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(71) Demandeur/Applicant:
CYTOMX THERAPEUTICS, LLC, US

(72) Inventeurs/Inventors:
STAGLIANO, NANCY E., US;
WEST, JAMES W., US;
KAMATH, KATHRYN, US;
BESSETTE, PAUL H., US;
GLUCK, FRED, US;
SAGERT, JASON, US;
DAUGHERTY, PATRICK, US

(74) Agent: RIDOUT & MAYBEE LLP

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(54) Title: MODIFIED ANTIBODY COMPOSITIONS, METHODS OF MAKING AND USING THEREOF

(57) **Abrégé/Abstract:**

The present disclosure provides modified antibodies which contain an antibody or antibody fragment (AB) modified with a masking moiety (MM). Such modified antibodies can be further coupled to a cleavable moiety (CM), resulting in activatable antibodies (AAs), wherein the CM is capable of being cleaved, reduced, photolysed, or otherwise modified. AAs can exhibit an activatable conformation such that the AB is more accessible to a target after, for example, removal of the MM by cleavage, reduction, or photolysis of the CM in the presence of an agent capable of cleaving, reducing, or photo lysing the CM. The disclosure further provides methods of making and using such modified antibodies and activatable antibodies.



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(71) Applicant (*for all designated States except US*): **CYTOMX THERAPEUTICS, LLC** [US/US]; 460 Ward Drive, Suite E1, Santa Barbara, CA 93111 (US).

(72) Inventors; and

(75) Inventors/Applicants (*for US only*): **STAGLIANO, Nancy, E.** [US/US]; 647 Por La Mar Circle, Unit C, Santa Barbara, CA (US). **WEST, James, W.** [US/US]; 748 Cieneguitas Rd Apt. A, Santa Barbara, CA 93110 (US). **KAMATH, Kathryn** [US/US]; 3230 Braemar Drive, Santa Barbara, CA 93109 (US). **BESSETTE, Paul, H.** [US/US]; 441 Avocado Place, Camarillo, CA 93010 (US). **GLUCK, Fred** [US/US]; 743 San Ysidro Road, Santa Barbara, CA 93108 (US). **SAGERT, Jason** [US/US]; 966 Cieneguitas Road, Santa Barbara, CA 93110 (US). **DAUGHERTY, Patrick** [US/US]; 917 N. Patterson Avenue, Santa Barbara, CA 93111 (US).

(74) Agents: **ROY, Madhuri** et al.; Wilson Sonsini Goodrich & Rosati, 650 Page Mill Road, Palo Alto, CA 94304-1050 (US).(81) Designated States (*unless otherwise indicated, for every kind of national protection available*): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PE, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.(84) Designated States (*unless otherwise indicated, for every kind of regional protection available*): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

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(54) Title: MODIFIED ANTIBODY COMPOSITIONS, METHODS OF MAKING AND USING THEREOF

(57) Abstract: The present disclosure provides modified antibodies which contain an antibody or antibody fragment (AB) modified with a masking moiety (MM). Such modified antibodies can be further coupled to a cleavable moiety (CM), resulting in activatable antibodies (AAs), wherein the CM is capable of being cleaved, reduced, photolysed, or otherwise modified. AAs can exhibit an activatable conformation such that the AB is more accessible to a target after, for example, removal of the MM by cleavage, reduction, or photolysis of the CM in the presence of an agent capable of cleaving, reducing, or photolysing the CM. The disclosure further provides methods of making and using such modified antibodies and activatable antibodies.



WO 2010/081173 A3

MODIFIED ANTIBODY COMPOSITIONS, METHODS OF MAKING AND USING THEREOF**CROSS-REFERENCE**

[0001] This application claims the benefit of U.S. Provisional Applications Nos. 61/144,110, filed January 12, 2009; 61/144,105, filed January 12, 2009; 61/249,441, filed October 7, 2009; and 61/249,416, filed October 7, 2009; which applications are incorporated herein by reference in their entirety.

BACKGROUND OF THE INVENTION

[0002] Protein-based therapies have changed the face of medicine, finding application in a variety of different diseases. In particular antibody-based therapies have proven effective treatments for some diseases but in some cases, toxicities due to broad target expression have limited their therapeutic effectiveness.

[0003] As with any drugs, however, the need and desire for drugs having improved specificity and selectivity for their targets is of great interest, especially in developing second generation of antibody-based drugs having known targets to which they bind. Increased targeting of antibody to the disease site could reduce systemic mechanism-based toxicities and lead to broader therapeutic utility.

[0004] In the realm of small molecule drugs, strategies have been developed to provide prodrugs of an active chemical entity. Such prodrugs are administered in a relatively inactive (or significantly less active) form. Once administered, the prodrug is metabolized *in vivo* into the active compound. Such prodrug strategies can provide for increased selectivity of the drug for its intended target and for a reduction of adverse effects. Drugs used to target hypoxic cancer cells, through the use of redox-activation, utilize the large quantities of reductase enzyme present in the hypoxic cell to convert the drug into its cytotoxic form, essentially activating it. Since the prodrug has low cytotoxicity prior to this activation, there is a markedly decreased risk of damage to non-cancerous cells, thereby providing for reduced side-effects associated with the drug. There is a need in the field for a strategy for providing features of a prodrug to antibody-based therapeutics.

SUMMARY OF THE INVENTION

[0005] The present disclosure provides for modified and activatable antibody compositions useful for therapeutics and diagnostics. The activatable antibody compositions exhibit increased bioavailability and biodistribution compared to conventional antibody therapeutics with prodrug features. Also provided are methods for use in diagnostics and therapeutics, as well as screening for and construction of such compositions.

[0006] In one aspect, the present disclosure provides a modified antibody comprising an antibody or antibody fragment (AB), capable of specifically binding its target, coupled to a masking moiety (MM), wherein the coupling of the MM reduces the ability of the AB to bind its target such that the dissociation constant (K_d) of the AB coupled to the MM towards the target is at least 100 times greater, at

least 1000 times greater, or at least 10,000 times greater than the K_d of the AB not coupled to the MM towards the target.

[0007] In another aspect, the present disclosure provides a modified antibody comprising an antibody or antibody fragment (AB), capable of specifically binding its target, coupled to a masking moiety (MM), wherein the coupling of the MM to the AB reduces the ability of the AB to bind the target by at least 90%, as compared to the ability of the AB not coupled to the MM to bind the target, when assayed *in vitro* using a target displacement assay. Such coupling of the MM to the AB reduces the ability of the AB to bind its target for at least 12 hours or for at least 24 hours or for at least 72 hours.

[0008] In another aspect, the modified antibody is further coupled to a cleavable moiety (CM). The CM is capable of being cleaved by an enzyme, or the CM is capable of being reduced by a reducing agent, or the CM is capable of being photolysed. The CM is capable of being specifically cleaved, reduced, or photolysed at a rate of about at least $1 \times 10^4 \text{ M}^{-1}\text{S}^{-1}$, or at least $5 \times 10^4 \text{ M}^{-1}\text{S}$, or at least $10 \times 10^4 \text{ M}^{-1}\text{S}$. In one embodiment, the CM of the modified antibody is be within the MM.

[0009] The dissociation constant (K_d) of the MM towards the AB in the modified antibodies provided herein is usually at least 100 times greater than the K_d of the AB towards the target. Generally, the K_d of the MM towards the AB is lower than 10nM, or lower than 5nM, or about 1nM.

[0010] In some embodiments, the MM of the modified antibody reduces the AB's ability to bind its target by specifically binding to the antigen-binding domain of the AB. Such binding can be non-covalent. The MM of the modified antibody can reduce the AB's ability to bind its target allosterically or sterically. In specific embodiments, the MM of the modified antibody does not comprise more than 50% amino acid sequence similarity to a natural binding partner of the AB.

[0011] In specific embodiments, the AB of the modified antibody is an antibody fragment that is selected from the group consisting of a Fab' fragment, a F(ab')₂ fragment, a scFv, a scAB a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.

[0012] In related embodiments, the AB of the modified antibody is selected from the group consisting of the antibodies in Table 2 or specifically the source of the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab. In a specific embodiment, the modified antibody is not alemtuzumab.

[0013] In related embodiments, the target of the AB is selected from the group consisting of the targets in Table 1. In exemplary embodiments, the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4. In one specific embodiment the target is not CD52.

[0014] In a specific embodiment, the modified antibody further comprises a second AB wherein the target for the second AB is selected from the group consisting of the targets in Table 1.

[0015] In related embodiments, the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3. In specific embodiments the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA. In such embodiments, where the modified AB comprises a CM, the AB is selected from the group consisting of the antibodies in Table 2; and specifically can be from cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab. In one exemplary embodiment, the AB is not alemtuzumab.

[0016] In one embodiment where the modified antibody comprises an AB, coupled to a CM and a MM, the target is selected from the group consisting of the targets in Table 1; or the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4. In one exemplary embodiment, the target is not CD52.

[0017] The modified antibody can be further coupled to a second cleavable moiety (CM), capable of being specifically modified by an enzyme. In this embodiment, the second cleavable is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

[0018] In another specific embodiment, the modified antibody further comprises a linker peptide, wherein the linker peptide is positioned between the AB and the MM; or the modified antibody further comprises a linker peptide, wherein the linker peptide is positioned between the MM and the CM; or the modified antibody further comprises a linker peptide, wherein the linker peptide is positioned between the AB and the CM; or the modified antibody further comprises two linker peptides, wherein the first linker peptide is between the AB and the CM and the second linker peptide is positioned between the MM and the CM. The linker is selected from the group consisting of a cleavable linker, a non-cleavable linker, and a branched linker.

[0019] In certain embodiments, the modified antibody further comprises a detectable moiety. In one specific embodiment, the detectable moiety is a diagnostic agent.

[0020] In one particular embodiment, the modified antibodies described herein further comprise an agent conjugated to the AB. In one aspect, the agent is a therapeutic agent, for example an antineoplastic agent. In such embodiments, the agent is conjugated to a carbohydrate moiety of the AB, wherein the carbohydrate moiety can be located outside the antigen-binding region of the AB. Alternatively the agent is conjugated to a sulfhydryl group of the AB.

[0021] The modified antibodies provided herein exhibit a serum half-life of at least 5 days when administered to an organism.

[0022] The consensus sequence of the MM of some of the modified antibodies provided herein is CISPRGC, C(N/P)H(HVF)(Y/T)(F/W/T/L)(Y/G/T/S)(T/S/Y/H)CGCISPRGCG, xCxxYQCLxxxxxx,XXQPxPPRVXX, PxPGFPYCxxxx, xxxxQxxPWPP, GxGxCYTILExxCxxxR,

GxxxCYxIxExxCxxxx, GxxxCYxIxExWCxxxx, xxxCCxxYxIxxCCxxx, or xxxxxYxILExxxxxx. In a specific embodiment, the consensus sequence is specific for binding to an anti-VEGF antibody, an anti-EGFR antibody, or an anti-CTLA-4 antibody.

[0023] In a related aspect, the present disclosure provides for an activatable antibody (AA) comprising an antibody or antibody fragment (AB), capable of specifically binding its target; a masking moiety (MM) coupled to the AB, capable of inhibiting the specific binding of the AB to its target; and a cleavable moiety (CM) coupled to the AB, capable of being specifically cleaved by an enzyme; wherein when the AA is not in the presence of sufficient enzyme activity to cleave the CM, the MM reduces the specific binding of the AB to its target by at least 90% when compared to when the AA is in the presence of sufficient enzyme activity to cleave the CM and the MM does not inhibit the specific binding of the AB to its target. In specific embodiments, the binding of the AB to its target is reduced for at least 12 hours, or for at least 24 hours, or for at least 72 hours.

[0024] In one embodiment, in the AA, the dissociation constant (K_d) of the AB coupled to the MM and CM towards the target is at least 100 times greater than the K_d of the AB not coupled to the MM and CM towards the target. In a related embodiment, the dissociation constant (K_d) of the MM towards the AB is at least 100 times greater than the K_d of the AB towards the target. Generally, the K_d of the MM towards the AB is lower than 10nM, or lower than 5nM, or about 1nM.

[0025] In some embodiments of the AA, the MM is capable of specifically binding to the antigen-binding domain of the AB.

[0026] In some embodiments of the AA the CM is capable of being specifically cleaved by an enzyme at a rate of about at least $1 \times 10^4 \text{ M}^{-1}\text{S}^{-1}$, or at least $5 \times 10^4 \text{ M}^{-1}\text{S}$, or at least $10 \times 10^4 \text{ M}^{-1}\text{S}$.

[0027] In certain embodiments, of the AA where the AB is an antibody fragment, the antibody fragment is selected from the group consisting of a Fab' fragment, a F(ab')₂ fragment, a scFv, a scAB, a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.

[0028] In certain embodiments, the AB of the AA is selected from the group consisting of the antibodies in Table 2. In specific embodiments, the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

[0029] In certain embodiments, the target of the AA is selected from the group consisting of the targets in Table 1. In specific embodiments, the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

[0030] In one specific embodiment the AB is not alemtuzumab and target is not CD52.

[0031] In certain embodiments, the CM of the AA is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA. In specific embodiments, the AA is further coupled to a second cleavable moiety (CM), capable of being specifically modified by an enzyme. In this embodiment, the second CM is a substrate for legumain,

plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

[0032] In some embodiments of the AAs provided herein, the CM is located within the MM.

[0033] In some embodiments of the AAs provided herein, the MM does not comprise more than 50% amino acid sequence similarity to a natural binding partner of the AB.

[0034] In some embodiments the AA further comprises a linker peptide, wherein the linker peptide is positioned between the MM and the CM. In specific embodiments, the linker peptide is positioned between the AB and the CM.

[0035] In certain embodiments, the AAs provided herein further comprise a detectable moiety or an agent conjugated to the AB.

[0036] In yet another aspect, the present disclosure provides for an activatable antibody complex (AAC) comprising: two antibodies or antibody fragments (AB1 and AB2), each capable of specifically binding its target; at least one masking moiety (MM) coupled to either AB1 or AB2, capable of inhibiting the specific binding of AB1 and AB2 to their targets; and at least one cleavable moiety (CM) coupled to either AB1 or AB2, capable of being specifically cleaved by an enzyme whereby activating the AAC composition; wherein when the AAC is in an uncleaved state, the MM inhibits the specific binding of AB1 and AB2 to their targets and when the AAC is in a cleaved state, the MM does not inhibit the specific binding of AB1 and AB2 to their targets.

[0037] In one embodiment, the AAC is bispecific, wherein AB1 and AB2 bind the same epitope on the same target; or the AB1 and AB2 bind to different epitopes on the same target; or the AB1 and AB2 bind to different epitopes on different targets.

[0038] In one embodiment of the AAC, the CM is capable of being specifically cleaved by an enzyme at a rate of about at least $1 \times 10^4 \text{ M}^{-1}\text{S}^{-1}$.

[0039] In the embodiments where AB1 or AB2 of the AAC is an antibody fragment, the antibody fragment is selected from the group consisting of a Fab' fragment, a F(ab')₂ fragment, a scFv, a scAB, a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.

[0040] In an embodiment of the AAC, the AB1 and/or AB 2 are selected from the group consisting of the antibodies in Table 2. In a specific embodiment, the AB1 and/or AB2 is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

[0041] In an embodiment of the AAC, the target for the AB1 and/or AB2 is selected from the group consisting of the targets in Table 1. In a related embodiment, the target of the AB1 and/or AB2 is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4. In a specific embodiment, the AB1 and AB2 are capable of binding to EGFR and VEGF, a Notch Receptor and EGFR, a Jagged ligand and EGFR or cMET and VEGF, respectively.

[0042] In a related AAC embodiment, the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3. In a specific embodiment, the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA. In yet another specific embodiment, the AAC is further coupled to a second cleavable moiety (CM), capable of being specifically cleaved by an enzyme and the second CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

[0043] In specific embodiments of the AAC, the MM does not comprise more than 50% amino acid sequence similarity to a natural binding partner of the AB.

[0044] In other specific embodiments of the AAC, the AAC further comprises a detectable moiety or is further conjugated to an agent.

[0045] Also provided herein is a method of treating or diagnosing a condition in a subject including administering to the subject a composition comprising: an antibody or antibody fragment (AB), capable of specifically binding its target; a masking moiety (MM) coupled to the AB, capable of inhibiting the specific binding of the AB to its target; and a cleavable moiety (CM) coupled to the AB, capable of being specifically cleaved by an enzyme; wherein upon administration to the subject, when the AA is not in the presence of sufficient enzyme activity to cleave the CM, the MM reduces the specific binding of the AB to its target by at least 90% when compared to when the AA is in the presence of sufficient enzyme activity to cleave the CM and the MM does not inhibit the specific binding of the AB to its target.

[0046] In this method, the AB is selected from the group consisting of a Fab' fragment, a F(ab')₂ fragment, a scFv, a scAB, a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.

[0047] In a specific embodiment, the condition is cancer.

[0048] In another specific embodiment, the MM is not the natural binding partner of the AB.

[0049] In various embodiments of the method, the AB is selected from the group consisting of the antibodies in Table 2. Specifically in some embodiments, the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

[0050] In various embodiments of the method, the target is selected from the group of targets in Table 1. In specific embodiments, the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

[0051] In a very specific embodiment of the method the AB is not alemtuzumab and the target is not CD52.

[0052] In various embodiments of the method, the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3. In specific embodiments, the CM is a substrate for legumain,

plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

[0053] Also provided herein is a method of inhibiting angiogenesis in a mammalian subject, the method comprising administering to a subject in need thereof a therapeutically effective amount of a pharmaceutical composition comprising a modified AB, an AA, an AAC, or an AACJ wherein the target is EGFR, TNF α , CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4. In a specific embodiment, the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab; the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

[0054] Also provided herein is a method of making an activatable antibody (AA) composition comprising: providing an antibody or antibody fragment (AB) capable of specifically binding its target; coupling a masking moiety (MM) to the AB, capable of inhibiting the specific binding of the AB to its target; and coupling a cleavable moiety (CM) to the AB, capable of being specifically cleaved by an enzyme; wherein the dissociation constant (K_d) of the AB coupled to the MM towards the target is at least 100 times greater than the K_d of the AB not coupled to the MM towards the target.

[0055] In one embodiment of the method, the AB is or is derived from an antibody selected from the group consisting of the antibodies in Table 2. In a specific embodiment, the AB is or is derived from cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

[0056] In one very specific embodiment, the AB is not alemtuzumab and the target is not CD52.

[0057] In another embodiment of the method, the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3. In a specific embodiment, the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

[0058] Also provided herein is a method of screening candidate peptides to identify a masking moiety (MM) peptide capable of binding an antibody or antibody fragment (AB) comprising: providing a library of peptide scaffolds, wherein each peptide scaffold comprises: a transmembrane protein (TM); and a candidate peptide; contacting an AB with the library; identifying at least one candidate peptide capable of binding the AB; and determining whether the dissociation constant (K_d) of the candidate peptide towards the AB is between 1-10 nM.

[0059] In various embodiments of the method, the library comprises viruses, cells or spores. Specifically in one embodiment, the library comprises E. coli. In another embodiment, the peptide scaffold further comprises a detectable moiety.

[0060] Also provided is another screening method to identify a masking moiety (MM) peptide capable of masking an antibody or antibody fragment (AB) with an optimal masking efficiency comprising: providing a library comprising a plurality of ABs, each coupled to a candidate peptide, wherein the ABs are capable of specifically binding a target; incubating each library member with the target; and comparing the binding affinity of each library member towards the target with the binding affinity of each AB not coupled to a candidate peptide towards the target. In a specific embodiment, the optimal binding efficiency is when the binding affinity of a library member to the target is 10% compared to the binding affinity of the unmodified AB to the target.

[0061] In one aspect, also provided herein is an antibody therapeutic having an improved bioavailability wherein the affinity of binding of the antibody therapeutic to its target is lower in a first tissue when compared to the binding of the antibody therapeutic to its target in a second tissue. In a related aspect, also provided herein is a pharmaceutical composition comprising: an antibody or antibody fragment (AB), capable of specifically binding its target; and a pharmaceutically acceptable excipient; wherein the affinity of the antibody or antibody fragment to the target in a first tissue is lower than the affinity of the antibody or antibody fragment to the target in a second tissue. In a specific embodiment, the affinity in the first tissue is 10-1,000 times lower than the affinity in the second tissue. In one embodiment, the AB is coupled to a masking moiety (MM), capable of reducing the binding of the AB to its target and a cleavable moiety (CM), capable of specifically being cleaved by an enzyme.

[0062] In related embodiments, the target is EGFR, TNF α , CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4. In related embodiments, the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA. In related embodiments, the antibody or antibody fragment is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

[0063] In a specific embodiment, the first tissue is a healthy tissue and the second tissue is a diseased tissue; or the first tissue is an early stage tumor and the second tissue is a late stage tumor; the first tissue is a benign tumor and the second tissue is a malignant tumor; or the first tissue and second tissue are spatially separated; or the first tissue is epithelial tissue and the second tissue is breast, head, neck, lung, pancreatic, nervous system, liver, prostate, urogenital, or cervical tissue.

[0064] In one embodiment, the antibody therapeutic is further coupled to an agent. In a specific embodiment, the the agent is an antineoplastic agent.

[0065] Also provided herein are specific compositions for diagnostic and therapeutic use. Provided herein is a composition comprising a legumain-activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a plasmin-activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a caspase-activatable

antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a TMPRSS-3/4 -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a PSA -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a cathepsin -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a human neutrophil elastase -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a beta-secretase -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an uPA -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a TMPRSS-3/4 -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising a MT1-MMP-activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable EGFR antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable TNFalpha antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable CD11a antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable CSFR antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable CTLA-4 antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable EpCAM antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable CD40L antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable Notch1 antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable Notch3 antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable Jagged1 antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable Jagged2 antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable cetuximab antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable vectibix antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable infliximab antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable adalimumab antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable efalizumab antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable ipilimumab antibody or antibody fragment (AB) coupled to a masking moiety (MM); a composition comprising an activatable tremelimumab antibody or antibody fragment (AB) coupled to a masking moiety (MM); or a composition comprising an activatable adecatumumab antibody or antibody fragment (AB) coupled to a masking moiety (MM).

INCORPORATION BY REFERENCE

[0066] All publications, patents, and patent applications mentioned in this specification are herein incorporated by reference to the same extent as if each individual publication, patent, or patent application was specifically and individually indicated to be incorporated by reference.

BRIEF DESCRIPTION OF THE DRAWINGS

[0067] The novel features of the invention are set forth with particularity in the appended claims. A better understanding of the features and advantages of the present invention will be obtained by reference to the following detailed description that sets forth illustrative embodiments, in which the principles of the invention are utilized, and the accompanying drawings of which:

[0068] **Figure 1** shows a protease-activated AA containing an antibody (an AB), a masking moiety (MM), and a cleavable moiety (CM).

[0069] **Figure 2** shows the activity of an exemplary AA *in vivo*. Panel A shows healthy tissue where the AA is not able to bind, side effects are minimal; Panel B shows diseased tissue where the AA is activated by a disease-specific protease/reducing agent allowing the AA to bind to target and be efficacious.

[0070] **Figure 3** illustrates a process to produce a protease-activated AA, involving: screening for MMs; screening for CMs; assembling the MM, CM, and an AB; expressing and purifying the assembled construct; and assaying the assembled construct for activity and toxicity *in vitro* and *in vivo*.

[0071] **Figure 4** provides an exemplary MMP-9 cleavable masked anti-VEGF scFv amino acid sequence.

[0072] **Figure 5** provides ELISA data showing the MMP-9 activation of the MBP:anti-VEGFscFv AAs with the MMs 306 and 314. Samples were treated with TEV to remove the MBP fusion partner and subsequently activated by MMP-9 digestion.

[0073] **Figure 6** provides ELISA data demonstrating the MMP-9-dependent VEGF binding of the anti-VEGFscFv His construct with the 306 MM.

[0074] **Figure 7** provides ELISA data demonstrating the MMP-9-dependent VEGF binding of anti-VEGFscFv-Fc AAs with the MMs 306 and 314 from HEK cell supernatants.

[0075] **Figure 8** provides ELISA data showing the MMP-9-dependent VEGF binding of anti-VEGF scFv-Fc AA constructs with the MMs 306 and 314 that were purified using a Protein A column.

[0076] **Figure 9** shows that the 306 MM, which binds to an anti-VEGF antibody with an affinity of > 600 nM, does not efficiently preclude binding to VEGF.

[0077] **Figure 10** shows light and heavy chains of anti-CTLA4 joined via SOE-PCR to generate scFv constructs in both orientations, V_HV_L and V_LV_H.

[0078] **Figure 11** shows the use of PCR to add sites for MM cloning, CM cleavage sequence, GGS2 linker on the N-terminus of the anti-CTLA4 scFv V_HV_L and V_LV_H constructs.

[0079] **Figure 12** shows the activation of an AA by MMP-9.

[0080] **Figure 13** shows that when the CM is cleaved to remove the MM, the binding of the AB is restored.

[0081] **Figure 14** shows the activation of an AA by a protease that leads to antibody binding indistinguishable from unmodified antibodies.

[0082] **Figure 15** illustrates that an AA comprising an AB with specific binding affinity to VEGF is inhibited; the activated AA binds VEGF with picomolar affinity.

[0083] **Figure 16** depicts that an AA comprising an AB with specific binding affinity to VEGF inhibits HUVEC proliferation.

[0084] **Figure 17** illustrates that cultured tumor cells demonstrate robust *in situ* activation of an AA comprising an AB with specific binding affinity to VEGF.

[0085] **Figure 18** illustrates that an AA is inactive in normal and cancer patient plasma

[0086] **Figure 19** illustrates the binding of anti-CTLA4 scFv to both murine and human CTLA4.

[0087] **Figure 20** shows a protease-activated AACJ-containing an antibody (containing an AB), a masking moiety (MM), a cleavable moiety (CM), and a conjugated agent. Upon cleavage of the CM and unmasking, the conjugated AB is released.

[0088] **Figure 21** shows that binding of the eCPX3.0 clones JS306, JS1825, JS1827, and JS1829 were analyzed on FACS at 3 different concentrations of DyLight labeled anti-VEGF. All three of the affinity matured peptides displayed at least 10 fold higher affinity than the JS306.

[0089] **Figure 22** shows the process for affinity maturation of some of the EGFR MM's.

[0090] **Figure 23** shows the binding curves for the on-cell affinity measurement of C225 Fab binding to MM's 3690, 3954 and 3957. MMs 3954 and 3957 displayed at least 100 fold higher affinity than 3690.

[0091] **Figure 24** displays the Target Displacement Assay and extent of equilibrium binding as a percent of parental antibody binding.

[0092] **Figure 25** shows that unlike the uPA control and substrate SM16, KK1203, 1204 and 1214 show resistance to cleavage by KLK5, KLK7 and Plasmin.

[0093] **Figure 26** shows that unlike a non-optimized substrate, the optimized substrates Plas1237, Plas129 and Plas 1254 show resistance to cleavage by KLK5, KLK7.

[0094] **Figure 27** Panel A shows activation of ScFv AAs containing legumain substrates AANL and PTNL following treatment with 5 mg/mL legumain. Panel B shows activation of an anti-VEGF IgG AA containing the legumain substrate PNTL.

[0095] **Figure 28** shows the ratio of activated AA to total AA at each time point in a legumain-activated AA. While the plasmin-activated AA is nearly completely activated at 7 days, both legumain- activatable AAs are only minimally activated. Legumain-activatable AAs isolated from serum up to 7 days following injection remain masked. (n=4).

[0096] **Figure 29** shows that masked single-chain Fv-Fc fusion pro-antibodies exhibit increased serum half-life.

[0097] **Figure 30** shows that the scFv-Fc serum concentration in healthy mice over 10 days. The AA concentration remained stable 7 days post dose, whereas the parent scFv-Fc concentration decreased after 3 days and was almost undetectable at 10 days.

[0098] **Figure 31** shows that AA scFv-Fc concentrations are elevated and persist longer in serum compared with parent scFv-Fc in tumor-bearing mice. A higher percentage of the initial AA dose was detected in the serum at 3 days (B) and 3 and 7 days (A).

[0099] **Figure 32** shows that AA scFv-Fcs persist at higher concentrations in a multidose study in Tumor-bearing mice. AAs maintained significantly higher serum concentrations than the parent throughout the study.

[00100] **Figure 33** shows that AAs persist at high levels in serum of normal mice as compared to the parental antibody not modified with a MM.

[00101] **Figure 34** shows protease-activated activatable antibody complexes (AACs) containing one or more antibodies or fragments thereof (in this figure the ABs are referred to as ABDs), a masking moiety (MM), and a cleavable moiety (CM), where ABD1 and ABD2 are arbitrary designations for first and second ABs. In such embodiments, the MM1 and MM2 bind the domains containing ABD1 and ABD2, respectively, and act as masking moieties to interfere with target binding to an uncleaved dual target-binding AAC. The target capable of binding the ABs may be the same or different target, or different binding sites of the same target. In some embodiments (Figures 1A, 1D, 1F), binding of MM1 to the domain containing ABD2 on the opposite molecule forms the complex capable of acting as a masking moiety of ABD1 and ABD2.

[00102] **Figure 35** shows an AAC with cross-masking occurring such that target binding by both ABs is attenuated in the uncleaved state, and target binding is increased in the presence of an agent that cleaves the CM allowing the complex to disassemble. In this figure the AB1 and AB2 are referred to as the ABD1 and ABD2, respectively.

[00103] **Figure 36** shows an AAC formed by covalent linkage of MM1 with ABD1 (AB1) such that target binding by ABD2 (AB2) is attenuated in the uncleaved state, and target binding by ABD2 (AB2) is increased in the presence of an agent that cleaves the CM allowing the complex to disassemble. In this figure the AB1 and AB2 are referred to as the ABD1 and ABD2, respectively.

DETAILED DESCRIPTION OF THE INVENTION

[00104] The present disclosure provides modified antibody compositions and that are useful for therapeutics and diagnostics. The compositions described herein allow for greater biodistribution and improved bioavailability.

Modified and Activatable Antibodies

[00105] The modified antibody compositions described herein contain at least an antibody or antibody fragment thereof (collectively referred to as AB throughout the disclosure), capable of specifically binding a target, wherein the AB is modified by a masking moiety (MM).

[00106] When the AB is modified with a MM and is in the presence of the target, specific binding of the AB to its target is reduced or inhibited, as compared to the specific binding of the AB not modified with an MM or the specific binding of the parental AB to the target.

[00107] The K_d of the AB modified with a MM towards the target can be at least 5, 10, 25, 50, 100, 250, 500, 1,000, 2,500, 5,000, 10,000, 50,000, 100,000, 500,000, 1,000,000, 5,000,000, 10,000,000, 50,000,000 or greater, or between 5-10, 10-100, 10-1,000, 10-10,000, 10-100,000, 10-1,000,000, 10-10,000,000, 100-1,000, 100-10,000, 100-100,000, 100-1,000,000, 100-10,000,000, 1,000-10,000, 1,000-100,000, 1,000-1,000,000, 1000-10,000,000, 10,000-100,000, 10,000-1,000,000, 10,000-10,000,000, 100,000-1,000,000, or 100,000-10,000,000 times greater than the K_d of the AB not modified with an MM or the parental AB towards the target. Conversely, the binding affinity of the AB modified with a MM towards the target can be at least 5, 10, 25, 50, 100, 250, 500, 1,000, 2,500, 5,000, 10,000, 50,000, 100,000, 500,000, 1,000,000, 5,000,000, 10,000,000, 50,000,000 or greater, or between 5-10, 10-100, 10-1,000, 10-10,000, 10-100,000, 10-1,000,000, 10-10,000,000, 100-1,000, 100-10,000, 100-100,000, 100-1,000,000, 100-10,000,000, 1,000-10,000, 1,000-100,000, 1,000-1,000,000, 1000-10,000,000, 10,000-100,000, 10,000-1,000,000, 10,000-10,000,000, 100,000-1,000,000, or 100,000-10,000,000 times lower than the binding affinity of the AB not modified with an MM or the parental AB towards the target.

[00108] The dissociation constant (K_d) of the MM towards the AB is generally greater than the K_d of the AB towards the target. The K_d of the MM towards the AB can be at least 5, 10, 25, 50, 100, 250, 500, 1,000, 2,500, 5,000, 10,000, 100,000, 1,000,000 or even 10,000,000 times greater than the K_d of the AB towards the target. Conversely, the binding affinity of the MM towards the AB is generally lower than the binding affinity of the AB towards the target. The binding affinity of MM towards the AB can be at least 5, 10, 25, 50, 100, 250, 500, 1,000, 2,500, 5,000, 10,000, 100,000, 1,000,000 or even 10,000,000 times lower than the binding affinity of the AB towards the target.

[00109] When the AB is modified with a MM and is in the presence of the target, specific binding of the AB to its target can be reduced or inhibited, as compared to the specific binding of the AB not modified with an MM or the specific binding of the parental AB to the target. When compared to the binding of the AB not modified with an MM or the binding of the parental AB to the target, the AB's ability to bind the target when modified with an MM can be reduced by at least 50%, 60%, 70%, 80%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% and even 100% for at least 2, 4, 6, 8, 12, 28, 24, 30, 36, 48, 60, 72, 84, 96, hours, or 5, 10, 15, 30, 45, 60, 90, 120, 150, 180 days, or 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 months or greater when measured *in vivo* or in a Target Displacement *in vitro* immunoabsorbant assay, as described herein.

[00110] The MM can inhibit the binding of the AB to the target. The MM can bind the antigen binding domain of the AB and inhibit binding of the AB to its target. The MM can sterically inhibit the binding of the AB to the target. The MM can allosterically inhibit the binding of the AB to its target. In these embodiments when the AB is modified or coupled to a MM and in the presence of target, there is no binding or substantially no binding of the AB to the target, or no more than .001%, .01%, .1%, 1%, 2%,

3%, 4%, 5%, 6%, 7%, 8%, 9%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, or 50% binding of the AB to the target, as compared to the binding of the AB not modified with an MM, the parental AB, or the AB not coupled to an MM to the target, for at least 2, 4, 6, 8, 12, 28, 24, 30, 36, 48, 60, 72, 84, 96, hours, or 5, 10, 15, 30, 45, 60, 90, 120, 150, 180 days, or 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 months or greater when measured *in vivo* or in a Target Displacement *in vitro* immunoabsorbant assay, as described herein.

[00111] When an AB is coupled to or modified by a MM, the MM can ‘mask’ or reduce, or inhibit the specific binding of the AB to its target. When an AB is coupled to or modified by a MM, such coupling or modification can effect a structural change which reduces or inhibits the ability of the AB to specifically bind its target.

[00112] An AB coupled to or modified with an MM can be represented by the following formulae (in order from an amino (N) terminal region to carboxyl (C) terminal region:

(MM)-(AB)
 (AB)-(MM)
 (MM)-L-(AB)
 (AB)-L-(MM)

where MM is a masking moiety, the AB is an antibody or antibody fragment thereof, and the L is a linker. In many embodiments it may be desirable to insert one or more linkers, e.g., flexible linkers, into the composition so as to provide for flexibility.

[00113] In certain embodiments the MM is not a natural binding partner of the AB. The MM may be a modified binding partner for the AB which contains amino acid changes that at least slightly decrease affinity and/or avidity of binding to the AB. In some embodiments the MM contains no or substantially no homology to the AB’s natural binding partner. In other embodiments the MM is no more than 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, or 80% similar to the natural binding partner of the AB.

[00114] The present disclosure also provides activatable antibodies (AAs) where the AB modified by an MM can further include one or more cleavable moieties (CM). Such AAs exhibit activatable/switchable binding, to the AB’s target. AAs generally include an antibody or antibody fragment (AB), modified by or coupled to a masking moiety (MM) and a modifiable or cleavable moiety (CM). In some embodiments, the CM contains an amino acid sequence that serves as a substrate for a protease of interest. In other embodiments, the CM provides a cysteine-cysteine disulfide bond that is cleavable by reduction. In yet other embodiments the CM provides a photolytic substrate that is activatable by photolysis.

[00115] A schematic of an exemplary AA is provided in Figure 1. As illustrated, the elements of the AA are arranged so that the CM is positioned such that in a cleaved (or relatively active state) and in the presence of a target, the AB binds a target, while in an uncleaved (or relatively inactive state) in the presence of the target, specific binding of the AB to its target is reduced or inhibited. The specific

binding of the AB to its target can be reduced due to the due to the inhibition or masking of the AB's ability to specifically bind its target by the MM.

[00116] The K_d of the AB modified with a MM and a CM towards the target can be at least 5, 10, 25, 50, 100, 250, 500, 1,000, 2,500, 5,000, 10,000, 50,000, 100,000, 500,000, 1,000,000, 5,000,000, 10,000,000, 50,000,000 or greater, or between 5-10, 10-100, 10-1,000, 10-10,000, 10-100,000, 10-1,000,000, 10-10,000,000, 100-1,000, 100-10,000, 100-100,000, 100-1,000,000, 100-10,000,000, 1,000-10,000, 1,000-100,000, 1,000-1,000,000, 1000-10,000,000, 10,000-100,000, 10,000-1,000,000, 10,000-10,000,000, 100,000-1,000,000, or 100,000-10,000,000 times greater than the K_d of the AB not modified with an MM and a CM or the parental AB towards the target. Conversely, the binding affinity of the AB modified with a MM and a CM towards the target can be at least 5, 10, 25, 50, 100, 250, 500, 1,000, 2,500, 5,000, 10,000, 50,000, 100,000, 500,000, 1,000,000, 5,000,000, 10,000,000, 50,000,000 or greater, or between 5-10, 10-100, 10-1,000, 10-10,000, 10-100,000, 10-1,000,000, 10-10,000,000, 100-1,000, 100-10,000, 100-100,000, 100-1,000,000, 100-10,000,000, 1,000-10,000, 1,000-100,000, 1,000-1,000,000, 1000-10,000,000, 10,000-100,000, 10,000-1,000,000, 10,000-10,000,000, 100,000-1,000,000, or 100,000-10,000,000 times lower than the binding affinity of the AB not modified with an MM and a CM or the parental AB towards the target.

[00117] When the AB is modified with a MM and a CM and is in the presence of the target but not in the presence of a modifying agent (for example an enzyme, protease, reduction agent, light), specific binding of the AB to its target can be reduced or inhibited, as compared to the specific binding of the AB not modified with an MM and a CM or the parental AB to the target. When compared to the binding of the parental AB or the binding of an AB not modified with an MM and a CM to its target, the AB's ability to bind the target when modified with an MM and a CM can be reduced by at least 50%, 60%, 70%, 80%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% and even 100% for at least 2, 4, 6, 8, 12, 28, 24, 30, 36, 48, 60, 72, 84, 96 hours, or 5, 10, 15, 30, 45, 60, 90, 120, 150, 180 days, or 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 months or greater when measured *in vivo* or in a Target Displacement *in vitro* immunoabsorbant assay, as described herein.

[00118] As used herein, the term cleaved state refers to the condition of the AA following modification of the CM by a protease and/or reduction of a cysteine-cysteine disulfide, bond of the CM, and/or photoactivation. The term uncleaved state, as used herein, refers to the condition of the AA in the absence of cleavage of the CM by a protease and/or in the absence reduction of a cysteine-cysteine disulfide bond of the CM, and/or in the absence of light. As discussed above, the term AA is used herein to refer to an AA in both its uncleaved (native) state, as well as in its cleaved state. It will be apparent to the ordinarily skilled artisan that in some embodiments a cleaved AA may lack an MM due to cleavage of the CM by protease, resulting in release of at least the MM (e.g., where the MM is not joined to the AA by a covalent bond (e.g., a disulfide bond between cysteine residues).

[00119] By activatable or switchable is meant that the AA exhibits a first level of binding to a target when in a inhibited, masked or uncleaved state (i.e., a first conformation), and a second level of binding to the

target in the uninhibited, unmasked and/or cleaved state (i.e., a second conformation), where the second level of target binding is greater than the first level of binding. In general, the access of target to the AB of the AA is greater in the presence of a cleaving agent capable of cleaving the CM than in the absence of such a cleaving agent. Thus, when the AA is in the uncleaved state, the AB is inhibited from target binding and can be masked from target binding (i.e., the first conformation is such the AB can not bind the target), and in the cleaved state the AB is not inhibited or is unmasked to target binding.

[00120] The CM and AB of the AA may be selected so that the AB represents a binding moiety for a target of interest, and the CM represents a substrate for a protease that is co-localized with the target at a treatment site in a subject. Alternatively or in addition, the CM is a cysteine-cysteine disulfide bond that is cleavable as a result of reduction of this disulfide bond. AAs contain at least one of a protease-cleavable CM or a cysteine-cysteine disulfide bond, and in some embodiments include both kinds of CMs. The AAs can alternatively or further include a photolabile substrate, activatable by a light source. The AAs disclosed herein find particular use where, for example, a protease capable of cleaving a site in the CM is present at relatively higher levels in target-containing tissue of a treatment site (for example diseased tissue; for example for therapeutic treatment or diagnostic treatment) than in tissue of non-treatment sites (for example in healthy tissue), as exemplified in Figure 2. The AAs disclosed herein also find particular use where, for example, a reducing agent capable of reducing a site in the CM is present at relatively higher levels in target-containing tissue of a treatment or diagnostic site than in tissue of non-treatment non-diagnostic sites. The AAs disclosed herein also find particular use where, for example, a light source, for example, by way of laser, capable of photolysing a site in the CM is introduced to a target-containing tissue of a treatment or diagnostic site.

[00121] In some embodiments AAs can provide for reduced toxicity and/or adverse side effects that could otherwise result from binding of the AB at non-treatment sites if the AB were not masked or otherwise inhibited from binding its target. Where the AA contains a CM that is cleavable by a reducing agent that facilitates reduction of a disulfide bond, the ABs of such AAs may be selected to exploit activation of an AB where a target of interest is present at a desired treatment site characterized by elevated levels of a reducing agent, such that the environment is of a higher reduction potential than, for example, an environment of a non-treatment site.

[00122] In general, an AA can be designed by selecting an AB of interest and constructing the remainder of the AA so that, when conformationally constrained, the MM provides for masking of the AB or reduction of binding of the AB to its target. Structural design criteria to be taken into account to provide for this functional feature.

[00123] In certain embodiments dual-target binding AAs are provided in the present disclosure. Such dual target binding AAs contain two ABs, which may bind the same or different target. In specific embodiments, dual-targeting AAs contain bispecific antibodies or antibody fragments. In one specific exemplary embodiment, the AA contains an IL17 AB and an IL23 AB. In other specific embodiments the AA contains a IL12 AB and a IL23 AB, or a EGFR AB and a VEGF AB, or a IGF1R AB and EGFR

AB, or a cMET AB and IGF1R AB, or a EGFR AB and a VEGF AB, or a Notch Receptor AB and a EGFR AB, or a Jagged ligand AB and a EGFR AB, or a cMET AB and a VEGF AB.

[00124] Dual target binding AAs can be designed so as to have a CM cleavable by a cleaving agent that is co-localized in a target tissue with one or both of the targets capable of binding to the ABs of the AA.

Dual target binding AAs with more than one AB to the same or different targets can be designed so as to have more than one CM, wherein the first CM is cleavable by a cleaving agent in a first target tissue and wherein the second CM is cleavable by a cleaving agent in a second target tissue, with one or more of the targets capable of binding to the ABs of the AA. The first and second target tissues can be spatially separated, for example, at different sites in the organism. The first and second target tissues can be the same tissue temporally separated, for example the same tissue at two different points in time, for example the first time point can be when the tissue is a healthy tumor, and the second time point can be when the tissue is a necrosed tumor.

[00125] AAs exhibiting a switchable phenotype of a desired dynamic range for target binding in an inhibited versus an uninhibited conformation are provided. Dynamic range generally refers to a ratio of (a) a maximum detected level of a parameter under a first set of conditions to (b) a minimum detected value of that parameter under a second set of conditions. For example, in the context of an AA, the dynamic range refers to the ratio of (a) a maximum detected level of target protein binding to an AA in the presence of protease capable of cleaving the CM of the AA to (b) a minimum detected level of target protein binding to an AA in the absence of the protease. The dynamic range of an AA can be calculated as the ratio of the equilibrium dissociation constant of an AA cleaving agent (e.g., enzyme) treatment to the equilibrium dissociation constant of the AA cleaving agent treatment. The greater the dynamic range of an AA, the better the switchable phenotype of the AA. AAs having relatively higher dynamic range values (e.g., greater than 1) exhibit more desirable switching phenotypes such that target protein binding by the AA occurs to a greater extent (e.g., predominantly occurs) in the presence of a cleaving agent (e.g., enzyme) capable of cleaving the CM of the AA than in the absence of a cleaving agent.

[00126] AAs can be provided in a variety of structural configurations. Exemplary formulae for AAs are provided below. It is specifically contemplated that the N- to C-terminal order of the AB, MM and CM may be reversed within an AA. It is also specifically contemplated that the CM and MM may overlap in amino acid sequence, e.g., such that the CM is contained within the MM.

[00127] For example, AAs can be represented by the following formula (in order from an amino (N) terminal region to carboxyl (C) terminal region:

(MM)-(CM)-(AB)

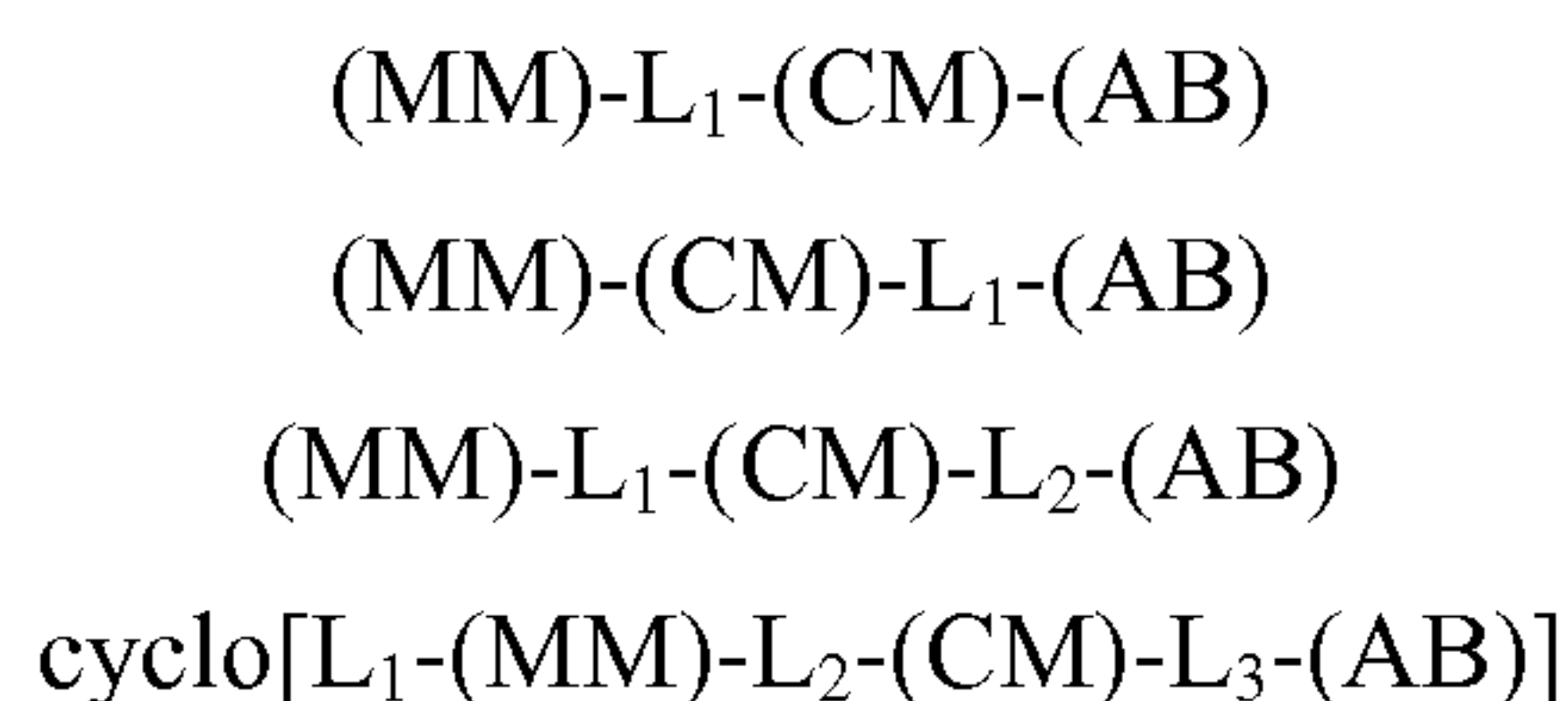
(AB)-(CM)-(MM)

where MM is a masking moiety, CM is a cleavable moiety, and AB is an antibody or fragment thereof. It should be noted that although MM and CM are indicated as distinct components in the formula above, in all exemplary embodiments (including formulae) disclosed herein it is contemplated that the amino acid sequences of the MM and the CM could overlap, e.g., such that the CM is completely or partially

contained within the MM. In addition, the formulae above provide for additional amino acid sequences that may be positioned N-terminal or C-terminal to the AA elements.

[00128] In many embodiments it may be desirable to insert one or more linkers, e.g., flexible linkers, into the AA construct so as to provide for flexibility at one or more of the MM-CM junction, the CM-AB junction, or both. For example, the AB, MM, and/or CM may not contain a sufficient number of residues (e.g., Gly, Ser, Asp, Asn, especially Gly and Ser, particularly Gly) to provide the desired flexibility. As such, the switchable phenotype of such AA constructs may benefit from introduction of one or more amino acids to provide for a flexible linker. In addition, as described below, where the AA is provided as a conformationally constrained construct, a flexible linker can be operably inserted to facilitate formation and maintenance of a cyclic structure in the uncleaved AA.

[00129] For example, in certain embodiments an AA comprises one of the following formulae (where the formula below represent an amino acid sequence in either N- to C-terminal direction or C- to N-terminal direction):



wherein MM, CM, and AB are as defined above; wherein L₁, L₂, and L₃ are each independently and optionally present or absent, are the same or different flexible linkers that include at least 1 flexible amino acid (e.g., Gly); and wherein cyclo where present, the AA is in the form of a cyclic structure due to the presence of a disulfide bond between a pair of cysteines in the AA. In addition, the formulae above provide for additional amino acid sequences that may be positioned N-terminal or C-terminal to the AA elements. It should be understood that in the formula cyclo[L₁-(MM)-L₂-(CM)-L₃-(AB)], the cysteines responsible for the disulfide bond may be positioned in the AA to allow for one or two tails, thereby generating a lasso or omega structure when the AA is in a disulfide-bonded structure (and thus conformationally constrained state). The amino acid sequence of the tail(s) can provide for additional AA features, such as binding to a target receptor to facilitate localization of the AA, increasing serum half-life of the AA, and the like. Targeting moieties (e.g., a ligand for a receptor of a cell present in a target tissue) and serum half-life extending moieties (e.g., polypeptides that bind serum proteins, such as immunoglobulin (e.g., IgG) or serum albumin (e.g., human serum albumin (HSA)).

Elements of Modified and Activatable Antibodies

(a) Antibodies or Antibody Fragments (collectively referred to as ABs)

[00130] According to the present invention, ABs directed against any antigen or hapten may be used. ABs used in the present invention may be directed against any determinant, e.g., tumor, bacterial, fungal, viral, parasitic, mycoplasmal, histocompatibility, differentiation and other cell membrane antigens,

pathogen surface antigens, toxins, enzymes, allergens, drugs, intracellular targets, and any biologically active molecules. Additionally, a combination of ABs reactive to different antigenic determinants may be used.

[00131] As used herein, the AB is a full length antibody or an antibody fragment containing an antigen binding domain, which is capable of binding, especially specific binding, to a target of interest, usually a protein target of interest. A schematic of an AA is provided in Figure 1. In such embodiments, the AB can be but is not limited to variable or hypervariable regions of light and/or heavy chains of an antibody (V_L , V_H), variable fragments (Fv), Fab' fragments, F(ab')₂ fragments, Fab fragments, single chain antibodies (scAb), single chain variable regions (scFv), complementarity determining regions (CDR), domain antibodies (dAbs), single domain heavy chain immunoglobulins of the BHH or BNAR type, single domain light chain immunoglobulins, or other polypeptides known in the art containing an AB capable of binding target proteins or epitopes on target proteins. In further embodiments, the AB may be a chimera or hybrid combination containing more than one AB, for example a first AB and a second AB such that each AB is capable of binding to the same or different target. In some embodiments, the AB is a bispecific antibody or fragment thereof, designed to bind two different antigens. In some embodiments there is a first MM and CM and/or a second MM and CM coupled to the first AB and the second AB, respectively, in the activatable form.

[00132] The origin of the AB can be a naturally occurring antibody or fragment thereof, a non-naturally occurring antibody or fragment thereof, a synthetic antibody or fragment thereof, a hybrid antibody or fragment thereof, or an engineered antibody or fragment thereof. The antibody can be a humanized antibody or fragment thereof.

[00133] In certain embodiments, more than one AB is contained in the AA. In some embodiments the ABs can be derived from bispecific antibodies or fragments thereof. In other embodiments the AA can be synthetically engineered so as to incorporate ABs derived from two different antibodies or fragments thereof. In such embodiments, the ABs can be designed to bind two different targets, two different antigens, or two different epitopes on the same target. An AB containing a plurality of ABs capable of binding more than one target site are usually designed to bind to different binding sites on a target or targets of interest such that binding of a first AB of the AA does not substantially interfere with binding of a second AB of the AA to a target. AAs containing multiple ABs can further include multiple AB-MM units, which may optionally be separated by additional CMs so that upon exposure to a modifying agent, the ABs are no longer inhibited from specifically binding their targets, or are 'unmasked'.

[00134] In some embodiments, use of antibody fragments as sources for the AB allow permeation of target sites at an increased rate. The Fab' fragments of IgG immunoglobulins are obtained by cleaving the antibody with pepsin [resulting in a bivalent fragment, (Fab')₂] or with papain [resulting in 2 univalent fragments, (2 Fab)]. Parham, 1983, J. Immunol. 131: 2895-2902; Lamoyi and Nisonoff, 1983, J. Immunol. Meth. 56: 235-243. The bivalent (Fab')₂ fragment can be split by mild reduction of one or a few disulfide bonds to yield univalent Fab' fragments. The Fab and (Fab')₂ fragments are smaller than a

whole antibody, still containing an AB and, therefore can permeate the target site or tissue more easily when used as the AB. This may offer an advantage for *in vivo* delivery in certain embodiments because many such fragments do not cross a placental barrier. As a result, using this embodiment of the present invention, an AA may be delivered at an *in vivo* site (such as a tumor) to a pregnant female without exposing the fetus.

[00135] Methods for generating an antibody (or fragment thereof) for a given target are well known in the art. The structure of antibodies and fragments thereof, variable regions of heavy and light chains of an antibody (V_H and V_L), F_v , $F(ab')_2$, Fab fragments, single chain antibodies (scAb), single chain variable regions (scFv), complementarity determining regions (CDR), and domain antibodies (dAbs) are well understood. Methods for generating a polypeptide having a desired antigen-binding domain of a target antigen are known in the art.

[00136] Methods for modifying antibodies or antibody fragments to couple additional polypeptides are also well-known in the art. For instance, peptides such as MMs, CMs or linkers may be coupled to modify antibodies to generate the modified ABs and AAs of the disclosure. AAs that contain protease-activated ABs can be developed and produced with standard methods, as described in the schematic in Figure 3.

[00137] The antibody or fragment thereof (collectively referred to as AB) is capable of specifically binding a protein target. An AB of the invention can specifically bind to its target with a dissociation constant (K_d) of no more than 1000 nM, 100nM, 50nM, 10nM, 5nM, 1nM, 500pM, 400pM, 350pM, 300pM, 250pM, 200pM, 150pM, 100pM, 50pM, 25pM, 10pM, 5pM, 1pM, .5pM, or .1pM.

[00138] Exemplary classes of targets of an AB include, but are not necessarily limited to, cell surface receptors and secreted binding proteins (e.g., growth factors), soluble enzymes, structural proteins (e.g. collagen, fibronectin) and the like. In some embodiments, AAs contemplated by the present disclosure are those having an AB capable of binding an extracellular target, usually an extracellular protein target. In other embodiments AAs can be designed such that they are capable of cellular uptake and are designed to be switchable inside a cell.

[00139] In exemplary embodiments, in no way limiting, the AB is a binding partner for any target listed in Table 1. In specific exemplary embodiments, the AB is a binding partner for EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4. In one specific embodiment the AB is not a binding partner for CD52.

[00140] In exemplary embodiments, in no way limiting, exemplary sources for ABs are listed in Table 2. In specific exemplary embodiments, the source for an AB of the invention is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab. In one specific embodiment, the source for the AB is not alemtuzumab or is not Campath™.

Table 1: Exemplary Targets

1-92-LFA-3	cMet	HGF	IL4	PSMA
Anti-Lewis-Y	Collagen	hGH	IL4R	RAAG12
Apelin J receptor	CSFR	Hyaluronidase	IL6	Sphingosine 1 Phosphate
C5 complement	CSFR-1	IFNalpha	Insulin Receptor	TGFbeta
CD11a	CTLA-4	IFNbeta	Jagged Ligands	TNFalpha
CD172A	CXCR4	IFNgamma	Jagged 1	TNFalpha
CD19	DL44	IgE	Jagged 2	TNFR
CD20	DLL4	IgE Receptor	MUC1	TRAIL-R1
CD22	EGFR	IGF	Na/K ATPase	TRAIL-R2
CD25	EpCAM	IGF1R	NGF	Transferrin
CD28	EPHA2	IL11	Notch Receptors	Transferrin receptor
CD3	ERBB3	IL12	Notch 1	TRK-A
CD30	F protein of RSV	IL13	Notch 2	TRK-B
CD33	FAP	IL15	Notch 3	VCAM-1
CD40	FGF-2	IL17	Notch 4	VEGF
CD40L	FGFR1	IL18	PDGF-AA	VEGF-A
CD41	FGFR2	IL1B	PDGF-BB	VEGF-B
CD44	FGFR3	IL1R	PDGFRalpha	VEGF-C
CD52	FGFR4	IL2	PDGFRalpha	VEGF-D
CD64	Folate receptor	IL21	PDGFRbeta	VEGFR1
CD80	GP IIb/IIIa receptors	IL23	PDGFRbeta	VEGFR2
CD86	Gp130	IL23R	Phosphatidylserine	VEGFR3
CLAUDIN-3	GPIIB/IIIA	IL29	PlGF	alpha4beta1 integrin
CLAUDIN-4	HER2/neu	IL2R	PSCA	alpha4beta7 integrin

Table 2: Exemplary sources for ABs

Antibody Trade Name (antibody name)	Target
Avastin™ (bevacizumab)	VEGF
Lucentis™ (ranibizumab)	VEGF
Erbitux™ (cetuximab)	EGFR
Vectibix™ (panitumumab)	EGFR
Remicade™ (infliximab)	TNFα
Humira™ (adalimumab)	TNFα
Tysabri™ (natalizumab)	Integrinα4
Simulect™ (basiliximab)	IL2R
Soliris™ (eculizumab)	Complement C5
Raptiva™ (efalizumab)	CD11a
Bexxar™ (tositumomab)	CD20
Zevalin™ (ibritumomab tiuxetan)	CD20
Rituxan™ (rituximab)	CD20
Zenapax™ (daclizumab)	CD25
Myelotarg™ (gemtuzumab)	CD33
Mylotarg™ (gemtuzumab ozogamicin)	CD33
Campath™ (alemtuzumab)	CD52

ReoPro™ (abiximab)	Glycoprotein receptor IIb/IIIa
Xolair™ (omalizumab)	IgE
Herceptin™ (trastuzumab)	Her2
Synagis™ (palivizumab)	F protein of RSV
(ipilimumab)	CTLA-4
(tremelimumab)	CTLA-4
Hu5c8	CD40L
(pertuzumab)	Her2-neu
(ertumaxomab)	CD3/Her2-neu
Orencia™ (abatacept)	CTLA-4
(tanezumab)	NGF
(bavituximab)	Phosphatidylserine
(zalutumumab)	EGFR
(mapatumumab)	EGFR
(matuzumab)	EGFR
(nimotuzumab)	EGFR
ICR62	EGFR
mAb 528	EGFR
CH806	EGFR
MDX-447	EGFR/CD64
(edrecolomab)	EpCAM
RAV12	RAAG12
huJ591	PSMA
Enbrel™ (etanercept)	TNF-R
Amevive™ (alefacept)	1-92-LFA-3
Antril™, Kineret™ (ankinra)	IL-1Ra
GC1008	TGFbeta
	Notch 1
	Jagged 1
(adecatumumab)	EpCAM
(figitumumab)	IGF1R
(tocilizumab)	IL-6

[00141] The exemplary sources for some of the ABs listed in Table 2 are detailed in the following references which are incorporated by reference herein for their description of one or more of the referenced AB sources: **Remicade™ (infliximab)**: U.S. patent 6,015,557, Nagahira K, Fukuda Y, Oyama Y, Kurihara T, Nasu T, Kawashima H, Noguchi C, Oikawa S, Nakanishi T. Humanization of a mouse neutralizing monoclonal antibody against tumor necrosis factor-alpha (TNF-alpha). J Immunol Methods. 1999 Jan 1;222(1-2):83-92.) Knight DM, Trinh H, Le J, Siegel S, Shealy D, McDonough M, Scallon B, Moore MA, Vilcek J, Daddona P, et al. Construction and initial characterization of a mouse-human chimeric anti-TNF antibody. Mol Immunol. 1993 Nov;30(16):1443-53. **Humira™ (adalimumab)**: Sequence in US Patent 6 258 562. **Raptiva™ (efalizumab)**: Sequence listed in Werther WA, Gonzalez TN, O'Connor SJ, McCabe S, Chan B, Hotaling T, Champe M, Fox JA, Jardieu PM, Berman PW, Presta LG. Humanization of an anti-lymphocyte function-associated antigen (LFA)-1 monoclonal antibody and reengineering of the humanized antibody for binding to rhesus LFA-1. J Immunol. 1996 Dec 1;157(11):4986-95. **Mylotarg™ (gemtuzumab ozogamicin)**: (Sequence listed in CO MS, Avdalovic NM, Caron PC, Avdalovic MV, Scheinberg DA, Queen C: Chimeric and humanized antibodies with

specificity for the CD33 antigen. *J Immunol* 148:1149, 1991) (Caron PC, Schwartz MA, Co MS, Queen C, Finn RD, Graham MC, Divgi CR, Larson SM, Scheinberg DA. Murine and humanized constructs of monoclonal antibody M195 (anti-CD33) for the therapy of acute myelogenous leukemia. *Cancer*. 1994 Feb 1;73(3 Suppl):1049-56). **Soliris™ (eculizumab)**: Hillmen P, Young N, Schubert J, Brodsky R, Socié G, Muus P, Röth A, Szer J, Elebute M, Nakamura R, Browne P, Risitano A, Hill A, Schrezenmeier H, Fu C, Maciejewski J, Rollins S, Mojcik C, Rother R, Luzzatto L (2006). The complement inhibitor eculizumab in paroxysmal nocturnal hemoglobinuria. *N Engl J Med* **355** (12): 1233–43. **Tysabri™ (natalizumab)**: Sequence listed in Leger OJ, Yednock TA, Tanner L, Horner HC, Hines DK, Keen S, Saldanha J, Jones ST, Fritz LC, Bendig MM. Humanization of a mouse antibody against human alpha-4 integrin: a potential therapeutic for the treatment of multiple sclerosis. *Hum Antibodies*. 1997;8(1):3-16. **Synagis™ (palivizumab)**: Sequence listed in Johnson S, Oliver C, Prince GA, Hemming VG, Pfarr DS, Wang SC, Dormitzer M, O'Grady J, Koenig S, Tamura JK, Woods R, Bansal G, Couchenour D, Tsao E, Hall WC, Young JF. Development of a humanized monoclonal antibody (MEDI-493) with potent *in vitro* and *in vivo* activity against respiratory syncytial virus. *J Infect Dis*. 1997 Nov; 176(5):1215-24. **Ipilimumab**: *J. Immunother*: 2007; 30(8): 825-830 Ipilimumab (Anti-CTLA4 Antibody) Causes Regression of Metastatic Renal Cell Cancer Associated With Enteritis and Hypophysitis; James C. Yang, Marybeth Hughes, Udai Kammula, Richard Royal, Richard M. Sherry, Suzanne L. Topalian, Kimberly B. Suri, Catherine Levy, Tamika Allen, Sharon Mavroukakis, Israel Lowy, Donald E. White, and Steven A. Rosenberg. **Tremelimumab**: *Oncologist* 2007;12;153-883; Blocking Monoclonal Antibody in Clinical Development for Patients with Cancer; Antoni Ribas, Douglas C. Hanson, Dennis A. Noe, Robert Millham, Deborah J. Guyot, Steven H. Bernstein, Paul C. Canniff, Amarnath Sharma and Jesus Gomez-Navarro.

(b) Masking Moiety (MM)

[00142] The masking moiety (MM) of the present disclosure generally refers to an amino acid sequence coupled to the AB and positioned such that it reduces the AB's ability to specifically bind its target. In some cases the MM is coupled to the AB by way of a linker.

[00143] When the AB is modified with a MM and is in the presence of the target, specific binding of the AB to its target is reduced or inhibited, as compared to the specific binding of the AB not modified with an MM or the specific binding of the parental AB to the target.

[00144] The K_d of the AB modified with a MM towards the AB's target is generally greater than the K_d of the AB not modified with a MM or the K_d of parental AB towards the target. Conversely, the binding affinity of the AB modified with a MM towards the target is generally lower than the binding affinity of the AB not modified with a MM or the parental AB towards the target.

[00145] The dissociation constant (K_d) of the MM towards the AB is generally greater than the K_d of the AB towards the target. Conversely, the binding affinity of the MM towards the AB is generally lower than the binding affinity of the AB towards the target.

[00146] When the AB is modified with a MM and is in the presence of the target, specific binding of the AB to its target can be reduced or inhibited, as compared to the specific binding of the AB not modified with an MM or the specific binding of the parental AB to the target. When the AB is modified with a CM and a MM and is in the presence of the target but not sufficient enzyme or enzyme activity to cleave the CM, specific binding of the modified AB to the target is reduced or inhibited, as compared to the specific binding of the AB modified with a CM and a MM in the presence of the target and sufficient enzyme or enzyme activity to cleave the CM.

[00147] The MM can inhibit the binding of the AB to the target. The MM can bind the antigen binding domain of the AB and inhibit binding of the AB to its target. The MM can sterically inhibit the binding of the AB to the target. The MM can allosterically inhibit the binding of the AB to its target. In these embodiments when the AB is modified or coupled to a MM and in the presence of target, there is no binding or substantially no binding of the AB to the target, or no more than .001%, .01%, .1%, 1%, 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, or 50% binding of the AB to the target, as compared to the binding of the AB not modified with an MM, the binding of the parental AB, or the binding of the AB not coupled to an MM to the target, for at least 2, 4, 6, 8, 12, 28, 24, 30, 36, 48, 60, 72, 84, 96 hours, or 5, 10, 15, 30, 45, 60, 90, 120, 150, 180 days, or 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 months or greater when measured *in vivo* or in a Target Displacement *in vitro* immunoabsorbant assay, as described herein.

[00148] In certain embodiments the MM is not a natural binding partner of the AB. The MM may be a modified binding partner for the AB which contains amino acid changes that at least slightly decrease affinity and/or avidity of binding to the AB. In some embodiments the MM contains no or substantially no homology to the AB's natural binding partner. In other embodiments the MM is no more than 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, or 80% similar to the natural binding partner of the AB.

[00149] When the AB is in a 'masked' state, even in the presence of a target for the AB, the MM interferes with or inhibits the binding of the AB to the target. However, in the unmasked state of the AB, the MM's interference with target binding to the AB is reduced, thereby allowing greater access of the AB to the target and providing for target binding.

[00150] For example, when the modified antibody is an AA and comprises a CM, the AB can be unmasked upon cleavage of the CM, in the presence of enzyme, preferably a disease-specific enzyme. Thus, the MM is one that when the AA is uncleaved provides for masking of the AB from target binding, but does not substantially or significantly interfere or compete for binding of the target to the AB when the AA is in the cleaved conformation. Thus, the combination of the MM and the CM facilitates the switchable/activatable phenotype, with the MM decreasing binding of target when the AA is uncleaved, and cleavage of the CM by protease providing for increased binding of target.

[00151] The structural properties of the MM will vary according to a variety of factors such as the minimum amino acid sequence required for interference with AB binding to target, the target protein-AB

binding pair of interest, the size of the AB, the length of the CM, whether the CM is positioned within the MM and also serves to mask the AB in the uncleaved AA, the presence or absence of linkers, the presence or absence of a cysteine within or flanking the AB that is suitable for providing a CM of a cysteine-cysteine disulfide bond, and the like.

[00152] One strategy for masking an antibody or fragment thereof (AB) in an AA is to provide the AA in a loop that sterically hinders access of target to the AB. In this strategy, cysteines are positioned at or near the N-terminus, C-terminus, or AB of the AA, such that upon formation of a disulfide bond between the cysteines, the AB is masked.

[00153] In some embodiments, the MM is coupled to the AA by covalent binding. In another embodiment, the AA composition is prevented from binding to the target by binding the MM to an N-terminus of the AA. In yet another embodiment, the AA is coupled to the MM by cysteine-cysteine disulfide bridges between the MM and the AA.

[00154] The MM can be provided in a variety of different forms. In certain embodiments, the MM can be selected to be a known binding partner of the AB, provided that the MM binds the AB with less affinity and/or avidity than the target protein to which the AB is designed to bind following cleavage of the CM so as to reduce interference of MM in target-AB binding. Stated differently, as discussed above, the MM is one that masks the AB from target binding when the AA is uncleaved, but does not substantially or significantly interfere or compete for binding for target when the AA is in the cleaved conformation. In a specific embodiment, the AB and MM do not contain the amino acid sequences of a naturally-occurring binding partner pair, such that at least one of the AB and MM does not have the amino acid sequence of a member of a naturally occurring binding partner

[00155] The efficiency of the MM to inhibit the binding of the AB to its target when coupled can be measured by a Masking Efficiency measure, using an immunoabsorbant Target Displacement Assay, as described herein in the Examples section of the disclosure. Masking efficiency of MMs is determined by at least two parameters: affinity of the MM for the antibody or fragment thereof and the spatial relationship of the MM relative to the binding interface of the AB to its target.

[00156] Regarding affinity, by way of example, an MM may have high affinity but only partially inhibit the binding site on the AB, while another MM may have a lower affinity for the AB but fully inhibit target binding. For short time periods, the lower affinity MM may show sufficient masking; in contrast, over time, that same MM may be displaced by the target (due to insufficient affinity for the AB).

[00157] In a similar fashion, two MMs with the same affinity may show different extents of masking based on how well they promote inhibition of the binding site on the AB or prevention of the AB from binding its target. In another example, a MM with high affinity may bind and change the structure of the AB so that binding to its target is completely inhibited while another MM with high affinity may only partially inhibit binding. As a consequence, discovery of an effective MM cannot be based only on affinity but can include an empirical measure of Masking Efficiency. The time-dependent target

displacement of the MM in the AA can be measured to optimize and select for MMs. A novel Target Displacement Assay is described herein for this purpose.

[00158] In some embodiments the MM can be identified through a screening procedure from a library of candidates AAs having variable MMs. For example, an AB and CM can be selected to provide for a desired enzyme/target combination, and the amino acid sequence of the MM can be identified by the screening procedure described below to identify an MM that provides for a switchable phenotype. For example, a random peptide library (e.g., from about 2 to about 40 amino acids or more) may be used in the screening methods disclosed herein to identify a suitable MM. In specific embodiments, MMs with specific binding affinity for an antibody or fragment thereof (AB) can be identified through a screening procedure that includes providing a library of peptide scaffolds consisting of candidate MMs wherein each scaffold is made up of a transmembrane protein and the candidate MM. The library is then contacted with an entire or portion of an AB such as a full length antibody, a naturally occurring antibody fragment, or a non-naturally occurring fragment containing an AB (also capable of binding the target of interest), and identifying one or more candidate MMs having detectably bound AB. Screening can include one more rounds of magnetic-activated sorting (MACS) or fluorescence-activated sorting (FACS). Screening can also include determination of the dissociation constant (K_d) of MM towards the AB and subsequent determination of the Masking Efficiency.

[00159] In this manner, AAs having an MM that inhibits binding of the AB to the target in an uncleaved state and allows binding of the AB to the target in a cleaved state can be identified, and can further provide for selection of an AA having an optimal dynamic range for the switchable phenotype. Methods for identifying AAs having a desirable switching phenotype are described in more detail below.

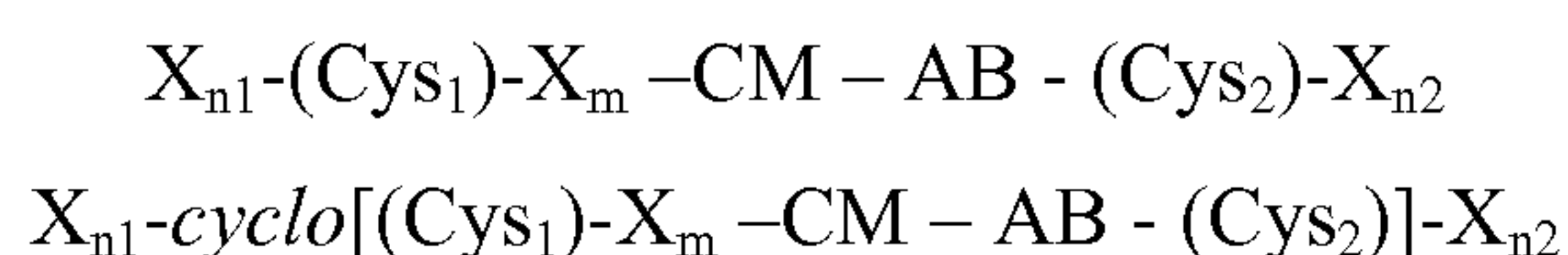
[00160] Alternatively, the MM may not specifically bind the AB, but rather interfere with AB-target binding through non-specific interactions such as steric hindrance. For example, the MM may be positioned in the uncleaved AA such that the tertiary or quaternary structure of the AA allows the MM to mask the AB through charge-based interaction, thereby holding the MM in place to interfere with target access to the AB.

[00161] AAs can also be provided in a conformationally constrained structure, such as a cyclic structure, to facilitate the switchable phenotype. This can be accomplished by including a pair of cysteines in the AA construct so that formation of a disulfide bond between the cysteine pairs places the AA in a loop or cyclic structure. Thus the AA remains cleavable by the desired protease while providing for inhibition of target binding to the AB. Upon cleavage of the CM, the cyclic structure is opened, allowing access of target to the AB.

[00162] The cysteine pairs can be positioned in the AA at any position that provides for a conformationally constrained AA, but that, following CM reduction, does not substantially or significantly interfere with target binding to the AB. For example, the cysteine residues of the cysteine pair are positioned in the MM and a linker flanked by the MM and AB, within a linker flanked by the MM and AB, or other suitable configurations. For example, the MM or a linker flanking an MM can

include one or more cysteine residues, which cysteine residue forms a disulfide bridge with a cysteine residue positioned opposite the MM when the AA is in a folded state. It is generally desirable that the cysteine residues of the cysteine pair be positioned outside the AB so as to avoid interference with target binding following cleavage of the AA. Where a cysteine of the cysteine pair to be disulfide bonded is positioned within the AB, it is desirable that it be positioned to as to avoid interference with AB-target binding following exposure to a reducing agent.

[00163] Exemplary AAs capable of forming a cyclic structure by disulfide bonds between cysteines can be of the general formula (which may be from either N-to C-terminal or from C- to N-terminal direction):



wherein

X_{n1} and X_{n2} are independently, optionally present or absent and, when present, independently represent any amino acid, and can optionally include an amino acid sequence of a flexible linker (e.g., at least one Gly, Ser, Asn, Asp, usually at least one Gly or Ser, usually at least one Gly), and n_1 and n_2 are independently selected from zero or any integer, usually not more than 1, 2, 3, 4, 5, 6, 7, 8, 9 or 10;

Cys_1 and Cys_2 represent first and second cysteines of a pair capable of forming a disulfide bond;

X_m represents amino acids of a masking motif (MM), where X is any amino acid, wherein X_m can optionally include a flexible linker (e.g., at least one Gly, Ser, Asn, Asp, usually at least one Gly or Ser, usually at least one Gly); and where m is an integer greater than 1, usually 2, 3, 4, 5, 6, 7, 8, 9, 10 or more (as described above);

CM represents a cleavable moiety (as described herein); and

AB represents an antibody or fragment thereof (as described herein).

[00164] As used in the formula above, *cyclo* indicates a disulfide bond in the AA that provides for a cyclic structure of the AA. Furthermore, the formula above contemplates dual target-binding AAs wherein MM refers to an AB1 and AB refers to AB2, where AB1 and AB2 are arbitrary designations for first and second ABs, and where the target capable of binding the ABs may be the same or different target, or the same or different binding sites of the same target. In such embodiments, the AB1 and/or AB2 acts as a masking moiety to interfere with target binding to an uncleaved dual target-binding AA.

[00165] As illustrated above, the cysteines can thus be positioned in the AA allow for one or two tails (represented by X_{n1} and X_{n2} above), thereby generating a lasso or omega structure when the AA is in a disulfide-bonded structure (and thus conformationally constrained state). The amino acid sequence of the tail(s) can provide for additional AA features, such as binding to a target receptor to facilitate localization of the AA.

[00166] In certain specific embodiments, the MM does not inhibit cellular entry of the AA.

(c) Cleavable Moiety (CM)

[00167] In some embodiments, the cleavable moiety (CM) of the AA may include an amino acid sequence that can serve as a substrate for a protease, usually an extracellular protease. In other embodiments, the CM comprises a cysteine-cysteine pair capable of forming a disulfide bond, which can be cleaved by action of a reducing agent. In other embodiments the CM comprises a substrate capable of being cleaved upon photolysis.

[00168] The CM is positioned in the AA such that when the CM is cleaved by a cleaving agent (e.g., a protease substrate of a CM is cleaved by the protease and/or the cysteine-cysteine disulfide bond is disrupted via reduction by exposure to a reducing agent) or by light-induced photolysis, in the presence of a target, resulting in a cleaved state, the AB binds the target, and in an uncleaved state, in the presence of the target, binding of the AB to the target is inhibited by the MM (Figure 2). It should be noted that the amino acid sequence of the CM may overlap with or be included within the MM, such that all or a portion of the CM facilitates masking of the AB when the AA is in the uninhibited or uncleaved or unmasked conformation.

[00169] The CM may be selected based on a protease that is co-localized in tissue with the desired target of the AB of the AA. A variety of different conditions are known in which a target of interest is co-localized with a protease, where the substrate of the protease is known in the art. In the example of cancer, the target tissue can be a cancerous tissue, particularly cancerous tissue of a solid tumor. There are reports in the literature of increased levels of proteases having known substrates in a number of cancers, e.g., solid tumors. See, e.g., La Rocca et al, (2004) British J. of Cancer 90(7): 1414-1421. Non-limiting examples of disease include: all types of cancers (breast, lung, colorectal, prostate, head and neck, pancreatic, etc), rheumatoid arthritis, Crohn's disease, melanomas, SLE, cardiovascular damage, ischemia, etc. Furthermore, anti-angiogenic targets, such as VEGF, are known. As such, where the AB of an AA is selected such that it is capable of binding an anti-angiogenic target such as VEGF, a suitable CM will be one which comprises a peptide substrate that is cleavable by a protease that is present at the cancerous treatment site, particularly that is present at elevated levels at the cancerous treatment site as compared to non-cancerous tissues. In one exemplary embodiment, the AB of an AA can bind VEGF and the CM can be a matrix metalloprotease (MMP) substrate, and thus is cleavable by an MMP. In other embodiments, the AB of an AA can bind a target of interest and the CM can be, for example, legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA. In other embodiments, the AA is activated by other disease-specific proteases, in diseases other than cancer such as multiple sclerosis or rheumatoid arthritis.

[00170] The unmodified or uncleaved CM can allow for efficient inhibition or masking of the AB by tethering the MM to the AB. When the CM is modified (cleaved, reduced, photolysed), the AB is no longer inhibited or unmasked and can bind its target.

[00171] The AA can comprise more than one CM such that the AA would comprise, for example, a first CM (CM1) and a second CM (CM2). The CM1 and CM2 can be different substrates for the same enzyme (for example exhibiting different binding affinities to the enzyme), or different substrates for

different enzymes, or CM1 can be an enzyme substrate and CM2 can be a photolysis substrate, or CM1 can be an enzyme substrate and CM2 can be a substrate for reduction, or CM1 can be a substrate for photolysis and CM2 can be a substrate for reduction, and the like.

[00172] The CM is capable of being specifically modified (cleaved, reduced or photolysed) by an agent (ie enzyme, reducing agent, light) at a rate of about $.001-1500 \times 10^4 \text{ M}^{-1}\text{S}^{-1}$ or at least .001, .005, .01, .05, .1, .5, 1, 2.5, 5, 7.5, 10, 15, 20, 25, 50, 75, 100, 125, 150, 200, 250, 500, 750, 1000, 1250, or $1500 \times 10^4 \text{ M}^{-1}\text{S}^{-1}$.

[00173] For specific cleavage by an enzyme, contact between the enzyme and CM is made. When the AA comprising an AB coupled to a MM and a CM is in the presence of target and sufficient enzyme activity, the CM can be cleaved. Sufficient enzyme activity can refer to the ability of the enzyme to make contact with the CM and effect cleavage. It can readily be envisioned that an enzyme may be in the vicinity of the CM but unable to cleave because of other cellular factors or protein modification of the enzyme.

[00174] Exemplary substrates can include but are not limited to substrates cleavable by one or more of the following enzymes or proteases in Table 3.

Table 3 –Exemplary Enzymes/Proteases

ADAM10	Caspase 8	Cathepsin S	MMP 8
ADAM12	Caspase 9	FAP	MMP 9
ADAM17	Caspase 10	Granzyme B	MMP-13
ADAMTS	Caspase 11	Guanidinobenzoate (GB)	MMP 14
ADAMTS5	Caspase 12	Hepsin	MT-SP1
BACE	Caspase 13	Human Neutrophil Elastase (HNE)	Neprilysin
Caspases	Caspase 14	Legumain	NS3/4A
Caspase 1	Cathepsins	Matriptase 2	Plasmin
Caspase 2	Cathepsin A	Meprin	PSA
Caspase 3	Cathepsin B	MMP 1	PSMA
Caspase 4	Cathepsin D	MMP 2	TACE
Caspase 5	Cathepsin E	MMP 3	TMPRSS 3 / 4
Caspase 6	Cathepsin K	MMP 7	uPA
Caspase 7	MT1-MMP		

[00175] Alternatively or in addition, the AB of an AA can be one that binds a target of interest and the CM can involve a disulfide bond of a cysteine pair, which is thus cleavable by a reducing agent such as, for example, but not limited to a cellular reducing agent such as glutathione (GSH), thioredoxins, NADPH, flavins, ascorbate, and the like, which can be present in large amounts in tissue of or surrounding a solid tumor.

(d) Linkers

[00176] Linkers suitable for use in compositions described herein are generally ones that provide flexibility of the modified AB or the AA to facilitate the inhibition of the binding of the AB to the target. Such linkers are generally referred to as flexible linkers. Suitable linkers can be readily selected and can be of any of a suitable of different lengths, such as from 1 amino acid (e.g., Gly) to 20 amino acids, from

2 amino acids to 15 amino acids, from 3 amino acids to 12 amino acids, including 4 amino acids to 10 amino acids, 5 amino acids to 9 amino acids, 6 amino acids to 8 amino acids, or 7 amino acids to 8 amino acids, and may be 1, 2, 3, 4, 5, 6, or 7 amino acids.

[00177] Exemplary flexible linkers include glycine polymers (G)_n, glycine-serine polymers (including, for example, (GS)_n, (GSGGS)_n and (GGGS)_n, where n is an integer of at least one), glycine-alanine polymers, alanine-serine polymers, and other flexible linkers known in the art. Glycine and glycine-serine polymers are relatively unstructured, and therefore may be able to serve as a neutral tether between components.

Glycine accesses significantly more phi-psi space than even alanine, and is much less restricted than residues with longer side chains (see Scheraga, Rev. Computational Chem. 11173-142 (1992)).

Exemplary flexible linkers include, but are not limited to Gly-Gly-Ser-Gly, Gly-Gly-Ser-Gly-Gly, Gly-Ser-Gly-Ser-Gly, Gly-Ser-Gly-Gly-Gly, Gly-Gly-Gly-Ser-Gly, Gly-Ser-Ser-Ser-Gly, and the like. The ordinarily skilled artisan will recognize that design of an AA can include linkers that are all or partially flexible, such that the linker can include a flexible linker as well as one or more portions that confer less flexible structure to provide for a desired AA structure.

(e) Additional Elements

[00178] In addition to the elements described above, the modified ABs and AAs can contain additional elements such as, for example, amino acid sequence N- or C-terminal of the AA. For example, AAs can include a targeting moiety to facilitate delivery to a cell or tissue of interest. Moreover, in the context of the AA libraries discussed further below, the AA can be provided in the context of a scaffold protein to facilitate display of the AA on a cell surface.

Exemplary Embodiments

[00179] The compositions and AAs provided here in can be useful for a variety of purposes including therapeutics and diagnostics.

[00180] An exemplary AA provided herein can be a legumain-activatable anti-EGFR coupled to a MM, plasmin-activatable anti-EGFR coupled to a MM, TMPRSS-3/4 activatable anti-EGFR coupled to a MM, legumain-activatable cetuximab coupled to a MM, plasmin-activatable cetuximab coupled to a MM, TMPRSS-3/4 activatable cetuximab coupled to a MM, legumain-activatable vectibix coupled to a MM, plasmin-activatable vectibix coupled to a MM, or a TMPRSS-3/4 activatable vectibix coupled to a MM. In some embodiments these AAs can be useful for the treatment of diagnosis of head and neck carcinomas, or colon, lung, or pancreatic carcinomas.

[00181] An exemplary AA provided herein can be a MMP9-activatable anti-TNFalpha coupled to a MM, MT1-MMP-activatable anti-TNFalpha coupled to a MM, cathepsin-activatable anti-TNFalpha coupled to a MM, MMP9-activatable infliximab coupled to a MM, MT1-MMP-activatable infliximab coupled to a MM, cathepsin-activatable infliximab coupled to a MM, MMP9-activatable adalimumab coupled to a MM, MT1-MMP-activatable adalimumab coupled to a MM, or a cathepsin-activatable adalimumab

coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of rheumatoid arthritis or multiple sclerosis.

[00182] An exemplary AA provided herein can be a legumain-activatable anti- CD11a coupled to a MM, plasmin-activatable anti- CD11a coupled to a MM, caspase-activatable anti- CD11a coupled to a MM, cathepsin-activatable anti- CD11a coupled to a MM, legumain-activatable efalizumab coupled to a MM, plasmin-activatable efalizumab coupled to a MM, caspase-activatable efalizumab coupled to a MM, cathepsin-activatable efalizumab coupled to a MM, legumain-activatable anti- CSFR coupled to a MM, plasmin-activatable anti- CSFR coupled to a MM, caspase-activatable anti- CSFR coupled to a MM, or a cathepsin-activatable anti- CSFR coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of tumor associated macrophages for carcinomas.

[00183] An exemplary AA provided herein can be a plasmin-activatable anti-CTLA-4 coupled to a MM, caspase-activatable anti-CTLA-4 coupled to a MM, MT1-MMP-activatable anti-CTLA-4 coupled to a MM, plasmin-activatable ipilimumab coupled to a MM, caspase-activatable ipilimumab coupled to a MM,

[00184] MT1-MMP-activatable ipilimumab coupled to a MM, plasmin-activatable tremelimumab coupled to a MM, caspase-activatable tremelimumab coupled to a MM, or a MT1-MMP-activatable tremelimumab coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of malignant melanomas.

[00185] An exemplary AA provided herein can be a PSA-activatable anti-EPCAM coupled to a MM, legumain-activatable anti-EPCAM coupled to a MM, PSA-activatable adecatumumab coupled to a MM or a legumain-activatable adecatumumab coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of prostate cancer.

[00186] An exemplary AA provided herein can be a human neutrophil elastase-activatable anti-CD40L coupled to a MM, or a human neutrophil elastase-activatable Hu5c8 coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of lymphomas.

[00187] An exemplary AA provided herein can be a beta-secretase -activatable anti-Notch1 coupled to a MM, legumain -activatable anti- Notch1 coupled to a MM, plasmin-activatable anti- Notch1 coupled to a MM, uPA-activatable anti- Notch1 coupled to a MM, beta-secretase -activatable anti-Notch3 coupled to a MM, legumain -activatable anti- Notch3 coupled to a MM, plasmin-activatable anti- Notch3 coupled to a MM, uPA-activatable anti- Notch3 coupled to a MM, beta-secretase -activatable anti-Jagged1 coupled to a MM, legumain -activatable anti- Jagged1 coupled to a MM, plasmin-activatable anti- Jagged1 coupled to a MM, uPA-activatable anti- Jagged1 coupled to a MM, beta-secretase -activatable anti-Jagged2 coupled to a MM, legumain -activatable anti- Jagged2 coupled to a MM, plasmin-activatable anti- Jagged2 coupled to a MM, or a uPA-activatable anti- Jagged2 coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of triple negative (ER, PR and Her2 negative) breast, head and neck, colon and other carcinomas.

[00188] An exemplary AA provided herein can be a MMP-activatable anti-CD52 coupled to a MM, or a MMP-activatable anti-campath coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of multiple sclerosis.

[00189] An exemplary AA provided herein can be a MMP-activatable anti-MUC1 coupled to a MM, legumain -activatable anti-MUC1 coupled to a MM, plasmin-activatable anti-MUC1 coupled to a MM, or a uPA-activatable anti-MUC1 coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of epithelial derived tumors.

[00190] An exemplary AA provided herein can be a legumain -activatable anti- IGF1R coupled to a MM, plasmin-activatable anti- IGF1R coupled to a MM, caspase-activatable anti- IGF1R coupled to a MM, legumain -activatable anti- figitumumab coupled to a MM, plasmin-activatable anti- figitumumab coupled to a MM, or a caspase-activatable anti- figitumumab coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of non-small cell lung, and other epithelial tumors.

[00191] An exemplary AA provided herein can be a legumain -activatable anti- transferrin receptor coupled to a MM, plasmin-activatable anti- transferrin receptor coupled to a MM, or a caspase-activatable anti- transferrin receptor coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of solid tumors, pancreatic tumors.

[00192] An exemplary AA provided herein can be a legumain -activatable anti- gp130 coupled to a MM, plasmin-activatable anti- gp130 coupled to a MM, or a uPA-activatable anti- gp130 coupled to a MM. In some embodiments these AAs can be useful for the treatment or diagnosis of solid tumors.

[00193] In certain other non-limiting exemplary embodiments, activatable antibody compositions include an legumain masked AB specific for Notch1, a uPA activatable masked AB specific for Jagged1, a plasmin activatable, masked anti-VEGF scFv, a MMP-9 activatable, masked anti-VCAM scFv, and a MMP-9 activatable masked anti-CTLA4.

[00194] These AAs are provided by way of example only and such enzyme activatable masked antibody AAs could be designed to any target as listed in but not limited to those in Table 1 and by using any antibody as listed in but not limited to those in Table 2.

Activatable Antibody Complexes

[00195] In one aspect of the invention, the AA exists as a complex (AAC) comprising two or more ABs, as depicted in Figures 34-36. The present disclosure provides complexes of activatable antibodies (AACs), which exhibit activatable/switchable binding to one or more target proteins. AACs generally include one or more antibodies or antibody fragments (ABs), masking moieties (MMs), and cleavable moieties (CMs). In some embodiments, the CM contains an amino acid sequence that serves as a substrate for a protease of interest. In other embodiments, the CM provides a cysteine-cysteine disulfide bond that is cleavable by reduction. The AAC exhibits an activatable conformation such that at least one AB is less accessible to target when unmodified than after modification of the CM, e.g., in the presence of

a cleavage agent (e.g., a protease that recognizes the cleavage site of the CM) or a reducing agent (e.g. a reducing agent that reduces disulfide bonds in the CM).

[00196] The CM and AB of the AAC may be selected so that the AB represents a binding moiety for a target of interest, and the CM represents a substrate for a protease that is co-localized with the target at a treatment site in a subject. In some embodiments AACs can provide for reduced toxicity and/or adverse side effects that could otherwise result from binding of the ABs at non-treatment sites if they were not masked. In some embodiments, the AAC can further comprise a detectable moiety or a diagnostic agent. In certain embodiments the AAC is conjugated to a therapeutic agent located outside the antigen binding region. AACs can also be used in diagnostic and/or imaging methods or to detect the presence or absence of a cleaving agent in a sample.

[00197] A schematic of an AAC is provided in Figure 43. As illustrated, the elements of the AAC are arranged so that the CM is positioned such that in a cleaved state (or relatively active state) and in the presence of a target, the AB binds a target, while in an uncleaved state (or relatively inactive state) in the presence of the target, binding of the ABs to the target is inhibited due to the masking of the ABs by the MM in the complex. As used herein, the term cleaved state refers to the condition of the AAC following cleavage of the CM by a protease and/or reduction of a cysteine-cysteine disulfide bond of the CM. The term uncleaved state, as used herein, refers to the condition of the AAC in the absence of cleavage of the CM by a protease and/or in the absence reduction of a cysteine-cysteine disulfide bond of the CM. As discussed above, the term AAC is used herein to refer to AAC in both its uncleaved (native) state, as well as in its cleaved state. It will be apparent to the ordinarily skilled artisan that in some embodiments a cleaved AAC may lack an MM due to cleavage of the CM by protease, resulting in release of at least the MM (e.g., where the MM is not joined to the AAC by a covalent bond.)

[00198] By activatable or switchable is meant that the AAC exhibits a first level of binding to a target when in a native or uncleaved state (i.e., a first conformation), and a second level of binding to the target in the cleaved state (i.e., a second conformation), where the second level of target binding is greater than the first level of binding. In general, access of target to the AB of the AAC is greater in the presence of a cleaving agent capable of cleaving the CM than in the absence of such a cleaving agent. Thus, in the native or uncleaved state the AB is masked from target binding (i.e., the first conformation is such that it interferes with access of the target to the AB), and in the cleaved state the AB is unmasked to target binding.

[00199] In general, an AAC can be designed by selecting an AB(s) of interest and constructing the remainder of the AAC so that, when conformationally constrained, the MM provides for masking of the AB. Dual target binding AACs contain two ABs, which may bind the same or different target. In specific embodiments, dual-targeting AACs contain bispecific antibodies or antibody fragments.

[00200] In certain embodiments, a complex is comprised of two activatable antibodies (AA), each containing an AB, CM, and MM such that cross-masking occurs – that is, the MM on one AA interferes with target binding by the AB on the other AA (Figure 34A). In other embodiments, a complex is

comprised of two AAs, with each AA containing an AB and one containing a CM and MM such that universal cross-masking occurs – that is, the MM effects formation of the complex and interferes with target binding by the ABs on both AAs (Figure 34B). In other embodiments, a complex is comprised of two AAs, each containing two ABs, CMs, and MMs such that cross-masking occurs – that is, the MMs on one AA interfere with target binding by the ABs on the other AA (Figure 34C). In other embodiments, a complex is comprised of two AAs, with one AA containing two ABs, CMs, and MMs such that universal cross-masking occurs – that is, the MMs interfere with target binding by the ABs on both AAs (Figure 34D). In other embodiments, a complex is comprised of two molecules of a bispecific AA where the bispecific AA contains two ABs, CMs, and MMs such that cross-masking occurs in the complex – that is, the MM1 interferes with target binding by the AB1 on the opposite molecule, and the MM2 interferes with target binding by the AB2 on the opposite molecule (Figure 34E). In other embodiments, a complex is comprised of two molecules of a bispecific AA where the bispecific AA contains two ABs, one CM, and one MM such that universal cross-masking occurs in the complex – that is, the MM interferes with target binding by both ABs (Figure 34F).

[00201] In general, disassembly of the AAC and access of targets to at least one of the ABs of the AACs are greater in the presence of a cleaving agent capable of cleaving the CMs than in the absence of such a cleaving agent (Figure 35). The two AAs of a complex may contain ABs that bind different targets, or that bind different epitopes on the same target.

[00202] One of the MM/AB pairs of the complex may be used for stable complex formation and have no therapeutic target on its own. A high affinity MM for the non-therapeutic AB allows a stable complex to form, even with a lower affinity MM for the therapeutic AB. The low affinity MM for the therapeutic AB, in the context of the multivalent complex, will be sufficient for masking the therapeutic AB, but after cleavage will more readily dissociate. For maximum target binding in the cleaved state, the difference in affinity of the MM and target for the AB should be maximized.

[00203] In other embodiments, an AB may form a covalent linkage to an MM on the opposite molecule of the complex. In the presence of a cleaving agent the complex disassembles such that at least one of the other ABs will bind its target (Figure 36). Such a covalent linkage may form between reactive amino acid side chains in the MM and AB, eg. disulfide bond between cysteines, or by chemical conjugation of reactive groups to the MM and a catalytic AB. For examples of covalent binding antibodies see Chmura A. J. et al., *Proc Natl Acad Sci U S A.* 2001 July 17, 98(15): 8480–8484 ; Rader, C. et al., *Proc Natl Acad Sci U S A.* 2003 April 29, 100(9): 5396–5400 ; Armentano, F. et al., *Immunology Letters* 2006 February 28, 103(1): 51-57.

[00204] It should be noted that although MM and CM are indicated as distinct components, it is contemplated that the amino acid sequences of the MM and the CM could overlap, e.g., such that the CM is completely or partially contained within the MM. In many embodiments it may be desirable to insert one or more linkers, e.g., flexible linkers, into the AAC construct so as to provide for flexibility at one or more of the MM-CM junction, the CM-AB junction, or both. In addition to the elements described

above, the AACs can contain additional elements such as, for example, amino acid sequence N- or C-terminal of the AAC.

Activatable Antibody Conjugates

[00205] In one aspect of the invention, the AB of the AA is further conjugated to an agent such as a therapeutic agent, thus producing activatable antibody conjugates (AACJs), a specific type of AA. The agent is attached either directly or via a linker to the AB. Such agents or linkers are selectively attached to those areas of ABs which are not a part of nor directly involved with the antigen binding site of the molecule. An exemplary AACJ is pictured in Figure 20.

[00206] According to one embodiment of the present invention, an agent may be conjugated to an AB. When delivery and release of the agent conjugated to the AB are desired, immunoglobulin classes that are known to activate complement are used. In other applications, carrier immunoglobulins may be used which are not capable of complement activation. Such immunoglobulin carriers may include: certain classes of antibodies such IgM, IgA, IgD, IgE; certain subclasses of IgG; or certain fragments of immunoglobulins, e.g., half ABs (a single heavy: light chain pair), or Fab, Fab' or (Fab')₂ fragments.

[00207] Exemplary AACJs are AAs coupled to a therapeutic agent wherein the AB is directed to EGFR, CD44, Notch1, 2, 3 or 4 Jagged1 or 2, EpCAM, or IGF-1R.

[00208] The chemical linking methods described herein allow the resulting AACJ to retain the ability to bind antigen and to activate the complement cascade (when the unconjugated AA also had such ability). As a result, when the AACJ is administered to an individual, the subsequent formation of immune complexes with target antigens in vivo can activate the individual's serum complement system. The linker is designed to be susceptible to cleavage by complement and so the agent can be cleaved at the target site by one or more of the enzymes of the complement cascade. The majority of the release of the agent occurs following delivery to the target site.

[00209] In an exemplary embodiment, it is known that all cells of a tumor do not each possess the target antigenic determinant. Thus, delivery systems which require internalization into the target cell will effect successful delivery to those tumor cells that possess the antigenic determinant and that are capable of internalizing the conjugate. Tumor cells that do possess the antigenic determinant or are incapable of this internalization, will escape treatment. According to the method of the present invention, AACJs deliver the agent to the target cells. More importantly, however, once attached to the target cell, the method described in the present invention allows the release or activation of the active or activatable therapeutic agent. Release or activation may be mediated by the individual's activated by but not limited to the following: complement enzymes, tissue plasminogen activator, urokinase, plasmin or another enzyme having proteolytic activity, or by activation of a photosensitizer or substrate modification. Once released, the agent is then free to permeate the target sites, e.g., tumor mass. As a result, the agent will act on tumor cells that do not possess the antigenic determinant or could not internalize the conjugate. Additionally, the

entire process is not dependent upon internalization of the conjugate.

(a) Methods for Conjugating Agents

[00210] The present invention utilizes several methods for attaching agents to ABs (which include antibodies and fragments thereof), two exemplary methods being attachment to the carbohydrate moieties of the AB, or attachment to sulfhydryl groups of the AB. In certain embodiments, the attachment does not significantly change the essential characteristics of the AB or the AA itself, such as immunospecificity and immunoreactivity. Additional considerations include simplicity of reaction and stability of the antibody conjugate produced. In certain embodiments the AB is first conjugated to one or more agents of interest followed by attachment of an MM and CM to produce an AACJ. In other embodiments the AB is first attached to a MM and CM following which an agent of interest is further conjugated producing an AACJ.

i. Attachment to Oxidized Carbohydrate Moieties

[00211] In certain embodiments, agents may be conjugated to the carbohydrate moiety of an AB. Some of the carbohydrate moieties are located on the Fc region of the immunoglobulin and are required in order for C1 binding to occur. The carbohydrate moiety of the Fc region of an immunoglobulin may be utilized in the scheme described herein in the embodiments where the AB is an antibody or antibody fragment that includes at least part of an Fc region. Alternatively, the Fab or Fab' fragments of any immunoglobulins which contain carbohydrate moieties may be utilized in the reaction scheme described herein. An example of such an immunoglobulin is the human IgM sequenced by Putnam et al. (1973, Science 182: 287).

[00212] The carbohydrate side chains of antibodies, Fab or Fab' fragments or other fragments containing an AB may be selectively oxidized to generate aldehydes. A variety of oxidizing agents can be used, such as periodic acid, paraperiodic acid, sodium metaperiodate and potassium metaperiodate. The resulting aldehydes may then be reacted with amine groups (e.g., ammonia derivatives such as primary amine, secondary amine, hydroxylamine, hydrazine, hydrazide, phenylhydrazine, semicarbazide or thiosemicarbazide) to form a Schiff base or reduced Schiff base (e.g., imine, enamine, oxime, hydrazone, phenylhydrazone, semicarbazone, thiosemicarbazone or reduced forms thereof). Chemical methods of oxidation of antibodies are provided in US 4,867,973 and this patent is incorporated by reference in its entirety. Oxidation of antibodies with these oxidizing agents can be carried out by known methods. In the oxidation, the AB is used generally in the form of an aqueous solution, the concentration being generally less than 100 mg/ml, preferably 1 to 20 mg/ml. When an oxygen acid or a salt thereof is used as the oxidizing agent, it is used generally in the form of an aqueous solution, and the concentration is generally 0.001 to 10 mM, sometimes 1.0 to 10 mM. The amount of the oxygen acid or salt thereof depends on the kind of AB, but generally it is used in excess, for example, twice to ten times as much as the amount of

the oxidizable carbohydrate. The optimal amount, however, can be determined by routine experimentation.

[00213] In the process for oxidizing ABs with oxygen acids or salts thereof, the optional ranges include a pH of from about 4 to 8, a temperature of from 0° to 37° C., and a reaction period of from about 15 minutes to 12 hours. During the oxidation with an oxygen acid or a salt thereof, the reaction can be carried in minimal light to prevent over oxidation.

[00214] Alternatively, the carbohydrate moiety of the AB may be modified by enzymatic techniques so as to enable attachment to or reaction with other chemical groups. One example of such an enzyme is galactose oxidase which oxidizes galactose in the presence of oxygen to form an aldehyde. Oxidation of the carbohydrate portion of ABs may also be done with the enzyme, galactose oxidase (Cooper et al., 1959, J. Biol. Chem. 234:445-448). The antibody is used in aqueous solution, the concentration being generally 0.5 to 20 mg/ml. The enzyme generally is used at about 5 to 100 units per ml of solution, at a pH ranging from about 5.5 to about 8.0. The influence of pH, substrate concentration, buffers and buffer concentrations on enzyme reaction are reported in Cooper et al., supra.

[00215] The AB conjugates, AA conjugates, or AB linker-intermediates of the invention may be produced by reacting the oxidized AB with any linker or agent having an available amine group selected from the group consisting of primary amine, secondary amine, hydrazine, hydrazide, hydroxylamine, phenylhydrazine, semicarbazide and thiosemicarbazide groups. In an exemplary method, a solution of the oxidized AB or AB linker at a concentration of from about 0.5 to 20 mg/ml is mixed with the agent or linker (molar ratios of reactive amine group to antibody aldehyde ranging from about 1 to about 10,000) and the solution incubated for from about 1 to 18 hours. Suitable temperatures are from 0° to 37° C. and pH may be from about 6 to 8. After the conjugates have been formed they can optionally be stabilized with a suitable reducing agent, such as sodium cyanoborohydride or sodium borohydride.

ii. Attachment to Sulfhydryl Groups

[00216] When the AB is a full-length antibody or includes at least part of the heavy chain, free sulfhydryl groups can be generated from the disulfide bonds of the immunoglobulin molecule. This is accomplished by mild reduction of the antibody. The disulfide bonds of IgG, which are generally susceptible to reduction, are those that link the two heavy chains. The disulfide bonds located near the antigen binding region of the antibody remain relatively unaffected. Such reduction results in the loss of ability to fix complement but does not interfere with antibody-antigen binding ability (Karush et al, 1979, Biochem. 18: 2226-2232). The free sulfhydryl groups generated in the intra-heavy chain region can then react with reactive groups of a linker or agent to form a covalent bond which will reduce interference with the antigen binding site of the immunoglobulin. Such reactive groups include, but are not limited to, reactive haloalkyl groups (including, for example, haloacetyl groups), p-mercuribenzoate groups and groups capable of Michael-type addition reactions (including, for example, maleimides and groups of the type

described in Mitra and Lawton, 1979, J. Amer. Chem. Soc. 101: 3097-3110). The haloalkyl can be any alkyl group substituted with bromine, iodine or chlorine.

[00217] Details of the conditions, methods and materials suitable for mild reduction of antibodies and antibody fragments as described generally herein may be found in Stanworth and Turner, 1973, In Handbook of Experimental Immunology, Vol. 1, Second Edition, Weir (ed.), Chapter 10, Blackwell Scientific Publications, London, which chapter is incorporated herein by reference.

[00218] AB-agent conjugates (or AB-linker intermediates) which are produced by attachment to free sulfhydryl groups of reduced immunoglobulin or reduced antibody fragments do not or negligibly activate complement. Thus, these conjugates may be used in *in vivo* systems where cleavage and release of the agent is not desirable (e.g., an enzyme that acts on a specific substrate). Such conjugates may also be used when non-complement mediated release is desired. In such an embodiment, the agent may be linked to sulfhydryl groups on the reduced AB via linkers which are susceptible to cleavage by enzymes having proteolytic activity, including but not limited to trypsin, urokinase, plasmin, tissue plasminogen activator and the like.

[00219] Although attachment of an agent to sulfhydryl groups of the AB reduces the complement fixation ability of the conjugate, such methods of attachment may be used to make AA conjugates for use in the complement-mediated release system. In such an embodiment, an agent joined to a complement-sensitive substrate linker can be attached to sulfhydryls of reduced ABs or AAs and delivered to the target in a mixture with non conjugated AAs that are capable of activating complement. The latter would activate complement which would cleave the agent from the former.

[00220] According to one embodiment of the present invention, for attachment to sulfhydryl groups of reduced ABs or AAs, the substrate linkers or the agents are modified by attaching an iodoalkyl group to one end of the linker. The unmodified site on the linker may or may not be covalently attached to an agent. For instance, the substrate linkers which are ester or amide linked to agents are modified by the addition of an iodoalkyl group thus forming an iodoalkyl derivative. As mentioned previously, the linker may be one that is susceptible or resistant to cleavage by activated complement, trypsin, plasmin, tissue plasminogen activator, urokinase or another specific enzyme having proteolytic activity.

(b) Agents for Conjugation to ABs

[00221] ABs may be attached to any agent which retains its essential properties after reaction with the AB, and which enables the AB to substantially retain immunospecificity and immunoreactivity allowing the AA to function as appropriate. The agent can include all chemical modifications and derivatives of agents which substantially retain their biological activity.

[00222] When it is desired to attach an aldehyde of the oxidized carbohydrate portion of an AB to an agent, the agent should contain an amine group selected from the group consisting of primary amine, secondary amine, hydrazine, hydrazide, hydroxylamine, phenylhydrazine, semicarbazide and thiosemicarbazide groups. If the agent does not contain any such amino group, the agent can be modified to introduce a suitable amine group available for coupling.

[00223] The agent to be attached to an AB for use in an AA is selected according to the purpose of the intended application (i.e, killing, prevention of cell proliferation, hormone therapy or gene therapy). Such agents may include but is not limited to, for example, pharmaceutical agents, toxins, fragments of toxins, alkylating agents, enzymes, antibiotics, antimetabolites, antiproliferative agents, hormones, neurotransmitters, DNA, RNA, siRNA, oligonucleotides, antisense RNA, aptamers, diagnostics, radioopaque dyes, radioactive isotopes, fluorogenic compounds, magnetic labels, nanoparticles, marker compounds, lectins, compounds which alter cell membrane permeability, photochemical compounds, small molecules, liposomes, micelles, gene therapy vectors, viral vectors, and the like. Non-limiting Table 4 lists some of the exemplary pharmaceutical agents that may be employed in the herein described invention but in no way is meant to be an exhaustive list. Finally, combinations of agents or combinations of different classes of agents may be used.

[00224] According to one embodiment of the present invention, photochemicals including photosensitizers and photothermolytic agents may be used as agents. Efficient photosensitizers include, but are not limited to porphyrins and modified porphyrins (e.g., hematoporphyrin, hematoporphyrin dihyddrazide, deuteroporphyrin dihydrazide and protoporphyrin dihydrazide), rose bengal, acridines, thiazines, xanthenes, anthraquinones, azines, flavin and nonmetal-containing porphyrins, porphyrin-like compounds, methylene blue, eosin, psoralin and the like. Other photosensitizers include, but are not limited to tetracyclines (e.g., dimethylchlor tetracycline) sulfonamides (e.g., sulfanilamide), griseofulvin, phenothiazines, (e.g., chlorpromazine), thiazides, sulfonylurea, and many others. Photochemicals may be designed or synthetically prepared to absorb light at specific wavelengths. Photothermolytic agents, such as Azure A, which are activated at the site of action by a light source (see Anderson and Parrish, 1983, Science 220: 524-527) may be utilized as agents.

[00225] According to another embodiment of the present invention, enzymes that catalyze substrate modification with the production of cytotoxic by-products may be used as agents. Examples of such enzymes include but are not limited to glucose oxidase, galactose oxidase, xanthene oxidase and the like.

Table 4: Exemplary Pharmaceutical Agents for Conjugation

NAME/CLASS	LINKAGE	MANUFACTURERS(S)
<u>I. ANTIBACTERIALS</u>		
<u>Aminoglycosides</u>		
Streptomycin	ester/amide	
Neomycin	ester/amide	Dow, Lilly, Dome, Pfipharmics
Kanamycin	ester/amide	Bristol
Amikacin	ester	Bristol
Gentamicin	ester/amide	Upjohn, Wyeth, Schering
Tobramycin	ester/amide	Lilly
Streptomycin B	ester/amide	Squibb
Spectinomycin	ester	Upjohn
Ampicillin	amide	Squibb, Parke-Davis, Comer, Wyeth, Upjohn, Bristol, SKF
Sulfanilamide	amide	Merrell-National

Polymyxin	amide	Burroughs-Wellcome, Dow, Parke-
Chloramphenicol	ester	Davis
		Parke-Davis
<u>II. ANTIVIRALS</u>		
Acyclovir	ester/amide	Burroughs-Wellcome
Vira A	amide	Parke-Davis
Symmetrel		Endo
<u>III. ANTIFUNGALS</u>		
Nystatin	ester	Squibb, Primo, Lederle, Pfizer, Holland-Rantor
<u>IV. ANTINEOPLASTICS</u>		
Adriamycin	ester/amide	Adria
Cerubidine	ester/amide	Ives
Bleomycin	ester/amide	Bristol
Alkeran	amide	Burroughs-Wellcome
Velban	ester	Lilly
Oncovin	ester	Lilly
Fluorouracil	ester	Adria, Roche, Herbert
Methotrexate	amide	Lederle
Thiotepa	—	Lederle
Bisantrene	—	Lederle
Novantrone	ester	Lederle
Thioguanine	amide	Burroughs-Wellcome
Procarabazine	—	Hoffman La Roche
Cytarabine	—	Upjohn
<u>V.</u>		
<u>RADIOPHARMACEUTICALS</u>		
¹²⁵ I		
¹³¹ I		
^{99m} Tc (Technetium)		
<u>VI. HEAVY METALS</u>		
Barium		
Gold		
Platinum		
<u>VII. ANTIMYCOPLASMA</u>		
Tylosine		
Spectinomycin		

(c) Linkers for Conjugating Agents

[00226] The present invention utilizes several methods for attaching agents to ABs: (a) attachment to the carbohydrate moieties of the AB, or (b) attachment to sulfhydryl groups of the AB. According to the invention, ABs may be covalently attached to an agent through an intermediate linker having at least two reactive groups, one to react with AB and one to react with the agent. The linker, which may include any compatible organic compound, can be chosen such that the reaction with AB (or agent) does not adversely affect AB reactivity and selectivity. Furthermore, the attachment of linker to agent might not destroy the activity of the agent. Suitable linkers for reaction with oxidized antibodies or oxidized antibody fragments include those containing an amine selected from the group consisting of primary amine, secondary amine, hydrazine, hydrazide, hydroxylamine, phenylhydrazine, semicarbazide and

thiosemicarbazide groups. Such reactive functional groups may exist as part of the structure of the linker, or may be introduced by suitable chemical modification of linkers not containing such groups.

[00227] According to the present invention, suitable linkers for attachment to reduced ABs include those having certain reactive groups capable of reaction with a sulfhydryl group of a reduced antibody or fragment. Such reactive groups include, but are not limited to: reactive haloalkyl groups (including, for example, haloacetyl groups), p-mercuribenzoate groups and groups capable of Michael-type addition reactions (including, for example, maleimides and groups of the type described by Mitra and Lawton, 1979, J. Amer. Chem. Soc. 101: 3097-3110).

[00228] The agent may be attached to the linker before or after the linker is attached to the AB. In certain applications it may be desirable to first produce an AB-linker intermediate in which the linker is free of an associated agent. Depending upon the particular application, a specific agent may then be covalently attached to the linker. In other embodiments the AB is first attached to the MM, CM and associated linkers and then attached to the linker for conjugation purposes.

(i) *Branched Linkers:*

[00229] In specific embodiments, branched linkers which have multiple sites for attachment of agents are utilized. For multiple site linkers, a single covalent attachment to an AB would result in an AB-linker intermediate capable of binding an agent at a number of sites. The sites may be aldehyde or sulfhydryl groups or any chemical site to which agents can be attached.

[00230] Alternatively, higher specific activity (or higher ratio of agents to AB) can be achieved by attachment of a single site linker at a plurality of sites on the AB. This plurality of sites may be introduced into the AB by either of two methods. First, one may generate multiple aldehyde groups and/or sulfhydryl groups in the same AB. Second, one may attach to an aldehyde or sulfhydryl of the AB a "branched linker" having multiple functional sites for subsequent attachment to linkers. The functional sites of the branched linker or multiple site linker may be aldehyde or sulfhydryl groups, or may be any chemical site to which linkers may be attached. Still higher specific activities may be obtained by combining these two approaches, that is, attaching multiple site linkers at several sites on the AB.

(ii) *Cleavable Linkers:*

[00231] Peptide linkers which are susceptible to cleavage by enzymes of the complement system, such as but not limited to urokinase, tissue plasminogen activator, trypsin, plasmin, or another enzyme having proteolytic activity may be used in one embodiment of the present invention. According to one method of the present invention, an agent is attached via a linker susceptible to cleavage by complement. The antibody is selected from a class which can activate complement. The antibody-agent conjugate, thus, activates the complement cascade and releases the agent at the target site. According to another method of the present invention, an agent is attached via a linker susceptible to cleavage by enzymes having a

proteolytic activity such as a urokinase, a tissue plasimogen activator, plasmin, or trypsin. Non-limiting examples of cleavable linker sequences are provided in Table 5.

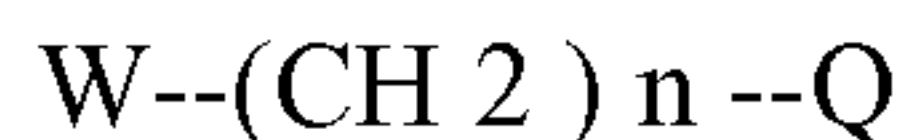
Table 5: Exemplary Linker Sequences for Conjugation

Types of Cleavable Sequences	Amino Acid Sequence
<u>Plasmin cleavable sequences</u>	
Pro-urokinase	PRFKIIGG PRFRIIGG
TGF β	SSRHRRALD
Plasminogen	RKSSIIIRMRDVVL
Staphylokinase	SSSFDKGKYKKGDDA SSSFDKGKYKRGDDA
<u>Factor Xa cleavable sequences</u>	
	IEGR IDGR GGSIDGR
<u>MMP cleavable sequences</u>	
Gelatinase A	PLGLWA
<u>Collagenase cleavable sequences</u>	
Calf skin collagen (α 1(I) chain)	GPQGIAGQ
Calf skin collagen (α 2(I) chain)	GPQGLLGA
Bovine cartilage collagen (α 1(II) chain)	GIAGQ
Human liver collagen (α 1(III) chain)	GPLGIAGI
Human α_2 M	GPEGLRVG
Human PZP	YGAGLGVV AGLGVVER AGLGISST
Rat α_1 M	EPQALAMS QALAMSAI
Rat α_2 M	AAYHLVSQ MDAFLESS
Rat α_1 I ₃ (2J)	ESLPVVAV
Rat α_1 I ₃ (27J)	SAPAVESE
Human fibroblast collagenase (autolytic cleavages)	DVAQFVLT VAQFVLTE AQFVLTEG PVQPIGPQ

[00232] In addition agents may be attached via disulfide bonds (for example, the disulfide bonds on a cysteine molecule) to the AB. Since many tumors naturally release high levels of glutathione (a reducing agent) this can reduce the disulfide bonds with subsequent release of the agent at the site of delivery. In certain specific embodiments the reducing agent that would modify a CM would also modify the linker of the conjugated AA.

(iii) *Spacers and Cleavable Elements:*

[00233] In still another embodiment, it may be necessary to construct the linker in such a way as to optimize the spacing between the agent and the AB of the AA. This may be accomplished by use of a linker of the general structure:



wherein

W is either --NH--CH₂-- or --CH₂--;

Q is an amino acid, peptide; and

n is an integer from 0 to 20.

[00234] In still other embodiments, the linker may comprise a spacer element and a cleavable element. The spacer element serves to position the cleavable element away from the core of the AB such that the cleavable element is more accessible to the enzyme responsible for cleavage. Certain of the branched linkers described above may serve as spacer elements.

[00235] Throughout this discussion, it should be understood that the attachment of linker to agent (or of spacer element to cleavable element, or cleavable element to agent) need not be particular mode of attachment or reaction. Any reaction providing a product of suitable stability and biological compatibility is acceptable.

(iv) *Serum Complement and Selection of Linkers:*

[00236] According to one method of the present invention, when release of an agent is desired, an AB that is an antibody of a class which can activate complement is used. The resulting conjugate retains both the ability to bind antigen and activate the complement cascade. Thus, according to this embodiment of the present invention, an agent is joined to one end of the cleavable linker or cleavable element and the other end of the linker group is attached to a specific site on the AB. For example, if the agent has an hydroxy group or an amino group, it may be attached to the carboxy terminus of a peptide, amino acid or other suitably chosen linker via an ester or amide bond, respectively. For example, such agents may be attached to the linker peptide via a carbodimide reaction. If the agent contains functional groups that would interfere with attachment to the linker, these interfering functional groups can be blocked before attachment and deblocked once the product conjugate or intermediate is made. The opposite or amino terminus of the linker is then used either directly or after further modification for binding to an AB which is capable of activating complement.

[00237] Linkers (or spacer elements of linkers) may be of any desired length, one end of which can be covalently attached to specific sites on the AB of the AA. The other end of the linker or spacer element may be attached to an amino acid or peptide linker.

[00238] Thus when these conjugates bind to antigen in the presence of complement the amide or ester bond which attaches the agent to the linker will be cleaved, resulting in release of the agent in its active form. These conjugates, when administered to a subject, will accomplish delivery and release of the agent at the target site, and are particularly effective for the *in vivo* delivery of pharmaceutical agents,

antibiotics, antimetabolites, antiproliferative agents and the like as presented in but not limited to those in Table 4.

(v) *Linkers for Release without Complement Activation:*

[00239] In yet another application of targeted delivery, release of the agent without complement activation is desired since activation of the complement cascade will ultimately lyse the target cell. Hence, this approach is useful when delivery and release of the agent should be accomplished without killing the target cell. Such is the goal when delivery of cell mediators such as hormones, enzymes, corticosteroids, neurotransmitters, genes or enzymes to target cells is desired. These conjugates may be prepared by attaching the agent to an AB that is not capable of activating complement via a linker that is mildly susceptible to cleavage by serum proteases. When this conjugate is administered to an individual, antigen-antibody complexes will form quickly whereas cleavage of the agent will occur slowly, thus resulting in release of the compound at the target site.

(vi) *Biochemical Cross Linkers:*

[00240] In other embodiments, the AA may be conjugated to one or more therapeutic agents using certain biochemical cross-linkers. Cross-linking reagents form molecular bridges that tie together functional groups of two different molecules. To link two different proteins in a step-wise manner, hetero-bifunctional cross-linkers can be used that eliminate unwanted homopolymer formation. Exemplary hetero-bifunctional cross-linkers are referenced in Table 6.

Table 6: Exemplary Hetero-Bifunctional Cross Linkers

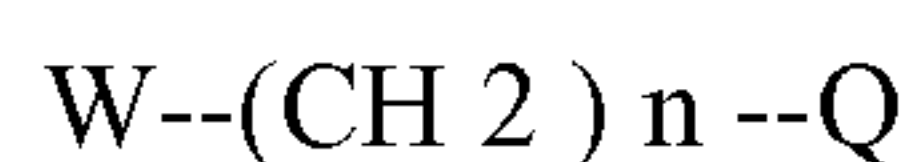
<u>HETERO-BIFUNCTIONAL CROSS-LINKERS</u>			
Linker	Reactive Toward	Advantages and Applications	Spacer Arm Length after cross-linking
SMPT	Primary amines Sulphydryls	Greater stability	11.2 A
SPDP	Primary amines Sulphydryls	Thiolation Cleavable cross-linking	6.8 A
LC-SPDP	Primary amines Sulphydryls	Extended spacer arm	15.6 A
Sulfo-LC-SPDP	Primary amines Sulphydryls	Extender spacer arm Water-soluble	15.6 A
SMCC	Primary amines Sulphydryls	Stable malcimide reactive group Enzyme-antibody conjugation Hapten-carrier protein conjugation	11.6 A
Sulfo-SMCC	Primary amines Sulphydryls	Stable maleimide reactive group Water-soluble Enzyme-antibody conjugation	11.6 A
MBS	Primary amines Sulphydryls	Enzyme-antibody conjugation Hapten-carrier protein conjugation	9.9 A
Sulfo-MBS	Primary amines Sulphydryls	Water-soluble	9.9 A
SIAB	Primary amines	Enzyme-antibody conjugation	10.6 A

Sulfo-SIAB	Sulphydryls Primary amines	Water-soluble	10.6 A
SMPB	Sulphydryls Primary amines	Extended spacer arm	14.5 A
Sulfo-SMPB	Sulphydryls Primary amines	Enzyme-antibody conjugation Extended spacer arm	14.5 A
EDE/Sulfo-NHS	Sulphydryls Primary amines	Water-soluble Hapten-Carrier conjugation	0
ABH	Carboxyl groups Carbohydrates Nonselective	Reacts with sugar groups	11.9 A

(vii) *Non-Cleavable Linkers or Direct Attachment:*

[00241] In still other embodiments of the invention, the conjugate may be designed so that the agent is delivered to the target but not released. This may be accomplished by attaching an agent to an AB either directly or via a non-cleavable linker.

[00242] These non-cleavable linkers may include amino acids, peptides, D-amino acids or other organic compounds which may be modified to include functional groups that can subsequently be utilized in attachment to ABs by the methods described herein. A general formula for such an organic linker could be



wherein W is either --NH--CH₂-- or --CH₂--;

Q is an amino acid, peptide; and

n is an integer from 0 to 20.

(viii) *Non-Cleavable Conjugates:*

[00243] Alternatively, a compound may be attached to ABs which do not activate complement. When using ABs that are incapable of complement activation, this attachment may be accomplished using linkers that are susceptible to cleavage by activated complement or using linkers that are not susceptible to cleavage by activated complement.

(d) Uses of Activatable Antibody Conjugates

[00244] The AA-agent conjugates (AACJs) of the invention are useful in therapeutics, diagnostics, substrate modification and the like.

[00245] The AACJs of the invention are useful in a variety of therapeutic *in vivo* applications such as but not limited to the treatment of neoplasms, including cancers, adenomas, and hyperplasias; certain immunological disorders, including autoimmune diseases, graft-versus-host diseases (e.g., after bone marrow transplantation), immune suppressive diseases, e.g., after kidney or bone marrow transplantation. Treatment of such cellular disorders involving, for example, bone marrow transplantation, may include purging (by killing) undesired cells, e.g., malignant cells or mature T lymphocytes.

[00246] Therapeutic applications center generally on treatment of various cellular disorders, including those broadly described above, by administering an effective amount of the antibody-agent conjugates of the invention. The properties of the antibody are such that it is immunospecific for and immunoreactive with a particular antigen render it ideally suited for delivery of agents to specific cells, tissues, organs or any other site having that particular antigen.

[00247] According to this aspect of the invention, the AACJ functions to deliver the conjugate to the target site.

[00248] The choice of ABs, linkers, and agents used to make the AACJs depends upon the purpose of delivery. The delivery and release or activation of agents at specific target sites may result in selective killing or inhibition of proliferation of tumor cells, cancer cells, fungi, bacteria, parasites, or virus. The targeted delivery of hormones, enzymes, or neurotransmitters to selected sites may also be accomplished. Ultimately the method of the present invention may have an application in gene therapy programs wherein DNA or specific genes may be delivered in vivo or in vitro to target cells that are deficient in that particular gene. Additionally, the conjugates may be used to reduce or prevent the activation of oncogenes, such as myc, ras and the like.

[00249] *In vivo* administration may involve use of agents of AACJs in any suitable adjuvant including serum or physiological saline, with or without another protein, such as human serum albumin. Dosage of the conjugates may readily be determined by one of ordinary skill, and may differ depending upon the nature of the cellular disorder and the agent used. Route of administration may be parenteral, with intravenous administration generally preferred.

(i) *Substrate Modification*

[00250] In an alternate embodiment of the present invention, substrate activation by the agent may be used to mediate formation of singlet oxygen or peroxides and induce cell killing. In this particular embodiment, the agent is an enzyme. For example, galactose oxidase will oxidize galactose and some galactose derivatives at the C 6 position. In the course of the oxidation reaction, molecular oxygen is converted into hydrogen peroxide which is toxic to neighboring cells. The enzyme glucose oxidase, a flavoenzyme, may also be used in the embodiment of this invention. This enzyme is highly specific for β -D-glucose and can act as an antibiotic due to peroxide formation. The enzyme may be attached to an AB either directly or via a non-cleavable linker. A subject is given an effective dosage of this AACJ and is then perfused with substrate. Cell killing is mediated through the formation of peroxides by the methods described above. The toxic effect of peroxides may be amplified by administration of a second enzyme, preferably of human origin, to convert its peroxide to a more toxic hypochlorous acid. Examples of suitable enzymes include but are not limited to myeloperoxidase, lactoperoxidase and chloroperoxidase.

Display Methods and Compositions for Identifying and/or Optimizing AAs

[00251] Methods for identifying and optimizing AAs, as well as compositions useful in such methods, are described below.

(a) Libraries of AAs or candidate AAs displayed on replicable biological entities

[00252] In general, the screening methods to identify an AA and/or to optimize an AA for a switchable phenotype can involve production of a library of replicable biological entities that display on their surface a plurality of different candidate AAs. These libraries can then be subjected to screening methods to identify candidate AAs having one or more desired characteristics of an AA.

[00253] The candidate AA libraries can contain candidate AAs that differ by one or more of the MM, linker (which may be part of the MM), CM (which may be part of the MM), and AB. In one embodiment the AAs in the library are variable for the MM and/or the linker, with the AB and CM being preselected. Where the AA is to include pairs of cysteine residues to provide a disulfide bond in the AA, the relative position of the cysteines in the AA can be varied.

[00254] The library for screening is generally provided as a library of replicable biological entities which display on their surface different candidate AAs. For example, a library of candidate AAs can include a plurality of candidate AAs displayed on the surface of population of a replicable biological entities, wherein each member of said plurality of candidate AAs comprises: (a) an antibody or fragment thereof (AB); (b) a cleavable moiety (CM); and (c) a candidate masking moiety (candidate MM), wherein the AB, CM and candidate MM are positioned such that the ability of the candidate MM to inhibit binding of the AB to a target in an uncleaved state and allow binding of the AB to the target in a cleaved state can be determined. Suitable replicable biological entities include cells (e.g., bacteria (e.g., *E. coli*), yeast (e.g., *S. cerevisiae*), protozoan cells, mammalian cells), bacteriophage, and viruses. Antibody display technologies are well known in the art.

(b) Display of candidate AAs on the surface of replicable biological entities

[00255] A variety of display technologies using replicable biological entities are known in the art. These methods and entities include, but are not limited to, display methodologies such as mRNA and ribosome display, eukaryotic virus display, and bacterial, yeast, and mammalian cell surface display. See Wilson, D. S., et al. 2001 *PNAS USA* 98(7):3750- 3755; Muller, O. J., et al. (2003) *Nat. Biotechnol.* 3:312; Bupp, K. and M. J. Roth (2002) *Mol. Ther.* 5(3):329 3513; Georgiou, G., et al., (1997) *Nat. Biotechnol.* 15(1):29 3414; and Boder, E. T. and K. D. Wittrup (1997) *Nature Biotech.* 15(6):553 557. Surface display methods are attractive since they enable application of fluorescence-activated cell sorting (FACS) for library analysis and screening. See Daugherty, P. S., et al. (2000) *J. Immunol. Methods* 243(1 2):211 2716; Georgiou, G. (2000) *Adv. Protein Chem.* 55:293 315; Daugherty, P. S., et al. (2000) *PNAS USA* 97(5):2029 3418; Olsen, M. J., et al. (2003) *Methods Mol. Biol.* 230:329 342; Boder, E. T. et al. (2000) *PNAS USA* 97(20):10701 10705; Mattheakis, L. C., et al. (1994) *PNAS USA* 91(19): 9022 9026; and Shusta, E. V., et al. (1999) *Curr. Opin. Biotech.* 10(2):117 122. Additional display methodologies which

may be used to identify a peptide capable of binding to a biological target of interest are described in U.S. Patent No. 7,256,038, the disclosure of which is incorporated herein by reference.

[00256] A display scaffold refers to a polypeptide which when expressed in a host cell is presented on an extracellularly accessible surface of the host cell and provides for presentation of an operably linked heterologous polypeptide. For example, display scaffolds find use in the methods disclosed herein to facilitate screening of candidate AAs. Display scaffolds can be provided such that a heterologous polypeptide of interest can be readily released from the display scaffold, e.g. by action of a protease that facilitates cleavage of the fusion protein and release of a candidate AA from the display scaffold.

[00257] Phage display involves the localization of peptides as terminal fusions to the coat proteins, e.g., pIII, pIIV of bacteriophage particles. See Scott, J. K. and G. P. Smith (1990) *Science* 249(4967):386 390; and Lowman, H. B., et al. (1991) *Biochem.* 30(45):10832 10838. Generally, polypeptides with a specific function of binding are isolated by incubating with a target, washing away non-binding phage, eluting the bound phage, and then re-amplifying the phage population by infecting a fresh culture of bacteria.

[00258] Exemplary phage display and cell display compositions and methods are described in U.S. Patent Nos. 5,223,409; 5,403,484; 7,118,159; 6,979,538; 7,208,293; 5,571,698; and 5,837,500.

[00259] Additional exemplary display scaffolds and methods include those described in U.S. Patent Application Publication No: 2007/0065158, published March 22, 2007.

[00260] Optionally, the display scaffold can include a protease cleavage site (different from the protease cleavage site of the CM) to allow for cleavage of an AA or candidate AA from a surface of a host cell.

[00261] In one embodiment, where the replicable biological entity is a bacterial cell, suitable display scaffolds include circularly permuted *Escherichia coli* outer membrane protein OmpX (CPX) described by Rice et al, *Protein Sci.* (2006) 15: 825-836. See also, U.S. Patent No. 7,256,038, issued August 14, 2007.

(c) Constructs encoding AAs

[00262] The disclosure further provides nucleic acid constructs which include sequences coding for AAs and/or candidate AAs. Suitable nucleic acid constructs include, but are not limited to, constructs which are capable of expression in a prokaryotic or eukaryotic cell. Expression constructs are generally selected so as to be compatible with the host cell in which they are to be used.

[00263] For example, non-viral and/or viral constructs vectors may be prepared and used, including plasmids, which provide for replication of an AA- or candidate AA-encoding DNA and/or expression in a host cell. The choice of vector will depend on the type of cell in which propagation is desired and the purpose of propagation. Certain constructs are useful for amplifying and making large amounts of the desired DNA sequence. Other vectors are suitable for expression in cells in culture. The choice of appropriate vector is well within the skill of the art. Many such vectors are available commercially. Methods for generating constructs can be accomplished using methods well known in the art.

[00264] In order to effect expression in a host cell, the polynucleotide encoding an AA or candidate AA is operably linked to a regulatory sequence as appropriate to facilitate the desired expression properties.

These regulatory sequences can include promoters, enhancers, terminators, operators, repressors, silencers, inducers, and 3' or 5' UTRs. Expression constructs generally also provide a transcriptional and translational initiation region as may be needed or desired, which may be inducible or constitutive, where the coding region is operably linked under the transcriptional control of the transcriptional initiation region, and a transcriptional and translational termination region. These control regions may be native to the species from which the nucleic acid is obtained, or may be derived from exogenous sources.

[00265] Promoters may be either constitutive or regulatable. In some situations it may be desirable to use conditionally active promoters, such as inducible promoters, e.g., temperature-sensitive promoters.

Inducible elements are DNA sequence elements that act in conjunction with promoters and may bind either repressors (e.g. lacO/LAC Iq repressor system in *E. coli*) or inducers (e.g. gal1/GAL4 inducer system in yeast). In such cases, transcription is virtually shut off until the promoter is de-repressed or induced, at which point transcription is turned-on.

[00266] Constructs, including expression constructs, can also include a selectable marker operative in the host to facilitate, for example, growth of host cells containing the construct of interest. Such selectable marker genes can provide a phenotypic trait for selection of transformed host cells such as dihydrofolate reductase or neomycin resistance for eukaryotic cell culture.

[00267] Expression constructs can include convenient restriction sites to provide for the insertion and removal of nucleic acid sequences encoding the AA and/or candidate AA. Alternatively or in addition, the expression constructs can include flanking sequences that can serve as the basis for primers to facilitate nucleic acid amplification (e.g., PCR-based amplification) of an AA-coding sequence of interest.

[00268] The above described expression systems may be employed with prokaryotes or eukaryotes in accordance with conventional ways, depending upon the purpose for expression. In some embodiments, a unicellular organism, such as *E. coli*, *B. subtilis*, *S. cerevisiae*, insect cells in combination with baculovirus vectors, or cells of a higher organism such as vertebrates, e.g. COS 7 cells, HEK 293, CHO, Xenopus Oocytes, etc., may be used as the expression host cells. Expression systems for each of these classes and types of host cells are known in the art.

(d) Methods of making libraries of AAs/candidate AAs displayed on replicable biological entities

[00269] The present disclosure contemplates methods of making the libraries of AAs and/or candidate AAs described herein.

[00270] In one embodiment, a method of making an AA library and/or candidate AA library comprises:

(a) constructing a set of recombinant DNA vectors as described herein that encode a plurality of AAs and/or candidate AAs; (b) transforming host cells with the vectors of step (a); and (c) culturing the host

cells transformed in step (b) under conditions suitable for expression and display of the fusion polypeptides.

(e) Production of nucleic acid sequences encoding candidate AAs

[00271] Production of candidate AAs for use in the screening methods can be accomplished using methods known in the art. Polypeptide display, single chain antibody display, antibody display and antibody fragment display are methods well known in the art. In general, an element of an AA e.g., MM, to be varied in the candidate AA library is selected for randomization. The candidate AAs in the library can be fully randomized or biased in their randomization, e.g. in nucleotide/residue frequency generally or in position of amino acid(s) within an element.

Methods of Screening for AAs

[00272] The present disclosure provides methods of identifying AAs, which can be enzymatically activated AAs, reducing agent-susceptible AAs, or an AA that is activatable by either or both of enzymatic activation or reducing agent-based activation. Generally, the methods include contacting a plurality of candidate AAs with a target capable of binding an AB of the AAs and a protease capable of cleaving a CM of the AAs, selecting a first population of members of said plurality which bind to the target when exposed to protease, contacting said first population with the target in the absence of the protease, and selecting a second population of members from said first population by depleting from said first population members that bind the target in the absence of the protease, wherein said method provides for selection of candidate AAs which exhibit decreased binding to the target in the absence of the protease as compared to target binding in the presence of the protease.

[00273] In general, the method for screening for candidate AAs having a desired switchable phenotype is accomplished through a positive screening step (to identify members that bind target following exposure to protease) and a negative screening step (to identify members that do not bind target when not exposed to protease). The negative screening step can be accomplished by, for example, depleting from the population members that bind the target in the absence of the protease. It should be noted that the library screening methods described herein can be initiated by conducting the negative screening first to select for candidates that do not bind labeled target in the absence of enzyme treatment (i.e., do not bind labeled target when not cleaved), and then conducting the positive screening (i.e., treating with enzyme and selecting for members which bind labeled target in the cleaved state). However, for convenience, the screening method is described below with the positive selection as a first step.

[00274] The positive and negative screening steps can be conveniently conducted using flow cytometry to sort candidate AAs based on binding of a detectably labeled target. One round or cycle of the screening procedure involves both a positive selection step and a negative selection step. The methods may be repeated for a library such that multiple cycles (including complete and partial cycles, e.g., 1.5 cycles, 2.5

cycles, etc.) are performed. In this manner, members of the plurality of candidate AAs that exhibit the switching characteristics of an AA may be enriched in the resulting population.

[00275] In general, the screening methods are conducted by first generating a nucleic acid library encoding a plurality of candidate AAs in a display scaffold, which is in turn introduced into a display scaffold for expression on the surface of a replicable biological entity. As used herein, a plurality of candidate AAs refers to a plurality of polypeptides having amino acid sequences encoding candidate AAs, where members of the plurality are variable with respect to the amino acid sequence of at least one of the components of an AA, e.g., the plurality is variable with respect to the amino acid sequence of the MM, the CM or the AB, usually the MM.

[00276] For example, the AB and CM of the candidate AAs are held fixed and the candidate AAs in the library are variable with respect to the amino acid sequence of the MM. In another example, a library can be generated to include candidate AAs having an MM that is designed to position a cysteine residue such that disulfide bond formation with another cysteine in the candidate AA is favored (with other residues selected to provide an MM having an amino acid sequence that is otherwise fully or at least partially randomized). In another example, a library can be generated to include candidate AAs in which the MM includes a fully randomized amino acid sequence. Such libraries can contain candidate AAs designed by one or more of these criterion. By screening members of said plurality according to the methods described herein, members having candidate MMs that provide a desired switchable phenotype can be identified.

[00277] In one embodiment of the methods, each member of the plurality of candidate AAs is displayed on the surface of a replicable biological entity (exemplified here by bacterial cells). The members of the plurality are exposed to a protease capable of cleaving the CM of the candidate AAs and contacted with a target which is a binding partner of the AB of the candidate AAs. Bacterial cells displaying members comprising ABs which bind the target after exposure to the protease are identified and/or separated via detection of target binding (e.g., detection of a target-AB complex). Members comprising ABs which bind the target after protease exposure (which can lead to cleavage of the CM) are then contacted with the target in the absence of the protease. Bacterial cells displaying members comprising ABs which exhibit decreased or undetectable binding to the target in the absence of cleavage are identified and/or separated via detection of cells lacking bound target. In this manner, members of the plurality of candidate AAs which bind target in a cleaved state and exhibit decreased or undetectable target binding in an uncleaved state are identified and/or selected.

[00278] As noted above, candidate AA libraries can be constructed so as to screen for one or more aspects of the AA constructs, e.g., to provide for optimization of a switchable phenotype for one or more of the MM, the CM, and the AB. One or more other elements of the AA can be varied to facilitate optimization. For example: vary the MM, including varying the number or position of cysteines or other residues that can provide for different conformational characteristics of the AA in the absence of cleaving agent (e.g., enzyme): vary the CM to identify a substrate that is optimized for one or more desired characteristics

(e.g., specificity of enzyme cleavage, and the like); and/or vary the AB to provide for optimization of switchable target binding.

[00279] In general, the elements of the candidate AA libraries are selected according to a target protein of interest, where the AA is to be activated to provide for enhanced binding of the target in the presence of a cleaving agent (e.g., enzyme) that cleaves the CM. For example, where the CM and AB are held fixed among the library members, the CM is selected such that it is cleavable by a cleaving agent (e.g., enzyme) that is co-localized with a target of interest, where the target of interest is a binding partner of the AB. In this manner, an AA can be selected such that it is selectively activated under the appropriate biological conditions, and thus at an appropriate biological location. For example, where it is desired to develop an AA to be used as an anti-angiogenic compound and exhibit a switchable phenotype for VEGF binding, the CM of the candidate AA is selected to be a substrate for an enzyme and/or a reducing agent that is co-localized with VEGF (e.g., a CM cleavable by a matrix-metalloprotease). By way of another example, where it is desired to develop an AA to be used as an anti-angiogenic compound and exhibit a switchable phenotype for Notch receptor binding, Jagged ligand binding, or EGFR binding, the CM of the candidate AA is selected to be a substrate for an enzyme and/or a reducing agent that is co-localized with the Notch receptor, Jagged ligand, or EGFR (e.g., a CM cleavable by a uPA or plasmin).

[00280] As discussed above, an AB is generally selected according to a target of interest. Many targets are known in the art. Biological targets of interest include protein targets that have been identified as playing a role in disease. Such targets include but are not limited to cell surface receptors and secreted binding proteins (e.g., growth factors), soluble enzymes, structural proteins (e.g. collagen, fibronectin), intracellular targets, and the like. Exemplary non-limiting targets are presented in Table 1, but other suitable targets will be readily identifiable by those of ordinary skill in the art. In addition, many proteases are known in the art which co-localize with targets of interest. As such, persons of ordinary skill in the art will be able to readily identify appropriate enzymes and enzyme substrates for use in the above methods.

(a) Optional enrichment for cell surface display prior to AA screening

[00281] Prior to the screening method, it may be desirable to enrich for cells expressing an appropriate peptide display scaffold on the cell surface. The optional enrichment allows for removal of cells from the cell library that (1) do not express peptide display scaffolds on the cell outer membrane or (2) express non-functional peptide display scaffolds on the cell outer membrane. A non-functional peptide display scaffold does not properly display a candidate AA, e.g., as a result of a stop codon or a deletion mutation.

[00282] Enrichment for cells can be accomplished by growing the cell population and inducing expression of the peptide display scaffolds. The cells are then sorted based on, for example, detection of a detectable signal or moiety incorporated into the scaffold or by use of a detectably-labeled antibody that binds to a shared portion of the display scaffold or the AA. These methods are described in greater detail in U.S. Patent Application Publication No: 2007/0065158, published March 22, 2007.

(b) Screening for target binding by cleaved AAs

[00283] Prior to screening, the candidate AA library can be expanded (e.g., by growth in a suitable medium in culture under suitable conditions). Subsequent to the optional expansion, or as an initial step, the library is subjected to a first screen to identify candidate AAs that bind target following exposure to protease. Accordingly, this step is often referred to herein as the positive selection step.

[00284] In order to identify members that bind target following protease cleavage, the candidate AA library is contacted with a protease capable of cleaving the CM of the displayed candidate AAs for an amount of time sufficient and under conditions suitable to provide for cleavage of the protease substrate of the CM. A variety of protease-CM combinations will be readily ascertainable by those of ordinary skill in the art, where the protease is one which is capable of cleaving the CM and one which co-localizes *in vivo* with a target of interest (which is a binding partner of the AB). For example, where the target of interest is a solid tumor associated target (e.g. VEGF), suitable enzymes include, for example, Matrix-Metalloproteases (e.g., MMP-2), A Disintegrin and Metalloprotease(s) (ADAMs)/ADAM with thrombospondin-like motifs (ADAMTS), Cathepsins and Kallikreins. The amino acid sequences of substrates useful as CMs in the AAs described herein are known in the art and, where desired, can be screened to identify optimal sequences suitable for use as a CM by adaptation of the methods described herein. Exemplary substrates can include but are not limited to substrates cleavable by enzymes listed in Table 3.

[00285] The candidate AA library is also exposed to target for an amount of time sufficient and under conditions suitable for target binding, which conditions can be selected according to conditions under which target binding to the AB would be expected. The candidate AA library can be exposed to the protease prior to exposure to target (e.g., to provide a population of candidate AAs which include cleaved AAs) or in combination with exposure to target, usually the latter so as to best model the expected *in vivo* situation in which both protease and target will be present in the same environmental milieu. Following exposure to both protease and target, the library is then screened to select members having bound target, which include candidate AAs in a target-AB complex.

[00286] Detection of target-bound candidate AAs can be accomplished in a variety of ways. For example, the target may be detectably labeled and the first population of target-bound candidate AAs may be selected by detection of the detectable label to generate a second population having bound target (e.g., a positive selection for target-bound candidate AAs).

(c) Screening for candidate AAs that do not bind target in the absence of protease cleavage

[00287] The population of candidate AAs selected for target binding following exposure to protease can then be expanded (e.g., by growth in a suitable medium in culture under suitable conditions), and the expanded library subjected to a second screen to identify members exhibiting decreased or no detectable

binding to target in the absence of protease exposure. The population resulting from this second screen will include candidate AAs that, when uncleaved, do not bind target significantly or to a detectable level. Accordingly, this step is often referred to herein as the negative selection step.

[00288] The population that resulted from the first screen is contacted with target in the absence of the protease for a time sufficient and under conditions suitable for target binding, which conditions can be selected according to conditions under which target binding to the AB would be expected. A negative selection can then be performed to identify candidate AAs that are relatively decreased for target binding, including those which exhibit no detectably target binding. This selection can be accomplished by, for example, use of a detectably labeled target, and subjecting the target-exposed population to flow cytometry analysis to sort into separate subpopulation those cells that display a candidate AA that exhibits no detectable target binding and/or which exhibit a relatively lower detectable signal. This subpopulation is thus enriched for cells having a candidate AA that exhibit decreased or undetectable binding to target in the absence of cleavage.

(d) Detectable labels

[00289] A detectable label and detectable moiety are used interchangeably to refer to a molecule capable of detection, including, but not limited to, radioactive isotopes, fluorescers, chemilumescers, chromophores, enzymes, enzyme substrates, enzyme cofactors, enzyme inhibitors, chromophores, dyes, metal ions, metal sols, ligands (e.g., biotin, avidin, strepavidin or haptens) and the like. The term fluorescer refers to a substance or a portion thereof which is capable of exhibiting fluorescence in the detectable range. Exemplary detectable moieties suitable for use as target labels include affinity tags and fluorescent proteins.

[00290] The term affinity tag is used herein to denote a peptide segment that can be attached to a target that can be detected using a molecule that binds the affinity tag and provides a detectable signal (e.g., a fluorescent compound or protein). In principal, any peptide or protein for which an antibody or other specific binding agent is available can be used as an affinity tag. Exemplary affinity tags suitable for use include, but are not limited to, a monocytic adaptor protein (MONA) binding peptide, a T7 binding peptide, a streptavidin binding peptide, a polyhistidine tract, protein A (Nilsson et al., EMBO J. 4:1075 (1985); Nilsson et al., Methods Enzymol. 198:3 (1991)), glutathione S transferase (Smith and Johnson, Gene 67:31 (1988)), Glu-Glu affinity tag (Grussenmeyer et al., Proc. Natl. Acad. Sci. USA 82:7952 (1985)), substance P, FLAG peptide (Hopp et al., Biotechnology 6:1204 (1988)), or other antigenic epitope or binding domain. See, in general, Ford et al., Protein Expression and Purification 2:95 (1991). DNA molecules encoding affinity tags are available from commercial suppliers (e.g., Pharmacia Biotech, Piscataway, N.J.).

[00291] Any fluorescent polypeptide (also referred to herein as a fluorescent label) well known in the art is suitable for use as a detectable moiety or with an affinity tag of the peptide display scaffolds described herein. A suitable fluorescent polypeptide will be one that can be expressed in a desired host cell, such as

a bacterial cell or a mammalian cell, and will readily provide a detectable signal that can be assessed qualitatively (positive/negative) and quantitatively (comparative degree of fluorescence). Exemplary fluorescent polypeptides include, but are not limited to, yellow fluorescent protein (YFP), cyan fluorescent protein (CFP), GFP, mRFP, RFP (tdimer2), HCRED, etc., or any mutant (e.g., fluorescent proteins modified to provide for enhanced fluorescence or a shifted emission spectrum), analog, or derivative thereof. Further suitable fluorescent polypeptides, as well as specific examples of those listed herein, are provided in the art and are well known.

[00292] Biotin-based labels also find use in the methods disclosed herein. Biotinylation of target molecules and substrates is well known, for example, a large number of biotinylation agents are known, including amine-reactive and thiol-reactive agents, for the biotinylation of proteins, nucleic acids, carbohydrates, carboxylic acids; see, e.g., chapter 4, Molecular Probes Catalog, Haugland, 6th Ed. 1996, hereby incorporated by reference. A biotinylated substrate can be detected by binding of a detectably labeled biotin binding partner, such as avidin or streptavidin. Similarly, a large number of haptenylation reagents are also known.

(e) Screening methods

[00293] Any suitable method that provides for separation and recovery of AAs of interest may be utilized. For example, a cell displaying an AA of interest may be separated by FACS, immunochromatography or, where the detectable label is magnetic, by magnetic separation. As a result of the separation, the population is enriched for cells that exhibit the desired characteristic, e.g., exhibit binding to target following cleavage or have decreased or no detectable binding to target in the absence of cleavage.

[00294] For example, selection of candidate AAs having bound detectably labeled target can be accomplished using a variety of techniques known in the art. For example, flow cytometry (e.g., FACS®) methods can be used to sort detectably labeled candidate AAs from unlabeled candidate AAs. Flow cyometry methods can be implemented to provide for more or less stringent requirements in separation of the population of candidate AAs, e.g., by modification of gating to allow for dimmer or to require brighter cell populations in order to be separated into the second population for further screening.

[00295] In another example, immunoaffinity chromatography can be used to separate target-bound candidate AAs from those that do not bind target. For example, a support (e.g., column, magnetic beads) having bound anti-target antibody can be contacted with the candidate AAs that have been exposed to protease and to target. Candidate AAs having bound target bind to the anti-target antibody, thus facilitating separation from candidate AAs lacking bound target. Where the screening step is to provide for a population enriched for uncleaved candidate AAs that have relatively decreased target binding or no detectable target binding (e.g., relative to other candidate AAs), the subpopulation of interest is those members that lack or have a relatively decreased detectably signal for bound target. For example, where an immunoaffinity technique is used in such negative selection for bound target, the subpopulation of interest is that which is not bound by the anti-target support.

(f) Screening for dual target-binding AAs

[00296] Methods for screening disclosed herein can be readily adapted to identify dual target-binding AAs having two ABs. In general, the method involves a library containing a plurality of candidate AAs, wherein each member of said plurality comprises a first AB, a second AB, a first CM and/or a second CM, a first MM, and/or a second MM. The library is contacted with target capable of binding at least the first AB and a cleaving agent capable of cleaving the first CM. A first population of members of the library is selected for binding the target in the presence of the cleaving agent (e.g., protease for the CM). This selected population is then subjected to the negative screen above, in which binding of target to the library members in the absence of the cleaving agent is assessed. A second population of members is then generated by depleting the subpopulation of members that bind to said target in the absence of the cleaving agent. This can be accomplished by, for example, sorting members that are not bound to target away from those that are bound to target, as determined by detection of a detectably labeled target. This method thus provides for selection of candidate AAs which exhibit decreased binding to the target in the absence of the cleaving agent as compared to binding to said target in the presence of the cleaving agent. This method can be repeated for both targets.

Exemplary variations of the screening methods to select for candidate AAs

[00297] The above methods may be modified to select for populations and library members that demonstrate desired characteristics.

(a) Determination of the masking efficiency of MMs

[00298] Masking efficiency of MMs is determined by at least two parameters: affinity of the MM for antibody or fragment thereof and the spatial relationship of the MM relative to the binding interface of the AB to its target.

[00299] Regarding affinity, by way of example, an MM may have high affinity but only partially inhibit the binding site on the AB, while another MM may have a lower affinity for the AB but fully inhibit target binding. For short time periods, the lower affinity MM may show sufficient masking; in contrast, over time, that same MM may be displaced by the target (due to insufficient affinity for the AB).

[00300] In a similar fashion, two MMs with the same affinity may show different extents of masking based on how well they promote inhibition of the binding site on the AB or prevention of the AB from binding its target. In another example, a MM with high affinity may bind and change the structure of the AB so that binding to its target is completely inhibited while another MM with high affinity may only partially inhibit binding. As a consequence, discovery of an effective MM cannot be based only on affinity but can include an empirical measure of masking efficiency. The time-dependent target displacement of the MM in the AA can be measured to optimize and select for MMs. A novel Target Displacement Assay (TDA) is described herein for this purpose.

[00301] The TDA assay can be used for the discovery and validation of efficiently masked AAs comprises empirical determination of masking efficiency, comparing the ability of the masked AB to bind the target in the presence of target to the ability of the unmasked and/or parental AB to bind the target in the presence of the target. The binding efficiency can be expressed as a % of equilibrium binding, as compared to unmasked/parental AB binding. When the AB is modified with a MM and is in the presence of the target, specific binding of the AB to its target can be reduced or inhibited, as compared to the specific binding of the AB not modified with an MM or the parental AB to the target. When compared to the binding of the AB not modified with an MM or the parental AB to the target, the AB's ability to bind the target when modified with an MM can be reduced by at least 50%, 60%, 70%, 80%, 90%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% and even 100% for at least 2, 4, 6, 8, 12, 28, 24, 30, 36, 48, 60, 72, 84, 96, hours, or 5, 10, 15, 30, 45, 60, 90, 120, 150, 180 days, or 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12 months or greater when measured *in vivo* or in a Target Displacement *in vitro* immunosorbant assay, as described herein.

(b) Iterative screens to identify and/or optimize AA elements

[00302] The methods and candidate AA libraries described herein can be readily adapted to provide for identification and/or optimization of one or more elements of an AA. For example, candidate AAs that vary with respect to any one or more of AB, CM, linkers, and the like can be produced and subjected to the screening methods described herein.

(c) Reducing agent-activatable AAs

[00303] While the methods above describe screening methods for identifying AAs, it should be understood that an AA or candidate AA with a CM that can facilitate formation of a cysteine-cysteine disulfide bond in an AA and can also be subjected to the screening methods disclosed herein. Such AAs may or may not further include a CM (which may be the same or different CM) that may or may not comprise a protease substrate. In these embodiments, the positive screen described above may be conducted by exposing an AA or candidate AA to a reducing agent (e.g., to reducing conditions) capable of cleaving the disulfide bond of the cysteine-cysteine pair of the AA. The negative screen can then be conducted in the absence of the reducing conditions. As such, a library produced having may be enriched for AAs which are activatable by exposure to disulfide bond reducing conditions.

(d) Photo-activatable AAs

[00304] While the methods above describe screening methods for identifying AAs, it should be understood that an AA or candidate AA with a CM that is photo-sensitive, and can be activated upon photolysis are also provided. In these embodiments, the positive screen described above may be conducted by exposing an AA or candidate AA to light. The negative screen can then be conducted in the absence of light. As such, a library produced having may be enriched for AAs which are activatable by

exposure to light.

(e) Number of cycles and scaffold free screening of AAs

[00305] By increasing the number of cycles of the above methods, populations and library members that demonstrate improved switching characteristics can be identified. Any number of cycles of screening can be performed.

[00306] In addition, individual clones of candidate AAs can be isolated and subjected to screening so as to determine the dynamic range of the candidate AA. Candidate AAs can also be tested for a desired switchable phenotype separate from the scaffold, i.e., the candidate AA can be expressed or otherwise generated separate from the display scaffold, and the switchable phenotype of the candidate AA assessed in the absence of the scaffold and, where desired, in a cell-free system (e.g., using solubilized AA).

(f) Optimization of AA components and switching activity

[00307] The above methods may be modified to optimize the performance of an AA, e.g., an AA identified in the screening method described herein. For example, where it is desirable to optimize the performance of the masking moiety, e.g., to provide for improved inhibition of target binding of the AB in the uncleaved state, the amino acid sequences of the AB and the CM may be fixed in a candidate AA library, and the MM varied such that members of a library have variable MMs relative to each other. The MM may be optimized in a variety of ways including alteration in the number and or type of amino acids that make up the MM. For example, each member of the plurality of candidate AAs may comprise a candidate MM, wherein the candidate MM comprises at least one cysteine amino acid residue and the remaining amino acid residues are variable between the members of the plurality. In a further example, each member of the plurality of candidate AAs may comprise a candidate MM, wherein the candidate MM comprises a cysteine amino acid residue and a random sequence of amino acid residues, e.g., a random sequence of 5 amino acids.

(g) Selection for expanded dynamic range

[00308] As noted above, AAs having a desired dynamic range with respect to target binding in the unmasked/cleaved versus masked/uncleaved state are also of interest. Such AAs are those that, for example, have no detectable binding in the presence of target at physiological levels found at treatment and non-treatment sites in a subject but which, once cleaved by protease, exhibit high affinity and/or high avidity binding to target. The greater the dynamic range of an AA, the better the switchable phenotype of the AA. Thus AAs can be optimized to select for those having an expanded dynamic range for target binding in the presence and absence of a cleaving agent.

[00309] The screening methods described herein can be modified so as to enhance selection of AAs having a desired and/or optimized dynamic range. In general, this can be accomplished by altering the concentrations of target utilized in the positive selection and negative selection steps of the method such

that screening for target binding of AAs exposed to protease (i.e., the screening population that includes cleaved AAs) is performed using a relatively lower target concentration than when screening for target binding of uncleaved AAs. Accordingly, the target concentration is varied between the steps so as to provide a selective pressure toward a switchable phenotype. Where desired, the difference in target concentrations used at the positive and negative selection steps can be increased with increasing cycle number.

[00310] Use of a relatively lower concentration of target in the positive selection step can serve to drive selection of those AA members that have improved target binding when in the cleaved state. For example, the screen involving protease-exposed AAs can be performed at a target concentration that is from about 2 to about 100 fold lower, about 2 to 50 fold lower, about 2 to 20 fold lower, about 2 to 10-fold lower, or about 2 to 5-fold lower than the K_d of the AB- target interaction. As a result, after selection of the population for target-bound AAs, the selected population will be enriched for AAs that exhibit higher affinity and/or avidity binding relative to other AAs in the population.

[00311] Use of a relatively higher concentration of target in the negative selection step can serve to drive selection of those AA members that have decreased or no detectable target binding when in the uncleaved state. For example, the screen involving AAs that have not been exposed to protease (in the negative selection step) can be performed at a target concentration that is from about 2 to about 100 fold higher, about 2 to 50 fold higher, about 2 to 20 fold higher, about 2 to 10-fold higher, or about 2 to 5-fold higher, than the K_d of the AB-target interaction. As a result, after selection of the population for AAs that do not detectably bind target, the selected population will be enriched for AAs that exhibit lower binding for target when in the uncleaved state relative to other uncleaved AAs in the population. Stated differently, after selection of the population for AAs that do not detectably bind target, the selected population will be enriched for AAs for which target binding to AB is inhibited, e.g., due to masking of the AB from target binding.

[00312] Where the AA is a dual target-binding AA, the screening method described above can be adapted to provide for AAs having a desired dynamic range for a first target that is capable of binding a first AB and for a second target that is capable of binding a second AB. Target binding to an AB that is located on a portion of the AA that is cleaved away from the AA presented on a display scaffold can be evaluated by assessing formation of target-AB complexes in solution (e.g., in the culture medium), e.g., immunochromatography having an anti-AA fragment antibody to capture cleaved fragment, then detecting bound, detectably labeled target captured on the column.

(h) Testing of soluble AAs

[00313] Candidate AAs can be tested for their ability to maintain a switchable phenotype while in soluble form. One such method involves the immobilization of target to support (e.g., an array, e.g., a Biacore™ CM5 sensor chip surface). Immobilization of a target of interest can be accomplished using any suitable techniques (e.g., standard amine coupling). The surface of the support can be blocked to reduce non-

specific binding. Optionally, the method can involve use of a control (e.g., a support that does not contain immobilized target (e.g., to assess background binding to the support) and/or contains a compound that serves as a negative control (e.g., to assess specificity of binding of the candidate AA to target versus non-target).

[00314] After the target is covalently immobilized, the candidate AA is contacted with the support under conditions suitable to allow for specific binding to immobilized target. The candidate AA can be contacted with the support-immobilized target in the presence and in the absence of a suitable cleavage agent in order to assess the switchable phenotype. Assessment of binding of the candidate AA in the presence of cleavage agent as compared to in the absence of cleavage agent and, optionally, compared to binding in a negative control provides a binding response, which in turn is indicative of the switchable phenotype.

(i) Screening for individual moieties for use in candidate AAs

[00315] It may be desirable to screen separately for one or more of the moieties of a candidate AA, e.g., an AB, MM or CM, prior to testing the candidate AA for a switchable phenotype. For example, known methods of identifying peptide substrates cleavable by specific proteases can be utilized to identify CMs for use in AAs designed for activation by such proteases. In addition a variety of methods are available for identifying peptide sequences which bind to a target of interest. These methods can be used, for example, to identify ABs which binds to a particular target or to identify a MM which binds to a particular AB.

[00316] The above methods include, for example, methods in which a moiety of a candidate AA, e.g., an AB, MM or CM, is displayed using a replicable biological entity.

(j) Automated Screening Methods

[00317] In certain embodiments the screening methods described herein are automated to provide convenient, real time, high volume methods of screening a library of AAs for a desired switchable activity. Automated methods can be designed to provide for iterative rounds of positive and negative selection, with the selected populations being separated and automatically subjected to the next screen for a desired number of cycles.

[00318] Assessing candidate AAs in a population may be carried out over time iteratively, following completion of a positive selection step, a negative selection step, or both. In addition, information regarding the average dynamic range of a population of candidate AAs at selected target concentrations in the positive and negative selection steps can be monitored and stored for later analysis, e.g. so as to assess the effect of selective pressure of the different target concentrations.

[00319] In some embodiments, a executable platform such as a computer software product can control operation of the detection and/or measuring means and can perform numerical operations relating to the above-described steps, and generate a desired output (e.g., flow cytometry analysis, etc.). Computer

program product comprises a computer readable storage medium having computer-readable program code means embodied in the medium. Hardware suitable for use in such automated apparatus will be apparent to those of skill in the art, and may include computer controllers, automated sample handlers, fluorescence measurement tools, printers and optical displays. The measurement tool may contain one or more photodetectors for measuring the fluorescence signals from samples where fluorescently detectable molecules are utilized. The measurement tool may also contain a computer-controlled stepper motor so that each control and/or test sample can be arranged as an array of samples and automatically and repeatedly positioned opposite a photodetector during the step of measuring fluorescence intensity.

[00320] The measurement tool (e.g., FACS) can be operatively coupled to a general purpose or application-specific computer controller. The controller can comprise a computer program produce for controlling operation of the measurement tool and performing numerical operations relating to the above-described steps. The controller may accept set-up and other related data via a file, disk input or data bus. A display and printer may also be provided to visually display the operations performed by the controller. It will be understood by those having skill in the art that the functions performed by the controller may be realized in whole or in part as software modules running on a general purpose computer system. Alternatively, a dedicated stand-alone system with application specific integrated circuits for performing the above described functions and operations may be provided.

Methods of Use of AAs in Therapy

[00321] AAs can be incorporated into pharmaceutical compositions containing, for example, a therapeutically effective amount of an AA of interest and a carrier that is a pharmaceutically acceptable excipient (also referred to as a pharmaceutically acceptable carrier). Many pharmaceutically acceptable excipients are known in the art, are generally selected according to the route of administration, the condition to be treated, and other such variables that are well understood in the art. Pharmaceutically acceptable excipients have been amply described in a variety of publications, including, for example, A. Gennaro (2000) Remington: The Science and Practice of Pharmacy, 20th edition, Lippincott, Williams, & Wilkins; Pharmaceutical Dosage Forms and Drug Delivery Systems (1999) H.C. Ansel et al., eds., 7th ed., Lippincott, Williams, & Wilkins; and Handbook of Pharmaceutical Excipients (2000) A.H. Kibbe et al., eds., 3rd ed. Amer. Pharmaceutical Assoc. Pharmaceutical compositions can also include other components such as pH adjusting and buffering agents, tonicity adjusting agents, stabilizers, wetting agents and the like. In some embodiments, nanoparticles or liposomes carry a pharmaceutical composition comprising an AA.

[00322] Suitable components for pharmaceutical compositions of AAs can be guided by pharmaceutical compositions that may be already available for an AB of the AA. For example, where the, the AA includes an antibody to EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130,

VCAM-1, CD44, DLL4, or IL4, for example, such AAs can be formulated in a pharmaceutical formulation according to methods and compositions suitable for use with that antibody.

[00323] In general, pharmaceutical formulations of one or more AAs are prepared for storage by mixing the AA having a desired degree of purity with optional physiologically acceptable carriers, excipients or stabilizers (Remington's Pharmaceutical Sciences 16th edition, Osol, A. Ed. (1980)), in the form of lyophilized formulations or aqueous solutions. Acceptable carriers, excipients, or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid and methionine; preservatives (such as octadecyldimethylbenzyl ammonium chloride; hexamethonium chloride; benzalkonium chloride, benzethonium chloride; phenol, butyl or benzyl alcohol; alkyl parabens such as methyl or propyl paraben; catechol; resorcinol; cyclohexanol; 3-pentanol; and m-cresol); low molecular weight (less than about 10 residues) polypeptide; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, histidine, arginine, or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugars such as sucrose, mannitol, trehalose or sorbitol; salt-forming counter-ions such as sodium; metal complexes (e.g., Zn-protein complexes); and/or non-ionic surfactants such as TWEENTM, PLURONICSTM or polyethylene glycol (PEG).

[00324] The formulations to be used for *in vivo* administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes. Pharmaceutical formulations may also contain more than one active compound as necessary for the particular indication being treated, where the additional active compounds generally are those with activities complementary to an AA. Such compounds are suitably present in combination in amounts that are effective for the purpose intended.

[00325] The pharmaceutical formulation can be provided in a variety of dosage forms such as a systemically or local injectable preparation. The components can be provided in a carrier such as a microcapsule, e.g., such as that prepared by coacervation techniques or by interfacial polymerization, for example, hydroxymethylcellulose or gelatin-microcapsule and poly-(methylmethacrylate) microcapsule, respectively, in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules) or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences 16th edition, Osol, A. Ed. (1980).

[00326] Sustained-release preparations are also within the scope of an AA-containing formulations. Exemplary sustained-release preparations can include semi-permeable matrices of solid hydrophobic polymers containing the AA, which matrices are in the form of shaped articles, e.g., films, or microcapsule. Examples of sustained-release matrices include polyesters, hydrogels (for example, poly(2-hydroxyethyl-methacrylate), or poly(vinylalcohol)), polylactides (U.S. Pat. No. 3,773,919), copolymers of L-glutamic acid and γ -ethyl-L-glutamate, non-degradable ethylene-vinyl acetate, degradable lactic acid-glycolic acid copolymers such as the LUPRON DEPOTTM (injectable microspheres composed of lactic acid-glycolic acid copolymer and leuprolide acetate), and poly-D-(-)-3-hydroxybutyric

acid. While polymers such as ethylene-vinyl acetate and lactic acid-glycolic acid enable release of molecules for over 100 days, certain hydrogels release proteins for shorter time periods.

[00327] When encapsulated AAs remain in the body for a long time, they may denature or aggregate as a result of exposure to moisture at physiological temperature ($\sim 37^{\circ}\text{C}$), resulting in decreased biological activity and possible changes in immunogenicity. Rational strategies can be devised for stabilization depending on the mechanism involved. For example, if the aggregation mechanism is discovered to be undesirable intermolecular S-S bond formation through thio-disulfide interchange, stabilization may be achieved by modifying sulfhydryl residues, lyophilizing from acidic solutions, controlling moisture content, using appropriate additives, and developing specific polymer matrix compositions.

[00328] AAs can be conjugated to delivery vehicles for targeted delivery of an active agent that serves a therapeutic purpose. For example, AAs can be conjugated to nanoparticles or liposomes having drugs encapsulated therein or associated therewith. In this manner, specific, targeted delivery of the drug can be achieved. Methods of linking polypeptides to liposomes are well known in the art and such methods can be applied to link AAs to liposomes for targeted and or selective delivery of liposome contents. By way of example, polypeptides can be covalently linked to liposomes through thioether bonds. PEGylated gelatin nanoparticles and PEGylated liposomes have also been used as a support for the attachment of polypeptides, e.g., single chain antibodies. See, e.g., Immordino et al. (2006) *Int J Nanomedicine*. September; 1(3): 297–315, incorporated by reference herein for its disclosure of methods of conjugating polypeptides, e.g., antibody fragments, to liposomes.

(a) Methods of treatment

[00329] AAs described herein can be selected for use in methods of treatment of suitable subjects according to the CM-AB combination provided in the AA. The AA can be administered by any suitable means, including oral, parenteral, subcutaneous, intraperitoneal, intrapulmonary, and intranasal, and, if desired for local injection (e.g., at the site of a solid tumor). Parenteral administration routes include intramuscular, intravenous, intraarterial, intraperitoneal, or subcutaneous administration.

[00330] The term treatment site or disease site is meant to refer to a site at which an AA is designed to be switchable, as described herein, e.g., a site at which a target for one or both ABs of an AA and a cleaving agent capable of cleaving a CM of the AA are co-localized, as pictorially represented in Figure 2. Treatment sites include tissues that can be accessed by local administration (e.g., injection, infusion (e.g., by catheter), etc.) or by systemic administration (e.g., administration to a site remote from a treatment site). Treatment sites include those that are relatively biologically confined (e.g., an organ, sac, tumor site, and the like).

[00331] The appropriate dosage of an AA will depend on the type of disease to be treated, the severity and course of the disease, the patient's clinical history and response to the AA, and the discretion of the attending physician. AAs can suitably be administered to the patient at one time or over a series of

treatments. AAs can be administered along with other treatments and modes of therapies, other pharmaceutical agents, and the like.

[00332] Depending on the type and severity of the disease, about 1 $\mu\text{g/kg}$ to 15 mg/kg (e.g., 0.1-20 mg/kg) of an AA can serve as an initial candidate dosage for administration to the patient, whether, for example, by one or more separate administrations, or by continuous infusion. A typical daily dosage might range from about 1 $\mu\text{g/kg}$ to 100 mg/kg or more, depending on factors such as those mentioned herein. For repeated administrations over several days or longer, depending on the condition, the treatment is sustained until a desired suppression of disease symptoms occurs. However, other dosage regimens may be useful.

[00333] The AA composition will be formulated, dosed, and administered in a fashion consistent with good medical practice. Factors for consideration in this context include the particular disorder being treated, the particular mammal being treated, the clinical condition of the individual patient, the cause of the disorder, the site of delivery of the AA, the method of administration, the scheduling of administration, and other factors known to medical practitioners. The therapeutically effective amount of an AA to be administered will be governed by such considerations, and is the minimum amount necessary to prevent, ameliorate, or treat a disease or disorder.

[00334] Generally, alleviation or treatment of a disease or disorder involves the lessening of one or more symptoms or medical problems associated with the disease or disorder. For example, in the case of cancer, the therapeutically effective amount of the drug can accomplish one or a combination of the following: reduce the number of cancer cells; reduce the tumor size; inhibit (i.e., to decrease to some extent and/or stop) cancer cell infiltration into peripheral organs; inhibit tumor metastasis; inhibit, to some extent, tumor growth; and/or relieve to some extent one or more of the symptoms associated with the cancer. In some embodiments, a composition of this invention can be used to prevent the onset or reoccurrence of the disease or disorder in a subject or mammal.

[00335] AAs can be used in combination (e.g., in the same formulation or in separate formulations) with one or more additional therapeutic agents or treatment methods (combination therapy). An AA can be administered in admixture with another therapeutic agent or can be administered in a separate formulation. Therapeutic agents and/or treatment methods that can be administered in combination with an AA, and which are selected according to the condition to be treated, include surgery (e.g., surgical removal of cancerous tissue), radiation therapy, bone marrow transplantation, chemotherapeutic treatment, certain combinations of the foregoing, and the like.

(b) Use of AAs in diseased tissue versus healthy tissue

[00336] The AAs of the present invention, when localized to a healthy tissue, show little or no activation and the AB remains in a 'masked' state, or otherwise exhibits little or no binding to the target. However, in a diseased tissue, in the presence of a disease-specific protease, for example, capable of cleaving the CM of the AA, the AB becomes 'unmasked' or can specifically bind the target.

[00337] A healthy tissue refers to a tissue that produces little or no disease-specific agent capable of specifically cleaving or otherwise modifying the CM of the AA, for example a disease-specific protease, a disease-specific enzyme, or a disease-specific reducing agent. A diseased tissue refers to a tissue that produces a disease-specific agent capable of specifically cleaving or otherwise modifying the CM of the AA, for example a disease-specific protease, a disease-specific enzyme, or a disease-specific reducing agent.

(c) Use of AAs in diseased tissue at different stages of a disease

[00338] In some embodiments, the AAs described herein are coupled to more than one CM. Such an AA can be activated in different stages of a disease, or activated in different compartments of the diseased tissue. By way of example, an AB coupled to both a MMP-9 cleavable CM and a cathepsin D-cleavable CM can be activated in an early stage tumor and in a late stage, necrosing tumor. In the early stage tumor, the CM can be cleaved and the AA unmasked by MMP-9. In the late stage tumor, the CM can be cleaved and the AA unmasked by cathepsin D which is upregulated in the dying center of late stage tumors. In another exemplary embodiment an AB coupled to an MM and to a MMP-9-activatable CM and a caspase-activatable CM can be cleaved at both early and late stage tumors. In another plasmin at active sites of angiogenesis (early stage tumor) can cleave a plasmin-cleavable CM and legumain in disease tissues with invading macrophages can cleave a leugamain-specific CM in a late stage tumor.

(d) Use of AAs in anti-angiogenic therapies

[00339] In an exemplary embodiment where the AA contains an AB that binds a mediator of angiogenesis such as EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4, the AA finds use in treatment of conditions in which inhibition of angiogenesis is desired, particularly those conditions in which inhibition of VEGF is of interest. VEGF-binding AAs can include dual target binding AAs having an AB that binds to VEGF as well as an AB that binds to a second growth factor, such as a fibroblast growth factor (e.g., FGF-2), and inhibits FGF activity. Such dual target binding AAs thus can be designed to provide for inhibition of two angiogenesis-promoting factors, and which are activatable by a cleaving agent (e.g., enzyme, such as a MMP or other enzymes such as one presented in Table 3) which co-localizes at a site of aberrant angiogenesis.

[00340] Angiogenesis-inhibiting AAs find use in treatment of solid tumors in a subject (e.g., human), particularly those solid tumors that have an associated vascular bed that feeds the tumor such that inhibition of angiogenesis can provide for inhibition or tumor growth. Anti-VEGF-based anti-angiogenesis AAs also find use in other conditions having one or more symptoms amenable to therapy by inhibition of abnormal angiogenesis.

[00341] In general, abnormal angiogenesis occurs when new blood vessels either grow excessively, insufficiently or inappropriately (e.g., the location, timing or onset of the angiogenesis being undesired

from a medical standpoint) in a diseased state or such that it causes a diseased state. Excessive, inappropriate or uncontrolled angiogenesis occurs when there is new blood vessel growth that contributes to the worsening of the diseased state or causes a diseased state, such as in cancer, especially vascularized solid tumors and metastatic tumors (including colon, lung cancer (especially small-cell lung cancer), or prostate cancer), diseases caused by ocular neovascularisation, especially diabetic blindness, retinopathies, primarily diabetic retinopathy or age-induced macular degeneration and rubeosis; psoriasis, psoriatic arthritis, haemangioblastoma such as haemangioma; inflammatory renal diseases, such as glomerulonephritis, especially mesangioproliferative glomerulonephritis, haemolytic uremic syndrome, diabetic nephropathy or hypertensive nephrosclerosis; various inflammatory diseases, such as arthritis, especially rheumatoid arthritis, inflammatory bowel disease, psoriasis, sarcoidosis, arterial arteriosclerosis and diseases occurring after transplants, endometriosis or chronic asthma and other conditions that will be readily recognized by the ordinarily skilled artisan. The new blood vessels can feed the diseased tissues, destroy normal tissues, and in the case of cancer, the new vessels can allow tumor cells to escape into the circulation and lodge in other organs (tumor metastases).

[00342] AA-based anti-angiogenesis therapies can also find use in treatment of graft rejection, lung inflammation, nephrotic syndrome, preeclampsia, pericardial effusion, such as that associated with pericarditis, and pleural effusion, diseases and disorders characterized by undesirable vascular permeability, e.g., edema associated with brain tumors, ascites associated with malignancies, Meigs' syndrome, lung inflammation, nephrotic syndrome, pericardial effusion, pleural effusion, permeability associated with cardiovascular diseases such as the condition following myocardial infarctions and strokes and the like.

[00343] Other angiogenesis-dependent diseases that may be treated using anti-angiogenic AAs as described herein include angiofibroma (abnormal blood of vessels which are prone to bleeding), neovascular glaucoma (growth of blood vessels in the eye), arteriovenous malformations (abnormal communication between arteries and veins), nonunion fractures (fractures that will not heal), atherosclerotic plaques (hardening of the arteries), pyogenic granuloma (common skin lesion composed of blood vessels), scleroderma (a form of connective tissue disease), hemangioma (tumor composed of blood vessels), trachoma (leading cause of blindness in the third world), hemophilic joints, vascular adhesions and hypertrophic scars (abnormal scar formation).

[00344] Amounts of an AA for administration to provide a desired therapeutic effect will vary according to a number of factors such as those discussed above. In general, in the context of cancer therapy, a therapeutically effective amount of an AA is an amount that is effective to inhibit angiogenesis, and thereby facilitate reduction of, for example, tumor load, atherosclerosis, in a subject by at least about 5%, at least about 10%, at least about 20%, at least about 25%, at least about 50%, at least about 75%, at least about 85%, or at least about 90%, up to total eradication of the tumor, when compared to a suitable control. In an experimental animal system, a suitable control may be a genetically identical animal not

treated with the agent. In non-experimental systems, a suitable control may be the tumor load present before administering the agent. Other suitable controls may be a placebo control.

[00345] Whether a tumor load has been decreased can be determined using any known method, including, but not limited to, measuring solid tumor mass; counting the number of tumor cells using cytological assays; fluorescence-activated cell sorting (e.g., using antibody specific for a tumor-associated antigen) to determine the number of cells bearing a given tumor antigen; computed tomography scanning, magnetic resonance imaging, and/or x-ray imaging of the tumor to estimate and/or monitor tumor size; measuring the amount of tumor-associated antigen in a biological sample, e.g., blood or serum; and the like.

[00346] In some embodiments, the methods are effective to reduce the growth rate of a tumor by at least about 5%, at least about 10%, at least about 20%, at least about 25%, at least about 50%, at least about 75%, at least about 85%, or at least about 90%, up to total inhibition of growth of the tumor, when compared to a suitable control. Thus, in these embodiments, effective amounts of an AA are amounts that are sufficient to reduce tumor growth rate by at least about 5%, at least about 10%, at least about 20%, at least about 25%, at least about 50%, at least about 75%, at least about 85%, or at least about 90%, up to total inhibition of tumor growth, when compared to a suitable control. In an experimental animal system, a suitable control may be tumor growth rate in a genetically identical animal not treated with the agent. In non-experimental systems, a suitable control may be the tumor load or tumor growth rate present before administering the agent. Other suitable controls may be a placebo control.

[00347] Whether growth of a tumor is inhibited can be determined using any known method, including, but not limited to, an *in vivo* assay for tumor growth; an *in vitro* proliferation assay; a ³H-thymidine uptake assay; and the like.

(e) Use of AAs in anti-inflammatory therapies

[00348] In another exemplary embodiment where the AA contains an AB that binds mediators of inflammation such as interleukins, the AA finds use in treatment of related conditions. Interleukin-binding AAs can include dual target binding AAs having an AB that binds to for example IL12 as well as an AB that binds to IL23, or an AA where a first AB binds to IL17 and a second AB binds to IL23. Such dual target binding AAs thus can be designed to provide for mediation of inflammation, and which are activatable by a cleaving agent (e.g., enzyme, such as a MMP or other enzyme such as one presented in Table 3) which co-localizes at a site of inflammation.

Non-Therapeutic Methods of Using AAs

[00349] AAs can also be used in diagnostic and/or imaging methods. For example, AAs having an enzymatically cleavable CM can be used to detect the presence or absence of an enzyme that is capable of cleaving the CM. Such AAs can be used in diagnostics, which can include *in vivo* detection (e.g., qualitative or quantitative) of enzyme activity (or, in some embodiments, an environment of increased reduction potential such as that which can provide for reduction of a disulfide bond) accompanied by

presence of a target of interest through measured accumulation of activated AAs in a given tissue of a given host organism.

[00350] For example, the CM can be selected to be a protease substrate for a protease found at the site of a tumor, at the site of a viral or bacterial infection at a biologically confined site (e.g., such as in an abscess, in an organ, and the like), and the like. The AB can be one that binds a target antigen. Using methods familiar to one skilled in the art, a detectable label (e.g., a fluorescent label) can be conjugated to an AB or other region of an AA. Suitable detectable labels are discussed in the context of the above screening methods and additional specific examples are provided below. Using an AB specific to a protein or peptide of the disease state, along with a protease whose activity is elevated in the disease tissue of interest, AAs will exhibit increased rate of binding to disease tissue relative to tissues where the CM specific enzyme is not present at a detectable level or is present at a lower level than in disease tissue. Since small proteins and peptides are rapidly cleared from the blood by the renal filtration system, and because the enzyme specific for the CM is not present at a detectable level (or is present at lower levels in non-diseased tissues), accumulation of activated AA in the diseased tissue is enhanced relative to non-disease tissues.

[00351] In another example, AAs can be used in to detect the presence or absence of a cleaving agent in a sample. For example, where the AA contains a CM susceptible to cleavage by an enzyme, the AA can be used to detect (either qualitatively or quantitatively) the presence of an enzyme in the sample. In another example, where the AA contains a CM susceptible to cleavage by reducing agent, the AA can be used to detect (either qualitatively or quantitatively) the presence of reducing conditions in a sample. To facilitate analysis in these methods, the AA can be detectably labeled, and can be bound to a support (e.g., a solid support, such as a slide or bead). The detectable label can be positioned on a portion of the AA that is released following cleavage. The assay can be conducted by, for example, contacting the immobilized, detectably labeled AA with a sample suspected of containing an enzyme and/or reducing agent for a time sufficient for cleavage to occur, then washing to remove excess sample and contaminants. The presence or absence of the cleaving agent (e.g., enzyme or reducing agent) in the sample is then assessed by a change in detectable signal of the AA prior to contacting with the sample (e.g., a reduction in detectable signal due to cleavage of the AA by the cleaving agent in the sample and the removal of an AA fragment to which the detectable label is attached as a result of such cleavage).

[00352] Such detection methods can be adapted to also provide for detection of the presence or absence of a target that is capable of binding the AB of the AA. Thus, the *in vitro* assays can be adapted to assess the presence or absence of a cleaving agent and the presence or absence of a target of interest. The presence or absence of the cleaving agent can be detected by a decrease in detectable label of the AA as described above, and the presence or absence of the target can be detected by detection of a target-AB complex, e.g., by use of a detectably labeled anti-target antibody.

[00353] As discussed above, the AAs disclosed herein can comprise a detectable label. In one embodiment, the AA comprises a detectable label which can be used as a diagnostic agent. Non-limiting

examples of detectable labels that can be used as diagnostic agents include imaging agents containing radioisotopes such as indium or technetium; contrasting agents for MRI and other applications containing iodine, gadolinium or iron oxide; enzymes such as horse radish peroxidase, alkaline phosphatase, or β -galactosidase; fluorescent substances and fluorophores such as GFP, europium derivatives; luminescent substances such as N-methylacrydium derivatives or the like.

[00354] The rupture of vulnerable plaque and the subsequent formation of a blood clot are believed to cause the vast majority of heart attacks. Effective targeting of vulnerable plaques can enable the delivery of stabilizing therapeutics to reduce the likelihood of rupture.

[00355] VCAM-1 is upregulated both in regions prone to atherosclerosis as well as at the borders of established lesions. Iiyama, et al. (1999) *Circulation Research, Am Heart Assoc.* 85: 199-207.

Collagenases, such as MMP-1, MMP-8 and MMP-13, are overexpressed in human atheroma which may contribute to the rupture of atheromatous plaques. Fricker, J. (2002) *Drug Discov Today* 7(2): 86-88.

[00356] In one example, AAs disclosed herein find use in diagnostic and/or imaging methods designed to detect and/or label atherosclerotic plaques, e.g., vulnerable atherosclerotic plaques. By targeting proteins associated with atherosclerotic plaques, AAs can be used to detect and/or label such plaques. For example, AAs comprising an anti-VCAM-1 AB and a detectable label find use in methods designed to detect and/or label atherosclerotic plaques. These AAs can be tested in animal models, such as ApoE mice.

Biodistribution Considerations

[00357] The therapeutic potential of the compositions described herein allow for greater biodistribution and bioavailability of the modified AB or the AA. The compositions described herein provide an antibody therapeutic having an improved bioavailability wherein the affinity of binding of the antibody therapeutic to the target is lower in a healthy tissue when compared to a diseased tissue. A pharmaceutical composition comprising an AB coupled to a MM can display greater affinity to the target in a diseased tissue than in a healthy tissue. In preferred embodiments, the affinity in the diseased tissue is 5-10,000,000 times greater than the affinity in the healthy tissue.

[00358] Generally stated, the present disclosure provides for an antibody therapeutic having an improved bioavailability wherein the affinity of binding of the antibody therapeutic to its target is lower in a first tissue when compared to the binding of the antibody therapeutic to its target in a second tissue. By way of example in various embodiments, the first tissue is a healthy tissue and the second tissue is a diseased tissue; or the first tissue is an early stage tumor and the second tissue is a late stage tumor; the first tissue is a benign tumor and the second tissue is a malignant tumor; the first tissue and second tissue are spatially separated; or in a specific example, the first tissue is epithelial tissue and the second tissue is breast, head, neck, lung, pancreatic, nervous system, liver, prostate, urogenital, or cervical tissue.

EXAMPLES

Example 1: Screening of Candidate Masking Moieties (MMs)

[00359] In order to produce compositions comprising antibodies and fragments thereof (AB) coupled to MMs with desired optimal binding and dissociation characteristics, libraries of candidate MMs are screened. MMs having different variable amino acid sequences, varying positions of the cysteine, various lengths, and the like are generated. Candidate MMs are tested for their affinity of binding to ABs of interest. Preferably, MMs not containing the native amino acid sequence of the binding partner of the AB are selected for construction of the modified antibodies.

[00360] Affinity maturation of MMs for ABs of interest to select for MMs with an affinity of about 1-10 nM is carried out.

Example 2: Screening of Modified Antibody and Activatable Antibody (AA) Libraries

[00361] In order to identify modified antibodies and AAs having desired switching characteristics (i.e., decreased target binding when in an masked and/or uncleaved conformation relative to target binding when in a masked and/or cleaved conformation), libraries of candidate modified antibodies and candidate AAs having different variable amino acid sequences in the masking moieties (MMs), varying positions of the cysteine in the MM, various linker lengths, and various points of attachment to the parent AB are generated.

[00362] A scheme for screening/sorting method to identify candidate AAs that display the switchable phenotype is provided here. The libraries are introduced via expression vectors resulting in display of the AAs on the surface of bacterial cells. After expansion of the libraries by culture, cells displaying the AA polypeptides are then treated with the appropriate enzyme or reducing agent to provide for cleavage or reduction of the CM. Treated cells are then contacted with fluorescently labeled target and the cells are sorted by FACS to isolate cells displaying AAs which bind target after cleavage/reduction. The cells that display target-binding constructs are then expanded by growth in culture. The cells are then contacted with labeled target and sorted by FACS to isolate cells displaying AAs which fail to bind labeled target in the absence of enzyme/reducing agent. These steps represent one cycle of the screening procedure. The cells can then be subjected to additional cycles by expansion by growth in culture and again subjecting the culture to all or part of the screening steps.

[00363] Library screening and sorting can also be initiated by first selecting for candidates that do not bind labeled target in the absence of enzyme/reduction agent treatment (i.e., do not bind target when not cleaved/reduced).

Example 3: In vitro Screening of Modified Antibodies to Determine Masking Efficiency of the MM

[00364] In order to screen modified antibodies and AAs that exhibit optimal characteristics when masked, for example, only 10% of binding to the target when in a masked state and in the presence of target, ABs

coupled to different MMs or ABs coupled to the same MMs at different points of attachment, or ABs coupled to the same MM via linkers of different lengths and/or sequences are generated.

[00365] The masking efficiency of MMs can be determined by the affinity of the MM for AB and the spatial relationship of the MM relative to the binding interface of the AB to its target. Discovery of an effective MM is based on affinity and as well optionally an empirical measure of masking efficiency. The time-dependent target displacement of the MM in the modified antibody or AA can be measured to optimize and select for MMs. A immunoabsorbant Target Displacement Assay (TDA) is described herein for the discovery and validation of efficiently masked antibodies

[00366] In the TDA assay, the ability of an MM to inhibit AB binding to its target at therapeutically relevant concentrations and times is measured. The assay allows for measurement of the time-dependent target displacement of the MM.

[00367] Briefly the antibody target is adsorbed to the wells of an ELISA plate overnight at about 4°C. The plate is blocked by addition of about 150 µl 2% non-fat dry milk (NFDM) in PBS, about 0.5% (v/v) Tween20 (PBST), and incubation at room temperature for about 1 hour. The plate is then washed about three times with PBST. About 50 µl superbloc is added (Thermo Scientific) and supplemented with protease inhibitors (Complete, Roche). About 50 µl of an AB coupled to a MM is dissolved in superbloc with protease inhibitors (Complete, Roch) and incubated at about 37°C for different periods of time. The plate is washed about three times with PBST. About 100 µl of anti-huIgG-HRP is added in about 2% NFDM/PBST and incubated at room temperature for about 1 hour. The plate is washed about four times with PBST and about twice with PBS. The assay is developed using TMB (Thermo Scientific) as per manufacturer's directions.

Example 4: AAs Comprising an scFv as the AB

[00368] Examples of AAs comprising an anti-Jagged1 scFv are described herein. These AAs are inactive (masked) under normal conditions due to the attached MM. When the scFv reaches the site of disease, however, a disease-specific enzyme such as ADAM17 will cleave a substrate linker connecting the peptide inhibitor to the scFv allowing it to bind to Jagged1. Bacterial cell surface display is used to find suitable MMs for the anti-Jagged1 scFv. In this example, selected MMs are combined with an enzyme substrate to be used as a trigger to create a scFv construct that becomes competent for targeted binding after protease activation.

Construction of Protease Activated Antibody

[00369] Genes encoding AAs comprising a Jagged1 antibody in single-chain form are produced by overlap extension PCR or total gene synthesis and ligated into a similarly digested expression plasmid or any other suitable bacterial, yeast, or mammalian expression vector familiar to one skilled in the art. Full length antibodies can be alternatively produced using commercially available expression vectors incorporating half-life extending moieties (e.g. the Fc region of an IgG, serum albumin, or transferrin)

and methods familiar to one skilled in the art. The expression plasmid is then transformed or transfected into an appropriate expression host such as BL21 for E. coli or HEK293t cells. Single chain antibodies are harvested from overnight cultures using a Periplasmic fraction extraction kit (Pierce), and purified by immobilized metal ion affinity chromatography, and by size exclusion chromatography.

Assay for antibody switching activity in vitro

[00370] Aliquots of protease-activated antibodies, at a concentration of 1 pM -1 μ M are incubated in a buffered aqueous solution separately with 0 and 50 nM enzyme for 3 hrs. The reaction mixtures are then assayed for binding using ELISA or surface Plasmon resonance with immobilized antigen Jagged1. An increase in binding activity for the AA after protease treatment is indicated by an increase in resonance units when using BIAcore™ SPR instrumentation. The change in apparent dissociation constant (K_d) as a result of cleavage can then be calculated according the instrument manufacturer's instructions (BIAcore, GE Healthcare).

Example 5: Cloning of the Anti-VEGF scFv AB

[00371] In this and following examples an AA containing a masked MMP-9 cleavable anti-VEGF scFv (target = VEGF; AB = anti-VEGF single chain Fv) was constructed. As a first step in the production of such an AA, constructs containing an anti-VEGF scFv were generated (the AB). An anti-VEGF scFv AB (V_L -linker L- V_H) was designed from the published sequence of ranibizumab (Genentech, Chen, Y., Wiesmann, C., Fuh, G., Li, B., Christinger, H., McKay, P., de Vos, A. M., Lowman, H. B. (1999) Selection and Analysis of an Optimized Anti-VEGF Antibody: Crystal Structure of an Affinity-matured Fab in Complex with Antigen *J. Mol. Biol.* 293, 865-881) and synthesized by Codon Devices (Cambridge, MA).

[00372] Ranibizumab is a monoclonal antibody Fab fragment derived from the same parent murine antibody as bevacizumab (Presta LG, Chen H, O'Connor SJ, et al Humanization of an anti-vascular endothelial growth factor monoclonal antibody for the therapy of solid tumors and other disorders. *Cancer Res*, 57: 4593-9, 1997). It is smaller than the parent molecule and has been affinity matured to provide stronger binding to VEGF-A. Ranibizumab binds to and inhibits all subtypes of vascular endothelial growth factor A (VEGF-A). A His6 tag at the N-terminus and a TEV protease cleavage site were included in the design. The TEV protease is a protease isolated from tobacco etch virus, is very specific, and is used to separate fusion proteins following purification. The anti-VEGF scFv nucleotide and amino acid sequences are provided below in Tables 7 and 8.

Table 7: anti-VEGF scFv AB nucleotide sequence

gatattcaactgaccagagcccttctccctgagtgccagcgtgggtgaccgtgttacgatcacttgctcggccagccaagatatttctaactacctgaatt ggtaccagcagaagccaggaaaggcaccaaaagtctgatctacttcacaagttcactgcattccggcgtaccgtcgcgctttagcggttctggcagtgg taccgacttcaccctgactatctcgagtctgcaacctgaggattttgctacatattactgtcagcaatattcgaccgtgccgtggacgttcgggcagggcac caaagtggagattaaggggggtggaggcagcgggggaggtggctcaggcgggtggagggtctggcgaggtccagctggtagaaagcggggggcgg actggtccaaccgggcggatccctgcgtctgagctgcgcggcctcgggttacgactttactactacggaatgaactgggttcgccaagcccttggtaaa

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gggtctggaatgggtcggatggattaatacatacactggagaacctacttatgctgctgattcaaacgtcgctttactttctctctggatacaagtaagtcaac
cgcctatctgcaaataaacagcctgcgtgcagaggacacggctgtgtactattgtgcgaaatattccttattattatggaacttcccactgggtatttcgatgtat
ggggccagggtactctggttacagtgtcg

```

Table 8: anti-VEGF scFv AB amino acid sequence

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DIQLTQSPSSLSASVGDRVITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGS
GTDFTLTISLQPEDFATYYCQQYSTVPWTFGQGTKVEIKGGGGSGGGGSGGGGSGGEVQLVESG
GGLVQPGGSLRLSCAASGYDFTHYGMNWVRQAPGKGLEWVGWINTYTGEPTYAADFKRRFTF
SLDTSKSTAYLQMNSLRAEDTAVYYCAKYPYYYGTSHWYFDVWGQGTLLVTVS

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Example 6: Screening and Identification of MMs for anti-VEGF scFv

[00373] Ranibizumab was used to screen a pooled random peptide library, consisting of peptides that are X_{15} (8.3×10^9), $X_4CX_7CX_4$ (3.6×10^9), or $X_{12}CX_3$ (1.1×10^9), where X is any amino acid and the number represents the total diversity of the library. The total diversity of the pooled library was 1.3×10^{10} . The screening consisted of one round of MACS and two rounds of FACS sorting. In the first round MACS screen, 1×10^{11} cells were probed with 150 nM biotinylated-ranibizumab, and 5.5×10^7 binding cells were isolated. In the first FACS screen, positive cells isolated in the MACS screen were probed with 500 nM biotinylated- ranibizumab, and visualized with neutrAvidin-PE (Molecular Probes, Eugene, OR). The second and third rounds of FACS selections were done with 500nM and then 100 nM Alexa-labeled ranibizumab in the presence of 20uM IgG. Individual clones were sequenced and subsequently verified for their ability to bind anti-VEGF scFv by FACS analysis. Amino acid sequences of MMs for anti-VEGF scFv are provided in Table 9 below. (These sequences will hereafter be referred to as 283MM, 292MM, 306MM, etc.)

Table 9: MMs for anti-VEGF scFv

JS283	ATAVWNSMVKQSCYMQG
JS292	GHGMCYTILEDHCDVR
JS306	PCSEWQSMVQPRCYGG
JS308	NVECCQNYNLWNCCGGR
JS311	VHAWEQLVIQELYHC
JS313	GVGLCYTILEQWCEMGR
JS314	RPPCCRDYSILECCKSD
JS315	GAMACYNIFEYWCSAMK

Example 7: Construction of the AA: MMP-9 Cleavable, Masked- Anti-VEGF scFv Vectors

[00374] A CM (substrate for MMP-9) was fused to the masked anti-VEGF scFv construct to provide a cleavable, masked AA. An exemplary construct is provided in Figure 4. Several exemplary AA constructs and sequences containing different CMs are described in great detail below. Primers utilized for construction of the exemplary constructs are represented in Table 10 below.

Table 10: Primers utilized for construction of MMP-9 Cleavable, masked- anti-VEGF scFv

CX0233 5'gaattcatgggccatcaccatcaccatcacggtgggg3'
CX0249 5'gtgagtaagctttattacgacactgtaaccagagtaccctgg3'
CX0270 5'gtggcatgtgcacttggccaccttggcccactcgagctggccagactggccctgaaaatacagattttccc3'
CX0271 5'gagtgggccaaggtggccaagtgcacatgccactgggcttctgggtccgggcggttctgatattcaactgaccagagcc3'
CX0288 5' ttcgagctcgaacaacaacaacaataacaataacaacaac3'
CX0289 5' gctttcaccgcaggtacttccgtagctggccagtctggcc3'
CX0290 5' cgtccatgggccaccttggccgctgccaccagaaccgcc3'
CX0308 5' gccagccggccatggccggcagctctggccagctcgagt3'
CX0310 5' ccagtccaagcttttagtggtgatggtgatgatgcgacactgtaaccagagtaccctggcc3'
CX0312 5'cttgacgaattcgggccagctctggccagctcgagt3'
CX0314 5'cagatctaaccatggcgccgctaccgccgacactgtaaccagagtaccctg3'

Cloning and expression of the AA: a MMP-9 cleavable, masked anti-VEGF scFv as a MBP fusion

[00375] *Cloning:* A MBP:anti-VEGF scFv AB fusion was cloned. The MBP (maltose binding protein) expresses well in E. coli, as a fusion protein, and can be purified on a maltose column, a method well known in the art to make fusion proteins. In this example, the MBP was used to separate the masked scFv. The His6 tagged Anti-VEGF scFv AB was cloned into the pMal-c4x vector (NEB) as a C-terminal fusion with MBP using the EcoRI and HindIII restriction sites in the multiple cloning site (MCS). The primers CX0233 and CX0249 (Table 10) were used to amplify the Anti-VEGF scFv AB and introduce the EcoRI and HindIII sites, respectively.

[00376] The accepting vector for the AA (the peptide MM, the anti-VEGF scFv AB and the MMP-9 CM) was synthesized using polymerase chain reaction (PCR) with the overlapping primers CX0271 and CX0270 which placed the cloning site for the peptide MM's, linker sequences, and MMP-9 CM protease site between the TEV protease site and the anti-VEGF scFv AB. The primers CX0271 and CX0249 (Table 10) were used to amplify the C-terminal portion of the construct, while the primers CX0270 and CX0288 (Table 10) were used to amplify the N-terminal portion. Products from both the above reactions were combined for a final PCR reaction using the outside primers, CX0249 and CX0288 (Table 10), and cloned into the pMal vector as an MBP fusion using the SacI and HindIII restriction sites.

Table 11: MBP/MM accepting site/MMP-9 CM/Anti-VEGF scFv AB vector nucleotide sequence

atgggccatcaccatcaccatcacggtggggaaaatctgtattttcagggccagctctggccagctcgagtgggccaaggtggccaagtgcacatgccac tgggcttctgggtccggcggttctgatattcaactgaccagagcccttcttcctgagtgccagcgtgggtgaccgtgttacgacttgcctggcca gccaagatatttcaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtctgatctacttcacaagttcactgcattccggcgtagcg tcgcgcttagcggttctggcagtggtaccgacttcacctgactatctcgagtctgaacctgaggattttgctacatattactgtcagcaatattcgaccgt gccgtggacgttcgggcagggcaccaaagtggagattaaggggggtggaggcagcgggggaggtggctcaggcggtggagggtctggcgaggtc cagctggtagaaagcggggcgactggtccaaccggcgatccctgcgtctgagctgcgcggcctcgggttacgactttactcactacggaatgaa ctgggttcgccaagcccttgtaaggtctggaatgggtcgatggattaatacactggagaacctacttatgctgctgatttcaaacgtcgctttact ttctcttgatacaagtaagtcaaccgcctatctgcaaatgaacagcctgcgtgcagaggacacggctgtgtactattgtcgaaatatccttattattatgg aactcccactggtatttcgatgtatggggccagggtactctggttacagtgcg

[00377] The 306MM and 314MM (Table 9) were amplified from the ecpX display vector using the primers CX0289 and CX0290 (Table 10), and directionally cloned into the N-terminally masked vector

using the SfiI restriction sites. The corresponding nucleotide and amino acid sequences are provided in Table 12 below.

Table 12: 306 or 314 MM / MMP-9 CM / Anti-VEGF scFv AB Sequences

MBP / 306 MM / MMP-9 CM / Anti-VEGF scFv AB nucleotide sequence
atgggccatcaccatcaccatcacgggtggggaaaatctgtattttcagggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgtt gctattatgggggcggttctggtggcagcggccaaggtggccaagtgcacatgccactgggcttctgggtccgggcggttctgatattcaactgaccca gagcccttctccctgagtgccagcgtgggtgaccgtgttacgatcacttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagcc aggaaaggcaccaaaaagtcctgatctacttcacaagttcactgcattccggcgtaccgtcgcgctttagecggttctggcagtggtaccgacttcacctga ctatctcgagtctgcaacctgaggattttgctacatattactgtcagcaatattcgaccgtgccgtggacgttcgggcagggcaccaaagtggagattaagg ggggtggaggcagcgggggaggtggctcaggcgggtggagggtctggcgaggtccagctggtagaaagcggggggcggactggtccaaccgggcg gatccctgcgtctgagctgcgcggcctcgggttacgactttactcactacggaatgaactgggttcgccaagcccctggtaaaggcttggaatgggtcgg atggattaatacatatactggagaacctacttatgctgctgatttcaaacgtcgtttactttctctctggatacaagtaagtcaaccgcctatctgcaaatga cagcctgcgtgcagaggacacggctgtgtactattgtgcgaaatataccttattattatggaacttcccactggtatttcgatgtatggggccagggtactctg gttacagtgtcg
MBP / 306 MM / MMP-9 CM / Anti-VEGF scFv AB amino acid sequence
MGHHHHHHGGENLYFQQQSGQPCSEWQSMVQPRCYGSGSGGSGQGGQVHMPLGFLGPGG SDIQLTQSPSSLSASVGDRVTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVP SRFSGSGSGTDFTLTISSLQPEDFATYYCQQYSTVPWTFGQGTKVEIKGGGGSGGGGSGG GGSGEVQLVESGGGLVQPGGSLRLSCAASGYDFTHYGMNWVRQAPGKGLEWVGWINTYTG EPTYAADFKRRFTFSLDTSKSTAYLQMNSLRAEDTAVYYCAKYPYYYGTSHWYFDVWGQG TLVTVS
MBP / 314 MM / MMP-9 CM / Anti-VEGF scFv AB nucleotide sequence
atgggccatcaccatcaccatcacgggtggggaaaatctgtattttcagggccagtctggccagcggccgctgttgcctgattatagtagtatttggagtgc tgaagagtgatggcggttctggtggcagcggccaaggtggccaagtgcacatgccactgggcttctgggtccgggcggttctgatattcaactgaccc agagcccttctccctgagtgccagcgtgggtgaccgtgttacgatcacttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagc caggaaaggcaccaaaaagtcctgatctacttcacaagttcactgcattccggcgtaccgtcgcgctttagecggttctggcagtggtaccgacttcacctg actatctcgagtctgcaacctgaggattttgctacatattactgtcagcaatattcgaccgtgccgtggacgttcgggcagggcaccaaagtggagattaag ggggtggaggcagcgggggaggtggctcaggcgggtggagggtctggcgaggtccagctggtagaaagcggggggcggactggtccaaccgggcg ggatccctgcgtctgagctgcgcggcctcgggttacgactttactcactacggaatgaactgggttcgccaagcccctggtaaaggcttggaatgggtcgg gatggattaatacatatactggagaacctacttatgctgctgatttcaaacgtcgtttactttctctctggatacaagtaagtcaaccgcctatctgcaaatga acagcctgcgtgcagaggacacggctgtgtactattgtgcgaaatataccttattattatggaacttcccactggtatttcgatgtatggggccagggtactct ggttacagtgtcg
MBP / 314 MM / MMP-9 CM / Anti-VEGF scFv AB amino acid sequence
MGHHHHHHGGENLYFQQQSGQRPPCCRDYSILECKSDGGSGSGGSGQGGQVHMPLGFLGPG GSDIQLTQSPSSLSASVGDRVTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGV PSRFSGSGSGTDFTLTISSLQPEDFATYYCQQYSTVPWTFGQGTKVEIKGGGGSGGGGSGG GGSGEVQLVESGGGLVQPGGSLRLSCAASGYDFTHYGMNWVRQAPGKGLEWVGWINTYT GEPTYAADFKRRFTFSLDTSKSTAYLQMNSLRAEDTAVYYCAKYPYYYGTSHWYFDVWGQ GTLVTVS

[00378] *Expression:* Expression of the MBP:AA fusions were conducted in a K12 TB1 strain of *E. coli*. An ampicillin-resistant colony containing the desired construct was used to inoculate a 5 ml overnight culture containing LB medium supplemented with 50 µg/mL Ampicillin. The entire overnight culture was used to inoculate 500 mL of fresh LB medium supplemented with 50 µg/mL ampicillin and 0.2% Glucose and allowed to grow at 37°C shaking at 250 rpm until an O.D. of 0.5 was reached. Isopropylthio-β-D-galactosidase was then added to a final concentration of 0.3 mM and the culture was allowed to grow for a further 3 hrs under the same conditions after which the cells were harvested by

centrifugation at 3000xg. Inclusion bodies were purified using standard methods. Briefly, 10mls of BPER II cell lysis reagent (Pierce). Insoluble material was collected by centrifugation at 14,000xg and the soluble proteins were discarded. The insoluble materials were resuspended in 5mls BPER II supplemented with 1mg/mL lysozyme and incubated on ice for 10 minutes after which 5mls of BPER II diluted in water 1:20 was added and the samples were spun at 14,000xg. The supernatant was removed and the pellets were wash twice in 1:20 BPERII. The purified inclusion bodies were solubilized in PBS 8 M Urea, 10 mM BME, pH 7.4.

[00379] The MBP fusion proteins were diluted to a concentration of approximately 1 mg/mL and refolded using a stepwise dialysis in PBS pH 7.4 from 8 to 0 M urea through 6, 4, 2, 0.5, and 0 M urea. At the 4, 2, and 0.5 M Urea steps 0.2 M Arginine, 2 mM reduced Glutathione, and 0.5 mM oxidized glutathione was added. The 0M Urea dialysis included 0.2 M Arginine. After removal of the urea, the proteins were dialyzed against 0.05 M Arginine followed by and extensive dialysis against PBS pH 7.4. All dialysis were conducted at 4°C overnight. To remove aggregates, each protein was subjected to size exclusion chromatography on a sephacryl S-200 column. Fractions containing the correctly folded proteins were concentrated using an Amicon Ultra centrifugal filter.

Cloning and expression of the AA: a MMP-9 cleavable, masked anti-VEGF scFv CHis tag

[00380] *Cloning:* The primers CX0308 and CX0310 (Table 10) were used to amplify and add a NcoI restriction site to the 5' end and a HindIII restriction site and His6 tag to the 3' end, respectively, of the (MM accepting site / MMP-9 CM / VEGFscFv AB) vector which was subsequently cloned into a vector containing the pelB signal peptide. Anti-VEGF scFv MMs were cloned as previously described. The corresponding nucleotide and amino acid sequences are provided in Table 13.

Table 13: 306 or 314 MM / MMP-9 CM / anti-VEGF scFv CHis AB Sequences

306 MM / MMP-9 CM / anti-VEGF scFv CHis AB nucleotide sequence
ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttctattatggggggcggttctggtggcagcggccaaggtggccaagt gcacatgccactgggcttctgggtccgggcggttctgatattcaactgacctagagcccttcttccctgagtccagcgtgggtgacctgttacgatca cttctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcactgcatt ccggcgtaccgtcgcgcttttagcggttctggcagtggtagcacttcacctgactatctcgagtctgcaacctgaggattttgctacatattactgtcagca atattcgacctgcccgtggacgttcggggcagggcaccaaagtggagattaaggggggtggaggcagcgggggaggtggctcaggcgggtggagggt ctggcgaggtccagctggtagaaagcgggggcggactggtccaaccgggcggatccctgcgtctgagctgcgcggcctcgggttacgactttactcac tacggaatgaactgggttcgccaagcccctggtaaaggtctggaatgggtcggtggtgattaatacatacactggagaacctacttatgctgctgatttcaa cgtcgctttactttctctctgatacaagtaagtcaaccgcctatctgcaaatgaacagcctgcgtgcagaggacacggctgtgtactattgtgcgaaatata cttattattatggaactcccactggtatttcgatgtatggggccagggtactctggttacagtgctgcacatcaccatcaccac
306 MM / MMP-9 CM / anti-VEGF scFv CHis AB amino acid sequence
GQSGQPCSEWQSMVQPRCYYGGGSGGSGQGQVHMPLGFLPGGSDIQLTQSPSSLSASV GDRVITITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTI SSLQPEDFATYYCQQYSTVPWTFGQGTKVEIKGGGSGGGGSGGGGSGEVQLVESGGGLV QPGGSLRLSCAASGYDFTHYGMNWVRQAPGKGLEWVGWINTYTGEPTYAADFKRRFTFSL DTSKSTAYLQMNSLRAEDTAVYYCAKYPYYYGTSHWYFDVWGQGTLLTVSHHHHHH
314 MM / MMP-9 CM / anti-VEGF scFv CHis AB nucleotide sequence
ggccagtctggccagcggccgccgtgttgccgtgattatagatattttggagtgcgtgaagagtgatggcggttctggtggcagcggccaaggtggccaa

gtgcacatgccactgggcttctgggtccgggcgggttctgatattcaactgaccagagcccttctccctgagtgccagcgtgggtgaccgtgttacgat
cacttctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcacaaaaagtcctgatctacttcacaagttcactgc
attcggcgtagcgtcgcgttttagcggttctggcagtggtaccgacttcaccctgactatctcgagtctgcaacctgaggattttgtacatattactgtcag
caatattcgaccgtgccgtggacgttcgggcagggcaccaaagtggagattaaggggggtggaggcagcgggggaggtggctcaggcgggtggagg
gtctggcgaggtccagctggtagaaagcggggcggaactggccaaccgggcggatccctgcgtctgagctgcgcggcctcgggttacgactttactc
actacggaatgaactgggttcgccaagcccttggtaaaggtctggaatgggtcggatggattaatacatacactggagaacctacttatgtctgtgattca
aacgtcgtttactttctcttgatacaagtaagtcaaccgcctatctgcaaataaacagcctgcgtgcagaggacacggctgtgtactattgtgcgaaata
tccttattattatggaactcccactggtatttcgatgtatggggccagggtactctggttacagtgtcgcatacaccataccactaa

314 MM / MMP-9 CM / anti-VEGF scFv CHis AB amino acid sequence

GQSGQRPPCCRDYSILECCKSDGGSGGSGQGGQVHMPLGFLGPGGSDIQLTQSPSSLSAS
VGDRVITITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLT
ISSLPEDFATYYCQQYSTVPWTFGQGTKVEIKGGGGSGGGGSGGGGSGGEVQLVESGGGL
VQPGGSLRLSCAASGYDFTHYGMNWVRQAPGKGLEWVGWINTYTGEPTYAADFKRRFTFS
LDTSKSTAYLQMNSLRAEDTAVYYCAKYPYYYGTSHWYFDVWGQGTLLTVSHHHHHH

[00381] *Expression:* Expression of the Anti-VEGF scFv His AAs was conducted in a K12 TB1 strain of *E. coli*. An ampicillin-resistant colony containing the desired construct was used to inoculate a 5 ml overnight culture containing LB medium supplemented with 50 µg/mL Ampicillin. 2.5 ml of overnight culture was used to inoculate 250 mL of fresh LB medium supplemented with 50 µg/mL ampicillin and 0.2% Glucose and allowed to grow at 37°C shaking at 250 rpm until an O.D. of 1.0 was reached. Isopropylthio-β-D-galactosidas was then added to a final concentration of 0.3 mM and the culture was allowed to grow for a further 5 hrs at 30°C after which the cells were harvested by centrifugation at 3000xg. The periplasmic fraction was immediately purified using the lysozyme/osmotic shock method. Briefly, the cell pellet was resuspended in 3mLs of 50mM Tris, 200 mM NaCl, 10mM EDTA, 20% Sucrose, pH 7.4 and 2 uL/mL ready-use lysozyme solution was added. After a 15 min. incubation on ice, 1.5 volumes of water (4.5mLs) was added and the cells were incubated for another 15 min. on ice. The soluble periplasmic fraction was recovered by centrifugation at 14,000xg.

[00382] The Anti-VEGF scFv His proteins were partially purified using Ni-NTA resin. Crude periplasmic extracts were loaded onto 0.5 ml of Ni-NTA resin and washed with 50 mM phosphate, 300 mM NaCl, pH 7.4. His tagged proteins were eluted with 50mM phosphate, 300 mM NaCl, 200 mM Imidazole, pH 6.0. Proteins were concentrated to approximately 600 µL and buffer exchanged into PBS using Amicon Ultra centrifugal concentrators.

Cloning and expression of the AA: a MMP-9 cleavable, masked anti-VEGF scFv as human Fc fusion

[00383] *Cloning:* The primers CX0312 and CX0314 (Table 10) were used to amplify the sequence encoding MMP-9 CM / Anti-VEGF scFv. The primers also included sequences for a 5' EcoRI restriction site and a 3' NcoI restriction site and linker sequence. Cutting the PCR amplified sequence with EcoRI and NcoI and subsequent cloning into the pFUSE-hIgG1-Fc2 vector generated vectors for the expression of Fc fusion proteins. Anti-VEGF scFv AB MMs were inserted into these vectors as previously described. Constructs containing 306MM, 313MM, 314MM, 315MM, a non-binding MM (100MM), as well as no MM were constructed and sequences verified. The corresponding nucleotide and amino acid sequences are provided below in Table 14.

Table 14: 306 MM / MMP-9 CM /anti-VEGF scFv-Fc AB sequences

306 MM / MMP-9 CM /anti-VEGF scFv-Fc AB nucleotide sequence
ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttctgattatggggggcggttctggtggcagcggccaaggtggccaagt cacatgccactgggcttctgggtccgggcggttctgatattcaactgacctagagcccttcttccctgagtgccagcgtgggtgacctgtgtacgatcactt gtcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcactgcattccg gcgtaccgtcgcgcttttagcggttctggcagtggtaccgacttcacctgactatctcgagtctgcaacctgaggattttgctacatattactgtcagcaatatt cgaccgtgccgtggacgttcgggcagggcaccaaagtggagattaaggggggtggaggcagcgggggaggtggctcaggcggtggagggtctggc gagggtccagctggtagaaagcgggggcggactggtccaaccgggcggatccctgcgtctgagctgcgcggcctcgggttacgactttactactacgg aatgaactgggttcgccaagccccctggtaaaggcttggaatgggtcggatggattaatacatacactggagaacctacttatgctgctgatttcaaacgtcg ctttactttctctctggatacaagtaagtcaaccgcctatctgcaaataaacagcctgcgtgcagaggacacggctgtgtactattgtgcgaaatatccttatta ttatggaaacttcccactggtatttcgatgtatggggccagggtactctggttacagtgtcgggcggttagcggcgccatggttagatctgacaaaactcacac atgccaccgtgccagcacctgaactcctgggggggaccgtcagtcttcttcttccccccaaaacccaaggacacctcatgatctcccgaccctgag gtcacatgcgtggtggtggacgtgagccacgaagacctgaggtcaagttcaactggtacgtggacggcgtggaggtgcataatgccaagacaaaagcc gcgggaggagcagtagacaacagcacgtaccgtgtggtcagcgtcctcaccgtcctgcaccaggactggctgaatggcaaggagtacaagtgaagggtct ccaacaaagccctcccagcccccatcgagaaaaccatctccaaagccaaagggcagccccgagaaccacaggtgtacacctgcccccatcccgga ggagatgaccaagaaccaggtcagcctgacctgctggtcaaaggcttctatcccagcgacatcgccgtggagtgggagagcaatgggcagccggag aacaactacaagaccacgcctcccgtgctggactccgacggctccttcttctctacagcaagctcaccgtggacaagagcaggtggcagcaggggaa cgtcttctcatgctccgtgatgcatgagggtctgcacaaccactacacgcagaagagcctctccctgtctccgggtaaa
306 MM / MMP-9 CM / anti-VEGF scFv-Fc AB amino acid sequence
GQSGQPCSEWQSMVQPRCYGSGSGGSGQGGQVHMPLGFLGPGGSDIQLTQSPSSLSASV GDRVITITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLT SSLQPEDFATYYCQQYSTVPWTFGQGTKVEIKGGGGSGGGGSGGGGSGEVQLVESGGGLV QPGGSLRLSCAASGYDFTHYGMNWVRQAPGKGLEWVGWINTYTGEPTYAADFKRRFTFSL DTSKSTAYLQMNSLRAEDTAVYYCAKYPYYYGTSHWYFDVWGQGLTVTVSGGSGAMVRS KTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDG VEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKG QPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD GSFFLYSKLTVDKSRWQQGNVFSCSVMHEGLHNHYTQKSLSLSPGK
314 MM / MMP-9 CM / anti-VEGF scFv-Fc AB nucleotide sequence
ggccagtctggccagcggccgcggtgttgccgtgattatagatattttggagtgtgtaagagtgtggcggttctggtggcagcggccaaggtggccaag tgcacatgccactgggcttctgggtccgggcggttctgatattcaactgacctagagcccttcttccctgagtgccagcgtgggtgacctgtgtacgatca cttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaaagtcctgatctacttcacaagttcactgcattc cggcgtaccgtcgcgcttttagcggttctggcagtggtaccgacttcacctgactatctcgagtctgcaacctgaggattttgctacatattactgtcagcaat attcgacctgcccgtggacgttcgggcagggcaccaaagtggagattaaggggggtggaggcagcgggggaggtggctcaggcggtggagggtctg gcgaggtccagctggtagaaagcgggggcggactggtccaaccgggcggatccctgcgtctgagctgcgcggcctcgggttacgactttactactac ggaatgaactgggttcgccaagccccctggtaaaggcttggaatgggtcggatggattaatacatacactggagaacctacttatgctgctgatttcaaacgt cgctttactttctctctggatacaagtaagtcaaccgcctatctgcaaataaacagcctgcgtgcagaggacacggctgtgtactattgtgcgaaatatcctta ttattatggaacttcccactggtatttcgatgtatggggccagggtactctggttacagtgtcgggcggttagcggcgccatggttagatctgacaaaactcac acatgccaccgtgccagcacctgaactcctgggggggaccgtcagtcttcttcttccccccaaaacccaaggacacctcatgatctcccgaccctg aggtcacatgcgtggtggtggacgtgagccacgaagacctgaggtcaagttcaactggtacgtggacggcgtggaggtgcataatgccaagacaaaag ccgcgggaggagcagtagacaacagcacgtaccgtgtggtcagcgtcctcaccgtcctgcaccaggactggctgaatggcaaggagtacaagtgaagggt ctccaacaaagccctcccagcccccatcgagaaaaccatctccaaagccaaagggcagccccgagaaccacaggtgtacacctgcccccatcccg gaggagatgaccaagaaccaggtcagcctgacctgcctggtcaaaggcttctatcccagcgacatcgccgtggagtgggagagcaatgggcagccgg agaacaactacaagaccacgcctcccgtgctggactccgacggctccttcttctctacagcaagctcaccgtggacaagagcaggtggcagcagggg aacgtcttctcatgctccgtgatgcatgagggtctgcacaaccactacacgcagaagagcctctccctgtctccgggtaaa
314 MM / MMP-9 CM / anti-VEGF scFv-Fc AB amino acid sequence
GQSGQRPPCCRDYSILECCKSDGGSGGSGQGGQVHMPLGFLGPGGSDIQLTQSPSSLSAS VGDRVITITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLT ISSLQPEDFATYYCQQYSTVPWTFGQGTKVEIKGGGGSGGGGSGGGGSGEVQLVESGGGL VQPGGSLRLSCAASGYDFTHYGMNWVRQAPGKGLEWVGWINTYTGEPTYAADFKRRFTFS LDTSKSTAYLQMNSLRAEDTAVYYCAKYPYYYGTSHWYFDVWGQGLTVTVSGGSGAMVRS DKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVD GVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAK

GQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDS
DGSFFLYSKLTVDKSRWQQGNVFCFSVMHEGLHNHYTQKSLSLSPGK

[00384] *Expression:* 10 µg of expression vectors for 306 MM / MMP-9 CM / anti-VEGFscFv-Fc, 314 MM / MMP-9 CM / anti-VEGFscFv-Fc or anti-VEGFscFv-Fc were introduced into 10⁷ HEK-293 freestyle cells (Invitrogen, CA) by transfection using transfectamine 2000 as per manufacturer's protocol (Invitrogen, CA). The transfected cells were incubated for an additional 72 hours. After incubation, the conditioned media was harvested and cleared of cells and debris by centrifugation. The conditioned media was assayed for activity by ELISA.

Example 8: Measurement of the Activation of a Masked MMP-9 Cleavable AA

[00385] To measure the activation of the masked MMP-9 cleavable anti-VEGF AAs by MMP-9, 100 µl of a 2 µg/ml PBS solution of VEGF was added to microwells (96 Well Easy Wash; Corning) and incubated overnight at 4°C. Wells were then blocked for 3 x 15 minute with 300 µL Superblock (Pierce). One hundred microliters of an AA (see below for details pertaining to each construct), treated or untreated with MMP-9, were then added to wells in PBST, 10% Superblock and incubated at room temperature (RT) for 1 hr. All wash steps were done three times and performed with 300 µl PBST. One hundred microliters of secondary detection reagent were then added and allowed to incubate at RT for 1 hr. Detection of HRP was completed using 100 µl of TMB one (Pierce) solution. The reaction was stopped with 100 µL of 1N HCL and the absorbance was measured at 450 nM.

ELISA assay of an AA construct containing: MBP / MM / MMP-9 CM / anti-VEGF scFv AB

[00386] Two hundred microliters of biotinylated AA in MMP-9 digestion buffer (50 mM Tris, 2 mM CaCl₂, 20 mM NaCl, 100 µM ZnCl₂, pH 6.8) at a concentration of 200 nM was digested with 20U TEV protease overnight at 4°C to remove the MBP fusion partner. Samples were then incubated for 3 hrs with or without ~3U of MMP-9 at 37°C, diluted 1:1 to a final concentration of 100 nM in PBST, 10% Superblock, and added to the ELISA wells. Detection of the AA was achieved with an Avidin-HRP conjugate at a dilution of 1:7500. MMP-9 activation of MMP-9 cleavable masked MBP:anti-VEGF scFv AA is presented in Figure 5.

ELISA assay of an AA construct containing: MM / MMP-9 CM / anti-VEGF scFv His

[00387] Crude periplasmic extracts dialyzed in MMP-9 digestion buffer (150 µL) were incubated with or without ~3U of MMP-9 for 3 hrs at 37°C. Samples were then diluted to 400 µL with PBST, 10% Superblock and added to the ELISA wells. Detection of the AA was achieved using an Anti-His6-HRP conjugate at a dilution of 1:5000. MMP-9 activation of MMP-9 cleavable masked anti-VEGF scFv His AA is presented in Figure 6.

ELISA assay of an AA construct containing: MM / MMP-9 CM / anti-VEGF scFv-Fc

[00388] Fifty microliters of HEK cell supernatant was added to 200 μ L MMP-9 digestion buffer and incubated with or without \sim 19U MMP-9 for 2 hrs at 37°C. Samples were then diluted 1:1 in PBST, 10% Superblock and 100 μ L were added to the ELISA wells. Detection of the AA was achieved using Anti-human Fc-HRP conjugate at a dilution of 1:2500. MMP-9 activation of MMP-9 cleavable masked anti-VEGF scFv-Fc is presented in Figure 7.

Purification and assay of an AA construct containing: MM / MMP-9 CM / anti-VEGF scFv-Fc

[00389] Anti-VEGF scFv Fc AAs were purified using a Protein A column chromatography. Briefly, 10 mLs of HEK cell supernatants were diluted 1:1 with PBS and added to 0.5 mL Protein A resin pre-equilibrated in PBS. Columns were washed with 10 column volumes of PBS before eluting bound protein with 170 mM acetate, 300 mL NaCl pH. 2.5 and immediately neutralized 1 mL fractions with 200 μ L of 2 M Tris pH 8.0. Fractions containing protein were then concentrated using Amicon Ultra centrifugal concentrators. ELISA was conducted as with HEK cell supernatants. ELISA data showing the MMP-9 dependent VEGF binding of Anti-VEGFscFv Fc AA constructs with the MMs 306 and 314 that were purified using a Protein A column are presented in Figure 8.

Example 9: Target Displacement Assay for the Discovery and Validation of Efficiently Masked Therapeutic Proteins

[00390] VEGF was adsorbed to the wells of a 96-well micro-titer plate, washed and blocked with milk protein. 25 ml of culture media containing anti-VEGF antibody or anti-VEGF AA's containing the MM JS306, was added to the coated wells and incubated for 1, 2, 4, 8 or 24 hours. Following incubation, the wells were washed and the extent of bound AA's was measured by anti-huIgG immunodetection. Figure 9 shows mask 306 can completely inhibit binding to VEGF at one hour; however, at 16 hours, >50% of the 306-antiVEGF AA is bound to its antigen, VEGF. The 306 mask, which binds to anti-VEGF antibody with an affinity of > 600 nM, does not efficiently preclude binding to VEGF.

Example 10: Library Screening and Isolation of anti-CTLA4 MMs

[00391] CTLA4 antibody masking moieties (MMs) were isolated from a combinatorial library of 10^{10} random 15mer peptides displayed on the surface of *E. coli* according to the method of Bessette et al (Bessette, P.H., Rice, J.J and Daugherty, P.S. Rapid isolation of high-affinity protein binding peptides using bacterial display. Protein Eng. Design & Selection. 17:10,731–739, 2004). Biotinylated mouse anti-CTLA4 antibody (clone UC4 F10-11, 25 nM) was incubated with the library and antibody-bound bacteria expressing putative binding peptides were magnetically sorted from non-binders using streptavidin-coated magnetic nanobeads. Subsequent rounds of enrichment were carried out using FACS. For the initial round of FACS, bacteria were sorted using biotinylated target (5 nM) and secondary labeling step with streptavidin phycoerythrin. In subsequent rounds of FACS, sorting was performed

with Dylight labeled antibody and the concentration of target was reduced (1 nM, then 0.1 nM) to avoid the avidity effects of the secondary labeling step and select for the highest affinity binders. One round of MACS and three rounds of FACS resulted in a pool of binders from which individual clones were sequenced. Relative affinity and off-rate screening of individual clones were performed using a ficin digested Dylight-labeled Fab antibody fragment to reduce avidity effects of the bivalent antibody due to the expression of multiple peptides on the bacterial surface. As an additional test of target specificity, individual clones were screened for binding in the presence of 20 uM E. Coli depleted IgG as a competitor. Amino acid and nucleotide sequences of the 4 clones chosen for MM optimization are shown in Table 15. These sequences will interchangeably referred to as 115MM, 184MM, 182MM, and 175MM. MM candidates with a range of off-rates were chosen, to determine the effects of off-rates on MM dissociation after cleavage. An MM that did not bind anti-CTLA4 was used as a negative control.

Table 15: Amino acid and nucleotide sequences for MMs that mask anti-CTLA4

KK115 MM																			
M	I	L	L	C	A	A	G	R	T	W	V	E	A	C	A	N	G	R	
ATGATTTTGTGTGCGCGGCGGGTCGGACGTGGGTGGAGGCTTGCGCTAA TGGTAGG																			
KK184 MM																			
A	E	R	L	C	A	W	A	G	R	F	C	G	S						
GCTGAGCGGTTGTGCGCGTGGGCGGGGCGGTTCTGTGGCAGC																			
KK182 MM																			
W	A	D	V	M	P	G	S	G	V	L	P	W	T	S					
TGGGCGGATGTTATGCCTGGGTCGGGTGTGTTGCCGTGGACGTCG																			
KK175 MM																			
S	D	G	R	M	G	S	L	E	L	C	A	L	W	G	R	F	C	G	S
AGTGATGGTCGTATGGGGAGTTTGGAGCTTTGTGCGTTGTGGGGGCGGTTCTGTGGCAGC																			
Negative control (does not bind anti-CTLA4)																			
P	C	S	E	W	Q	S	M	V	Q	P	R	C	Y	Y					
GCCGTGTTCTGAGTGGCAGTCGATGGTGCAGCCGCGTTGCTATTA																			

Example 11: Cloning of Anti-CTLA4 scFv

[00392] Anti-CTLA4 ScFv was cloned from the HB304 hybridoma cell line (American Type Culture Collection) secreting UC4F10-11 hamster anti-mouse CTLA4 antibody according to the method of Gilliland et al. (Gilliland L. K., N. A. Norris, H. Marquardt, T. T. Tsu, M. S. Hayden, M. G. Neubauer, D. E. Yelton, R. S. Mittler, and J. A. Ledbetter.. Rapid and reliable cloning of antibody variable regions and generation of recombinant single chain antibody fragments. Tissue Antigens 47:1, 1-20, 1996). A detailed version of this protocol can be found at the website located by placing [http://www](http://www.ibms.sinica.edu.tw/~sroff/protocols/scFv.htm) in front of [.ibms.sinica.edu.tw/~sroff/protocols/scFv.htm](http://www.ibms.sinica.edu.tw/~sroff/protocols/scFv.htm). In brief, total RNA was isolated from hybridomas using the RNeasy total RNA isolation kit (Qiagen). The primers IgK1 (gtyttrtngngtnacyterca) and IgH1 (acdatytyttrtenacyttngt) (Gilliland et al. referenced above) were used for first strand synthesis of the variable light and heavy chains, respectively. A poly G tail was added with terminal transferase, followed by PCR using the 5' ANCTAIL primer (Gilliland et al. referenced above)

(cgtcgatgagctctagaattcgcatgtgcaagtccgatgggtcccccccccccccc) containing EcoRI, SacI and XbaI sites for both light and heavy chains (poly G tail specific) and the 3' HBS-hIgK (cgtcgatgtcgacggatccaagcttacyttccayttnacrttdatrtc) and HBS-hIgH (cgtcgatgtcgacggatccaagcttrcangcngngcnarnnggrtanac) derived from mouse antibody constant region sequences and containing HindIII, BamHI and Sall sites for light and heavy chain amplification, respectively (Gilliland et al. referenced above). Constructs and vector were digested with HindIII and SacI, ligated and transformed into E.Coli. Individual colonies were sequenced and the correct sequences for V_L and V_H (Tables 16 and 17 respectively) were confirmed by comparison with existing mouse and hamster antibodies. The leader sequences, as described for anti-CTLA4 in the presented sequence is also commonly called a signal sequence or secretion leader sequence and is the amino acid sequence that directs secretion of the antibody. This sequence is cleaved off, by the cell, during secretion and is not included in the mature protein. Additionally, the same scFv cloned by Tuve et al (Tuve, S. Chen, B.M., Liu, Y., Cheng, T-L., Toure, P., Sow, P.S., Feng, Q., Kiviat, N., Strauss, R., Ni, S., Li, Z., Roffler, S.R. and Lieber, A. Combination of Tumor Site –Located CTL-Associated Antigen-4 Blockade and Systemic Regulatory T-Cell Depletion Induces Tumor Destructive Immune Responses. Cancer Res. 67:12, 5929-5939, 2007) was identical to sequences presented here.

Table 16: Hamster anti-mouse CTLA4 V_L

Leader																			
M	E	S	H	I	H	V	F	M	S	L	F	L	W	V	S	G	S	C	A
L	S	V	S	A	G	E	K	A	T	I	S	C	K	S	S	Q	S	L	F
L	N	W	Y	L	Q	K	P	G	Q	S	P	K	L	L	I	Y	Y	A	S
R	F	R	G	S	G	T	D	F	T	L	T	I	S	S	V	Q	D	E	D
W	Y	D	Y	P	Y	T	F	G	A	G	T	K	V	E	I	K			
atggaatcacatatccatgtcttcattgtccttgggtgtctggttcctgtgcagacatcatgatgacccagtccttcattccctga																			
gtgtgtcagcgggagagaaaagccactatcagctgcaagtccagtcagagtcctttcaacagtaacgccaaaacgaactactgaactgg																			
tatttgcagaaaccagggcagtcctcctaaactgctgatctattatgcattcactagggcactggtggtcctgatcgcttcagaggcagtg																			
gatctgggacggatttcacttcaccatcagcagtgccaggatgaagacctggcattttattactgtcagcagtggtatgactaccata																			
cacgttcggagctgggaccaaggtggaaatcaaa																			

Table 17: Hamster anti mouse CTLA4 V_H

Leader																			
K	M	R	L	L	G	L	L	Y	L	V	T	A	L	P	G	V	L	S	Q
L	V	N	P	S	Q	S	L	S	L	S	C	S	V	T	G	Y	S	I	T
Q	F	P	G	Q	K	V	E	W	M	G	F	I	Y	Y	E	G	S	T	Y
I	S	I	T	R	D	T	S	K	N	Q	F	F	L	Q	V	N	S	V	T
A	R	Q	T	G	Y	F	D	Y	W	G	Q	G	T	M	V	T	V	S	S
aagatgagactgttgggtcttctgtacctggtgacagcccttctggtgtcctgtccagatccagcttcaggagtcaggacctggcctggt																			
gaaccctcacaatcactgtcctctcttctgtcactggttactccatcaccagtgggtatggatggaactggatcaggcagttccag																			
ggcagaaggtggagtggatgggattcatatattatgagggttagcactactacaacccttccatcaagagccgcattccatcaccagag																			
acacatcgaagaaccagttcttctgcaggtgaattctgtgaccactgaggacacagccacatattactgtgcgagacaaactgggtact																			
ttgattactggggccaaggaacatggtcaccgtctcctca																			

Example 12: Construction of the anti-CTLA4 scFv with MMs and CMs

[00393] To determine the optimal orientation of the anti-CTLA4 scFv for expression and function, primers were designed to PCR amplify the variable light and heavy chains individually, with half of a (GGGS)₃ linker at either the N- or C-terminus for a subsequent ‘splicing by overlapping extension’ PCR (SOE-PCR; Horton, R.M., Hunt, H.D., Ho, S.N., Pullen, J.K. and Pease, L.R. (1989) Engineering hybrid genes without the use of restriction enzymes: gene splicing by overlap extension. Gene 77, 61-68) with either V_H or V_L at the N-terminus. An NdeI restriction site was engineered at the N-terminus to generate a start codon in frame at the beginning of the nucleotide sequence and a His tag and stop codon were added to the C-terminus. Light and heavy chains were then joined via sewing PCR using the outer primers to generate ScFvs in both V_HV_L and V_LV_H (Figure 10). Primers are shown below in Table 18.

Table 18: Primers to generate scFvs V_HV_L and V_LV_H

VL for1	caaggaccatagcatatggacatcatgatgacccagtct
VL linker	
rev1	acttcgcctccacctgatccaccaccacctttgattccaccttggtcc
linker VH	
for2	ggatcaggtggaggcgggaagtggaggtggcgggtcccagatccagcttcaggagtcagga
VH his rev2	ggccggatccaagcttttagtggtgatggtgatgatgtgaggagacggtgacctggttcc
VH for3	acaaggaccatagcatatgcagatccagcttcaggagtca
VH linker	
rev3	acttcgcctccacctgatccaccaccacctgaggagacggtgacctggttcc
linker VL for4	ggtggatcaggtggaggcgggaagtggaggtggcgggtccgacatcatgatgacccagtctcct
VL his rev4	cggccggatccaagcttttagtggtgatggtgatgatgtttgattccaccttggtcccagc

[00394] Next, a set of overlapping primers were designed to add sfi and xho1 sites for MM cloning followed by the MMP-9 cleavage sequence and (GGS)₂ linker on the N-terminus of the ScFv constructs. These primers are presented in Table 19 and shown schematically in Figure 10.

Table 19: Primers MM and CM cloning

for 1c linker	gccagtctggccggtagggtcgcagcggccaagtgcacatgccactgggcttctgggtc
for 1d linker VL	gccactgggcttctgggtccgggtggaagcggcggtcagacatcatgatgacccagtc
for 1e linker VH	gccactgggcttctgggtccgggtggaagcggcggtcagacatccagcttcaggagtca
for 1a	ttaccaacaaggaccatagcatatgggccagtctggccggtagggc
VH his rev2	ggccggatccaagcttttagtggtgatggtgatgatgtgaggagacggtgacctggttcc
VH linker rev3	acttcgcctccacctgatccaccaccacctgaggagacggtgacctggttcc

[00395] Linker containing ScFvs were PCR amplified, digested with NdeI and EcoRI (an internal restriction site in V_H) and gel purified. The PCR fragments were ligated into the vectors and transformed into E. coli. The nucleotide and amino acid sequences are presented in Table 20..

Table 20: Sequence of MM linker–CM–anti-CTLA4 scFv linker

Amino acid sequence:	
(-----MM Linker-----)	(-----CM-----) (---scFv Linker---)
G G S G G S G G S S	G Q V H M P L G F L G P G G S G G S

Nucleotide sequence:

GGCGGTTCTGGTGGCAGCGGTGGCTCGAGCGGCCAAGTGCACATGCCACTGGGCTT
CCTGGGTCCGGGTGGAAGCGGCGGCTCA

[00396] MM sequences were PCR amplified, digested at *sf*I and *xho*I sites, ligated into linker anti-CTLA4 scFv constructs, transformed into *E. coli* and sequenced. The complete nucleotide and amino acid sequences of the MM115-CM-AB are shown below in Tables 21 and 22 respectively.

Table 21: Amino acid sequence of MM115-anti-CTLA4 ScFv AB

M	I	L	L	C	A	A	G	R	T	W	V	E	A	C	A	N	G	R	G	G	S	G	G	S	G	G	S	S	G	Q	V	H	M		
P	L	G	F	L	G	P	G	G	S	G	G	S	Q	I	Q	L	Q	E	S	G	P	G	L	V	N	P	S	Q	S	L	S	L	S	C	S
V	T	G	Y	S	I	T	S	G	Y	G	W	N	W	I	R	Q	F	P	G	Q	K	V	E	W	M	G	F	I	Y	Y	E	G	S	T	
Y	Y	N	P	S	I	K	S	R	I	S	I	T	R	D	T	S	K	N	Q	F	F	L	Q	V	N	S	V	T	T	E	D	T	A	T	Y
Y	C	A	R	Q	T	G	Y	F	D	Y	W	G	Q	G	T	M	V	T	V	S	S	G	G	G	G	S	G	G	G	G	S	G	G	G	G
G	G	S	D	I	M	M	T	Q	S	P	S	S	L	S	V	S	A	G	E	K	A	T	I	S	C	K	S	S	Q	S	L	F	N	S	
N	A	K	T	N	Y	L	N	W	Y	L	Q	K	P	G	Q	S	P	K	L	L	I	Y	Y	A	S	T	R	H	T	G	V	P	D	R	
R	F	R	G	S	G	S	G	T	D	F	T	L	T	I	S	S	V	Q	D	E	D	L	A	F	Y	Y	C	Q	Q	W	Y	D	Y	P	
Y	T	F	G	A	G	T	K	V	E	I	K																								

Table 22: Nucleotide sequence of MM115-anti-CTLA4 ScFv AB

atgattttgtgtgcgcggcggtcgacgtgggtggaggttgcgctaattgtagggcggttctggtggcagcgggtggctcgagcggccaagtgc catgccactgggcttctgggtccgggtggaagcggcggtcacagatccagcttcaggagtcaggacctggcctggtgaaccctcacaatcactgc cctctcttgcctgtcactggttactccatcaccagtgggttatggatggaactggatcaggcagttcccagggcagaaggtggagtggatgggattcatata ttatgagggtagcacctactacaaccctccatcaagagccgcattccatcaccagagacacatcgaagaaccagttcttctgcaggtgaattctgtgac cactgaggacacagccacatattactgtgcgagacaaactgggtactttgattactggggccaaggaaccatggtcaccgtctcctcaggtgggtgga tcaggtggagggcgaagtggaggtggcggtccgacatcatgatgaccagtccttcctcagtgagtggtgcagcgggagagaaagccactatcagc tgcaagtccagtcagagtcctttcaacagtaacgccaaaacgaactactgaactggattttgcagaaaccagggcagtcctcctaaactgctgatctattatg catccactaggcactactggggtccctgatcgcttcagaggcagtggtatctgggacggattcactctcaccatcagcagtgccaggatgaagacctggc atcttattactgtcagcagtggtatgactaccatacacgttcggagctgggaccaaggtggaaatcaaacatcaccatcaccactaa

[00397] To generate MM-CM-anti-CTLA4 scFv-Fc fusions, the following primers listed in Table 23 were designed to PCR amplify the constructs for cloning into the pfuse Fc vector via the in fusion system (Clontech). Plasmids were transformed into *E. coli*, and the sequence of individual clones was verified.

Table 23: Primers to generate MM-CM-anti-CTLA4 scFv-Fc fusions

HLCTLA4ScFv pFuse reverse	tcagatctaaccatggctttgattccaccttggtcc
LHCTLA4ScFv pFuse reverse	tcagatctaaccatggctgaggagacggtgacctgg
p115CTLA4 pfuse forward	cacttgtcacgaattcgatgattttgtgtgcgcggc
p182CTLA4 pfuse forward	cacttgtcacgaattcggtggcggtatgtatgcctg
p184CTLA4 pfuse forward	cacttgtcacgaattcggtgagcgggtgtgcgcgtg
p175CTLA4 pfuse forward	cacttgtcacgaattcgagtgtggtcgtatggggag
pnegCTLA4 pfuse forward	cacttgtcacgaattcgctgtgtctgagtggcagtcg

Example 13: Expression and Assay of masked/MMP-9/ anti-CTLA4 scFv-Fc in HEK-293 cells

[00398] 10 ug of expression vectors for p175CTLA4pfuse, p182CTLA4pfuse, p184CTLA4pfuse, p115CTLA4pfuse, or pnegCTLA4pfuse were introduced into 10^7 HEK-293 freestyle cells (Invitrogen) by transfection using transfectamine 2000 as per manufacturer's protocol (Invitrogen). The transfected cells were incubated for an additional 72 hours. After incubation the conditioned media was harvested and cleared of cells and debris by centrifugation. The conditioned media was assayed for activity by ELISA as described below.

[00399] Fifty microliters of conditioned media from HEK-293 expressing MM175-anti-CTLA4 scFv, MM182-anti-CTLA4 scFv, MM184-anti-CTLA4 scFv, MM115-anti-CTLA4 scFv, or MMneg-anti-CTLA4 scFv was added to 200 μ L MMP-9 digestion buffer and incubated with or without \sim 19U MMP-9 for 2 hrs at 37°C. Samples were then diluted 1:1 in PBS, 4% non fat dry milk (NFDM) and assayed for binding activity by competition ELISA.

[00400] 100 ul of 0.5 mg/ml solution of murine CTLA4-Fc fusion protein (R & D systems) in PBS was added to wells of 96 well Easy Wash plate (Corning) and incubated overnight at 4°C. Wells were then blocked for one hour at room temperature (RT) with 100 ul of 2% non-fat dry milk (NFDM) in PBS and then washed 3X with PBS; 0.05% Tween-20 (PBST). 50 ul of conditioned media from cultures of transfected HEK-293 cells expressing MM175-anti-CTLA4 scFv, MM182-anti-CTLA4 scFv, MM184-anti-CTLA4 scFv, MM115-anti-CTLA4 scFv, or MMneg-anti-CTLA4 scFv that had previously been untreated or treated with MMP-9, were added to wells and incubated RT for 15 minutes. Following incubation, 50 ul of PBS containing 0.5 ug/ml biotinylated murine B71-Fc (R & D systems) was added to each well. Following a further incubation at RT of 30 minutes the wells were washed 5X with 150 ul PBST. 100 ul of PBS containing 1:3000 dilution of avidin-HRP was added and the plate incubated at RT for 45 minutes and then washed 7X with 150 ul PBST. The ELISA was developed with 100 ul of TMB (Pierce), stopped with 100 uL of 1N HCL and the absorbance was measured at 450 nM.

Example 14: Construction of an anti-CTLA4

[00401] Tables 24 and 25 display nucleotide and amino acid sequences for anti-human CTLA-4 scFv, respectively. M13 bacteriophage capable of binding human CTLA were supplied (under contract, by Creative Biolabs, 21 Brookhaven Blvd., Port Jefferson Station, NY 11776). Phage were produced in *E. coli* TG-1 and purified by PEG;NaCl precipitation.

Table 24: anti-human CTLA4 scFv AB nucleotide sequence

```
gaaattgtgtgacacagtctccaggcaccctgtctttgtctccaggggaaagagccaccctctcctgcagggccagtcagagtgttagcagcagctactt
agcctggtaccagcagaaacctggccaggctcccaggctcctcatctatggtgcatccagcagggccactggcatcccagacaggttcagtggcagtg
ggtctgggacagacttcaacttcaccatcagcagactggagcctgaagattttgcagtgtattactgtcagcagtatggtagctcaccgctcactttcggcg
gagggaccaaggtggaaatcaaacttccggagggtcgaccataactcgtataatgtatactatacgaagttatcctcgagcgggtaccaggtgcagct
ggtgcagactgggggagggcgtggtccagcctgggaggtccctgagactctctgtgcagcctctggatccaccttagcagctatgccatgagctgggt
ccgccaggctccagggaaggggctggagtgggtctcagctattagtggtagtggttagcacatactacgcagactccgtgaagggccggttcacca
tctccagagacaattccaagaacacgctgtatctgaaatgaacagcctgagagccgaggacacggccgtatattactgtgcgacaaactccctttactg
gtacttcgatctctggggccgtggcaccctggcactgtctcttcagctagc
```


Table 25: anti-human CTLA4 scFv AB amino acid sequence

EIVLTQSPGTLSPGERATLSCRASQSVSSSYLAWEYQQKPGQAPRLLIYGASSRATGIPDRFSGSG
 SGTDFTLTISRLEPEDFAVYYCQQYGSSPLTFGGGGTKVEIKRSGGSTITSYNVYYTKLSSSGTQV
 QLVQTGGGVVQPGRSLRLSCAASGSTFSSYAMSWVRQAPGKGLEWVSAIAGSGGSTYYADSVK
 GRFTISRDNSKNTLYLQMNSLRAEDTAVYYCATNSLYWYFDLWGRGTLVTVSSAS

[00402] *Phage ELISA measurement of CTLA-4 binding:* To measure the binding of anti-CTLA-4 scFv-C2, 100 ul of a 0.5 ug/ml Human CTLA-4-IgG or murine CTLA-4-IgG (R&D Systems) in PBS was added to microwells (96 Well Easy Wash; Corning) and incubated overnight at 4°C. Wells were then blocked for 1 hour at room temperature (RT) with 150 ul of 2% non-fat dry milk (NFDM) in PBST (PBS, pH 7.4, 0.5% Tween-20). The wells were then washed 3X with 300 ul PBST. Following washing 100 ul of purified anti-CTLA-4 scFv phage in PBST were added to triplicate wells and incubated RT for 1 hr. The wells were then washed 3X with 300 ul PBST. One hundred microliters of anti-M13 HRP-conjugated antibody was then added and incubated at RT for 1 hr. Detection of HRP was completed using 100 ul of TMB one (Pierce) solution. The reaction was stopped 100 ul of 1N HCL and the absorbance was measured at 450 nM. Figure 19 shows the binding of anti-CTLA4 scFv to both murine and human CTLA4.

AAs comprising an IgG as the AB

[00403] Examples of AAs comprising an anti-EGFR and anti-VEGF in the human IgG are described in the following sections. These AAs are masked and inactive under normal conditions. When the AAs reach the diseased tissue, they are cleaved by a disease-specific protease and can then bind their target. Bacterial display is used to discover suitable MMs for the anti-EGFR and anti-VEGF antibodies. In, these examples, selected MMs are combined with an enzyme substrate to be used as a trigger to create AAs that become competent for specific binding to target following protease activation. Furthermore, bacterial display is used to alter the discovered peptides to increase affinity for the ABs and enhance the inhibition of targeted binding in the un-cleaved state. The, increased MM affinity and enhanced inhibition is important for appropriate AA function.

Example 15: Construction of an anti-VEGF IgG AA

Construction of the anti-VEGF IgG antibody

[00404] The anti-VEGF light chain variable region was PCR amplified with primers CX0311 and CX0702 using the anti-VEGF mmp-9 306 scFv (described above) as template and then cloned into the pFIL2-CL-hk vector using the EcoRI and BsiWI restriction sites (pFIL2-VEGF-Lc). The 306 mmp-9 light chain was PCR amplified with primers CX0325 and CX0702 using the anti-VEGF mmp-9 scFv as template and cloned as above (pFIL2-306mVEGF-Lc). The anti-VEGF heavy chain variable regions

were PCR amplified using primers CX0700 and CX0701 using the 306 MM / MMP-9 CM / anti-VEGFscFv (described above) as template and cloned into the pFIL-CHIg-hG1 vector using the EcoRI and NheI restriction sites (pFIL-VEGF-Hc). The primers are provided below in Table 26.

Table 26: Primers for Construction of an anti-VEGF IgG antibody

CX0311 cttgtcacgaattcggatattcaactgacccagagc
CX0702 gtgcagccaccgtacgcttaatctccactttggtg
CX0325 tgettgtcaactctacgtc
CX0289 gctttcaccgcaggtacttccgtagctggccagtctggcc
CX0687 cgctccatggggccaccttggccgctgccaccgctcgagcc
CX0700 cacttgtcacgaattcggaggtccagctggtagaaag
CX0701 ggcccttggtgctagcgtcgacactgtaaccagagtac

Table 27: Sequences for heavy and light chain anti-VEGF antibody

pFIL2-CL-hk anti-VEGF Lc (pFIL2-VEGF-Lc)

gatattcaactgacccagagcccttctccctgagtgccagcgtgggtgaccgtgttacgateacttgctcggccagccaagatatttctaactacctgaatt
ggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcactgcattccggcgtaccgtcgcgctttagecggttctggcagtgg
taccgacttcacctgactatctcgagtctgcaacctgaggattttgctacatattactgtcagcaatattcgaccgtgccgtggacgttcgggcagggcac
caaagtggagattaagcgtacggttggtgcaccatctgtcttcattctcccgccatctgatgagcagttgaaatctggaactgcctctgttgtgtgcctgctg
aataactctatcccagagaggccaaagtacagtgggaaggtggataacgccctccaatcgggtaactcccaggagagtgtcacagagcaggacagcaa
ggacagcacctacagcctcagcagcacctgacgctgagcaagcagactacgagaaacacaaagtctacgcctgcgaagtcacccatcagggcctg
agctcgcccgtcacaaagagcttcaacaggggagagtgttag

DIQLTQSPSSLSASVGDRVITITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGS
GTDFTLTISLQPEDFATYYCQQYSTVPWTFGQGKVEIKRTVAAPSVFIFPPSDEQLKSGTASVV
CLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSSTLSSTLTLSKADYEKHKVYACEV
THQGLSSPVTKSFNRGEC

[00405] As described above, the mask 306, used for anti-VEGF AA development did not efficiently mask the target binding over long exposure to target, due to low affinity of the MM for the AB. One approach to increasing the affinity of the MM is to subject the peptide to affinity maturation as described below.

Library construction for Affinity Maturation

[00406] The 306 anti-VEGF MM was affinity matured by using a soft randomization approach. An ecpX cell display library was constructed in with the nucleotide ratios shown in Table 28. The final library diversity (306 SR) was approximately 2.45×10^8 .

Table 28

Original Base	Ratio of Bases
G	G=70%; T=8%; A=11%; C=11%
T	T=70%; G=8%; A=11%; C=11%
A	A=80%; G=5%; T=6%; C=9%
C	C=80%; G=5%; T=6%; A=9%

306 SR library screening

[00407] An initial MACS round was performed with protein-A labeled magnetic beads and a number of cells that provided greater than 100X oversampling of the library. Prior to magnetic selection the cells were incubated with 100 nM anti-VEGF IgG and 10 μ M 306 peptide (306P, PCSEWQSMVQPRCYYG), to reduce the binding of variants with equal or lower affinity than the original 306 sequence. Magnetic selection resulted in the isolation of 2×10^7 cells.

[00408] The first round of FACS sorting was performed on cells labeled with 1 nM DyLight (fluor 530 nM)-anti-VEGF. To apply selective pressure to the population, the second and third round of FACS was performed on cells labeled with 1 nM DyLight-anti-VEGF in the presence of 100 nM 306P. Selection gates were set so that only 5% of cells with the strongest binding were collected. The population of cells sorted in the third round were first incubated with 10 nM DyLight-anti-VEGF followed by addition of 306P to a final concentration of 100 nM and incubated at 37°C for 20 minutes. The brightest 2% of the positive population was collected, representing binding that was not competed by 306P. FACS rounds 5 through 7 were done as follows; the populations were labeled with 10 nM DyLight labeled anti-VEGF and then competed off with unlabeled VEGF (100 nM) at 37°C for 7, 10, and 15 minutes, respectively. The brightest 1% were sorted in FACS rounds 5 through 7.

Table 29

306SR M1F7 peptide sequences
JS306 PCSEWQSMVQPRCYYG
JS1825 SCTAWQSMVEQRCYFG 3X
JS1826 PCSKWESMVEQRCYFA
JS1827 PCSAWQSMVEQRCYFG 2X
JS1829 PCSKWESMVLQSCYFG 4X
JS1830 TCSAWQSMVEQRCYFG 2X
JS1837 TCSQWESMVEPRCYYG

306SR affinity matured peptide analysis

[00409] Binding of the eCPX3.0 clones 306, JS1825, JS1827, and JS1829 were analyzed on FACS at 3 different concentrations of DyLight labeled anti-VEGF. The binding curves are shown in Figure 21. All three of the affinity matured peptides displayed at least 10 fold higher affinity than 306P.

Construction of anti-VEGF AAs

[00410] Affinity matured ecpX3.0 clones (JS1825, JS1827, and JS1829) were PCR amplified using primers CX0289 and CX0687 and cloned into pFIL2-306mVEGF-Lc using the SfiI restriction sites to produce the vectors pFIL2-1825mVEGF-Lc, pFIL2-1827mVEGF-Lc, and pFIL2-1829mVEGF-Lc. The nucleotide and amino acid sequences are provided in the tables following. Parentheses delineate the demarcations between the various sequence domains: (Linker)(MM)(Linker)(CM)(Linker)(AB).

Table 30: Sequences of anti-VEGF AA: pFIL2-CL-hk anti-VEGF mmp-9 306 Lc (pFIL2-306mVEGF-Lc)

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttctattatgggggcggttctggtggcagcggccaaggtggccaagt
 gcacatgccactgggcttctgggtccgggcggttctgatattcaactgaccagagcccttcttccctgagtgccagcgtgggtgaccgtgttacgatca
 cttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcactgcatt
 ccggcgtaccgtcgcgcttagcgggttctggcagtggtaccgacttcaccctgactatctcagttctgcaacctgaggattttgctacatattactgtcagca
 atattcgaccgtgccgtggacgttcgggcagggcaccaaagtggagattaagcgtacgggtggctgcaccatctgtcttcatcttcccgccatctgatgagc
 agttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaaaggtggataacgccctccaatcgggt
 aactcccaggagagtgacacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgagaaaca
 caaagtctacgctgcgaagtcacccatcagggcctgagctcggcgcacaaagagcttcaacaggggagagtgtag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSEWQSMVQPRCYYG)	(GGSGGSGQGG)	(QVHMPLGFLGP)	(GGS)	(DIQLTQSPSSLSA
SVGDRV	TITCSASQDISNYLNWYQQKPGKAPKVL	IYFTSSLHSGVPSRFS	SGSGSGTDFTL	TISSLQP	EDFATYYCQQYSTVPWTFGQG
TKVEIKRTVAAPS	VFIFPPSDEQLKSGTASVVCLLN	NFY	PREAK	VQWKVDNALQSGNSQESVTEQ	DSKDYSLSTLTL
SKADYEKHKVYACEVTHQGLSSPVT	KSF	NRGEC)			

Table 31: Sequences of anti-VEGF AA: pFIL2-CL-hk anti-VEGF mmp-9 1825 Lc (pFIL2-1825mVEGF-Lc)

ggccagtctggccagtcgtgtacggcgtggcagtcgatggtggagcagcgttgctattttgggggctcgagcgggtggcagcggccaaggtggccaag
 tgcacatgccactgggcttctgggtccgggcggttctgatattcaactgaccagagcccttcttccctgagtgccagcgtgggtgaccgtgttacgatc
 acttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcactgcatt
 tccggcgtaccgtcgcgcttagcgggttctggcagtggtaccgacttcaccctgactatctcagttctgcaacctgaggattttgctacatattactgtcagc
 aatattcgaccgtgccgtggacgttcgggcagggcaccaaagtggagattaagcgtacgggtggctgcaccatctgtcttcatcttcccgccatctgatgag
 cagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaaaggtggataacgccctccaatcggg
 taactcccaggagagtgacacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgagaaaca
 caaagtctacgctgcgaagtcacccatcagggcctgagctcggcgcacaaagagcttcaacaggggagagtgtag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(SCTAWQSMVEQRCYFG)	(GSSGGSGQGGQ)	(VHMPLGFLGP)	(GGS)	(DIQLTQSPSSLSA
SVGDRV	TITCSASQDISNYLNWYQQKPGKAPKVL	IYFTSSLHSGVPSRFS	SGSGSGTDFTL	TISSLQP	EDFATYYCQQYSTVPWTFGQG
TKVEIKRTVAAPS	VFIFPPSDEQLKSGTASVVCLLN	NFY	PREAK	VQWKVDNALQSGNSQESVTEQ	DSKDYSLSTLTL
SKADYEKHKVYACEVTHQGLSSPVT	KSF	NRGEC)			

Table 32: Sequences of anti-VEGF AA: pFIL2-CL-hk anti-VEGF mmp-9 1827 Lc (pFIL2-1827mVEGF-Lc)

ggccagtctggccagccgtgttctgcgtggcagtcgatggtggagcagcgttgctattttgggggctcgagcgggtggcagcggccaaggtggccaagt
 gcacatgccactgggcttctgggtccgggcggttctgatattcaactgaccagagcccttcttccctgagtgccagcgtgggtgaccgtgttacgatca
 cttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcactgcatt
 ccggcgtaccgtcgcgcttagcgggttctggcagtggtaccgacttcaccctgactatctcagttctgcaacctgaggattttgctacatattactgtcagca
 atattcgaccgtgccgtggacgttcgggcagggcaccaaagtggagattaagcgtacgggtggctgcaccatctgtcttcatcttcccgccatctgatgagc
 agttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaaaggtggataacgccctccaatcgggt
 aactcccaggagagtgacacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgagaaaca
 caaagtctacgctgcgaagtcacccatcagggcctgagctcggcgcacaaagagcttcaacaggggagagtgtag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSAWQSMVEQRCYFG)	(GSSGGSGQGG)	(QVHMPLGFLGP)	(GGS)	(DIQLTQSPSSLSA
SVGDRV	TITCSASQDISNYLNWYQQKPGKAPKVL	IYFTSSLHSGVPSRFS	SGSGSGTDFTL	TISSLQP	EDFATYYCQQYSTVPWTFGQG
TKVEIKRTVAAPS	VFIFPPSDEQLKSGTASVVCLLN	NFY	PREAK	VQWKVDNALQSGNSQESVTEQ	DSKDYSLSTLTL
SKADYEKHKVYACEVTHQGLSSPVT	KSF	NRGEC)			

Table 33: Sequences of anti-VEGF AA: pFIL-CL-hk anti-VEGF mmp-9 1829 Lc (pFIL2-1829mVEGF-Lc)

ggccagtctggccagccgtgttctaagtgggaatcgatggtgctgcagagttgctattttggcggctcgagcgggtggcagcggccaaggtggccaagtg
cacatgccactgggcttctgggtccgggcgggttctgatattcaactgaccagagcccttcttcctgagtgccagcgtgggtgaccgtgttacgacact
tgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcactgcattcc
ggcgtaccgtcgcgctttagcgggttctggcagtggtaccgacttcaccctgactatctcgagctgcaacctgaggattttgctacatattactgtcagcaat
attcgaccgtgccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacgggtggctgcaccatctgtcttcattctccgccatctgatgagca
gttgaaatctggaactgcctctgtgtgtgctgctgaataacttctatcccagagaggccaaagtacagtggaaaggtggataacgccctccaatcgggta
actcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgagaaacac
aaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgtcacaaagagcttcaacaggggagagtgttag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSKWESMVLQSCYFG)	(GSSGGSGQGG)	(QVHMPLGFLGP)	(GGS)	(DIQLTQSPSSLAS
VGDRVTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTISSLQPE					
DFATYYCQQYSTVPWTFGQGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKV					
QWKVDNALQSGNSQESVTEQDSKSTYSLSTLTLSKADYEKHKVYACEVTHQGLSPVTKSFN					
RGEC)					

Expression and purification of anti-VEGF antibody and AA

[00411] 3 µg of pFIL-VEGF-Hc and 3 µg pFIL2-VEGF-Lc were co-transfected into CHO-S cells (Invitrogen) using Lipofectamine 200 (Invitrogen) according to manufacturers protocol. Transfected cells were cultured in Freestyle CHO media (Invitrogen) and selected for resistance to zeocin and blasticidin. Individual clones were isolated by limiting dilution and selected for expression of human IgG capable of binding EGFR by ELISA. All antibodies and AAs are purified by Protein-A chromatography using standard techniques.

[00412] Likewise, 3 µg of each expression vector for AA light chains pFIL2-306mVEGF-Lc, pFIL2-1825mVEGF-Lc, pFIL2-1827mVEGF-Lc, or pFIL2-1829mVEGF-Lc was co-transfected into CHO-S cells with 3 µg pFIL-VEGF-Hc. Transfected cells were cultured in Freestyle CHO media (Invitrogen) and selected for resistance to zeocin and blasticidin. Individual clones were isolated by limiting dilution and selected for expression of human IgG capable of binding EGFR by ELISA.

Target Displacement Assay of anti-VEGF antibody and AA

[00413] VEGF is adsorbed to the wells of a 96 well micro-titer plate, washed and blocked with milk protein. About 25 µl of culture media containing anti-VEGF antibody or anti-VEGF AA's containing the MM's JS306, JS1825, JS1827 and JS 1829 is added to the coated wells and incubated for about 1, 2, 4, 8 or 24 hours. Following incubation the wells are washed and the extent of bound AA's measured by anti-huIgG immunodetection.

Example 16: Construction of an anti-EGFR IgG AA

Construction of an anti-EGFR IgG antibody

[00414] The C225 light chain variable region gene was synthesized by assembly PCR using oligos CX638-CX655 as in Bessette et al., Methods in Molecular Biology, vol. 231. The resulting product was

digested with BamHI/NotI and ligated to the large fragment of pXMal digested with BamHI/NotI to create plasmid pX-scFv225-Vk. Similarly, the C225 heavy chain variable region gene was synthesized by assembly PCR using oligos CX656-CX677, digested with BglII/NotI and ligated to pXMal BamHI/NotI to create plasmid pX-scFv225-Vh. The variable light chain gene was then cloned from pX-scFv225-Vk as a BamHI/NotI fragment into the pX-scFv225-Vh plasmid at BamHI/Not to create the plasmid pX-scFv225m-HL, containing the scFv gene based on C225.

[00415] The IL2 signal sequence was moved from pINFUSE-hIgG1-Fc2 (InvivoGen) as a Kasi/NcoI fragment to pFUSE2-CLIg-hk (InvivoGen) digested with Kasi/NcoI, resulting in plasmid pFIL2-CL-hk. The IL2 signal sequence was also moved from pINFUSE-hIgG1-Fc2 as a Kasi/EcoRI fragment to pFUSE-CHlg-hG1 (InvivoGen) digested with Kasi/EcoRI (large and medium fragments) in a three-way ligation, resulting in plasmid pFIL-CHlg-hG1.

[00416] The human IgG light chain constant region was site specifically mutated by amplification from plasmid pFIL2-CL-hk with oligos CX325/CX688, digestion with BsiWI/NheI, and cloning into pFIL2-CL-hk at BsiWI/NheI, resulting in plasmid pFIL2-CL₂₂₅.

[00417] The human IgG heavy chain constant region was site specifically mutated by amplification from plasmid pFIL-CHlg-hG1 in three segments with oligos CX325/CX689, CX690/CX692, and CX693/CX694, followed by overlap PCR of all three products using outside primers CX325/CX694. The resulting product was digested with EcoRI/AvrII and cloned into pINFUSE-hIgG1-Fc2 at EcoRI/NheI, resulting in plasmid pFIL-CH₂₂₅.

[00418] The variable light chain gene segment was amplified from pX-scFv225m-HL with oligos CX695/CX696, digested with BsaI, and cloned into pFIL2-CL₂₂₅ at EcoRI/BsiWI, resulting in the C225 light chain expression vector pFIL2-C225-light.

[00419] The variable heavy chain gene segment was amplified from pX-scFv225m-HL with oligos CX697/CX698, digested with BsaI, and cloned into pFIL-CH₂₂₅ at EcoRI/NheI, resulting in the C225 heavy chain expression vector pFIL-C225-heavy.

Table 33: Primers Used in the Construction of anti-EGFR IgG antibody

CX268	ccgcaggtacctgagcgctagccagtctggccag
CX325	tgcttgctcaactctacgtc
CX370	aactgtttattgcagctt
CX448	gagttttgtcggatccaccagagccaccgctgccaccgctcgagcc
CX638	gcgtatgcaggatccggcgccgatattctgctgaccaga
CX639	cacgctcagaatcaccgggctctgggtcagcagaatatcg
CX640	gcccgggtgattctgagcgtgagcccgggcgaacgtgtgag
CX641	ggctcgcgcggcagctaaagctcacacgttcgccgggct
CX642	cttagctgccgcgcgagccagagcattggcaccaacatt
CX643	gtgcgctgctgataccaatgaatgttggtgccaatgctct
CX644	cattggtatcagcagcgcaccaacggcagcccgcgcctgc
CX645	ttcgtcgcataattaatcagcagggcgggctgccgttg
CX646	tgattaaatatgcgagcgaagcattagcggcattccgag
CX647	tgccgctgccgctaaagcggtcgggaatgccgctaattgt

CX648	ccgctttagcggcagcggcagcggcaccgattttaccctg
CX649	ctttccacgctgttaatgctcagggtaaaatcgggtgccgc
CX650	agcattaacagcgtggaaagcgaagatattgcggattatt
CX651	gttggtgttctgctggcaataataatccgcaatatcttcg
CX652	attgccagcagaacaacaactggccgaccacctttggcgc
CX653	tcagttccagtttggtgcccgcgccaaggtggtcggcca
CX654	gggcaccaaactggaactgaaacgcggccgccatcaccat
CX655	ctcccacgcgtatggtgatgatggtgatggcggccgcgtt
CX656	cgtatgcaagatctggtagcggtagccaggtgcagctgaa
CX657	ccaggcccggggcgcgtctgtttcagctgcacctgggtacc
CX658	acagagcggccccgggcctggtgcagccgagccagagcctg
CX659	ctcacggtgcaggtaatgctcaggctctggctcggctgca
CX660	agcattacctgcaccgtgagcggctttagcctgaccaact
CX661	gcgcacccaatgcacgccatagttggtcaggctaaagccg
CX662	atggcgtgcattgggtgcgccagagccccgggcaaaggcct
CX663	aaatcacgcccagccattccaggcctttggccgggctctg
CX664	ggaatggctgggcgtgatttgagcggcggcaaacaccgat
CX665	ctggtaaacggggtgttataatcgggtgttgcggccgctcc
CX666	tataacaccccgtttaccagccgctgagcattaacaaag
CX667	cacctggcttttgcgttatctttgttaatgctcaggcgg
CX668	ataacagcaaaagccagggtgtttttaaaatgaacagcct
CX669	tcgcggtatcgttgctttgcaggctgttcattttaaaaaa
CX670	gcaaagcaacgataccgcgattttattattgcgcgcgcgcg
CX671	tcataatcataataggtcagcgcgcgcgcgcgaataataaa
CX672	ctgacctattatgattatgaatttgcgtattggggccagg
CX673	gctcacggtcaccagggtgccctggccccaatacgc aaat
CX674	gcacctgggtgaccgtgagcgcgggtggtagcggtagcgg
CX675	taccggcgcctccagatcctccgctaccgctaccacccgc
CX676	aggatctggaggcggcggtagtagtggtggaggatccggt
CX677	tggtagtgccggccgcggccaccggatcctccaccactac
CX688	cgaagtagctccctctacgctccccctgttgaagctcttg
CX690	acaagegcgttgagcccaaatcttg
CX692	cagttcatcccgggatgggggcagggtg
CX693	ccccatcccgggatgaactgaccaagaaccagggtcagc
CX694	ctggccacctaggactcatttacc
CX695	gcactggtctcgaattcggatattctgctgaccagag
CX696	ggtgcggtctccgtacgtttcagttccagtttggtg
CX697	gcactggtctcgaattcgcagggtgcagctgaaacagag
CX698	gagacggtctcgtagccgcgctcaggtcaccag
CX730	tgcgtatgcaagatctggtagcggtagcattctgctgaccagag
CX731	actactaccgccgcctccagatcctccgctaccgctaccaccttcagttccagtttggtg
CX732	tctggaggcggcggtagtagtggtggaggctcaggcggccagggtgcagctgaaacagag
CX733	gatggtgatggcggccgcgcgcgtcacggtcaccag
CX735	tgtcggtaccaccgctaccgcccgcgtcacggtcaccag
CX740	tcacgaattcgaaggccagctggccagggtcgcgggtggcagcgggtggctctggtggatccggcgggtggca
CX741	tggtagatccggcgggtggcagcgggtgggtggctccggcggtagcggtagatctgacaaaactcacac
CX747	gateccccgtctccgccagtc aaaatgatgccggaaggcggtag
CX748	cgccttccggcatcattttgactggcggagacggg

Construction of expression vectors for anti-EGFR AAs

[00420] Plasmid pX-scFv225m-HL was PCR amplified in separate reactions with primers CX730/CX731 and CX732/CX733, and the resulting products were amplified by overlap PCR with outside primers

CX730/CX733, digested with BglII/NotI, and cloned into pXMal at BamHI/NotI, resulting in plasmid pX-scFv225m-LH.

[00421] Linker sequence was added to the N-terminal side of the human IgG Fc fragment gene by PCR amplification of pFUSE-hIgG-Fc2 in a reaction with overlapping forward primers CX740, CX741 and reverse primer CX370. The resulting product was digested with EcoRI/BglII, and the ~115bp fragment was cloned into pFUSE-hIgG-Fc2 at EcoRI/BglII. The resulting plasmid was digested with KpnI/BglII, and the large fragment was ligated to the KpnI/BamHI-digested PCR product of amplifying pX-scFv225m-LH with oligos CX736/CX735, resulting in plasmid pPHB3734.

[00422] The resulting plasmid was digested with SfiI/XhoI, and masking peptide 3690 was cloned in as an SfiI/XhoI fragment of pPHB3690, resulting in plasmid pPHB3783.

[00423] The protease substrate SM984 was added by digesting the resulting plasmid with BamHI/KpnI and ligating the product of annealing the phosphorylated oligos CX747/CX748, resulting in plasmid pPHB3822.

[00424] The tandem peptide mask was constructed by digesting the resulting plasmid with XhoI, dephosphorylating the 5' ends, and cloning in the XhoI-digested PCR product of amplifying pPHB3579 with primers CX268/CX448, resulting in plasmid pPHB3889.

[00425] The masking region, linker, substrate, and light chain variable region of pPHB3783, pPHB3822, and pPHB3889 were amplified by PCR with primers CX325/CX696, digested with EcoRI/BsiWI, and cloned into pFIL2-CL₂₂₅ at EcoRI/BsiWI, resulting in the AA light chain expression vectors pPHB4007, pPHB3902, and pPHB3913 respectively.

[00426] Affinity matured masking peptides were swapped into the AA light chain expression vectors by cloning as SfiI/XhoI fragments. Protease substrates were swapped in as BamHI/KpnI compatible fragments.

Expression and purification of the anti-EGFR antibody and AAs

[00427] 3 µg of pFIL-CH₂₂₅-HL and 3 µg pFIL2-CH₂₂₅-light were co-transfected into CHO-S cells (Invitrogen) using Lipofectamine 200 (Invitrogen) according to manufacturers protocol. Transfected cells were cultured in Freestyle CHO media (Invitrogen) and selected for resistance to zeocin and blasticidin. Individual clones were isolated by limiting dilution and selected for expression of human IgG capable of binding EGFR by ELISA. All antibodies and AAs are purified by Protein-A chromatography using standard techniques.

[00428] Likewise, 3 µg of each expression vector for AA light chains was co-transfected into CHO-S cells with 3 µg pFIL-CH₂₂₅-HL. Transfected cells were cultured in Freestyle CHO media (Invitrogen) and selected for resistance to zeocin and blasticidin. Individual clones were isolated by limiting dilution and selected for expression of human IgG capable of binding EGFR by ELISA.

Screening of the affinity matured anti-EGFR MM library.

[00429] An initial MACS round was performed with SA dynabeads and 1.4×10^8 cells from the ecpX3-755 library. Prior to magnetic selection the cells were incubated with 3 nM biotin labeled C225Mab. Magnetic selection resulted in the isolation of 6×10^6 cells. The first round of FACS sorting was performed on 2×10^7 cells labeled with 0.1 nM DyLight (fluor 530 nM)-C225Mab and resulted in isolation of 1.5×10^5 cells with positive binding. To apply increased selective pressure to the population, the second round of FACS was performed on cells labeled with 10 nM DyLight-C225Mab in the presence of 100 uM 3690 peptide (CISPRGC) at 37°C. To further increase the selection pressure the 3rd and 4th rounds were performed on cells labeled with 100 nM DyLight-C225Fab in the presence of 100 uM 3690 peptide (CISPRGC) at 37°C. The brightest 1% of the positive population were collected, representing binding that was not competed by 3690 peptide. On cell affinity measurements of individual clones isolated from the above screen revealed three peptides, 3954(CISPRGCPDGPYVM), 3957(CISPRGCEPGTYVPT) and 3958(CISPRGCPGQIWHPP) with affinities for C225 at least 100 fold greater than 3690 (CISPRGC). These three MMs were incorporated into anti-EGFR AAs. Figure 22 shows the process for affinity maturation of some of the EGFR MM's.

Affinity measurement for C225 MMs

[00430] On-cell affinity measurement of C225 Fab binding to MM's 3690, 3954 and 3957. Binding of the eCPX3.0 clones 3690, 3954 and 3957 were analyzed on FACS at 3 different concentrations of DyLight labeled anti-EGFR Fab. The binding curves are shown in Figure 23. MMs 3954 and 3957 displayed at least 100 fold higher affinity than 3690.

Target displacement assay for anti-EGFR AAs

[00431] EGFR was adsorbed to the wells of a 96 well micro-titer plate, washed and blocked with milk protein. 25 ml of culture media containing 2 nM anti-EGFR antibody or anti-EGFR AA's containing the MM's 3690, 3957, 3954 and 3960/3579 was added to the coated wells and incubated for 1, 2, 4, 8 or 24 hours. Following incubation the wells were washed and the extent of bound AA's measured by anti-huIgG immunodetection. Anti-EGFR AA binding was normalized to anti-EGFR antibody binding (100%) for direct comparison of the masking efficiency in the AA context. The extents of equilibrium binding as a percent of parental or unmodified antibody binding are shown in Table 34 and Figure 24. Whereas MMs 3954 and 3957 display the same affinity, 100 times higher than 3609, 3954 is at least 2 times more efficient at inhibiting target binding. The sequences of the C225 heavy and light chains, MMs, and AAs are provided in the tables following. Nucleotide and amino acid sequences provided in the tables following. Parentheses delineate the demarcations between the various sequence domains: (Linker)(MM)(Linker)(CM)(Linker)(AB).

Table 34: C225 TDA: Percent of parental antibody binding \pm SEM at each time point

Time (hours)	3690 AA	3954 AA	3975 AA	3690/3579 AA
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1	15.5 ± 4.2	4.4 ± 1.8	7.3 ± 2.0	3.6 ± 1.2
2	19.3 ± 6.0	6.0 ± 2.0	9.3 ± 2.8	2.1 ± 0.6
4	21.5 ± 5.0	7.6 ± 1.7	12.8 ± 2.3	3.3 ± 1.2
8	27.6 ± 7.4	9.7 ± 0.4	14.9 ± 0.03	3.0 ± 1.6
24	20.0 ± 9.1	13.4 ± 1.2	22.3 ± 2.6	2.8 ± 0.1

Table 35: C225 Heavy Chain

caggtgcagctgaaacagagcggccccgggcctggtgcagccgagccagagcctgagcattacctgcaccgtgagcggcttagcctgaccaactatg
gcgtgcattgggtgcgccagagccccgggcaaaggcctggaatggctgggcgtgatttgagcggcggcaacaccgattataacaccccgttaccag
ccgcctgagcattaacaaagataacagcaaaagccaggtgtttttaaaatgaacagcctgcaaagcaacgataccgcgattattattgcgcgcgcgcg
ctgacctattatgattatgaatttgctattggggccagggcaccctggtgacctgagcgcggctagcaccaagggccccatcggtcttccccctggcac
cctctccaagagcacctctgggggcacagcggccctgggctgcttggtcaaggactacttccccgaaccggtgacggtgtcgtggaactcaggcgc
cctgaccagcggcgtgcacaccttccccggctgtctacagtcctcaggactctactccctcagcagcgtggtgacctgacctccagcagcttgggcac
ccagacctacatctgcaacgtgaatcacaagcccagcaacaccaaggtggacaagcgcgttgagcccaaatcttgacaaaactcacacatgccac
cgtgcccagcacctgaactcctggggggaccgtcagtcttcttcttccccccaaaacccaaggacacctcatgatctcccggaccttgaggtcacat
gcgtggtggtgacgtgagccacgaagacctgaggtcaagtcaactggtacgtggacggcgtggaggtgcataatgccaaagacaaagccgcggg
aggagcagtacaacagcacgtacctgtggtcagcgtctcaccgtctgcaccaggactggctgaatggcaaggagtacaagtgaaggtctccaac
aaagccctcccagcccccatcgagaaaacctctccaaagccaaagggcagccccgagaaccacaggtgtacacctgcccccatcccgggatgaa
ctgaccaagaaccaggtcagcctgacctgcctggtcaaaggttctatcccagcgacatcgccgtggagtgggagagcaatgggcagccggagaaca
actacaagaccacgcctcccgtgctggactccgacggctccttcttctctacagcaagctcaccgtggacaagagcaggtggcagcaggggaacgtc
ttctcatgctcctgatgatgaggtctctgcacaaccactacacgcagaagagcctctcctgtctccgggtaaa

QVQLKQSGPGLVQPSQSL SITCTVSGFSLTNYGVHWVRQSPGKGLEWLGVIWSGGNTDYNTPFT
SRLSINKDNSKSQVFFKMNSLQSNDAIYYCARALTYDYEFAYWGQGLVTVSAASTKGPSVF
PLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSS
LGTQTYICNVNHKPSNTKVDKRVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPE
VTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEY
KCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSRDELTKNQVSLTCLVKGFYPSDIAVEWESN
GQPENNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK

Table 36: Sequence of 3690-SM984-C225 Light Chain

Caaggccagtctggccaggttcacattgtctcattctattaacatgggcgcgcctcatcgggctcgagcgggtggcagcgggtggctctggtggatccccgtctccgccagtcaaaat
gatgccggaaggcgttaccagatcttctgacctagagccccgggtgattctgagcgtgagccccgggcgaacgtgtgagcttagctgccgcgcgagc
cagagcattggcaccaacattcattggtatcagcagcgcaccaacggcagccccgcgcctgctgattaaatatgcgagcgaaagcattagcggcattccg
agccgcttagcggcagcggcagcggcaccgattttaccctgagcattaacagcgtggaaagcgaagatattgcggattattattgccagcagaacaac
aactggccgaccaccttggcgcgggcaccaaactggaactgaaacgtacgggtggctgcaccatctgtcttcatcttcccgccatctgatgagcagttga
aatctggaactgcctctgtgtgtgctgctgaataacttctatcccagagaggccaaagtacagtggaaaggtggataacgccctccaatcgggtaactcc
caggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgagaaacacaaagt
ctacgcctgcgaagtcacccatcagggcctgagctcgcccgtcacaagagcttcaacagggggagcg

Linker MM Linker CM Linker AB
(QGQSGQ)(CISPRGC)(GGSSGGSGGSGGS)(PSPPVKMMPE)(GG)(TQILLTQSPVILSVSPGERVSFS
CRASQSIGTNIHWYQQR TNGSPRL LIKYASESISGIPSRFSGSGSGTDFTLSINSVESEDIADYYCQQ
NNNWPTTFGAGTKLELKR TVAAPS VFIFPPSDEQLKSGTASVVCLLN NFYPREAKVQWKVDNAL
QSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGA)

Table 37: Sequence of 3579-NSUB-C225 Light Chain

caaggccagtctggccaggttcacattgtctcattctattaacatgggcgcgcctcatcgggctcgagcgggtggcagcgggtggctctggtggatccg
gcgggtggcagcgggtgggtgctccggcggtaccagatcttctgacctagagccccgggtgattctgagcgtgagccccgggcgaacgtgtgagcttagc
tgccgcgcgagccagagcattggcaccaacattcattggtatcagcagcgcaccaacggcagccccgcgcctgctgattaaatatgcgagcgaaagcat

tagcggcattccgagccgctttagcggcagcggcagcggcaccgattttaccctgagcattaacagcgtggaaagcgaagatattgcggattattattgc
cagcagaacaacaactggccgaccacctttggcgcgggcaccaaactggaactgaaacgtacgggtggtgcaccatctgttctcatcttcccgccatctg
atgagcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaaggtggataacgccctcaa
tcgggtaactcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgag
aaacacaaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgtcacaaagagcttcaacaggggagcg

Linker	MM	Linker	AB
(QQQSGQ)	(GSHCLIPINMGAPSC)	(GSSGGSGGSGGSGGGSGGGSGG)	(TQILLTQSPVILSVSPGER
VSFSCRASQSIGTNIHWYQ	QRTNGSPRLLIK	YASESISGIPSRFSGSGSGTDFTLSINSVESEDIADY	YCQQNNNWPTTFGAGTKLELKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKV
DNALQSGNSQESVTEQDSK	DSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVT	KSFNRGA)	

Table 38: Sequence of 3690-3579-SM984-C225 Light Chain

caaggccagtctggccagtgcattctgccccgtggtgtggaggctcgagcgttagccagtctggccagggttcacattgtctcattcctattaacatggg
cgcgccgtcatcggtctgagcgggtggcagcgggtggtctggtgatccccgtctccgccagtcaaatgatccggaaggcgggtaccagatcttgc
tgaccagagccccgtgattctgagcgtgagccccggcgaacgtgtgagctttagctgccgcgcgagccagagcattggcaccaacattcattggtatc
agcagcgcaccaacggcagccccgcgcctgctgattaaatgcgagcgaagcattagcggcattccgagccgctttagcggcagcggcagcggca
ccgattttaccctgagcattaacagcgtggaaagcgaagatattgcggattattattgccagcagaacaacaactggccgaccacctttggcgcgggcac
caaactggaactgaaacgtacgggtggctgcaccatctgttctcatcttcccgccatctgatgagcagttgaaatctggaactgcctctgttgtgtgcctgctg
aataacttctatcccagagaggccaaagtacagtggaaaggtggataacgccctccaatcgggtaactcccaggagagtgtcacagagcaggacagcaa
ggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgagaaacacaaagtctacgcctgcgaagtcacccatcagggcctg
agctcgcccgtcacaaagagcttcaacaggggagcg

Linker	MM	Linker	CM
(QQQSGQ)	(CISPRGCGGSSASQSGQGSHCLIPINMGAPSC)	(GSSGGSGGSGGSGG)	(PSPPVKMMPE)
Linker	AB		
(GG)	(TQILLTQSPVILSVSPGERVSFSCRASQSIGTNIHWYQ	QRTNGSPRLLIK	YASESISGIPSRFSG
SGSGTDFTLSINSVESEDIADY	YCQQNNNWPTTFGAGTKLELKRTVAAPSVFIFPPSDEQLKSGTA	SVVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSK	DSTYSLSSTLTLSKADYEKHKVYA
CEVTHQGLSSPVT	KSFNRGA)		

Table 39: Sequence of 3954-NSUB-C225 Light Chain

caaggccagtctggccagtgcattctacctcgtggtgtccggacggcccatagctcatgtacggctcgagcgggtggcagcgggtggtctggtgatcc
ggcgggtggcagcgggtggtggctccggcgggtaccagatcttctgaccagagccccgtgattctgagcgtgagccccggcgaacgtgtgagcttta
gctgccgcgcgagccagagcattggcaccaacattcattggtatcagcagcgcaccaacggcagccccgcgcctgctgattaaatgcgagcgaagc
attagcggcattccgagccgctttagcggcagcggcagcggcaccgattttaccctgagcattaacagcgtggaaagcgaagatattgcggattattattg
ccagcagaacaacaactggccgaccacctttggcgcgggcaccaaactggaactgaaacgtacgggtggtgcaccatctgttctcatcttcccgccatct
gatgagcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaaaggtggataacgccctcca
atcgggtaactcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacga
gaaacacaaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgtcacaaagagcttcaacaggggagcg

Linker	MM	Linker	AB
(QQQSGQ)	(CISPRGCPDGPYVMY)	(GSSGGSGGSGGSGGGSGGGSGG)	(TQILLTQSPVILSVSPGER
VSFSCRASQSIGTNIHWYQ	QRTNGSPRLLIK	YASESISGIPSRFSGSGSGTDFTLSINSVESEDIADY	YCQQNNNWPTTFGAGTKLELKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKV
DNALQSGNSQESVTEQDSK	DSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVT	KSFNRGA)	

Table 40: Sequence of 3957-NSUB-C225 Light Chain

caaggccagtctggccagtgcattctacctcgtggtgtgagcctggcacctatgttccaacaggctcgagcgggtggcagcgggtggtctggtgatcc
ggcgggtggcagcgggtggtggctccggcgggtaccagatcttctgaccagagccccgtgattctgagcgtgagccccggcgaacgtgtgagcttta
gctgccgcgcgagccagagcattggcaccaacattcattggtatcagcagcgcaccaacggcagccccgcgcctgctgattaaatgcgagcgaagc
attagcggcattccgagccgctttagcggcagcggcagcggcaccgattttaccctgagcattaacagcgtggaaagcgaagatattgcggattattattg

ccagcagaacaacaactggccgaccacctttggcgcgggcaccaaactggaactgaaacgtacgggtggctgcaccatctgtcttcatcttcccgccatct
gatgagcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaaggtggataacgccctcca
atcgggtaactcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacga
gaaacacaaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgtcacaaagagcttcaacaggggagcg

Linker	MM	Linker	AB
(QGQSGQ)	(CISPRGCEPGTYVPT)	(GSSGGSGGSGGSGGGSGGGSGG)	(TQILLTQSPVILSVSPGER
VSFSCRASQSIGTNIHWYQ	QRTNGSPRLLIK	YASESISGIPSRFSGSGSGTDFTLSINSVESEDIADY	
YCQQNNNWPTTFGAGTKLELKRTVA	APSVFIFPPSDEQLKSGTASVVCLLNNFYPBREAKVQWKV		
DNALQSGNSQESVTEQDSKDY	SLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGA)		

Table 41: Sequence of 3958-NSUB-C225 Light Chain

caaggccagtctggccagtgcattctacctcgtggtgtccgggcaaatgtgcatccacctggctcgagcgggtggcagcgggtggctctggtggatcc
ggcgggtggcagcgggtgggtggctccggcggtaccagatcttctgaccagagcccgggtgattctgagcgtgagcccgggcgaacgtgtgagcttta
gctgccgcgcgagccagagcattggcaccaacattcattggtatcagcagcgcaccaacggcagcccgcgctgctgattaaatgcgagcgaagc
attagcggcattccgagccgctttagcggcagcggcagcggcaccgattttaccctgagcattaacagcgtggaaagcgaagatattgcggattattattg
ccagcagaacaacaactggccgaccacctttggcgcgggcaccaaactggaactgaaacgtacgggtggctgcaccatctgtcttcatcttcccgccatct
gatgagcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaaggtggataacgccctcca
atcgggtaactcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacga
gaaacacaaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgtcacaaagagcttcaacaggggagcg

Linker	MM	Linker	AB
(QGQSGQ)	(CISPRGCPGQIWHPP)	(GSSGGSGGSGGSGGGSGGGSGG)	(TQILLTQSPVILSVSPGER
VSFSCRASQSIGTNIHWYQ	QRTNGSPRLLIK	YASESISGIPSRFSGSGSGTDFTLSINSVESEDIADY	
YCQQNNNWPTTFGAGTKLELKRTVA	APSVFIFPPSDEQLKSGTASVVCLLNNFYPBREAKVQWKV		
DNALQSGNSQESVTEQDSKDY	SLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGA)		

Table 42: Sequences of C225 MMs:

3690	CISPRGCG
3579	GSHCLIPINMGAPSC
3690-3579	CISPRGCGGSSASQSGQGSHCLIPINMGAPSC
3954	CISPRGCPDGPYVMY
3957	CISPRGCEPGTYVPT
3958	CISPRGCPGQIWHPP
4124	CNHHYFYTCGCISPRGCPG
4125	ADHVFWGSYGCISPRGCPG
4127	CHHVYWGHCISPRGCPG
4133	CPHFTTTSCGCISPRGCPG
4137	CNHHYHYCYGCISPRGCPG
4138	CPHVSGSCGCISPRGCPG
4140	CPYYTSLSYGCISPRGCPG
4141	CNHVYFGTCGCISPRGCPG
4143	CNHFTLTTCGCISPRGCPG
4148	CHHFTLTTCGCISPRGCPG
4157	YNPCATPMCCISPRGCPG

EGFR MM Consensus Sequences

[00432] The consensus sequences for the EGFR MMs are provided below. The 3690 MM consensus (CISPRGC) is one major consensus sequence.

Table 43: C225 EGFR MM Consensus Sequences

PHB4124 CNHHYFYTCGCISPRGCG
 PHB4137 CNHHYHYTCGCISPRGCG
 PHB4141 CNHIVYEGTCGCISPRGCG
 PHB4127 CHHIVYWGHCISPRGCG
 PHB4133 CPHFTTTSCGCISPRGCG
 PHB4143 CNHFTLTTCGCISPRGCG
 PHB4148 CHHFTLTTCGCISPRGCG
 PHB4140 CPY YTLSYCGCISPRGCG
 PHB4138 CPHVSEFGSCGCISPRGCG
 PHB4125 ADHVFWGSYGCISPRGCG
 PHB4157 YNPCATPMCCISPRGCG
 PHB4127 CHHIVYWGHCISPRGCG

EGFR consensus Sequences from the 2nd round of screening for higher affinity masks

C(N/P)H(H/V/F)(Y/T)(F/W/T/L)(Y/G/T/S)(T/S/Y/H)CGCISPRGCG

CISPRGCGQPIPSVK
 CISPRGCTQPYHYSR
 CISPRGCNAVSGLGS

Example 17: Selective Substrate/CM Discovery and Testing

[00433] The section below the process for selective substrate discovery and testing for a number of exemplary enzymes.

uPA selective substrate discovery

uPA-selective substrates were isolated from an 8eCLiPS bacterial library consisting of $\sim 10^8$ random 8-mer substrates expressed as N-terminal fusions on the surface of E. coli. Alternating rounds of positive and negative selections by FACS were used to enrich for substrates optimized for cleavage by uPA and resistant to cleavage by the off-target serine proteases klk5 and 7. The naive library was incubated with 8ug/ml uPA for 1 h at 37°C followed by labeling with SAPE(red) and yPET mona (green). Cleavage by uPA results in loss of the SAPE tag and allows for sorting of bacteria expressing uPA substrates (green only, positive selection) from bacteria expressing uncleaved peptides (red + green). uPA substrates were sorted by FACS and the enriched pool was amplified and then incubated with 5 ng/ml KLK5 and 7 for 1 h at 37°C, labeled with SAPE and yPET mona, and sorted for lack of cleavage by these off-target proteases (red+ green, negative selection). The pool was amplified and sorted with 4 additional alternating rounds of positive and negative FACS using decreasing concentrations of uPA (4ug/ml, 2ug/ml) and increasing concentrations of klk5 and 7 (5 ng/ml, 10ng/ml). Individual clones from the last 3 rounds of FACS were sequenced and grouped into several consensus (Table 44). Clones from each consensus were then analyzed individually for cleavage by a range of concentrations of uPA, klk5 and 7

and plasmin for specificity of cleavage by on versus off-target proteases in Table 44. Figure 25 shows that unlike the uPA control and substrate SM16, KK1203, 1204 and 1214 show resistance to cleavage by KLK5, KLK7 and Plasmin.

Table 44: uPA Consensus sequences

	(1)	1	9
kk1203(1)	T	T	G
kk1206(1)	S	A	P
kk1216(1)	T	A	R
Consensus(1)	T	A	R
	(1)	1	11
kk1204 (1)	---	L	S
kk1208 (1)	G	G	W
kk1211 (1)	---	H	T
kk1214 (1)	--	P	L
Consensus (1)		L	T
	(1)	1	10
kk1217 (1)	A	A	R
kk1219 (1)	--	R	G
kk1196 (1)	S	S	R
kk1201 (1)	--	R	G
Consensus (1)		R	G

Plasmin selective substrate discovery

[00434] Plasmin-selective substrates were isolated from a second generation plasmin 10eCLiPS bacterial library consisting of $\sim 10^8$ random 10-mer substrates expressed as N-terminal fusions on the surface of *E. coli* (ref). Alternating rounds of positive and negative selections by FACS were used to enrich for substrates optimized for cleavage by plasmin and resistant to cleavage by the off-target matrix metalloproteinases (represented by MMP-9) and serine proteases (represented by klk5 and klk7).

[00435] The second generation plasmin 10eCLiPS library was based on a consensus sequence identified in-house by selecting the naïve 8eCLiPS for rapidly cleaved plasmin substrates using concentrations as low as 30pM plasmin for selection. Individual residues within the 10mer were either random ($n=20$), restricted ($1 < n < 20$) or fixed ($n=1$) to bias the peptide toward the consensus sequence while allowing flexibility to down-select away from unfavorable off-target sequences.

The second generation plasmin 10eCLiPS library was incubated with 300pM plasmin for 1 h at 37°C followed by labeling with SAPE(red) and yPET mona (green). Cleavage by plasmin results in loss of the SAPE tag and allows for sorting of bacteria expressing plasmin substrates (green only, positive selection) from bacteria expressing uncleaved peptides (red + green). plasmin substrates were sorted by FACS and the enriched pool was amplified and then incubated with 80U/ml MMP-9 2 h at 37°C, labeled with SAPE and yPET mona, and sorted for lack of cleavage by these off-target proteases (red+ green, negative selection). The pool was amplified and sorted with 4 additional alternating rounds of positive and

negative FACS using plasmin (round three at 100pM or 300pM, round five at 100pM or 300pM) and klk5 and 7 (Round four at 100ng/ml, round six at 200ng/ml). Individual clones from each the last 2 rounds of FACS were sequenced (Table 45). Clones from each consensus were then analyzed individually for cleavage by plasmin, MMP-9, klk5 and klk7 for specificity of cleavage by on versus off-target proteases. Representative data showing increased specificity towards Plasmin cleavage is shown in Figure 26. Figure 26 shows that unlike a non-optimized substrate, the optimized substrates Plas1237, Plas129 and Plas 1254 show resistance to cleavage by KLK5, KLK7.

Table 45: Peptide sequences derived from three rounds of Positive selection for Plasmin cleavage and negative selection for MMP9, KLK5 and KLK7

SM1191	EHPRVKVVSE
SM1197	PPDMLKLFPG
SM1200	PPVLKLLEW
SM1203	VLPELRSVFS
SM1206	APPSFKLVNA
SM1212	PPPEVRSFSV
SM1214	ALPSVKMVSE
SM1215	ETPSVKTMGR
SM1219	AIPRVRLFDV
SM1224	GLGTPRGLFA
SM1276	DRPKVKTMDF
SM1275	RVPKVKVMLD
SM1274	APPLVKSMVV
SM1272	REPFMKSLPW
SM1270	PVPRLKLIKD
SM1269	KGPKVKVVTL
SM1268	ERPGVKSLVL
SM1267	NZPRVRLVLP
SM1265	PRPFVKSVDQ
SM1263	RFPSLKSFPL
SM1261	ESPVMKSMAL
SM1260	VAPQLKSLVP
SM1255	APPLVKSMVV
SM1254	NMPSFKLVTG
SM1245	DRPEMKSLSG
SM1244	EQPEVKMVKG
SM1243	AVPKVRVVPE
SM1241	DLPLVKSLPS
SM1240	EAPKVKALPK
SM1239	GFPHMKTFQH
SM1238	YDPZVKVLA
SM1237	ASPTMKTVGL
SM1236	DVPPMKTLRP
SM1235	AFPDMRSVRS
SM1234	SAPYFRMMDM
SM1233	EKPRMKLFQG
SM1231	YVPRVKALEM

uPA Enzyme Activated AA Sequences

[00436] Nucleotide and amino acid sequences of uPA enzyme-activated anti VEGF light chain AAs are provided in the tables below. Parentheses delineate the demarcations between the various sequence domains: (Linker)(MM)(Linker)(CM)(Linker)(AB).

Table 46: PFIL2-CLIg-HK-anti-VegF 306 KK1203 LC

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttgctattatgggggcggttctggtggcagcggccaaggtggccaagg
tactggccgtggtccaagctgggtggcagtagcggcgggttctgatattcaactgacctagagcccttcttccctgagtgccagcgtgggtgacctgtta
cgatacattgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcacaaaaagtcctgatctacttcacaagttca
ctgcattccggcgtaccgtcgcgcttttagcgggttctggcagtggtaccgacttcacctgactatctcgagtetgcaacctgaggattttgctacatattactg
tcagcaatattcgacctgcccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacgggtggctgcaccatctgttcttcttcccgccatctg
atgagcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtgggaaggtggataacgccctccaa
tcgggtaactcccaggagagtggtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgag
aaacacaaaagtcctgcgaagtcacctatcagggcctgagctcgcccgctcacaagagcttcaacaggggagagtgttag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSEWQSMVQPRCYYG)	(GGSGGSGQGGQ)	(GTGRGPSWVGSS)	(GGS)	(DIQLTQSPSSL
SASVGDRVTITCSASQDISNYLNWYQQKPGKAPKVL					YFTSSLHSGVPSRFS
QPEDFATYYCQQYSTV					SGSGSGTDFTLT
AKVQWKVDNALQSGNSQESVTEQDSK					STLTLTKADYEKHKVYACEVTHQGLSSPVT
KSFNRGEC)					

Table 47: PFIL2-CLIg-HK-antiVegF 306 KK1204 LC

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttgctattatgggggcggttctggtggcagcggccaaggtggccaagg
tctgagcggccgttccgataatcatggcagtagcggcgggttctgatattcaactgacctagagcccttcttccctgagtgccagcgtgggtgacctgtta
cgatacattgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcacaaaaagtcctgatctacttcacaagttca
ctgcattccggcgtaccgtcgcgcttttagcgggttctggcagtggtaccgacttcacctgactatctcgagtetgcaacctgaggattttgctacatattactg
tcagcaatattcgacctgcccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacgggtggctgcaccatctgttcttcttcccgccatctg
atgagcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtgggaaggtggataacgccctccaa
tcgggtaactcccaggagagtggtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgag
aaacacaaaagtcctgcgaagtcacctatcagggcctgagctcgcccgctcacaagagcttcaacaggggagagtgttag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSEWQSMVQPRCYYG)	(GGSGGSGQGGQ)	(GLSGRSDNHGSS)	(GGS)	(DIQLTQSPSSL
SASVGDRVTITCSASQDISNYLNWYQQKPGKAPKVL					YFTSSLHSGVPSRFS
QPEDFATYYCQQYSTV					SGSGSGTDFTLT
AKVQWKVDNALQSGNSQESVTEQDSK					STLTLTKADYEKHKVYACEVTHQGLSSPVT
KSFNRGEC)					

Table 48: PFIL2-CLIg-HK-antiVegF 306 KK1214 LC

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttgctattatgggggcggttctggtggcagcggccaaggtggccaacc
actgactggtcgtagcgggtggtggaggaagtagcggcgggttctgatattcaactgacctagagcccttcttccctgagtgccagcgtgggtgacctgtt
acgatacattgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcacaaaaagtcctgatctacttcacaagttc
actgcattccggcgtaccgtcgcgcttttagcgggttctggcagtggtaccgacttcacctgactatctcgagtetgcaacctgaggattttgctacatattact
gtcagcaatattcgacctgcccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacgggtggctgcaccatctgttcttcttcccgccatctg
gatgagcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtgggaaggtggataacgccctcca
atcgggtaactcccaggagagtggtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacga
gaaacacaaaagtcctgcgaagtcacctatcagggcctgagctcgcccgctcacaagagcttcaacaggggagagtgttag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)(PCSEWQSMVQPRCYYG)(GGSGGSGQGGQ)(PLTGRSGGGSS)(GGS)(DIQLTQSPSSL SASVGDRVTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTISL QPEDFATYYCQQYSTVPWTFGQGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPRE AKVQWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVT KSFNRGEC)					

Table 49: PFIL2-CLIg-HK-antiVegF 306 SM1215 LC

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttctattatgggggcggttctggtggcagcggccaaggtggccaaga
aactccatctgtaaagactatgggccgtagtagcggcggttctgatattcaactgaccagagcccttctccctgagtgccagcgtgggtgaccgtgttac
gatcacttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcact
gcattccggcgtaccgtcgcgttttagcgggttctggcagtgggtaccgacttcaccctgactatctcagtgctgcaacctgaggattttgctacatattactgtc
agcaatattcgaccgtgccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacgggtggctgcaccatctgtcttcattctcccgccatctgat
gagcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtgggaaggtggataacgccctccaatc
gggtaactcccaggagagtgacagagcaggacagcaaggacagcacctacagcctcagcagcaccctgacgctgagcaaagcagactacgaga
aacacaaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgctcacaagagcttcaacaggggagagtgtag

Linker	MM	Linker	CM	Linker	AB
GQSGQ)(PCSEWQSMVQPRCYYG)(GGSGGSGQGGQ)(ETPSVKTMGRSS)(GGS)(DIQLTQSPSSLS ASVGDRVTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTISLQ PEDFATYYCQQYSTVPWTFGQGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREA KVQWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVT KSFNRGEC)					

Plasmin -activated AA Sequences

[00437] Nucleotide and amino acid sequences of plasmin enzyme-activated anti VEGF light chain AAs are provided in the tables below. Parentheses delineate the demarcations between the various sequence domains: (Linker)(MM)(Linker)(CM)(Linker)(AB).

Table 50: PFIL2-CLIg-HK-antiVegF 306 SM1239 LC

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttctattatgggggcggttctggtggcagcggccaaggtggccaagg
ttcccatatgaaaactttccagcatagtagcggcggttctgatattcaactgaccagagcccttctccctgagtgccagcgtgggtgaccgtgttacg
atcacttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtcctgatctacttcacaagttcactg
cattccggcgtaccgtcgcgttttagcgggttctggcagtgggtaccgacttcaccctgactatctcagtgctgcaacctgaggattttgctacatattactgtc
gcaatattcgaccgtgccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacgggtggctgcaccatctgtcttcattctcccgccatctgatg
agcagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtgggaaggtggataacgccctccaatc
ggtaactcccaggagagtgacagagcaggacagcaaggacagcacctacagcctcagcagcaccctgacgctgagcaaagcagactacgagaa
acacaaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgctcacaagagcttcaacaggggagagtgtag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)(PCSEWQSMVQPRCYYG)(GGSGGSGQGGQ)(GFPHMKTFQHSS)(GGS)(DIQLTQSPSSL SASVGDRVTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTISL QPEDFATYYCQQYSTVPWTFGQGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPRE AKVQWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVT KSFNRGEC)					

Legumain-activated AAs

[00438] The sequences for the legumain substrates AANL and PTNL are known in the art (Liu, et al. 2003. Cancer Research 63, 2957-2964; Mathieu, et al 2002. Molecular and Biochemical Parasitology 121, 99-105). Nucleotide and amino acid sequences of legumain enzyme-activated anti VEGF light chain AAs are provided in the tables below. Parentheses delineate the demarcations between the various sequence domains: (Linker)(MM)(Linker)(CM)(Linker)(AB).

Table 51: PFIL2-CLIg-HK-antiVEGF 306 AANL Light Chain

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttgctattatgggggcggttctggtggcagcggccaaggtggccaagc
agctaactctgggcagcggaggaagtagcggcggttctgatattcaactgaccagagcccttcttccctgagtgccagcgtgggtgaccgtgttacgac
acttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtctgatctacttcacaagttcactgcat
tccggcgtaccgtcgcgctttagcgggttctggcagtggtaccgacttcacctgactatctcagagtctgcaacctgaggattttgctacatattactgtcagc
aatattcgaccgtgccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacggtggctgcaccatctgttctcatcttcccgccatctgatgag
cagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaggtggataacgccctccaatcggg
taactcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcaccctgacgctgagcaaagcagactacgagaaaca
caaagtctacgcctgcgaagtcacccatcagggcctgagctcgcctgcacaaagagcttcaacaggggagagtgttag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSEWQSMVQPRCYYG)	(GGSGGSGQGGQ)	(AANLGSGGSS)	(GGS)	(DIQLTQSPSSLSA
SVGDRVITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTIISSLQP					
EDFATYYCQQYSTVPWTFGQGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAK					
VQWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSF					
NRGEC)					

Table 52: PFIL2-CLIg-HK- antiVEGF 306 PTNL Light Chain

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttgctattatgggggcggttctggtggcagcggccaaggtggccaacc
gactaatctgggcagcggaggaagtagcggcggttctgatattcaactgaccagagcccttcttccctgagtgccagcgtgggtgaccgtgttacgac
acttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtctgatctacttcacaagttcactgcat
tccggcgtaccgtcgcgctttagcgggttctggcagtggtaccgacttcacctgactatctcagagtctgcaacctgaggattttgctacatattactgtcagc
aatattcgaccgtgccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacggtggctgcaccatctgttctcatcttcccgccatctgatgag
cagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaggtggataacgccctccaatcggg
taactcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcaccctgacgctgagcaaagcagactacgagaaaca
caaagtctacgcctgcgaagtcacccatcagggcctgagctcgcctgcacaaagagcttcaacaggggagagtgttag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSEWQSMVQPRCYYG)	(GGSGGSGQGGQ)	(PTNLGSGGSS)	(GGS)	(DIQLTQSPSSLSAS
VGDRVITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTIISSLQPE					
DFATYYCQQYSTVPWTFGQGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKV					
QWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFN					
RGEC)					

Table 53: PFIL2-CLIg-HK- antiVEGF 306 PTN Light Chain

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttgctattatgggggcggttctggtggcagcggccaaggtggccaacc
gactaatggtggcagcggaggaagtagcggcggttctgatattcaactgaccagagcccttcttccctgagtgccagcgtgggtgaccgtgttacgac
acttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcaccaaaagtctgatctacttcacaagttcactgcat
tccggcgtaccgtcgcgctttagcgggttctggcagtggtaccgacttcacctgactatctcagagtctgcaacctgaggattttgctacatattactgtcagc
aatattcgaccgtgccgtggacgttcgggcagggcaccaaaagtgagattaagcgtacggtggctgcaccatctgttctcatcttcccgccatctgatgag
cagttgaaatctggaactgcctctgttgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtggaggtggataacgccctccaatcggg

taactcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgagaaaca
caaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgtcacaagagcttcaacaggggagagtgttag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSEWQSMVQPRCYYG)	(GGSGGSGQGGQ)	(PTNGGSGGSS)	(GGS)	(DIQLTQSPSSLSA
SVGDRVTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTISLQP					
EDFATYYCQQYSTVPWTFGQGKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAK					
VQWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSF					
NRGEC)					

Caspase activated AAs

[00439] Nucleotide and amino acid sequences of caspase enzyme-activated anti VEGF light chain AAs are provided in the tables below. Parentheses delineate the demarcations between the various sequence domains: (Linker)(MM)(Linker)(CM)(Linker)(AB). The caspase substrate, sequence DEVD, is known in the art.

Table 54: PFIL2-CLIg-HK-antiVegF 306 DEVD LC

ggccagtctggccagccgtgttctgagtggcagtcgatggtgcagccgcgttctattatgggggcggttctggtggcagcggccaaggtggccaaga
cgaagtcgatggcagcggaggaagtagcggcggttctgatattcaactgaccagagcccttctccctgagtgccagcgtgggtgaccgtgttacgatc
acttgctcggccagccaagatatttctaactacctgaattggtaccagcagaagccaggaaaggcacaaaaagtcctgatctacttcacaagttcactgcat
tccggcgtaccgtcgcgctttagcggttctggcagtggtaccgacttcacctgactatctcagctctgcaacctgaggattttgctacatattactgtcagc
aatattcgaccgtgccgtggacgttcgggcagggcaccaaagtggagattaagcgtacggtggctgcaccatctgttctcatcttcccgccatctgatgag
cagttgaaatctggaactgcctctgtgtgtgcctgctgaataacttctatcccagagaggccaaagtacagtgggaaggtggataacgccctccaatcggg
taactcccaggagagtgtcacagagcaggacagcaaggacagcacctacagcctcagcagcacctgacgctgagcaaagcagactacgagaaaca
caaagtctacgcctgcgaagtcacccatcagggcctgagctcgcccgtcacaagagcttcaacaggggagagtgttag

Linker	MM	Linker	CM	Linker	AB
(GQSGQ)	(PCSEWQSMVQPRCYYG)	(GGSGGSGQGGQ)	(DEVDGSGGSS)	(GGS)	(DIQLTQSPSSLSA
SVGDRVTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTISLQP					
EDFATYYCQQYSTVPWTFGQGKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAK					
VQWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSF					
NRGEC)					

Construction of Legumain and Caspase Activated AA expression Vectors

[00440] Substrates were constructed in a two step process. First, two products were PCR amplified using the CX0325 forward primer with a substrate specific reverse primer (CX0720 AANL, CX0722 PTNL, CX0724 PTN, and CX0758 DEVD), the other PCR amplified using the CX0564 reverse primer with a substrate specific forward primer (CX0721 AANL, CX0723 PTNL, CX0725 PTN, and CX0754 DEVD). In both cases the substrate for the PCR was the anti-VEGF mmp-9 306 scFv. Second, the two products were combined and PCR amplified using the outside primers CX0325 and CX0564. The final products were cloned into the pFIL2-CL-anti-VEGF Lc using the EcoRI and XhoI restriction sites.

Table 55: Primers for Construction of Legumain and Caspase Activated AA expression Vectors
CXO564 aggttgcagactcgagatagtcagggtgaagtc

CX0720	tcctccgctgcccagattagctgcttggccaccttggccgctgccac
CX0721	gcagctaactctgggcagcggaggaagtagcggcggttctgatattcaactg
CX0722	tcctccgctgcccagattagctggttggccaccttggccgctgccac
CX0723	ccgactaatctgggcagcggaggaagtagcggcggttctgatattcaactg
CX0724	tcctccgctgccaccattagctggttggccaccttggccgctgccac
CX0725	ccgactaatggtggcagcggaggaagtagcggcggttctgatattcaactg
CX0754	gacgaagtcgatggcagcggaggaagtagcggcggttctgatattcaactg
CX0758	tcctccgctgccatcgacttcgttcttggccaccttggccgctgccac

Expression and purification of legumain activated AAs

[00441] 3 µg of pFIL-VEGF-HL and 3 µg pFIL2-306-substrate-VEGF-light were co-transfected into CHO-S cells (Invitrogen) using Lipofectamine 200 (Invitrogen) according to manufacturers protocol. Transfected cells were cultured in Freestyle CHO media (Invitrogen) and selected for resistance to zeocin and blasticidin. Individual clones were isolated by limiting dilution and selected for expression of human IgG capable of binding EGFR by ELISA. All antibodies and AAs are purified by Protein-A chromatography using standard techniques.

Assay description for the scFv AA digest

[00442] ScFv AAs were diluted to 200nM in assay buffer and combined with rhLegumain diluted in assay buffer at 2ug/ml. Digests were incubated overnight at room temperature. IgG AAs were diluted to 200nM in assay buffer and combined with rhLegumain diluted in assay buffer at concentrations from 2-40 mg/mL (final rhLegumain concentrations 1ug/ml, 5ug/ml, 20ug/ml. Digests were incubated overnight at 37°C. Following digestion, the extent of activation was measured by the extent of AA binding to VEGF on ELISA plates, visualized with anti-human-Fc. Figure 27 Panel A shows activation of ScFv AAs containing legumain substrates AANL and PTNL following treatment with 5 mg/mL legumain. Panel B shows activation of an anti-VEGF IgG AA containing the legumain substrate PNTL.

in vivo Stability of Legumain Activated AAs

Four 12 week old Balb/C mice were each given a single bolus injection of 100 µg of a plasmin activated AA, AA^{PLAS}VEGF, or one of the legumain activated AAs, AA^{AANL}VEGF or AA^{PTNL}VEGF. At 15 minutes, 1 day, 3 days, and 7 days following injection, serum was collected. Total AA concentration was calculated from ELISA measurement of total human Fc in the serum. The concentration of activated antibody was calculated from a human VEGF binding ELISA measurement and is shown in Figure 28. Legumain activated AAs isolated from serum up to 7 days following injection remain masked. (n=4). The ratio of activated AA to total AA at each time point is shown in Figure 28 as the average of measurements from individual animals and is expressed as per cent activated. While the plasmin activated AA is nearly completely activated at 7 days both legumain activated AAs are only minimally activated.

Example 18: Serum Half Lives of AAs

[00446] Figure 32 shows that AAs persist at higher concentrations in a multidose study: Tumor-bearing Balb/c nu/nu mice were injected with 5 mg/kg of parental VEGF scFv-Fc, AA1, 2 or 3 every 3 days. Serum was collected at the indicated times and the concentration of AA or parent scFv-Fc was measured by ELISA. All three AAs maintained significantly higher serum concentrations than the parent throughout the study.

Amino Acid Sequences of VEGF scFv-Fcs AAs

Table 56: The amino acid sequence of Anti-VEGF scFv-Fc from which AA scFvs were derived

DIQLTQSPSSLSASVGDRVTTITCSASQDISNYLNWYQQKPGKAPKVLIIYFTSSSLHSGVPSRFSGSGS
GTDFTLTISLQPEDFATYYCQQYSTVPWTFGQGTKVEIKGGGSGGGSGGGSGGGSGGGSGGGSGG
GGSGEVQLVESGGGLVQPGGSLRLSCAASGYDFTHYGMNWVRQAPGKGLEWVGWINTYTGEPT
TYAADFKRRFTFSLDTSKSTAYLQMNSLRAEDTAVYYCAKYPYYGTSHWYFDVWGQGTLLVT
VSGGSGAMVRSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVK
FNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTIS
KAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD
GSFFLYSKLTVDKSRWOOQGNVVFSCSVMHEGLHNHYTQKSLSLSPGK

Table 57: AA^{MMP}VEGF: AAs contain a masking peptide and MMP substrate attached by a short linker as shown

masking peptide **substrate**
GQSGQPCSEWQSMVQPRCYYGGGSGGSGQGGQVHMPLGFLGPGGSDIQLTQSPSSLSASVGDR
 VTITCS
 ASQDISNYLNWYQQKPGKAPKVLIIYFTSSLHSGVPSRFSGSGSGTDFTLTISSLQPEDFATYYCQQ
 YSTVPWTFGOGTKVEIKGGGSGGGSGGGSGGGGSGGGGSGGGGSGGEVOLVESGGGLVOPGGSL

107

[00448] Greater than 80% of the patients typically administered a conventional EGFR antibody therapeutic exhibit toxicity of the skin, the largest organ of the body. When patients are administered AAs directed against EGFR it is expected that there will be little or no toxicity of the skin, as the AA will not be activated in the skin, due to lack of disease specific CM. As such, it is expected that the anti-EGFR AB of the AA will not be able to specifically bind the EGFR target. Additionally it is expected that in such patients, because the AA will not be active in the skin, the AA will not be sequestered and it is expected that the serum levels of the AA will remain high, thereby increasing the concentration of the AA in the diseased tissue, effectively raising the effective dose. Hydrolysis of the CM in the diseased tissue based on the disease environment will lead to an activated AA allowing for unmasking and specific binding of the AB to the EGFR target, and will lead to the desired therapeutic effect.

[00449] While the present invention has been described with reference to the specific embodiments thereof, it should be understood by those skilled in the art that various changes may be made and equivalents may be substituted without departing from the true spirit and scope of the invention. In addition, many modifications may be made to adapt a particular situation, material, composition of matter, process, process step or steps, to the objective, spirit and scope of the present invention. All such modifications are intended to be within the scope of the claims appended hereto.

CLAIMS

WHAT IS CLAIMED IS:

1. A modified antibody comprising: an antibody or antibody fragment (AB), capable of specifically binding its target, coupled to a masking moiety (MM), wherein the coupling of the MM reduces the ability of the AB to bind its target such that the dissociation constant (K_d) of the AB coupled to the MM towards the target is at least 100 times greater than the K_d of the AB not coupled to the MM towards the target.
2. The modified antibody of claim 1 wherein the K_d of the AB coupled to the MM towards the target is at least 1000 times greater than the K_d of the AB not coupled to the MM towards the target.
3. The modified antibody of claim 1 wherein the K_d of the AB coupled to the MM towards the target is at least 10,000 times greater than the K_d of the AB not coupled to the MM towards the target.
4. The modified antibody of claim 1 wherein in the presence of target, the coupling of the MM to the AB reduces the ability of the AB to bind its target by at least 90%, as compared to the ability of the AB not coupled to the MM to bind its target, when assayed *in vitro* using a target displacement assay.
5. The modified antibody of claim 4 wherein the coupling of the MM to the AB reduces the ability of the AB to bind its target for at least 12 hours.
6. The modified antibody of claim 4 wherein the coupling of the MM to the AB reduces the ability of the AB to bind its target for at least 24 hours.
7. The modified antibody of claim 4 wherein the coupling of the MM to the AB reduces the ability of the AB to bind its target for at least 72 hours.
8. The modified antibody of claim 1 wherein the dissociation constant (K_d) of the MM towards the AB is at least 100 times greater than the K_d of the AB towards the target.
9. The modified antibody of claim 8 wherein the K_d of the MM towards the AB is lower than 10nM.
10. The modified antibody of claim 8 wherein the K_d of the MM towards the AB is lower than 5nM.
11. The modified antibody of claim 8 wherein the K_d of the MM towards the AB is about 1nM.
12. The modified antibody of claim 1 further coupled to a cleavable moiety (CM).
13. The modified antibody of claim 12 wherein the CM is capable of being cleaved by an enzyme.
14. The modified antibody of claim 12 wherein the CM is capable of being reduced by a reducing agent.
15. The modified antibody of claim 12 wherein the CM is capable of being photolysed.

16. The modified antibody of claim 1 wherein the MM is capable of specifically binding to the antigen-binding domain of the AB.

17. The modified antibody of claim 16 wherein the binding of the MM to the antigen-binding domain is non-covalent.

18. The modified antibody of claim 1 wherein the MM reduces the AB's ability to bind its target allosterically.

19. The modified antibody of claim 1 wherein the MM reduces the AB's ability to bind its target sterically.

20. The modified antibody of claim 12 wherein the CM is capable of being specifically cleaved by an enzyme at a rate of about at least $1 \times 10^4 \text{ M}^{-1}\text{S}^{-1}$.

21. The AB of claim 20 wherein the CM is capable of being specifically cleaved by an enzyme at a rate of at least $5 \times 10^4 \text{ M}^{-1}\text{S}$.

22. The modified antibody of claim 20 wherein the CM is capable of being specifically cleaved by an enzyme at a rate of at least $10 \times 10^4 \text{ M}^{-1}\text{S}$.

23. The modified antibody of claim 1 wherein the antibody fragment is selected from the group consisting of a Fab' fragment, a F(ab')₂ fragment, a scFv, a scAB a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.

24. The modified antibody of claim 1 wherein the AB is selected from the group consisting of the antibodies in Table 2.

25. The modified antibody of claim 24 wherein the AB is not alemtuzumab.

26. The modified antibody of claim 24 wherein the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

27. The modified antibody of claim 1 wherein the target is selected from the group consisting of the targets in Table 1.

28. The modified antibody of claim 1 wherein the target is not CD52.

29. The modified antibody of claim 27 wherein the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

30. The modified antibody of claim 1 further comprising a second AB wherein the target for the second AB is selected from the group consisting of the targets in Table 1.

31. The modified antibody of claim 12 wherein the CM is located within the MM.

32. The modified antibody of claim 12 wherein the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3.

33. The modified antibody of claim 32 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

34. The modified antibody of claim 32 wherein the AB is selected from the group consisting of the antibodies in Table 2.

35. The modified antibody of claim 32 wherein the AB is not alemtuzumab.

36. The modified antibody of claim 34 wherein the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

37. The modified antibody of claim 32 wherein the target is selected from the group consisting of the targets in Table 1.

38. The modified antibody of claim 37 wherein the target is not CD52.

39. The modified antibody of claim 37 wherein the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

40. The modified antibody of claim 12 further coupled to a second cleavable moiety (CM), capable of being specifically modified by an enzyme.

41. The modified antibody of claim 40 wherein the second CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA

42. The modified antibody of claim 1 wherein the MM does not comprise more than 50% amino acid sequence similarity to a natural binding partner of the AB.

43. The modified antibody of claim 1 further comprising a linker peptide, wherein the linker peptide is positioned between the AB and the MM.

44. The modified antibody of claim 12 further comprising a linker peptide, wherein the linker peptide is positioned between the MM and the CM.

45. The modified antibody of claim 12 further comprising a linker peptide, wherein the linker peptide is positioned between the AB and the CM.

46. The modified antibody of claim 12 further comprising two linker peptides, wherein the first linker peptide is between the AB and the CM and the second linker peptide is positioned between the MM and the CM.

47. The modified antibody of claim 43 wherein the linker is selected from the group consisting of a cleavable linker, a non-cleavable linker, and a branched linker.

48. The modified antibody of claim 44 wherein the linker is selected from the group consisting of a cleavable linker, a non-cleavable linker, and a branched linker.

49. The modified antibody of claim 45 wherein the linker is selected from the group consisting of a cleavable linker, a non-cleavable linker, and a branched linker.

50. The modified antibody of claim 46 wherein the linkers are selected from the group consisting of a cleavable linker, a non-cleavable linker, and a branched linker.

51. The modified antibody of claim 1 further comprising a detectable moiety.

52. The modified antibody of claim 47 further comprising a cleavable moiety (CM) coupled to the AB, capable of being specifically modified by an enzyme.

53. The modified antibody of claim 47 wherein the detectable moiety is a diagnostic agent.

54. The modified antibody of claim 1 further comprising an agent conjugated to the AB.

55. The modified antibody of claim 54 wherein the agent is a therapeutic agent.

56. The modified antibody of claim 54 further comprising a cleavable moiety (CM) coupled to the AB, capable of being specifically cleaved.

57. The modified antibody of claim 56 wherein the agent is an antineoplastic agent.

58. The modified antibody of claim 56 wherein the antibody or antibody fragment is selected from the group consisting of the antibodies in Table 2.

59. The modified antibody of claim 56 wherein the AB is not alemtuzumab.

60. The modified antibody of claim 58 wherein the antibody or antibody fragment is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

61. The modified antibody of claim 56 wherein the target is selected from the group consisting of the targets in Table 1.

62. The modified antibody of claim 56 wherein the target is not CD52.

63. The modified antibody of claim 61 wherein the target is target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

64. The modified antibody of claim 56 wherein the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3.

65. The modified antibody of claim 64 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

66. The modified antibody of claim 56 wherein the CM is capable of being cleaved by photolysis.

67. The modified antibody of claim 54 wherein the agent is conjugated to a carbohydrate moiety of the AB.

68. The modified antibody of claim 67 wherein the carbohydrate moiety is located outside the antigen-binding region of the AB.

69. The modified antibody of claim 54 wherein the agent is conjugated to a sulfhydryl group of the AB.

70. The modified antibody of claim 12 wherein the serum half-life of the composition is at least 5 days when administered to an organism.

71. The modified antibody of claim 1 wherein the consensus sequence of the MM is CISPRGC, C(N/P)H(HVF)(Y/T)(F/W/T/L)(Y/G/T/S)(T/S/Y/H)CGCISPRGCG, xCxxYQCLxxxxxx,XXQPxPPRVXX, PxPGFPYCxxxx, xxxxQxxPWPP, GxGxCYTILExxCxxR, GxxxCYxIxExxCxxxx, GxxxCYxIxExWCxxxx, xxxCCxxYxIxxCCxxx, or xxxxxYxILExxxxx.

72. The modified antibody of claim 57 wherein the consensus sequence is specific for binding to an anti-VEGF antibody, an anti-EGFR antibody, or an anti-CTLA-4 antibody.

73. A modified antibody comprising an antibody or antibody fragment (AB), capable of specifically binding its target, coupled to a masking moiety (MM), wherein the coupling of the MM to the AB reduces the ability of the AB to bind the target by at least 90%, as compared to the ability of the AB not coupled to the MM to bind the target, when assayed *in vitro* using a target displacement assay.

74. The modified antibody of claim 73 wherein the binding of the AB to the target is reduced for at least 12 hours.

75. The modified antibody of claim 73 wherein the binding of the AB to the target is reduced for at least 24 hours.

76. The modified antibody of claim 73 wherein the binding of the AB to the target is reduced for at least 72 hours.

77. The modified antibody of claim 73 wherein the dissociation constant (K_d) of the AB coupled to the MM towards the target is at least 100 times greater than the K_d of the AB not coupled to the MM towards the target.

78. The modified antibody of claim 73 wherein the dissociation constant (K_d) of the MM towards the AB is at least 100 times greater than the K_d of the AB towards the target.

79. The modified antibody of claim 78 wherein the K_d of the MM towards the AB is lower than 10nM.

80. The modified antibody of claim 78 wherein the K_d of the MM towards the AB is lower than 5nM.

81. The modified antibody of claim 78 wherein the K_d of the MM towards the AB is about 1nM.

82. The modified antibody of claim 73 further comprising a linker between the AB and the MM.

83. The modified antibody of claim 82 wherein the linker comprises a cleavable moiety (CM).

84. The modified antibody of claim 73 wherein the MM is capable of specifically binding to the antigen-binding domain of the AB.

85. The modified antibody of claim 83 wherein the CM is capable of being specifically cleaved by an enzyme at a rate of at least $1 \times 10^4 \text{ M}^{-1} \text{ S}^{-1}$.

86. The modified antibody of claim 83 wherein the CM is capable of being specifically cleaved by an enzyme at a rate of at least $5 \times 10^4 \text{ M}^{-1} \text{ S}^{-1}$.

87. The modified antibody of claim 83 wherein the CM is capable of being specifically cleaved by an enzyme at a rate of at least $10 \times 10^4 \text{ M}^{-1} \text{ S}^{-1}$.

88. The modified antibody of claim 83 wherein the CM is capable of being cleaved by a reducing agent.

89. The modified antibody of claim 83 wherein the CM is capable of being cleaved by photolysis.

90. The modified antibody of claim 73 wherein the antibody fragment is selected from the group consisting of: a Fab' fragment, a F(ab')_2 fragment, a scFv, a scAB, a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.

91. The modified antibody of claim 73 wherein the AB is selected from the group consisting of the antibodies in Table 2.

92. The modified antibody of claim 91 wherein the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

93. The modified antibody of claim 73 wherein the target is selected from the group consisting of the targets in Table 1.

94. The modified antibody of claim 93 wherein the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

95. The modified antibody of claim 73 wherein the AB is not alemtuzumab.

96. The modified antibody of claim 73 wherein the target is not CD52.

97. The modified antibody of claim 83 wherein the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3.

98. The modified antibody of claim 97 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

99. The modified antibody of claim 83 further coupled to a second cleavable moiety (CM), capable of being specifically modified by an enzyme.

100. The modified antibody of claim 99 wherein the second CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA

101. The modified antibody of claim 73 wherein the MM does not comprise more than 50% amino acid sequence similarity to a natural binding partner of the AB.

102. The modified antibody of claim 73 further comprising a detectable moiety.

103. The modified antibody of claim 73 further comprising an agent conjugated to the AB.

104. An activatable antibody (AA) comprising:

- (a) an antibody or antibody fragment (AB), capable of specifically binding its target;
- (b) a masking moiety (MM) coupled to the AB, capable of inhibiting the specific binding of the AB to its target; and
- (c) a cleavable moiety (CM) coupled to the AB, capable of being specifically cleaved by an enzyme;

wherein when the AA is not in the presence of sufficient enzyme activity to cleave the CM, the MM reduces the specific binding of the AB to its target by at least 90% when compared to when the AA is in the presence of sufficient enzyme activity to cleave the CM and the MM does not inhibit the specific binding of the AB to its target.

105. The AA of claim 104 wherein the binding of the AB to its target is reduced for at least 12 hours.

106. The AA of claim 104 wherein the binding of the AB to its target is reduced for at least 24 hours.

107. The AA of claim 104 wherein the binding of the AB to its target is reduced for at least 72 hours.

108. The AA of claim 104 wherein the dissociation constant (K_d) of the AB coupled to the MM and CM towards the target is at least 100 times greater than the K_d of the AB not coupled to the MM and CM towards the target.

109. The AA of claim 104 wherein the dissociation constant (K_d) of the MM towards the AB is at least 100 times greater than the K_d of the AB towards the target.

110. The modified antibody of claim 109 wherein the K_d of the MM towards the AB is lower than 10nM.

111. The modified antibody of claim 109 wherein the K_d of the MM towards the AB is lower than 5nM.

112. The modified antibody of claim 109 wherein the K_d of the MM towards the AB is about 1nM.

113. The AA of claim 104 wherein the MM is capable of specifically binding to the antigen-binding domain of the AB.

114. The AA of claim 104 wherein the CM is capable of being specifically cleaved by an enzyme at a rate of about at least $1 \times 10^4 \text{ M}^{-1}\text{S}^{-1}$.

115. The AA of claim 104 wherein the CM is capable of being specifically reduced by an enzyme at a rate of at least $5 \times 10^4 \text{ M}^{-1}\text{S}$.

116. The AA of claim 104 wherein the CM is capable of being specifically reduced by an enzyme at a rate of at least $10 \times 10^4 \text{ M}^{-1}\text{S}$.

117. The AA of claim 104 wherein the antibody fragment is selected from the group consisting of a Fab' fragment, a F(ab')₂ fragment, a scFv, a scAB a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.

118. The AA of claim 104 wherein the AB is selected from the group consisting of the antibodies in Table 2.

119. The AA of claim 118 wherein the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

120. The AA of claim 104 wherein the target is selected from the group consisting of the targets in Table 1.

121. The AA of claim 120 wherein the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

122. The AA of claim 104 wherein the AB is not alemtuzumab.

123. The AA of claim 104 wherein the target is not CD52.

124. The AA of claim 104 wherein the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3.

125. The AA of claim 124 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

126. The AA of claim 104 further coupled to a second cleavable moiety (CM), capable of being specifically modified by an enzyme.

127. The AA of claim 126 wherein the second CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

128. The AA of claim 104 wherein the CM is located within the MM.

129. The AA of claim 104 wherein the MM does not comprise more than 50% amino acid sequence similarity to a natural binding partner of the AB.

130. The AA of claim 104 further comprising a linker peptide, wherein the linker peptide is positioned between the MM and the CM.

131. The AA of claim 104 further comprising a linker peptide, wherein the linker peptide is positioned between the AB and the CM.

132. The AA of claim 104 further comprising a detectable moiety.

133. The AA of claim 104 further comprising an agent conjugated to the AB.

134. An activatable antibody complex (AAC) comprising:

(a) two antibodies or antibody fragments (AB1 and AB2), each capable of specifically binding its target;

(b) at least one masking moiety (MM) coupled to either AB1 or AB2, capable of inhibiting the specific binding of AB1 and AB2 to their targets; and

(c) at least one cleavable moiety (CM) coupled to either AB1 or AB2, capable of being specifically cleaved by an enzyme whereby activating the AAC composition;

wherein when the AAC is in an uncleaved state, the MM inhibits the specific binding of AB1 and AB2 to their targets and when the AAC is in a cleaved state, the MM does not inhibit the specific binding of AB1 and AB2 to their targets.

135. The composition of claim 134 wherein the complex is bispecific.

- 136.** The composition of claim 134 wherein AB1 and AB2 bind the same epitope on the same target.
- 137.** The composition of claim 134 wherein AB1 and AB2 bind to different epitopes on the same target.
- 138.** The composition of claim 134 wherein AB1 and AB2 bind to different epitopes on different targets.
- 139.** The composition of claim 134 wherein the CM is capable of being specifically cleaved by an enzyme at a rate of about at least $1 \times 10^4 \text{ M}^{-1}\text{S}^{-1}$.
- 140.** The composition of claim 134 wherein the antibody fragment is selected from the group consisting of a Fab' fragment, a F(ab')₂ fragment, a scFv, a scAB, a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.
- 141.** The composition of claim 134 wherein AB1 and/or AB 2 are selected from the group consisting of the antibodies in Table 2.
- 142.** The composition of claim 134 wherein AB1 and/or AB2 is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.
- 143.** The composition of claim 134 wherein the target is selected from the group consisting of the targets in Table 1.
- 144.** The composition of claim 134 wherein AB1 and/or AB2 is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.
- 145.** The composition of claim 134 wherein AB1 and AB2 are capable of binding to EGFR and VEGF, a Notch Receptor and EGFR, a Jagged ligand and EGFR or cMET and VEGF, respectively.
- 146.** The composition of claim 134 wherein the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3.
- 147.** The composition of claim 134 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.
- 148.** The composition of claim 134 further coupled to a second cleavable moiety (CM), capable of being specifically cleaved by an enzyme.
- 149.** The composition of claim 148 wherein the second CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

150. The composition of claim 134 wherein the MM does not comprise more than 50% amino acid sequence similarity to a natural binding partner of the AB.

151. The composition of claim 134 further comprising a detectable moiety.

152. The composition of claim 134 further comprising an agent conjugated to the AB.

153. A method of treating or diagnosing a condition in a subject including administering to the subject a composition comprising:

(a) an antibody or antibody fragment (AB), capable of specifically binding its target;

(b) a masking moiety (MM) coupled to the AB, capable of inhibiting the specific binding of the AB to its target; and

(c) a cleavable moiety (CM) coupled to the AB, capable of being specifically cleaved by an enzyme;

wherein upon administration to the subject, when the AA is not in the presence of sufficient enzyme activity to cleave the CM, the MM reduces the specific binding of the AB to its target by at least 90% when compared to when the AA is in the presence of sufficient enzyme activity to cleave the CM and the MM does not inhibit the specific binding of the AB to its target.

154. The method of claim 153 wherein the AB is selected from the group consisting of a Fab' fragment, a F(ab')₂ fragment, a scFv, a scAB a dAb, a single domain heavy chain antibody, and a single domain light chain antibody.

155. The method of claim 153 wherein the condition is cancer.

156. The method of claim 153 wherein the MM is not the natural binding partner of the AB.

157. The method of claim 153 wherein the AB is selected from the group consisting of the antibodies in Table 2.

158. The method of claim 157 wherein the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

159. The method of claim 153 wherein the target is selected from the group of targets in Table 1.

160. The method of claim 159 wherein the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

161. The method of claim 153 wherein the AB is not alemtuzumab.

162. The method of claim 153 wherein the target is not CD52.

163. The method of claim 153 wherein the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3.

164. The method of claim 163 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

165. A method of inhibiting angiogenesis in a mammalian subject, said method comprising administering to a subject in need thereof a therapeutically effective amount of a pharmaceutical composition of claim 12.

166. The method of claim 165 wherein the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

167. The method of claim 165 wherein the AB is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

168. The method of claim 165 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

169. A method of making an activatable antibody (AA) composition comprising:

- (a) providing an antibody or antibody fragment (AB) capable of specifically binding its target;
- (b) coupling a masking moiety (MM) to the AB, capable of inhibiting the specific binding of the AB to its target; and
- (c) coupling a cleavable moiety (CM) to the AB, capable of being specifically cleaved by an enzyme;

wherein the dissociation constant (K_d) of the AB coupled to the MM towards the target is at least 100 times greater than the K_d of the AB not coupled to the MM towards the target.

170. The method of claim 166 wherein the AB is or is derived from an antibody selected from the group consisting of the antibodies in Table 2.

171. The method of claim 166 wherein the AB is or is derived from cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

172. The method of claim 166 wherein the AB is not alemtuzumab.

173. The method of claim 166 wherein the target is not CD52.

174. The method of claim 166 wherein the CM is a substrate for an enzyme selected from the group consisting of the enzymes in Table 3.

175. The method of claim 166 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

176. A method of screening candidate peptides to identify a masking moiety (MM) peptide capable of binding an antibody or antibody fragment (AB) comprising:

- (a) providing a library of peptide scaffolds, wherein each peptide scaffold comprises:
 - (i) a transmembrane protein (TM); and
 - (ii) a candidate peptide;
- (b) contacting an AB with the library;
- (c) identifying at least one candidate peptide capable of binding the AB; and
- (d) determining whether the dissociation constant (K_d) of the candidate peptide towards the AB is between 1-10 nM.

177. The method of claim 176 wherein the library comprises viruses, cells or spores.

178. The method of claim 176 wherein the library comprises *E. coli*

179. The method of claim 176 wherein the peptide scaffold further comprises a detectable moiety.

180. A method of screening candidate peptides to identify a masking moiety (MM) peptide capable of masking an antibody or antibody fragment (AB) with an optimal masking efficiency comprising:

- (a) providing a library comprising a plurality of ABs, each coupled to a candidate peptide, wherein the ABs are capable of specifically binding a target;
- (b) incubating each library member with the target; and
- (c) comparing the binding affinity of each library member towards the target with the binding affinity of each AB not coupled to a candidate peptide towards the target.

181. The method of claim 180 step c wherein optimal binding efficiency is when the binding affinity of a library member to the target is 10% compared to the binding affinity of the unmodified AB to the target.

182. The method of claim 180 wherein the ABs are the same.

183. An antibody therapeutic having an improved bioavailability wherein the affinity of binding of the antibody therapeutic to its target is lower in a first tissue when compared to the binding of the antibody therapeutic to its target in a second tissue.

184. The antibody therapeutic of claim 183 wherein the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

185. The antibody therapeutic of claim 183 wherein the first tissue is a healthy tissue and the second tissue is a diseased tissue.

186. The antibody therapeutic of claim 183 wherein the first tissue is an early stage tumor and the second tissue is a late stage tumor.

187. The antibody therapeutic of claim 183 wherein the first tissue is a benign tumor and the second tissue is a malignant tumor.

188. The antibody therapeutic of claim 183 wherein the first tissue and second tissue are spatially separated.

189. The antibody therapeutic of claim 183 wherein the first tissue is epithelial tissue and the second tissue is breast, head, neck, lung, pancreatic, nervous system, liver, prostate, urogenital, or cervical tissue.

190. The antibody therapeutic of claim 183 further coupled to an agent.

191. The antibody therapeutic of claim 190 wherein the agent is an antineoplastic agent.

192. A pharmaceutical composition comprising:

- (a) an antibody or antibody fragment (AB), capable of specifically binding its target; and
- (b) a pharmaceutically acceptable excipient;

wherein the affinity of the antibody or antibody fragment to the target in a first tissue is lower than the affinity of the antibody or antibody fragment to the target in a second tissue.

193. The composition of claim 192 wherein the affinity in the first tissue is 10-1.000 times lower than the affinity in the second tissue.

194. The composition of claim 192 wherein the AB is coupled to a masking moiety (MM), capable of reducing the binding of the AB to its target and a cleavable moiety (CM), capable of specifically being cleaved by an enzyme.

195. The composition of claim 192 wherein the target is EGFR, TNFalpha, CD11a, CSFR, CTLA-4, EpCAM, VEGF, CD40, CD20, Notch 1, Notch 2, Notch 3, Notch 4, Jagged 1, Jagged 2, CD52, MUC1, IGF1R, transferrin, gp130, VCAM-1, CD44, DLL4, or IL4.

196. The composition of claim 192 wherein the first tissue is a healthy tissue and the second tissue is a diseased tissue.

197. The composition of claim 192 wherein the first tissue is an early stage tumor and the second tissue is a late stage tumor.

198. The composition of claim 192 wherein the first tissue is a benign tumor and the second tissue is a malignant tumor.

199. The composition of claim 192 wherein the first tissue and second tissue are spatially separated.

200. The composition of claim 192 wherein the first tissue is epithelial tissue and the second tissue is breast, head, neck, lung, pancreatic, nervous system, liver, prostate, urogenital, or cervical tissue.

201. The composition of claim 194 wherein the CM is a substrate for legumain, plasmin, TMPRSS-3/4, MMP-9, MT1-MMP, cathepsin, caspase, human neutrophil elastase, beta-secretase, uPA, or PSA.

202. The composition of claim 192 wherein the antibody or antibody fragment is cetuximab, panitumumab, infliximab, adalimumab, efalizumab, ipilimumab, tremelimumab, adecatumumab, Hu5c8, alemtuzumab, ranibizumab, tositumomab, ibritumomab tiuxetan, rituximab, infliximab, bevacizumab, or figitumumab.

203. The composition of claim 192 further comprising an agent coupled to the AB.

204. The composition of claim 203 wherein the agent is an antineoplastic agent.

205. A composition comprising a legumain-activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

206. A composition comprising a plasmin-activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

207. A composition comprising a caspase-activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

208. A composition comprising a TMPRSS-3/4 -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

209. A composition comprising a PSA -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

210. A composition comprising a cathepsin -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

211. A composition comprising a human neutrophil elastase -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

212. A composition comprising a beta-secretase -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

213. A composition comprising an uPA -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

214. A composition comprising a TMPRSS-3/4 -activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

215. A composition comprising a MT1-MMP-activatable antibody or antibody fragment (AB) coupled to a masking moiety (MM).

216. A composition comprising an activatable EGFR antibody or antibody fragment (AB) coupled to a masking moiety (MM).

217. A composition comprising an activatable TNFalpha antibody or antibody fragment (AB) coupled to a masking moiety (MM).

218. A composition comprising an activatable CD11a antibody or antibody fragment (AB) coupled to a masking moiety (MM).

219. A composition comprising an activatable CSFR antibody or antibody fragment (AB) coupled to a masking moiety (MM).

220. A composition comprising an activatable CTLA-4 antibody or antibody fragment (AB) coupled to a masking moiety (MM).

221. A composition comprising an activatable EpCAM antibody or antibody fragment (AB) coupled to a masking moiety (MM).

222. A composition comprising an activatable CD40L antibody or antibody fragment (AB) coupled to a masking moiety (MM).

223. A composition comprising an activatable Notch1 antibody or antibody fragment (AB) coupled to a masking moiety (MM).

224. A composition comprising an activatable Notch3 antibody or antibody fragment (AB) coupled to a masking moiety (MM).

225. A composition comprising an activatable Jagged1 antibody or antibody fragment (AB) coupled to a masking moiety (MM).

226. A composition comprising an activatable Jagged2 antibody or antibody fragment (AB) coupled to a masking moiety (MM).

227. A composition comprising an activatable cetuximab antibody or antibody fragment (AB) coupled to a masking moiety (MM).

228. A composition comprising an activatable vectibix antibody or antibody fragment (AB) coupled to a masking moiety (MM).

229. A composition comprising an activatable infliximab antibody or antibody fragment (AB) coupled to a masking moiety (MM).

230. A composition comprising an activatable adalimumab antibody or antibody fragment (AB) coupled to a masking moiety (MM).

231. A composition comprising an activatable efalizumab antibody or antibody fragment (AB) coupled to a masking moiety (MM).

232. A composition comprising an activatable ipilimumab antibody or antibody fragment (AB) coupled to a masking moiety (MM).

233. A composition comprising an activatable tremelimumab antibody or antibody fragment (AB) coupled to a masking moiety (MM).

234. A composition comprising an activatable adecatumumab antibody or antibody fragment (AB) coupled to a masking moiety (MM).

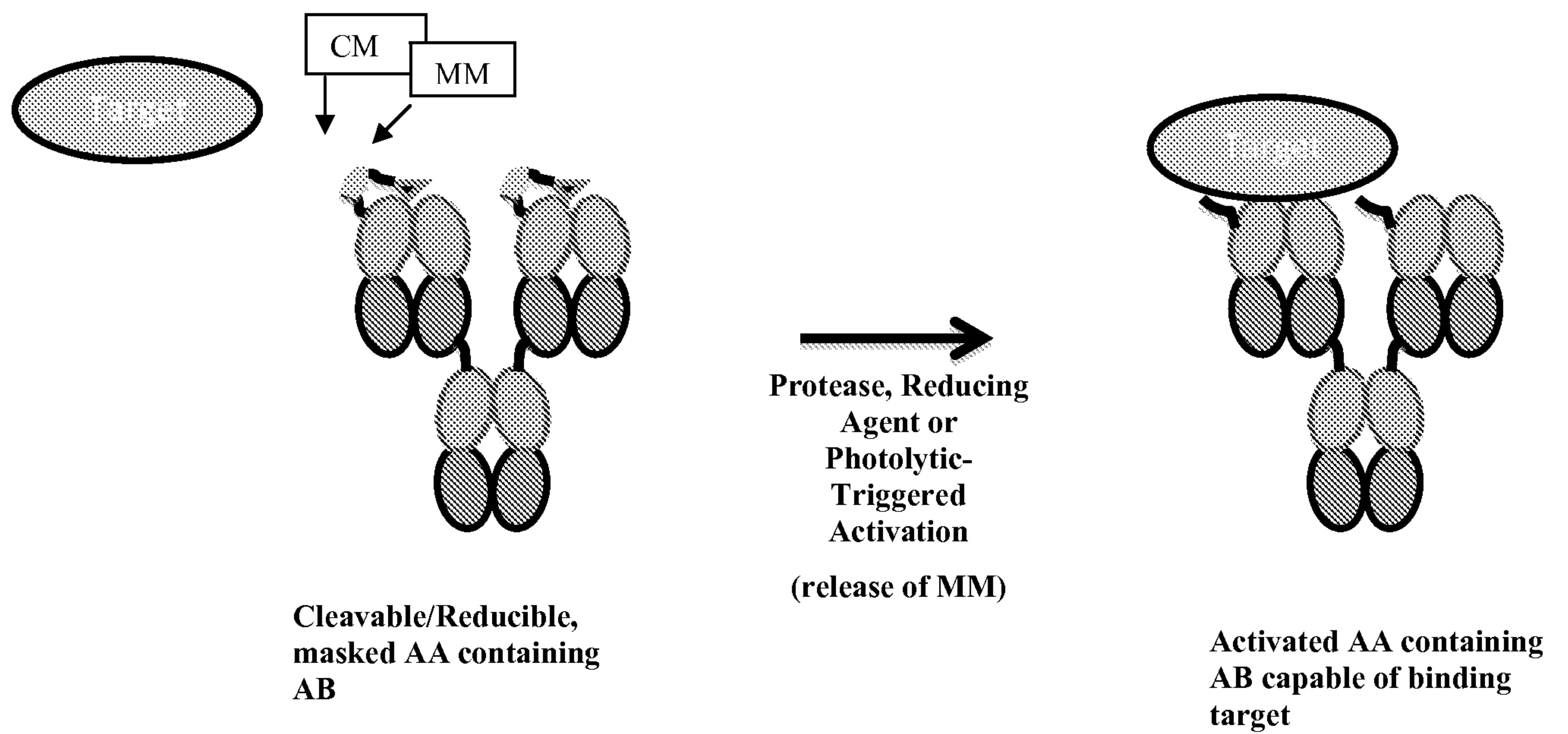
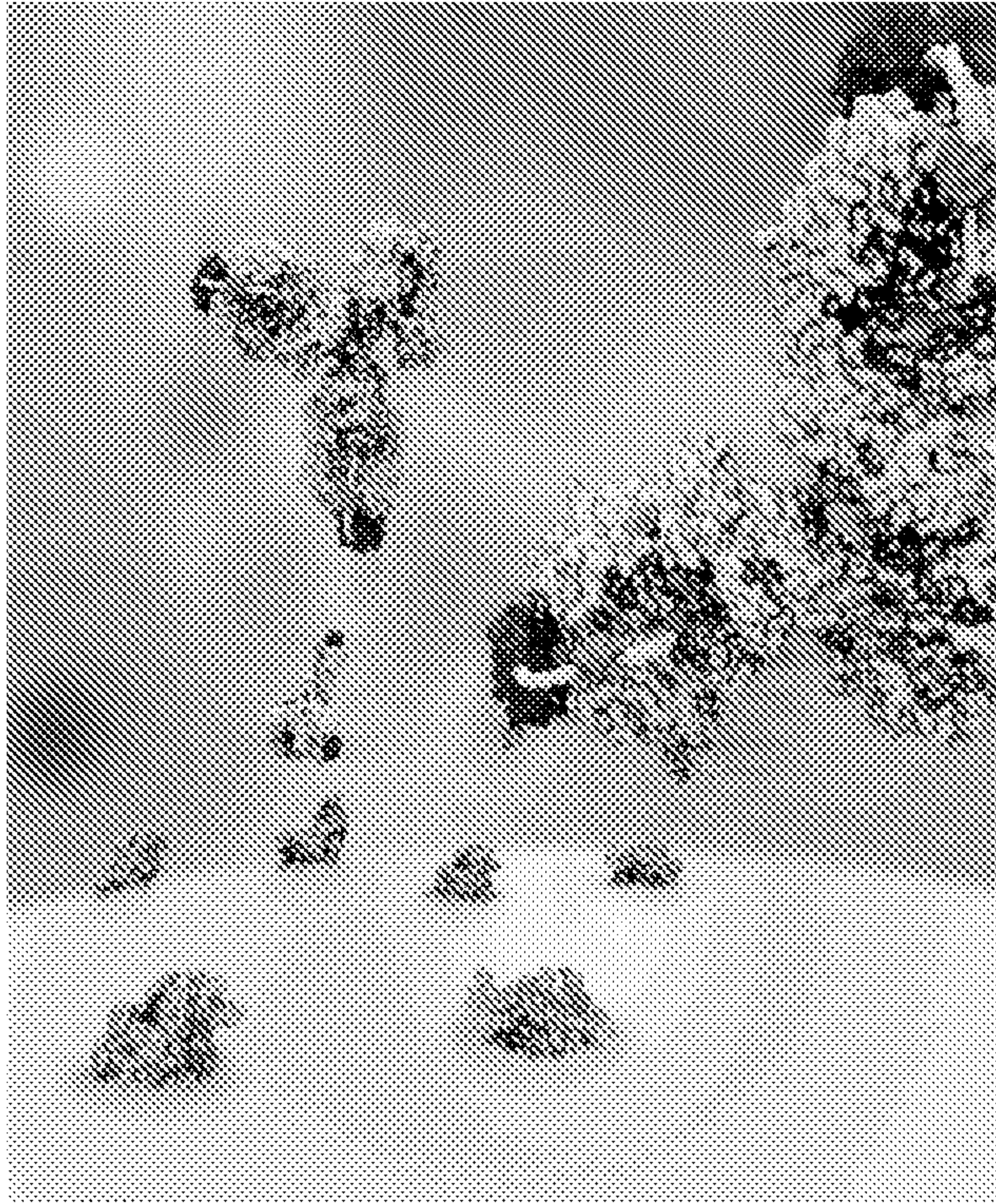
Figure 1

Figure 2

A



B

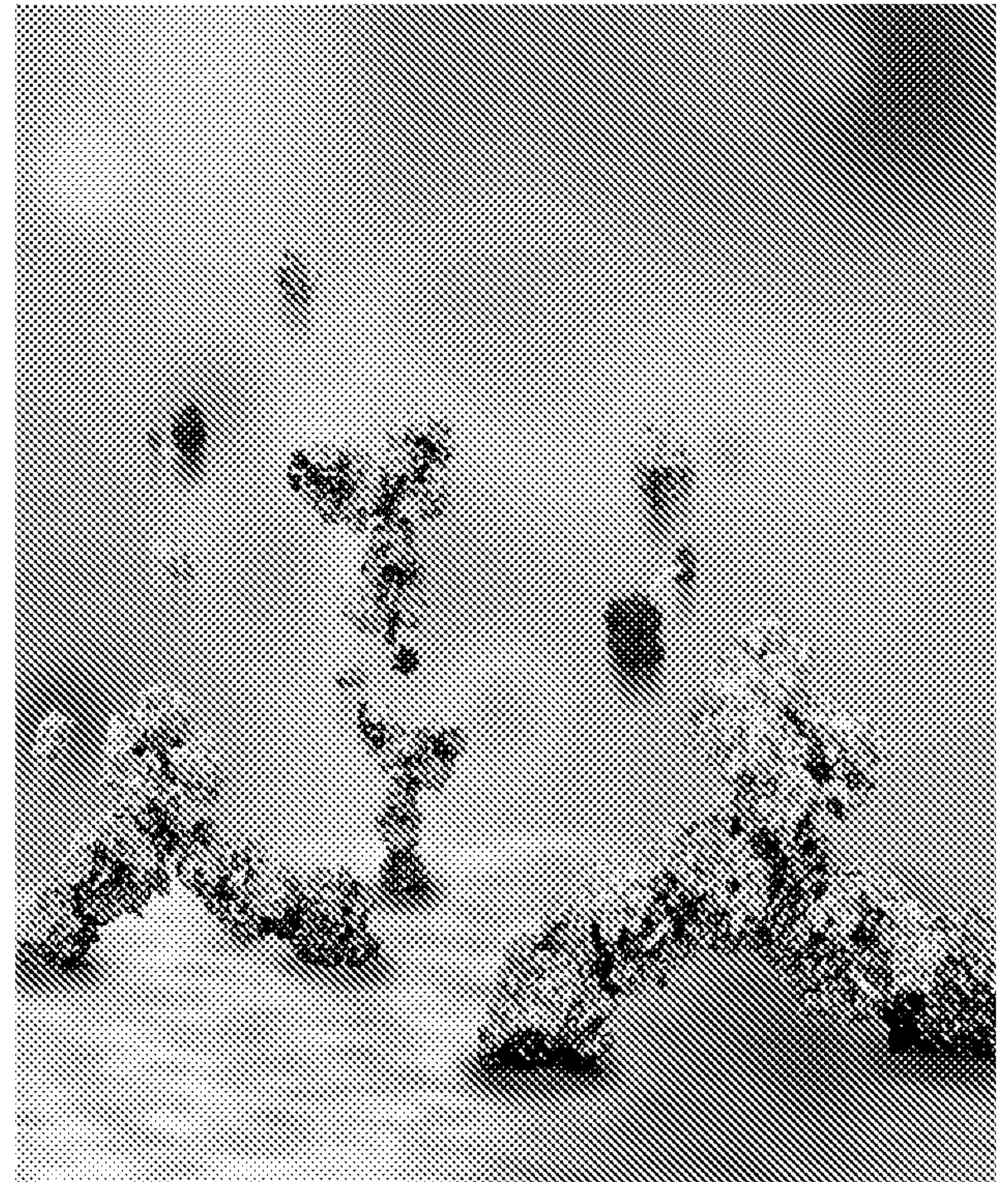


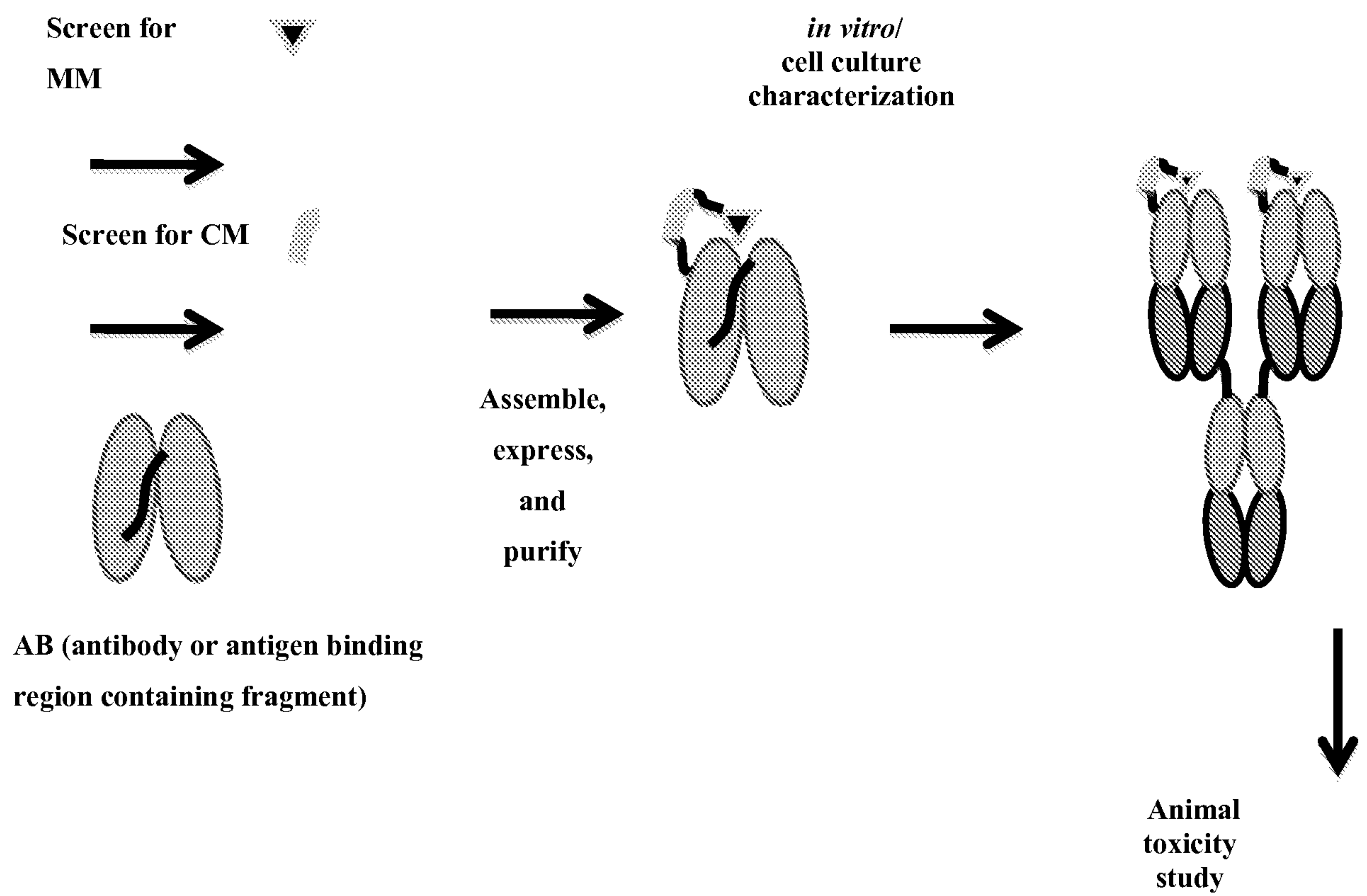
Figure 3

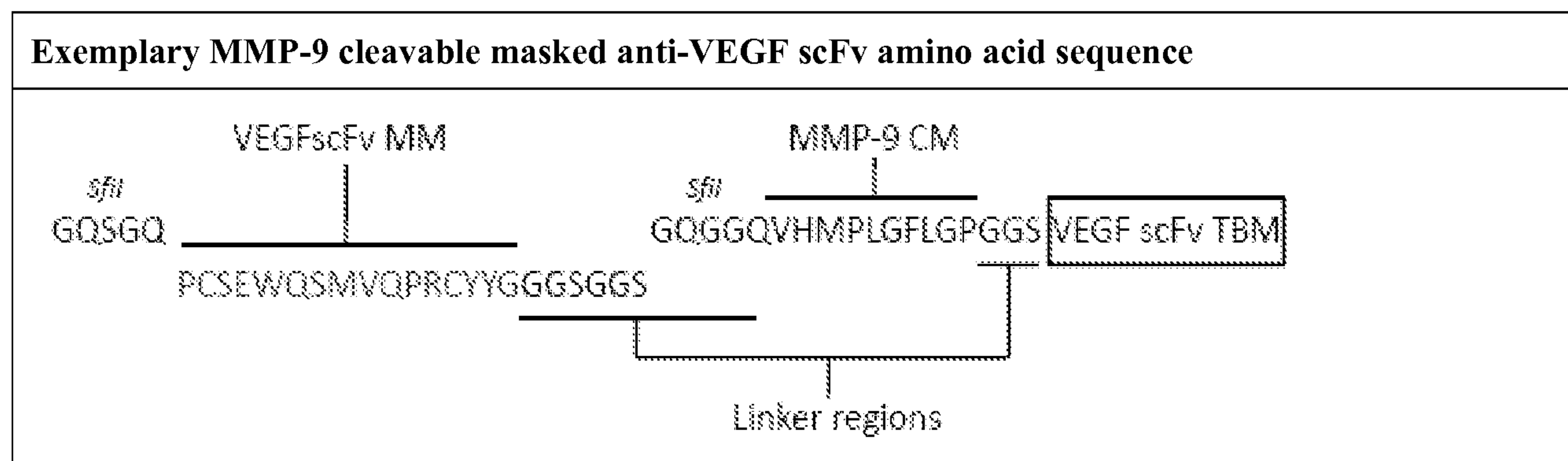
Figure 4

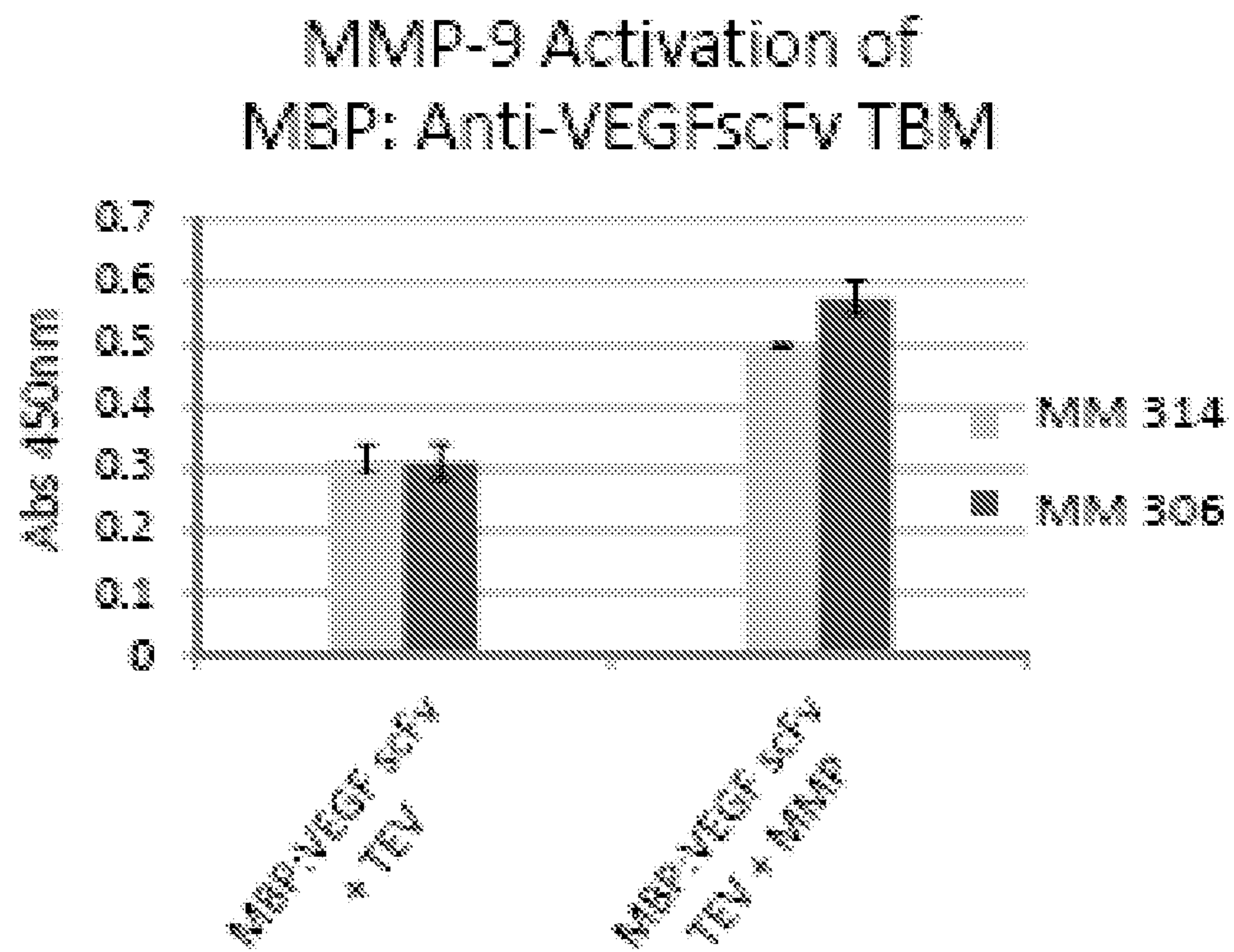
Figure 5

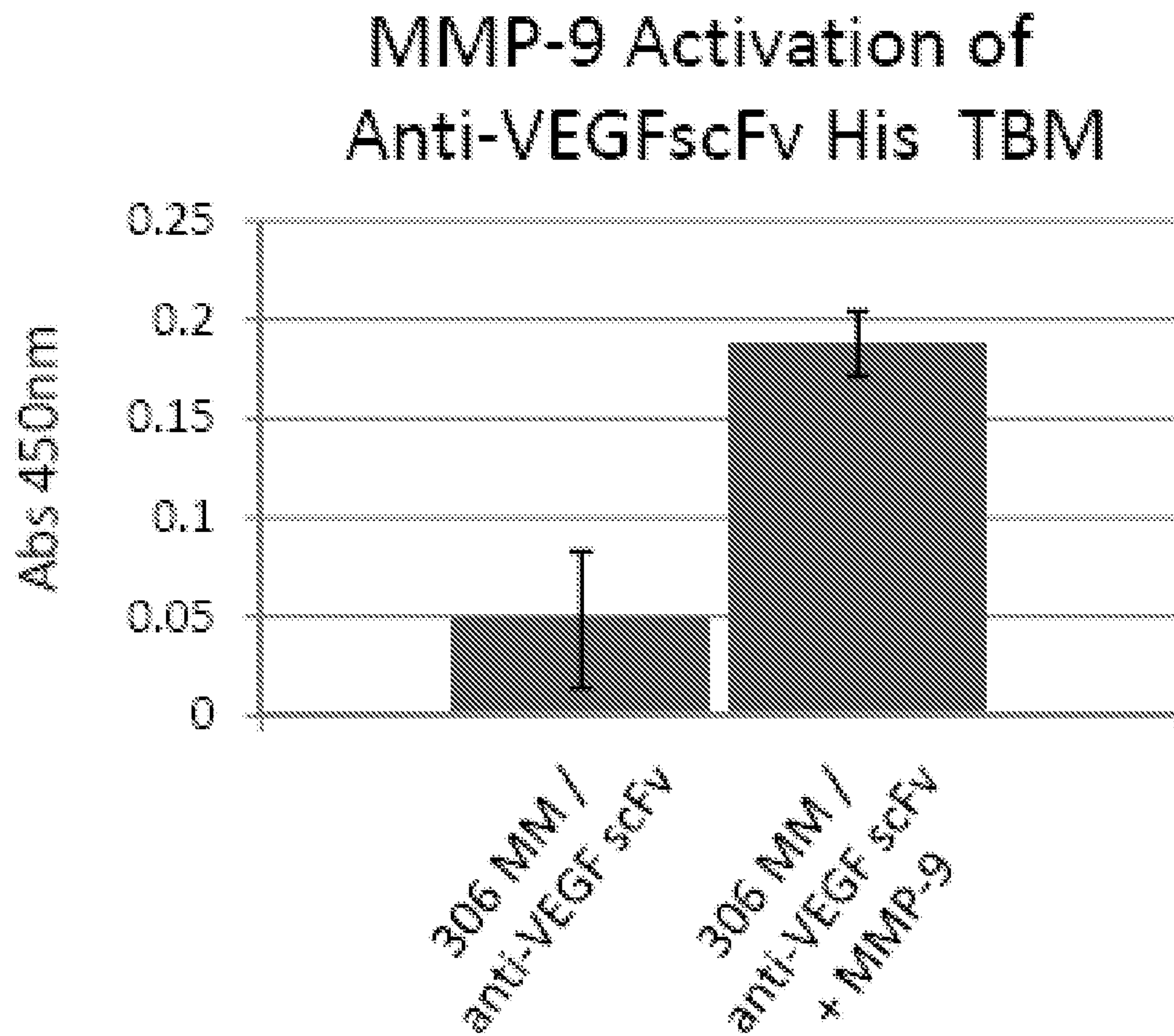
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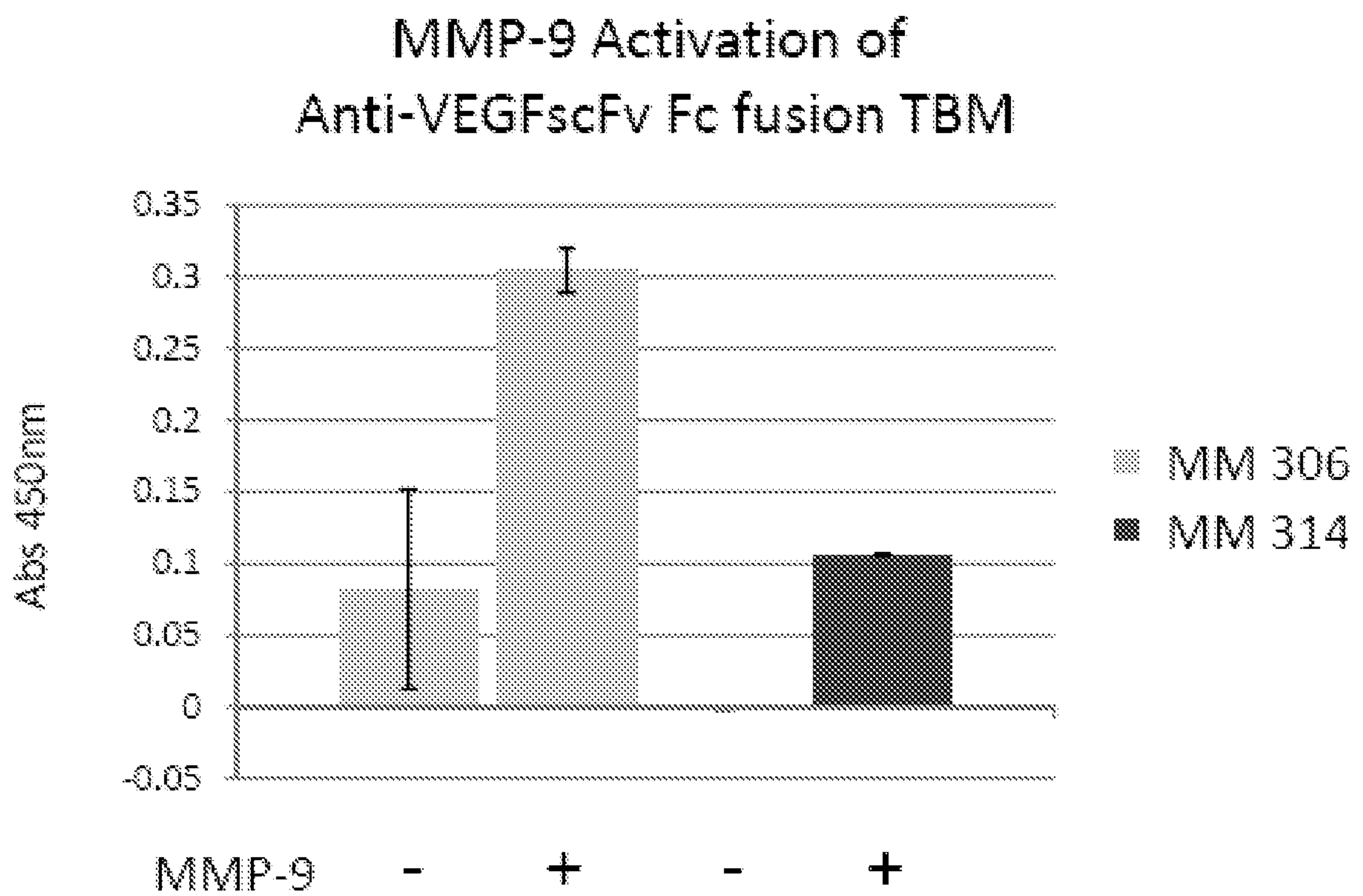
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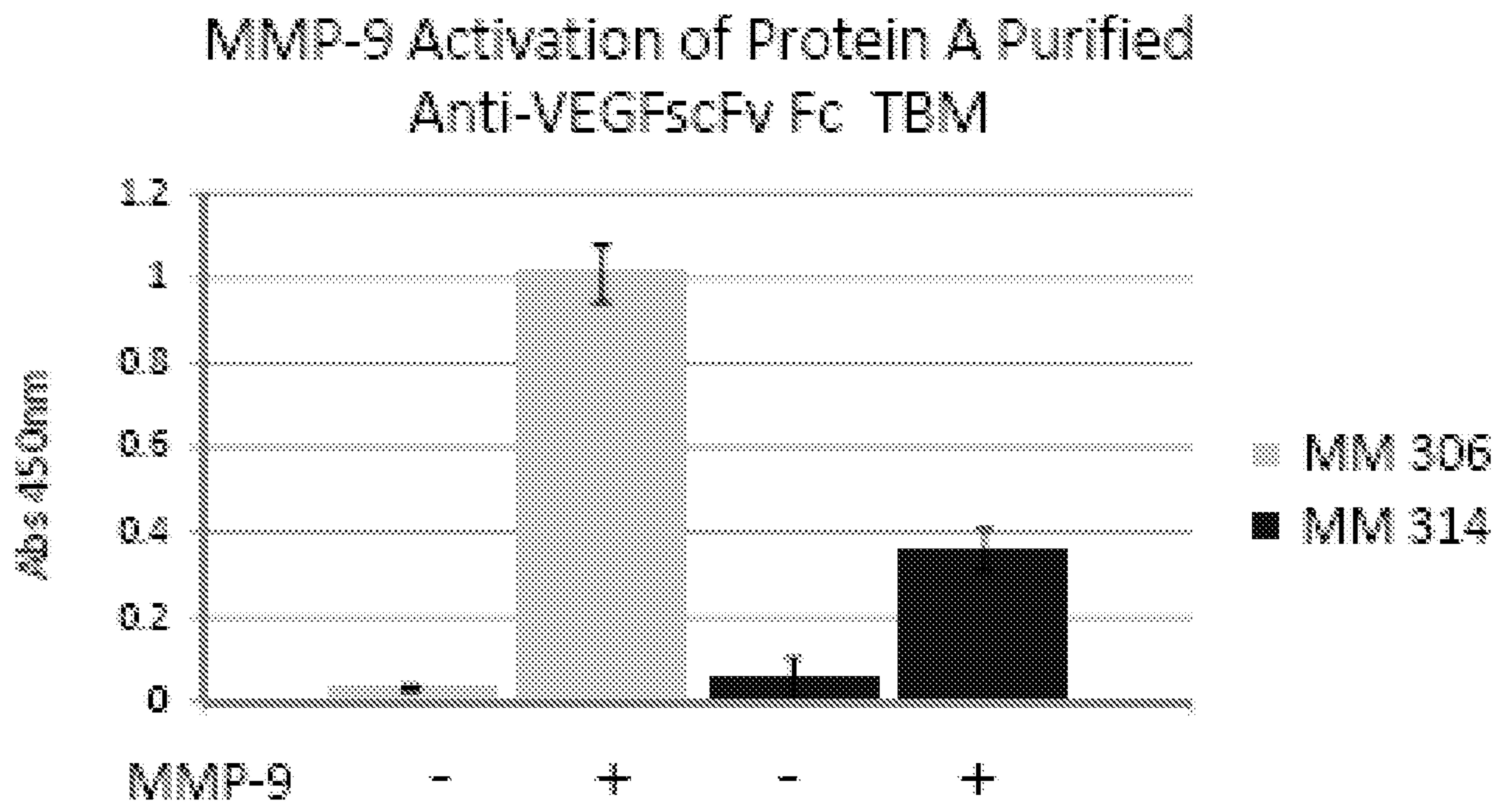
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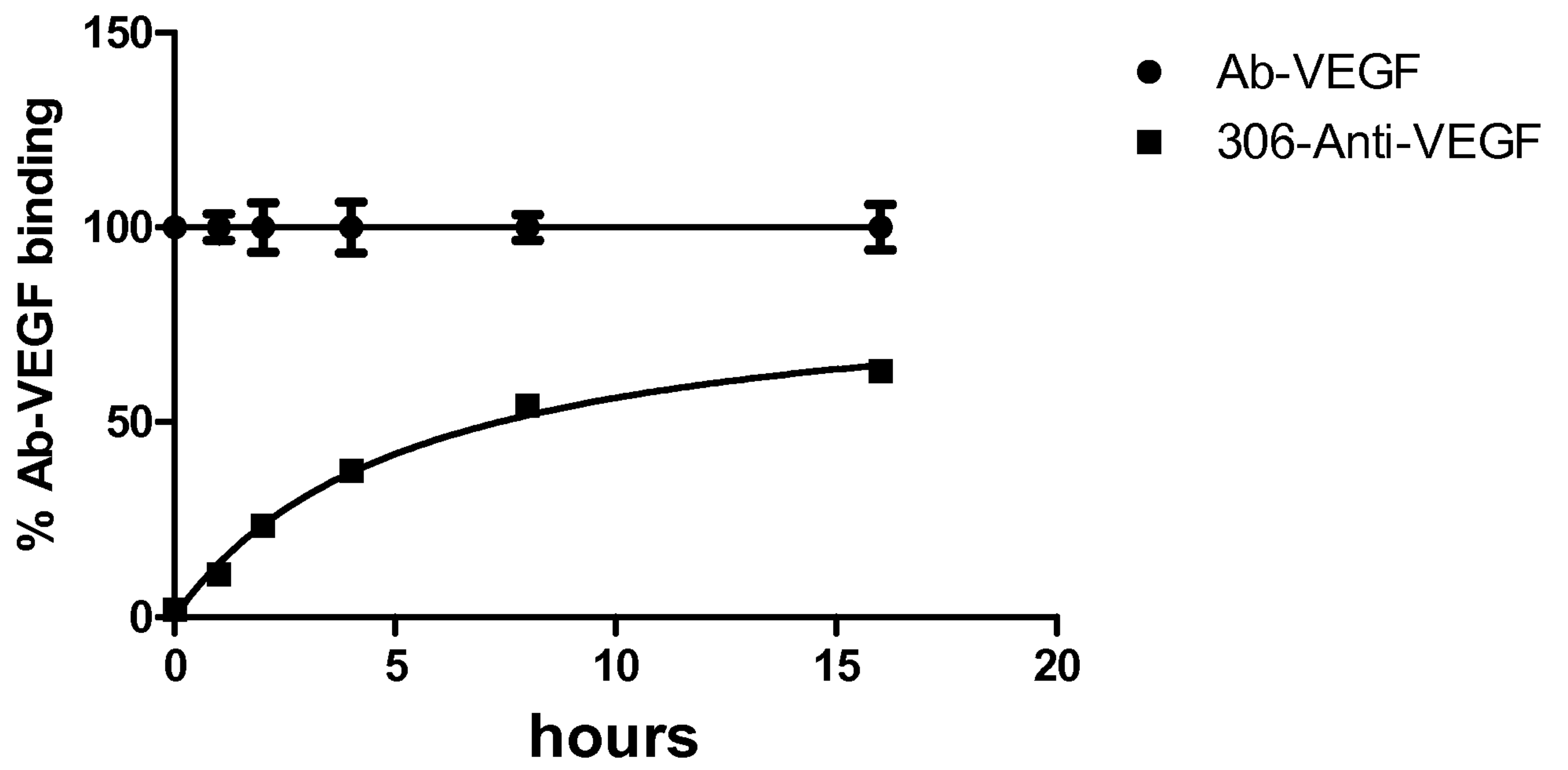
Figure 9

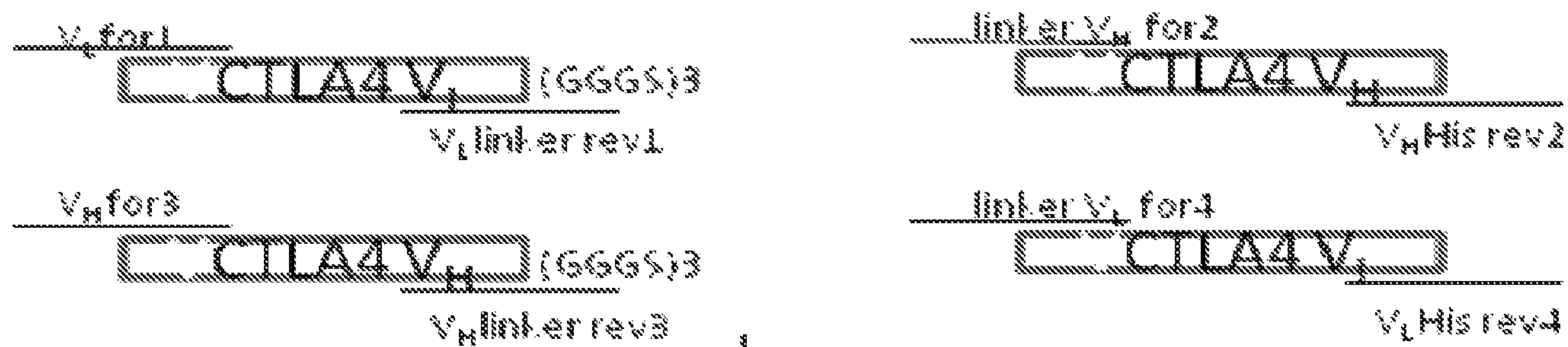
Figure 10**1st round PCR****2nd round PCR**

Figure 11

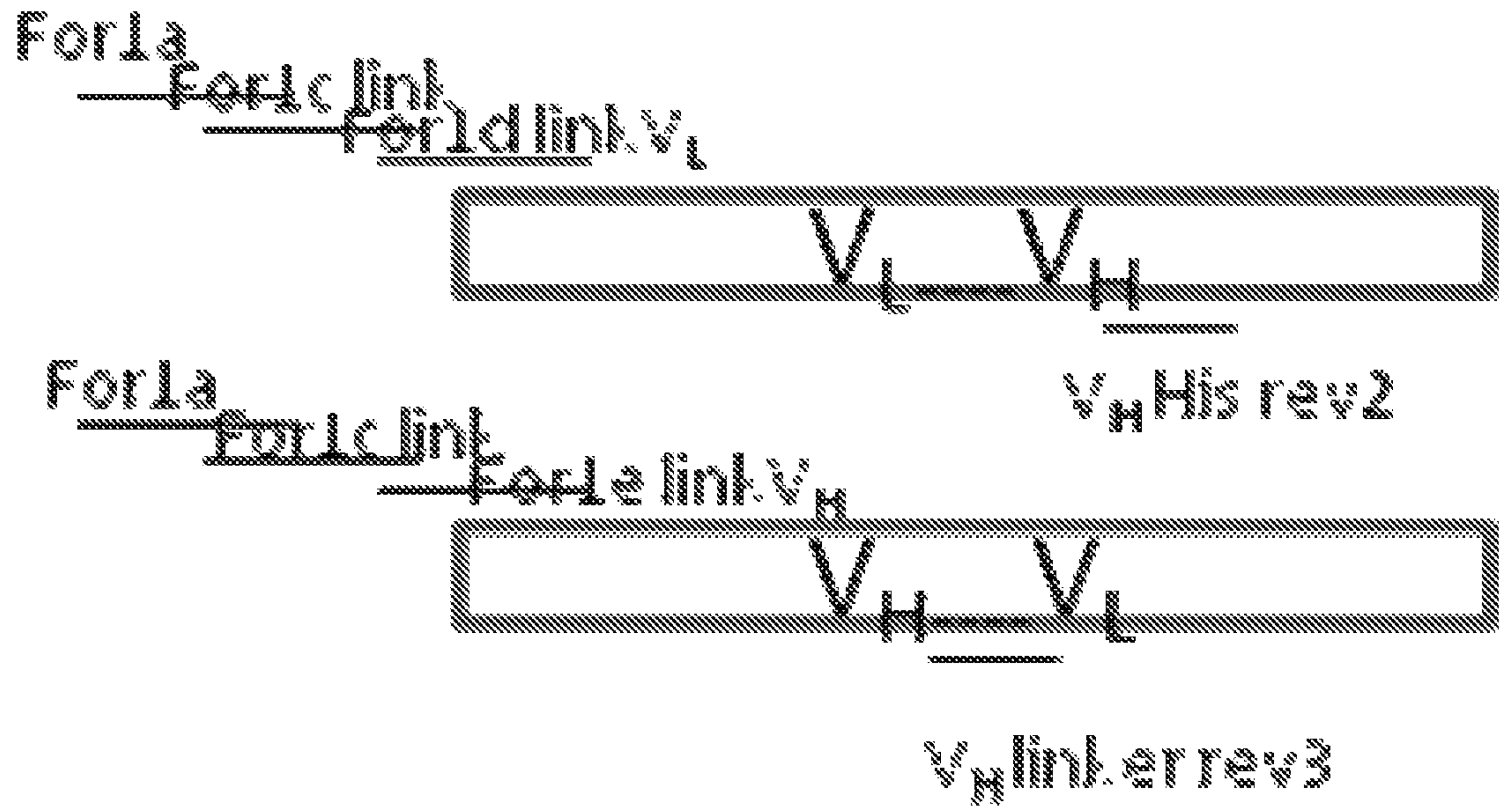


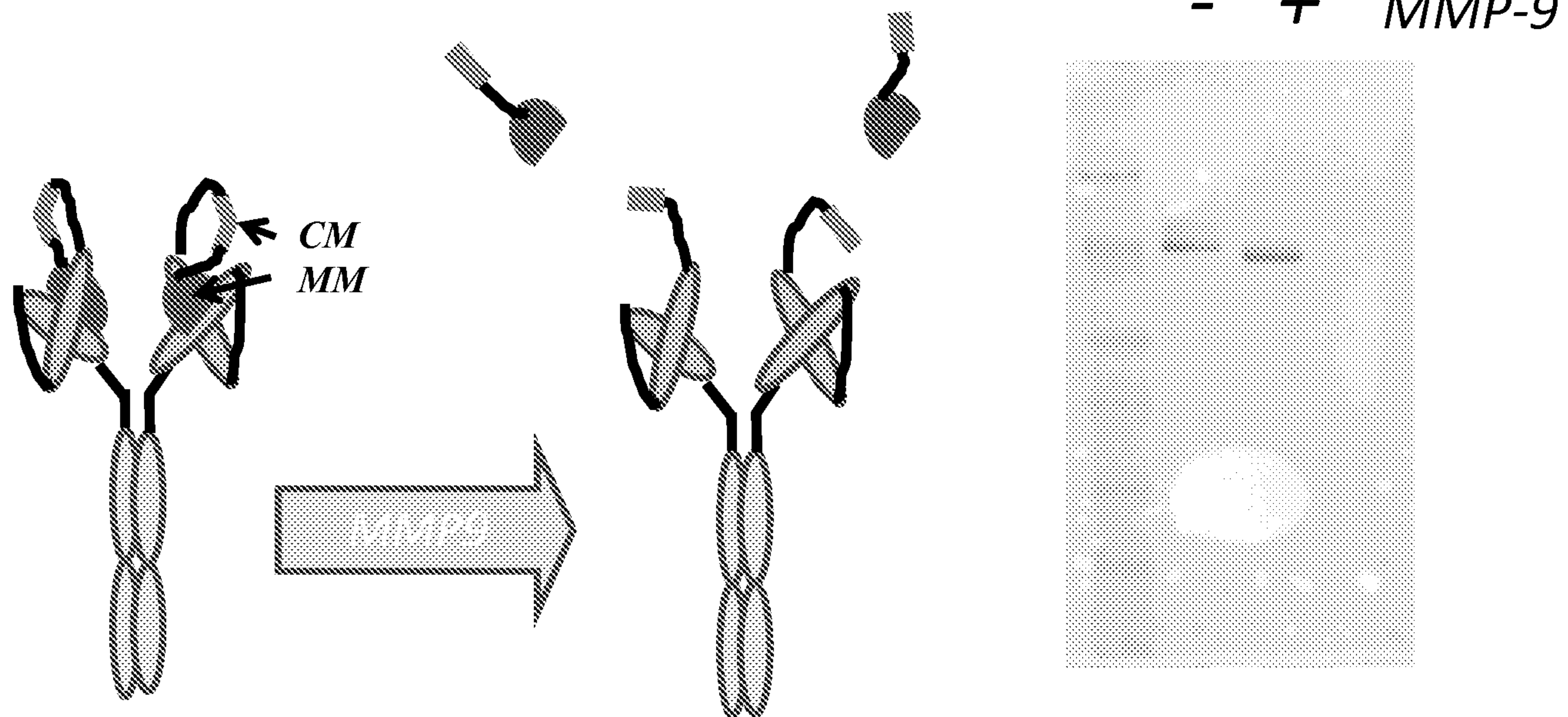
Figure 12

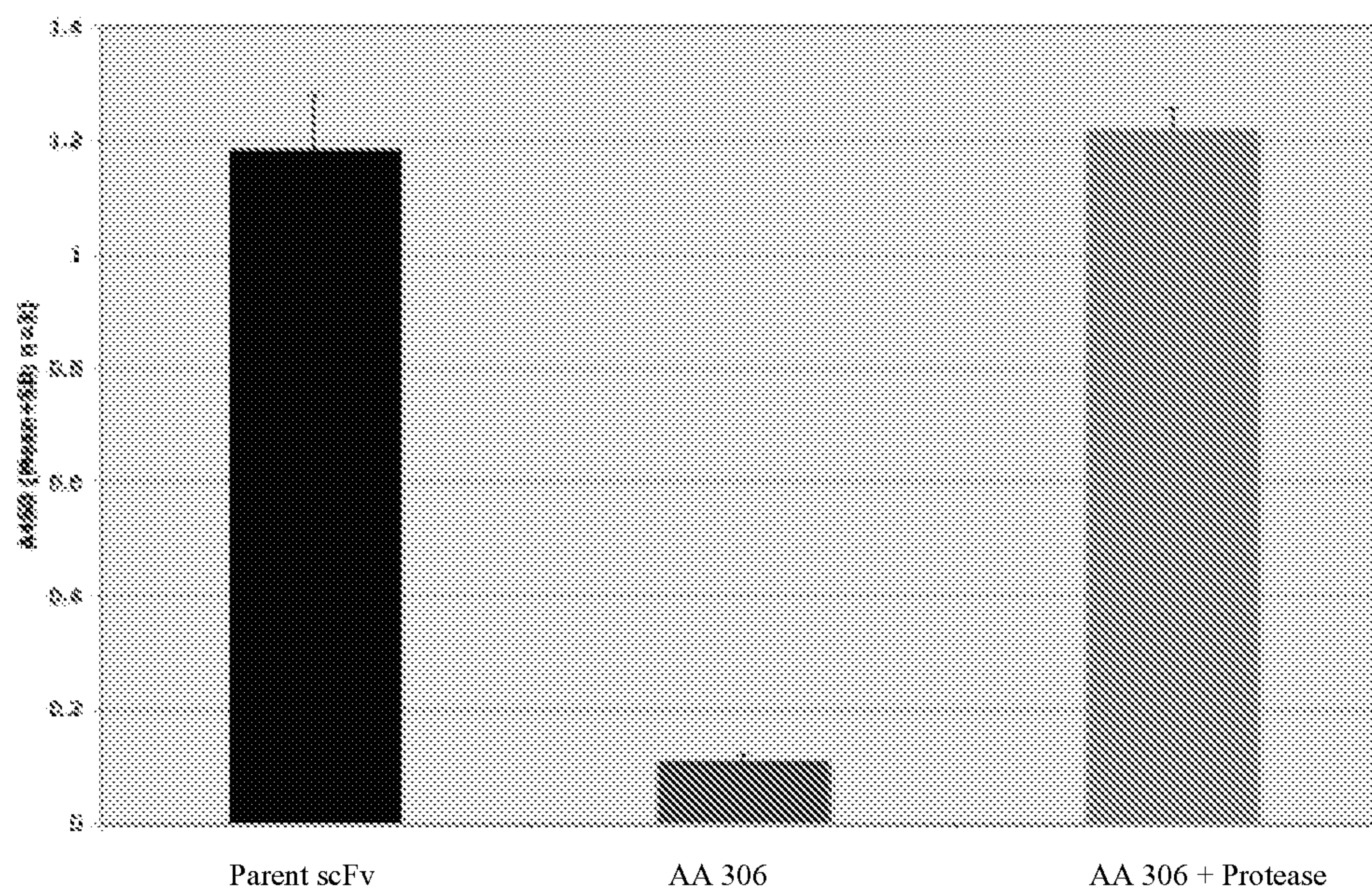
Figure 13

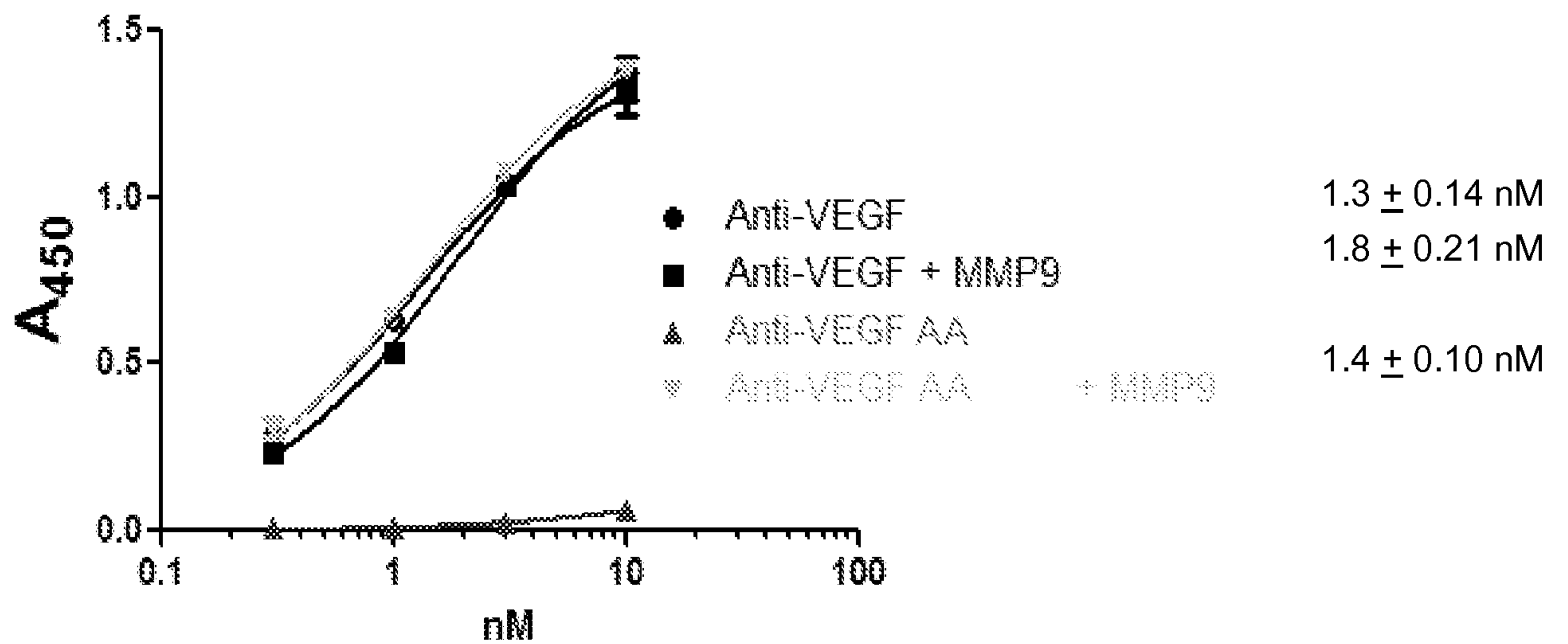
Figure 14

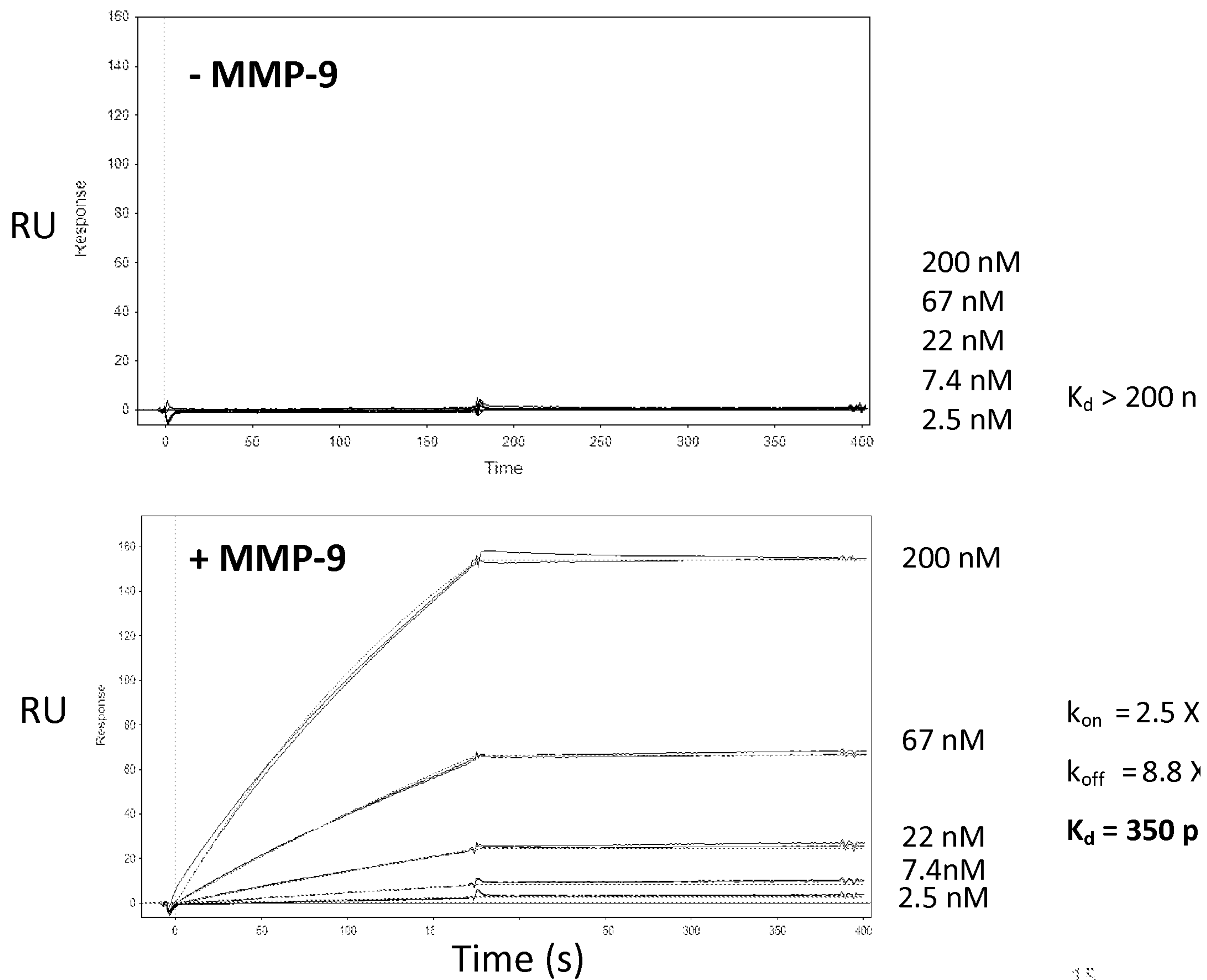
Figure 15

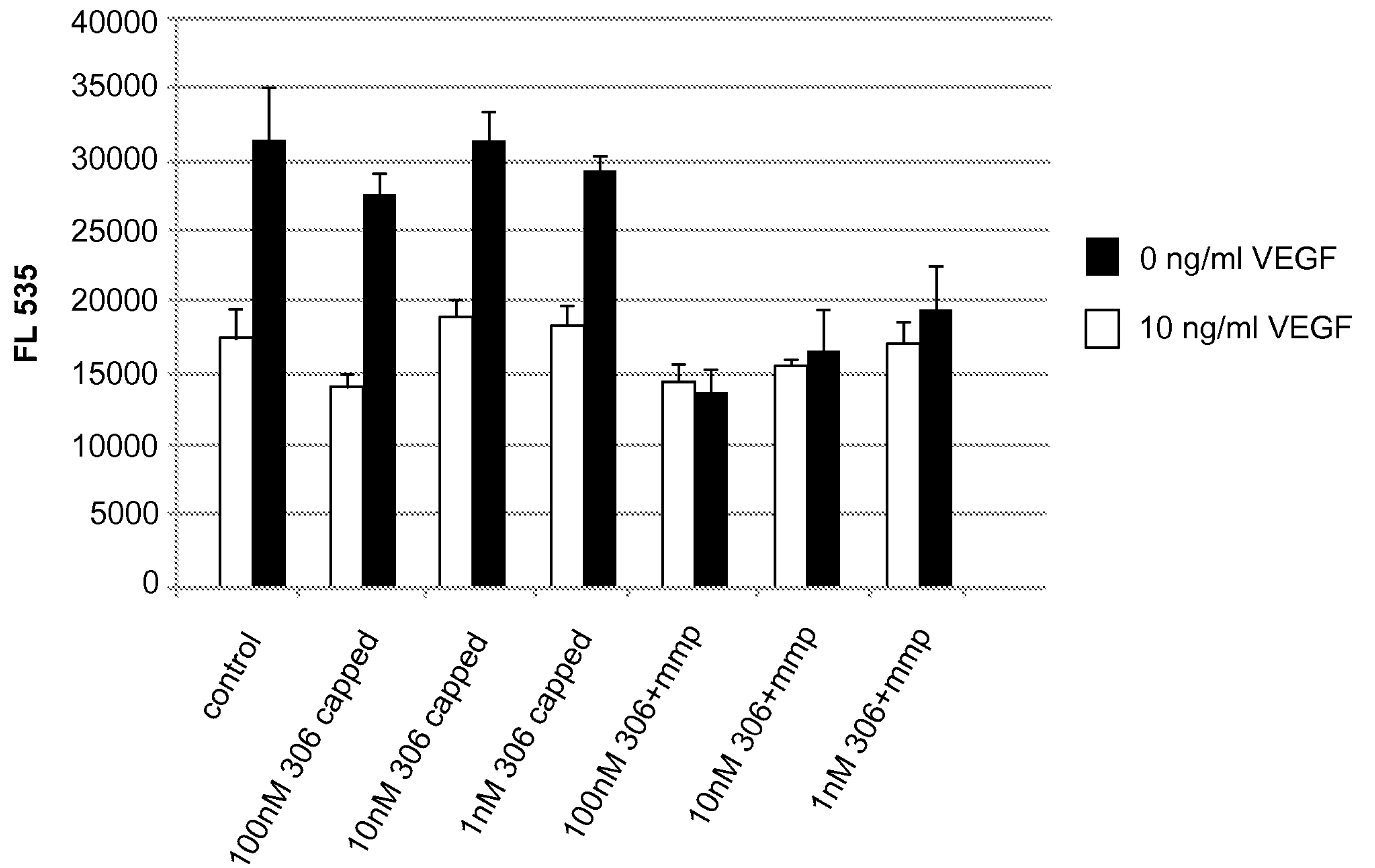
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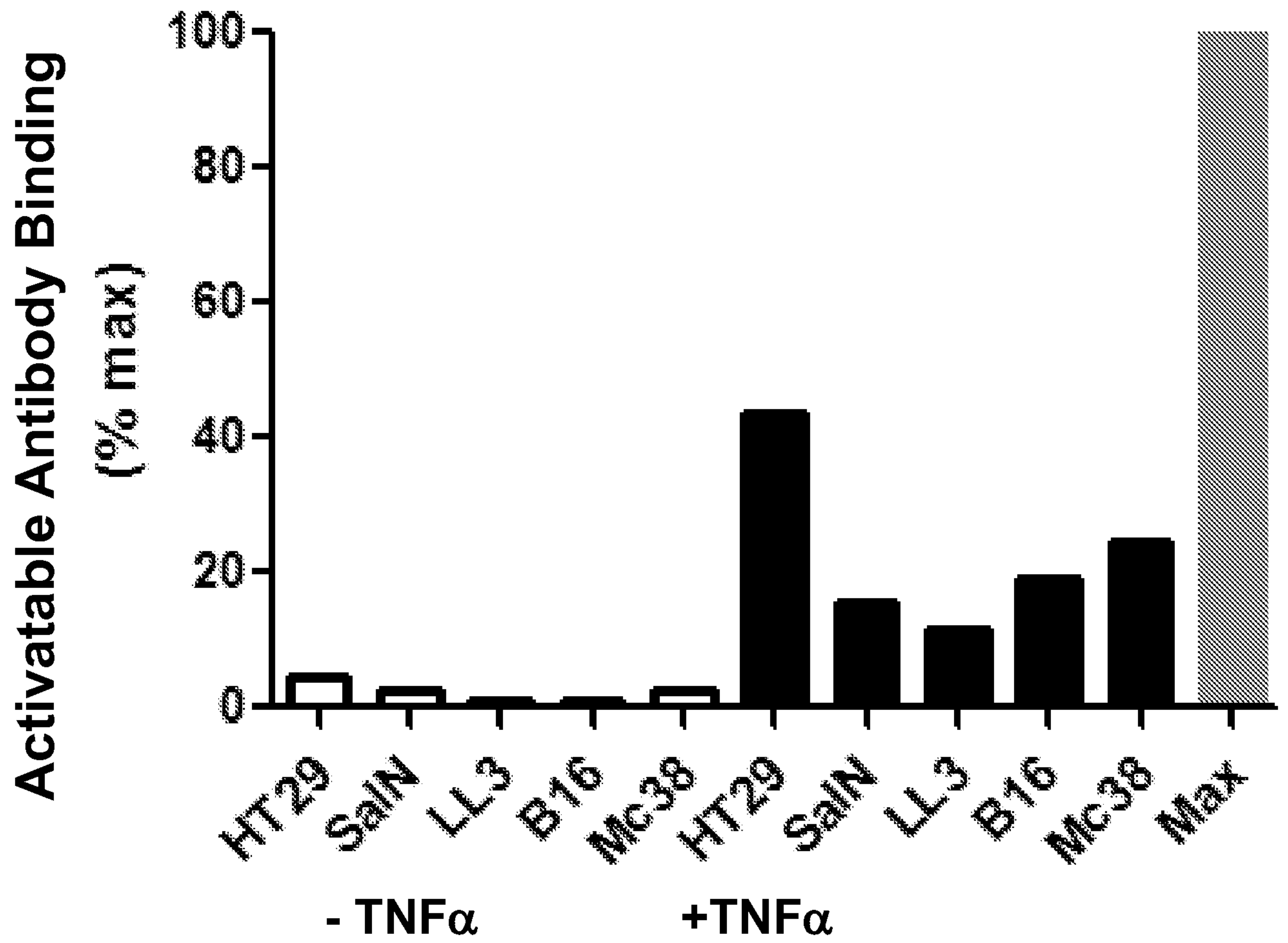
Figure 17

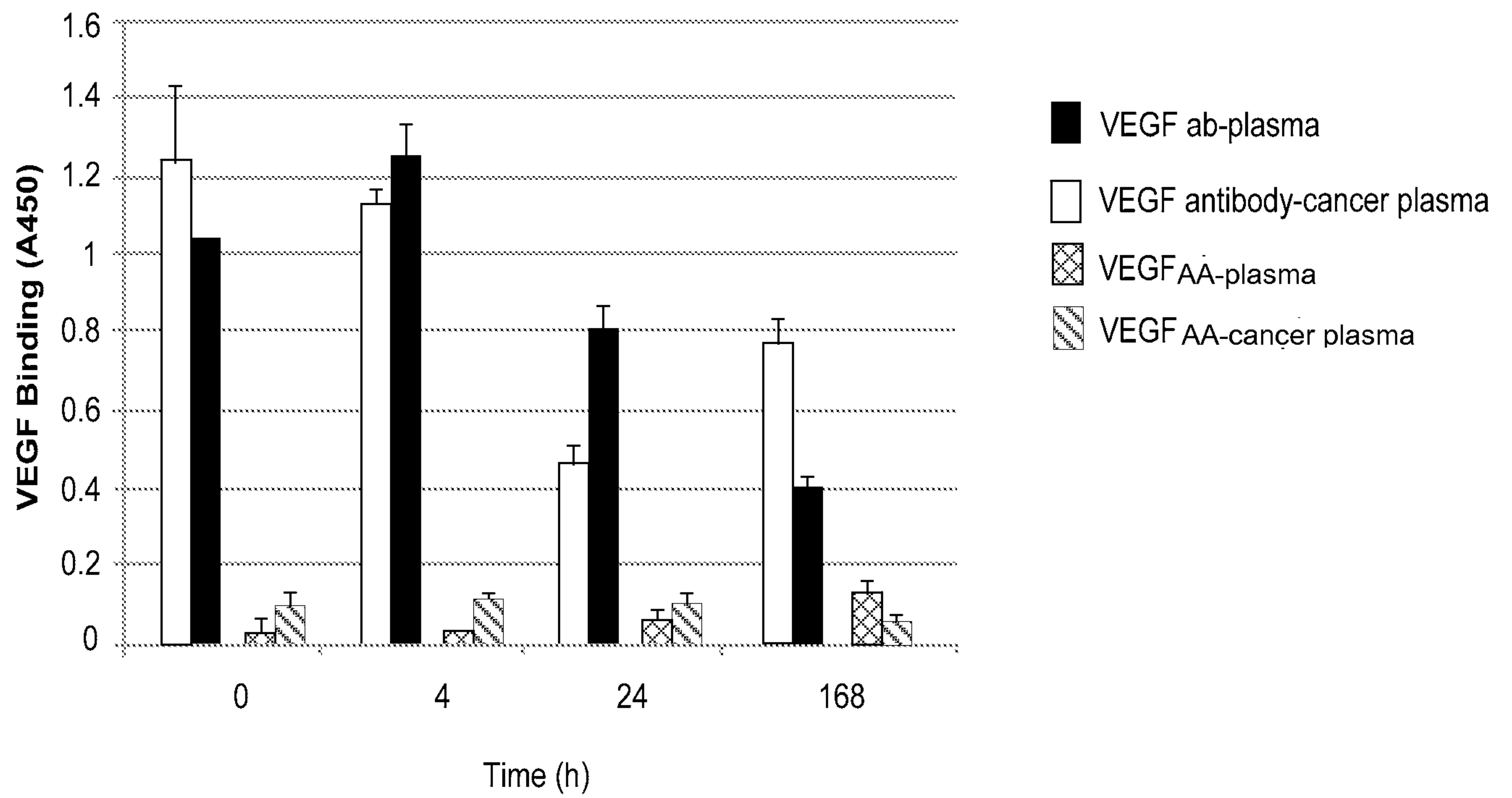
Figure 18

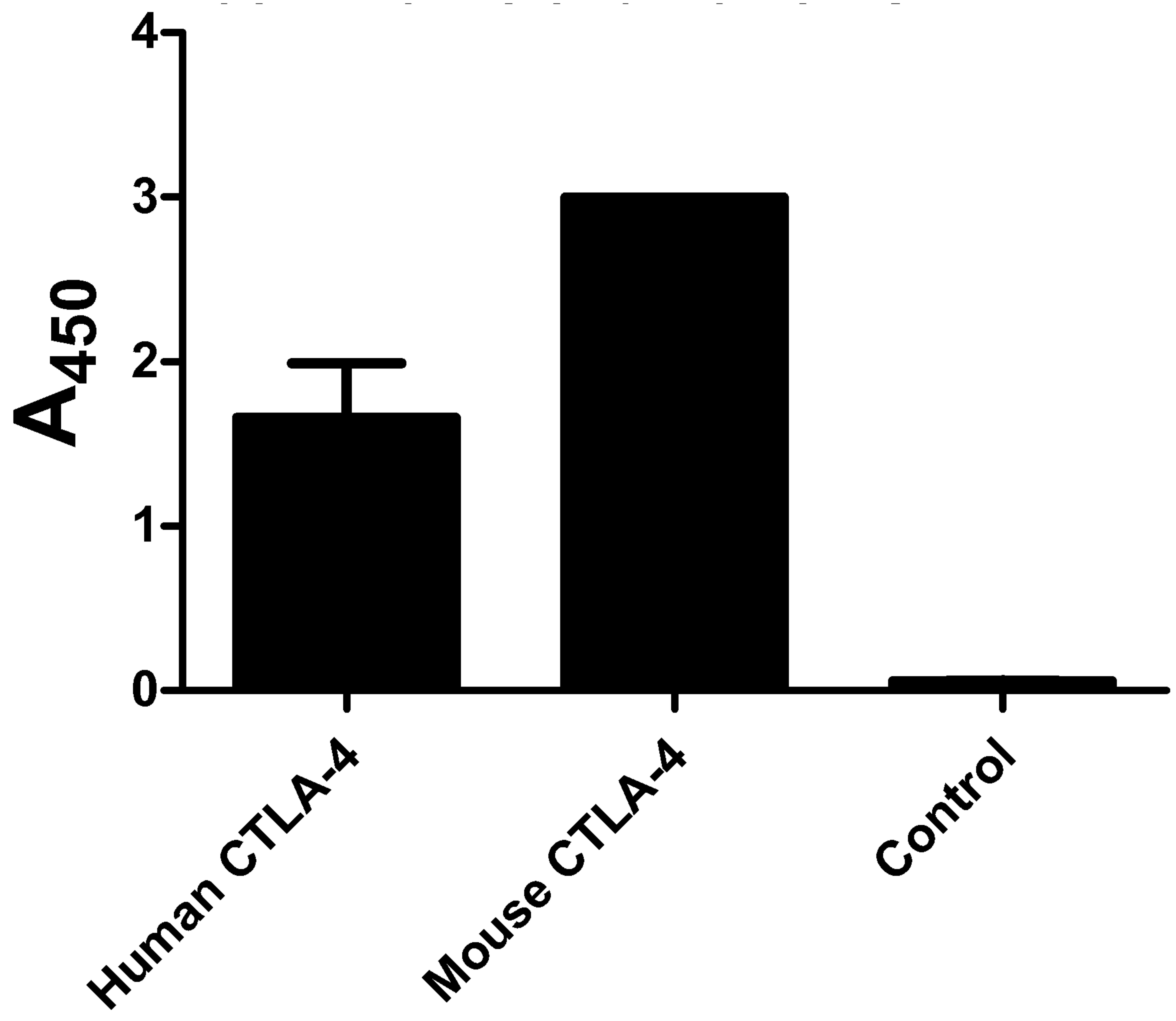
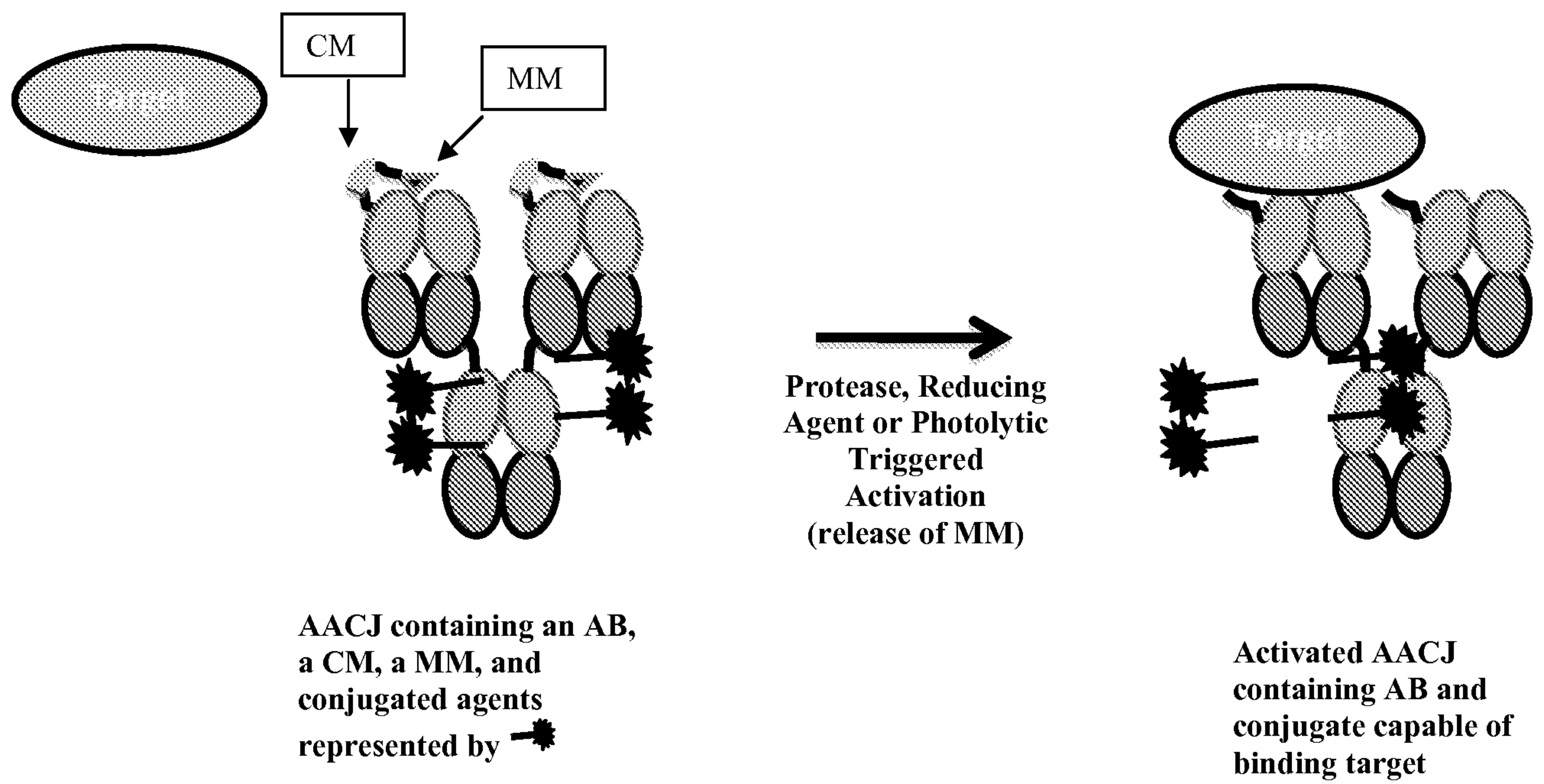
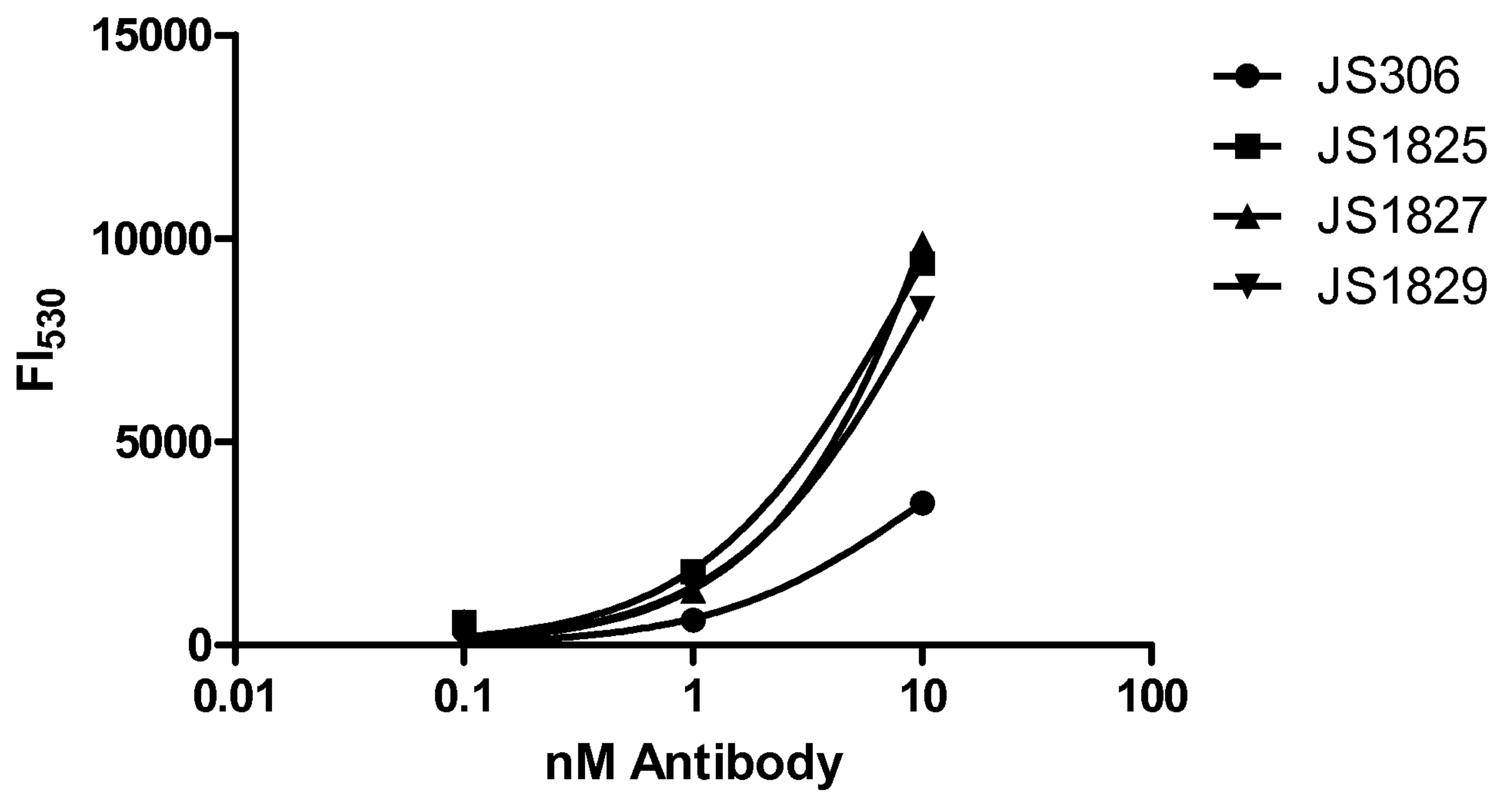
Figure 19

Figure 20

21/36
Figure 21



Fully
random
eCPX
library

Consensus
binding motif

XXXXXXXXXXXXXXXXXX

- 4 rounds of sorting
for binding to C225
- 1nM mAb in final round

CISPRGC GQPIPSVK
CISPRGC TQPYHVS
CISPRGC NAVSGLGS

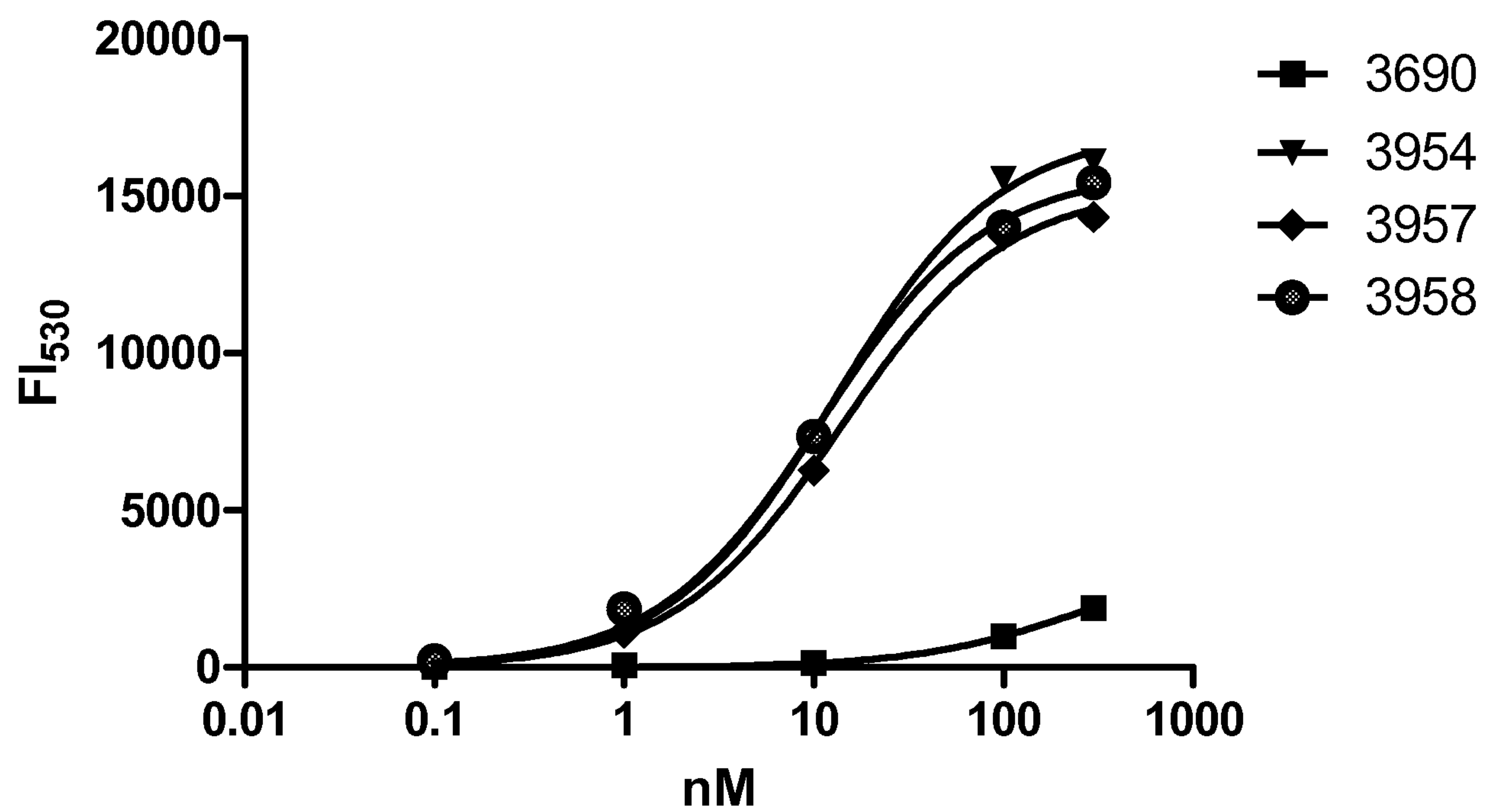
- Screen for KLK5/7
resistance

CISPRGC GQPIPSVK
GSHCLIPINMGAPS C

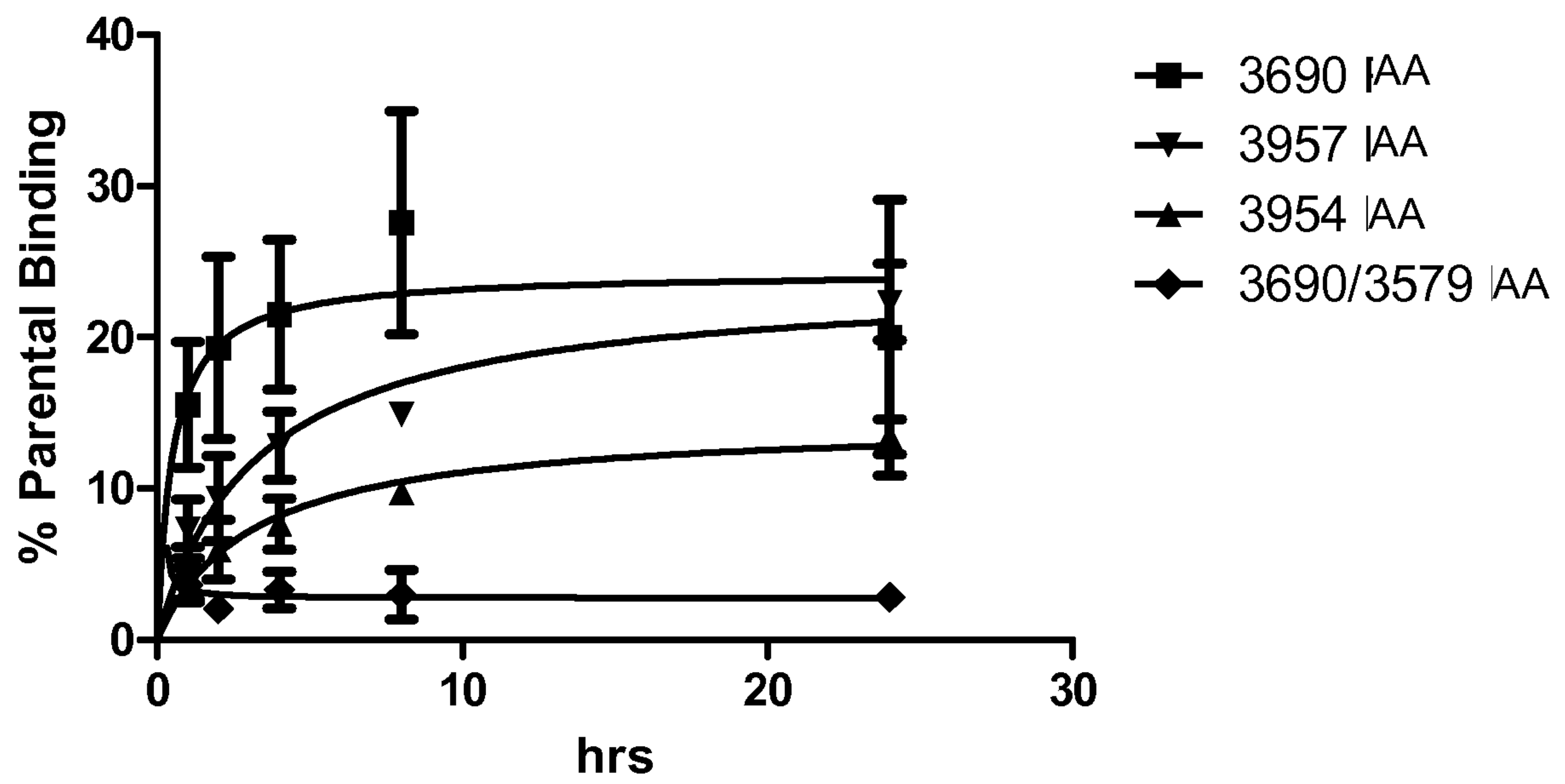
C225 binders

CISPRGC GQPIPSVK	8
CISPRGC TQPYHVS	5
CISPRGC NAVSGLGS	3
CPICFMRPAHEE	2
CPICGFQNNRA	2
CPMSSVRLCYEFNQE	6
TLTPENTRQWYLEKY	4
GSHCLIPINMGAPS C	3
CTPTLIRDGWLH CFS	2
WCRPTQSYEHI CPKE	2
LICDLYPTVNATR CK	2
HVHENEKEWCBNIRC	
PIYDYAFYQSDAYS	

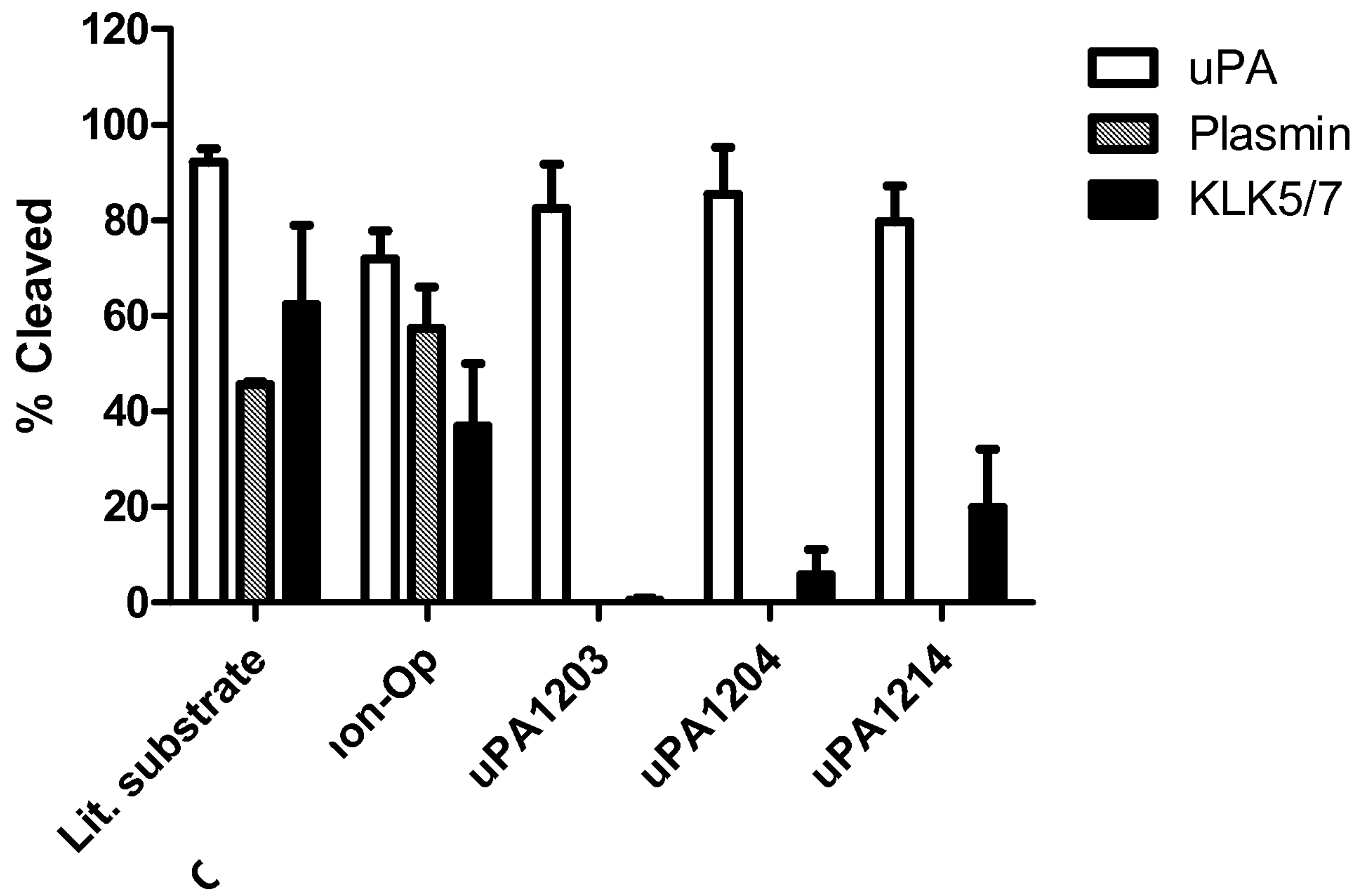
23/36
Figure 23



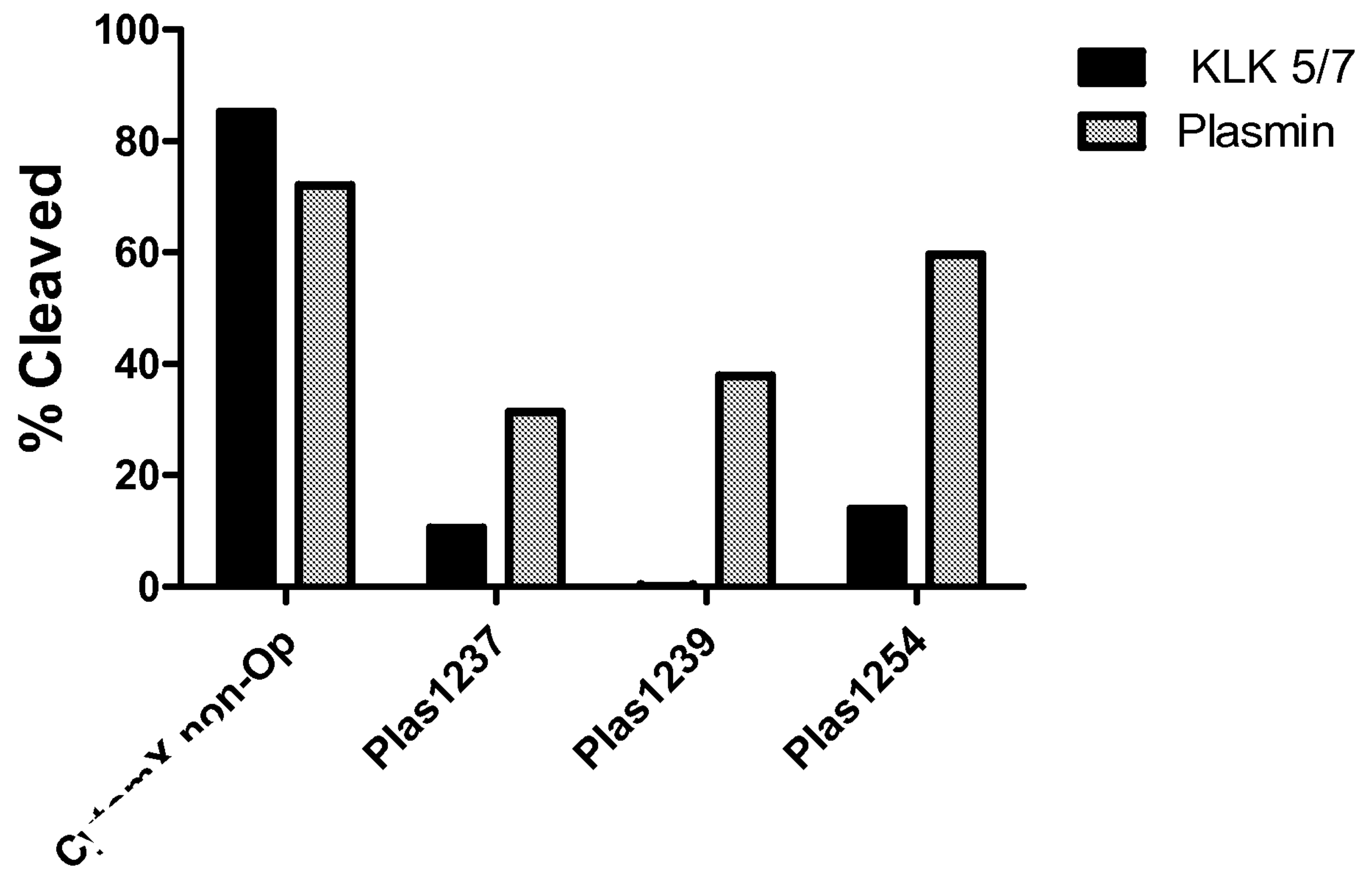
24/36
Figure 24



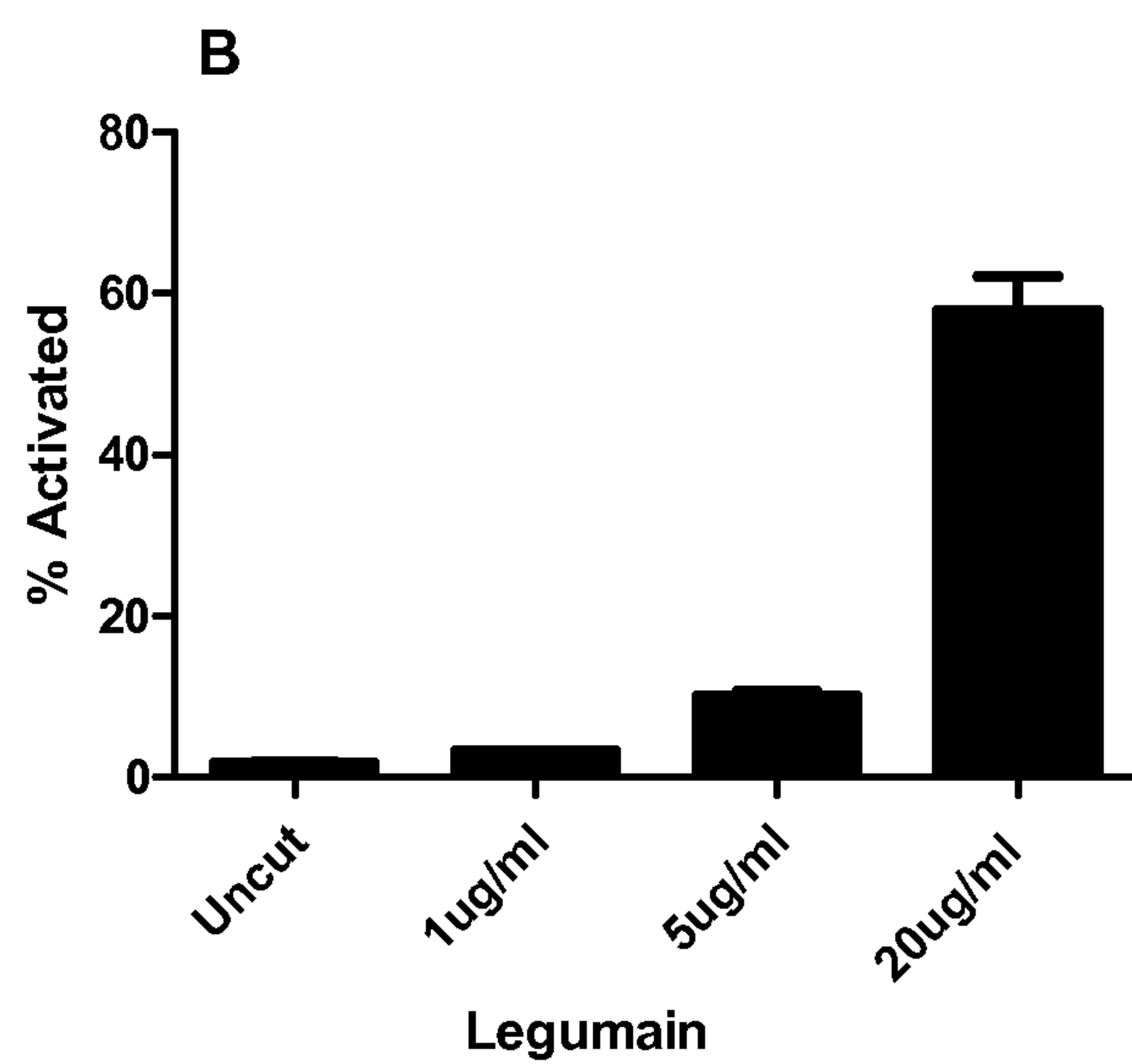
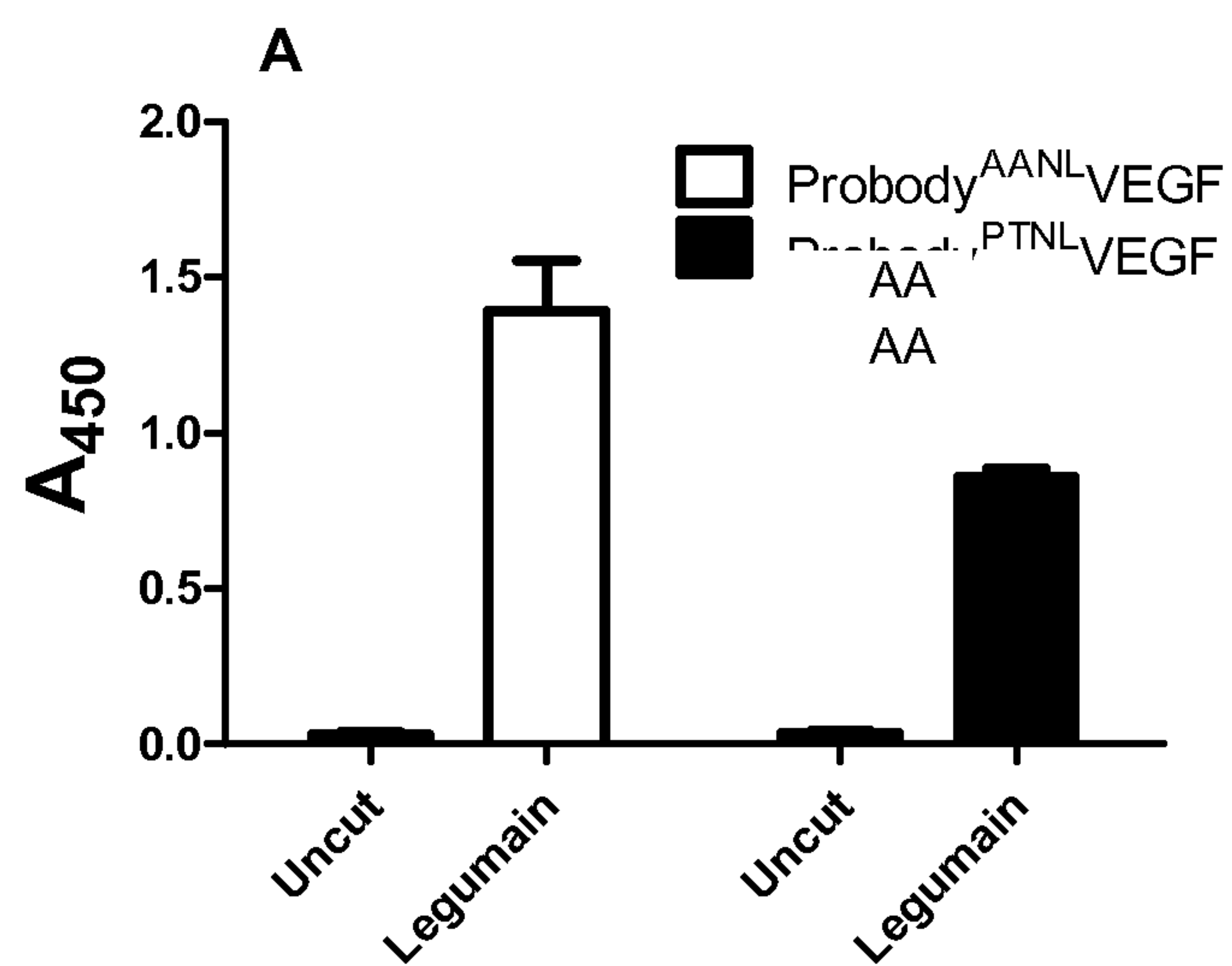
25/36
Figure 25



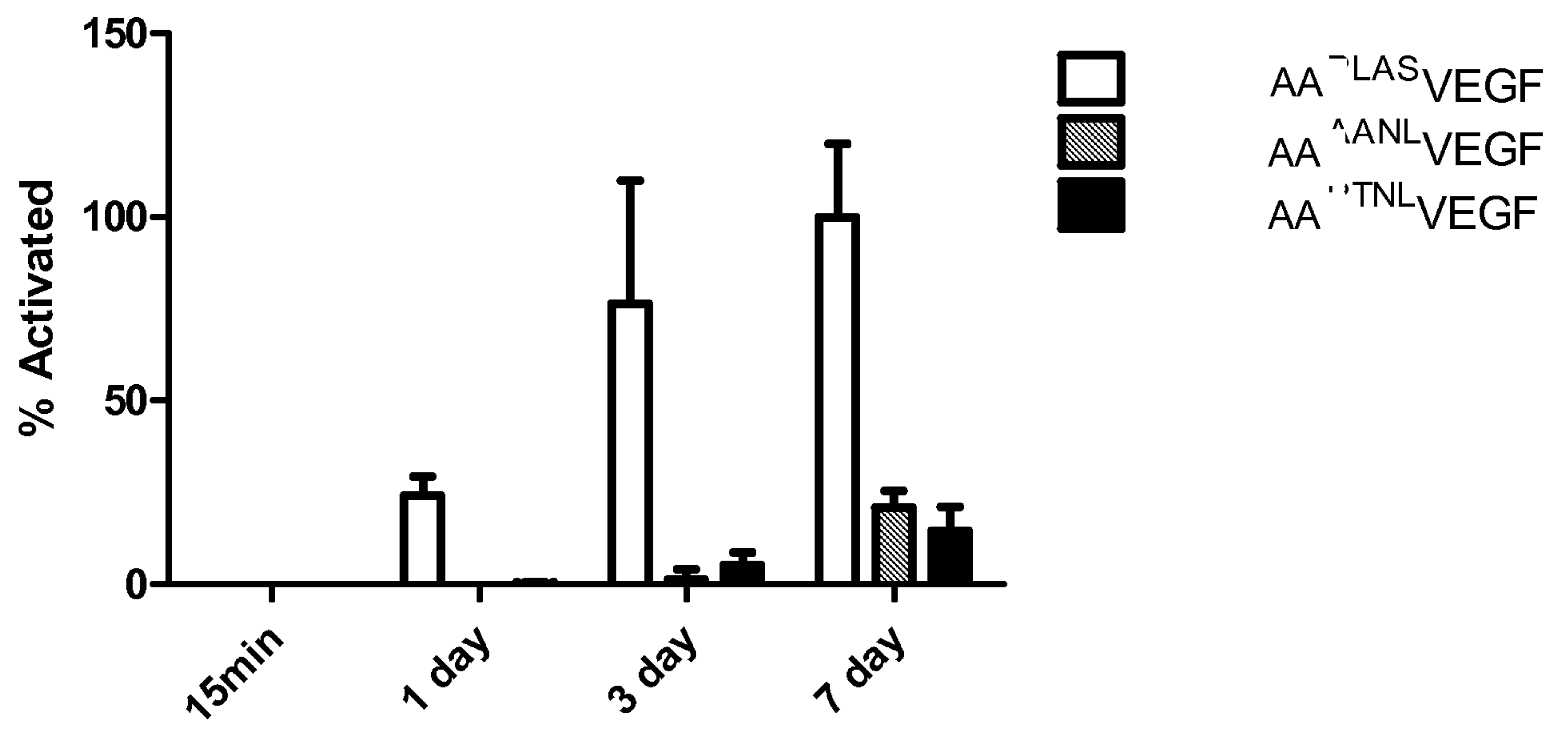
26/36
Figure 26



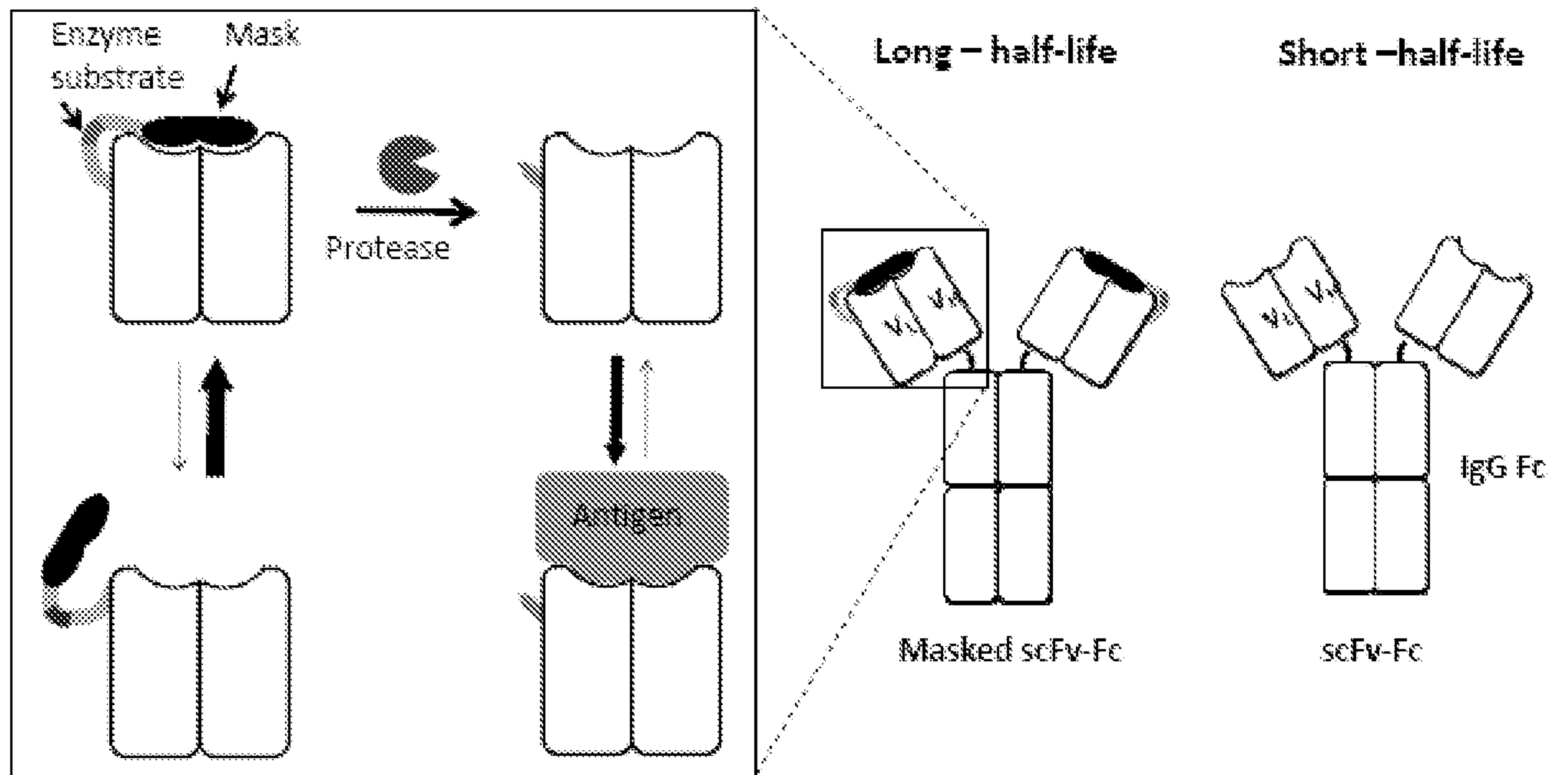
27/36

Figure 27

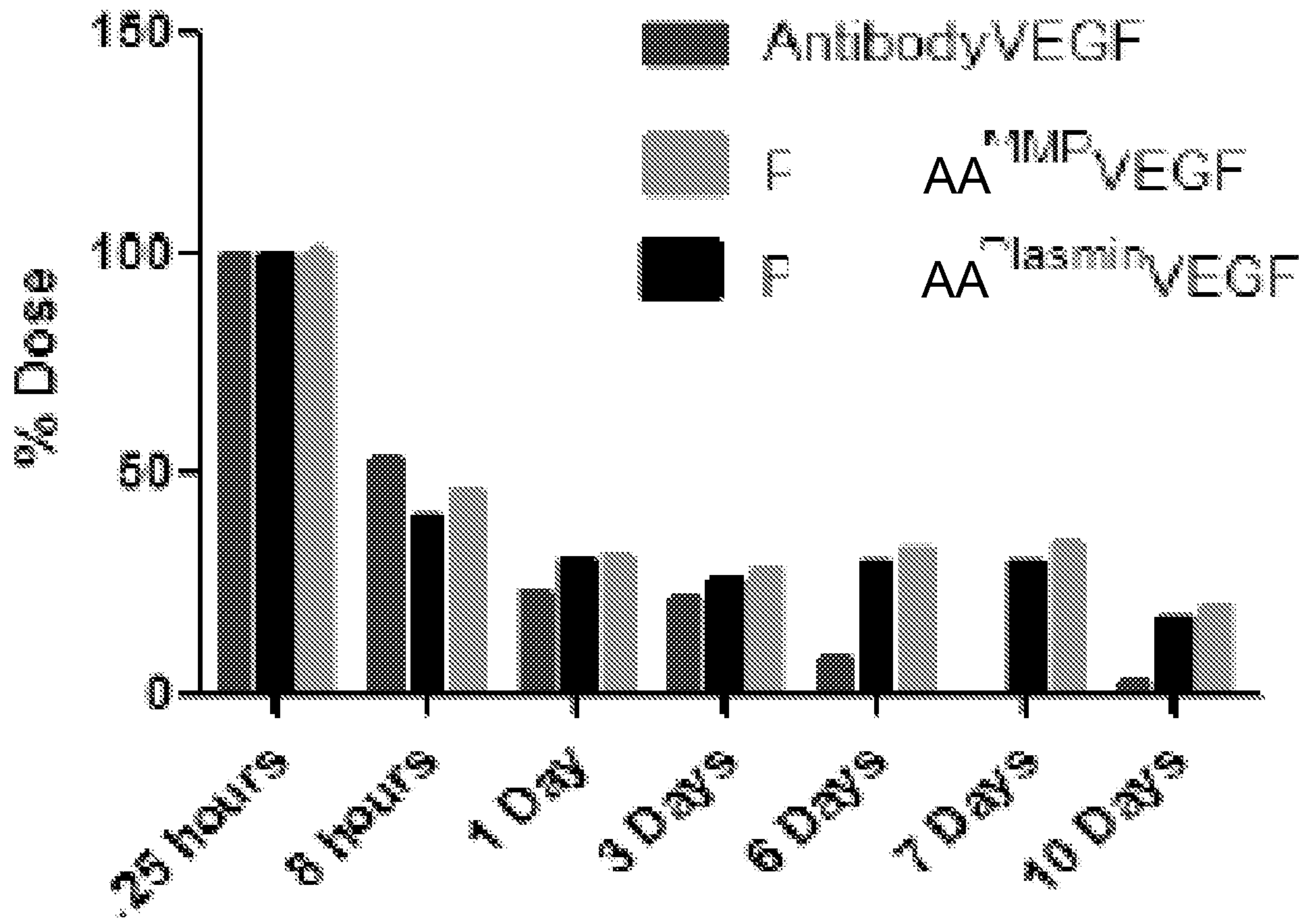
28/36
Figure 28

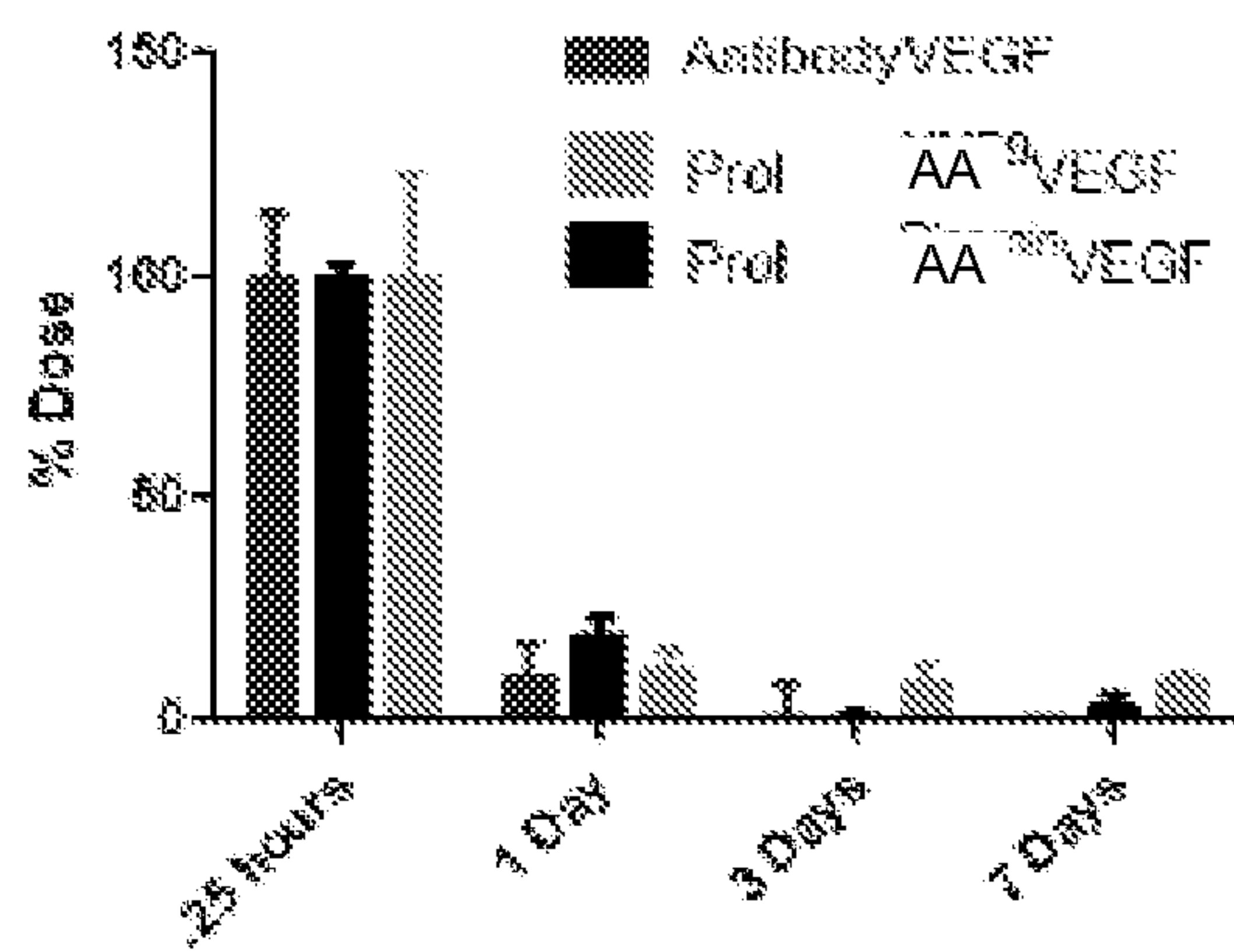
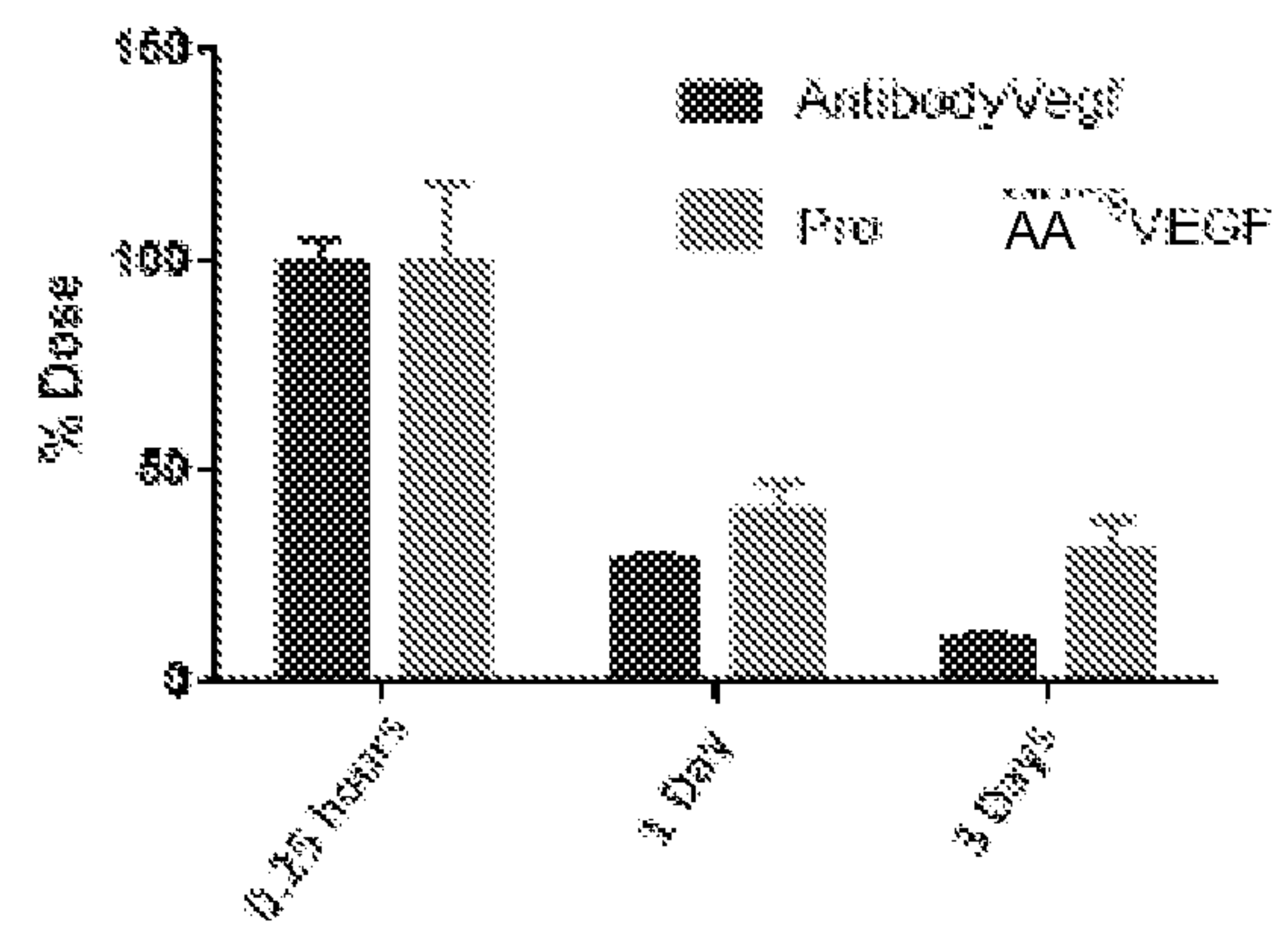


29/36
Figure 29

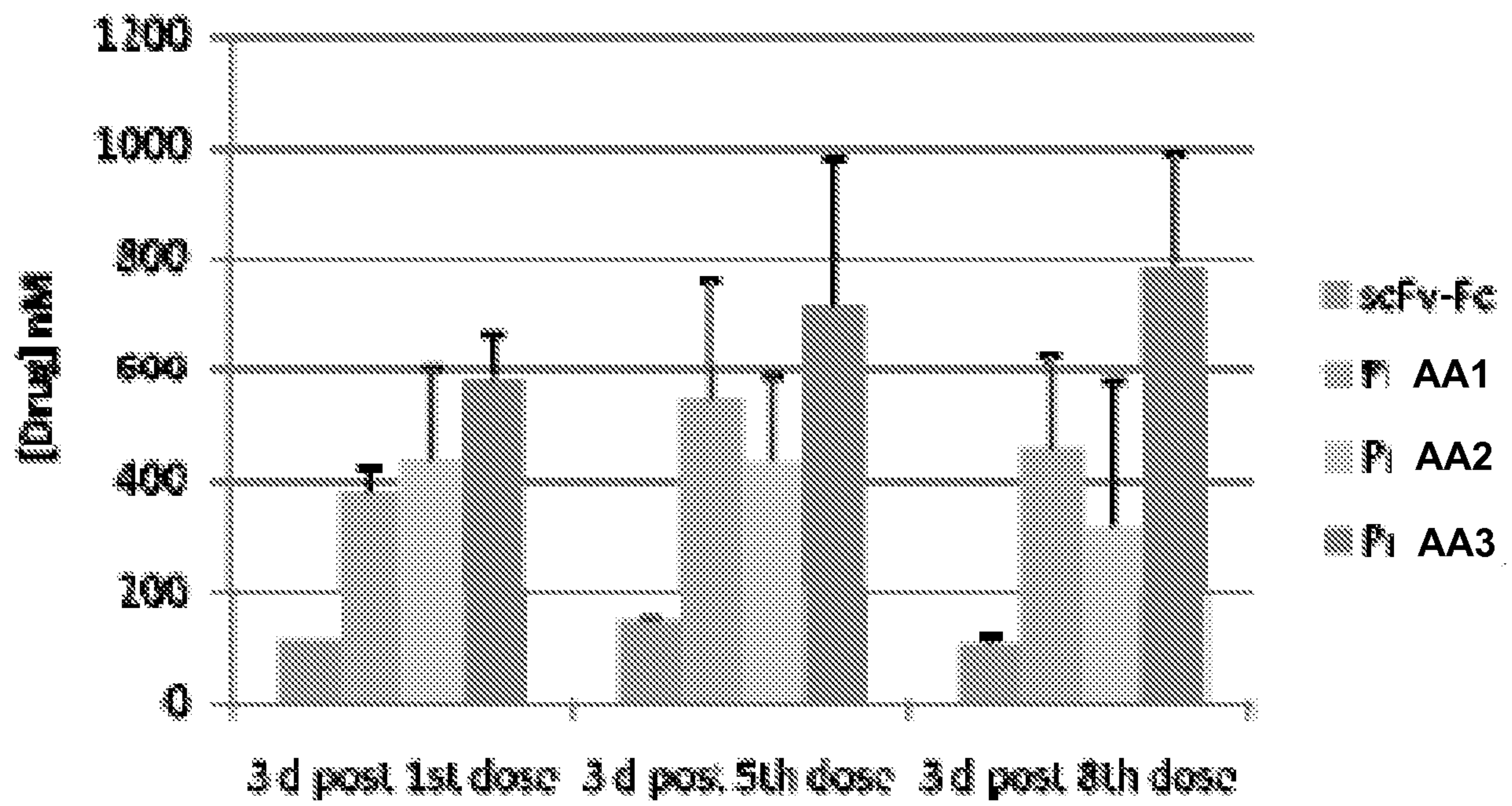


30/36

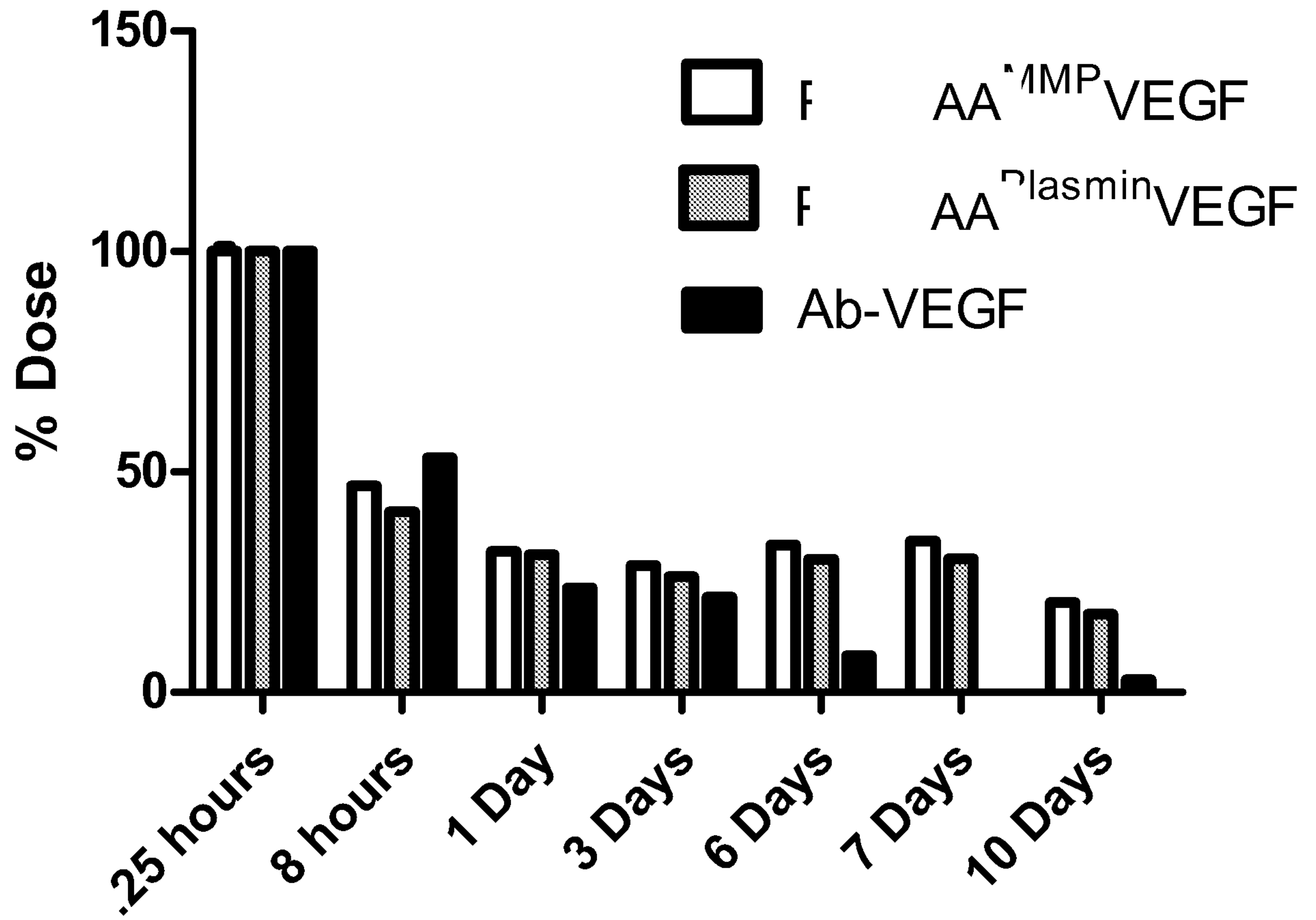
Figure 30

A Nude mice with HT-29 tumors**B** Nude mice with MDA-MB 231 tumors

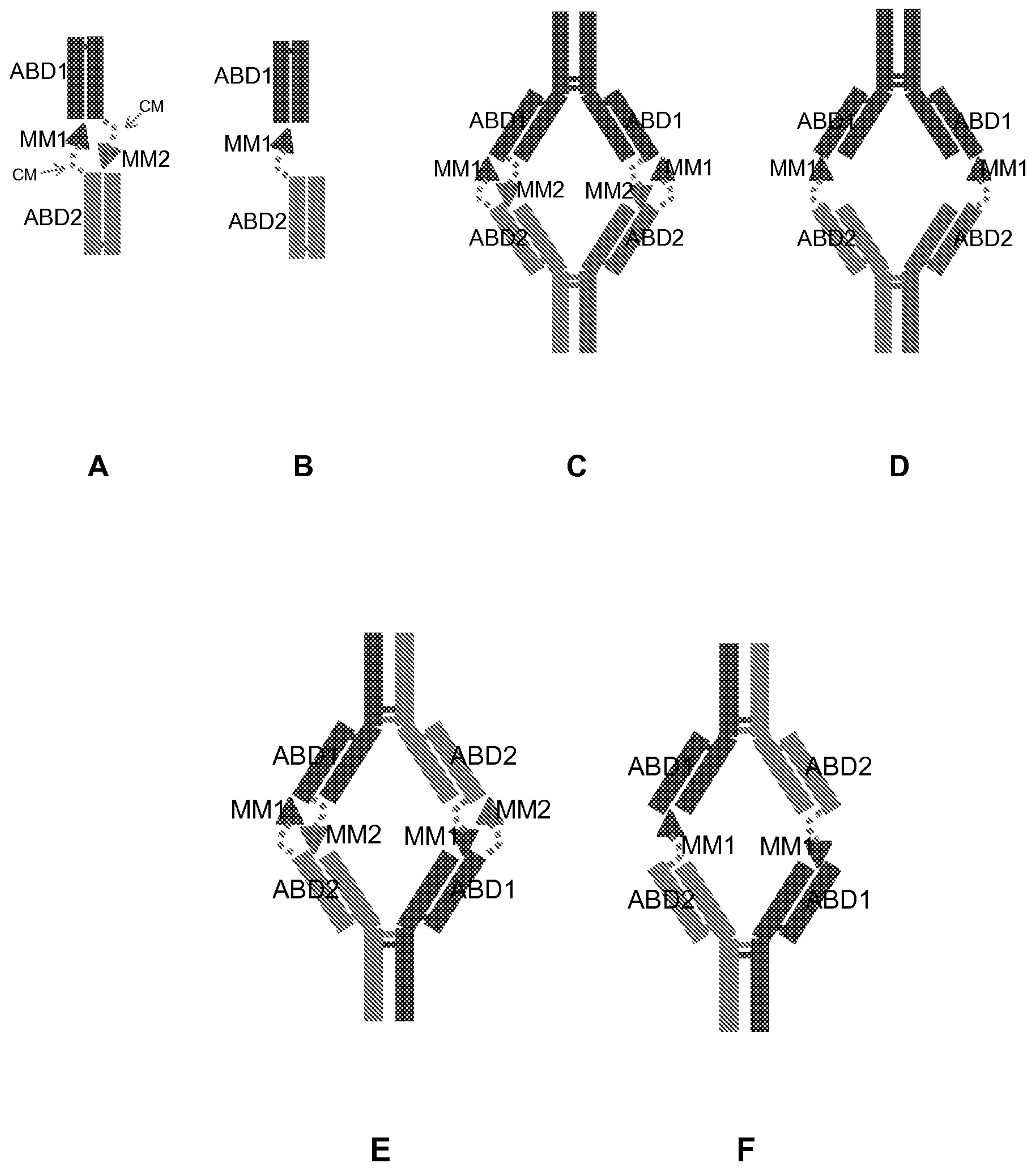
32/36

Figure 32

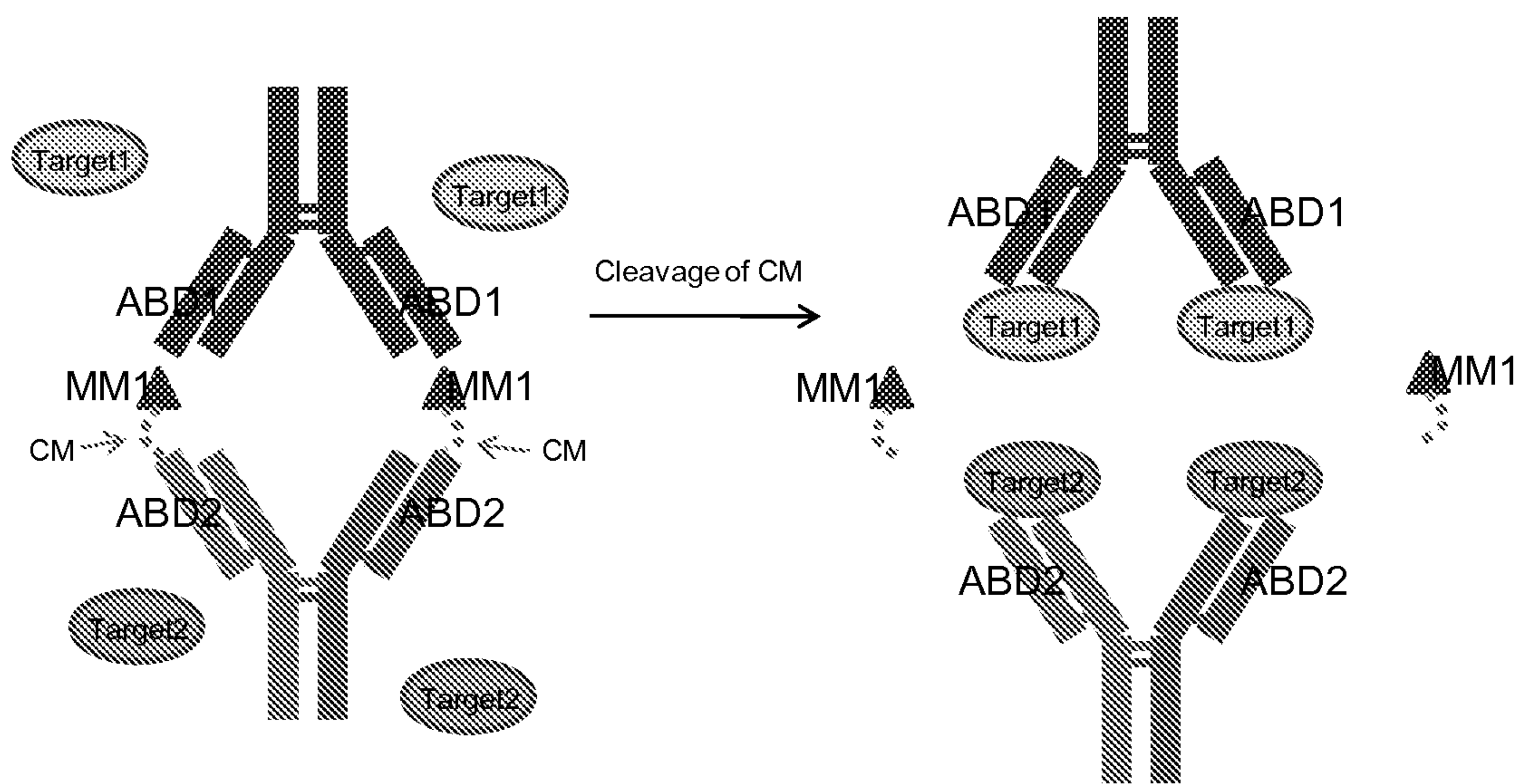
33/36

Figure 33

34/36
Figure 34



35/36
Figure 35



36/36
Figure 36

