypeptide or protein. "Synthetic" nucleic acids (including oligonucleotides), polypeptides or proteins of the invention include those prepared by any chemical synthesis, *e.g.*, as described, below. Solid-phase chemical peptide synthesis methods can also be used to synthesize the polypeptide or fragments of the invention. Such method have been known in the art since the early 1960's (Merrifield, R. B., *J. Am. Chem. Soc.*, 85:2149-2154, 1963) (See also Stewart, J. M. and Young, J. D., *Solid Phase Peptide Synthesis*, 2nd Ed., Pierce Chemical Co., Rockford, Ill., pp. 11-12)) and have recently been employed in commercially available laboratory peptide design and synthesis kits (Cambridge Research Biochemicals). Such commercially available laboratory kits have generally utilized the teachings of H. M. Geysen et al, *Proc. Natl. Acad. Sci., USA*, 81:3998 (1984) and provide for synthesizing peptides upon the tips of a multitude of "rods" or "pins" all of which are connected to a single plate. When such a system is utilized, a plate of rods or pins is inverted and inserted into a second plate of corresponding wells or reservoirs, which contain solutions for attaching or anchoring an appropriate amino acid to the pin's or rod's tips. By repeating such a process step, *i.e.*, inverting and inserting the rod's and pin's tips into appropriate solutions, amino acids are built into desired peptides. In addition, a number of available Fmoc peptide synthesis systems are available. For example, assembly of a polypeptide or fragment can be carried out on a solid support using an Applied Biosystems, Inc. Model 431A automated peptide synthesizer. Such equipment provides ready access to the peptides of the invention, either by direct synthesis or by synthesis of a series of fragments that can be coupled using other known techniques.

[0464] "Fragments" or "enzymatically active fragments" as used herein are a portion of an amino acid sequence (encoding a protein) which retains at least one functional activity of the protein to which it is related. Fragments can have the same or substantially the same amino acid sequence as the naturally

occurring protein. "Substantially the same" means that an amino acid sequence is largely, but not entirely, the same, but retains at least one functional activity of the sequence to which it is related. In general two amino acid sequences are "substantially the same" or "substantially homologous" if they are at least about 85% identical. Fragments which have different three dimensional structures as the naturally occurring protein are also included. An example of this, is a "pro-form" molecule, such as a low activity proprotein that can be modified by cleavage to produce a mature enzyme with significantly higher activity.

[0465] The peptides and polypeptides of the invention, as defined above, include all "mimetic" and "peptidomimetic" forms. The terms "mimetic" and "peptidomimetic" refer to a synthetic chemical compound which has substantially the same structural and/or functional characteristics of the polypeptides of the invention. The mimetic can be either entirely composed of synthetic, non-natural analogues of amino acids, or, is a chimeric molecule of partly natural peptide amino acids and partly non-natural analogs of amino acids. The mimetic can also incorporate any amount of natural amino acid conservative substitutions as long as such substitutions also do not substantially alter the mimetic's structure and/or activity. As with polypeptides of the invention which are conservative variants, routine experimentation will determine whether a mimetic is within the scope of the invention, i.e., that its structure and/or function is not substantially altered. Thus, in one aspect, a mimetic composition is within the scope of the invention if it has a xylanase activity.

[0466] Polypeptide mimetic compositions of the invention can contain any combination of non-natural structural components. In alternative aspect, mimetic compositions of the invention include one or all of the following three structural groups: a) residue linkage groups other than the natural amide bond ("peptide bond") linkages; b) non-natural residues in place of naturally occurring amino acid residues; or c) residues which induce secondary structural mimicry, i.e., to induce or stabilize a secondary structure, e.g., a beta turn, gamma turn, beta sheet, alpha helix conformation, and the like. For example, a polypeptide of the invention can be characterized as a mimetic when all or some of its residues are joined by chemical means other than natural peptide bonds. Individual peptidomimetic residues can be joined by peptide bonds, other chemical bonds or coupling means, such as, e.g., glutaraldehyde, N-hydroxysuccinimide esters, bifunctional maleimides, N,N'-dicyclohexylcarbodiimide (DCC) or N,N'-diisopropylcarbodiimide (DIC). Linking groups that can be an alternative to the traditional amide bond ("peptide bond") linkages include, e.g., ketomethylene (e.g., $-C(=O)-CH_2-$ for $-C(=O)-NH-$), aminomethylene (CH_2-NH), ethylene, olefin ($CH=CH$), ether (CH_2-O), thioether (CH_2-S), tetrazole (CN_4-), thiazole, retroamide, thioamide, or ester (see, e.g., Spatola (1983) in *Chemistry and Biochemistry of Amino Acids, Peptides and Proteins*, Vol. 7, pp 267-357, "Peptide Backbone Modifications," Marcell Dekker, NY).

[0467] A polypeptide of the invention can also be characterized as a mimetic by containing all or some non-natural residues in place of naturally occurring amino acid residues. Non-natural residues are well described in the scientific and patent literature; a few exemplary non-natural compositions useful as mimetics of natural amino acid residues and guidelines are described below. Mimetics of aromatic amino acids can be generated by replacing by, e.g., D- or L- naphylalanine; D- or L- phenylglycine; D- or L-2 thieneylalanine; D- or L-1, - 2, 3-, or 4- pyreneylalanine; D- or L-3 thieneylalanine; D- or L-(2-pyridinyl)-alanine; D- or L-(3-pyridinyl)-alanine; D- or L-(2-pyrazinyl)-alanine; D- or L-(4-isopropyl)-phenylglycine; D-(trifluoromethyl)-phenylglycine; D-(trifluoromethyl)-phenylalanine; D-p-fluoro-phenylalanine; D- or L-p-biphenylphenylalanine; D- or L-p-methoxy-phenylphenylalanine; D- or L-2-indole(alkyl)alanines; and, D- or L-alkylalanines, where alkyl can be substituted or unsubstituted methyl, ethyl, propyl, hexyl, butyl, pentyl, isopropyl, iso-butyl, sec-isotyl, isopentyl, or a non-acidic amino acids. Aromatic rings of a non-natural amino acid include, e.g., thiazolyl, thiophenyl, pyrazolyl, benzimidazolyl, naphthyl, furanyl, pyrrolyl, and pyridyl aromatic rings.

[0468] Mimetics of acidic amino acids can be generated by substitution by, e.g., non-carboxylate amino acids while maintaining a negative charge; (phosphono)alanine; sulfated threonine. Carboxyl side groups (e.g., aspartyl or glutamyl) can also be selectively modified by reaction with carbodiimides (R'-N-C-N-R') such as, e.g., 1-cyclohexyl-3(2-morpholinyl-(4-ethyl) carbodiimide or 1-ethyl-3(4-azonia-4,4-dimetholpentyl) carbodiimide. Aspartyl or glutamyl can also be converted to asparaginyl and glutaminyl residues by reaction with ammonium ions. Mimetics of basic amino acids can be generated by substitution with, e.g., (in addition to lysine and arginine) the amino acids ornithine, citrulline, or (guanidino)-acetic acid, or (guanidino)alkyl-acetic acid, where alkyl is defined above. Nitrile derivative (e.g., containing the CN-moiety in place of COOH) can be substituted for asparagine or glutamine. Asparaginyl and glutaminyl residues can be deaminated to the corresponding aspartyl or glutamyl residues. Arginine residue mimetics can be generated by reacting arginyl with, e.g., one or more conventional reagents, including, e.g., phenylglyoxal, 2,3-butanedione, 1,2-cyclo-hexanedione, or ninhydrin, preferably under alkaline conditions. Tyrosine residue mimetics can be generated by reacting tyrosyl with, e.g., aromatic diazonium compounds or tetranitromethane. N-acetylimidizol and tetranitromethane can be used to form O-acetyl tyrosyl species and 3-nitro derivatives, respectively. Cysteine residue mimetics can be generated by reacting cysteinyl residues with, e.g., alpha-haloacetates such as 2-chloroacetic acid or chloroacetamide and corresponding amines; to give carboxymethyl or carboxyamidomethyl derivatives. Cysteine residue mimetics can also be generated by reacting cysteinyl residues with, e.g., bromo-trifluoroacetone, alpha-bromo-beta-(5-imidazolyl) propionic acid; chloroacetyl phosphate, N-alkylmaleimides, 3-nitro-2-pyridyl disulfide; methyl 2-pyridyl disulfide; p-chloromercuribenzoate; 2-chloromercuri-4 nitrophenol; or, chloro-7-nitrobenzo-oxa-1,3-diazole. Lysine mimetics can be generated (and amino terminal residues can be altered) by reacting lysinyl with, e.g., succinic or other carboxylic acid anhydrides. Lysine and other alpha-amino-containing residue mimetics can also be generated by reaction with imidoesters, such as methyl picolinimide, pyridoxal phosphate, pyridoxal, chloroborohydride, trinitro-benzenesulfonic acid, O-methylisourea, 2,4-pentanedione, and transamidase-catalyzed reactions with glyoxylate. Mimetics of methionine can be generated by reaction with, e.g., methionine sulfoxide. Mimetics of proline include, e.g., pipecolic acid, thiazolidine carboxylic acid, 3- or 4- hydroxy proline, dehydropyrolidine, 3- or 4-methylproline, or 3,3,-dimethylproline. Histidine residue mimetics can be generated by reacting histidyl with, e.g., diethylprocarbonate or para-bromophenacyl bromide. Other mimetics include, e.g., those generated by hydroxylation of proline and lysine; phosphorylation of the hydroxyl groups of seryl or threonyl residues; methylation of the alpha-amino groups of lysine, arginine and histidine; acetylation of the N-terminal amine; methylation of main chain amide residues or substitution with N-methyl amino acids; or amidation of C-terminal carboxyl groups.

[0469] A residue, e.g., an amino acid, of a polypeptide of the invention can also be replaced by an amino acid (or peptidomimetic residue) of the opposite chirality. Thus, any amino acid naturally occurring in the L-configuration (which can also be referred to as the R or S, depending upon the structure of the chemical entity) can be replaced with the amino acid of the same chemical structural type or a peptidomimetic, but of the opposite chirality, referred to as the D- amino acid, but also can be referred to as the R- or S- form.

[0470] The polypeptides of the invention may be modified by either natural processes, such as post-translational processing (e.g., phosphorylation, acylation, etc), or by chemical modification techniques. Modifications can occur anywhere in the polypeptide, including the peptide backbone, the amino acid side-chains and the amino or carboxyl termini. It will be appreciated that the same type of modification may be present in the same or varying degrees at several sites in a given polypeptide. Also a given polypeptide may have many types of modifications. Modifications include acetylation, acylation, ADP-ribosylation, amidation, covalent attachment of flavin, covalent attachment of a heme moiety, covalent attachment of a nucleotide or nucleotide derivative, covalent attachment of a lipid or lipid derivative, covalent attachment of a phosphatidylinositol, cross-linking cyclization, disulfide bond formation, demethylation, formation of covalent cross-links, formation of cysteine, formation of pyroglutamate, formylation, gamma-carboxylation,

glycosylation, GPI anchor formation, hydroxylation, iodination, methylation, myristoylation, oxidation, pegylation, proteolytic processing, phosphorylation, prenylation, racemization, selenoylation, sulfation, and transfer-RNA mediated addition of amino acids to protein such as arginylation. See, e.g., Creighton, T.E., *Proteins - Structure and Molecular Properties* 2nd Ed., W.H. Freeman and Company, New York (1993); *Posttranslational Covalent Modification of Proteins*, B.C. Johnson, Ed., Academic Press, New York, pp. 1-12 (1983).

[0471] Solid-phase chemical peptide synthesis methods can also be used to synthesize the polypeptide or fragments of the invention. Such method have been known in the art since the early 1960's (Merrifield, R. B., *J. Am. Chem. Soc.*, 85:2149-2154, 1963) (See also Stewart, J. M. and Young, J. D., *Solid Phase Peptide Synthesis*, 2nd Ed., Pierce Chemical Co., Rockford, Ill., pp. 11-12)) and have recently been employed in commercially available laboratory peptide design and synthesis kits (Cambridge Research Biochemicals). Such commercially available laboratory kits have generally utilized the teachings of H. M. Geysen et al, *Proc. Natl. Acad. Sci., USA*, 81:3998 (1984) and provide for synthesizing peptides upon the tips of a multitude of "rods" or "pins" all of which are connected to a single plate. When such a system is utilized, a plate of rods or pins is inverted and inserted into a second plate of corresponding wells or reservoirs, which contain solutions for attaching or anchoring an appropriate amino acid to the pin's or rod's tips. By repeating such a process step, i.e., inverting and inserting the rod's and pin's tips into appropriate solutions, amino acids are built into desired peptides. In addition, a number of available Fmoc peptide synthesis systems are available. For example, assembly of a polypeptide or fragment can be carried out on a solid support using an Applied Biosystems, Inc. Model 431A™ automated peptide synthesizer. Such equipment provides ready access to the peptides of the invention, either by direct synthesis or by synthesis of a series of fragments that can be coupled using other known techniques.

[0472] The invention includes xylanases of the invention with and without signal. The polypeptide comprising a signal sequence of the invention can be a xylanase of the invention or another xylanase or another enzyme or other polypeptide.

[0473] The invention includes immobilized xylanases, anti-xylanase antibodies and fragments thereof. Methods for inhibiting xylanase activity may use dominant negative mutants or anti-xylanase antibodies of the invention. Heterocomplexes, e.g., fusion proteins, heterodimers, etc., may comprise the xylanases of the invention.

[0474] Polypeptides of the invention can have a xylanase activity under various conditions, e.g., extremes in pH and/or temperature, oxidizing agents, and the like. The invention provides methods leading to alternative xylanase preparations with different catalytic efficiencies and stabilities, e.g., towards temperature, oxidizing agents and changing wash conditions. In one aspect, xylanase variants can be produced using techniques of site-directed mutagenesis and/or random mutagenesis. Directed evolution can be used to produce a great variety of xylanase variants with alternative specificities and stability.

[0475] The proteins of the invention are also useful as research reagents to identify xylanase modulators, e.g., activators or inhibitors of xylanase activity. Briefly, test samples (compounds, broths, extracts, and the like) are added to xylanase assays to determine their ability to inhibit substrate cleavage. Inhibitors identified in this way can be used in industry and research to reduce or prevent undesired proteolysis. As with xylanases, inhibitors can be combined to increase the spectrum of activity.

[0476] The enzymes of the invention are also useful as research reagents to digest proteins or in protein sequencing. For example, the xylanases may be used to break polypeptides into smaller fragments for sequencing using, e.g. an automated sequencer.

[0477] The invention also provides methods of discovering new xylanases using the nucleic acids, polypeptides and antibodies of the invention. In one aspect, phagemid libraries are screened for expression-based discovery of xylanases. In another aspect, lambda phage libraries are screened for expression-based discovery of xylanases. Screening of the phage or phagemid libraries can allow the detection of toxic clones; improved access to substrate; reduced need for engineering a host, by-passing the potential for any bias resulting from mass excision of the library; and, faster growth at low clone densities. Screening of phage or phagemid libraries can be in liquid phase or in solid phase. In one aspect, the invention provides screening in liquid phase. This gives a greater flexibility in assay conditions; additional substrate flexibility; higher sensitivity for weak clones; and ease of automation over solid phase screening.

[0478] Screening methods may use the proteins and nucleic acids of the invention and robotic automation to enable the execution of many thousands of biocatalytic reactions and screening assays in a short period of time, e.g., per day, as well as ensuring a high level of accuracy and reproducibility (see discussion of arrays, below). As a result, a library of derivative compounds can be produced in a matter of weeks. For further teachings on modification of molecules, including small molecules, see PCT/US94/09174.

[0479] Another aspect of the invention is an isolated or purified polypeptide comprising the sequence of one of the invention and sequences substantially identical thereto, or fragments comprising at least about 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof. As discussed above, such polypeptides may be obtained by inserting a nucleic acid encoding the polypeptide into a vector such that the coding sequence is operably linked to a sequence capable of driving the expression of the encoded polypeptide in a suitable host cell. For example, the expression vector may comprise a promoter, a ribosome binding site for translation initiation and a transcription terminator. The vector may also include appropriate sequences for amplifying expression.

[0480] Another aspect of the invention is polypeptides or fragments thereof which have at least about 95%, or more than about 95% homology to one of the polypeptides of the invention and sequences substantially identical thereto, or a fragment comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof. Homology may be determined using any of the programs described above which aligns the polypeptides or fragments being compared and determines the extent of amino acid identity or similarity between them. It will be appreciated that amino acid "homology" includes conservative amino acid substitutions such as those described above.

[0481] The polypeptides or fragments having homology to one of the polypeptides of the invention, or a fragment comprising at least about 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof may be obtained by isolating the nucleic acids encoding them using the techniques described above.

[0482] Alternatively, the homologous polypeptides or fragments may be obtained through biochemical enrichment or purification procedures. The sequence of potentially homologous polypeptides or fragments may be determined by xylan hydrolase digestion, gel electrophoresis and/or microsequencing. The sequence of the prospective homologous polypeptide or fragment can be compared to one of the polypeptides of the invention, or a fragment comprising at least about 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof using any of the programs described above.

[0483] The assay for determining if fragments of variants retain the enzymatic activity of the polypeptides of the invention includes the steps of: contacting the polypeptide fragment or variant with a substrate molecule under conditions which allow the polypeptide fragment or variant to function and detecting either

a decrease in the level of substrate or an increase in the level of the specific reaction product of the reaction between the polypeptide and substrate.

[0484] The polypeptides of the invention or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof may be used in a variety of applications, for example, for hydrolyzing glycosidic linkages. In such procedures, a substance containing a glycosidic linkage (e.g., a starch) is contacted with one of the polypeptides of the invention, or sequences substantially identical thereto under conditions which facilitate the hydrolysis of the glycosidic linkage.

[0485] This exploits the unique catalytic properties of enzymes. Whereas the use of biocatalysts (i.e., purified or crude enzymes, non-living or living cells) in chemical transformations normally requires the identification of a particular biocatalyst that reacts with a specific starting compound, here biocatalysts and reaction conditions may be selected that are specific for functional groups that are present in many starting compounds, such as small molecules. Each biocatalyst is specific for one functional group, or several related functional groups and can react with many starting compounds containing this functional group.

[0486] The biocatalytic reactions produce a population of derivatives from a single starting compound. These derivatives can be subjected to another round of biocatalytic reactions to produce a second population of derivative compounds. Thousands of variations of the original small molecule or compound can be produced with each iteration of biocatalytic derivatization.

[0487] Enzymes react at specific sites of a starting compound without affecting the rest of the molecule, a process which is very difficult to achieve using traditional chemical methods. This high degree of biocatalytic specificity provides the means to identify a single active compound within the library. The library is characterized by the series of biocatalytic reactions used to produce it, a so called "biosynthetic history". Screening the library for biological activities and tracing the biosynthetic history identifies the specific reaction sequence producing the active compound. The reaction sequence is repeated and the structure of the synthesized compound determined. This mode of identification, unlike other synthesis and screening approaches, does not require immobilization technologies and compounds can be synthesized and tested free in solution using virtually any type of screening assay. It is important to note, that the high degree of specificity of enzyme reactions on functional groups allows for the "tracking" of specific enzymatic reactions that make up the biocatalytically produced library.

[0488] Many of the procedural steps are performed using robotic automation enabling the execution of many thousands of biocatalytic reactions and screening assays per day as well as ensuring a high level of accuracy and reproducibility. As a result, a library of derivative compounds can be produced in a matter of weeks which would take years to produce using current chemical methods.

[0489] The disclosure provides a method for modifying small molecules, comprising contacting a polypeptide encoded by a polynucleotide described herein or enzymatically active fragments thereof with a small molecule to produce a modified small molecule. A library of modified small molecules is tested to determine if a modified small molecule is present within the library which exhibits a desired activity. A specific biocatalytic reaction which produces the modified small molecule of desired activity is identified by systematically eliminating each of the biocatalytic reactions used to produce a portion of the library and then testing the small molecules produced in the portion of the library for the presence or absence of the modified small molecule with the desired activity. The specific biocatalytic reactions which produce the modified small molecule of desired activity is in one aspect (optionally) repeated. The biocatalytic reactions are conducted with a group of biocatalysts that react with distinct structural moieties found within the structure of a small molecule, each biocatalyst is specific for one structural moiety or a group of related structural moieties; and each biocatalyst reacts with many different small molecules which contain the

distinct structural moiety.

Xylanase signal sequences, prepro and catalytic domains

[0490] Xylanase signal sequences (e.g., signal peptides (SPs)), prepro domains and catalytic domains (CDs) can be isolated, synthetic or recombinant peptides or can be part of a fusion protein, e.g., as a heterologous domain in a chimeric protein. The disclosure provides nucleic acids encoding these catalytic domains (CDs), prepro domains and signal sequences (SPs, e.g., a peptide having a sequence comprising/ consisting of amino terminal residues of a polypeptide of the invention). The disclosure provides a signal sequence comprising a peptide comprising/ consisting of a sequence as set forth in residues 1 to 12, 1 to 13, 1 to 14, 1 to 15, 1 to 16, 1 to 17, 1 to 18, 1 to 19, 1 to 20, 1 to 21, 1 to 22, 1 to 23, 1 to 24, 1 to 25, 1 to 26, 1 to 27, 1 to 28, 1 to 28, 1 to 30, 1 to 31, 1 to 32, 1 to 33, 1 to 34, 1 to 35, 1 to 36, 1 to 37, 1 to 38, 1 to 39, 1 to 40, 1 to 41, 1 to 42, 1 to 43, 1 to 44, 1 to 45, 1 to 46, 1 to 47, 1 to 48, 1 to 49 or 1 to 50, of a polypeptide of the invention.

[0491] The xylanase signal sequences (SPs) and/or prepro sequences can be isolated peptides, or, sequences joined to another xylanase or a non-xylanase polypeptide, e.g., as a fusion (chimeric) protein. Polypeptides comprise xylanase signal sequences of the invention. Polypeptides comprise xylanase signal sequences SPs and/or prepro comprise sequences heterologous to a xylanase of the invention (e.g., a fusion protein comprising an SP and/or prepro and sequences from another xylanase or a non-xylanase protein). Xylanases of the invention may have heterologous SPs and/or prepro sequences, e.g., sequences with a yeast signal sequence. A xylanase of the invention can comprise a heterologous SP and/or prepro in a vector, e.g., a pPIC series vector (Invitrogen, Carlsbad, CA).

[0492] In one aspect, SPs and/or prepro sequences are identified following identification of novel xylanase polypeptides. The pathways by which proteins are sorted and transported to their proper cellular location are often referred to as protein targeting pathways. One of the most important elements in all of these targeting systems is a short amino acid sequence at the amino terminus of a newly synthesized polypeptide called the signal sequence. This signal sequence directs a protein to its appropriate location in the cell and is removed during transport or when the protein reaches its final destination. Most lysosomal, membrane, or secreted proteins have an amino-terminal signal sequence that marks them for translocation into the lumen of the endoplasmic reticulum. More than 100 signal sequences for proteins in this group have been determined. The signal sequences can vary in length from between about 11 to 41, or between about 13 to 36 amino acid residues. Various methods of recognition of signal sequences are known to those of skill in the art. For example, in one aspect, novel xylanase signal peptides are identified by a method referred to as SignalP. SignalP uses a combined neural network which recognizes both signal peptides and their cleavage sites; see, e.g., Nielsen (1997) "Identification of prokaryotic and eukaryotic signal peptides and prediction of their cleavage sites." Protein Engineering 10:1-6.

[0493] It should be understood that in some aspects xylanases of the invention may not have SPs and/or prepro sequences, or "domains." In one aspect, the invention provides the xylanases of the invention lacking all or part of an SP and/or a prepro domain. In one aspect, the invention provides a nucleic acid sequence encoding a signal sequence (SP) and/or prepro from one xylanase operably linked to a nucleic acid sequence of a different xylanase or, in one aspect (optionally), a signal sequence (SPs) and/or prepro domain from a non-xylanase protein may be desired.

[0494] The invention also provides isolated, synthetic or recombinant polypeptides comprising signal sequences (SPs), prepro domain and/or catalytic domains (CDs) of the invention and heterologous

sequences. The heterologous sequences are sequences not naturally associated (e.g., to a xylanase) with an SP, prepro domain and/or CD. The sequence to which the SP, prepro domain and/or CD are not naturally associated can be on the SP's, prepro domain and/or CD's amino terminal end, carboxy terminal end, and/or on both ends of the SP and/or CD. In one aspect, the invention provides an isolated, synthetic or recombinant polypeptide comprising (or consisting of) a polypeptide comprising a signal sequence (SP), prepro domain and/or catalytic domain (CD) of the invention with the proviso that it is not associated with any sequence to which it is naturally associated (e.g., a xylanase sequence). Similarly in one aspect, the invention provides isolated, synthetic or recombinant nucleic acids encoding these polypeptides. Thus, in one aspect, the isolated, synthetic or recombinant nucleic acid of the invention comprises coding sequence for a signal sequence (SP), prepro domain and/or catalytic domain (CD) of the invention and a heterologous sequence (i.e., a sequence not naturally associated with the a signal sequence (SP), prepro domain and/or catalytic domain (CD) of the invention). The heterologous sequence can be on the 3' terminal end, 5' terminal end, and/or on both ends of the SP, prepro domain and/or CD coding sequence.

Hybrid (chimeric) xylanases and peptide libraries

[0495] Hybrid xylanases and fusion proteins, including peptide libraries, may comprise sequences of the invention. The peptide libraries can be used to isolate peptide modulators (e.g., activators or inhibitors) of targets, such as xylanase substrates, receptors, enzymes. The peptide libraries can be used to identify formal binding partners of targets, such as ligands, e.g., cytokines, hormones and the like. Chimeric proteins may comprise a signal sequence (SP), prepro domain and/or catalytic domain (CD) of the disclosure or a combination thereof and a heterologous sequence (see above).

[0496] The fusion proteins (e.g., the peptide moiety) may be conformationally stabilized (relative to linear peptides) to allow a higher binding affinity for targets. Fusions may be of xylanases of the invention and other peptides, including known and random peptides. They can be fused in such a manner that the structure of the xylanases is not significantly perturbed and the peptide is metabolically or structurally conformationally stabilized. This allows the creation of a peptide library that is easily monitored both for its presence within cells and its quantity.

[0497] Amino acid sequence variants of the invention can be characterized by a predetermined nature of the variation, a feature that sets them apart from a naturally occurring form, e.g., an allelic or interspecies variation of a xylanase sequence. In one aspect, the variants of the invention exhibit the same qualitative biological activity as the naturally occurring analogue. Alternatively, the variants can be selected for having modified characteristics. In one aspect, while the site or region for introducing an amino acid sequence variation is predetermined, the mutation per se need not be predetermined. For example, in order to optimize the performance of a mutation at a given site, random mutagenesis may be conducted at the target codon or region and the expressed xylanase variants screened for the optimal combination of desired activity. Techniques for making substitution mutations at predetermined sites in DNA having a known sequence are well known, as discussed herein for example, M13 primer mutagenesis and PCR mutagenesis. Screening of the mutants can be done using, e.g., assays of xylan hydrolysis. In alternative aspects, amino acid substitutions can be single residues; insertions can be on the order of from about 1 to 20 amino acids, although considerably larger insertions can be done. Deletions can range from about 1 to about 20, 30, 40, 50, 60, 70 residues or more. To obtain a final derivative with the optimal properties, substitutions, deletions, insertions or any combination thereof may be used. Generally, these changes are done on a few amino acids to minimize the alteration of the molecule. However, larger changes may be tolerated in certain circumstances.

[0498] The invention provides xylanases where the structure of the polypeptide backbone, the secondary or the tertiary structure, e.g., an alpha-helical or beta-sheet structure, has been modified. In one aspect, the charge or hydrophobicity has been modified. In one aspect, the bulk of a side chain has been modified. Substantial changes in function or immunological identity are made by selecting substitutions that are less conservative. For example, substitutions can be made which more significantly affect: the structure of the polypeptide backbone in the area of the alteration, for example an alpha-helical or a beta-sheet structure; a charge or a hydrophobic site of the molecule, which can be at an active site; or a side chain. The invention provides substitutions in polypeptide of the invention where (a) a hydrophilic residues, e.g. seryl or threonyl, is substituted for (or by) a hydrophobic residue, e.g. leucyl, isoleucyl, phenylalanyl, valyl or alanyl; (b) a cysteine or proline is substituted for (or by) any other residue; (c) a residue having an electropositive side chain, e.g. lysyl, arginyl, or histidyl, is substituted for (or by) an electronegative residue, e.g. glutamyl or aspartyl; or (d) a residue having a bulky side chain, e.g. phenylalanine, is substituted for (or by) one not having a side chain, e.g. glycine. The variants can exhibit the same qualitative biological activity (i.e. xylanase activity) although variants can be selected to modify the characteristics of the xylanases as needed.

[0499] In one aspect, xylanases of the invention comprise epitopes or purification tags, signal sequences or other fusion sequences, etc. In one aspect, the xylanases of the invention can be fused to a random peptide to form a fusion polypeptide. By "fused" or "operably linked" herein is meant that the random peptide and the xylanase are linked together, in such a manner as to minimize the disruption to the stability of the xylanase structure, e.g., it retains xylanase activity. The fusion polypeptide (or fusion polynucleotide encoding the fusion polypeptide) can comprise further components as well, including multiple peptides at multiple loops.

[0500] In one aspect, the peptides and nucleic acids encoding them are randomized, either fully randomized or they are biased in their randomization, e.g. in nucleotide/residue frequency generally or per position. "Randomized" means that each nucleic acid and peptide consists of essentially random nucleotides and amino acids, respectively. In one aspect, the nucleic acids which give rise to the peptides can be chemically synthesized, and thus may incorporate any nucleotide at any position. Thus, when the nucleic acids are expressed to form peptides, any amino acid residue may be incorporated at any position. The synthetic process can be designed to generate randomized nucleic acids, to allow the formation of all or most of the possible combinations over the length of the nucleic acid, thus forming a library of randomized nucleic acids. The library can provide a sufficiently structurally diverse population of randomized expression products to affect a probabilistically sufficient range of cellular responses to provide one or more cells exhibiting a desired response. Thus, the invention provides an interaction library large enough so that at least one of its members will have a structure that gives it affinity for some molecule, protein, or other factor.

[0501] Xylanases are multidomain enzymes that may consist of a signal peptide, a carbohydrate binding module, a xylanase catalytic domain, a linker and/or another catalytic domain.

[0502] Chimeric polypeptides may encode biologically active hybrid polypeptides (e.g., hybrid xylanases). The original polynucleotides may encode biologically active polypeptides. New hybrid polypeptides may be produced by utilizing cellular processes which integrate the sequence of the original polynucleotides such that the resulting hybrid polynucleotide encodes a polypeptide demonstrating activities derived from the original biologically active polypeptides. For example, the original polynucleotides may encode a particular enzyme from different microorganisms. An enzyme encoded by a first polynucleotide from one organism or variant may, for example, function effectively under a particular environmental condition, e.g. high salinity. An enzyme encoded by a second polynucleotide from a different organism or variant may function effectively under a different environmental condition, such as extremely high temperatures. A hybrid

polynucleotide containing sequences from the first and second original polynucleotides may encode an enzyme which exhibits characteristics of both enzymes encoded by the original polynucleotides. Thus, the enzyme encoded by the hybrid polynucleotide may function effectively under environmental conditions shared by each of the enzymes encoded by the first and second polynucleotides, *e.g.*, high salinity and extreme temperatures.

[0503] Glycosidase hydrolases were first classified into families in 1991, see, *e.g.*, Henrissat (1991) *Biochem. J.* 280:309-316. Since then, the classifications have been continually updated, see, *e.g.*, Henrissat (1993) *Biochem. J.* 293:781-788; Henrissat (1996) *Biochem. J.* 316:695-696; Henrissat (2000) *Plant Physiology* 124:1515-1519. There are 87 identified families of glycosidase hydrolases. The xylanases of the invention may be categorized in families 8, 10, 11, 26 and 30. In one aspect, the invention also provides xylanase-encoding nucleic acids with a common novelty in that they are derived from a common family, *e.g.*, 11.

[0504] A hybrid polypeptide may exhibit specialized enzyme activity not displayed in the original enzymes. For example, following recombination and/or reductive reassortment of polynucleotides encoding hydrolase activities, the resulting hybrid polypeptide encoded by a hybrid polynucleotide can be screened for specialized hydrolase activities obtained from each of the original enzymes, *i.e.* the type of bond on which the hydrolase acts and the temperature at which the hydrolase functions. Thus, for example, the hydrolase may be screened to ascertain those chemical functionalities which distinguish the hybrid hydrolase from the original hydrolases, such as: (a) amide (peptide bonds), *i.e.*, xylanases; (b) ester bonds, *i.e.*, esterases and lipases; (c) acetals, *i.e.*, glycosidases and, for example, the temperature, pH or salt concentration at which the hybrid polypeptide functions.

[0505] Sources of the original polynucleotides may be isolated from individual organisms ("isolates"), collections of organisms that have been grown in defined media ("enrichment cultures"), or, uncultivated organisms ("environmental samples"). The use of a culture-independent approach to derive polynucleotides encoding novel bioactivities from environmental samples is most preferable since it allows one to access untapped resources of biodiversity.

[0506] "Environmental libraries" are generated from environmental samples and represent the collective genomes of naturally occurring organisms archived in cloning vectors that can be propagated in suitable prokaryotic hosts. Because the cloned DNA is initially extracted directly from environmental samples, the libraries are not limited to the small fraction of prokaryotes that can be grown in pure culture. Additionally, a normalization of the environmental DNA present in these samples could allow more equal representation of the DNA from all of the species present in the original sample. This can dramatically increase the efficiency of finding interesting genes from minor constituents of the sample which may be under-represented by several orders of magnitude compared to the dominant species.

[0507] For example, gene libraries generated from one or more uncultivated microorganisms are screened for an activity of interest. Potential pathways encoding bioactive molecules of interest are first captured in prokaryotic cells in the form of gene expression libraries. Polynucleotides encoding activities of interest are isolated from such libraries and introduced into a host cell. The host cell is grown under conditions which promote recombination and/or reductive reassortment creating potentially active biomolecules with novel or enhanced activities.

[0508] Additionally, subcloning may be performed to further isolate sequences of interest. In subcloning, a portion of DNA is amplified, digested, generally by restriction enzymes, to cut out the desired sequence, the desired sequence is ligated into a recipient vector and is amplified. At each step in subcloning, the

portion is examined for the activity of interest, in order to ensure that DNA that encodes the structural protein has not been excluded. The insert may be purified at any step of the subcloning, for example, by gel electrophoresis prior to ligation into a vector or where cells containing the recipient vector and cells not containing the recipient vector are placed on selective media containing, for example, an antibiotic, which will kill the cells not containing the recipient vector. Specific methods of subcloning cDNA inserts into vectors are well-known in the art (Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory Press (1989)). The enzymes of the invention may be subclones. Such subclones may differ from the parent clone by, for example, length, a mutation, a tag or a label.

[0509] It should be understood that some of the xylanases of the invention may or may not contain signal sequences. It may be desirable to include a nucleic acid sequence encoding a signal sequence from one xylanase operably linked to a nucleic acid sequence of a different xylanase or, in one aspect (optionally), a signal sequence from a non-xylanase protein may be desired.

[0510] The microorganisms from which the polynucleotide may be prepared include prokaryotic microorganisms, such as *Eubacteria* and *Archaeobacteria* and lower eukaryotic microorganisms such as fungi, some algae and protozoa. Polynucleotides may be isolated from environmental samples in which case the nucleic acid may be recovered without culturing of an organism or recovered from one or more cultured organisms. In one aspect, such microorganisms may be extremophiles, such as hyperthermophiles, psychrophiles, psychrotrophs, halophiles, barophiles and acidophiles. Polynucleotides encoding enzymes isolated from extremophilic microorganisms can be used. Such enzymes may function at temperatures above 100°C in terrestrial hot springs and deep sea thermal vents, at temperatures below 0°C in arctic waters, in the saturated salt environment of the Dead Sea, at pH values around 0 in coal deposits and geothermal sulfur-rich springs, or at pH values greater than 11 in sewage sludge. For example, several esterases and lipases cloned and expressed from extremophilic organisms show high activity throughout a wide range of temperatures and pHs.

[0511] Polynucleotides selected and isolated as hereinabove described are introduced into a suitable host cell. A suitable host cell is any cell which is capable of promoting recombination and/or reductive reassortment. The selected polynucleotides are preferably already in a vector which includes appropriate control sequences. The host cell can be a higher eukaryotic cell, such as a mammalian cell, or a lower eukaryotic cell, such as a yeast cell, or preferably, the host cell can be a prokaryotic cell, such as a bacterial cell. Introduction of the construct into the host cell can be effected by calcium phosphate transfection, DEAE-Dextran mediated transfection, or electroporation (Davis *et al.*, 1986).

[0512] As representative examples of appropriate hosts, there may be mentioned: bacterial cells, such as *E. coli*, *Streptomyces*, *Salmonella typhimurium*; fungal cells, such as yeast; insect cells such as *Drosophila* S2 and *Spodoptera Sf9*; animal cells such as CHO, COS or Bowes melanoma; adenoviruses; and plant cells. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein.

[0513] With particular references to various mammalian cell culture systems that can be employed to express recombinant protein, examples of mammalian expression systems include the COS-7 lines of monkey kidney fibroblasts, described in "SV40-transformed simian cells support the replication of early SV40 mutants" (Gluzman, 1981) and other cell lines capable of expressing a compatible vector, for example, the C127, 3T3, CHO, HeLa and BHK cell lines. Mammalian expression vectors will comprise an origin of replication, a suitable promoter and enhancer and also any necessary ribosome binding sites, polyadenylation site, splice donor and acceptor sites, transcriptional termination sequences and 5' flanking nontranscribed sequences. DNA sequences derived from the SV40 splice and polyadenylation sites may be used to provide the required nontranscribed genetic elements.

[0514] Methods can be used to generate novel polynucleotides encoding biochemical pathways from one or more operons or gene clusters or portions thereof. For example, bacteria and many eukaryotes have a coordinated mechanism for regulating genes whose products are involved in related processes. The genes are clustered, in structures referred to as "gene clusters," on a single chromosome and are transcribed together under the control of a single regulatory sequence, including a single promoter which initiates transcription of the entire cluster. Thus, a gene cluster is a group of adjacent genes that are either identical or related, usually as to their function. An example of a biochemical pathway encoded by gene clusters are polyketides.

[0515] Gene cluster DNA can be isolated from different organisms and ligated into vectors, particularly vectors containing expression regulatory sequences which can control and regulate the production of a detectable protein or protein-related array activity from the ligated gene clusters. Use of vectors which have an exceptionally large capacity for exogenous DNA introduction are particularly appropriate for use with such gene clusters and are described by way of example herein to include the f-factor (or fertility factor) of *E. coli*. This f-factor of *E. coli* is a plasmid which affects high-frequency transfer of itself during conjugation and is ideal to achieve and stably propagate large DNA fragments, such as gene clusters from mixed microbial samples. Cloning vectors, referred to as "fosmids" or bacterial artificial chromosome (BAC) vectors may be used. These are derived from *E. coli* f-factor which is able to stably integrate large segments of genomic DNA. When integrated with DNA from a mixed uncultured environmental sample, this makes it possible to achieve large genomic fragments in the form of a stable "environmental DNA library." Another type of vector for use is a cosmid vector. Cosmid vectors were originally designed to clone and propagate large segments of genomic DNA. Cloning into cosmid vectors is described in detail in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory Press (1989). Once ligated into an appropriate vector, two or more vectors containing different polyketide synthase gene clusters can be introduced into a suitable host cell. Regions of partial sequence homology shared by the gene clusters will promote processes which result in sequence reorganization resulting in a hybrid gene cluster. The novel hybrid gene cluster can then be screened for enhanced activities not found in the original gene clusters.

[0516] A method produces a biologically active hybrid polypeptide and screens such a polypeptide for enhanced activity by:

1. 1) introducing at least a first polynucleotide in operable linkage and a second polynucleotide in operable linkage, the at least first polynucleotide and second polynucleotide sharing at least one region of partial sequence homology, into a suitable host cell;
2. 2) growing the host cell under conditions which promote sequence reorganization resulting in a hybrid polynucleotide in operable linkage;
3. 3) expressing a hybrid polypeptide encoded by the hybrid polynucleotide;
4. 4) screening the hybrid polypeptide under conditions which promote identification of enhanced biological activity; and
5. 5) isolating the a polynucleotide encoding the hybrid polypeptide.

[0517] Methods for screening for various enzyme activities are known to those of skill in the art and are discussed throughout the present specification. Such methods may be employed when isolating the polypeptides and polynucleotides of the invention.

Screening Methodologies and "On-line" Monitoring Devices

[0518] In practicing the methods disclosed herein, a variety of apparatus and methodologies can be used to in conjunction with the polypeptides and nucleic acids of the invention, e.g., to screen polypeptides for xylanase activity (e.g., assays such as hydrolysis of casein in zymograms, the release of fluorescence from gelatin, or the release of p-nitroanalide from various small peptide substrates), to screen compounds as potential modulators, e.g., activators or inhibitors, of a xylanase activity, for antibodies that bind to a polypeptide of the invention, for nucleic acids that hybridize to a nucleic acid of the invention, to screen for cells expressing a polypeptide of the invention and the like. In addition to the array formats described in detail below for screening samples, alternative formats can also be used. Such formats include, for example, mass spectrometers, chromatographs, e.g., high-throughput HPLC and other forms of liquid chromatography, and smaller formats, such as 1536-well plates, 384-well plates and so on. High throughput screening apparatus can be adapted and used to practice the methods, see, e.g., U.S. Patent Application No. 20020001809.

Capillary Arrays

[0519] Nucleic acids or polypeptides of the invention can be immobilized to or applied to an array. Arrays can be used to screen for or monitor libraries of compositions (e.g., small molecules, antibodies, nucleic acids, etc.) for their ability to bind to or modulate the activity of a nucleic acid or a polypeptide of the invention. Capillary arrays, such as the GIGAMATRIX™, Diversa Corporation, San Diego, CA; and arrays described in, e.g., U.S. Patent Application No. 20020080350 A1; WO 0231203 A; WO 0244336 A, provide an alternative apparatus for holding and screening samples. The capillary array may include a plurality of capillaries formed into an array of adjacent capillaries, wherein each capillary comprises at least one wall defining a lumen for retaining a sample. The lumen may be cylindrical, square, hexagonal or any other geometric shape so long as the walls form a lumen for retention of a liquid or sample. The capillaries of the capillary array can be held together in close proximity to form a planar structure. The capillaries can be bound together, by being fused (e.g., where the capillaries are made of glass), glued, bonded, or clamped side-by-side. Additionally, the capillary array can include interstitial material disposed between adjacent capillaries in the array, thereby forming a solid planar device containing a plurality of through-holes.

[0520] A capillary array can be formed of any number of individual capillaries, for example, a range from 100 to 4,000,000 capillaries. Further, a capillary array having about 100,000 or more individual capillaries can be formed into the standard size and shape of a Microtiter® plate for fitment into standard laboratory equipment. The lumens are filled manually or automatically using either capillary action or microinjection using a thin needle. Samples of interest may subsequently be removed from individual capillaries for further analysis or characterization. For example, a thin, needle-like probe is positioned in fluid communication with a selected capillary to either add or withdraw material from the lumen.

[0521] In a single-pot screening assay, the assay components are mixed yielding a solution of interest, prior to insertion into the capillary array. The lumen is filled by capillary action when at least a portion of the array is immersed into a solution of interest. Chemical or biological reactions and/or activity in each capillary are monitored for detectable events. A detectable event is often referred to as a "hit", which can usually be distinguished from "non-hit" producing capillaries by optical detection. Thus, capillary arrays allow for massively parallel detection of "hits".

[0522] In a multi-pot screening assay, a polypeptide or nucleic acid, e.g., a ligand, can be introduced into a first component, which is introduced into at least a portion of a capillary of a capillary array. An air bubble

can then be introduced into the capillary behind the first component. A second component can then be introduced into the capillary, wherein the second component is separated from the first component by the air bubble. The first and second components can then be mixed by applying hydrostatic pressure to both sides of the capillary array to collapse the bubble. The capillary array is then monitored for a detectable event resulting from reaction or non-reaction of the two components.

[0523] In a binding screening assay, a sample of interest can be introduced as a first liquid labeled with a detectable particle into a capillary of a capillary array, wherein the lumen of the capillary is coated with a binding material for binding the detectable particle to the lumen. The first liquid may then be removed from the capillary tube, wherein the bound detectable particle is maintained within the capillary, and a second liquid may be introduced into the capillary tube. The capillary is then monitored for a detectable event resulting from reaction or non-reaction of the particle with the second liquid.

Arrays, or "Biochips"

[0524] Nucleic acids and/or polypeptides of the invention can be immobilized to or applied to an array, e.g., a "biochip". Arrays can be used to screen for or monitor libraries of compositions (e.g., small molecules, antibodies, nucleic acids, etc.) for their ability to bind to or modulate the activity of a nucleic acid or a polypeptide of the invention. For example, in one aspect of the invention, a monitored parameter is transcript expression of a xylanase gene. One or more, or, all the transcripts of a cell can be measured by hybridization of a sample comprising transcripts of the cell, or, nucleic acids representative of or complementary to transcripts of a cell, by hybridization to immobilized nucleic acids on an array, or "biochip." By using an "array" of nucleic acids on a microchip, some or all of the transcripts of a cell can be simultaneously quantified. Alternatively, arrays comprising genomic nucleic acid can also be used to determine the genotype of a newly engineered strain. Polypeptide arrays can also be used to simultaneously quantify a plurality of proteins. Any known "array," also referred to as a "microarray" or "nucleic acid array" or "polypeptide array" or "antibody array" or "biochip," or variation thereof may be used. Arrays are generically a plurality of "spots" or "target elements," each target element comprising a defined amount of one or more biological molecules, e.g., oligonucleotides, immobilized onto a defined area of a substrate surface for specific binding to a sample molecule, e.g., mRNA transcripts.

[0525] The terms "array" or "microarray" or "biochip" or "chip" as used herein is a plurality of target elements, each target element comprising a defined amount of one or more polypeptides (including antibodies) or nucleic acids immobilized onto a defined area of a substrate surface, as discussed in further detail, below.

[0526] Any known array and/or method of making and using arrays can be used in whole or in part, or variations thereof, as described, for example, in U.S. Patent Nos. 6,277,628; 6,277,489; 6,261,776; 6,258,606; 6,054,270; 6,048,695; 6,045,996; 6,022,963; 6,013,440; 5,965,452; 5,959,098; 5,856,174; 5,830,645; 5,770,456; 5,632,957; 5,556,752; 5,143,854; 5,807,522; 5,800,992; 5,744,305; 5,700,637; 5,556,752; 5,434,049; see also, e.g., WO 99/51773; WO 99/09217; WO 97/46313; WO 96/17958; see also, e.g., Johnston (1998) Curr. Biol. 8:R171-R174; Schummer (1997) Biotechniques 23:1087-1092; Kern (1997) Biotechniques 23:120-124; Solinas-Toldo (1997) Genes, Chromosomes & Cancer 20:399-407; Bowtell (1999) Nature Genetics Supp. 21:25-32. See also published U.S. patent applications Nos. 20010018642; 20010019827; 20010016322; 20010014449; 20010014448; 20010012537; 20010008765.

Antibodies and Antibody-based screening methods

[0527] The invention provides isolated, synthetic or recombinant antibodies that specifically bind to a xylanase of the invention. These antibodies can be used to isolate, identify or quantify the xylanases of the invention or related polypeptides. These antibodies can be used to isolate other polypeptides within the scope the invention or other related xylanases. The antibodies can be designed to bind to an active site of a xylanase.

[0528] The invention provides fragments of the enzymes of the invention, including immunogenic fragments of a polypeptide of the invention. The invention provides compositions comprising a polypeptide or peptide of the invention and adjuvants or carriers and the like.

[0529] The antibodies can be used in immunoprecipitation, staining, immunoaffinity columns, and the like. If desired, nucleic acid sequences encoding for specific antigens can be generated by immunization followed by isolation of polypeptide or nucleic acid, amplification or cloning and immobilization of polypeptide onto an array. Alternatively, the structure of an antibody can be modified, e.g., an antibody's affinity can be increased or decreased. Furthermore, the ability to make or modify antibodies can be a phenotype engineered into a cell as disclosed herein.

[0530] The term "antibody" includes a peptide or polypeptide derived from, modeled after or substantially encoded by an immunoglobulin gene or immunoglobulin genes, or fragments thereof, capable of specifically binding an antigen or epitope, see, e.g. Fundamental Immunology, Third Edition, W.E. Paul, ed., Raven Press, N.Y. (1993); Wilson (1994) J. Immunol. Methods 175:267-273; Yarmush (1992) J. Biochem. Biophys. Methods 25:85-97. The term antibody includes antigen-binding portions, i.e., "antigen binding sites," (e.g., fragments, subsequences, complementarity determining regions (CDRs)) that retain capacity to bind antigen, including (i) a Fab fragment, a monovalent fragment consisting of the VL, VH, CL and CH1 domains; (ii) a F(ab')₂ fragment, a bivalent fragment comprising two Fab fragments linked by a disulfide bridge at the hinge region; (iii) a Fd fragment consisting of the VH and CH1 domains; (iv) a Fv fragment consisting of the VL and VH domains of a single arm of an antibody, (v) a dAb fragment (Ward et al., (1989) Nature 341:544-546), which consists of a VH domain; and (vi) an isolated complementarity determining region (CDR). Single chain antibodies are also included by reference in the term "antibody."

[0531] Methods of immunization, producing and isolating antibodies (polyclonal and monoclonal) are known to those of skill in the art and described in the scientific and patent literature, see, e.g., Coligan, CURRENT PROTOCOLS IN IMMUNOLOGY, Wiley/Greene, NY (1991); Stites (eds.) BASIC AND CLINICAL IMMUNOLOGY (7th ed.) Lange Medical Publications, Los Altos, CA ("Stites"); Goding, MONOCLONAL ANTIBODIES: PRINCIPLES AND PRACTICE (2d ed.) Academic Press, New York, NY (1986); Kohler (1975) Nature 256:495; Harlow (1988) ANTIBODIES, A LABORATORY MANUAL, Cold Spring Harbor Publications, New York. Antibodies also can be generated *in vitro*, e.g., using recombinant antibody binding site expressing phage display libraries, in addition to the traditional *in vivo* methods using animals. See, e.g., Hoogenboom (1997) Trends Biotechnol. 15:62-70; Katz (1997) Annu. Rev. Biophys. Biomol. Struct. 26:27-45.

[0532] The polypeptides of the invention or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof, may also be used to generate antibodies which bind specifically to the polypeptides or fragments. The resulting antibodies may be used in immunoaffinity chromatography procedures to isolate or purify the polypeptide or to determine whether the polypeptide is present in a biological sample. In such procedures, a protein preparation, such as an extract, or a biological sample is contacted with an antibody capable of specifically binding to one of the polypeptides of the invention, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof.

[0533] In immunoaffinity procedures, the antibody is attached to a solid support, such as a bead or other column matrix. The protein preparation is placed in contact with the antibody under conditions in which the antibody specifically binds to one of the polypeptides of the invention, or fragment thereof. After a wash to remove non-specifically bound proteins, the specifically bound polypeptides are eluted.

[0534] The ability of proteins in a biological sample to bind to the antibody may be determined using any of a variety of procedures familiar to those skilled in the art. For example, binding may be determined by labeling the antibody with a detectable label such as a fluorescent agent, an enzymatic label, or a radioisotope. Alternatively, binding of the antibody to the sample may be detected using a secondary antibody having such a detectable label thereon. Particular assays include ELISA assays, sandwich assays, radioimmunoassays and Western Blots.

[0535] Polyclonal antibodies generated against the polypeptides of the invention, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 or more consecutive amino acids thereof can be obtained by direct injection of the polypeptides into an animal or by administering the polypeptides to an animal, for example, a nonhuman. The antibody so obtained will then bind the polypeptide itself. In this manner, even a sequence encoding only a fragment of the polypeptide can be used to generate antibodies which may bind to the whole native polypeptide. Such antibodies can then be used to isolate the polypeptide from cells expressing that polypeptide.

[0536] For preparation of monoclonal antibodies, any technique which provides antibodies produced by continuous cell line cultures can be used. Examples include the hybridoma technique (Kohler and Milstein, *Nature*, 256:495-497, 1975), the trioma technique, the human B-cell hybridoma technique (Kozbor et al., *Immunology Today* 4:72, 1983) and the EBV-hybridoma technique (Cole, et al., 1985, in *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96).

[0537] Techniques described for the production of single chain antibodies (U.S. Patent No. 4,946,778) can be adapted to produce single chain antibodies to the polypeptides of The invention, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof. Alternatively, transgenic mice may be used to express humanized antibodies to these polypeptides or fragments thereof.

[0538] Antibodies generated against the polypeptides of the invention, or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof may be used in screening for similar polypeptides from other organisms and samples. In such techniques, polypeptides from the organism are contacted with the antibody and those polypeptides which specifically bind the antibody are detected. Any of the procedures described above may be used to detect antibody binding. One such screening assay is described in "Methods for Measuring Cellulase Activities", *Methods in Enzymology*, Vol 160, pp. 87-116.

Kits

[0539] Kits may comprise the compositions, e.g., nucleic acids, expression cassettes, vectors, cells, transgenic seeds or plants or plant parts, polypeptides (e.g., xylanases) and/or antibodies of the invention. The kits also can contain instructional material teaching the methodologies and industrial, research, medical, pharmaceutical, food and feed and food and feed supplement processing and other applications and processes as described herein.

Whole cell engineering and measuring metabolic parameters

[0540] The methods of the disclosure provide whole cell evolution, or whole cell engineering, of a cell to develop a new cell strain having a new phenotype, e.g., a new or modified xylanase activity, by modifying the genetic composition of the cell. The genetic composition can be modified by addition to the cell of a nucleic acid of the invention, e.g., a coding sequence for an enzyme of the invention. See, e.g., WO0229032; WO0196551.

[0541] To detect the new phenotype, at least one metabolic parameter of a modified cell is monitored in the cell in a "real time" or "on-line" time frame. A plurality of cells, such as a cell culture, may be monitored in "real time" or "on-line." A plurality of metabolic parameters may be monitored in "real time" or "on-line." Metabolic parameters can be monitored using the xylanases of the invention.

[0542] Metabolic flux analysis (MFA) is based on a known biochemistry framework. A linearly independent metabolic matrix is constructed based on the law of mass conservation and on the pseudo-steady state hypothesis (PSSH) on the intracellular metabolites. In practicing these methods, metabolic networks are established, including the:

- identity of all pathway substrates, products and intermediary metabolites
- identity of all the chemical reactions interconverting the pathway metabolites, the stoichiometry of the pathway reactions,
- identity of all the enzymes catalyzing the reactions, the enzyme reaction kinetics,
- the regulatory interactions between pathway components, e.g. allosteric interactions, enzyme-enzyme interactions etc,
- intracellular compartmentalization of enzymes or any other supramolecular organization of the enzymes, and,
- the presence of any concentration gradients of metabolites, enzymes or effector molecules or diffusion barriers to their movement.

[0543] Once the metabolic network for a given strain is built, mathematic presentation by matrix notion can be introduced to estimate the intracellular metabolic fluxes if the on-line metabolome data is available. Metabolic phenotype relies on the changes of the whole metabolic network within a cell. Metabolic phenotype relies on the change of pathway utilization with respect to environmental conditions, genetic regulation, developmental state and the genotype, etc. After the on-line MFA calculation, the dynamic behavior of the cells, their phenotype and other properties may be analyzed by investigating the pathway utilization. For example, if the glucose supply is increased and the oxygen decreased during the yeast fermentation, the utilization of respiratory pathways will be reduced and/or stopped, and the utilization of the fermentative pathways will dominate. Control of physiological state of cell cultures will become possible after the pathway analysis. The methods can help determine how to manipulate the fermentation by determining how to change the substrate supply, temperature, use of inducers, etc. to control the physiological state of cells to move along desirable direction. In practicing the methods, the MFA results can also be compared with transcriptome and proteome data to design experiments and protocols for metabolic engineering or gene shuffling, etc.

[0544] In practicing the methods, any modified or new phenotype can be conferred and detected, including new or improved characteristics in the cell. Any aspect of metabolism or growth can be

monitored.

Monitoring expression of an mRNA transcript

[0545] The engineered phenotype may comprise increasing or decreasing the expression of an mRNA transcript (e.g., a xylanase message) or generating new (e.g., xylanase) transcripts in a cell. This increased or decreased expression can be traced by testing for the presence of a xylanase of the invention or by xylanase activity assays. mRNA transcripts, or messages, also can be detected and quantified by any method known in the art, including, e.g., Northern blots, quantitative amplification reactions, hybridization to arrays, and the like. Quantitative amplification reactions include, e.g., quantitative PCR, including, e.g., quantitative reverse transcription polymerase chain reaction, or RT-PCR; quantitative real time RT-PCR, or "real-time kinetic RT-PCR" (see, e.g., Kreuzer (2001) Br. J. Haematol. 114:313-318; Xia (2001) Transplantation 72:907-914).

[0546] The engineered phenotype may be generated by knocking out expression of a homologous gene. The gene's coding sequence or one or more transcriptional control elements can be knocked out, e.g., promoters or enhancers. Thus, the expression of a transcript can be completely ablated or only decreased.

[0547] The engineered phenotype may comprise increasing the expression of a homologous gene. This can be effected by knocking out of a negative control element, including a transcriptional regulatory element acting in cis- or trans- , or, mutagenizing a positive control element. One or more, or, all the transcripts of a cell can be measured by hybridization of a sample comprising transcripts of the cell, or, nucleic acids representative of or complementary to transcripts of a cell, by hybridization to immobilized nucleic acids on an array.

Monitoring expression of a polypeptides, peptides and amino acids

[0548] The engineered phenotype may comprise increasing or decreasing the expression of a polypeptide (xylanase) or generating new polypeptides in a cell. This increased or decreased expression can be traced by determining the amount of xylanase present or by xylanase activity assays. Polypeptides, peptides and amino acids also can be detected and quantified by any method known in the art, including, e.g., nuclear magnetic resonance (NMR), spectrophotometry, radiography (protein radiolabeling), electrophoresis, capillary electrophoresis, high performance liquid chromatography (HPLC), thin layer chromatography (TLC), hyperdiffusion chromatography, various immunological methods, e.g. immunoprecipitation, immunodiffusion, immuno-electrophoresis, radioimmunoassays (RIAs), enzyme-linked immunosorbent assays (ELISAs), immuno-fluorescent assays, gel electrophoresis (e.g., SDS-PAGE), staining with antibodies, fluorescent activated cell sorter (FACS), pyrolysis mass spectrometry, Fourier-Transform Infrared Spectrometry, Raman spectrometry, GC-MS, and LC-Electrospray and cap-LC-tandem-electrospray mass spectrometries, and the like. Novel bioactivities can also be screened using methods, or variations thereof, described in U.S. Patent No. 6,057,103. Furthermore, as discussed below in detail, one or more, or, all the polypeptides of a cell can be measured using a protein array.

Industrial, Energy, Pharmaceutical, Medical, Food Processing and other Applications

[0549] Polypeptides of the invention can be used in any industrial, agricultural, food and feed and food and

feed supplement processing, pharmaceutical, medical, research (laboratory) or other process. The invention provides industrial processes using enzymes of the invention, e.g., in the pharmaceutical or nutrient (diet) supplement industry, the energy industry (e.g., to make "clean" biofuels), in the food and feed industries, e.g., in methods for making food and feed products and food and feed additives. Processes using enzymes of the invention in the medical industry are, e.g., to make pharmaceuticals or dietary aids or supplements, or food supplements and additives. In addition, the enzymes of the invention may be used in biofuel production, including, e.g., a bioalcohol such as bioethanol, biomethanol, biobutanol or biopropanol, thus comprising a "clean" fuel production. Enzymes of the invention can be added to industrial processes continuously, in batches or by fed-batch methods. In another aspect, enzymes of the invention can be recycled in the industrial processes, thereby lowering enzyme requirements.

[0550] For example, xylanases can be used in the biobleaching and treatment of chemical pulps, for example, as described in U.S. Pat. No. 5,202,249; or for biobleaching and treatment of wood or paper pulps, for example, as described in U.S. Pat. Nos. 5,179,021, 5,116,746, 5,407,827, 5,405,769, 5,395,765, 5,369,024, 5,457,045, 5,434,071, 5,498,534, 5,591,304, 5,645,686, 5,725,732, 5,759,840, 5,834,301, 5,871,730 and 6,057,438; or for reducing lignin in wood and modifying wood, for example, as described in U.S. Pat. Nos. 5,486,468 and/or 5,770,012; or for use as flour, dough and bread improvers, for example, as described in U.S. Pat. Nos. 5,108,765 and/or 5,306,633; or for use as feed additives and/or supplements, for example, as described in U.S. Pat. Nos. 5,432,074; 5,429,828; 5,612,055; 5,720,971; 5,981,233; 5,948,667; 6,099,844; 6,132,727 and/or 6,132,716; or in manufacturing cellulose solutions, for example, as described in U.S. Pat. No. 5,760,211; or in detergent compositions; or used for fruit, vegetables and/or mud and clay compounds, for example, as described in U.S. Pat. No. 5,786,316. Xylanases of this invention also can be used in hydrolysis of hemicellulose, for example, as described in U.S. Pat. No. 4,725,544.

[0551] The xylanase enzymes of the invention can be highly selective catalysts. They can catalyze reactions with exquisite stereo-, regio- and chemo- selectivities that are unparalleled in conventional synthetic chemistry. Moreover, enzymes are remarkably versatile. The xylanase enzymes of the invention can be tailored to function in organic solvents, operate at extreme pHs (for example, high pHs and low pHs) extreme temperatures (for example, high temperatures and low temperatures), extreme salinity levels (for example, high salinity and low salinity) and catalyze reactions with compounds that are structurally unrelated to their natural, physiological substrates.

Wood, paper and pulp treatments

[0552] The xylanases of the invention can be used in any wood, wood product, wood waste or by-product, paper, paper product, paper or wood pulp, Kraft pulp, or wood or paper recycling treatment or industrial process, e.g., any wood, wood pulp, paper waste, paper or pulp treatment or wood or paper deinking process. In one aspect, xylanases of the invention can be used to treat/ pretreat paper pulp, or recycled paper or paper pulp, and the like. In one aspect, enzyme(s) of the invention are used to increase the "brightness" of the paper via their use in treating/ pretreating paper pulp, or recycled paper or paper pulp, and the like. The higher the grade of paper, the greater the brightness; paper brightness can impact the scan capability of optical scanning equipment; thus, the enzymes and processes of the invention can be used to make high grade, "bright" paper for, e.g., use in optical scanning equipment, including inkjet, laser and photo printing quality paper.

[0553] For example, the enzymes of the invention can be used in any industrial process using xylanases

known in the art, e.g., treating waste paper, as described in, e.g., USPN 6,767,728 or 6,426,200; seasoning wood, e.g., for applications in the food industry, as described in, e.g., USPN 6,623,953; for the production of xylose from a paper-grade hardwood pulp, as described in, e.g., USPN 6,512,110; treating fibrous lignocellulosic raw material with a xylanase in an aqueous medium as described in, e.g., USPN 6,287,708; dissolving pulp from cellulosic fiber, as described in, e.g., USPN 6,254,722; deinking and decolorizing a printed paper or removing color from wood pulp, as described in, e.g., USPN 6,241,849, 5,834,301 or 5,582,681; bleaching a chemical paper pulp or lignocellulose pulp using a xylanase, as described in, e.g., USPN 5,645,686 or 5,618,386; for treating wood pulp that includes incompletely washed brownstock, as described in, e.g., USPN 5,591,304; purifying and delignifying a waste lignocellulosic material, as described in, e.g., in USPN 5,503,709; manufacturing paper or cardboard from recycled cellulose fibers, as described in, e.g., in USPN 5,110,412; debarking of logs, as described in, e.g., in USPN 5,103,883; producing fluff pulp with improved shredding properties, as described in, e.g., in USPN 5,068,009, and the like. The xylanases of the invention can be used to process or treat any cellulosic material, e.g., fibers from wood, cotton, hemp, flax or linen.

[0554] Wood, wood pulp, paper, paper pulp, paper waste or wood or paper recycling treatment processes may use a xylanase of the invention. In one aspect, the xylanase of the invention is applicable both in reduction of the need for a chemical decoloring (e.g., bleaching) agent, such as chlorine dioxide, and in high alkaline and high temperature environments. Most lignin is solubilized in the cooking stage of pulping process. The residual lignin is removed from the pulp in the bleaching process. In one aspect, xylanase bleaching of pulp (e.g., using an enzyme of the invention) is based on the partial hydrolysis of xylan, which is the main component of the hemicellulose. The enzymatic action (e.g., of an enzyme of the invention) releases hemicellulose-bound lignin and increases the extractability of lignin from the pulp in the subsequent bleaching process, e.g. using chlorine and oxygen chemicals. In one aspect, xylanases of the invention can be used to increase the final brightness of the pulp at a fixed level of bleaching chemicals. In another aspect, xylanases of the invention can be used to decrease the kappa number of the pulp.

[0555] Wood, wood pulp, paper, paper pulp, paper waste or wood or paper recycling treatment processes (methods) may use a xylanase of the invention where the treatment time (the amount of time the xylanase is in contact with the pulp, paper, wood, etc.) and/or retention time can be anywhere from between about 1 minute to 12 hours, or between about 5 minutes to 1 hour, or between about 15 to 30 minutes; or the treatment and/or retention time can be any time up to about 0.1, 0.25, 0.5, 0.75, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 or more hours.

[0556] In one aspect, the xylanase of the invention is a thermostable alkaline endoxylanase which in one aspect can effect a greater than 25% reduction in the chlorine dioxide requirement of kraft pulp with a less than 0.5% pulp yield loss. In one aspect, boundary parameters are pH 10, 65-85°C and treatment time of less than 60 minutes at an enzyme loading of less than 0.001 wt%; in alternative aspects the treatment and/or retention time is less than about 0.1, 0.25, 0.5, 0.75, 1, 1.5, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11 or 12.

[0557] A pool of xylanases may be tested for the ability to hydrolyze dye-labeled xylan at, for example, pH 10 and 60°C. The enzymes that test positive under these conditions may then be evaluated at, for example pH 10 and 70°C. Alternatively, enzymes may be tested at pH 8 and pH 10 at 70°C. In discovery of xylanases desirable in the pulp and paper industry libraries from high temperature or highly alkaline environments were targeted. Specifically, these libraries were screened for enzymes functioning at alkaline pH and a temperature of approximately 45°C. In another aspect, the xylanases of the invention are useful in the pulp and paper industry in degradation of a lignin-hemicellulose linkage, in order to release the lignin.

[0558] Enzymes of the invention can be used for deinking printed wastepaper, such as newspaper, or for

deinking noncontact-printed wastepaper, e.g., xerographic and laser-printed paper, and mixtures of contact and noncontact-printed wastepaper, as described in USPN 6,767,728 or 6,426,200; Neo (1986) J. Wood Chem. Tech. 6(2):147. Enzymes of the invention can be used in processes for the production of xylose from a paper-grade hardwood pulp by extracting xylan contained in pulp into a liquid phase, subjecting the xylan contained in the obtained liquid phase to conditions sufficient to hydrolyze xylan to xylose, and recovering the xylose, where the extracting step includes at least one treatment of an aqueous suspension of pulp or an alkali-soluble material a xylanase enzyme, as described in, e.g., USPN 6,512,110. Enzymes of the invention can be used in processes for dissolving pulp from cellulosic fibers such as recycled paper products made from hardwood fiber, a mixture of hardwood fiber and softwood fiber, waste paper, e.g., from unprinted envelopes, de-inked envelopes, unprinted ledger paper, de-inked ledger paper, and the like, as described in, e.g., USPN 6,254,722.

[0559] The xylanases of the invention can also be used in any wood, wood product, paper, paper product, paper or wood pulp, Kraft pulp, or wood or paper recycling treatment or industrial process, e.g., any wood, wood pulp, paper waste, paper or pulp treatment or wood or paper deinking process as an antimicrobial or microbial repellent. Alternatively, the xylanases of the invention can be part of a wood, wood product, paper, paper product, paper or wood pulp, Kraft pulp, or recycled paper composition, and/or a composition comprising one or more wood, wood product, paper, paper product, paper or wood pulp, Kraft pulp, or recycled paper compositions, wherein the xylanases of the invention act as an antimicrobial or microbial repellent in the composition.

Treating fibers and textiles

[0560] The xylanases of the invention can be used in any fiber- or fabric-treating method, which are well known in the art, see, e.g., U.S. Patent No. 6,261,828; 6,077,316; 6,024,766; 6,021,536; 6,017,751; 5,980,581; US Patent Publication No. 20020142438 A1. For example, xylanases of the invention can be used in fiber and/or fabric desizing. In one aspect, the feel and appearance of a fabric is improved by a method comprising contacting the fabric with a xylanase of the invention in a solution. In one aspect, the fabric is treated with the solution under pressure. For example, xylanases of the invention can be used in the removal of stains.

[0561] The xylanases of the invention can be used to treat any cellulosic material, including fibers (e.g., fibers from cotton, hemp, flax or linen), sewn and unsewn fabrics, e.g., knits, wovens, denims, yarns, and toweling, made from cotton, cotton blends or natural or manmade cellulose (e.g. originating from xylan-containing cellulose fibers such as from wood pulp) or blends thereof. Examples of blends are blends of cotton or rayon/viscose with one or more companion material such as wool, synthetic fibers (e.g. polyamide fibers, acrylic fibers, polyester fibers, polyvinyl alcohol fibers, polyvinyl chloride fibers, polyvinylidene chloride fibers, polyurethane fibers, polyurea fibers, aramid fibers), and cellulose-containing fibers (e.g. rayon/viscose, ramie, hemp, flax/linen, jute, cellulose acetate fibers, lyocell).

[0562] The textile treating processes using xylanases of the invention can be used in conjunction with other textile treatments, e.g., scouring and bleaching. Scouring is the removal of non-cellulosic material from the cotton fiber, e.g., the cuticle (mainly consisting of waxes) and primary cell wall (mainly consisting of pectin, protein and xyloglucan). A proper wax removal is necessary for obtaining a high wettability. This is needed for dyeing. Removal of the primary cell walls by the processes of the invention improves wax removal and ensures a more even dyeing. Treating textiles can improve whiteness in the bleaching process. The main chemical used in scouring is sodium hydroxide in high concentrations and at high temperatures. Bleaching comprises oxidizing the textile. Bleaching typically involves use of hydrogen

peroxide as the oxidizing agent in order to obtain either a fully bleached (white) fabric or to ensure a clean shade of the dye.

[0563] The invention also provides alkaline xylanases (xylanases active under alkaline conditions). These have wide-ranging applications in textile processing, degumming of plant fibers (e.g., plant bast fibers), treatment of pectic wastewaters, paper-making, and coffee and tea fermentations. See, e.g., Hoondal (2002) Applied Microbiology and Biotechnology 59:409-418.

[0564] In another aspect of the invention, the xylanases of the invention can also be used in any fiber- and/or fabric-treating process as an antimicrobial or microbial repellent. Alternatively, the xylanases of the invention can be part of a fiber- and/or fabric-composition, where the xylanases of the invention act as an antimicrobial or microbial repellent in the fiber and/or fabric.

Detergent, Disinfectant and Cleaning Compositions

[0565] Detergent, disinfectant or cleanser (cleaning or cleansing) compositions may comprise one or more xylanases of the invention. For methods of making and using detergent, disinfectant or cleanser compositions, see, e.g., U.S. Patent No. 6,413,928; 6,399,561; 6,365,561; 6,380,147. The detergent, disinfectant or cleanser compositions can be a one and two part aqueous composition, a non-aqueous liquid composition, a cast solid, a granular form, a particulate form, a compressed tablet, a gel and/or a paste and a slurry form. The xylanases of the invention can also be used as a detergent, disinfectant or cleanser additive product in a solid or a liquid form. Such additive products are intended to supplement or boost the performance of conventional detergent compositions and can be added at any stage of the cleaning process.

[0566] The actual active enzyme content depends upon the method of manufacture of a detergent, disinfectant or cleanser composition and is not critical, assuming the detergent solution has the desired enzymatic activity. In one aspect, the amount of xylanase present in the final solution ranges from about 0.001 mg to 0.5 mg per gram of the detergent composition. The particular enzyme chosen for use in the process and products of this invention depends upon the conditions of final utility, including the physical product form, use pH, use temperature, and soil types to be degraded or altered. The enzyme can be chosen to provide optimum activity and stability for any given set of utility conditions. In one aspect, the xylanases of the present invention are active in the pH ranges of from about 4 to about 12 and in the temperature range of from about 20°C to about 95°C. The detergents of the invention can comprise cationic, semi-polar nonionic or zwitterionic surfactants; or, mixtures thereof.

[0567] Xylanases of the invention can be formulated into powdered and liquid detergents, disinfectants or cleansers having pH between 4.0 and 12.0 at levels of about 0.01 to about 5% (preferably 0.1% to 0.5%) by weight. These detergent, disinfectant or cleanser compositions can also include other enzymes such as xylanases, cellulases, lipases, esterases, proteases, or endoglycosidases, endo-beta.-1,4-glucanases, beta-glucanases, endo-beta-1,3(4)-glucanases, cutinases, peroxidases, catalases, laccases, amylases, glucoamylases, pectinases, reductases, oxidases, phenoloxidases, ligninases, pullulanases, arabinanases, hemicellulases, mannanases, xyloglucanases, xylanases, pectin acetyl esterases, rhamnogalacturonan acetyl esterases, polygalacturonases, rhamnogalacturonases, galactanases, pectin lyases, pectin methylesterases, cellobiohydrolases and/or transglutaminases. These detergent, disinfectant or cleanser compositions can also include dyes, colorants, odorants, bleaches, buffers, builders, enzyme "enhancing agents" (see, e.g., U.S. Patent application no. 20030096394) and stabilizers.

[0568] The addition of xylanases of the invention to conventional cleaning compositions does not create any special use limitation. In other words, any temperature and pH suitable for the detergent is also suitable as long as the enzyme is active at or tolerant of the pH and/or temperature of the intended use. In addition, the xylanases of the invention can be used in a cleaning composition without detergents, again either alone or in combination with builders and stabilizers.

[0569] Relevant cleaning compositions include detergent compositions for cleaning hard surfaces, detergent compositions for cleaning fabrics, dishwashing compositions, oral cleaning compositions, denture cleaning compositions, and contact lens cleaning solutions.

[0570] A method for washing an object comprises contacting the object with a polypeptide of the invention under conditions sufficient for washing. A xylanase of the invention may be included as a detergent, disinfectant or cleanser additive. The detergent, disinfectant or cleanser composition may, for example, be formulated as a hand or machine laundry detergent, disinfectant or cleanser composition comprising a polypeptide of the invention. A laundry additive suitable for pre-treatment of stained fabrics can comprise a polypeptide of the invention. A fabric softener composition can comprise a xylanase of the invention. Alternatively, a xylanase of the invention can be formulated as a detergent, disinfectant or cleanser composition for use in general household hard surface cleaning operations. In alternative aspects, detergent, disinfectant or cleanser additives and detergent, disinfectant or cleanser compositions may comprise one or more other enzymes such as a xylanase, a lipase, a protease, a cutinase, an esterase, another xylanase, a carbohydrase, a cellulase, a pectinase, a mannanase, an arabinase, a galactanase, a xylanase, an oxidase, e.g., a lactase, and/or a peroxidase (see also, above). The properties of the enzyme(s) of the invention are chosen to be compatible with the selected detergent (i.e. pH-optimum, compatibility with other enzymatic and non-enzymatic ingredients, etc.) and the enzyme(s) is present in effective amounts. In one aspect, xylanase enzymes of the invention are used to remove malodorous materials from fabrics. Various detergent compositions and methods for making them that can be used in practicing the invention are described in, e.g., U.S. Patent Nos. 6,333,301; 6,329,333; 6,326,341; 6,297,038; 6,309,871; 6,204,232; 6,197,070; 5,856,164.

[0571] When formulated as compositions suitable for use in a laundry machine washing method, the xylanases of the invention can comprise both a surfactant and a builder compound. They can additionally comprise one or more detergent components, e.g., organic polymeric compounds, bleaching agents, additional enzymes, suds suppressors, dispersants, lime-soap dispersants, soil suspension and anti-redeposition agents and corrosion inhibitors. Laundry compositions can also contain softening agents, as additional detergent components. Such compositions containing carbohydrase can provide fabric cleaning, stain removal, whiteness maintenance, softening, color appearance, dye transfer inhibition and sanitization when formulated as laundry detergent compositions.

[0572] The density of the laundry detergent, disinfectant or cleanser compositions can range from about 200 to 1500 g/liter, or, about 400 to 1200 g/liter, or, about 500 to 950 g/liter, or, 600 to 800 g/liter, of composition; this can be measured at about 20°C.

[0573] In one aspect, the "compact" form of laundry detergent, disinfectant or cleanser compositions is best reflected by density and, in terms of composition, by the amount of inorganic filler salt. Inorganic filler salts are conventional ingredients of detergent compositions in powder form. In conventional detergent compositions, the filler salts are present in substantial amounts, typically 17% to 35% by weight of the total composition. In one aspect of the compact compositions, the filler salt is present in amounts not exceeding 15% of the total composition, or, not exceeding 10%, or, not exceeding 5% by weight of the composition. The inorganic filler salts can be selected from the alkali and alkaline-earth-metal salts of sulphates and chlorides, e.g., sodium sulphate.

[0574] Liquid detergent compositions can also be in a "concentrated form." The liquid detergent, disinfectant or cleanser compositions can contain a lower amount of water, compared to conventional liquid detergents, disinfectants or cleansers. The water content of the concentrated liquid detergent may be less than 40%, or, less than 30%, or, less than 20% by weight of the detergent, disinfectant or cleanser composition. Detergent, disinfectant or cleanser compounds can comprise formulations as described in WO 97/01629.

[0575] Xylanases of the invention can be useful in formulating various detergent, cleaning, disinfectant or cleanser compositions. A number of known compounds are suitable surfactants including nonionic, anionic, cationic, or zwitterionic detergents, can be used, e.g., as disclosed in U.S. Patent Nos. 4,404,128; 4,261,868; 5,204,015. In addition, xylanases can be used, for example, in bar or liquid soap applications, dish care formulations, contact lens cleaning solutions or products, peptide hydrolysis, waste treatment, textile applications, as fusion-cleavage enzymes in protein production, and the like. Xylanases may provide enhanced performance in a detergent composition as compared to another detergent xylanase, that is, the enzyme group may increase cleaning of certain enzyme sensitive stains such as grass or blood, as determined by usual evaluation after a standard wash cycle. Xylanases can be formulated into known powdered and liquid detergents having pH between 6.5 and 12.0 at levels of about 0.01 to about 5% (for example, about 0.1% to 0.5%) by weight. These detergent cleaning compositions can also include other enzymes such as known xylanases, xylanases, proteases, amylases, cellulases, mannanases, lipases or endoglycosidases, redox enzymes such as catalases and laccases, as well as builders, stabilizers, fragrances and pigments.

[0576] Detergent, disinfectant or cleanser compositions having xylanase activity, comprising a xylanase of the invention may be for use with fruit, vegetables and/or mud and clay compounds (see, for example, U.S. Pat. No. 5,786,316).

[0577] In another aspect of the invention, the xylanases of the invention can also be used in any detergent, disinfectant or cleanser (cleaning solution) manufacturing process, wherein the xylanase is used as an antimicrobial or microbial repellent. Alternatively, the xylanases of the invention can be used in any cleaning or washing process, wherein the xylanase is used as an antimicrobial or microbial repellent. In another aspect of the invention, the xylanase of the invention can be included in any detergent or cleanser composition, wherein the xylanases of the invention act as an antimicrobial or microbial repellent in the composition.

Treating foods and food processing

[0578] The xylanases of the invention have numerous applications in food processing industry. For example, in one aspect, the xylanases of the invention are used to improve the extraction of oil from oil-rich plant material, e.g., oil-rich seeds, for example, soybean oil from soybeans, olive oil from olives, rapeseed oil from rapeseed and/or sunflower oil from sunflower seeds.

[0579] The xylanases of the invention can be used for separation of components of plant cell materials. For example, xylanases of the invention can be used in the separation of xylan-rich material (e.g., plant cells) into components. In one aspect, xylanases of the invention can be used to separate xylan-rich or oil-rich crops into valuable protein and oil and hull fractions. The separation process may be performed by use of methods known in the art.

[0580] The xylanases of the invention can be used in the preparation of fruit or vegetable juices, syrups, extracts and the like to increase yield. The xylanases of the invention can be used in the enzymatic treatment (e.g., hydrolysis of xylan-comprising plant materials) of various plant cell wall-derived materials or waste materials, e.g. from cereals, grains, wine or juice production, or agricultural residues such as vegetable hulls, bean hulls, sugar beet pulp, olive pulp, potato pulp, and the like. The xylanases of the invention can be used to modify the consistency and appearance of processed fruit or vegetables. The xylanases of the invention can be used to treat plant material to facilitate processing of plant material, including foods, facilitate purification or extraction of plant components. The xylanases of the invention can be used to improve feed value, decrease the water binding capacity, improve the degradability in waste water plants and/or improve the conversion of plant material to ensilage, and the like.

[0581] In one aspect, xylanases of the invention are used in baking applications, e.g., cookies and crackers, to hydrolyze xylans such as arabinoxylans. In one aspect, xylanases of the invention are used to create non-sticky doughs that are not difficult to machine and to reduce biscuit size. Xylanases of the invention can be used to hydrolyze arabinoxylans to prevent rapid rehydration of the baked product resulting in loss of crispiness and reduced shelf-life. In one aspect, xylanases of the invention are used as additives in dough processing. In one aspect, xylanases of the invention are used in dough conditioning, wherein in one aspect the xylanases possess high activity over a temperature range of about 25-35°C and at near neutral pH (7.0 - 7.5). In one aspect, dough conditioning enzymes can be inactivated at the extreme temperatures of baking (>500°F). The enzymes of the invention can be used in conjunction with any dough processing protocol, e.g., as in U.S. Patent App. No. 20050281916.

[0582] In one aspect, xylanases of the invention are used as additives in dough processing to perform optimally under dough pH and temperature conditions. In one aspect, an enzyme of the invention is used for dough conditioning. In one aspect, a xylanase of the invention possesses high activity over a temperature range of 25-35°C and at near neutral pH (7.0 - 7.5). In one aspect, the enzyme is inactivated at the extreme temperatures of baking, for example, >500°F.

[0583] In another aspect of the invention, the xylanases of the invention can also be used in any food or beverage treatment or food or beverage processing process, wherein the xylanase is used as an antimicrobial or microbial repellent. In another aspect of the invention, the xylanase of the invention can be included in any food or beverage composition, wherein the xylanases of the invention act as an antimicrobial or microbial repellent in the composition.

Animal feeds and food or feed or food additives (supplements)

[0584] Methods for treating animal feeds and foods and food or feed additives (supplements) using xylanases of the invention, animals including mammals (e.g., humans), birds, fish and the like are disclosed. Treating animal feeds, foods and additives using xylanases of the invention can help in the availability of nutrients, e.g., starch, protein, and the like, in the animal feed or additive (supplements). By breaking down difficult to digest proteins or indirectly or directly unmasking starch (or other nutrients), the xylanase makes nutrients more accessible to other endogenous or exogenous enzymes. The xylanase can also simply cause the release of readily digestible and easily absorbed nutrients and sugars.

[0585] When added to animal feed, xylanases of the invention improve the *in vivo* break-down of plant cell wall material partly due to a reduction of the intestinal viscosity (see, e.g., Bedford et al., Proceedings of the 1st Symposium on Enzymes in Animal Nutrition, 1993, pp. 73-77), whereby a better utilization of the plant nutrients by the animal is achieved. Thus, by using xylanases of the invention in feeds the growth

rate and/or feed conversion ratio (i.e. the weight of ingested feed relative to weight gain) of the animal is improved.

[0586] The animal feed additive may be a granulated enzyme product which may readily be-mixed with feed components. Alternatively, feed additives can form a component of a pre-mix. The granulated enzyme product may be coated or uncoated. The particle size of the enzyme granulates can be compatible with that of feed and pre-mix components. This provides a safe and convenient mean of incorporating enzymes into feeds. Alternatively, the animal feed additive of the invention may be a stabilized liquid composition. This may be an aqueous or oil-based slurry. See, e.g., U.S. Patent No. 6,245,546.

[0587] Xylanases of the present invention, in the modification of animal feed or a food, can process the food or feed either *in vitro* (by modifying components of the feed or food) or *in vivo*. Xylanases can be added to animal feed or food compositions containing high amounts of xylans, e.g. feed or food containing plant material from cereals, grains and the like. When added to the feed or food the xylanase significantly improves the *in vivo* break-down of xylan-containing material, e.g., plant cell walls, whereby a better utilization of the plant nutrients by the animal (e.g., human) is achieved. In one aspect, the growth rate and/or feed conversion ratio (i.e. the weight of ingested feed relative to weight gain) of the animal is improved. For example a partially or indigestible xylan-comprising protein is fully or partially degraded by a xylanase of the invention, e.g. in combination with another enzyme, e.g., beta-galactosidase, to peptides and galactose and/or galactooligomers. These enzyme digestion products are more digestible by the animal. Thus, xylanases of the invention can contribute to the available energy of the feed or food. Also, by contributing to the degradation of xylan-comprising proteins, a xylanase of the invention can improve the digestibility and uptake of carbohydrate and non-carbohydrate feed or food constituents such as protein, fat and minerals.

[0588] In another aspect, xylanase of the invention can be supplied by expressing the enzymes directly in transgenic feed crops (as, e.g., transgenic plants, seeds and the like), such as grains, cereals, corn, soy bean, rape seed, lupin and the like. As discussed above, the invention provides transgenic plants, plant parts and plant cells comprising a nucleic acid sequence encoding a polypeptide of the invention. In one aspect, the nucleic acid is expressed such that the xylanase of the invention is produced in recoverable quantities. The xylanase can be recovered from any plant or plant part. Alternatively, the plant or plant part containing the recombinant polypeptide can be used as such for improving the quality of a food or feed, e.g., improving nutritional value, palatability, and rheological properties, or to destroy an antinutritive factor.

[0589] Oligosaccharides may be removed from feed prior to consumption by an animal subject using a xylanase of the invention. In this process a feed is formed having an increased metabolizable energy value. In addition to xylanases of the invention, galactosidases, cellulases and combinations thereof can be used. In one aspect, the enzyme is added in an amount equal to between about 0.1% and 1 % by weight of the feed material. In one aspect, the feed is a cereal, a wheat, a grain, a soybean (e.g., a ground soybean) material. See, e.g., U.S. Patent No. 6,399,123.

[0590] Xylanase of the invention may be used as a nutritional supplement in the diets of animals by preparing a nutritional supplement containing a recombinant xylanase enzyme comprising at least thirty contiguous amino acids of a sequence of the invention, and administering the nutritional supplement to an animal to increase the utilization of xylan contained in food ingested by the animal.

[0591] An enzyme delivery matrix readily releases a xylanase enzyme, such as one having an amino acid sequence of the invention, or at least 30 contiguous amino acids thereof, in aqueous media, such as, for example, the digestive fluid of an animal. The enzyme delivery matrix is prepared from a granulate edible

carrier selected from such components as grain germ that is spent of oil, hay, alfalfa, timothy, soy hull, sunflower seed meal, corn meal, soy meal, wheat midd, and the like, that readily disperse the recombinant enzyme contained therein into aqueous media. In use, the edible pelletized enzyme delivery matrix is administered to an animal to deliver xylanase to the animal. Suitable grain-based substrates may comprise or be derived from any suitable edible grain, such as wheat, corn, soy, sorghum, alfalfa, barley, and the like. An exemplary grain-based substrate is a corn-based substrate. The substrate may be derived from any suitable part of the grain, but is preferably a grain germ approved for animal feed use, such as corn germ that is obtained in a wet or dry milling process. The grain germ preferably comprises spent germ, which is grain germ from which oil has been expelled, such as by pressing or hexane or other solvent extraction. Alternatively, the grain germ is expeller extracted, that is, the oil has been removed by pressing.

[0592] The enzyme delivery matrix is in the form of discrete plural particles, pellets or granules. By "granules" is meant particles that are compressed or compacted, such as by a pelletizing, extrusion, or similar compacting to remove water from the matrix. Such compression or compacting of the particles also promotes intraparticle cohesion of the particles. For example, the granules can be prepared by pelletizing the grain-based substrate in a pellet mill. The pellets prepared thereby are ground or crumbled to a granule size suitable for use as an adjuvant in animal feed. Since the matrix is itself approved for use in animal feed, it can be used as a diluent for delivery of enzymes in animal feed.

[0593] The enzyme delivery matrix can be in the form of granules having a granule size ranging from about 4 to about 400 mesh (USS); more preferably, about 8 to about 80 mesh; and most preferably about 14 to about 20 mesh. If the grain germ is spent via solvent extraction, use of a lubricity agent such as corn oil may be necessary in the pelletizer, but such a lubricity agent ordinarily is not necessary if the germ is expeller extracted. The matrix may alternatively be prepared by other compacting or compressing processes such as, for example, by extrusion of the grain-based substrate through a die and grinding of the extrudate to a suitable granule size.

[0594] The enzyme delivery matrix may further include a polysaccharide component as a cohesiveness agent to enhance the cohesiveness of the matrix granules. The cohesiveness agent is believed to provide additional hydroxyl groups, which enhance the bonding between grain proteins within the matrix granule. It is further believed that the additional hydroxyl groups so function by enhancing the hydrogen bonding of proteins to starch and to other proteins. The cohesiveness agent may be present in any amount suitable to enhance the cohesiveness of the granules of the enzyme delivery matrix. Suitable cohesiveness agents include one or more of dextrans, maltodextrins, starches, such as corn starch, flours, cellulose, hemicellulose, and the like. For example, the percentage of grain germ and cohesiveness agent in the matrix (not including the enzyme) is 78% corn germ meal and 20% by weight of corn starch.

[0595] Because the enzyme-releasing matrix is made from biodegradable materials and contains moisture, the matrix may be subject to spoilage, such as by molding. To prevent or inhibit such molding, the matrix may include a mold inhibitor, such as a propionate salt, which may be present in any amount sufficient to inhibit the molding of the enzyme-releasing matrix, thus providing a delivery matrix in a stable formulation that does not require refrigeration.

[0596] The xylanase enzyme contained in the enzyme delivery matrix is preferably a thermostable xylanase, as described herein, so as to resist inactivation of the xylanase during manufacture where elevated temperatures and/or steam may be employed to prepare the pelletized enzyme delivery matrix. During digestion of feed containing the invention enzyme delivery matrix, aqueous digestive fluids will cause release of the active enzyme. Other types of thermostable enzymes and nutritional supplements that are thermostable can also be incorporated in the delivery matrix for release under any type of

aqueous conditions.

[0597] A coating can be applied to the invention enzyme matrix particles for many different purposes, such as to add a flavor or nutrition supplement to animal feed, to delay release of animal feed supplements and enzymes in gastric conditions, and the like. Or, the coating may be applied to achieve a functional goal, for example, whenever it is desirable to slow release of the enzyme from the matrix particles or to control the conditions under which the enzyme will be released. The composition of the coating material can be such that it is selectively broken down by an agent to which it is susceptible (such as heat, acid or base, enzymes or other chemicals). Alternatively, two or more coatings susceptible to different such breakdown agents may be consecutively applied to the matrix particles.

[0598] A process for preparing an enzyme-releasing matrix may comprise providing discrete plural particles of a grain-based substrate in a particle size suitable for use as an enzyme-releasing matrix, wherein the particles comprise a xylanase enzyme encoded by an amino acid sequence of the invention or at least 30 consecutive amino acids thereof. Preferably, the process includes compacting or compressing the particles of enzyme-releasing matrix into granules, which can be accomplished by pelletizing. The mold inhibitor and cohesiveness agent, when used, can be added at any suitable time, and can be mixed with the grain-based substrate in the desired proportions prior to pelletizing of the grain-based substrate. Moisture content in the pellet mill feed can be in the ranges set forth above with respect to the moisture content in the finished product, and can be about 14-15%. In one aspect, moisture is added to the feedstock in the form of an aqueous preparation of the enzyme to bring the feedstock to this moisture content. The temperature in the pellet mill can be brought to about 82°C with steam. The pellet mill may be operated under any conditions that impart sufficient work to the feedstock to provide pellets. The pelleting process itself is a cost-effective process for removing water from the enzyme-containing composition.

[0599] In one aspect, the pellet mill is operated with a 1/8 in. by 2 inch die at 100 lb./min. pressure at 82°C to provide pellets, which then are crumbled in a pellet mill crumbler to provide discrete plural particles having a particle size capable of passing through an 8 mesh screen but being retained on a 20 mesh screen.

[0600] The thermostable xylanases of the invention can be used in the pellets. They can have high optimum temperatures and high heat resistance such that an enzyme reaction at a temperature not hitherto carried out can be achieved. The gene encoding the xylanase according to the present invention (e.g. as set forth in any of the sequences in the invention) can be used in preparation of xylanases (e.g. using GSSM as described herein) having characteristics different from those of the xylanases of the invention (in terms of optimum pH, optimum temperature, heat resistance, stability to solvents, specific activity, affinity to substrate, secretion ability, translation rate, transcription control and the like). Furthermore, a polynucleotide of the invention may be employed for screening of variant xylanases prepared by the methods described herein to determine those having a desired activity, such as improved or modified thermostability or thermotolerance. For example, U.S. Patent No. 5,830,732, describes a screening assay for determining thermotolerance of a xylanase.

[0601] The xylanases of the invention can also be used in any animal feed, animal food or feed additive production process, wherein the xylanase is used as an antimicrobial or microbial repellent. The xylanase of the invention can be included in any animal feed, animal food or feed additive composition, wherein the xylanases of the invention act as an antimicrobial or microbial repellent in the composition.

Waste treatment

[0602] The xylanases of the invention can be used in a variety of other industrial applications, e.g., in waste treatment. For example, a solid waste digestion process may use xylanases of the invention. The methods can comprise reducing the mass and volume of substantially untreated solid waste. Solid waste can be treated with an enzymatic digestive process in the presence of an enzymatic solution including xylanases of the invention at a controlled temperature. This results in a reaction without appreciable bacterial fermentation from added microorganisms. The solid waste is converted into a liquefied waste and any residual solid waste. The resulting liquefied waste can be separated from said any residual solidified waste. See e.g., U.S. Patent No. 5,709,796.

[0603] The xylanases of the invention can also be used in any waste treatment process, wherein the xylanase is used as an antimicrobial or microbial repellent. The xylanase of the invention can be included in any waste treatment composition, wherein the xylanases of the invention act as an antimicrobial or microbial repellent in the composition.

Oral care products

[0604] Oral care product may comprise xylanases of the invention, including the enzyme mixtures or "cocktails" of the invention. Oral care products include toothpastes, dental creams, gels or tooth powders, odontics, mouth washes, pre- or post brushing rinse formulations, chewing gums, lozenges, or candy. See, e.g., U.S. Patent No. 6,264,925.

[0605] The xylanases of the invention, including the enzyme mixtures or "cocktails" of the invention, can also be used in any oral care manufacturing process, wherein the xylanase is used as an antimicrobial or microbial repellent. The xylanase of the invention, including the enzyme mixtures or "cocktails" of the invention, can be included in any oral care composition, wherein the xylanases of the invention act as an antimicrobial or microbial repellent in the composition.

Brewing and fermenting

[0606] Methods of brewing (e.g., fermenting) beer may comprise xylanases of the invention, including the enzyme mixtures or "cocktails" of the invention. Starch-containing raw materials may be disintegrated and processed to form a malt. A xylanase of the invention is used at any point in the fermentation process. For example, xylanases of the invention can be used in the processing of barley malt. The major raw material of beer brewing is barley malt. This can be a three stage process. First, the barley grain can be steeped to increase water content, e.g., to around about 40%. Second, the grain can be germinated by incubation at 15 to 25°C for 3 to 6 days when enzyme synthesis is stimulated under the control of gibberellins. In one aspect, xylanases of the invention are added at this (or any other) stage of the process. Xylanases of the invention can be used in any beer or alcoholic beverage producing process, as described, e.g., in U.S. Patent No. 5,762,991; 5,536,650; 5,405,624; 5,021,246; 4,788,066.

[0607] In one aspect, an enzyme of the invention is used to improve filterability and wort viscosity and to obtain a more complete hydrolysis of endosperm components. Use of an enzyme of the invention would also increase extract yield. The process of brewing involves germination of the barley grain (malting) followed by the extraction and the breakdown of the stored carbohydrates to yield simple sugars that are used by yeast for alcoholic fermentation. Efficient breakdown of the carbohydrate reserves present in the barley endosperm and brewing adjuncts requires the activity of several different enzymes.

[0608] In one aspect, an enzyme of the invention has activity in slightly acidic pH (e.g., 5.5-6.0) in, e.g., the 40°C to 70°C temperature range; and, in one aspect, with inactivation at 95°C. Activity under such conditions would be optimal, but are not an essential requirement for efficacy. In one aspect, an enzyme of the invention has activity between 40-75° C, and pH 5.5-6.0; stable at 70° for at least 50 minutes, and, in one aspect, is inactivated at 96-100 °C. Enzymes of the invention can be used with other enzymes, e.g., beta-1,4-endoglucanases and amylases.

[0609] The xylanases of the invention, including the enzyme mixtures or "cocktails" of the invention, can also be used in any brewing or fermentation process, wherein the xylanase is used as an antimicrobial or microbial repellent. The xylanase of the invention can be included in any brewed or fermented composition, wherein the xylanases of the invention act as an antimicrobial or microbial repellent in the composition.

Biomass conversion and biofuel production

[0610] Methods and processes for biomass conversion, e.g., to a biofuel, such as bioethanol, biomethanol, biopropanol and/or biobutanol and the like, may use enzymes of the invention, including the enzyme mixtures or "cocktails" of the invention. Fuels, e.g., biofuels, such as bioethanols, may comprise a polypeptide of the invention, including the enzyme mixtures or "cocktails" of the invention, or a polypeptide encoded by a nucleic acid of the invention. The fuel may be derived from a plant material, which optionally comprises potatoes, soybean (rapeseed), barley, rye, corn, oats, wheat, beets or sugar cane, and optionally the fuel comprises a bioethanol or a gasoline-ethanol mix.

[0611] Methods for making a fuel may comprise contacting a composition comprising a xylan, hemicellulose, cellulose or a fermentable sugar with a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention, or any one of the mixtures or "cocktails" or products of manufacture of the invention. In alternative embodiments, the composition comprising a xylan, hemicellulose, a cellulose or a fermentable sugar comprises a plant, plant product or plant derivative, and the plant or plant product can comprise cane sugar plants or plant products, beets or sugarbeets, wheat, corn, soybeans, potato, rice or barley. In alternative embodiments, the polypeptide has activity comprising catalyzing hydrolysis of internal P-1,4-xylosidic linkages or endo- β -1,4-glucanase linkages; and/or degrading a linear polysaccharide beta-1,4-xylan into xylose. In one aspect, the fuel comprises a bioethanol or a gasoline-ethanol mix, or a biopropanol or a gasoline-propanol mix, or a biobutanol or a gasoline-butanol mix, or a biomethanol or a gasoline-methanol mix, or any combination thereof.

[0612] Methods for making bioethanol, biobutanol, biomethanol and/or a biopropanol may comprise contacting a composition comprising a xylan, hemi-cellulose, cellulose or a fermentable sugar with a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention, or any one of the mixtures or "cocktails" or products of manufacture of the invention. The composition comprising a cellulose or a fermentable sugar may comprise a plant, plant product or plant derivative, and the plant or plant product can comprise cane sugar plants or plant products, beets or sugarbeets, wheat, corn, soybeans, potato, rice or barley.

[0613] The invention provides enzyme ensembles, or "cocktails", for depolymerization of cellulosic and hemicellulosic polymers, xylans, and polysaccharides to metabolizeable carbon moieties comprising a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention. In alternative embodiments, the polypeptide has activity comprising catalyzing hydrolysis of internal β -1,4-xylosidic linkages or endo- β -1,4-glucanase linkages; and/or degrading a linear polysaccharide beta-1,4-xylan into

xylose. The enzyme ensembles, or "cocktails", of the invention can be in the form of a composition (e.g., a formulation, liquid or solid), e.g., as a product of manufacture. The invention further enzymes, enzyme ensembles, or "cocktails" for depolymerization of cellulosic and hemicellulosic polymers, xylans and polysaccharides, to simpler moieties, such as sugars, which are then microbially fermented to generate products such as succinic acid, lactic acid, or acetic acid.

[0614] The invention provides compositions (including products of manufacture, enzyme ensembles, or "cocktails") comprising a mixture (or "cocktail") of hemicellulose- and cellulose-hydrolyzing enzymes, wherein the xylan-hydrolyzing enzymes comprise at least one of each of a xylanase of the invention and at least one, several or all of a cellulase, glucanase, a cellobiohydrolase and/or a β -glucosidase. In alternative embodiments, the xylan-hydrolyzing and/or hemicellulose-hydrolyzing mixtures of the invention comprise at least one of each of a xylanase of the invention and at least one or both of a β -xylosidase and/or an arabinofuranosidase.

[0615] The invention provides compositions (including products of manufacture, enzyme ensembles, or "cocktails") comprising a mixture (or "cocktail") of xylan-hydrolyzing, hemicellulose- and/or cellulose-hydrolyzing enzymes comprising at least one, several or all of a cellulase, a glucanase, a cellobiohydrolase and/or an arabinofuranosidase, and a xylanase of this invention.

[0616] The invention provides compositions (including products of manufacture, enzyme ensembles, or "cocktails") comprising mixture (or "cocktail") of xylan-hydrolyzing, hemicellulose- and/or cellulose-hydrolyzing enzymes comprising at least one, several or all of a cellulase, a glucanase; a cellobiohydrolase; an arabinofuranosidase; a xylanase; a β -glucosidase; a β -xylosidase; and at least one enzyme of the invention.

[0617] The invention provides compositions (including products of manufacture, enzyme ensembles, or "cocktails") comprising mixture (or "cocktail") of enzymes comprising, in addition to at least one enzyme of the invention: (1) a glucanase which cleaves internal β -1,4 linkages resulting in shorter glucooligosaccharides, (2) a cellobiohydrolase which acts in an "exo" manner processively releasing cellobiose units (β -1,4 glucose - glucose disaccharide), and/or (3) a β -glucosidase for releasing glucose monomer from short cellooligosaccharides (e.g. cellobiose).

Biomass conversion and production of clean bio fuels

[0618] Enzymes of this invention, including mixtures, or "cocktails" of enzymes of the invention, may be used in compositions and processes for the conversion of a biomass, or any organic material, e.g., any xylan-comprising or lignocellulosic material (e.g., any composition comprising a xylan, cellulose, hemicellulose and/or lignin), to a fuel, such as a biofuel (e.g., bioethanol, biobutanol, biomethanol and/or a biopropanol), including biodiesels, in addition to feeds, foods, food or feed supplements (additives), pharmaceuticals and chemicals. These compositions and methods of the invention provide effective and sustainable alternatives or adjuncts to use of petroleum-based products, e.g., as a mixture of a biofuel (e.g., bioethanol, biobutanol, biomethanol and/or a biopropanol) and gasoline and/or diesel fuel.

[0619] Cells and/or organisms expressing enzymes of the invention (e.g., wherein the cells or organisms comprise as heterologous nucleic acids a sequence of this invention) are suitable for participation in chemical cycles involving natural biomass (e.g., plant) conversion. Enzymes and methods for the conversion may be used in enzyme ensembles (or "cocktails") for the efficient depolymerization of xylan-comprising compositions, or xylan, cellulosic and hemicellulosic polymers, to metabolizeable carbon

moieties. Methods for discovering and implementing the most effective of enzymes enable these important new "biomass conversion" and alternative energy industrial processes.

[0620] Enzymes and mixtures of enzymes or "cocktails" of the invention may be used in methods, for processing a material, e.g. a biomass material, comprising a celooligosaccharide, an arabinoxylan oligomer, a lignin, a lignocellulose, a xylan, a glucan, a cellulose and/or a fermentable sugar comprising contacting the composition with a polypeptide of the invention, or a polypeptide encoded by a nucleic acid of the invention, wherein optionally the material is derived from an agricultural crop (e.g., wheat, barley, potatoes, switchgrass, poplar wood), is a byproduct of a food or a feed production, is a lignocellulosic waste product, or is a plant residue or a waste paper or waste paper product, and optionally the plant residue comprise stems, leaves, hulls, husks, corn or corn cobs, corn stover, corn fiber, hay, straw (e.g. rice straw or wheat straw), sugarcane bagasse, sugar beet pulp, citrus pulp, and citrus peels, wood, wood thinnings, wood chips, wood pulp, pulp waste, wood waste, wood shavings and sawdust, construction and/or demolition wastes and debris (e.g. wood, wood shavings and sawdust), and optionally the paper waste comprises discarded or used photocopy paper, computer printer paper, notebook paper, notepad paper, typewriter paper, newspapers, magazines, cardboard and paper-based packaging materials, and recycled paper materials. In addition, urban wastes, e.g. the paper fraction of municipal solid waste, municipal wood waste, and municipal green waste, along with other materials containing sugar, starch, and/or cellulose can be used. Optionally the processing of the material, e.g. the biomass material, generates a bioalcohol, e.g., a bioethanol, biomethanol, biobutanol or biopropanol.

[0621] Alternatively, the polypeptide of the invention may be expressed in the biomass plant material or feedstock itself.

[0622] These methods also include taking the converted biomass (e.g., lignocellulosic) material (processed by enzymes of the invention) and making it into a fuel (e.g. a biofuel such as a bioethanol, biobutanol, biomethanol, a biopropanol, or a biodiesel) by fermentation and/or by chemical synthesis. The produced sugars may be fermented and/or the non-fermentable products gasified.

[0623] Methods of converting algae, virgin vegetable oils, waste vegetable oils, animal fats and greases (e.g. tallow, lard, and yellow grease), or sewage, may use enzymes of the invention, and making it into a fuel (e.g. a bioalcohol, e.g., a bioethanol, biomethanol, biobutanol or biopropanol, or biodiesel) by fermentation and/or by chemical synthesis or conversion.

[0624] The enzymes of the invention (including, for example, organisms, such as microorganisms, e.g., fungi, yeast or bacteria, and plants and plant cells and plant parts, e.g., seeds, making and in some aspects secreting recombinant enzymes of the invention) can be used in or included/ integrated at any stage of any organic matter/ biomass conversion process, e.g., at any one step, several steps, or included in all of the steps, or all of the following methods of biomass conversion processes, or all of these biofuel alternatives:

- Direct combustion: the burning of material by direct heat and is the simplest biomass technology; can be very economical if a biomass source is nearby.
 1. 1 Pyrolysis: is the thermal degradation of biomass by heat in the absence of oxygen. In one aspect, biomass is heated to a temperature between about 800 and 1400 degrees Fahrenheit, but no oxygen is introduced to support combustion resulting in the creation of gas, fuel oil and charcoal.
 2. 2 Gasification: biomass can be used to produce methane through heating or anaerobic digestion. Syngas, a mixture of carbon monoxide and hydrogen, can be derived from biomass.

- Landfill Gas: is generated by the decay (anaerobic digestion) of buried garbage in landfills. When the organic waste decomposes, it generates gas consisting of approximately 50% methane, the major component of natural gas.
- Anaerobic digestion: converts organic matter to a mixture of methane, the major component of natural gas, and carbon dioxide. In one aspect, biomass such as wastewater (sewage), manure, or food processing waste, is mixed with water and fed into a digester tank without air.
- Fermentation
 - Alcohol Fermentation: fuel alcohol is produced by converting cellulosic mass and/or starch to sugar, fermenting the sugar to alcohol, then separating the alcohol water mixture by distillation. Feedstocks such as dedicated crops (e.g., wheat, barley, potatoes, switchgrass, poplar wood), agricultural residues and wastes (e.g. rice straw, corn stover, wheat straw, sugarcane bagasse, rice hulls, corn fiber, sugar beet pulp, citrus pulp, and citrus peels), forestry wastes (e.g. hardwood and softwood thinnings, hardwood and softwood residues from timber operations, wood shavings, and sawdust), urban wastes (e.g. paper fraction of municipal solid waste, municipal wood waste, municipal green waste), wood wastes (e.g. saw mill waste, pulp mill waste, construction waste, demolition waste, wood shavings, and sawdust), and waste paper or other materials containing sugar, starch, and/or cellulose can be converted to sugars and then to alcohol by fermentation with yeast. Alternatively, materials containing sugars can be converted directly to alcohol by fermentation.
- Transesterification: An exemplary reaction for converting oil to biodiesel is called transesterification. The transesterification process reacts an alcohol (like methanol) with the triglyceride oils contained in vegetable oils, animal fats, or recycled greases, forming fatty acid alkyl esters (biodiesel) and glycerin. The reaction requires heat and a strong base catalyst, such as sodium hydroxide or potassium hydroxide.
- Biodiesel: Biodiesel is a mixture of fatty acid alkyl esters made from vegetable oils, animal fats or recycled greases. Biodiesel can be used as a fuel for vehicles in its pure form, but it is usually used as a petroleum diesel additive to reduce levels of particulates, carbon monoxide, hydrocarbons and air toxics from diesel-powered vehicles.
- Hydrolysis: includes hydrolysis of a compound, e.g., a biomass, such as a lignocellulosic material, catalyzed using an enzyme of the instant invention.
- Cogeneration: is the simultaneous production of more than one form of energy using a single fuel and facility. In one aspect, biomass cogeneration has more potential growth than biomass generation alone because cogeneration produces both heat and electricity.

[0625] In one aspect, the polypeptides of the invention have sufficient xylanase activity, for, or can be used with other enzymes in a process for, generating a biodiesel or a fuel, (e.g. a bioalcohol, e.g., a bioethanol, biomethanol, biobutanol or biopropanol, or biodiesel) from an organic material, e.g., a biomass, such as compositions derived from plants and animals, including any agricultural crop or other renewable feedstock, an agricultural residue or an animal waste, the organic components of municipal and industrial wastes, or construction or demolition wastes or debris, or microorganisms such as algae or yeast.

[0626] Polypeptides of the invention may be used in processes for converting an organic material, e.g., a biomass, such as a lignocellulosic biomass, to a biofuel, such as a bioethanol, biobutanol, biomethanol, a biopropanol, or otherwise are used in processes for hydrolyzing or digesting biomaterials such that they can be used as a biofuel (including biodiesel or bioethanol, biobutanol, biomethanol or biopropanol), or for making it easier for the biomass to be processed into a fuel. Polypeptides of the invention may be used in processes for a transesterification process reacting an alcohol (like methanol, butanol, propanol, ethanol) with a triglyceride oil contained in a vegetable oil, animal fat or recycled greases, forming fatty acid alkyl

esters (biodiesel) and glycerin. Biodiesel may be made from soybean oil or recycled cooking oils. Animal's fats, other vegetable oils, and other recycled oils can also be used to produce biodiesel, depending on their costs and availability. Blends of all kinds of fats and oils may be used to produce a biodiesel fuel of the invention.

[0627] Enzymes of the invention can also be used in glycerin refining. The glycerin by-product contains unreacted catalyst and soaps that are neutralized with an acid. Water and alcohol are removed to produce 50% to 80% crude glycerin. The remaining contaminants include unreacted fats and oils, which can be processed using the polypeptides of the invention. In a large biodiesel plants, the glycerin can be further purified, e.g., to 99% or higher purity, for the pharmaceutical and cosmetic industries.

[0628] Fuels (including bioalcohols such as bioethanols, biomethanols, biobutanols or biopropanols, or biodiesels) made using the polypeptides of the invention, including the mixture of enzymes or "cocktails" of the invention, can be used with fuel oxygenates to improve combustion characteristics. Adding oxygen results in more complete combustion, which reduces carbon monoxide emissions. This is another environmental benefit of replacing petroleum fuels with biofuels (e.g., a fuel of the invention). A biofuel made using these compositions and/or methods can be blended with gasoline to form an E10 blend (about 5% to 10% ethanol and about 90% to 95% gasoline), but it can be used in higher concentrations such as E85 or in its pure form. A biofuel so made can be blended with petroleum diesel to form a B20 blend (20% biodiesel and 80% petroleum diesel), although other blend levels can be used up to B100 (pure biodiesel).

[0629] The polypeptides of this invention may be used in processes for converting organic material, e.g., a biomass, such as a lignocellulosic biomass, to methanol, butanol, propanol and/or ethanol, including processes for making ethanol ("bioethanol") methanol, butanol and/or propanol from compositions comprising organic material, e.g., a biomass, such as a lignocellulosic biomass. The organic material, e.g., a biomass, such as a lignocellulose biomass material, can be obtained from agricultural crops, as a byproduct of food or feed production, or as biomass waste products, such as plant residues and waste paper or construction and/or demolition wastes or debris. Examples of suitable plant residues for treatment with polypeptides of the invention include grains, seeds, stems, leaves, hulls, husks, corn cobs, corn stover, straw, grasses (e.g., Indian grass, such as *Sorghastrum nutans*; or, switch grass, e.g., *Panicum species*, such as *Panicum virgatum*), and the like, as well as wood, wood chips, wood pulp, and sawdust. Examples of paper waste suitable for treatment with polypeptides of the invention include discard photocopy paper, computer printer paper, notebook paper, notepad paper, typewriter paper, and the like, as well as newspapers, magazines, cardboard, and paper-based packaging materials. Examples of construction and demolition wastes and debris include wood, wood scraps, wood shavings and sawdust.

[0630] The enzymes of the invention can be used in conjunction with more "traditional" means of making methanol, butanol, propanol and/or ethanol from biomass, e.g., as methods comprising hydrolyzing biomass (e.g., lignocellulosic materials) by subjecting dried biomass material in a reactor to a catalyst comprised of a dilute solution of a strong acid and a metal salt; this can lower the activation energy, or the temperature, of cellulose hydrolysis to obtain higher sugar yields; see, e.g., U.S. Patent Nos. 6,660,506; 6,423,145.

[0631] Another method that incorporates use of enzymes of the invention comprises hydrolyzing biomass (e.g., lignocellulosic materials) containing xylan, hemicellulose, cellulose and/or lignin by subjecting the material to a first stage hydrolysis step in an aqueous medium at a temperature and a pressure chosen to effect primarily depolymerization of hemicellulose without major depolymerization of cellulose to glucose. This step results in a slurry in which the liquid aqueous phase contains dissolved monosaccharides resulting from depolymerization of hemicellulose and a solid phase containing cellulose and lignin. A

second stage hydrolysis step can comprise conditions such that at least a major portion of the cellulose is depolymerized, such step resulting in a liquid aqueous phase containing dissolved/ soluble depolymerization products of cellulose. See, e.g., U.S. Patent No. 5,536,325. Enzymes of the invention can be added at any stage of this process.

[0632] Another method that incorporates use of enzymes of the invention comprises processing a biomass material by one or more stages of dilute acid hydrolysis with about 0.4% to 2% strong acid; and treating an unreacted solid lignocellulosic component of the acid hydrolyzed biomass material by alkaline delignification to produce precursors for biodegradable thermoplastics and derivatives. See, e.g., U.S. Patent No. 6,409,841. Enzymes of the invention can be added at any stage of this process.

[0633] Another method that incorporated use of enzymes of the invention comprises prehydrolyzing biomass (e.g., lignocellulosic materials) in a prehydrolysis reactor; adding an acidic liquid to the solid lignocellulosic material to make a mixture; heating the mixture to reaction temperature; maintaining reaction temperature for time sufficient to fractionate the lignocellulosic material into a solubilized portion containing at least about 20% of the lignin from the lignocellulosic material and a solid fraction containing cellulose; removing a solubilized portion from the solid fraction while at or near reaction temperature wherein the cellulose in the solid fraction is rendered more amenable to enzymatic digestion; and recovering a solubilized portion. See, e.g., U.S. Patent No. 5,705,369. Enzymes of the invention can be added at any stage of this process.

[0634] Motor fuel compositions (e.g., for spark ignition motors) based on liquid hydrocarbons blended with a fuel grade alcohol may be made by using an enzyme of the invention. The fuels made by use of an enzyme of the invention comprise, e.g., coal gas liquid- or natural gas liquid-ethanol blends. A co-solvent may be biomass-derived 2-methyltetrahydrofuran (MTHF). See, e.g., U.S. Patent No. 6,712,866.

[0635] Enzymatic degradation of biomass (e.g., lignocellulosic materials), e.g., for production of a biofuel, e.g., an ethanol, from a biomass or any organic material, can also comprise use of ultrasonic treatment of a biomass material; see, e.g., U.S. Patent No. 6,333,181.

[0636] A biofuel, e.g., an ethanol (a bioethanol) produced from a biomass (e.g., a cellulosic) substrate by providing a reaction mixture in the form of a slurry comprising biomass (e.g., a cellulosic) substrate, an enzyme of this invention and a fermentation agent (e.g., within a reaction vessel, such as a semi-continuously solids-fed bioreactor), and the reaction mixture is reacted under conditions sufficient to initiate and maintain a fermentation reaction (as described, e.g., in U.S. Pat. App. No. 20060014260). Experiment or theoretical calculations can determine an optimum feeding frequency. Additional quantities of the biomass (e.g., a cellulosic) substrate and the enzyme are provided into the reaction vessel at an interval(s) according to the optimized feeding frequency.

[0637] One process for making a biofuels and biodiesels is described in U.S. Pat. App. Pub. Nos. 20050069998; 20020164730; and comprises stages of grinding the biomass (e.g., lignocellulosic material) (e.g., to a size of 15-30 mm), subjecting the product obtained to steam explosion pre-treatment (e.g., at a temperature of 190-230°C) for between 1 and 10 minutes in a reactor; collecting the pre-treated material in a cyclone or related product of manufacture; and separating the liquid and solid fractions by filtration in a filter press, introducing the solid fraction in a fermentation deposit and adding one or more enzymes of the invention, and optionally, another enzyme is also added, e.g., a cellulase and/or beta-glucosidase enzyme (e.g., dissolved in citrate buffer pH 4.8).

[0638] Another process for making a biofuels and biodiesels comprising methanol, butanol, propanol and/or ethanol using enzymes of the invention comprises pretreating a starting material comprising a

biomass (e.g., a lignocellulosic) feedstock comprising at least a xylan, a hemicellulose and/or a cellulose. The starting material may comprise potatoes, soybean (rapeseed), barley, rye, corn, oats, wheat, beets or sugar cane or a component or waste or food or feed production byproduct. The starting material ("feedstock") is reacted at conditions which disrupt the plant's fiber structure to effect at least a partial hydrolysis of the biomass (e.g., hemicellulose and/or cellulose). Disruptive conditions can comprise, e.g., subjecting the starting material to an average temperature of 180°C to 270°C at pH 0.5 to 2.5 for a period of about 5 seconds to 60 minutes; or, temperature of 220°C to 270°C, at pH 0.5 to 2.5 for a period of 5 seconds to 120 seconds, or equivalent. This generates a feedstock with increased accessibility to being digested by an enzyme of the invention. U.S. Patent No. 6,090,595.

[0639] Exemplary conditions for hydrolysis of biomass (e.g., a lignocellulosic material) by an enzyme of this invention include reactions at temperatures between about 30°C and 48°C, and/or a pH between about 4.0 and 6.0. Other exemplary conditions include a temperature between about 30°C and 60°C and a pH between about 4.0 and 8.0.

Biofuels and biologically produced alcohols

[0640] Biofuels and synthetic fuels, including liquids and gases (e.g., syngas) and biologically produced alcohols, may be made, using the enzyme and nucleic acids, and transgenic plants, animal, seeds and microorganisms of the invention. Biofuels and biologically produced alcohols may comprise enzymes, nucleic acids, transgenic plants, animals (e.g., microorganisms, such as bacteria or yeast) and/or seeds of the invention. These biofuels and biologically produced alcohols may be produced from a biomass.

[0641] Biologically produced alcohols, such as ethanol, methanol, propanol and butanol produced by methods of the disclosure include by the action of microbes and enzymes of the invention through fermentation (hydrolysis) to result in an alcohol fuel.

Biofuels as a liquid or a gas gasoline

[0642] Enzymes of the invention may be used for chemical cycles for natural biomass conversion, e.g., for the hydrolysis of a biomass to make a biofuel, e.g., a bioethanol, biopropanol, bio-butanol or a biomethanol, or a synthetic fuel, in the form of a liquid or as a gas, such as a "syngas".

[0643] This biofuel gas of the invention may be mixed with a natural gas (can also be produced from biomass), e.g., a hydrogen or a hydrocarbon-based gas fuel. Methods for processing biomass may produce a synthetic fuel, e.g., a syngas, such as a syngas produced from a biomass by gasification. An ethanol, propanol, butanol and/or methanol gas may be made from a sugar cane, e.g., a bagasse. This fuel, or gas, may be used as motor fuel, e.g., an automotive, truck, airplane, boat, small engine, etc. fuel. Ethanol, propanol, butanol and/or methanol may be made from a plant, e.g., corn, or a plant product, e.g., hay or straw (e.g., a rice straw or a wheat straw, or any the dry stalk of any cereal plant), or an agricultural waste product. Cellulosic ethanol, propanol, butanol and/or methanol can be manufactured from a plant, e.g., corn, or plant product, e.g., hay or straw, or an agricultural waste product (e.g., as processed by Iogen Corporation of Ontario, Canada).

[0644] The ethanol, propanol, butanol and/or methanol so made can be used as a fuel (e.g., a gasoline) additive (e.g., an oxygenator) or in a direct use as a fuel. An ethanol, propanol, butanol and/or methanol, including a fuel, so made can be mixed with ethyl tertiary butyl ether (ETBE), or an ETBE mixture such as

ETBE containing 47% ethanol as a biofuel, or with MTBE (methyl tertiary-butyl ether). An ethanol, propanol, butanol and/or methanol, including a fuel, so made can be mixed with:

IUPAC name	Common name
but-1-ene	α-butylene
<i>cis</i> -but-2-ene	<i>cis</i> - β -butylene
<i>trans</i> -but-2-ene	<i>trans</i> - β -butylene
2-methylpropene	isobutylene

[0645] A butanol and/or ethanol made using an enzyme of the invention can be further processed using "A.B.E." (Acetone, Butanol, Ethanol) fermentation; butanol being the only liquid product. This butanol and/or ethanol may be burned "straight" in existing gasoline engines (without modification to the engine or car), produces more energy and is less corrosive and less water soluble than ethanol, and can be distributed via existing infrastructures.

[0646] Mixed alcohols may include one, several or all of the alcohols made by processes using an enzyme of the invention e.g., comprising a mixture of ethanol, propanol, butanol, pentanol, hexanol, and heptanol, such as ECALENE™ (Power Energy Fuels, Inc., Lakewood, CO), e.g.:

Exemplary Fuel	
Component	Weight %
Methanol	0%
Ethanol	75%
Propanol	9%
Butanol	7%
Pentanol	5%
Hexanol & Higher	4%

[0647] One, several or all of these alcohols may be made by a process using an enzyme of the invention, and the process can further comprise a biomass-to-liquid technology, e.g., a gasification process to produce syngas followed by catalytic synthesis, or by a bioconversion of biomass to a mixed alcohol fuel.

[0648] Processes using an enzyme of the invention may incorporate (or, be incorporated into) "gas to liquid", or GTL; or "coal to liquid", or CTL; or "biomass to liquid" or BTL; or "oilsands to liquid", or OTL, processes; and these processes of the invention may be used to make synthetic fuels. One of these processes comprises making a biofuel (e.g., a synfuel) out of a biomass using, e.g., the so-called "Fischer Tropsch" process (a catalyzed chemical reaction in which carbon monoxide and hydrogen are converted into liquid hydrocarbons of various forms; typical catalysts used are based on iron and cobalt; the principal purpose of this process is to produce a synthetic petroleum substitute for use as synthetic lubrication oil or as synthetic fuel). This synthetic biofuel can contain oxygen and can be used as additive in high quality diesel and petrol.

[0649] Various pretreatments, which can be grouped into three categories: physical, chemical, and multiple (physical + chemical) can be used. Any chemicals can be used as a pretreatment agent, e.g., acids, alkalis, gases, cellulose solvents, alcohols, oxidizing agents and reducing agents. Among these

chemicals, alkali is the most popular pretreatment agent because it is relatively inexpensive and results in less cellulose degradation. The common alkalis sodium hydroxide and lime also can be used as pretreatment agents. Although sodium hydroxide increases biomass digestibility significantly, it is difficult to recycle, is relatively expensive, and is dangerous to handle. In contrast, lime has many advantages: it is safe and very inexpensive, and can be recovered by carbonating wash water with carbon dioxide.

[0650] A multi-enzyme system (including at least one enzyme of this invention) can hydrolyze polysaccharides in a biomass, e.g. sugarcane, e.g., bagasse, a component of sugarcane processed in sugar mills. The biomass may be processed by an enzyme of the invention made by an organism (e.g., transgenic animal, plants, transformed microorganism) and/or byproduct (e.g., harvested plant, fruit, seed) expressing an enzyme of the invention. The enzyme may be a recombinant enzyme made by the plant or biomass which is to be processed to a fuel, e.g., a transgenic sugarcane bagasse comprising an enzyme of the invention. These compositions and products may comprise chemical cycles for natural biomass conversion, e.g., for the hydrolysis of a biomass to make a biofuel, e.g., bioethanol, biopropanol, bio-butanol, biomethanol, a synthetic fuel in the form of a liquid or a gas, such as a "syngas".

[0651] A biofuel, e.g., a biogas, may be produced by the process of anaerobic digestion of organic material by anaerobes, wherein the process comprises use of an enzyme of the invention. This biofuel, e.g., a biogas, can be produced either from biodegradable waste materials or by the use of energy crops fed into anaerobic digesters to supplement gas yields. The solid output, digestate, can also be used as a biofuel.

[0652] A biofuel, e.g., a biogas, comprising a methane, may be produced in a process comprising use of an enzyme of the invention. This biofuel, e.g., a biogas, can be recovered in industrial anaerobic digesters and mechanical biological treatment systems. Landfill gas can be further processed using an enzyme of this invention; before processing landfill gas can be a less clean form of biogas produced in landfills through naturally occurring anaerobic digestion. Paradoxically if landfill gas is allowed to escape into the atmosphere it is a potent greenhouse gas.

[0653] Methods for making biologically produced oils and gases from various wastes, comprises use of an enzyme of the invention. These methods can comprise thermal depolymerization of waste to extract methane and other oils similar to petroleum; or, e.g., a bioreactor system that utilizes nontoxic photosynthetic algae to take in smokestacks flue gases and produce biofuels such as biodiesel, biogas and a dry fuel comparable to coal, e.g., as designed by GreenFuel Technologies Corporation, of Cambridge, MA.

[0654] Methods for making biologically produced oils, including crude oils, and gases that can be used in diesel engines, comprise use of an enzyme of the invention. These methods can refine petroleum, e.g., crude oils, into kerosene, petroleum, diesel and other fractions.

[0655] Enzymes of the invention are used in methods for making biologically produced oils from:

- Straight vegetable oil (SVO).
- Waste vegetable oil (WVO) - waste cooking oils and greases produced in quantity mostly by commercial kitchens.
- Biodiesel obtained from transesterification of animal fats and vegetable oil, directly usable in petroleum diesel engines.
- Biologically derived crude oil, together with biogas and carbon solids via the thermal depolymerization of complex organic materials including non oil based materials; for example, waste products such as old tires, offal, wood and plastic.

- Pyrolysis oil; which may be produced out of biomass, wood waste etc. using heat only in the flash pyrolysis process (the oil may have to be treated before using in conventional fuel systems or internal combustion engines).
- Wood, charcoal, and dried dung.

Medical and research applications

[0656] Xylanases of the invention, including the enzyme mixtures or "cocktails" of the invention, can be used as antimicrobial agents due to their bacteriolytic properties. Xylanases of the invention can be used to eliminating or protecting animals from salmonellae, as described in e.g., PCT Application Nos. WO0049890 and WO9903497. The xylanases of the invention can also be used as an antimicrobial surface cleanser or microbial repellent.

Other industrial and medical applications

[0657] As discussed above, xylanases of the invention, including the enzyme mixtures or "cocktails" of the invention, can be used, e.g., in a wide variety of industrial processes, medical and research (laboratory) applications, and food, animal feed and beverage applications. New xylanases are discovered by screening existing libraries and DNA libraries constructed from diverse mesophilic and moderately thermophilic locations as well as from targeted sources including digestive flora, microorganisms in animal waste, soil bacteria and highly alkaline habitats. Biotrap and primary enrichment strategies using arabinoxylan substrates and/or non-soluble polysaccharide fractions of animal feed material are also useful.

[0658] Two screening formats (activity-based and sequence-based) are used in the discovery of novel xylanases. The activity-based approach is direct screening for xylanase activity in agar plates using a substrate such as azo-xylan (Megazyme). Alternatively a sequence-based approach may be used, which relies on bioinformatics and molecular biology to design probes for hybridization and biopanning. See, for example, U.S. Patents No. 6,054,267, 6,030,779, 6,368,798, 6,344,328. Hits from the screening are purified, sequenced, characterized (for example, determination of specificity, temperature and pH optima), analyzed using bioinformatics, subcloned and expressed for basic biochemical characterization. These methods may be used in screening for xylanases useful in a myriad of applications, including dough conditioning and as animal feed additive enzymes.

[0659] In characterizing enzymes obtained from screening, the exemplary utility in dough processing and baking applications may be assessed. Characterization may include, for example, measurement of substrate specificity (xylan, arabinoxylan, CMC, BBG), temperature and pH stability and specific activity. A commercial enzyme may be used as a benchmark. The enzymes of the invention may have significant activity at pH ≥ 7 and 25-35° C, are inactive on insoluble xylan, are stable and active in 50-67% sucrose.

[0660] In another aspect, utility as feed additives may be assessed from characterization of candidate enzymes. Characterization may include, for example, measurement of substrate specificity (xylan, arabinoxylan, CMC, BBG), temperature and pH stability, specific activity and gastric stability. The feed may be designed for a monogastric animal or designed for a ruminant animal. The enzymes of the invention may have significant activity at pH 2-4 and 35-40°C, a half-life greater than 30 minutes

in gastric fluid, formulation (in buffer or cells) half-life greater than 5 minutes at 85°C and are used as a monogastric animal feed additive. The enzymes of the invention may have one or more of the following characteristics: significant activity at pH 6.5-7.0 and 35-40°C, a half-life greater than 30 minutes in rumen fluid, formulation stability as stable as dry powder and are used as a ruminant animal feed additive.

[0661] Enzymes are reactive toward a wide range of natural and unnatural substrates, thus enabling the modification of virtually any organic lead compound. Moreover, unlike traditional chemical catalysts, enzymes are highly enantio- and regio-selective. The high degree of functional group specificity exhibited by enzymes enables one to keep track of each reaction in a synthetic sequence leading to a new active compound. Enzymes are also capable of catalyzing many diverse reactions unrelated to their physiological function in nature. For example, peroxidases catalyze the oxidation of phenols by hydrogen peroxide. Peroxidases can also catalyze hydroxylation reactions that are not related to the native function of the enzyme. Other examples are xylanases which catalyze the breakdown of polypeptides. In organic solution some xylanases can also acylate sugars, a function unrelated to the native function of these enzymes.

[0662] The present invention exploits the unique catalytic properties of enzymes. Whereas the use of biocatalysts (i.e., purified or crude enzymes, non-living or living cells) in chemical transformations normally requires the identification of a particular biocatalyst that reacts with a specific starting compound, the present invention uses selected biocatalysts and reaction conditions that are specific for functional groups that are present in many starting compounds. Each biocatalyst is specific for one functional group, or several related functional groups and can react with many starting compounds containing this functional group. The biocatalytic reactions produce a population of derivatives from a single starting compound. These derivatives can be subjected to another round of biocatalytic reactions to produce a second population of derivative compounds. Thousands of variations of the original compound can be produced with each iteration of biocatalytic derivatization.

[0663] Enzymes react at specific sites of a starting compound without affecting the rest of the molecule, a process which is very difficult to achieve using traditional chemical methods. This high degree of biocatalytic specificity provides the means to identify a single active compound within the library. The library is characterized by the series of biocatalytic reactions used to produce it, a so-called "biosynthetic history". Screening the library for biological activities and tracing the biosynthetic history identifies the specific reaction sequence producing the active compound. The reaction sequence is repeated and the structure of the synthesized compound determined. This mode of identification, unlike other synthesis and screening approaches, does not require immobilization technologies and compounds can be synthesized and tested free in solution using virtually any type of screening assay. It is important to note, that the high degree of specificity of enzyme reactions on functional groups allows for the "tracking" of specific enzymatic reactions that make up the biocatalytically produced library.

[0664] Many of the procedural steps are performed using robotic automation enabling the execution of many thousands of biocatalytic reactions and screening assays per day as well as ensuring a high level of accuracy and reproducibility. As a result, a library of derivative compounds can be produced in a matter of weeks which would take years to produce using current chemical methods. (For further teachings on modification of molecules, including small molecules, see PCT/US94/09174).

[0665] A composition may comprise at least one mucoadhesive polymer that is capable of forming a hydrogel and at one least water soluble polymer, and one or more enzymes of the invention. This formulation can be used in any industrial, food or feed processing or medical or research application of the invention, i.e., any application using an enzyme or nucleic acid of the invention. The formulation forms a hydrogel in aqueous solution that has mucoadhesive properties; this can be capable of releasing enzymes, microorganisms capable of generating enzymes of the invention, or antibodies of the invention, over an

extended period of time. Alternatively, the hydrogel can entrap enzymes, microorganisms capable of generating enzymes of the invention, or antibodies of the invention and release them over a defined (e.g., an extended) period of time.

[0666] The invention will be further described with reference to the following examples; however, it is to be understood that the invention is not limited to such examples.

EXAMPLES

EXAMPLE 1: Xylanase assay with wheat arabinoxylan as substrate

[0667] The following example describes an exemplary xylanase assay that can be used, for example, to determine if an enzyme is within the scope of the invention. Enzymes of the invention, e.g., SEQ ID NO:2 having one or more amino acid residue changes (mutations) as described herein, can be subjected to an assay at pH 8 (Na-phosphate buffer) and 70°C using wheat arabinoxylan as a substrate.

EXAMPLE 2: Determination of melting temperature and xylanase activity

Differential Scanning Calorimetry (DSC)

[0668] The melting temperature transition midpoint (T_m) for each enzyme of the invention, can be determined by Differential Scanning Calorimetry (DSC). Baseline subtracted DSC data can be normalized for protein concentration.

[0669] In one assay, calorimetry can be performed using a Model 6100 NANO II DSC™ apparatus (Calorimetry Sciences Corporation, American Fork, UT) using the DSCRUN™ (DSCRun) software package for data acquisition, CPCALC™ (CpCalc) for analysis, CPCONVERT™ (CpConvert) for conversion into molar heat capacity from microwatts and CPDECONVOLUTE™ (CpDeconvolute) for deconvolution. Analysis can be carried out with 1 mg/ml recombinant protein in 20 mM potassium phosphate (pH 7.0) and 100 mM KCl at a scan rate of 1°C/min. A constant pressure of 5 atm can be maintained during all DSC experiments to prevent possible degassing of the solution on heating. The instrumental baseline can be recorded routinely before the experiments with both cells filled with buffer. Reversibility of the thermally induced transitions can be tested by reheating the solution in the calorimeter cell immediately after cooling the first run.

[0670] Alternatively, DSC measurements can be made using a VP-DSC microcalorimeter (Micro-Cal) in duplicate. In one aspect, a required sample volume is 540 μ L. The concentrations of the protein can be between 0.1 to 0.5 mg/mL in 50mM HEPES, pH 7.2; a sample of the dialysis buffer can be retained for base line controls. Each sample can be heated from 40°C to 110°C. Samples and/or buffer can be heated and cooled at a scan rate of 90°C/h. Buffer baselines were recorded multiple times until the system reached a stable state. The T_m value was the temperature where maximum heat was released.

Xylanase activity assays

[0671] Enzymatic activities can be determined using 400 μL of 2% Azo-xylan as substrate in 550 μL of CP (citrate-phosphate) buffer, pH 6.0 at the indicated temperatures. Activity measurements as a function of pH can be determined using 50 mM Britton and Robinson buffer solutions (pH 3.0, 5.0, 6.0, 7.0, 8.0 and 9.0) prepared by mixing solutions of 0.1 M phosphoric acid solution, 0.1 M boric acid and 0.1 M acetic acid followed by pH adjustment with 1 M sodium hydroxide. Reactions can be initiated by adding 50 μL of 0.1 mg/ml of purified enzyme. Time points can be taken from 0 to 15 minutes where 50 μL of reaction mixture are added to 200 μL of precipitation solution (100% ethanol). When all time points have been taken, samples are mixed, incubated for 10 minutes and centrifuged at 3000 g for 10 minutes at 4°C. Supernatant (150 μL) can be aliquoted into a fresh 96 well plate and absorbance is measured at 590 nm. A590 values can be plotted against time and the initial rate is determined from the slope of the line.

Polysaccharide Fingerprinting.

[0672] Polysaccharide fingerprints can be determined by polysaccharide analysis using carbohydrate gel electrophoresis (PACE). Beechwood xylan (0.1 mg/mL, 100 μL , Sigma, Poole, Dorset, UK) or xylooligosaccharides (1 mM, 20 μL , Megazyme, Wicklow, Ireland) can be treated with enzyme (1 - 3 μg) in a total volume of 250 μL for 16 hours. The reaction is buffered in 0.1 M ammonium acetate pH 5.5. Controls without substrates or enzymes are performed under the same conditions to identify any unspecific compounds in the enzymes, polysaccharides/oligosaccharides or labeling reagents. The reactions are stopped by boiling for 20 min. Assays can be independently performed at least 2 times for each condition. Derivatization using ANTS (8-aminonaphthalene-1,3,6-trisulfonic acid, Molecular Probes, Leiden, The Netherlands), electrophoresis and imaging are carried out as described (Goubet, F., Jackson, P., Deery, M. and Dupree, P. (2002) Anal. Biochem. 300, 53-68).

Fitness Calculation.

[0673] The fitness (F_n), for a given enzyme variant, n , can be calculated by equally weighting increase in denaturation temperature transition midpoint (T_m) and increase (or decrease) in enzymatic activity relative to the largest difference in each parameter across all variants: $F_n = F_{Tn} + F_{Vn}$, where F_{Tn} = T_m fitness factor of the variant and F_{Vn} = activity fitness factor of the variant. The fitness factors for each (T_m and activity) are relative to the largest difference in T_m or rate across all of the variants. $F_{Tn} = (T_m - T_{mL}) / (T_{mH} - T_{mL})$ where T_{mn} is the T_m for the given variant, n , and T_{mL} is the lowest T_m across all variants and T_{mH} the highest T_m across all variants and $F_{Vn} = (V_n - V_L) / (V_H - V_L)$ where V_n is the relative rate for the given variant, n , and V_L is the lowest rate across all variants and V_H the highest rate across all variants.

EXAMPLE 3: Pre-treating paper pulp with xylanases of the invention

[0674] Xylanases of the invention are used to treat/ pretreat paper pulp, or recycled paper or paper pulp, waste wood or wood chips, and the like. Enzyme(s) of the invention are used to increase the "brightness" of the paper via their use in treating/ pretreating paper pulp, or recycled paper or paper pulp, and the like.

[0675] Xylanases of the invention are used to treat/ pretreat paper pulp, or recycled paper or paper pulp, and the like to reduce the Kappa number. Kappa number is defined as a numerical value indicating a

paper's relative lignin content - the higher the Kappa number, the higher the lignin content. Reduction in Kappa # has benefits when treating unbleached pulp (kappa # 70 - 90), when then is used for, e.g., processing, such as in board manufacture. A reduction in Kappa across the X stage allows lower alkali use in cooking or cooking to a higher target Kappa #. This results in higher pulp strength, less machine refining and higher machine speeds. Such results are seen using digester additives (surfactants) in linerboard mills; this can allow for better liquor penetration, and allow lower effective alkali charge leading to higher pulp strength, lower refining and a 200 fpm (feet per minute) increase in machine speed.

[0676] This example describes an exemplary routine screening protocol to determine whether a xylanase is useful in pretreating paper pulp; e.g., in reducing the use of bleaching chemicals (e.g., chlorine dioxide, ClO_2) when used to pretreat Kraft paper pulp.

[0677] The screening protocol has two alternative test parameters: Impact of xylanase treatment after an oxygen delignification step (post- O_2 pulp); and, impact of xylanase in a process that does not include oxygen delignification (pre- O_2 brownstock).

[0678] Pulp or paper treatment conditions that simulate process conditions in industrial situations, e.g., factories: for example, at about pH 8.0; 70°C; 60 min duration. For example, an exemplary process of the invention is schematically depicted in the Flow Diagram of Figure 5; see also Figure 6. However, the conditions of a process of method of the invention can be adjusted to any temperature, time duration and/or pH, depending on the exemplary enzyme(s) of the invention used and the objective of the process; for example, there are a variety of ways to adjust pH in the various pulp and paper processes of the invention:

- adding acid and/or base:
- Hydrochloric acid (HCl)
- Sodium hydroxide (NaOH)
- H_2SO_4 (sulfuric acid)
- NaHSO_4 (sodium hydrogen sulfate)
- H_2SO_3 (sulfurous acid)
- H_3PO_4 (phosphoric acid)
- HF (hydrofluoric acid)
- $\text{CH}_3\text{CO}_2\text{H}$ (acetic acid)
- H_2CO_3 (carbonic acid)
- H_2S (hydrogen sulfide)
- NaH_2PO_4 (sodium dihydrogen phosphate)
- NH_4Cl (ammonium chloride)
- HCN (hydrocyanic acid)
- Na_2SO_4 (sodium sulfate)

- NaCl (sodium chloride)
- NaCH₃CO₂ (sodium acetate)
- NaHCO₃ (sodium bicarbonate)
- Na₂HPO₄ (sodium hydrogen phosphate)
- Na₂SO₃ (sodium sulfite)
- NaCN (sodium cyanide)
- NH₃ (aqueous ammonia)
- Na₂CO₃ (sodium carbonate)
- Na₃PO₄ (sodium phosphate)
- bubbling in gas, e.g. CO₂ (which forms an acid with water when dissolved)

[0679] Dose Response Determination for Xylanases on Pre-O₂ Brownstock

[0680] Conditions for xylanase stage (X-stage) as follows:

pH 8

Temperature 70°C

Time 60 min

Kappa factor 0.24

For no-enzyme control, kappa factor was 0.30

Pretreatment of Intercontinental Pre-O₂ Brownstock Xylanase

Determination of ClO₂ Dose Response in D₀

Experimental outline

Pre-O₂ Brownstock

Initial kappa 31.5

X stage conditions

Xylanase charge 0.7 U/gm

Temperature 70°C

pH 8

Treatment time 1 hr

Pulp consistency 10%

Bleach sequence XDE_p

Kappa factor 0.22, 0.26 and 0.30 (%D on pulp: 2.63, 3.12 and 3.60)

Determination of ClO₂ Dose Response in D₀:

[0681] Xylanase 0.7 U/g, pH 8.0, 70 °C, 1 hr

[0682] Pulp: Pre-O₂ Brownstock, initial kappa 31.5

[0683] Percentage saving of ClO₂ is of little significance to the industry. Their primary concern is lbs of ClO₂ required per ton OD pulp. This makes sense when one considers that a lower percentage saving seen with a high initial kappa brownstock can be more valuable in terms of lbs of ClO₂ saved than a higher percentage reduction for a low initial kappa pulp which will require a lower total charge of ClO₂ to reach target brightness.

Relationship between Brightness, Yield and Kappa Factor for Bleached Control Pulp:

[0684] Bleaching with increasing doses of ClO₂ to achieve higher target brightness results in increased loss of pulp yield. This is an issue because pulp at this stage of the process has a value of almost \$400 per ton and loss of cellulose costs money.

[0685] A benefit of xylanase (e.g., a xylanase of the invention) is that use of a lower ClO₂ dose can reduce yield losses as long as the action of the xylanase itself doesn't cancel out the gain.

Dose Response Data for Pretreatment of Pre-O₂ Brownstock with Xylanase

[0686] Experimental outline

- Northwood Pre-O₂ Brownstock
 - Initial kappa 28.0
 - Initial consistency 32.46%
 - Initial brightness 28.37
- X stage conditions
 - Xylanase charge 0 to 2.70 U/gm
 - Temperature 58°C to 61°C
 - pH 8.2 to 8.5
 - Treatment time 1hr
- Bleach sequence XDE_p
 - Kappa factor 0.24
- ClO₂ saving calculated for Kappa factors between 0.24 and 0.30

[0687] The purpose of this experiment is to evaluate xylanases on unwashed SPF brownstock. Results can show dose-dependent increases in final brightness for pulp treated with XYLB (E.c), with brightness achieved in presence of xylanase at lower Kf of 0.24, approaching brightness achieved at higher Kf of 0.30 asymptotically.

Example 4: Novel Biobleaching Assay for assessing xylanase performance in enhancing the brightness of pulp

[0688] This example describes an exemplary protocol, a "biobleaching assay," that can be used to determine if a polypeptide has xylanase activity and is within the scope of the invention. This assay can be used to assess the performance of an exemplary enzyme of the invention in enhancing the brightness of a pulp, e.g., a Kraft Pulp.

[0689] The invention provides biobleaching procedures, e.g., a three-stage biobleaching procedure that closely simulates the conditions of an actual pulp mill bleach plant, as illustrated in Figure 5; including a process as illustrated in Figure 6. This bleach sequence is designated by (X)DoEp, in which X represents the xylanase treatment stage (using, e.g., an enzyme of the invention), D for chlorine dioxide bleaching stage, and Ep for alkaline peroxide extraction stage. Many different feedstocks may be used, for example, Southern Softwood Kraft Brownstock (without oxygen delignification) and hardwood Kraft pulp (e.g., maple and aspen). Upon completion of each biobleaching round, ensuing pulp can be used to produce TAPPI (Technical Association of Pulp and Paper Industries, the technical association for the worldwide pulp, paper and converting industry) - standard handsheets. The GE% brightness of each handsheet can be measured, and the brightness values can be used as the indication of how well each enzyme performs on the pulp during the enzymatic pretreatment stage (X).

[0690] Pulp biobleaching: Pulp was bleached in 10-g batches in sealed plastic bags using a 3-stage (X)DoEp sequence, as illustrated in Figure 5. The treatment conditions at the three stages can be summarized as follows:

- ❖ X stage: 10% (w/v) consistency at 65°C and pH=8 for 60 min
- ❖ Do stage: 4% (w/v) consistency at 60°C for 30 min; Kappa Factor=0.18 for enzyme treated samples, and 0.18 and 0.21 for no-enzyme controls.
- ❖ Ep stage: 10% (w/v) consistency at 75°C for 90 min; caustic charge: 1.7% (w NaOH/w OD pulp) and H₂O₂ charge: 0.5% (w/w)

[0691] As noted in Figure 5, in one aspect, raw pulp is washed to reduce pH to pH 8.5; pulp is filter pressed and divided into bags. At each stage, bags can be incubated in a water bath at the desired temperature and each bag is taken out and kneaded thoroughly every 10 min to ensure uniform mass and heat transfer within the pulp mass. After each treatment, pulp can be filtered, washed with 2 L of DI water and filtered again before receiving the next treatment. The moisture content of the pulp can be measured using a Mettler-Toledo moisture analyzer (Fisher Scientific, USA).

[0692] As noted in Figure 5, in one aspect, after the pulp is filter pressed and divided into bags, in the X

stage, the pulp can be resuspended, filter pressed, the pH adjusted; and then, incubated with enzyme at 10% solids, 65°C, 1 hour; then kneaded for 10 minutes. At the Do stage the pulp can be resuspended, washed, pH set to 4.0, and filter pressed; then, impregnated with ClO_2 at 4% solids (i.e., 4% (w/v) consistency) at 60°C for 30 min; then kneaded for 10 minutes. At the Do stage the Kappa Factor=0.18 for enzyme treated samples, and 0.18 and 0.21 for no-enzyme controls. At the Ep stage the pulp can be resuspended, washed, and filter pressed; then, incubated with NaOH and H_2O_2 at 10% solids (i.e., 10% (w/v) consistency) at 75°C for 90 min; then kneaded for 10 minutes. The caustic charge: 1.7% (w NaOH/w OD pulp) and H_2O_2 charge: 0.5% (w/w). After kneading, handsheets were formed.

[0693] Handsheets: As noted in Figure 5, in one aspect, handsheets can be formed (4 m pulp, pH about 6.5); handsheets can be made from unbleached and bleached pulp using TAPPI standard equipment (Kalamazoo Paper Chemicals, Richland, MI) according to TAPPI method T-272 sp-97. The GE% brightness of each handsheet can be measured using a BRIGHTMETER MICRO S-5/BC™ (Technidyne Corp., New Albany, IN) according to TAPPI method T-452 om-98 (reference at 457 nm).

Example 5: Novel Biobleaching Process

[0694] This example describes a novel biobleaching process of the invention, as illustrated in Figure 6. This process can be practiced using any xylanase enzyme, including a polypeptide of the invention. This exemplary process of the invention can have a starting material comprising "brownstock," which can be described as: 1) feedstock preparation - logs coming into the paper mill are debarked, chipped and screened to remove overthick chips, fines, knots and foreign matter, 2) pulping - wood chips are cooked at 160°C to 190°C under pressure for several hours in a concentrated liquor of sodium hydroxide and sodium sulfide to separate cellulose fibers and increase cellulose content by extracting the majority of unwanted lignin. The output of this step is referred to as "brownstock".

[0695] This process of the invention comprises a "Bleaching Step" - a multistage process by which residual lignin and other chromophores are removed to whiten the pulp to target brightness in preparation for making paper or other products. Pulp is treated with oxidizing chemicals, for example chlorine and chlorine dioxide, that attack lignin preferentially. In one aspect the process comprises a bleaching sequence where pulp is reacted with chlorine dioxide, the "Do" stage (see also Figure 5); extracted with alkali in the presence of hydrogen peroxide, the "Ep" stage (see also Figure 5, the "Ep" stage); reacted with chlorine dioxide a second time, a "D1" stage; extracted with alkali and hydrogen peroxide, an Ep stage; and, reacted with chlorine dioxide a third time, a D2 stage. In practicing this process, bleaching can be subject to many variations with respect to type and quantity of oxidizing chemicals used and the number of process steps (however, chlorine dioxide is currently the most widely used chemical oxidant). In one aspect, this process comprises pretreatment of cooked pulp with oxygen under pressure; the oxygen reactor can be at high pressure - at about 200 to 230°F and pH 12 to 14 (this is a common first step in bleaching, known as "oxygen delignification").

[0696] In one aspect, this process comprises refining. For example, prior to papermaking bleached pulp is mechanically fined to collapse the cellulose fibers into flat ribbons, fibrilate their surfaces and improve their physical characteristics for papermaking. At any stage of the process following pulping, the pulp may be dewatered, washed and adjusted to a predetermined consistency by the addition of clean water or recycled streams.

[0697] Xylanase (e.g., an enzyme of the invention) can be just added after pulping, in the oxygen reactor or in the storage container just before the oxygen reactor. Xylanase (e.g., an enzyme of the invention) can

be added at multiple points (one or more or all points) in the bleaching process. In one aspect, a laccase is added to catalyze break-down of lignin. The laccase may be added at any stage of the process, including in the oxygen reactor. Pulp may release various components that self-mediate the laccase. Alternatively, in one aspect, organic or inorganic mediators can be added (see, e.g., DE 19723890 describing an oxidation system comprising an organic mediator and a laccase; alternative exemplary mediators include 2,2'-azinobis(3-ethylbenzthiazoline-5-sulphonate) (ABTS) as an exemplary organic mediator and potassium octacyanomolybdate $[K_4Mo(CN)_8]$ as an exemplary inorganic mediator). Mediators as described in U.S. patent application no. 20030096394, can also be used in the processes of the invention, including any compound capable of enhancing the activities of laccase and laccase-related enzymes.

[0698] In one aspect, an esterase, e.g. lipase, or oxidoreductase, e.g. peroxidase is added. In addition, pH and/or temperature can be modified in the reactor. In monitoring reactions of the invention, any lignin content-measuring technique can be used, e.g., see U.S. Patent Application No. 20020144795, describing a method to measure kappa number or lignin content of kraft pulps based on the voltammetric measurement of catalytic reactions involving lignin and redox mediators.

[0699] Enzymes of the invention can also be used in with alkali-oxygen bleaching (oxygen delignification) processes as described, e.g., in U.S. Patent No. 6,824,646, the process comprising bleaching lignocellulose pulp in aqueous alkali solution with oxygen and treating the pulp with a hemicellulase, while a liquid fraction delivered from the enzyme treatment step is separated from the hemicellulase treated reaction mixture, and subjected to a penetration treatment through a separation membrane, for example, reverse osmosis membrane, to separate a permeated fraction from a non-permeated fraction; and then the permeated fraction is fed to the alkali-oxygen bleaching (oxygen delignification) step comprising use of an enzyme of the invention.

[0700] In alternative aspects of this or any other process (method) of the invention xylanases (e.g., enzymes of the invention) are used to reduce bleaching chemicals, e.g., chlorine, chlorine dioxide, caustic, peroxide, or any combination thereof; and in alternative aspects, a reduction of up to about 1%, 5%, 10%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95% or more, or 100%, of chemicals can be seen in practicing the methods and using the enzymes of the invention. In one aspect, a 100% reduction in chemicals can be achieved when the xylanase is used in combination with a laccase or other enzyme, e.g., by use of enzyme cocktails; noting the the invention provides enzyme mixtures, or "cocktails" comprising at least one enzyme of the invention and one or more other enzyme(s), which can be another xylanase, or any other enzyme.

[0701] In one aspect xylanases of the invention are used to reduce chlorine dioxide to allow recycling of water in the process; thus, there is less water used and less water dumped into the sewer. In one aspect xylanases of the invention are used to allow more lignin-rich pulp to enter the bleaching plant, allowing for better pulp yield and better quality pulp (i.e., less destruction during the cooking process). In one aspect, xylanases of the invention are used to increase the overall brightness of the paper. In one aspect, xylanases of the invention are used to lower the kappa number of the pulp.

[0702] Xylanases of the invention can be used, and the processes of the invention can be practiced, on all wood types, including, for example, on hard wood with, e.g., oxygen delignification, hard wood without oxygen delignification, soft wood with oxygen delignification and soft wood without oxygen delignification, and the like. Xylanases of the invention can be used, and the processes of the invention can be practiced for processing of recycled paper and/or pulp.

[0703] Oxygen delignification typically requires the addition of a reaction tower between a brownstock

washer and a bleach plant. Typically, oxygen and sodium hydroxide are added to brownstock. Reduction of bleaching chemistry by 50% can be achieved in the bleaching process if preceded by oxygen delignification. Washing follows oxygen delignification; effluent can be recovered or discharged. Ozone delignification can be used in place of oxygen delignification.

Example 6: Novel Biobleaching Assay

[0704] This example describes assays that can demonstrate xylanase activity in polypeptides of the invention.

[0705] These xylanase activity studies can be based on those described by Nelson (1944) J. Biol. Chem. 153:375-380, "Reducing Sugar Assay for Xylanase"; and, Somogyi (1952) J. Biol. Chem. 195:19-23. This "Nelson- Somogyi" assay can be used to determine units of activity; data from "Nelson- Somogyi" assays demonstrating xylanase activity in polypeptides of the invention by determining units of activity is set forth, below.

[0706] Enzyme unit determinations also can be determined using the Nelson-Somogyi assay. Biobleaching assays can be based on methods from TAPPI ((Technical Association of Pulp and Paper Industries, see above). Below a description along with references to the TAPPI protocols.

[0707] *Pulp*: In one aspect, two batches of southern softwood Kraft brownstock are obtained, e.g., from the Department of Wood and Fiber Science at North Carolina State University (Raleigh, NC). The pulp Kappa Numbers can be determined, e.g., typically are or are between 21.4 or 29.7, as analyzed using TAPPI method T-236 om-99; see e.g., TAPPI Test Methods (2000-2001, 2003 173).

[0708] *Pulp biobleaching*: Pulp can be pretreated with xylanase and bleached in 10g batches in sealed plastic bags using a 3-stage xylanase/chlorine dioxide/alkaline peroxide sequence: (X)DoEp (see explanation above). The treatment conditions at the three stages can be:

X stage: 10% (w/v) consistency at 65°C and pH 8 for 60 min.

[0709] Do stage: 4% (w/v) consistency at 60 °C for 30 min; a Kappa Factor of 0.18 was used for enzyme treated samples, and 0.18 and 0.21 for no-enzyme control samples. The concentration of chlorine dioxide used during the Do stage was calculated using equation (1):

$$ClO_2\% = \frac{KF \times K\#}{2.63} \quad (1)$$

[0710] Where $ClO_2\%$ is equal to g pure chlorine dioxide per 100 g oven-dried (OD) pulp

[0711] KF is the Kappa Factor and K# is the Kappa Number of the pulp as determined by TAPPI method T-236 om-99, TAPPI Test Methods (2000-2001, 2003 173), Ep stage: 10% (w/v) consistency at 75 °C for 90 min; caustic charge is 1.7% on pulp (w/w) and H_2O_2 charge is 0.5% on pulp (w/w).

[0712] At each stage, replicate bags can be incubated in a water bath at the desired temperature and then removed and kneaded thoroughly every 10 min to ensure uniform mass and heat transfer within the pulp mass. After each stage, pulp can be filtered, e.g., through a Buchner funnel lined with a hard polypropylene filter (297-micron mesh, Spectrum Labs, Ft. Lauderdale, FL). The filtrate can be recycled once to catch the fines, and the pulp cake can be washed, e.g., with 2 L of DI water. The pulp cake can

then be resuspended, e.g., in 1.5 L of DI water and pH can be adjusted, e.g., to pH 8 and pH 4 prior to X and Do stages, respectively. The moisture content of the pulp can be measured using a Mettler-Toledo moisture analyzer (Fisher Scientific, USA).

[0713] Handsheets can be made from the bleached pulp using TAPPI standard equipment (Kalamazoo Paper Chemicals, Richland, MI) according to TAPPI method T-272 sp-97, TAPPI Test Methods (2000-2001, 2003 173). The GE% brightness of each handsheet TAPPI Test Methods (2000-2001, 2003 173) can be measured, e.g., using a Technidyne BRIGHTMETER MICRO S-5/BC™ (Technidyne Corp., New Albany, IN) according to TAPPI method T-452 om-98.

COMPONENTS used in assay	(1)
• 1M NaOH	• Solution 1 : 12g K ⁺ /Na ⁺ tartrate; 24g Na ₂ CO ₃ ; 16g NaHCO ₃ ; 144g Na ₂ SO ₄ in 800 mL H ₂ O
• 0.5 M Sodium phosphate buffer pH 8	
• 1% Arabinoxylan - (Megazyme #P-WAXYM) prepared according to the manufacturer's instructions	
• Xylose - prepare standards 0.15 mM-2mM using D-xylose dissolved in H ₂ O	• Solution 2 : 4g CuSO ₄ *5H ₂ O; 36 g Na ₂ SO ₄ in 200 mL H ₂ O
• 96 well PCR plate (Fisher 05 500-48)	• Reagent A : Mix 4 volumes of solution 1 with 1 volume of solution 2. Note-make fresh daily
• PCR plate seals	
• Standard 96 well clear plates	
• 1 mL tubes (E&K 671511-RC) for the 96 well block	• Reagent B : 25g (NH ₄) ₂ MoO ₄ in 450 mL H ₂ O; add 21 mL conc. H ₂ SO ₄ , mix. Dissolve 3g Na ₂ HAsO ₄ *7H ₂ O in 25 mL dH ₂ O; mix with ammonium molybdate solution and incubate reagent at 37°C for 24-48 h. Store solution in a dark bottle i.e. away from light at room temperature.

Procedure

[0714]

1. 1. Prepare reagent A
2. 2. Pipet 5 uL of 1 M NaOH into each well of a 96 well PCR plate. Keep plate on ice.
3. 3. Prepare reaction mixture. Alternatively, you can make a master mix for multiple samples. Here is the IX mix. Add to the 1 mL tubes and place into the 96 well block.
 1. a. 50 uL pH8 Na-phosphate buffer
 2. b. 250 uL of 1% substrate (to make a final concentration of 0.5%)
 3. c. 150 uL H₂O
4. 4. Preheat reaction mixture to desired temperature for 3 minutes.
5. 5. Dilute the 0.5 M phosphate buffer to 5 mM pH 8 and make enzyme dilutions using this buffer.
6. 6. Pipet 75 uL of diluted enzyme into a well of a 96 well microtiter plate
7. 7. Pipet 50 uL of diluted enzyme into the 1 mL tube containing the reaction mix.
8. 8. At the desired timepoint, pipet 50 uL from each reaction mixture into tubes containing the NaOH (the NaOH will raise the pH to 12, quenching the reaction).
9. 9. Add 50 uL of each standard to separate tubes also containing NaOH. Standards are linear within the range of 0.25 mM xylose to 2.0 mM. Use at least 4 standards to generate the standard curve.

10. 10. Add 50 μ L of Reagent A to each well. Seal plate using the Microseal™ 'A' Film.
11. 11. Heat the plate for 20 min. at 100°C in a PCR machine. Set the machine to cool down to 4°C after heating the samples.
12. 12. Add 50 μ L of reagent B to each tube, mix.
13. 13. -note a significant amount of CO₂ is formed after addition of reagent B. Care should be taken so sample does not contaminate adjacent wells.
14. 14. Pipet 100 μ L of each sample or standard into separate wells of a 96 well microtiter plate.
15. 15. Read plate at 560 nm.
16. 16. Plot standard curve data and express standards as umoles of xylose i.e. 50 μ L of 2.5 mM xylose is .125 μ moles of xylose.
17. 17. Subtract buffer control from sample data for each timepoint and plot the data
18. 18. Divide timepoint curve slope value by the xylose standard curve slope value
19. 19. Multiply by 10 (accounts for the 50 μ L samples (1/10 of the total assay volume)
20. 20. Divide by the volume used in the assay (0.05) to get μ moles of xylose released per min per mL of enzyme or U/mL of enzyme.
21. 21. Divide this number by the protein concentration to get U/mg.

[0715] "Units of Activity" data from the "Nelson- Somogyi" assays can be used to determine dosing in biobleaching assays (based on TAPPI methods).

[0716] As noted above, the enzymes and processes of the invention can also be used in conjunction with a second approach to enzymatic bleaching using oxidative enzymes such as laccase and/or manganese peroxidase (MnP) to delignify pulp. In one aspect of this second approach, of these enzymes, laccase is preferred, because MnP requires hydrogen peroxide, manganese (II) ions and a chelator. Laccase can cause delignification of pulp under slight oxygen pressure, but is considerably more effective when mediators are added, as discussed above.

[0717] Catalyst improved delignification methods can also be used in conjunction with the methods of the invention, for example, polysulfide or anthraquinone. Anthraquinone is a pulping reaction catalyst which can increase the speed of pulping, increase yield, and reduce pulping chemical usage by up to 10%. It is possible to use both anthraquinone and polysulfide together.

[0718] In one aspect, laccase is used in conjunction with the methods of the invention, as discussed above. For example, laccase is used in an oxygen reactor in a process of the invention, where the laccase breaks down the lignin in the oxygen reactor. While pulp may release various components that self-mediate the laccase, in one aspect organic or inorganic mediators are added (see discussion above, e.g., alternative exemplary mediators include 2,2'-azinobis(3-ethylbenzthiazoline-5-sulphonate) (ABTS) as an exemplary organic mediator and potassium octacyanomolybdate [K₄Mo(CN)₈] as an exemplary inorganic mediator, or mediators as described in U.S. patent application no. 20030096394). In one aspect, another hydrolase, such as an esterase (e.g., a lipase) and/or an oxidoreductase (e.g., a peroxidase) is also added. In alternative aspects, pH and/or temperature are modified in the reactor.

EXAMPLE 7: Studies demonstrating the enzymatic activity of enzymes of the invention

[0719] This example describes studies demonstrating the enzymatic activity of the exemplary xylanase enzymes of the invention, which demonstrates that polypeptides of this invention have xylanase activity.

[0720] An exemplary assay for evaluating these xylanases:

1. 1. Initial Screen - using an azo-xylan (solution-based) substrate
 1. a. Enzymatic activity of enzymes can be determined by an azo-xylan assay using MEGAZYME® substrate Birchwood Azo-xylan in 100 mM sodium phosphate, pH 8, according to manufacturer's recommended assay protocol. The concentrations of enzyme samples can be adjusted such that they had equal amounts of xylanase activity at pH8.
 2. b. The azo-xylan assay are then repeated with normalized samples in 100 mM sodium borate buffer at pH 10.4.
2. 2. Initial Screen - ENZ-CHEK ULTRA XYLANASE ASSAY KIT™ (Invitrogen)
 1. a. Xylanase enzyme samples can be prepared in the same manner as for the azo-xylan assay (section 1, above).
 2. b. The level of enzymatic activity of enzymes can be measured by employing commercially available assay kit, e.g., sold by Invitrogen under the name ENZ-CHEK ULTRA XYLANASE ASSAY KIT™ (Product number E33650). The ENZ-CHEK™ kit substrate produces fluorescent signal in the presence of xylanases, which can be used to quantify xylanase activities using kit-supplied standards. The protocol used for testing xylanase enzymes can be slightly modified from any manufacturer-recommended protocol. The modifications can primarily involve, e.g., testing xylanases at different pH and temperature that what is recommended by the manufacturer.
3. 3. Secondary Screen - Exemplary Pulp Assays
 1. a. The enzymes from azo-xylan assay can be tested for activity on wheat arabinoxylan using, e.g., a Nelson-Somogyi assay as already described herein. They can be then tested in a laboratory scale bleaching assays to determine the amount of chemical savings each can achieved for a given pulp type and chlorine dioxide loading. The ones that meet desired performance characteristics can be tested in TAPPI bag biobleaching assay (e.g., in triplicate) at a range of loadings and pH levels.
4. 4. Exemplary enzyme characterization screen - Temperature profile
 1. a. Thermotolerance of xylanases can be assayed using azo-xylan assay at pH 8 and pH 10.4 at progressively more elevated temperatures; and enzymes of the invention were tested using this assay. The initial rates of reaction at each temperature can be recorded and plotted to determine optimal performance temperature of xylanases.
 2. b. Residual activity - Another exemplary assay that can be employed for testing thermostability of enzymes is the residual activity method, whereby a sample of enzyme is treated at an elevated temperature at a particular pH for a specific period of time, and then assayed under standard conditions under permissive temperature (typically 37°C). A half-life at a particular temperature is then determined and provides a measure of a given enzyme fitness under those temperature conditions.

EXAMPLE 8: Studies demonstrating the enzymatic activity of enzymes of the invention

[0721] This example describes studies demonstrating the enzymatic activity of the exemplary xylanase enzymes of the invention, including the enzymatic activity of any polypeptide of this invention have xylanase activity.

[0722] The evolution of endoxylanase SEQ ID NO:2 (Xyl 11) utilizing GSSM technology and xylanase

screening identified point mutations (Xyl 11 mutants) having increased xylanase activity, as well increased sugar release from alkaline pretreated corn stover, when used in combination with 7 other cellulosic enzymes (Table 2, below) after 36 hrs in saccharification cocktail assays at 50°C. These assays contain alkaline pretreated dry corn cobs at 5% (w/v) with a total enzyme loading of 10.2 mg / g cellulose in the solids.

Table 2: Composition of enzyme cocktail

Enzyme	SEQ ID NOs:	Conc. mg/g cellulose
Endoglucanase [^]	SEQ ID NO:4 (encoded by SEQ ID NO:3)	1.7
Oligomerase I (beta-glucosidase) [^]	SEQ ID NO:6 (encoded by SEQ ID NO:5)	0.5
CBH1 (GH family 7) [^]	SEQ ID NO:8 (encoded by SEQ ID NO:7)	5
CBH2 (GH family 6) [^]	SEQ ID NO: 10 (encoded by SEQ ID NO:9)	1
Xylanase (GH family 11)	VARIES (control*, Xyl 11 or Xyl 11 mutants)	0.6
Arabinofuranosidase [^]	SEQ ID NO:14 (encoded by SEQ ID NO:13)	0.25
Xylanase (GH family 10) [^]	SEQ ID NO:16 (encoded by SEQ ID NO:15)	0.15
Oligomerase II (beta-xylosidase) [^]	SEQ ID NO:18 (encoded by SEQ ID NO:17)	1
*control xylanase is SEQ ID NO:12 (encoded by SEQ ID NO:11)		
[^] previously described in PCT Publication No. WO 07/094852		

[0723] The new xylanase mutants improved xylose release over the wild type at 0.6 mg / g cellulose as well as 0.2 mg / g cellulose loading (Figure 2). At the standard loading of 0.6 mg / g cellulose these new variants achieved conversion rates of up to 90% monomeric xylose released vs. 63% with the wild type. Some of the polypeptides of the invention (the mutants of SEQ ID NO:2), in particular, Xyl 11 mutant 11 and Xyl 11 mutant 14, also achieved greater than 90% xylose release even at the reduced loading of 0.2 mg cellulose. These novel polypeptides of the invention (the mutants of SEQ ID NO:2) therefore not only improve the rate of xylose release but also can do so at a reduced enzyme loading. Similar positive effects on xylose release and enzyme loading could be envisioned for comparable saccharification reactions using different feed stocks (switch grass, hard and soft woods, energy cane, bagasse etc.) applied to alkaline or acidic pretreatments and with different initial enzyme loadings (1mg - 100 mg / g cellulose) and different ratios of cocktail components.

[0724] The enzymes of this invention can be used to process/ treat cellulosic material for, e.g., biological alcohol (e.g., EtOH, or ethanol) fermentation; cellulosic material that is processed using compositions and methods of the invention can be mainly composed of cellulose (containing glucose), and hemicellulose - which is mostly containing xylose. In one aspect, glucose as well as xylose can be used as a sugar source for EtOH fermentation. In one aspect, xylanases of the invention are active in the enzymatic breakdown of the hemicellulose portion of cellulosic material, releasing a monomeric xylose. In one aspect, the improved xylanase activity of polypeptides of the invention increases the amount of xylose available for fermentation.

[0725] In one aspect, by removing the hemicellulose the cellulose becomes more accessible to cellulases, which can also increase the conversion of cellulose to glucose. Using xylanases of the invention, e.g., the sequence variations of the exemplary endoxylanase Xyl 11 (SEQ ID NO:2), including the exemplary 18 amino acid substitutions described herein, an increased specific activity can be achieved over the "wild type" xylanase, as described in Table 1, above. Note: in Table 1 tertiary assay activity is indicated as the Absorbance at 560 nm measured in the BCA assay reached after 9.5 h of hydrolysis. Referencing Table 1, when each of these clones (xylanases of the invention) was evaluated in cocktail saccharification assays with the xylanase as the variable, fourteen of these clones (xylanases of the invention) improved xylose conversion rates when compared to assays with the wild type at the same loading, as noted in Table 3 (see Table 1 for the sequence referenced in Table 3, e.g., Table 1 sets for the sequence of Xyl 11 mutant 5, Xyl 11 mutant 5, etc., based on the exemplary SEQ ID NO:2; note also, "Xyl 11 (WT)" refers to the "wild type" exemplary SEQ ID NO:2):

Table 3: Xylose conversion by the cocktail shown in Table 2 (above). Note the xylanase component (Xyl 11 WT or Xyl 11 mutant) varies in each cocktail.

Xylose Conversion @ 0.2mg			Xylose Conversion @ 0.6mg		
Xyl 11 or Xyl 11 mutant used in cocktail	36hr	STDEV	Xyl 11 or Xyl 11 mutant used in cocktail	36hr	STDEV
Xyl 11 (WT)	51.57%	0.01	Xyl 11 WT)	62.84%	0.02
Xyl 11 mutant 5	55.98%	0.00	Xyl 11 mutant 16	68.10%	0.01
Xyl 11 mutant 16	57.43%	0.01	Xyl 11 mutant 5	69.69%	0.00
Xyl 11 mutant 12	59.03%	0.02	Xyl 11 mutant 12	71.75%	0.01
Xyl 11 mutant 4	59.46%	0.00	Xyl 11 mutant 7	74.25%	0.00
Xyl 11 mutant 9	60.34%	0.01	Xyl 11 mutant 9	74.45%	0.00
Xyl 11 mutant 17	60.45%	0.01	Xyl 11 mutant 17	74.76%	0.01
Xyl 11 mutant 7	61.23%	0.02	Xyl 11 mutant 4	74.94%	0.01
Xyl 11 mutant 2	61.73%	0.02	Xyl 11 mutant 13	75.30%	0.01
Xyl 11 mutant 6	62.31%	0.00	Xyl 11 mutant 2	78.24%	0.05
Xyl 11 mutant 13	63.58%	0.01	Xyl 11 mutant 15	80.03%	0.00
Xyl 11 mutant 15	65.95%	0.00	Xyl 11 mutant 6	80.36%	0.09
Xyl 11 mutant 10	66.34%	0.00	Xyl 11 mutant 10	80.61%	0.01
Xyl 11 mutant 11	71.76%	0.01	Xyl 11 mutant 14	84.74%	0.01
Xyl 11 mutant 14	73.69%	0.03	Xyl 11 mutant 11	90.36%	0.06

[0726] Accordingly, the invention provides an enzyme cocktail comprising, or consisting of, the enzymes: Endoglucanase, Oligomerase I (beta-glucosidase), CBH1 (GH family 7), CBH2 (GH family 6), Xylanase (GH family 11), Arabinofuranosidase, Xylanase (GH family10) and an Oligomerase II (beta-xylosidase); wherein one, two, three, four, five, six, seven and/or all eight of these enzyme are a polypeptide of this invention, and methods for treating polysaccharide compositions using these cocktails, or any cocktail of this invention, for, e.g., treating/ processing wood, pulp, paper, waste(s) and the like, or making biofuels or foods or feeds, or any other industrial process or method, e.g., as described herein.

Screens and assays for identifying enzymes of the invention

[0727] The following screens and assays were used in identifying exemplary enzymes of the invention, and in one aspect, these screens and assays can be applied to determine if any polypeptide has sufficient xylanase activity to fall within the scope of this invention - assuming of course it also has the requisite sequence identity, as described herein:

[0728] Xylanase evolution screen: Utilizing the GSSM technology (Verenium Corporation, US Patent No. 6,171,820) an evolution library for endoxylanase Xyl 11 (SEQ ID NO:2) representing all possible amino acid exchanges for each of the 194 residues of this enzyme was created. Point mutations were introduced using degenerate oligonucleotides, one amino acid position at a time, so that each original codon could be substituted with each of the 20 naturally encoded amino acids. The mutated variants were transformed into XL1-Blue (recA- strain, Stratagene) and then into *Pseudomonas fluorescens* MB214 (Dow Global Technologies Inc., US Patent Publication No. 20050130160), using vector pWZ82T (SEQ ID NO:25). All variants were grown and expressed (from *Pseudomonas fluorescens* MB214) and lysed in 96 well plates. Hydrolysis reactions with the lysates were carried out in 96 well plates (200 μ l of 200mM citrate buffer, pH 5.5, 0.5% dried and milled alkaline pretreated corn stover - CP-15, 50C). Aliquots were removed from the reaction at 1, 3, 5 and 10hrs and added to 800mM carbonate buffer pH 10 to stop the reaction. The extent of hydrolysis at each time point was evaluated via a reducing ends assay (BCA), as described by Johnston et al. 1998 (see below), recording absorption at 560nm (A560). In addition a quantitative ELISA utilizing Xyl 11 (SEQ ID NO:2) specific antibodies was used to normalize activity to protein expression. Both functional and quantitative assays were automated for high through put. In the primary screen, clones exhibiting normalized activity exceeding Xyl 11 (SEQ ID NO:2) controls on the plate by at least 2 standard deviations ($> 1.0 + 2 \text{ STDV wt}$) were moved on to a secondary screen. In the secondary screen, all primary hits were re-screened in duplicate applying the same assay and hit criteria as in the primary screen. Clones that confirmed for both duplicates were then moved on to a tertiary screen. In tertiary screens, these clones again were assayed in duplicate using the BCA assay, but this time with different defined concentrations of protein (0.1, 0.05 and 0.025 mg/ml). Total protein of lysates was determined via Bradford assays (e.g., as described in Bradford 1976, see below) the relative content of xylanase then was determined via densitometry of SDS PAGE gels after running defined amounts of total protein. All clones exceeding wt activity, recorded as absorption at 560 nm (A560), for at least one enzyme concentration in the tertiary screen were then assayed in saccharification assays.

[0729] Saccharification / cocktail assay: Cocktail reactions were set up in capped 10 ml glass vials containing two metal ball bearings. The reaction volume was 5 ml (200 mM Sodium Citrate- 1 mM Sodium Azide pH 5.5) with 5% solids (size 40 grit milled alkaline pretreated corn stover). Enzyme composition and loadings were according to Table 2, above, only varying the family 11 endoxylanase. Reaction vials were incubated for 36h at 50C under shaking at 300 rpm. The concentration of xylose monomers released was determined by HPLC (RI detector, Shodex SP-0810 column, flow rate of 0.5 ml/min) using a set of standards and calibration curves.

- Johnston, D.B.; Shoemaker, S.P.; Smith, G.M. and Whitaker, J.R.: Kinetic Measurement of Cellulase Activity on Insoluble Substrates Using Disodium 2,2' Bicinchoninate. *Journal of Food Biochemistry* (22) Issue 4 pp. 301-319, 1998
- Bradford, M. M. (1976) A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Protein Utilizing the Principle of Protein-Dye Binding. *Anal. Biochem.* 72:248-25

SEQUENCE LISTING

[0730]

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<141> 2013-08-23

<150> 60/977,348

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Leu Leu Glu Gly Gly Arg Tyr Thr Ser Asn Trp Ser Gly Ile Asn Asn
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Trp Val Gly Gly Lys Gly Trp Gln Thr Gly Ser Arg Arg Asn Ile Thr
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Asn Val Thr Trp Pro Lys Gly Ser Thr Asp Gly Ser Pro Gln Thr Arg
740 745 750

Ile Ala Ala Gly Gly Ala Pro Gly Gly Asn Pro Gln Leu Trp Asp Val
755 760 765

Leu Phe Lys Val Glu Ala Thr Ile Thr Asn Thr Gly His Val Ala Gly
770 775 780

Asp Glu Val Ala Gln Ala Tyr Ile Ser Leu Gly Gly Pro Asn Asp Pro
785 790 795 800

Lys Val Leu Leu Arg Asp Phe Asp Arg Leu Thr Ile Lys Pro Gly Glu
805 810 815

Ser Ala Val Phe Thr Ala Asn Ile Thr Arg Arg Asp Val Ser Asn Trp
820 825 830

Asp Thr Val Ser Gln Asn Trp Val Ile Thr Glu Tyr Pro Lys Thr Ile
835 840 845

His Val Gly Ala Ser Ser Arg Asn Leu Pro Leu Ser Ala Pro Leu Asp
850 855 860

Thr Ser Ser Phe Arg
865

<210> 7

<211> 1527

<212> DNA

<213> Unknown

<220>

<223> Obtained from environmental sample

<400> 7

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tgcagcaacg tccgcggatc tgtcgtggtt gacgccaaact ggcgatggac ccatagcacc	180
tccagcagca ccaactgcta caccggcaac acctgggaca agactctctg ccccgatgga	240
aagacctgcg ctgacaagtg ctgtcttgat ggtgccgact actctggcac ctacggagtc	300
acctcgagcg gcaaccagct caacctcaag ttgttgactg ttggaccata cagcaccaat	360
gttggcagcc gtctctacct catggaggat gagaacaact accagatgtt cgacctcctg	420
ggcaacgaat tcacctttga tgtcgtatgc aacaacatcg gatgcggcct gaacggcgcc	480
ctctacttcg tctccatgga caaggatggt ggcaagagcc gcttcagcac caacaaggct	540
ggtgccaaat acggaactgg ctactgcgat gccagtgcc ctccgatgt caagttcatc	600
aacggagttg ccaactccga cgactggcag ccctccgcca gcgacaagaa cgccggtgtt	660
ggcaagtacg gcacctgctg ccctgagatg gatctctggg aggcacaaca gatctccacg	720
gcttacactc cccatccctg caagagcctc acccagcagt cctgcgaggg cgatgcctgc	780
ggtggcacct actcttctac tcgctatgct ggaacttgcg atcccgatgg ttgcgatttc	840
aacctttacc gccagggcaa ccacaccttc tacggtcccg gctccggctt caacgttgat	900
accaccaaga aggtgactgt cgtgaccag ttcatcaagg gcagcgacgg caagctctct	960
gagatcaagc gtctctatgt tcagaacggc aaggtcattg gcaaccccca gtccgagatt	1020

```

gccacaacc ccggcagctc cgtcaccgac agcttctgca aggccagaa ggttgcatc 1080
aacgaccccg atgacttcaa caagaagggt ggctggagcg gcatgaacga cgccctcgcc 1140
aagcccatgg ttctcgtcat gagcctgtgg caccgacct acgccaacat gctctggctc 1200
gactctacct accccaaggg ctccaagact cccggctctg ctctgggctc ttgccctgag 1260
gactctgggtg tccccgccac tctcgagaag gaggtcccca actccagcgt cagcttctcc 1320
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cccgaggagc ctgaggagcc cgaggagcct gtcggcaccg tccccagtg gggccagtg 1440
ggcggcatca actacagcgg cccaccgcc tgcgtgtctc cctacaagtg caacaagatc 1500
aacgactact actcccagtg ctactag 1527

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<210> 8

<211> 508

<212> PRT

<213> Unknown

<220>

<223> Obtained from environmental sample

<220>

<221> SIGNAL

<222> (1) ... (20)

<220>

<221> DOMAIN

<222> (19) ... (453)

<223> Glycosyl hydrolase family 7

<220>

<221> DOMAIN

<222> (476) ... (504)

<223> Fungal cellulose binding domain

<400> 8

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Met Tyr Arg Ile Leu Ala Thr Ala Ser Ala Leu Leu Ala Thr Ala Arg
1           5           10          15

```

```

Ala Gln Gln Ala Cys Thr Leu Asn Ala Glu Ser Lys Pro Ala Leu Thr
20          25          30

```

```

Trp Ser Lys Cys Thr Ser Ser Gly Cys Ser Asn Val Arg Gly Ser Val
35          40          45

```

```

Val Val Asp Ala Asn Trp Arg Trp Thr His Ser Thr Ser Ser Ser Thr
50          55          60

```

```

Asn Cys Tyr Thr Gly Asn Thr Trp Asp Lys Thr Leu Cys Pro Asp Gly
65          70          75          80

```

```

Lys Thr Cys Ala Asp Lys Cys Cys Leu Asp Gly Ala Asp Tyr Ser Gly
85          90          95

```

```

Thr Tyr Gly Val Thr Ser Ser Gly Asn Gln Leu Asn Leu Lys Phe Val
100         105         110

```

```

Thr Val Glv Pro Trp Ser Thr Asn Val Glv Ser Arg Leu Trp Leu Met

```

```

115      120      125

Glu Asp Glu Asn Asn Tyr Gln Met Phe Asp Leu Leu Gly Asn Glu Phe
130      135      140

Thr Phe Asp Val Asp Val Asn Asn Ile Gly Cys Gly Leu Asn Gly Ala
145      150      155      160

Leu Tyr Phe Val Ser Met Asp Lys Asp Gly Gly Lys Ser Arg Phe Ser
165      170      175

Thr Asn Lys Ala Gly Ala Lys Tyr Gly Thr Gly Tyr Cys Asp Ala Gln
180      185      190

Cys Pro Arg Asp Val Lys Phe Ile Asn Gly Val Ala Asn Ser Asp Asp
195      200      205

Trp Gln Pro Ser Ala Ser Asp Lys Asn Ala Gly Val Gly Lys Tyr Gly
210      215      220

Thr Cys Cys Pro Glu Met Asp Ile Trp Glu Ala Asn Lys Ile Ser Thr
225      230      235      240

Ala Tyr Thr Pro His Pro Cys Lys Ser Leu Thr Gln Gln Ser Cys Glu
245      250      255

Gly Asp Ala Cys Gly Gly Thr Tyr Ser Ser Thr Arg Tyr Ala Gly Thr
260      265      270

Cys Asp Pro Asp Gly Cys Asp Phe Asn Pro Tyr Arg Gln Gly Asn His
275      280      285

Thr Phe Tyr Gly Pro Gly Ser Gly Phe Asn Val Asp Thr Thr Lys Lys
290      295      300

Val Thr Val Val Thr Gln Phe Ile Lys Gly Ser Asp Gly Lys Leu Ser
305      310      315

Glu Ile Lys Arg Leu Tyr Val Gln Asn Gly Lys Val Ile Gly Asn Pro
325      330      335

Gln Ser Glu Ile Ala Asn Asn Pro Gly Ser Ser Val Thr Asp Ser Phe
340      345      350

Cys Lys Ala Gln Lys Val Ala Phe Asn Asp Pro Asp Asp Phe Asn Lys
355      360      365

Lys Gly Gly Trp Ser Gly Met Asn Asp Ala Leu Ala Lys Pro Met Val
370      375      380

Leu Val Met Ser Leu Trp His Asp His Tyr Ala Asn Met Leu Trp Leu
385      390      395      400

Asp Ser Thr Tyr Pro Lys Gly Ser Lys Thr Pro Gly Ser Ala Arg Gly
405      410      415

Ser Cys Pro Glu Asp Ser Gly Val Pro Ala Thr Leu Glu Lys Glu Val
420      425      430

Pro Asn Ser Ser Val Ser Phe Ser Asn Ile Lys Phe Gly Pro Ile Gly
435      440      445

Ser Thr Tyr Ser Gly Thr Gly Gly Asn Asn Pro Asp Pro Glu Glu Pro

```

450 455 460
 Glu Glu Pro Glu Glu Pro Val Gly Thr Val Pro Gln Trp Gly Gln Cys
 465 470 475 480
 Gly Gly Ile Asn Tyr Ser Gly Pro Thr Ala Cys Val Ser Pro Tyr Lys
 485 490 495
 Cys Asn Lys Ile Asn Asp Tyr Tyr Ser Gln Cys Tyr
 500 505

<210> 9

<211> 1413

<212> DNA

<213> Unknown

<220>

<223> Obtained from an environmental sample

<400> 9

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accctctatg gacaatgtgg tggtcagggc tactccggac tcaccagctg cgtggcggga      120
gcaacatgct ccaccgtaaa tgaatactac gctcagtgtg cgcacgacgc aggcagcgcc      180
acttccacca ccttgaagac aactacgacc accgctgggg cgacgacgac gacgactagc      240
aagacttctg ctteccagac gtctactact aaaacctcaa ccagtaccgc ctcaacaacc      300
acggctacaa ccacggccag cgcgagcggc aaccctgtca gtgggtacca gctctacgtg      360
aaccctact actcctcga agtggcctcc ctggctatcc catccctcac ggggacactt      420
tcctcgctcc aggtgcgac cacagccgca gccaaagtgc cctctttcgt ctggctggac      480
gtggctgcca aggtgccgac gatggccacc tacctggccg acatcaaagc ccagaatgca      540
gcgggagcca acccccccgt cgcgggccag tttgtgtgtc acgacctccc tgaccgcgac      600
tgccgcgcgc tggccagcaa cggcgagtac tccatcgcca acaacggtgt ggccaactac      660
aaggcctaca tcgactccat ccgcaaggtc ctggtgcagt actcggatgt gcacaccatt      720
ctggtgatcg agcccgacag tctcgccaac ctggtgacca acctcaatgt ggccaaatgt      780
gccaacgctc agagcgccct cctcgaatgc accaactatg ccctggagca gctgaacctc      840
cccaacgtgg ccatgtatct tgatgccgga cagcccggtt ggctcggctg gcccgcgaaac      900
cagcaaccgg ccgccaatct gtacgcgagc gtgtacaaga acgccagctc gcccgccgca      960
gtgcgcggcc tggccacgaa cgtcgccaac tacaacgctt tcaccatcgc ctcggtccccg     1020
tcgtacaccc agggcaacag cgtctgcgac gagcagcagt acatcaacgc gatcgccccg     1080
ctcctgtcag cgcagggtct caacgcccac ttcatcgtcg acaccggccg caacggcaaa     1140
cagcccaccg gccacaagc ctggggcgac tggtgcaacg tcatcaacac ggggttcggc     1200
gtgcgccccg ccaccaaacac gggcgacgcg ctcgctcgacg ccttcgtctg ggtcaagccc     1260
ggcggcgaga gcgacggcac ctccgatagc tcggcgaccc gctacgacgc ccactgcggg     1320
tacagcgatg ccttgacgcc ggcgccggag gcggggacct ggttccaggc ctacttcgta     1380
caattgtctc cgaacgcaaa tccggctttc tag                                     1413

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<210> 10

<211> 470

<212> PRT

<213> Unknown

<220>

<223> Obtained from an environmental sample

<220>

<221> SIGNAL

<222> (1) ... (18)

<220>

<221> DOMAIN

<222> (22) ... (50)

<223> Fungal cellulose binding domain

<220>

<221> DOMAIN

<222> (120) ... (437)

<223> Glycosyl hydrolases family 6

<220>

<221> SITE

<222> (26) ... (53)

<223> CBM1 (carbohydrate binding type-1) domain signature. Prosite id = PS00562

<220>

<221> SITE

<222> (240) ... (249)

<223> Glycosyl hydrolases family 6 signature 2. Prosite id = PS00656

<220>

<221> SITE

<222> (314) ... (317)

<223> N-glycosylation site. Prosite id = PS00001

<400> 10

Met	Arg	Tyr	Thr	Trp	Ser	Val	Ala	Ala	Ala	Leu	Leu	Pro	Cys	Ala	Ile
1			5						10				15		

Gln	Ala	Gln	Gln	Thr	Leu	Tyr	Gly	Gln	Cys	Gly	Gly	Gln	Gly	Tyr	Ser
		20					25					30			

Gly	Leu	Thr	Ser	Cys	Val	Ala	Gly	Ala	Thr	Cys	Ser	Thr	Val	Asn	Glu
	35					40						45			

Tyr	Tyr	Ala	Gln	Cys	Thr	Pro	Ala	Ala	Gly	Ser	Ala	Thr	Ser	Thr	Thr
	50					55					60				

Leu	Lys	Thr	Thr	Thr	Thr	Thr	Ala	Gly	Ala	Thr	Thr	Thr	Thr	Thr	Ser
65					70				75						80

Lys	Thr	Ser	Ala	Ser	Gln	Thr	Ser	Thr	Thr	Lys	Thr	Ser	Thr	Ser	Thr
			85					90						95	

Ala	Ser	Thr	Thr	Thr	Ala	Thr	Thr	Thr	Ala	Ser	Ala	Ser	Gly	Asn	Pro
			100					105						110	

Phe Ser Gly Tyr Gln Leu Tyr Val Asn Pro Tyr Tyr Ser Ser Glu Val
 115 120 125
 Ala Ser Leu Ala Ile Pro Ser Leu Thr Gly Thr Leu Ser Ser Leu Gln
 130 135 140
 Ala Ala Ala Thr Ala Ala Ala Lys Val Pro Ser Phe Val Trp Leu Asp
 145 150 155 160
 Val Ala Ala Lys Val Pro Thr Met Ala Thr Tyr Leu Ala Asp Ile Lys
 165 170 175
 Ala Gln Asn Ala Ala Gly Ala Asn Pro Pro Val Ala Gly Gln Phe Val
 180 185 190
 Val Tyr Asp Leu Pro Asp Arg Asp Cys Ala Ala Leu Ala Ser Asn Gly
 195 200 205
 Glu Tyr Ser Ile Ala Asn Asn Gly Val Ala Asn Tyr Lys Ala Tyr Ile
 210 215 220
 Asp Ser Ile Arg Lys Val Leu Val Gln Tyr Ser Asp Val His Thr Ile
 225 230 235 240
 Leu Val Ile Glu Pro Asp Ser Leu Ala Asn Leu Val Thr Asn Leu Asn
 245 250 255
 Val Ala Lys Cys Ala Asn Ala Gln Ser Ala Tyr Leu Glu Cys Thr Asn
 260 265 270
 Tyr Ala Leu Glu Gln Leu Asn Leu Pro Asn Val Ala Met Tyr Leu Asp
 275 280 285
 Ala Gly His Ala Gly Trp Leu Gly Trp Pro Ala Asn Gln Gln Pro Ala
 290 295 300
 Ala Asn Leu Tyr Ala Ser Val Tyr Lys Asn Ala Ser Ser Pro Ala Ala
 305 310 315 320
 Val Arg Gly Leu Ala Thr Asn Val Ala Asn Tyr Asn Ala Phe Thr Ile
 325 330 335
 Ala Ser Cys Pro Ser Tyr Thr Gln Gly Asn Ser Val Cys Asp Glu Gln
 340 345 350
 Gln Tyr Ile Asn Ala Ile Ala Pro Leu Leu Ser Ala Gln Gly Phe Asn
 355 360 365
 Ala His Phe Ile Val Asp Thr Gly Arg Asn Gly Lys Gln Pro Thr Gly
 370 375 380
 Gln Gln Ala Trp Gly Asp Trp Cys Asn Val Ile Asn Thr Gly Phe Gly
 385 390 395 400
 Val Arg Pro Thr Thr Asn Thr Gly Asp Ala Leu Val Asp Ala Phe Val
 405 410 415
 Trp Val Lys Pro Gly Gly Glu Ser Asp Gly Thr Ser Asp Ser Ser Ala
 420 425 430
 Thr Arg Tyr Asp Ala His Cys Gly Tyr Ser Asp Ala Leu Gln Pro Ala
 435 440 445

Pro Glu Ala Gly Thr Trp Phe Gln Ala Tyr Phe Val Gln Leu Leu Ser
 450 455 460

Asn Ala Asn Pro Ala Phe
 465 470

<210> 11

<211> 594

<212> DNA

<213> Artificial Sequence

<220>

<223> Synthetically generated

<400> 11

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ttctggaagg	acagtccggg	cacggtgaat	ttttgcctgc	agtccggcgg	ccgttacaca	120
tcgaactgga	gcggcatcaa	caactgggtg	ggcggcaagg	gatggcagac	cggttcacgc	180
cggaacatca	cgtactcggg	cagcttcaat	tcaccgggca	acggctacct	ggcgccttac	240
ggatgggacca	ccaatccact	cgtcagtagc	tacgtcgtcg	atagctgggg	gagctggcgt	300
ccgccggggt	cggacggaac	gttcctgggg	acggatcaaca	gcgatggcgg	aacgtatgac	360
atctatcgcg	cgcagcgggt	caacgcgccg	tccatcatcg	gcaacgccac	gttctatcaa	420
tactggagcg	ttcggcagtc	gaagcgggta	ggtgggacga	tcaccaccgg	aaaccacttc	480
gacgcgtggg	ccagcgtggg	cctgaacctg	ggcactcaca	actaccagat	catggcgacc	540
gagggctacc	aaagcagcgg	cagctccgac	atcacggtga	gtgaaggcgg	ttga	594

<210> 12

<211> 197

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthetically generated

<400> 12

Met	Ala	Gln	Thr	Cys	Leu	Thr	Ser	Ser	Gln	Thr	Gly	Thr	Asn	Asn	Gly	
1				5					10					15		
Phe	Tyr	Tyr	Ser	Phe	Trp	Lys	Asp	Ser	Pro	Gly	Thr	Val	Asn	Phe	Cys	
			20					25					30			
Leu	Gln	Ser	Gly	Gly	Arg	Tyr	Thr	Ser	Asn	Trp	Ser	Gly	Ile	Asn	Asn	
		35					40					45				
Trp	Val	Gly	Gly	Lys	Gly	Trp	Gln	Thr	Gly	Ser	Arg	Arg	Asn	Ile	Thr	
	50					55					60					
Tyr	Ser	Gly	Ser	Phe	Asn	Ser	Pro	Gly	Asn	Gly	Tyr	Leu	Ala	Leu	Tyr	
65					70				75					80		
Gly	Trp	Thr	Thr	Asn	Pro	Leu	Val	Glu	Tyr	Tyr	Val	Val	Asp	Ser	Trp	
				85					90					95		
Gly	Ser	Trp	Arg	Pro	Pro	Gly	Ser	Asp	Gly	Thr	Phe	Leu	Gly	Thr	Val	
			100					105					110			
Asn	Ser	Asp	Gly	Gly	Thr	Tyr	Asp	Ile	Tyr	Arg	Ala	Gln	Arg	Val	Asn	
		115					120					125				
Ala	Pro	Ser	Ile	Ile	Gly	Asn	Ala	Thr	Phe	Tyr	Gln	Tyr	Trp	Ser	Val	
		130				135						140				
Arg	Gln	Ser	Lys	Arg	Val	Gly	Gly	Thr	Ile	Thr	Thr	Gly	Asn	His	Phe	
145					150				155					160		
Asp	Ala	Trp	Ala	Ser	Val	Gly	Leu	Asn	Leu	Gly	Thr	His	Asn	Tyr	Gln	
			165						170					175		
Ile	Met	Ala	Thr	Glu	Gly	Tyr	Gln	Ser	Ser	Gly	Ser	Ser	Asp	Ile	Thr	
		180						185					190			
Val	Ser	Glu	Gly	Gly												
		195														

<210> 13

<211> 2637

<212> DNA

<213> Unknown

<220>

<223> Obtained from environmental sample

<400> 13

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cggttggacc aaccgggagc ggtgatcaat cgcaacatct acggccagtt tgcggagcat      180
ctcggacggt tgatctacga cgggctctgg gttggtgaag gatcgtcgat ccggaacacg      240
cgcggattgc gtaacgacgt cgttacggcg ttaaaagaat tgcattgtgc tgtgctgcgt      300
tggcccgcg gctgttttgc cgacgagtat cactggcgtg acggcattgg accacgcgac      360
aagcgtccgc ggcggccgaa cgcgagtttg ggcggcgtcg attcgaatgc gtttggcacg      420
catgagttca tggagctgtg cgagatgttg ggcgcagacg cttatatcaa tggcaacgtc      480
ggcagcggca cgcgcagga gatgatggaa tggatcgagt acatgacttc cgattccgat      540
tcggatctcg ccaacctgcg ccgtcgcaat ggcgcgaca agccgtggaa ggtgccgtat      600

ttcgcctcg gcaatgagac gtggggctgt ggtggaaata tgcggccgga gttttacgcc      660
gacgtgtatc gccagtagc cacttcacg aagaaccatt caggcaatcg cattcagaaa      720
ctcgcgagcg gtggttacga caacaattac aactggaccg aggtgctgat ggcgcaggcg      780
gcgaagcaga tcgatggcct gtcgttgac tattacacgc tgcccaccgg caactgggac      840
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aagcgcgtcg gtttgatggt tgacgagtgg ggcacgtggt acgaccgcga cgaggccgc      1020
gacatggggc cgttttatca gcagaacacg ttgcccgatg cggttgcggc cggtatcaat      1080
ctcaatatct ttcacaagta tgccgatcgc gtgcgcattg cgaacatcgc gcagatgggt      1140
aacgtgttgc aggcgatggt gttgacggac aaagagaaaa tgggtgctgac gccgacgtat      1200
cacgttttcc ggatgtatcg cgtgcacag ggagcgacgc tgatcccggt cgaggttagt      1260
gcgcgcgagt acacgctggg tgggtgcgtct gtgccgtcgt tgagcgtgtc ggcttcgcgt      1320
gacggtgaag gacgggtgca tctgtcgatc gtgaatctcg atccagcgcg ggcggcgag      1380
atcgatgcga acggaccgtt cagcagtgtc aagggagaag tattgactgc gccggcggtg      1440
aatgcgtga atactttcga tcacccgat agtgtcaagc ccgtgtcttt taatggatat      1500
aaattagaag gctctaaatt aatcctgaat attccggcga aatccgtggt ggtgttgaa      1560
cttgaccac agaaacaagc aacgctcaaa gatgcattca aaaacgattt catgatcggc      1620
gcggcgctca accggcgaca gttcttcgaa gaagacgctc gcggcgcaga gatcgtgcgc      1680
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atgttcgtcg tcggacacac gctcgtcttg cataacaaaa cgccgcgctg ggtttttgaa      1860
gacgaaaaga aacagccgct cgatcgcgag acgttgctga aacgaatcg cgatcacatc      1920
ttcaccgtcg tcggcgctta caagggacgc attaaaggct gggacgtagt caacgaggcg      1980
ctgaatcagg atggcacgat gcggcagtcg ccgtggttca agatcatcgg cgaggattat      2040
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gactacgac tcgagctgcc ggcgaagcgc gcaggcgccg tcgaactgct gaagaaactg      2160
aaagcccgcg gtgtgtcgt tgcgtggtgt ggattgcaga accacagtct catggagtgg      2220
ccgtcagccg cagatgtgga tgcgacgac gcggcgcttc cgaatctggg tttgaagggt      2280
cacatcacgg aactcgacgt cgacgtgctg ccgcgcacga cgaaaccgg tgcggattac      2340
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cgacagtcgg cgttggcgag gcgttatgcg gagctgttcc acgtgttttag aaaacatcgc 2460
gacgcgatcg agcgtgtgac gttctgggga gttgcggacg gcgattcgtg gttgaacaac 2520
tggtcccatcc gcggcaggac aaactatccg ctgctcttcg atcgttccgg ccaaccgaaa 2580
ccggcggttag cgtcggtgat cgaaaccgct aattattcaa cggaacgtcg acggtga 2637

<210> 14

<211> 878

<212> PRT

<213> Unknown

<220>

<223> Obtained from environmental sample

<220>

<221> SIGNAL

<222> (1) ... (28)

<220>

<221> DOMAIN

<222> (328) ... (515)

<223> Alpha-L-arabinofuranosidase C-terminus

<220>

<221> DOMAIN

<222> (528) ... (870)

<223> Glycosyl hydrolase family 10

<220>

<221> SITE

<222> (127) ... (130)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (205) ... (208)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (232) ... (235)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (251) ... (254)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (734) ... (737)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (757) ... (767)

<223> Glycosyl hydrolases family 10 active site. Prosite id = PS00591

<220>

<221> SITE

<222> (871) ... (874)

<223> N-glycosylation site. Prosite id = PS00001

<400> 14

Met	Thr	Ser	Gly	Arg	Asn	Thr	Cys	Val	Cys	Leu	Leu	Leu	Ile	Val	Leu
1			5						10					15	

Ala	Ile	Gly	Leu	Leu	Ser	Lys	Pro	Pro	Ala	Ser	Ala	Gln	Asn	Glu	Ala
		20					25						30		

Pro	Tyr	Lys	Ala	Thr	Leu	Thr	Ile	Arg	Leu	Asp	Gln	Pro	Gly	Ala	Val
		35					40					45			

Ile	Asn	Arg	Asn	Ile	Tyr	Gly	Gln	Phe	Ala	Glu	His	Leu	Gly	Arg	Leu
	50					55					60				

Ile	Tyr	Asp	Gly	Leu	Trp	Val	Gly	Glu	Gly	Ser	Ser	Ile	Pro	Asn	Thr
65					70					75					80

Arg	Gly	Leu	Arg	Asn	Asp	Val	Val	Thr	Ala	Leu	Lys	Glu	Leu	His	Val
			85						90					95	

Pro	Val	Leu	Arg	Trp	Pro	Gly	Gly	Cys	Phe	Ala	Asp	Glu	Tyr	His	Trp
		100						105					110		

Arg	Asp	Gly	Ile	Gly	Pro	Arg	Asp	Lys	Arg	Pro	Arg	Arg	Pro	Asn	Ala
	115						120					125			

Ser	Trp	Gly	Gly	Val	Asp	Ser	Asn	Ala	Phe	Gly	Thr	His	Glu	Phe	Met
	130					135					140				

Glu	Leu	Cys	Glu	Met	Leu	Gly	Ala	Asp	Ala	Tyr	Ile	Asn	Gly	Asn	Val
145				150						155					160

Gly	Ser	Gly	Thr	Pro	Gln	Glu	Met	Met	Glu	Trp	Ile	Glu	Tyr	Met	Thr
				165					170					175	

Ser	Asp	Ser	Asp	Ser	Asp	Leu	Ala	Asn	Leu	Arg	Arg	Arg	Asn	Gly	Arg
		180						185					190		

Asp	Lys	Pro	Trp	Lys	Val	Pro	Tyr	Phe	Ala	Val	Gly	Asn	Glu	Thr	Trp
		195					200					205			

Gly	Cys	Gly	Gly	Asn	Met	Arg	Pro	Glu	Phe	Tyr	Ala	Asp	Val	Tyr	Arg
	210					215					220				

Gln	Tyr	Ala	Thr	Phe	Ile	Lys	Asn	His	Ser	Gly	Asn	Arg	Ile	Gln	Lys
225					230					235					240

Leu	Ala	Ser	Gly	Gly	Tyr	Asp	Asn	Asn	Tyr	Asn	Trp	Thr	Glu	Val	Leu
			245						250					255	

Met	Ala	Gln	Ala	Ala	Lys	Gln	Ile	Asp	Gly	Leu	Ser	Leu	His	Tyr	Tyr
			260					265					270		

Thr	Leu	Pro	Thr	Gly	Asn	Trp	Asp	Lys	Lys	Gly	Ser	Ala	Thr	Glu	Phe
		275					280					285			

Gly Glu Ser Glu Trp His Ala Thr Leu Ala Arg Thr Leu Arg Ile Glu
 290 295 300
 Glu Phe Ile Gln Lys His Ser Ala Ile Met Asp Lys His Asp Pro Gln
 305 310 315 320
 Lys Arg Val Gly Leu Met Val Asp Glu Trp Gly Thr Trp Tyr Asp Arg
 325 330 335
 Asp Glu Gly Arg Asp Met Gly Ala Leu Tyr Gln Gln Asn Thr Leu Arg
 340 345 350
 Asp Ala Val Ala Ala Gly Ile Asn Leu Asn Ile Phe His Lys Tyr Ala
 355 360 365
 Asp Arg Val Arg Met Ala Asn Ile Ala Gln Met Val Asn Val Leu Gln
 370 375 380
 Ala Met Val Leu Thr Asp Lys Glu Lys Met Val Leu Thr Pro Thr Tyr
 385 390 395 400
 His Val Phe Arg Met Tyr Arg Val His Gln Gly Ala Thr Leu Ile Pro
 405 410 415
 Val Glu Val Ser Ala Pro Gln Tyr Thr Leu Gly Gly Ala Ser Val Pro
 420 425 430
 Ser Leu Ser Val Ser Ala Ser Arg Asp Gly Glu Gly Arg Val His Leu
 435 440 445
 Ser Ile Val Asn Leu Asp Pro Ala Arg Ala Ala Glu Ile Asp Ala Asn
 450 455 460
 Gly Pro Phe Ser Ser Val Lys Gly Glu Val Leu Thr Ala Pro Ala Val
 465 470 475 480
 Asn Ala Leu Asn Thr Phe Asp His Pro Asp Ser Val Lys Pro Val Ser
 485 490 495
 Phe Asn Gly Tyr Lys Leu Glu Gly Ser Lys Leu Ile Leu Asn Ile Pro
 500 505 510
 Ala Lys Ser Val Val Val Leu Glu Leu Gly Pro Gln Lys Gln Ala Thr
 515 520 525
 Leu Lys Asp Ala Phe Lys Asn Asp Phe Met Ile Gly Ala Ala Leu Asn
 530 535 540
 Arg Arg Gln Phe Phe Glu Glu Asp Ala Arg Gly Ala Glu Ile Val Arg
 545 550 555 560
 Met His Phe Asn Ser Ile Thr Pro Glu Asn Val Leu Lys Trp Gly Leu
 565 570 575
 Val His Pro Glu Pro Asn Lys Tyr Asp Phe Thr Ala Pro Asp Arg Phe
 580 585 590
 Val Glu Phe Gly Glu Lys His Gly Met Phe Val Val Gly His Thr Leu
 595 600 605
 Val Trp His Asn Gln Thr Pro Arg Trp Val Phe Glu Asp Glu Lys Lys
 610 615 620

Gln Pro Leu Asp Arg Glu Thr Leu Leu Lys Arg Met Arg Asp His Ile
625 630 635 640

Phe Thr Val Val Gly Arg Tyr Lys Gly Arg Ile Lys Gly Trp Asp Val
645 650 655

Val Asn Glu Ala Leu Asn Gln Asp Gly Thr Met Arg Gln Ser Pro Trp
660 665 670

Phe Lys Ile Ile Gly Glu Asp Tyr Leu Val Lys Ala Phe Glu Phe Ala
675 680 685

His Glu Ala Asp Pro Ala Ala Glu Leu Tyr Tyr Asn Asp Tyr Asp Leu
690 695 700

Glu Leu Pro Ala Lys Arg Ala Gly Ala Val Glu Leu Leu Lys Lys Leu
705 710 715 720

Lys Ala Ala Gly Val Ser Leu Ala Gly Val Gly Leu Gln Asn His Ser
725 730 735

Leu Met Glu Trp Pro Ser Ala Ala Asp Val Asp Ala Thr Ile Ala Ala
740 745 750

Phe Ala Asn Leu Gly Leu Lys Val His Ile Thr Glu Leu Asp Val Asp
755 760 765

Val Leu Pro Arg Thr Thr Lys Pro Gly Ala Asp Tyr Ala Val Asp Val
770 775 780

Lys Val Thr Pro Gln Leu Asn Pro Tyr Leu Asp Gly Leu Pro Glu Ala
785 790 795 800

Arg Gln Ser Ala Leu Ala Arg Arg Tyr Ala Glu Leu Phe His Val Phe
805 810 815

Arg Lys His Arg Asp Ala Ile Glu Arg Val Thr Phe Trp Gly Val Ala
820 825 830

Asp Gly Asp Ser Trp Leu Asn Asn Trp Pro Ile Arg Gly Arg Thr Asn
835 840 845

Tyr Pro Leu Leu Phe Asp Arg Ser Gly Gln Pro Lys Pro Ala Leu Ala
850 855 860

Ser Val Ile Glu Thr Ala Asn Tyr Ser Thr Glu Arg Arg Arg
865 870 875

<210> 15

<211> 1725

<212> DNA

<213> Clostridium thermocellum

<400> 15

gtgtggaagc cggattgtg gaatttcctt caaatggcag atgaagccgg attgacgagg 60

gatggaaaca ctccggttcc gacaccocagt ccaaagccgg ctaacacacg tattgaagcg 120

gaagattatg acggtattaa ttcttcaagt attgagataa taggtgttcc acctgaagga 180

ggcagaggaa taggttatat taccagtggg gattatctgg tatacaagag tatagacttt 240

.....

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ggaaacggag caacgtcgtt taaggccaag gttgcaaatg caaatacttc caatattgaa 300
cttagattaa acgggtccgaa tgggtactctc ataggcacac tctcggtaaa atccacagga 360
gattggaata catatgagga gcaaacttgc agcattagca aagtcaccgg aataaatgat 420
ttgtacttgg tattcaaagg ccctgtaaac atagactggg tcacttttgg cgttgaaagc 480
agttccacag gtctggggga tttaaatggg gacggaaata ttaactcgtc ggaccttcag 540
gcgttaaaga ggcatttgct cggatatatca ccgcttacgg gagaggctct ttaagagcg 600
gatgtaaata ggagcggcaa agtggattct actgactatt cagtgtgtaa aagatatata 660
ctccgcatta ttacagagtt ccccgacaa ggtgatgtac agacaccaa tccgtctgtt 720
actccgacac aaactcctat cccacgatt tcgggaaatg ctcttaggga ttatgcggag 780
gcaaggggaa taaaaatcgg aacatgtgtc aactatccgt tttacaacaa ttcagatcca 840

acctacaaca gcattttgca aagagaattt tcaatggttg tatgtgaaaa tgaaatgaag 900
tttgatgctt tgcagccgag acaaaacgtt tttgattttt cgaaaggaga ccagttgctt 960
gcttttgca aaagaaacgg tatgcagatg aggggacata cgttgatttg gcacaatcaa 1020
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aatcacatta ccactgttat gaccattac aaaggtaaaa ttgttgagtg ggatgtggca 1140
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gcggaaatag gcgttatagt atcctttacc gaaatagata tacgcatacc tcagtcggaa 1500
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ctggcaaaacc ccaattgcaa tacctttgta atgtggggat tcacagataa atacacatgg 1620
attccgggaa ctttcccagg atatggcaat ccattgattt atgacagcaa ttacaatccg 1680
aaaccggcat acaatgcaat aaaggaagct cttatgggct attga 1725

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<210> 16

<211> 574

<212> PRT

<213> Clostridium thermocellum

<220>

<221> DOMAIN

<222> (39) ... (158)

<223> Carbohydrate binding module (family 6)

<220>

<221> DOMAIN

<222> (167) ... (187)

<223> Dockerin type I repeat

<220>

<221> DOMAIN

<222> (201) ... (221)

<223> Dockerin type I repeat

<220>

<221> DOMAIN

<222> (254) ... (571)

<223> Glycosyl hydrolase family 10

<220>

<221> SITE

<222> (47) ... (50)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (95) ... (98)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (107) ... (110)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (167) ... (179)

<223> EF-hand calcium-binding domain. Prosite id = PS00018

<220>

<221> SITE

<222> (167) ... (186)

<223> Clostridium cellulosome enzymes repeated domain signature. Prosite id = PS00448

<220>

<221> SITE

<222> (175) ... (178)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (201) ... (213)

<223> EF-hand calcium-binding domain. Prosite id = PS00018

<220>

<221> SITE

<222> (201) ... (220)

<223> Clostridium cellulosome enzymes repeated domain signature. Prosite id = PS00448

<220>

<221> SITE

<222> (203) ... (206)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (276) ... (279)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (484) ... (494)

<223> Glycosyl hydrolases family 10 active site. Prosite id = PS00591

<400> 16

Met Trp Lys Pro Gly Leu Trp Asn Phe Leu Gln Met Ala Asp Glu Ala
 1 5 10 15

Gly Leu Thr Arg Asp Gly Asn Thr Pro Val Pro Thr Pro Ser Pro Lys
 20 25 30

Pro Ala Asn Thr Arg Ile Glu Ala Glu Asp Tyr Asp Gly Ile Asn Ser
 35 40 45

Ser Ser Ile Glu Ile Ile Gly Val Pro Pro Glu Gly Gly Arg Gly Ile
 50 55 60

Gly Tyr Ile Thr Ser Gly Asp Tyr Leu Val Tyr Lys Ser Ile Asp Phe
 65 70 75 80

Gly Asn Gly Ala Thr Ser Phe Lys Ala Lys Val Ala Asn Ala Asn Thr
 85 90 95

Ser Asn Ile Glu Leu Arg Leu Asn Gly Pro Asn Gly Thr Leu Ile Gly
 100 105 110

Thr Leu Ser Val Lys Ser Thr Gly Asp Trp Asn Thr Tyr Glu Glu Gln
 115 120 125

Thr Cys Ser Ile Ser Lys Val Thr Gly Ile Asn Asp Leu Tyr Leu Val
 130 135 140

Phe Lys Gly Pro Val Asn Ile Asp Trp Phe Thr Phe Gly Val Glu Ser
 145 150 155 160

Ser Ser Thr Gly Leu Gly Asp Leu Asn Gly Asp Gly Asn Ile Asn Ser
 165 170 175

Ser Asp Leu Gln Ala Leu Lys Arg His Leu Leu Gly Ile Ser Pro Leu
 180 185 190

Thr Gly Glu Ala Leu Leu Arg Ala Asp Val Asn Arg Ser Gly Lys Val
 195 200 205

Asp Ser Thr Asp Tyr Ser Val Leu Lys Arg Tyr Ile Leu Arg Ile Ile
 210 215 220

Thr Glu Phe Pro Gly Gln Gly Asp Val Gln Thr Pro Asn Pro Ser Val
 225 230 235 240

Thr Pro Thr Gln Thr Pro Ile Pro Thr Ile Ser Gly Asn Ala Leu Arg
 245 250 255

Asp Tyr Ala Glu Ala Arg Gly Ile Lys Ile Gly Thr Cys Val Asn Tyr
 260 265 270

Pro Phe Tyr Asn Asn Ser Asp Pro Thr Tyr Asn Ser Ile Leu Gln Arg
 275 280 285

210 200 200
 Glu Phe Ser Met Val Val Cys Glu Asn Glu Met Lys Phe Asp Ala Leu
 290 295 300
 Gln Pro Arg Gln Asn Val Phe Asp Phe Ser Lys Gly Asp Gln Leu Leu
 305 310 315 320
 Ala Phe Ala Glu Arg Asn Gly Met Gln Met Arg Gly His Thr Leu Ile
 325 330 335
 Trp His Asn Gln Asn Pro Ser Trp Leu Thr Asn Gly Asn Trp Asn Arg
 340 345 350
 Asp Ser Leu Leu Ala Val Met Lys Asn His Ile Thr Thr Val Met Thr
 355 360 365
 His Tyr Lys Gly Lys Ile Val Glu Trp Asp Val Ala Asn Glu Cys Met
 370 375 380
 Asp Asp Ser Gly Asn Gly Leu Arg Ser Ser Ile Trp Arg Asn Val Ile
 385 390 395 400
 Gly Gln Asp Tyr Leu Asp Tyr Ala Phe Arg Tyr Ala Arg Glu Ala Asp
 405 410 415
 Pro Asp Ala Leu Leu Phe Tyr Asn Asp Tyr Asn Ile Glu Asp Leu Gly
 420 425 430
 Pro Lys Ser Asn Ala Val Phe Asn Met Ile Lys Ser Met Lys Glu Arg
 435 440 445
 Gly Val Pro Ile Asp Gly Val Gly Phe Gln Cys His Phe Ile Asn Gly
 450 455 460
 Met Ser Pro Glu Tyr Leu Ala Ser Ile Asp Gln Asn Ile Lys Arg Tyr
 465 470 475 480
 Ala Glu Ile Gly Val Ile Val Ser Phe Thr Glu Ile Asp Ile Arg Ile
 485 490 495
 Pro Gln Ser Glu Asn Pro Ala Thr Ala Phe Gln Val Gln Ala Asn Asn
 500 505 510
 Tyr Lys Glu Leu Met Lys Ile Cys Leu Ala Asn Pro Asn Cys Asn Thr
 515 520 525
 Phe Val Met Trp Gly Phe Thr Asp Lys Tyr Thr Trp Ile Pro Gly Thr
 530 535 540
 Phe Pro Gly Tyr Gly Asn Pro Leu Ile Tyr Asp Ser Asn Tyr Asn Pro
 545 550 555 560
 Lys Pro Ala Tyr Asn Ala Ile Lys Glu Ala Leu Met Gly Tyr 565 570

<210> 17

<211> 2232

<212> DNA

<213> *Cochliobolus heterostrophus* ATCC 48331

<400> 17

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ataaacctca tcaataaata caaaggtata tcaagattag atcttcacac atataactaa	180

tggggcggaag	ctctacacgg	tgtagctgga	gcgccaggaa	tcaaattcgt	cgaaccttat	240
aaaaacgcta	cttcgtttcc	tatgccaatc	cttatgtcgg	cagcttttga	tgatgatctc	300
attttcaaaa	ttgccaatat	tatcgggaac	gaggcccgag	ccttcggaaa	tggtggagtc	360
gctcctatgg	actattggac	ccctgacatc	aatcccgtcc	gcgatatacg	atggggccga	420
gccagtgaat	cacccggaga	ggacattcga	cgaataaaag	ggtacaccaa	ggctctgctt	480
gctggcctcg	aaggtgacca	agcccaaagg	aagatcattg	caacatgcaa	acactatgtg	540
ggttacgaca	tggaagcttg	gggaggatac	gatcgacaca	acttcagtgc	aaagatcacc	600
atgcaagacc	tcgcagagta	ctacatgccg	ccattccagc	aatgtgcgcg	tgactcgaag	660
gtcgggtcat	tcatgtgcag	ctacaatgca	gtcaacggtg	ttccaacatg	cgctgacacc	720
tacgttcttc	aaacaatcct	gagggaccac	tggaaactgga	cagatagcaa	taactacatt	780
actagcgatt	gcgaagccgt	tgcggatata	tctgagaacc	acaaatatgt	cgaaacccctt	840
gcgcaaggca	ccgcacttgc	ttttgccaaag	ggtatggatc	ttagctgtga	atacagtgga	900
tcgtcagata	tcccaggagc	ttggtcacaa	ggtcttctga	atctttctgt	tatcgacaaa	960
gcattgactc	gacaatatga	aggcttagtc	catgccggct	actttgatgg	cgcgaaaggcg	1020
acttacgcaa	acttgagtta	taatgacatc	aacacacccg	aagcacgaca	gctatccttg	1080
caagttacct	ctgaaggttt	ggtcatgcta	aagaacgata	acacacttcc	attgcctctc	1140
acgaagggat	caaagggtgc	tatgataggt	ttctgggccca	acgactcttc	caaactccag	1200
ggcatctaca	gcggtccacc	tccttaccgg	cactctccag	tattcgctgg	tgaacaaatg	1260
ggattagata	tggccatagc	ctggggccca	atgattcaga	actcaagtgt	gcccgacaac	1320
tggactacca	acgcgcctcg	cgcgcccgag	aagtccgact	atatctctta	ctttggtggt	1380
caagactgga	cagtggcgca	agaaggctac	gatcgcata	caatcagttt	tcctcaagtg	1440
caaatcgacc	ttcttgccaa	actggctaaa	cttggcaagc	cgcttggtgt	catcacgctt	1500
ggtgatatga	ctgatcactc	ccctctcttg	tccatggaag	gcatacaatc	aattatctgg	1560
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agcgaagaag	gtctaacctc	cgatatccag	gaaaccttgg	atagctgtac	acagcagtac	1860
aaggatttgt	gtgaggttgc	accactggag	gtcaccgttg	caaacaaggg	taaccgaaca	1920
tcggatttgc	tcgctctcgc	tttcatcaag	ggcgagggtg	gacctaaagg	ataccactta	1980
aagactctga	ttacgtacgg	gaggctcaga	gatattccatg	ggggcgcgaa	gaagtcggcg	2040
tcacttccgc	ttacacttgg	agaattggcc	agagtggatc	aatcaggcaa	caccgttatc	2100
tatcccggcg	aatacacccct	gtcctttgac	gagcctactc	aggetgagct	gaaattgact	2160
attacggggc	aggagacaat	tctggacaaa	tggccccagc	cgccaaacgg	aggcaatcgg	2220
accgtccttt	ga					2232

<210> 18

<211> 743

<212> PRT

<213> *Cochliobolus heterostrophus* ATCC 48331

<220>

<221> DOMAIN
<222> (47) ... (297)
<223> Glycosyl hydrolase family 3 N terminal domain

<220>
<221> DOMAIN
<222> (367) ... (594)
<223> Glycosyl hydrolase family 3 C terminal domain

<220>
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<222> (82) ... (85)
<223> N-glycosylation site. Prosite id = PS00001

<220>
<221> SITE
<222> (252) ... (255)
<223> N-glycosylation site. Prosite id = PS00001

<220>
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<222> (314) ... (317)
<223> N-glycosylation site. Prosite id = PS00001

<220>
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<222> (344) ... (347)
<223> N-glycosylation site. Prosite id = PS00001

<220>
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<222> (394) ... (397)
<223> N-glycosylation site. Prosite id = PS00001

<220>
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<222> (434) ... (437)
<223> N-glycosylation site. Prosite id = PS00001

<220>
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<222> (440) ... (443)
<223> N-glycosylation site. Prosite id = PS00001

<220>
<221> SITE
<222> (580) ... (583)
<223> N-glycosylation site. Prosite id = PS00001

<220>
<221> SITE
<222> (638) ... (641)
<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (739) ... (742)

<223> N-glycosylation site. Prosite id = PS00001

<400> 18

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 20 25 30

Ala Met Glu Pro Gln Glu Lys Leu Asp Asn Leu Val Ser Lys Ser Lys
 35 40 45

Gly Val Ser Arg Leu Gly Leu Pro Ala Tyr Asn Trp Trp Gly Glu Ala
 50 55 60

Leu His Gly Val Ala Gly Ala Pro Gly Ile Lys Phe Val Glu Pro Tyr
 65 70 75 80

Lys Asn Ala Thr Ser Phe Pro Met Pro Ile Leu Met Ser Ala Ala Phe
 85 90 95

Asp Asp Asp Leu Ile Phe Lys Ile Ala Asn Ile Ile Gly Asn Glu Ala
 100 105 110

Arg Ala Phe Gly Asn Gly Gly Val Ala Pro Met Asp Tyr Trp Thr Pro
 115 120 125

Asp Ile Asn Pro Val Arg Asp Ile Arg Trp Gly Arg Ala Ser Glu Ser
 130 135 140

Pro Gly Glu Asp Ile Arg Arg Ile Lys Gly Tyr Thr Lys Ala Leu Leu
 145 150 155 160

Ala Gly Leu Glu Gly Asp Gln Ala Gln Arg Lys Ile Ile Ala Thr Cys
 165 170 175

Lys His Tyr Val Gly Tyr Asp Met Glu Ala Trp Gly Gly Tyr Asp Arg
 180 185 190

His Asn Phe Ser Ala Lys Ile Thr Met Gln Asp Leu Ala Glu Tyr Tyr
 195 200 205

Met Pro Pro Phe Gln Gln Cys Ala Arg Asp Ser Lys Val Gly Ser Phe
 210 215 220

Met Cys Ser Tyr Asn Ala Val Asn Gly Val Pro Thr Cys Ala Asp Thr
 225 230 235 240

Tyr Val Leu Gln Thr Ile Leu Arg Asp His Trp Asn Trp Thr Asp Ser
 245 250 255

Asn Asn Tyr Ile Thr Ser Asp Cys Glu Ala Val Ala Asp Ile Ser Glu
 260 265 270

Asn His Lys Tyr Val Glu Thr Leu Ala Gln Gly Thr Ala Leu Ala Phe
 275 280 285

Ala Lys Gly Met Asp Leu Ser Cys Glu Tyr Ser Gly Ser Ser Asp Ile
 290 295 300

Pro Gly Ala Trp Ser Gln Gly Leu Leu Asn Leu Ser Val Ile Asp Lys
 305 310 315 320
 Ala Leu Thr Arg Gln Tyr Glu Gly Leu Val His Ala Gly Tyr Phe Asp
 325 330 335
 Gly Ala Lys Ala Thr Tyr Ala Asn Leu Ser Tyr Asn Asp Ile Asn Thr
 340 345 350
 Pro Glu Ala Arg Gln Leu Ser Leu Gln Val Thr Ser Glu Gly Leu Val
 355 360 365
 Met Leu Lys Asn Asp His Thr Leu Pro Leu Pro Leu Thr Lys Gly Ser
 370 375 380
 Lys Val Ala Met Ile Gly Phe Trp Ala Asn Asp Ser Ser Lys Leu Gln
 385 390 395 400
 Gly Ile Tyr Ser Gly Pro Pro Pro Tyr Arg His Ser Pro Val Phe Ala
 405 410 415
 Gly Glu Gln Met Gly Leu Asp Met Ala Ile Ala Trp Gly Pro Met Ile
 420 425 430
 Gln Asn Ser Ser Val Pro Asp Asn Trp Thr Thr Asn Ala Leu Asp Ala
 435 440 445
 Ala Glu Lys Ser Asp Tyr Ile Leu Tyr Phe Gly Gly Gln Asp Trp Thr
 450 455 460
 Val Ala Gln Glu Gly Tyr Asp Arg Thr Thr Ile Ser Phe Pro Gln Val
 465 470 475 480
 Gln Ile Asp Leu Leu Ala Lys Leu Ala Lys Leu Gly Lys Pro Leu Val
 485 490 495
 Val Ile Thr Leu Gly Asp Met Thr Asp His Ser Pro Leu Leu Ser Met
 500 505 510
 Glu Gly Ile Asn Ser Ile Ile Trp Ala Asn Trp Pro Gly Gln Asp Gly
 515 520 525
 Gly Pro Ala Ile Leu Asn Val Ile Ser Gly Val His Ala Pro Ala Gly
 530 535 540
 Arg Leu Pro Ile Thr Glu Tyr Pro Ala Asp Tyr Val Lys Leu Ser Met
 545 550 555 560
 Leu Asp Met Asn Leu Arg Pro His Ala Glu Ser Pro Gly Arg Thr Tyr
 565 570 575
 Arg Trp Phe Asn Glu Ser Val Gln Pro Phe Gly Phe Gly Leu His Tyr
 580 585 590
 Thr Thr Phe Glu Ala Gly Phe Ala Ser Glu Glu Gly Leu Thr Tyr Asp
 595 600 605
 Ile Gln Glu Thr Leu Asp Ser Cys Thr Gln Gln Tyr Lys Asp Leu Cys
 610 615 620
 Glu Val Ala Pro Leu Glu Val Thr Val Ala Asn Lys Gly Asn Arg Thr
 625 630 635 640

Ser Asp Phe Val Ala Leu Ala Phe Ile Lys Gly Glu Val Gly Pro Lys

645

650

655

Pro Tyr Pro Leu Lys Thr Leu Ile Thr Tyr Gly Arg Leu Arg Asp Ile
660 665 670

His Gly Gly Ala Lys Lys Ser Ala Ser Leu Pro Leu Thr Leu Gly Glu
675 680 685

Leu Ala Arg Val Asp Gln Ser Gly Asn Thr Val Ile Tyr Pro Gly Glu
690 695 700

Tyr Thr Leu Leu Leu Asp Glu Pro Thr Gln Ala Glu Leu Lys Leu Thr
705 710 715 720

Ile Thr Gly Glu Glu Thr Ile Leu Asp Lys Trp Pro Gln Pro Pro Asn
725 730 735

Gly Gly Asn Arg Thr Val Leu
740

<210> 19

<211> 1362

<212> DNA

<213> Bacteria

<400> 19

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ggcaaagtag cgaatgggga taatggcgat atcgcaattg atcattacaa tcgttatgtt	180
gaagacatcg cattaatgaa agcgccttcat ttgaaagcgt atcgattttc gactagttag	240
gcgagacttt attgtgaaac gccagggaag tttaacgaaa aaggtttaga tttttataag	300
cgtcttgtag atgaattgct agagaacggt atcgagccaa tgttgaccat ttatcattgg	360
gatatgccac aagctcttca agagaaaggt ggctgggaaa atcgtgatat cgttcactac	420
ttccaagaat acgctgcttt cctttacgag aatcttgagg atgtcgtgaa aaaatggatt	480
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gggattcaaa actttacatc atttttaaaa gcagcacatc atgtttcttct ctacacggg	600
gaagcggtaa aagcgtttcg agcaatcggg tcgaaagatg gggaaattgg tattacgttg	660
aatttgacac ctggatatgc ggtcgatccg aaagatgaaa aagcagttga tgccgctcga	720
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cctgtgacgg atatgaattg ggaagtgaat ccaaaccggt tgtttgattt aatgggtcga	1020
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<210> 20

<211> 453

<212> PRT

<213> Bacteria

<220>

<221> DOMAIN

<222> (1) ... (447)

<223> Glycosyl hydrolase family 1

<220>

<221> SITE

<222> (8) ... (22)

<223> Glycosyl hydrolases family 1 N-terminal signature. Prosite id = PS00653

<220>

<221> SITE

<222> (184) ... (187)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (350) ... (358)

<223> Glycosyl hydrolases family 1 active site. Prosite id = PS00572

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Tyr Gln Ile Glu Gly Thr Ala Thr Gly Glu Asp Lys Ile Tyr Ser Ile
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Trp Asp His Phe Ser Arg Ile Pro Gly Lys Val Ala Asn Gly Asp Asn
35 40 45

Gly Asp Ile Ala Ile Asp His Tyr Asn Arg Tyr Val Glu Asp Ile Ala
50 55 60

Leu Met Lys Ala Leu His Leu Lys Ala Tyr Arg Phe Ser Thr Ser Trp
65 70 75 80

Ala Arg Leu Tyr Cys Glu Thr Pro Gly Lys Phe Asn Glu Lys Gly Leu
85 90 95

Asp Phe Tyr Lys Arg Leu Val His Glu Leu Leu Glu Asn Gly Ile Glu
100 105 110

Pro Met Leu Thr Ile Tyr His Trp Asp Met Pro Gln Ala Leu Gln Glu
115 120 125

Lys Gly Gly Trp Glu Asn Arg Asp Ile Val His Tyr Phe Gln Glu Tyr
130 135 140

Ala Ala Phe Leu Tyr Glu Asn Leu Gly Asp Val Val Lys Lys Trp Ile
145 150 155 160

Thr His Asn Glu Pro Trp Val Val Thr Tyr Leu Gly Tyr Gly Asn Gly
 165 170 175
 Glu His Ala Pro Gly Ile Gln Asn Phe Thr Ser Phe Leu Lys Ala Ala
 180 185 190
 His His Val Leu Leu Ser His Gly Glu Ala Val Lys Ala Phe Arg Ala
 195 200 205
 Ile Gly Ser Lys Asp Gly Glu Ile Gly Ile Thr Leu Asn Leu Thr Pro
 210 215 220
 Gly Tyr Ala Val Asp Pro Lys Asp Glu Lys Ala Val Asp Ala Ala Arg
 225 230 235 240
 Lys Trp Asp Gly Phe Met Asn Arg Trp Phe Leu Asp Pro Val Phe Lys
 245 250 255
 Gly Gln Tyr Pro Ala Asp Met Leu Glu Val Tyr Lys Asp Tyr Leu Pro
 260 265 270
 Asp Val Tyr Lys Glu Gly Asp Leu Gln Thr Ile Gln Gln Pro Ile Asp
 275 280 285
 Phe Phe Gly Phe Asn Tyr Tyr Ser Thr Ala Thr Leu Lys Asp Trp Lys
 290 295 300
 Thr Gly Asp Arg Glu Pro Ile Val Phe Glu His Val Ser Thr Gly Arg
 305 310 315 320
 Pro Val Thr Asp Met Asn Trp Glu Val Asn Pro Asn Gly Leu Phe Asp
 325 330 335
 Leu Met Val Arg Leu Lys Lys Asp Tyr Gly Asp Ile Pro Leu Tyr Ile
 340 345 350
 Thr Glu Asn Gly Ala Ala Tyr Lys Asp Arg Val Asn Glu Gln Gly Glu
 355 360 365
 Val Glu Asp Asp Glu Arg Val Ala Tyr Ile Arg Glu His Leu Ile Ala
 370 375 380
 Cys His Arg Ala Ile Glu Gln Gly Val Asn Leu Lys Gly Tyr Tyr Val
 385 390 395 400
 Trp Ser Leu Phe Asp Asn Phe Glu Trp Ala Phe Gly Tyr Asp Lys Arg
 405 410 415
 Phe Gly Ile Val Tyr Val Asp Tyr Glu Thr Leu Glu Arg Ile Pro Lys
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 Lys Ser Ala Leu Trp Tyr Lys Glu Thr Ile Ile Asn Asn Gly Leu Gln
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 Val Asp Asn Asp Lys
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<210> 21

<211> 930

<212> DNA

<213> *Cochliobolus heterostrophus* ATCC 48331

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<210> 22

<211> 309

<212> PRT

<213> *Cochliobolus heterostrophus* ATCC 48331

<220>

<221> DOMAIN

<222> (3) ... (276)

<223> Glycosyl hydrolase family 62

<220>

<221> SITE

<222> (43) ... (46)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (192) ... (195)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (287) ... (290)

<223> N-glycosylation site. Prosite id = PS00001

<400> 22

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20           25           30

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Lys Glu Ala Gly Tyr Asn Leu Val Tyr Phe Asn Phe Thr Asp Phe Ser
35 40 45

Arg Ala Asn Gln Ala Pro Phe Phe Tyr Leu Asp Gln Ser Gly Ile Gly
50 55 60

Thr Gly Tyr Arg Ala Ala Pro Gln Val Phe Tyr Phe Ala Pro Gln Lys
65 70 75 80

Leu Trp Tyr Leu Ile Tyr Gln Asn Gly Asn Ala Ala Tyr Ser Thr Asn
85 90 95

Pro Asp Ile Ser Asn Pro Arg Gly Trp Thr Ala Pro Gln Val Phe Tyr
100 105 110

Pro Asn Gly Thr Pro Gln Thr Ile Gln Asn Gly Leu Gly Thr Thr Gly
115 120 125

Tyr Trp Val Asp Met Trp Val Ile Cys Asp Thr Ala Leu Cys His Leu
130 135 140

Tyr Ser Ser Asp Asp Asn Gly Gly Leu Tyr Arg Ser Gln Thr Pro Val
145 150 155 160

Ser Gln Phe Pro Arg Gly Met Asn Glu Pro Val Val Thr Leu Lys Ala
165 170 175

Asn Lys Asn Asp Leu Phe Glu Ala Ser Thr Val Tyr Asn Ile Val Asn
180 185 190

Thr Ser Thr Tyr Leu Leu Met Val Glu Cys Ile Gly Ser Gly Asn Ser
195 200 205

Pro Gly Gly Leu Arg Tyr Phe Arg Ser Trp Thr Thr Gln Ser Leu Thr
210 215 220

Ser Asp Lys Trp Thr Pro Leu Ala Ala Ser Gln Gln Thr Pro Phe Leu
225 230 235 240

Gly Ala Ala Asn Thr Gln Phe Pro Ala Gly Arg Trp Ser Gln Ser Leu
245 250 255

Ser His Gly Glu Leu Val Arg Thr Asn Val Asp Gln Arg Leu Gln Ile
260 265 270

Arg Pro Cys Glu Met Arg Tyr Leu Tyr Gln Gly Ile Asp Pro Asn Ala
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Thr Gly Thr Tyr Asn Ala Leu Pro Trp Lys Leu Ala Leu Ala Thr Gln
290 295 300

Thr Asn Ser Lys Cys
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<210> 23

<211> 2163

<212> DNA

<213> Unknown

<220>

<223> Obtained from environmental sample

<400> 23

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aacatgtaca tcggagtgga agacgagcag cgcaccttcc attgccttcc cttttttggg      180
gatgctgctg cagggggcca ggaagcactg cgctacgatg tggaaggcag ccaatccagt      240

gacgatccac tggccggcgc ctatgtcgga caccagagg atgcgccatc gctgcctccg      300
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gacttgacgg tggatcatct ctactgggaa ctgcgcctga atccctgggt tgtgaaaaat     1140
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catcttgaag cgtgctaaa acccggtatc tgtctgtttg aagatggtgg ctggaagtta     1920
agttcaacct ctgataaact gtggctgagc aaaatctacc tgtgccagtt cgtcgcgcgc     1980
cagatattgg gccgtgaacg cgatgacatt gacaagcgcg ccgatgccgc gcacgtgggc     2040
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tga                                                                 2163

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<210> 24

<211> 720

<212> PRT

<213> Unknown

<220>

<223> Obtained from environmental sample

<220>

<221> DOMAIN

<222> (19) ... (456)

<223> Glycosyl hydrolase family 52

<220>

<221> SITE

<222> (165) ... (168)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (229) ... (232)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (314) ... (317)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (522) ... (525)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (571) ... (574)

<223> N-glycosylation site. Prosite id = PS00001

<220>

<221> SITE

<222> (582) ... (585)

<223> N-glycosylation site. Prosite id = PS00001

<400> 24

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			20					25					30		

Glu	Leu	Gly	Gly	Pro	Ala	Asn	His	Asn	Met	Tyr	Ile	Gly	Val	Glu	Asp
		35					40					45			

Glu	Gln	Arg	Thr	Phe	His	Cys	Leu	Pro	Phe	Phe	Gly	Asp	Ala	Ala	Ala
	50					55					60				

Gly	Ala	Glu	Glu	Ala	Leu	Arg	Tyr	Asp	Val	Glu	Gly	Ser	Gln	Ser	Ser
65					70					75				80	

Asp Asp Pro Leu Ala Gly Ala Tyr Val Gly His Pro Glu Asp Ala Pro
 85 90 95
 Ser Leu Pro Pro Ala Lys Leu Arg Ala Leu Asp Gln Ser Ala Ile Ser
 100 105 110
 Arg Asp Phe Gln Leu Thr Thr Asp Thr Trp Thr Ala Pro Asp Phe Ser
 115 120 125
 Leu Thr Ile Tyr Ser Pro Val Arg Gly Val Pro Asp Pro Thr Thr Ala
 130 135 140
 Ala Glu Asp Glu Leu Lys Ala Ile Leu Val Pro Ala Val Leu Cys Glu
 145 150 155 160
 Leu Thr Val Asp Asn Ser Ser Gly Gln Gln Ser Arg Arg Ala Leu Phe
 165 170 175
 Gly Phe Thr Gly Asn Asp Pro Tyr Trp Gly Thr Arg Arg Leu Asp Asp
 180 185 190
 Val Ala Asn Ser Ala Phe Val Gly Val Gly Glu Gly Asn His Leu Ala
 195 200 205
 Ile Ala Ser Arg Asp Glu Gly Val Thr Ala Ala Leu Gly Phe Asn Ile
 210 215 220
 Asn Gly Val Ile Asn Glu Thr Leu Pro Glu Asn Tyr Ala Phe Gly Leu
 225 230 235 240
 Gly Lys Cys Ala Val Leu Leu Cys Glu Val Pro Ala Gly Glu Lys Arg
 245 250 255
 Thr Phe His Ile Ala Val Cys Phe His Arg Ser Gly Ile Ala Thr Thr
 260 265 270
 Gly Leu Lys Met Arg Tyr Tyr Tyr Thr Arg Phe Phe Pro Asp Ile Glu
 275 280 285
 Ser Val Ala Ala Tyr Ala Leu Glu Gln Phe Asp Ser Leu Lys Ser Ala
 290 295 300
 Ala Leu Gln Asp Asn Gln Leu Val Glu Asn Ala Ser Leu Ser Glu Asp
 305 310 315 320
 Gln Lys Trp Met Phe Cys His Ala Val Arg Ser Tyr Tyr Gly Ser Thr
 325 330 335
 Glu Leu Leu Glu Tyr Asn Asp Asn Pro Val Trp Val Val Asn Glu Gly
 340 345 350
 Glu Tyr Arg Met Met Asn Thr Phe Asp Leu Thr Val Asp His Leu Tyr
 355 360 365
 Trp Glu Leu Arg Leu Asn Pro Trp Val Val Lys Asn Gln Leu Asp Trp
 370 375 380
 Phe Val Asp Arg Tyr Ser Tyr Glu Asp Lys Val Arg Phe Pro Gly Asp
 385 390 395 400
 Lys Thr Glu Tyr Pro Cys Gly Leu Ser Phe Thr His Asp Met Gly Val
 405 410 415

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Thr Asn Val Trp Ser Arg Pro Gly Tyr Ser Ser Tyr Glu Lys Gln Gly
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Leu Lys Gly Val Phe Ser Tyr Met Thr His Glu Gln Leu Val Asn Trp
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Leu Cys Cys Ala Thr Val Tyr Val Glu Gln Thr Gly Asp Gln Glu Trp
    450                      455                      460

Leu Glu Gln Arg Trp Pro Ile Phe Asn Arg Cys Phe Glu Ser Leu Leu
    465                      470                      475                      480

Asn Arg Asp His Pro Asp Pro Glu Lys Arg Arg Gly Leu Met Gln Leu
    485                      490                      495

Asp Ser Thr Arg Cys Ala Gly Gly Ala Glu Ile Thr Thr Tyr Asp Ser
    500                      505                      510

Leu Asp Val Ser Leu Gly Gln Ser Arg Asn Asn Thr Tyr Leu Gly Gly
    515                      520                      525

Lys Ile Trp Ala Ser Tyr Leu Ala Leu Glu Lys Leu Phe Arg Glu Arg
    530                      535                      540

Gly Asp Val Glu Arg Ala Gln Val Ala His Gln Gln Ala His Arg Thr
    545                      550                      555                      560

Ala Gln Thr Leu Leu Glu Asn Val Gly Glu Asn Gly Thr Ile Pro Ala
    565                      570                      575

Val Leu Glu Gly Ser Asn Gln Ser Arg Ile Ile Pro Val Ile Glu Gly
    580                      585                      590

Leu Ile Phe Pro Tyr Phe Thr Gly Arg Lys Asp Val Leu Ser Ser Asp
    595                      600                      605

Gly Asp Phe Gly Glu Met Phe Ser Ala Leu Lys Arg His Leu Glu Ala
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Val Leu Lys Pro Gly Ile Cys Leu Phe Glu Asp Gly Gly Trp Lys Leu
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Ser Ser Thr Ser Asp Asn Ser Trp Leu Ser Lys Ile Tyr Leu Cys Gln
    645                      650                      655

Phe Val Ala Arg Gln Ile Leu Gly Arg Glu Arg Asp Asp Ile Asp Lys
    660                      665                      670

Arg Ala Asp Ala Ala His Val Gly Trp Leu Leu Asp Glu Arg Asn Ala
    675                      680                      685

Tyr Phe Ala Trp Ser Asp Gln Met Leu Ala Gly Phe Ala Glu Gly Ser
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Lys Tyr Tyr Pro Arg Gly Val Thr Ser Ala Leu Trp Leu Leu Glu Gly
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<210> 25

<211> 6564

<212> DNA

<213> Artificial sequence

<220>

<223> Synthetically generated vector

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Patentkrav

- 1.** Isoleret, syntetisk eller rekombinant nukleinsyre omfattende en nukleinsyre der koder for et polypeptid med en xylanaseaktivitet, hvor nukleinsyren omfatter en
- 5 sekvens med mindst 95%, 96%, 97%, 98%, 99%, eller mere, eller fuldstændig (100%), sekvensidentitet med en sekvens, der er SEQ ID NO: 1 med mutationen, at nukleinsyrekodonet der koder for aminosyrerest 38 af SEQ ID NO: 2 er ændret fra CGT til CAC, og aminosyrerest 38 Arginin, Arg (eller "R") er ændret til Histidin, His (eller "H"), hvor polypeptidet viser en forbedret rate af xylosefrigivelse på
- 10 mere end 90% ved sammenligning med assays der viser 63% frigivelse med SEQ ID NO: 2, med den samme enzymladning på 0,6 mg / g cellulose, og viser en forbedret rate af xylosefrigivelse ved sammenligning med assays med SEQ ID NO: 2, ved en reduceret enzymladning.
- 15 **2.** Isoleret, syntetisk eller rekombinant nukleinsyre ifølge krav 1:
- A) hvor nukleinsyren har en eller flere yderligere nukleotidrestændringer valgt fra
 - a) nukleinsyrekodonet, der koder for aminosyrerest 4 af SEQ ID NO: 2, er ændret fra ACC til AAC, og aminosyrerest 4 er ændret fra T til N;
 - 20 b) nukleinsyrekodonet, der koder for aminosyrerest 4 af SEQ ID NO: 2, er ændret fra ACC til CGC, og aminosyrerest 4 er ændret fra T til R;
 - c) nukleinsyrekodonet, der koder for aminosyrerest 4 af SEQ ID NO: 2, er ændret fra ACC til CAC, og aminosyrerest 4 er ændret fra T til H;
 - d) nukleinsyrekodonet, der koder for aminosyrerest 9 af SEQ ID NO: 2, er
 - 25 ændret fra CCC til GAC, og aminosyrerest 9 er ændret fra P til D;
 - e) nukleinsyrekodonet, der koder for aminosyrerest 17 af SEQ ID NO: 2, er ændret fra TTC til GTC, og aminosyrerest 17 er ændret fra F til V;
 - f) nukleinsyrekodonet, der koder for aminosyrerest 21 af SEQ ID NO: 2, er ændret fra TTC til TAC, og aminosyrerest 21 er ændret fra F til Y;
 - 30 g) nukleinsyrekodonet, der koder for aminosyrerest 33 af SEQ ID NO: 2, er ændret fra CTG til GCG, og aminosyrerest 33 er ændret fra L til A;
 - h) nukleinsyrekodonet, der koder for aminosyrerest 44 af SEQ ID NO: 2, er ændret fra AGC til ACG, og aminosyrerest 44 er ændret fra S til T;
 - i) nukleinsyrekodonet, der koder for aminosyrerest 63 af SEQ ID NO: 2, er
 - 35 ændret fra ATC til GTC, og aminosyrerest 63 er ændret fra I til V;

- j) nukleinsyrekodonet, der koder for aminosyrerest 73 af SEQ ID NO: 2, er ændret fra GGC til TAC, og aminosyrerest 73 er ændret fra G til Y;
- k) nukleinsyrekodonet, der koder for aminosyrerest 73 af SEQ ID NO: 2, er ændret fra GGC til GAG, og aminosyrerest 73 er ændret fra G til E;
- 5 l) nukleinsyrekodonet, der koder for aminosyrerest 73 af SEQ ID NO: 2, er ændret fra GGC til GTC, og aminosyrerest 73 er ændret fra G til V;
- m) nukleinsyrekodonet, der koder for aminosyrerest 108 af SEQ ID NO: 2, er ændret fra TTC til AAG, og aminosyrerest 108 er ændret fra F til K;
- 10 n) nukleinsyrekodonet, der koder for aminosyrerest 125 af SEQ ID NO: 2, er ændret fra CAG til TAC, og aminosyrerest 125 er ændret fra Q til Y;
- o) nukleinsyrekodonet, der koder for aminosyrerest 150 af SEQ ID NO: 2, er ændret fra GTA til GCC, og aminosyrerest 150 er ændret fra V til A;
- p) nukleinsyrekodonet, der koder for aminosyrerest 188 af SEQ ID NO: 2, er ændret fra AGC til GAG, og aminosyrerest 188 er ændret fra S til E; og
- 15 q) nukleinsyrekodonet, der koder for aminosyrerest 189 af SEQ ID NO: 2, er ændret fra TCC til CAG, og aminosyrerest 189 er ændret fra S til Q; eller
- B) hvor nukleinsyren koder for mindst et polypeptid med mindst en yderligere aminosyrerestændring valgt fra:
- 20 a) aminosyreresten 4 er ændret fra T til N;
- b) aminosyreresten 4 er ændret fra T til R;
- c) aminosyreresten 4 er ændret fra T til H;
- d) aminosyreresten 9 er ændret fra P til D;
- e) aminosyreresten 17 er ændret fra F til V;
- 25 f) aminosyreresten 21 er ændret fra F til Y;
- g) aminosyreresten 33 er ændret fra L til A;
- h) aminosyreresten 44 er ændret fra S til T;
- i) aminosyreresten 63 er ændret fra I til V;
- j) aminosyreresten 73 er ændret fra G til Y;
- 30 k) aminosyreresten 73 er ændret fra G til E;
- l) aminosyreresten 73 er ændret fra G til V;
- m) aminosyreresten 108 er ændret fra F til K;
- n) aminosyreresten 125 er ændret fra Q til Y;
- o) aminosyreresten 150 er ændret fra V til A;
- 35 p) aminosyreresten 188 er ændret fra S til E; og

- q) aminosyreresten 189 er ændret fra S til Q;
 hvor nukleinsyren af A) eller B) omfatter en sekvens med mindst 95%
 sekvensidentitet med SEQ ID NO: 1, og den koder for et polypeptid, der viser en
 forbedret rate af xylosefrigivelse på mere end 90% ved sammenligning med
 5 assays, der viser 63% frigivelse med SEQ ID NO: 2, ved den samme
 enzymladning på 0,6 mg / g cellulose, og viser en forbedret rate af
 xylosefrigivelse ved sammenligning med assays med SEQ ID NO: 2, ved en
 reduceret enzymladning.
- 10 **3.** Isoleret, syntetisk eller rekombinant nukleinsyre ifølge krav 1 eller 2, hvor
 nukleinsyren endvidere omfatter en eller flere nukleotidændringer, således at
 nukleinsyren koder for et polypeptid med mindst en konservativ
 aminosyresubstitution og bevarer dets xylanaseaktivitet; og, eventuelt, hvor den
 mindst ene konservative aminosyresubstitution omfatter at substituere en
 15 aminosyre med en anden aminosyre med tilsvarende karakteristika; eller, en
 konservativ substitution omfatter: erstatning af en alifatisk aminosyre med en
 anden alifatisk aminosyre; erstatning af en serin med en threonin eller vice versa;
 erstatning af en sur rest med en anden sur rest; erstatning af en rest der bærer
 en amidgruppe med en anden rest der bærer en amidgruppe; udskiftning af en
 20 basisk rest med en anden basisk rest; eller erstatning af en aromatisk rest med
 en anden aromatisk rest; og hvor nukleinsyren omfatter en sekvens med mindst
 95% sekvensidentitet med SEQ ID NO: 1, og den koder for et polypeptid, der
 viser en forbedret rate af xylosefrigivelse på mere end 90% ved sammenligning
 med assays der viser 63% frigivelse med SEQ ID NO: 2, ved den samme
 25 enzymladning på 0,6 mg / g cellulose, og viser en forbedret rate af
 xylosefrigivelse ved sammenligning med assays med SEQ ID NO: 2, ved en
 reduceret enzymladning.
- 4.** Isoleret, syntetisk eller rekombinant nukleinsyre ifølge et hvilket som helst
 30 foregående krav,
 A) hvor nukleinsyren endvidere omfatter en heterolog nukleinsyresekvens
 eller endvidere omfatter en heterolog nukleinsyresekvens der koder for en
 heterolog polypeptidsekvens;
 B) hvor nukleinsyren er fra (A), og hvor den heterologe polypeptidsekvens
 35 omfatter, eller består af:

- (i) en heterolog signalsekvens, et heterologt kulhydratbindende modul (CBM), et heterologt dockerin-domæne, et heterologt katalytisk domæne (CD), eller en kombination deraf;
- (ii) sekvensen fra (i), hvor den heterologe signalsekvens, det
- 5 kulhydratbindende modul eller det katalytiske domæne (CD) er afledt fra et heterologt enzym; eller,
- (iii) et mærke, en epitop, et målretningspeptid, en spaltelig sekvens, en detekterbar del eller et enzym;
- C) hvor polypeptidet eller peptidet er fra (B), og hvor det heterologe
- 10 kulhydratbindende modul (CBM) omfatter, eller består af, et xylanbindende modul, et cellulosebindende modul, et ligninbindende modul, et xylosebindende modul, et mannansebindende modul, et xyloglucan-specifikt modul og/eller et arabinofuranosidasebindende modul; eller
- D) hvor polypeptidet eller peptidet er fra (C), og hvor den heterologe
- 15 signalsekvens målretter det kodede protein til en vakuole, det endoplasmatiske reticulum, en kloroplast eller et stivelsesgranulat;
- og hvor nukleinsyren koder for et polypeptid der viser en forbedret rate af xylosefrigivelse på mere end 90% ved sammenligning med assays der viser 63% frigivelse med SEQ ID NO: 2, ved den samme enzymladning på 0,6 mg /g
- 20 cellulose, og viser en forbedret rate af xylosefrigivelse ved sammenligning med assays med SEQ ID NO: 2, ved en reduceret enzymladning.

5. Isoleret, syntetisk eller rekombinant nukleinsyresekvens, hvilken er helt (fuldstændigt) komplementær med sekvensen ifølge et hvilket som helst af

25 kravene 1-4, over hele længden af den kodende region.

6. Ekspressionskassette, vektor, eller kloningsvehikel omfattende en nukleinsyre omfattende sekvensen ifølge et hvilket som helst af kravene 1 til 5,

hvor kloningsvehiklen eventuelt omfatter en virusvektor, et plasmid, en

30 fag, et fagmid, et cosmid, et fosmid, en bakteriofag, eller et kunstigt kromosom, eller virusvektoren eventuelt omfatter en adenovirusvektor, en retrovirusvektor eller en adeno-associeret virusvektor, eller, det kunstige kromosom omfatter et kunstigt bakteriekromosom (BAC), en bakteriofag PI-afledt vektor (PAC), et kunstigt gærkromosom (YAC), eller et kunstigt

35 padderdyrskromosom (MAC).

7. Transformeret celle omfattende som transgen en nukleinsyre ifølge et hvilket som helst af kravene 1 til 5, eller omfattende ekspressionskassetten, vektoren
5 eller kloningsvehiklen ifølge krav 6,

hvor cellen eventuelt er en bakteriecelle, en pattedyrscelle, en
svampecelle, en gærcele, en insektcelle eller en plantecelle.

8. Transgen plante, transgen plantedel eller transgent plantefrø, eller transgent
10 ikke-humant dyr, omfattende som transgen en nukleinsyre ifølge et hvilket som
helst af kravene 1 til 5, eller omfattende ekspressionskassetten, vektoren eller
kloningsvehiklen ifølge krav 6, eller den transformerede celle ifølge krav 7,

hvor planten eventuelt er en majsplante, en sorghumplante, en
kartoffelplante, en tomatplante, en hvedeplante, en oliefrøplante, en
15 rapsfrøplante, en sojabønneplante, en risplante, en bygplante, en græs, en
bomuldsplante, en bomuldsfrøplante, en palme, en sesamplante, en
jordnøddeplante, en solsikkeplante eller en tobaksplante;
eller dyret eventuelt er en mus, en rotte, en kanin, et får, en gris, en
kylling, en ged, en fisk eller en ko.

20

9. Isoleret, syntetisk eller rekombinant polypeptid eller peptid med en
xylanaseaktivitet, hvor polypeptidet eller peptidet omfatter en sekvens med
mindst 95%, 96%, 97%, 98%, 99%, eller mere, eller fuldstændig (100%),
sekvensidentitet med en sekvens, hvilken er SEQ ID NO:2 med mutationen, at
25 aminosyreresten 38 Arginin, Arg (eller "R") er ændret til Histidin, His (eller "H"),
hvor polypeptidet med xylanaseaktivitet viser en forbedret rate af xylosefrigivelse
på mere end 90% ved sammenligning med assays der viser 63% frigivelse med
SEQ ID NO: 2, ved den samme enzymladning på 0,6 mg / g cellulose, og viser en
forbedret rate af xylosefrigivelse ved sammenligning med assays med SEQ ID NO:
30 2, ved en reduceret enzymladning.

10. Isoleret, syntetisk eller rekombinant polypeptid eller peptid ifølge krav 9,
A) hvor polypeptidet eller peptidet har mindst en af følgende yderligere
aminosyrerestændringer:

35 a) aminosyreresten 4 er ændret fra T til N;

- b) aminosyreresten 4 er ændret fra T til R;
- c) aminosyreresten 4 er ændret fra T til H;
- d) aminosyreresten 9 er ændret fra P til D;
- e) aminosyreresten 17 er ændret fra F til V;
- 5 f) aminosyreresten 21 er ændret fra F til Y;
- g) aminosyreresten 33 er ændret fra L til A;
- h) aminosyreresten 44 er ændret fra S til T;
- i) aminosyreresten 63 er ændret fra I til V;
- j) aminosyreresten 73 er ændret fra G til Y;
- 10 k) aminosyreresten 73 er ændret fra G til E;
- l) aminosyreresten 73 er ændret fra G til V;
- m) aminosyreresten 108 er ændret fra F til K;
- n) aminosyreresten 125 er ændret fra Q til Y;
- o) aminosyreresten 150 er ændret fra V til A;
- 15 p) aminosyreresten 188 er ændret fra S til E; og
- q) aminosyreresten 189 er ændret fra S til Q

hvor polypeptidet eller peptidet omfatter en sekvens med mindst 95% identitet med SEQ ID NO: 2 og viser en forbedret rate af xylosefrigivelse på mere end 90% ved sammenligning med assays der viser 63% frigivelse med SEQ ID NO: 2, ved
 20 den samme enzymladning på 0,6 mg /g cellulose, og viser en forbedret rate af xylosefrigivelse ved sammenligning med assays med SEQ ID NO: 2, ved en reduceret enzymladning.

11. Isoleret, syntetisk eller rekombinant polypeptid eller peptid ifølge krav 9 eller
 25 10, hvor polypeptidet endvidere omfatter mindst en konservativaminosyresubstitution og bevarer dets xylanaseaktivitet; hvor, eventuelt, den mindst ene konservativeaminosyresubstitution omfatter at substituere en aminosyre med en anden aminosyre med tilsvarende karakteristika; eller, en konservativsubstitution omfatter: erstatning af en alifatisk
 30 aminosyre med en anden alifatisk aminosyre; erstatning af en serin med en threonin eller vice versa; erstatning af en sur rest med en anden sur rest; erstatning af en rest der bærer en amidgruppe med en anden rest der bærer en amidgruppe; udskiftning af en basisk rest med en anden basisk rest; eller erstatning af en aromatisk rest med en anden aromatisk rest; hvor polypeptidet
 35 eller peptidet omfatter en sekvens med mindst 95% identitet med SEQ ID NO: 2

og viser en forbedret rate af xylosefrigivelse på mere end 90% ved sammenligning med assays der viser 63% frigivelse med SEQ ID NO: 2, ved den samme enzymladning på 0,6 mg /g cellulose, og viser en forbedret rate af xylosefrigivelse ved sammenligning med assays med SEQ ID NO: 2, ved en
5 reduceret enzymladning.

12. Isoleret, syntetisk eller rekombinant polypeptid eller peptid ifølge krav 9, hvor polypeptidet eller peptidet er kodet af nukleinsyren ifølge krav 1.

10 **13.** Isoleret, syntetisk eller rekombinant polypeptid eller peptid ifølge et hvilket som helst af kravene 9 til 12, hvor

A) polypeptidet eller peptidet endvidere omfatter en heterolog amino- eller polypeptid- eller peptidsekvens;

15 B) hvor polypeptidet eller peptidet er af (A), og hvor den heterologe polypeptid- eller peptidsekvens omfatter, eller består af:

(a) en heterolog signalsekvens, et heterologt kulhydratbindende modul (CBM), et heterologt dockerin-domæne, et heterologt katalytisk domæne (CD), eller en kombination deraf;

20 (b) sekvensen fra (a), hvor den heterologe signalsekvens, det kulhydratbindende modul eller det katalytiske domæne (CD) er afledt fra et heterologt enzym; eller,

(c) et mærke, en epitop, et målretningspeptid, en spaltelig sekvens, en detekterbar del eller et enzym;

25 C) hvor polypeptidet eller peptidet er fra (B), og hvor det heterologe kulhydratbindende modul (CBM) omfatter, eller består af, et xylanbindende modul, et cellulosebindende modul, et ligninbindende modul, et xylosebindende modul, et mannansebindende modul, et xyloglucan-specifikt modul og/eller et arabinofuranosidasebindende modul; eller

30 D) hvor polypeptidet eller peptidet er fra (C), og hvor den heterologe signalsekvens målretter det kodede protein til en vakuole, det endoplasmatiske reticulum, en kloroplast eller et stivelsesgranulat;

hvor polypeptidet eller peptidet viser en forbedret rate af xylosefrigivelse på mere end 90% ved sammenligning med assays der viser 63% frigivelse med SEQ ID NO: 2, ved den samme enzymladning på 0,6 mg /g cellulose, og viser en forbedret
35 rate af xylosefrigivelse ved sammenligning med assays med SEQ ID NO: 2, ved en

reduceret enzymladning.

14. Proteinpræparat, eller et immobiliseret polypeptid, omfattende polypeptidet ifølge et hvilket som helst af kravene 9 til 13, hvor proteinpræparatet omfatter en væske, et fast stof eller en gel.

15. Isoleret, syntetisk eller rekombinant antistof der specifikt binder til polypeptidet ifølge et hvilket som helst af kravene 9 til 13, hvor antistoffet eventuelt er et monoklonalt eller et polyklonalt antistof, eller er et enkeltkædet antistof.

16. Industriel proces eller en fremgangsmåde til anvendelse af et polypeptid med en xylanaseaktivitet, hvor polypeptidet omfatter en aminosyresekvens ifølge et hvilket som helst af kravene 9 til 13, eller polypeptidet er kodet af en nukleinsyre ifølge et hvilket som helst af kravene 1 til 5; omfattende:

- (a) en fremgangsmåde til at hydrolysere, flydendegøre, opbryde eller fraktionere en xylan-, cellulose- eller hemicellulose-holdig sammensætning omfattende at bringe polypeptidet i kontakt med en sammensætning omfattende en xylan, en cellulose eller en hemicellulose under forhold, hvor polypeptidet hydrolyserer, flydendegør, opbryder eller fraktionereden xylan-, cellulose- eller hemicellulose-holdige sammensætning, hvor sammensætningen eventuelt omfatter en plantecelle eller en bakteriecelle,
- (b) en fremgangsmåde til at reducere mængden af lignin (delignificering), eller opløsning af en lignin, i et papir eller papirprodukt, et træ, træmasse eller træprodukt, eller en træ- eller papirgenanvendelsessammensætning, omfattende at bringe papiret eller papirproduktet, træet, træmassen eller træproduktet, eller træ- eller papirgenanvendelsessammensætningen i kontakt med polypeptidet;
- (c) en fremgangsmåde til at hydrolysere celluloser, hemicelluloser eller xylaner i en biomasse, et træ, et træprodukt, en papirmasse, papirprodukt eller papiraffald omfattende at bringe træet, træproduktet, papirmassen, papirproduktet eller papiraffaldet i kontakt med polypeptidet;
- (d) en fremgangsmåde til enzymatisk affarvning af papir-, hamp- eller hørmasse omfattende at bringe papir-, hamp- eller hørmassen i kontakt med polypeptidet og et blegemiddel, hvor affarvningsmidlet eventuelt

omfatter oxygen eller hydrogenperoxid under forhold som er egnede til enzymatisk affarvning;

(e) en fremgangsmåde til affarvning af en lignocellulosemasse omfattende at bringe lignocellulosemassen i kontakt med polypeptidet under forhold som er egnede til enzymatisk affarvning;

(f) en fremgangsmåde til enzymatisk afsværtning af papir, papiraffald, genbrugspapirprodukt, afsværnings-toner fra berøringsfrit trykt affaldspapir eller blandinger af berøringsfrit og berørings-trykt affaldspapir, omfattende at bringe papiret, paperaffaldet, genbrugspapirproduktet, det berøringsfrie trykte affaldspapir eller berøringsstrykte affaldspapir i kontakt med polypeptidet, under forhold som er egnede til enzymatisk afsværtning;

(g) en fremgangsmåde til affarvning eller afsværtning af avispapir omfattende at bringe avispapiret i kontakt med polypeptidet under forhold som er egnede til enzymatisk affarvning eller afsværtning; eller

(h) fremgangsmåde til at reducere lignin i en sammensætning valgt fra et træ eller et træprodukt, en træmasse, en kraftpapirmasse, papir, et papirprodukt eller en papirmasse, omfattende at bringe sammensætningen i kontakt med polypeptidet under forhold som er egnede til enzymatisk reduktion af ligninen.

17. Enzymblanding eller -cocktail omfattende

(a) mindst et polypeptid ifølge et hvilket som helst af kravene 9 til 13, eller et polypeptid kodet af en nukleinsyre ifølge et hvilket som helst af kravene 1 til 5, og et eller flere andre enzym(er); eller

(b) blandingen eller cocktailen fra (a), hvor det ene eller flere andre enzym(er) er en anden xylanase, en mannanase og/eller en glucanase, cellulaser, lipaser, esteraser, proteaser, eller endoglycosidaser, endo-beta-1,4-glucanaser, beta-glucanaser, endo-beta-1,3(4)-glucanaser, cutinaser, peroxidaser, catalaser, laccaser, amylaser, glucoamylaser, pectinaser, reductaser, oxidaser, phenoloxidaser, ligninaser, pullulanaser, arabinanaser, hemicellulaser, mannanaser, xyloglucanaser, xylanase, en mannanase og/eller en glucanase, pektinacetyleraser, rhamnogalacturonacetyleraser, polygalacturonaser, rhamnogalacturonaser, galactanaser, pektinlyaser, pektinmethylesteraser, cellobiohydrolaser og/eller transglutaminaser.

18. Proces til hydrolysering af xylaner, celluloser eller hemicelluloser i en hvilken som helst organisk forbindelse, plante eller træ eller træprodukt eller biprodukt, træaffald, papirmasse, papirprodukt eller papiraffald eller biprodukt omfattende
5 anvendelse af et polypeptid ifølge et hvilket som helst af kravene 9 til 13, eller et polypeptid kodet af en nukleinsyre ifølge et hvilket som helst af kravene 1 til 5, eller enzymblandingen eller cocktailen ifølge krav 17, eller en kombination deraf.

19. Flydende sammensætning omfattende

- 10 (a) (i) en alkohol og (ii) et polypeptid ifølge et hvilket som helst af kravene 9 til 13, eller et polypeptid kodet af en nukleinsyre ifølge et hvilket som helst af kravene 1 til 5, eller enzymblandingen eller cocktailen ifølge krav 17, eller en kombination deraf;
- (b) den flydende sammensætning fra (a), hvor alkoholen er eller omfatter
15 ethanol, propanol, butanol og/eller methanol; eller
- (c) den flydende sammensætning fra (a) eller (b) omfattende eller indeholdt i et brændstof, et biobrændstof, en syntetisk væske eller gas eller en syngas.

- 20 **20.** Enzymblanding eller cocktail ifølge krav 17, omfattende en endo-glucanase, en oligomerase I (beta-glucosidase), en CBHI (GH-familie 7), en CBH2 (GH-familie 6), en xylanase (GH-familie 11), en arabinofuranosidase, en xylanase (GH-familie 10) og en oligomerase II (beta-xylosidase), hvor mindst et af disse
25 enzymer er et polypeptid ifølge et hvilket som helst af kravene 9 til 13, eller et polypeptid kodet af en nukleinsyre ifølge et hvilket som helst af kravene 1 til 5.

DRAWINGS

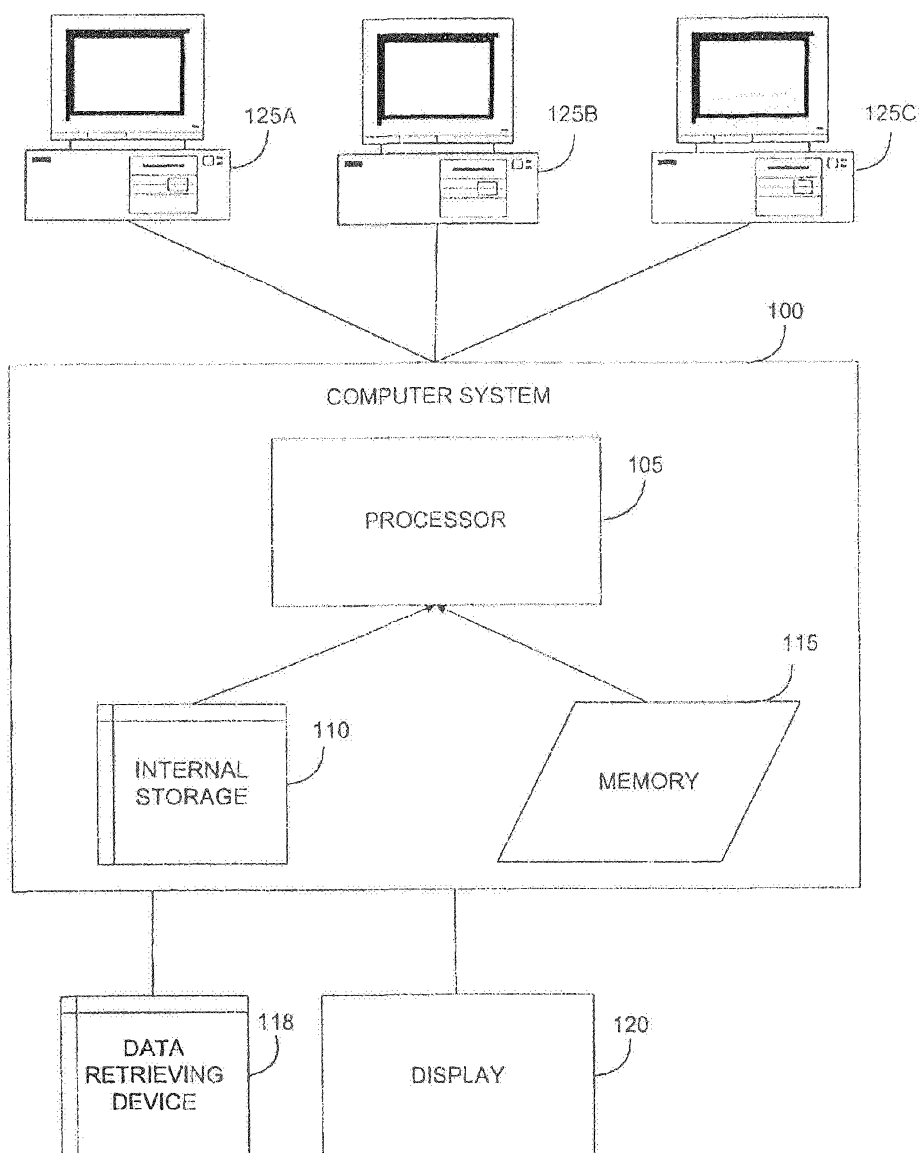


FIG. 1

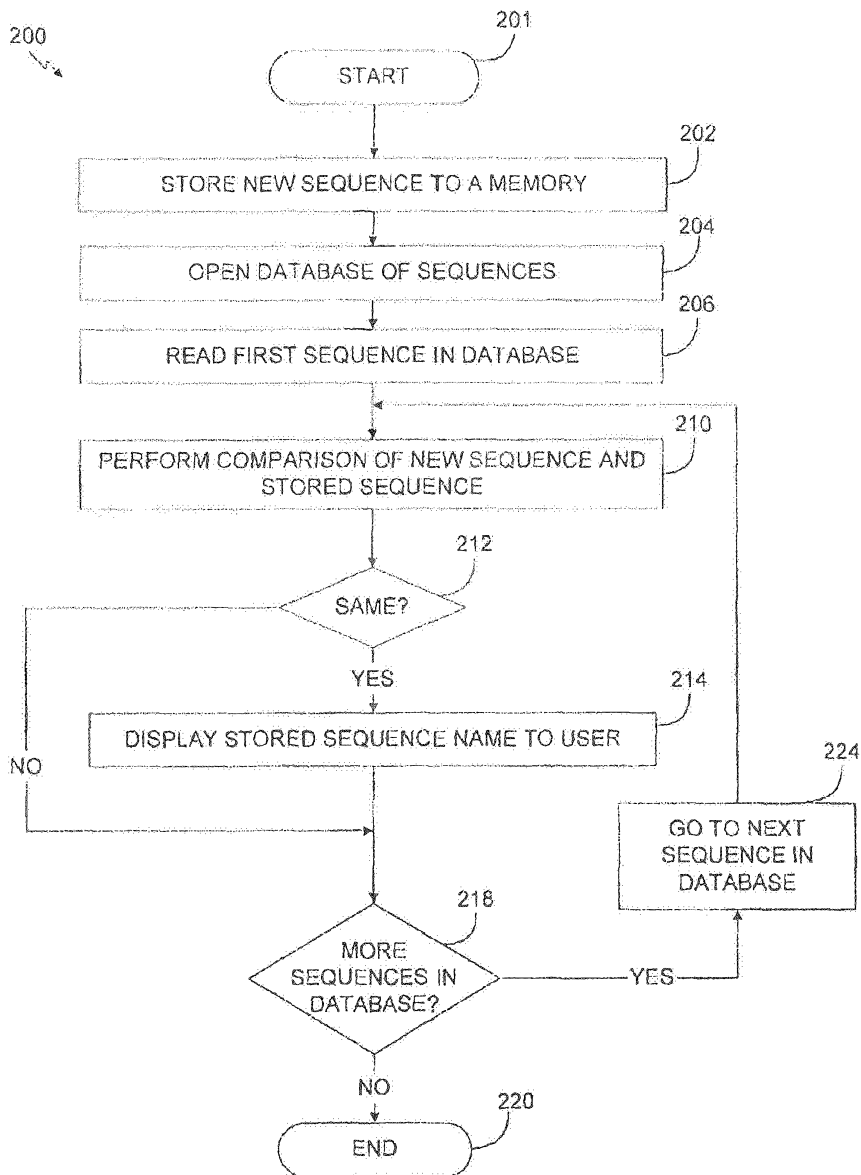


FIG. 2

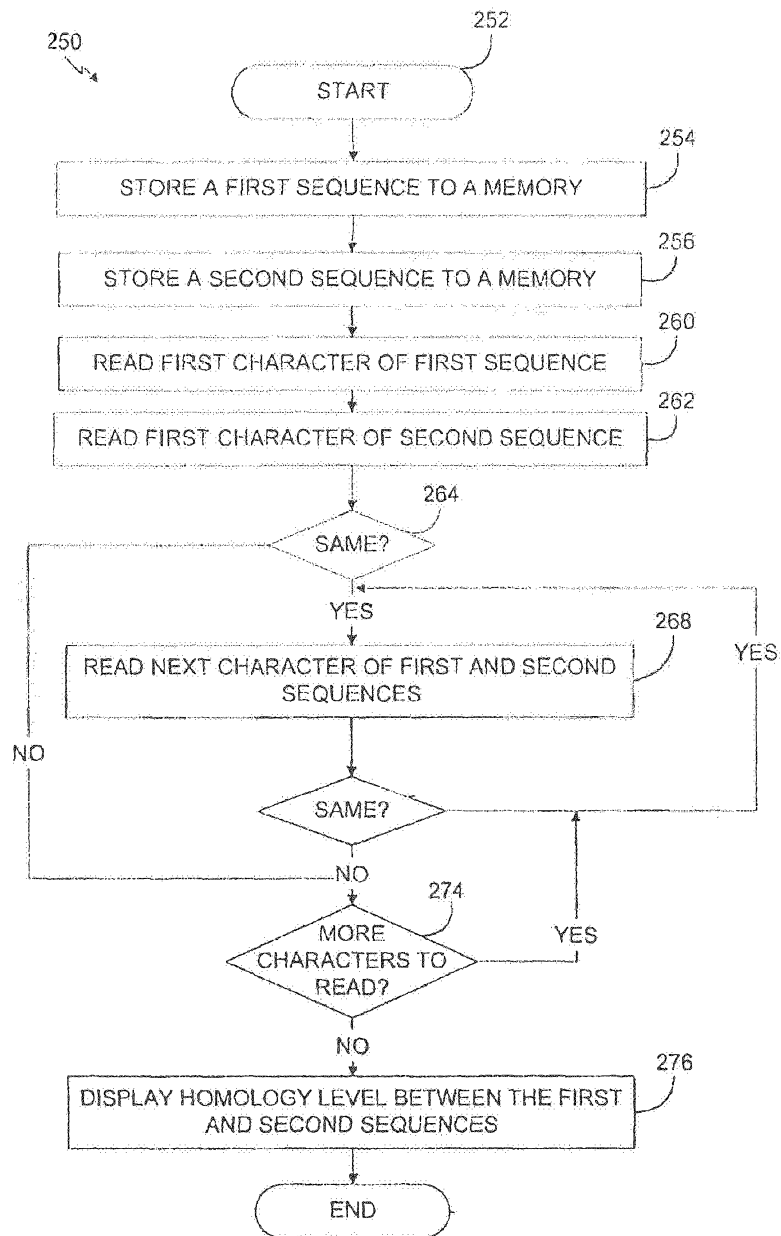


FIG. 3

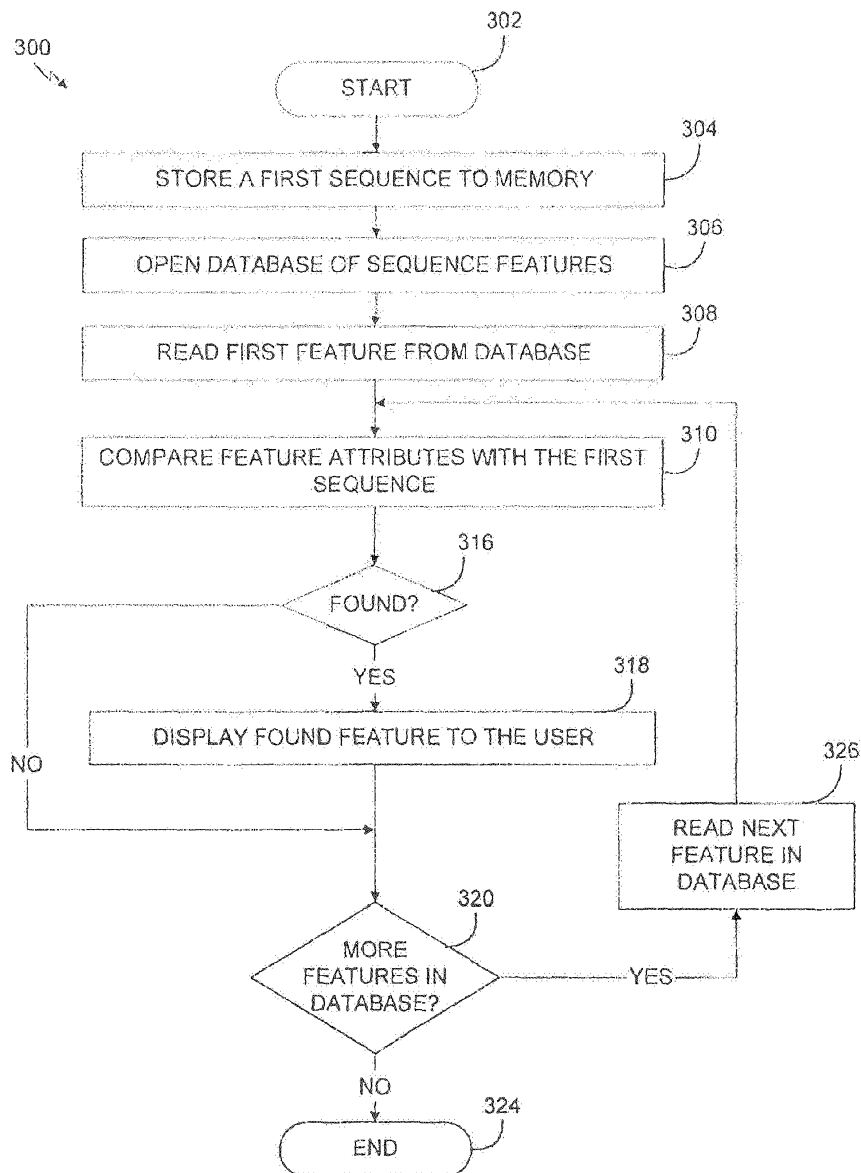


FIG. 4

Fig. 5

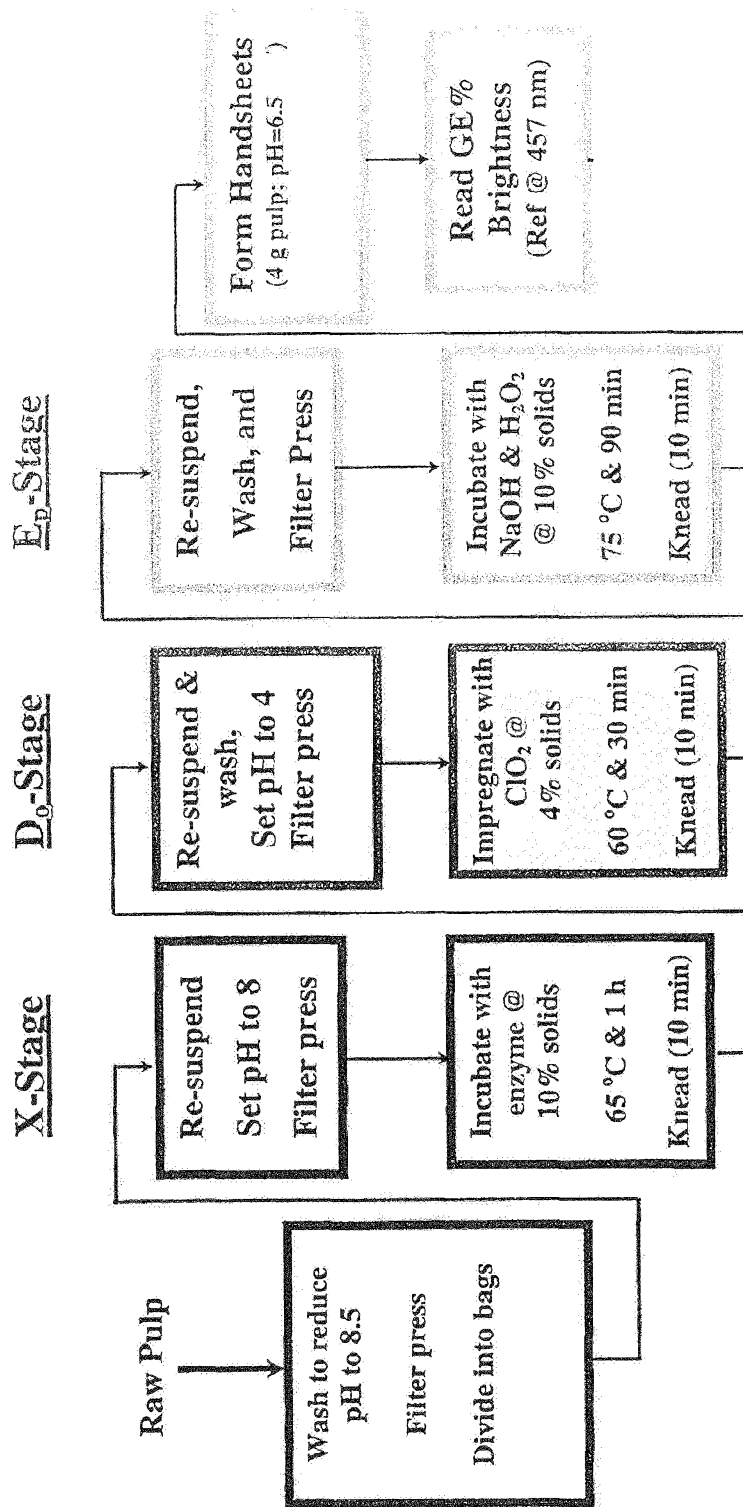


Fig. 6

