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54	TITLE OF INVENTION

Diagnosis and treatment of multiple sulfatase deficiency and other using a Formylglycine Generating Enzyme (FGE)

57	ABSTRACT (NOT MORE THAN 150 WORDS)

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The sheet(s) containing the abstract is/are attached.

If no classification is furnished, Form P.9 should accompany this form.
The figure of the drawing to which the abstract refers is attached.

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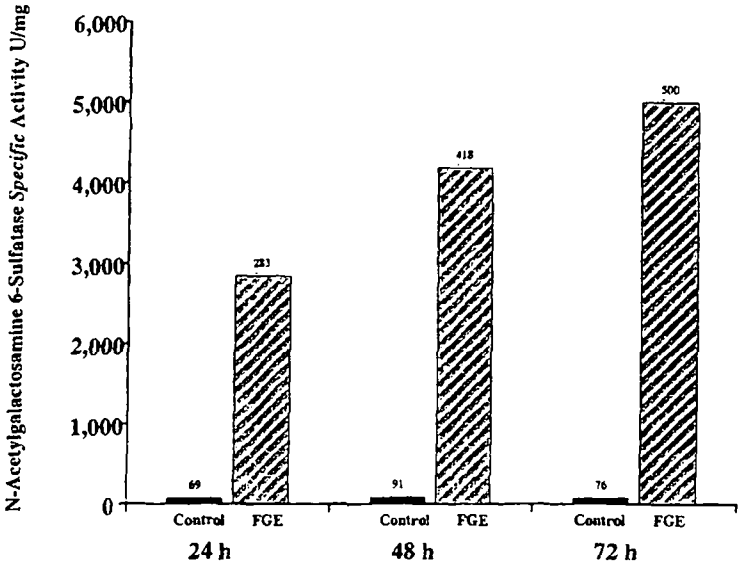
[Continued on next page]

(54) Title: DIAGNOSIS AND TREATMENT OF MULTIPLE SULFATASE DEFICIENCY AND OTHER USING A FORMYL-GLYCINE GENERATING ENZYME (FGE)



(57) Abstract: This invention relates to methods and compositions for the diagnosis and treatment of Multiple Sulfatase Deficiency (MSD) as well as other sulfatase deficiencies. More specifically, the invention relates to isolated molecules that modulate post-translational modifications on sulfatases. Such modifications are essential for proper sulfatase function.

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DIAGNOSIS AND TREATMENT OF MULTIPLE SULFATASE DEFICIENCY AND OTHER SULFATASE DEFICIENCIES

Field of the Invention

This invention relates to methods and compositions for the diagnosis and treatment of Multiple Sulfatase Deficiency (MSD) as well as other sulfatase deficiencies. More specifically, the invention relates to isolated molecules that modulate post-translational modifications on sulfatases. Such modifications are essential for proper sulfatase function.

Background of the Invention

Sulfatases are members of a highly conserved gene family, sharing extensive sequence homology (Franco, B., et al., *Cell*, 1995, 81:15-25; Parenti, G., et al., *Curr. Opin. Gen. Dev.*, 1997, 7:386-391), a high degree of structural similarity (Bond, C.S., et al., *Structure*, 1997, 5:277-289; Lukatela, G., et al., *Biochemistry*, 1998, 37:3654-64), and a unique post-translational modification that is essential for sulfate ester cleavage (Schmidt, B., et al., *Cell*, 1995, 82:271-278; Selmer, T., et al., *Eur. J. Biochem.*, 1996, 238:341-345). The post-translational modification involves the oxidation of a conserved cysteine (in eukaryotes) or serine (in certain prokaryotes) residue, at C_β, yielding L-C_α-formylglycine (a.k.a. *FGly*; *2-amino-3-oxopropanoic acid*) in which an aldehyde group replaces the thiomethyl group of the side chain. The aldehyde is an essential part of the catalytic site of the sulfatase and likely acts as an aldehyde hydrate. One of the geminal hydroxyl groups accepts the sulfate during sulfate ester cleavage leading to the formation of a covalently sulfated enzyme intermediate. The other hydroxyl is required for the subsequent elimination of the sulfate and regeneration of the aldehyde group. This modification occurs in the endoplasmic reticulum during, or shortly after, import of the nascent sulfatase polypeptide and is directed by a short linear sequence surrounding the cysteine (or serine) residue to be modified. This highly conserved sequence is hexapeptide L/V-C(S)-X-P-S-R (SEQ ID NO:32), present in the N-terminal region of all eukaryotic sulfatases and most frequently carries a hydroxyl or thiol group on residue X (Dierks, T., et al., *Proc. Natl. Acad. Sci. U. S. A.*, 1997, 94:11963-11968).

To date thirteen sulfatase genes have been identified in humans. They encode enzymes with different substrate specificity and subcellular localization such as lysosomes, Golgi and ER. Four of these genes, *ARSC*, *ARSD*, *ARSE*, and *ARSF*, encoding arylsulfatase

C, D, E and F, respectively, are located within the same chromosomal region (Xp22.3). They share significant sequence similarity and a nearly identical genomic organization, indicating that they arose from duplication events that occurred recently during evolution (Franco B, et al., *Cell*, 1995, 81:15-25; Meroni G, et al., *Hum Mol Genet*, 1996, 5:423-31).

The importance of sulfatases in human metabolism is underscored by the identification of at least eight human monogenic diseases caused by the deficiency of individual sulfatase activities. Most of these conditions are lysosomal storage disorders in which phenotypic consequences derive from the type and tissue distribution of the stored material. Among them are five different types of mucopolysaccharidoses (MPS types II, IIIA, IIID, IVA, and VI) due to deficiencies of sulfatases acting on the catabolism of glycosaminoglycans (Neufeld and Muenzer, 2001, The mucopolysaccharidoses, *In The Metabolic and Molecular Bases of Inherited Disease*, C.R. Scriver, A.L. Beaudet, W.S. Sly, D. Valle, B. Childs, K.W. Kinzler and B. Vogelstein, eds. New York: Mc Graw-Hill, pp. 3421-3452), and metachromatic leukodystrophy (MLD), which is characterized by the storage of sulfolipids in the central and peripheral nervous systems leading to severe and progressive neurologic deterioration. Two additional human diseases are caused by deficiencies of non-lysosomal sulfatases. These include X-linked ichthyosis, a skin disorder due to steroid sulfatase (STS/ARSC) deficiency, and chondrodysplasia punctata, a disorder affecting bone and cartilage due to arylsulfatase E (ARSE) deficiency. Sulfatases are also implicated in drug-induced human malformation syndromes, such as Warfarin embryopathy, caused by inhibition of ARSE activity due to *in utero* exposure to warfarin during pregnancy.

In an intriguing human monogenic disorder, multiple sulfatase deficiency (MSD), all sulfatase activities are simultaneously defective. Consequently, the phenotype of this severe multisystemic disease combines the features observed in individual sulfatase deficiencies. Cells from patients with MSD are deficient in sulfatase activities even after transfection with cDNAs encoding human sulfatases, suggesting the presence of a common mechanism required for the activity of all sulfatases (Rommerskirch and von Figura, *Proc. Natl. Acad. Sci., USA*, 1992, 89:2561-2565). The post-translational modification of sulfatases was found to be defective in one patient with MSD, suggesting that this disorder is caused by a mutation in a gene, or genes, implicated in the cysteine-to-formylglycine conversion machinery (Schmidt, B., et al., *Cell*, 1995, 82:271-278). In spite of intense biological and medical interest, efforts aimed at the identification of this gene(s) have been hampered by the rarity of MSD patients and consequent lack of suitable familial cases to perform genetic mapping.

Summary of the Invention

This invention provides methods and compositions for the diagnosis and treatment of Multiple Sulfatase Deficiency (MIM 272200), and the treatment of other sulfatase deficiencies. More specifically, we have identified a gene that encodes Formylglycine Generating Enzyme (FGE), an enzyme responsible for the unique post-translational modification occurring on sulfatases that is essential for sulfatase function (formation of L-C_α-formylglycine; a.k.a. *FGly* and/or *2-amino-3-oxopropanoic acid*). It has been discovered, unexpectedly, that mutations in the FGE gene lead to the development of Multiple Sulfatase Deficiency (MSD) in subjects. It has also been discovered, unexpectedly, that FGE enhances the activity of sulfatases, including, but not limited to, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6. In view of these discoveries, the molecules of the present invention can be used in the diagnosis and treatment of Multiple Sulfatase Deficiency as well as other sulfatase deficiencies.

Methods for using the molecules of the invention in the diagnosis of Multiple Sulfatase Deficiency, are provided.

Additionally, methods for using these molecules *in vivo* or *in vitro* for the purpose of modulating *FGly* formation on sulfatases, methods for treating conditions associated with such modification, and compositions useful in the preparation of therapeutic preparations for the treatment of Multiple Sulfatase Deficiency, as well as other sulfatase deficiencies, are also provided.

The present invention thus involves, in several aspects, polypeptides modulating *FGly* formation on sulfatases, isolated nucleic acids encoding those polypeptides, functional modifications and variants of the foregoing, useful fragments of the foregoing, as well as therapeutics and diagnostics, research methods, compositions and tools relating thereto.

According to one aspect of the invention, an isolated nucleic acid molecule selected from the group consisting of: (a) nucleic acid molecules which hybridize under stringent conditions to a molecule consisting of a nucleotide sequence set forth as SEQ ID NO:1 and which code for a Formylglycine Generating Enzyme (FGE) polypeptide having C_α-formylglycine generating activity, (b) nucleic acid molecules that differ from the nucleic acid molecules of (a) in codon sequence due to the degeneracy of the genetic code, and (c) complements of (a) or (b), is provided. In certain embodiments, the isolated nucleic acid molecule comprises the nucleotide sequence set forth as SEQ ID NO:1. In some

embodiments, the isolated nucleic acid molecule consists of the nucleotide sequence set forth as SEQ ID NO:3 or a fragment thereof.

The invention in another aspect provides an isolated nucleic acid molecule selected from the group consisting of (a) unique fragments of a nucleotide sequence set forth as SEQ ID NO:1, and (b) complements of (a), provided that a unique fragment of (a) includes a sequence of contiguous nucleotides which is not identical to any sequence selected from the sequence group consisting of: (1) sequences identical to SEQ ID NO. 4 and/or nucleotides 20-1141 of SEQ ID NO. 4, and (2) complements of (1). In any of the foregoing embodiments, complements refer to full-length complements.

In one embodiment, the sequence of contiguous nucleotides is selected from the group consisting of (1) at least two contiguous nucleotides nonidentical to the sequence group, (2) at least three contiguous nucleotides nonidentical to the sequence group, (3) at least four contiguous nucleotides nonidentical to the sequence group, (4) at least five contiguous nucleotides nonidentical to the sequence group, (5) at least six contiguous nucleotides nonidentical to the sequence group, and (6) at least seven contiguous nucleotides nonidentical to the sequence group.

In another embodiment, the fragment has a size selected from the group consisting of at least: 8 nucleotides, 10 nucleotides, 12 nucleotides, 14 nucleotides, 16 nucleotides, 18 nucleotides, 20, nucleotides, 22 nucleotides, 24 nucleotides, 26 nucleotides, 28 nucleotides, 30 nucleotides, 40 nucleotides, 50 nucleotides, 75 nucleotides, 100 nucleotides, 200 nucleotides, 1000 nucleotides and every integer length therebetween.

According to another aspect, the invention provides expression vectors, and host cells transformed or transfected with such expression vectors, comprising the nucleic acid molecules described above.

According to still another aspect, the invention provides cells expressing activated forms of the endogenous FGE gene. In one embodiment, activation of the endogenous FGE gene occurs via homologous recombination.

According to another aspect of the invention, an isolated polypeptide is provided. The isolated polypeptide is encoded by the foregoing nucleic acid molecules of the invention. In some embodiments, the isolated polypeptide is encoded by the nucleic acid of SEQ ID NO:1, giving rise to a polypeptide having the sequence of SEQ ID NO:2 that has C_α-formylglycine generating activity. In other embodiments, the isolated polypeptide may be a fragment or variant of the foregoing of sufficient length to represent a sequence unique within the human genome, and identifying with a polypeptide that has C_α-formylglycine generating

activity, provided that the fragment includes a sequence of contiguous amino acids which is not identical to any sequence encoded for by a nucleic acid sequence having SEQ ID NO. 4. In another embodiment, immunogenic fragments of the polypeptide molecules described above are provided. The immunogenic fragments may or may not have C_α-formylglycine generating activity.

According to another aspect of the invention, isolated binding polypeptides are provided which selectively bind a polypeptide encoded by the foregoing nucleic acid molecules of the invention. Preferably the isolated binding polypeptides selectively bind a polypeptide which comprises the sequence of SEQ ID NO:2, fragments thereof, or a polypeptide belonging to the family of isolated polypeptides having C_α-formylglycine generating activity described elsewhere herein. In preferred embodiments, the isolated binding polypeptides include antibodies and fragments of antibodies (e.g., Fab, $F(ab)_2$, Fd and antibody fragments which include a CDR3 region which binds selectively to the FGE polypeptide). In certain embodiments, the antibodies are human. In some embodiments, the antibodies are monoclonal antibodies. In one embodiment, the antibodies are polyclonal antisera. In further embodiments, the antibodies are humanized. In yet further embodiments, the antibodies are chimeric.

According to another aspect of the invention, a family of isolated polypeptides having C_α-formylglycine generating activity, are provided. Each of said polypeptides comprises from amino terminus to carboxyl terminus: (a) an amino-terminal subdomain 1; a subdomain 2; a carboxy-terminal subdomain 3 containing from 35 to 45 amino acids; and wherein subdomain 3 has at least about 75% homology and a length approximately equal to subdomain 3 of a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78. In important embodiments, subdomain 2 contains from 120 to 140 amino acids. In further important embodiments, at least 5% of the amino acids of subdomain 2 are Tryptophans. In some embodiments, subdomain 2 has at least about 50% homology to subdomain 2 of a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78. In certain embodiments, subdomain 3 of each of the polypeptides has at least between about 80% and about 100% homology to subdomain 3 of a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

According to a further aspect of the invention, a method for determining the level of FGE expression in a subject, is provided. The method involves measuring expression of FGE

in a test sample from a subject to determine the level of FGE expression in the subject. In certain embodiments, the measured FGE expression in the test sample is compared to FGE expression in a control containing a known level of FGE expression. Expression is defined as FGE mRNA expression, FGE polypeptide expression, or FGE C_α-formylglycine generating activity as defined elsewhere herein. Various methods can be used to measure expression. Preferred embodiments of the invention include PCR and Northern blotting for measuring mRNA expression, FGE monoclonal antibodies or FGE polyclonal antisera as reagents to measure FGE polypeptide expression, as well as methods for measuring FGE C_α-formylglycine generating activity.

In certain embodiments, test samples such as biopsy samples, and biological fluids such as blood, are used as test samples. FGE expression in a test sample of a subject is compared to FGE expression in control.

According to another aspect of the invention, a method for identifying an agent useful in modulating C_α-formylglycine generating activity of a molecule, is provided. The method involves (a) contacting a molecule having C_α-formylglycine generating activity with a candidate agent, (b) measuring C_α-formylglycine generating activity of the molecule, and (c) comparing the measured C_α-formylglycine generating activity of the molecule to a control to determine whether the candidate agent modulates C_α-formylglycine generating activity of the molecule, wherein the molecule is a nucleic acid molecule having the nucleotide sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87, or an expression product thereof (e.g., a peptide having a sequence selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78). In certain embodiments, the control is C_α-formylglycine generating activity of the molecule measured in the absence of the candidate agent.

According to still another aspect of the invention, a method of diagnosing Multiple Sulfatase Deficiency in a subject, is provided. The method involves contacting a biological sample from a subject suspected of having Multiple Sulfatase Deficiency with an agent, said agent specifically binding to a molecule selected from the group consisting of: (i) a FGE nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, 3, or 4, (ii) an expression product of the nucleic acid molecule of (i), or (iii) a fragment of the expression product of (ii); and measuring the amount of bound agent and determining therefrom if the

expression of said nucleic acid molecule or of an expression product thereof is aberrant, aberrant expression being diagnostic of the Multiple Sulfatase Deficiency in the subject.

According to still another aspect of the invention, a method for diagnosing a condition characterized by aberrant expression of a nucleic acid molecule or an expression product thereof, is provided. The method involves contacting a biological sample from a subject with an agent, wherein said agent specifically binds to said nucleic acid molecule, an expression product thereof, or a fragment of an expression product thereof; and measuring the amount of bound agent and determining therefrom if the expression of said nucleic acid molecule or of an expression product thereof is aberrant, aberrant expression being diagnostic of the condition, wherein the nucleic acid molecule has the nucleotide sequence of SEQ ID NO:1 and the condition is Multiple Sulfatase Deficiency.

According to another aspect of the invention, a method for determining Multiple Sulfatase Deficiency in a subject characterized by aberrant expression of a nucleic acid molecule or an expression product thereof, is provided. The method involves monitoring a sample from a patient for a parameter selected from the group consisting of (i) a nucleic acid molecule having the nucleotide sequence of SEQ ID NO:1, 3, 4, or a nucleic acid molecule having a sequence derived from the FEG genomic locus, (ii) a polypeptide encoded by the nucleic acid molecule, (iii) a peptide derived from the polypeptide, and (iv) an antibody which selectively binds the polypeptide or peptide, as a determination of Multiple Sulfatase Deficiency in the subject. In some embodiments, the sample is a biological fluid or a tissue as described in any of the foregoing embodiments. In certain embodiments the step of monitoring comprises contacting the sample with a detectable agent selected from the group consisting of (a) an isolated nucleic acid molecule which selectively hybridizes under stringent conditions to the nucleic acid molecule of (i), (b) an antibody which selectively binds the polypeptide of (ii), or the peptide of (iii), and (c) a polypeptide or peptide which binds the antibody of (iv). The antibody, polypeptide, peptide, or nucleic acid can be labeled with a radioactive label or an enzyme. In further embodiments, the method further comprises assaying the sample for the peptide.

According to another aspect of the invention, a kit is provided. The kit comprises a package containing an agent that selectively binds to any of the foregoing FGE isolated nucleic acids, or expression products thereof, and a control for comparing to a measured value of binding of said agent any of the foregoing FGE isolated nucleic acids or expression products thereof. In some embodiments, the control is a predetermined value for comparing to the measured value. In certain embodiments, the control comprises an epitope of the

expression product of any of the foregoing FGE isolated nucleic acids. In one embodiment, the kit further comprises a second agent that selectively binds to a polypeptide selected from the group consisting of Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, or a peptide thereof, and a control for comparing to a measured value of binding of said second agent to said polypeptide or peptide thereof.

According to a further aspect of the invention, a method of treating Multiple Sulfatase Deficiency, is provided. The method involves administering to a subject in need of such treatment an agent that modulates C_α-formylglycine generating activity, in an amount effective to treat Multiple Sulfatase Deficiency in the subject. In some embodiments, the method further comprises co-administering an agent selected from the group consisting of a nucleic acid molecule encoding Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, or HSulf-6, an expression product of the nucleic acid molecule, and a fragment of the expression product of the nucleic acid molecule. In certain embodiments, the agent that modulates C_α-formylglycine generating activity is an isolated nucleic acid molecule of the invention (e.g., a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87). In important embodiments, the agent that modulates C_α-formylglycine generating activity is a peptide of the invention (e.g., a peptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78). The agent that modulates C_α-formylglycine generating activity may be produced by a cell expressing an endogenous and/or exogenous FGE nucleic acid molecule. In important embodiments, the endogenous FGE nucleic acid molecule may be activated.

According to one aspect of the invention, a method for for increasing C_α-formylglycine generating activity in a subject, is provided. The method involves administering an isolated FGE nucleic acid molecule of the invention (e.g., a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71,

73, 75, 77, and 80-87), and/or an expression product thereof, to a subject, in an amount effective to increase C_α-formylglycine generating activity in the subject.

According to one aspect of the invention, a method for treating a subject with Multiple Sulfatase Deficiency, is provided. The method involves administering to a subject in need of such treatment an agent that modulates C_α-formylglycine generating activity, in an amount effective to increase C_α-formylglycine generating activity in the subject. In some embodiments, the agent that modulates C_α-formylglycine generating activity is a sense nucleic acid of the invention (e.g., a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87). In certain embodiments, the agent that modulates C_α-formylglycine generating activity is an isolated polypeptide of the invention (e.g., a polypeptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78).

According to still another aspect of the invention, a method for increasing C_α-formylglycine generating activity in a cell, is provided. The method involves contacting the cell with an isolated nucleic acid molecule of the invention (e.g., a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87), or an expression product thereof, in an amount effective to increase C_α-formylglycine generating activity in the cell. In important embodiments, the method involves activating the endogenous FGE gene to increase C_α-formylglycine generating activity in the cell.

According to a further aspect of the invention, a pharmaceutical composition is provided. The composition comprises an agent comprising an isolated nucleic acid molecule of the invention (e.g., an isolated nucleic acid molecule as claimed in any one of Claims 1-8, an FGE nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87), or an expression product thereof, in a pharmaceutically effective amount to treat Multiple Sulfatase Deficiency, or an expression product thereof, in a pharmaceutically effective amount to treat Multiple Sulfatase Deficiency, and a pharmaceutically acceptable carrier.

According to one aspect of the invention, a method for identifying a candidate agent useful in the treatment of Multiple Sulfatase Deficiency, is provided. The method involves determining expression of a set of nucleic acid molecules in a cell or tissue under conditions

which, in the absence of a candidate agent, permit a first amount of expression of the set of nucleic acid molecules, wherein the set of nucleic acid molecules comprises at least one nucleic acid molecule selected from the group consisting of: (a) nucleic acid molecules which hybridize under stringent conditions to a molecule consisting of a nucleotide sequence set forth as SEQ ID NO:1 and which code for a polypeptide having C_α-formylglycine generating activity (FGE), (b) nucleic acid molecules that differ from the nucleic acid molecules of (a) or (b) in codon sequence due to the degeneracy of the genetic code, (c) a nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87, and (d) complements of (a) or (b) or (c), contacting the cell or tissue with the candidate agent, and detecting a test amount of expression of the set of nucleic acid molecules, wherein an increase in the test amount of expression in the presence of the candidate agent relative to the first amount of expression indicates that the candidate agent is useful in the treatment of the Multiple Sulfatase Deficiency.

According to a further aspect of the invention, methods for preparing medicaments useful in the treatment of Multiple Sulfatase Deficiency and/or other sulfatase deficiencies, are provided.

According to still another aspect of the invention, a solid-phase nucleic acid molecule array, is provided. The array consists essentially of a set of nucleic acid molecules, expression products thereof, or fragments (of either the nucleic acid or the polypeptide molecule) thereof, each nucleic acid molecule encoding for a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, fixed to a solid substrate. In some embodiments, the solid-phase array further comprises at least one control nucleic acid molecule. In certain embodiments, the set of nucleic acid molecules comprises at least one, at least two, at least three, at least four, or even at least five nucleic acid molecules, each selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6.

According to a further aspect of the invention, a method for treating a sulfatase deficiency in a subject, is provided. The method involves administering to a subject in need of such treatment a sulfatase that has been produced according to the invention, in an amount effective to treat the sulfatase deficiency in the subject and the sulfatase deficiency is not Multiple Sulfatase Deficiency. In important embodiments, the sulfatase is produced by a cell that has been contacted with an an agent that modulates C_α-formylglycine generating activity. In certain embodiments, the sulfatase deficiency includes, but is not limited to, Mucopolysaccharidosis II (MPS II; Hunter Syndrome), Mucopolysaccharidosis IIIA (MPS IIIA; Sanfilippo Syndrome A), Mucopolysaccharidosis VIII (MPS VIII), Mucopolysaccharidosis IVA (MPS IVA; Morquio Syndrome A), Mucopolysaccharidosis VI (MPS VI; Maroteaux-Lamy Syndrome), Metachromatic Leukodystrophy (MLD), X-linked Recessive Chondrodysplasia Punctata 1, or X-linked Ichthyosis (Steroid Sulfatase Deficiency). In certain embodiments, the agent that modulates C_α-formylglycine generating activity can be a nucleic acid molecule or peptide of the invention. In one embodiment, the sulfatase and the agent that modulates C_α-formylglycine generating activity are co-expressed in the same cell. The sulfatase and/or the agent that modulates C_α-formylglycine generating activity can be endogenous or exogenous in origin. If endogenous in origin it can be activated (e.g., by insertion of strong promoter and/or other elements at the appropriates places known in the art). If exogenous, its expression can be driven by elements on the expression vector, or it can be targeted to appropriated places within the cell genome that will allow for its enhanced expression (e.g., downstream of a strong promoter).

According to another aspect of the invention, a pharmaceutical composition, is provided. The composition comprises an agent comprising an isolated nucleic acid molecule of the invention, or an expression product thereof, in a pharmaceutically effective amount to treat a sulfatase deficiency, and a pharmaceutically acceptable carrier.

According to a still further aspect of the invention, a method for increasing sulfatase activity in a cell, is provided. The method involves contacting a cell expressing a sulfatase with an isolated nucleic acid molecule of of the invention (e.g., an isolated nucleic acid molecule as claimed in any one of Claims 1-8, an FGE nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87), or an expression product thereof (e.g., a polypeptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78), in an amount effective to increase sulfatase activity in the cell. The cell may

express an endogenous and/or an exogenous sulfatase. In important embodiments, the endogenous sulfatase is activated. In certain embodiments, the sulfatase is Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and/or HSulf-6. In certain embodiments the cell is a mammalian cell.

According to another aspect of the invention, a pharmaceutical composition, is provided. The composition comprises a sulfatase that is produced by cell, in a pharmaceutically effective amount to treat a sulfatase deficiency, and a pharmaceutically acceptable carrier, wherein said cell has been contacted with an agent comprising an isolated nucleic acid molecule of the invention (e.g., as claimed in Claims 1-8, or a nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87), or an expression product thereof (e.g., a peptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78).

According to still another aspect of the invention, an isolated variant allele of a human FGE gene which encodes a variant FGE polypeptide, is provided. The isolated variant allele comprises an amino acid sequence comprising at least one variation in SEQ ID NO:2, wherein the at least one variation comprises: Met1Arg; Met1Val; Leu20Phe; Ser155Pro; Ala177Pro; Cys218Tyr; Arg224Trp; Asn259Ile; Pro266Leu; Ala279Val; Arg327Stop; Cys336Arg; Arg345Cys; Ala348Pro; Arg349Gln; Arg349Trp; Arg349Trp; Ser359Stop; or a combination thereof.

According to yet another aspect of the invention, an isolated variant human FGE polypeptide, is provided. The isolated variant human FGE polypeptide comprises an amino acid sequence comprising at least one variation in SEQ ID NO:2, wherein the at least one variation comprises: Met1Arg; Met1Val; Leu20Phe; Ser155Pro; Ala177Pro; Cys218Tyr; Arg224Trp; Asn259Ile; Pro266Leu; Ala279Val; Arg327Stop; Cys336Arg; Arg345Cys; Ala348Pro; Arg349Gln; Arg349Trp; Arg349Trp; Ser359Stop; or a combination thereof.

Antibodies having any of the foregoing variant human FGE polypeptides as an immunogen are also provided. Such antibodies include polyclonal antisera, monoclonal, chimeric, and can also be detectably labeled. A detectable label may comprise a radioactive element, a chemical which fluoresces, or an enzyme.

According to another aspect of the invention, a sulfatase-producing cell wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased, is provided. The

cell comprises: (i) a sulfatase with an increased expression, and (ii) a Formylglycine Generating Enzyme with an increased expression, wherein the ratio of active sulfatase to total sulfatase (i.e., the specific activity of the sulfatase) produced by the cell is increased by at least 5% over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme. In certain embodiments, the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 10%, 15%, 20%, 50%, 100%, 200%, 500%, 1000%, over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme.

According to a further aspect of the invention, an improved method for treating a sulfatase deficiency in a subject is provided. The method involves administering to a subject in need of such treatment a sulfatase in an effective amount to treat the sulfatase deficiency in the subject, wherein the sulfatase is contacted with a Formylglycine Generating Enzyme in an amount effective to increase the specific activity of the sulfatase. In an important embodiment, the sulfatase is selected from the group consisting of Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6. In certain embodiments, the Formylglycine Generating Enzyme is encoded by a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87. In some embodiments, the Formylglycine Generating Enzyme is a peptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

These and other objects of the invention will be described in further detail in connection with the detailed description of the invention.

Brief Description of the Sequences

SEQ ID NO:1 is the nucleotide sequence of the human FGE cDNA.

SEQ ID NO:2 is the predicted amino acid sequence of the translation product of human FGE cDNA (SEQ ID NO:1).

SEQ ID NO:3 is the nucleotide sequence of the human FGE cDNA encoding the polypeptide of SEQ ID NO:2 (i.e., nucleotides 20-1141 of SEQ ID NO:1).

SEQ ID NO:4 is the nucleotide sequence of GenBank Acc. No. AK075459.

SEQ ID NO:5 is the predicted amino acid sequence of the translation product of SEQ ID NO:4, an unnamed protein product having GenBank Acc.No. BAC11634.

SEQ ID NO:6 is the nucleotide sequence of the human Iduronate 2-Sulfatase cDNA (GenBank Acc. No. M58342).

SEQ ID NO:7 is the predicted amino acid sequence of the translation product of human Iduronate 2-Sulfatase cDNA (SEQ ID NO:6).

SEQ ID NO:8 is the nucleotide sequence of the human Sulfamidase cDNA (GenBank Acc. No. U30894).

SEQ ID NO:9 is the predicted amino acid sequence of the translation product of human Sulfamidase cDNA (SEQ ID NO:8).

SEQ ID NO:10 is the nucleotide sequence of the human N-Acetylgalactosamine 6-Sulfatase cDNA (GenBank Acc. No. U06088).

SEQ ID NO:11 is the predicted amino acid sequence of the translation product of human N-Acetylgalactosamine 6-Sulfatase cDNA (SEQ ID NO:10).

SEQ ID NO:12 is the nucleotide sequence of the human N-Acetylglucosamine 6-Sulfatase cDNA (GenBank Acc. No. Z12173).

SEQ ID NO:13 is the predicted amino acid sequence of the translation product of human N-Acetylglucosamine 6-Sulfatase cDNA (SEQ ID NO:12).

SEQ ID NO:14 is the nucleotide sequence of the human Arylsulfatase A cDNA (GenBank Acc. No. X52151).

SEQ ID NO:15 is the predicted amino acid sequence of the translation product of human Arylsulfatase A cDNA (SEQ ID NO:14).

SEQ ID NO:16 is the nucleotide sequence of the human Arylsulfatase B cDNA (GenBank Acc. No. J05225).

SEQ ID NO:17 is the predicted amino acid sequence of the translation product of human Arylsulfatase B cDNA (SEQ ID NO:16).

SEQ ID NO:18 is the nucleotide sequence of the human Arylsulfatase C cDNA (GenBank Acc. No. J04964).

SEQ ID NO:19 is the predicted amino acid sequence of the translation product of human Arylsulfatase C cDNA (SEQ ID NO:18).

SEQ ID NO:20 is the nucleotide sequence of the human Arylsulfatase D cDNA (GenBank Acc. No. X83572).

SEQ ID NO:21 is the predicted amino acid sequence of the translation product of human Arylsulfatase D cDNA (SEQ ID NO:20).

SEQ ID NO:22 is the nucleotide sequence of the human Arylsulfatase E cDNA (GenBank Acc. No. X83573).

SEQ ID NO:23 is the predicted amino acid sequence of the translation product of human Arylsulfatase E cDNA (SEQ ID NO:22).

SEQ ID NO:24 is the nucleotide sequence of the human Arylsulfatase F cDNA (GenBank Acc. No. X97868).

SEQ ID NO:25 is the predicted amino acid sequence of the translation product of human Arylsulfatase F cDNA (SEQ ID NO:24).

SEQ ID NO:26 is the nucleotide sequence of the human Arylsulfatase G cDNA (GenBank Acc.No. BC012375).

SEQ ID NO:27 is the predicted amino acid sequence of the translation product of the human Arylsulfatase G (SEQ ID NO:26).

SEQ ID NO:28 is the nucleotide sequence of the HSulf-1 cDNA (GenBank Acc.No. AY101175).

SEQ ID NO:29 is the predicted amino acid sequence of the translation product of HSulf-1 cDNA (SEQ ID NO:28).

SEQ ID NO:30 is the nucleotide sequence of the HSulf-2 cDNA (GenBank Acc.No. AY101176).

SEQ ID NO:31 is the predicted amino acid sequence of the translation product of HSulf-2 cDNA (SEQ ID NO:30).

SEQ ID NO:32 is the highly conserved hexapeptide L/V-FGly-X-P-S-R present on sulfatases.

SEQ ID NO:33 is a synthetic *FGly* formation substrate; its primary sequence is derived from human Arylsulfatase A.

SEQ ID NO:34 is scrambled oligopeptide PVSLPTRSCAALLTGR.

SEQ ID NO:35 is Ser69 oligopeptide PVSLSTPSRAALLTGR.

SEQ ID NO:36 is human FGE-specific primer 1199nc.

SEQ ID NO:37 is human FGE-specific forward primer 1c.

SEQ ID NO:38 is human FGE-specific reverse primer 1182c.

SEQ ID NO:39 is human 5'- FGE-specific primer containing EcoRI site.

SEQ ID NO:40 is a HA-specific primer.

SEQ ID NO:41 is a c-myc -specific primer.

SEQ ID NO:42 is a RGS-His_6 - specific primer.

SEQ ID NO:43 is tryptic oligopeptide SQNTPDSSASNLGFR from a human FGE preparation.

SEQ ID NO:44 is tryptic oligopeptide MVPIPAGVFTMGTDDPQIK from a human FGE preparation.

SEQ ID NO:45 is the nucleotide sequence of the human FGE2 paralog (GenBank GI: 24308053).

SEQ ID NO:46 is the predicted amino acid sequence of the translation product of the human FGE2 paralog (SEQ ID NO:45).

SEQ ID NO:47 is the nucleotide sequence of the mouse FGE paralog (GenBank GI: 26344956).

SEQ ID NO:48 is the predicted amino acid sequence of the translation product of the mouse FGE paralog (SEQ ID NO:47).

SEQ ID NO:49 is the nucleotide sequence of the mouse FGE ortholog (GenBank GI: 22122361).

SEQ ID NO:50 is the predicted amino acid sequence of the translation product of the mouse FGE ortholog (SEQ ID NO:49).

SEQ ID NO:51 is the nucleotide sequence of the fruitfly FGE ortholog (GenBank GI: 20130397).

SEQ ID NO:52 is the predicted amino acid sequence of the translation product of the fruitfly FGE ortholog (SEQ ID NO:51).

SEQ ID NO:53 is the nucleotide sequence of the mosquito FGE ortholog (GenBank GI: 21289310).

SEQ ID NO:54 is the predicted amino acid sequence of the translation product of the mosquito FGE ortholog (SEQ ID NO:53).

SEQ ID NO:55 is the nucleotide sequence of the closely related S. coelicolor FGE ortholog (GenBank GI: 21225812).

SEQ ID NO:56 is the predicted amino acid sequence of the translation product of the S. coelicolor FGE ortholog (SEQ ID NO:55).

SEQ ID NO:57 is the nucleotide sequence of the closely related C. efficiens FGE ortholog (GenBank GI: 25028125).

SEQ ID NO:58 is the predicted amino acid sequence of the translation product of the C. efficiens FGE ortholog (SEQ ID NO:57).

SEQ ID NO:59 is the nucleotide sequence of the N. aromaticivorans FGE ortholog (GenBank GI: 23108562).

SEQ ID NO:60 is the predicted amino acid sequence of the translation product of the N. aromaticivorans FGE ortholog (SEQ ID NO:59).

SEQ ID NO:61 is the nucleotide sequence of the M. loti FGE ortholog (GenBank GI: 13474559).

SEQ ID NO:62 is the predicted amino acid sequence of the translation product of the M. loti FGE ortholog (SEQ ID NO:61).

SEQ ID NO:63 is the nucleotide sequence of the B. fungorum FGE ortholog (GenBank GI: 22988809).

SEQ ID NO:64 is the predicted amino acid sequence of the translation product of the B. fungorum FGE ortholog (SEQ ID NO:63).

SEQ ID NO:65 is the nucleotide sequence of the S. meliloti FGE ortholog (GenBank GI: 16264068).

SEQ ID NO:66 is the predicted amino acid sequence of the translation product of the S. meliloti FGE ortholog (SEQ ID NO:65).

SEQ ID NO:67 is the nucleotide sequence of the Microscilla sp. FGE ortholog (GenBank GI: 14518334).

SEQ ID NO:68 is the predicted amino acid sequence of the translation product of the Microscilla sp. FGE ortholog (SEQ ID NO:67).

SEQ ID NO:69 is the nucleotide sequence of the P. putida KT2440 FGE ortholog (GenBank GI: 26990068).

SEQ ID NO:70 is the predicted amino acid sequence of the translation product of the P. putida KT2440 FGE ortholog (SEQ ID NO:69).

SEQ ID NO:71 is the nucleotide sequence of the R. metallidurans FGE ortholog (GenBank GI: 22975289).

SEQ ID NO:72 is the predicted amino acid sequence of the translation product of the R. metallidurans FGE ortholog (SEQ ID NO:71).

SEQ ID NO:73 is the nucleotide sequence of the P. marinus FGE ortholog (GenBank GI: 23132010).

SEQ ID NO:74 is the predicted amino acid sequence of the translation product of the P. marinus FGE ortholog (SEQ ID NO:73).

SEQ ID NO:75 is the nucleotide sequence of the C. crescentus CB15 FGE ortholog (GenBank GI: 16125425).

SEQ ID NO:76 is the predicted amino acid sequence of the translation product of the C. crescentus CB15 FGE ortholog (SEQ ID NO:75).

SEQ ID NO:77 is the nucleotide sequence of the M. tuberculosis Ht37Rv FGE ortholog (GenBank GI: 15607852).

SEQ ID NO:78 is the predicted amino acid sequence of the translation product of the M. tuberculosis Ht37Rv FGE ortholog (SEQ ID NO:77).

SEQ ID NO:79 is the highly conserved heptapeptide present on subdomain 3 of FGE orthologs and paralogs.

SEQ ID NO:80 is the nucleotide sequence of FGE ortholog EST fragment having GenBank Acc. No.: CA379852.

SEQ ID NO:81 is the nucleotide sequence of FGE ortholog EST fragment having GenBank Acc. No.: AI721440.

SEQ ID NO:82 is the nucleotide sequence of FGE ortholog EST fragment having GenBank Acc. No.: BJ505402.

SEQ ID NO:83 is the nucleotide sequence of FGE ortholog EST fragment having GenBank Acc. No.: BJ054666.

SEQ ID NO:84 is the nucleotide sequence of FGE ortholog EST fragment having GenBank Acc. No.: AL892419.

SEQ ID NO:85 is the nucleotide sequence of FGE ortholog EST fragment having GenBank Acc. No.: CA064079.

SEQ ID NO:86 is the nucleotide sequence of FGE ortholog EST fragment having GenBank Acc. No.: BF189614.

SEQ ID NO:87 is the nucleotide sequence of FGE ortholog EST fragment having GenBank Acc. No.: AV609121.

SEQ ID NO:88 is the nucleotide sequence of the HSulf-3 cDNA.

SEQ ID NO:89 is the predicted amino acid sequence of the translation product of HSulf-3 cDNA (SEQ ID NO:88).

SEQ ID NO:90 is the nucleotide sequence of the HSulf-4 cDNA.

SEQ ID NO:91 is the predicted amino acid sequence of the translation product of HSulf-4 cDNA (SEQ ID NO:90).

SEQ ID NO:92 is the nucleotide sequence of the HSulf-5 cDNA.

SEQ ID NO:93 is the predicted amino acid sequence of the translation product of HSulf-5 cDNA (SEQ ID NO:92).

SEQ ID NO:94 is the nucleotide sequence of the HSulf-6 cDNA.

SEQ ID NO:95 is the predicted amino acid sequence of the translation product of HSulf-6 cDNA (SEQ ID NO:94).

Brief Description of the Drawings

Fig. 1: A MALDI-TOF mass spectra schematic of P23 after incubation in the absence (A) or presence (B) of a soluble extract from bovine testis microsomes.

Fig. 2: A phylogenetic tree derived from an alignment of human FGE and 21 proteins of the PFAM-DUF323 seed.

Fig. 3: Organisation of the human and murine FGE gene locus. Exons are shown to scale as boxes and bright boxes (murine locus). The numbers above the intron lines indicate the size of the introns in kilobases.

Fig. 4: Diagram showing a map of FGE Expression Plasmid pXMG.1.3

Fig. 5: Bar graph depicting N-Acetylgalactosamine 6-Sulfatase Activity in 36F Cells Transiently Transfected with FGE Expression Plasmid.

Fig. 6: Bar graph depicting N-Acetylgalactosamine 6-Sulfatase *Specific* Activity in 36F Cells Transiently Transfected with FGE Expression Plasmid.

Fig. 7: Bar graph depicting N-Acetylgalactosamine 6-Sulfatase Production in 36F Cells Transiently Transfected with FGE Expression Plasmid.

Fig. 8: Graph depicting Iduronate 2-Sulfatase Activity in 30C6 Cells Transiently Transfected with FGE Expression Plasmid.

Fig. 9: Depicts a kit embodying features of the present invention.

Detailed Description of the Invention

The invention involves the discovery of the gene that encodes Formylglycine Generating Enzyme (FGE), an enzyme responsible for the unique post-translational modification occurring on sulfatases that is essential for sulfatase function: the formation of L-C_α-formylglycine (a.k.a. *FGly* and/or *2-amino-3-oxopropanoic acid*). It has been discovered, unexpectedly, that mutations in the FGE gene lead to the development of Multiple Sulfatase Deficiency (MSD) in subjects. It has also been discovered, unexpectedly, that FGE enhances the activity of sulfatases, including, but not limited to, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, and sulfatases described in U.S. Provisional applications with publication numbers 20030073118, 20030147875, 20030148920, 20030162279, and 20030166283 (the contents of which are expressly incorporated herein). In view of these discoveries, the molecules of

the present invention can be used in the diagnosis and/or treatment of Multiple Sulfatase Deficiency, as well as the treatment of other sulfatase deficiencies.

Methods for using the molecules of the invention in the diagnosis of Multiple Sulfatase Deficiency are provided.

Additionally, methods for using these molecules *in vivo* or *in vitro* for the purpose of modulating *FGly* formation on sulfatases, methods for treating conditions associated with such modification, and compositions useful in the preparation of therapeutic preparations for the treatment of Multiple Sulfatase Deficiency as well as other sulfatase deficiencies, are also provided.

The present invention thus involves, in several aspects, polypeptides modulating *FGly* formation on sulfatases, isolated nucleic acids encoding those polypeptides, functional modifications and variants of the foregoing, useful fragments of the foregoing, as well as therapeutics and diagnostics, research methods, compositions and tools relating thereto.

"C_α-formylglycine generating activity" refers to the ability of a molecule to form, or enhance the formation of, *FGly* on a substrate. The substate may be a sulfatase as described elsewhere herein, or a synthetic oligopeptide (see, e.g., SEQ ID NO:33, and the Examples). The substrate preferably contains the conserved hexapeptide of SEQ ID NO:32 [L/V-C(S)-X-P-S-R]. Methods for assaying *FGly* formation are as described in the art (see, e.g., Dierks, T., et al., *Proc. Natl. Acad. Sci. U. S. A.*, 1997, 94:11963-11968), and elsewhere herein (see, e.g., the Examples). A "molecule," as used herein, embraces both "nucleic acids" and "polypeptides." FGE molecules are capable of forming, or enhancing/increasing formation of, *FGly* both *in vivo* and *in vitro*.

"Enhancing (or "increasing")" C_α-formylglycine generating activity, as used herein, typically refers to increased expression of FGE and/or its encoded polypeptide. Increased expression refers to increasing (i.e., to a detectable extent) replication, transcription, and/or translation of any of the nucleic acids of the invention (FGE nucleic acids as described elsewhere herein), since upregulation of any of these processes results in concentration/amount increase of the polypeptide encoded by the gene (nucleic acid). Enhancing (or increasing) C_α-formylglycine generating activity also refers to preventing or inhibiting FGE degradation (e.g., *via* increased ubiquitinization), downregulation, etc., resulting, for example, in increased or stable FGE molecule $t_{1/2}$ (half-life) when compared to a control. Downregulation or decreased expression refers to decreased expression of a gene and/or its encoded polypeptide. The upregulation or downregulation of gene expression can be directly determined by detecting an increase or decrease, respectively, in the level of

mRNA for the gene (e.g, FGE), or the level of protein expression of the gene-encoded polypeptide, using any suitable means known to the art, such as nucleic acid hybridization or antibody detection methods, respectively, and in comparison to controls. Upregulation or downregulation of FGE gene expression can also be determined indirectly by detecting a change in C_α-formylglycine generating activity.

"Expression," as used herein, refers to nucleic acid and/or polypeptide expression, as well as to activity of the polypeptide molecule (e.g., C_α-formylglycine generating activity of the molecule).

One aspect of the invention involves the cloning of a cDNA encoding FGE. FGE according to the invention is an isolated nucleic acid molecule that comprises a nucleic acid molecule of SEQ ID NO:1, and codes for a polypeptide with C_α-formylglycine generating activity. The sequence of the human FGE cDNA is presented as SEQ ID NO:1, and the predicted amino acid sequence of this cDNA's encoded protein product is presented as SEQ ID NO:2.

As used herein, a subject is a mammal or a non-human mammal. In all embodiments human FGE and human subjects are preferred.

The invention thus involves in one aspect an isolated FGE polypeptide, the cDNA encoding this polypeptide, functional modifications and variants of the foregoing, useful fragments of the foregoing, as well as diagnostics and therapeutics relating thereto.

As used herein with respect to nucleic acids, the term "isolated" means: (i) amplified *in vitro* by, for example, polymerase chain reaction (PCR); (ii) recombinantly produced by cloning; (iii) purified, as by cleavage and gel separation; or (iv) synthesized by, for example, chemical synthesis. An isolated nucleic acid is one which is readily manipulated by recombinant DNA techniques well known in the art. Thus, a nucleotide sequence contained in a vector in which 5' and 3' restriction sites are known or for which polymerase chain reaction (PCR) primer sequences have been disclosed is considered isolated but a nucleic acid sequence existing in its native state in its natural host is not. An isolated nucleic acid may be substantially purified, but need not be. For example, a nucleic acid that is isolated within a cloning or expression vector is not pure in that it may comprise only a tiny percentage of the material in the cell in which it resides. Such a nucleic acid is isolated, however, as the term is used herein because it is readily manipulated by standard techniques known to those of ordinary skill in the art.

As used herein with respect to polypeptides, the term "isolated" means separated from its native environment in sufficiently pure form so that it can be manipulated or used for any

one of the purposes of the invention. Thus, isolated means sufficiently pure to be used (i) to raise and/or isolate antibodies, (ii) as a reagent in an assay, (iii) for sequencing, (iv) as a therapeutic, etc.

According to the invention, isolated nucleic acid molecules that code for a FGE polypeptide having C_α-formylglycine generating activity include: (a) nucleic acid molecules which hybridize under stringent conditions to a molecule consisting of a nucleic acid of SEQ ID NO:1 and which code for a FGE polypeptide having C_α-formylglycine generating activity, (b) deletions, additions and substitutions of (a) which code for a respective FGE polypeptide having C_α-formylglycine generating activity, (c) nucleic acid molecules that differ from the nucleic acid molecules of (a) or (b) in codon sequence due to the degeneracy of the genetic code, and (d) complements of (a), (b) or (c). "Complements," as used herein, includes "full-length complementary strands or 100% complementary strands of (a), (b) or (c).

Homologs and alleles of the FGE nucleic acids of the invention also having C_α-formylglycine generating activity are encompassed by the present invention. Homologs, as described herein, include the molecules identified elsewhere herein (see e.g., SEQ ID NOs:4, 5, 45-78, and 80-87) i.e. orthologs and paralogs. Further homologs can be identified following the teachings of the present invention as well as by conventional techniques. Since the FGE homologs described herein all share C_α-formylglycine generating activity, they can be used interchangeably with the human FGE molecule in all aspects of the invention.

Thus, an aspect of the invention is those nucleic acid sequences which code for FGE polypeptides and which hybridize to a nucleic acid molecule consisting of the coding region of SEQ ID NO:1, under stringent conditions. In an important embodiment, the term "stringent conditions," as used herein, refers to parameters with which the art is familiar. With nucleic acids, hybridization conditions are said to be stringent typically under conditions of low ionic strength and a temperature just below the melting temperature (T_m) of the DNA hybrid complex (typically, about 3°C below the T_m of the hybrid). Higher stringency makes for a more specific correlation between the probe sequence and the target. Stringent conditions used in the hybridization of nucleic acids are well known in the art and may be found in references which compile such methods, e.g. *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989, or *Current Protocols in Molecular Biology*, F.M. Ausubel, et al., eds., John Wiley & Sons, Inc., New York. An example of "stringent conditions" is hybridization at 65°C in 6 x SSC. Another example of stringent conditions is hybridization at 65°C in hybridization buffer that consists of 3.5 x SSC, 0.02%

Ficoll, 0.02% polyvinyl pyrolidone, 0.02% Bovine Serum Albumin, 2.5mM NaH_2PO_4[pH7], 0.5% SDS, 2mM EDTA. (SSC is 0.15M sodium chloride/0.15M sodium citrate, pH7; SDS is sodium dodecyl sulphate; and EDTA is ethylenediaminetetracetic acid). After hybridization, the membrane upon which the DNA is transferred is washed at 2 x SSC at room temperature and then at 0.1 x SSC/0.1 x SDS at temperatures up to 68°C. In a further example, an alternative to the use of an aqueous hybridization solution is the use of a formamide hybridization solution. Stringent hybridization conditions can thus be achieved using, for example, a 50% formamide solution and 42°C. There are other conditions, reagents, and so forth which can be used, and would result in a similar degree of stringency. The skilled artisan will be familiar with such conditions, and thus they are not given here. It will be understood, however, that the skilled artisan will be able to manipulate the conditions in a manner to permit the clear identification of homologs and alleles of FGE nucleic acids of the invention. The skilled artisan also is familiar with the methodology for screening cells and libraries for expression of such molecules which then are routinely isolated, followed by isolation of the pertinent nucleic acid molecule and sequencing.

In general homologs and alleles typically will share at least 40% nucleotide identity and/or at least 50% amino acid identity to SEQ ID NO:1 and SEQ ID NO:2, respectively, in some instances will share at least 50% nucleotide identity and/or at least 65% amino acid identity and in still other instances will share at least 60% nucleotide identity and/or at least 75% amino acid identity. In further instances, homologs and alleles typically will share at least 90%, 95%, or even 99% nucleotide identity and/or at least 95%, 98%, or even 99% amino acid identity to SEQ ID NO:1 and SEQ ID NO:2, respectively. The homology can be calculated using various, publicly available software tools developed by NCBI (Bethesda, Maryland). Exemplary tools include the heuristic algorithm of Altschul SF, et al., (*J Mol Biol*, 1990, 215:403-410), also known as BLAST. Pairwise and ClustalW alignments (BLOSUM30 matrix setting) as well as Kyte-Doolittle hydropathic analysis can be obtained using public (EMBL, Heidelberg, Germany) and commercial (e.g., the MacVector sequence analysis software from Oxford Molecular Group/enetics Computer Group, Madison, WI). Watson-Crick complements of the foregoing nucleic acids also are embraced by the invention.

In screening for FGE related genes, such as homologs and alleles of FGE, a Southern blot may be performed using the foregoing conditions, together with a radioactive probe. After washing the membrane to which the DNA is finally transferred, the membrane can be placed against X-ray film or a phosphoimager plate to detect the radioactive signal.

Given the teachings herein of a full-length human FGE cDNA clone, other mammalian sequences such as the mouse cDNA clone corresponding to the human FGE gene can be isolated from a cDNA library, using standard colony hybridization techniques.

The invention also includes degenerate nucleic acids which include alternative codons to those present in the native materials. For example, serine residues are encoded by the codons TCA, AGT, TCC, TCG, TCT and AGC. Thus, it will be apparent to one of ordinary skill in the art that any of the serine-encoding nucleotide triplets may be employed to direct the protein synthesis apparatus, *in vitro* or *in vivo*, to incorporate a serine residue into an elongating FGE polypeptide. Similarly, nucleotide sequence triplets which encode other amino acid residues include, but are not limited to: CCA, CCC, CCG and CCT (proline codons); CGA, CGC, CGG, CGT, AGA and AGG (arginine codons); ACA, ACC, ACG and ACT (threonine codons); AAC and AAT (asparagine codons); and ATA, ATC and ATT (isoleucine codons). Other amino acid residues may be encoded similarly by multiple nucleotide sequences. Thus, the invention embraces degenerate nucleic acids that differ from the biologically isolated nucleic acids in codon sequence due to the degeneracy of the genetic code.

The invention also provides isolated unique fragments of SEQ ID NO:1 or SEQ ID NO:3 or complements of thereof. A unique fragment is one that is a 'signature' for the larger nucleic acid. For example, the unique fragment is long enough to assure that its precise sequence is not found in molecules within the human genome outside of the FGE nucleic acids defined above (and human alleles). Those of ordinary skill in the art may apply no more than routine procedures to determine if a fragment is unique within the human genome. Unique fragments, however, exclude fragments completely composed of the nucleotide sequences selected from the group consisting of SEQ ID NO:4, and/or other previously published sequences as of the filing date of this application.

A fragment which is completely composed of the sequence described in the foregoing GenBank deposits is one which does not include any of the nucleotides unique to the sequences of the invention. Thus, a unique fragment according to the invention must contain a nucleotide sequence other than the exact sequence of those in the GenBank deposits or fragments thereof. The difference may be an addition, deletion or substitution with respect to the GenBank sequence or it may be a sequence wholly separate from the GenBank sequence.

Unique fragments can be used as probes in Southern and Northern blot assays to identify such nucleic acids, or can be used in amplification assays such as those employing PCR. As known to those skilled in the art, large probes such as 200, 250, 300 or more

nucleotides are preferred for certain uses such as Southern and Northern blots, while smaller fragments will be preferred for uses such as PCR. Unique fragments also can be used to produce fusion proteins for generating antibodies or determining binding of the polypeptide fragments, as demonstrated in the Examples, or for generating immunoassay components. Likewise, unique fragments can be employed to produce nonfused fragments of the FGE polypeptides, useful, for example, in the preparation of antibodies, immunoassays or therapeutic applications. Unique fragments further can be used as antisense molecules to inhibit the expression of FGE nucleic acids and polypeptides respectively.

As will be recognized by those skilled in the art, the size of the unique fragment will depend upon its conservancy in the genetic code. Thus, some regions of SEQ ID NO:1 or SEQ ID NO:3 and complements will require longer segments to be unique while others will require only short segments, typically between 12 and 32 nucleotides long (e.g. 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31 and 32 bases) or more, up to the entire length of the disclosed sequence. As mentioned above, this disclosure intends to embrace each and every fragment of each sequence, beginning at the first nucleotide, the second nucleotide and so on, up to 8 nucleotides short of the end, and ending anywhere from nucleotide number 8, 9, 10 and so on for each sequence, up to the very last nucleotide, (provided the sequence is unique as described above). Virtually any segment of the region of SEQ ID NO:1 beginning at nucleotide 1 and ending at nucleotide 1180, or SEQ ID NO:3 beginning at nucleotide 1 and ending at nucleotide 1122, or complements thereof, that is 20 or more nucleotides in length will be unique. Those skilled in the art are well versed in methods for selecting such sequences, typically on the basis of the ability of the unique fragment to selectively distinguish the sequence of interest from other sequences in the human genome of the fragment to those on known databases typically is all that is necessary, although *in vitro* confirmatory hybridization and sequencing analysis may be performed.

As mentioned above, the invention embraces antisense oligonucleotides that selectively bind to a nucleic acid molecule encoding a FGE polypeptide, to decrease FGE activity.

As used herein, the term "antisense oligonucleotide" or "antisense" describes an oligonucleotide that is an oligoribonucleotide, oligodeoxyribonucleotide, modified oligoribonucleotide, or modified oligodeoxyribonucleotide which hybridizes under physiological conditions to DNA comprising a particular gene or to an mRNA transcript of that gene and, thereby, inhibits the transcription of that gene and/or the translation of that mRNA. The antisense molecules are designed so as to interfere with transcription or

translation of a target gene upon hybridization with the target gene or transcript. Those skilled in the art will recognize that the exact length of the antisense oligonucleotide and its degree of complementarity with its target will depend upon the specific target selected, including the sequence of the target and the particular bases which comprise that sequence. It is preferred that the antisense oligonucleotide be constructed and arranged so as to bind selectively with the target under physiological conditions, i.e., to hybridize substantially more to the target sequence than to any other sequence in the target cell under physiological conditions. Based upon SEQ ID NO:1 or upon allelic or homologous genomic and/or cDNA sequences, one of skill in the art can easily choose and synthesize any of a number of appropriate antisense molecules for use in accordance with the present invention. In order to be sufficiently selective and potent for inhibition, such antisense oligonucleotides should comprise at least 10 and, more preferably, at least 15 consecutive bases which are complementary to the target, although in certain cases modified oligonucleotides as short as 7 bases in length have been used successfully as antisense oligonucleotides (Wagner et al., *Nat. Med,* 1995, 1(11):1116-1118; *Nat. Biotech.*, 1996, 14:840-844). Most preferably, the antisense oligonucleotides comprise a complementary sequence of 20-30 bases. Although oligonucleotides may be chosen which are antisense to any region of the gene or mRNA transcripts, in preferred embodiments the antisense oligonucleotides correspond to N-terminal or 5' upstream sites such as translation initiation, transcription initiation or promoter sites. In addition, 3'-untranslated regions may be targeted by antisense oligonucleotides. Targeting to mRNA splicing sites has also been used in the art but may be less preferred if alternative mRNA splicing occurs. In addition, the antisense is targeted, preferably, to sites in which mRNA secondary structure is not expected (see, e.g., Sainio et al., *Cell Mol. Neurobiol.* 14(5):439-457, 1994) and at which proteins are not expected to bind. Finally, although, SEQ ID No:1 discloses a cDNA sequence, one of ordinary skill in the art may easily derive the genomic DNA corresponding to this sequence. Thus, the present invention also provides for antisense oligonucleotides which are complementary to the genomic DNA corresponding to SEQ ID NO:1. Similarly, antisense to allelic or homologous FGE cDNAs and genomic DNAs are enabled without undue experimentation.

In one set of embodiments, the antisense oligonucleotides of the invention may be composed of "natural" deoxyribonucleotides, ribonucleotides, or any combination thereof. That is, the 5' end of one native nucleotide and the 3' end of another native nucleotide may be covalently linked, as in natural systems, via a phosphodiester internucleoside linkage. These oligonucleotides may be prepared by art recognized methods which may be carried out

manually or by an automated synthesizer. They also may be produced recombinantly by vectors.

In preferred embodiments, however, the antisense oligonucleotides of the invention also may include "modified" oligonucleotides. That is, the oligonucleotides may be modified in a number of ways which do not prevent them from hybridizing to their target but which enhance their stability or targeting or which otherwise enhance their therapeutic effectiveness.

The term "modified oligonucleotide" as used herein describes an oligonucleotide in which (1) at least two of its nucleotides are covalently linked via a synthetic internucleoside linkage (i.e., a linkage other than a phosphodiester linkage between the 5' end of one nucleotide and the 3' end of another nucleotide) and/or (2) a chemical group not normally associated with nucleic acids has been covalently attached to the oligonucleotide. Preferred synthetic internucleoside linkages are phosphorothioates, alkylphosphonates, phosphorodithioates, phosphate esters, alkylphosphonothioates, phosphoramidates, carbamates, carbonates, phosphate triesters, acetamidates, carboxymethyl esters and peptides.

The term "modified oligonucleotide" also encompasses oligonucleotides with a covalently modified base and/or sugar. For example, modified oligonucleotides include oligonucleotides having backbone sugars which are covalently attached to low molecular weight organic groups other than a hydroxyl group at the 3' position and other than a phosphate group at the 5' position. Thus modified oligonucleotides may include a 2'-O-alkylated ribose group. In addition, modified oligonucleotides may include sugars such as arabinose instead of ribose. The present invention, thus, contemplates pharmaceutical preparations containing modified antisense molecules that are complementary to and hybridizable with, under physiological conditions, nucleic acids encoding FGE polypeptides, together with pharmaceutically acceptable carriers. Antisense oligonucleotides may be administered as part of a pharmaceutical composition. Such a pharmaceutical composition may include the antisense oligonucleotides in combination with any standard physiologically and/or pharmaceutically acceptable carriers which are known in the art. The compositions should be sterile and contain a therapeutically effective amount of the antisense oligonucleotides in a unit of weight or volume suitable for administration to a patient. The term "pharmaceutically acceptable" means a non-toxic material that does not interfere with the effectiveness of the biological activity of the active ingredients. The term "physiologically acceptable" refers to a non-toxic material that is compatible with a biological system such as a cell, cell culture, tissue, or organism. The characteristics of the

carrier will depend on the route of administration. Physiologically and pharmaceutically acceptable carriers include diluents, fillers, salts, buffers, stabilizers, solubilizers, and other materials which are well known in the art.

The invention also involves methods for increasing C_α-formylglycine generating activity in a cell. In important embodiments, this is accomplished by the use of vectors ("expression vectors" and/or "targeting vectors").

"Vectors," as used herein, may be any of a number of nucleic acids into which a desired sequence may be inserted by restriction and ligation for transport between different genetic environments or for expression in a host cell. Vectors are typically composed of DNA although RNA vectors are also available. Vectors include, but are not limited to, plasmids, phagemids and virus genomes. A cloning vector is one which is able to replicate in a host cell, and which is further characterized by one or more endonuclease restriction sites at which the vector may be cut in a determinable fashion and into which a desired DNA sequence may be ligated such that the new recombinant vector retains its ability to replicate in the host cell. In the case of plasmids, replication of the desired sequence may occur many times as the plasmid increases in copy number within the host bacterium or just a single time per host before the host reproduces by mitosis. In the case of phage, replication may occur actively during a lytic phase or passively during a lysogenic phase. An "expression vector" is one into which a desired DNA sequence (e.g., the FGE cDNA of SEQ ID NO:3) may be inserted by restriction and ligation such that it is operably joined to regulatory sequences and may be expressed as an RNA transcript. Vectors may further contain one or more marker sequences suitable for use in the identification of cells which have or have not been transformed or transfected with the vector. Markers include, for example, genes encoding proteins which increase or decrease either resistance or sensitivity to antibiotics or other compounds, genes which encode enzymes whose activities are detectable by standard assays known in the art (e.g., β-galactosidase or alkaline phosphatase), and genes which visibly affect the phenotype of transformed or transfected cells, hosts, colonies or plaques (e.g., green fluorescent protein).

A "targeting vector" is one which typically contains targeting constructs/sequences that are used, for example, to insert a regulatory sequence within an endogenous gene (e.g., within the sequences of an exon and/or intron), within the endogenous gene promoter sequences, or upstream of the endogenous gene promoter sequences. In another example, a targeting vector may contain the gene of interest (e.g., encoded by the cDNA of SEQ ID NO:1) and other sequences necessary for the targeting of the gene to a preferred location in

the genome (e.g., a trascriptionally active location, for example downstream of an enogenous promoter of an unrelated gene). Construction of targeting constructs and vectors are described in detail in U.S. Patents 5,641,670 and 6,270,989, and which are expressly incorporated herein by reference.

Virtually any cells, prokaryotic or eukaryotic, which can be transformed with heterologous DNA or RNA and which can be grown or maintained in culture, may be used in the practice of the invention. Examples include bacterial cells such as *Escherichia coli*, insect cells, and mammalian cells such as human, mouse, hamster, pig, goat, primate, etc. They may be primary or secondary cell strains (which exhibit a finite number of mean population doublings in culture and are not immortalized) and immortalized cell lines (which exhibit an apparently unlimited lifespan in culture). Primary and secondary cells include, for example, fibroblasts, keratinocytes, epithelial cells (e.g., mammary epithelial cells, intestinal epithelial cells), endothelial cells, glial cells, neural cells, formed elements of the blood (e.g., lymphocytes, bone marrow cells), muscle cells and precursors of these somatic cell types including embryonic stem cells. Where the cells are to be used in gene therapy, primary cells are preferably obtained from the individual to whom the manipulated cells are administered. However, primary cells can be obtained from a donor (other than the recipient) of the same species. Examples of immortalized human cell lines which may be used with the DNA constructs and methods of the present invention include, but are not limited to, HT-1080 cells (ATCC CCL 121), HeLa cells and derivatives of HeLa cells (ATCC CCL 2, 2.1 and 2.2), MCF-7 breast cancer cells (ATCC BTH 22), K-562 leukemia cells (ATCC CCL 243), KB carcinoma cells (ATCC CCL 17), 2780AD ovarian carcinoma cells (Van der Blick, A. M. et al., *Cancer Res*, 48:5927-5932 (1988), Raji cells (ATCC CCL 86), WiDr colon adenocarcinoma cells (ATCC CCL 218), SW620 colon adenocarcinoma cells (ATCC CCL 227), Jurkat cells (ATCC TIB 152), Namalwa cells (ATCC CRL1432), HL-60 cells (ATCC CCL 240), Daudi cells (ATCC CCL 213), RPMI 8226 cells (ATCC CCL 155), U-937 cells (ATCC CRL 1593), Bowes Melanoma cells (ATCC CRL 9607), WI-38VA13 subline 2R4 cells (ATCC CLL 75.1), and MOLT-4 cells (ATCC CRL 1582), CHO cells, and COS cells, as well as heterohybridoma cells produced by fusion of human cells and cells of another species. Secondary human fibroblast strains, such as WI-38 (ATCC CCL 75) and MRC-5 (ATCC CCL 171) may also be used. Further discussion of the types of cells that may be used in practicing the methods of the present invention are described in U.S. Patents 5,641,670 and 6,270,989. Cell-free transcription systems also may be used in lieu of cells.

The cells of the invention are maintained under conditions, as are known in the art, which result in expression of the FGE protein or functional fragments thereof. Proteins expressed using the methods described may be purified from cell lysates or cell supernatants. Proteins made according to this method can be prepared as a pharmaceutically-useful formulation and delivered to a human or non-human animal by conventional pharmaceutical routes as is known in the art (e.g., oral, intravenous, intramuscular, intranasal, intratracheal or subcutaneous). As described elsewhere herein, the recombinant cells can be immortalized, primary, or secondary cells, preferably human. The use of cells from other species may be desirable in cases where the non-human cells are advantageous for protein production purposes where the non-human FGE produced is useful therapeutically.

As used herein, a coding sequence and regulatory sequences are said to be "operably" joined when they are covalently linked in such a way as to place the expression or transcription of the coding sequence under the influence or control of the regulatory sequences. If it is desired that the coding sequences be translated into a functional protein, two DNA sequences are said to be operably joined if induction of a promoter in the 5' regulatory sequences results in the transcription of the coding sequence and if the nature of the linkage between the two DNA sequences does not (1) result in the introduction of a frame-shift mutation, (2) interfere with the ability of the promoter region to direct the transcription of the coding sequences, or (3) interfere with the ability of the corresponding RNA transcript to be translated into a protein. Thus, a promoter region would be operably joined to a coding sequence if the promoter region were capable of effecting transcription of that DNA sequence such that the resulting transcript might be translated into the desired protein or polypeptide.

The precise nature of the regulatory sequences needed for gene expression may vary between species or cell types, but shall in general include, as necessary, 5' non-transcribed and 5' non-translated sequences involved with the initiation of transcription and translation respectively, such as a TATA box, capping sequence, CAAT sequence, and the like. Especially, such 5' non-transcribed regulatory sequences will include a promoter region which includes a promoter sequence for transcriptional control of the operably joined gene. Regulatory sequences may also include enhancer sequences or upstream activator sequences as desired. The vectors of the invention may optionally include 5' leader or signal sequences. The choice and design of an appropriate vector is within the ability and discretion of one of ordinary skill in the art.

Expression vectors containing all the necessary elements for expression are commercially available and known to those skilled in the art. See, e.g., Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory Press, 1989. Cells are genetically engineered by the introduction into the cells of heterologous DNA (RNA) encoding FGE polypeptide or fragment or variant thereof. That heterologous DNA (RNA) is placed under operable control of transcriptional elements to permit the expression of the heterologous DNA in the host cell.

Preferred systems for mRNA expression in mammalian cells are those such as pRc/CMV (available from Invitrogen, Carlsbad, CA) that contain a selectable marker such as a gene that confers G418 resistance (which facilitates the selection of stably transfected cell lines) and the human cytomegalovirus (CMV) enhancer-promoter sequences. Additionally, suitable for expression in primate or canine cell lines is the pCEP4 vector (Invitrogen, Carlsbad, CA), which contains an Epstein Barr virus (EBV) origin of replication, facilitating the maintenance of plasmid as a multicopy extrachromosomal element. Another expression vector is the pEF-BOS plasmid containing the promoter of polypeptide Elongation Factor 1α, which stimulates efficiently transcription *in vitro*. The plasmid is described by Mishizuma and Nagata (*Nuc. Acids Res.* 18:5322, 1990), and its use in transfection experiments is disclosed by, for example, Demoulin (*Mol. Cell. Biol.* 16:4710-4716, 1996). Still another preferred expression vector is an adenovirus, described by Stratford-Perricaudet, which is defective for E1 and E3 proteins (*J. Clin. Invest.* 90:626-630, 1992). The use of the adenovirus as an Adeno.P1A recombinant is disclosed by Warnier et al., in intradermal injection in mice for immunization against P1A (*Int. J. Cancer*, 67:303-310, 1996).

The invention also embraces so-called expression kits, which allow the artisan to prepare a desired expression vector or vectors. Such expression kits include at least separate portions of each of the previously discussed coding sequences. Other components may be added, as desired, as long as the previously mentioned sequences, which are required, are included.

It will also be recognized that the invention embraces the use of the above described, FGE cDNA sequence containing expression vectors, to transfect host cells and cell lines, be these prokaryotic (e.g., *Escherichia coli*), or eukaryotic (e.g., CHO cells, COS cells, yeast expression systems and recombinant baculovirus expression in insect cells). Especially useful are mammalian cells such as human, mouse, hamster, pig, goat, primate, etc. They may be of a wide variety of tissue types, and include primary cells and immortalized cell lines as described elsewhere herein. Specific examples include HT-1080 cells, CHO cells,

dendritic cells, U293 cells, peripheral blood leukocytes, bone marrow stem cells, embryonic stem cells, and insect cells. The invention also permits the construction of FGE gene "knock-outs" in cells and in animals, providing materials for studying certain aspects of FGE activity.

The invention also provides isolated polypeptides (including whole proteins and partial proteins), encoded by the foregoing FGE nucleic acids, and include the polypeptide of SEQ ID NO:2 and unique fragments thereof. Such polypeptides are useful, for example, alone or as part of fusion proteins to generate antibodies, as components of an immunoassay, etc. Polypeptides can be isolated from biological samples including tissue or cell homogenates, and can also be expressed recombinantly in a variety of prokaryotic and eukaryotic expression systems by constructing an expression vector appropriate to the expression system, introducing the expression vector into the expression system, and isolating the recombinantly expressed protein. Short polypeptides, including antigenic peptides (such as are presented by MHC molecules on the surface of a cell for immune recognition) also can be synthesized chemically using well-established methods of peptide synthesis.

A unique fragment of a FGE polypeptide, in general, has the features and characteristics of unique fragments as discussed above in connection with nucleic acids. As will be recognized by those skilled in the art, the size of the unique fragment will depend upon factors such as whether the fragment constitutes a portion of a conserved protein domain. Thus, some regions of SEQ ID NO:2 will require longer segments to be unique while others will require only short segments, typically between 5 and 12 amino acids (e.g. 5, 6, 7, 8, 9, 10, 11 and 12 amino acids long or more, including each integer up to the full length, 287 amino acids long).

Unique fragments of a polypeptide preferably are those fragments which retain a distinct functional capability of the polypeptide. Functional capabilities which can be retained in a unique fragment of a polypeptide include interaction with antibodies, interaction with other polypeptides or fragments thereof, interaction with other molecules, etc. One important activity is the ability to act as a signature for identifying the polypeptide. Those skilled in the art are well versed in methods for selecting unique amino acid sequences, typically on the basis of the ability of the unique fragment to selectively distinguish the sequence of interest from non-family members. A comparison of the sequence of the fragment to those on known databases typically is all that is necessary.

The invention embraces variants of the FGE polypeptides described above. As used herein, a "variant" of a FGE polypeptide is a polypeptide which contains one or more

modifications to the primary amino acid sequence of a FGE polypeptide. Modifications which create a FGE polypeptide variant are typically made to the nucleic acid which encodes the FGE polypeptide, and can include deletions, point mutations, truncations, amino acid substitutions and addition of amino acids or non-amino acid moieties to: 1) reduce or eliminate an activity of a FGE polypeptide; 2) enhance a property of a FGE polypeptide, such as protein stability in an expression system or the stability of protein-ligand binding; 3) provide a novel activity or property to a FGE polypeptide, such as addition of an antigenic epitope or addition of a detectable moiety; or 4) to provide equivalent or better binding to a FGE polypeptide receptor or other molecule. Alternatively, modifications can be made directly to the polypeptide, such as by cleavage, addition of a linker molecule, addition of a detectable moiety, such as biotin, addition of a fatty acid, and the like. Modifications also embrace fusion proteins comprising all or part of the FGE amino acid sequence. One of skill in the art will be familiar with methods for predicting the effect on protein conformation of a change in protein sequence, and can thus "design" a variant FGE polypeptide according to known methods. One example of such a method is described by Dahiyat and Mayo in *Science* 278:82-87, 1997, whereby proteins can be designed *de novo*. The method can be applied to a known protein to vary only a portion of the polypeptide sequence. By applying the computational methods of Dahiyat and Mayo, specific variants of the FGE polypeptide can be proposed and tested to determine whether the variant retains a desired conformation.

Variants can include FGE polypeptides which are modified specifically to alter a feature of the polypeptide unrelated to its physiological activity. For example, cysteine residues can be substituted or deleted to prevent unwanted disulfide linkages. Similarly, certain amino acids can be changed to enhance expression of a FGE polypeptide by eliminating proteolysis by proteases in an expression system (e.g., dibasic amino acid residues in yeast expression systems in which KEX2 protease activity is present).

Mutations of a nucleic acid which encodes a FGE polypeptide preferably preserve the amino acid reading frame of the coding sequence, and preferably do not create regions in the nucleic acid which are likely to hybridize to form secondary structures, such a hairpins or loops, which can be deleterious to expression of the variant polypeptide.

Mutations can be made by selecting an amino acid substitution, or by random mutagenesis of a selected site in a nucleic acid which encodes the polypeptide. Variant polypeptides are then expressed and tested for one or more activities to determine which mutation provides a variant polypeptide with the desired properties. Further mutations can be made to variants (or to non-variant FGE polypeptides) which are silent as to the amino acid

sequence of the polypeptide, but which provide preferred codons for translation in a particular host, or alter the structure of the mRNA to, for example, enhance stability and/or expression. The preferred codons for translation of a nucleic acid in, e.g., *Escherichia coli*, mammalian cells, etc. are well known to those of ordinary skill in the art. Still other mutations can be made to the noncoding sequences of a FGE gene or cDNA clone to enhance expression of the polypeptide.

The skilled artisan will realize that conservative amino acid substitutions may be made in FGE polypeptides to provide functionally equivalent variants of the foregoing polypeptides, i.e, the variants retain the functional capabilities of the FGE polypeptides. As used herein, a "conservative amino acid substitution" refers to an amino acid substitution which does not significantly alter the the tertiary structure and/or activity of the polypeptide. Variants can be prepared according to methods for altering polypeptide sequence known to one of ordinary skill in the art, and include those that are found in references which compile such methods, e.g. *Molecular Cloning: A Laboratory Manual*, J. Sambrook, et al., eds., Second Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1989, or *Current Protocols in Molecular Biology*, F.M. Ausubel, et al., eds., John Wiley & Sons, Inc., New York. Exemplary functionally equivalent variants of the FGE polypeptides include conservative amino acid substitutions of SEQ ID NO:2. Conservative substitutions of amino acids include substitutions made amongst amino acids within the following groups: (a) M, I, L, V; (b) F, Y, W; (c) K, R, H; (d) A, G; (e) S, T; (f) Q, N; and (g) E, D.

Thus functionally equivalent variants of FGE polypeptides, i.e., variants of FGE polypeptides which retain the function of the natural FGE polypeptides, are contemplated by the invention. Conservative amino-acid substitutions in the amino acid sequence of FGE polypeptides to produce functionally equivalent variants of FGE polypeptides typically are made by alteration of a nucleic acid encoding FGE polypeptides (SEQ ID NOs:1, 3). Such substitutions can be made by a variety of methods known to one of ordinary skill in the art. For example, amino acid substitutions may be made by PCR-directed mutation, site-directed mutagenesis according to the method of Kunkel (Kunkel, *Proc. Nat. Acad. Sci. U.S.A.* 82: 488-492, 1985), or by chemical synthesis of a gene encoding a FGE polypeptide. The activity of functionally equivalent fragments of FGE polypeptides can be tested by cloning the gene encoding the altered FGE polypeptide into a bacterial or mammalian expression vector, introducing the vector into an appropriate host cell, expressing the altered FGE polypeptide, and testing for a functional capability of the FGE polypeptides as disclosed herein (e.g., C_{α}-formylglycine generating activity, etc.).

The invention as described herein has a number of uses, some of which are described elsewhere herein. First, the invention permits isolation of FGE polypeptides. A variety of methodologies well-known to the skilled practitioner can be utilized to obtain isolated FGE molecules. The polypeptide may be purified from cells which naturally produce the polypeptide by chromatographic means or immunological recognition. Alternatively, an expression vector may be introduced into cells to cause production of the polypeptide. In another method, mRNA transcripts may be microinjected or otherwise introduced into cells to cause production of the encoded polypeptide. Translation of FGE mRNA in cell-free extracts such as the reticulocyte lysate system also may be used to produce FGE polypeptides. Those skilled in the art also can readily follow known methods for isolating FGE polypeptides. These include, but are not limited to, immunochromatography, HPLC, size-exclusion chromatography, ion-exchange chromatography and immune-affinity chromatography.

The invention also provides, in certain embodiments, "dominant negative" polypeptides derived from FGE polypeptides. A dominant negative polypeptide is an inactive variant of a protein, which, by interacting with the cellular machinery, displaces an active protein from its interaction with the cellular machinery or competes with the active protein, thereby reducing the effect of the active protein. For example, a dominant negative receptor which binds a ligand but does not transmit a signal in response to binding of the ligand can reduce the biological effect of expression of the ligand. Likewise, a dominant negative catalytically-inactive kinase which interacts normally with target proteins but does not phosphorylate the target proteins can reduce phosphorylation of the target proteins in response to a cellular signal. Similarly, a dominant negative transcription factor which binds to a promoter site in the control region of a gene but does not increase gene transcription can reduce the effect of a normal transcription factor by occupying promoter binding sites without increasing transcription.

The end result of the expression of a dominant negative polypeptide in a cell is a reduction in function of active proteins. One of ordinary skill in the art can assess the potential for a dominant negative variant of a protein, and use standard mutagenesis techniques to create one or more dominant negative variant polypeptides. See, e.g., U.S. Patent No. 5,580,723 and Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Second Edition, Cold Spring Harbor Laboratory Press, 1989. The skilled artisan then can test the population of mutagenized polypeptides for diminution in a selected activity and/or for

retention of such an activity. Other similar methods for creating and testing dominant negative variants of a protein will be apparent to one of ordinary skill in the art.

The isolation of the FGE cDNA also makes it possible for the artisan to diagnose a disorder characterized by an aberrant expression of FGE. These methods involve determining expression of the FGE gene, and/or FGE polypeptides derived therefrom. In the former situation, such determinations can be carried out via any standard nucleic acid determination assay, including the polymerase chain reaction, or assaying with labeled hybridization probes as exemplified below. In the latter situation, such determination can be carried out via any standard immunological assay using, for example, antibodies which bind to the secreted FGE protein. A preferred disorder that can be diagnosed according to the invention is Multiple Sulfatase Deficiency.

The invention also embraces isolated peptide binding agents which, for example, can be antibodies or fragments of antibodies ("binding polypeptides"), having the ability to selectively bind to FGE polypeptides. Antibodies include polyclonal and monoclonal antibodies, prepared according to conventional methodology. In certain embodiments, the invention excludes binding agents (e.g., antibodies) that bind to the polypeptides encoded by the nucleic acids of SEQ ID NO:4.

Significantly, as is well-known in the art, only a small portion of an antibody molecule, the paratope, is involved in the binding of the antibody to its epitope (see, in general, Clark, W.R. (1986) The Experimental Foundations of Modern Immunology Wiley & Sons, Inc., New York; Roitt, I. (1991) Essential Immunology, 7th Ed., Blackwell Scientific Publications, Oxford). The pFc' and Fc regions, for example, are effectors of the complement cascade but are not involved in antigen binding. An antibody from which the pFc' region has been enzymatically cleaved, or which has been produced without the pFc' region, designated an $F(ab')_2$ fragment, retains both of the antigen binding sites of an intact antibody. Similarly, an antibody from which the Fc region has been enzymatically cleaved, or which has been produced without the Fc region, designated an Fab fragment, retains one of the antigen binding sites of an intact antibody molecule. Proceeding further, Fab fragments consist of a covalently bound antibody light chain and a portion of the antibody heavy chain denoted Fd. The Fd fragments are the major determinant of antibody specificity (a single Fd fragment may be associated with up to ten different light chains without altering antibody specificity) and Fd fragments retain epitope-binding ability in isolation.

Within the antigen-binding portion of an antibody, as is well-known in the art, there are complementarity determining regions (CDRs), which directly interact with the epitope of

the antigen, and framework regions (FRs), which maintain the tertiary structure of the paratope (see, in general, Clark, 1986; Roitt, 1991). In both the heavy chain Fd fragment and the light chain of IgG immunoglobulins, there are four framework regions (FR1 through FR4) separated respectively by three complementarity determining regions (CDR1 through CDR3). The CDRs, and in particular the CDR3 regions, and more particularly the heavy chain CDR3, are largely responsible for antibody specificity.

It is now well-established in the art that the non-CDR regions of a mammalian antibody may be replaced with similar regions of conspecific or heterospecific antibodies while retaining the epitopic specificity of the original antibody. This is most clearly manifested in the development and use of "humanized" antibodies in which non-human CDRs are covalently joined to human FR and/or Fc/pFc' regions to produce a functional antibody. See, e.g., U.S. patents 4,816,567, 5,225,539, 5,585,089, 5,693,762 and 5,859,205. Thus, for example, PCT International Publication Number WO 92/04381 teaches the production and use of humanized murine RSV antibodies in which at least a portion of the murine FR regions have been replaced by FR regions of human origin. Such antibodies, including fragments of intact antibodies with antigen-binding ability, are often referred to as "chimeric" antibodies.

Thus, as will be apparent to one of ordinary skill in the art, the present invention also provides for $F(ab')_2$, Fab, Fv and Fd fragments; chimeric antibodies in which the Fc and/or FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences; chimeric $F(ab')_2$ fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences; chimeric Fab fragment antibodies in which the FR and/or CDR1 and/or CDR2 and/or light chain CDR3 regions have been replaced by homologous human or non-human sequences; and chimeric Fd fragment antibodies in which the FR and/or CDR1 and/or CDR2 regions have been replaced by homologous human or non-human sequences. The present invention also includes so-called single chain antibodies.

Thus, the invention involves polypeptides of numerous size and type that bind specifically to FGE polypeptides, and complexes of both FGE polypeptides and their binding partners. These polypeptides may be derived also from sources other than antibody technology. For example, such polypeptide binding agents can be provided by degenerate peptide libraries which can be readily prepared in solution, in immobilized form, as bacterial flagella peptide display libraries or as phage display libraries. Combinatorial libraries also

can be synthesized of peptides containing one or more amino acids. Libraries further can be synthesized of peptides and non-peptide synthetic moieties.

Phage display can be particularly effective in identifying binding peptides useful according to the invention. Briefly, one prepares a phage library (using e.g. m13, fd, or lambda phage), displaying inserts from 4 to about 80 amino acid residues using conventional procedures. The inserts may represent, for example, a completely degenerate or biased array. One then can select phage-bearing inserts which bind to the FGE polypeptide or a complex of FGE and a binding partner. This process can be repeated through several cycles of reselection of phage that bind to the FGE polypeptide or complex. Repeated rounds lead to enrichment of phage bearing particular sequences. DNA sequence analysis can be conducted to identify the sequences of the expressed polypeptides. The minimal linear portion of the sequence that binds to the FGE polypeptide or complex can be determined. One can repeat the procedure using a biased library containing inserts containing part or all of the minimal linear portion plus one or more additional degenerate residues upstream or downstream thereof. Yeast two-hybrid screening methods also may be used to identify polypeptides that bind to the FGE polypeptides. Thus, the FGE polypeptides of the invention, or a fragment thereof, or complexes of FGE and a binding partner can be used to screen peptide libraries, including phage display libraries, to identify and select peptide binding partners of the FGE polypeptides of the invention. Such molecules can be used, as described, for screening assays, for purification protocols, for interfering directly with the functioning of FGE and for other purposes that will be apparent to those of ordinary skill in the art.

An FGE polypeptide, or a fragment thereof, also can be used to isolate their native binding partners. Isolation of binding partners may be performed according to well-known methods. For example, isolated FGE polypeptides can be attached to a substrate, and then a solution suspected of containing a FGE binding partner may be applied to the substrate. If the binding partner for FGE polypeptides is present in the solution, then it will bind to the substrate-bound FGE polypeptide. The binding partner then may be isolated. Other proteins which are binding partners for FGE, may be isolated by similar methods without undue experimentation. A preferred binding partner is a sulfatase.

The invention also provides methods to measure the level of FGE expression in a subject. This can be performed by first obtaining a test sample from the subject. The test sample can be tissue or biological fluid. Tissues include brain, heart, serum, breast, colon, bladder, uterus, prostate, stomach, testis, ovary, pancreas, pituitary gland, adrenal gland, thyroid gland, salivary gland, mammary gland, kidney, liver, intestine, spleen, thymus, blood

vessels, bone marrow, trachea, and lung. In certain embodiments, test samples originate from heart and blood vessel tissues, and biological fluids include blood, saliva and urine. Both invasive and non-invasive techniques can be used to obtain such samples and are well documented in the art. At the molecular level both PCR and Northern blotting can be used to determine the level of FGE mRNA using products of this invention described herein, and protocols well known in the art that are found in references which compile such methods. At the protein level, FGE expression can be determined using either polyclonal or monoclonal anti-FGE sera in combination with standard immunological assays. The preferred methods will compare the measured level of FGE expression of the test sample to a control. A control can include a known amount of a nucleic acid probe, a FGE epitope (such as a FGE expression product), or a similar test sample of a subject with a control or 'normal' level of FGE expression.

FGE polypeptides preferably are produced recombinantly, although such polypeptides may be isolated from biological extracts. Recombinantly produced FGE polypeptides include chimeric proteins comprising a fusion of a FGE protein with another polypeptide, e.g., a polypeptide capable of providing or enhancing protein-protein binding, sequence specific nucleic acid binding (such as GAL4), enhancing stability of the FGE polypeptide under assay conditions, or providing a detectable moiety, such as green fluorescent protein. A polypeptide fused to a FGE polypeptide or fragment may also provide means of readily detecting the fusion protein, e.g., by immunological recognition or by fluorescent labeling.

The invention also is useful in the generation of transgenic non-human animals. As used herein, "transgenic non-human animals" includes non-human animals having one or more exogenous nucleic acid molecules incorporated in germ line cells and/or somatic cells. Thus the transgenic animals include "knockout" animals having a homozygous or heterozygous gene disruption by homologous recombination, animals having episomal or chromosomally incorporated expression vectors, etc. Knockout animals can be prepared by homologous recombination using embryonic stem cells as is well known in the art. The recombination may be facilitated using, for example, the cre/lox system or other recombinase systems known to one of ordinary skill in the art. In certain embodiments, the recombinase system itself is expressed conditionally, for example, in certain tissues or cell types, at certain embryonic or post-embryonic developmental stages, is induced by the addition of a compound which increases or decreases expression, and the like. In general, the conditional expression vectors used in such systems use a variety of promoters which confer the desired gene expression pattern (e.g., temporal or spatial). Conditional promoters also can be

operably linked to FGE nucleic acid molecules to increase expression of FGE in a regulated or conditional manner. *Trans*-acting negative regulators of FGE activity or expression also can be operably linked to a conditional promoter as described above. Such *trans*-acting regulators include antisense FGE nucleic acids molecules, nucleic acid molecules which encode dominant negative FGE molecules, ribozyme molecules specific for FGE nucleic acids, and the like. The transgenic non-human animals are useful in experiments directed toward testing biochemical or physiological effects of diagnostics or therapeutics for conditions characterized by increased or decreased FGE expression. Other uses will be apparent to one of ordinary skill in the art.

The invention also contemplates gene therapy. The procedure for performing *ex vivo* gene therapy is outlined in U.S. Patent 5,399,346 and in exhibits submitted in the file history of that patent, all of which are publicly available documents. In general, it involves introduction *in vitro* of a functional copy of a gene into a cell(s) of a subject which contains a defective copy of the gene, and returning the genetically engineered cell(s) to the subject. The functional copy of the gene is under operable control of regulatory elements which permit expression of the gene in the genetically engineered cell(s). Numerous transfection and transduction techniques as well as appropriate expression vectors are well known to those of ordinary skill in the art, some of which are described in PCT application WO95/00654. *In vivo* gene therapy using vectors such as adenovirus, retroviruses, herpes virus, and targeted liposomes also is contemplated according to the invention.

The invention further provides efficient methods of identifying agents or lead compounds for agents active at the level of a FGE or FGE fragment dependent cellular function. In particular, such functions include interaction with other polypeptides or fragments. Generally, the screening methods involve assaying for compounds which interfere with FGE activity (such as C_α-formylglycine generating activity), although compounds which enhance FGE C_α-formylglycine generating activity also can be assayed using the screening methods. Such methods are adaptable to automated, high throughput screening of compounds. Target indications include cellular processes modulated by FGE such as C_α-formylglycine generating activity.

A wide variety of assays for candidate (pharmacological) agents are provided, including, labeled *in vitro* protein-ligand binding assays, electrophoretic mobility shift assays, immunoassays, cell-based assays such as two- or three-hybrid screens, expression assays, etc. The transfected nucleic acids can encode, for example, combinatorial peptide libraries or cDNA libraries. Convenient reagents for such assays, e.g., GAL4 fusion proteins, are known

in the art. An exemplary cell-based assay involves transfecting a cell with a nucleic acid encoding a FGE polypeptide fused to a GAL4 DNA binding domain and a nucleic acid encoding a reporter gene operably linked to a gene expression regulatory region, such as one or more GAL4 binding sites. Activation of reporter gene transcription occurs when the FGE and reporter fusion polypeptide binds such as to enable transcription of the reporter gene. Agents which modulate a FGE polypeptide mediated cell function are then detected through a change in the expression of reporter gene. Methods for determining changes in the expression of a reporter gene are known in the art.

FGE fragments used in the methods, when not produced by a transfected nucleic acid are added to an assay mixture as an isolated polypeptide. FGE polypeptides preferably are produced recombinantly, although such polypeptides may be isolated from biological extracts. Recombinantly produced FGE polypeptides include chimeric proteins comprising a fusion of a FGE protein with another polypeptide, e.g., a polypeptide capable of providing or enhancing protein-protein binding, sequence specific nucleic acid binding (such as GAL4), enhancing stability of the FGE polypeptide under assay conditions, or providing a detectable moiety, such as green fluorescent protein or Flag epitope.

The assay mixture is comprised of a natural intracellular FGE binding target capable of interacting with FGE. While natural FGE binding targets may be used, it is frequently preferred to use portions (e.g., peptides –see e.g., the peptide of SEQ ID NO:33- or nucleic acid fragments) or analogs (i.e., agents which mimic the FGE binding properties of the natural binding target for purposes of the assay) of the FGE binding target so long as the portion or analog provides binding affinity and avidity to the FGE fragment measurable in the assay.

The assay mixture also comprises a candidate agent. Typically, a plurality of assay mixtures are run in parallel with different agent concentrations to obtain a different response to the various concentrations. Typically, one of these concentrations serves as a negative control, i.e., at zero concentration of agent or at a concentration of agent below the limits of assay detection. Candidate agents encompass numerous chemical classes, although typically they are organic compounds. Preferably, the candidate agents are small organic compounds, i.e., those having a molecular weight of more than 50 yet less than about 2500, preferably less than about 1000 and, more preferably, less than about 500. Candidate agents comprise functional chemical groups necessary for structural interactions with polypeptides and/or nucleic acids, and typically include at least an amine, carbonyl, hydroxyl or carboxyl group, preferably at least two of the functional chemical groups and more preferably at least three of

the functional chemical groups. The candidate agents can comprise cyclic carbon or heterocyclic structure and/or aromatic or polyaromatic structures substituted with one or more of the above-identified functional groups. Candidate agents also can be biomolecules such as peptides, saccharides, fatty acids, sterols, isoprenoids, purines, pyrimidines, derivatives or structural analogs of the above, or combinations thereof and the like. Where the agent is a nucleic acid, the agent typically is a DNA or RNA molecule, although modified nucleic acids as defined herein are also contemplated.

Candidate agents are obtained from a wide variety of sources including libraries of synthetic or natural compounds. For example, numerous means are available for random and directed synthesis of a wide variety of organic compounds and biomolecules, including expression of randomized oligonucleotides, synthetic organic combinatorial libraries, phage display libraries of random peptides, and the like. Alternatively, libraries of natural compounds in the form of bacterial, fungal, plant and animal extracts are available or readily produced. Additionally, natural and synthetically produced libraries and compounds can be modified through conventional chemical, physical, and biochemical means. Further, known (pharmacological) agents may be subjected to directed or random chemical modifications such as acylation, alkylation, esterification, amidification, etc. to produce structural analogs of the agents.

A variety of other reagents also can be included in the mixture. These include reagents such as salts, buffers, neutral proteins (e.g., albumin), detergents, etc. which may be used to facilitate optimal protein-protein and/or protein-nucleic acid binding. Such a reagent may also reduce non-specific or background interactions of the reaction components. Other reagents that improve the efficiency of the assay such as protease, inhibitors, nuclease inhibitors, antimicrobial agents, and the like may also be used.

The mixture of the foregoing assay materials is incubated under conditions whereby, but for the presence of the candidate agent, the FGE polypeptide specifically binds a cellular binding target, a portion thereof or analog thereof. The order of addition of components, incubation temperature, time of incubation, and other parameters of the assay may be readily determined. Such experimentation merely involves optimization of the assay parameters, not the fundamental composition of the assay. Incubation temperatures typically are between 4°C and 40°C. Incubation times preferably are minimized to facilitate rapid, high throughput screening, and typically are between 0.1 and 10 hours.

After incubation, the presence or absence of specific binding between the FGE polypeptide and one or more binding targets is detected by any convenient method available

to the user. For cell free binding type assays, a separation step is often used to separate bound from unbound components. The separation step may be accomplished in a variety of ways. Conveniently, at least one of the components is immobilized on a solid substrate, from which the unbound components may be easily separated. The solid substrate can be made of a wide variety of materials and in a wide variety of shapes, e.g., microtiter plate, microbead, dipstick, resin particle, etc. The substrate preferably is chosen to maximum signal to noise ratios, primarily to minimize background binding, as well as for ease of separation and cost.

Separation may be effected for example, by removing a bead or dipstick from a reservoir, emptying or diluting a reservoir such as a microtiter plate well, rinsing a bead, particle, chromatograpic column or filter with a wash solution or solvent. The separation step preferably includes multiple rinses or washes. For example, when the solid substrate is a microtiter plate, the wells may be washed several times with a washing solution, which typically includes those components of the incubation mixture that do not participate in specific bindings such as salts, buffer, detergent, non-specific protein, etc. Where the solid substrate is a magnetic bead, the beads may be washed one or more times with a washing solution and isolated using a magnet.

Detection may be effected in any convenient way for cell-based assays such as two- or three-hybrid screens. The transcript resulting from a reporter gene transcription assay of FGE polypeptide interacting with a target molecule typically encodes a directly or indirectly detectable product, e.g., β-galactosidase activity, luciferase activity, and the like. For cell free binding assays, one of the components usually comprises, or is coupled to, a detectable label. A wide variety of labels can be used, such as those that provide direct detection (e.g., radioactivity, luminescence, optical or electron density, etc), or indirect detection (e.g., epitope tag such as the FLAG epitope, enzyme tag such as horseseradish peroxidase, etc.). The label may be bound to a FGE binding partner, or incorporated into the structure of the binding partner.

A variety of methods may be used to detect the label, depending on the nature of the label and other assay components. For example, the label may be detected while bound to the solid substrate or subsequent to separation from the solid substrate. Labels may be directly detected through optical or electron density, radioactive emissions, nonradiative energy transfers, etc. or indirectly detected with antibody conjugates, streptavidin-biotin conjugates, etc. Methods for detecting the labels are well known in the art.

The invention provides FGE-specific binding agents, methods of identifying and making such agents, and their use in diagnosis, therapy and pharmaceutical development.

For example, FGE-specific pharmacological agents are useful in a variety of diagnostic and therapeutic applications, especially where disease or disease prognosis is associated with altered FGE binding characteristics such as in Multiple Sulfatase Deficiency. Novel FGE-specific binding agents include FGE-specific antibodies, cell surface receptors, and other natural intracellular and extracellular binding agents identified with assays such as two hybrid screens, and non-natural intracellular and extracellular binding agents identified in screens of chemical libraries and the like.

In general, the specificity of FGE binding to a specific molecule is determined by binding equilibrium constants. Targets which are capable of selectively binding a FGE polypeptide preferably have binding equilibrium constants of at least about 10^7 M^{-1}, more preferably at least about 10^8 M^{-1}, and most preferably at least about 10^9 M^{-1}. A wide variety of cell based and cell free assays may be used to demonstrate FGE-specific binding. Cell based assays include one, two and three hybrid screens, assays in which FGE-mediated transcription is inhibited or increased, etc. Cell free assays include FGE-protein binding assays, immunoassays, etc. Other assays useful for screening agents which bind FGE polypeptides include fluorescence resonance energy transfer (FRET), and electrophoretic mobility shift analysis (EMSA).

According to another aspect of the invention, a method for identifying an agent useful in modulating C_α-formylglycine generating activity of a molecule of the invention, is provided. The method involves (a) contacting a molecule having C_α-formylglycine generating activity with a candidate agent, (b) measuring C_α-formylglycine generating activity of the molecule, and (c) comparing the measured C_α-formylglycine generating activity of the molecule to a control to determine whether the candidate agent modulates C_α-formylglycine generating activity of the molecule, wherein the molecule is an FGE nucleic acid molecule of the invention, or an expression product thereof. "Contacting" refers to both direct and indirect contacting of a molecule having C_α-formylglycine generating activity with the candidate agent. "Indirect" contacting means that the candidate agent exerts its effects on the C_α-formylglycine generating activity of the molecule via a third agent (e.g., a messenger molecule, a receptor, etc.). In certain embodiments, the control is C_α-formylglycine generating activity of the molecule measured in the absence of the candidate agent. Assaying methods and candidate agents are as described above in the foregoing embodiments with respect to FGE.

According to still another aspect of the invention, a method of diagnosing a disorder characterized by aberrant expression of a nucleic acid molecule, an expression product thereof, or a fragment of an expression product thereof, is provided. The method involves contacting a biological sample isolated from a subject with an agent that specifically binds to the nucleic acid molecule, an expression product thereof, or a fragment of an expression product thereof, and determining the interaction between the agent and the nucleic acid molecule or the expression product as a determination of the disorder, wherein the nucleic acid molecule is an FGE molecule according to the invention. The disorder is Multiple Sulfatase Deficiency. Mutations in the FGE gene that cause the aberrant expression of FGE molecules result in the following amino acid changes on SEQ ID NO:2: Met1Arg; Met1Val; Leu20Phe; Ser155Pro; Ala177Pro; Cys218Tyr; Arg224Trp; Asn259Ile; Pro266Leu; Ala279Val; Arg327Stop; Cys336Arg; Arg345Cys; Ala348Pro; Arg349Gln; Arg349Trp; Arg349Trp; Ser359Stop; or a combination thereof.

In the case where the molecule is a nucleic acid molecule, such determinations can be carried out via any standard nucleic acid determination assay, including the polymerase chain reaction, or assaying with labeled hybridization probes as exemplified herein. In the case where the molecule is an expression product of the nucleic acid molecule, or a fragment of an expression product of the nucleic acid molecule, such determination can be carried out via any standard immunological assay using, for example, antibodies which bind to any of the polypeptide expression products.

"Aberrant expression" refers to decreased expression (underexpression) or increased expression (overexpression) of FGE molecules (nucleic acids and/or polypeptides) in comparison with a control (i.e., expression of the same molecule in a healthy or "normal" subject). A "healthy subject", as used herein, refers to a subject who, according to standard medical standards, does not have or is at risk for developing Multiple Sulfatase Deficiency. Healthy subjects also do not otherwise exhibit symptoms of disease. In other words, such subjects, if examined by a medical professional, would be characterized as healthy and free of symptoms of a Multiple Sulfatase Deficiency. These include features of metachromatic leukodystrophy and of a mucopolysaccharidosis, such as increased amounts of acid mucopolysaccharides in several tissues, mild 'gargoylism', rapid neurologic deterioration, excessive presence of mucopolysaccharide and sulfatide in the urine, increased cerebrospinal fluid protein, and metachromatic degeneration of myelin in peripheral nerves.

The invention also provides novel kits which could be used to measure the levels of the nucleic acids of the invention, or expression products of the invention.

In one embodiment, a kit comprises a package containing an agent that selectively binds to any of the foregoing FGE isolated nucleic acids, or expression products thereof, and a control for comparing to a measured value of binding of said agent any of the foregoing FGE isolated nucleic acids or expression products thereof. In some embodiments, the control is a predetermined value for comparing to the measured value. In certain embodiments, the control comprises an epitope of the expression product of any of the foregoing FGE isolated nucleic acids. In one embodiment, the kit further comprises a second agent that selectively binds to a polypeptide selected from the group consisting of Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, or a peptide thereof, and a control for comparing to a measured value of binding of said second agent to said polypeptide or peptide thereof.

In the case of nucleic acid detection, pairs of primers for amplifying a nucleic acid molecule of the invention can be included. The preferred kits would include controls such as known amounts of nucleic acid probes, epitopes (such as Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, expression products) or anti-epitope antibodies, as well as instructions or other printed material. In certain embodiments the printed material can characterize risk of developing a sulfatase deficiency condition based upon the outcome of the assay. The reagents may be packaged in containers and/or coated on wells in predetermined amounts, and the kits may include standard materials such as labeled immunological reagents (such as labeled anti-IgG antibodies) and the like. One kit is a packaged polystyrene microtiter plate coated with FGE protein and a container containing labeled anti-human IgG antibodies. A well of the plate is contacted with, for example, a biological fluid, washed and then contacted with the anti-IgG antibody. The label is then detected. A kit embodying features of the present invention, generally designated by the numeral 11, is illustrated in Figure 25. Kit 11 is comprised of the following major elements: packaging 15, an agent of the invention 17, a control agent 19 and instructions 21. Packaging 15 is a box-like structure for holding a vial (or number of vials) containing an agent of the invention 17, a vial (or number of vials) containing a control agent 19, and instructions 21. Individuals skilled in the art can readily modify packaging 15 to suit individual needs.

The invention also embraces methods for treating Multiple Sulfatase Deficiency in a subject. The method involves administering to a subject in need of such treatment an agent that modulates C_α-formylglycine generating activity, in an amount effective to increase C_α-formylglycine generating activity in the subject. In some embodiments, the method further comprises co-administering an agent selected from the group consisting of a nucleic acid molecule encoding Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, an expression product of the nucleic acid molecule, and/or a fragment of the expression product of the nucleic acid molecule.

"Agents that modulate expression" of a nucleic acid or a polypeptide, as used herein, are known in the art, and refer to sense and antisense nucleic acids, dominant negative nucleic acids, antibodies to the polypeptides, and the like. Any agents that modulate exression of a molecule (and as described herein, modulate its activity), are useful according to the invention. In certain embodiments, the agent that modulates C_α-formylglycine generating activity is an isolated nucleic acid molecule of the invention (e.g., a nucleic acid of SEQ ID NO.3). In important embodiments, the agent that modulates C_α-formylglycine generating activity is a peptide of the invention (e.g., a peptide of SEQ ID NO.2). In some embodiments, the agent that modulates C_α-formylglycine generating activity is a sense nucleic acid of the invention.

According to one aspect of the invention, a method for for increasing C_α-formylglycine generating activity in a subject, is provided. The method involves administering an isolated FGE nucleic acid molecule of the invention, and/or an expression product thereof, to a subject, in an amount effective to increase C_α-formylglycine generating activity in the subject.

According to still another aspect of the invention, a method for increasing C_α-formylglycine generating activity in a cell, is provided. The method involves contacting the cell with an isolated nucleic acid molecule of the invention (e.g., a nucleic acid of SEQ ID NO.1), or an expression product thereof (e.g., a peptide of SEQ ID NO.2), in an amount effective to increase C_α-formylglycine generating activity in the cell. In important embodiments, the method involves activating the endogenous FGE gene to increase C_α-formylglycine generating activity in the cell.

In any of the foregoing embodiments the nucleic acid may be operatively coupled to a gene expression sequence which directs the expression of the nucleic acid molecule within a eukaryotic cell such as an HT-1080 cell. The "gene expression sequence" is any regulatory nucleotide sequence, such as a promoter sequence or promoter-enhancer combination, which facilitates the efficient transcription and translation of the nucleic acid to which it is operably linked. The gene expression sequence may, for example, be a mammalian or viral promoter, such as a constitutive or inducible promoter. Constitutive mammalian promoters include, but are not limited to, the promoters for the following genes: hypoxanthine phosphoribosyl transferase (HPTR), adenosine deaminase, pyruvate kinase, α-actin promoter and other constitutive promoters. Exemplary viral promoters which function constitutively in eukaryotic cells include, for example, promoters from the simian virus, papilloma virus, adenovirus, human immunodeficiency virus (HIV), Rous sarcoma virus, cytomegalovirus, the long terminal repeats (LTR) of moloney leukemia virus and other retroviruses, and the thymidine kinase promoter of herpes simplex virus. Other constitutive promoters are known to those of ordinary skill in the art. The promoters useful as gene expression sequences of the invention also include inducible promoters. Inducible promoters are activated in the presence of an inducing agent. For example, the metallothionein promoter is activated to increase transcription and translation in the presence of certain metal ions. Other inducible promoters are known to those of ordinary skill in the art.

In general, the gene expression sequence shall include, as necessary, 5' non-transcribing and 5' non-translating sequences involved with the initiation of transcription and translation, respectively, such as a TATA box, capping sequence, CAAT sequence, and the like. Especially, such 5' non-transcribing sequences will include a promoter region which includes a promoter sequence for transcriptional control of the operably joined nucleic acid. The gene expression sequences optionally includes enhancer sequences or upstream activator sequences as desired.

Preferably, any of the FGE nucleic acid molecules of the invention is linked to a gene expression sequence which permits expression of the nucleic acid molecule in a cell of a specific cell lineage, e.g., a neuron. A sequence which permits expression of the nucleic acid molecule in a cell such as a neuron, is one which is selectively active in such a cell type, thereby causing expression of the nucleic acid molecule in these cells. The synapsin-1 promoter, for example, can be used to express any of the foregoing nucleic acid molecules of the invention in a neuron; and the von Willebrand factor gene promoter, for example, can be used to express a nucleic acid molecule in a vascular endothelial cell. Those of ordinary skill

in the art will be able to easily identify alternative promoters that are capable of expressing a nucleic acid molecule in any of the preferred cells of the invention.

The nucleic acid sequence and the gene expression sequence are said to be "operably linked" when they are covalently linked in such a way as to place the transcription and/or translation of the nucleic acid coding sequence (e.g, in the case of FGE, SEQ ID NO. 3) under the influence or control of the gene expression sequence. If it is desired that the nucleic acid sequence be translated into a functional protein, two DNA sequences are said to be operably linked if induction of a promoter in the 5' gene expression sequence results in the transcription of the nucleic acid sequence and if the nature of the linkage between the two DNA sequences does not (1) result in the introduction of a frame-shift mutation, (2) interfere with the ability of the promoter region to direct the transcription of the nucleic acid sequence, and/or (3) interfere with the ability of the corresponding RNA transcript to be translated into a protein. Thus, a gene expression sequence would be operably linked to a nucleic acid sequence if the gene expression sequence were capable of effecting transcription of that nucleic acid sequence such that the resulting transcript might be translated into the desired protein or polypeptide.

The molecules of the invention can be delivered to the preferred cell types of the invention alone or in association with a vector (see also earlier discussion on vectors). In its broadest sense (and consistent with the description of expression and targeting vectors elsewhere herein), a "vector" is any vehicle capable of facilitating: (1) delivery of a molecule to a target cell and/or (2) uptake of the molecule by a target cell. Preferably, the delivery vectors transport the molecule into the target cell with reduced degradation relative to the extent of degradation that would result in the absence of the vector. Optionally, a "targeting ligand" can be attached to the vector to selectively deliver the vector to a cell which expresses on its surface the cognate receptor for the targeting ligand. In this manner, the vector (containing a nucleic acid or a protein) can be selectively delivered to a neuron. Methodologies for targeting include conjugates, such as those described in U.S. Patent 5,391,723 to Priest. Another example of a well-known targeting vehicle is a liposome. Liposomes are commercially available from Gibco BRL. Numerous methods are published for making targeted liposomes.

In general, the vectors useful in the invention include, but are not limited to, plasmids, phagemids, viruses, other vehicles derived from viral or bacterial sources that have been manipulated by the insertion or incorporation of the nucleic acid sequences of the invention, and additional nucleic acid fragments (e.g., enhancers, promoters) which can be attached to

the nucleic acid sequences of the invention. Viral vectors are a preferred type of vector and include, but are not limited to, nucleic acid sequences from the following viruses: adenovirus; adeno-associated virus; retrovirus, such as moloney murine leukemia virus; harvey murine sarcoma virus; murine mammary tumor virus; rouse sarcoma virus; SV40-type viruses; polyoma viruses; Epstein-Barr viruses; papilloma viruses; herpes virus; vaccinia virus; polio virus; and RNA virus such as a retrovirus. One can readily employ other vectors not named but known in the art.

A particularly preferred virus for certain applications is the adeno-associated virus, a double-stranded DNA virus. The adeno-associated virus is capable of infecting a wide range of cell types and species and can be engineered to be replication-deficient. It further has advantages, such as heat and lipid solvent stability, high transduction frequencies in cells of diverse lineages, including hematopoietic cells, and lack of superinfection inhibition thus allowing multiple series of transductions. Reportedly, the adeno-associated virus can integrate into human cellular DNA in a site-specific manner, thereby minimizing the possibility of insertional mutagenesis and variability of inserted gene expression. In addition, wild-type adeno-associated virus infections have been followed in tissue culture for greater than 100 passages in the absence of selective pressure, implying that the adeno-associated virus genomic integration is a relatively stable event. The adeno-associated virus can also function in an extrachromosomal fashion.

In general, other preferred viral vectors are based on non-cytopathic eukaryotic viruses in which non-essential genes have been replaced with the gene of interest. Non-cytopathic viruses include retroviruses, the life cycle of which involves reverse transcription of genomic viral RNA into DNA with subsequent proviral integration into host cellular DNA. Adenoviruses and retroviruses have been approved for human gene therapy trials. In general, the retroviruses are replication-deficient (i.e., capable of directing synthesis of the desired proteins, but incapable of manufacturing an infectious particle). Such genetically altered retroviral expression vectors have general utility for the high-efficiency transduction of genes *in vivo*. Standard protocols for producing replication-deficient retroviruses (including the steps of incorporation of exogenous genetic material into a plasmid, transfection of a packaging cell lined with plasmid, production of recombinant retroviruses by the packaging cell line, collection of viral particles from tissue culture media, and infection of the target cells with viral particles) are provided in Kriegler, M., "Gene Transfer and Expression, A Laboratory Manual," W.H. Freeman C.O., New York (1990) and Murry, E.J. Ed. "Methods in Molecular Biology," vol. 7, Humana Press, Inc., Cliffton, New Jersey (1991).

Another preferred retroviral vector is the vector derived from the moloney murine leukemia virus, as described in Nabel, E.G., et al., *Science*,1990, 249:1285-1288. These vectors reportedly were effective for the delivery of genes to all three layers of the arterial wall, including the media. Other preferred vectors are disclosed in Flugelman, et al., *Circulation*, 1992, 85:1110-1117. Additional vectors that are useful for delivering molecules of the invention are described in U.S. Patent No. 5,674,722 by Mulligan, et. al.

In addition to the foregoing vectors, other delivery methods may be used to deliver a molecule of the invention to a cell such as a neuron, liver, fibroblast, and/or a vascular endothelial cell, and facilitate uptake thereby.

A preferred such delivery method of the invention is a colloidal dispersion system. Colloidal dispersion systems include lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, and liposomes. A preferred colloidal system of the invention is a liposome. Liposomes are artificial membrane vessels which are useful as a delivery vector *in vivo* or *in vitro*. It has been shown that large unilamellar vessels (LUV), which range in size from 0.2 - 4.0 μm can encapsulate large macromolecules. RNA, DNA, and intact virions can be encapsulated within the aqueous interior and be delivered to cells in a biologically active form (Fraley, et al., *Trends Biochem. Sci.*, 1981, 6:77). In order for a liposome to be an efficient gene transfer vector, one or more of the following characteristics should be present: (1) encapsulation of the gene of interest at high efficiency with retention of biological activity; (2) preferential and substantial binding to a target cell in comparison to non-target cells; (3) delivery of the aqueous contents of the vesicle to the target cell cytoplasm at high efficiency; and (4) accurate and effective expression of genetic information.

Liposomes may be targeted to a particular tissue, such as the myocardium or the vascular cell wall, by coupling the liposome to a specific ligand such as a monoclonal antibody, sugar, glycolipid, or protein. Ligands which may be useful for targeting a liposome to the vascular wall include, but are not limited to the viral coat protein of the Hemagglutinating virus of Japan. Additionally, the vector may be coupled to a nuclear targeting peptide, which will direct the nucleic acid to the nucleus of the host cell.

Liposomes are commercially available from Gibco BRL, for example, as LIPOFECTIN™ and LIPOFECTACE™, which are formed of cationic lipids such as N-[1-(2, 3 dioleyloxy)-propyl]-N, N, N-trimethylammonium chloride (DOTMA) and dimethyl dioctadecylammonium bromide (DDAB). Methods for making liposomes are well known in the art and have been described in many publications. Liposomes also have been reviewed by Gregoriadis, G. in *Trends in Biotechnology*, V. 3, p. 235-241 (1985). Novel liposomes for

the intracellular delivery of macromolecules, including nucleic acids, are also described in PCT International application no. PCT/US96/07572 (Publication No. WO 96/40060, entitled "Intracellular Delivery of Macromolecules").

In one particular embodiment, the preferred vehicle is a biocompatible micro particle or implant that is suitable for implantation into the mammalian recipient. Exemplary bioerodible implants that are useful in accordance with this method are described in PCT International application no. PCT/US/03307 (Publication No. WO 95/24929, entitled "Polymeric Gene Delivery System", claiming priority to U.S. patent application serial no. 213,668, filed March 15, 1994). PCT/US/0307 describes a biocompatible, preferably biodegradable polymeric matrix for containing an exogenous gene under the control of an appropriate promoter. The polymeric matrix is used to achieve sustained release of the exogenous gene in the patient. In accordance with the instant invention, the nucleic acids described herein are encapsulated or dispersed within the biocompatible, preferably biodegradable polymeric matrix disclosed in PCT/US/03307. The polymeric matrix preferably is in the form of a micro particle such as a micro sphere (wherein a nucleic acid is dispersed throughout a solid polymeric matrix) or a microcapsule (wherein a nucleic acid is stored in the core of a polymeric shell). Other forms of the polymeric matrix for containing the nucleic acids of the invention include films, coatings, gels, implants, and stents. The size and composition of the polymeric matrix device is selected to result in favorable release kinetics in the tissue into which the matrix device is implanted. The size of the polymeric matrix devise further is selected according to the method of delivery which is to be used, typically injection into a tissue or administration of a suspension by aerosol into the nasal and/or pulmonary areas. The polymeric matrix composition can be selected to have both favorable degradation rates and also to be formed of a material which is bioadhesive, to further increase the effectiveness of transfer when the devise is administered to a vascular surface. The matrix composition also can be selected not to degrade, but rather, to release by diffusion over an extended period of time.

Both non-biodegradable and biodegradable polymeric matrices can be used to deliver the nucleic acids of the invention to the subject. Biodegradable matrices are preferred. Such polymers may be natural or synthetic polymers. Synthetic polymers are preferred. The polymer is selected based on the period of time over which release is desired, generally in the order of a few hours to a year or longer. Typically, release over a period ranging from between a few hours and three to twelve months is most desirable. The polymer optionally is

in the form of a hydrogel that can absorb up to about 90% of its weight in water and further, optionally is cross-linked with multi-valent ions or other polymers.

In general, the nucleic acids of the invention are delivered using the bioerodible implant by way of diffusion, or more preferably, by degradation of the polymeric matrix. Exemplary synthetic polymers which can be used to form the biodegradable delivery system include: polyamides, polycarbonates, polyalkylenes, polyalkylene glycols, polyalkylene oxides, polyalkylene terepthalates, polyvinyl alcohols, polyvinyl ethers, polyvinyl esters, poly-vinyl halides, polyvinylpyrrolidone, polyglycolides, polysiloxanes, polyurethanes and co-polymers thereof, alkyl cellulose, hydroxyalkyl celluloses, cellulose ethers, cellulose esters, nitro celluloses, polymers of acrylic and methacrylic esters, methyl cellulose, ethyl cellulose, hydroxypropyl cellulose, hydroxy-propyl methyl cellulose, hydroxybutyl methyl cellulose, cellulose acetate, cellulose propionate, cellulose acetate butyrate, cellulose acetate phthalate, carboxylethyl cellulose, cellulose triacetate, cellulose sulphate sodium salt, poly(methyl methacrylate), poly(ethyl methacrylate), poly(butylmethacrylate), poly(isobutyl methacrylate), poly(hexylmethacrylate), poly(isodecyl methacrylate), poly(lauryl methacrylate), poly(phenyl methacrylate), poly(methyl acrylate), poly(isopropyl acrylate), poly(isobutyl acrylate), poly(octadecyl acrylate), polyethylene, polypropylene, poly(ethylene glycol), poly(ethylene oxide), poly(ethylene terephthalate), poly(vinyl alcohols), polyvinyl acetate, poly vinyl chloride, polystyrene and polyvinylpyrrolidone.

Examples of non-biodegradable polymers include ethylene vinyl acetate, poly(meth) acrylic acid, polyamides, copolymers and mixtures thereof.

Examples of biodegradable polymers include synthetic polymers such as polymers of lactic acid and glycolic acid, polyanhydrides, poly(ortho)esters, polyurethanes, poly(butic acid), poly(valeric acid), and poly(lactide-cocaprolactone), and natural polymers such as alginate and other polysaccharides including dextran and cellulose, collagen, chemical derivatives thereof (substitutions, additions of chemical groups, for example, alkyl, alkylene, hydroxylations, oxidations, and other modifications routinely made by those skilled in the art), albumin and other hydrophilic proteins, zein and other prolamines and hydrophobic proteins, copolymers and mixtures thereof. In general, these materials degrade either by enzymatic hydrolysis or exposure to water *in vivo*, by surface or bulk erosion.

Bioadhesive polymers of particular interest include bioerodible hydrogels described by H.S. Sawhney, C.P. Pathak and J.A. Hubell in Macromolecules, 1993, 26, 581-587, the teachings of which are incorporated herein, polyhyaluronic acids, casein, gelatin, glutin, polyanhydrides, polyacrylic acid, alginate, chitosan, poly(methyl methacrylates), poly(ethyl

methacrylates), poly(butylmethacrylate), poly(isobutyl methacrylate), poly(hexylmethacrylate), poly(isodecyl methacrylate), poly(lauryl methacrylate), poly(phenyl methacrylate), poly(methyl acrylate), poly(isopropyl acrylate), poly(isobutyl acrylate), and poly(octadecyl acrylate). Thus, the invention provides a composition of the above-described molecules of the invention for use as a medicament, methods for preparing the medicament and methods for the sustained release of the medicament *in vivo*.

Compaction agents also can be used in combination with a vector of the invention. A "compaction agent", as used herein, refers to an agent, such as a histone, that neutralizes the negative charges on the nucleic acid and thereby permits compaction of the nucleic acid into a fine granule. Compaction of the nucleic acid facilitates the uptake of the nucleic acid by the target cell. The compaction agents can be used alone, i.e., to deliver an isolated nucleic acid of the invention in a form that is more efficiently taken up by the cell or, more preferably, in combination with one or more of the above-described vectors.

Other exemplary compositions that can be used to facilitate uptake by a target cell of the nucleic acids of the invention include calcium phosphate and other chemical mediators of intracellular transport, microinjection compositions, and electroporation.

The invention embraces methods for increasing sulfatase activity in a cell. Such methods involve contacting a cell expressing a sulfatase with an isolated nucleic acid molecule of of the invention (e.g., an isolated nucleic acid molecule as claimed in any one of Claims 1-8, an FGE nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87), or an expression product thereof (e.g., a polypeptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78), in an amount effective to increase sulfatase activity in the cell. "Increasing" sulfatase activity, as used herein, refers to increased affinity for, and/or conversion of, the specific substrate for the sulfatase, typically the result of an increase in *FGly* formation on the sulfatase molecule. In one embodiment, the cell expresses a sulfatase at levels higher than those of wild type cells. By "increasing sulfatase activity in a cell" also refers to increasing activity of a sulfatase that is secreted by the cell. The cell may express an endogenous and/or an exogenous sulfatase. Said contacting of the FGE molecule also refers to activating the cells's endogenous FGE gene. In important embodiments, the endogenous sulfatase is activated. In certain embodiments, the sulfatase is Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase

C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and/or HSulf-6. In certain embodiments the cell is a mammalian cell.

According to another aspect of the invention, a pharmaceutical composition, is provided. The composition comprises a sulfatase that is produced by cell, in a pharmaceutically effective amount to treat a sulfatase deficiency, and a pharmaceutically acceptable carrier, wherein said cell has been contacted with an agent comprising an isolated nucleic acid molecule of the invention (e.g., as claimed in Claims 1-8, or a nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87), or an expression product thereof (e.g., a peptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78). In important embodiments, the sulfatase is expressed at higher levels than normal/control cells.

The invention also embraces a sulfatase producing cell wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased. The cell comprises: (i) a sulfatase with an increased activity compared to a control, and (ii) a Formylglycine Generating Enzyme with an increased activity compared to a control, wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 5% over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme. It is known in the art that overexpression of sulfatases can decrease the activity of endogenous sulfatases (Anson et al., *Biochem. J.*, 1993, 294:657-662). Furthermore, only a fraction of the recombinant sulfatases is active. We have discovered, unexpectedly, that increased expression/activity of FGE in a cell with increased expression/activity of a sulfatase results in the production of a sulfatase that is more active. Since the presence of *FGly* on a sulfatase molecule is associated with sulfatase activity, "active sulfatase" can be quantitated by determining the presence of *FGly* on the sulfatase cell product using MALDI-TOF mass spectrometry, as described elsewhere herein. The ratio with total sulfatase can then be easily determined.

The invention also provides methods for the diagnosis and therapy of sulfatase deficiencies. Such disorders include, but are not limited to, Multiple Sulfatase Deficiency, Mucopolysaccharidosis II (MPS II; Hunter Syndrome), Mucopolysaccharidosis IIIA (MPS IIIA; Sanfilippo Syndrome A), Mucopolysaccharidosis VIII (MPS VIII), Mucopolysaccharidosis IVA (MPS IVA; Morquio Syndrome A), Mucopolysaccharidosis VI (MPS VI; Maroteaux-Lamy Syndrome), Metachromatic Leukodystrophy (MLD), X-linked

Recessive Chondrodysplasia Punctata 1, and X-linked Ichthyosis (Steroid Sulfatase Deficiency).

The methods of the invention are useful in both the acute and the prophylactic treatment of any of the foregoing conditions. As used herein, an acute treatment refers to the treatment of subjects having a particular condition. Prophylactic treatment refers to the treatment of subjects at risk of having the condition, but not presently having or experiencing the symptoms of the condition.

In its broadest sense, the terms "treatment" or "to treat" refer to both acute and prophylactic treatments. If the subject in need of treatment is experiencing a condition (or has or is having a particular condition), then treating the condition refers to ameliorating, reducing or eliminating the condition or one or more symptoms arising from the condition. In some preferred embodiments, treating the condition refers to ameliorating, reducing or eliminating a specific symptom or a specific subset of symptoms associated with the condition. If the subject in need of treatment is one who is at risk of having a condition, then treating the subject refers to reducing the risk of the subject having the condition.

The mode of administration and dosage of a therapeutic agent of the invention will vary with the particular stage of the condition being treated, the age and physical condition of the subject being treated, the duration of the treatment, the nature of the concurrent therapy (if any), the specific route of administration, and the like factors within the knowledge and expertise of the health practitioner.

As described herein, the agents of the invention are administered in effective amounts to treat any of the foregoing sulfatase deficiencies. In general, an effective amount is any amount that can cause a beneficial change in a desired tissue of a subject. Preferably, an effective amount is that amount sufficient to cause a favorable phenotypic change in a particular condition such as a lessening, alleviation or elimination of a symptom or of a condition as a whole.

In general, an effective amount is that amount of a pharmaceutical preparation that alone, or together with further doses, produces the desired response. This may involve only slowing the progression of the condition temporarily, although more preferably, it involves halting the progression of the condition permanently or delaying the onset of or preventing the condition from occurring. This can be monitored by routine methods. Generally, doses of active compounds would be from about 0.01 mg/kg per day to 1000 mg/kg per day. It is expected that doses ranging from 50μg-500 mg/kg will be suitable, preferably orally and in one or several administrations per day.

Such amounts will depend, of course, on the particular condition being treated, the severity of the condition, the individual patient parameters including age, physical condition, size and weight, the duration of the treatment, the nature of concurrent therapy (if any), the specific route of administration and like factors within the knowledge and expertise of the health practitioner. Lower doses will result from certain forms of administration, such as intravenous administration. In the event that a response in a subject is insufficient at the initial doses applied, higher doses (or effectively higher doses by a different, more localized delivery route) may be employed to the extent that patient tolerance permits. Multiple doses per day are contemplated to achieve appropriate systemic levels of compounds. It is preferred generally that a maximum dose be used, that is, the highest safe dose according to sound medical judgment. It will be understood by those of ordinary skill in the art, however, that a patient may insist upon a lower dose or tolerable dose for medical reasons, psychological reasons or for virtually any other reasons.

The agents of the invention may be combined, optionally, with a pharmaceutically-acceptable carrier to form a pharmaceutical preparation. The term "pharmaceutically-acceptable carrier" as used herein means one or more compatible solid or liquid fillers, diluents or encapsulating substances which are suitable for administration into a human. The term "carrier" denotes an organic or inorganic ingredient, natural or synthetic, with which the active ingredient is combined to facilitate the application. The components of the pharmaceutical compositions also are capable of being co-mingled with the molecules of the present invention, and with each other, in a manner such that there is no interaction which would substantially impair the desired pharmaceutical efficacy. In some aspects, the pharmaceutical preparations comprise an agent of the invention in an amount effective to treat a disorder.

The pharmaceutical preparations may contain suitable buffering agents, including: acetic acid in a salt; citric acid in a salt; boric acid in a salt; or phosphoric acid in a salt. The pharmaceutical compositions also may contain, optionally, suitable preservatives, such as: benzalkonium chloride; chlorobutanol; parabens or thimerosal.

A variety of administration routes are available. The particular mode selected will depend, of course, upon the particular drug selected, the severity of the condition being treated and the dosage required for therapeutic efficacy. The methods of the invention, generally speaking, may be practiced using any mode of administration that is medically acceptable, meaning any mode that produces effective levels of the active compounds without causing clinically unacceptable adverse effects. Such modes of administration

include oral, rectal, topical, nasal, intradermal, transdermal, or parenteral routes. The term "parenteral" includes subcutaneous, intravenous, intraomental, intramuscular, or infusion. Intravenous or intramuscular routes are not particularly suitable for long-term therapy and prophylaxis. As an example, pharmaceutical compositions for the acute treatment of subjects having a migraine headache may be formulated in a variety of different ways and for a variety of administration modes including tablets, capsules, powders, suppositories, injections and nasal sprays.

The pharmaceutical preparations may conveniently be presented in unit dosage form and may be prepared by any of the methods well-known in the art of pharmacy. All methods include the step of bringing the active agent into association with a carrier which constitutes one or more accessory ingredients. In general, the compositions are prepared by uniformly and intimately bringing the active compound into association with a liquid carrier, a finely divided solid carrier, or both, and then, if necessary, shaping the product.

Compositions suitable for oral administration may be presented as discrete units, such as capsules, tablets, lozenges, each containing a predetermined amount of the active compound. Other compositions include suspensions in aqueous liquids or non-aqueous liquids such as a syrup, elixir or an emulsion.

Compositions suitable for parenteral administration conveniently comprise a sterile aqueous preparation of an agent of the invention, which is preferably isotonic with the blood of the recipient. This aqueous preparation may be formulated according to known methods using suitable dispersing or wetting agents and suspending agents. The sterile injectable preparation also may be a sterile injectable solution or suspension in a non-toxic parenterally-acceptable diluent or solvent, for example, as a solution in 1,3-butane diol. Among the acceptable vehicles and solvents that may be employed are water, Ringer's solution, and isotonic sodium chloride solution. In addition, sterile, fixed oils are conventionally employed as a solvent or suspending medium. For this purpose any bland fixed oil may be employed including synthetic mono-or di-glycerides. In addition, fatty acids such as oleic acid may be used in the preparation of injectables. Formulations suitable for oral, subcutaneous, intravenous, intramuscular, etc. administrations can be found in Remington's Pharmaceutical Sciences, Mack Publishing Co., Easton, PA.

According to one aspect of the invention, a method for increasing C_{α}-formylglycine generating activity in a cell, is provided. The method involves contacting the cell with an isolated nucleic acid molecule of the invention (e.g., a nucleic acid of SEQ ID NO.1), or an expression product thereof (e.g., a peptide of SEQ ID NO.2), in an amount effective to

increase C_α-formylglycine generating activity in the cell. In important embodiments, the method involves activating the endogenous FGE gene to increase C_α-formylglycine generating activity in the cell. In some embodiments, the contacting is performed under conditions that permit entry of a molecule of the invention into the cell.

The term "permit entry" of a molecule into a cell according to the invention has the following meanings depending upon the nature of the molecule. For an isolated nucleic acid it is meant to describe entry of the nucleic acid through the cell membrane and into the cell nucleus, where upon the "nucleic acid transgene" can utilize the cell machinery to produce functional polypeptides encoded by the nucleic acid. By "nucleic acid transgene" it is meant to describe all of the nucleic acids of the invention with or without the associated vectors. For a polypeptide, it is meant to describe entry of the polypeptide through the cell membrane and into the cell cytoplasm, and if necessary, utilization of the cell cytoplasmic machinery to functionally modify the polypeptide (e.g., to an active form).

Various techniques may be employed for introducing nucleic acids of the invention into cells, depending on whether the nucleic acids are introduced *in vitro* or *in vivo* in a host. Such techniques include transfection of nucleic acid-$CaPO_4$ precipitates, transfection of nucleic acids associated with DEAE, transfection with a retrovirus including the nucleic acid of interest, liposome mediated transfection, and the like. For certain uses, it is preferred to target the nucleic acid to particular cells. In such instances, a vehicle used for delivering a nucleic acid of the invention into a cell (e.g., a retrovirus, or other virus; a liposome) can have a targeting molecule attached thereto. For example, a molecule such as an antibody specific for a surface membrane protein on the target cell or a ligand for a receptor on the target cell can be bound to or incorporated within the nucleic acid delivery vehicle. For example, where liposomes are employed to deliver the nucleic acids of the invention, proteins which bind to a surface membrane protein associated with endocytosis may be incorporated into the liposome formulation for targeting and/or to facilitate uptake. Such proteins include capsid proteins or fragments thereof tropic for a particular cell type, antibodies for proteins which undergo internalization in cycling, proteins that target intracellular localization and enhance intracellular half life, and the like. Polymeric delivery systems also have been used successfully to deliver nucleic acids into cells, as is known by those skilled in the art. Such systems even permit oral delivery of nucleic acids.

Other delivery systems can include time-release, delayed release or sustained release delivery systems. Such systems can avoid repeated administrations of an agent of the present invention, increasing convenience to the subject and the physician. Many types of release

delivery systems are available and known to those of ordinary skill in the art. They include polymer base systems such as poly(lactide-glycolide), copolyoxalates, polycaprolactones, polyesteramides, polyorthoesters, polyhydroxybutyric acid, and polyanhydrides. Microcapsules of the foregoing polymers containing drugs are described in, for example, U.S. Patent 5,075,109. Delivery systems also include non-polymer systems that are: lipids including sterols such as cholesterol, cholesterol esters and fatty acids or neutral fats such as mono- di- and tri-glycerides; hydrogel release systems; sylastic systems; peptide based systems; wax coatings; compressed tablets using conventional binders and excipients; partially fused implants; and the like. Specific examples include, but are not limited to: (a) erosional systems in which an agent of the invention is contained in a form within a matrix such as those described in U.S. Patent Nos. 4,452,775, 4,675,189, and 5,736,152, and (b) diffusional systems in which an active component permeates at a controlled rate from a polymer such as described in U.S. Patent Nos. 3,854,480, 5,133,974 and 5,407,686. In addition, pump-based hardware delivery systems can be used, some of which are adapted for implantation.

Use of a long-term sustained release implant may be desirable. Long-term release, as used herein, means that the implant is constructed and arranged to deliver therapeutic levels of the active ingredient for at least 30 days, and preferably 60 days. Long-term sustained release implants are well-known to those of ordinary skill in the art and include some of the release systems described above. Specific examples include, but are not limited to, long-term sustained release implants described in U.S. Patent No. 4,748,024, and Canadian Patent No. 1330939.

The invention also involves the administration, and in some embodiments co-administration, of agents other than the FGE molecules of the invention that when administered in effective amounts can act cooperatively, additively or synergistically with a molecule of the invention to: (i) modulate C_{α}-formylglycine generating activity, and (ii) treat any of the conditions in which C_{α}-formylglycine generating activity of a molecule of the invention is involved (e.g., a sulfatase deficiency including MSD). Agents other than the molecules of the invention include Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, or HSulf-6, (nucleic acids and polypeptides, and/or fragments thereof), and/or combinations thereof.

"Co-administering," as used herein, refers to administering simultaneously two or more compounds of the invention (e.g., an FGE nucleic acid and/or polypeptide, and an agent known to be beneficial in the treatment of, for example, a sulfatase deficiency -e.g., Iduronate 2-Sulfatase in the treatment of MPSII-), as an admixture in a single composition, or sequentially, close enough in time so that the compounds may exert an additive or even synergistic effect.

The invention also embraces solid-phase nucleic acid molecule arrays. The array consists essentially of a set of nucleic acid molecules, expression products thereof, or fragments (of either the nucleic acid or the polypeptide molecule) thereof, each nucleic acid molecule selected from the group consisting of FGE, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, fixed to a solid substrate. In some embodiments, the solid-phase array further comprises at least one control nucleic acid molecule. In certain embodiments, the set of nucleic acid molecules comprises at least one, at least two, at least three, at least four, or even at least five nucleic acid molecules, each selected from the group consisting of FGE, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6. In preferred embodiments, the set of nucleic acid molecules comprises a maximum number of 100 different nucleic acid molecules. In important embodiments, the set of nucleic acid molecules comprises a maximum number of 10 different nucleic acid molecules.

According to the invention, standard hybridization techniques of microarray technology are utilized to assess patterns of nucleic acid expression and identify nucleic acid expression. Microarray technology, which is also known by other names including: DNA chip technology, gene chip technology, and solid-phase nucleic acid array technology, is well known to those of ordinary skill in the art and is based on, but not limited to, obtaining an array of identified nucleic acid probes (e.g., molecules described elsewhere herein such as of FGE, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and/or HSulf-6) on a fixed substrate, labeling target molecules with reporter molecules (e.g., radioactive, chemiluminescent, or fluorescent tags such as

fluorescein, Cye3-dUTP, or Cye5-dUTP), hybridizing target nucleic acids to the probes, and evaluating target-probe hybridization. A probe with a nucleic acid sequence that perfectly matches the target sequence will, in general, result in detection of a stronger reporter-molecule signal than will probes with less perfect matches. Many components and techniques utilized in nucleic acid microarray technology are presented in *The Chipping Forecast*, Nature Genetics, Vol.21, Jan 1999, the entire contents of which is incorporated by reference herein.

According to the present invention, microarray substrates may include but are not limited to glass, silica, aluminosilicates, borosilicates, metal oxides such as alumina and nickel oxide, various clays, nitrocellulose, or nylon. In all embodiments a glass substrate is preferred. According to the invention, probes are selected from the group of nucleic acids including, but not limited to: DNA, genomic DNA, cDNA, and oligonucleotides; and may be natural or synthetic. Oligonucleotide probes preferably are 20 to 25-mer oligonucleotides and DNA/cDNA probes preferably are 500 to 5000 bases in length, although other lengths may be used. Appropriate probe length may be determined by one of ordinary skill in the art by following art-known procedures. In one embodiment, preferred probes are sets of two or more of the nucleic acid molecules set forth as SEQ ID NOs: 1, 3, 4, 6, 8, 10, and/or 12. Probes may be purified to remove contaminants using standard methods known to those of ordinary skill in the art such as gel filtration or precipitation.

In one embodiment, the microarray substrate may be coated with a compound to enhance synthesis of the probe on the substrate. Such compounds include, but are not limited to, oligoethylene glycols. In another embodiment, coupling agents or groups on the substrate can be used to covalently link the first nucleotide or olignucleotide to the substrate. These agents or groups may include, but are not limited to: amino, hydroxy, bromo, and carboxy groups. These reactive groups are preferably attached to the substrate through a hydrocarbyl radical such as an alkylene or phenylene divalent radical, one valence position occupied by the chain bonding and the remaining attached to the reactive groups. These hydrocarbyl groups may contain up to about ten carbon atoms, preferably up to about six carbon atoms. Alkylene radicals are usually preferred containing two to four carbon atoms in the principal chain. These and additional details of the process are disclosed, for example, in U.S. Patent 4,458,066, which is incorporated by reference in its entirety.

In one embodiment, probes are synthesized directly on the substrate in a predetermined grid pattern using methods such as light-directed chemical synthesis,

photochemical deprotection, or delivery of nucleotide precursors to the substrate and subsequent probe production.

In another embodiment, the substrate may be coated with a compound to enhance binding of the probe to the substrate. Such compounds include, but are not limited to: polylysine, amino silanes, amino-reactive silanes (Chipping Forecast, 1999) or chromium (Gwynne and Page, 2000). In this embodiment, presynthesized probes are applied to the substrate in a precise, predetermined volume and grid pattern, utilizing a computer-controlled robot to apply probe to the substrate in a contact-printing manner or in a non-contact manner such as ink jet or piezo-electric delivery. Probes may be covalently linked to the substrate with methods that include, but are not limited to, UV-irradiation. In another embodiment probes are linked to the substrate with heat.

Targets are nucleic acids selected from the group, including but not limited to: DNA, genomic DNA, cDNA, RNA, mRNA and may be natural or synthetic. In all embodiments, nucleic acid molecules from subjects suspected of developing or having a sulfatase deficiency, are preferred. In certain embodiments of the invention, one or more control nucleic acid molecules are attached to the substrate. Preferably, control nucleic acid molecules allow determination of factors including but not limited to: nucleic acid quality and binding characteristics; reagent quality and effectiveness; hybridization success; and analysis thresholds and success. Control nucleic acids may include, but are not limited to, expression products of genes such as housekeeping genes or fragments thereof.

To select a set of sulfatase deficiency disease markers, the expression data generated by, for example, microarray analysis of gene expression, is preferably analyzed to determine which genes in different categories of patients (each category of patients being a different sulfatase deficiency disorder), are significantly differentially expressed. The significance of gene expression can be determined using Permax computer software, although any standard statistical package that can discriminate significant differences is expression may be used. Permax performs permutation 2-sample t-tests on large arrays of data. For high dimensional vectors of observations, the Permax software computes t-statistics for each attribute, and assesses significance using the permutation distribution of the maximum and minimum overall attributes. The main use is to determine the attributes (genes) that are the most different between two groups (e.g., control healthy subject and a subject with a particular sulfatase deficiency), measuring "most different" using the value of the t-statistics, and their significance levels.

Expression of sulfatase deficiency disease related nucleic acid molecules can also be determined using protein measurement methods to determine expression of SEQ ID NOs: 2, e.g., by determining the expression of polypeptides encoded by SEQ ID NOs: 1, and/or 3. Preferred methods of specifically and quantitatively measuring proteins include, but are not limited to: mass spectroscopy-based methods such as surface enhanced laser desorption ionization (SELDI; e.g., Ciphergen ProteinChip System), non-mass spectroscopy-based methods, and immunohistochemistry-based methods such as 2-dimensional gel electrophoresis.

SELDI methodology may, through procedures known to those of ordinary skill in the art, be used to vaporize microscopic amounts of protein and to create a "fingerprint" of individual proteins, thereby allowing simultaneous measurement of the abundance of many proteins in a single sample. Preferably SELDI-based assays may be utilized to characterize multiple sulfatase deficiency as well as stages of such conditions. Such assays preferably include, but are not limited to the following examples. Gene products discovered by RNA microarrays may be selectively measured by specific (antibody mediated) capture to the SELDI protein disc (e.g., selective SELDI). Gene products discovered by protein screening (e.g., with 2-D gels), may be resolved by "total protein SELDI" optimized to visualize those particular markers of interest from among SEQ ID NOs: 1, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, and/or 28. Predictive models of a specific sulfatase deficiency from SELDI measurement of multiple markers from among SEQ ID NOs: 1, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, and/or 28, may be utilized for the SELDI strategies.

The use of any of the foregoing microarray methods to determine expression of a sulfatase deficiency disease related nucleic acids can be done with routine methods known to those of ordinary skill in the art and the expression determined by protein measurement methods may be correlated to predetermined levels of a marker used as a prognostic method for selecting treatment strategies for sulfatase deficiency disease patients.

The invention also embraces a sulfatase-producing cell wherein the ratio of active sulfatase to total sulfatase produced (i.e., the specific activity) by the cell is increased. The cell comprises: (i) a sulfatase with an increased expression, and (ii) a Formylglycine Generating Enzyme with an increased expression, wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 5% over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme.

A "sulfatase with an increased expression," as used herein, typically refers to increased expression of a sulfatase and/or its encoded polypeptide compared to a control.

Increased expression refers to increasing (i.e., to a detectable extent) replication, transcription, and/or translation of any of the sulfatase nucleic acids (sulfatase nucleic acids of the invention as described elsewhere herein), since upregulation of any of these processes results in concentration/amount increase of the polypeptide encoded by the gene (nucleic acid). This can be accomplished using a number of methods known in the art, also described elsewhere herein, such as transfection of a cell with the sulfatase cDNA, and/or genomic DNA encompassing the sulfatase locus, activating the endogenous sulfatase gene by placing, for example, a strong promoter element upstream of the endogenous sulfatase gene genomic locus using homologous recombination (see, e.g., the gene activation technology described in detail in U.S. Patents Nos. 5,733,761, 6,270,989, and 6,565,844, all of which are expressly incorporated herein by reference), etc. A typical control would be an identical cell transfected with a vector plasmid(s). Enhancing (or increasing) sulfatase activity also refers to preventing or inhibiting sulfatase degradation (e.g., *via* increased ubiquitinization), downregulation, etc., resulting, for example, in increased or stable sulfatase molecule $t_{1/2}$ (half-life) when compared to a control. Downregulation or decreased expression refers to decreased expression of a gene and/or its encoded polypeptide. The upregulation or downregulation of gene expression can be directly determined by detecting an increase or decrease, respectively, in the level of mRNA for the gene (e.g, a sulfatase), or the level of protein expression of the gene-encoded polypeptide, using any suitable means known to the art, such as nucleic acid hybridization or antibody detection methods, respectively, and in comparison to controls. Upregulation or downregulation of sulfatase gene expression can also be determined indirectly by detecting a change in sulfatase activity.

Similarily, a "Formylglycine Generating Enzyme with an increased expression," as used herein, typically refers to increased expression of an FGE nucleic acid of the invention and/or its encoded polypeptide compared to a control. Increased expression refers to increasing (i.e., to a detectable extent) replication, transcription, and/or translation of any of the FGE nucleic acids of the invention (as described elsewhere herein), since upregulation of any of these processes results in concentration/amount increase of the polypeptide encoded by the gene (nucleic acid). This can be accomplished using the methods described above (for the sulfatases), and elsewhere herein.

In certain embodiments, the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 10%, 15%, 20%, 50%, 100%, 200%, 500%, 1000%, over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme.

The invention further embraces an improved method for treating a sulfatase deficiency in a subject. The method involves administering to a subject in need of such treatment a sulfatase in an effective amount to treat the sulfatase deficiency in the subject, wherein the sulfatase is contacted with a Formylglycine Generating Enzyme in an amount effective to increase the specific activity of the sulfatase. As described elsewhere herein, "specific activity" refers to the ratio of active sulfatase to total sulfatase produced. "Contacted," as used herein, refers to FGE post-translationally modifying the sulfatase as described elsewhere herein. It would be apparent to one of ordinary skill in the art that an FGE can contact a sulfatase and modify it if nucleic acids encoding FGE and a sulfatase are co-expressed in a cell, or even if an isolated FGE polypeptide contacts an isolated sulfatase polypeptide *in vivo* or *in vitro*. Even though an isolated FGE polypeptide can be co-administered with an isolated sulfatase polypeptide to a subject to treat a sulfatase deficiency in the subject, it is preferred that the contact between FGE and the sulfatase takes place *in vitro* prior to administration of the sulfatase to the subject. This improved method of treatment is beneficial to a subject since lower amounts of the sulfatase need to be administered, and/or with less frequency, since the sulfatase is of higher specific activity.

The invention will be more fully understood by reference to the following examples. These examples, however, are merely intended to illustrate the embodiments of the invention and are not to be construed to limit the scope of the invention.

Examples

Example 1:

Multiple Sulfatase Deficiency is caused by mutations in the gene encoding the human C_α-formylglycine generating enzyme (FGE)

Experimental Procedures
Materials and Methods
In vitro assay for FGE

For monitoring the activity of FGE, the N-acetylated and C-amidated 23mer peptide P23 (MTDFYVPVSLCTPSRAALLTGRS) (SEQ ID NO:33) was used as substrate. The conversion of the Cysteine residue in position 11 to *FGly* was monitored by MALDI-TOF mass spectrometry. A 6μM stock solution of P23 in 30% acetonitrile and 0.1% trifluoroacetic acid (TFA) was prepared. Under standard conditions 6 pmol of P23 were incubated at 37°C with up to 10 μl enzyme in a final volume of 30 μl 50 mM Tris/HCl, pH 9.0, containing 67 mM NaCl, 15 μM $CaCl_2$, 2 mM DTT, and 0.33 mg/ml bovine serum

albumin. To stop the enzyme reaction 1.5 µl 10% TFA were added. P23 then was bound to ZipTip C18 (Millipore), washed with 0.1% TFA and eluted in 3µl 50% acetonitrile, 0.1% TFA. 0.5 µl of the eluate was mixed with 0.5 µl of matrix solution (5 mg/ml a-cyano-4-hydroxy-cinnamic acid (Bruker Daltonics, Billerica, MA) in 50% acetonitrile, 0.1% TFA) on a stainless steel target. MALDI-TOF mass spectrometry was performed with a Reflex III (Bruker Daltonics) using reflectron mode and laser energy just above the desorption/ionization threshold. All spectra were averages of 200-300 shots from several spots on the target. The mass axis was calibrated using peptides of molecular masses ranging from 1000 to 3000 Da as external standards. Monoisotopic MH^+ of P23 is 2526.28 and of the *FGly* containing product 2508.29. Activity (pmol product / h) was calculated on the basis of the peak height of the product divided by the sum of the peak heights of P23 and the product.

Purification of FGE from bovine testis

Bovine testes were obtained from the local slaughter house and stored for up to 20 h on ice. The parenchyme was freed from connective tissue and homogenized in a waring blendor and by three rounds of motor pottering. Preparation of rough microsomes (RM) by cell fractionation of the obtained homogenate was performed as described (Meyer et al., *J. Biol. Chem.*, 2000, 275:14550-14557) with the following modifications. Three differential centrifugation steps, 20 minutes each at 4°C, were performed at 500 g (JA10 rotor), 3000 g (JA10) and 10000 g (JA20). From the last supernatant the RM membranes were sedimented (125000 g, Ti45 rotor, 45 min, 4°C), homogenized by motor pottering and layered on a sucrose cushion (50 mM Hepes, pH 7.6, 50 mM KAc, 6 mM $MgAc_2$, 1 mM EDTA, 1.3 M sucrose, 5 mM β-mercaptoethanol). RMs were recovered from the pellet after spinning for 210 minutes at 45000 rpm in a Ti45 rotor at 4°C. Usually 100000-150000 equivalents RM, as defined by Walter and Blobel (*Methods Enzymol.*,1983, 96:84-93), were obtained from 1 kg of testis tissue. The reticuloplasm, i.e. the luminal content of the RM, was obtained by differential extraction at low concentrations of deoxy Big Chap, as described (Fey et al., *J. Biol. Chem.*, 2001, 276:47021-47028). For FGE purification, 95 ml of reticuloplasm were dialyzed for 20 h at 4 °C against 20 mM Tris/HCl, pH 8.0, 2.5 mM DTT, and cleared by centrifugation at 125000 g for 1 h. 32 ml-aliquots of the cleared reticuloplasm were loaded on a MonoQ HR10/10 column (Amersham Biosciences, Piscataway, NJ) at room temperature, washed and eluted at 2 ml/min with a linear gradient of 0 to 0.75 M NaCl in 80 ml of the Tris buffer. The fractions containing FGE activity, eluting at 50-165 mM NaCl, of three runs were pooled (42 ml) and mixed with 2 ml of Concanavalin A-Sepharose (Amersham Biosciences) that had been washed with 50 mM Hepes buffer, pH 7.4, containing 0.5 M KCl, 1 mM

$MgCl_2$, 1 mM $MnCl_2$, 1 mM $CaCl_2$, and 2.5 mM DTT. After incubation for 16 h at 4 °C, the Concanavalin A-Sepharose was collected in a column and washed with 6 ml of the same Hepes buffer. The bound material was eluted by incubating the column for 1 h at room temperature with 6 ml 0.5 M a-methylmannoside in 50 mM Hepes, pH 7.4, 2.5 mM DTT. The elution was repeated with 4 ml of the same eluent. The combined eluates (10 ml) from Concanavalin A-Sepharose were adjusted to pH 8.0 with 0.5 M Tris/HCl, pH 9.0, and mixed with 2 ml of Affigel 10 (Bio-Rad Laboratories, Hercules, CA) that had been derivatized with 10 mg of the scrambled peptide (PVSLPTRSCAALLTGR) (SEQ ID NO:34) and washed with buffer A (50 mM Hepes, pH 8.0, containing 0.15 M potassium acetate, 0.125 M sucrose, 1 mM $MgCl_2$, and 2.5 mM DTT). After incubation for 3 h at 4 °C the affinity matrix was collected in a column. The flow through and a wash fraction with 4 ml of buffer A were collected, combined and mixed with 2 ml of Affigel 10 that had been substituted with 10 mg of the Ser69 peptide (PVSLSTPSRAALLTGR) (SEQ ID NO:35) and washed with buffer A. After incubation overnight at 4°C, the affinity matrix was collected in a column, washed 3 times with 6 ml of buffer B (buffer A containing 2 M NaCl and a mixture of the 20 proteinogenic amino acids, each at 50 mg/ml). The bound material was eluted from the affinity matrix by incubating the Affigel twice for 90 min each with 6 ml buffer B containing 25 mM Ser69 peptide. An aliqout of the eluate was substituted with 1 mg/ml bovine serum albumin, dialyzed against buffer A and analyzed for activity. The remaining part of the activity (11.8 ml) was concentrated in a Vivaspin 500 concentrator (Vivascience AG, Hannover, Germany), and solubilized at 95 °C in Laemmli SDS sample buffer. The polypeptide composition of the starting material and preparations obtained after the chromatographic steps were monitored by SDSPAGE (15% acrylamide, 0.16% bisacrylamide) and staining with SYPRO Ruby (Bio-Rad Laboratories).

Identification of FGE by mass spectrometry

For peptide mass fingerprint analysis the purified polypeptides were in-gel digested with trypsin (Shevchenko et al., *Anal. Chem.*, 1996, 68:850-855), desalted on C18 ZipTip and analyzed by MALDI-TOF mass spectrometry using dihydrobenzoic acid as matrix and two autolytic peptides from trypsin (m/z 842.51 and 2211.10) as internal standards. For tandem mass spectrometry analysis selected peptides were analyzed by MALDI-TOF post-source decay mass spectrometry. Their corresponding doubly charged ions were isolated and fragmented by offline nano-ESI ion trap mass spectrometry (EsquireLC, Bruker Daltonics). The mass spectrometric data were used by Mascot search algorithm for protein identification in the NCBInr protein database and the NCBI EST nucleotide database.

Bioinformatics

Signal peptides and clevage sites were described with the method of von Heijne (von Heijne, *Nucleic Acids Res.*, 1986, 14:4683-90) implemented in EMBOSS (Rice et al., *Trends in Genetics*, 2000, 16:276-277). N-glycosylation sites were predicted using the algorithm of Brunak (Gupta and Brunak, *Pac. Symp. Biocomput.*, 2002, 310-22). Functional domains were detected by searching PFAM-Hidden-Markov-Models (version 7.8) (Sonnhammer et al., *Nucleic Acids Res.*, 1998, 26:320-322). To search for FGE homologs, the databases of the National Center for Biotechnology Information (Wheeler et al., *Nucleic Acids Res.*, 2002, 20:13-16) were queried with BLAST (Altschul et al., *Nucleic Acids Res.*, 1997, 25:3389-3402). Sequence similarities were computed using standard tools from EMBOSS. Genomic loci organisation and synteny were determined using the NCBI's human and mouse genome resources and the Human-Mouse Homology Map also form NCBI, Bethesda, MD).

Cloning of human FGE cDNA

Total RNA, prepared from human fibroblasts using the RNEASY™ Mini kit (Qiagen, Inc., Valencia, CA) was reverse transcribed using the OMNISCRIPT RT™ kit (Qiagen, Inc., Valencia, CA) and either an oligo(dT) primer or the FGE-specific primer 1199nc (CCAATGTAGGTCAGACACG) (SEQ ID NO:36). The first strand cDNA was amplified by PCR using the forward primer 1c (ACATGGCCCGCGGGAC) (SEQ ID NO:37) and, as reverse primer, either 1199nc or 1182nc (CGACTGCTCCTTGGACTGG) (SEQ ID NO:38). The PCR products were cloned directly into the pCR4-TOPO™ vector (Invitrogen Corporation, Carlsbad, CA). By sequencing multiple of the cloned PCR products, which had been obtained from various individuals and from independent RT and PCR reactions, the coding sequence of the FGE cDNA was determined (SEQ ID NOs:1 and 3).

Mutation detection, genomic sequencing, site-directed mutagenesis and Northern blot analysis

Standard protocols utilized in this study were essentially as described in Lübke et al. (*Nat. Gen.*, 2001, 28:73-76) and Hansske et al. (*J. Clin. Invest.*, 2002, 109:725-733). Northern blots were hybridized with a cDNA probe covering the entire coding region and a β-actin cDNA probe as a control for RNA loading.

Cell lines and cell culture

The fibroblasts from MSD patients 1-6 were obtained from E. Christenson (Rigshospitalet Copenhagen), M. Beck (Universitätskinderklinik Mainz), A. Kohlschütter (Universitätskrankenhaus Eppendorf, Hamburg), E. Zammarchi (Meyer Hospital, University of Florence), K. Harzer (Institut für Hirnforschung, Universität Tübingen), and A. Fensom

(Guy's Hospital, London), respectively. Human skin fibroblasts, HT-1080, BHK21 and CHO cells were maintained at 37°C under 5% CO_2 in Dulbecco's modified Eagle's medium containing 10% fetal calf serum.

Transfection, indirect immunofluorescence, Western blot analysis and detection of FGE activity

The FGE cDNA was equipped with a 5' EcoRI-site and either a 3' HA-, c-Myc or RGS-His_6-tag sequence, followed by a stop-codon and a HindIII site, by add-on PCR using Pfu polymerase (Stratagene, La Jolla, CA) and the following primers: GGAATTCGGGACAACATGGCTGCG (EcoRI) (SEQ ID NO:39), CCCAAGCTTATGC GTAGTCAGGCACATCATACGGATAGTCCATGGTGGGCAGGC(HA)(*SEQ ID NO:40*), CCCAAGCTTACAGGTCTTCTTCAGAAATCAGCTTTTGTTCGTCCATGGTGGGCAG GC (c-Myc) (SEQ ID NO:41), CCCAAGCTTAGTGATGGTGATGGTGATGCGATC CTCTGTCCATGGTGGGCAGGC (RGS-His_6) (SEQ ID NO:42). The resulting PCR products were cloned as EcoRI/HindIII fragments into pMPSVEH (Artelt et al., *Gene*, 1988, 68:213-219). The plasmids obtained were transiently transfected into HT-1080, BHK21 and CHO cells, grown on cover slips, using EFFECTENE™ (Qiagen) as transfection reagent. 48h after transfection the cells were analyzed by indirect immunofluorescence as described previously (Lübke et al., *Nat. Gen.*, 2001, 28:73-76; Hansske et al., *J. Clin. Invest.*, 2002, 109:725-733), using monoclonal IgG1 antibodies against HA (Berkeley Antibody Company, Richmond, CA), c-Myc (Santa Cruz Biotechnology, Inc., Santa Cruz, CA) or RGS-His (Qiagen) as primary antibodies. The endoplasmic reticulum marker protein proteindisulfide isomerase (PDI) was detected with a monoclonal antibody of different subtype (IgG2A, Stressgen Biotech., Victoria BC, Canada). The pimary antibodies werde detected with isotype-specific goat secondary antibodies coupled to CY2 or CY3, respectively (Molecular Probes, Inc., Eugene, OR). Immunofluorescence images were obtained on a Leica TCS Sp2 AOBS laser scan microscope. For Western blot analysis the same monoclonal antibodies and a HRP-conjugated anti-mouse IgG as secondary antibody were used. For determination of FGE activity, the trypsinised cells were washed with phosphate buffered saline containing a mixture of proteinase inhibitors (208 μM 4-(2- aminoethyl)benzene sulfonyl fluoride hydrochloride, 0.16 μM aprotinin, 4.2 μM leupeptin, 7.2 μM bestatin, 3 μM pepstatin A, 2.8 μM E-64), solubilized in 10 mM Tris, pH 8.0, containing 2.5 mM DTT, the proteinase inhibitors and 1% Triton X-100, and cleared by centrifugation at 125,000 g for 1h. The supernatant was subjected to chromatography on a MonoQ PC 1.6/5 column using the conditions described above. Fractions eluting at 50-200 mM NaCl were pooled, lyophilised

and reconstituted in one tenth of the original pool volume prior determination of FGE activity with peptide P23.

Retroviral transduction

cDNAs of interest were cloned into the Moloney murine leukemia virus based vector pLPCX and pLNCX2 (BD Biosciences Clontech, Palo Alto, CA). The transfection of ecotropic FNX-Eco cells (ATCC, Manassas, VA) and the transduction of amphotropic RETROPACK™ PT67 cells (BD Biosciences Clontech) and human fibroblasts was performed as described (Lübke et al., *Nat. Gen.*, 2001, 28:73-76; Thiel et al., *Biochem. J.*, 2002, 376, 195-201). For some experiments pLPCX-transduced PT67 cells were selected with puromycin prior determination of sulfatase activities.

Sulfatase assays

Activity of ASA, STS and GalNAc6S were determined as described in Rommerskirch and von Figura, *Proc. Natl. Acad. Sci., USA*, 1992, 89:2561-2565; Glössl and Kresse, *Clin. Chim. Acta*, 1978, 88:111-119.

Results

A rapid peptide based assay for FGE activity

We had developed an assay for determining FGE activity in microsome extracts using *in vitro* synthesized [^{35}S] ASA fragments as substrate. The fragments were added to the assay mixture as ribosome-associated nascent chain complexes. The quantitation of the product included tryptic digestion, separation of the peptides by RP-HPLC and identification and quantitation of the [^{35}S]-labeled *FGly* containing tryptic peptide by a combination of chemical derivatization to hydrazones, RP-HPLC separation and liquid scintillation counting (Fey et al., *J. Biol. Chem.*, 2001, 276:47021-47028). For monitoring the enzyme activity during purification, this cumbersome procedure needed to be modified. A synthetic 16mer peptide corresponding to ASA residues 65-80 and containing the sequence motif required for *FGly* formation inhibited the FGE activity in the *in vitro* assay. This suggested that peptides such as ASA65-80 may serve as substrates for FGE. We synthesized the 23mer peptide P23 (SEQ ID NO:33), which corresponds to ASA residues 60-80 with an additional N-acetylated methionine and a C-amidated serine residue to protect the N- and C-terminus, respectively. The cysteine and the *FGly* containing forms of P23 could be identified and quantified by matrix-assisted laser desorption/ionisation time of flight (MALDI-TOF) mass spectrometry. The presence of the *FGly* residue in position 11 of P23 was verified by MALDI-TOF post source decay mass spectrometry (see Peng et al., *J. Mass Spec.*, 2003, 38:80-86). Incubation of P23 with extracts from microsomes of bovine pancreas or bovine testis converted up to

95% of the peptide into a *FGly* containing derivative (Fig. 1). Under standard conditions the reaction was proportional to the amount of enzyme and time of incubation as long as less than 50% of the substrate was consumed and the incubation period did not exceed 24 h. The k_m for P23 was 13 nM. The effects of reduced and oxidized glutathione, Ca^{2+} and pH were comparable to those seen in the assay using ribosome-associated nascent chain complexes as substrate (Fey et al., *J. Biol. Chem.*, 2001, 276:47021-47028).

Purification of FGE

For purification of FGE the soluble fraction (reticuloplasm) of bovine testis microsomes served as the starting material. The specific activity of FGE was 10-20 times higher than that in reticuloplasm from bovine pancreas microsomes (Fey et al., *J. Biol. Chem.*, 2001, 276:47021-47028). Purification of FGE was achieved by a combination of four chromatographic steps. The first two steps were chromatography on a MonoQ anion exchanger and on Concanavalin A-Sepharose. At pH 8 the FGE activity bound to MonoQ and was eluted at 50-165 mM NaCl with 60-90% recovery. When this fraction was mixed with Concanavalin A-Sepharose, FGE was bound. 30-40% of the starting activity could be eluted with 0.5 M a-methyl mannoside. The two final purification steps were chromatography on affinity matrices derivatized with 16mer peptides. The first affinity matrix was Affigel 10 substituted with a variant of the ASA65-80 peptide, in which residues Cys69, Pro71 and Arg73, critical for *FGly* formation, were scrambled (scrambled peptide PVSLPTRSCAALLTGR -SEQ ID NO:34). This peptide did not inhibit FGE activity when added at 10 mM concentration to the *in vitro* assay and, when immobilized to Affigel 10, did not retain FGE activity. Chromatography on the scrambled peptide affinity matrix removed peptide binding proteins including chaperones of the endoplasmic reticulum. The second affinity matrix was Affigel 10 substituted with a variant of the ASA65-80 peptide, in which the Cys69 was replaced by a serine (Ser69 peptide PVSLSTPSRAALLTGR-SEQ ID NO:35). The Ser69 peptide affinity matrix efficiently bound FGE. The FGE activity could be eluted with either 2 M KSCN or 25 mM Ser69 peptide with 20-40% recovery. Prior to activity determination the KSCN or Ser69 peptide had to be removed by dialysis. The substitution of Cys69 by serine was crucial for the elution of active FGE. Affigel 10 substituted with the wildtype ASA65-80 peptide bound FGE efficiently. However, nearly no activity could be recovered in eluates with chaotropic salts (KSCN, $MgCl_2$), peptides (ASA65-80 or Ser69 peptide) or buffers with low or high pH. In Fig. 2 the polypeptide pattern of the starting material and of the active fractions obtained after the four chromatographic steps of a typical

purification is shown. In the final fraction 5% of the starting FGE activity and 0.0006% of the starting protein were recovered (8333-fold purification).

The purified 39.5 and 41.5 kDa polypeptides are encoded by a single gene

The 39.5 and 41.5 kDa polypeptides in the purified FGE preparation were subjected to peptide mass fingerprint analysis. The mass spectra of the tryptic peptides of the two polypeptides obtained by MALDI-TOF mass spectrometry were largely overlapping, suggesting that the two proteins originate from the same gene. Among the tryptic peptides of both polypeptides two abundant peptides MH^+ 1580.73, SQNTPDSSASNLGFR (SEQ ID NO:43), and MH^+ 2049.91, MVPIPAGVFTMGTDDPQIK -SEQ ID NO:44 plus two methionine oxidations) were found, which matched to the protein encoded by a cDNA with GenBank Acc. No. AK075459 (SEQ ID NO:4). The amino acid sequence of the two peptides was confirmed by MALDI-TOF post source decay spectra and by MS/MS analysis using offline nano-electrospray ionisation (ESI) iontrap mass spectrometry. An EST sequence of the bovine ortholog of the human cDNA covering the C-terminal part of the FGE and matching the sequences of both peptides provided additional sequence information for bovine FGE.

Evolutionary conservation and domain structure of FGE

The gene for human FGE is encoded by the cDNA of (SEQ ID NOs:1 and/or 3) and located on chromosome 3p26. It spans ~105 kb and the coding sequence is distributed over 9 exons. Three orthologs of the human FGE gene are found in mouse (87% identity), *Drosophila melanogaster* (48% identity), and *Anopheles gambiae* (47% identity). Orthologous EST sequences are found for 8 further species including cow, pig, *Xenopus laevis*, *Silurana tropicalis*, zebra fish, salmon and other fish species (for details see Example 2). The exon-intron structure between the human and the mouse gene is conserved and the mouse gene on chromosome 6E2 is located within a region syntenic to the human chromosome 3p26. The genomes of *S. cerevisiae* and *C. elegans* lack FGE homologs. In prokaryotes 12 homologs of human FGE were found. The cDNA for human FGE is predicted to encode a protein of 374 residues (Fig. 3 and SEQ ID NO:2). The protein contains a cleavable signal sequence of 33 residues, which indicates translocation of FGE into the endoplasmic reticulum, and contains a single N-glycosylation site at Asn141. The binding of FGE to concanavalin A suggests that this N-glycosylation site is utilized. Residues 87-367 of FGE are listed in the PFAM protein motif database as a domain of unknown function (PFAM: DUF323). Sequence comparison analysis of human FGE and its eukaryotic

orthologs identified in data bases indicates that this domain is composed of three distinct subdomains.

The N-terminal subdomain (residues 91-154 in human FGE) has a sequence identity of 46% and a similarity of 79% within the four known eukaryotic FGE orthologs. In human FGE, this domain carries the N-glycosylation site at Asn 141, which is conserved in the other orthologs. The middle part of FGE (residues 179-308 in human FGE) is represented by a tryptophan-rich subdomain (12 tryptophans per 129 residues). The identity of the eukaryotic orthologs within this subdomain is 57%, the similarity is 82%. The C-terminal subdomain (residues 327-366 in human FGE) is the most highly conserved sequence within the FGE family. The sequence identity of the human C-terminal subdomain with the eukaryotic orthologs (3 full length sequences and 8 ESTs) is 85%, the similarity 97%. Within the 40 residues of the subdomain 3 four cysteine residues are fully conserved. Three of cysteins are also conserved in the prokaryotic FGE orthologs. The 12 prokaryotic members of the FGE-family (for details see Example 2) share the subdomain structure with eukaryotic FGEs. The boundaries between the three subdomains are more evident in the prokaryotic FGE family due to non-conserved sequences of variable length separating the subdomains from each other. The human and the mouse genome encode two closely related homologs of FGE (SEQ ID NOs:43 and 44, GenBank Acc. No. NM_015411, in man, and SEQ ID NOs:45 and 46, GenBank Acc. No. AK076022, in mouse). The two paralogs are 86% identical. Their genes are located on syntenic chromosome regions (7q11 in human, 5G1 in mouse). Both paralogs share with the FGE orthologs the subdomain structure and are 35% identical and 47% similar to human FGE. In the third subdomain, which is 100% identical in both homologs, the cysteine containing undecamer sequence of the subdomain 3 is missing.

Expression, subcellular localization and molecular forms

A single transcript of 2.1 kb is detectable by Northern blot analysis of total RNA from skin fibroblasts and poly A^+ RNA from heart, brain, placenta, lung, liver, skeletal muscle, kidney and pancreas. Relative to β-actin RNA the abundance varies by one order of magnitude and is highest in pancreas and kidney and lowest in brain. Various eukaryotic cell lines stably or transiently expressing the cDNA of human FGE or FGE derivatives C-terminally extended by a HA-, Myc- or His_6-tag were assayed for FGE activity and subcellular localization of FGE. Transient expression of tagged and non-tagged FGE increased the FGE activity 1.6 – 3.9-fold. Stable expression of FGE in PT67 cells increased the activity of FGE about 100-fold. Detection of the tagged FGE form by indirect immunofluorescence in BHK 21, CHO, and HT1080 cells showed a colocalization of the

variously tagged FGE forms with proteindisulfide isomerase, a lumenal protein of the endoplasmic reticulum. Western blot analysis of extracts from BHK 21 cells transiently transfected with cDNA encoding tagged forms of FGE showed a single immunoreactive band with an apparent size between 42 to 44 kDa.

The FGE gene carries mutations in MSD

MSD is caused by a deficiency to generate *FGly* residues in sulfatases (Schmidt, B., et al., *Cell*, 1995, 82:271-278). The FGE gene is therefore a candidate gene for MSD. We amplified and sequenced the FGE encoding cDNA of seven MSD patients and found ten different mutations that were confirmed by sequencing the genomic DNA (Table 1).

Table 1: Mutations in MSD patients

Mutation	Effect on Protein	Remarks	Patient
1076C>A	S359X	Truncation of the C-terminal 16 residues	1*
IVS3+5-8 del	Deletion of residues 149-173	In-frame deletion of exon 3	1, 2
979C>T	R327X	Loss of subdomain 3	2
1045C>T	R349W	Substitution of a conserved residue in subdomain 3	3, 7
1046G>A	R349Q	Substitution of a conserved residue in subdomain 3	4
1006T>C	C336R	Substitution of a conserved residue in subdomain 3	4
836C>T	A279V	Substitution of a conserved residue in subdomain 2	5
243delC	frameshift and truncation	Loss of all three subdomains	5
661delG	frameshift and truncation	Loss of the C-terminal third of FGE including subdomain 3	6**
IVS6-1G>A	Deletion of residues 281-318	In-frame deletion of exon 7	5

*Patient 1 is the MSD patient Mo. in Schmidt, B., et al., *Cell*, 1995, 82:271-278 and Rommerskirch and von Figura, *Proc. Natl. Acad. Sci., USA*, 1992, 89:2561-2565.
**Patient 6 is the MSD patient reported by Burk et al., *J. Pediatr.*, 1984, 104:574-578.
The other patients represent unpublished cases.

The first patient was heterozygous for a 1076C>A substitution converting the codon for serine 359 into a stop codon (S359X) and a mutation causing the deletion of the 25 residues 149-173 that are encoded by exon 3 and space the first and the second domain of the protein. Genomic sequencing revealed a deletion of nucleotides +5-8 of the third intron (IVS3+5-8 del) thereby destroying the splice donor site of intron 3. The second patient was heterozygous for the mutation causing the loss of exon 3 (IVS3+5-8 del) and a 979C>T

substitution converting the codon for arginine 327 into a stop codon (R327X). The truncated FGE encoded by the 979C>T allele lacks most of subdomain 3. The third patient was homozygous for a 1045C>T substitution replacing the conserved arginine 349 in subdomain 3 by tryptophan (R349W). The fourth patient was heterozygous for two missense mutations replacing conserved residues in the FGE domain: a 1046>T substitution replacing arginine 349 by glutamine (R349Q) and a 1006T>C substitution replacing cysteine 336 by arginine (C336R). The fifth patient was heterozygous for a 836 C>T substitution replacing the conserved alanine 279 by valine (A279V). The second mutation is a single nucleotide deletion (243delC) changing the sequence after proline 81 and causing a translation stop after residue 139. The sixth patient was heterozygous for the deletion of a single nucleotide (661delG) changing the amino acid sequence after residue 220 and introducing a stop codon after residue 266. The second mutation is a splice acceptor site mutation of intron 6 (IVS6-1G>A) causing an in-frame deletion of exon 7 encoding residues 281-318. In the seventh patient the same 1045C>T substitution was found as in the third patient. In addition we detected two polymorphisms in the coding region of 18 FGE alleles from controls and MSD patients. 22% carried a 188G>A substitution, replacing serine 63 by asparagine (S63N) and 28% a silent 1116C>T substitution.

Transduction of MSD fibroblasts with wild type and mutant FGE cDNA

In order to confirm the deficiency of FGE as the cause of the inactivity of sulfatases synthesized in MSD, we expressed the FGE cDNA in MSD fibroblasts utilizing retroviral gene transfer. As a control we transduced the retroviral vector without cDNA insert. To monitor the complementation of the metabolic defect the activity of ASA, steroid sulfatase (STS) and N-acetylgalactosamine 6-sulfatase (GalNAc6S) were measured in the transduced fibroblasts prior or after selection. Transduction of the wild type FGE partially restored the catalytic activity of the three sulfatases in two MSD-cell lines (Table 2) and for STS in a third MSD cell line. It should be noted that for ASA and GalNAc6S the restoration was only partial after selection of the fibroblasts reaching 20 to 50% of normal activity. For STS the activity was found to be restored to that in control fibroblasts after selection. Selection increased the activity of ASA and STS by 50 to 80%, which is compatible with the earlier observation that 15 to 50% of the fibroblasts become transduced (Lübke et al., *Nat. Gen.*, 2001, 28:73-76). The sulfatase activities in the MSD fibroblasts transduced with the retroviral vector alone (Table 2) were comparable to those in non-transduced MSD fibroblasts (not shown). Transduction of FGE cDNA carrying the IVS3+5-8del mutation failed to restore the sulfatase activities (Table 2).

Table 2: Complementation of MSD fibroblasts by transduction of wild type or mutant FGE cDNA

Fibroblasts	FGE-insert	Sulfatase		
		ASA[1]	STS[1]	GalNAc6S[1]
MSD 3°	-	1.9 ± 0.2	< 3	56.7 ± 32
	FGE^{+}	7.9	13.5	n. d.
	FGE^{++}	12.2 ± 0.2	75.2	283 ± 42
	FGE-IVS3+5-8del^{+}	1.8	< 3	n. d.
	FGE-IVS3+5-8del^{++}	2.1	< 3	98.5
MSD 4°	-	1.1 ± 0.3	< 3	n. d.
	FGE^{+}	4.7	17.0	n. d.
Control fibroblasts		58 ± 11	66 ± 31	828 ± 426

[1]The values give the ratio between ASA (mU/mg cell protein), STS (μU/mg cell protein), GalNAc6S (μU/mg cell protein) and that of β-hexosaminidase (U/mg cell protein). For control fibroblasts the mean and the variation of 6-11 cell lines is given. Where indicated the range of two cultures transduced in parallel is given for MSD fibroblasts.

° The number of MSD fibroblasts refers to that of the patient in Table 1.

+ Activity determination prior to selection.

++ Activity determination after selection.

n.d.: not determined

Discussion

FGE is a highly conserved glycoprotein of the endoplasmic reticulum.

Purification of FGE from bovine testis yielded two polypeptides of 39.5 and 41.5 kDa which originate from the same gene. The expression of three differently tagged versions of FGE in three different eukaryotic cell lines as a single form suggests that one of the two forms observed in the FGE preparation purified from bovine testis may have been generated by limited proteolysis during purification. The substitution of Cys69 in ASA65-80 peptide by serine was critical for the purification of FGE by affinity chromatography. FGE has a cleavable signal sequence that mediates translocation across the membrane of the endoplasmic reticulum. The greater part of the mature protein (275 residues out of 340) defines a unique domain, which is likely to be composed of three subdomains (see Example 2), for none of the three subdomains homologs exist in proteins with known function. The recognition of the linear *FGly* modification motif in newly synthesized sulfatase polypeptides (Dierks et al., *EMBO J.*, 1999, 18:2084-2091) could be the function of a FGE subdomain. The catalytic domain could catalyse the *FGly* formation in several ways. It has been proposed that FGE abstracts electrons from the thiol group of the cysteine and transfers them to an acceptor. The resulting thioaldehyde would spontaneously hydrolyse to *FGly* and H_2S

(Schmidt, B., et al., *Cell*, 1995, 82:271-278). Alternatively FGE could act as a mixed-function oxygenase (monooxygenase) introducing one atom of O_2 into the cysteine and the other in H_2O with the help of an electron donor such as $FADH_2$. The resulting thioaldehyde hydrate derivative of cysteine would spontaneously react to *FGly* and H_2S. Preliminary experiments with a partially purified FGE preparation showed a critical dependence of the *FGly* formation on molecular oxygen. This would suggest that FGE acts as a mixed-function oxygenase. The particular high conservation of subdomain 3 and the presence of three fully conserved cysteine residues therein make this subdomain a likely candidate for the catalytic site. It will be interesting to see whether the structural elements mediating the recognition of the *FGly* motif and the binding of an electron acceptor or electron donor correlate with the domain structure of FGE.

Recombinant FGE is localized in the endoplasmic reticulum, which is compatible with the proposed site of its action. *FGly* residues are generated in newly synthesized sulfatases during or shortly after their translocation into the endoplasmic reticulum (Dierks et al., *Proc. Natl. Acad. Sci. U.S.A.*, 1997, 94:11963-11968; Dierks et al., *FEBS Lett.*, 1998, 423:61-65). FGE itself does not contain an ER-retention signal of the KDEL type. Its retention in the endoplasmic reticulum may therefore be mediated by the interaction with other ER proteins. Components of the translocation/ N-glycosylation machinery are attractive candidates for such interacting partners.

Mutations in FGE cause MSD

We have shown that mutations in the gene encoding FGE cause MSD. FGE also may interact with other components, and defects in genes encoding the latter could equally well cause MSD. In seven MSD patients we indeed found ten different mutations in the FGE gene. All mutations have severe effects on the FGE protein by replacing highly conserved residues in subdomain 3 (three mutations) or subdomain 2 (one mutation) or C-terminal truncations of various lengths (four mutations) or large inframe deletions (two mutations). For two MSD-cell lines and one of the MSD mutations it was shown that transduction of the wild type, but not of the mutant FGE cDNA, partially restores the sulfatase activities. This clearly identifies the FGE gene as the site of mutation and the disease causing nature of the mutation. MSD is both clinically and biochemically heterogenous. A rare neonatal form presenting at birth and developing a hydrocephalus, a common form resembling initially to an infantile metachromatic leukodystrophy and subsequently developing ichthyosis- and mucopolysaccharidosis-like features, and a less frequent mild form in which the clinical features of a mucopolysaccharidosis prevail, have been differentiated. Biochemically it is

characteristic that a residual activity of sulfatases can be detected, which for most cases in cultured skin fibroblasts is below 10% of controls (Burch et al., *Clin. Genet.*, 1986, 30:409-15; Basner et al., *Pediatr. Res.*, 1979, 13:1316-1318). However, in some MSD cell lines the activity of selected sulfatases can reach the normal range (Yutaka et al., *Clin. Genet.*, 1981, 20:296-303). Furthermore, the residual activity has been reported to be subject to variations depending on the cell culture conditions and unknown factors. Biochemically, MSD has been classified into two groups. In group I the residual activity of sulfatases is below 15% including that of ASB. In group II the residual activity of sulfatases is higher and particularly that of ASB may reach values of up to 50-100% of control. All patients reported here fall into group I except patient 5, which falls into group II (ASB activity in the control range) of the biochemical phenotype. Based on clinical criteria patients 1 and 6 are neonatal cases, while patients 2-4 and 7 have the common and patient 5 the mucopolysaccharidosis-like form of MSD.

The phenotypic heterogeneity suggests that the different mutations in MSD patients are associated with different residual activities of FGE. Preliminary data on PT67 cells stably expressing FGE IVS3+5-8del indicate that the in-frame deletion of exon 3 abolishes FGE activity completely. The characterization of the mutations in MSD, of the biochemical properties of the mutant FGE and of the residual content of *FGly* in sulfatases using a recently developed highly sensitive mass spectrometric method (Peng et al., *J. Mass Spec.*, 2003, 38:80-86) will provide a better understanding of the genotype-phenotype correlation in MSD.

Example 2:
The human FGE gene defines a new gene family modifying sulfatases which is conserved from prokaryotes to eukaryotes

Bioinformatics

Signal peptides and cleavage sites were described with the method of von Heijne (*Nucleic Acids Res.*, 1986, 14:4683) implemented in EMBOSS (Rice et al., *Trends in Genetics*, 2000, 16:276-277), and the method of Nielsen et al. (*Protein Engineering*, 1997, 10:1-6). N-glycosylation sites were predicted using the algorithm of Brunak (Gupta and Brunak, *Pac. Symp. Biocomput.*, 2002, 310-22).

Functional domains were detected by searching PFAM-Hidden-Markov-Models (version 7.8) (Sonnhammer et al., *Nucleic Acids Res.*, 1998, 26:320-322). Sequences from the PFAM DUF323 seed were obtained from TrEMBL (Bairoch, A. and Apweiler, R., *Nucleic Acids Res.*, 2000, 28:45-48). Multiple alignments and phylogenetic tree constructions were

performed with Clustal W (Thompson, J., et al., *Nucleic Acids Res.*, 1994, 22:4673-4680). For phylogenetic tree computation, gap positions were excluded and multiple substitutions were corrected for. Tree bootstraping was performed to obtain significant results. Trees were visualised using Njplot (Perriere, G. and Gouy, M., *Biochimie*, 1996, 78:364-369). Alignments were plotted using the pret- typlot command from EMBOSS.

To search for FGE homologs, the databases NR, NT and EST of the National Center for Biotechnology Information (NCBI) (Wheeler et al., *Nucleic Acids Res.*, 2002, 20:13-16), were queried with BLAST (Altschul et al., *Nucleic Acids Res.*, 1997, 25:3389-3402). For protein sequences, the search was performed using iterative converging Psi-Blast against the current version of the NR database using an expectation value cutoff of 10^{-40}, and default parameters. Convergence was reached after 5 iterations. For nucleotide sequences, the search was performed with Psi-TBlastn: using NR and the protein sequence of human FGE as input, a score matrix for hFGE was built with iterative converging Psi-Blast. This matrix was used as input for blastall to query the nucleotide databses NT and EST. For both steps, an expectation value cutoff of 10^{-20} was used.

Protein secondary structure prediction was done using Psipred (Jones, D., *J Mol Biol.*, 1999, 292:1950-202; McGuffin, L., et al., *Bioinformatics*, 2000, 16:404-405).

Similarity scores of the subdomains were computed from alignments using the cons algorithm form EMBOSS with default parameters. The metaalignments were generated by aligning consensus sequences of the FGE-family subgroups. Genomic loci organisation and synteny were determined using the NCBI's human and mouse genome resources at NCBI (Bethesda, MD) and Softberry's (Mount Kisco, NY) Human- Mouse-Rat Synteny. Bacterial genome sequences were downloaded from the NCBI-FTP-server. The NCBI microbial genome annotation was used to obtain an overview of the genomic loci of bacterial FGE genes.

Results and Discussion

Basic features and motifs of human FGE and related proteins

The human FGE gene (SEQ ID NOs:1, 3) encodes the FGE protein (SEQ ID NO:2) which is predicted to have 374 residues. A cleavage signal between residues 22-33 (Heijne-Score of 15.29) and a hydropathy-score (Kyte, J. and Doolittle, R., *J Mol Biol.*, 1982, 157:105-132) of residues 17-29 between 1.7 and 3.3 indicate that the 33 N-terminal residues are cleaved off after ER-translocation. However with the algorithm of Nielsen et al. (*Protein Engineering*, 1997, 10:1-6), cleavage of the signal sequence is predicted after residue 34. The protein has a single potential N-glycosylation site at Asn 141.

A search with the FGE protein sequence against the protein motif database PFAM (Sonnhammer et al., *Nucleic Acids Res.*, 1998, 26:320-322) revealed that residues 87-367 of human FGE can be classified as the protein domain DUF323 ("domain of unknown function", PF03781) with a highly significant expectation value of $7{:}9{*}10^{-114}$. The PFAM-seed defining DUF323 consists of 25 protein sequences, of which the majority are hypothetical proteins derived from sequencing data. To analyse the relationship between human FGE and DUF323, a multiple alignment of FGE with the sequences of the DUF323 seed was performed. Based on this, a phylogenetic tree was constructed and bootstraped. Four of the hypothetical sequences (TrEMBL-IDs Q9CK12, Q9I761, O94632 and Q9Y405) had such a strong divergence from the other members of the seed that they prevented successfull bootstraping and had to be removed from the set. Figure 2 shows the bootstraped tree displaying the relationship between human FGE and the remaining 21 DUF323 seed proteins. The tree can be used to subdivide the seed members into two categories: homologs closely related to human FGE and the remaining, less related genes.

The topmost 7 proteins have a phylogenetic distance between 0.41 and 0.73 to human FGE. They only contain a single domain, DUF323. The homology within this group extends over the whole amino acid sequence, the greater part of which consists of the DUF323 domain. The DUF323 domain is strongly conserved within this group of homologs, while the other 15 proteins of the seed are less related to human FGE (phylogenetic distance between 1.14 and 1.93). Their DUF323 domain diverges considerably from the highly conserved DUF323-domain of the first group (cf. section "Subdomains of FGE and mutations in the FGE gene"). Most of these 15 proteins are hypothetical, six of them have been further investigated. One of them, a serine/threonine kinase (TrEMBL:O84147) from C. trachomatis contains other domains in addition to DUF323: an ATP-binding domain and a kinase domain. The sequences from R. sphaeroides (TrEMBL: Q9ALV8) and Pseudomonas sp.(TrEMBL: O52577) encode the protein NirV, a gene cotranscribed with the copper-containing nitrite reductase nirK (Jain, R. and Shapleigh, J., *Microbiology*, 2001, 147:2505-2515). CarC (TrEMBL: Q9XB56) is an oxygenase involved in the synthesis of a β-lactam antibiotic from E. carotovora (McGowan, S., et al., *Mol Microbiol.*, 1996, 22:415-426; Khaleeli N, T. C., and Busby RW, *Biochemistry*, 2000, 39:8666-8673). XylR (TrEMBL: O31397) and BH0900 (TrEMBL: Q9KEF2) are enhancer binding proteins involved in the regulation of pentose utilisation (Rodionov, D., et al., *FEMS Microbiol Lett.*, 2001, 205:305-314) in bacillaceae and clostridiaceae. The comparison of FGE and DUF323 led to the establishment of a homology threshold differentiating the FGE family from distant DUF323-containing

homologs with different functions. The latter include a serine/threonine kinase and XylR, a transcription enhancer as well as FGE, a *FGly* generating enzyme and CarC, an oxygenase. As discussed in elsewhere herein, FGE might also exert its cysteine modifying function as an oxygenase, suggesting that FGE and non-FGE members of the DUF323 seed may share an oxygenase function.

Homologs of FGE

The presence of closely related homologs of human FGE in the DUF323 seed directed us to search for homologs of human FGE in NCBI's NR database (Wheeler et al., *Nucleic Acids Res.*, 2002, 20:13-16). The threshold of the search was chosen in such a way that all 6 homologs present in the DUF323 seed and other closely related homologs were obtained without finding the other seed members. This search led to the identification of three FGE orthologs in eukaryotes, 12 orthologs in prokaryotes and two paralogs in man and mouse (Table 3).

Table 3: The FGE gene family in eukaryotes and prokaryotes

SEQ ID NOs: NA, AA [GI]	SPECIES	LENGTH [AA]	SUBGROUP
1/3, 2	Homo sapiens	374	E1
49, 50 [22122361]	Mus musculus	372*f*	E1
51, 52 [20130397]	Drosophila melanogaster	336	E1
53, 54 [21289310]	Anopheles gambiae	290	E1
47, 48 [26344956]	Mus musculus	308	E2
45, 46 [24308053]	Homo sapiens	301	E2
55, 56 [21225812]	Streptomyces coelicolor A3(2)	314	P1
57, 58 [25028125]	Corynebacterium efficiens YS-314	334	P1
59, 60 [23108562]	Novosphingobium aromaticivorans	338	P2
61, 62 [13474559]	Mesorhizobium loti	372	P2
63, 64 [22988809]	Burkholderia fungorum	416	P2
65, 66 [16264068]	Sinorhizobium meliloti	303	P2
67, 68 [14518334]	Microscilla sp.	354	P2
69, 70 [26990068]	Pseudomonas putida KT2440	291	P2
71, 72	Ralstonia metallidurans	259	P2

[22975289]			
73, 74 [23132010]	Prochlorococcus marinus	291	P2
75, 76 [16125425]	Caulobacter crescentus CB15	338	P2
77, 78 [15607852]	Mycobacterium tuberculosis Ht37Rv	299	P2

GI- GenBank protein identifier

NA- nucleic acid AA - amino acids,

E1 - eukaryotic orthologs E2 - eukaryotic paralogs

P1 - closely related prokaryotic orthologs P2 - other prokaryotic orthologs

f- protein sequence mispredicted in GenBank

Note that the mouse sequence GI 22122361 is predicted in GenBank to encode a protein of 284 aa, although the cDNA sequence NM 145937 encodes for a protein of 372 residues. This misprediction is based on the omission of the first exon of the murine FGE gene. All sequences found in the NR database are from higher eukaryotes or prokaryotes. FGE-homologs were not detected in archaebacteriae or plants. Searches with even lowered thresholds in the fully sequenced genomes of *C. elegans* and *S. cerevisiae* and the related ORF databases did not reveal any homologs. A search in the eukaryotic sequences of the NT and EST nucleotide databases led to the identification of 8 additional FGE orthologous ESTs with 3'-terminal cDNA sequence fragments showing a high degree of conservation on the protein level which are not listed in the NR database. These sequences do not encompass the full coding part of the mRNAs and are all from higher eukaryotes (Table 4).

Table 4: FGE ortholog EST fragments in eukaryotes

SEQ ID NOs: NA [GB]	SPECIES
80 [CA379852]	Oncorhynchus mykiss
81 [AI721440]	Danio rerio
82 [BJ505402]	Oryzias latipes
83 [BJ054666]	Xenopus laevis
84 [AL892419]	Silurana tropicalis
85 [CA064079]	Salmo salar
86 [BF189614]	Sus scrofa
87 [AV609121]	Bos taurus

GB- GenBank Accession No; NA- nucleic acid

Multiple alignment and construction of a phylogenetic tree (using ClustalW) of the coding sequences from the NR database allowed the definition of four subgroups of homologs: eukaryotic orthologs (human, mouse, mosquito and fruitfly FGE, eukaryotic paralogs (human and mouse FGE paralog), prokaryotic orthologs closely related to FGE (Streptomyces and Corynebacterium and other prokaryotic orthologs (Caulobacter, Pseudomonas, Mycobacterium, Prochlorococcus, Mesorhizobium, Sinorhizobium, Novosphingobium, Ralstonia, Burkholderia, and Microscilla) . The eukaryotic orthologs show an overall identity to human FGE of 87% (mouse), 48% (fruitfly) and 47% (anopheles). While FGE orthologs are found in prokaryotes and higher eukaryotes, they are missing in the completely sequenced genomes of lower eukaryotes phylogenetically situated between S. cerevisiae and D. melanogaster. In addition, FGE homologs are absent in the fully sequenced genomes of E. coli and the pufferfish.

As discussed elsewhere herein, the FGE paralogs found in human and mouse may have a minor *FGly*-generating activity and contribute to the residual activities of sulfatases found in MSD patients.

Subdomains of FGE

The members of the FGE gene family have three highly conserved parts/domains (as described elsewhere herein). In addition to the two non-conserved sequences separating the former, they have non-conserved extensions at the N- and C- terminus. The three conserved parts are considered to represent subdomains of the DUF323 domain because they are spaced by non-conserved parts of varying length. The length of the part spacing subdomains 1 and 2 varies between 22 and 29 residues and that spacing subdomains 2 and 3 between 7 to 38 amino acids. The N- and C-terminal non-conserved parts show an even stronger variation in length (N-terminal: 0-90 AA, Cterminal: 0-28 AA). The sequence for the FGE gene from Ralstonia metallidurans is probably incomplete as it lacks the first subdomain.

To verify the plausibility of defining subdomains of DUF323, we performed a secondary structure prediction of the human FGE protein using Psipred. The hydrophobic ER-signal (residues 1-33) is predicted to contain helix-structures confirming the signal prediction of the von-Heijne algorithm. The N-terminal non-conserved region (aa 34-89) and the spacing region between subdomains 2 and 3 (aa 308-327) contain coiled sections. The region spacing subdomains 1 and 2 contains a coil. The α-helix at aa 65/66 has a low predicition confidence and is probably a prediction artefact. The subdomain boundaries are situated within coils and do not interrupt α-helices or β-strands. The first subdomain is made up of several β-strands and an α-helix, the second subdomain contains two β-strands and four

α-helices. The third subdomain has a α-helix region flanked by a sheet a the beginning and the end of the subdomain. In summary, the secondary structure is in agreement with the proposed subdomain structure as the subdomain boundaries are situated within coils and the subdomains contain structural elements α-helices and β-strands).

It should be noted that none of the subdomains exists as an isolated module in sequences listed in databases. Within each of the four subgroups of the FGE family, the subdomains are highly conserved, with the third subdomain showing the highest homology (Table 5). This subdomain shows also the strongest homology across the subgroups.

Table 5: Homology (% similarity) of the FGE family subdomains

Subfamily	Members	Subdomain 1	2	3
E1	4	79	82	100
E2	2	90	94	100
P1	2	70	79	95
P2	10	59	79	80

E1 - eukaryotic orthologs; E2 - eukaryotic paralogs

P1 - closely related prokaryotic orthologs; P2 - other prokaryotic orthologs

The first subdomain of the FGE-family shows the weakest homology across the subgroups. In the eukaryotic orthologs it carries the N-glycosylation site: at residue Asn 141 in human, at Asn 139 in the mouse and Asn 120 in the fruit fly. In anopheles, no asparagine is found at the residue 130 homologous to D. melanogaster Asn 120. However, a change of two nucleotides would create an N-glycosylation site Asn 130 in anopheles. Therefore, the sequence encompassing residue 130 needs to be resequenced. The second subdomain is rich in tryptophans with 12 Trp in 129 residues of human FGE. Ten of these tryptophans are conserved in the FGE family.

High conservation of subdomain 3: subdomain 3 between eukaryotic orthologs are 100% similar and 90% identical. The importance of the third subdomain for the function of the protein is underlined by the observation that this subdomain is a hot spot for disease causing mutations in MSD patients. Seven of nine mutations identified in six MSD patients described in Example 1 are located in sequences that encode the 40 residues of subdomain 3. The residues contain four cysteines, three of which are conserved among the pro- and eukaryotic orthologs. The two eukaryotic paralogs show the lowest homology to the other members of the FGE-family, e.g. they lack two of the three conserved cysteines of subdomain 3. Features conserved between subdomain 3 sequences of orthologs and paralogs

are the initial RVXXGG(A)S motif (SEQ ID NO:79), a heptamer containing three arginines (residues 19-25 of the subdomain consensus sequence) and the terminal GFR motif. A comparison with the DUF323 domain of the 15 seed sequences that are no close homologs of FGE shows marked sequence differences: the 15 seed sequences have a less conserved first and second subdomain, although the overall subdomain structure is also visible. Subdomain 3, which is strongly conserved in the FGE family, is shorter and has a significantly weaker homology to the eukaryotic subdomain 3 (similarity of about 20%) as compared to the prokaryotic FGE family members (similarity of about 60%). Thus they lack all of the conserved cysteine residues of subdomain 3. The only conserved features are the initial RVXXGG(A)S motif (SEQ ID NO:79) and the terminal GFR motif.

Genomic organisation of the human and murine FGE gene

The human FGE gene is located on chromosome 3p26. It encompasses 105 kb and 9 exons for the translated sequence. The murine FGE gene has a length of 80 Kb and is located on chromosome 6E2. The 9 exons of the murine FGE gene have nearly the same size as the human exons (Figure 3). Major differences between the human and the mouse gene are the lower conservation of the 3'-UTR in exon 9 and the length of exon 9, which is 461 bp longer in the murine gene. Segment 6E2 of mouse chromosome 6 is highly syntenic to the human chromosome segment 3p26. Towards the telomere, both the human and the murine FGE loci are flanked by the genes coding for LMCD1, KIAA0212, ITPR1, AXCAM, and IL5RA. In the centromeric direction, both FGE loci are flanked by the loci of CAV3 and OXTR.

Genomic organisation of the prokaryotic FGE genes

In prokaryotes the sulfatases are classified either as cysteine- or serine-type sulfatases depending on the residue that is converted to *FGly* in their active center (Miech, C., et al., *J Biol Chem.*, 1998, 273:4835-4837; Dierks, T., et al., *J Biol Chem.*, 1998, 273:25560-25564). In Klebsiella pneumoniae, E. coli and Yersinia pestis, the serine-type sulfatases are part of an operon with AtsB, which encodes a cytosolic protein containing iron-sulfur cluster motifs and is critical for the generation of *FGly* from serine residues (Marquordt, C., et al., *J Biol Chem.*, 2003, 278:2212-2218; Szameit, C., et al., *J Biol Chem.*, 1999, 274:15375-15381).

It was therefore of interest to examine whether prokaryotic FGE genes are localized in proximity to cysteine-type sulfatases that are the substrates of FGE. Among the prokaryotic FGE genes shown in Table 3, seven have fully sequenced genomes allowing a neighbourhood analysis of the FGE loci. Indeed, in four of the 7 genomes (C. efficiens: PID 25028125, P. putida: PID 26990068, C. crescentus: PID 16125425 and M. tuberculosis: PID 15607852) a cysteine-type sulfatase is found in direct vicinity of FGE compatible with a cotranscription of

FGE and the sulfatase. In two of them (C. efficiens and P. putida), FGE and the sulfatase have even overlapping ORFs, strongly pointing to their coexpression. Furthermore, the genomic neighbourhood of FGE and sulfatase genes in four prokaryotes provides additional evidence for the assumption that the bacterial FGEs are functional orthologs.

The remaining three organisms do contain cysteine-type sulfatases (S. coelicolor: PID 24413927, M. loti: PID 13476324, S. meliloti: PIDs 16262963, 16263377, 15964702), however, the genes neighbouring FGE in these organisms neither contain a canonical sulfatase signature (Dierks, T., et al., *J Biol Chem.*, 1998, 273:25560-25564) nor a domain that would indicate their function. In these organims the expression of FGE and cysteine-type sulfatases is therefore likely to be regulated *in trans*.

Conclusions

The identification of human FGE whose deficiency causes the autosomal-recessively transmitted lysosomal storage disease Multiple Sulfatase Deficiency, allows the definition of a new gene family which comprises FGE orthologs from prokaryotes and eukaryotes as well as an FGE paralog in mouse and man. FGE is not found in the fully sequenced genomes of E. coli, S. cerevisiae, C. elegans and Fugu rubripes. In addition, there is a phylogenetic gap between prokaryotes and higher eukaryotes with FGE lacking in any species phylogenetically situated between prokaryotes and D. melanogaster. However, some of these lower eukaryotes, e.g. C. elegans, have cysteine-type sulfatase genes. This points to the existence of a second *FGly* generating system acting on cysteine-type sulfatases. This assumption is supported by the observation that E. coli, which lacks FGE, can generate *FGly* in cysteine-type sulfatases (Dierks, T., et al., *J Biol Chem.*, 1998, 273:25560-25564).

Example 3:

FGE expression causes significant increases in sulfatase activity in cell lines that overexpress a sulfatase

We wanted to examine the effects of FGE on cells expressing/overexpressing a sulfatase. To this end, HT-1080 cells expressing human sulfatases Iduronate 2-Sulfatase (I2S) or N-Acetylgalactosamine 6-Sulfatase (GALNS) were transfected in duplicate with either a FGE expression construct, pXMG.1.3 (Table 7 and Fig. 4) or a control plasmid, pXMG.1.2 (FGE in antisense orientation incapable of producing functional FGE, Table 7). Media samples were harvested 24, 48, and 72 hours following a 24 hour post-electroporation medium change. The samples of medium were tested for respective sulfatase activity by activity assay and total sulfatase protein level estimated by ELISA specific for either Iduronate 2-Sulfatase or N-Acetylgalactosamine 6-Sulfatase.

Table 6. Transfected Cell Lines Expressing Sulfatases Used as Substrates for Transfection

Cell Strain	Plasmid	Sulfatase Expressed
36F	pXFM4A.1	N-Acetylgalactosamine 6-Sulfatase
30C6	pXI2S6	Iduronate 2-Sulfatase

Table 7. FGE and Control Plasmids Used to Transfect Iduronate 2-Sulfatase and N-Acetylgalactosamine 6-Sulfatase Expressing HT-1080 Cells

Plasmid	Configuration of Major DNA Sequence Elements*
pXMG.1.3 (FGE expression)	>1.6 kb CMV enhancer/promoter > 1.1 kb FGE cDNA>hGH3' untranslated sequence <amp <DHFR cassette < Cdneo cassette (neomycin phosphotransferase)
pXMG.1.2 (control, FGE reverse orientation)	>1.6 kb CMV enhancer/promoter < 1.1 kb FGE cDNA<hGH3' untranslated sequence <amp <DHFR cassette < Cdneo cassette (neomycin phosphotransferase)

* > denotes orientation 5' to 3'

Experimental Procedures
Materials and Methods

Transfection of HT-1080 cells producing Iduronate 2-Sulfatase and N-Acetylgalactosamine 6-Sulfatase

HT-1080 cells were harvested to obtain 9-12 x 10^6 cells for each electroporation. Two plasmids were transfected in duplicate: one to be tested (FGE) and a control; in this case the control plasmid contained the FGE cDNA cloned in the reverse orientation with respect to the CMV promoter. Cells were centrifuged at approximately 1000 RPM for 5 minutes. Cells were suspended in 1X PBS at 16x10^6 cells/mL. To the bottom of electroporation cuvette, 100 μg of plasmid DNA was added, 750 μL of cell suspension (12x10^6 cells) was added to the DNA solution in the cuvette. The cells and DNA were mixed gently with a plastic transfer pipette, being careful not to create bubbles. The cells were electroporated at 450 V, 250 μF (BioRad Gene Pulser). The time constant was recorded.

The electroporated cells were allowed to sit undisturbed for 10-30 minutes. 1.25 mL of DMEM/10% calf serum was then added to each cuvette, mixed, and all the cells transferred to a fresh T75 flask containing 20 mL DMEM/10. After 24 hours, the flask was re-fed with 20 mL DMEM/10 to remove dead cells. 48-72 hours after transfection, media samples were collected and the cells harvested from duplicate T75 flasks.

Medium Preparation

1L DMEM/10 (contains: 23ml of 2mM L Glutamine,115mL calf serum)

Cells were transfected in media without methotrexate (MTX). 24 hours later cells were re-fed with media containing the appropriate amounts of MTX (36F = 1.0 μM MTX, 30C6 = 0.1M MTX). Medium was harvested and cells collected 24, 48, and 72 hours after re-feed.

Activity Assays

Iduronate 2-Sulfatase (I2S). NAP5 Desalting columns (Amersham Pharmacia Biotech AB, Uppsala, Sweden) were equilibrated with Dialysis Buffer (5 mM sodium acetate, 5 mM tris, pH 7.0). I2S-containing sample was applied to the column and allowed to enter the bed. The sample was eluted in 1 mL of Dialysis Buffer. Desalted samples were further diluted to approximately 100 ng/mL I2S in Reaction Buffer (5 mM sodium acetate, 0.5 mg/L BSA, 0.1 % Triton X-100, pH 4.5). 10 μL of each I2S sample was added to the top row of a 96-well Fluormetric Plate (Perkin Elmer, Norwalk, CT) and pre-incubated for 15 minutes at 37°C. Substrate was prepared by dissolving 4-methyl-umbelliferyl sulfate (Fluka, Buchs, Switzerland) in Substrate Buffer (5 mM sodium acetate, 0.5 mg/mL BSA, pH 4.5) at a final concentration of 1.5 mg/mL. 100 μL of Substrate was added to each well containing I2S sample and the plate was incubated for 1 hour at 37°C in the dark. After the incubation 190 μL of Stop Buffer (332.5 mM glycine, 207.5 mM sodium carbonate, pH 10.7) was added to each well containing sample. Stock 4-methylumbelliferone (4-MUF, Sigma, St. Louis, MO) was prepared as the product standard in reagent grade water to a final concentration of 1 μM. 150 μL of 1 μM 4-MUF Stock and 150 μL Stop Buffer were added to one top row well in the plate. 150 μL of Stop Buffer was added to every remaining well in the 96-well plate. Two fold serial dilutions were made from the top row of each column down to the last row of the plate. The plate was read on a Fusion Universal Microplate Analyzer (Packard, Meriden, CT) with an excitation filter wavelength of 330 nm and an emission filter wavelength of 440 nm. A standard curve of μmoles of 4-MUF stock versus fluorescence was generated, and unknown samples have their fluorescence extrapolated from this curve. Results are reported as Units/mL where one Unit of activity was equal to 1 μmole of 4-MUF produced per minute at 37°C.

N-Acetylgalactosamine 6-Sulfatase (GALNS). The GALNS activity assay makes use of the fluorescent substrate, 4-methylumbelliferyl-β-D-galactopyranoside-6-sulfate (Toronto Research Chemicals Inc., Catalogue No. M33448). The assay was comprised of two-steps. At the first step, 75 μL of the 1.3 mM substrate prepared in reaction buffer (0.1M sodium acetate, 0.1M sodium chloride, pH 4.3) was incubated for 4 hours at 37°C with 10 μL of

media/protein sample or its corresponding dilutions. The reaction was stopped by the addition of 5 μL of 2M monobasic sodium phosphate to inhibit the GALNS activity. Following the addition of approximately 500 U of β-galactosidase from Aspergillus oryzae (Sigma, Catalogue No. G5160), the reaction mixture was incubated at 37°C for an additional hour to release the fluorescent moiety of the substrate. The second reaction was stopped by the addition of 910 μL of stop solution (1% glycine, 1% sodium carbonate, pH 10.7). The fluorescence of the resultant mixture was measured by using a measurement wavelength of 359 nm and a reference wavelength of 445 nm with 4-methylumbelliferone (sodium salt from Sigma, Catalogue No. M1508) serving as a reference standard. One unit of the activity corresponds to nmoles of released 4-methylumbelliferone per hour.

Immunoassays (ELISA)

Iduronate 2-Sulfatase (I2S). A 96-well flat bottom plate was coated with a mouse monoclonal anti-I2S antibody diluted to 10 μg/mL in 50 nM sodium bicarbonate pH 9.6 for 1 hour at 37°C. The mouse monoclonal anti-I2S antibody was developed under contract by Maine Biotechnology Services, Inc. (Portland, ME) to a purified, recombinantly-produced, full-length, human I2S polypeptide using standard hybridoma-producing technology. The plate was washed 3 times with 1X PBS containing 0.1% Tween-20 and blocked for 1 hour with 2% BSA in wash buffer at 37°C. Wash buffer with 2% BSA was used to dilute samples and standards. I2S standard was diluted and used from 100 ng/mL to 1.56 ng/mL. After removal of the blocking buffer, samples and standards were applied to the plate and incubated for 1 hour at 37°C. Detecting antibody, horseradish peroxidase-conjugated mouse anti-I2S antibody, was diluted to 0.15 μg/mL in wash buffer with 2% BSA. The plate was washed 3 times, detecting antibody added to the plate, and it was incubated for 30 minutes at 37°C. To develop the plate, TMB substrate (Bio-Rad, Hercules, CA) was prepared. The plate was washed 3 times, 100 μL of substrate was added to each well and it was incubated for 15 minutes at 37°C. The reaction was stopped with 2 N sulfuric acid (100 μL/well) and the plate was read on a microtiter plate reader at 450 nm, using 655 nm as the reference wavelength.

N-Acetylgalactosamine 6-Sulfatase (GALNS). Two mouse monoclonal anti-GALNS antibodies provided the basis of the GALNS ELISA. The mouse monoclonal anti- GALNS antibodies were also developed under contract by Maine Biotechnology Services, Inc. (Portland, ME) to a purified, recombinantly-produced, full-length, human GALNS polypeptide using standard hybridoma-producing technology. The first antibody, for capture of GALNS was used to coat a F96 MaxiSorp Nunc-Immuno Plate (Nalge Nunc, Catalogue No. 442404) in a coating buffer (50 mM sodium bicarbonate, pH 9.6). After incubation for

one hour at 37°C and washing with a wash buffer, the plate was blocked with blocking buffer (PBS, 0.05% Tween-20, 2% BSA) for one hour at 37°C. Experimental and control samples along with GALNS standards were then loaded onto the plate and further incubated for one hour at 37°C. After washing with a wash buffer, the second, detection antibody conjugated to HRP was applied in blocking buffer followed by 30 minute incubation at 37°C. After washing the plate again, the Bio-Rad TMB substrate reagent was added and incubated for 15 minutes. 2N sulfuric acid was then added to stop the reaction and results were scored spectrophotometrically by using a Molecular Device plate reader at 450 nm wavelength.

Discussion

Effect of FGE on Sulfatase Activity

GALNS. An approximately 50-fold increase in total GALNS activity was observed over the control levels (Figure 5). This level of increased activity was observed with all three medium sampling time points. Moreover, the GALNS activity was accumulated linearly over time with a four-fold increase between 24 and 48 hours and a two-fold increase between the 48 hour and 72 hour timepoints.

I2S. Although of smaller absolute magnitude, a similar effect was observed for total I2S activity where an approximately 5-fold increase in total I2S activity was observed over the control levels. This level of increased activity was sustained for the duration of the experiment. I2S activity accumulated in the medium linearly over time, similar to the results seen with GALNS (2.3-fold between 24 and 48 hours, and 1.8-fold between 48 and 72 hours).

Effect of FGE on Sulfatase *Specific* Activity

GALNS. Expression of FGE in 36F cells enhanced apparent *specific* activity of GALNS (ratio of enzyme activity to total enzyme estimated by ELISA) by 40-60 fold over the control levels (Figure 6). The increase in specific activity was sustained over the three time points in the study and appeared to increase over the three days of post-transfection accumulation.

I2S. A similar effect was seen with I2S, where a 6-7-fold increase in specific activity (3-5 U/mg) was observed over the control values (0.5-0.7 U/mg).

The ELISA values for both GALNS (Figure 7) and I2S were not significantly affected by transfection of FGE. This indicates that expression of FGE does not impair translational and secretory pathways involved in sulfatase production.

In sum, all of these results for both sulfatases indicate that FGE expression dramatically increases sulfatase *specific* activity in cell lines that overexpress GALNS and I2S.

Co-expression of FGE (SUMF1) and other sulfatase genes

To test the effect of FGE (SUMF1) on additional sulfatase activities in normal cells we overexpressed ARSA (SEQ ID NO:14), ARSC (SEQ ID NO:18) and ARSE (SEQ ID NO:22) cDNAs in various cell lines with and without co-transfection of the FGE (SUMF1) cDNA and measured sulfatase activities. Overexpression of sulfatase cDNAs in Cos-7 cells resulted in a moderate increase of sulfatase activity, while a striking synergistic increase (20 to 50 fold) was observed when both a sulfatase gene and the FGE (SUMF1) gene were co-expressed. A similar, albeit lower, effect was observed in three additional cell lines, HepG2, LE293, and U2OS. Simultaneous overexpression of multiple sulfatase cDNAs resulted in a lower increase of each specific sulfatase activity as compared to overexpression of a single sulfatase, indicating the presence of competition of the different sulfatases for the modification machinery.

To test for functional conservation of the FGE (SUMF1) gene during evolution we overexpressed ARSA, ARSC and ARSE cDNAs in various cell lines with and without co-transfection of the MSD cDNA and measured sulfatase activities. Both the murine and the Drosophila FGE (SUMF1) genes were active on all three human sulfatases, with the Drosophila FGE (SUMF1) being less efficient. These data demonstrate a high degree of functional conservation of FGE (SUMF1) during evolution implicating significant biological importance to cellular function and survival. A similar and consistent, albeit much weaker, effect was observed by using the FGE2 (SUMF2) gene, suggesting that the protein encoded by this gene also has a sulfatase modifying activity. These data demonstrate that the amount of the FGE (SUMF1)-encoded protein is a limiting factor for sulfatase activities, a finding with important implications for the large scale production of active sulfatases to be utilized in enzyme replacement therapy.

Example 4:
Identification of the gene mutated in MSD by means of functional complementation using microcell mediated chromosome transfer.

In a separate experiment using microcell mediated chromosome transfer by means of functional complementation we confirmed that the gene mutated in MSD is FGE. Our findings provide further insight into a novel biological mechanism affecting an entire family of proteins in distantly related organisms. In addition to identifying the molecular basis of a rare genetic disease, our data further confirms a powerful enhancing effect of the *FGE* gene product on the activity of sulfatases. The latter finding has direct clinical implications for the therapy of at least eight human diseases caused by sulfatase deficiencies.

The gene for MSD maps to chromosome 3p26

To identify the chromosomal location of the gene mutated in MSD we attempted to rescue the deficient sulfatase enzymes by functional complementation via microcell mediated chromosome transfer. A panel of human/mouse hybrid cell lines, containing individual normal human chromosomes tagged with the dominant selectable marker HyTK, was used as the source of donor human chromosomes and fused to an immortalized cell line from a patient with MSD. All 22 human autosomes were transferred one by one to the patient cell line and hybrids were selected in hygromycin. Approximately 25 surviving colonies were picked in each of the 22 transfer experiments. These were grown separately and harvested for subsequent enzymatic testing. ArylsulfataseA (ARSA) (SEQ ID NO:15), ArylsulfataseB (ARSB) (SEQ ID NO:17), and ArylsulfataseC (ARSC) (SEQ ID NO:19) activities were tested for each of the approximately 440 clones (20 x 22). This analysis clearly indicated that sulfatase activities of several clones deriving from the chromosome 3 transfer was significantly higher compared to that of all the other clones. A striking variability was observed when analyzing the activities of each individual clone from the chromosome 3 transfer. To verify whether each clone had an intact human chromosome 3 from the donor cell line, we used a panel of 23 chromosome 3 polymorphic genetic markers, evenly distributed along the length of the chromosome and previously selected on the basis of having different alleles between the donor and the patient cell lines. This allowed us to examine for the presence of the donor chromosome and to identify possible loss of specific regions due to incidental chromosomal breakage. Each clone having high enzymatic activity retained the entire chromosome 3 from the donor cell line, whereas clones with low activities appeared to have lost the entire chromosome on the basis of the absence of chromosome 3 alleles from the donor cell line. The latter clones probably retained a small region of the donor chromosome containing the selectable marker gene that enabled them to survive in hygromycin containing medium. These data indicate that a normal human chromosome 3 was able to complement the defect observed in the MSD patient cell line.

To determine the specific chromosomal region containing the gene responsible for the complementing activity we used Neo-tagged chromosome 3 hybrids which were found to have lost various portions of the chromosome. In addition, we performed irradiated microcell-mediated chromosome transfer of HyTK-tagged human chromosomes 3. One hundred and fifteen chromosome 3 irradiated hybrids were tested for sulfatase activities and genotyped using a panel of 31 polymorphic microsatellite markers spanning the entire chromosome. All clones displaying high enzymatic activities appeared to have retained

chromosome 3p26. A higher resolution analysis using additional markers from this region mapped the putative location for the complementing gene between markers *D3S3630* and *D3S2397*.

Identification of the gene mutated in MSD

We investigated genes from the 3p26 genomic region for mutations in MSD patients. Each exon including splice junctions were PCR-amplified and analyzed by direct sequencing. Mutation analysis was performed on twelve unrelated affected individuals; five previously described MSD patients and seven unpublished cases. Several mutations were identified from our MSD cohort in the expressed sequence tag (EST) AK075459 (SEQ ID NOs:4,5), corresponding to a gene of unknown function, strongly suggesting that this was the gene involved in MSD. Each mutation was found to be absent in 100 control individuals, thus excluding the presence of a sequence polymorphism. Additional confirmatory mutation analysis was performed on reverse transcribed patients' RNAs, particularly in those cases in which genomic DNA analysis revealed the presence of a mutation in or near a splice site, possibly affecting splicing. Frameshift, nonsense, splicing, and missense mutations were also identified, suggesting that the disease is caused by a loss of function mechanism, as anticipated for a recessive disorder. This is also consistent with the observation that almost all missense mutations affect amino acids that are highly conserved throughout evolution (see below).

Table 8: Additional MSD Mutations identified

Case	reference	phenotype	exon	nucleotide change	amino acid change
1. BA426	Conary et al, 1988	moderate	3	463T>C	S155P
			3	463T>C	S155P
2. BA428	Burch et al, 1986	severe neonatal	5	661delG	frameshift
3. BA431	Zenger et al, 1989	moderate	1	2T>G	M1R
			2	276delC	frameshift
4. BA799	Burk et al, 1981	mild-moderate	3	463T>C	S155P
			3	463T>C	S155P
5. BA806	unpublished	severe neonatal	9	1045T>C	R349W
6. BA807	Schmidt et al, 1995	unknown	3	c519+4delGTAA	ex 3 skipping
			9	1076C>A	S359X
7. BA809	Couchot et al, 1974	mild-moderate	1	1A>G	M1V
			9	1042G>C	A348P
8. BA810	unpublished	severe	8	1006T>C	C336R

			9	1046G>A	R349Q
9. BA811	unpublished	severe neonatal	3 8	c519+4delGTAA 979C>T	ex 3 skipping R327X
10. BA815	unpublished	moderate	5 6	c.603-6delC 836C>T	ex 6 skipping A279V
11. BA919	unpublished	mild-moderate	9 9	1033C>T 1033C>T	R345C R345C
12. BA920	unpublished	moderate	5 9	653G>A 1033C>T	C218Y R345C

Mutations were identified in each MSD patient tested, thus excluding locus heterogeneity. No obvious correlation was observed between the types of mutations identified and the severity of the phenotype reported in the patients, suggesting that clinical variability is not caused by allelic heterogeneity. In three instances different patients (case 1 and 4, case 6 and 9, and case 11 and 12 in Table 6) were found to carry the same mutation. Two of these patients (case 11 and 12) originate from the same town in Sicily, suggesting the presence of a founder effect that was indeed confirmed by haplotype analysis. Surprisingly, most patients were found to be compound heterozygotes, carrying different allelic mutations, while only a few were homozygous. Albeit consistent with the absence of consanguinity reported by the parents, this was a somehow unexpected finding for a very rare recessive disorder such as MSD.

The FGE gene and protein

The consensus cDNA sequence of the human FGE (also used interchangeably herein as SUMF1) cDNA (SEQ ID NO:1) was assembled from several expressed sequence tag (EST) clones and partly from the corresponding genomic sequence. The gene contains nine exons and spans approximately 105 kb (see Example 1). Sequence comparison also identified the presence of a FGE gene paralog located on human chromosome 7 that we designated FGE2 (also used interchangeably herein as SUMF2) (SEQ ID NOs: 45, 46).

Functional complementation of sulfatase deficiencies

Fibroblasts from two patients (case 1 and 12 in Table 8) with MSD in whom we identified mutations of the FGE (SUMF1) gene (cell lines BA426 and BA920) were infected with HSV viruses containing the wild type and two mutated forms of the FGE (SUMF1) cDNA (R327X and Δex3). ARSA, ARSB, and ARSC activities were tested 72 hrs after infection. Expression of the wild type FGE (SUMF1) cDNA resulted in functional complementation of all three activities, while mutant FGE (SUMF1) cDNAs did not (Table 9). These data provide conclusive evidence for the identity of FGE (SUMF1) as the MSD

gene and they prove the functional relevance of the mutations found in patients. The disease-associated mutations result in sulfatase deficiency, thus demonstrating that FGE (SUMF1) is an essential factor for sulfatase activity.

Table 9: Functional complementation of sulfatase deficiencies

Recipient MSD cell line	construct	ARSA[(1)]	ARSB[(1)]	ARSC[(1)]
BA426	HSV amplicon	24.0	22.5	0.15
	SUMF1-Δex3	42.0	23.8	0.29
	SUMF1-R327X	33.6	24.2	0.16
	SUMF1	119.5 (4.9 x)	37.8 (1.7 x)	0.62(4.1 x)
BA920	HSV amplicon	16.6	11.3	0.15
	SUMF1-Δex3	17.2	14.4	0.07
	SUMF1-R327X	36.0	13.5	0.13
	SUMF1	66.5 (4.0 x)	21.6 (1.9 x)	0.42(2.8 x)
Control range		**123.7-394.6**	**50.6-60.7**	**1.80-1.58**

[(1)]All enzymatic activities are expressed as nmoles 4-methylumbelliferone liberated · mg protein $^{-1}$ · 3 hrs. MSD cell lines BA426 and BA920 were infected with the HSV amplicon alone, and with constructs carrying either mutant or wild-type SUMF1 cDNAs. The increase of single arylsulfatase activities in fibroblasts infected with the wild-type SUMF1 gene, as compared to those of cells infected with the vector alone, is indicated in parentheses. Activities measured in uninfected control fibroblasts are indicated.

Molecular basis of MSD

Based on the hypothesis that the disease gene should be able to complement the enzymatic deficiency in a patient cell line, we performed microcell-mediated chromosome transfer to an immortalized cell line from a patient with MSD. This technique has been successfully used for the identification of genes whose predicted function could be assessed in cell lines (e.g. by measuring enzymatic activity or by detecting morphologic features). To address the problem of stochastic variability of enzyme activity we measured the activities of three different sulfatases (ARSA, ARSB and ARSC) in the complementation assay. The results of chromosome transfer clearly indicated mapping of the complementing gene to chromosome 3. Subregional mapping was achieved by generating a radiation hybrid panel for chromosome 3. Individual hybrid clones were characterized both at the genomic level, by typing 31 microsatellite markers displaying different alleles between donor and recipient cell lines, and at the functional level by testing sulfatase activities. The analysis of 130 such hybrids resulted in the mapping of the complementing region to chromosome 3p26.

Once the critical genomic region was defined, the FGE (SUMF1) gene was also identified by mutation analysis in patients' DNA. Mutations were found in all patients tested, proving that a single gene is involved in MSD. The mutations found were of different types, the majority (e.g. splice site, start site, nonsense, frameshift) putatively result in a loss function of the encoded protein, as expected for a recessive disease. Most missense mutations affect codons corresponding to amino acids that have been highly conserved during evolution, suggesting that also these mutations cause a loss of function. No correlations could be drawn between the type of mutation and the severity of the phenotype, indicating that the latter is due to unrelated factors. Unexpectedly for a rare genetic disease, many patients were found to be compound heterozygotes, carrying two different mutations. However, a founder effect was identified for one mutation originating from a small town in Sicily.

FGE (SUMF1) gene function

The identity of the FGE (SUMF1) gene as the "complementing factor" was demonstrated definitively by rescuing the enzymatic deficiency of four different sulfatases upon expression of exogenous FGE (SUMF1) cDNA, inserted into a viral vector, in two different patient cell lines. In each case a consistent, albeit partial, restoration of all sulfatase activities tested was observed, as compared to control patient cell lines transfected with empty vectors. On average, the increase of enzyme activities ranged between 1.7 to 4.9 fold and reached approximately half of the levels observed in normal cell lines. Enzyme activity correlates with the number of virus particles used in each experiment and with the efficiency of the infection as tested by marker protein (GFP) analysis. In the same experiments vectors containing FGE (SUMF1) cDNAs carrying two of the mutations found in the patients, R327X and Δex3, were used and no significant increase of enzyme activity was observed, thus demonstrating the functional relevance of these mutations.

As mentioned elsewhere herein, Schmidt et al. first discovered that sulfatases undergo a post-translational modification of a highly conserved cysteine, that is found at the active site of most sulfatases, to Cα-formylglycine. They also showed that this modification was defective in MSD (Schmidt, B., et al., *Cell*, 1995, 82:271-278). Our mutational and functional data provide strong evidence that FGE (SUMF1) is responsible for this modification.

The FGE (SUMF1) gene shows an extremely high degree of sequence conservation across all distantly related species analyzed, from bacteria to man. We provide evidence that that the Drosophila homologue of the human FGE (SUMF1) gene is able to activate overexpressed human sulfatases, proving that the observed high level of sequence similarity

of the FGE (SUMF1) genes of distantly related species correlates with a striking functional conservation. A notable exception is yeast, which appears to lack the FGE (SUMF1) gene as well as any sulfatase encoding genes, indicating that sulfatase function is not required by this organism and suggesting the presence of a reciprocal influence on the evolution of FGE (SUMF1) and sulfatase genes.

Interestingly, there are two homologous genes, FGE (SUMF1) and FGE2 (SUMF2), in the genomes of all vertebrates analyzed, including humans. As evident from the phylogenetic tree, the FGE2 (SUMF2) gene appears to have evolved independently from the FGE (SUMF1) gene. In our assays the FGE2 (SUMF2) gene is also able to activate sulfatases, however it does it in a much less efficient manner compared to the FGE (SUMF1) gene. This may account for the residual sulfatase activity found in MSD patients and suggests that a complete sulfatase deficiency would be lethal. At the moment we cannot rule out the possibility that the FGE2 (SUMF2) gene has an additional, yet unknown, function.

Impact on the therapy of diseases due to sulfatase deficiencies

A strong increase, up to 50 fold, of sulfatase activities was observed in cells overexpressing FGE (SUMF1) cDNA together with either ARSA, ARSC, or ARSE cDNAs, compared to cells overexpressing single sulfatases alone. In all cell lines a significant synergic effect was found, indicating that FGE (SUMF1) is a limiting factor for sulfatase activity. However, variability was observed among different sulfatases, possibly due to different affinity of the FGE (SUMF1)-encoded protein with the various sulfatases. Variability was also observed between different cell lines which may have different levels of endogenous formylglycine generating enzyme. Consistent with these observations, we found that the expression of the MSD gene varies among different tissues, with significantly high levels in kidney and liver. This may have important implications as tissues with low FGE (SUMF1) gene expression levels may be less capable of effectively modifying exogenously delivered sulfatase proteins (see below). Together these data suggest that the function of the FGE (SUMF1) gene has evolved to achieve a dual regulatory system, with each sulfatase being controlled by both an individual mechanism, responsible for the mRNA levels of each structural sulfatase gene, and a common mechanism shared by all sulfatases. In addition, FGE2 (SUMF2) provides partial redundancy for sulfatase modification.

These data have profound implications for the mass production of active sulfatases to be utilized in enzyme replacement therapy. Enzyme replacement studies have been reported on animal models of sulfatase deficiencies, such as a feline model of mucopolysaccharidosis VI, and proved to be effective in preventing and curing several symptoms. Therapeutic trials

in humans are currently being performed for two congenital disorders due to sulfatase deficiencies, MPSII (Hunter syndrome) and MPSVI (Maroteaux-Lamy syndrome) and will soon be extended to a large number of patients.

Example 5:

Enzyme Replacement Therapy with FGE-activated GALNS for Morquio Disease MPS IVA

The primary cause of skeletal pathology in Morquio patients is keratan sulfate (KS) accumulation in epiphyseal disk (growth plate) chondrocytes due to deficiency of the lysosomal sulfatase, GALNS. The primary objective of *in vivo* research studies was to determine whether intravenously (IV) administered *FGE-activated* GALNS was able to penetrate chondrocytes of the growth plate as well as other appropriate cell types in normal mice. Notwithstanding a general lack of skeletal abnormalities, a GALNS deficient mouse model (Morquio Knock-In -MKI, S. Tomatsu, St. Louis University, MO) was also used to demonstrate *in vivo* biochemical activity of repeatedly administered *FGE-activated* GALNS. The lack of skeletal pathology in mouse models reflects the fact that skeletal KS is either greatly reduced or absent in rodents (Venn G, & Mason RM., *Biochem J.*, 1985, 228:443-450). These mice did, however, demonstrate detectable accumulation of GAG and other cellular abnormalities in various organs and tissues. Therefore, the overall objective of the studies was to demonstrate that *FGE-activated* GALNS penetrates into the growth plate (biodistribution study) and show functional GALNS enzyme activity directed towards removal of accumulated GAG in affected tissues (pharmacodynamic study).

The results of these studies demonstrated that IV injected *FGE-activated* GALNS was internalized by chondrocytes of the growth plate, albeit at relatively low levels compared to other tissues. In addition, *FGE-activated* GALNS injection over the course of 16 weeks in MKI mice effectively cleared accumulated GAG and reduced lysosomal biomarker staining in all soft tissues examined. In sum, the experiments successfully demonstrated GALNS delivery to growth plate chondrocytes and demonstrated biochemical activity in terms of GAG clearance in multiple tissues.

Biodistribution Study

Four-week-old ICR (normal) mice were given a single IV injection of 5 mg/kg *FGE-activated* GALNS. Liver, femur (bone), heart, kidney and spleen were collected two hours after injection and prepared for histological examination. A monoclonal anti-human GALNS antibody was used to detect the presence of injected GALNS in the various tissues. GALNS was detected in all tissues examined as compared to the vehicle controls. Moreover, GALNS was readily observed in all tissues examined using a horseradish-peroxidase reporter system,

with the exception of bone. Demonstration of GALNS uptake in the growth plate required the use of a more sensitive fluorescein-isothiocyanate (FITC) reporter system and indicates that although GALNS penetrates the growth plate, it is less readily available to growth plate chondrocytes than to cells of soft tissues. Notwithstanding the requirement of a more sensitive fluorescent detection method, GALNS delivery to bone growth plate chondrocytes was observed in all growth plate sections examined as compared to the vehicle controls.

Pharmacodynamic Study in MKI Mice

Four-week-old MKI or wild-type mice were given weekly IV injections (n=8 per group) through 20 weeks of age. Each weekly injection consisted of either 2 mg/kg *FGE-activated* GALNS or vehicle control (no injection for wild-type mice). All mice were sacrificed for histological examination at 20 weeks of age and stained using the following methods: hematoxylin and eosin for cellular morphology, alcian blue for detection of GAGs.

Clearance of accumulated GAG was demonstrated by reduced or absent alcian blue staining in all soft tissues examined (liver, heart, kidney and spleen). This was observed only in the GALNS injected mice. Although the growth plate in the MKI mice functioned normally as evidenced by normal skeletal morphology, there were more subtle cellular abnormalities observed (including vacuolization of chondrocytes without apparent pathological effect). The vacuolized chondrocytes of the hypertrophic and proliferating zones of the growth plate were unaffected by GALNS administration. This was in contrast to the chondrocytes in the calcification zone of the growth plate where a reduction of vacuolization was observed in GALNS injected mice. The vacuolization of chondrocytes and accumulation of presumed non-KS GAG in the growth plate in MKI mice was, in general, surprising and unexpected due to the known lack of KS in the growth plate of mice. These particular observations likely reflect the fact that, in the knock-in mice, high levels of mutant GALNS are present (as opposed to knock-out mice where there is no residual mutant GALNS, no growth plate chondrocyte vacuolization and no GAG accumulation- Tomatsu S. et al., *Human Molecular Genetics*, 2003, 12:3349-3358). The vacuolization phenomenon in the growth plate may be indicative of a secondary effect on a subset of cells expressing mutant GALNS. Nonetheless, enzyme injection over the course of 16 weeks demonstrated strong evidence of multiple tissue *FGE-activated* GALNS delivery and *in vivo* enzymatic activity.

Detailed Description of the Drawings

Fig. 1: MALDI-TOF mass spectra of P23 after incubation in the absence (A) or presence (B) of a soluble extract from bovine testis microsomes. 6 pmol of P23 were incubated under standard conditions for 10 min at 37°C in the absence or presence of 1 μl

microsomal extract. The samples were prepared for MALDI-TOF mass spectrometry as described in Experimental Procedures. The monoisotopic masses MH^+ of P23 (2526.28) and its FGly derivative (2508.29) are indicated.

Fig. 2: Phylogenetic tree derived from an alignment of human FGE and 21 proteins of the PFAM-DUF323 seed. The numbers at the branches indicate phylogenetic distance. The proteins are designated by their TrEMBL ID number and the species name. hFGE - human FGE. Upper right: scale of the phylogenetic distances. A asterisk indicates that the gene has been further investigated. The top seven genes are part of the FGE gene family.

Fig. 3: Organisation of the human and murine FGE gene locus. Exons are shown to scale as dark boxes (human locus) and bright boxes (murine locus). The bar in the lower right corner shows the scale. The lines between the exons show the introns (not to scale). The numbers above the intron lines indicate the size of the introns in kilobases.

Fig. 4: Diagram showing a map of FGE Expression Plasmid pXMG.1.3

Fig. 5: Bar graph depicting N-Acetylgalactosamine 6-Sulfatase Activity in 36F Cells Transiently Transfected with FGE Expression Plasmid. Cells were transfected with either a control plasmid, pXMG.1.2, with the FGE cDNA in the reverse oreintation, or a FGE expression plasmid, pXMG.1.3 in media without methotrexate (MTX). 24 hours later cells were re-fed with media containing 1.0 μM MTX. Medium was harvested and cells collected 24, 48, and 72 hours after re-feed. N-Acetylgalactosamine 6-Sulfatase activity was determined by activity assay. Each value shown is the average of two separate transfections with standard deviations indicated by error bars.

Fig. 6: Bar graph depicting N-Acetylgalactosamine 6-Sulfatase *Specific* Activity in 36F Cells Transiently Transfected with FGE Expression Plasmid. Cells were transfected with either a control plasmid, pXMG.1.2, with the FGE cDNA in the reverse oreintation, or a FGE expression plasmid, pXMG.1.3 in media without methotrexate (MTX). 24 hours later cells were re-fed with media containing 1.0 μM MTX. Medium was harvested and cells collected 24, 48, and 72 hours after re-feed. N-Acetylgalactosamine 6-Sulfatase specific activity was determined by activity assay and ELISA and is represented as a ratio of N-Acetylgalactosamine 6-Sulfatase activity per mg of ELISA-reactive N-Acetylgalactosamine 6-Sulfatase. Each value shown is the average of two separate transfections.

Fig. 7: Bar graph depicting N-Acetylgalactosamine 6-Sulfatase Production in 36F Cells Transiently Transfected with FGE Expression Plasmid. Cells were transfected with either a control plasmid, pXMG.1.2, with the FGE cDNA in the reverse oreintation, or a FGE expression plasmid, pXMG.1.3 in media without methotrexate (MTX). 24 hours later cells

were re-fed with media containing 1.0 μM MTX. Medium was harvested and cells collected 24, 48, and 72 hours after re-feed. N-Acetylgalactosamine 6-Sulfatase total protein was determined by ELISA. Each value shown is the average of two separate transfections with standard deviations indicated by error bars.

Fig. 8: Graph depicting Iduronate 2-Sulfatase Activity in 30C6 Cells Transiently Transfected with FGE Expression Plasmid. Cells were transfected with either a control plasmid, pXMG.1.2, with the FGE cDNA in the reverse oreintation, or a FGE expression plasmid, pXMG.1.3 in media without methotrexate (MTX). 24 hours later cells were re-fed with media containing 0.1μM MTX. Medium was harvested and cells collected 24, 48, and 72 hours after re-feed. Iduronate 2-Sulfatase activity was determined by activity assay. Each value shown is the average of two separate transfections.

Fig. 9: Depicts a kit embodying features of the present invention.

All references disclosed herein are incorporated by reference in their entirety. What is claimed is presented below and is followed by a Sequence Listing.

We claim:

Claims

1. An isolated nucleic acid molecule selected from the group consisting of:

(a) nucleic acid molecules which hybridize under stringent conditions to a molecule consisting of a nucleotide sequence set forth as SEQ ID NO:1 and which code for a polypeptide having C_α-formylglycine generating activity (FGE),

(b) nucleic acid molecules that differ from the nucleic acid molecules of (a) in codon sequence due to the degeneracy of the genetic code, and

(c) complements of (a) or (b).

2. The isolated nucleic acid molecule of claim 1, wherein the isolated nucleic acid molecule comprises the nucleotide sequence set forth as SEQ ID NO:1.

3. The isolated nucleic acid molecule of claim 1, wherein the isolated nucleic acid molecule consists of the nucleotide sequence set forth as SEQ ID NO:3 or a fragment thereof.

4. An isolated nucleic acid molecule selected from the group consisting of

(a) unique fragments of a nucleotide sequence set forth as SEQ ID NO:1, and

(b) complements of (a).

5. The isolated nucleic acid molecule of claim 4, wherein the unique fragment has a size selected from the group consisting of at least: 8 nucleotides, 10 nucleotides, 12 nucleotides, 14 nucleotides, 16 nucleotides, 18 nucleotides, 20, nucleotides, 22 nucleotides, 24 nucleotides, 26 nucleotides, 28 nucleotides, 30 nucleotides, 50 nucleotides, 75 nucleotides, 100 nucleotides, and 200 nucleotides.

6. The isolated nucleic acid molecule of claim 4, wherein the molecule encodes a polypeptide which is immunogenic.

7. An expression vector comprising the isolated nucleic acid molecule of claim 1, 2, 3, 4, 5, or 6, operably linked to a promoter.

8. An expression vector comprising the isolated nucleic acid molecule of claim 4 operably linked to a promoter.

9. A host cell transformed or transfected with the expression vector of claim 7.

10. A host cell transformed or transfected with the expression vector of claim 8.

11. An isolated polypeptide encoded by a nucleic acid molecule of claim 1, 2, 3, or 4, wherein the polypeptide, or fragment of the polypeptide, has C_α-formylglycine generating activity.

12. The isolated polypeptide of claim 11, wherein the polypeptide is encoded by the nucleic acid molecule of claim 2.

13. The isolated polypeptide of claim 12, wherein the polypeptide comprises a polypeptide having the sequence of amino acids 1-374 of SEQ ID NO:2.

14. An isolated polypeptide encoded by a nucleic acid molecule of claim 1, 2, 3, or 4, wherein the polypeptide, or fragment of the polypeptide, is immunogenic.

15. The isolated polypeptide of claim 14, wherein the fragment of the polypeptide, or portion of the fragment, binds to a human antibody.

16. An isolated binding polypeptide which binds selectively a polypeptide encoded by an isolated nucleic acid molecule of claim 1, 2, 3, or 4.

17. The isolated binding polypeptide of claim 16, wherein the isolated binding polypeptide binds to a polypeptide having the sequence of amino acids of SEQ ID NO:2.

18. The isolated binding polypeptide of claim 17, wherein the isolated binding polypeptide is an antibody or an antibody fragment selected from the group consisting of a Fab fragment, a $F(ab)_2$ fragment or a fragment including a CDR3 region.

19. A family of isolated polypeptides having C_α-formylglycine generating activity, each of said polypeptides comprising from amino terminus to carboxyl terminus:

(a) an amino-terminal subdomain 1;

(b) a subdomain 2 containing from 120 to 140 amino acids comprising at least 8 Tryptophans;

(c) a carboxy-terminal subdomain 3 containing from 35 to 45 amino acids;

wherein subdomain 2 has at least about 50% homology to subdomain 2 of a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78; and

wherein subdomain 3 has at least about 75% homology and a length approximately equal to subdomain 3 of a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

20. The polypeptides of claim 19, wherein subdomain 3 of each of said polypeptides has at least between about 80% and about 100% homology to subdomain 3 of a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

21. A method for determining the level of FGE expression in a subject, comprising measuring expression of FGE in a test sample from the subject to determine the level of FGE expression in the subject.

22. The method of claim 21, wherein the measured FGE expression in the test sample is compared to FGE expression in a control containing a known level of expression.

23. The method of claim 21, wherein the expression of FGE is FGE mRNA expression.

24. The method of claim 21, wherein the expression of FGE is FGE polypeptide expression.

25. The method of claim 21, wherein the test sample is tissue.

26. The method of claim 21, wherein the test sample is a biological fluid.

27. The method of claim 23, wherein FGE mRNA expression is measured using PCR.

28. The method of claim 23, wherein FGE mRNA expression is measured using Northern blotting.

29. The method of claim 24, wherein FGE polypeptide expression is measured using monoclonal antibodies to FGE.

30. The method of claim 24, wherein FGE polypeptide expression is measured using polyclonal antisera to FGE.

31. The method of claim 24, wherein expression of FGE is measured using C_α-formylglycine generating activity.

32. A method for identifying an agent useful in modulating C_α-formylglycine generating activity, comprising:

(a) contacting a molecule having C_α-formylglycine generating activity with a candidate agent,

(b) measuring C_α-formylglycine generating activity of the molecule, and

(c) comparing the measured C_α-formylglycine generating activity of the molecule to a control to determine whether the candidate agent modulates C_α-formylglycine generating activity of the molecule,

wherein the molecule is a nucleic acid molecule having a nucleotide sequence as the one set forth as SEQ ID NO:1, or an expression product thereof.

33. A method of diagnosing Multiple Sulfatase Deficiency in a subject, said method comprising:

(a) contacting a biological sample from a subject suspected of having Multiple Sulfatase Deficiency with an agent, said agent specifically binding to a molecule selected from the group consisting of: (i) a nucleic acid molecule having a nucleotide sequence as the one set forth as SEQ ID NO:1, (ii) an expression product of the nucleic acid molecule of (i), or (iii) a fragment of the expression product of (ii); and

b) measuring the amount of bound agent and determining therefrom if the expression of said nucleic acid molecule or of an expression product thereof is aberrant, aberrant expression being diagnostic of the Multiple Sulfatase Deficiency in the subject.

34. A method of diagnosing a condition characterized by aberrant expression of a nucleic acid molecule or an expression product thereof, said method comprising:

a) contacting a biological sample from a subject with an agent, wherein said agent specifically binds to said nucleic acid molecule, an expression product thereof, or a fragment of an expression product thereof; and

b) measuring the amount of bound agent and determining therefrom if the expression of said nucleic acid molecule or of an expression product thereof is aberrant, aberrant expression being diagnostic of the condition;

wherein the nucleic acid molecule has a nucleotide sequence as the one set forth as SEQ ID NO:1 and the condition is Multiple Sulfatase Deficiency.

35. A method for determining Multiple Sulfatase Deficiency in a subject characterized by aberrant expression of a nucleic acid molecule or an expression product thereof, comprising:

monitoring a sample from a patient for a parameter selected from the group consisting of

(i) a nucleic acid molecule having a nucleotide sequence as the one set forth as SEQ ID NO:1,

(ii) a polypeptide encoded by the nucleic acid molecule,

(iii) a peptide derived from the polypeptide, and

(iv) an antibody which selectively binds the polypeptide or peptide,

as a determination of Multiple Sulfatase Deficiency in the subject.

36. The method of claim 35, wherein the sample is a biological fluid or a tissue.

37. The method of claim 35, wherein the step of monitoring comprises contacting the sample with a detectable agent selected from the group consisting of

(a) an isolated nucleic acid molecule which selectively hybridizes under stringent conditions to the nucleic acid molecule of (i),

(b) an antibody which selectively binds the polypeptide of (ii), or the peptide of (iii), and

(c) a polypeptide or peptide which binds the antibody of (iv).

38. The method of claim 37, wherein the antibody, the polypeptide, the peptide or the nucleic acid is labeled with a radioactive label or an enzyme.

39. The method of claim 35, comprising assaying the sample for the peptide.

40. A kit, comprising a package containing:

an agent that selectively binds to the isolated nucleic acid of claim 1 or an expression product thereof, and

a control for comparing to a measured value of binding of said agent to said isolated nucleic acid of claim 1 or expression product thereof.

41. The kit of claim 40, wherein the control is a predetermined value for comparing to the measured value.

42. The kit of claim 40, wherein the control comprises an epitope of the expression product of the nucleic acid of claim 1.

43. The kit of claim 40, further comprising a second agent that selectively binds to a polypeptide selected from the group consisting of Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, or a peptide thereof, and

a control for comparing to a measured value of binding of said second agent to said polypeptide or peptide thereof.

44. Use of an agent that modulates C_{α}-formylglycine generating activity in the manufacture of a preparation for treating Multiple Sulfatase Deficiency.

45. Use of claim 44, wherein an agent selected from the group consisting of a nucleic acid molecule encoding Iduronate 2-Sulfatse, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, or HSulf-6, an expression product of the nucleic acid molecule, and a fragment of the expression product of the nucleic acid molecule is co-administrable with said preparation.

46. Use of claim 44, wherein the agent that modulates C_{α}-formylglycine generating activity is a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77 and 80-87.

47. Use of claim 44, wherein the agent that modulates C_α-formylglycine generating activity is a peptide as claimed in Claims 11-15, 19, 20 or a peptide having a sequence selected from the group consisting of SEQ ID NO: 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

48. Use of claim 44, wherein the agent that modulates C_α-formylglycine generating activity is produced by a cell expressing an FGE nucleic acid molecule as claimed in Claims 1-8, or an FGE nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77 and 80-87.

49. Use of claim 48, wherein the cell expressing an FGE nucleic acid molecule expresses an exogenous FGE nucleic acid molecule.

50. Use of claim 48, wherein the cell expressing an FGE nucleic acid molecule expresses an endogenous FGE nucleic acid molecule.

51. A method for increasing C_α-formylglycine generating activity in a subject, comprising:

administering an isolated FGE nucleic acid molecule of the invention or an expression product thereof to a subject, in an amount effective to increase C_α-formylglycine generating activity in the subject.

52. Use of an agent that modulates C_α-formylglycine generating activity in the manufacture of a preparation for treating a subject with Multiple Sulfatase Deficiency.

53. Use of claim 52, wherein the agent that modulates C_α-formylglycine generating activity is a sense nucleic acid as claimed in Claims 1-8, or an FGE nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77 and 80-87.

54. Use of claim 52, wherein the agent that modulates C_α-formylglycine generating activity is an isolated polypeptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO: 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

55. A method of increasing C_α-formylglycine generating activity in a cell, comprising:

contacting the cell with an isolated nucleic acid molecule of claim 1 or an expression product thereof, in an amount effective to increase C_α-formylglycine generating activity in the cell.

56. A pharmaceutical composition, comprising:

an agent comprising an isolated nucleic acid molecule as claimed in any one of Claims 1-8, an FGE nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77 and 80-87, or an expression product thereof, in a pharmaceutically effective amount to treat Multiple Sulfatase Deficiency, and

a pharmaceutically acceptable carrier.

57. A method for identifying a candidate agent useful in the treatment of Multiple Sulfatase Deficiency, comprising:

determining expression of a set of nucleic acid molecules in a cell or tissue under conditions which, in the absence of a candidate agent, permit a first amount of expression of the set of nucleic acid molecules, wherein the set of nucleic acid molecules comprises at least one nucleic acid molecule selected from the group consisting of

(a) nucleic acid molecules which hybridize under stringent conditions to a molecule consisting of a nucleotide sequence set forth as SEQ ID NO: 1 and which code for a polypeptide having C_α-formylglycine generating activity (FGE),

(b) nucleic acid molecules that differ from the nucleic acid molecules of (a) or (b) in codon sequence due to the degeneracy of the genetic code,

(c) a nucleic acid molecule having a sequence selected from the group consisting of SEQ NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87, and

(d) complements of (a) or (b) or (c),

contacting the cell or tissue with the candidate agent, and

detecting a test amount of expression of the set of nucleic acid molecules, wherein an increase in the test amount of expression in the presence of the candidate agent relative to the first amount of expression indicates that the candidate agent is useful in the treatment of the Multiple Sulfatase Deficiency.

58. A solid-phase nucleic acid molecule array consisting essentially of a set of nucleic acid molecules, expression products thereof, or fragments thereof, each nucleic acid molecule encoding for a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6, fixed to a solid substrate.

59. The solid-phase nucleic acid molecule array of claim 58, further comprising at least one control nucleic acid molecule.

60. The solid-phase nucleic acid molecule array of claim 58, wherein the set of nucleic acid molecules comprises at least one nucleic acid molecule encoding for a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6.

61. The solid-phase nucleic acid molecule array of claim 58, wherein the set of nucleic acid molecules comprises at least two nucleic acid molecules, each nucleic acid molecule encoding for a polypeptide selected from the group consisting of SEQ ID NO. 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78, Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6.

62. A method for increasing sulfatase activity in a cell, comprising:

contacting a cell expressing a sulfatase with an isolated nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87, or an expression product thereof, in an amount effective to increase sulfatase activity in the cell.

63. The method of claim 62, wherein the cell expresses endogenous sulfatase.

64. The method of claim 62, wherein the cell expresses exogenous sulfatase.

65. The method of claim 63, wherein the endogenous sulfatase is activated.

66. The method according to any one of claims 62-66, wherein the sulfatase is selected from the group consisting of Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6.

67. The method of claim 62, wherein the cell is a mammalian cell.

68. A pharmaceutical composition, comprising:

a sulfatase that is produced by cell, in a pharmaceutically effective amount to treat a sulfatase deficiency, and

a pharmaceutically acceptable carrier,

wherein said cell has been contacted with an agent comprising an isolated nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87), or an expression product thereof .

69. An isolated variant allele of a human FGE gene, which encodes a variant FGE polypeptide, comprising:

an amino acid sequence comprising at least one variation in SEQ ID NO:2, wherein the at least one variation comprises: Met1Arg; Met1Val; Ser155Pro; Cys218Tyr; Ala279Val;

Arg327Stop; Cys336Arg; Arg345Cys; Arg349Trp; Arg349Trp; Arg349Gln; Ser359Stop; or a combination thereof.

70. An isolated variant human FGE polypeptide, comprising:

an amino acid sequence comprising at least one variation in SEQ ID NO:2, wherein the at least one variation comprises: Met1Arg; Met1Val; Ser155Pro; Cys218Tyr; Ala279Val; Arg327Stop; Cys336Arg; Arg345Cys; Arg349Trp; Arg349Trp; Arg349Gln; Ser359Stop; or a combination thereof.

71. An antibody having the variant human FGE polypeptide of claim 69 as an immunogen.

72. The antibody or claim 71, which is a polyclonal antibody.

73. The antibody of claim 71, which is a monoclonal antibody.

74. The antibody of claim 71, which is a chimeric antibody.

75. The antibody of claim 71, detectably labeled.

76. The antibody of claim 75, wherein said detectable label comprises a radioactive element, a chemical which fluoresces, or an enzyme.

77. A sulfatase-producing cell wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased, the cell comprising:

(i) a sulfatase with an increased expression, and

(ii) a Formylglycine Generating Enzyme with an increased expression,

wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 5% over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formyglycine Generating Enzyme.

78. The cell of claim 77, wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 10% over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme.

79. The cell of claim 77, wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 20% over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme.

80. The cell of claim 77, wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 50% over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme.

81. The cell of claim 77, wherein the ratio of active sulfatase to total sulfatase produced by the cell is increased by at least 100% over the ratio of active sulfatase to total sulfatase produced by the cell in the absence of the Formylglycine Generating Enzyme.

82. Use of a sulfatase contacted with a Formylglycine Generating Enzyme in the manufacture of a preparation for treating a sulfatase deficiency with a sulfatase in a subject.

83. Use of claim 82, wherein the sulfatase is selected from the group consisting of Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6.

84. Use of claim 82, wherein the Formylglycine Generating Enzyme is encoded by a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87.

85. Use of claim 82, wherein the Formylglycine Generating Enzyme is a peptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO: 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76,

and 78.

86. Use of an isolated FGE nucleic acid molecule of the invention or an expression product thereof in the manufacture of a preparation for increasing C_α-formylglycine generating activity in a subject.

87. Use of an isolated nucleic acid molecule of claim 1 or an expression product thereof in the manufacture of a preparation for increasing C_α-formylglycine generating activity in a cell.

88. A substance or composition for use in a method for treating Multiple Sulfatase Deficiency in a subject, said substance or composition comprising an agent that modulates C_α-formylglycine generating activity and said method comprising administering to a subject in need of such treatment said substance or composition in an amount effective to treat Multiple Sulfatase Deficiency in the subject.

89. A substance or composition for use in a method of treatment of claim 88, further comprising co-administering an agent selected from the group consisting of a nucleic acid molecule encoding Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, or HSulf-6, an expression product of the nucleic acid molecule, and a fragment of the expression product of the nucleic acid molecule.

90. A substance or composition for use in a method of treatment of claim 88, wherein the agent that modulates C_α-formylglycine generating activity is a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87.

91. A substance or composition for use in a method of treatment of claim 88, wherein the agent that modulates C_α-formylglycine generating activity is a peptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO: 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

92. A substance or composition for use in a method of treatment of claim 88, wherein the agent that modulates C_α-formylglycine generating activity is produced by a cell expressing an FGE nucleic acid molecule as claimed in Claims 1-8, or an FGE nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87.

93. A substance or composition for use in a method of treatment of claim 92, wherein the cell expressing an FGE nucleic acid molecule expresses an exogenous FGE nucleic acid molecule.

94. A substance or composition for use in a method of treatment of claim 92, wherein the cell expressing an FGE nucleic acid molecule expresses an endogenous FGE nucleic acid molecule.

95. A substance or composition for use in a method for increasing C_α-formylglycine generating activity in a subject, said substance or composition comprising an isolated FGE nucleic acid molecule of the invention or an expression product thereof, and said method comprising administering said substance or composition to a subject, in an amount effective to increase C_α-formylglycine generating activity in the subject.

96. A substance or composition for use in a method for treating a subject with Multiple Sulfatase Deficiency, said substance or composition comprising an agent that modulates C_α-formylglycine generating activity, and said method comprising administering to a subject in need of such treatment said substance or composition in an amount effective to increase C_α-formylglycine generating activity in the subject.

97. A substance or composition for use in a method of treatment of claim 96, wherein the agent that modulates C_α-formylglycine generating activity is a sense nucleic acid as claimed in Claims 1-8, or an FGE nucleic acid molecule having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87.

98. A substance or composition for use in a method of treatment of claim 96, wherein the agent that modulates C_α-formylglycine generating activity is an isolated polypeptide as claimed in Claims 11-15,19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO: 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

99. A substance or composition for use in a method for increasing C_α-formylglycine generating activity in a cell, said substance or composition comprising an isolated nucleic acid molecule of claim 1 or an expression product thereof, and said method comprising contacting the cell with said substance or composition in an amount effective to increase C_α-formylglycine generating activity in the cell.

100. A substance or composition for use in a method for treating a sulfatase deficiency with a sulfatase, said substance or composition comprising a sulfatase contacted with a Formyglycine Generating Enzyme, and said method comprising administering to a subject in need of such treatment said substance or composition in an amount effective to increase the specific activity of the sulfatase.

101. A substance or composition for use in a method of treatment of claim 100, wherein the sulfatase is selected from the group consisting of Iduronate 2-Sulfatase, Sulfamidase, N-Acetylgalactosamine 6-Sulfatase, N-Acetylglucosamine 6-Sulfatase, Arylsulfatase A, Arylsulfatase B, Arylsulfatase C, Arylsulfatase D, Arylsulfatase E, Arylsulfatase F, Arylsulfatase G, HSulf-1, HSulf-2, HSulf-3, HSulf-4, HSulf-5, and HSulf-6.

102. A substance or composition for use in a method of treatment of claim 100, wherein the Formylglycine Generating Enzyme is encoded by a nucleic acid molecule as claimed in Claims 1-8, or a nucleic acid having a sequence selected from the group consisting of SEQ ID NO: 1, 3, 4, 45, 47, 49, 51, 53, 55, 57, 59, 61, 63, 65, 67, 69, 71, 73, 75, 77, and 80-87.

103. A substance or composition for use in a method of treatment of claim 100, wherein the Formylglycine Generating Enzyme is a peptide as claimed in Claims 11-15, 19, 20, or a peptide having a sequence selected from the group consisting of SEQ ID NO: 2, 5, 46, 48, 50, 52, 54, 56, 58, 60, 62, 64, 66, 68, 70, 72, 74, 76, and 78.

104. A nucleic acid molecule of any one of claims 1 to 6, substantially as herein described and illustrated.

105. A vector of claim 7 or claim 8, substantially as herein described and illustrated.

106. A host cell of claim 9 or claim 10, substantially as herein described and illustrated.

107. A polypeptide of any one of claims 11 to 20 or 70, substantially as herein described and illustrated.

108. A method of any one of claims 21 to 31, substantially as herein described and illustrated.

109. A method of claims 32, substantially as herein described and illustrated.

110. A method of any one of claims 33 to 39, substantially as herein described and illustrated.

111. A kit of any one of claims 40 to 43, substantially as herein described and illustrated.

112. Use of any one of claims 44 to 50, 52 to 54 or 82 to 87, substantially as herein described and illustrated.

113. A method of claim 51, substantially as herein described and illustrated.

114. A method of claim 55, substantially as herein described and illustrated.

115. A composition of claim 56 or claim 68, substantially as herein described and illustrated.

116. A method or claim 57, substantially as herein described and illustrated.

117. A nucleic acid molecule array of any one of claims 58 to 61, substantially as herein described and illustrated.

118. A method of any one of claims 62 to 67, substantially as herein described and illustrated.

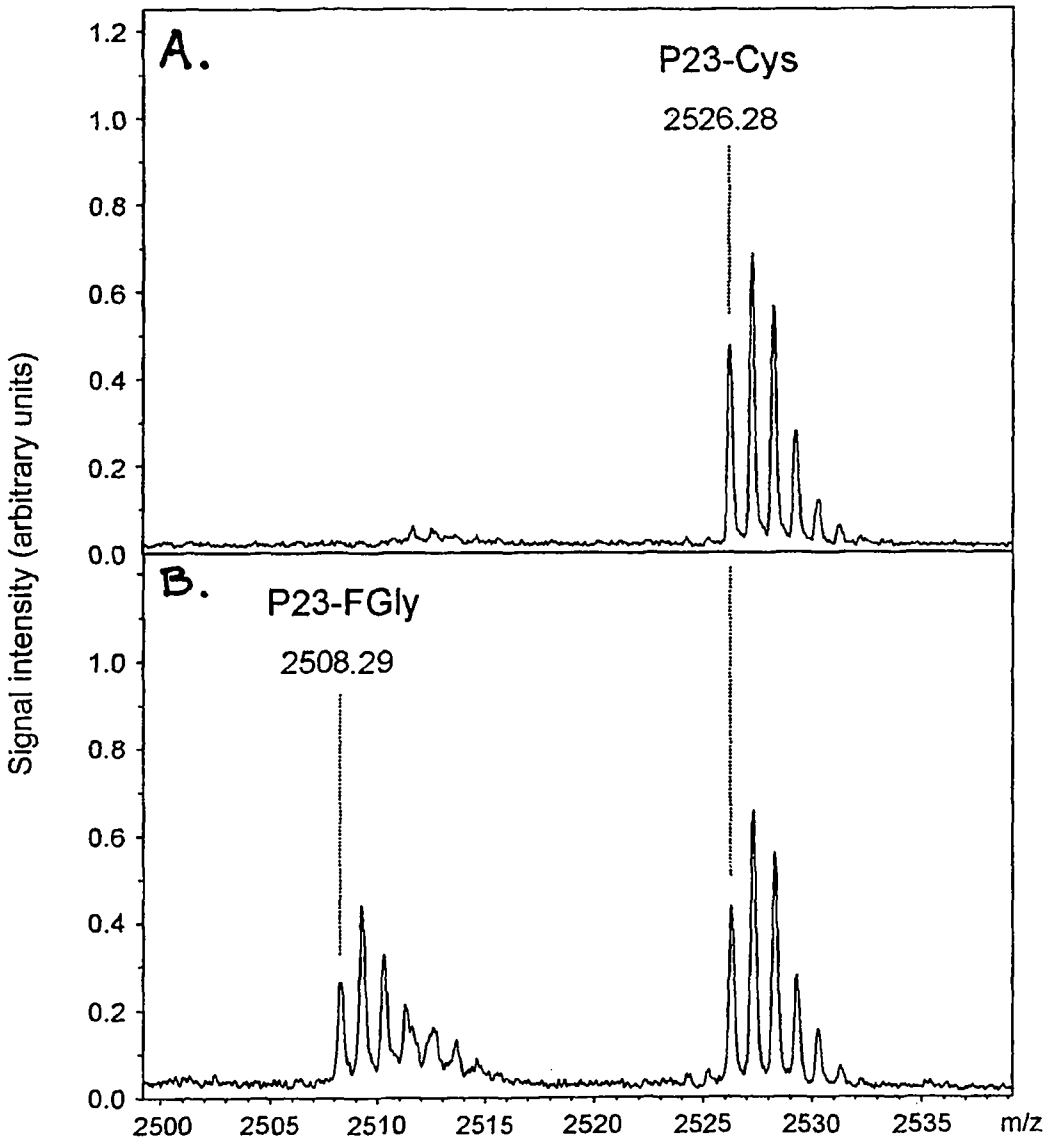
119. A variant allele of claim 69, substantially as herein described and illustrated.

120. An antibody of any one of claims 71 to 76, substantially as herein described and illustrated.

121. A cell of any one of claims 77 to 85, substantially as herein described and illustrated.

122. A substance or composition for use in a method of treatment of any one of claims 88 to 103, substantially as herein described and illustrated.

123. A new nucleic acid molecule, a new vector, a new host cell, a new polypeptide, a new in vitro method for determining the level of FGE expression in a subject, a new method for identifying an agent, a new in vitro method of diagnosis, a new in vitro method for determining a condition in a subject, a new kit, a new use of an agent that modulates C_α-formylglycine generating activity, a new use of an isolated FGE nucleic acid molecule of the invention or an expression product thereof, a new non-therapeutic method of treatment, a new nucleic acid molecule array, a new composition, a new variant allele, a new antibody, a new cell, a new use of an isolated nucleic acid molecule of claim 1 or an expression product thereof, a new use of a sulfatase contacted with a Formylglycine Generating Enzyme, or a substance or composition for a new use in a method of treatment, substantially as herein described and illustrated.


A.
P23-Cys
2526.28
B.
P23-FGly
2508.29
Signal intensity (arbitrary units)
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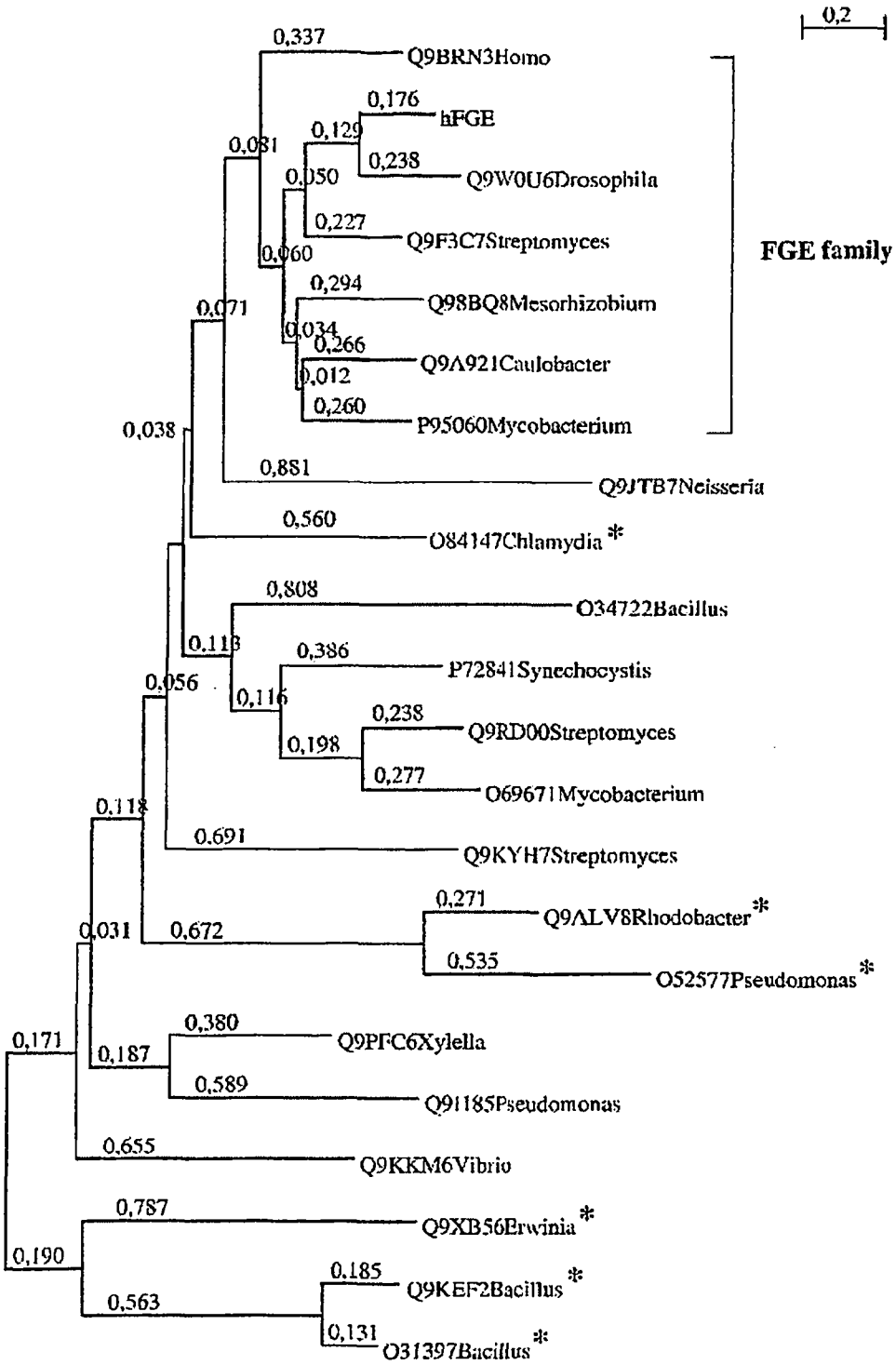
1.0
0.8
0.6
0.4
0.2
0.0
2500
2505
2510
2515
2520
2525
2530
2535
m/z

Fig. 1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0,238
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0,227
Q9F3C7Streptomyces
0,060
FGE family
0,294
Q98BQ8Mesorhizobium
0,071
0,034
0,266
Q9A921Caulobacter
0,012
0,260
P95060Mycobacterium
0,038
0,881
Q9JTB7Neisseria
0,560
O84147Chlamydia*
0,808
O34722Bacillus
0,113
0,386
P72841Synechocystis
0,056
0,116
0,238
Q9RD00Streptomyces
0,198
0,277
O69671Mycobacterium
0,118
0,691
Q9KYH7Streptomyces
0,271
Q9ALV8Rhodobacter*
0,031
0,672
0,535
O52577Pseudomonas*
0,171
0,380
Q9PFC6Xylella
0,187
0,589
Q91185Pseudomonas
0,655
Q9KKM6Vibrio
0,787
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0,185
Q9KEF2Bacillus*
0,563
0,131
O31397Bacillus*

Fig. 2

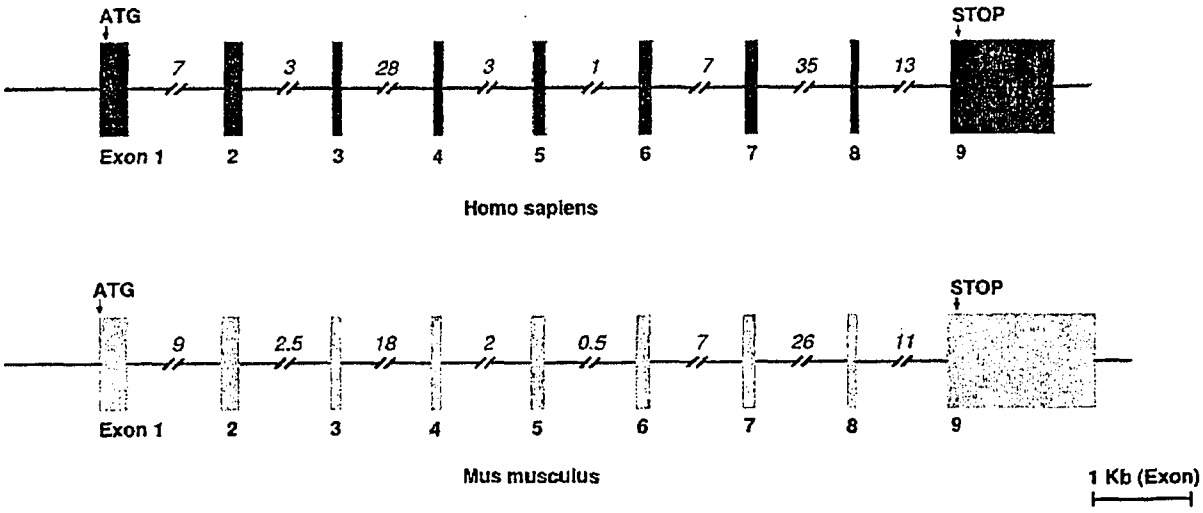
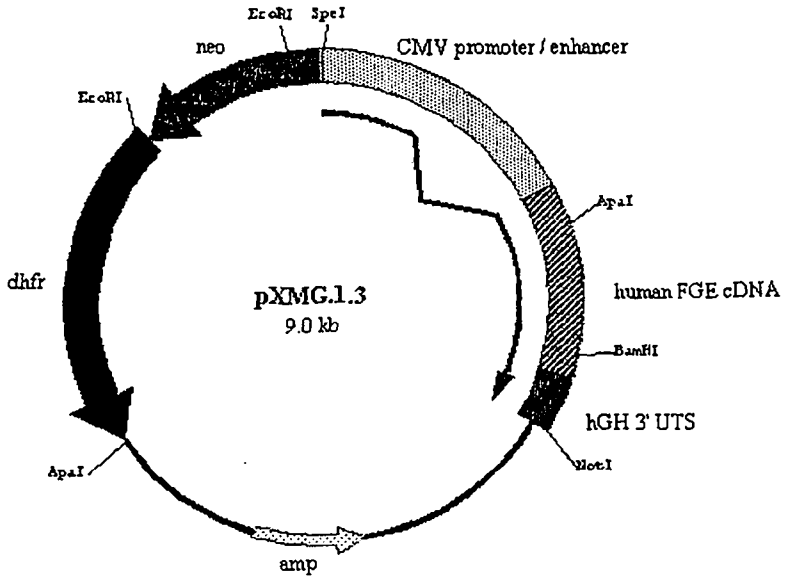

ATG
STOP
7
3
28
3
1
7
35
13
Exon 1
2
3
4
5
6
7
8
9
Homo sapiens
ATG
STOP
9
2.5
18
2
0.5
7
26
11
Exon 1
2
3
4
5
6
7
8
9
Mus musculus
1 Kb (Exon)

Fig. 3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SpeI
neo
CMV promoter / enhancer
EcoRI
ApaI
dhfr
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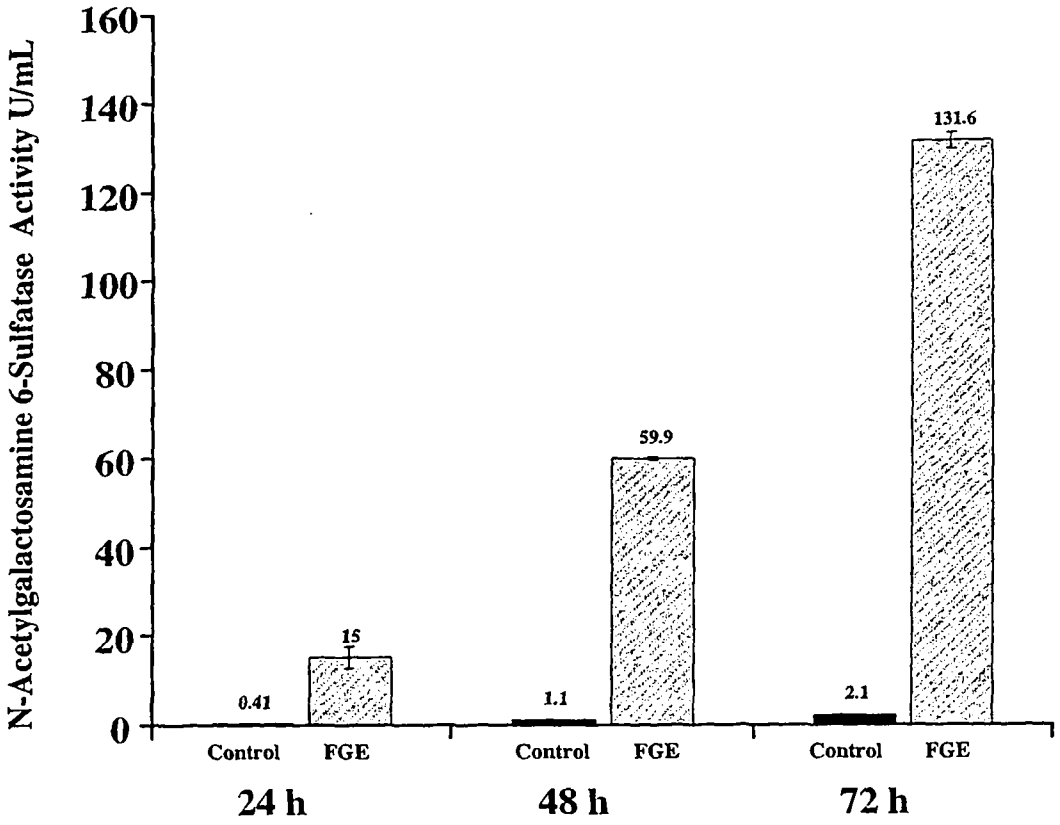
9.0 kb
human FGE cDNA
BamHI
hGH 3' UTS
NotI
ApaI
amp

Fig. 4


N-Acetylgalactosamine 6-Sulfatase Activity U/mL
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140
120
100
80
60
40
20
0
0.41
15
1.1
59.9
2.1
131.6
Control
FGE
Control
FGE
Control
FGE
24 h
48 h
72 h
Post Transfection Accumulation
(Hours)

Fig. 5

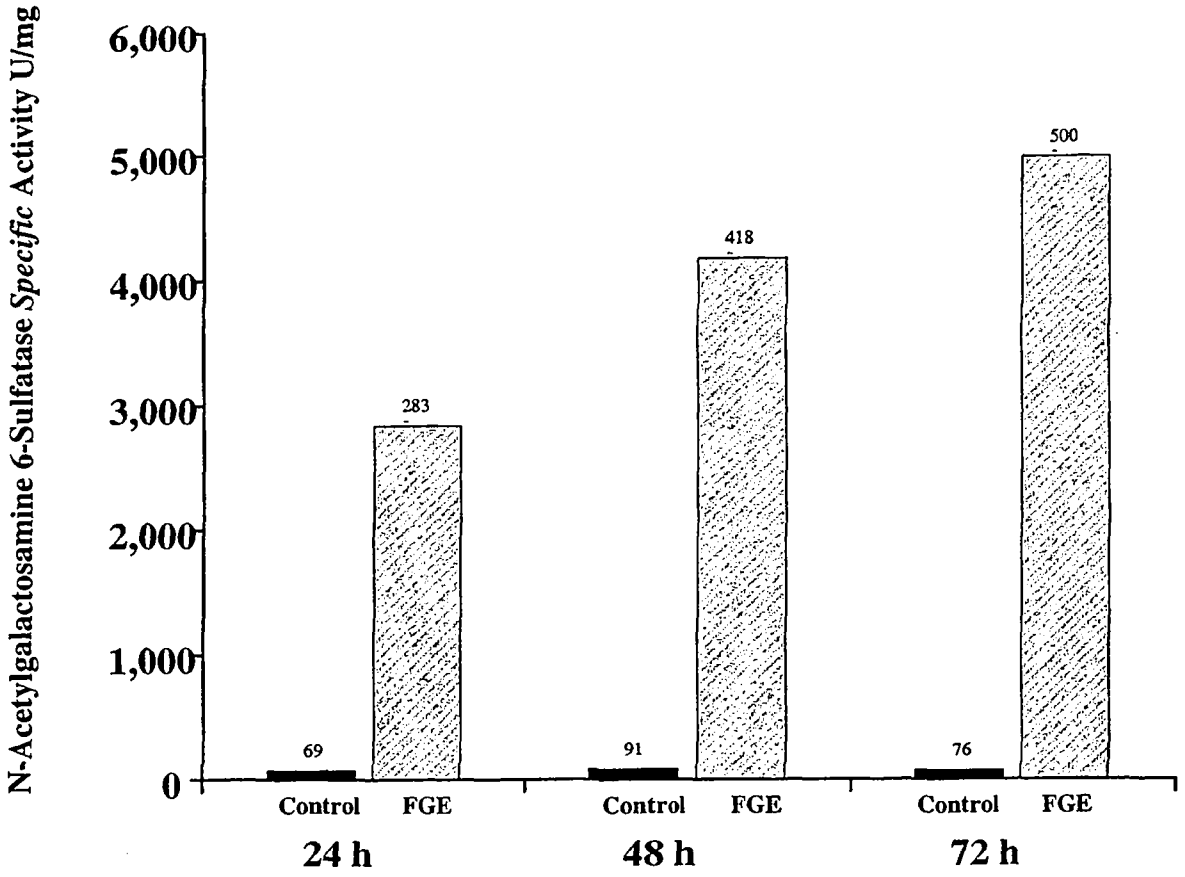
N-Acetylgalactosamine 6-Sulfatase *Specific* Activity U/mg
6,000
5,000
4,000
3,000
2,000
1,000
0
69
283
91
418
76
500
Control
FGE
Control
FGE
Control
FGE
24 h
48 h
72 h
Post Transfection Accumulation (Hours)

Fig. 6

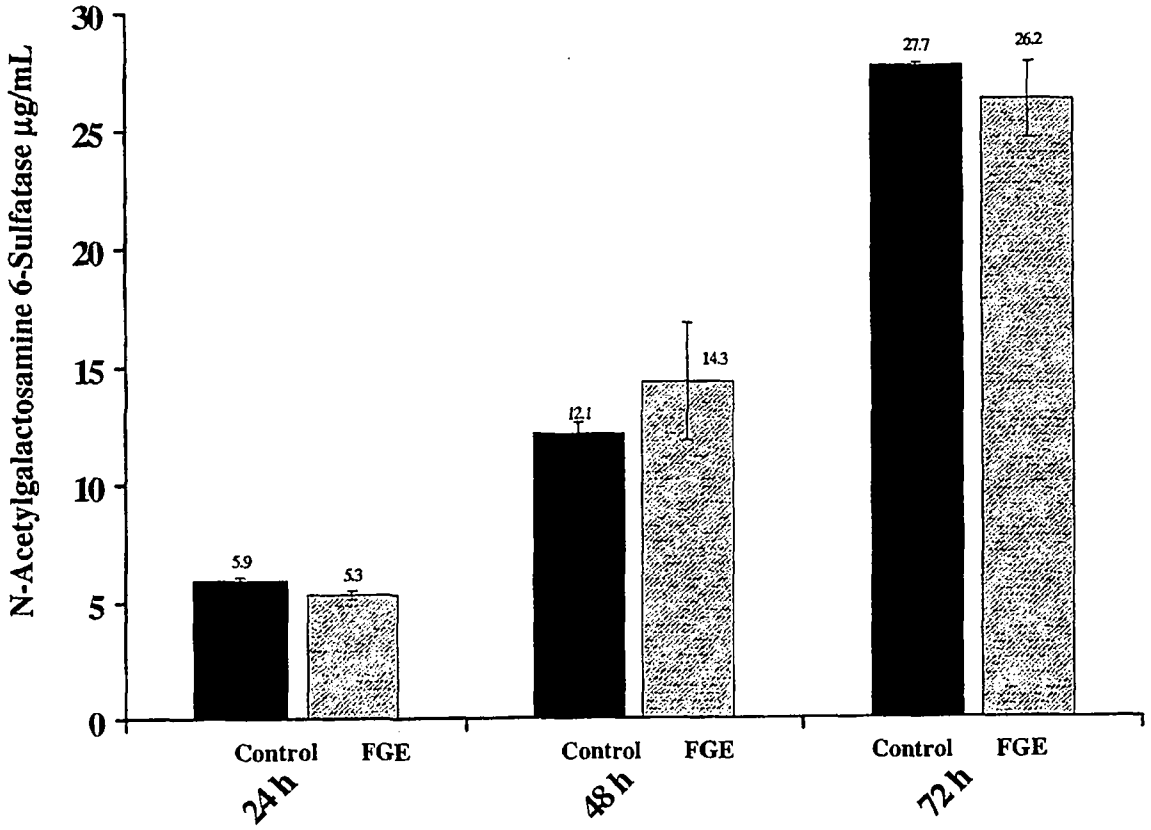
N-Acetylgalactosamine 6-Sulfatase µg/mL
30
25
20
15
10
5
0
5.9
5.3
12.1
14.3
27.7
26.2
Control
FGE
Control
FGE
Control
FGE
24 h
48 h
72 h
Post Transfection Accumulation
(Hours)

Fig. 7

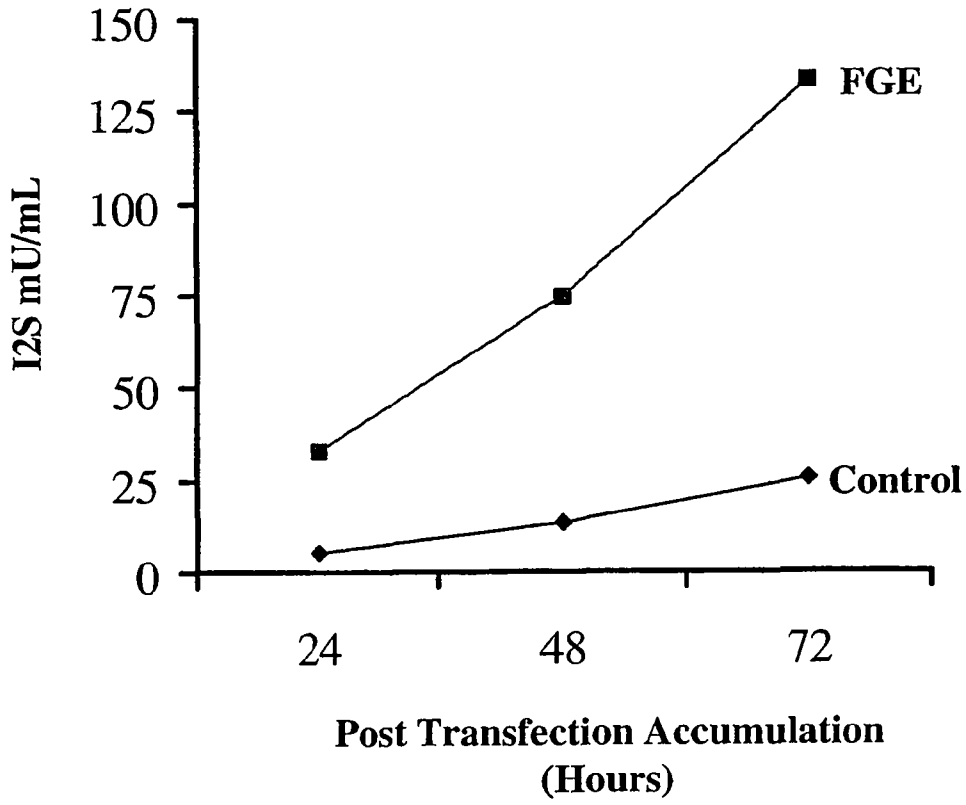
150
125
100
75
50
25
0
I2S mU/mL
FGE
Control
24
48
72
Post Transfection Accumulation
(Hours)

Fig. 8

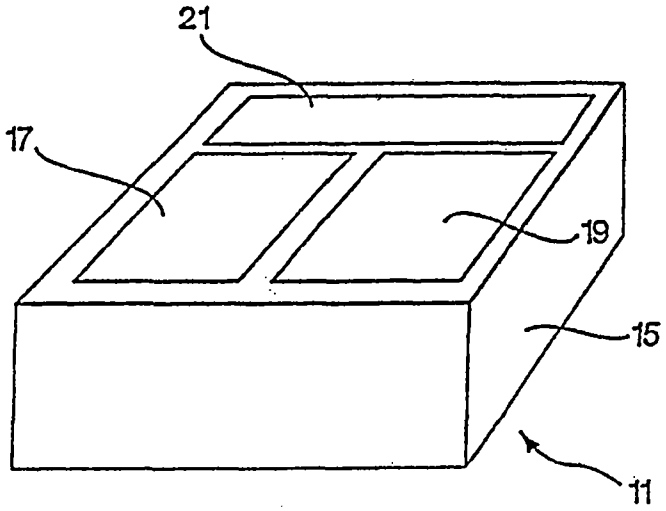

Fig. 9