Title: USE OF 13-HODE AS A REGULATOR OF VASCULAR BIOCOMPATIBILITY AND AN INHIBITOR OF CELL HYPERPLASIA

Abstract: This invention relates to the regulation of vascular endothelium biocompatibility and to the inhibition of vessel wall cell and other types of cell hyperplasia following vessel wall dysfunction and/or injury. More particularly, the invention relates to the dietary and pharmaceutical preparations of 13-hydroxyoctadec-9-Z, 11E-dienoic acid (13-HODE) and its use in reducing or inhibiting vessel wall hyperplasia and restoring vessel wall biocompatibility.
USE OF 13-HODE AS A REGULATOR OF VASCULAR BIOCOMPATIBILITY AND AN INHIBITOR OF CELL HYPERPLASIA

Field of the Invention

This invention relates to the regulation of vascular endothelium biocompatibility, as well as to the inhibition of vessel wall (VW) cell and other types of cell hyperplasia following vessel wall (VW) dysfunction and/or injury. Specifically, this invention relates to dietetic and pharmaceutical preparations of 13-hydroxyoctadeca-9Z, 11E-dienoic acid (13-HODE) and its use to restore vascular endothelial cell biocompatibility, thereby rendering the vasculature less reactive to circulating blood constituents during acute pathophysiological responses and decreasing chronic hyperplastic cell responses during and/or following VW stimulation, dysfunction or injury.

Background of the Invention

Cell cell interactions play a fundamental role in the genesis of most diseases including cardiovascular disease, cancer and metatasis, and infection and inflammation. The disadvantages and limitations of current antithrombotic therapies and the advantages of the present invention are discussed below in the context of cardiovascular disease, however these discussions are also relevant to other disease states.

Cardiovascular Disease: Treatment and Prevention - State of the Art

Cardiovascular disease is a major cause of morbidity and death in Western societies. It is exacerbated by smoking, hyperlipidemia, hypertension and diabetes. Over the last 40 years, our society has taken multiple steps to reduce cardiovascular disease such as promoting a healthier lifestyle, particularly in regard to smoking and diet. Nonetheless, each year, there are > 600,000 percutaneous transluminal
coronary angioplasty (PTCA) and surgically invasive procedures, e.g. coronary artery bypass grafting (CABG) in N. America alone, performed in cardiovascular disease patients to improve (cardio)vascular blood flow. While these procedures are beneficial to many patients, the benefits are finite and short-lived, and VW stenosis will reoccur. (RITA Trial Participants. 1993; Kirklin JW et al. 1989). For example, restenosis occurs in 25-30% of patients within 6 months of PTCA despite acute heparin treatment, followed by continuous aspirin (ASA) treatment ± oral anticoagulants throughout the 6 month post PTCA period. Heparin is given to accelerate thrombin inhibition by antithrombin III (ATIII), thereby preventing fibrinogen cleavage to fibrin and subsequent fibrin clot formation; ASA is given to acetylitate platelet cyclooxygenase, thereby inhibiting thromboxane A2 (TXA₂) synthesis which renders platelets less reactive to prothrombotic stimuli; an oral anticoagulant, e.g. coumadin, is given to decrease the level of vitamin K-dependent procoagulants, thereby decreasing the amounts of procoagulant substrates available for thrombus formation. Thus, the current approach to treat cardiovascular disease is to impair platelet function and/or coagulation as a means to prevent (re)occurrence of heart and blood vessel disease. It does not repair the underlying defect, the latter of which if attempted, might return the patient to a normal healthier state.

The only approaches currently proposed to reverse cardiovascular disease, are the use of lipid lowering agents which decrease the risk of atherosclerotic lesion formation, and gene therapy. The former approach also has provided some benefit, but again, it does not 'fix' the underlying problem. Gene therapy may, in fact, address the issue of repairing the underlying defect(s), but gene therapy for cardiovascular disease is still in its infancy, and not without the risk of complex side effects (Libby P and Ganz P. 1997).

A treatment process, which not only corrects the underlying cause of the disease problem but also prevents its onset, is therefore needed. The present invention, which relates to the use of 13-HODE in the regulation of blood cell/VW
compatibility per se, offers a more effective approach than do current antithrombotic therapies to both treating and preventing diseases like cardiovascular disease.

**Rationale for Regulating VW Biocompatibility**

In order to better understand the rationale for regulating VW biocompatibility and its benefits over current antithrombotic treatment practices, the rationale behind the current antithrombotic strategies and their obvious limitations is set out below. This, in turn, will highlight some insights which have led to the concept of regulating VW biocompatibility and the novel approach of using 13-HODE to treat and prevent cardiovascular disease of the present invention.

**Regulating VW Biocompatibility**

1. **Vessel Wall Stenosis & Occlusion:**

   The problem of vascular stenosis and subsequent occlusion is one of the most important of all medical problems and can produce a very wide range of diseases, the best known of which are coronary, cerebral and peripheral arterial blockage. It is, of course, very difficult and perhaps impossible to study the earliest development of such arterial blockages in humans. People who feel healthy are not inclined to submit to invasive procedures, which, in turn, may detect the onset of the disease before it manifests clinical symptoms. However, it is generally accepted that the processes of restenosis after an artery has been cleared or partially cleared of the occlusive material by a procedure such as angioplasty, is likely in many aspects to be similar to the processes involved in the original development of the problem. Vascular restenosis and occlusion after angioplasty or after vessel wall injury is therefore widely used as a model of the whole series of events involved in both primary and secondary arterial occlusion.

   Vascular restenosis is thought to occur as a result of a combination of intimal smooth muscle cell (SMC) proliferation, SMC synthesis and secretion of extracellular matrix, and VW remodelling. (Schwartz SM et al. 1995; Strauss BH et al. 1994;
Chervu A, Moore WS. 1990; Bocan TMA, Guyton JR. 1985). SMC proliferation per se, occurs in response to the mitogenic effects of thrombin generated at the time of VW injury, to platelet-derived growth factor (PDGF) secreted by platelets which adhere at the site of VW injury, and to mitogens secreted by activated endothelial cells (Bocan TMA, Guyton JR. 1985; Chen LB, Buchanan JM. 1975; Ross R. 1993; Bretschneider E et al. 1997; Fischman DL et al. 1994; Grandaliano G et al. 1998; Stouffer GA et al. 1998).

Polymorphonuclear leukocytes (PMNs) and monocytes/macrophages also invade the VW injury site, activating both coagulation and platelets, thereby augmenting the hyperplasia process (Alexander RW. 1994; Mallory GA et al. 1939; Mehta JL et al. 1998). Moreover, invading monocytes differentiate into macrophages, ingest lipids, calcium and other blood-derived constituents, thereby forming a more complex atherosclerotic plaque (Chervu A, Moore WS. 1990; Bocan TMA, Guyton JR. 1985). Thus, there is a multiplicity of cell cell interactions, which trigger and sustain intimal hyperplasia and subsequent restenosis (Ross R. 1993; Schwartz RS. 1998; Cicala C, Cirino G. 1997).

All of these events involve the interactions of blood components with the VW, which under 'healthy conditions' occur in response to injury and infection - but do not lead to (cardio)vascular disease. However, when these blood component/VW interactions are exaggerated such as with induced SMC proliferation, platelet/fibrin thrombus formation and VW hyperplasia, (cardio)vascular disease is initiated.

2. VW Injury, Repair and Remodelling:

Recent studies debate the relative contributions of intimal VW hyperplasia per se versus VW remodelling after injury, to subsequent VW restenosis in the clinical setting. Lafont, Post, Mintz et al argue that VW remodelling associated with internal elastic lamina dilation or constriction, contributes more to restenosis after PTCA than intimal hyperplasia (Lafont A et al. 1995; Post MJ et al. 1994). The results of the
Benestent and STRESS studies are said to be consistent with that argument since increasing the coronary artery diameter with a stent, decreases the need for revascularization (Fischman DL et al. 1994; Grandaliano G et al. 1998). Coats et al agree since there is more SMC-derived collagen (and presumably more SMCs) in non-stenosed VWs than in stenosed VWs (Coats WD et al. 1997; McGee MP et al. 1995). The opposite might be expected if hyperplasia was the predominate cause for restenosis. Coats et al suggested that the failure of our current antithrombotic therapy to inhibit restenosis as effectively as expected, is because that therapy focuses predominantly on inhibiting SMC proliferation. These conclusions, however, do not consider the heterogeneity of proliferating SMCs and their capacity to synthesise various matrices (Frid MG et al. 1997), or the fact that SMC collagen synthesis is affected by the presence (or absence) of the endothelium. Specifically, endothelial cells inhibit SMC protein synthesis, particular type III collagen (Myers PR, Tanner MA. 1998). The opposite is not true. It is also known that the extracellular matrix within a hyperplastic intima of a 1st injury VW, is rich in elastin while the extracellular matrix within the hyperplastic intima of a 2nd injury VW, is rich in collagen (Buchanan MR, Brister SJ. 1998), (Capron Let al. 1997.). Moreover, the clinical studies cited above were performed with patients who also required a stent due to the complex nature of their lesions. The restenosis rate in those patients is > 4 x's the restenosis rate in PTCA patients who do not require a stent (Antoniucci D et al. 1998). The treatment of PTCA patients who require a stent also differs significantly from the treatment of PTCA patients who do not require a stent (Antoniucci D et al. 1998). These differences are likely to affect subsequent outcome, both at the basic and the clinical end point levels. It is more likely that the relative roles of SMC hyperplasia and VW remodelling in restenosis varies depending on the type of injury and the type of the antithrombotic therapy use.

3. Blood Cell/Injured Vessel Wall Interactions:
Normally, the VW is nonthrombogenic and, therefore, biocompatible with the circulating blood. When the VW is injured, it becomes highly thrombogenic. Injured veins and arteries express tissue factor in both their media and intima. This expression increases over time after injury. Tissue factor expression is minimal in uninjured VWs (Channon KM et al. 1997). Tissue factor expression is enhanced further by PMNs and/or monocyte/macrophages, which invade the injury site. This enhancement is dependent on PMN and/or monocyte/macrophage CD18 integrin expression (Channon KM et al. 1997; McGee MP et al. 1995). VW tissue factor expression activates prothrombin, which is widely distributed throughout VW tissue rich in SMCs (McBane RD et al. 1997). Thrombin upregulates endothelial cell PDGF receptor expression, thereby facilitating SMC proliferation (Grandaliano G et al. 1998; DiCorleto PE, Bowen-Pope DF. 1983), and platelet activation. Activated platelets secrete TxA₂ (which is vasoconstrictive), PDGF (which is mitogenic for SMCs) and procoagulants, which exacerbate coagulation (Pakala R et al. 1997). Platelet-related factor Xa/Va activity bound to the injured VW also renders it highly thrombogenic. This latter effect persists for > 96 hours (Ghigliotti G et al. 1998). Thus, the multiplicity of these events could be addressed through use of an antithrombotic therapy which targets coagulation, platelet function and inflammation, and which also targets VW thrombogenicity per se. To date, this latter approach is virtually non-existent.

4. Anticoagulant Therapy and VW Hyperplasia:

A number of studies demonstrate that heparin can inhibit experimentally-induced SMC proliferation in vitro and in vivo (Castellot JJ Jr et al. 1984; Clowes AW, Clowes MM. 1986; Ferrell M et al. 1992; Hanke H et al. 1992). This suggests that heparin should prevent SMC hyperplasia and subsequent restenosis clinically. However, restenosis occurs clinically despite heparin treatment. It is now recognized that thrombin is protected from inhibition by ATIII and the acceleration of that effect by heparin when thrombin binds to fibrin or other constituents on the injured VW
surface (Okwusidi JL et al. 1991; Okwusidi JL et al. 1990; Hogg PJ, Jackson CM. 1989; Bar-Shavit R et al. 1989). Moreover, the surface-bound thrombin remains active, contributing to systemic hypercoagulation despite anticoagulant therapy (Ghigliotti G et al. 1998; Brister SJ et al. 1993; Wells J et al. 1994; Gill JB et al. 1993). Consequently, surface-bound thrombin can activate platelets, SMC proliferation and further coagulation unchecked. There also is evidence that SMCs, which proliferate in response to repeated injury, are less sensitive to the heparin treatment than SMCs, which proliferate in response to a first injury (Capron L et al. 1997; Geary RL et al. 1985).

5. Antiplatelet Therapy and Hyperplasia

There is little evidence that antiplatelet therapy per se reduces SMC hyperplasia. Clearly, ASA is beneficial in reducing the risks of stroke, myocardial infarction and transient ischemic attacks in patients with a variety of cardiovascular diseases (Aspirin Trialists’ Collaboration. 1994). However, the overall risk reduction with ASA, is only 25% (Aspirin Trialists’ Collaboration. 1994). While this risk reduction is statistically significant, the reduction is modest at best.

Also, ASA may benefit only certain subgroup of patients (Buchanan MR, Brister SJ. 1995; Grottemeyer K-H et al. 1993; Grottemeyer KH. 1991). This may be due, in part, to the wide variation in platelet responsiveness to assorted stimuli after ASA ingestion (Mueller MR et al. 1997). The effect of ASA is also finite and has little benefit after 2 years (Aspirin Trialists’ Collaboration. 1994).

Alternate antiplatelet agents which block the platelet glycoprotein IIb/IIIa (GPIIb/IIIa) receptor have been proposed as superior alternates to ASA. The EPILOG study demonstrated that blocking the GPIIb/IIIa receptor with c7E3 decreases acute ischemic complications in patients undergoing PTCA (The EPILOG Investigators. 1997). Similar results were obtained in the PRISM study using Aggrastat, a non-peptide GPIIb/IIIa antagonist (The Platelet Receptor Inhibition in
Ischemic Syndrome Management (PRISM) Study Investigators. 1998). It also has been suggested that the short $t^{1/2}$ (half-life) of these compounds may circumvent any bleeding side effect as compared to ASA. However, the bleeding issue still remains controversial. More importantly, like with aspirin, there is little clinical evidence to suggest that long-term hyperplasia is inhibited by these compounds.

Given recent studies, it is not surprising that platelet function inhibitors have little effect on preventing hyperplasia and restenosis. Specifically, Sirois et al made animals thrombocytopenic and then injured their arteries. Thrombocytopenia was sustained for short or long periods of time, and then their platelet counts were restored to normal levels. While the onset of VW hyperplasia was delayed in the long-term thrombocytopenic animals, the potential for SMC proliferation was not inhibited at all. Thus, medial SMC PDGR-β receptor expression was upregulated in all animals despite their being or not being thrombocytopenic. As a result, when the platelet count was restored to normal, SMC proliferation and subsequent intimal hyperplasia were initiated (Sirois MG et al. 1997). These data not only emphasise the need to regulate acute platelet/VW interactions to inhibit chronic intimal hyperplasia, but also suggest that platelet inhibition alone for any finite period of time is unlikely to have a lasting effect.

6. Limitations with the Current Antithrombotic Therapies

While all of the studies cited above, both experimental and clinical clearly indicate clinical benefits with the varied approaches to attenuate the different stages in the development of atherosclerosis, none of these approaches prevent disease onset or facilitate disease regression. Moreover, all of the therapeutic approaches mentioned above act indirectly by compromising coagulation, platelet function and/or injured vessel wall repair. As a result, all patients receiving any form of the currently recommended antithrombotic therapies, are rendered hemostatically dysfunctional, and therefore, at a significant hemorrhagic risk. Thus, there is a clear need for a
better antithrombotic approach which leads to the prevention and/or reversal of vascular disease, and which achieves these effects without any adverse side effects.

7. 13-HODE VW Biocompatibility and Hyperplasia

The concept of preventing VW hyperplasia by altering VW biocompatibility has not been considered directly, except perhaps, from the perspective of reducing fat and cholesterol intake in an attempt to reduce VW lipid accumulation and fatty streak formation on the VW. Most attempts have focussed more on the isolation and recombinant synthesis and subsequent utilization of VW constituents to alter blood component properties. For example, there is both experimental and clinical data to suggest that endothelial cell-derived nitric oxide, tissue plasminogen activator and prostacyclin are useful in the treatment of patients at risk of acute thromboembolic events (Gershlick AH et al. 1994; Zerkowski H-R et al. 1993; The GUSTO Investigators. 1993). Their effects, like the antiplatelet and anticoagulant therapies, target platelet function, vessel wall calibre and thrombolysis, thereby compromising hemostasis and coagulation. Moreover, it should be noted that all of these are only produced by the VW following injury or activation, and have little effect on regulating the innate biocompatible properties of a healthy, injured or diseased VW per se.

13-HODE is produced in various cells and tissues of the body, particularly by vascular endothelial cells in healthy vessel walls and by dermal epithelial cells. 13-HODE is formed by the action of an enzyme known as 15-lipoxygenase on the dietary essential fatty acid, linoleic acid. The first step is oxidation of the linoleic acid to give 13-hydroperoxyoctadeca-9Z, 11E-dienoic acid (13-HODE). This is then reduced to 13-HODE. 13-HODE is an important signal transduction molecule which is short-lived and whose synthesis is activated by a variety of different stimuli (Buchanan MR et al. 1985; Haas TA et al. 1990; Weber E et al. 1990; Bertomeu M-C et al. 1990; Brister SJ et al. 1990; Buchanan MR, Bastida E. 1988; Cho Y, Ziboh VA. 1994; Mari I. 1998; Kang L-T et al. 1999; Pongracz J, Lund JM. 1999; Friedrichs et
al. 1999; Cho Y, Ziboh VA. 1994). Many of the effects of 13-HODE are mediated by inhibition of protein kinases (PK), particularly PKC and mitogen-activated PK (MAP kinase).

13-HODE which is an oily liquid can be incorporated in much the same way as its parent fatty acid, linoleic acid, into a range of complex molecules including phospholipids and triglycerides (Spiteller G. 1998; Fang X et al. 1999). 13-HODE which is not incorporated into complex lipids is rapidly metabolized by hydrogenation and beta-oxidation (Bronstein JC, Bull AW. 1993; Hecht, Spiteller G. 1998).

Almost all of the studies designed to investigate the effects of 13-HODE involve measuring the effects of altering endogenous 13-HODE production or by adding exogenous 13-HODE (in various forms) to cultured cells in vitro. In the past, there have been few studies, which measure the effects of 13-HODE when given orally or parenterally to animals or humans. This limitation has been due, in part, to the difficulties of making large quantities of 13-HODE and its availability to the scientific community. Consequently, the amount of 13-HODE needed for in vivo studies has been extremely expensive. Second, generally it has been believed that 13-HODE is unstable and readily metabolized, like many signal molecules. As such, it has been thought that it would be a waste of time and money to perform studies involving the oral administration of 13-HODE since none of the orally administered material would be expected to reach its target site of action.

However, there are a few studies which suggest that orally administered 13-HODE has biological relevant effects in vivo. Streber's patent describes the use of 13-HODE and other related fatty acids to inhibit aromatase enzymes, which convert androgens to estrogens. The purpose of the treatment is to act on any disease, which is induced by estrogen such as breast cancer, and possibly some types of benign prostatic hyperplasia. However, it should be noted that all of the evidence provided in Streber's patent is based on data obtained in vitro. There are no experiments, which demonstrate that that invention actually works in vivo. Moreover,
Streber does not provide any details regarding the methods of administration or any practical details as to how the materials might be formulated (although it is stated that 'tablets or capsules or dragees' may be used). Finally, the daily dose specified ranges from 100 to 1,000 mg, preferably in the 200 to 500 mg range. (Streber AS. Hydroxy-octadecadienoic acid for the treatment of estrogen-dependent disease. US Patent # 5,102,912, April 1992).

The only study known to us which actually describes any experiments whereby a hydroxy derivative of linoleic acid has been administered orally outside of the experiences with the present invention discussed below, is that of Kaminakai et al (Japanese patent, #7-291862, November 7, 1995). However, they only mention 13-HODE in passing. The actual hydroxy derivative of linoleic acid manufactured for patent use in their experiments, is a different fatty acid; namely, 9-hydroxy-10(E)-12(Z) octadecadienoic acid (9-HODE) which is described in the NMR spectrum shown in Figure 2 of their patent, and which is stated in the text to be the material actually manufactured and studied in their experiments. These experiments involved the use of 9-HODE given orally to influence the action of Sarcoma 180 tumors implanted in the abdominal cavity of mice. They report an inhibitory effect on the rate of tumour growth, but the minimum effective dose required is 55 mg/kg/day.

The only 13-HODE studies which focus specifically on altering VW biocompatibility to prevent thrombogenesis have been reported by Buchanan et al. Their earlier studies demonstrate that healthy VW cells continuously turn over linoleic acid; i.e. at a time when the endothelium is nonthrombogenic or biocompatible with the circulating blood (Buchanan MR et al. 1985). Intracellular linoleic acid is metabolized to 13-HODE via the lipoxygenase pathway (Haas TA et al. 1990).

They also reported that:

i) endogenous VW 13-HODE plays an important role in regulating VW biocompatibility under both healthy and thrombogenic situations. For example, VW cell thrombogenicity varies inversely with VW 13-HODE levels in both animals and
humans. Therefore, increasing endogenous levels of 13-HODE in both animals and humans results in a decrease in VW cell thrombogenicity (Weber E et al. 1990; Bertomeu M-C et al. 1990; Brister SJ et al. 1990; Buchanan MR, Brister SJ. 1994); and a decrease in platelet/VW interactions following injury. (Weber E et al. 1990; Bertomeu M-C et al. 1990; Buchanan MR, Brister SJ. 1994); and

ii) 13-HODE down regulates the ability of the vitronectin receptor to recognise its ligands, thereby decreasing its adhesivity for vitronectin, fibronectin and fibrin(ogen) (Buchanan MR et al. 1998).

The mechanisms underlying these protective effects of 13-HODE are thought to involve inhibition of protein kinases (PK), particularly PKC and mitogen-activated PK (MAP kinase).

It is important to emphasize that in all of these studies, the aim was always to raise the level of endogenous production of 13-HODE by manipulating its endogenous synthesis or breakdown. None of these studies considered manipulating vessel wall 13-HODE levels by the exogenous administration of 13-HODE. This is clearly demonstrated by a series of experiments involving the administration of Persantine (dipyridamole) which is a phosphodiesterase inhibitor and which was thought might regulate endogenous 13-HODE metabolism.

In these experiments, it was demonstrated that an antithrombotic therapy which involved increasing VW 13-HODE levels, decreased SMC hyperplasia. Rabbits were treated with Persantine (1 mg/kg/day for 7 days) before a 1st or a 2nd VW injury, and then 4 weeks later, intimal SMC hyperplasia was measured. Persantine was given on the basis that it inhibits phosphodiesterase, thereby increasing VW cAMP levels (Weber E et al. 1990; Haas TA et al. 1990). Increasing VW cAMP increased VW linoleic acid turnover and subsequent VW 13-HODE synthesis, which, in turn, was associated with decreased platelet/VW interactions at the time of injury (Weber E et al. 1990). The Persantine treatment inhibited SMC hyperplasia 4 weeks after VW injury. Platelet function in these animals was unchanged.
These studies are consistent with the discovery that decreasing VW thrombogenicity by increasing VW 13-HODE at the time of injury will attenuate long-term intimal hyperplasia and subsequent VW restenosis. Moreover, this approach did not compromise normal hemostasis and coagulation like the currently used clinical approaches do.

13-HODE, Anti-inflammatory Therapy and VW Hyperplasia

Inflammation has been recognized as an integral part of the thrombotic process as early as 1939 (Mallory GA et al. 1939), yet it is not considered in the rationale for our current antithrombotic therapies. However, there is convincing evidence that attenuating certain inflammatory responses provide a significant benefit. For example, monocytes/macrophages and PMNs express the integrin CD11/CD18 (ICAM), and they release cytokines when activated (Peracchia R et al. 1997; Yasukawa H et al. 1997; Turek JJ et al. 1998), which, in turn, stimulate i) $\beta_3$ integrin expression in other cells such as platelets, endothelial cells and SMCs (Blanks J E et al. 1998; Golino P et al. 1997); ii) tissue factor activation (McGee MP et al. 1995); and iii) PDGF expression (Rubin P et al. 1998; Panek RL et al. 1997). Lipid fractions derived from platelets augment these responses by inducing monocyte/macrophage differentiation and growth (Ammon C et al. 1998). Macrophages interacting with the injured vessel wall, accumulate lipid, leading to the formation of a more complex atherosclerotic lesion (Ross R. 1993; Post MJ et al. 1994). Blocking monocyte/macrophage ICAM expression reduces VW hyperplasia significantly (Golino P et al. 1997; Nageh MR et al. 1997; Natori S et al. 1997). Others have found that radiation ($^{89}$Sr/Y or $^{102}$Ir) at doses, which selectively impair monocyte/macrophage function also, decreases VW hyperplasia in both rodent and rabbit models (Rubin P et al. 1998; Panek RL et al. 1997; Williams DO. 1998; Kipshidze N et al. 1998). In a preliminary clinical study, the SCRIPPS trial using endovascular radiation, the restenosis rate in 35 patients undergoing PTCA who also
required a stent was 11%, significantly less than the 37% restenosis rate seen in comparable non-irradiated controls (Williams DO, 1998). These data provide direct evidence, which suggests that altering inflammatory responses also affect intimal hyperplasia and subsequent VW restenosis.

Studies by Buchanan et al suggest that the culprit inflammatory cell is not the PMN. In fact, PMNs may attenuate the vessel wall thrombogenicity by providing a source of 13-HODE at the site of blood cell/VW interactions at the time of VW injury (Buchanan MR, 1989; Buchanan MR et al. 1993). Others have argued that PMN-derived oxygen radicals promote ischemia-related damage (Shen J et al. 1996), but this has not been linked to long-term hyperplasia. PMNs also secrete a nitric oxide-like factor, which inhibits platelet function and vasoconstriction (Cerletti C et al. 1992). Monocytes/macrophages, on the other hand, normally do not synthesize 13-HODE (Shen J et al. 1996; Shen J et al. 1995). Interestingly however, Shen et al upregulated 15-lipoxygenase in differentiated macrophages and found that 13-HODE synthesis increased. This increase was associated with decreased macrophage lipid accumulation. Fan et al also found that macrophages enriched with linoleic and gamma linolenic acids (substrates for 13-HODE and PGE₁, respectively), stimulate intracellular SMC cAMP which, in turn, decreases SMC proliferation (Fan YY et al. 1997). Finally, 13-HODE inhibits PAF (platelet activating factor)-induced PMN and monocyte/macrophage degranulation and ICAM expression (Cerletti C et al. 1992), thereby preventing further integrin-dependent cell cell and cell ligand interactions.

Recent Studies with 13-HODE

The above data suggest that endogenous VW 13-HODE plays an important role in regulating VW biocompatibility under both healthy and thrombogenic situations, and that 13-HODE is therefore a useful antithrombotic agent. However, any progress in developing that concept has been thwarted by the lack or absence of any agent which would directly upregulate 13-HODE synthesis by VW cells, PMNs or
other relevant cells. While Persantine has been a useful tool to generate preliminary data, it is a weak and reversible inhibitory of phosphodiesterase. Supplementing a diet of cardiovascular diseased patients with linoleic acid (the substrate for 13-HODE) also has been useful to demonstrate the benefit of elevating VW 13-HODE levels. However, that approach requires the patients to ingest a daily dose of 20 grams or more of linoleic acid-rich capsules, and the treatment is not without its unwanted side effects, including an increased caloric intake.

**Surprising Results**

To date, researchers working in the field have concentrated on the idea of maintaining healthy endothelial cell function either by regulating the endogenous production of 13-HODE, avoiding factors which suppress its synthesis and/or providing agents containing linoleic acid which may enhance the synthesis of 13-HODE. It was thought that only trivial amounts of purified 13-HODE would reach the target site of the VW endothelium if administered orally, and would, therefore, be biologically inactive. Recently, these ideas were tested and, surprisingly, proven wrong. Specifically, it was found that orally administered 13-HODE does reach its intended targets and is biologically active. In addition, suitable vehicles in which 13-HODE can be administered orally were identified. And most amazingly, the beneficial effects of orally administered 13-HODE are achieved with unexpectedly low doses.

These and other objects and advantages of the invention will be apparent to those skilled in the art from a reading of the following description and appended claims.

**Summary of the Invention**

This invention relates to a method of reducing or inhibiting cell and vessel wall hyperplasia and restoring vessel wall biocompatibility, comprising administering to an
animal or human in need of such treatment an amount of 13-hydroxyoctadeca-9Z, 11E-dienoic acid (13-HODE) effective to reduce or inhibit vessel wall thrombogenicity wherein the compound is administered orally.

This invention also relates to the method described above wherein the pharmaceutical composition comprises 13-HODE either in its free form, or with a pharmaceutically acceptable carrier, auxiliary or excipient.

The carrier, auxiliary or excipient may be mono-, di- or triglyceride oil, corn, sunflower, safflower, cottonseed, grape seed, olive, evening primrose, borage, fish body and fish liver oils, or an ester of a fatty acid containing 16-26 carbon atoms and one or more double bonds. The estermay be ethyl-eicosapentaenoic (ethyl-EPA), oleic, linoleic, alpha-linoleic, stearidonic, gamma-linolenic, dihomogammalinolenic, arachidonic, docosapentaenoic or docosahexaenoic (DHA).

This invention also includes a pharmaceutical composition comprising 13-HODE and a fat-soluble antioxidant, such as, ascorbyl palmitate, tocopherols, and ascorbic acid in the presence of lecithin.

Furthermore, this invention relates to a pharmaceutical composition comprising 13-HODE and an additive selected from the group consisting of aggregants, disaggregants, osmotic pressure regulating salts, buffers, sweeteners, and coloring agents.

The pharmaceutical composition or 13-HODE of this invention may be administered in the form of a dietetic composition, or as a formulation selected from the group consisting of tablets, dragees, capsules, granules, suppositories, solutions, suspensions and lyophilized compositions. A pharmaceutical composition as described above wherein the daily dose of 13-HODE is equal to or less than 100 mg is also part of this invention.
The invention further relates to a pharmaceutical composition comprising 13-HODE and an omega-3 fatty acid (such as EPA, DHA, or any a derivative of EPA or DHA, such as ethyl-EPA or ethyl-DHA).

The invention includes a method of correcting the inhibition of endogenous 13-HODE synthesis by omega-3 fatty acids by incorporating 13-HODE into formulations of omega-3 fatty acids.

The method and the pharmaceutical composition of 13-HODE of this invention may be used to treat cardiovascular or cerebrovascular disease, inflammatory or autoimmune disease, infection with bacteria, viruses, fungi, or protozoa, respiratory disease, gastrointestinal disease, renal or urinary tract disease, skin disease, neurological or psychiatric disease, disease of the reproductive system, diabetes, syndrome X or any complication of diabetes, diseases associated with overactive protein kinases, and diseases associated with endothelial dysfunction.

The method and the pharmaceutical composition of 13-HODE of this invention may also be used to treat and/or prevent cancer or the metastatic spread of cancer.

Other and further advantages and features of the invention will be apparent to those skilled in the art from the following detailed description of the invention taken in conjunction with the accompanying drawings.

**Brief Description of the Drawings**

In the accompanying drawings:

FIG. 1 illustrates the cell free plasma 13-HODE levels in rabbits treated orally with 13-HODE (suspended in corn oil, upper; or EPA, lower) at 100 to 1000 μg/kg/day on days 1 to 7.
FIG. 2 illustrates 13-HODE levels in vessel walls obtained from rabbits treated orally with 0 to 100 µg/kg/day for 7 days. 13-HODE was suspended in corn oil (upper) or EPA (lower) and given on days 1 to 7.

FIG. 3 illustrates the effect of 13-HODE, suspended in corn oil, on increase of vessel wall area of injured vessel walls from rabbits treated orally with 0 or 1000 µg/kg/day for 7 days.

FIG. 4 illustrates cell free plasma 13-HODE levels in rabbits treated orally with 13-HODE (suspended in corn oil upper; or EPA, lower) at 100 to 1000 µg/kg/day on days 1 to 7.

FIG. 5 illustrates effect of 13-HODE, suspended in corn oil (upper) or EPA (lower), on increase of vessel wall area of injured vessel walls from rabbits treated orally with 0 to 1000 µg/kg/day for 7 days.

FIG. 6 illustrates the properties of 13-hydroxyoctadeca-9Z, 11E-dienoic acid (13-HODE).

**Detailed Description of the Invention and Preferred Embodiments**

The present invention demonstrates a method for reducing or inhibiting vessel wall hyperplasia and restoring vessel wall biocompatibility in a mammal or human in need of such treatment, by administering an amount of 13-hydroxyoctadeca-9Z, 11E-dienoic acid (13-HODE) effective to reduce or inhibit vessel wall thrombogenicity.

The present invention also specifically demonstrates the beneficial effects of exogenously administered 13-HODE in animals and humans of vascular response to
VW injury, including the prevention and treatment of vessel wall hyperplasia, as well as the facilitation of vessel wall disease regression.

As discussed earlier, 13-HODE is one of the factors, which regulates vessel wall biocompatibility, thereby attenuating untoward blood component/vessel wall interactions. It may do this in several ways, one of which probably is to reduce the expression and activation of the vitronectin receptor. In recent years, it has been found that abnormalities of vessel wall biocompatibility are associated with a remarkable number of illnesses, including infections, cardiovascular problems of many types, as well as to problems relating to inflammation, fibrosis and loss of normal metabolic control (Vallance et al. 1997), and tumour cell metastasis. In all of these illnesses, vascular endothelial cells are activated, leading to the loss of the normal vascular permeability barrier, expression of leukocyte adhesion molecules, change in VW surface thromboreactivity, the production of a wide range of cytokines and the upregulation of HLA antigens (Hunt BJ, Jurd KM.). A wide range of illnesses may be caused or exacerbated by endothelial cell activation and damage.

The present invention through the regulation of endothelial function in a favourable direction by 13-HODE, will therefore have a desirable effect in a wide variety of diseases encompassing almost every aspect of medicine. Of particular interest is the fact that the administration of exogenous 13-HODE in this invention is able to compensate for any suppression of endogenous 13-HODE synthesis which can occur as a result of administration of omega-3 fatty acids such as alpha-linoleic acid, docoapentaenoic acid and particularly EPA and docosahexaenoic acid (DHA) (Miller and Ziboh, as above). EPA and DHA have many desirable actions but sometimes the clinical results of administering EPA and DHA are less favourable than expected, as in the case with attempts to prevent reocclusion after angioplasty (Cairns et al, as above). The co-administration of 13-HODE with EPA or DHA or other omega-3 fatty acids is therefore of particular value.
The composition according to the present invention can be formulated for administration orally. Thus the composition may be in the form of tablets, capsules, suspensions, emulsions and solutions.

Formulations for oral use include tablets, which contain the active ingredient in admixture with non-toxic pharmaceutically acceptable excipients. The excipients may be, for example, inert diluents, such as calcium carbonate, sodium chloride, lactose, calcium phosphate or sodium phosphate; granulating and disintegrating agents, for example, potato starch or alginic acid; binding agents, for example, starch, gelatin, or acacia; and lubricating agents, for example, magnesium stearate, stearic acid or talc. Other pharmaceutically acceptable excipients can be colorants, flavoring agents, plasticizers, humectants etc. The tablets may be uncoated or they may coated by known techniques, optionally to delay disintegration and absorption in the gastrointestinal tract and thereby provide a sustained action over a longer period. For example, a time delay material such as glyceryl monostearate or glyceryl distearate may be employed.

Formulations for oral use may also be presented as chewing tablets, or as hard gelatin capsules wherein the active ingredient is mixed with an inert solid diluent, for example, calcium carbonate, calcium phosphate or kaolin, or as soft gelatin capsules wherein the active ingredient is mixed with water or an oil medium, for example, peanut oil, liquid paraffin or olive oil. Powders, dispersible powders or granules suitable for preparation of an aqueous suspension by addition of water are also convenient dosage forms of the present invention. Formulation as a suspension provide the active ingredient in admixture with a dispersing or wetting agent, suspending agent and one or more preservatives. Suitable dispersion or wetting agents are, for example, naturally-occurring phosphatides, as e.g. lecithin, or condensation products of ethylene oxide with e.g. a fatty acid, a long chain aliphatic alcohol or a partial ester derived from fatty acids and a hexitol or hexitol anhydrides, for example, polyoxyethylene stearate, polyoxyethylene sorbitol monooleate,
polyoxyethylene sorbitan monooleate etc. Suitable suspending agents are, for example, sodium carboxymethylcellulose, methylcellulose, sodium alginate etc.

The pharmaceutically acceptable carriers or excipients may include emulsifying agents, antioxidants, buffering agents, preservatives, humectants, penetration enhancers, chelating agents, gelforming agents, ointment bases, perfumes and skin protective agents. Examples of emulsifying agents are naturally occurring gums, e.g. gum acacia or gum tragacanth, naturally occurring phosphatides, e.g. soybean lecithin and sorbitan monooleate derivatives. Examples of antioxidants are butylated hydroxy anisole (BHA), ascorbic acid and derivatives thereof, tocopherol and derivatives thereof and cysteine. Examples of preservative are parabens and benzalkonium chloride. Examples of humectants are glycerin, propylene glycol, sorbitol and urea. Examples of penetration enhancers are propylene glycol, DMSO, triethanolamine, N,N-dimethylacetamide, N,N-dimethylformamide, 2-pyrrolidone and derivatives thereof, tetrahydrofurfuryl alcohol and Azone. Examples of chelating agents are sodium EDTA, citric acid and phosphoric acid. Examples of gel forming agents are Carbopol, cellulose derivatives, bentonit, alginates, gelatin and PVP. Examples of ointment bases are beeswax, paraffin, cetyl palmitate, vegetable oil, sorbitan esters of fatty acids (Span), polyethyleneglycols and condensation products between sorbitan esters of fatty acids and ethylene oxide, e.g. polyoxyethylene sorbitan monooleate (Tween). The formulation and preparation of the above-mentioned compositions is well known to those skilled in the art of pharmaceutical formulation. Specific formulation can be found in "Remington's Pharmaceutical Sciences".

Dietetic compositions of the present invention may be made up in the form of emulsions, for example, sauces, mayonnaise or margarine.

One embodiment of the invention relates to the administration of a pharmaceutical composition, in conjunction with a pharmaceutically acceptable carrier, for the therapeutic effects, which are discussed below.
In a preferred embodiment of the invention, the pharmaceutical composition of 13-HODE is administered orally, and comprises a combination product containing 13-HODE in combination with a carrier from the group consisting of corn, sunflower, safflower, cottonseed, grapeseed, olive, evening primrose, borage, fish body, fish liver oils, ethyl-eicosapentaenoic, oleic, linoleic, alpha-linolenic, stearidonic, gamma-linolenic, dihomogammalinolenic, arachidonic, docosapentaenoic or docosahexaenoic (DHA).

In another preferred embodiment, the pharmaceutical composition of the present invention comprises a combination product containing 13-HODE in combination with corn oil in a ratio between about 1:3 to about 1:100. For example, 50mg of 13-HODE can be mixed with 450 mg corn oil.

In another embodiment, the pharmaceutical composition of the present invention comprises a combination product containing 13-HODE in combination with an ester carrier in a ratio of about 1:3 to about 1:100.

In yet another embodiment, the pharmaceutical composition of the present invention comprises a combination product containing 13-HODE in combination with an ethyl ester of a 16-26 carbon fatty acid with one or more double bonds, such as ethyl-oleate, ethyl-linolate, ethyl-EPA or ethyl-DHA.

The pharmaceutical composition in a further embodiment of the present invention comprises 13-HODE is incorporated into the Sn1 or Sn2 positions of an appropriate phospholipid prior to mixing with a carrier.

In another preferred embodiment of the invention, the pharmaceutical composition comprises 13-HODE and omega-3 fatty acids, like EPA, DHA, derivatives of EPA and DHA, ethyl-EPA and ethyl-DHA.

The compositions are useful when administered in methods of medical treatment or prophylaxis of a disease, disorder or abnormal physical state associated with abnormalities of vessel wall biocompatibility, including cardiovascular or cerebrovascular, inflammatory or auto-immune, respiratory, gastrointestinal, renal,
skin, neurological and psychiatric diseases and cancers. Examples of diseases, which can be treated or prevented by administering the pharmaceutical composition of the present invention include both Type I, Type II and the precursor Type II diabetes, syndrome X (Cosentino F, Lucher TF. 1998; Steinberg AD. 1997), many types of inflammatory disorders including rheumatoid arthritis and osteoarthritis and autoimmune diseases (Perretti M. 1997), infections with bacteria and protozoa like malaria and sleeping sickness, and fungi which can generate endotoxins (Gerrity et al. 1976), sickle cell disease and related haemoglobins disorders (Lubin BH. 1997), kidney disease (Clausen et al. 1999), inflammatory bowel disease (Binion DG et al. 1998) pregnancy hypertension and pre-eclampsia (Endresen MJR et al. 1998), normal aging (Hashimoto M et al. 1999), dementias (Iadecola C et al. 1999), retinal ischemia and age-related macular degeneration (Gidday JM, Zhu Y. 1998; Wada M et al. 1999), cancer and especially cancer metastasis and angiogenesis (Pinedo HM et al. 1998; Shureigi I et al. 1999; Baron Ja et al. 1998; Hazelton D et al. 1999), all types of cardiovascular diseases (Kanani PM et al. 1999), hyperlipidemias and atherosclerosis of all types (Lefer DJ, Granger DN. 1999; Mombouli J-V. 1999; Blann AD. 1999; Brown BG, Zhao X-Q. 1998; Freedman JE, Loscalzo J. 1997; Abe Y. 1998), transplantation (Labarrere CA et al. 1997), pulmonary hypertension (Higenbottom TW, Laude EA. 1998), hypertension and heart failure (Boulanger CM. 1999), and in Raynaud’s syndrome (Edwards CM et al. 1999). In addition, smoking can be treated with the present invention as endothelial function also is impaired by smoking (Motoyama T et al., 1997) in normal people who have a high fat meal (Plotnick GD et al., 1997), and in apparently healthy people who had a low birth weight or who are at risk of cardiac disease (Goodfellow J et al.) Endothelial function is impaired in fit young adults of low birth weight. (Ridker PM et al., 1998) Plasma concentrations of soluble intercellular adhesion molecule 1 and risks of future myocardial infarction in apparently healthy men. (Lancet, 1998).
Dosages to be administered depend on individual patient condition, indication of the drug, physical and chemical stability of the drug, toxicity of the desired effect and the chosen route of administration (Robert Rakel, ed., Conn's Current Therapy (1995, W.B. Saunders Company, USA)).

The following Examples are presented as specific illustrations of the present invention. It should be understood, however, that the invention is not limited to the specific details set forth in the Examples.

Example 1

Identifying the Appropriate vehicle for 13-HODE

13-HODE is a colourless or very pale yellow oily liquid. Within the body 13-HODE can be metabolized as described earlier or transferred intact between various possible complex lipids including triglycerides, diacylglycerols (diacylglycerols), monoglycerides, cholesterol esters and phospholipids of many different types. As a pharmaceutical, 13-HODE may be used as is, or be dissolved in various carriers, or be incorporated into glyceride, ester, phospholipid or other molecules with any appropriate carrier. Glycerides, esters of propane diol, ethyl esters and phospholipids to which 13-HODE is co-valently bound and any other molecules or vehicles which can release13-HODE in a biologically active form within the body all lie within this invention.

A real problem is presented by the fact that active daily doses of 13-HODE are in the 50 to 1,000 µg/kg/day range. The lower end of this range, 100 µg/kg/day, was shown to be highly biologically active in rabbits. That translates into a 7 mg/day dose for a 70 kg person. Since doses for humans are often considerably lower than doses for animals because of a weight/body surface area scaling, a daily dose of as little as 5 mg or less is really possible. Formulating an oily liquid is such small doses is a problem. It could be absorbed into tabletting materials and tableted and coated,
or micro-encapsulated by methods well known to those skilled in the art. However, the most convenient and preferred dosage forms of 13-HODE, and the ones in which it was found to be stable without degradation, was to dissolve the 13-HODE into a triglyceride oil carrier or an ester. It was found that corn oil is a particularly useful diluent, and that 13-HODE can be readily and conveniently mixed with corn oil in a ratio, for example, from 1:3 to 1:100. Other triglyceride oils such as vegetable oil, including sunflower, safflower, soy, evening primrose, borage, coconut or palm oil, or cottonseed, rapeseed, olive, fish body or fish liver oils may all be used for this purpose. Particularly useful esters for this purpose are esters of fatty acids with 16-26 carbon atoms and one or more double bonds in the chain. Ethyl ester of EPA was found to be particularly appropriate, but equivalent esters of fatty acids such as oleic, linoleic, alpha-linoleic, stearidonic, gamma-linoleic, dihomogammapalnineoleic, arachidonic, docosapentaenoic and DHA are all examples of esters which could be useful to carry the 13-HODE.

The preparations can then be further processed to give a final dosage form. The oils can be ingested directly, or appropriate antioxidants or flavours can be added, or they can be converted into palatable, flavoured emulsions by the use of emulsifying agents or flavouring well known to those skilled in the art. A particularly valuable dosage form is a soft gelatin or bonded hard gelatin capsule, or a similar capsule made with agar or other appropriate materials.

At present, the only readily available supplies of 13-HODE are in very expensive mg quantities for use as laboratory reagents. The properties of the pure material are shown in Figure 6. Larger quantities can now be prepared using soybean lipoxygenase or an enzyme of similar specificity. This enzyme metabolizes linoleic acid to 13-HODE. The reaction can be carried out in an appropriate vessel filled with a cooling and stirring system and pH, oxygen (dissolved oxygen content, DOC), and temperature probes. The reaction is first charged with 0.1 M borate buffer which is then chilled to below 100 ° C. The buffer is then purged with oxygen until
the DOC reaches 100%. Soybean lipoxygenase is then added at the rate of about 2500 U/litre and the mixture is stirred and regularly purged with oxygen to keep the DOC at 100%. Octa-deca-9Z, 12Z-dienoic acid, dissolved 1/1 in ethanol is then added at a rate of about 10 g/litre. The reaction is then pressurized with an overblanket of oxygen, and vigorously stirred. The reaction is allowed to proceed, and monitored at 15 minute intervals by ultraviolet analysis and thin layer chromatography analysis to confirm the conversion of the linoleic acid to 13-HODE.

On completion of the reaction, the vessel is flushed with nitrogen and reduced by adding sodium borohydride at the rate of about 3.3 g/litre. On completion of the reduction process, the mixture is acidified to pH 6 with citric acid. Reverse phase silica (OD53) is then added and stirred and the mixture is allowed to continue to stir overnight under nitrogen at room temperature. The silica absorbs the 13-HODE, which is then recovered by filtering the silica, washing it with water, and then eluting out the product by multiple washing with acetonitrile. The solvent is then washed off and the crude oil is purified by column chromatography with diethyl ester and methylene chloride to yield pure 13-HODE as a viscous pale yellow oily substance. This material can then be formulated as discussed above.

In some of the experiments conducted, corn oil was used because it had previously been found that suspending linoleic acid in corn oil facilitated its selective uptake by the VW (Bertomeu M-C et al. 1990). In other experiments, marine fish oil was used (specifically marine menhaden or 97% pure ethyl eicosapentaenoic (ethyl-EPA)) since marine fish oils such as tuna, sardine or other oils also are suitable to maintain 13-HODE stability. Ethyl-EPA was of great interest therapeutically because EPA has many desirable actions other than on the VW endothelium such as the inhibition of platelet aggregation, the lowering of triglyceride levels and the attenuation of inflammation. However, it also can inhibit the formation of endogenous
13-HODE, which might be a possible negative effect (Miller CC, Ziboh, VA. 1990; Gimenez-Arnau A et al. 1997).

This inhibitory effect of EPA may help explain why the expected desirable cardiovascular effects of EPA have not been realized in practice. For example, a continuous course of EPA treatment failed to reduce restenosis after occluded coronary arteries had been opened by angioplasty (Cairns JA et al. 1996).

Materials and methods:

New Zealand white rabbits (half males/half females; 2.5 - 2.9 kg) were used throughout. Rabbits were treated with 100, 400 or 1000 μg/kg/day of purified 13-HODE suspended in corn oil or ethyl-eicosapentaenoic acid (EPA), or with an equi-volume of either suspending vehicle (total volume 1 ml) for 7 days. Serial blood samples were collected before, during and after treatment to assess the levels of 13-HODE in plasma. On day 7, the treatments were stopped. At that time, the rabbits were anaesthetised with a combination of Atravet, Ketamine, Vetrepham and glycopyrolate, given subcutaneously. Both carotid arteries of anaesthetised rabbits were isolated between 2 temporary ligatures, first by applying the proximal ligature, then allowing the blood to drain from the segment, and then applying the distal ligature. A 24-gauge angiocath attached to tubing filled with sterile saline and connected to a pressure manometer, was inserted into the isolated segment. Then the segment was filled with the saline to a pressure of 600 mm Hg which was maintained for 5 minutes. The pressure was relieved, and the angiocath and ligatures were removed, thereby allowing for blood flow restoration. Cessation of bleeding from the needle puncture site was achieved within 3 minutes without any manual manipulation. The incisions were sutured closed with 000 proline. The rabbits were injected intramuscularly with Temgesic to minimize any pain and with 12.5 mg Baytril as an antibiotic, and then allowed to recover. This injury procedure results in endothelial denudation and the exposure of a thrombogenic surface within
1 hour of restoration of blood flow, followed by SMC proliferation and intimal hyperplasia, which plateaus at 4 weeks and which is sustained for ≥12 weeks (Buchanan MR, Brister SJ: 1998; Buchanan MR et al. 1999).

Two, four or twelve weeks later, the rabbits were re-anaesthetised and their injured carotid arteries were again isolated. Segments of injured and uninjured carotid artery were harvested and processed histologically to assess VW hyperplasia. Other VW segments and blood samples were collected and processed for VW and plasma 13-HODE levels.

Results

**Plasma 13-HODE Levels:** There was a three-fold increase in the plasma 13-HODE levels after 7 days of treatment with 100 µg/kg/day of 13-HODE suspended in corn oil (Fig 1, upper panel). Increasing the 13-HODE dose to 1,000 µg/kg/day had no further effect. The plasma 13-HODE levels returned back to almost control levels within 14 to 21 days. Similar results were seen when the 13-HODE was suspended in ethyl-EPA (Fig 1, lower panel), although the absolute levels of plasma 13-HODE were lower in all treatment levels tested. Notwithstanding, there were no significant differences in the dose-related increases in plasma 13-HODE levels between the two suspending vehicle treatment groups.

**VW 13-HODE Levels:** There also was a dose-related increase in VW wall 13-HODE levels in the rabbits treated with 13-HODE suspended in corn oil and measured 28 days after stopping the treatment (Fig 2, upper panel). These data indicated the VW 13-HODE levels remain elevated despite stopping the treatment. This may be due to the 13-HODE being incorporated into complex lipids as demonstrated by Fang et al (1999).

Similar results were seen when the 13-HODE was suspended in ethyl-EPA (Fig 2, lower panel), but again, the absolute levels of VW 13-HODE were somewhat
lower at all treatment levels tested. Since the VW 13-HODE levels were of course a measurement of both endogenous- and exogenously-derived 13-HODE, it is possible that the lower VW 13-HODE levels seen in the ethyl-EPA treated group reflect a suppression of endogenous 13-HODE synthesis as described by Miller and Ziboh (1990).

**Biological Effects:** In the initial studies with the purified 13-HODE, the marked increases in VW 13-HODE were associated with a significant decrease in intimal hyperplasia measured 2 and 4 weeks after injury. Thus, intimal hyperplasia in rabbits treated with 1,000 μg/kg/day of 13-HODE, was 8 and 16% at 2 and 4 weeks respectively, compared to 18 and 38% hyperplasia seen at 2 and 4 weeks, respectively in the placebo treated rabbits, p < 0.002 (Fig 3). Moreover, the intimal hyperplasia regressed in the 13-HODE treated animals such that intimal hyperplasia was barely detectable at 12 weeks, > 3%, p < 0.01. This is the first direct evidence demonstrating that purified 13-HODE inhibits VW intimal hyperplasia. It was interesting to note that the 13-HODE level in the cell-free plasmas of those animals not only was elevated significantly within 3 days of treatment, but remained elevated for 4 weeks despite our stopping the 13-HODE treatment after 7 days (Fig 4). These latter data suggest that orally administered 13-HODE is well absorbed, has a long half life, and can be monitored relatively easily in a clinical setting. These latter data also suggested to us that a markedly lower dose of 13-HODE could be as effective in preventing VW intimal hyperplasia.

**Example 2**

Rabbits were administered a 7 day course of 100 μg/kg/day of 13-HODE suspended in either corn oil or ethyl-EPA, significantly inhibited intimal VW hyperplasia, p < 0.001 (Fig 5). The rises in VW 13-HODE and the inhibition of VW hyperplasia were considerably greater than could be achieved by attempting to raise
13-HODE levels indirectly, for example by giving its precursor linoleic acid or by giving Persantine. For comparison, when patients ingest 3.2 gm of linoleic acid daily (20 capsules) for 30 days, VW 13-HODE levels only increases 2-fold (Buchanan MR, Brister SJ. 1994), and when rabbits were treated with Persantine in a dose comparable to that used clinically, VW 13-HODE only increased 30 to 50 % (Weber E et al. 1990). Platelet/injured VW interactions were decreased > 2-fold in both cases.

These recent data demonstrate that the effect; i.e. inhibition of intimal hyperplasia, is achieved with an amazingly low dose of 13-HODE, and because hyperplasia regresses. No other current antithrombotic treatment has this capability. This effect is achieved without impairing platelet function and coagulation, as based on earlier studies (Weber E et al. 1990; Bertomeu M-C et al. 1990; Brister et al. 1990), and the observation that there was no detectable hemorrhagic defect in any of the experimental animals.

*Experimental Conclusions:* The following conclusions can be made from these experiments:

1) 13-HODE prevents VW hyperplasia effective when administered orally since the 13-HODE will reach the vascular tissue and other tissues in concentrations, which are biologically, active and which restore VW biocompatibility.

2) The doses of 13-HODE, which are biologically active, are surprisingly low.

3) Triglyceride and ester oils are appropriate vehicles for the 13-HODE.

4) The biological effects of 13-HODE on the vessel wall are highly desirable. These include both the prevention and treatment of vessel wall hyperplasia, as well as the facilitation of vessel wall disease regression.
Example 3

13-HODE is used in a ratio of between 1:3 and 1:100 or even up to 1:1000 with triglyceride oil, particularly corn oil. For example, 50 mg of 13-HODE could be mixed with 450 mg corn oil in a soft gelatin or bonded hard gelatin capsule, or 5 mg could be mixed with 100 mg of evening primrose oil or any other appropriate oil in similar types of capsules.

Example 4

Compositions as in Example 3 are used, but in which the oil is for direct administration as a liquid and is flavoured in an appropriate way, for example with lemon.

Example 5

Compositions as in Example 3 are used, but in which the oil is mixed with water to form a 5 to 40% oil in water emulsion, using an appropriate emulsifier and appropriate flavourings.

Example 6

Compositions as in Examples 3-5 are used, but in which the oil is mixed with an ester, particularly an ethyl ester of a 16-18 carbon fatty acid with one or more double bonds. Ethyl oleate, ethyl-linolate, ethyl-EPA and ethyl-DHA are examples of appropriate carriers for 13-HODE.

Example 7

As in Examples 3-6 but in which the 13-HODE is incorporated itself into a mono-, di- or tri-glyceride prior to mixing with the carrier.
As in Examples 3-6, but in which the 13-HODE is incorporated itself into the Sn1 or Sn2 positions of an appropriate phospholipid prior to mixing with the carrier.

Example 9

As in Examples 3-6, but in which the 13-HODE is incorporated into any other appropriate carrier molecule which will allow the 13-HODE to be delivered to target sites where 13-HODE is biologically active, such as the vascular endothelium.

Although preferred embodiments of the invention have been described herein, it will be understood by those skilled in the art that variations may be made thereto without departing from the spirit of the invention or the scope of the appended claims.
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The embodiments of the invention in which an exclusive property or privilege is claimed are defined as follows:

1. A method of reducing or inhibiting cell hyperlasia and restoring vessel wall biocompatibility in a mammal or human in need of such treatment, comprising administering orally an amount of 13-hydroxyoctadeca-9Z, 11E-dienoic acid (13-HODE) effective to reduce or inhibit vessel wall thrombogenicity.

2. The method of claim 1, wherein 13-HODE is administered as a pharmaceutical composition comprising 13-HODE and a pharmaceutically acceptable carrier, auxiliary, or excipient.

3. The method of claim 2, wherein the carrier is a mono-, di- or triglyceride oil.

4. The method of claim 2, wherein the carrier is selected from the group consisting of corn, sunflower, safflower, cottonseed, grape seed, olive, evening primrose, borage, fish body and fish liver oils.

5. The method of claim 2, wherein the carrier is an ester of a fatty acid containing 16-26 carbon atoms and one or more double bonds.

6. The method of claim 2, wherein the ester is selected from the group consisting of ethyl-eicosapentaenoic (ethyl-EPA), oleic, linoleic, alpha-linoleic, stearidonic, gamma-linolenic, dihomogammarinolenic, arachidonic, docosapentaenoic and docosahexaenoic (ethyl-DHA).

7. The method of claim 2, wherein the composition further comprises a fat-soluble antioxidant selected from the group consisting of ascorbyl palmitate, tocopherols, and ascorbic acid in the presence of lecithin.

8. The method of claim 2, wherein the composition further comprises an additive selected from the group consisting of aggregants, disaggregants, osmotic pressure regulating salts, buffers, sweeteners, and coloring agents.

9. The method of claim 2, wherein the composition is administered as a formulation selected from the group consisting of tablets, dragees, capsules, granules, solution, suspensions, and lyophilized compositions.
10. A method of correcting the inhibition of endogenous 13-HODE synthesis by omega-3 fatty acids by incorporating 13-HODE into formulations of omega-3 fatty acids.

11. The method of claim 1, wherein 13-HODE is administered as a pharmaceutical composition comprising 13-HODE and omega-3 fatty acids.

12. The method of claim 10 or 11, wherein the omega-3 fatty acid is selected from the group consisting of EPA, DHA, a derivative of EPA and a derivative of DHA.

13. The method of claim 10 or 11, wherein the omega-3 fatty acid is ethyl-EPA or ethyl-DHA.


15. A pharmaceutical composition of 13-hydroxyoctadeca-9Z, 11E-dienoic acid (13-HODE) for oral administration, comprising, 13 HODE and a pharmaceutically acceptable carrier.

16. The pharmaceutical composition of claim 14 or 15 wherein the daily dose of 13-HODE is equal to or less than 100 mg.

17. The pharmaceutical composition of claim 15, wherein the carrier is a mono-, di- or triglyceride oil.

18. The pharmaceutical composition of claim 15, wherein the carrier is selected from the group consisting of corn, sunflower, safflower, cottonseed, grape seed, olive, evening primrose, borage, fish body, and fish liver oils.

19. The pharmaceutical composition of claim 15, wherein the carrier is an ester of a fatty acid containing 16-26 carbon atoms and one or more double bonds.

20. The pharmaceutical composition of claim 15, wherein the carrier is selected from the group consisting of ethyl-eicosapentaenoic (ethyl-EPA), oleic, linoleic, alpha-linolenic, stearidonic, gamma-linolenic, dihomogammalinolenic, arachidonic, docosapentaenoic and docosahexaenoic (ethyl-DHA).

21. The pharmaceutical composition of claim 14 or 15, wherein the composition is administered in the form selected from the group consisting of tablets,
dragees, capsules, granules, solutions, suspensions and lyophilized compositions.

22. The pharmaceutical composition of claim 14 or 15 wherein the composition further comprises a fat-soluble antioxidant selected from the group consisting of ascorbyl palmitate, tocopherols, and ascorbic acid in the presence of lecithin.

23. The pharmaceutical composition of claim 14 or 15 wherein the composition further comprises an additive selected from the group consisting of aggregants, disaggregants, osmotic pressure regulating salts, buffers, sweeteners, and coloring agents.


25. The pharmaceutical composition of claim 24, wherein the omega-3 fatty acid is selected from the group consisting of EPA, DHA, a derivative of EPA and a derivative of DHA.

26. The pharmaceutical composition of claim 24, wherein the omega-3 fatty acid is selected from the group consisting of ethyl-EPA and ethyl-DHA.

27. The use of the pharmaceutical composition of claim 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25 or 26 to treat:
   (a) cardiovascular or cerebrovascular disease
   (b) inflammatory or autoimmune disease
   (c) infection with bacteria, viruses, fungi, or protozoa,
   (d) respiratory disease
   (e) gastrointestinal disease
   (f) renal or urinary tract disease
   (g) skin disease
   (h) neurological or psychiatric disease
   (i) disease of the reproductive system
   (j) diabetes, syndrome A or any complication of diabetes

28. The use of the pharmaceutical composition of claim 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25 or 26 to treat a disease or condition associated with overactive protein kinases.
29. The use of claim 28 wherein the disease or condition is associated with increase in Protein Kinase C activity and/or an increase in Mitogen Activated Protein Kinase activity.

30. The use of the pharmaceutical composition of claim 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25 or 26 to treat a disease or condition where endothelial function is disordered.

31. The use of the pharmaceutical composition of claim 14, 15, 16, 17, 18, 19, 20, 21, 22 or 23 to treat cancer or the metastatic spread of cancer.

32. The use of the pharmaceutical composition of claim 14, 15, 16, 17, 18, 19, 20, 21, 22 or 23 to prevent cancer or the metastatic spread of cancer.
FIGURE 2

2/6
SUBSTITUTE SHEET (RULE 26)
FIGURE 5

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SUBSTITUTE SHEET (RULE 26)
Proton nmr spectrum (270MHz; CDCl₃)

δ(PPM)  
6.5(1H, dd, H₁₁, J₁₁,₁₂=11Hz, J₁₁,₁₂=15.2Hz), 6.0(1H, t, H₁₀) 
J₁₀,₁₁=J₁₀,₁₂=11Hz), 5.7(1H, dd, H₁₂, J₁₂,₁₃=15.2Hz, J₁₂,₁₃=6.8Hz), 5.4(1H, dd, H₇, J₇,₈=7.7Hz, J₈,₉=10.8Hz), 4.1(1H, m, H₁₄), 2.4(2H, t, H₂) 
J₂₃=7.3Hz), 2.2(2H, m, H₃), 1.6(4H, m, H₅, H₆, H₇, H₈, H₉, H₁₀, H₁₁, H₁₂) and 0.9 (3H, t, H₁₃, J₁₃,₁₄=6.7Hz).

Carbon-13 nmr spectrum (67.8MHz, CDCl₃)

δ(ppm)  
179.3(C₁), 135.6(C₁₂), 132.6(C₁₃), 127.8(C₁₄), 125.8(C₁₁), 72.9(C₁₅), 37.1-22.4(C₁₇-C₁₆, C₁₈, C₁₉, C₂₀, C₁, C₁₀, C₂, C₃, C₄, C₅, C₆, C₇, C₈, C₉) and 13.9(C₁₀).

Infrared spectrum

3500-2500 cm⁻¹ (broad O-H stretch) and 1709 cm⁻¹ (C=O stretch)

Ultraviolet spectrum (ethanolic solution)

λₘₐₓ=232 nm (ε=25,000 mol⁻¹ cm⁻¹)

Soluble in ethanol, dichloromethane

Insoluble in hexane, water

FIGURE 6

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