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(54) **Titre : BACTERIES MODIFIEES GENETIQUEMENT**  
 (54) **Title: GENETICALLY MODIFIED BACTERIA**

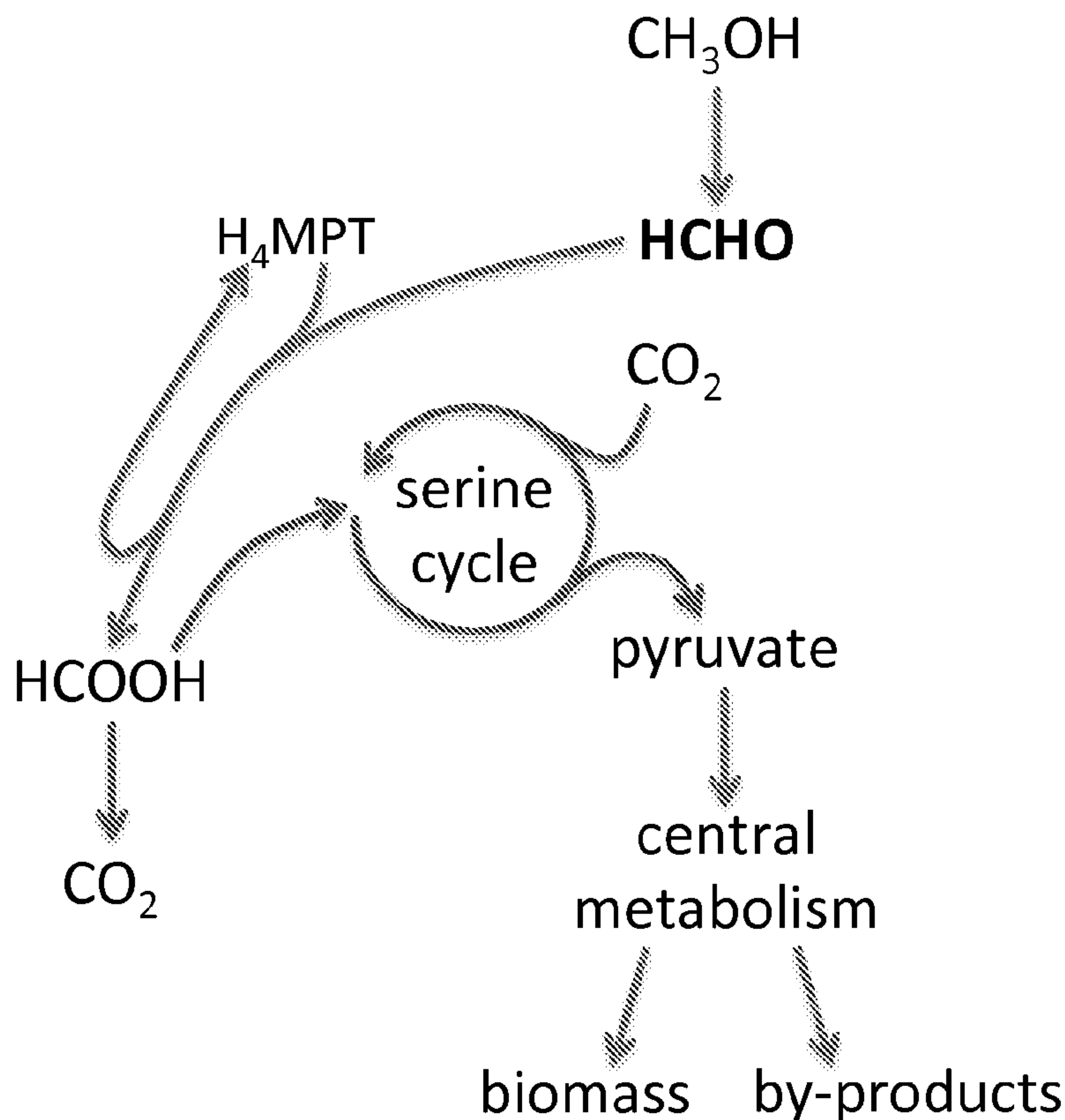


Figure 1

(57) **Abrégé/Abstract:**  
 The present invention relates to genetically modified methylotrophic bacteria.

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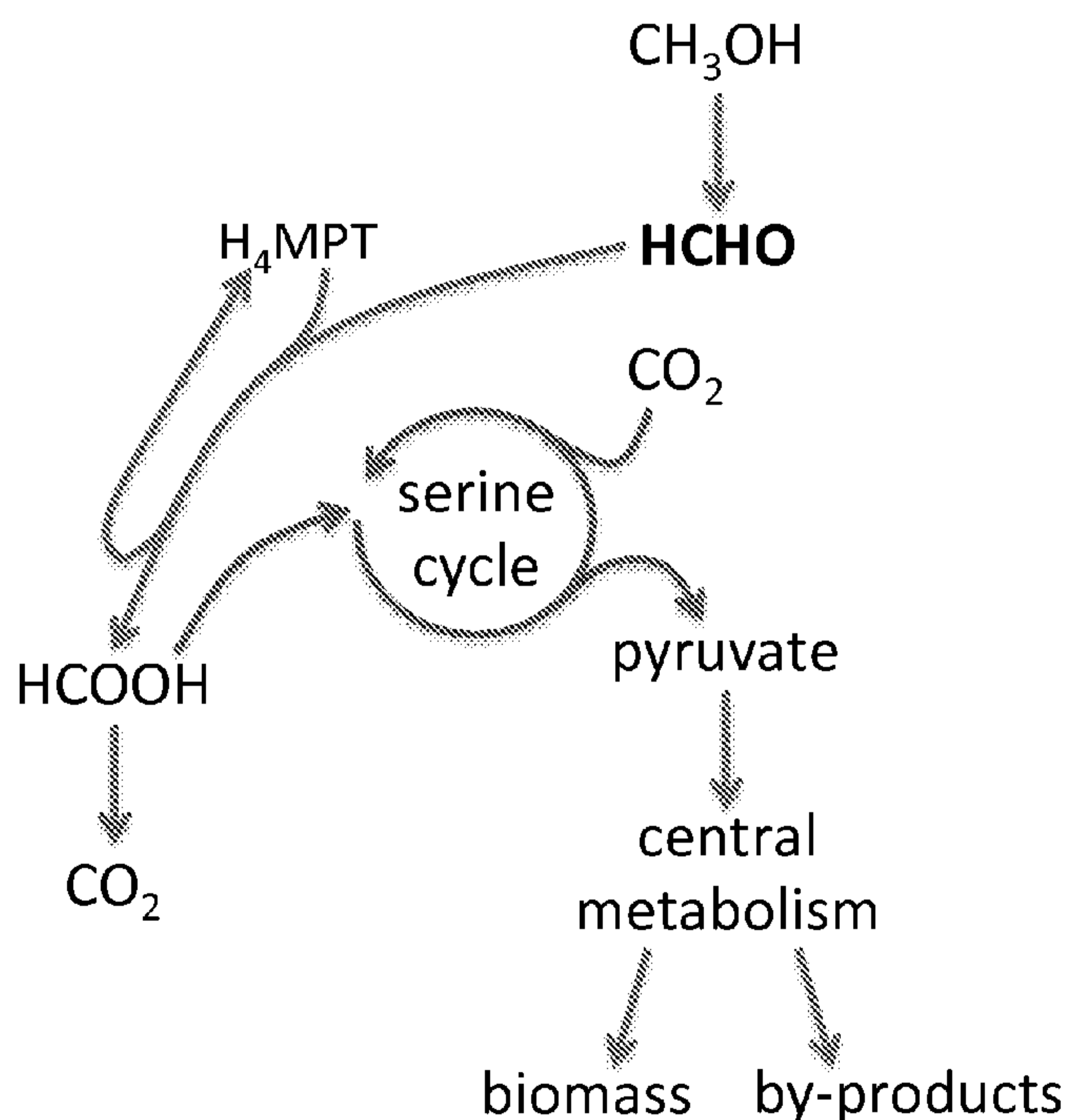
(54) **Title:** GENETICALLY MODIFIED BACTERIA(57) **Abstract:** The present invention relates to genetically modified methylotrophic bacteria.

Figure 1

## GENETICALLY MODIFIED BACTERIA

### RELATED APPLICATION DATA

This application claims priority to U.S. Provisional Patent Application No. 61/782,141 filed on  
5 March 14, 2013 and is hereby incorporated herein by reference in its entirety for all purposes.

### FIELD

The present invention relates in general to genetically modified bacteria, such as  
*Methylobacterium*, having an increased biomass producing capability. Such genetically modified  
bacteria can be used to produce useful chemical compounds from single-carbon (C<sub>1</sub>) compounds  
10 such as methanol (CH<sub>3</sub>OH) and methane (CH<sub>4</sub>).

### BACKGROUND

Methylotrophy is the ability of microorganisms, such as bacteria and certain fungi, to grow on  
reduced C<sub>1</sub> compounds like CH<sub>4</sub> and CH<sub>3</sub>OH as the sole carbon and energy source. As some C<sub>1</sub>  
assimilation pathways are highly inefficient, metabolic pathways diverting less feedstock to carbon  
15 dioxide (CO<sub>2</sub>) would be useful for production of beneficial carbon-based compounds. Therefore,  
genetically modified microorganisms optimized to reduce CO<sub>2</sub> waste are desirable.

### SUMMARY

Embodiments of the present disclosure are directed to the modification of pathways within bacteria  
to increase overall retention of carbon in central metabolism versus obligate waste to CO<sub>2</sub> when  
20 grown on C<sub>1</sub> compounds such as methanol. According to one aspect, genetically modified bacteria  
described herein have an increased substrate yield (grams carbon into biomass vs. grams carbon to  
CO<sub>2</sub>) compared to bacteria without the genetic modifications, including wild-type bacteria. In this  
aspect, biomass production is increased per unit substrate due to the increased efficiency of the  
genetically modified bacteria. According to an alternate aspect, genetically modified bacteria  
25 described herein produce less carbon dioxide and more carbon-based biomass compared to bacteria  
without the genetic modification, including wild-type bacteria. In this aspect, biomass production  
is increased per substrate.

According to certain aspects of the present disclosure, a recombinant bacterium is provided which  
has been genetically modified to disrupt the bacterium's H<sub>4</sub>MPT (tetrahydromethanopterin)

pathway for dissimilation of C<sub>1</sub> compounds like CH<sub>3</sub>OH. According to this aspect, one or more genes within the bacterium's H<sub>4</sub>MPT (tetrahydromethanopterin) pathway for C<sub>1</sub> dissimilation are removed, altered, inhibitor-bound, or otherwise prevented from being expressed. In this manner, the bacterium's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation is disrupted, inhibited, silenced or otherwise rendered ineffective for C<sub>1</sub> dissimilation. According to this aspect, C<sub>1</sub> dissimilation need not be completely prevented by disruption of the H<sub>4</sub>MPT pathway. Instead, genetic modification may result in some operation of the pathway, however, the genetic modification is intended to disable the H<sub>4</sub>MPT pathway relative to the unmodified H<sub>4</sub>MPT pathway.

According to certain aspects of the present disclosure, a recombinant bacterium is provided which has been genetically modified to disrupt the bacteria's serine cycle for lower carbon assimilation, such as C<sub>1</sub> assimilation. According to this aspect, one or more genes within the bacterium's serine cycle for C<sub>1</sub> assimilation are removed, altered, inhibitor-bound, or otherwise prevented from being expressed. In this manner, the bacterium's serine cycle for C<sub>1</sub> assimilation is disrupted, inhibited, silenced or otherwise rendered ineffective for C<sub>1</sub> assimilation. According to this aspect, C<sub>1</sub> assimilation need not be completely prevented by disruption of the serine cycle. Instead, genetic modification may result in some operation of the serine cycle, however, the genetic modification is intended to disable the serine cycle relative to the unmodified serine cycle.

According to certain aspects of the present disclosure, a bacterium may be genetically modified to disrupt the bacterium's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation or to disrupt the bacterium's serine cycle for C<sub>1</sub> assimilation. According to certain aspects of the present disclosure, a bacterium may be genetically modified to disrupt the bacterium's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation and to disrupt the bacterium's serine cycle for C<sub>1</sub> assimilation.

According to certain aspects, a bacterium may be genetically modified to include one or more genes from a RuMP (ribulose monophosphate) pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation. One or more genes from the RuMP pathway encode for certain C<sub>1</sub> assimilation enzymes and when included into the bacterium, the enzymes are expressed to carry out C<sub>1</sub> assimilation. One or more genes from the RuMP pathway encode for certain C<sub>1</sub> dissimilation enzymes and when included into the bacterium, the enzymes are expressed to carry out C<sub>1</sub> dissimilation. It is to be understood that not all genes attributed by those of skill in the art to the RuMP pathway need to be included into the genome of a bacteria to have the bacteria express the enzymes responsible for either C<sub>1</sub> assimilation or C<sub>1</sub> dissimilation.

According to one aspect, a bacterium may be genetically modified to disrupt the bacterium's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation and to include one or more genes from a RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation. According to a further aspect, a bacterium may be genetically modified to disrupt the bacterium's serine cycle for C<sub>1</sub> assimilation and to include one or more genes from an RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation. According to a still further aspect, a bacterium may be genetically modified to disrupt the bacterium's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation and to disrupt the bacterium's serine cycle for C<sub>1</sub> assimilation and to include one or more genes from an RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation.

According to certain aspects, the bacteria may be any which contain the serine cycle or the Calvin Cycle for C<sub>1</sub> assimilation, and either the H<sub>4</sub>MPT or thiol-based (e.g., glutathione, mycothiol) C<sub>1</sub> dissimilation pathway. According to the present disclosure, such bacteria may be improved by disabling the serine cycle, the Calvin Cycle, the H<sub>4</sub>MPT C<sub>1</sub> dissimilation pathway or the thiol-based (e.g., glutathione, mycothiol) C<sub>1</sub> dissimilation pathway and enabling the RuMP pathway for either C<sub>1</sub> assimilation, C<sub>1</sub> dissimilation or both. According to certain aspects, the bacteria may be a *Methylobacterium*. According to certain aspects, the bacteria may be a *Methylobacterium extorquens*. According to certain aspects, the bacteria may be a *Methylobacterium extorquens* AM1. According to certain aspects, the bacteria may be a *Methylobacterium extorquens* PA1.

According to one aspect, a *Methylobacterium extorquens* PA1 cell is genetically modified to disrupt the cell's H<sub>4</sub>MPT (tetrahydromethanopterin) pathway for C<sub>1</sub> dissimilation and to include a recombinant RuMP pathway for C<sub>1</sub> dissimilation.

According to an alternate aspect, a *Methylobacterium extorquens* PA1 cell is genetically modified to disrupt the cell's serine cycle for C<sub>1</sub> assimilation and to include a recombinant RuMP pathway for C<sub>1</sub> assimilation.

According to an alternate aspect, a *Methylobacterium extorquens* PA1 cell is genetically modified to disrupt the cell's H<sub>4</sub>MPT (tetrahydromethanopterin) pathway for C<sub>1</sub> dissimilation, to disrupt the cell's serine cycle for C<sub>1</sub> assimilation and to include a recombinant RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation.

## BRIEF DESCRIPTION OF THE DRAWINGS

The foregoing and other features and advantages of the present invention will be more fully understood from the following detailed description of illustrative embodiments taken in conjunction with the accompanying drawing in which:

- 5 Figure 1 is the metabolic pathways present in an unaltered serine cycle methylotroph. Native pathways contributing to both C<sub>1</sub> assimilation and dissimilation are shown in purple; pathways solely for C<sub>1</sub> dissimilation in red; pathways for C<sub>1</sub> assimilation in turquoise; pathways of central metabolism in grey.

- 10 Figure 2 is the metabolic pathways present in a serine cycle methylotroph in which the H<sub>4</sub>MPT-dependent formaldehyde oxidation pathway has been disabled (bright red X), and an RuMP cycle has been introduced, such as through the introduction and expression of HPS and PHI (dark blue). These introduce novel reactions for C<sub>1</sub> assimilation and dissimilation (brown), novel C<sub>1</sub> dissimilation in orange, and novel C<sub>1</sub> assimilation in green. Remaining colors are as described in Figure 1.

- 15 Figure 3 is the metabolic pathways present in a serine cycle methylotroph in which the native serine cycle for C<sub>1</sub> assimilation has been disabled (bright red X), and an RuMP cycle has been introduced, such as through the introduction and expression of HPS and PHI. Remaining colors are described in Figures 1 and 2.

- 20 Figure 4 is the metabolic pathways present in a serine cycle methylotroph in which the H<sub>4</sub>MPT-dependent formaldehyde oxidation pathway and the serine cycle have been disabled (bright red Xs), eliminating both C<sub>1</sub> assimilation and dissimilation functions, and an RuMP cycle has been introduced, such as through the introduction and expression of HPS and PHI. Remaining colors are described in Figures 1 and 2.

- 25 Figure 5 shows the plasmid map of the construct made to knockout *hprA* in the serine cycle and to replace it with the genes encoding hexulose 6-phosphate synthase (HPS) and hexulose 6-phosphate isomerase (PHI) from *Methylococcus capsulatus* Bath. The construct was made on the backbone of pCM433 [GenBank:EU118176] which is a 8081 bp long Cam<sup>R</sup>, Tet<sup>R</sup>, Amp<sup>R</sup> vector with a ColE1 origin of replication and an IncP origin of transfer (Marx, *BMC Research Notes* 1:1 (2008)).

Figure 6 shows the plasmid map of the construct made to knockout *hprA* in the serine cycle and to replace it with the genes encoding hexulose 6-phosphate synthase (HPS) and hexulose 6-phosphate isomerase (PHI) from *Methylococcus capsulatus* Bath. The construct was made on the backbone of pCM433 [*GenBank*:EU118176] which is a 8081 bp long Cam<sup>R</sup>, Tet<sup>R</sup>, Amp<sup>R</sup> vector with a ColE1  
5 origin of replication and an IncP origin of transfer (Marx, *BMC Research Notes* 1:1 (2008)).

Figure 7 shows the plasmid map of the construct made to knockout *mptG* in *M. extorquens* PA1. The construct was made on the backbone of pCM433 [*GenBank*:EU118176] which is a 8081 bp long Cam<sup>R</sup>, Tet<sup>R</sup>, Amp<sup>R</sup> vector with a ColE1 origin of replication and an IncP origin of transfer (Marx, *BMC Research Notes* 1:1 (2008)).

10

## DETAILED DESCRIPTION

Embodiments of the present disclosure include a recombinant host microorganism that includes one or more genetic modifications which result in reduced carbon dioxide production and concomitant increase in flow to biomass or by-product production. Embodiments of the present disclosure include a recombinant host microorganism that includes one or more genetic  
15 modifications that result in increased rate of cell proliferation. Embodiments of the present disclosure include a recombinant host microorganism that includes one or more genetic modifications that result in increased biomass production per substrate.

Embodiments of the present disclosure include recombinant host microorganisms described herein that have been further genetically modified to include a biosynthetic pathway for a target carbon-  
20 containing compound. According to one aspect, the recombinant host microorganisms have been genetically modified to reduce use of a carbon feedstock to produce carbon dioxide resulting in an increased production of carbon biomass by the recombinant host microorganisms. When a recombinant host microorganism has been further genetically modified to include a biosynthetic pathway for a target carbon-containing compound, the recombinant host microorganism produces  
25 the target compound in a greater amount compared to a host organism that has not been recombinantly modified as described herein.

Aspects of the present disclosure relate to recombinant methylotrophic microorganisms being genetically modified to increase overall biomass production from lower carbon-containing compounds, such as C<sub>1</sub> compounds. Methylotrophic microorganisms are those, such as bacteria  
30 and certain fungi, which grow on C<sub>1</sub> compounds like CH<sub>4</sub> and CH<sub>3</sub>OH as the carbon and energy

source. Methylophilic microorganisms may also include those which, in addition to C<sub>1</sub> compounds, grow on lower carbon compounds such as C<sub>2</sub> to C<sub>5</sub> carbon compounds.

Standard recombinant DNA and molecular cloning techniques used herein are well known in the art and are described in Sambrook, J., Fritsch, E.F. and Maniatis, T., *Molecular Cloning: A Laboratory Manual*, 2<sup>nd</sup> ed.; Cold Spring Harbor Laboratory: Cold Spring Harbor, N.Y., (1989) and by Silhavy, T.J., Bennis, M.L. and Enquist, L.W., *Experiments with Gene Fusions*; Cold Spring Harbor Laboratory: Cold Spring Harbor, N.Y., (1984); and by Ausubel, F.M. et. al., *Current Protocols in Molecular Biology*, Greene Publishing and Wiley-Interscience (1987) each of which are hereby incorporated by reference in their entireties.

Additional useful methods are described in manuals including Advanced Bacterial Genetics (Davis, Roth and Botstein, Cold Spring Harbor Laboratory, 1980), Experiments with Gene Fusions (Silhavy, Berman and Enquist, Cold Spring Harbor Laboratory, 1984), Experiments in Molecular Genetics (Miller, Cold Spring Harbor Laboratory, 1972) Experimental Techniques in Bacterial Genetics (Maloy, in Jones and Bartlett, 1990), and A Short Course in Bacterial Genetics (Miller, Cold Spring Harbor Laboratory 1992) each of which are hereby incorporated by reference in their entireties.

Methylophilic microorganisms may be genetically modified to delete genes or incorporate genes by methods known to those of skill in the art. Genes within the methylophilic microorganisms to be inhibited are known to those of skill in the art or may be determined using methods known to those of skill in the art. Genes, or homologs thereof, to be added to the genome of the methylophilic microorganisms are known to those of skill in the art or may be identified and obtained using methods known to those of skill in the art. Vectors and plasmids useful for transformation of some host cells may be common and commercially available from companies such as Invitrogen Corp. (Carlsbad, CA), Stratagene (La Jolla, CA) and New England Biolabs, Inc. (Beverly, MA). Other organisms will require broad-host-range vectors such as the pCM series generated from a minimal IncP replicon (Marx and Lidstrom, *Microbiology* 147-2065-2075 (2001)), or that take advantage of conjugal transfer to introduce suicide vectors (Marx and Lidstrom, *BioTechniques* 33:1062-1067 (2002); Marx, *BMC Research Notes* 1:1 (2008)).

Typically, the vector or plasmid contains sequences directing transcription and translation of a relevant gene, a selectable marker, and sequences allowing autonomous replication or chromosomal integration. Suitable vectors comprise a region 5' of the gene which harbors

transcriptional initiation controls and a region 3' of the DNA fragment which controls transcription termination. Both control regions may be derived from genes homologous to the transformed host cell, although it is to be understood that such control regions may also be derived from genes that are not native to the species chosen as a production host.

5 Initiation control regions or promoters, which are useful to drive expression of the relevant pathway coding regions in the desired host cell are numerous and familiar to those skilled in the art. Virtually any promoter capable of driving these genetic elements is suitable for the present invention including, but not limited to, *lac*, *ara*, *tet*, *trp*, *IP<sub>L</sub>*, *IP<sub>R</sub>*, *T7*, *tac*, and *trc* (useful for expression in *Escherichia coli* and *Pseudomonas*); or *mxoF*, and *tacA* for *Methylobacterium* spp.  
10 (Marx and Lidstrom, *Microbiology* 147:2065-2075 (2001); Chou et al., *PLoS Genet.* 5:e1000652 (2009)). Termination control regions may also be derived from various genes native to the preferred hosts, or introduced from *E. coli*, such as *t<sub>rrnB</sub>* or *t<sub>T7</sub>* (Marx and Lidstrom, *Microbiology* 150:9-19 (2004)).

Certain vectors are capable of replicating in a broad range of host bacteria and can be transferred  
15 by conjugation. The complete and annotated sequence of pRK404 and three related vectors- pRK437, pRK442, and pRK442(H) are available. These derivatives have proven to be valuable tools for genetic manipulation in Gram-negative bacteria (Scott et al., *Plasmid* 50(1):74-79 (2003)). Several plasmid derivatives of broad-host-range Inc P4 plasmid RSF1010 are also available with promoters that can function in a range of Gram-negative bacteria. Plasmid pAYC36 and pAYC37,  
20 have active promoters along with multiple cloning sites to allow for the heterologous gene expression in Gram-negative bacteria.

Vectors useful for the transformation of *E. coli* are common and commercially available. For example, the desired genes may be isolated from various sources, cloned onto a modified pUC19 vector and transformed into *E. coli* host cells. Alternatively, the genes encoding a desired  
25 biosynthetic pathway may be divided into multiple operons, cloned onto expression vectors, and transformed into various *E. coli* strains.

The various genes for a desired biosynthetic or other desired pathway may be assembled into any suitable vector, such as those described above. The codons can be optimized for expression based on the codon index deduced from the genome sequences of the host strain, such as for *M.*  
30 *extorquens* (Agashe et al., *Mol. Biol. Evol.* 30:549-560 (2013)).

Methylotrophic microorganisms which may serve as host cells and which may be genetically modified to produce recombinant methylotrophic microorganisms include methylotrophic bacteria and methylotrophic fungi.

5 Methylotrophic microorganisms which may serve as host cells and which may be genetically modified to produce recombinant methylotrophic microorganisms as described herein may include one or members of the genera *Clostridium*, *Rhodobacter*, *Xanthobacter*, *Brevibacterium*, *Mycobacterium*, *Amycolaptosis*, *Bacillus*, *Methylobacillus*, *Methylomicrobium*, *Methylotenera*, *Paracoccus*, *Methylocella*, *Methylacidiphium*, *Methylobacterium*, *Methylococcus*, *Methylobacter*, *Methylibium*, *Leisingera*, *Methylophilus*, *Methylosulfonomonas*, *Hyphomicrobium*, *Methylocystis*,  
10 *Methylosinus*, *Methylomirabilis*, *Methylophaga* and *Methyloversatilis*.

Methylotrophic microorganisms which may serve as host cells and which may be genetically modified to produce recombinant methylotrophic microorganisms as described herein include bacteria including the serine cycle or the Calvin cycle for C<sub>1</sub> assimilation or the H<sub>4</sub>MPT pathway or the thiol-based (e.g., glutathione, mycothiol) C<sub>1</sub> dissimilation pathway.

15 Methylotrophic microorganisms which may serve as host cells and which may be genetically modified to produce recombinant methylotrophic microorganisms as described herein may include *Methylobacterium* spp. such as *M. extorquens* AM1, *M. extorquens* PA1, *M. extorquens* DM4, *M. extorquens* CM4, *M. populi* BJ001, *M. nodulans* ORS 2060, *Methylobacterium* spp. 4-46, *M. radiotolerans* JCM 2831, *Paracoccus denitrificans* PD122, *Xanthobacter autotrophicus* Py2, and  
20 *Hyphomicrobium denitrificans*.

*Methylobacterium extorquens* AM1 is a well-characterized bacterium and is described in (Peel and Quayle, *Biochem. Journal* 81(3): 465-469 (1961)). The genes encoding enzymes involved in C<sub>1</sub> growth of *Methylobacterium extorquens* AM1 are described in (Chistoserdova *et al.*, *J. Bacteriol.* 185(10): 2980-2987 (2003)).

25 *Methylobacterium extorquens* PA1 is a well-characterized bacterium and is described in (Knief *et al.*, *Microb. Ecol.* 60: 440-452 (2010)). The genes encoding enzymes involved in C<sub>1</sub> growth of *Methylobacterium extorquens* PA1 are described in (Marx *et al.*, *J. Bacteriol.* 194: 4746-4748 (2012)).

30 According to certain aspects, a carbon-containing compound is used as a feedstock or growth substrate for the recombinant microorganisms described herein. Suitable carbon containing

compounds useful as a feedstock include any C<sub>1</sub> compounds, or multi-C compounds that contain C<sub>1</sub> units such as methylated amines, methylated sulfhydryls, methoxy groups, etc. Such carbon-containing compounds include methane, methanol, methylamine hydrochloride, sodium formate, betaine, sarcosine, methanethiol, and formaldehyde. Exemplary carbon-containing compounds  
5 useful as a feedstock include methane and methanol.

According to certain aspects of the present disclosure a *Methylobacterium*, such as *M. extorquens* PA1, is genetically modified to produce a recombinant *Methylobacterium*, such as a recombinant *M. extorquens* PA1 cell. According to certain aspects, a *Methylobacterium* is genetically modified to disrupt the bacterial cell's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation. The bacterial cell's H<sub>4</sub>MPT  
10 pathway for C<sub>1</sub> dissimilation is well characterized as provided in (Marx et al., *J. Bacteriol.* 185:7160-7168 (2003)). A summary of the genes and proteins characterizing the H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation is detailed in the text below and shown in Figure 1.

According to this aspect, disruption of the cell's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation includes altering the pathway to alter or otherwise eliminate one or more genes within the pathway in a  
15 manner to prevent C<sub>1</sub> dissimilation by the pathway. One or more genes within the H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation that can be altered or otherwise eliminated include *fae*, *mtdB*, *fhcABCD*, *mptG*, *mch*, *dmrA*, ORF5, ORF7, ORF9, ORF17, ORF19, ORF20, ORF21, ORF22, and ORFY. According to one aspect, one or more genes within the H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation can be altered or otherwise eliminated using methods known to those of skill in the  
20 art. For example, such methods include cloning the upstream and downstream flanks of the gene to be deleted, altered, or modified by PCR, restriction enzyme mediated DNA digestion, and subsequent DNA ligation, in an allelic exchange vector that cannot replicate in the host strain; introducing the allelic exchange vector in the host through a tri-parental mating using a pRK2073 containing helper strain as described in Marx, *BMC Research Notes*, 1:1 (2008) hereby  
25 incorporated by reference in its entirety for all purposes.

According to certain aspects of the present disclosure a *Methylobacterium*, such as *M. extorquens* PA1, is genetically modified to produce a recombinant *Methylobacterium*, such as a recombinant *M. extorquens* PA1 cell. According to certain aspects, a *Methylobacterium* is genetically modified to disrupt the bacterial cell's serine cycle for C<sub>1</sub> assimilation. The bacterial cell's serine cycle for  
30 C<sub>1</sub> assimilation is well characterized as provided in (Chistoserdova et al., *J. Bacteriol.* 185(10): 2980-2987 (2003)). A summary of the genes and proteins characterizing the serine cycle for C<sub>1</sub> assimilation is detailed in the text below and shown in Figure 1.

According to this aspect, disruption of the cell's serine cycle for C<sub>1</sub> assimilation includes altering the pathway to alter or otherwise eliminate one or more genes within the pathway in a manner to prevent C<sub>1</sub> assimilation by the pathway. One or more genes within the serine cycle for C<sub>1</sub> assimilation that can be altered or otherwise eliminated include *glyA*, *hprA*, *sga*, *glc*, *eno*, *ppc*, *mdh*,  
5 and *mtkAB*. According to one aspect, one or more genes within the serine cycle for C<sub>1</sub> assimilation can be altered or otherwise eliminated using methods known to those of skill in the art. For example, such methods include cloning the upstream and downstream flanks of the gene to be deleted, altered, or modified by PCR, restriction enzyme mediated DNA digestion, and subsequent DNA ligation, in an allelic exchange vector that cannot replicate in the host strain; introducing the  
10 allelic exchange vector in the host through a tri-parental mating using a pRK2073 containing helper strain as described in Marx, *BMC Research Notes*, 1:1 (2008)

According to certain aspects of the present disclosure a *Methylobacterium*, such as *M. extorquens* PA1, is genetically modified to produce a recombinant *Methylobacterium*, such as a recombinant *M. extorquens* PA1 cell. According to certain aspects, a *Methylobacterium* is genetically modified  
15 to include one or more genes to express proteins and enzymes responsible for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation. The genes (HPS and PHI) that need to be incorporated encode two enzymes (hexulose 6-phosphate synthase and hexulose 6-phosphate isomerase) that catalyze the first two steps of the RuMP pathway. While these two genes are generally found as two separate ORFs (Open Reading Frames) in methylotrophs using the RuMP pathway for C<sub>1</sub> assimilation and /or C<sub>1</sub>  
20 dissimilation, certain methylotrophs also use a single enzyme encoding a fused polypeptide with two domains – one with hexulose 6-phosphate synthase activity and the other with a hexulose 6-phosphate isomerase activity in the RuMP pathway for C<sub>1</sub> assimilation and/or C<sub>1</sub> dissimilation. According to one aspect, genes encoding the RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation and inserted into the genome of the *Methylobacterium*. The RuMP pathway for C<sub>1</sub>  
25 assimilation and C<sub>1</sub> dissimilation is well characterized as provided in (Ward et al., *PLoS Biology* 2:e303 (2004)). A summary of the genes and proteins characterizing the RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation is provided in Table 3 below.

Genes/Enzymes	Function
Hexulose 6-phosphate synthase (HPS)	Catalyzes the reaction of ribulose monophosphate and formaldehyde to hexulose 6-phosphate
Hexulose 6-phosphate isomerase (PHI)	Catalyzes the isomerization of hexulose 6-phosphate to fructose 6-phosphate
Phosphofructokinase (PFK)	Catalyzes the phosphorylation of fructose 6-phosphate to fructose 1,6-bisphosphate
Fructose bisphosphate aldolase (FBA)	Catalyzes the cleavage of fructose 1,6-bisphosphate to dihydroxyacetone 3-phosphate and glyceraldehyde 3-phosphate
Transketolase (TK)	Catalyzes the rearrangement of C <sub>3</sub> -C <sub>7</sub> phosphorylated sugars
Pentose phosphate epimerase	Catalyzes the epimerization of xylulose 5-phosphate to ribulose 5-phosphate
Pentose phosphate isomerase	Catalyzes the isomerization of ribose 5-phosphate to ribulose 5-phosphate
Glucose phosphate isomerase (GPI)	Catalyzes the isomerization of fructose 6-phosphate to glucose 6-phosphate
Glucose 6-phosphate dehydrogenase	Catalyzes the oxidation of glucose 6-phosphate to 6-phosphogluconate
Phosphogluconate dehydrase	Catalyzes the dehydration of 6-phosphogluconate to 2-keto, 3-deoxy, 6-phosphogluconate (KDPG)

2-keto, 3-deoxy, 6-phosphogluconate (KDPG) aldolase	Catalyzes the cleavage of 2-keto, 3-deoxy, 6-phosphogluconate (KDPG) to glyceraldehyde 3-phosphate and pyruvate
Sedoheptulose biphosphatase	Catalyzes the dephosphorylation of sedoheptulose 1,7-bisphosphate to sedoheptulose 7-phosphate
Phosphogluconate dehydrogenase	Catalyzes the decarboxylation of 6-phosphogluconate to ribulose 5-phosphate

According to this aspect, insertion of the RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation into the genome of the *Methylobacterium* includes inserting one or more genes within the pathway in a manner to promote C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation. One or more genes within the RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation that can promote C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation include a gene (HPS) encoding a hexulose 6-phosphate synthase and a gene (PHI) encoding a hexulose 6-phosphate isomerase or a gene encoding a fusion polypeptide containing a hexulose 6-phosphate synthase domain as well as a hexulose 6-phosphate isomerase domain. According to one aspect, one or more genes within the RuMP pathway for C<sub>1</sub> assimilation and C<sub>1</sub> dissimilation can be inserted within the genome of the *Methylobacterium* using methods known to those of skill in the art. For example, such methods include cloning the promoter, RBS (Ribosome Binding Site) and entire ORF (Open Reading Frame) as well as the transcription termination site of the genes to be inserted by PCR, restriction enzyme mediated DNA digestion, and subsequent DNA ligation, in an allelic exchange vector that cannot replicate in the host strain; introducing the allelic exchange vector in the host through a tri-parental mating using a pRK2073 containing helper strain as described in Marx, *BMC Research Notes*, 1:1 (2008).

According to certain aspects, recombinant host microorganisms described herein have been further genetically modified to include a biosynthetic pathway for a target carbon-containing compound. Certain useful target carbon-containing compounds include but are not limited to polyhydroxyalkanoates and related storage polymers (Borque et al., *Appl. Microbiol. Biotechnol.* 44:367-376 (1995)), polysaccharides (Oh et al., *Biotechnol. Bioeng.* 54:115-121 (1997)), amino acids such as serine (Sirirote et al., *J. Ferment. Technol.* 66:291-297 (1988)), liquid biofuels such as

ethanol and butanol, 1,4-butanediol, isoprenoids (Van Dien et al., *Appl. Environ. Microbiol.* 69:7563-7566 (2003)), and plant growth hormones such as indole-3-acetic acid (Omer et al., *Plant Growth Regul.* 43:93-96 (2004)) *trans*-zeatin (Koenig et al., *J. Bacteriol.* 184:1832-1842 (2002)). Furthermore, C<sub>1</sub>-based protein production could be enhanced due to higher biomass efficiencies for  
 5 active polypeptides such as enterocin P (Gutierrez et al., *FEMS Microbiol. Lett.* 248:125-131 (2005)), haloalkane dehalogenase (FitzGerald and Lidstrom, *Biotechnol. Bioeng.* 81:263-268 (2003)), and esterase (Choi et al., *Appl. Environ. Microbiol.* 72:753-759 (2006)).

Biosynthetic pathways for useful target carbon-containing compounds are known to those of skill in the art and include the pathway for the generation and incorporation of  $\beta$ -hydroxybutyryl-CoA  
 10 into polyhydroxybutyrate or related polymers (Korotkova and Lidstrom, *J. Bacteriol.* 184:1750-1758 (2001)), the biosynthetic pathway for conversion of isopentyl pyrophosphate into carotenoids (Van Dien et al., *Appl. Environ. Microbiol.* 69:7563-7566 (2003)), the isopentylation of adenines of some tRNAs by the *miaA* gene product to produce *trans*-zeatin (Koenig et al., *J. Bacteriol.* 184:1832-1842 (2002)), the pathway for indole-3-acetic acid from tryptophan (Omer et al., *Plant*  
 15 *Growth Regul.* 43:93-96 (2004)), enzymes such as pyruvate decarboxylase and alcohol dehydrogenase to generate ethanol (Deng and Coleman, *Appl. Environ. Microbiol.* 65:523-528 (1999)), butanol production from crotonyl-CoA via expression of enzymes such as Bcd, EtfAB, and AdhE2 from *Clostridium acetobutylicum* (Shen et al., *Appl. Environ. Microbiol.* 77:2905-2915 (2011)), or 1,4-butanediol from succinyl-CoA or  $\alpha$ -ketoglutarate via CoA-dependent succinate  
 20 semialdehyde dehydrogenase or 2-oxoglutarate decarboxylase, then 4-hydroxybutyrate dehydrogenase, 4-hydroxybutyryl-CoA transferase, 4-hydroxybutyryl-CoA reductase, and alcohol dehydrogenase (Yim et al., *Nat. Chem. Biol.* 7:445-452 (2011)). Protein production can be accomplished via strong expression systems for methylotrophic bacteria, such as those dependent upon the methanol dehydrogenase promoter ( $P_{mxaF}$ ) of *M. extorquens* AM1 such as pCM80,  
 25 pCM110, pCM160 (Marx and Lidstrom, *Microbiology* 147-2065-2075 (2001)), or inducer-regulated versions such as pH112 (Chou and Marx, *Cell Reports* 1:1-8 (2012)).

## EXAMPLE I

### RuMP Pathway for C<sub>1</sub> metabolism

The Ribulose Monophosphate pathway for C<sub>1</sub> metabolism is present in several methylotrophs such  
 30 as the obligate methylotroph *Methylobacillus flagellatus* KT and the methanotroph *Methylococcus capsulatus* Bath. Methylotrophs growing on methanol using the RuMP Pathway assimilate carbon

at the oxidation level of formaldehyde in comparison to formate, which is the intermediate that gets assimilated in methylotrophs using the serine cycle. Formaldehyde is condensed with ribulose monophosphate by an enzyme that is unique to this pathway called hexulose-6-phosphate synthase (HPS) to produce hexulose 6-phosphate. Hexulose 6-phosphate is isomerized into fructose 6-phosphate by another enzyme that is unique to this pathway called hexulose 6-phosphate isomerase (PHI). The fructose 6-phosphate can be assimilated either with certain enzymes that are involved in the pentose phosphate pathway or with certain enzymes that are involved in the Entner-Doudoroff pathway depending on the organism as listed in Table 1. If the fructose 6-phosphate is assimilated through the pentose phosphate pathway then two molecules of fructose 6-phosphate are converted to two molecules of fructose bisphosphate each of which is cleaved to give four molecules of triose phosphate in total. One of these four molecules of triose phosphate is assimilated into biomass by conversion to pyruvate, which leads to the production of one NADPH and one ATP. The three molecules of triose phosphate are rearranged with one more fructose 6-phosphate molecule through a series of reactions with transketolase, transaldolase/sedoheptulose phosphatase, and finally pentose phosphate isomerases to regenerate three molecules of ribulose monophosphate as listed in Table 1. If the fructose 6-phosphate is assimilated through enzymes of the Entner-Doudoroff pathway then one molecule of fructose 6-phosphate is cleaved by a series of reactions to pyruvate and a triose phosphate. The triose phosphate along with two other molecules of fructose 6-phosphate are rearranged through a series of reactions with transketolase, transaldolase/sedoheptulose phosphatase, and finally pentose phosphate isomerases to regenerate three molecules of ribulose monophosphate as listed in Table 1. The RuMP pathway used for the assimilation of C<sub>1</sub> compounds with fructose bis-phosphate aldolase (an enzyme also involved in the pentose phosphate pathway) and transaldolase as the key enzymes involved in cleavage and rearrangement respectively is extremely efficient. It is to be understood that although some bacteria may require only HPS and PHI, other bacteria may include additional enzymes or directed alteration of their expression. For one molecule of pyruvate produced 1 molecule of NADPH and 1 molecule of ATP are also produced. On the contrary, the serine cycle leads to the net consumption of 1 molecule of NADPH and 1 molecule of ATP per molecule of pyruvate produced.

The Ribulose monophosphate pathway can also be used for the dissimilation of C<sub>1</sub> compounds. The fructose 6-phosphate generated by HPS and PHI is isomerized to glucose monophosphate by a hexose phosphate isomerase. Glucose monophosphate is oxidized to 6-phosphogluconate coupled with the reduction of NADP<sup>+</sup> to NADPH. The 6-phosphogluconate thus formed is oxidized to

ribulose monophosphate with the release of CO<sub>2</sub> and reduction of another molecule of NADP<sup>+</sup> to NADPH by an enzyme called phosphogluconate dehydrogenase as listed in Table 1.

While the RuMP pathway produces 2 molecules of NADPH per molecule of formaldehyde dissimilated, the H<sub>4</sub>MPT pathway produces 1 molecule of NADPH and ~1 molecule of NADH per molecule of formaldehyde dissimilated. Purely thinking in terms of energetics of dissimilation, neither one has a higher energy output per unit carbon. However, the RuMP pathway for dissimilation of C<sub>1</sub> compounds involves fewer enzymes and does not require a complex cofactor like H<sub>4</sub>MPT that is used by the H<sub>4</sub>MPT dissimilatory pathway for C<sub>1</sub> compounds.

## EXAMPLE II

### 10 RuMP Pathway for C<sub>1</sub> metabolism: Strain 1

Strain 1- Replacing the native H<sub>4</sub>MPT (tetrahydromethanopterin) pathway for C<sub>1</sub> dissimilation with the RuMP pathway. The serine cycle will be kept intact. However, since no formate will be produced during dissimilation of C<sub>1</sub> by the RuMP pathway, the serine cycle will become redundant and the RuMP pathway will take over C<sub>1</sub> assimilation as well as shown in Figure 2.

15 As described above, the RuMP pathway and the H<sub>4</sub>MPT pathway are almost equivalent in terms of the net energetic output per molecule of methanol completely oxidized to CO<sub>2</sub>. However, taking into account the fact that there are fewer enzymes involved in C<sub>1</sub> dissimilation using the RuMP pathway than the H<sub>4</sub>MPT pathway as well as incorporating the cost of production of the complex cofactor in the H<sub>4</sub>MPT pathway, we postulate that the RuMP pathway might carry out dissimilation of C<sub>1</sub> compounds more efficiently. Dissimilation of C<sub>1</sub> compounds through the introduced dissimilatory RuMP pathway, however, cannot be coupled to the native serine cycle for assimilation. This is because the key metabolic intermediate that gets fed into the serine cycle – formate – is not produced during dissimilation of methanol through the RuMP pathway. Hence, despite being intact in strain 1, the serine cycle should play a very minor role in assimilation of C<sub>1</sub> compounds and flux to biomass should mainly be routed through the RuMP pathway as well. Strain 1 was constructed by eliminating the native H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation by deleting a gene involved in the biosynthesis of the cofactor – tetrahydromethanopterin- and replacing it with a genes encoding hexulose 6-phosphate synthase (HPS) as well as a hexulose 6-phosphate isomerase (PHI) from *M. capsulatus* Bath. It should be noted that only two genes – those encoding HPS and PHI- are needed in the *Methylobacterium* genome to allow the RuMP pathway to

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dissimilate C<sub>1</sub> compounds since all other genes in this pathway are already present in the host strain.

The strain was made with a construct as shown in Figure 5. The construct was generated as follows. The specific gene in the biosynthetic pathway of tetrahydromethanopterin that was knocked out is *mptG* (Mext\_1828 on the *M. extorquens* PA1 chromosome, ref) which is a ribofuranosylaminobenzene 5'-phosphate (RFAP) synthase (Chistoserdova et al., *J. Bacteriol.* 187:2508-2512 (2005)). The ORF encoding *mptG* is 1023 bases long and is found on the positive strand between bases 2049113 and 2050135 on the chromosome. The upstream flank of *mptG* for the construct (2048710-2049237) was amplified by a forward primer containing the recognition sequence for the restriction enzyme *Xba*I at the 5' end (ATCTAGACTTCCTCACCGTCGCTTCTTCG) and a reverse primer containing the recognition sequence for the restriction enzyme *Nde*I at the 5' end (TCATATGCGTTGAGATCGAGGAAGCCGA). The downstream flank of *mptG* for the construct (2049864-2050304) was amplified by a forward primer containing the recognition sequence for the restriction enzyme *Apa*I at the 5' end (ACTCGAGGCATCACCGCGATCCAGAAGC) and a reverse primer containing the recognition sequence for the restriction enzyme *Sac*I at the 5' end (TGAGCTCACGTCGAACGGGCTCATGT).

On investigating the genome sequence of *M. extorquens* PA1, it was evident that the only genes needed to introduce the dissimilatory RuMP cycle in the *M. extorquens* PA1 metabolic network were HPS and PHI. These genes were introduced from *M. capsulatus* Bath. In *M. capsulatus* Bath there are three genes encoding a HPS (MCA2738, MCA3043, and MCA3049 on the main chromosome of *Me. capsulatus* Bath), as well as three genes encoding a PHI (MCA2738, MCA3044, and MCA3050 on the main chromosome of *M. capsulatus* Bath). Previous work (Ward et al., *PLoS Biology* 2:e303 (2004)) has shown that most carbon flux enters the RUMP cycle through MCA3049 and MCA3050. Hence, these two genes were introduced in place of the *mptG* gene in *M. extorquens* PA1 for making Strain1. As stated above, MCA3049 encodes a hexulose 6-phosphate synthase (HPS) gene that is 648 bp long on the positive strand from position 3236945 to position 3237592 on the chromosome. MCA3050 encodes the hexulose 6-phosphate isomerase (PHI) gene that is 534 bp long on the positive strand from position 3237605 to position 3238138 on the chromosome. A forward primer with a recognition sequence for the restriction enzyme *Nde*I at the 5' end (TCATATGACCGGTGTCGCCAGGTACA) and a reverse primer with a recognition

sequence for the restriction enzyme *ApaI* at the 5' end (TGGGCCACGGACAACATGCCACGC) were used to amplify a region between position 3236701 and 3238196 on the *M. capsulatus* Bath chromosome containing the promoter, the RBS (ribosome binding site) as well as the entire ORF for MCA3049 and MCA 3050.

5 The final construct was made on the backbone of an allelic exchange vector (pCM433), as described in (Marx, *BMC Research Notes* 1:1 (2008)), in the order described below. The vector pCM433 was digested with restriction enzymes *NdeI* and *XbaI*, as was the insert, the *mptG* upstream flank. The 5' end of the digested vector was dephosphorylated using an Antarctic Phosphatase manufactured by NEB (New England Biolabs). The digested vector and insert were  
10 gel extracted using the Gel Extraction kit manufactured by Zymo Research Inc. A 3:1 molar ratio of the insert to vector was ligated using the Quick Ligase Kit manufactured by NEB (New England Biolabs). 1  $\mu$ l of the ligated mixture was transformed in 50  $\mu$ l of 10 $\beta$  chemically competent cells manufactured by NEB (New England Biolabs). The transformed cells were plated on LB + tetracycline (12.5  $\mu$ g/ml) + 1.2% agar plates and screened by PCR for the insert. Colonies that  
15 screened positive for the insert were grown in LB+ tetracycline (12.5  $\mu$ g/ml). The resulting plasmid with the *mptG* upstream flank in pCM433 was named pDN117. Plasmid extraction was done using the standard Miniprep kit manufactured by Qiagen. For the next round, a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of pDN117 was used as the vector and a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of the pCRII- Blunt- Topo vector containing *mptG* downstream flank as the insert. The same protocol described above was repeated.  
20 The resulting plasmid was named pDN128. For the final round, a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of pDN128 was used as the vector and a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of the pCRII- Blunt- Topo vector with the region of the *Methylococcus capsulatus* Bath containing the MCA3049 and MCA3050 ORF, promoter and RBS as the insert and the same protocol described above was repeated. The resulting plasmid was named pDN131. All plasmids generated were verified by Sanger Sequencing and an *E. coli* strain  
25 containing each plasmid was cryopreserved with 10% DMSO in a -80 °C freezer.

The final construct generated (pDN131) was introduced in a  $\Delta$ *cel* *M. extorquens* PA1 strain using a three-way mating protocol. The *E. coli* strain containing pDN131 was grown in 10 ml of LB + tetracycline (12.5  $\mu$ g/ml), a strain containing the a helper plasmid with the conjugal transfer genes of the broad host-range plasmid RK2 called pRK2073 was grown 10ml in LB + Streptomycin  
30 (50.0  $\mu$ g/ml), and *M. extorquens* PA1 was grown in 10 ml Hypho minimal media with 3.5 mM succinate as the carbon source. The *E. coli* strains were harvested in early stationary phase and *M. extorquens* PA1 was harvested in mid-exponential phase (OD<sub>600</sub> ~0.3). Centrifuged *E. coli* cells

were resuspended in 1 ml of fresh LB media without any antibiotic. Centrifuged *M. extorquens* PA1 cells were resuspended in 1 ml of Hypho minimal media. 50 µl of the pRK2073 resuspended in LB was added to the resuspended *E. coli* strain with pDN131 and incubated in a 37 °C shaking incubator for one hour. 10 µl of this culture was spotted on a Nutrient Agar plate and allowed to air dry. 50 µl of the *M. extorquens* PA1 culture was spotted on top of the *E. coli* culture on the Nutrient Agar plate and allowed to air dry. The plate was incubated at 30 °C to overnight and at 4 °C for a day. The colony growth was suspended in 100 µl of fresh hypho and plated on a hypho + 3.5 mM succinate + 1.2% agar + tetracycline (10 µg/ml) minimal media plate. Colonies from the first tetracycline plate were streaked on another hypho + 3.5 mM succinate + 1.2% agar + tetracycline (10 µg/ml) minimal media plate to ensure tetracycline resistance. Tetracycline resistant colonies are those that had integrated the allelic exchange vector in the chromosome. These colonies were then grown in hypho + 3.5 mM succinate overnight without any antibiotic. The overnight growth was plate on hypho + succinate + 1.2% agar + 5% sucrose minimal media plates. Since sucrose is a counter-selection agent, this step was used to select for colonies that had excised the allelic exchange vector out of the chromosome through homologous recombination. Unintended mutational inactivation of the *sacB* (responsible for sucrose sensitivity) gene on the allelic exchange vector was screened out by testing the sucrose<sup>+</sup> colonies for tetracycline resistance. Those colonies that had a sucrose<sup>+</sup> and tetracycline sensitive phenotype were tested for the ancestral or modified version of the chromosome at the *mptG* locus using PCR. Colonies that had replaced the *mptG* locus with the HPS and PHI genes of the RuMP pathway were verified by Sanger Sequencing and cyro-preserved in a -80 °C freezer with 10% DMSO.

### EXAMPLE III

#### RuMP Pathway for C<sub>1</sub> metabolism: Strain 2

Strain 2- Replacing the native serine cycle for C<sub>1</sub> assimilation with the RuMP pathway. The H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation will be kept intact however, due to the absence of a key gene in the serine cycle, carbon will not be assimilated from the formate produced during dissimilation as shown in Figure 3.

As described above the RuMP pathway used for the assimilation of C<sub>1</sub> compounds with fructose bis-phosphate aldolase (an enzyme also involved in the pentose phosphate pathway) and transaldolase as the key enzymes involved in cleavage and rearrangement respectively is extremely efficient. For one molecule of pyruvate produced 1 molecule of NADPH and 1 molecule of ATP

are also produced. On the contrary, the serine cycle leads to the net consumption of 1 molecule of NADPH and 1 molecule of ATP per molecule of pyruvate produced as well as production of increased CO<sub>2</sub> per unit substrate. Hence replacing the native serine cycle in *Methylobacterium* with the RuMP pathway will not only increase the efficiency of carbon assimilation but also the amount of carbon assimilated into biomass per unit substrate. Strain 2 was constructed by eliminating the native serine cycle for C<sub>1</sub> assimilation by deleting a gene involved in serine cycle and replacing it with a genes encoding hexulose 6-phosphate synthase (HPS) as well as a hexulose 6-phosphate isomerase (PHI) from *M. capsulatus* Bath. It should be noted that only two genes – those encoding HPS and PHI- are needed in the *Methylobacterium* genome to allow the RuMP pathway to assimilate C<sub>1</sub> compounds since all other genes in this pathway are already present in the host strain.

The construct used for the generation of this strain is shown in Figure 6. The construct was made as follows. The specific gene in the serine cycle that was knocked out is *hprA* (Mext\_1796 on the *M. extorquens* PA1 chromosome, ref), which is an NADPH dependent hydroxypyruvate reductase described in (Chistoserdova *et al.*, *J. Bacteriol.* 185(10): 2980-2987 (2003)). The ORF encoding *hprA* is 945 bases long and is found on the positive strand between bases 20161384 and 2017078 on the chromosome. The upstream flank of *hprA* for the construct (2015764-2016107) was amplified by a forward primer containing the recognition sequence for the restriction enzyme *Xba*I at the 5' end (TCTAGACTC ATC GAC AAC GGC GTG AAG G) and a reverse primer containing the recognition sequence for the restriction enzyme *Nde*I at the 5' end (CATATGGGGCAATCGTGTCGCTCAC). The downstream flank of *mptG* for the construct (2017019-2017465) was amplified by a forward primer containing the recognition sequence for the restriction enzyme *Apa*I at the 5' end (GGGCCCCCTCGTGGACAACGTCGAAGC) and a reverse primer containing the recognition sequence for the restriction enzyme *Sac*I at the 5' end (GAGCTCTCT TCA CCG CCT CGA ACA CC).

On investigating the genome sequence of *M. extorquens* PA1, it was evident that the only genes needed to introduce the dissimilatory RuMP cycle in the *M. extorquens* PA1 metabolic network were HPS and PHI. These genes were introduced from *M. capsulatus* Bath. In *M. capsulatus* Bath there are three genes encoding a HPS (MCA2738, MCA3043, and MCA3049 on the main chromosome of *Me. capsulatus* Bath), as well as three genes encoding a PHI (MCA2738, MCA3044, and MCA3050 on the main chromosome of *M. capsulatus* Bath). Previous work (Ward *et al.*, *PLoS Biology* 2:e303 (2004)) has shown that most carbon flux enters the RUMP cycle through MCA3049 and MCA3050. Hence, these two genes were introduced in place of the *mptG*

gene in *M. extorquens* PA1 for making Strain1. As stated above, MCA3049 encodes a hexulose 6-phosphate synthase (HPS) gene that is 648 bp long on the positive strand from position 3236945 to position 3237592 on the chromosome. MCA3050 encodes the hexulose 6-phosphate isomerase (PHI) gene that is 534 bp long on the positive strand from position 3237605 to position 3238138 on the chromosome. A forward primer with a recognition sequence for the restriction enzyme *NdeI* at the 5' end (TCATATGACCGGTGTCGCCAGGTACA) and a reverse primer with a recognition sequence for the restriction enzyme *ApaI* at the 5' end (TGGGCCACGGACAACATGCCACGC) were used to amplify a region between position 3236701 and 3238196 on the *M. capsulatus* Bath chromosome containing the promoter, the RBS (ribosome binding site) as well as the as the entire ORF for MCA3049 and MCA 3050.

The final construct was made on the backbone of an allelic exchange vector (pCM433), as described in (Marx, *BMC Research Notes* 1:1 (2008)), in the order described below. The vector pCM433 was digested with restriction enzymes *NdeI* and *XbaI*, as was the insert, the *hprA* upstream flank. The 5' end of the digested vector was dephosphorylated using an Antarctic Phosphatase manufactured by NEB (New England Biolabs). The digested vector and insert were gel extracted using the Gel Extraction kit manufactured by Zymo Research Inc. A 3:1 molar ratio of the insert to vector was ligated using the Quick Ligase Kit manufactured by NEB (New England Biolabs). 1 µl of the ligated mixture was transformed in 50 µl of 10<sup>8</sup> chemically competent cells manufactured by NEB (New England Biolabs). The transformed cells were plated on LB + tetracycline (12.5 µg/ml) + 1.2% agar plates and screened by PCR for the insert. Colonies that screened positive for the insert were grown in LB+ tetracycline (12.5 µg/ml). The resulting plasmid was named pDN123. Plasmid extraction was done using the standard Miniprep kit manufactured by Qiagen. For the next round, a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of pDN123 was used as the vector and a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of the pCRII- Blunt- Topo vector containing *hprA* downstream flank as the insert. The same protocol described above was repeated. The resulting plasmid was named pDN127. For the final round, a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of pDN127 was used as the vector and a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of the pCRII- Blunt- Topo vector with the region of the *Methylococcus capsulatus* Bath containing the MCA3049 and MCA3050 ORF, promoter and RBS as the insert and the same protocol described above was repeated. The resulting plasmid was named pDN132. All plasmids generated were verified by Sanger Sequencing and an *E. coli* strain containing each plasmid was cryopreserved with 10% DMSO in a -80 °C freezer.

The final construct generated (pDN132) was introduced in a  $\Delta cel$  *M. extorquens* PA1 strain using a three-way mating protocol. The *E. coli* strain containing pDN132 was grown in 10 ml of LB + tetracycline (12.5  $\mu\text{g/ml}$ ), a strain containing the a helper plasmid with the conjugal transfer genes of the broad host-range plasmid RK2 called pRK2073 was grown 10 ml in LB + streptomycin (50.0  $\mu\text{g/ml}$ ), and *M. extorquens* PA1 was grown in 10 ml Hypho minimal media with 3.5 mM succinate as the carbon source. The *E. coli* strains were harvested in early stationary phase and *M. extorquens* PA1 was harvested in mid-exponential phase ( $\text{OD}_{600} \sim 0.3$ ). Centrifuged *E. coli* cells were resuspended in 1 ml of fresh LB media without any antibiotic. Centrifuged *M. extorquens* PA1 cells were resuspended in 1 ml of Hypho minimal media. 50  $\mu\text{l}$  of the pRK2073 resuspended in LB was added to the resuspended *E. coli* strain with pDN131 and incubated in a 37 °C shaking incubator for one hour. 10  $\mu\text{l}$  of this culture was spotted on a Nutrient Agar plate and allowed to air dry. 50  $\mu\text{l}$  of the *M. extorquens* PA1 culture was spotted on top of the *E. coli* culture on the Nutrient Agar plate and allowed to air dry. The plate was incubated at 30 °C to overnight and at 4 °C for a day. The colony growth was suspended in 100  $\mu\text{l}$  of fresh hypho and plated on a hypho + 3.5 mM succinate + 1.2% agar + tetracycline (10  $\mu\text{g/ml}$ ) minimal media plate. Colonies from the first tetracycline plate were streaked on another hypho + 3.5 mM succinate + 1.2% agar + tetracycline (10  $\mu\text{g/ml}$ ) minimal media plate to ensure tetracycline resistance. Tetracycline resistant colonies are those that had integrated the allelic exchange vector in the chromosome. These colonies were then grown in hypho + 3.5 mM succinate overnight without any antibiotic. The overnight growth was plate on hypho + succinate + 1.2% agar + 5% sucrose minimal media plates. Since sucrose is a counter-selection agent, this step was used to select for colonies that had excised the allelic exchange vector out of the chromosome through homologous recombination. Unintended mutational inactivation of the *sacB* (responsible for sucrose sensitivity) gene on the allelic exchange vector was screened out by testing the sucrose<sup>+</sup> colonies for tetracycline resistance. Those colonies that had a sucrose<sup>+</sup> and tetracycline sensitive phenotype were tested for the ancestral or modified version of the chromosome at the *hprA* locus using PCR. Colonies that had replaced the *hprA* locus with the HPS and PHI/PHI genes of the RuMP pathway were verified by Sanger Sequencing and cyro-preserved in a -80 °C freezer with 10% DMSO.

## EXAMPLE IV

RuMP Pathway for C<sub>1</sub> metabolism: Strain 3

Strain 3 – Replacing the native H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation and the native serine cycle for C<sub>1</sub> assimilation with the RuMP pathway. This engineered strain should solely use the RuMP pathway to grow on methanol as shown in Figure 4.

As described above the RuMP pathway used for the assimilation of C<sub>1</sub> compounds with fructose bis-phosphate aldolase (an enzyme also involved in the pentose phosphate pathway) and transaldolase as the key enzymes involved in cleavage and rearrangement respectively is extremely efficient. For one molecule of pyruvate produced 1 molecule of NADPH and 1 molecule of ATP are also produced. On the contrary, the serine cycle leads to the net consumption of 1 molecule of NADPH and 1 molecule of ATP per molecule of pyruvate produced as well as production of increased CO<sub>2</sub> per unit substrate. Hence replacing the native serine cycle in *Methylobacterium* with the RuMP pathway will not only increase the efficiency of carbon assimilation but also the amount of carbon assimilated into biomass per unit substrate. Similarly, taking into account the fact that there are fewer enzymes involved in C<sub>1</sub> dissimilation using the RuMP pathway than the H<sub>4</sub>MPT pathway as well as incorporating the cost of production of the complex cofactor in the H<sub>4</sub>MPT pathway, we postulate that the RuMP pathway might carry out dissimilation of C<sub>1</sub> compounds more efficiently. Hence a strain of *Methylobacterium* that uses the RuMP pathway for assimilation and dissimilation of C<sub>1</sub> compounds might be able to grow faster, assimilate carbon more efficiently and also channel more carbon to biomass and reduce CO<sub>2</sub> production per unit substrate.

Strain 3 was constructed by first deleting the H<sub>4</sub>MPT (tetrahydromethanopterin) pathway for C<sub>1</sub> dissimilation in *Methylobacterium* and then eliminating the native serine cycle for C<sub>1</sub> assimilation by deleting a gene involved in serine cycle and replacing it with a gene encoding hexulose 6-phosphate synthase (HPS) as well as a hexulose 6-phosphate isomerase (PHI) from *M. capsulatus* Bath. It should be noted that only two genes – those encoding HPS and PHI- are needed in the *Methylobacterium* genome to allow the RuMP pathway to assimilate C<sub>1</sub> compounds since all other genes in this pathway are already present in the host strain.

The construct for deleting *mptG* is shown in Figure 7. The specific gene in the biosynthetic pathway of tetrahydromethanopterin that was knocked out is *mptG* (Mext\_1828 on the *M. extorquens* PA1 chromosome), which is a ribofuranosylaminobenzene 5'-phosphate (RFAP) (Chistoserdova et al., *J. Bacteriol.* 187:2508-2512 (2005)). The ORF encoding *mptG* is 1023 bases

long and is found on the positive strand between bases 2049113 and 2050135 on the chromosome. A forward primer containing a 30 bp overhang (ATGGATGCATATGCTGCAGCTCGAGCGGCCGCGAT CTC GGC GAT CAG CTC ACC) corresponding to bases 1029 through 1060 on the positive strand of pCM433 and a reverse primer (GCC GTT GAG ATC GAG GAA GCC) were used to amplify the upstream flank from position 2048391 to 2049259 on the *M. extorquens* PA1 chromosome. A forward primer containing a 30 bp overhang (CTGCATTTTCGGCTTCCTCGATCTCAACGGCCTC GCA GAA GGA GGC GGA) spanning the region from 2049230 through 2049259 on the positive strand of the *M. extorquens* PA1 chromosome and a reverse primer (GGTTAACACCGCGTACGTAGGGCCCCGCGGCCGCGGT GAA GGC GAT CTT CGA GAC G) with a 30 bp overhang spanning the region from 1053 to 1084 on the negative strand of pCM433 was used to amplify the downstream flank from position 2050027 to 2050749 on the *M. extorquens* PA1 chromosome. The final construct was made on the backbone of the pCM433 allelic exchange vector (Marx, *BMC Research Notes* 1:1 (2008)) using the protocol described in (Gibson et al., *Nature Methods* 6:343-345(2009)). The construct containing the upstream and downstream flanks of *mptG* was named pDN68 and verified by Sanger sequencing and cryo-preserved in a NEB10 $\beta$  competent *E. coli* strain with 10% DMSO in a -80 °C freezer. pDN68 was introduced in a  $\Delta cel$  *M. extorquens* PA1 strain using a three-way mating protocol. The *E. coli* strain containing pDN68 was grown in 10 ml of LB + tetracycline (12.5  $\mu$ g/ml), a strain containing the a helper plasmid with the conjugal transfer genes of the broad host-range plasmid RK2 called pRK2073 was grown 10 ml in LB + Streptomycin (50.0  $\mu$ g/ml), and *M. extorquens* PA1 was grown in 10 ml Hypho minimal media with 3.5 mM succinate as the carbon source. The *E. coli* strains were harvested in early stationary phase and *M. extorquens* PA1 was harvested in mid-exponential phase (OD<sub>600</sub> ~0.3). Centrifuged *E. coli* cells were resuspended in 1 ml of fresh LB media without any antibiotic. Centrifuged *M. extorquens* PA1 cells were resuspended in 1 ml of Hypho minimal media. 50  $\mu$ l of the pRK2073 resuspended in LB was added to the resuspended *E. coli* strain with pDN68 and incubated in a 37 °C shaking incubator for one hour. 10  $\mu$ l of this culture was spotted on a Nutrient Agar plate and allowed to air dry. 50  $\mu$ l of the *M. extorquens* PA1 culture was spotted on top of the *E. coli* culture on the Nutrient Agar plate and allowed to air dry. The plate was incubated at 30 °C overnight and at 4 °C for a day. The colony growth was suspended in 100  $\mu$ l of fresh hypho and plated on a hypho + 3.5 mM succinate + 1.2% agar + tetracycline (10  $\mu$ g/ml) minimal media plate. Colonies from the first tetracycline plate were streaked on another hypho + 3.5 mM succinate + 1.2% agar + tetracycline (10  $\mu$ g/ml) minimal media plate to ensure tetracycline resistance. Tetracycline resistant colonies are those that had integrated the allelic exchange vector

in the chromosome. These colonies were then grown in hypho + 3.5 mM succinate overnight without any antibiotic. The overnight growth was plate on hypho + succinate + 1.2% agar + 5% sucrose minimal media plates. Since sucrose is a counter-selection agent, this step was used to select for colonies that had excised the allelic exchange vector out of the chromosome through  
 5 homologous recombination. Unintended mutational inactivation of the *sacB* (responsible for sucrose sensitivity) gene on the allelic exchange vector was screened out by testing the sucrose<sup>+</sup> colonies for tetracycline resistance. Those colonies that had a sucrose<sup>+</sup> and tetracycline sensitive phenotype were tested for the ancestral or knockout version of the chromosome at the *mptG* locus using PCR. Colonies that deleted the *mptG* locus were verified by Sanger Sequencing and cyro-  
 10 preserved in a -80° C freezer with 10% DMSO.

The specific gene in the serine cycle that was knocked out is *hprA* (Mext\_1796 on the *M. extorquens* PA1 chromosome, ref), which is a NADPH dependent hydroxypyruvate reductase, which is described in (Chistoserdova *et al.*, *J. Bacteriol.* 185(10): 2980-2987 (2003)). The ORF encoding *hprA* is 945 bases long and is found on the positive strand between bases 20161384 and  
 15 2017078 on the chromosome. The upstream flank of *hprA* for the construct (2015764-2016107) was amplified by a forward primer containing the recognition sequence for the restriction enzyme *XbaI* at the 5' end (TCTAGACTC ATC GAC AAC GGC GTG AAG G) and a reverse primer containing the recognition sequence for the restriction enzyme *NdeI* at the 5' end (CATATGGGGCAATCGTGTCGCTCAC). The downstream flank of *mptG* for the construct  
 20 (2017019-2017465) was amplified by a forward primer containing the recognition sequence for the restriction enzyme *ApaI* at the 5' end (GGGCCCCTCGTGGACAACGTCGAAGC) and a reverse primer containing the recognition sequence for the restriction enzyme *SacI* at the 5' end (GAGCTCTCT TCA CCG CCT CGA ACA CC).

On investigating the genome sequence of *M. extorquens* PA1, it was evident that the only genes  
 25 needed to introduce the dissimilatory RuMP cycle in the *M. extorquens* PA1 metabolic network were HPS and PHI. These genes were introduced from *M. capsulatus* Bath. In *M. capsulatus* Bath there are three genes encoding a HPS (MCA2738, MCA3043, and MCA3049 on the main chromosome of *Me. capsulatus* Bath), as well as three genes encoding a PHI (MCA2738, MCA3044, and MCA3050 on the main chromosome of *M. capsulatus* Bath). Previous work (Ward  
 30 *et al.*, *PLoS Biology* 2:e303 (2004)) has shown that most carbon flux enters the RUMP cycle through MCA3049 and MCA3050. Hence, these two genes were introduced in place of the *mptG* gene in *M. extorquens* PA1 for making Strain1. As stated above, MCA3049 encodes a hexulose 6-phosphate synthase (HPS) gene that is 648 bp long on the positive strand from position 3236945 to

position 3237592 on the chromosome. MCA3050 encodes the hexulose 6-phosphate isomerase (PHI) gene that is 534 bp long on the positive strand from position 3237605 to position 3238138 on the chromosome. A forward primer with a recognition sequence for the restriction enzyme *NdeI* at the 5' end (TCATATGACCGGTGTCGCCAGGTACA) and a reverse primer with a recognition sequence for the restriction enzyme *ApaI* at the 5' end (TGGGCCACGGACAACATGCCACGC) were used to amplify a region between position 3236701 and 3238196 on the *M. capsulatus* Bath chromosome containing the promoter, the RBS (ribosome binding site) as well as the entire ORF for MCA3049 and MCA 3050.

The final construct was made on the backbone of an allelic exchange vector (pCM433), as described in (Marx, *BMC Research Notes* 1:1 (2008)), in the order described below. The vector pCM433 was digested with restriction enzymes *NdeI* and *XbaI*, as was the insert, the *hprA* upstream flank. The 5' end of the digested vector was dephosphorylated using an Antarctic Phosphatase manufactured by NEB (New England Biolabs). The digested vector and insert were gel extracted using the Gel Extraction kit manufactured by Zymo Research Inc. A 3:1 molar ratio of the insert to vector was ligated using the Quick Ligase Kit manufactured by NEB (New England Biolabs). 1  $\mu$ l of the ligated mixture was transformed in 50  $\mu$ l of 10 $\beta$  chemically competent cells manufactured by NEB (New England Biolabs). The transformed cells were plated on LB + tetracycline (12.5  $\mu$ g/ml) + 1.2% agar plates and screened by PCR for the insert. Colonies that screened positive for the insert were grown in LB+ tetracycline (12.5  $\mu$ g/ml). The resulting plasmid was named pDN123. Plasmid extraction was done using the standard Miniprep kit manufactured by Qiagen. For the next round, a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of pDN123 was used as the vector and a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of the pCRII- Blunt- Topo vector containing *hprA* downstream flank as the insert. The same protocol described above was repeated. The resulting plasmid was named pDN127. For the final round, a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of pDN127 was used as the vector and a *dam*<sup>-</sup>/*dcm*<sup>-</sup> version of the pCRII- Blunt- Topo vector with the region of the *Methylococcus capsulatus* Bath containing the MCA3049 and MCA3050 ORF, promoter and RBS as the insert and the same protocol described above was repeated. The resulting plasmid was named pDN132. All plasmids generated were verified by Sanger Sequencing and an *E. coli* strain containing each plasmid was cryopreserved with 10% DMSO in a -80 °C freezer.

The final construct generated (pDN132) was introduced in a  $\Delta$ *cel*  $\Delta$ *mptG* *M. extorquens* PA1 strain using a three-way mating protocol. The *E. coli* strain containing pDN132 was grown in 10 ml of LB + tetracycline (12.5  $\mu$ g/ml), a strain containing the a helper plasmid with the conjugal transfer

genes of the broad host-range plasmid RK2 called pRK2073 was grown 10 ml in LB + Streptomycin (50.0 µg/ml), and *M. extorquens* PA1 was grown in 10 ml Hypho minimal media with 3.5 mM succinate as the carbon source. The *E. coli* strains were harvested in early stationary phase and *M. extorquens* PA1 was harvested in mid-exponential phase (OD<sub>600</sub> ~0.3). Centrifuged  
5 *E. coli* cells were resuspended in 1 ml of fresh LB media without any antibiotic. Centrifuged *M. extorquens* PA1 cells were resuspended in 1 ml of Hypho minimal media. 50 µl of the pRK2073 resuspended in LB was added to the resuspended *E. coli* strain with pDN131 and incubated in a 37 °C shaking incubator for one hour. 10 µl of this culture was spotted on a Nutrient Agar plate and allowed to air dry. 50 µl of the *M. extorquens* PA1 culture was spotted on top of the *E. coli* culture  
10 on the Nutrient Agar plate and allowed to air dry. The plate was incubated at 30 °C to overnight and at 4 °C for a day. The colony growth was suspended in 100 µl of fresh hypho and plated on a hypho + 3.5 mM succinate + 1.2% agar + tetracycline (10 µg/ml) minimal media plate. Colonies from the first tetracycline plate were streaked on another hypho + 3.5 mM succinate + 1.2% agar + tetracycline (10 µg/ml) minimal media plate to ensure tetracycline resistance. Tetracycline resistant  
15 colonies are those that had integrated the allelic exchange vector in the chromosome. These colonies were then grown in hypho + 3.5 mM succinate overnight without any antibiotic. The overnight growth was plate on hypho + succinate + 1.2% agar + 5% sucrose minimal media plates. Since sucrose is a counter-selection agent, this step was used to select for colonies that had excised the allelic exchange vector out of the chromosome through homologous recombination.  
20 Unintended mutational inactivation of the *sacB* (responsible for sucrose sensitivity) gene on the allelic exchange vector was screened out by testing the sucrose<sup>+</sup> colonies for tetracycline resistance. Those colonies that had a sucrose<sup>+</sup> and tetracycline sensitive phenotype were tested for the ancestral or modified version of the chromosome at the *hprA* locus using PCR. Colonies that had replaced the *hprA* locus with the HPS and PHI genes of the RuMP pathway were verified by  
25 Sanger Sequencing and cyro-preserved in a -80 °C freezer with 10% DMSO.

#### EXAMPLE V

Subsequent evolution to improve growth properties

of strains resulting from Examples II-IV

The above strains generate methylotrophs with improved yield on C<sub>1</sub> compounds due to less CO<sub>2</sub>  
30 production. This results from the stoichiometric advantage of incorporating all carbon at the level of formaldehyde, rather than from a combination of formate and CO<sub>2</sub> for the serine cycle, or purely

CO<sub>2</sub> for autotrophs. However, cells can be further optimized. To further develop strains in which central metabolism is to be engineered to produce valuable chemicals, strains are first evolved in batch culture to select for increased growth rate. This follows precedent with *M. extorquens* AM1 where the H<sub>4</sub>MPT-dependent C<sub>1</sub> dissimilation pathway was replaced with the analogous, but non-homologous glutathione-dependent pathway from *Paracoccus denitrificans* (Marx et al., *J. Bacteriol.* 185:7160-7168 (2003)). Subsequent evolution in methanol-containing media resulted in dramatic increases in growth (Chou et al., *Science* 332:1190-1192 (2011)). Much of the improvement occurred in the first couple hundred generations of these experiments, and the extent of improvement varied considerably across replicates (Lee and Marx, *Genetics* 193:943-952 (2013)). Briefly, the desired starting strain to be used as an ancestor is streaked for individual colonies, each of which is used to found an independent replicate population. Cultures are then periodically diluted ( $1/2^n$ ) into fresh medium containing the C<sub>1</sub> substrate of interest, such as methanol, resulting in  $n$  net generations per transfer. Samples of the remaining culture are cryopreserved to provide a living fossil record of the experiment. Populations or isolates from them can be screened for improved growth under the desired conditions.

The contents of all references, patents and published patent applications cited throughout this application are hereby incorporated by reference in their entirety for all purposes.

#### EQUIVALENTS

Other embodiments will be evident to those of skill in the art. It should be understood that the foregoing description is provided for clarity only and is merely exemplary. The spirit and scope of the present invention are not limited to the above example, but are encompassed by the claims. All publications, patents and patent applications cited above are incorporated by reference herein in their entirety for all purposes to the same extent as if each individual publication or patent application were specifically indicated to be so incorporated by reference.

What is claimed is:

1. A recombinant bacterium cell  
genetically modified to disrupt the cell's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation and to include a recombinant RuMP pathway for C<sub>1</sub> dissimilation.  
5
2. The recombinant bacterium cell of claim 1 being a *Methylobacterium*.
3. The recombinant bacterium cell of claim 1 being a *Methylobacterium extorquens*.
- 10 4. The recombinant bacterium cell of claim 1 being a *Methylobacterium extorquens* PA1.
5. A recombinant bacterium cell  
genetically modified to disrupt the cell's serine cycle for C<sub>1</sub> assimilation and to include a recombinant RuMP pathway for C<sub>1</sub> assimilation.  
15
6. The recombinant bacterium cell of claim 5 being a *Methylobacterium*.
7. The recombinant bacterium cell of claim 5 being a *Methylobacterium extorquens*.
- 20 8. The recombinant bacterium cell of claim 5 being a *Methylobacterium extorquens* PA1.
9. A recombinant bacterium cell  
genetically modified to disrupt the cell's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation, to disrupt the cell's serine cycle for C<sub>1</sub> assimilation and to include a recombinant RuMP pathway for C<sub>1</sub>  
25 assimilation and C<sub>1</sub> dissimilation.
10. The recombinant bacterium cell of claim 9 being a *Methylobacterium*.
11. The recombinant bacterium cell of claim 9 being a *Methylobacterium extorquens*.  
30
12. The recombinant bacterium cell of claim 9 being a *Methylobacterium extorquens* PA1.
13. A method of making a genetically modified bacterium comprising

genetically modifying a target bacterium to disrupt the target bacterium's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation and to include a recombinant RuMP pathway for C<sub>1</sub> dissimilation.

14. The method of claim 13 wherein the target bacterium is a *Methylobacterium*.

5

15. The method of claim 13 wherein the target bacterium is a *Methylobacterium extorquens*.

16. The method of claim 13 wherein the target bacterium is a *Methylobacterium extorquens* PA1.

10

17. A method of making a genetically modified bacterium comprising genetically modifying a target bacterium to disrupt the target bacterium's serine cycle for C<sub>1</sub> assimilation and to include a recombinant RuMP pathway for C<sub>1</sub> assimilation.

15

18. The method of claim 17 wherein the target bacterium is a *Methylobacterium*.

19. The method of claim 17 wherein the target bacterium is a *Methylobacterium extorquens*.

20. The method of claim 17 wherein the target bacterium is a *Methylobacterium extorquens* PA1.

20

21. A method of making a genetically modified bacterium comprising genetically modifying a target bacterium to disrupt the target bacterium's H<sub>4</sub>MPT pathway for C<sub>1</sub> dissimilation, to disrupt the target bacterium's serine cycle for C<sub>1</sub> assimilation and to include a recombinant RuMP pathway for C<sub>1</sub> assimilation and C<sub>2</sub> dissimilation.

25

22. The method of claim 21 wherein the target bacterium is a *Methylobacterium*.

23. The method of claim 21 wherein the target bacterium is a *Methylobacterium extorquens*.

30

24. The method of claim 21 wherein the target bacterium is a *Methylobacterium extorquens* PA1.

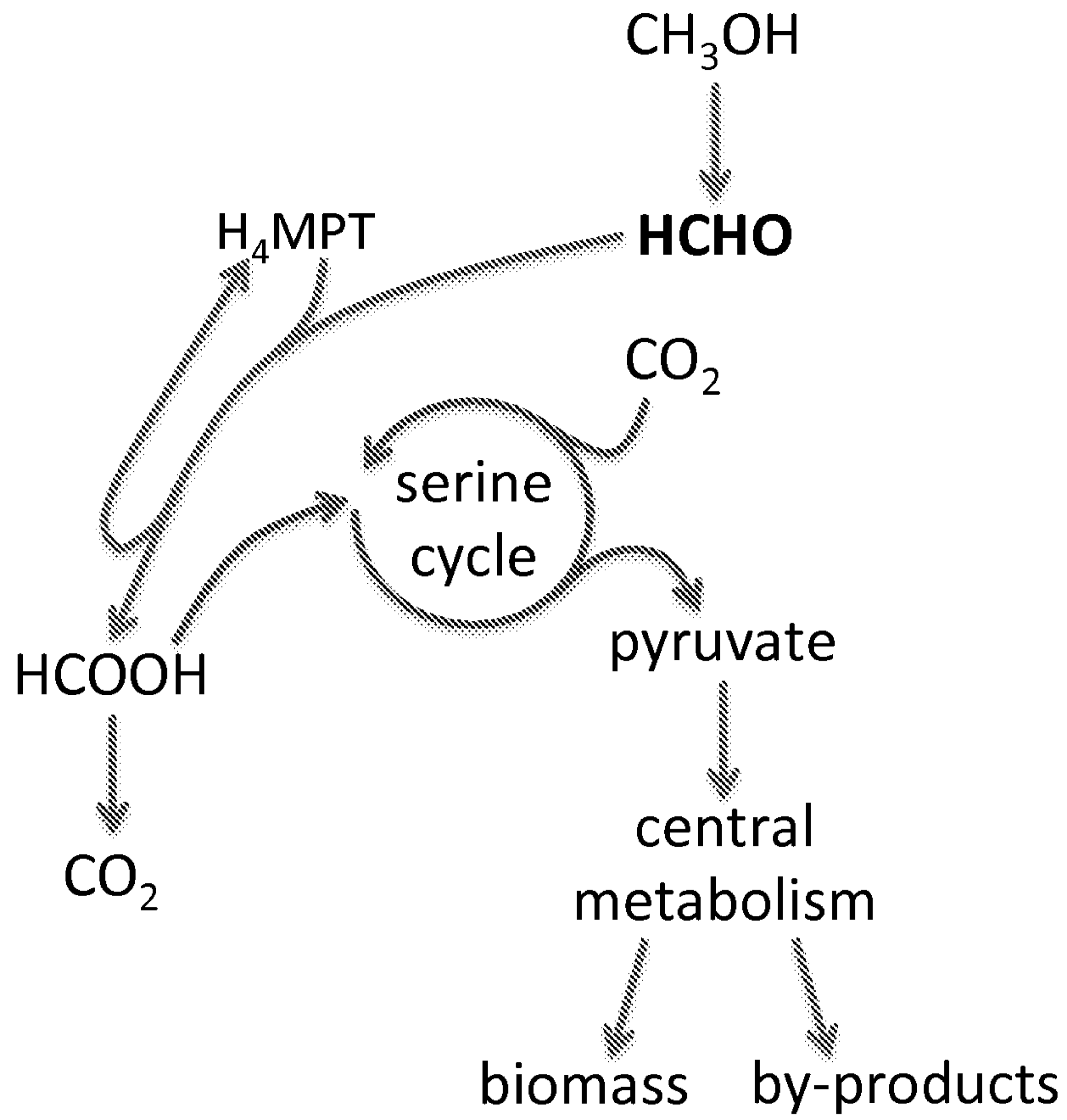


Figure 1

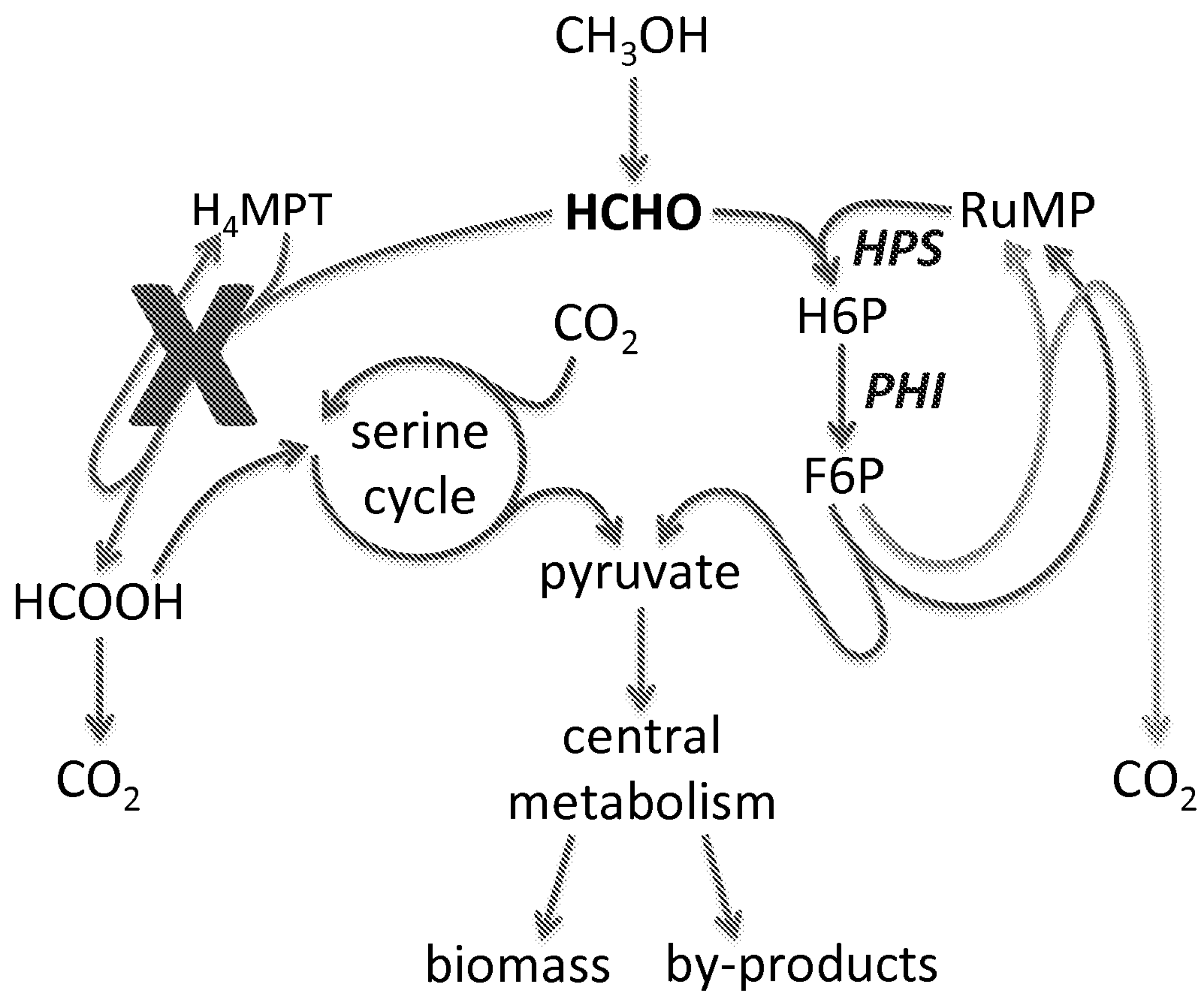


Figure 2

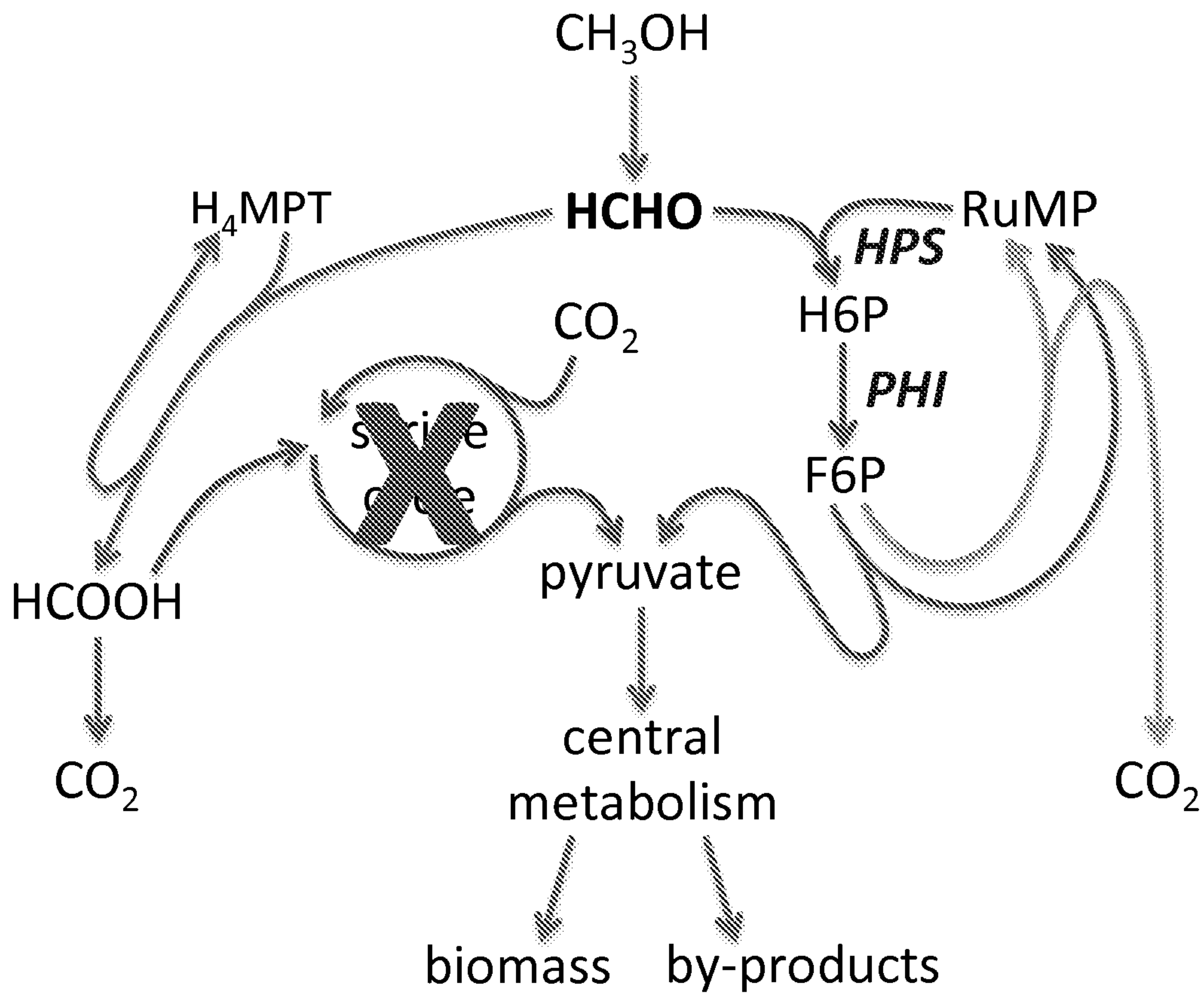


Figure 3

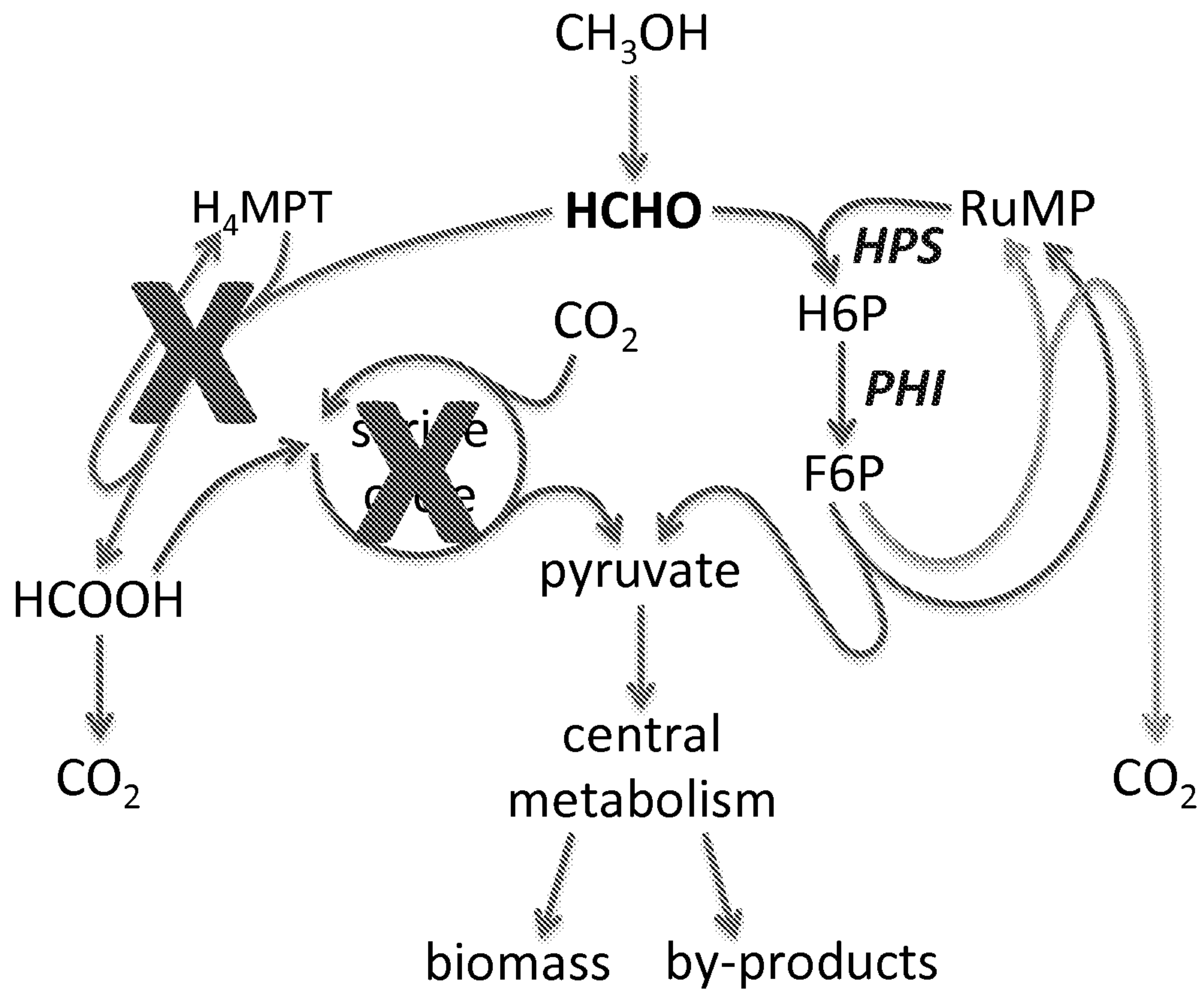


Figure 4

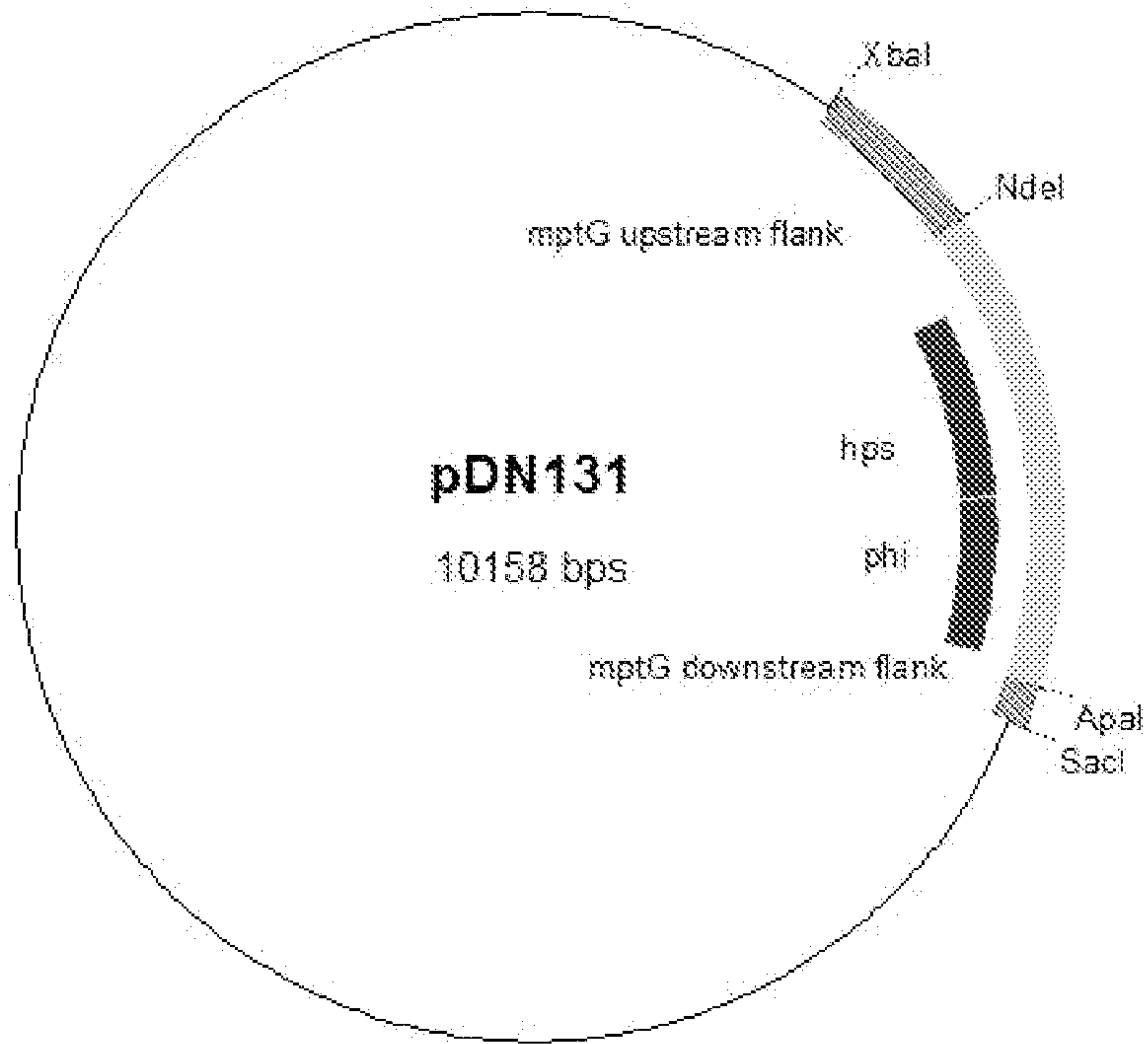


Figure 5

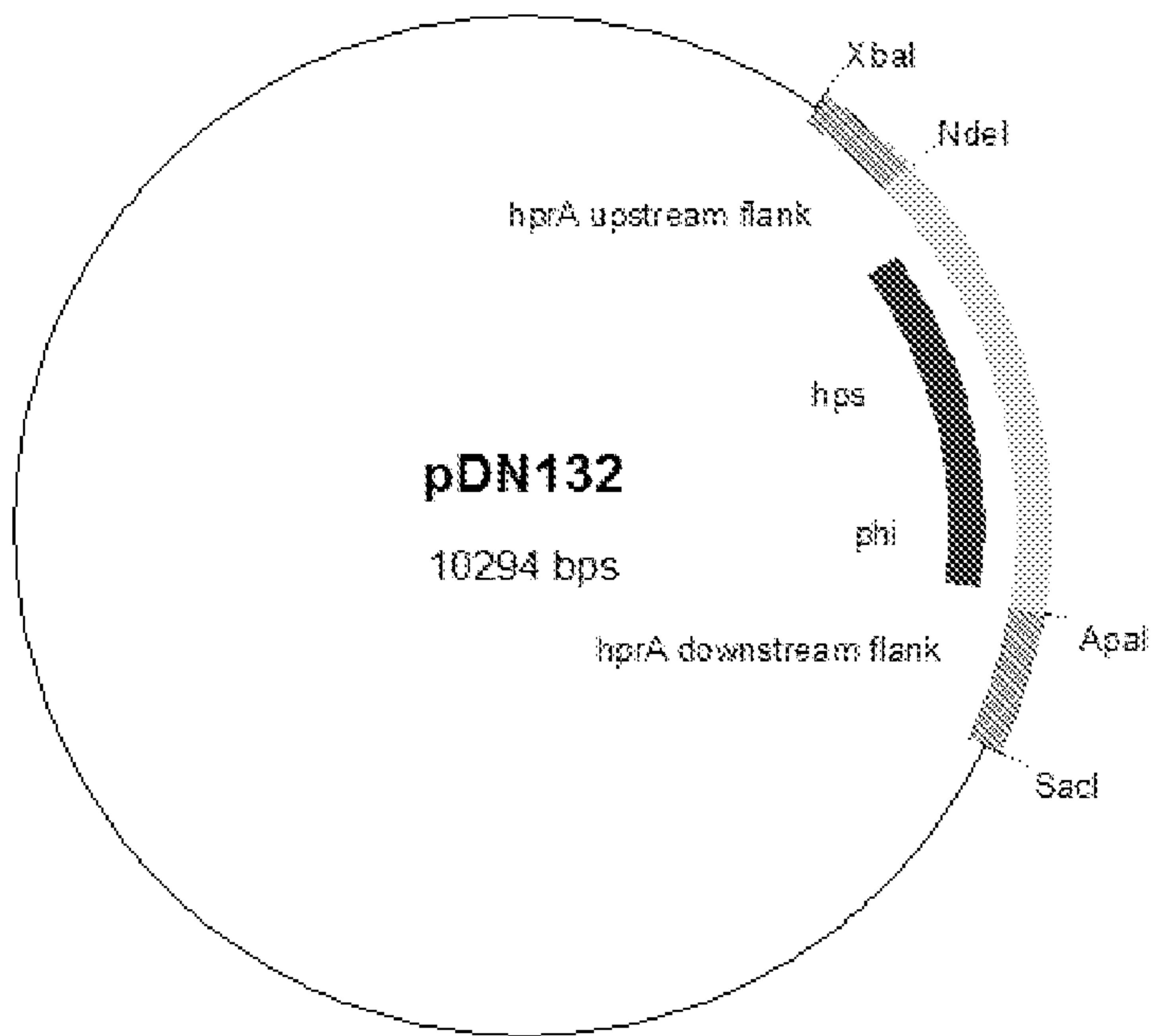


Figure 6

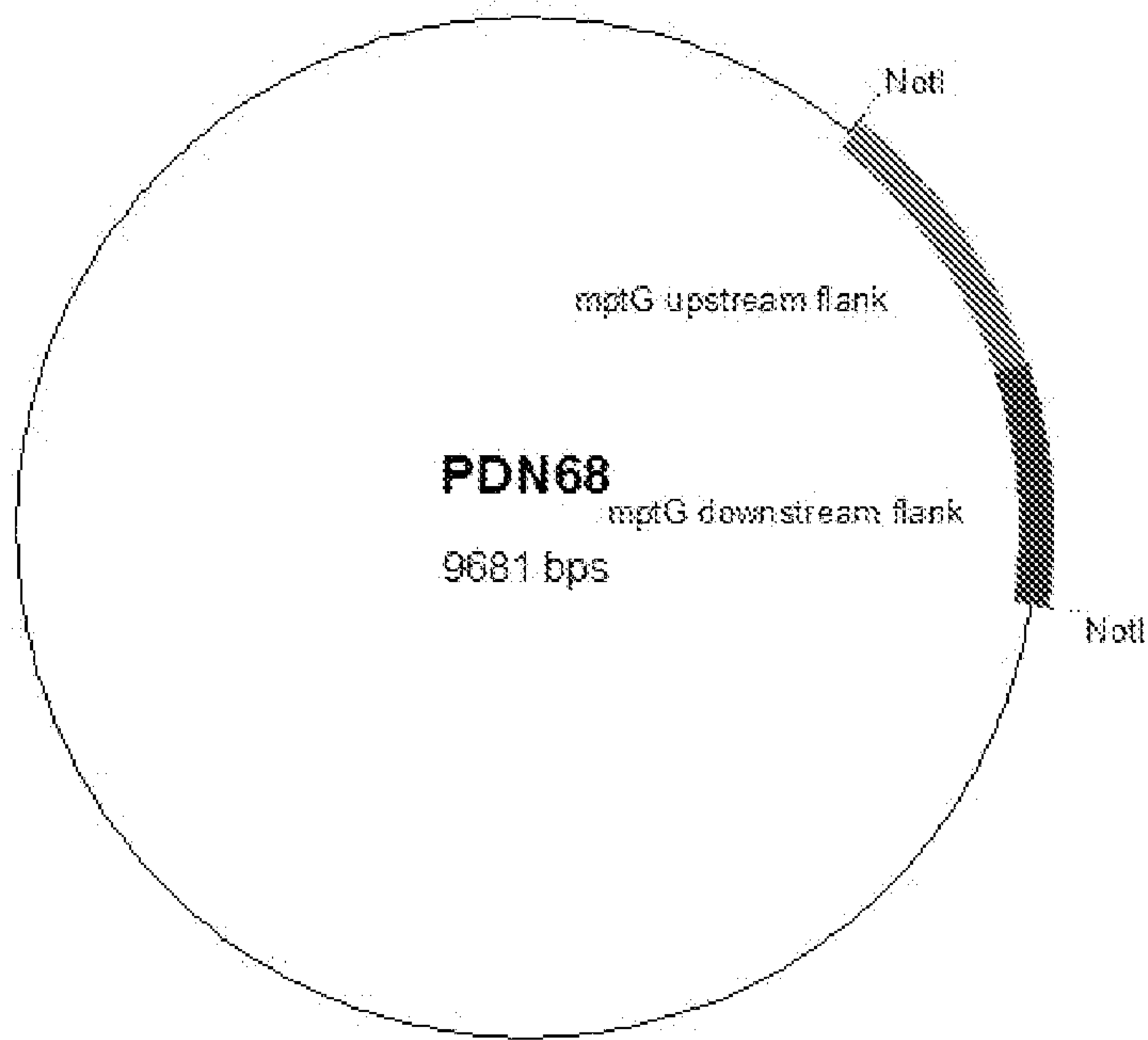


Figure 7

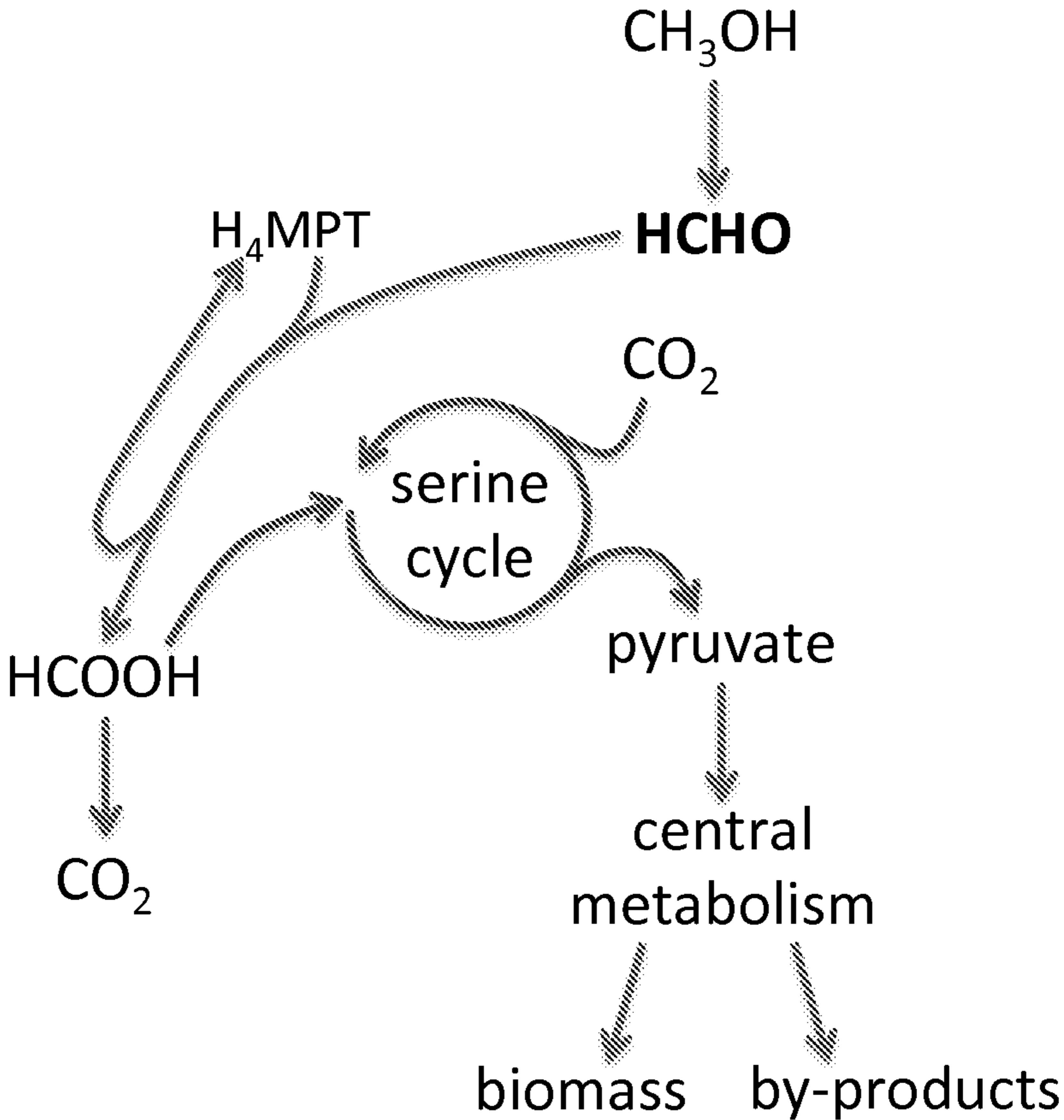


Figure 1