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(54) Title: DEVELOPMENT OF A VECTOR TO TARGET GENE EXPRESSION TO THE EPIDERMIS OF TRANSGENIC ANIMALS

(57) Abstract

The keratin K1 vector for expression of a nucleic acid cassette in the epidermis. The vector has a 5′ flanking region of the keratin K1 gene including a 5′ flanking sequence, a keratin K1 promoter, 5′ transcribed but untranslated region and a first intron and an intron/exon boundary, all in sequential and positional relationship for expression of a nucleic acid cassette. A 3′ flanking region in the K1 keratin gene containing regulatory sequences including a 3′ transcribed but untranslated region and contiguous noncoding DNA containing the transcriptional termination region. The 3′ flanking region and a 5′ flanking region are linked by a polylinker having a plurality of restriction endonuclease sites for the insertion of a nucleic acid cassette. The nucleic acid cassette encodes a sequence which can be used to express any protein, polypeptide or antisense RNA. The vector can be inserted both in vivo and ex vivo into epidermal cells. Further, it can be used for making transgenic animals or bioreactors of epidermal cells. The vector has been found useful for gene therapy in treatment of a variety of diseases including skin ulcers, wound healing, surgical incisions, psoriasis and cancer.

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DEVELOPMENT OF A VECTOR TO TARGET GENE EXPRESSION TO THE EPIDERMIS OF TRANSGENIC ANIMALS

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FIELD OF THE INVENTION

The present invention relates to expression vectors for use in expressing polypeptides in epidermal cells of transgenic animals. More particularly it relates to vectors containing the K1 keratin gene promoter, its 5′ flanking region, its 5′ transcribed but untranslated region, its first intron and intron/exon boundary, its 3′ transcribed but untranslated region, its contiguous non-coding DNA containing the gene's natural transcriptional termination region and its 3′ flanking region.

BACKGROUND OF THE INVENTION

The ability to stably introduce genes into the germline of mice has greatly enhanced prospects for the generation of animal models of human diseases (Palmiter and Brinster, Ann. Rev. Genet., Vol. 20, pp. 465-499 (1986)). The need for such animal models is becoming increasingly apparent as novel pharmaceuticals are developed which are specifically

designed to inhibit expression of human viruses or counteract the effect of mutated genes that occur in human diseases. Current efficacy assessments of these new therapeutic agents are restricted to *in vitro* models which do not allow evaluation of delivery routes nor assessment of other factors known to affect disease processes *in vivo*, such as blood supply, an intact immune system, humoral and cell-mediated growth controls and physical barriers to disease progression. In addition, the prospects for utilizing gene therapy to treat human disorders are coming closer to reality. Therefore, animal models of human diseases would be useful to assess the therapeutic potential of these approaches. The epidermis is an attractive tissue for the development animal models since it serves as a general model for other squamous epithelia and its accessibility allows macroscopic observation of pathological events and easy assessment of therapeutic potential. The development of a vector which specifically targets gene expression to the epidermis of transgenic animals is the subject of this invention.

The epidermis is a continuously regenerating stratified squamous epithelium. Differentiated epidermal cells are the progeny of proliferative cells located in the basal cell layer and there is substantial evidence suggesting that the regeneration process occurs in proliferative units composed of slowly cycling, self-renewing stem cells, proliferative but non-renewing transit amplifying cells, and post-mitotic maturing epidermal cells (Iversen, et al., Cell Tissue Kinet., Vol. 1, pp. 351-367, (1968); MacKenzie, et al., Nature, Vol. 226, pp. 653-655, (1970); Christophers, et al., J. Invest. Dermatol., Vol. 56, pp. 165-170, (1971); Potten, In Stem Cells: Their Identification and Characterization, pp. 200-232, (1983); Cotsarelis, et al., Cell, Vol. 61, pp. 1329-1337, (1990)). The maturation process (terminal differentiation) is initiated when epidermal cells withdraw from the cell cycle and migrate from the basal layer into the spinous layer. Maturation continues as spinous cells migrate into the

granular layer and terminates with the formation of the stratum corneum. Morphological and biochemical studies have shown that terminal differentiation occurs in stages. (Matoltsy, J. Invest. Dermatol., Vol. 65, pp. 127-142, (1975)). Keratins K5 and K14 are major products of basal epidermal cells (Woodcock-Mitchell, et al., J. Cell Biol., Vol. 95, pp. 580-588, (1982)). These proteins assemble into 10 nm filaments (intermediate filaments [IF]) and, together with microtubules (tubulin) and microfilaments (actin), comprise the cytoskeleton of epidermal cells (Steinert, P.M., et al., Cell, Vol. 42, pp. 411-419, (1985)). One of the earliest changes associated with the commitment to differentiation and migration into the spinous layer is the induction of another differentiation-specific pair of keratins (K1 and K10). IF containing K1 and K10 replace those containing K5 and K14 as the major products of cells in the spinous layer (Woodcock-Mitchell, et al., J. Cell Biol., Vol. 95, pp. 580-588, (1982); Roop, et al., Proc. Natl. Acad. Sci., USA, Vol. 80, pp. 716-720, (1983); Schweizer, et al, Cell, Vol. 37, pp. 159-170, (1984)). The keratin IF formed by these proteins assemble into bundles. In the granular layer, another high molecular weight non-IF protein is synthesized, which is processed into filaggrin, and is thought to promote keratin filament aggregation and disulfide-bond formation (Dale, B.A., et al., Nature, Vol. 276, pp. 729-731, (1978); Harding, C.R., et al., J. Mol. Biol., Vol. 170, pp. 651-673, (1983)). In the final stage of epidermal cell maturation, transglutaminase catalyzes the crosslinking of involucrin and loricrin, by the formation of (γ-glutamyl) lysine isopeptides, into a highly insoluble cornified envelope which is located just beneath the plasma membrane (Rice and Green, Cell Vol. II, pp. 417-422 (1977) Mehrel, et al., Cell, Vol. 61, pp. 1103-1112, (1990)).

Genes or cDNAs encoding the major keratins expressed in epidermal cells have now been cloned: K5 (Lersch, et al., Mol. and Cell Biol., Vol. 8, pp. 486-493, (1988), K14 (Marchuk, et al., Proc. Natl. Acad.

Sci, USA, Vol. 82, pp. 1609-1613, (1985); Knapp, et al., J. Biol. Chem, Vol. 262, pp. 938-945, (1987); Roop, et al., Cancer Res., Vol. 48, pp. 3245-3252, (1988), K1 (Steinert, et al., J. Biol. Chem., Vol. 260, pp. 7142-7149, (1985) and K10 (Krieg, et al., J. Biol. Chem., Vol. 260, pp. 5867-5870, (1985)). Northern blot analysis and *in situ* hybridization studies suggest that keratin genes K5 and K14 are predominantly transcribed in the proliferating basal layer and transcription of keratin genes K1 and K10 is induced as cells migrate into the spinous layer (Lersch, et al., Mol. and Cell Biol., Vol. 8, pp. 486-493, (1988); Knapp, et al., J. Biol. Chem., Vol. 262, pp. 938-945, (1987); Roop, et al., Cancer Res., Vol. 48, pp. 3245-3252, (1988)). Genes encoding rat (Haydock, et al., J. Biol. Chem., Vol. 261, pp. 12520-12525, (1986)) and mouse (Rothnagel, et al., J. Biol. Chem., Vol. 262, pp. 15643-15648, (1987)) filaggrin have now been identified and *in situ* hybridization experiments have confirmed that transcription of this gene is restricted to the granular layer (Rothnagel, et al, J. Biol. Chem., Vol. 262, pp. 15643-15648, (1987); Fisher, et al. J. Invest. Dermatol., Vol. 88, pp. 661-664, (1987)). To date, loricrin is the only gene encoding a component of the cornified envelope to be studied at the molecular level by *in situ* hybridization and transcripts of this gene are restricted to the granular layer (Mehrel, et al., Cell, Vol. 61, pp. 1103-1112, (1990)).

From this description of gene expression in the epidermis, there would appear to be many candidate genes from which to choose for targeting to the epidermis. However, this is not the case. Keratins K5 and K14, expressed in the proliferative compartment of the epidermis, are not only expressed in the epidermis but in all squamous epithelia. Furthermore, these genes are expressed early in development (Dale and Holbrook, In: Current Topics in Developmental Biology, pp. 127-151, (1987)) and this could cause lethality *in utero*. The generation of animal models of hyperproliferative diseases such as cancer and psoriasis would most likely require expression in the basal compartment in cells with

proliferative potential, therefore, genes expressed post-mitotically such as keratins K1 and K10 and those encoding filaggrin and components of the cell envelope (involucrin and loricrin) would be excluded on this basis. The present invention, however, demonstrates that a 12 kb fragment of the human keratin gene K1 (HK1) contains sequences regulating tissue and developmental specific expression in transgenic mice. This fragment lacks sequences responsive to negative control of differentiation specific expression resulting in expression of the HK1 gene in some cells of the basal cell compartment of the epidermis. Although regulatory elements of the HK1 gene fail to completely mimic the expression pattern of the endogenous mouse K1 gene, they are ideally suited for targeting gene expression for the following reasons: (1) expression only occurs in the epidermis and not other squamous epithelia; (2) expression occurs at a late stage of development (day 15) and, therefore, is unlikely to result in lethality *it utero*; (3) expression occurs in a large proportion of basal cells that have proliferative potential.

SUMMARY OF THE INVENTION

An object of the present invention is a keratin K1 vector for expressing nucleic acid sequences in the epidermis.

An additional object of the present invention is a keratin K1 vector containing an oncogene.

A further object of the present invention is a bioreactor for producing proteins, polypeptides and antisense RNA in transduced epidermal cells.

An additional object of the present invention is an *in vivo* method of transducing epidermal cells with a keratin K1 vector.

A further object of the present invention is provision of a transgenic animal containing the keratin K1 epidermal vector.

An additional object of the present invention is the provision of a transgenic animal for the study of cancer.

Another object of the present invention is a method of treating skin ulcers.

A further object of the present invention is an enhanced method of wound healing or healing of surgical incisions.

An additional object of the present invention is a method of treating psoriasis.

An additional object of the present invention is a method of treating skin cancer.

An additional object of the present invention is a vaccination procedure using the keratin K1 epidermal vector.

Thus, in accomplishing the foregoing objects, there is provided in accordance with one aspect of the present invention a keratin K1 vector for expression of a nucleic acid cassette in the epidermis comprising: a 5′ flanking region of the keratin K1 gene, said 5′ flanking sequence including a keratin K1 promoter, a 5′ transcribed but untranslated region and a first intron and an intron/exon boundary all in sequential and positional relationship for expression of a nucleic acid cassette; a 3′ flanking region of the keratin K1 gene containing regulatory sequences, said 3′ flanking region including a 3′ transcribed but untranslated region and contiguous noncoding DNA containing a transcriptional termination region; and a polylinker having a plurality of restriction endonuclease sites, said polylinker connecting the 5′ flanking region to the 3′ flanking region and said polylinker further providing a position for insertion of the nucleic acid cassette.

In specific embodiments of the present invention the keratin K1 vector has a 5′ flanking region of approximately 1.2 kb and a 3′ flanking region of approximately 3.9 kb.

In alternative embodiments of the present invention there is a further addition of approximately 8.0 kb of 5' flanking sequence from the 18 kb Eco RV fragment onto the end of the vector.

In another alternative embodiment the Vitamin D_3 regulatory element within the human K1 keratin gene is identified and utilized to suppress expression of the keratin K1 vector.

In the specific embodiments of the present invention the keratin K1 vector is used to transduce epidermal cells to form bioreactors. The bioreactors produce a variety of proteins, polypeptides and RNAs. Additionally, the vector can be used to form transgenic animals. The transgenic animals can be used to study cancer, drug reactions and treatments.

The keratin K1 vector can also be used for the treatment of a variety of diseases, including wounds, surgical incisions, psoriasis, skin ulcers and skin cancer and can be used for the production of vaccines.

Other and further objects, features and advantages will be apparent from the following description of the presently preferred embodiments of the invention which are given for the purposes of disclosure when taken in conjunction with the accompanying drawings.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a schematic drawing of the human keratin K1 gene (HK1) and the expression vector derived from its regulatory sequences.

Figure 2 shows the expression characteristics of the HK1 vector *in vivo* in transgenic mice utilizing a reporter gene encoding E. coli β-galactosidase.

Figure 3 demonstrates the suppression of the SV40 promoter by a novel negative regulatory element from the HK1 gene (HK1.NRE) in the presence of Vitamin D_3.

Figure 4 is a schematic drawing of the HK1 vector containing the coding sequence of v-ras^{Ha} protein of Harvey Murine Sarcoma Virus.

Figure 5 is a schematic drawing of the HK1 vector containing the coding sequence of the v-fos protein from a FBJ/FBR chimeric plasmid.

Figure 6 is a schematic drawing of the HK1 expression vector containing the coding sequences of the E6 and E7 proteins from human papilloma virus 18.

Figure 7 is a schematic drawing of the HK1 vector containing the coding sequence of TGF-α.

Figure 8 is a schematic drawing of the HK1 vector containing the coding sequence of the trans-regulatory protein *tat*, from human immunodeficiency virus.

Figure 9 is a schematic drawing of an 18kb Eco RV fragment containing the HK1 gene.

Figure 10 is a schematic drawing of a derivative of the HK1 vector containing additional 5′ flanking sequences which restrict expression to differentiated epidermal cells.

The drawings are not necessarily to scale, and certain features of the invention may be exaggerated in scale and shown in schematic form in the interest of clarity and conciseness.

DETAILED DESCRIPTION OF THE INVENTION

It will be readily apparent to one skilled in the art that varying substitutions and modifications may be made to the invention disclosed herein without departing from the scope and spirit of the invention.

The term "transformed" as used herein refers to the process or mechanism of inducing changes in the characteristics (expressed phenotype) of a cell by the mechanism of gene transfer whereby DNA is introduced into a cell in a form where it expresses a specific gene product or alters expression of endogenous gene products.

The term "transduction" as used herein refers to the process of introducing a DNA expression vector into a cell. Various methods of

transduction are possible, including microinjection, $CaPO_4$, lipofection (lysosome fusion), use of a gene gun and DNA vector transporter. The human keratin K1 vector can be transduced into the epidermal cells by any of the variety of ways described above.

The term "DNA vector transporter" as used herein refers to those molecules which bind to DNA vectors and are capable of being taken up by epidermal cells. DNA transporter is a molecular complex capable of non-covalent binding to DNA and efficiently transporting the DNA through the cell membrane. Although not necessary, it is preferable that the transporter also transport the DNA through the nuclear membrane.

The term "transient" as used in transient transfection, transient transduction or transiently transformed relates to the introduction of genes into the epidermal cells to express specific proteins, polypeptides and RNA wherein the introduced genes are not integrated into the host cell genome and accordingly are eliminated from the cell over a period of time. Transient expression relates to the expression of gene products during the period of transient transfection. Additionally, transient can refer to a stable transfection or transduction into cells, where the cells die and are sloughed off from the skin. Thus, the transformed cells are only transiently available for the expression of the incorporated genes.

The term "stable" as used in stable transfection, stable transduction or stably transformed refers to the introduction of genes into the chromosome of the targeted cell where it integrates and becomes a permanent component of the genetic material in that cell. Gene expression after stable transduction can permanently alter the characteristics of the cell leading to stable transformation. An episomal transformation is a variant of stable transformation in which the introduced gene is not incorporated into the host cell chromosomes but rather is replicated as an extrachromosomal element. This can lead to apparently stable transformation of the characteristics of the cell. As

indicated above, in epidermal cells, which are sloughed from the body through the skin, stable transformation can become transient transformation because the cells are lost.

The term "nucleic acid cassette" as used herein refers to the genetic material of interest which can express a protein, polypeptide or RNA and which is capable of being incorporated into the epidermal cells. The nucleic acid cassette is positionally and sequentially oriented within the keratin K1 vector such that the nucleic acid in the cassette can be transcribed into RNA or antisense RNA and, when necessary, translated into proteins or polypeptides in the transformed epidermal cells. A variety of proteins and polypeptides can be expressed in the transformed epidermal cells by the sequence in the nucleic acid cassette. These proteins or polypeptides which can be expressed include hormones, growth factors, enzymes, clotting factors, apolipoproteins, receptors, drugs, tumor antigens, viral antigens, parasitic antigens, bacterial antigens and oncogenes. Specific examples of these compounds include proinsulin, insulin, growth hormone, insulin-like growth factor I, insulin-like growth factor II, insulin growth factor binding protein, epidermal growth factor TGF-α, dermal growth factor PDGF, angiogenesis factor, for instance, acid fibroblast and basic fibroblast growth factors and angiogenin, matrix protein, such as, Type IV collagen, Type VII collagen, laminin, nidogen and proteins from viral, bacterial and parasitic organisms which can be used to induce an immunologic response.

In addition, the nucleic acid cassette can encode a "transforming gene" which encompasses viral oncogenes, endogenous proto-oncogenes and activated proto-oncogenes. A variety of oncogenes are known in the art. The term "oncogene" means those genes which cause cancer and include both viral and cellular oncogenes, many of which are homologous to DNA sequences endogenous to rodents and/or humans. The term

oncogene includes both the viral sequence and the homologous endogenous sequences. Some examples of transforming genes are listed in Table 1.

Table 1.

Transforming Genes

ABBREVIATION	NAME
Ha-*ras*	Harvey Murine Sarcoma Virus
Ki-*ras*	Kirsten Murine Sarcoma Virus
N-*ras*	Neuroblastoma oncogene
fos	FBJ or FBR osteosarcoma virus
myc	Avian MC29 myelocytomatosis virus
src	Rous sarcoma virus
sis	Simian sarcoma virus/PDGF β chain
*erb*A	Avian erythroblastosis virus/Thyroxine T3 receptor
*erb*B	Avian erythroblastosis virus/Truncated EGF receptor
jun	Avian sarcoma virus 17
p Large T	Polyomavirus transforming gene
p Middle T	Polyomavirus transforming gene
HPV E7	Early region transforming gene from human papillomavirus 6, 11, 16, 18
HPV E6	Early region transforming gene from human papilloma virus 6, 11, 16, 18

HPV E5	Early region transforming gene from human papilloma virus 6, 11, 16, 18
tat	HIV transforming gene
EIA	Adenovirus early region 1A
Rb	Mutated retinoblastoma gene
p53	Mutated p53 anti-oncogene
WT1	Mutated Wilms tumor anti-oncogene
TGF-α	Transforming growth factor α
TGF-β	Transforming growth factor β
EGFR	Mutated epidermal growth factor receptor
RAR	Mutated retinoic acid receptor
VD_3R	Mutated vitamin D_3 receptor
PKC	Mutated protein kinase C

The genetic material which is incorporated into the epidermal cells using the keratin K1 vector includes DNA not normally found in epidermal cells, DNA which is normally found in epidermal cells but not expressed at physiologically significant levels, DNA normally found in epidermal cells and normally expressed at physiological desired levels, any other DNA which can be modified for expression in epidermal cells, and any combination of the above.

The term "keratin K1 vector" or "HK1 vector" as used herein is a vector which is useful for expression of a nucleic acid sequence in epidermal cells. The keratin K1 vector comprises a 5′ flanking region of

the keratin K1 gene, said flanking region including a promoter, a first intron and an intron/exon boundary all in sequential and positional relationship for the expression of a nucleic acid cassette; a 3′ flanking sequence of a keratin K1 gene; and a poly-linker. The poly-linker includes a plurality of restriction endonuclease sites. The polylinker connects the 5′ flanking region to the 3′ flanking sequence and further provides a position for insertion of the nucleic acid cassette.

The sequence for the 3′ flanking region of the human keratin K1 gene contains regulator elements and is used for preparing the keratin K1 vector. It is shown in SEQ. ID No. 1. The keratin K1 vector has a 5′ flanking region comprising nucleotides 1 to 1246 of SEQ. ID. No. 1; a 3′ flanking sequence containing regulatory sequences comprises nucleotides 6891 to 10747 of SEQ. ID. No. 1; and a poly linker comprising nucleotides 2351 to 2376 of SEQ. ID. No. 2 (the HK1 expression vector).

The keratin K1 vector has a 5′ flanking region of approximately 1.2 kb, an intron and intron/exon boundary of approximately 1.0 kb and a 3′ flanking sequence of approximately 3.9 kb.

The restriction endonuclease sites found in the linker and poly-linker of the keratin K1 vector can be any restriction endonucleases which will allow insertion of the nucleic acid cassette. In the preferred embodiment they are usually selected from the group consisting of Bam HI, Kpn I, Cla I, Not I, Xma I, and Bgl II.

One skilled in the art will readily recognize that there are a variety of ways to introduce the keratin K1 vector into epidermal cells. The vectors can be inserted either *in vivo* or *ex vivo*. The mode of insertion will, to a certain degree, determine the available methods for the insertion. The *in vivo* insertion is preferred for gene therapy. In this procedure the human keratin K1 vector is contacted with epidermal cells for sufficient time to transform the epidermal cells.

One embodiment of the present invention includes a bioreactor. A bioreactor is comprised of transformed epidermal cells which contain the keratin K1 vector. Once the vector is inserted in the epidermal cells, the epidermal cells will express the nucleic cassette and produce the protein, polypeptide or antisense RNA of interest. This can be done either *in vivo* or *ex vivo*. Any compound which can be encoded in, and expressed by, the nucleic acid cassette can be produced by the bioreactor.

One method for *ex vivo* introduction of the keratin K1 vector into epidermal cells includes a cotransfection of the vector with a selectable marker. The selectable marker is used to select those cells which have become transformed. The cells can then be used in any of the methods described in the present invention.

Another embodiment of the present invention is a method of making transgenic animals comprising the steps of inserting the human keratin K1 vector into the embryo of the animal. The transgenic animal can include the resulting animal in which the vector has been inserted into the embryo or any progeny. The term progeny as used herein includes direct progeny of the transgenic animal as well as any progeny of succeeding progeny. Thus, one skilled in the art will readily recognize that if two different transgenic animals have been made using different genes in the nucleic acid cassette and they are mated, the possibility exists that some of the resulting progeny will contain two or more introduced sequences. One skilled in the art will readily recognize that by controlling the matings, transgenic animals with multiple vectors can be made.

In the transgenic animals that contain the human keratin K1 vector in its germ and somatic cells, the nucleic acid cassette of the said vector is only expressed in the epidermal cells. This is a distinct advantage over other transgenic animal models where there is not as much control over the expression of the sequence in the tissues.

In the preferred embodiment, the transgenic animal will contain an oncogene sequence in the nucleic acid cassette. Preferably the animal is a rodent. The transgenic animal can be used in any method for studying a variety of diseases including the origin of cancer, the treatment of cancer, interaction of the cancer with the environment as well as for looking at drugs, pharmaceuticals and other chemical interactions. The transgenic animals are useful in any assay in which the skin cells of the animal can be used.

One specific embodiment of the present invention is a method for the enhanced healing of a wound or surgical incision. This method comprises the *in vivo* transduction of epidermal cells with a keratin K1 vector. The nucleic acid cassette of said vector contains a nucleic acid sequence for a growth factor.

In the preferred embodiment for the treatment of wounds or surgical incisions, a plurality of vectors are introduced into the epidermal cells. In the plurality of vectors, the cassette of at least one vector contains a nucleic acid sequence for an epidermal growth factor (TGF-α), the cassette of at least one vector contains a dermal growth factor (PDGF), a cassette of at least one vector contains a nucleic acid sequence for a matrix protein to anchor the epidermis to the dermis, and a cassette of at least one vector contains a nucleic acid sequence for an angiogenesis factor. The sequence for matrix proteins can be selected from any sequences useful for the anchoring of the epidermis to the dermis but are usually selected from the group consisting of Type IV collagen, laminin, nidogen, and Type VII collagen. The angiogenesis factor is usually selected from the group consisting of acid fibroblast and basic fibroblast growth factors, and angiogenin. The combination of the vectors provides all of the necessary elements for quick and rapid enhancement of healing of wounds or surgical incisions. This procedure is very helpful in the case of plastic or reconstructive surgery. Furthermore, skin ulcers can be

treated by following similar procedures as described for wound healing or surgical incision. These procedures for healing of wounds, surgical incisions and skin ulcers are useful in animals and humans.

In the *ex vivo* approach for treating or healing wounds, surgical incisions and skin lesions, the vectors are first transduced into the epidermal cells *ex vivo*. The transformed epidermal cells are transplanted onto the animal or human to be treated.

Another embodiment of the present invention is a method for treating psoriasis. In this method, epidermal cells are transduced in *in vivo* with a keratin K1 vector. A nucleic acid cassette in said vector contains a nucleic acid sequence for a protein or polypeptide selected from the group consisting of TGF-β, a soluble form of cytokine receptor, and an antisense RNA. The cytokine receptor can be selected from the group consisting of IL-1, IL-6, and IL-8. The antisense RNA sequence is selected from the group consisting of TGF-α, IL-1, IL-6, and IL-8.

In another embodiment of the present invention there is a method of treating skin cancer. This method comprises the steps of *in vivo* transduction of epidermal cells with a keratin K1 vector. The nucleic acid cassette of either vector contains the nucleic acid sequence coding for antisense RNA for the E6 or E7 genes of the human papilloma virus or coding for the normal p53 protein.

It has been found that the keratin K1 vector contains a novel negative regulatory element in its 3′ flanking sequence which can be suppressed by Vitamin D_3. With the Vitamin D_3 regulatory element in the vector, the expression of a nucleic acid cassette can be regulated by Vitamin D, a commonly used substance in animals and humans.

The human keratin K1 vector can also be modified by the insertion of additional 5′ flanking sequences from an 18 kb Eco RV fragment to its 5′ end (nucleotides 6090 to 14180 of SEQ. ID. No. 3). The addition of these sequences allows the human keratin K1 vector to be expressed

exactly like the endogenous K1 gene, that is, post mitotically in cells committed to terminal differentiation. Since these cells are programmed to die and will eventually slough into the environment, this is another way of producing transient expression in cells.

An additional embodiment of the present invention is a method for vaccination comprising the step of *in vivo* introduction of a keratin K1 vector into epidermal cells. The nucleic acid cassette in the vectors usually codes for a polypeptide which induces an immunological response. An example of this is the viral capsid from the human papilloma virus. One skilled in the art will readily recognize that any other variety of proteins can be used to generate a immunologic response and thus produce antibodies for vaccination.

The following examples are offered by way of illustration and are not intended to limit the invention in any manner.

EXAMPLE 1

Construction and Characterization of a Vector From the HK1 Gene

To target the expression of exogenous DNA to the epidermis a vector from the human keratin K1 gene was constructed. Among its many uses, it is useful in making transgenic animals.

A schematic showing the structure of the human keratin K1 gene is shown in Figure 1. The 12 kb EcoRI fragment containing the entire human keratin K1 gene was originally isolated from lambda clone c55 (Johnson, et al., PNAS, USA, Vol. 82, pp. 1896-1900, (1985)). In constructing the targeting vector, most of the first exon including the ATG was removed, leaving only the 5′ non-coding sequences, the first intron and the intron-exon boundaries. In addition, the remainder of the gene up to the termination codon was deleted. A poly linker containing the following unique restriction sites (Bam HI, Xma I, Kpn I, Not I, and Cla I) was engineered into a site 3′ of the first intron to allow easy insertion

of exogenous DNA. These manipulations were performed through the use of polymerase chain reactions (PCR). The unique EcoRI sites were conserved at the ends of the vector to allow easy amplification in pGEM vectors and excision for purification from plasmid sequences prior to injection into embryos.

The rationale for constructing the vector in this manner was as follows. Since the specific elements responsible for the expression characteristics of the 12 kb human keratin K1 fragment have not been defined, the entire 5′ and 3′ flanking regions were included in the vector construct. One skilled in the art will readily recognize that as these elements are further defined the flanking sequences can be changed accordingly. In addition, sequences within the 3′ non-coding region were retained since these may confirm stability to transcripts of exogenous DNA in epidermal cells. The first intron was retained to potentially enhance expression efficiency (Brinster, et al., PNAS, USA, Vol. 82, pp. 1896-1900 (1988).

EXAMPLE 2

HK1 Expression in Epidermal Keratinocytes

To assess the human keratin K1 targeting vector for exclusive expression in epidermal keratinocytes, the β-galactosidase reporter gene was cloned into Bam HI and Cla I restriction sites located in the polylinker region of the expression vector (Figure 1). The β-galactosidase gene has frequently been used as a reporter gene to assess targeting specificity (MacGregor et al., In: Methods in Molecular Biology Vol. 7, pp. 217-235 (1991). This construct was designated pHK1.β-gal. To determine if expression of this construct resulted in the production of a functional protein, and to determine whether the vector retained cell type specificity, this construct was transfected into primary epidermal keratinocytes and primary dermal fibroblasts. At seventy-two hours post transfection cells

were stained with a solution containing the substrate 5-bromo-4-chloro-3-indoyl-β-galactosidase (X-gal). β-galactosidase activity, indicated by a blue coloration, was detected in keratinocytes but not fibroblasts. Thus, expression of the HK1.β-gal construct was cell type specific and resulted in the production of a functional protein.

EXAMPLE 3

Transgene Mice

The same pHK1.β-gal construct utilized in the *in vitro* studies discussed in Example 2 was used in the production of transgenic mice. This construct was digested with EcoRI (see Figure 1) and subjected to preparative agarose gel electrophoresis to purify the pHK1.β-gal expression construct away from plasmid sequences (pGEM 3) which might interfere with expression. The separated expression construct sequences were purified and recovered using NA 45 DEAE membrane (Schleicher & Schuell). DNA was precipitated and resuspended at 1-3 ng/ul. ICR outbred female mice (Sasco) were given PMS and HCG to stimulate superovulation, mated to FVB males (Taconic) and the resulting early fertilized embryos (most preferably on cell stage) were collected from the oviducts. DNA was micro-injected into the pronuclei and the embryos were surgically transferred to pseudopregnant recipient females (the result of mating ICR females with vasectomized $B_6D_2F_1$ males (Taconic))

In the initial experiments, 40 mice were born. In order to quickly determine if the pHK1.β-gal transgene was being exclusively expressed in the epidermis of these mice, these animals were sacrificed at birth. A small amount of tissue was removed for extraction of DNA and the remainder of the neonate was rapidly frozen in Tissue-Tek O.C.T. for frozen sections. PCR analysis was performed on the extracted DNA using oligonucleotide primers specific for the intron within the HK1 vector and

this demonstrated that 5 of the 40 neonates contained the HK1.β-gal construct.

To assess whether expression of the HK1β-gal construct was restricted to the epidermis or expressed in other squamous epithelia, frozen longitudinal sections were cut from several PCR positive and PCR negative embedded neonates and these were stained with X-Gal. Typical results are shown in Figure 2 where PCR positive animal, #30, expressed high levels of β-galactosidase in the epidermis (Figure 2A and a PCR negative sibling, #29, was completely negative (Figure 2B), indicating that endogenous murine β-galactosidase was not expressed at sufficient levels in the epidermis to cause false positives in this assay. Staining of the intestine was observed in both the positive (#30) and negative (#29) neonates. This may represent endogenous enzyme activity or the production of β-galactosidase by bacteria in the intestine. X-gal staining was detected in the basal compartment, although it is not as intense as in the differentiated layers (Figure 2D). Thus, the human keratin K1 expression vector is also expressed in a substantial number of proliferating basal cells.

The most important finding from these initial transgenic experiments in that the vector constructed from the human keratin K1 gene can target the expression of an exogenous coding sequence exclusively to the epidermis of transgenic mice. This specificity of targeting can be readily seen in Figure 2A. This low power exposure of the skin of #30 demonstrates intense staining with X-Gal. In addition, there are numerous hair follicles and sebaceous glands in this section which are marked by arrows and these do not stain with X-Gal. Keratins K5 and K14 are not only expressed in the epidermis, but in all squamous epithelia, including hair follicles and sebaceous glands. The expression pattern for keratin K14 (Figure 2C) is revealed by immunofluorescence with a specific K14 antiserum of an area from a consecutive section that

is comparable to that in Figure 2A. Note staining of the epidermis, as well as, hair follicles and sebaceous glands. If the strategy used in construction the human keratin K1 expression vector had altered its targeting specificity in transgenic mice, then X-Gal staining would have been observed in hair follicles, sebaceous glands, other squamous epithelia, and perhaps even other tissue types. However, expression of the HK1.β-gal transgene, like the keratin K1 gene itself is restricted to the epidermis.

EXAMPLE 4

Regulation of Keratin K1 Vector by Vitamin D_3

A novel Vitamin D_3 responsive element was used to modulate expression levels in the epidermis. Although all of the regulatory elements of the human keratin K1 gene have not been identified, a novel negative regulatory element from the human keratin K1 gene (HK1.NRE) has been identified and this example demonstrates that it is able to suppress a heterologous promoter in response to Vitamin D_3. The HK1.NRE is 70 nucleotides in length (nucleotides 9134 to 9204 of SEQ. ID. No. 1). PCR technology was used to generate Bam HI and Bgl II sites at opposite ends of this fragment. This facilitates generating multiple copies of this fragment since ligation and digestion with Bam HI and Bgl II will select for oligomers which have ligated head to tail. Four tandem copies of the HK1.NRE were inserted into the Bgl II cloning site of pA10.CAT. In the absence of Vitamin D_3 this construct is highly expressed when transfected into primary mouse epidermal cells (Figure 3). The addition of increasing concentrations of Vitamin D_3 to the culture medium completely suppresses transcription of this heterologous promoter. This observation indicates that the activity of the human keratin K1 expression vector can be modulated in the epidermis. The activity of the human keratin K1 vector is suppressed in the epidermis by topical application of Vitamin D_3, or an analogue, to the skin.

EXAMPLE 5

Development of Transgenic Animal Models for Skin Carcinogenesis

The ability to stably introduce genes into the germline of mice has greatly enhanced prospects for generation of animal models of human disease (Leder and Stewart U.S. Patent No. 4,736,866 issued April 12, 1988 and Palmiter and Brinster, Ann. Rev. Genet., Vol. 20, pp. 465-499). When such genes are combined with regulatory sequences that target their expression to specific tissues, it provides a model to not only study diseases in the context of living organisms, but also in specific tissues suspected of being the targets of these genes. Thus, transgenic mice offer the possibility to determine the influence of factors such as blood supply, an intact immune system, humoral and cell-mediated growth controls and physical barriers on disease progression. The epidermis is an attractive tissue for targeted gene expression; not only is it a model for epithelial diseases in general but the accessibility of the epidermis allows easy detection of progressive pathological changes that result from transgene expression as well as the assessment of the potential role played by environmental factors in these processes. In addition, the prospects for utilizing gene therapy to treat cancer are coming closer to reality. Therefore, animal models of human cancers would be useful to assess the therapeutic potential of these approaches. The development of animal models of skin disease is dependent upon the ability to specifically target gene expression to the epidermis. The human keratin K1 targeting vector described in Example 1 is ideally suited for this purpose.

EXAMPLE 6

Targeting the v-*ras*^{Ha} Oncogene to the Epidermis

One family of proto-oncogenes, the *ras* family (*ras*^{Ha}, *ras*^{K1}, *ras*^{N}) has been identified in approximately 20% of human tumors by virtue of specific point mutations at codons 12, 13, and 61 which activate their

transforming potential. The mechanisms whereby *ras* genes become activated are currently unknown but there is widespread evidence that environmental agents play pivotal roles in the etiology of *ras* mutations. To date few studies have undertaken to study *ras* activation in human skin malignancies. However recent reports have identified ras^{Ha} activation in basal and squamous cell carcinomas appearing on sun exposed body sites, interestingly at potential pyrimidine dimer sites possibly derived from skin exposure to UV irradiation. In the mouse skin model of chemical carcinogenesis where the three distinct stages of initiation, promotion and malignant conversion have been defined, ras^{Ha} activation has been found in benign squamous papillomas, the end point of initiation and promotion suggesting an early role for ras^{Ha} in skin carcinogenesis. Taken collectively, the above experimental evidence suggests the importance of developing an animal model to further study the mechanism of ras^{Ha}-induced skin carcinogenesis. Toward this end, the sequence encoding the v-ras^{Ha} protein of Harvey Murine Sarcoma Virus (Dhar, et al., Science, Vol. 217, pp. 934-937, (1982) was cloned into the Bam HI and Cla I sites of the human keratin K1 expression vector (Figure 4) To discriminate expression of the v-ras^{Ha} transgene from that of the endogenous *ras* gene, a sequence encoding the human keratin K6 epitope SEQ. ID. No 4 was engineered onto the 5′ end of the v-ras^{Ha} cassette.

HK1 *ras* transgenic mice exhibit the following phenotype: 1) Newborn transgenic mice expressing v-ras^{Ha} (HK1 *ras*) exclusively in the epidermis show distinct wrinkled skin at 48 hours and are smaller than litter mates. 2) Juvenile HK1 *ras* transgenic mice exhibit progressive keratinization which peaks at 14 days. 3) The histotype of newborn HK1 *ras* mice reveals massive epidermal hyperplasia with up to 20 fold thickening of the epidermis. 4) By day 14 this progresses to massive hyperkeratosis. Both histotypes are pre-neoplastic, papillomatous, non dysplastic and exhibit few appendages.

The HK1 *ras* transgenic mice develop benign tumors. Typical lesions appear within 10-12 weeks at single sites. The histotype of these tumors reveals a well differentiated squamous papilloma. Papillomas often appear at sites after wounding. Many of these papillomas are prone to regression. This regression phenomenon suggests that ras^{Ha} alone is insufficient to maintain even a benign phenotype and requires further events which may involve roles for additional oncogenes/antioncogenes.

EXAMPLE 7

Targeting the *fos* Oncogene to the Epidermis of Transgenic Mice

Recent *in vitro* studies have shown that the v-*fos* gene can convert to malignancy primary keratinocytes or papilloma cell lines which expressed an activated ras^{Ha} (Greenhalgh, et al., PNAS, USA, Vol. 87, pp. 643-647, (1990); Greenhalgh and Yuspa, Mol. Carcinogen., Vol. 1, pp. 134-143 (1988). This suggested that *fos* could play a later role in epidermal carcinogenesis and cooperate with the benign phenotype imparted by activated ras^{-Ha} expression. Although this alone was sufficient to initiate the establishment of HK1 *fos* transgenic mice with a view to mate with HK1 *ras* mice, two further studies have identified a role for *fos* in normal epidermal differentiation and thus highlights *fos* as an attractive target for perturbation. Using a c-*fos*/β-gal fusion gene Curran and co-workers (Smeyne et al, Neuron, Vol. 8, pp. 13-23 (1992)) have shown significant *fos* expression in the differentiated layers of the epidermis and (Fisher, et al., Development, Vol. III, pp. 253-258, (1991)) have localized c-*fos* expression to a specific subset of granular cells. Thus *fos* may have an important role in the control of the final stages of keratinocyte differentiation. The putative perturbations of this normal role for c-*fos* in such specialized cells by v-*fos* can only be explored in the context of targeted expression in transgenic mice. In addition the c-*fos* proto-oncogene is known to function as a transcriptional regulator in conjunction with the c-*jun*/AP1 gene

product and thus, while targeting *ras*^{Ha} represents studies of membrane signalling on neoplasia, targeting *fos* explores the role of transcriptional control on this process.

Thus, the *fos* protein coding sequence from the FBJ/FBR chimeric v-*fos* plasmid pFBRJ was inserted into the human keratin K1 targeting vector (Figure 5). To discriminate expression of the v-*fos* transgene from that of the endogenous *fos* gene, a sequence encoding the human keratin K1 epitope (SEQ. ID. No. 5) was engineered onto the 5′ end of the v-*fos* cassette.

HK1 *fos* transgenic mice exhibit the following phenotype: 1) A specific ear phenotype typically appears at 3-4 months initially in the wounded (tagged) ear and then becomes bilateral. 2) In several animals expressing severe phenotypes, the wounded ear lesion can grossly resemble a benign keratoacanthoma. 3) Alopecia and hyperkeratosis of the axilla often develop in older animals (approximately 1 year of age).

The histotypes of the HK1 *fos* mice are as follows: 1) The histotype of the initial ear lesions exhibits hyperplasia and hyperkeratosis, a pre-neoplastic pathology with few dysplastic cells and little evidence of further neoplastic progression. 2) At later stages the massive hyperkeratotic histotype resembles a benign keratoacanthoma.

Three HK1 *fos* transgenic mice lines have been established which develop an obvious pre-neoplastic ear phenotype at 3-4 months. The promotion stimulus derived from wounding (i.e. ear tag) appears to accelerate the appearance of this phenotype which eventually becomes bilateral. Also, it appears that friction in the axilla and inguinal area may also promote a pre-neoplastic hyperplastic/hyperkeratotic response after a significant latent period. Collectively these data support a fundamental role for the *fos* gene in normal keratinocyte differentiation and perturbation by v-*fos* results in pre-neoplastic differentiation disorders. In several HK1 *fos* mice severe ear lesions appear to progress to resemble

benign keratoacanthomas. Although numbers are low at this time, that this is the resultant tumor type is consistent with a role for *fos* in the latter stages of terminal differentiation, and low numbers and latency suggest a requirement for additional events.

EXAMPLE 8

Targeting HPV 18 E6 and E7 Gene Expression to the Epidermis

There is widespread evidence from clinical and epidemiological studies which implicate human papilloma viruses (HPV) in the etiology of certain squamous epithelial tumors in humans. HPV's have a specific tropism for squamous epithelial cells and different types of HPVs have specificity for the anatomic site that they infect. Additionally, within a specific subgroup of HPVs, certain types are associated with development of either benign (e.g. HPV6 and 11) or malignant (e.g., HPV-16 and 18) disease and this may center on the properties of the E6 and E7 genes. Through adaptation to the differentiation programs of the epithelia that they infect, HPVs have evolved a clever strategy for the production of infectious progeny. HPVs infect basal epithelial cells but do not undergo lytic replication in this compartment, thus, the germinative pool of cells is not subjected to the cytopathic effects of late viral gene expression. Production of virus only occurs in terminally differentiated cells that have lost proliferative potential and will be desquamated into the environment. This strategy not only provides for the spread of mature viral particles, but ensures their continuous production by replenishment with cells from the basal compartment. Since the life cycle of the virus is so tightly linked to all stages of differentiation of squamous epithelial cells, establishment of successful culture systems has been difficult. To date, these host factors, coupled with regulatory mechanisms present within papilloma virus genomes themselves have also hindered attempts to observe pathological effects of HPV gene expression in squamous epithelia in

transgenic mice. These restrictions on utilization of the transgenic mouse model have been overcome with the ability to specifically target HPV gene expression to squamous epithelia using the human keratin K1 targeting vector. In the example provided, the coding sequence for the E6 and E7 genes of HPV 18 were inserted into the human keratin K1 targeting vector at the Bgl II and Cla I sites (Figure 6).

HK1 E6/E7 mice exhibit the following phenotypes and histotypes: 1) One mouse exhibited a subtle skin lesion at 7 months characterized by skin rigidity, thickening and roughness underlying the fur which later progresses to a wart like structure by 10 months. 2) The histotype of this lesion exhibits hyperplasia, hyperkeratosis and the beginnings of verrucous formation. 3) The histotype of a lesion from another mouse at 11 months, is characteristic of a typical wart induced by HPV.

To date three HK1 E6/E7 transgenic mouse lines have been established which develop HPV-like lesions with low frequency and long latent periods. At this time it is unclear whether this limited appearance of phenotypes reflects the subtle nature of the lesion, and its requirement for a long latency period, or the complex nature of HPV biology. However it is noteworthy that our result is consistent with the epidemiology of HPV infections in humans, e.g. although a large percentage of a given female population can test positive for cervical infection by HPV 6, 11, 16, and 18 relatively few progress to develop overt lesions. It may be therefore that the apparent delay and low phenotype frequency exhibited by these mice provides a relevant background to study the consequences of HPV expression during epithelial differentiation. In addition, these mice can be useful in assessing the efficacy of novel antisense pharmaceuticals which have been designed to inhibit expression of the E6 and E7 genes of HPV 18.

EXAMPLE 9

Production of Transgenic Mice Expressing TGF-α in the epidermis

Transforming growth factor alpha (TGF-α) is a cytokine with structural and functional characteristics similar to epidermal growth factor (EGF). Both TGF-α and EGF bind to the epidermal growth factor receptor (EGF-R) and stimulate the tyrosine kinase cascade. TGF-α is expressed by both normal and transformed cells and causes proliferation of cultured keratinocytes. *In vivo*, TGF-α induces angiogenesis and is more potent than EGF in accelerating wound healing. In normal human skin, expression of TGF-α occurs in all layers of the epidermis and in certain areas of the appendages. Several cutaneous diseases such as psoriasis, squamous cell carcinoma, and congenital bullous ichthyosiform erythroderma have been associated with altered expression of TGF-α.

To determine whether altered expression of TGF-α plays a role in the pathogenesis of these diseases, the protein coding sequence of human TGF-α was inserted into the human keratin K1 targeting vector (Figure 7). Injection of the HK1.TGF-α construct into embryos resulted in phenotypic founders that were quite similar to that of ras^{Ha}. The histotype was also similar with epidermal hyperplasia, hyperkeratosis and relative alopecia. One founder (2 1/2 months of age) has developed multiple papillomas. Histologically these appear to be squamous papillomas. To date, none of these lesions have converted to a malignant phenotype.

EXAMPLE 10

Production of Transgenic Mice Expressing the HIV *tat* gene in the epidermis.

Patients infected with the human immunodeficiency virus (HIV) are at high risk for the development of specific AIDS-associated cutaneous disorders. Often patients manifesting symptoms have skin lesions ranging

from hyperproliferative conditions such as psoriasis to Kaposi's sarcoma and metastatic basal cell carcinoma. The precise role of HIV genes, the cells of origin and hence etiology of such skin lesions remains unknown. It may be that specific HIV genes, e.g., the trans-regulatory protein *tat*, play a role directly or indirectly on the homeostatic mechanisms of host cells and tissues. Alternatively, the HIV *tat* gene may interact with or activate other viral genes present from latent or opportunistic infections, e.g., human papilloma virus (HPV). To directly assess the role of keratinocytes in the development of AIDS-associated cutaneous disorders, the HIV *tat* gene is targeted to the epidermis of transgenic mice. Targeting of the *tat* gene and exclusive expression in keratinocytes is achieved by the use of the human keratin K1 vector (Figure 8). The development of strains of mice which develop cutaneous lesions with predictable kinetics as a result of expression of the HIV *tat* gene alone or in combination with other oncogenes serves as a useful model for assessing therapeutic potential of antisense pharmaceuticals designed to inhibit expression of the HIV *tat* gene.

EXAMPLE 11

Utilization of the HK1 Vector for Gene Therapy Applications

Where exclusive expression in epidermal cells is desirable and for transient expression the HK1 vector is an excellent choice for gene therapy. Unlike the human keratin K1 gene itself, the human keratin K1 vector derived from the 12 kb fragment is expressed in proliferating basal cells in the epidermis. In more recent transgenic experiments, it has been determined that a larger fragment containing the human keratin K1 gene, a 18 kb Eco RV fragment (Shown schematically in Figure 9), is expressed exactly like the endogenous mouse K1 gene, i.e. post mitotically in cells committed to terminal differentiation. These cells are programmed to die and will eventually slough into the environment. Therefore, for human

applications where transient expression is desired, it is possible to design a vector that will only be expressed in cells after they commit to terminal differentiation and begin moving upward toward the outer layers of the epidermis. The vector will be expressed approximately 10-14 days prior to being shed into the environment. This can be accomplished by inserting additional 5′ flanking sequences from the 18 kb Eco RV fragment onto the end of the original human keratin K1 vector (See Fig. 10).

EXAMPLE 12

Detection of Carcinogens and Tumor Promoters

Short-term tests (STTs) for genotoxic chemicals were originally developed as fast, inexpensive assays to assess the potential hazard of chemicals to humans. However, a recent report summarizing the results of a project initiated by the National Toxicology Program to evaluate the ability of STT's to predict rodent carcinogenicity questions the validity of relying solely on STT's. Three of the most potent carcinogens, detected in the rodent assays, produced no genetic toxicity in any of the four STTs evaluated (Tennant et al., Science, Vol. 236, pp. 933-941, (1987). Thus, to receive EPA/FDA approval for new compounds, chemical, agricultural, food and drug companies are currently required to perform two year animal tests costing up to $2 million. The development of new transgenic strains of mice that have been genetically engineered to rapidly detect carcinogens and tumor promoters would substantially reduce the overhead cost of long-term animal studies. The suitability of the transgenic mouse lines claimed in this patent application for rapid detection of carcinogens is initially determined with a known skin carcinogen, DMBA, to determine whether benign lesions appear earlier than in control non-treated litter mates. To determine suitability for detecting tumor promoters, a known promoter, 12-O-tetra-decanoylphorbol-13-acetate (TPA) is applied to *ras*^{Ha},

and *fos* mice. Since benign lesions in ras^{Ha} mice and hyperplasia in *fos* mice appeared at sites of wounding (i.e., tagged ears), and wounding can promote tumor formation, these lines are useful for these studies.

All patents and publications mentioned in this specification are indicative of the levels of those skilled in the art to which the invention pertains. All patents and publications which are incorporated herein by reference are incorporated to the same extent as if each individual publication was specifically and individually indicated to be incorporated by reference.

One skilled in the art will readily appreciate that the present invention is well adapted to carry out the objects and obtain the ends and advantages mentioned, as well as those inherent therein. The bioreactors, nucleic acid sequences, transformed epidermal cells, transgenic animals and human keratin K1 vector, along with the methods, procedures, treatments, molecules of specific compounds, are exemplary and are not intended as limitations on the scope of the invention. Changes therein and other uses will occur to those skilled in the art which are encompassed within the spirit of the invention as defined by the scope of the claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

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(ii) TITLE OF INVENTION: DEVELOPMENT OF A VECTOR TO TARGET GENE EXPRESSION TO THE EPIDERMIS OF TRANSGENIC ANIMALS

(iii) NUMBER OF SEQUENCES: 5

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(C) OPERATING SYSTEM: PC-DOS/MS-DOS
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(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10747 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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TTCAACCCCT GTGTCTCTGC TTCCTCATTT TACAAATGGG GAGAAAAATA GCATCTATCT 180
CAAAGTTGTG AAAATTAAGC AAGTTAATAC ATATGTGCTA CGTAGAACAG TGCCTGGTAC 240
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(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 6693 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 24979 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: double
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Met Ser Arg Lys Ser Tyr Lys His
1 5

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 16 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: peptide

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

Met Ser Ser Val Lys Phe Val Ser Thr Thr Tyr Ser Gly Val Thr Arg
1 5 10 15

CLAIMS

What we claim is:

1. A keratin K1 vector for expression of a nucleic acid cassette in the epidermis comprising:

a 5′ flanking region of the keratin K1 gene, said 5′ flanking sequence including a keratin K1 promoter, a 5′ transcribed but untranslated region and a first intron all in sequential and positional relationship for expression of the nucleic acid cassette;

a 3′ flanking region of the keratin K1 gene containing Vitamin D_3 regulatory sequences, said 3′ flanking region including a 3′ transcribed but untranslated region and contiguous non-coding DNA containing the transcriptional termination region; and

a polylinker having a plurality of restriction endonuclease sites, said polylinker connecting the 5′ flanking region to the 3′ flanking region and said polylinker further providing a position for insertion of the nucleic acid cassette.

2. The keratin K1 vector of claim 1, wherein the 5′ flanking region is approximately 1.2 kb and the 3′ flanking region is approximately 3.9 kb.

3. The vector of claim 1, wherein the nucleic acid cassette includes a nucleic acid sequence coding for a protein, polypeptide or antisense RNA.

4. The vector of claim 1, wherein the cassette includes a nucleic acid sequence coding for an oncogene.

5. The vector of claim 4, wherein the oncogene is selected from the group consisting of *ras*, *fos*, *myc*, *erb*, *src*, *sis* and *jun*.

6. The vector of claim 1 wherein the cassette includes a nucleic acid sequence coding for a transforming gene.

7. The vector of claim 1, wherein the restriction endonucleases are selected from the group consisting of Bam HI, Xma I, Kpn I, Not I Cla I and Bgl II.

8. The vector of claim 1, wherein the nucleic acid cassette contains the E6 or E7 transforming sequence of human papilloma virus.

9. The vector of claim 1, wherein the nucleic acid cassette contains the TGF-α sequence.

10. The vector of claim 1, further comprising an additional 5′ flanking sequence from the 18 kb Eco RV fragment on the end of the vector.

11. A bioreactor comprising transformed epidermal cells including the vector of claim 3.

12. The bioreactor according to claim 11 wherein the vector includes a cassette having a nucleic acid sequence coding for a protein or polypeptide selected from the group consisting of a hormone, a growth factor, an enzyme, a drug, a tumor suppressor, a receptor, an apolipoprotein, a clotting factor, a tumor antigen, a viral antigen, an insect antigen, a bacterial antigen and a parasitic antigen.

13. The bioreactor of claim 12, wherein the nucleic acid sequence encodes proinsulin or insulin.

14. The bioreactor of claim 12, wherein the nucleic acid sequence encodes growth hormone.

15. The bioreactor of claim 12, wherein the nucleic acid sequence encodes insulin-like growth factor I, insulin-like growth factor II or insulin growth factor binding protein.

16. The bioreactor of claim 12, wherein the nucleic acid sequence encodes a clotting factor.

17. The bioreactor of claim 12, wherein the nucleic acid sequence encodes an epidermal growth factor (TGF-α), a dermal growth factor (PDGF) or an angiogenesis factor.

18. The bioreactor of claim 12, wherein the nucleic acid sequence encodes a Type IV collagen, laminin, nidogen, or Type VII collagen.

19. The bioreactor of claim 12 for vaccine production, wherein the cassette includes a protein which induces an immunological response.

20. A method for *ex vivo* introduction of a keratin K1 vector into epidermal cells comprising the steps of contransfecting the vector with a selectable marker and selecting the transformed cells.

21. A noncoding fragment of a human keratin K1 gene containing regulatory sequences of SEQ. ID. No. 1.

22. A human keratin K1 vector having:

a 5′ flanking region comprising nucleotides 1 to 1246 of SEQ. ID. No. 1;

a 3′ flanking region comprising nucleotides 6891 to 10747 of SEQ. ID. No. 1; and

a linker comprising nucleotide 2351 to 2376 SEQ. ID. No. 2

23. A method of making transgenic animals comprising the steps of:

collecting very early fertilized eggs;

inserting the vector of claim 3 into said fertilized eggs by micro-injecting the vector into pronuclei; and

transferring said injected eggs into pseudopregnant recipient females.

24. A transgenic animal containing the vector of claim 3 in its germ and somatic cells, wherein said vector was introduced into said animal or an ancestor of said animal at an embryonic stage and the nucleic acid cassette of said vector is only expressed in the epidermis.

25. The transgenic animal of claim 24, wherein the nucleic acid cassette contains a transforming gene sequence.

26. The transgenic animal of claim 25, wherein the oncogene is selected from the group in table 1.

27. The transgenic animal of claim 24 wherein the animal is a rodent.

28. A method of studying the origin of or treatment for cancer comprising the steps of:

making a transgenic animal by injecting an embryo with a human keratin K1 vector containing an oncogene in the nucleic acid cassette;

using the resultant animal or the progeny in studies of cancer.

29. The method of claim 28 wherein the nucleic acid cassette contains the *fos* oncogene sequence.

30. The method of claim 28 wherein the nucleic acid cassette contains the *ras* oncogene sequence.

31. The method of claim 28 wherein the test animal contains more than one oncogene.

32. A method for *in vivo* transduction of epidermal cells with a keratin K1 vector comprising the step of contacting the vector with epidermal cells for sufficient time to transform the epidermal cells.

33. A method for transient introduction of a nucleic acid cassette into a human, comprising the step of contacting a keratin K1 vector containing said cassette with an epidermal cell for sufficient time to transduce the cells with the vector.

34. A method of enhanced healing of a wound or surgical incision comprising the step of *in vivo* transduction of epidermal cells with a keratin K1 vector, wherein said vector includes a nucleic acid cassette having a nucleic acid sequence for a growth factor.

35. The method of claim 34, wherein the epidermal cells are transduced with a plurality of vectors and wherein the cassette of at least one vector includes the nucleic acid sequence of epidermal growth factor

(TGF-α), the cassette of at least one vector includes dermal growth factor (PDGF), the cassette of at least one vector includes the nucleic acid sequence for a matrix protein to anchor the epidermis to the dermis and the cassette of at least one vector includes the nucleic acid sequence for an angiogenesis factor.

36. The method of claim 35, wherein the sequence for the matrix protein is selected from sequences coding for a protein selected from the group consisting of Type IV collagen, laminin, nidogen, and Type VII collagen.

37. The method of claim 35, wherein the angiogenesis factor is selected from the group consisting of acid fibroblast growth factor, basic fibroblast growth factor and angiogenin.

38. A method of treating skin ulcers comprising the steps of *in vivo* transduction of epidermal cells with a keratin K1 vector, wherein said vectors include a nucleic acid cassette having a nucleic acid sequence for a growth factor.

39. The method of claim 38, wherein the epidermal cells are transduced with a plurality of vectors and wherein the cassette of at least one vector includes the nucleic acid sequence of epidermal growth factor (TGF-α), the cassette of at least one vector includes dermal growth factor (PDGF), the cassette of at least one vector includes the nucleic acid sequence for a matrix protein to anchor the epidermis to the dermis and the cassette of at least one vector includes the nucleic acid sequence for an angiogenesis factor.

40. The method of claim 39, wherein the sequence for the matrix protein is selected from sequences coding for a protein selected from the group consisting of Type IV collagen, laminin, nidogen, and Type VII collagen.

41. The method of claim 39, wherein the angiogenesis factor is selected from the group consisting of acid fibroblast growth factor, basic fibroblast growth factor and angiogenin.

42. A method of enhanced healing of a wound, surgical incision or skin ulcers in humans and animals, comprising the steps of:

ex vivo transduction of epidermal cells with a keratin K1 vector, wherein said vector includes a nucleic acid cassette having a nucleic acid sequence for a growth factor; and

transplanting said transduced epidermal cells into the animal or human to be treated.

43. The method of claim 42, wherein the epidermal cells are transduced with a plurality of vectors and wherein the cassette of at least one vector includes the nucleic acid sequence of epidermal growth factor (TGF-α), the cassette of at least one vector includes dermal growth factor (PDGF), the cassette of at least one vector includes the nucleic acid sequence for a matrix protein to anchor the epidermis to the dermis and the cassette of at least one vector includes the nucleic acid sequence for an angiogenesis factor.

44. The method of claim 43, wherein the sequence for the matrix protein is selected from sequences coding for a protein selected from the group consisting of Type IV collagen, laminin, nidogen, and Type VII collagen.

45. The method of claim 43, wherein the angiogenesis factor is selected from the group consisting of acid fibroblast growth factor, basic fibroblast growth factor and angiogenin.

46. A method for treating psoriasis comprising the step of *in vivo* transduction of epidermal cells with a keratin K1 vector, wherein said vector includes a nucleic acid cassette having a nucleic acid sequence coding for a protein or polypeptide selected from the

group consisting of TGF-β, a soluble form of cytokine receptor, and an antisense RNA.

47. The method of claim 46 wherein the cassette contains the sequence for TGF-β.

48. The method of claim 46 wherein the cassette contains a soluble form of cytokine receptor selected from the group consisting of IL-1, IL-6 and IL-8.

49. The method of claim 46, wherein the cassette contains an antisense RNA to a sequence selected from the group consisting of TGF-α, IL-1, IL-6 and IL-8.

50. A method for treating skin cancer comprising the step of *in vivo* transduction of epidermal cells with a keratin K1 vector, wherein said vector includes a nucleic acid cassette having a nucleic acid sequence coding for an antisense RNA for the E6 or E7 gene of human papilloma virus.

51. A method for treating skin cancer comprising the step of *in vivo* transduction of epidermal cells with a keratin K1 vector, wherein said vector includes a nucleic acid cassette having a nucleic acid sequence coding for the normal p53 protein.

52. A method for vaccination comprising the step of the *in vivo* transduction of epidermal cells with a keratin K1 vector, wherein said vector includes a nucleic acid cassette having a nucleic acid sequence coding for a protein or polypeptide which induces an immunological response.

53. The method of claim 52, wherein the cassette includes a sequence for a viral capsid protein.

54. The method of claim 53, wherein the capsid protein is from the human papilloma virus.

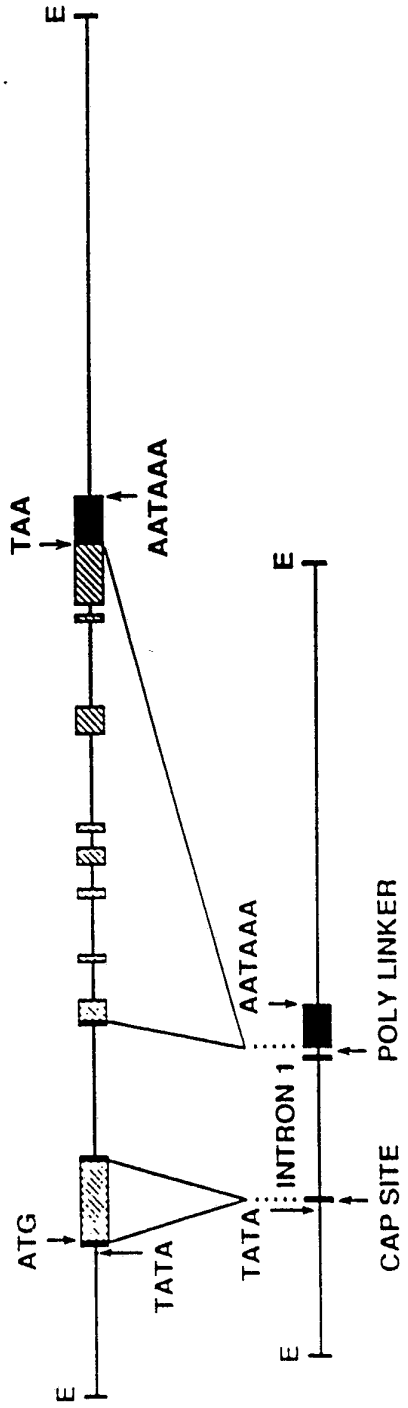

E
TAA
AATAAA
E
AATAAA
POLY LINKER
INTRON 1
CAP SITE
ATG
TATA
TATA
E
E

FIGURE 1

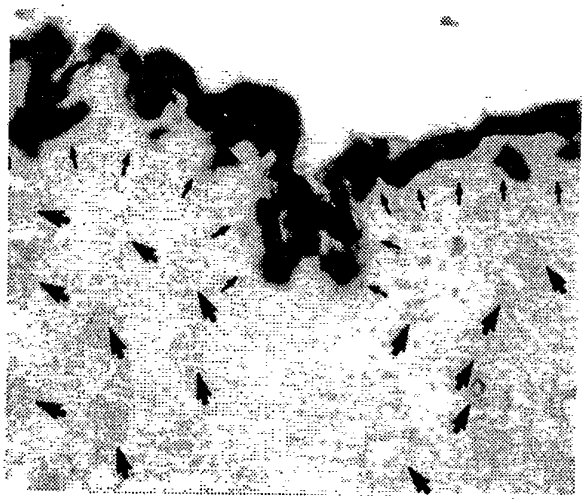


FIGURE 2A

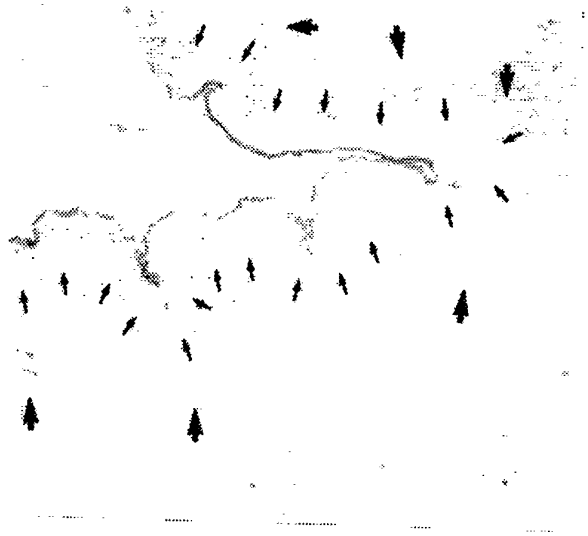


FIGURE 2B

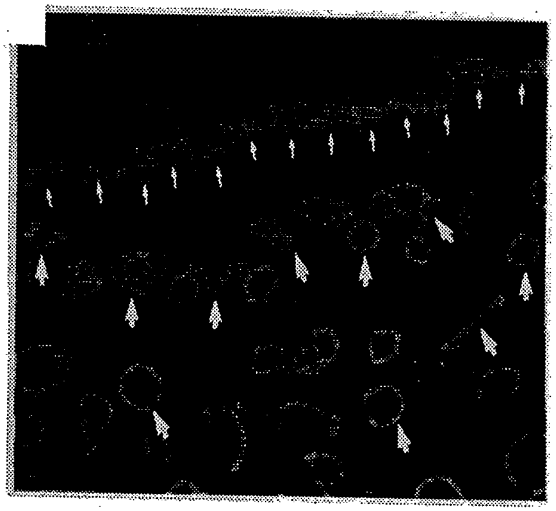


FIGURE 2C

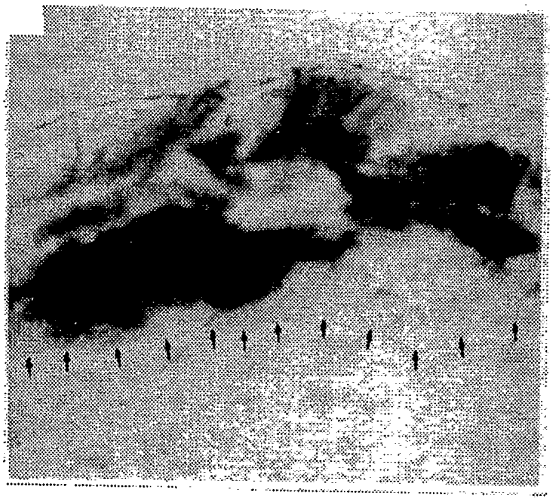
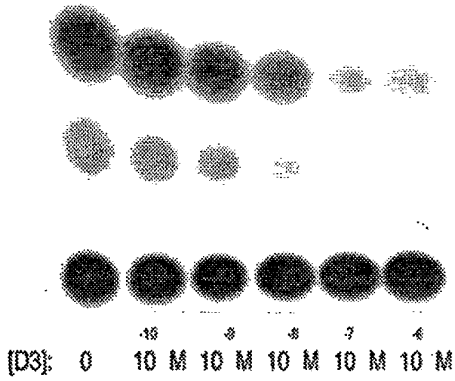


FIGURE 2D



Sequence of HK1.NRE

TGGCCTTGAG$_{10}$GGAGATGATT$_{10}$CACTCTCCTT$_{10}$CACAGAAGAG$_{10}$CTGACCTCTG$_{10}$GGGTCAACAG$_{10}$ATATAGCAGG$_{10}$

FIGURE 3

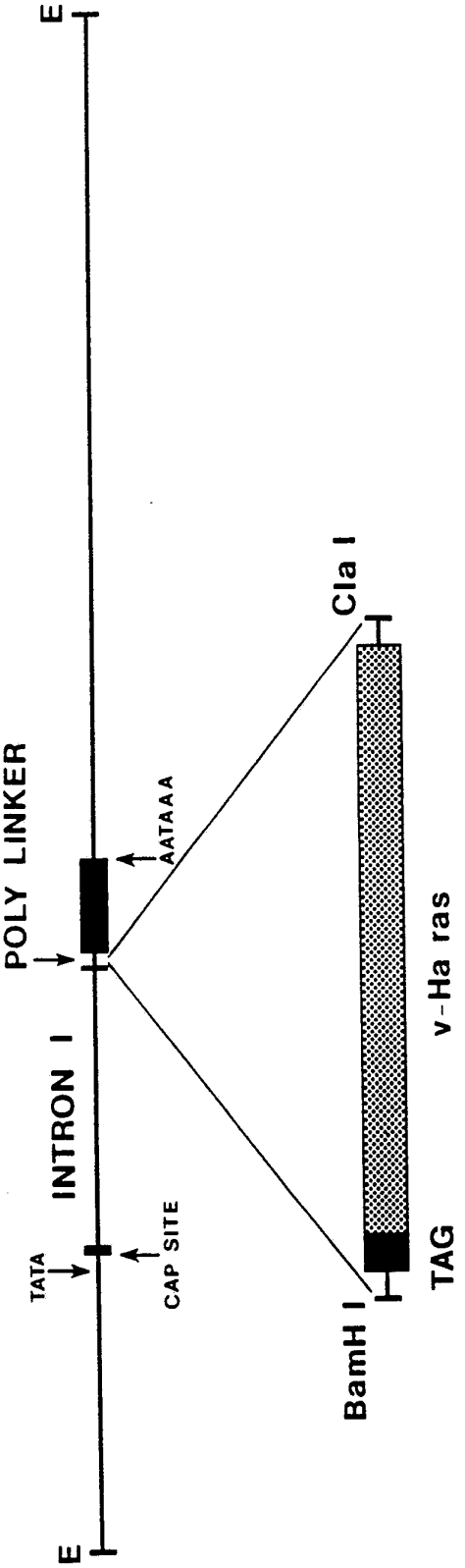

E
POLY LINKER
AATAAA
INTRON I
TATA
CAP SITE
E
Cla I
v-Ha ras
TAG
BamH I
mRNA
AATAAA
3'nc
TAA
3'
ATG
5'
CAP

FIGURE 4

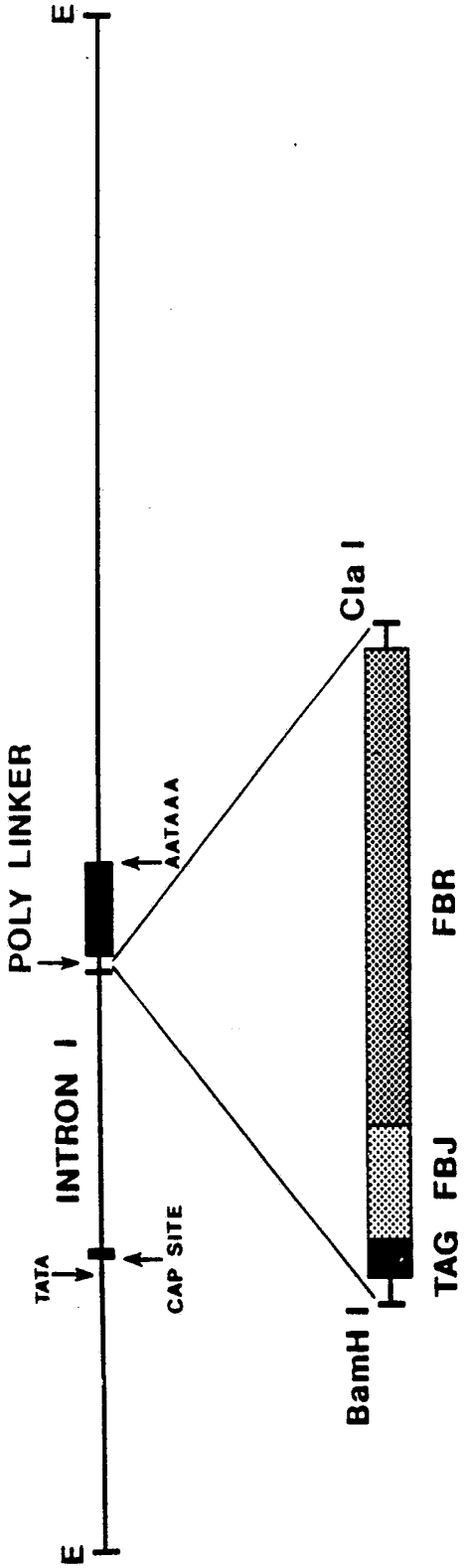
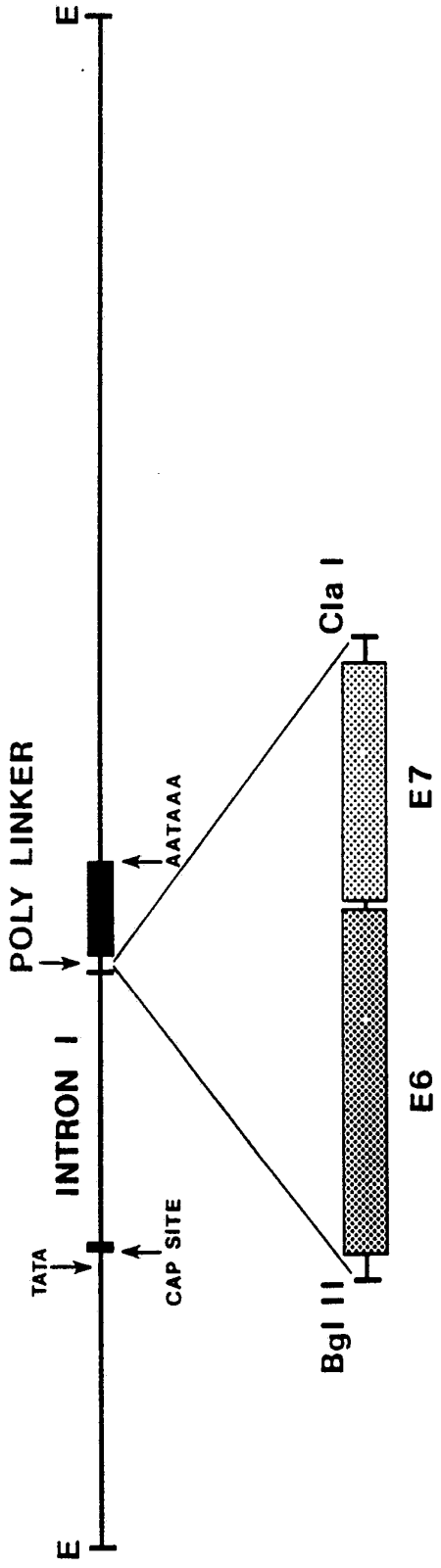

E
POLY LINKER
AATAAA
INTRON I
TATA
CAP SITE
E
Cla I
FBR
TAG FBJ
BamH I
mRNA
AATAAA
3' nc
TAA
3'
ATG
5'
CAP

FIGURE 5


E
POLY LINKER
AATAAA
INTRON I
TATA
CAP SITE
E
Cla I
E7
E6
Bgl II

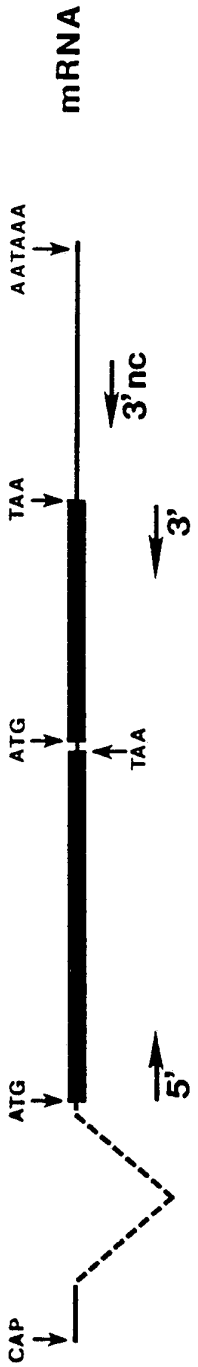

mRNA
AATAAA
3'nc
TAA
3'
ATG
TAA
ATG
5'
CAP

FIGURE 6

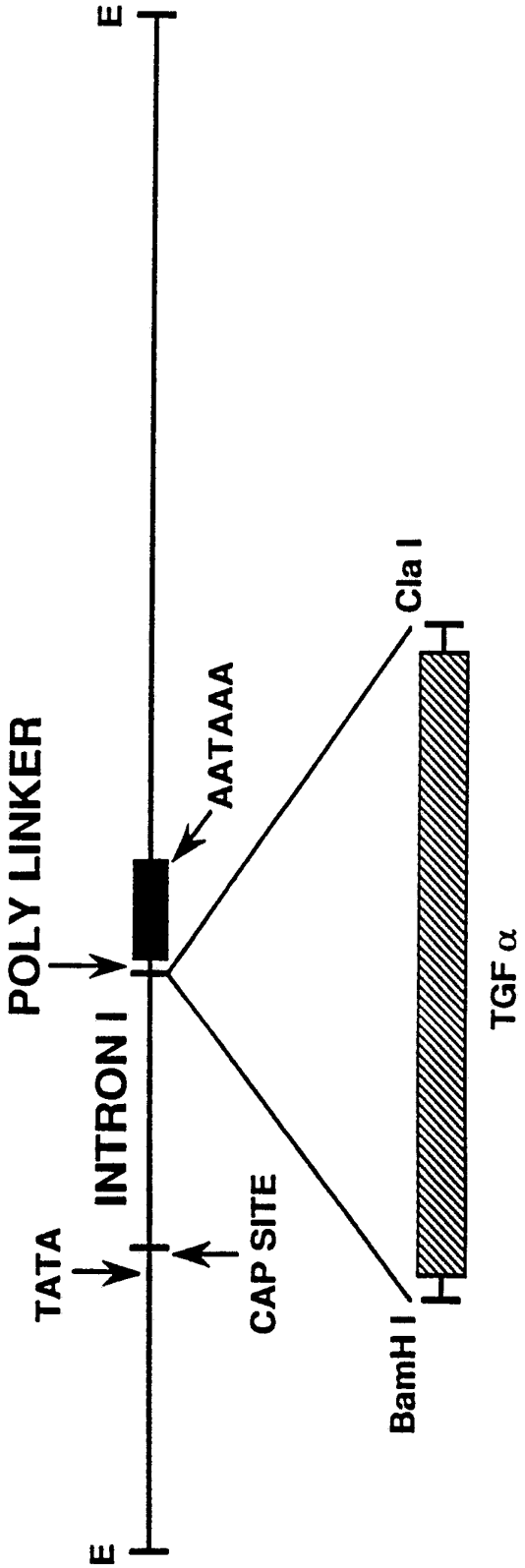


FIGURE 7

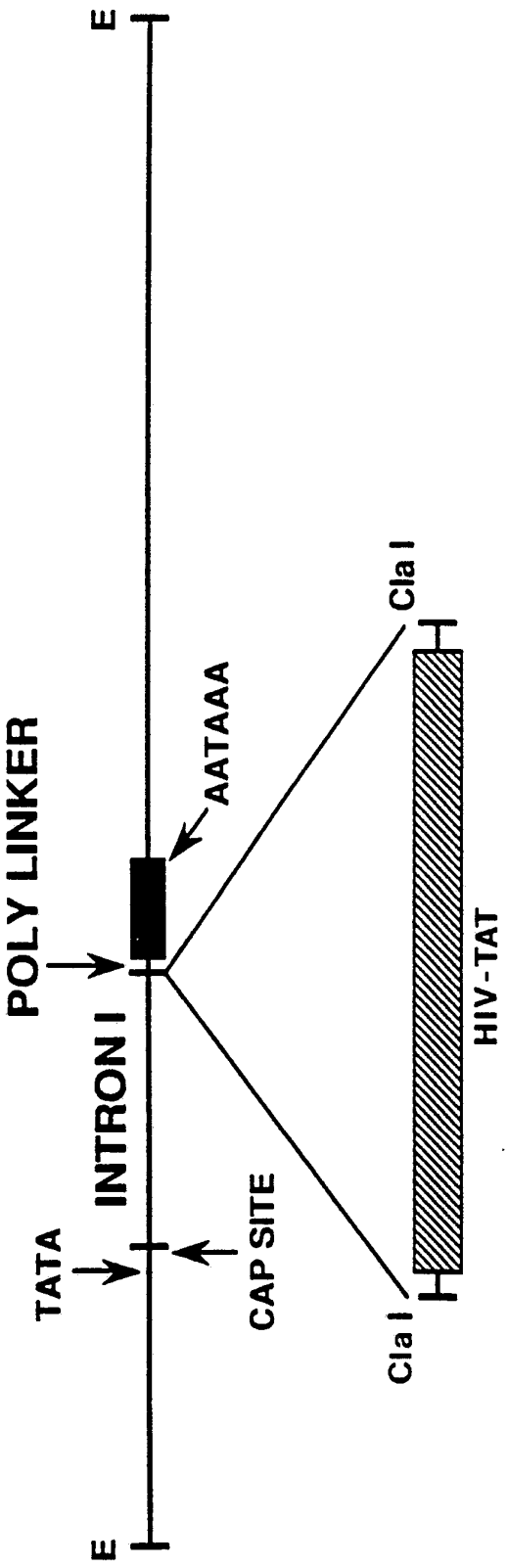

E
POLY LINKER
AATAAA
Cla I
INTRON I
HIV-TAT
TATA
CAP SITE
Cla I
E

FIGURE 8

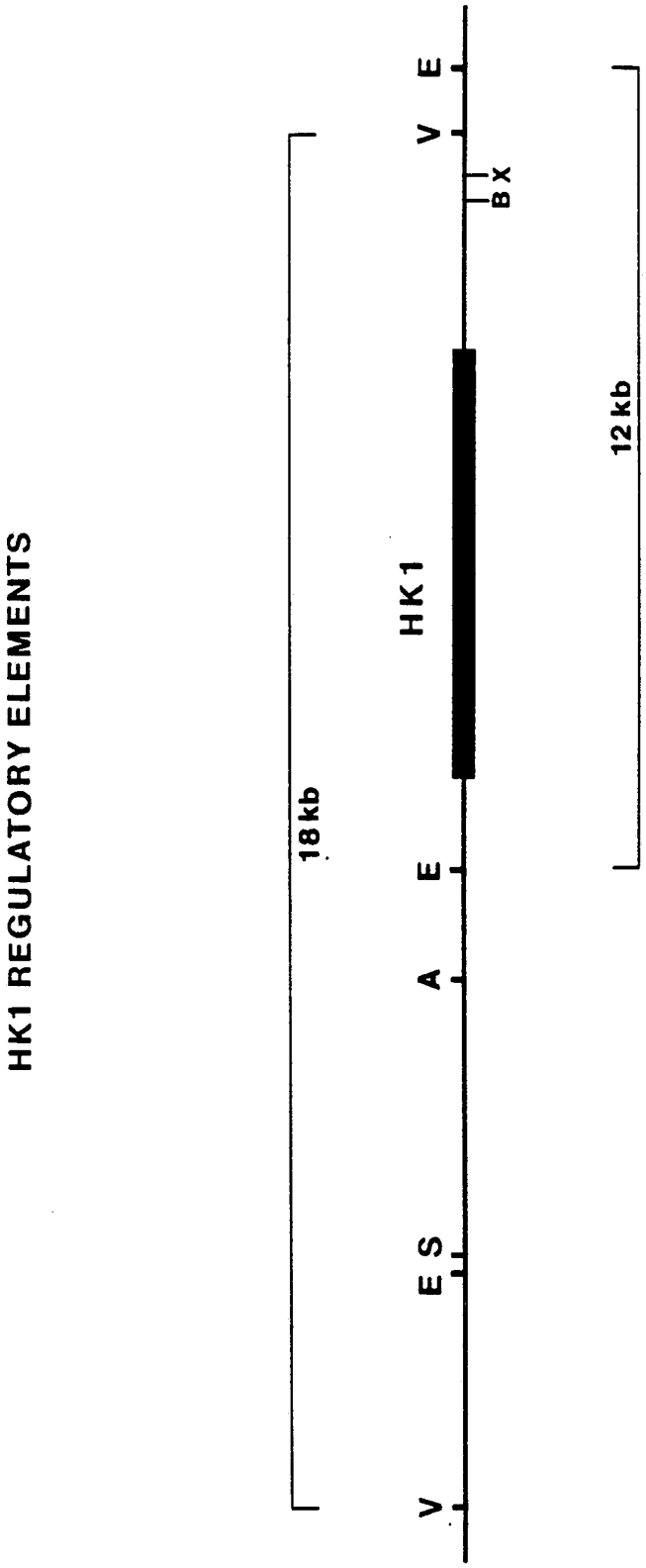


FIGURE 9

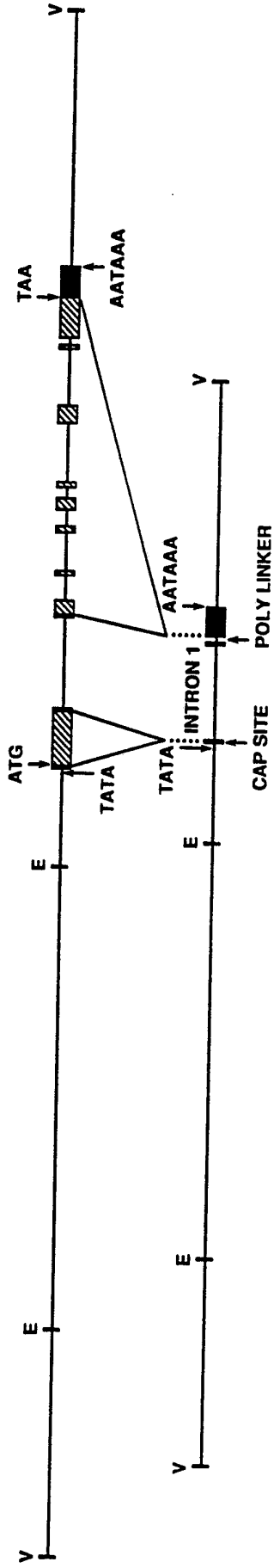

V
TAA
AATAAA
V
AATAAA
POLY LINKER
INTRON 1
CAP SITE
ATG
TATA
TATA
E
E
E
E
V
V

FIGURE 10

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US93/03985

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :C12N 15/00; C12N 5/00; A61K 49/00, A61K 35/00, A61K 31/00
US CL :435/172.3, 240.2, 320.1, 800/2; 424/9, 93B; 514/44
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/172.3, 240.2, 320.1, 800/2; 424/9, 93B; 514/44; 935/22, 55, 62, 70, 71, 111

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Please See Extra Sheet.

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Cell Growth & Differentiation, Volume 2, Issued February 1991, Rosenthal et al., "A human epidermal differentiation-specific keratin gene is regulated by calcium but not negative modulators of differentiation in transgenic mouse keratinocytes", pages 107-113, see the entire document.	1-54
Y	Proc. Natl. Acad. Sci. USA, Volume 82, Issued April 1985, Johnson et al., "Structure of a gene for the human epidermal 67-kDa keratin", pages 1896-1900, see the entire document.	1-54

[X] Further documents are listed in the continuation of Box C. [ ] See patent family annex.

*	Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A"	document defining the general state of the art which is not considered to be part of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E"	earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L"	document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"&"	document member of the same patent family
"O"	document referring to an oral disclosure, use, exhibition or other means		
"P"	document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search 04 JUNE 1993	Date of mailing of the international search report 13 JUL 1993
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. NOT APPLICABLE	Authorized officer JASEMINE C. CHAMBERS Telephone No. (703) 308-0196

Form PCT/ISA/210 (second sheet)(July 1992)*

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US93/03985

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Science, Volume 217, issued 03 September 1982, Dhar et al., "Nucleotide sequence of the p21 transforming protein of Harvey murine sarcoma virus", pages 934-937, see the entire document.	3-6, 8, 9, 30
Y	Virology, Volume 145, Issued 1985, Seedorf et al., "Human papillomavirus type 16 DNA sequence," pages 181-185, see the entire document.	3-6, 8, 9, 50, 52-54
Y	Journal of Virology, Volume 61, No. 4, Issued April 1987, Pirisi et al., "Transformation of human fibroblasts and keratinocytes with human papillomavirus type 16 DNA", pages 1061-1066, see the entire document.	11-20
Y	US,A, 4,736,866 (Leder et al.), 12 April 1988, see the entire document.	23-31
Y	US,A, 4,497,796 (Salser et al.), 05 February 1985, see the entire document.	42-45
Y	Nature, Volume 352, Issued 29 August 1991, Acsadi et al., "Human dystrophin expression in mdx mice after intramuscular injection of DNA constructs", pages 815-818, see the entire document.	32-41, 46-54
Y	Science, Volume 254, Issued 06 December 1991, Dhawan et al., "Systemic delivery of human growth hormone by injection of genetically engineered myoblasts", pages 1509-1512, see the entire document.	42-45

B. FIELDS SEARCHED
Electronic data bases consulted (Name of data base and where practicable terms used):

DIALOG (Files 154, 55, 311, 312), U.S. Automated Patent System (File USPAT, 1975-1993).
Search terms: keratin, K1, clone, gene, vector, transgenic, epidermis, therapy, transduction, oncogene, transforming, HPV, ras, TGF, inventors' names.

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING
This ISA found multiple inventions as follows:

I. Claims 1-22 and 42-45, drawn to a keratin K1 vector, transformed epidermal cells containing the same and a method of wound healing which comprises the ex vivo transduction of epidermal cells with a keratin K1 vector, classified in Classes 435 and 424, subclasses 320.1 and 93B, respectively, for example.

II. Claims 23-31, drawn to a transgenic animal containing a keratin K1 vector and a method of using the same, classified in Classes 800 and 424, subclasses 2 and 9, respectively, for example.

III. Claims 32-41 and 46-54, drawn to methods of wound healing, treating skin ulcers, treating psoriasis, treating skin cancer or vaccination, all of which comprise the in vivo transduction of epidermal cells with a keratin K1 vector, classified in Class 514, subclass 44, for example.