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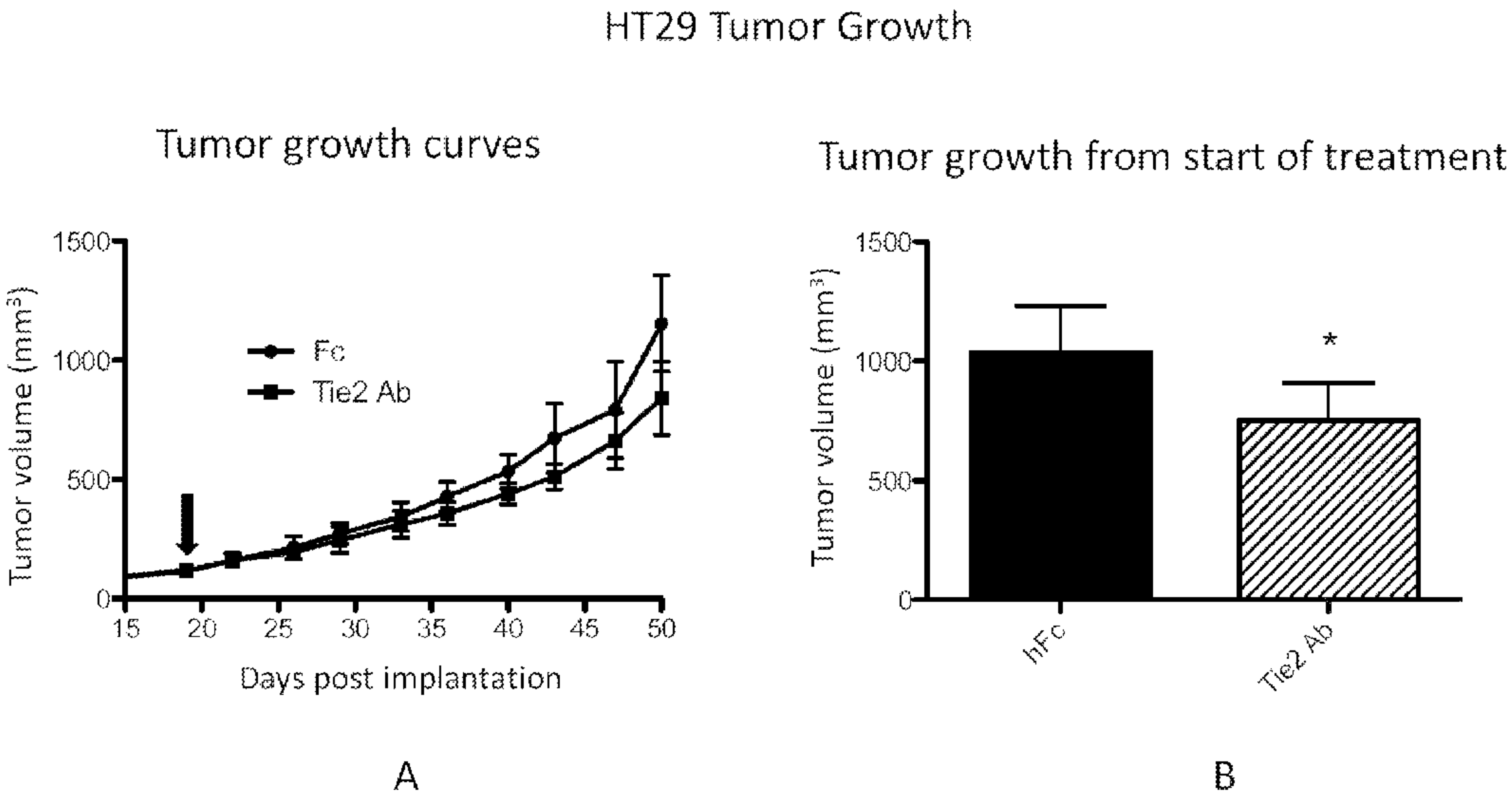
(54) **Titre : ANTICORPS ANTI-TIE2 ET UTILISATIONS ASSOCIEES**
(54) **Title: ANTI-TIE2 ANTIBODIES AND USES THEREOF**


HT29 Tumor Growth

Tumor growth curves

Tumor growth from start of treatment

Figure 4

(57) **Abrégé/Abstract:**
The present invention provides antibodies that bind to Tie2 and methods of using same. According to certain embodiments of the invention, the antibodies are fully human antibodies that bind to human Tie2 and block the interaction between Tie2 and one or more Tie2 ligands such as angtopotetin 1 (Ang1), angtopoietin 2 (Ang2), angiopoietin 3 (Ang3) and/or angtopoietin 4 (Ang4). The antibodies of the invention are useful, inter alia, for the treatment of d and disorders associated with one or more Tie2 biological activities including angiogenesis.

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(54) **Title**: ANTI-TIE2 ANTIBODIES USES THEREOF

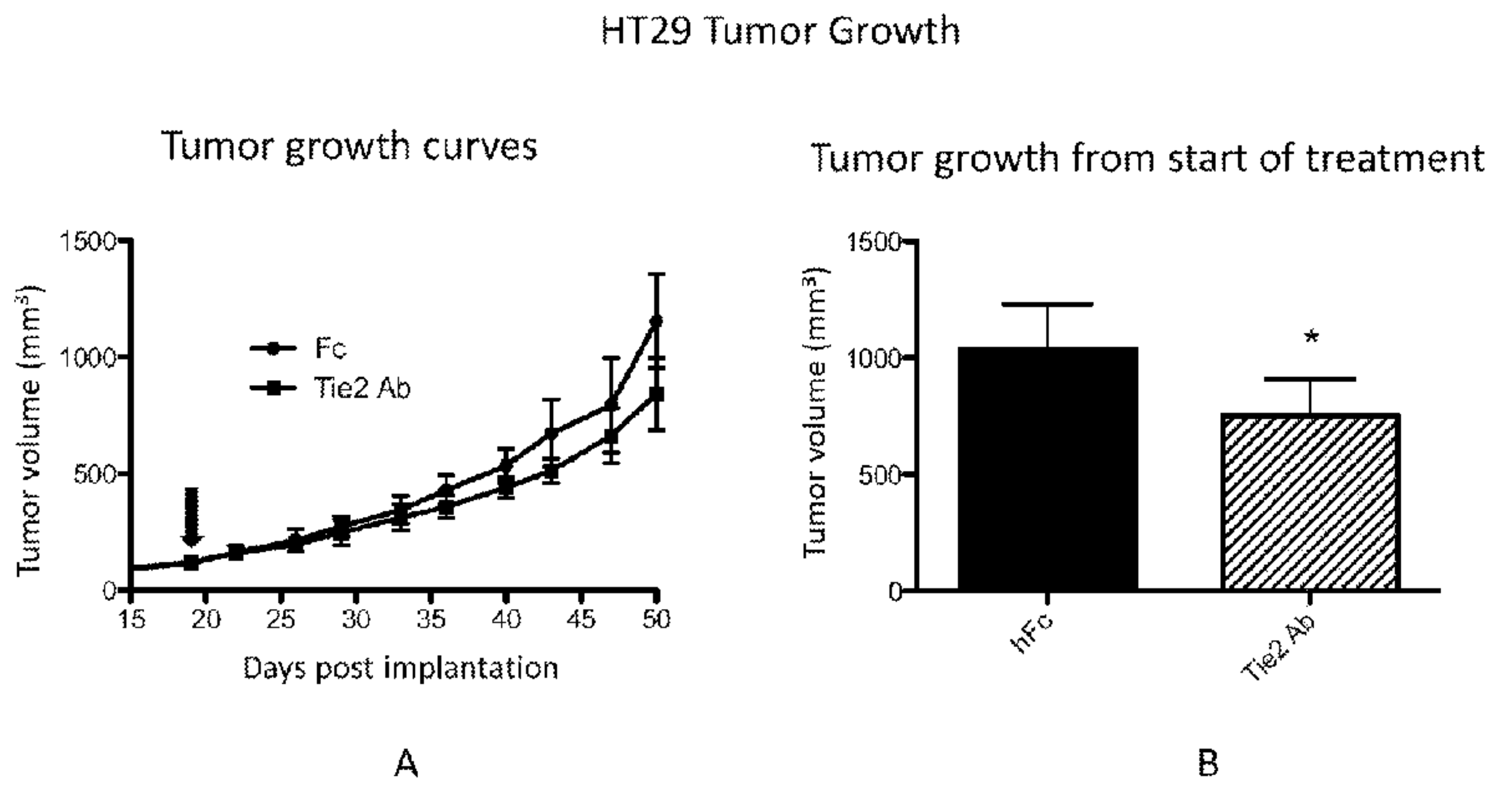


Figure 4

(57) **Abstract**: The present invention provides antibodies that bind to Tie2 and methods of using same. According to certain embodiments of the invention, the antibodies are fully human antibodies that bind to human Tie2 and block the interaction between Tie2 and one or more Tie2 ligands such as angtopotetin 1 (Ang1), angtopoietin 2 (Ang2), angiopoietin 3 (Ang3) and/or angtopoietin 4 (Ang4). The antibodies of the invention are useful, *inter alia*, for the treatment of d and disorders associated with one or more Tie2 biological activities including angiogenesis.

WO 2013/028442 A1

ANTI-TIE2 ANTIBODIES AND USES THEREOF

FIELD OF THE INVENTION

[0001] The present invention relates to antibodies, and antigen-binding fragments thereof, which are specific for human Tie2.

BACKGROUND

[0002] Angiogenesis is the biological process whereby new blood vessels are formed. Aberrant angiogenesis is associated with several disease conditions including, *e.g.*, proliferative retinopathies, rheumatoid arthritis and psoriasis. In addition, it is well established that angiogenesis is critical for tumor growth and maintenance. Tie2 is a single transmembrane tyrosine kinase receptor that has been localized to the endothelial cells of forming blood vessels and has been shown to play a role in angiogenesis. Tie2 ligands include the angiopoietins (*e.g.*, Ang1, Ang2, Ang3 and Ang4). Blocking the interaction between Tie2 and one or more of its ligands is expected to have beneficial therapeutic effects in settings where it is advantageous to limit or block angiogenesis.

[0003] Antibodies to Tie2 are mentioned, *e.g.*, in US Patent Nos. 6,365,154 and 6,376,653. Nonetheless, there remains a need in the art for novel molecules capable of binding to Tie2, especially anti-Tie2 antibodies that can block the interaction of Tie2 with one or more Tie2 ligands such as Ang2. Such molecules would be useful for various therapeutic and diagnostic purposes.

BRIEF SUMMARY OF THE INVENTION

[0004] The present invention provides antibodies that bind human Tie2. The antibodies of the invention are useful, *inter alia*, for inhibiting Tie2-mediated signaling and for treating diseases and disorders caused by or related to Tie2 activity and/or signaling. According to certain embodiments, the antibodies of the invention block the interaction between Tie2 and one or more Tie2 ligands such as Ang1, Ang2, Ang3, and/or Ang4.

[0005] The antibodies of the invention can be full-length (for example, an IgG1 or IgG4 antibody) or may comprise only an antigen-binding portion (for example, a Fab, $F(ab')_2$ or scFv fragment), and may be modified to affect functionality, e.g., to eliminate residual effector functions (Reddy et al., 2000, J. Immunol. 164:1925-1933).

[0006] The present invention provides anti-Tie2 antibodies that have substantially the same binding characteristics as any of the exemplary anti-Tie2 antibodies described herein. The present invention includes cell lines that produce the anti-Tie2 antibodies described herein. As non-limiting examples, cell lines which produce the exemplary antibodies H1M2055N and H2aM2760N were deposited under terms in accordance with the Budapest Treaty with the American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, Va. 20110-2209 on December 2, 2011. The deposited cell lines have been assigned the following accession numbers: PTA-12295 (H1M2055N) and PTA-12296 (H2aM2760N).

[0007] The present invention provides nucleic acid molecules encoding the exemplary anti-Tie2 antibodies described herein. Recombinant expression vectors carrying the nucleic acids of the invention, and host cells into which such vectors have been introduced, are also encompassed by the invention, as are methods of producing the antibodies by culturing the host cells under conditions permitting production of the antibodies, and recovering the antibodies produced.

[0008] The present invention includes antibodies and antigen-binding fragments thereof comprising the heavy and light chain CDR amino acid sequences (HCDR1, HCDR2, HCDR3, LCDR1, LCDR2 and LCDR3) found within any of the exemplary anti-Tie2 antibodies described herein, including the antibodies produced by the deposited cell lines PTA-12295 and PTA-12296. The present invention also includes antibodies and antigen-binding fragments thereof comprising the heavy and light chain variable domain amino acid sequences (HCVR and LCVR) found within any of the exemplary anti-Tie2 antibodies described herein, including the antibodies produced by the deposited cell lines PTA-12295 and PTA-12296.

[0009] The present invention includes any of the exemplary anti-Tie2 antibodies described herein having a modified glycosylation pattern. In some applications, modification to remove undesirable glycosylation sites may be useful, or an antibody lacking a fucose moiety present on the oligosaccharide chain, for example, to increase antibody dependent cellular cytotoxicity (ADCC) function (see Shield et al. (2002) JBC 277:26733). In other applications, modification of galactosylation can be made in order to modify complement dependent cytotoxicity (CDC).

[0010] In another aspect, the invention provides pharmaceutical compositions comprising an anti-Tie2 antibody as described herein and a pharmaceutically acceptable carrier. In a related aspect, the invention features compositions which comprise a combination of an anti-Tie2 antibody and a second therapeutic agent. Exemplary agents that may be advantageously combined with an anti-Tie2 antibody include, without limitation, other agents that inhibit anti-Tie2 activity (including other antibodies or antigen-binding fragments thereof, peptide inhibitors, small molecule antagonists, etc.) and/or agents which interfere with Tie2 upstream or downstream signaling.

[0011] In yet another aspect, the invention provides methods for inhibiting Tie2 activity using an anti-Tie2 antibody or antigen-binding portion of an antibody of the invention, wherein the therapeutic methods comprise administering a therapeutically effective amount of a pharmaceutical composition comprising an antibody or antigen-binding fragment of an antibody of the invention. The disorder treated is any disease or condition which is improved, ameliorated, inhibited or prevented by removal, inhibition or reduction of Tie2 activity. The anti-Tie2 antibody or antibody fragment of the invention may function to block the interaction between Tie2 and a Tie2 binding partner (*e.g.*, Ang1, Ang2, Ang3, and/or Ang4), or otherwise inhibit the signaling activity of Tie2.

[0012] The present invention also includes the use of an anti-Tie2 antibody or antigen binding portion of an antibody of the invention in the manufacture of a medicament for the treatment of a disease or disorder related to or caused by Tie2 activity in a patient.

[0013] Other embodiments will become apparent from a review of the ensuing detailed description.

BRIEF DESCRIPTION OF THE FIGURES

[0014] **Fig. 1.** Linear depiction of full-length human Tie2 and various deletion constructs used to map the epitope of the anti-Tie2 antibodies of the present invention. Numbers above the constructs indicate the amino acid boundaries of the constructs relative to the full-length Tie2 molecule (SEQ ID NO:1).

[0015] **Fig. 2.** Histogram showing the extent of angiopoietin (hAng1, hAng2, mAng3 or hAng4) binding to a hTie2-coated sensor tip pre-treated with anti-Tie2 antibody H4H2055N or Control I.

[0016] **Fig. 3.** Histogram showing the extent of anti-Tie2 antibody (H4H2055N or Control I) binding to a hTie2-coated sensor tip pre-treated with 100 nM of angiopoietin (hAng1, hAng2, mAng3 or hAng4).

[0017] **Fig. 4.** Panel A shows the H29 tumor growth curves in mice following administration of anti-Tie2 antibody H2M2055N (■) or Fc control (●). Downward arrow indicates the treatment start date. Panel B shows the tumor growth from the start of treatment in mice treated with anti-Tie2 antibody H2M2055N or Fc control. Asterisk (*) indicates $p<0.05$ Mann Whitney non-parametric two-tailed t-test.

[0018] **Fig. 5.** H29 tumor vessel density measured at the end of the experiment in mice treated with anti-Tie2 antibody H2M2055N or Fc control. Asterisk (*) indicates $p<0.05$ Mann Whitney non-parametric two-tailed t-test.

[0019] **Fig. 6.** Panel A shows the Colo305 tumor growth curves in mice following administration of anti-Tie2 antibody H2M2055N (■) or Fc control (●). Downward arrow indicates the treatment start date. Panel B shows the tumor growth from the start of treatment in mice treated with anti-Tie2 antibody H2M2055N or Fc control. Asterisk (*) indicates $p<0.05$ Mann Whitney non-parametric two-tailed t-test.

[0020] **Fig. 7.** Colo205 tumor vessel density measured at the end of the experiment in mice treated with anti-Tie2 antibody H2M2055N or Fc control. Asterisk (*) indicates $p<0.05$ Mann Whitney non-parametric two-tailed t-test.

DETAILED DESCRIPTION

[0021] Before the present invention is described, it is to be understood that this invention is not limited to particular methods and experimental conditions described, as such methods and conditions may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

[0022] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. As used herein, the term "about," when used in reference to a particular recited numerical value, means that the value may vary from the recited value by no more than 1%. For example, as used herein, the expression "about 100" includes 99 and 101 and all values in between (*e.g.*, 99.1, 99.2, 99.3, 99.4, etc.).

[0023] Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods and materials are now described.

Definitions

[0024] The expressions "Tie2" and "Tie2 fragment," as used herein refer to the human Tie2 protein or fragment unless specified as being from a non-human species (*e.g.*, "mouse Tie2," "mouse Tie2 fragment," "monkey Tie2," "monkey Tie2 fragment," etc.). A "Tie2 fragment" is any portion of Tie2 having fewer amino acids than the full-length Tie2 molecule and which is capable of binding to a Tie2 ligand. Human Tie2 has the amino acid sequence set forth in SEQ ID NO:1. Amino acid sequences of Tie2 molecules from non-human species (*e.g.*, mouse, monkey, rabbit, dog, pig, etc.) are available from public sources such as GenBank (*e.g.*, GenBank accession numbers NP_038718.2 (mouse); NP_001099207.1 (rat); etc).

[0025] The term "Tie2 ligand," as used herein, means a protein with which the Tie2 protein interacts to transmit a biological signal *in vivo*. The term "Tie2 ligand " includes any of the angiopoietins, including, *e.g.*, Ang1, Ang2, Ang3 and/or Ang4. The term "Ang1," as used herein, means a protein comprising the amino acid sequence of SEQ ID NO:15, or a portion thereof which is capable of interacting with Tie2. The term "Ang2," as used herein, means a protein comprising the amino acid sequence of SEQ ID NO:16, or a portion thereof which is capable of interacting with Tie2. The term "Ang3," as used herein, means a protein comprising the amino acid sequence of SEQ ID NO:17, or a portion thereof which is capable of interacting with Tie2. The term "Ang4," as used herein, means a protein comprising the amino acid sequence as set forth in of SEQ ID NO:18, or a portion thereof which is capable of interacting with Tie2.

[0026] The term "antibody", as used herein, is intended to refer to immunoglobulin molecules comprising four polypeptide chains, two heavy (H) chains and two light (L) chains interconnected by disulfide bonds, as well as multimers thereof (*e.g.*, IgM). Each heavy chain comprises a heavy chain variable region (abbreviated herein as HCVR or V_H) and a heavy chain constant region. The heavy chain constant region comprises three domains, C_H1, C_H2 and C_H3. Each light chain comprises a light chain variable region (abbreviated herein as LCVR or V_L) and a light chain constant region. The light chain constant region comprises one domain (C_L1). The

V_H and V_L regions can be further subdivided into regions of hypervariability, termed complementarity determining regions (CDRs), interspersed with regions that are more conserved, termed framework regions (FR). Each V_H and V_L is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. In different embodiments of the invention, the FRs of the anti-Tie2 antibody (or antigen-binding portion thereof) may be identical to the human germline sequences, or may be naturally or artificially modified. An amino acid consensus sequence may be defined based on a side-by-side analysis of two or more CDRs.

[0027] The term "antibody," as used herein, also includes antigen-binding fragments of full antibody molecules. The terms "antigen-binding portion" of an antibody, "antigen-binding fragment" of an antibody, and the like, as used herein, include any naturally occurring, enzymatically obtainable, synthetic, or genetically engineered polypeptide or glycoprotein that specifically binds an antigen to form a complex. Antigen-binding fragments of an antibody may be derived, *e.g.*, from full antibody molecules using any suitable standard techniques such as proteolytic digestion or recombinant genetic engineering techniques involving the manipulation and expression of DNA encoding antibody variable and optionally constant domains. Such DNA is known and/or is readily available from, *e.g.*, commercial sources, DNA libraries (including, *e.g.*, phage-antibody libraries), or can be synthesized. The DNA may be sequenced and manipulated chemically or by using molecular biology techniques, for example, to arrange one or more variable and/or constant domains into a suitable configuration, or to introduce codons, create cysteine residues, modify, add or delete amino acids, etc.

[0028] Non-limiting examples of antigen-binding fragments include: (i) Fab fragments; (ii) F(ab')2 fragments; (iii) Fd fragments; (iv) Fv fragments; (v) single-chain Fv (scFv) molecules; (vi) dAb fragments; and (vii) minimal recognition units consisting of the amino acid residues that mimic the hypervariable region of an antibody (*e.g.*, an isolated complementarity determining region (CDR) such as a CDR3 peptide), or a constrained FR3-CDR3-FR4 peptide. Other engineered molecules, such as domain-specific antibodies, single domain antibodies, domain-deleted antibodies, chimeric antibodies, CDR-grafted antibodies, diabodies, triabodies, tetrabodies, minibodies, nanobodies (*e.g.* monovalent nanobodies, bivalent nanobodies, etc.), small modular immunopharmaceuticals (SMIPs), and shark variable IgNAR domains, are also encompassed within the expression "antigen-binding fragment," as used herein.

[0029] An antigen-binding fragment of an antibody will typically comprise at least one variable domain. The variable domain may be of any size or amino acid composition and will generally comprise at least one CDR which is adjacent to or in frame with one or more framework sequences. In antigen-binding fragments having a V_H domain associated with a V_L domain, the V_H and V_L domains may be situated relative to one another in any suitable arrangement. For example, the variable region may be dimeric and contain V_H-V_H, V_H-V_L or V_L-V_L dimers. Alternatively, the antigen-binding fragment of an antibody may contain a monomeric V_H or V_L

domain.

[0030] In certain embodiments, an antigen-binding fragment of an antibody may contain at least one variable domain covalently linked to at least one constant domain. Non-limiting, exemplary configurations of variable and constant domains that may be found within an antigen-binding fragment of an antibody of the present invention include: (i) V_H-C_H1; (ii) V_H-C_H2; (iii) V_H-C_H3; (iv) V_H-C_H1-C_H2; (v) V_H-C_H1-C_H2-C_H3; (vi) V_H-C_H2-C_H3; (vii) V_H-C_L; (viii) V_L-C_H1; (ix) V_L-C_H2; (x) V_L-C_H3; (xi) V_L-C_H1-C_H2; (xii) V_L-C_H1-C_H2-C_H3; (xiii) V_L-C_H2-C_H3; and (xiv) V_L-C_L. In any configuration of variable and constant domains, including any of the exemplary configurations listed above, the variable and constant domains may be either directly linked to one another or may be linked by a full or partial hinge or linker region. A hinge region may consist of at least 2 (*e.g.*, 5, 10, 15, 20, 40, 60 or more) amino acids which result in a flexible or semi-flexible linkage between adjacent variable and/or constant domains in a single polypeptide molecule. Moreover, an antigen-binding fragment of an antibody of the present invention may comprise a homo-dimer or hetero-dimer (or other multimer) of any of the variable and constant domain configurations listed above in non-covalent association with one another and/or with one or more monomeric V_H or V_L domain (*e.g.*, by disulfide bond(s)).

[0031] As with full antibody molecules, antigen-binding fragments may be monospecific or multispecific (*e.g.*, bispecific). A multispecific antigen-binding fragment of an antibody will typically comprise at least two different variable domains, wherein each variable domain is capable of specifically binding to a separate antigen or to a different epitope on the same antigen. Any multispecific antibody format, including the exemplary bispecific antibody formats disclosed herein, may be adapted for use in the context of an antigen-binding fragment of an antibody of the present invention using routine techniques available in the art.

[0032] The antibodies of the present invention may function through complement-dependent cytotoxicity (CDC) or antibody-dependent cell-mediated cytotoxicity (ADCC). "Complement-dependent cytotoxicity" (CDC) refers to lysis of antigen-expressing cells by an antibody of the invention in the presence of complement. "Antibody-dependent cell-mediated cytotoxicity" (ADCC) refers to a cell-mediated reaction in which nonspecific cytotoxic cells that express Fc receptors (FcRs) (*e.g.*, Natural Killer (NK) cells, neutrophils, and macrophages) recognize bound antibody on a target cell and thereby lead to lysis of the target cell. CDC and ADCC can be measured using assays that are well known and available in the art. (*See*, *e.g.*, U.S. 5,500,362 and 5,821,337, and Clynes *et al.* (1998) Proc. Natl. Acad. Sci. (USA) *95*:652-656). The constant region of an antibody is important in the ability of an antibody to fix complement and mediate cell-dependent cytotoxicity. Thus, the isotype of an antibody may be selected on the basis of whether it is desirable for the antibody to mediate cytotoxicity.

[0033] The term "human antibody", as used herein, is intended to include antibodies having variable and constant regions derived from human germline immunoglobulin sequences. The human antibodies of the invention may include amino acid residues not encoded by human germline immunoglobulin sequences (e.g., mutations introduced by random or site-specific mutagenesis *in vitro* or by somatic mutation *in vivo*), for example in the CDRs and in particular CDR3. However, the term "human antibody", as used herein, is not intended to include antibodies in which CDR sequences derived from the germline of another mammalian species, such as a mouse, have been grafted onto human framework sequences.

[0034] The term "recombinant human antibody", as used herein, is intended to include all human antibodies that are prepared, expressed, created or isolated by recombinant means, such as antibodies expressed using a recombinant expression vector transfected into a host cell (described further below), antibodies isolated from a recombinant, combinatorial human antibody library (described further below), antibodies isolated from an animal (e.g., a mouse) that is transgenic for human immunoglobulin genes (see e.g., Taylor et al. (1992) Nucl. Acids Res. 20:6287-6295) or antibodies prepared, expressed, created or isolated by any other means that involves splicing of human immunoglobulin gene sequences to other DNA sequences. Such recombinant human antibodies have variable and constant regions derived from human germline immunoglobulin sequences. In certain embodiments, however, such recombinant human antibodies are subjected to *in vitro* mutagenesis (or, when an animal transgenic for human Ig sequences is used, *in vivo* somatic mutagenesis) and thus the amino acid sequences of the V_H and V_L regions of the recombinant antibodies are sequences that, while derived from and related to human germline V_H and V_L sequences, may not naturally exist within the human antibody germline repertoire *in vivo*.

[0035] Human antibodies can exist in two forms that are associated with hinge heterogeneity. In one form, an immunoglobulin molecule comprises a stable four chain construct of approximately 150-160 kDa in which the dimers are held together by an interchain heavy chain disulfide bond. In a second form, the dimers are not linked via inter-chain disulfide bonds and a molecule of about 75-80 kDa is formed composed of a covalently coupled light and heavy chain (half-antibody). These forms have been extremely difficult to separate, even after affinity purification.

[0036] The frequency of appearance of the second form in various intact IgG isotypes is due to, but not limited to, structural differences associated with the hinge region isotype of the antibody. A single amino acid substitution in the hinge region of the human IgG4 hinge can significantly reduce the appearance of the second form (Angal et al. (1993) Molecular Immunology 30:105) to levels typically observed using a human IgG1 hinge. The instant invention encompasses antibodies having one or more mutations in the hinge, C_H2 or C_H3 region which may be desirable, for example, in production, to improve the yield of the desired antibody form.

[0037] An "isolated antibody," as used herein, means an antibody that has been identified and separated and/or recovered from at least one component of its natural environment. For example, an antibody that has been separated or removed from at least one component of an organism, or from a tissue or cell in which the antibody naturally exists or is naturally produced, is an "isolated antibody" for purposes of the present invention. An isolated antibody also includes an antibody *in situ* within a recombinant cell. Isolated antibodies are antibodies that have been subjected to at least one purification or isolation step. According to certain embodiments, an isolated antibody may be substantially free of other cellular material and/or chemicals.

[0038] The term "specifically binds," or the like, means that an antibody or antigen-binding fragment thereof forms a complex with an antigen that is relatively stable under physiologic conditions. Methods for determining whether an antibody specifically binds to an antigen are well known in the art and include, for example, equilibrium dialysis, surface plasmon resonance, and the like. For example, an antibody that "specifically binds" human Tie2, as used in the context of the present invention, includes antibodies that bind human Tie2 or portion thereof with a K_D of less than about 1000 nM, less than about 500 nM, less than about 300 nM, less than about 200 nM, less than about 100 nM, less than about 90 nM, less than about 80 nM, less than about 70 nM, less than about 60 nM, less than about 50 nM, less than about 40 nM, less than about 30 nM, less than about 20 nM, less than about 10 nM, less than about 5 nM, less than about 4 nM, less than about 3 nM, less than about 2 nM, less than about 1 nM or less than about 0.5 nM, as measured in a surface plasmon resonance assay. (See, *e.g.*, Example 2, herein). An isolated antibody that specifically binds human Tie2 may, however, have cross-reactivity to other antigens, such as Tie2 molecules from other (non-human) species.

[0039] A "neutralizing" or "blocking" antibody, as used herein, is intended to refer to an antibody whose binding to Tie2: (i) interferes with the interaction between Tie2 or a Tie2 fragment and a Tie2 ligand (*e.g.*, an angiopoietin), and/or (ii) results in inhibition of at least one biological function of Tie2. The inhibition caused by a Tie2 neutralizing or blocking antibody need not be complete so long as it is detectable using an appropriate assay. Exemplary assays for detecting TIE2 inhibition are described herein.

[0040] The anti-Tie2 antibodies disclosed herein may comprise one or more amino acid substitutions, insertions and/or deletions in the framework and/or CDR regions of the heavy and light chain variable domains as compared to the corresponding germline sequences from which the antibodies were derived. Such mutations can be readily ascertained by comparing the amino acid sequences disclosed herein to germline sequences available from, for example, public antibody sequence databases. The present invention includes antibodies, and antigen-binding fragments thereof, which are derived from any of the amino acid sequences disclosed herein, wherein one or more amino acids within one or more framework and/or CDR regions are mutated to the corresponding residue(s) of the germline sequence from which the antibody was derived, or to the corresponding residue(s) of another human germline sequence, or to a conservative amino acid substitution of the corresponding germline residue(s) (such sequence changes are referred to herein collectively as "germline mutations"). A person of ordinary skill in the art, starting with the heavy and light chain variable region sequences disclosed herein, can easily produce numerous antibodies and antigen-binding fragments which comprise one or more individual germline mutations or combinations thereof. In certain embodiments, all of the framework and/or CDR residues within the V_H and/or V_L domains are mutated back to the residues found in the original germline sequence from which the antibody was derived. In other embodiments, only certain residues are mutated back to the original germline sequence, *e.g.*, only the mutated residues found within the first 8 amino acids of FR1 or within the last 8 amino acids of FR4, or only the mutated residues found within CDR1, CDR2 or CDR3. In other embodiments, one or more of the framework and/or CDR residue(s) are mutated to the corresponding residue(s) of a different germline sequence (*i.e.*, a germline sequence that is different from the germline sequence from which the antibody was originally derived). Furthermore, the antibodies of the present invention may contain any combination of two or more germline mutations within the framework and/or CDR regions, *e.g.*, wherein certain individual residues are mutated to the corresponding residue of a particular germline sequence while certain other residues that differ from the original germline sequence are maintained or are mutated to the corresponding residue of a different germline sequence. Once obtained, antibodies and antigen-binding fragments that contain one or more germline mutations can be easily tested for one or more desired property such as, improved binding specificity, increased binding affinity, improved or enhanced antagonistic or agonistic biological properties (as the case may be), reduced immunogenicity, etc. Antibodies and antigen-binding fragments obtained in this general manner are encompassed within the present invention.

[0041] The present invention also includes anti-Tie2 antibodies comprising variants having one or more conservative substitutions as compared with the HCVR, LCVR and/or CDR amino acid sequences found within the exemplary anti-Tie2 antibodies disclosed herein. For example, the present invention includes anti-Tie2 antibodies having HCVR, LCVR, and/or CDR amino acid sequences with, *e.g.*, 10 or fewer, 8 or fewer, 6 or fewer, 4 or fewer, etc. conservative amino acid substitutions relative to any of the HCVR, LCVR, and/or CDR amino acid sequences of the anti-Tie2 antibodies disclosed herein.

[0042] The term "surface plasmon resonance", as used herein, refers to an optical phenomenon that allows for the analysis of real-time interactions by detection of alterations in protein concentrations within a biosensor matrix, for example using the BIAcore™ system (Biacore Life Sciences division of GE Healthcare, Piscataway, NJ).

[0043] The term "K_D ", as used herein, is intended to refer to the equilibrium dissociation constant of a particular antibody-antigen interaction.

[0044] The term "epitope" refers to an antigenic determinant that interacts with a specific antigen binding site in the variable region of an antibody molecule known as a paratope. A single antigen may have more than one epitope. Thus, different antibodies may bind to different areas on an antigen and may have different biological effects. Epitopes may be either conformational or linear. A conformational epitope is produced by spatially juxtaposed amino acids from different segments of the linear polypeptide chain. A linear epitope is one produced by adjacent amino acid residues in a polypeptide chain. In certain circumstance, an epitope may include moieties of saccharides, phosphoryl groups, or sulfonyl groups on the antigen.

[0045] The term "substantial identity" or "substantially identical," when referring to a nucleic acid or fragment thereof, indicates that, when optimally aligned with appropriate nucleotide insertions or deletions with another nucleic acid (or its complementary strand), there is nucleotide sequence identity in at least about 95%, and more preferably at least about 96%, 97%, 98% or 99% of the nucleotide bases, as measured by any well-known algorithm of sequence identity, such as FASTA, BLAST or Gap, as discussed below. A nucleic acid molecule having substantial identity to a reference nucleic acid molecule may, in certain instances, encode a polypeptide having the same or substantially similar amino acid sequence as the polypeptide encoded by the reference nucleic acid molecule.

[0046] As applied to polypeptides, the term "substantial similarity" or "substantially similar" means that two peptide sequences, when optimally aligned, such as by the programs GAP or BESTFIT using default gap weights, share at least 95% sequence identity, even more preferably at least 98% or 99% sequence identity. Preferably, residue positions which are not identical differ by conservative amino acid substitutions. A "conservative amino acid substitution" is one in which an amino acid residue is substituted by another amino acid residue having a side chain (R group) with similar chemical properties (e.g., charge or hydrophobicity). In general, a conservative amino acid substitution will not substantially change the functional properties of a protein. In cases where two or more amino acid sequences differ from each other by conservative substitutions, the percent sequence identity or degree of similarity may be adjusted upwards to correct for the conservative nature of the substitution. Means for making this adjustment are well-known to those of skill in the art. *See, e.g.*, Pearson (1994) Methods Mol. Biol. 24: 307-331. Examples of groups of amino acids that have side chains with similar chemical properties include (1) aliphatic side chains: glycine, alanine, valine, leucine and isoleucine; (2) aliphatic-hydroxyl side chains: serine and threonine; (3) amide-containing side chains: asparagine and glutamine; (4) aromatic side chains: phenylalanine, tyrosine, and tryptophan; (5) basic side chains: lysine, arginine, and histidine; (6) acidic side chains: aspartate and glutamate, and (7) sulfur-containing side chains are cysteine and methionine. Preferred conservative amino acids substitution groups are: valine-leucine-isoleucine, phenylalanine-tyrosine, lysine-arginine, alanine-valine, glutamate-aspartate, and asparagine-glutamine. Alternatively, a conservative replacement is any change having a positive value in the PAM250 log-likelihood matrix disclosed in Gonnet *et al.* (1992) Science 256: 1443-1445. A "moderately conservative" replacement is any change having a nonnegative value in the PAM250 log-likelihood matrix.

[0047] Sequence similarity for polypeptides, which is also referred to as sequence identity, is typically measured using sequence analysis software. Protein analysis software matches similar sequences using measures of similarity assigned to various substitutions, deletions and other modifications, including conservative amino acid substitutions. For instance, GCG software contains programs such as Gap and Bestfit which can be used with default parameters to determine sequence homology or sequence identity between closely related polypeptides, such as homologous polypeptides from different species of organisms or between a wild type protein and a mutein thereof. *See, e.g.*, GCG Version 6.1. Polypeptide sequences also can be compared using FASTA using default or recommended parameters, a program in GCG Version 6.1. FASTA (e.g., FASTA2 and FASTA3) provides alignments and percent sequence identity of the regions of the best overlap between the query and search sequences (Pearson (2000) supra). Another preferred algorithm when comparing a sequence of the invention to a database containing a large number of sequences from different organisms is the computer program BLAST, especially BLASTP or TBLASTN, using default parameters. *See, e.g.*, Altschul *et al.* (1990) J. Mol. Biol. 215:403-410 and Altschul *et al.* (1997) Nucleic Acids Res. 25:3389-402.

Biological Characteristics of the Antibodies

[0048] The present invention includes antibodies that block the interaction between Tie2 and a Tie2 ligand. As used herein, the expression "blocks the interaction between Tie2 and a Tie2 ligand" means that, in an assay in which the physical interaction between Tie2 and a Tie2 ligand can be detected and/or quantified, the addition of an antibody of the invention reduces the interaction between Tie2 and the Tie2 ligand (*e.g.*, Ang1, Ang2, Ang3 and/or Ang4) by at least 50%. A non-limiting, exemplary assay that can be used to determine if an antibody blocks the interaction between human Tie2 and a Tie2 ligand is illustrated in Example 5, herein. In one exemplary embodiment of this assay format, antibodies are mixed with Tie2 protein, and then the antibody/Tie2 mixture is applied to a surface coated with a Tie 2 ligand (in this case, Ang2 protein). After washing away unbound molecules, the amount of Tie2 bound to the Ang2-coated surface is measured. By using varying amounts of antibody in this assay format, the amount of antibody required to block 50% of Tie2 binding to Ang2 can be calculated and expressed as an IC_{50} value. The format of this assay can be reversed such that Tie2 is coated to surface, antibody is added to the Tie2-coated surface, unbound antibody is washed away, and then a Tie2 ligand is added to the antibody-treated Tie2 surface. The present invention includes anti-Tie2 antibodies that exhibit an IC_{50} of less than about 100 nM when tested in a Tie2/Tie2 ligand binding assay as illustrated in Example 5, or a substantially similar assay. For example, the invention includes anti-Tie2 antibodies that exhibit an IC_{50} of less than about 100, 90, 80, 70, 60, 50, 40, 30, 20, 19, 18, 17, 16, 15, 14, 12, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1, 0.5, 0.4, 0.3, or 0.2 nM when tested in a Tie2/Tie2 ligand binding assay as illustrated in Example 5, or a substantially similar assay.

[0049] Another assay format that can be used to determine whether an antibody blocks the interaction between human Tie2 and a Tie2 ligand is illustrated in Example 6, herein. In this assay format, a cell line is used which is engineered to express human Tie2 on its surface, and which also includes a reporter construct that causes a detectable signal to be expressed when Tie2 interacts with a Tie2 ligand. The engineered cells are treated with anti-Tie2 antibodies and with Tie2 ligand and the reporter signal is measured. By using varying amounts of antibody in this assay format, the amount of antibody required to inhibit 50% of the reporter signal observed in the absence of antibody can be calculated and expressed as an IC_{50} value. The present invention includes anti-Tie2 antibodies that exhibit an IC_{50} of less than about 20 nM when tested in a Tie2/Tie2 ligand binding assay as illustrated in Example 6, or a substantially similar assay. For example, the invention includes anti-Tie2 antibodies that exhibit an IC_{50} of less than about 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1, 0.5, 0.4, 0.3, 0.2 or 0.1 nM when tested in a Tie2/Tie2 ligand binding assay as described in Example 6, or a substantially similar assay.

Epitope Mapping and Related Technologies

[0050] The human Tie2 protein contains the following domains: an Ig1 domain, an Ig2 domain, an EGF repeat domain, an Ig3 domain, and a fibronectin repeat (FN) domain (including FN1, FN2 and FN3). These domains are graphically depicted in Figure1. The present invention includes anti-Tie2 antibodies which bind specifically to an epitope within one or more of the following regions: (a) the Ig1-Ig2-EGF domains (SEQ ID NO:7); (b) the Ig2-EGF domains (SEQ ID NO:8); (c) the EGF domain (SEQ ID NO:9); (d) the Ig3-FN domains (SEQ ID NO:10); and/or (e) the FN domains (SEQ ID NO:11). (See Examples 3 and 4).

[0051] According to certain embodiments, the present invention provides anti-Tie2 antibodies which interact with one or more amino acids found within the Ig1 and/or Ig2 domains of Tie2. The epitope(s) may consist of one or more contiguous sequences of 3 or more (*e.g.*, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20 or more) amino acids located within the Ig1 and/or Ig2 domains of Tie2. Alternatively, the epitope may consist of a plurality of non-contiguous amino acids (or amino acid sequences) located within the Ig1 and/or Ig2 domains of Tie2. According to certain embodiments of the present invention, anti-Tie2 antibodies are provided which interact with one or more amino acids located within one or more amino acid segments selected from the group consisting of amino acids 96-106 of SEQ ID NO:7, amino acids 139-152 of SEQ ID NO:7; and amino acids 166-175 of SEQ ID NO:7. For example, the present invention includes anti-Tie2 antibodies which interact with at least one amino acid within each of the aforementioned segments (*i.e.*, within each of amino acids 96-106, 139-152, and 166-175 of SEQ ID NO:7). According to certain embodiments of the present invention, antibodies which interact with amino acids 139-152 and/or 166-175 of SEQ ID NO:7 are capable of blocking the interaction between Tie2 and one or more Tie2 ligands, such as, *e.g.*, Ang2 (see Examples 4-6, herein).

[0052] Various techniques known to persons of ordinary skill in the art can be used to determine whether an antibody "interacts with one or more amino acids" within a polypeptide or protein. Exemplary techniques include, *e.g.*, routine cross-blocking assay such as that described Antibodies, Harlow and Lane (Cold Spring Harbor Press, Cold Spring Harb., NY), alanine scanning mutational analysis, peptide blots analysis (Reineke, 2004, Methods Mol Biol 248:443-463), and peptide cleavage analysis. In addition, methods such as epitope excision, epitope extraction and chemical modification of antigens can be employed (Tomer, 2000, Protein Science 9:487-496). Another method that can be used to identify the amino acids within a polypeptide with which an antibody interacts is hydrogen/deuterium exchange detected by mass spectrometry. (See, *e.g.*, Example 4 herein). In general terms, the hydrogen/deuterium exchange method involves deuterium-labeling the protein of interest, followed by binding the antibody to the deuterium-labeled protein. Next, the protein/antibody complex is transferred to water to allow hydrogen-deuterium exchange to occur at all residues except for the residues protected by the antibody (which remain deuterium-labeled). After dissociation of the antibody, the target protein is subjected to protease cleavage and mass spectrometry analysis, thereby revealing the deuterium-labeled residues which correspond to the specific amino acids with which the antibody interacts. *See, e.g.*, Ehring (1999) *Analytical Biochemistry* 267(2):252-259; Engen and Smith (2001) *Anal. Chem. 73*:256A-265A.

[0053] The present invention includes anti-Tie2 antibodies that bind to the same epitope as any of the specific exemplary antibodies described herein (*e.g.*, antibodies H1M2055N, and H2aM2760N, produced from deposited cell lines PTA-12295 and PTA-12296, respectively). Likewise, the present invention also includes anti-Tie2 antibodies that compete for binding to Tie2 or a Tie2 fragment with any of the specific exemplary antibodies described herein (*e.g.*, antibodies H1M2055N, and H2aM2760N, produced from deposited cell lines PTA-12295 and PTA-12296, respectively).

[0054] One can easily determine whether an antibody binds to the same epitope as, or competes for binding with, a reference anti-Tie2 antibody by using routine methods known in the art. For example, to determine if a test antibody binds to the same epitope as a reference anti-Tie2 antibody of the invention, the reference antibody is allowed to bind to a Tie2 protein or peptide under saturating conditions. Next, the ability of a test antibody to bind to the Tie2 molecule is assessed. If the test antibody is able to bind to Tie2 following saturation binding with the reference anti-Tie2 antibody, it can be concluded that the test antibody binds to a different epitope than the reference anti-Tie2 antibody. On the other hand, if the test antibody is not able to bind to the Tie2 molecule following saturation binding with the reference anti-Tie2 antibody, then the test antibody may bind to the same epitope as the epitope bound by the reference anti-Tie2 antibody of the invention. Additional routine experimentation (*e.g.*, peptide mutation and binding analyses) can then be carried out to confirm whether the observed lack of binding of the test antibody is in fact due to binding to the same epitope as the reference antibody or if steric blocking (or another phenomenon) is responsible for the lack of observed binding. Experiments of this sort can be performed using ELISA, RIA, Biacore, flow cytometry or any other quantitative or qualitative antibody-binding assay available in the art. In accordance with certain embodiments of the present invention, two antibodies bind to the same (or overlapping) epitope if, *e.g.*, a 1-, 5-, 10-, 20- or 100-fold excess of one antibody inhibits binding of the other by at least 50% but preferably 75%, 90% or even 99% as measured in a competitive binding assay (see, e.g., Junghans et al., Cancer Res. 1990:50:1495-1502). Alternatively, two antibodies are deemed to bind to the same epitope if essentially all amino acid mutations in the antigen that reduce or eliminate binding of one antibody reduce or eliminate binding of the other. Two antibodies are deemed to have "overlapping epitopes" if only a subset of the amino acid mutations that reduce or eliminate binding of one antibody reduce or eliminate binding of the other.

[0055] To determine if an antibody competes for binding with a reference anti-Tie2 antibody, the above-described binding methodology is performed in two orientations: In a first orientation, the reference antibody is allowed to bind to a Tie2 molecule under saturating conditions followed by assessment of binding of the test antibody to the Tie2 molecule. In a second orientation, the test antibody is allowed to bind to a Tie2 molecule under saturating conditions followed by assessment of binding of the reference antibody to the Tie2 molecule. If, in both orientations, only the first (saturating) antibody is capable of binding to the Tie2 molecule, then it is concluded that the test antibody and the reference antibody compete for binding to Tie2. As will be appreciated by a person of ordinary skill in the art, an antibody that competes for binding with a reference antibody may not necessarily bind to the same epitope as the reference antibody, but may sterically block binding of the reference antibody by binding an overlapping or adjacent epitope.

Preparation of Human Antibodies

[0056] Methods for generating monoclonal antibodies, including fully human monoclonal antibodies are known in the art. Any such known methods can be used in the context of the present invention to make human antibodies that specifically bind to human Tie2.

[0057] Using VELOCIMMUNE™ technology or any other known method for generating monoclonal antibodies, high affinity chimeric antibodies to Tie2 are initially isolated having a human variable region and a mouse constant region. As in the experimental section below, the antibodies are characterized and selected for desirable characteristics, including affinity, selectivity, epitope, etc. The mouse constant regions are replaced with a desired human constant region to generate the fully human antibody of the invention, for example wild-type or modified IgG1 or IgG4. While the constant region selected may vary according to specific use, high affinity antigen-binding and target specificity characteristics reside in the variable region.

Bioequivalents

[0058] The anti-Tie2 antibodies and antibody fragments of the present invention encompass proteins having amino acid sequences that vary from those of the described antibodies but that retain the ability to bind human Tie2. Such variant antibodies and antibody fragments comprise one or more additions, deletions, or substitutions of amino acids when compared to parent sequence, but exhibit biological activity that is essentially equivalent to that of the described antibodies. Likewise, the anti-Tie2 antibody-encoding DNA sequences of the present invention encompass sequences that comprise one or more additions, deletions, or substitutions of nucleotides when compared to the disclosed sequence, but that encode an anti-Tie2 antibody or antibody fragment that is essentially bioequivalent to an anti-Tie2 antibody or antibody fragment of the invention.

[0059] Two antigen-binding proteins, or antibodies, are considered bioequivalent if, for example, they are pharmaceutical equivalents or pharmaceutical alternatives whose rate and extent of absorption do not show a significant difference when administered at the same molar dose under similar experimental conditions, either single does or multiple dose. Some antibodies will be considered equivalents or pharmaceutical alternatives if they are equivalent in the extent of their absorption but not in their rate of absorption and yet may be considered bioequivalent because such differences in the rate of absorption are intentional and are reflected in the labeling, are not essential to the attainment of effective body drug concentrations on, e.g., chronic use, and are considered medically insignificant for the particular drug product studied.

[0060] In one embodiment, two antigen-binding proteins are bioequivalent if there are no clinically meaningful differences in their safety, purity, and potency.

[0061] In one embodiment, two antigen-binding proteins are bioequivalent if a patient can be switched one or more times between the reference product and the biological product without an expected increase in the risk of adverse effects, including a clinically significant change in immunogenicity, or diminished effectiveness, as compared to continued therapy without such switching.

[0062] In one embodiment, two antigen-binding proteins are bioequivalent if they both act by a common mechanism or mechanisms of action for the condition or conditions of use, to the extent that such mechanisms are known.

[0063] Bioequivalence may be demonstrated by in vivo and in vitro methods. Bioequivalence measures include, e.g., (a) an in vivo test in humans or other mammals, in which the concentration of the antibody or its metabolites is measured in blood, plasma, serum, or other biological fluid as a function of time; (b) an in vitro test that has been correlated with and is reasonably predictive of human in vivo bioavailability data; (c) an in vivo test in humans or other mammals in which the appropriate acute pharmacological effect of the antibody (or its target) is measured as a function of time; and (d) in a well-controlled clinical trial that establishes safety, efficacy, or bioavailability or bioequivalence of an antibody.

[0064] Bioequivalent variants of anti-Tie2 antibodies of the invention may be constructed by, for example, making various substitutions of residues or sequences or deleting terminal or internal residues or sequences not needed for biological activity. For example, cysteine residues not essential for biological activity can be deleted or replaced with other amino acids to prevent formation of unnecessary or incorrect intramolecular disulfide bridges upon renaturation. In other contexts, bioequivalent antibodies may include anti-Tie2 antibody variants comprising amino acid changes which modify the glycosylation characteristics of the antibodies, *e.g.*, mutations which eliminate or remove glycosylation.

Species Selectivity and Species Cross-Reactivity

[0065] According to certain embodiments of the invention, the anti-Tie2 antibodies bind to human Tie2 but not to Tie2 from other species. The present invention also includes anti-Tie2 antibodies that bind to human Tie2 and to Tie2 from one or more non-human species. For example, the anti-Tie2 antibodies of the invention may specifically bind to human Tie2 as well as to a rodent Tie2 (*e.g.*, Tie2 from mouse or rat). An exemplary construct that can be used to determine whether an antibody specifically binds mouse Tie2 is the construct having the amino acid sequence of SEQ ID NO:6; an exemplary construct that can be used to determine whether an antibody specifically binds rat Tie2 is the construct having the amino acid sequence of SEQ ID NO:5. The use of these constructs to assess anti-Tie2 antibody binding is illustrated in Example 2, herein.

Immunoconjugates

[0066] The invention encompasses anti-Tie2 monoclonal antibodies conjugated to a therapeutic moiety ("immunoconjugate"), such as a cytotoxin, a chemotherapeutic drug, an immunosuppressant or a radioisotope. Cytotoxic agents include any agent that is detrimental to cells. Examples of suitable cytotoxic agents and chemotherapeutic agents for forming immunoconjugates are known in the art, (see for example, WO 05/103081).

Multispecific Antibodies

[0067] The antibodies of the present invention may be monospecific, bi-specific, or multispecific. Multispecific antibodies may be specific for different epitopes of one target polypeptide or may contain antigen-binding domains specific for more than one target polypeptide. See, e.g., Tutt et al., 1991, J. Immunol. 147:60-69; Kufer *et al.*, 2004, Trends Biotechnol. 22:238-244. The anti-Tie2 antibodies of the present invention can be linked to or co-expressed with another functional molecule, e.g., another peptide or protein. For example, an antibody or fragment thereof can be functionally linked (e.g., by chemical coupling, genetic fusion, noncovalent association or otherwise) to one or more other molecular entities, such as another antibody or antibody fragment to produce a bi-specific or a multispecific antibody with a second binding specificity. For example, the present invention includes bi-specific antibodies wherein one arm of an immunoglobulin is specific for human Tie2 or a fragment thereof, and the other arm of the immunoglobulin is specific for a second therapeutic target or is conjugated to a therapeutic moiety such as a trypsin inhibitor.

[0068] An exemplary bi-specific antibody format that can be used in the context of the present invention involves the use of a first immunoglobulin (Ig) C_H3 domain and a second Ig C_H3 domain, wherein the first and second Ig C_H3 domains differ from one another by at least one amino acid, and wherein at least one amino acid difference reduces binding of the bispecific antibody to Protein A as compared to a bi-specific antibody lacking the amino acid difference. In one embodiment, the first Ig C_H3 domain binds Protein A and the second Ig C_H3 domain contains a mutation that reduces or abolishes Protein A binding such as an H95R modification (by IMGT exon numbering; H435R by EU numbering). The second C_H3 may further comprise a Y96F modification (by IMGT; Y436F by EU). Further modifications that may be found within the second C_H3 include: D16E, L18M, N44S, K52N, V57M, and V82I (by IMGT; D356E, L358M, N384S, K392N, V397M, and V422I by EU) in the case of IgG1 antibodies; N44S, K52N, and V82I (IMGT; N384S, K392N, and V422I by EU) in the case of IgG2 antibodies; and Q15R, N44S, K52N, V57M, R69K, E79Q, and V82I (by IMGT; Q355R, N384S, K392N, V397M, R409K, E419Q, and V422I by EU) in the case of IgG4 antibodies. Variations on the bi-specific antibody format described above are contemplated within the scope of the present invention.

Therapeutic Formulation and Administration

[0069] The invention provides pharmaceutical compositions comprising the anti-Tie2 antibodies or antigen-binding fragments thereof of the present invention. The pharmaceutical compositions of the invention are formulated with suitable carriers, excipients, and other agents that provide improved transfer, delivery, tolerance, and the like. A multitude of appropriate formulations can be found in the formulary known to all pharmaceutical chemists: Remington's Pharmaceutical Sciences, Mack Publishing Company, Easton, PA. These formulations include, for example, powders, pastes, ointments, jellies, waxes, oils, lipids, lipid (cationic or anionic) containing vesicles (such as LIPOFECTIN™), DNA conjugates, anhydrous absorption pastes, oil-in-water and water-in-oil emulsions, emulsions carbowax (polyethylene glycols of various molecular weights), semi-solid gels, and semi-solid mixtures containing carbowax. See also Powell et al. "Compendium of excipients for parenteral formulations" PDA (1998) J Pharm Sci Technol 52:238-311.

[0070] The dose of antibody administered to a patient may vary depending upon the age and the size of the patient, target disease, conditions, route of administration, and the like. The preferred dose is typically calculated according to body weight or body surface area. When an antibody of the present invention is used for treating a condition or disease associated with Tie2 activity in an adult patient, it may be advantageous to intravenously administer the antibody of the present invention normally at a single dose of about 0.01 to about 20 mg/kg body weight, more preferably about 0.02 to about 7, about 0.03 to about 5, or about 0.05 to about 3 mg/kg body weight. Depending on the severity of the condition, the frequency and the duration of the treatment can be adjusted. Effective dosages and schedules for administering Tie2 antibodies may be determined empirically; for example, patient progress can be monitored by periodic assessment, and the dose adjusted accordingly. Moreover, interspecies scaling of dosages can be performed using well-known methods in the art (*e.g.*, Mordenti *et al.*, 1991, *Pharmaceut. Res. 8*:1351).

[0071] Various delivery systems are known and can be used to administer the pharmaceutical composition of the invention, e.g., encapsulation in liposomes, microparticles, microcapsules, recombinant cells capable of expressing the mutant viruses, receptor mediated endocytosis (see, e.g., Wu et al., 1987, J. Biol. Chem. 262:4429-4432). Methods of introduction include, but are not limited to, intradermal, intramuscular, intraperitoneal, intravenous, subcutaneous, intranasal, epidural, and oral routes. The composition may be administered by any convenient route, for example by infusion or bolus injection, by absorption through epithelial or mucocutaneous linings (e.g., oral mucosa, rectal and intestinal mucosa, etc.) and may be administered together with other biologically active agents. Administration can be systemic or local.

[0072] A pharmaceutical composition of the present invention can be delivered subcutaneously or intravenously with a standard needle and syringe. In addition, with respect to subcutaneous delivery, a pen delivery device readily has applications in delivering a pharmaceutical composition of the present invention. Such a pen delivery device can be reusable or disposable. A reusable pen delivery device generally utilizes a replaceable cartridge that contains a pharmaceutical composition. Once all of the pharmaceutical composition within the cartridge has been administered and the cartridge is empty, the empty cartridge can readily be discarded and replaced with a new cartridge that contains the pharmaceutical composition. The pen delivery device can then be reused. In a disposable pen delivery device, there is no replaceable cartridge. Rather, the disposable pen delivery device comes prefilled with the pharmaceutical composition held in a reservoir within the device. Once the reservoir is emptied of the pharmaceutical composition, the entire device is discarded.

[0073] Numerous reusable pen and autoinjector delivery devices have applications in the subcutaneous delivery of a pharmaceutical composition of the present invention. Examples include, but are not limited to AUTOPEN™ (Owen Mumford, Inc., Woodstock, UK), DISETRONIC™ pen (Disetronic Medical Systems, Bergdorf, Switzerland), HUMALOG MIX 75/25™ pen, HUMALOG™ pen, HUMALIN 70/30™ pen (Eli Lilly and Co., Indianapolis, IN), NOVOPEN™ I, II and III (Novo Nordisk, Copenhagen, Denmark), NOVOPEN JUNIOR™ (Novo Nordisk, Copenhagen, Denmark), BD™ pen (Becton Dickinson, Franklin Lakes, NJ), OPTIPEN™, OPTIPEN PRO™, OPTIPEN STARLET™, and OPTICLIK™ (sanofi-aventis, Frankfurt, Germany), to name only a few. Examples of disposable pen delivery devices having applications in subcutaneous delivery of a pharmaceutical composition of the present invention include, but are not limited to the SOLOSTAR™ pen (sanofi-aventis), the FLEXPEN™ (Novo

Nordisk), and the KWIKPEN™ (Eli Lilly), the SURECLICK™ Autoinjector (Amgen, Thousand Oaks, CA), the PENLET™ (Haselmeier, Stuttgart, Germany), the EPIPEN (Dey, L.P.), and the HUMIRA™ Pen (Abbott Labs, Abbott Park IL), to name only a few.

[0074] In certain situations, the pharmaceutical composition can be delivered in a controlled release system. In one embodiment, a pump may be used (see Langer, supra; Sefton, 1987, CRC Crit. Ref. Biomed. Eng. 14:201). In another embodiment, polymeric materials can be used; see, Medical Applications of Controlled Release, Langer and Wise (eds.), 1974, CRC Pres., Boca Raton, Florida. In yet another embodiment, a controlled release system can be placed in proximity of the composition's target, thus requiring only a fraction of the systemic dose (see, *e.g.*, Goodson, 1984, in Medical Applications of Controlled Release, supra, vol. 2, pp. 115-138). Other controlled release systems are discussed in the review by Langer, 1990, Science 249:1527-1533.

[0075] The injectable preparations may include dosage forms for intravenous, subcutaneous, intracutaneous and intramuscular injections, drip infusions, etc. These injectable preparations may be prepared by methods publicly known. For example, the injectable preparations may be prepared, *e.g.*, by dissolving, suspending or emulsifying the antibody or its salt described above in a sterile aqueous medium or an oily medium conventionally used for injections. As the aqueous medium for injections, there are, for example, physiological saline, an isotonic solution containing glucose and other auxiliary agents, etc., which may be used in combination with an appropriate solubilizing agent such as an alcohol (e.g., ethanol), a polyalcohol (e.g., propylene glycol, polyethylene glycol), a nonionic surfactant [e.g., polysorbate 80, HCO-50 (polyoxyethylene (50 mol) adduct of hydrogenated castor oil)], etc. As the oily medium, there are employed, e.g., sesame oil, soybean oil, etc., which may be used in combination with a solubilizing agent such as benzyl benzoate, benzyl alcohol, etc. The injection thus prepared is preferably filled in an appropriate ampoule.

[0076] Advantageously, the pharmaceutical compositions for oral or parenteral use described above are prepared into dosage forms in a unit dose suited to fit a dose of the active ingredients. Such dosage forms in a unit dose include, for example, tablets, pills, capsules, injections (ampoules), suppositories, etc. The amount of the aforesaid antibody contained is generally about 5 to about 500 mg per dosage form in a unit dose; especially in the form of injection, it is preferred that the aforesaid antibody is contained in about 5 to about 100 mg and in about 10 to about 250 mg for the other dosage forms.

Therapeutic Uses of the Antibodies

[0077] The antibodies of the invention are useful, *inter alia*, for the treatment, prevention and/or amelioration of any disease or disorder associated with Tie2 activity, including diseases or disorders associated with angiogenesis. The antibodies and antigen-binding fragments of the present invention may be used to treat, *e.g.*, primary and/or metastatic tumors arising in the brain and meninges, oropharynx, lung and bronchial tree, gastrointestinal tract, male and female reproductive tract, muscle, bone, skin and appendages, connective tissue, spleen, immune system, blood forming cells and bone marrow, liver and urinary tract, and special sensory organs such as the eye. In certain embodiments, the antibodies and antigen-binding fragments of the invention are used to treat one or more of the following cancers: renal cell carcinoma, pancreatic carcinoma, breast cancer, prostate cancer, malignant gliomas, osteosarcoma, colorectal cancer, malignant mesothelioma, multiple myeloma, ovarian cancer, small cell lung cancer, non-small cell lung cancer, synovial sarcoma, thyroid cancer, or melanoma.

Combination Therapies

[0078] The present invention includes therapeutic administration regimens which comprise administering an anti-Tie2 antibody of the present invention in combination with at least one additional therapeutically active component. Non-limiting examples of such additional therapeutically active components include, for example, another Tie2 antagonist (*e.g.*, an anti-Tie2 antibody), an antagonist of epidermal growth factor receptor (EGFR) (*e.g.*, anti-EGFR antibody [*e.g.*, cetuximab or panitumumab] or small molecule inhibitor of EGFR activity [*e.g.*, gefitinib or erlotinib]), an antagonist of another EGFR family member such as Her2/ErbB2, ErbB3 or ErbB4 (*e.g.*, anti-ErbB2, anti-ErbB3 or anti-ErbB4 antibody or small molecule inhibitor of ErbB2, ErbB3 or ErbB4 activity), an antagonist of EGFRvIII (*e.g.*, an antibody that specifically binds EGFRvIII), a cMET anagonist (*e.g.*, an anti-cMET antibody), an IGF1R antagonist (*e.g.*, an anti-IGF1R antibody), a B-raf inhibitor (*e.g.*, vemurafenib, sorafenib, GDC-0879, PLX-4720), a PDGFR-α inhibitor (*e.g.*, an anti-PDGFR-α antibody), a PDGFR-β inhibitor (*e.g.*, an anti-PDGFR-β antibody), a VEGF antagonist (*e.g.*, a VEGF-Trap, *see*, *e.g.*, US 7,087,411 (also referred to herein as a "VEGF-inhibiting fusion protein"), anti-VEGF antibody (*e.g.*, bevacizumab), a small molecule kinase inhibitor of VEGF receptor (*e.g.*, sunitinib, sorafenib or pazopanib)), a DLL4 antagonist (*e.g.*, an anti-DLL4 antibody disclosed in US 2009/0142354 such as REGN421), an Ang2 antagonist (*e.g.*, an anti-Ang2 antibody disclosed in US 2011/0027286 such as H1H685P), etc. Other agents that may be beneficially administered in combination with the anti-Tie2 antibodies of the invention include cytokine inhibitors, including small-molecule cytokine inhibitors and antibodies that bind to cytokines such as IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-8, IL-9, IL-11, IL-12, IL-13, IL-17, IL-18, or to their respective receptors.

[0079] The present invention also includes therapeutic combinations comprising any of the anti-Tie2 antibodies mentioned herein and an inhibitor of one or more of VEGF, Ang2, DLL4, ErbB2, ErbB3, ErbB4, EGFRvIII, cMet, IGF1R, B-raf, PDGFR-α, PDGFR-β, or any of the aforementioned cytokines, wherein the inhibitor is an aptamer, an antisense molecule, a ribozyme, an siRNA, a peptibody, a nanobody or an antibody fragment (*e.g.*, Fab fragment; $F(ab')_2$ fragment; Fd fragment; Fv fragment; scFv; dAb fragment; or other engineered molecules, such as diabodies, triabodies, tetrabodies, minibodies and minimal recognition units). The anti-Tie2 antibodies of the invention may also be administered in combination with antivirals, antibiotics, analgesics, corticosteroids and/or NSAIDs. The anti-Tie2 antibodies of the invention may also be administered as part of a treatment regimen that also includes radiation treatment and/or conventional chemotherapy.

[0080] The additional therapeutically active component(s) may be administered just prior to, concurrent with, or shortly after the administration of an anti-Tie2 antibody of the present invention; (for purposes of the present disclosure, such administration regimens are considered the administration of an anti-Tie2 antibody "in combination with" an additional therapeutically active component). The present invention includes pharmaceutical compositions in which an anti-Tie2 antibody of the present invention is co-formulated with one or more of the additional therapeutically active component(s) as described elsewhere herein.

Diagnostic Uses of the Antibodies

[0081] The anti-Tie2 antibodies of the present invention may also be used to detect and/or measure Tie2 in a sample, *e.g.*, for diagnostic purposes. For example, an anti-Tie2 antibody, or fragment thereof, may be used to diagnose a condition or disease characterized by aberrant expression (*e.g.*, over-expression, under-expression, lack of expression, etc.) of Tie2. Exemplary diagnostic assays for Tie2 may comprise, *e.g.*, contacting a sample, obtained from a patient, with an anti-Tie2 antibody of the invention, wherein the anti-Tie2 antibody is labeled with a detectable label or reporter molecule. Alternatively, an unlabeled anti-Tie2 antibody can be used in diagnostic applications in combination with a secondary antibody which is itself detectably labeled. The detectable label or reporter molecule can be a radioisotope, such as ^{3}H, ^{14}C, ^{32}P, ^{35}S, or ^{125}I; a fluorescent or chemiluminescent moiety such as fluorescein isothiocyanate, or rhodamine; or an enzyme such as alkaline phosphatase, beta-galactosidase, horseradish peroxidase, or luciferase. Specific exemplary assays that can be used to detect or measure Tie2 in a sample include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence-activated cell sorting (FACS).

[0082] Samples that can be used in Tie2 diagnostic assays according to the present invention include any tissue or fluid sample obtainable from a patient which contains detectable quantities of Tie2 protein, or fragments thereof, under normal or pathological conditions. Generally, levels of Tie2 in a particular sample obtained from a healthy patient (*e.g.*, a patient not afflicted with a disease or condition associated with abnormal Tie2 levels or activity) will be measured to initially establish a baseline, or standard, level of Tie2. This baseline level of Tie2 can then be compared against the levels of Tie2 measured in samples obtained from individuals suspected of having a Tie2 related disease or condition.

EXAMPLES

[0083] The following examples are put forth so as to provide those of ordinary skill in the art with a complete disclosure and description of how to make and use the methods and compositions of the invention, and are not intended to limit the scope of what the inventors regard as their invention. Efforts have been made to ensure accuracy with respect to numbers used (e.g., amounts, temperature, etc.) but some experimental errors and deviations should be accounted for. Unless indicated otherwise, parts are parts by weight, molecular weight is average molecular weight, temperature is in degrees Centigrade, and pressure is at or near atmospheric.

Control Constructs Used in the Following Examples

[0084] An exemplary control construct (anti-Tie2 antibody) was included in several experiments described below for comparative purposes. The antibody, referred to as Control I, is a chimeric anti-Tie2 antibody with mouse heavy and light chain variable domains having the amino acid sequences of the corresponding domains of "12H8", as set forth in US Patent 6,376,653. The constant domain of this antibody is human IgG4.

Example 1. Generation of Human Antibodies to Human Tie2

[0085] Several human anti-Tie2 antibodies were generated by immunizing a VELOCIMMUNE® mouse with human Tie2 antigen according to standard methods (*see, e.g.*, US Patent No. 6,596,541). Using this technique, several anti-Tie2 antibodies were obtained; exemplary antibodies generated in this manner , and their corresponding biological characteristics, are described in detail in the following Examples and include the antibodies designated H2aM2760N, H2aM2761N, H1M2055N, H1H2304B, H1H2317B, H1H2322B, H1H2324B, H1H2331B, H1H2332B, H1H2333S, H1H2337B, H1H2338B, H1H2339B, H1H2340B, and H4H2055N. The H1M, H2M, H1H, etc. prefixes on the antibody designations used herein indicate the particular Fc region of the antibody. For example, an "H2M" antibody has a mouse IgG2 Fc, whereas an "H1H" antibody has a human IgG1 Fc. As will be appreciated by a person of ordinary skill in the art, an Fc region of an antibody can be modified or replaced with a different Fc region, but the variable domains (including the CDRs) will remain the same.

[0086] Hybridomas which produce the anti-Tie2 antibodies H1M2055N, and H2aM2760N were deposited under terms in accordance with the Budapest Treaty with the American Type Culture Collection (ATCC), 10801 University Blvd., Manassas, Va. 20110-2209 on December 2, 2011, under accession numbers PTA-12295 (H1M2055N) and PTA-12296 (H2aM2760N).

Example 2. Surface Plasmon Resonance Derived Binding Affinities and Kinetic Constants of Human Monoclonal Anti-Tie2 Antibodies

[0087] Binding affinities and kinetic constants of human monoclonal anti-Tie2 antibodies were determined by surface plasmon resonance at 25°C and 37°C (Tables 1-4). Measurements were conducted on a Biacore 2000 or T200 instrument.

[0088] For antibodies with a mouse constant region (designated H1M or H2M), antibodies were immobilized onto an anti-mouse Fc sensor surface and different concentrations of human, mouse or rat Tie2 constructs (hTie2-His, mTie2-hFc and rTie2-hFc) were injected over the antibody captured surface. For antibodies in the human IgG format (designated H1H or H4H), either an anti-human Fc sensor surface (hTie2-His and hTie2-mFc) or anti-human Fab sensor surface (mTie2-hFc and rTie2-hFc) was employed depending on the Tie-2 construct applied to the antibody captured surface. Amino acid sequences of the constructs used in this Example are as follows: hTie2-His (SEQ ID NO:2); hTie2-hFc (SEQ ID NO:3); hTie2-mFc (SEQ ID NO:4); rTie2-hFc (SEQ ID NO:5); and mTie2-hFc (SEQ ID NO:6).

[0089] Kinetic rate constants - association rate (ka) and dissociation rate (kd) were determined by fitting the real time binding sensorgrams to a 1:1 binding model with mass transport limitation using Scrubber 2.0 curve fitting software. The equilibrium dissociation constant (K_D) and dissociative half-life (T½) were calculated from the kinetic rate constants as: K_D (M) = kd / ka; and T½ (min) = (ln2/(60*kd).). As shown in Tables 1-2, several antibodies demonstrated high affinity binding to hTie2 at both temperatures tested. In addition H1M2055N, H1H2332B, H1H2337B, H1H2340B and H4H2055N exhibited significant binding to mouse and rat Tie2 (Tables 3-4).

Table 1: Biacore binding affinities of human mAbs to hTie2 at 25°C

Binding at 25° Mab capture format					
Antibody	Analyte	ka (Ms^{-1})	kd (s^{-1})	K_D (Molar)	T½ (min)
H2aM2760N	hTie2-His	2.91E+04	1.84E-04	6.32E-09	63
	hTie2-hFc	NT	NT	NT	NT
H2aM2761N	hTie2-His	3.33E+04	1.01E-04	3.02E-09	115
	hTie2-hFc	NT	NT	NT	NT
H1M2055N	hTie2-His	5.75E+05	1.34E-04	2.33E-10	86
	hTie2-hFc	NT	NT	NT	NT
H1H2304B	hTie2-His	1.99E+05	1.32E-03	6.66E-09	9
	hTie2-mFc	5.80E+05	3.75E-05	6.46E-11	308
H1H2317B	hTie2-His	1.43E+05	9.26E-04	6.50E-09	12
	hTie2-mFc	3.90E+05	2.36E-05	6.10E-11	489
H1H2322B	hTie2-His	1.67E+05	9.76E-04	5.84E-09	12
	hTie2-mFc	4.80E+05	1.39E-05	2.90E-11	829
H1H2324B	hTie2-His	2.40E+05	9.25E-04	3.86E-09	12
	hTie2-mFc	6.30E+05	5.92E-05	9.39E-11	195
H1H2331B	hTie2-His	3.63E+04	1.17E-03	3.22E-08	10
	hTie2-mFc	8.20E+04	1.24E-04	1.52E-09	93
H1H2332B	hTie2-His	8.89E+04	2.23E-03	2.51E-08	5
	hTie2-mFc	1.25E+05	2.13E-04	1.1E-09	54
H1H2333S	hTie2-His	5.92E+04	3.09E-04	5.21E-09	37

	hTie2-mFc	NB	NB	NB	NB
H1H2337B	hTie2-His	6.47E+04	6.49E-04	1.00E-08	18
	hTie2-mFc	1.20E+05	1.11E-04	9.30E-10	104
H1H2338B	hTie2-His	4.93E+04	3.44E-04	6.99E-09	34
	hTie2-mFc	NB	NB	NB	NB
H1H2339B	hTie2-His	5.58E+04	1.25E-03	2.24E-08	9
	hTie2-mFc	1.51E+05	1.02E-04	6.70E-10	114
H1H2340B	hTie2-His	7.01E+04	3.45E-03	4.92E-08	3
	hTie2-mFc	1.65E+05	1.91E-04	1.16E-09	60
H4H2055N	hTie2-His	3.82E+05	4.84E-04	1.27E-09	24
	hTie2-mFc	1.27E+06	9.92E-05	7.80E-11	116
Control I	hTie2-His	6.53E+04	4.57E-04	7.00E-09	25
	hTie2-mFc	1.45E+05	1.16E-04	8.03E-10	99

NB = no binding under the conditions tested
NT = not tested

Table 2: Biacore binding affinities of human mAbs to hTie2 at 37°C

Binding at 37° Mab capture format					
Antibody	Analyte	ka (Ms^{-1})	kd (s^{-1})	K_D (Molar)	T½ (min)
H2aM2760N	hTie2-His	4.04E+04	3.06E-04	7.60E-09	38
	hTie2-hFc	NT	NT	NT	NT
H2aM2761N	hTie2-His	4.26E+04	2.87E-04	6.74E-09	40
	hTie2-hFc	NT	NT	NT	NT
H1M2055N	hTie2-His	1.08E+06	3.36E-04	3.10E-10	34
	hTie2-hFc	NT	NT	NT	NT
H1H2304B	hTie2-His	3.50E+05	3.67E-03	1.05E-08	3
	hTie2-mFc	6.60E+05	1.30E-04	1.96E-10	89
H1H2317B	hTie2-His	1.74E+05	3.46E-03	1.98E-08	3
	hTie2-mFc	4.60E+05	9.94E-05	2.15E-10	116
H1H2322B	hTie2-His	2.12E+05	4.74E-03	2.23E-08	2
	hTie2-mFc	5.70E+05	9.72E-05	1.72E-10	119
H1H2324B	hTie2-His	2.78E+05	2.26E-03	8.12E-09	5
	hTie2-mFc	7.40E+05	1.47E-04	1.99E-10	79
H1H2331B	hTie2-His	5.48E+04	4.74E-03	8.65E-08	2
	hTie2-mFc	1.28E+05	1.99E-04	1.56E-09	58
H1H2332B	hTie2-His	9.58E+04	8.19E-03	8.55E-08	1
	hTie2-mFc	1.81E+05	6.52E-04	3.60E-09	18
H1H2333S	hTie2-His	6.20E+04	9.83E-04	1.58E-08	12
	hTie2-mFc	NB	NB	NB	NB
H1H2337B	hTie2-His	8.35E+04	2.41E-03	2.89E-08	5
	hTie2-mFc	1.92E+05	2.55E-04	1.33E-09	45
H1H2338B	hTie2-His	5.55E+04	9.93E-04	1.79E-08	12
	hTie2-mFc	NB	NB	NB	NB
H1H2339B	hTie2-His	6.78E+04	4.88E-03	7.20E-08	2

	hTie2-mFc	1.93E+05	2.95E-04	1.53E-09	39
H1H2340B	hTie2-His	9.75E+04	9.98E-03	1.02E-07	1
	hTie2-mFc	2.00E+05	5.15E-04	2.58E-09	22
H4H2055N	hTie2-His	5.49E+05	8.01E-04	1.46E-09	14
	hTie2-mFc	1.67E+06	1.55E-04	9.30E-11	74
Control I	hTie2-His	7.48E-04	1.12E-03	1.49E-08	10
	hTie2-mFc	1.80E+05	1.70E-04	9.40E-10	68

NB = no binding under the conditions tested
NT = not tested

Table 3: Biacore binding affinities of human mAbs to mTie2 at 25°C

Binding at 25° Mab capture format					
Antibody	Analyte	ka (Ms^{-1})	kd (s^{-1})	K_D (Molar)	T½ (min)
H2aM2760N	mTie2-hFc	NB	NB	NB	NB
H2aM2761N	mTie2-hFc	NB	NB	NB	NB
H1M2055N	mTie2-hFc	1.07E+06	1.85E-05	1.73E-11	625
H1H2304B	mTie2-hFc	NB	NB	NB	NB
H1H2317B	mTie2-hFc	NB	NB	NB	NB
H1H2322B	mTie2-hFc	NB	NB	NB	NB
H1H2324B	mTie2-hFc	NB	NB	NB	NB
H1H2331B	mTie2-hFc	4.81E+04	2.72E-03	5.65E-08	4
H1H2332B	mTie2-hFc	5.39E+04	5.76E-03	1.07E-07	2
H1H2333S	mTie2-hFc	NB	NB	NB	NB
H1H2337B	mTie2-hFc	1.87E+05	1.84E-02	9.83E-08	1
H1H2338B	mTie2-hFc	NB	NB	NB	NB
H1H2339B	mTie2-hFc	4.68E+04	2.63E-03	5.63E-08	4
H1H2340B	mTie2-hFc	1.36E_05	6.72E-03	4.96E-08	2
H4H2055N	mTie2-hFc	1.24E+06	6.19E-06	5.01E-12	1867
Control I	mTie2-hFc	NB	NB	NB	NB

NB = no binding under the conditions tested

Table 4: Biacore binding affinities of human mAbs to rTie2 at 25°C

Binding at 25°C Mab capture format					
Antibody	Analyte	ka (Ms^{-1})	kd (s^{-1})	K_D (Molar)	T½
H2aM2760N	rTie2-hFc	NB	NB	NB	NB
H2aM2761N	rTie2-hFc	NB	NB	NB	NB
H1M2055N	rTie2-hFc	9.43E+05	2.55E-05	2.70E-11	454
H1H2304B	rTie2-hFc	NB	NB	NB	NB
H1H2317B	rTie2-hFc	NB	NB	NB	NB
H1H2322B	rTie2-hFc	NB	NB	NB	NB
H1H2324B	rTie2-hFc	NB	NB	NB	NB
H1H2331B	rTie2-hFc	NB	NB	NB	NB
H1H2332B	rTie2-hFc	1.12E+05	6.18E-03	5.53E-08	2

H1H2333S	rTie2-hFc	NB	NB	NB	NB
H1H2337B	rTie2-hFc	1.45E+05	8.46E-03	5.85E-08	1
H1H2338B	rTie2-hFc	NB	NB	NB	NB
H1H2339B	rTie2-hFc	NB	NB	NB	NB
H1H2340B	rTie2-hFc	1.43E+05	3.71E-03	2.59E-08	3
H4H2055N	rTie2-hFc	1.07E+06	1.00E-06	9.31E-13	11550
Control I	rTie2-hFc	NB	NB	NB	NB

NB = no binding under the conditions tested

Example 3. Epitope Mapping of Tie2 mAbs using Luminex Beads

[0090] To determine domain binding for the anti-Tie2 antibodies, several hTie2 receptor extracellular domain-deletion constructs were covalently linked to luminex xMAP beads (10µg/ml of each protein per 10^7 beads). The constructs are depicted in Figure 1 and are designated as follows: hTie2 (Ig1-Ig2-EGF) (SEQ ID NO:7); hTie2 (Ig2-EGF) (SEQ ID NO:8); hTie2 (EGF) (SEQ ID NO:9); hTie2 (Ig3-FN) (SEQ ID NO:10); and hTie2 (FN) (SEQ ID NO:11). Also tested in this Example were full-length hTie2-mFc (SEQ ID NO:4) and hTie1-hFc (SEQ ID NO:12) ectodomain constructs.

[0091] For binding, 25 µl of anti-Tie2 antibody (25 µg/ml) was added to 75 µl of the above created Luminex bead mixture (3x10^3 beads per construct) in binding buffer (PBS, 0.05% Tween 20, 1 mg/ml BSA, 0.05% sodium azide) into a 96 well filter plate (Millipore). Incubation was at room temperature (RT) for 90 min or overnight at 4°C with shaking. The beads were washed 3x with washing buffer (PBS + 0.05% Tween 20) resuspended in 100 µl binding buffer containing PE (phycoerythrine)-labeled anti human kappa or PE-labeled anti mouse Fab secondary antibody and incubated at RT for 45 min with shaking. Samples were washed 2X more and binding signal (MFI) for each bead was determined using a Luminex L200 or FLEXMAP3D instrument. Bead linked human Tie2 was used as positive control and bead linked human Tie1 was used to measure family cross-reactivity. In general MFI signals greater than 500 units represent significant binding. Results are summarized in Table 5.

Table 5

Antibody	hTie2-mFc	hTie1-hFc	hTie2 (EGF)	hTie2 (Ig1,Ig2,EGF)	hTie2 (Ig2,EGF)	hTie2 (FN)-hFc	hTie2 (Ig3,FN)-hFc	Predicted Binding Domain
H2aM2760N	1668	15	4	9392	6245	341	330	Ig1, Ig2
H2aM2761N	8503	135	10205	11657	8568	3403	14509	EGF,Ig3
H1M2055N	753	112	90	2022	873	43	40	Ig1, Ig2
H1H2304B	4250	40	8901	8776	7648	5	6	EGF repeat
H1H2317B	4853	37	9611	8966	8325	9	11	EGF repeat
H1H2322B	4040	47	9901	9161	9167	10	13	EGF repeat
H1H2324B	4506	31	9773	9438	8686	7	10	EGF repeat
H1H2331B	5766	118	8	6	10	8097	6568	FN repeat
H1H2332B	4783	32	3	3	4	6711	6044	FN repeat
H1H2333S	1875	10	3	2	2	6	6	ND

H1H2337B	5208	48	9	7	9	6425	7368	FN repeat
H1H2338B	1385	9	4	5	5	6	8	ND
H1H2339B	2913	62	6	5	6	4529	3838	FN repeat
H1H2340B	3009	22	4	4	6	5149	4599	FN repeat
Control I	4901	7	3	4	5	6105	4554	FN repeat

ND = not determined

[0092] From the above results, it can be concluded that H2aM2760N and H1M2055N bind to the Ig1/Ig2 domains of Tie2; H2aM2761N binds to the EGF and Ig3 domains of Tie2; H1H2304B, H1H2317B, H1H2322B and H1H2324B bind to the EGF domain of Tie2; and H1H2331B, H1H2332B, H1H2337B, H1H2339B, H1H2340B, and Control I bind to the FN domain of Tie2.

Example 4. Epitope Mapping of H4H2055N Binding to Tie2 by H/D Exchange

[0093] Experiments were conducted to more precisely define the amino acid residues of Tie2 with which H4H2055N interacts. (H4H2055N is a fully human IgG4 version of the antibody H1M2055N produced from hybridoma PTA-12295.) For this purpose H/D exchange epitope mapping was carried out. A general description of the H/D exchange method is set forth in *e.g.*, Ehring (1999) *Analytical Biochemistry* 267(2):252-259; and Engen and Smith (2001) *Anal. Chem. 73*:256A-265A.

[0094] To map the binding epitope(s) of antibody H4H2055N on Tie2 via H/D exchange, a recombinant construct consisting of the Ig1, Ig2 and EGF extracellular domains of human Tie2 (hTie2 (Ig1-Ig2-EGF); SEQ ID NO:7) was used. Antibody H4H2055N was covalently attached to N-hydroxysuccinimide (NHS) agarose beads (GE Lifescience).

[0095] In the 'on-solution/off-beads' experiment (on-exchange in solution followed by off-exchange on beads), the ligand (hTie2 Ig1-Ig2-EGF) was deuterated for 5 min or 10 min in PBS buffer prepared with D_2O, and then bound to H4H2055N beads through a 2 min incubation. The Tie2-bound beads were washed with PBS aqueous buffer (prepared with H_2O) and incubated for half of the on-exchange time in PBS buffer. After the off-exchange, the bound Tie2 was eluted from beads with an ice-cold low pH TFA solution. The eluted Tie2 was then digested with immobilized pepsin (Thermo Scientific) for 5 min. The resulting peptides were desalted using ZipTip® chromatographic pipette tips and immediately analyzed by UltrafleXtreme matrix assisted laser desorption ionization time of flight (MALDI-TOF) mass spectrometry (MS).

[0096] In the 'on-beads/off-beads' experiment (on-exchange on beads followed by off-exchange on beads), Tie2 was first bound to H4H2055N beads and then incubated for 5 min or 10 min in D_2O for on-exchange. The following steps (off-exchange, pepsin digestion, and MS analysis) were carried out as described for the 'on-solution/off-beads' procedure. The centroid values or average mass-to-charge ratios (m/z) of all the detected peptides were calculated and compared between these two sets of experiments.

[0097] The results are summarized in Table 6 which provides a comparison of the centroid m/z values for all the detected peptides identified by liquid chromatography-matrix assisted laser desorption ionization (LC-MALDI) MS following the H/D exchange and peptic digest procedure. Likely due to disulfide bonds, the Tie2 sequence coverage detected from a single MALDI-TOF spectrum is relatively low. Nevertheless, more than half of the detected peptic peptides gave similar centroid values for both the on-solution/off-beads and on-beads/off-beads protocols. Three segments with corresponding residues 88-106, 139-152, and 166-175 had delta centroid values ≥ 0.20 m/z in both experiments. For purposes of the present Example a positive difference (Δ) of at least 0.20 m/z in both experiments indicates amino acids protected by antibody binding. Segments meeting this criterion are indicated by bold text and an asterisk (*) in Table 6.

Table 6 : H4H2055N Binding to Tie2

	Experiment I 5 min on- / 2.5 min off-exchange			Experiment II 10 min on- / 5 min off-exchange		
Residues (of SEQ ID NO:7)	On-solution / Off Beads	On-Beads / Off-Beads	Δ	On-solution / Off Beads	On-Beads / Off-Beads	Δ
20-33	1568.70	1568.63	0.07	1568.63	1568.63	0.00
26-33	1036.17	1036.16	0.01	1036.27	1036.21	0.07
38-50	1572.91	1572.93	-0.02	1572.78	1572.98	-0.20
39-50	1425.38	1425.39	0.00	1425.54	1425.45	0.08
42-53	1440.52	1440.44	0.09	1440.62	1440.52	0.10
42-58	2041.43	2041.42	0.01	2041.58	2041.41	0.17
78-87	1124.17	1124.15	0.03	1124.20	1124.14	0.07
88-95	1049.49	1049.46	0.02	1049.46	1049.54	-0.08
88-101*	1767.78	1767.51	**0.27**	1767.79	1767.57	**0.22**
88-102*	1881.08	1880.66	**0.42**	1881.14	1880.69	**0.45**
93-101*	1127.80	1127.57	**0.24**	1127.80	1127.61	**0.19**
93-102*	1241.46	1240.83	**0.63**	1241.34	1240.83	**0.51**
96-102*	850.54	850.22	**0.32**	850.55	850.25	**0.30**
96-106*	1233.29	1232.66	**0.64**	1233.43	1232.65	**0.78**
139-151*	1514.22	1513.16	**1.06**	1514.49	1513.21	**1.28**
139-152*	1643.17	1642.02	**1.16**	1643.44	1642.06	**1.38**
152-165	1498.87	1498.73	0.14	1498.88	1498.77	0.11
152-166	1662.09	1662.01	0.07	1662.10	1662.08	0.02
153-165	1369.70	1369.64	0.06	1369.77	1369.66	0.11
166-175*	1115.07	1114.59	**0.47**	1115.04	1114.65	**0.39**
167-175*	951.73	951.30	**0.42**	951.74	951.33	**0.41**

[0098] Since the peptide fragment (m/z of 1049.4) corresponding to residues 88-95 did not show deuteron retention (Δ = 0) after off-exchange, the first protected segment (88-106) can be reduced to only include residues 96-106. Thus the three regions 96-106, 139-152, and 166-175 of SEQ ID NO:7 are protected from full off-exchange by H4H2055N binding to Tie2 after on-exchange.

[0099] The Ig1 domain of Tie2 consists of amino acids 1-97 of SEQ ID NO:7; the Ig2 domain of Tie 2 consists of amino acids 98-186 of SEQ ID NO:7; and the EGF repeat domain consists of amino acids 187-321 of SEQ ID NO:7. Thus the first binding region (amino acids 96-106 of SEQ ID NO:7) includes the last two amino acids of the Ig1 domain and the first 9 amino acids of the Ig2 domain. The second and third binding regions (amino acids 139-152 and 166-175 of SEQ ID NO:7) are located entirely within the Ig2 domain.

[0100] It should also be noted that the second and third binding regions (amino acids 139-152 and 166-175 of SEQ ID NO:7) lie within the amino acid region of Tie 2 that directly contacts its cognate ligand, Angiopoietin-2, based on the co-crystal structure by Barton et al. (2006, Nat Struct Mol Biol 13(6) 524-532). Therefore, this example suggests that antibody H4H2055N binds a discontinuous epitope primarily within the Ig2 domain of human Tie2 (with one or two potential amino acid contacts within the C-terminal portion of the Ig1 domain), and that binding to these regions correlates with the ability of H4H2055N to block the interaction between Tie2 and Ang2 as illustrated in Examples 5 and 6 herein.

Example 5. Assessment of the Ability of Anti-Tie2 Antibodies to Block the Interaction Between Tie2 and Angiopoietins

[0101] Tie2 is known to interact with angiopoietins (Ang1, Ang2, Ang3 and Ang4). An initial set of experiments was therefore conducted to evaluate the ability of anti-Tie2 antibodies to block Tie2 binding to Ang2. In this first set of experiments, a quantitative blocking immunoassay was utilized. Briefly, solutions of 0.8 nM biotinylated ecto-Tie2 fused to 6xHis (SEQ ID NO:2) were premixed with anti-Tie2 antibody ranging from ~ 50 nM to 0 nM in serial dilutions. After a 1-hour incubation at room temperature, the amount of Tie2-His bound to plate coated Bow-Ang2 was measured by sandwich ELISA. Bow-Ang2 (SEQ ID NO:13), an Ang2 construct that comprises the Fc domain of hIgG1 flanked by one FD domain of Ang2 at both termini, was coated at 2 µg/ml on a 96 well microtiter plate and blocked with BSA. Plate-bound biotin-Tie2-His was detected using HRP conjugated streptavidin, and developed using BD OptEIA™ (BD Biosciences Pharmingen, San Diego, CA). Signals of OD_{450} nm were recorded and data was analyzed using GraphPad Prism to calculate IC_{50} values. Results are summarized in Table 7. IC_{50}s were defined as the amount of antibody required to reduce 50% of biotin-Tie2-His detectable to plate bound Bow-Ang2 ligand. In Table 7, an antibody is designated an "enhancer" if the addition of the antibody increases the amount of Tie2-His bound to the Bow-Ang2-coated surface relative to the no antibody control.

Table 7: IC_{50} values for anti-Tie2 Mabs blocking of hTie2 to plate bound Bow-Ang2

Antibody	Result in Blocking ELISA	Blocking of binding to Bow-Ang2 IC_{50} (M)
H2aM2760N	Enhancer	N/A
H2aM2761N	Enhancer	N/A
H1M2055N	Enhancer	N/A
H1H2304B	Enhancer	N/A
H1H2317B	Enhancer	N/A

H1H2322B	Enhancer	N/A
H1H2324B	Enhancer	N/A
H1H2331B	Enhancer	N/A
H1H2332B	Enhancer	N/A
H1H2333S	Enhancer	N/A
H1H2337B	Enhancer	N/A
H1H2338B	Enhancer	N/A
H1H2339B	Blocker	1.89E-10
H1H2340B	Blocker	1.03E-09
H4H2055N	Blocker	1.06E-09
Control I	Enhancer	N/A

[0102] This first set of experiments demonstrates that anti-Tie2 antibodies H1H2339B, H1H2340B and H4H2055N are able to block the interaction between Ang2 and Tie2 in an ELISA format.

[0103] A further set of experiments was conducted to evaluate the ability of H4H2055N to block Tie2 binding to Ang2 and other members of the angiopoietin family (Ang1, Ang3 and Ang4). In this set of experiments, an Octet Red biosensor was employed using two experimental formats. In the first format, hTie2.mFc (SEQ ID NO:4; 10 µg/ml) or a negative control was captured on anti-mFc Octet senor tips for 5min. The captured sensor tip surfaces were then saturated with H4H2055N or Control I antibody by dipping into wells containing 300 nM of respective monoclonal antibodies for 10 min. Finally the sensor tips were placed into wells containing 100nM of hAng1 (R&D Systems, Inc., Minneapolis, MN, Cat 923-AN/CF; Accession #Q5HYA0), hAng2 (R&D Systems, Inc., Minneapolis, MN, Cat 623-AN/CF; Accession #O15123), mAng3 (R&D Systems, Inc., Minneapolis, MN, Cat 738-AN/CF; Accession #Q9WVH6) or hAng4 (R&D Systems, Inc., Minneapolis, MN, Cat 964-AN/CF; Accession Q9Y264) for 5 min. Binding response at each step of the experiment was monitored and the binding of different angiopoietins to the monoclonal antibody saturated receptor surfaces was plotted (Figure 2). As illustrated by Figure 2, H4H2055N blocked the binding of all four angiopoietins to the hTie2.mFc surface. The Control I antibody did not block binding of any of the angiopoietins to the Tie2-coated surfaces.

[0104] In the second assay format, hTie2.mFc (10 µg/ml) or negative control was captured on anti-mFc coated Octet sensor tips (5 min), followed by placing the tips into different wells containing 100 nM of angiopoietin (hAng1, hAng2, mAng3 or hAng4) for 10 min. Lastly, the senor tips were placed into wells containing 300nM of H4H2055N or Control I (5min). As in format 1, the binding response at each step was monitored and the binding response was plotted (Figure 3). This format demonstrated that, unlike Control I, the binding of H4H2055N is diminished by the already bound angiopoietins on the Tie2-coated surfaces. Thus, H4H2055N competes with the angiopoietins (Ang1, Ang2, Ang3 and Ang4) for binding to Tie2.

Example 6. Assessment of the Ability of Anti-Tie2 Antibodies to Block Angiopoietin-Mediated Tie2 Signaling

[0105] To further characterize anti-hTie2 mAbs of the invention, the ability of the antibodies to block Tie2-induced luciferase activity was explored. Briefly, HEK293 cells were stably transfected with hTie2, and the top 10% of Tie2 expressing cells were isolated via FACS. These cells were then transduced with a serum response element (SRE)-dependent luciferase reporter lentivirus (SA Biosciences), and a clonal population (293-TIE2/SRE-Luc) with robust response to tetrameric forms of angiopoietin-1 (Bow-Ang1, SEQ ID NO:14) and angiopoietin-2 (Bow-Ang2, SEQ ID NO:13) was isolated.

[0106] For assays, 293-Tie2/SRE-Luc cells were seeded (30,000 cells per well) in 96-well plates one day prior to treatment. Dose-response curves were generated with serial dilutions of Bow-Ang1 (BA1) or Bow-Ang2 (BA2) incubated with cells for 4h before measurement of luciferase activity. For Tie2 inhibition, cells were treated with serially diluted Tie2 mAbs in the presence of a 1 nM constant dose of BA1 or BA2 for 4 h. Luciferase activity was measured on a Victor luminometer following incubation with One-Glo™ (Promega) reagent. Three classes of antibodies were observed: <u>Blockers</u>, defined as a reduction in Tie2 dependent luciferase activity upon addition of antibody; <u>Activators</u>, defined as an increase in Tie2 dependent luciferase activity upon addition of antibody; and <u>Not Active</u>, defined as no significant change in signal upon their addition. For "Not Active" antibodies, no EC_{50} or IC_{50} value was calculated. Results are summarized in Table 8.

Table 8: IC_{50} values for Anti-Tie2 antibodies

Antibody	Mab Class (BowAng1)	EC_{50} or IC_{50} ± SEM (10^{-9} M) BowAng1	Mab Class (BowAng2)	EC_{50} or IC_{50} ± SEM (10^{-9} M) BowAng2
H2aM2760N	Blocker	11.6 ± .81	Blocker	2.8 ± .14
H2aM2761N	Blocker, not to baseline	5.2 ± 1.3	Blocker, not to baseline	1.1 ± .11
H4H2055N	Blocker	0.69 ± .16	Blocker	0.23 ± .03
H1H2304B	No activity	NA	No activity	NA
H1H2317B	No activity	NA	No activity	NA
H1H2322B	No activity	NA	No activity	NA
H1H2324B	No activity	NA	No activity	NA
H1H2331B	Activator	13.6 ± 3.1	Activator	5.1 ± 1.9
H1H2332B	Activator	2.5 ± 1.2	Activator	3.7 ± .15
H1H2333B	No activity	NA	No activity	NA
H1H2337B	Activator	1.1 ± .02	Activator	4.7 ± .80
H1H2338B	No activity	NA	No activity	NA
H1H2339B	Activator	10.3 ± .23	Activator	13.0 ± 3.5
HIH2340B	Activator	1.41 ± .01	Activator	2.1 ± .04
Control I	Activator	0.76 ± .50	Activator	1.4 ± .03

IC_{50} values are reported for Blockers, and EC_{50} values are reported for Activators

[0107] Anti-Tie2 antibodies H2aM2760N, H2aM2761N and H4H2055N were identified as blockers in this experimental system.

Example 7. Assessment of the Anti-Tumor Activity of an Anti-Tie2 Antibody *in vivo*

[0108] An exemplary anti-Tie2 antibody (H2M2055N) was tested for its ability to inhibit the growth of human tumor xenografts in immunocompromised mice. Briefly, either human colorectal HT29 or Colo205 tumors were grown in SCID mice. When tumors were palpable (100 mm^3 for HT29 and 400mm^3 for Colo205) animals were treated biweekly with Fc protein (10 mg/kg) or anti-Tie2 antibody (H1M2055N; 10 mg/kg). At the end of treatment (31 days for HT29 & 14 days for Colo205), the percent tumor growth inhibition (%TGI) was determined (Table 9) and tumor tissue was harvested and utilized for vessel density analysis. Vascular density was assessed in 30µm thick OCT tumor sections by CD31 immunohistochemistry. NIH Image software was utilized to determine the % area vessel density in the tumor tissue sections. Results are illustrated graphically in Figures 4-7.

Table 9: Percent Decrease in Tumor Growth with Anti-Tie2 Mab Treatment

	HT29		Colo205	
Antibody	Avg Tumor Growth (mm3) from start of treatment	% Decrease in Tumor Growth	Avg Tumor Growth (mm3) from start of treatment	% Decrease in Tumor Growth
Fc protein (10 mg/kg)	100		400	
H2M2055N (10 mg/kg)	100	27.5	400	77

[0109] As shown in this Example, the anti-Tie2 Ab, H1M2055N, not only decreased tumor growth (Table 9), but also significantly decreased vessel density in both HT29 (25%, Figure 5) and Colo205 (40%, Figure 7) xenograft models.

[0110] The present invention is not to be limited in scope by the specific embodiments describe herein. Indeed, various modifications of the invention in addition to those described herein will become apparent to those skilled in the art from the foregoing description and the accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

What is claimed is:

1. An isolated antibody or antigen-binding fragment thereof that specifically binds human Tie2 and blocks the interaction between Tie2 and a Tie2 ligand.

2. The isolated antibody or antigen-binding fragment of claim 1, wherein the Tie2 ligand is selected from the group consisting of Ang1, Ang2, Ang3 and Ang4.

3. The isolated antibody or antigen-binding fragment of claim 1, wherein antibody or antigen-binding fragment thereof blocks the interaction between Tie2 and all four of Ang1, Ang2, Ang3 and Ang4.

4. The isolated antibody or antigen-binding fragment of any one of claims 1 to 3, wherein the antibody or antigen-binding fragment thereof specifically binds a polypeptide consisting of Ig1-Ig2-EGF domains of human Tie2 (SEQ ID NO:7), or a polypeptide consisting of Ig2-EGF domains of human Tie2 (SEQ ID NO:8).

5. The antibody or antigen-binding fragment of any one of claims 1 to 4, wherein the antibody or antigen-binding fragment interacts with one or more amino acids located within one or more amino acid segments selected from the group consisting of amino acids 96-106 of SEQ ID NO:7, amino acids 139-152 of SEQ ID NO:7; and amino acids 166-175 of SEQ ID NO:7.

6. The antibody or antigen-binding fragment of claim 5, wherein the antibody or antigen-binding fragment interacts with amino acids 96-106 of SEQ ID NO:7, amino acids 139-152 of SEQ ID NO:7; and amino acids 166-175 of SEQ ID NO:7, as determined by hydrogen/deuterium exchange.

7. The isolated antibody or antigen-binding fragment of any one of claims 1 to 6, wherein the antibody or antigen-binding fragment thereof specifically binds a rodent Tie2 and human Tie2.

8. The isolated antibody or antigen-binding fragment of claim 7, wherein the rodent Tie2 is mouse Tie2 or rat Tie2.

9. The isolated antibody or antigen-binding fragment of any one of claims 1 to 6 wherein the antibody or antigen-binding fragment thereof specifically binds human, mouse and rat Tie2.

10. The isolated antibody or antigen-binding fragment of any one of claims 1 to 9, wherein the antibody or antigen-binding fragment thereof is produced from a cell line deposited with the American Type Culture Collection under accession number PTA-12295 or PTA-12296.

11. An isolated antibody or antigen-binding fragment thereof that binds the same epitope

on human Tie2 as the antibody produced from a cell line deposited with the American Type Culture Collection under accession number PTA-12295 or PTA-12296.

12. An isolated antibody or antigen-binding fragment thereof that competes for binding to human Tie2 with an antibody produced from a cell line deposited with the American Type Culture Collection under accession number PTA-12295 or PTA-12296.

13. A pharmaceutical composition comprising the antibody or antigen-binding fragment of any one of claims 1 to 12 and a pharmaceutically acceptable carrier or diluent.

14. A pharmaceutical composition for use in inhibiting the growth of a tumor in a patient, the pharmaceutical composition comprising an antibody or antigen-binding fragment thereof that specifically binds human Tie2 and blocks the interaction between Tie2 and a Tie2 ligand.

15. The pharmaceutical composition of claim 14, wherein the Tie2 ligand is selected from the group consisting of Ang1, Ang2, Ang3 and Ang4.

16. The pharmaceutical composition of claim 14, wherein antibody or antigen-binding fragment thereof blocks the interaction between Tie2 and all four of Ang1, Ang2, Ang3 and Ang4.

17. The pharmaceutical composition of claim 14, wherein the antibody or antigen-binding fragment thereof is produced from a cell line deposited with the American Type Culture Collection under accession number PTA-12295 or PTA-12296.

18. The pharmaceutical composition of claim 14, wherein the antibody or antigen-binding fragment thereof binds the same epitope on human Tie2 as the antibody produced from a cell line deposited with the American Type Culture Collection under accession number PTA-12295 or PTA-12296.

19. The pharmaceutical composition of claim 14, wherein the antibody or antigen-binding fragment thereof competes for binding to human Tie2 with an antibody produced from a cell line deposited with the American Type Culture Collection under accession number PTA-12295 or PTA-12296.

20. A method for inhibiting the growth of a tumor in a patient, the method comprising administering to the patient a pharmaceutical composition comprising an antibody or antigen-binding fragment thereof that specifically binds human Tie2 and blocks the interaction between Tie2 and a Tie2 ligand.

hTie2 Deletion Constructs

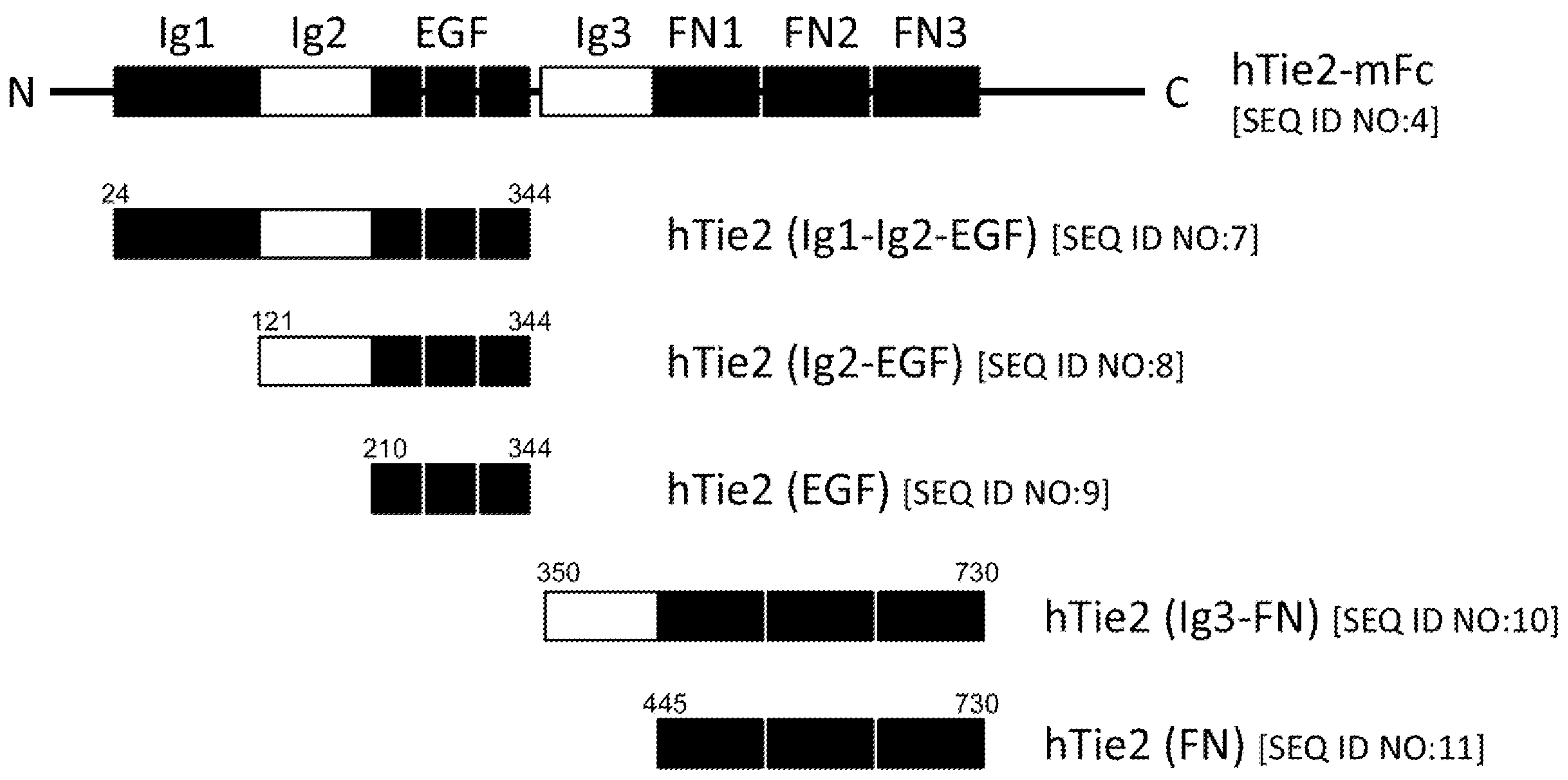


Figure 1

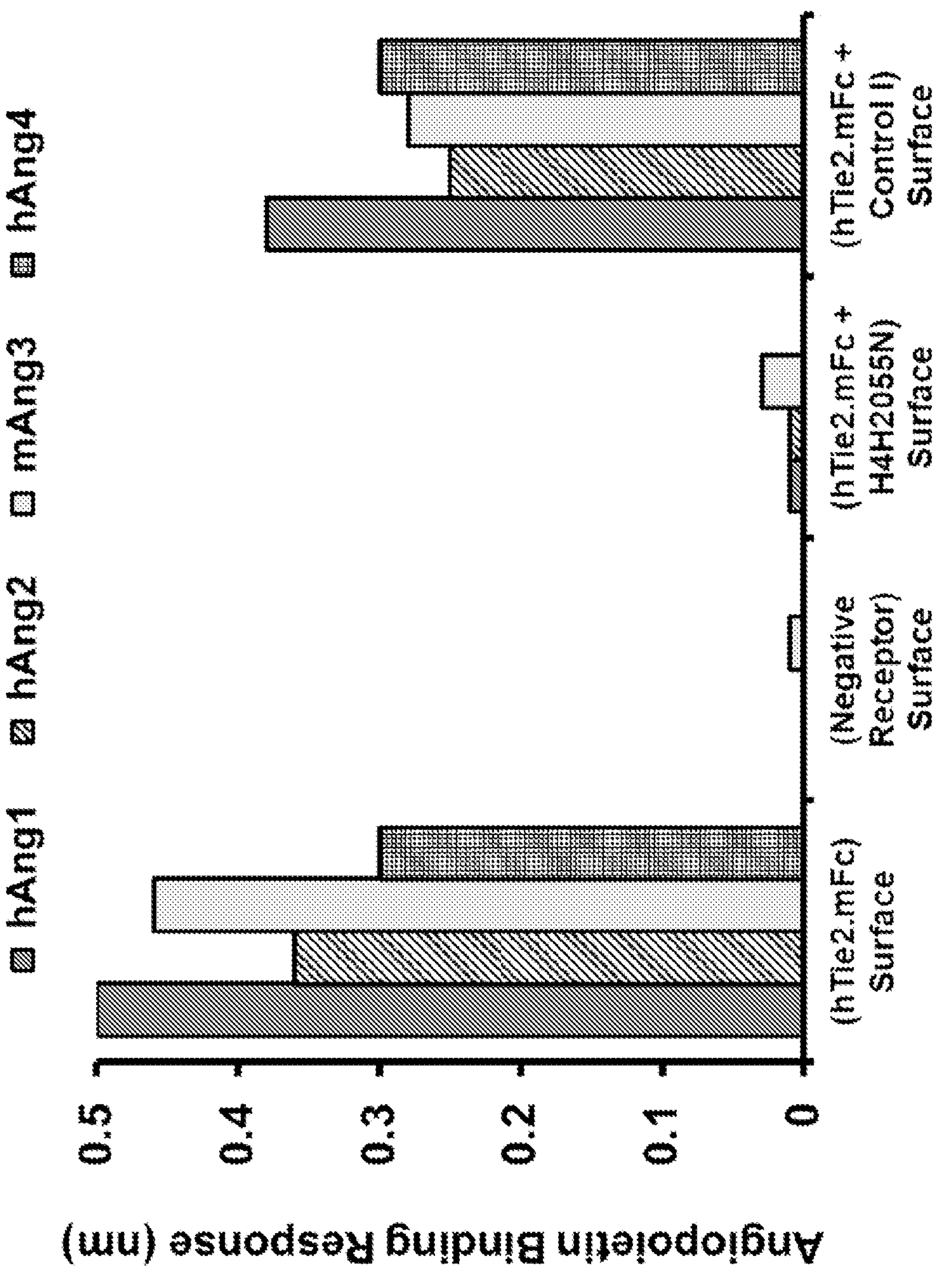

▨ hAng1
▨ hAng2
☐ mAng3
▦ hAng4
Angiopoietin Binding Response (nm)
0.5
0.4
0.3
0.2
0.1
0
(hTie2.mFc) Surface
(Negative Receptor) Surface
(hTie2.mFc + H4H2055N) Surface
(hTie2.mFc + Control I) Surface

Figure 2

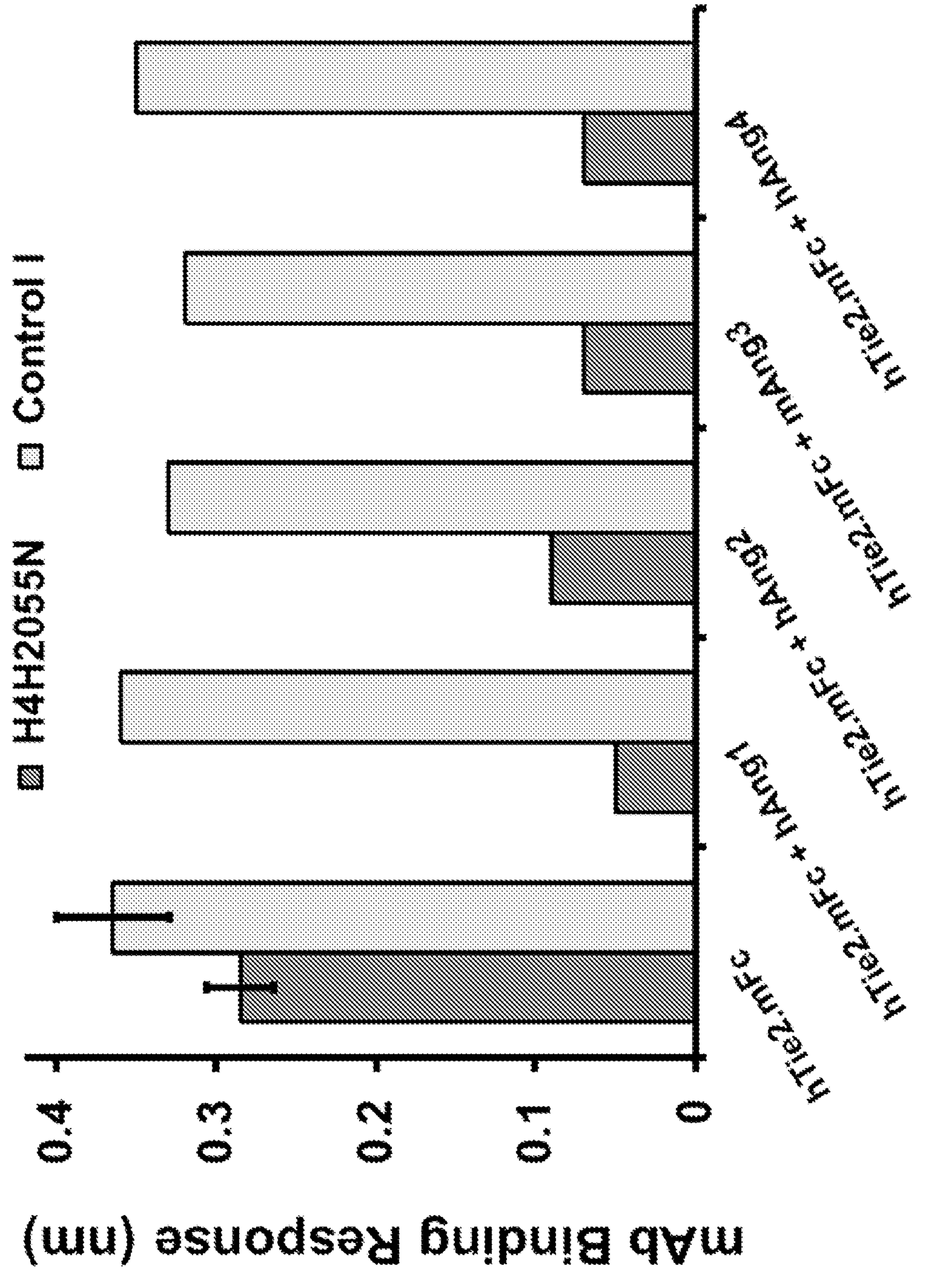

▨ H4H2055N
☐ Control I
mAb Binding Response (nm)
0.4
0.3
0.2
0.1
0
hTie2.mFc
hTie2.mFc + hAng1
hTie2.mFc + hAng2
hTie2.mFc + mAng3
hTie2.mFc + hAng4

Figure 3

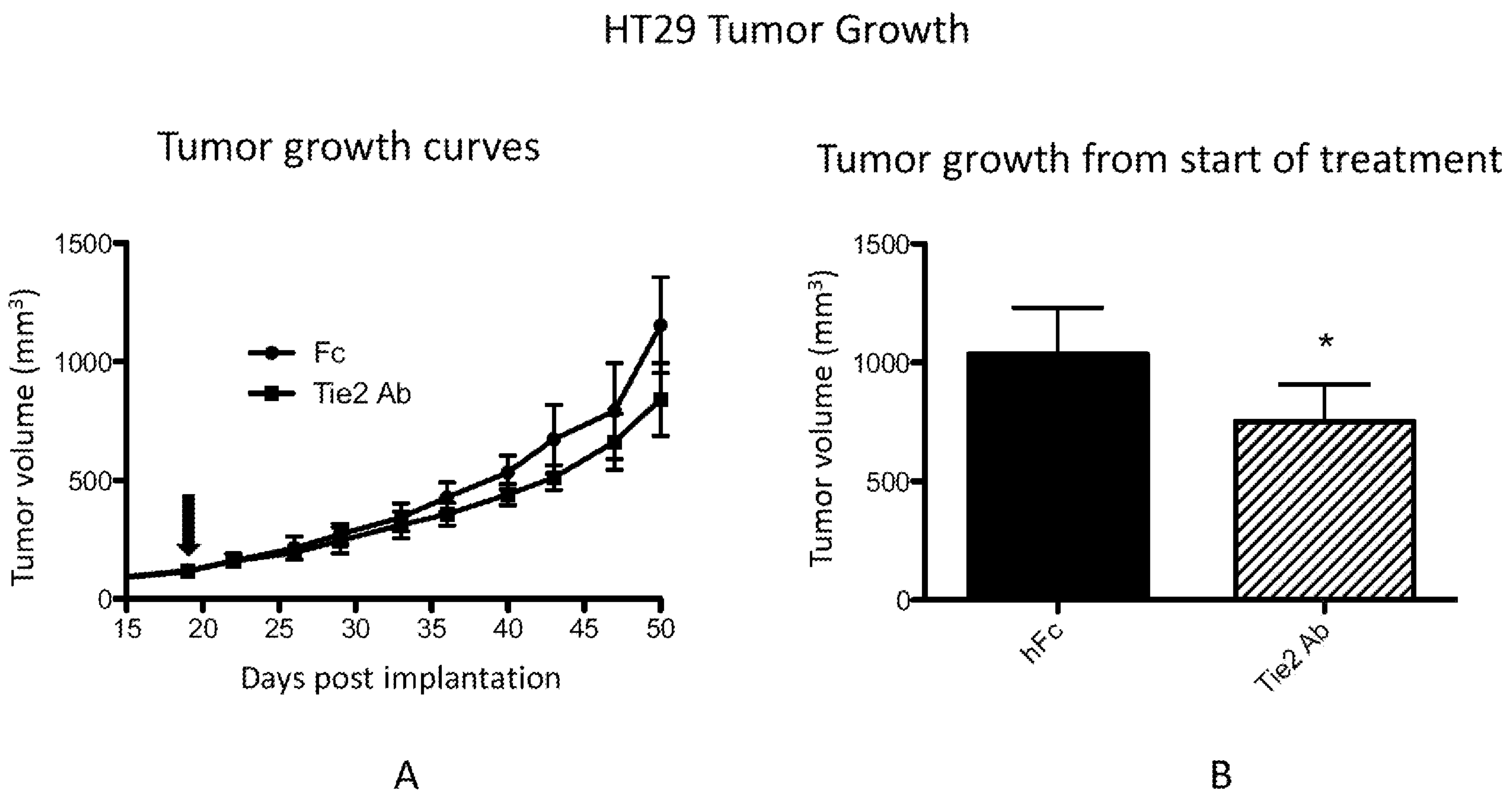

HT29 Tumor Growth
Tumor growth curves
Tumor volume (mm³)
1500
1000
500
0
Fc
Tie2 Ab
15
20
25
30
35
40
45
50
Days post implantation
A
Tumor growth from start of treatment
Tumor volume (mm³)
1500
1000
500
0
*
hFc
Tie2 Ab
B

Figure 4

HT29 Vessel Density

Tumor vessel density at end of experiment

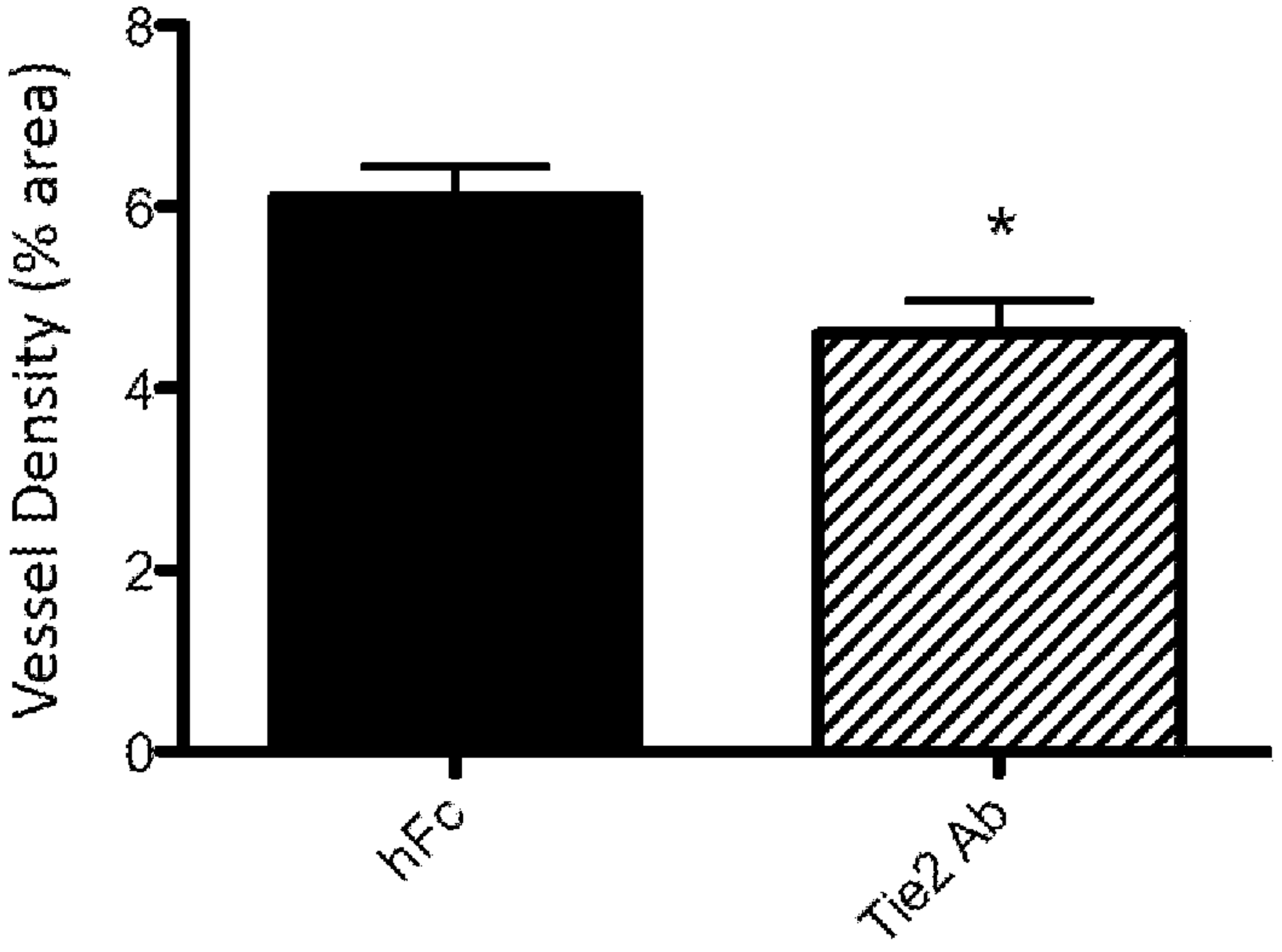
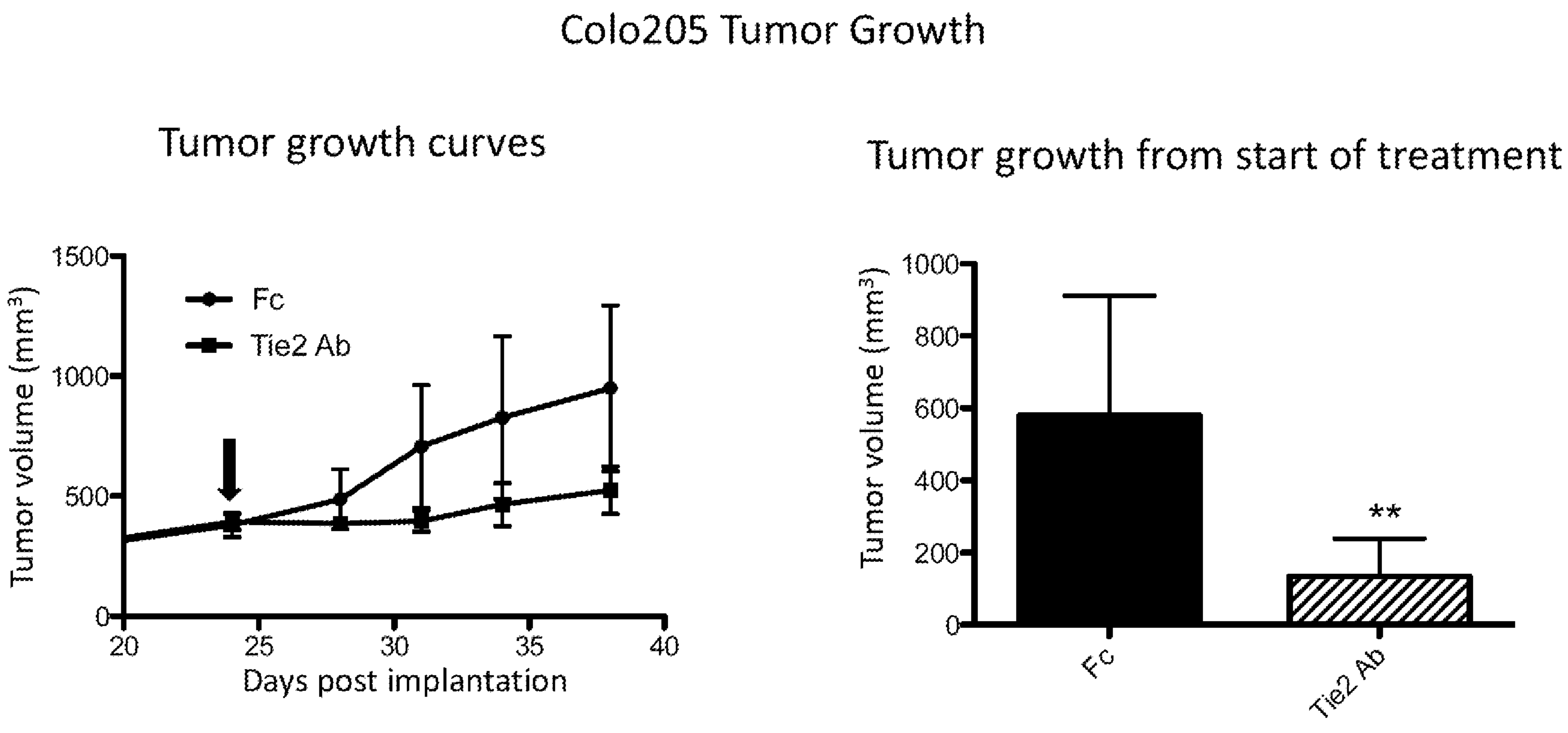


Figure 5


Colo205 Tumor Growth
Tumor growth curves
Tumor volume (mm³)
1500
1000
500
0
20
25
30
35
40
Days post implantation
Fc
Tie2 Ab
Tumor growth from start of treatment
Tumor volume (mm³)
1000
800
600
400
200
0
**
Fc
Tie2 Ab

A

B

Figure 6

Colo205 Vessel Density

Tumor vessel density at end of experiment (21-28 days of treatment)

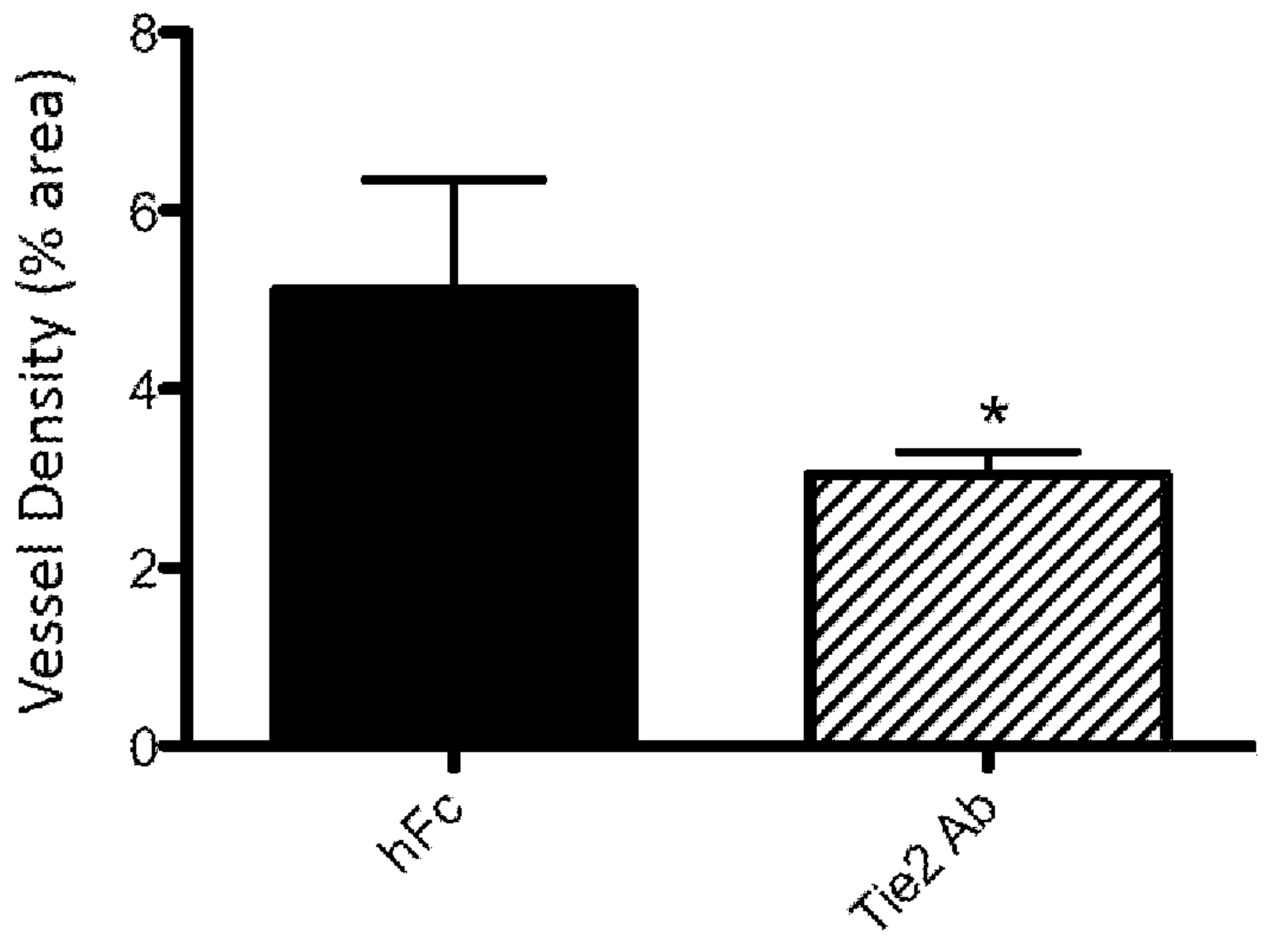


Figure 7

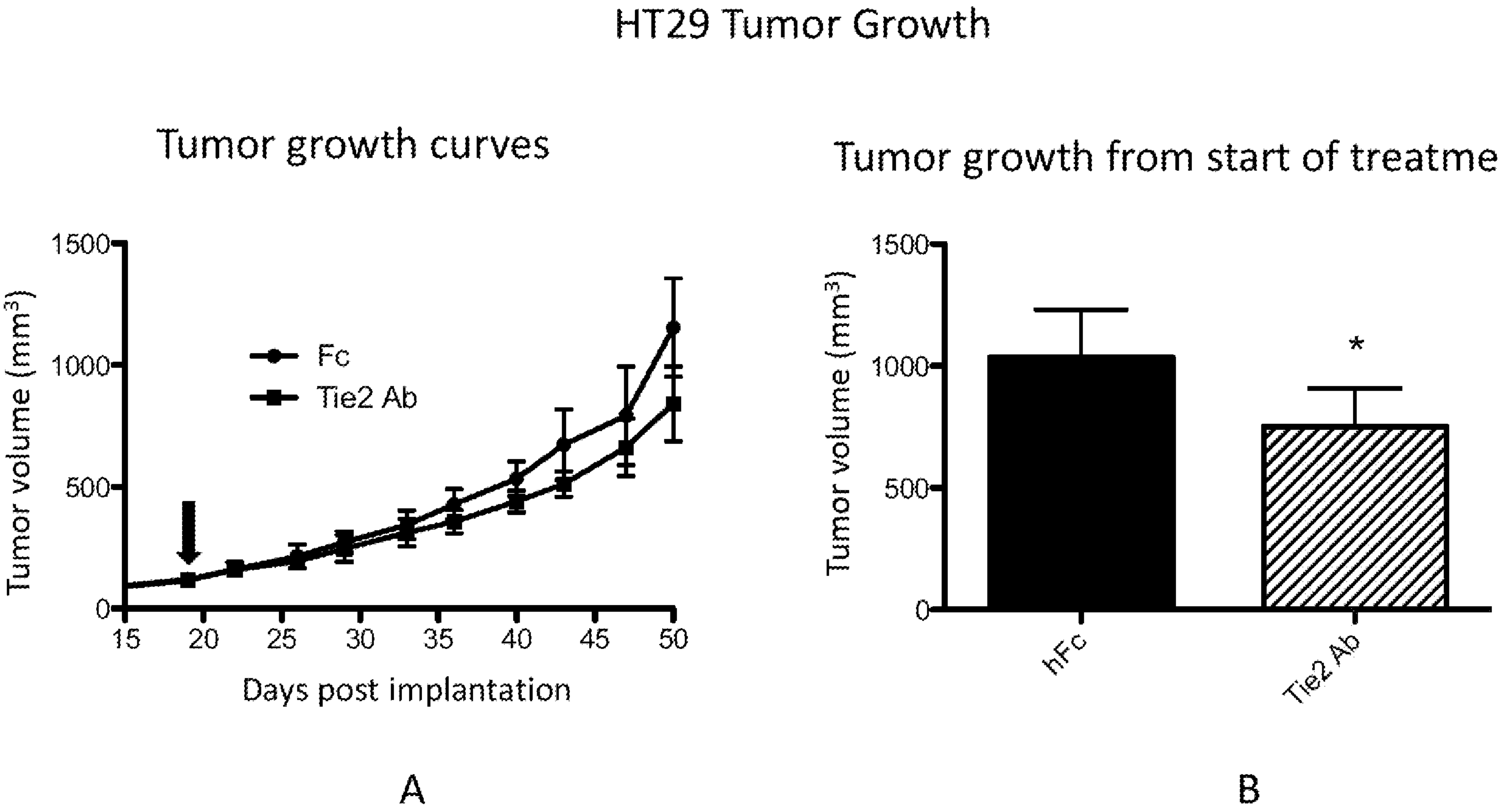

HT29 Tumor Growth
Tumor growth curves
Tumor volume (mm³)
1500
1000
500
0
Fc
Tie2 Ab
15
20
25
30
35
40
45
50
Days post implantation
A
Tumor growth from start of treatment
Tumor volume (mm³)
1500
1000
500
0
*
hFc
Tie2 Ab
B

Figure 4